Schwann-like cell differentiation from rat bone marrow stem cells

Iraj Ragerdi Kashani, Zolikha Golipoor, Mohammad Akbari, Reza Mahmoudi, Shahram Azari, Reza Shirazi, Mohammad Bayat, Soudabeh Ghasemi

Abstract

Introduction: The main purpose of this study was differentiation of bone marrow stem cells (BMSCs) into Schwann-like cells and to determine the intensity of apoptosis in BMSCs during the differentiation process.

Material and methods: Bone marrow stem cells were isolated from the femur of adult rats and the identity of the undifferentiated BMSCs was confirmed by the detection of specific cell surface markers. The BMSCs were differentiated by sequential administration of β-mercaptoethanol and all-trans-retinoic acid as pre-inducer factors and a mixture of forskolin, basic fibroblast growth factor, platelet-derived growth factor-AA and heregulin-b1 as inducer factors. The immunocytochemical properties of differentiated Schwann-like cells were examined at a specified time point. Reverse transcription-polymerase chain reaction (RT-PCR) was used to investigate the gene expression of the undifferentiated and differentiated BMSCs. Cell apoptosis and viability were assessed with annexin V and propidium iodide double staining and dimethylthiazol-2-yl-2, 5-diphenyltetrazolium bromide (MTT) assay.

Results: Immunocytochemistry staining and RT-PCR analysis revealed that the induced BMSCs exhibited Schwann cell-specific markers such as S-100, P75 and glial fibrillary acidic protein (GFAP) at the 14th day of differentiation. MTT assay and flow cytometry revealed that of the total BMSCs in the differentiation medium, 40% to 50% of the cells died by apoptosis, but the remaining cell population remained strongly attached to the substrate and differentiated.

Conclusions: These findings indicated that BMSCs could differentiate into Schwann-like cells. As a side effect of differentiation an increased cell death rate was noted and our findings indicate that the principle mode of cell death is by apoptosis.

Key words: bone marrow, stem cells, apoptosis, Schwann cells.

Introduction

The transplantation of cultured Schwann cells (SCs) into bio-engineered conduits has been shown to improve nerve regeneration [1]. The ideal “transplantable cell” should be easily accessible, capable of rapid expansion in culture, immunologically inert, and capable of long-term survival and integration in the host tissue [2]. An alternative candidate may be
bone marrow stem cells (BMSCs) [3]. Recent transplantation studies have suggested that transplantation of undifferentiated BMSCs in rats showed therapeutic benefit after traumatic brain injury [4, 5] or spinal cord injury [6]. However, in these post-injury transplantation studies, generally less than 20% of transplanted cells were immunoreactive for specific antigens [7]. In a different paradigm, in vitro differentiation of BMSCs into cells expressing Schwann cell antigens, followed by in vivo transplantation, was shown to have a regenerative effect on damaged sciatic nerve [8]. This finding suggests that in vitro differentiation of BMSCs into cells of central and peripheral nervous systems, followed by in vivo transplantation, may be a potential therapy for nervous system repair. For cell-based regenerative therapies BMSCs are expanded in vitro and differentiated toward Schwann-like cells by applying culture media contain β-mercaptoethanol (BME), all-trans-retinoic acid (RA), forskolin, basic fibroblast growth factor, platelet-derived growth factor-AA and heregulin-b1 [9-11]. However, in these in vitro studies, generally less than 50% of BMSCs were differentiated into Schwann-like cells [11].

Beta-mercaptoethanol [12] and RA [13] are toxic chemicals that were used as pre-inducer factors. Evidence has been presented suggesting that RA is more active in inhibiting proliferation and inducing differentiation in relatively undifferentiated systems, i.e., neuroblastoma and embryonal stem cells [14]. In contrast, in other systems, such as keratinocytes [15] and embryonal lung cells [16], proliferation is stimulated. Moreover, at specific concentrations in culture medium, BME increase BMSCs apoptosis [13]. While the use of specific chemical compounds to induce neurogenic differentiation of BMSCs in culture has been well documented, the potential effects on cell death have not been elucidated [11, 13].

The main purpose of this study was differentiation of BMSCs into Schwann-like cells and to determine the intensity of apoptosis in BMSCs during the differentiation process.

Material and methods

**Isolation and culture of bone marrow stem cells**

About 6- to 8-week-old male Wistar rats of the Albino strain were killed using diethyl ether and the bones were collected under sterile conditions; then all the bones were cut at both ends. The bone marrow from each bone was collected by flushing the bone with α-Minimum Essential Medium Eagle (αMEM) (Sigma, USA) containing 1000 U/ml Penicillin G (Sigma, USA). After filtering, the cells were centrifuged at 1000 x g for 5 min. The purified cells were finally dispersed in αMEM with 15% fetal bovine serum (Sigma, USA) containing 100 U/ml penicillin and 100 µg/ml streptomycin (Sigma, USA) [17].

**Cell culture and expansion**

The isolated BMSCs were plated in T75 tissue culture flasks containing appropriate stem cell medium (Sigma, USA) at a density of 10 x 10^5 cells per flask. The flasks were maintained in a tissue culture incubator at 37°C and 5% carbon dioxide. The medium was replaced every third day afterwards. Cell viability was confirmed by continued cell division and the cells were subcultured using 3 ml of trypsin/EDTA (Sigma, USA) when the flasks reached 90% confluence [17].

**Primary culture of Schwann cells**

Primary Schwann cells were obtained by methods similar to those first described by Brockes et al. [18].

**Flow cytometry**

Rat BMSCs within 3-5 passages after the initial plating of the primary culture were harvested by trypsinization, and then the cells were fixed in neutralized 2% paraformaldehyde solution (Sigma, USA) for 30 min. The fixed cells were washed twice with phosphate buffer saline (PBS) (Sigma, USA) and incubated with antibodies to the following antigens: CD31, CD45, CD90 and CD44 (all from Chemicon, CA) for 30 min. Primary antibodies were directly conjugated with FITC. Flow cytometry was performed with a FACScan flow cytometer (Becton Dickinson, CA) [19].

**BMSCs multilineage differentiation potential**

The adipogenic and osteogenic differentiation capacity of BMSCs was confirmed according to the previously published protocols [11].

**Differentiation of BMSCs to SC-like cells**

After subculturing at the concentration of 10 x 10^5 cells/cm^2, BMSCs were incubated in αMEM containing 1 mM BME without serum for 24 h. The culture medium was then replaced with αMEM containing 10% FBS and 35 ng/ml RA (Sigma, USA). Three days later, cells were finally transferred to inducer medium containing αMEM, 10% FBS and trophic factors of 5 µM FSK (Calbiochem, CA), 10 ng/ml bFGF (Peprotech, UK), 5 ng/ml PDGF (Peprotech, UK), and 200 ng/ml HRG (R&D systems, USA) and cultured for 10 days [9].

**Immunostaining of cultured cells**

Differentiated BMSCs cultured on chamber slides (Lab-Tek, Denmark) were fixed in 4% (w/v)
paraformaldehyde at 4°C for 20 min. Cell nuclei were labelled with 6 diamidino-2-phenylindole dihydrochloride (DAPI, Sigma-Aldrich) (1 µg/ml) for 60 min at room temperature. Cells were then incubated overnight at 4°C with primary antibodies to S100 (rabbit polyclonal; 1 : 200; Dako, Denmark). The following day, slides were incubated for 2 hour with fluorescein isothiocyanate (FITC)-conjugated secondary antibody (horse anti-mouse or goat anti-rabbit; 1 : 100; Vector Labs., USA). Slides were examined under a fluorescence microscope (Olympus BX60). Cultures of Schwann cells were similarly stained as positive controls according to the antibodies used [11].

**RNA extraction and reverse transcription polymerase chain reaction (RT-PCR) analysis of gene expression**

After the pre-induction and the induction periods, the total RNA was extracted from the induced BMSCs, the non-induced cells and Schwann cells using RNX plus (CinnaGen) and was stored at –70°C. The cDNA synthesis was carried out from 5 µg total RNA using Fermentas kit (Fermentas) according to the manufacturer’s instructions. The cDNA was amplified (the primers of the genes in this study are listed in Table I). The 25 µl PCR product contained the following components: 0.2 pmol of each primer, 0.3 mM dNTP , 1.5 mM MgCl2, 1 U Taq DNA polymerase and 1 × PCR buffer (Fermentas). The PCR reactions were conducted in a programmable thermocycler (Bio-Rad) with the following temperature profile: 94°C for 5 min, 35 cycles at 94°C for 45 s, 55°C for 45 s and 72°C for 45 s, and a final extension at 72°C for 10 min; 10 µg of the PCR product was separated, run on a 1.5% agarose gel and stained with ethidium bromide [10].

**Cytotoxicity assay**

The BMSCs were incubated with β-mercaptoethanol, all-trans-retinoic acid and inducer medium respectively. Prior to measuring viability, treatment media were removed and replaced with 100 µl fresh 10% FBS/DMEM medium and 100 µl 3-(4,5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) (Sigma, USA). Cells were then returned to the incubator for an additional 4 h before 50 µl of 10% SDS was added to stop the reaction. Optical density (OD) at 570 nm was read on a microplate reader to indirectly reflect the number of viable cells. The inhibition rate (IR, IR = [1-ODtest/ODcontrol] × 100) was used to evaluate the cytotoxicity of pre-inducer and inducer media [20].

**Annexin V/PI assays**

To confirm apoptosis induction, annexin V, and propidium iodide (PI) double staining was performed by using the Annexin V-FITC Apoptosis Detection Kit as described by the manufacturer (Cat. No. 556420; BD Biosciences Pharmingen, San Jose, CA). The cells stained with annexin V-FITC were analysed with a flow cytometer (FACScan; Becton Dickinson, Mountain View, CA) using Cell Quest Software (BD Biosciences), which was also used to determine the percentage of apoptotic and/or necrotic cells [21].

**Statistical analysis**

The data were analysed with one-way ANOVA and Tukey's test.

**Results**

**Rat BMSCs characterization**

The BMSCs appeared as a monolayer of large, fibroblast-like flattened cells (Figure 1.A) which when treated with appropriate induction media

**Table I. Primer sequences used for RT-PCR**

| Gene  | Primers                      |
|-------|------------------------------|
| P75   | Forward 5'-CATCTCTTGTGGACAGCAGCAG-3'  |
|       | Reverse 5'-CTCTCTTGTGGACAGCAGCAG-3' |
| S100  | Forward 5'-ATAGCACTTCCTGCTGCTGAGCAG-3'  |
|       | Reverse 5'-CTGTTTGGCAAGAGGAAAG-3' |
| GFAP  | Forward 5'-GGTGAGTCATTGCTTTGTA-3'  |
|       | Reverse 5'-GGTGAGTCATTGCTTTGTA-3' |
| Beta actin | Forward 5'-CTGGCAACCCAGCACAATG-3'  |
|       | Reverse 5'-CTGGCAACCCAGCACAATG-3' |
where able to adopt an osteogenic (Figure 1. B) or adipogenic (Figure 1. C) phenotype.

Flow cytometry analysis of rat BMSCs within 3-5 passages showed that rat BMSCs were CD90 and CD44 positive, but CD31 and CD45 negative (Figure 2).

**Immunocytochemistry of BMSCs after induction**

We detected the expression of SC marker S-100 to evaluate the nature of SC-like cells. Almost all the differentiated BMSCs and Schwann cells were also positive to the S-100 antibody (Figure 3). After 14 days of induction, expression of S-100 protein was demonstrated in the cytoplasm of SC-like cells.

**RT-PCR results**

The presence of the glial cell marker gene transcripts was detected by RT-PCR methodology (Figure 4). Before differentiation induction, BMSCs were shown to be negative for S100, P75 and GFAP. After 14 days of induction, BMSCs expressed the transcripts for S100, P75 and GFAP compared to the undifferentiated BMSCs. The intensity of S100 expression was higher than that of GFAP and similar to the Schwann cell group. RT-PCR amplification efficacy of the mRNA was confirmed by amplification of the constitutively expressed beta actin housekeeping gene (Figure 4).

**Cytotoxicity assay**

To study the cytotoxicity effect of BME, RA and inducer medium on BMSCs during differentiation into Schwann-like cells, we evaluated the viability of BMSCs by MTT assay. This assay usually shows high correlation with the number of living cells, cell proliferation and release of mitochondrial matrix enzymes. The BME, RA and inducer medium significantly reduced BMSCs viability as assessed by MTT assay (Figure 5).

**Quantification of apoptosis by flow cytometry**

By staining cells with annexin V-FITC and PI, FACS was used to distinguish and quantitatively determine the percentage of viable cells (lower
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left quadrant), apoptotic cells (lower right quadrant), late apoptotic necrotic cells (upper right quadrant) and necrotic cells (upper left quadrant) after sequential administration of β-mercaptoethanol, all-trans-retinoic acid and a mixture of FSK, bFGF, PDGF and HRG was used to induce differentiation of BMSCs into Schwann-like cells (Figure 6. A). The proportions of viable cells, early apoptotic cells, late apoptotic/necrotic and necrotic cells for the control group were 94.34%, 2.34%, 1.62% and 1.7% respectively. For cells treated with β-mercaptoethanol, the proportions were 78%, 14%, 6% and 2% respectively. For cells treated with all-trans-retinoic acid, the proportions were 75.84%, 19.31%, 3.53% and 1.32% respectively. For cells treated with inducer medium proportions were 80.84%, 13.93%, 5% and 0.23% (Figure 6. B).

Discussion

Over the past decade, stem cells from adult bone marrow have been exploited as therapeutic vectors in the treatment of a wide variety of diseases [4, 5]. This study clearly shows the plasticity of BMSCs by their differentiation into Schwann-like cells.

Figure 3. Immunocytochemistry of S100 in differentiated BMSCs (A-C) and rat Schwann cell-line (D-F). After the induction, differentiated BMSCs became positive for S100 antibody. Rat Schwann cell-line was used as a positive control. DAPI staining (A, D); S100 antibody (B, E); merge (C, F).
The previous reports showed that the addition of BME to cultured BMSCs induced the formation of neurite-like processes [14]. RA is a morphogenic factor, and was reported to induce differentiation of neural stem cells into nerve cells [15]. Therefore, BME and RA are presumed to work as triggering factors, inducing changes in the morphological and transcriptional characteristics of BMSCs.

Previous studies reported that bFGF functions as a mitogen for BMSCs, and that MAP kinases are activated by PDGF and bFGF in MSCs [16]. HRG, a subtype of neuregulin, instructively influences the decision of cell fates and was reported to induce neural crest cells to develop selectively into Schwann cells [17].

FSK increases the level of intracellular cyclic adenosine monophosphate (cAMP), which increases mitogenic responses at an early time [18]. As cAMP elevation is reported to enhance the responsiveness of cells to trophic factors [19], FSK together with bFGF, PDGF, and HRG could have a synergistic effect in enhancement of these factors to BMSCs [20].

MTT assay and flow cytometry analysis for annexin V and PI indicated that during the chemical induction process there was an increase in cell death. Putative early apoptotic cells increased after exposure to BME and RA.

Measures of cell death during this specific induction process are indicative of toxicity of the chemical compounds used. Here we show that almost 50% of cells die within 48 h of chemical induction, in agreement with other studies [8, 10, 21].

Our results indicate that RA exerts two apparently opposite effects on cultured ADSCs, i.e., a differentiating effect and induction of cell death. Flow cytometry analysis for annexin V and PI indicated that putative early apoptotic cells increased after exposure to RA. These data agreed with previous reports where all-trans-retinoic acid produced similar effects in cancers [22] and stroke [23].

The mechanism of the apoptotic effects of β-mercaptoethanol and all-trans-retinoic acid is unknown and there are several pathways that need to be investigated.

Oliva et al. reported that antiproliferative activity of retinoic acid is associated with (and probably due to) the up-regulation of two pivotal cdk inhibitors, and subsequent cdk2 activity decrease and retinoblastoma protein (pRb) hypophosphorylation [24].

pRb is a tumour suppressor protein that is dysfunctional in many types of cancer [25]. In the hypophosphorylated state, pRb is active and carries out its role as a tumour suppressor by inhibiting cell cycle progression [26].

However, there is evidence indicating that either direct contact with axons or survival factors secreted by neurons are necessary for the survival
and development of SC precursors as well as of mature Schwann cell [8].

In conclusion, these findings indicated that BMSCs could differentiate into Schwann-like cells in terms of morphology and phenotype. As a side effect of differentiation an increased cell death rate was noted and our findings indicate that the principle mode of cell death is by apoptosis.

Hence, a considerable amount of BMSCs would die during differentiation into Schwann-like cells, depending on the method used. This may compromise the optimal outcome of the engraftment and subsequently nerve regeneration and repair.

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