Increased Gastric IL-1β Concentration and Iron Deficiency Parameters in *H. pylori* Infected Children

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Abstract

Association between *H. pylori* infection, iron deficiency and iron deficiency anaemia has been described, but the mechanisms involved have not been established. We hypothesized that in *H. pylori* infected children increased gastric concentrations of IL-1β and/or TNF-α, both potent inhibitors of gastric acid secretion that is essential for iron absorption, are predictors for low blood concentrations of ferritin and haemoglobin, markers of early depletion of iron stores and anaemia, respectively. We evaluated 125 children undergoing endoscopy to clarify the origin of gastrointestinal symptoms. Gastric specimens were obtained for *H. pylori* status and cytokine evaluation and blood samples for determination of iron deficiency/iron deficiency anaemia parameters and IL1 cluster and TNFA polymorphisms that are associated with increased cytokine secretions. Higher IL-1β and TNF-α gastric concentrations were observed in *H. pylori*-positive (*n* = 47) than in negative (*n* = 78) children. Multiple linear regression models revealed gastric IL-1β, but not TNF-α, as a significant predictor of low ferritin and haemoglobin concentrations; results were reproduced in young children in whom *IL1RN* polymorphic genotypes associated with higher gastric IL-1β expression and lower blood ferritin and haemoglobin concentrations. In conclusion, high gastric levels of IL-1β can be the link between *H. pylori* infection and iron deficiency/iron deficiency anaemia in childhood.

Introduction

Anaemia is a major public health problem in developing countries and approximately half of all cases are due to iron deficiency (ID) [1,2]. Iron deficiency anaemia (IDA) is the final stage in the spectrum of a persistent negative iron balance, being preceded by an iron-restricted erythropoiesis characterized by low iron stores. The greater demand for iron due to growth, expansion of red cell mass and menstrual blood loss in female adolescents favors the development of ID/IDA in childhood/adolescence [2]. Factors which contribute to the high frequency of ID/IDA in developing countries include poor iron intake, low dietary iron bioavailability and blood loss due to gastrointestinal parasitic infections [2,3].

*Helicobacter pylori* colonizes the stomach of more than a half of the world’s population. Concomitant high prevalence of *H. pylori* infection and ID/IDA in some areas, particularly in developing countries, has led to the hypothesis that *H. pylori* may contribute to ID/IDA. Potential mechanisms proposed include increased blood loss due to *H. pylori*-induced gastric lesions [4], iron uptake by *H. pylori* [5], deficient iron absorption due to decreased gastric acidity [6,7], that is essential for the reduction and solubilization of non-heme iron, and reduced gastric juice ascorbic acid concentrations [6].

The acute phase of *H. pylori* infection is accompanied by transient hypochlorhydria of variable duration [8–10]. Similar perturbations in gastric acid secretion occur in animals following infection with gastric *Helicobacter* species [11] or *H. pylori* [12]. In the latter study, reversal of the hypochlorhydria induced by *H. pylori* infection in gerbils by treatment with recombinant IL-1 receptor antagonist implicated the IL-1β gene cluster in hypochlorhydric response to *H. pylori*. As *H. pylori* infection is mainly acquired in childhood [13], it is biologically plausible that infected children are at increased risk of developing ID/IDA as a consequence of hypochlorhydria.

Concentrations of IL-1β and TNF-α, potent inhibitors of gastric acid secretion [14], are increased in the gastric mucosa of *H. pylori* infected adults and children [15–17]. As both cytokines are capable of directly inhibiting gastric acid secretion by parietal cells, they might be one of the links between ID and *H. pylori* infection in...
childhood [13]. Furthermore, *H. pylori* infected adults with functional polymorphisms in the IL1 gene cluster associated with over expression of IL-1β have increased hypochlohydria [17] that could, theoretically, interfere with intestinal iron absorption leading to ID.

We hypothesized that in *H. pylori* infected children without the known common causes of ID, increased gastric concentrations of IL-1β and/or TNF-α could be predictors for low blood concentrations of ferritin and haemoglobin, markers of early depletion of iron stores and anaemia, respectively.

**Patients and Methods**

This study was approved by the Ethics Committee of the Universidade Federal de Minas Gerais, Belo Horizonte, Brazil and the National Ethics Committee on Research from the Health Ministry of Brazil. Signed informed consent to participate was obtained from the children (whenever possible) and adolescents and their parents.

**Patients**

Between June 2007 and July 2010 125 children and adolescents (74 girls and 51 boys, mean age 11.1 ± 2.9 years, range 4–16 years) undergoing gastrointestinal endoscopy to clarify the origin of symptoms related to the upper gastrointestinal tract were prospectively studied. All the patients were from Minas Gerais state, localized in the Southeast Brazil. To avoid confounding factors which can modify the gastritis classification and the diagnosis of *H. pylori* infection, or which independently can modify iron stores, rigorous exclusion criteria were used. Exclusions included children who had received antimicrobial drugs, anti-cholinergic and steroid and non-steroidal anti-inflammatory agents for at least 30 days, or proton pump inhibitors for at least 15 days before endoscopy; children with peptic ulcer disease, coeliac disease and intestinal parasitic infections; children with gastrooesophageal varices, coagulation disorders, acquired or congenital immunosuppression, inflammatory diseases, renal failure, hematological disorders and neoplasias. Following endoscopy, additional exclusion criteria were previously undiagnosed coeliac disease or any histological non-specific duodenal inflammation in the absence of duodenal gastric metaplasia. From each evaluated patient (PCR) for *H. pylori* status, histological scores were graded as absent (0), mild (1), moderate (2), or marked (3).

**Blood Haemoglobin, Haematocrit and Ferritin Values**

Blood haemoglobin concentrations and haematocrit values were determined by using an automated electronic counter, Sysmex XT 1800i (Sysmex Corporation, Kobe, Japan). The serum ferritin concentration was determined by a chemiluminescence method employing the ADVIA Centaur® Immunoassay CP System (Siemens Healthcare, Erlangen, Germany).

**Statistical Analysis**

Data were analysed with SPSS statistical software package version 17.0 (SPSS Inc., Chicago, IL). Hardy-Weinberg equilibrium of alleles at individual loci was tested by χ²-test with Yate’s correction or Fisher’s exact test. The Kolmogorov-Smirnov goodness-of-fit was used to assess the normality of the data. When significant departures from normality were detected, the data were log-transformed. The histopathological scores in binary variables were transformed by combining absent/mild and moderate/severe scores to compare histopathological data with IL-1β and TNF-α gastric concentrations. The degree of gastric chronic (mononuclear) and active (polymorphonuclear) in *H. pylori*-positive and -negative children was compared by the two-tailed Mann Whitney U test. Correlations were evaluated by the Pearson’s correlation (continuous normally distributed data) or Spearman’s correlation (scores). The comparisons between antral and corpus cytokine concentrations were undertaken by two-tailed paired Student’s t test or Wilcoxon test. The level of significance was set at p≤0.05. Multiple linear regression analyses (“enter option”) were used in order to quantify the simultaneous and mutually independent contribution, of selected relevant predictor candidates, e.g. IL-1β and TNF-α gastric concentrations, for low ferritin
and haemoglobin blood concentration (dependent variables) while controlling for confounders such as gender and age. Variables with p values ≤0.20 in the univariate analyses were selected for the multivariate analyses. The optimum sample size, based on a significant level of 0.05 and a statistical power of 0.80 (type II error 0.02) is 125 cases, even when only a small effect size (f = 0.10) is expected.

**Results**

Among the 125 children, 47 (37.6%) were *H. pylori*-positive (mean age 11.7±2.7 years; 29 girls) and 78 (62.4%) were *H. pylori*-negative (mean age 10.7±2.9 years, 45 girls).

**Gastric Cytokine Concentrations**

A 17.2-fold increase in corpus and antral gastric concentration of IL-1β as well as a 7.0-fold increase in corpus and 57.7-fold increase in antral gastric TNF-α concentrations were observed in infected children when compared with non-infected children (p<0.001). Comparisons between *H. pylori*-negative and -positive children are shown in Figure 1A and 1B. The concentrations of IL-1β were higher in the corpus than in the antral gastric mucosa (2.5-fold increase) in both *H. pylori*-positive (p<0.001) and *H. pylori*-negative (p = 0.02) children. Conversely, the TNF-α concentration was higher in the antral than in the corpus gastric mucosa in *H. pylori*-positive children (2.5-fold increase, p<0.001) [Figure 1A and 1B].

**Blood IDA Parameters and Gastric Cytokine Concentrations**

The multiple regression analyses revealed the gastric concentration of IL-1β, but not TNF-α, as a significant independent predictor for low serum ferritin and haemoglobin concentrations (Table 1). In addition, the male gender was an independent predictor of ferritin increase and the age was an independent predictor of haemoglobin increase. As *H. pylori* infection is mainly acquired by young children and we have previously observed differences in the inflammatory and immune response between children younger and older than 10–12 years [23], we stratified children by age: 12 years old or younger, and those older than 12 years of age. IL-1β remained a predictor of low ferritin concentration and was a stronger predictor of low haemoglobin concentration in the younger age group. The male gender was an independent predictor of ferritin increasing (Table 1).

When *H. pylori*-positive and -negative children were separately analyzed, significant correlations were observed only in the *H. pylori*-positive group [ferritin (r = −0.42, p = 0.004) and haemoglobin (r = −0.35, p = 0.02)] (Figure 2). The values of haematocrit were also negatively correlated with gastric IL-1β concentrations (r = −0.40, p = 0.007). In contrast, corpus and antral TNF-α concentrations neither correlated with serum ferritin (p≥0.48), nor with haemoglobin (p≥0.74) or haematocrit (p≥0.40) values.

**Gastric Inflammation and Cytokine Concentrations**

The degrees of antral and corpus chronic and active inflammation were significantly higher (p<0.001 for all) in *H. pylori*-positive than in -negative children (Table 2). In *H. pylori*-positive children, the corpus concentration of IL-1β was significantly higher in children with a mild/moderate degree of corpus chronic inflammation than in those without corpus inflammation (672.48±300.37 pg/mg vs. 504.39±28.81 pg/mg, respectively, p = 0.002). Furthermore, the corpus concentration of TNF-α was higher (p<0.001) in infected children with a mild/moderate degree of corpus active inflammation than in those without corpus PMN cell infiltration (369.2±75.3 pg/mg vs. 277.8±75.3 pg/mg).

**IL1B, IL1RN and TNFA Polymorphisms**

*IL1B* (p = 0.45), *IL1RN* (p = 0.36) and *TNFA* (p = 0.78) polymorphic genotypes did not associate with *H. pylori* status (Table 3). No association between *IL1B* and *TNFA* polymorphic alleles and blood ID/IDA parameters (p≥0.17) was observed.

In the *H. pylori*-positive group, carriers of the polymorphic *IL1RN* allele 2 had increased corpus inflammatory scores (p = 0.02) and significantly higher (p = 0.01 for both) IL-1β concentrations than the non-carriers (antrum 354.4±189.3 pg/mg vs. 253.5±74.3 pg/mg and corpus 873.4±466.6 pg/mg vs.

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**Figure 1. Box plots representing the comparison of gastric IL-1β (A) and TNF-α (B) concentrations (pg/mg of protein) between *H. pylori*-positive (HP+, n = 47) and -negative (HP-, n = 78) children, and between antral and corpus concentration in *H. pylori*-positive and -negative groups.** The upper and lower limits of the boxes represent the 75th and 25th percentiles, respectively. The horizontal bar across the box indicates the median and the capped bars indicate the minimum and maximum data values. Statistical analysis by Student's t test after log transformation in the case of IL-1β; *p<0.001 and **p = 0.02.

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In the *H. pylori*-positive group of children of 12 years of age or younger (16 non-carriers vs. 9 carriers), a significant negative association between *IL1RN* polymorphic genotype and haemoglobin (13.0 ± 0.9 g/dL vs. 11.9 ± 1.3 g/dL, p = 0.03) or haematocrit (39.5 ± 2.2% vs. 36.0 ± 4.2%, p = 0.01) was observed.

The mean gastric concentration of antral (p = 0.12) and corpus *IL-1β* (p = 0.15) and antral (p = 0.32) and corpus *TNF-α* (p = 0.28) did not differ between carriers and non-carriers of *IL1B* and *TNFA* polymorphic genotypes, independently of the *H. pylori*-status.

### Discussion

ID is a very common micronutrient deficiency that affects individuals globally, but is a particular problem in developing countries. Children are considered to be at high-risk of ID that associates with deficits of immune, cognitive and motor functions [24, 25], in addition to the possible development of anaemia.

Accumulating evidence from epidemiological studies and clinical trials implicates *H. pylori* infection in the aetiology of ID/IDA [22, 26, 27]. Notably, ID in *H. pylori* infected subjects is resistant to iron supplementation, but this resistance is reversible by *H. pylori* eradication [28, 29]. However, the mechanism, or mechanisms, by which the infection might cause ID/IDA are still not determined. In the present study, we demonstrate that in *H. pylori*-infected children without common known causes of ID/IDA increased gastric *IL-1β* concentration is an independent predictor for low blood concentration of ferritin and haemoglobin.

Although it would be expected that in *H. pylori* infected children gastric *IL-1β* concentrations would be higher in the antrum, which is more inflamed than the corpus, a 2.5-fold increase in corpus *IL-1β* was observed. This could be explained either by increased corpus *IL-1β* production, or by a higher number of *IL-1β* receptors in the gastric corpus. The mean gastric concentration of antral (p > 0.12) and corpus *IL-1β* (p > 0.15) and antral (p > 0.32) and corpus *TNF-α* (p > 0.28) did not differ between carriers and non-carriers of *IL1B* and *TNFA* polymorphic genotypes, independently of the *H. pylori*-status.

Table 1. Multiple linear regression model including ferritin or haemoglobin as dependent variables and *IL-1β* and *TNF-α* gastric corpus concentrations, gender and age as independent variables.

|                      | Univariate analysis | Multivariate analysis |
|----------------------|---------------------|-----------------------|
|                      | Beta coefficient    | P value               |
|                      | Beta coefficient    | P value               |
| FERRITIN             |                     |                       |
| Children of all ages (n = 125) |                     |                       |
| age                  | 0.119               | 0.19                  |
| male gender          | 0.280               | 0.002                 |
| *IL-1β*              | −0.200              | 0.006                 |
| *TNF-α*              | −0.075              | 0.41                  |
| Children ≤12 years of age (n = 84) |                     |                       |
| age                  | −0.040              | 0.32                  |
| male gender          | 0.208               | 0.20                  |
| *IL-1β*              | −0.202              | 0.03                  |
| *TNF-α*              | −0.106              | 0.35                  |
| HAEMOGLOBIN          |                     |                       |
| Children of all ages (n = 125) |                     |                       |
| age                  | 0.285               | 0.001                 |
| male gender          | 0.151               | 0.09                  |
| *IL-1β*              | −0.220              | 0.02                  |
| *TNF-α*              | −0.135              | 0.14                  |
| Children ≤12 years of age (n = 84) |                     |                       |
| age                  | 0.145               | 0.19                  |
| male gender          | −0.170              | 0.12                  |
| *IL-1β*              | −0.399              | <0.001                |
| *TNF-α*              | −0.167              | 0.13                  |

### Table 1. Multivariate regression model including ferritin or haemoglobin as dependent variables and *IL-1β* and *TNF-α* gastric corpus concentrations, gender and age as independent variables.

Figure 2. Correlations between corpus *IL-1β* concentrations and concentrations of ferritin and haemoglobin in *H. pylori*-positive children (n = 47). Statistical analysis by Pearson’s correlation after log transformation in the case of *IL-1β* and ferritin.

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Increased gastric *IL-1β* concentration is an independent predictor for low blood concentration of ferritin and haemoglobin.
The negative correlation between gastric IL-1β and ID/IDA blood parameters observed in this study could be due to the powerful capability of IL-1β in inhibiting gastric acid secretion [14]. Gastric acid is essential for iron absorption by reducing the ferric iron to a more soluble and absorbable ferrous iron form [14,15]. The results of this study would support the argument that in the early phase of H. pylori infection, high gastric secretion of IL-1β that inhibits acid secretion impairs the absorption of iron. IL-1β would also participate in the impairment of iron absorption by up-regulating hepcidin as demonstrated in vivo [31,32]. However, in a recent study, Schwarz et al. [33] did not observe associations between the serum concentrations of hepcidin and H. pylori infection.

In chronic H. pylori infection in adulthood, gastric atrophy leading to hypochlorhydria is considered a mechanism of iron deficiency. Gastric corpus atrophic changes are more frequently observed in adult carriers of the IL1β gene cluster polymorphisms [17]. In the present study, IL1RN, but not IL1B polymorphism, was associated with increased gastric IL-1β concentration in concordance with studies in our adult population [34]. The latter study demonstrated that polymorphism of IL1RN but not IL1B is associated with increased risk of atrophic gastric changes and gastric carcinoma in this region of Brazil. In the current study we demonstrate that in the group of the H. pylori-positive youngest children, the haemoglobin and haematocrit values are lower in carriers of IL1RN polymorphic alleles than in children with the wild genotype. The high production of IL-1β in the former group might account for a more severe hypochlorhydria in the acute phase of H. pylori infection that is mainly acquired in early childhood.

In conclusion, we provide additional and convincing evidence of a role of gastric IL-1β induced by H. pylori infection in decreasing iron absorption in children. Thus, H. pylori infected children with ID/IDA may benefit from H. pylori eradication therapy.

**Author Contributions**

Conceived and designed the experiments: DMMQ JEC. Performed the experiments: AMCR GAR FFM KNT SDC PFSB LPFC. Analyzed the data: DMMQ JEC AMCR. Contributed reagents/materials/analysis tools: DMMQ JEC AMCR GAR FFM KNT SDC PFSB LPFC. Wrote the paper: DMMQ JEC AMCR.

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**Table 2.** Histological comparison of antral and corpus gastric mucosa of H. pylori (HP)-positive (n = 47) and -negative (n = 78) children*.

| Inflammation | Absent n (%) | Mild n (%) | Moderate n (%) | Marked n (%) | P value |
|--------------|--------------|------------|----------------|-------------|---------|
| **Antrum**   |              |            |                |             |         |
| Chronic inflammation | HP-positive 01 (2.2) 09 (19.5) 35 (76.1) 01 (2.2) | HP-negative 34 (47.9) 37 (52.1) 00 0 0 <0.001 |
| Active inflammation | HP-positive 04 (8.7) 26 (56.5) 16 (34.8) 0 0 <0.001 |
| HP-negative 66 (93.0) 5 (7.0) 00 0 <0.001 |
| **Corpus**   |              |            |                |             |         |
| Chronic inflammation | HP-positive 03 (6.8) 38 (86.4) 03 (6.8) 0 0 0 | HP-negative 42 (54.5) 35 (45.5) 00 0 <0.001 |
| Active inflammation | HP-positive 15 (34.1) 29 (65.9) 0 0 0 0 | HP-negative 74 (96.1) 03 (3.9) 0 0 <0.001 |

*One antral and 3 corpus gastric biopsy specimens from HP-positive and 7 antral and 1 corpus biopsy specimens from HP-negative children were deemed to be inadequate for histology assessment; n, number. Neither atrophy, nor intestinal metaplasia was observed.

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**Table 3.** IL1B-31, IL1RN and TNFA-307 genotypic frequencies in H. pylori-positive (n = 47) and -negative children (n = 78).

| All children n (%) | H. pylori-positive H. pylori-negative n (%) |
|--------------------|-------------------------------|
| IL1B-31*           |                               |
| T/T                | 37 (29.8)                     | 16 (34.8) 21 (26.9) |
| T/C                | 63 (50.8)                     | 20 (43.5) 42 (55.2) |
| C/C                | 24 (19.4)                     | 10 (21.7) 14 (17.9) |
| IL1RN VNTR*        |                               |
| 1/1                | 82 (66.2)                     | 30 (65.2) 52 (66.7) |
| 1/2                | 35 (28.2)                     | 15 (32.6) 20 (25.6) |
| 2/2                | 07 (5.6)                      | 01 (2.2) 06 (7.7)  |
| TNFA-307           |                               |
| G/G                | 87 (69.6)                     | 32 (68.1) 55 (70.5) |
| G/A                | 38 (30.4)                     | 15 (31.9) 23 (29.5) |

*It was not possible to genotype 1 H. pylori-positive children for IL1B-31 and 1 for IL1RN.

†It indicates all the long alleles and 2 the short allele. The loci did not deviate significantly from the expected Hardy-Weinberg distribution (P = 0.90 for IL1B-31, P = 0.26 for IL1RN and P = 0.08 for TNFA-307) and all segregated independently.

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