Maturation of active zone assembly by Drosophila Bruchpilot

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Synaptic vesicles fuse at active zone (AZ) membranes where Ca²⁺ channels are clustered and that are typically decorated by electron-dense projections. Recently, mutants of the Drosophila melanogaster ERC/CAST family protein Bruchpilot (BRP) were shown to lack dense projections (T-bars) and to suffer from Ca²⁺ channel–clustering defects. In this study, we used high resolution light microscopy, electron microscopy, and intravital imaging to analyze the function of BRP in AZ assembly. Consistent with truncated BRP variants forming shortened T-bars, we identify BRP as a direct T-bar component at the AZ center with its N terminus closer to the AZ membrane than its C terminus. In contrast, Drosophila Liprin-α, another AZ-organizing protein, precedes BRP during the assembly of newly forming AZs by several hours and surrounds the AZ center in few discrete punctae. BRP seems responsible for effectively clustering Ca²⁺ channels beneath the T-bar density late in a protracted AZ formation process, potentially through a direct molecular interaction with intracellular Ca²⁺ channel domains.

Introduction

The arrival of action potentials mediates Ca²⁺ influx through strategically localized clusters of voltage-operated Ca²⁺ channels at the synaptic active zone (AZ) membrane. Ca²⁺ triggers exocytosis of synaptic vesicles, and tight coupling between release-ready vesicles and Ca²⁺ channels seems important for efficient neurotransmitter release (Neher and Sakaba, 2008). AZs are further characterized by macromolecular cytomatrices named dense bodies (Zhai and Bellen, 2004; Sikou et al., 2007). The role of these electron-dense specializations and of AZ-enriched proteins in the assembly of the AZ and/or the synaptic vesicle exo-endocytosis cycle is under intense investigation (Owald and Sigrist, 2009). Protein architectures constituting and controlling dense bodies remain to be revealed, and their contributions to AZ assembly in general and Ca²⁺ channel clustering in particular must be defined genetically.

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Abbreviations used in this paper: au, arbitrary units; AZ, active zone; BRP, Bruchpilot; Cac, Cacophony; DGluR, Drosophila glutamate receptor subunit; DLiprinα, Drosophila Liprin-α; ESM, ethyl methyl sulfonate; FS, freeze substitution; HPF, high pressure freezing; IP, immunoprecipitation; mStraw, mStrawberry; NMJ, neuromuscular junction; PSD, postsynaptic density; PSF, point spread function; STED, stimulated emission depletion; UAS, upstream activator sequence.

The most straightforward approach would be to use immunolabeling combined with light microscopy. However, individual AZs measure only a few hundred nanometers in diameter, rendering this approach difficult. Recently, stimulated emission depletion (STED) microscopy (Hell, 2007) has proven valuable for high resolution light microscopic studies of synapse architectures (Kittel et al., 2006; Jin and Garner, 2008; Westphal et al., 2008).

The Drosophila melanogaster neuromuscular junction (NMJ) is a leading model for genetic analyses of synapse structure and assembly (Featherstone et al., 2000; Koh et al., 2000; Collins and DiAntonio, 2007). In this preparation, dense body structures termed T-bars may take part in activity-dependent changes of synaptic performance (Prokop and Meinerzhagen, 2006). Furthermore, the Ca²⁺ channel α1 subunit Cacophony (Cac) was shown to dominate neurotransmitter release at NMJ synapses (Kawasaki et al., 2000).

Proteins of the conserved CAST (CAZ-associated structural protein)/ERC (ELKS–Rab6-interacting protein CAST)

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densities were compromised within AZs (Kittel et al., 2006). However, whether BRP performs an essential signaling role in T-bar formation or whether the protein itself is an essential building block of T-bars remained to be clarified. Thus, we entered into a structure-function analysis of BRP.

mAb Nc82 is derived from a Drosophila head extract–directed library (Hofbauer et al., 2009) and allowed the first identification of the BRP protein. mAb Nc82 is a widely used marker in Drosophila, both for neuropil in general and for AZs in particular (Wucherpfennig et al., 2003; Kittel et al., 2006; Wagh et al., 2006). Previously, we had loosely mapped the epitope of Nc82 to the region between aa 635 and the end of the 1,740-aa BRP protein (based on cDNA AT09405; Wagh et al., 2006). To define the Nc82 epitope more precisely, various BRP fragments (Fig. 1 A and Table I) were ectopically expressed in wing discs using dpp-Gal4 (Fig. 1 B). In this manner, the mAb Nc82 epitope (hereafter BRPNc82) could be mapped to the region between aa 1,227 and 1,740. Additionally, an antibody directed against an N-terminal peptide (BRPN-Term antibody; aa 62–75; Fig. 1 B) was produced.

Table I. BRP-reexpressing constructs

| BRP fragments | Start (aa) | End (aa) |
|---------------|------------|---------|
| Domain 1 [D1] | 1          | 320     |
| Domain 2 [D2] | 268        | 617     |
| Domain 3 [D3] | 473        | 1,226   |
| Domain 4 [D4] | 1,152      | 1,740   |

The predicted lengths in aa of the UAS-BRP fragments (relative to full-length BRP [1,740 aa]) are shown.

family are generic AZ proteins. In mice, CAST/ERC proteins have been shown to localize to AZs of various synapses and to bind other AZ proteins such as RIM (Rab3a-interacting molecule) and Liprin-α (Ohtsuka et al., 2002; Wang et al., 2002; Ko et al., 2003; Deguchi-Tawarada et al., 2004). In Caenorhabditis elegans, the CAST/ERC family member ELKS (glutamine-, leucine-, lysine-, and serine-rich protein) appears to operate genetically downstream of Syd-2/Liprin-α during the assembly of AZs at vulval synapses (Dai et al., 2006; Patel et al., 2006). Recently, the CAST/ERC family member Bruchpilot (BRP), a coiled-coil rich protein of nearly 200 kD, was identified via its localization to Drosophila AZs. Mutants of brp lacked T-bars, and Ca2+ channels were mislocalized at AZs, leading to inefficient vesicle release and changes in synaptic short-term plasticity (Kittel et al., 2006; Wagh et al., 2006).

In this study, we provide evidence that BRP takes up an elongated conformation and is a direct component of the T-bar. The N terminus of BRP is found superimposed on the Ca2+ channel clusters at the AZ center. BRP and Cac arrive at an advanced stage of the protracted synapse assembly process, and both proteins interact in vitro. In contrast, another AZ-organizing protein, Drosophila Liprin-α (DLiprin-α), localizes to a different subcompartment of the AZ and enters nascent AZs substantially earlier than BRP. Thus, the assembly of the T-bar is instructed by BRP, which seems essential for clustering higher numbers of Ca2+ channels at an advanced stage of AZ maturation.

Results

The AZ protein BRP was recently shown to be crucial for efficient neurotransmission at Drosophila NMJs. Presynaptic AZs missing BRP lacked dense bodies (T-bars), and Ca2+ channel densities were compromised within AZs (Kittel et al., 2006). However, whether BRP performs an essential signaling role in T-bar formation or whether the protein itself is an essential building block of T-bars remained to be clarified. Thus, we entered into a structure-function analysis of BRP.
Structure-function analysis of BRP in T-bar formation

So far, the analysis of BRP function was based on the brp69 allele in which most of the protein-coding sequence (corresponding to aa 283–1,740) is deleted (Fig. 1 C, green). As previously reported, T-bars were missing at brp69 AZs (Fig. 2, compare A and B), and the BRP Nc82 label (Fig. 2, A and B) was absent (Kittel et al., 2006). The BRP N-Term label was also completely absent (Fig. 2 B), indicating that the predicted residual protein (corresponding to aa 1–282) is unstable or at least does not localize to the NMJ. To ensure that brp69 reflects a true null phenotype, we produced the deletion mutant brp6.1 (Fig. 1 C, green) in which all genomic sequences of brp were removed (see Materials and methods). This led to a complete loss of BRP Nc82/BRP N-Term labels and T-bars, whereas some traces of residual electron-dense material appeared at the same frequency as in brp69 (Kittel et al., 2006; unpublished data). As brp6.1 and brp69 (Kittel et al., 2006) behaved identically in all aspects, our previous analysis based on brp69 reflected a true null situation.

BRP is a large protein (1,740 aa). To enter into a structure-function analysis of BRP in T-bar assembly, additional brp alleles were looked into. First, a piggyBac-transposon insert (brp c04298; Fig. 1 C, blue; Bellen et al., 2004) located toward the middle of the locus was characterized. At brp69 NMJs, the BRP Nc82 label was absent, whereas the BRP N-Term label was dramatically reduced (Fig. 2 C). Comparable with our observations...
for brp<sup>69</sup> (Kittel et al., 2006), electron microscopic analysis of brp<sup>o4298</sup> showed a complete lack of T-bars (Fig. 2 C), and only traces of electron-dense material remained at AZ membranes. Thus, as this allele is a site-specific insertion but not a deletion (which in principle might eliminate control elements of genes other than brp), this allele provides further proof that BRP is essential for T-bar assembly. However, as the molecular alterations of brp<sup>o4298</sup> cannot be predicted easily, we sought to analyze aa point mutations in brp. To do so, a chemical mutagenesis screen (ethyl methyl sulfonate [EMS]) selecting for reduced viability over brp<sup>detected at Fig. S1, B and C</sup>). Despite extensive analysis, T-bars were not for brp<sup>5.45</sup> NMJs (Fig. 2 E). T-bar–like structures were observed at brp<sup>5.45</sup> null alleles was performed. However, as the molecular alterations of brp<sup>o4298</sup> cannot be predicted easily, we sought to analyze aa point mutations in brp. To do so, a chemical mutagenesis screen (ethyl methyl sulfonate [EMS]) selecting for reduced viability over brp-null alleles was performed.

The brp<sup>5.45</sup> allele is characterized by a stop codon at aa position 867 (∼50% protein length), which leads to pupal lethality over brp null with weak escapers (Fig. 1 C). As expected, the BRPN<sup>c82</sup> label was absent from brp<sup>5.45</sup> NMJs. Although the number of BRPN<sup>N-Term</sup> clusters was reduced over the whole NMJ (Fig. S1 A), those remaining in brp<sup>5.45</sup> were slightly smaller, although of comparable intensity as in controls (Fig. 2 D; and Fig. S1, B and C). Despite extensive analysis, T-bars were not detected at brp<sup>5.45</sup> NMJs (Fig. 2 D).

The EMS allele brp<sup>1.3</sup> delivered paralyzed adult escapers over brp null as the result of a premature stop codon at aa 1,390 (generating a protein 523 aa longer than predicted for brp<sup>5.45</sup>; Fig. 1 C). Although the number of BRPN<sup>N-Term</sup> clusters was reduced to ∼40% (Fig. S1 A), their sizes and intensities were comparable with controls. At the same time, the BRPN<sup>c82</sup> label was absent (Fig. 2 E). T-bar–like structures were observed at brp<sup>5.45</sup> NMJs (Fig. 2 E), although at lower frequency than in controls (not depicted). However, upon closer inspection, the T-bar–like structures typically appeared truncated (Fig. 2).

For EM, conventional room temperature embedding procedures, including aldehyde fixation and dehydration of the tissue (Fig. 2, A–E), are prone to shrinkage artifacts. To use an alternative conservation method for the analysis of brp<sup>1.3</sup>, we introduced high pressure freezing (HPF)/freeze substitution (FS) EM (Gray et al., 2006; Rostaing et al., 2006; Siksoo et al., 2007) to larval NMJs. With HPF/FS, NMJ tissue appeared well preserved, as judged by the smooth membrane surfaces of, for example, mitochondria (Fig. S2, A and B) or presynaptic boutons (not depicted). Furthermore, electron-dense structures appeared taller, which was likely caused by a reduced loss of material during HPF/FS embedding (e.g., the synaptic cleft; Fig. 2 F). Unlike T-bars visualized in conventionally embedded tissues, HPF/FS-processed T-bars were characterized by filamentous elements at their distal ends (Fig. 2 F, arrowheads). At brp<sup>1.3</sup> NMJs, HPF/FS EM (similarly to our observations obtained with standard EM) typically revealed shortened T-bars (Fig. S2 C, quantification). In conclusion, elimination of aa 1,390–1,740 of BRP did not prevent the formation of T-bar–like assemblies per se. However, these assemblies were significantly smaller than in controls. This result is in line with the assumption that BRP operates as a building block shaping the T-bar.

To both confirm and extend our results, BRP-encoding cDNA fragments (C-terminally GFP tagged for in vivo visualization) were expressed in motoneurons of brp<sup>69</sup> larvae (Fig. 2, G and H). As expected (Kittel et al., 2006), full-length BRP localized to AZs and restored T-bar formation (not depicted). The C-terminally truncated fragment D1-3GFP (BRPD1-3GFP; Δ aa 1,227–1,740; Fig. 1 A and Table I) localized to presynaptic sites but was not sufficient for T-bar formation (Fig. 2 G). Thus, expression of BRPD1-3GFP further suggests that the C-terminal region of BRP (distal of aa 1,226; Table I) is important for T-bar formation.

By expressing D2-4GFP (BRPD2-4GFP; Δ aa 1–267; Fig. 1 A and Table I), the role of the N-terminal region of BRP was tested. The expressed protein was found close to individual AZs (Fig. 2 H). Although T-bars were never observed, small electron-dense aggregates (clearly smaller than T-bars) localized to the AZ membrane at high frequency (Fig. 2 H). Thus, the N-terminal region of BRP seems important for the formation of proper, full-sized T-bars, whereas the C-terminal region is required for the assembly of T-bars by itself. Thus, the entire BRP protein appears to take part in configuring the T-bar structure.

**BRP epitopes reside at the electron-dense T-bar matrix**

To monitor whether BRP epitopes are associated with the electron-dense T-bar matrix, HPF/FS samples were subjected to immuno-EM. The antibodies to both epitopes, BRPN<sup>N-Term</sup> and BRPN<sup>c82</sup>, bound to the T-bar matrix (Fig. 3). Notably, the BRPN<sup>N-Term</sup> antibody showed higher labeling efficacy (Fig. 3, compare B with C) and was found throughout cross-sectional views of T-bars in vertical sections (Fig. 3 B, right).

In contrast, BRPN<sup>c82</sup>-conjugated gold particles were typically found at the most distal edge of electron-dense structures in vertical sections (Fig. 3 C, right). From the expression of ectopic BRP fragments (Fig. 1, A and B) and brp<sup>1.3</sup> (Fig. 2 E), it is clear that the Nc82 epitope must be located distal to aa 1,390 in the C-terminal quarter of the protein. When comparing BRPN<sup>N-Term</sup> and BRPN<sup>c82</sup>, the BRPN<sup>N-Term</sup> label appeared significantly closer to the AZ membrane than the BRPN<sup>c82</sup> label (Fig. 3 D). Thus, the N-terminal region of BRP seems to reside closer to the AZ membrane than the C-terminal region. However, it should be noted that because of the moderate membrane contrast possible with the immuno-EM technique used, the angle at which AZs are observed cannot be exactly determined. This complicates a quantitative analysis. Collectively, both BRP epitopes are found at the T-bar matrix, and the ultrastructural analysis supports the notion that BRP is a direct component of the T-bar.

**BRPN<sup>c82</sup> and BRPN<sup>N-Term</sup> epitopes are vertically segregated relative to theAZ membrane**

To validate and extend our findings concerning the localization of BRPN<sup>N-Term</sup> and BRPN<sup>c82</sup>, further light microscopic experiments were performed. Conventional fluorescence microscopy is highly compatible with protein-specific labeling and enables the processing of high sample numbers. Importantly, confocal sectioning of Drosophila NMJ boutons allows for a reliable definition of the orientation of synapses relative to the optical axis because bouton surfaces are nearly spherical. Hereafter, tangentially imaged AZs are called planar AZs, whereas vertically imaged AZs are referred to as vertical AZs.

We were interested in visualizing the BRP protein with its distinct epitopes in relation to the remainder of the AZ and
Our measurements were performed using sandwiches of primary antibodies and labeled secondary antibodies. For distances in the double-digit nanometer range, the size of individual Ig molecules (used for the detection of epitopes) might be relevant. Thus, we sought to independently validate the distance between BRP N and C termini. To do so, BRPD1-4GFP was expressed in the brp background, and the distance between the BRP C terminus (endogenous GFP fluorescence) and the N terminus (BRPN-Term antibody) was determined (Fig. 4 A). Again, \( \sim 70 \) nm was measured. Finally, the center to center distance between BRP N-Term and CacGFP was measured as \( \sim 60 \) nm (Fig. 4 A). Collectively, we conclude that the BRPNc82 and BRPN-Term epitopes are segregated along an axis perpendicular to the AZ membrane.

The BRP N terminus displays a confined distribution close to the AZ membrane

We proceeded to study the molecular organization of AZs at the Drosophila NMJ with STED microscopy to obtain improved optical resolution in xy coordinates. Previously, it was demonstrated that BRPc82 forms doughnut-shaped structures when visualized at AZs arranged planar to the optical axis (Kittel et al., 2006). BRPc82 doughnuts were reproduced from planar AZs (Fig. 5, A [arrow] and B) with a resolution displaying an
and BRPNc82 was similar to that observed at control AZs (Fig. 5 E). This suggests that individual BRP molecules can adopt an elongated conformation.

The N terminus of BRP overlays the Ca2+ channels at the AZ core

How does the molecular architecture of BRP relate to Ca2+ channels at AZs? Ca2+ channel spots (CacGFP) imaged at standard confocal resolution were found to cocenter with the BRPN-Term label and BRPNc82 doughnuts at planar AZs. At vertical AZs, the CacGFP signal localized toward the AZ membrane relative to both BRPN-Term (Fig. 5 C) and BRPNc82 (Fig. 5, F and G). When imaged with STED resolution, Ca2+ channels consistently localized to small, typically slightly elliptical patches (∼100–150 nm effective point-spread function (PSF) of 80-nm full-width half-maximum. Other than BRPNc82, BRPN-Term did not show a doughnut-shaped distribution when imaged with STED (Fig. 5 D). The combination of STED resolution (confined to one channel in our experiments) for BRPNc82 and confocal resolution for BRPN-Term revealed a polarized and funnel-like distribution of BRP epitopes (Fig. 5, A–C). Notably, the BRPNc82 signal did not appear fully continuous but instead consisted of discrete foci (Fig. 5, B and F) within an overall circular array. In an additional experiment, we expressed full-length BRPD1-4 (Wagh et al., 2006) in brp69 mutants. The distance between BRPN-Term and BRPNc82 was similar to that observed at control AZs (Fig. 5 E). This suggests that individual BRP molecules can adopt an elongated conformation.
Figure 5. STED analysis of AZ organization at Drosophila NMJ synapses. (A) Overview of a bouton stained for BRP\textsuperscript{N-Term} (confocal; magenta) and BRP\textsuperscript{Nc82} (STED; green) showing planar (arrow) and vertical (arrowhead) AZs. (B and C) Magnifications of individual planar (left) and vertical (right) AZs stained for BRP\textsuperscript{Nc82} (STED) and BRP\textsuperscript{N-Term} (confocal; B) and BRP\textsuperscript{N-Term} (STED) and Cac\textsuperscript{GFP} (confocal; C). (D) Mean normalized planar BRP\textsuperscript{N-Term} (magenta) and BRP\textsuperscript{Nc82} (green) arrangement shown with STED resolution (BRP\textsuperscript{N-Term}, n = 14; BRP\textsuperscript{Nc82}, n = 47). (right) The merge superimposed with the intensity profile along one axis through the midpoint for BRP\textsuperscript{N-Term} (magenta) and BRP\textsuperscript{Nc82} (green) is shown. Error bars indicate ± SEM. (E) BRP\textsuperscript{Nc82} (STED) and BRP\textsuperscript{N-Term} (confocal) after expression of full-length BRP cDNA in brp\textsuperscript{69} background (BRP\textsuperscript{D1-4}). (F and G) Individual planar (left) and vertical (right) AZs stained for BRP\textsuperscript{Nc82} (STED) and Cac\textsuperscript{GFP} (confocal; F) and Cac\textsuperscript{GFP} (STED) and BRP\textsuperscript{Nc82} (confocal; G). All images were deconvolved using Improper software. Bars: (A) 1 µm; (G) 100 nm.
along the longest axis) at the AZ center (Fig. 5 G). Thus, the T-bar organized by BRP overlays the field of Ca²⁺ channels at the AZ center.

**DLiprin-α localizes to discrete compartments surrounding the AZ center**

Liprin-α localizes to the AZ and has been shown to be important for the formation of AZs in both *Drosophila* and *C. elegans* (Kaufmann et al., 2002; Dai et al., 2006; Patel et al., 2006). To extend our "AZ map," DLiprin-α<sub>GFP</sub> was expressed in motoneurons and visualized via αGFP stainings with STED microscopy (Fig. 6 A). DLiprin-α localized to presynaptic AZs opposite DGluRIID-positive PSDs. However, as opposed to BRP, DLiprin-α clustered somewhat lateral from the AZ center. STED resolution revealed that DLiprin-α formed discrete "quantal" clusters at the edge of a single AZ co-labeled with BRP<sub>Nc82</sub> (Fig. 6 B) or DGluRIID (Fig. 6 C).

Discrete DLiprin-α clusters were still observable at *brp<sup>69</sup>* NMJs, suggesting that the presence of BRP is not essential for
the recruitment of DLiprin-α to the AZ (Fig. 6 D). However, the localization of BRP, as imaged with STED, appeared aberrant at dliprin-α NMJs (Fig. 6 E). Strikingly, individual BRP doughnuts seemed interconnected, which is directly consistent with the previous observation of complex, multi–T-bar AZs at dliprin-α mutant NMJs (Kaufmann et al., 2002).

BRP and Ca²⁺ channels accumulate late during AZ assembly

So far, we have provided evidence that BRP operates as an essential building block of the T-bar. Notably, fast assembly of T-bars might drive experience-dependent changes of synaptic transmission in the fly central nervous system (Brandstatter et al., 1991; Rybak and Meinertzhagen, 1997). Thus, to learn about the T-bar assembly process in the frame of synapse reorganization, we visualized BRP accumulation in vivo during the developmental assembly of individual synapses (Rasse et al., 2005; Fuger et al., 2007; Schmid et al., 2008). Previously, we found that neuromuscular accumulation of glutamate receptors (as DGluRIIAs) in PSDs typically form at a distance from existing PSDs and then grow over several hours before reaching a final mature size (Rasse et al., 2005; Schmid et al., 2008). Thus, for the analysis of AZ assembly in vivo, DGluRIIA was coimaged to serve as a reference point for our temporal analysis (Fig. 7, A, B, and D; and Table II).

Larvae coexpressing two fluorescently tagged synaptic proteins were imaged (Fig. 7), and quantitative data were obtained to analyze the temporal sequence of protein arrival at developing AZs. For a given larval NMJ, two in vivo images were acquired with a time interval of 12 h. Sites were regarded as new synapses if protein labels exceeded the mean background by a factor of 2.5 at the second (t = 12 h) but not at the first time point (t = 0 h). This way, a temporal sequence of molecular AZ assembly was extracted.

We first compared BRP and DGluRIIA accumulation. For visualization of BRP, we used a fragment of the protein (BRP-short), which delivered a label that fully matched the label of endogenous BRP (Schmid et al., 2008). When BRP-short was examined with STED, doughnuts were detected that resembled those found with BRP (unpublished data). As previously described (Schmid et al., 2008), the accumulation of DGluRIIA clearly preceded BRP arrival in vivo (Fig. 7 A and Table II). Moreover, all postsynaptic DGluRIIA accumulations eventually incorporated presynaptic BRP, demonstrating that DGluRIIA accumulation reliably indicates the formation of new synapses.

As described in the previous section, presynaptic DLiprin-α localization seems to be largely independent of BRP (Fig. 6 D), which is compatible with DLiprin-α functioning upstream of BRP in AZ assembly. Consistently, DLiprin-α incorporation invariably preceded BRP accumulation (Fig. 7 C and Table II).

BRP is crucial for either the initial formation or the maintenance of Ca²⁺ channel clusters. Thus, we analyzed Cac localization at individual developing AZs. Cac and BRP appeared highly correlated, and the timing of Cac accumulation at AZs was typically very close to the advent of BRP with a slight tendency of Cac to precede BRP (Fig. 7 E and Table II).

Collectively, we show that newly forming AZs, similar to PSDs (Rasse et al., 2005), are small to begin with and then increase in size over many hours in vivo before accumulating detectable levels of BRP and reaching a final mature size at developing NMJs. This assembly process of individual new synapses is protracted over hours and is characterized by the contribution of presynaptic and postsynaptic proteins in a defined, overlapping sequence. DLiprin-α appears to be a very early player involved in initializing AZ assembly, whereas BRP, together with Cac, follows only after postsynaptic DGluRIIA incorporation is already detectable (Fig. 7, A and D).

Table II. Quantification of protein accumulation during AZ assembly using NMJ in vivo imaging

| Coexpressed proteins (A × B) | Both A and B | A before B | B before A |
|-----------------------------|--------------|------------|------------|
| BRP × DGluRIIA | BRP⁺/IIA⁺ | BRP⁺/IIA⁺ | BRP⁺/IIA⁺ |
| 17/39 (44%) | 0/39 (0%) | 22/39 (56%) |
| DLiprin-α × DGluRIIA | DLiprin-α⁺/IIA⁺ | DLiprin-α⁺/IIA⁺ | DLiprin-α⁺/IIA⁺ |
| 16/39 (41%) | 23/39 (59%) | 0/39 (0%) |
| BRP × DLiprin-α | BRP⁺/DLiprin-α⁺ | BRP⁺/DLiprin-α⁺ | BRP⁺/DLiprin-α⁺ |
| 8/31 (26%) | 0/31 (0%) | 23/31 (74%) |
| DGluRIIA × Cac | II⁺/Cac⁺ | II⁺/Cac⁺ | II⁺/Cac⁺ |
| 36/62 (58%) | 21/62 (34%) | 5/62 (8%) |
| BRP × Cac | BRP⁺/Cac⁺ | BRP⁺/Cac⁺ | BRP⁺/Cac⁺ |
| 42/62 (68%) | 7/62 (11%) | 13/62 (21%) |

Quantification of the relative accumulation of the indicated synaptic proteins at newly forming AZs (Δt = ∼12 h). A synaptic site was scored positive (+) or negative (−) for a specific protein depending on whether protein fluorescence signals exceeded the mean background level by >2.5-fold. For example, when comparing BRP and DGluRIIA (at synapses forming newly over 12 h), 44% were positive for both proteins, 0% for BRP only, and 36% for DGluRIIA only.
reaching their fully mature size. Thus, we wondered whether BRP may be more important for maturation and potentially less so for initialization of Cac clustering. To address this, we visualized Cac in intact, living brp c04298 mutant larvae together with DGluRIIA as a reference for synapse maturation (Fig. 8 B). In brp c04298 larvae, nascent synapses, identified by their small DGluRIIA accumulations, seemed to accumulate Cac at normal density (Fig. 8, B and C). However, larger, more mature synapses those observed in brp69 (Fig. S3; Kittel et al., 2006). Thus, several independent alleles clearly demonstrate that loss of BRP results in defective clustering of Cac at the AZ, which in turn provokes defects in transmitter release.

Notably, the mislocalization of Cac is not absolute, but instead, a certain degree of Cac remained clustered at AZs in brp null. As our in vivo imaging experiments show (Fig. 7), AZs go through a long, protracted assembly process before finally reaching their fully mature size. Thus, we wondered whether BRP may be more important for maturation and potentially less so for initialization of Cac clustering. To address this, we visualized Cac in intact, living brp104298 mutant larvae together with DGluRIIA as a reference for synapse maturation (Fig. 8 B). In brp104298 larvae, nascent synapses, identified by their small DGluRIIA accumulations, seemed to accumulate Cac at normal density (Fig. 8, B and C). However, larger, more mature synapses...
Alternatively, the Ca\(^{2+}\) channel–clustering defect in \textit{brp} could be caused by impaired Cac transport. To address this, Cac trafficking dynamics were measured using FRAP experiments. Notably, Cac FRAP was slow, with a recovery half-time of \(\approx 12\) h (Fig. 8 D), which is similar to other synaptic membrane proteins such as DGluRIIA (Schmid et al., 2008). Importantly, however, Cac FRAP was essentially unaltered at \textit{brp} boutons, indicating that the long-range transport of Cac to the AZ is not affected by the loss of BRP (Fig. 8 D). Thus, BRP seems to be directly required for Ca\(^{2+}\) channel clustering at AZs but not for its recruitment to the terminal. We aimed to further address the relation between T-bar assembly and Ca\(^{2+}\) channel clustering. Notably, with more substantial DGluRIIA accumulations showed reduced Cac density in the absence of BRP (Fig. 8, B and C). In clear contrast, the recruitment of Cac\(^{GFP}\) to AZs appears to be independent of presynaptic DLiprin-\(\alpha\), as the localization of Ca\(^{2+}\) channels appears normal in \textit{dliprin-}\(\alpha\) mutants (Fig. S4).

Our data imply the following scenario: BRP, an integral component of the T-bar, is recruited to AZs later than DLiprin-\(\alpha\). BRP seems largely dispensable for the initial Ca\(^{2+}\) channel accumulation. However, the matrix organized by BRP is required for maintaining the continuous clustering of Ca\(^{2+}\) channels beneath the T-bar base during the maturation process, which lasts several hours.
brp<sup>1.3</sup> (Fig. 8 A) showed partially restored Cac clustering. Moreover, those AZs positive for BRP<sup>N-Term</sup> in brp<sup>1.3</sup> corresponded to the AZs of restored Cac clustering (Fig. 8 E). Thus, AZ accumulation of the C-terminally truncated but N-terminally intact BRP protein seems to permit the assembly of a distally truncated T-bar, which still functions in clustering Cac.

To further explore this, we prepared larval NMJ AZs via HPF/FS. In this study, a gap between the AZ membrane and the T-bar pedestal (Fig. 8 F, arrows) was observed, where peg-like (Harlow et al., 2001) structures extended from the AZ membrane and distally contacted (or even slightly penetrated) the T-bar pedestal (Fig. 8 F, top, arrowheads). Such pegs were also observed at brp<sup>1.3</sup> AZs (Fig. 8 F, middle, arrowheads) but not at AZ membranes of the brp<sup>69</sup>-null allele (Fig. 8 F, bottom). Thus, these data further support the notion that Cac clustering beneath the T-bar pedestal is restored at brp<sup>1.3</sup> AZs. Moreover, the cytoplasmic domains of Ca<sup>2+</sup> channels might well extend from the AZ membrane, possibly allowing for a direct interaction with BRP.

**Possible direct interaction between the C terminus of Cac and the N terminus of BRP**

To address a possible interaction between BRP and Ca<sup>2+</sup> channels, we tested for protein–protein interactions in vitro. First, all intracellular loops of Cac were tested with BRP constructs in a yeast two-hybrid assay. Only the C-terminal domain of Cac and the N-terminal domain of BRP showed positive for interaction (Fig. 9 A). Thus, in agreement with our hypothesis concerning the orientation of BRP within the T-bar, the BRP N terminus may interact with the Cac C-terminal domain. To further investigate this interaction, we double transfected Schneider S2R+ cells with a GFP-tagged N-terminal construct of BRP (aa 1–617) and Myc-tagged Cac<sup>C-Term</sup>. Western blotting shows the pull-down of the BRP construct in the anti-Myc IP at ~100 kD (arrow). The corresponding band is not detected in the control lanes (IgGs). The slightly different migration of the band in the input lane and in the IP lanes is the result of differences in sample buffer. MW, molecular weight. (C) Spatiotemporal model of AZ assembly and organization at Drosophila NMJs. SVs, synaptic vesicles.
Discussion

Efficient neurotransmission is believed to crucially depend on the structural and functional integrity of the presynaptic AZ compartment (Schoch and Gundelfinger, 2006). An ancestral set of AZ components is conserved between Drosophila, C. elegans, and mammals (Stryker and Johnson, 2007; Jin and Garner, 2008). The strong phenotype we observed at Drosophila NMJs in the absence of the ERC/CAST member BRP lacking T-bar–dense bodies and defective Ca^{2+} channel clustering (Kittel et al., 2006; Wagh et al., 2006), forms an entry point for studying AZ assembly, which is often complicated by redundant and cooperative interactions between AZ components (Jin and Garner, 2008).

BRP and dense body formation

We addressed whether BRP signals T-bar formation (without being a direct component of the T-bar) or whether the protein itself is an essential building block of this electron-dense structure. In this study, we provide evidence that BRP is a direct T-bar component. Immuno-EM identifies the N terminus of BRP throughout the whole cross section of the T-bar (Fig. 3, A and B), and genetic approaches show that a truncated BRP, lacking the C-terminal 30% of the protein’s sequence, forms truncated T-bars (Fig. 2, E and F). Immuno-EM and light microscopy consistently demonstrate that N- and C-terminal epitopes of BRP are segregated along an axis vertical to the AZ membrane and suggest that BRP is an elongated protein, which directly shapes the T-bar structure (Fig. 9 C).

In brp^{141} (predicted as aa 1–866), T-bars were not detected, whereas brp^{21} (aa 1–1,389) formed T-bar–like structures, although fewer and smaller than normal (Fig. 2, E and F; and Fig. S2 C). Moreover, the BRP^{D1-3GFP} construct (1–1,226) did not rescue T-bar assembly. Thus, domains between aa 1,226 and 1,390 of BRP may also be important for the formation of T-bars. Clearly, however, the assembly scheme for T-bars is expected to be controlled at several levels (e.g., by phosphorylation) and might involve further protein components. Nonetheless, it is highly likely that the C-terminal half of BRP plays a crucial role.

BRP, the dense body, and Ca^{2+} channel clustering

As BRP represents an essential component of the electron-dense T-bar subcompartment at the AZ center, it might link Ca^{2+} channel–dependent release sites to the synaptic vesicle cycle (Nehler and Sakaba, 2008). Interestingly, light and electron microscopic analysis has located CAST at mammalian synapses both with and without ribbons (tom Dieck et al., 2005; Deguchi-Tawarada et al., 2006; Sikou et al., 2007). Overall, this study is one of the first to genetically identify a component of an electron-dense synaptic specialization and thus paves the way for further genetic analyses of this subcellular structure.

The N terminus of BRP is found significantly closer to the AZ membrane than the C terminus, where it covers a confined area very similar to the area defined by the CacGFP epitope. Electron tomography of frog NMJs suggested that the cytoplasmic domains of Ca^{2+} channels, reminiscent of pegs, are concentrated directly beneath a component of an electron-dense AZ matrix resembling ribs (Harlow et al., 2001). In addition, freeze-fracture EM identified membrane-associated particles at flesh fly AZs, which, as judged by their dimensions, might well be Ca^{2+} channels (Feeney et al., 1998). We observed peg-like structures beneath the T-bar pedestal. Similar to fly T-bars, the frog AZ matrix extends up to 75 nm into the presynaptic cytoplasm. Based on the amount of cytoplasmic Ca^{2+} channel protein (Catterall, 1998), Harlow et al. (2001) concluded that Ca^{2+} channels are likely to extend into parts of the ribs. Thus, physical interactions between cytoplasmic domains of Ca^{2+} channels and components of ribs/T-bars might well control the formation of Ca^{2+} channel clusters at the AZ membrane. However, a short N-terminal fragment of BRP (aa 1–320) expressed in the brp-null background was unable to localize to AZs efficiently and consistently failed to restore Cac clustering (unpublished data).

The mean Ca^{2+} channel density at AZs is reduced in brp-null alleles. In vitro assays indicate that the N-terminal 20% of BRP can physically interact with the intracellular C terminus of Cac. Notably, we found that the GFP epitope at the very C terminus of CacGFP was closer to the AZ membrane than the N-terminal epitope of BRP (Fig. 4 A). It is conceivable that parts of the Cac C terminus extend into the pedestal region of the T-bar cytomatrix to locally interact with the BRP N terminus. This interaction might play a role in clustering Ca^{2+} channels beneath the T-bar pedestal.

Clearly, additional work will be needed to identify the contributions of discrete protein interactions in the potentially complex AZ protein interaction scheme. Our study should pave the way for a genetic analysis of spatial relationships and structural linkages within the AZ organization. Moreover, we aim to integrate our findings in the framework of mechanisms for Ca^{2+} channel trafficking, clustering, and functional modulation (Cao et al., 2004; Evans and Zamponi, 2006; Catterall and Few, 2008).

Timing of AZ assembly, Ca^{2+} channel accumulation, and synapse maturation

Our imaging assays allowed a temporally resolved analysis of AZ assembly in vivo (Fig. 7). BRP is a late player in AZ assembly, arriving hours after DLiprin-α and also clearly after the postsynaptic accumulation of DGluRIIA. Accumulation of Cac was late as well, although it slightly preceded the arrival of BRP, and impaired Cac clustering at AZs lacking BRP became apparent only from a certain synapse size onwards (Fig. 8, B and C). In this study, we report that new AZs, similar to PSDs (Rasse et al., 2005), form at sites distant from preexisting ones and grow to reach a mature, fixed size. Thus, the late, BRP-dependent formation of the T-bar seems to be required for maintaining high Ca^{2+} channel levels at maturing AZs but not for initializing Ca^{2+} channel clustering at newly forming sites. As the dominant fraction of neuromuscular AZs is mature at a given time point, the overall impression is that of a general clustering defect in brp mutants. In reverse, it will be of interest to further differentiate the molecular mechanisms governing early Ca^{2+} channel clustering. Pre- to postsynaptic communication via neurexin–neuroligin (Missler et al., 2003; Li et al., 2007; Zeng et al., 2007) interactions might well contribute to this process. A further candidate involved in early Ca^{2+} channel clustering is the Fuseless protein, which was recently shown to be crucial for proper Cac localization at AZs (Long et al., 2008).
In summary, during the developmental formation of *Drosophila* NMJ synapses, the emergence of a presynaptic dense body, which is involved in accumulating Ca\(^{2+}\) channels, appears to be a central aspect of synapse maturation. This is likely to confer mature release probability to individual AZs (Kittel et al., 2006) and contribute to matching pre- and postsynaptic assembly by regulating glutamate receptor composition (Schmid et al., 2008). Whether similar mechanisms operate during synapse formation and maturation in mammals remains an open question.

**Outlook: dense body architectures and synaptic plasticity**

In this study, we concentrated on developmental synapse formation and maturation. The question arises whether similar mechanisms to those relevant for AZ maturation might control activity-dependent plasticity as well and whether maturation-dependent changes might be reversible at the level of individual synapses. Notably, experience-dependent, bidirectional changes in the size and number of T-bars (occurring within minutes) were implied at *Drosophila* photoreceptor synapses by ultrastructural means (Brandstatter et al., 1991; Rybak and Meinertzhagen, 1997). Moreover, in the crayfish NMJ, multiple complex AZs with double-dense body architecture were produced after stimulation and were associated with higher release probability (Wojtowicz et al., 1994).

In fact, a recent study has correlated the ribbon size of inner hair cell synapses with Ca\(^{2+}\) microdomain amplitudes (Frank et al., 2003). Either w1 or w1118 was used as background for transgenesis (Kaufmann et al., 2002). Whether similar mechanisms operate during synapse formation in either w1 or w1118 is an open question in future studies.

**Materials and methods**

**Genetics**

All fly strains were reared under standard laboratory conditions (Sigrist et al., 2003). Either w1 or w1118 was used as background for transgenesis (BestGene Inc.).

**Chemical mutagenesis**

The EMS screen was performed according to standard protocols. In brief, isogenic w1118 males were mutagenized with 25 mM EMS solution and crossed to virgins carrying a second chromosomal balancer. For initial mapping, male F1 offspring were crossed with Df(2R)BSC29 or truncated *cac* or truncated *brp* mutant lines. Genomic DNA was extracted from positive candidate flies, and PCR amplicons containing *brp* exon clusters were double-strand sequenced to identify the mutations.

**Generation of *brp* deletions**

*brp* chromosomal deletions were constructed using the Flp recombinase system as previously described (Parks et al., 2004). The different parental lines were provided by the Exelixis collection at Harvard Medical School (Boston, MA).

The following genotypes were used for the ectopic expression of the *brp* constructs (Fig. 1 A; Fig. 2, G and H; and Fig. 5): for wing disc expression, upstream activator sequence (UAS)-BRPΔN-TermGFP/+; UAS-BRPΔN-TermGFP/dpp-Gal4. UAS-BRPΔN-TermGFP/dpp-Gal4. For expression of *brp* constructs at NMJs in the *brp* mutant background, UAS-BRPΔN-TermGFP/+; Df(2R)BSC29/brp\(^{69}\), ok6-Gal4. Df(2R)BSC29/brp\(^{69}\), ok6-Gal4; UAS-BRPΔN-TermGFP/+; Df(2R)BSC29/brp\(^{69}\), ok6-Gal4; UAS-BRPΔN-TermGFP/+; Df(2R)BSC29/brp\(^{69}\), ok6-Gal4. The Gal4 lines and Df(2R)BSC29 were provided by the Bloomington *Drosophila* Stock Center. For experiments in the *dilprina* mutant background (Fig. 6 D), we used *dilprina* \(^{66D}\)/*dilprina* \(^{23}c^{15}\) animals (Kauffman et al., 2002).

**Antibodies**

For the N-terminal BRP (BRPΔN-Term) antibody, a rabbit polyclonal antibody was raised against *Drosophila* BRP (provided by E. Buchner, Universität Würzburg, Würzburg, Germany) antibodies. For immunocytochemistry, 85-nm (silver gold) sections were cut using an EM AFS 6 (Leica). Sections were collected on formvar-coated 100 mesh grids. For immunocytochemistry, sections were dried and poststained with uranyl acetate and lead citrate as described previously (Schmid et al., 2006).

For immunochemistry, grids were placed in droplets of PBS, pH 7.2, until labeling procedure started. Immunocytochemistry was performed previously (Rostaing et al., 2006; Sisikou et al., 2007). The antibody containing serum was affinity purified versus the same protein as used for immunization.

**Transmission EM**

For EM, about one to three *Drosophila* late second/early third instar larvae were placed in aluminum specimen carrier of 200-\(\mu\)m depth (type A; Bal-Tec), filled with yeast paste, and covered with a lid. The samples were frozen immediately in an HPK machine (HPK 010; Bal-Tec) and rapidly transferred to liquid nitrogen for storage.

FS and embedding were performed in acetone in either an EM ATS 6 (Leica). Two separate protocols were used for morphological and immunocytochemical analysis (Rostaing et al., 2006; Sisikou et al., 2007). For immunocytochemistry, the substitution was performed in pure acetone without uranyl acetate.

For morphology, 55–60-nm (gray silver) sections, and for immunocytochemistry, 85-nm (silver gold) sections were cut using an EM AFS 6 (Leica). Sections were collected on formvar-coated 100 mesh grids. For immunocytochemistry, sections were dried and poststained with uranyl acetate and lead citrate as described previously (Schmid et al., 2006). For immunocytochemistry, grids were placed in droplets of PBS, pH 7.2, until labeling procedure started. Immunocytochemistry was performed previously (Rostaing et al., 2006; Sisikou et al., 2007). The antibody containing serum was affinity purified versus the same protein as used for immunization.

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**Molecular cloning**

For expression constructs of DlPrima-αGFP, a 3.6-kb fragment of pOT2 ID27334 was subcloned into pBluescript KS(+) (Agilent Technologies) using the *Sal* and EcoRI restriction sites introduced by PCR primers and subsequently double-strand sequenced. The insert was excised and inserted into pENTR4 (Invitrogen) via *Sal* and Not site. The final expression construct of DlPrima-αGFP was obtained using the Gateway system (Invitrogen). In brief, pENTR4 DlPrima-α was recombinated with pTWG (a *Drosophila* pUAST Gateway vector developed in the laboratory of T. Murphy, The Carnegie Institution of Washington, Baltimore, MD).

The BRP constructs used for expression in *Drosophila* flies and *Drosophila* Schneider cell culture were obtained by PCR using the corresponding cDNA as template (brp cDNA; Wagh et al., 2006) and cloned into pCRII, a modified version of pENTR4 using the SpeI and Asp718 restriction sites.

**pUBiP Gateway destination vectors, used for coexpression experiments in S2R+ cell culture, were obtained from A. Herzig (Max Planck Institute, Göttingen, Germany). These contained a ubiquitin promoter and either a N- or C-terminal tag.**

**Yeast two-hybrid constructs for cac and brp were obtained by PCR using the corresponding cDNA as template (cac cDNA was provided by R.W. Ordway, The Pennsylvania State University, Philadelphia, PA; brp cDNA; Wagh et al., 2006) and cloned into the bait vector pGBK7 (Clontech Laboratories, Inc.) or the prey vector pGAD77 (Clontech Laboratories, Inc.) using the restriction sites introduced with the PCR primers.**

**Vectors used for fusion protein expression in *Drosophila* Schneider cell culture were made from respective modified pENTR4 clones containing truncated brp or truncated cac using the Gateway system.**

**Molecular cloning in detail**

**pCRII.** The multiple cloning site of pCRII (Invitrogen) was modified by oligonucleotide annealing using two primers with a 5′ phosphate modification (MWG-Biotech AG): 5′-CATGGGAACCTGTTCCGCCGCGCGCGCGCGG-
CCCGGTACCGC-3 and 5'-TCGAGCTGTAACGGGGCCCGC-3

The annealed oligonucleotides were ligated into a previously cut pENTR4 vector using the Ncol and XhoI restriction sites. This modification resulted in a loss of the cCAB gene and a new multiple cloning site.

**pTMwStrawberry.** In short, the *Drosophila* Gateway vector pTWG containing an EGFP tag placed downstream of the Gateway cassette was used as a template to replace the EGFP by pStrawberry (mStraw; Invitrogen) for antibody fragments were amplified by PCR using either pTWG (Gateway vectors.html#_Copyright_2003,_Carnegie. The following

La Jolla, CA). For general information about the mStraw was provided by R.Y. Tsien, University of California, San Diego, used as a template to replace the EGFP by mStrawberry (mStraw; pRSETB

were used. The PCR product was cut using NcoI–EcoRI.

were used. The PCR product was cut using NcoI–SalI.

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Mowiol. The following antibody dilutions were used: Mx-Nc82 (provided by E. Buchner), 1:200; Mx-DGLuRIIA (B84D2; Developmental Studies Hybridoma Bank), 1:100; Rb-BRPN-Term, 1:250; Rb-DGLuRIID (Qin et al., 2005), 1:500; Mx-GFP (Invitrogen), 1:500; Rb-GFP (Invitrogen), 1:500; and HRP-Cy5 (Dianova), 1:250. For standard immunostainings, secondary antibodies were diluted 1:500. For STED staining, dyes ([Atto647N] conjugation to secondary antibodies giving sheep α-MAtto647N and sheep α-RbAtto647N were performed according to producer protocols [http://www.atto-tec.com] and used 1:100. Embryos (T[11E]/ and elav-Gal4, F[1]/ T[13C1H129]; Kawasaki et al., 2002) were staged by time (22–24 h after egg laying) and morphology, dissected, and stained as described for larvae.

Live imaging
The following strains were used for in vivo imaging experiments: for Ca2+ channels, ok-Gal4/+; Ca2+SPG, DGLuRIIA/FRT10a (+); control, Df(2R)BSC29, ok-Gal4/brp; Ca2+SPG, DGLuRIIA/FRT10a (+); brp mutant background. For temporal analysis of AZ assembly, ok-Gal4/+; BRF-short/FRT10a, ok-Gal4/+/; DIPrin-GFP/FRT10a, ok-Gal4/+/; and DIPrin-short/FRT10a, ok-Gal4/+, DIPrin-GFP/FRT10a (+). Imaging of intact Drosophila larvae along with larval anaesthesia was performed as described previously (Rasse et al., 2005; Fuger et al., 2007; Schmid et al., 2008). For FRAP experiments, intense 488-nm laser light was applied inside a region of interest of ~10 μm of edge length (zoom, 25) bleaching both green and red fluorescent dyes. After an incubation of 12 h at 25°C, the junctions were imaged and compared with the prebleached pictures. Control regions were conserved at the junctions for internal control of intensity levels.

Image processing
Images were acquired using Application Suite Advanced Fluorescence (LAS-AF; Leica) software.

Confocal imaging
Confocal stacks were processed with ImageJ software (National Institutes of Health). Single slices and confocal stacks were deconvoluted using the ImageJ plug-ins iterative deconvolution and iterative deconvolution 3D, respectively (provided by B. Dougherty, OptiNav, Inc., Redmond, WA). To generate the PSF for deconvolution, the ImageJ plug-in diffraction PSF 3D (provided by B. Dougherty) was used. The PSF settings were adjusted according to our hardware parameters, emission wavelengths, and image dimensions.

For quantification of areas and intensities, maximum projections, acquired with comparable confocal settings, were thresholded at 30 arbitrary units (au), and remaining areas were measured via the analyze particle function of ImageJ.

To obtain unbiased mean BRP Nc82 and BRP N-Term distributions with STED resolution (Fig. 5 D), we used STED images of BRP Nc82–Atto647N (or BRP N-Term–Atto647N) that were simultaneously recorded with confocal images of the BRF GFP–Alexa Fluor 488 (or BRFna42–Alexa Fluor 488). From these images, AZs were selected, which appeared planar to the optical slice. The confocal channel of each image of a planar AZ was automatically fitted with a 2D Gaussian function with Mathematica (version 5.0; Wolfram Research). The peaks of the Gaussian functions were used to automatically align and subsequently average the corresponding STED signal. The averaged STED images (BRP Nc82 and BRP N-Term) were finally aligned according to the corresponding confocal counterpart. The BRP Nc82 signal was fitted with a single Gaussian (standard deviation = 52 nm), and the BRP N-Term signal was fitted with the sum of two Gaussians (standard deviation = 48 nm and peak distance Δx = 189 nm).

STED imaging
STED images were processed using a linear deconvolution software integrated into the Inspector Software package (Max-Planck Innovations GmbH). Regularization parameters ranged from 1e-2 to 1e-12. The PSF was generated by using a 2D Lorentzian function with its half-width and half-length and fitted to the half-width and half-length obtained by images of 25-nm crimson beads conjugated to Atto647N.

Electrophysiology
Two electrode voltage clamp recordings were essentially conducted as previously described (Kittel et al., 2006). All experiments were performed on male third instar larval NMJs (muscle 6; segments A2 and A3) in HL3 (70 mM NaCl, 5 mM KCl, 20 mM MgCl2, 10 mM NaHCO3, 5 mM trehalose, 115 mM sucrose, 5 mM Hepes, and 1 mM CaCl2, pH 7.2). Muscle cells had an input resistance ≥4 MΩ. Intracellular electrodes were filled with 3 M KCl, and resistances ranged from 10–25 MΩ. Stimulation artifacts of evoked excitatory junctional currents were removed for clarity. Genotypes used were brpΔbrp/GFP/brpΔbrp and w1118 as controls.

Statistics
Data were analyzed using the Mann-Whitney rank sum test for linear independent data groups (Prism; GraphPad Software, Inc.). Means are annotated ± SEM. Asterisks are used to denote significance (*, P < 0.05; **, P < 0.01; ***, P < 0.005; not significant, P > 0.05).

Online supplemental material
Fig. S1 shows a quantification of BRP Nc82 expression levels in several brp mutants. Fig. S2 shows a comparison of mitochondrial ultrastructure either preserved using conventional room temperature embedding with aldehyde fixation and dehydration or HPF/FS along with T-bar size quantification of conventional and HPF/FS-prepared control and brpΔ larvae. Fig. S3 shows amplitude and rise time of evoked excitatory junctional current recorded from brpΔ larvae. Fig. S4 shows that dIPrin-Δ2s A2s do not suffer from Cac-clustering defects. Fig. S5 shows that BRP clustering is not disturbed at cac A2s. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200812150/DC1.

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