Clinicopathologic significance and race-specific prognostic association of MYB overexpression in ovarian cancer

Orlandric Miree1,2, Sanjeev Kumar Srivastava1,2, Mohammad Aslam Khan1,2, Fnu Sameeta1, Srijan Acharya1,2, Harrison Ndettan3, Karan Pal Singh3, Kate Louise Hertweck4, Santanu Dasgupta1,2,5, Luciana Madeira da Silva6, Rodney Paul Rocconi6, James Elliot Carter1, Seema Singh1,2,5 & Ajay Pratap Singh1,2,5*

Late diagnosis, unreliable prognostic assessment, and poorly-guided therapeutic planning result in dismal survival of ovarian cancer (OC) patients. Therefore, identifying novel functional biomarker(s) is highly desired for improved clinical management. MYB is an oncogenic transcription factor with emerging functional significance in OC. Here we examined its clinicopathologic significance by immunohistochemistry and TCGA/GTex data analyses. Aberrant MYB expression was detected in 94% of OC cases (n = 373), but not in the normal ovarian tissues (n = 23). MYB was overexpressed in all major epithelial OC histological subtypes exhibiting the highest incidence (~97%) and overall expression in serous and mucinous carcinomas. MYB expression correlated positively with tumor grades and stages. Moreover, MYB exhibited race-specific prognostic association. Moderate-to-high MYB levels were significantly associated with both poor overall- (p = 0.02) and progression-free (p = 0.02) survival in African American (AA), but not in the Caucasian American (CA) patients. Consistent with immunohistochemistry data, we observed significantly higher MYB transcripts in OC cases (n = 426) than normal ovary (n = 88). MYB transcripts were significantly higher in all epithelial OC subtypes, compared to normal, and its greater levels predicted poor survival in AA OC, but not CA OC, patients. Thus, MYB appears to be a useful clinical biomarker for prognostication, especially in AA patients.

Ovarian cancer (OC) is the fifth most common cancer among women in the United States1. It is also the second most common gynecological malignancy in American women, causing more deaths than any other cancer of the female reproductive system2. American Cancer Society estimates that 21,410 new OC cases will be diagnosed in 2021, and it will claim approximately 13,770 lives this year3. It is also reported that OC disproportionately affects women of certain racial and ethnic backgrounds, especially African American (AA) women. AA women experience higher mortality than Caucasian American (CA) women regardless of tumor stage, grade, and histology despite having slightly lower incidence (9.5 per 100,000 versus 12.9 per 100,000)3–5. While the five-year survival rate has improved for CA women in the past decade, it has worsened in AA women with an ovarian cancer diagnosis6. Underlying causes of such disparity are unclear, but a late diagnosis and the lack of therapeutic options for the advanced disease are considered prime reasons for the overall grim prognosis of OC.

OC is a highly heterogeneous group of diseases, of which most (~90%) are of epithelial origin. The epithelial OC (EOC) is subdivided into four major histological subtypes: serous (high- and low-grade), endometrioid, clear cell, and mucinous. Serous carcinoma (SC) is the most common histological subtype (~70%), followed by endometrioid (EC), mucinous (MC), and clear cell carcinoma (CCC)6–9. Increasing evidence suggests that these EOC histotypes have different cells of origin and also vary in genetic susceptibilities and clinical responses to
therapy. Significant molecular differences also exist among histological subtypes, which remain to be characterized fully for clinical exploitation in biomarker and therapeutic development.

MYB is a proto-oncogene that encodes for a transcription factor protein. Defects in the chromosomal region harboring MYB were first reported in human acute myelogenous leukemia. Since then, its overexpression and other abnormalities have been detected in several other malignancies. MYB plays an essential role in maintaining the undifferentiated proliferative state of immature hematopoietic cells, and its knockout causes the loss of blood cell lineages in mice and is embryonically lethal. Information on MYB functions in OC pathobiology has begun to emerge recently, underscoring the need to study its expression pattern and clinicopathologic significance in OC.

In the present study, we investigated MYB expression in situ by immunohistochemistry in normal and malignant ovarian tumors. We also evaluated MYB mRNA expression in OC and its clinicopathologic correlation using publicly available transcriptomic databases. Our findings demonstrate significant MYB overexpression in all EOC histological subtypes, with the most pronounced expression detected in OC and MC. MYB at the protein level correlated positively with increasing tumor-grade and clinical stages. Moderate-to-high MYB protein levels also predicted worse progression-free survival. Although no significant difference in MYB expression was observed between AA and CA OC, high MYB expression was a stronger predictor of poor overall and progression-free survival in AA women. Thus, our study suggests that MYB can be a useful diagnostic and prognostic biomarker for ovarian malignancy, particularly in AA patients.

**Results**

**MYB is overexpressed in ovarian cancer.** To analyze MYB expression in OC, we performed immunohistochemical analysis on tissue sections from the normal ovary (n = 23) and ovarian tumors (n = 373). As expected, we observed predominant nuclear staining for MYB with some diffuse staining in the cytoplasm. No expression of MYB was detected in any of the normal ovarian tissues examined, whereas 94% of the OC cases stained positive for MYB. Of the positively stained OC cases, 60% (225/373) exhibited strong (3+) staining, whereas moderate (2+) and weak (1+) staining was detected in 23% (85/373) and 11% (40/373) cases, respectively (Fig. 1A). The overall mean composite score, reflective of percent positivity and staining intensity, was 7.7 ± 3.7 for all OC cases combined. Among the different histological subtypes, the highest incidence of MYB positivity was recorded in serous carcinoma (97%, 249/256) and mucinous carcinoma (97%, 36/37), followed by clear-cell carcinoma (83%, 10/12), and endometrioid carcinoma (75%, 9/12). The composite score distribution in these subtypes revealed the highest mean composite score for serous carcinoma (8.12 ± 3.6) followed by mucinous (8.05 ± 3.3), clear cell (6.5 ± 4.4), and endometrioid carcinoma (4.5 ± 3.7) (Fig. 1B). We also examined if the MYB expression varied between OC cases from CA and AA patients but did not observe any significant difference (Supplementary Figure S1).

**MYB exhibited a positive association with progressive clinical stages and histological grades.** We next examined changes in MYB expression pattern in progressive histological grades and clinical stages. Our cohort had 14% (52/373) tumors from grade I, 28% (106/373) from grade II, and 35% (131/373) from grade III. Similarly, 47% (174/373) tumors were from stage I, 11% (43/373) from stage II, 18% (69/373) from stage III, and 8% (29/373) from stage IV. Overall, we observed a positive association of MYB expression (mean composite score) with the increasing histological grade of ovarian tumors. Specifically, an increase in MYB expression significantly correlated with the grade, and it changes from grade I to II (OR = 2.579, 95% CI = 1.278–5.207, p = 0.008) and from II to III (OR = 3.751, 95% CI 2.165–6.499, p = <0.001) (Fig. 2A). Furthermore, the stage-wise progression was also found to be positively associated with increased MYB expression. While the cancer stages change from stage I to II (OR = 3.230, 95% CI 1.393–7.492, p = 0.006), II to III (OR = 5.255, 95% CI 1.866–14.800, p = 0.002), and III to IV (OR = 2.521, 95% CI 1.013–6.275, p = 0.047), respectively (Fig. 2B).

**MYB overexpression is a predictor of poor disease outcomes, especially in African American ovarian cancer patients.** Since MYB expression exhibited a positive association with increasing tumor grade and stage, we next examined if MYB levels correlated with the survival of OC patients by performing Kaplan–Meier analysis. Survival data were available for a total of 74 OC cases, of which 36 and 38 belonged to AA and CA women with OC, respectively. We first used a mean composite score cut-off value of ≤ 6 as low MYB and ≥ 6 as high MYB to examine a survival association. Our data demonstrated a trend of poor -overall and -progression-free survival in AA OC patients with high MYB expression; however, it failed to reach the statistical significance (Supplementary Figure S2). Next, we examined the survival association by keeping the mean composite score of ≤ 4 as a cut-off for the low MYB-expressing group, and cases with a composite score of > 4 were categorized as moderate to high MYB-expressing group. Based on this criterion, we observed a significantly worse progression-free survival (p = 0.04) of the OC patients with moderate/high MYB expression than those with low MYB expression. In addition, a clear trend towards poor overall survival of the OC patients with moderate to high MYB expression was also recorded compared to the low MYB group (p = 0.05) (Fig. 3A). Since women of AA racial background have greater mortality from OC despite having a lower incidence than the CA women, we examined the prognostic association of MYB separately in these groups. We observed a significantly worst overall (p = 0.02) and progression-free (p = 0.02) survival of the AA subjects with moderate to high MYB expression when compared to the low MYB expressing group (Fig. 3B). However, we did not observe a significant change in overall and progression-free survival of the CA subjects with mod/high MYB expression compared to the low MYB expressing group (Fig. 3C). Thus, our data suggest that MYB could be a stronger predictor of survival in AA women with OC diagnosis.
Expression pattern of MYB transcript and its prognostic association in OC patients. To further generate support for our observations, we next analyzed the transcriptomic data available in multiple public databases, including GTEx (Genotype-Tissue Expression), TCGA-ovarian cancer project (The Cancer Genome Atlas), and individual laboratory studies. These databases were searched by using interactive analytical platforms, GEPIA, UALCAN, and ONCOMINE. The data mining revealed significantly higher expression of MYB transcript (false discovery rate-adjusted) in a large ovarian cancer cohort (n = 426) compared to the normal
ovarian tissues (n = 88) (Fig. 4A). A differential expression of MYB transcripts among histological OC subtypes was also recorded (Fig. 4B, C). No significant differences in MYB expression were recorded in AA and CA OC cases in the TCGA database (Fig. 4D). Similarly, no significant association of MYB transcript levels with clinical stages or histological grade was observed (Fig. 4E, F). MYB also did not significantly associate with overall or progression-free survival (Fig. 5A, B). When examined in AA and CA groups, a significant association of high MYB with lower survival was recorded in AA women with OC (p = 0.021) but not in CA women (p = 0.57) (Fig. 5C, D). These data suggest that MYB is aberrantly expressed at both RNA and protein levels in OC and could serve as a predictor of survival, especially in AA women with OC. Further, MYB should also be explored as a functional target associated with racially disparate clinical outcomes in AA women.

Discussion
The scarcity of reliable screening tools for early-stage diagnosis is one of the major challenges associated with the poor survival of OC patients. Further, lack of clarity about the cell of origin and molecular and histological complexities make the therapeutic planning and prognostication of epithelial OC clinically challenging. Identification of functional biomarkers associating with disease aggressiveness and clinical outcomes could allow for better therapeutic planning, disease monitoring, and the development of novel targeted therapeutics.
Indeed, existing and emerging studies continue to demonstrate the strength of molecular markers in accurate histopathological and clinical diagnosis, prognosis, and treatment guidance. The present study identified MYB as a potentially useful clinical biomarker for OC. Like other transcription factors, activated MYB is localized in the nucleus, where it regulates the expression of genes associated with different cellular processes\(^1\)\(^{,}\)\(^{11}\)\(^{,}\)\(^{17}\)\(^{,}\)\(^{25}\). The earliest documentation of aberrant MYB expression in OC came from a study that examined MYB transcripts in OC cell lines and a few clinical cases\(^2\)\(^{,}\)\(^{26}\). Our study examined MYB expression in a larger cohort of normal and ovarian cancer cases and found its widespread overexpression in all EOC histological subtypes. Staining intensity varied (weak to strong) between and within cases suggestive of the heterogeneous disease or the cells in various progression stages. The variable expression of MYB in OC cases could also represent molecular heterogeneity resulting from genetic differences or epigenetic alterations in cell populations due to their exposure to the differing tumor microenvironment.

MYB expression was not detected in any of the normal ovary cases suggesting that its dysregulation occurs following malignant transformation, and thus it could be of potential diagnostic significance. As expected, MYB expression was predominantly nuclear, although diffuse staining in the cytoplasm was also detected. Therefore, it will be interesting to pursue if MYB also has any non-nuclear functions besides gene regulation. Further, among all histological subtypes, SC and MC exhibited the highest expression. \(^{p53}\) mutations have been reported in both these subtypes\(^1\)\(^{,}\)\(^{10}\). Thus, molecular crosstalk between MYB and \(p53\) can aid SC and MC, requiring further validation in preclinical and clinical settings.

Irrespective of the histological subtypes, MYB protein expression positively correlated with OC's progressive histological grades and clinical stages. This is in line with an established role of MYB in cellular differentiation and promotion of malignant behavioral features\(^2\)\(^{,}\)\(^{16}\)\(^{,}\)\(^{17}\)\(^{,}\)\(^{25}\)\(^{,}\)\(^{27}\). It also underscores the need to delineate the molecular mechanisms involved in MYB dysregulation in OC to develop disease prevention and therapy strategies. Prediction of worst progression-free survival by the moderate-to-high MYB protein expression in OC cases further strengthens the notion that MYB plays an important role in disease aggressiveness. Moreover, stronger MYB association with poor survival in AA patients is also exciting and warrants further investigations. This is quite interesting and suggests that MYB likely acts in concert with other molecular factors in AA OC to impact tumor biology leading to racially disparate clinical outcomes. In fact, we have previously observed that increased expression of indoleamine 2.3-dioxygenase in AA OC patients associated with poor survival\(^2\)\(^8\). Thus, it will be interesting to examine a mechanistic link of MYB with other proteins exhibiting racially disparate expression in OC. It is also interesting to note that a lower cutoff of MYB expression levels provided a better correlation with

---

**Figure 4.** Transcriptomic analysis of MYB in ovarian cancer patients. (A) Significantly higher MYB mRNAs were present in ovarian cancer cases than in the normal ovary. (B, C) A variable but increased expression of MYB transcripts was also observed in different histological subtypes. (D) MYB transcript levels were not significantly different in AA (n = 26) and CA (n = 252) OC cases. (E, F) Similarly, no significant association of MYB mRNA levels was observed with clinical stages and histological grades. A \(p\) value of < 0.05 was considered significant.
survival outcomes. It suggests that MYB being a transcription factor can significantly impact the transcriptome and, as a result, cellular phenotypes upon slight increases in its expression.

Aberrant expression of MYB was also apparent at the mRNA level, suggesting that its dysregulation in OC likely occurs at the genomic (gene amplification), transcriptional and/or post-transcriptional levels. Indeed, in other cancers, both gene amplification and transcriptional upregulation of MYB have been reported\(^{11,14}\). Further, post-transcriptional regulation via microRNAs is also documented\(^{21,29}\). Unlike protein, the MYB transcript did not significantly associate with histological grades and clinical stages due to the smaller sample size. A statistically significant association of higher MYB mRNA levels with poor survival outcomes in AA patients further supports its role in racial disparity requiring further investigations.

In conclusion, our study establishes the clinical significance of MYB in OC that should be explored further in combination with other biomarkers. Functional and in-depth mechanistic studies including identification of transcriptional targets of MYB in OC should also be pursued. These efforts will yield information that could be exploited to develop MYB-targeted therapeutic strategies and clinical utilization of MYB as a useful biomarker of diagnostic and prognostic significance.

**Materials and methods**

**Tissue samples, antibodies and reagents.** A total of 373 ovarian cancer and 23 normal ovary tissues were used in this study. The de-identified formalin-fixed paraffin-embedded tissue sections (5 μm) were either obtained through the commercial source (US Biomax, Inc., Rockville, MD, USA) or the University of South Alabama Health Biobank under an Institutional Review Board-approved protocol. Polyclonal anti-MYB rabbit antibody was obtained from Abcam (Cambridge, MA). Betazoid DAB Chromogen Kit, Da Vinci Green diluent, peroxidased 1, MACH 3 Rabbit HRP Polymer Detection, Background Sniper, Diva Decloaker, wash buffer (Tris-buffered saline with Tween 20), Tacha’s bluing solution (Biocare Medical, Concord, CA), ethanol (Thermo Fisher Scientific, Waltham, MA).

**Immunohistochemistry.** IHC was performed on formalin-fixed, paraffin-embedded normal or ovarian tumor tissue sections as previously described\(^{25}\). In brief, tissue sections were deparaffinized, hydrated, and subjected to antigen retrieval in Decloaking Chamber (Biocare Medical, Concord, CA). After that, peroxide
quenching was done with 3% H$_2$O$_2$ in methanol at room temperature for 30 min followed by noise blocking for 10 min with Background Sniper. Sections were then incubated with anti-MYB antibody overnight at 4 °C, washed with Tris-containing buffer, and incubated at room temperature with respective polymer and probe for 10 min each. Immunoreactivity was visualized by treating the sections with DAB chromogen followed by hematoxylin counterstaining.

**Quantification of MYB staining.** All the slides were viewed under a Zeiss Axioskop 40 (White Plains, NY) and scored for the nuclear staining of MYB on a three-point scale, 1+ (weak); 2+ (moderate); and 3+ (strong), and the percent positivity on a four-point scale, 0–25% (‘1’), 26–50% (‘2’), 51–75% (‘3’), and 76–100% (‘4’) using standard scoring recommendation $^{10}$. Weak (1+) staining in less than 10% of cells was considered negligible and given a value of ‘0’. A composite score was calculated for each section by multiplying the staining intensity and percent positivity values, ranging from 0 to 12 $^{10}$.

**In silico analysis for differential MYB transcript estimation.** To examine the differential expression of MYB at the transcript level in normal ovary versus ovarian cancer, using different public databases, including GTEx (Genotype-Tissue Expression), TCGA-ovarian cancer project (The Cancer Genome Atlas), and individual laboratory studies. These databases were searched by using interactive analytical platforms, GEPIA (http://gepia.cancer-pku.cn/), UALCAN (http://ualcan.path.uab.edu/), and ONCOMINE (https://www.oncomine.org). Besides, an association of MYB mRNA with race, clinical stages, histological grades, and patient survival was also studied.

**Statistical analysis.** We employed Wilcoxon, Mann–Whitney, Anova, Chi-square, Fisher’s exact tests, and logistic regression model as appropriate for all statistical data comparisons between various experimental groups. Overall survival curves were developed using the Kaplan–Meier method and compared using the stratified log-rank test. All p-values were two-sided, and all confidence intervals were at the 95% level. A P-value of < 0.05 was considered statistically significant. Computation for all the analyses was performed using the Statistical Analysis System (SAS).

**Data availability**
The data presented in this article is available for sharing upon a reasonable request.

Received: 20 April 2021; Accepted: 3 June 2021
Published online: 18 June 2021

**References**

1. Siegel, R. L., Miller, K. D., Fuchs, H. E. & Jemal, A. Cancer statistics, 2021. *CA Cancer J. Clin.* 71, 7–33. https://doi.org/10.3322/caac.21654 (2021).

2. Stewart, C., Ralyea, C. & Lockwood, S. Ovarian cancer: An integrated review. *Semin. Oncol. Nurs.* 35, 151–156. https://doi.org/10.1016/j.socn.2019.02.001 (2019).

3. Srivastava, S. K. *et al.* Racial health disparities in ovarian cancer: Not just black and white. *J. Ovarian Res.* 10, 58. https://doi.org/10.1186/s13048-017-0355-y (2017).

4. Dilley, S. *et al.* Do differences in medical comorbidities and treatment impact racial disparities in epithelial ovarian cancer? *Gynecol. Oncol.* 149, 49–52. https://doi.org/10.1016/j.ygyno.2017.10.035 (2018).

5. Ross, J. *et al.* Unraveling the etiology of ovarian cancer racial disparity in the deep south: Is it nature or nurture?. *Gynecol. Oncol.* 145, 329–333. https://doi.org/10.1016/j.ygyno.2017.02.025 (2017).

6. Singh, A. P. *et al.* Clinical potential of mycin in diagnosis, prognosis, and therapy of ovarian cancer. *Lancet Oncol.* 9, 1076–1085. https://doi.org/10.1016/S1470-2045(08)70277-8 (2008).

7. Bell, D. A. Origins and molecular pathology of ovarian cancer. *Mod. Pathol.* 18(Suppl 2), S19-32. https://doi.org/10.1038/modpathol.3800306 (2005).

8. Lheureux, S., Gourley, C., Vergote, I. & Oza, A. M. Epithelial ovarian cancer. *CA Cancer J. Clin.* 69, 49–52. https://doi.org/10.3322/caac.21689–21694. https://doi.org/10.1073/pnas.0907643106 (2012).

9. Chauhan, S. C. *et al.* Ablation expression of MUC4 in ovarian carcinoma: Diagnostic significance alone and in combination with MUC1 and MUC16 (CA125). *Mod. Pathol.* 19, 1386–1394. https://doi.org/10.1038/modpathol.3800646 (2006).

10. Dion, L. *et al.* The landscape and therapeutic implications of molecular profiles in epithelial ovarian cancer. *J. Clin. Med.* 9, 66. https://doi.org/10.3390/jcm9022359 (2020).

11. Araz, S. *et al.* Deep sequencing and in silico analyses identify MYB-regulated gene networks and signaling pathways in pancreatic cancer. *Sci. Rep.* 6, 28446. https://doi.org/10.1038/srep28446 (2016).

12. Gonda, T. J. & Metcalf, D. Expression of myb, myc and fos proto-oncogenes during the differentiation of a murine myeloid leukemia. *Biochim. Biophys. Acta*. 710, 193–199. https://doi.org/10.1016/0005-2736(82)90141-9 (1982).

13. Braddock, M. C. *et al.* c-Myb and Bcl-x overexpression predicts poor prognosis in colorectal cancer: clinical and experimental findings. *Am. J. Pathol.* 158, 1289–1299. https://doi.org/10.1016/S0002-9440(10)60408-1 (2001).

14. Greco, C. *et al.* Activation of c-MYC and c-MYB proto-oncogenes is associated with decreased apoptosis in tumor colon progression. *Anticancer Res.* 21, 3185–3192 (2001).

15. Persson, M. *et al.* Recurrent fusion of MYB and NFIb transcription factor genes in carcinomas of the breast and head and neck. *Proc. Natl. Acad. Sci. USA* 106, 18740–18744. https://doi.org/10.1073/pnas.0909114106 (2009).

16. Srivastava, S. K. *et al.* MYB is a novel regulator of pancreatic tumour growth and metastasis. *Br. J. Cancer* 113, 1694–1703. https://doi.org/10.1038/bjc.2015.400 (2015).

17. Srivastava, S. K. *et al.* Myb overexpression overrides androgen depletion-induced cell cycle arrest and apoptosis in prostate cancer cells, and confers aggressive malignant traits: Potential role in castration resistance. *Carcinogenesis* 33, 1149–1157. https://doi.org/10.1093/carcin/bgp1134 (2012).

18. Lies, Y. K. & Reddy, E. P. Conditional c-myc knockout in adult hematopoietic stem cells leads to loss of self-renewal due to impaired proliferation and accelerated differentiation. *Proc. Natl. Acad. Sci. USA* 106, 21689–21694. https://doi.org/10.1073/pnas.0907623106 (2009).
19. Soza-Ried, C., Hess, I., Netuschil, N., Schorpp, M. & Boehm, T. Essential role of c-myb in definitive hematopoiesis is evolutionarily conserved. *Proc. Natl. Acad. Sci. USA* **107**, 17304–17308, https://doi.org/10.1073/pnas.1009464107 (2010).

20. Zhang, X. Y., Li, Y. F., Ma, H. & Gao, Y. H. Regulation of MYB mediated cisplatin resistance of ovarian cancer cells involves miR-21-wnt signaling axis. *Sci. Rep.* **10**, 6893, https://doi.org/10.1038/s41598-020-63396-8 (2020).

21. Tung, C. H. et al. MicroRNA-150-5p promotes cell motility by inhibiting c-Myb-mediated Slug suppression and is a prognostic biomarker for recurrent ovarian cancer. *Oncogene* **39**, 862–876, https://doi.org/10.1038/s41388-019-1025-x (2020).

22. Cronin, K. A. et al. Racial disparities in the receipt of guideline care and cancer deaths for women with ovarian cancer. *Cancer Epidemiol. Biomark. Prev.* **28**, 539–545, https://doi.org/10.1158/1055-9965.EPI-18-0285 (2019).

23. Arora, T., Mullangi, S. & Lekkala, M. R. Ovarian cancer. [Updated 2021 Feb 22]. In *StatPearls* [Internet] (StatPearls Publishing, Treasure Island, FL, 2021). Available from: https://www.ncbi.nlm.nih.gov/books/NBK567760/.

24. Lockwood-Rayermann, S., Donovan, H. S., Rambo, D. & Kuo, C. W. Women's awareness of ovarian cancer risks and symptoms. *A.P. Am. J. Nurs.* **109**, 36–45, https://doi.org/10.1097/01.NAJ.0000360339.08701.73 (2009).

25. Bhardwaj, A. et al. MYB promotes desmolipia in pancreatic cancer through direct transcriptional up-regulation and cooperative action of sonic hedgehog and adrenomedullin. *J. Biol. Chem.* **291**, 16263–16270, https://doi.org/10.1074/jbc.M116.732651 (2016).

26. Barletta, C. et al. C-MYB activation and the pathogenesis of ovarian cancer. *Eur. J. Gynaecol. Oncol.* **13**, 53–59 (1992).

27. Zubair, H. et al. Proteomic analysis of MYB-regulated secretome identifies functional pathways and biomarkers: Potential pathobiological and clinical implications. *J. Proteome Res.* **19**, 794–804, https://doi.org/10.1021/acs.jproteome.9b00641 (2020).

28. Madeira da Silva, L., Starenki, D., Scalici, J. M. & Rocconi, R. P. Molecular determinants of immune response in ovarian cancer racial disparity: can selective immunotherapy optimize therapy and close the disparity gap? *Gynecol. Oncol.* **154**(1), 3, https://doi.org/10.1016/j.ygyno.2019.04.036 (2019).

29. Sun, X., Zhang, C., Cao, Y. & Liu, E. miR-150 suppresses tumor growth in melanoma through downregulation of MYB. *Oncol. Res.* **28**, 539–545, https://doi.org/10.1158/1055-9965.EPI-18-0285 (2019).

30. Remmelle, W. & Stegner, H. E. Recommendation for uniform definition of an immunoreactive score (IRS) for immunohistochemical estrogen receptor detection (ER-ICA) in breast cancer tissue. *Pathol. Oncol. Res.* **8**, 138–140 (1987).

Acknowledgements
The authors would like to acknowledge the funding from NIH/NCI [R01CA224306, R01CA175772, and U01CA185490 (to APS)].

Author contributions
A.P.S. conceived the idea; O.M., S.K.S., M.A.K., S.S. designed the experiments; O.M., S.K.S., S.D. collected and analyzed the data; H.N., K.P.S. performed statistical analysis; K.L.H. performed bioinformatics analysis; J.E.C., F.S. reviewed and scored immunostaining in clinical specimens; A.P.S., R.P.R., L.M.S. procured the resources; M.A.K., O.M., and S.D. wrote the manuscript; S.S., A.P.S., R.P.R., L.M.S. reviewed the manuscript; and A.P.S. supervised the study. All the authors have read and approved the manuscript.

Competing interests
The authors declare no competing interests.

Additional information
Supplementary Information The online version contains supplementary material available at https://doi.org/10.1038/s41598-021-92352-3.

Correspondence and requests for materials should be addressed to A.P.S.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

© The Author(s) 2021