Identification of a 13-gene-based classifier as a potential biomarker to predict the effects of fluorouracil-based chemotherapy in colorectal cancer

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Abstract. The aim of the current study was to develop a predictor classifier for response to fluorouracil-based chemotherapy in patients with advanced colorectal cancer (CRC) using microarray gene expression profiles of primary CRC tissues. Using two expression profiles downloaded from the Gene Expression Omnibus database, differentially expressed genes (DEGs) between responders and non-responders to fluorouracil-based chemotherapy were identified. A total of 791 DEGs, including 303 that were upregulated and 488 that were downregulated in responders, were identified. Functional enrichment analysis revealed that the DEGs were primarily involved in ‘cell mitosis’, ‘DNA replication’ and ‘cell cycle’ signaling pathways. Following feature selection using two methods, a random forest classifier for response to fluorouracil-based chemotherapy with 13 DEGs was constructed. The accuracy of the 13-gene classifier was 0.930 in the training set and 0.810 in the validation set. The receiver operating characteristic curve analysis revealed that the area under the curve was 1.000 in the training set and 0.873 in the validation set (P=0.227). The 13-gene-based classifier described in the current study may be used as a potential biomarker to predict the effects of fluorouracil-based chemotherapy in patients with CRC.

Introduction

Colorectal cancer (CRC) is the third most commonly diagnosed cancer in males and the second in females, and it is one of the most common causes of cancer mortality (1). Localized CRCs are amenable to curative surgical resection, however, ~25% of patients present with metastatic disease and ~50% of patients will develop metastases (2). Fluorouracil-based chemotherapy remains the primary treatment for metastatic CRC (3). 5-fluorouracil (5-FU) alone has an objective response rate of ~20% (4). The addition of irinotecan or oxaliplatin to 5-FU increases the objective response rate to ~50% (5). The effects of 5-FU/leucovorin combined with irinotecan (FOLFIRI) or oxaliplatin (FOLFOX) in the first-line treatment of metastatic CRC are comparable (6). In the last decade, the addition of targeted therapies based on these chemotherapy regimens has improved the therapeutic approach and significantly increased progression-free survival and overall survival times (7-9). Fluorouracil-based chemotherapy remains the primary treatment for metastatic CRC. However, ~50% of patients are resistant to fluorouracil-based chemotherapy. In addition, the side effects of systemic chemotherapy, including neurotoxicity, myelotoxicity and gastrointestinal toxicity, may have a major impact on the quality of life of the patients and may lead to life-threatening complications (3). Therefore, identifying effective strategies that predict response to chemotherapy are required. Using these strategies, patients that are predicted to not respond to chemotherapy may receive other potentially effective treatments as early as possible and avoid unnecessary side effects. Gene expression profiling is used to predict the clinical outcome of patients with CRC (10-12). Previous studies have revealed that gene expression profiling may be used to predict cancer response to chemotherapy, including breast cancer and CRC (13-15).

The aim of the present study was to develop a predictor classifier for response to fluorouracil-based chemotherapy in patients with advanced CRC using microarray gene expression profiles of primary CRC tissues.
Materials and methods

Data processing. The raw microarray data (CEL files) of three datasets [GSE52735 (16), GSE62080 (15) and GSE69657 (17)] and corresponding clinical data were downloaded from the Gene Expression Omnibus database (www.ncbi.nlm.nih.gov/geo). The microarray data of the 3 datasets were based on the GPL570 Affymetrix Human Genome U133 Plus 2.0 Array platform (Affymetrix; Thermo Fisher Scientific Inc., Waltham, MA, USA). The GSE52735 set contained 37 advanced CRC samples treated with a fluoropyrimidine-based chemotherapy regimen (specific chemotherapy regimens were not available). A total of 23 of the samples were classified as responders and 14 samples were classified as non-responders to the chemotherapy regimen according to Response Evaluation Criteria in Solid Tumors (RECISt) (18). The GSE62080 dataset contained 21 advanced CRC samples treated with the FOLFI R1 regimen. A total of 9 samples were classified as responders and 12 samples were classified as non-responders according to the World Health Organization (WHO) criteria (19). The GSE69657 dataset contained 30 advanced CRC samples treated with the FOLFOX4 regimen. However, the raw microarray data was available for only 16 samples. A total of 7 of these samples were classified as responders and 9 samples were classified as non-responders according to RECIST. Two different evaluation criteria used in these three studies due to long time intervals between the studies. Previous studies have revealed that the RECIST criteria are comparable with the WHO criteria in evaluating the response of solid tumors (20-23). Preprocessing and normalization of the raw data were analyzed using the ‘affy’ (version 3.8) package (24) in R (www.r-project.org; version 3.5), using robust multi-array average for background correction and quantiles for normalization. Kernel and nearest neighbor averaging methods were used to impute the missing values using the ‘impute’ package (bioconductor.org/packages/impute; version 3.8) in R. The ComBat function in the ‘sva’ (version 3.8) package (25) was applied to remove batch effects. If one gene matched multiple probes, the average value of the probes was calculated as the expression of the corresponding gene. To build a robust predictive classifier, the GSE52735 and GSE62080 datasets were used as the training set (n=58), while the GSE69657 dataset was used as the validation set (n=16).

Screening of differentially expressed genes (DEGs) and enrichment analysis. Following preprocessing of the raw expression data, the DEGs between responders and non-responders in the training set were screened using the unpaired t-test in the ‘limma’ (version 3.8) package (26) in R. A DEG was defined as |log2 fold change (FC)|≥0.263 and P<0.05. The Gene Ontology (GO; http://geneontology.org/) and Kyoto Encyclopedia of Genes and Genomes (KEGG; https://www.genome.jp/kegg/) pathway enrichment analyses of DEGs were performed using the ‘clusterProfiler’ (version 3.8) package (27) in R with a cut-off of q<0.01.

Principal component analysis (PCA) prior to and following feature selection using the least absolute shrinkage and selection operator (LASSO) method. The expression values of DEGs in each sample were extracted. The LASSO logistic regression model analysis was performed using the ‘glmnet’ package (CRAN.R-project.org/package=glmnet; version 2.0-16) in R. The LASSO method is used to select optimal features in high-dimensional microarray data with a powerful predictive value and a low correlation between each other to prevent over-fitting (28). In the training set, the LASSO logistic regression model was used to select the optimal predictive markers. PCA using the expression profiles of the DEGs was performed prior to feature selection using the LASSO method. PCA was subsequently performed using the expression profiles of the optimal DEGs identified using the LASSO method. Samples were plotted in two-dimensional plots across the first two principal components.

Feature selection using Boruta and random forest classifier construction. A lower-dimensional model may reduce costs and is more likely to be used by clinicians (29). Following DEGs selection by the LASSO method, a feature selection was performed using the ‘Boruta’ package (www.jstatsoft.org/article/v036i11; version 6.0.0) in R. Boruta is a random forest-based feature selection method, which provides an unbiased and stable selection of important and non-important attributes from an information system. A variable importance (VIMP) measure may be calculated and visualized based on Boruta. In the current study, DEGs selected by Boruta were used to develop a gene-based classifier for response to fluorouracil-based chemotherapy in advanced CRCs. The random forest classifier was developed using the ‘randomForest’ package (CRAN.R-project.org/package=randomForest; version 4.6-14) in R. The validation set (GSE69657) was used to confirm the robustness and transferability of the classifier. The performance of the classifier was assessed by accuracy, sensitivity (Se), specificity (Sp), positive predictive value (PPV), negative predictive value (NPV) and receiver operating characteristic (ROC) curves in the training and validation sets. The ROC curves were drawn and compared using the ‘pROC’ (version 1.13.0) package (30) in R.

Results

DEGs in responders and non-responders and enrichment analysis. The training set included 32 responders and 26 non-responders. According to the cut-off criteria (log2_FC|≥0.263 and P<0.05), 791 genes were identified as differentially expressed between responders and non-responders. A total of 303 genes were upregulated and 488 genes were downregulated in responders. Functional enrichment analysis revealed that the biological process of DEGs were primarily involved in ‘cell mitosis’, ‘DNA replication’ and ‘cell cycle’ signaling pathways. The results of enrichment analysis are presented in Fig. 1.

PCA and feature selection using LASSO. For the first feature selection, LASSO logistic regression was performed using the expression data of DEGs in the training set. The group-wise classifications in 10-fold cross-validations were computed as default. A total of 31 DEGs were identified as optimal genes (Fig. 2A) with non-zero regression coefficients (Table I).
Fig. 2B presents the results of PCA prior to feature selection using LASSO and Fig. 2C presents the results of PCA following feature selection using LASSO. As demonstrated in Fig. 2C, responders and non-responders are easily distinguished using the 31 DEGs selected by LASSO.

Features selection using Boruta and construct of the random forest classifier. The Boruta function was used to further select features among the 31 DEGs. A total of 13 genes were confirmed as important, 7 genes were rejected and 11 tentative genes remained (Table I). Fig. 3 presents the variables' importance. These 13 important DEGs included small muscle protein X-linked, pleckstrin homology like domain family A member 1 (PHLDA1), prostaglandin reductase 2 (PTGR2), chitinase 1 (CHIT1), histone cluster 1 H2B family member c, formin homology 2 domain containing 3, OTUD6B antisense RNA 1 (head to head), cholinergic receptor nicotinic β1 subunit (CHRNB1), RPA interacting protein, DNA ligase 4 (LIG4), ASAP1 intronic transcript 2, small integral membrane protein 30 and c-Maf inducing protein. A random forest classifier was constructed using these 13 important DEGs.

Performance of the gene-based classifier. The accuracy of the 13-gene classifier was 0.930 in the training set and 0.810 in the validation set. Based on accuracy, Se, Sp, PPV, NPV and area under curve (AUC) values, the sample recognition efficiency of the classifier was high (Table II). ROC curve analysis revealed that the AUC was 1.000 in the training set and 0.873 in the validation set (P=0.227; Fig. 4).
Table I. Overview of the 31 optimal genes.

| Gene                                           | Log2 fold change (Responder/non-responder) | P-value | Coefficients provided by least absolute shrinkage and selection operator | Variable importance provided by Boruta |
|------------------------------------------------|-------------------------------------------|---------|------------------------------------------------------------------------|---------------------------------------|
| Matrix metallopeptidase 12                     | -1.069                                    | 0.002   | -0.154                                                                 | Tentative                             |
| C-X-C motif chemokine ligand 11                | -1.016                                    | 0.015   | -0.184                                                                 | Rejected                              |
| Forkhead box P2                                | 0.957                                     | 0.003   | 0.032                                                                  | Tentative                             |
| Small muscle protein X-linked                  | 0.766                                     | 0.003   | 0.575                                                                  | Confirmed                             |
| Pleckstrin homology like domain family A member 1 | -0.625                                   | 0.000   | -0.584                                                                 | Confirmed                             |
| Prostaglandin reductase 2                      | -0.602                                    | 0.000   | -0.792                                                                 | Confirmed                             |
| Chitinase 1                                    | 0.569                                     | 0.002   | 0.976                                                                  | Confirmed                             |
| S100 calcium binding protein A2                | -0.541                                    | 0.039   | -0.091                                                                 | Rejected                              |
| Histone cluster 1 H2B family member c          | 0.539                                     | 0.001   | 0.927                                                                  | Confirmed                             |
| RP1-74M1.3                                     | -0.515                                    | 0.005   | -0.023                                                                 | Tentative                             |
| Formin homology 2 domain containing 3          | 0.469                                     | 0.013   | 0.855                                                                  | Confirmed                             |
| RNA binding motif protein 3                    | -0.451                                    | 0.001   | -0.555                                                                 | Tentative                             |
| Tubulin polymerization promoting protein family member 3 | 0.412                      | 0.011   | 0.442                                                                  | Rejected                              |
| Cadherin related family member 2               | 0.411                                     | 0.047   | 0.913                                                                  | Tentative                             |
| OTUD6B antisense RNA 1 (head to head)          | 0.387                                     | 0.012   | 0.651                                                                  | Confirmed                             |
| Teashirt zinc finger homeobox 1                | 0.384                                     | 0.004   | 0.347                                                                  | Tentative                             |
| Cholinergic receptor nicotinic β1 subunit      | -0.365                                    | 0.000   | -3.574                                                                 | Confirmed                             |
| Stromal antigen 3-like 4 (pseudogene)          | -0.364                                    | 0.005   | -0.554                                                                 | Rejected                              |
| RPA interacting protein                        | -0.343                                    | 0.000   | -0.825                                                                 | Confirmed                             |
| Leucine rich repeat neuronal 1                 | -0.334                                    | 0.017   | -0.331                                                                 | Rejected                              |
| Heparan-α-glucosaminide N-acetyltransferase    | 0.334                                     | 0.006   | 1.374                                                                  | Rejected                              |
| MINDY lysine 48 deubiquitinase 3               | -0.320                                    | 0.001   | -0.253                                                                 | Tentative                             |
| THAP domain containing 5                       | -0.308                                    | 0.016   | -0.432                                                                 | Rejected                              |
| DNA ligase 4                                   | 0.298                                     | 0.002   | 1.692                                                                  | Confirmed                             |
| Zinc finger protein 2                          | -0.291                                    | 0.004   | -1.885                                                                 | Tentative                             |
| ASAP1 intronic transcript 2                    | 0.289                                     | 0.004   | 0.045                                                                  | Confirmed                             |
| Small integral membrane protein 30             | -0.287                                    | 0.001   | -0.973                                                                 | Confirmed                             |
| c-Maf inducing protein                         | 0.282                                     | 0.001   | 0.208                                                                  | Confirmed                             |
| ADAMTS like 2                                  | 0.278                                     | 0.005   | 1.088                                                                  | Tentative                             |
| Nucleoporin 133                                | -0.273                                    | 0.011   | -1.718                                                                 | Tentative                             |
| DEAD-box helicase 28                           | -0.267                                    | 0.003   | -0.063                                                                 | Tentative                             |
Personalized treatment may improve the treatment outcome of patients with tumors (31). In CRC, the gene expression levels of vascular endothelial growth factor (VEGF) and epidermal growth factor receptor (EGFR) provide the basis for selecting EGFR and VEGF inhibitor combinations (32-36). Monoclonal antibodies against VEGF and EGFR have been approved for treatment of metastatic CRC in combination with 5-FU-based regimens (3). The identification of subsets of patients that respond to specific chemotherapy regimens remains a challenge (3). A previous study demonstrated that tumors with microsatellite instability (MSI) respond well to 5-FU-based therapies; however, further studies are required to substantiate these results (37). Another previously published study suggested that MSI status does not affect the outcome of the treatment (38). Therefore, effective tools for predicting the outcome of chemotherapy are currently lacking. The present study identified 13 genes from 791 DEGs using two feature selection algorithms and developed a 13-gene predictor classifier for response to fluorouracil-based chemotherapy in CRC. The predictor classifier demonstrated high accuracy in the training and validation sets. The training set included two datasets from different centers, and the validation set was from an additional independent center. ROC curve analysis revealed that the AUC was 1.000 in the training set and 0.873 in the validation set, and their difference was not significant (P=0.227). These results suggested that the classifier was robust. The study established a foundation for further research into personalized treatment of CRC.

Previous studies have attempted to identify a single biomarker to predict response to fluorouracil-based chemotherapy in CRC (17,39-42). However, there is currently no single biomarker that is routinely applied in clinical practice. CRC is a heterogeneous disease, which is compounded by changes in the molecular profile of the tumor as it progresses (3). An in vitro study demonstrated that the measurement of multiple, rather than single marker genes, may provide a more accurate assessment of drug response in colon carcinoma (43). Previous studies have been designed to identify a pattern of gene expression capable of predicting response to fluorouracil-based chemotherapy in CRC (15,16). One study identified a set of 14 genes for predicting response to the FOLFIRI regimen based on 21 samples (15), and an expression profile of 7 genes was identified in another study (16). Compared with the two aforementioned studies, the current study performed a comprehensive analysis of more samples (n=58) from two centers and validated the predictor classifier in an independent dataset (n=16). Furthermore, to the best of the authors' knowledge, the current study is the first to construct a random forest classifier to predict response to chemotherapy in CRC.

### Table II. Performance of the 13-gene classifier.

| Cohort      | Sensitivity | Specificity | Positive predictive value | Negative predictive value | Accuracy | Area under the curve |
|-------------|-------------|-------------|---------------------------|---------------------------|----------|-----------------------|
| Training set| 0.970       | 0.960       | 0.910                     | 0.960                     | 0.930    | 1.000                 |
| Validation set| 0.860   | 0.880       | 0.750                     | 0.880                     | 0.810    | 0.873                 |

Figure 3. Z score evolution during Boruta run. Green lines correspond to confirmed attributes, yellow to tentative, red to rejected ones; and blue lines correspond to respectively minimal, average and maximal shadow attribute importance.

Figure 4. Receiver operating characteristic curves for training and validation sets. AUC, area under the curve.
Considering the limited ability of Cox regression analysis to process high-dimensional data (44), it was not performed in the current study. A random forest algorithm was used to construct the classifier, which was subsequently validated with an independent dataset. The results obtained in the current study suggest that the robust classifier developed warrants further investigation.

Functional enrichment analysis revealed that certain DEGs identified in the present study are involved in DNA replication and cell cycle pathways; however, none of the 13 genes were involved in these two signaling pathways. A previous study suggested that PHLD1A may be associated with CRC progression (45). A previous study demonstrated that PTGR2-knockdown gastric cancer cells rendered them more sensitive to cisplatin and 5-FU compared with the PTGR2-overexpressing cells (46). In addition, two variants of CHIT1, rs61745299 and rs35920428, may increase expression of the gene and have been associated with CRC (47). CHRNA1 may be a biomarker for the detection of relapsed and early relapsed CRC (48). In addition, LIG4 may mediate Wnt signaling-induced radioresistance in CRC (49). With the exception of the aforementioned studies, the association between the 13 genes identified in the current study and CRC or chemotherapy has not been investigated. Therefore, it is not clear whether these genes are causal or merely markers for response to fluorouracil-based chemotherapy in CRC.

Although the current study provides novel insights into the treatment of CRC, it has some limitations. The present study was based on a relatively small sample size; however, it is worth noting that the sample size in our study is relatively large compared with previous studies (15,16). Future studies are required to verify and improve the 13-gene signature in a larger independent cohort of patients.

In conclusion, the current study identified a 13-gene predictor classifier for the response to fluorouracil-based chemotherapy in patients with advanced CRC.

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Availability of data and materials

All data generated or analyzed during the present study are included in this published article.

Authors’ contributions

YFL designed the study and revised the manuscript. ZHG and QYZ analyzed the data and wrote the manuscript. YL, ZHX, ZH, and ZCC assisted with analyzing the data and writing the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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