Fifty million years of beetle evolution along the Antarctic Polar Front

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Abstract

The hypothesis that diversification has proceeded similarly in Antarctic marine and terrestrial groups has not been tested. While the extinction of a diverse continental Antarctic biota is well established (13), mounting evidence of significant and biogeographically structured Antarctic terrestrial diversity (8, 14, 15) with a long evolutionary history (6, 16) suggests the possibility of broadly similar diversification processes across marine and terrestrial Antarctic systems. If valid for some taxa, further tests should then be sought across a wider variety of organisms. Here, we therefore evaluate the terrestrial applicability of the paradigm emerging for Antarctic marine biodiversity—that a major cooling phase from the mid-Miocene climatic transition (14 Ma) onwards, and subsequent habitat restructuring, have led to significant and ongoing diversification for many taxa, including those with much older origins in the region (2, 4, 17). We do so by using one of the most well-known and speciose groups from the sub-Antarctic, the herbivorous Ectemnorhinini weevils (Coleoptera: Curculionidae) (18–20).

Preliminary molecular studies indicate that the Ectemnorhinini, along with numerous other terrestrial taxa, have long histories in Antarctica’s paleoclimate and glaciation cycles since Antarctica’s isolation. Here, we demonstrate that these weevils colonized the sub-Antarctic islands from Africa at least 50 Ma ago and repeatedly dispersed among them. As the climate cooled from the mid-Miocene, warming and cooling periods resulted in the remarkable diversification of a unique marine fauna (1, 2). The investigation of marine evolutionary histories demonstrates the potential of a general biodiversity paradigm that should be sought broadly for the region’s taxa.

Significance

The Antarctic environment is famously inhospitable to most terrestrial biodiversity, traditionally viewed as a driver of species extinction. Combining population- and species-level molecular data, we show that beetles on islands along the Antarctic Polar Front diversified in response to major climatic events over the last 50 Ma in surprising synchrony with the region’s marine organisms. Unique algae- and moss-feeding habits enabled beetles to capitalize on cooling conditions, which resulted in a decline in flowering plants—the typical hosts for beetles elsewhere. Antarc-tica’s cooling paleoclimate thus fostered the diversification of both terrestrial and marine life. Climatically driven evolutionary processes since the Miocene may underpin much of the region’s diversity, are still ongoing, and should be further investigated among Antarctic biota.

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The authors declare no competing interest.

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3On the first page of the article, "Antarctica’s isolation, cooling, and glacial–interglacial cycles since the Cenozoic have resulted in the remarkable diversification of a unique marine fauna (1, 2). The investigation of marine radiations in Antarctica has reshaped modern understanding of biodiversity processes, for example, by revealing a surprising inverse relationship between diversity and latitude (3–5). In contrast, Antarctica’s paleoclimatic legacy for terrestrial communities has long been considered one of widespread extinction due to glaciation. Evidence of terrestrial species surviving in Antarctic glacial refugia (6) and discoveries of substantial endemic diversity and biogeographic structuring in some groups (7, 8) is changing this narrative, indicating extended evolutionary histories on land. Yet, such evolutionary histories remain obscured by a lack of large-scale molecular phylogenetic work, with most Antarctic terrestrial research focused on small subsets of species or populations (9, 10). The few studies that have taken a multilocus phylogenetic approach have uncovered hidden terrestrial diversity and signals of long-term allopatric divergence (e.g., refs. 11 and 12), hinting that Cenozoic climatic processes may have driven terrestrial diversification in ways similar to that for marine life.

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the sub-Antarctic, extending to the Miocene or earlier [e.g., beetles (21), midges (22), and springtails (11)]. This enables a comparison of their evolution throughout the same periods of environmental change that drove the diversification of Antarctic marine taxa. Moreover, the sub-Antarctic islands overlap spatially with the Southern Ocean, with climates that reflect oceanic conditions both past and present (23, 24). While in some respects quite different to the continental Antarctic, the islands are in other ways quite similar, providing a window into diversification processes that might be sought for continental groups, especially given their age and biogeographic structuring. Both regions share many higher taxa (e.g., ref. 25), a dynamic geo-climatic history (6, 26), a profound degree of isolation, and indications that climatic events likely structured their biota (6, 8, 27). The terrestrial habitat on the continent and its surrounding islands is fragmented by large expanses of ice or ocean, respectively, and has been further isolated by the Antarctic Circumpolar Current for at least 34 Ma (28, 29). Cyclic growth and contraction of ice sheets throughout the Plio–Pleistocene, though typically associated with the continent, has also had extensive impacts on the sub-Antarctic islands (26). The more intensively surveyed sub-Antarctic faunas thus provide an opportunity to investigate terrestrial diversification processes for the wider Antarctic while recognizing that for many groups on the continent, the main legacy of change has been extinction. To test the hypothesis that a major phase of cooling from the mid-Miocene onwards and subsequent habitat restructuring has led to the diversification of Antarctic terrestrial taxa, we integrate three tiers of molecular data to reveal a comprehensive evolutionary history for the Ectemnorhinini weevils. This additionally allows us to resolve the geographic, taxonomic, and temporal origins of the Ectemnorhinini and the role of dispersal and colonization in the development of the region’s biogeography. We first resolve the controversial origins of these weevils (19, 30) with a phylogenomic approach using anchored hybrid enrichment (AHE) for up to 515 genes across 12 representative species of Ectemnorhinini and a worldwide sample of 87 species of putative relatives and known outgroups, mostly from the beetle subfamily Entiminae (18, 30, 31). We then build on these outcomes by exploring the timing and patterns of taxonomic diversification, including divergence times and proposed dispersal events, using a multilocus phylogenetic dataset (three mitochondrial and two nuclear genes) for an extensive sample of Ectemnorhinini in which each is sequenced at every position of which they are known to occur. Finally, we reveal contemporary limits to gene flow and examine the population structure of the littoral-dwelling ectemnorhinine weevil Palirhoeus eatoni using phylogeographic methods applied to a library of 5,859 genome-wide single-nucleotide polymorphisms (SNPs). This unusually widespread species is found on all four archipelagos of the Kerguelen Province known to host Ectemnorhinini: Crozet, Kerguelen, Prince Edward Islands (PEI), and Heard Island and McDonald Islands (HIMI).

Results

Phylogenomics. We generated a fossil-calibrated Bayesian timescale for 12 Ectemnorhinini species and 87 phylogenetically relevant taxa from all major biogeographic regions worldwide. Our analyses were based on data from the first and second codon positions of 515 genes (111,294 base pairs [bp] total) recovered by AHE. The resulting timetree revealed that Ectemnorhinini weevils diverged from their most recent common ancestor with other weevils ~55 Ma ago (Fig. 1, 95% highest posterior density [HPD]: 44 to 65 Ma; SI Appendix, Fig. S1). The closest relatives of Ectemnorhinini are three African genera, Syntaphocerus, Disker, and Syntaphocerini (Fig. 1), the latter two representing the African tribe Embritini and the first representing an African group often confused with the tribe Peritelini. Marginal likelihood estimates obtained via stepping stone analysis were used to test alternative hypotheses for the nearest extant relatives of Ectemnorhinini. These analyses provided additional support for a sister relationship with the aforementioned African genera (SI Appendix, Text S1 and Figs. S2 and S3). Tribes previously proposed to be the nearest living relatives of Ectemnorhinini based on morphology, such as the Phyllobiini (30) and the Leptopiini (18), represented in our analysis by Phyllobius and Leptopus, were not found to be close relatives (Fig. 1 and SI Appendix, Figs. S2 and S3). However, in agreement with morphological hypotheses, Christensenia from the Crozet archipelago was recovered as the most basal genus of the Ectemnorhinini (18), estimated to have diverged from the rest of the tribe in the Eocene (95% HPD: 37 to 57 Ma; SI Appendix, Fig. S1), followed by the monospecific genera Canonopsis and Disker. The two most diverse clades of Ectemnorhinini, the Ectemnorhinini and the Palirhoeus–Bothrometopus groups, diverged in the Miocene (Fig. 1).

Phylogenetics. A broader taxon sample of 24 Ectemnorhinini species (two-thirds of known species in the group) was sequenced for five genes (2,812 bp total), with phylogenetic analysis resulting in congruent Bayesian and maximum likelihood phylogenies that challenge traditionally recognized diversity in the tribe (SI Appendix, Fig. S4). In the molecular phylogenies, distinct clades were evident for island populations of some morphologically determined species, whereas other morphologically determined species failed to show clear delineation; we therefore proposed molecular operational taxonomic units (MOTUs) based on these phylogenies (SI Appendix, Designation of Molecular Operational Taxonomic Units (MOTUs)). The phylogenetic trees were generally well supported, although some nodes in the genera Bothrometopus and Ectemnorhinus had relatively low support, possibly reflecting incomplete lineage sorting due to recent radiation (32) (SI Appendix, Fig. S4). Bayesian timescales calibrated using the geological emergence of the PEI archipelago had largely congruent topologies and date estimates, whether individual gene trees were linked in BEAST (Bayesian Evolutionary Analysis Sampling Trees) or coestimated within a shared species tree in starBEAST and whether or not secondary constraints from the phylogenomic tree were applied (SI Appendix, Figs. S5–S7). These trees were also reasonably concordant with the phylogenomic timetree, with error bars for most key divergence dates overlapping. However, date estimates obtained via the phylogenomic analyses were typically somewhat older (SI Appendix, Supplementary phylogenomic results), and unsampled Ectemnorhinini species could potentially affect these estimates.

The crown age estimate for Ectemnorhinini based on the most thoroughly calibrated BEAST timetree was 38 Ma (Fig. 2, 95% HPD: 29 to 48 Ma; SI Appendix, Fig. S5), which is compatible with maximum age estimates for subaerial land on the Kerguelen Plateau (26), although divergence of the group from its African relatives necessarily occurred earlier than this (Fig. 2). The divergence of the two species-rich clades of Ectemnorhinini, each of which broadly represents a feeding niche (cryptogams versus angiosperms) (33), occurred in the mid-Miocene (14 Ma, 95% HPD: 9 to 19 Ma; SI Appendix, Fig. S5), with most of the diversification in these groups (at least of extant and sampled species) occurring throughout the Pliocene and Pleistocene (Fig. 2). Estimation of diversification rates in BAMM (Bayesian Analysis of Macroevolutionary Mixtures) identified a single significant rate shift (increase) occurring in the Miocene just before the crown divergence of these species-rich clades (Fig. 2 and SI Appendix, Diversification analyses). Furthermore, there was substantial evidence of an inverse relationship between BAMM-generated diversification rates and paleoclimate data (an increase in diversification with decreasing temperature) based on Pearson’s correlations and, more importantly, detrended cross-correlation analysis (Fig. 2). This correlation was significant whether temperature trends were smoothed over either million-year or 20,000-y windows (Fig. 2, cf. SI Appendix, Fig. S9). Furthermore, in comparing among constant-rate, time-dependent, and temperature-dependent diversification models, RPANDA (Phylogenetic Analyses of
Diversification) allocated the highest support to the model in which speciation rate increases exponentially with decreasing temperature, although other diversification processes may also have played a role (SI Appendix, Table S3).

Ancestral range reconstruction using a range of biogeographic models consistently designated the older archipelagos of Crozet or Kerguelen (or both) as the likely range for ectemnorhinine early ancestors, with Crozet remaining the likely ancestral range at most nodes, even after the emergence of Heard Island and the PEI (dispersal–extinction–cladogenesis [DEC]+J+X model: Fig. 2; simpler models: SI Appendix, Figs. S10 and S11). Biogeographic stochastic mapping (BSM) thus unsurprisingly inferred a high degree of speciation in, and dispersal from, the Crozet archipelago (Fig. 2 and SI Appendix, Historical biogeography inference). The estimate for parameter “X” (a distance-based modifier of dispersal) was ~2 (SI Appendix, Table S4), indicating an inverse square function of dispersal probability with increasing geographic distance as commonly predicted by ecological theory (34). Nonetheless, at least 25 interarchipelago dispersal events (mostly reflecting anagenetic range expansion: SI Appendix, Fig. S12) were estimated throughout the phylogeny by BSM, the majority of which occurred in an east–west direction against the prevailing winds and currents of the Antarctic Circumpolar Current (Fig. 2; and for a comparison of all models, SI Appendix, Tables S5–S7). Regardless of the biogeographic model employed, within-archipelago speciation was inferred as the dominant mode of cladogenesis (SI Appendix, Table S4 and Fig. S12), occurring primarily on the Crozet and Kerguelen archipelagos (SI Appendix, Fig. S13).
Phylogeography. The population structure in the marine weevil *Palirhoeus eatoni* based on the analysis of 5,859 SNPs showed a deep genetic break between the PEI archipelago (“west” clade) and all other islands (“east” clade), explaining over 65% of the variation in principal coordinate analysis (PCoA) (Fig. 3). Between the two clades, \(F_{ST}\) values ranged from 0.77 to 0.88, and up to one-third of SNPs were fixed for different alleles (SI Appendix, Tables S8 and S9), indicating a complete absence of contemporary
gene flow and the likelihood that these populations are undergoing allopatric speciation. The magnitude of this division supports our designation of MOTUs to capture additional diversity in the phylogenetic timetree, in which the same east and west clades were observed for *P. eatoni* (SI Appendix, Fig. S4). A further subdivision into two clusters was evident within the east clade—the Crozet archipelago formed one cluster, and the Kerguelen and HIMI archipelagos formed another (Fig. 3 and SI Appendix, Fig. S14). Considerable transoceanic dispersal between Prince Edward and Marion islands and between Kerguelen and Heard islands (despite separation by >500 km open ocean in the latter case) was demonstrated by their admixture in Bayesian cluster analysis (Fig. 3B). Nonetheless, a strong isolation-by-distance (IBD) correlation ($r = 0.85$; Fig. 3D), which remained significant when correcting for cluster in partial Mantel tests (SI Appendix, Table S10), indicates that dispersal in *P. eatoni* is distance dependent in agreement with similar findings from the historical biogeographic analysis of our phylogenetic timetree. Compared to archipelago-level structure, intraisland structure was weak (Fig. 3A) though detectable on Kerguelen Island ($F_{ST} = 0.17$ to 0.37; SI Appendix, Table S8). None of the populations were significantly different in terms of genetic diversity or inbreeding (SI Appendix, Table S11).

**Discussion**

**Antarctic Life Diversified as the Planet Cooled.** Despite growing recognition that extant terrestrial life survived in the Antarctic throughout glacial cycles (6, 14, 16), little rigorous molecular evidence is available for the tempo or mode of terrestrial diversification, nor for prolonged terrestrial evolution throughout multiple epochs (35). Here, we have integrated ~120,000 bp of genomic data with multiple calibrations and analyses to demonstrate that Ectemnorhinini beetles arrived in the sub-Antarctic from Africa around 50 Ma ago, repeatedly dispersing along the Antarctic Polar Front and diversifying as temperatures decreased from the mid-Miocene throughout the Plio–Pleistocene and up to the present day. This beetle diversification is synchronous with that of several marine groups that have radiated primarily across the Southern Ocean, including penguins (36), fishes (17), limpets (37), and cryptic-species complexes of octocorals (38) and amphipods (39). Although such a pattern may reflect a variety of extinction and speciation scenarios (40), its persistence across diverse taxa and over a period of pronounced environmental transitions is unlikely to be coincidental and aligns predictably with evolutionary theory (e.g., refs. 41 and 42).

Significant global cooling, ice sheet growth, and oceanographic changes during the middle and late Miocene climate transitions (~14 and ~7 Ma ago) finalized the isolation of Antarctica and the establishment of permanent polar conditions both marine and terrestrial (43). The emergence of most Antarctic marine biodiversity around this time has been attributed to these profound environmental changes, which provided ecological opportunities (e.g., new habitat and diet niches, extinction of predators) for marine life to exploit (2, 17, 36). This also seems to be the case for Ectemnorhinini weevils. On land, the major Miocene ecosystem transition in Antarctica was the loss of angiosperm plant diversity (44), leaving cryptogam flora and fellfield biotopes dominant on the continent (45) and even in the sub-Antarctic, where angiosperm communities waxed and waned in glacial refugia (46). The Ectemnorhinini, which appear to have undergone an acceleration in diversification from the mid-Miocene, represent one of the few beetle groups worldwide to have radiated on a primarily cryptogam (rather than angiosperm) diet (47). Most ectemnorhinines are also particularly well adapted to the epilithic setting provided by fellfield habitat (33). Therefore, a Miocene transition to polar terrestrial conditions likely promoted the diversification of these weevils (especially the cryptogam-feeding *Bothrometopus* clade), which capitalized on novel polar niches, mirroring the region’s marine scenario. Vacant ecological niches may have also been contributed by earlier extinctions, promoting “rebound” speciation in the Miocene (4).

Following a Miocene increase in diversification rate, the most significant accumulation of ectemnorhinine lineages occurred over the last 5 Ma, including angiosperm-feeding lineages, reflecting the persistence of at least some angiosperms throughout the region’s cooling (27, 46). Indeed, diversification appears to be ongoing as indicated by significant phylogeographic structure for the widespread species *P. eatoni*. A similar pattern of continued or accelerated diversification over the Plio–Pleistocene has been documented in many Antarctic marine groups (36, 38, 48). Moreover, preliminary evidence indicates the same timing of diversification processes for other terrestrial groups, such as mites (49), mosses (50), springtails (16), and diatoms (12), with diversification ongoing in some of these groups too (51). Ongoing speciation across such a fragmented terrestrial habitat should not be surprising, yet Antarctica has only recently been acknowledged as a dynamic region actively generating diversity and, even then, mainly for marine life (4, 5). As deeper molecular attention is paid to lesser-known terrestrial taxa from the Antarctic continent (e.g., bacteria, tardigrades), additional endemic diversity is invariably uncovered, implying extended histories in the region (15, 52). Given the island-like habitat available to terrestrial biota on the continent reduced to fellfield during the Miocene and further reshaped by ~5 Ma of glacial cycles, these lineages have likely undergone allopatric diversification in a manner similar to sub-Antarctic groups. Indeed, this has already been anticipated by phylogeographic studies (53, 54).

Macroevolutionary theory predicts that synchronous diversification across varied taxa should reflect allopatric processes driven by acute changes in the physical environment (41, 42). Indeed, mounting evidence indicates that cooling and habitat restructuring owing to Plio–Pleistocene glacial cycles promoted the remarkable diversification of marine fauna through population fragmentation and recolonization processes (1, 2). Similar processes are likely for the Ectemnorhinini, with increased diversification over the Plio–Pleistocene largely attributed to speciation within (rather than between) the Crozet and Kerguelen archipelagos—both of which experienced substantial glaciation (26, 55). Population divergence across known historical glacial barriers (55) was evident for the weevil *P. eatoni* on Kerguelen Island, and has been demonstrated for numerous other terrestrial species throughout the region (see ref. 10). Founder events arising from long-distance dispersal have additionally played a role as exemplified by the pronounced east–west divergence of *P. eatoni*. The mode and timing of diversification for sub-Antarctic Ectemnorhinini, alongside emerging circumstantial evidence for similar diversification tempos in other Antarctic terrestrial groups, provide strong initial support that the diversification processes common to Antarctic marine taxa are more general. Geo-climatically driven diversification since the mid-Miocene (1, 2) should thus be considered a potential general biodiversity paradigm that warrants explicit testing among additional marine and terrestrial Antarctic taxa.

**Sub-Antarctic Biogeography Characterized by Repeated Colonization.** Unveiling one of the longest Antarctic evolutionary histories to date for an extant terrestrial group not only provides insights about general diversification processes in the region but also informs long-standing biogeographic debates about faunal origins and colonization dynamics in the sub-Antarctic (56–58). African affinities have previously been suggested for invertebrates of the Kerguelen Province islands based on biogeographic and genetic evidence (30, 49, 59). Yet, these origins have been widely debated and clearly do not apply to several taxa (46, 60). Similarly, both the Kerguelen and Crozet archipelagos have been the source of much speculation about their geological origins and roles in the diversification of the Southern Hemisphere fauna (30, 59, 61).
Our analyses strongly support an African origin hypothesis for the Ectemnorhinini, settling this century-long debate. An estimated divergence from African relatives 44 to 65 Ma ago suggests that the ancestors of present-day weevils inhabited the Southern Ocean before the oldest currently subaerial island in the Kerguelen Province. Although historical biogeographic analysis could not distinguish between an ancestral distribution on the Kerguelen or Crozet islands, the former has a maximum age estimate of 39 Ma (26), and the latter has an even younger maximum estimate of 9 Ma (62). However, terrestrial sediments and fossils drilled from the Kerguelen Plateau indicate that submerged components were previously subaerial as early as the Cretaceous (63), with similar proposals made for the age of the Crozet Plateau (59). These ancient lost islands have been implicated as stepping stones for the diversification of Kerguelen weevils.

Fig. 3. Phylogeographic structure in the marine weevil *Palirhoeus eatoni* (Ectemnorhinini). Principal Coordinate Analysis (A) demonstrates the deeply divergent west and east clades. Bayesian cluster analysis (B) was conducted separately on these clades to explore finer-level structuring (cluster results based on the entire dataset are provided in SI Appendix, Figure S14). Diagrammatic representation of the sites on each island and the population connectivity requiring transoceanic dispersal is shown below the cluster analysis, while an image of *P. eatoni* (from Marion Island) is provided in panel C. A significant isolation-by-distance correlation was found (D)—for statistics regarding this correlation, see SI Appendix, Table S10. A photo of the littoral zone on Marion Island (E) demonstrates the dynamic rockface habitat of *P. eatoni* with abundant kelp for rafting.
stones for dispersal between Africa and other continents during the early Eocene (64, 65), and our findings certainly imply a potential role for them in the early evolution of the region’s insect fauna. While our dating estimates were based on an incomplete sampling of the Ectemnorhinini, unsampled extant species almost entirely belong to the recently diverged species-rich clades. Taxon sampling is therefore unlikely to significantly affect age estimates for deeper nodes such as the divergence between Ectemnorhinini and African sister taxa.

Our data suggest that long-distance transoceanic dispersal has been integral to both the initial colonization of the sub-Antarctic and the ongoing evolution of Ectemnorhinini weevils. More than 20 interarchipelago dispersal events (not necessarily involving speciation) were inferred throughout the region, with additional dispersal likely having occurred among islands within archipelagos (i.e., between subaerial and since-submerged islands) and involving unsampled lineages. Transoceanic dispersal in flightless weevils is thought to be feasible via rafting on oceanographic currents (66). Initial dispersal of weevils from Africa across the Southern Ocean could certainly have been aided by the Agulhas Return Current and Antarctic Circumpolar Current (Fig. 2) as proposed for other terrestrial taxa (67).RAFTing is also a likely dispersal mechanism for P. eatoni given its littoral habitat and genetic signals of IBD since shorter rafting journeys are likely to occur most frequently (68). Bird-mediated dispersal, however, is strongly favored for at least some ectemnorhiniine species based on our inferences of significant dispersal against the region’s prevailing westward currents. Antarctic seabirds can cover several hundred kilometers in a single day when migrating among sub-Antarctic islands (69), and soil invertebrates are known to attach to their feathers (70). Indeed, physiological data (SI Appendix, Physiological data) demonstrate that most Ectemnorhinini weevils could survive more than a week of bird-mediated dispersal before succumbing to dehydration or starvation. Irrespective of the mechanisms, our results support the growing paradigm that colonization processes are important in the biogeography of the far Southern Hemisphere (10, 58, 71).

Materials and Methods

Samples. We refer to morphologically described species using the nomenclature established by Kuschel and Chown (18), acknowledging that the taxonomy of the Ectemnorhinini remains contested (20). Ectemnorhinini were collected from Possession Island (Crozet Islands archipelago); Grande Terre Island, or commonly “Kerguelen Island” (Kerguelen Islands archipelago); Prince Edward Island (PEI archipelago); Marion Island (PEI archipelago) and Heard Island (HIMI archipelago), all situated between 46° and 53° south in the Southern Ocean (Fig. 2 and Table S13). Weevils were collected opportunistically from 2000 to 2018. Uplift remote inaccessible nature of these islands and the rarity of many of the species, specimens were collected opportunistically from 2000 to 2018. Ultimately, 24 of the 36 putative species in the group were sampled (SI Appendix, Table S13). Weevils were preserved in ethanol until genomic DNA was extracted from leg or thoracic muscle tissue. DNA was extracted using the QIAamp DNA Micro Kit. The resulting fasta files, which contained all taxa for each gene, were saved in a folder with files organized by OrthoDB IDs. Multiple sequence alignments (MSA) were then undertaken for each gene using the LINS-i option (an iterative refinement method) in MAFFT (Multiple Alignment using Fast Fourier Transform) v.7.130b (80). The alignments were checked with Aliscore v.1.2 (81, 82) and for outliers as described in ref. B3. To reconstruct a phylogeny with nucleotide data corresponding to the protein alignments, we employed PaLdNal v.14 (84), again as outlined in ref. B3. After processing MSAs for outliers and with Aliscore, the filtered fasta files were concatenated to form a supermatrix of 515 genes using FASconCAT v.1.0 (85). The final dataset analyzed contained nucleotides from first and second codon positions (“C12”) and is deposited on the Zenodo Digital Repository (doi:10.5281/zenodo.3955188). All code used in preparing the dataset can be found in ref. B6.

Phylogenomic Inference and Molecular Dating. Detailed analytical methodology is provided in SI Appendix, Text S4. Briefly, we used PartitionFinder v.1.1.1 (87) to find the best-fit cluster of partitions for the C12 dataset. The dataset was partitioned using PartitionFinder and analyzed under the GTR+G+I model using maximum likelihood (ML) phylogenetic inference conducted in RAxML (Randomized Accelerated Maximum Likelihood) v.8 (88) on the University of Memphis high-performance computing (HPC) cluster. The Phylogenet Analysis by Maximum Likelihood (PAML) package (89) was employed to generate a timetree and estimate divergence times using a Markov chain Monte Carlo (MCMC) approach. We used the data from first and second codon positions and the partitioned (best) ML tree as input for our analysis. A total of seven fossil calibrations (SI Appendix, Text S5) were used as soft minimum ages of the islands within their distribution, and for most species, multiple individuals were sequenced from each island on which they are known to occur (SI Appendix, Table S13).

Phylogenetics. We explored interspecific relationships, divergence times, and dispersal events within Ectemnorhinini by sequencing three mitochondrial and two nuclear gene regions for all 24 morphologically determined species sampled from the tribe. All but one of these species were sequenced from all of the islands within their distribution, and for most species, multiple individuals were sequenced from each island on which they are known to occur (SI Appendix, Table S13).

Multilocus Sanger Sequencing. Fragments of five genes—cytochrome c oxidase I (COI), cytochrome b, elongation factor 1-alpha, 16S ribosomal RNA, and 28S ribosomal RNA—were amplified using the Qiagen Taq DNA Polymerase Kit (details in SI Appendix, Table S13). The PCR product was purified by ExoSAP and sequenced in both directions on an Applied Biosystems 3730xl sequencer at Macrogen, Inc. Sequence chromatograms were aligned using the ClustalW algorithm (91) to create consensus sequences for each
individual. These were visually checked for indels and translated to amino acids to check for stop codons. Due to the presence of indels, the ribosomal genes 16S and 28S were aligned using the G-INS-i algorithm (an iterative refinement method) in MAFFT, with ambiguously aligned regions deleted using Gblocks v.0.91 and the options set for a stringent selection. We “RY-coded” purines and pyrimidines for the third codon of COI (see ref. 92) to ameliorate stationarity saturation (SI Appendix, Table S16 and Fig. S15). The raw sequences for each gene are deposited in GenBank (SI Appendix, Table S13).

Phylogenetic Inference and Molecular Dating. Detailed methodology regarding phylogenetic analyses and molecular dating, including a schematic overview of all analyses and how they provided cross-checks, is provided in SI Appendix Text S6 and Fig. S16). In brief, we initially explored relationships among Ectemnorhinini species using Bayesian inference (BI) and ML analyses of a concatenated dataset of the five genes. BI analysis was carried out in MrBayes on the CIPRES (Cyberinfrastructure for Phylogenetic Research) Science Gateway 3.3 (93) using best-fitting substitution models applied independently to partitions of the sequence data as determined by PartitionFinder (SI Appendix, Table S17). ML analysis was conducted with RAxML on the CIPRES Gateway using the GTR+G substitution model applied independently to data partitions as determined by PartitionFinder (SI Appendix, Table S17). BI and ML phylogeny estimates were compared to assess congruence—with one another, with the phylogenetic tree generated by AHe, and with morphological species designations. As some phylogenetic clades were inconsistent with morphological species designations, these species themselves have been reanalyzed (see refs. 18 and 20), and ML and BI trees were used to designate MOTUs for subsequent analyses as detailed in SI Appendix, Designation of Molecular Operational Taxonomic Units (MOTUs). We additionally performed all analyses using morphological species to ensure our use of MOTUs did not significantly affect our interpretations, and the results were highly similar (SI Appendix, Supplementary results based on morphological species rather than MOTUS). Thus, we only refer to MOTU-derived results in the main text.

Divergence dates among Ectemnorhinini were estimated from our five-gene concatenated dataset, as implemented in BEAST v2.5 (94), and by costimating gene trees embedded in a shared species tree, as implemented in starBEAST (95). We randomly selected one sequence to represent each MOTU (or morphological species) for analysis in BEAST. For starBEAST, which performs optimally with multiple samples per lineage, we retained the entire sequence dataset. Uncorrelated relaxed lognormal clocks allowed for rate variation among lineages and a birth–death process was used for the tree prior. For calibration, geological (island formation) constraints were applied as uniform upper bounds, and secondary fossil-derived constraints were applied as normal priors (see SI Appendix, Text S7 and Fig. S17). A second analysis was run in BEAST only with secondary constraints excluded but with substitution rates calculated previously for Coleoptera provided as lognormal priors for the mean rates of relaxed clocks (SI Appendix, Text S7).

Diversification and Historical Biogeography Inference. Diversification rates and potential rate shifts for the Ectemnorhinini (based on the most thoroughly calibrated phylogeny generated by BEAST; see SI Appendix, Fig. S16) were assessed using a MCMC approach implemented in BAMM v2.5.0 (96). Sampling bias was accounted for using an input file providing the taxonomic coverage for each genus (as per SI Appendix, Table S13). We ran four chains of 20 million generations each and analyzed the output with the BAMMMtoolkit package for R, checking log likelihood traces and effective sample size values for convergence. For each of the 9,000 simulations of diversification rate retained by BAMM (after a burn-in of 10%), we also tested for a correlation between the estimated speciation rate and paleoclimate (specifically temperature). Deep-sea benthic oxygen isotope data smoothed in both 20,000-y and million-year windows (from ref. 97) were converted to temperature as per ref. 98 to reconstruct paleoclimate records reflecting shorter- and longer-term trends. The correlations were analyzed in R (see ref. 99 for details). We used Pearson's correlation coefficients to account for autocorrelation of the two timeseries, detrended cross-correlation analysis (100). A Student's t test was used to determine if the mean correlation coefficient in each case was significantly different from zero. To further examine the relationship between paleotemperature and speciation rate, we used the RPANDA package for R (101) to assess the fit of various temperature-dependent, time-dependent, and constant-rate diversification models to our phylogenetic tree (using the million-year smoothed paleoclimate data). We used Akaike information criterion correction values and weights to identify the diversification model with the best fit. We restricted comparisons to exponentially variable models because of ongoing difficulties with the way RPANDA handles linear dependencies. Input files and the R script for the RPANDA analyses have been deposited on Figshare (doi:10.26180/14446023).

Biogeographic history was inferred from our BEAST timetree using ML and a conservative range estimation carried out with BioGeoBEARS (BioGeography with Bayesian (and likelihood) Evolutionary Analysis in R Scripts) (102). We applied the DEC biogeographic model (103), which includes more biologically relevant assumptions of speciation such as subset sympathy, than, for instance, the Dispersal–Vicariance Analysis model, which assumes vicariance as a default explanation (104). We also ran the model with or without two additional free parameters: founder-event speciation (“r”) and a distance modifier for dispersal probability (“h”), both of which are prudent in the context of widely dispersed islands (105). Statistically comparing the likelihood of each model fit has been deemed inappropriate (106), and thus, we focused on the most biologically relevant model which includes all parameters (DEC+J+X) and compared outcomes from the simpler models. In each instance, we carried out 100 iterations of BS (107) to infer the average number and type of cladogenesis and anagenetic dispersal and speciation events throughout the evolutionary history of the phylogeny. Analyses were time stratified such that Heard Island and the PEI could not be ancestral ranges before their subaerial appearance 22 (108) and 0.475 Ma ago (109), respectively. The ranges were designated at the archipelago level (i.e., the geographically proximal Marion and Prince Edward islands were considered a single range or the PEI) so that inferences to separate islands or assemblage ranges before their subaerial appearance could not be considered as long-distance among-archipelago events. Geographic ranges and the distances between each archipelago (used as input data) are provided in SI Appendix, Input data for historical biogeographic inference and on Figshare, along with all associated R scripts (doi:10.26180/14446023).

Phylogeography. We explored contemporary gene flow among islands using genome-wide SNPs for P. raihiceus eatoni, the most widespread and only supralittoral-dwelling ectemnorhinine species. We sampled P. eatoni from one site each on Marion and Heard islands and from multiple sites on Prince Edward, Possession, and Kerguelen islands, enabling exploration of intraspecies as well as interisland population genetic structure (SI Appendix, Table S14). Marion Island was sampled at two different time points, though preliminary analyses failed to demonstrate any significant temporal difference (regarding allele frequencies or overall diversity); therefore, these samples were pooled to create a single population for Marion.

SNP Library Preparation, Genotyping, and Quality Filtering. A genome-wide SNP library for P. eatoni was developed using Diversity Arrays Technology sequencing (DART-Seq) (110) at DArT Pty. Ltd. DArt-Seq is a reduced-representation next-generation sequencing method, similar to double-digest restriction-associated DNA sequencing. A customized combination of restriction enzymes was used to digest genomic DNA, adaptors were ligated to restriction enzyme overhangs, fragments were PCR amplified and sequenced on an Illumina HiSeq 2500, and raw Illumina data files were processed with a proprietary DArT pipeline to provide a quality-filtered SNP dataset. The SNP dataset generated by DArT was further filtered using the “dartsr” package v.1.0.5 (111) in R. A detailed explanation of DArt-Seq SNP library preparation, quality control, and filtering pipeline can be found in SI Appendix, Supplementary methods for phylogeography, and the final filtered SNP dataset is available on Figshare, together with code for all analyses performed in R (doi:10.26180/14446023).

Phylogeographic Analyses. The final dataset was tested for evidence of selection using the Fu’s outlier approach and a false discovery rate of α = 0.01 in BayScan 2.1 (112). SNPs were also tested for deviations from the Hardy–Weinberg Equilibrium (HWE) in each population, using exact tests conducted in dartsr and adjusted with the Bonferroni correction. SNPs putatively under selection or found to violate HWE in any single population each represented less than 5% of the total dataset, and only 15 SNPs (0.3% of the dataset) violated HWE in more than one population. Therefore, while a conservative dataset was prepared with all of these SNPs removed, it was only used for a subset of analyses to check that the removal of these loci did not affect results. Indeed, results were congruent with those for the full dataset (SI Appendix, Phylogeographic results for conservative SNP dataset); thus, we only refer to the full dataset in the main text.

Genetic diversity and population structure were assessed within and among sites as detailed in SI Appendix, Text S9. In summary, we calculated observed and expected heterozygosity, inbreeding coefficients, and pairwise FST estimations among all populations using GenoDive v.2.0 (113). PCA was conducted in dartsr to observe the clustering of individuals due to genetic similarity, and a Bayesian clustering algorithm was also used to infer population structure as implemented in fastSTRUCTURE (114). It was clear that...
there were two pronounced genetic clusters of the data (“east” and “west”), which swapped all finer-level genetic structure; therefore we ad-
dition for conducting fastSTRUCTURE analysis within each of these clusters independently. As the identification of population clusters can be con-
fronted by an underlying pattern of IBD and vice versa (115), we explored the dataset for a linear relationship between genetic distance (FST) and geographic distance between sites (input data: SI Appendix, Table S25), which would indicate IBD. We carried out partial Mantel tests using the “ncf” package v.1.2-8 (116) in R, testing whether IBD patterns remained significant while controlling for clusters as a covariate and whether clusters remained significant after correcting for geographic distance. Finally, we explored the dataset for fixed allelic differences between populations (when no alleles are shared for a given SNP), which indicates a complete absence of contemporary gene flow (117), using dartR.

Data Availability. Data on genetic material contained in this paper are published for noncommercial use only. Utilization by third parties for purposes other than noncommercial scientific research may infringe the conditions under which the genetic resources were originally accessed and should not be un-
taken without obtaining consent from the corresponding author of the paper and/or obtaining permission from the original provider of the genetic material. Phylogenomic (AHE) data have been deposited in Zenodo (DOI: 10.5281/zenodo.3955188) (118); phylogeographic (SNP) data have been deposited in Figshare (DOI: 10.26180/14446023) (119) and all phylogenetic sequences have been deposited in GenBank (accessions MT701045 and MT701503).

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