Resveratrol activates endogenous cardiac stem cells and improves myocardial regeneration following acute myocardial infarction

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Abstract. Stem cell antigen-1-positive (Sca-1⁺) cardiac stem cells (CSCs) therapy for myocardial regeneration following acute myocardial infarction (AMI) is limited by insufficient cell viability and a high rate of apoptosis, due to the poor regional microenvironment. Resveratrol, which is a compound extracted from red wine, has been reported to protect myocardial tissue post-AMI by increasing the expression of angiogenic and chemotactic factors. The present study aimed to investigate the effects of resveratrol on Sca-1⁺ CSCs, and to optimize Sca-1⁺ CSCs therapy for myocardial regeneration post-AMI. C57/BL6 mice (age, 6 weeks) were divided into two groups, which received intragastric administration of PBS or 2.5 mg/kg.d resveratrol. The endogenous expression of Sca-1⁺ CSCs in the heart was assessed on day 7. Furthermore, C57/BL6 mice underwent left anterior descending coronary artery ligation for the construction of an AMI model, and received an injection of 1x10⁶ CSCs into the peri-ischemic area (n=8/group). Mice received intragastric administration of PBS or resveratrol (2.5 mg/kg.d) for 4 weeks after cell transplantation. Echocardiography was used to evaluate cardiac function 4 weeks after cell transplantation. Capillary density and cardiomyocyte apoptosis in the peri-ischemic myocardium were assessed by cluster of differentiation 31 immunofluorescent staining and terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling assay, respectively. Western blot analysis was conducted to detect the protein expression levels of vascular endothelial growth factor (VEGF) and stromal cell-derived factor (SDF)-1α in the myocardium. Treatment with resveratrol increased the number of endogenous Sca-1⁺ CSCs in heart tissue after 7 days (PBS vs. Res, 1.85±0.41/field vs. 3.14±0.26/field, P<0.05). Furthermore, intragastric administration of resveratrol significantly increased left ventricle (LV) function 4 weeks after AMI, as determined by an increase in LV fractional shortening (CSCs vs. Res + CSCs, 28.82±1.58% vs. 31.18±2.02%, P<0.05), reduced LV end-diastolic diameter (CSCs vs. Res + CSCs, 0.37±0.01 mm vs. 0.35±0.02 mm, P<0.05), and reduced LV end-systolic diameter (CSCs vs. Res + CSCs, 0.26±0.01 mm vs. 0.23±0.02 mm, P<0.05). These protective effects were predominantly achieved via an increase in capillary density (CSCs vs. Res + CSCs, 281.02±24.08/field vs. 329.75±36.69/field, P<0.05) and a reduction in cardiomyocyte apoptosis (CSCs vs. Res + CSCs, 1.5±0.54/field vs. 0.83±0.40/field, P<0.05) in peri-ischemic myocardium. Western blot analysis indicated that VEGF and SDF-1α were upregulated in resveratrol-treated myocardium after a 7 day treatment or 4 weeks after AMI (7 days VEGF PBS vs. Res, 0.89±0.07 vs. 1.21±0.02, P<0.05; SDF-1α PBS vs. Res, 0.66±0.04 vs. 1.33±0.04, P<0.05; 4 weeks VEGF CSCs vs. Res + CSCs, 0.54±0.03 vs. 0.93±0.13, P<0.05; SDF-1α CSCs vs. Res + CSCs, 0.53±0.03 vs. 0.93±0.03, P<0.05). Resveratrol activated endogenous CSCs, increased capillary density and decreased cardiomyocyte apoptosis in the peri-ischemic myocardium, and augmented the effects of CSCs transplantation. These effects may be caused by the upregulation of VEGF and SDF-1α.

Introduction

The adult heart is no longer considered a terminally differentiated organ, due to the presence of cardiac stem cells (CSCs) (1). CSCs have been considered a prospective cell source for myocardial repair following acute myocardial infarction (AMI) due to their self-renewing properties, and their multipotent ability to differentiate into cardiomyocytes and endothelial cells (2,3). CSCs are positive for several stem cell antigens, including stem cell antigen (Sca)-1 and c-Kit (4).
CSC-based therapy may be considered a novel strategy to repair infarcted myocardium post-AMI; however, this method is limited by low regenerative efficiency, due to insufficient cell viability and a high rate of apoptosis following transplantation (5).

AMI is associated with inflammation and may result in the release of high concentrations of cytokines and inflammatory factors. These molecules may act as chemotactic factors that induce the homing of several progenitor cells into the infarcted areas to participate in myocardial repair (6,7). The paracrine ability of transplanted cells is considered an important factor in the improvement of cardiac function (8,9). Previous studies have indicated that the predominant effect of mesenchymal stem cells in myocardial repair was associated with the release of several cytokines and chemotactic factors involved in neovascularization or chemotaxis, including vascular endothelial growth factor (VEGF) and stromal derived factor (SDF)-1α (10,11). However, whether transplanted CSCs may also act in the same way and increase the release of VEGF and SDF-1α, resulting in the induction of microvascular sprouting, the formation of new blood vessels and an increase in progenitor cell homing remains unknown.

Resveratrol, which is a compound extracted from red wine, is considered a strong antioxidant and protective molecule in the cardiovascular system (12). A previous in vitro study demonstrated that resveratrol was able to reduce oxidative stress in several cell types, including endothelial cells, smooth muscle cells, cardiomyocytes and macrophages (13). Besides its antioxidative effects, resveratrol has been reported to exert cardioprotective effects against ischemia-reperfusion injury in animal models. Pretreatment of AMI rats with resveratrol resulted in an increase in microvessel density, and preservation of left ventricle (LV) function and blood flow (14,15). In addition, resveratrol has been shown to increase the protein levels of VEGF, angiotensin II and their receptors, in order to fulfill its angiogenic effect (16). The angiogenic effect of resveratrol was achieved through increasing the bioviability and production of nitrogen monoxide, in order to modulate the VEGF signaling pathway (17). Furthermore, upregulation of VEGF may directly influence the expression of SDF-1α and CXC chemokine receptor type 4 (CXCR-4), which are expressed on the surface of several types of stem cells, and are required for signal transduction in stem cell-associated chemotaxis, migration and homing (18,19).

Therefore, the present study hypothesized that pretreatment with resveratrol may influence the angiogenic and homing ability of CSCs via the VEGF/SDF-1α pathway, and may improve their transplantation efficiency following AMI. An AMI mouse model was generated and the effects of CSC regenerative therapy with resveratrol pretreatment were determined.

**Materials and methods**

**Ethics statement.** The present study was approved by the Soochow University Scientific and Animal Ethics Committee (Suzhou, China) and it was conducted in compliance with the Chinese national regulations on the use of experimental animals. Procedures for the animal studies were performed in accordance with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (revised in 1996). All animals were purchased from the Laboratory Animal Center of Soochow University. They were maintained on a standard diet with access to water, under a 12-h light/dark cycle at the Animal Center of the First Affiliated Hospital of Soochow University (Suzhou, China).

**CSCs culture and resveratrol administration.** CSCs were isolated from hearts harvested from male C57/BL6 mice (age, 5 weeks; weight, 10-15 g; two groups n=11 per group) with 0.1% collagenase B (Sigma-Aldrich; Merck Millipore, Darmstadt, Germany) and 0.2% trypsin (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) followed euthanasia with carbon dioxide. Cells were labeled with antigen-presenting cell-conjugated anti-Sca-1 (cat. no. 130092529; Miltenyi Biotec, Inc., Auburn, CA, USA) and were separated using a magnetic selection system employing anti-antigen-presenting microbeads (Miltenyi Biotec, Inc.). Selected cells were cultured in Dulbecco’s modified Eagle’s medium/F12 supplemented with 10% fetal bovine serum (Gibco; Thermo Fisher Scientific, Inc.), 10 ng/ml basic fibroblast growth factor, 10 ng/ml leukemia inhibitory factor, 10 ng/ml cardiotropin, 10 ng/ml epidermal growth factor (PeproTech, Rocky Hill, NJ, USA) and a 1% antibiotic solution of penicillin and streptomycin (Hyclone; GE Healthcare, Logan, UT, USA). The C57/BL6 mice (age, 5 weeks, two groups; n=8 per group) were treated with 2.5 mg/kg/d resveratrol (Baile Company, China) by intragastric administration with a stomach needle. The drug was dissolved in 0.5 ml PBS. The control group received the same volume of PBS. All mice (age, 5 weeks) received intragastric administration of resveratrol or PBS. Some (n=3 per group) were sacrificed in order to quantify CSCs after 7 days. Other mice (n=8 per group) underwent AMI construction and cell transplantation on day 7 and continued to be treated with administration of resveratrol or PBS for 4 weeks.

**Measurement of CSCs after resveratrol pretreatment.** A total of 7 days after resveratrol administration, mice were sacrificed with CO2 frozen cardiac tissue specimens (n=3/group) were obtained and sliced into 2 µm sections to quantify the number of Sca-1+ CSCs in the myocardium using an anti-Sca-1 immuno-fluorescent antibody (1:50; eBioscience, Inc., San Diego, CA, USA; cat. no. 11-5981-81). Positive cells were stained red and were counted in 10 random 100x fields using a fluorescence microscope. The average number was obtained for statistical analysis.

**AMI construction and cell transplantation.** An AMI model was constructed by surgically ligating the left anterior descending coronary artery (LAD) with a prolene suture. Successful ligation was verified by observation of a color change from red to white in the infarct area. C57/BL6 mice (age, 6 weeks; weight, 15-20 g) were randomly assigned into two groups (n=8/group), which received either PBS or 2.5 mg/kg/d resveratrol via intra-gastric administration for 7 days prior to and 4 weeks after LAD ligation. All mice received an injection with 1x106 CSCs into the peri-ischemic area and were sacrificed with CO2 4 weeks after cell transplantation for histological analysis.

**Assessment of cardiac function with echocardiography.** A total of 4 weeks after LAD ligation, transthoracic echocardiography was performed.
(SONOS 5500, Philips Medical Systems International BV, Eindhoven, The Netherlands) was used to evaluate the cardiac function of experimental mice by an observer blinded to the experiment. LV end-diastolic diameter (LVEDD), LV end-systolic diameter (LVESD), interventricular septal thickness in diastole (IVST), LV posterior wall thickness (LVPWT) and the percentage of LV fractional shortening (FS) were detected. All measurements were repeated for at least three consecutive pulsation cycles and the data were averaged for statistical analysis.

Capillary density assessment. Cluster of differentiation (CD)31 immunofluorescent staining was conducted to determine capillary density. Frozen tissue specimens of the infarcted area were sliced into 2 µm sections and were labeled with rat anti-mouse CD31 (1:800; Abcam, Cambridge, MA, USA; cat. no. ab8365) and Alexa Fluor 594 anti-rat immunoglobulin G (Invitrogen; Thermo Fisher Scientific, Inc.; cat. no. SA000064). CD31-positive cells were stained red and the number of capillaries was counted using Image Pro Plus Software version 6.0 (Media Cybernetics, Inc., Rockville, MD, USA) in 10 random 100x fields using a fluorescence microscope and averaged for statistical analysis.

Terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) assay. The present study detected cardiomyocyte apoptosis in the peri-ischemic myocardium using the TUNEL assay (Promega Corporation, Madison, WI, USA), according to the manufacturer’s protocol. Nuclei of apoptotic cardiomyocytes were stained dark brown. The number of TUNEL-positive cardiomyocyte nuclei per 200x field was counted and used for statistical analysis.

Western blot analysis. Cardiac tissues were lysed using lysis buffer with phosphatase inhibitor (Roche Diagnostics, Basel, Switzerland). A bicinchoninic acid protein assay kit (Pierce; Thermo Fisher Scientific, Inc.) was used to determine protein concentrations. Proteins (30 µg samples) were separated by 10% SDS-PAGE and were transferred to a polyvinylidene fluoride membrane (EMD Millipore, Billerica, MA, USA). The membrane was blocked with 5% nonfat dried milk dissolved in Tris-buffered saline containing 0.1% Tween-20 at room temperature for 2 h. Subsequently, the membrane was incubated with primary antibodies against VEGF (1:2,000; Abcam; cat. no. ab2350) and SDF-1α (1:1,500; Abcam; cat. no. ab9797) overnight at 4°C, followed by an incubation with peroxidase-conjugated secondary antibodies for 2 h at room temperature. Bands were detected using a chemiluminescent western blot detection system (Pierce; Thermo Fisher Scientific, Inc.). Protein expression was normalized to GAPDH. Assays were repeated three times and band intensities were quantified using the Photo-Image system (Siemens AG, Munich, Germany).

Statistical analysis. Data are presented as the mean ± standard deviation and were analyzed using SPSS 17.0 statistical software (SPSS, Inc., Chicago, IL, USA). Comparisons between two groups were performed using an unpaired Student’s t-test. P<0.05 was considered to indicate a statistically significant difference.
Results

Pretreatment with resveratrol increases the number of Sca-1+ CSCs in myocardium. The present study examined the number of Sca-1+ CSCs following resveratrol pretreatment. Sca-1+ CSCs were stained red and the nuclei were stained blue. Pretreatment with resveratrol for 7 consecutive days resulted in an increase in the number of Sca-1+ CSCs in the myocardium (PBS vs. resveratrol, 1.85±0.41/field vs. 3.14±0.26/field, P<0.05, Fig. 1).

Figure 2. Stem cell antigen-1+ CSCs transplantation combined with resveratrol treatment improved cardiac performance 4 weeks after acute myocardial infarction, as determined by measuring (A) fractional shortening (B) LVESD (C) and LVEDD (P<0.05 vs. the CSCs group). There was no significant difference in (D) IVST and (E) LVPWT between the groups. CSCs, cardiac stem cells; LVESD, left ventricular end-systolic diameter; LVEDD, left ventricular end-diastolic diameter; IVST, interventricular septal thickness in diastole; LVPWT, LV posterior wall thickness.

Figure 3. Stem cell antigen-1+ CSCs transplantation combined with resveratrol treatment increased capillary density in infarcted myocardium 4 weeks after acute myocardial infarction. *P<0.05 vs. the CSCs group. Scale bar, 100 µm. CSCs, cardiac stem cells.

Figure 4. Stem cell antigen-1+ CSCs transplantation combined with resveratrol treatment decreased cardiomyocyte apoptosis in infarcted myocardium 4 weeks after acute myocardial infarction. *P<0.05 vs. the CSCs group. Scale bar, 100 µm. CSCs, cardiac stem cells; Tunel, terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling.
CSCs combined with resveratrol protect cardiac function post-AMI. A total of 4 weeks after LAD ligation, cardiac function of the mice was evaluated by echocardiography; the results are presented in Fig. 2. CSCs transplantation combined with resveratrol administration exerted a protective effect on mice, as compared with CSCs transplantation only. Resveratrol and CSCs transplantation preserved LV FS, and reduced LVEDD and LVESD (CSCs vs. resveratrol + CSCs, P<0.05, Fig. 2). The detailed data are presented in Table I.

CSCs combined with resveratrol increase capillary density in infarcted myocardium. CD31 immunofluorescent staining was used to measure capillary density in the infarcted myocardium. As presented in Fig. 3, capillaries were stained red. A total of 4 weeks after LAD ligation, an increased number of CD31-positive cells was detected in the resveratrol-treated myocardium (CSCs vs. resveratrol + CSCs, 281.02±24.08/field vs. 329.75±36.69/field, P<0.05).

CSCs combined with resveratrol reduce cardiomyocyte apoptosis. Cardiomyocyte apoptosis was measured by TUNEL assay. TUNEL-positive cells were stained brown. As presented in Fig. 4, the number of TUNEL-positive cells in the peri-ischemic area was reduced in the resveratrol-treated myocardium (CSCs vs. resveratrol + CSCs, 1.5±0.54/field vs. 0.83±0.40/field, P<0.05).

VEGF and SDF-1α expression is upregulated in resveratrol-treated myocardium. Western blotting was used to evaluate the protein expression levels of VEGF and SDF-1α in myocardium 7 days after resveratrol administration, and 4 weeks after AMI. Western blot analysis indicated that VEGF and SDF-1α were upregulated in resveratrol-treated myocardium after 7 days of treatment, and 4 weeks after AMI (7 days VEGF PBS vs. resveratrol, 0.89±0.07 vs. 1.21±0.02, P<0.05; SDF-1α PBS vs. resveratrol, 0.66±0.04 vs. 1.33±0.04, P<0.05; 4 weeks VEGF CSCs vs. resveratrol + CSCs, 0.54±0.03 vs. 0.93±0.13, P<0.05; SDF-1α CSCs vs. resveratrol + CSCs, 0.53±0.03 vs. 0.93±0.03, P<0.05, Fig. 5).

Discussion

Since the adult heart was recognized as containing its own stem cells (20), CSCs therapy for AMI has been considered as a novel treatment strategy. CSCs are able to regenerate several types of cell, including cardiomyocytes, endothelial cells and smooth muscle cells, and directly participate in myocardial and endothelial repair (21,22). Cell therapy of AMI with CSCs has been reported to ameliorate cardiac function and tissue remodeling (23). However, just like other stem cell-based therapies for AMI, CSCs-based therapy is hindered by limited transplantation efficiency, and researchers have aimed to solve this problem with appropriate cell modification (24). Resveratrol is a compound that exerts numerous effects, including anti-inflammatory, antitumor and immunomodulatory activities. Resveratrol has also been reported to exert strong cardioprotective effects in the cardiovascular system (25,26). The present study combined this cardioprotective agent with CSCs-based therapy. A mouse model of AMI treated with CSCs transplantation plus resveratrol exhibited preserved cardiac function, increased capillary density and decreased cardiomyocyte apoptosis in the peri-ischemic myocardium. In addition, VEGF and SDF-1α expression was upregulated. These results suggested that this combination therapy may augment the effects of CSCs-mediated myocardial regeneration, and may fulfill the potential of CSCs cell therapy for AMI.

VEGF is widely known as an angiogenic molecule that serves an important role in neovascularization and angiogenesis.
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