The citrus flavanone naringenin impairs dengue virus replication in human cells

Sandra Frabasile¹, Andrea Cristine Koishi², Diogo Kuczera², Guilherme Ferreira Silveira², Waldiceu Aparecido Verri Jr.³, Claudia Nunes Duarte dos Santos² & Juliano Bordignon²

Dengue is one of the most significant health problems in tropical and sub-tropical regions throughout the world. Nearly 390 million cases are reported each year. Although a vaccine was recently approved in certain countries, an anti-dengue virus drug is still needed. Fruits and vegetables may be sources of compounds with medicinal properties, such as flavonoids. This study demonstrates the anti-dengue virus activity of the citrus flavanone naringenin, a class of flavonoid. Naringenin prevented infection with four dengue virus serotypes in Huh7.5 cells. Additionally, experiments employing subgenomic RepDV-1 and RepDV-3 replicon systems confirmed the ability of naringenin to inhibit dengue virus replication. Antiviral activity was observed even when naringenin was used to treat Huh7.5 cells 24 h after dengue virus exposure. Finally, naringenin anti-dengue virus activity was demonstrated in primary human monocytes infected with dengue virus serotype-4, supporting the potential use of naringenin to control dengue virus replication. In conclusion, naringenin is a suitable candidate molecule for the development of specific dengue virus treatments.

Dengue virus (DENV) infection is a rapidly spreading viral disease worldwide and is transmitted by the Aedes aegypti and Aedes albopictus mosquitoes. Two and a half billion people live in dengue-endemic regions in tropical and subtropical countries in Southeast Asia, the Pacific and the Americas. Approximately 390 million dengue cases are estimated each year. In the Americas alone, 2.3 million cases were reported between 2008 and 2012. Dengue is caused by infection with one of four antigenically related viruses (DENV-1, -2, -3 and -4). The virus, belonging to the genus Flavivirus and family Flaviviridae, is a small, enveloped virion with a single 11-kb positive RNA strand. The RNA genome comprises 10 genes, 5′-C-prM-E-NS1-NS2A/B-NS3-NS4A/B-NS5-3′, which are flanked by a 5′ and a 3′ UTR (untranslated region). Infection with DENV leads to dengue fever (DF) without warning signs, DF with warning signs or severe dengue with capillary leakage and hemorrhagic manifestations.

Despite the development and approval (in certain countries) of a tetravalent dengue vaccine, antiviral research is still necessary to develop treatments for individuals that are unable to receive the vaccine and for unvaccinated infected patients. The use of plant-derived extracts and their purified compounds as possible antiviral therapies has been explored. Among various natural products, flavonoids are an important source of compounds possessing antiviral activity against HIV-1, rotavirus, norovirus, adenovirus, herpesvirus (1 and 2), dengue and influenza. The basic structure of a flavonoid consists of a flavan nucleus with 15 carbon atoms arranged in three rings (C-6–C-3–C-6), referred to as A, B, and C. Flavonoid classes include flavones, flavanone, flavonols, flavanolol, isoflavones, flavan-3-ols, anthocyanidins, chalcones and auroines, which differ in terms of their oxidation levels and C ring substitution patterns. Substitutions in the A and B rings differentiate each compound within a class. Flavonoids are widely found in fruits (grapefruit and oranges), vegetables, nuts, seeds, flowers, teas and wine. One flavanone, naringenin, possesses anticancer, antimutagenic, anti-inflammatory, analgesic, antioxidant, antiproliferative and antiatherogenic properties. Importantly, naringenin exerts antiviral effects against Hepatitis C virus (HCV), another flavivirus, by inhibiting NS2 activity.

¹Sección Virologia, Facultad de Ciencias, Universidad de La República, 11400, Montevideo, Uruguay. ²Laboratório de Virologia Molecular, Instituto Carlos Chaques, ICC/FIOCRUZ/PR, Curitiba, Paraná, Brazil. ³Departamento de Ciências Patológicas, Centro de Ciências Biológicas, Universidade Estadual de Londrina, Paraná, Brazil. *These authors contributed equally to this work. Correspondence and requests for materials should be addressed to J.B. (email: julianobordignon@fioiculto.br)
Using flow cytometry, virus titration and replicon assays, this study demonstrated the ability of naringenin to inhibit the replication of four DENV serotypes in Huh7.5 cells. Antiviral activity was evident even when naringenin was used to treat Huh7.5 cells 24 h after DENV infection. To extend our observations to a more relevant system, the antiviral activity of naringenin was also demonstrated in primary human monocytes (CD14+) after DENV-4 infection.

Results

Naringenin inhibits infection with four different DENV serotypes in Huh7.5 cells. The cytotoxicity of naringenin is cell line-dependent. Different results have been observed in Vero and Hep2 cells. Therefore, prior to the in vitro assessment of antiviral activity in Huh7.5 cells, cytotoxicity was evaluated using the neutral red assay. Plotting cell viability against eight different concentrations of naringenin revealed concentration-dependent toxicity in Huh7.5 cells (Supplementary Figure 1). Also, a naringenin concentration of 311.3 μM inhibited 50% of cell viability (CC50). The non-toxic concentration (NTC) of naringenin was 250 μM, which did not differ statistically from untreated control cells.

The anti-dengue activity of naringenin was evaluated using four DENV serotypes and recombinant IFN-α 2A as the positive control. Naringenin (250 μM) was added to Huh7.5 cells during and after DENV infection to investigate the entire life cycle of the virus. After 72 h, a flow cytometry assay (FACS) revealed a reduction in the number of DENV-infected cells compared to non-treated controls (Fig. 1A,B). Additionally, the titration of DENV viable particles in cell culture supernatants using a foci-forming immunodetection assay confirmed...
the anti-DENV activity of naringenin (Fig. 1C). To further explore the anti-DENV effects of naringenin, seven other DENV strains belonging to the four DENV serotypes were tested. As shown in Fig. 2, the percentage of 4G2-positive Huh7.5 cells was reduced after treatment with either naringenin or IFN-α2A for all strains tested (Fig. 2). Based on these results, the anti-DENV activity of naringenin is independent of the virus serotype or strain.

Furthermore, a concentration-response curve was performed to establish whether the activity in the antiviral assay was dependent on the amount of naringenin. According to our results, the anti-DENV activity of naringenin was concentration-dependent (Supplementary Figure 2 and Table 1). The 50% inhibitory concentration (IC50) was calculated, as well as the selectivity index for each DENV serotype (SI = CC50/IC50).

Table 1. Naringenin half maximal inhibitory concentration (IC50) and selectivity index (SI) for anti-DENV activity in Huh7.5 cells. SI = CC50/IC50.

| DENV     | IC50  | SI   |
|----------|-------|------|
| DENV-1/FGA | 35.81 | 8.60 |
| DENV-2/ICC265 | 17.97 | 17.32|
| DENV-3/5532 | 117.1 | 2.66 |
| DENV-4/TVP360 | 177.5 | 1.75 |

the anti-DENV activity of naringenin (Fig. 1C). To further explore the anti-DENV effects of naringenin, seven other DENV strains belonging to the four DENV serotypes were tested. As shown in Fig. 2, the percentage of 4G2-positive Huh7.5 cells was reduced after treatment with either naringenin or IFN-α2A for all strains tested (Fig. 2). Based on these results, the anti-DENV activity of naringenin is independent of the virus serotype or strain.

Furthermore, a concentration-response curve was performed to establish whether the activity in the antiviral assay was dependent on the amount of naringenin. According to our results, the anti-DENV activity of naringenin was concentration-dependent (Supplementary Figure 2 and Table 1). The 50% inhibitory concentration (IC50) was calculated, as well as the selectivity index for each DENV serotype (SI = CC50/IC50).

Naringenin inhibits DENV replication in Huh7.5 cells. After demonstrating the antiviral effects of naringenin, we determined the stage of the DENV life cycle during which this compound exerts its effects. Initially, we tested the virucidal effects of naringenin and observed its inability to destroy DENV particles (Supplementary Figure 3). Next, we used the time-of-drug addition approach to observe whether the compound interfered with the early or late phases of the DENV life cycle. In most cases, naringenin treatment before...
and during infection did not reduce the percentage of Huh7.5 DENV-infected cells, although decreases were observed in the percentages of cells infected with DENV-2/ICC265. Overall, naringenin treatment was most effective either during and after infection or only after infection with the four DENV serotypes. These results were similar to those observed with IFN-α 2A treatment after infection (Fig. 3) and suggest the ability of naringenin to impair DENV replication and/or virus maturation. To determine if naringenin affects viral replication, DENV-1 and -3 replicons (RepDV1 and RepDV3) were employed. These subgenomic RNA systems contain DENV non-structural viral proteins (NS) that enable RNA replication and translation without viable DENV particle assembly. These features make DENV replicons good tools to study virus replication for the purpose of antiviral drug development. The DENV replicon assay is used to identify antiviral compounds that specifically impair DENV replication and/or translation27,28. According to our results, Huh7.5 cells were successfully transfected, and naringenin reduced DENV replication with an efficiency similar to IFN-α 2A and ribavirin treatments (Fig. 4A–C). Furthermore, DENV-RNA transfection of Huh7.5 cells did not affect cell viability (data not shown).

To determine the efficiency of naringenin after Huh7.5 cells were infected with DENV, treatment was administered at different time points after challenge with the DENV-1 serotype (FGA/89). Naringenin efficiently reduced the percentage of Huh7.5 DENV-1 infected cells even when added 24 h after the infection (Fig. 5). Also, the naringenin treatment 6 h after the DENV-1 infection reduced the DENV-titer at the cell culture supernatant (Fig. 5). Treatment with IFN-α 2A was no longer effective when added 4 h after infection, confirming the potential anti-DENV activity of naringenin (Fig. 5).

**Naringenin decreases DENV infection in human monocytes.** Once naringenin anti-DENV activity was confirmed in a human cell line (Huh7.5 cells), the ability of naringenin to impair DENV replication in primary human monocytes from healthy donors was investigated. First, it was established the non-toxic concentration of naringenin for human PBMCs using annexin V/7-AAD assay and monocyte quantification (CD14+).
Unlike the results in Huh7.5 cells, naringenin was toxic to monocytes at concentrations of 250 and 125 μM but not at 62.5 μM (Supplementary Figure 4C,D). Thus, as a proof of concept, peripheral blood mononuclear cells (PBMCs) were infected with DENV-4/TVP360 and treated with the non-toxic concentration of naringenin (62.5 μM). Importantly, even at lower concentrations (relative to those employed in Huh7.5 cells), naringenin reduced the number of infected CD14+ cells (monocytes) in human PBMCs as well as the number of infectious viral particles in the culture supernatant (Fig. 6A–D).

Discussion
Flavonoids constitute a class of naturally occurring compounds that possess multiple pharmacological characteristics such as antioxidant, anti-inflammatory, analgesic, anti-carcinogenic, antibacterial infection, antifungal and antiviral properties. The antiviral activity of flavonoids or flavonoid-derived compounds has been demonstrated against several viruses affecting humans, including norovirus, rotavirus, adenovirus, herpes simplex, HCV, dengue, chikungunya and HIV-1. Here, we evaluated the anti-DENV activity of the flavanone naringenin.

Flavonoids are amply present in fruits and vegetables. Importantly, flavonoids present low toxicity in different cell lines. Here, the CC50 of naringenin was determined to be 311.3 μM (84.75 μg/mL) in Huh7.5 cells, similar to that reported by Zandi et al. in Vero cells (CC50 of 304.85 μM; 83 μg/mL). We also established the NTC of naringenin in Huh7.5 cells (250 μM; 68.06 μg/mL). In a previous study by Khachatoorian et al., naringenin was not toxic to Huh7.5 cells at concentrations between 25 and 125 μM. However, for primary human monocytes, a lower concentration of naringenin had to be used (62.5 μM; 16.94 μg/mL). Additionally, Zanello et al. studying anti-dengue virus activity of quinic acid derivatives, had demonstrated that primary human PBMC seems to be more sensible than cell lines, confirming our observation.

The flavonoid fisetin inhibits DENV-2 infection in Vero cells, whereas naringenin and rutin do not. Additionally, naringenin has virucidal effects on DENV-2. Surprisingly, using flow cytometry and virus titration, we demonstrated the ability of naringenin to efficiently inhibit infection by eleven different DENV strains.
representing the four different virus serotypes in the Huh7.5 cell line. In contrast to results reported by Zandi et al., our RT-PCR assay results did not reveal any virucidal effects of naringenin on any of the four DENV serotypes. Zandi et al. only used the DENV-2 strain (New Guinea C), while we employed an RT-PCR assay and recent clinical isolates of the four different DENV serotypes to investigate the virucidal effects of naringenin. These differences may explain the contradictory results.

To determine the stage of the DENV life cycle during which naringenin exerts its antiviral effects, time-of-drug addition experiments were carried out. Aside from the partial inhibition observed with DENV-2, naringenin did not affect binding and entry into Huh7.5 cells. Indeed, naringenin appeared to target other stages of the DENV life cycle, including replication and/or maturation. To determine if naringenin impairs DENV replication, we used the DENV-1 and DENV-3 replicon systems. In accordance with quinic acid derivatives and 2-bromo-α-ergocriptine, naringenin appears to reduce DENV replication. Recently, a stable BHK-21 cell line carrying the chikungunya replicon was employed to demonstrate the ability of naringenin to impair chikungunya virus (CHIKV) replication. It seems that some flavonoids could impair the activity of viral proteins important for virus replication, like the serine protease activity (NS2B-NS3) of dengue and Zika virus, and also the protease (NS2) of HCV. Furthermore, naringenin inhibits intracellular HCV protein production and viral assembly, in agreement with our findings. Flavonoids also impair influenza A replication and spread. C2 group replacement in the chemical bond between the C2 and C3 rings may interfere with the cellular response to influenza A infection via MAPK signaling pathway modulation.

Even when naringenin was added to Huh7.5 cell cultures several hours after infection (6 h and 24 h), the synthesis of viable virus particles and the percentage of infected cells were significantly reduced. Naringenin controlled DENV replication as efficiently as IFN-α 2A, a known antiviral cytokine. 2-bromo-α-ergocriptine impairs DENV translation and/or RNA synthesis as late as 6 h after infection. Treatment of DENV-2-infected cells with NITD-618, a benzomorphan core structure, reduces infection even when administered 12 h after infection. The effects of naringenin 24 h after DENV infection in Huh7.5 cells demonstrated the potential of this flavanone as an anti-DENV compound.

In conclusion, data from multiple assays (flow cytometry, viral titration and replicon system) employed to assess infection with eleven different strains representing the four DENV serotypes in two cell types (the Huh7.5 cell line and primary human monocytes) support the ability of naringenin to target DENV replication, making naringenin a suitable candidate for the treatment of DENV infection. Our results provide novel insights for the development of specific anti-DENV drugs to treat infected patients.
Materials and Methods

Cells, DENV and Naringenin. *Aedes albopictus* mosquito cells C6/36 (ATCC; CLR-1660) were grown in Leibovitz’s Medium (L-15; Gibco, Grand Island, NY, USA) supplemented with 0.26% tryptose (Sigma-Aldrich, St. Louis, MO, USA) and 25 μg/mL gentamicin (Sigma-Aldrich, St. Louis, MO, USA) at 28 °C. The human-derived Huh7.5 hepatoma cell line (ATCC PTA-8561) was grown in Dulbecco’s Modified Eagle Medium/nutrient mixture F-12 (D-MEM/F-12; Gibco, Grand Island, NY, USA) supplemented with 100 IU/μg/mL penicillin/streptomycin (Gibco, Grand Island, NY, USA) at 37 °C in a humidified, 5% CO2 controlled atmosphere. PBMCs were isolated from the whole blood of healthy volunteers (approved by the FIOCRUZ Committee of Ethics in Research, number 514/09), by density gradient centrifugation in lymphocyte separation medium (Lonza, Walkersville, MD, USA). PBMCs were cultured in 24-well plates with Roswell Park Memorial Institute medium-1640 (RPMI-1640; Lonza, Walkersville, MD, USA) supplemented with 2 mM glutamine (Gibco, Grand Island, NY, USA), 100 IU/μg/mL penicillin/streptomycin, 2.5 μg/mL amphotericin B (Sigma-Aldrich, St. Louis, MO, USA), 100 mM sodium pyruvate (Sigma-Aldrich, St. Louis, MO, USA) and 10% FBS in a humidified, 37 °C atmosphere with 5% CO2. The authors confirm that all experiments were performed in accordance with relevant guidelines and regulations.

DENV-1/FGA/89 was isolated in 1989 from a South American patient suffering from DF (GenBank: AF226687). DENV-1/ICC-265 was isolated from a DF patient in Brazil in 2009. DENV3/5532 was isolated in 2007 from a fatal case of dengue with visceral manifestations in a patient in Paraguay (GenBank: HG235027). DENV-4/TVP360 is a laboratory strain that was kindly provided by Dr. Ricardo Galler (Fundação Oswaldo Cruz, Rio de Janeiro, Brazil; GenBank: KU513442). Other DENV virus strains from Brazilian patients with DF between 1990 and 2014 were also used: DENV-1/BR/90 (GenBank AF226685.2), DENV-1 209, DENV-2 Jamaica,
DENV-2/ICC-266, DENV-3/97 (GenBank EF629367), and DENV-3/98 (GenBank EF629368). Additionally, a clinical isolate from a non-fatal case of DENV-4 with hemorrhagic manifestations was employed (DENV-4/LRV13/422; GenBank: KU513441). Virus stocks were propagated in C6/36 cells and titrated in a foci-forming immunodetection assay44.

Naringenin (>95% purity) was purchased from Santa Cruz Biotechnology and prepared in a 100% solution of dimethyl sulfoxide (DMSO, Sigma-Aldrich, St. Louis, MO, USA).

**Cytotoxicity Assays.** Huh7.5 cell viability was tested using a neutral red (NR; Sigma-Aldrich, Irvine, UK) uptake assay as previously described25. Serial dilutions of naringenin (50 mM stock) in DMEM-F12 medium and 1% DMSO were added to 2 × 10^4 Huh7.5 cells/well in a 96-well plate and incubated for 72 h. The supernatant was removed, and a solution containing neutral red (33 μg/mL) was applied for 2 h. After incubation, the supernatant was removed, and an acified ethanol solution was used to extract the dye and perform optical density (OD) quantification at an absorbance of 540 nm using a spectrophotometer. Data from three independent experiments were normalized using the following equation: cell viability (%) = (OD sample value − OD blank control)/(OD cell control − OD blank control) × 100. The NTC was defined as the highest concentration that did not differ statistically from untreated control cells. The concentration that inhibited viability in 50% of cells (CC50) was obtained by performing nonlinear regression followed by the construction of a sigmoidal concentration-response curve (variable slope; GraphPad Prism; La Jolla, CA, USA).

**Antiviral Activity Assays.** The antiviral activity of naringenin was assessed in two different assays. Huh7.5 cells at concentration of 2 × 10^4 cells/well in 96-well plates were infected with one representative strain of each of the four DENV serotypes (DENV-1/FGA/89, DENV-2/ICC265, DENV-3/5532 and DENV-4/TVP360) at a multiplicity of infection (MOI) of 10 for 90 minutes. Naringenin at its NTC was added during and after infection. Positive and negative controls for antiviral activity were included using IFN-α 2A (200 IU/mL; Blau Farmacéutica, Cotia, SP, Brazil) and mock-infected cells, respectively. After 72 h of incubation, the cell culture supernatants were stored at −86 °C for virus titration. The cells were detached and stained for flow cytometry33. A BD FACS Canto II (BD Biosciences, USA) was used to quantify cells infected by DENV. Cell culture supernatants were also employed for titration in a foci-forming immunodetection assay in C6/36 cells to confirm the FACS results44.

Additionally, dose response curves were obtained with a serial dilution starting at the NTC of naringenin. The concentration that inhibited 50% of virus infection (IC50) was obtained using nonlinear regression, followed by the construction of a sigmoidal concentration-response curve (variable slope; GraphPad) and calculation of the selectivity index (SI = CC50/IC50). All assays were performed in triplicate.

Antiviral activity was further confirmed for other DENV strains representing the four serotypes (DENV-1 209, DENV-1/BR/90, DENV-2/ICC-266, DENV-2 Jamaica, DENV-3/97, DENV-3/98 and DENV-4/LRV13/422) using the FACS assay as described above.

**Virucidal Assay.** A virucidal assay was performed as previously described with minor modifications45. Briefly, a sample of each DENV serotype (2 × 10^6 ffu/mL; DENV-1/FGA/89; DENV-2/ICC-265; DENV3/5532 and DENV-4/TVP360) was treated with naringenin (250 μM) in the presence or absence of 150 μg/mL RNase A (USB-Affymetrix; Santa Clara, CA, USA) for 1 h at 37 °C. After treatment, viral RNA was extracted using a QIaAmp Viral RNA Mini Kit (QIAGEN; Hilden, Germany). The RNA was reverse transcribed using 250 pmol of a random primer (Gibco, Grand Island, NY, USA) and Improp II Reverse Transcriptase (Promega, Fitchburg, WI, USA). PCR amplification was performed as described by Lanciotti et al. with a D1 (5′-TCAATATGCTGAAACGCGCGAGAAACCG-3′) and D2 (5′-ATTGACCAGCAGTCAACGTCTCGGTTTC-3′) primer pair. DENV RNA samples treated with RNase or left untreated were used as the positive and negative control, respectively.

**Time-of-Drug Addition Assay.** Initially, Huh7.5 cells at a density of 2 × 10^4 cells/well in 96-well plates were (i) treated with naringenin for 1 h prior to DENV infection; (ii) treated with naringenin during DENV infection; (iii) treated with naringenin after DENV infection; or (iv) treated with naringenin during and after DENV infection. An MOI of 10 was used for DENV infection, and naringenin was tested at a concentration of 250 μM. The percentage of infected cells was determined using FACS as previously described.

In another experiment, cells were infected with DENV-1 strain FGA/89 (MOI: 10) as previously described and treated with naringenin (250 μM) or with IFN-α 2A (200 IU/mL) after infection (0, 1, 2, 4, 6, 24, 30 and 48 h). The percentage of infected cells was determined using FACS. The supernatants were titrated in a foci-forming immunodetection assay 72 h post-infection.

**Naringenin Impairs DENV Replication in Huh7.5 Cells.** After confirming the anti-DENV effects of naringenin after viral entry into Huh7.5 cells, we determined if this flavanone impairs DENV replication. Two subgenomic replicons derived from DV1-1/BR/90 (RepDV1) and BR DENV 290-02 (RepDV3) were used in these experiments27,28. DENV replicon RNAs were obtained and used to transfect Huh7.5 cells (2 μg of RNA/2 × 10^6 cells) in a Nucleofector 2B device (Lonza; Cologne, Germany) according to the manufacturer’s instructions33. After 1 h of transfection, the Huh7.5 cells were treated with naringenin (250 μM). The plates were further incubated for 72 h. After incubation, the cells were recovered and subjected to FACS analysis, as described previously. As a negative control, Huh7.5 cells were transfected without RNA and were not treated. Huh7.5 cells transfected with RNA (from the DENV-1 or DENV-3 replicon) and untreated cells were used as positive controls for virus replication. The reference control consisted of treatment with recombinant IFN-α 2A (200 IU/mL) and 20 μM ribavirin (Sigma-Aldrich, St. Louis, MO, USA).
Antiviral Effects in Primary Human Cells. PBMCs obtained from healthy donors who provided signed informed consents were purified using Ficoll-histopaque and a classical protocol16. After purification, PBMCs were plated onto 24-well plates at a density of $1 \times 10^5$ cells/well and treated with 200 μM of naringenin for 5 days and then the exposure of annexin V/FITC-PE was evaluated. Also, we quantified the number of human monocytes (CD14$^+$) after treatment with naringenin (250, 125 and 62.5 μM) for five days. After it was established that the NTC of naringenin was 62.5 μM, infection with dengue was performed. Human PBMCs were plated onto 24-well plates at a density of $1 \times 10^6$ cells/well and infected with DENV-4 TVP/360 (MOI: 10) for 2h. The inoculum was removed, and cells were treated with naringenin (62.5 μM), IFN-α 2A (200 IU/mL) or RPMI media. After incubation for 5 days at 37°C and 5% CO₂, cells were stained for DENV E protein (mAb 4G2-FITC conjugated) and with a mouse anti-human CD14 PE-Cy7 antibody67 (BD Biosciences, USA). CD14 expression was used to gate the monocytes. The percentage of DENV E-positive cells (4G2$^+$) in this population was calculated.

Data Analysis. Statistical analysis consisted of one-way ANOVA followed by Dunnett's test for multiple comparisons with a significance of p < 0.05. Statistical analyses were performed using Prism software (GraphPad version 5.0; La Jolla, CA, USA). Primary human monocyte infection was analyzed using two-way ANOVA and the Bonferroni correction for multiple comparisons with a significance level of p < 0.05. Flow cytometry data were analyzed using FlowJo version X software (Tree Star Inc.; Ashland, OR, USA).

References

1. WHO. Dengue: guidelines for diagnosis, treatment, prevention, and control. Spec. Program. Res. Train. Trop. Dis. 147, doi: WHO/HTM/NTD/DEN/2009.1 (2009).
2. Bhatt, S. et al. The global distribution and burden of dengue. Nature 496, 504–507 (2013).
3. Henchel, E. A. & Putnak, J. R. The Dengue Viruses. Clinical Microbiology Reviews 3, 376–396 (1990).
4. Chambers, T. J., Hahn, C. S., Galler, R. & Rice, C. M. Flavivirus genome organization, expression, and replication. Annu. Rev. Microbiol. 44, 649–688 (1990).
5. Hadiningsarto, S. R. et al. Efficacy and Long-/Term Safety of a Dengue Vaccine in Regions of Endemic Disease. N. Engl. J. Med. 373, 1195–1206 (2015).
6. Jassim, S. A. A. & Najj, M. A. Novel antiviral agents: A medicinal plant perspective. J. Appl. Microbiol. 95, 412–423 (2003).
7. Cushnie, T. P. T. & Lamb, A. J. Antimicrobial activity of flavonoids. Int. J. Antimicrob. Agents 26, 343–356 (2005).
8. Debiaggi, M., Tatoe, F., Pagani, L., Luini, M. & Romero, E. Effects of propolis flavonoids on virus infectivity and replication. Microbiologica 13, 207–213 (1990).
9. Lulu, S. S., Thabitha, A., Vino, S., Priya, M. & Rout, M. Naringenin and quercetin – potential anti-HCV agents for NS2 protease targets. Natural Product Research 30(4), 464–8 (2016).
10. Verri, W. A. et al. Flavonoids as anti-inflammatory and analgesic drugs: Mechanisms of action and perspectives in the development of pharmaceutical forms. Stud. Nat. Prod. Chem. 36, 297–330 (2012).
11. Havsteen, B. Flavonoids, a class of natural products of high pharmacological potency. Biochem. Pharmacol. 32, 1141–1148 (1983).
12. Macdonald, J. E. & Kandaswami, C. The impact of plant flavonoids on mammalian biology: implications for immunity, inflammation and cancer. The flavonoids–Advances in research since 1986, doi: 10.1007/978-1-4899-2911-2_15 (Springer, US, 1993).
13. Grange, J. M. & Davey, R. W. Antibacterial properties of propolis (bee glue). J. R. Soc. Med. 83, 159–60 (1990).
14. Harborne, J. B. & Baxter, H. Handbook of Natural Flavonoids (1999).
15. Lin, X. & D’O Souza, D. H. Naturally Occurring Flavonoids Against Human Norovirus Surrogates. Food Environ. Virol. 5, 97–102 (2013).
16. Ginneken, U. et al. Discovery of prenylated flavonoids with dual activity against influenza virus and Streptococcus pneumoniae. Sci. Rep. 6, 27156 (2016).
17. Verri, W. A. et al. Flavonoids as anti-inflammatory and analgesic drugs: Mechanisms of action and perspectives in the development of pharmaceutical forms. Stud. Nat. Prod. Chem. 36, 297–330 (2012).
18. Li, B. Q. et al. Antiviral activity of Fisetin, Rutin and Naringenin against Dengue virus type-2. J. Med. Plants Res. 5, 5534–5539 (2011).
19. Su, X. & D’O Souza, D. H. Naturally Occurring Flavonoids Against Human Norovirus Surrogates. Food Environ. Virol. 5, 97–102 (2013).
20. Lulu, S. S., Thabitha, A., Vino, S., Priya, M. A. & Rout, M. Naringenin reduces inflammatory pain in mice. Neuropharmacology 105, 508–519 (2016).
21. Pinho-Ribeiro, F. A. et al. Naringenin reduces inflammatory pain in mice. Neuropharmacology 7668–7674 (2010).
22. Pinho-Ribeiro, F. A. et al. Antiviral Res 30(4), 464–8 (2016).
23. Pinho-Ribeiro, F. A. et al. Flavonoid baicalin inhibits HIV-1 infection at the level of viral entry. Antiviral Res 87(1), 70–76 (2009).
24. Cotrin, S. et al. Eight flavonoids and their potential as inhibitors of human cytomegalovirus replication. Antiviral Res. 96, 181–6 (2012).
25. Repetto, G., del Peso, A. & Zurita, J. L. Neutral red uptake assay for the estimation of cell viability/cytotoxicity. Nat. Protoc. 3, 1125–31 (2008).
26. Koishi, A. C., Zanello, P. R., Bianco, É. M. & Bordignon, J. & Nunes Duarte dos Santos, C. Screening of Dengue Virus Antiviral Activity of Marine Seaweeds by an In Situ Enzyme-Linked Immunoassorbent Assay. PLoS One 7 (2012).
27. Suzuki, R., de Borba, L., Duarte dos Santos, C. N. & Mason, P. W. Construction of an infectious cDNA clone for a Brazilian prototype strain of dengue virus type 1: Characterization of a temperature-sensitive mutation in NS1. Virology 362, 374–383 (2007).
28. Mosimann, A. L. P., de Borba, L., Bordignon, J., Mason, P. W. & Santos, C. N. D. os. Construction and characterization of a stable subgenomic replicon system of a Brazilian dengue virus type 3 strain (BR DENV 290-02). J. Virol. Methods 163, 147–152 (2010).
29. Li, R. Q. et al. Flavonoid baicalin inhibits HIV-1 infection at the level of viral entry. Biochem. Biophys. Res. Commun. 276, 534–538 (2000).
30. Lani, R. et al. Antiviral activity of selected flavonoids against Chikungunya virus. Antiviral Res. 133, 50–61 (2016).
31. Pohjala, L. et al. Inhibitors of alphavirus entry and replication identified with a stable Chikungunya replicon cell line and virus-based assays. PLoS One 6 (2011).
32. Kachatoorian, R. et al. Divergent antiviral effects of bioflavonoids on the hepatitis C virus life cycle. Virology 433, 346–355 (2012).
33. Zanello, P. R. et al. Quinic acid derivatives inhibit dengue virus replication in vitro. J. Virol. 1, 1–13, doi: 10.1186/12285-015-0443-9 (2015).
34. de Burghgraeven, T. et al. An analogue of the antibiotic teicoplanin prevents flavivirus entry in vitro. PLoS One 7, 1–9 (2012).
35. Kato, F. et al. Novel antiviral activity of bromocriptine against dengue virus replication. Antiviral Res. 131, 141–147 (2016).
36. de Sousa, L. R. F. et al. Flavonoids as noncompetitive inhibitors of Dengue virus NS2B-NS3 protease: Inhibition kinetics and docking studies. Bioorg. Med. Chem. 23, 466–470 (2015).
37. Lim, H. et al. Inhibitory effect of flavonoids against NS2B-NS3 protease of ZIKV virus and their structure activity relationship. Biotechnol. Lett. In press, doi: 10.1007/s10529-016-2261-6 (2016).
38. Dong, W. et al. A dual character of flavonoids in influenza A virus replication and spread through modulating cell-autonomous immunity by MAPK signaling pathways. Sci. Rep. 4, 7237 (2014).
39. Isaacs, A. & Lindenmann, J. Virus interference. I. The interferon. Proc. R. Soc. London. Ser. B, Biol. Sci. 147, 258–67 (1957).
40. Wang, Q. Y. et al. A translation inhibitor that suppresses dengue virus in vitro and in vivo. Antimicrob. Agents Chemother. 55, 4072–4080 (2011).
41. Xie, X. et al. Inhibition of dengue virus by targeting viral NS4B protein. J. Virol. 85, 11183–95 (2011).
42. Brandt, W. E., McCown, J. M., Top, F. H., Bancroft, W. H. & Russell, P. K. Effect of passage history on dengue-2 virus replication in subpopulations of human leukocytes. Infect. Immun. 26, 534–541 (1979).
43. Kuczera, D. et al. Isolation of dengue virus serotype 4 genotype II from a patient with high viral load and a mixed Th1/Th17 inflammatory cytokine profile in South Brazil. Virol. J. 13, 93 (2016).
44. Gould, E. A. & Clegg, C. Growth, titration and purification of togaviruses virology: a practical approach in Virology: a practical approach. (1985).
45. Lanzetti, R. S., Calisher, C. H., Gubler, D. I., Chang, G. J. & Vorndam, A. V. Rapid detection and typing of dengue viruses from clinical samples by using reverse transcriptase–polymerase chain reaction. J. Clin. Microbiol. 30, 545–551 (1992).
46. Büyum, A. Isolation of mononuclear cells and granulocytes from human blood. Isolation of mononuclear cells by one centrifugation, and of granulocytes by combining centrifugation and sedimentation at 1 g. Scand. J. Clin. Lab. Invest. Suppl. 97, 77–89 (1968).
47. Silveira, G. F. et al. Dengue virus type 3 isolated from a fatal case with visceral complications induces enhanced proinflammatory responses and apoptosis of human dendritic cells. J. Virol. 85, 5374–83 (2011).

Acknowledgements
The authors thank the Program for Technological Development in Tools for Health-PDTIS-FIOCRUZ for the use of their facilities (RPT07C; Microscopy Facility and RPT08 L; Flow Cytometry Facility at the Carlos Chagas Institute/Fiocruz-PR, Brazil). The authors thank CNPq (PROCAD/Casadinho, Universal/2013, PAPES/Fiocruz) and the Brazilian Ministry of Health (PPSUS-2013)/Fundação Araucária for their financial support. CNDS and WAVJr are CNpq scholars, and JB is a Fundação Araucária scholar.

Author Contributions
S.F., A.C.K., D.K. and G.F.S. carried out the experiments and performed statistical analysis. S.F., A.C.K., C.N.D.S., W.A.VJr., J.B. design the experiments and the conception of the study. S.F., A.C.K., C.N.D.S., W.A.VJr., J.B. drafted and review the manuscript. All authors have read and approved the final manuscript.

Additional Information
Supplementary information accompanies this paper at http://www.nature.com/srep

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Frabasile, S. et al. The citrus flavanone naringenin impairs dengue virus replication in human cells. Sci. Rep. 7, 41864; doi: 10.1038/srep41864 (2017).

Publisher’s note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.
Corrigendum: The citrus flavanone naringenin impairs dengue virus replication in human cells

Sandra Frabasile, Andrea Cristine Koishi, Diogo Kuczera, Guilherme Ferreira Silveira, Waldiceu Aparecido Verri Jr., Claudia Nunes Duarte dos Santos & Juliano Bordignon

Scientific Reports 7:41864; doi: 10.1038/srep41864; published online 03 February 2017; updated on 14 March 2017

In the original version of this Article, Claudia Nunes Duarte dos Santos and Juliano Bordignon were incorrectly affiliated with 'Sección Virologia, Facultad de Ciencias, Universidad de La República, 11400, Montevideo, Uruguay'. The correct affiliations are listed below:

Claudia Nunes Duarte dos Santos
Laboratório de Virologia Molecular, Instituto Carlos Chagas, ICC/FIOCRUZ/PR, Curitiba, Paraná, Brazil

Juliano Bordignon
Laboratório de Virologia Molecular, Instituto Carlos Chagas, ICC/FIOCRUZ/PR, Curitiba, Paraná, Brazil

These errors have now been corrected in the PDF and HTML versions of the Article.

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/

© The Author(s) 2017