Elucidation of Olive Oil Oxidation Mechanisms by Analysis of Triacylglycerol Hydroperoxide Isomers Using LC-MS/MS

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Abstract: Despite the importance of the insight about the oxidation mechanisms (i.e., radical and singlet oxygen ($^1\text{O}_2$) oxidation) in extra virgin olive oil (EVOO), the elucidation has been difficult due to its various triacylglycerol molecular species and complex matrix. This study tried to evaluate the mechanisms responsible for EVOO oxidation in our daily use by quantitative determination of triacylglycerol hydroperoxide (TGOOH) isomers using LC-MS/MS. The standards of dioleoyl-(hydroperoxy octadecadienoyl)-triacylglycerol and dioleoyl-(hydroperoxy octadecamonoenoyl)-triacylglycerol, which are the predominant TGOOHs contained in EVOO, were prepared. Subsequently, fresh, thermal-, and photo-oxidized EVOO were analyzed. The obtained results mostly agreed with the previously reported characteristics of the radical and $^1\text{O}_2$ oxidation of linoleic acid and oleic acid. This suggests that the methods described in this paper should be valuable in understanding how different factors that determine the quality of EVOO (e.g., olive species, cultivation area, cultivation timing, and extraction methods) contribute to its oxidative stability.

Keywords: radical oxidation; singlet oxygen oxidation; olive oil; triacylglycerol hydroperoxide; mass spectrometry; oxidation mechanisms; hydroperoxide positional isomers

1. Introduction

Extra virgin olive oil (EVOO) is rich in monounsaturated fatty acids and contains characteristic components, such as phenolics, tocopherol, carotenoids, and chlorophyll [1–13]. These components contribute to its unique characteristics (e.g., taste, color, and flavor) and, accordingly, EVOO is used widely in the food and cosmetics industries. However, these characteristics are often impaired by oxidation. Therefore, to provide guidance on preventing oxidation, extensive efforts have been made to reveal the oxidation mechanisms in EVOO [1–14]. Nevertheless, the mechanisms that contribute to the oxidation of EVOO in our daily use are not yet fully understood.

EVOO is mainly composed of linoleic acid (FA 18:2(9Z,12Z)) and oleic acid (FA 18:1(9Z)). These fatty acids can be oxidized by radical oxidation (e.g., auto- and thermal-oxidation) and/or singlet oxygen ($^1\text{O}_2$) oxidation (e.g., photo-oxidation) during our daily use [15–17]. Radical oxidation is initiated by the abstraction of H· from an allylic hydrogen within the fatty acid structure. It is well-known that FA 18:1(9Z) is more resistant to radical oxidation than FA 18:2(9Z,12Z) due to the absence of a bis-allylic hydrogen. Alternatively, in a typical $^1\text{O}_2$ oxidation reaction, the irradiation of light to photosensitizers yields $^1\text{O}_2$, which reacts with the double bond of fatty acids via the ene reaction. Hence, in $^1\text{O}_2$ oxidation, the rate of hydroperoxide formation is proportional to the number of double bonds [18]. Unlike other edible oils, EVOO contains a high amount of chlorophyll...
(which acts both as a photosensitizer [1,4,8,19] and a radical scavenger [7,9,20]), various polyphenols [4,7,9] and tocopherols [4,7,9,17,21] (which act as radical scavengers) and carotenoids [4,17,22] (which act as $^1$O$_2$ quenchers). Consequently, this complex composition hinders the identification of the mechanisms (i.e., radical or $^1$O$_2$ oxidation) that oxidize EVOO in our daily use.

Primary oxidation of lipids affords lipid hydroperoxide (LOOH) isomers, whose structures (i.e., hydroperoxyl group binding positions) depend on oxidation mechanisms (i.e., radical and $^1$O$_2$ oxidation; Figure 1) [15,17]. In other words, triacylglycerol (TG) oxidation mechanisms can be identified by characterizing TG hydroperoxide (TGOOH) isomers. However, analyzing TGOOH isomers has been a great challenge, even using the latest analytical techniques and instruments [23–27]. To overcome such issues, we recently developed methods to analyze the positional isomers of various LOOHs by utilizing sodium ions during electrospray ionization (ESI)-LC-MS/MS [28,29]. Using this method, we analyzed the major TGOOH contained in canola oil (TG 18:1_18:1_18:2;OOH) and identified that canola oil was predominantly oxidized by $^1$O$_2$ oxidation during storage [29]. The study led us to believe that our LC-MS/MS method can further be applied to elucidate the oxidation mechanisms of EVOO that possess a more complex matrix than canola oil.

Figure 1. Triacylglycerol (TG) oxidation mechanisms and chemical structures of TG 18:1_18:1_18:2;OOH isomers (A) and TG 18:1_18:1_18:1;OOH isomers (B). Isomeric structure of TGOOH depends on oxidation mechanisms (radical and $^1$O$_2$ oxidation). The shorthand notation of lipids was in accordance with LIPID MAPS [30].
In this study, to elucidate the oxidation mechanisms of EVOO, we analyzed TG 18:1_18:1_18:2;OOH isomers in fresh, thermally oxidized, and photo-oxidized EVOO using our LC-MS/MS method described above. Oxidation was performed under conditions that resembled our daily use (e.g., storage and cooking). Additionally, because the susceptibility of FA 18:1(9Z) towards both radical and $^{13}\text{O}_2$ oxidation is different from that of FA 18:2(9Z,12Z), we also aimed to analyze the oxidation mechanisms of FA 18:1(9Z). Hence, the predominant TGOOH molecular species in EVOO that possesses FA 18:1;OOH, TG 18:1_18:1_18:1;OOH isomers were also analyzed. The insights obtained in this study should be helpful to comprehensively understand the mechanisms underlying the oxidation of EVOO.

2. Materials and Methods

2.1. Materials

TG 18:1_18:1_18:1 was purchased from Sigma-Aldrich (St. Louis, MO, USA). 2-Methoxypropene (MxP) and 2,2′-azobis-(4-methoxy-2,4-dimethylvaleronitrile) (MeO-AMVN) were obtained from FUJIFILM Wako Pure Chemical Corporation (Osaka, Japan). Fresh EVOO was collected immediately after production at J-Oil MILLS, Inc. (Tokyo, Japan) and stored under N$_2$ gas and shading until use. All other reagents were of the highest grade available.

2.2. Preparation of TG 18:1_18:1_18:2;OOH and TG 18:1_18:1_18:1;OOH Standards

Individual standards of the twelve TG 18:1_18:1_18:2;OOH isomers (Figure 1A) were prepared previously [29]. TG 18:1_18:1_18:1;OOH, as a mixture of the isomers (Figure 1B), was prepared as follows. As an initiator of radical oxidation, 10 µL of MeO-AMVN (500 mg/mL in chloroform) was added to 1 g of TG 18:1_18:1_18:1. To obtain a crude mixture containing TG 18:1_18:1_18:1;OOH isomers, chloroform was evaporated under a N$_2$ gas stream and TG 18:1_18:1_18:1 was oxidized under heating at 40–50 °C for 20 h. The hydroperoxyl group of TG 18:1_18:1_18:1;OOH was protected with MxP using a method described previously [28,29,31]. The protected TG 18:1_18:1_18:1;OOH was isolated by semi-preparative HPLC (LC-6AD (Shimadzu, Kyoto, Japan)) using an Inertsil ODS-3 column (10 µm, 20 × 250 mm) at 40 °C with a mobile phase consisting of methanol/2-propanol (3:2, v/v). The flow rate was set at 20 mL/min and the eluent was monitored with a UV-detector (SPD-20A (Shimadzu, Kyoto, Japan)) at 210 nm. The obtained protected TG 18:1_18:1_18:1;OOH was deprotected and purified as described previously [28,29,31]. A portion of the purified TG 18:1_18:1_18:1;OOH was methyl esterified, subjected to gas chromatography (GC) analysis, and intact acyl residues (i.e., FA 18:1) were measured to determine the concentration of TG 18:1_18:1_18:1;OOH [29]. The prepared TG 18:1_18:1_18:1;OOH was dissolved in 2-propanol and stored at −80 °C until use.

The shorthand notations of TGOOH isomers (and other lipids) described in this study follow the LIPID MAPS nomenclature (Table 1) [30].

2.3. MS/MS and LC-MS/MS Analysis of TGOOH Isomers

Q1 mass and product ion mass spectra in the TG 18:1_18:1_18:1;OOH isomer mixture were obtained using a 4000 QTRAP mass spectrometer (SCIEX, Tokyo, Japan). Standard TG 18:1_18:1_18:1;OOH was diluted in methanol (0.5 µM) and directly infused into the MS at a flow rate of 10 µL/min. Positive ESI was used as the ion source. MS spectra were obtained in a range of m/z 100–1000. Analytical parameters were optimized using the Analyst software (ver. 1.6.2, SCIEX, Tokyo, Japan) (Supplementary Materials).
TGOOH

| Molecular Species Level | Hydroperoxyl Group Positional Isomer Level | EZ Isomer Level of Hydroperoxyl Fatty Acid | Fatty Acid Positional Isomer Level | Causative Oxidation Mechanism | Compound Number (Figures 2–4) |
|-------------------------|------------------------------------------|------------------------------------------|----------------------------------|-------------------------------|-----------------------------|
| TG 18:1_18:1_18:2:OOH  | TG 18:1_18:1_18:2(10E,12Z);9OOH          | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 1                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 2                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 3                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 4                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 5                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 6                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 7                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 8                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 9                             |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 10                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 11                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 12                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 13                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 14                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 15                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical                           | 16                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 17                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 18                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 19                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 20                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 21                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 22                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 23                            |
|                         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | TG 18:1_18:1_18:2(10E,12Z);9OOH         | Radical and $^1$O$_2$             | 24                            |
| Molecular Species Level | Hydroperoxyl Group Positional Isomer Level | EZ Isomer Level of Hydroperoxy Fatty Acid | Fatty Acid Positional Isomer Level | Causative Oxidation Mechanism | Compound Number (Figures 2–4) |
|------------------------|-------------------------------------------|---------------------------------------------|----------------------------------|--------------------------------|--------------------------------|
| FA 18:2;1OMe,OOH       | FA 18:2;1OMe,9OOH                         | FA 18:2(10E,12Z);1OMe,9OOH                 |                                 |                                |                                |
| FA 18:2;1OMe,13OOH     | FA 18:2(9E,11E);1OMe,13OOH               | FA 18:2(9E,11E);1OMe,13OOH                 |                                 |                                |                                |
| FA 18:1;1OMe,8OOH      | FA 18:1(9Z);1OMe,8OOH                    | FA 18:1(9Z);1OMe,8OOH                     |                                 |                                |                                |
| FA 18:1;1OMe,10OOH     | FA 18:1(8E);1OMe,10OOH                   | FA 18:1(8E);1OMe,10OOH                    |                                 |                                |                                |
| FA 18:1;1OMe,11OOH     | FA 18:1(9Z);1OMe,11OOH                   | FA 18:1(9Z);1OMe,11OOH                    |                                 |                                |                                |
| TG 18:1,18:1,18:2      |                                            |                                             |                                 |                                |                                |
| TG 18:1,18:1,18:1      |                                            |                                             |                                 |                                |                                |
| FA 18:1                |                                            |                                             |                                 |                                |                                |
| FA 18:2                |                                            |                                             |                                 |                                |                                |
TG 18:1_18:1_18:2;OOH and TG 18:1_18:1_18:1;OOH isomers were analyzed in multiple-reaction-monitoring (MRM) mode. The MRM transitions described in the Supplementary Materials were used. LC-MS/MS analysis was conducted using an ExionLC HPLC system (SCIEX, Tokyo, Japan) equipped with a 4000 QTRAP mass spectrometer. An Inertsil SIL-10A column (5 µm, 2.1 x 250 mm, GL Sciences Inc., Tokyo, Japan) was eluted with hexane/2-propanol/acetic acid (100:0.6:0.5, v/v/v) at 0.2 mL/min (40 °C). A post-column solvent consisting of methanol/2-propanol (1:1, v/v) containing 0.2 mM sodium acetate was mixed with the eluent at 0.2 mL/min to promote ionization [29]. TG 18:1_18:1_18:2;OOH and TG 18:1_18:1_18:1;OOH isomers were quantitated with external standard curves.

2.4. Oxidation of EVOO

Fresh EVOO (400 mL) was thermally oxidized (radical oxidation) in an amber 500 mL glass beaker under gentle stirring. The beaker was heated in an oil bath kept at 150 °C in the dark.

Photo-oxidation (¹⁰₂ oxidation) of fresh EVOO (400 mL) was performed in a clear 500 mL beaker under gentle stirring. The beaker was irradiated with light-emitting diode (LED) light (5000 lux) at 26 ± 1 °C.

Oxidized EVOO samples were collected at 20 min intervals until 240 min (n = 3). Portions of the collected samples were diluted 10,000-fold in hexane and analyzed with LC-MS/MS (10 µL).

3. Results and Discussion

3.1. Target TGOOH to Determine EVOO Oxidation Mechanisms

Edible oils, such as EVOO, contain various TG molecular species. Moreover, their oxidation results in an even greater number of hydroperoxyl group positional isomers. Therefore, the analysis of TGOOH isomers in edible oils remains challenging. Meanwhile, because different oxidation mechanisms (e.g., radical and ¹⁰₂ oxidation) afford different hydroperoxide isomers [15–17], analysis of TGOOH isomers enables the evaluation of oxidation mechanisms. In this study, we focused on the oxidation of FA 18:2(9Z,12Z) and FA 18:1(9Z). Of the TG molecular species that contain these fatty acids, TG 18:1_18:1_18:2 and TG 18:1_18:1_18:1 are the most predominant in olive oil [26]. Hence, we sought to analyze their hydroperoxides (TG 18:1_18:1_18:2;OOH isomers and TG 18:1_18:1_18:1;OOH isomers) contained in EVOO (Figure 1).

As described above, determining hydroperoxyl group positions is pivotal to evaluate the mechanisms responsible for EVOO oxidation. Thus, previous studies analyzed TGOOH hydroperoxyl group positions after derivatization reactions (e.g., reduction, trimethylsilylation, and methyl esterification). However, because the hydroperoxyl group is relatively unstable, artifacts can be formed during derivatization. Therefore, a direct analysis should be favored over derivatization methods. Meanwhile, most of the previous studies that directly analyzed TGOOH depended solely on molecular weight (i.e., intact TG molecular weight + 32 Da) and, hence, their isomers were not analyzed [23–27]. Under these circumstances, we discovered that the collision-induced dissociation (CID) of the sodium adducts of LOOH provide hydroperoxyl group-position-specific product ions based on α-cleavage [28,29]. Using this method, we analyzed hydroperoxyl group positions for the main TGOOH in canola oil (TG 18:1_18:1_18:2;OOH isomers) and found that FA 18:2(9Z,12Z) in canola oil was oxidized predominantly by ¹⁰₂ oxidation during storage [29]. Therefore, in this study, we aimed to apply the above method to determine the oxidation mechanisms of FA 18:2(9Z,12Z) in EVOO. Additionally, to obtain further insight into the oxidation of EVOO (i.e., oxidation mechanisms of FA 18:1(9Z)), TG 18:1_18:1_18:1;OOH isomers in EVOO were also analyzed.
3.2. Analysis of TG 18:1_18:1_18:2;OOH Isomers in Fresh, Thermally Oxidized, and Photo-Oxidized EVOO

TG 18:1_18:1_18:2;OOH isomers in fresh EVOO were analyzed using our previously developed LC-MS/MS method [29]. Typical chromatograms are shown in Figure 2A. Despite being analyzed immediately after opening, TG 18:1_18:1_18:2;OOH isomers were clearly detected from fresh EVOO. Their concentrations were as follows: 647.3 ± 138.4 µM for TG 18:1_18:1_18:2(10E,12Z);9OOH, 89.1 ± 19.3 µM for TG 18:1_18:1_18:2(10E,12E);9OOH, 5.6 ± 1.1 µM for TG 18:1_18:1_18:2(9Z,11E);13OOH, and 65.3 ± 12.3 µM for TG 18:1_18:1_18:2(9E,11E);13OOH (Figure 2B). We previously reported that the concentrations of TG 18:1_18:1_18:2;OOH isomers in fresh canola oil were ~0.8 µM [29]. We also identified that O2 oxidation mainly contributed to canola oil oxidation based on isomer analysis [29]. The higher content of these isomers in EVOO, identified in this study, agrees with the fact that EVOO generally has higher peroxide values than other refined oils (e.g., soybean oil and canola oil) [7–9,11,12]. EVOO demonstrates higher peroxide values because it is, unlike other oils, typically obtained only by mechanical means and lacks chemical purification processes (e.g., decidification and deodorizing) to maintain its characteristics (e.g., taste, color, and flavor) [32]. Regarding isomer compositions, concentrations of TG 18:1_18:1_18:2;9OOH and TG 18:1_18:1_18:2;13OOH notably increased by thermal oxidation, whereas concentrations of TG 18:1_18:1_18:2(10E,12Z);9OOH and TG 18:1_18:1_18:2(9Z,11E);13OOH were higher than those of TG 18:1_18:1_18:2(9E,11E);13OOH (Figure 2D). It is well known that the composition of EZ and EE isomers depends on oxidation temperatures [16,29]. Additionally, studies have shown that the composition of EZ and EE isomers also depends on the presence of antioxidants because hydroperoxyl radicals isomerize to the thermodynamically favored EE form in the absence of proton donors [33]. Therefore, the above composition of EE and EZ isomers should have reflected the oxidation temperature and the concentration of antioxidants in the EVOO used in this study. Interestingly, the concentration of TG 18:1_18:1_18:2(10E,12E);9OOH was higher than that of TG 18:1_18:1_18:2(10E,12Z);9OOH and TG 18:1_18:1_18:2(9E,11E);13OOH, even though their decomposition rates were similar. Meanwhile, in a previous study that investigated the oxidation of FA 18:2(9Z,12Z);1OMe, no difference was observed in the amount of FA 18:2;1OMe;9OOH and FA 18:2;1OMe;13OOH formed [15,17]. Therefore, a preference as to where the hydroperoxyl group is inserted, not seen in the fatty acid form, and may exist in the TG form. Concentrations of TG 18:1_18:1_18:2;10OOH and TG 18:1_18:1_18:2;12OOH did not significantly change during thermal oxidation (Figure 2D).

Photo-oxidation of EVOO was conducted at 5000 lux to represent storage under cloudiness. A typical chromatogram of EVOO, photo-oxidized for 240 min, is shown in Figure 2E. Photo-oxidation of EVOO resulted in a slight increase in the concentrations of TG 18:1_18:1_18:2(10E,12Z);9OOH (5.6 ± 1.1 µM (0 min) → 11.8 ± 0.5 µM (240 min)) and TG 18:1_18:1_18:2;12OOH (5.1 ± 0.9 µM (0 min) → 13.8 ± 0.5 µM (240 min)), which are the isomers formed only by O2 oxidation (Figure 2F). O2 oxidation also yields TG 18:1_18:1_18:2(10E,12Z);9OOH and TG 18:1_18:1_18:2(9Z,11E);13OOH, in addition to the above two isomers [15–17,29]. However, changes in their concentrations were not observed during photo-oxidation, presumably due to their relatively high initial concentrations in the EVOO used in this study.
As we expected, analysis of TG 18:1_18:1_18:2;OOH isomers enabled the elucidation of the oxidation mechanisms of EVOO that possess a complex matrix. On the other hand, the most abundant fatty acid in EVOO is FA 18:1(9Z). Therefore, to obtain further insights into the oxidation mechanisms of EVOO, TG 18:1_18:1_18:1;OOH isomers were next analyzed.

3.3. MS/MS and LC-MS/MS Analysis of TG 18:1_18:1_18:1;OOH Standards

Oxidation of TG 18:1_18:1_18:1 yields TG 18:1_18:1_18:1;OOH isomers, whose structures depend on oxidation mechanisms (i.e., radical and $^{1}$O$_{2}$ oxidation; Figure 1) [15–17]. In this study, to accurately analyze TG 18:1_18:1_18:1;OOH isomers and oxidation mechanisms, a standard mixture of the isomers was prepared. We initially attempted to synthesize...
TG 18:1_18:1_18:1;OOH isomers in a way similar to TG 18:1_18:1_18:2;OOH isomers [29], i.e., by first synthesizing each FA 18:1;OOH isomer, then esterifying them to DG 18:1_18:1. However, because we were unable to separate each FA 18:1;OOH isomer using HPLC, a mixture of TG 18:1_18:1_18:1;OOH isomers was prepared via the direct oxidation of TG 18:1_18:1_18:1. TG 18:1_18:1_18:1 was oxidized with MeO-AMVN, a radical initiator, as all TG 18:1_18:1_18:1;OOH isomers can be formed by radical oxidation [15–17]. Then, by selectively protecting the hydroperoxyl group of the crude radical oxidation product with MxP [28,29,31], a mixture of TG 18:1_18:1_18:1;OOH isomers was obtained with high purity. Q1 mass analysis of the prepared standard mixture demonstrated a clear single peak at m/z 940 ([M+Na]+; Figure 3A).

Concentrations of each TGOOH isomer in the prepared mixture were then calculated. Firstly, the total concentration of TG 18:1_18:1_18:1;OOH isomers was determined by GC. Then, we approximated that the mixture contained hydroperoxyl group positional isomers in equal amounts (i.e., TG 18:1_18:1_18:1;8OOH (25%), TG 18:1_18:1_18:1;9OOH (25%), TG 18:1_18:1_18:1;10OOH (25%), and TG 18:1_18:1_18:1;11OOH (25%).) This approximation reflected the radical oxidation pathway of FA 18:1(9Z); theoretically, the hydroperoxyl group should be equally distributed among C8–C11 positions. Indeed, a previous study on FA 18:1(9Z);1OMe demonstrated that there is barely a preference in hydroperoxide group positions during radical oxidation (i.e., FA 18:1;1OMe,8OOH (26–28%), FA 18:1;1OMe,9OOH (22–25%), FA 18:1;1OMe,10OOH (22–25%), and FA 18:1;1OMe,11OOH (26–28%)) [15–17].

Subsequently, product ion analysis of the prepared mixture was performed. Sodium adducts were analyzed based on our previous findings that the CID of LOOH sodium adducts induces hydroperoxyl group position-specific product ions based on α-cleavage [34]. As we expected, each TG 18:1_18:1_18:1;OOH isomer afforded the following specific product ions: m/z 755 for TG 18:1_18:1_18:1;8OOH, m/z 769 for TG 18:1_18:1_18:1;9OOH, m/z 810 for TG 18:1_18:1_18:1;10OOH, and m/z 824 for TG 18:1_18:1_18:1;11OOH (Figure 3B,C). These product ions were used as MRM transitions (Supplementary Materials) Under optimized LC-MS/MS conditions, clear peaks were detected from the prepared mixture of TG 18:1_18:1_18:1;OOH isomers. MRM of 940 > 755 (TG 18:1_18:1_18:1;8OOH) and 940 > 824 (TG 18:1_18:1_18:1;11OOH) each detected four peaks (Figure 3D), corresponding to the number of fatty acid positional isomers and EZ isomers (Figure 1, Table 1). On the other hand, MRM of 940 > 769 (TG 18:1_18:1_18:1;9OOH) and 940 > 810 (TG 18:1_18:1_18:1;10OOH) each detected two peaks corresponding to the number of fatty acid positional isomers (Table 1). Regarding their quantitation, we assumed, based on our previous study [29], that the ionization efficiencies among the fatty acid positional isomers and EZ isomers are the same. The prepared calibration curves demonstrated good linearity within a range of 0.0147 to 0.29 pmol for TG 18:1_18:1_18:1;8OOH (r² = 0.9889), TG 18:1_18:1_18:1;9OOH (r² = 0.9854), and TG 18:1_18:1_18:1;11OOH (r² = 0.9888) (Figure 3E).

3.4. Analysis of TG 18:1_18:1_18:1;OOH Isomers in Fresh, Thermally Oxidized, and Photo-Oxidized EVOO

TG 18:1_18:1_18:1;OOH isomers were analyzed to determine the oxidation mechanisms of TG 18:1_18:1_18:1 in EVOO (Figure 4A). TG 18:1_18:1_18:1;OOH concentrations in fresh EVOO were as follows: TG 18:1_18:1_18:1;8OOH (3.1 ± 0.6 µM), TG 18:1_18:1_18:1;9OOH (16.5 ± 3.1 µM), TG 18:1_18:1_18:1;10OOH (19.7 ± 3.4 µM), and TG 18:1_18:1_18:1;11OOH (3.1 ± 0.6 µM) (Figure 4B). Concentrations of TG 18:1_18:1_18:1;9OOH and TG 18:1_18:1_18:1;10OOH were notably higher than those of other isomers, suggesting that TG 18:1_18:1_18:1 in fresh EVOO was oxidized mainly by O₂ oxidation rather than radical oxidation. These results agreed with the well-known fact that FA 18:1 is tolerant to radical oxidation. Regarding unoxidized TG, the concentration of TG 18:1_18:1_18:1 (567 ± 10 mM) was 2.5-times higher than that of TG 18:1_18:1_18:2 (213 ± 3 mM). However, the concentration of TG 18:1_18:1_18:1;OOH was significantly lower than that of TG 18:1_18:1_18:2;OOH.
This suggested that the radical oxidation of TG 18:1_18:1_18:2 was the main factor that contributed to the oxidation of the fresh EVOO analyzed in this study.

**Figure 3.** Q1 mass (A) and product ion mass (B) spectra of prepared TG 18:1_18:1_18:1;OOH isomers. A mixture of TG 18:1_18:1_18:1;OOH isomers (0.5 µM in methanol) was directly infused to the MS/MS system. The ion m/z 940 [M+Na]^+ was used as the precursor ion. Proposed fragmentation patterns of TG 18:1_18:1_18:1;OOH isomers (C). LC-MS/MS chromatograms of TG 18:1_18:1_18:1;OOH isomers (D). A mixture of TG 18:1_18:1_18:1;OOH isomers (0.29 pmol each) were analyzed. Refer to Table 1 for peak numbers. Calibration curves of reference TG 18:1_18:1_18:1;OOH isomers (E). Different amounts of TG 18:1_18:1_18:1;OOH isomers (0.015–0.29 pmol) were analyzed by optimized LC-MS/MS. Mean ± SD (n = 3).
Figure 4. MRM chromatograms of TG 18:1_18:1_18:1;OOH isomers in fresh EVOO (A). Refer to Table 1 for peak numbers. Concentration of TG 18:1_18:1_18:1;OOH isomers in fresh EVOO (B). MRM chromatograms (C) and concentration (D) of TG 18:1_18:1_18:1;OOH isomers in thermal-oxidized EVOO. MRM chromatograms (E) and concentration (F) of TG 18:1_18:1_18:1;OOH isomers in photo-oxidized EVOO. Mean ± SD (n = 3).

Subsequently, TG 18:1_18:1_18:1;OOH isomers in heated EVOO (150 °C) were analyzed. A typical chromatogram of EVOO, thermally oxidized for 240 min, is shown in Figure 4C. Despite the tolerance of FA 18:1 to radical oxidation, all TG 18:1_18:1_18:1;OOH isomers increased with heating time. Isomer compositions followed the radical oxidation pathway of FA 18:1 (Figure 4D); concentrations of each TG 18:1_18:1_18:1;OOH isomer equally increased, as opposed to the case of TG 18:1_18:1_18:2;OOH. This result was consistent with a previous study on the radical oxidation of FA 18:1(9Z);1OMe [15–17].

In photo-oxidized EVOO (Figure 4E), the concentration of TG 18:1_18:1_18:1;OOH isomers specific to $^1$O$_2$ oxidation (i.e., TG 18:1_18:1_18:1;9OOH and TG 18:1_18:1_18:1;10OOH) increased with irradiation time: TG 18:1_18:1_18:1;9OOH (16.5 ± 3.1 μM (0 min) → 48.3 ± 1.8 μM (240 min)) and TG 18:1_18:1_18:1;10OOH (19.7 ± 3.4 μM (0 min) → 52.9 ± 1.5 μM (240 min)) (Figure 4F). These increments were five-times higher than those of TG 18:1_18:1_18:2;10OOH and TG 18:1_18:1_18:2;12OOH (Figure 2F). Considering that the concentration of TG 18:1_18:1_18:1 (567 ± 10 mM) was only 2.5-times higher than that of TG 18:1_18:1_18:2 (213 ± 3 mM), this suggests that the oxidation of FA 18:1(9Z) contributes to the photo-oxidation of EVOO. However, $^1$O$_2$ oxidation of TG 18:1_18:1_18:2 also affords endoperoxyl-
hydroperoxides [16,35] and TG 18:1_18:1_18:2;OOH that were not analyzed in this study. Hence, the analysis of such hydroperoxides should provide further insights into the 1O2 oxidation of EVOO.

In summary, TG 18:1_18:1_18:2;OOH and TG 18:1_18:1_18:1;OOH isomers were directly analyzed from fresh, thermally oxidized, and photo-oxidized EVOO. To the best of our knowledge, this is the first study reporting analysis of TG 18:1_18:1_18:1;OOH isomers. The obtained results, in most cases, agreed with the previously reported characteristics of the radical and 1O2 oxidation of FA 18:2(9Z,12Z) and FA 18:1(9Z). Hence, the LC-MS/MS methods reported herein were advantageous in determining oxidation mechanisms of EVOO that possess a complex matrix (i.e., the presence of chlorophyll, tocopherol, various polyphenols, and carotenoids). The methods described in this paper should also be valuable in understanding how different factors that determine the quality of EVOO (e.g., olive species, cultivation area, cultivation timing, and extraction methods) contribute to its oxidative stability.

Supplementary Materials: The following are available online at https://www.mdpi.com/article/10.3390/molecules27165282/s1, Supplementary Materials: MS and MS/MS parameters for the analysis of TG 18:1_18:1_18:1;OOH and TG 18:1_18:1_18:2;OOH isomers.

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Sample Availability: Samples of the compounds are available from the authors.

Abbreviations

MeO-AMVN 2,2′-azobis-(4-methoxy-2,4-dimethylvaleronitrile)
CID collision induced dissociation
ESI electrospray ionization
EVOO extra virgin olive oil
LED light-emitting diode
LOOH lipid hydroperoxide
MRM multiple reaction monitoring
MxP 2-methoxypropene
1O2 singlet oxygen
TG triacylglycerol
TGOOH triacylglycerol hydroperoxide

References
1. Kiritsakis, A.; Dugan, L.R. Studies in photooxidation of olive oil. J. Am. Oil Chem. Soc. 1985, 62, 892–896. [CrossRef]
2. Psomiadou, E.; Tsimidou, M. Simultaneous HPLC determination of tocopherols, carotenoids, and chlorophylls for monitoring their effect on virgin olive oil oxidation. J. Agric. Food Chem. 1998, 46, 5132–5138. [CrossRef]
3. Aparicio, R.; Roda, L.; Albi, M.A.; Gutiérrez, F. Effect of various compounds on virgin olive oil stability measured by rancimat. J. Agric. Food Chem. 1999, 47, 4150–4155. [CrossRef] [PubMed]
4. Morales, M.T.; Przybylski, R. Olive Oil Oxidation. In Handbook of Olive Oil; Harwood, J., Aparicio, R., Eds.; Springer: Boston, MA, USA, 2000; Volume 13, pp. 459–490. [CrossRef]

5. Psomiadou, E.; Tsimidou, M. Pigments in Greek virgin olive oils: Occurrence and levels. J. Sci. Food Agric. 2001, 81, 640–647. [CrossRef]

6. Gimeno, E.; Castellote, A.I.; Lamuela-Raventós, R.M.; De la Torre, M.C.; López-Sabater, M.C. The effects of harvest and extraction methods on the antioxidant content (phenolics, α-tocopherol, and β-carotene) in virgin olive oil. Food Chem. 2002, 78, 207–211. [CrossRef]

7. Psomiadou, E.; Tsimidou, M. Stability of virgin olive oil 1. Autoxidation studies. J. Agric. Food Chem. 2002, 50, 716–721. [CrossRef]

8. Psomiadou, E.; Tsimidou, M. Stability of virgin olive oil 2. Photo-oxidation studies. J. Agric. Food Chem. 2002, 50, 722–727. [CrossRef] [PubMed]

9. Hrncirik, K.; Fritsche, S. Relation between the endogenous antioxidant system and the quality of extra virgin olive oil under accelerated storage conditions. J. Agric. Food Chem. 2005, 53, 2103–2110. [CrossRef]

10. Tena, N.; García-González, D.L.; Aparicio, R. Evaluation of virgin olive oil thermal deterioration by fluorescence spectroscopy. J. Agric. Food Chem. 2009, 57, 10505–10511. [CrossRef]

11. Blasi, F.; Rocchetti, G.; Montesano, D.; Lucini, L.; Chiiodelli, G.; Ghisoni, S.; Baccolo, G.; Simonetti, M.S.; Cossignani, L. Changes in extra-virgin olive oil added with Lycium barbarum L. carotenoids during frying: Chemical analysis and metabolomic approach. Food Res. Int. 2018, 105, 507–516. [CrossRef]

12. Kishimoto, N. Influence of exposure to sunlight on the oxidative deterioration of extra virgin olive oil during storage in glass bottles. Food Sci. Technol. Res. 2019, 25, 539–544. [CrossRef]

13. Esposto, S.; Taticchi, A.; Servili, M.; Urbani, S.; Sordini, B.; Veneziani, G.; Daidone, L.; Selvaggini, R. Overall quality evolution of extra virgin olive oil exposed to light for 10 months in different containers. Food Chem. 2021, 351, 129297. [CrossRef]

14. Gómez-Alonso, S.; Salvador, M.D.; Fregapane, G. Evolution of the oxidation process in olive oil triacylglycerol under accelerated storage conditions (40–60 °C). J. Am. Oil Chem. Soc. 2004, 81, 177–184. [CrossRef]

15. Franken, E.N. Lipid oxidation. Prog. Lipid Res. 1980, 19, 1–22. [CrossRef]

16. Franken, E.N. Chemistry of free radical and singlet oxidation of lipids. Prog. Lipid Res. 1985, 23, 197–221. [CrossRef]

17. Choe, E.; Min, D.B. Mechanisms and factors for edible oil oxidation. Compr. Rev. Food Sci. Food Saf. 2006, 5, 169–186. [CrossRef]

18. Gollmer, A.; Regensburger, J.; Maisch, T.; Bäumler, W. Luminescence spectroscopy of singlet oxygen enables monitoring of oxygen consumption in biological systems consisting of fatty acids. Phys. Chem. Chem. Phys. 2013, 15, 11386–11393. [CrossRef]

19. Endo, Y.; Usuki, R.; Kaneda, T. Prooxidant activities of chlorophylls and their decomposition products on the photooxidation of methyl linoleate. J. Am. Oil Chem. Soc. 1984, 61, 781–784. [CrossRef]

20. Endo, Y.; Usuki, R.; Kaneda, T. Antioxidant effects of chlorophylls and phycophytin on the autoxidation of oils in the dark. J. Sci. Food Agric. 1985, 62, 1375–1378. [CrossRef]

21. Tanno, R.; Kato, S.; Shimizu, N.; Ito, J.; Sato, S.; Ogura, Y.; Sakaino, M.; Sano, T.; Etsuoka, T.; Kuwahara, S.; et al. Analysis of oxidation products of α-tocopherol in extra virgin olive oil using liquid chromatography-tandem mass spectrometry. Food Chem. 2019, 306, 125582. [CrossRef]

22. Yang, W.T.; Lee, J.H.; Min, D.B. Quenching mechanisms and kinetics of α-tocopherol and β-carotene on the photosensitizing effect of synthetic food colorant FD&C Red No. 3. J. Food Sci. 2006, 67, 507–510. [CrossRef]

23. Hui, S.-P.; Sakurai, T.; Takeda, S.; Jin, S.; Fuda, H.; Kurosawa, T.; Chiba, H. Analysis of triacylglycerol hydroperoxides in human lipoproteins by Orbitrap mass spectrometer. J. Agric. Food Chem. 2019, 67, 10174–10184. [CrossRef] [PubMed]

24. Gómez-Alonso, S.; Fregapane, G. Evolution of the oxidation process in olive oil triacylglycerol under accelerated storage conditions (40–60 °C). J. Am. Oil Chem. Soc. 2004, 81, 177–184. [CrossRef]

25. Giuliani, F.; Destaillats, F.; Skibsted, L.H.; Dionisi, F. Structural analysis of hydroperoxy- and epoxy-triacylglycerols by liquid chromatography mass spectrometry. Chem. Phys. Lipids 2004, 131, 41–49. [CrossRef]

26. Zeb, A.; Murkovic, M. Carotenoids and triacylglycerols interactions during thermal oxidation of refined olive oil. Food Chem. 2011, 127, 1584–1593. [CrossRef]

27. Zeb, A. Triacylglycerols composition, oxidation and oxidation compounds in camellia oil using liquid chromatography-mass spectrometry. Chem. Phys. Lipids 2012, 165, 608–614. [CrossRef]

28. Kato, S.; Nakagawa, K.; Suzuki, Y.; Asai, A.; Nagao, M.; Nagashima, K.; Oikawa, S.; Miyazawa, T. Liquid chromatography-tandem mass spectrometry determination of human plasma 1-palmitoyl-2-hydroperoxooyctadecadienoyl-phosphatidylcholine isomers via promotion of sodium adduct formation. Anal. Biochem. 2015, 471, 51–60. [CrossRef] [PubMed]

29. Kato, S.; Shimizu, N.; Hanzawa, Y.; Otoki, Y.; Ito, J.; Kimura, F.; Takekoshi, S.; Sakaino, M.; Sano, T.; Etsuoka, T.; et al. Determination of triacylglycerol oxidation mechanisms in canola oil using liquid chromatography-tandem mass spectrometry. nplj Sci. Food Chem. 2018, 2, 11–11. [CrossRef]

30. Liebsch, G.; Fahy, E.; Aoki, J.; Dennis, E.A.; Durand, T.; Ejsing, C.S.; Fedorova, M.; Feusner, I.; Griffiths, W.J.; Köfeler, H.; et al. Update on LIPID MAPS classification, nomenclature, and shorthand notation for MS-derived lipid structures. J. Lipid Res. 2020, 61, 1539–1555. [CrossRef]

31. Kato, S.; Nakagawa, K.; Suzuki, Y.; Suzuki, K.; Mizuochi, S.; Miyazawa, T. Preparation of 13 or 9-hydroperoxy-9Z,11E (9E,11E) or 10E,12Z (10E,12E)-octadecadienoic phosphatidylcholine hydroperoxide. J. Oleo Sci. 2014, 63, 431–437. [CrossRef]
32. Bhosle, B.M.; Subramanian, R. New approaches in deacidification of edible oils-a review. *J. Food Eng.* 2005, 69, 481–494. [CrossRef]

33. Milne, G.L.; Seal, J.R.; Havrilla, C.M.; Wijtmans, M.; Porter, N.A. Identification and analysis of products formed from phospholipids in the free radical oxidation of human low density lipoproteins. *J. Lipid Res.* 2005, 46, 307–319. [CrossRef]

34. Kato, S.; Shimizu, N.; Ogura, Y.; Otoki, Y.; Ito, J.; Sakaino, M.; Sano, T.; Kuwahara, S.; Takekoshi, S.; Imag, J.; et al. Structural analysis of lipid hydroperoxides using mass spectrometry with alkali metals. *J. Am. Soc. Mass Spectrom.* 2021, 32, 2399–2409. [CrossRef] [PubMed]

35. Frankel, E.N. Secondary products of lipid oxidation. *Chem. Phys. Lipids* 1987, 44, 73–85. [CrossRef]