Effect of freezing-thawing on weight loss, melanosis, and microbial growth in mildly cooked snow crab (Chionoecetes opilio) clusters

Grete Lorentzena,*, Federico Liana, Amalie Agersborg Røhmea,b, Eirin Johannessena,b, Karen Vanglo Grastveita,b, Adrian Eyser Gripa,b, Sten I. Siikavuopioa

a NOFIMA – Norwegian Institute of Food, Fisheries and Aquaculture Research, P.O. Box 6122, NO-9291, Tromsø, Norway
b NTNU, NO-7491, Trondheim, Norway

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ABSTRACT

Different mild cooking treatments of snow crab clusters in combination with immediate refrigeration (IR) or freezing-thawing before refrigeration (FBR) have been performed. The clusters were cooked in either fresh or salted (5 g NaCl/100 ml) water at two temperature/time conditions (87 °C/430 s and 96 °C/148 s) both targeting a core temperature of 81 °C in the largest cluster leg. The freezing-thawing affected quality parameters such as weight loss, melanosis and microbial growth (total viable psychrotrophic counts (TVC) and Pseudomonas spp.) during refrigeration. During refrigeration, FBR clusters showed a higher weight loss compared to the IR clusters, and the development of melanosis in the FBR clusters was faster and more severe compared to the IR clusters. Melanosis was also affected by the cooking treatments as the clusters cooked in water at 96 °C obtained better visual quality scores compared to those cooked in water at 87 °C (P < 0.05). In the FBR clusters, the levels of TVC were lower compared to the IR clusters. In summary, melanosis was the dominant shelf-life limiting factor irrespective of freezing-thawing and cooking combinations. Application of such combinations in processing of snow crabs is therefore not recommended unless measures to reduce melanosis are adopted.

1. Introduction

Snow crab (Chionoecetes opilio) is widely distributed in the North Pacific, the Arctic, the Northwest Atlantic, and in the Barents Sea (Siikavuopio et al., 2017). Its commercial fishery is rapidly growing under the increasing demand for snow crab products, especially in the EU, USA, Japan, and South Korea (Norwegian Seafood Council, 2019). In 2018, 2697 tons of snow crabs were captured in Norway (Norwegian Fishermen’s Sales Organization, 2019).

Nowadays, the entire volume of snow crabs landed in Norway is processed into cooked and frozen clusters, i.e., sections of four walking legs and one claw attached to a shoulder (Lian et al., 2018; Norwegian Fishermen’s Sales Organization, 2019). Usually, the crabs are processed on-board close to the locations of capture. Upon arrival at the destination market, the clusters are either thawed or kept frozen when displayed at points of sale (Lorentzen et al., 2018).

Cooking of snow crab clusters is usually conducted in boiling water to a target core temperature of at least 91–92 °C in the largest leg of the cluster (Lorentzen et al., 2018). However, it may be desirable to apply milder cooking conditions to reduce the risk of overcooking, which is often associated with a lower yield and a poorer eating quality (Martínez-Alvarez, López-Caballero, Gómez-Guillén, & Montero, 2009). Also, reducing total processing time and saving of energy costs are important drivers for a milder cooking treatment.

Freezing of seafood is an effective long-term preservation method. However, freezing and frozen storage may lead to denaturation and aggregation, especially of myofibrillar proteins, resulting in changes in textural attributes, such as reduced juiciness and water holding capacity (Burgaard, 2010). The extent of this quality loss depends on the rate of freezing, duration of frozen storage, speed of thawing, storage temperature, and temperature fluctuations during storage (Boonsumrej, Chaivanichsiri, Tantratian, Suzuki, & Takai, 2007). In shrimp and other shellfish, loss of quality during frozen storage is predominately caused by oxidation, denaturation of proteins, sublimation, and ice recrystallization (Löndahl, 1991). These quality losses often lead to off-flavors, rancidity, dehydration, weight loss, loss of juiciness, and toughening (Bhobe & Pai, 1986).

From time to time, snow crab clusters may develop dark blue spots (i.e., melanosis), that can appear shortly after slaughtering (Lian et al., 2018), leading to product rejection by consumers (García-Carreño, Cota, & Navarrete del Toro, 2008). The occurrence of melanosis is linked to the presence of enzymatic phenoloxidase (PO) activity in crab melanocytes.
tissues including the hemolymph (Gonçalves & de Oliveira, 2016; Lindberg, Sikavuopio, Øverbo, Lorentzen, & Whitaker, 2017). Melanosis is a potential challenge in snow crabs and other crustaceans, especially in the context of mild cooking treatments (Manheem, Benjakul, Kijroongrojana, & Vissessanguan, 2012), as they contain PO enzymes that are highly thermostable (D90 = 3.3–12.0 min) (Huang et al., 2014; Williams, Mamo, & Davidson, 2007).

Psychrotrophic microorganisms such as Pseudomonas spp., Moraxella, Achromobacter and Acinetobacter spp. have been reported to be major contributors to spoilage in blue crab (Callinectes sapidus) (Alford, Tobin, & McClesky, 1942), Dungeness crab (Cancer magister) (Lee & Pfeifer, 1975) and edible crab (Cancer pagurus) (McDermott et al., 2018) during refrigerated storage. During freezing, microorganisms suffer damage which may cause inactivation (Lund, 2000) or death. Death rates are high at the start of freezing, followed by lower inactivation rates during frozen storage. At the same time, microbial growth does not occur at temperatures below −8°C (Farkas, 2007).

Quality parameters related to the frozen storage of raw mud crab (Scylla serrata) (Mishra & Dora, 2008) as well as raw (Paparella & Tatro, 1971) and cooked blue crab (Webb, Tate, Thomas, Carawan, & Monroe, 1976) have been reported. To our knowledge, corresponding studies for cooked snow crab clusters have not been published. Thus, different mild cooking treatments of snow crab clusters in combination with either immediate refrigeration (IR) or freezing-thawing before refrigeration (FBR) was performed. The clusters were evaluated for weight loss, melanosis, and microbial growth during refrigeration.

2. Material and methods

2.1. Raw material

In March 2017, male snow crabs were harvested using snow crab pots in the Barents Sea (75°30.372 N / 33°14.957 E) at depths from 230 to 350 m and stored live onboard in holding tanks. The crabs were transported to the Aquaculture Research Station in Tromso (Norway) and immediately placed in 6 m³ tanks containing seawater at 3°C. After a week, the snow crabs were transported live in a dry state, in polystyrene boxes covered with gel ice (Cold Inc., Oakland, CA, USA), from the Research Station to Nofima in Tromso (N = 76). The crabs were processed at Nofima within 15 h of arrival; they were in good condition and vital before processing. The average weight and carapace length of the crabs was 604.6 ± 164.0 g and 112.8 ± 8.6 mm, respectively.

2.2. Processing, freezing and storage

Initially, experiments were performed to calibrate two temperature/time cooking conditions achieving a common core temperature of 80.8°C in the largest cluster leg. The temperature was monitored using thermocouples connected to data loggers (model 175HI, Testo Ltd., Hampshire, UK). The core temperature of 80.8°C represents a mild cooking treatment as compared with a core temperature of about 91–92°C which is commonly targeted in conventional industrial cooking protocols.

The processing and subsequent storage (IR and FBR) followed an experimental setup as illustrated (Fig. 1). The first step was slaughtering, followed by manual removal of remaining gills using a knife. The 76 cleaned left clusters were placed in one cage, and the corresponding 76 right clusters were placed in another one. To facilitate drainage of the hemolymph (i.e., de-bleeding), both cages with the clusters were soaked in fresh water at 2.5°C for 60 min followed by draining in 10 min (Fig. 1, level 3). The de-bleeding water containing 4-hexyl-resorcinol (0.01 g/100 ml water) (4-HR; Sigma Aldrich Co., Darmstadt, Germany), was used as a melanosis inhibitor being a competitive binder of PO (Lian et al., 2018; Sae-leaw & Benjakul, 2019). Afterward, the clusters were cooked by immersion in either fresh or salted water (5 g NaCl/100 ml) (Sigma Aldrich) at 87°C for 430 s or at 96°C for 148 s, resulting in four different cooking treatments (Fig. 1, level 4). The two temperature/time conditions for the cooking treatments are hereafter denoted as “87°C/430 s” and “96°C/148 s”.

The average weight of the clusters cooked at “87°C/430 s” was 240.0 ± 60.5 g, while for clusters cooked at “96°C/148 s”, the average weight was 183.4 ± 53.9 g. After cooking, the clusters were cooled immediately in fresh water with ice (50 kg ice mixed with 50 l water) until a corresponding core temperature of 1.5°C was reached. In each cooling session, the total snow crab cluster weight was about 5 kg.

The clusters that underwent the same cooking treatment were then further separated into two subgroups (Fig. 1, level 5). In the IR group, 19 clusters from each of the four cooking treatments, were placed individually in plastic bags and closed with paper clips followed by storage at 4°C in a climate chamber (Binder GmbH, Tutlingen, Germany). The FBR group consisted of the remaining 76 clusters, which were evenly distributed on trays and placed in an air-blast freezer at −40°C with an air velocity at 2.39 m/s for 75 min. Each cluster was then individually weighed, packed in plastic bags as described above, and stored for 3 months at −20°C before storage at 4°C in the climate chamber.

The core temperature of the largest legs of two selected clusters in each of the eight subgroups (Fig. 1, level 5) was monitored from cooking up to 12 days of refrigeration using thermocouples connected to data loggers (Testo 175HI). T-bar labels (Floy Tag Inc., Seattle, WA, USA) were used to keep track of each cluster from slaughtering to sample analyses.

2.3. Cooking loss

The cooking loss was calculated as the percentage weight change of the clusters after de-bleeding, cooling, cooling and draining, relative to the weight of the corresponding raw clusters (Lorentzen, Lian, & Sikavuopio, 2019). The calculations were based on the data from at least 15 clusters from each of the four cooking treatments (Fig. 1, level 4).

2.4. Drip loss, thawing/drip loss, and freezing loss

Weight loss in both IR and FBR clusters was monitored for 5 days during storage at 4°C. In RI clusters, the weight loss (i.e., drip loss) was calculated as the percentage cluster weight change during refrigeration relative to the cluster weight after cooking and draining. In FBR clusters, the weight loss included both thawing and drip loss (hereafter denoted as “thawing/drip loss”), and it was calculated as the percentage cluster weight change during refrigeration, relative to the cluster weight after the 3-month storage at −20°C. To obtain uniform conditions for weight loss during refrigeration, all clusters were oriented vertically in the plastic bags with the shoulder joint pointing downwards to allow optimum conditions for drainage.

Freezing loss during the frozen storage was calculated as the percentage cluster weight change during refrigeration. Weight loss and weight gain during storage at −20°C was monitored for 3 months. In each 24-h period, the weight loss included both freezing and drip loss.

In the FBR group, the clusters were placed individually in plastic bags and stored for 3 months at −20°C before storage at 4°C in the climate chamber. The core temperature of the largest legs of two selected clusters in each of the eight subgroups (Fig. 1, level 5) was monitored from cooking up to 12 days of refrigeration using thermocouples connected to data loggers (Testo 175HI). T-bar labels (Floy Tag Inc., Seattle, WA, USA) were used to keep track of each cluster from slaughtering to sample analyses.

2.5. Meat content

At the end of the refrigeration period, for both IR and FBR clusters, cross sections in the middle of the largest legs in 12 selected clusters were performed to evaluate the meat content. The meat content was evaluated as the area of meat in percentage of the total inner area of the cross-section as previously described (Lorentzen et al., 2019). Average values of meat content of the clusters were calculated.

2.6. Melanosis

Melanosis was monitored during the first 48 h of refrigeration in six clusters from each of the eight subgroups (Fig. 1, level 5) by acquiring digital images in standard light condition as described by Lian et al. (2018). Four experienced judges trained at Nofima performed the evaluation independently using the digital images of the clusters.
The rate of melanosis was evaluated using a 5-point visual quality scale, in which “1” referred to remarkable melanosis in the merus, shoulder and joints of the cluster; “2” referred to extensive melanosis in the shoulder and/or in the joints; “3” referred to a slight presence of melanosis in the space between the segments of the shoulder and/or the shoulder edge; “4” referred to the absence of melanosis, but slight darkening of the area around the shoulder joints; and “5” referred to freshly-cooked appearance and absence of melanosis (Lian et al., 2018). Score “3” was defined as the cut-off value for sensory acceptability in accordance with a previous study on visual quality of food products undergoing enzymatic discoloration (López-Gálvez et al., 2015).

2.7. Microbiological analyses

Analyses of total viable psychrotrophic counts (TVC) and Pseudomonas spp. were performed as previously described (Lorentzen, Vorre Skuland, Sone, Johansen, & Rotabakk, 2014). The largest legs of three selected clusters from each of the eight subgroups (Fig. 1, level 5) were sampled at day 4, 5, 7, 9, and 12 of refrigeration. The microbial counts were expressed as log10 CFU/g (mean ± standard deviation).

2.8. Salt content

The salt content was measured in leg meat samples, obtained from the four different cooking treatments, by standard procedure (AOAC, 1990). From each cooking group, a pooled sample of meat was collected from the legs of six selected clusters.

2.9. Statistical analyses

Regression analysis, Scheffe’s and Tukey’s test at a 5% significance level (SPSS windows version 25, SPSS Inc., Chicago, IL, USA) were used to study differences between the eight treatment subgroups (Fig. 1, level 5). Due to an unequal number of samples in the groups, Scheffe’s test was applied to results for weight change, drip loss and thawing/drip loss of IR and FBR clusters during the refrigerated storage. Tukey’s test was applied to the microbial and melanosis results as the number of samples was equal in these subgroups. In the case of microbial results below the detection level (<1.7 log10 CFU/g), Tukey’s test was not performed.

3. Results and discussion

3.1. Weight change after cooking and after freezing

Weight change after cooking varied among the four cooking treatments (Table 1). The weight change of “87°C/430s” with and without 5% NaCl differed significantly (P < 0.05). In the corresponding “96°C/148s” with and without 5% NaCl, the weight change did not differ significantly (P > 0.05), showing that the effect of salt was only present in water at 87°C. However, the salt content in the leg meat was on average equal to 1.65 ± 0.15%, and it did not differ significantly (P > 0.05) between cooking treatment groups.

In a previous study, the weight change of king crab clusters after cooking (i.e., cooking loss), was coherent with the meat content (Lorentzen et al., 2019). In more detail, a low cooking loss corresponded to a high meat content and vice versa. In the present study, a low weight change was observed (Table 1) and the meat content was calculated to about 90% or more. Consequently, a minor weight change in the cooking step can be used as an indicator of a high meat content in snow crabs as well.

For FBR clusters, the weight change after freezing (Table 1) varied among the four cooking treatments (Fig. 1, level 4), although not significantly (P > 0.05). The highest freezing loss was observed in clusters cooked in salted water, irrespective of water temperature and cooking time.

3.2. Drip loss and thawing/drip loss

The weight loss in terms of drip loss and thawing/drip loss during refrigeration was remarkably lower in the IR clusters compared to the corresponding FBR clusters (Table 2). After 5 days of refrigeration, the drip loss in IR clusters was about 2–3%, while the thawing/drip loss of
disrupts the cells, allowing previously compartmentalized contents to storage (Le Bris et al., 2016). Furthermore, the freezing-thawing process allows their ability to cause melanosis even after several months of frozen storage. Hemocyanin-derived POs are stable at freezing temperatures and retain their activity when frozen, as demonstrated by the ability of Melostoma melanaster hemocyanin, an oxygen carrier protein present in the hemolymph, to cause melanosis in fish (Kijroongrojana, & Visessanguan, 2013). In crustaceans, melanosis is caused by the formation of melanin, a pigment produced by the activation of POs (Manheem, Benjakul, Litopenaeus vannamei), freezing and thawing induced higher melanosis levels, likely mediated by increased PO activity (Manheem, Benjakul, & Visessanguan, 2013). In crustaceans, melanosis is mediated by the activation of POs (Manheem, Benjakul, & Visessanguan, 2013).

FBR clusters were about 7%, with no significant differences (P > 0.05) between the four cooking groups (Fig. 1, level 4). The high standard deviation may indicate individual differences, which may, in turn, be ascribed to challenges in orientating the clusters into a vertical position during the daily handling, since the main source of weight loss is the liquid loss through the shoulder of the cluster. Similar differences in weight loss between refrigerated and frozen-thawed meat of blue crab were observed by Webb et al. (1976).

It should be noted that in the case of FBR clusters, the time to obtain a core temperature at 4°C in the largest leg varied from 24 to 36 h, depending on the position in the climate chamber and the cluster size. Thus, a low thawing/drip loss in FBR clusters after one day of refrigeration is explained by the fact that these clusters were still in a partially-frozen state.

The observed differences in weight loss between IR and FBR clusters are probably due to the physical damage caused to the muscle cell wall by the pressure exerted by the ice crystals formed in the freezing process and during frozen storage (Löndahl, 1991). Notably, almost all the claws of the clusters presented visible cracks after air-blast freezing, probably due to the high meat content and freezing rate. Indeed, the cracks could have contributed to the thawing/drip loss.

3.3. Melanosis

The first spots of melanosis were observed as early as only a few hours after cooking, and this was observed in proximity to the cluster joints. Subsequently, it also occurred in other parts of the cluster.

The development of melanosis was faster and more severe in FBR clusters than in the IR clusters. It should be noted that, considering that FBR clusters were still in a partially-frozen state after 24 h of refrigeration, the effect of freezing-thawing on melanosis should be evaluated by comparing the visual quality scores obtained for IR clusters after 24 h of refrigeration with the scores for FBR clusters after 48 h of refrigeration. At this time, the visual quality scores in FBR clusters were below the acceptability threshold except for the clusters cooked in salted water at 96°C (Table 3).

Similarly, in a study on pre-cooked Pacific white shrimps (Litopenaeus vannamei), freezing and thawing induced higher melanosis levels, likely mediated by increased PO activity (Manheem, Benjakul, Kijroongrojana, & Visessanguan, 2013). In crustaceans, melanosis is catalyzed by enzymes with PO activity which can also be displayed by hemocyanin, an oxygen carrier protein present in the hemolymph. Hemocyanin-derived POs are stable at freezing temperatures and retain their ability to cause melanosis even after several months of frozen storage (Le Bris et al., 2016). Furthermore, the freezing-thawing process disrupts the cells, allowing previously compartmentalized contents to react together (García-Carreño et al., 2008), thus exacerbating the development of melanosis.

Regarding the effect of different mild cooking treatments on melanosis, after 24 h of refrigeration, the visual quality score for IR clusters was significantly higher (P < 0.05) in the clusters cooked in water at 96°C compared with their counterparts cooked in water at 87°C, irrespective of the presence of salt in the cooking water. The same trend was also observed for FBR clusters, although significant differences (P < 0.05) between cooking treatments were present only for the clusters cooked at 96°C in salted water.

The difference in visual quality scores observed between cooking treatment groups indicates that the melanosis rate is more a function of the heat exposure of the cluster surface rather than the core temperature of the leg muscle. This might be ascribed to the presence of POs in the joints and in the subcuticular membrane in proximity to the calcareous layer of the shell (Gonzalves & de Oliveira, 2016). These parts of the cluster reach the temperature of the cooking water very quickly, allowing a higher PO inactivation. However, irrespective of the temperature/time conditions applied, the thermal load was not sufficient to provide adequate inactivation of POs to inhibit or satisfactorily delay the melanosis (Williams et al., 2007).

3.4. Microbial growth

The microorganisms analyzed were below the level of detection (<1.7 log10 CFU/g) up to 4 days of refrigeration in both IR and FBR

| Temperature of cooking water (°C)/Cooking time (s) | Cluster group | Drip loss | Visual quality score (±standard deviation) of cooked snow crab clusters immediately refrigerated (IR) after processing or after freezing before refrigeration (FBR). N = 11. | Thawing/drip loss |
|--------------------------------------------------|--------------|----------|-------------------------------------------------------------------------------------------------|------------------|
| 87°C/430 s                                      | Yes          | IR       | Day 1 of refrigeration: −2.19 ± 0.53a, −2.64 ± 0.58a, −2.84 ± 0.64a, −2.71 ± 1.41a, −3.65 ± 1.10a | No −2.72 ± 0.86a |
| 96°C/148 s                                      | Yes          | FBR      | −1.84 ± 1.80a, −5.02 ± 1.59ab, −6.33 ± 1.84ab, −6.95 ± 1.95a, −6.15 ± 2.42a | −2.21 ± 0.71a    |
| 96°C/148 s                                      | No           | FBR      | −2.25 ± 2.18a, −6.51 ± 2.48a, −8.32 ± 3.30ab, −8.34 ± 2.68a, −7.25 ± 2.79a | −0.29 ± 0.94a    |
| 96°C/148 s                                      | No           | IR       | −0.22 ± 1.04a, −4.22 ± 1.90b, −6.13 ± 2.36b, −7.39 ± 2.66a, −8.30 ± 2.53a | −4.58 ± 1.80a    |

Different letters (a or b) within the same column and group (IR or FBR) indicate a significant difference (P < 0.05).

Table 2

| Temperature of cooking water (°C)/Cooking time (s) | Cluster group | IR (refrigeration temperature) | FBR (refrigeration temperature) |
|--------------------------------------------------|--------------|-------------------------------|--------------------------------|
| 87°C/430 s                                      | Yes          | 4.7 ± 0.5a, 3.1 ± 0.9a, 3.2 ± 0.7a, 2.1 ± 0.7a | 4.5 ± 0.6a, 3.1 ± 0.9a, 3.8 ± 0.5ab, 2.4 ± 0.9a |
| 96°C/148 s                                      | Yes          | 4.8 ± 0.4a, 4.5 ± 0.7b, 4.4 ± 0.5ab, 3.6 ± 0.5b | 4.9 ± 0.3a, 4.4 ± 0.7b, 3.9 ± 0.6ab, 2.6 ± 0.8b |

Different letters (a or b) within the same column indicate a significant difference (P < 0.05).

- Clusters were air-blast frozen, and then stored at −20°C for 3 months before refrigeration.
- All clusters thawed.
clusters (data not shown). In IR clusters, no significant differences were observed in TVC (P > 0.05) between cooking treatments at day 7, whereas at day 9 and 12 of refrigeration, the treatment “96 °C/148 s” with 5% NaCl resulted in significantly lower levels of Pseudomonas spp. (P < 0.05) compared to the other cooking treatments.

In FBR clusters, at day 9 and 12 of refrigeration, the cooking treatment “96 °C/148 s” resulted in significantly lower levels of TVC (P < 0.05) compared to “87 °C/430 s” irrespective of the presence of salt in the cooking water (Table 4). Notably, in FBR clusters, Pseudomonas spp. was detected at low levels and only sporadically, except for the clusters cooked at 87 °C for 430 s in fresh water.

In the FBR clusters, lower levels of TVC and Pseudomonas spp. were observed as compared with the IR clusters. Considering the time required for the frozen clusters to thaw during refrigeration (24–36 h), the results of microbial growth at day 5, 7, and 9 in IR clusters can reasonably be compared with the counts at day 7, 9, and 12 in FBR clusters. When performing such a comparison, the TVC levels were within the same log unit for “87 °C/430 s” samples, while the corresponding differences for “96 °C/148 s” were in the range of 2–3 log units. Thus, the freezing-thawing process had a higher limiting effect on TVC during refrigeration in clusters cooked at 96 °C compared to their counterparts cooked at 87 °C. The reduction in TVC level observed in IR clusters, no significant differences were observed in TVC (P > 0.05) between cooking treatments at day 7, whereas at day 9 and 12 of refrigeration, the treatment “96 °C/148 s” with 5% NaCl resulted in significantly lower levels of Pseudomonas spp. (P < 0.05) compared to the other cooking treatments.

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The differences in the microbial growth between IR and FBR clusters could be due to stress imposed by cooking in combination with freezing-thawing, leading to sublethal cell damage, and therefore, limiting the survival and growth of microorganisms in the subsequent refrigeration. Moreover, the freezing-thawing process was more effective in delaying the microbial growth, especially of Pseudomonas spp., in the clusters cooked with the treatment “96 °C/148 s” as compared with “87 °C/430 s”. This suggests that, when applying a mild cooking treatment, a short cooking time at a higher temperature is more effective in delaying microbial growth than a long cooking time at a lower temperature.

3.5. Shelf-life considerations

In literature, there is a lack of studies describing the shelf-life of cooked snow crab clusters. A shelf-life of 10 days was reported for snow crab clusters processed according to the conventional industrial protocol in boiling water to a core temperature of minimum 91 °C in the claw and stored at 4 °C (Lorentzen, Rotabakk, Olsen, Vorre Skuland, & Siikavuopio, 2016). Melanosis was not observed, and the shelf-life was defined as the time to reach the acceptability threshold for off-odors, in turn, corresponding to TVC and Pseudomonas spp. levels of approximately 5–6 and 3–4 log10 CFU/g, respectively.

In the present study, melanosis appeared very early and severely during storage at 4 °C in both IR and FBR clusters. The occurrence of melanosis was therefore the shelf-life limiting factor, as it resulted in visual quality scores below the acceptability threshold as early as after 48 h of refrigeration. Consequently, when applying the mild cooking treatments of the present study, further measures should be implemented to inhibit melanosis and thereby prolong shelf-life. In this regard, considering that oxygen is a key factor in the melanosis cascade (Sae-leaw & Benjakul, 2019), potential solutions may include mild cooking in combination with oxygen-free packaging (i.e., sous-vide cooking, vacuum or modified atmosphere during storage) (Mohan, Ravishankar, & Srinivasa Gopal, 2017).

Furthermore, it should be noted that while the exoskeleton of the cluster was severely affected by melanosis in the early refrigeration period, the leg meat did not present any discoloration in the first 4–6 days of storage, irrespective of the cooking treatment and storage conditions. At points of sale, consumers make their purchasing choice on the basis of the visual appearance of the outer part of the cluster. Therefore, another possibility for prolonging the shelf-life of mildly cooked snow crab clusters is to perform the extraction of the meat in the legs and claws immediately after the cooking treatment (Jun, Jung, Jeong, Kim, & Kim, 2019).

4. Conclusion

The freezing-thawing clearly affected the quality parameters of mildly cooked clusters of snow crab. The weight loss during refrigeration was significantly higher in frozen-thawed clusters compared to the clusters that were refrigerated immediately after cooking. However, neither drip loss or thawing/drip loss was influenced by the different temperature/time conditions of mild cooking.

The freezing-thawing delayed the microbial growth during storage at 4 °C, as suggested by the lower levels of both total viable count and
Pseudomonas spp. observed in frozen-thawed clusters compared to the clusters that were refrigerated immediately after cooking. In addition, in frozen-thawed clusters, the microbial growth was influenced also by the temperature/time conditions applied for mild cooking, with a short treatment time at a higher temperature (96 °C/148 s) resulting in a better quality score of melanosis compared to those cooked in water at “96 °C/148 s”, a better quality score of melanosis was obtained compared to those cooked in water at “87 °C/430 s”). Melanosis was the dominant shelf-life limiting factor irrespective of freezing-thawing and cooking combinations. The development of melanosis was faster and more severe in the clusters that were subjected to freezing-thawing before refrigeration. In clusters cooked in water at “96 °C/148 s”, a better quality score of melanosis was obtained compared to those cooked in water at “87 °C/430 s”).

Declaration of interest

None.

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References

Alford, J. A., Tobin, L., & McClesky, C. S. (1942). Bacterial spoilage of iced fresh crabmeat. Journal of Food Science, 70(5), 353-359.

AOAC (1990). (15th ed.). Official methods for analysis Vol 937, Arlington, VA: Association of Official Analytical Chemists.

Bhobe, A. M., & Pai, J. S. (1986). Study of the properties of frozen shrimps. Journal of Food Science & Technology, 23, 145-147.

Boonsumrej, S., Chaiwanichsiri, S., Tantratian, S., Suzuki, T., & Takai, R. (2007). Effects of previous frozen storage on chemical, microbiological and sensory changes during chilled storage of Mediterranean hake (Merluccius merluccius) after thawing. European Food Research and Technology, 226(1–2), 287-293.

Bhobe, A. M., & Pui, J. S. (1986). Study of the properties of frozen shrimps. Journal of Food Science & Technology, 23, 145-147.

Boonsunrej, S., Chaiwanichsiri, S., Tantratian, S., Suzuki, T., & Takai, R. (2007). Effects of freezing and thawing on the quality changes of tiger shrimp (Penaeus monodon) frozen by air-blast and cryogenic freezing. Journal of Food Engineering, 80(1), 292-299.

Burgard, M. G. (2010). Effect of frozen storage temperature on quality-related changes in fish muscle: Changes in physical, chemical and biochemical quality indicators during short- and long-term storage PhD thesis. Kgs. Lyngby, Denmark: Technical University of Denmark.

Farkas, J. (2007). Physical methods of food preservation. In M. P. Doyle, & L. R. Beuchat (Eds.). Food microbiology. Fundamentals and frontiers (pp. 685–712). Washington, DC: ASM Press.

García-Carreño, F. L., Cota, K., & Navarrete del Toro, M. A. (2008). Phenoloxidase activity of hemocyanin in whitelip shrimp Penaeus vannamei: Conversion, characterization of catalytic properties, and role in postmortem melanosis. Journal of Agricultural and Food Chemistry, 56(15), 6454-6459.

Gonçalves, A. A., & de Oliveira, A. R. M. (2016). Melanosis in crustaceans: A review. Lebensmittel-Wissenschaft und -Technologie- Food Science and Technology, 65, 791-799.

Huang, W., Ji, H., Liu, S., Zhang, C., Chen, Y., Guo, M., et al. (2014). Inactivation effects of freezing-thawing and cooking combinations. The development of melanosis in frozen-thawed clusters, the microbial growth was influenced also by the temperature/time conditions applied for mild cooking, with a short treatment time at a higher temperature (96 °C/148 s) resulting in a better quality score of melanosis compared to those cooked in water at “96 °C/148 s”, a better quality score of melanosis was obtained compared to those cooked in water at “87 °C/430 s”). Melanosis was the dominant shelf-life limiting factor irrespective of freezing-thawing and cooking combinations. The development of melanosis was faster and more severe in the clusters that were subjected to freezing-thawing before refrigeration. In clusters cooked in water at “96 °C/148 s”, a better quality score of melanosis was obtained compared to those cooked in water at “87 °C/430 s”).

Lóndahl, G. (1991). Freezing of crustaceans and mollusks. Infoshift International, 3, 53-56.

Lee, J. S., & Pfeifer, D. K. (1975). Microbiological characteristics of Dungeness crab (Cancer magister). Applied Microbiology, 30(1), 72-78.

Lian, F., Måge, I., Lorentzen, G., Sihlkuvuo, S. I., Øverbe, K., Vang, B., et al. (2018). Exploring the effect of inhibitors, cooking and freezing on melanosis in snow crab (Chionoecetes opilio) clusters. Food Control, 92, 255-266.

Lindberg, D., Sihlkuvuo, S. I., Øverbe, K., Lorentzen, G., & Whitaker, R. D. (2017, March). Evaluating the efficiency of commercial inhibitors in preventing bluing in snow crab. BIOPROS'17. The 8th international conference on marine biotechnology, Tromsø, Norway.

Manheem, K., Benjakul, S., Kijnoonrojanakul, K., & Vissesuan, W. (2010). The effects of the different factors on the shelf-life and quality of frozen shrimp. Food Chemistry, 124(4), 1370-1375.

Manheem, K., Benjakul, S., Kijnoonrojanakul, K., & Vissesuan, W. (2013). Impacts of freezing-thawing on melanosis and quality change of pre-cooked Pacific white shrimp during refrigerated storage. Fish Chemistry, 131(4), 1370-1375.

Marshall, S., Benjakul, S., Kijnoonrojanakul, K., & Vissesuan, W. (2010). The effect of temperature during cold storage on melanosis and quality change of pre-cooked Pacific white shrimp. International Food Research Journal, 20(3), 1277-1283.

Martínez-Alvarez, O., López-Caballero, M. E., Gómez-Guillén, M. C., & Montero, P. (2009). Influence of different freezing methods on the shelf-life of frozen shrimps. Lübeck: Lebensmittel-Wissenschaft und -Technologie- Food Science and Technology, 42(8), 1335-1344.

McDermott, A., Whyte, P., Brunton, N., Lyng, J., Fagan, J., & Bolton, D. J. (2018). The effect of organic acid and sodium chloride dips on the shelf-life of refrigerated Irish brown crab (Cancer pagurus) meat. Lebensmittel-Wissenschaft und -Technologie- Food Science and Technology, 98, 141-147.

Mishra, R., & Dora, K. C. (2008). Shelf life of frozen stored mud crab (Scylla serrata) meat. Indian Journal of Fisheries, 55(4), 333-336.

Mohan, C. O., Ravishankar, C. N., & Srinivasan Gopal, T. K. (2017). Effect of vacuum packaging and sous vide processing on the quality of Indian white shrimp (Penaeus indicus) during chilled storage. Journal of Aquatic Food Product Technology, 26(10), 1280-1293.

Norwegian Fishermen’s Sales Organization (2019). (in Norwegian) https://www.rakislaget.no/, Accessed date: 4 February 2019.

Norwegian Seafood Council (2019). (in Norwegian) https://seafood.no/, Accessed date: 4 February 2019.

Paparella, M. W., & Tatro, M. C. (1971). Freezing and storage life of Blue crab claws. Chesapeake Science, 12(2), 119-126.

Rebach, S., Stribling, J., & Wilber, M. (1990). Frozen storage quality changes in whole Jonah crabs. Journal of Food Quality, 13(3), 203-208.

Sae-leaw, T., & Benjakul, S. (2019). Prevention of melanosis in crustaceans by plant polyphenols: A review. Trends in Food Science & Technology, 85, 1–9.

Sihlkuvuo, S. I., James, P., Olsen, B. R., Evensen, T., Mortensen, A., & Olsen, S. H. (2013). Impacts of freezing-thawing on melanosis and quality change of pre-cooked Pacific white shrimp. Lebensmittel-Wissenschaft und -Technologie- Food Science and Technology, 42(8), 1335-1344.

Subramanian, T. A. (2007). Effect of processing on microbial population of cuttle fish and crab and determination of bacterial spoilage and rancidity developing on frozen storage. Journal of Food Processing and Preservation, 31(1), 13–31.

Webb, N. B., Tate, J. W., Thomas, F. B., Carawan, R. E., & Monroe, R. J. (1976). Effect of freezing, additives, and packaging techniques on the quality of processed blue crab meat. Journal of Milk and Food Technology, 39(5), 345–350.

Williams, H. G., Mano, J., & Davidson, G. W. (2007). Polyphenoloxidase and its thermal deactivation in western rock lobster (Palinurus cygnus) processing. Journal of Aquatic Food Product Technology, 16(1), 87–102.