Transfection of Endothelial Nitric Oxide Synthase Gene Improves Angiogenic Efficacy of Endothelial Progenitor Cells in Rabbits with Hindlimb Ischemia

Savneet Kaur 1,6, VS Harikrishnan 2, Sachin J Shenoy 2, NS Radhakrishnan 3, Akira Ur uno 4, Akira Sugawara 4 and Chandrasekharan C Kartha 5*

1Division of Cellular and Molecular Cardiology, Sree Chitra Tirunal Institute for Medical Sciences and Technology, Thiruvananthapuram 695011, India
2Biomedical Technology Wing, Sree Chitra Tirunal Institute for Medical Sciences and Technology, Thiruvananthapuram 695014, India
3Division of Pathology, Sree Chitra Tirunal Institute for Medical Sciences and Technology, Thiruvananthapuram 695011, India
4Department of Medicine, Tohoku University Graduate School of Medicine, Sendai 980-8574, Japan
5Disease Biology and Molecular Medicine, Rajiv Gandhi Center for Biotechnology, Thiruvananthapuram 695014, India
6School of Biotechnology, Gautam Buddha University, Greater Noida, UP- 201308, India

Abstract

Background: The present study explored the effect of endothelial nitric oxide synthase (eNOS) gene transfer on the angiogenic potential of ex vivo expanded endothelial progenitor cells (EPCs) in a rabbit model of hindlimb ischemia.

Methods: Rabbit peripheral blood EPCs were cultured and transfected with mammalian expression vector pcDNA3.1-eNOS containing full-length human eNOS gene. Ischemia was induced in the right hind limb of three groups of rabbits by ligation of the distal external iliac artery and excision of the common and superficial femoral arteries. In one group of animals, ten days after the surgery, autologous eNOS-EPCs were transplanted intramuscularly in the ischemic limb. Two other groups received an equivalent number of unmodified EPCs or phosphate buffered saline (PBS) respectively.

Results: Two weeks after cell transplantation, the in vivo expression of eNOS was detected in limb tissue sections of eNOS-EPCs treated animals. Animals treated with eNOS-EPCs had a significant reduction in ischemic muscle necrosis and inflammation, augmentation in the capillary density (P< 0.05) and angiographic scores demonstrating distal arterial reconstitution and enhanced angiogenesis in comparison to animals transplanted with EPCs or PBS (P< 0.05).

Conclusion: We conclude that modification of EPCs by eNOS constitutes an effective strategy to improve the efficacy of EPCs for therapeutic angiogenesis.

Introduction

Therapeutic angiogenesis represents an innovative strategy to treat vascular insufficiency. It is aimed at improving the compensatory mechanisms of angiogenesis and arteriogenesis by stimulating the growth and development of collateral vessels in critical peripheral or myocardial ischemia. Bone marrow-derived endothelial progenitor cells (EPCs) harvested from peripheral blood have been recently identified for their contribution to angiogenesis and revascularization of ischemic tissues [1]. EPCs proliferate and migrate in response to angiogenic growth factors and differentiate into mature endothelial cells in situ for blood vessel formation [2]. In recent years, transplantation of ex vivo expanded autologous EPCs has emerged as a novel therapeutic strategy that could provide a robust source of viable endothelial cells to supplement the contribution of endothelial cells resident in the adult vasculature. The regenerative potential of ex vivo expanded EPCs has been well demonstrated in animal models of limb and myocardial ischemia [3-7].

Despite the promising applications for tissue regeneration, the limited endogenous pool, the possible functional impairment associated with a variety of physiological and pathological phenotypes of clinical patients, and the limited replicative potential of EPCs for ex vivo expansion, largely impede their use for autologous transplantation [8, 9]. Given these findings, genetic modification of EPCs to over express angiogenic growth factors, enhance signaling activity of the angiogenic response, rejuvenate the bioactivity, and/or extend the life span of EPCs constitute various potential strategies that might address the limitations of EPC transplantation and thereby optimize therapeutic angiogenesis [10]. Recent studies have employed genetic modification strategies of EPCs with vascular endothelial growth factor (VEGF), telomerase reverse transcriptase, fibroblast growth factor-1 and stromal-derived factor-1 alpha genes to enhance their vasculogenic function [11-14].

Endothelial nitric oxide synthase (eNOS) has been shown to elicit multiple beneficial effects within the cardiovascular system. In addition to its vaso-dilatory properties, eNOS-derived nitric oxide (NO) has been reported to play a critical role in angiogenesis and arteriogenesis in response to tissue ischemia [15-17]. In our previous in vitro study, we demonstrated enhanced migration, differentiation and angiogenic potential of EPCs over expressing eNOS [18]. Accordingly, in the present study, we further evaluated the efficacy of genetically modified autologous EPCs over expressing eNOS vis-à-vis unmodified EPCs in inducing angiogenesis in a rabbit model of experimentally induced hind limb ischemia. The study indicates that eNOS modified EPCs show

*Corresponding author: Dr. Chandrasekharan C Kartha, Department of Disease Biology & Molecular Medicine, Rajiv Gandhi Center for Biotechnology, Thiruvananthapuram 695014, India, Tel: 91- 471-2529448; Fax: 91- 471-2348086; E-mail: cckartha@yahoo.com

Received May 26, 2011; Accepted July 06, 2011; Published July 11, 2011

Citation: Kaur S, Harikrishnan VS, Shenoy S J, Radhakrishnan NS, Ur uno A, et al. (2011) Transfection of Endothelial Nitric Oxide Synthase Gene Improves Angiogenic Efficacy of Endothelial Progenitor Cells in Rabbits with Hindlimb Ischemia. J Clinic Experiment Cardiol 2:140. doi:10.4172/2155-9880.1000140

Copyright: © 2011 Kaur S, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.
enhanced angiogenic properties than their unmodified counterparts in the ischemic animals.

Materials and Methods

Rabbit hind limb ischemia model

In total, twenty-eight New Zealand White rabbits of either sex weighing 2.2-3.1 kg were studied. Food and water were provided ad libitum. Ten rabbits were used for the isolation and characterization of EPCs. In the remaining, unilateral hindlimb vascular insufficiency was induced surgically by previously described methods [19]. Briefly, rabbits were anesthetized with intramuscular (in) injections of 0.5mg/kg atropine, 5mg/kg xylazine and 50mg/kg ketamine. Under sterile surgical conditions, a longitudinal incision was made on the medial thigh of one hind limb, extending from the inguinal ligament to a point just proximal to the patella. After the skin incision, the entire femoral artery and all its major branches including the inferior epigastria, deep femoral, lateral circumflex and superficial epigastria arteries were dissected free. The external iliac artery and all of the arteries listed were ligated. Finally, the femoral artery was excised from its proximal origin as a branch of the external iliac artery, to the point distally where it bifurcates into the saphenous and popliteal arteries. The incision was closed in three layers with 2.0 braided silk. All rabbits were closely monitored by the veterinary staff and received in injections of ampicillin-clavulanic acid [10mg/kg body weight twice daily] and paracetamol [10mg/kg body weight once daily for 5 days]. EPCs for treatment were given after 10 days of surgery. Two weeks after cell transplantation, the rabbits were taken for angiogram analysis and thereafter euthanized with a single intravenous overdose of 1% sodium thiopentone and muscle tissue was extracted from the ischemic limb. The mid-portions of the tissue sections were placed in phosphate buffered saline (PBS), mounted in cross section in OCT compound (Leica) and snap-frozen in liquid nitrogen. Cryostat sections (5µm) were prepared on microscope slides for histological analysis. The remaining tissue was snap-frozen in liquid nitrogen for RNA extraction. All animals have received humane care in compliance with the international standards on animal welfare and Guide for the Care and Use of Laboratory Animals of the Institutional Animal Ethics Committee.

Assessment of limb necrosis

All the rabbits were clinically evaluated for incidence of muscle atrophy and distal limb necrosis/ulcers by macroscopic examination at day 10 post-surgery (before treatment) and at day 20 post-surgery (after treatment). Muscle atrophy and necrosis was also evaluated histologically by staining the ischemic muscle tissue sections by hematoxylin and eosin staining at day 20 post-surgery in all the animal groups.

EPC isolation and characterization

Samples of peripheral blood (12-15 ml) from the central auricular artery of rabbits were harvested 1 day before surgery and anti-coagulated with heparin. The isolation and culture of EPCs was done as previously reported [5]. Briefly, peripheral blood mononuclear cells (PBMCs) were isolated by density centrifugation (Histopaque 1077, Sigma-Aldrich, USA) and after purification with PBS and RBC lysis, 1 x 10^7 cells/cm^2 were seeded on fibronectin-coated, 6-well plates (Nunc) in M-199 (Sigma-Aldrich, USA), 20% FBS (Sigma-Aldrich, USA), 10 ng/ml vascular endothelial growth factor (VEGF) (US Biologicals, USA) and antibiotics, 100 U/ml benzyl penicillin and 100 µg/ml streptomycin. Medium change was done every three days.

The identity of EPC-CFUs at day 7 was confirmed by immunocytochemistry with 1:50 dilution of monoclonal anti human von Willebrand factor (vWF, Sigma-Aldrich, USA), and anti-human platelet endothelial cell adhesion molecule (PECAM/CD31,Dako,Germany) as previously described [20]. After incubation with the primary antibodies, cells were incubated with 1:500 dilution of anti-mouse IgG rhodamine conjugate (ProteinProbes, Netherlands) and were counterstained for 5 minutes with 10 µg of nuclear dye Hoechst 33342 (Sigma-Aldrich, USA). Functionality and viability of EPCs were assessed by 1,1'-dioctadecyl-3,3',3-tetramethyl-indocarbocyanine perchlorate labeled acetylated low-density lipoprotein (DiI-ac-LDL, Invitrogen, USA) uptake and lectin (UEA-1, Sigma-Aldrich, USA) staining after 7 days in culture [8]. The stained cells were visualized under an inverted fluorescent microscope.

eNOS gene transfer in EPCs

Human full-length wild-type eNOS cDNA (3.6 kb) was isolated by RT-PCR from the total RNAs of human dermal microvascular endothelial cells with the use of specific primers (forward: 5'-CCCAAAGCTTACATGGGGAAGAGGACGGT-3'; reverse: 5'-GGAATTCCAGGGGCTGTGTGGTCTGTA-3') [18]. The isolated cDNA was confirmed to be identical to the previously reported clone (NM_000603) by sequencing and was sub-cloned into pcDNA3.1 expression vector (Invitrogen, USA) (pcDNA3-eNOS). After characterization, rabbit EPCs after 7 days were transfected with 2 µg of plasmid DNA, pcDNA3.1-eNOS using lipofectamine and plus reagent in serum-free OptiMEM medium (Sigma-Aldrich, USA) as per the manufacturer’s instructions for six hours (Sigma-Aldrich, USA). Six hours after transfection, medium containing M199 + 10% FBS + VEGF was added. EPCs were transplanted in the rabbits after three days of transfection.

Treatment groups and EPC transplantation

After femoral artery excision, the rabbits were randomly assigned to one of the three groups: (a) control group (receiving PBS, n = 6); (b) EPC group (receiving cultured EPCs, n = 6); (c) eNOS-EPC group (receiving cultured eNOS-EPCs, n = 6). At day 10, post-surgery, EPCs or eNOS-EPCs in culture were trypsinized (0.05% trypsin-0.02% EDTA) and collected for transplantation. Cells in 600µL of PBS were injected in into 5-6 different sites of the ischemic thigh skeletal muscles with a 22-gauge needle. The control group received 600 µL PBS. Overall, a mean of 1 x 10^5±0.35 x 10^5 EPC were injected per rabbit.

Tissue procurement, histological section preparation, and mRNA extraction

Muscle samples were weighed and total RNA was extracted with TRIzol reagent (Invitrogen, USA) as per the instructions. RNA concentration was determined by spectrophotometry at 260 nm and 1 µg RNA was reverse transcribed into complementary DNA (cDNA) using MMLV Reverse transcriptase (Promega, USA). The transcribed cDNA, 500 ng was used for the PCR to detect the expression of human eNOS transgene. The RT-PCR primer sequence and annealing temperature used were as reported earlier: endothelial nitric oxide synthase, forward: 5'-AAGATCTCCCGCTCGCTCA-3'; reverse: 5'-GCTGTAGAAGCGGATCTTAA-3'; product size: 336 bp and annealing temp: 58°C [20].

Immunohistochemistry

Immunohistochemical detection of human eNOS in the tissue sections was performed by incubating 5µm-thick frozen sections in
1:100 dilution of monoclonal anti-human eNOS antibody (Sigma-aldrich, USA). Sections were then incubated in 1:200 dilution of FITC-labeled anti-mouse secondary antibody (Molecular probes, Netherlands) for 30 min in dark. The slides were washed, mounted with 50% glycerol and examined under a fluorescence microscope. Some of the cryosections were also stained with hematoxylin and visualized under a phase contrast microscope.

Measurement of nitrite levels

The NO production in the tissue homogenates was assessed by measuring the levels of nitrite by a one-step colorimetric assay using modified Griess reagent (Sigma-aldrich, USA). Nitrite concentration (nmoles/ml) was determined using a calibration curve with sodium nitrite standards.

Analysis of capillary density

The collateral micro-vascular formation was evaluated by measuring the number of CD31 positive capillaries in two muscle tissue sections from each animal. Frozen sections were stained with antibody to CD31 followed by incubation with 1:500 dilution of biotinylated secondary antibody (Novocastra, UK). Sections were treated further with an avidin-biotin conjugated to horseradish peroxidase, developed with DAB (Novocastra, UK), and counterstained with hematoxylin. The number of capillaries was counted under a 20X objective lens using an image analysis system (Image Pro-plus 5.1, Media Cybernetics, Inc) in 5 randomly selected fields. To ensure that capillary densities were not overestimated as a consequence of myocyte atrophy or underestimated because of interstitial edema, the capillary/muscle fiber ratio was determined.

Angiographic analysis

The animals were pre-medicated with atropine [0.05mg/kg body weight (Bwt)] and xylazine [5mg/kg Bwt]. Ketamine [50 mg/kg Bwt] was used to induce the anesthesia. When the animals were under deep surgical anesthesia, under aseptic conditions, with a mid-ventral incision, laparotomy was performed and the abdominal aorta was exposed below the level of kidneys. Afterlooping the aorta with umbilical tape, arterial cannulation was done using 20G i/v cannula with injection valve (Medifon, Eastern Medikit Ltd., India) and the cannula was secured at site with braided silk sutures. Aorta was ligated anteriorly to prevent the back flow of the dye while the angiography was performed. The animal was positioned on its back (dorsoventral positioning) to prevent the back flow of the dye while the angiography was performed. The ischemic limbs of the eNOS-EPC treated groups showed complete absence of muscle atrophy and tissue necrosis at day 20 post-surgery. Histological studies of the ischemic muscle sections at day 20 post-surgery also revealed evidence of severe inflammation, necrosis and atrophy in the PBS treated animals in contrast to the EPC and eNOS-EPC treated animals which showed visible reduction in muscle necrosis and atrophy (Figure 1A-C).

Characterization of EPCs

Under the culture conditions used in the present study, large round adherent cells appeared from the plated PBMCs at day 2 - day 3 (Supp Figure 1A), which then assumed a spindle-shaped, endothelial cell-like morphology at day 7 - day 8 (Supp Figure 1B). More than 90% of the adherent cells took up Di-acetylated LDL (red fluorescence) and showed concurrent binding to FITC-UEA-1 lectin (green fluorescence) (Supp Figure 2A). After 7 days in culture, majority of the cells also expressed endothelial markers vWF (Supp Figure 2B) and CD31 (Supp Figure 2C).

eNOS gene transfer and expression In vivo

Immunohistochemical staining of skeletal muscle sections with human specific eNOS antibody showed positive FITC-fluorescence in the tissues treated with eNOS-EPCs (Figure 2A, Figure 2B). No fluorescence was observed in the sections from EPC or PBS transplanted tissues (Supp Figure 3A-D). Human eNOS mRNA expression was detected in tissues homogenates of the eNOS-EPC treated rabbits whereas there was no mRNA expression in the limb tissue homogenates of EPC or PBS treated animals (Figure 2C).

Table 1: Macroscopic features observed in different groups of rabbits.

| Control n | EPC-treated rabbits n | eNOS-EPC treated rabbits n |
|-----------|------------------------|-----------------------------|
| Day 10 (Before Treatment) | Day 20 (After Treatment) | Day 10 (Before Treatment) | Day 20 (After Treatment) | Day 10 (Before Treatment) | Day 20 (After Treatment) |
| Muscle atrophy | 4 | 4 | 4 | 2 | 4 | 0 |
| Tissue necrosis | 2 | 2 | 2 | 1 | 2 | 0 |

n= no. of animals
Discussion

The present study demonstrates that eNOS gene transfer in ex vivo expanded EPCs improves angiogenesis in a rabbit model of experimentally induced hind limb ischemia. In a hypoxic state brought about by tissue ischemia, angiogenesis is induced by an up regulation of VEGF. This is followed by changes in shear stress in preexisting collaterals that activate eNOS and induce its gene expression after chronic changes in blood flow [22]. Consequently, a certain amount of basal revascularization occurs in response to the ischemic conditions. In comparison with earlier studies which observed more than two-

### Capillary/myocyte ratio

Representative anti CD31 immunostained sections of skeletal muscle tissue from various animal groups, two weeks after cell transplantation are shown in Figure 3A-C. Quantitative analysis revealed that capillary vessel numbers were significantly increased in rabbits receiving EPCs than those receiving PBS (2.7-fold, P< 0.05). Neo-capillary formation in rabbits receiving eNOS-EPCs was even more pronounced (4.5-fold relative to PBS treated group, P= 0.03 and 1.6-fold relative to EPC treated group, P= 0.05) (Figure 3D).

### Angiographic analysis

In the contrast angiography, there was restored perfusion in ischemic limbs in rabbits treated with eNOS-EPCs than those treated with EPCs or PBS (Figure 4A-C). Also, collateral vessel numbers and hence the angiographic score were significantly increased by 2.4 fold in eNOS-EPC treated group (0.7 ± 0.09) as compared to that observed in PBS (0.29 ± 0.03, P= 0.007) treated group and by 1.7 fold in comparison to EPC treated group (0.4 ± 0.04, P= 0.01) (Figure 4D). In eNOS-EPC treated group, a few of the growing collateral vessels were shown to extend all the way from their original source to near-the ankle, where they seem to reconstitute the distal arterial tree.
fold increase in angiogenic response after EPC therapy, we observed that the transplantation of ex vivo expanded autologous EPCs in the ischemic sites resulted in about two-fold increase in the angiogenic response over normal response to tissue ischemia after two weeks of EPC transplantation. This may be because in our study, EPC therapy was applied 10 days after the hind limb surgery in contrast to other studies that administered the cells one day after the surgery. Moreover, the dosage of EPCs used in the present study was about five to ten times less than that used in earlier studies [3-5]. The angiogenic effect was further enhanced by about four-fold over the normal response by an over expression of eNOS in culture-expanded EPCs, which could be robustly identified in the muscle tissues by RT-PCR, immunohistochemistry and increased nitrite levels in eNOS-EPC treated animals. This suggests that a combinatorial approach using eNOS and EPCs constitute an effective strategy to address the issue of quantitative and qualitative impairment of EPCs before ex vivo expansion and subsequent autologous readministration. The specific effects of eNOS transfection on EPCs are guarded by the fact that GFP transfection of EPCs did not result in any significant change/improvement in the functional properties of these cells under in vitro conditions [18]. It should also be noted that we didn’t observe any macroscopic adverse effects of the EPC or eNOS-EPC therapy on other organs such as liver, spleen, lungs and contra-lateral limbs. The treatment effect was also observed in terms of significant improvement in the macroscopic and histological features of the ischemic muscle such as higher reduction in the muscle atrophy and necrosis in the animals transplanted with eNOS-EPCs as compared to that observed in animals treated with saline and unmodified EPCs. We preferred to deliver the cells locally in the ischemic region as the efficiency of recruitment is reduced if cells are introduced into the systemic venous circulation due to sequestration in the lung and other tissues.

The formation of sustainable and functional new blood vessels in vivo comprises of three important processes: vasculogenesis, angiogenesis, and arteriogenesis [23]. Capillary density of ischemic muscle reflects vasculogenesis and angiogenesis because they are processes of capillary formation from endothelial cells or EPCs. On the other hand, the angiographic score represents the number of arteries with a relatively large diameter because angiograms provide images of vessels of >50 μm in diameter. In our study, both histological and angiographic evidences demonstrate a significant improvement in the angiogenesis and arteriogenesis, respectively of eNOS-EPC treated animals as compared to the EPC or PBS treated animals, implicating that eNOS may be enhancing several reactions that involve both angiogenesis and arteriogenesis leading to the formation of collateral vessels.

A plethora of studies in recent years have reported the putative potential of eNOS in angiogenesis. In mouse hindlimb models, eNOS knockout mice show a severe form of critical limb ischemia [16]. Further, studies have demonstrated an enhancement in the ischemia-induced angiogenesis by a local overexpression of eNOS [17,24,25]. In addition, eNOS exerts a second messenger role in VEGF signaling and has been shown to be vital for VEGF-induced endothelial cell proliferation, migration and capillary formation both in vitro and in vivo [15,26]. Studies from our lab have also suggested that in presence of VEGF, eNOS over expression improves the angiogenic properties of EPCs including proliferation, migration and integration into tube-like structures in vitro [18]. Given a crucial role of eNOS-NO in the induction of angiogenesis and the fact that ex vivo expanded EPCs are preferentially recruited to the ischemic foci [4], it is logical to presume that EPCs transfected with eNOS may be leading to an increased local overexpression of NO that in conjunction with other angiogenic factors such as VEGF may be directly or indirectly promoting angiogenic potential of the transplanted EPCs and/or differentiated ECs resident in the target ischemic tissue. The increased local bioavailability of NO may also augment the endogenous mobilization and functional activity of resident EPCs in these animals [27].

An added advantage of our therapy is that we have used a non-viral method of gene delivery that represents an important factor that may potentially enhance the safety profile of applications of gene therapy. Moreover, the need for long-term gene expression, which may be a prerequisite for certain types of inherited genetic defects, appears not to be a requirement for gene transfer strategies designed to promote neovascularization. Two to three weeks of gene expression appears to be sufficient to promote neovascularization and persistent blood flow.

In conclusion, the present study shows that autologous transplantation of eNOS modified ex vivo expanded EPCs enhances the angiogenic benefits of EPC therapy. As the viability of endogenous EPCs and also fully differentiated ECs is recurrently subjected to a variety of individual and environmental stress factors in peripheral and myocardial ischemic diseases, the concept of augmenting the regenerative and reparative potential of EPCs before in vivo transplantation constitutes a lucrative clinical approach for improving therapeutic angiogenesis in these diseases.

Acknowledgements

The study was financially supported by the Department of Biotechnology, Government of India.

References

1. Asahara T, Murohara T, Sullivan A, Silver M, van der Zee R, et al. (1997) Isolation of putative progenitor endothelial cells for angiogenesis. Science 275: 964-967.
2. Takahashi T, Kalka C, Masuda H, Chen D, Silver M, et al. (1999) Ischemia- and cytokine-induced mobilization of bone marrow-derived endothelial progenitor cells for neovascularization. Nat Med 5: 434-438.
3. Kalka C, Masuda H, Takahashi T, Kalka-Moll WM, Silver M, et al. (2000) Transplantation of ex vivo expanded endothelial progenitor cells for therapeutic neovascularization. Proc Natl Acad Sci U S A 97: 3422-3427.
4. Kawamoto A, Gwon HC, Iwaguro H, Yamaguchi JI, Uchida S, et al. (2001) Therapeutic potential of autologous transplantation of ex vivo expanded endothelial progenitor cells for myocardial ischemia. Circulation 103: 634-637.
5. Fan CL, Gao PJ, Che ZZ, Liu JJ, Wei J, et al. (2005) Therapeutic neovascularization by autologous transplantation with expanded endothelial progenitor cells from peripheral blood into ischemic hind limbs. Acta Pharmacol Sin 26: 1069-1075.
6. Tongers J, Roncalli JG, Losordo DW (2010) Role of endothelial progenitor cells during ischemia-induced vasculogenesis and collateral formation. Microvasc Res 79: 200-206.
7. Kirton JP, Xu Q (2010) Endothelial precursors in vascular repair. Microvasc Res 79: 193-199.
8. Vasa M, Fichtlscherer S, Aicher A, Adler K, Urbich C, et al. (2001) Number and migratory activity of circulating endothelial progenitor cells inversely correlate with risk factors for coronary artery disease. Circ Res 89: E1-7.
9. Kaur S, Jayakumar K, Kartha CC (2000) The potential of circulating endothelial progenitor cells to form colonies is inversely proportional to total vascular risk score in patients with coronary artery disease. Indian Heart J 59: 475-481.
10. Asahara T (2007) Cell therapy and gene therapy using endothelial progenitor cells for vascular regeneration. Handb Exp Pharmacol 180: 181-194.
11. Iwaguro H, Yamaguchi J, Kalka C, Murasawa S, Masuda H, et al. (2002) Endothelial progenitor cell vascular endothelial growth factor gene transfer for vascular regeneration. Circulation 105: 732-738.
12. Murasawa S, Llevadot J, Silver M, Isner JM, Losordo DW, et al. (2002) Constitutive human telomerase reverse transcriptase expression enhances regenerative properties of endothelial progenitor cells. Circulation 106: 2133-2139.

13. Chen SY, Wang F, Yan XY, Zhou Q, Ling Q, et al. (2009) Autologous transplantation of EPCs encoding FGF1 gene promotes neovascularization in a porcine model of chronic myocardial ischemia. Int J Cardiol 135: 223-232.

14. Yu JX, Huang XF, Lv WM, Ye CS, Peng XZ, et al. (2009) Combination of stromal-derived factor-1 alpha and vascular endothelial growth factor gene-modified endothelial progenitor cells is more effective for ischemic neovascularization. J Vasc Surg 50: 608-616.

15. Fukumura D, Gohongi T, Kadambi A, Izumi Y, Ang J, et al. (2001) Predominant role of endothelial nitric oxide synthase in vascular endothelial growth factor-induced angiogenesis and vascular permeability. Proc Natl Acad Sci U S A 98: 2604-2609.

16. Murohara T, Asahara T, Silver M, Bauters C, Masuda H, et al. (1998) Nitric oxide synthase modulates angiogenesis in response to tissue ischemia. J Clin Invest 101: 2567-2578.

17. Yu J, deMunck ED, Zhuang Z, Drnane M, Kauser K, et al. (2005) Endothelial nitric oxide synthase is critical for ischemic remodeling, mural cell recruitment, and blood flow reserve. Proc Natl Acad Sci U S A 102: 10999-11004.

18. Kaur S, Kumar TR, Uruno A, Sugawara A, Jayakumar K, et al. (2009) Genetic engineering with endothelial nitric oxide synthase improves functional properties of endothelial progenitor cells from patients with coronary artery disease: an in vitro study. Basic Res Cardiol 104: 739-749.

19. Pu LQ, Sniderman AD, Brassard R, Lachapelle KJ, Graham AM, et al. (1993) Enhanced revascularization of the ischemic limb by angiogenic therapy. Circulation 88: 208-215.

20. Kong D, Melo LG, Mangi AA, Zhang L, Lopez-Iliasaca M, et al. (2004) Enhanced inhibition of neointimal hyperplasia by genetically engineered endothelial progenitor cells. Circulation 109: 1769-1775.

21. Tsurumi Y, Takehisa S, Chen D, Kearney M, Rossow ST, et al. (1996) Direct intramuscular gene transfer of naked DNA encoding vascular endothelial growth factor augments collateral development and tissue perfusion. Circulation 94: 3281-3290.

22. Isner JM (2000) Tissue responses to ischemia: local and remote responses for preserving perfusion of ischemic muscle. J Clin Invest 106: 615-619.

23. Carmeliet P (2000) Mechanisms of angiogenesis and arteriogenesis. Nat Med 6: 389-395.

24. Smith RS Jr, Lin KF, Agata J, Chao L, Chao J (2002) Human endothelial nitric oxide synthase gene delivery promotes angiogenesis in a rat model of hindlimb ischemia. Arterioscler Thromb Vasc Biol 22: 1279-1285.

25. Amano K, Matsubara H, Iba O, Okigaki M, Fujiyama S, et al. (2003) Enhancement of ischemia-induced angiogenesis by eNOS overexpression. Hypertension 41: 156-162.

26. Papapetropoulos A, Garcia-Cardenas G, Madri JA, Sessa WC (1997) Nitric oxide production contributes to the angiogenic properties of vascular endothelial growth factor in human endothelial cells. J Clin Invest 100: 3131-3139.

27. Aicher A, Heeschen C, Mildner-Rihm C, Urbich C, Ihling C, et al. (2003) Essential role of endothelial nitric oxide synthase for mobilization of stem and progenitor cells. Nat Med 9: 1370-1376.