Movements of southern elephant seals (*Mirounga leonina*) from Davis Base, Antarctica: combining population genetics and tracking data

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Abstract

Marine animals such as the southern elephant seal (*Mirounga leonina*) rely on a productive marine environment and are vulnerable to oceanic changes that can affect their reproduction and survival rates. Davis Base, Antarctica, acts as a moulting site for southern elephant seals that forage in Prydz Bay, but the mitochondrial haplotype diversity and natal source populations of these seals have not been characterized. In this study, we combined genetic and animal tracking data on these moulting seals to identify levels of mitochondrial haplotype diversity, natal source population, and movement behaviours during foraging and haul-out periods. Using partial sequences of the mitochondrial control region, we identified two major breeding mitochondrial lineages of seals at Davis Base. We found that the majority of the seals originated from breeding stocks within the South Atlantic Ocean and South Indian Ocean. One seal was grouped with the Macquarie Island breeding stock (South Pacific Ocean). The Macquarie Island population, unlike the other two stocks, is decreasing in size. Tracking data revealed long-distance foraging activity of the Macquarie Island seal around Crozet Islands. We speculate that changes to the Antarctic marine environment can result in a shift in foraging and movement strategies, which subsequently affects seal population growth rates.

Keywords Southern elephant seals · Genetic diversity · Mitochondrial DNA · Population genetics · Moulting · Animal tracking

Introduction

Warmer oceans result in a loss of sea ice, which is likely to affect reproduction and survival rates in species that depend on sea ice for foraging (Bryndum-Buchholz et al. 2019). In other species, loss of sea ice has reduced movement corridors, which has resulted in loss of connectivity pathways for some marine animal populations (Laidre et al. 2018). As global temperatures continue to increase, changes to habitat structure in ice-locked regions are likely to cause shifts in marine animal movements, effective dispersal, foraging behaviours, and population numbers (Hindell et al. 2017, 2020). These changes can lead to population restructuring or even loss of genetic diversity (Laidre et al. 2018; Siegert et al. 2019).

Over a 40-year data record from the late 1970s, the highest peak in Antarctic sea ice was evident in 2014, but large declines occurred in 2017 and 2018, possibly caused by the El Niño Southern Oscillation (Parkinson 2019). Polar marine animals, such as pinnipeds, occupy the upper trophic levels in the Antarctic region and therefore integrate large environmental signals (Bestley et al. 2020). Sea ice is crucial to the survival of these predators, especially those that require sea ice for breeding, foraging, and moulting (Bestley et al. 2020). A recent study used a climate model to project that colonies of the Emperor Penguin (*Aptenodytes forsteri*) would become quasi-extinct by 2100 because of the reduced availability of sea ice and foraging habitat (Jenouvrier et al. 2018).
is situated near Vestfold Hills, covering roughly 400 km² (Rodríguez et al. 2017). Davis Base is an ice-free area that S, 77°58′ E) act as moulting sites for some seal colonies (Rodríguez et al. 2017). Davis Base is an ice-free area that is situated near Vestfold Hills, covering roughly 400 km² (Australian Antarctic Program 2020). Previous studies indicated that the male seals at Davis Base originated from the Kerguelen Islands, Heard Island, and some of the smaller populations in the Kerguelen stock at Marion and Crozet islands (Tierney 1977; Bester 1988). The seals’ annual visits might be due to the abundance of food along Prydz Bay during the austral summer (Bester 1988). The underlying genetic make-up of this and indeed other moulting aggregations can potentially be identified through genetic analysis. The results of such an analysis would allow comparison with traditional capture-mark-recapture studies that formed the basis of much previous work on population composition at Davis Station.

The genetic structure and genetic diversity of the breeding colonies of SES have been investigated previously using mitochondrial DNA (mtDNA) and microsatellites (Hoelzel et al. 1993; Slade et al. 1998; Fabiani 2002; Chauke 2008; Bogdanowicz et al. 2013). Previous studies have shown lower genetic diversity in the Península Valdés and Macquarie Island breeding stocks in both mtDNA and microsatellites (Slade et al. 1998; Fabiani 2002; Chauke 2008), whereas the study by Hoelzel et al. (1993) investigated only mtDNA. In contrast, genetic diversity was higher within the South Georgia stock (South Georgia, Elephant Island, Sea Lion Island, and Falkland Islands seals), and the Kerguelen and Heard Island stock (Heard Island and Marion Island seals), as seen from mitochondrial studies (Hoelzel et al. 1993) and other studies that investigated genetic diversity in both mtDNA and microsatellites (Slade et al. 1998; Chauke 2008; Bogdanowicz et al. 2013). Evidence of sex-biased dispersal was found in previous studies where male-biased gene flow and male dispersal occurred more than in females (Slade et al. 1998; Fabiani 2002; Chauke 2008; Bogdanowicz et al. 2013; Corrigan et al. 2016). However, there remains a gap in our knowledge regarding the natal origins of seals at moulting sites such as Davis Base, where seals potentially from multiple breeding sites aggregate (Bester 1988).

Southern elephant seals are polygynous and display philopatry, so breeding colonies have high female-to-male ratios (one male beachmaster to a female harem of up to 100 seals) (de Bruyn et al. 2011). Therefore, the reproductive success and dispersal of a single male can affect disproportionately the genetic structure of an entire breeding colony (Fabiani 2002; Chauke 2008). Nuclear DNA markers such as microsatellites are commonly used to infer male gene flow because these markers are inherited by both males and females. However, the natal source population of seal moulting aggregations can be inferred using molecular markers such as mtDNA, which is maternally inherited. Sequences of the mitochondrial control region effectively delineate the major breeding stocks of SES (Hoelzel et al. 1993; Slade et al. 1998; Bogdanowicz et al. 2013).

In this study, we aim to resolve the natal origins of SES that haul out at the moulting site at Davis Base, Antarctica. We analysed a partial sequence of the mitochondrial control region from blood samples and used tracking data from these seals to determine their mtDNA diversity, infer natal source location, and explore movement from Davis Base. We compared these data with previous studies of the genetic structure of surrounding breeding colonies. Our study shows that genetic and physical tracking data provide complementary information on the natal affiliations of the seals and their resource usage around the Kerguelen Plateau.
Materials and methods

Blood sample collection and animal tracking tags

We captured, collected blood samples from, and attached animal tracking devices to 12 male SES (10 subadults and 2 juveniles) at Davis Base in 2016 and 2017. Animal tracking data were collected over one year of the seals’ lifecycle, from the start of their trip (moulting at Davis Base) to returning to moult the following year. After collection, blood from each sampled seal was spotted onto Whatman FTA cards (Stowell et al. 2018) and stored at −20 °C until required for DNA extraction. The 12 blood samples were then sent to the University of Sydney for the purpose of this study.
DNA library assembly

We prepared seal blood samples using the standard sterile technique, which involved taking a sterilized (Bunsen flame) standard office one-hole punch from the centre of the dried blood sample (Stowell et al. 2018). We extracted and purified the sampled DNA using the Qiagen DNeasy Blood & Tissue Kit following the Spin-Column protocol for blood with non-nucleated erythrocytes. To amplify the mitochondrial control region, we used the primers ancF (5'-GCTGACATTTACTAAAAT-3') and mdbR (5'-CAGTAT AGAAACCCCCCATGA-3') (de Bruyn et al. 2009). We ran PCRs for approximately 3 h using the following cycle protocol: 10 min at 95 °C followed by 35 cycles of 30 s at 52 °C, 30 s at 72 °C, 1 min at 72 °C, followed by an additional extension step for 10 min, and then 4 °C for cooldown. After successful PCR, we cleaned up the PCR product following the ExoSAP-IT Express PCR Product Cleanup kit and standard protocol. The cleaned PCR products were stored at 4 °C prior to DNA sequencing by Macrogen (Seoul, South Korea).

We collected a total of 201 additional sequences of the mitochondrial control region from GenBank (Hoelzel et al. 1993; Slade et al. 1998; Fabiani 2002; Chauke 2008; Curtis et al. 2009; Bogdanowicz et al. 2013; Zappes et al. 2017). These comprised 191 sequences from southern elephant seals and 10 sequences from Weddell seals (Slade et al. 2009; Bogdanowicz et al. 2013; Zappes et al. 2017). We aligned the 12 newly generated sequences and the 201 published sequences using Geneious Prime (Kearse et al. 2012).

Genetic analyses

Uncorrected pairwise distances were calculated from sampled sequences of the mitochondrial control region using MEGA X (Kumar et al. 2018). We used pairwise deletion to account for gaps in sequences. We used DnaSP version 6.12 (Rozas et al. 2017) to calculate the number of segregating sites, number of haplotypes and haplotype diversity, nucleotide diversity and average nucleotide differences, and neutrality tests using Fu’s F_S (Fu 1997). Rarefaction was used to correct for unequal sample sizes by comparing haplotype richness between the samples from Davis Base and from all other populations. The sampled haplotype sequences were rarefied to generate the expected haplotype richness using the function rarefy (Hurlbert 1971; Heck et al. 1975) in R package vegan (Oksanen et al. 2020).

To visualize the relationships among mitochondrial haplotypes, we constructed a median-joining haplotype network using the software package POPART (Leigh and Bryant 2015). We assigned each sequence to one of nine geographical locations: Marion Island (MR), King George Island (KG), Macquarie Island (MQ), Elephant Island (EI), Falkland Islands (FI), Peninsula Valdés (PV), Heard Island (HI), South Georgia (SG), and Davis Base. The sequence data collected from GenBank were assigned locations according to where they were collected for the referenced study.

Animal tracking

We immobilized the 12 seals as part of an integrated oceanography and animal behaviour study (McMahon et al. 2021). From each seal, we took morphometric measurements including standard body length, maximum girth, and weight (Field et al. 2002). Each seal was anaesthetised using Zoletil 100, or available combinations of Tiletamine and Zolazepam (McMahon et al. 2000). We then attached identification tags to the hind flippers using Dalton Jumbo Robotags (Wilkinson and Bester 1997). We tracked the 12 seals over a year of their lifecycle from post-moulting to the next moulting haul-out season the following year (Table 1). Conductivity-Temperature-Depth Satellite Data Relay Loggers (CTD-SRDL, Sea Mammal Research Unit, University of St Andrews, UK) were glued to the top of the seal’s head for improved satellite reception and transmission at sea (Horning et al. 2019). Based on a previous study, the tag attachments to the seals were believed to have had no impact.

### Table 1

| Sample label | Age class | Mass (kg) | Tag date (d-m-y) |
|--------------|-----------|-----------|-----------------|
| 42_ SES      | Subadult  | 338       | 17-01-2016     |
| 44_ SES      | Subadult  | 320       | 15-02-2016     |
| 46_ SES      | Subadult  | 406       | 17-02-2016     |
| 48_ SES      | Subadult  | 392       | 20-02-2016     |
| 50_ SES      | Subadult  | 299       | 07-02-2017     |
| 52_ SES      | Subadult  | 309       | 16-02-2017     |
| 54_ SES      | Juvenile  | NA        | 16-02-2017     |
| 56_ SES      | Subadult  | 265       | 16-02-2017     |
| 58_ SES      | Subadult  | 345       | 20-02-2017     |
| 60_ SES      | Subadult  | 237       | 21-02-2017     |
| 62_ SES      | Subadult  | 380       | 23-02-2017     |
| 64_ SES      | Juvenile  | NA        | 25-02-2017     |

Data were collected at Davis Base, Antarctica (68°34′ 36″ S, 77°58′ 03″ E)
on their reproduction or survival patterns (McMahon et al. 2008). Tracking data received from the tags, such as location and diving behaviour, were transmitted via the ARGOS satellite network (Myers et al. 2006; Henderson et al. 2020).

We analysed the tracking data using the R package foieGras (Jonsen and Patterson 2020). We first used the fit_ssm function to fit a continuous-time correlated random walk state-space model (SSM; Jonsen et al. 2020) to the ARGOS satellite-derived locations. This model accounted for well-known measurement errors in the ARGOS locations and predicted locations at regular 12-h time intervals along the seal tracks (as per Jonsen et al. 2019). We then used the fit_mpm function to fit a movement persistence model (Jonsen et al. 2019) to the predicted locations to infer changes in the seals’ movement behaviour, possibly arising in response to stimuli such as changes in prey density or ice concentration, along their estimated tracks. Movement persistence ($\gamma_t$) is the autocorrelation in both speed and direction (scaled from 0 to 1) between successive displacements along a movement pathway. Low $\gamma_t$ values represent low speed and/or directionality that are typical of resident or area-restricted searching behaviours, whereas high $\gamma_t$ values represent higher speed and/or directionality that are typical of directed travel associated with dispersal or migration. Using the SSM-predicted locations, we also calculated the following track summary statistics: maximum displacement from deployment location; total deployment duration; maximum displacement scaled by deployment duration; and path tortuosity (mean vector of turning angles along each seal’s track).

Results

Population genetic diversity

Nucleotide sequences of the mitochondrial control region (348 bp), sampled from the 12 SES at Davis Base, showed extensive divergence between sample 48_SES and the other 11 samples (the latter were all more closely related to each other, and do not include 48_SES). Analysis of these 12 sequences and 191 published southern elephant seal sequences revealed a close relationship between sample 48_SES and sequences from Macquarie Island seals. The largest pairwise genetic distance (0.065 substitutions site$^{-1}$) is seen between seals from Macquarie Island and Península Valdés (Table 2). This is expected, given that the two locations are geographically the farthest apart. The pairwise distance between the Davis Base and Macquarie Island seals (0.049 substitutions site$^{-1}$) is greater than that between the Davis Base seals and other population groups. This confirms the divergence between sample 48_SES and the other 11 Davis Base seals.

We identified a total of 65 haplotypes in the combined data set of 203 mitochondrial sequences (Table 3). From the 12 sequences from Davis Base, 11 haplotypes were identified (two individual samples 42_SES and 62_SES shared the same haplotype). A single haplotype was carried by 26 individual seals that were previously sampled from King George Island, Marion Island, Falkland Islands, Heard Island, and one individual from Elephant Island. Six haplotypes from Davis Base seals were shared with those from Marion Island, and three haplotypes with those from the Falkland Islands. The final Davis Base haplotype grouped with the Macquarie Island population as described above. The seals from Davis Base had high haplotype diversity (0.99 ± 0.04; $n = 12$) and had higher nucleotide diversity (2.52 ± 0.004; $n = 12$) than the seals from the other populations (Macquarie Island, Marion Island, Heard Island, King George Island, and Península Valdés). Haplotype richness and average number of nucleotide differences were elevated in the Davis Base seals compared with all other populations.

Haplotype network

We assigned locations to the 12 sequences from Davis Base seals based on their haplotype grouping (Fig. 2). For the

| Table 2 | Pairwise mitochondrial genetic distances (nucleotide substitutions site$^{-1}$) between seals from eight population groups and Davis Base |
|--------|----------------------------------------------------------|
|        | MQ  | MR  | FI  | EI  | SG  | HD  | KG  | PV  |
| MR     | 0.050 |     |     |     |     |     |     |     |
| FI     | 0.053 | 0.030 |     |     |     |     |     |     |
| EI     | 0.049 | 0.031 | 0.033 |     |     |     |     |     |
| SG     | 0.052 | 0.032 | 0.035 | 0.033 |     |     |     |     |
| HD     | 0.050 | 0.027 | 0.030 | 0.032 | 0.033 |     |     |     |
| KG     | 0.047 | 0.027 | 0.029 | 0.029 | 0.030 | 0.027 |     |     |
| PV     | **0.065** | 0.042 | 0.043 | 0.047 | 0.039 | 0.039 | 0.040 |     |
| Davis Base | 0.049 | 0.030 | 0.033 | 0.034 | 0.036 | 0.031 | 0.030 | 0.044 |

Estimated standard errors for all values were approximately 0.01 ($n = 12$). Bold font denotes the largest nucleotide pairwise distance, between seals from Macquarie Island and Península Valdés. Location groups are: Macquarie Island (MQ), Marion Island (MR), Falkland Islands (FI), Elephant Island (EI), South Georgia (SG), Heard Island (HD), King George Island (KG), Península Valdés (PV)
Table 3  Statistical summary data for all southern elephant seal populations, including Davis Base seals

| Population          | N   | $H_d \pm SD$ | $\pi(\%) \pm SD$ | $k$ | $F_S$ | $r_{12}$ |
|---------------------|-----|--------------|------------------|-----|-------|---------|
| Macquarie Island    | 53  | 0.92 ± 0.02  | 2.50 ± 0.002     | 5.73| −1.00 | 7.24    |
| Marion Island       | 50  | 0.98 ± 0.01  | 2.51 ± 0.002     | 5.71| −24.87| 7.53    |
| Falkland Islands    | 16  | 1.00 ± 0.02  | 3.16 ± 0.002     | 7.24| −10.51| 10.40   |
| Elephant Island     | 12  | 1.00 ± 0.03  | 3.29 ± 0.002     | 7.53| −6.13 | 8.00    |
| South Georgia       | 28  | 0.99 ± 0.01  | 2.84 ± 0.002     | 6.51| −18.42| 9.13    |
| Heard Island        | 6   | 1.00 ± 0.10  | 2.42 ± 0.005     | 5.53| −1.96 | 4.00    |
| King George Island  | 23  | 0.91 ± 0.05  | 1.03 ± 0.002     | 2.22| −9.55 | 8.56    |
| Península Valdés    | 32  | 1.00 ± 0.27  | 0.58 ± 0.002     | 1.33| −1.22 | 2.00    |
| Davis Base          | 12  | 0.99 ± 0.04  | 2.52 ± 0.004     | 8.73| −3.31 | 11.00   |
| All combined        | 203 | 0.96 ± 0.01  | 1.96 ± 0.001     | 4.20| −60.75|         |

Statistics included are the following: the number of individuals ($N$), haplotype diversity ($H_d$), nucleotide diversity ($\pi$), average number of nucleotide differences ($k$), Fu’s $F_S$ statistic ($F_S$), and haplotype richness based on a sample $n = 12$ ($r_{12}$).

Fig. 2  Median-joining network of mitochondrial haplotypes shared between southern elephant seals from Davis Base and all other major populations. Sizes of circles represent the number of individuals per haplotype. Each colour in the pie charts represents a unique population group. The South Georgia breeding stocks are indicated as shades of green (dark blue for Península Valdés). The Kerguelen and Heard Island breeding stocks are indicated as shades of red. The Macquarie Island breeding stock are indicated by yellow. Davis Base samples are indicated as purple. Black dots represent unobserved haplotypes that have one mutational step from adjacent haplotypes. Hatch marks on lines connecting haplotypes indicate mutations. ‘$n = x$’ represents the number of Davis Base individuals that carry the specific haplotype.
seal sequences from GenBank, we assigned locations based on their sampling information. Of the 12 Davis Base seals, three had unique mitochondrial haplotypes. Two seals had haplotypes that were shared with Marion Island seals, three seals had haplotypes that were shared with seals from breeding colonies in the South Atlantic and South Indian Ocean, three seals had haplotypes shared with those from Marion Island and the South Atlantic Ocean breeding colonies, and one seal shared a haplotype with Macquarie Island seals (see below). From the combined analysis of all population groups and the Davis Base seals, 12 haplotypes were unique to the Macquarie Island stock. One Davis Base seal (48_SES) shared a haplotype with four seals from Macquarie Island. Eleven haplotypes were unique among seals from Marion Island. Out of the 11 Marion Island haplotypes, four haplotypes were shared by seals from the Falkland Islands, three shared by seals from each of Elephant Island, Heard Island, and King George Island, and two shared by seals from South Georgia. Islands in the South Atlantic Ocean region mostly shared haplotypes with one another, with only 7.7% of haplotypes shared with populations on islands in the South Indian Ocean. We found no common haplotypes between Macquarie Island (South Pacific Ocean), the South Atlantic Ocean, and South Indian Ocean islands.

**Animal tracking analyses**

We analysed track summaries, estimated move persistence, and predicted locations of the 12 seal samples from Davis Base (Table 4). Between 2005 and 2017, we estimated that the 12 tagged seals account for approximately 9.7% of the total moulting population (see Online Resource 1). Track summaries indicated that the average maximum displacement of the seals from Davis Base was 1330 km, with 44_SES travelling the greatest distance (3056 km). The average maximum displacement scaled by duration was found to be 8.1 km day\(^{-1}\), with the greatest being 14.5 km day\(^{-1}\) (62_SES), and the smallest being 1.30 km day\(^{-1}\) (seal 60). The average path tortuosity was found to be 0.078, with 58_SES having the highest path tortuosity (0.188), and 44_SES and 54_SES with the lowest (0.020 and 0.021, respectively).

The sampled seals showed high move persistence during their outbound trips into open waters, and low move persistence when approaching areas with high sea-ice coverage or when returning to breeding and moulting areas (Fig. 3). Predicted locations have indicated visits to Kerguelen Islands, Peninsula Valdés, and Crozet Island, Kerguelen and Heard Islands stocks, but excluding South Georgia Island and the Falkland Islands. We also identified two distinct mtDNA lineages, representing three of the four main breeding stocks (Macquarie Island, South Georgia, and Kerguelen and Heard Islands stocks, but excluding Península Valdés). Our tracking data showed that all seals, despite their varied lineages, remained within the vicinity of Crozet Island, Kerguelen Islands, and Heard Island for the duration of the tracking study. Seals that showed short displacements and less move persistence travelled more frequently around areas of high ice concentration. Overall, our integrated genetic and telemetry analyses provided longer-term insights into the seals’ natal affiliations and migration strategies, and additional short-term information on their annual life-cycle movements and foraging ecology.

The genetic data from 12 SES collected at Davis Base suggested high nucleotide diversity and haplotypes shared with elephant seals from other breeding colonies within the South Atlantic Ocean, South Indian Ocean, and South Pacific Ocean. However, given the small sample size, our study might not be representative of the larger population of all male SES at Davis Base. Replication of this study with a larger sample size and analysing nuclear markers, such as microsatellites, to clarify male gene flow and levels of

**Table 4** Southern elephant seal track summaries by maximum displacement from the track start \((D_{\text{max}})\), tracking deployment days (duration), maximum displacement scaled by deployment duration \((D_{\text{max}} \times t)\), and path tortuosity \((\tau)\).

| Sample  | \(D_{\text{max}}\) (km) | duration (days) | \(D_{\text{max}} \times t\) (km day\(^{-1}\)) | \(\tau\) |
|---------|----------------|----------------|-----------------------------------|-------|
| 42_SES  | 634.6          | 143.0          | 4.4                               | 0.070 |
| 44_SES  | 3055.9         | 270.0          | 11.3                              | 0.020 |
| 46_SES  | 1013.3         | 74.0           | 13.7                              | 0.058 |
| 48_SES  | 2967.4         | 305.0          | 9.7                               | 0.044 |
| 50_SES  | 2584.3         | 246.0          | 10.5                              | 0.126 |
| 52_SES  | 691.6          | 59.5           | 11.6                              | 0.102 |
| 54_SES  | 251.0          | 57.5           | 4.4                               | 0.021 |
| 56_SES  | 2234.8         | 216.0          | 10.3                              | 0.048 |
| 58_SES  | 142.96         | 48.0           | 3.0                               | 0.188 |
| 60_SES  | 28.7           | 22.0           | 1.3                               | 0.144 |
| 62_SES  | 2205.9         | 152.5          | 14.5                              | 0.065 |
| 64_SES  | 147.0          | 60.5           | 2.4                               | 0.050 |
| Average | 1329.8         | 137.8          | 8.1                               | 0.078 |

**Discussion**

By combining genetic and animal tracking data, we determined the natal locations and at-sea movements of moulting seals at Davis Base, Antarctica. Our analysis of genetic data revealed a mixed sample of likely natal locations from the 12 Davis Base seals, including some of the main breeding stocks (South Georgia, Macquarie Island, and Marion Island) and two of the smaller breeding colonies (King George Island and the Falkland Islands). We also identified two distinct mtDNA lineages, representing three of the four main breeding stocks (Macquarie Island, South Georgia, and Kerguelen and Heard Islands stocks, but excluding Península Valdés). Our tracking data showed that all seals, despite their varied lineages, remained within the vicinity of Crozet Island, Kerguelen Islands, and Heard Island for the duration of the tracking study. Seals that showed short displacements and less move persistence travelled more frequently around areas of high ice concentration. Overall, our integrated genetic and telemetry analyses provided longer-term insights into the seals’ natal affiliations and migration strategies, and additional short-term information on their annual life-cycle movements and foraging ecology.
interbreeding would assist in describing more comprehensively the genetic make-up of this moulting population. Our mitochondrial evidence shows that one seal had natal source affinities with the Macquarie Island stock, approximately 4207 km away, whilst the other 11 Davis Base seals showed affinities with the South Georgia stock (South Georgia, Elephant Island, Falkland Island, and King George Island), and the Kerguelen and Heard Island stock (Heard Island and Marion Island).

Animal tracking analyses

The male seals from our study spent more time in areas of high ice concentration and typically involved intense search behaviours exemplified by highly tortuous movements. Similar patterns have been observed previously in male seals that adopted sea-ice foraging strategies rather than pelagic foraging (Labrousse et al. 2017; Rodriguez et al. 2017; Jonsen et al. 2019; Hindell et al. 2021). Moreover, the preferences of foraging locations along the Kerguelen Plateau were shown by seal 50_SES, which had less move persistence near Kerguelen Islands. Five other seal tracks (44_SES, 48_SES, 50_SES, 56_SES, and 62_SES) showed patterns of inter-island movements by outbound trips made towards the Kerguelen Plateau and Crozet Islands, which might indicate pelagic foraging trips away from the ice shelf areas. Similarly, seal 48_SES showed less move persistence and had the second-longest maximum displacement per day around Crozet Islands (which is approximately 6900 km from its natal affinity, Macquarie Island). This suggests a long-distance movement event to gain foraging (and potentially future breeding) advantage away from the seal’s natal location.

SES are philopatric to breeding and foraging sites, so long-distance movements are rare, particularly for a juvenile seal (Hindell and McMahon 2000; Reisinger and Bester 2010). Our findings show that the only two male juvenile seals, out of the 12 seals included in this study, remained close to the ice shelf region to forage close to Davis Base, which was indicated by the reduced move persistence. Seals that remain close to the ice shelf region might have easier access to sea-ice polynyas where their prey (myctophids and ice fish) may be more easily found (Labrousse et al. 2017). However, a previous study found that a juvenile female from Macquarie Island was sighted at Peter 1 Øy, which is approximately 5,200 km to the east (Hindell and McMahon 2000). This finding corresponds with that observed here of 48_SES (Macquarie Island natal affinity), which foraged around Crozet Islands, as discussed above. The parallel finding between this study and previous studies might indicate a lack of resource availability around Macquarie Island where the population has been in decline in recent decades (McMahon et al. 2005a), which could be forcing seals from this location to travel farther to find food.
Changes to foraging site and the consequences on populations

The Kerguelen Plateau is a popular foraging location for male SES. However, there are high levels of predation on SES here (Hindell et al. 2021), which might have caused terminations of seal tracking signals. The Kerguelen Plateau lies within the Antarctic Polar Front boundary, which is an area of elevated productivity for most Antarctic marine species due to the decrease in water temperature down to 2 °C at 200 m, and the distribution of water masses and associated abundance of primary producers (O’Toole et al. 2014; Cristofari et al. 2018). SES will often forage on the Antarctic Continental Shelf and Polar Front, and because they are a deep-diving species, sea-surface temperatures have less of an effect on their diving behaviours (Hindell et al. 1991; O’Toole et al. 2014). Alternatively, previous studies have found that seals have greater foraging opportunities in colder waters, particularly along the Antarctic shelf; temperatures slow down the movements of prey, allowing their capture with less energy expenditure from the seals (Bailleul et al. 2007).

Popular foraging locations around the Kerguelen Plateau, such as Crozet Islands and Kerguelen Islands, have been reported to show either a slight increase or a stabilization in SES population numbers over the last decade (Guinet et al. 2004). The stabilized population numbers might have been caused by the seals’ inter-island movements and foraging between islands in the Kerguelen province, including Marion Island, Crozet Islands, Kerguelen Islands, and Heard Island (Oosthuizen et al. 2011). However, despite showing philopatry to foraging sites, the seals’ foraging strategies and movements might shift based on the changes to oceanic conditions that will influence where resources are available (Bailleul et al. 2007). Moreover, poor foraging success by females has led to a decrease in first-year pup survival and would thus reduce reproductive success in populations such as Macquarie Island (Arnbom et al. 1997; McMahon et al. 2003; Clausius et al. 2017; Mestre et al. 2020). As oceanic conditions continue to change with climate, the entire marine ecosystem will also shift. Therefore, the lack of resources around Macquarie Island is likely to continue causing decreases in population size (McMahon et al. 2005a; Clausius et al. 2017).
Population declines impacted by climate change

Broad climate events such as El Niño can affect the oceanic structure of the Southern Ocean. The Southern Ocean ecosystem relies heavily on phytoplankton and krill abundance for other species’ survival, previously reported for Antarctic whales, King Penguins (Aptenodytes patagonicus), and SES (McIntyre et al. 2014; Cristofari et al. 2018; Bestley et al. 2020; Rogers et al. 2020; Agrelo et al. 2021; Volzke et al. 2021). Over the last four decades, the global SES population has seen dramatic declines in some populations due to the changes in food availability (McMahon et al. 2005a; Volzke et al. 2021). Populations of the four main breeding stocks, South Georgia, Kerguelen Islands and Heard Island, Macquarie Island, and Peninsula Valdés, have all decreased since the 1970s. However, presently all the populations barring the Macquarie population are either stable or increasing (McMahon et al. 2005b; Hindell et al. 2016). A more thorough understanding of seal foraging behaviour, the selection and variations in foraging sites, and how this is expressed and transmitted within a population is central to understanding how foraging site selection affects population growth and ultimately population viability.

In summary, we were able to identify the likely natal locations of the 12 Davis Base seals through genetic data. We combined findings from genetic data with satellite telemetry tracking data and identified that the majority of the seals are spending most of the time foraging along the Kerguelen Plateau. Our data suggest that a seal from Macquarie Island has travelled a long distance, probably to gain a foraging advantage over its conspecifics. Long-range migrations and movements to distant feeding grounds might be one way for seals to maximize foraging efficiency, which may affect population growth rates through changes in survival and reproduction.

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Author contributions CRM conducted Antarctic fieldwork and collected data. MC did the molecular lab work, conducted molecular and tracking data analyses, and wrote the first draft of the manuscript. JJ provided guidance on and contributed to tracking data analysis. SYWH, CRM, and MdB supervised the research project. All authors revised and contributed to the writing of the final manuscript.

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Data availability The data sets generated during and/or analysed during the current study are available from the corresponding author on reasonable request. DNA sequences have been deposited in GenBank, with accession numbers ON571492–ON571503.

Declarations

Conflict of interest The authors declare that they have no conflicts of interest.

Ethical approval The animal capture, handling and tag deployment procedures were reviewed and approved by Macquarie University under permit number: ARA 2014_057.

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