Basic Study

Protective effect of bicyclol against bile duct ligation-induced hepatic fibrosis in rats

Yong-Zhan Zhen, Na-Ren Li, Hong-Wei He, Shuang-Shuang Zhao, Guang-Ling Zhang, Xiao-Fang Hao, Rong-Guang Shao

Correspondence to: Hong-Wei He, Associate Professor, Institute of Medicinal Biotechnology, Chinese Academy of Medical Sciences and Peking Union Medical College, No. 1, Tian Tan Xi Li, Dongcheng District, Beijing 100050, China. hehwei@imb.pumc.edu.cn
Telephone: +86-10-83166673
Fax: +86-10-63017302

Received: January 3, 2015
Peer-review started: January 4, 2015
First decision: January 22, 2015
Revised: February 13, 2015
Accepted: April 9, 2015
Article in press: April 9, 2015
Published online: June 21, 2015

Abstract

AIM: To evaluate the protective effect of bicyclol against bile duct ligation (BDL)-induced hepatic fibrosis in rats.

METHODS: Sprague-Dawley male rats underwent BDL and sham-operated animals were used as healthy controls. The BDL rats were divided into two groups which received sterilized PBS or bicyclol (100 mg/kg per day) orally for two consecutive weeks. Serum, urine and bile were collected for biochemical analysis and a whole genome oligonucleotide microarray assay. Reverse transcription-polymerase chain reaction and Western blotting were used to verify the expression of liver fibrosis-related genes.

RESULTS: Treatment with bicyclol significantly reduced liver fibrosis and bile duct proliferation after BDL. The levels of alanine aminotransferase (127.7 ± 72.3 vs 230.4 ± 69.6, \( P < 0.05 \)) and aspartate aminotransferase...
transferase (696.8 ± 232.6 vs 1032.6 ± 165.8, \( P < 0.05 \)) were also decreased by treatment with bicyclol in comparison to PBS. The expression changes of 45 fibrogenic genes and several fibrogenesis-related pathways were reversed by bicyclol in the microarray assay. Bicyclol significantly reduced liver mRNA and/or protein expression levels of collagen 1a1, matrix metalloproteinase 2, tumor necrosis factor, tissue inhibitors of metalloproteinases 2, transforming growth factor-\( \beta 1 \) and \( \alpha \)-smooth muscle actin.

**CONCLUSION:** Bicyclol significantly attenuates BDL-induced liver fibrosis by reversing fibrogenic gene expression. These findings suggest that bicyclol might be an effective anti-fibrotic drug for the treatment of cholestatic liver disease.

**Key words:** Bicyclol; Rat; Bile duct ligation; Liver fibrosis; Gene expression profile

© The Author(s) 2015. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Cholestasis often fails to respond to medical therapy, resulting in liver necrosis, fibrosis, cirrhosis and subsequent liver failure. Bicyclol has significant liver protective effects but little is known about the effect of this drug on cholestatic liver fibrosis. Therefore, the present study was designed to investigate the protective effects of bicyclol in a rat model of liver fibrosis induced by bile duct ligation (BDL). Bicyclol can improve liver function after BDL and decrease bile duct proliferation, inflammation and fibrosis. Bicyclol might be an effective anti-fibrotic drug for the treatment of cholestatic liver disease.

Zhen YZ, Li NR, He HW, Zhao SS, Zhang GL, Hao XF, Shao RG. Protective effect of bicyclol against bile duct ligation-induced hepatic fibrosis in rats. *World J Gastroenterol* 2015; 21(23): 7155-7164 Available from: URL: http://www.wjgnet.com/1007-9327/full/v21/i23/7155.htm DOI: http://dx.doi.org/10.3748/wjg.v21.i23.7155

**INTRODUCTION**

Biliary obstruction is characterized by portal tract expansion, bile duct proliferation, hepatic stellate cell activation, extracellular matrix accumulation, liver fibrosis and subsequent cirrhosis. Cirrhosis, which is the end stage of progressive fibrosis, is a major health problem worldwide and is characterized by the accumulation of extracellular matrix proteins and the distortion of the hepatic architecture. Current evidence suggests that the process of hepatic fibrosis is driven by a complex network of cytokines. In particular, the fibrogenic cytokine transforming growth factor-beta (TGF-\( \beta \)) is central to the development of cirrhosis due to its stimulating effect on matrix protein generation and its inhibitory effect on matrix protein removal. TGF-\( \beta \) plays a pivotal role in the development of hepatic fibrosis and cirrhosis, therefore, the inhibition of TGF-\( \beta \) has been used in several therapeutic approaches to prevent hepatic fibrosis. However, no clinically effective treatment for liver fibrosis is available and an effective therapeutic approach that blocks the development of hepatic fibrosis is needed.

Although much has been learned in recent decades about the molecular basis of cholestasis and the pathophysiology of hepatic fibrosis, the mechanisms that underlie the development of cholestatic liver disease have not been well elucidated and new therapeutic approaches have been limited. Few effective treatments for cholestasis are currently approved by the Food and Drug Administration; these treatments include ursodeoxycholic acid (UDCA) which achieves limited benefits during the early stages of primary biliary cirrhosis. Alternative therapies, including the combination of UDCA with immunomodulatory drugs, have been attempted but have not been successful in limited clinical trials. Therefore, new therapeutic agents are needed for these chronic cholestatic diseases.

Bicyclol (4,4’-dimethoxy-5,6,5′,6′-dimethylenedioxy-2-hydroxymethyl-2′-carbonyl biphenyl) is a synthetic drug that was developed by Chinese scientists and is widely used in the clinic to treat patients with chronic hepatitis B and C. Pharmacologically, bicyclol exhibits anti-hepatitis virus activity in duck viral hepatitis and in the 2.2.15 cell line. Furthermore, bicyclol also exhibits a protective effect against liver injury induced by hepatotoxins in mice and rats. Recent studies demonstrated that bicyclol markedly attenuates experimental liver fibrosis induced by chemical toxins, including carbon tetrachloride (CCL\(_4\)) and dimethylnitrosamine, in mice and significantly protects against chemotherapeutic agent-induced liver damage in elderly patients with cancer. These results suggest that bicyclol has a chemopreventive effect on liver injury and fibrogenesis. Therefore, the present study was designed to investigate the effect of bicyclol on cholestatic liver fibrosis induced by bile duct ligation (BDL) and the possible mechanisms that underlie this effect in rats.

**MATERIALS AND METHODS**

**Materials**

Bicyclol was purchased from Beijing Union Pharmaceutical Factory (Beijing, China). Antibodies that target TGF-\( \beta 1/2 \) were purchased from Epitomics (Burlingame, CA). Antibodies specific for \( \alpha \)-smooth muscle actin (\( \alpha \)-SMA) and GAPDH were purchased from Sigma-Aldrich (St. Louis, MO). The TaqMan real-time polymerase chain reaction (RT-PCR) master mix was purchased from Roche (Indianapolis, IN). ABI TaqMan primer/probes were purchased from Applied
Biosystems (Foster City, CA).

Animal experiments
Male adult Sprague-Dawley rats weighing 180-220 g were purchased from the Institute of Laboratory Animal Science (Chinese Academy of Medical Sciences, Beijing, China). The rats were maintained in a temperature-controlled room (22 ± 2 °C) with a 12 h light/dark cycle and a relative humidity of 40%-60%. The rats were given access to food and water ad libitum. All animal experiments were approved by the Institutional Animal Care and Use Committee of Hebei United University and the animal protocol was designed to minimize the pain and discomfort of the animals. For experiments using BDL, the rats were randomly divided into three groups of six animals. Each day, six animals underwent BDL and three sham-operated animals were used as a healthy control. Twenty-four hours after surgery, the BDL animals were randomly assigned to receive a daily gavage consisting of PBS (i.e., the treatment control and solvent) or 100 mg/kg of bicyclol suspended in PBS for 14 d. The sham controls also received PBS by gavage. All BDL rats received 0.1 mL of vitamin K (100 μg/mL) every other day via subcutaneous injection. The body weight was measured daily. The animals were sacrificed in random order between 9:00 am and 11:00 am after an overnight fast. Samples of serum, bile (from bile duct cysts in BDL animals), urine (the urine of each rat was collected during an overnight fast using a metabolic cage) and liver were collected for further analyses, as described previously.

Serum biochemistry and liver histology
Serum alanine transaminase (ALT), aspartate transaminase (AST), total cholesterol, high-density lipoprotein (HDL) cholesterol and low-density lipoprotein (LDL) cholesterol, total bile acid and total bilirubin of serum, bile and urine were analyzed using a Hitachi 7170 chemistry analyzer and kits from Zhongsheng Biotechnology (Beijing, China). Formalin-fixed liver tissue was embedded in paraffin and sections were stained with hematoxylin and eosin and Sirius red. Histological evaluation of the liver sections was performed by a single pathologist who assessed for bile duct proliferation, inflammation and fibrosis on a 1 to 4+ scale in a blinded manner. Hydroxyproline was assayed. Calculations were then made and statistical analysis was performed between the groups.

Western blot analysis
The homogenized liver tissues were lysed in RIPA buffer (50 mmol/L Tris-HCl, pH 7.5, 1% NP-40, 0.1 mmol/L NaCl, 1 mg/mL of aprotinin, 1 mmol/mL of leupeptin, 1 mmol/L NaVO₄ and 1 mmol/L NaF) for 30 min at 4 °C. Cell debris was removed by centrifugation at 12000 × g for 20 min at 4 °C. Protein concentrations were determined using the BCA assay. Equal amounts of lysate were resolved via SDS-polyacrylamide gel electrophoresis and transferred to a PVDF membrane (Millipore, Bedford, MA). The membranes were blocked with 5% nonfat milk in TBS-T buffer at room temperature for 1 h and then incubated for 2 h or overnight with primary antibodies. After three 10 minute washes with 0.1% TBS-T buffer, the membranes were incubated for 1 h at room temperature with a species-specific horseradish-peroxidase-linked secondary antibody (Cell Signaling Technology). After three washes with 0.1% TBS-T buffer, the membranes were soaked in enzyme-linked chemiluminescence detection reagent (Millipore) and protein bands were visualized using the Fluorchem HD2 imaging system (Protein simple, Santa Clara, CA).

Statistical analysis
The data are presented as the mean ± SD. The significance of differences between experimental groups was analyzed using the Lowess method and only those genes that exhibited a consistent alteration tendency (both ≥ 1.5-fold) in both microarrays were selected as differentially expressed genes. Fibrogenesis-related genes were selected from the differentially expressed genes (Ratio ≥ 1.5) by searching in PubMed using the gene description/symbol and liver fibrosis as keywords.
Bicyclol significantly improves the hepatic morphology of BDL rat livers

After hematoxylin and eosin staining, the histological structure of the liver was observed to be normal in the sham group, as shown in Figure 1A. Portal inflammation, parenchymal necrosis and ductal proliferation, including bridging of the portal tracts, were observed in the BDL group. Portal inflammation and ductal proliferation were markedly reduced in the bicyclol-treated group compared to the BDL group. A blinded assessment also revealed significantly lower scores for bile duct proliferation and inflammation in the bicyclol-treated groups compared to the BDL group (Figure 1B).

**Bicyclol reduces the level of liver fibrosis in BDL rat livers**

After Sirius red staining, liver damage was visualized based on the hyperplasia of the lattice fibers and collagenous fibers in the portal area; this damage extended outwards in BDL group rats. These liver histopathological changes were improved in bicyclol-treated rats (Figure 2A). A blinded assessment also revealed significantly lower scores for the level of fibrosis in the bicyclol-treated groups compared to the BDL group. This finding was further confirmed using a hydroxyproline assay (Figure 2B and C).

**Whole genome analysis of the effect of bicyclol on liver fibrosis**

To analyze genome-wide alterations in gene expression, total RNA was extracted from sham, BDL and bicyclol-treated group rat livers for whole genome microarray experiments. For our analysis, a 1.5-fold alteration in the signal intensity was regarded as a significant change in mRNA expression. We found that 1060 genes were down-regulated and 1334 genes were up-regulated in BDL rat livers compared with sham rat livers. In addition, 236 genes were down-regulated and 149 genes were up-regulated in bicyclol-treated rat livers compared with BDL rat livers (Figure 3A). We found that a total of 180 genes were included in both sets of significantly changed genes. In addition to analyzing the effects of bicyclol treatment, we performed a gene functional enrichment analysis using the DAVID Bioinformatics Resources[22,23] to further characterize those 180 genes. The results demonstrated that these genes are related to ECM-receptor interactions, fatty acid metabolism, the PPAR signaling pathway, focal adhesion, the biosynthesis of unsaturated fatty acids, oxidation/reduction and the integrin signaling pathway, among other processes (Table 2). We also observed that among those 180 genes, 45 genes were reported to be associated with liver fibrosis, including Spp1, collagen 1a1 (Col1a1), matrix metalloproteinase 2 (Mmp2), α-SMOOTH MUSCLE ACTIN (α-Sma, Acta2) and other genes (Figure 3B).

**Bicyclol represses the expression of liver fibrogenic genes**

To verify the anti-fibrotic effects of bicyclol and the microarray data, some fibrogenic genes were assayed using TaqMan real-time PCR and/or Western blot. Our results demonstrated that bicyclol significantly

---

**Table 1  Serum, urine and bile biochemistry of bile duct ligation rats**

|                      | Sham-PBS (n = 6) | BDL-PBS (n = 6) | BDL-Bic100 (n = 6) |
|----------------------|------------------|-----------------|-------------------|
| Serum ALT (U/L)      | 29.3 ± 3.8       | 230.4 ± 69.6    | 127.7 ± 72.3**    |
| Serum AST (U/L)      | 98.3 ± 18.1      | 1032.6 ± 165.8* | 696.8 ± 232.6**   |
| Serum total bilirubin (μmol/L) | 0.1 ± 0.2 | 161.5 ± 19.2    | 152.4 ± 41.8*     |
| Serum total cholesterol (mmol/L) | 1.5 ± 0.5 | 3.0 ± 0.6*      | 3.1 ± 0.7*        |
| Serum LDL (mmol/L)   | 0.2 ± 0.0        | 1.5 ± 0.3*      | 1.8 ± 0.3*        |
| Serum HDL (mmol/L)   | 0.9 ± 0.2        | 0.3 ± 0.2*      | 0.3 ± 0.1*        |
| Serum total bile acid (μmol/L) | 8.0 ± 3.0 | 173.0 ± 38.0*   | 172.6 ± 39.1*     |
| Urine total bilirubin (μmol/L) | 3.4 ± 1.5 | 247.2 ± 30.9*   | 246.5 ± 59.6*     |
| Urine total cholesterol (μmol/L) | 0.3 ± 0.7 | 100.4 ± 27.5*   | 105.8 ± 58.0*     |
| Bile total bile acid (μmol/L) | Not applicable | 3183.5 ± 908.6  | 3261.0 ± 488.8    |
| Bile total bilirubin (μmol/L) | Not applicable | 96.3 ± 35.9     | 126.3 ± 46.9      |

In the sham group, rats underwent laparotomy. In the bile duct ligation (BDL) and bicyclol groups, rats were submitted to undergo BDL operative manipulation and treated with PBS or bicyclol for two weeks. The levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST) in serum were expressed as U/L. The levels of total bilirubin, total bile acid in the serum, bile and urine were significantly increased after 2 wk of BDL. In addition, the concentration of HDL in the serum was reduced. These changes are classical of BDL rat livers significantly reduced these values.

The concentration of serum total cholesterol and LDL, total bilirubin and total bile acid in the serum, bile and urine were significantly increased after 2 wk of BDL. In addition, the concentration of LDL in the serum was reduced. These changes are classical markers of cholestasis in BDL animals. Unexpectedly, pharmacological treatment with bicyclol failed to change the levels of these markers (Table 1).

was assessed using the two-tailed Student’s t-test. The value of statistical significance was set at P < 0.05.

**RESULTS**

**Effect of bicyclol on the serum, urine and bile biochemistry of BDL rats**

The results of the serum, urine and bile biochemistry are shown in Table 1. The activities of ALT and AST in the sham-operated rats remained within control values. Serum levels of ALT and AST were increased by approximately eight- and ten-fold, respectively, after bile duct ligation in comparison to the values of sham-operated rats. A 100 mg/kg dose of bicyclol in BDL rats significantly reduced these values.

The concentration of serum total cholesterol and LDL, total bilirubin and total bile acid in the serum, bile and urine were significantly increased after 2 wk of BDL. In addition, the concentration of HDL in the serum was reduced. These changes are classical markers of cholestasis in BDL animals. Unexpectedly, pharmacological treatment with bicyclol failed to change the levels of these markers (Table 1).

**Bicyclol significantly improves the hepatic morphology of BDL rat livers**

After hematoxylin and eosin staining, the histological structure of the liver was observed to be normal in the sham group, as shown in Figure 1A. Portal inflammation, parenchymal necrosis and ductal proliferation, including bridging of the portal tracts, were observed in the BDL group. Portal inflammation and ductal proliferation were markedly reduced in the bicyclol-treated group compared to the BDL group. A blinded assessment also revealed significantly lower scores for bile duct proliferation and inflammation in the bicyclol-treated groups compared to the BDL group (Figure 1B).
reduces liver mRNA expression of Col1a1, Mmp2, tumor necrosis factor (Tnf) and tissue inhibitors of metalloproteinases 2 (Timp2). Bicyclol also reduced the mRNA levels of Spp1, Timp1 and Tgf-β1, but these differences failed to reach statistical significance (Figure 4A). Based on Western blots, the levels of TGF-β1/2 and α-SMA were increased in the BDL group and decreased in the bicyclol-treated groups (Figure 4B).

DISCUSSION

Cholestasis is caused by dysfunction of the hepatobiliary system which results from either a functional defect in bile formation or an impairment in bile secretion and flow, leading to the intracellular accumulation of bilirubin, toxic bile salts and cholesterol in the blood causing hepatocellular injury[24]. Cholestasis has been shown to stimulate hepatic stellate cells to express collagen, leading to the accumulation of extracellular matrix protein and progressive fibrogenesis. Chronic cholestasis eventually results in liver fibrosis, cirrhosis and subsequent liver failure[25].

In this report, we assessed the therapeutic effects of bicyclol in a BDL cholestatic rat model. Our results demonstrated that bicyclol significantly protects against BDL-induced liver injury and fibrosis, as evidenced by the reduction of elevated serum AST and ALT activities and the reduction of liver inflammation, fibrosis and bile duct proliferation.

The presence of cirrhosis indicates a perturbation of liver homeostasis and the intracellular release of cytokines and signaling molecules occurs during
the development of cirrhosis. The mechanisms that underlie the development of cholestatic liver disease are not well understood. Current evidence suggests that the process of hepatic fibrosis is driven by a complex network of cytokines\cite{26-30}. TGF-\(\beta\)1 was demonstrated to play an important role in the progression of liver fibrosis\cite{31,32}. The release of TGF-\(\beta\)1 may be one of the first signals to activate quiescent hepatic stellate cells (HSCs), resulting in the transdifferentiation of these cells into proliferative, fibrogenic and contractile myofibroblasts. Activated HSCs, which express myogenic markers such as \(\alpha\)-SMA, are considered to be central extracellular matrix (ECM)-producing cells within the injured liver. Concomitant with the increased activity of TGF-\(\beta\) that occurs during liver fibrogenesis, HSCs increase the production and deposition of collagen, leading to progressive scarring and the loss of organ function\cite{33-37}. Thus, a reduction in TGF-\(\beta\) indicates a reduction in liver damage and the down-regulation of \(\alpha\)-SMA indicates the inactivation of HSCs. The results of our whole genome oligonucleotide microarray revealed that some fibrosis-related pathways are regulated by bicyclol. These pathways include ECM-receptor interactions, the PPAR signaling pathway, focal adhesion, oxidation/reduction, the integrin signaling pathway and other pathways. Interestingly, fatty acid metabolism and the biosynthesis of unsaturated fatty acids were also regulated in this assay; this finding indicates that bicyclol might improve fatty acid metabolism in some types of liver disease, such as non-alcoholic fatty liver disease. The microarray data also demonstrated that

**Figure 2** Effects of bicyclol on liver fibrosis of bile duct ligation rats. Bile duct ligation (BDL) group consisted of animals submitted to undergo BDL operative manipulation and sham-operated animals were used as healthy control. BDL rats were divided into two groups, which orally received sterilized PBS or bicyclol [100 mg/(kg·d)] for consecutive 14 d. Liver sections were stained by Sirius red-stained sections (magnification × 200) (A) and quantified by blinded assessment (B). Hydroxyproline was assayed with frozen liver tissues (C). Data are expressed as mean ± SD of 6 animals per group. \(P < 0.05\) vs Sham group; \(P < 0.05\) vs BDL group. Sham: Sham group; BDL: BDL group; BDL-Bic: Bicyclol group.
numerous fibrogenic genes are reversed by bicyclol; some of these genes are potential anti-fibrotic targets, including Spp1\(^{[38,39]}\), Fgfr2\(^{[40,41]}\) and others.

RT-PCR and Western blot verified that bicyclol significantly down-regulates the mRNA and/or protein expression of liver fibrogenic genes, such as Col1a1, Mmp2, Tnf, Timp2 and the protein level of TGF-β1 and α-SMA.

Although our study demonstrated that bicyclol has a protective effect against bile duct ligation (BDL)-induced hepatic injury in rats, pharmacological treatment with bicyclol has no effect on total bile acid and total bilirubin in the serum, urine and bile. Microarray assays also demonstrated that bicyclol failed to regulate the bile acid synthesis pathway and transport protein expression; these findings were consistent with the data obtained from biochemical assays that indicated that bicyclol represses liver fibrosis through fibrogenesis-related pathways or targets but not through bile acid-related pathway or targets. This drug has the potential to repress liver fibrogenesis in the context of most types of chronic liver injury.

In summary, we demonstrated that bicyclol treatment improves liver condition in vivo in BDL rats. We also reported the whole genome mRNA expression profile of bicyclol-treated rat livers, which was verified using Taqman real-time PCR and Western blot. These findings suggest that bicyclol might be a potential therapy for liver fibrosis in patients with chronic cholestatic disorders. Further research is needed to clarify the molecular targets and related pathways of bicyclol. Such studies might identify potential novel therapeutic targets for liver fibrosis.

---

![Figure 3](https://www.wjgnet.com/)

**Figure 3** Microarray analysis of rat liver tissues. Total RNA was isolated from the liver tissues, then each group of total RNA was mixed with the same amount of total RNA from each animal, then labeled with Cy3/Cy5 and measured by microarray assay. Red: increase in gene expression; green: decrease in gene expression; A: Number of genes significant changed in microarray assays; B: 45 fibrogenesis related genes reversed by bicyclol treatment.
Table 2  Bile duct ligation-induced pathways reversed by bicyclol treatment

| Pathway                                      | Relate genes                                      | P value   |
|----------------------------------------------|---------------------------------------------------|-----------|
| ECM-receptor interaction                     | Lamc1, Col4a2, Spp1, Col3a1, Col1a1, Lamb2, Col4a1, Agrn, Itgb4, Tlbs1 | 0.0000003 |
| Fatty acid metabolism                        | Acadsb, Ehhadh, Cyp4a3/4a2, Acod1, Cpt1a, Acot1b | 0.00012   |
| PPAR signaling pathway                       | Ehhadh, Scd, Cyp4a3/4a2, Acod1, Cpt1a, Acot1b    | 0.00017   |
| Focal adhesion                               | Lamc1, Col4a2, Spp1, Col3a1, Col1a1, Lamb2, Col4a1, Itgb4, Tlbs1 | 0.00192   |
| Biosynthesis of unsaturated fatty acids       | Scdf1, Acot2, Scd, Acot1b                         | 0.00267   |
| Oxidation reduction                          | Acadsb, Dicer24, Gprc3, Scd, Rdh12, Akrlb1, Cyp4a3/4a2, Steap4, Cyp3a23/3a1, Loxl11, Ehhadh, Scdf1, Aldh1a2, Cyp3a12,  | 0.00575   |
| Integrin signaling pathway                   | Lamc1, Col4a2, Col3a1, Col1a1, Lamb2, Col4a1, Itgb4, Col12a1 | 0.00793   |
| Retinol metabolism                           | Cyp3a23/3a1, Ugt2a3, Cyp4a3/4a2, Aldh1a2           | 0.03196   |

ECM: Extracellular matrix.

Figure 4  Effects of bicyclol on the fibrogenic gene expression. The mRNA expression levels were assayed by reverse transcription quantitative-polymerase chain reaction with TaqMan gene expression kits (A), Western blot analysis of α-smooth muscle actin (α-SMA) and transforming growth factor (TGF)-β1/2 expression in each bile duct ligation (BDL) rat liver (B) and normalized against Gapdh. Data are expressed as mean ± SD of 6 animals per group. *P < 0.05 vs Sham group; **P < 0.05 vs BDL group. Sham: Sham group; BDL: BDL group; BDL-bic: Bicyclol group.
Bicyclol improves cholestatic liver fibrosis

Zhen YZ et al.

Background
Liver fibrosis is a histological hallmark of liver injury characterized by abnormal deposition of extracellular matrix with subsequent destruction of the normal liver architecture, which has become an important target for pre-cancer prevention and treatment of liver diseases. There are a variety of pathogens that can trigger liver fibrogenesis, including cholestasis, hepatic viral infection, nonalcoholic steatohepatitis and drug toxicity. Among these pathogens, cholestasis is the major cause of liver fibrosis in children and can be induced by bile duct ligation (BDL) in animal models. Current studies have shown that a synthetic drug bicyclol has effective anti-hepatitis virus activity and improves liver injury and fibrogenesis induced by hepatotoxins and chemical toxins. However, no previous studies have evaluated whether bicyclol has any effects on cholestatic liver fibrosis. In this report, we assessed the therapeutic effect of bicyclol after BDL and verified this effect.

Research frontiers
Considerable progress has been made in the research of cholestatic liver diseases. Pharmacological therapy for cholestasis is limited and ursodeoxycholic acid is the only Food and Drug Administration approved medicine for disease-modifying therapy with evidence of efficacy. So, exploring and screening effective and reliable drugs and therapies is urgent.

Innovations and breakthroughs
No studies have previously evaluated the influence of bicyclol on cholestatic liver injury in BDL rats and the molecular targets or related pathways. This study is the first to investigate and confirm the protective effect of bicyclol after BDL. These results indicate that bicyclol is a potential anti-fibrotic drug. Furthermore, this study demonstrated that bicyclol attenuates BDL-induced liver injury by reversing fibrogenic gene expression but not bile acid synthesis and transport. This provides a guide for further exploring molecular targets and/or mechanisms that underlie this effect.

Applications
The study results suggest that bicyclol significantly protected against BDL-induced liver injury and fibrosis, as evidenced by the reduction of elevated serum aspartate transaminase and alanine transaminase activities and reducing liver inflammation, necrosis and bile duct proliferation. The chemopreventive effect of bicyclol is by repressing fibrogenesis-related markers but it failed to regulate bile acid metabolism. It implies that bicyclol might be a potential anti-fibrotic drug for treatment of cholestatic liver disease.

Terminology
BDL is an established model of cholestasis, with complete biliary obstruction and accumulation of multiple primary bile acids, bilirubin and circulating endotoxins, parallel to those that occur in humans with cholestasis. Chronic cholestasis can result in bile duct proliferation, liver fibrosis, cirrhosis and eventually liver failure. Therefore, BDL was used to screen and evaluate drugs for cholestatic liver diseases. Bicyclol is a synthetic drug and is widely used in the clinic to treat patients with chronic hepatitis B and C due to its anti-hepatitis virus activity. Besides, bicyclol exhibits a chemopreventive effect on liver injury and fibrogenesis induced by hepatotoxins and chemical toxins.

Peer-review
The authors evaluated the protective effect of bicyclol on bile duct ligation-induced hepatic fibrosis in rats. The authors report that 14 d post-BDL, rats treated with 100 mg/kg bicyclol had significantly less liver fibrosis and bile duct proliferation. Liver function was partially restored and liver fibrosis was reversed by down-regulation of fibrogenic genes assessed by real-time polymerase chain reaction and microarray assay. This report is well written, the experiments were well done and they corroborate the effect of bicyclol in a different animal model. Minor language polishing is needed.

REFERENCES
1. Nieto JC, Sánchez E, Román E, Vidal S, Oliva L, Guarnier-Arregúe C, Poca M, Torras X, Juárez C, Guarnier C, Soriano G. Cytokine production in patients with cirrhosis and TLR4 polymorphisms. World J Gastroenterol 2014; 20: 17516-17524 [PMID: 25516666 DOI: 10.3748/wjg.v20.i46.17516]
2. Mitra A, Satelii A, Yan J, Xueqing X, Gagea M, Hunter CA, Mishra L, Li S. IL-30 (II72p28) attenuates liver fibrosis through inducing NGK2D-rac1 interaction between NKT and activated hepatic stellate cells in mice. Hepatology 2014; 60: 2027-2039 [PMID: 25351459 DOI: 10.1002/hep.23792]
3. Yang Y, Kim B, Park YK, Koo SI, Lee JY. Astaxanthin prevents TGFβ1-induced pro-fibrogenic gene expression by inhibiting Smad3 activation in hepatic stellate cells. Biochim Biophys Acta 2015; 1850: 178-185 [PMID: 25450180 DOI: 10.1016/j.bbadgen.2014.10.014]
4. Puhe JE, Saiman Y, Friedman SL. Hepatic stellate cells and liver fibrosis. Compr Physiol 2013; 3: 1473-1492 [PMID: 24265236 DOI: 10.1002/cphy.c120035]
5. Lee JH, Jang EJ, Seo HL, Ku SK, Lee JR, Shin SS, Park SD, Kim SC, Kim YW. Sauchinone attenuates liver fibrosis and hepatic stellate cell activation through TGF-β/Smad signaling pathway. Chem Biol Interact 2014; 224C: 58-67 [PMID: 25451574 DOI: 10.1016/j.cbi.2014.10.005]
6. Huang W, Li L, Tian X, Yan J, Yang X, Wang X, Liao G, Qiu G. Astragalus and Paeoniae radix rubra extract inhibits liver fibrosis by modulating the transforming growth factor-β/Smad pathway in rats. Mol Med Rep 2015; 11: 805-814 [PMID: 25373883 DOI: 10.3892/mmr.2014.2868]
7. Reyes-Gordillo K, Segura J, Shibayama M, Tsutsumi V, Vergara P, Moreno MG, Muriel P. Cureumin prevents and reverses cirrhosis induced by bile duct obstruction or CC4 in rats: role of TGF-beta modulation and oxidative stress. Fundam Clin Pharmacol 2008; 22: 417-427 [PMID: 18705752 DOI: 10.1111/j.1472-8206.2008.08611.x]
8. Hohenester S, Oude-Elferink RP, Beuers U. Primary biliary cirrhosis. Semin Immunopathol 2009; 31: 283-307 [PMID: 19603170 DOI: 10.1007/s00281-009-0164-5]
9. Silveira MG, Lindor KD. Treatment of primary biliary cirrhosis: therapy with choleretic and immunosuppressive agents. Clin Liver Dis 2008; 12: 425-443; x-xi [PMID: 18456189 DOI: 10.1016/j.cld.2008.02.008]
10. Chapman R, Fevery J, Kallolo A, Nagorney DM, Boberg KM, Shneider B, Gores GJ. Diagnosis and management of primary sclerosing cholangitis. Hepatology 2010; 51: 660-678 [PMID: 19603170 DOI: 10.1007/s00281-009-0164-5]
11. Zein CO, Lindor KD. Latest and emerging therapies for primary biliary cirrhosis and primary sclerosing cholangitis. Curr Gastroenterol Rep 2010; 12: 13-22 [PMID: 20425480 DOI: 10.1007/s11894-009-0079-2]
12. Tang Y, Hu W, Li Y, Zhang CZ. [Synthesis of metabolites of bicyclol]. Yaoxue Xuebao 2001; 42: 1054-1057 [PMID: 18229611]
13. Xie W, Shi GF, Zhang HF, Zhao GM, Yu ZJ, Lang ZW, Zhao H, Yan J, Cheng J. A randomized, multi-center, controlled study of patients with hepatitis B e antigen-positive chronic hepatitis B treated by adefovir dipivoxil or adefovir dipivoxil plus bicyclol. J Hepatol 2016; 64: 606-612 [PMID: 26194731 DOI: 10.1002/hep.29316]
14. Liu GT. Bicyclol: a novel drug for treating chronic viral hepatitis B and C. Med Chem 2009; 5: 29-43 [PMID: 19149648]
15. Li M, Liu GT. Inhibition of Fas/FasL mRNA expression and TNF-α release in concanavalin A-induced liver injury in mice by bicyclol. World J Gastroenterol 2004; 10: 1775-1779 [PMID: 15188504]
16. Zhao D, Liu G. [Protective effect of bicyclol on concanavalin A-induced liver nuclear DNA injury in mice]. Zhonghua Yi Xue Za Zhi 2001; 81: 844-848 [PMID: 11702677]
17. Yu HY, Wang BL, Zhao J, Yao XM, Gu Y, Li L. Protective effect of bicyclol on tetracycline-induced fatty liver in mice. Toxicology 2009; 261: 112-118 [PMID: 19427351 DOI: 10.1016/j.tox.2009.04.051]
18. Hu QQ, Liu GT. Effects of bicyclol on dimethylthiourea-induced liver fibrosis in mice and its mechanism of action. Life Sci 2006; 79: 606-612 [PMID: 16603200 DOI: 10.1016/j.lfs.2006.02.025]
19. Gu Y, Zhao J, Yao XM, Li Y. Effects of bicyclol on immunological liver fibrosis in rats. J Asian Nat Prod Res 2010; 12: 388-398 [PMID: 20496196 DOI: 10.1080/12220010.2013.8079047]
20. Li XY, Zhou JF, Chen SC, Guan M, Wang YY, Zhao L, Ying HY,
Zhou YP. Role of bicyclol in preventing chemotherapeutic agent-induced liver injury in patients over 60 years of age with cancer. *J Int Med Res* 2014; 42: 906-914 [PMID: 24903556 DOI: 10.1177/0300060514527058]

He H, Mennone A, Boyer JL, Cai SY. Combination of retinoic acid and ursoodeoxycholic acid attenuates liver injury in bile duct-ligated rats and human hepatic cells. *Hepatology* 2011; 53: 548-557 [PMID: 21274875 DOI: 10.1002/hep.24047]

Huang DW, Sherman BT, Tan Q, Kir J, Liu D, Bryant D, Guo Y, Stephens R, Baseler MW, Lane HC, Lemppicki RA. DAVID Bioinformatics Resources: expanded annotation database and novel algorithms to better extract biology from large gene lists. *Nucleic Acids Res* 2007; 35: W169-W175 [PMID: 17576678 DOI: 10.1093/nar/gkm415]

Huang da W, Sherman BT, Lemppicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc* 2009; 4: 44-57 [PMID: 19131956 DOI: 10.1038/nprot.2008.211]

Maillette de Buy Wenniger L, Beuers U. Bile salts and cholestasis. *Dig Liver Dis* 2010; 42: 409-418 [PMID: 20434968 DOI: 10.1016/j.dld.2010.03.015]

Jüngst C, Berg T, Cheng J, Green RM, Jia J, Mason AL, Lammert F. Intrahepatic cholestasis in common chronic liver diseases. *Eur J Clin Invest* 2013; 43: 1069-1083 [PMID: 23927644 DOI: 10.1111/eci.12128]

Pérez-Vargas JE, Zarchi N, Shibayama M, Segovia J, Tsutsumi V, Ornitz DM, Werner S. FGF receptors 1 and 2 control chemically induced injury and compound detoxification in regenerating livers of mice. *Hepatology* 2006; 44: 1267-1277 [PMID: 17058265 DOI: 10.1002/hep.21375]

Lee UE, Friedman SL. Mechanisms of hepatic fibrogenesis. *Best Pract Res Clin Gastroenterol* 2011; 25: 195-206 [PMID: 21497738 DOI: 10.1016/j.bpcg.2011.02.005]

Bataller R, Brenner DA. Liver fibrosis. *J Clin Invest* 2005; 115: 209-218 [PMID: 15690074 DOI: 10.1172/JCI24282]

Whittington PF, Malladi P, Melin-Aldana H, Azzam R, Mack CL, Sahai A. Expression of osteopontin correlates with portal biliary proliferation and fibrosis in biliary atresia. *Pediatr Res* 2005; 57: 837-844 [PMID: 15845635 DOI: 10.1203/01.PDR.0000161414.99181.61]

Lorena D, Darby IA, Gadeau AP, Leen LL, Rittling S, Porto LC, Rosenbaum J, Desmoulière A. Osteopontin expression in normal and fibrotic liver. altered liver healing in osteopontin-deficient mice. *J Hepatol* 2006; 44: 383-390 [PMID: 16221502 DOI: 10.1016/j.jhep.2005.07.024]

Böhm F, Speicher T, Hellerbrand C, Dickson C, Partanen JM, Ornitz DM, Werner S. FGF receptors 1 and 2 control chemically induced injury and compound detoxification in regenerating livers of mice. *Gastroenterology* 2010; 138: 1385-1396 [PMID: 20603121 DOI: 10.1053/j.gastro.2010.06.069]

Nakamura I, Zakharia K, Banini BA, Mikhail DS, Kim TH, Yang JD, Moser CD, Shalhe L, Bowers J, Walters I, Roberts LR. Brivanib attenuates hepatic fibrosis in vivo and stellate cell activation in vitro by inhibition of FGF, VEGF and PDGF signaling. *PLoS One* 2014; 9: e92273 [PMID: 24710173 DOI: 10.1371/journal.pone.0092273]

P- Reviewer: Atta H, Gorrell MD, Panduro A S- Editor: Ma YJ L- Editor: Roennele A E- Editor: Liu XM
