A 55-Day-Old Female Infant Infected With 2019 Novel Coronavirus Disease: Presenting With Pneumonia, Liver Injury, and Heart Damage

Yuxia Cui1,4, Maolu Tian2,4 Dong Huang1, Xike Wang1, Xuying Huang4, Li Fan5, Liang Wang1, Yun Chen1, Wenwu Liu1, Kai Zhang1, Yue Wu1, Zhenhong Yang1, Jing Tao2, Jie Feng1, Kaiyu Liu1, Xianwei Ye2, Rongpin Wang4, Xiangyan Zhang3 and Yan Zha2

1Department of Pediatrics, Guizhou Provincial People’s Hospital, Guiyang, Guizhou, China, 2Department of Nephrology, Institute of Nephritic and Urinary Disease, Guizhou Provincial People’s Hospital, Guiyang, Guizhou, China, 3Department of Pulmonary Medicine, NHC Key Laboratory of Pulmonary Immunological Diseases, Guizhou Provincial People’s Hospital, Guiyang, China, 4Department of Radiology, Guizhou Provincial People’s Hospital, Guiyang, China, 5Department of Pediatrics, Guizhou Provincial People’s Hospital, Guiyang, China

Keywords. COVID-19 pneumonia; heart damage; liver injury.

A 55-day-old otherwise healthy female infant of mixed feeding became ill with rhinorrhea and a dry cough on January 28, 2020. She was admitted to our hospital on February 2, 2020. Before the onset of symptoms, she had been taken to Lu’an, Hubei Province by her parents for a family party between January 16 and January 24, 2020. At that party, the infant’s uncle and aunt, Wuhan residents, presented with cough and fever. Then, the child’s parents were diagnosed with coronavirus disease 2019 (COVID-19) on January 31, 2020 based on their symptoms, chest imaging, and viral ribonucleic acid (RNA) in pharyngeal swabs. The nasopharyngeal swab obtained from the infant also tested positive for severe acute respiratory syndrome coronavirus (SARS-CoV-2) on real-time, reverse-transcription polymerase-chain-reaction (RT-PCR) assay.

PATIENTS AND METHODS

On admission, the physical examination showed that vital signs were within normal ranges except for pharyngeal hyperemia. A chest computed tomographic (CT) scan was reported as showing patchy shadows and ground-glass opacity in the right lung (Figure 1A). Laboratory results reflected there were alterations in hepatic function measures and mildly abnormal myocardial zymogram (Table 1). Lymphocyte count, platelet count, CD8+ T lymphocyte count, and serum immunoglobulin (Ig)M level were all mildly elevated. Other laboratory examinations including hemoglobin, D-dimer, activated partial thromboplastin time, prothrombin time, C-reactive protein, erythrocyte sedimentation rate, and renal function in this patient showed no abnormalities. Both rotavirus in stool and cytomegalovirus deoxyribonucleic acid in blood were negative. Based on the definite contact history and the above test results, the infant was diagnosed as COVID-19 (ordinary type). She was isolated and treated empirically with the inhaled interferon α-1b (15 μg, bid), amoxicillin potassium clavulanate (30 mg/kg, q8h, intravenous glucose tolerance test [IVGTT]), reduced glutathione, ursodeoxycholic acid, and traditional Chinese medicine lotus qingwen.

On days 2 through 6 of hospitalization (days 7 through 11 of illness), the patient became sicker, presenting as a frequently productive cough accompanied by occasional tachycardia (150–170 beats per minute). Decreased arterial oxygen partial pressures (lowest PaO₂, 56 mmHg) with elevated lactic acid while breathing ambient air were found in the morning of hospital day 4 (Figure 2). She received sputum suctioning. Meanwhile, oxygen was supplied through a nasal cannula immediately, and ambroxol was administrated to eliminate phlegm. Because there was a trend of aggravation in the disease grade, a second chest CT scan was performed in the evening of hospital day 4 (illness day 9), which showed evidence of progressive pneumonia (Figure 1B). These radiographic findings coincided with a progress in respiratory status. Abnormal myocardial zymogram on admission and increased troponin I (0.025 μg per liter) on hospital day 4 indicated myocardia injury, so intravenous sodium creatine phosphate was added to protect her heart. During this period, the baby still had a good appetite, without diarrhea, oliguria, or shock. On day 11
Figure 1. Chest computed tomographic (CT) images of a 55-day-old female infant with COVID-19. NOTE: The scanning condition was low-dose CT with 80-kV tube voltage and 23- to 26-mAs tube current (automatic). (A) The CT images on admission (illness day 6): multiple patchy shadows and ground-glass opacity in the upper and lower lobes of the right lung (black arrow). (B) The CT images on hospital day 4 (illness day 9): compared with day 6 of illness, the lesion progressed and the range widened (gray arrow), accompanied with small consolidation shadow and a few stripe shadows (black arrow). (C) The CT images on hospital day 11 (illness day 16): compared with day 9 of illness, the inflammation of the right lung was obviously absorbed. Small patchy shadows with increased density and a few strip-like shadows were still observed in the upper lobe of the right lung (black arrow).
of the patient's illness, a stool specimen was collected for the first time to detect the viral RNA, and the PCR result showed positive.

**RESULTS**

On days 7 through 12 of hospitalization (days 12 through 17 of illness), the patient's clinical condition gradually improved.
and the respiratory symptoms disappeared on hospital day 11. Supplemental oxygen was discontinued, and her oxygen saturation values were 94% to 100% while breathing ambient air. The heart rate fluctuated between 120 and 140 beats per minute. The laboratory examinations on hospital day 10 showed elevated myocardial zymogram and abnormal liver function both recovered. On hospital day 11 (illness day 16), a third chest CT scan showed patchy shadows and ground-glass opacity were obviously absorbed (Figure 1C). In view of the patient's clinical manifestations and other laboratory findings, the *Citrobacter freundii* in 2 sputum cultures was considered as a colonized bacteria of the respiratory tract and was not being treated. Three pharyngeal swabs obtained from hospital day 10 to hospital day 13 were found to be negative for SARS-CoV-2 on RT-PCR assays. Amoxicillin potassium clavulanate was discontinued on hospital day 11, and interferon α-1b was discontinued on the following day. According to the latest interim guidance of the COVID-19 diagnosis and treatment in China, the patient met the discharge criteria. However, we found that her anal swabs collected on hospital day 11 and 13 were still positive for SARS-CoV-2 RNA. Thus, the patient remained hospitalized. From February 16, 2020, she was transferred to another hospital for continued isolation and observation. In the process of manuscript revision, we have learned that she was still asymptomatic during this period, and the anal swab collected on February 28, 2020 was negative for this viral RNA.

Regarding the patient's mother, we have learned that this viral RNA had been detected in the mother's stool sample on February 4, 2020 (illness day 8) and anal swab on February 7, 2020 (illness day 11), although she also had no abdominal symptoms. In addition, 3 consecutive tests of SARS-CoV-2 RNA in the monther's breastmilk were negative between February 2, 2020 and February 4, 2020. The mother's urine specimen collected on February 6, 2020 was negative for SARS-CoV-2.

**DISCUSSION**

The SARS-CoV-2 infection can lead to acute resolved or fatal pneumonia. Currently, the main source of infection is COVID-19 patients. The route of human-to-human transmission of SARS-CoV-2 is mainly through respiratory droplets and contacts. The fecal-oral route and maternal-infant transmission have not been confirmed or ruled out. Although the number of cases has increased rapidly, information on the clinical characteristics of affected pediatric patients is rare. We have learned that Middle East respiratory syndrome-coronavirus (MERS-CoV) in children is less frequent and seems to be associated with less mortality unless the patient has underlying comorbidities [1, 2]. This impression in children with MERS-CoV was also reported during the SARS-CoV infections in pediatric patients, in which symptoms were milder, there were few hospitalizations, and resulted in no death [3, 4]. Possible explanations include low exposure, presence of asymptomatic or mildly symptomatic patients, the immature immune system, and the presence of unidentified factors. The SARS-CoV-2 strains are less genetically similar to SARS-CoV (with 79% identity) and
MERS-CoV (approximately 50%) [5], so regarding the infection of SARS-CoV-2 in children, there are still many gaps in our understanding, including route of transmission, susceptibility, clinical course of patients, the disease pathogenesis, pharmacological therapies, prognosis, etc.

Wei et al [6] reported 9 infants diagnosed with COVID-19, including 1 baby at the age of 1 month, 26 days. However, there was no detailed information about the patients’ examination and treatment in this letter. Our report illustrated the full clinical course of the infected infant. The patient had

Table 1. Laboratory Indicators on Admission and the Changes After Treatment

| Measure                                | Reference Range | Illness Day 6, Hospital Day 1 | Illness Day 9, Hospital Day 4 | Illness Day 15, Hospital Day 10 |
|----------------------------------------|-----------------|-----------------------------|-----------------------------|-----------------------------|
| White cell count (×10⁹/L)              | 6–18            | 7.96                        | 10.04                       | 9.46                        |
| Lymphocyte count (×10⁹/L)              | 1.1–3.2         | 5.22*                       | 6.59*                       | 6.25*                       |
| Neutrophil count (×10⁹/L)              | 1.8–6.3         | 1.87                        | 2.44                        | 2.01                        |
| Platelet count (×10⁹/L)                | 125–350         | 408*                        | 449*                        | 604*                        |
| Hemoglobin (g/L)                       | 95–145          | 112                         | 91b                         | 101                         |
| Erythrocyte sedimentation rate (mm/h)  | 0–20            | 7                           | 2                           | -                           |
| C-reactive protein (mg/L)              | 0–5             | 0.56                        | 0.63                        | 0.32                        |
| Procalcitonin (ng/mL)                  | 0–0.046         | 0.15*                       | 0.11*                       | -                           |
| Álalanine aminotransferase (U/L)       | 7–40            | 84*                         | 49*                         | 33                          |
| Aspartate aminotransferase (U/L)       | 13–35           | 100*                        | 47*                         | 35                          |
| Total bilirubin (μmol/L)               | 3.4–20.5        | 33.7*                       | 20.1*                       | 10.9                        |
| Direct bilirubin (μmol/L)              | 0–8.6           | 25.2*                       | 16.6*                       | 7                           |
| Total bile acid (μmol/L)               | 0–10            | 154.4*                      | 89.8*                       | 46.4*                       |
| γ-hydroxybutyrate dehydrogenase (U/L)  | 0–25            | 46*                         | -                           | 25                          |
| Troponin I (μg/L)                      | 0–0.0156        | -                           | 0.025*                      | -                           |
| Creatinine (μmol/L)                    | 15–45           | 20                          | 19                          | 21                          |
| Blood urea nitrogen (mmol/L)           | 1.8–6           | 3.61                        | 2.09                        | 2.15                        |
| Serum immunoglobulin M (g/L)           | 0.06–0.21       | 0.66*                       | -                           | -                           |
| CD8⁺ T-cell count (cell/μL)            | 400–1700        | 2208*                       | -                           | -                           |
| D-dimer (μg/mL)                        | 0–1.5           | 0.54                        | -                           | -                           |
| Activated partial thromboplastin time (seconds) | 21.1–36.5   | 30.6                        | -                           | -                           |
| Prothrombin time (seconds)             | 9.2–12.2        | 9.7                         | -                           | -                           |
| Rotavirus in stool                     | Negative         | Negative                     | -                           | Negative                    |
| Cytomegalovirus DNA in blood           | Negative         | Negative                     | -                           | Negative                    |

Abbreviations: DNA, deoxyribonucleic acid.

aThe value in the patient was above normal.

bThe value in the patient was below normal.

Figure 2. Disease course of the infected infant and important information about her parents.
traveled to Hubei, China and had the sick contacts during her stay in Hubei. Although signs and symptoms caused by SARS-CoV-2 infection at the prodromal phase was atypical and nonspecific, the chest CT scan showed bilateral patchy shadows, in line with the atypical pneumonia. Acute liver injury and cardiac damage were also observed in this patient. Due to the difficulty of specimen collecting and the risk of cross-infection when going out for examination, other etiological tests of cardiac and hepatic abnormalities, except for rotavirus and cytomegalovirus detection, were not performed. Given the patient’s definite contact history of COVID-19, positive SARS-CoV-2 RNA, and good response to treatment, the 2 injuries were thought to be related to COVID-19. Elevated CD8+ cell count in peripheral blood and increased serum IgM level on admission might indicate that both cellular immune and humoral immune in acute infection have been activated to produce cytotoxic T lymphocyte and antibodies to kill and neutralize the virus. Huang et al [7] found that SARS-CoV-2 infection appears to be initially associated with an increased T helper 2 cell response, which might reflect a physiological reaction to suppress overt inflammatory responses.

Our case patient initially presented with mild dry cough and no fevers. However, from day 7 of her illness, her symptoms were gradually getting worse, especially on day 9 of her illness, coinciding with results of the second chest CT scan. The timing of our patient’s progression is consistent with that reported in adults [8, 9]. Due to the lack of effective verbal communication with infants, more frequent and careful clinical monitorings should be performed to find disease progression. When the patient’s status was worsening, symptomatic support was strengthened. Corticosteroids were not used in this patient, because her clinical condition was not completely out of control, and there is still no conclusive evidence of net benefit in the treatment of respiratory infection due to several viruses including SARS-CoV and MERS-CoV [10, 11]. Therefore, the timely and appropriate symptomatic support treatment and the patients’ own immune system may be the best means to combat the SARS-CoV-2 in pediatric patients.

CONCLUSIONS

We noticed that 3 consecutive tests of SARS-CoV-2 RNA in the breastmilk of the infant’s monther were negative. Whether the virus could not enter the milk due to some barrier and whether a confirmed case could continue to provide her breastmilk for baby deserve further study. In addition, the SARS-CoV-2 RNAs have been detected in the stool specimens or anal swabs of both the patient and her mother, indicating a potential alternative route of fecal-oral transmission, even in patients without gastrointestinal symptoms. However, the clinical significance of this detection of viral RNA outside the respiratory tract is unknown at this stage. Fecal RNA appeared later than that of pharyngeal swab in the mother, which might indicate that the virus is swallowed into the digestive tract during the process of its elimination from the respiratory tract. Moreover, we also observed that it takes longer for a fecal SARS-CoV-2 RNA to turn negative than that of a pharyngeal specimen in the infant. This phenomenon might indicate that the digestive tracts clear the virus more slowly than the respiratory tracts. Whether the patient isolation should not be terminated until SARS-CoV-2 RNA in the stool turns negative is worth exploring. Just like the other 2 studies [9, 12], we found no evidence of viral shedding in urine of the patient’s monther. However, improved systematic serial collection and testing of an increased number of urine samples is warranted to exclude this route of transmission. Finally, our understanding of COVID-19 is changing on a daily basis, and with continuous efforts from all sides, we hope and believe that this abominable disease can be controlled soon.

Notes

Acknowledgments. We thank the infant and her parents involved in the study. We acknowledge all healthcare workers involved in the diagnosis and treatment of patients with 2019 Novel Coronavirus Disease in Guizhou Provincial People’s Hospital.

Financial support. This study was supported by the Cultivation Fund from National Natural Science Foundation of Guizhou people’s Hospital (Number: [2018]5764-02) to Dr Cui.

Potential conflicts of interest. All authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. The funder had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication. There were also no any other financial interests or connections, direct or indirect, or other situations which had raised the question of bias in the work reported or the conclusions, implications or opinions stated.

References

1. Arabi YM, Balkhy HH, Hayden FG, et al. Middle East respiratory syndrome. N Engl J Med 2017; 376:584–94.
2. Memish ZA, Al-Tawfiq JA, Assiri A, et al. Middle East respiratory syndrome coronavirus disease in children. Pediatr Infect Dis J 2014; 33:904–6.
3. Hon KL, Leung CW, Cheng WT, et al. Clinical presentations and outcome of severe acute respiratory syndrome in children. Lancet 2003; 361:1701–3.
4. Li AM, Ng PC. Severe acute respiratory syndrome (SARS) in neonates and children. Arch Dis Child Fetal Neonatal Ed 2005; 90:F461–5.
5. Lu R, Zhao X, Li J, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. Lancet 2020; 395:1565–74.

6. Wei M, Yuan J, Liu Y, et al. Novel coronavirus infection in hospitalized infants under 1 year of age in China. JAMA 2020; DOI: 10.1001/jama.2020.2131.

7. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 coronavirus in Wuhan, China. Lancet 2020; 395:497–506.

8. Wang D, Hu B, Hu C, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus–infected pneumonia in Wuhan, China. JAMA 2020; DOI: 10.1001/jama.2020.1585.

9. Holshue ML, DeBolt C, Lindquist S, et al. First case of 2019 novel coronavirus in the United States. N Engl J Med 2020; 382:929–36.

10. Russell CD, Millar JE, Baillie JK. Clinical evidence does not support corticosteroid treatment for 2019-nCoV lung injury. Lancet 2020; DOI: 10.1016/S0140-6736(20)30317-2.

11. Shang L, Zhao J, Hu Y, Du R, Cao B. On the use of corticosteroids for 2019-nCoV pneumonia. Lancet 2020; 395:683–4–5.

12. Fuk-Woo Chan J, Yuan S, Kok K-H, et al. A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster. Lancet 2020; 395:514–23.