Commentary: Response of Human Macrophages to Clinically Applied Wound Dressings Loaded With Silver

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A Commentary on

Response of Human Macrophages to Clinically Applied Wound Dressings Loaded With Silver by Varela, P., Marlinghaus, L., Sartori, S., Viebahn, R., Salber, J., and Ciardelli, G. (2020). Front. Bioeng. Biotechnol. 8:124. doi: 10.3389/fbioe.2020.00124

INTRODUCTION

The authors Varela et al. (2020) pose interesting, relevant questions related to silver-containing wound dressing toxicity, antimicrobial effects, effect on the transition from the M1 to the M2 macrophage phenotypes, and effect on wound inflammation overall. They designed creative studies to answer these questions using six primary dressings: Atrauman, Biatain, and PolyMem WIC, all with and without silver (Varela et al., 2020). Four bacterial strains known to infect open wounds (Staphylococcus aureus, Staphylococcus epidermidis, Escherichia coli, and Pseudomonas aeruginosa), and three human cell types (fibroblasts, keratinocytes, and macrophages) were cultured. Test dressing discs were placed over two-dimensional bacteria and cell culture colonies, which were then incubated (Varela et al., 2020). Wells with test dressings covered with 2 ml of a bacterial broth were also incubated (Varela et al., 2020). The descriptions of wound healing, materials used, tests, and rationale are clear (Varela et al., 2020).

However, the study results are unlikely to be clinically meaningful because of two major design flaws: (1) Contrary to the Instructions for Use, the researchers did not provide secondary (cover) dressings to prevent desiccation with any of the six study dressings (Hess, 2020; Atrauman, 2020). Atrauman (with and without silver) is intended to keep relatively dry wounds moist, making it somewhat suitable for testing on agar (Atrauman, 2020). In contrast, PolyMem WIC and Biatain (with and without silver) are designed for cavity and heavily exudating wounds, making them unsuitable (Hess, 2020). Because all six dressings were tested without moisture-retentive coverings (Varela et al., 2020), the probability that many of the

ANTIBACTERIAL AND CYTOTOXICITY TESTS FOR ALL TEST DRESSINGS CONFOUNDED BY DESICCATION

All six test dressings are designed to be covered with a secondary dressing to maintain appropriate wound moisture (Hess, 2020; Atrauman, 2020). Atrauman (with and without silver) is intended to keep relatively dry wounds moist, making it somewhat suitable for testing on agar (Atrauman, 2020). In contrast, PolyMem WIC and Biatain (with and without silver) are designed for cavity and heavily exudating wounds, making them unsuitable (Hess, 2020). Because all six dressings were tested without moisture-retentive coverings (Varela et al., 2020), the probability that many of the
plated bacterial and human cells died due to desiccation, rather than due to the toxicity of any particular dressing components, cannot be discounted.

PolyMem WIC dressings gradually release glycerol, which draws fluid from the body into the wound bed (Cutting et al., 2015; Benskin, 2016b, 2018). From there, the fluid is absorbed by the superabsorbent within the dressings and the dressing substrate itself (Cutting et al., 2015; Benskin, 2016b, 2018). Without a body to donate fluid, both the standard and the silver PolyMem WIC entrapped a part of the agar in the dressing, providing evidence of desiccation and a failure of the experimental model (Benskin, 2016a,b; Varela et al., 2020). Desiccation as an unmeasured confounding variable would explain why Atrauman, with and without silver, the test dressings with the greatest ability to preserve wound moisture in vitro without a cover dressing, appeared to be less cytotoxic than the other test dressings, including the other two without silver (Atrauman, 2020; Varela et al., 2020).

CYTOTOXICITY TEST OF POLYMEM DRESSINGS CONFOUNDED BY NON-INTERACTIVE IN VITRO SETTING

As the authors noted, rather than distributing cytotoxic silver into the wound bed like the other silver test dressings, PolyMem Silver’s continuous wound cleansing system interacts with the body to dislodge wound pathogens and pull them into contact with silver locked in the dressing (Burd et al., 2007; Benskin, 2012, 2018; Varela et al., 2020). Rather, in vitro, PolyMem WIC had no body from which to draw new moisture to replace what was absorbed, impairing the ordinarily robust moisture-balancing system and leading to cell death by desiccation (Benskin, 2016a, 2018).

ANTIBACTERIAL TESTS OF POLYMEM DRESSINGS CONFOUNDED BY NON-INTERACTIVE IN VITRO SETTING

PolyMem Silver cannot demonstrate its antibacterial benefits on a petri dish because virtually all of the silver is retained safely in the dressing substrate (Burd et al., 2007). Failure to take PolyMem’s powerful continuous wound cleansing system into account also confounded the antibacterial tests. In the disc diffusion test, the entrapped agar is evidence of this study design failure (Varela et al., 2020). In the broth medium experiment, after PolyMem WIC Silver was completely saturated, bacteria remaining in the broth could not be pulled into contact with the silver in the PolyMem (Varela et al., 2020).

WOUND INFLAMMATION TESTS OF POLYMEM DRESSINGS CONFOUNDED BY NON-INTERACTIVE IN VITRO SETTING

The statement that PolyMem is “defined” as an “anti-inflammatory dressing” is a gross over-simplification (Varela et al., 2020). Controlled inflammation is beneficial throughout the wound healing process (Davies and White, 2011). PolyMem dressings (with and without silver) focus inflammation, permitting the robust concentration of macrophages and neutrophils at the injury site that is necessary for brisk healing to take place, while virtually eliminating the secondary inflammation that causes poor perfusion (Beitz et al., 2004; Davies and White, 2011; Benskin, 2012; Weissman et al., 2013). This highly desirable alteration in the inflammatory response is not attributable to any individual component, such as the silver, glycerol, surfactant, or superabsorbent, but rather, it is the synergistic effect of PolyMem’s multiple components interacting with the body’s nociceptor response (Kahn et al., 2000; Beitz et al., 2004; Weissman et al., 2013; Benskin, 2016a).
Varela et al. measured the total number and phenotypes of the macrophages and inflammatory markers, failing to examine their location within the wound (Varela et al., 2020). Beitz et al. applied PolyMem dressings to a stab wound, finding that the inflammatory cells deep in the muscle tissue lined up along the incision line, with almost no inflammatory cells in the surrounding tissue (see Figure 1) (Beitz et al., 2004). And, applying PolyMem to distal open or closed tissue injuries altered inflammation markers in the spinal cord laminae (Beitz et al., 2004). Kahn et al., found that bruising and edema were dramatically reduced when PolyMem was applied to intact skin immediately following deep tissue mechanical trauma (Kahn et al., 2000). Varela et al. could not measure these critical distinctions between PolyMem and other wound dressings with respect to inflammation because in vitro models cannot include an intact central nervous system (Benskin, 2018; Varela et al., 2020).

DISCUSSION

The absence of secondary dressings in this study design would result in varying degrees of cell desiccation, a confounding variable that could have influenced bacterial and human cell death quite apart from the effects of the silver in the test dressings (Hess, 2020; Atrauman, 2020). This renders the results of the bactericidal and cytotoxicity aspects of this study clinically irrelevant. And, testing any configuration of PMDs in vitro (or ex vivo) will fail to capture all the benefits of PolyMem dressings which are achieved through their complex synergistic interaction with the patient’s body, such as controlling and focusing inflammation, continuously cleansing wounds, and balancing moisture (Weissman et al., 2013; Benskin, 2016a,b; Benskin, 2018).

Perhaps new devices will be invented to allow non-invasive dynamic wound inflammation evaluation, similar to current oxygen saturation monitors. Until these are available, inflammatory markers in the exudate from PolyMem and other spent dressings could be compared.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

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REFERENCES

Atrauman (2020). HARTMANN GROUP. Available online at: https://www.hartmann.info/en-gb/our-products/wound-management/wound-contact-layers/impregnated-wound-contact-layers/atrauman (accessed April 26, 2020).

Beitz, A. J., Newman, A., Kahn, A. R., Ruggles, T., and Eikmeier, L. (2004). A polymeric membrane dressing with antinociceptive properties: analysis with a rodent model of stab wound secondary hyperalgesia. J. Pain 5, 38–47. doi: 10.1016/j.jpain.2003.09.003

Benskin, L. L. (2016a). Limitations of in vitro antimicrobial dressings study. Burns 42, 1147–1148. doi: 10.1016/j.burns.2016.01.034

Benskin, L. L. (2016b). Polymeric membrane dressings for topical wound management of patients with infected wounds in a challenging environment: a protocol with 3 case examples. Ostomy Wound Manage. 62, 42–50. doi: 10.25270/owm.2016.6.4250

Benskin, L. L. (2018). Methods used in the study. Evaluation of a polyurethane foam dressing impregnated with 3% povidone-iodine (Betafoam) in a rat wound model, led to unreliable results. Ann. Surg. Treat. Res. 95, 230–232. doi: 10.4174/astr.2018.95.4.230

Benskin, L. L. (2012). PolyMem Wic Silver rope: a multifunctional dressing for decreasing pain, swelling, and inflammation. Adv. Wound Care 1, 44–47. doi: 10.1089/wound.2011.0285

Burd, A., Kwok, C. H., Hung, S. C., Chan, H. S., Gu, H., Lam, W. K., et al. (2007). A comparative study of the cytotoxicity of silver-based dressings in monolayer cell, tissue explant, and animal models. Wound Repair Regen. 15, 94–104. doi: 10.1111/j.1524-475X.2006.00190.x

Cutting, K. F., Vowden, P., and Wiegand, C. (2015). Wound inflammation and the role of a multifunctional polymeric dressing. Wounds Int. J. 6, 41–46.

Davies, S. L., and White, R. J. (2011). Defining a holistic pain-relieving approach to wound care via a drug free polymeric membrane dressing. J. Wound Care. 20, 250, 252, 254 passim. doi: 10.12968/jowc.2011.20.5.250

Hess, C. T. (2020). Product Guide to Skin and Wound Care, 8th Edn. Philadelphia, PA: Wolters Kluwer, 491.

Kahn, A. R., Sessions, R. W., and Apasova, E. V. (2000). A superficial cutaneous dressing inhibits pain, inflammation and swelling in deep tissues. Pain Med. 1:187. doi: 10.1046/j.1526-4637.2000.00024.x

Varela, P., Marlinghaus, L., Sartori, S., Viebahn, R., Salber, J., and Ciardelli, G. (2020). Response of human macrophages to clinically applied wound dressings loaded with silver. Front. Bioeng. Biotechnol. 8:124. doi: 10.3389/fbioe.2020.00124

Weissman, O., Hundeshagen, G., Harats, M., Farber, N., Millet, E., Winkler, E., et al. (2013). Custom-fit polymeric membrane dressing masks in the treatment of second degree facial burns. Burns 39, 1316–1320. doi: 10.1016/j.burns.2013.03.005

Conflict of Interest: LB is an employee of Ferris Mfg. Corp., makers of PolyMem and PolyMem Silver dressings.

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