2013

**Oxidative Modification of Lipoic Acid by HNE in Alzheimer Disease Brain**

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Repository Citation  
Hardas, Sarita S.; Sultana, Rukhsana; Clark, Amy M.; Beckett, Tina L.; Szweda, Luke I.; Murphy, M. Paul; and Butterfield, D. Allan, "Oxidative Modification of Lipoic Acid by HNE in Alzheimer Disease Brain" (2013). *Chemistry Faculty Publications*. 6.  
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Digital Object Identifier (DOI)
http://dx.doi.org/10.1016/j.redox.2013.01.002

Notes/Citation Information
Published in Redox Biology, v. 1, no. 1, p. 80-85.

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Short Communication

Oxidative modification of lipoic acid by HNE in Alzheimer disease brain

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Keywords: Alzheimer disease, Lipid peroxidation, 4-hydroxy-2-trans nonenal (HNE), Lipoic acid, Lipoamide dehydrogenase

Abstract

Alzheimer disease (AD) is an age-related neurodegenerative disease characterized by the presence of three pathological hallmarks: synapse loss, extracellular senile plaques (SP) and intracellular neurofibrillary tangles (NFTs). The major component of SP is amyloid β-peptide (Aβ), which has been shown to induce oxidative stress. The AD brain shows increased levels of lipid peroxidation products, including 4-hydroxy-2-nonenal (HNE). HNE can react covalently with Cys, His, or Lys residues on proteins, altering structure and function of the latter. In the present study we measured the levels of the HNE-modified lipoic acid in brain of subjects with AD and age-matched controls. Lipoic acid is a key co-factor for a number of proteins including pyruvate dehydrogenase and α-ketoglutarate dehydrogenase, key complexes for cellular energetics. We observed a significant decrease in the levels of HNE-lipoic acid in the AD brain compared to that of age-matched controls. To investigate this phenomenon further, the levels and activity of lipoamide dehydrogenase (LADH) were measured in AD and control brains. Additionally, LADH activities were measured after in-vitro HNE-treatment to mice brains. Both LADH levels and activities were found to be significantly reduced in AD brain compared to age-matched control. HNE-treatment also reduced the LADH activity in mice brain. These data are consistent with a two-hit hypothesis of AD: oxidative stress leads to lipid peroxidation that, in turn, causes oxidative dysfunction of key energy-related complexes in mitochondria, triggering neurodegeneration. This study is consonant with the notion that lipoic acid supplementation could be a potential treatment for the observed loss of cellular energetics in AD and potentiate the antioxidant defense system to prevent or delay the oxidative stress in and progression of this devastating dementing disorder.

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Introduction

Oxidative stress occurs due to an imbalance in the levels of antioxidant defense systems and production of reactive oxygen/reactive nitrogen species. Oxidative stress has reported to be important in the pathophysiology of a number of age-related diseases, including Alzheimer disease (AD). AD is characterized by the presence of three principal pathological hallmarks: synapse loss, extracellular senile plaques (SP), and intracellular neurofibrillary tangles (NFTs). The major component of SP is amyloid β-peptide (Aβ), a 40–42 amino acid peptide that is derived from proteolytic cleavage of an integral membrane protein, i.e., amyloid precursor protein (APP), by the action of beta- and gamma-secretases [1,2].

Shown to induce oxidative stress, Aβ1–42 can insert as oligomers into the lipid bilayer and initiate lipid peroxidation [3–8], resulting in the formation of lipid peroxidation products including 4-hydroxy-2-nonenal (HNE), malondialdehyde, F2-isoprostanes, and 2-propen-1-al (acrolein), among others. Protein-bound HNE is one of the important markers used for studying the lipid peroxidation process, and has been known to be involved in...
depleting cellular nucleophilic compounds such as thioredoxin, glutathione, lipoic acid, etc.[5,9–13].

Protein-bound HNE as well as free HNE, TBARS, MDA, and isoprostanes (F₂-isop) levels are increased in plasma, urine, and CSF in AD and amnestic mild cognitive impairment (aMCI—arguably the earliest form of AD), compared to healthy controls [5,8,14–17]. The increased HNE formation of covalent Michael adducts account for one means of increased formation of protein carbonyls [18]. Brains from both AD and aMCI subjects show increased levels of protein carbonyls in AD in affected brain regions, while the cerebellum, largely devoid of Aβ pathology, remained relatively untouched [19]. The anti-oxidant system, both enzymatic and non-enzymatic, shows an inverse correlation with increased oxidative stress markers suggesting that free radicals are important in the progression and pathogenesis of AD [20–22].

Lipoic acid is an important co-factor for multi-enzyme complexes such as α-ketoglutarate dehydrogenase (KGDH), pyruvate dehydrogenase (PDH), branched o xo-acid or α-ketoacid dehydrogenase complex, and glycine decarboxylase complex or glycine cleavage system [23]. Lipoic acid also is unique among endogenous antioxidants, in that it can scavenge free radicals in aqueous as well as in lipid or membrane phase [23]. Lipoic acid in its endogenous form contains a disulfide bond and it remains inactive unless reduced to dihydrolipoic acid (DHLA) in a reaction catalyzed by lipoamide dehydrogenase (LADH), using NADH as the source of reducing equivalents [24]. The antioxidant and functional activity of lipoic acid as a co-factor is attributed to its reduced DHLA form [23]. The sulphydryl groups on DHLA act similar to −SH group from glutathione or cysteine and help in reduction of free radicals by providing –H or an electron. Hence, covalent modification of reduced lipoic acid by HNE could have detrimental effects on cellular energetics and the antioxidant defense system [9].

In the present study, we determined the levels of HNE-bound lipoic acid and also measured the levels and activity of the enzyme LADH in AD and age-matched control brain. In addition, we investigated the alterations in LADH enzyme activity in presence of the lipid peroxidation end product-HNE.

Materials and methods

Chemicals

All chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA) unless otherwise stated. Nitrocellulose membranes and the electrophoresis transfer system employing a Trans-Blot semi-dry transfer cell were obtained from Bio-Rad (Hercules, CA, USA).

Control and AD brains

Frozen inferior parietal lobule (IPL) specimens from ten control and twelve AD subjects were obtained from the University of Kentucky Rapid Autopsy Program of the Alzheimer’s Disease Clinical Center (UK ADC). All the subjects were longitudinally followed and underwent neurological and physical examinations and annual neuropsychological testing. The control subjects used in the current study were without history of dementia or other neurological disorders and with intact activities of daily living (ADLs).

Sample preparation

Briefly, the frozen brain tissue samples were weighed, and homogenized via polytron at 200 mg/mL in ice-cold PBS with a complete cocktail of protease inhibitors (from Amresco, in Solon, OH). An aliquot of the raw homogenate was centrifuged at 20,800 × g for 30 mins at 4 °C. The supernatants were collected and stored at − 80 °C until the time of assay, which was then used for slot blot analysis. For Western blot and enzyme assays raw homogenates were used.

HNE-bound lipoic acid (HNE-LA) levels

Levels of HNE-LA were measured by the slot blot technique as previously described with some modification in the immunochromatographic detection part of the technique [25,26]. Protein (1 µg) from AD-IPL and age matched control samples were loaded onto nitrocellulose membranes using the slot blot technique. After the transfer of proteins on the nitrocellulose membrane, the membrane was incubated in a solution of 250 mM sodium borohydride in 100 mM MOPS buffer at pH 8.0 for 15 min. Sodium borohydride chemically reduced the aldehyde moiety on HNE to an alcohol, since the primary antibody would recognize only 4-hydroxy-2-trans-nonanal bound lipoic acid. The reducing solution was prepared fresh and the pH of the MOPS was adjusted to 8.0 prior to addition of sodium borohydride. After 15 min, the membrane was washed three times with ultra pure H₂O followed by washing three times with PBS for 5 min each and blocked for 30 min in 5% milk in 1X PBS. Then the membrane was subsequently incubated with primary antibody (1:2000) followed by HRP-conjugated secondary antibody as described previously [25,26] for the detection of the HNE-LA levels.

Lipoamide dehydrogenase (LADH) levels

LADH levels were estimated by the Western blot technique as described previously [20]. Protein (75 µg) from controls and AD samples were loaded side by side on SDS-PAGE gels. Separated proteins were then transferred to the nitrocellulose membrane and detected immunochemically using the specific primary and secondary antibodies.

In-vitro HNE treatment to mice brain

Whole brains from male CD-1/129 background mice (n=4) were homogenized separately in isolation buffer (0.32 M sucrose, 2 mM EDTA, 2 mM EGTA, 20 mM HEPES, 0.2 mM PMSF, 4 µg/ml leupeptin, 4 µg/ml pepstatin, 5 µg/ml aprotinin, pH 7.4). The raw homogenates were centrifuged at 2500xg, for 10 min at 4 °C and supernatants were collected. The supernatant solutions were treated with HNE as described previously [8]. 1 mg of the supernatant from a sample was incubated with 10 µM of HNE (prepared in 10 mM, pH 8.0 PBS) and for control, another 1 mg was incubated with the same volume of PBS alone. The HNE concentration was chosen based on previous studies [8,27–31]. The incubation was carried out for 3 h at 37 °C on an orbital shaker. After the incubation, control and HNE-treated samples were centrifuged at 10,000 × g for 10 min at 4 °C. The resultant pellets were separately washed with sucrose buffer three times and finally suspended in ~150 µL of the same sucrose buffer. Protein estimation was performed prior to measuring LADH enzyme activity and HNE-LA levels (as described above) in these samples.

Lipoamide dehydrogenase activity assay

The enzymatic activity of LADH (EC 1.8.1.4) was measured by decreased absorbance caused by oxidation of NADH. The assay protocol was provided by Sigma-Aldrich. In brief, a mixture of 2% bovine serum albumin with 30 mM EDTA, 2 mM NAD+ , 1.5 mM NADH disodium salt, along with 20 mM lipoic acid as a substrate was
added to 20 μg of sample. Sodium phosphate buffer (50 mM, pH 6.5) was used as an assay buffer. The rate of the reaction was monitored spectrophotometrically at 340 nm wavelength for 10 min.

Statistical and data analysis

All the data obtained from different biochemical assays were subjected to statistical analysis. Significant difference in AD-IPL and age-matched control samples as well as HNE-treated and age-matched controls was determined by employing the Student’s unpaired t-test with significance at p < 0.05.

Results

The levels of HNE-bound lipoic acid (HNE-LA) were detected in the IPL region of AD and age-matched control brain samples using a specialized antibody against HNE-LA modification (produced in Dr. Luke Szweda’s laboratory). Significantly decreased HNE-LA levels in AD samples were observed (Fig. 1-left). Previous studies from our laboratory and other showed that HNE levels are elevated in AD, and the oxidative modification of the proteins affects the structure and function of proteins [3,8,17,32,33]. HNE can react only with the reduced form of lipoic acid-DHLA [34], and the enzyme responsible for reduction of LA to DHLA is lipoamide dehydrogenase. Therefore, to test the hypothesis that reduced levels of HNE bound to lipoic acid could be due to alterations in lipoamide dehydrogenase, we measured activities and levels of LADH in AD and control IPL brain region. Both the levels and activities of LADH were significantly reduced in AD-IPL compared to age-matched controls (Fig. 1-center, Fig. 2-left), consistent with the notion that LADH has decreased ability to reduce LA in AD brain. To investigate further if HNE itself can inhibit enzyme activity of LADH, an in-vitro HNE treatment was performed using mice brain homogenates. LADH enzyme activity in HNE-treated mice brain samples was significantly decreased compared to their age-matched controls (Fig. 2-right), consistent with the notion that HNE can covalently modify LADH. However, HNE-LA levels were increased in mice brain after HNE-treatment (Fig. 1-right).

Discussion

Glucose metabolism is essential for proper brain function; a minimum interruption of glucose metabolism causes brain dysfunction and memory loss [35]. Positron emission tomography [36] scanning shows a consistent pattern of reduced cerebral glucose utilization in AD brain [37]. Mitochondria are a main source for free radical production and are also critically important for production of the cellular currency, ATP. AD brain shows abnormal mitochondrial morphology and impaired mitochondrial energy metabolism [38–43]. A number of studies suggest that the rate of cerebral metabolism is reduced in AD, MCI and early AD [44–47], and redox proteomics studies from our laboratory identified a large number of glycolytic, TCA, and mitochondrial enzymes as oxidatively dysfunctional [48]. A recent study employing a triple transgenic mouse model of AD showed that mitochondrial bioenergetics deficits precede AD pathology [49].

Decreased cerebral energy metabolism correlates with the altered expression and decreased activity of mitochondrial energy-related proteins such as the pyruvate dehydrogenase complex, and alpha-ketoglutarate dehydrogenase complex (KGDHC). As mentioned earlier, both these proteins have lipoic acid as a co-factor that is critical for proper function of the protein. The modification of lipoic acid by HNE would likely lead to altered energy metabolism, consequently leading to decreased levels of cellular ATP at nerve terminals promoting loss of synapses and synaptic function. Detrimental downstream consequences of ATP loss include decreased propagation of action potentials, decreased long-term potentiation, and loss of memory and learning, likely contributing to these clinical observations exhibited by AD and MCI patients. As noted above, synapse loss is considered an early feature of AD progression.

The treatment of rat mitochondria with HNE leads to selective inhibition of KGDH and PDH leading to the loss of activity. These results were echoed when purified forms of KGDH and PDH were treated with HNE suggesting that HNE modification of lipoic acid might contribute to their reduced activity [34]. Further, Korotchkin...
et al. [50] showed that HNE inactivates KGDH and PDH in human HepG2 cells. However, successive treatment with lipoic acid and other thiol supplements protected these complexes from HNE modification. Further, the same study reported that inhibition of PDH was observed at its subunit E2E3-BP. More extensive damage was in the subunit E3-dihydrodipioamide dehydrogenase (reduced form of LADH) when NADH was present and sulfhydryl groups on lipoic acid were in the reduced form [50].

Similar to lipoic acid, LADH is also shared as a subunit E3 by four multi-enzyme complexes: KGDH, PDH, the branched oxo acid or α-ketoacid dehydrogenase complex, and the glycine decarboxylase complex or glycine cleavage system [51–53]. Inhibition of the levels and activity of LADH may reflect the loss of enzymatic activity of these multi-enzyme complexes in AD. Our results are consonant with previous findings of decreased LADH levels and activity in AD brain [53]. Furthermore, decreased LADH activity after in-vitro HNE-treatment suggests oxidative inhibition of LADH can be induced by HNE as observed previously in case of other TCA enzymes [34]. There is substantial evidence that alterations in the enzymes related to mitochondrial energetics play a role in pathogenesis of neurodegenerative diseases [54]. Studies with caucasians [55] and Ashkenazi Jewish populations [56] have associated the gene for LADH with reduced risk of AD. In other research, mice deficient in LADH had altered vulnerability to the dopaminergic neurotransmitter MPTP used in Parkinson disease models and to malonate and 3-nitropropionic acid, which have been used in models of Huntington disease [36]. However, post HNE-treatment increased HNE-LA levels are contrary to our observation in AD brain. While we do not have a definitive explanation for this observation, several differences in mouse vs. human brain may be involved, three of which may include: (a) involvement of multiple signaling pathways that could affect HNE-LA levels; (b) involvement of some but not all lipoic acid dependent enzyme complexes in mouse brain following HNE addition, which would lead to elevated HNE-LA; and/or (c) because of the extensive oxidative stress present in AD brain a less reductive environment in AD brain exists than in mouse brain, the latter perhaps permitting more disulfide bond breakage of LA via means other than LADH with consequent elevated HNE-LA. Such possibilities are the subject of future studies in our laboratory.

In AD, oxidative stress can alter the functions of TCA enzymes as well as induce intracellular accumulation of calcium, which may lead to cellular death [33,57,58]. Dysfunction of TCA enzymes may compromise cellular energetics and elevate oxidative stress further [55,59]. If not intervened, this feed-forward loop may accelerate the progression of the disease. Various studies have proposed using antioxidant therapeutics to prolong the time of onset of AD or retard the rate of its progression [60–65]. Treatment with lipoic acid reduced the lipid peroxidation marker-HNE and apoptotic markers in AD fibroblasts cultures [66]. In the same study, co-treatment of cells with N-acetyl cysteine showed much better protection from oxidative stress than lipoic acid or N-acetyl cysteine alone [66]. Apart from acting as an antioxidant itself, lipoic acid in the reduced form can also reinforce other water-or lipid-soluble antioxidants such as glutathione, ascorbate, and vitamin E by scavenging their radicals [23]. A previous study from our laboratory showed that pretreatment of cortical neurons with acetyl l-carnitine and lipoic acid protected cells against HNE-mediated oxidative stress and neurotoxicity by inducing GSH levels and reducing protein and lipid oxidation and inducing protective pathways involving ERK1/2 [67]. The current studies have provided a rationale (Fig. 3) for studies underway in our laboratory to better understand the mechanism(s) and develop therapeutic(s) to prevent Aβ-induced oxidative stress and neurotoxicity, relevant to slowing or inhibiting of the progression and pathogenesis of AD.

Finally, the results of this paper fit the two-hit hypothesis of AD [68–72], namely that oxidative stress may combine with other events to push the patient to AD. In this case, the well-known oxidative stress in AD, which occurs early in the disease, i.e., at the aMCI stage if not earlier [3,4], could lead to loss of ATP production by oxidatively modifying numerous enzymes of the glycolytic and TCA cycles and the ETC, among which is LADH. The consequent decreased levels of DHLA would decrease ATP production, while simultaneously having a less reduced environment with consequent more free radical oxidative stress. The resulting loss of neurons could then contribute to the clinical presentation, progression, and pathology of AD. Studies to test this idea are underway in our laboratory.

**Fig. 3.** The NADH-dependent oxido-reductase enzyme lipoamide dehydrogenase (LADH) is also an important member of the mitochondrial energy generation complex. Alteration of the structure and activity of LADH by elevated reactive oxygen species (ROS) may hamper energy metabolism and ATP production. Lipoic acid (LA) must be in the reduced form as part of its co-factor function for mitochondrial TCA complexes such as α-ketoglutarate dehydrogenase. However, oxidized LADH is unable to reduce LA to DHLA, and therefore HNE is unable to bind to DHLA efficiently. Consequently, in AD brain decreased LA-HNE binding was observed. Severe effects on learning, memory, and higher executive functioning, all significantly lost in AD patients would be expected. Supplementation of LA conceivably may protect LADH from ROS or end products of ROS (e.g., HNE) by self-sacrifice mechanism, potentially providing protection against dementia or slowing the rate of progression of AD.

**Conflict of interest statement**

No conflict of interest exists for any of the authors.

**Acknowledgments**

This work was supported, in part, by a NIH grant to D. A. Butterfield (AG-05119). We are grateful to the Neuropathology Core of the University of Kentucky Alzheimer Disease Clinical Center for providing us well-characterized specimens to carry out this research.
References

[1] C. Duyckaerts, R. Delatorre, M.C. Potier, Classification and basic pathology of Alzheimer disease, Acta Neuropathology 118 (1) (2009) 5–36.

[2] I. Grundke-Iqbal, K. Iqbal, M. Quinlan, Y.C. Tung, M.S. Zaidi, H.M. Wisniewski, Microtubule-associated protein tau. A component of Alzheimer paired helical filaments, Journal of Biological Chemistry 261 (13) (1986) 6084–6089.

[3] W.R. Markesbery, Oxidative stress hypothesis in Alzheimer’s disease, Free Radical Biology and Medicine 23 (1) (1997) 134–147.

[4] D.A. Butterfield, J. Drake, C. Pocernich, A. Castegna, Evidence of oxidative damage in Alzheimer’s disease brain: central role for amyloid beta-peptide, Trends in Molecular Medicine 7 (12) (2001) 548–554.

[5] D.A. Butterfield, A. Castegna, C.M. Lauderback, J. Drake, Evidence that amyloid beta-peptide-induced lipid peroxidation and its sequelae in Alzheimer’s disease brain contribute to neuronal death, Neurobiology of Aging 23 (5) (2002) 655–664.

[6] R.J. Mark, M.A. Lovell, W.R. Markesbery, K. Uchida, M.P. Mattson, A role for 4-hydroxynonenal, an aldehyde product of lipid peroxidation, in disruption of ion homeostasis and neuronal death induced by amyloid beta-peptide, Journal of Neurochemistry 68 (1) (1997) 255–264.

[7] M.P. Mattson, Cellular actions of beta-amyloid precursor protein and its soluble and fibrilligenic derivatives, Physiological Reviews 77 (4) (1997) 1081–1132.

[8] C.M. Lauderback, J.M. Hackett, F.F. Huang, J.N. Keller, L.L. Szwed, W.R. Markesbery, et al., The glial glutamate transporter, GLT-1, is oxidatively modified by 4-hydroxy-2-nonenal in the Alzheimer’s disease brain: the role of a beta-1-42, Journal of Neurochemistry 78 (2) (2001) 413–416.

[9] A.N. Higdon, A. Landar, S. Barnes, V.M. Darley-Usmar, The electrophile-responsive pro tease: integrating proteomics and lipiddomics with cellular function, Antioxidants and Redox Signaling 17 (11) (2012) 1580–1589.

[10] C.F. Mello, R. Sultana, M. Piroddi, J. Cai, W.M. Pierce, J.B. Klein, et al., Acrolein induces selective protein carbonylation in synaptosomes, Toxicology 204 (2–3) (2004) 209–218.

[11] J.N. Keller, F.R. Schmitt, S.W. Scheff, Q. Ding, Q. Chen, D.A. Butterfield, et al., Evidence of increased oxidative damage in subjects with mild cognitive impairment, Neurology 64 (7) (2005) 1152–1156.

[12] Y. Yao, C.M. Clark, J.Q. Trojanowski, V.M. Lee, D. Pratico, Elevation of 12/15 lipoxigenase products in AD and mild cognitive impairment, Annals of Neurology 58 (4) (2005) 623–626.

[13] M.A. Lovell, W.D. Ehmann, M.P. Mattson, W.R. Markesbery, Elevated 4-hydroxynonenal, a lipid peroxidation product in the Alzheimer’s disease brain, Neurobiology of Aging 18 (5) (1997) 457–461.

[14] W.R. Markesbery, M.A. Lovell, Four-hydroxynonenal, a product of lipid peroxidation, is increased in the brain in Alzheimer’s disease, Neurobiology of Aging 19 (1) (1998) 17–26.

[15] D.A. Butterfield, E.R. Stadman, Protein Oxidation Processes in Aging Brain, in: M.A. Lovell, W.D. Ehmann, M.P. Mattson, W.R. Markesbery, Elevated 4-hydroxynonenal and pyruvate dehydrogenase: reaction of lipoyc acid with 4-hydrox-2-nonenal, Biochemistry 37 (45) (1998) 15835–15841.

[16] W. Weser-Ruge, C. Bertoni-Freddari, P. Iwangoff, Changes in brain glucose metabolism as a key to the pathogenesis of Alzheimer’s disease, Gerontology 40 (5) (1994) 246–252.

[17] P. Klivenyi, A.A. Starkov, N.Y. Calingasan, G. Gardian, S.E. Brownne, L. Yang, et al., Mice deficient in dihydrodipicolinate dehydrogenase show increased vulnerability to MPTP and 3-nitropropionic acid neurotoxicity, Journal of Neurochemistry 88 (6) (2004) 1352–1360.

[18] S. Hoyer, Glucose metabolism and insulin receptor signal transduction in Alzheimer disease, European Journal of Pharmacology 490 (1-3) (2004) 115–125.

[19] M. Manzack, B.S. Park, Y. Jung, P.H. Reddy, Differential expression of oxidative phosphorylation genes in patients with Alzheimer’s disease: implications for early mitochondrial dysfunction and oxidative damage, NeuroMolecular Medicine 5 (2) (2004) 122–131.

[20] X. Wang, B. Su, S.L. Siedlak, P.I. Moreira, H. Fujikoa, Y. Wang, et al., Amyloid-beta overproduction causes abnormal mitochondrial dynamics via differential modulation of mitochondrial fission/fusion proteins, Proceedings of the National Academy of Sciences USA 105 (48) (2008) 19318–19323.

[21] A.M. Aleardi, G. Benard, O. Augereau, M. Malgat, J.C. Talbot, J.P. Mazet, et al., Gradual alteration of mitochondrial structure and function by beta-amyloids: importance of membrane viscosity changes, energy deprivation, reactive oxygen species production, and cytochrome c release, Journal of Bioenergetics and Biomembranes 37 (4) (2005) 207–225.

[22] E. Perez-Gracia, B. Torrejon-Escribano, I. Ferrer, Dystrophic neurites of senile plaques in Alzheimer’s disease are deficient in cytochrome c oxidase, Acta Neuropathologica 116 (3) (2008) 261–268.

[23] S.H. Kim, R. Vilikonis, N. Cairns, M. Fountoulakis, G. Lubec, The reduction of NADH ubiquinone oxidoreductase 24- and 75-kDa subunits in brains of patients with Down syndrome and Alzheimer’s disease, Life Science 68 (24) (2001) 2741–2750.

[24] R. Sultana, D.A. Butterfield, Oxidatively modified, mitochondria-relevant brain proteins in subjects with Alzheimer disease and mild cognitive impairment, Journal of Bioenergetics and Biomembranes 41 (5) (2009) 441–446.

[25] L. Mosconi, Brain glucose metabolism in the early and specific diagnosis of Alzheimer’s disease, FDG-PET studies in MCI and AD, European Journal of Nuclear Medicine and Molecular Imaging 31 (4) (2005) 840–510.

[26] S. Damsbo, C. Lindbo, O.C. Toft, A. Christensen, Brain energy metabolism, cognitive function and down-regulated oxidative phosphorylation in Alzheimer disease, Neurogerentation 5 (4) (1996) 473–476.

[27] H. Attems, W.H. Frey 2nd, Mechanisms of mitochondrial dysfunction and energy deficiency in Alzheimer’s disease, Mitochondrion 7 (5) (2007) 297–310.

[28] L. Mosconi, A. Pupi, M.J. De Leon, Brain glucose hypometabolism and oxidative stress precede clinical Alzheimer’s disease, Annals of the New York Academy of Sciences 1147 (2008) 180–195.

[29] D.A. Butterfield, M. Perlugio, T. Reed, T. Muharib, C.P. Hughes, R.A. Robinson, et al., Redox proteomics in selected neuromdegenerative disorders: from its discovery to future applications, Antioxidants and Redox Signaling 17 (17) (2012) 1610–1655.

[30] J.J. Yee, R.W. Irwin, L. Zhai, J. Nisen, R.T. Hamilton, R.D. Brighton, Mitochondrial bioenergetic deficit precedes Alzheimer’s pathology in female mouse model of Alzheimer’s disease, Proceedings of the National Academy of Sciences USA 108 (34) (2009) 14670–14675.

[31] L.G. Korotchkina, H.-S. Yang, O. Tirosh, L. Packer, M.S. Patel, Protection by thiols of the mitochondrial complexes from 4-hydroxy-2-nonenal, Free Radical Biology and Medicine 51 (1) (2016) 992–999.

[32] J.G. Wiltfang, Targeting of 2-oxo acid dehydrogenase complexes to the mitochondria, Annals of the New York Academy of Sciences 573 (1) (1989) 254–266.
A. Hunter, J.G. Lindsay, Immunological and biosynthetic studies on the mammalian 2-oxoglutarate dehydrogenase multienzyme complex, European Journal of Biochemistry 155 (1) (1986) 103–109.

F. Mastrogiacomo, J.G. Lindsay, L. Bettendorf, J. Rice, S.J. Kish, Brain protein and α-ketoglutarate dehydrogenase complex activity in Alzheimer’s disease, Annals of Neurology 39 (5) (1996) 592–598.

G.E. Gibson, L.C.H. Park, K.-F.R. Sheu, J.P. Blass, N.Y. Calingasan, The α-ketoglutarate dehydrogenase complex in neurodegeneration, Neurochemistry International 36 (2) (2000) 97–112.

K.-F.R. Sheu, A.J.L. Cooper, K. Koike, M. Koike, J.G. Lindsay, J.P. Blass, Abnormality of the α-ketoglutarate dehydrogenase complex in fibroblasts from familial Alzheimer’s disease, Annals of Neurology 35 (3) (1994) 312–318.

A.M. Brown, D. Gordon, H. Lee, M. Caudy, J. Hardy, V. Haroutunian, et al., Association of the dihydrolipoamide dehydrogenase gene with Alzheimer’s disease in an Ashkenazi Jewish population, American Journal of Medical Genetics Part B: Neuropsychiatric Genetics 131B (1) (2004) 60–66.

W. Fu, H. Luo, S. Parthasarathy, M.P. Mattson, Catecholamines potentiate amyloid β-peptide neurotoxicity: involvement of oxidative stress, mitochondrial dysfunction, and perturbed calcium homeostasis, Neurobiology of Disease 5 (4) (1998) 229–243.

G.E. Gibson, H. Zhang, H. Xu, L.C.H. Park, T.M. Jeitner, Oxidative stress increases internal calcium stores and reduces a key mitochondrial enzyme, Biochimica Biophysica Acta 1502 (1) (2000) 139–144.

M.A. Smith, C.A. Rottkamp, A. Nunomura, A.K. Raina, G. Perry, Oxidative stress in Alzheimer’s disease, Biochimica Biophysica Acta 1502 (1) (2000) 139–144.

W.O. Opip, G. Joshi, E. Head, N.W. Milgram, B.A. Muggenburg, J.B. Klein, et al., Proteomic identification of brain proteins in the canine model of human aging following a long-term treatment with antioxidants and a program of behavioral enrichment: relevance to Alzheimer’s disease, Neurobiology of Aging 29 (1) (2008) 51–70.

C. Mancuso, T.E. Bates, D.A. Butterfield, S. Calafato, C. Cornelius, A.D. Lorenzo, et al., Natural antioxidants in Alzheimer’s disease, Expert Opinion on Orphan Drugs 16 (12) (2007) 1921–1931.

S.M. Yatin, S. Varadarajan, D.A. Butterfield, Vitamin E prevents Alzheimer’s amyloid β-peptide (1–42)-induced neuronal protein oxidation and reactive oxygen species production, Journal of Alzheimer’s Disease 2 (2) (2000) 123–131.

C. Behl, B. Moosmann, Antioxidant neuroprotection in Alzheimer’s disease as preventive and therapeutic approach, Free Radical Biology and Medicine 33 (2) (2002) 182–191.

D. Pratico, Evidence of oxidative stress in Alzheimer’s disease brain and antioxidant therapy, Annals of the New York Academy of Sciences 1147 (1) (2008) 70–78.

S.S. Pitchumoni, P.M. Doraiswamy, Current status of antioxidant therapy for Alzheimer’s Disease, Journal of the American Geriatrics Society 46 (12) (1998) 1566–1572.

P.I. Moreira, P.L.R. Harris, X. Zhu, M.S. Santos, C.R. Oliveira, M.A. Smith, et al., Lipidic acid and N-acetyl cysteine decrease mitochondrial-related oxidative stress in Alzheimer disease patient fibroblasts, Journal of Alzheimer Disease 12 (2) (2007) 195–206.

M.A. Hafiz, D.A. Butterfield, Involvement of PI3K/PKG/ERK1/2 signaling pathways in cortical neurons to trigger protection by cotreatment of acetyl-L-carnitine and α-lipoic acid against HNE-mediated oxidative stress and neurotoxicity: Implications for Alzheimer’s disease, Free Radical Biology and Medicine 42 (3) (2007) 371–384.

X. Zhu, H.-g Lee, G. Perry, M.A. Smith, Alzheimer disease, the two-hit hypothesis: an update, Biochimica Biophysica Acta 1772 (4) (2007) 494–502.

V.P. Reddy, X. Zhu, G. Perry, M.A. Smith, Oxidative stress in diabetes and Alzheimer’s disease, Journal of Alzheimer Disease 16 (4) (2009) 763–774.

Z. Nagy, The last neuronal division: a unifying hypothesis for the pathogenesis of Alzheimer’s disease, Journal of Cellular and Molecular Medicine 9 (3) (2005) 531–541.

X. Zhu, R.J. Castellani, A. Takeda, A. Nunomura, C.S. Atwood, G. Perry, et al., Differential activation of neuronal ERK, JNK/SAPK and p38 in Alzheimer disease: the ‘two hit’ hypothesis, Mechanisms of Ageing and Development 123 (1) (2001) 39–46.

X. Zhu, A.K. Raina, G. Perry, M.A. Smith, Alzheimer’s disease: the two-hit hypothesis, Lancet Neurology 3 (4) (2004) 219–226.