Research Article

Potential Protective Effects of Bioactive Constituents from Chinese Propolis against Acute Oxidative Stress Induced by Hydrogen Peroxide in Cardiac H9c2 Cells

Liping Sun,1,2 Kai Wang,1,3 Xiang Xu,1,2 Miaomiao Ge,1,2 Yifan Chen,3 and Fuliang Hu3

1 Institute of Apicultural Research, Chinese Academy of Agricultural Sciences, Beijing 100093, China
2 National Research Center of Bee Product Processing, Ministry of Agriculture, Beijing 100093, China
3 College of Animal Sciences, Zhejiang University, Hangzhou 310058, China

Correspondence should be addressed to Liping Sun; caasun@126.com

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Chinese propolis (CP) is known as a health food but its beneficial effects in protecting cardiomyocytes remain elusive. Here, we investigated the effects of CP and its active compounds on hydrogen peroxide (H2O2) induced rats cardiomyocytes (H9c2) oxidative injury. Cell viability decreases induced by H2O2 were mitigated by different CP extracts using various solvents. From these active fractions, six active compounds were separated and identified. Among tested isolated compound, the cytoprotective activities of three caffeates, caffeic acid phenethyl ester (CAPE), benzyl caffeate (BZC), and cinnamyl caffeate (CNC), exerted stronger effects than chrysin, pinobanksin, and 3,4-dimethoxycinnamic acid (DMCA). These three caffeates also increased H9c2 cellular antioxidant potential, decreased intracellular calcium ion ([Ca2+]i) level, and prevented cell apoptosis. Overall, the cardiovascular protective effects of the CP might be attributed to its caffeates constituents (CAPE, BZC, and CNC) and provide evidence for its usage in complementary and alternative medicine.

1. Introduction

Cardiovascular disease (CV) is responsible for 30% of deaths worldwide, surpassed other diseases, and is projected to account for 25 million deaths annually by 2030 [1]. The cost of CV estimated U.S. $863 billion (in 2010) [2]. Myocardial ischemia (MI), commonly known as angina, is one of the major clinical indications of CV and mainly caused by intraluminal coronary thrombosis and ruptured atherosclerotic plaque [3]. Ischemic damages to the cardiac cells are known to be related to reactive oxygen species (ROS) produced during tissue ischemia, which will lead to cardiomyocytes' oxidative stress and further lead to apoptotic cell death [4]. Now one major interesting area is to understand and to prevent cardiac cell death associated with oxidative stress, and several antioxidants have been shown with promising therapeutic effects [5].

Chinese propolis (CP) is an important hive product collected by honeybees (Apis mellifera) from buds of plants [6]. CP is widely used as a natural antioxidant and is as a well-known functional food for its biological activities, like anti-inflammatory, antimicrobial, liver detoxifying, and cardioprotective effects [7, 8]. Moreover, the antioxidant basis of CP might attribute to its abundant polyphenolic compounds, mainly flavonoids, quercetin, chrysin, kaempferol, pinocembrin, or phenolic acids and its esters, including caffeic acid and CAPE, p-coumaric acid [8]. Recently, cardioprotective effect of propolis extract has been investigated both in vitro and in vivo [9]. Although the mechanisms of action beyond these polyphenolic compounds from propolis are not well-defined, it is still convincible that some of their protective effect by propolis can be attributed to direct scavenging properties of these polyphenolic compounds in CP [10].

The study of the material basis and mechanism of natural medicine including propolis always becomes a critical issue for its potential clinical application and modern medicine development [11]. However, mechanisms underlying the cardioprotective effects of CP and its bioactive constituent
basis are still lacking. In the present study, we aim to investigate the protective effects of CP against H$_2$O$_2$ induced rat cardiac H9c2 injury and to explore the potential bioactive compounds of CP.

2. Material and Methods

2.1. Regents. HPLC grade methanol was purchased from Merck (Darmstadt, Germany) analytical grade solvents were purchased from Beijing Chemical Works (Beijing, China). Quercetin, Fura-2/AM probe, Annexin V-FITC/PI apoptosis kit and the standards used in the chromatography analysis were purchased from Sigma (St. Louis, MO, USA). Other chemicals were of analytical grade and purchased from Sangon Biotechnology (Shanghai, China).

2.2. Cell Culture and Cell Viability Assay. Rat H9c2 cardiomyocyte cell line was obtained from ATCC (American Type Culture Collection). Cells were cultured using high glucose Dulbecco's modified Eagle's medium with 4.0 mM L-glutamine (Thermo Scientific) supplemented with 1% penicillin/streptomycin (Solarbio, Beijing, China) and 10% fetal bovine serum (Gibco, Carlsbad, CA, USA). Cells were split when a confluence of ~70% was achieved using trypsin-EDTA (Solarbio, Beijing, China) and subcultured at a ratio of 1:3. H9c2 cardiomyocytes were seeded at a density of 1.2 × 10$^6$ cells/well.

H$_2$O$_2$ (30%, 3 μL) was diluted by cell culture medium (high glucose DMEM, 3 mL) to obtain 10 mM stock and further diluted to specific working solutions. The H9c2 cells were cultured in DMEM without FBS for 12 h and then treated with various concentrations of H$_2$O$_2$ for different periods. Cell viability assay was then performed using a CCK-8 kit. Cellular morphology was observed under an inverted microscope.

Cell viability assay was performed using CCK-8 kit (Solarbio, Beijing, China) according to the manufacturer’s instruction. Then the optical density (OD) at 450 nm for each well was measured by a microplate reader (Synergy HT, BioTek Instruments, Winooski, VT). Cell viability was also confirmed by trypan blue exclusion and microscopy examination during the following experiments.

2.3. CP Active Compounds Separation, Determination, and Selection

2.3.1. CP Sample Preparation. CP was collected from Shandong, China, and the botanical origin was poplar (Populus sp.). A voucher specimen was deposited at Institute of Agricultural Research, Chinese Academy of Agricultural Sciences, China. The propolis stored at −20°C was grinded into powder with a grinder and extracted by 40% ethanol at 30°C, and then the propolis sample was filtered and concentrated under vacuum using a rotary evaporator. The residues were washed and dried to obtain 40% ethanol upper fraction (40EU), and the supernatants were rotated and dried to obtain 40% propolis extracts (40EL). The 70% ethanol and 95% ethanol were further used to extract from the residues to get 70% ethanol extracted propolis (70E) and 95% ethanol extracted propolis (95E).

2.3.2. Fractional Extraction of CP. Propolis sample was mixed by petroleum ether, ethyl acetate, acetone, and methyl alcohol (the proportion of solid to liquid is 1:10). After standing delaminating and evaporation of the solvent, paste-like extraction was obtained. TLC (thin-layer chromatography) condition as followed, chloroform: methanol: formic acid = 8.8: 0.5: 1.0, ferric trichloride (2%) as the chromogenic agent.

2.4. Separation, Purification, and Determination of Propolis Extracts

2.4.1. Column Chromatography. Column was packed with 200–300 mesh silica gel. After eluting with dichloromethane-acetone system in normal pressure, six components were obtained. Column chromatography was performed by silicaSphere C18 chromatography column (packed with 300–400 mesh silica gel), eluted with chloroform-methanol, dichloromethane-methanol, petroleum ether-ethyl acetate, and methanol-H$_2$O system with a solvent flow rate of 10 mL/min in medium-pressure.

2.4.2. High-Performance Liquid Chromatography (HPLC) Analysis. HPLC analyses were carried out on an Agilent HPLC system. The separation was performed on Agilent C18 column (250 mm × 4.6 mm, 5 μm) with a flow rate of 1.0 mL/min at 40°C, eluted with methanol-H$_2$O system. The preparative HPLC was equipped with an Agilent SB C18 column (150 mm × 21.2 mm, 5 μm) with a flow rate of 20 mL/min at 40°C.

2.4.3. LC-MS Determination. An Agilent SB C18 column (2.1 mm × 50 mm, 1.8 μm) was used for the separation at a flow rate of 0.2 mL/min. The mobile phase comprised aqueous 60% methanol. The column was maintained at 40°C. Mass spectrometer operated in negative and positive full-scan mode in the range 100–1000 Da. The capillary voltage set to 4.0 kV and the desolvation temperature was 350°C. The cone gas flow was set at 6 L/h, while the desolvation gas flow was set to 9 L/h, and the collision energy at 20 V.

2.5. Cellular Antioxidant Activity Determination

2.5.1. Measurement of Cellular SOD Activity. The H9c2 cells were seeded into 24-well plates at a density of 1 × 10$^5$ cells/mL. Then the DMEM of propolis treated group was removed and 1% Triton× 100 was added into cells and incubated for 30 min. After this incubation, cells were collected by centrifugation at 2500 rpm for 10 min. The supernatants separated were used 100 ul for measurement of SOD activities according to the manufacturer’s instructions (Jiancheng Bioengineering Institute, Nanjing, China).

2.5.2. Measurement of MDA Activity. Cell culture and treatment methods were as described. We collected 150 μL supernatants of cell lysate for MDA levels measurement according to the manufacturer’s instructions (Jiancheng Bioengineering Institute).
2.5.3. Measurement of GSH-Px Activity. Supernatants (200 μL) were used for measurement of GSH-Px content according to the manufacturer's instructions (Jiancheng Bioengineering Institute).

2.6. Determination of Intracellular Calcium Ion ([Ca^{2+}]_i). H9c2 cells were digested and seeded into culture plate (10^5 cells/mL) at 37°C in a 5% CO₂ atmosphere. The cell medium was discarded and washed cells with HBSS buffer solution 3 times, then added Fura-2/AM and incubated 45 min at 37°C in the dark. After the incubation, the cells were washed 2-3 times with HBSS solution and then 2 mL EBSS buffer solution was loaded. Fura-2 fluorescence was excited alternately at 340 and 380 nm and the 340/380 ratio was obtained. Values for R_min, F 380 max, R 380 min, R_max and Kd were obtained using the Fura-2 Calcium Imaging Calibration Kit (Molecular Probes).

2.7. Cell Apoptosis Analysis Using Flow Cytometry. Cardiomyocytes were labeled with Annexin V-FITC and PI, and apoptosis rate was measured by flow cytometry using a Cell Lab Quanta™ SC flow cytometer (Beckman Coulter Inc., Miami, FL). H9c2 cells were digested and seeded into culture plate to a density of 5 × 10^5 cells/mL at 37°C, 5% CO₂ before the experiment. Cells were then centrifuged at 1000 rpm 5 min and washed 2 times with PBS and 500 μL of conjugation buffer was added to resuspend the cells. Then 5 μL Annexin V-FITC and 10 μL PI were loaded and reacted for 10 min before FCM analysis.

2.8. Statistical Analysis. All experiments were performed in triplicate, and each experiment was repeated at least three times. All values are presented as mean ± SD. The data were analyzed by one-way analysis of variance followed by post hoc Dunnett’s t-test for multiple comparisons. Values of P < 0.05 were considered to be statistically significant.

3. Results and Discussion

3.1. Oxidative Damage Model Establishment in H9c2 Cardiomyocytes. Myocardial damage is largely due to the generation of ROS. ROS-induced effects of ischemia–reperfusion and myocardial dysfunction could be alleviated by treating the tissue with antioxidants or blocking signaling-related ROS generation. Based on this concept, several practices have been performed to evaluate the effects of antioxidants, including propolis, on myocardial injuries in animals and patients [12, 13].

We challenged H9c2 cardiomyocytes with various concentrations of H₂O₂ (0–900 mM) for different time periods (0 to 6 h) to induce in vitro oxidative damage. As shown in Figure 1(a), H9c2 cell viability decreased significantly in a time- and dose-dependent manner after H₂O₂ treatment. Six hours after H₂O₂ challenge, more than 50% cell viability losses were observed in 700 and 900 mM H₂O₂ H9c2 cells (P < 0.01 compared with control cells). In parallel, damaged cell morphology was observed using an inverted microscope, shown as broken cellular membranes, swelling, and vacuole degeneration in 700 mM H₂O₂ treated H9c2 cells (Figure 1(b)), which were quite similar to several previous studies using H9c2 cells [14].

Propolis has abundant polyphenolic constituents, like flavonoids and phenolic acids [15]. These constituents are known with good antioxidant, iron-chelating, and carbonyl reductase-inhibitory effects, which act as new protective compounds against cardiotoxicity [16–18]. It has been reported that quercetin, an important flavonoid abundantly found in fruits, vegetables, wine, and tea (also found in CP [10, 19]), exerts protective effects against H₂O₂ cardiotoxicity in H9c2 cardiomyocytes [20, 21]. We used 5 μM quercetin as a positive control in this study, which rescued cell viability from 53.9% to 86.0% in 700 mM H₂O₂ treated H9c2 cells for 4 h (Figure 1(c)), whereas 900 mM H₂O₂ insult with 5 μM quercetin pretreatment can only rescue H9c2 cell viability to 60%, suggesting that oxidative damage induced by this concentration H₂O₂ was irreversible. Therefore, in our system, 700 μM H₂O₂ challenge for 4 h was chosen in the subsequent experiments as an oxidative damage model.

3.2. CP Extracts and Its Fractions Inhibit H₂O₂-Induced Cell Death in H9c2 Cardiomyocytes. It has been known that different solvents will affect yield of bioactive constituents, like flavonoids or phenolic acids [22]. To find active cardioprotective fractions from CP, different ethanol CP extracts were screened in H₂O₂ challenged H9c2 cells. As shown in Figure 2(a), among all CP fractions tested, 70% ethanol fraction (70E) showed strongest protective effects against H₂O₂ challenge (with a cell viability of 51.3 ± 1.0%), in which the cell viability significantly increased to 75.8 ± 0.2% (P < 0.01). Nevertheless, both 40% ethanol upper fraction and lower fraction (40EU and 40EL) showed less potent protective effects against H9c2 cell viability decreases, which were significant from 70E fraction. Since the protective effects of 90% ethanol fraction (90E) were quite similar to 70E fraction, we merged them for the following fractionation.

Further, merged 70E and 90E fractions were sequentially fractionated into five subextracts explicitly, namely, petroleum ether (PE), dichloromethane (DCM), ethyl acetate (EtOAc), and acetone (thin layer chromatography, TLC). profile of these fractions was shown in Supplemental Figure 1 in Supplementary Material available online at https://doi.org/10.1155/2017/7074147. As shown in Figure 2(b), EtOAc subfraction and acetone subfraction showed most effective protective effects and kept for next fractionation.

Subsequently, combined EtOAc and acetone subfractions were separated over a silica gel column and six subfractions were obtained (TLC profile was shown in Supplemental Figure 2). As shown in Figure 2(c), the cytoprotective effect by fraction 3 was the strongest among all tested fractions, which was comparable to the quercetin positive control. Based on these results, we chosen this fraction for the active compound isolation and structure elucidation.

3.3. Active Compounds Isolation and Characterization of CP Extracts Responsible for Its Cardioprotective Activities. Fraction 3 from EtOAc and acetone sub-fraction was repeatedly separated by silica gel column chromatography, purified by reverse phase HPLC (Figure 3(a)), six known compounds, which is (1) 3,4-dimethoxycinnamic acid (DMCA) [23], (2) pinobanksin [24], (3) benzyl caffeate (BZC) [25], (4) chrysin
Figure 1: Oxidative damage model establishment in H9c2 cardiomyocytes. (a) H9c2 cells were treated with various concentrations of H$_2$O$_2$ (100 μM, 300 μM, 500 μM, 700 μM, and 900 μM) for different time periods (2, 4, and 6 h). (b) H9c2 cell morphology from 700 μM H$_2$O$_2$ stimulation for 0 h (A), 2 h (B), 4 h (C), and 6 h (D). (c) Comparisons of cell survival rates between quercetin treatment group and oxidative injury group. H9c2 cardiomyocytes were pretreated with 5 μM quercetin or not for 12 h and then challenged with 700 or 900 μM H$_2$O$_2$ for 2 h, 4 h, and 6 h. ** P < 0.01 and *** P < 0.001 compared with H$_2$O$_2$ controls.

[23], (5) caffeic acid phenethyl ester (CAPE) [23, 26] and (6) cinnamyl caffeate (CNC) [23, 27].

Next, we examined different concentrations of these isolated active compounds on H$_2$O$_2$ induced H9c2 cell viability decreases. Compared to oxidative damage control group, 10 μM of tested compounds showed significant protective effects (P < 0.01). These results provided further evidence which previously stated that chrysin can alleviate acute cardiotoxicity in rats [28].

Caffeic acid esters, major active compounds widely found in propolis, were reported to have a wide range of biological effects, which were also quite similar to propolis, like antitumour, antioxidant, and anti-inflammatory activities [13]. We noticed that three major caffeate derivatives, BZC, CAPE, and CNC, exerted stronger protective effects than the remaining three polyphenolic acids (DMCA, chrysin, and pinobanksin) in damaged H9c2 cells (Figure 3(b)). These caffeate derivatives were further investigated regarding their cardiac cell protective mechanisms.

3.4. Caffeate Derivatives in CP Increased Cellular Antioxidant Activities in H9c2 Cardiomyocytes. Oxidative stress was considered as a major challenge during the cardiac ischemic damages, which can be reflected by measurement of the products of free-radical attack on biological substrates (MDA) as well as intracellular and extracellular anti-oxidant
Figure 2: (a) Effects of different alcoholic extracts of CP on H9c2 cardiomyocytes cell viability decreases induced by H$_2$O$_2$ (700 μM for 6 h). H9c2 cardiomyocytes were pretreated with 20 μg/mL alcoholic extracts of propolis, that is, 70% ethanol fraction (70E), 95% ethanol fraction (95E), 40% ethanol upper fraction (40EU), and 40% ethanol lower fraction (40EL). Quercetin (5 μM) served as a positive control. **$P<0.01$ versus oxidative injury group, #P < 0.01 versus control group, and $\xi$P < 0.01 versus 70E group. (b) Effects of different fractions from CP using different solvents on H9c2 cardiomyocytes cell viability decreases induced by H$_2$O$_2$ (700 μM for 6 h), that is, petroleum ether (PE), dichloromethane (DCM), ethyl acetate (EtOAc), and acetone fractions. Quercetin (5 μM) served as a positive control. **$P<0.01$ versus oxidative injury group, #P < 0.01 versus control group, and $\xi$P < 0.01 versus EtOAc group. (c) Effects of different subfractions (fractions 1 to 7) from CP EtOAc/acetone fraction on H9c2 cardiomyocytes cell viability decreases induced by H$_2$O$_2$ (700 μM for 6 h). **$P<0.01$ versus oxidative injury group, #P < 0.01 versus control group, and $\xi$P < 0.01, $\zeta$P < 0.05 versus fraction 3 group.

3.5. Caffeate Derivatives in CP Decreased the H$_2$O$_2$ Induced Elevation of [Ca$^{2+}$]$_i$ and Inhibited Cell Apoptosis in H9C2 Cardiomyocytes. Calcium is frequently played in the oxidative stress induced cellular injury [31, 32]. In order to further investigate the protective effects by CP active constituents, [Ca$^{2+}$]$_i$ were determined in H9c2 cells (Figure 5(a)). All of three caffeate derivatives (5 and 10 μM) pretreatment significantly brought back myocardial ionizable calcium to near positive quercetin control. These data are consistent with a previous study in human endothelial cells (HUVEC) that cytosolic [Ca$^{2+}$]$_i$ were also increased by CAPE [33]. Also, these results provided novel evidence for the other two caffeates, which have promising potential in regulating cellular calcium homeostasis [34].

Myocardial ischemic damages can be affected by apoptosis through at least three potential mechanisms: (1) reducing myocardial cell numbers, which will directly decrease the heart pumping function; (2) damaging the heart’s conduction function; and (3) initiating myocardial remodeling and inducing other cardiac pathological changes [29, 35]. For quantitative analyses of myocardial apoptosis, flow cytometry (FCM) with Annexin V-FITC and PI staining was used to detect H9c2 cellular apoptosis. Compared to H$_2$O$_2$ damage group (16.8%–18.8% apoptotic cell rates), significant
FIGURE 3: Continued.
Figure 3: Active compounds isolation and characterization of CP extracts responsible for its cardioprotective activities. (a) HPLC chromatogram of CP fraction 3. Peaks present are as follows: (1) 3,4-dimethoxycinnamic acid (DMCA); (2) pinobanksin; (3) benzyl caffeate (BZC); (4) chrysin; (5) caffeic acid phenethyl ester (CAPE); and (6) cinnamyl caffeate (CNC). Their UV (left) and MS profiles (left) are shown individually as (b) DMCA; (c) pinobanksin; (d) BZC; (e) chrysin; (f) CAPE; (g) CNC.

Figure 4: Caffeate derivatives in CP increased cellular antioxidant activities in H9c2 cardiomyocytes. Effects of CP bioactive compounds (BZC, CAPE, and CNC) on cellular SOD (a), MDA, and GPx activities in H2O2 induced injured H9c2 cardiomyocytes. CP bioactive compounds (1, 5, and 10 µM) were pretreated for 12 h before 6 h H2O2 (700 µM) insult. **P < 0.01 versus oxidative injury group and ##P < 0.01 versus control group.
lower apoptotic cells were observed in caffeate derivatives pretreated cells (5 and 10 \( \mu M \)). Based on these results, we demonstrated that caffetates from CP could reduce the myocardial injury by inhibiting \( \text{H}_2\text{O}_2 \)-induced cell apoptosis and restored cytosolic \([\text{Ca}^{2+}]_i\) pool.

4. Conclusion

In summary, our study provides an important basis for the use of Chinese propolis for the prevention and treatment of cardiovascular diseases. The crude Chinese propolis extract as well as its isolated compounds caffeate derivatives (CAPE, BZC, and CNC) could mostly possibly be useful for the development of new antimyocardial ischemia drugs, depending on their in vitro activity. However, further in vivo pharmacological and toxicity studies are necessary for its potential clinical usages.

Competing Interests

All the authors declare that they have no conflict of interests.

Authors’ Contributions

Liping Sun and Xiang Xu conceived and designed the experiments; Liping Sun, Xiang Xu and Miaomiao Ge performed the experiments; Kai Wang and Yifan Chen analyzed the data; Liping Sun, Xiang Xu, and Miaomiao Ge contributed to the reagents/materials/analysis tools; Liping Sun, Kai Wang, Yifan Chen, and Fuliang Hu wrote the paper.

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References

[1] A. S. Savchenko, J. I. Borissoff, K. Martinod et al., "VWF-mediated leukocyte recruitment with chromatin decondensation by PAD4 increases myocardial ischemia/reperfusion injury in mice," Blood, vol. 123, no. 1, pp. 141–148, 2014.
Evidence-Based Complementary and Alternative Medicine

[2] F. K. Swirski and M. Nahrrendorf, "Leukocyte behavior in atherosclerosis, myocardial infarction, and heart failure," Science, vol. 339, no. 6166, pp. 166–169, 2013.

[3] E. G. Nabel and E. Braunwald, "A tale of coronary artery disease and myocardial infarction," The New England Journal of Medicine, vol. 366, no. 1, pp. 54–63, 2012.

[4] D. Fraccarollo, P. Galuppo, J. Neuser, J. Bauersachs, and J. D. Widder, "Pentacythritol tetranitrate targeting myocardial reactive oxygen species production improves left ventricular remodeling and function in rats with ischemic heart failure," Hypertension, vol. 66, no. 5, pp. 978–987, 2015.

[5] V. Braunersreuther and V. Jaquet, "Reactive oxygen species in myocardial reperfusion injury: from physiopathology to therapeutic approaches," Current Pharmaceutical Biotechnology, vol. 13, no. 1, pp. 97–114, 2012.

[6] K. Wang, S. Ping, S. Huang et al., "Molecular mechanisms underlying the in vitro anti-inflammatory effects of a flavonoid-rich ethanol extract from Chinese propolis (poplar type)," Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 127672, 11 pages, 2013.

[7] J. M. Sforcin and V. Bankova, "Propolis: is there a potential for the development of new drugs?" Journal of Ethnopharmacology, vol. 133, no. 2, pp. 253–260, 2011.

[8] K. Wang, J. Zhang, S. Ping et al., "Anti-inflammatory effects of ethanol extracts of Chinese propolis and buds from poplar (Populus canadensis)," Journal of Ethnopharmacology, vol. 155, no. 1, pp. 300–311, 2014.

[9] J. B. Daleprane and D. S. Abdalla, "Emerging roles of propolis: Antioxidant, cardioprotective, and antiangiogenic actions," Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 175135, 8 pages, 2013.

[10] K. Wang, L. Hu, X.-L. Jin et al., "Polyphenol-rich propolis extracts from China and Brazil exert anti-inflammatory effects by modulating ubiquitination of TRAF6 during the activation of NF-κB," Journal of Functional Foods, vol. 19, pp. 464–478, 2015.

[11] Z. Fang, B. Lu, M. Liu et al., "Evaluating the pharmacological mechanism of Chinese medicine Si-Wu-Tang through multi-level data integration," PLoS ONE, vol. 8, no. 11, Article ID e72334, 2013.

[12] S.-R. Hsieh, W.-C. Cheng, Y.-M. Su, C.-H. Chiu, and Y.-M. Liou, "Molecular targets for anti-oxidative protection of green tea polyphenols against myocardial ischemic injury," BioMedicine, vol. 4, no. 4, pp. 7–16, 2014.

[13] D.-A. Chou, Y.-H. Kuo, M.-S. Jan et al., "Caffeine derivatives induce apoptosis in COLO 205 human colorectal carcinoma cells through Fas- and mitochondria-mediated pathways," Food Chemistry, vol. 131, no. 4, pp. 1460–1465, 2012.

[14] J. H. Ha, H. S. Noh, J. W. Shin, J. R. Hahn, and D. R. Kim, "Mitigation of H2O2-induced autophagic cell death by propofol in H9c2 cardiomyocytes," Cell Biology and Toxicology, vol. 28, no. 1, pp. 19–29, 2012.

[15] K. Wang, X. Jin, Y. Chen et al., "Polyphenol-rich propolis extracts strengthen intestinal barrier function by activating AMPK and ERK signaling," Nutrients, vol. 8, article 272, 2016.

[16] S. Udai, V. Gopi, S. P. Simna, A. Parthasarathy, S. M. J. Yousuf, and V. Elangovan, "Studies on the cardio protective role of gallic acid against age-induced cell proliferation and oxidative stress in H9C2 (2-1) cells," Cardiovascular Toxicology, vol. 12, no. 4, pp. 304–311, 2012.

[17] S. M. Nadtochiy and E. K. Redman, "Mediterranean diet and cardioprotection: the role of nitrite, polysaturated fatty acids, and polyphenols," Nutrition, vol. 27, no. 7–8, pp. 733–744, 2011.

[18] H. Kaiserová, T. Šimunek, W. J. F. van der Vijgh, A. Bast, and E. Krasnićková, "Flavonoids as protectors against doxorubicin cardiotoxicity: role of iron chelation, antioxidant activity and inhibition of carbonyl reductase," Biochimica et Biophysica Acta—Molecular Basis of Disease, vol. 1772, no. 9, pp. 1065–1074, 2007.

[19] J. Zhou, Y. Li, J. Zhao, X. Xue, L. Wu, and F. Chen, "Geographical traceability of propolis by high-performance liquid-chromatography fingerprints," Food Chemistry, vol. 108, no. 2, pp. 749–759, 2008.

[20] J. Daubney, P. L. Bonner, A. J. Hargreaves, and J. M. Dickenson, "Cardioprotective and cardiotoxic effects of quercetin and two of its in vivo metabolites on differentiated H9c2 cardiomyocytes," Basic and Clinical Pharmacology and Toxicology, vol. 116, no. 2, pp. 96–109, 2015.

[21] C. Angeloni, J. P. E. Spencer, E. Leoncini, P. L. Biagi, and S. Hrelia, "Role of quercetin and its in vivo metabolites in protecting H9c2 cells against oxidative stress," Biochimie, vol. 89, no. 1, pp. 73–82, 2007.

[22] Z. Shouqin, X. Jun, and W. Changzheng, "High hydrostatic pressure extraction of flavonoids from propolis," Journal of Chemical Technology and Biotechnology, vol. 80, no. 1, pp. 50–54, 2005.

[23] A. H. Banskota, T. Nagaoka, L. Y. Sumioka et al., "Antiproliferative activity of the Netherlands propolis and its active principles in cancer cell lines," Journal of Ethnopharmacology, vol. 80, no. 1, pp. 67–73, 2002.

[24] T. Usia, A. H. Banskota, Y. Tezuka, K. Midorioka, K. Matsushige, and S. Kadota, " Constituents of Chinese propolis and their antiproliferative activities, " Journal of Natural Products, vol. 65, no. 5, pp. 673–676, 2002.

[25] H. Yang, Y. Dong, H. Du, H. Shi, Y. Peng, and X. Li, "Antioxidant compounds from propolis collected in Anhui, China," Molecules, vol. 16, no. 4, pp. 3444–3455, 2011.

[26] R. Yamauchi, K. Kato, S. Oida et al., "Benzyl caffeate, an antioxidant compound isolated from propolis," Bioscience, Biotechnology, and Biochemistry, vol. 56, no. 8, pp. 1321–1322, 1992.

[27] S. Kumazawa, K. Hayashi, K. Kajiya, T. Ishii, T. Hamasaka, and T. Nakayama, "Studies of the constituents of Uruguayan propolis, " Journal of Agricultural and Food Chemistry, vol. 50, no. 17, pp. 4777–4782, 2002.

[28] E. M. Mantawy, W. M. El-Bakly, A. Esmat, A. M. Badr, and E. El-Demerdash, "Chrysín alleviates acute doxorubicin cardiotoxicity in rats via suppression of oxidative stress, inflammation and apoptosis," European Journal of Pharmacology, vol. 728, no. 1, pp. 107–118, 2014.

[29] R. Chang, Y. Li, X. Yang et al., "Protective role of deoxycholic acid against myocardial ischemia-reperfusion injury in rats," PLoS ONE, vol. 8, no. 4, Article ID e61590, 2013.

[30] R. C. Chen, G. B. Sun, J. Wang, H. J. Zhang, and X. B. Sun, "Naringin protects against anoxia/reoxygenation-induced apoptosis in H9c2 cells via the Nrf2 signaling pathway," Food and Function, vol. 6, no. 4, pp. 1331–1344, 2015.

[31] J. J. Lemasters, T. P. Theruvath, Z. Zhong, and A.-L. Nieminen, "Mitochondrial calcium and the permeability transition in cell death," Biochimica et Biophysica Acta—Bioenergetics, vol. 1787, no. 11, pp. 1395–1401, 2009.
[32] D. M. Ansley and B. Wang, "Oxidative stress and myocardial injury in the diabetic heart," Journal of Pathology, vol. 229, no. 2, pp. 232–241, 2013.

[33] M. Kamil Burgazli, N. Aydogdu, A. Rafiq, M. Mericliler, R. Chasan, and A. Erdogan, "Effects of caffeic acid phenethyl ester (CAPE) on membrane potential and intracellular calcium in human endothelial cells," European Review for Medical and Pharmacological Sciences, vol. 17, no. 6, pp. 720–728, 2013.

[34] O. Ilkun and S. Boudina, "Cardiac dysfunction and oxidative stress in the metabolic syndrome: an update on antioxidant therapies," Current Pharmaceutical Design, vol. 19, no. 27, pp. 4806–4817, 2013.

[35] Y. Lee and Á. B. Gustafsson, "Role of apoptosis in cardiovascular disease," Apoptosis, vol. 14, no. 4, pp. 536–548, 2009.