Effects of Low-Level Autonomic Stimulation on Prevention of Atrial Fibrillation Induced by Acute Electrical Remodeling

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Background. Rapid atrial pacing (RAP) can induce electrical and autonomic remodeling and facilitate atrial fibrillation (AF). Recent reports showed that low-level vagosympathetic nerve stimulation (LVNS) can suppress AF, as an antiarrhythmic effect. We hypothesized that LVNS can reverse substrate heterogeneity induced by RAP. Methods and Results. Mongrel dogs were divided into (LVNS+RAP) and RAP groups. Electrode catheters were sutured to multiple atrial sites, and LVNS was applied to cervical vagosympathetic trunks with voltage 50% below the threshold slowing sinus rate by ≤30 msec. RAP induced a significant decrease in effective refractory period (ERP) and increase in the window of vulnerability at all sites, characterized by descending and elevated gradient differences towards the ganglionic plexi (GP) sites, respectively. The ERP dispersion was obviously enlarged by RAP and more significant when the ERP of GP-related sites was considered. Recovery time from AF was also prolonged significantly as a result of RAP. LVNS could reverse all these changes induced by RAP and recover the heterogeneous substrate to baseline. Conclusions. LVNS can reverse the electrical and autonomic remodeling and abolish the GP-central gradient differences induced by RAP, and thus it can recover the homogeneous substrate, which may be the underlying mechanism of its antiarrhythmic effect.

1. Introduction

Atrial fibrillation (AF) is the most common cardiac rhythm disorder and contributes to thromboembolism. The presence of AF is an independent risk factor for thromboembolism; especially stroke in association with AF increases mortality and morbidity, leading to greater disability, longer hospital stays, and worse quality of life [1, 2]. AF affects approximately 10 million people in China alone [3]. AF is a complex disease that is initiated by a specific trigger and maintained in the presence of a vulnerable substrate. The autonomic nervous system (ANS), especially the vagosympathetic/vagal nerve, is an important participant in the mechanism of AF initiation.

The cardiac ANS consists of extrinsic and intrinsic components [4, 5]. The extrinsic component comprises cervical and thoracic stellate ganglia, which contribute to complex sympathetic innervation of the heart. Most of the vagosympathetic trunks of the extrinsic system contain vagal nerve fibers; they serve as parasympathetic innervation and converge at the epicardial fat pads around the superior vena cava, pulmonary veins (PVs), and the aorta, postulated to be “integration centers.” The extrinsic component modulates
cardiac electrophysiology and the inducibility of AF [6, 7]. The intrinsic cardiac ANS is a complex neural network composed of ganglionated plexi (GP) concentrated within epicardial fat pads, and interconnected ganglia and axons [8–10]. Stimulation of GP can convert the firing from PVs into AF in dogs and in humans [11–13], then promote maintenance of AF due to markedly reduced atrial effective refractory period (ERP), widen the window of vulnerability (WOV), and increase gradient differences in ERP and WOV towards GP, which serve as heterogeneous substrate caused by GP hyperactivity [14]. This mechanism is largely based on the observation that vagosympathetic nerve stimulation (VNS) can greatly facilitate initiation and maintenance of AF by activating the “integration centers of GP” [7, 15]. Once initiated, AF leads to atrial remodeling, which involves electrical and structural remodeling and even promotes autonomic remodeling; this facilitates maintenance and recurrence of AF.

Ablation of the autonomic substrate suppresses or eliminates focal AF originating from PVs [11]. In particular, GP ablation can result in successful autonomic denervation, thus prevent the induction of AF, attenuate complex fractionated atrial potentials, and suppress vagally induced AF [16–18]. Until recently, the success rate of a single or multiple circumferential PV ablations for paroxysmal AF was 70–90% (followup for about 1 year) [19], and refractory AF was reported in some of the patients. Moreover, although GP ablation significantly reduced the vagal stimulation effect, AF inducibility was strongly augmented by vagal stimulation 4 weeks after GP ablation [20]. It was also reported that, 8 weeks after ARGP and IRGP ablation, AF was induced easily by atrial RAP [21]. Therefore, the long-term effects of GP ablation are debatable. Although the autonomic effect is eliminated after GP ablation, atrial autonomic intervention remodeling and neurohormonal disturbances in the atrium may also be responsible for AF vulnerability: for example, atrial natriuretic peptide, a substrate for arrhythmia, is associated with shortened atrial conduction time and effective refractory period (ERP) [21]. Therefore, there is a need for new therapeutic strategies.

Low-level VNS (LLVNS) is being explored as another strategy. Strong-level VNS that produces >60% prolongation of the sinus cycle length facilitates AF; it produces long pauses and even sinus arrest, which are generally required to induce and maintain AF [16, 22–24]. However, moderate-level VNS (MLVNS) that produces <40% prolongation of the sinus cycle length appears not to induce AF. It has been reported that MLVNS that slowed the sinus rate by 10% for 4 weeks did not induce AF; and was not associated with arrhythmicogenic risk [23]. Some recent evidence suggests that LLVNS, with voltage levels 10–50% below threshold, which does not slow the sinus rate, has an antiarrhythmic effect [25–28].

Based on all these reported results, we hypothesized that the antiarrhythmic effect of LLVNS is brought about by elimination of the heterogeneous substrate surrounding the GP areas, which is induced by rapid atrial pacing (RAP). This study was specifically designed to test the previous hypothesis in dogs with acute electrical remodeling.

2. Methods

2.1. Animal Preparation. The Institutional Animal Care and Use Committee of JINAN University of Experimental Animal Management Centre reviewed and approved the design of all animal experiments. All animal studies were reviewed and approved by the animal experimental administration of JINAN University of China. A total of 20 adult mongrel dogs weighing 13–17 kg were anesthetized with sodium pentobartal (initial bolus, 30 mg/kg body weight, i.v.), with an additional dose of 2 mg/kg given at the end of each hour. All dogs were ventilated with room air using a positive pressure respirator, and oxygen saturation was maintained at 95–100%. The animals were fixed on an operating table, the temperature of which thermostatically controlled at 37°C. The chest was entered via bilateral thoracotomy at the bilateral fourth intercostal space. Several 10-bipolar electrodes were sutured using a noninjurious method to allow recording and stimulation at the left superior pulmonary vein (LSPV), left inferior pulmonary vein (LIPV), left appendage (LAA, catheter: LA1,2), left atrium (LA, catheter: LA5,6), area around the Marshall ligament (MSL, catheters: LA9,10) [15,29], right superior pulmonary vein (RSPV), right inferior pulmonary vein (RIPV), right appendage (RAA, catheter: RA1,2), right atrium (RA, catheter: RA5,6), and margin of the anterior right ganglionated plexi (ARGPM, catheter: RA9,10) (Figure 1). The capture threshold of the electrodes was tested and stabilized at 1–2V. The incisions were sutured, and then the thoracic cavity was pumped till negative pressure was achieved and closed completely.

Standard ECG and electrophysiological channels (IECG) were continuously recorded and filtered at 0.05–100 Hz and 200–1200 Hz, respectively. All tracings from the electrode catheters were amplified and digitally recorded using a computer-based electrophysiological system (Lead2000B; Jingjiang Inc., China).

2.2. Programmed Stimulation. In both groups, RAP was delivered (1200 bpm, 4 V, 1 msec in duration) at the LA. After each pacing hour, RAP was temporarily stopped to measure the electrophysiological data.

Programmed stimulation was performed with Lead2000B stimulator. Regular atrial pacing as $S_1-S_1$ interval was set at 330 msec. ERP was determined using $S_2-S_2$ programmed stimulation ($S_1-S_1 : S_2-S_2 = 8:1$), which decreased from 150 msec by 10 msec decrements; as the $S_1-S_2$ intervals approached the ERP, the last $S_1-S_2$ interval increased by 8 msec, and the decrement was reduced by 1 msec again, until precise ERP was reached. ERP dispersion (ERPD) was defined as the coefficient of variation (standard deviation/mean) of the ERP at all 8 or 10 sites [30], and was calculated using MATLAB-R2008. ERPD was calculated for RSPV, RIPV, RA, RAA, LSPV, LIPV, LA, and LAA. When adding the ERP of ARGPM and MSL into calculation together with the previous 8 sites, we would get the values of ERPD2.

The WOV was used as a quantitative measure of AF inducibility. During ERP measurements, if AF or >2 echo was induced by decremental $S_1-S_2$ stimulation, the difference
between the longest and the shortest $S_1$-$S_2$ interval was designated as the WOV. The $\sum$ WOV was counted as the sum of WOV at each site [14].

The stimulation of the atrium was delivered at the LA site with $S_1$-$S_1$ intervals set at 100 msec and sustained for 10 seconds, which induced atria tachycardia or AF easily when it was continued or stopped. The recovery time (RT) was represented as the duration from AF triggered by persistent $S_1$-$S_1$ stimulation to sinus rhythm.

2.3. Low-Level Vagosympathetic Nerve Stimulation. Bilateral cervical vagosympathetic trunks were decentralized by surgical procedures. Two pairs of electrodes insulated with surrounding tissue were embedded in the vagosympathetic trunks located adjacent to the cervical artery for LLVNS [31]. LLVNS was induced by applying high-frequency electrical stimulation (HFS, 20 Hz, 0.1 msec in duration, square waves, voltage 0.1–0.5 V) to vagosympathetic nerves via an electronic stimulator (BL-420E Experimental System; Tai Meng, Chengdu, China) (Figure 2). The lowest voltage of VNS that prolonged the cycle length of sinus rhythm (A-A intervals) by no more than 30 msec was considered the threshold. The LLVNS voltage was approximately 50% below the threshold. Prior to each hour of LLVNS, the threshold of VNS was determined again to adjust the LLVNS voltages for the next hour [25, 26, 28].

2.4. Experimental Protocol. Twenty dogs were randomly divided into the experimental group ($n = 10$), which underwent RAP remodeling concomitant with LLVNS for 3 hours (Figure 2), and the control group ($n = 10$), which only underwent RAP remodeling for 3 hours. In the experimental group, LLVNS was delivered to bilateral cervical vagosympathetic nerves by applying HFS as described previously. The ERP and WOV were measured at LSPV, LIPV, LA, LAA, MSL, RSPV, RIPV, ARGPM, RA, and RAA at baseline levels and the end of every hour during the 3 hours pacing period.

2.5. Statistical Analysis. All values were expressed as the mean ± standard deviation of the mean. Paired $t$-test was used for comparisons of ERP and WOV of baseline levels and each pacing period. Analysis of variance (ANOVA) by SPSS20 was used to compare ERP, WOV, ERP dispersion, and RT in both groups, and graphs were drawn with Microsoft Excel2010. $P$ values $\leq 0.05$ were considered to indicate statistical significance.

3. Results

The systolic and diastolic blood pressures were stable during the entire experimental period, with no sign of heart failure throughout. Oxygen saturation was maintained at 95–100% during the experiments.

3.1. Effective Refractory Period. In the control group, the ERP at RSPV, RIPV, RA, RAA, ARGPM, LSPV, LIPV, LA, LAA, and MSL was markedly shortened in the second and third hour (Figure 3). The decrease in ERP at ARGPM,
3.2. Window of Vulnerability. In the control group, the WOV at each site of the atrium showed a significantly progressive increase during each hour of RAP (Figure 4). Moreover, a significant WOV gradient was observed in the atrium, and gradients were not observed at the end of each hour, as observed at the baseline.

MSL, RSPV, RIPV, and LSPV was more significant compared to other sites of the atrium. Moreover, there were apparent ERP gradients in the atrium at the second and third hour.

In contrast, in the experimental group, the ERP of each site of the atrium showed no significant variance at the end of each hour, compared to baseline levels. Moreover, ERP
3.3. Dispersion of the Effective Refractory Period. In the control group, ERPD1 and ERPD2 showed progressive increase and reached statistical significance in the second and third hour of RAP (Figure 5). In contrast, the experimental group showed no significant difference in ERPD1 and ERPD2 for each RAP period. Moreover, ERPD2 was more obviously enlarged than ERPD1 in the control group, but there was no significant difference between ERPD1 and ERPD2 in the experimental group during each period.

3.4. Recovery Time and Low-Level Voltage. The recovery time (RT) was represented as the duration from atrial arrhythmia that is facilitated by fast pacing triggers in atrium to sinus rhythm. In control group, RAP remodeling progressively increased RT (Figure 6). While RAP was persistent for 3 hours, the RT was obviously longer than that of baseline (5.078 ± 2.266 seconds versus 1.886 ± 1.059 seconds, \( P < 0.01 \)). In experimental group, combining RAP with LLVNS for 3 hours, the RT at each period has no difference from that of baseline levels. During the judgments of threshold, the voltage at 0.5 V that slowed the sinus rate apparently can lead to atrial premature and then facilitate AF at baseline (Figure 7). The AF was persistent during the VNS. While the VNS was removed, AF still maintained (Figure 8). When RAP that is persistent for 3 hours concomitant with LLVNS (0.25 V) was stopped, the AF progressively terminated even though the LLVNS still continued (Figure 9). The low-level voltages for LLVNS that is 50% below the threshold that prolonged the A-A intervals of sinus rhythm by no more than 30 msec had not changed, prior to the first, second, and third remodeling (Pre-RMD 1h, 0.154 ± 0.058 V; Pre-RMD 2h, 0.22 ± 0.071 V; Pre-RMD 3h, 0.225 ± 0.115 V, \( P > 0.05 \)).

4. Discussion

We have successfully proved our hypothesis about the benefits of LLVNS for treating AF. This is clear from the results, which show that application of LLVNS to canine models of RAP remodeling markedly reversed the decrease in ERP and increase in WOV at each site. Moreover, it abolished gradient differences in the electrophysiological substrate surrounding
GP and thus maintained ERP dispersion at baseline levels. It also reduced the RT from AF.

In our research, we measured the threshold, the VNS with the voltage at 0.5 V triggered atrial premature and then facilitated AF at baseline. The AF was persistent during the VNS. While the VNS was removed, AF still maintained. These results suggest that vagosympathetic trunks as extrinsic components play an important role in the initiation and maintenance of AF. The arrhythmic effect of VNS may be mediated by activating the “integration centers of GP” [7, 15].

The hyperactivity of the autonomic element in MSL may contribute to the initiation of AF and even ventricular tachyarrhythmia [32]. It is reported that MSL participates in the interaction between integration centers and GP [15]. High-level stimulation of GP can progressively reduce atrial ERP, widen the WOV, and lead to gradient differences in ERP and WOV towards GP [14], and it is even associated with complex fractionated atrial potentials [33]. Similar to these reports, in our study too, RAP remodeling for 2-3 hours led to a significant decrease in ERP and caused an obvious decrease in the ERP gradient from the appendages towards PV, MSL, and ARGP, which serve as GP areas. Moreover, ERP dispersion progressively increased and reached a significant difference at the end of RAP, and when the ERP dispersion values for ARGP and MSL were added, ERP dispersion was further augmented. The WOV progressively increased during RAP, and it was noted that the increase in WOV at PV, MSL, and ARGP was more apparent, which caused an increase in the gradient difference from the non-GP area towards the GP sites. Based on these results, we concluded that the ERP and WOV gradients were reflective of the electrophysiological substrate surrounding GP, since autonomic remodeling is characterized by hyperactive GP caused by RAP.

In the RAP remodeling, when electrical remodeling occurs concomitantly with autonomic remodeling, it is believed to indicate progressive enhancement of neural activity [34]; in this study, heterogeneous differences in both ERP and WOV were observed at atrial sites, especially around the GP, so it is possible that RAP resulted in obvious hyperactivity of GP and thus increased vulnerability to AF because of the increased heterogeneity of the substrate.

It has been reported that LLVNS that is 10–50% below the threshold voltage, required to slow the sinus rate or atrioventricular conduction, may prevent episodic AF caused by rapid PV and non-PV firing, due to a progressive increase in AF threshold at all PVs and atrial appendages sites, particularly RSPV, RIPV, LSPV, and RAA. Moreover, this type of antiarrhythmic effect is not dependent on the activation of the aferent vagal nerve fibers that project to the brain [28]. The activation of neural elements within the ARGP, SLGP, and stellate ganglion was attenuated by LLVNS application to both [28, 35] or one vagosympathetic trunk [26, 27], auricular branch of the vagus nerve of the right ear (low-level tragus stimulation) [36], or superior vena cava [34]. Therefore, the antiarrhythmic effect of LLVNS may be brought about via its action on GP sites. When the voltage of LLVNS was further reduced to 80% below the threshold, it still induced a similarly antiarrhythmic effect [36], LLVNS at these values eliminated the spatial gradient of ERP and WOV induced by ARGP stimulation from the PV-atrial junction toward the atrial appendage [25]. Moreover, it could prevent and reverse atrial remodeling induced by RAP as well as suppress AF induced by strong cholinergic stimulation, for example, by injection of acetylcholine (10 mM) into the ARGP or RAA.

The duration and cycle length of AF decreased obviously during LLVNS [35]. In contrast, stimulation of ARGP and SLGP for 6 hours with a voltage that causes a 10% decrease in sinus rate does not appear to have an antiarrhythmic effect [37]. In our study, we used LLVNS values at 50% below the threshold (which slowed the sinus rate by no more than 30 msec). We found that this reversed the electrical and autonomic remodeling, restoring ERP and WOV to baseline levels. LLVNS maintained the two kinds of ERP dispersion at baseline levels. Also, the ERP and WOV around ARGP, PV, and MSL were not significantly different from those in other areas. Thus, LLVNS resulted in substrate homogeneity. Thus, they are in agreement with previously published results. Based on our current study and previous studies, we think that hyperactivation of intrinsic nerves or “integration centers” was suppressed by LLVNS. It brought about this effect by abolishing gradient differences in the substrate surrounding GP, which contributed to the homogeneity of ERP and WOV in the atrium. More importantly, the duration of AF promoted by fast-pacing triggers at the end of each period was significantly decreased. During the process of RAP and LLVNS at the third period, when the RAP was removed, the AF induced by RAP recovered to sinus rhythm immediately, even though LLVNS was still continued.

Based on our results, the mechanism of action of LLVNS may be explained as follows: (1) suppression of the neural activity of GP, which abolishes gradient differences in the substrate surrounding GP and (2) enhanced recovery of the homogeneous substrate with baseline susceptibility to AF. In addition, there are other mechanisms that have been proposed by researchers: (3) decrease in transient intracellular Ca$^{2+}$ levels, owing to reduced release of sympathetic neurotransmitters via inhibition of activity of the stellate ganglion [27, 38], (4) desensitization of autonomic receptors, such as β-adrenergic receptors [25, 39, 40], (5) increase...
in nitric oxide availability mediated by upregulation of the PI3K/NO signaling pathway [41], which subsequently leads to inhibition of GP function, (6) increase in the levels of neuropeptides or neurotransmitters inhibiting GP function, for instance the neuropeptide Y2 and vasostatin-1, which exert strong antiadrenergic and anticholinergic effects [41–43]. Future studies should be conducted to integrate and explain these mechanisms in relation to one another.

A limitation of this study is that we have no direct evidence indicating inhibition of neuronal firing within GP and MSL, and the results are therefore limited to short-term RAP and LLVNS. This hypothesis will have to be verified in the future by using RAP and LLVNS of longer duration. This will help evaluate the antiarrhythmic effects of LLVNS in chronic AF models. Moreover, research will be required to determine the optimal parameters and sites for LLVNS that will have the greatest degree of AF inhibition with minimal side effects. We hope to devise future treatment methods to treat chronic AF, in which autonomic nerve stimulators can be inserted using noninvasive methods.

5. Conclusions

LLVNS can reverse the electrical and autonomic remodeling induced by RAP. The mechanism involves abolishing gradient differences in the substrate surrounding GP and recovery of the homogeneous substrate.

Disclosure

The authors take responsibility for all aspects of the reliability and freedom from bias of the data presented and their discussed interpretation.

Conflict of Interests

The authors declare that they have no conflict of interests.

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References

[1] T. Watson, E. Shantsila, and G. Y. Lip, “Mechanisms of thrombogenesis in atrial fibrillation: virchow’s triad revisited,” *The Lancet*, vol. 373, no. 9658, pp. 155–166, 2009.

[2] C. Steger, A. Pratter, M. Martinke-Bregel et al., “Stroke patients with atrial fibrillation have a worse prognosis than patients without: data from the Austrian Stroke registry,” *European Heart Journal*, vol. 25, no. 19, pp. 1734–1740, 2004.

[3] C. Ma and W. Qi, “Management of atrial fibrillation in Chinese patients,” *CVD Prevention and Control*, vol. 4, no. 1, pp. 79–83, 2009.

[4] J. A. Armour, “Cardiac neuronal hierarchy in health and disease,” *American Journal of Physiology*, vol. 287, no. 2, pp. R262–R271, 2004.

[5] J. A. Armour, “Functional anatomy of intrathoracic neurons innervating the atria and ventricles,” *Heart Rhythm*, vol. 7, no. 7, pp. 994–996, 2010.

[6] C.-W. Chiou, J. N. Eble, and D. P. Zipes, “Efferent vagal innervation of the canine atria and sinus and atrioventricular nodes: the third fat pad,” *Circulation*, vol. 95, no. 11, pp. 2573–2584, 1997.

[7] Y. Hou, B. J. Scherlag, J. Lin et al., “Ganglionated plexi modulate extrinsic cardiac autonomic nerve input. effects on sinus rate, atrioventricular conduction, refractoriness, and inducibility of atrial fibrillation,” *Journal of the American College of Cardiology*, vol. 50, no. 1, pp. 61–68, 2007.

[8] J. A. Armour, D. A. Murphy, B. X. Yuan, S. Macdonald, and D. A. Hopkins, "Gross and microscopic anatomy of the human intrinsic cardiac nervous system," *The Anatomical Record*, vol. 247, no. 2, pp. 289–298, 1997.

[9] B.-X. Yuan, J. L. Ardell, D. A. Hopkins, A. M. Losier, and J. A. Armour, "Gross and microscopic anatomy of the canine intrinsic cardiac nervous system," *The Anatomical Record*, vol. 239, no. 1, pp. 75–87, 1994.

[10] D. H. Pauza, V. Skripka, and N. Pauziene, "Morphology of the intrinsic cardiac nervous system in the dog: a whole-mount study employing histochemical staining with acetylcholinesterase," *Cells Tissues Organs*, vol. 172, no. 4, pp. 297–320, 2002.

[11] Z. Lu, B. J. Scherlag, J. Lin et al., "Autonomic mechanism for initiation of rapid firing from atria and pulmonary veins: evidence by ablation of ganglionated plexi," *Cardiovascular Research*, vol. 84, no. 2, pp. 245–252, 2009.

[12] B. J. Scherlag, W. Yamanashi, U. Patel, R. Lazzara, and W. M. Jackman, "Autonomically induced conversion of pulmonary vein focal firing into atrial fibrillation," *Journal of the American College of Cardiology*, vol. 45, no. 11, pp. 1878–1886, 2005.

[13] P. B. Lim, L. C. Malcolm-Lawes, T. Stub et al., "Intrinsic cardiac autonomic stimulation induces pulmonary vein ectopy and triggers atrial fibrillation in humans," *Journal of Cardiovascular Electrophysiology*, vol. 22, no. 6, pp. 638–646, 2011.

[14] J. Zhou, B. J. Scherlag, J. Edwards, W. M. Jackman, R. Lazzara, and S. S. Po, "Gradients of atrial refractoriness and inducibility of atrial fibrillation due to stimulation of ganglionated plexi," *Journal of Cardiovascular Electrophysiology*, vol. 18, no. 1, pp. 83–90, 2007.

[15] J. Lin, B. J. Scherlag, G. Niu et al., "Autonomic elements within the ligament of marshall and inferior left ganglionated plexus mediate functions of the atrial neural network," *Journal of Cardiovascular Electrophysiology*, vol. 20, no. 3, pp. 318–324, 2009.

[16] K. Lemola, D. Chartier, Y.-H. Yeh et al., "Pulmonary vein region ablation in experimental vagal atrial fibrillation : role of pulmonary veins versus autonomic ganglia," *Circulation*, vol. 117, no. 4, pp. 470–477, 2008.

[17] C. Pappone, V. Santinelli, F. Manguso et al., "Pulmonary vein denervation enhances long-term benefit after circumferential ablation for paroxysmal atrial fibrillation," *Circulation*, vol. 109, no. 3, pp. 327–334, 2004.

[18] Z. Lu, B. J. Scherlag, J. Lin et al., "Autonomic mechanism for complex fractionated atrial electrograms: evidence by fast Fourier transform analysis," *Journal of Cardiovascular Electrophysiology*, vol. 19, no. 8, pp. 835–842, 2008.

[19] J. P. Piccini, R. D. Lopes, M. H. Kong, V. Hasselblad, K. Jackson, and S. M. Al-Khatib, "Pulmonary vein isolation for the maintenance of sinus rhythm in patients with atrial fibrillation a meta-analysis of randomized, controlled trials," *Circulation: Arrhythmia and Electrophysiology*, vol. 2, no. 6, pp. 626–633, 2009.

[20] S. Oh, Y. Zhang, S. Bibeviski, N. F. Marrouche, A. Natale, and T. N. Mazgalev, "Vagal denervation and atrial fibrillation inducibility: epicardial fat pad ablation does not have long-term effects," *Heart Rhythm*, vol. 3, no. 6, pp. 701–708, 2006.

[21] Q. Y. Zhao, H. Huang, S. D. Zhang et al., "Atrial autonomic innervation remodelling and atrial fibrillation inducibility after epicardial ganglionic plexi ablation," *Europace*, vol. 12, no. 6, pp. 805–810, 2010.

[22] P. Schauerte, B. J. Scherlag, J. Pitha et al., "Catheter ablation of cardiac autonomic nerves for prevention of vagal atrial fibrillation," *Circulation*, vol. 102, no. 22, pp. 2774–2780, 2000.

[23] Y. Zhang, I. Ilsar, H. N. Sabbah, T. Ben David, and T. N. Mazgalev, "Relationship between right cervical vagus nerve stimulation and atrial fibrillation inducibility: therapeutic intensities do not increase arrhythmogenesis," *Heart Rhythm*, vol. 6, no. 2, pp. 244–250, 2009.

[24] R. Nadeau, R. Cardinal, J. A. Armour et al., "Cervical vagosympathetic and mediastinal nerves activation effects on atrial arrhythmia formation," *Anadolu Kardiyoloji Dergisi*, vol. 7, no. 1, pp. 34–36, 2007.

[25] L. Yu, B. J. Scherlag, S. Li et al., "Low-level vagosympathetic nerve stimulation inhibits atrial fibrillation inducibility: direct evidence by neural recordings from intrinsic cardiac ganglia," *Journal of Cardiovascular Electrophysiology*, vol. 22, no. 4, pp. 455–463, 2011.

[26] Y. Sha, B. J. Scherlag, L. Yu et al., "Low-level right vagal stimulation: anticholinergic and antiadrenergic effects," *Journal of Cardiovascular Electrophysiology*, vol. 22, no. 10, pp. 1147–1153, 2011.
[27] M. J. Shen, T. Shinohara, H.-W. Park et al., “Continuous low-level vagus nerve stimulation reduces stellate ganglion nerve activity and paroxysmal atrial tachyarrhythmias in ambulatory canines,” Circulation, vol. 123, no. 20, pp. 2204–2212, 2011.

[28] S. Li, B. J. Scherlag, L. Yu et al., “Low-level vagosympathetic stimulation a paradox and potential new modality for the treatment of focal atrial fibrillation,” Circulation: Arrhythmia and Electrophysiology, vol. 2, no. 6, pp. 645–651, 2009.

[29] D. T. Kim, A. C. Lai, C. Hwang et al., “The ligament of Marshall: a structural analysis in human hearts with implications for atrial arrhythmias,” Journal of the American College of Cardiology, vol. 36, no. 4, pp. 1324–1327, 2000.

[30] Z. Lu, B. J. Scherlag, J. Lin et al., “Atrial fibrillation begets atrial fibrillation: autonomic mechanism for atrial electrical remodeling induced by short-term rapid atrial pacing,” Circulation: Arrhythmia and Electrophysiology, vol. 1, no. 3, pp. 184–192, 2008.

[31] D. Chang, S. Zhang, D. Yang et al., “Effect of epicardial fat pad ablation on acute atrial electrical remodeling and inducibility of atrial fibrillation,” Circulation Journal, vol. 74, no. 5, pp. 885–894, 2010.

[32] J. Lin, B. J. Scherlag, Z. Lu et al., “Inducibility of atrial and ventricular arrhythmias along the ligament of marshall: role of autonomic factors,” Journal of Cardiovascular Electrophysiology, vol. 19, no. 9, pp. 955–962, 2008.

[33] J. Lin, B. J. Scherlag, J. Zhou et al., “Autonomic mechanism to explain complex fractionated atrial electrograms (CFAE),” Journal of Cardiovascular Electrophysiology, vol. 18, no. II, pp. 1197–1205, 2007.

[34] L. Yu, B. J. Scherlag, Y. Sha et al., “Interactions between atrial electrical remodeling and autonomic remodeling: how to break the vicious cycle,” Heart Rhythm, vol. 9, no. 5, pp. 804–809, 2012.

[35] X. Sheng, B. J. Scherlag, L. Yu et al., “Prevention and reversal of atrial fibrillation inducibility and autonomic remodeling by low-level vagosympathetic nerve stimulation,” Journal of the American College of Cardiology, vol. 57, no. 5, pp. 563–571, 2011.

[36] L. Yu, B. J. Scherlag, S. Li et al., “Low-level transcutaneous electrical stimulation of the auricular branch of the vagus nerve: a noninvasive approach to treat the initial phase of atrial fibrillation,” Heart Rhythm, vol. 10, no. 3, pp. 428–435.

[37] L. Wu, Z. Lu, H. Jiang et al., “Low-level ganglionated plexus stimulation facilitates atrial fibrillation: in vivo and in vitro studies,” Autonomic Neuroscience: Basic and Clinical, vol. 168, no. 1-2, pp. 58–65, 2012.

[38] E. Patterson, S. S. Po, B. J. Scherlag, and R. Lazzara, “Triggered firing in pulmonary veins initiated by in vitro autonomic nerve stimulation,” Heart Rhythm, vol. 2, no. 6, pp. 624–631, 2005.

[39] T. Iwao, M. Ito, N. Takahashi, T. Saikawa, and T. Sakata, “Sustained left ansae subclaviae stimulation for a 5-hour period inhibits cesium-induced ventricular arrhythmogenesis in rabbits,” Journal of Molecular and Cellular Cardiology, vol. 30, no. II, pp. 2237–2245, 1998.

[40] E. Patterson, X. Yu, S. Huang, M. Garrett, and D. C. Kem, “Suppression of autonomic-mediated triggered firing in pulmonary vein preparations, 24 hours postcoronary artery ligation in dogs,” Journal of Cardiovascular Electrophysiology, vol. 17, no. 7, pp. 763–770, 2006.

[41] S. Stavrakis, B. J. Scherlag, Y. Fan et al., “Inhibition of atrial fibrillation by low-level vagus nerve stimulation: the role of the nitric oxide signaling pathway,” Journal of Interventional Cardiac Electrophysiology, vol. 36, no. 3, pp. 199–208, 2012.