**ABSTRACT**

The elusive nature of *Trichomonas vaginalis*, the most common, non-viral, sexually transmitted pathogen has hampered our knowledge of its significance for human health for over 150 years. The combination of epidemiology, molecular cell biology, immunology and more recently genomics and other allied omics data, are all contributing at shedding new light onto what is increasingly recognised as a significant human pathogen leading to important health sequelae due to multifaceted interactions with its human host, the human microbiota, bacterial pathogens and viruses. The integrations of these various data are contributing in important ways to refining our understanding of the parasite pathobiology and virulent factors. Indeed, it is increasingly recognised that to rationalise the development of effective prophylactic and therapeutic treatments for human pathogens it is important to integrate the broadest possible spectrum of human-microbial-parasite-virus interactions in relation to qualitative and quantitative variations in the human innate and adaptive defence responses. This short review aims at providing an integrative overview of *T. vaginalis* virulent factors by taking into account the importance of the human-microbiota-parasite-virus interplay in human health. It also highlights selected cellular characteristics of the parasite often overlooked in the biological and medical literature.

**INTRODUCTION**

Ever since its discovery in 1836 by the French medical doctor and microbiologist, Alfred F. Donné, the sexually transmitted obligate extracellular mucosal parasite *Trichomonas vaginalis* (*TV*) has progressively grown, rather sluggishly, from a perceived insignificant commensal to an important pathogen, inducing significant health sequelae in both men and women, and with adverse pregnancy outcomes. Some of the most recent developments in our understanding of *TV* pathobiology include its epidemiological association with HIV over and above high-risk members of the population in terms of exposure to sexually transmitted infections (STI), and a positive association with aggressive prostate cancers. The dramatic impact on human health of the HIV pandemic and resource-limiting conditions experienced by hundreds of millions of people worldwide have combined in highlighting the pathogenic significance of this common human parasite—one that is facilitating the spread of HIV and is associated with a number of reproductive complications including sterility, preterm birth, underweight newborn babies and cervical cancer. Significantly, *TV* infects a large fraction of individuals, in particular men, without overt symptoms, complicating its diagnosis and control.

Furthermore, and particularly relevant in the context of a discussion on *TV* virulent factors, the parasite’s elusive nature also renders the characterisation of the molecular and cellular basis of its pathobiology less straightforward, as highlighted for similarly elusive bacterial pathogens. Indeed, there is growing recognition that the outcome of human–symbiont interactions (encompassing mutualists, commensals and parasite/pathogens) cannot be fully understood without considering the host immunological status and host-associated microbiota. There is a need to integrate the full range of human-microbe interactions in all its diversity to comprehend their different contributions and influence on each other, and how these impact both positively and negatively on human health. These considerations also underscore the limitations of the reductive approach implied by the original four Koch postulates and their molecular children, used to characterise pathogens and their virulence factors. Incorporating the host response to exposures to microbes is now considered to be an integral part of defining pathogens and their virulence factors.

Indeed, these issues are increasingly recognised by the research community working on *TV* as illustrated in a number of recent reviews. Hence, an integrative overview of *TV* virulence factors is presented here, acknowledging the complex host-microbiota-parasite-virus interplay in influencing the outcomes of human-*TV* interactions.

**CELLULAR AND MOLECULAR BASIS OF TV VIRULENCE**

A number of recent reviews have discussed various aspects of well-characterised and more speculative virulence factors of *TV*.

Only a selection of *TV* virulence factors will be considered here and integrated with some of the most recent data on *TV* molecular cell biology and human innate and adaptive responses to *TV*. Some important gaps in our knowledge of *TV* pathobiology will also be highlighted. The evolutionary origin of the best-studied organelle of the parasite, the hydrogenosome, and the roles of hydrogenosomal enzymes in the parasite’s physiology and as a drug target were recently reviewed and are not covered here.

*TV* is a flagellated microbial eukaryote known to exist in several cellular forms (figure 1). The two best characterised forms are the trophozoite, a free-swimming, flagellated, pear-like cell, and an amoeboid form, with a pancake shape characterised by an important increase in surface contact; this is rapidly induced upon trophozoite contact in vivo with epithelial cells from the vagina, cervix, urethra, prostate and extracellular matrix (ECM) proteins, allowing for the infection to occur.
Various cellular forms of *Trichomonas vaginalis* (TV). The TV trophozoite is characterised with four anterior and a fifth recurrent flagella. Upon contact with human tissue, TV can rapidly differentiate into an amoeboid form. In vitro trophozoites can differentiate into pseudocysts upon exposure to cold. Pseudocysts are spherical cells without visible flagella (they are internalised), they can divide and redifferentiate into trophozoites. It is not known if amoeboid form can differentiate into pseudocysts (?). Trophozoites are also known to form large cellular aggregates, in a process called swarming. Such groups of cells are thought to have stronger binding and cytotoxicity properties towards human tissues, and could also represent a form of defensive reaction to the host immune responses.

Trophozoites are typically considered as the infective form. A third cellular form called pseudocysts can be induced in vitro upon exposure of trophozoites to cold and other stressors. However, the significance of this form during the infection process is currently unknown. Data for the related bovine parasite *Trichomonas foetus*, which leads to various reproductive complications including abortion and sterility, suggest that pseudocysts are highly relevant corresponding to aggressive forms facilitating transmission to, and the initial colonisation of, a new host. Additionally, aggregates of parasites are also known to form (figure 1). Very little is known concerning the triggers and cellular signalling that orchestrate TV differentiations into these different forms. Identifying specific molecular markers for the different TV cellular forms and aggregates will be an essential prerequisite to investigate their relevance during the infection process. The draft genome sequence and its annotation represent an invaluable resource to investigate the molecular cell biology of TV during the infection process. The draft genome sequence and its annotation represent an invaluable resource to investigate the molecular cell biology of TV by providing specific molecular leads and allowing comparative transcriptomics and proteomics investigations. An initial study of the phosphoproteome of TV trophozoites, amoeba and pseudocysts grown in vitro suggested differential protein phosphorylation profiles, consistent with specific signalling mechanisms occurring in the different cellular forms of the parasite that are induced upon specific environmental triggers.

Clinical isolates of TV have been accumulated over the years from many regions of the world. Comparisons of their capacity to bind and kill human (and other species) cell lines in vitro have demonstrated important variations between TV isolates. Accordingly, these isolates are considered important virulent traits for TV. One of the currently best-characterised adhesins mediating parasite binding to host tissue are TV lipoglycans (TvLG), the most abundant surface molecules of the parasite. Investigating TvLG also led to the identification of galectin-1, the only identified human receptor for TV so far. TvLGs are also known to modulate inflammatory responses of epithelial cells and macrophages. A proteomics survey of TV surface proteins identified a total of 411 proteins, confirming a number of in silico-predicted surface proteins, and also a host of novel and important candidate virulent factors. By contrasting strains with low versus high level of adhesion to vaginal epithelial cells (VECs) in vitro, 11 proteins were shown to be more abundant on the cell surface of the highly adhering strains. These include proteins annotated as hypotheticals—which have no sequence similarity to any other proteins in protein databases. A high level of expression of two of these hypothetical proteins increased the binding to VECs of a poorly binding TV strain. Other novel candidate cell surface virulence factors identified in this proteomics survey included three tetraspanins, which are membrane proteins involved in signalling modulating adhesion, motility and tissue invasion in other systems; all these are key processes underlying TV pathobiology. The characterisation of one TV tetraspanin demonstrated that its expression and cellular localisation was modulated upon TV binding to VEC, and that it plays a role in regulating migration of the parasite through a surrogate ECM gel, strongly supporting tetraspanins as important TV virulence factors. These studies highlight how little we know about key aspects of TV biology, and how genomics and allied omics provide important tools to investigate host-parasite interactions.

### TV Interactions with Human-Associated Bacteria and Viruses

The phagocytosis of human microbiota by TV, including bacteria and fungal cells, has the potential to induce dysbiosis...
Table 1 Example of Trichomonas vaginalis (TV) candidate transmembrane proteins potentially mediating endocytosis*

| Protein family | RefSeq accession | Protein length | [FY]NPX(FY) motif | Acidic cluster |
|----------------|------------------|----------------|-------------------|----------------|
| BspA           | XP_001308117     | 735            | NENPIF           | DDPFAADFMDS    |
| BspA           | XP_001584393     | 1092           | FTNFLP           | DDPFDSDPDp     |
| BspA           | XP_003121489     | 600            | YDNPFF           | DDPFDSDQND     |
| BspA           | XP_003121493     | 582            | YDNPFF           | DDPFDSDQND     |
| BspA           | XP_003121496     | 594            | FDNPFL           | DDPFDSDQNE     |
| BspA           | XP_003121499     | 497            | FDNPFL           | DDPFDSDQNE     |
| BspA           | XP_003121508     | 733            | FDNPFL           | DDPFADEEFEK    |
| BspA           | XP_003123867     | 549            | FSNPFL           | EDPPADDEEEG    |
| BspA           | XP_003123873     | 1142           | YINPFL           | DDPFDSDEDH     |
| BspA           | XP_003123876     | 834            | FTNFLP           | DDPFDSDDEE     |
| BspA           | XP_003132366     | 703            | ADNPIF           | DDPFAQFDNDD    |
| Pmp            | XP_001579443     | 593            | NDNPFL           | DDPFRKDEFE     |
| Pmp            | XP_003133544     | 742            | DNPFLP           | DDPFRKDEEV     |
| Pmp1           | XP_00325298      | 624            | NDNPFL           | DDPFRKDEE     |
| Pmp1           | XP_003130614     | 569            | IDNPFL           | DDPFRKDEE     |
| Pmp1           | XP_003158285     | 712            | NDNPFL           | DDPFRKDEE     |
| Pmp1           | XP_003130974     | 812            | NDNPFL           | DDPFRKDEE     |
| Pmp1           | XP_00328616      | 605            | YTNPFL           | DDPVDQFEOQ     |
| Pmp1           | XP_003158104     | 680            | NDNPFL           | DDPFRKDEE     |
| Pmp1           | XP_003252882     | 803            | NDNPFL           | DDPFRKDNFEI    |
| Pmp1           | XP_00325146      | 593            | QENPFL           | DDPFDQFEOQ     |
| Pmp1           | XP_003203030     | 526            | NVNPFL           | DDPFRDDEE     |
| Pmp1           | XP_00320335      | 1819           | NDNPFL           | DDPFRNDEE     |
| Pmp1           | XP_003129921     | 652            | NENPFL           | ENPLFNDDE     |

*Entries with at least one inferred transmembrane domain from two distinct TV protein families (BspA: Bacteroides surface protein A-like extracellular domain and Pmp: Chlamydia polymorphic membrane protein-like extracellular domain) but with a shared cytoplasmic tails possessing two motifs ([FY]NPX(FY) and acidic clusters) representing potentially signals for endocytosis—for details see ref. 29.†Entries supported by cell surface proteomics.22

Table 2 List of enzymes involved in glycan metabolism derived from bacterial lateral gene transfers (LGTs)*

| EC number | Enzyme name | RefSeq accession | Nearest neighbour in phylogeny |
|-----------|-------------|------------------|-------------------------------|
| 3.2.1.18  | Exo-α-sialidase | XP_001319692 | Bacteroidetes                 |
| 3.2.1.23  | β-galactosidase  | XP_001581038 | Bacteroidetes                 |
| 3.2.1.52  | β-N-acetylhexosaminidase | XP_00322989 | Bacteroidetes                 |
| 3.2.1.24  | α-mannosidase   | XP_001579222 | Prokaryotes                   |
| 3.2.1.25  | β-mannosidase   | XP_001322689 | Proteobacteria                 |
| 3.2.1.51  | α-fucosidase   | XP_001316088 | Bacteria                      |
| 4.1.3.3   | N-acetylneuraminic lyase | XP_00323296 | Pasteurellaceae               |
| 5.1.3.8   | Acylglucosamine | XP_001308218 | Bacteroidetes                 |
| 3.2.1.45  | Glucosylceramidase | XP_001279673 | Bacteroidetes                 |

*For details about inference of LGT methodology and gene and enzyme characteristics see ref. 26.†Note that some of the listed enzymes could target O-glycans, N-glycans and/or gangliosides.‡EC enzyme commission number.

(imbalanced microbial community?) in the human microbiota, for example, as observed during bacterial vaginosis (BV). Consistent with this possibility, a correlation between the presence of TV and a vaginal microbiota characterised with low abundance of protective lactobacilli and higher proportions of Mycoplasma, Prevotella and other bacteria typically observed in BV, has been observed.9 Future work will be required to establish the temporality and causality between TV and BV and their contribution (possibly synergistic) to increase host susceptibility to HIV.9 In addition, antibiotics can significantly disturb the microbial community7 and facilitate HIV transmission through damaging mucosal surfaces, and their consequence of studying a pathogen in the broader context of human–microbes interactions.7

TV has been shown to form a symbiosis with Mycoplasma hominis in in vitro cultures, and Mycoplasma cells carried by TV could infect human cells, suggesting that TV could be a Trojan horse for the bacterium.28 Intriguingly, coinfections with TV-Mycoplasma were shown to consume larger amounts of free arginine than TV alone, which could contribute to reducing nitric oxide production by macrophages through depletion of free arginine in the vagina, thus potentially interfering with an important host defence mechanism.25 Another likely consequence of TV actively feeding on host microbiota, is the acquisition of a number of bacterial genes through lateral gene transfer (LGT). Of particular interest is an almost complete set of enzymes of bacterial origin capable of degrading glycans, and mostly acquired from members of the bacteroidetes,26 one of the most abundant bacterial lineages of mucosal microbiota and known to encode a rich set of glycan-targeting enzymes.27 (Table 2). These TV enzymes of bacterial origins are thought to play important roles in degrading mucins and other human glycans, including those from the glycocalyx epitheial cells, which represent the initial barrier the parasite needs to deal with to be able to colonise the host mucosal surfaces.28 Glycan degradation is also likely to provide an important source of energy for the parasite.

Tissue damage and inflammatory responses due to TV infections are thought to facilitate HIV entry and, in TV-HIV dually infected patients, to stimulate HIV production.1 Additionally, TV can also internalise human viruses through endocytosis.11 Viruses carried by TV could contribute to their transmission to a new host, as viruses internalised by TV were shown to be infectious for human cells for 2–6 days. The viruses are either released upon parasite death or secreted from TV-loaded viruses through the recycling route of the endocytic pathway.11 However, there is no evidence that any human virus can replicate in TV. Hence, as suggested for Mycoplasma, TV could be a Trojan horse for HIV and possibly other viruses.

These different considerations clearly illustrate the importance of studying a pathogen in the broader context of human–microbes interactions (figure 2).7

TV VIRUSES

Adding to its capacity to physically interact with human viruses and facilitate HIV transmission through damaging mucosal surfaces, TV can be infected by four dsRNA viruses (TVV) members of the Totiviridae, thought to be transmitted mainly vertically.29 Various data suggest, collectively, that TVV modulate the parasite virulence by influencing TV gene expression; however, additional studies are required to firmly establish this fascinating possibility.29 Two recent developments are of particular interest in relation to a potential role of TVV in TV pathobiology. A study investigating TV genetic diversity and population structure identified two major genotypes for the

*Example of transmembrane proteins potentially mediating endocytosis*

†Entries with at least one inferred transmembrane domain from two distinct TV protein families (BspA: Bacteroides surface protein A-like extracellular domain and Pmp: Chlamydia polymorphic membrane protein-like extracellular domain) but with a shared cytoplasmic tails possessing two motifs ([FY]NPX(FY) and acidic clusters) representing potentially signals for endocytosis—for details see ref. 11.‡Entries supported by cell surface proteomics.22
The pathobiology of *Trichomonas vaginalis* (TV) is multifaceted involving direct and indirect interactions with host tissue, bacteria, human viruses and *Trichomonas* viruses.

- The genome sequence data of TV is being exploited to facilitate the study of the molecular basis of the parasite pathobiology.
- The immune response to TV is still poorly understood, and it will be essential to study its variations to rationalise the outcome of host–parasite interactions.

**Key messages**

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