Epigenome-wide association study of biomarkers of liver function identifies albumin-associated DNA methylation sites among male veterans with HIV

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Background: Liver disease (LD) is an important cause of morbidity and mortality for people with HIV (PWH). The molecular factors linked with LD in PWH are varied and incompletely characterized. We performed an epigenome-wide association study (EWAS) to identify associations between DNA methylation (DNAm) and biomarkers of liver function—aspartate transaminase, alanine transaminase, albumin, total bilirubin, platelet count, FIB-4 score, and APRI score—in male United States veterans with HIV.

Methods: Blood samples and clinical data were obtained from 960 HIV-infected male PWH from the Veterans Aging Cohort Study. DNAm was assessed using the Illumina 450K or the EPIC 850K array in two mutually exclusive subsets. We performed a meta-analysis for each DNAm site

Abbreviations: AA, Age acceleration; ALT, Alanine amino transferase; APRI, AST to platelet ratio index; ART, Antiretroviral therapy; AST, Aspartate amino transferase; BMI, Body mass index; CpG, 5′-Cytosine—phosphate—Guanine—3′; DNAm, Deoxyribonucleic acid methylation; EEAA, Extrinsic epigenetic age acceleration; EWAS, Epigenome-wide association study; FIB, Fibrosis; FKBP, FK506 binding protein; HBV, Hepatitis B virus; HCC, Hepatocellular carcinoma; HCV, Hepatitis C virus; HIV, Human immunodeficiency virus; IEAA, Intrinsic epigenetic age acceleration; LD, Liver Disease; NACD, Non-AIDS related chronic disease; NAFLD, Non-Alcohol fatty liver disease; PWH, People with HIV; SOCS, suppressor of cytokine signaling; STAT, Signal transducer and activator of transcription; TGF, Tissue growth factor; TMEM, transmembrane; TNFa, tumor necrosis factor- alpha; VACS, Veterans aging cohort; ZEB, Zinc Finger E-Box Binding Homeobox 1.
measured by either platform. We also examined the associations between four measures of DNAm age acceleration (AA) and liver biomarkers.

**Results:** Nine DNAm sites were positively associated with serum albumin in the meta-analysis of the EPIC and 450K EWAS after correcting for multiple testing. Four DNAm sites (cg16936953, cg18942579, cg01409343, and cg12054453), annotated within the TMEM49 and four of the remaining five sites (cg18181703, cg03546163, cg20995564, and cg23966214) annotated to SOCS3, FKBP5, ZEB2, and SAMD14 genes, respectively. The DNAm site, cg12992827, was not annotated to any known coding sequence. No significant associations were detected for the other six liver biomarkers. Higher PhenoAA was significantly associated with lower level of serum albumin ($\beta = -0.007$, $p$-value = $8.6 \times 10^{-4}$, CI: -0.011116, -0.002884).

**Conclusion:** We identified epigenetic associations of both individual DNAm sites and DNAm AA with liver function through serum albumin in men with HIV. Further replication analyses in independent cohorts are warranted to confirm the epigenetic mechanisms underlying liver function and LD in PWH.

**KEYWORDS**
EWAS, liver disease, HIV, biomarkers, albumin (ALB)
Introduction

The widespread availability and use of effective combination antiretroviral therapy to treat HIV has transformed the outlook for people with HIV (PWH), many of whom now live longer and are facing the challenges of aging with HIV. The burden of non-AIDS related chronic disease (NACD) is increasingly recognized in this patient population as an urgent concern that needs to be addressed to improve care for PWH.

Liver disease (LD) accounts for an estimated 13%–18% all-cause mortality in PWH, and PWH have a high prevalence of traditional causes of LD (Palella et al., 2006; Smith et al., 2010; Smith et al., 2014; Kaspar and Sterling, 2017). Co-infection with hepatitis B and C viruses is estimated between 5%–30% based on cohort studies (Konopnicki et al., 2005; Shepard et al., 2005), and up to 30%–40% of PWH have signs of non-alcoholic fatty liver disease (NAFLD) (Lemoine et al., 2012). Poorly controlled HIV-infection itself is an independent risk factor for liver fibrosis (Kim et al., 2016a), which is the most common outcome of chronic liver injury. Even in individuals with well-controlled HIV (undetectable HIV RNA) and immunologic recovery (CD4+ cell counts > 500 cells/mm³), oxidative stress, mitochondrial injury, toxic metabolite accumulation, gut microbial translocation, systemic inflammation, senescence, and nodular regenerative hyperplasia [reviewed in (Kim et al., 2016a)] contribute to liver injury and fibrosis. Given all of the factors contributing to LD among PWH, there is a need to improve prevention, risk prediction, and treatment of LD.

Progress in elucidating the biological mechanisms underlying LD has been challenging due to the complexity of the multiple roles played by the liver in homeostasis and metabolic functions. Epigenome-wide association study (EWAS) have the potential to identify novel mechanisms of LD and generate new hypotheses that may provide insights into prevention and treatment of LD. Previous studies on the role of DNA methylation (DNAm) in LD have focused on non-alcoholic fatty LD (NAFLD), since determinants of NAFLD are not as easily identifiable as LD resulting from e.g., alcoholism or viral hepatitis [reviewed in (Zhang et al., 2021)]. Studies on the association between alcoholic liver disease and DNAm have identified genes which may be associated with the incidence and progression of hepatocellular carcinoma (HCC) (French, 2013; Varela-Rey et al., 2013; Zakhari, 2013; Rosen et al., 2018). Other EWAS have uncovered DNAm sites and genes associated with the development of HCC in patients with chronic hepatitis B infections (Su et al., 2007; Nishida et al., 2008; Zhao et al., 2014). PWH are significantly underrepresented in these studies. Moreover, most studies have focused on LD as an overall clinical diagnosis and have not assessed potential associations between DNAm and specific metabolic functions of the liver. Biomarkers are useful surrogates of the liver’s metabolic functions and may reflect its capacity to perform specific functions. The absence of data on the relationship between DNAm and specific markers of liver function, particularly among PWH, represents an important gap in the literature which we hope the present study will begin to fill.

DNAm age acceleration (AA) is a novel biomarker of biological aging, and predicts age-related disease outcomes and mortality (Bell et al., 2019). Several studies have also shown that PWH exhibit elevated DNAm AA above the average level exhibited by non-infected people of equivalent chronological age. (Horvath and Levine, 2015; Rickabaugh et al., 2015). These observations suggest that DNAm AA may partially explain the early onset of chronic comorbid conditions among PWH (Levine et al., 2015a). Relative acceleration of DNAm age from blood-based assays is often measured by four metrics: intrinsic epigenetic age acceleration (IEAA) (Levine et al., 2015a; Horvath and Levine, 2015), extrinsic epigenetic age acceleration (EEAA) (Hannum et al., 2013; Levine et al., 2015b), phenotypic age acceleration (PhenoAA) (Levine et al., 2018), and Grim age acceleration (GrimAA) (Lu et al., 2019).

The present study included available markers of liver function from the Veterans Aging Cohort Study (VACS): aspartate transaminase (AST), alanine transaminase (ALT), serum albumin, total bilirubin, platelet count, fibrosis-4 (FIB-4) score, and the AST-to-platelet-ratio index (APRI) score. We conducted epigenetic association analyses between individual DNAm sites, measures of DNAm AA (IEAA, EEAA, PhenoAA and GrimAA) and these seven biomarkers of liver function in a cohort of male veterans with HIV infection in the United States.

Methods

Study population

A total of 960 male veterans with HIV at the time of blood collection were included from the VACS, having both phenotypic and epigenetic data available from previous studies (Justice et al., 2006; Chen et al., 2020). Participants with cancer at the time of blood collection were excluded because the cancer treatment and disease can influence liver function and DNAm which is beyond the scope of the present study. The VACS consists of electronic medical record data from patients with HIV receiving care at Veterans Health Administration (VA) medical facilities across the United States. Each PWH is matched on age, sex, race/ethnicity, and site to two persons without HIV. Data include hospital and outpatient diagnoses (recorded using International Classification of Diseases, Ninth Revision [ICD-9] and Tenth [ICD-10] codes), procedures (recorded...
using Current Procedural Terminology [CPT] codes), laboratory results, and pharmacy data (Justice et al., 2006). Information on age, race, smoking status, body mass index (BMI), diabetes status (glucose level ≥ 200 mg/dl on 2 separate occasions or glucose level ≥ 200 mg/dl on 1 occasion plus treatment with an oral hypoglycemic or insulin for ≥ 30 days) (Mathur et al., 2019), ever infection with HBV (defined as HBV surface antigen positive, acute resolved HBV or ICD-9 code for HBV diagnosis), ever infection with HCV (defined as HCV antibody positive regardless of HCV RNA, ICD-9 code for HCV diagnosis, or HCV genotype), and antiretroviral (ART) use at time of blood draw were obtained from VA’s clinical data warehouse. Alcohol use status was obtained from the VACS survey (Justice et al., 2006). At the visit of blood draw, AST, ALT, serum albumin, total bilirubin, platelet count, CD4+ T-cell count, and plasma HIV-1 RNA viral load were measured for each patient using approved clinical assays from certified clinical laboratories. Participants were categorized as HIV suppressed if their HIV RNA was < 200 copies/mm³ or as unsuppressed VL otherwise. A fibrosis-4 (FIB-4) score and an AST-to-platelet ratio index (APRI) score were calculated for each participant following the standard equations (Wai et al., 2003; Sterling et al., 2006). All participants provided written consent for the use of their data.

**DNAm data generation, processing, and quality control**

The genome-wide DNAm profiles were measured using genome DNA samples extract from whole blood. Genomic DNA extraction, epityping, data processing, and quality control were performed as previously described. (Chen et al., 2020). Genome-wide DNAm levels for 473 of the participants were assessed using the Infinium HumanMethylation450 (450K) array platform (Illumina), while those for the remaining 487 participants were assessed using the Infinium HumanMethylationEPIC (EPIC) array platform (Illumina) at the Yale Center for Genomic Analysis. Quality control procedures were followed as previously described in published studies (Zhang et al., 2017; Solomon et al., 2018; Chen et al., 2020). We compared the characteristics between the 450K and EPIC subsets using t-tests for continuous variables and chi-square tests for categorical variables.

DNAm sites for 412,583 and 846,604 DNAm sites were included in the analyses for the dataset acquired with the 450K and EPIC arrays, respectively. Differences in the proportions of the six main leukocyte cell types present in whole blood (CD4+ T cells, CD8+ T cells, monocytes, B cells, granulocytes, and natural killer cells) across samples are well described confounders of associations between DNAm in the blood and many phenotypes. To account for this, the proportions of these six cell types for each participant were determined based on the top cell-type-specific DNAm sites in a reference panel of known proportions following the standard algorithm through the miniF package in R (Houseman et al., 2012; Moran et al., 2016). The estimated cell type proportions were then controlled for in all EWAS analyses.

The association between each covariate included in the EWAS model—race, smoking status, BMI, diabetes status, hazardous alcohol use (Freiberg et al., 1999), ever HCV infection, ever HBV infection, ART use, CD4 count, viral suppression, and leukocyte cell-type proportions, top ten principal components—and each liver marker was assessed using a linear model controlling for chronological age as in the following model.

**Individual liver marker ~ covariate + age**

Associations between each liver marker and chronological age were also assessed using a univariate model.

**Individual liver marker ~ age**

Regardless of statistical significance, all covariates listed above were included in the final EWAS model to account for the reasonable possibility that they might confound the relationship between DNAm and the liver markers as in the following model.

**Individual liver marker ~ DNAm + age + race +current smoking + BMI + diabetes + hazardous alcohol use + ever infection with HCV + ever infection with HBV + ART use + CD4 T-cell count + VL + leukocyte cell type proportions + top ten principal components.**

**Correlations among liver biomarkers**

To identify correlations among the biomarkers of liver function included in this study, a Spearman correlation coefficient and corresponding p-value were calculated for every pair of selected biomarkers using the R statistical software package. To normalize the distributions of strongly right-skewed biomarkers for this and all subsequent analyses, a natural log transformation was performed on the measured ALT, AST, and calculated FIB-4 and APRI scores.

**Associations of DNAm age acceleration with liver biomarkers**

DNAm AA for each participant was measured using the IEAA, EEAA, PhenoAA, and GrimAA metrics as specified by
IEAA is calculated as the residual of a person’s DNAm level in blood cells across a set of DNAm sites proposed by Horvath after regressing DNAm level on chronological age and controlling for proportions of different leukocytes within the sample (Levine et al., 2015a; Horvath and Levine, 2015). EEAA is calculated similarly, except that a different set of DNAm sites proposed by Hanum are used while controlling for proportions of a different set of cell types (Hannum et al., 2013; Levine et al., 2018). PhenoAA, in contrast, is calculated by first estimating phenotypic age (in years) with a linear regression model that uses clinical variables as inputs, then estimating DNAm age (in years) based on methylation levels at a set of DNAm sites proposed by Levine et al. (2018) and finally calculating the residual that results from regressing calculated DNAm age on calculated phenotypic age. GrimAA is calculated in a similar way as PhenoAA but utilizes levels of plasma biomarkers indicative of physiological stress to estimate phenotypic age and a different set of DNAm sites to assess DNAm age (Lu et al., 2019). Each of these measures of DNAm AA perform differently depending on the outcome being evaluated and can complement each other to assess the biological aging process.

The association between DNAm AA, as measured by each of these four metrics, and each of the selected biomarkers of liver function was then assessed by linear regression while controlling for all covariates included in the final EWAS model.

### TABLE 1 Characteristics of participants grouped by platform used for genome-wide DNA methylation.

| Characteristic                  | 450K (N = 473) | EPIC (N = 488) | Overall (N = 961) | p-value |
|--------------------------------|----------------|----------------|-------------------|---------|
| ALT (units/L)                  | 38.8 (24.9)    | 34.1 (21.2)    | 36.4 (23.2)       | 1.9 × 10⁻^3 |
| AST (units/L)                  | 43.4 (28.1)    | 37.0 (20.1)    | 40.2 (24.6)       | 7.3 × 10⁻^5 |
| Serum Albumin (mg/dl)          | 3.84 (0.489)   | 3.95 (0.457)   | 3.90 (0.476)      | 2.9 × 10⁻^6 |
| Total Serum Bilirubin (mg/dl)  | 0.78 (0.485)   | 0.76 (0.513)   | 0.77 (0.500)      | 0.61    |
| Platelet Count (x10 [Zhang et al., 2021]/L) | 224 (71.3) | 232 (69.4) | 228 (70.4) | 8.0 × 10⁻⁴ |
| FIB-4 Score                    | 1.81 (1.22)    | 1.57 (0.943)   | 1.69 (1.09)       | 8.5 × 10⁻⁴ |
| APRI Score                     | 0.58 (0.602)   | 0.47 (0.393)   | 0.52 (0.509)      | 8.4 × 10⁻⁴ |
| Age (years)                    | 51.8 (7.53)    | 50.7 (7.48)    | 51.2 (7.52)       | 3.1 × 10⁻² |
| Race                           |                |                |                   | 1.00    |
| Black                          | 402 (85.0%)    | 391 (80.1%)    | 793 (82.5%)       | 1.00    |
| Non-Black                      | 71 (15.0%)     | 97 (19.9%)     | 168 (17.5%)       | 1.00    |
| Smoking Status                 |                |                |                   | 1.00    |
| Not current                    | 203 (42.9%)    | 210 (43.0%)    | 413 (43.0%)       | 1.00    |
| Current                        | 270 (57.1%)    | 278 (57.0%)    | 548 (57.0%)       | 1.00    |
| Diabetes Status                |                |                |                   | 1.00    |
| No                             | 390 (82.5%)    | 406 (83.2%)    | 796 (82.8%)       | 1.00    |
| Yes                            | 83 (17.5%)     | 82 (16.8%)     | 165 (17.2%)       | 1.00    |
| BMI                            | 25.4 (4.44)    | 25.9 (4.43)    | 25.6 (4.44)       | 0.14    |
| Missing                        | 8 (1.7%)       | 9 (1.8%)       | 17 (1.8%)         | 1.00    |
| Alcohol Use                    |                |                |                   | 1.00    |
| Non-Hazardous                  | 195 (41.2%)    | 191 (39.1%)    | 386 (40.2%)       | 1.00    |
| Hazardous                      | 276 (58.4%)    | 297 (60.9%)    | 573 (59.8%)       | 1.00    |
| Ever Hepatitis C Infection     |                |                |                   | 1.00    |
| No                             | 241 (51.0%)    | 312 (63.9%)    | 553 (57.5%)       | 1.00    |
| Yes                            | 232 (49.0%)    | 176 (36.1%)    | 408 (42.5%)       | 1.00    |
| Ever Hepatitis B Infection     |                |                |                   | 1.00    |
| No                             | 399 (84.4%)    | 430 (88.1%)    | 829 (86.3%)       | 1.00    |
| Yes                            | 52 (11.0%)     | 40 (8.2%)      | 92 (9.6%)         | 1.00    |
| CD4⁺ Cell Count                | 411 (252)      | 442 (263)      | 427 (258)         | 6.4 × 10⁻⁷ |
| HIV viral Load                 |                |                |                   | 1.00    |
| Suppressed                     | 196 (41.4%)    | 204 (41.8%)    | 400 (41.6%)       | 1.00    |
| Unsuppressed                   | 274 (57.9%)    | 284 (58.2%)    | 558 (58.1%)       | 1.00    |

Statistics for numeric variables are presented as mean (SD), while those for categorical variables are presented as count (%). Counts across levels of some categorical variables may not sum to the corresponding total due to missing values. N= number; P-value, level of statistical significance.
infection with HCV + ever infection with HBV + ART use + CD4+ T-cell count + VL + leukocyte cell type proportions + top ten principal components.

Meta-analysis of EWAS for liver biomarkers

To investigate the association between each biomarker of liver function and individual DNAm sites across the autosomal chromosomes, data from the 450K dataset were analyzed in parallel with data from the EPIC dataset using the same model, and a fixed effect model meta-analysis was conducted using inverse variance weighted effect size method. The final model was adjusted for age, race, current smoking, BMI, diabetes, hazardous alcohol use, ever infection with HCV, ever infection with HBV, ART use, CD4+ T-cell count, VL, leukocyte cell type proportions, and the top ten principal components within the group being analyzed. Where necessary for any given combination of DNAm site and biomarker, participants missing values required for the model were excluded from the analysis for that combination.

A total of 385,062 DNAm sites measured by both the 450K, and EPIC arrays were included in the EWAS meta-analysis. For DNAm sites not covered by both platforms, results were obtained only from the cohort profiled by the covering platform and no meta-analysis was conducted. Separate false-discovery rate adjustments (q < 0.05) were conducted for each liver biomarker. All analyses were performed in R (v4.0.3).

Results

Participant characteristics

After data processing and quality control, the analysis dataset for those profiled using the 450K and EPIC platforms included observations from 473 to 487 individuals, respectively. Characteristics of the two EWAS subsets are summarized in Table 1. All participants were male veterans with HIV who were never diagnosed with cancer and had an average age of 51.2 ± 7.5 years. The chronological age between the 450K and EPIC sub-cohorts was moderately different (Δ = 1.1 years, p-value = 0.031). The distribution of two sub-cohorts did not differ significantly (p-value > 0.05) in race (Black vs. non-Black) or in prevalence of current smoking, diabetes, hazardous alcohol use, ever infection with HBV, ever infection with HCV, or viral load suppression. They also did not differ significantly in their average BMI or CD4+ T-cell count. Average biomarker values from the 450K cohort differed significantly from those from the EPIC cohort for ALT (Δ = 4.7 units/L, p-value = 1.93 × 10^{-3}), AST (Δ = 6.4 units/L, p-value = 7.34 × 10^{-4}), serum albumin (Δ = −0.11 mg/dl, p-value = 2.89 × 10^{-3}), FIB-4 score (Δ = 0.24, p-value = 8.49 × 10^{-4}), and APRI score (Δ = 0.112, p-value = 8.35 × 10^{-4}). Total bilirubin and platelet count did not differ significantly between the two sub-cohorts.

Correlations among liver biomarkers

To illustrate the overall correlation between seven liver function markers, we summarized the pair-wise correlation in Figure 1. Observed values for ALT, AST, FIB-4 score, and APRI score all correlated positively and strongly with each other (Spearman correlation coefficient ρ > 0.6), except for ALT and FIB-4 score with moderate correlation (ρ = 0.36) (Figure 1). Platelet count correlated negatively with AST, ALT, FIB-4 score, and APRI score to varying degrees and weakly with albumin (ρ = 0.13), but not with total bilirubin (ρ = -0.06). Serum albumin correlated negatively with AST, APRI score, and FIB-4 score (−0.25 ≤ ρ ≤ −0.18) but not with ALT (ρ = -0.01). Total bilirubin did not correlate significantly with serum albumin, platelet count, FIB-4 score, or APRI score (ρ > 0.05) and correlated only weakly with AST and ALT (ρ < 0.14). Distributions of the biomarkers before and after transformation, are presented in Supplementary Figure S1.
Covariate associations with liver biomarkers

Associations between covariates included in the final EWAS model and the selected liver biomarkers are presented in Table 2. Only "ever infection with HCV" was associated with all seven selected liver markers when controlling for age, while diabetes status was not associated with any of the selected markers. All liver markers associated with at least four of the covariates included in the model excluding cell type proportions. Regardless of the p-value for the observed association between each covariate and each liver biomarker, all covariates were consistently included in the final epigenetic analysis model for each liver biomarker. Participants with complete data of all covariates were analyzed.

Associations of DNAm age acceleration with liver biomarkers

Serum albumin was significantly associated with PhenoAA (beta = −0.014, p-value = 1.6 × 10−13) in an unadjusted model (Figure 2). That association remained significant after adjusting for all covariates in the final regression model (beta = −0.007, p-value = 8.6 × 10−4) (Table 3). No other metric of DNAm AA (i.e., IEAA, EEAA, and GrimAA) was associated with any other

TABLE 2 Associations between DNA methylation age acceleration and selected biomarkers of liver function.

| Liver Marker | PhenoAA | GrimAA | IEAA | EEAA |
|--------------|---------|--------|------|------|
| ln (AST) Beta (SE) | 2.5 × 10⁻⁴ (2.1 × 10⁻⁴) | 7.9 × 10⁻⁷ (4.0 × 10⁻⁴) | 2.6 × 10⁻⁴ (2.8 × 10⁻⁴) | 1.9 × 10⁻⁴ (2.8 × 10⁻⁴) |
| p | 0.23 | 0.05 | 0.36 | 0.51 |
| ln (ALT) Beta (SE) | 1.9 × 10⁻⁴ (2.5 × 10⁻⁴) | 3.4 × 10⁻⁵ (4.7 × 10⁻⁵) | 1.3 × 10⁻⁴ (3.3 × 10⁻⁵) | 5.5 × 10⁻⁴ (3.3 × 10⁻⁵) |
| p | 0.94 | 0.47 | 0.69 | 0.87 |
| ALB Beta (SE) | -7.0 × 10⁻⁶ (2.1 × 10⁻⁴) | -7.2 × 10⁻⁶ (4.0 × 10⁻⁶) | -4.1 × 10⁻⁶ (2.8 × 10⁻⁶) | -4.3 × 10⁻⁶ (2.8 × 10⁻⁶) |
| p | 8.6 × 10⁻⁴ | 0.07 | 0.15 | 0.12 |
| TBILI Beta (SE) | -4.9 × 10⁻⁵ (2.3 × 10⁻⁵) | -1.8 × 10⁻⁵ (4.4 × 10⁻⁶) | 2.1 × 10⁻⁵ (3.2 × 10⁻⁵) | 2.0 × 10⁻⁵ (3.1 × 10⁻⁵) |
| p | 0.04 | 0.69 | 0.50 | 0.52 |
| PLT Beta (SE) | -0.48 (0.32) | -0.12 (0.61) | -0.2 (0.44) | -0.72 (0.43) |
| p | 0.14 | 0.84 | 0.64 | 0.09 |
| ln (FIB-4) Beta (SE) | 1.4 × 10⁻⁴ (2.2 × 10⁻⁴) | -5.5 × 10⁻⁴ (4.1 × 10⁻⁵) | -1.4 × 10⁻⁴ (2.9 × 10⁻⁵) | 1.8 × 10⁻⁴ (2.9 × 10⁻⁵) |
| p | 0.53 | 0.89 | 0.63 | 0.54 |
| ln (APRI) Beta (SE) | 1.3 × 10⁻⁴ (2.9 × 10⁻⁴) | 3.2 × 10⁻⁴ (5.5 × 10⁻⁵) | 2.6 × 10⁻⁴ (3.9 × 10⁻⁵) | 3.7 × 10⁻⁴ (3.9 × 10⁻⁵) |
| p | 0.64 | 0.56 | 0.95 | 0.34 |

Biomarker values were ln-transformed when needed to achieve a more normal distribution as indicated on the left and then regressed on each DNA methylation age acceleration metric in linear models that control for the covariates included in the EWAS model. Associations that remained significant after FDR adjustment at Q < 0.05 are highlighted in yellow. Abbreviations: AST, aspartate aminotransferase (units/L); ALT, alanine aminotransferase (units/L); ALB, serum albumin (mg/dl); TBILI, total bilirubin level (mg/dl); PLT, platelet count (cells/µL); FIB-4, FIB-4 score; APRI, APRI score; PhenoAA, phenotype age acceleration; GrimAA, Grim age acceleration; IEAA, intrinsic epigenetic age acceleration; EEAA, extrinsic epigenetic age acceleration.

FIGURE 2
Scatter plot of serum albumin values across values for phenotypic age acceleration (PhenoAA) among the entire cohort. Red dots indicate participants from 450K and green dots indicate participants from EPIC. An unadjusted best-fit line is included (b = −0.01, p-value = 1.6 × 10⁻¹³).
TABLE 3 DNAm sites are significantly associated with selected liver markers after meta-analysis and FDR correction.

| Chr | Pos | Gene       | Beta (SE) | p   | FDR | Beta (SE) | p   | 450K Start Site |
|-----|-----|------------|-----------|-----|-----|-----------|-----|----------------|
| 1   | 158 | cg16936953 | 1.2 (0.21) | 4.2 × 10^-4 | 0.001 | 1.0 (0.29) | 6.6 × 10^-4 | 1.4 (0.29) |
| 57  | 57915665 | TMEM49     | 1.9 (0.35) | 9.5 × 10^-4 | 0.009 | 2.3 (0.45) | 9.9 × 10^-4 | 1.3 (0.55) |
| 7   | 35654363 | FKBP5      | 1.1 (0.20) | 1.1 × 10^-7 | 0.009 | 1.2 (0.26) | 9.2 × 10^-4 | 0.94 (0.33) |
| 17  | 57915773 | TMEM49     | 1.4 (0.26) | 1.1 × 10^-7 | 0.009 | 1.2 (0.34) | 7.3 × 10^-4 | 1.7 (0.41) |
| 17  | 57915740 | TMEM49     | 1.7 (0.32) | 1.2 × 10^-7 | 0.009 | 1.7 (0.43) | 1.4 × 10^-4 | 1.8 (0.48) |
| 17  | 57915717 | TMEM49     | 1.0 (0.20) | 5.0 × 10^-7 | 0.024 | 0.7 (0.29) | 1.1 × 10^-4 | 1.3 (0.28) |
| 17  | 101901234| None Mapped| 1.9 (0.38) | 8.0 × 10^-7 | 0.034 | 2.2 (0.47) | 4.0 × 10^-4 | 1.3 (0.66) |

Chr: chromosome number; Pos: basepair position (human genome build hg19); Beta: beta coefficient; SE: standard error; p: p-value, i.e. level of statistical significance. Results for DNAm sites where DNA methylation remained significantly associated with serum albumin after meta-analysis and FDR correction are presented, including statistics from the meta-analysis and from the separate EWAS of the two cohorts. No significant associations were identified after meta-analysis and FDR correction for AST, ALT, total serum albumin, platelet count, FIB-4 score, or APRI score, so those markers are excluded from the table. Coefficients represent the average change in serum albumin (mg/dl) expected for an increase in DNA methylation from 0% to 100%.

Discussion

In this study we evaluated the epigenetic associations with seven clinically relevant biomarkers of liver function—aspartate aminotransferase (AST), alanine aminotransferase (ALT), total serum albumin, platelet count, fibrosis-4 (FIB-4) score, and aspartate aminotransferase-to-platelet ratio-index (APRI). Our meta-analysis of the EWAS results from the EPIC and 450K datasets included in both platforms identified that hypermethylation at nine DNAm sites was significantly associated with increased serum albumin among male veterans with HIV after adjusting for all covariates (Table 3; Figure 3A). A regional plot for the section of chromosome 17 that contains five significant DNAm sites is presented in Figure 3B. Quantile-quantile analysis of the expected and observed p-values from the meta-analysis for serum albumin showed very moderate global inflation (inflation factor of 1.02) of the nominal p-values (Figure 3C), so no further corrections were applied. Notably, among the top one hundred DNAm sites most strongly associated with (i.e., with the lowest p-values) serum albumin in the meta-analysis, the beta coefficients obtained from the two EWAS sub-cohorts (the EPIC and 450K) show a strong, positive correlation (beta = 0.89, p-value < 0.001) (Figure 3D).

Four of the remaining five DNAm sites—cg18181703, cg03546163, cg16936953, and cg19842579—were significantly associated with serum albumin in the adjusted model (Table 3).
transaminase, alanine transaminase, serum albumin, total bilirubin, platelet count, FIB-4 score, and APRI score—among male United States veterans with HIV. We identified that nine DNA methylation sites mapped to the genes TMEM49, SOCS3, FKBP5, ZEB2, and SAMD14 were significantly associated with serum albumin levels. We also demonstrate an association between higher PhenoAA (an aging marker for DNA methylation) and low serum levels of albumin.

Serum albumin is a highly abundant protein primarily synthesized by the liver. Synthesis of albumin occurs in polysomes of hepatocytes at a rate of 10 g/day–15 g/day and accounts of 10% of liver protein synthesis (Garcovich et al., 2009). Albumin is not stored in the liver and therefore is not released on demand. Instead in situations of increased need in healthy adults, hepatocytes can increase albumin synthesis by up to 300% (Garcovich et al., 2009). Impaired albumin synthesis and function have been reported in many liver diseases (Carvalho and Machado, 2018), and low serum levels of albumin may be a useful indicator of LD. Several studies have shown that serum albumin is an important prognostic factor and a significant predictor of death in patients with cirrhosis (Carvalho and Machado, 2018) and also a predictor of serious non-AIDS events in PWH (Ronit et al., 2018a; Ronit et al., 2018b).

Four of the nine serum albumin associated DNA methylation sites are located near the TMEM49 gene (also known as VMP1). DNA methylation site TMEM49-cg16936953 was also associated with HIV infection, C-reactive protein and inflammatory bowel disease (Gross et al., 2016; Ligthart et al., 2016; Venthem et al., 2016). TMEM49 encodes a transmembrane protein that drives cellular

FIGURE 3
(A) Manhattan plot of unadjusted p-values from meta-analysis of EWAS results for serum albumin. The red line indicates the unadjusted p-value that corresponds to a threshold for FDR significance at Q < 0.05. DNA methylation was not significantly associated with serum albumin at any DNA methylation site after a more restrictive Bonferroni correction to an overall α = 0.05. (B) Regional Manhattan plot of unadjusted p-values from meta-analysis of EWAS results for CpG sites in chromosome 17 and serum albumin. The red line indicates the unadjusted p-value that corresponds to a threshold for FDR significance (among all CpG sites from across entire genome) at Q < 0.05. DNA methylation was not significantly associated with serum for any CpG site after a more restrictive Bonferroni correction to an overall α = 0.05. (C) Quantile-quantile plot of unadjusted p-values from meta-analysis of EWAS results for serum albumin. The global inflation factor was 1.02. Red lines represent a distribution of null associations with a 95% confidence interval. (D) Correlation of beta coefficients obtained from separate EWAS of serum albumin among the EPIC and 450K cohorts for the 100 DNA methylation sites with the smallest unadjusted p-values after meta-analysis. Beta coefficients from the separate analyses are positively correlated (r = 0.89, p-value < 0.001), indicating that the signs of observed beta coefficient are consistent between the two datasets. Circles representing p-value pairs that were significant after meta-analysis and FDR correction to Q < 0.05 are highlighted in red.
autophagy (Molejon et al., 2013) and is overexpressed in pancreatitis-affected acinar cells (Vaccaro et al., 2003). Cellular autophagy, therefore, may be linked to regulation of serum albumin. Previous studies have shown that serum albumin suppresses autophagy via mTOR activation and that depriving cultured cells of serum albumin induces autophagy which is protective against the accumulation of harmful reactive oxygen species (ROS).

SOCS3 encodes a protein that inhibits cytokine signaling in the STAT pathway in response to elevated cytokine levels (Jo et al., 2005; Carow and Rottenberg, 2014). Suppression of SOCS3 contributes to liver fibrosis by increasing fibrogenic signaling via STAT3-mediated upregulation of tissue growth factor (TGF)-β (Ogata et al., 2006; Jadid et al., 2018; Dees et al., 2020). In addition, obesity has been associated with downregulation of SOCS3, which raises the possibility that an association between DNAm near SOCS3 and liver dysfunction could result from the well described relationship between obesity and liver dysfunction. DNAm site SOCS3-cg18181703 was associated with BMI in several recent EWAS (Ali et al., 2016; Mendelson et al., 2017; Wahl et al., 2017; Xu et al., 2018; Sun et al., 2019). In this cohort of PWH, SOCS3-cg18181703 was associated with BMI (p-value:0.044) further supporting this hypothesis. Epigenetic associations between SOCS3-cg18181703 and C-reactive protein, inflammatory bowel disease, type-2 diabetes and cognitive abilities (Chambers et al., 2015; Ligthart et al., 2016; Marioni et al., 2018; Juvinao-Quintero et al., 2021) also suggest potential pleiotropic effects of SOCS3 gene.

FKBP5 encodes an immunoregulatory protein that contributes to basic protein folding and trafficking (Zannas et al., 2016). FKBP5 has been demonstrated to positively regulate the stress response and drive acquisition of metabolic disorders including obesity, insulin resistance, and diabetes (Sidibeh et al., 2018). Moreover, FKBP5 is thought to contribute to liver dysfunction (Kusumanchi et al., 2021) and in one study, deletion of FKBP5 protected knock-out mice from fatty liver disease despite high fat diets (Stechschulte et al., 2016). ZEB2 encodes a zinc-finger DNA-binding protein expressed in hepatocytes (Cai et al., 2012). Studies have demonstrated that repression of ZEB2 expression by microRNAs induces apoptosis in hepatocytes (Zhao et al., 2018), suggesting that ZEB2 might influence preservation of liver integrity in the face of hepatocellular damage. The remaining DNAm site associated positively with serum albumin levels was located near the SMAD14 gene. The function of the SMAD14 gene is unknown, but it has been linked to gastric cancer and hypothesized to act as a tumor suppressor gene (Xu et al., 2020). No studies have associated SMAD14 or its tumor suppressor properties to either general liver health and function or specifically to serum albumin regulation. Despite the association between PhenoAA and serum albumin, no other significant associations were noted between the four assessed metrics of DNAm AA—PhenoAA, GrimAA, IEAA, and EEAA—and other biomarkers. Compared to the other metrics of DNAm aging, PhenoAA is unique in that it is highly predictive of morbidity and mortality even after adjusting for chronological age (Liu et al., 2019). This may explain its significant association with albumin, which is also a prognostic marker of survival in LD (Carvalho and Verdelho Machado, 2018). It is plausible that specific DNAm sites may regulate the expression of genes in a manner that influences behavior of the selected biomarkers of liver health, either by impacting the progression of conditions that damage the liver and inhibit function—like fibrosis or hepatocellular carcinoma—or by affecting pathways that are specifically involved in regulation of a specific liver biomarkers. Low serum level of albumin, though an important prognostic biomarker for LD (Carvalho and Verdelho Machado, 2018), is not specific. Thus, the associations we have uncovered in this study may be mediated by other non-hepatic factors in PWH. For example, hypoalbuminemia is associated with inflammation and synthesis of albumin is suppressed by pro-inflammatory cytokines TNF-α and Interleukin-1 (Dinarello, 1984; Perlmutter et al., 1986), both of which are important mediators of chronic inflammation in PWH (Deeks et al., 2013; De Pablo-Bernal et al., 2014). It is thus presumptuous to infer that the association between DNAm and albumin reflects underlying LD alone and is not combination of multiple interacting factors.

Strengths and limitations

This study combines data from two different microarray platforms to assess consistent epigenetic associations of overlapping DNAm sites. This enabled a robust meta-analysis of epigenetic associations with a large sample size. The significant associations are likely to be true positive findings, and should be replicated in future studies. Secondly, we examine multiple biomarkers which when combined reflect overall liver function and individually inform on the liver’s capacity to perform specific metabolic functions. This allowed us to examine DNAm sites across a wider range of genes that may link to specific aspects of liver function. Thirdly, unlike other EWAS of liver function or LD, our cohort included exclusively people with HIV, a group which has a higher burden of liver dysfunction and related morbidity. Our findings contribute to the identification of genes that may be relevant to liver health and function in the context of HIV infection.

One limitation of our study is that EWAS discovery was sub-optimally powered for a subset of DNAm sites included in only one of the platforms. Meta-analysis results from DNAm
sites measured on both platforms were highly consistent, suggesting that combining data from the two platforms successfully augments the power to detect significant associations. Future replication and meta-analysis with extended epigenomic coverage would improve the discovery of liver function-associated DNAm sites. Secondly, our study population was restricted to male veterans with HIV, limiting the generalizability of our findings to women with HIV. Thirdly, we used DNAm levels measured in blood cells to assess associations between DNAm and biomarkers of liver function, which may not represent DNAm patterns in the liver cells that are responsible for regulating these biomarkers. The lack of significant associations between DNAm and AST, ALT, total serum bilirubin, platelet count, FIB-4 score and, APRI score might result from an absence of epigenetic associations with liver functions of large effects, suggesting better powered future studies are needed to identify epigenetic associations with liver functions with smaller effects. Also, the potential for misclassifying liver fibrosis with the use of clinical scores like FIB-4 and APRI as has been shown in hepatitis B infection (Kim et al., 2016b) and may have limited our ability to detect an association with these markers.

Conclusion

We identified epigenetic associations of both individual DNAm sites and DNAm AA with liver function through serum albumin in men with HIV. EWAS may inform on disease pathogenesis and generate new hypotheses for predicting disease progression and treatment of LD in HIV. Identifying specific genes linked with liver function among PWH can inform disease progression to improve the health of this patient population. Further replication analyses in independent cohorts are warranted to confirm and improve the discovery of the epigenetic markers of liver function and LD in PWH.

Data availability statement

The data analyzed in this study is subject to the following licenses/restrictions: Due to US Department of Veterans Affairs (VA) regulations and our ethics agreements, the analytic data sets used for this study are not permitted to leave the VA firewall without a Data Use Agreement. This limitation is consistent with other studies based on VA data. However, VA data are made freely available to researchers with an approved VA study protocol. For more information, please visit https://www.virec.research.va.gov or contact the VA Information Resource Center at VIREC@va.gov. Requests to access these datasets should be directed to yan.v.sun@emory.edu.

Ethics statement

The datasets for this article are not publicly available due to concerns of ethics agreement with the US Department of Veterans Affairs (VA). These data are part of the VACS database. Requests to access these datasets should be directed to the corresponding author, YS, yan.v.sun@emory.edu. The institutional review board approved the original VACS cohort, and all participants provided written informed consent.

Author contributions

BT and ML—Original manuscript draft and data analysis ZW, JC, QH—Data analysis VR, KA, KX, MF, MG, VM, AJ—Manuscript revision YS—Study design, data and, analysis and, manuscript revision.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene.2022.1020871/full#supplementary-material
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