Comprehensive role of SARS-CoV-2 spike glycoprotein in regulating host signaling pathway

Shuvomoy Banerjee1 | Xinyu Wang2 | Shujuan Du2 | Caixia Zhu2 | Yuping Jia3 | Yuyan Wang2 | Qiliang Cai2

1Department of Biotechnology and Bioengineering, Koba Institutional Area, Gandhinagar, Gujarat, India
2MOE&NHC&CAMS Key Laboratory of Medical Molecular Virology, Shanghai Institute of Infections Disease and Biosecurity, & School of Basic Medical Science, Shanghai Medical College, Fudan University, Shanghai, China
3Shandong Academy of Pharmaceutical Sciences, Jinan, China

Correspondence
Shuvomoy Banerjee, Department of Biotechnology and Bioengineering, Institute of Advanced Research, Koba institutional area, Gandhinagar, Gujarat 382426, India.
Email: shuvomoy.banerjee@iar.ac.in
Yuping Jia, Shandong Academy of Pharmaceutical Sciences, Jinan 250100, China.
Email: jiayupingygs@163.com
Yuyan Wang and Qiliang Cai, MOE&NHC&CAMS Key Laboratory of Medical Molecular Virology, Shanghai Institute of Infections Disease and Biosecurity, & School of Basic Medical Science, Shanghai Medical College, Fudan University, Shanghai 200032, China.
Email: yuyanss@fudan.edu.cn and qiliang@fudan.edu.cn

Funding Information
National Key Research and Development Program of China

Abstract
Since the outbreak of coronavirus disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection, global public health and the economy have suffered unprecedented damage. Based on the increasing related literature, the characteristics and pathogenic mechanisms of the virus, and epidemiological and clinical features of the disease are being rapidly discovered. The spike glycoprotein (S protein), as a key antigen of SARS-CoV-2 for developing vaccines, antibodies, and drug targets, has been shown to play an important role in viral entry, tissue tropism, and pathogenesis. In this review, we summarize the molecular mechanisms of interaction between S protein and host factors, especially receptor-mediated viral modulation of host signaling pathways, and highlight the progression of potential therapeutic targets, prophylactic and therapeutic agents for prevention and treatment of SARS-CoV-2 infection.

KEYWORDS
ACE2 receptor, S protein, SARS-CoV-2, signaling pathway

1 | INTRODUCTION

Acute respiratory diseases (ARD) are considered as the major cause of all acute morbidities and mortalities worldwide.1 Among these, acute viral respiratory tract infection, including the influenza virus, respiratory syncytial virus, coronavirus, adenovirus, and rhinovirus, is the leading cause of ARD. In the case of adenovirus and rhinovirus, they are associated with lower mortality but significant morbidity, and cause vast health and economic burdens.2 The observation of the mode of virus transmission from a natural vertebrate animal host to the human host, leading to cause of “zoonotic infection” spread in the human being, could be the case with COVID-19 (The Coronavirus Disease 2019) (Figure 1A).4 The zoonotic virus spillover or transfer to human beings can be attributed to various factors, including global warming, climate changing, and loss of biodiversity due to deforestation, poaching, decreased habitat quality, effects of expanded global food production on natural environments.5 Therefore, emerging infectious diseases have an intense impact in terms of the global burden, threat, and

Shuvomoy Banerjee and Xinyu Wang contributed equally to this study.
FIGURE 1  Evolutionary origin and transmission routes of SARS-CoV-2. (A) The evolutionary origin and journey of SARS-CoV-2 through different primary and intermediate hosts. Different paths of potential viral transmissions are indicated with yellow arrows. (B) The circular phylogenetic tree illustrates the positions of human coronavirus HKU1 (HCoV-HKU1), human coronavirus OC43 (HCoV-OC43), Middle East respiratory syndrome coronavirus (MERS-CoV), Severe acute respiratory syndrome (SARS)-associated coronavirus (SARS-CoV and SARS-CoV-2) in different host groups. The complete genome sequences were collected from National Center for Biotechnology Information (NCBI) database and analyzed in The Interactive Tree Of Life (https://itol.embl.de) online software tool. A separate phylogenetic tree of HCoV-HKU1, HCoV-OC43, MERS-CoV, SARS-CoV, and SARS-CoV-2 is shown on the right panel.

1.1 | SARS-CoV-2: A glimpse of evolutionary perspectives

Coronavirus is enveloped single-stranded positive-sense RNA virus, and belongs to the family of Coronaviridae, subfamily Coronavirinae. As shown in Figure 1B, based on genotype and serological tests, CoVs are classified into four genera assigned as alpha, beta, gamma, and delta coronavirus. Seven viruses are currently identified to infect human beings within the Coronaviridae family, including NL63 and 229E from the genus alpha, OC43, HKU1, SARS-CoV, MERS-CoV, and SARS-CoV-2 from the genus beta. The SARS-CoV-2 is clustered with SARS-CoVs in the species tree of severe acute respiratory syndrome-related coronavirus in the Sarbecovirus of genus beta-coronavirus.

In the past few decades, the appearance of MERS-CoV, SARS-CoV, and SARS-CoV-2 with epidemic capabilities has become the third coronavirus. Of these, SARS-CoV-2 causes a lower fatality rate than the other two, but with a higher transmission rate
results in a pandemic shape of COVID-19. So far, analysis by various bioinformatics tools has provided a deep insight into the genome of COVID-19 causative pathogen. The SARS-CoV-2 genome analysis has revealed high similarity with bat coronavirus genome, and a highly similar receptor-binding domain (RBD) of the spike protein as Malayan pangolin coronavirus. The genetic proximity indicates that SARS-CoV-2 is likely to have originated from bat-derived CoV, as a consequence of the accumulation of mutations and the acquisition of new genomic regions by recombination events between bat and pangolin coronaviruses (Figure 1A). However, the exact mechanisms involved with the evolution of this coronavirus as a human pathogen are not fully understood yet. The missing links in the evolution of SARS-CoV-2 include the exact immediate reservoir and all other possible hosts. In addition, the spillover at the animal–human interface leading to new human infection is influenced by several factors, such as pathogen pressure human and reservoir host behavior, and the factors linked with human infection susceptibility. In these human-dominated landscapes, primates, and bats are reservoir hosts of more viruses than other mammal species, which increases the possibility of new infections in humans. For instance, the potential risk of SARS-CoV spillover from horseshoe bat population to humans was demonstrated by Menachery et al., using a reverse genetics system entailing a chimeric virus expressing the bat SHCO14 in a mouse-adapted backbone. This study illustrates scenarios for the emergence of bat SARS-CoV in humans, probably infection of an intermediate nonhuman host might be followed by human infection. Direct bat–human transmission could be followed by selection in the human population. In another scenario, the circulation of quasi-species pools in the animal reservoirs (camel or civet cat) might maintain multiple virus strains, some of which capable of causing infection in humans without the need for additional mutations. Both alpha-coronaviruses and beta-coronaviruses are identified in free-ranging bats from Myanmar, indicating that the potential for zoonotic virus emergence in humans in close contact with sylvatic animals in forest areas are disturbed by an ongoing process of changes in land use.

1.2 Viral entry in cells

SARS-CoV-2 infects human cells via S protein binding to the receptor ACE2 on the host cell that allows the virus entry into the cell. As shown in Figure 2A, the entry of coronaviruses into target cells depends on the binding of the S1 subunit of S protein to specific cellular receptor ACE2. The viral protein–receptor binding helps viral attachment to the cell surface. In addition, viral entry requires an important step known as “protein priming” by cellular proteases (TMPRSS2), which involves the cleavage of S protein at the S1/S2 and the S2′ sites enabling the fusion of viral and cellular membranes. Although the binding of viral S protein and ACE2 receptors has been explained in detail, and ACE2 is considered as the prime determinant of SARS-CoV transmissibility, it is still not clear whether SARS-CoV-2-S follows the same mechanisms as SARS-CoV-S which employs ACE2 and TMPRSS2 for cellular entry. In addition to depending on ACE2 for host cell entry, both SARS-CoV and SARS-CoV-2 depend on entry activation by host cell proteases at the S1/S2 and S2′ sites, regardless of whether entry occurs by fusion or endocytosis. In contrast to SARS-CoV, SARS-CoV-2 also has a furin cleavage site that can be recognized by neuropilin-1 for infectivity. Like other trimeric class I fusion proteins, S proteins from SARS-CoV, SARS-CoV-2, and MERS-CoV undergo dramatic structural changes to fuse membranes, which occurs after proteolytic activation at the S1/S2 boundary. S1 dissociates, and conformational changes allow the fusion peptide (FP), which is hydrophobic residues-rich and inserted into the host cell membrane, S2 forms an elongated structure, and the two heptad repeats, HR1 and HR2, eventually form a six-helix bundle to complete the fusion process and deliver the viral genome into the cytoplasm.

1.3 Cell and tissue tropism of SARS-CoV-2

In general, viral tropism is considered the ability of a given virus to effectively infect a particular cell, tissue, or host species. The degree and establishment of viral tropism mainly depend on the susceptibility and permissiveness of a specific host cell. Host cellular receptors play key roles in the determination of virus tropism and pathogenesis. The widespread distribution and expression of ACE2 across multiple organs are critical to helping understand the varied clinical outcomes of COVID-19. COVID-19 patients often showed respiratory-like illnesses leading to severe pneumonia, suggesting that the SARS-CoV-2 mainly targets the lung as the primary site of tropism. Many studies have also demonstrated that important mutations within the RBD of SARS-CoV-2 spike protein make additional adjacent contact points with ACE2, establishing the viral characteristics of higher binding affinity and increased infectivity. In the context of cellular tropism of SARS-CoV-2, researchers identified that airway and alveolar epithelial cells, vascular endothelial cells, and alveolar macrophages are mainly targeted by the virus during the respiratory tract infection. The cells in the respiratory tract are suitable for establishing SARS-CoV-2 early infection and subsequent replication due to their expression of ACE2. Moreover, it has been assumed that the respiratory cells in the lung may be contingent on the regulation of ACE2 expressions at the transcriptional and protein levels, favoring SARS-CoV-2 entry, as several studies showed that SARS-CoV-2 can proficiently infect the nasopharyngeal and oropharyngeal tissues, possibly due to its higher affinity for ACE2 which is also expressed in those tissues. SARS-CoV-2 internalizes inside the cells via ACE2, the downregulation of ACE2 expression can lead to altered tissue function and exacerbate chronic diseases. ACE2 has also been identified to be critical for maintaining tissue homeostasis, as it negatively regulates the renin–angiotensin–aldosterone system (RAAS), which is extremely important for regulating normal functions in different organs including, the lungs, heart, kidney,
and vasculatures. In addition, several studies have shown the involvement of ACE2 in ARDS in the animal model, which may be triggered by multiple diseases including the infection of SARS-CoV and SARS-CoV-2. In particular, ACE2 knockout mice showed severe onset of ARDS and could be rescued from ARDS condition if ACE2 knockout mice are treated with AR1R blocker, implicating the vital role of ACE2 and the critical balance between protective and proinflammatory immune reaction influenced by RAAS.

Despite the respiratory route is dominant for SARS-CoV-2 infection, the highest levels of ACE2 expression are found in the small intestine, testis, kidney, heart muscle, colon, and thyroid gland (Figure 2B). The phenomenon of ACE2 and TMPRSS2 abundant expression in the gastrointestinal tract is consistent with the observation that many other coronaviruses are transmitted via the fecal–oral route and respiratory route, and may be maintained for a long time in the intestinal tissue (Figure 2C). Supporting this speculation, gastrointestinal disorders are frequently found in

FIGURE 2 Regulation of host cell receptors by SARS-CoV-2-Spike protein. (A) Sequential events of viral–host cell membrane fusion were diagrammatically represented involving SARS-CoV-2 spike protein, ACE2, and TMPRSS2 receptors. (B) The expression patterns of human ACE2 and TMPRSS2 receptors in normal tissues were shown by heatmap. The expression patterns were analyzed by the Open Access FunRich functional enrichment analysis tool (http://www.funrich.org/index.html). (C) Cell and tissue tropism of SARS-CoV-2. After successful infection, the virus disseminates in different organ tissues of the human body including the lung, pancreas, liver, brain, intestine, heart, and skin.
patients with COVID-19, and nearly 20% of patients are identified with detectable SARS-CoV-2 mRNA in fecal samples.\textsuperscript{51} Cardiac infection by SARS-CoV-2 is frequently found in autopsy cases, and the presence of ACE2 in colon and kidney cells has been suggested as an explanation for gastrointestinal and renal complications of SARS-CoV-2 infection.\textsuperscript{52} Results from organoid models showed that the SARS-CoV-2 can directly attack the liver tissue and cause liver damage, and can also destroy cells that control blood sugar in pancreatic organoids, which adds to mounting evidence that the virus can trigger diabetes in some people. In addition, like other coronaviruses, SARS-CoV-2 can enter the nervous system by crossing the neural–mucosal interface in olfactory mucosa, exploiting the close vicinity of olfactory mucosal, endothelial, and nervous tissue, including delicate olfactory and sensory nerve endings.\textsuperscript{53} The tissue expression pattern of coronavirus receptors (ACE2, BSG, NRP1) and proteases (TMPRSS2, TMPRSS11A, TMPRSS11B, and Furin) in excitatory and inhibitory neurons, astrocytes, oligodendrocytes, and microglia, as well as in cranial vascular endothelial cells, is an indicator for SARS-CoV-2’s vulnerability for these cells.\textsuperscript{54} RNA-seq profiling of the expression pattern of ACE2 in adult human testis indicated that ACE2 is predominantly enriched in spermatogonia and Leydig and Sertoli cells.\textsuperscript{55} SARS-CoV-2 infection might damage the male reproductive system via ACE2 receptor, which is required to identify potential short-term and long-term effects of SARS-CoV-2 on male fertility.\textsuperscript{56}

Interestingly, low expression of ACE2 mRNA was detected in human and other mammalian lung tissues (bat, ferret, cat, dog, etc.) compared with extrapulmonary tissues.\textsuperscript{57} ACE2 alone cannot explain the multiorgan tropism of SARS-CoV-2 nor the clinical differences between SARS-CoV-2 and SARS-CoV, suggesting the involvement of other receptors(s) or additional cell-intrinsic factors.\textsuperscript{58} Several studies have demonstrated that TMPRSS2 (transmembrane serine protease-2) plays an important role in viral entry, as the nearly untraceable level of ACE2 in lung tissue still provides support for SARS-CoV entry as long as TMPRSS2 is present.\textsuperscript{59} Gu et al. identified that ASGR1 and KREMEN1, as alternative functional receptors, play essential roles in ACE2-independent virus entry, providing other insight into SARS-CoV-2 tropism and pathogenesis.\textsuperscript{60} In addition, higher mRNA expression of several cellular genes, including gene members of endosomal sorting complex required for transport machinery (i.e., CHMP3, CHMP5, CHMP1A, and VPS37B which are important for the early SARS-CoV-2 lifecycle), are identified in human type II alveolar cells with abundant ACE2, compared to ACE2-deficient cells.\textsuperscript{61} Such experimental evidence indicates the way by which SARS-CoV-2 employs to takeover a small population of type II alveolar cells with high expression of ACE2, as well as regulates other pro-viral genes for its successful replication inside the host cells.

### 1.4 | SARS-CoV-2 spike protein: Structure–function variability

SARS-CoV-2 S protein has two functional subunits (S1 and S2). As shown in Figure 3, the S1 subunit contains the N-terminal domain (NTD) and RBD, which can bind with the cellular receptor. Compared to S1, the S2 subunit contains a bit enriched structure, including FP, heptad repeat-1 (HR1), central helix (CH), connector domain (CD), heptad repeat-2 (HR2), transmembrane domain (TM), and cytoplasmic tail (CT).\textsuperscript{62} The furin cleavage site at the border between the S1 and S2 subunits is called the “S1/S2 protease cleavage site.” Importantly, for all the CoVs, host proteases cut the spike glycoprotein at the S2’ cleavage site to activate proteins for membrane fusion through the irreversible conformational changes.\textsuperscript{63} The mutation rate analysis of the S protein domain shows that S1/S2 protease cleavage site is associated with the maximum mutation density, which is much higher than that of the RBD region directly binding to the host receptor.\textsuperscript{64} This suggests that mutations at this site in the S protein may be of advantage for the virus to undergo proteolytic cleavage by a large number of host enzymes during evolution.\textsuperscript{65} S protein undergoes significant structural rearrangement important for ACE2 receptor binding and subsequent entry of virus by membrane fusion.\textsuperscript{66} At the prefusion conformational stage, S1 and S2 subunits bind non-covalently. However, some reports have shown that coronaviruses use “special domains” in the S1 subunit to identify different cellular receptors for viral entry. In particular, SARS-CoV and SARS-CoV-2 recognize the ACE2 via the RBD.\textsuperscript{67} Structural data reveals that the S protein has two forms of structural conformations such as the “closed” and “open” state. In the open state conformation, the RBD remains in

![Image](image-url)  
**Figure 3** Structural overview of SARS-CoV-2 Spike protein. Diagrammatic representation of SARS-CoV-2 genome, and spike glycoprotein (S) structures. Illustration of full-length SARS-CoV-2 S-protein domains (N-terminal, receptor binding domain, C-terminal), furin cleavage site, fusion peptide, and Heptad repeat regions (HR1 and HR2) are highlighted.
the “up” position. While in the closed state conformation, the three recognition motifs do not project from the interface. The open state conformation is extremely important for the fusion of the virus with the host cell membranes, leading to SARS-CoV-2 entry in the host cells. Importantly, both SARS-CoV-2 S and SARS-CoV S proteins converge on an ACE2 epitope (17-residue long) with the receptor-binding motif (RBM). Studies have also revealed the finer molecular architecture of RBM itself is made up of a concave surface stabilized by two β-hairpins. The RBD of S protein is responsible for further stabilization of the concave structure. On the other hand, the ACE2-long N-terminal helix embraced on top of the RBM, and the ACE2 residues K31 and K353 are critical for interacting with SARS-CoV S protein. In contrast, SARS-CoV-2 S protein residues including Y449, Q493, Q498, and N501 have been identified to be of immense importance for ACE2 binding. Upon mutations of these residues, SARS-CoV-2 exhibits altered affinity towards S-ACE2 complex formation. Therefore, SARS-CoV-2 Spike protein shows conformational dynamics and its RBD has been observed as very important for virus-host membrane fusion and cellular entry. Interestingly, S protein can only bind with ACE2 when the RBD remains in the upstate conformation, while heparan sulfate interaction with the RBD maintains the open state conformation for ACE2 binding, and the closed state conformation is ideally observed at endosomal pH 5.5. Although different structures of the SARS-CoV-2 S-ACE2 complex have been identified, both SARS-CoV-2 and SARS-CoV bound with the ACE2 receptor are similar.

SARS-CoV-2 S protein is highly glycosylated, each protomer in the trimeric spike has 22 N-linked glycosylation sites. N-linked glycans are crucial for proper protein folding, making neutralizing antibodies, and arranging the spike protein trimers extensively. Glycosylation modification of S protein will create a glycan shield which complicates antibody neutralization. However, some of the spike glycans may directly regulate the interaction between virus and receptor. For example, elegant molecular dynamics simulations show that N-glycans at sites N165 and N234 of S protein in modulating the conformational dynamics of the spike’s RBD, which is responsible for ACE2 recognition, while glycans at N90, N322, and N546 of ACE2 directly mediate the interaction with S trimer. Aptamer studies also show that glycosylation of the RBD protein has significant effects on values of dissociation constants and the relative efficacy of the aptamer binding. Thus, variations in glycan occupancy or processing at these sites could alter the affinity of the SARS-CoV-2–ACE2 interaction and modulate infectivity. Furthermore, S protein can interact with SARS-CoV-2 attachment receptor type C lectin through a high-mannose N-glycan structure, and this interaction can be blocked by glycosylation inhibitors. In addition, an in silico study shows that the S1 subunit of S protein has a high affinity with toll-like receptor TLR-4, which may also be caused by the complex glycoconjugate structure of S protein. Therefore, glycosylation modification is diverse, which is varied by tissue cell type, host race, and age, and each glycosylation site can be modified with one of several glycan structures to produce site-specific glycosylation combinations. Glycosylation of S protein will increase the heterogeneity of its structure, affect the binding of SARS-CoV-2 to its receptor, and the individual susceptibility. Interestingly, studies have shown that D614G mutation may increase glycosylation of neighboring asparagine 616, and glycosylation mutant N331 and N343 will significantly reduce SARS-CoV-2 infectivity, indicating that the mutation of the S protein will also affect its glycosylation level.

### 1.5 | SARS-CoV-2 spike protein-mediated signaling: A molecular-level analysis

To be more efficient for spreading infections in the respiratory tract, it has been reported that SARS-CoV-2 often uses different attachment factors presented on the host cell surface (Figure 4). Emerging evidence from in vitro and in silico experiments indicates that there are a series of molecular interactions between SARS-CoV-2 S protein and receptors associated with immune functions, including neuropilin-1 (NRP1), C-lectin type receptors (CLR)-mannose receptor (MR), dendritic cell-specific intracellular adhesion molecule-3-grabbing non-integrin receptor (DC-SIGN), homolog dendritic cell-specific intercellular adhesion molecule-3-grabbing nonintegrin related receptor (L-SIGN), and macrophage galactose-type lectin (MGL) and toll-like receptors including TLR1, TLR4, and TLR6. Researchers have also identified the association of nonimmune receptor glucose-regulated protein 78 (GRP78) with SARS-CoV-2 S protein. Likewise, TIM1 and AXL as members of phosphatidylserine receptor families were also suggested to be alternative SARS-CoV-2 receptors, respectively. Such interactions greatly improve their adhesion as well as the virus’s concentration to the cell surface. S protein-other attachment factors association was found very crucial for potentiating access and final engagement with their prime entry receptors. Such molecular interactions not only augment SARS-CoV-2 infection of target cells but also potentiate the viral entry in nonpermissive cells by the process of “trans infection.” Importantly, these host factors have their unique functions in cells. The interaction between S protein and host factors often triggers the expression changes of downstream signal pathways, resulting in a series of pathological reactions.

### 1.6 | Spike protein mediates hypertension and inflammatory factor storm via interaction with ACE2 receptor

ACE2, as the most important receptor for SARS-CoV-2 to enter cells, is also the master regulator of the local and central renin–angiotensin system (RAS). ACE2 is widely expressed in lung, kidney, testis, intestine, adipose tissue, and brain, and catalyzes the conversion of angiotensin I to angiotensin-(1–9) and angiotensin II (Ang II) to angiotensin-(1–7). To serve its regulatory role in the RAS, membrane-bound ACE2 (mACE2) is the first required to be transported to the cell surface, where it is cleaved by host proteases, to release an enzymatically active soluble...
FIGURE 4  The SARS-CoV-2-Spike protein mediates signaling transduction pathways. The diagram shows SARS-CoV-2-Spike glycoprotein (S) interacts with several host cell receptor proteins mainly with ACE2 by its RBD region. TMPRSS2 initiates the activation of S protein for viral–host cell membrane fusion and subsequent internalization of viral particles inside the cell. SARS-CoV-2-S and ACE2 interaction elicit NF-κB signaling to regulate different cellular functions. S protein also shows the interaction with cytokine receptors (TLR protein (GRP78), mineralocorticoid receptor (MR), and important growth factor receptor neuropilin-1 (NRP1). Apart from these, SARS-CoV-2-Spike glycoprotein was shown to interact with dendritic cell-specific intercellular adhesion molecule-3-grabbing non-integrin (DC-SIGN) for regulating different immunological functions in the host cell.

form of ACE2 (sACE2) into the plasma. As sACE2 preserves the binding site for SARS-CoV-2, sequestration of SARS-CoV-2 by sACE2 may enable cell entry of tissues where cACE2 is poorly expressed.\(^{77}\) Research shows that SARS-CoV-2 infection may lead to ACE2 shedding and the concentrations of sACE2 may correlate with the level of systemic inflammation that occurs.\(^{98}\) The binding of SARS-CoV-2 S protein to ACE2 will reduce the number of membrane-bound ACE2 and affect its normal physiological function, resulting in the imbalance of the ACE2/ACE ratio.\(^{79}\) Consequently, elevated ACE/Ang II/AT1R axis activity or decreased ACE2/Ang-(1–7)/Masr axis activity can lead to the pulmonary artery and neurogenic hypertension, which may lead to thrombosis, cardiovascular complications, and ultimately lung injury, heart failure, multiple organ failure, and bleeding.\(^{100,101}\) which supporting clinical evidence that RAS imbalance is associated with the development of multiple organ damage during SARS-CoV-2 infection.\(^{102}\)

The interaction between SARS-CoV-2 spike protein and ACE2 receptor is also related to the excessive inflammatory response. On the one hand, the spike protein promoted an angiotensin II type 1 receptor (AT1) mediated signaling cascade, induced the transcriptional regulatory molecules NF-κB and AP-1/c-Fos via MAPK activation, and increased IL-6 release.\(^{103}\) On the other hand, the interaction of the ACE2 receptor with the SARS-CoV-2 spike protein directly activates the Nlrp3 inflammasome in human VSELs and HSCs, which, if hyperactivated, may lead to cell death by pyroptosis.\(^{104}\) Meanwhile, once SARS-CoV-2 spike protein interacts with ACE2, macrophages/microglia cells can potentiate the immune response through the cleavage of fragments complement component 3a and 5a (C3a and C5a, respectively) and non-lytic C5b-C9 membrane attack complex by ComC, thus activating the NLRP3 inflammasome.\(^{105}\) Barreda et al. found the expression of ACE2 during the differentiation of human monocytes to mature DCs and interdonor is different. Both SARS-CoV-2 S protein and its RBD fragments could promote an inflammatory response in dendritic cells, and increase the expression of maturation markers (including MHC molecules and costimulatory receptors, and active signaling molecules of MAPK, AKT, STAT1, and NF-κB), which correlates with the expression and secretion of distinctive proinflammatory cytokines.\(^{106}\) Therefore, upon S protein binding, DC maturation is expected to be a key step to induce long-lasting immunity against SARS-CoV-2. In addition, the study of SARS-CoV-2 infection in the nervous system shows that virus infection causes the release of cellular ATP, and extracellular ATP can robustly trigger activation of the NLRP3 inflammasome through P2X7 receptor activation.\(^{107}\) With NLRP3 inflammasome activation, ATP is also released into the extracellular milieu, which promotes a strong positive feedback loop.\(^{108}\) In this scenario, proinflammatory factors are intensely released, the so-called “cytokine storm,” and enter the bloodstream and reach other organs, causing extensive complications.\(^{109}\)

1.7 | Spike protein mediates inflammatory response via interaction with immune function related receptors

TLR can identify a repertoire of pathogen-associated molecular patterns (PAMP),\(^{110}\) and induce a robust inflammatory response
through the myeloid differentiation factor-88 (MyD88)-dependent pathway, or the Toll/IL-1-domain-containing adapter-inducing interferon-beta (TRIF)-dependent pathway. Molecular docking studies have demonstrated that SARS-CoV-2 S protein can directly bind to TLR1, TLR4, and TLR6. Biochemical studies revealed that SARS-CoV-2 S protein triggers inflammation via activation of the NF-κB pathway in a MyD88-dependent manner. Khan et al. demonstrated that SARS-CoV-2 S protein is a potent viral PAMP that sense dimers of TLR2/TLR1 or TLR2/TLR6 to activate the NF-κB pathway, leading to the expression of inflammatory mediators in innate immune and epithelial cells. Recent studies have also shown that the S1 subunit of SARS-CoV-2 S protein may play a role as a pathogen-related molecular model (PAMP) to induce a neuroinflammatory process independent of virus infection. For example, S1 treatment could increase gene expression of microglia/brain macrophage activation markers (Iba1, Cd11b, MHC-II), astrocyte activation markers (Gfap), TLRs (Tlr4), inflammasomes (Nlpr3), and proinflammatory cytokines (IL-1β); and S1 could activate expression of TLR2 and TLR4 in HEK293 cells and TLR2 and TLR4 in microglia. Due to microglia do not express ACE2, this indicates that S1 may signal through these TLRs to produce a neuroinflammatory response in microglia. Following this hypothesis, Shirato et al. demonstrated that the SARS-CoV-2 spike protein S1 subunit activates TLR4 signaling to induce proinflammatory responses in murine and human macrophages. Utilizing the microglia cell line BV-2, Olajide et al. reported that the S1 subunit of the S protein activates NF-κB and p38 MAPK signaling pathways and in turn induces TNF-α, IL-6, IL-1β, and iNOS/NO production by targeting uses TLR4. Likewise, the S protein binds to TLR4 with high affinity and induces a proinflammatory response in THP-1 and RAW 264.7 cells, which is blocked by a TLR4 inhibitor.

In addition, several studies have also shown that DC-SIGN and L-SIGN promote SARS-CoV-2 transfer to permissive Vero E6 and Calu-3 cells with ACE2 expression. Apart from the sole contribution of Spike protein for inducing DC/L-SIGN expression, it has been demonstrated that other cellular factors are equally important for their functional activation. For example, researchers observed that proinflammatory cytokines, including IL-4, IL-6, IL-10, and IL-13 (which are known to be overexpressed in severe COVID-19 cases), may activate DC/L-SIGN. It is, therefore, speculated that DC-SIGN binding to SARS-CoV-2 and increased IL-10 levels in severe COVID-19 patients may serve as an immune-inhibitory mechanism stimulated by the rapid accumulation of inflammatory cytokines, via a negative feedback loop. While on the other hand, the significant increase of IL-10 will stimulate the production of other cytokine storm mediators and potentially lead to hyperactivation of the immune system exacerbating COVID-19 severity.

In addition, mannose-binding lectin (MBL) has been reported to associate with SARS-CoV-2 infection severity. The interaction of MBL with SARS-CoV-2 spike protein requires a trimeric conformation of the viral protein, which does not involve direct recognition of the RBD but is glycan dependent. Unlike other receptors, it has been shown that the interaction between MBL and S protein may prevent SARS-CoV-2 from entering cells and activate the complement pathway.

1.8 Spike protein mediates the signaling pathway via other cofactors

Despite host cellular receptors playing key roles in the determination of virus tropism and pathogenesis, however, ACE2 alone cannot explain the multiorgan tropism of SARS-CoV-2 nor the clinical differences between SARS-CoV-2 and SARS-CoV, suggesting the involvement of other receptors (Figure 4). In addition to the above receptors, SARS-CoV-2 spike protein has been shown to interact with glucose-regulated protein 78 (GRP78), cluster of differentiation 147 (CD147), Neutrophil-1 (NRP1), and other molecules. These molecules are not yet clear in SARS’s entry into cells and pathogenesis, so they are classified as cofactors. These molecules are not yet clear in the role of SARS-CoV-2 entry into cells and pathogenesis, so they are classified as cofactors. GRP78 is a nonimmune receptor and essential endoplasmic reticulum chaperone protein, which has been identified as a DAMP for specific TLRs. In silico study has demonstrated that it is preferred potential binding of RBD regions III and IV of S glycoprotein to substrate-binding domain-beta of GRP78. Therefore, GRP78 can cooperate with TLRs to enhance the inflammatory reaction caused by SARS infection. In another case, NRP1 is a transmembrane polypeptide, which acts as a coreceptor for different growth factors, and regulating angiogenesis, gangliogenesis, and vascular permeability. Recent reports have demonstrated that the SARS-CoV-2 spike protein can bind to the b1b2 domain of the NRP1 and thereby blocking VEGF-A/NRP-1 signal transduction and affecting pain behavior. Given the facts that KREMEN1 as an alternative functional receptor of SARS-CoV-2, and KREMEN1/2 plus FUT8 are all negative regulators of the Wnt/β-catenin signaling pathway which is critical in taste bud cell renewal and behavioral taste perception, as well as loss of smell and taste has frequently been observed in COVID-19 patients, suggesting that SARS-CoV-2 may act through these factors to affect Wnt/β-catenin signaling and in turn, promote taste loss.

In addition, some studies have also shown that SARS-CoV-2 spike protein may mediate cellular signals with an unknown receptor. For example, Suzuki et al. found that the S protein region without the RBD structure of SARS-CoV-2 alone can induce the cellular signals of human pulmonary vascular smooth muscle and endothelial cells, especially the activation of the MEK/ERK pathway. Given the MEK/ERK pathway is a well-known signaling pathway to facilitate viral replication, SARS-CoV-2 spike protein-mediated signaling could promote the hyperplasia and/or hypertrophy of vascular smooth muscle and endothelial cells, which contributes to the complex cardiovascular outcomes in COVID-19.
1.9 Therapeutic strategies to target SARS-CoV-2 spike protein

During the COVID-19 pandemic, the main approach employed by physicians to treat COVID-19 patients is to control the COVID-pneumonia and inhibit the deadly cytokine storm. Therefore, the therapeutic approach for completely curing and eradicating this viral disease is a major challenge to scientists worldwide. Fortunately, researchers have learned from other RNA virus, including human immunodeficiency virus or Ebola virus, SARS-CoV-2 infection may be controlled by using the same strategies which previously applied for curing other RNA virus-associated infections. Several clinical trials have been recently undergoing combination of antiretroviral drugs along with remdesivir. Other repurposed drugs are also applied in COVID-19 treatment including chloroquine and Tocilizumab, although there are controversies for their significant side effects on COVID-19 patients. Researchers also realized that besides newly synthesized or discovered antiviral drugs, alternative approaches, including monoclonal antibodies and repurposing drugs, should be taken with the rapid COVID-19 spread in the global population. Regardless of tremendous efforts by scientific communities, no specific antiviral drugs or vaccines have been discovered to combat this deadly virus, albeit intensive research continues to seek efficient therapeutic interventions for fighting against the COVID-19 pandemic. We will summarize some important therapeutic strategies from the perspective of S protein and its related receptors against SARS-CoV-2 infection below.

1.10 Vaccines and antibodies targeting SARS-CoV-2 spike protein

Many researchers have identified the importance of SARS-CoV-2 spike protein as the primary antigenic target for developing potential antiviral therapeutics and vaccine development. So far, there are three major types of SARS-CoV-2 vaccines based on the S protein: (1) recombinant protein vaccines that express S proteins in vitro and are used for immunization, including full-length S protein (FLSP) recombinant vaccines, RBD recombinant vaccines, and virus-like particle vaccines that carry S proteins on their surfaces; (2) recombinant vector vaccines that use viral vectors to carry S protein genes, including replication-incompetent and replication-competent vector vaccines; (3) nucleic acid vaccines that introduce the S protein gene into vaccinated individual, including DNA and RNA vaccines. Recombinant protein vaccines are currently the most developed, accounting for 34% of vaccines entering the clinical phase, and studies have shown that the FLSP, RBD domain, S1 subunit, S2 subunit, NTD and FP of S proteins can all be potential components of antigens. Recombinant vector vaccine technology is relatively mature and a wide range of vectors are available, including modified adenovirus, Ankara poxvirus, human parainfluenza virus, influenza virus, and Sendai virus. Most advanced candidate is ChAdOx1 nCoV-19 (AZD1222), developed by AstraZeneca and the University of Oxford, which induces a balanced cellular and humoral immune response, and has been approved by EUA in England. RNA vaccines also present greater promising future, with features of prompt modification of the encoded immunogen and quick manufacturing process. Moderna (mRNA-1273) or Pfizer-BioNTech (BNT162b2) vaccines in Phase III clinical trials, elicited monoclonal antibodies (mAbs) potently neutralize SARS-CoV-2. However, the development of S protein-based vaccines is still facing many dilemmas. First, full-length recombinant S proteins are difficult to express and have poor stability, while recombinant RBD vaccines recognize fewer neutralizing epitopes compared to FLSP vaccines. Second, the safety of the vaccine remains controversial, as clinical studies show that mRNA vaccines may induce immune thrombotic thrombocytopenia. In addition, the continuous emergence of SARS-CoV-2 mutant strains requires researchers to detect S protein mutations and update the vaccine in a timely manner.

A wide range of neutralizing antibodies have been found for targeting RBD and the NTD of the SARS-CoV-2 S1 subunit to prevent viral infections. As the RBD of SARS-CoV-2 and other CoVs structurally resemble each other, the possible cross-neutralizing effects of current SARS-CoV neutralizing antibodies against SARS-CoV-2 infection have been thoroughly examined. For example, scientists have discovered the SARS-CoV RBD-specific monoclonal antibody (CR3022) is capable of binding with a conserved epitope of the SARS-CoV-2 RBD region, and human 47D11 antibodies are able to neutralize both SARS-CoV-2 and SARS-CoV in the experimental settings with Vero-E6 cell lines. Convaлексnt plasma sera from the SARS-CoV infected patients are also observed to cross-neutralize SARS-CoV-2 particles. Therefore, using antibody cocktails containing the antibodies for targeting the different epitopes of the spike protein could be a promising therapeutic intervention. In terms of the S2 subunit that contains an N-terminal FP with heptad repeat 1 and 2 (HR1, HR2) domains, a transmembrane domain, and a cytoplasmic domain, is required for the virus-cell membrane fusion, in vitro and in vivo studies have indicated that the EK1C4 (an inhibitor of coronavirus-host cell membrane fusion) inhibits SARS-CoV-2 infection by potentially targeting the HR1 domain.

1.11 Specific drugs targeting SARS-CoV-2 spike protein

The effective strategy for preventing SARS-CoV-2 infection is to stop the viral entry into the host cell. Therefore, extensive studies have been undergoing on specific proteins, peptides, or small molecule compounds that can bind to the S protein, thereby preventing the interaction between virus and host cell membrane (Table 1). For example, ivermectin and arbidol can block viral membrane fusion by competitively inhibiting the interaction between S protein RBD and ACE2, while lipopeptide EK1 and its derivative eK1c4, as a pan coronavirus fusion inhibitor, inhibits membrane fusion by
### Inhibitory drugs targeting SARS-CoV-2 spike protein-related receptors

The inhibitors used to downregulate the activity of the SARS receptor and cofactor and block the binding of S protein to the receptor are also one of the strategies to combat SARS-CoV-2 infection (Table 2). As the main receptor for SARS-CoV-2 to enter host cells, ACE2 is an important target for many COVID-19 treatment strategies. Importantly, the critical residues for ACE2 receptors have been identified to binding with the C-terminal SD-1 domain (CTD) of the SARS-CoV-2 S protein, which could be a promising target for antivirus therapeutics. Some studies have shown the use of ACE2 antibodies, which antagonize the binding of SARS-CoV-2 to ACE2 without interfering with the normal function of ACE2, may crosslink and downregulate ACE2, and thereby destroy the RAS. Similarly, ACE2 mimetics are also ideal therapeutic drugs, such as ACE2 ectodomain, its Fc-fusion form, and their variants selected for higher affinity can prevent the initial step of virus binding to the receptor without interfering with the RAS. In addition, some drugs can affect the binding of SARS-CoV-2 to ACE2 through different mechanisms. For instance, amitrityline and other FIASMAs inhibit ASM and the formation of ceramide-enriched membrane domains, which serve viral entry and infection by clustering ACE2, thereby preventing infection with SARS-CoV-2. Diltiazem, a blocker of L-type calcium channel calcium, can interact and colocalizes with SARS-CoV-2 spike protein and ACE2, thereby affecting cell attachment and internalization of SARS-CoV-2. Recent studies have also shown that peptide-based drugs could be used to compete for binding of ACE2 receptor with the SARS-CoV-2 spike protein by computational prediction of drug repurposing strategy. For example, Gu et al. discovered two small compounds protoporphyrin IX and verteporfin, which had been approved by the FDA for the treatment of cancers and age-related macular degeneration respectively, sharing a porphyrin ring structure, are able to bind viral receptor ACE2 and interfere with the interaction between ACE2 and the RBD of viral S protein. The compounds present an effective
| Target | Compound | Approval for clinical diseases | Clinical phase | Related mechanisms |
|--------|----------|------------------------------|----------------|--------------------|
| ACE2   | Dalbavancin | Anti-cooperative ligand        | 4              | Reduce SARS-CoV-2 load and pneumonia in both mouse and rhesus macaque models.\(^{186}\) |
|        |          |                              |                | Inhibit SARS-CoV-2 cell attachment and internalization and decrease the viral infection in mouse lung.\(^{183}\) |
|        | Diltiazem | Acute heart rate control      | 3              | Prevent SARS-CoV-2 infection in mice adenovirally transduced with ACE2.\(^{185}\) |
|        | Protoporphyrin IX | Cancer screening biomarkers | 1              | Prevent SARS-CoV-2 infection in mice adenovirally transduced with ACE2.\(^{185}\) |
|        | Verteporfin | Treatment for choroidal neovascularization | –              | Inhibit of the infection of nasal epithelial cells from volunteers with pp-VSV-SARS-CoV-2 spike particle.\(^{181}\) |
|        | FIASMA amitriptyline | Antidepressant | –              | |
| TMPRS-S2 | Bromhexine | Mucolytic agent               | 4              | Early oral therapy reduces the ICU transfer, intubation, and the mortality rate in patients with COVID-19.\(^{187}\) |
|        | Camostat mesylate | Acute pancreatitis | 3              | Block SARS-CoV-2 infection of lung cells.\(^{59,188}\) |
|        | Nafamostat | Anticoagulant                 | 3              | Reduce SARS-CoV-2 pulmonary infection in mouse models.\(^{188}\) |
|        | α1-antitrypsin (α1AT) | –                         | –              | Inhibit SARS-CoV-2 entry and suppress viral replication in cell lines and primary cells.\(^{189}\) |
|        | N-0385 | –                             | –              | Afford a high level of prophylactic and therapeutic benefit in the K18-human ACE2 transgenic mouse model.\(^{190}\) |
| CatB/L | Amantadine | Anti-influenza                | 3              | Inhibit SARS-CoV-2 cell entry with little cytotoxicity.\(^{191}\) |
|        | E-64d | Treatment for Alzheimer's disease | 2              | Block SARS-CoV-2 infection of lung cells with Camostat mesylate.\(^{59}\) |
|        | K777 | Antiparasite drug             | –              | Block SARS-CoV-2 infection of human and monkey cells.\(^{192}\) |
| Furin  | Decanoyl-RVKR-chloromethylketone (CMK) | –                        | –              | Inhibit SARS-CoV-2 infection in Vero E6 cell.\(^{193}\) |
antiviral concentration and a broad margin of safety in the cell and mouse model, but it is required for clinical evaluation in vivo.

Given the activation of SARS-CoV-2 S protein depends on the host protease cleavage of S1/S2 and S2 sites,\(^9\) it is another group of potential targets for developing inhibitors against SARS-CoV-2. In this aspect, the relevant drugs supported by experimental data include furan inhibitors,\(^{194}\) Tmprss2 inhibitors (camostat mesylate,\(^{59}\) Nafamostat\(^{195}\)), and cathepsin L inhibitors (E-64\(^{19}\)). It is worthy to mention that inhibitors of S protein cofactors also have antiviral effects. For example, PM26, a glycomimetric antagonist of DC-SIGN, is able to inhibit the interaction of the S protein with the lectin receptor and blocks DC-SIGN-mediated SARS-CoV-2 infection.\(^{80}\) However, if these drugs are used for the prevention or treatment of SARS-CoV-2 infection, they still need to determine the time point and dosage of the best curative effect, clinical therapeutic effect, and side effects.

### 2 | CONCLUSION

In summary, even though several COVID-19 vaccines are currently in distribution worldwide, new threats are posed by the emerging circulating variants of SARS-CoV-2. The high mortality rate of critical illness is still a serious threat, due to the lack of safe and effective drugs for clinical treatments. We review and highlight the signal pathway modulation by SARS-CoV-2 S protein and its receptors, particularly current therapies and vaccines targeting the viral entry machinery. In-depth research is still required to comprehend more details of the SARS-CoV-2 spike protein mutations and mode of interactions with receptors and cofactors by innovative structural biology techniques and will provide valuable information for successful antiviral therapeutics and vaccination against COVID-19.

### AUTHOR CONTRIBUTIONS

Shuvomoy Banerjee drafted review parts of introduction, evolution, tropism, and structure, as well as preparing analysis. Xinyu Wang, Yuping Jia, and Yuyan Wang drafted review parts of host factors, signaling, therapeutic, and conclusion. Shujuan Du and Caixia Zhu modified figures and critical reading. Yuyan Wang and Qiliang Cai supervised and editing paper.

### ACKNOWLEDGMENTS

This study was supported by the National Key Research and Development Program (2021YFA1300803), National Natural Science Foundation of China (81971930, 32120103001, 82102386), Shanghai Municipal Science and Technology Major Project (ZD2021CY001), and Jinan University and Institute Innovation Program (2020GXRC043). Qiliang Cai is a scholar of New Century Excellent Talents at the University of China.

### CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

### DATA AVAILABILITY STATEMENT

Data sharing not applicable to this article as no datasets were generated or analysed during the current study. The data that support the findings of this study are openly available in National Center for Biotechnology Information (NCBI) database and analyzed in The Interactive Tree Of Life online software tool at https://itol.embl.de; the Open Access FunRich functional enrichment analysis tool at http://www.funrich.org/index.html.

### ORCID

Qiliang Cai http://orcid.org/0000-0002-7147-0953

### REFERENCES

1. Madjid M, Miller CC, Zarubaev VV, et al. Influenza epidemics and acute respiratory disease activity are associated with a surge in autopsy-confirmed coronary heart disease death: results from 8 years of autopsies in 34,892 subjects. Eur Heart J. 2007;28(10):1205-1210.

2. Ljubin-Sternak S, Mestrovic T, Luksic I, Mijac M, Vranes J. Seasonal coronaviruses and other neglected respiratory viruses: a global perspective and a local snapshot. Front Public Health. 2021;9:691163.

3. Letunic I, Bork P. Interactive Tree Of Life (iTOl) v5: an online tool for phylogenetic tree display and annotation. Nucleic Acids Res. 2021;49(W2):W293-W296.

4. Haider N, Rothman-Ostrow P, Osman AY, et al. COVID-19—zoonosis or emerging infectious disease? Front Public Health. 2020;8:596944.

5. El-Sayed A, Kamel M. Climatic changes and their role in emergence and re-emergence of diseases. Environ Sci Pollut Res Int. 2020;27(18):22336-22352.

6. Liu J, Xie W, Wang Y, et al. A comparative overview of COVID-19, MERS and SARS: review article. Int J Surg. 2020;81:1-8.

7. Mouyra DT, Yadav PD, Ullas PT, et al. Emerging/re-emerging viral diseases & new viruses on the Indian horizon. Indian J Med Res. 2019;149(4):447-467.

8. Sharma A, Tiwari S, Deb MK, Marty JL. Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2): a global pandemic and treatment strategies. Int J Antimicrob Agents. 2020;56(2):106054.

9. Chakraborty I, Malty P. COVID-19 outbreak: migration, effects on society, global environment and prevention. Sci Total Environ. 2020;728:138882.

10. Huang Y, Yang C, Xu XF, Xu W, Liu SW. Structural and functional properties of SARS-CoV-2 spike protein: potential antivirus drug development for COVID-19. Acta Pharmacol Sin. 2020;41(9):1141-1149.

11. Belouzard S, Millet JK, Licitara BN, Whittaker GR. Mechanisms of coronavirus cell entry mediated by the viral spike protein. Viruses. 2012;4(6):1011-1033.

12. Shang J, Wan Y, Luo C, et al. Cell entry mechanisms of SARS-CoV-2. Proc Natl Acad Sci U S A. 2020;117(21):11727-11734.

13. Lu B, Yan Y, Dong L, et al. Integrated characterization of SARS-CoV-2 genome, microbiome, antibiotic resistance and host response from single throat swabs. Cell Discov. 2021;7(1):19.

14. Schuhenn J, Meister TL, Todt D, et al. Differential interferon-alpha subtype induced immune signatures are associated with suppression of SARS-CoV-2 infection. Proc Natl Acad Sci U S A. 2022;119(8):e2111600119.

15. Suzuki YJ, Nikolaienko SI, Dibrova VA, et al. SARS-CoV-2 spike protein-mediated cell signaling in lung vascular cells. Vascul Pharmacol. 2021;137:106823.
16. Suzuki YJ. The viral protein fragment theory of COVID-19 pathogenesis. Med Hypotheses. 2020;144:110267.

17. Sternberg A, Naujokat C. Structural features of coronavirus SARS-CoV-2 spike protein: targets for vaccination. Life Sci. 2020;205:118056.

18. Abdelghany TM, Ganash M, Bakri MM, Qanash H, Al-Rajhi AMH, Elhussieny NI. SARS-CoV-2, the other face to SARS-CoV and MERS-CoV: future predictions. Biomed J. 2021;44(1):86-93.

19. Coronaviridae Study Group of the International Committee on Taxonomy of Viruses. The species Severe acute respiratory syndrome-related coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2. Nat Microbiol. 2020;5(4):536-544.

20. Zhang S, Qiao S, Yu J, et al. Bat and pangolin coronavirus spike glycoprotein structures provide insights into SARS-CoV-2 evolution. Nat Commun. 2021;12:1607.

21. Zhou P, Yang XL, Wang XG, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature. 2020;579(7798):270-273.

22. Touati R, Haddad-Boubaker S, Ferchichi I, et al. Comparative genomic signature representations of the emerging COVID-19 coronavirus and other coronaviruses: high identity and possible recombination between Bat and Pangolin coronaviruses. Genomics. 2020;112(6):4189-4202.

23. Sironi M, Hasnain SE, Rosenthal B, et al. SARS-CoV-2 and COVID-19: a genetic, epidemiological, and evolutionary perspective. Infect Genet Evol. 2020;84:104384.

24. Johnson CK, Hitchens PL, Pandit PS, et al. Global shifts in mammalian population trends reveal key predictors of virus spillover risk. Proc Biol Sci. 2020;287(1924):20192736.

25. Cui J, Li F, Shi ZL. Origin and evolution of pathogenic coronaviruses. Nat Rev Microbiol. 2019;17(3):181-192.

26. Valitutto MT, Aung O, Tun KYN, et al. Detection of novel coronavirus in bats in Myanmar. PLoS One. 2020;15(4):e0230802.

27. Fonseka P, Pathan M, Chitti SV, Kang T, Mathivanan S. FunRich enables enrichment analysis of OMICS datasets. J Mol Biol. 2021;433(11):166747.

28. Jackson CB, Farzan M, Chen B, Choe H. Mechanisms of SARS-CoV-2 entry into cells. Nat Rev Mol Cell Biol. 2021;23:3-20.

29. Zipeto D, da Fonseca Palmeira JD, Argaranza GA, Argaranza ER. ACE2/ADAM17/TMPRSS2 interplay may be the main risk factor for COVID-19. Front Immunol. 2020;11:576745.

30. V’Kovski P, Kretzal A, Steiner S, Stalder H, Thiel V. Coronavirus biology and replication: implications for SARS-CoV-2. Nat Rev Microbiol. 2021;19(3):155-170.

31. Tang T, Jaimes JA, Bidon MK, Straus MR, Daniel S, Whittaker GR. Proteolytic activation of SARS-CoV-2 spike at the S1/S2 boundary: potential role of proteases beyond furin. ACS Infect Dis. 2021;7(2):264-272.

32. Cantuti-Castelvetri L, Ojha R, Pedro LD, et al. Neuroplin-1 facilitates SARS-CoV-2 cell entry and infectivity. Science. 2020;370(6518):856-860.

33. Finkelstein MT, Mermelstein AG, Miller EP, Seth PC, Stancofski ED, Fera D. Structural analysis of neutralizing epitopes of the SARS-CoV-2 spike to guide therapy and vaccine design strategies. Viruses. 2021;13(1):134.

34. Xia X. Domains and functions of spike protein in SARS-CoV-2 in the context of vaccine design. Viruses. 2021;13(1):109.

35. McFadden G, Mohamed MR, Rahman MM, Bartee E. Cytokine determinants of viral tropism. Nat Rev Immunol. 2009;9(9):645-655.

36. Harrison AG, Lin T, Wang P. Mechanisms of SARS-CoV-2 transmission and pathogenesis. Trends Immunol. 2020;41(12):1100-1115.

37. Ni W, Yang X, Yang D, et al. Role of angiotensin-converting enzyme 2 (ACE2) in COVID-19. Crit Care. 2020;24(1):422.
58. Trougakos IP, Stamatelopoulos K, Terpos E, et al. Insights to SARS-CoV-2 life cycle, pathophysiology, and rationalized treatments that target COVID-19 clinical complications. J Biomed Sci. 2021;28(1):9.

59. Hoffmann M, Kleine-Weber H, Schroeder S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell. 2020;181(2):271-280.e8.

60. Gu Y, Cao J, Zhang X, et al. Receptor profiling identifies KREMEN1 and ASGR1 as alternative functional receptors of SARS-CoV-2. Cell Res. 2022;32(1):24-37.

61. Zhao Y, Zhao Z, Wang Y, Zhou Y, Ma Y, Zuo W. Single-cell RNA expression profiling of ACE2, the receptor of SARS-CoV-2. Am J Respir Crit Care Med. 2020;202(5):756-759.

62. Wang Q, Zhang Y, Wu L, et al. Structural and functional basis of SARS-CoV-2 entry using human ACE2. Cell. 2020;181(4):894-904.e9.

63. Peacock TP, Goldhill DH, Zhou J, et al. The furin cleavage site in the SARS-CoV-2 spike protein is required for transmission in ferrets. Nat Microbiol. 2021;6(7):899-909.

64. Guruprasad L. Human SARS CoV‐2 spike protein mutations. Proteins. 2021;89(5):569-576.

65. Shang J, Ye G, Shi K, et al. Structural basis of receptor recognition by SARS-CoV-2. Nature. 2020;581(7807):221-224.

66. Ke Z, Oton J, Qu K, et al. Structures and distributions of SARS-CoV-2 spike proteins on intact virions. Nature. 2020;588(7838):498-502.

67. Yi C, Sun X, Ye J, et al. Key residues of the receptor binding motif in the spike protein of SARS-CoV-2 that interact with ACE2 and neutralizing antibodies. Cell Mol Immunol. 2020;17(6):621-630.

68. Lim H, Baek A, Kim J, et al. Hot spot profiles of SARS-CoV-2 and human ACE2 receptor protein interaction obtained by density functional tight binding fragment molecular orbital method. Sci Rep. 2020;10:16862.

69. Laurini E, Marson D, Aulic S, Feregglia A, Pricl S. Computational mutagenesis at the SARS-CoV-2 spike protein/angiotensin-converting enzyme 2 binding interface: comparison with experimental evidence. ACS Nano. 2021;15(4):6929-6948.

70. Ghorbani M, Brooks BR, Klauda JB. Critical sequence hotspots for binding of novel coronavirus to angiotensin converter enzyme as evaluated by molecular simulations. J Phys Chem B. 2020;124(45):10034-10047.

71. Gur M, Taka E, Yilmaz SZ, Kilinc C, Aktas U, Golcuk M. Conformational transition of SARS-CoV-2 spike glycoprotein between its closed and open states. J Chem Phys. 2020;153(7):075101.

72. Zhang Q, Chen CZ, Swaroop M, et al. Heparan sulfate assists SARS-CoV-2 in cell entry and can be targeted by approved drugs in vitro. Cell Discov. 2020;6(1):80.

73. Finkelsstein MT, Mermelstein AG, Miller EP, Seth PC, Stancofski ESD, Fera D. Structural analysis of neutralizing epitopes of the SARS-CoV-2 spike to guide therapy and vaccine design strategies. Viruses. 2021;13(1):134.

74. Watanabe Y, Allen JD, Wrapp D, McLellan JS, Crispin M. Site-specific glycan analysis of the SARS-CoV-2 spike. Science. 2020;369(6501):330-333.

75. Zhao P, Praissman JL, Grant OC, et al. Virus-receptor interactions of glycosylated SARS-CoV-2 spike and human ACE2 receptor. Cell Host Microbe. 2020;28(4):586-601.e6.

76. Basu A, Sarkar A, Maulik U. Molecular docking study of potential phytochemicals and their effects on the complex of SARS-CoV2 spike protein and human ACE2. Sci Rep. 2020;10:17699.

77. Casalino L, Gaib Z, Goldsmith JA, et al. Beyond shielding: the roles of glycans in the SARS-CoV-2 spike protein. ACS Cent Sci. 2020;6(10):1722-1734.

78. Grabovenko F, Nikiforova L, Yanenko B, et al. Glycosylation of receptor binding domain of SARS-CoV-2 S-protein influences on binding to immobilized DNA aptamers. Int J Mol Sci. 2022;23(1):557.

79. Lempp FA, Sorriaga LB, Montiel-Ruiz M, et al. Lectins enhance SARS-CoV-2 infection and influence neutralizing antibodies. Nature. 2021;598(7880):342-347.

80. Thépaut M, Luczkoziak J, Vivès C, et al. DC/L-SIGN recognition of spike glycoprotein promotes SARS-CoV-2 trans-infection and can be inhibited by a glycomimetic antagonist. PLoS Pathog. 2021;17(5):e1009576.

81. Guo L, Liang Y, Li H, et al. Epigenetic glycosylation of SARS-CoV-2 impact viral infection through DC-L-SIGN receptors. iScience. 2021;24(12):103426.

82. Choudhury A, Mukherjee S. In silico studies on the comparative characterization of the interactions of SARS-CoV-2 spike glycoprotein with ACE-2 receptor homologs and human TLRs. J Med Virol. 2020;92(10):2105-2113.

83. Gebrehiwot AG, Helika DS, Kassaye YM, et al. Healthy human serum N-glycan profiling reveals the influence of ethnic variation on the identified cancer-relevant glycan biomarkers. PLoS One. 2018;13(12):e0209515.

84. Kriścić J, Vučković F, Menni C, et al. Glycans are a novel biomarker of chronological and biological ages. J Gerontol A Biol Sci Med Sci. 2014;69(7):779-789.

85. Korber B, Fischer WM, Gnanakaran S, et al. Tracking changes in SARS-CoV-2 spike: evidence that D614G increases infectivity of the COVID-19 virus. Cell. 2020;182(4):812-827.e19.

86. Li Q, Wu J, Nie J, et al. The impact of mutations in SARS-CoV-2 spike on viral infectivity and antigenicity. Cell. 2020;182(5):1284-1294.e9.

87. Gadaneck LK, Mcsweeney KR, Qaradakhi T, Ali B, Zulli A, Apostolopoulos V. Can SARS-CoV-2 virus use multiple receptors to enter host cells? Int J Mol Sci. 2021;22(3):992.

88. Sabiri R, Koseler A, Goren T, Turkuc I, Kurt O. High GRP78 levels in Covid-19 infection: a case-control study. Life Sci. 2021;265:118781.

89. Mori Y, Fink C, Ichimura T, et al. KIM-1/TIM-1 is a receptor for SARS-CoV-2 in lung and kidney. medRxiv. 2022.

90. Wang S, Qiu Z, Hou Y, et al. AXL is a candidate receptor for SARS-CoV-2 that promotes infection of pulmonary and bronchial epithelial cells. Cell Res. 2021;31(2):126-140.

91. Yang ZY, Huang Y, Ganesh L, et al. pH-dependent entry of severe acute respiratory syndrome coronavirus is mediated by the spike glycoprotein and enhanced by dendritic cell transfer through DC-SIGN. J Virol. 2004;78(11):5642-5650.

92. Navarro-Sanchez E, Altmeyer R, Amara A, et al. Dendritic-cell-specific ICAM3-grabbing non-integрин is essential for the productive infection of human dendritic cells by mosquito-cell-derived dengue viruses. EMBO Rep. 2003;4(7):723-728.

93. Ghebraeli M, Wang K, Viveiros A, et al. Angiotensin-converting enzyme 2: SARS-CoV-2 receptor and regulator of the renin-angiotensin system: celebrating the 20th anniversary of the discovery of ACE2. Circ Res. 2020;126(10):1456-1474.

94. Donoghue M, Hsieh F, Baronas E, et al. A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9. Circ Res. 2000;87(5):E1-E9.

95. Turner AJ, Hooper NM. The angiotensin-converting enzyme gene family: genomics and pharmacology. Trends Pharmacol Sci. 2002;23(4):177-183.

96. Lambert DW, Yarski M, Warner FJ, et al. Tumor necrosis factor-alpha convertase (ADAM17) mediates regulated ectodomain shedding of the severe-acute respiratory syndrome-coronavirus (SARS-CoV) receptor, angiotensin-converting enzyme-2 (ACE2). J Biol Chem. 2005;280(34):30113-30119.

97. Yeung ML, Teng JL, Jia L, et al. Soluble ACE2-mediated cell entry of SARS-CoV-2 via interaction with proteins related to the renin-angiotensin system. Cell. 2021;184(8):2212-2228.e12.
98. Kornilov SA, Lucas I, Jade K, Dai CL, Lovejoy JC, Magis AT. Plasma levels of soluble ACE2 are associated with sex, metabolic syndrome, and its biomarkers in a large cohort, pointing to a possible mechanism for increased severity in COVID-19. Crit Care. 2020;24(1):452.

99. Kulkarni PG, Sakharkar A, Banerjee T. Understanding the role of nACE2 in neurogenic hypertension among COVID-19 patients. Hypertens Res. 2022;45(2):254-269.

100. Srimulata S, Xia H, Xu P, Lazartigues E. Brain-targeted angiotensin-converting enzyme 2 overexpression attenuates neurogenic hypertension by inhibiting cyclooxygenase-mediated inflammation. Hypertension. 2015;65(3):577-586.

101. Moriguchi T, Harii N, Goto J, et al. A first case of meningitis/encephalitis associated with SARS-CoV-2. Int J Infect Dis. 2020;94:55-58.

102. Liu Y, Yang Y, Zhang C, et al. Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. Sci China Life Sci. 2020;63(3):364-374.

103. Patra T, Meyer K, Geerling L, et al. SARS-CoV-2 infection in mice: a matter of SARS-CoV-2 spike protein promotes IL-6 trans-signaling by activation of angiostatin II receptor signaling in epithelial cells. PLoS Pathog. 2020;16(12):e1009128.

104. Ratajczak MZ, Bujko K, Ciechanowicz A, et al. SARS-CoV-2 spike protein activates the Nlrp3 inflammasome. Stem Cell Res Rep. 2021;17(1):266-277.

105. Swanson KV, Deng M, Ting JP. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. Nat Rev Immunol. 2019;19(8):477-489.

106. Sun X, Wang T, Cai D, et al. Cytokine storm intervention in the early stages of COVID-19 pneumonia. Cytokine Growth Factor Rev. 2020;53:38-42.

107. Ratajczak MZ, Bujko K, Ciechanowicz A, et al. SARS-CoV-2 entry receptor ACE2 is expressed on very small CD45-precursors of hematopoietic and endothelial cells and in response to virus spike protein activates the Nlrp3 inflammasome. Stem Cell Res Rep. 2021;17(1):266-277.

108. Swanson KV, Deng M, Ting JP. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. Nat Rev Immunol. 2019;19(8):477-489.

109. Barreda D, Santiago C, Rodríguez JR, et al. SARS-CoV-2 spike protein and its receptor binding domain promote a proinflammatory activation profile on human dendritic cells. Cells. 2021;10(12):3279.

110. Rabeckzak MZ, Bujko K, Ciechanowicz A, et al. SARS-CoV-2 entry receptor ACE2 is expressed on very small CD45-precursors of hematopoietic and endothelial cells and in response to virus spike protein activates the Nlrp3 inflammasome. Stem Cell Res Rep. 2021;17(1):266-277.

111. Bagchi A, Herrup EA, Warren HS, et al. MyD88-independent toll-like receptor signaling pathway. Mol Psychiatry. 2021;26(4):1044-1059.

112. Albalawi F, Lu W, Beckel JM, Lin JC, McCaughey SA, Mitchell CH. The P2X7 receptor primes IL-1β and the NLRP3 inflammasome in astrocytes exposed to mechanical strain. Front Cell Neurosci. 2017;11:227.

113. Sun X, Wang T, Cai D, et al. Cytokine storm intervention in the early stages of COVID-19 pneumonia. Cytokine Growth Factor Rev. 2020;53:38-42.

114. Takeda K, Akira S. Toll-like receptors in innate immunity. Int Immunol. 2005;17(1):1-14.

115. Bagchi A, Herrup EA, Warren HS, et al. MyD88-dependent and MyD88-independent pathways in synergy, priming, and tolerance between TLR agonists. J Immunol. 2007;178(2):1164-1171.

116. Yamamoto M, Sato S, Hemmi H, et al. Role of adaptor TRIF in the MyD88-independent toll-like receptor signaling pathway. Science. 2003;301(5633):640-643.

117. Khan S, Shafiei MS, Longoria C, Schoggins JW, Savani RC, Zaki H. SARS-CoV-2 spike protein induces inflammation via TLR2-dependent activation of the NF-κB pathway. eLife. 2021;10:e68563.

118. Frank MG, Nguyen KH, Ball JB, et al. SARS-CoV-2 spike S1 subunit induces neuroinflammation, microglial and behavioral sickness responses: evidence of PAMP-like properties. Brain Behav Immun. 2022;100:267-277.

119. Shirato K, Kiizaki T. SARS-CoV-2 spike protein S1 subunit induces pro-inflammatory responses via toll-like receptor 4 signaling in murine and human macrophages. Heilinon. 2021;7(2):e06187.

120. Olajide OA, Iwuananywu VU, Adegbola OD, Al-Hindawi AA. SARS-CoV-2 spike glycoprotein S1 induces neuroinflammation in BV-2 microglia. Mol Neurobiol. 2022;59(1):445-458.

121. Zhao Y, Kuang M, Li J, et al. SARS-CoV-2 spike protein interacts with and activates TLR4. Cell Res. 2021;31(7):818-820.

122. Katz DH, Tahir UA, Ngo D, et al. Proteomic profiling in biracial cohorts implicates DC-SIGN as a mediator of genetic risk in COVID-19. medRxiv. 2020.

123. Relloso M, Puig-Kroger A, Pello OM, et al. DC-SIGN (CD209) expression is IL-4 dependent and is negatively regulated by IFN, TGF-β, and anti-inflammatory agents. J Immunol. 2002;168(6):2634-2643.

124. Lu L, Zhang H, Dauphars DJ, He YW. A potential role of interleukin 10 in COVID-19 pathology. Trends Immunol. 2021;42(1):3-5.

125. Trbojević-Akmacić I, Petrović T, Lauc G. SARS-CoV-2 S glycoprotein binding to multiple host receptors enables cell entry and infection. Glycoconjug J. 2021;38(5):611-623.

126. Stravalaci M, Pagani I, Paraboschi EM, et al. Recognition and inhibition of SARS-CoV-2 by humoral innate immunity pattern recognition molecules. Nature Immunol. 2022;23(2):275-286.

127. Evans JP, Liu S-L. Role of host factors in SARS-CoV-2 entry. J Biol Chem. 2021;297(1):100847.

128. Lee AS. The ER chaperone and signaling regulator GRP78/BiP as a monitor of endoplasmic reticulum stress. Methods. 2005;35(4):373-381.

129. Shimasaki S, Koga T, Shuto T, et al. Endoplasmic reticulum stress increases the expression and function of toll-like receptor-2 in epithelial cells. Biochem Biophys Res Commun. 2010;402(2):235-240.

130. Ibrahım IM, Abdelmalek DH, Elshehate ME, Elfiky AA. COVID-19 spike-host cell receptor GRP78 binding site prediction. J Infect. 2020;80(5):554-562.

131. Huang X, Ye Q, Chen M, et al. N-glycosylation-defective spike variants of neuruplin-1 promote metastasis by activating endosomal signals. Nat Commun. 2019;10(1):3708.

132. Maden CH, Gomes J, Schwarz Q, Davidson K, Tinker A, Ruhrberg C. NRP1 and NRP2 cooperate to regulate gangliogenesis, axon guidance and target innervation in the sympathetic nervous system. Dev Biol. 2012;369(2):277-285.

133. Cantuti-Castelvetri L, Ojha R, Pedro LD, et al. Neuruplin-1 facilitates SARS-CoV-2 cell entry and infectivity. Science. 2020;370(6518):856-860.

134. Daly JL, Simonetti B, Klein K, et al. Neuruplin-1 is a host factor for SARS-CoV-2 infection. Science. 2020;370(6518):861-865.

135. Bonjardim CA. Viral exploitation of the MEK/ERK pathway in reality and challenges. J Microbiol Immunol Infect. 2019;52(1):27-37.

136. Jean SS, Lee PI, Hsueh PR. Treatment options for COVID-19: a tale of vaccinia virus and other viruses. Virology. 2017;507:267-275.

137. Jean SS, Lee PI, Hsueh PR. Treatment options for COVID-19: the reality and challenges. J Microbiol Immunol Infect. 2019;52(1):27-37.

138. Hills M, Bajic VB, Forrest J, et al. Identification of vaccinia virus and other viruses. Virology. 2017;507:267-275.

139. Sun DX. Remdesivir for treatment of COVID-19: combination of pulmonary and IV administration may offer additional benefit. AAPS J. 2020;22(4):77.

140. Khuroo MS. Chloroquine and hydroxychloroquine in coronavirus disease 2019 (COVID-19). Facts, fiction and the hype: a critical appraisal. Int J Antimicrob Agents. 2020;56(3):106101.
140. Sameeh H, Mohsenzadeeghan M, Ala S, Maroufi SS, Moradimaj P, Tolcilizumab for treatment patients with COVID-19: recommended medication for novel disease. Int Immunopharmacol. 2020;89:107018.

141. Saha RP, Sharma AR, Singh MK, et al. Repurposing drugs, ongoing vaccine, and new therapeutic development initiatives against COVID-19. Front Pharmacol. 2020;11:1258.

142. Taylor PC, Adams AC, Hufford MM, de la Torre I, Winthrop K, Gottlieb RL. Neutralizing monoclonal antibodies for treatment of COVID-19. Nat Rev Immunol. 2021;21(6):382-393.

143. Malik JA, Mulla AH, Farooqi T, Pottou FH, Anwar S, Rengasamy KRR. Targets and strategies for vaccine development against SARS-CoV-2. Biomed Pharmacother. 2021;137:11254.

144. van Doremalen N, Lambe T, Spencer A, et al. ChAdOx1 nCoV-19 vaccine prevents SARS-CoV-2 pneumonia in rhesus macaques. Nature. 2020;586(7830):578-582.

145. Voysey M, Clemens SAC, Madhi SA, et al. Safety and efficacy of the ChAdOx1 nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. Lancet. 2021;397(10269):99-111.

146. Voysey M, Costa Clemens SA, Madhi SA, et al. Single-dose administration and the influence of the timing of the booster dose on immunogenicity and efficacy of ChAdOx1 nCoV-19 (AZD1222) vaccine: a pooled analysis of four randomised trials. Lancet. 2021;397(10277):881-891.

147. Muhammed Y, Yusuf Nadabo A, Pius M, et al. SARS-CoV-2 spike protein and RNA dependent RNA polymerase as targets for drug and vaccine development: a review. Biosaf Health. 2021;3(5):249-263.

148. Wang Z, Schmidt F, Weisblum Y, et al. mRNA vaccine elicited antibodies to SARS-CoV-2 and circulating variants. Nature. 2021;592(7855):616-622.

149. Krammer F. SARS-CoV-2 vaccines in development. Nature. 2020;586(7830):516-527.

150. Gallo K, Goede A, Mura C, et al. A comparative analysis of COVID-19 vaccines based on over 580,000 cases from the vaccination adverse event reporting system. Vaccines. 2022;10(3):408.

151. Cines DB, Bussel JB. SARS-CoV-2 vaccine-induced immune thrombotic thrombocytopenia. N Engl J Med. 2020;383(23):2254-2256.

152. Fu Y, Pan Y, Li Z, Li Y. The utility of specific antibodies against SARS-CoV-2 in laboratory diagnosis. Front Microbiol. 2020;11:603058.

153. Rattanapisit K, Shanmugaraj B, Manopwisedjaroen S, et al. Rapid production of SARS-CoV-2 receptor binding domain (RBD) and spike specific monoclonal antibody CR3022 in Nicotiana benthamiana. Sci Rep. 2020;10(1):17698.

154. Wang C, Li W, Drabek D, et al. A human monoclonal antibody blocking SARS-CoV-2 infection. Nat Commun. 2020;11(1):2251.

155. Chen RE, Zhang X, Case JB, et al. Resistance of SARS-CoV-2 variants to neutralization by monoclonal and serum-derived polyclonal antibodies. Nat Med. 2021;27(4):717-726.

156. Baum A, Fulton BO, Wloda E, et al. Antibody cocktail to SARS-CoV-2 spike protein prevents rapid mutational escape seen with individual antibodies. Science. 2020;369(6506):1014-1018.

157. Tang T, Bidon M, Jaimes WA, Whitaker GR, Daniel S. Coronavirus membrane fusion mechanism offers a potential target for antiviral development. Antivir Res. 2020;178:104792.

158. Xia S, Liu M, Wang C, et al. Inhibition of SARS-CoV-2 (previously 2019-nCoV) infection by a highly potent pan-coronavirus fusion inhibitor targeting its spike protein that harbors a high capacity to mediate membrane fusion. Cell Res. 2020;30(4):343-355.

159. Oliveira OV, Rocha GB, Paluch AS, Costa LT. Repurposing approved drugs as inhibitors of SARS-CoV-2: S-protein from molecular modeling and virtual screening. J Biol Mol Struct Dyn. 2021;39(11):3924-3933.

160. Vankadari N. Arbidol: a potential antiviral drug for the treatment of SARS-CoV-2 by blocking trimerization of the spike glycoprotein. Int J Antimicrob Agents. 2020;56(2):105998.
182. Carpinteiro A, Gripp B, Hoffmann M, et al. Inhibition of acid sphingomyelinase by ambroxol prevents SARS-CoV-2 entry into epithelial cells. J Biol Chem. 2021;296:100701.

183. Wang X, Luo J, Wen Z, et al. Diltiazem inhibits SARS-CoV-2 cell attachment and internalization and decreases the viral infection in mouse lung. PLoS Pathog. 2022;18(2):e1010343.

184. Egieyeh S, Egieyeh E, Malan S, Christofells A, Fielding B. Computational drug repurposing strategy predicted peptide-based drugs that can potentially inhibit the interaction of SARS-CoV-2 spike protein with its target (humanACE2). PLoS One. 2021;16(1):e0245258.

185. Gu C, Wu Y, Guo H, et al. Protoporphyrin IX and verteporfin potently inhibit SARS-CoV-2 infection in vitro and in a mouse model expressing human ACE2. Sci Bull. 2021;66(9):925-936.

186. Wang G, Yang M-L, Duan Z-L, et al. Dalbavancin binds ACE2 to block its interaction with SARS-CoV-2 spike protein and is effective in inhibiting SARS-CoV-2 infection in animal models. Cell Res. 2021;31(1):17-24.

187. Ansarin K, Tolouian R, Ardalan M, et al. Effect of bromhexine on clinical outcomes and mortality in COVID-19 patients: A randomized clinical trial. Bioimpacts. 2020;10(4):209-215.

188. Li K, Meyerholz DK, Bartlett JA, McCray PB. The TMPRSS2 Inhibitor Nafamostat Reduces SARS-CoV-2 Pulmonary Infection in Mouse Models of COVID-19. mBio. 2021;12(4):e0097021.

189. Wettstein L, Weil T, Conzelmann C, et al. Alpha-1 antitrypsin inhibits TMPRSS2 protease activity and SARS-CoV-2 infection. Nat Commun. 2021;12(1):1726.

190. Shapira T, Monreal IA, Dion SP, et al. A TMPRSS2 inhibitor acts as a pan-SARS-CoV-2 prophylactic and therapeutic. Nature. 2022.

191. Zhao M-M, Yang W-L, Yang F-Y, et al. Cathepsin L plays a key role in SARS-CoV-2 infection in humans and humanized mice and is a promising target for new drug development. Signal Transduct Target Ther. 2021;6(1):134.

192. Mellott DM, Tseng C-T, Drellich A, et al. A clinical-stage cysteine protease inhibitor blocks SARS-CoV-2 infection of human and monkey cells. ACS Chem Biol. 2021;16(4):642-650.

193. Cheng Y-W, Chao T-L, Li C-L, et al. Furin inhibitors block SARS-CoV-2 spike protein cleavage to suppress virus production and cytopathic effects. Cell Rep. 2020;33(2):108254.

194. Bollavaram K, Leeman TH, Lee MW, et al. Multiple sites on SARS-CoV-2 spike protein are susceptible to proteolysis by cathepsins B, K, L, S, and V. Protein Sci. 2021;30(6):1131-1143.

195. Hoffmann M, Schroeder S, Kleine-Weber H, Müller MA, Drosten C, Pöhlmann S. Nafamostat mesylate blocks activation of SARS-CoV-2: new treatment option for COVID-19. Antimicrob Agents Chemother. 2020;64(6):e00754-20.

How to cite this article: Banerjee S, Wang X, Du S, et al. Comprehensive role of SARS-CoV-2 spike glycoprotein in regulating host signaling pathway. J Med Virol. 2022;94:4071-4087. doi:10.1002/jmv.27820