Adenoviruses-mediated RNA interference targeting cytosolic phospholipase A2α attenuates focal ischemic brain damage in mice

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Abstract. Cerebral ischemia injury is a clinical, frequently occurring disease, which causes a heavy burden on society and families. It has been demonstrated that cytosolic phospholipase A2α (cPLA2α) is significant in neurological injury caused by ischemic brain injury, and inhibition of cPLA2α may reduce stroke injury. In the present study, the role of cPLA2α was investigated in a mouse model of middle cerebral artery occlusion and/or reperfusion (MCAO/R) using an effective cPLA2α inhibitor and adenoviruses-mediated RNA interference. The most effective recombinant adenovirus encoding cPLA2α, small interfering RNA (pAd-siRNA-cPLA2α) was constructed and selected. MCAO/R surgery is used to construct the model of focal ischemic brain damage in mice. Adenoviruses-mediated RNA interference targeting cPLA2α was administered by stereotactic surgery 2 h before the MCAO/R. The expression/activity of cPLA2α and cPLA2α-derived injurious lipid mediators was assessed. pAd-siRNA-cPLA2α-treated animals (RNA interference; RNAi group) were compared with pAd-siRNA-control-treated animals (negative group) with regard to neurological deficit, motor function, pathological changes, apoptosis, and infarct volume. The RNAi group animals reduced the expression level of cPLA2α, as determined by western blotting and reverse transcription-quantitative polymerase chain reaction, the improvement of locomotor function was evaluated by rotarod test, and the decrease of apoptosis was evaluated by terminal deoxynucleotidyl transferase dUTP nick end-labeling staining. The decreased infarct areas were evaluated by 2,3,5-triphenyltetrazolium chloride staining. The expression levels of prostaglandin E2, leukotrienes B4, lysophosphatidylcholine and free fatty acids were reduced in the RNAi group when compared with the negative control group. Thus, the data indicates that the expression level of cPLA2α was effectively controlled by pAd-siRNA-cPLA2α treatment. pAd-siRNA-cPLA2α treatment, in reducing the levels of inflammatory factors, neurological deficit and tissue damage, represents an effective potential therapeutic strategy. pAd-siRNA-cPLA2α reduces cPLA2α expression levels with long-term efficacy, thereby improving functional deficits and effectively attenuating ischemic brain damage. Thus, pAd-siRNA-cPLA2α shows potential value for therapeutic evaluation in ischemic brain damage.

Introduction

Ischemic brain damage has long been associated with neuronal injury (1-4), and it is a result of combined action of acute necrosis with delayed injury mechanisms. The damage cascade is the recognized theory of the cerebral ischemia physiological and pathological mechanism, predominantly including excitotoxicity, peri-infarct depolarization, inflammation and apoptosis (5). Metabolism of arachidonic acid (AA) and its metabolic products have been implicated to be associated with these pathophysiological mechanisms (4). To a great extent, cellular levels of AA are under the control of phospholipase A2 (PLA2) enzymes. PLA2 catalyze the membrane phospholipid hydrolysis, and the resulting released AA is metabolized into eicosanoids. PLA2 is a family of esterases that catalyze the hydrolysis of phospholipids in the sn-2 position, and it is the crucial enzyme of AA, prostaglandin (PG) and platelet activating factor (PAF) (6). Previous studies have indicated that PLA2 is crucial in ischemic cell death (1,7,8).

Almost all human cells contain PLA2, and numerous isotypes have been identified. There are three well known major subfamilies: Secretory PLA2α, Ca2+-independent PLA2, and cytosolic Ca2+-dependent PLA2 (cPLA2α). The cPLA2α (cPLA2α) form is a member of the cPLA2 class, and has unique characteristics, including its preference to catalyze the hydrolysis of sn-2 position of phospholipid substrate molecules to AA. Numerous studies demonstrated that cPLA2α is a key component associated with the development of stroke injury (2,3,9,10), and it has been shown that cPLA2α has a marked role in neurological injury caused by ischemic brain injury, and the inhibitor of cPLA2α may reduce stroke injury (11).

RNA interference (RNAi) is an experimental technique with a wide range of applications in gene function research and gene therapy, and there is increasing focus on developing...
therapeutic strategies based on RNAi (12). Since the discovery of mammalian RNAi, >50,000 studies have described the use of small interfering RNA (siRNA) (13). RNAi is a gene silencing mechanism triggered by double-stranded RNA with 9-23 base pairs, it activates an RNA-induced silencing complex (RISC) and guides the degradation of homologous RNA by functional RISC (13). Compared with other antisense strategic tools, RNAi is a perfect technique for gene silencing with sensitivity and specificity (14).

Silencing of target genes in dividing and non-dividing cells is effectively via RNAi technology (15). In mammalian cells, RNAi efficiently blocks specific gene expression to treat diseases caused by excess protein expression. Due to the poor efficiency of retrovirus vectors and liposomes in transfection, adenovirus-mediated gene transfer was developed (16). The transfection capacity of adenovirus vectors was identified to be highly effective in vivo and in vitro. The expression of foreign genes in brain tissues peaked 7 days after transfection and terminated after 2 weeks (17). It has been demonstrated that a low degree virus, limited within a certain range, prevents the body's immune response. Low dosage adenovirus transfection to ischemic brain tissue with is efficient and stable with low toxicity. To the best of our knowledge, there are no studies reporting the effects of cPLA2α RNAi on the development of ischemic brain damage in an in vivo animal model. Therefore, the present study employed the adenovirus-delivered RNAi technique to evaluate the effects of cPLA2α knockdown in vivo in a mouse model, and hypothesized that it may be used in further adenovirus intervention experimental research.

Materials and methods

Cell culture. L929 and HEK293 cells were purchased from Shanghai Bioleaf Biotech Co., Ltd. (Shanghai, China). Cells were maintained in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (both from Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) and 100 U/ml penicillin/streptomycin (AppliChem GmbH, Darmstadt, Germany). At confluency, cells were split 1:3. Cultures were incubated at 37°C in a humidified atmosphere with 5% CO₂. The growth media was replaced every other day. The engineered, stable L929-cPLA2α-siRNA cells were maintained in the above-mentioned conditions. The L929 cells stably expressed wild-type human cPLA2α.

Reagents and siRNA. If not otherwise stated, reagents were supplied by Sigma-Aldrich (Merck KGaA, Darmstadt, Germany). Arachidonyl trifluoromethyl ketone (ATK), an inhibitor of cPLA2α, was purchased from Cayman Chemical Co. (Ann Arbor, MI, USA). The siRNAs targeting the mouse cPLA2α gene and the sequences of RNAi were designed and synthesized by Guangzhou Ribobio Co., Ltd. (Guangzhou, China). Three gene sequences were selected for RNAi, which were verified by experiments to suppress the expression of mouse cPLA2α in L929 cells (18). The siRNA sequences (sense strand) were as follows: No. 1, 5'-GGCGAAC GAGACCTTCAAT-3'; no. 2, 5'-GGTGCTAATCCTCAC GTG-3'; no. 3, 5'-GACACCTTGAGAATGAC-3'. 5'-TTC TCCGAACTGTCACT-3' served as a negative control. A BLAST search on the NCBI website was used to confirm that the sequences did not have homology to any other mouse genes.

**Generation and appraisal of recombinant adenovirus encoding cPLA2α siRNA (pAd-siRNA-cPLA2α).** To construct a plasmid expressing siRNA/cPLA2α, the target sequences were subcloned into shuttle vector, pShuttle-H1, then sequencing was performed by Thermo Fisher Scientific, Inc. (19). The replication-deficient adenovirus that contained the full-length cDNA of the RNAi sequence was generated by homologous recombination through co-transfection of plasmid, pShuttle-H1-RNAi with pAdEasy into HEK293 cells that were grown in DMEM to 70% confluence in 60 mm wells using DOTAP liposome reagent. The cells were incubated for 48 h at 37°C. Following several rounds of plaque assay for purification (20), the adenovirus was purified from cell lysates via banding twice in CsCl density gradients. The harvested viral supernatant was desalted and added to phosphate-buffered saline (PBS) containing 10% glycerol (v/v). The infectious titer was determined via a standard plaque assay and the titers averaged 2×10⁷ pfu/ml. The mRNA and protein expression levels were detected by reverse transcription-quantitative polymerase chain reaction and western blot analyses.

**RNAi target screening.** At 24 h prior to transfection, L929 cells (90% confluent) were digested for 5 min at 37°C with trypsin/EDTA solution (Invitrogen; Thermo Fisher Scientific, Inc.). The cells were inoculated in 12-well tissue culture plates with 1.5 ml cell culture medium (DMEM containing 10% FBS and 100 U/ml penicillin/streptomycin) and 5×10⁵ cells/well, and incubated in culture medium overnight at 37°C. The cells were transfected with Ad-siRNA-cPLA2α or negative control (NC) for detecting the efficiency of silencing. The transfection efficiency of the adenovirus had plateaued at 7 days post-transfection, and cPLA2α protein and mRNA expression levels were analyzed by western blotting and RT-qPCR 4 days after transfection. All transfections were performed in triplicate.

**Animals and treatment.** Experimental mice (male C57BL/6 mice, aged 12-16 weeks) were purchased from Better Biotechnology Co., Ltd. (Jiangsu, China). The mice were housed in a constant-temperature facility with a 12-h dark/light cycle and given access to water and food freely. All procedures performed with mice were according to the National Institutes of Health Guide for the Care and Use of Laboratory Animals (21), and the experimental procedures were approved by the Ethics Committee of Hebei, Cangzhou Central Hospital (Cangzhou, China).

The mice were randomly allocated to the sham, middle cerebral artery occlusion and/or reperfusion (MCAO/R), NC and RNAI groups. At 2 h before unilateral ischemia, freshly prepared pAd-siRNA-cPLA2α and pAd-siRNA-control were injected into the RNAi and NC animals, respectively. The sham and MCAO/R group animals received only the carrier solution. Mice were sacrificed by cardiac puncture following anesthesia by intraperitoneal injection of pentobarbital sodium (45 mg/kg) 2 h, 1, 3, 7 or 14 days post-surgery. Blood (500 μl) was collected by direct cardiac puncture and indomethacin (10 μM) was added to inhibit PG synthase activity.
immediately after the blood was drawn. Brains were removed from the skull and prepared for the experience. Brain tissue samples from each group were divided into two parts, some (n=3 of each group) were fixed in 4% paraformaldehyde in PBS, and the others (n=3 of each group) were stored at -80°C until use. At 14 days post-surgery, the mice (n=3 from each group) were anesthetized and perfused through the left ventricle with 0.01 M PBS. Brains were removed from the skull and the rhinencephalon, cerebellum and brain stem were removed. The coronary brain regions of brain tissue samples were divided into slices (~2 mm thick) and prepared for triphenyltetrazolium chloride (TTC) staining.

**Stereotactic surgery.** Prior to the MCAO/R surgery, animals were anaesthetized with 2% halothane in a mixture of 60% N2O and 40% O2 using a face mask and fixed in stereotactic apparatus, where a longitudinal incision was made in the middle of the head. The skull was drilled into and the epidural thorn was opened. A microsyringe (World Precision Instruments, LLC, Sarasota, FL, USA) was used to slowly puncture to a corresponding depth (0.4-0.6 mm), and pAd-siRNA-cPLA2α and pAd-siRNA-control, were injected at a constant slow speed (200 nl/min; dose, 200 nl). The wounds were stitched following surgery.

**Focal cerebral ischemia model.** The transient focal ischemic brain damage in mice was performed using the intraluminal occlusion technique, as described previously (22). Occlusion of the right MCA was removed after 1 h, and the surgery in the sham group mice was performed through a vertical cervical incision. Thermostatically controlled heating pads and a heating lamp were used to maintain a normal body temperature until the mouse recovered from anesthesia. In addition, a laser Doppler perfusion monitor was used to monitor the regional blood flow. The cerebral blood flow (CBF) measurements in the mice were performed with the probe affixed with glue to the surface of the cerebral cortex perfused by the MCA.

Neurological deficit in the mice was evaluated using a 5-point neurological deficit score (NDS) following surgery and prior to euthanasia. Neurological impairment was assessed using the scores between 0 and 4 as follows: 0, no neurological deficit; 1, forelimb weakness; 2, circling to the affected side; 3, falling to the affected side; and 4, unable to walk spontaneously (23). Animals that were scored 1-3 following reperfusion were used for subsequent experiments, and the required number of mice were randomly selected to the experimental group. The mice were scored on day 1, 3, 7 and 14 day after reperfusion.

**Evaluation of locomotor function.** The rotarod test was used to analyze the motor coordination and anti-fatigue characteristics of the mice. The rotarod machine (Med Associates, Inc., Fairfax, VT, USA) adjusts the initial rotating and accelerating speeds. The detecting parameters in the present study were the latency time to the first fall and the number of falls in 5 min. The experimental mice were trained at 2-20 rpm in the days prior to commencing testing. Assessment of their motor ability was performed 1 day before surgery and on days 1, 3, 7 and 14 after surgery.

**Enzyme-linked immunosorbent assay (ELISA).** The levels of leukotriene B4 (LTB4) and prostaglandin E2 (PGE2) were examined using commercially-available ELISA kits (cat nos. 10009292-96S and 500141-96; Cayman Chemical Co.). The experiment required 50 μl serum samples from the mice, and all procedures were performed according to the manufacturer's instructions.

**High-performance thin-layer chromatography (HPTLC).** Free fatty acids (FFAs) and lysophosphatidylcholine (LPC) were determined and quantified using HPTLC. The required lipids samples were extracted from the brain tissue samples using the Folch method (24). FFA and LPC levels were determined and quantified using HPTLC plates according to previously described methods (25,26).

**Histological examination.** The tissue samples were fixed and embedded in paraffin (4 µm sections) and processed in an alcohol series (50-100%), stained with hematoxylin for 10 min and eosin for 5 min at 25°C (H&E). The stained sections were observed under inverted microscope (magnification, x100) for histological examination.

**Terminal deoxynucleotidyl transferase dUTP nick end-labeling (TUNEL) staining.** To evaluate the degree of apoptosis, TUNEL staining was performed on brain sections using a TUNEL Apoptosis Assay kit (cat. no. 11684795910; Roche Diagnostics, Basel, Switzerland). The tissue samples were fixed in 4% paraformaldehyde for 24 h at 4°C, paraffin-embedded, dewaxed and the TUNEL assay was performed. All procedures were conducted according to the manufacturer's instructions. DAPI Fluoromount-G™ anti-fluorescence quenching sealant (cat. no. 36308ES11; Shanghai Yeasen Biotechnology Co., Ltd., Shanghai, China) was used to seal slices. The observed results were recorded using a fluorescence microscope equipped with AxiowVision software (version 4.1; Carl Zeiss AG, Oberkochen, Germany). Four brain samples from each experimental group were used for the analysis. At least four microscopic fields within the ischemic penumbra of each brain sample were imaged, and a double-blinded manner was used to count the TUNEL-positive cells. The mean value of the TUNEL-positive cells counted from these areas was calculated to represent each brain section.

**Western blotting.** Total proteins were extracted from the cells or frozen tissue samples using radioimmunoprecipitation assay lysis buffer (Sigma-Aldrich; Merck KGaA). The brain tissue lysates were prepared as described previously (27). Briefly, brain tissue samples were homogenized, centrifuged and the supernatants were collected. The Bradford method was used to determine the protein concentrations (28), and bovine serum albumin standard protein served as the standard. Western blotting was performed according to standard protocols. The proteins (40 μg) were separated by 12% SDS-PAGE, then transferred to a polyvinylidene fluoride membrane and blocked with 5% non-fat dried milk at 37°C for 2 h. Membranes were incubated with the following primary antibodies for 16 h at 4°C: Anti-cPLA2α antibody (cat. no. sc-137069; 1:200; Santa Cruz Biotechnology, Inc., Dallas, TX, USA) and anti-β-actin antibody (cat. no. sc-8432; 1:200; Santa Cruz Biotechnology, Inc., Dallas, TX, USA) and anti-β-actin antibody (cat. no. sc-8432; 1:200; Santa Cruz Biotechnology, Inc., Dallas, TX, USA).
RNA isolation and RT-qPCR. Total RNA extraction, and RT-qPCR were performed as previously described (29). Total RNA was isolated from the tissue samples or cells using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's instructions. First-strand cDNA was synthesized using PrimeScript reverse transcriptase (Takara Biotechnology Co., Ltd., Dalian, China) and oligo (dT). Following RT, qPCR was performed according to the manufacturer's instructions of the SYBR Premix Ex Taq™ II PCR kit (Takara Bio, Inc., Otsu, Japan). The thermocycling conditions were as follows: 94°C for 3 min, followed by 40 amplification cycles of 30 sec at 94°C, 30 sec at 58°C and 30 sec at 72°C. The relative expression levels of cPLA2α mRNA were calculated using SYBR green on the ABI Prism 7300 Sequence Detection System (Applied Biosystems; Thermo Fisher Scientific, Inc.) using β-actin as the endogenous reference gene amplified from the samples. The relative amounts of mRNA were calculated using the 2^{△△Ct} method (30). The primers were as follows: cPLA2α forward, 5′-GGTGGGAGAGAGAAAGATGC-3′ and reverse, 5′-AGGGAATTTGAGATGGACC-3′; β-actin forward, 5′-GCTATGCTCTCTCCACGCCT-3′ and reverse, 5′-TCACGGACGATTTCCTCAG-3′.

TTC staining. The infarct volume was evaluated by TTC staining as previously described (31). The 2 mm brain sections were incubated with 2% TTC at 37°C for 30 min with gentle shaking, then were fixed with 10% formalin in PBS. The stained slices were photographed, and the size of the infarct volume was determined by subtracting the area of the non-infarcted ipsilateral hemisphere from that of the intact contralateral hemisphere. The percentage of infarct volume was calculated by dividing the sum of the area from all sections of infarction by the total of that of contralateral hemisphere to avoid the influence of tissue edema (32).

Statistical analysis. All values are presented as the mean ± standard deviation. Each experiment was repeated at least three times. Statistical analysis was performed by comparing two groups using Student's t-test and P<0.05 was considered to indicate a statistically significant difference.

Results

Selection of RNAi sequence for knockdown of mouse cPLA2α in L929 cells. We examined the inhibitory effects of the three RNAi sequences for mouse cPLA2α in L929, and attempted to establish the most effective silencer. ATK effectively inhibited cPLA2α in vitro and in vivo (33-37). The treatment of L929 cells with 10 nM ATK every 24 h served as a positive control. The level of cPLA2α in L929-cPLA2α-RNAi cells was examined by qPCR and western blotting. The effects of three predesigned RNAi sequences for mouse cPLA2α (silencer nos. 1-3) on cPLA2α protein and activity (Fig. 1) were confirmed. Treatment with silencer-2 or -3 decreased the expression level and activity of cPLA2α markedly, while Silencer-1 only exerted a partial inhibitory effect. Results showed that cells under silencer-3 treatment almost entirely inhibited the expression level of cPLA2α protein without altering β-actin expression. These results indicate that the Silencer-3 sequence is an effective RNAi sequence for inhibiting mouse cPLA2α expression. Subsequently, the most effective RNAi sequence was used to knockdown the expression of cPLA2α in mice.

Neurological function and motor function evaluation in animals following MCAO/R surgery. To investigate the role of cPLA2α in focal ischemic brain damage, MCAO/R surgery was performed on male C57BL/6 mice (age, 12-16 weeks). Neurological deficits of these mice were assessed and 2 h after reperfusion, all mice exhibited neurological injury. Neurological deficit was evaluated again on days 1, 3, 7 and 14 after reperfusion. The RNAi group demonstrated no significant difference compared with the NC group and MCAO/R group at 2 h post-surgery (P>0.05); however, the NDSs of the RNAi group animals were significantly lower than those of the control group mice at 7 and 14 days after stroke (Fig. 2B; P<0.01).

Ischemic brain damage patients are prone to motor function impairment, losing partial or complete motor ability...
To monitor the motor function change of mice following ischemic stroke, rotarod tests were performed at 1, 3, 7 and 14 days after ischemic stroke surgery. Prior to surgery, the mice demonstrated no differences during the rotarod test (Fig. 2). However, the number of falls in 5 min was significantly higher in the MCAO/R surgery mice compared with the sham mice when evaluated following surgery (Fig. 2A). Seven days after surgery, the RNAi group exhibited a good recovery as assessed by the number of falls in 5 min, whereas the MCAO/R group and NC group mice continued to demonstrate evident deficits at that time-point. These data indicate that pAd-siRNA-cPLA2α treatment in mice alleviates exacerbated motor dysfunction induced by ischemic brain damage.

Effects of pAd-siRNA-cPLA2α on the expression levels of cPLA2α, FFA, LPC, PGE2 and LTB4. The role of cPLA2α in exacerbating transient focal ischemic brain damage has been previously demonstrated in cPLA2α-knockout mice (11). In the present study, whether the use of pAd-siRNA-cPLA2α attenuates ischemic brain damage was evaluated in a mouse model of MCAO/R. A laser Doppler perfusion monitor was used to monitor the regional blood flow and this was continued until 20 min after reperfusion. No difference in blood flow was detected in the mice from the different groups at any time (Fig. 6). Compared with the NC group, the RNAi group effectively decreased the cPLA2α expression levels in the ischemic damage mice at 2 h, 1, 3 and 7 days after surgery (Fig. 3; P<0.01).

To evaluate whether pAd-siRNA-cPLA2α treatment decreased the levels of phospholipid degradation products, cPLA2α-derived injurious lipid mediators in the brain tissue samples were assessed. The results indicated that pAd-siRNA-cPLA2α treatment significantly reduced the levels of FFA and LPC, while the levels of FFA and LPC exhibited a marked increase in the NC group (Fig. 4A and B). At the measurement time-points, the levels of FFA and LPC in the RNAi group did not recover to the normal levels of the sham group (data not shown). cPLA2α tend to catalyze the hydrolysis of phospholipids in the sn-2 position (35), and it is the crucial enzyme of AA, PG and PAF. PGE2 and LTB4 are proinflammatory eicosanoids metabolized from free AA. Serum from
the RNAi group mice exhibited a reduced level of PGE2 compared with the sham, MCAO/R and NC group animals at 24 h following surgery (Fig. 4C; P<0.001). Similarly, reduced levels of LTB4 were observed in the serum of the RNAi group animals (Fig. 4D; P<0.001). PGE2 and LTB4 in the serum were significantly decreased following pAd-siRNA-cPLA2α treatment (Fig. 4).

Effects of cPLA2α on pathological changes. H&E staining effectively demonstrated the pathological changes. The examination of these pathological sections demonstrated that the morphological features of the brain tissue samples were normal 14 days after surgery in the sham group (Fig. 5A), while the morphological features of the brain tissue samples from the other three groups exhibited significant changes. MCAO/R tissue samples exhibited nerve cell necrosis, loose structures and visible intercellular edema to different degrees, which validated that the MCAO/R surgery had been successfully performed. However, compared with the MCAO/R group or NC group, the histological structure changes of the brain tissue samples were less marked in the RNAi group. The results demonstrate that the treatment of cPLA2α RNAi ameliorated pathological changes following ischemic brain damage in mice.

Effects of cPLA2α on reducing ischemia-induced brain infarct volume. Brain infarct was assessed following the experiment using laboratory standard volumetric analysis of the anterior and posterior views of coronal slabs stained with TTC, and corrected for swelling. No infarction was observed in the sham group, whereas extensive lesions were observed in the model and NC groups. RNAi treatment effectively decreased the cerebral injury with a significant reduction in infarct volume of the ischemic hemisphere when compared with the NC or model group mice (Fig. 5B; P<0.01).

Effects of cPLA2α on apoptosis in brain cells. TUNEL staining was performed to evaluate whether pAd-siRNA-cPLA2α treatment alleviates cell apoptosis by decreasing cPLA2α expression levels (Fig. 5C and E). TUNEL+ cell number in the brain tissue of mice following MCAO/R surgery exhibited a significant increase when compared with the
The RNAi group had a smaller number of TUNEL-positive cells in comparison to the NC group on days 7 and 14. Such results indicate that pAd-siRNA-cPLA2α treatment may improve pathological changes by inhibiting apoptosis in mice with focal ischemic brain damage.

Discussion

The treatment of cerebral ischemia should be selected according to the principle of damage cascades. The aim of clinical treatment of cerebral ischemia is to recover the blood and oxygen supply, suppress inflammation of the ischemic area, and maintain the integrity of the neuron structure and function. In the present study, it was validated and confirmed that cPLA2α significantly influences the speed of recovery from cerebral ischemia. In addition, cPLA2α-knockout effectively suppresses inflammation and helps to maintain the integrity of the neuron structure and function. The present results provide a reference for the potential use of adenoviruses-mediated RNAi targeting cPLA2α in the clinical setting.

Numerous studies demonstrated that the level of cPLA2α was closely associated with the functional recovery in stroke (11), Alzheimer's disease (38) and multiple sclerosis (34), which indicated the causative role of cPLA2α in neurodegeneration. cPLA2α has a major role in rat cerebral ischemia, and upregulated expression of cPLA2α following trauma may be significant in secondary injury, such as inflammation, nociception and functional deficits (11). Certain studies indicated that certain inhibitors of cPLA2α inhibit the activity of cPLA2α, but not the protein expression and that the effects are short term (22,39). In the present study, three adenoviruses-mediated RNAi sequences were established and their effects in L929 cells were identified. An RNAi sequence that inhibits the expression and activity of mouse cPLA2α was successfully identified (Fig. 1). The MCAO/R mice were treated with pAd-siRNA-cPLA2α prior to injury, and it was observed that it effectively reduced the expression levels of cPLA2α (Fig. 3). pAd-siRNA-cPLA2α affects the expression of cPLA2α in the long-term (14 days) following only one injection (Fig. 3C).

The fact that inhibition of cPLA2α attenuates focal ischemic brain damage in mice indicates that the activity of cPLA2α contributes markedly to the injury cascade following MCAO/R surgery (9,10). The effects of cPLA2α on focal ischemic brain damage go beyond the phase of acute injury. Indeed, the present and previous studies have demonstrated that the secondary injury mechanism is closely associated with cPLA2α (11), as the cPLA2α RNAi treatment reduced the injury following 3 days of reperfusion. It has been reported previously that upregulation of cPLA2α is relevant to blood-brain barrier disruption in rats at 24 h of reperfusion (40).

A total of 75% of patients with cerebral ischemia exhibit varying degrees of neurological deficit (41), and motor function deficits are an important manifestation of neurological deficits (42). Neurological deficits are a characteristic of cerebral ischemia. Thus, motor function recovery is an important index for evaluating the therapeutic efficacy of medications. An apparent neurological deficit was observed from 2 h after MCAO/R surgery in the present study, which validated the cerebral ischemia model. However, the finding that the NDS was reduced in the RNAi group compared with the NC group...

**Figure 4. Effects of siRNA-cPLA2α on the expression of FFA, LPC, PGE2 and LTB4 in mice after MCAO/R. Expression levels of (A) FFA and (B) LPC in the brain tissue from mice 24 h after MCAO/R surgery were measured by high performance thin layer chromatography. Expression levels of (C) PGE2 and (D) LTB4 in the serum of mice 24 h after MCAO/R surgery were measured using ELISA. pAd-siRNA-cPLA2α treatment decreased the expression levels of FFA, LPC, PGE2 and LTB4. Data are presented as mean ± standard deviation of three independent experiments. **P<0.01 and ***P<0.001 (Student's t-test). siRNA, small interfering RNA; cPLA2α, cytosolic phospholipase A2α; FFA, free fatty acids; LPC, lysophosphatidylcholine; PGE2, prostaglandin E2; LTB4, leukotriene B4; MCAO/R, middle cerebral artery occlusion and/or reperfusion; NC, negative control.
supports the conclusion that pAd-siRNA-cPLA2α treatment protects against functional injury (Fig. 2A). The motor function recovery was restored faster in the RNAi group, which was assessed by the number of falls by mice following MCAO/R surgery (Fig. 2B).

Inhibition of cPLA2α to treat cerebral ischemia has certain anti-inflammatory and neuroprotective effects, expediting the processes involved in physiological function recovery. Previous studies regarding the link between cPLA2α and cerebral ischemia have revealed that levels of cyclooxygenase (COX)-2 in the basal and stimulated central nervous system are reduced in cPLA2α knock-out animals (43, 44). Proinflammatory lipid mediators, such as PGE2, LTB4, LPC and FFA, which are regulated by the activity of cPLA2α are closely associated with ischemic brain damage. COX-2 and 5-lipoxygenase (5-LOX), as well as their products, were involved in neuron damage and neurodegeneration. FFA represents predominantly free AA, and was catalyzed to prostaglandins and leukotrienes by COX-2 and 5-LOX, respectively. In the present study, PGE2 was a representative product of COX-2, and LTB4 was a representative product of 5-LOX. LPC is an important signaling molecule correlated with chronic inflammation and...
tissue damage (45). In the present study, MCAO/R surgery mice treated with pAd-siRNA-cPLA2α demonstrated effectively reduced expression levels of those proinflammatory lipid mediators (Fig. 4), indicating that they originated from cPLA2α. H&E staining demonstrated that the RNAi group changes were less marked when compared with the NC group. Similarly, mice treatment with pAd-siRNA-cPLA2α exhibited a decreased number of TUNEL+ cells and the brain infarct volume was reduced. These findings indicate that treatment with pAd-siRNA-cPLA2α may alleviate the pathological damage by reducing inflammation and inhibiting apoptosis following MCAO/R surgery. Thus, these data suggest that treatment with pAd-siRNA-cPLA2α inhibits cPLA2α effectively and efficiently protects cells from ischemic-induced adverse effects.

In conclusion, the present study demonstrates the therapeutic potential of the adenoviruses-mediated RNAi targeting cPLA2α in a cerebral ischemia animal model. The efficacy of the treatment was predominantly attributed to the inhibition of cPLA2α and the reduction of cPLA2α-derived proinflammatory lipid mediators. The neuroprotective effects of pAd-siRNA-cPLA2α treatment in the MCAO/R mice indicated that a long-term effect cPLA2α inhibitor may be used in the future as a therapeutic strategy for cerebral ischemia. However, further investigations are required to evaluate the adenoviruses-mediated RNAi targeting cPLA2α in human cerebral ischemia patients.

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Availability of data and materials

The analyzed datasets generated during the study are available from the corresponding author on reasonable request.

Authors’ contributions

HW and LZ conceived and designed the study. HW, HL and FZ performed the experiments. HW wrote the study. LZ reviewed and edited the study. All authors read and approved the study.

Ethics approval and consent to participate

All procedures performed with mice were according to the National Institutes of Health Guide for the Care and Use of Laboratory Animals, and the experimental procedures were approved by the Ethics Committee of Cangzhou Central Hospital (Hebei, China).

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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