Domestic Exposure to Fungi and Total Serum IgE Levels in Asthmatic Children

Huey-Jen Jenny Su,1 Pei-Chih Wu,1 Huan-Yao Lei,2 and Jiu-Yao Wang3

1Department of Environmental and Occupational Health, Medical College, National Cheng Kung University, 138 Sheng Li Road, 70428 Tainan, Taiwan
2Department of Microbiology and Immunology, Medical College, National Cheng Kung University, 138 Sheng Li Road, 70428 Tainan, Taiwan
3Department of Pediatrics, Medical College, National Cheng Kung University, 138 Sheng Li Road, 70428 Tainan, Taiwan

Received 9 March 2005; accepted 21 March 2005

We measured the number of airborne, viable fungi and house dust mite (HDM) allergen levels in the homes of a group of asthmatic children. Blood samples were drawn and the amounts of total and specific serum IgE were determined. The association between the number of fungal colonies, dust mite allergen exposure, and specific and total IgE was evaluated. The number of viable airborne fungi was high (20 543 CFU/m3) in those investigated houses. Der p1 concentrations on child’s mattress exceeding 2 µg/g were found in 78.6% of the houses. A quantitative dose-response relationship was demonstrated between the exposure to viable, airborne molds and the amount of total IgE ($r = 0.4399$ and $P = .0249$) and the level was further increased in children with coexposure to viable fungi and HDM.

INTRODUCTION

Fungi are part of the normal environment but exposure levels much over normal may occur indoors in buildings with humidity problems, either due to climate or due to local water damage [1, 2]. Several studies have reported that children living in damp and moldy dwellings have a larger prevalence of respiratory symptoms, such as cough, phlegm, wheeze, sore throat and runny nose, as well as asthma [3, 4]. There are however only limited data demonstrating dose relationships between mold exposure and the extent of symptoms or clinical markers of inflammation or asthma. In one field study a relation was reported between the extent of upper respiratory symptoms and the exposure to molds, expressed as the amount of airborne (1 → 3)-β-D-glucan [5].

Taiwan is located in a subtropical region with optimal environmental characteristics, such as high temperature and relative humidity that sustain the growth of molds and indoor aeroallergens. Our previous work demonstrated that 65% of children’s mattresses in Taiwan’s homes had Der p1 allergen levels higher than that 2 µg/g, a critical level suggested for atopic sensitization [6, 7].

In this study, we measured the number of airborne, viable fungi and house dust mite (HDM) allergen levels in the homes of a group of asthmatic children. Blood samples were drawn and the amounts of total and specific serum IgE were determined. The association between the number of fungal colonies, dust mite allergen exposure, and specific and total IgE was evaluated.

MATERIALS AND METHODS

Study subjects

A randomly selected subgroup of asthmatic children was chosen from our previous environmental survey of domestic exposures to dust mite allergens and airborne fungi in a metropolitan city of southern Taiwan [7]. A total of 28 subjects, aged 10–12 years, were invited through phone contact for clinical confirmation conducted by participating pediatricians for their asthmatic status and blood samples were drawn. The clinical visits took place in winter (December, January, and February) at the same time as the environmental measurements.

Airborne fungi sampling and identification

Airborne fungal concentrations were measured in the subject’s bedroom once a month during the season. Duplicate samples of airborne fungi were collected using Burkard sampler, a single-stage hole-to-agar impactor with 100 holes (Model PASA/B, Burkard Mfg Co, Rickmansworth, UK) with Malt Extract Agar plates...
(MEA) at a flow rate of 10 LPM. After field sampling, the plates were incubated at 25°C for 5 days.

The yields of colonies were corrected by using positive-hole conversion table before the calculation of final concentrations, colony forming unit per cubic meter (CFU/m³). The average concentrations of monthly measurements (December, January, and February) were used to express the airborne fungal exposure levels.

**House dust sampling and Der p 1 analysis**

Dust samples from the child’s mattress were collected by a hand-held vacuum cleaner (National MC 33000) with a glass fiber filter and collected concurrently along the airborne samples. Samples were collected by vacuuming for 3 minutes on 1 m² of the study area. After the sampling, the samples were sealed with parafilm and stored at 4°C before handling. After weighting, all samples were stored with desiccating agents until extraction. Samples were expressed in µg/g dust by weighing for 3 minutes on 1 m² of the study area. After the sampling, the samples were sealed with parafilm and stored at 4°C before handling. After weighting, all samples were stored with desiccating agents until extraction. Samples were expressed in µg/g dust.

**Total serum IgE and specific IgE measurement**

Total serum IgE and specific IgE antibodies to Dermatophagoides pteronyssinus, Cladosporium herbarum, and Aspergillus fumigatus were measured by a quantitative fluoroimmunoassay (CAP-FEIA; Pharmacia Up&John, Sweden, detection limit 0.35 kU/L for total IgE and specific IgE). The standard curve was examined in triplicate samples and within the range of 10% of coefficient of variation. Children were considered as sensitized if the IgE concentrations exceeded 0.35 kU/L.

**RESULTS**

The number of viable airborne fungi was high (medium: 19359, 25th–75th quartile: 15350–24525 CFU/m³) in those investigated houses. The predominant indoor genera were Cladosporium, Aspergillus, Penicillium, Alternaria, nonsporulating fungi and yeast. Cladosporium accounted for almost 85% of the colonies from all indoor samples and was present in high amounts (17691 CFU/m³). Der p1 concentrations on child’s mattress exceeding 2 µg/g were found in 78.6% of the houses. The average concentration of Der p1 was 6.72 µg/g (Table 1). No significant relationship was found between levels of Der p1 and viable fungi (r = 0.13).

Among the 28 children, 23 were found to sensitiz to HDM. One child was sensitiz to C herbarum allergen, and two to A fumigatus extract; all three of them were also HDM-sensitiz. Total serum IgE levels ranged from 18.8 to 2000.0 kU/L, and the average concentration was 772.3 kU/L.

As illustrated in Figure 1, there was a positive and significant correlation between the number of airborne fungi and the amount of total IgE (r = 0.4399 and P = 0.0249); yet, such a relationship could not be detected between Der p1 levels and total IgE. Study subjects were grouped by the median of viable fungi and Der p1 concentrations, respectively. Logistic regression identified that the only significant relationship was between the dichotomized level of viable fungi and IgE with OR = 5.06 (95% CI: 1.02–30.06) in univariant analysis. Moreover, an increased OR of 6.67 (95% CI: 1.19–55.16) between the exposure and response, and the IgE levels, was found when the exposures to airborne fungi and dust Der p1 were concurrently examined in a multivariate analysis.

**DISCUSSION**

Although the sample size was relatively small in this pilot investigation, the sample was selected randomly from the general asthmatic population in the study city. Therefore, these results still may reflect the real phenomenon of asthmatic population in this region.

The major finding in the study was the relation between the number of viable fungi and the total amount of IgE in serum. Similar findings are reported from a study conducted in a mold-contaminated school building where elevated IgE levels were significantly more common among children in mold-contaminated buildings [9]. Another study was performed on persons living in Sweden, detection limit 0.35 kU/L for total IgE and specific IgE. The standard curve was examined in triplicate samples and within the range of 10% of coefficient of variation. Children were considered as sensitized if the IgE concentrations exceeded 0.35 kU/L.

**RESULTS**

The number of viable airborne fungi was high (medium: 19359, 25th–75th quartile: 15350–24525 CFU/m³) in those investigated houses. The predominant indoor genera were Cladosporium, Aspergillus, Penicillium, Alternaria, nonsporulating fungi and yeast. Cladosporium accounted for almost 85% of the colonies from all indoor samples and was present in high amounts (17691 CFU/m³). Der p1 concentrations on child’s mattress exceeding 2 µg/g were found in 78.6% of the houses. The average concentration of Der p1 was 6.72 µg/g (Table 1). No significant relationship was found between levels of Der p1 and viable fungi (r = 0.13).

Among the 28 children, 23 were found to sensitiz to HDM. One child was sensitiz to C herbarum allergen, and two to A fumigatus extract; all three of them were also HDM-sensitiz. Total serum IgE levels ranged from 18.8 to 2000.0 kU/L, and the average concentration was 772.3 kU/L.

As illustrated in Figure 1, there was a positive and significant correlation between the number of airborne fungi and the amount of total IgE (r = 0.4399 and P = 0.0249); yet, such a relationship could not be detected between Der p1 levels and total IgE. Study subjects were grouped by the median of viable fungi and Der p1 concentrations, respectively. Logistic regression identified that the only significant relationship was between the dichotomized level of viable fungi and IgE with OR = 5.06 (95% CI: 1.02–30.06) in univariant analysis. Moreover, an increased OR of 6.67 (95% CI: 1.19–55.16) between the exposure and response, and the IgE levels, was found when the exposures to airborne fungi and dust Der p1 were concurrently examined in a multivariate analysis.

**DISCUSSION**

Although the sample size was relatively small in this pilot investigation, the sample was selected randomly from the general asthmatic population in the study city. Therefore, these results still may reflect the real phenomenon of asthmatic population in this region.

The major finding in the study was the relation between the number of viable fungi and the total amount of IgE in serum. Similar findings are reported from a study conducted in a mold-contaminated school building where elevated IgE levels were significantly more common among children in mold-contaminated buildings [9]. Another study was performed on persons living in...
houses with mold problems, using the mold cell wall agent (1→3)-β-D-glucan as a marker for mold cell biomass. Total IgE levels were dose-related to the amount of (1→3)-β-D-glucan in the homes, independent of atopic sensitization (Beijer et al, personal communication).

The presence of a relation between mold exposure and total IgE increase and the very low prevalence of sensitization to mold among the children studied suggests that a major effect of mold exposure is a general stimulation of the IgE production without specific sensitization. An increased level of total IgE on the other hand implies a larger risk for sensitization to other agents, as evidenced by the high prevalence of sensitization to other antigens in general and possibly also increases the sensitivity to inflammagenic agents.

ACKNOWLEDGMENTS

The authors wish to thank Ms Chu-Yun Huang and Ms Fang-Chun Lee for their friendly and laboratory assistance during the study. Most of all, our appreciation goes to all participants and their families for the patience and cooperation. We are eternally in great debt to this support. This study was supported in part by a grant from the Taiwan National Science Council, NSC 86-2621-P-006-013.

REFERENCES

[1] Garrett MH, Rayment PR, Hooper MA, Abramson MJ, Hooper BM. Indoor airborne fungal spores, house dampness and associations with environmental factors and respiratory health in children. Clin Exp Allergy. 1998;28(4):459–467.

[2] Verhoeven AP, van Wijnen JH, Brunekreef B, Fischer P, van Reenen-Hoekstra ES, Samson RA. Presence of viable mould propagules in indoor air in relation to house damp and outdoor air. Allergy. 1992;47(pt 1):83–91.

[3] Hu FB, Persky V, Flay BR, Richardson J. An epidemiological study of asthma prevalence and related factors among young adults. J Asthma. 1997;34(1):67–76.

[4] Cuipers CE, Swaen GM, Wesseling G, Sturmans F, Wouters EF. Adverse effects of the indoor environment on respiratory health in primary school children. Environ Res. 1995;68(1):11–23.

[5] Rylander R, Persson K, Goto H, Yuasa K, Tanaka S. Airborne Beta-1,3-glucan may be related to symptoms in sick buildings. Indoor Environment. 1992;1:263–267.

[6] Su HJ, Wu PC, Chen HL, Lee FC, Lin LL. Exposure assessment of indoor allergens, endotoxin, and airborne fungi for homes in southern Taiwan. Environ Res. 2001;85(2):135–144.

[7] Platts-Mills TA, Vervloet D, Thomas WR, Aalberse RC, Chapman MD. Indoor allergens and asthma: report of the third international workshop. J Allergy Clin Immunol. 1997;100(pt 1):S2–S24.

[8] Luczynska CM, Arruda LK, Platts-Mills TA, Miller JD, Lopez M, Chapman MD. A two-site monoclonal antibody ELISA for the quantification of the major Dermatophagoides spp allergens, Der p I and Der f I. J Immunol Methods. 1989;118(2):227–235.

[9] Savilathi R, Uitti J, Roto P, Liappala P, Husman T. Increased prevalence of atopy among children exposed to mold in a school building. Allergy. 2001;56(2):175–179.
[10] Pollart SM, Chapman MD, Fiocco GP, Rose G, Platts-Mills TA. Epidemiology of acute asthma: IgE antibodies to common inhalant allergens as a risk factor for emergency room visits. *J Allergy Clin Immunol.* 1989;83(5):875–882.

[11] Sarsfield JK. Environmental, clinical, and immunological study of house dust mite in childhood asthma. *Arch Dis Child.* 1973;48(8):655.

[12] Tang RB, Tsai LC, Hwang HM, Hwang B, Wu KG, Hung MW. The prevalence of allergic disease and IgE antibodies to house dust mite in schoolchildren in Taiwan. *Clin Exp Allergy.* 1990;20(1):33–38.

[13] Rylander R, Fogelmark B. Inflammatory responses by inhalation of endotoxin and (1→3)-beta-D-glucan. *Am Ind Med.* 1994;25(1):101–102.

[14] Beijer L, Thorn J, Rylander R. Effects after inhalation of (1→3)-beta-D-glucan and relation to mould exposure in the home. *Mediators Inflamm.* 2002;11(3):149–153.