INTRODUCTION

The number of microorganisms that coexist in the human body is 100 trillion, which is 100 times more than the number in human cells. Further, the number of genes in microorganisms is known to be over 100 times. The intestinal microbiome plays a physiological role in digestive function, including the digestion of polysaccharides in the human body, development and differentiation of the intestinal epithelium and immune system, maintenance of tissue homeostasis, resistance to infection, synthesis of vitamins, and balancing of intestinal microbes. Presently, functional and pathological changes occur because of alterations in composition, richness, and/or diversity of the intestinal microbiota, or “dysbiosis”. Although changes in the composition and imbalance of intestinal microflora occur in various diseases, whether these changes are the cause or effect of the disease is unclear. Recently, various attempts, including fecal microbiota transplantation (FMT), have been made to prevent and treat various diseases by controlling intestinal microbial imbalance.

INFLAMMATORY BOWEL DISEASE

Isolation of microbes from the gastrointestinal tract of patients with inflammatory bowel disease (IBD) reveals that the diversity of microbiota is reduced compared to that in healthy people. This phenomenon is seen in both ulcerative colitis (UC) and Crohn’s disease, which are two major IBDs. UC is associated with decreased Akkermansia muciniphila, and Crohn’s disease is associated with decreased Faecalibacterium prausnitzii. In animal IBD models, intestinal microbes cause disease. Further, in Crohn’s disease, if stool flow alters, inflammation of the lower intestine improves. Probiotics are effective in preventing pouchitis in UC, suggesting that the gut microbiota plays an important role in the development of IBD.

In a recent meta-analysis of four double-blind randomized
controlled trials (RCTs) involving patients with UC, clinical remission was observed in 28% of cases in the stool transplant group and 9% of cases in the placebo group. These results suggest that stool transplantation has a moderate therapeutic effect in patients with active UC, but applying the results directly to clinical practice is difficult because of differences in methods across studies.\textsuperscript{10}

The persistence of response to FMT in UC is unclear, with a few studies reporting long-term outcomes.\textsuperscript{11} Two RCTs evaluating remission induction in UC reported 1-year outcomes in patients with primary outcomes at week 8. In a study by Costello et al.,\textsuperscript{12} five of 12 (42%) patients with steroid-free remission (clinical and endoscopic) after donor FMT showed maintenance of remission for 12 months. In a study by Moayyedi et al., eight of nine patients showed maintenance of clinical remission at 9–12 months, but some patients received FMT monthly.\textsuperscript{13} There is no consensus on FMT as a UC treatment, and a recent consensus addressed that FMT should be performed for UC in a clinical trial setting.

**IRRITABLE BOWEL SYNDROME**

The mechanism of irritable bowel syndrome (IBS) development is unclear. A hypothesis to be explained is intestinal dysbiosis.\textsuperscript{14,15} In animal experiments, intestinal bacteria play an important role in digestive tract motility. Germ-free mice show delayed gastric emptying time and intestinal pain, which is normalized by the administration of normal intestinal bacteria.\textsuperscript{16,17} The expressions of Escherichia, Shigella, and Aeromonas increase and those of Actinobacter, Citrobacter, and Microvagula decrease in patients with IBS.\textsuperscript{18} After FMT in 45 patients with IBS with chronic constipation, 60% of patients recovered, and 60% returned to a normal state 9–19 months later.\textsuperscript{19}

A recent systematic review and meta-analysis analyzed five RCTs including 267 patients with IBS, in which 92% of patients had IBS with diarrhea or mixed bowel habits.\textsuperscript{20} FMT failed to improve the symptoms of IBS (relative risk [RR] , 0.98; 95% confidence interval [CI], 0.58–1.66). FMT using donor stool via colonoscopy was superior in two RCTs (RR, 0.63; 95% CI, 0.43–0.93) and via a nasojejunal tube in one RCT (RR, 0.69; 95% CI, 0.46–1.02) compared to those using autologous stool. Further large-scale RCTs are required to reach definitive conclusions.

**NON-ALCOHOLIC FATTY LIVER DISEASE**

The pathogenesis of non-alcoholic fatty liver disease (NAFLD) is unknown, but hepatic focal deposition, liver damage, and imbalance of intestinal bacteria may be involved. Studying microorganisms in the patient's feces using pyrosequencing or quantitative polymerase chain reaction confirmed that the Ruminococcaceae family or Bacteroidetes phylum increased in patients compared to healthy individuals.\textsuperscript{21–23} A randomized controlled blinding method is underway to determine the effect of FMT on regression.\textsuperscript{19} Further research is warranted to identify the effect of FMT on NAFLD.

**METABOLIC SYNDROME AND OBESITY**

Alterations in the composition and metabolic capacity of gut microbiota in obesity promote adiposity and influence metabolic processes in the peripheral organs, such as the brain, liver, adipose tissue, and muscle.\textsuperscript{24} Metabolic syndrome occurs in more than 50% of patients with obesity and is associated with chronic inflammation. Translocation of bacteria in the intestine has been suggested as the cause of this metabolic endotoxemia.\textsuperscript{25} Several studies have revealed that obesity is associated with changes in the gut microbiota. The first evidence reported identified that the microbiota of leptin-deficient ob/ob mice increases Firmicutes and decreases Bacteroidetes.\textsuperscript{26} In addition, when wild-type mice are administered with a high-fat diet, Firmicutes increases, and Bacteroidetes decreases.\textsuperscript{27} With respect to obesity, the ratio of Firmicutes/Bacteroidetes increased in the gut microbiota of patients with obesity, and fraction of Bacteroidetes increased when a low-carb or low-fat diet was fed.\textsuperscript{28}

When FMT was performed on 18 patients with metabolic syndrome using their own or the underweighted person's stool, nine patients who received stool from people with underweight body mass index (BMI) had noticeably reduced triglyceride levels and increased insulin sensitivity compared to the control group.\textsuperscript{29} In another study, 6 weeks after infusing microbiota from people with underweight BMI, the recipient's insulin sensitivity increased along with the level of intestinal microbiota producing butyrate.\textsuperscript{30}

**DIABETES**

Regarding the role of intestinal microbes in the etiology of type 2 diabetes characterized by insulin resistance, administering a high-fat diet to mice increases intestinal permeability, resulting in elevated lipopolysaccharide levels, an outer membrane component of microbes, in the blood. Plasma lipopolysaccharide concentration could set the tone for insulin sensitivity and onset of diabetes and obesity.\textsuperscript{31} Lactobacillus and Bifidobacterium were increased in diabetes-resistant mice,
and *Clostridiaeacea* and *Ruminococcus* were increased in diabetic-prone mice. In a randomized controlled, double-blind study including 18 men with insulin resistance who had not received previous treatment, 6 weeks after FMT, the insulin sensitivity and level of butyrate-producing intestinal microbes of recipients increased.

**CONCLUSIONS**

Gut microbes are related to various diseases, with the breakthrough development of analytical methods. Inducing changes in gut microbes through FMT or probiotics has been actively researched to prevent and treat various diseases. Together with these studies, research is warranted on the method of safely performing FMT or best target diseases, and evidence needs to be obtained for use in clinical practice.

**Conflicts of Interest**

The authors have no potential conflicts of interest.

**Funding**

None.

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