Current advances on the phytochemical composition, pharmacologic effects, toxicology, and product development of *Phyllanthi Fructus*

Xiaoyu Yan¹, Qiuju Li²,³, Lin Jing¹, Shuangyue Wu¹, Wei Duan¹, Yan Chen¹,³, Dayi Chen¹* and Xiaoqi Pan¹,²*

¹School of Public Health, Chengdu University of Traditional Chinese Medicine, Chengdu, China, ²State Key Laboratory of Southwestern Chinese Medicine Resources, Chengdu University of Traditional Chinese Medicine, Chengdu, China, ³School of Pharmacy, Chengdu University of Traditional Chinese Medicine, Chengdu, China

*Phyllanthi Fructus* (PF), the edible fruits of *Phyllanthus emblica* L., serves as an important resource for some health products, foods and drugs due to its high safety and sufficient nutritional value. In recent years, in vivo and in vitro experiments have been conducted to reveal the active components of PF. More than 180 compounds have been isolated and identified from the PF so far, primarily including tannins, phenolic acids, flavonoids, terpenoids, polysaccharides, fatty acids and amino acids. In traditional Chinese medicine (TCM), PF is used to cure several diseases such as bronchitis, asthma, diabetes, peptic ulcer, hepatopathy, leprosy, and jaundice. Consistent with ethnopharmacology, numerous modern studies have demonstrated that the extracts or monomeric compounds derived from PF exhibit various pharmacological effects including anti-oxidation, anti-bacteria, anti-inflammation, anti-tumour, anti-virus, immunity improvement, hypoglycemic and hypolipidemic effects, and multiple organ protective protection. Toxicological studies on PF indicated the absence of any adverse effects even at a high dose after oral administration. Due to strict quality control, these pharmacological activities and the safety of PF greatly improve the development and utilization of products. Our comprehensive review aims to summarize the phytochemistry, pharmacological effects, toxicology, and product development of PF to provide theoretical guidance and new insights for further research on PF in the future.

**KEYWORDS**

phyllanthi fructus, traditional Chinese medicine, phytochemistry, pharmacological effects, toxicology, product development
1 Introduction

One-quarter of the drugs in the world are directly extracted from plants or prepared with plants as raw materials, such as cocaine in coca leaves and digitalis toxin in digitalis leaves (Na and Yang, 2002). The prevalence of medicinal plants in clinical application is well documented in the pharmacopeias of various countries. Over 1,300 kinds of medicinal plants with anti-inflammatory and analgesic properties have been used by European physicians to treat renal and digestive system diseases in clinics (Wang et al., 2020). In addition, the United States has screened more than 20,525 kinds of medicinal plants to discover novel drugs against cancer and acquired immune deficiency syndrome (AIDS) (Xiao and Liu, 1983). Medicinal plants have been used in China for at least 2000 years. According to records, there are 35,784 species of plants in China, with nearly 12,000 species having medicinal value (Wang, 2022).

*Phyllanthus emblica* L., which is also known as amla, nelli, annole, or dhatriphala in different countries, belongs to the Phyllanthaceae family that contains more than 1,000 species widely distributed around the world. And it has been promoted and planted by the United Nations Health Organization throughout the world as three healthy plants, together with *Pinus massoniana* Lamb. and *Pueraria thomsonii* Benth. (Wu et al., 2017). Although all parts of *Phyllanthus Emblica* L. are used for medicinal purposes, the fruits (*Phyllanthi Fructus, PF*) are more widely used in traditional Chinese medicine (TCM), either alone or in combination with other traditional herbs for the treatment of many infectious and non-infectious diseases. PF was listed as a dual-purpose medicinal material by China’s Ministry of Health in 1998 (Wu et al., 2017). In Ayurveda and Unani systems of medicine, PF is also one of the key constituents used in various herbal formulations including patented drugs (Rai et al., 2012). Meanwhile, PF has been used as nutritious tonic, possessing vital amino acids and vitamins. It is particularly the main source of ascorbic acid and minerals compared to other citrus fruits. More than 180 compounds, such as tannins, phenolic acids, flavonoids, fatty acids, polysaccharides, terpenoids, and amino acids, have so far been identified in PF. Various biological activities of these compounds have been reported. Modern pharmacology research has confirmed that PF has many pharmacological effects including anti-inflammation, anti-oxidation, anti-tumour, anti-viral and lowering blood lipid, blood glucose and blood pressure. A recent study reported the role of PF against skin aging (Chaikul et al., 2021). Furthermore, no obvious adverse effects have been observed during PF application, which means that PF presents high security in clinical treatment (Li, 2001).

Previous studies focused on pharmacological activities of PF without providing a comprehensive critical analysis of other relevant information, such as phytochemistry and product development. In this review, a comprehensive understanding of PF as drugs or food, such as phytochemistry, pharmacological effects, toxicology, product application and quality control were summarized. Our review provides scientific evidence for fully exploiting the nutritional value of PF products and developing high-value-added products in depth to promote the sustainable and healthy development of the PF production industry.

2 Literature search methods

Available information on PF was collected from published materials, including monographs on medicinal plants, ancient and modern recorded classics, pharmacopeias, and electronic databases, such as Web of Science, PubMed, CNKI, Wanfang Database, Baidu Scholar, Flora of China (FOR), from 1952 up to June 2022. The following keywords including “Phyllanthus emblica”, “the botany of Phyllanthus emblica”, “pharmacological activity of Phyllanthus emblica”, “antioxidant of Phyllanthus emblica”, “biological activity of Phyllanthus emblica”, “quality control of Phyllanthus emblica”, “traditional uses of Phyllanthus emblica”, or “toxicity of Phyllanthus emblica” were used to search relevant literature. The titles and abstracts were screened for relevancy, and articles related to roots and leaves of *Phyllanthus emblica* L., or repetitive articles were eliminated. *In vitro* or *in vivo* studies and clinical trials of PF were included. The search strategy is shown in Figure 1. The IUPAC names of the known chemical compounds were checked using the PubChem database and

![FIGURE 1](image-url)
their chemical structures were drawn using ChemDraw Pro 20.0 software.

3 Botanical description and geographic distribution

PF, the ripe fruit of *Phyllanthus Emblica* L., is recorded in various versions of Chinese Pharmacopoeia. The plant is easy to adapt to the external environment and climate, which often grows in sparse forests, scrub, wastelands or sunny places in ravines at 200–2,300 m above sea level (Yang et al., 2018). The size of the tree is small to medium with a range of 8–18 m in height (Ganesan, 2003). The flowers are greenish-yellow. Flowering between April and June (Wei et al., 2002; Arora et al., 2012). The fruit is nearly spherical, diameter ranges between 1.8 and 2.5 cm, light greenish yellow or brick-red, quite smooth and hard on appearance, with six vertical stripes or furrows (Sangeetha et al., 2010). The fruiting period is from July to September. The fruit, as a highly nutritious drug-food homologous substance, has a sweet and sour taste. It is often divided into wild and fruity types in the market. And the fruity types are characterized by larger and sweeter fruits, smaller kernels, less fiber and higher yields compared with the wild types. Fresh fruit is often processed as jam, preserved, and canned fruit. The seeds are slightly reddish, 5–6 mm long, and 2–3 mm wide (Flora of China Editorial Committee, 1998). The seedlings start bearing fruits 7–8 years after planting, while the budded clones start bearing fruits after 5 years (Kumar et al., 2012).

PF is distributed in most tropical and subtropical countries mainly in China, Indonesia, and the Malay Peninsula. PF is native to tropical southeastern Asia, particularly in central and southern India, Pakistan, Nepal, Bhutan, Bangladesh, Malaya, Myanmar and the Mascarene Islands (Supplementary Figure S1) (Xia et al., 1997). Originally, it was cultivated in Madagascar (Ganesan, 2003). Subsequently, PF is also cultivated in South America. In China, PF is widely growing in Sichuan, Guizhou, Jiangxi, Fujian, Guangdong, Hainan, Taiwan, Guangxi and Yunnan provinces (Supplementary Figure S2) (Yang et al., 2014). The fruit from Guizhou and Yunnan provinces are most suitable for use as medicine and health products, which contain high levels of gallic acid, corilagin and ascorbic acid (Mao, 2019).

4 Phytochemical composition of *Phyllanthus Fructus*

PF, like most botanical medicines, contains numerous natural products with different structural patterns, such as tannins, phenolic acids, flavonoids, and ascorbic acid, which are the main active components in pharmacodynamic activity (Pareek et al., 2018; Zhu et al., 2018). Isocorilagin (27), emblicanin A (44) and B (45), etc. are unique phytochemicals of PF with antioxidant and anti-tumour properties. Major chemical components isolated from PF are listed in Table 1. Different pharmacological activities of PF are often attributed to these active ingredients.

4.1 Tannins

Tannins are a kind of polyphenolic compounds with a complex structure, whose molecular structure has many active phenolic hydroxyl groups. These active phenolic hydroxyl groups are easily oxidised into quinone structures which can scavenge free radicals, thus rendering their antioxidant, anti-inflammatory, anti-mutagenic and anti-cancer activities (Shao et al., 2014). According to the chemical structure, tannins include hydrolyzed and condensed types (Guo, 2013). Tannins content is up to 45% and 14% in fresh and dried PF respectively, most of which are hydrolysable tannins (Chen et al., 2016). Tannin compounds mainly take D-glucose as glycosyl, and various monomers are derived at the substitutions of R1, R2, R3, R4 and R5, respectively. Approximately 49 types of tannins have been isolated from PF thus far, including chebulinic acid (1), elaeocarpusin (15), chebulagic acid (17), and geraniin (23), etc (Figure 2). 3,4,6-tri-O-galloyl-β-D-glucose was isolated from PF for the first time in 2015 (Olennikov et al., 2015). Methyl chebulagulate (13) and dimethyl neochebulagulate (14) as novel compounds isolated from PF have not been fully studied in terms of pharmacological activity (Yang et al., 2020).

4.2 Phenolic acids

Phenolic acids are generated by replacing a hydrogen atom in an aromatic ring with a carboxylic acid group and at least one hydroxyl group. On the one hand, the hydrogen atoms given by the breakage of the -OH bond of the phenolic hydroxyl group can bind directly to free radical molecules. On the other hand, phenolic compounds can be converted into polyphenol anions by ionizing H+ in polar solvents or under alkaline conditions, which converts oxygen radicals directly into anions (Chen, 2019). Therefore, phenolic acids have a wide range of physiological activities, such as antioxidant, anti-UV radiation, anti-bacterial and anti-viral effects, which are used in food additives, pharmaceuticals, and cosmetic ingredients. 30 phenolic acid (51–81) components have been isolated from PF (Figure 3). Olennikov et al. isolated two new mucic acid derivatives, mucic acid 2,5-di-O-gallate 56) and mucic acid 2,5-di-O-gallate1,4-lactone (63), both of which have antioxidant activity (Olennikov et al., 2015). Chebulic acid trimethyl ester (72) and aryl-tetralin-type lignan (77) in PF were first identified in 2020. (Yang et al., 2020).
| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|----------------------|-------------------|----------|-----------------------|------------|
| 1   | Chebulinic acid      | C41H32O27         | Methanol | Anti-inflammatory, anti-hypertension | Avula et al., 2013; Lu et al., 2020 |
| 2   | Neochebulic acid     | C41H32O28         | Methanol | Antioxidant            | Avula et al. (2013) |
| 3   | 1,2,3,6-tetra-O-galloyl-β-D-glucose | C34H28O21 | Methanol | Antioxidant            | Xu et al. (2016) |
| 4   | 1,2,4,6-tetra-O-galloyl-β-D-glucose | C34H28O22 | Ethanol  | Anti-virus             | Xiang et al. (2011), Yang et al. (2010) |
| 5   | 3,4,6-tri-O-galloyl-β-D-glucose | C32H4O18 | Methanol | Antioxidant            | Xu et al. (2016) |
| 6   | 1,2,3,6-penta-O-galloyl-β-D-glucose | C34H28O22 | Methanol | Antioxidant            | Xu et al. (2016) |
| 7   | 1-O-galloyl-β-D-glucose | C34H28O22 | Aqueous  | Antioxidant            | Zhang et al. (2001) |
| 8   | 1,6-di-O-galloyl-β-D-glucose | C20H20O14 | Methanol | Antioxidant            | Majeeed et al. (2009) |
| 9   | 1-O-galloyl-β-D-glucose | C13H15O10 | Ethanol  | Antioxidant            | Zhang et al. (2001) |
| 10  | Terchebin            | C41H30O27         | Ethanol  | Antioxidant, anti-bacteria | Luo et al. (2012) |
| 11  | Mallotusinin         | C41H30O27         | Methanol | Antioxidant, anti-inflammatory | Luo et al. (2012) |
| 12  | Phyllanthunin A      | C22H30O17         | Ethanol  | _                      | Yang et al. (2020) |
| 13  | Mullenin             | C22H30O17         | Ethanol  | _                      | Xiang et al. (2011), Yang et al. (2010) |
| 14  | Mallotonin A         | C22H30O17         | Methanol | _                      | Xu et al., 2016; Bobasa et al., 2021 |
| 15  | Putranjivain A       | C41H30O27         | Methanol | Antioxidant, anti-inflammatory, anti-virus | Xu et al., 2016; Du et al., 2021 |
| 16  | Geraniin             | C41H30O27         | Methanol | Antioxidant, immunomodulatory, anti-tumour | Xu et al., 2016; Liu et al., 2008 |
| 17  | Punicafolin          | C41H30O27         | Methanol | Anti-diabetic           | Xu et al., 2016 |
| 18  | Carpinusin           | C41H30O27         | Methanol | Anti-diabetic           | Xu et al., 2016 |
| 19  | Chebulanin           | C41H30O27         | Methanol | Anti-diabetic           | Xu et al., 2016 |
| 20  | Corilagin            | C42H30O28         | Methanol | Anti-diabetic           | Xu et al., 2016 |
| 21  | Isocorilagin*        | C42H30O28         | Methanol | _                      | Liu et al. (2008) |
| 22  | Furosin              | C42H30O28         | Methanol | Antioxidant            | Liu et al. (2008) |
| 23  | Chebulanicin         | C42H30O28         | Methanol | Anti-diabetic           | Liu et al. (2008) |
| 24  | 2.3-(S)-hexahydroxy-diphen-oyl-β-D-glucose | C20H20O14 | Ethanol  | Anti-inflammatory       | Xu et al., 2016; Liu et al., 2008 |
| 25  | 3,4,6-tri-O-galloyl-β-D-glucose | C27H24O18 | Methanol | Anti-inflammation, anti-tumour | Xu et al., 2016; Yang and Liu, 2014 |
| 26  | 1β,4-di-O-galloyglucose | C20H20O14 | Ethanol  | _                      | Luo et al. (2012) |
| 27  | 2-(2-methylbutyryl)-phloroglucinol-1-O-β-D-glucopyranoside | C20H20O14 | Ethanol  | _                      | Xu, 2009; Wu et al., 2017 |
| 28  | Phyllanthin          | C22H30O17         | Ethanol  | _                      | Yang et al. (2007) |
| 29  | 2-(2-methylbutyryl)-phloroglucinol-1-O-(6″-O-galloyl)-β-D-glucopyranoside | C22H30O14 | Ethanol  | _                      | Xu, 2009; Wu et al., 2017 |

(Continued on following page)
| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|----------------------|-------------------|----------|-----------------------|------------|
| 40  | 3,3′-dimethylellagic acid-4′-O-α-L-rhamnopyranoside | C_{30}H_{32}O_{12}N_{1}B_{1} | Ethanol | _ | Xu, (2009) |
| 41  | 3-O-methyllellagic acid-4′-O-α-L-rhamnopyranoside | C_{22}H_{20}O_{12} | Ethanol | _ | Xu, (2009) |
| 42  | Isotricinnin | C_{20}H_{22}O_{18} | Ethanol | Hepatoprotective activity | Zhou et al. (2018) |
| 43  | Chebulic acid | C_{14}H_{12}O_{11} | Ethanol | Anti-neuroinflammatory, antioxidant | Xu et al. (2016) |
| 44  | Emblicanin A | C_{14}H_{12}O_{12} | Methanol | Antioxidant | Bhattacharya et al. (2000a) |
| 45  | Emblicanin B | C_{14}H_{12}O_{12} | Methanol | Antioxidant | Bhattacharya et al. (2000b) |
| 46  | Prodelphinidin B2 | C_{16}H_{32}O_{14} | Ethanol | Anti-tumour | Zhou et al. (2018) |
| 47  | Epicatechin-(4β→8)-epigallocatechin-3-O-gallate* | C_{23}H_{36}O_{12} | Ethanol | _ | Xu, (2009) |
| 48  | Epicatechin-(4β→8)-gallocatechin | C_{23}H_{36}O_{12} | Acetone | Hepatoprotective effects | Xu, (2009) |
| 49  | Prodelphinidin B1 | C_{34}H_{22}O_{14} | Ethanol | Anti-tumour | Zhou et al. (2018) |
| 50  | Epicatechin-(2β→7,4β→8)-gallocatechin | C_{30}H_{26}O_{14} | Ethanol | _ | Xu, (2009) |

**Phenolic acids**

| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|----------------------|-------------------|----------|-----------------------|------------|
| 51  | Mucic acid | C_{6}H_{10}O_{8} | Aqueous | Antioxidant | Majeed et al. (2009) |
| 52  | Mucic acid-1-methyl ester-6-ethyl ester | C_{16}H_{22}O_{12} | Methanol | Antioxidant | Zhang et al. (2017) |
| 53  | Mucic acid-1-methyl ester-2-O-gallate | C_{16}H_{22}O_{12} | Methanol | Antioxidant | Zhang et al. (2017) |
| 54  | Mucic acid-6-methyl ester-2-O-gallate | C_{16}H_{20}O_{12} | Methanol | Antioxidant, anti-virus | Zhang et al. (2017) |
| 55  | Mucic acid dimethyl ester-2-O-gallate* | C_{16}H_{20}O_{12} | Ethanol | _ | Xu et al., 2016 |
| 56  | Mucic acid-2-O-gallate* | C_{16}H_{20}O_{12} | Ethanol | _ | Olennikov et al. (2015) |
| 57  | Mucic acid-2,5-di-O-gallate | C_{16}H_{20}O_{12} | Ethanol | _ | Xu, 2009; Liu et al., 2017 |
| 58  | 1,4-lactone-3,5-di-O-gallate | C_{21}H_{16}O_{15} | Acetone | Antioxidant | Xu, 2009; Zhang et al., 2011 |
| 59  | Mucic acid-3,4,5-trihydroxybenzene-1-O-galloyl-β-D-glucopyranoside | C_{21}H_{16}O_{15} | Acetone | Antioxidant, anti-virus | Zhang et al., 2018 |
| 60  | Mucic acid-3,4,5-trihydroxybenzene-1,4-lactone-3,5-di-O-gallate | C_{21}H_{16}O_{15} | Acetone | Antioxidant, anti-virus | Zhang et al., 2018 |
| 61  | Mucic acid-1,4-lactone-5-O-gallate* | C_{14}H_{12}O_{11} | Ethanol | _ | Xu, 2009; Liu et al., 2017 |
| 62  | Mucic acid-1,4-lactone-6-methyl-ester-5-O-gallate | C_{14}H_{12}O_{11} | Ethanol | _ | Xu, 2009; Zhang et al., 2011 |
| 63  | Mucic acid-1,4-lactone-2-O-gallate | C_{16}H_{20}O_{12} | Ethanol | _ | Xu, 2009; Zhang et al., 2011 |
| 64  | Mucic acid-1,4-lactone-6-methyl-ester-5-O-gallate | C_{16}H_{20}O_{12} | Ethanol | _ | Xu, 2009; Zhang et al., 2011 |
| 65  | Mucic acid-2,5-di-O-gallate | C_{16}H_{20}O_{12} | Ethanol | _ | Olennikov et al. (2015) |
| 66  | Pyrogallol | C_{6}H_{6}O_{5} | Ethanol | Antioxidant | Xu, 2009; Liu et al., 2017 |
| 67  | 1,3,5-trihydroxybenzene-1,4-lactone-β-D-glucopyranoside | C_{14}H_{12}O_{8} | Ethanol | _ | Xu, 2009; Zhang et al., 2011 |
| 68  | Gallic acid | C_{6}H_{6}O_{5} | Methanol | Antioxidant, anti-inflammatory | Xu et al., 2009 |
| 69  | Protocatechuic acid | C_{6}H_{6}O_{5} | Methanol | Antioxidant | Xu, 2009 |
| 70  | Methyl gallate | C_{6}H_{6}O_{5} | Methanol | Hypoglycemic activity | Xu, 2009; Liu et al., 2017 |
| 71  | Ethyl gallate | C_{6}H_{6}O_{5} | Methanol | Antioxidant | Zhang et al. (2016) |
| 72  | 3-ethoxy-4,5-dihydroxybenzoic acid | C_{14}H_{12}O_{5} | Acetone | _ | Zhang et al. (2003) |
| 73  | 2-carboxymethylphenol-1,4-β-glucopyranoside* | C_{14}H_{12}O_{5} | Acetone | _ | Zhang et al. (2003) |
| 74  | 2,6-dimethoxy-4-(2-hydroxyethyl)phenol-1,4-β-glucopyranoside* | C_{14}H_{12}O_{5} | Acetone | _ | Zhang et al. (2003) |
| 75  | Chebulic acid trimethyl ester | C_{14}H_{12}O_{5} | Ethanol | Hepatoprotective effects | Yang et al. (2020) |
| 76  | 3,3′-dihydroxy-4,4′-2-propenyl-2,2′-biphenylcarboxylic acid | C_{20}H_{10}O_{4} | Ethanol | Antioxidant | Yang et al. (2020) |
| 77  | 3,4,8,9-tetrahydroxybenzo [b] pyran-6-one | C_{14}H_{12}O_{3} | Ethanol | _ | Zhang et al. (2016) |
| 78  | 1,6-di-O-galloylglucose | C_{14}H_{12}O_{3} | Ethanol | _ | Noor et al. (2018) |

(Continued on following page)
| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|-----------------------|-------------------|----------|----------------------|------------|
| 79  | Digallic acid          | C_{14}H_{10}O_{9}  | Methanol | Antioxidant, lipid-lowering activity | Xu et al. (2016) |
| 80  | Ellagic acid           | C_{14}H_{6}O_{8}   | Acetone  | Antioxidant, anti-inflammatory | Xu et al. (2016) |
| 81  | Aryl-tetralin-type lignan$^a$ | C_{31}H_{33}O_{15} | Ethanol  | _                     | Yang et al. (2020) |
| 82  | Eriodictyol            | C_{15}H_{12}O_{6}  | Methanol | Antioxidant, anti-tumour | Xu et al. (2016) |
| 83  | Eriodictyol-7-O-glaceside | C_{21}H_{22}O_{11} | Methanol | _                     | Xu et al. (2016) |
| 84  | (S)-eriodictyol-7-O-(6″-O-galloyl)-β-D-glucopyranoside | C_{22}H_{17}O_{10} | Ethanol  | _                     | Zhang et al. (2002) |
| 85  | (-)-epigallocatechin-3-O-gallate | C_{22}H_{18}O_{11} | Methanol | Antioxidant, anti-inflammatory | Xu et al. (2016) |
| 86  | (-)-epicatechin-3-O-gallate | C_{22}H_{18}O_{10} | Methanol | Antioxidant, anti-inflammatory | Xu et al. (2016) |
| 87  | (-)-epiafzelechin      | C_{15}H_{14}O_{5}  | Methanol | Antioxidant           | Xu et al. (2016) |
| 88  | (-)-epigallocatechin   | C_{15}H_{14}O_{5}  | Methanol | Antioxidant, anti-virus | Xu et al. (2016) |
| 89  | (-)-epicatechin        | C_{15}H_{14}O_{5}  | Methanol | Antioxidant           | Xu et al. (2016) |
| 90  | (-)-gallolechin        | C_{15}H_{14}O_{5}  | Methanol | Antioxidant, anti-tumour | Xu et al. (2016) |
| 91  | (+)-catechin           | C_{15}H_{14}O_{5}  | Ethanol  | Antioxidant, anti-inflammatory | Wang et al. (2019) |
| 92  | Delphinidin            | C_{15}H_{14}O_{6}  | Methanol | Antioxidant           | Xu et al., 2009 |
| 93  | Wogonin                | C_{15}H_{14}O_{6}  | Methanol | Antioxidant, anti-tumour | Nisar et al. (2018) |
| 94  | 5,7-dihydroxy-2-(4-hydroxyphenyl)-4H-chromen-4-one | C_{15}H_{14}O_{5} | Methanol | Antioxidant, anti-tumour | Xu et al., 2009 |
| 95  | Naringenin             | C_{15}H_{10}O_{5}  | Ethanol  | Antioxidant           | Wang et al. (2019) |
| 96  | Naringenin-7-O-glucoside | C_{21}H_{22}O_{10} | Methanol | Hypoglycemic activity | Xu et al. (2016) |
| 97  | Naringenin-7-O-(6″-O-galloyl)-glucoside | C_{22}H_{22}O_{17} | Ethanol  | _                     | Xu et al. (2016) |
| 98  | Dihydrokaempferol      | C_{15}H_{10}O_{4}  | Methanol | Antioxidant, anti-inflammatory | Xu et al. (2016) |
| 99  | Rutin                  | C_{15}H_{10}O_{4}  | Methanol | Antioxidant, anti-tumour | Xu et al. (2016) |
| 100 | Kaempferol 3-O-rhamnioside | C_{15}H_{10}O_{16} | Aqueous  | Antioxidant           | Cui et al. (2010) |
| 101 | Quercetin              | C_{15}H_{10}O_{5}  | Aqueous  | _                     | Cui et al. (2010) |
| 102 | Myricetin-3-O-rhamnioside | C_{21}H_{16}O_{12} | Methanol | Antioxidant, anti-inflammatory, antiviral, anti-tumour | Liu et al. (2007b) |
| 103 | Quercetin-3-rhamnoside | C_{21}H_{20}O_{11} | Ethanol  | _                     | Liu et al. (2007a) |
| 104 | Isoquercitrin           | C_{21}H_{20}O_{11} | Aqueous  | _                     | Liu et al. (2007b) |
| 105 | Quercetin-7-O-β-D-glucopyranoside | C_{21}H_{20}O_{12} | Aqueous  | Antioxidant           | Liu et al. (2007a) |
| 106 | Myricetin              | C_{21}H_{10}O_{8}  | Aqueous  | Antioxidant           | Liu et al. (2007b) |
| 107 | 2-(2-methylbutryl) phloroglucinol (multifidol) glucoside$^*$ | C_{21}H_{20}O_{10} | Methanol | _                     | Xu et al. (2016) |
| 108 | 2-(2-methylbutylyl) phloroglucinol-1-O-(6″-O-β-D-apiofuranosyl)-β-D-glucopyranoside$^*$ | C_{22}H_{22}O_{13} | Methanol | _                     | Xu et al. (2016) |
| 109 | Avicularin             | C_{20}H_{16}O_{3}  | Ethanol  | _                     | Zhou et al. (2018) |
| 110 | Naringenin-7-O-(6″-O-trans-pcoumaroyl)-glucoside | C_{20}H_{18}O_{12} | Methanol | _                     | Xu, (2009) |
| 111 | (S)-eriodictyol-7-O-(6″-O-trans-pcoumaroyl)-β-D-glucopyranoside$^*$ | C_{20}H_{18}O_{13} | Ethanol  | _                     | Zhang et al. (2002) |

**Terpenoids**

(Continued on following page)
| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|----------------------|-------------------|----------|-----------------------|------------|
| 113 | β-amyrin             | C_{30}H_{50}O      | Ethanol  | Anti-inflammatory, anti-virus | Zhou et al. (2018) |
| 114 | Betulin              | C_{30}H_{50}O_{2}  | Ethanol  | Anti-inflammatory, anti-virus | Zhou et al. (2018) |
| 115 | β-amyrone            | C_{30}H_{52}O      | Ethanol  | Antiviral              | Zhou et al. (2018) |
| 116 | Lupeol               | C_{30}H_{52}O      | Methanol | Anti-inflammatory, anti-virus | Xu et al. (2016) |
| 117 | Phyllaemblinola      | C_{20}H_{24}O_{9}  | Ethanol  | Antioxidant            | Zhang et al. (2016) |

**Polysaccharides**

| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|----------------------|-------------------|----------|-----------------------|------------|
| 118 | Pectin               | C_{5}H_{10}O_{5}   | Methanol | Antioxidant, anti-aging | Wang et al. (2019) |
| 119 | Glucose              | C_{6}H_{12}O_{6}   | Methanol | Antioxidant, anti-aging | Wang et al. (2019) |
| 120 | Fructose             | C_{6}H_{12}O_{6}   | Methanol | Antioxidant, anti-aging | Wang et al. (2019) |
| 121 | Saccharose            | C_{12}H_{22}O_{11} | Methanol | Antioxidant, anti-aging | Wang et al. (2019) |

**Fatty acids**

| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|----------------------|-------------------|----------|-----------------------|------------|
| 122 | Lauric acid          | C_{12}H_{24}O_{2}  | Ethanol  | Immuno-enhancement, anti-virus | Kumaran and Karunasagar, (2006) |
| 123 | Palmitic acid        | C_{16}H_{32}O_{2}  | Ethanol  | Antioxidant            | Xu, (2009) |
| 124 | Hexacosanoic acid    | C_{26}H_{52}O_{2}  | Ethanol  | _                      | Xu, (2009) |
| 125 | Palmitoleic acid     | C_{16}H_{30}O_{2}  | Ethanol  | Hypoglycemic, hypolipidemic activities | Zhao et al. (2007) |
| 126 | Heptadecanoic acid   | C_{17}H_{36}O_{2}  | Ethanol  | _                      | Zhao et al. (2007) |
| 127 | Oleic acid           | C_{18}H_{34}O_{2}  | Ethanol  | Hypoglycemic, hypolipidemic activities | Zhao et al. (2007) |
| 128 | Stearic acid         | C_{18}H_{36}O_{2}  | Ethanol  | _                      | Zhao et al. (2007) |
| 129 | Linolenic acid (a-linolenic acid, b-γ-linolenic acid) | C_{18}H_{30}O_{2} | Ethanol  | Hypoglycemic, hypolipidemic activities and antioxidant | Zhao et al. (2007) |
| 130 | Myristic acid        | C_{14}H_{28}O_{2}  | Ethanol  | Anti-tumour, anti-bacteria | Zhao et al. (2007) |
| 131 | Arachidonic acid     | C_{20}H_{34}O_{2}  | Ethanol  | Immuno-enhancement, antioxidant | Zhao et al. (2007) |
| 132 | Docosanoic acid/Behenic acid | C_{22}H_{44}O_{2} | Ethanol  | _                      | Meng et al. (2011) |
| 133 | Pentacosanoic acid   | C_{30}H_{60}O_{2}  | Ethanol  | _                      | Zhao et al. (2007) |
| 134 | Melissic acid        | C_{30}H_{60}O_{2}  | Ethanol  | Anti-bacteria          | Zhao et al. (2007) |
| 135 | Lignoceric acid      | C_{24}H_{48}O_{2}  | Ethanol  | _                      | Meng et al. (2011) |

**Amino acids**

| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|----------------------|-------------------|----------|-----------------------|------------|
| 136 | Glutamic acid        | C_{6}H_{5}NO_{4}   | Aqueous  | Regulation of protein and glucose metabolism | Yuan et al. (2021) |
| 137 | Proline acid         | C_{6}H_{5}NO_{4}   | Aqueous  | Immunomodulatory effect | Yuan et al. (2021) |
| 138 | Alanine acid         | C_{3}H_{7}NO_{2}   | Aqueous  | Hypoglycemic, hypolipidemic activities, and immuno-enhancement | Yuan et al. (2021) |

(Continued on following page)
4.3 Flavonoids

Flavonoids are formed by two benzene rings with hydroxyl groups attached to the A and B ring linked via three central carbon atoms, thus having 2-benzylchromone basic parent nucleus structures. The biological activity of flavonoids is closely related to the chemical structure. Flavonoids terminate free radical chain reactions by binding to peroxyl radicals, which is one of the reasons for the antioxidant properties. The antioxidant activity is also related to the following structural features, the substitution position and number of phenolic hydroxyl groups, the C-2,3 double bond, and the spatial structure of the compound (Chen et al., 2013). The antitumour activity of flavonoids is attributed to the difference in parent nucleus structure, hydroxylation pattern and degree, and whether the hydroxyl group is substituted or not (Zhao et al.,

| NO. | Chemical constituents | Molecular formula | Extracts | Biological activities | References |
|-----|----------------------|-------------------|----------|-----------------------|------------|
| 139 | Lysine acid          | C₆H₁₄N₂O₂         | Aqueous  | Neuroprotective effect | Yuan et al. (2021) |
| 140 | Tryptophan acid      | C₁₁H₁₃N₂O₂        | Aqueous  | Neuroprotective effect | Yuan et al. (2021) |
| 141 | Histidine acid       | C₆H₁₀N₂O₄         | Aqueous  | Antioxidant and anti-inflammatory | Yuan et al. (2021) |
| 142 | Tyrosine acid        | C₆H₁₁NO₂          | Aqueous  | Hypoglycemic activity | Wu et al. (2003) |
| 143 | Leucine acid         | C₆H₁₁NO₂          | Aqueous  | Hypoglycemic activity | Wu et al. (2003) |
| 144 | Methionine acid      | C₆H₁₁NO₂S         | Aqueous  | Immunomodulatory effect | Wu et al. (2003) |
| 145 | Isoleucine acid      | C₆H₁₁NO₂          | Aqueous  | Hypoglycemic activity | Wu et al. (2003) |
| 146 | Arginine acid        | C₆H₁₁NO₂          | Aqueous  | Immuno-enhancement     | Wu et al. (2003) |
| 147 | Serine acid          | C₆H₁₁NO₂          | Aqueous  | Immunomodulatory effect | Wu et al. (2003) |
| 148 | Glycine acid         | C₆H₁₁NO₂          | Aqueous  | Immunomodulatory and Neuroprotective effect | Wu et al. (2003) |
| 149 | Threonine acid       | C₆H₁₁NO₂          | Aqueous  | Immunomodulatory effect and hypolipidemic activity | Wu et al. (2003) |
| 150 | Phenylalanine acid   | C₆H₁₁NO₂          | Aqueous  | Hypoglycemic activity and antioxidant | Wu et al. (2003) |
|     | **Other compounds**  |                   |          |                       |            |
| 151 | Vitamin B₁           | C₁₂H₁₇CIN₄OS      | Aqueous  | Hypoglycemic activity and neuroprotective effect | Chen et al. (2003) |
| 152 | Vitamin B₂           | C₁₇H₂₀N₄O₆       | Aqueous  | Regulation of metabolism | Chen et al. (2003) |
| 153 | Ascorbic acid        | C₆H₈O₆            | Methanol | Antioxidant and anti-inflammatory | Chen et al. (2003) |
| 154 | α-carotene           | C₁₇H₁₄          | Methanol | Anti-tumour | Wang et al. (2019) |
| 155 | β-carotene           | C₁₇H₁₄          | Methanol | Immuno-enhancement and antioxidant | Wang et al. (2019) |
| 156 | Nicotinic acid       | C₆H₅NO₂         | Ethanol  | Anti-glycation and skin Protection | Wang et al. (2019) |
| 157 | 24-propylcholesterol | C₂₀H₄₀O        | Ethanol  | Hypoglycemic activity, antioxidant, anti-inflammatory and anti-tumour | Kumar and Karunakaran, (2006) |
| 158 | Sitosterol           | C₂₃H₄₆O₄        | Ethanol  | Antioxidant, anti-inflammatory and anti-tumour | Xu et al. (2016) |
| 159 | β-sitosterol-D-glucoside | C₂₅H₄₈O₆    | Ethanol  | Antioxidant and anti-inflammatory | Xu et al. (2016) |

*aThis compound is unique to PF.*
In addition, its anti-inflammatory and antiviral activity is influenced by the above-mentioned constitutive relationships. So far, over 30 flavonoids (82–112) have been reported in PF (Figure 4). Wogonin (93), rutin (99), and quercetin (102) are typical representatives of the flavonoids in PF.

### 4.4 Terpenoids

Terpenoids belong to the olefins, whose molecular formula is usually an integral multiple of isoprene. The biological activities of terpenoids are determined by variations in structure. For
example, its anti-inflammatory, antitumour and antiviral activities are affected by the substituents at C-4, C-8, and C-10 positions and the parent nucleus. At present, terpenoids have become an important source for studying natural products and developing new drugs. PF contains a variety of terpenoids with strong antiviral and antitumour activities (Figure 5). β-amyrin (113), betulin (114), and β-amyrone (115) are the most fully researched terpenoids to date (Zhou et al., 2018., Xu et al., 2009). A new sesquiterpene namely phyllaemblinol (116) from PF ethanol extracts was separated by high performance liquid chromatography (HPLC) in 2016 (Zhang et al., 2016).

4.5 Polysaccharides

Polysaccharides play a positive role in immune regulation and cell recognition (Du et al., 2017). It has a variety of biological activities such as anti-tumour, anti-viral and hypoglycaemic. However, the biological activity of polysaccharides is influenced by many factors including the composition of monosaccharides, various isomers, sequence of glycogen linkage, and position and content of substituents (Bai, 2019). The content of polysaccharides in dry powder is 47.76 ± 0.37 mg/g, which accounts for 5% of the dry powder of PF, and that of fresh fruit pulp is 2.67 ± 0.02 mg/g, which accounts for 4.38% of the fresh fruit of PF (Liu B. et al., 2015). PF is rich in polysaccharides thereby characterizing it with a unique flavour, which is initially sour and astringent and then sweet. There are four main types of polysaccharides in PF (Figure 5), including pectin (118), glucose (119), fructose (120), and sucrose (121).

4.6 Fatty acids

PF includes a variety of unsaturated (80.36%) and saturated fatty acids (19.64%) (Figure 5) (Zhang et al., 2013). The saturated fatty acids in PF mainly include lauric (122), palmitic (123), hexacosanoic
(124), melissic (134) (Kumaran and Karunakaran, 2006; Zhao et al., 2007; Xu, 2009), and lignoceric acid (135) (Li et al., 2015), etc. Palmitoleic (125), oleic (127), linolenic (129), are unsaturated fatty acids in PF (Zhao et al., 2007). Among them, the proportion of \( \alpha \)-linolenic acid belonging to \( \omega-3 \) polyunsaturated fatty acids is more than 50\%, which is beneficial to the brain and neural development, cardiovascular health, and tumour suppression. Importantly, \( \alpha \)-linolenic acid is an essential fatty acid that cannot be synthesised by the human body (Wang et al., 2007).

### 4.7 Amino acids

Amino acids are involved in maintaining the body’s normal physiological, biochemical, and immune functions, as well as growth, development, metabolism, and other life activities. The antioxidant properties of amino acids are due to the ability to directly provide hydrogen atoms to bind to free radicals (Chen, 2019). PF contains 18 kinds of amino acids (Figure 6), among which 8 essential amino acids, such as glutamic acid (136), proline (137), alanine (138) and lysine (139), are required by the human body. And the amount of total amino acid is 1.851 mg/g. (Xia et al., 1997). Other amino acids include tryptophan (140), histidine (141), tyrosine (142), leucine (143), methionine (144), isoleucine (145), arginine (146), serine (147), glycine (148), threonine (149), and phenylalanine (150) (Wu et al., 2003).

### 4.8 Others

PF contains 18 kinds of trace elements such as Zn, Ge, Se, Gr, Fe, Cu, and Mn, and the major elements include K, Na, Ca and P.
It is worth mentioning that the amount of Se in PF is up to 0.24–0.73 mg/100g, which is a Se-rich fruit with an extremely high health value (Jose et al., 1995). Moreover, about 12 vitamins such as vitamin B1 (151), vitamin B2 (152), ascorbic acid (153), and carotene (154, 155) are isolated from PF (Chen et al., 2003). Moreover, PF also contains alkaloid compounds, superoxide dismutase (SOD), steroids, protein, and other compounds (Kumaran and Karunakaran, 2006).

5 Pharmacological effects of *Phyllanthi Fructus*

Many *in vitro*, *in vivo* and clinical studies have confirmed the pharmacological properties and functional activities of PF. The protective effect of PF on the body based on its biological activities were summarized in Figure 7.

5.1 Ethnopharmacology

PF has been utilized in Ayurveda, Siddha, and Unani systems in India, Sri Lanka, and China to treat various diseases. In the Chinese Pharmacopoeia (version 2020), PF is described as being cool in property, and sweet, sour, and astringent in flavour. It affects the stomach and lung meridians and acts to clear heat and cool blood, improve throat conditions, remove phlegm and promote good digestion. Therefore, PF can be applied in the treatment of blood heat and blood stasis, indigestion and abdominal distension. The recommended dosage is 3–9 g.
Because PF is cool in property, and sour and astringent in flavour, individuals with deficiency-cold of the spleen and stomach, and pregnant women should use it with caution. In China, different ethnic groups use PF in slightly different ways. For example, Tibetan people mainly use the fruit to treat “Blood disease”, “Chiba disease” and “Bacon disease”, which are equivalent to the syndrome of blood stasis, liver and gallbladder disease, hypertension and digestive system disease (Luo, 2004), Dai people directly eat fresh fruits for curing tonsillitis, Uygur people use PF to treat high blood pressure, Zhuang people use dried fruit to treat cold, fever, cough, pharyngitis, enteritis and abdominal pain, Miao and Buyi people like to use wine or saltwater to soak PF to invigorate the stomach and relieve cough. Bai (Luo et al., 2000), Bulang, Jinuo and Lahu nationalities treat sore throat with fruit chewing, and Naxi people believed that PF could prevent influenza (Higby and King, 2013).

PF is considered one of the most important medicines in Ayurveda, which is called immortal medicine in Indian mythology. According to statistics, the frequency of use of PF in Ayurveda’s commonly used pharmaceutical system is up to 195 times. It is often used in the treatment of various heat syndromes and cancer (Thakur et al., 1993). PF is used in the traditional Turkish medicine system to treat diarrhoea, dysentery and gastroenteritis (Ernst, 1999). In North America, it is mainly used as a diaphoretic prescription and
cathartic. In general, there is a surprising consistency of PF in the traditional medicine of different nationalities in China, India, and Southeast Asia, as a tonic medicine for longevity (Higby and King, 2013).

As a traditional medicine, PF is widely used in modern medical practice with compound prescriptions combined with other botanical drugs (Table 5), and some of them are even used directly in their traditional form. PF as an aphrodisiac helps to...
increase sperm count (Udupa, 1985). PF is also used as a hair tonic in traditional formulations for enriching hair growth and pigmentation due to the presence of fixed oil (Thakur et al., 1993). They are applied as skin lighteners loved by women (Chaudhuri, 2004). Herbal formulations of burnt seeds and oils of PF have been used to treat skin affictions (Dastur, 1952). Furthermore, PF helps to rejuvenate all body organ systems, provides strength and wellness, and boosts the immune system. The traditional efficacy of PF is an irreplaceable compass for researchers developing modern applications. Due to widely traditional usage, PF has also been developed in modern applications as drugs, health care products, and skincare products.

5.2 Biological activities

5.2.1 Anti-oxidant activity

Before the 1990s, it was believed that PF contained a high level of ascorbic acid which is mainly responsible for antioxidant potency (Kapoor, 1989). A study later contradicted this claim, stating that PF did not contain any ascorbic acid at all and hydrolyzable tannins like emblicanin A and emblicanin B were responsible for its antioxidant function (Ghosal et al., 1996). However, a study in 2006 suggested that high levels of ascorbic acid may still be the main reason for the antioxidant activity of PF (Scartezzini et al., 2006). Assays including iron (III) reductive activity, iron (II) chelating activity, 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging, superoxide anion free radical scavenging, and hydroxyl free radical scavenging were conducted to evaluate the antioxidative properties of PF collected in four areas. The results showed that four PF samples exhibited varying degrees of antioxidant potential, regardless of different contents of ellagic acid, corilagin, gallic acid and ascorbic acid. Interestingly, the PF samples still had consistently potential in the antioxidant screens although the ascorbic acid level is not the highest (Poltanov et al., 2010). Bhattacharyya et al. investigated the effect of PF on rat brain frontal cortical and striatal oxidative free radical scavenging enzyme levels and reported that the antioxidant activity of PF could be due to tannins including emblicanin A, emblicanin B, pedunculagin, and punigluconin, showing ascorbic acid like properties (Bhattacharya A. et al., 2000). Overall, it is not difficult to conclude that strong antioxidant activity of PF is owed to the influence of various phytochemicals, such as ascorbic acid, hydrolyzed tannins, and phenolic acids, etc.

The antioxidative activity of PF mainly manifests as a scavenging effect on free radicals. Oxygen-derived free radicals, especially reactive oxygen species (ROS) including hydrogen peroxide, superoxide and hydroxyl compounds, are strongly inhibited by the polyphenols, flavonoids and polysaccharides in PF extracts (Shi and Yang, 1994; Li et al., 2010). ROS has been reported to be associated with diseases such as diabetes, Alzheimer’s disease, coronary heart disease, nephritis, cancer, arteriosclerosis, and diseases related to ageing (Chen and Kan, 2018). Related scholars evaluated the free radical scavenging ability in vitro anti-oxidant tests, which showed that PF extracts have strong metal chelating, reducibility, 2,2′-azinobis-(3-ethylbenzthiazoline-6-sulfonic acid (ABTS) scavenging and DPPH scavenging ability (Wei et al., 2011; Wang et al., 2019). PF also has a scavenging effect on hydroxyl radicals and superoxide radical O2 via Fenton Reaction (Zhang et al., 2011). Malondialdehyde (MDA) is the main product of lipid peroxidation in the free radical membrane, whereas superoxide dismutase (SOD) is an important antioxidant enzyme that defends ROS (Liu et al., 2019). Therefore, increasing SOD activity and decreasing MDA content can reduce oxidative stress. In vitro tests have confirmed that PF ethanolic extracts protected RAW264.7 cells from oxidative damage by increasing glutathione content and total SOD activity and suppressing MDA content (Li et al., 2020). The active extracts of PF have been shown to possess antioxidant properties in several experimental models, which are shown in Table 2. In addition, another study reported the antioxidant activity of PF kernels methanol extracts using the DPPH assay (IC50 15 μg/ml) and H2O2 scavenging assay (IC50 32 μg/ml) (Gupta et al., 2012). While the antioxidant activity of PF was explained by several studies using different solvent extracts, there are only a few in vitro studies on the antioxidant activity of the kernels and seeds coat. Thus, further research work should focus on the antioxidant activity of PF kernels and seeds’ coat waste.

The anti-aging effects of PF is also attribute to its strong antioxidant ability. PF polyphenols showed a strong protective effect against the aging process in the Caenorhabditis elegans model through increasing thermal resistance, extending lifespan by 18.35%, reducing the activity of enzymes acetylicholinesterase (AchE) by 34.71% and butyrylcholinesterase by 45.38% (Wu et al., 2022). The life span of drosophila can be extended by ingesting certain concentrations of PF powder, nutrient solutions and juices (Zhao et al., 2018). Moreover, the extraordinary effect of PF in reducing the toxicity of heavy metals is also attributed to the antioxidant effects, such as its inhibitive effect on Ar-induced oxidative damage and apoptosis of mouse splenocytes, thymocytes and liver cells (Singh et al., 2013; Singh et al., 2014). The remarkable antioxidant and anti-aging potential of PF could be implemented in the food and pharmaceutical industry.

Although PF has proven its free radical scavenging activity in vivo and in vitro, it is difficult to achieve the desired outcome in the human body because of its low bioavailability like other herbal formulations. Researchers are developing various nanoformulations of PF extracts to overcome such bottlenecks. Rosarin et al. synthesize silver nanoparticles (AgNPs) loading with the aqueous extracts of PF, and it is reported that the IC50 value of PF (alone) and AgNPs is 30μg/mL and 20μg/mL.
TABLE 2 Anti-oxidant active components of PF extracts.

| Extracts            | Active compounds/ monomer | Models and dosage range | Positive control | Antioxidant performances | References                      |
|---------------------|---------------------------|-------------------------|------------------|-------------------------|---------------------------------|
| Aqueous Tannins/- Flavonoids/- | RAW 264.7 cells stimulated with LPS (5 µg/ml) for 6 h. 0.125-2 mg/ml | Ascorbic acid (100 µM) | Dose-dependently decrease of DPPH, ROS, iNOS and COX-2 production | Wang et al. (2019) |
| Ethanol | Tannins/gorilagin Phenolic acids/gallic acid and ellagic acid | RAW264.7 cells stimulated with H2O2 (100-2,400 µmol/L) for 24 h. 0.16-1.56 mg/ml | 1-Acetic acid (0.16-2.56 mg/ml) | Increase of glutathione content and total superoxide dismutase activity, and decrease of MDA content | Li et al. (2020) |
| Aqueous/ethyl acetate | Flavonoids/quinic acid, gallic acid, and quercetin | Red blood cells (physiological) pretreated with glucose (5 or 50 mM) for 24 h. 50-200 µg/mL | _ | Improvement of the plasma MDA, protein carbonylation, total protein, and albumin levels | Packirisamy et al. (2018) |
| Ethanol | Flavonoids/- | C. elegans stimulated with H2O2 solution (20 mMol/L). 1-4 mg/ml | _ | Enhancement of thermal resistance, exercise capacity and lifespan | Wang et al. (2018) |
| Ethanol | Phenolics/gallic acid, chebulic acid and ellagic acid | C. elegans stimulated with H2O2 sol. 0-1.2 mg/ml | Ascorbic acid (0-0.15 mg/ml) | Enhancement of thermal resistance, extension, of lifespan and inhibition of AChE and BuChE activity, increase of SOD and CAT and decrease of MDA. | Wu et al. (2022) |
| Ethanol | Phenolics/gallic acid and ellagic acid. Tannins/isocorilagin, chebulanin and chebulagic acid | Mice were exposed to arsenic (sodium arsenite 3 mg/kg p.o.) for 28 days. 500 mg/kg p.o | _ | Decrease of the lipid peroxidation, ROS production, the activity of caspase-3; increase of SOD,GSH, and CAT. | Singh et al. (2013) |
| Aqueous | Phenolics/gallic acid Flavonoids/- | Male Sprague Dawley rats induced by iodinated contrast agents (1,600 mg iodine/kg). 125 or 250 or 500 mg/kg/day for 5 days | Ascorbic acid and gallic acid (6.25, 12.5, 25, 50 and 100 µg/ml). Quercetin (25, 50, 100, 200 and 400 µg/ml) | Significantly decrease of MDA, and increase of total antioxidant capacity, SOD and CAT. | Tasanarong et al. (2014) |
| Methanol | Phenolics/- | Sprague Dawley rats induced by indomethacin (20 mg/kg p.o.) for 2 days. 100 mg/kg body-weight for 10 days | _ | PF (100 mg/kg) significantly decreased the level of MDA and SOD. | Randypadhyay et al. (2000) |
| Aqueous | Tannins/- | 0.39-12.5 µg/mL, The DPPH radical scavenging and reactive oxygen species inhibition assay | Ascorbic acid (100 µM) | A free radical scavenging activity (IC50 = 2.37 µg/ml), and protection of cells from ROS (IC501.77 µg/ml) | Majeed et al. (2019) |

respectively (Rosarin et al., 2013). However, studies on the use of these nano-formulations in clinical applications are very limited so far. Thus the use of nanotechnology to enhance the therapeutic value of PF may be one of the future research prospects for scientists and clinicians.

5.2.2 Analgesic, antipyretic, and anti-inflammatory activities

Phenolics, flavonoids and ascorbic acid in PF interfere with multiple processes in the pathogenesis of inflammation, such as the production of pro-inflammatory mediators, the expression of adhesion molecules, the adhesion of circulating leukocytes to the endothelium, and NF-κB activation (Lampronti et al., 2008; Saito et al., 2008; Muthuraman et al., 2011). In vitro studies showed that PF alleviated lipopolysaccharide (LPS)-induced inflammation in RAW 264.7 cells by decreasing the release of pro-inflammatory mediators, the expression of inducible nitric oxide synthase, cyclooxygenase-2 (COX-2) and NF-κB (Shi and Yang, 1994; Li et al., 2020). Another in vitro study found that PF prevented the further development of the inflammatory response in PAO1-induced IB3-1 CF bronchial epithelial cells by inhibiting the expression of the PAO1-dependent neutrophil chemokines interleukin-8 (IL-8), growth-regulating oncogene alpha (GRO-α) and GRO-γ, the adhesion molecule intercellular adhesion molecule (ICAM) 1 and the pro-inflammatory cytokine interleukin-6 (IL-6) (Nicoli et al., 2008). Furthermore, PF has a therapeutic effect on a variety of inflammatory diseases. PF aqueous extracts have a significant protective effect on the cartilage in patients with osteoarthritis by inhibiting hyaluronidase and type II collagenase activities (Sumantran et al., 2011). PF extracts inhibited the inflammatory response to benzo[a]pyrene (BaP)-induced acute lung injury by significantly reducing the levels of tumour necrosis factor-α (TNF-α), IL-6, and macrophage inflammatory protein 2 (MIP-2) in BaP-induced lung cancer mice (Wang et al., 2017). PF extracts also have a good therapeutic effect on...
acute necrotizing pancreatitis. It can reduce serum lipase and IL-10 levels, and significantly increase the nucleic acid content, DNA synthesis rate, pancreatic proteins and pancreatic amylase contents in rats with sexual necrotizing pancreatitis (Sidhu et al., 2011).

Modern pharmacological studies have confirmed that the ethyl acetate, petroleum ether and n-butanol components in the methanol extracts of PF are potent anti-inflammatory and analgesic agents. These anti-inflammatory factors treat gouty arthritis by inhibiting the release of the inflammatory factors including prostaglandin E2 (PGE2) and TNF-α, which may relieve the symptoms of redness, fever and severe pain that occur during gout attacks (Zeng and Cen, 2012). Dong Wook Lim et al. established a postoperative pain model (PI) produced by plantar incisions and a neuropathic pain nerve injury (SNI) rat model. Results found that PF extracts reduced the number of ultrasonic vocalizations in response to PI-related post-operative pain and hypersensitivity in response to von Frey stimulation of the hind paw in the PI model rats, as evidenced by an increased mechanical withdrawal threshold in the SNI-related neuropathic pain model rats. And PF extracts also significantly decreased pain-related pro-inflammatory cytokine levels in the dorsal root ganglion caused by neuropathic pain in SNI rats (Lim et al., 2016). The inhibitory effect of PF on the synthesis and/or release of pain and inflammatorymediators is similar to that of non-steroidal anti-inflammatory drugs (NSAIDs) (Golechha et al., 2014). Although the anti-inflammatory and analgesic effects of PF have been well studied, its inflammatory response in allergic reactions and immune-related diseases have not been carefully investigated. Besides, more research is needed to develop greener, safer, and more effective nano-anti-inflammatory drugs using PF as a raw material.

5.2.3 Antimicrobial activity

Many in vivo and in vitro experimental studies have confirmed the powerful antibacterial activity of PF. Alkaloid components of the PF alcoholic extracts have significant antibacterial activity against Gram-positive (G+) bacteria, such as *Bacillus cereus*, *Bacillus subtilis*, *Staphylococcus aureus*, *Salmonella typhi*, *Salmonella paratyphi*, *Pseudomonas aeruginosa*, *Shigella boydii*, and some fungi, *Candida albicans*, *Sacharomyces cerevisiae*, and *Aspergillus niger*, were also included (Rahman et al., 2009). Tannoids, saponins, flavonoids, and terpenoids from the aqueous extracts of PF also exhibited potent antimicrobial activity against *Enterobacter cloacae*, *Escherichia coli*, and *Klebsiella pneumonia* (Saeed and Tariq, 2007; Kumar A. et al., 2011). The chloroform soluble fraction of the PF methanolic extracts exhibited an inhibitory effect against some G+ and Gram-negative (G−) pathogenic bacteria and strong cytotoxicity with an IC50 of 10.257 ± 0.770 μg/ml (Rahman et al., 2009). MgO nanoparticles and silver nano-particles loading with the extracts of PF which were synthesized through green synthesis showed potential anti-microbial activity against various pathogenic bacteria such as *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Escherichia coli* and *Bacillus subtilis* (Ramanujam and Sundrarajan, 2014; Ramesh et al., 2015). Due to the effects against resistant strains, nano-formulations synthesized using PF has the potential to be developed as the carriers for antibiotics. In traditional medicine, Basant (a mixture of herbs containing PF extracts, curcumin, soapberry saponin, aloxe vera and rose water) inhibits the growth of strains and clinical isolates of Neisseria gonorrhoeae, among which are resistant to penicillin, tetracycline, nalidixic acid or ciprofloxacin (Dong et al., 2014). Moreover, PF essential oil extracted by Zhao et al. using the supercritical CO2 extraction technique showed good inhibition of common food contaminating bacteria (*Bacillus subtilis* (G+), *Staphylococcus aureus* (G+), *Salmonella* (G−)), and the inhibition effect on G+ bacteria was greater than that on G− bacteria (Zhao et al., 2007). Together, these results suggest that PF can be used as a plant-derived antimicrobial agent in the clinical control of drug-resistant pathogens and has the potential to control food-contaminating bacteria. However, the specific antibacterial components and mechanism of PF are not yet clear and need in-depth study. Similarly, there are few studies on the antifungal properties of PF, so further studies are required to find out the antifungal active ingredient.

5.2.4 Anti-viral activity

Previous investigations revealed that PF has inhibitory effects against human immunodeficiency virus (HIV), H1N1 influenza virus, herpes simplex virus (HSV), human papillomavirus (HPV), hepatitis B Virus (HBV), and severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). PF methanol extracts and water extracts have a good inhibitory effect on HIV reverse transcriptase and H1N1 infection (Sun et al., 2002; Kong et al., 2016). 1,2,4,6-tetra-O-galloyl-β-D-glucose from PF has anti-HSV activity in vitro (Xiang et al., 2011). The anti-HSV-1 and anti-HSV-2 effects of polyphenols isolated from PF extracts were confirmed in a guinea pig cutaneous HSV-1 infection model, cytopathic effect observation method, and thrombocytopenia assay. And the mechanism may be direct inactivation of the virus and inhibition of early viral DNA replication, thereby inhibiting HSV virus-induced cytopathy (Qu et al., 2010). PF also has a good inhibitory effect on HPV-16 and HPV-18 viruses by dose-and time-dependent reduction of DNA binding activity of constitutively active activator protein-1 in HeLa cervical cancer cells (Mahata et al., 2013). Elaeocarpususin and digallic acid in PF ethanolic extracts showed remarkable anti-HBV activity. These compounds screened by HBV-DNA transfected HepG2 2.1.5 cell model showed strong inhibition for the secretion of HBeAg and HSeAg with IC50 of 10.2, 79.0 μg/ml and 5.5, 101.0 μg/ml, respectively (Hou, 2006). Moreover, numerous computational screening studies have mapped out the potential bioactive compounds from medicinal plants against the targets of severe acute respiratory syndrome coronavirus 2
(SARS-CoV-2) (Siddiqui et al., 2020; Natesh et al., 2021). PF extracts contain bioactive compounds such as flavonoids, phenols, tannins, and alkaloids with good antiviral activity against SARS-CoV-2, which may guide the development of novel anti-neoplastic pneumonia prophylactic drugs (Krupanidhi et al., 2020; Kumar et al., 2020). Murugesan’s study also found that PF quercetin has a good binding effect on SARS-CoV-2 main protease and could be developed as a promising inhibitor of SARS-CoV-2 main protease (Murugesan et al., 2021). However, the technical difficulties in synthesizing complex natural compounds with good antiviral activity from PF need to be solved and the antiviral activity of PF needs to be validated in more animal and clinical studies.

### 5.2.5 Anti-tumour activity

Nowadays, plant-originated natural products constitute a considerable proportion of commercial anticancer drugs (Shynu et al., 2011). The anticancer properties of some molecules identified by high performance liquid chromatography (HPLC) from PF extracts have been repeatedly demonstrated (Table 3). Corilagin has significant growth inhibition on human nasopharyngeal carcinoma cells kB, ovarian carcinoma cells A2780, osteosarcoma cells OS-732, human lung adenocarcinoma cells A549, mouse liver cancer H22 tumour, murine Lewis tumour and human HCT-8 (Liu et al., 2002). Geraniin can induce the apoptosis of human melanoma cell A2058 and inhibit the proliferation of human lung adenocarcinoma cell A549 (Lee et al., 2008; Li et al., 2013). Chebulic acid has obvious anti-leukemia, anti-cervical cancer and anti-colon cancer HT-29 cell effects and can inhibit the growth of malignant tumour (Yi et al., 2004). Chebulagic acid also has a good inhibitory effect on the growth of eye cancer cells and HSC-T6 hepatocytes by inhibiting vascular endothelial growth factors (Athira et al., 2013; Chuang et al., 2011; Kumar, N. et al., 2014). These results suggested that tannins might be responsible for the growth inhibition of cancer cells. However, animal and clinical trials of PF active monomers as complementary drugs for cancer treatment and prevention are lacking. Interestingly, crude PF extracts are also capable of tumour inhibition. PF extracts enriched with polyphenols or aqueous extracts have shown cytotoxic activity against cervical and ovarian cancer cell (De et al., 2013; Zhu et al., 2013). In a murine model of skin carcinogenesis, continuous administration of PF extracts at 100 mg/kg reduced tumour incidence by ~60% (Sancheti et al., 2005). The pre-clinical experiments of various cancer cell lines (A549 [lung], HepG2 [liver], HeLa [cervix], MDA-MB-231 [breast], SK-OV-3 [ovary] and SW620 [colorectal]) showed that the water extracts of PF (50–100 μg/ml) can induce apoptosis by activating caspase-8 and caspase-3/7 and upregulating Fas expression (Ngamkitidechakul et al., 2010). PF extracts also sensitized refractory high-grade serous ovarian cancer (HGSOC) cells with high drug resistance by regulating key signalling molecules of angiogenesis and metastasis (De et al., 2020).

How do these extracts exert anti-tumour effects? There are several possibilities. First of all, PF can prevent DNA damage and tumour occurrence caused by ROS (Majeed et al., 2009; Hazra et al., 2010). However, it is unclear from the current findings to what extent ROS contribute to the underlying pathology. Secondly, many of the anticancer properties of PF extracts may be brought about through inhibition of this transcription factor binding with its cognate DNA binding elements (Hoesel and Schmid, 2013). Thirdly, PF extracts have anti-inflammatory activity that may prevent related cancers caused by inflammation (Golechha et al., 2014). Fourth, the antitumour activity of PF may be due to the blocking cell cycle in the G2/M phase (Huang et al., 2017). Either crude PF extracts or purified components show effective cytotoxicity to most cancer cell lines, but its resistance mechanism still exists. Remarkable results in a study were not reported when PF extracts were examined for liver tumours induced by initiation with diethylnitrosamine followed by promotion with 2-acetylaminofluorene (Sultana et al., 2008). Although a large number of studies have reported the beneficial effects of PF for the treatment of cancer, these results are still in the cell and animal experimental stage. The anti-tumour effects of PF are yet to be explored in more clinical trials to develop effective new anti-cancer drugs.

### 5.2.6 Immunomodulatory and anti-fatigue activities

The activation of the immune system is an effective protection mechanism against internal and external threats. In vitro and in vivo studies have been performed on the extracts of PF and bioactive constituents to evaluate their immunomodulating effects. The tannins and polyphenols responsible for immunomodulatory properties of PF included gallic acid, ellagic acid, corilagin, chebulagic acid, and quercetin (Scartezzini and Speroni, 2000). PF has been demonstrated to regulate the immune system via various mechanisms. Chebulagic acid found in PF has been reported to inhibit the NF-κB signaling pathway. Impaired regulation of NF-κB may lead to various immune/non-immune related disorders including cancer, inflammatory, and autoimmune diseases (Hamada et al., 1997). Thus, the immunomodulatory effects of PF could be its potential to regulate the NF-κB signaling pathway. Diethyl ether extracts of PF potentially suppressed calcium ionophore A23187-induced LTβ4 release from human neutrophils and TXB2 release in platelets (Vormisto et al., 1997). Ram et al. reported that Chromium (VI) induced cellular stress and immunosuppressive effects on lymphocyte proliferation were ameliorated following the treatment with PF and it also restored the altered levels of IL-2 and γ-IFN (Ram et al., 2002). Luteolin and apigenin are important flavones found in PF and were able to modulate the immune system by attenuating LPS-induced splenocyte proliferation and enhancing humoral immune responses (Cui et al., 2020).
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Diseases because of the immunomodulatory activities.

Application values for the treatment and prevention of various diseases because of the immunomodulatory activities.

PF. The research results showed that a low-dose extracts of PF was established to observe the effect of reducing glycogen consumption and controlling fatty energy metabolism (0.167 g/kg), only liver glycogen significantly increased, whereas in the high-dose group (11.67 g/kg), there was a significant increase in exhaustive swimming time (Zhang et al., 2011). These results indicate that PF extracts have broad application prospects for improving the working ability of the plateau.

**5.2.7 Hypoglycemic and hypolipidemic activities**

There is increasing evidences that PF may have a positive role in controlling disorders of blood glucose and lipid metabolism. Polysaccharides in PF extracts can significantly improve insulin resistance or β-cell dysfunction, thus, having a therapeutic effect on type 2 diabetes (Dong et al., 2009). Polysaccharides also remove OH, O₂-, and DPPH, which shows down the oxidative stress of patients with diabetes and effectively prevents diabetic complications (Wang et al., 2013; Guo et al., 2014). Favoronoids from PF exhibited significant hypoglycemic activity. Its main mechanisms of action are to maintain β-cell activity, inhibit β-cell apoptosis and insulin-like effects, promote peripheral tissue regulation of glucose metabolism, maintain biofilm stability through antioxidation, protect pancreatic tissue and regulate the release and activity of enzymes related to glucose metabolism (Kang et al., 2011).

**TABLE 3 The anti-tumour mechanisms of active components in PF extracts.**

| Extracts | Active compounds/monomer | Anti-tumour mechanisms | Anti-tumour indicators | References |
|----------|--------------------------|------------------------|------------------------|------------|
| Ethanol  | Phenolics/gallic acid     | 1. Down-regulation of ERK/p-ERK signalling pathway | 1.↑ Proliferation and tubule formation of human cervical cancer cells HeLa and HTB-35 | Zhao and Hu., 2013 |
|          |                          | 2. The activation level of ERK1/2 was down-regulated | 2.↑ The proliferation of human osteosarcoma cell lines U-2OS and MNG/HOS | Liang et al. (2012) |
| Methanol | Tannins/1,2,3,4,6-penta-O-galloyl-β-glucose | 3. Activation of the MAPK8/9/10 and JNK signalling pathways | 3.↑ Autophagy occurred in HepG2, MCF-7 and A549 tumour cells | Dong et al. (2014) |
| Ethanol  | Flavonoids/progallin A Tannins/... | 4. G₁/M and G₂/M cell cycle arrest was induced | 4.↑ The proliferation of human liver cancer cell BEL-7404 | Zhong et al. (2011) |
| Ethanol  | Phenolics/penta-1,2,3,4,6-O-galloyl-β-o-glucose and geraniin | 1. The expression of key enzymes MMP-2 and MMP-9 in tumour cells were inhibited | 1.↑ Proliferation, migration, invasion and adhesion of human fibrosarcoma HT1080 cells | Sancheti et al. (2005) |
|          |                          | 2. Inhibition of Akt and ERK1/2 activities | 2.↑ Proliferation and migration of rat glioma cell C6 and human glioma cell U373 | Li et al. (2020) |
| Ethanol  | Phenolics/gallic acid     | 3. Inhibition of tumour angiogenesis | 3.↑ Angiogenesis and growth of new blood vessels (Human Placental Vein Angiogenesis Model) | Ji. (2010) |
| Ethanol  | Ascorbic acid             | 1. NK cell activity and antibody-dependent cell-mediated cytotoxicity were significantly enhanced | 1.↑ The lifespan of immunodeficient mice vaccinated with Dalton’s lymphoma acutes | Suresh and Vasudevan, 1994 |
|          | Tannins/corilagin         | 2. The expression of the E3 ubiquitin ligase RING finger protein 8 were attenuated | 2.↑ Proliferation and migration of esophageal squamous cell carcinoma | Qiu et al. (2019) |
| Aqueous  | Tannins/corilagin         | 1. The conduction of the TGFβ/AKT/ERK/Smad signalling pathway was targeted to inhibit | 1.↑ The growth of ovarian cancer cells | Liu et al. (2002) |

et al., 2010; Wang et al., 2014). The investigation has also shown the ability of these compounds to enhance natural killer (NK) cells and cytotoxic T lymphocytes (CTL) activities. Thus, the immunomodulatory properties of PF extracts could be due to the synergistic effects of these cells. In TCM, PF in combination with *Radix Plantarum*, *Astragalus spp.*, *Salvia miltiorrhiza*, *Panax notoginseng*, *Ginkgo biloba* and *Radix Puerariae* can improve immunity, reduce blood lipid and protect the heart and cerebrovascular integrity (Zhu et al., 2018). PF also has an excellent protective effect on immune organs while inhibiting tumour growth in mice via the research methods of TCM serum pharmacology (Luo, 2010). PF showed a wide range of application values for the treatment and prevention of various diseases because of the immunomodulatory activities.

PF has an anti-fatigue effect by increasing glycogen stores, reducing glycogen consumption and controlling fatty energy supply. An anti-fatigue evaluation model of the simulated plateau-exposed mice was established to observe the effect of PF. The research results showed that a low-dose extracts of PF (0.383 g/kg) can significantly prolong the exhausted swimming time of mice, promote the elimination of lactic acid after exercise and increase liver glycogen reserves. In the middle dose group (0.167 g/kg), only liver glycogen significantly increased, whereas
the elevation of lecithin-cholesterol acyltransferase by flavonoids of PF (Anila and Vijayalakshmi, 2002). PF extracts have the same effect as the anti-diabetic drug chlorpropamide in reducing serum glucose and glycosylated haemoglobin (HbA1C) (Daisy et al., 2009). When PF was used in combination with hypoglycaemic drugs, it was significantly more effective in controlling blood glucose and lipid levels than hypoglycaemic drugs alone (Linda and Mary, 2014). A 21-day clinical study showed that PF reduced fasting and 2-h postprandial blood glucose levels and cholesterol and triglyceride (TG) levels in patients with diabetes (Akhtar et al., 2011). The alcohol-extracted part of PF could increase the biosynthesis of GLUT4 by enhancing the expression of GLUT4 mRNA in skeletal muscle and accelerating the transmembrane transfer of glucose and the utilization of glucose by peripheral tissues, thereby improving insulin resistance and treating diabetes (Kang et al., 2011). Moreover, some studies showed that a regular intake of PF powder can effectively reduce levels of total cholesterol, LDLs, TGs, and very-low-density lipoproteins (VLDLs) and increase levels of high-density lipoproteins (HDLs) in patients with type II hyperlipidemia by mechanisms similar to that of simvas-tatin (Gopa et al., 2012). A study by Balusamy et al. suggested that the digallic acid of PF showed significant anti-lipolytic activity compared to the standard drug orlistat. In the mature adipocytes, digallic acid significantly reduces TG accumulation by down-regulating adiponectin, peroxisome proliferator-activated receptor γ (PPARγ), CCAAT/enhancer-binding protein α, and fatty acid-binding protein 4 (FABP4), and triggers adipocyte apoptosis by up-regulating the expression of Bax/Bcl-2 (Balusamy et al., 2020). Hence, PF negatively regulates adipogenesis by initiating fat cell apoptosis and therefore it can be considered a potential herbal medicinal product for treating obesity.

Hyperglycaemia and hyperlipidemia interact with each other and exacerbate each other. Both endanger the health of the heart, brain, kidneys, and other organs. Many scholars have found through animal studies and clinical trials that the bioactive components from PF can prevent complications caused by diabetes and dyslipidemia through different mechanisms (Table 4). At present, most of the existing literature focuses on research into the therapeutic mechanisms of certain compositions or extracts of PF, which may show the synergistic effects of multiple compositions. Although PF contains hundreds of compounds, current reports focus on polysaccharides, ellagic acid, gallic acid, chebulic acid, etc. However, it does not mean that other compounds have no effects. Moreover, PF has preventive and therapeutic effects on diabetes and hyperlipidemia, but there is still a lack of sufficient evidence in as food and dietary supplement. We hope that PF and related products can provide a new healthy diet reference for high-risk groups and patients with diabetes in the future.

5.2.8 Neuroprotective activity

PF has been considered to exert therapeutic effects on neurological ailments due to its antioxidant, cholesterol-lowering and anti-inflammatory properties. The ethanol extracts of PF can increase the levels of SOD, CAT, and glutathione peroxidase (GSH-Px) in rat brain tissue homogenates and decrease the level of tributyltin compounds and the activity of AchE. Therefore, PF can improve the cognitive ability of rats (Uddin et al., 2016). Ellagic acid and gallic acid from PF methanol extract improved the performance of Alzheimer’s disease rodents in neuro-behavioural tests by inhibiting AchE activity (IC50 < 100 μg/ml) and scavenging ability of DPPH (IC50 < 10 μg/ml) (Mathew and Subramanian, 2014; Uddin et al., 2016). A previous study established a depression mouse model using a tail suspension test and forced swimming test, then this research observed that PF has significant anti-depressant activity, and its efficacy is comparable to imipramine and fluoxetine. In addition, a reduction in the monoamine oxidase-A enzyme in the mouse brain after using antagonists of the α1 adrenergic, serotonin, dopaminergic D2, and γ-aminobutyric acid-b receptors caused the anti-depressant activity of PF to be ineffective. Hence, PF might interact with these receptors to exhibit anti-depressant activity (Dhingra et al., 2012). Moreover, PF ethanol extracts have antiepileptic activity by reducing glucose and cortisol levels in the brain tissue of mice in a restraint stress model and improving the performance in the elevated plus-maze test (Kumar, C.P. et al., 2014). Kainic acid of PF attenuated global tonic seizures and persistent status epilepticus in pentyleneterazole-induced epileptic rats, thereby improving the cognitive dysfunction, which may be related to the decrease of TNF-α levels in rat brain tissue (Golechha et al., 2010; Golechha et al., 2011). Furthermore, PF can resist the toxicity and cognitive impairment induced by aluminum chloride in rats by regulating oxidative stress and reducing the expression of apoptosis markers including Bax, caspases-3, caspases-9, cytosolic cytochrome c and p-Tau (Thenmozhi et al., 2016; Bharathi et al., 2019). PF also has a significant neuroprotective effect on oxidative DNA damage of neurons induced by H2O2 (Ramakrishna et al., 2014). The combined treatment with PF, Arrowshaped Tinospora Root, and Ocimum tenuiflorum also improved learning and memory disorders caused by scopolamine, diazepam and cyclosporin (Malve et al., 2014). In short, PF extracts have been hypothesized to act through multiple neuroprotective pathways. To date, most of the studies have shown neuroprotective effects of PF crude extracts. However, only a few research focuses on the molecular mechanisms by which the active monomers of PF act on neural cells. Further studies on the neuroprotective effects of PF extracts and monomers are crucial to uncovering the molecular mechanisms and gene regulation underlying these effects. Furthermore, given the neuroprotective properties exhibited by PF, more researches will focus on the clinical
| Complications                        | Extracts                      | Active compounds/monomer | Positive control | Mechanisms                                                                 | Improvement indicators                                                                 | References                          |
|-------------------------------------|-------------------------------|--------------------------|------------------|--------------------------------------------------------------------------|----------------------------------------------------------------------------------------|-------------------------------------|
| **Cardiovascular diseases**         | Aqueous                       | Phenolics/ellagic acid and gallic acid (20 mg p.o.) | Simvastatin capsule | 1. Reduction of oxidative stress and ROS production in cardiomyocytes  
2. Regulation of adipokines and improvement of lipid metabolism disorders  
3. Reduction of the advanced glycation end products (AGEs) production | Animal experiment: LDH ↓, creatinine kinase MB (CK-MB) ↓, SOD ↓, GSH ↓, serum creatinine (Scr) ↓, left ventricular wet LV/BW and HW/BW ratios ↓, blood pressure (BP) levels ↓ | Gopa et al. (2012)                                                                 |
|                                    | Aqueous                       | Phenolics/Tannins/gallotannin |                 | 4. Regulation of cardiac autonomic dysfunction  
5. Improvement of glucose utilization and maintaining of glucose homeostasis by restore and regenerate β-cell structure | Clinical experiments: TC ↓, LDL ↓, TG ↓, VLDL↓, HDL↑ | Patel and Goyal, (2011)                                                               |
| **Nephropathy**                     | Methanol                      | Tannins/gallotannin, Phenolics/ellagic acid | Insulin (3 IU/kg, s.c.) | 1. Oxidative stress-induced apoptosis was reduced  
2. The production of AGES was reduced  
3. Over-activation of poly-ADP-ribose polymerase and mRNA expression of aldose reductase in the kidney were inhibited  
4. The release of renal inflammatory cytokines was inhibited  
5. The activity of renal aldose reductase (AR) was inhibited | Animal experiment: serum urea ↓, uric acid ↓, Scr ↓, total protein (TP) ↓, albumin (Alb) ↓, aldose reductase (AR) ↓, sorbitol dehydrogenase (ID)↓, HbA1c ↓, urinary glycosylated albumin ↓, renal carboxymethyl lysine ↓, pentoside ↓, sorbitol ↓, fructose level ↓, IL-6↓, IL-1β ↓, TNF-α↓ and MCP-1 ↓ in kidney tissue | Chandak et al., 2009; Chao et al., 2010; Latha and Daisy, 2011 |
| **Neuropathy** (neuropathic pain)  | Aqueous                       | Phenolics/gallic acid     | Insulin (10 IU/kg, s.c.) | 1. Scavenging free radicals and anti-oxidation  
2. Inhibition of hyperglycemia leading to reactive dicarbonyl formation of AGES  
3. Regulation of lipid metabolism and axon degeneration in the brain | Animal experiment: J TNF-α, IL-1β and TGF-β1 levels in the serum and sciatic nerve of diabetic rats, regulation of oxidative-nitrosation stress in diabetic rats, neuropathic pain | Tiwari et al. (2011)                                                                 |
| **Diabetic cataract**               | Aqueous                       | Tannins/β-glucogallin, Flavonoids/quercetin (50 µg/ml p.o.) | Quercetin | 1. Aldose reductase activity and sorbitol formation were significantly inhibited | Animal experiment: J eye's lens high pressure, ↓ the accumulation of galactitol, sorbitol in red blood cells, lens and sciatic nerve | Suryanarayana et al. (2004)            |
|                                    | Methanol                      | Phenolics/ellagic acid    |                 | 2. Polyol-induced oxidative stress pathways in the lens are altered  
3. Inhibition of hyperglycemia-induced aggregation and insolubility of lens proteins | Rat lens organ culture model: normalization of the altered a-crystallin profile, and preservation of a-crystallin chaperone activity | Kumar et al. (2011b)                                                               |
| **Protein wasting**                 | Methanol                      | Phenolics/ellagic acid, and gallic acid | Insulin (3 IU/kg, s.c.) | 1. Inhibition of oxidative stress caused by hyperglycemia  
2. Improvement of insulin insensitivity or insulin resistance to maintain glucose homeostasis  
3. Inhibition of α-glucosidase | Animal experiment: J total hemoglobin (HB), maintaining food and water intake, and maintaining body weight | D’Souza et al., 2014; Patel and Goyal, 2011; Wang, 2017 |

(Continued on following page)
effects of PF on depression, epilepsy, and neurodegenerative disorders in future.

5.2.9 Multiorgan functional protective activity

Over 40 years of scientific research on PF have confirmed the protective effect on many organs. Inflammation and oxidative stress play a major role in liver injury. And the phytoconstituents viz. ascorbic acid and gallic acid present in PF protect the hepatocellular environment against toxic moieties generated from oxidative stress or inflammation (Chen et al., 2011). PF extracts significantly increased hepatic SOD and GSH-Px activities, and inhibited MDA and ROS production to protect the liver by regulating protein kinase (AMPK) and acetyl coenzyme a carboxylase signaling pathways (Malar and Mettilda, 2009; Yang et al., 2016). PF also reduced liver coenzyme a carboxylase signaling pathways (Malar and the liver by regulating protein kinase (AMPK) and acetyl activities, and inhibited MDA and ROS production to protect antioxidative stress or inflammation. And the phytoconstituents protective effect on many organs. In TCM, PF along with the combination of other herbs is widely used to treat wheezing coughs, eruptions and other respiratory disorders (Sai et al., 2010). Flavonoids in PF extracts attenuate the development of lung inflammation in B(a) p or H2N1 influenza virus-infected mice by reducing the levels of pro-inflammatory cytokines MIP-2, TNF-α, IL-6, and IL-1β in lung tissue and the number of lymph nodes on the lung surface. (Kong et al., 2016; Wang et al., 2017). PF mixture improves the clinical efficacy and lung function of patients with acute exacerbation of chronic obstructive pulmonary disease (Chen and Jing, 2012).

PF showed in numerous animal studies to be cardioprotective and anticoagulant (McCord, 1985; Bhatia et al., 2011). The cardioprotective effect of PF is attributed to its high content of antioxidant active substances (Bhatia et al., 2011). For example, ascorbic acid reacts with singlet oxygen radicals and hydrogen radicals to produce semi-dehydroascorbic acid, which inhibits oxidative modification of LDL and improves atherosclerosis-induced endothelial damage (Villacorta et al., 2007). PF inhibited the accumulation of intracellular oxidants and the conversion of phosphorylated P53 protein (phospho-P53) in the nucleus by upregulating the PI3K/Akt/GSK3β-catenin cardioprotective signaling pathway and downstream cascade reactions, thereby reducing endothelial cell apoptosis induced by oxidative load and inhibiting cardiomyocyte necrosis and fibrosis (Liu et al., 2007a; Thirunavukkarasu et al., 2015). In TCM research, the Tibetan medicine prescription Wwwei Yugenzi plays a role in protecting the myocardium by regulating the expression of Bcl-2 and Bax, increasing the ratio of Bcl-2/Bax expression and the level of NO in the myocardium of IRI rats (Tan et al., 2015). For example, the expression of OXPHOS complex and α1/2 adrenergic receptor gene, and the level of NO in the myocardium of IRI rats (Tan et al., 2015). 

Preclinical studies have shown the beneficial effect of PF in the treatment and/or prevention of various GI problems like ulcerative colitis and Crohn’s disease (Romano et al., 2014; Saxena et al., 2014). The ethanolic extracts of PF had a protective effect against NSAID-induced gastropathy by significantly increasing the levels of mucin and PGE2, decreasing the levels of pro-inflammatory cytokines (TNF-α, IL-1β) and up-regulating the levels of anti-inflammatory cytokines (IL-10) (Chatterjee et al., 2011). The gallic acid-rich fraction of PF extracts exerted therapeutic effects on indomethacin-induced gastric ulcer in mice by increasing the

### Table 4 (Continued) The possible mechanisms underlying PF extracts against hyperlipidaemia and diabetic complications.

| Complications | Extracts | Active compounds/monomer | Positive control | Mechanisms | Improvement indicators | References |
|---------------|---------|--------------------------|------------------|------------|------------------------|-----------|
| Hepatopathy   | Methanol | Flavonoids/epigallocatechin gallate | Simvastatin (1.77 mg/kg p.o.) | 1. Antioxidant activity in liver; anti-vascular oxidative stress | Animal experiment: [mRNA expression of IL-1β, IL-6, TNF-a, and monocyte chemoattractant protein-1 (MCP-1), [serum resistin levels, [plasma total cholesterol and triglyceride level, [plasma ALT and AST levels, [hepatic triglyceride and cholesterol content | Yoshimura et al., 2013; Liu et al., 2015b; Ding et al., 2015 |

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levels of pro-angiogenic factors like PGE2, vascular endothelial growth factor (VEGF), hepatocyte growth factor, von Willebrand Factor VIII and endothelial NOS (Chatterjee et al., 2012). In addition, the chloroform part and ethyl acetate part of PF extracts can increase gastrointestinal motility and act as a laxative by activating cholinergic receptors; therefore, it is used to treat indigestion and constipation (Mehmood et al., 2013). PF is reported to have diuretic potential. PF has a nephroprotective effect on producing nephrogenesis by significantly reducing arterial blood pressure, blood urea nitrogen, and creatinine levels and decreasing the expression of iNOS, COX-2, and NF-κB in aged rats (Yokozawa et al., 2007).

5.2.10 Others

PF contains a high concentration of ascorbic acid, which can prevent and treat scurvy (Jain et al., 2015). PF extracts significantly antagonized the Vipera russelli and Naja kaouthia venom-induced lethality in vivo and in vitro experiments (Alamand and Gomes, 2003). Furthermore, its combination with high levels of ascorbic acid and polyphenols can promote wound healing by regulating collagen synthesis, extracellular matrix protein synthesis and oxidative stress (Sumitra et al., 2009).

6 Toxicology of Phyllanthi Fructus

Since ancient times, PF has been used for the treatment of diseases. In vivo and in vitro studies have verified its role in the inhibition of various pathogenesis. The results of the safety evaluation showed that the acute oral maximum tolerable dose of PF was greater than 20.0 g/kg b. w., which was practically non-toxic according to acute toxicity classification (Wen et al., 2018). Acute toxicity experiment showed no behavioural changes or symptoms of toxicity in rats given up to 2000 mg/kg b. w. of PF ethanol extract orally for 14 days (Uddin et al., 2016). A previous study has administrated rats with PF juice (up to 9 ml/kg/day) for 60 days. As a result, no significant changes in body weight, internal organs, or biochemical and hematological parameters were found (Chaiyasut et al., 2018). Chronic toxicity studies in rats with daily oral administration at doses of 300, 600, and 1,200 mg/kg for 270 days also did not observe obvious histopathology changes (Jaijoy et al., 2010). Moreover, both the micronucleus test in the bone marrow cells of mice and the mouse sperm aberration test suggested that PF had no genotoxicity to somatic or germ cells (Wen et al., 2018). In brief, no toxic effects of PF were observed as demonstrated by hematological examination, behavioral observation, and biochemical parameters suggesting the safe utilization of PF. Thus, PF has high safety for animals and humans under reasonable application, which is very suitable for drugs or healthy product development.

7 Quality control of Phyllanthi Fructus

Although PF has shown unique efficacy for the treatment of several diseases, there is still a big gap in the industrialization and standardization of PF. Currently, the Chinese Pharmacopoeia (2020 edition) requires that gallic acid in PF (anhydrous substance) is more than 1.2% to ensure medicinal quality (Chinese Pharmacopoeia Commission, 2020), which is still somewhat inadequate. First, gallic acid, found in many plants, is not a chemical component unique to PF. Second, the hydrolyzed tannins contained in PF can be hydrolyzed to gallic acid by heat, affecting the accuracy of the test results (Huang et al., 2019). Therefore, the measurement of gallic acid is not an adequate way to control the quality of PF. Depending on more adequate phytochemical and pharmacological studies of PF, more accurate quality control criteria will be reestablished after the exact active components of PF must be fully studied.

Actually, the quality of PF is closely related to multiple factors, such as germplasm, origin, harvesting, processing, preparation, storage and extraction conditions. There are differences in the quality of PF from different varieties and production areas. HPLC fingerprints were used to confirm that there were significant differences in gallic acid and quercetin in methanol extracts of PF among 6 producing areas (Guo et al., 2013). Salt soaking is one of the traditional processing methods for PF. The contents of gallic acid, ellagic acid and epicatechin were affected by the amount of water, the proportion of salt, and temperature during the salting process (Li et al., 2019). Modern processing methods of PF are mainly based on the removal of impurities and drying. Different drying methods, such as vacuum freeze drying and hot air drying at different temperatures, affected the content of gallic acid, corilagin and polysaccharides in PF (Mao, 2019). In addition, the gallic acid content of PF showed a trend of first increasing and then decreasing during storage. Hence, the single indicator is no longer sufficient to meet the needs of the whole industry chain. The quality evaluation of PF also should pay close attention to the place of origin, planting, production, and processing.

Quality control analysis methods, including Ultra-high performance liquid chromatography-multistage mass spectrometry (UPLC-MSn), ultraviolet spectrophotometry, HPLC, and gas chromatogram (GC)/mass spectrometry (MS) are used to analyze the active ingredients of PF. Wu et al. used the UPLC-MSn method to determine the content of tannins and luteolin in PF (Zhang et al., 2019; Wu et al., 2021). This method is high separation efficiency, but it has the disadvantage of time-consuming and costly. The content of gallic acid, epigallocatechin, corilagin, terminalia biphenyl acid, and ellagic acid in PF eluted by ethanol were also measured simultaneously using the HPLC method (Li et al., 2018). Wang et al. identified 43 volatile oil components in PF by GC-MS (Wang et al., 2009). Although the above methods have the advantages of high precision, accuracy, sensitivity, short analysis time, strong separation ability, good selectivity, and wide application range, most methods have the
| Table 5 | Traditional Chinese formulations containing PF. |
|---|---|
| **Formulation names** | **Main composition** | **Formulations, usage, dosage** | **Diseases or symptoms therapy** | **Prescription sources** |
| Er Shi Wu Wei Zheng Zhu Wan (二十五味珍珠丸) | *Pernulo* (20 g), *Myristicae Semen* (40 g), *Travertine* (100 g), *Fructus of Amomum tsao-ko* (30 g), the flower of *Syringa oblata* Lindl. (50 g), the center of the trunk of *Dulbergia adorfera* T. Chen (100 g), *Amomi Rotundus Fructus* (15 g), *Elettaria cardamomum* White et Maton (40 g), *Fructus of Choerospondias axillaris* (Roeh.), *Burr et Hill* (130 g), *Santalum album* L. (50 g), *Phyllanthi Fructus* (100 g), *Aquilaria spr.* (80 g), *Cinnamomum Cortis* | *Pill/oral administration/bid-tid/1.2–1.5 g each time* | Used for cerebral hemorrhage, cerebral infarction, poststroke syndrome, myocardial infarction and angina pectoris | Si Bu Yi Dian. AD773–783. Chinese Pharmacopoeia (2020) |
| Shi Ba Wei Du Juan Wan (十八味杜鹃丸) | *Bloododendron anthropogonoides* Maxim. (75 g), *Fructus of Amomum tsao-ko* (15 g), *Cevrot et Lemarie* (15 g), *Fructus of Terminalia chebula Retz.* (70 g), *Fructus of Terminalia bellirica* (Gaertn.) (Roeh.), *Phyllanthi Fructus* (60 g), the lumps in the stalks of *Bambusa textilis* McClure/Schizostachyum chinense Rendle (35 g), The dried flower of *Carthamus tinctorius* L. (50 g), the dried seed of *Myristica fragrans* Houtt. (15 g), the dried flower buds *Eugenia caryophyllata* Thunb. (20 g), *Fructus of Anomumkravanh* Pierre ex Gagnep. (15 g), the center of the trunk of a *Santalum album* L. (30 g), the trunk of *Pterocarpus santalinus* L.f. (100 g), the leaves of *Symplcostrum sumunita* Buch.-Ham. ex D. Don (40 g), *Rubia cordifolia* L. (35 g), the secretion of *Lacciferlacca* Kerr. (40 g), *Gentiana straminea* Maxim. (35 g), *Glycyrrhiza uraleensis* Fischer. (40 g), the wood of *Aquilaria sinensis* (Lour.) (55 g), *Gymnema* (10 g) | *Pill/oral administration/bid-tid/2g~2.5 g each time* | Used for muscular dystrophy, numbness of limbs, facial distortion, dysmenorrhea and amenorrhea | NMPA*. Tibetan Medicin* |
| Shi Wu Wei Cheng Xiang Wan (十五味沉香丸) | *Aquilaria spp.* (100 g), *Inula graveolens* (150 g), *Santalinum album* L. (50 g), *Pterocarpus indicus* wildl. (150 g), the flower of *Carthamus tinctorius* L. (100 g), *Myristica fragrans* (25 g), *Rhizome of Pogostephyton scaphifolium* (Hook.f.Thom.) Marq.et Shaw (150 g), *Stem of Rubus L.* (200 g), *Falllopia aubertii* (L. Henry) Holub (peeled) (100 g), *Asarum canadense* (50 g), *Travertine* (100 g), *Fructus of Choerospondias axillaris* (Roeh.) *Burr et Hill* (150 g), *Fructus of Terminalia chebula Retz.* (80 g), *Phyllanthi Fructus* (100 g) | *Pill/oral administration/bid-tid/1.5 g each time* | Used for coordinating qi and blood, relieving cough, and soothing the nerves. Treatment of qi stagnation and blood stasis, dry cough, shortness of breath, and insomnia | Chinese Pharmacopoeia (2020) |
| Ba Wei An Ming San (八味安宁散) | *Gypsum/Calcite* (400 g), *limestone* (100 g), the seed of *Soupis alba* L. (150 g), *Dracaucophalam tanguticum* Maxim (10 g), the root of *Aconitum pendulum* Buch (10 g), *Fructus of Choerospondias axillaris* (Roeh.) *Burr et Hill* (10 g), *Fructus of Terminalia chebula Retz.* (10 g), *Phyllanthi Fructus* (10 g) | *Decoction/oral administration/bid-tid/1 g each time* | Used for menorrhagia, metrorrhagia and metrostaxis, dyspepsia | NMPA*. Tibetan Medicin* |

(Continued on following page)
### TABLE 5 (Continued) Traditional Chinese formulations containing PF.

| Formulation names | Main composition | Formulations, usage, dosage | Diseases or symptoms therapy | Prescription sources |
|-------------------|------------------|-----------------------------|-----------------------------|----------------------|
| Si Wei Jang Huang Tang San. (四味姜黄散) | Curcuma longa (15 g), The dry bark of Berberis kansuensis Schneid. (12.5 g), Phyllanthus Fructus (25 g), Fructus of Cenchrus echinatus Linn. (25 g) | Decoction/oral administration/bid/3.2–4.8 g each time | Used for urethritis, frequent urination and urgency. Clearing away heat and diuresis | Chinese Pharmacopoeia (2020). NMPA* |
| Liu Wei Yu Gan Zi Tang San (六味余甘子汤散) | Phylanthi Fructus (200 g), Fructus of Coriandrum sativum L. (75 g), Fructus of Malva verticillata var. Crepe Linnaeus (75 g), the root of Pedicularis reuipina L. (40 g), Oxytropis kansuensis Bunge (100 g), the root and stem of Glycyrhiza uralensis Fisch./Glycyrrhiza inflata Bat./Glycyrrhiza glabra L. (50 g) | Decoction/oral administration/bid/3 g each time | Used for clearing heat and diuresis | Shang Han Lan Ji Yi. AD1801 |
| Ba Wei Xiao Bo Pi San (八味小檗皮散) | The roots and stems of Berberis silvaratoucana Schneid. (150 g), the fruit ear Piper longum L. (20 g), Phyllanthi Fructus (125 g), the root and stem of Glycyrhiza uralensis Fisch./Glycyrrhiza inflata Bat./Glycyrrhiza glabra L. (50 g), Cori aces sativus L. (75 g), Urti Feliis Pulpis (6 g), the secretion of Moschus berceovusii/M. fuscus Przewalski/ M. Moschiferus Linnaeus (6 g), Jing Mo (12 g) | Decoction/oral administration/bid/1 g each time | Used for urinary tract infection, odyuria, hemorrhoid, gonorrea and spermatorrhea | Chinese Pharmacopoeia (2020). Tibetan Medicon® |
| Qi Zhen Tang San (七珍散) | The root of Inula helenium L. (70 g), the stem of Rubus L. (225 g), the stem of Tinospora sinensis (Lour.) Merr. (125 g), the root and stem of Zonghe a quinquelina Rinc. (50 g), Fructus of Choeropousa auxiliaris (Rosb.) Burtt et Hill (90 g), Fructus of Terminalia chebula Retz. (50 g), Phyllanthi Fructus (100 g) | Decoction/oral administration/bid/3 g each time | Used for common cold due to wind-cold, fever and arthralgia | NMPC* Tibetan Medicon® |
| Qi Wei Kuan Jin Teng Tang San (七味宽筋藤汤散) | The vine stem of Tinospora sinensis (Lour.) Merr. (30 g), Fructus of Choeropousa auxiliaris (Rosb.) Burtt et Hill (25 g), Fructus of Terminalia chebula Retz. (80 g), Phyllanthi Fructus (25 g), Swertia bimaculata (Sieb. et Zucc.) Hook. f. et Thom. Ex C. B. Clark (25 g), Corydalis impatiens (pall.) Fisch. (25 g), Rhodiola crenulata (Hook. f. et Thom.) H. Ohba (25 g) | Decoction/oral administration/bid/3 g each time | Used for heat-clearing and detoxifying, variola | Tibetan Medicon® |
| Jie Ke Bai Bei Wan (解渴百贝丸) | Fructus of Chamaemoles speciosa (Sweet) Nakai (200g), Prunus mume (Sieb.) Sch.-et-Zucc. (500 g), the root and stem of Glycyrhriza uralensis Fisch./Glycyrrhiza inflata Bat./Glycyrrhiza glabra L. (375 g), the root of Pueraria lobata (Wild) Ohwi/ Pueraria thomunii Benth. (100 g), Ligusticum chuanxiong hort (25 g), Phyllanthi Fructus (25 g), the leaves of Perilla/r/Mescews (L.) Britt. (25 g), Mass Galla chinensis et camelliae Fermentata (50 g), Sal (500 g) | Pill/oral administration/1 pill each time | Used for clearing heat and detoxication | Pu Ji Fang. AD1406 |
| Qi Wei Xiao Zhong Wan (七味消肿丸) | Phyllanthi Fructus (300 g), the flower of Carthamus tinctorius L. (150 g), the seed of Herpetospermum polyanthodium (Ser.) C.B.Clarke (40 g), Dracaecophalum tanguticum Maxim (200 g), Veronica erige Me H.Winkl. (80 g), Aconitum napaulare (Brull.) Stapf (100 g), the flower of Maconopsis Vig. (150 g) | Pill/oral administration/ bid-tid/2g–2.5 g each time | Used for edema, thirst, oliburia, asthma and ascites | NMPC*. Tibetan Medicon® |
| Shi Wei Ke Zi Tang San (十味开子汤散) | Fructus of Choeropousa auxiliaris (Rosb.) Burtt et Hill (15 g), Fructus of Terminalia chebula Retz. (10 g), Phyllanthi Fructus (15 g), the stem of Tinospora sinensis (Lour.) Merr. (15 g), Swertia bimaculata (Sieb. et Thom.) H. Ohba (25 g), Glycyrrhiza uralensis Fisch./Glycyrrhiza inflata Bat./Glycyrrhiza glabra L. (50 g), Cori aces sativus L. (75 g), Urti Feliis Pulpis (6 g), the secretion of Moschus berceovusii/M. fuscus Przewalski/ M. Moschiferus Linnaeus (6 g), Jing Mo (12 g) | Decoction/oral administration/bid/1.6 g each time | Used for hepatitis B | Tibetan Medicon® |

(Continued on following page)
disadvantages of complex sample pretreatment process, expensive equipment, and not being environmentally-friendly. A recent study hypothesized that gallic acid, quercetin, ellagic acid, and corilagin could be used as candidate compounds for PF quality markers using predictive analysis with a multi-directional Q-marker (Guan et al., 2022). In the future, Q-marker-based predictive analysis can be studied in depth during the quality control of PF, and methods such as metabolomics and network pharmacology can be applied to it to establish safer, more scientific and effective quality assessment methods.

### 8 Products development of *Phyllanthi Fructus*

PF has been developed into various products in many fields due to its various pharmacological activities, including drugs, health products, foods, and cosmetics as shown in Figure 8.

#### 8.1 Drugs

PF has been developed into a variety of medicines due to its richness in bioactive substances. For example, PF capsules can reduce the content of TG and TC, increase the content of HDL and the activity of SOD in red blood cells and brain tissue, and reduce the content of lipid peroxides in serum and brain tissue (Li et al., 2002). PF ointment is mainly used for indigestion, abdominal distension, cough, sore throat, and toothache (Yu and Gongbu, 1987). In addition to singular use, PF is often used as a compound prescription. Based on the theory of TCM, it plays different roles when it is compatible with different Chinese herbal medicines. For example, Hepatitis B Kangtai pills are processed and formulated from Chinese medicines including PF, Morus mulberry, Turmeric, Kochia scoparia, Cao Guo and Coptis chinensis, which can effectively protect the liver and inhibit HBV (He, 2010). The common formulations of PF and traditional Chinese prescriptions that have been clinically tested and approved by the National Medical Products Administration (NMPA) were collected in Table 5.

PF is an herbal remedy with multiple healing properties for a wide range of diseases. After extensive basic research, researchers have conducted several clinical trials to scientifically validate the remarkable therapeutic effects of PF. In clinical trials for different diseases, it is often used alone or as the main ingredient in combination formulations. Several clinical trials of PF for different disease were listed in Table 6.

#### 8.2 Health products

A wide range of health products has been developed using PF as a raw material. First, health care wines and beverages,
including PF health wine, PF health ginger wine (Ming and Wang, 2018), Sanzi health drink, spiral health drink, and PF functional oral liquid. Wine prevents or reduces the development of atherosclerosis and coronary heart disease by increasing the level of HDL in the body. Brewing PF into health care wine could exert better therapeutic efficacy. Second, fruit powders. PF nutritional powders retain the SOD and over 70% of the vitamin C contained in the fresh fruit. Fruit powder is

| Diseases                     | Investigational drug and extracts | Formulations, dosage and administration | Positive/placebo control | Clinical trials                                                                 | Outcome indicators | References               |
|------------------------------|----------------------------------|------------------------------------------|--------------------------|----------------------------------------------------------------------------------|--------------------|--------------------------|
| Dyslipidemia                 | PF extracts capsule (35% polyphenols, 8% triterpenoids and 10% oil). Ethyl acetate. Amlamax™ (30% gallo ellagi tannins). Aqueous | Capsule/oral administration/bid/ 500mg each time for 6 months | Roasted rice powder (bid/ 500mg each time p.o. ) | A randomized, double-blind, placebo, controlled, multicenter clinical trial. Total Subjects: 98 (45 males and 55 females). PF group: 49 placebo groups: 49. Pilot Clinical study. Total Subjects: 51. Group I: 26 (Amlamax 500mg; Group II: 25 (Amlamax 1000mg) | TC↓, TG↓, VLDL-C↓, atherogenic index of the plasma (AIP), the ratio of Apo B to Apo A1, Group I and II total and LDL cholesterol, HDL cholesterol | Upadhyay et al. (2019), Antony et al. (2008) |
| Type 2 diabetic              | PF dried powder (Undefined Proportion). | Tablet/oral administration/bid/1, 2, or 3g each time for 21 days | Gilbenclamide (bid/5mg each time p.o.) | Comparative randomized clinical trial. Total Subjects: 32. Normal group: three subgroups (1,2,3g) and control group (carboxymethyl cellulose fiber). Diabetic group: three subgroups and control group (gilbenclamide orally) | 1.2,3g subgroups: fasting and 2-h postprandial blood glucose levels ↓, 2.3g subgroup: total cholesterol and TG↓, HDL-C↑, LDL-C↓ on day 21. 3g group: total lipids↓ | Akhtar et al. (2011) |
| Chronic periodontitis        | PF gel (10% sustained-release gel, 20% Poloxamer 407, 4% Carbopol 940 polymers and 10% PF extracts). Ethanol | Gel/ext. administration/ qd-tid/subgingival application for 3 months | Placebo gel. (qd-tid/subgingival application for 3 months) | A Randomized Placebo-controlled Clinical Trial. Total Subjects: 46 patients (528 sites). Control group (23, 264): SRP + placebo gel test group; Test group (23, 264): SRP + 10%PF gel application | Mean probing pocket depth↓, modified sulcus bleeding index↓, clinical attachment attachment | Grover et al. (2016) |
| Melasma                      | Three herb cream (PF, Licorice and Belides depigmenting complex 7%). Ethanol | Cream/ext. administration/bid/ 60 days | Hydroquinone cream 2% (qd/ext.) | Mono-blind clinical study. Total subjects: 56 women,18–60 years of age. Group A: 23, cream with Belides, Emblica and Licorice, applied twice a day; Group B: 23, cream with Hydroquinone 2%, used at night | Group A showed fewer skin adverse events | Costa et al. (2010) |
| Essential hypertension       | PF dry powder capsule (Undefined Proportion). Aqueous | Capsule/oral administration/bid/ 500mg each time for 12 weeks | Maize starch IP grade (bid/500mg each time p.o.) | A randomized double-blind placebo-controlled Trial. Total subjects:150; Test group:75; Placebo group : 75, maize starch IP grade | No significant change in systolic and/or diastolic blood pressure levels, lipid profile, HBA1C, hs-CRP levels | Shanmugarajan et al. (2021) |
| Symptomatic knee osteoarthritis | SGC® and SGG® formulations (Undefined Proportion). | Capsule/oral administration/qd/ 400mg each time for 24 weeks | Glucosamine sulphate (qd/2g each time p.o.) and celecoxib (qd/ 200mg each time p.o.) | A randomized, double-blind, controlled equivalence drug trial. Total subjects:440; Group 1:110; SGC®; Group 2: SGC®. Group 3:10, glucosamine sulphate. Group 4: 110, celecoxib | Knee pain↓, knee function↑ | Chopra et al. (2013) |

*Each SGC, capsule (400mg) contained Zingiber officinale, Tinospora cordifolia, PF, and B. serrata. The SGC, capsule (400mg) was similar to SGG® (both for content and quantity) except for the absence of B. serrata extracts and a higher quantity of excipients.¹ "¹ the experimental study has no positive/placebo control.
also suitable for long-term consumption by the elderly. It has the effects of clearing heat, lowering blood pressure, preventing liver and gallbladder diseases, and strengthening immunity. Third, lozenges, such as PF lozenge and PF lemon health-care lozenge, are rich in vitamins, amino acids, and a variety of trace elements. It has positive effects on the treatment of cough, obesity and cardiovascular diseases (Xu, 2007).

8.3 Foods

Ready-to-eat food prepared from fresh PF, including PF jam, preserved fruit, fructose, fruit cake, jelly, and yogurt. These products are more acceptable to the public because they retain the unique flavour and nutrients of PF while removing the sourness and astringency. A range of compound drinks and teas made from PF, including PF juice, lemon drink, Rosa roxburghii fruit drink, fruit tea, instant oolong tea, and herbal tea. These compound drinks taste good and achieve specific health benefits (Cao et al., 2019). PF when incorporated with ice cream in processed form resulted in increasing melting resistance and decreasing overrun, thereby enhancing the functional property and nutritional value of ice cream (Goraya and Bajwa, 2015). In addition, PF is made into condiments, health-care noodles and other products, retaining some of its medicinal properties.

8.4 Cosmetics

Herbs have long been used in medicines and cosmetics due to their potential to treat skin ailments and improve skin appearance. Antioxidants, flavonoids, and phenolic compounds present in PF play critical roles to counteract free radicals, the main cause of various unfavorable skin changes. The results of Fujii et al. revealed stimulated proliferation of fibroblast and controlled collagen metabolism confirmed the mitigative, cosmetic, and therapeutic application of PF (Fujii et al., 2008). Therefore, PF has been added to sunscreens, anti-aging and whitening skincare products and cosmetics. For example, a cosmetic developed from the raw materials of PF, lotus, tuckahoe, schisandra, and mulberry bark extract can inhibit the formation of melanin (Chen, 2019). These cosmetics meet the skincare needs of people for facial skin whitening, anti-aging, and wrinkle removal.

9 Discussion and perspective

The value of phytomedicine is well recognized in China and South East Asia followed by the continuously increasing demand worldwide. PF, rich in tannins, phenolic acids, flavonoids, polysaccharides, and vitamin, has been shown to have a variety of pharmacological effects on the respiratory, digestive, nervous, and cardiovascular systems, which are attributed to its anti-inflammatory, antioxidant, antiviral, and anti-apoptotic properties. Modern pharmacological studies have gradually confirmed some traditional effects of PF, such as moistening the lung for arresting cough, clearing heat and detoxifying, lowering blood pressure and glucose, etc. Moreover, some novel effects of PF were identified with modern pharmacology, including anti-anxiety, anti-depression, and neuroprotective effects. These pharmacological and toxicologic studies provide an important scientific basis for the development of PF-related products. In addition to being developed as a variety of drugs, PF has also been developed in the food, health care products, and cosmetics products industries.

Although it has been widely studied using modern technology, there are still challenges in the development of PF in a sustainable and maximally beneficial way. For instance, both cultivated and wild PF have the problem of varying quality. PF that does not meet the medicinal standard is not fully utilized, which easily results in a huge waste of resources. Therefore, it is of great significance to establish the medicinal and food grading utilization system of PF. At present, the research and application of Phyllanthus Emblica L. mainly focus on mature fruits, but the research on its roots, stems and leaves is less. Through deepening the research on the chemical composition and pharmacological activity of other medicinal parts of Phyllanthus Emblica L., the comprehensive utilization of Phyllanthus Emblica L. plants as a viable resource can eventually effectively promote the economic development of Phyllanthus Emblica L. production area. In addition, gallic acid is specified as a quality control index of PF in the Chinese Pharmacopoeia (version 2020). The single quality control index of PF is not representative of the overall quality of PF. A more scientific, simple, and easily replicable quality evaluation method needs to be sought in the future for PF industrialization.

Author contributions

XP and XY conceived the review. XY, QL, LJ, SW, WD, YC, and DC collected the literatures. XP, DC, and XY wrote and edited the manuscript.

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Conflict of interest
The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Chen, Z. Y., Liu, X. M., and Wu, J. J. (2003). Biological characteristics and nutritional components of Glycyrrhiza sinensis. Zhangzhou Nanfang Guoshu 032 (006), 71–73. doi:10.3969/j.issn.1007–1431.2003.06.045

Chinese Pharmacopoeia Commission (2020). Chinese Pharmacopoeia. Beijing: China Medical And Technology Press.

Chopra, A., Sahuj, M., Tilla, G., Sarmukkaddam, S., Venugopalan, A., Narsimulu, G., et al. (2013). Ayurvedic medicine offers a good alternative to glucosamine and celecoxib in the treatment of symptomatic knee osteoarthritis: A randomized, double-blind, placebo-controlled equivalence drug trial. Rheumatol. Oxf. 52 (8), 1408–1417. doi:10.1093/rheumatology/kez414

Chuang, H. Y., Ng, L. T., Lin, L. T., Chang, J. S., Chen, Y. J., Lin, T. C., et al. (2011). Hydrolyzable tannins of tropical almond show antibiotic effects in TGF-β1-induced hepatic stellate cells. J. Sci. Food Agric. 91 (13), 2777–2784. doi:10.1002/jsfa.4521

Costa, A., Mioses, T. A., Cordero, T., Alves, C., and Marmorei, J. (2010). Association of emblica, licorice and belides as an alternative to hydroquinone in the clinical treatment of melasma. An. Bras. Dermatol. 85 (5), 613–620. doi:10.1590/S0365–03262010000500003

Cui, B. Q., He, Z. Y., and Yang, Z. M. (2010). Effects of Phyllanthus emblica extracts on immune function of mice. Lishizhen Med. Mater. Medica Res. 21 (008), 1920–1922. doi:10.3969/j.issn.1008–0805.2010.08.033

Daisy, P., Babu, A. S., and Modial, R. (2009). Anti-hyperglycemic effects of Phyllanthus extracts in alloxan-induced diabetic rats. D’Souza, J. J., D’Souza, P. P., Faizal, F., Kumar, A., Bhat, H. P., Baliga, M. S., et al. (2014). Antimicrobial effects of the Indian indigenous fruit emblica officinalis gaertn: Active constituents and modes of action. Food Funct. 5 (4), 635–644. doi:10.1039/c3fo60366k

Dastur, J. F. (1952). Medicinal plants of India and Pakistan. Bombay: D. B. Taraporevala Sons & Co., Ltd.

De, A., De, A., Sharma, R., Sao, W., and Sharma, M. (2020). Sensitization of carboplatin-treated and -taxol-resistant high grade serous ovarian cancer cells carrying p53, BCA1/2 mutations by emblica officinalis (amla) via multiple targets. J. Cancer 11 (7), 1927–1939. doi:10.7150/jca.36019

De, A., Papasian, C., Hentges, S., Banerjee, I., Hentges, S. K., et al. (2013). Emblica officinalis extract induces autophagy and inhibits human ovarian cancer cell proliferation, angiogenesis, growth of mouse xenograft tumors. PLoS One 8 (8), e27428. doi:10.1371/journal.pone.002748

Dhingra, D., Joshi, P., Gupta, A., and Chhillar, R. (2012). Possible involvement of monosynaptic neurotransmission in antidepressant-like activity of Emblica officinalis fruits in mice. CNS Neurosci. Ther. 18 (5), 419–425. doi:10.1111/j.1745–1594.2011.08256.x

Ding, Y., Sun, X., Chen, Y., Deng, Y., and Qian, K. (2015). Epigallocatechin gallate attenuated non-alcoholic steatohepatitis induced by methionine- and choline-deficient diet. Eur. J. Pharmacol. 761, 405–412. doi:10.1016/j.ejphar.2015.05.005

Dong, K., Li, C. W., and Qiu, H. (2009). Effect of phyllanthus emblica on the expression of protein kinase B and glucose transporter4 mRNA in the skeletal muscle of insulin resistance rat. J. Fourth Mil. Med. Univ. 30 (6), 2352–2355.

Dong, Y. H., Yin, S. T., Jiang, C., Luo, X. H., Gao, X., Zhao, C., et al. (2014). Involvement of autophagy induction in penta-1, 2, 3, 4, 6-O-galloyl-beta-D-galactose induced-Senessence-like growth arrest in human cancer cells. Autophagy 10 (2), 296–310. doi:10.4161/auto.27210

Du, Q., Xu, X. H., Lin, P. C., Yong, C. L., and Ju, Y. E. (2017). Research progress and developmental prospect of plant polysaccharide. Electron. J. Transl. Med. doi:10.3969/j.issn.1673–8989.2017.04.020

Du, R., Cooper, L., Chen, Z., Lee, H., Rong, L., and Cui, Q. (2021). Discovery of chebulagic acid and punicalagin as novel allosteric inhibitors of HIV-1 fusion process of Phyllanthus emblica. J. Dispensatory of the United States of America. 2nd edition.

Ehsel, B., and Schmid, J. A. (2013). The complexity of NF-kB signaling in inflammation and cancer. Mol. Cancer 12 (1), 86–15. doi:10.1186/1476–4598-12-86

Hou, H. Y. (2006). Studies on anti-HBV principles of Phyllanthus emblicaL. Chinese People’s Liberation Army Academy of Military Medical Sciences.

Hou, K. W. (2002). The compositions and application of phylanthus emblica in the traditional medicine. Chin. J. Ethn. Folk Med. 11 (6), 345–348.

Huang, C. Z., Tung, Y. T., Hsu, S. M., Wu, C. H., and Yen, G. C. (2017). The hepatoprotective effect of Phyllanthus emblica L. fruit on high fat diet-induced non-alcoholic fatty liver disease (NAFLD) in SD rats. Food Funct. 8 (2), 842–850. doi:10.1039/c6fo01585a

Huang, H. Z., Wei, L. X., Lin, J. Z., Tan, P., Fan, S. H., Han, L., et al. (2019). Discussion on the rationality of tannin transformation and Pharmacopoeia determination method in the Re duction process of Phyllanthus emblica. Chin. Pharm. J. 54 (7), 581–587.

Jaisoy, K., Sonthornchareonnon, N., Nertprasertsuke, N., Panthong, A., and Sureeratwong, S. (2010). Acute and chronic oral toxicity of standardized water extracts from the fruit of Phyllanthus emblica Linn. Int. J. Appl. Res. Nat. Prod. 3 (1), 48–58.
Yan et al. 10.3389/fphar.2022.1017268

Jain, I., Jain, P., Bisht, D., Sharma, A., Srivastava, B., Gupta, N., et al. (2015). Use of traditional Indian plants in the inhibition of caries-causing bacteria—Streptococcus mutans. Braz. Dent. J. 26 (2), 110–115. doi:10.1016/j.bjodent.2013.10.012

Li, X. H. (2010). The screening and mechanism study of Phyllanthus urinaria L. which have anti-angiogenic effect. Nanjing University of Traditional Chinese Medicine.

Jose, K. J., Kuttan, R., and Joc, B. (1995). Antioxidant activity of Emblica officinalis. J. Clin. Biochem. Nutr. 19 (2), 63–70. doi:10.3164/jcbn.19.63

Kang, W. J., Zhang, G. M., and Zhao, X. H. (2011). Hypoglycemic effects of different solvent extracts from phyllanthus fructus. Arch. Hau. Med. Sci. 39 (30), 18545–18547. doi:10.3969/j.issn.0517-6611.2011.03.002

Kapoor, L. (1989). CRC handbook of ayurvedic medicinal plants. CRC Press.

Kong, X. Y., Yu, R., and Liu, J. X. (2016). The influence of flavone extracts of Phyllanthus emblica on influenza H1N1 virus infected pneumonia mice. Her. Traditional Chin. Med. 255 (105), 64–65.

Krupanidhi, S., Peele, K. A., Venkateswaralu, T. C., Ayyagiri, V. S., Aishwarya, G., John Babu, D., et al. (2020). Screening of phytochemical compounds of tinospora cordifolia for their inhibitory activity on sars-cov-2: An in silico study. J. Biomol. Struct. Dyn. 39 (15), 5799–5803. doi:10.1080/07391102.2019.1715772

Kumar, A., Tantry, B. A., Rahman, S., and Gupta, U. (2011a). Comparative study of antimicrobial activity and phytochemical analysis of methanolic and aqueous extracts of the fruit of Emblica officinalis against pathogenic bacteria. J. Tradit. Chin. Med. 31 (03), 246–250. doi:10.1016/j.jtcm.2010.07.002

Kumar, C. P., Sowthi, B. S., Kumar, D. Y., and Neelima, K. (2014). Effect of emblica officinalis on stress induced biochemical and psychological changes in mice.

Kumar, D., Gunra, A., Abbirami, E., Preethi, J., Sivasadula, T., and Thilagar, S. (2020). Cissus quadrangularis (veldt grape) attenuates disease progression and inflammatory properties. J. Ethnopharmacol. 254, 112740. doi:10.1016/j.jep.2020.112740

Li, W., Zhang, X., Chen, R., Li, Y., Miao, J., Liu, G., et al. (2020). HPLC fingerprint analysis of Phyllanthus emblica ethanol extract and their antioxidant and anti-inflammatory properties. J. Ethnopharmacol. 254, 112740. doi:10.1016/j.jep.2020.112740

Li, Z. W., Ma, S. B., and Zeng, Y. Q. (2010). Extraction of Phyllanthus emblica flavone and its antioxidant activity. Jinguo Sci. Agric. 300 (082), 287–289. doi:10.3969/j.issn.1001-0408.2010.11.074

Li, C. L. (2001). Pharmacological studies on Phyllanthus emblica L. Fruit Plants Hun. Nutr. 64 (1), 1–5. doi:10.1111/j.1399-6001.2001.00001-0

Lim, D. W., Kim, J. G., and Kim, Y. T. (2016). Analgesic effect of Indian gooseberry (emblica officinalis fruit) extracts on postoperative and neuropathic pain in rats. Nutrients 8 (12), 760. doi:10.3390/nu8120760

Luo, D. S. (2004). Newly repaired jingzhu materia medica. Beijing: Science and Technology Press.

Luo, C. L. (2010). Immunoregulative action and inhibitive effect of Phyllanthus emblica on tumour cells. Chin. J. Exp. Traditional Med. Formulae 16 (13), 155–158. doi:10.3969/j.issn.1005-9903.2010.13.047

Luo, D. S. (2004). Newly repaired jingzhu materia medica. Sichuan Science and Technology Press.

Lu, W., Wen, L. R., Zhao, M. M., Yang, B., Ren, J., Shen, G., et al. (2012). Structural identification of nosomaltosin and other phenolics in Phyllanthus emblica L. fruit hull. Food Chem. 132 (3), 1527–1533. doi:10.1016/j.foodchem.2011.11.146
emblica against paracetamol induced hepatic damage in wister albino rats on spatial learning and memory in rats.

Maged, M., Bhat, B., Idrishan, A. N., Srivastava, J. S., and Nagabhushanam, K. (2009). Ascorbic acid and tannins from Emblica officinalis Gaertn. Fruits.—a revisit. J. Agric. Food Chem. 57 (1), 220–225. doi:10.1021/jf809200b

Maled, H., and Mettula, S. M. (2009). Hepato-protective activity of Phyllanthus emblica against paracetamol induced hepatic damage in wister albino rats.

Malve, H. O., Raut, S. B., Parmate, P. A., and Rege, N. N. (2014). Effect of combination of Phyllanthus emblica, Tinospora cordifolia, and Ocimum sanctum on spatial learning and memory in rats. J. Ayurveda Integr. Med. 5 (4), 209–215. doi:10.4103/0975-9476.146656

Mao, S. N. (2019). Study on the establishment conditions and quality characteristics of Euphorbia. Chengdu University of Traditional Chinese Medicine. doi:10.26988/cnki.gcdz.2019.00113

Mathew, M., and Subramanian, S. (2014). In vitro screening for anti-cholinesterase and antioxidant activity of methanolic extracts of ayurvedic medicinal plants used for cognitive disorders. PLoS One 9 (1), e88064. doi:10.1371/journal.pone.0088064

McCord, J. M. (1985). Oxygen-derived free radicals in postischemic tissue injury. N. Engl. J. Med. 312 (3), 159–163. doi:10.1056/NEJM198501173120305

Mehrood, M. H., Rehman, A., Rehman, N. U., and Gilani, A. H. (2013). Studies on prokinetic, laxative and spasmodic activities of Phyllanthus emblica in experimental animals. Phytotest. Res. 27 (7), 1054–1066. doi:10.1002/phtr.4823

Meng, Z. Y., Zhu, Y. L., and Wang, G. W. (2011). Extraction process and molecular docking and simulation study. Phytother. Res. 25 (3), 327–334. doi:10.1002/ptr.3127

Minging, Y. Q., and Wang, S. M. (2018). The brewing method of Phyllanthus emblica healthy ginger wine.

Murugesan, S., Kottedak, S., Crasta, I., Sreevathasan, S., Usarhani, D., Perumal, M. K., et al. (2021). Targeting COVID-19 (SARS-CoV-2) main protease through active phytoconstituents of ayurvedic medicinal plants – emblica officinalis (Amla), Phyllanthus niruri Linn (Bhumli Amla) and Tinospora cordifolia (Giloy) – a molecular docking and simulation study. Comput. Biol. Med. 136, 104683. doi:10.1016/j.compbiomed.2021.104683

Muthuraman, A., Sood, S., and Singla, S. K. (2011). The anti-inflammatorily potential of phenolic compounds from Emblica officinalis L. in rat. Inflammopharmacology 19 (6), 327–334. doi:10.1007/s10787-010-0041-9

Na, Y. X., and Yang, F. R. (2002). Overview of research on development and utilization of medicinal plant resources in foreign countries. For. Investigation Des. Sci. 312 (3), 159–163. doi:10.1391/11346.312(3).3127

Natesh, I., Mondal, P., Penta, D., Ajeer, A., and Meenan, S. M. (2021). Culinary spice bioactives as potential therapeutics against sars-cov-2. Computational investigation. Comput. Biol. Med. 128, 101402. doi:10.1016/j.compbiomed.2020.101402

Ngamkitdechatkul, C., Jaijoy, K., Hansakul, P., Soonthorncharoenon, N., and Sireeratawong, S. (2010). Antitumour effects of Phyllanthus emblica L. Induction of cancer cell apoptosis and inhibition of in vivo tumour promotion and in vitro invasion of human cancer cells. Phytotest. Res. 24 (9), 1405–1413. doi:10.1002/phtr.3127

Nicolas, E., Lampronni, I., Drehcicci, M. C., Borgatti, M., Tannamini, A., Bianchi, N., et al. (2008). Pyrogallol, an active compound from the medicinal plant Emblica officinalis, regulates expression of pro-inflammatory genes in bronchial epithelial cells. Int. Immunopharmacol. 8 (12), 1672–1680. doi:10.1016/j.intimp.2008.08.001

Nisar, M. F., He, J. W., Ahmed, A., Yang, Y. X., Li, M. X., and Wang, C. P. (2018). Chemical components and biological activities of the genus Phyllanthus: A review of the recent literature. Molecules 23 (10), 2567. doi:10.3390/molecules23102567

Oleniuk, D. N., Kochchenko, N. I., Schwabl, H., Vennos, C., and Leepfe, C. (2015). New mucic acid gallates from Phyllanthus emblica. Chem. Nat. Compd. 51 (4), 666–670. doi:10.1007/s10501-015-1380-y

Packiramasam, R. M., Bobby, Z., Panneerselvam, S., Koshy, S. M., and Jacob, S. E. (2018). Metabolomic analysis and antioxidant effect of amla (emblica officinalis) extract in preventing oxidative stress-induced red cell damage and plasma protein alterations. An in vitro study. J. Med. Food 21 (1), 81–89. doi:10.1089/jmf.2017.3942
Traditional medicine with antioxidant activity. J. Ethnopharmacol. 71 (1-2), 23–43. doi:10.1016/s0378-8741(00)00213-0

Shanmugarajan, D., Girish, C., Harivenkatesh, N., Chavanerrappa, B., and Lakshma, N. (2021). Antihypertensive and pleotropic effects of Phyllanthus emblica extract as an add-on therapy in patients with essential hypertension-A randomized double-blind placebo-controlled trial. Phytother. Res. 35 (6), 3275–3285. doi:10.1007/s11101-020-01178-7

Shao, Y. Y., Zhu, D., Yang, G. H., Mao, X., and Xia, Q. (2014). Health effects of tannin-containing herbs society of traditional Chinese medicine. Shi, Y. P., and Yang, Z. M. (1994). The study of nutritive components of flash of Phyllanthus emblica fruit in GuangZhou. J. Guizhou Normal Univ. (2), 1–6.

Shmyr, M., P. Gupta, K., and Saini, M. (2011). Antineoplastic potential of medicinal plants. Recent Pat. Biotechnol. 5 (2), 85–94. doi:10.2174/18722081179365662

Siddiqua, A. J., Danciu, C., Ashraf, S. A., Moin, A., Adnan, M., Alreshid, M., et al. (2020). Plants-derived biomolecules as potent antiradical phytomedicines: New insights on ethnobotanical evidences against coronaviruses. J. Ethnopharmacol. 230.

Thenmozhi, A. J., Dhivyabharathi, M., Raja, T. R. W., Manivasagam, T., and Essa, M. M. (2016). Tannoid principles of Emblica of Phyllanthus emblica extract prevents contrast-induced acute kidney injury. Mol. Vis. 22 (10), 148–155. doi:10.3390/mi1229193

Tan, G. S., Jiang, R. F., Li, Y. P., Li, F. A., and Zhang, D. J. (2015). Protective effect of Wu Wei Yu Gan Zi San on myocardial ischemia-reperfusion injury in rats. J. Ethnopharmacol. 168 (1), 148–155. doi:10.1016/j.jep.2015.01.009

Uddin, M. S., Mamun, A. A., Hossain, M. S., Akter, F., Ishbal, M. A., and Asaduzzaman, M. (2016). Exploring the effect of Phyllanthus emblica L. On cognitive performance, antioxidant markers and acetylcholinesterase activity in rats: Promising natural gift for the mitigation of alzheimer’s disease. Ann. Neurosci. 23 (4), 218–229. doi:10.1055/s-004494482

Udupa, K. N. (1985). Ayurveda for promotion of health. J. Ayurveda 3.

Upadhy, H., Prabhu, S., Prasad, A., Subramanian, D., Gupta, S., and Goel, A. (2015). A randomized, double-blind, placebo-controlled, multicenter clinical trial to assess the efficiency and safety of Emblica officinalis extract in patients with dyslipidemia. BMC Complement. Altern. Med. 19 (1), 27. doi:10.1186/s12906-019-2430-y

Vallacotta, L., Arzi, A., and Zingg, J. M. (2007). Regulatory role of vitamins E and C on extracellular matrix components of the vascular system. Mol. Asp. Med. 28 (5–6), 507–537. doi:10.1016/j.mam.2007.05.002

Vormisto, A. I., Summanen, J., Kankaanranta, H., Vuorela, H., Asmawi, Z. M., and Moilanen, E. (1997). Anti-inflammatory activity of extracts liom leaves of Phyllanthus emblica. Planta Med. 63 (6), 518–524. doi:10.1055/s-2006-977754

Wang, C. C., Yuan, J. R., Wang, C. F., Yang, N., Chen, J., Liu, D., et al. (2016). Anti-inflammatory effects of Phyllanthus emblica L. on benzoylperoxide-induced precancorous lung lesion by regulating the IL-12/IFN-γ signaling pathway. Cancer Med. Ther. 16 (4), 505–515. doi:10.1159/000453676

Wang, H. M., Fu, L., Cheng, C. C., Gao, R., Lin, M. Y., Su, H. L., et al. (2019). Inhibition of LPS-induced oxidative damages and potential anti-inflammatory effects of Phyllanthus emblica extract via down-regulating NF-κB, COX-2, and iNOS in RAW 264.7 cells. Antioxidants (Basel) 8 (8), 270. doi:10.3390/antiox8080270

Wang, H. Z., Shan, M., and Sun, J. Y. (2007). Progress of pharmacological research on alpha-linolenic acid.

Wang, J., Liu, Y. T., Xiao, L., Zhu, L., Wang, Q., and Yan, T. (2014). Anti-inflammatory effects of apigenin in lipopolysaccharide-induced inflammatory in acute lung injury by suppressing COX-2 and NF-κB pathway. Inflammation 37 (6), 2085–2090. doi:10.1007/s10753-014-9715-y

Wang, L. P., Cui, Y., and Xu, J. (2018). Anti-aging effect of Phyllanthus emblica. On Cuernorhabditis elegans. J. Ilirn Univ. 56 (3), 276–280. doi:10.1341/jckdxblxb.2018.03.45

Wang, M., Sun, P. Y., Liang, W., Dai, Y. Y., and Cai, S. (2020). Critical success factors for the entry of compound Chinese herbal medicines into the European market. Chin. J. Pharmacol. Toxicol. 34 (2), 81–94. doi:10.3867/j.issn.1000-3002.2020.02.001

Wang, R. (2017). Inhibition effect on β-glucosidase and antioxidant activity for polyphenol extracts from Phyllanthus emblica L. Food Res. Dev. 38 (11), 13–16. doi:10.3969/j.issn.1000-3002.2017.11.004

Wang, S. H., Cheng, J. T., Guo, C., Cui, W. J., Shi, J., and Liu, A. (2019). Chemical constituents of Phyllanthus emblica and its anti-inflammation activities. Wang, S. P., Ma, Y., and Wang, S. H. (2009). Analysis of chemical composition of volatile oil of Phyllanthus emblica L from Sichuan by GC-MS. West China Pharmaceutical Journal, 279–281.

Wang, W. (2022). Book review of Chinese medicinal plants. Chin. J. Traditional Chin. Med. 47 (9), 2556.

Wang, Z. X., Xu, G. L., and Wang, Y. F. (2013). The possible role of oxidative stress and antioxidants in the occurrence and development of diabetic complications. J. Med. Res. 42 (004), 12–15.

Wei, L., Zhao, M., Bao, Y., Ren, J., Shen, G., and Rao, G. J. (2011). Antioxidant and antiproliferative capacities of phenolics purified from Phyllanthus emblica L. fruit. Food Chem. x. 126 (11), 277–282. doi:10.1016/j.foodchem.2010.11.018

Wei, Z. X., Qiu, H. X., and Zhou, L. H. (2002). Studies on pollen morphology of Phyllanthus Emblica. J. Ayurveda Prev. Med. Forum 19 (6), 269–278. doi:10.1159/000449482

Wu, L. F., Ye, T., and Liang, L. J. (2017). Research on stability of tannin part in Phyllanthus emblica L. In artificial gastric and intestinal juice. World Sci. Technology-Modernization Traditional Chin. Med. (08), 136–140. doi:10.11404/wst.2017.08.023

Wu, L., Zhang, Q., Liang, W., Ma, Y., Niu, L., and Zhang, L. (2021). Phytochemical analysis using upl-cms combined with network pharmacology approaches to explore the biomarkers for the quality control of the anticancer

Scartezzini, P., and Speroni, E. (2000). Review on some plants of Indian traditional medicine with antioxidant activity. J. Ethnopharmacol. 71 (1-2), 23–43. doi:10.1016/s0378-8741(00)00213-0

Shanmugarajan, D., Girish, C., Harivenkatesh, N., Chavanerrappa, B., and Lakshma, N. (2021). Antihypertensive and pleotropic effects of Phyllanthus emblica extract as an add-on therapy in patients with essential hypertension-A randomized double-blind placebo-controlled trial. Phytother. Res. 35 (6), 3275–3285. doi:10.1007/s11101-020-01178-7

Shao, Y. Y., Zhu, D., Yang, G. H., Mao, X., and Xia, Q. (2014). Health effects of tannin-containing herbs society of traditional Chinese medicine. Shi, Y. P., and Yang, Z. M. (1994). The study of nutritive components of flash of Phyllanthus emblica fruit in GuangZhou. J. Guizhou Normal Univ. (2), 1–6.

Shmyr, M., P. Gupta, K., and Saini, M. (2011). Antineoplastic potential of medicinal plants. Recent Pat. Biotechnol. 5 (2), 85–94. doi:10.2174/18722081179365662

Siddiqua, A. J., Danciu, C., Ashraf, S. A., Moin, A., Adnan, M., Alreshid, M., et al. (2020). Plants-derived biomolecules as potent antiradical phytomedicines: New insights on ethnobotanical evidences against coronaviruses. J. Ethnopharmacol. 230.
The four medical classics. Beijing People’s Medical Publishing House.

Yuan, J. M., Yang, X. Q., Xu, Z. P., Kong, W. X., Zhao, Q. L., and Lei, B. (2021). Evaluation of amino acid composition and nutritional value of Ya Ganzhi fruit in the dry and hot valley region of Yunnan. *Jiangsu J. Agric. 33* (10), 9.

Zeng, X. X., and Cen, Z. F. (2012). Anti-inflammatory and analgesic effects of Phyllanthus emblica extracts. *Guangdong Med. 33* (023), 3533–3536. doi:10.3969/j.issn.1001-9448.2012.23.008

Zhang, G., Zhou, S. M., and Tian, H. J. (2011). An experimental study of Phyllanthus Emblica. On the anti-fatigue effect on mice in simulated plateau environment. *Pharm. J. Chin. People’s Liberation Army* (03), 208–211. doi:10.3969/j.issn.1008-9926.2011.03.007

Zhang, J., Miao, D., Zhu, W. F., Xu, J., Liu, W. Y., Kidambrongtham, W., et al. (2017). Biological activities of phenolics from the fruits of Phyllanthus emblica L. *Phytochemistry* 141 (12), e1790040. doi:10.1002/phcs.201790040

Zhang, L. Z., Zhao, W. H., Guo, Y. J., Tu, G. x., Lin, S., and Xin, L. G. (2003). [Studies on chemical constituents in fruits of Tibetan medicine Phyllanthus emblica]. *J. Chin. Materia Medica 28* (10), 940–943.

Zhang, W. L., Zhang, J. Y., Zhang, K. L., Huang, Y. C., Lin, W. B., Liang, Z. W., et al. (2014). Determination of gallic acid and ellagic acid in extracts of Phyllanthus emblica by HPLC. *China Pharm. 28* (2), 26–29.

Zhang, W. W., Zhang, H., Zheng, H. L., and Feng, Y. (2013). Microwave-assisted extraction and fatty acid composition analysis of oil from Phyllanthus emblica L. seeds. *Food Sci. 34* (6), 20. doi:10.7506/spkx/0002-6650-201320003

Zhang, X. M., Liu, X. F., and Gao, Y. T. (2011). On the Antioxidation Effects of the extracts of Phyllanthus emblica L. *Res. Trace Elem. Health 28* (003), 29–30.

Zhang, Y. J., Tanaka, Y., Yang, C. R., and Kouno, I. (2001). New phenolic constituents from the fruit juice of Phyllanthus emblica. *Chem. Pharm. Bull. 49* (5), 537–540. doi:10.1248/cpb.49.537

Zhang, Y. J., Abe, T., Tanaka, Y., Yang, C. R., and Kouno, I. (2002). Two new acylated flavanone glycosides from the leaves and branches of Phyllanthus emblica. *Chem. Pharm. Bull. 50* (6), 841–843. doi:10.1248/cpb.50.841

Zhang, Y., Zhao, L. J., Guo, X. J., Li, C., Li, H. Z., Lou, H. X., et al. (2016). Chemical constituents from Phyllanthus emblica L and the cytoprotective effects on H2O2-induced PC12 cell injuries. *Arch. Pharm. Res. 39* (9), 1202–1211. doi:10.1007/s12272-014-0433-2

Zhao, B., and Hu, M. (2013). Gallic acid reduces cell viability, proliferation, invasion and angiogenesis in human cervical cancer cells. * Oncol. Lett.* 6 (6), 1749–1755. doi:10.3892/ol.2013.1652

Zhao, K. H., Wang, J., and Lv, X. M. (2018). Pharmacological study of Tibetan medicine emblica in the prevention and treatment of high altitude polycythemia. *J. Traditional Chin. Med. 033* (003), 934–939.

Zhao, M. L., Liu, X. L., Cui, C., and Luo, W. (2007). Composition and antimicrobial activity of essential oil from Phyllanthus emblica L. By supercritical CO2 extraction. *J. South China Univ. Technol. 035* (012), 116–120. doi:10.3321/j.issn:1000-565x.2007.12.023

Zhou, K., Jian, P., Liang, W. Y., Liang, L. J., Cui, Y. P., Ye, T., et al. (2018). Research progress on the chemical constituents and pharmacological effects of the Tibetan medicine Fructus mongolicum. *World Sci. Technol. - Med. TCM 20* (9), 1608–1614

Zhu, X. X., Wang, J. J., Ou, Y., Han, W. W., and Li, F. F. (2013). Polyphenol extract of Phyllanthus emblica (PEEP) induces inhibition of cell proliferation and triggers apoptosis in cervical cancer cells. *Eur. J. Med. Res. 18* (1), 46. doi:10.1186/2047-783X-18-46

Zhu, Z. M., Jin, J. W., and Zhao, D. P. (2018). *A Chinese medicinal composition capable of boosting immunity, lowering blood lipids, and protecting the heart and brain and its preparation method.*
Glossary

**ABTS**+ 2,2′-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid)

**AchE** acetylcholinesterase; AgNPs, synthesize silver nanoparticles

**AIDS** anti-acquire immunodeficiency syndrome

**ALT** alanine aminotransferase

**AMPK** adenosine 5′-monophosphate-activated protein kinase

**AR** Amalaki Rasayana

**AST** aspartate aminotransferase

**CAT** catalase

**COX-2** cyclooxygenase-2

**DPPH** 2, 2 diphenyl-1-picrylhydrazyl

**GC/MS** gas chromatogram/mass spectrometry

**GRO-α** growth-regulated oncogene alpha

**GSH-Px** glutathione peroxidase

**HDL** high-density lipoprotein

**HFD** high-fat diet

**HGSOC** high-grade serous ovarian cancer

**HIV** human immunodeficiency virus

**HPLC** high performance liquid chromatography

**HPV** human papillomavirus

**HSV-1** herpes simplex virus type 1

**IL-6** interleukin-6

**IL-8** interleukin-8

**INF-α** tumour necrosis factor-α

**IRI** ischaemia-reperfusion injury

**LDH** lactate dehydrogenase

**LDL** low-density lipoprotein

**LPS** Lipopolysaccharide

**MDA** malondialdehyde

**MgO** magnesium oxide

**MIP-2** macrophage inflammatory protein 2

**NF-κB** nuclear factor kappa-B

**NK** natural killer

**NSAIDs** non-steroidal anti-inflammatory drugs

**OXPHOS** oxidative phosphorylation

**PAO1** pseudomonas aeruginosa 1

**PF** Phyllanthi Fructus

**PGE2** prostaglandin E2

**ROS** reactive oxygen species

**SARS-CoV-2** severe acute respiratory syndrome coronavirus 2

**SNI** a neuropathic pain nerve injury

**SOD** super oxide dismutase

**STZ** streptozotocin

**TC** total cholesterol

**TCM** traditional Chinese medicine

**TG** triglyceride

**UPLC-MSn** Ultra-high performance liquid chromatography-multistage mass spectrometry; triglyceride

**VLDLs** very-low-density lipoproteins