Re-Expression of AKAP12 Inhibits Progression and Metastasis Potential of Colorectal Carcinoma In Vivo and In Vitro

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Abstract

Background: AKAP12/Gravin (A kinase anchor protein 12) is one of the A-kinase scaffold proteins and a potential tumor suppressor gene in human primary cancers. Our recent study demonstrated the highly recurrent loss of AKAP12 in colorectal cancer and AKAP12 reexpression inhibited proliferation and anchorage-independent growth in colorectal cancer cells, implicating AKAP12 in colorectal cancer pathogenesis.

Methods: To evaluate the effect of this gene on the progression and metastasis of colorectal cancer, we examined the impact of overexpressing AKAP12 in the AKAP12-negative human colorectal cancer cell line LoVo, the single clone (LoVo-AKAP12) compared to mock-transfected cells (LoVo-CON).

Results: pCMV6-AKAP12-mediated AKAP12 re-expression induced apoptosis (3% to 12.7%, p<0.01), migration (89.6±7.5 cells to 31.0±4.1 cells, p<0.01) and invasion (82.7±5.2 cells to 24.7±3.3 cells, p<0.01) of LoVo cells in vitro compared to control cells. Nude mice injected with LoVo-AKAP12 cells had both significantly reduced tumor volume (p<0.01) and increased apoptosis compared to mice given AKAP12-CON. The quantitative human-specific Alu PCR analysis showed overexpression of AKAP12 suppressed the number of intravasated cells in vivo (p<0.01).

Conclusion: These results demonstrate that AKAP12 may play an important role in tumor growth suppression and the survival of human colorectal cancer.

Introduction

Colorectal cancer is the third most common form of cancer in the world and the leading cause of cancer mortality[1–3]. Better understanding of the mechanisms underlying colorectal cancer formation and progression is greatly needed because only modest improvements in the survival of colorectal cancer patients have been achieved over the last decade.

A-kinase anchor protein 12 (AKAP12/Gravin) was first isolated as a protein recognized by the serum of myasthenia gravis patients[4]. It is one of the A-kinase anchoring proteins (AKAPs) that belong to a family of scaffold proteins, and organizes the protein kinase A (PKA) and C (PKC)[5]. It is also an important regulator of the β2-adrenergic receptor complex, which controls cell signaling, cell adhesion, mitogenesis and differentiation[6,7]. AKAP12 has been mapped to chromosome 6q24–25.2, a cancer deletion hotspot [8]. DNA hypermethylation in the AKAP12 promoter region, and the accompanied underexpression of the corresponding gene, has been noted in a variety of human cancers, including gastric cancer, esophageal cancer and lung cancer, and in myeloma cells and myeloid malignancies[6,8–11]. Downregulation of AKAP12 expression suggests that the inactivation of AKAP12 expression may be linked to oncogenesis. A recent report using microarray data from in vivo genetic searches indicated that methylation is associated with the downregulation of AKAP12 in colon cancer and identified AKAP12 as a potential tumor suppressor gene candidate[12]. In our previous study, downregulation or loss of AKAP12 mRNA expression was detected in 68.9% (31/45) of colorectal carcinoma tissues and methylation of the AKAP12 promoter region was detected in 77.8% (35/45) of these tissues compared with 13.3% (6/454) in the adjacent tissue[13]. Complete loss of AKAP12 and hypermethylation of the promoter was detected in colorectal cancer cell lines, LoVo, SW40 and COLO320. However, it remains unclear if AKAP12 plays an inhibitory role in the progression or metastasis of colorectal cancer.

In this report, we analyzed the effect of AKAP12 on the tumorigenesis and metastasis of LoVo cells in vitro and in vivo. Our data demonstrate that the re-expression of AKAP12 could inhibit the progression and metastatic potential of colorectal carcinoma.
Therefore, we propose that AKAP12 functions as a tumor suppressor of this solid cancer.

Results

AKAP12 inhibited cell growth and induced apoptosis

To examine the impact of the AKAP12 re-expression in colorectal cancer cells, we constructed a vector constitutively expressing AKAP12 (pCMV6-AKAP12) to restore AKAP12 expression in LoVo cells, which lack AKAP12 expression in our previous study [13]. We stably transfected LoVo cells with the pCMV6-AKAP12 or empty control vector, respectively. Significant AKAP12 mRNA and protein re-expression in LoVo-AKAP12 cells compared with LoVo-CON was observed [13]. We also analyzed cell proliferation using a WST assay and the growth of LoVo cells in soft agar. Strongly inhibited the growth of LoVo cells and a significant decrease in the number of colonies was observed when tumor cells were transfected with pCMV6-AKAP12 [13].

Given the effects of AKAP12 on cell proliferation and viability, using PI and Annexin V staining, we next examined whether pCMV6-AKAP12 transfection-induced apoptosis of LoVo cells. Flow cytometry revealed that transfection of LoVo cells with pCMV6-AKAP12 induced a marked increase in the percentage of apoptotic (annexin V+ PI+) cells compared with the LoVo cells with control vectors (12.7% versus 3.0%, respectively; p < 0.01) (Fig. 1A). We also evaluated the expression of cleaved-caspase-3 (the active form of caspase-3, Fig. 1B) in these two cell lines. The results indicate that cells transfected with the Lovo-AKAP12 can significantly increase cleaved-caspase-3 level.

Figure 1. Effects of AKAP12 on apoptosis of LoVo cells transfected with pCMV6-AKAP12. (A) Effects of AKAP12 expression on apoptosis in AKAP12-transfected LoVo cells. LoVo-AKAP12 cells and LoVo-CON cells were analyzed using FACS and PI/Annexin V staining. The legends show the percentage of cells undergoing apoptosis. (B) Western blot analysis of cleaved-caspase-3 in Lovo-CON and Lovo-AKAP12 cells. AKAP12 increase cleaved-caspase-3 in cultured Lovo cells.

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Oncosuppressive effect of AKAP12 on LoVo cells

Given that migration and invasion are key components of the metastatic cascade, we conducted in vitro migration and invasion assays to evaluate the migration and invasion potential of LoVo-AKAP12 and LoVo-CON cells. Overexpression of AKAP12 lead to an average decrease in migration of an average of 65.4% compared to the control cells (31.0±4.1 versus 89.6±7.5 cells per field, respectively; p<0.01) (Fig. 2A, B). Furthermore, overexpression of AKAP12 lead to an average of 70.1% (24.7±3.3 versus 82.7±5.2 cells per field, respectively; p<0.01) decrease invasiveness compared with the control cells (Fig. 2C, D). These results suggest that AKAP12 expression is inversely associated with the migration and invasion ability of colorectal cell lines LoVo in vitro.

AKAP12 inhibits tumorigenesis, metastatic and induced apoptosis in vivo

Because AKAP12 inhibits LoVo cell growth in vitro, we further assessed its effect on tumor formation in vivo. The anti-tumor activity of AKAP12 was evaluated in a primary xenograft model using nude mice. LoVo-AKAP12 cells, LoVo-CON cells and non-transfected LoVo-LIPO cells were injected into the flank of nude mice. Four weeks later, mice were sacrificed and examined for the presence of metastases in the lungs. Injection of LoVo-AKAP12 cells reduced tumor volume significantly compared with the LoVo-CON and LoVo-LIPO cells four weeks-post injection (p<0.01) (Fig. 3A, B). To assess if AKAP12 could reduce the formation of lung metastases, we determined the presence of Alu sequences in the lungs of nude mice injected with each type of LoVo cells using real-time PCR. The human tumor cell metastasis
Figure 2. AKAP12 suppresses \textit{in vitro} migration and invasion ability of LoVo cells. (A) LoVo-AKAP12 and LoVo-CON cells penetrated through the transwell chambers and were photographed at ×100 magnification. (B) The \textit{in vitro} migration ability of LoVo-AKAP12 cells and LoVo-CON cells was measured by determining the number of cells that migrated through the transwell chamber. Columns, mean values; bar, SD. * \(p<0.01\). (C) LoVo-AKAP12 and LoVo-CON cells that migrated through the transwell chambers and photographed at ×100 magnification. (D) The \textit{in vitro} migration ability of LoVo-AKAP12 cells and LoVo-CON cells was measured by determining the number of matrigel-coated cells that penetrated through the transwell chambers. Columns, mean values; bar, SD. * \(p<0.01\).
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formation in mouse lung did not differ between LoVo-CON and LoVo-LIPO recipients. However, the formation of lung metastases in mice given LoVo-AKAP12 cells (6/10) was significantly lower than the mice receiving either LoVo-CON (10/10) or LoVo-LIPO (10/10) cells (Table 1). Additionally, lung tissue from mice injected with LoVo-AKAP12 cells displayed significantly less human Alu expression than LoVo-CON or LoVo-LIPO control group (Fig. 3C). We also evaluated the expression of AKAP12 and cleaved-caspase-3 level in the tumor tissues of LoVo-CON or LoVo-AKAP12 group (Fig. 3D). The results indicate that LoVo-AKAP12 group can significantly increase AKAP12 and cleaved-caspase-3 level. These data indicated that ectopic expression of AKAP12 in LoVo cells decreases their metastatic ability and induced apoptosis in vivo.

Discussion

The biological role of AKAP12 in colorectal cancer progression and metastasis is poorly understood. In the present study, we examined this question through re-expression of AKAP12 in AKAP12-null cancer cells to determine if this would influence cancer cell growth and metastatic potential. AKAP12 inhibited cell growth, migration, invasion, anchorage-independent growth and induced apoptosis when transfected into LoVo cells. Furthermore, injection of LoVo-AKAP12, LoVo-CON or LoVo-LIPO cells into nude mice demonstrated that, in vivo, induced AKAP12 expression in the colorectal cancer cells suppressed both tumor growth and metastasis and induced apoptosis.

Cancer develops when the balance between cell proliferation and cell death is disturbed, and aberrant cell proliferation leads to tumor growth. In human gastric cancer cells, re-expression of AKAP12 lead to reduced colony formation and apoptotic cell death[8]. To examine whether modulation of AKAP12 expression influenced the tumorigenic properties of the colorectal cancer cells, we tested in vitro LoVo cell proliferation and apoptosis. As shown by both the WST assay and FACS analysis, AKAP12 displays tumor-suppressive activity in vitro. AKAP12-mediated suppression of tumor growth may be achieved through direct or indirect interaction with multiple apoptotic proteins and associated signaling pathways in cancer cells[14]. These include several signaling molecules that participate
in cell proliferation and cytoskeletal organization, as well as protein kinase C (PKC), protein kinase A (PKA), cyclin D1 and calmodulin[15–17]. In HT1080 cells, AKAP12 was found to suppress tumor cell viability by inducing apoptosis via caspase-3 and was associated with a decreased expression of Bcl-2 and increased expression of Bax[18]. Moreover, AKAP12 induced the expression of Cipl/p21 and Kipl/p27 and decreased the expression of cyclin D1[18]. SSeCKS, is the orthologue of human AKAP12 gene, re-expression was reported to result in the attenuation of critical Src-induced proliferative and pro-angiogenic gene expression, including Afp and Cdc20a, and cell cycle regulatory genes such as Ptpn11 and Gadd45a[19].

Cancer cells are capable of anchorage-independent growth and are able to travel and settle in a new site in which the microenvironment may be completely different[20,21], a process crucial for oncogenesis and cancer metastasis. The re-expression of AKAP12 in LoVo cells reduced colony formation and inhibited anchorage-independent growth ability, which may explain the mortality in human colorectal cancer.

AKAP12 in LoVo cells reduced colony formation and inhibited anchorage-independent growth ability, which may explain the hypothesis, our results demonstrated that, in vitro, induced expression of AKAP12 could reduce the invasion or migration ability of oncogenic cells and significantly suppress the metastatic potential of the colorectal cancer cells. AKAP12 also could inhibit colorectal cancer tumorgenesis and metastasis in vivo and suggests that AKAP12 may hold great promise for designing novel therapeutic strategies against this solid cancer.

Our study indicates that AKAP12 is a suppressive factor in colorectal cancer cells, inhibiting tumor cell growth and metastases in vitro and in vivo. Collectively, these findings demonstrated that AKAP12 may play an important role in the development of malignancies. Therefore, AKAP12 could serve as an effective target for the development of novel anti-cancer therapeutics and may be a useful biomarker for human colorectal cancer.

### Materials and Methods

#### Cell lines

The colorectal carcinoma cell line, LoVo, was obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA). Cells were cultured in F12K media (GibcoBRL, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (Sigma, St. Louis, MO, USA) and penicillin-streptomycin (GibcoBRL). Cells were incubated in 5% CO2 at 37°C, and only cells with a passage number of <10 were used in the experiments.

#### Recombinant vectors and stable transfection

pCMV6-XL4-AKAP12 (Cat No. SC110078) was purchased from Origene (Rockville, MD). The plasmids were purified, digested with XbaI and subcloned into the pCMV6-Neo expression vector. LoVo cells were transfected with the pCMV6-Neo-AKAP12 construct (LoVo-AKAP12), or the control empty vector (LoVo-CON). For stable transfections, 10 μg of plasmid DNA was resuspended in 500 μl of serum-free DMEM and mixed with 60 μl Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) in 500 μl of serum-free DMEM. The mixture was incubated for 20 min at room temperature and added to a 70%-80% confluent 100-mm tissue culture dishes. After 2 h, the transfection medium was changed to F12K medium containing 10% FBS. Transfection efficiency was typically 60–80% as determined by GFP expression. After 48 h, the transfection medium was changed to F12K medium containing 10% FBS and antibiotic geneticin (G418) (400 μg/ml) (Invitrogen Life Technologies, Inc., Carlsbad, CA). Clones were screened by limited dilution cloning 4 days after selection. AKAP12 expression levels using Western blots, and one clone with vector alone (LoVo-CON) and a clone overexpressing AKAP12 (LoVo-AKAP12) were selected for further experiments.

#### Flow cytometry assay

Analysis of apoptosis was determined by flow cytometry. LoVo-AKAP12 and LoVo-CON cells were seeded in six-well plates in 10% FBS/F12K and then incubated at 37°C. FACS analysis using propidium iodide (PI) staining combined with Annexin V-FTTC (BD, Franklin Lakes, N, USA) was performed. First, 106 cells were washed twice with cold PBS and resuspended in 1× binding buffer. Then, 105 cells were transferred to a 5 mL tube, 5 μL of Annexin V-FTTC and PI were added and the cells were incubated in the dark for 15 min at room temperature. The experiment was repeated three times.

#### Western blot analysis

Western blot analysis was performed as described previously[26]. Equal mounts of proteins from cells and tissues were separated by SDS-PAGE gels and then transferred to PVDF...
membranes. After blocking, the membranes were incubated with the appropriate primary antibodies overnight (anti-cleaved-caspase-3, 1:500, Cell Signaling; anti-β-actin, 1:500, Cell Signaling; anti-human AKAP12, 1:500, Sigma). Following three washes with TBST, the blots were incubated with the secondary horseradish peroxidase-conjugated antibody (1:30000) at room temperature for 1 h. Immunocomplexes were visualized by using enhanced chemiluminescence (BioRad) following the manufacturer’s instructions. The bands were subsequently analyzed densitometrically with Quantity One Software (BioRad).

In vitro cell migration and invasion assay

Cell migration was evaluated using the QCM 24-Well Colorimetric Cell Migration Assay Kit (Chemicon). In brief, LoVo-AKAP12 and LoVo-CON cells were plated at a density of 1 × 10^6 cells/mL on 8 μm inserts and cultured for 24 h. The non-migrating cells were removed from the upper surface of the insert with a cotton-tipped swab, and cells that have migrated are incubated with stain solution. Transfer the stained insert to a clean well containing 200 μL of Extraction Buffer for 15 minutes at room temperature. Extract the stain from the underside by gently tilting the insert back and forth several times during incubation. Remove the insert from the well. The insert could be counted manually through a microscope and five fields were evaluated for migration. The experiment was repeated three times.

The invasive capability of the transfected cells was assessed using Cell Invasion Assay Kit (Chemicon), a similar chemotaxis chamber as above but with a few modifications. In brief, LoVo-AKAP12 and LoVo-CON cells were plated at a density of 10^5 cells/mL on ECMatrix gel-coated polycarbonate membrane inserts with an 8 μm pore size and cultured for 24 h. Invasive cells migrate through the ECM layer and cling to the bottom of the polycarbonate membrane. The non-invading cells and the ECMatrix gel were removed from the upper surface of the insert with a cotton-tipped swab. Add staining solution to the unoccupied wells of the plate and stain invasive cells on lower surface of the membrane by dipping inserts in the staining solution. Dip inserts in a beaker of water several times to rinse. Count cells by photographing the membrane through the microscope and five fields were evaluated for invading cells. The experiment was repeated three times.

Primary xenografts

To evaluate in vivo tumorigenesis, a colorectal cancer xenografting mouse model was used. Female BALB/c nude mice of four to six weeks old were prepared for tumor implantation. Mice were allowed to acclimate for one week after arrival. All animals were maintained in a sterile environment on a daily 12-h light/dark cycle. After resuspension in PBS, LoVo-AKAP12, LoVo-CON cells and LoVo-LIPO (transfected with Lipofectamine only), 5×10^6 cells/mouse were injected subcutaneously into the flank of nude mice (n = 10/group). Tumor volume was calculated weekly for four weeks according to the formula, TV (cm^3) = length × width2 × 0.5. The primary xenografts and lungs were harvested, weighed, and snap-frozen[27]. All of the animal experiments were conducted in strict accordance with the National Institute of Health Guide for the Care and Use of Laboratory Animals.

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Metastasis assay in vivo

The detection of the presence of human tumor cells in the lungs of mice was achieved using the quantitative detection of human Alu sequences present in total lung genomic DNA preparations[28]. Whole lungs were harvested and genomic DNA was isolated using the DNeasy Tissue kit (Qiagen). Lung genomic DNA was quantified in human tumor cells from the lungs using a PCR-based detection of the human Alu sequences and, as a control, GAPDH, and the following primers: Alu-F, 5'-CACCTGTAATCCAGGACCTTTT-3'; Alu-R, 5'-CCCAAGCAGTGGACTGC-3'; GAPDH-F, 5'-GACAGATGGGCCGGAAT-3' and GAPDH-R, 5'-GCCCTTCTCCATTGTTGTTGAA-3'. PCR was performed under the following conditions: 95°C for 2 min, 30 cycles at 95°C for 30 s, 65°C for 20 s, and 72°C for 20 s. The SYBR Green PCR Master Mix (Qiagen) was used for the real-time PCR amplification of the Alu sequences. A quantitative measure of amplifiable mouse DNA was obtained through amplification of the mouse GAPDH genomic DNA sequence. The fluorescence emitted by the reporter dye was detected using the SYBR Green and the threshold cycle (Ct) for each sample was recorded as a quantitative measure of the amount of product in each sample. When indicated, the Alu signal was normalized against the relative quantity of GAPDH and expressed as ΔCt = (CtAlu–CtGAPDH). The changes in Alu signal relative to the total amount of genomic DNA were expressed as 2−ΔΔCt, indicating the changes in the quantity of human DNA in the mouse lung tissue. Data are shown as relative to the expression of Alu and levels from the LoVo-LIPO group were normalized to one. The specificity of human Alu PCR was verified by the lack of amplification from 100% mouse DNA.

Statistical analysis

The Student’s t-test was used to compare the obtained values with those from the corresponding control experiments and p-values < .05 was considered statistically significant.

Ethical Treatment of Animals

This study was carried out in strict accordance with the recommendations in the relevant national and international guidelines for the Care and Use of Laboratory Animals. The protocol was approved by the Committee on the Ethics of Animal Experiments of the Department of Laboratory Animal Science, Shanghai Jiao Tong University School Of Medicine (Permit Number: 2009014). All efforts were made to minimize suffering.

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Author Contributions

Conceived and designed the experiments: WL, MG. Performed the experiments: WL, MG TH XG. Analyzed the data: WL, MG TH XG. Contributed reagents/materials/analysis tools: WL, MG TH XG. Wrote the paper: WL, MG YL.

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