Characterization of potential drug targeting folate transporter proteins from Eukaryotic Pathogens [version 2; peer review: 2 approved with reservations]

Previously titled: Genome-wide characterization of folate transporter proteins of eukaryotic pathogens

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Abstract

Background: Medically important pathogens are responsible for the death of millions every year. For many of these pathogens, there are limited options for therapy and resistance to commonly used drugs is fast emerging. The availability of genome sequences of many eukaryotic microbes is providing critical biological information for understanding parasite biology and identifying new drug and vaccine targets.

Methods: We developed automated search strategies in the Eukaryotic Pathogen Database Resources (EuPathDB) to construct a protein list and retrieve protein sequences of folate transporters encoded in the genomes of 200 eukaryotic microbes. The folate transporters were categorized according to features including mitochondrial localization, number of transmembrane helix, and protein sequence relatedness.

Results: We identified 234 folate transporter proteins associated with 63 eukaryotic microbes including 48 protozoa, 13 fungi the others being algae and bacteria. Phylogenetic analysis placed 219 proteins into a major clade and 15 proteins into a minor clade. All the folate transporter sequences from the malaria parasite, Plasmodium, belonged to the major clade. The identified folate transporters include folate-binding protein YgfZ, folate/pteridine transporter, folate/biopterin transporter, reduced folate carrier family protein and folate/methotrexate transporter FT1. About 60% of the identified proteins are reported for the first time. Phylogeny computation shows the similarity of the proteins identified.

Conclusion: These findings offer new possibilities for potential drug development targeting folate-salvage proteins in eukaryotic pathogens.

Open Peer Review

Invited Reviewers

version 2
(revision)
13 Jul 2017

1

2

version 1
12 Jan 2017

1

2

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Any reports and responses or comments on the article can be found at the end of the article.
Keywords
Folate transporter, Eukaryotic pathogens, Drug discovery, Putative homologues

This article is included in the Neglected Tropical Diseases collection.

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Author roles: Falade MO: Conceptualization, Data Curation, Formal Analysis, Methodology, Writing – Original Draft Preparation, Writing – Review & Editing; Otarigho B: Conceptualization, Data Curation, Formal Analysis, Methodology, Project Administration, Supervision, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: B.O. was supported by a TWAS-CNPq fellowship (FP number: 3240274297)

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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How to cite this article: Falade MO and Otarigho B. Characterization of potential drug targeting folate transporter proteins from Eukaryotic Pathogens [version 2; peer review: 2 approved with reservations] F1000Research 2017, 6:36
https://doi.org/10.12688/f1000research.10561.2

First published: 12 Jan 2017, 6:36 https://doi.org/10.12688/f1000research.10561.1
Antifolate drugs target this pathway and are the most important and successful antimicrobial chemotherapies targeting a range of bacterial and eukaryotic pathogens. While most parasitic protozoa can synthesize folates from simple precursors, such as GTP, p-aminobenzoic acid (pABA) and glutamate, higher animals and humans cannot\textsuperscript{30}. Additionally, a few of these parasites can also salvage folate as nutrient from their host\textsuperscript{31}. These folate compounds are important for synthesis of DNA, RNA and membrane lipids and are transported via receptor-mediated or and carrier-mediated transmembrane proteins; folate transporters\textsuperscript{20,22}. Importantly, antifolate chemotherapies that target the biosynthesis and processing of folate cofactors have been effective in the chemotherapy of bacterial and protozoan parasites\textsuperscript{25}. More importantly, the folate pathway has also been confirmed as being essential in some eukaryotic pathogens such as \textit{Plasmodium}, \textit{trypanosomes} and \textit{Leishmania}\textsuperscript{39}.

In addition to the folate biosynthesis pathway, proteins that mediate transport of useful nutrients such as folic acid have been identified as important chemotherapeutic drug targets\textsuperscript{34,35,36}. Hence, the folate pathway, metabolites and transporters continue to be extensively studied for identification of new enzymes including transporters, which may serve as new drug targets\textsuperscript{22}. Recent estimates have ascribed eight different membrane transporters to eukaryotes\textsuperscript{41}.

Proteins that mediate transportation of folates have been well studied in a few parasites such as \textit{Plasmodium falciparum}, \textit{Trypanosoma brucei, Leishmania donovani} and \textit{Leishmania major}\textsuperscript{37,38}. These studies have provided information on mode of action of drugs\textsuperscript{53,27,28} in addition to studies describing mechanisms of parasite drug resistance\textsuperscript{26,32}. However, folate transport proteins remain unidentified and uncharacterized in many other eukaryotic pathogens. This is despite the sequencing of the genomes of most eukaryotic microbes, which has produced a vast wealth of data that could aid in identification of druggable pathogen-specific proteins\textsuperscript{35–39}. It is therefore imperative to search and identify from these parasite genomes additional proteins such as folate transporters that may serve as novel drug targets\textsuperscript{40,41}.

Therefore, in an attempt to identify and characterize targets for novel therapeutics, we report herein an extensive search of folate transporters from pathogen genomes. In addition, we investigated the evolutionary relationship of these transporters in a bid to determine similarities and differences that make them attractive drug targets. The knowledge provided may assist in the design of new antifolates for protozoan parasites.

**Methods**

We extracted protein sequences of approximately 200-pathogens that mediate transportation or salvage of folates from Eukaryotic Pathogen Genome Database Resources (http://eupathdb.org/eupathdb/), and from the literature using a key-word search. We also searched the 200-pathogen genome sequences archived at the Eukaryotic Pathogen Genome Database Resources (http://eupathdb.org/eupathdb/). The search was for all proteins that mediate transportation or folate salvage alone or folate salvation and related compounds (such as pteridine, bipterin and methotrexate) together. This database gives public access to most sequenced emerging/re-emerging infectious pathogen genomes\textsuperscript{42}. We utilized
the word “folate” for search on the gene text and “folic acid” was used to confirm the hits. Hit results containing proteins annotated as folate-binding protein YgfZ, folate/pteridine transporter, folate/biopterin transporter, reduced folate carrier family protein, folate/methotrexate transporter FT1, Folate transporters alone and other folate related proteins were retrieved. The complete list of proteins extracted from EuPathdb is presented in Dataset 1⁴. The folate transporters were classified based on type of transporter, number of transmembrane helix (TMH) and localization (either cell or mitochondrial membrane) of transporter. Gene sequences were obtained in FASTA format for transporter proteins using the sequence download tool on EuPathDB (http://eupathdb.org/eupathdb/).

To ensure that most of the retrieved proteins had not been previously studied, we performed a literature search on PubMed (http://www.ncbi.nlm.nih.gov/pubmed/?term=) and Google Scholar (https://scholar.google.com) using the query “folate transporter + Parasite name”. The protein sequence information (Dataset 1⁴) obtained from literature search was used for a BLAST search on EupathDB (http://eupathdb.org/eupathdb/), UniprotDB (http://www.uniprot.org) and GeneDB (http://www.genedb.org/Homepage). The protein information are included in Dataset 1⁴ and summarised in Dataset 2⁴, which are marked as either identified in other literature or in this research work. Sequence data were edited on textEdit mac version and uploaded to Molecular Evolutionary Genetics Analysis (MEGA) platform version 7.0 obtained from http://www.megasoftware.net⁵. The 234 sequences were aligned using muscle tools with large alignment (Max iterations = 2) selected while other settings were left at defaults. Evolutionary history was inferred using the Neighbor-Joining method⁶. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) was also analysed⁷. The tree was drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were computed using the number of differences method⁸. While uniform rate and complete deletion was selected for substitution rates and data subset, respectively. Other parameters were at default settings. All positions containing gaps and missing data were eliminated. The newick format of the tree was exported and opened on FigTree 1.4.2 platform downloaded from http://tree.bio.ed.ac.uk/software/figtree⁹. The final tree was constructed using radial tree layout. Additional consistency of sub-phylogenies based on the transporter type. Since folate-binding protein YgfZ, folate/pteridine transporter, folate/biopterin transporter, putative, reduced folate carrier family protein, folate/methotrexate transporter FT1, putative folate transporters alone and others have 10, 25, 132, 2, 7, 49 and 9. So we decided to reconstruct the phylogeny based folate transporter, folate-biopterin transporter after considering the identification number, the species diversity in each category.

Results
A methodological search for folate transporters in all eukaryotic microbe genomes examined under EuPathDB with validation via GenBank, GeneDB and UniProt contained a total of 234 proteins (detail features of proteins are presented in Dataset 1⁴). We identified these transporters in 28 pathogen species (containing 63 strains) cutting across 12 phyla (Dataset 2⁴). The parasites with the highest number of folate transporters are Phytophthora parasitica INRA-310, P. infestans T30-4 and Leptomonas pyrrocorys H10 with 20, 16 and 16 proteins, respectively. While Aspergillus clavatus NRRL 1, A. flavus NRRL3357, A. macrognys ATCC 38327, Crithidia fasciculata strain CF-CI and others have one folate transporter protein each (Dataset 2⁴). The different proteins identified to be involved in folate salvage or related molecules were folate-binding protein YgfZ, folate/pteridine transporter, folate/biopterin transporter, reduced folate carrier family protein, folate/methotrexate transporter FT1 and folate transporters having a 4%, 11%, 56%, 1%, 3% and 21% identity, respectively. Proteins that did not belong to these groups were classified as others (4%) (Figure 1A). A good number of the proteins identified had predicted transmembrane helixes with a few having none (Figure 1B). Furthermore, a number of the transporters possess signal peptides (Dataset 1⁴), which may be required for targeting to cellular locations. Deciphering the sequence of the targeting signal may indicate its product destination.

Our literature search for parasite folate transporters on PubMed and Google Scholar indicated 60% (38 out 63) of the proteins were identified for the first time as presented in Dataset 2⁴, while 40% have been previously investigated. Besides, the Leishmania folate transporters we came across were not found on the EupathDB resource. We thus performed a BLAST search of Kinetoplastida on EupathDB, the returned hits were folate/biopterin transporter for L. infantum. The only Plasmodium species with results for proteins that salvage folate was P. falciparum. Our study, however, describes for the first time the presence of these transporters in other Plasmodium species. There were no transporter proteins deposited in EupathDB for P. malariae and P. ovale. However, folate transporters I and II were retrieved from our search of GeneDB for P. malariae and P. ovale curtisi, respectively.

Our analysis of folate transporters indicate the presence in Plasmodium species of two proteofoms; folate transporter I and II (Dataset 1⁴). All Leishmania species identified possess folate/biopterin transporters and not folate transporters. Trypanosome species have both folate/pteridine and folate transporters; T. cruzi Dm28c, T. cruzi Sylvio X10/1 and T. cruzi CL Brener Esmeraldo-like, T. cruzi CL Brener Non-Emesraldo-like and T. cruzi marinkellei strain B7 all have folate/pteridine transporter while T. brucei TREU927, T. brucei Lister strain 427, T. brucei gambiensse DAL972, T. congolense IL3000 possess folate transporters. Eukaryotic parasites like Eimeria acervulina Houghton, E. brunetti Houghton, E. maxima Weybridge, E. necatrix Houghton, E. praecox Houghton, E. tenella strain Houghton and Neospora caninum Liverpool all boast folate/methotrexate transporter FT1. The folate-binding protein YgfZ was found in the fungus, Allomyces macrogynys ATCC 38327, the protist C. fasciculata strain CF-CI, C. immitis RS, the feline protozoan, Hammondia hammondi strain H.H.34, Sarco- cystis neurona SN3, S. punctatus DAOM BR117, T. gondii GT1, T. gondii ME49, T. gondii VEG and T. brucei TREU927. Parasites such as Microsporidium daphniae UGP3 and the amoeba Naegle- ria fowleri ATCC 30863 possess the reduced folate carrier family protein. We observed that 7% of the identified proteins are localized on the mitochondrial membrane of some pathogens such as the fungi Aspergillus clavatus NRRL 1, A. flavus NRRL3357,
Figure 1. Categorization of proteins identified based on [A] Type of transporters and [B] Number of TMHs.
C. immitis RS, the yeast Cryptococcus neoformans var. grubii H99, Fusarium graminearum PH-1, A. capsulatus G186AR, Leptomonas pyrrhocoris H10, the food fungus Neosartorya fischeri NRRL 181, Phytophthora parasitica INRA-310 and P. ultimum DAOM BR144. The remaining proteins are localized on the plasma membrane (Dataset 1).

Approximately 15% (34/234) of the folate transporters identified possess signal peptides with the tryptophanases with the most signal peptides. Deductions can be made of the probable destination within the cell of any transporter by its signal peptide sequence; thus, further work may seek to decipher the sequence of the targeting signal to determine its localization. The proteins identified all have transmembrane helixes with the exception of the alveolate Chromera velia CCMP2878, apicomplexan P. berghei ANKA, S. neurona SN3, the kinetoplastid T. brucei TREU927, T. grayi ANR4 and protist Vitrella brassicaformis CCMP3155 with Gene ID’s Cvel_17766, PBANKA_0713700, SN3_01500005, Tb927.8.6480, Tgr.2739.1000 and Vbra_15327, respectively (Dataset 1).

The phylogenetic tree (Figure 2) shows the evolutionary position, history and relationship of all the folate transporters identified in this work. The type of transporter or species/strain was used for constructing phylogenic trees, with the 234 proteins identified forming two clades, a major and minor. The major clade lacked a sub-clade, while the minor clade possessed a sub-clade. All proteins identified were distributed between the two major clades; except for folate/methotrexate transporter and mitochondrial folate transporter, with the latter present on the major clade and the former on the minor clade exclusively. All the species are represented on both clades, however, V. brassicaformis CCMP3155, Plasmodium species, A. clavatus NRRL, A. flavus NRRL3357, A. macrogyrus ATCC 38327, C. fasciculata strain CI-CI, C. immitis RS, C. immitis RS, C. mutis RN66, C. neoforms var. grubii H99, C. neoforms var. grubii H99, Leishmania species, N. bombycis CQ1, N. caninum Liverpool, F. graminearum PH-1 and H. hammodi strain H.H.34 are exclusively on the major clade. There are some parastases that were identified once, as shown in Dataset 1; these are mostly in the large clade. Some of these pathogens include P. ultimum DAOM BR144, which has mitochondrial folate transporter/carrider proteins similar to Homo sapiens, E. cuniculi GB-M1, which has proteins similar to folate transporter, and S. punctatus DAOM BR117, which has folate-binding protein YgfZ. These were the only proteins of the aforementioned species identified in this work. However, M. daphniae UGP3, which had reduced folate carrier domain containing protein, was the only parastase that was found in the small clade. Improving on our phylogenetic analysis, we performed a sub-phylogenetic reconstruction (Figure 3–Figure 5) based on the substrate type of the transport proteins. After phylogenetic analysis each sub-phylogeny show a clear characterization except for folate-biopterin transporters (Figure 4), which fell in a different clade save for Leptomonas species and C. velia. The newick formats of the phylogenetic trees in Figure 2–Figure 5 are presented as Supplementary Dataset 1–Supplementary Dataset 4.

Discussion

Folate transporters are important proteins involved in the salvage of folate, cofactors and related molecules in eukaryotic pathogens important for metabolism and survival in their respective hosts. We identified proteins that could mediate the salvage of folates into cells and/or mitochondria from eukaryotic microbe genomes in EuPathDB. Many of these proteins are involved in folate biosynthesis or transport and are present in most of the eukaryotic microbe genomes we queried. In this study, 234 genes encoding homologues of folate salvaging proteins were identified in the genome of 64 strains, representing 28 species of eukaryotic microorganisms. Some of the pathogens among the microbes queried include P. falciparum 3D7 and IT, P. knowlesi H, P. berghei ANKA, P. chabaudi chabaudi, T. brucei Lister 427, T. brucei TREU927, T. brucei gambiensis DAL972, Encephalitozoon cuniculi GB-M1. The pathogens range from bacteria through to fungi, intracellular parasitises such as Plasmodium and Leishmania species, to extracellular parasitises such as trypanosome species. This suggests a widespread presence of the proteins cutting across a range of pathogens that infect humans and animals.

About 40% of the proteins we identified have been previously identified and characterized in parasites such as Plasmodium falciparum, Trypanosome species, Leishmania species and Toxoplasma gondii. It has been estimated that over half of the drugs currently on the market target integral membrane proteins ion channel blockers like verapamil, and serotonin transporter inhibitors feature prominently in the WHO model list of essential medicines. Unfortunately, a great number of these transporters have not been adequately explored as drug targets. Folate transporters therefore represent attractive drug targets for treatment of infectious diseases. Thus their identification from other eukaryotic pathogens could open a window for novel chemotherapeutics for disease control.

In Plasmodium two folate transporters have been identified, namely PfFT1 and PfT2. These transporters have been shown to mediate the salvage of folate derivatives and precursors in P. falciparum and proposed blocking of their salvage activities may improve the antimalarial efficacy of several classes of antimalarial drugs. In our work we identified folate transporters for other plasmodial
Figure 2. Phylogenetic tree showing the relatedness of all the proteins identified to transport folate in different pathogen species. Tree was inferred using the Neighbor-Joining method with bootstrap test at 1000 replicates.
Figure 3. Phylogenetic tree showing the relatedness of folate transporters alone in different pathogen species. Tree was inferred using the Neighbor-Joining method with bootstrap test at 1000 replicates.
Figure 4. Phylogenetic tree showing the relatedness of folate/biopterin transporter alone in different pathogen species. Tree was inferred using the Neighbor-Joining method with bootstrap test at 1000 replicates.
**Figure 5.** Phylogenetic tree showing the relatedness of folate-binding protein YgfZ alone in different pathogen species. Tree was inferred using the Neighbor-Joining method with bootstrap test at 1000 replicates.
species, which, like \textit{P. falciparum}, may also be chemotherapeutic targets. Transport of folate in higher eukaryotes is made possible by a high affinity folate-bioterin transporters FBT or BT1 family\cite{22,30}. In the trypanosomes and related kinetoplastids, a member of these transporters, the folate bioterin transporter (FBT) family of proteins was identified in \textit{Leishmania}\cite{35}. It is thought that MFS proteins are related to the FBT. These proteins have been characterized in a few protozoa and cyanobacteria\cite{46}. Results from our study describing the presence of these transporters across several phyla corroborate results from other works establishing the conservation of folate transport function among FBT family proteins from plants and protists\cite{12,48}.

Malaria parasites encode transporters belonging to the organo anion transporter (OAT) folate-bioterin transporter (FBT), glycoside-pentoside-hexuronide: cation symporter (GPH), families, which are closely related to the major facilitator super-family of membrane proteins\cite{62}. The inhibition of these transporters by blockers of organic anion transporters such as probenecid has been implicated in sensitization of \textit{Plasmodium} resistant parasites to antifolates\cite{49,50}. Thus, in \textit{Plasmodium} chemotherapy, identification of folate transporters could lead to screening for compounds that interfere with folate transport and salvage for antimalarial chemotherapy\cite{22,30}.

We identified several types of folate transporters that have been described and functionally characterized in \textit{Leishmania} with some implicated in the import of the antifolate methotrexate\cite{56,63}. Thus far, only protozoan transporters in \textit{Plasmodium}, \textit{Leishmania}, and \textit{Trypanosoma brucei} have been characterized and these are known to mediate the uptake of the vitamins folate and/or bioterin\cite{22,56,63}. Thus in parasites species of medical importance folate transporter proteins may provide new targets for therapy.

We also identified folate salvaging proteins from fungi such as \textit{Coccidioides immitis} and \textit{A. clavatus}, fungi found in soil\cite{44-46}, vegetable\cite{64} and waters in tropical and subtropical areas\cite{67}. These fungi are known to occasionally become pathogenic and act as opportunistic pathogens for animals and man\cite{66}. Coccidioidomycosis caused by \textit{C. immitis} in association with AIDS has been known to be a fatal disease\cite{68}. Treatment of acute and chronic infections with antifungals such as amphotericin B have not been adequate, hence folate transporters may present new targets in this group of pathogens. Identification was also made on pathogens such as \textit{C. fasciculata} that parasitize several species of insects including mosquitoes and has been widely used to test new therapeutic strategies against parasitic infections\cite{69}. As a model organism, folate transporters identified in \textit{C. fasciculata} may be useful in research for developing new drugs in medically important Kinetoplastids as has been shown for other targets in this protozoan parasite\cite{35}.

We noticed that \textit{P. parasitica} INRA-310 and \textit{L. pyrrochoris} H10 had the highest number of folate transporters identified. Their utility as model fungal (\textit{P. parasitica}) and monoxenous kinetoplast may provide models instrumental for developing new antifoliates for fungal and protozoan diseases. The relatedness of these proteins across the different pathogens show that there are two major

phylogenetically distinct clades in the eukaryotic pathogens examined. The clustering of these proteins suggests that these transport proteins have highly conserved regions often required for basic cellular function or stability\cite{69,71,47}. Thus, antifolate chemotherapy drugs that are effective against one pathogen might have some effect on others. However, the converse may be the case for the free-living non-parasitic photosynthetic algae, \textit{Chromera velia} and \textit{Vitrella brassicaformis}, protists related to apicomplexans\cite{40,89}.

These groups of algae live freely in their environment, which unlike apicomplexans that depend on a host animal to survive\cite{88}. This adaptation may explain the difference in the clustering of their transporters after phylogenetic analysis, which separated on the minor clade from other apicomplexans that separated on the major clade. This suggests a high level of evolutionary divergence between folate transporters in both the apicomplexans and these algae based on life-style adaptations.

**Conclusion**

In summary, we have retrieved information on 234 folate transporter proteins from Eukaryotic Pathogen Database (EuPathDB) resources. The folate transporter proteins were categorized into potential drug targeting features including mitochondrial localization, number of transmembrane helix, and protein sequence relatedness. The identification of folate salvage proteins in diverse eukaryotes extend the evolutionary diversity of these proteins and suggests they might offer new possibilities for potential drug development targeting folate-salvaging routes in eukaryotic pathogens.

**Data availability**

**Dataset 1:** Complete list of proteins extracted from Eupathdb and literature search, including their properties. These data are available in a .xlsx file. Doi, 10.5256/f1000research.10561. d167325\cite{43}

**Dataset 2:** Eukaryotic microbes from which folate transporters were identified. These data are available in a .xlsx file. Doi, 10.5256/f1000research.10561.d167326\cite{44}

**Author contributions**

M.O.F. and B.O. Conceptualized and Designed the study. M.O.F. and B.O. structured methodology. B.O. performed analysis. M.O.F. and B.O. wrote manuscript.

**Competing interests**

No competing interests were disclosed.

**Grant information**

B.O. was supported by a TWAS-CNPq fellowship (FP number: 3240274297).

*The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.*
Supplementary material

Supplementary Dataset 1: Newick format for the phylogenetic tree showing the relatedness of all the proteins identified to transport folate in different pathogen species. Tree was inferred using the Neighbor-Joining method with bootstrap test at 1000 replicates.

Click here to access the data.

Supplementary Dataset 2: Newick format for the phylogenetic tree showing the relatedness of folate transporters alone in different pathogen species. Tree was inferred using the Neighbor-Joining method with bootstrap test at 1000 replicates.

Click here to access the data.

Supplementary Dataset 3: Newick format for the phylogenetic tree showing the relatedness of folate/biotin transporter alone in different pathogen species. Tree was inferred using the Neighbor-Joining method with bootstrap test at 1000 replicates.

Click here to access the data.

Supplementary Dataset 4: Newick format for the phylogenetic tree showing the relatedness of folate-binding protein YgfZ alone in different pathogen species. Tree was inferred using the Neighbor-Joining method with bootstrap test at 1000 replicates.

Click here to access the data.

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General Comments
1. Several revisions are noted especially the datasets and the phylogenetic trees. However, the narrative contain sections that needs to be improved for accuracy.
2. Correct abbreviations. For example “EuPathdb” should be “EuPathDB”

Specific Comments. Suggested revisions are provided.

INTRODUCTION
1. The statement of objectives of the research needs to be clear. “Therefore, in an attempt to identify and characterize targets for novel therapeutics, we report herein an extensive search of folate transporters from pathogen genomes. In addition, we investigated the evolutionary relationship of these transporters in a bid to determine similarities and differences that make them attractive drug targets. The knowledge provided may assist in the design of new antifolates for protozoan parasites.”

Suggested Revision:
“The objectives of the research reported in this article were to (1) characterize folate transporters encoded in genomes of eukaryotic pathogens; and (2) determine evolutionary relationship of folate transporters in genomes of eukaryotic genomes. These research objectives will help advance the development of effective anti-folate drugs to reduce the morbidity and mortality associated with eukaryotic pathogens.”

METHODS
1. Divide the methods section according to the two objectives.
2. approximately 200-pathogens

Suggested revision: delete “-“. The purpose of the dash is not justified.
3. The folate transporters were classified based on type of transporter, number of
transmembrane helix (TMH) and localization (either cell or mitochondrial membrane) of transporter. Gene sequences were obtained in FASTA format for transporter proteins using the sequence download tool on EuPathDB (http://eupathdb.org/eupathdb/).

Suggested revision:
“The folate transporters were characterized according to type of transporter, number of transmembrane helix (TMH) and localization (either cell or mitochondrial membrane) of transporter. Gene sequences were obtained in FASTA format for transporter proteins using the sequence download tool on EuPathDB (http://eupathdb.org/eupathdb/).”

**Dataset 1**
1. Dataset 1. Complete list of proteins extracted from Euphadb and literature search, including their properties.
   Correct the spelling of EuPathDB.

**Dataset 2**
1. Actinobacillus is included in Dataset 2. Correct to Ajellomyces since the Strain/Species field is A. capsuleus G186AR. Provide the correct taxonomic lineage (http://www.uniprot.org/taxonomy/5037). Update the abstract, phylogenetic tree and discussion since no bacterial folate transport was characterized in the research. The protein sequence mislabeled clusters with Allomyces.
2. Provide the purpose of the color coding.
3. Label the eukaryotic microbe group according to the higher level classification of all living organisms e.g. fungi, protozoa and chromista.
4. To facilitate secondary analysis of Dataset 2, move legend to another sheet and remove blank rows and blank columns.

**RESULTS**
1. The results section should have subsection headings to help with readability and understanding of the article.
2. Replace “Methodological search” with “Systematic search”.
3. Replace “we came across” with “we observed”.
4. “A good number of the proteins identified had predicted transmembrane helixes with a few having none (Figure 1B).”
   Specify a quantity or percentage for the “good number”.
5. “Furthermore, a number of the transporters possess signal peptides (Dataset 1), which may be required for targeting to cellular locations. Deciphering the sequence of the targeting signal may indicate its product destination.”
   How many transporters possess the signal peptide sequence?
6. “We identified these transporters in 28 pathogen species (containing 63 strains) cutting across 12 phyla”

Suggested revision:
“We obtained information on folate transporters encoded in genomes of 63 eukaryotic microbes consisting of 26 pathogenic genera and two genera of photosynthetic algae closely related to the apicomplexan pathogens. The genera are Ajellomyces, Allomyces, Aspergillus, Chromera, Coccidioides, Crithidia, Cryptococcus, Cryptosporidium, Eimeria,”
**DISCUSSION**

1. Replace “were identified in the genome of 64 strains” with “were identified in the genomes of 63 strains”.

**References**

1. Ruggiero MA, Gordon DP, Orrell TM, Bailly N, et al.: A higher level classification of all living organisms. *PLoS One*. 2015; 10(4): e0119248 PubMed Abstract | Publisher Full Text

**Competing Interests:** No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

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**Reviewer Report 26 July 2017**

https://doi.org/10.5256/f1000research.12807.r24209

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**Gajinder Singh**

Molecular Medicine Group, International Centre for Genetic Engineering and Biotechnology (ICGEB), New Delhi, Delhi, India

I am disappointed to see that most of the issues identified in the original manuscript remain unaddressed including grammatical mistakes. e.g.

1. The current literature on folate transporters as drug targets has not been discussed, including information on their essentiality.

2. Using keyword and BLAST based searches will miss and misidentify folate transporters, thus more sensitive HMM profile based methods should be employed or please provide a justification for not doing the same.

3. Many sentences identified as unclear in the previous report remain unchanged.

I would request authors to provide a point by point response to my previous comments. I would be very happy to reconsider their response.
Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

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**Version 1**

Reviewer Report 06 March 2017

https://doi.org/10.5256/f1000research.11380.r20419

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Gajinder Singh
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Below are my major concerns:

1. The abstract does not provides an adequate summary of the article.

   The authors have claimed much higher scope of their work than actually reported. While their work is restricted to folate transporters, they have claimed to work on whole folate pathway. In the Abstract: "We applied a combination of bioinformatics methods to examine the genomes of pathogens in the EupathDB for genes encoding homologues of proteins that mediate folate salvage in a bid to identify and assign putative functions." "These findings offer new possibilities for potential drugConclusion: development targeting folate-salvage proteins."

2. No proper justification for the work is provided.

   a) While folate pathway is well established as drug target, the authors have only identified folate transporters. So please give examples of drugs (with names) which are known to target folate transporters in any organism. If no sufficient information is provided, the usefulness of the work is severely reduced.

   b) How many of folate transporters are essential in species where essentiality data is available such as P. berghei and T. gondii?

3. The methodology to identify transporters is not comprehensive.

   Since authors have used key-word searches to identify folate transporters, they are likely to miss many transporters not labelled as such. An appropriate methodology will incorporate
profile (such as HMM) based searches. Thus the number of transporters identified by authors is most likely to be an underestimate.

4. The methodology is not clear.

Authors write that "we utilized the word “folate” for search on the gene text and “folic acid” was used to confirm the hits", then how only transporters were retrieved? The Figure 1 is confusing. Where is BLAST used here?

5. The manuscript is written very poorly with so many scientific and grammatical mistakes that it is very difficult for the reader to follow the manuscript. Below are some examples.

a) "The mitochondrion is the predicted location of the majority of the proteins, with 15% possessing signal peptides." - how can mitochondria be majority location if only 15% have signal peptides and even less with mitochondrial signal peptide? Shouldn't the majority then be cytoplasmic?

b) "We identified 234 proteins to be involved in folate transport".

c) "Since folate-binding protein YgfZ, folate/pteridine transporter, folate/biopterin transporter, putative, reduced folate carrier family protein, folate/methotrexate transporter FT1, putative folate transporters alone and others have 10, 25, 132, 2, 7, 49 and 9." What are these numbers?

d) "So we decided to reconstruct the phylogeny based folate transporter, folate-biopterin transporter after considering the identification number, the species diversity in each category."

e) "The different proteins identified to be involved in folate salvage or related molecules were folate-binding protein YgfZ, folate/pteridine transporter, folate/biopterin transporter, reduced folate carrier family protein, folate/methotrexate transporter FT1 and folate transporters having a 4%, 11%, 56%, 1%, 3% and 21% identity, respectively." What does this statement mean?

f) Does Table 1 really need to be 4 page long?

g) "The only Plasmodium species with results for proteins that salvage folate was P. falciparum"

h) "However, folate transporters I and II were retrieved from our search of GeneDB for P. malariae and P. ovale curtisi, respectively." What are these transporter classes?

i) "Some of these pathogens include P. ultimum DAOM BR144, which has mitochondrial folate transporter/carrier proteins similar to Homo sapiens, E. cuniculi GB-M1, which has proteins similar to folate transporter, and S. punctatus DAOM BR117, which has folate-binding protein YgfZ."

j) "After phylogenetic analysis each sub-phylogeny show a clear characterization except for
folate-biopterin transporters

h) "In this study, 234 genes encoding homologues of folate salvaging proteins were identified in the genome of 64 strains, representing 28 species of eukaryotic pathogens. Some of the pathogens include P. falciparum 3D7 and IT, P. knowlesi H, P. berghei ANKA, P. chabaudi chabaudi, T. brucei Lister 427, T. brucei TREU927, T. brucei gambiense DAL972, Encephalitozoon cuniculi GB-M1. The pathogens range from bacteria through to fungi, intracellular parasites such as Plasmodium and leishmania species, to extracellular parasites such as trypanosome species" Which bacteria was included in the study?

i) "It has been estimated that over half of the drugs currently on the market target integral membrane proteins of which membrane transporters are a part, but unfortunately, these transporters have not been adequately explored as drug targets. Folate transporters therefore represent attractive drug targets for treatment of infectious diseases." Please tell us how many drugs are available in the market which target folate transporters, which is a more relevant statistic with respect to this study.

j) "In the trypanosomes and related kinetoplastids, a member of these transporters, the folate biopterin transporter (FBT) family of proteins was identified in Leishmania."

k) "It is thought that MFS proteins are related to the FBT." What is MFS?

l) "Results from our study describing the presence of these transporters across several phyla corroborate results other researches, establishing the conservation of folate transport function among FBT family proteins from species from plants and protists".

m) "The clustering of these proteins suggests that these transport proteins have highly conserved regions often required for basic cellular function or stability". The clustering does not suggest that these transport proteins have highly conserved regions often required for basic cellular function or stability.

n) " We also performed phylogenetic comparisons of identified proteins. .".

**Competing Interests:** No competing interests were disclosed.

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.**
Summary of Referee's Report
The manuscript presents a strong justification for research on folate transporter proteins as drug targets for diseases caused by eukaryotic pathogens including the malaria parasite. The manuscript reports a data curation effort that involves the use of the Eukaryotic Pathogens Database (EuPathDB) Resource. Several novel results to guide future research are included such as (i) a list of 234 folate transporter proteins from 63 eukaryotic microbes including eukaryotic pathogens; and (ii) phylogenetic trees of relatedness of the protein sequences. The authors observed the clustering of the protein sequences that indicate the possibility that antifolate drugs could be effective for multiple eukaryotic pathogens.

Major concerns are:
(i) The need for a clearer description of the workflow for the construction of the protein list.
(ii) There is inadequate support for the statement that “60% of the proteins were identified for the first time”.
(iii) Confusion between retrieval and identification of protein sequences. The workflow diagram indicates retrieval of sequences but narrative text describes identification in multiple sections. The potential drug targeting categorization of the retrieved protein sequences is a key contribution of the research study.

Some minor concerns include (i) typographic errors such as spelling of abbreviations (e.g. EuPathDB, PubMed and UniProt); (ii) classification of *A. (Ajellomyces) capsulatus* G186AR as a bacteria; and (iii) need to quantify the statistics associated with observations (Examples: in abstract “Many of the genomes”; in discussion: “A few of the proteins” ).

Title and Abstract:
*Is the title appropriate for the content of the article?*
The manuscript title is “Genome-wide characterization of folate transporter proteins of eukaryotic pathogens”. “Genome-wide characterization” does not effectively describe the accomplishments of the research reported. The manuscript in the conclusion section (Page 15) states “we have identified and classified 234 proteins...”. The workflow (Figure 1) provides categorization of proteins by features such as cellular location, presence of signal peptide and number of transmembrane helices. Figure 2 has the title “Categorization of proteins identified”. A suggested revised title is “Categorization of potential drug targeting folate transporter proteins from eukaryotic pathogens”. The “potential drug targeting” is obtained from the conclusion section.

*Does the abstract represent a suitable summary of the work?*
There are sentences in the abstract that should be revised to accurately represent a suitable summary of the research performed. Comments/suggestions are provided below.
1. Quantify the observations described as many. For example “genome sequences of many”: How many?
2. “eukaryotic protozoa” > “eukaryotic microbes”. The term “eukaryotic microbes” encompasses pathogens and non-pathogens (e.g. *Chromera velia* and *Vitrella brassicaformis*).
3. “important data” > “critical biological information”

4. “pathway are important for” > “pathways are necessary for”

5. “Methods: We applied a combination of bioinformatics methods to examine the genomes of pathogens in the EupathDB for genes encoding homologues of proteins that mediate folate salvage in a bid to identify and assign putative functions.” > “Methods: We developed automated search strategies in the Eukaryotic Pathogen Database Resources (EuPathDB) to construct a protein list and retrieve protein sequences of folate transporters encoded in the genomes of 200 eukaryotic microbes. The folate transporters were categorized according to features including mitochondrial localization, number of transmembrane helix, and protein sequence relatedness.

6. Provide key result(s) of the protein list retrieval and phylogenetic comparison of the retrieved proteins. For example, We constructed a list of 234 folate transporter proteins associated with 63 eukaryotic microbes including ??? algae, ??? fungi and ??? protozoa. Seven percent of the proteins were predicted to localize on the mitochondrial membrane. Phylogenetic tree revealed major (??? proteins) and minor (??? Proteins) clades. All the folate transporter sequences from the malaria parasite, *Plasmodium*, belonged to the major clade.

7. “The mitochondrion is the predicted location of the majority of the proteins”. This statement is not supported by Figure 2D, where 7% of the protein sequences are labelled as Mitochondrial folate transporters.

**Article content:**
*Have the design, methods and analysis of the results from the study been explained and are they appropriate for the topic being studied?*

**Design and Methods:**
1. Figure 1 presents a conceptual hierarchical methodology. The rectangle labelled “Protein names/sequences verification” has arrows to PubMed, UniProt, Membranetransporters.org, NCBI, GeneDB, Google Scholar and Phylogenetic analyses. It appears that the integrated results from the search strategies in the databases provided the input for the phylogenetic analyses. Please clarify.

2. Which step of the workflow resulted in the list of 234 proteins?

3. How many proteins were retrieved from the initial search using EuPathDB?

4. Protein Features Retrieved rectangle: Was the retrieval of protein features performed on only the 234 proteins?

5. There is adequate explanation of the methods for phylogenetic analyses. Please provide the Newick format phylogenetic tree as a supplementary dataset.

**Analysis of the Results:**
1. Table 1 is a major curation effort presented in the manuscript.
a. The title of Table 1 should be updated to “Eukaryotic microbes from which folate transporters were identified”. The list includes non-pathogens.
b. The content of the table (especially the Kingdom entries) should be checked for accuracy. The Kingdom column could be updated to Eukaryotic Microbe Group with entries as algae, protozoa or fungi.
c. The column entries for *A. capsulatus* G186AR (mislabelled as bacteria) should be updated as the organism name is for a fungus (genus *Ajellomyces*). This update will also affect the Phylogenetic Tree (Figure 3). The node labelled *Actinobacillus* clusters with *Ajellomyces macrogynus*.
d. An updated Table 1 should be presented as Dataset 2 in a spreadsheet file. This would enable secondary data analysis by other researchers.
e. A new Table 1 could consist of columns for Eukaryotic Microbe Group, Genera of Eukaryotic Microbe, List of Species/Strain and Number of Folate Transport Proteins. This will provide reader with an overview of how the 234 proteins is distributed into the genera of the eukaryotic microbes.
f. References listed for confirmation searches. Page 8, Paragraph 1, Sentence 1: “Our literature search for parasite folate transporters on PubMed and Google Scholar indicated 60% (38 out 63) of the proteins were identified for the first time as presented in Table 1. Comment: Among the references included in Table 1, only eight references (22, 73 to 77 and 82) on the basis of the article title provide experimental assessments of the folate transporters. Reference 85 is a reference for MEGA7 software. In the sentence “proteins” should be eukaryotic microbes. The proportion of eukaryotic microbes whose folate transporter(s) have been previously investigated with functional assays should be revised.

2. Figure 2. Categorization of proteins identified.
Authors should consider representing Figure 2A and 2B as bar graphs. Figures 2C, 2D and 2E have only two categories that can be described in the Results section.

3. Discussion
a. “genomes of 64 strains”. Table 1 has 63 eukaryotic microbes.
b. Discuss *Chromera velia* and *Vitrella brassicaformis* as organismal systems for investigating folate transporter function. See Woo et al.¹

4. Conclusion
Consider revising “In summary, we have identified and classified 234 proteins...” to “In summary, we have retrieved information on 234 folate transporter proteins from the Eukaryotic Pathogen Database (EuPathDB) resources. The folate transporter proteins were categorized into potential drug targeting features including mitochondrial localization, number of transmembrane helix, and protein sequence relatedness.”

**Data (if applicable):**
Has enough information been provided to be able to replicate the experiment? Are the data in a usable format/structure and have all the data been provided?
1. Table 1 needs to be revised and converted to a Dataset.
2. Please provide the Newick format phylogenetic tree as a supplementary dataset.
3. The Gene Identifiers [Gene ID] in Dataset 1 can be used to retrieve the protein sequences
from EuPathDB.

References
1. Woo YH, Ansari H, Otto TD, Klinger CM, et al.: Chromerid genomes reveal the evolutionary path from photosynthetic algae to obligate intracellular parasites. *Elife*. 2015; 4: e06974 PubMed

*Abstract* | *Publisher Full Text*

**Competing Interests:** No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

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