Role of LncRNA-AF085935, IL-10 and IL-17 in Rheumatoid Arthritis Patients With Chronic Hepatitis C

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Abstract

Background: The current study aimed at testing the effect of corticosteroid therapy on serum levels of interleukin-10 (IL-10) and IL-17 as well as LncRNA-AF085935 in patients of rheumatoid arthritis (RA) associated with hepatitis C virus (HCV) and evaluating the usefulness of using these parameters to predict the therapeutic efficacy of steroids in these patients.

Methods: Thirty healthy control subjects and 65 chronic HCV patients with RA were included in our study. Patients were subjected to clinical examination, abdominal ultrasound, and liver biopsy and received 6-methyl-prednisolone (PDN) 16 mg/day for 48 weeks. Blood samples were collected from all subjects and serum was separated to assess IL-10 and IL-17 by ELISA and HCV RNA and LncRNA-AF085935 by qRT-PCR.

Results: Our study revealed that there were significant increases in serum levels of IL-10, IL-17 and LncRNA-AF085935 in RA patients associated with HCV compared with healthy control subjects. Also there were significant increases in serum levels of IL-10 and HCV RNA and a significant decrease in serum level of IL-17 in patients after corticosteroid therapy, while LncRNA-AF085935 is not significantly changed.

Conclusion: LncRNA-AF085935 might be a useful candidate biomarker for the early detection of RA associated with HCV, providing potential new strategies for early screening and therapy of these patients. IL-17 is a non-invasive prognostic marker to predict the efficacy of corticosteroid therapy in RA patients associated with chronic hepatitis C.

Keywords: Rheumatoid arthritis; HCV; IL-10; IL-17; LncRNA-AF085935

Introduction

Hepatitis C virus (HCV) is a hepatotropic non-cytopathic positive strand RNA virus, which is considered the main cause of chronic hepatitis, cirrhosis, and hepatocellular carcinoma (HCC), affecting over 150 million people worldwide. In Egypt, the condition is even worse as just about every family is touched by hepatitis C. It kills an estimated 40,000 Egyptian per year where at least 1 in every 10 of the population aged 15 - 59 is infected [1]. Egypt has the top ranks in prevalence of HCV globally (15%) [2].

HCV infection is more than just a liver disease; several extrahepatic manifestations have been associated with chronic HCV infection. These include numerous rheumatic, hematologic, renal, dermatologic and autoimmune manifestations [3-5]. Rheumatic manifestations were found in 31% subjects infected with HCV in an Israeli study [6]. Rheumatic manifestations associated with HCV infection include arthralgia, myalgia, arthritis, vasculitis and fibromyalgia [6-9]. In addition, associations have been reported between HCV infection and other rheumatic and autoimmune diseases including systemic lupus, Sjogren’s syndrome and anti-phospholipid syndrome [8, 10].

As the prevalence of HCV infection is high in the general population, it is frequent to use corticosteroids in HCV-infected patients for different diseases which sometimes are associated to the virus itself. In chronic hepatitis C related autoimmune hepatitis, the clinician must face the challenge of treating with corticosteroids versus treating HCV with combined therapy. Interferon may worsen the autoimmune component [11, 12] and corticosteroids might increase HCV RNA and worsen the clinical evolution of chronic hepatitis C. However, the effect of steroids on chronic hepatitis C has not been fully determined, and there is not enough information to consider steroids as beneficial or harmful drugs on natural history of chronic hepatitis C [13].
Several articles have been published on this issue. Most of the reported studies were performed with low numbers of subjects using high-dose corticosteroids and seemed to reduce aspartate aminotransferase (AST) and alanine aminotransferase (ALT) at the expense of increasing HCV RNA [14, 15]. However, the effect of low-dose corticosteroids on histological activity and HCV RNA kinetics might be different [13].

Rheumatoid arthritis (RA) is a chronic, systemic, autoimmune disorder of unknown cause, which primarily affects the peripheral joints causing inflammation (arthritis) in a symmetric pattern [16]. Constitutional symptoms of the disease include fatigue, malaise and morning stiffness. Extra-articular involvement of organs such as the skin, heart, lungs and eyes can be significant. Moreover, RA causes joint destruction and thus often leads to considerable morbidity and mortality [17].

The worldwide incidence of RA is approximately three cases per 10,000 and the prevalence rate is approximately 1% of adults, but its prevalence rate in Egypt is approximately 0.2% of adults. Moreover, the frequency of RA increases with age and peaks in persons aged 35 - 50 years [18]. Egyptian RA patients were significantly younger, had more active disease, and were more functionally disabled. They showed more frequent use of NSAIDs, methotrexate and steroids when compared to non-Egyptians. Egyptian patients had the highest ever HCV prevalence [19].

Interleukin-10 (IL-10) is a multifunctional cytokine with potent immunoregulatory and anti-inflammatory properties. It prevents the release and function of a number of pro-inflammatory cytokines. Elevated serum levels of IL-10 have been observed in patients with untreated chronic HCV infection [20, 21]. IL-10 is also a T-helper (Th)-2-type cytokine, and it has been suggested that an imbalance in the Th1/Th2 immune response could influence the clinical outcome and disease progression [22].

Pro-inflammatory cytokines, TNF-α and IL-1 are overproduced by macrophages, synovial fibroblasts and neutrophils and play an important role in the process of chronic inflammation and joint destruction [23]. Synovial fluid from patients with RA also contains detectable levels of anti-inflammatory cytokine IL-10, but they are insufficient to counterbalance the effect of pro-inflammatory cytokines. The cellular sources of IL-10 in the synovial tissue are macrophages and T cells [24].

RA patients showed significantly reduced IL-10 levels in serum in comparison to healthy donors, suggesting that IL-10 synthesis is depressed in RA [25]. In addition, patients with advanced RA had the lowest values of IL-10 in serum [26]. On the contrary, some studies have shown that concentration of IL-10 in sera of patients with RA is higher as compared to controls [27, 28].

IL-17A (IL-17) is one of T cell-derived cytokines produced by Th17 cells [29]. IL-17 is a powerful chemoattractant for neutrophils and has been reported to be involved in many immune processes, most notably in inducing and mediating pro-inflammatory responses, e.g. several autoimmune diseases, allergic diseases, asthma and pulmonary infection [30-32].

Compared with controls, serum IL-17 levels were significantly higher in chronically HCV infected patients [33]. This result goes with what has been reported previously that serum IL-17 level was increased in liver injuries following chronic hepatitis and cirrhosis supporting a role for IL-17 as a chronic disease inducer in the pathogenesis and/or progression of liver fibrosis [34].

Among the complex network of inflammatory cells involved in the pathogenesis of RA, Th17 cells have recently been identified as key cells in the promotion of autoimmune processes, joint destruction and angiogenesis [35]. The hallmark of the Th17 subset is the production of IL-17A and IL-17F, which share strong homology [36]. Therefore, IL-17A and IL-17F may play an important role in T cell-triggered inflammation by upregulating some of the gene products involved in cell activation, proliferation and growth, and it is an important inductor of various cytokines and chemokines that are crucial in regulating the inflammatory response [37].

LncRNAs were a class of newfound non-coding RNAs, shorter than 200 nucleotides in length and were widely depicted in the genome currently [38]. As the function of most lncRNAs remains largely unknown, through our study, we investigated the correlation between lncRNA-AF085935 expression level and the RA HCV sufferers with corticosteroid therapy.

**Patients and Methods**

**Total enrolment**

Thirty control healthy subjects and 65 chronic HCV patients with RA were recruited from outpatient clinic at rheumatology and tropical departments, faculty of medicine according to the following inclusion and exclusion criteria.

Inclusion criteria were: 1) adult male and female patients aged 38 - 40 years old; 2) chronic HCV infection for > 6 months with detectable serum quantitative HCV RNA and serum ALT above the laboratory defined upper limit of normal; 3) positive anti-CCP; 4) compensated liver disease and albumin within normal limits; 5) liver biopsy confirming a histological diagnosis consistent with HCV necroinflammatory score (META VIR) ≥ 1, and fibrosis score ≥ 1.

Exclusion criteria were: 1) co-infection with HBV or HIV; 2) decompensated liver disease (Child-Pugh classification B or C); 3) autoimmune or metabolic liver disease; 4) renal or parathyroid disease; 5) patients with a value of alpha-fetoprotein > 100 ng/mL to exclude presence of HCC.

**Study design**

The study population consisted of 30 healthy control and 65 HCV patients. The study was approved by the institutional ethical committee and written informed consent was obtained from all patients. Sixty-five HCV patients were assigned to receive 6-methyl-prednisolone (PDN) 16 mg/day for 48 weeks [39]. All subjects were reviewed as outpatients at 4-week intervals and were monitored for 6 months after treatment.

Natural interferon-alpha versus its combination with PDN in the therapy of type II mixed cryoglobulinemia: a long-term,
randomized, controlled study.

**Blood samples collection**

Five milliliter blood sample was driven from each subject enrolled in study and divided into two parts. The first part was collected into vacationer EDTA tubes to assess biochemical parameters as follows: AST, ALT, total bilirubin (T. Bil), direct bilirubin (D. Bil), albumin, alkaline phosphatase (ALKP), creatinine, and random blood sugar (RBS) were detected using Roche Hitachi 911 Chemistry Analyzer (Bunker Lake Blvd. Greater Minneapolis, St. Paul Area, USA). Hemoglobin (Hb), total leucocytes count (TLC), absolute neutrophil counts (ANC), and platelets were detected by cell counter (Sysmex XT-4000i Automated Hematology Analyzer, Lincolnshire, IL, USA). Thyroid-stimulating hormone (TSH) and alpha-fetoprotein (AFP) were detected using ELISA kits (DRG International Inc., Springfield, NJ, USA). Hepatic fibrosis metavir score was used to evaluate liver inflammation A (0-3), fibrosis F (0-4) and size S (0-4). The second part was collected into another was used to evaluate liver inflammation A (0-3), fibrosis F (0-4) and size S (0-4). The second part was collected into another

**IL-10 and IL-17 levels assessment**

IL-10 and IL-17 serum levels were assessed using ELISA kit (DRG International Inc., Springfield, NJ, USA). All protocols followed the manufacturer’s instructions. Each ELISA test was performed in triplicate, with an illustrative standards curve done in the same run and the mean absorbance computed from the average for three wells normalized to a zero calibrator well.

**LncRNA-AF085935 expression**

Real-time qRT-PCR was used to detect LncRNA-AF085935 level by the SensiFAST™ SYBR® Hi-ROX One-Step Kit, catalog no. PI-50217 V, UK. Ten microliters RNA was prepared as templates for the reverse transcription reaction and the PCR reaction in one-step RT-PCR reaction. Samples were analyzed in duplicate, and non-template controls were included. The thermal cycling conditions were as follows: 50 °C for 15 min one cycle (for reverse transcriptase), 95 °C at 10 min for RT enzyme inactivation, then 45 cycles at 93 °C for 15 s, 54 °C for 30 s, and 72 °C for 30 s. β-actin was used as control. The sequence of LncRNA-AF085935 primers was forward 5'-CAGGGCAAGGTGTTTTTC-3' and reverse 5'-TTGTTGGTGCCTGATACC-3' (Genebank accession no. NG009286.1) and β-actin was 5'-forward 5'-GGCGGCACCACCATTGCTC-3' and reverse 5'-AGG GGCCGACTCGTCTACT-3' (Genebank accession no. NM001101.3).

The amplification specific was confirmed by melting curve analysis. The relative level of LncRNA was normalized to β-actin and calculated by using the -ΔCt method.

**Statistical analysis**

Data were collected and coded to facilitate data manipulation and double entered into Microsoft Access and data analysis was performed using SPSS software version 18 under windows 7. Simple descriptive analysis was in the form of numbers and percentages for qualitative data, and arithmetic means as central tendency measurement, standard deviations as measure of dispersion for quantitative parametric data, and inferential statistic test. Independent Student’s t-test was used to compare measures of two independent groups of quantitative data. Paired t-test was used to compare two dependent quantitative data. Chi-square test was used to compare two of more than two qualitative groups. The level P ≤ 0.05 was considered the cut-off value for significance.

**Results**

There is no statistically significant difference with P-value > 0.05 between cases and controls as regards age, sex and BMI, which indicates proper matching between two study groups (Table 1). There is statistically significant difference with P-value < 0.05 between cases and controls as regards liver biopsy and ultrasound findings were illustrated in Figures 1 and 2. There is statistically significant difference with P-value < 0.05 between cases before and after treatment as regards AST, ALT, total bilirubin, ALKP, PC, AFP, Hb, TLC, and platelets with high mean of ALT, AST, AFP, Hb, TLC, and platelets and low mean of total bilirubin, ALKP and PC among cases. On the other hand, there is no statistically significant difference with P-value > 0.05 as regards level of direct bilirubin, albumin, creatinine, HbA1c, TSH, RBS, and INR level (Table 2).

**Table 1.** Demographic Characters in Different Study Groups

| Variables       | Cases (n = 65), mean ± SD | Control (n = 30), mean ± SD | P-value |
|-----------------|---------------------------|----------------------------|---------|
| Age (years)     | 39.4 ± 11.7               | 41.5 ± 8.4                 | 0.4     |
| BMI             | 27.6 ± 2.1                | 26.1 ± 3                   | 0.1     |
| Sex, no. (%)    |                           |                            |         |
| Male            | 47 (72.3%)                | 19 (63.3%)                 | 0.5     |
| Female          | 18 (27.7%)                | 11 (36.7%)                 |         |

Percentage of liver biopsy and ultrasound findings were illustrated in Figures 1 and 2. There is statistically significant difference with P-value < 0.05 between cases before and after treatment as regards HCV RNA with high mean after treatment. There is statistically significant difference with P-value < 0.05 between cases before and after treatment and controls as regards level of IL-10 and
IL-17. There is statistically significant difference with P-value < 0.05 between cases before treatment and controls and between cases after treatment and controls as regards lncRNA-AF085935 with high mean among cases. On the other hand, there is no statistically significant difference with P-value > 0.05 as regards lncRNA-AF085935 before and after treatment (Table 3).

There is no statistically significant correlation with P-value > 0.05 between HCV RNA level and all parameters before and after treatment (Table 4).

This table shows that IL-17 after treatment had the highest sensitivity, specificity, and accuracy in predicting the efficacy of steroid therapy in chronic HCV patients with RA (Table 5 and Figs. 3, 4 and 5).

**Discussion**

RA is a chronic inflammatory disorder in which the articular joints are gradually destroyed [40].

The relationship between HCV and rheumatic diseases is complex and represents both a diagnostic and therapeutic problem to rheumatologists [41]. Management of RA is complicated with the highly significant difference regarding

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**Table 2. Laboratory Investigations in Different Studied Groups**

| Variables     | Cases (n = 65), mean ± SD | Control (n = 30), mean ± SD | P-value |
|---------------|---------------------------|-----------------------------|---------|
| AST           | 45.1 ± 14.6               | 27.1 ± 7.5                  | < 0.001 |
| ALT           | 52.7 ± 10.6               | 26.3 ± 7.8                  | < 0.001 |
| Total bilirubin| 0.74 ± 0.16               | 0.96 ± 0.21                 | < 0.001 |
| Direct bilirubin| 0.23 ± 0.11               | 0.26 ± 0.14                 | 0.2     |
| Albumin       | 4.1 ± 0.48                | 4.3 ± 0.59                  | 0.06    |
| Alkaline phosphate| 68.9 ± 24.6               | 81.9 ± 28.2                 | 0.02    |
| PC            | 0.88 ± 0.07               | 0.91 ± 0.06                 | 0.02    |
| INR           | 1.08 ± 0.10               | 1.1 ± 0.12                  | 0.3     |
| Creatinine    | 0.93 ± 0.29               | 0.93 ± 0.29                 | 0.9     |
| HbA1c         | 5.4 ± 0.78                | 5.33 ± 0.94                 | 0.8     |
| TSH           | 1.63 ± 0.98               | 1.89 ± 1.5                  | 0.3     |
| AFP           | 13.2 ± 6.3                | 3.33 ± 2.1                  | < 0.001 |
| RBS           | 87.6 ± 23.3               | 95.7 ± 26.2                 | 0.1     |
| Hb (g)        | 13.2 ± 2.3                | 10.8 ± 1.1                  | < 0.001 |
| TLC (× 1,000) | 6.4 ± 1.9                 | 4.2 ± 1.2                   | < 0.001 |
| Platelets     | 273.6 ± 103.5             | 181.1 ± 69.4                | < 0.001 |

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**Figure 1. Liver biopsy findings among cases.**
prevalence of HCV in Egyptians. Such a finding was expected as HCV is a global health threat in Egypt, and the country is the largest reservoir of HCV in the world [19]. Patients with rheumatic diseases have an increased susceptibility to infections including HCV, due to immunosuppression caused by the disease itself and by drugs used in their treatment [42], in addition to their high exposure to numerous invasive diagnostic and therapeutic procedures, including blood transfusions [43].

The current study aimed at testing the effect of corticosteroid therapy on serum levels of IL-10 and IL-17 as well as lncRNA-AF085935 in patients of RA associated with HCV and evaluating the usefulness of using these parameters to predict the therapeutic efficacy of steroids in these patients.

In this study, it was found that our patients significantly had lower pretreatment HCV RNA level which increased markedly after the administration of systemic steroid therapy. This agrees with Cansu et al who demonstrated that glucocorticoids and immunosuppressive drugs that are commonly used in the management of rheumatic diseases can increase HCV replication [44]. In another study, corticosteroids rose HCV RNA levels when used in treatment of HCV patients [13]. There is one study in which corticosteroid treatment was applied at clinically relevant concentrations (1 - 10 nM) to the HCV replicon model. Not only did it not enhance but resulted in a slight reduction of HCV replication (over 41% reduction in HCV RNA levels) [45].

Our results revealed that the serum levels of IL-10 were significantly higher in our patients of RA associated with HCV than in healthy individuals. This increase was in partial agreement with Lacki et al [27] and St Clair [28] who detected that concentration of IL-10 in sera of patients with RA is higher as compared to controls and in partial agreement with what has been reported by Jia et al who reported that increased serum level of IL-10 is present in chronic HCV patients [21]. Moreover, a significant high level of serum IL-10 was found in our patients after steroid therapy.

The result of the current study was in contrast with the study done by Lapadula et al who detected that RA patients showed significantly reduced IL-10 levels in serum in comparison to healthy donors, suggesting that IL-10 synthesis is depressed in RA [25]. In addition, patients with advanced RA had the lowest values of IL-10 in serum [26]. On the other hand, a significant decrease in plasma IL-10 level was reported

| Table 3. HCV RNA, IL-10, IL-17 and lncRNA-AF085935 in Different Study Groups |
|---------------------------------|---------------------------------|---------------------------------|---------------------------------|
| Variables                        | G1: before treatment (n = 65), mean ± SD | G2: after treatment (n = 65), mean ± SD | G3: control (n = 30), mean ± SD |
| HCV RNA                          | 323,955 ± 337,313.6                  | 1418,729.7 ± 2,380,203.2         | 0.001a                          |
| IL-10                            | 68.5 ± 24.7                         | 201.4 ± 31.9                    | < 0.001b, c                     |
| IL-17                            | 227.9 ± 35.5                        | 112.4 ± 28.1                    | < 0.001b, c                     |
| LncRNA-AF085935                  | 4.12 ± 0.67                         | 4.17 ± 0.66                     | < 0.001b, c                     |

aSignificance between G1 and G2. bSignificance between G1 and G3. cSignificance between G2 and G3.

| Table 4. Correlation Between HCV RNA Level With Other Parameters Before and After Treatment Among Study Group |
|---------------------------------------------------------------------------------|--------------------------------------------------------------------------------|--------------------------------------------------------------------------------|
| Variables | HCV-PCR before treatment | HCV-PCR after treatment |
| IL-10     | 0.02 (0.8)               | 0.01 (0.9)                |
| IL-17     | 0.05 (0.7)               | -0.17 (0.2)               |
| LncRNA-AF085935 | -0.033 (0.9) | 0.04 (0.8) |
The cellular sources of IL-10 in synovial tissue are macrophages and T cells [24]. The pro-inflammatory cytokines themselves appear to trigger the synthesis of IL-10 as evidenced by experiments in which adding TNF-α or IL-1 has been shown to augment IL-10 production [47]. IL-10 acts to curb the inflammatory response, so it serves to inhibit synovial inflammation [28]. Synovial fluid from patients with RA contains detectable levels of IL-10 mRNA and protein [24]. The predominant source of IL-10 in the synovial fluid is the mononuclear cell (MNC) [48]. The blood MNCs from patients with RA spontaneously produce IL-10 in culture [49] and represent the main source of circulatory IL-10 so the median serum level of IL-10 is increased in patients with RA compared with that of healthy controls [50].

IL-10 is an important anti-inflammatory cytokine that is secreted from Th2 cells. The levels of IL-10 production determine the regulation of the immune response and the balance between the inflammatory and humoral responses [51]. The Th1 cell-mediated immune response is a key factor in the protection against HCV infection [52]. Conversely, patients presenting with a Th2 response evolve into chronicity [53] and are less likely to eliminate viremia following therapy [54]. Because it has been shown that the Th1 and Th2 responses are not completely independent, Th2 cytokines such as IL-10 can downregulate the cell-mediated immune mechanisms that

**Table 5. Sensitivity and Specificity of Different Parameters in Predicting the Efficacy of Steroid Therapy in Patients**

| Variable          | Sensitivity | Specificity | Accuracy | Cut-off point |
|-------------------|-------------|-------------|----------|---------------|
| IL-10             |             |             |          |               |
| Before            | 77.3%       | 23.3%       | 51%      | 45            |
| After             | 72.7%       | 30.2%       | 53.9%    | 175.1         |
| IL-17             |             |             |          |               |
| Before            | 72.7%       | 25.6%       | 42%      | 198.5         |
| After             | 81.8%       | 23.3%       | 62.1%    | 196.8         |
| LncRNA-AF085935   |             |             |          |               |
| Before            | 63.6%       | 27.2%       | 48.5%    | 3.74          |
| After             | 68.2%       | 20.9%       | 38.4%    | 3.63          |

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**Figure 3.** ROC curve for IL-10 before and after treatment.
are important for host defense against viral infections [55].

The persistent production of IL-10 as a result of stimulation by HCV proteins leads to skewing from a protective Th1 response, which results in viral evasion and chronic infection [56]. A shift toward a Th1 cytokine profile is implicated in a more benign and favorable course of HCV infection [57].

Our results regarding the increase of IL-10 following steroid therapy was in harmony with the study done by Peek et al who reported that secretion of endogenous IL-10 might be augmented by glucocorticoids (GCS) [58].
Our results revealed that the serum levels of IL-17 were significantly higher in our patients than in healthy individuals. Moreover, the serum IL-17 level was observed to be significantly lower in patients after steroid therapy than in them before treatment. Th17 cells produce the cytokine IL-17, which induces the production of several pro-inflammatory cytokines, such as TNF-α and IL-6, chemokines, and some other additional novel factors responsible for RA and other autoimmune diseases [59]. IL-17 has also been found in the synovial membrane and synovial fluid of RA patients [60]. So in chronic inflammatory conditions such as RA, the levels of pro-inflammatory cytokines are increased [61], possibly to provide additional energy resources for the activated immune system [62]. On the other hand, it was found that serum IL-17 levels were significantly higher in chronically HCV infected patients than controls [33]. This higher serum IL-17 in patients with chronic liver disease goes with what has been reported previously that serum IL-17 level was increased in liver injuries following chronic hepatitis and cirrhosis supporting a role for IL-17 as a chronic disease inducer in the pathogenesis and/or progression of liver fibrosis [34]. Administration of corticosteroid therapy is associated with a decrease in inflammatory markers levels due to its anti-inflammatory effects [63]. Our results detected that the high expression of IncRNA-AF085935 in our patients of RA associated with HCV is highly significant than in healthy individuals who show underexpression of this IncRNA. The high upregulated AF085935 was not changed in patients following steroid therapy. We suggest that IncRNA-AF085935 may serve as a potential biomarker for the detection of RA associated with HCV patients. More studies about the biological regulatory roles of IncRNAs have demonstrated that some IncRNAs can influence the roles of their neighboring protein-coding genes by acting as biological enhancers [64]. In addition, some IncRNAs have been tightly linked between gene regulation and nuclear organization [65].

In conclusion, corticosteroid treatment caused a significant decrease in the serum levels of IL-17 and a significant increase in IL-10 and HCV RNA. However, our findings provide evidence that serum levels of IL-10 are non-invasive prognostic markers to predict the efficacy of corticosteroid therapy in RA patients associated with chronic hepatitis C. A member of IncRNAs, IncRNA-AF085935 was highly expressed in the serum of RA patients associated with HCV. This leads to an indication that IncRNA-AF085935 might be a useful candidate biomarker for the early detection of RA associated with HCV which may provide potential new strategies for early screening and therapy of these patients. Further studies are needed to clarify these conclusions. This knowledge could be used to develop new treatment options. Also, the population of enrolled patients and controls was relatively small, which needs a larger sample to further study to verify our results.

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