GATA4 as a novel regulator involved in the development of the neural crest and craniofacial skeleton via Barx1

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Abstract
The role of GATA-binding protein 4 (GATA4) in neural crest cells (NCCs) is poorly defined. Here we showed that mouse NCCs lacking GATA4 exhibited developmental defects in craniofacial bone, teeth, and heart. The defects likely occurred due to decreased cell proliferation at the developmental stage. The in vitro results were consistent with the mouse model. The isobaric tags for relative and absolute quantitation assay revealed that BARX1 is one of the differentially expressed proteins after GATA4 knockdown in NCCs. On the basis of the results of dual-luciferase, electro-mobility shift, and chromatin immunoprecipitation assays, Barx1 expression is directly regulated by GATA4 in NCCs. In zebrafish, gata4 knockdown affects the development of NCCs derivatives. However, the phenotype in zebrafish could be partly rescued by co-injection of gata4 morpholino oligomers and barx1 mRNA. This study identified new downstream targets of GATA4 in NCCs and uncovered additional evidence of the complex regulatory functions of GATA4 in NCC development.

Introduction
GATA-binding protein 4 (GATA4) is a transcription factor that has been extensively studied in heart development and has multiple essential roles in cell proliferation, survival, and differentiation [1]. Depletion of GATA4 in the mouse embryos leads to lethal cardiovascular defects [2, 3]. Moreover, knockdown of gata4 expression in zebrafish by morpholinos (MO) resulted in an unlooped heart tube [4], suggesting that GATA4 has a conserved role in heart development from fish to mammals.

The neural crest (NC) is a multipotent cell population that originates from the dorsal neural tube and gives rise to numerous cell types [5]. Defective NC development is usually associated with many congenital birth abnormalities, including syndromes such as DiGeorge, Treacher–Collins, CHARGE, and Hirschsprung’s disease [6–8]. The development of neural crest cells (NCCs) can be broadly divided into three stages: formation, migration, and differentiation [9]. The inadequate development of NCCs during these stages may result in any of the aforementioned syndromes. The NC derivatives include bone and cartilage of the skull as well as tendons, muscles, and connective tissues of the ear, eye, teeth, and heart [5]. Due to their remarkable plasticity, the development of NCCs requires a complex landscape of transcriptional control [10].

It has been reported that GATA4 is widely expressed in developing embryos [11]. Early GATA4 expression is detected in intra-embryonic cells of the blastocyst. Moreover, GATA4 is also expressed within migratory NCCs.
The specific NCCs marker Sox10 is co-expressed with GATA4 in a large subset of cranial NCCs during embryo development [11]. Our previous microarray data (unpublished data) showed that the GATA4 expression level in the mouse maxillofacial tissues was significantly higher at embryonic day 13.5 (E13.5) than at E18.5. This may indicate that GATA4 plays an essential role in NCCs during the embryo maxillofacial development stage. Moreover, GATA4 also functions as a pioneer factor in osteoblasts [12] and is vital for bone mineralization [13]. These findings...
Fig. 1 Mice lacking GATA4 in NCC-derived tissue exhibit craniofacial and cardiac defects. a Localization of GATA4 expression during mouse development at embryonic day 14.5 (E14.5) and at postnatal day 1 (P1) and P14 (scale bar: 50 μm). Brown staining corresponds to anti-GATA4 immunoreactivity. b Photographic analysis of 3-week-old Wnt1-cre;Gata4fl/fl (Mutant) mice and Gata4fl/fl (WT) mice (n = 5). (c, Upper) Wnt1-cre;Gata4fl/fl and Gata4fl/fl P1 skulls stained with Alizarin red and Alcian blue. (c, Middle) Magnification of the calvaria with widened cranial frontal sutures in mutant mice. (c, Lower) Sagittal view shows the mandibular retrogrowthism of the mutant mice. Scale bar, 500 μm. d Quantitative analysis of the length of the mandible (n = 5). e Three-dimensional micro-CT image of the mandible and teeth at P21. Mutant mice displayed a drastic decrease in mineralization and short root abnormalities. Yellow arrow indicates the decreased mineralization area. Scale bar, 500 μm. g Three-dimensional micro-CT image of the skull (Left) and palate bone (Right) at P21. The right pictures of each group show the thickness of the frontal bone and palate bone. h Mandible tissue stains of von Kossa, total collagen, and Collagen I at P1; scale bars: 100, 200, and 100 μm, respectively. i-k Quantitative analysis of the positive area (the whole image section was analyzed for the mineralization area quantification) of von Kossa, total collagen, and Collagen I staining, respectively (n = 5). i Sagittal and coronal view of the heart in mutant mice showed VSD and enlarged hearts; scale bar: 200 μm. The data are presented as the mean ± S.E.M. from at least three independent experiments. **P < 0.01. cdp condylar process, CT computed tomography, la labial, li lingual, NCC neural crest cell, VSD ventricular septal defect

prompted our interest in exploring whether GATA4 is involved in the development of NCC derivatives. Although great progress has been made toward understanding the gene regulatory networks underlying NC development, the function of GATA4 in NC development remains poorly understood.

To date, there have been no reports on the specific mechanism of GATA4’s role in NC development due to the early lethality of GATA4-knockout embryos at around E8.5–E9.5 [14, 15]. Therefore, conditional loss-of-function models for GATA4 genes could be applied to reveal the role of GATA4 in NCCs. In this study, we generated and examined the Wnt1-Cre;Gata4fl/fl conditional knockout mouse model in which GATA4 was conditionally ablated in the NCCs. In addition, the phenotype associated with MO-induced knockdown of gata4 in zebrafish was assessed and observed to be consistent with that obtained in mice. Taken together, these results obtained in the two independent animal models described herein offer interesting new insights into the key role of GATA4 in regulating NC development.

Results

Mutant mice exhibit craniofacial and cardiac defects

According to a previous study [11], GATA4 transcripts are found within most of the migratory NCCs. Thus, we examined the expression patterns of GATA4 in the mouse NCC-derived craniofacial tissue using immunohistochemical analysis at E14.5, postnatal day 1 (P1), and P14 (Fig. 1a; detailed descriptions and magnified images of each panel can be found in the Supplementary Figure S1a–c). In the mandible, GATA4 was expressed in the osteoblasts, and the positively stained cells were distributed along the surface of the bone trabecular (Supplementary Figure S1a). In the teeth, GATA4 was expressed in the dental mesenchyme, odontoblasts, and ameloblasts (Supplementary Figure S1b, red arrows, black arrows, and yellow arrows, respectively). In the palate, GATA4 was expressed both in the palate epithelium and palate mesenchyme at E14.5 and P1. In the palate at P14, the GATA4-positive cells (osteoblasts) were distributed along the surface of bone trabecular (Supplementary Figure S1c).

To investigate whether GATA4 controls craniofacial development, we ablated GATA4, specifically in NCCs and NCC-derived tissues, by crossing a floxed Gata4 allele with a Wnt1-Cre driver. The Wnt1-Cre;Gata4fl/fl (mutant) mice were born in the predicted Mendelian ratio and survived into adulthood. Compared with control Gata4fl/fl (wild-type, WT) littermates, 3-week-old Wnt1-Cre;Gata4fl/fl mutants exhibited short stature (Fig. 1b and Supplementary Figures S1d–f). We subsequently performed Alcian Blue and Alizarin Red staining at P1 to examine the role of GATA4 in cranial development. On the basis of the results, the mutants exhibited widened cranial frontal sutures and mandibular retrogrowthism (Fig. 1c–e). Micro-computed tomography (micro-CT) scanning analysis revealed a drastic decrease in mineralization of the mandible, frontal bone, and palate in mutants (Fig. 1f and g and Supplementary Figure S1g–j). In addition, the mutants’ teeth were grossly smaller than those of controls (Fig. 1f). Von Kossa, total collagen, and collagen I staining revealed a substantial decrease in mineralization in the mandible of P1 Wnt1-cre; Gata4fl/fl mice compared with that in control mice (Fig. 1h–k). Strikingly, the absence of GATA4 in NCCs induced severe cardiac defects with a prominent ventricular septal defect (VSD) and enlarged heart (Fig. 1i). However, cardiac development was beyond the scope of the present study, which focused on craniofacial development.

Lack of GATA4 in NCCs affects proliferation at E14.5

Given that the loss of GATA4 in NCCs impaired both bone and tooth growth during development (Fig. 1), we hypothesized that GATA4 may have a direct effect on early NCC development, in addition to its roles in migratory NCC [11]. To validate our hypothesis, we analyzed the pharyngeal arch 1 (pa1) by staining for SOX9, a marker of early migrating NCCs, in E9.5 embryos. Our results showed no differences in NCC migration into the pa1 between WT and mutant mice (Fig. 2a, b). Additionally, phosphohistone H3
(PHH3) and terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) staining at E10.5 also showed no significant differences in cell proliferation and death between Wnt1-cre;Gata4\textsuperscript{fl/fl} (Mutant) embryos at embryonic day 9.5 (E9.5) were stained with SOX9 antibody, a marker of early migrating NCCs. There were no significant differences between the two groups. The dotted line shows the pa1; scale bar: 200 \( \mu \)m. 

The results showed that the specific loss of GATA4 in NCCs decreased cell proliferation in the mandible (Fig. 2h, i); however, the apoptosis levels were comparable between mutant and WT mice (Fig. 2j, k). Taken together, these observations suggest that GATA4 is essential for the development of NCC-derived craniofacial derivatives through its role in promoting proliferation at the developmental stage.

**GATA4 regulates the function of NCCs in vitro**

To provide further evidence of the function of GATA4 in vitro, GATA4 expression was reduced in primary embryonic NCCs collected from adult WT mice by using a small hairpin RNA (shRNA)-mediated knockdown approach. The results indicated that GATA4 knockdown decreased the proliferative ability of NCCs, as measured by...
cell counting kit-8 (CCK-8) assay, compared with that of the control group on culture day 4 (Fig. 3a). However, Annexin V-fluorescein isothiocyanate (FITC)/propidium iodide staining showed no significant difference in the percentages of apoptotic cells between the two groups (Fig. 3b, c). Moreover, shGATA4-treated NCCs exhibited slower scratch closure, indicating that the migratory ability was attenuated (Fig. 3d). The osteogenic differentiation ability of NCCs treated with shGATA4 followed by mineralization for 14 days was significantly decreased compared with the control group (Fig. 3e, f). Furthermore, we extracted the mRNA from the pa1 of WT and mutant mice at E10.5 and performed real-time quantitative polymerase chain reaction (qPCR) analysis for nine key genes of interest including Sox9, Snail2, Ets1, Msx1, AP-2, Twist, Nestin, Pax3, and HNK-1, which are known to play important roles in the development of NCCs. As demonstrated in Fig. 3g,h, the mRNA expression levels of these key genes (except Nestin, Pax3, and HNK-1) were decreased dramatically after GATA4 knockdown. Taken together, these in vitro findings are consistent with the in vivo observations (animal model phenotype) and the hypothesis that GATA4 is critical for the function of NCCs.
BARX1 as a distinct protein identified by isobaric tags for relative and absolute quantitation (iTRAQ) analysis in NCCs after GATA4 knockdown

Based on the aforementioned in vivo and in vitro results, we sought to examine the potential mechanism underlying the function of GATA4 in NCCs. Thus, iTRAQ analysis was used to identify proteomic changes after GATA4 knockdown in NCCs (Fig. 4a). Among the identified proteins, a group of 43 proteins was significantly upregulated, whereas 19 proteins were significantly downregulated after GATA4 knockdown (Fig. 4b, c). Our results revealed that BARX1 was among the downregulated proteins after GATA4 knockdown. BARX1, was examined in NCCs using qRT-PCR ($n = 5$). e BARX1 was also examined in p1 of mice at E10.5 using qRT-PCR ($n = 5$). f, g Protein expression of BARX1 in the NCCs was tested by Western blotting ($n = 3$). The data are presented as the mean ± S.E.M. from at least three independent experiments. *$P < 0.05$, **$P < 0.01$. iTRAQ isobaric tags for relative and absolute quantitation, NCC neural crest cell, qRT-PCR quantitative reverse transcription-polymerase chain reaction, shCTRL short hairpin control (control lentivirus).

GATA4 binds to the Barx1 promoter

Several putative GATA4-binding sites upstream of the transcription start site of the mouse Barx1 promoter were acquired from the JASPAR database, and two predicted binding sites with high scores were selected for further investigation (Fig. 5a). Both a dual-luciferase assay and electrophoretic mobility shift assay (EMSA) were performed to analyze the respective promoter regions in vitro. In the dual-luciferase assay, we constructed several reporter vectors, including WT GATA4-binding sites or mutant GATA4-binding sites (Fig. 5b). Overexpression of GATA4 enhanced the transcription of Barx1 compared with that in the controls (Fig. 5c). The mutant GATA4 core binding sequence of the Barx1 promoter reduced the induction via both binding sites (Fig. 5c). As revealed by EMSA, in the first predicted binding site, a retarded band was observed in
the lane containing the GATA4-Barx1 and the biotin-labeled oligonucleotide probe (Fig. 5d) that encompassed the putative GATA4-binding sequence. Since we did not find a binding activity in the second binding site by EMSA assay, we selected the first binding site for further analysis.

To identify direct targets of GATA4 in vivo, a chromatin immunoprecipitation (ChIP) assay was carried out using NCCs. We analyzed the first predicted promoter region and designed primers accordingly. ChIP analysis was performed using anti-GATA4 antibody, and the ChIP input value represented the binding efficiency. The ChIP assay demonstrated a significant increase in the binding efficiency, confirming the in vivo binding of GATA4 to the Barx1 promoter (Fig. 5e, f).

Full images of EMSA and ChIP assay results are presented in Supplementary Figure S2.

Knockdown of gata4 in zebrafish affects NC derivatives

To confirm that gata4 expression promotes the development of NC derivatives via barx1, we adopted a loss-of-function analysis of gata4 in zebrafish, since both zebrafish embryos and mouse embryos lacking the gene exhibit similar
cardiomyopathies [4, 19]. To this end, we examined the phenotype of NC derivatives in zebrafish embryos.

To knockdown the expression of gata4 in zebrafish embryos, we used gata4 MO. Zebrafish embryos injected with a standard control MO and gata4 MO, respectively. The gata4 MO embryos showed a shorter body length, mandibular deficiency (black dotted line and blue arrow), edema around the heart (red dotted line and black arrow) and regression of the yolk stalk extension (red arrow). Alcian blue staining revealed a defective mandibular arch compared with controls. The dotted line represents the length of mandible.

**Fig. 6** gata4 is required for the development of the neural crest derivatives in zebrafish. a The qRT-PCR analysis showed that gata4 MO knocked down gata4 efficiently (n = 5). b At 72 hpf, zebrafish larvae were imaged with transmitted light. Control embryos and gata4 MO embryos were injected with a standard control MO and gata4 MO, respectively. The gata4 MO embryos showed a shorter body length, mandibular deficiency (black dotted line and blue arrow), edema around the heart (red dotted line and black arrow) and regression of the yolk stalk extension (red arrow). Alcian blue staining revealed a defective mandibular arch compared with controls. The dotted line represents the length of mandible. d Quantitative analysis of the length of mandibular arch (n = 5). e At 72 hpf, the distribution and amount of iridophores were reduced in gata4 morphants. f, g Quantitative analysis of the number of iridophores in the eyes and the body (n = 5). h At 96 hpf, the distribution and amount of iridophores were reduced in gata4 MO embryos. i, j Quantitative analysis of the number of iridophores in the eyes and the body (n = 5). The data are presented as the mean ± S.E.M. from at least three independent experiments. **P < 0.01. hpf hours post-fertilization, MO morpholino, NCC neural crest cell, qRT-PCR quantitative reverse transcription-polymerase chain reaction**
established that iridophores and the mandibular arch are both NC derivatives. Taken together, these results indicate that gata4 is required for the normal development of NC derivatives, which is consistent with our results in the mouse model.

**barx1 mRNA partly restores the defects in gata4 morphants**

Barx1 is expressed in the pharyngeal arches of both mice and zebrafish. barx1 mutant zebrafish exhibit a severely reduced lower jaw [18], which is similar to our observation in the gata4 MO group, indicating that gata4 may promote the development of NC derivatives via barx1. Thus, we examined the expression of barx1 in zebrafish. Whole-mount in situ hybridization showed that barx1 expression in the control group at 24 hpf was normal in each of the pharyngeal arches within the dorsal and ventral domains that developed into cartilage elements. However, the gata4 MO group displayed a significantly lower expression of barx1 in these areas (Fig. 7a). These results were further confirmed by qRT-PCR analysis, which indicated a nearly 30% decrease in the expression of barx1 upon the down-regulation of gata4 (Fig. 7b).

To determine whether barx1 expression was regulated by gata4, rescue experiments were carried out by co-injecting gata4 MO and barx1 mRNA. Notably, administration of barx1 mRNA could partially rescue the defects caused by gata4 deficiency (Fig. 7c). The length of the mandible and the number of the iridophores in the eyes and the body were also partly rescued compared with those in the gata4 MO group (Fig. 7d–h).

In summary, our findings using two independent animal models indicated that Barx1 is a downstream effector of GATA4, which thereby trans-activates Barx1 expression and regulates NC development.
Discussion

NC developmental defects contribute to clinical pathologies of many human congenital malformations. Understanding the origins of these diseases and the molecular mechanisms governing NC development will help improve current diagnostics and therapeutics. Our study is the first to characterize the relationship between GATA4 and Barx1 in NC development. Our results demonstrated that GATA4 is essential for normal NC development by promoting NCC proliferation at the developmental stage through direct regulation of Barx1 at the transcriptional level.

GATA4 has highly conserved zinc finger domains and has been widely studied in the context of early embryonic cardiac development in various species [20, 21, 22]. The structure of the heart is also derived from NCCs. In our study, GATA4 knockout in NCCs resulted in ventricular septal defects and enlarged hearts. The knockdown of gata4 in our zebrafish model also induced heart failure, which is consistent with previous findings [4]. However, the specific function of GATA4 in NC development has never been studied due to the early lethality in GATA4-deficient mice. Thus, the NCC-specific knockout mouse Wnt1-Cre;Gata4fl/fl in our study was a useful animal model for studying NC development.

GATA4 was reported to be expressed in migratory NCCs [11], which indicates that it may influence the migration of NCCs. However, in our study, conditional knockout of GATA4 had no effect on the migratory activity of NCCs. Instead, the absence of GATA4 decreased cell proliferation in NCC-derived tissues at developmental time points, potentially leading to the phenotypes of decreased bone mineralization, mandibular retrognathism, and short root deformity of molars. Several studies have revealed positive regulation of GATA4 in the function of osteoblasts [12, 13], which supports our conclusion.

GATA4 as a transcription factor regulates several downstream molecules in different tissues, including the bone morphogenetic protein 4 (BMP4) gene, which was identified as a direct downstream target for GATA4 in mesoderm differentiation [23]. However, GATA4 and its downstream molecular events in NC development were still unclear. Through dual-luciferase, EMSA, and ChIP assays, our study indicated that Barx1 may be one of the direct downstream targets of GATA4 in NCCs. Barx1 is a homeobox-containing transcription factor that is implicated in the processes of craniofacial, tooth, articular development, and stomach organogenesis [18, 24–27]. Microdeletion of the Barx1 gene in the human chromosome 9q22.32 is related to craniofacial developmental disorders such as microstomia deformity (small mouth) and mandibular retrusion [28, 29]. Moreover, barx1 morphants in zebrafish exhibited mandibular arch lengthening [27]. Notably, the phenotypes in humans and zebrafish are similar to the phenotype reported in the present study. This supports our results that Barx1 may serve as a direct downstream factor of GATA4 involved in NC development.

Taken together, these findings demonstrate that the regulation of GATA4 in NCCs was required for normal NCC development and differentiation. Furthermore, the loss of GATA4 led to NCC-derived craniofacial bone, teeth, and heart hypoplasia characterized by decreased ossification and aberrant cell proliferation. Uncovering the relationship between GATA4 and Barx1 in NCCs during development is critical to understand the complex morphogenic event. It may also provide direct insight into the causes and potential treatments of similar disorders in humans.

Materials and Methods

Animals

Wnt1-Cre and Gata4fl/fl mice have been described elsewhere [9, 19]. To knockout GATA4 specifically in NCCs, we crossed Wnt1-Cre;Gata4+/fl males with Gata4+/fl females (C57/B6). We used Gata4+/fl mice and Wnt1-Cre mice as controls and Wnt1-Cre;Gata4fl/fl as mutants. The mice were obtained from the Model Animal Research Center of Nanjing University (MARC). Zebrafish were raised on a 14/10 h light/dark cycle at 28.5 °C in the zebrafish facility of the Model Animal Research Center, Nanjing University. All experiments were performed with the approval of the Ethics Committee of the Stomatological School of Nanjing Medical University. All procedures were performed according to the guidelines of the Animal Care Committee of Nanjing Medical University.

Skeletal preparations

According to standard protocols [30], mice were skinned, eviscerated, then fixed in 95% (vol/vol) ethanol, and finally cleared in acetone. Sections were stained with Alcian blue and Alizarin red, and the cartilage and mineralized bone were identified by blue and red staining, respectively. To measure the length of the mouse mandible, the front most part of the mandible and the condylar process were used as the endpoints.

Micro-CT analysis

Mouse heads were harvested at P21, soft tissues were removed, and the remaining tissues were stored in 70% ethanol overnight. Scanning was performed with a micro-CT scanner (Skyscan, Kontich, Belgium). Slice thickness was 18 μm at 50 kV and 456 μA [31]. Finally, images were
reconstructed and analyzed using NRecon v1.6 and CTAn v1.13.8.1 software.

**Histology and immunostaining**

*Wnt1-Cre;Gata4fl/fl* and *Gata4fl/fl* mice were harvested at E9.5, E10.5, E14.5, P1, and P14. Embryos and tissues were carefully dissected and fixed in freshly prepared 4% paraformaldehyde (PFA). The tissue blocks were cut into 5-μm thick sections and mounted on glass slides. Sections were stained with hematoxylin and eosin using standard methods [32]. Von Kossa staining (1% silver nitrate) and total collagen staining (1% Sirius Red) were performed to detect the areas of mineralization [9, 33]. For whole-mount immunostaining, the embryos were blocked with protein block (0.2% bovine serum albumin, 0.2% Triton X-100 in phosphate-buffered saline [PBS]), and stained with rabbit anti-Sox9 (Millipore, #ABE571, 1:1000) [9]. Other antibodies used in this section were rabbit anti-GATA4 antibody (Abcam, #ab84593, 1:300), rabbit anti-collagen I (Abcam, #ab21286, 1:300) and phosphohistone H3 (PHH3; Cell Signaling, #9017, 1:100). The TUNEL assay was performed with the In Situ Cell Death Detection Kit following the manufacturer’s instructions (Roche, #12156792910). Nuclei were stained with 4',6-diamidino-2-phenylindole (DAPI) at 1:1000. Staining was subsequently visualized by using a laser confocal scanning microscopy (Carl Zeiss, Heidenheim, Germany) and Leica DM 4000 Fluorescence System.

**Culture of NCCs**

Embryos were harvested at gestational day E9.5, and deciduas were subsequently removed using microscissors and scalpels. The first branchial arch was extracted in PBS and dissociated in 0.25% trypsin-ethylenediaminetetraacetic acid (Gibco, #25200056) for 30 min at 37 °C, and the dissociated cells were plated into a 6-well dish [34, 35]. Cells were cultured in Dulbecco’s Modified Eagle Medium: Nutrient Mixture F-12 (1:1) media containing 15% fetal bovine serum supplemented with 100 U/mL penicillin, 100 mg/mL streptomycin, 100 mg/mL L-glutamate, 0.1 mM minimum essential medium non-essential amino acids, 1 mM sodium pyruvate, 55 mM β-mercaptoethanol, 25 mg/mL basic fibroblast growth factor (Gibco, #AA10155), and 1000 U/mL leukemia inhibitory factor (PeproTech, #25002), and then filtered with a 0.22-μm pore size filter.

**Lentiviral transfection**

Recombinant lentivirus of shRNA to knockdown Gata4 expression (shGATA4; 5'-CCAAGCAGGACTTTG-GAA-3') and control lentivirus (shCTRL; 5'-TTCTCCGA-ACGTGTACGT-3') were purchased from GenePharma (GenePharma, Shanghai, China). The NCCs were seeded in 6-well culture plates at a density of 1.5 × 10^5 cells per well and cultured overnight. When cells grew to 60–70% confluence, they were infected with the lentivirus (multiplicity of infection = 50) in the presence of 5 μg/mL Polybrene [36]. The medium was replaced with fresh medium after 24 h. The efficiency of the lentiviral infection was measured by fluorescence microscopy 3 days later (Leica Microsystems, Ontario, Canada).

**Apoptosis analysis**

For the analysis of cell apoptosis, cells were collected and stained with the Annexin V-FITC/Propidium Iodide Kit (KeyGen Biotech, Nanjing, China) according to the manufacturer’s instructions. NCCs were incubated in the dark for 30 min at room temperature followed by the analysis of the apoptotic cells by flow cytometry [37]. FlowJo V7 software (Tree Star, Oregon, USA) was used to analyze the data.

**Wound scratch assay**

The NCCs were transfected with lentivirus and seeded at a density of 1.5 × 10^5 cells per well in 6-well plates. When cells reached 95–100% confluency, a wound was created using a pipette tip, and the debris and floating cells were subsequently removed by washing the cells once with 0.1 mol/L PBS [38]. Wound closure was captured at 24 h using a Leica DMIRE 2 microscope in phase contrast mode and Leica FW 4000 software.

**CCK8 assay**

Transfected cells were seeded into 96-well plates and CCK8 reagent (Dojindo, #CK04) was added to each well for 1 h incubation at 37 °C at five different time points (0, 24, 48, 72, and 96 h). The cell proliferation rate was measured at a wavelength of 450 nm by a microplate reader [39]. Cells infected with a control lentivirus were used as controls.

**Alizarin red staining**

Cells were cultured in mineralization-inducing medium for 2 weeks, and the calcification of osteoblast was subsequently analyzed by Alizarin Red staining. After fixation in 10% formaldehyde and rinsing with distilled water, cells were stained with 1% Alizarin Red (Beyotime, Shanghai, China) at room temperature for 20 min [40]. Alizarin red was eluted using 10% cetylpyridinium chloride in 10 mM sodium phosphate for 30 min at room temperature, and the calcium mineral content was subsequently evaluated by
measuring the absorbance at 562 nm on a 96-well plate reader.

**Western blotting**

Proteins were extracted by using a cell lysis reagent containing the protease inhibitor phenylmethylsulfonyl fluoride [41]. Protein samples were boiled for 5 min and then separated on a denatured SDS-polyacrylamide gel before transfer onto a polyvinylidene difluoride membrane at 300 mA for 1 h. The blotting membrane was blocked with bovine serum albumin for 2 h and then incubated overnight at 4 °C in Barx1 (Abcam, #ab181851, 1:1000) and β-Actin (Santa Cruz Biotechnology, #sc-81178, 1:500). After incubation with secondary antibody at room temperature for 1 h, the membranes were rinsed three times in Tris-buffered saline with Tween-20 and visualized by enhanced chemiluminescence. Semi-quantitative measurements were carried out using ImageJ software.

**Quantitative reverse transcription PCR for mRNA analysis**

Total cell RNA was isolated by using TRIZol reagent (Life Technologies, #15596018) according to the manufacturer’s protocol [42]. The mRNA was employed to generate cDNA using a PrimeScript RT reagent kit (TaKaRa, #RR047A). The gene expression level was analyzed by qRT-PCR using the ABI-7300 Real-Time PCR System (Applied Biosystems). The primers used are listed in Supplemental Table S1.

**Protein extraction and iTRAQ labeling**

Total proteins of each sample were extracted from the transfected NCCs. After reduction and alkylation by 10 mM dithiothreitol and 55 mM iodoacetamide, each group proteins were digested overnight with trypsin at 37 °C. Finally, samples were labeled with iTRAQ reagents [43]. The samples from both experimental conditions were pooled and subjected to fractionation.

**Plasmid construction**

The predicted GATA4-binding site (obtained from the JASPAR database), with the SpeI/XbaI enzyme site and the oligonucleotide sequence (Forward: 5'-CAGCCTCCGGACTCTAGC-3'; Reverse: 5'-TAATACGACTCACTATAGGG-3') was synthesized and cloned into the SpeI/XbaI site of the pEZX-FR03 luciferase vector after reannealing. The pEZX-FR03 luciferase vector containing the GATA4-binding site was constructed at the same time.

**Dual-luciferase assay**

For the luciferase reporter experiments [44], 293T cells were cultured in 24-well plates at a density of 1.0 × 10^5 cells per well. When cells grew to 80%, they were co-transfected with pEZ-GATA4, pEZ-Barx1 promoter-luciferase (100 ng per well), and plasmid pEZ-SV40 (10 ng per well) using Polyetherimide. 48 h after transfection, cells were collected and lysed in 1 x passive lysis buffer. Luciferase assays were performed using the Dual-Luciferase Reporter Assay System (Promega, Madison, WI, USA). All samples were tested in triplicate.

**Electro-mobility shift assay (EMSA)**

Briefly, nuclear proteins extracted from NCCs were incubated with a biotin-labeled GATA4-binding site DNA probe in binding buffer for 30 min at room temperature. The probe used for the reaction contained the GATA4-binding site of the Barx1 promoter with a sequence of 5’-CTGAGAGGCGCAGATGGAAATACA-3’. To determine the binding specificity, “cold probe” or “mutant probe”, which lacked 5’-biotin labeling, were also used in the competitive EMSA to assess the involvement of GATA4 (Supplemental Table S2). A probe lacking nuclear extracts was used as a negative control. The protein–DNA complexes were then electrophoresed on a 6% acrylamide gel and analyzed by autoradiography. Anti-GATA4 polyclonal antibody (0.1 μg/μL; Santa Cruz Biotechnology, #sc-25310 × ) was used for supershift assays. All the experiments were performed in triplicate.

**The ChIP assay**

The ChIP assays were performed using EZ-ChIP (Millipore, #17371) according to the manufacturer’s protocol [45]. Briefly, NCCs were incubated in 5 mM dimethyl 3,3-dithiobispropionimidate-HCl (DTBP) for 10 min at room temperature, and cross-linked with 1% formaldehyde for 10 min at 37 °C. Lysates diluted with ChIP dilution buffer were immunoprecipitated with anti-GATA4 (Abcam, #ab86371) or rabbit immunoglobulin as an internal control. Precipitated DNA samples (with or without antibody) as a template were quantified with qPCR to amplify the fragment of Barx1 promoter. The primers are listed in Supplemental Table S3. Products from qPCR were electrophoresed on 2% agarose gel stained with ethidium bromide and visualized under UV light.

**Microinjection of MO and mRNA**

The MO antisense oligonucleotides were synthesized by Gene Tools (Philomath, USA); MO against gata4 were used [4] as follows: gata4 MO1 (5'-TCCACAGGTGAGCTTATTGCTCC-3'), and standard control MO (5'-CCTCTACTCCAGTTACAATTATA-3'). All the experiments in this study
used 10 ng MO per injection. Total RNA was extracted with Trizol reagent (Ambion) and reverse-transcribed with the Reverse Transcription System (Takara). The sequence of barx1 cDNA was cloned and ligated to the pGEM plasmid, linearized by BamHI, and transcribed with the mMESSAGE mMACHINE T7 kit (Ambion, #AM1344). The mRNA encoding barx1 was injected into one-cell stage zebrafish embryos at 0.8 ng/embryo.

Alcian blue staining

Zebrafish embryos were fixed in 4% PFA at 96 hpf before staining with Alcian blue (Sigma, #A5268) [46]. Embryos were transferred to 30:70 glycerol/1% potassium hydroxide and subsequently to 60:40 glycerol/1% potassium hydroxide and incubated for 2–3 days until they were sufficiently translucent. Specimens were stored and analyzed in 100% glycerol. All the staining procedures were the same in each group. The vertical distance from the beginning to the end of the mandibular arch was used to quantify the mandibular length.

Whole-mount in situ hybridization

Whole-mount in situ hybridizations were performed using a standard protocol with modifications [47]. Complementary DNA probes were synthesized using a DIG RNA labeling kit (Roche, #11175025910). The primers listed below were designed by Primer 5.0 software. Whole-mount in situ hybridization of barx1 was done according to the manufacturer’s protocol. Briefly, embryos at 24 hpf were collected, fixed in 4% PFA overnight at 4 °C. After rinsing three times in PBS at room temperature, embryos were maintained in absolute methanol at −20 or −80 °C for further use. Embryos were rehydrated at gradient dilutions using methanol/PBS (75, 50, and 25%). The embryos were treated with proteinase K (10 μg/mL) at room temperature, refixed with 4% PFA for 20 min, washed three times with PBST (PBS + 0.1% Tween-20), and then prehybridized for 2 h with the hybridization solution without the probe. Subsequently, the digoxin-labeled probe was done according to the standard protocols. The following primers were used (forward/reverse): barx1 (5’-CACTCTGGTCT CCTCCGT-3’/5’-ATTCTCCGTTCTCTTCC-3’).

Statistical analysis

All experiments in this study were performed in triplicate to test the reliability of the results. Experimental values were expressed as the mean ± S.E.M. unless otherwise stated. The results in the control and experimental groups were assessed by Student’s t test or ANOVA (GraphPad Prism-6 software; San Diego, CA, USA). A P value < 0.05 was considered statistically significant.

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Author contributions: SG, YZ, DW, YW, and TZ performed the experiments and analyzed the data. JM, Y-PL, SG, and LW designed the study. JM, Y-PL, SG, YZ, and QC wrote the manuscript. All authors reviewed and revised the manuscript.

Compliance with ethical standards

Conflict of Interest The authors declare that they have no conflict of interest.

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