A novel UGGT1 and p97-dependent checkpoint for native ectodomains with ionizable intramembrane residue

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ABSTRACT Only native polypeptides are released from the endoplasmic reticulum (ER) to be transported at the site of activity. Persistently misfolded proteins are retained and eventually selected for ER-associated degradation (ERAD). The paradox of a structure-based protein quality control is that functional polypeptides may be destroyed if they are architecturally unfit. This has health-threatening implications, as shown by the numerous “loss-of-function” proteopathies, but also offers chances to intervene pharmacologically to promote bypassing of the quality control inspection and export of the mutant, yet functional protein. Here we challenged the ER of human cells with four modular glycopolypeptides designed to alert luminal and membrane protein quality checkpoints. Our analysis reveals the unexpected collaboration of the cytosolic AAA-ATPase p97 and the luminal quality control factor UDP-glucose:glycoprotein glucosyltransferase (UGGT1) in a novel, BiP- and CNX-independent checkpoint. This prevents Golgi transport of a chimera with a native ectodomain that passes the luminal quality control scrutiny but displays an intramembrane defect. Given that human proteopathies may result from impaired transport of functional polypeptides with minor structural defects, identification of quality checkpoints and treatments to bypass them as shown here upon silencing or pharmacologic inhibition of UGGT1 or p97 may have important clinical implications.

INTRODUCTION The protein quality control machineries operating in the mammalian endoplasmic reticulum (ER) lumen examine soluble proteins or the ectodomain of membrane polypeptides in search of nonnative determinants such as hydrophobic patches, unpaired cysteine residues, and nonnative peptidyl-prolyl bonds. Most polypeptides entering the ER are modified with core oligosaccharides (glucose-α1,2-mannose-N-acetylg glucosamine2-), which are covalently bound to asparagine side chains (N-linked glycosylation). Their processing is a crucial part of protein quality control. Cycles of deglucosylation/reglucosylation catalyzed by glucosidase II and UDP-glucose:glycoprotein glucosyltransferase (UGGT1) determine retention in the calnexin (CNX) chaperone system and promote formation of native disulfide and peptidyl-prolyl bonds (Caramelo and Parodi, 2008). Complete deglucosylation allows secretion from the ER and transport at the site of activity. Extensive demannosylation catalyzed by members of the glycosyl hydrolase 47 family of α1,2-mannosidases (Olivari and Molinari, 2007) triggers selection for dislocation into the cytosol and proteasome-mediated clearance (ER-associated degradation [ERAD]; Brodsky, 2012; Braakman and Hebert, 2013; Merulla et al., 2013).

Structural defects that may prevent polypeptide transport through the secretory pathway include potentially charged (ionizable) residues in the intramembrane domain of membrane-anchored...
polypeptides (Klausner et al., 1990). Such residues (aspartic and glutamic acids, lysines, and arginines) lower the hydrophilicity of transmembrane (TM) regions (Hessa et al., 2007) and play crucial roles in biogenesis of several oligomeric signaling complexes by ensuring retention of orphan subunits to complete complex assembly and degradation of subunits produced in nonstoichiometric amount (Klausner et al., 1990; Call and Wucherpfennig, 2005; Feng et al., 2006; Tyler et al., 2012). Mutations that introduce ionizable residues in the intramembrane domain have been linked to loss-of-function diseases such as idiopathic epilepsy (Cossette et al., 2002; Gallagher et al., 2005) or Ehlers–Danlos syndrome (Bin et al., 2014), in which an alanine- or glycine-to-aspartic acid mutation in the TM domain inappropriately tags for degradation functional γ-aminobutyric acid type A (GABA<sub>A</sub>) receptors or ZIP13 proteins, respectively. Idiopathic epilepsy and Ehlers–Danlos syndrome are only two of the hundreds of loss-of-function diseases in which the quality control operating in the secretory pathway, which is based on detection of structural defects, inappropriately prevents transport of functional polypeptides at their final destination. In such cases, the therapeutic interventions that enhance folding, stabilize the mutated polypeptide, or promote bypassing of the transport block are highly sought after (Powers et al., 2009; Guerriero and Brodsky, 2012; Noack et al., 2014).

To understand in more detail the mechanisms regulating protein quality control in the secretory pathway, to understand the consequences of the presence of intramembrane ionizable residues, and to evaluate pharmacologic interventions that promote secretion of inappropriately retained proteins, we monitored the fate of model type I membrane proteins for which a folding-defective (null Hong Kong variant [NHK] of human α1-antitrypsin [α1AT]) or a folding-competent ectodomain (α1ATc) were fused to the C-terminal domain of CD36. This domain contains a naturally occurring integration-competent intramembrane region with an ionizable aspartic acid residue at position 6 (model chimeras NHKc and α1ATc), which was replaced by an uncharged alanine residue in the NHKc<sub>D6A</sub> and α1ATc<sub>D6A</sub> chimeras (Figure 1A). Because ionizable residues in single-pass membrane-anchored proteins increase the free energy of membrane integration (∆G<sub>TM</sub>) to levels that may prevent insertion of newly synthesized polypeptides in the lipid bilayer (Shin et al., 1993; Fayadat and Kopito, 2003; Hessa et al., 2007; Feige and Hendershot, 2013), we first controlled membrane association of the four chimeras used in this study. To this end, we performed subcellular fractionation to separate cytosolic (Supplemental Figure S1A, lanes 1 and 2, p97 as marker), luminal (lanes 3 and 4, calreticulin [CRT]), and membrane-associated proteins (lanes 5 and 6, thioredoxin-related transmembrane protein 1 [TMX1]). Efficient membrane integration was confirmed (Figure 1, B–E, lane 5), and only a negligible fraction of the four chimeras was detected in the cytosolic (lane 1) or luminal fractions (lane 3), unless cells were exposed to the proteasome inhibitor PS341 (lanes 2, 4, and 6; see later discussion). This was in good agreement with the calculated ∆G<sub>TM</sub> (http://dgpred.cbr.su.se), which predicts efficient membrane integration for the ionizable CD36 TM, which anchors NHKc and α1ATc at the membrane (∆G<sub>TM</sub> = −0.76), and for the nonionizable CD36 TM, which anchors NHKc<sub>D6A</sub> and α1ATc<sub>D6A</sub> (∆G<sub>TM</sub> = −2.26).

An ionizable residue in the TM domain determines higher extractability from the lipid bilayer and faster disposal of a folding-defective ectodomain

NHKc is a soluble ERAD substrate, which is dislocated across the ER membrane and degraded by cytosolic proteasomes (Liu et al., 1997). Addition of a membrane anchor with or without an ionizable residue in the TM domain (NHKc and NHKc<sub>D6A</sub>, respectively; Figure 1A) did not change the fate of the folding-defective ectodomain. In fact, both chimeras were efficiently cleared from the ER (Figure 2, A and B, lanes 1–5, and C). To study this, HEK293 cells expressing NHKc or NHKc<sub>D6A</sub> were pulsed with <sup>35</sup>S)methionine and cysteine and chased for up to 120 min in the absence of radioactivity. The decay of the immunolabeled chimeras was quantified after their separation in SDS–PAGE (Figure 2, A–C). NHKc had a
half-life of ∼60 min (Figure 2, A, lanes 1–5, and C), which was substantially prolonged by the proteasomal inhibitor PS341, as expected for ERAD substrates (Figure 2A, lane 6). Replacement of the intramembrane ionizable aspartic acid with an alanine delayed the decay of NHKcD6A (half-life of ∼90 min; Figure 2, B, lanes 1–5, and C). Consistent with their retention in the ER during preparation for ERAD, both NHKc and NHKcD6A displayed endoglycosidase H (EndoH)–sensitive oligosaccharides (Figure 2D) and colocalized with the ER marker CNX in immunofluorescence (IF; Figure 2E).

Analysis of the subcellular compartmentalization of the misfolded proteins upon inactivation of the proteasomes revealed the substantial accumulation of partially and fully deglycosylated NHKc in the cell cytosol (Figure 1B, lane 2, and Supplemental Figure S1B) to a much larger extent that what was observed for NHKcD6A (Figure 1C, lane 2, and Supplemental Figure S1B). This is consistent with an enhanced extractability from the lipid bilayer of the folding-defective chimera displaying an ionizable residue in the intramembrane space.

**An ionizable residue in the TM domain prevents Golgi transport of the folding-competent α1AT ectodomain**

The foregoing pulse-chase analysis was also used to determine the fate of the chimeras displaying the folding-competent α1AT ectodomain tethered at the ER membrane with an anchor containing (α1ATc) or not containing (α1ATcD6A) the ionizable aspartic acid residue. Analysis of radiolabeled α1ATc revealed a major polypeptide band (l for immature; Figure 3A, lanes 1–5) and a very faint polypeptide with slower electrophoretic mobility appearing at longer chase times (M for mature; Figure 3A, lanes 4 and 5). In the case of α1ATcD6A, the fast-migrating polypeptide shown with I was efficiently converted, starting at 30 min of chase, into the slowly migrating M band (Figure 3B, lanes 2–5). Reduction in a glycoprotein’s electrophoretic mobility occurs upon release of native proteins from the ER and arrival in the Golgi compartment, where N-linked oligosaccharides are modified to become much larger and resistant to EndoH cleavage (Rothman *et al*., 1984). An EndoH assay confirmed that α1ATc oligosaccharides remained EndoH sensitive throughout the chase (Figure 3, C and E), an indication that this protein was not delivered to the Golgi compartment. In contrast, α1ATcD6A oligosaccharides became resistant to EndoH cleavage (Figure 3, D and E), implying that this chimera was efficiently transported to the Golgi. The different intracellular fate of these two chimeras was confirmed in IF. Both proteins colocalized poorly with the ER marker CNX (Figure 3, F and I) and, more significantly, especially α1ATcD6A, with intermediate compartment/Golgi markers such as βCOP (Figure 3, G and J). In agreement with the biochemical analysis showing that only α1ATcD6A displays EndoH-resistant oligosaccharides (Figure 3, D and E), this chimera showed a substantial colocalization with the Golgi marker giantin (Figure 3K), whereas α1ATc was virtually excluded from the Golgi (Figure 3H).
Chimeras with folding-defective ectodomains are persistently reglucosylated and show prolonged association with the ER retention-factors CNX and BiP

Before selection for ERAD, folding-defective polypeptides are retained in the ER upon association with CNX and BiP (Molinari et al., 2002). Here the quality control factor UGGT1 plays a crucial role by persistently reglucosylating the oligosaccharides displayed on misfolded conformers, thereby preventing their release from the CNX chaperone system (Caramelo and Parodi, 2008). Persistent protein glucosylation is assessed by a jack bean α-mannosidase (JBαM) assay. In this assay, JBαM removes eight mannoses from deglucosylated oligosaccharides (Figure 4A, left) but only five from oligosaccharides where branch A is protected by a terminal glucose residue (Figure 4A, middle). This allows distinction of polypeptides displaying nonglucosylated oligosaccharides, for which the mobility shift upon JBαM treatment is similar to the shift caused by EndoH treatment (Figure 4A, right), from polypeptides displaying glucosylated oligosaccharides, which are characterized by a smaller shift in electrophoretic mobility upon JBαM treatment (Hebert et al., 1995).

JBαM treatment performed on NHKc immediately after synthesis (10-min chase, Figure 4B, lanes 1 and 2) or 120 min after synthesis (lanes 3 and 4) caused a smaller shift in electrophoretic mobility than EndoH treatment (lanes 5 and 6). Hence, throughout the chase, NHKc mannoses were protected from complete removal by JBαM. This shows that NHKc is persistently reglucosylated by the UGGT1 or fails to be efficiently deglucosylated by the glucosidase II.
Pharmacologic inactivation of p97 promotes α1ATc transport to the Golgi

To characterize the protein quality checkpoint preventing Golgi transport of α1ATc, we exposed cells for 4 h to low temperature (30°C), chemical chaperones (glycerol), or proteasomal (PS341) or p97 inhibitors (N²,N²-dibenzylquinazoline-2,4-diamine [DBeQ]), that is, to treatments that bypass retention-based quality control of disease-related polypeptides such as the cystic fibrosis conductance regulator, dystrophin, GABA receptor, ZIP13, and other mutant polypeptides, thereby alleviating disease phenotypes (Denning et al., 1992; Brown et al., 1996; Bonuccelli et al., 2003; Chou et al., 2011; Bin et al., 2014; Han et al., 2015). Only cell treatment with DBeQ substantially enhanced α1ATc secretion to the Golgi, as shown by the appearance of the mature (M in Figure 5, A, lane 6, and B, lane 2 and arrow in lane 5), EndoH-resistant form of α1ATc (Figure 5B, lane 6). IF analysis confirmed that in cells exposed to DBeQ, α1ATc escapes retention-based quality control (Figure 5C) and travels to the Golgi compartment (Figure 5D and quantification). It is worth mentioning that α1ATc retention was not affected by the inactivation of access to the CNX chaperone system with castanospermine (CST; Figure 5A, lane 4), a specific inhibitor of glucosidase I and glucosidase II (Hammond et al., 1994).

To evaluate the selectivity of DBeQ, we compared the fate of NHKcD6A and NHKc (subjected to the conventional ER quality control), α1ATcD6A (efficiently exported to the Golgi), and α1ATc (halted by the DBeQ-sensitive checkpoint) in mock-, PS341-, or...
DBeQ-treated cells. The chimeras were immunosolated from cell lysates to evaluate changes in the association with the ER-retention factors CNX and BiP (Molinari et al., 2002; Figure 5E, top and middle), as well as variations in their electrophoretic mobility as a straightforward readout for glycoprotein release to the Golgi compartment (Figure 5E, bottom).

The newly synthesized, radiolabeled chimeras with the misfolded ectodomain show persistent association with CNX and BiP (Figure 4, F and G). Consistently, at steady state, NHKcD6A and NHKc showed abundant association with the ER-retention factors (Figure 5E, top and middle, lanes 1–6), whereas α1ATcD6A and α1ATc associated poorly, if at all (lanes 7–12). PS341 and DBeQ had little effect on the coprecipitation of CNX and BiP with NHKcD6A and NHKc (Figure 5E, top and middle, lane 1 vs. lanes 2 and 3, and lane 4 vs. lanes 5 and 6). The unchanged electrophoretic mobility of the two chimeras with the folding-defective ectodomain confirmed that DBeQ did not impair the conventional CNX/BiP-mediated quality control that retains them in the ER (Figure 5E, bottom, lane 1 vs. lanes 2 and 3, and lane 4 vs. lanes 5 and 6). DBeQ did not affect transport to the Golgi of α1ATcD6A, as shown by the unchanged attainment of the mature form (Figure 5, A–D and E, bottom, arrow in lane 12). Thus, pharmacologic inactivation of p97 selectively inactivates the checkpoint alerted by the chimera with the native ectodomain tethered at the membrane with an anchor containing an ionizable residue. Of interest, a similar alanine-to–aspartic acid mutation in the TM domain of the GABA receptor is linked to defective surface transport of the mutant protein causing idiopathic epilepsy (Cossette et al., 2002; Gallagher et al., 2005). The disease...
phenotype is alleviated by pharmacologic inactivation or silencing of p97. The recover in mutant GABA_α receptor secretion has been attributed to the inhibition of the p97 function in ERAD (Han et al., 2015). However, as in the case of α1ATc (Figures 5, A, lane 4, and E, lane 11), proteasome inhibition does not restore secretion of the mutant GABA_α receptor. This leads us to postulate that inhibition of a role of p97 in quality control, rather than in ERAD, may play a role in recovering from loss-of-function diseases as well.

**Pharmacologic inactivation of p97 restores transport to the Golgi of α1ATc by promoting α1ATc dissociation from p97 and UGGT1**

To confirm the involvement of the DBeQ target (i.e., p97) and verify possible involvement in the checkpoint preventing α1ATc transport to the Golgi of the quality control factor UGGT1, which cycles in post-ER boundaries (Zuber et al., 2001; Mezzacasa and Helenius, 2002; Gilchrist et al., 2006) and participates in p97-containing complexes (Haines et al., 2012), we analyzed NHKc, α1ATc, and α1ATc_Δ\(_{D6A}\) associations with p97 and UGGT1 in the presence of the chemical cross-linker dithiobis succinimidyl propionate (DSP) to stabilize weak interactions. This test showed that NHKc and α1ATc (Figure 5F, lanes 1–4), but not α1ATc_Δ\(_{D6A}\) (lanes 5 and 6), entered complexes with both quality control factors (Figure 5F). Cell incubation with DBeQ did not significantly affect the association of p97 and UGGT1 with the ERAD substrate NHKc (lane 1 vs. lane 2), but it substantially decreased the coprecipitation of p97 and UGGT1 with α1ATc (lane 3 vs. lane 4). We postulate that in the case of NHK, UGGT1 regulates the reglucosylation that prolongs CNX association, and p97 intervenes in membrane extraction of this ERAD substrate from the ER. In the case of α1ATc, which is rapidly released from CNX and is not degraded, the two quality control factors might participate in a novel functional retention complex preventing α1ATc secretion. Note, p97 and UGGT1 have been identified as major interactors of the UBXD1 adaptor protein (Haines et al., 2012) as an indication that they might coexist in functional complexes. Even though we did not detect UBXD1 among the α1ATc interactors (unpublished data), the immunoprecipitation of endogenous UGGT1 revealed the coprecipitation of p97 (Figure 5G). DBeQ did not disassemble the UGGT1:p97 complex, as shown by the unchanged presence of p97 in immunocomplexes isolated with UGGT1 as bait (Figure 5G, middle, lane 2 vs. lane 3). Instead, the pharmacologic inactivation of p97 caused α1ATc release from the two retention factors (Figure 5G, bottom, lane 2 vs. lane 3), thus restoring transport of this chimera to the Golgi.

**p97 and UGGT1 silencing selectively restores transport to the Golgi of α1ATc**

Silencing the expression of a series of early secretory pathway proteins involved in quality control, in ERAD, or in vesicular transport revealed that only p97 and UGGT1 silencing correlated with the complex modification of oligosaccharides, showing transport of α1ATc to the Golgi compartment (Figure 5H, lanes 2 and 12, and Supplemental Figure S3). Silencing of p97 and of UGGT1 did not affect the ER retention of the chimeras displaying the misfolded ectodomain (Figure 5I). For p97, this is consistent with the fact that pharmacologic p97 inactivation did not interfere with conventional, retention-based ER quality control (Figure 5E, bottom, lanes 3 and 6, showing the lack of mature NHKc_Δ\(_{D6A}\) and NHKc forms in cells treated with DBeQ).

**DISCUSSION**

Human proteopathies may result from inappropriate elimination of functional polypeptides displaying minor structural defects or delayed attainment of the native structure as a consequence of gene mutations (Lindquist and Kelly, 2011). Thus, the identification and comprehension at the molecular level of the quality checkpoints surveying production of the human proteome is instrumental for designing interventions that counteract their progression. Our data reveal a novel druggable quality checkpoint alerted when the polypeptide’s ectodomain fulfills quality control requirements for ER exit but displays other structural defects (in the chimera analyzed in this study, an intramembrane ionizable residue). By showing that cell exposure to DBeQ and silencing of p97 restore transport to the Golgi of α1ATc, our study identifies p97 as a pharmacologic target to treat loss-of-function proteopathies and underscores the importance of developing function-specific inhibitors of p97 (Chou and Deshaies, 2011).

The cytosolic AAA ATPase p97 is a multifunctional protein (Yamanaka et al., 2012). It participates in a growing number of poorly characterized supramolecular complexes, some of which are involved in ERAD, whereas others contain quality control and vesicular transport factors. The data presented here led us to exclude that transport to the Golgi of α1ATc is restored upon inactivation of the function of p97 in ERAD. In fact, proteasome inactivation does not lead to α1ATc secretion. Instead, it must be ascribed to a role of p97 in regulating retention of proteins displaying intramembrane defects.

That silencing of p97 would restore α1ATc transport to the Golgi was expected because pharmacologic inactivation of p97 does. In contrast, that also UGGT1 silencing would restore α1ATc transport was at least partially surprising. In fact, α1ATc displays a native ectodomain whose oligosaccharides rapidly lose their terminal glucoses, is rapidly released from CNX, and is therefore most likely not a target of the enzymatic activity of UGGT1. The unperturbed retention of α1ATc in cells treated with CST (Figure 5A) confirms that generation of the monoglycosylated form of the α1ATc oligosaccharides by the glucosidase I and glucosidase II and/or the persistence of monoglycosylated forms engaging the ER-resident lectin chaperones upon UGGT1 activity is dispensable for the tightness of this novel quality control checkpoint. Nevertheless, UGGT1 cycles in post-ER boundaries (Zuber et al., 2001; Mezzacasa and Helenius, 2002; Gilchrist et al., 2006), might participate in p97-containing supramolecular complexes (Haines et al., 2012), and, as our data imply, participate in DBeQ-sensitive complexes that control α1ATc retention. Intervention of p97 and of the UGGT1 in the protein quality checkpoint described in this work seems to elucidate their conventional functions of major components of the ERAD machinery and the CNX cycle, respectively.

The UGGT1/p97 checkpoint complex must contain additional components and, possibly, one or more membrane-spanning proteins that bridge the cytosolic p97 and the luminal UGGT1 (Figure 6). Like p97, the multispanning protein BAP31 is involved in ERAD (Wang et al., 2008) but also in vesicular transport in the early secretory pathway (Bell et al., 2001; Wakana et al., 2008). Significantly, BAP31 shows preferential action on proteins characterized by the presence of ionizable residues in the TM domain (Geiger et al., 2011). As such, BAP31 was a likely candidate to play a role in this novel, DBeQ-sensitive protein quality checkpoint that prevents α1ATc transport to the Golgi. However, BAP31 silencing did not restore transport to the Golgi of α1ATc (Figure 5H, lane 8). This shows that the residual amount (Supplemental Figure S3) of this very abundant protein warrants retention of α1ATc or that an elusive, surrogate retention factor can intervene to replace BAP31. Similarly, silencing of several other factors did not reestablish α1ATc transport to the Golgi (Figure 5H). Here as well, these factors are not involved in the DBeQ-sensitive checkpoint, their silencing was not efficient.

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FIGURE 6: The chimeras analyzed in this study and their fate. (A) The chimeras with the misfolded ectodomain (NHKcDM and NHKc) are retained in the ER by the conventional quality control relying on UGGT1, CNX, and BiP intervention. These proteins are eventually destined to ERAD. (B) The chimera with the native ectodomain, characterized by an ionizable residue in the TM (α1ATc), fulfills the quality control requirement for release from CNX and BiP, but its transport to the Golgi is halted upon p97 and UGGT1 intervention. This protein quality checkpoint is bypassed upon pharmacologic inhibition of p97 or silencing of p97 or UGGT1 expression. (C) The folding-competent chimera (α1ATcD6A) is efficiently transported to the Golgi compartment.
enough to reveal a phenotype, or other factors intervene upon their silencing.

Retention-quality control is alerted in the early secretory pathway by structural defects that may not compromise the function of the mutant polypeptide yet can prevent the transport of the mutant polypeptide at the site of activity. Secretion of mutant polypeptides linked to cystic fibrosis, muscular dystrophy, idiopathic epilepsy, and lysosomal storage diseases, to name only a few, can be promoted in cultured cells and/or in the organism by chemical or substrate-specific pharmacologic chaperones (Guerriero and Brodsky, 2012; Noack et al., 2014). The development of treatments that alleviate loss-of-function phenotypes caused by hampered trafficking of functional yet structurally defective polypeptides has important medical implications. It relies on the characterization of the mechanisms that regulate quality control and transport at the intracellular or extracellular site of activity of newly synthesized polypeptides. The characterization of a novel, p97-regulated, DBeQ sensitive quality checkpoint for native ectodomains with intramembrane defects highlights a novel therapeutic avenue to counter the progression of loss-of-function diseases caused by defective transport of functional polypeptides at their final destination.

**MATERIALS AND METHODS**

**Expression plasmids, antibodies, and inhibitors**

The pCIneo plasmids encoding for CD38 was provided by S. Fang (University of Maryland Biotechnology Institute, Baltimore, MD). The membrane-tethered variants of α1ATc (α1ATCD38) and NHKc (NHKCD38) (Bernasconi et al., 2010) were generated by inserting at the C-terminus of α1AT or NHK, both cloned in pcDNA3, residues 97–173 of mouse CD38 (VeldsGMAGFIDILATLALGYYCFA-GHETGRPSGAAEVALLKNEQLY QPLRDRDETYQYSLRLGNWPRNKK, transmembrane domain in italic). Commercially available purification kits (Sigma-Aldrich, Buchs, Switzerland) were used for DNA sample preparations. The uncharged versions of α1ATCD38 and NHKCD38 were produced by replacing the aspartic acid residue (D) of the TM sequence (marked in bold) with alanine (A). An Agilent Technologies (Basel, Switzerland) mutagenesis kit was used to prepare the constructs. All proteins were tagged with hemagglutinin (HA) epitope (YPYDVPDYA), and the nucleotide sequences of plasmids were verified on both strands. Antibodies to HA, TMX1, SEL1L, derlin-1 were obtained from Sigma-Aldrich; antibodies to CNX, GP78, HPRD1, HERP, derlin-2 and UGTT1 were kind gifts of A. Helenius (ETH, Zurich, Switzerland), S. Fang and R. Wojcikiewicz (SUNY- Upstate Medical University, Syracuse, NY), K. Koke (Cerebatal Cardiovascular Center, Osaka, Japan), Y. Ye (National Institutes of Health, Bethesda, MD), and A. Parodi (Fundación Instituto Leloir, Buenos Aires, Argentina), respectively. KDEL (BiP-GRP94) was provided by Stressgen (Lausen, Switzerland), BAP31 by Alexis Biochemicals (Lausen, Switzerland), JcOP by ABR (Lausen, Switzerland), and ubiquitin by DAKO (Baar, Switzerland). P-341 (Millennium Pharmaceuticals, Cambridge, MA), DBeQ, CST, and apyrase (all from Sigma-Aldrich) were used at final concentrations of 10 μM, 10 μM, 1 mM, and 18 U/ml, respectively.

**Cell lines, transient and stable transfections, and RNA interference**

HEK293 cells were grown in DMEM/10% fetal bovine serum (FBS). Transfections were performed with Lipofectamine 2000 (Invitrogen) according to the instructions of the manufacturer. For generation of stable cell lines, cells were selected for 2 wk in DMEM supplemented with 10% FBS and 2% Geneticin. RNA interferences were performed in HEK293 cells plated at 50–60% confluence on polylsine-coated 3.5-cm Petri dishes. Cells were cotransfected with siRNA duplex (50 pmol/dish; Ambion, Lucerne, Switzerland) and the expression plasmid of interest (0.5 μg/dish) using Lipofectamine 2000 (Invitrogen, Lucerne, Switzerland) according to the instructions of the manufacturer. Experiments were performed 48 h after transfection.

**Metabolic labeling, immunoprecipitations, Western blots, and analysis of data**

HEK293 cells transiently transfected (18 h after transfection) and stable cell lines were pulsed with 0.05 mCi of [35S]methionine/ cysteine mix and chased for the indicated time points with DMEM supplemented with 5 mM cold methionine and cysteine. Cell lysates were prepared by solubilization of cells with ice-cold 2% [3-(3-cholamidopropyl)dimethylammonio]-1-propanesulfonate (CHAPS; or RIPA) in 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid–buffered saline (HBS) and 20 mM N-ethylmaleimide (NEM) and then centrifuged 10 min at 10,000 × g. Native immunoprecipitations were performed by adding Protein A beads (Sigma-Aldrich; 1:10 [wt/vol] swollen in HBS) and the selected antibody to cell lysates. Incubations were performed for 2 h at 4°C. The immunoprecipitates were extensively washed (3× 10 min) with 0.5% CHAPS in HBS or 0.5% Triton X-100, resuspended in sample buffer, incubated at 65°C for 10 min, and separated in SDS–PAGE. To assess chimeras association with CNX and BiP, after immunosolubilation of the ER chaperones with the specific antibodies, proteins were denatured with 1% SDS and incubated at 65°C for 10 min, and 1% Triton X-100 was added to the samples. The supernatant obtained after centrifugation (2 min at 10,000 × g) was used for immunoprecipitation with anti-HA. Relevant bands were quantified by Image Quant software (Molecular Dynamics). Gels were also exposed to BioMax (Kodak) films and scanned with an AGFA scanner. For EndoH (New England Biolabs, Alschwil, Switzerland) treatment, immunosolubilated chimeras were incubated for 2 h at 37°C. Samples were then analyzed in reducing SDS–PAGE. Western blots were performed with blocking polyvinylidene difluoride (PVDF) membranes with 10% (wt/vol) milk (Bio-Rad, Cressier, Switzerland) and using primary antibodies at 1:1000 to 1:3000 dilution. Secondary antibodies were horseradish peroxidase conjugated and used at 1:20,000 to 1:40,000 dilution. The Luminata Forte ECL detection system was from Millipore (Schaffhausen, Switzerland), and signal was detected using ImageQuant LAS 4000 (GE Healthcare, Glatbrugg, Switzerland).

**Jack bean α-mannosidase assay**

Protein glucosylation state was monitored by JBeαM assay as described in Hebert et al. (1995).

**Carbonate extraction**

For carbonate extraction, cells were seeded in 6-cm dishes (polylsine-coated) and washed in PBS and PBS-NEM. Cells were then resuspended in 10 mM triethanolamine, 10 mM acetic acid, 250 mM sucrose, 1 mM EDTA, pH 7.4, and 20 mM NEM and were broken by four to six passages through a 1-ml, 25-gauge needle syringe. Nuclei were removed by 2× 1000 × g centrifugation. The supernatant was centrifuged at 200,000 × g in TLA-120.2 rotor for 45 min. The supernatant obtained (cytosol) was recovered, and the pellet fraction was resuspended in 500 μl of 100 mM Na2CO3 and incubated for 25 min on ice. Samples were then centrifuged for 45 min at 200,000 × g. The supernatant (lumen) was harvested and the pellet was rinsed with 500 μl of 100 mM Na2CO3 and subjected to further centrifugation at 200,000 × g for 35 min. The pellet was solubilized in RIPA on ice (20 min), and the membrane-bound
fraction was rescued after further centrifugation for 10 min at 200,000 x g. Normalized quantities of fractions were loaded on SDS–PAGE and transferred on PVDF membranes, and immunoblot was performed as previously described.

**Indirect immunofluorescence microscopy**

Stable HEK293 cells were plated 17 h before fixation on polylysine-coated coverslips. Cells were washed twice with PBS and fixed at room temperature for 20 min with 3.7% formaldehyde diluted in PBS. Cells were washed (3x 5 min) with PBS and incubated in permeabilization solution (0.1% Triton X-100 in PBS) for 15 min to improve antigen accessibility. Washings in PBS (3x 5 min) were performed before 30-min incubation with 10% FBS diluted in PBS (blocking solution). Cells were incubated for at least 1 h, 30 min with primary antibodies diluted 1:50 to 1:100 in blocking solution, washed three times (PBS), and incubated with Alexa Fluor–conjugated secondary antibodies (Invitrogen) diluted 1:200 in blocking solution for at least 45 min. Cells were rinsed with PBS (three for 5 min each) and H2O and mounted with Vectorshield (Vector Laboratories) supplemented with 4’,6-diamidino-2-phenylindole. Images were collected using a laser scanning confocal microscope (Leica DI6000 microscope stand connected to a SP5 scan head).

**Cross-linking assay**

HEK293 cells were seeded in 3.5-cm polylysine-coated dishes 17 h before treatment. Immediately before use, DSP (Sigma-Aldrich) was prepared at 100x final concentration in dimethyl sulfoxide. Cells were washed once with PBS and incubated for 30 min at room temperature in PBS supplemented with 1 mM DSP. The reaction was quenched by incubating cells with PBS and 20 mM Tris, pH 7.8, for 15 min.

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