Features of the Course of Non-alcoholic Fatty Liver Disease in Experimental Animals at High Altitudes

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Introduction

Non-alcoholic fatty liver disease (NAFLD) is the most common liver disease worldwide, affecting nearly 25% of adults. As a rule, changes in the liver begin with ordinary steatosis, which rapidly progresses to non-alcoholic steatohepatitis and its complicated forms such as fibrosis and cirrhosis. Ultimately, towards the terminal stage of liver disease, hepatocellular carcinoma develops, which has a high morbidity and mortality [1]. The obvious differences between steatohepatitis and steatosis are the presence of an inflammatory infiltrate and an increase in serum levels of many inflammatory markers such as C-reactive protein, tumor necrosis factor (TNF-α), and interleukins IL-6 and IL-8 [2].

Experimental modeling of NAFLD plays a key role in the study of the etiology and mechanisms of pathogenesis. A method for modeling fatty liver degeneration in laboratory rats was proposed in 2005 by Ackermann et al. It is known that hepatic fructose metabolism stimulates lipogenesis regardless of insulin load [3]. Based on studies of the primary metabolism of fructose in the liver [4], [5], the development of NAFLD due to excess consumption of fructose should be considered a public health crisis. The increase in total bilirubin levels in the group of animals in diet rich in fructose is consistent with early studies. [6]. It is believed that the pathogenesis of NAFLD is based on a severe imbalance of lipid metabolism with the formation of hypercholesterolemia and hypertriglyceridemia [7], [8]. There are very few data in the literature on the development of NAFLD in high-altitude conditions. Of great interest is the study of shifts in an organism located in high altitude conditions, where, in addition to the action of the main factor - a decrease in the partial pressure of oxygen, a number of other factors act on the organism: daily temperature fluctuations, increased radiation of the ultraviolet part of the spectrum, etc. There are no data on the association of IL-4 and TNF-α with NAFLD developed in high mountains. Therefore, it was decided to study the disturbances caused by high-altitude hypoxia in a pressure chamber at a constant temperature. It is known that liver cells rank second after brain cells in sensitivity to hypoxia. The latest publications indicating the protective effect of chronic high-altitude hypoxia on the progression of NAFLD began to arrive from China only in July 2020 [9].

The role of pro- and anti-inflammatory cytokines in NAFLD in high-altitude conditions is not found in the...
literature. Thus, additional pathophysiological studies in animal models will help find answers to these questions. The aim of the study was to study the features of the course of NAFLD in experimental animals at high altitudes.

Materials and Methods

The study was carried out in accordance with the principles of the humanity of the directives of the European Community (86/609/EEC) and the Declaration of Helsinki. Prior to the experiment, the research plan, standardized procedures, and accompanying documentation were ethically reviewed and approved by the Ethics Committee of the Scientific and Production Association “Preventive Medicine” of the Ministry health care of the Kyrgyz Republic (conclusion No. 6 dated October 08, 2019).

The study was carried out in the field of pathophysiology. The experiments were carried out from March 14, 2020, for 10 weeks.

The study was conducted on 180 male Wistar rats weighing 150–200 g at the time of inclusion in the experiment. The rats were randomly assigned to control and experimental groups. The rats of the control group (CG) were kept on a standard diet (food “Eshka,” Russia, total caloric content of 3000 kcal/kg). NAFLD was modeled by the formation of metabolic syndrome in laboratory rats (Ackermann et al. 2005). During the study period, the animals received a special diet containing 20.7% proteins (casein), 5% fats (beef lard), 60% carbohydrates (fructose), 8% cellulose, 5% minerals, 1% vitamins [10]. This model makes it possible to reproduce NAFLD in an extremely short time, taking into account all the pathogenetic aspects of this pathology: hypercholesterolemia, triglyceridemia, insulin resistance. Animals were divided into 3 CG and 4 main groups (MG): “Control 1” (n = 20) - intact, healthy rats kept in low mountains (Bishkek, 800 m above sea level) on a standard diet; “Control 2” (n = 20) - hypobaric hypoxia (6000 m above sea level in a pressure chamber, daily exposure for 6 h for 5 weeks) on a standard diet; “Control 3” (n = 20) - hypobaric hypoxia (6000 m above sea level in a pressure chamber, 6 h–10 weeks) on a standard diet; “Main group 1’ (n = 30) - low-mountain group of rats for 5 weeks on a diet enriched with fructose and fat equally tended to suppress the synthetic function of the liver in low mountains and under pressure chamber hypoxia (57.0 ± 0.7 and 57.7 ± 0.18, respectively). Liver enzyme levels were uncertain. AST levels were high in all major groups, with a similar upward trend at weeks 5 and 10 of the fructose-fortified diet. Under conditions of pressure chamber hypoxia, the greatest shift was observed in ALT, the growth of which was statistically significant lower than in the low-mountain groups with p < 0.001. The opposite picture was observed in pigment metabolism. The total bilirubin in the high-altitude groups on a diet rich in fructose and fat was statistically significant higher relative to the low-altitude groups on the identical diet (15.3 ± 0.2 and 15.8 ± 0.2 versus 7.1 ± 0.14 and 4.1 ± 0.06). It should be noted that the initial levels of TC and LDL in the high mountain group

Results

Biochemical blood parameters in the studied groups are presented in Table 1. It should be noted that the CG-3 control group, after 10 weeks at altitude, had a higher total protein level than the low altitude group. The presence of animals on a diet enriched with fructose and fat equally tended to suppress the synthetic function of the liver in low mountains and under pressure chamber hypoxia (57.0 ± 0.7 and 57.7 ± 0.18, respectively). Liver enzyme levels were uncertain. AST levels were high in all major groups, with a similar upward trend at weeks 5 and 10 of the fructose-fortified diet. Under conditions of pressure chamber hypoxia, the greatest shift was observed in ALT, the growth of which was statistically significant lower than in the low-mountain groups with p < 0.001. The opposite picture was observed in pigment metabolism. The total bilirubin in the high-altitude groups on a diet rich in fructose and fat was statistically significant higher relative to the low-altitude groups on the identical diet (15.3 ± 0.2 and 15.8 ± 0.2 versus 7.1 ± 0.14 and 4.1 ± 0.06). It should be noted that the initial levels of TC and LDL in the high mountain group

The results were analyzed using the SPSS 16.0 statistical software package for Windows (IBM, USA). To assess the probabilities that the analyzed samples belong to general populations with a normal distribution, the Kolmogorov - Smirnov test was used. Taking into account the normal distribution of sample data (P > 0.05), Student’s t-test was used to compare them. The significance of differences between the groups was determined by the methods of nonparametric statistics, and within the group at 5 and 10 weeks by the method of parametric statistics. Descriptive treatment results were presented as mean and standard deviation (M ± SD). The Pearson correlation test was used to see the correlation between variables. A p < 0.05 was considered statistically significant at the 95% confidence level.
Table 1: Levels of pro- and anti-inflammatory cytokines

| Indicator                      | CG-1 (n = 10) | MG-1 (n = 20) | MG-2 (n = 20) | CG-2 (n = 10) | MG-3 (n = 20) | CG-3 (n = 10) | MG-4 (n = 20) | P-value |  
|-------------------------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------|  
| Mean total protein ± SD (g/l) | 66 ± 1.5     | 60 ± 0.5     | 57 ± 0.7     | 64.4 ± 0.21  | 61.2 ± 0.24  | 68 ± 0.1     | 57.7 ± 0.18  |        |  
| Mean ALT ± SD (IU/L)          | 80           | 76-84.1      | 127.7        | 69           | 87.6         | 68           | 102.9       |        |  
| Mean AST ± SD (IU/L)          | 68.8 ± 0.1   | 263.6 ± 5.4  | 347.5 ± 4.3  | 55.6 ± 0.2   | 254.1 ± 2.8  | 55.1 ± 0.1   | 266.3 ± 2.9  |        |  
| Mean Ritis coefficient ± SD   | 0.86 ± 0.1   | 2.3 ± 0.4    | 1.2 ± 0.2    | 0.8 ± 0.1    | 3.1 ± 0.5    | 0.8 ± 0.2    | 2.7 ± 0.4    |        |  
| Mean Total bilirubin ± SD (umol/l) | 1.4 ± 0.2 | 7.1 ± 0.14   | 4.1 ± 0.06   | 4.2 ± 0.06   | 15.3 ± 0.2   | 10.7 ± 0.1   | 15.8 ± 0.2   |        |  
| Mean TC ± SD (mmol/l)         | 1.2 ± 0.1    | 1.7 ± 0.2    | 2.1 ± 0.1    | 3.7 ± 0.1    | 6.6 ± 0.8    | 3.2 ± 0.1    | 5.3 ± 0.4    |        |  
| Mean LDL ± SD (mmol/l)        | 0.2 ± 0.1    | 0.2 ± 0.1    | 1.2 ± 0.1    | 2.5 ± 0.1    | 4.0 ± 0.5    | 2.0 ± 0.1    | 3.5 ± 0.2    |        |  
| Mean IL-4 ± SD (pg/ml)        | 4.4 ± 0.4    | 9.9 ± 0.6    | 19.8 ± 0.16  | 27.5 ± 0.55  | 34.5 ± 0.3   | 19.0 ± 0.1   | 22.6 ± 0.2   |        |  
| Mean TNF-α ± SD (pg/ml)       | 6.9 ± 0.7    | 17.8 ± 1.1   | 40.6 ± 0.6   | 49.5 ± 0.25  | 72.3 ± 0.3   | 36.5 ± 0.25  | 40.5 ± 0.3   |        |  

were higher compared to the low mountain group. The total cholesterol index also almost doubled in the main high-mountain groups and was significantly higher than the low-mountain animals with *p* < 0.001. The same tendency is observed after 5 weeks for LDL, an increase from 2.5 ± 0.1 to 4.0 ± 0.5 mmol/L in MG-3 versus MG-1 0.2 ± 0.1 and 0.2 ± 0.1 mmol/L, respectively, with *p* < 0.001. At the 10th week of ascent to altitude, the levels of total cholesterol and LDL cholesterol remained statistically significant high in the TG 4 group, although there was a downward trend. The cytokine activity in the high-altitude group increased sharply at 5 weeks. Thus, on the 35th day of stay at altitude, the content of IL-4 increased statistically significant from 27.5 ± 0.5 to 34.5 ± 0.3 pg/ml as compared with low mountains (*p* < 0.001). TNF-α statistically significant increased from 49.5 ± 0.25 to 72.3 ± 0.3 versus the low-mountain group on a fat diet. At 10 weeks of ascent to altitude in animals (“MG 4”), the levels of pro- and anti-inflammatory cytokines did not have statistically significant differences from the low-mountain group, which were also on FED (“MG 2”).

![Image](https://oamjms.eu/index.php/mjms/index)

The role of a diet rich in fructose in inflammation is well known [12]. The vulnerability of hepatocytes to cellular stress increases with the ectopic accumulation of fat, which activates the inflammatory process [13]. Experiments have shown that fructose promotes the synthesis of saturated fatty acids, which are able to activate Toll-like receptors 4 in liver Kupffer cells, activating cellular stress and aggravating inflammation [14]. The activation of the pro-inflammatory cytokine production pathway by Kupffer cells is confirmed by an increase in the proportion of “conventional” pro-inflammatory macrophages M1 compared to “alternative” anti-inflammatory macrophages M2 [15].

Thus, the balance of pro- and anti-inflammatory cytokines plays a key role in the development of NAFLD [16]. Our results demonstrate higher values of both pro-inflammatory and anti-inflammatory cytokines in animals on a diet enriched with fructose for 5 weeks under the influence of pressure chamber hypoxia. Hypoxia associated with chronic inflammation can induce pro-inflammatory cytokines [17]. It is possible that the increase in pro- and anti-inflammatory cytokines in the high-altitude group in our experiment is associated with hypoxia of adipose tissue. The results also demonstrate a gradual impairment of the synthetic and pigment functions of the liver under conditions of hypobaric hypoxia associated with a fructose diet.
Under conditions of hypobaric hypoxia, low ALT values are more likely associated with hypoergosis, which is the essence of any form of hypoxia and causes qualitatively the same type of metabolic and structural changes in various organs and tissues [18]. Although the literature lacks data on the effect of altitude hypoxia on biochemical parameters in NAFLD, several studies show that, in contrast to the lean cohort, obese animals and people are more susceptible to insulin resistance in a hypoxic environment at high altitude [19], [20], [21], [22]. As we know, insulin resistance plays a key role in the pathogenesis of NAFLD. The results of another group of scientists demonstrate that intermittent hypobaric hypoxia, such as chronic hypoxia, affects the levels of lipids in plasma and liver. Basically, the levels of TG and HDL are elevated in plasma, while the levels of TG and TC are elevated in liver tissue [23]. The behavior of the lipid spectrum under hypoxic conditions in patients with NAFLD has not been studied anywhere. In our work, we demonstrated a violation of the lipid profile with statistically significant positive correlation between hyperenzymemia, hypercholesterolemia, LDL values with TNF-α, and IL-4 values in NAFLD. This confirms the role and significance of pro- and anti-inflammatory markers in the pathogenesis of this disease.

The results of the study indicate the need for a comprehensive study of all factors affecting the development of NAFLD, such as genetic and epigenetic mechanisms, intestinal microbiota, etc.

Conclusion

In the high-altitude group of animals on a diet enriched with fructose for 5 and 10 weeks, statistically significant low ALT values are noted in comparison with the low-altitude groups on an identical diet. TNF-α and IL-4 in high-altitude animals on a diet enriched with fructose for 5 weeks has a higher value than in low-altitude group. The levels of pro-and anti-inflammatory cytokines statistically significantly correlate with cytolysis syndromes, hypercholesterolemia and impaired synthetic function against the background of liver hypoergosis.

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