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Case No 161: A 42-year-old journalist with fatigue, elevated liver function tests, hyperglycemia and pruritus

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Key words Autoimmune pancreatitis type 1 · IgG4 · Cholestasis · Total pancreatectomy · Leukocytoclastic vasculitis

Presentation of case

Dr. C. Putz-Bankuti: The patient, a 42-year-old white male, had been admitted to another teaching hospital for renal colic and subsequent shockwave lithotripsy 2 years before first presenting at the Outpatient Clinic for Gastroenterology and Hepatology at the University Medical Center in Graz. At that time, elevated liver function tests had been known for years and were attributed to hepatic steatosis: total bilirubin 2.5 mg/dl (normal: 0.1–1.2 mg/dl), direct bilirubin 1.8 mg/dl (normal: 0–1.0 mg/dl), alkaline phosphatase 813 U/l (normal: 35–105 U/l), gamma-glutamyl transferase (GGT) 1153 U/l (normal: <38 U/l), aspartate amino transferase (AST) 581 U/l (normal: <30 U/l), alanine amino transferase (ALT) 1294 U/l (normal: <35 U/l), lactate dehydrogenase (LDH) 334 U/l (normal: 120–250 U/l). Computed tomography (CT) of the abdomen revealed liver parenchymal fibrosis and steatosis, cholecytolithiasis, localized thickening of the wall of the common bile duct probably following previous cholangitis and...
enlarged lymph nodes along the celiac trunk and the transverse fissure.

The patient was again admitted to the same hospital for fatigue one year before the current admission for elevated liver function tests and hyperglycemia. Physical examination was unremarkable (weight 71 kg, height 170 cm, pulse 80/min, blood pressure 100/70 mmHg), but the patient complained of pruritus. Laboratory data: erythrocytes 4.4 T/L (normal: 4.50–5.90 T/L), hemoglobin (Hb) 13.2 g/dl (normal: 12.0–15.3 g/dl), hematocrit 36.5% (normal: 35.0–45.0%), eosinophils 9.9% (normal: <5%), AST 659 U/l, ALT 981 U/l, GGT 2030 U/l, AP 1260 U/l, LDH 385 U/l, total bilirubin 2.75 mg/dl (normal: 0.2–1.3 mg/dl), glucose 110 mg/dl (normal: 70–115 mg/dl), HbA1c 10% (normal: 4.3–5.9%), ferritin 530 ng/ml (normal: 30–400 ng/ml). Further laboratory data were all normal or negative: creatinine kinase, serum myoglobin, C-reactive protein, total protein, albumin, C-peptide, antibodies against insulin, tyrosine phosphatase and glutamic acid decarboxylase 65 were normal.

The medical history further revealed meningitis in his early childhood with hydrocephalus, shunt implantation with later revision, surgery for strabismus, tonsillectomy, and surgery following right shoulder plantation with later revision, surgery for strabismus, acid decarboxylase 65 were normal.

Answer insulin, tyrosine phosphatase and glutamic normal: <8 pmol/l), insulin, C-peptide, antibodies against insulin, tyrosine phosphatase and glutamic acid decarboxylase 65 were normal.

The patient sought a second opinion when someone brought up the subject of liver transplantation. Since the consulted physician did not see an indication for liver transplantation and the patient had not responded to ursodeoxycholic acid therapy, he was referred to the University Medical Center in Graz to be enrolled in a clinical trial studying 24-nor-ursodeoxycholic acid in patients with PSC. Except for several bland diverticula, colonoscopy was unremarkable. Follow-up MRCP and MRI showed slight accentuation of the first-order intrahepatic bile ducts but no significant biliary retention. The common bile duct was thin with slight periductal fibrosis but was not beaded. The pancreatic duct appeared unremarkable. Contrast enhancement showed a subsepted pseudo-cyst (7.0 × 3.6 cm) primarily localized in the tail of the pancreas but also extending to the omental bursa. The pancreas appeared grossly altered (Fig. 1) and several enlarged lymph nodes (diameter up to 1 cm) were visible along the mesentry. A panel of genetic tests was negative for mutations causing hereditary pancreatitis. Tumor markers including CA 19-9, CEA and CA 72-4 were unremarkable.

The further management of the patient led to the diagnosis.

Differential diagnosis

Dr. M. Peck-Radosavljevic: This complex case of a probably liver-associated disease was first recognized when the patient was admitted to hospital due to renal colic. At that time, elevated liver function tests had been known for years. Thus, it is not surprising that abdominal CT revealed pathological changes of the liver, such as damage to the liver parenchyma consistent with steatosis and fibrosis, cholecystolithiasis and local thickening of the wall of the common duct. After this first hospitalization the patient's medical history was unremarkable for 1 year, but he then had to be readmitted for fatigue, elevated liver function tests and hyperglycemia. Cholestasis had increased and hyperglycemia was prominent. Increased serum ferritin could be due to hemochromatosis causing both hepatic abnormalities and diabetes; however, hemochromatosis can be ruled out here because other iron parameters, such as transferrin and transferrin saturation, which are typically increased in iron storage diseases, were normal. Further laboratory data focusing on liver disease (e.g. albumin, C-reactive protein, prothrombin time, thyroid stimulating hormone, alpha1-antitrypsin and ceruloplasmin)
were also normal. Thus, it can be suggested that in the presence of cholestasis, the metabolic function of the liver had hardly been affected by the underlying disease. The IgM, IgG4, ANA and SMA levels were normal, so there was no indication of autoimmune liver disease.

According to the protocol, MRCP showed moderate distension of the intrahepatic bile ducts in the left lobe of the liver with irregular contours, localized beading, and alternating strictures and dilatations. The narrowing of the common bile duct that was more prominent in the central portion could explain the failure to visualize it by ERCP. The ERCP further detected an edematous papilla and an irregular pancreatic duct repeatedly releasing white viscous secretion, which might be an important clue for the underlying problem in this case. Histopathology of the liver biopsy revealed PSC and ruled out an overlap syndrome; however, in PSC one would expect higher inflammatory markers than found in this patient. Actually, laboratory data are more compatible with an overlap syndrome than with PSC in this case, but this was not confirmed by liver histology. The patient received ursodeoxycholic acid for PSC, cholestyramine, an antihistamine and an SSRI for treatment of pruritus, along with insulin for hyperglycemia. Under this treatment blood glucose, cholestasis, inflammation and liver function tests significantly improved, and according to the MRCP/MRI report, the beaded configuration of bile ducts disappeared; however, after this treatment the pancreas appeared grossly abnormal and contained a subsepted cyst, presumably a pseudocyst.

In summary, this case is very complex and there is no specific finding suggesting a definite diagnosis. Differential diagnosis of elevated liver function tests includes a wide range of diseases (Table 1); however, chronic hepatitis C and B, acute viral hepatitis, steatosis/steatohepatitis, hemochromatosis, alpha1-antitrypsin deficiency and Wilson’s disease can be ruled out as diagnoses because they were not confirmed by laboratory data. According to the protocol, hepatotoxic substances (medication/toxins) can also...
Table 1  Differential diagnoses of elevated liver function tests [4]

| Moderate elevation of transaminases (<5 × ULN) | Significant elevation of transaminases (>15 × ULN) | Elevation of alkaline phosphatase |
|-----------------------------------------------|-----------------------------------------------|----------------------------------|
| **Hepatic: ALT-predominant**                  |                                               |                                  |
| Acute viral hepatitis (A–E, herpes)          |                                               | Hepatobiliary                    |
| Medication/toxins                             |                                               |                                  |
| Ischemic hepatitis                            |                                               |                                  |
| Wilson’s disease                              |                                               |                                  |
| Acute bile duct obstruction                   |                                               |                                  |
| Medications                                   |                                               |                                  |
| **Hemochromatosis**                           |                                               |                                  |
| Acute Budd-Chiari syndrome                    |                                               | Infiltrating diseases of the liver|
| Medication/toxins                             |                                               |                                  |
| **Autoimmune hepatitis**                      |                                               |                                  |
| Alpha1-antitrypsin deficiency                 |                                               |                                  |
| Wilson’s disease                              |                                               |                                  |
| **Celiac disease**                            |                                               |                                  |
| **Hepatic: AST-predominant**                  |                                               |                                  |
| Alcohol-related liver injury                  |                                               |                                  |
| Steatosis/steatohepatitis                     |                                               |                                  |
| **Cirrhosis**                                 |                                               |                                  |
| **Non-hepatic**                               |                                               |                                  |
| Hemolysis                                     |                                               |                                  |
| Myopathy                                      |                                               |                                  |
| Thyroid disease                               |                                               |                                  |
| Strenuous exercise                            |                                               |                                  |
| Macro-AST                                     |                                               |                                  |

ULN upper limit of normal, EBV Epstein-Barr virus, CMV cytomegaly virus, AST aspartate-amino transferase, ALT alanine-amino transferase

be excluded. Autoimmune hepatitis may sometimes cause significantly elevated liver function tests and cholestasis (overlap syndrome), as present here. Negative serology does not necessarily rule out the disease but also does not really favor a suspicion of autoimmune hepatitis. The ALT level can also be elevated due to celiac disease, but this can be excluded because of the negative medical history for this disease. Since this situation was not AST-predominant, alcohol-related liver injury, steatosis/steatohepatitis and cirrhosis can also be ruled out as diagnoses. Non-hepatic causes given in Table 1 are also unlikely to have caused the patient’s problems, and the medical history does not hint at an ischemic event. Although the bile duct had been obstructed, an acute bile duct obstruction could not have been present because cholestasis was not as severe as expected in acute biliary obstruction and bilirubin was not as high as it usually is in this situation. Finally, nothing in the protocol suggests acute Budd-Chiari syndrome. Taking into account additional diagnoses that are related to elevated alkaline phosphatase (Table 1), primary biliary cirrhosis, infiltrating diseases of the liver, hepatic metastases, vanishing bile duct syndrome and benign recurrent cholestasis can be excluded due to lack of evidence. Although the patient was diagnosed with PSC, the rapid and significant improvement of cholestasis and liver function tests under treatment is very unusual for PSC and makes the diagnosis highly questionable. Except for infection, there is no evidence that the non-hepatic causes of elevated alkaline phosphatase summarized in Table 1 were present in this case. Laboratory data indicated increased eosinophils, possibly due to a parasitic infection. This suggestion is further strengthened by the diagnosis of splenomegaly, which among other causes such as portal hypertension (not present here) can be due to infections. Moderately enlarged lymph nodes along the portal area may also suggest more an infectious than a malignant cause of lymphadenopathy. The finding of a subsepted cyst in the pancreas provides an additional clue for the diagnosis of parasitic infection leading to cyst formation, i.e. echinococcosis, also known as hydatid disease or hydatidosis. Echinococcosis is caused by larval infection with Echinococcus granulosus and is characterized by long-term growth of metacestode (hydatid) cysts, occurring mainly in the liver and lungs but also in other organs of the intermediate host [1]. Hydatid cysts can be classified differently according to their appearance [2]. Subsepted cysts may be due to formation of daughter cysts that are frequent in echinococcosis [3].

Dr. M. Peck-Radosavljevic’s diagnosis

Echinococcosis with hydatid cyst (with daughter cysts) in the pancreas.

Discussion of case

Dr. P. Fickert: When the patient was referred to the outpatient clinic of the Division of Gastroenterology
and Hepatology at the University Medical Center in Graz, there was no evidence for pancreatitis in his medical history, while the radiologic findings revealed a subsepted pancreatic cyst of unclear origin. Thus, different causes such as intraductal papillary mucinous neoplasia (IPMN), probably associated with autoimmune pancreatitis (AIP), and mucous cystadenoma or carcinoma were considered. The finding of repeated release of white viscous secretion from the pancreatic duct actually did make us think of a mucous cystadenoma or carcinoma; however, the intermittent cholestasis resulting in significantly elevated liver function tests is atypical for cystadenoma or carcinoma and could also be due to different hepatic diseases as already discussed by Dr. Peck-Radosavljevic. Cyst formation in AIP is rare. Currently, only a small number of cases have been reported [5–10]. In the patient under discussion no specific diagnostic criteria for AIP were found; however, this does not necessarily rule out the diagnosis. An IPMN may be rarely associated with AIP [11–15]. Thus, it was thought that only histology would lead to the final diagnosis in this case. Performing transabdominal or transgastric biopsy of the cystic lesion was, however, not advisable because of vascular convolutes in the vicinity. Furthermore, adhesions of intestinal loops as documented in this patient are typical for tumors.

Since the radiologic findings did not provide a clear diagnosis and biopsy was not possible, the attending physicians suggested laparotomy to obtain tissue from the cystic lesion and to establish the diagnosis histologically.

**Dr. W. Fuerst:** Dr. Peck-Radosavljevic suspected echinococcosis; however, parasite screening was negative.

**Dr. G.J. Krejs:** Parasite screening tests are known to be negative sometimes, even though the patient is infected. Recently, we saw a case where the results had been negative for hookworm (*Ancylostoma duodenale*) three times although the patient was infected. Screening should always be repeated when a parasitic infection is suspected, even when one or two tests have been negative.

**Dr. H. Mueller:** Prior to surgery, the finding of a subsepted pancreatic pseudocyst of unclear origin by MRI was additionally investigated by CT, which suggested a cystic primary neoplasm, either cystadenoma or cystadenocarcinoma of the pancreas. At surgery, the chronically inflamed pancreas was found grossly enlarged with loss of its lobulated contour (“sausage-shaped”), very hard and in some areas shaped irregularly because of connective tissue attached to the pancreatic surface (Fig. 2). Macroscopically, the pancreas suggested carcinoma to the surgeon; however, intraoperative frozen sections revealed non-neoplastic chronic pancreatitis. Due to the discrepancy between macroscopic and microscopic appearance and the fact that the patient had already developed insulin-dependent diabetes, a total pancreatectomy was performed.

**Dr. P. Fickert:** In this case both MRI and CT could not clearly distinguish chronic pancreatitis from an intraductal pancreatic neoplasm, cystadenoma or carcinoma, which may sometimes even occur concomitantly. Recently, Roch et al. found that IPMNs are associated with a significantly higher rate of autoimmune diseases than expected in the general population [16]. About 82% of patients with AIP also have pancreatic intraepithelial neoplasms (Pan-IN), suggesting that AIP might be associated with an el-
Most of the pancreatic parenchyma (lower left corner) is replaced by fibrosis with a storiform pattern. Numerous inflammatory cells infiltrate the wall of a venous vessel (venolitis). Pancreatic duct surrounded by storiform fibrosis and inflammatory infiltrates. The periductal infiltrate contains lymphocytes and numerous plasma cells.

Evated risk for malignancy [17]. Furthermore, about 10% of cases with pancreatic ductal adenocarcinoma additionally show histological features of AIP [18]. Shiokawa et al. found that in some patients AIP develops as a paraneoplastic syndrome and described dermatomyositis as a concomitant condition [19]. Among the few reported cases of concurrent AIP and pancreatic cancer, some described AIP surrounding the cancer, whereas other reports described AIP in sites remote from the cancer. Further investigation is needed to determine whether AIP induces pancreatic cancer due to chronic inflammation, Pan-IN leads to AIP-like stromal tissue, or the occurrence of concurrent Pan-IN and AIP is coincidental [20]. Histological examination is essential to clearly distinguish AIP from intraductal pancreatic neoplasia, cystadenoma or carcinoma and possible overlaps. Biopsy of the suspected mass could not be done transabdominally or transgastrically for the reasons mentioned above. Resected specimens were examined histologically by Dr. Lackner.

Dr. C. Lackner: Macropscopically, the resected specimens of the duodenum and the grossly fibrotic pancreas were 29 × 3.8 cm and 12 × 5.5 × 3.6 cm in size. The pancreatic head was 6 × 4 × 3.6 cm and showed a whitish grey mass (2.5 × 2.5 × 2 cm) with a diffuse margin towards the duodenum and the peripancreatic fatty tissue. The common bile duct and the pancreatic duct both presented with high-grade stenosis. More than 100 enlarged lymph nodes were found in the specimens of small intestine and pancreas. The spleen (14.5 × 12 × 4.6 cm) appeared unremarkable. Histopathological findings included abundant areas with storiform fibrosis replacing parts of the pancreatic parenchyma with extension into the papilla, and partially surrounded the main and interlobular pancreatic ducts, which also showed irregular outlines (Fig. 3). The fibrous tissue was infiltrated mainly by lymphocytes and plasma cells, which were often centered around the pancreatic ducts. However, there was no granulocytic infiltration and IgG4 staining was negative. Further, histology of the pancreatic specimens revealed obliterator venulitis, neuritis and...
and lymphadenopathy with interfollicular reactive hyperplasia and atrophic lymph follicles. Storiform fibrosis, stenosis and IgG4-negative periductal lymphoplasmacytic inflammation without granulocytic infiltration also affected the common bile duct. These histopathological findings supported the diagnosis of autoimmune pancreatitis (AIP) type 1, even without elevated numbers of IgG4-positive plasma cells, associated with fibrosis and atrophy of the exocrine tissue, and with stenosis of the pancreatic and common bile duct.

The AIP is a distinct form of pancreatitis with autoimmune features and frequently presents with painless obstructive jaundice (60–75%), a pancreatic mass or focal enlargement of the pancreas. Pancreatic insufficiency in the form of new or worsened hyperglycemia and steatorrhea may be present and, in a rare case, acute pancreatitis as well [21]. The disease is characterized by abnormalities of histology, imaging, serology, other organ involvement, and response to (steroid) therapy (HISORt criteria; [22]), or different combinations of these cardinal features [23]. The international consensus diagnostic criteria (ICDC) for AIP describe two subtypes: type 1 lymphoplasmacytic sclerosing pancreatitis (LPSP), and type 2 idiopathic duct-centric pancreatitis (IDCP) or AIP with granulocytic epithelial lesion (GEL) [24]. The characteristic histological features of both types of AIP are summarized in Table 2. Massive infiltration by IgG4 positive lymphocytes is a histological hallmark of AIP [25, 26] but is not pathognomonic for the disease and was not found in this case. Histological review is essential for diagnosis of AIP and differentiation from other forms of chronic pancreatitis such as chronic pancreatitis-not otherwise specified (CP-NOS), follicular pancreatitis, hereditary chronic pancreatitis, paraudodenal chronic pancreatitis and lobular fibrosis with low-grade Pan-IN.

**Dr. G.J. Krejs:** In the past, AIP has been described by various terms including chronic inflammatory sclerosis of the pancreas, chronic sclerosing pancreatitis, non-alcoholic duct-destructive chronic pancreatitis, lymphoplasmacytic sclerosing pancreatitis, idiopathic tumefactive chronic pancreatitis and idiopathic duct-centric chronic pancreatitis [27]. With an estimated prevalence of <1–2.2 per 100,000 in the general population, AIP is a rare disorder [28, 29]. Type 1 is the most prevalent subtype worldwide, accounting for up to 80% of cases in the USA and AIP type 2 is relatively more common in Europe but type 1 still remains the more prevalent subtype [30]. This disease is typically diagnosed later in life (80% of patients are over 50 years of age at diagnosis) with type 1 diagnosed an average of 16 years later than type 2 [31, 32]. In type 1 AIP males are more often affected than females (3:1; [21]), but there is no gender predilection documented for type 2 [33]. While type 2 AIP is a pancreas-specific disease mostly without extrapancreatic organ involvement and negative serology for IgG4 and autoantibodies, type 1 AIP is suggested to be a pancreatic manifestation of an IgG4-related disease characterized by systemic inflammation of unknown origin [27]. Despite the frequent association with elevated levels of IgG4 in serum and plasma cells in tissue, the role of IgG4 in AIP remains unclear. In fact, it is even unclear whether IgG4 type antibodies in AIP behave like tissue-destructive antibodies, i.e. autoantibodies, or are overexpressed in response to an unknown stimulus [23]. In healthy individuals IgG4 comprises 4–6% of the total IgG [34] and is considered a non-inflammatory antibody because of its relative inability to fix complement and its poor capacity to bind Fc receptors [35, 36]. Thus, IgG4 may be secondarily induced to dampen the extensive immune reaction in AIP [23]. In contrast, IgG4 has been demonstrated to be self-reactive in patients with IgG4-related disease [37], which supports the suggestion of primary IgG4 tissue-destructive properties in AIP [23]. A positive test for IgG4 is not pathognomonic for AIP or IgG4-related diseases and can occur in benign inflammatory processes such as allergic diseases, parasitic infestations and pemphigus vulgaris [34], as well as in malignancies such as pancreatic and bile duct cancers [23]. Although not disease-specific, IgG4 has the highest diagnostic value as a single serological test in AIP because elevated serum IgG4 is found in up to 75% of patients [38]; however, a subset of type 1 AIP patients who are seronegative for IgG4 should not be classified as type 2 [39]. Other serological findings in AIP include hypergammaglobulinemia, elevated IgG, antinuclear antibodies, anti-smooth muscle antibodies, carbonic anhydrase II antibodies, lactoferrin antibodies and rheumatoid factor [40, 41].

**Dr. P. Fickert:** The pathogenesis of AIP is not completely understood. Autoimmune processes are suggested to play a pivotal role in genetically susceptible patients. Human leukocyte antigen (HLA) DRB1*0405-DQB1*0401 haplotype, substitution of aspartic acid at DQB1 46 [42, 43] and single nucleotide polymorphism in cytotoxic T lymphocyte-associated antigen-4 (CTLA-4), which is expressed on CD4+ and CD8+ T cells and important for the regulation of T cell stimulation [44, 45], are discussed as key genetic factors in AIP. Moreover, Helicobacter pylori could probably trigger AIP via molecular mimicry in genetically predisposed individuals due to the significant homology between human carbonic anhydrase II and alpha-carbonic anhydrase of Helicobacter pylori with the homologous segments containing the binding motif of the HLA molecule DRB1*0405 [46]. Complement activation [47], Th2 cells and regulatory T cells (Treg) [48] are also hypothesized to be involved in the pathogenesis of AIP.

The clinical presentation of AIP is non-specific, sharing overlapping features with other diseases such as pancreatic cancer. Diagnosis requires a variable
### Table 2  Criteria for autoimmune pancreatitis (AIP) types 1 and 2 [24]

| Criterion                      | AIP type 1 | Level 1 |                  | Level 2 |                  |
|-------------------------------|------------|---------|------------------|---------|------------------|
| **Parenchymal imaging**       |            |         | Typical: diffuse enlargement with delayed enhancement (sometimes associated with rim-like enhancement) | Indeterminate (including atypical): Segmental/focal enlargement with delayed enhancement |
| **Ductal imaging (ERP)**      |            |         | Long (>1/3 length of the main pancreatic duct) or multiple strictures without marked upstream dilatation | Segmental/focal narrowing without marked upstream dilatation (duct size <5 mm) |
| **Serology**                  |            |         | IgG4 > 2 × upper limit of normal value | IgG4 1–2 × upper limit of normal value |
| **Other organ involvement**   |            |         | a or b | a or b |
|                               |            |         | a. Histology of extrapancreatic organs, any 3 of the following: | a. Histology of extrapancreatic organs including endoscopic biopsies of bile duct, both of the following: |
|                               |            |         | (I) Marked lymphoplasmacytic infiltration with fibrosis and without granulocytic infiltration | (I) Marked lymphoplasmacytic infiltration without granulocytic infiltration |
|                               |            |         | (II) Storiform fibrosis | (II) Abundant (>10 cells/HPF) IgG4-positive cells |
|                               |            |         | (III) Obliterative phlebitis | (III) Abundant (>10 cells/HPF) IgG4-positive cells |
|                               |            |         | (IV) Abundant (>10 cells/HPF) IgG4-positive cells | |
|                               |            |         | Typical radiological evidence | b. Physical or radiological evidence |
|                               |            |         | At least 1 of the following: | At least 1 of the following: |
|                               |            |         | (I) Segmental/multiple proximal (hilar/intrahepatic) or proximal and distal bile duct stricture | (I) Symmetrically enlarged salivary/lachrymal glands |
|                               |            |         | (II) Retroperitoneal fibrosis | (II) Radiological evidence of renal involvement described in association with AIP |
| **Histology of the pancreas** |            |         | LPSP and at least 3 of the following: | LPSP and any 2 of the following: |
|                               |            |         | (I) Periductal lymphoplasmacytic infiltrate without granulocytic infiltration | (I) Periductal lymphoplasmacytic infiltrate without granulocytic infiltration |
|                               |            |         | (II) Obliterative phlebitis | (II) Obliterative phlebitis |
|                               |            |         | (III) Storiform fibrosis | (III) Storiform fibrosis |
|                               |            |         | (IV) Abundant (>10 cells/HPF) IgG4-positive cells | (IV) Abundant (>10 cells/HPF) IgG4-positive cells |
| **Response to steroids**      |            |         | Diagnostic steroid trial | Rapid (≤ 2 weeks) radiologically demonstrable resolution or marked improvement in pancreatic/extrapancreatic manifestations |

**AIP** autoimmune pancreatitis, **ERP** endoscopic retrograde pancreatography, **LPSP** lymphoplasmacytic sclerosing pancreatitis, **HPF** high power field, **IDCP** idiopathic duct-centric pancreatitis

1 atypical; some AIP cases may show low-density mass, pancreatic duct dilatation or distal atrophy. Such atypical imaging findings in patients with obstructive jaundice and/or pancreatic mass are highly suggestive of pancreatic cancer. Such patients should be managed as pancreatic cancer unless there is strong collateral evidence for AIP and a thorough work-up for cancer is negative

2 Endoscopic biopsy of the duodenal papilla is a useful adjunctive method because the ampulla is often pathological in AIP

3 Diagnostic steroid trial should be conducted carefully by pancreatologists with caveats only after negative workup for cancer including endoscopic ultrasound-guided fine needle aspiration
process, there are a number of associated extrapancreatic manifestations that occur in up to 77% of patients [52]. Extrapancreatic manifestations can be useful in the diagnosis of AIP and, therefore, are part of the HISORt criteria. The most common are hilar lymphadenopathy, sclerosing cholangitis, retroperitoneal fibrosis, salivary and lacrimal gland involvement and tubulointerstitial nephritis (summarized by [53]), worsening or onset of diabetes mellitus or symptoms such as back pain, weight loss and fatigue [40, 52]. Conditions less frequently reported are hypophysitis and chronic thyroiditis [54]. Abdominal pain is more common in patients with type 2 AIP (68%) than in those with type 1 (41%); inflammatory bowel disease is only observed in type 2 [24, 31, 40]. Biliary disease associated with jaundice is one of the most common extrapancreatic manifestations of AIP and mainly attributable to obstruction at the level of the intrapancreatic portion of the common bile duct, associated with an inflammatory pancreatic head mass. This condition occurring in 20–88% of cases of AIP has been termed IgG4-associated cholangitis (IAC) [55]. An overlap between IAC and PSC can be suggested because serum IgG4 levels are increased in 9–36% of patients with PSC but only in 1% of patients with other liver diseases [56, 57]. Interestingly, PSC patients with elevated serum IgG4 levels have a more rapid disease progression than those with normal levels [55].

This patient showed several extrapancreatic manifestations including lymphadenopathy, sclerosing cholangitis, jaundice and fatigue. Moreover, he complained of pruritus and skin manifestations that had been seen by the consulting dermatologist, Dr. Wutte.

**Dr. N. Wutte:** The patient was first seen at the outpatient dermatological clinic of the University Medical Center in Graz a few days prior to total pancreatectomy because he had rapidly developed exanthematic lesions on the lower extremities. The purpuric plaques were sharply delineated at the sock line, had a diameter of up to 3 cm and were reaching up the legs, but without vesication or necrosis (Fig. 4). The patient was diagnosed with leukocytoclastic vasculitis and successfully treated with local corticosteroids. As there were repeated relapses during the following year, a biopsy was performed to confirm the diagnosis of leukocytoclastic vasculitis (Fig. 5). Leukocytoclastic vasculitis is an immune complex mediated disorder with degradation of leukocytes and subsequent destruction of small postcapillary venules. Clinically, leukocytoclastic vasculitis is characterized by palpable purpuric lesions. In children the most common type of vasculitis is IgA-mediated, in adults IgM and IgG-mediated vasculitis is more prevalent. The disease can be caused by specific medication, infections, tumors or hematologic diseases and is frequently observed in autoimmune diseases; however, in up to 50% of patients the underlying cause of immune-me-

**Fig. 4** Purpuric, partly confluent plaques up to 1.5 cm in size on the lower extremities with central bullous transformation in larger lesions

**Fig. 5** Skin biopsy showing leukocytoclastic vasculitis. Perivascular infiltrate of neutrophilic and eosinophilic granulocytes (gc) with focal leukocytolysis in the superficial dermis, fibroid necrosis of the vessel walls (n)
diated leukocytoclastic vasculitis remains unclear [58, 59]. Cutaneous small vessel vasculitis in association with AIP is rare and was first documented by Garg et al. in a patient with IgG4 positive AIP type 1 [60]. General evidence in support of the association between extrapancreatic manifestations and AIP include (1) shared characteristic histopathological findings of lymphoplasmacytic infiltration, IgG4-positive plasma cell infiltration, obliterator phlebitis and storiform fibrosis, (2) frequent co-existence or occurrence, (III) predictably favorable response to steroids and (4) differentiation from the lesion of the corresponding organ [27]. The immunopathological mechanism between AIP and cutaneous purpuric leukocytoclastic vasculitis is not clear but an increased deposition of immune complexes formed in response to the underlying systemic disease is a possible explanation; however, leukocytoclastic vasculitis may even occur without demonstrable histological immunoreactivity [61]. The discussed IgG4 negative patient reported repeated relapses of the purpuric rash with intermittent improvement. This might have been due to physical activity consistently identified as a precipitating and/or aggravating factor in leukocytoclastic vasculitis [61].

**Dr. P. Fickert:** At a follow-up visit one year after total pancreatectomy, the patient’s liver function tests were all normal.

**Dr. M. Peck-Radosavljevic:** This case shows that making the correct diagnosis and differentiating AIP from other diseases, especially pancreatic cancer, is of utmost importance. Use of standard diagnostic criteria will usually allow AIP to be distinguished from pancreatic cancer. In some patients, pancreatic biopsy, a therapeutic trial with steroids, or surgery (as in the discussed case) will be needed to clarify the diagnosis of AIP.

### Final diagnosis

Autoimmune pancreatitis type 1 (IgG4 negative) with associated leukocytoclastic vasculitis.

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### Conflict of interest

E. Fabian, M. Peck-Radosavljevic, E. Krones, H. Mueller, C. Lackner, C. Spreizer, C. Putz-Bankuti, W. Fuerst, N. Wutte, P. Fickert, H. Mischinger and G.J. Krejs declare that they have no competing interests.

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### References

1. Moro P, Schantz PM. Echinococcosis: A review. Int J Infect Dis. 2008;13:125–33.
2. World Health Organization, Informal Working Group (IFWG). International classification of ultrasound images in cystic echinococcosis for application in clinical and field epidemiological settings. Acta Trop. 2003;85:253–61.
3. Fabian E, Madl C, Horn S, Kornprat P, Maderthaner R, Aigelsreiter A, Krause R, Fickert P, Krejs GJ. Clinical-pathological conference series from the Medical University of Graz: case no. 151: 19-year-old student from Albania with emergency admission due to shock. Wien Klin Wochenschr. 2015;127(3–4):151–9.
4. Green RM, Flamm S. AGA technical review on the evaluation of liver chemistry tests. Gastroenterology. 2002;123(4):1367–84.
5. Nishimura T, Masaoaka T, Suzuki H, Aiura K, Nagata H, Ishii H. Autoimmune pancreatitis with pseudocysts. J Gastroenterol. 2004;39(10):1005–10.
6. Welsch T, Kleeff J, Esposito I, Büchler MW, Friess H. Autoimmune pancreatitis associated with a large pancreatic pseudocyst. World J Gastroenterol. 2006;12(36):5904–6.
7. Kawakami H, Kuvatani M, Shinada K, Yamato H, Hirano S, Kondo S, Yonemori A, Itoh T, Matsuno Y, Asaka M. Autoimmune pancreatitis associated with hemorrhagic pseudocysts: A case report and literature review. Intern Med. 2008;47(7):603–8.
8. Chang KA, Kim TN, Lee SH. Autoimmune pancreatitis complicated by an infected pseudocyst. Clin J Gastroenterol. 2010;3(3):168–73.
9. Sohn JW, Cho CM, Jung MK, Park SY, Jeon SW. A Case of autoimmune pancreatitis manifested by a pseudocyst and IgG4-associated cholangitis. Gut Liver. 2012;6(1):132–5.
10. Gompertz M, Morales C, Aldana H, Castillo J, Berger Z. Cystic lesions in autoimmune pancreatitis. Case Rep Gastroenterol. 2015;9(3):366–74.
11. Tabata T, Kamisawa T, Hara S, Kuruma S, Chiba K, Kuwata G, Fujwara T, Egashira H, Koizumi S, Endo Y, Koizumi K, Fujiwara J, Arakawa T, Momma K, Horiuchi S, Hishima T, Kurata M, Honda G, Kloppe G. Intraductal papillary mucinous neoplasm of the pancreas and IgG4-related disease: A coincidental association. Pancreatology. 2013;13(4):379–83.
12. Nakaji S, Hirata N, Fujii H, Iwaki K, Shiratori T, Kobayashi M, Wakisugi S, Ishii E, Takeyama H, Hoshi K. A case of focal autoimmune pancreatitis (AIP) mimicking an intraductal papillary mucinous neoplasm (IPMN). Clin J Gastroenterol. 2013;6(4):329–33.
13. Urata T, Naito Y, Izumi Y, Takekuma Y, Yokomizo H, Nagamine M, Fukuda S, Notohara K, Hifumi M. Localized autoimmune pancreatitis type 1 superimposed upon pre-existing intraductal papillary mucinous neoplasms. World J Gastroenterol. 2013;19(7):9127–32.
14. Vaquero EC, Salcedo MT, Cuatrecasas M, De León H, Merino X, Navarro S, Ginés A, Abu-Suboh M, Balsells J, Fernández-Cruz L, Molero X. Autoimmune pancreatitis type-1 associated with intraductal papillary mucinous neoplasm: Report of two cases. Pancreatology. 2014;14(4):316–8.
15. Hedayat AA, Lisovsky M, Suriawinata AA, Longnecker DS. Association of IgG4 response and autoimmune pancreatitis with intraductal papillary-mucinous neoplasms. Pancreatology. 2017;17(2):263–6.
16. Roch AM, Rosati CM, Cioffi JL, Ceppa EP, DeWitt JM, Al-Haddad MA, House MG, Zyromski NJ, Nakeeb A, Schmidt CM.
Intraductal papillary mucinous neoplasm of the pancreas, one manifestation of a more systemic disease? Am J Surg. 2016;211(3):512–8.

17. Gupta R, Khosroshahi A, Shinagare S, Fernandez C, Ferrone C, Lauwers GY, Stone JH, Deshpande V. Does autoimmune pancreatitis increase the risk of pancreatic carcinoma?: A retrospective analysis of pancreatic resections. Pancreas. 2013;42(3):506–10.

18. Zhang X, Liu X, Joseph L, Zhao L, Hart J, Xiao SY. Pancreatic ductal adenocarcinoma with autoimmune pancreatitis-like histologic and immunohistochemical features. Hum Pathol. 2014;45(3):621–7.

19. Shiokawa M, Kodama Y, Yoshimura K, Kawanami C, Mimura J, Yamashita Y, Asada M, Kikuyama M, Okabe Y, Inokuma T, Ohana M, Kukury H, Takeda K, Tsuji Y, Minami R, Sakuma Y, Kuriyama K, Ota Y, Tanabe W, Maruno T, Kuriya A, Sawai Y, Uza N, Watanabe T, Haga H, Chiba T. Risk of cancer in patients with autoimmune pancreatitis. Am J Gastroenterol. 2013;108(4):610–7.

20. Takano Y, Nagahama M, Yamamura E, Maruoka N, Yokomizo K, Mizukami H, Tanaka J, Ohike N. A case of concurrent pancreatic intraepithelial neoplasia and type 1 autoimmune pancreatitis with marked pancreatic duct dilatation. Clin J Gastroenterol. 2016;9(4):266–71.

21. Hart PA, Kamisawa T, Bruggie WR, Chung JB, Culver EL, Chung DD, Topazian MD, Takahashi Y, Yokomizo K, Mizukami H, Shimosegawa T, Soetikno R, Lee KT, Liao WC, Löhr M, Okazaki K, Ryu JK, Schleinitz N, Shimizu K, Shimosegawa T, Soetikno R, Webster G, Yadav D, Zen Y, Chari ST. Long-term outcomes of autoimmune pancreatitis: A multicentre, international analysis. Gut. 2013;62(12):1771–6.

22. Chari ST, Smyrk TC, Levy MJ, Petersen BT, Topazian MD, Takahashi Y, Czako L, Frulloni L, Gu V L, Gress TM, Kim MH, Kawa S, Lee KT, Lohr M, Okazaki K, Ryu JK, Schleinitz N, Shimizu K, Shimosegawa T, Soetikno R, Webster G, Yadav D, Zen Y, Chari ST. Long-term outcomes of autoimmune pancreatitis: A multicentre, international analysis. Gut. 2013;62(12):1771–6.

23. Zhang L, Chari ST, Smyrk TC, Levy MJ, Topazian MD, Takahashi N, Zhang L, Clain JE, Pearson RK, Petersen BT, Vege SS, Farnell MR. Diagnosis of autoimmune pancreatitis: The Mayo Clinic experience. Clin Gastroenterol Hepatol. 2006;4(8):1010–6.

24. Hart PA, Zen Y, Chari ST. Recent advances in autoimmune pancreatitis. Gastroenterology. 2015;149(1):39–51.

25. Shimosegawa T, Chari ST, Frulloni L, Kamisawa T, Kawa S, Mino-Kenudson M, Kim MH, Kloppegl G, Lorch LM, Lohr M, Notohara K, Okazaki K, Schneider A, Zhang L, International Association of Pancreatology. International consensus diagnostic criteria for autoimmune pancreatitis: Guidelines of the International Association of Pancreatology. Pancreas. 2011;40(3):352–8.

26. Zhang L, Chari ST, Smyrk TC, Levy MJ, Topazian MD, Takahashi N, Zhang L, Clain JE, Pearson RK, Petersen BT, Vege SS, Farnell MR. Diagnosis of autoimmune pancreatitis: The Mayo Clinic experience. Clin Gastroenterol Hepatol. 2006;4(8):1010–6.

27. Hart PA, Zen Y, Chari ST. Recent advances in autoimmune pancreatitis. Gastroenterology. 2015;149(1):39–51.

28. Shiokawa M, Kodama Y, Yoshimura K, Kawanami C, Mimura J, Yamashita Y, Asada M, Kikuyama M, Okabe Y, Inokuma T, Ohana M, Kukury H, Takeda K, Tsuji Y, Minami R, Sakuma Y, Kuriyama K, Ota Y, Tanabe W, Maruno T, Kuriya A, Sawai Y, Uza N, Watanabe T, Haga H, Chiba T. Risk of cancer in patients with autoimmune pancreatitis. Am J Gastroenterol. 2013;108(4):610–7.

29. Takano Y, Nagahama M, Yamamura E, Maruoka N, Yokomizo K, Mizukami H, Tanaka J, Ohike N. A case of concurrent pancreatic intraepithelial neoplasia and type 1 autoimmune pancreatitis with marked pancreatic duct dilatation. Clin J Gastroenterol. 2016;9(4):266–71.

30. Sugumar A, Klöppel G, Chari ST. Autoimmune pancreatitis: Pathologic subtypes and their implications for its diagnosis. Am J Gastroenterol. 2009;104(9):2308–10.

31. Kamisawa T, Chari ST, Giday SA, Kim MH, Chung JB, Lee Kt, Werner J, Bergmann F, Lorch LM, Mayerle J, Pickartz T, Lohr M, Schneider A, Frulloni L, Webster GL, Reddy DN, Liao WC, Wang HP, Okazaki K, Shimosegawa T, Kloppegl G, Go VL. Clinical profile of autoimmune pancreatitis and its histological subtypes: An international multicenter survey. Pancreas. 2011;40(6):808–184.

32. Hamano H, Kawa S, Horiuichi A, Unno H, Furuya N, Akamatsu T, Fukushima M, Nikaido T, Nakayama K, Usuda N, Kiyosawa K. High serum IgG4 concentrations in patients with sclerosing pancreatitis. N Engl J Med. 2001;344(10):732–8.

33. Zamboni G, Lüttges J, Capelli P, Frulloni L, Cavallini G, Pederzoli P, Leins A, Longnecker D, Klöppel G. Histopathological features of diagnostic and clinical relevance in autoimmune pancreatitis: A study on 53 resection specimens and 9 biopsy specimens. Virchows Arch. 2004;455(6):552–63.

34. Kawa S, Okazaki K, Kamisawa T, Kubo K, Ohara H, Hasebo O, et al. Amendment of the Japanese consensus guidelines for autoimmune pancreatitis, 2013 II. Extrapancreatic lesions, differential diagnosis. J Gastroenterol. 2014;49(5):765–84.

35. Aalberse RC, Stapel SO, Schuurman J, Rispens T. Immunoglobulin G4: An odd antibody. Clin Exp Allergy. 2009;39(4):469–77.

36. Nirula A, Glaser SM, Kalled SL, Taylor FR. What is IgG4?: A review of the biology of a unique immunoglobulin subtype. Curr Opin Rheumatol. 2011;23(1):119–24.

37. Matteo H, Mahajan VS, Della-Torre E, Sekigami Y, Car ruthers M, Wallace ZS, Deshpande V, Stone JH, Pillai S. De novo oligoclonal expansions of circulating plasmablasts in active and relapsing IgG4-related disease. J Allergy Clin Immunol. 2014;134(3):679–87.

38. Choi EK, Kim MH, Lee TY, Kwon S, Oh HC, Hwang CY, Seo DW, Lee SS, Lee SK. The sensitivity and specificity of serum immunoglobulin G and immunoglobulin G4 levels in the diagnosis of autoimmune chronic pancreatitis: Korean experience. Pancreas. 2007;35(2):156–61.

39. Sah RP, Chari ST. Autoimmune pancreatitis: An update on classification, diagnosis, natural history and management. Curr Gastroenterol Rep. 2012;14(2):95–105.

40. Okazaki K, Kawa S, Kamisawa T, Ito T, Inui K, Irie H, Nishino T, Notohara K, Nishimori I, Tanaka S, Nishiyama T, Suda K, Shiratori K, Tanaka M, Shimosegawa T, Working Committee of the Japan Pancreas Society and the Research Committee for Intractable Pancreatic Disease. Amendment of the Japanese consensus guidelines for autoimmune pancreatitis, 2013 I. Concept and diagnosis of autoimmune pancreatitis. J Gastroenterol. 2014;49(4):567–88.

41. Kamisawa T, Okamoto A. Autoimmune pancreatitis: Proposal of IgG4-related sclerosing disease. J Gastroenterol. 2006;41(7):613–25.
polymorphisms in Japanese patients. Am J Gastroenterol. 2008;103(3):588–94.
45. Chang MC, Chang YT, Tien YW, Liang PC, Jan IS, Wei SC, Wong JM. T-cell regulatory gene CTLA-4 polymorphism/haplotype association with autoimmune pancreatitis. Clin Chem. 2007;53(9):1700–5.
46. Guarneri F, Guarneri C, Benvenga S. Helicobacter pylori and autoimmune pancreatitis: Role of carbonic anhydrase via molecular mimicry? J Cell Mol Med. 2005;9(3):741–4.
47. Muraki T, Hamano H, Ochi Y, Komatsu K, Komiyama Y, Arakura N, Yoshizawa K, Ota M, Kawa S, Kiyosawa K. Autoimmune pancreatitis and complement activation system. Pancreas. 2006;32(1):16–21.
48. Zen Y, Fujii T, Harada K, Kawano M, Yamada K, Takahira M, Nakanuma Y. Th2 and regulatory immune reactions are increased in immunoglobulin G4-related sclerosing pancreatitis and cholangitis. Hepatology. 2007;45(6):1538–46.
49. Hong SP, Park SW, Chung JP, Chung JB, Kang JH, Kim J, Kim MJ, Park YN, Song SY. Autoimmune pancreatitis with effective steroid therapy. Yonsei Med J. 2003;44(3):534–8.
50. Ito T, Nakanou I, Koyanagi S, Miyahara T, Migita Y, Ogoshi K, Sakai H, Matsunaga S, Yasuda O, Sumii T, Nawata H. Autoimmune pancreatitis as a new clinical entity. Three cases of autoimmune pancreatitis with effective steroid therapy. Yonsei Med J. 1997;38(7):1458–68.
51. Sugumar A, Chari ST. Diagnosis and treatment of autoimmune pancreatitis. Curr Opin Gastroenterol. 2010;26(5):513–8.
52. Xin L, He YX, Zhu XJ, Zhang QH, Hu LH, Zou DW, Jin ZD, Chang XJ, Zheng JM, Zuo CJ, Shao CW, Jin G, Liao Z, Li ZS. Diagnosis and treatment of autoimmune pancreatitis: Experience with 100 patients. Hepatobiliary Pancreat Dis Int. 2014;13(6):642–8.
53. O’Reilly DA, Malde DJ, Duncan T, Rao M, Fillobbos R. Review of the diagnosis, classification and management of autoimmune pancreatitis. World J Gastrointest Pathophysiol. 2014;5(2):71–81.
54. Stone JH, Zen Y, Deshpande V. IgG4-related disease. N Engl J Med. 2012;366(6):539–51.
55. Kalaitzakis E, Webster GJ. Review article: Autoimmune pancreatitis—management of an emerging disease. Aliment Pharmacol Ther. 2011;33(3):291–303.
56. Ko SB, Mizuno N, Yatabe Y, Yoshikawa T, Ishiguro H, Yamamoto A, Azuma S, Naruse S, Yamao K, Muallem S, Goto H. Corticosteroids correct aberrant CFTR localization in the duct and regenerate acinar cells in autoimmune pancreatitis. Gastroenterology. 2010;138(5):1988–96.
57. Moon SH, Kim MH, Park DH, Hwang CY, Park SJ, Lee SS, Seo DW, Lee SK. Is a 2-week steroid trial after initial negative investigation for malignancy useful in differentiating autoimmune pancreatitis from pancreatic cancer? A prospective outcome study. Gut. 2008;57(12):1704–12.
58. Sunderkötter C, Pappelbaum KI, Ehrchen J. Cutaneous symptoms of various vasculitides. Hautarzt. 2015;66(8):589–98.
59. Chen KR, Carlson JA. Clinical approach to cutaneous vasculitis. Am J Clin Dermatol. 2008;9(2):71–92.
60. Garg M, Mackay S, Hill PA, Katsoulis J, McLaughlin S, Desmond PV, Chen Ry. Leucocytoelastic and renal vasculitis in a patient with autoimmune pancreatitis: New associations. Intern Med J. 2010;40(5):376–80.
61. Garg A, Goldberg D. Relapsing leucocytoelastic vasculitis triggered by activity in a young woman. Arch Dermatol. 2009;145(5):601–2.