MODY patients exhibit shorter telomere length than non-diabetic subjects

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Abstract

Background: Given the increasing evidence supporting the association between telomere shortening and diabetes, the aim of the present work was to establish whether MODY patients suffer a reduction in telomere length (TL) due to oxidative stress produced by chronic hyperglycemia, despite not presenting insulin resistance or inflammation.

Methods: We analysed clinical and biochemical parameters in 35 MODY2 and 12 MODY3 patients compared with 48 control subjects. The absolute telomere length (aTL) of peripheral blood leukocytes was measured using the quantitative polymerase chain reaction (qPCR).

Results: A significant negative correlation was observed between aTL and age in the whole population, among MODY patients and in each subtype studied, MODY2 and MODY3, which allowed us to validate the method. We found, for the first time, that MODY patients have shorter aTL with respect to non-diabetic controls (6.49 ± 3.31 kbp vs 11.13 ± 7.82 kbp, \( p = .006 \)). However, no differences were found between MODY2 and MODY3. In addition, aTL showed a negative correlation with duration of the disease and fasting plasma glucose (FPG) levels in MODY patients in general and also with HbA1c in MODY2 patients in particular.

Conclusions: Both MODY2 and MODY3 types present telomere shortening, which, at least partly, responds to HbA1c and FPG levels. These findings suggest comparable mechanisms underlying the attrition of TL. Taken together, our results on aTL in MODY patients may provide a parameter relatively easy and inexpensive to quantify in order to measure the impact of high glucose levels and potentially carry out anti-diabetic treatment with stricter targets.

KEYWORDS
hyperglycemia, MODY, oxidative stress, telomere length

1 INTRODUCTION

Maturity-onset diabetes of the young (MODY) is a monogenic disorder characterized by autosomal dominant inheritance, which usually develops in childhood, adolescence or young adulthood.1,2 It has been established that MODY represents 1% to 5% of all cases of diabetes and is often misdiagnosed as type 1 diabetes (T1D) or type 2 diabetes (T2D).1,3,4
MODY is caused by mutations in different genes, resulting in pancreatic \( \beta \)-cell dysfunction.\(^5\)\(^6\) There are 14 subtypes of MODY, depending on the genes involved, MODY2 and MODY3 being the most frequent genotypes. MODY2 is caused by mutations in the glucokinase (GCK) gene, while MODY3 is caused by mutations in the hepatic nuclear factor (HNF1A) gene.\(^2\)

The clinical characteristics of the different subtypes oscillate between a mild hyperglycemic state with good clinical prognosis (MODY2) to a more severe hyperglycemic state accompanied by early chronic complications (MODY3). Both subtypes present alterations in insulin secretion as a primary defect, without insulin resistance.\(^7\)-\(^9\)

Telomeres are nucleo-protein complexes located at the end of eukaryotic chromosomes, composed of several tandem repeats of a non-coding DNA hexanucleotide (TTAGGG in mammals) and associated proteins. Telomeres play an essential role in the integrity and stability of the chromosome and cell survival.\(^10\) During DNA replication in the process of cell division, telomere length (TL) can undergo a reduction of about 20 to 40 kilobase pairs (kbp) per year in peripheral blood leukocytes. Due to this mechanism, a negative correlation was observed between age and TL.\(^11\)-\(^13\) TL shortening is an important marker of the replicative capacity of the cell, which makes it a suitable marker of cellular aging.\(^14\) In a pathological context, inflammation and oxidative stress present a negative association with TL.\(^15\),\(^16\)

Hyperglycemia is one of the most important factors that determine the production of oxidative stress and its consequent reactive oxygen species (ROS), all of which produces chronic complications in diabetes, including the shortening of TL.\(^17\),\(^18\)

Although several studies have reported a negative correlation between TL and T1D or T2D,\(^19\) no studies have been conducted so far on the possible association between TL and MODY patients. Therefore, the hypothesis of the present work was that MODY patients may suffer a reduction in TL due to oxidative stress produced by chronic hyperglycemia, despite not presenting insulin resistance or inflammation. In reference to this, the promoter of the C-reactive protein (CRP) gene requires hepatocyte nuclear factor 1 alpha (HNF1A) for its expression. Mutations present in the HNF1A gene (MODY3) are associated with reductions in CRP levels, an important marker of inflammation, compared to other types of diabetes and non-diabetic individuals.\(^20\),\(^21\)

To test this hypothesis, we evaluated absolute TL (aTL) in genetically diagnosed MODY2 and MODY3 individuals and non-diabetic controls. We also assessed the correlation between aTL and other clinical or metabolic factors and possible differences in TL between MODY2 and MODY3 patients.

## 2 MATERIALS AND METHODS

### 2.1 Study population

The size of the sample was calculated from a previous report, which informed a 30% shorter mean telomere length in patients with type 1 diabetes when compared with controls.\(^18\) Assuming a mean telomere length of 13 kbp (highest mean value), with an effect size of 1.25 (delta), corresponding with a 50% shorter telomere length in MODY subjects, an alpha error of 0.05 and a power of 80%, a total sample size of 30 individuals was considered sufficient.

We analysed 35 patients with genetic diagnosis of MODY2, 12 with MODY3 and 48 unrelated non-diabetic control individuals, matched by sex and age from the general population of the Autonomous City of Buenos Aires (BA) and its metropolitan area, Argentina. None of the controls had clinical components of MODY or a family history of MODY (self-reported). The individuals recruited had normal results in the medical examination and blood counts, and were not taking any medications. In this respect, the diabetic status was ruled out in accordance with the American Diabetes Association recommendation.\(^6\)

Clinical characterization of MODY was done following the ADA (American Diabetes Association) recommendation.\(^5\) Genetic studies consisted in the purification of genomic DNA from peripheral blood by MagNA Pure system (Roche), PCR amplification of the coding regions of both genes was carried out as published by Kaisaki P. et al and Stoffel M. et al,\(^22\),\(^23\) followed by Sanger sequencing. Mutations present in all participants were analysed by bioinformatic tools to determine the possible effect of the alterations found. Among them, the service provided by mutation taster was preferred for being more accurate.

Glutamic acid decarboxylase autoantibodies (GADA), insulin autoantibodies (IAA) and anti-phosphatase autoantibodies (IA2A) were measured by radioimmunoassays in all MODY patients, being in all cases negative.

Demographic data including age and gender of the patients were collected. The anthropometric measures (height, weight and waist circumference [CC]) and systolic and diastolic blood pressure (SysBP and DiasBP, respectively) were determined using a standardized protocol. The body mass index (BMI) was calculated as weight (kg)/height (m)\(^2\). Fasting plasma glucose (FPG) and HA1c levels were performed by the usual standardized biochemical procedures.

### 2.2 Measurement of leukocyte aTL

The estimation of aTL was carried out using an optimized methodology based on real-time polymerase chain reaction (qPCR), using a previously described protocol.\(^24\) Genomic DNA samples were purified from peripheral blood leukocytes and aTL was determined using SYBR green as an intercalating agent. This method allows us to determine the repetitive sequences of telomeres compared to a single copy gene using standard curves. For the PCR of the single-copy gene, a 75-bp oligonucleotide of the RPLP0 (ribosomal protein, large, subunit P0) gene was used as a standard and allowed the genome/reaction number (S) to be exported from the standard curve. The standard for telomere PCR is a synthetic oligonucleotide of 84 bp in length formed by the tandem repeats of telomeric DNA and the standard curve is used to measure the kbp of telomeric sequence/reaction (T).

The calculation of the T/S ratio allows to determine the kbp of telomeric sequence per cell for each individual. The PCR reactions...
were performed in duplicate for all the study samples in a StepOne Real-Time PCR System (Applied Biosystems). DNA (20 ng) was amplified in a 20 μL reaction volume containing 10 μL of SYBR Select Master Mix, and 250 nM of primers were added for the RPLP0 gene or 100 nM of primers for the telomeric sequence.

The PCR conditions consisted of a denaturation of 10 minutes at 95°C followed by 40 cycles at 95°C for 15 seconds, 60°C for 1 minute and the melting curve with 1 cycle of 15 seconds at 95°C, 1 minute at 60°C and 15 seconds at 98°C with a temperature ramp of 0.3°C/second.

2.3 | Statistical analysis

The statistical analysis was carried out through Statistical Package for Social Sciences software (SPSS version 20.0) with a level of significance of 0.05. Normal distribution of continuous variables was examined by Kolmogorov-Smirnov normality test.

The relationship between aTL and clinical-biochemical and anthropometric characteristics in whole samples was assessed with partial correlation analysis using age and gender as covariates.

Comparison of clinical-biochemical and anthropometric characteristics between groups is evaluated by univariate analysis of covariance (ANCOVA), followed by Bonferroni post hoc test for multiple comparisons.

Among MODY patients, the relationship between clinical-biochemical and anthropometric characteristics is evaluated by linear regression model or partial correlation analysis controlling for the effect of age, gender or type of MODY.

| TABLE 1 | Clinical-biochemical and anthropometric characteristics of control and patients with MODY2 and MODY3 |
|----------|-------------------------------------------------------------|
|          | Control | MODY2 | MODY3 | P   | p Control vs MODY2 | p Control vs MODY3 | p MODY2 vs MODY3 |
| n        | 48      | 35    | 12    |     |                   |                   |                  |
| Age (years) | 24 ± 9.66 | 20.11 ± 14.43 | 30.25 ± 18.42 | 0.035 | 0.028 | 0.049 | 0.161 |
| aTL (Kpb) | 11.13 ± 7.82 | 6.64 ± 3.50 | 6.05 ± 2.78 | <0.001 | <0.001 | 0.002 | 0.645 |
| Weight (kg) | 65.43 ± 12.00 | 45.38 ± 17.52 | 67.19 ± 14.49 | <0.001 | <0.001 | 0.029 | <0.001 |
| WC (cm) | 77.38 ± 10.42 | 68 ± 23.42 | 86.33 ± 22.05 | 0.180 | 0.343 | 0.156 | 0.634 |
| BMI (kg/m2) | 23.45 ± 3.93 | 19.82 ± 3.34 | 24.02 ± 3.56 | <0.001 | <0.001 | 0.687 | <0.001 |
| FPG (mg/dL) | 85.36 ± 8.98 | 117.40 ± 11.59 | 129.37 ± 14.12 | <0.001 | <0.001 | 0.095 |
| TG (mg/dL) | 77.15 ± 34.17 | 87.31 ± 62.26 | 125.62 ± 93.22 | 0.046 | 0.446 | 0.050 | 0.181 |
| TC (mg/dL) | 157.58 ± 29.03 | 170.35 ± 52.24 | 178.5 ± 27.52 | 0.222 | 0.144 | 0.179 | 0.318 |
| LDL-C (mg/dL) | 87.04 ± 22.81 | 98.37 ± 39.34 | 101.37 ± 22.63 | 0.150 | 0.125 | 0.311 | 0.423 |
| HDL-C (mg/dL) | 86.33 ± 22.05 | 60.25 ± 14.49 | 225.62 ± 93.22 | 0.383 | 0.432 | 0.309 | 0.451 |
| TG/HDL | 1.44 ± 0.78 | 1.75 ± 1.27 | 2.25 ± 1.85 | 0.031 | 0.186 | 0.004 | 0.270 |
| HbA1c (%) | 6.32 ± 0.42 | 6.92 ± 1.56 | - | - | - | - | 0.139 |
| SysBP (mmHg) | 113.18 ± 8.53 | 107.42 ± 8.39 | 118.50 ± 11.36 | 0.049 | 0.105 | 0.567 | 0.008 |
| DiasBP (mmHg) | 70.68 ± 6.78 | 60.33 ± 6.21 | 77 ± 12.49 | 0.001 | 0.001 | 0.350 | 0.021 |

Note: Values are expressed as mean ± standard deviation (SD). P and p values were obtained from univariate analysis adjusted by age and gender (ANCOVA). Data with significant differences within groups (p < .05) are in bold.

Abbreviations: aTL, absolute telomere length; BMI, body mass index; DiasBP, diastolic blood pressure; FPG, fasting plasma glucose; HbA1c, glycated haemoglobin A1c; HDL-C, high density cholesterol; LDL-C, low density cholesterol; SysBP, systolic blood pressure; TC, total cholesterol; TG, triglycerides; WC, waist circumference.

3 | RESULTS

The 95 participants of this study presented a mean age of 23.61 ± 13.11 years. Table 1 shows the mean clinical-biochemical and anthropometric characteristics of control, MODY2 and MODY3 patients. The groups differed significantly in weight and, in particular, MODY2 patients showed significant differences in BMI, FPG levels and DiasBP compared to controls. MODY3 patients, instead, differed significantly in FPG levels and TG/HDL with respect to controls.

The complete sample showed a negative correlation between aTL and age (r = −0.311, p = .002, Figure 1A). Among the different clinical biochemical variables analysed in the complete sample, aTL only showed a significant negative correlation with FPG (r = −0.241, p = .026). This correlation remained significant even when age and gender were used as covariates (r = −0.347, p = .001).

Most importantly, MODY patients had significantly shorter mean aTL than controls (6.49 ± 3.31 kpb vs 11.13 ± 7.82 kpb, p = .006, Figure 1B), a difference which remained significant even after adjustment for non-modifiable factors, gender and age (p < .001).

Among MODY patients, a significant negative correlation was observed between aTL and age (r = −0.447, p = .009, covariate = gender), FPG levels (r = −0.586, p = .008, covariates = age, gender, and type of MODY) and duration of the disease (r = −0.385, p = .027,
covariates = gender and type of MODY). Furthermore, we found a significant positive correlation between disease duration and weight ($r = 0.504, p = .005$) and BMI ($r = 0.504, p = .001$) using gender and MODY type as covariates. No significant correlations were found with metabolic parameters.

Regarding the characteristics of each MODY type, MODY3 patients exhibited significantly higher weight ($p < .001$), BMI ($p < .001$), SysBP ($p = .008$) and DiasBP ($p = .021$) than MODY2 patients (Table 1). In turn, no significant differences were found between MODY2 and MODY3 individuals in terms of aTL (Figure 2A), whereas a negative correlation was observed between aTL and age in both MODY2 ($r = −0.369, p = .029$) and MODY3 ($r = −0.754, p = .023$) individuals (Figure 2B). Worth highlighting, MODY2 patients revealed a significant negative correlation between aTL and HbA1c ($r = −0.388, p = .030$) with gender and age as covariates (Figure 3), while MODY3 patients showed a trend toward lower aTL at higher HbA1c levels ($r = −0.760, p = .07$) with gender as a covariate.

4 | DISCUSSION

TL is an important parameter that has increasingly gained importance due to its proven relationship to a variety of diseases, among them metabolic diseases such as diabetes. Given their chemical composition, telomeres are highly vulnerable to oxidative damage and inflammation, which might accelerate telomere shortening. In addition, hyperglycemia induces high oxidative stress, and consequent telomere shortening. Therefore, telomere shortening mechanisms may vary depending on the pathogenicity of diabetes.

Previous studies have shown more significant mononuclear cell telomere shortening in T1D than in T2D patients. In this context, the hypothesis of the present work was that MODY patients may suffer a reduction in TL due to oxidative stress produced by chronic hyperglycemia, despite not presenting insulin resistance or inflammation. To test this hypothesis, we evaluated aTL in MODY2 and MODY3 patients compared with control subjects.
We observed a negative correlation between aTL and age in the whole population. This result was expected, considering that age is the main factor involved in the shortening of telomeres and allowed us to validate the method. Furthermore, we found that aTL was negatively correlated with FPG levels. In agreement, TL was recently shown to be negatively associated with glucose concentrations and HbA1c within the normal non-diabetic range, which shows an important contribution of hyperglycemia to telomere shortening.

We found, for the first time, that MODY patients have shorter aTL with respect to non-diabetic controls, which may be a consequence of MODY-associated hyperglycemia. We also observed a significant negative association between aTL and age among MODY patients, and in both subtypes studied, MODY2 and MODY3.

In addition, aTL showed a negative correlation with duration of the disease and FPG levels in MODY patients in general and also with HbA1c in MODY2 patients in particular, after adjustment for age, gender and type of MODY. On the one hand, the absence of correlation between aTL and HbA1c in MODY3 patients may be explained by the small cohort size. On the other hand, these results find support in studies by Rosa et al, who recently showed that relative telomere length was inversely associated with FPG and HbA1c levels in patients newly diagnosed with T2D, after adjustment for age, gender and BMI, and also with duration of the disease. In addition, a study by Januszewski et al revealed shorter TL in 199 T1D patients as compared to controls and a negative correlation between TL and duration of the disease and age. On the whole, our findings show a significant contribution of hyperglycemia and disease duration to telomere shortening even in the absence of insulin resistance or inflammation.

As opposed to what may have been expected in newly diagnosed MODY patients that have significant differences in the glucose level, we found no differences in aTL between our MODY2 and MODY3 patients, whose glucose levels were controlled through treatment. It can be thus inferred that both MODY types present telomere shortening which, at least partly, responds to HbA1c and FPG levels, which suggests comparable mechanisms underlying the attrition of TL.

The main limitation is that the sample size was relatively small; therefore, the results of this study should be verified using a larger sample size. We cannot exclude that some clinical characteristics, such as inflammation or lifestyle markers, that were not available for analysis could affect aTL. Furthermore, being a cross-sectional study, the present work cannot address the rate of telomere attrition in MODY patients.

Taken together, our findings on aTL in MODY patients may provide a parameter relatively easy and inexpensive to quantify the impact of the high glucose level, in order to carry out antidiabetic treatments with strict targets.

**CONFLICT OF INTEREST**

We wish to confirm that there are no known conflicts of interest associated with this publication and there has been no financial support for this work that could have influenced its outcome.

**AUTHOR CONTRIBUTIONS**

Andrea L. Millan performed the most part of the laboratory work, designed the research study, analysed the data and wrote the paper. Sofía I. Trobo provided the control individuals samples and the clinical data for the study. Martina C. García performed part of the laboratory work related to the control individuals. Alejandro de Dios provided the patients samples and the clinical data for the study. Gloria E. Cerrone assisted in the design of the research study and analysed the data. Gustavo D. Frechtel designed the research study and supervised the writing of the paper. Ariel P. López provided the funds, designed the research study and supervised the writing of the paper. All authors declare to have been revising the paper critically and have been approved the final version.

**ETHICS STATEMENT**

This work has been approved by the Institutional Review Committee and written informed consent was obtained from all individuals involved or from responsible family members after full explanation of the purpose and nature of all procedures used. All human investigations were conducted according to the principles expressed in the Declaration of Helsinki as revised in 1983. Informed consent was obtained from all individual participants included in the study.

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