Pharmacogenetics of anticancer drugs in non-Hodgkin lymphomas

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Summary The variability of tumour responses to chemotherapeutic agents is a topic of major interest in current oncology research. Advances in the knowledge of molecular pathology of cancer make available strategies by which tumour cells can be profiled for their genetic background in order to select anticancer agents that might selectively kill cells in a molecular context that matches the mechanism of action of drugs. The next generation of anticancer treatments might thus be tailored on the basis of the numerous molecular alterations identified in tumour cells of a particular patient. However, to exploit these alterations, it is necessary to understand how they influence the cellular pathways that control the sensitivity or, conversely, resistance to chemotherapeutic agents. The aim of this article is to outline major genetic abnormalities in non-Hodgkin lymphomas that can be used to streamline anticancer drug selection and to underscore the major role of pharmacogenetics, which studies the interactions between genetic background and drug activity, to the prediction of likelihood of response and identification of potential new targets for pharmacological intervention.

Keywords: pharmacogenetics; chemosensitivity; drug resistance; non-Hodgkin lymphomas

NON-HODGKIN LYMPHOMAS: CLINICAL RELEVANCE AND THERAPEUTIC MANAGEMENT

Non-Hodgkin lymphomas (NHLs) represent a clinically heterogeneous group of malignancies arising from B, T or NK-lymphoid cells. Approximately 50 000 new cases of NHLs are diagnosed every year in the United States, with 20 500 estimated deaths, representing 4% of cancer incidence and the seventh cause of death for malignancies in the United States (Groves et al, 2000). For clinical purposes, the division into low, intermediate- and high-grade NHLs is commonly used. Low-grade NHLs are initially treated with radiation therapy or single-agent chemotherapy, including alkylators or nucleoside analogues. However, the treatment of advanced-stage, relapsed low-grade NHLs requires more aggressive combination chemotherapy, including first-generation protocols (cyclophosphamide, vincristine and prednisone (CVP) or cyclophosphamide, doxorubicin, vincristine and prednisone (CHOP)) (Fisher, 2000). The use of monoclonal antibodies, targeted against selected antigens (i.e., CD20), is associated with lower toxicity and higher response rates (Fisher, 2000). Intermediate- and high-grade NHLs are managed with radiotherapy and chemotherapy with CHOP, or second- and third-generation regimens, including MACOP-B (methotrexate, doxorubicin, cyclophosphamide, vincristine, prednisone and bleomycin), m-BACOD (methotrexate, bleomycin, doxorubicin, cyclophosphamide, vincristine and dexamethasone), or ProMACE-CytaBOM (cyclophosphamide, etoposide, cytarabine, vincristine, bleomycin, methotrexate and prednisone), resulting in long-term survival rates of more than 80% (Fisher, 2000).

Despite the advances in the knowledge of molecular pathology of NHLs, histologic criteria still represent the mainstay of classification of NHLs and the choice of systemic chemotherapy is mostly empiric. However, the genetic abnormalities of NHLs will permit in the future the outlook for individual patients to be assessed at diagnosis and treatment may be personalised, if we will be able to establish a relationship between gene expression, disease-specific molecular abnormalities and drug activity. Therefore, the present work reviews the recent advances in the field of molecular genetics of NHLs in order to describe selected molecular pathways involved in the response to anticancer drugs and how pharmacogenetics is going to play a role in rational drug choice in the future.

MOLECULAR DETERMINANTS OF DRUG SENSITIVITY AND RESISTANCE

Drug metabolism

The nucleoside analogues fludarabine, gemcitabine, 2-chloro-deoxyadenosine and cytarabine have significant activity in the treatment of NHLs. These prodrugs are phosphorylated by the rate-limiting enzyme deoxycytidine kinase to generate metabolites that inhibit ribonucleotide reductase, resulting in a decrease of endogenous deoxynucleotides with enhanced formation of active drug metabolites and incorporation into the DNA (Dumontet et al, 1999). Drug inactivation may occur by dephosphorylation of active metabolites by cellular 5'-nucleotidase, a major nucleoside dephosphorylating enzyme (Dumontet et al, 1999) or by deamination by cytidine deaminase, whose overexpression confers drug resistance in vitro to CD34 + haematopoietic progenitor cells (Schröder et al, 1996). Prolonged exposure to fludarabine, gemcitabine, 2-chloro-deoxyadenosine and cytarabine resulted in the selection of human leukaemia K562 cells displaying a high degree of cross-resistance to all deoxynucleotide analogues, depending on strong increase in cellular 5'-nucleotidase activity and deoxycytidine kinase down-regulation, resulting in poor accumulation of triphosphate metabolites.
of expression of topoisomerase I gene can be informative in planning a chemotherapeutic regimen. High levels of topoisomerase I are associated with increase in drug sensitivity; on the contrary, drug resistance of CCRF-CEM cells to topoisomerase I inhibitors results from point mutations of the gene sequence and reduction in the content of nuclear topoisomerase I (Fujimori et al, 1995). Likewise, drug resistance to topoisomerase II inhibitors, including anthracyclines and epipodophyllotoxins, is dependent on point mutations and gene deletions of topoisomerase IIα as well as low expression and alterations in subcellular distribution of the enzyme, as shown in CLL cells (Valkov and Sullivan, 1997).

Proteins encoded by the multi-drug resistance mdr1 and mdr associated (mrp) proteins induce chemoresistance in cancer cells to a large number of anticancer agents, including topoisomerase II inhibitors (anthracyclines and epipodophyllotoxins), and vinca alkaloids but not to platinum compounds or alkylating agents. On the contrary, mdr1-positive cells are more sensitive or as sensitive as non-mdr cells to cytarabine and fludarabine (Michelutti et al, 1997). Patients exposed to high doses of P-glycoprotein transportable drugs show high tumour expression of mdr1 and mrp, as a result of selective pressure of the drugs (Webb et al, 1998). In this setting, chemosensitivity may be restored by P-glycoprotein antagonists, including cyclosporine A and PSC833 (valspodar), although myelosuppression at the bone marrow precursor cell level may be enhanced, as observed with etoposide and cyclosporine A (Lum et al, 2000).

Bryostatin 1, a macrocyclic lactone isolated from the marine bryozoa Bugula neritina, is an inhibitor of protein kinase C endowed with antitumour, immune-modulating and differentiating effects on B-cell NHLs. Bryostatin 1 improves the antitumour activity of vincristine in a murine model of diffuse large cell lymphoma, possibly through the down-regulation of mdr1/P-glycoprotein and bcl-2 gene expression and up-regulation of p53 (Al-Katib et al, 1998).

It has been demonstrated that the down-regulation of p53 in human leukaemia/lymphoma cell lines resistant to vincristine, may influence mdr1 overexpression via up-regulation of Wilms tumour gene 1 (WT-1) (Hirose and Kuroda, 1998). In patients with NHLs given bifunctional alkylating agents, a significant correlation was observed between complete response to chemotherapy and low expression of glutathione-S-transferase (GST)-α (Ribrag et al, 1996). On the contrary, in patients with CLL, similar levels of GST-α, -μ and -π have been found in normal and neoplastic cells, but patients with low GST-π showed enhanced response to chlorambucil (Ribrag et al, 1996). Increased GST-α and -π isoenzyme expression was reported in CLL resistant to CHOP regimen, compared to CLL sensitive to chlorambucil, while no correlation was found between GST-π expression and P-glycoprotein positivity in CLL (Ribrag et al, 1996). Finally, low levels of the DNA repair enzyme O’-alkylguanine-DNA alkyltransferase in neoplastic T lymphocytes from patients with mycosis fungoides may be predictive of improved clinical response to the new alkylating agent temozolomide, while increased expression and activity of the enzyme represents a marker of resistance (Dolan et al, 1999).

Oncogenes and regulators of cell cycle – The Bcl-2 family

Chronic lymphocytic leukaemia cells display higher levels of bax, bcl-2 and mdm-2 than normal B cells (Johnston et al, 1997). Following incubation of CLL cells with 2-chloro-deoxyadenosine,
fludarabine and chlorambucil, an increase in p53 and mdm-2 occurs in cells with wild-type p53, but not in p53-mutated cells highly resistant to chlorambucil and nucleoside analogues, while dexamethasone and vincristine had no effect on mdm-2 (Johnston et al, 1997). Therefore, 2-chloro-deoxyadenosine, fludarabine and chlorambucil induce cytotoxicity in chronic lymphocytic leukaemia cells through a p53-dependent pathway, whereas dexamethasone and vincristine do not (Johnston et al, 1997). This finding may be explained taking into account that p53-dependent induction of apoptosis occurs as a consequence of DNA damage induced by nucleoside analogues and chlorambucil, while vincristine targets microtubules and dexamethasone affects gene transcription (Johnston et al, 1997). An overview of genetic factors involved in drug response is presented in Table 1.

The human pre-B leukaemia cell line 697 overexpressing bcl-2 exhibited strikingly prolonged survival and markedly reduced apoptotic DNA fragmentation when exposed to a large number of drugs, including dexamethasone, methotrexate, cytarabine, etoposide, vincristine, cisplatin and cyclophosphamide (Reed et al, 1994). In addition to this, bcl-2 overexpression induces resistance to nitrogen mustards and camptothecin (Walton et al, 1993), while up-regulation of bcl-xl is associated with resistance to bleomycin, cisplatin, etoposide and vincristine in FL5.12 prolymphoid human cell line (Minn et al, 1995), despite the different mechanisms of action of each agent. In FL5.12 lymphoid cells transduced with bcl-xl and bcl-2, both members provided similar protection against vincristine and vinblastine, whereas bcl-xl was more effective than bcl-2 in preserving cells against etoposide, teniposide, methotrexate, fluorouracil, hydroxyurea and cisplatin cytotoxicity (Simonian et al, 1997). These results indicate that bcl-xl and bcl-2 provide a differential protection against chemotherapy-induced cell death. Finally, the myeloma cell lines 8226, IM-9 and U266 overexpressing bcl-2 are able to survive to doxorubicin and etoposide and resume their proliferation after the drugs are removed (Tu et al, 1996).

Fludarabine down-regulates the expression of bcl-2 in peripheral blood malignant lymphocytes from patients with CD5+ B-chronic lymphocytic leukaemia and mantle cell lymphoma; in vitro, drug-induced bcl-2 down-regulation and apoptosis predicted an objective response to fludarabine (Gottardi et al, 1997). Moreover, high bcl-2/bax ratios may be predictive of a drug-resistant phenotype in B-CLL cells and modulation of these proteins by fludarabine is essential for the induction of cell death (Pepper et al, 1999).

In patients with CLL and hairy cell leukaemia (HCL), who were consecutively selected for treatment with fludarabine and 2-chlorodeoxyadenosine, respectively, bcl-2 oncoprotein expression was evaluated in marrow leukaemia cells before treatment. All samples were found to be bcl-2 positive; 83% of CLL and 100% of HCL patients were responsive to purine analogues. These findings show that bcl-2 is overexpressed in almost all cases of CLL and HCL and that bcl-2 overexpression does not predict a poor response to purine analogues in these diseases (Zaja et al, 1998).

Paclitaxel and docetaxel induce bcl-2 phosphorylation, which in turn results in increased levels of free proapoptotic bax protein, and apoptosis in follicular and Burkitt lymphomas, docetaxel being 100-fold more potent than paclitaxel. Both drugs are able to kill tumour cells, even if they express high levels of bcl-2, provided they are in G2–M cycle phase; however, phosphorylation of bcl-2 and apoptosis do not occur in resting CLL cells (Haldar et al, 1997). H9 cells derived from a T-cell lymphoma selected for resistance to high concentrations of azidothymidine (250 μM) displayed overexpression of bcl-2 and were 2 to 10-fold less sensitive to the toxic effects of cisplatin, vincristine, doxorubicin and etoposide, when compared to parental H9 cells, while they retained sensitivity to nucleoside analogues, including cytarabine (Cinatl et al, 1998).

High levels of the antiapoptotic proteins mcl-1 and bag-1 (a bcl-2-binding protein that inhibits apoptosis) have been related to failure to achieve complete remission in patients with chronic lymphocytic leukaemia treated with chlorambucil, fludarabine and 2-chloro-deoxyadenosine (Kitada et al, 1998). In vitro experiments with combinations of cyclophosphamide, fludarabine and mitoxantrone demonstrated that a decrease in mcl-1 and increase in p53 levels correlated with apoptosis in B-chronic lymphocytic leukaemia cells, while the levels of bcl-2 and bax were not modified (Bellossillo et al, 1999). The down-regulation of egr-1 (early growth response gene-1), c-myc, bcl-xl and NF-kB by curcumin, a phenolic extract of the spice turmeric, causes inhibition of cell proliferation of BKS-2 neoplastic B cells (Han et al, 1999). Microenvironmental factors, including activating anti-CD40 antibody, vascular cellular adhesion molecule-1 (VCAM-1), are epigenetic determinants of resistance to etoposide in JLP119 Burkitt lymphoma cells in vitro. Indeed, the activation of surface protein CD40 and interleukin 4 (IL-4) increases bcl-xl protein levels, while cell signalling mediated by VCAM-1 and IL-4 diminished conformational changes in bax protein and prevented the etoposide-induced release of bax from the constitutive bax–bcl-xl complex and occurrence of apoptosis (Taylor et al, 2000). These interactions provide a paradigm for epigenetically induced drug resistance in lymphoma. Overexpression of bax protein in Burkitt lymphoma cells accelerates cell death induced by camptothecin, etoposide, and vinblastine but is without effect on cisplatin and paclitaxel. These results suggest that cell death by selected anticancer drugs correlates with bax expression (Schmitt et al, 1998).

B-chronic lymphocytic leukaemia cells expressing mcl-1, X-linked inhibitor of apoptosis (antiapoptosis protein-XIAP), bag-1 and bcl-2 are highly sensitive to the cyclin-dependent kinase inhibitors, flavopiridol and 7-hydroxystaurosporine. Flavopiridol and 7-hydroxystaurosporine strongly decreased the expression of proapoptotic genes; on the contrary, expression of the proapoptotic proteins bax and bak was not affected by flavopiridol (Kitada et al, 2000). Flavopiridol-induced decrease in X-linked inhibitor of apoptosis and mcl-1 precedes apoptosis and occurs independently of caspase activation. Finally, in vitro studies on human chronic lymphocytic leukaemia cells incubated with theophylline, a methylxanthine derivative and phosphodiesterase inhibitor, as well as 2-chloro-deoxyadenosine and chlorambucil, demonstrated that bcl-2 was down-regulated and apoptosis was induced by inhibition of intracellular cyclic adenosine monophosphate (cAMP) degradation (Byrd et al, 2000).

Retinoids are able to modulate cell growth and differentiation in a wide variety of human tumour cells. All-trans retinoic acid (ATRA) inhibits proliferation of NHL-B cells; after ATRA exposure, a 50% reduction in the expression of bcl-2 protein was observed, while bax protein levels were up-regulated in ATRA-sensitive NHL-B cells (Sundaresan et al, 1997).

Cell-cycle-related factors

p53 gene mutations are associated with decreased sensitivity of human lymphoma cells to DNA-damaging agents (see Table 1). Burkitt lymphoma and lymphoblastoid cell lines, treated with etoposide, nitrogen mustards and cisplatin are arrested in G1 phase if a wild-type p53 is present, while the same agents fail to induce
Table 1 Genetic abnormalities in non-Hodgkin lymphomas affecting drug response

| Genetic abnormality | Drug sensitivity | References | Drug resistance | References |
|---------------------|------------------|------------|-----------------|------------|
| **p53 mutations**   | VCR, DEX (B-CLL) | Johnston et al, 1997 | VP-16, HN₂, CDDP (BL) | Fan et al, 1994 |
|                     | Paclitaxel, VCR (BL) | Fan et al, 1998 | CLB, F-ara, CPTs (B-CLL) | Silber et al, 1994 |
|                     |                  |            | 2-CdA, F-ara, CLB (B-CLL) | Johnston et al, 1997 |
| **p53 deletions**   |                  |            | F-ara, 2-CdA (B-CLL) | Döhner et al, 1995 |
|                     |                  |            | DOX, CTX (BL) | Schmitt et al, 1999 |
| **INK4a/ARF mutations** |                  |            | CTX (BL) | Schmitt et al, 1999 |
| **ras overexpression** | DEX, VP-16 (Thymic lymphoma) | Kobzdej et al, 2000 |                |            |
| **ras mutations**   |                  |            | DEX, DOX, melphalan (MM) | Rowley et al, 2000 |
| **bcl-2 overexpression** | Paclitaxel, docetaxel (FL, BL) | Haldar et al, 1997 | Nitrogen mustards, CPT (FL, B-DLCL) | Walton et al, 1993 |
|                     | F-ara (B-CLL, MCL) | Gottardi et al, 1997 | DEX, MTX, ARA-C, VP-16, VCR, CDDP, CTX (FL, B-DLCL) | Reed et al, 1994 |
|                     | ATRA (FL, B-DLCL) | Sundaresan et al, 1997 | DOX, VP-16 (MM) | Tu et al, 1996 |
|                     | F-ara, 2-CdA (B-CLL, HCL) | Zaja et al, 1998 | VCR, VLB (FL, B-DLCL) | Simonian et al, 1997 |
|                     |                  |            | CDDP, VCR, DOX, VP-16 (T-cell lymphoma) | Cinat et al, 1998 |
|                     |                  |            | DEX (Thymic lymphoma) | Kobzdej et al, 2000 |
| **Mcl-1 overexpression** | F-ara, CTX, DHAD (B-CLL) | Bellosillo et al, 1999 | CLB, F-ara, 2-CdA (B-CLL) | Kitada et al, 1998 |
| **Bag-1 overexpression** |                  |            |               | Kitada et al, 1998 |
| **Bcl-xl overexpression** |                  |            | CLB, F-ara, 2-CdA (B-CLL) | Kitada et al, 1998 |
| **bax overexpression** | ATRA (FL, B-DLCL) | Sundaresan et al, 1997 | BLM, CDDP, VP-16, VCR (FL, B-DLCL) | Minn et al, 1995 |
|                     | CPT, VP-16, VBL (BL) | Schmitt et al, 1998 | VCR, VLB, VP-16, VM-26, MTX, hydroxyurea, CDDP, fluorouracil (FL, B-DLCL) | Simonian et al, 1997 |
|                     |                  |            | VP-16 (BL) | Taylor et al, 2000 |
| **c-myc overexpression** | HN₂ (BL) | O'Connor et al, 1991 | CDDP, paclitaxel (BL) | Schmitt et al, 1998 |
|                     | Rapamycin (BL) | Muthukkumar et al, 1995 |                |            |
| **ATIC-ALK translocation** | MTX (ALCL) | Ma et al, 2000 |                |            |

ALCL: anaplastic large cell lymphoma; B-CLL: B-chronic lymphocytic leukaemia; BL: Burkitt lymphoma; B-DLCL: B-diffuse large cell lymphoma; FL: follicular lymphoma; MCL: mantle cell lymphoma; HCL: hairy cell leukaemia; MM: multiple myeloma; ARA-C: cytarabine; ATRA: all-trans-retinoic acid; BLM: bleomycin; 2-CdA: 2-chloro-deoxyadenosine; CDDP: cisplatin; CLB: chlorambucil; CPT(s): camptothecin(s); CTX: cyclophosphamide; DEX: dexamethasone; DHAD: mitoxantrone; DOX: doxorubicin; F-ara: fludarabine; HN₂: mechlorethamine; MTX: methotrexate; VCR: vincristine; VLBl: vinblastine; VM-26: teniposide; VP16: etoposide.
The degree of G1 arrest observed with these agents correlated with the rate of p53 and p21\textsuperscript{Ntr/Cip1} protein accumulation; etoposide induced rapid increase of both p53 and p21\textsuperscript{Ntr/Cip1}, while nitrogen mustards and cisplatin induced slow accumulation of p53 and no substantial changes in p21\textsuperscript{Ntr/Cip1} levels (Fan et al, 1994). Regardless of the differences in G1 arrest and kinetics of p53 or p21\textsuperscript{Ntr/Cip1} accumulation, all agents induced apoptosis to a greater extent in the wild-type than in the mutant p53 cells (Fan et al, 1994). An inverse relationship between chemosensitivity to nitrogen mustards/cisplatin and etoposide was observed in the mutant p53 lines and this finding correlated with topoisomerase II mRNA levels in the cells (Fan et al, 1994).

The INK4a/ARF locus encodes 2 tumour suppressor genes, designated p16INK4a and p19ARF, which are upstream regulators of the retinoblastoma (Rb) and p53 genes (Schmitt et al, 1999). INK4a/ARF null lymphomas display low p53 expression despite the presence of a wild-type p53 gene; they are highly aggressive and resistant to cytotoxic agents, including cyclophosphamide and doxorubicin, although high drug doses might induce p53-independent apoptosis in vitro (Schmitt et al, 1999). Lymphocytes from patients with B-chronic lymphocytic leukaemia with p53 gene mutations and increased expression of the ERCC1 gene (excision repair cross complementing-1) are able to repair drug-induced DNA-damaged genes and are cross-resistant to chlorambucil, fludarabine, and camptothecins in vitro (Silber et al, 1994). p53 gene deletion is involved in the lack of response to purine analogues, including fludarabine and 2-chloro-deoxyadenosine, and poor survival in B-chronic lymphocytic leukaemia (Döhner et al, 1995), while paclitaxel and vincristine are capable of inducing apoptosis independent of p53 status in human lymphoblastoid cell lines (Fan et al, 1998).

Protein kinase C inhibitors suppress cell proliferation and are useful in overcoming drug resistance by inhibiting mdr-mediated drug efflux. They increase the cytotoxicity of DNA-damaging agents, including platinum complexes, by interfering with cellular repair mechanisms. 7-Hydroxystaurosporine (UCN-01) is a protein kinase C inhibitor that blocks cells in G1 phase by promoting accumulation of dephosphorylated retinoblastoma Rb protein as a consequence of inhibition of cyclin-dependent kinases and increase in the expression of cyclin-dependent kinase inhibitor proteins. Moreover, UCN-01 induces the expression of apoptosis-related surface markers such as annexin and Fas (CD95). UCN-01 prevents the G1 checkpoint arrest in human lymphoma CA46 cells lacking normal p53 function and sensitises cells to the effects of DNA-damaging agents, including cisplatin, etoposide, cyclophosphamide, doxorubicin, as well as prednisone and vincristine (Wilson et al, 2000). Finally, UCN-01 increases fludarabine-induced mitochondrial damage, caspase activation, apoptosis, and suppresses clonogenic survival in bcl-2 overexpressing human leukaemia U937 cells by altering bcl-2 phosphorylation (Harvey et al, 2001).

Cells of thymic lymphoma overexpressing ras/raf genes, developed resistance to dexamethasone as a consequence of the overexpression of bcl-2, while retaining their sensitivity to p53-dependent apoptosis by etoposide (Kobzdej et al, 2000). This finding suggests that glucocorticoid-induced apoptosis involves bcl-2 pathway but not p53 activity. The IL-6-dependent myeloma cell line ANBL6 is sensitive to dexamethasone, doxorubicin, and melphalan. IL-6-driven cell proliferation involves ras-signalling pathways; however, N- or K-ras mutations at codon 12 (N-ras12 and K-ras12) lead to a constitutively active ras protein and IL-6 independent growth and protect ANBL6 cells from apoptosis induced by dexamethasone, doxorubicin and melphalan (Rowley et al, 2000).

Overexpression of c-myc is associated with induction of apoptosis by mechloretamine in the JLP116 Burkitt cell line (O’Connor et al, 1991) and by the immunosuppressive drug rapamycin in the immature B cell lymphoma BKS-2 (Muthukumar et al, 1995). Finally, fludarabine, is able to target non-dividing cells through a specific decrease in the expression of STAT, (signal transducer and activator of transcription), a key mediator of cellular responses to cytokines and oncoproteins (Frank, 1999).

**CONCLUSIONS**

The developments in molecular medicine suggest that pharmacogenetics will influence the decisions concerning the treatment of selected diseases. With the available information, the likelihood of response to fludarabine of a NHL might be dependent, at least in part, on the following pharmacogenetic profile: (1) low bcl-2/bax ratio (Pepper et al, 1999); (2) high cellular deoxycytidine kinase ratio (Pepper et al, 1999); (2) high cellular deoxycytidine kinase and low 5′-nucleotidase activities (Dumontet et al, 1999); and (3) wild-type p53 (Feng et al, 2000), while a favourable profile predicting response to CHOP regimen would include the following: (1) wild-type p53 (Navaratnam et al, 1998); (2) presence of bcl-2 translocation, in particularly if CHOP is associated with rituximab (Vose et al, 2001); and (3) presence of NPM/ALK chimeric protein (Muegerian-Bedoyan et al, 1997).

Today’s specialized techniques of bioinformatics are likely to become routine diagnostic tools in the near future to predict the likelihood of response based on the genome-wide profiling of disease and to select anticancer agents based on genetic abnormalities of the neoplasm to be treated. This prospective implies that the classic approach of standard drug combinations for the treatment of a specific tumour, irrespective of its molecular characteristics, may be abandoned in the future in favour of individually tailored cancer chemotherapy, based on the genetic pattern of disease. Novel technologies provide new avenues of investigation, including real-time PCR and cDNA microarray, and offer powerful tools to elucidate the in vivo molecular events involved in the development and progression of NHLs. Other intriguing issues will be the identification of the optimal drug sequence of treatment in combination regimens and the pharmacological monitoring of drug disposition to prevent possible drug–drug interactions and unexpected toxicities due to polymorphism in drug-metabolizing enzymes. It may be thus hypothesized that bioinformatics may help in the future clinicians in the integration of multiple variables, including disease-specific factors and the mechanism of action of drugs, in the formulation of optimized chemotherapeutic protocols.

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