Antiangiogenic cancer treatment: The great discovery and greater complexity (Review)

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Abstract. The discovery of tumor angiogenesis opened a new path in fighting cancer. The approval of different antiangiogenic agents, most targeting vascular endothelial growth factor (VEGF) signaling, has either increased the effectiveness of standard chemotherapy or even replaced it by offering better patient outcomes. However, an increasing number of preclinical and clinical observations have shown that the process of angiogenesis is far from clearly understood. Apart from targeting the VEGF pathway, novel strategies aim to influence other molecular factors that are involved in tumor angiogenesis. In addition, naturally occurring compounds seem to offer additional agents for influencing angiogenesis. The first concept of antiangiogenic therapy aimed to destroy tumor vessels, while it turned out that, paradoxically, antiangiogenic drugs normalized vasculature and as a result offered an improvement in chemotherapeutic delivery. In order to design an effective treatment schedule, methods for detecting the time window of normalization and biomarkers predicting patient response are needed. The initial idea that antiangiogenic therapy would be resistance-free failed to materialize and currently we still face the obstacle of resistance to antiangiogenic therapy.

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1. Introduction

The concept of antiangiogenic therapy in cancer patients started after observations performed by Judah Folkman approximately 45 years ago. He noticed that in order to grow beyond 1-2 mm³ tumors require blood supply and for that reason induce the generation of new vessels in the process of angiogenesis. Based on such observations, it was proposed that inhibition of tumor vessel formation could suppress tumor growth and that concept was called antiangiogenic therapy (1,2). The next step in the field of discovering angiogenesis was the isolation and characterization of the vascular endothelial growth factor (VEGF), initially termed the vascular permeability factor (VPF) by Senger et al (3) and Ferrara (4). VEGF is the best characterized angiogenic factor. The function of VEGF is to modulate vessel permeability, remodeling, endothelial cell (EC) survival, proliferation and migration (5,6). VEGF is overexpressed in cancer cells (7,8). Very high levels of VEGF and other proangiogenic factors result in the formation of new vessels, but their architecture and function is abnormal. Tumor vessels are dilated, tortuous, and disorganized with haphazard patterns (lack microvascular hierarchy) and their pore sizes are 100 times bigger than is physiologically normal. ECs forming tumor vessels are loosely connected with each other and have an irregular morphology. Perivascular cells, i.e. pericytes and vascular smooth muscle cells that normally stabilize blood vessels by covering ECs, within tumors are absent or poorly attached to vessels. The vascular basement membrane is also abnormal: thick in some tumors or thin or even absent in others. These structural abnormalities cause functional aberrations. Tumor vessels are hyperpermeable and hence intravascular fluid and plasma proteins extravasate, causing an increase in interstitial fluid pressure (IFP). The blood supply is heterogeneous, some areas are hyper- whilst other are hypo-vascularized. As a consequence, hypoxia and acidosis occur within a tumor. Moreover, hypoxia is one of the mechanisms regulating VEGF expression; therefore, the formation of abnormal vasculature intensifies in a self-reinforcing vicious cycle. The chronic imbalance of the pro- and anti-angiogenic factors in cancer, i.e. excess of pro- and deficiency of antiangiogenic factors, leads to abnormal angiogenesis (9,10). Thus, VEGF, as a main agent involved in angiogenesis and signaling pathway engaged in the regulation of the function of ECs, became a target in developing antiangiogenic therapies (11).

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was approved as a monotherapy in second-line treatment of metastatic CRC (16). Within the next few years, bevacizumab showed that bevacizumab, a humanized monoclonal antibody binding specifically to VEGF-A alone, when combined with chemotherapy in metastatic colorectal cancer improved progression-free survival (PFS) (10.6 vs. 6.2 months) and overall survival (OS) (23 vs. 15.3 months) compared to chemotherapy alone. Based on those results, the US Food and Drug Administration (FDA) accelerated in 2008 approval of bevacizumab in combination with paclitaxel in metastatic breast cancer. Further trials, AVADO and RIBBON-1, confirmed the improvement of PFS by bevacizumab, but neither demonstrated any improvement of OS. In 2011, FDA withdrew approval for bevacizumab in metastatic breast cancer (19). In 2014, bevacizumab was approved for the treatment of patients with platinum-resistant recurrent epithelial ovarian, fallopian tube, or primary peritoneal cancer in combination with paclitaxel, pegylated liposomal doxorubicin or topotecan, based on the results of AURELIA clinical trials comparing bevacizumab plus chemotherapy with chemotherapy alone (20,21). Also in 2014, bevacizumab was approved in combination with paclitaxel and cisplatin or paclitaxel and topotecan in persistent, recurrent or metastatic cervical cancer (22,23).

Tyrosine kinase inhibitors (TKIs) are small-molecular-weight drugs that inhibit the kinase activity of different receptors. The mechanism of action of TKIs relies on binding around the ATP binding site of a given receptor and thus hindering phosphorylation of the tyrosine residue of that receptor and subsequent transmission of signaling down the intercellular pathway (2). There are 28 small-molecule kinase inhibitors (tyrosine kinase, serine/threonine kinase or dual protein kinase inhibitors) approved by the FDA. Among these, there are some agents that target VEGF receptors (VEGFR) and these are used to treat different types of cancer, e.g. sunitinib, sorafenib, axitinib and pazopanib (Table I) (24,25). Compared to VEGF neutralizing antibodies, TKIs do not interfere with the binding of VEGF to its receptors and they usually target not only VEGFR but additionally other kinases, such as PDGFR, FGFR and c-KIT (9).

Another strategy developed to inhibit angiogenesis is a human recombinant fusion protein called aflibercept, acting as a decoy receptor of angiogenic factors. Aflibercept, unlike bevacizumab, targets not only VEGF-A, but also VEGF-B and placental growth factor (PIGF). This is a fusion protein of the 2nd immunoglobulin domain of VEGFR1, 3rd immunoglobulin domain of VEGFR2 and constant region Fc of human IgG1. In 2012, FDA approved aflibercept in the treatment of metastatic colorectal cancer (CRC) with infusional fluorouracil, leucovorin and irinotecan, based on phase III trial results (26).

Ramucirumab is another human monoclonal antibody developed to inhibit angiogenesis. It blocks the interaction of VEGF with its receptor by binding to the extracellular domain of VEGFR2. Preclinical studies showed that ramucirumab binds selectively to VEGFR2 with a greater efficacy than its natural ligand VEGF-A. It is approved in second-line treatment in gastric, NSCLC and colon cancer. Based on the RAISE study, ramucirumab was approved in combination with FOLFIRI (folinic acid, 5-fluorouracil and irinotecan) in metastatic CRC patients, if disease progressed after therapy with bevacizumab, oxaliplatin and fluoropyrimidine. In NSCLC,
ramucirumab was approved in combination with docetaxel after platinum-based chemotherapy. In gastric cancer patients, FDA approved ramucirumab as a monotherapy in advanced or metastatic or gastroesophageal junction carcinoma patients for whom 1st line chemotherapy had failed (27,28). Other strategies in preclinical and clinical studies. Apart from the above already approved antiangiogenic agents, additional strategies have been developed aimed at inhibiting tumor angiogenesis, directly or indirectly, that are being tested in preclinical and clinical trials. These include agents that target angiogenesis directly: PlGF, angiopoietin-Tie2 axis, integrins or agents targeting angiogenesis indirectly by inhibiting oncogenic pathways (e.g. HER2, PI3k/AkT/mTOR and mutated EGFR) or hormone signaling (9). Moreover, some known anticancer drugs designed for specified mechanisms of action may reveal previously unknown antiangiogenic activity, due to interaction with other signaling pathways that had not initially been considered. For example, Calero et al (30) showed that sunitinib, TKI designed to inhibit VEGF receptor activity, decreased VEGF secretion from SK-N-BE(2) neuroblastoma cells. The lowered VEGF expression correlated with both PI3k/AkT signaling pathway inhibition after sunitinib treatment and increased Myc protein degradation. In turn, in a lung cancer model, treatment with imatinib resulted in downregulation of VEGF expression in A549 tumors, which was accompanied by upregulation of p53 expression (31).

### Table I. FDA approved tyrosine kinase inhibitors with known anti-VEGFR activity.

| TKI              | Activity                              | Initial US approval | Indicationsa |
|------------------|---------------------------------------|---------------------|--------------|
| Axitinib         | VEGFR 1-3                             | 2012                | Advanced RCC |
| Cabozantinib     | RET, MET, VEGFR 1-3, KIT, TRKB, FLT-3, AXL, TIE-2 | 2012                | Progressive, metastatic medullary thyroid cancer |
| Lenvatinib       | VEGFR 1-3, FGFR 1-3, PDGFRα, KIT, RET | 2015                | Locally recurrent or metastatic, progressive, radioactive iodine-refractory thyroid cancer |
| Nintedanib       | FGFR 1-3, PDGFRα/β, VEGFR 1-3, FLT3   | 2014                | Idiopathic pulmonary fibrosis |
| Pazopanib        | VEGFR 1-3, PDGFRα/β, FGFR 1/3, KIT, LCK, FMS, Itk | 2009                | Advanced RCC, advanced soft tissue carcinoma |
| Ponatinib        | BCR-ABL, BCR-ABL T315I, VEGFR, PDGFR, FGFR, EPFR, EPFR, SRC family kinases, KIT, RET, TIE2, FLT3 | 2012                | Adult patients with T315I+ CML (chronic phase, accelerated phase, or blast phase) or T315I+ Ph+ ALL; adult patients with chronic phase, accelerated phase, or blast phase CML or Ph+ ALL for whom no other tyrosine kinase inhibitor (TKI) therapy is indicated |
| Regorafenib      | VEGFR 1-3, BCR-ABL, B-RAF, B-RAF(V600E), KIT, PDGFRα/β, RET, FGFR1/2, TIE2, Eph2A | 2012                | Metastatic CRC treated previously with fluoropyrimidine, oxaliplatin and irinotecan; locally advanced, unresectable or metastatic GIST treated previously with imatinib or sunitinib |
| Sorafenib        | B/C-RAF, B-RAF(V600E), KIT, FLT3, RET, VEGFR 1-3, PDGFRβ | 2005                | Unresectable hepatocellular carcinoma, advanced RCC, locally recurrent or metastatic, progressive, differentiated TC refractory to radioactive iodine treatment |
| Sunitinib        | PDGFRα/β, VEGFR 1-3, KIT, FLT3, CSF-1R, RET | 2006                | GIST after disease progression on or intolerance to imatinib mesylate, advanced RCC, progressive, well-differentiated pNET |
| Vandetanib       | EGFRs, VEGFRs, RET, BRK, TIE2, EPFRs, SRC family kinases | 2011                | Symptomatic or progressive medullary TC |

a Data collected from FDA website (29). RCC, renal cell carcinoma; CML, chronic myeloid leukemia; ALL, acute lymphoblastic leukemia; Ph+, Philadelphia chromosome positive; CRC, colorectal carcinoma; GIST, gastrointestinal stromal tumor; TC, thyroid cancer; pNET, pancreatic neuroendocrine tumor.
tolerated dose (MTD) schedule (33-35). Given in a metronomic schedule, docetaxel downregulated VEGF expression in gastric cancer BGC-823 cells and VEGF, bFGF, matrix metalloproteinase (MMP)-2 and MMP-9 in colon adenocarcinoma LS174T cells, while it upregulated TSP-1 expression in HUVCE cells and decreased microvessel density (MVD) and VEGF and increased TSP-1 in tumor tissue of a BGC-823 model (36,37).

Another concept in anticancer treatment related to angiogenesis is the inhibition of metastatic potential through an influence on the interaction between cancer cells and ECs and platelets with compounds targeting cyclooxygenase-2/prostaglandin pathways. The use of 1,4-dimethylpyridinium chloride (1,4-DMP) has been shown to decrease the number of metastases in combination with cyclophosphamide in a 4T1 breast cancer model (38).

There are also some interesting studies on the anticancer activity of naturally occurring products and their potential usefulness as chemopreventive agents. These are also being tested for their antiangiogenic activity. The antiangiogenic effect of plant extracted compounds involves: EC proliferation and migration inhibition, preventing sprout formation, MMP inhibition and modulation of angiogenic signaling pathways. Plant-based agents demonstrate synergism when used in combination with chemotherapy and many of them reveal low levels of undesired side-effects, or even limit side-effects caused by chemotherapy (14,39,40). These agents have also been tested in the context of ocular diseases, where excessive ocular neoangiogenesis is, like cancer neovascularization, the consequence of an imbalance between pro- and antiangiogenic factors. It has been shown that different natural compounds, e.g. curcumin, genistein, luteolin and resveratrol, suppress retinal neovascularization in different in vitro and in vivo models (41). One of the plant-derived compounds extensively studied for anticancer and antiangiogenic activity is soy isoflavon genistein, first isolated in 1899. The antiangiogenic activity of genistein was revealed as the ability to decrease microvessel density, lower VEGF and increase endostatin plasma level (42,43). However, the exact results of genistein treatment depend on the doses used: at high and medium concentrations of soy isoflavones (10-150 µM) antiangiogenic activity has been observed, while at lower doses (<10 µM) genistein tended to increase VEGF secretion from breast cancer cells (44). The antiangiogenic properties of genistein have also shown its ability to prevent metastasis or lower blood supply measured in Lewis lung cancer and B16 melanoma models (45-47). Another natural agent generating general interest in anticancer research is resveratrol, a phytochemical of grapes, berries and peanuts. On human ovarian cancer cells it has been shown that resveratrol attenuates the induction of HIF-1α and VEGF by lipopolysaccharide LPS (42,48). Resveratrol also inhibits mediation of tumor necrosis factor α (TNFα) MMP-9 expression in HepG2 hepatocellular carcinoma cells and also NF-κB expression and invasion of HepG2 cells (49). The identified mechanisms of antiangiogenic activity of natural, plant derived agents so far include: interfering with signaling of VEGF and FGF, decreased vascular permeability by inhibiting NO release from ECs, modulation of NF-κB activity, and inhibition of HIF-1α expression and its downstream targets (VEGF, TNFα, COX2, IL-6 and IL-8) (41).

Another important naturally occurring compound that is intensively studied in cancer research is vitamin D, a steroid hormone regulating calcium and phosphate homeostasis (50). Vitamin D is synthesized in the skin upon UVB exposure from 7-dehydrocholesterol and next hydroxylated at C-25 and C-1 in the liver and kidney, respectively. Vitamin D can also be obtained from the diet, especially from food of animal origin, fish, meat, eggs and milk products, but it has also been shown that vitamin D can be found in plants (51). Calcitriol, the active form of vitamin D₃, regulates many cellular and tissue processes involved in carcinogenesis: proliferation, differentiation, apoptosis, inflammation, invasiveness and metastasis by transcriptional regulation of gene expression (genomic action) or by influencing different signaling pathways (rapid, non-genomic action) (50,52,53). One of the first pieces of evidence for the antiangiogenic activity of calcitriol was a study on 4.5-day old chick embryos, where it was shown that calcitriol and vitamin D₃ analog, 22-oxa-1,25-dihydroxyvitamin D₃, inhibited angiogenesis in chorioallantoic membranes (54). Calcitriol reduces the expression of proangiogenic factors, e.g. VEGF, and IL-8, and inhibits the proliferation of ECs derived from tumors through the induction cell cycle arrest and apoptosis (55-57). In azoxymethane-induced colon cancer in rats, vitamin D derivative administration resulted in decreased immunohistochemical staining of VEGF and microvessel counts (58). In LLC cells, vitamin D derivatives reduced MMP-2, MMP-9 and VEGF expression and in in vivo Matrigel assays inhibited angiogenesis induced by bFGF (59). In another study using the LLC model, it was shown that calcitriol and its analog PRL-2191 inhibit growth and metastasis of LLC cells transplanted subcutaneously. Moreover, a tendency to decrease blood vessel diameter, without influencing their number, was observed. This observation may suggest the influence of calcitriol and its analog on tumor growth not only directly, but also through normalization of tumor vasculature (60). Therefore, a number of in vitro and in vivo studies have proved that vitamin D has a significant impact on the process of angiogenesis (50). Unfortunately, the biological anticancer activity of calcitriol can only be obtained when administered in high doses, which limits its use due to the risk of hypercalcemia. This has inspired many scientists to synthesize vitamin D analogs in order to dissociate calcemic from anti-proliferative activity of vitamin D. These analogs were then tested alone or in combinations in cancer research (53,61-63). No vitamin D compounds are currently used in clinical practice for cancer treatment, although in preclinical animal cancer models several vitamin D analogs have appeared to be potent drugs, especially in combination with known chemotherapy (31,63-66). To date, a small number of studies have assessed the influence of vitamin D and its derivatives on angiogenesis (63). In clinical trials, vitamin D supplementation has been studied with the aim of reducing the risk of cancer. In addition, vitamin D has been studied in combination with chemotherapy, mainly in prostate cancer patients. However, results are still inconsistent and no clear conclusions have been made in this field; therefore, more studies are required (67-71).

The uncovered anticancer and antiangiogenic activity of many natural compounds may offer a great opportunity...
in cancer prevention or may strengthen existing anticancer treatment options. It is believed that the use of such natural health products may be beneficial, as they may act through many signaling pathways and reduce the development of resistance by cancer cells and therefore improve patient outcomes (39,40,42).

3. Vessel disruption or normalization

The rationale behind antiangiogenic therapy was the concept that blocking blood vessel formation in tumors or its regression would deprive cancer cells of nutrients and oxygen and finally starve tumors to death or induce tumor dormancy. One of the first preclinical studies clearly showed that treatment with an anti-VEGF monoclonal antibody caused a significant vascular density reduction and tumor growth delay in mice bearing xenografts of glioblastoma multiforme, leiomyosarcoma and rhabdomyosarcoma (72). It turned out, however, that anti-VEGF monotherapy in clinical trials of human solid tumors showed only modest objective response rates and lacked noticeable survival benefits for patients (73). After many years of clinical trials, anti-VEGF agents appeared to be active as single agents only in a limited number of cancers, e.g. renal cell carcinoma, hepatocellular carcinoma, ovarian, neuroendocrine tumors and glioblastoma. In contrast, in other studied cancers, CRC, NSCLC and breast cancer, the administration of anti-VEGF drugs was effective only when combined with chemotherapy, leading to significant improvements of PFS and OS compared to chemotherapy alone (73,74).

Such clinical results thus generated some confusion. It is known that the efficacy of chemotherapy depends on efficient delivery of cytotoxic agents to tumor cells through efficient blood flow, whilst antiangiogenic therapy, according to the theory, should destroy blood vessels and thus prevent drug delivery. In order to elucidate these seemingly counterintuitive observations the hypothesis of ‘vessel normalization’ was proposed in 2001, that is many years after the importance of inhibiting tumor angiogenesis had been identified (10). It was assumed that judicious administration of antiangiogenic drugs reverts the abnormal structure and function of the tumor vessels towards normal state (Fig. 2). In this regard, treatment with antiangiogenic agents would correct the arrangement of vasculature towards a more organized structure leading to increased homogeneity in blood flow. Furthermore, improvement in junctions between ECs, increases in pericyte content around vessels and also better connections between ECs and pericytes would decrease vascular permeability with a simultaneous decline in intratumoral fluid pressure. As a result, cytotoxic drugs would be effectively delivered to cancer cells owing to increased and efficient blood perfusion within the tumor (75,76). Indeed, many preclinical and clinical studies have shown that antiangiogenic therapy results in vascular normalization. For example, it has been shown in preclinical studies with different human tumors that administration of
an anti-VEGF A4.6.1 antibody results in a reduction in vessel diameter and tortuosity, a significant decline in vascular permeability to plasma proteins, providing evidence that after neutralizing tumor cell-derived VEGF abnormalities of tumor vasculature could be reversed (77). Similar results have been achieved in studies with the use of bevacizumab in combination with chemotherapy or ionizing radiation. After bevacizumab administration of mice bearing neuroblastoma or rhabdomyosarcoma xenografts, a substantial decrease in tumor microvessel density and improved pericyte coverage in tumors has been observed with a concomitant decrease in vascular permeability, a drop in intratumoral fluid pressure and an increase in intratumoral oxygen pressure (78,79).

Tumor vessel normalization could also be observed after using tyrosine kinase inhibitors. In a murine Lewis lung cancer model, treatment with axitinib resulted in reduction in microvessel density and vascular sprouting (80). Similarly, the administration of sunitinib in a human glioma model resulted in a decrease in MVD and collagen IV density (but no effect on α-SMA density) and an improvement in tamoxifen/loxitane penetration into brain tumors (81,82). Some clinical studies have also demonstrated the occurrence of vascular normalization in cancer patients after treatment with antiangiogenic agents. A study in patients with locally advanced rectal adenocarcinoma receiving bevacizumab 7 weeks before surgical resection, first dose was given alone and after 2 weeks with 5-fluorouracil, showed a decline in tumor microvessel density, a reduction of intratumoral fluid pressure, and an increase in the content of pericytes covering vessels (83). More examples of vessel normalization as a consequence of antiangiogenic therapy have been reviewed elsewhere (9).

Hypoxia induced by abnormal tumor vascularity influences the immune response in cancer. This contributes to immune tolerance by inhibiting the proliferation and activity of T lymphocytes and inducing accumulation and polarization of immune cells towards suppressive phenotypes. It has been proposed that normalizing tumor vascularity could enhance the adoptive cell transfer efficacy (84). In 2012, Huang et al (85) demonstrated that angiogenic treatment influences the effectiveness of immunotherapy by the modulatory activity of antiangiogenic agents on tumor microenvironment. In a breast cancer model, the authors studied the influence of administration of anti-VEGFR2 antibody on anticancer vaccine therapy in immune-tolerant and immunogenic mice. The study showed that treatment with an anti-VEGFR2 antibody enhanced anticancer activity of whole cancer cell vaccine in a CD8+ T-cell-dependent manner in both murine models. Additionally, the efficacy of the tested therapy depended on the dose of the anti-VEGF agent: lower doses of anti-angiogenic agent were superior to higher doses in augmenting the infiltration of tumor with CD4+ and CD8+ T-cells and in polarizing tumor-associated macrophages, from immunosuppressive M2 towards immunostimulatory M1 phenotypes.

Data obtained in preclinical and clinical studies have shown that after cessation of the antiangiogenic therapy rapid revascularization occurs in tumors, which can be followed by rapid regrowth of the tumor (86–88). It turned out that vessel normalization obtained as a result of antiangiogenic therapy is transient. The period when the vessel normalization is present is called the ‘time window’ or ‘window of opportunity’. The temporariness of normal features of tumor vessels may result from discontinuation of therapy or rest periods in therapy schedules, but also as a consequence of excessive doses or prolonged administration of antiangiogenic drugs (9,89).

Studies have shown that abnormalities of tumor vessels are reversed as early as 24-72 h after starting the therapy and are sustained for different periods, from few to dozens of days, or sometimes longer (78,90,91). The existence of a time window is important for appropriate scheduling of combined treatment. Many studies have shown that the efficacy of combined antiangiogenic and chemotherapy treatment is schedule-dependent. Since successful activity of cytotoxic drugs depends on efficient drug delivery to cancer cells, which can be obtained after vessel normalization, it has been proposed that chemotherapy or radiotherapy should be applied after administration of antiangiogenic agents (78,92,93). In a study of 2 patients with metastatic breast cancer, Chen et al (89) analyzed the time of appearance of vessel normalization after bevacizumab administration by means of three-dimensional power Doppler ultrasonography and observed that the window was open 20-24 h after bevacizumab injection. Additionally, it was shown that sequential treatment of bevacizumab: on days 1 and 15, and paclitaxel: on days 2, 9 and 16 resulted in rapid reduction of tumors in brain, as observed in computed tomography.

The optimal timing of administration of anti-VEGF agents before cytotoxic drugs is required to achieve the highest anticancer response of treatment used. The challenge is the method of determination of the normalization window in a patient, and what is more non-invasively. There have been some studies aimed at probing the time window of vessel normalization. Vangestel et al (94) used 99mTc-tricarbonyl His-annexin A5, radiotracer of apoptosis to explore the timing between administration of bevacizumab and irinotecan in a colon cancer model that would result in the greatest tumor cell death. Hernandez-Agudo et al (95) used 18F-misonidazole ((18F)-FMISO) PET as a hypoxia tracer in order to explore vessel normalization after administration of divotinib in pancreas and breast cancer models. After a decrease in hypoxia, and therefore vessel normalization, the delivery of chemotherapy was improved and so was the cytotoxic effect. Data obtained in that study suggested that (18F)-FMISO mirrors the dynamic of hypoxia and changes in vessel normality/abnormality in response to a short course of antiangiogenic therapy. Other candidate biomarkers tested to assess the response to anti-VEGF therapy are: tumor biopsy, measuring plasma protein concentration e.g. VEGF, or level of circulating ECs and progenitor cells, also imaging diagnostic methods (CT, PET, MRI) (73).

4. Toxicity

As angiogenesis in adults is a rather rare process, it was thought that anti-VEGF therapies would be free of toxicity. However, clinical practice showed that anti-angiogenic therapy is accompanied with a number of side-effects, including hemorrhage, hypertension, proteinuria, impaired wound healing, thrombosis and others (11). Preclinical studies with non-tumor bearing mice administered with anti-angiogenic agents have shown that the treatment alters the density and architecture of vessels in multiple tissues and organs, especially in endocrine organs that had fenestrated vessels as a result of antiangiogenic therapy (96).
Yang et al (97) showed that systemic administration of anti-VEGF and anti-VEGFR neutralizing antibodies affected the vasculature of multiple organs, with the greatest vessel regression in endocrine glands, intestine and uterus. On the other hand, high levels of VEGF produced by cancer cells correlated with abnormal hepatic sinusoidal blood vessels and high mortality in a VEGF expressing melanoma model (98). Cancer patients, mostly in the advanced stage of the disease, experience so-called cancer-associated systemic syndrome (CASS) or paraneoplastic syndrome as a result of production and secretion excess amounts of different peptides and hormones that affect diverse systems, most frequently the endocrine, gastrointestinal, neurologic, dermatologic and hematologic systems. Physiologically, these factors are paracrine hormones, but when overproduced by malignant cells they enter the circulation and influence distant tissues and organs deregulating homeostasis (96,99,100). Elevated levels of VEGF expressed by cancer cells induced CASS in mice, manifesting with severe anemia, ascites, hepatic dysfunction, and decreases in serum corticosterone levels, whilst the use of anti-VEGFR agents resulted in vessel normalization in healthy tissues and improved survival of animals (98,100). These surprising observations show that antiangiogenic therapy may also have an impact on improving healthy tissue and organ function in cancer patients.

5. Predictive biomarkers

Efforts are being made to identify some biomarkers that could predict the clinical benefits of antiangiogenic therapy for a given patient. Since the monoclonal antibody bevacizumab specifically targets VEGF, it was assumed that measuring serum VEGF levels could serve as a predictive marker for patient selection. Unfortunately, so far it has not been proved that VEGF level, in blood or in tumor biopsies, could fulfill the requirements of a predictive biomarker (101,102). Studies on circulating VEGF levels in cancer patients have shown the importance of VEGF as a prognostic rather than predictive biomarker (11). On the other hand, it has turned out that some of the adverse effects related to antiangiogenic agents appeared to be positively correlated with response to therapy. For example, it was shown that hypertension associated with the bevacizumab or TKIs correlated with clinical response in patients with breast, colorectal and NSCLC, whilst skin rashes correlated with drug response in patients with colorectal and hepatocellular carcinoma (96). Another approach in predicting the response to antiangiogenic therapy was the imaging of tumor vasculature, with the use of CT, MRI or PET. In a study of glioblastoma patients treated with cediranib vessel normalization was shown by means of MRI as a decrease in vessel diameter and permeability (10). Some studies have shown a correlation between changes in vascular features and patient outcome, but there are some limitations and obstacles that need to be challenged: an understanding of detected characteristics of vasculature with the biology of tumor, also the methodologies require standardization (11).

6. Resistance to antiangiogenic therapy

Despite the great success of antiangiogenic therapy, as for anticancer drugs, resistance to antiangiogenic treatment is also an important issue. Introduction of anti-VEGF drugs to anticancer therapy augmented PFS, causing transient disease stabilization, but improvement in OS can not always be achieved. What is more, withdrawal of an antiangiogenic drug from a therapy is followed by rapid regrowth of the tumor. It turned out that some types of cancer can be intrinsically refractory to antiangiogenic therapy or during the treatment acquire resistance to anti-VEGF agents (103,104). The intrinsic resistance may result e.g. from elevated levels of circulating, soluble VEGFR1 (sVEGFR1) before therapy. sVEGFR1 acts as an intrinsic VEGF decoy receptor and thus adding an external anti-VEGF drug has no biological effect. It has been shown that patients with rectal carcinoma, hepatocellular carcinoma, and metastatic colorectal carcinoma who had elevated levels of sVEGFR1 did not benefit from adding bevacizumab to chemotherapy (102). Acquired resistance to antiangiogenic therapy may result from a few possible mechanisms: activation of alternative signaling pathways, recruitment of bone-marrow derived cells, stromal cells of tumor microenvironment, vessel co-option and vessel mimicry and increased invasiveness and metastasis (Fig. 3) (105).

There are a number of other pro-angiogenic pathways and factors that can stimulate blood vessel growth and survival when the VEGF-mediated pathway is inhibited. Some of these pro-angiogenic factors are: angiopoietins (Ang), epidermal growth factor (EGF), fibroblast growth factors (FGF1 and FGF2), hepatocyte growth factor (HGF), interleukin 8 (IL-8), platelet-derived growth factor C (PDGF-C) and placental growth factor (PIGF) (11,105). The upregulation of PIGF, FGF2, IL-8 expression could be observed in colorectal, glioblastoma, renal cell and hepatocellular cancer patients after anti-VEGF therapy (106-109). Proteomic analysis performed in bevacizumab treated breast cancer xenograft showed upregulation of several compensatory signaling pathways with persistent mTOR signaling. It was next hypothesized that targeting the PI3K pathway would increase the efficacy of therapy consisting of bevacizumab and indeed combining an mTOR inhibitor with bevacizumab increased the effectiveness of such treatment. Therefore, exploring the mechanisms activated upon VEGF signaling inhibition and next their attenuation could avoid failure and simultaneously improve the efficacy of therapy (110).

Growth factors released by cancer cells also recruit bone marrow-derived cells into the tumor microenvironment: monocytes/macrophages, endothelial precursor cells (EPCs), myeloid-derived suppressor cells (MDSCs) and cancer-associated fibroblasts (CAFs). These cells contribute to the induction of resistance to antiangiogenic therapy (105). Anti-VEGF agents have been shown to induce the expression of factors such as stromal derived factor-1 (SDF-1), PIGF, stem cell factor (SCF), interleukin 6 (IL-6) and others that are involved in recruitment of bone marrow-derived cells (111). The resistance to sunitinib was correlated in studies of metastatic renal cell carcinoma (mRCC) with infiltration of the tumor tissue with CD11b+Gr1+ myeloid cells that apart from sustaining suppression of immune cells produce proangiogenic factors (112). A study with the use of an anti-Gr1 antibody and anti-VEGF treatment showed improved tumor growth inhibition compared to anti-VEGF alone and delayed the onset of refractoriness (113). It was found that CD11b+Gr1+ cells, upon G-CSF stimulation,
express the Bv8 protein, known as prokineticin-2, mediator of VEGF-independent angiogenesis. Blocking Bv8 with neutralizing antibody caused angiogenesis inhibition and tumor growth and together with anti-VEGF antibodies exhibit an additive effect (114). Angiogenesis and the immune system are bidirectionally dependent; therefore, appropriate knowledge of their relationship may help in developing new effective treatment strategies (115). Tumor-associated macrophages (TAMs) contribute to resistance to antiangiogenic therapy by secretion of a number of proangiogenic factors. Moreover, these cells secrete MMPs that degrade extracellular matrix with concomitant release of matrix-sequestered growth factors that contribute to tumor growth and angiogenesis (116).

Tumors are also infiltrated with stromal cells, such as CAFs or pericytes, which are also engaged in resistance to antiangiogenic agents (105). CAFs contribute to tumor angiogenesis through secretion of angiogenic factors, whilst via production of SDF-1 they recruit bone-marrow endothelial progenitor cells (VEGF-independent mechanism) (117). It has also been shown that in tumors resistant to anti-VEGF therapy CAFs expressed pro-angiogenic PDGF-C. Blocking PDGF-C by neutralizing antibodies inhibited angiogenesis and in combination with anti-VEGF antibody revealed an additive effect (118). In turn, pericytes are recruited in response to PDGF-B released by ECs, and are responsible for vessel stabilization and maturation (119). The role of pericytes is to protect ECs from antiangiogenic agents, as well as to inhibit EC proliferation. After the treatment with angiogenesis inhibitors, an increase in pericyte coverage microvessels could be observed (105). On the other hand, enhancing tumor vessel covering by pericytes, the vessel maturation and resultant decreased leakiness may improve chemotherapy delivery. Therefore, the role of pericytes and PDGF-B mediated signaling in resistance to antiangiogenic therapy requires further study (103).

Tumor vascularization may be a result of a few different potential mechanisms. Apart from angiogenesis, cancer may achieve new vasculature by vessel co-option (using existing vessels), vascular mimicry (the process of forming vessels from tumor cells) and vasculogenesis (involving bone marrow-derived progenitor cells) (120). For example, in glioblastoma multiforme it was shown that VEGF signaling inhibition caused more invasive tumors and it was proposed that activation of

Figure 3. Resistance to antiangiogenic therapy. Acquired resistance to antiangiogenic therapy may result from a few possible mechanisms: cancer cells produce multiple proangiogenic factors and activate alternative signaling pathways other than VEGF/VEGFR, recruit bone-marrow derived cells (MDSCs, EPCs) and stromal cells [cancer-associated fibroblasts (CAFs), pericytes] into tumor microenvironment, or use other than angiogenesis mechanisms such as vessel co-option and vessel mimicry.
MET (the cellular receptor for HGF) after inhibition of VEGF signaling, as well as tumor-derived EC-induced angiogenesis and vasculogenic mimicry, could be engaged in anti-VEGF therapy resistance (121).

New possible mechanisms of tumor escape from antiangiogenic therapy include: EC heterogeneity, antiangiogenic VEGF, extracellular vesicles, lysosomal sequestration, glycosylation-dependent resistance and genetic polymorphism (reviewed in ref. 105).

7. Conclusion

The discovery of tumor angiogenesis and the subsequent concept of antiangiogenic therapy was a great breakthrough in anticancer treatment and improved our knowledge of the biology of cancer. In many cases, antiangiogenic agents when added to standard chemotherapy offered an improvement in therapeutic efficacy with different cancers: colorectal, breast, non-small cell lung cancer and hepatocellular carcinoma. However, a decade after approval of the first antiangiogenic agents, today all the above issues and obstacles related to antiangiogenic therapy in solid tumors have to be reconsidered in order to offer appropriate treatment for patients. Combining knowledge of the mechanisms of resistance to antiangiogenic therapy, the relationship between angiogenesis and immunity in cancer, validation of prognostic and predictive biomarkers, and targeting multiple signaling molecules, but with rationally designed schedule, may advance anticancer therapy and offer new promising results in the future.

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