Patterns of Symmetry Expression in Angiosperms: Developmental and Evolutionary Lability

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Variation in flower symmetry is a remarkable aspect of flowering plant diversity. Multiple evolutionary transitions from an ancestral form of radial symmetry to bilateral symmetry and reversals from bilateral to radial symmetry have occurred during angiosperm evolution. The high incident of symmetry transitions poses the question concerning the homology of underlying developmental processes. I conducted a comprehensive study of developmental expression of symmetry across angiosperms covering 72 families and 39 orders. A wide range of timing for the expression of symmetry was found from organ initiation, to enlargement, to late differentiation of organs. In many studied genera, the symmetry pattern was transient during development. Mapping the floral symmetry patterns on the phylogenetic tree indicated the scattered distribution of patterns across angiosperms. Mid to late onset of actinomorphy/zygomorphy was found in all major clades. The expression of symmetry was also versatile within families. No strong relationship was found between the degree of zygomorphy at mature stage and its time of expression. The developmental constraints exerted by the inflorescence, bracts, neighbor flowers or neighbor organs within a flower could provide a possible explanation for the observed variation of symmetry expression.

Keywords: symmetry expression, angiosperms, developmental constraints, actinomorphy, zygomorphy

INTRODUCTION

One of the fascinating aspects of the floral organization is the balanced and harmonic arrangement of organs within a flower creating specific symmetry patterns. Actinomorphy (i.e., polysymmetry or radial symmetry with several symmetry planes) and zygomorphy (i.e., monosymmetry or bilateral symmetry with one symmetry plane) are the most common types of symmetry among flowering plants (Endress, 1999, 2010). Zygomorphy has diverse expressions in flowers, affecting organ categories in various combination (all organs, or only one organ category) and to various degrees (by differential shape, size, curvature, number) (Citerne et al., 2010; Endress, 2012). A weak degree of monosymmetry which is sometimes expressed due to the positional effect (positional monosymmetry, e.g., bending of stamens and styles due to gravity), may provide a precondition for the evolution of elaborate constitutional monosymmetry in polsymmetric groups (Endress, 1999). Except for Lamiales in which both corolla and androecium are remarkably affected, monosymmetry is commonly concentrated on or restricted to the perianth (Endress, 2012).
The reconstruction of the flower symmetry on the phylogenetic tree of angiosperms indicated actinomorphy as the ancestral state which is in agreement with its prevalence in flowers of basal angiosperms (Endress and Doyle, 2009). While relatively elaborate forms of zygomorphy are found in basal angiosperms (Aristolochiaceae of Piperales) and monocots (Orchidaceae of Asparagales, Commelinaceae of Commelinales), zygomorphy is mainly concentrated among rosids especially in Fabales and Malvales and among asterids especially in Lamiales, Asterales, and Dipsacales (Citerne et al., 2010; Endress, 2012). The analysis of symmetry evolution on the phylogenetic tree of angiosperms indicated frequent transitions from the ancestral actinomorphy to zygomorphy at least two times among the basal angiosperms, 29 times in monocots, and 99 times in the eudicots (Reyes et al., 2016). The high incident of zygomorphy and its homoplastic evolution poses the question concerning the homology of underlying developmental processes.

Floral symmetry is established through the activity of the floral meristem during organ initiation, enlargement, and subsequent differentiation. Several authors studied the manifestation of floral symmetry during ontogeny (Mair, 1977; Endress, 1999; Tucker, 1999). They reported a wide range of timing for the expression of symmetry, from organ initiation, to later enlargement of floral organs, to late differentiation of structures. In many studied genera, the symmetry pattern is transient during development. The symmetry established by the ordered initiation of floral organs at the beginning of development can be reduced, enhanced or changed later through differential growth of floral sectors. The developmental or evolutionary constraints underlying such labile expression of symmetry is poorly understood. It is suggested that the phylogenetic background of taxa and the degree of zygomorphy in mature flowers can influence the time of zygomorphy expression (Endress, 1999; Tucker, 1999). In many families with prevalent elaborate zygomorphy (e.g., Fabaceae), zygomorphy is manifested early at organs initiation and persists throughout development (Tucker, 2003). On the other hand, in zygomorphic taxa embedded in groups with predominantly actinomorphic flowers (e.g., Aconitum in Ranunculaceae), the flower is actinomorphic through most of its development and the manifestation of zygomorphy occurs toward the end of the ontogeny (Mair, 1977). However, the reverse condition also occurs. Some lamiaceous flowers with elaborate zygomorphy have shown a preceding actinomorphy in early development (Naghiloo et al., 2013a,b, 2014b). The weakly zygomorphic flowers of Capparis spinosa from the actinomorphic family Capparaceae express signs of zygomorphy at organ initiation (Naghiloo et al., 2015).

I conducted a comprehensive study of developmental expression of symmetry across angiosperms covering 72 families and 39 orders. Focusing on corolla symmetry as the most prominent manifestation of symmetry, I traced the evolution of developmental changes on a phylogenetic background. The objectives were (1) to determine patterns of symmetry expression within angiosperms, (2) to map the distribution of patterns on a phylogenetic tree, and (3) to infer evolutionary and developmental constraints underlying diversity of symmetry patterns.

### MATERIALS AND METHODS

#### Data Collection

The pattern of symmetry expression of 227 species belonging to 72 families and 39 orders of angiosperms was extracted from literature (for details of species and list of literature see Supplementary Appendix S1). The corolla symmetry in anthetic flowers was identified as zygomorphic (79 species) or actinomorphic (148 species) based on the available descriptions and images of the flower. Focusing on the corolla symmetry, I described tetramerous flowers of Brassicaceae and Oleaceae as actinomorphic while they are otherwise described as dissymmetric considering the whole flower.

I then used SEM micrographs from the literature to indicate the expression of symmetry in three developmental stages: (1) The early/initiation phase in which the first trace of organ primordia was detectable. (2) The mid/enlargement phase in which all primordia enlarged as clear bulges. (3) The late/differentiation phase during which the growth of primordia was accelerated and they started to cover the inner organs. The literatures in which the related stages could not be detected were excluded from the study.

The symmetry pattern in early phase was evaluated based on the order of primordia emergence; the simultaneous appearance was interpreted as actinomorphy, while the sequential appearance with a single dorsoventral axis was described as zygomorphy. The decision about organ initiation was based on clear separation from the floral meristem usually forming a furrow. In mid and late phases, the size difference was used as a criterion to indicate symmetry pattern; the size equality was described as actinomorphy while the size gradient along the dorsoventral axis was interpreted as zygomorphy.

#### Micrographs

Upon identifying the symmetry patterns, representative micrographs for each developmental pattern were extracted from the author’s previous publications and presented. For a description of the microscopic procedure, see the related publications (Dadpour et al., 2011a,b, 2012a; Naghiloo et al., 2012, 2013b, 2014a; Naghiloo and Classen-Bockhoff, 2016; Naghiloo and Claßen-Bockhoff, 2017).

#### Phylogenetic Framework and Character Optimization

I used the angiosperm phylogeny created by Soltis et al. (2011) to infer the relationships among the 72 families examined in the current study. This well-resolved tree was based on data from 17 DNA regions representing all three plant genomes, i.e., nucleus, plastid, and mitochondrion (Soltis et al., 2011). All branches were set equal with a value of 1.

To infer the evolutionary change in symmetry pattern, character optimization was carried out with the Maximum Parsimony method implemented in Mesquite 3.04 (Maddison and Maddison, 2015). All character changes were treated as an unordered and equally weighted.
The Maximum Likelihood method (ML) was not employed here as this method does not accept polymorphisms which were numerous in my dataset.

RESULTS

Patterns of Symmetry Expression

(A) Actinomorphic Species: The evaluation of symmetry expression in 148 actinomorphic species indicated the occurrence of four developmental patterns (Figures 1, 2):

1. Early expression: The actinomorphy first appeared during petal initiation (S1) due to the simultaneous emergence of primordia (Figures 1A,D). It was maintained during development due to synchronous enlargement (S2) and differentiation (S3) (Figures 1B,C,E,F). The flowers were, therefore, actinomorphic throughout development.

2. Mid expression: The corolla started development with a zygomorphic, non-simultaneous initiation (Figures 1G,J). However, the initial zygomorphy soon disappeared due to the synchronous enlargement and growth of corolla members after initiation (Figures 1H,I,K,L).

3. Late expression: The corolla development started with the zygomorphic, non-simultaneous initiation of primordia (Figures 2A,D). The zygomorphy was still evident during the enlargement of organ primordia (Figures 2B,E). The synchronization of petal growth only happened during organ differentiation (Figures 2C,F).

4. Very late expression: The corolla development started with a zygomorphic, non-simultaneous initiation (Figures 2G,J). The zygomorphy maintained during enlargement and differentiation phases due to asynchronous growth (Figures 2H,I,K,L). The flowers were, therefore, zygomorphic throughout development and actinomorphy only appeared due to equalization of organ growth in pre-anthetic flowers.

(B) Zygomorphic Species: I could distinguish three developmental patterns among 79 zygomorphic species:

1. Early expression: The zygmomorphy first appeared at organ initiation due to the sequential emergence of petal primordia (Figures 3A,D). The first initiated primordium showed the highest growth rate, followed by the second and third initiated primordia (Figures 3B,E). The size gradient was evident during enlargement and differentiation phases (Figures 3C,F). The flowers were therefore zygomorphic throughout development.

2. Late expression: The corolla development started with an actinomorphic, simultaneous initiation (Figures 3G,J). The petals developed synchronously during enlargement and early differentiation phases (Figures 3H,I,K,L). The zygmomorphy only appeared at the late differentiation phase prior to anthesis.

3. The double shift of symmetry: The corolla development started with a zygomorphic, non-simultaneous initiation of primordia (Figures 4A,D,G). However, the initial zygmomorphy was masked due to the equalization of primordia during enlargement phase (Figures 4B,E,H). After this period of actinomorphy, zygmomorphy appeared again during the differentiation phase due to asynchronous organ growth (Figures 4C,F,I).

Evolution of Symmetry Patterns

Tracing the evolution of symmetry indicated that the ancestral state for angiosperms has been an actinomorphic flower which manifested actinomorphy at mid to late stage of development (Figure 5 and Supplementary Appendixes S2–S4). All major angiosperm clades (rosids, asterids, and monocots) were also characterized by an ancestral actinomorphic flower with a preceding early zygmomorphy. The transition to early actinomorphy occurred several times in rosids (Fabaceae, Euphorbiaceae, Malvaceae-Neuradaceae clade, Cleomaceae, Rutaceae, Geraniaceae-Myrtaceae clade, and Vitaceae), asterids (Oleaceae, Rubiaceae, Solanaceae, Apiaceae, and Caprifoliaceae), eudicots (Caryophyllaceae), and basal angiosperms (Annonaceae). On the other hand, the transition to late actinomorphy also happened frequently by a shift to zygmomorphy at the enlargement or differentiation phase. Such a transition was found in several families among rosids (Elaeagnaceae, Passifloraceae, and Zygophyllaceae), asterids (Plantaginaceae, Rubiaceae, Apiaceae, Caprifoliaceae, and Hydrangeaceae), eudicots (Portulacaceae, Nyctaginaceae, Phytolaccaceae, Berberidaceae, Loranthaceae, Olacaceae, and Ranunculaceae) monocots (Heliconiaceae, Arecaceae) and basal angiosperms (Lauraceae, Nymphaeaceae).

The majority of zygmomorphic flowers were derived from the ancestral state by the transition to zygmomorphy during both enlargement and differentiation phases. Examples of such early zygmomorphic flowers were found among rosids (Fabaceae, Bataceae, Cleomaceae, and Sapindaceae), asterids (Orobanchaceae, Paulowniaceae, Gesneriaceae, Solanaceae, Caprifoliaceae, Lecythidaceae, and Balsaminaceae), eudicots (Dilleniaceae, Sabiaceae), monocots (Xyridaceae, Zingiberaceae). However, transition to mid-late zygmomorphy also happened several times among rosids (Fabaceae, Brassicaceae, and Cleomaceae), asterids (Orobanchaceae, Phrymaceae, Lamiaceae, Scrophulariaceae, Calceolariaceae, Asteraceae, and Caprifoliaceae), monocots (Commelinaceae, Orchidaceae), and basal angiosperms (Aristolochiaceae).

DISCUSSION

Developmental Flexibility and Variation of Symmetry Patterns

Tracing developmental expression of symmetry indicated that it does not follow a fixed pattern and that the symmetry expression at each stage could be independent of the previous or the following stages. The symmetry in mature flowers is often independent of the early expression of symmetry. Regardless
FIGURE 1 | Early to mid-stage establishment of actinomorphy in corolla. (A–F) Simultaneous initiation followed by synchronous enlargement and differentiation in Syringa vulgaris from Oleaceae (A–C) and Alcea rosea from Malvaceae (D–F). (G–I) Non-simultaneous initiation followed by synchronous enlargement and differentiation in Zygophyllum fabago (Zygophyllaceae). (J–L) Non-simultaneous initiation of petals within each whorl followed by synchronous enlargement and differentiation in Crocus sativus (Iridaceae). P: petal, S1: Initiation phase, S2: Enlargement phase, S3: Differentiation phase.
Late establishment of actinomorphy in corolla. (A–F) Non-simultaneous initiation followed by asynchronous enlargement and later synchronization of growth during differentiation in *Rubia tinctorium* from Rubiaceae (A–C) and *Bougainvillea spectabilis* from Nyctaginaceae (D–F). (G–L) Non-simultaneous initiation followed by asynchronous enlargement and differentiation in *Larrea divaricata* from Zygophyllaceae (G–I) and *Patrinia gibbosa* from Caprifoliaceae (J–L). P: petal, S1: initiation phase, S2: enlargement phase, S3: differentiation phase. Note the size difference between the largest and the smallest corolla members during asynchronous enlargement and differentiation (shown by stars).
FIGURE 3 | Early and late establishment of zygomorphy in corolla. (A–F) Non-simultaneous initiation followed by asynchronous enlargement and differentiation in *Salvia nemorosa* from Lamiales (A–C) and ray florets of *Osteospermum ecklonis* from Asteraceae (D–F). (G–L) Simultaneous initiation followed by synchronous enlargement and asynchronous differentiation in *Verbascum thapsus* from Scrophulariaceae (G–I) and *Succisa pratensis* from Caprifoliaceae (J–L). P: petal, S1: initiation phase, S2: enlargement phase, S3: differentiation phase. Note the size difference between the largest and the smallest corolla members during asynchronous enlargement and differentiation (shown by stars).
of being actinomorphic or zygomorphic, the majority of flowers were characterized by zygomorphic initiation which was supported as the ancestral state for the angiosperms. The results, therefore, suggest the existence of three independent steps for the establishment of symmetry in the course of ontogeny: organ initiation, enlargement, and differentiation. Hypothetically, if we consider both possible symmetry expressions (zygomorphy or actinomorphy) at each step, the combination of steps will yield eight developmental patterns (Figure 6). In agreement with this hypothesis, examples of all predicted symmetry patterns were found among angiosperms. It seems that flexibility in the developmental regulation of floral symmetry provides a broad scope for the evolution of symmetry pattern.

The first insights into the genetic mechanism underlying symmetry expression came from the study of model species *A. majus*. Two recently duplicated TCP family transcription factors, CYCLOIDEA (*CYC*) and DICHOTOMA (*DICH*), specify the dorsal floral identity (Luo et al., 1996, 1999). The expression of *CYC* and *DICH* is restricted to the dorsal region of the floral meristem, beginning from organ initiation and maintaining throughout enlargement and differentiation (Luo et al., 1996, 1999; Almeida et al., 1997). The function of both genes

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**FIGURE 4** | Double change of symmetry in zygomorphic corollas. (A–I) Non-simultaneous initiation followed by synchronous enlargement and asynchronous differentiation causing a double shift in symmetry in Astragalus compactus from Fabaceae (A–C), Rhynnocorys elephas from Orobanchaceae (D–F), and Zaiphora fenuior from Lamiaceae (G–I). P: petal, S1: initiation phase, S2: enlargement phase, S3: differentiation phase. Note the size difference between the largest and the smallest corolla members during asynchronous differentiation (shown by stars).
FIGURE 5 | The evolution of corolla symmetry reconstructed on the phylogenetic tree of angiosperms (extracted from Soltis et al., 2011). The ancestral character stats (obtained under Maximum Parsimony method implemented in Mesquite) are shown for the nodes of interest.
has been shown to be preserved in zygomorphic flowers across different clades of angiosperms (Hileman, 2014). The expression of CYC homologues has been studied in relation to the evolutionary transition from zygomorphic to actinomorphic symmetry. In Plantago, one CYC-like paralog is expressed across the dorsoventral flower axis, while the other paralog has been lost (Reardon et al., 2009; Preston et al., 2011). The combination of expanded CYC expression and loss of CYC expression, therefore,
mediates the transition to a derived actinomorphy. In Cadia and Tengia, a similar expansion of one CYC paralog across the dorsoventral axis in parallel with the downregulation and loss of expression of the other paralog is suggested to be responsible for the evolution of actinomorphic flowers from zygomorphic ancestors (Citerne et al., 2006; Pang et al., 2010). In contrast, the reversal to radial symmetry in *Psychopterys* and *Sphedannocarpus* involves the entire loss of both CYC-like paralogs (Zhang et al., 2013). An specific pattern of CYC-like paralogs is reported in derived actinomorphic flowers of *Bournea* (Zhou et al., 2008). Despite expanded expression of one CYC-like paralog, the other paralog is neither lost nor downregulated, but retains dorsal-specific expression at early stage (Zhou et al., 2008). Such transient dorsal-specific expression can explain the initial zygomorphic pattern in *Bournea*. Considering all mentioned examples, I would suggest that changes in the timing, duration, and localization of CYC-like gene expression are implicated in the developmental and evolutionary transition of floral symmetry.

**Evolution of Symmetry Patterns and Developmental Constraints**

Mapping the floral symmetry patterns on the phylogenetic tree indicated the scattered distribution of patterns across angiosperms. Mid-late onset of actinomorphy were found in all major clades including basal angiosperms (Lauraceae), monocots (Arecales and Heliconiaceae), eudicots (Ranunculaceae, Olacaceae, Loranthaceae, Berberidaceae, Phytolaccaceae, Nyctaginaceae, and Portulacaceae), asterids (Hydrangeaceae, Lecythidaceae, Caprifoliaceae, Apiaceae, Rubiaceae, and Plantaginaceae), and rosids (Zygophyllaceae, Passifloraceae, and Elaeagnaceae). The same was true for the mid-late onset of zygomorphy with representative examples in eudicots (Ranunculaceae), asterids (Caprifoliaceae, Dipsacaceae, Asteraceae, Calceolariaceae, and Scrophulariaceae), and rosids (Cleomaceae). The expression of symmetry was also versatile within families. The onset of actinomorphy in families such as Ranunculaceae, Oleaceae, Apiaceae, and Zygophyllaceae ranged from early to mid or late development depending on the taxa (see Supplementary Appendix S1).

A similar variation was found within zygomorphic families. In Lamiaceae, some species like *Marrubium parviflorum*, *Stachys byzantine*, and *Salvia nemorosa* were zygomorphic from petal inception till anthesis, while in *Phlomis arméniana* a double change in symmetry pattern was found (i.e., from early zygomorphy at initiation time to actinomorphy at enlargement and back to zygomorphy at late development) (Naghiloo et al., 2013b, 2014b). Fabaceae represented another zygomorphic family in which both early expression (*Amherstia nobilis*, *Duparquetia orchidacea*) as well as the double change in symmetry pattern (*Astragalus compactus*, *Wisteria sinensis*) were found (Tucker, 2000; Prenner and Klitgaard, 2008; Naghiloo and Dadpour, 2010; Naghiloo et al., 2012). In zygomorphic flowers of Caprifoliaceae, examples of early zygomorphy (*Dipsacus folliculum*, *Knautia arvensis*, and *Morina persica*), late zygomorphy (*Succisa pratensis*, *Lonicera periclymenum*) and shift in symmetry pattern (e.g., *Lomelosia palaestina*, *Succisella inflexa*, *Cephalaria transsylvenica*) were found (Naghiloo and Claßen-Bockhoff, 2017). The time of zygomorphy expression in ray flowers of Asteraceae depended on the taxa, ranging from early (e.g., *Osteospermum ecklonis*, *Xeranthemum squarrosum*) to mid or late stages of development (*Senecio glabellus*, *Mutisia coccinea*) (Harris, 1995; Dadpour et al., 2011a, 2012b).

In zygomorphic species of the otherwise radially symmetric family Ranunculaceae, the zygomorphy either expressed at an early stage through precocious initiation of some petal members (e.g., *Delphinium*, Naghiloo Unpublished Data) or at a late stage of development through the formation of a petal spur (*Aconitum*, Mair, 1977; Jabbour et al., 2009). Other examples of zygomorphy in families with predominantly actinomorphic flowers are *Schizanthus* and *Schwenckia* from Solanaceae, and *Iberis amara* from Brassicaceae. In the two first species, zygomorphy is apparent from the onset of organ initiation (Ampornan and Armstrong, 1988, 1989), while in *Iberis amara* zygomorphy establish through heterogeneous growth of petals at the late stage of development (Busch and Zachgo, 2007).

Given the parallel occurrence of symmetry patterns (early, mid or late expression) in different angiosperm clades and their variation among closely related species, the phylogenetic signal (although not tested here) seems unlikely to be the main determinant of the symmetry pattern. I would rather suggest that developmental constraints exerted by the entire inflorescence, bracts, neighbor flowers or neighbor organs within a flower could provide a possible explanation for the observed variation of symmetry expression. As an example, I refer to the correlation found between cymose monochasial inflorescences and late expression of actinomorphy in some taxa (*Centranthus*, *Valerianaceae*, *Qualea*, and *Vochysiaceae*) (Eichler, 1878). In *Patrinia*, which expresses actinomorphy at a very late stage of development, the flowers are arranged in dichasias with monochasial peripheral braches. In early floral development, the two petals and stamens on the side toward the weaker subsequent ramification of the dichasium appear earlier than their counterparts (Hofmann and Gottmann, 1990). Beside the ramification pattern of inflorescence, the mechanical forces generated by subtending leaves or bracts can also influence the order of organ initiation (Naghiloo et al., 2012). An association between order of initiation and the presence of bracteoles has been suggested for some members of Papilionoideae (Prenner, 2004).

Another example of developmental constraints which can influence the pattern of symmetry expression is found in species with a double change of floral symmetry (i.e., from zygomorphy to actinomorphy and back to zygomorphy). The size gradient in mature zygomorphic flowers often follows the initiation sequence; organs that are initiated first are usually the largest at maturity and the pattern of early symmetry is maintained throughout development. However, in some taxa (e.g., *Papilionoideae*, *Lamiaceae*, *Orobanchaceae*, *Dipsacaceae*) the size gradient in mature petals is opposite to the direction of their initiation; the last initiated petal is the largest one at maturity. In such cases, a period of synchronization is required...
before the new size gradient establishes through differential growth. The mismatch between the direction of zygomorphy in early and late stages could, therefore, explain double changes of symmetry during development.

DATA AVAILABILITY STATEMENT

The datasets generated for this study is available as Supplementary Material online.

REFERENCES

Almeida, J., Rocheta, M., and Galembo, L. (1997). Genetic control of flower shape in Antirrhinum majus. Development 124, 1387–1392.

Ampornpoom, L., and Armstrong, J. (1988). The floral ontogeny of Schizanthus, a zygomorphic member of the Solanaceae. Am. J. Bot. 75:54.

Ampornpoom, L., and Armstrong, J. (1989). The floral ontogeny of Salpiglossis, a zygomorphic member of the Solanaceae. Am. J. Bot. 76:64.

Busch, A., and Zachgo, S. (2007). Control of corolla monosymmetry in the Brassicaceae Iberis amara. Proc. Natl. Acad. Sci. U.S.A. 104, 16714–16719. doi: 10.1073/pnas.0705338104

Citerne, H., Jabou, F., Natod, S., and Damerval, C. (2010). “The evolution of floral symmetry,” in Advances in Botanical Research, eds J.-C. Kader and M. Delseny (Cambridge, MA: Academic Press), 85–137. doi: 10.1016/s0065-2296(10)53403-5

Citerne, H., Toby Pennington, R., and Cronk, Q. (2006). An apparent reversal in growth. The mismatch between the direction of zygomorphy in before the new size gradient establishes through differential growth.

Jubb, F., Ronse De Craene, L. P., Nadot, S., and Damerval, C. (2009). Establishment of zygomorphy on an ontogenetic spiral and evolution of perianth in the tribe Delphinieae (Ranunculaceae). Am. J. Bot. 104, 809–822. doi: 10.1093/ajb/mcp162

AUTHOR CONTRIBUTIONS

SN performed all aspects of this research.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fevo.2020.00104/full#supplementary-material

Luo, D., Carpenter, R., Copsey, L., Vincent, C., Clark, J., and Coen, E. (1999). Control of organ asymmetry in flowers of Antirrhinum. Cell 99, 367–376. doi: 10.1016/s0092-8674(00)81523-8

Luo, D., Carpenter, R., Vincent, C., Copsey, L., and Coen, E. (1996). Origin of floral asymmetry in Antirrhinum. Nature 383, 794–799. doi: 10.1038/383794a0

Maddison, W. P., and Maddison, D. R. (2015). Mesquite: A Modular System for Evolutionary Analysis. Available online at: Version 3.4. http://mesquiteproject.org (accessed July, 2019).

Mair, O. (1977). Zur Entwicklungsgeschichte monosymmetrischer Dicotyleton-Blüten. Diss. Bot. 58, 260–274.

Naghiloo, S., and Classen-Bockhoff, R. (2016). Developmental analysis of merosity and sexual morphs in Rubiaceae: A case study in Rubia and Crucca. Flora 222, 52–59. doi: 10.1016/j.flora.2016.03.010

Naghiloo, S., and Claßen-Bockhoff, R. (2017). Developmental changes in time and space promote evolutionary diversification of flowers: A case study in Dipsacaleae. Front. Plant Sci. 8:1665–1665. doi: 10.3389/fpls.2017.01665

Naghiloo, S., and Dadpour, M. (2010). Floral ontogeny in Wisteria sinensis (Fabaceae: Fabioideae: Millettieae) and its systematic implications. Aust. Syst. Bot. 23, 393–400. doi: 10.1071/SB10027

Naghiloo, S., Dadpour, M. R., and Movafeghi, A. (2012). Floral ontogeny in Astragalus compactus (Leguminosae: Papilionoideae: Galegaeae): variable occurrence of bracteoles and variable patterns of sepall initiation. Planta 235, 793–805. doi: 10.1007/s00014-011-1388-1

Naghiloo, S., Esmailloo, Z., and Dadpour, M. R. (2014a). Comparative floral ontogeny of single-flowered and double-flowered phenotypes of Alcea rosea: Flora 27, 257–263. doi: 10.1016/j. flora.2012.02.001

Naghiloo, S., Dadpour, M. R., and Neycharan, S. F. (2012b). The development of pistillate and perfect flowers in Xenarthrum squarrosum (Asteraceae). Plant Biol. 14, 234–243. doi: 10.1111/j.1438-8671.2011.00469.x

Eichler, A. W. (1878). Blüthenformen. II. Leipzig: Engelmann.

Endress, P. (2010). Evolution of floral symmetry. Curr. Opin. Plant Biol. 4, 86–91. doi: 10.1016/j.scienco.2010.12.004

Endress, P. K. (1999). Symmetry in flowers: diversity and evolution. Int. J. Plant Sci. 160, S3–S23.

Endress, P. K. (2012). The immense diversity of floral monosymmetry and asymmetry across angiosperms. Bot. Rev. 78, 345–397. doi: 10.1007/s12229-012-9106-3

Endress, P. K., and Doyle, J. A. (2009). Reconstructing the ancestral angiosperm flower and its initial specializations. Am. J. Bot. 96, 22–66. doi: 10.3732/ajb.0800047

Harris, E. M. (1995). Inflorescence and floral ontogeny in Asteraceae: A synthesis of historical and current concepts. Bot. Rev. 61:93. doi: 10.1086/50887192

Hileman, L. C. (2014). Trends in flower symmetry evolution revealed through phylogenetic and developmental genetic advances. Philos. Trans. R. Soc. Lond. B Biol. Sci. 369:20130348. doi: 10.1098/rstb.2013.0348

Hoffmann, U., and Gottmann, J. (1990). Morina l. und Triplostegia Wall. ex DC. im Vergleich mit Valerianacia und Dipsacaceae. Bot. Jahrh. Syst. 111, 499–553.

Jabbour, F., Ronse De Craene, L. P., Natod, S., and Damerval, C. (2009). Establishment of zygomorphy on an ontogenetic spiral and evolution of perianth in the tribe Delphinieae (Ranunculaceae). Am. J. Bot. 104, 809–822. doi: 10.1093/ajb/mcp162
Preston, J. C., Martinez, C. C., and Hileman, L. C. (2011). Gradual disintegration of the floral symmetry gene network is implicated in the evolution of a wind-pollination syndrome. Proc. Natl. Acad. Sci. U.S.A. 108, 2343–2348. doi: 10.1073/pnas.1011361108

Reardon, W., Fitzpatrick, D. A., Fares, M. A., and Nugent, J. M. (2009). Evolution of flower shape in Plantago lanceolata. Plant Mol. Biol. 71, 241–250. doi: 10.1007/s11103-009-9520-z

Reyes, E., Sauquet, H., and Nadot, S. (2016). Perianth symmetry changed at least 199 times in angiosperm evolution. Taxon 65, 945–964. doi: 10.12705/655.1

Soltis, D. E., Smith, S. A., Cellinese, N., Wurdack, K. J., Tank, D. C., Brockington, S. F., et al. (2011). Angiosperm phylogeny: 17 genes, 640 taxa. Am. J. Bot. 98, 704–730. doi: 10.3732/ajb.1000404

Tucker, S. C. (1999). Evolutionary lability of symmetry in early floral development. Int. J. Plant Sci. 160, S25–S39.

Tucker, S. C. (2000). Floral development in Tribe Detarieae (Leguminosae: Caesalpinioideae): Amherstia, Brownea, and Tamarindus. Am. J. Bot. 87, 1385–1407. doi: 10.2307/2656867

Tucker, S. C. (2003). Floral development in legumes. Plant Physiol. 131, 911–926. doi: 10.1104/pp.102.017459

Zhang, W., Steinmann, V. W., Nikolov, L., Kramer, E. M., and Davis, C. C. (2013). Divergent genetic mechanisms underlie reversals to radial floral symmetry from diverse zygomorphic flowered ancestors. Front. Plant. Sci. 4:302. doi: 10.3389/fpls.2013.00302

Zhou, X. R., Wang, Y. Z., Smith, J. F., and Chen, R. (2008). Altered expression patterns of TCP and MYB genes relating to the floral developmental transition from initial zygomorphy to actinomorphy in Bournea (Gesneriaceae). New Phytol. 178, 532–543. doi: 10.1111/j.1469-8137.2008.02384.x

Conflict of Interest: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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