Abstract
Varicocele is considered as one of the main etiologies of male infertility. Along with altered semen parameters, increased DNA fragmentation is believed to play an important role in varicocele-induced infertility. DNA damage may result from intra- or extra-testicular factors. Among these, apoptosis, abnormal chromatin packaging and oxidative stress are the most researched and are addressed in this review. Significant evidence suggests that varicoceles have a harmful effect on testicular function and a varicocelectomy not only prevents progressive decline in testicular function, but also reverses the damage. However, the degree to which varicocele repair improves pregnancy rates and the success of assisted reproductive technology (ART) remains controversial. Therefore, the role of varicocele repair on DNA fragmentation is also discussed.

Keywords: Varicocele, DNA Damage, Repair

Introduction
Abnormally dilated veins in the pampiniform plexus define varicocele, which is historically associated with male factor infertility (1). Even though varicoceles have been detected in boys as young as ten years of age, it is commonly believed that this condition may begin with the onset of puberty, at around the age of 15 (2, 3). The incidence of varicocele in adolescents is approximately 15% and, in the general population, it ranges from 4.4 to 22.6% (4). What distinguishes varicocele is its higher incidence in the infertile population; 21-41% in men with primary infertility and 75-81% in those with secondary infertility (5, 6). In addition, it causes a decrease in both semen quality and testicular volume.

The effects of varicocele are long-term and progressive, therefore the fertility potential of individuals afflicted with varicocele declines by the time they desire to achieve fatherhood (7). However, what has made the role of varicocele in fertility enigmatic is the fact that fertile men with varicocele are able to be father while infertile men with varicocele have lower chances for fertility. Surgical correction of varicocele improves their fertility potential or assisted reproductive technology (ART) outcome post-varicocelectomy (8). A likely explanation is the heterogeneity observed in the varicocele population, which includes different grades of varicocele, clinical diversity, location (bilateral vs. unilateral), duration of varicocele, individual genetic background, and social, economic, and geographical status of these individuals (6, 9, 10). Moreover, the pathophysiology of varicocele remains unclear.

Despite the availability of different procedures to diagnose varicocele, physical examinations and scrotal ultrasounds remain the most commonly used methods. Varicocele is graded at the time of the initial physical examination according to the Dubin grading system (I-III), where grade II is palpable without the Valsalva maneuver and grade I is not visualized, rather it is only palpable by the Valsalva maneuver (11-13). The term clinical varicocele refers to varicoceles detectable by palpation or visual inspection.

Treatment
Considering the enigma regarding varicocele, its treatment is also controversial. There are two
classes of thought regarding the treatment of varicocele. One group believes that its treatment improves semen parameters and fertility whereas the other believes there is no significant improvement post-varicocelectomy (6, 14, 15).

What is important in these studies are the control groups and sample sizes. Some studies have shown that the percentage of pregnancies in varicocele individuals after varicocelectomy was not statistically higher in comparison to those who underwent no treatment (14, 16). However, the current guidelines or the general consensus among the clinicians is that varicocele individuals with decreased testicular size and abnormal seminal parameters must be treated (17, 18). A recent meta-analysis suggests that the spontaneous pregnancy rate increases following repair of the varicocele. However, if confounding variables or heterogeneity between the clinical trials are taken into consideration, this effect would become insignificant or less clear (6).

In addition, the rate of spontaneous abortion in the spouses of post-varicocelectomy responders has been reported at approximately 15% and remains an issue of concern (19, 20). Increased levels of sperm DNA fragmentation have been proposed as a possible cause for low-quality embryos and spontaneous abortion. In this meta-analysis, the authors have shown that varicocele repair significantly improved sperm concentration and motility, yet they were unable to reach a conclusion on sperm morphology due to variations between different studies (6). These controversies suggest that more robust parameters with threshold should be defined to detect individuals who may benefit from varicocelectomy. To achieve this, different authors have assessed sperm functional parameters, such as DNA fragmentation, which will be discussed in the course of this review.

**DNA damage and varicocele**

Spermatogenesis is a complex process by which the male germ cell proliferates and matures through meiosis from diploid spermatagonia to haploid spermatozoa. Although a small percentage of spermatozoa from fertile men possesses detectable levels of DNA damage, which can occur at any step of spermatogenesis (21, 22), a higher degree of DNA fragmentation is clearly associated with male infertility. Many factors, including intra- or extratesticular factors, may be involved in this process (21, 23). Abnormal chromatin packaging, abortive apoptosis, and extreme production of reactive oxygen species (ROS) are factors which may lead to DNA damage (24-26). In addition, extra-testicular factors such as age, drugs, cigarette smoking, genital tract inflammation, hormonal factors, varicocele, and testicular hyperthermia are among the main reasons for DNA damage (24).

A variety of methods have been used to evaluate the integrity of sperm chromatin and DNA. For evaluation of major proteins associated with DNA, researchers commonly use chromomycin A3 (CMA3) and aniline blue staining. Deoxynucleotidyl transferase-mediated dUTP nick-end labeling (TUNEL), the comet assay, sperm chromatin dispersion (SCD), acridine orange (AO), and sperm chromatin structure assay (SCSA) are used for DNA breaks. Recently our group presented a review article that discussed the methodologies, advantages, and disadvantages of these tests that was published in the International Journal of Fertility and Sterility (IJFS) (25). The current review focuses on the relation between internal testicular factors involved in DNA damage and their relation with varicocele.

**Apoptosis and varicocele**

Germ cell death occurs during normal spermatogenesis in mammals and it is estimated to be responsible for the loss of up to 75% of potential spermatozoa. Spermatogonia are eliminated by apoptosis and only 25% of the theoretically expected number of primary spermatocytes are produced from the original population of spermatogonia (27, 28). In addition, 20% of spermatocytes and spermatids are frequently removed by selective apoptosis (29). Therefore, the presentation of apoptotic markers such as phosphatidylserine externalization, DNA fragmentation, and Fas expression are considered a part of this process (30, 31). Increased apoptosis, necrosis, and degeneration of sperm in individuals with varicocele may suggest that a redundancy of spermatogenic cells by apoptosis is facilitated in these patients. At least three pathways have been proposed: i. excess heat, ii. androgen deprivation at the testicular level, and iii. accumulation of toxic agents, including the products of cigarette smoke, such as cadmium (10).

A higher population of sperm with fragile DNA has been observed upon heat treatment. According to research, heat treatment is associated with poor ca-
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Citation characteristics and apoptosis (32). Therefore, it is plausible to expect that exposure to poorly regulated genital temperatures may cause spermatozoa to undergo significant apoptosis or necrosis during spermatogenesis in varicocele individuals, and at its extreme it may lead to hypospermatogenesis and spermatogeneretic arrest. At the spermatid level, a predominant phenotype has been reported by Lin et al. in varicocele individuals. This is the reason for improvement in semen parameters in azoospermic men with maturation arrest post-varicocelectomy (33, 34).

It has been shown that Sertoli cells can begin and regulate germ cell apoptosis via the apoptosis-stimulating Fas system, as characterized by the interaction between Fas and its ligand (30, 35). Del Giudice et al. evaluated FasL mRNA levels using reverse transcription and real-time quantitative polymerase chain reaction (RQ-PCR) in adolescents with and without varicocele. They concluded that FasL mRNA levels were higher in varicocele individuals compared to those without varicocele. In addition, they observed a significant negative correlation between sperm concentration and FasL mRNA levels. These findings suggested that individuals with low sperm concentrations have high FasL mRNA levels (36).

Wu et al. also evaluated apoptosis markers in varicocele individuals. Their findings showed increased externalization of phosphatidylserine, mitochondrial dysfunction, and nuclear DNA damage in fertile and varicocele individuals. This has suggested that an association may exist between certain apoptotic mechanisms and varicocele, which possibly originates in the mitochondria of spermatocytes, resulting in DNA anomalies or fragmentation in the nucleus of these cells (37).

Protamine deficiency and varicocele

Spermatogenesis is considered a continual process. However, in order to make this process more comprehensible, it has been divided into three components: spermatocytogenesis, reduction division, and spermiogenesis (38). During normal spermiogenesis, 85% of histones are replaced with protamines, which results in sperm chromatin condensation (39). According to Oliva, chromatin condensations are essential for: i. protecting paternal genes from nucleases, mutagens, toxins, and heavy metals in the testis or spermatozoa, ii. the generation of a hydrodynamic nucleus to facilitate genome transportation and fertilization, and iii. the removal of proteins and transcription factors from the spermatid, which can lead to the formation of a blank paternal genetic message free of epigenetic information and ready for reprogramming by the oocyte (40). Different authors have shown that varicocele adversely affects these processes. For reason El-Segini et al. evaluated chromatin condensation by aniline blue in infertile patients with varicocele and showed that they have significantly more impaired chromatin condensation than either the fertile group or those fertile individuals with varicocele (41). Thus, they concluded that one of the major spermiogenic insults affecting fertility in varicocele patients was a defect in the histone/protamine exchange (40, 41).

This defect has also been verified by other authors using different approaches, such as assessing protamine deficiency by CMA3 or showing the presence of excessive histone by anilineblue or toluidine blue (42, 43). Our studies in varicocele individuals also confirmed defects in histone/protamine exchange and have further shown that this insult is rectified by varicocele repair (8, 44). However, by assessing protamine deficiency in pregnant and non-pregnant partners of individuals who underwent varicocelectomies, our results have indicated no significant improvement in one group over the other. We can thus conclude that the defect in his tone/protamine exchange may be one of the mechanisms associated with varicocele, it may not be the major factor in the pathogenesis of varicocelectomy (8).

Hyperthermia is believed to play a key role in the pathophysiology of varicocele and the reduction of fertility rates (45). One of the responses of eukaryotic organisms to hyperthermia is production of heat-shock proteins (HSPs) (46). HSPA2, a member of the 70-kDa family, is testis tissue-specific and has been shown to function as a chaperone protein that aids in protein folding (47). The deficiency of this gene in knockout mice or a mutation in this gene results in the arrest of primary spermatocytes at stage I of meiosis, leading to azoospermic or infertility in mice. This result shows an important role for HSPA2 in spermatogenesis (48). HSPA2 has been evaluated as a sperm maturity and function index (49). Our study showed that chronic hyperthermia induced by varicocele not only resulted in impaired semen parameters but also reduced HSPA2 expression when compared to the fertile population. Removal of this stress by varicocelectomy has been shown to improve semen pa-
parameters and increase HSPA2 expression (50). These improvements are likely achieved by the proper folding proteins involved in spermatogenesis including protamine, which is involved in DNA packaging and protection of DNA from damage.

**Reactive oxygen species (ROS) and varicocele**

ROS are free radicals produced in biological systems that have an important role in capacitation, hyper-activation, and sperm oocyte fusion (51). In the presence of polyunsaturated fatty acids in the plasma membrane of sperm, ROS triggers a chain of chemical reactions called lipid peroxidation (51). It has been shown that ROS can damage DNA by causing deletions, mutations, and other lethal genetic effects (52). Moein et al. compared ROS levels in the seminal fluid of infertile men with varicocele and idiopathic infertility to fertile donors and showed ROS levels to be significantly higher in the former group. In addition, it has been shown that elevated scrotal temperature due to impaired circulation results in the accumulation of toxic metabolites, which is believed to be the source for ROS production in these individuals (53).

Apart from xenobiotics and the accumulation of toxic metabolites, the other key sources of ROS in semen are leukocytes and immature spermatozoa (sperm with residual cytoplasm) (54). Thus DNA damage in sperm may have been induced by ROS sources. Kemal Duru et al. have shown that exposing spermatozoa to artificially produced ROS significantly increases DNA damage (55). However, the effect of ROS on DNA fragmentation should only be considered in light of cellular antioxidant mechanisms and the antioxidants present in the secretom. In association with increased ROS and DNA fragmentation, a decrease in the level of antioxidants (including superoxide dismutase, glutathione peroxidase, catalase, and ascorbic acid) and total antioxidant capacity has been reported both in infertile and varicocele individuals (56). Some authors have reported a clear association between ROS levels and antioxidant capacity to sperm parameters in infertile and varicocele individuals (57, 58). In addition, they believe that these aberrant levels of ROS and antioxidants are involved in the occurrence of oligospermia, sperm motility defects, and/or abnormal sperm morphology (59). Simsek et al. have suggested that this effect is mediated via cellular apoptosis (60). A meta-analysis by Agarwal et al. (5) has shown significantly higher ROS and lower total antioxidant capacity levels in the varicocele population compared to fertile men, which may have resulted from abortive apoptosis and improper protamination. Chen et al. in a prospective study, have documented a reduction in ROS level following varicocele repair. Thus, varicocele repair may restore spermatogenesis, and improve semen parameters and decreases DNA damage via decreases in ROS levels (52).

**DNA damage and varicocele repair**

Evaluation of DNA damage has been proposed as extra information regarding sperm quality and is a forecaster of fertility potential. In the previous section, various factors involved in DNA damage have been discussed. The management of infertile men with increased sperm DNA damage remains high, particularly in varicocele individuals. Some studies have reported that sperm DNA fragmentation to be significantly higher in varicocele individuals compared to those with normal semen parameters (50, 61). Furthermore, DNA damage is associated with fertilization rate, spontaneous pregnancy, and pregnancy outcome post ART (62, 63). It is important to confirm whether varicocele repair may resolve varicocele-induced DNA damage or factors involved in sperm DNA damage.

Many studies suggest that varicocele repair should be performed in infertile men who have clinical varicocele and abnormal semen analyses (8, 64). Despite this consensus, the question of whether varicocele should be repaired in individuals with high DNA damage remains controversial. In a prospective trial, Smit et al. have stated that the percentage of DNA index by SCSA was reduced three months post-varicocele repair (14). In addition, Azadi et al. (44) and Zini et al. (65) showed the beneficial effect of varicocele repair on human sperm DNA damage. Previous studies have found no significant reduction in DNA damage three months post-varicocele repair. They propose that the duration of spermatogenic cycle is about 64 days, thus the reduction of DNA fragmentation should be assessed six months after varicocele repair.

Considering this overview of the research papers, we suggest that varicocele repair should be performed in varicocele individuals who have high DNA fragmentation in their semen samples prior to surgery.
Conclusion
According to the literatures, this review suggested that varicocele has a detrimental effect on testis function and mainly spermatogenesis. Therefore, varicocelectomy could improve testicular function and sperm production.

References
1. World Health Organization. The influence of varicocele on parameters of fertility in a large group of men presenting to infertility clinics. Fertil Steril. 1992; 57(6): 1289-1293.
2. Steeno O, Knops J, Declerck L, Adimoeja A, van de Voorde H. Prevention of fertility disorders by detection and treatment of varicocele at school and college age. Andrologia.1976; 8(1): 47-53.
3. Abbasi H, Ghanbarian A, Khoozani S, Nasr-Esfahani MH. Evaluation of varicocele frequency in adolescents in the city of Isfahan. Int J Fertil Steril. 2007; 1(3): 107-112.
4. Dobanovacki D. Varicocele in adolescents. Med Pregl. 2010; 63(11-12): 741-746.
5. Agarwal A, Deepinder F, Coccuzza M, Agarwal R, Short RA, Sabanegh E, et al. Efficacy of varicocelectomy in improving semen parameters: new meta-analytical approach. Urology. 2007; 70(3): 532-538.
6. Baazeem A, Belzile E, Ciampi A, Dohle G, Jarvi K, Salonia A, et al. Varicocele and male factor infertility treatment: a new meta-analysis and review of the role of varicocele repair. Eur Urol. 2011; 60(4): 796-808.
7. Naughton CK, Nangia AK, Agarwal A. Varicocele and male infertility: Part II: Pathophysiology of varicoceles in male infertility. Hum Reprod Update. 2001; 7(5): 473-481.
8. Nasr Esfahani MH, Abasi H, Razavi S, Ashrafi S, Tavalaee M. Varicocelectomy: semen parameters and prostatic fluid. Int J Androl. 2009; 32(2): 115-122.
9. Harrison RM, Lewis RW, Roberts JA. Pathophysiology of varicocele in nonhuman primates: long-term seminal and testicular changes. Fertil Steril. 1986; 46(3): 500-510.
10. Benoff SH, Millan C, Hurley IR, Napolitano B, Marmar JL. Bilateral increased apoptosis and bilateral accumulation of cadmium in infertile men with left varicocele. Hum Reprod. 2004; 19(3): 616-627.
11. Hirsh AV, Cameron KM, Tyler JP, Simpson J, Pryor JP. The Doppler assessment of varicoceles and internal spermatic vein reflux in infertile men. Br J Urol. 1980; 52(1): 50-56.
12. Lee J, BinSaleh S, Lo K, Jarvi K. Varicoceles: the diagnostic dilemma. J Androl. 2008; 29(2): 143-146.
13. Will MA, Swain J, Fode M, Sonksen J, Christman GM, Ohl D. The great debate: varicocele treatment and impact on fertility. Fertil Steril. 2011; 95(3): 841-852.
14. Smit M, Romijn JC, Wildhagen MF, Veldhoven JL, Weiber RF, Dohle GR. Decreased sperm DNA fragmentation after surgical varicocelectomy is associated with increased pregnancy rate. J Urol. 2010; 183(1): 270-274.
15. Yamamoto M, Hibi H, Hirata Y, Miyake K, Ishigaki T. Effect of varicocelectomy on sperm parameters and pregnancy rate in patients with subclinical varicocele: a randomized prospective controlled study. J Urol. 1996; 155(5): 1636-1638.
16. Krause W, Muller HH, Schafer H, Weidner W. Does treatment of varicocele improve male fertility? Results of the 'Deutsche Varikozelenstudie', a multicentre study of 14 collaborating centres. Andrologia. 2002; 34(3): 164-171.
17. Dubin L, Amelar RD. Varicocelectomy as therapy in male infertility: a study of 504 cases. Fertil Steril. 1975; 26(3):217-220.
18. Zini A, Buckspan M, Berardinucci D, Jarvi K. The influence of clinical and subclinical varicocele on testicular volume. Fertil Steril. 1997; 68(4) 671-674.
19. Shamsa A, Nademi M, Aqaei M, Fard A N, Molaie M. Complications and the effect of varicocelectomy on semen analysis, fertility, early ejaculation and spontaneous abortion. Saudi J Kidney Dis Transpl. 2010; 21(6): 1100-1105.
20. Negri L, Levi-Setti PE. Pregnancy rate after varicocele repair: how many miscarriages? J Androl. 2011; 32(1): 1.
21. Sakkas D, Mariethoz E, Manicardi G, Bizzaro D, Bianchi PG, Bianchi U. Origin of DNA damage in ejaculated human spermatozoa. Rev Reprod. 1999; 4(1): 31-37.
22. O’Brien J, Zini A. Sperm DNA integrity and male infertility. Urology. 2005; 65(1): 18-22.
23. Babazadeh Z, Razavi S, Tavalaee M, Deemeh MR, Shahidi M, Nasr-Esfahani MH. Sperm DNA damage and its relation with leukocyte DNA damage. Reprod Toxicol. 2010; 29(1): 120-124.
24. Zini A, Libman J. Sperm DNA damage: clinical significance in the era of assisted reproduction. CMAJ. 2006; 175(5): 495-500.
25. Tavalaee M, Nasr Esfahani MH, Deemeh MR. Etiology and evaluation of sperm chromatin anomalies. Int J Fertil Steril. 2008; 2(1): 1-8.
26. Moustafa MH, Sharma RK, Thornton J, Mascha E, Abdel-Hafez MA, Thomas AJ Jr, et al. Relationship between ROS production, apoptosis and DNA denaturation in spermatozoa from patients examined for infertility. Hum Reprod. 2004; 19(1): 129-138.
27. Sakkas D, Seli E, Bizzaro D, Tarozzi N, Manicardi GC. Abnormal spermatozoa in the ejaculate: abortive apoptosis and faulty nuclear remodelling during spermatogenesis. Reprod Biomed Online. 2003; 7(4): 428-432.
28. Rodriguez I, Ody C, Araki K, Garcia I, Vassalli P. An early and massive wave of germinal cell apoptosis is required for the development of functional spermatogenesis. EMBO J. 1997; 16(9): 2262-2270.
29. Angelopoulou R, Plastira K, Msouel P. Spermatozoal sensitive biomarkers to defective proteasomis and fragmented DNA. Reprod Biol Endocrinol. 2007; 5: 36.
30. Soleimani M, Tavalaee M, Aboutorabi R, Adib M, Bahramian H, Jannamian E, et al. Evaluation of Fas positive sperm and complement mediated lysis in subfertile individuals. J Assist Reprod Genet. 2010; 27(8): 477-482.
31. Zhang HB, Lu SM, Ma CY, Wang L, Li X, Chen ZJ. Early apoptotic changes in human spermatozoa and their relationships with conventional semen parameters and sperm DNA fragmentation. Asian J Androl. 2008; 10(2): 227-235.
32. Mann SL, Patton WC, King A, Chan PJ. Comparative genomic hybridization analysis of sperm DNA apoptosis after exposure to heat shock. J Assist Reprod
33. Lin WW, Lamb DJ, Wheeler TM, Abrams J, Lipschultz LI, Kim ED. Apoptotic frequency is increased in spermatogenic maturation arrest and hypospermatogenic states. J Urol. 1997; 158(5): 1791-1793.
34. Lin WW, Lamb DJ, Wheeler TM, Lipschultz LI, Kim ED. In situ end-labeling of human testicular tissue demonstrates increased apoptosis in conditions of abnormal spermatogenesis. Fertil Steril. 1997; 68(6): 1065-1069.
35. D'Alessio A, Riccioli A, Lauretti P, Padula F, Muciaccia B, De Cesaris P, et al. Testicular Fasl is expressed by sperm cells. Proc Natl Acad Sci US A. 2001; 98(6): 3316-3321.
36. Del Giudice PT, Lima SB, Cenedese MA, Pacheco-Silva A, Bertolla RP, Cedenho AP. Expression of the Fas-ligand gene in ejaculated sperm from adolescents with and without varicocele. J Assist Reprod Genet. 2010; 27: 103-108.
37. Wu GJ, Chang FW, Lee SS, Cheng YY, Chen CH, Chen IC. Apoptosis-related phenotype of ejaculated spermatozoa in patients with varicocele. Fertil Steril. 2009; 91(3): 831-837.
38. Sharma R, Agarwal A. Spermatogenesis: an overview. In: Zini A; Agarwal A, editors. Sperm chromatin: biological and clinical applications in male infertility and assisted reproduction. 1st ed. New York: Springer; 2011; 19-44.
39. Balhorn R. A model for the structure of chromatin in mammalian sperm. J Cell Biol. 1982; 93(2): 298-305.
40. Oliva R. Protamines and male infertility. Hum Reprod Update. 2006; 12(4): 417-435.
41. El-Segini Y, Schill WB, Köhn FM, Zeid SA, Kamshushy AA, Marzouk S. Assessment of sperm functions in infertile patients with varicoceles. Andrologia. 2002; 34(5): 291-295.
42. Sadek A, Almohamdy AS, Zaki A, Aref M, Ibrahim SM, Mostafa T. Sperm chromatin condensation in infertile men with varicocele before and after surgical repair. Fertil Steril. 2011; 95(5): 1705-1708.
43. Talebi AR, Moein MR, Tabibnejad N, Ghasemzadeh J. Effect of varicoceles on chromatin condensation and DNA integrity of ejaculated spermatozoa using cytological examinations. Andrologia. 2008; 40(4): 245-251.
44. Azadi L, Abbasi H, Deemeh MR, Tavalaee M, Arbabian M, Pilevarian AA, et al. Zaditen (Ketotifen), as mast cell blocker, improves sperm quality, chromatin integrity and pregnancy rate after varicocelectomy. Int J Androl. 2011; 34(5 Pt 1): 446-452.
45. Marmar JL. The pathophysiology of varicoceles in the light of current molecular and genetic information. Hum Reprod Update. 2001; 7(5): 461-472.
46. Neuer A, Spandorfer SD, Giraldo P, Dieterle S, Rosenwaks Z, Wilkins SK. The role of heat shock proteins in reproduction. Hum Reprod Update. 2000; 6(2): 149-159.
47. Naaby-Hansen S, Herr JC. Heat shock proteins on the human sperm surface. J Reprod Immunol. 2010; 84(1): 32-40.
48. Eddy EM. Role of heat shock protein HSP70-2 in spermatogenesis. Rev Reprod. 1999; 4(1): 23-30.
49. Huszár G, Jakab A, Szakas D, Ozenç CC, Cayli S, Delpiano E. Fertility testing and ICSI sperm selection by hyaluronic acid binding: clinical and genetic aspects. Reprod Biomed Online. 2007; 14(5): 650-663.
50. Nasr Esfahani MH, Abbasi H, Mirhosseini Z, Ghasemi N, Razavi SH, Tavalaee M, et al. Can altered expression of HSPA2 in varicocele patients lead to abnormal spermatogenesis? Int J Fertil Steril. 2010; 4(3): 104-113.
51. Agarwal A, Prabakaran SA, Said TM. Prevention of oxidative stress injury to sperm. J Androl. 2005; 26(6): 654-660.
52. Chen SS, Huang WJ, Chang LS, Wei YH. Attenuation of oxidative stress after varicocelectomy in subfertile patients with varicocele. J Urol. 2008; 179: 639-642.
53. Moein MR, Dehghani VO, Tabibnejad N, Vahidi SAD. Reactive oxygen species (ROS) level in seminal plasma of infertile men and healthy donors. Iranian J Reprod Med. 2007; 5: 51-56.
54. Oborna I, Fingerova H, Novotny J, Brezinova J, Svobodova M, Aziz N. Reactive oxygen species in human semen in relation to leukocyte contamination. Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub. 2009; 153(1): 53-57.
55. Kemal Duru N, Morshedli M, Oehninger S. Effects of hydrogen peroxide on DNA and plasma membrane integrity of human spermatozoa. Fertil Steril. 2000; 74(6): 1200-1207.
56. Abd-Elmoaty MA, Saleh R, Sharma R, Agarwal A. Increased levels of oxidants and reduced antioxidants in semen of infertile men with varicocele. Fertil Steril. 2010; 94(4): 1531-1534.
57. Razi M, Sadirkhanlo RA, Malekinejad H, Sarafzadeh-Rezaei F. Varicocele time-dependently affects DNA integrity of sperm cells: evidence for lower in vitro fertilization rate in varicocele-positive rats. Int J Fertil Steril. 2011; 5(3): 122-196.
58. Sakamoto Y, Ishikawa T, Kondo Y, Yamaguchi K, Fujisawa M. The assessment of oxidative stress in infertile patients with varicocele. BJU Int. 2008; 101(12): 1547-1552.
59. Hammadeh ME, Filippos AA, Hamad MF. Reactive oxygen species and antioxidant in seminal plasma and their impact on male fertility. Int J Fertil Steril. 2009; 3(3): 87-110.
60. Simşek F, Türkeri L, Cevik I, Bircan K, Akdaş A. Role of apoptosis in testicular tissue damage caused by varicocele. Arch Esp Urol. 1998; 51(9): 947-950.
61. La Vignera S, Condorelli R, Vicari E, D’Agata R, Caloger AE. Effects of varicocelectomy on sperm DNA fragmentation, mitochondrial function, chromatin condensation, and apoptosis. J Androl. 2011; 32(3): 389-396.
62. Simon L, Brunborg G, Stevenson M, Lutton D, McManus J, Lewis SE. Clinical significance of sperm DNA damage in assisted reproduction outcome.Hum Reprod. 2010; 25(7): 1594-1608.
63. Tavalaee M, Razavi S, Nasr Esfahani MH. Influence of sperm chromatin anomalies on assisted reproductive technology outcome. Fertil Steril. 2009; 91(4): 1119-1126.
64. Cocuzza M, Cocuzza MA, Bragais FM, Agarwal A. The role of varicocele repair in the new era of assisted reproductive technology. Clinics (Sao Paolo). 2008; 63(3): 385-404.
65. Zini A, Azhar R, Baazeem A, Gabriel MS. Effect of microsurgical varicocelectomy on human sperm chromatin and DNA integrity: a prospective trial. Int J Androl. 2011; 34(1): 14-19.