Combination Effect of Hemostatic and Disinfecting Agents on Micro-leakage of Restorations Bonded with Different Bonding Systems

Farhadpour H, Sharafeddin F, Akbarian S, Azarian B

Department of Dental Materials, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran.
Department of Operative Dentistry and Biomaterials Research Center, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran.
Department of Operative Dentistry, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran.
Tohid high school, Shiraz, Iran.

Abstract

Statement of Problem: Hemostatic agents may affect the micro-leakage of different adhesive systems. Also, chlorhexidine has shown positive effects on micro-leakage. However, their interaction effect has not been reported yet.

Objectives: To evaluate the effect of contamination with a hemostatic agent on micro-leakage of total- and self-etching adhesive systems and the effect of chlorhexidine application after the removal of the hemostatic agent.

Materials and Methods: Standardized Class V cavity was prepared on each of the sixty caries free premolars at the cemento-enamel junction, with the occlusal margin located in enamel and the gingival margin in dentin. Then, the specimens were randomly divided into 6 groups (n = 10) according to hemostatic agent (H) contamination, chlorhexidine (CHX) application, and the type of adhesive systems (Adper Single Bond and Clearfil SE Bond) used. After filling the cavities with resin composite, the root apices were sealed with utility wax. Furthermore, all the surfaces, except for the restorations and 1mm from the margins, were covered with two layers of nail varnish. The teeth were immersed in a 0.5% basic fuschin dye for 24 hours, rinsed, blot-dried and sectioned longitudinally through the center of the restorations bucco-lingually. The sections were examined using a stereomicroscope and the extension of dye penetration was analyzed according to a non-parametric scale from 0 to 3. Statistical analysis was performed using Kruskal-Wallis test and Mann-Whitney U-test.

Results: While ASB group showed no micro-leakage in enamel, none of the groups showed complete elimination of micro-leakage from the dentin. Regarding micro-leakage at enamel, and dentin margins, there was no significant difference between groups 1 and 2, 1 and 3, and 2 and 3 (p > 0.05). A significantly lower micro-leakage at the enamel and dentin margins was observed in group 3, compared to group 6. No significant difference was observed between groups 4 and 5 in enamel (p = 0.35) and dentin (p = 0.34). Group 6 showed significantly higher micro-leakage, compared to group 4 and 5 (p < 0.05).

Conclusions: Hemostatic agent contamination had no significant effect on micro-leakage of total- and self-etching adhesive systems. Application of chlorhexidine after the removal of hemostatic agent increased micro-leakage in self-etching adhesives but did not affect when total-etching was used.

Key words: Chlorhexidine, Hemostatic Agent, Micro-leakage, Self-etching Adhesive, Total-etching Adhesive

Corresponding Author:
Sahar Akbarian
Department of Operative Dentistry, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran.
Email: drakbarian@yahoo.com
Tel:+98-71-36263193

Cite this article as: Farhadpour H, Sharafeddin F, Akbarian S, Azarian B. Combination Effect of Hemostatic and Disinfecting Agents on Micro-leakage of Restorations Bonded with Different Bonding Systems. J Dent Biomater, 2016;3(3):292-298.
Introduction

Micro-leakage is the transition of fluids, bacteria, ions or molecules between a cavity wall and the restorative material [1]. In composite restorations, micro-leakage can be caused by polymerization shrinkage, the differences in the coefficient of thermal expansion between tooth structure and the material, and occlusal loads [2]. Contamination in different stages of bonding procedure may also cause micro-leakage [3-5]. Clinically, it can lead to staining around the margins of restorations, margin deterioration, poor aesthetics, post-operative sensitivity, secondary caries, restoration failure, pulpal pathology, and unfavorable effects on clinical performance of composite restorations [6-8]. In most of the current literature on adhesive techniques and materials, the focus is on the elimination of leakage because of its importance for the long-term success of restorations [1].

In order to achieve a durable bond, moisture control is essential. However, isolation against saliva and blood contamination is sometimes difficult, especially at and below the gingival margin [9]. Saliva and blood contamination can reduce the dentin bond strength of total- and self-etching adhesive systems [10-12]. Clinicians may rely on hemostatic agents to avoid blood contamination, particularly near or at the gingival margin where blood contamination is more probable [13]. Hemostatic agents are substances that can be used on hemorrhagic gingival tissues before placing restorations [14-15]. Aluminium chloride with a concentration between 5 and 25% is one of the most frequently used hemostatic agents which, with its minimal systemic effects, can precipitate proteins, constrict blood vessels and extract fluid from tissues [14]. Hemostatic agents are highly acidic and are able to remove smear layer and cause some degree of demineralization [15].

Although the effect of hemostatic agents on micro-leakage and bond strength has been evaluated in several studies, they have reached different results [13,15,16]. If viable microorganisms remain after cavity preparation, micro-leakage may cause more problems which can result in secondary caries [6]. According to some bacteriologic and histologic studies, only a small portion of teeth is sterile after cavity preparation [7]. Proliferation of residual bacteria from the smear layer occurs even when there is a good seal from the oral cavity [17]. This problem can be magnified by micro-leakage of composite restoration at margins not ending on enamel [18]. Treating the cavity preparation with chlorhexidine may solve this problem to some extent. Chlorhexidine has a broad spectrum of action, mostly against Gram-positive bacteria, particularly Streptococcus mutans, which seems to be more sensitive [19]. Currently, chlorhexidine is used not only as an anti-microbial agent, but also as a potential adjuvant to establish a better bond to dentin [20]. However, owing to the lack of consensus existing over the effect of chlorhexidine [20,21], further studies are required in this regard.

Generally, bond degradation occurs because of resin elution and collagen fibrils alteration. Exposed collagen fibrils that are exposed in the hybrid bond layer, and are not completely infiltrated by resin are susceptible to degradation by matrix metalloproteinase (MMP) enzymes of the saliva and dentin [20,22].

MMPs that are host cell-derived proteolytic enzymes have a major role in tissue-destructive inflammatory diseases and are capable of degrading most of the extracellular matrix components, including different types of collagen [23,24]. Chlorhexidine can be used to prevent or minimize the auto-degradation of collagen matrices in incompletely resin-infiltrated dentin [25,26].

Considering the contrasting effects of hemostatic agents on micro-leakage, and the positive effects of chlorhexidine mentioned above, this study aimed to evaluate the effect of hemostatic agent contamination on micro-leakage of total- and self-etching adhesive systems and the effect of chlorhexidine application after the removal of the hemostatic agent.

The null hypothesis was that hemostatic agent contamination would not affect the micro-leakage of total- and self-etching adhesive systems and the application of chlorhexidine after the removal of the hemostatic agent.

Materials and Methods

Sixty extracted non-carious human premolar teeth collected and stored at 4°C were used within one month of extraction. Standardized Class V
cavities (2 mm depth, 2 mm mesiodistal, 3 mm occlusogingival) were prepared at the cemento-enamel junction, with the occlusal margin located in enamel and the gingival margin in dentin. In the occlusal cavo surface margin, a bevel was made. Cavities were prepared using a #556 diamond fissure bur in a high-speed handpiece with water coolant. Then, they were randomly divided into 6 groups (n = 10) according to the following factors: Hemostatic agent contamination, chlorhexidine application, and the adhesive systems used. Group 1- Adper Single Bond system (ASB) (3M,ESPE,USA) was applied as recommended by the manufacturer (Table 1). Group 2- One drop of hemostatic agent solution (Hemostop,Dentsply. Argentina) was applied on the dentin surface for 2 minutes and rinsed for 30s and air-dried. Then, the cavity surface was etched with 37% phosphoric acid gel for 15s. After rinsing and air-drying, ASB was applied (H + ASB). Group 3- One drop of the hemostatic agent solution was applied into the cavity for 2 minutes, rinsed for 30s and air-dried. After that, CHX 0.2% solution was applied with a microbrush. The next steps were similar to group 1 (H+CHX+ASB). Group 4- Clearfil SE Bond system (CSB) (Kuraray,Japan) was applied as recommended by the manufacturer (Table 1). Group 5- One drop of hemostatic agent solution was applied into the cavity for 2 minutes and rinsed for 30s and air-dried. Then, CSB was applied (H+CSB). Group 6- One drop of hemostatic agent solution was applied into the cavity for 2 minutes and rinsed for 30s and air-dried. After that, CHX 0.2% solution was applied with a microbrush. Then, CSB was applied (H+CHX+CSB).

After the adhesive application and light curing for 20s with an LED light curing system (Demetron,Kerr)(1200 mW/cm²), the cavities were bulk filled with one increment of resin composite (Filtek Z250, 3M, ESPE, St Paul, MN, USA). Excess composite was removed with an explorer and then light cured with an LED for 20s. All procedures were performed by one operator at room temperature and during procedures, the cavities were kept moist. After the restoration, the samples were stored for 24 hours in distilled water.

Afterwards, the root apices were sealed with utility wax, and all the surfaces, except for the restorations and 1mm from the margins, were covered with two layers of nail varnish. The teeth were immersed in a 0.5% basic fuschin dye for 24 hours. Then, they were rinsed, blot-dried and sectioned longitudinally through the center of the restorations from the buccal to lingual surface with a water-cooled diamond saw (Leitz 1600, Wetzlar, Germany).

The sections were examined for dye penetration by two independent researchers using a stereomicroscope (Carl ZiiessInc, Oberkochen, Germany) at 20× magnification. The extent of the dye penetration was analyzed according to a non-parametric scale from 0 to 3 (0 = no dye penetration, 1 = dye penetration less than 1/2 of the cavity depth, 2 = dye penetration more than 1/2 of the cavity depth, 3 = dye penetration spreading along the axial wall).

Statistical analysis was performed using Kruskal-Wallis H test and Mann-Whitney U-test. Significance level for all statistical tests was predetermined at $p < 0.05$.

| Table 1: Adhesive Systems, Composition, and Application Mode |
|----------------------------------|----------------------------------|----------------------------------|
| Adhesive Systems                 | Composition                      | Application mode                 |
| Adper Single Bond (3M, ESPE, St Paul, MN, USA) | Scotchbond etchant:37% phosphoric acid Adhesive:Bis-GMA, HEMA, dimethacrylates, polyalkenoic acid copolymer, initiators, water, and ethanol | 1. Apply acid etch for 15s 2. Rinse for 15s 3. Blot dry for 30s 4. Apply one coat of adhesive for 10s 5. Airdry for 10s at 20cm 6. Light cure. |
| Clearfil SE Bond (Kuraray, Okayama, Japan) | Primer:MDP,HEMA,dimethacrylate Monomer,water,catalyst Bond:MDP,HEMA, dimethacrylate Monomer,microfiller,catalyst | 1. Apply primer and leave for 20s 2. Dry thoroughly with mild air flow 3. Apply bond 4. Gentle air flow 5. Light cure. |
Results

Dye penetration scores for the enamel and dentinal margins are presented in Table 2. Whereas ASB group showed no micro-leakage in enamel, none of the groups showed complete elimination of micro-leakage in dentin. Pairwise multiple comparisons of the six groups were performed using the Mann-Whitney test between treatment groups for both enamel and dentinal margins.

There was no significant difference between groups 1 and 2 neither in enamel \((p = 0.51)\) nor in dentin \((p = 0.41)\). Moreover, there was no significant difference between groups 1 and 3 concerning enamel \((p = 0.51)\), and dentin \((p = 0.26)\). Also, no significant difference was seen between groups 2 and 3 regarding enamel \((p = 0.97)\) and dentin \((p = 0.52)\) margins. However, significantly lower micro-leakage at enamel and dentin margins was observed in group 3, compared to group 6 \((p < 0.05)\).

No significant difference was observed between groups 4 and 5 in enamel \((p = 0.35)\) and dentin \((p = 0.34)\) margins. Group 6 showed significantly higher micro-leakage, compared to group 4 \((p < 0.05)\) and group 5 \((p < 0.05)\).

Discussion

In the first part of this study, we evaluated the effect of hemostatic agent contamination on the micro-leakage of total- and self-etching adhesive systems. These findings are in line with a previous study performed by Arslan et al. that reported no significant changes in micro-leakage scores after the application of the hemostatic agent, with self-etching adhesives [13]. In another study, the researchers used Viscostat as a hemostatic agent and contrary to our findings; they reported significant increase in micro-leakage [3]. The difference in the results of that study and the present study may be attributed to differences in the hemostatic agents used.

The increased micro-leakage might be because of the fact that Viscostat, contrary to Hemostop, is a viscous gel composed of ferric sulphate and the viscosity makes its removal from the dentin surface harder, and may lead to coagulation of proteins present in the dentinal fluid. The coagulated proteins and residues of ferric sulfate may result in the inhibition of the infiltration of the bonding agent into the etched enamel surfaces and dentinal tubules.

Evaluation of the effect of chlorhexidine application after the removal of hemostatic agent revealed that the use of chlorhexidine after astringent removal did not have any significant effects on micro-leakage when ASB was used as adhesive system and the second part of the null hypothesis was accepted for ASB.

But regarding CSB, chlorhexidine application increased micro-leakage and the null hypothesis was rejected in this regard.

For deciding the step in which chlorhexidine is applied, in some studies, chlorhexidine has been used after etching [26,27]. The rationale for the application of chlorhexidine in this step is the
ability of chlorhexidine to inhibit enzyme activities directly; inhibiting the inherent collagenolytic activity of mineralized dentin would reinforce the bond of resin composite to dentin. In some other investigations, chlorhexidine has been used before etching with the purpose of the disinfection and cleansing of the dentin surface [21,28]. Also, according to some other investigators, there is no significant difference between the application of chlorhexidine before or after etching of dentin [29,30]. In this study, chlorhexidine was used after removing the hemostatic agent and before etching in order to disinfect the dentin surface and remove the microorganisms and thus to reduce the probability of recurrent caries and restoration failure as well as eliminating the remnants of hemostatic agent that could interfere with the bond of composite to dentin.

In a study by Sung et al., the researchers concluded that application of chlorhexidine had no significant effect on micro-leakage of total-etching system [31]. Siso et al. reported that chlorhexidine solution had no effect on micro-leakage of composite restorations [32]. Although a number of studies obtained different results after chlorhexidine use [31-33], some other studies reported findings similar to ours. For instance, Tulunoglu et al. concluded that chlorhexidine solution had an adverse effect on self-etching adhesive systems and produced higher micro-leakage [33]. Singla et al. also reported higher micro-leakage when using chlorhexidine with self-etching adhesives [34].

Further in vitro and in vivo studies are recommended to improve the understanding of the interaction effect of different hemostatic agents and chlorhexidine with bonding systems which have different acidities.

**Conclusions**

Considering the limitations of this in vitro study, we found that hemostatic agent contamination had no significant effect on micro-leakage of total- and self-etching adhesive systems. Application of chlorhexidine after the removal of hemostatic agent could increase micro-leakage in self-etching adhesives but did not affect the micro-leakage when using total-etching adhesive systems.

**Acknowledgements**

The authors thank the Vice-Chancellory of Shiraz University of Medical Science and Biomaterial Research Center of Shiraz University of Medical Sciences for supporting this research (Grant# 5455). The authors also thank Dr. M. Vossoughi for the statistical analysis and Dr. Ehya Amalsaleh for improving the use of English in the manuscript.

**Conflict of Interest:** None declared.

**References**

1. Kidd EA. Microleakage: a review. J Dent. 1976;4:199-206.
2. Ben-Amar A. Microleakage of composite resin restorations. A status report. Am J Dent. 1989;2:175-180.
3. Kumar P, Shenoy A, Joshi S. The effect of various surface contaminants on the microleakage of two different generation bonding agents: A stereomicroscopic study. J Conserv Dent. 2012;15:265-269.
4. Sharafeddin F, Koohpeima F, Palizian B. Evaluation of Microleakage in Class V Cavities Filled with Methacrylate-based versus Silorane-based Composites. J Dent Biomater. 2015;2:67-72.
5. Sharafeddin F, Darvishi F, Malekzadeh P. The Effect of Incorporation of Polyethylene and Glass Fiber on the Microleakage of Silorane-based and Methacrylate-based Composites in Class II Restorations: An in Vitro Study. J Dent Biomater. 2015;2:39-46.
6. Moreira Jr G, Sobrinho AP, Nicoli JR, et al. Evaluation of microbial infiltration in restored cavities—An alternative method. J Endod. 1999;25:605-608.
7. Kihn P, Spanganberg P, von Fraunhofer J. The role of cavity preparation and conditioning in the leakage of restorations. J Adhes Dent. 2004;6:287-291.
8. Eick JD, Welch FH. Polymerization shrinkage of posterior composite resins and its possible influence on postoperative sensitivity. Quintessence Int. 1986;17:103-111.
9. Yoo H, Pereira P. Effect of blood contamination with 1-step self-etching adhesives on micro-
Effect of Hemostatic and Disinfectants on Micro-leakage

10. de Carvalho Mendonça EC, Vieira SN, Kawaguchi FA, et al. Influence of blood contamination on bond strength of a self-etching system. Eur J Dent. 2010;4:280-286.
11. Chang S, Cho B, Lim R, et al. Effects of blood contamination on microtensile bond strength to dentin of three self-etch adhesives. Oper Dent. 2010;35:330-336.
12. Neelagiri K, Kundabala M, Shashi RA, et al. Effects of saliva contamination and decontamination procedures on shear bond strength of self-etch dentine bonding systems: An in vitro study. J Conserv Dent. 2010;13:71-75.
13. Arslan S, Ertaş H, Zorba YO. Effect of a plant-based hemostatic agent on microleakage of self-etching adhesives. Med Oral Patol Oral Cir Bucal. 2013;18:124-129.
14. Gupta G, Kumar S, Rao H, et al. Astringents in dentistry: a review. Asian J Pharm Hea Sci. 2012;2:428-432.
15. Harmirattisai C, Kuphasuk W, Senawongse P, et al. Bond strengths of resin cements to astringent-contaminated dentin. Oper Dent. 2009;34:415-422.
16. Sharafeddin F, Farhadpour H. Evaluation of Shear Bond Strength of Total- and Self-etching Adhesive Systems after Application of Chlorhexidine to Dentin Contaminated with a Hemostatic Agent. J Dent (Shiraz). 2015;16:175-181.
17. Brannstrom M. The cause of postrestorative sensitivity and its prevention. J Endod. 1986;12:475-481
18. Boston D, Graver H. Histological study of an acid red caries-disclosing dye. Oper Dent. 1989;14:186-192
19. Fardal O, Turnbull R. A review of the literature on use of chlorhexidine in dentistry. J Am Dent Assoc. 1986;112:863-869.
20. Herénio SS, de Carvalho NMP, Lima DM. Influence of chlorhexidine digluconate on bond strength durability of a self-etching adhesive system. Rev Sulbras Odontol. 2011;8:417-424.
21. Ercan E, Erdemir A, Zorba YO, et al. Effect of different cavity disinfectants on shear bond strength of composite resin to dentin. J Adhes Dent. 2009;11:343-346.
22. Brackett WW, Tay FR, Brackett MG, et al. The effect of chlorhexidine on dentin hybrid layers in vivo. Oper Dent. 2007;32:107-111.
23. Moon PC, Weaver J, Brooks CN. Review of matrix metalloproteinases’ effect on the hybrid dentin bond layer stability and chlorhexidine clinical use to prevent bond failure. Open Dent J. 2010;4:147-152.
24. Gendron R, Grenier D, Sorsa T, et al. Inhibition of the activities of matrix metalloproteinases 2, 8, and 9 by chlorhexidine. Clin Diagn Lab Immunol. 1999;6:437-439.
25. Hebling J, Pashley DH, Tjäderhane L, et al. Chlorhexidine arrests subclinical degradation of dentin hybrid layers in vivo. J Dent Res. 2005;84:741-746.
26. Komori PC, Pashley DH, Tjäderhane L, et al. Effect of 2% chlorhexidine digluconate on the bond strength to normal versus caries-affected dentin. Oper Dent. 2009;34:157-165.
27. Loguercio AD, Stanislawczuk R, Polli LG, et al. Influence of chlorhexidine digluconate concentration and application time on resin-dentin bond strength durability. Eur J Oral Sci. 2009;117:587-596.
28. Breschi L, Cammelli F, Visintini E, et al. Influence of chlorhexidine concentration on the durability of etch-and-rinse dentin bonds: a 12-month in vitro study. J Adhes Dent. 2009;11:191-198.
29. de Castro FL, de Andrade MF, Duarte Júnior SL, et al. Effect of 2% chlorhexidine on microtensile bond strength of composite to dentin. J Adhes Dent. 2003;5:129-138.
30. Campos EA, Correr GM, Leonardi DP, et al. Influence of chlorhexidine concentration on microtensile bond strength of contemporary adhesivesystems. Braz Oral Res. 2009;23:340-345.
31. Sung EC, Chan SM, Tai ET, et al. Effects of Various irrigation solutions on microleakage of Class V composite restoration. J Prostheth Dent. 2004;91:265-267
32. Siso HS, Kustarci A, Göktolga EG. Microleakage in resin composite restorations after antimicrobial pretreatments: effect of KTPlaser, chlorhexidine gluconate and Clearfil
33. Tulunoglu O, Ayhan H, Olmez A, et al. The effect of cavity disinfectants on microleakage in dentin bonding systems. J Clin Pediatr Dent. 1998;22:299-305

34. Singla M, Aggarwal V, Kumar N. Effect of chlorhexidine cavity disinfection on microleakage in cavities restored with composite using a self-etching single bottle adhesive. J Conserv Dent. 2011;14:374-377.