Abstract. MicroRNAs (miRNAs) are involved in the development of non-small cell lung cancer (NSCLC). However, the biological roles of several aberrantly expressed miRNAs have not been explored yet. In the present study, miR-4491 was identified as a novel upregulated miRNA in NSCLC tissues and cell lines. Downregulation of miR-4491 by a miR-4491 inhibitor inhibited the proliferation and triggered the apoptosis of NSCLC cells. Tripartite motif containing 7 (TRIM7), a tumor suppressor gene expressed in NSCLC, was demonstrated in the present study to be directly targeted by miR-4491. This finding was verified by bioinformatics analysis, reverse transcription-quantitative PCR, western blotting and dual luciferase reporter assays. Furthermore, downregulation of miR-4491 inactivated nuclear factor-κB signaling via induction of TRIM7. In addition, TRIM7 silencing attenuated the effect of miR-4491 inhibitor in NSCLC cells. The decreased TRIM7 level in NSCLC tissues was negatively correlated with miR-4491 expression in NSCLC tissues. In conclusion, the findings from this study demonstrated that miR-4491 expression was upregulated in NSCLC tissues and cells and that miR-4491 may promote NSCLC progression via targeting TRIM7.

Introduction

Lung cancer (LC) is a life-threatening disease with 1,735,350 new cancer cases diagnosed and 609,640 mortality cases in 2018 in the United States and ranks second among all types of cancer worldwide (1,2). The major risk factor for LC is smoking, which accounts for 75-80% of LC-associated mortality (3). Non-small cell lung cancer (NSCLC) accounts for 85% of LC cases and can be further divided into two main subtypes, including lung squamous cell carcinoma (LUSC) and lung adenocarcinoma (LUAD) (4). The occurrence and development of NSCLC involve a multistep carcinogenic process, which includes numerous variations of gene expression, for instance, V-Ki-ras2 Kirsten rat sarcoma viral oncogene homolog, and signal transduction, including nuclear factor-κB (NF-κB), ERK1/2 and AKT (5). The combination of surgery and chemotherapy remains the primary treatment method for patients with NSCLC (6). Despite significant improvements in the detection, diagnosis and targeted therapy, the 5-year survival rate of patients with NSCLC is only ~15% (7). The underlying mechanisms promoting the development of NSCLC are very specific and require further investigation in order to develop novel therapeutic strategies.

MicroRNAs (miRNAs) are non-coding RNA molecules of ~20 nucleotides in length that bind to the 3' untranslated region (UTR) of the mRNA. This miRNA-mRNA interaction results in mRNA degradation or translation repression, which regulates target gene expression (8). Furthermore, it has been verified that miRNAs serve as regulators in multiple physiological and pathological conditions (9). The expression of specific miRNAs is commonly dysregulated in tumor tissues, indicating that miRNAs may serve as oncogenes or tumor suppressors according to their target genes (10), leading to the regulation of cell proliferation, cell death and metastasis (11). Based on their functions, miRNAs are considered as therapeutic agents (8,12).

Numerous miRNAs have been reported to be involved in the development of NSCLC. For example, miR-486 was found to be associated with the overall survival of patients with advanced-stage NSCLC (13), whereas miRNA-148a can inhibit NSCLC cell invasion and migration in vitro as well as cancer metastasis in vivo (14). Furthermore, based on the analysis of the miRNA expression profile from the peripheral blood of healthy controls and patients with LC, miR-4491 was shown to be increased (15). However, its role in the development of NSCLC remains unknown.

Materials and methods

Collection of tissue samples. Tumor and matched normal tissues were collected from 78 patients (62 men and 16 women)

MicroRNA-4491 enhances cell proliferation and inhibits cell apoptosis in non-small cell lung cancer via targeting TRIM7

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diagnosed with NSCLC at the Shanxi Provincial Cancer Hospital between March 2016 and October 2018. The age range of patients was 45-76 years, 33 patients were ≤65 years and 45 patients were >65 years. The clinicopathological characteristics of patients are listed in Table I. A total of 52 patients were at I-II stages, 26 patients were at III-IV stages, 44 patients were diagnosed with LUAD, 34 patients were diagnosed with LUSC, 37 patients had no lymph node metastasis and 41 patients exhibited lymph node metastasis. All patients provided written informed consent prior to study enrollment. The present study was approved by the Ethics Committee of Shanxi Provincial Cancer Hospital (IRB no. SXPCP20160302).

**Cell lines.** The bronchial epithelium transformed cell line BEAS-2B and the LUAD cell lines A549 and NCI-H1650 were purchased from the American Type Culture Collection. The cells were maintained in RPMI-1640 medium (HyClone; Cytiva), which was supplemented with 10% fetal bovine serum (HyClone) and placed at 37˚C in a humidified incubator containing 5% CO₂.

**Cell transfections.** miR-negative control mimic (NC, 50 nM, 5'-UAGUCUGGAGACUCACUCAC-3'), miR-NC inhibitor (50 nM, 5'-UAACGGAUCACAGUUGCCUA-3'), miR-4491 inhibitor (50 nM, 5'-UUUGUCACACCAGUCCA CAUU-3'), miR-4491 mimic (50 nM, 5'-AAUGUGACUGGUGUCCAAA-3'), si-control (100 nM, 5'-TTCCTCGGA CGTGTCACTTTT-3') and si-Tripartite motif containing 7 (TRIM7, 100 nM, 5'-CGGAAAAGAAGGAGAGCA-3') were synthesized by GenePharma. Cell transfections were performed by Lipofectamine 2000® solution (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's instructions. Transfections were performed at 37˚C for 48 h and cells were collected for subsequent experiments.

**miR-4491 expression profile.** The Starbase V2.0 project (http://starbase.sysu.edu.cn/) was used for the analysis of miR-4491 expression levels in NSCLC and normal tissues from The Cancer Genome Atlas (TCGA) datasets (TCGA-LUAD and TCGA-LUSC) (16).

**mRNA and miRNA quantification.** Total RNA was isolated from tumor and normal tissues derived from patients with NSCLC and from BEAS-2B, A549 and NCI-H1650 cells using TRIzol® (Invitrogen; Thermo Fisher Scientific, Inc.). cDNA synthesis was performed using TaqMan microRNA reverse transcription kit (Applied Biosystems; Thermo Fisher Scientific, Inc.) and TransScript First-Strand cDNA Synthesis SuperMix (Beijing TransGen Biotech Co., Ltd.). Reverse transcription-quantitative PCR (RT-qPCR) experiments were performed on an ABI7500 system (Applied Biosystems; Thermo Fisher Scientific, Inc.) under the following conditions: 95˚C for 5 min, followed by 40 cycles of 95˚C for 10 sec, 60˚C for 30 sec and 72˚C for 1 sec. miR-4491 levels were normalized to those of U6, TRIM7 levels were normalized to those of GAPDH. The primers were synthesized by GenScript. The sequences of primers were as follows: GAPDH, forward 5'-GAAGTTGAGCTCGG AGTC-3', reverse 5'-GAAGTGATGTGGAGGTTC-3', U6, forward 5'-GCTCGAGGAGGATCAT-3', reverse 5'-GCAACAACACACATCTCCCA-3', TRIM7, forward 5'-ATTATATAGGTGTCCACATA-3', reverse 5'–TATGTC GACACCTATATAAT-3', miR-4491, forward 5'-AAT GCAATCTGTTGTAACCCAA-3' and reverse 5'-TTTGGT CACCCATGTCACAT-3'. The relative expression levels were normalized to endogenous control and were expressed as 2^ΔΔCq (17).

**Western blotting.** A549 and NCI-H1650 cells were lysed using RIPA on ice buffer (Sigma-Aldrich; Merck KGaA). The protein concentration was determined using the BCA method (Sigma-Aldrich; Merck KGaA). Proteins (20 μg) were separated by 8% SDS-PAGE and were transferred onto PVDF membranes. Membranes were blocked by 5% skimmed milk for 2 h at room temperature and incubated with primary antibodies against TRIM7 (cat. no. ab105330; 1:1,000; Abcam), β-actin (cat. no. 4970; 1:1,000; Cell Signaling Technology, Inc.) overnight at 4˚C. Membranes were then incubated with goat-anti-rabbit secondary antibody (cat. no. 7074; 1:2,000, Cell Signaling Technology, Inc.) for 2 h at room temperature. The protein blots were developed by ECL Western Blot Kit (Pierce; Thermo Fisher Scientific, Inc.). Band intensities were analyzed by Image Lab™ (version 4.0; Bio-Rad Laboratories, Inc.). Relative expression levels were normalized to endogenous control β-actin using ImageLab™ (version 3.0; Bio-Rad Laboratories, Inc.).

**Cell proliferation assay.** The proliferation of A549 and NCI-H1650 cells was evaluated using the Cell Counting Kit-8 (CCK-8; Dojindo Molecular Technologies, Inc.). Briefly, A549 and NCI-H1650 cells (3x10^3) were seeded in 96-well plates and incubated at 37˚C for 48 h. Subsequently, cells were incubated with CCK-8 reagent (10 μl) for 3 h at 37˚C. The absorbance was measured at 450 nm by spectrophotometry.

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**Table I. Clinicopathological characteristics of patients with non-small cell lung cancer.**

| Characteristics          | Patient number |
|--------------------------|----------------|
| Age ≤65                  | 33             |
| Age >65                  | 45             |
| Sex                      |                |
| Male                     | 62             |
| Female                   | 16             |
| TNM stage                |                |
| I-II                     | 52             |
| III-IV                   | 26             |
| Pathological type        |                |
| Adenocarcinoma           | 44             |
| Squamous cell carcinoma  | 34             |
| Lymph node metastasis    |                |
| No                       | 37             |
| Yes                      | 41             |

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**Patients with non-small cell lung cancer.**

- **Sex:** 62 males, 45 females
- **Age:** 33 patients ≤65 years, 45 patients >65 years
- **Pathological Type:**
  - 44 patients Adenocarcinoma
  - 34 patients Squamous cell carcinoma
- **TNM Stage:**
  - I-II: 52 patients
  - III-IV: 26 patients
- **Lymph Node Metastasis:**
  - No: 37 patients
  - Yes: 41 patients

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**Abbreviations:**
- NSCLC: non-small cell lung cancer
- LUAD: lung adenocarcinoma
- LUSC: lung squamous cell carcinoma
- miRNA: micro RNA
Cell apoptosis assay. A549 and NCI-H1650 cell apoptosis was assessed with the Annexin V-FITC/propidium (PI) apoptosis detection kit (Sigma-Aldrich; Merck KGaA). A549 and NCI-H1650 cells (1x10⁶) were seeded in 6-well plates and incubated at 37°C for 48 h. Subsequently, cells were stained with Annexin V (5 µl) and PI (1 µl) for 10 min at room temperature in the dark. The number of apoptotic cells was estimated by a FACSCalibur flow cytometer (BD Biosciences). The data were analyzed by the CellQuest software (version 3.3; BD Biosciences).

Prediction of target genes. miRWalk (http://mirwalk.umm.uni-heidelberg.de/) and TargetScan (http://www.targetscan.org/vert_72/) softwares were used to predict miR-4491 targets. The targets predicted by miRWalk were further selected for score analysis. A score of 1.0 was considered as optimal. The 50 top ranked targets of miR-4491 predicted by TargetScan were also selected. The two gene sets were overlapped by online Vein Map (http://bioinformatics.psb.ugent.be/webtools/Venn/). The functions of the 7 overlapped genes were subsequently investigated by literature review.

Dual luciferase reporter assays. Wild-type or mutant TRIM7 3’UTR were ligated into pGL3-luciferase reporter plasmids (Promega Corporation), co-transfected with miR-4491 mimic or miR-NC into A549 and NCI-H1650 cells by Lipofectamine™ 2000 solution (Invitrogen; Thermo Fisher Scientific, Inc.) and incubated for 48 h at 37°C. Subsequently, A549 and NCI-H1650 cells were trypsinized and transferred to 24-well plates. Luciferase reporter activities were examined by the Dual Luciferase Assay System (Promega Corporation). The luciferase reporter activities were normalized to the Renilla luciferase activity (Promega Corporation).

Statistical analysis. The data were analyzed by GraphPad Prism 6 (GraphPad Software, Inc.) and were expressed as the means ± standard deviation. The differences between two
groups from tissues and cell lines were analyzed using paired or unpaired Student's t-test, respectively. The differences between three groups were analyzed by one-way ANOVA and Newman Keuls analysis. The correlation between miR-4491 and TRIM7 expression was analyzed by Pearson Correlation analysis. P<0.05 was considered to indicate a statistically significant difference.

Results

miR-4491 levels are increased in patients with NSCLC. The data derived from TCGA indicated that miR-4491 levels were significantly increased in cancer tissues from patients with LUAD (n=512) compared with adjacent normal tissues from cancer patients (n=20; Fig. 1A). In addition, miR-4491 levels were significantly increased in cancer tissues from patients with LUSC (n=475) compared with those in adjacent normal tissues from cancer patients (n=38; Fig. 1B). The data derived from patient tissues collected in the present study indicated that the miR-4491 levels in NSCLC tissues (n=78) were significantly higher compared with those in the adjacent normal tissues (Fig. 1C). A similar expression profile of miR-4491 was also observed in patients with LUAD (n=42; Fig. 1D) and LUSC (n=36; Fig. 1E).

Downregulation of miR-4491 negatively affects the proliferation and induces the apoptosis of NSCLC cell lines. To detect the expression profile of miR-4491 in NSCLC cell lines, A549 and NCI-H1650 cells were selected. The results indicated that miR-4491 expression was significantly increased in A549 and NCI-H1650 cells compared with BEAS-2B cells (Fig. 2A).
Furthermore, the effects of the downregulation of miR-4491 in NSCLC cell lines were determined. The downregulation of miR-4491 expression by the miR-4491 inhibitor was confirmed in A549 and NCI-H1650 cells (Fig. 2B). Subsequently, transfection of A549 and NCI-H1650 cell with miR-4491 inhibitor resulted in a decrease in cell proliferation (Fig. 2C and D) in a time-dependent manner compared with that of the miR-NC group. In addition, miR-4491 inhibitor induced apoptosis of A549 and NCI-H1650 cells (Fig. 2E and F).

In addition, the effects of miR-4491 overexpression were determined in NSCLC cell lines. The overexpression of miR-4491 in A549 and NCI-H1650 cells using miR-4491 mimic was verified by RT-qPCR analysis (Fig. 2G). The effects caused by the miR-4491 mimic were opposite to those caused by the miR-4491 inhibitor. miR-4491 mimic increased A549 and NCI-H1650 cell proliferation (Fig. 2H and I) in a time-dependent manner.

TRIM7 is targeted by miR-4491 in NSCLC cell lines. The gene sets from miRWalk (n=2,231) and TargetScan (n=51) were retrieved and 7 overlapping genes (Fig. 3A) were identified. The complementary sites between TRIM7 and miR-4491 are presented in Fig. 3B.
Cell transfection with miR-4491 inhibitor induced a significant upregulation of TRIM7 mRNA and protein levels (Fig. 3C and D) and a concomitant significant downregulation of the protein levels of p65 (Fig. 3E) in A549 and NCI-H1650 cells. Furthermore, miR-4491 mimic significantly decreased the relative luciferase activity of TRIM7 wild-type (Fig. 3F) but not TRIM7 mutant (Fig. 3G) in A549 and NCI-H1650 cells. Downregulation of miR-4491 decreases p65 levels by targeting TRIM7 in NSCLC cells.

Downregulation of miR-4491 decreases p65 levels by targeting TRIM7 in NSCLC cells. TRIM7 was successfully downregulated using siRNA transfection in A549 and NCI-H1650 cells (Fig. 4A).

In A549 and NCI-H1650 cells transfected with miR-4491 inhibitor, p65 protein expression was significantly decreased, an effect that was rescued following co-transfection with si-TRIM7 (Fig. 4B).

Downregulation of miR-4491 negatively affects the proliferation and triggers the apoptosis of NSCLC cells by targeting TRIM7. The miR-4491 inhibitor inhibited the proliferation of A549 and NCI-H1650 cells (Fig. 5A and B) in a time-dependent manner compared with that of the miR-NC + si-control group, which was rescued following the co-transfection with si-TRIM7. In addition, the miR-4491 inhibitor induced A549 and NCI-H1650 cell apoptosis (Fig. 5C and D), which was reversed following the co-transfection with si-TRIM7.

miR-4491 expression is negatively correlated with TRIM7 expression in NSCLC tissues. The data derived from patient tissues collected in the present study indicated that TRIM7 expression in NSCLC tissues (n=78) was significantly lower compared with that in the adjacent normal tissues (Fig. 6A). A similar expression profile of TRIM7 was also observed in patients with LUAD (n=42; Fig. 6B) and LUSC (n=36; Fig. 6C). In addition, miR-4491 expression was negatively correlated with TRIM7 expression in NSCLC tissues (Fig. 6D), LUAD tissues (Fig. 6E) and LUSC tissues (Fig. 6F).

Discussion

miRNAs are emerging biomarkers used in the diagnosis and treatment of patients with NSCLC (18,19). In particular, miR-21 (20), miR-9 (21) and miR-143 (22) have been used as biomarkers of NSCLC. miR-4491 has shown potential diagnostic value for several diseases, as it was reported to be downregulated in ischemic stroke (23), while it is upregulated in gastric cancer (24) and chronic heart failure (25). A recent study reported a diagnostic value of miR-4491 in LC (15). However, its therapeutic value in NSCLC has yet to be reported.

In the present study, the data from TCGA and from the collected tumor tissues indicated that miR-4491 expression was increased in tumor tissues of patients with NSCLC compared with normal matched tissues. Furthermore, miR-4491 downregulation negatively affected the proliferation and triggered the apoptosis of NSCLC cells. These findings suggested that miR-4491 may have an oncogenic role in NSCLC.

The miRNA-mRNA interaction regulates target gene expression by inducing mRNA degradation or translational repression (8). This interaction might therefore regulate cell proliferation, cell death and metastasis (11). The involvement of
Figure 5. Downregulation of miR-4491 inhibited cell proliferation and induced cell apoptosis by targeting TRIM7 in non-small cell lung cancer cell lines. Effect of si-TRIM7 was rescued by miR-4491 inhibitor and resulted in (A and B) decreased proliferation and (C and D) increased apoptosis of A549 and NCI-H1650 cells. ***P<0.001 vs. miR-NC + si-control, &P<0.05, &&P<0.01 and &&&P<0.001 vs. miR-4491 inhibitor + si-control. miR, microRNA; TRIM7, tripartite motif containing 7; NC, negative control; si, small interfering.

Figure 6. miR-4491 expression is negatively correlated with TRIM7 expression in patients with NSCLC. TRIM7 expression level in tumor tissues from patients with (A) NSCLC, (B) LUAD and (C) LUSC was significantly lower than those in adjacent normal tissues. There was a negative correlation between miR-4491 and TRIM7 expression in (D) NSCLC tissues, (E) LUAD tissues and (F) LUSC tissues. ***P<0.001 and ***P<0.001, vs. normal. miR, microRNA; TRIM7, tripartite motif containing 7; NSCLC, non-small cell lung cancer; LUSC, lung squamous cell carcinoma; LUAD, lung adenocarcinoma.
miR-4491 target genes in NSCLC requires thus further investigation in order to determine the function of miR-4491 in NSCLC.

In the present study, miRWalk and TargetScan were used to identify 7 overlapping genes, including INTS3 and NABP interacting protein (INIP), protein O-mannose kinase (POMK), muscle RAS oncogene homolog (MRAS), alpha-1,3-mannosylglycoprotein 4-beta-N-acetylglucosaminyltransferase B (MGAT4B), angiotensin II receptor type 1 (AGTR1), chromosome 1 open reading frame 116 (Clorf116) and TRIM7, which could all be targeted by miR-4491. Among these genes, there has not been any reports about the function of INIP or POMK in the development of cancer. MRAS and MGAT4B however were reported as oncogenic genes, and MRAS can initiate tumor formation in lungs (26). MGAT4B transcripts are also upregulated in diethyltritosamine-induced mouse model for hepatocellular carcinoma (27). There are only few reports about the expression profile of AGTR1 and Clorf116 in patients with lung cancer. AGTR1 expression is shown to be decreased in LUAD tissues compared with adjacent normal tissues (28), and downregulation of Clorf116 is associated with a poor prognosis of patients with lung cancer (29). TRIM7 belongs to the TRIM protein family, which is involved in cell proliferation (30), cell apoptosis (31) and immunity (32), and is a well-known tumor suppressor involved in the development of various types of cancer. For example, TRIM7 inhibits the progression of hepatocellular carcinoma by negatively regulating Src (33), and TRIM7 expression is decreased in tumor tissues from patients with LC (34). Subsequently, TRIM7 was selected for the subsequent experiments in the present study.

In the present study, TRIM7 was targeted and negatively regulated by miR-4491. However, the downstream targets of TRIM7 were not detected.

NF-κB consists of 5 subunits, including NF-κB1 (p50 and its precursor p105), NF-κB2 (p52 and its precursor p100), RelA (p65), RelB and c-Rel. In addition, the p50/65 heterodimer is enriched in nearly all types of cells (35,36). NF-κB can target genes that stimulate cell proliferation, inflammation, angiogenesis, metastasis and cancer cell resistance to chemotherapy and radiotherapy (36). NF-κB p65 is commonly activated in LC (37). TRIM7 has been initially identified to degrade p65 in LC (34). The present study demonstrated that miR-4491 inhibitor decreased p65 protein expression in NSCLC cells. This effect was reversed following transfection with si-TRIM7. These observations were consistent with previous findings.

However, whether TRIM7 could affect the role of miR-4491 in the proliferation and apoptosis of NSCLC cells remains unknown. Cell transfection with miR-4491 inhibitor negatively affected cell proliferation and triggered apoptosis, whereas these effects were reversed by si-TRIM7 in NSCLC cells.

The present study demonstrated also that TRIM7 expression in cancer tissues from patients with NSCLC, including LUAD and LUSC, was downregulated compared with the adjacent normal tissues. In addition, miR-4491 expression was negatively correlated with TRIM7 expression in NSCLC tissues, including LUAD and LUSC.

Taken together, the results from the present study suggested that miR-4491 may enhance cell proliferation and resistance to apoptosis as well as the activation of p65 in NSCLC by targeting TRIM7.

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Availability of data and materials
The data and materials are available from the corresponding author on reasonable request.

Authors' contributions
FH, GC, YG, BL, YS, XQ, HT and XZ conducted the experimentations, performed the data interpretation and data analysis, and wrote the article. FH, GC, YG, BL, YS, XQ, HT, XZ and HZ confirmed the authenticity of all the raw data. All authors read and approved the final manuscript.

Ethics approval and consent to participate
The present study was approved by the Ethics Committee of Shanxi Provincial Cancer Hospital (IRB no. SXCPCP20160302). All patients provided written informed consent prior to study enrollment.

Patient consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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