To Established Hematological Reference Intervals Among Full-Term Newborn Babies at University of Gondar Comprehensive Specialized Hospital, Northwest Ethiopia: A Cross-Sectional Study

Tegenaw Tiruneh (tegenawt@gmail.com)
College of Health Science, Department of Hematology and Immunohematology, Debre Tabor University

Teklehymanot Kiros
College of Health Science, Department of Medical Laboratory Science, Debre Tabor University, Debre Tabor, Ethiopia

Sisay Getu
Debre Tabor University

Research article

Keywords: Hematological reference interval, newborn babies

DOI: https://doi.org/10.21203/rs.3.rs-30807/v1

License: This work is licensed under a Creative Commons Attribution 4.0 International License.
Read Full License
Abstract

Background: Hematological reference intervals are used for medical decision tools for interpretation of numerical test results. Establishing of hematological interval among newborn babies is very important for the diagnosis of malignancy, anemia, bleeding disorders, and various infections. There is no locally established hematological reference intervals in Ethiopia. Thus the aim of this study is to established locally determined hematological reference interval among full-term newborn babies.

Methods: A cross sectional study was conducted from February to May 2019 among 151 apparently healthy full-term newborn babies at Gondar University Hospital. About 3ml of cord blood was obtained for analysis of Hematological parameters and determined by using Sysmex KX-21N (Sysmex Corporation Kobe, Japan) automated analyzer. Median, 2.5th and 97.5th percentiles were computed.

Result: Males to female's ratio was almost equal. All hematological parameter had no statistically significant difference between males and females. The delivery types were not influenced its hematological values.

Conclusion: Hematological reference interval was established from full-term newborn babies at University of Gondar hospital and was different from other studies in African and Europeans. Therefore, own determined reference value is very important for the clinicians to correctly diagnosis the patients at health facility levels.

Introduction

Establishing hematological reference intervals among full-term newborn babies at University of Gondar comprehensive specialized hospital, Northwest Ethiopia: a cross-sectional study

Background

Umbilical cord blood (UCB) is the blood remaining after in the excluding of the placenta. It is considered as biological waste products. However, it has many advantages like stem cell transplantation (rich in hematopoietic progenitor and stem cells), RH blood typing of the newborn babies, assessment of the neonatal thrombocytopenia or thrombosis, screening and diagnosis of anemia, polycythemia, bacterial sepsis and determination of hematological reference interval (RI) (1–3). The other main advantage of UCB is it minimizes blood draw from very ill newborn babies that cause increased morbidity and mortality during neonatal life. The UCB is the alternative source to eliminate unwanted repeated blood drawn for laboratory tests. The collection of UCB is safe for the patients and technically easy as compared to other blood collection sites (4).

The UCB is composed of white blood cells (WBC), red blood cells (RBC), plasma, platelets and is also rich in hematopoietic stem cells, which have immense potential to cure the malignant and genetic disorder. Therefore, establishing of hematological RI among newborn babies from UCB is very important for
neonatologist for the diagnosis RBC disorders (like anemia and polycythemia), hematological malignancy, platelet disorders (like thrombosis and thrombocytopenia), autoimmune disease, certain genetic disorders problems, monitoring of the patients during in a therapeutic regimen and diagnosis of various bacterial infections (5–7). It is widely known that newborn hematological parameters are different from those of infants or adults. Therefore, separated hematological RI is mandatory for newborn babies (8).

RI is the range between values from the lower reference limit to the upper reference limits (9). They are ideally defined on apparently healthy individuals and should be distinguished from clinical decision limits that are derived from known diseased patients. It is a distribution of numerical test results expected in a representative population of healthy individuals. It is mainly used to make medical decisions about hematological disease diagnosis, treatment & monitoring of disease prognosis of the patients (10). According to Clinical Laboratory Standards Institute guideline, the RIs are defined in relation to a healthy population to include the values in which 95% of apparently healthy individuals would fall and in which 2.5% of results in the lower range are out of the RI and 2.5% of values in the upper range will be out of the RI (11, 12).

Hematological RIs are the most common medical decision-supporting tools used for the interpretation of numerical hematological test results. Accurate patient results interpreting within the correct RI is mandatory to minimize the patient risks from disease, increase recovery rates, and improve monitoring therapy of various hematological disorders (13). Almost 80% of physicians’ medical decisions are based on information provided by laboratory reports. Therefore, locally established hematological RI among newborn babies is very important for the physician to correctly diagnosis and early treatment of the patients (14).

In Ethiopia there is no locally determined hematological RI among healthy full-term newborn babies in the country, particularly is in the study area. They used in Western and American RI. Since hematological RI is varying through age, race, ethnicity, altitude, sex, drug intake, time of sampling and socioeconomic status. It is commonly known that newborn hematological parameters are different from those of neonates, infants or adults (8). It need locally determined RI. Due to these reason we motivate to conduct this study. The aim of this study was to establish hematological RI by using UCB of apparently healthy, full-term newborn babies at University of Gondar compressive specialized hospital.

**Method**

**Study Setting, design and population**

An institution-based cross-sectional study was conducted from February 1 to May 30, 2019, at the gynecology and obstetrics department of University of Gondar comprehensive specialized hospital, Northwest Ethiopia. The hospital is located in Amhara region, 180 km far from Bahir Dar, the capital city of Amhara regional state and 727 km from Addis Ababa, the capital city of Ethiopia. The town located
2,133 meters’ elevation above sea level. The hospital has a range of specialties including pediatrics, surgery, gynecology, psychiatry, human immunodeficiency virus (HIV) care, and an outpatient clinic.

The newborn babies were selected based on the following criteria: Birth at full-term (39–42 weeks of gestation), and absence of any congenital anomalies. All selected newborn babies were physically examined at birth and found normal and apparently healthy. The premature newborn (delivered less than 37 weeks of gestation), twin newborns, the pregnancy complicated with diabetics, preeclampsia, hypertension, HIV/AIDS, chronic kidney, liver disease, malaria, anemia, and hematological malignancy were excluded from the study. On the other hand, a mother who had bleeding during pregnancy, maternal drinking of alcohol during pregnancy, cigarette smoking during pregnancy, and no antenatal care also were excluded from being sampled. A systematic random sampling technique was employed to select study participants. A total of 151 newborn babies with their respective mothers were included. Maternal hemoglobin (Hgb) values less than 11 g/dl will be excluded from the study.

**Data Collection Method**

**Socio-demographic and clinical data collection**

A pre-tested structured questionnaire prepared in English and translated to the local language (Amharic) was used to obtain newborn gender, birth weight, and presence of bleeding during pregnancy, alcohol consumption habits, and cigarette smoking during pregnancy via face-to-face interviews. The presence of maternal complications like malignancy, hypertension, diabetics, tuberculosis, HIV/AIDS, chronic kidney, and liver disease were retrieved from maternal medical records with the aid of data extraction sheet.

**Blood collection and laboratory analysis**

About 3 ml of UCB specimen were obtained from each study participants after delivery from the clamped umbilical cord. The two trend Midwifery professionals collected the cord blood sample from the clamped cord through excluding of the placenta. The collected sample was immediately poured into tri-potassium ethylene diamine tetraacetic acid (K3-EDTA) test tube and gently mixed to prevent blood clotting. In addition, 3 ml of venous blood was collected from the mother after delivery with a sterile and disposable syringe. Hematological parameters: total white cell count (WBC), differential white cell count (neutrophils, lymphocytes and mixed which contains eosinophils, monocytes. and basophiles), platelet count, red blood cell count (RBC), Hgb, hematocrit (%), mean cell volume (MCV), mean cell hemoglobin (MCH), mean cell hemoglobin concentration (MCHC), and red cell distribution width (RDW) were determined by using the using Sysmex KX-21N (Sysmex Corporation Kobe, Japan) automated hematological whole blood analyzer based on direct current principle. Three experienced laboratory technologist performs the complete blood count (CBC) by strictly adhering standard operating procedures.

**Data quality control**
In order to increase the reliability of data, training was given for the data collector prior to data collection. During laboratory data collection standard operating procedures were strictly followed and implemented from prior to specimen collection up to recording and interpreting of the laboratory results. After sample collection immediately dispensed to the wall of the EDTA test tube slowly and properly mixed by inverting the tube gently 8–10 times to prevent hemolysis of blood samples. Then label of the sample and the request paper with the same identification number in order to avoid any mix up of errors. The expired date of reagent was checked before analysis of patient samples. Daily installations and background run were done to minimize any background errors. Repeated analysis of randomly selected samples for reproducibility check (delta check) was carried out to evaluate instrument performance consistently and precisely.

**Statistical analysis**

The data were cleaned, edited, checked for completeness, and entered into Epi-info version 7.2.1.0. Then it was exported into SPSS version 20 for analysis. Outliers were detected by using Z score and Tukey method. Before any analysis, normal distribution of numerical data was checked using the Kolmogorov-Smirnov and Shapiro-Wilk test. Since all data are not Gaussian distributed we used non-parametric tests to determine the hematological RI as recommended as the clinical laboratory standard institutes (12). Median with interquartile range (IQR), and 95% confidence interval were computed. The 97.5th percentile and 2.5th percentiles were the upper and the lower reference limit of the study populations, respectively. Mann-Whitney U test also used to test for the mean difference of variance between gender (male versus female), and delivery type (vaginal versus cesarean section). According to the guideline of clinical laboratory standard institutes, we determine the 95% hematological RI by computing 97.5th percentile and 2.5th percentile.

**Results**

**Sociodemographic and obstetric data of study participants**

A total of 151 healthy newborn babies (76 females and 75 males) were included in this study. Majority of study participant (87.4%) were born from mother came from urban residence with normal birth weight (90.1%). About 81.5% of study participants were born through spontaneous vaginal delivery route and the remaining were by cesarean sections (Table 1).
Table 1
Sociodemographic characteristics of study participants at University of Gondar compressive specialized hospital, 2019 (N = 151)

| Variable                  | Frequency | Percentage (%) |
|----------------------------|-----------|----------------|
| Gender                     |           |                |
| Male                       | 75        | 49.7           |
| Female                     | 76        | 50.3           |
| Residence of mother        |           |                |
| Urban                      | 132       | 87.4           |
| Rural                      | 19        | 12.6           |
| Occupation of the mother   |           |                |
| Employed                   | 54        | 35.8           |
| House wife                 | 57        | 64.2           |
| Birth weight               |           |                |
| Normal (≥ 2.5 kg)          | 136       | 90.1           |
| Under birth (< 2.5 kg)     | 15        | 9.9            |
| Gravidia                   |           |                |
| Primigravidia              | 65        | 43             |
| Multigravidia              | 86        | 57             |

The minimum vs maximum value of cord WBC, cord RBC, and cord platelet among study participants were (3.6 vs 27) x10^6/L, (3.02 vs 5.7) x10^{12}/L, (132 vs 500) x10^9/l, respectively. As a result, summarized in Table 2 sex and delivery type had no influence for cord hematological values (p-value > 0.05). The median value of cord WBC, RBC, Hgb, and Hct among male newborn babies were 12.6, 4.7, 16.2 and 49.6, respectively. On the other hand, the median value WBC, RBC, Hgb, and Hct among female newborn babies were 13.15, 4.6, 15.75, and 48.65, respectively (Table 2).
Table 2
The influence of sex and delivery type on hematological parameter in newborn babies based on median value

| Hematological parameter | Sex | Delivery type | P-value | Vaginal | Cesarean | p-value |
|-------------------------|-----|---------------|---------|---------|----------|---------|
|                         | Male| Female | P-value | Vaginal | Cesarean | p-value |
| WBC(x10^9/l)            | 12.6| 13.15  | 0.98    | 12.6    | 13.1     | 0.64    |
| RBC(x10^12/l)           | 4.7 | 4.6    | 0.51    | 4.61    | 4.73     | 0.27    |
| Hgb (g/dl)              | 16.2| 15.75  | 0.47    | 15.8    | 16.6     | 0.09    |
| Hct (%)                 | 49.6| 48.65  | 0.44    | 48.6    | 50.45    | 0.05    |
| Platelet (x10^9/l)      | 292 | 271    | 0.17    | 276     | 274      | 0.62    |
| Lymphocyte absolute (x10^9/l) | 4.5 | 4.6    | 0.12    | 4.6     | 4.55     | 0.91    |
| Neutrophil absolute (x10^9/l) | 6.4 | 7.25   | 0.19    | 7       | 6.95     | 0.57    |

Hematological reference interval

The newborn babies hematological RI is determined based on non-parametric estimation methods. Based on the SPSS statistical analysis of newborn laboratory data did not require partition by sex groups. As clearly mentioned above Table 2 showed that all hematological value did not have a statistically significant difference of means among sex groups as well as the route of delivery types. According to clinical laboratory approve guideline partitioning was required when the difference between the observed means of two subclasses is statistically significant at the 5% probability level (9). The median and 95% of RI of RBC, Hgb, Hct, MCV, MCH and MCHC were 4.64 (3.69–5.47 × 10^12/l), 16 (13.32–19.64), 49.1% (39.42–58.06%), 105.85 fl (91.6–113.22 fl), 35 pg (30.48–38.02 pg) and 33.1 g/dl (31.48–36.5 g/dl), respectively. The lower and upper limit WBC and platelets were 7.64, 22.16, and 132.74, 413.4, respectively (Table 3).
Table 3
Complete blood count parameter among full-term newborn babies at gynecology and obstetrics, University of Gondar compressive specialized hospital, 2019 (N = 151)

| CBC parameter                     | Median (IQR)     | Lower limit (2.5th) | Upper limit (97.5th) | RI (95th percentile) |
|-----------------------------------|------------------|---------------------|----------------------|----------------------|
| Total WBC (x10^9/l)               | 12.8 (10.5–15.4) | 7.64                | 22.16                | 7.64–22.16           |
| RBC (x10^{12}/l)                  | 4.64 (4.25–4.89) | 3.69                | 5.47                 | 3.69–5.47            |
| Hgb (g/dl)                        | 16 (14.8–17)     | 13.32               | 19.64                | 13.32–19.64          |
| Hct (%)                           | 49.1 (44.9–51.8) | 39.42               | 58.06                | 39.42–58.06          |
| MCV (fl)                          | 105.85 (102.6–109.6) | 91.6            | 113.22               | 91.6–113.22          |
| MCH (pg)                          | 35 (33.7–35.7)   | 30.48               | 38.02                | 30.48–38.02          |
| MCHC (g/dl)                       | 33.1 (32.5–33.5) | 31.48               | 36.5                 | 31.48–36.5           |
| Platelet (x10^9/l)                | 276 (233–314)    | 132.74              | 413.4                | 132.74–413.4         |
| Lymphocyte percentage (%)         | 35.9 (30–41)     | 14.22               | 58.32                | 14.22–58.32          |
| Mix percentage (%)                | 10.3 (7.9–12.5)  | 3.34                | 18.64                | 3.34–18.64           |
| Neutrophil percentage (%)         | 53.5 (48.5–60.7) | 36.74               | 79.48                | 36.74–79.48          |
| Lymphocyte absolute (x10^9/l)     | 4.6 (3.6–5.5)    | 2.16                | 10.3                 | 2.16–10.3            |
| Mix absolute (x10^9/l)            | 1.3 (0.9–1.7)    | 0.4                 | 2.94                 | 0.4–2.94             |
| Neutrophil absolute (x10^9/l)     | 7 (5.4–8.9)      | 2.96                | 13.54                | 2.96–13.54           |
| RDW-SD                            | 68.1 (64.7–72.1) | 55.48               | 81.3                 | 55.48–81.3           |
| Platelet distribution width (PDW) | 11.3 (10.5–12.2) | 8.8                 | 15.68                | 8.8–15.68            |
| Mean platelet volume (MPV)        | 9.5 (9.1–10)     | 7.88                | 11.04                | 7.88–11.04           |

Discussion
The RI has a great role in the interpretation of hematological test results. The aim of this study was to establish hematological parameters among full-term newborn babies by using UCB specimens which are the first time reported in Ethiopia. The present study showed that mode of delivery had no influence of hematological parameters of the newborn babies which agrees to a study done by Fady M et al reported that there was no significant difference in the MCV, MCH, MCHC, RDW, lymphocytes, and monocytes (15). However, this result is contrasting to a study done by L Glasser et al showed that mode of delivery had influenced to hematological test values.

In the current study, there was no statistically significant mean difference of hematological parameters between males and females which was consistent with a study reported in Sokoto, Northern Nigeria (16) and Lagos, Nigeria (17). On the other hand, Our finding of the 95% RI of RBC values were comparable to a study reported in Nepal (18), Iran (19), and Nigeria (20) reported. The of Hgb values of this study agrees with a study reported in Saudi Arabia, however, the 95th percentiles of platelet value the present study was greater than as compared to the Saudi reported (21). The 95% RI of WBC in our study similar to the study conducted in Turkey (7.64–22.16 × 10⁹/l versus 71.90–25.44 × 10⁹/l) (22).

The lower limit WBC value of this study was lower but the MCH RI were similar to a study done in Saudi Arabia (21), Nepal (18), and Pakistan (23). The Hct value of the current study was comparable to a study conducted in Logos, Nigeria and Iran. However, the lower limit of WBC was higher than a study reported in Sagamu, Nigeria, and Iran (Table 4). This may be due to the variation in geographical location, and race or genetic factors may be contributing to the presence of this difference.
Table 4
Comparison of the current study to other similar findings based on cord blood hematological parameters

| Hematological parameter | RI | Current study | Saudi Arabia (21) | Nepal (18) | Pakistan (23) | Lagos, Nigeria (17) | Mashhad, Iran (19) | Iraq (24) | Sagamu, Nigeria (20) |
|-------------------------|----|---------------|-------------------|------------|---------------|---------------------|-------------------|-----------|---------------------|
| Total WBC               |    | 7.64–22.16    | 10.9–21.5         | 10.49–19.4 | 9.7–17.7      | 7.9–18.3            | 5.16–18.2        | 7.32–12.92 | 4.2–25.8            |
| RBC                     |    | 3.69–5.47     | 5.1–16.3          | 3.67–4.93  | -             | 3.52–4.62           | 3.61–5.29        | 3.53–4.47  | 4.3–5.9             |
| Hgb                     |    | 13.3–19.6     | 15.7–19.7         | 13.28–17.2 | 13.5–17.3     | 11.8–14.8           | 13–18.8          | 12.3–15.22 | 13–17.8             |
| Hct                     |    | 39.4–58.1     | 46.9–59.5         | -          | -             | 39–50.6             | 39.6–56.9        | 39.7–49.2  | 39.1–53.5            |
| MCV                     |    | 91.6–113.2    | 97.9–112.3        | 95.2–107.2 | 103.8–108     | 98.5–122.2          | 97.5–119.8       | 105.5–117.7 | 84.4–98.8           |
| MCH                     |    | 30.5–38.02    | 30.2–40.6         | 31.6–36.2  | 32.2–35.4     | 28.5–36.7           | 31.7–40          | 32.1–36.8  | 27.9–33.1            |
| MCHC                    |    | 31.48–36.5    | 27–40.2           | 31.7–34.9  | -             | 28.1–31.4           | 30.1–35.2        | 29.03–32.86 | 31.7–34.5           |
| Platelet                |    | 132.7–413     | 178.2–297         | 165.6–288.2 | 223–347     | 152.9–297.3         | 131–383          | 207–328   | 129–607             |
| Lymphocyte %            |    | 14.2–58.3     | 16.5–37.1         | 23.7–46.7  | -             | -                   | -                | 29.6–49.98 | 27.8–49.6           |
| Neutrophil percentage   |    | 36.7–79.5     | 50.3–74.1         | 51.5–74.7  | -             | -                   | -                | 39.8–62.2  | 45.9–67.7           |
| Lymphocyte absolute     |    | 2.16–10.3     | -                  | -          | 3.3–6.9       | -                   | 1.14–8.46        | -          | -                   |
| Neutrophil absolute     |    | 2.96–13.54    | -                  | -          | 4.7–10.7      | -                   | 1.12–10.2        | -          | -                   |

The 95th percentile RI of the current study lower limit of RBC was lower than studies conducted in Saudi Arabia and Sagamu Nigeria. On the contrarily, the higher limit of 95th percentile RI of Hgb value of this study was higher than is a study was done in Logas Nigeria, Iran, Iraq and Sagamu Nigeria. The difference of Hgb value might be due to method variation, inclusion criteria of the study participants and
time of cord clamping. Similarly, the higher limit of 95th percentile RI of both WBC and platelet values were higher than in the study conducted in Nepal, Pakistan, Logas Nigeria, Iran and Iraq (17–19, 23, 24).

The platelet counts of this study range from (132.7–413) x10^9/L. Based on our finding less than 132 × 10^9/L is thrombocytopenia and greater than 413 × 10^9/L is thrombocytosis for newborn babies. The lower limit of platelet counts in the current study were consistent to a study reported from Mashhad, Iran (19) (131 × 10^9/L). However, the higher limit of the current study was higher in this study done by Marwaha et al (142 × 10^9/L) (25). The major limitation of this study was nucleated RBC is not corrected when total WBC was counted at the time of analysis.

**Conclusion**

Hematological RI was established from full-term newborn babies at University of Gondar compressive specialized hospital Northwest Ethiopia were different from African and Europeans. Each Hematological RI was no statistically significant among male and female newborn babies. The current finding of RI is very helpful for the physicians to accurately diagnose and monitoring of therapies of the patients.

**Abbreviations**

CBC
Complete Blood Count, Hct:Hematocrit, Hgb:Hemoglobin, MCH:Mean Cell Hemoglobin, MCHC:Mean Cell Hemoglobin Concentration, MCV:Mean Cell Volume, MPV:Mean platelet volume, PDW:Platelet distribution width, RBC:Red Blood Cell, RDW:Red Cell Distribution Width, RI:Reference Interval, WBC:White Blood Cell.

**Declarations**

**Ethical approval and consent to participate**

The research was conducted after obtaining an ethical clearance letter from the School of Biomedical and Laboratory Science Ethical Review Committee. A permission letter was obtained from the University of Gondar compressive specialized hospital. Written informed consent was obtained from each selected mother after explaining the aim of the study with the potential adverse effects during sample collection. The results of the study participants were used without personal identifiers and only used for research purpose.

**Consent for publication**

Not applicable

**Availability of data and material**
The datasets used and/or analyzed during the current study available from the corresponding author on reasonable request.

**Competing of interest**

The authors declare that they have no competing interests.

**Funding**

The author(s) received no specific funding for this work.

**Authors contribution**

TT involved in the study design, data collection, laboratory analysis, result interpretation and drafting of the manuscript. TK involved in statistical analysis, editing and reviewing of the manuscript. All authors read and approved the final manuscript.

**Acknowledgments**

The authors would like to acknowledge University of Gondar comprehensive specialized hospital, study participants, data collectors, staffs of gynecology and obstetrics ward for their collaboration and support during the process of data collection.

**References**

1. Katsares V, Paparidis Z, Nikolaidou E, Karvounidou I, Ardelean K-A, Drossas N, et al. Reference ranges for umbilical cord blood hematological values. Laboratory Medicine. 2009;40(7):437–9.

2. Wiedmeier SE, Henry E, Sola-Visner M, Christensen R. Platelet reference ranges for neonates, defined using data from over 47 000 patients in a multihospital healthcare system. Journal of perinatology. 2009;29(2):130–6.

3. Wall D. Umbilical cord blood: importance of supporting public banks. 2011. p. 638.

4. Carroll P, Nankervis C, Iams J, Kelleher K. Umbilical cord blood as a replacement source for admission complete blood count in premature infants. J Perinatol. 2012;32(2):97.

5. Slhessarenko N, Andriolo A. The importance of determining reference intervals for Laboratory Medicine. Jornal Brasileiro de Patologia e Medicina Laboratorial. 2016;52(2):68–9.

6. Zech NH, Broer N, Ribitsch I, Zech MH, Broer K-h, Ertan K, et al. The rationale behind collecting umbilical cord blood. Journal of the Turkish German Gynecological Association. 2010;11(2):99.
7. Xanthou M. Leucocyte blood picture in healthy full-term and premature babies during neonatal period. Arch Dis Child. 1970;45(240):242–9.

8. Noguera NI, Detarsio G, Perez SM, Bragos IM, Lanza O, Rodriguez JH, et al. Hematologic study of newborn umbilical cord blood. MEDICINA-BUENOS AIRES-. 1999;59:446–8.

9. Sasse EA. How to define and determine reference intervals in the clinical laboratory; approved guideline. NCCLS documents C28-A2. 2000.

10. Friedrichs KR, Harr KE, Freeman KP, Szladovits B, Walton RM, Barnhart KF, et al. ASVCP reference interval guidelines: determination of de novo reference intervals in veterinary species and other related topics. Veterinary clinical pathology. 2012;41(4):441–53.

11. Castellone D. Establishing reference intervals in the coagulation laboratory. International journal of laboratory hematology. 2017;39:121–7.

12. Horowitz G, Altaie S, Boyd J, Ceriotti F, Garg U, Horn P. Defining, establishing, and verifying reference intervals in the clinical laboratory; approved guideline, CLSI document C28-A3. Clinical and Laboratory Standards Institute, Wayne, PA. 2008.

13. Jones G, Barker A. Reference intervals. The Clinical Biochemist Reviews. 2008;29(Suppl 1):93.

14. Katayev A, Balciza C, Seccombe DW. Establishing reference intervals for clinical laboratory test results: is there a better way? Am J Clin Pathol. 2010;133(2):180–6.

15. El Gendy FM, Allam AA, Allam MM, Allam RK. Haematological parameters of newborns delivered vaginally versus caesarean section. Menoufia Medical Journal. 2016;29(2):259.

16. Imoru M, Momodu I, Buhari H, Nuhu A. Haematological Reference Values for Full Term Healthy Neonates Delivered Within 24 Hours in Sokoto, Northern Nigeria. International Journal of Hematology Therapy. 2016;2(2):0-.

17. Adewumi A, Adeyemo TA, Akinsegun AA, Abidoye G, Ebele U, Sulaimon AA. Cord blood full blood count parameters in Lagos, Nigeria. Pan African Medical Journal. 2014;17(1).

18. Basnet S, Singh SK, Sathian B, Mishra R. Reference Ranges for Hematological Values in Umbilical Cord Blood in Pokhara, Nepal. Journal of Nepal Paediatric Society. 2016;36(2):160–4.

19. Keramati MR, Mohammadzadeh A, Farhat AS, Sadeghi R. Determination of Hematologic Reference Values of Neonates in Mashhad-Iran. International Journal of Hematology Oncology. 2011;28(4):101–5.

20. Ogundeyi M, Olarewaju D, Njokanma O, Ogunlesi T. Haematological profile of apparently healthy term babies aged one day, three days and six weeks delivered in Sagamu, Nigeria. Nigerian Journal of Paediatrics. 2011;38(3):125–30.

21. Alharbi S, Alkhotani A. Hematological reference values for full-term, healthy newborns of Jeddah, Saudi Arabia. Journal of Clinical Neonatology. 2017;6(1):19.

22. Özyürek E, Cetintaş S, Ceylan T, Öğüş E, Haberal A, Gürakan B, et al. Complete blood count parameters for healthy, small-for-gestational-age, full-term newborns. Clinical Laboratory Haematology. 2006;28(2):97–104.
23. Pasha W, Ali W, Ahmed N, Khattak AL, Idris M, Nayyer ZA. Reference haematological values for full term healthy newborns from rural Sindh, Pakistan. Journal of Ayub Medical College Abbottabad. 2015;27(2):375–7.

24. Al-Marzoki JM, Al-Maaroof ZW, Kadhum AH. Determination of reference ranges for full blood count parameters in neonatal cord plasma in Hilla, Babil, Iraq. Journal of blood medicine. 2012;3:113.

25. Marwaha R, Narang A, Thusu K, Garewal G, Bhakoo O. Routine hematological values in term newborns. Hemoglobin. 1992;16:12–21.

**Supplementary Files**

This is a list of supplementary files associated with this preprint. Click to download.

- Additionalfile1.docx