Case Report: Imaging Features of Gallbladder Sessile Polyp Confirmed by Contrast-Enhanced Ultrasonography and Dynamic Computed Tomography in a Dog With Asymptomatic Chronic Cholecystitis

Jeongmin Lee¹, Jinsu Kang², Suyoung Heo², Kichang Lee¹ and Hakyoung Yoon¹*

¹ Department of Veterinary Medical Imaging, College of Veterinary Medicine, Jeonbuk National University, Iksan-si, South Korea, ² Department of Surgery, College of Veterinary Medicine, Jeonbuk National University, Iksan-si, South Korea

A 6-year-old dog was presented for health screening. It never suffered from any disease. Ultrasonography confirmed mass-like thickening with irregular margins protruding toward the gallbladder (GB) lumen. On contrast-enhanced ultrasonography (CEUS) and dynamic computed tomography (CT), contrast enhancement of the corresponding structures was confirmed in arterial phase. After cholecystectomy, cauliflower-like sessile polyps were identified. Histopathological examination revealed chronic lymphoplasmacytic cholecystitis. Bile culture revealed Escherichia coli growth. Our novel findings suggest that chronic cholecystitis should be considered as a differential diagnosis if contrast-enhanced sessile polyps of the GB are found on CEUS and dynamic CT.

Keywords: biliary system, canine, coprophagia, gallbladder polyp, ultrasound

INTRODUCTION

In veterinary medicine, there are few reports on the imaging features of bacterial cholecystitis (1, 2). Furthermore, a previous study showed that gallbladder (GB) polyps or mass can indicate neoplasia including GB adenoma or adenocarcinoma (3, 4); cholecystitis may also be involved (5). Therefore, although it is important to diagnose polyp in the GB, it is difficult to accurately distinguish it from wall-attached sludge only by ultrasonography. In this regard, there were studies using contrast-enhanced ultrasonography (CEUS) to diagnose GB polyp or mass (4, 5). However, based on our literature review, there are no reports that describe the imaging features of GB polyps using dynamic computed tomography (CT) and there are no reports to simultaneously describe CEUS and dynamic CT imaging features of chronic cholecystitis with sessile polyps of the dog. Therefore, we aimed to describe a case wherein GB polyp was diagnosed using CEUS and dynamic CT and we compared the advantages of the two modalities.

CASE DESCRIPTION

A 6-year-old, spayed female beagle dog was admitted to the Jeonbuk National University Animal Medical Center for health screening. It showed no clinical signs and had never suffered from any disease but practiced coprophagia. No abnormal findings were observed in blood chemistry test,
but a complete blood count revealed leukocytopenia (3.5 × 10^9 cells/L; reference range, 6–17 × 10^9 cells/L).

There were no remarkable findings except for splenomegaly in the chest (four radiographic views) and abdominal radiography (right lateral and ventrodorsal views) (HF-525PLUS, Ecoray, Seoul, Korea). On abdominal ultrasonography (Aplio 300, Canon Medical System, Europe B.V., Zoetermeer, Netherlands), mass-like thickening with irregular margins protruding into the lumen was noted, affecting large portions of the GB, particularly in the region of the neck. These were more echogenic than the contents of the GB lumen, which comprised some anechoic fluid surrounding a rounded organized content that is slightly more echogenic (Figure 1A). Additionally, hypoechoic splenic nodules were observed. Subsequently, CEUS of the GB was performed using SonoVue® (Bracco, Milan, Italy) at a dosage of 0.03 ml/kg. The wash-in for the protruding structures within the GB occurred 9 s after contrast medium (CM) injection. Maximum contrast enhancement (CE) was observed 13 s after CM injection (Figure 1B, left panel). Wash-out occurred 55 s after CM injection. One week later, dynamic CT scan was performed on the GB using a 16-row multi-detector CT scanner (Axelion, TSX-034A, Canon medical system Europe B.V., Zoetermeer, Netherlands) under general anesthesia with isoflurane. As contrast agent, iohexol (Omnipaque 300, GE Healthcare, Shanghai, China) was used at a dosage of 750 mg/kg. CT scan was taken in the following order: pre-contrast CT scan, dynamic CT scan on GB (imaging during 0–34 s after CM administration, four cross-sections), portal phase CT scan (imaging starts 40 s after CM administration, whole body), and delayed phase CT scan (imaging starts 140 s after CM administration, whole body) (kV: 100, mAs: 120, slice thickness: 1 mm). In pre-contrast images, the Hounsfield unit (HU) of the protruding structures within the GB was ∼30. Wash-in was confirmed for that structure (HU: 50) more than 20 s after CM injection. More intense and heterogeneous CE was confirmed 34 s after CM injection (HU: 160–270). The dynamic CT scan was stopped due to prolonged breath-holding, and portal and delayed phase CT scans were then performed. CE was confirmed to weaken over time; HU was 120 after 58 s since CM injection and 107 after 159 s (Figure 2). As per diagnostic imaging, vascularized GB polyps were considered. Differential diagnoses for this lesion included inflammatory polyps, mucosal hyperplasia, or neoplasia (adenoma, adenocarcinoma, and carcinoid in situ) (4–8). Laparotomy and cholecystectomy were performed considering the fact that the possibility that the lesion observed in the GB was neoplasia could not be excluded, and in human medicine, that asymptomatic gangrenous cholecystitis or emphysematous cholecystitis, which can be life-threatening, may occur. The bile was collected aseptically after the CT scans. At the time of surgery, there were no abnormal findings in the abdomen, including GB rupture or bile leakage. After GB incision, viscous substances and cauliflower-like sessile polyps were confirmed around the cystic duct (Figure 3A). Bile cytology revealed a number of bacilli (Figure 3B). GB wall was collected and fixed in 10% buffered formalin for histopathological analysis. No gross lesions were found in the liver; therefore, a biopsy was performed at one random site in the quadrate lobe. Histopathological examination of the GB revealed chronic lymphoplasmacytic cholecystitis and mucocele (Figure 4A). Additionally, mildly vacuolated hepatocytes were observed in the liver (Figure 4B). Escherichia coli was identified as a result of bile culture. The splenic nodules were diagnosed as benign nodular hyperplasia using fine needle aspiration.

The patient was discharged after 1 week of postoperative management. Antibiotics were prescribed for cultured biliary E. coli for 1 week: amoxicillin-clavulanic acid (13.75 mg/kg, twice daily) and metronidazole (15 mg/kg, twice daily). Except for mild increase in liver enzyme levels immediately after liver biopsy, no abnormalities were observed on physical examination, laboratory blood tests, or ultrasonography for 3 months.

**DISCUSSION**

In dogs, echogenic contents within the dog's GB are often diagnosed as sludge or mucocele (9). However, in this case, hypoechoic mass-like protruding structures were observed, which were distinguished from the rounded echogenic organized contents. Therefore, it was necessary to ascertain whether this structure was a tissue receiving blood flow or if it was a sludge or mucocele. Consequently, lack of CE of the GB wall, or a finding of rupture or necrosis, was not observed in CEUS (10). However, GB neoplasia was possible because CE of the protruding structure of the GB was observed in arterial phase (4, 5); it was necessary to determine whether the mass invaded into the adjacent tissues around the GB such as liver, or had metastasized to other organs (11). If the CT revealed findings including infiltration of the mass to the organs outside the GB, extensive resection and biopsy of insulated organs would have been necessary. However, in the arterial phase of the dynamic CT scan, the GB wall and mass-like structures of the GB were clearly distinguished from the surrounding tissue, and therefore, the possibility of invasion into the adjacent tissue such as liver or mesenteric fat was considered low. In the portal and delayed phase, this distinction was unclear, but there were no suspicious findings of metastasis; thus, only the GB was resected.

In the previous literature characterizing imaging features of cholecystitis through CT, it is explained that in acute biliary inflammation, a thickened GB wall and bile ducts are enhanced in the late arterial phase, while in chronic biliary inflammation, a thickened or mineralized GB wall is enhanced in delayed phase (12). Additionally, neoplastic lesions can show strong CE in arterial phase in dogs (5, 13); there is also a report that human adenocarcinoma shows strong CE in the arterial phase in CEUS (4). GB adenoma is reported in a human with cauliflower-like sessile polyps (3). However, herein, the GB sessile polyps showed clear CE in arterial phase on CEUS and dynamic CT scans, but were diagnosed as chronic cholecystitis upon histopathological examination, not as acute inflammation.

**Abbreviations:** GB, gallbladder; CEUS, contrast-enhanced ultrasonography; CT, computed tomography; CM, contrast medium; CE, contrast enhancement; E. coli, Escherichia coli.
FIGURE 1 | Ultrasonography (A) and contrast-enhanced ultrasonography (CEUS) (B) of the gallbladder (GB). (A) The hyperechoic structure (arrowheads) protruding toward the GB lumen appear more echogenic than GB contents (asterisk). (B) Strong CE of this structure is confirmed in CEUS 13 s after contrast medium injection. GB contents were not enhanced.
or neoplasia. Thus, chronic cholecystitis should be included in the differential diagnosis, if arterial CE with GB polyps protruding toward the lumen is confirmed. It was also reported that GB mucocele has a significantly higher HU value than GB sludge (14). However, herein, GB contents had low HU close to sludge, but as a result of histopathology, it was diagnosed as GB mucocele. This false negative was because although GB mucocele has a high HU value, the deviation is very large, and the HU value between sludge and mucocele has a large overlap (14).

CE of the GB sessile polyps was observed using two modalities. CEUS is more advantageous than dynamic CT as the lesion can be imaged without anesthesia. Additionally, it took longer in CT for CE of the lesion, presumably because cardiovascular function was suppressed by isoflurane (15). When dynamic CT scan was performed, the heart rate decreased from 80 to 46 beats per minute (BPM) during anesthesia. However, there was no bradycardia, and heart rate increased to 120 BPM after atropine administration before surgery. Another limitation of dynamic CT was that the motion artifact due to diaphragm movement had to be reduced through breath control, which stopped exhalation for an extended time period. However, unlike CEUS, which evaluates one cross-section, dynamic CT has the advantage of simultaneously evaluating multiple cross-sections and can evaluate wide area around the GB. CT also has the advantage of being able to evaluate metastases in the whole body.

Herein, chronic cholecystitis did not cause clinical signs, and blood tests were normal except for leukocytopenia. In humans, there are case reports of asymptomatic gangrenous cholecystitis and emphysematous cholecystitis (16, 17). However, CEUS and dynamic CT provide good imaging for the GB sessile polyps; thus, they could help diagnose asymptomatic polyloid chronic cholecystitis.

Escherichia coli was detected in the bile, and the patient had practiced coprophagia for several years. To the best of our knowledge, no studies have clearly established the relationship between coprophagia and chronic cholecystitis. Additionally, the etiology of bacterial cholecystitis and its correlation with cholangitis are not clearly established (1). However, there are studies suggesting that ascending infection from the gastrointestinal tract (GIT) and hematogenous infection from sepsis cause bacterial cholecystitis (1, 18). Fecal pathogens can infect the dog through coprophagia (19). Moreover, there was a case report wherein Cyniclomyces guttulatus was cultured in the bile of a dog with cholecystitis (2). This yeast is a normal flora of rabbit GIT, and coprophagia preserves it (20). It can also cause opportunistic infections in the GIT of dogs ingesting rabbit feces (20). Because she had no history of diseases including sepsis, ascending E. coli infection from the GIT is the likely cause of her chronic cholecystitis. The possibility that coprophagia may have contributed to this infection cannot be completely excluded, however.

The patient had mild leukocytopenia of unknown cause, which may have caused cholecystitis. Although dogs with neutropenia may not display obvious clinical signs, they are known to be susceptible to bacterial and fungal infections (21).

The liver showed mild vacuolar degeneration. Considering that cholecystitis may be associated with decreased GB contractility [(22), #17], vacuolar hepatopathy could be caused...
FIGURE 3 | Gross lesion of the inner surface of the resected gall bladder (A), and cytology of the bile (B). Diff quick staining, 1,000× magnification. (A) Cauliflower-like sessile polyps are observed. (B) A number of bacilli observed in the bile. Bile culture revealed these to be E. coli.
by cholecystitis. However, vacuolar hepatopathy is a non-specific change in the liver, and may not be clinically significant. This change can be caused by diabetes mellitus, hypothyroidism, hyperadrenocorticism, and stress conditions (23–25); laboratory blood and urine tests ruled out the above endocrinopathy in this case.

This study has several limitations. First, wash-out of the CM could not be observed on dynamic CT due to prolonged breath-holding. A method for reducing motion artifacts using the predictive respiratory gating system in humans is studied (26); however, to the best of our knowledge, there are no studies on GB imaging using this technique in dogs; further studies are needed to overcome these artifacts. Second, the etiology for this patient is incompletely elucidated. The dog had no history of systemic disease; thus, an ascending infection of *E. coli* from the GIT might have caused cholecystitis. However, we cannot clarify whether coprophagia was involved in the occurrence of chronic cholecystitis; further investigations are warranted.
In conclusion, chronic cholecystitis should be considered when contrast-enhanced structures protruding toward the GB lumen are confirmed through CEUS or dynamic CT scan in arterial phase. In this regard, CEUS has the advantage of imaging the lesion without anesthesia, whereas dynamic CT has the advantage of evaluating a wider area including the gallbladder and adjacent structures. This is expected to help in planning the surgical site and selecting the target organ for biopsy. Since both modalities image lesions well, a more useful modality can be selected and used in consideration of various circumstances, including the patient’s condition. Additionally, sessile polyps caused by chronic cholecystitis can mimic neoplasia and cholecystitis, which can cause GB wall thickening or CE in arterial phase (3, 5, 12). Therefore, it appears that histology is necessary for differential diagnosis.

**DATA AVAILABILITY STATEMENT**

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author(s).

**REFERENCES**

1. Tamborini A, Jahns H, McAllister H, Kent A, Harris B, Procoli F, et al. Bacterial cholangitis, cholecystitis, or both in dogs. *J Vet Intern Med.* (2016) 30:1046–55. doi: 10.1111/jvim.13974
2. Neel JA, Tarigo J, Grindem CB. Gallbladder aspirate from a dog. *Vet Clin Pathol.* (2006) 35:467–70. doi: 10.1111/j.1939-165X.2006.tb00167.x
3. Petrović B, Petrović A, Živković V, Malina M. Intestinal type of villous adenoma of gallbladder. *Acta Med Med.* (2010) 49:50–4. Available online at: https://publisher.medfak.ni.ac.rs/AMM_1/amm-stari/2010-3-broj/Bratislava%20Petrovic-Intestinal%20type%20of%20villous%20adenoma...50-54.pdf
4. Wang H, Ling W, Luo YJM. Contrast-enhanced ultrasound findings of gallbladder adenocarcinoma with sarcomatoid carcinoma accompanied by intrahepatic metastasis: a case report and literature review. *Medicine.* (2018) 97:e10773. doi: 10.1097/MD.0000000000010773
5. Bargellini P, Orlandi R, Paloni C, Rubini G, Fonti P, Righi C, et al. Contrast-enhanced ultrasound complements two-dimensional ultrasonography in diagnosing gallbladder diseases in dogs. *Vet Radiol Ultrasound.* (2018) 59:345–56. doi: 10.1111/vru.12601
6. Mellinck VM, Menias CO, Sandrasagar K, Hara AK, Kielar AZ, Brunt EM, et al. Polyoid lesions of the gallbladder: disease spectrum with pathological correlation. *Radiographics.* (2015) 35:387–99. doi: 10.1148/rg.3521400995
7. Bhandal J, Head LL, Francis DA, Foster RA, Berrington A. Use of color flow Doppler ultrasonography to diagnose a bleeding neuroendocrine tumor in chronically instrumented dogs. *Vet Pathol.* (2019) 60:689–95. doi: 10.1111/vru.12806
8. Morrell C, Volk M, Mankowski JJ. A carcinoid tumor in the gallbladder of a dog. *J Am Anim Hosp Assoc.* (2008) 44:195–8. doi: 10.1586/10799360800000010773
9. Bhandal J, Head LL, Francis DA, Foster RA, Berrington A. Use of color flow Doppler ultrasonography to diagnose a bleeding neuroendocrine tumor in chronically instrumented dogs. *Vet Pathol.* (2019) 60:689–95. doi: 10.1111/vru.12806
10. Aissi A, Slimani CJ. Ultrasound diagnosis of cholecystitis in a dog (A case report). *Glob Vet.* (2009) 3:514–5. Available online at: http://www.idosi.org/gv/v3i6/09/13.pdf
11. Baan M, Kidder AC, Johnson SE, Sherding RG. Rhinoscopic diagnosis of Equus caballus boehmi infection in a dog. *J Am Anim Hosp Assoc.* (2011) 47:60–3. doi: 10.5326/JAAHA-MS-5707
12. Bertolini G. *Body MDCT in Small Animals: Basic Principles, Technology, and Clinical Applications.* Cham: Springer (2017). p. 134–5. doi: 10.1007/978-3-319-46904-1
13. Brand EM, Lim CK, Heng HG, Gross FV, Hanlon J, Jones-Hall Y. Computed tomographic features of confirmed gallbladder pathology in 34 dogs. *Vet Radiol Ultrasound.* (2020) 61:667–79. doi: 10.1111/vru.12909
14. Fuerst IA, Hostnik ET. CT attenuation values and mineral distribution can be used to differentiate dogs with and without gallbladder mucocceles. *Vet Radiol Ultrasound.* (2019) 60:689–95. doi: 10.1111/vru.12601
15. Bernard J-M, Wouters PF, Doursout M-F, Florence B, Chey JE, Merin RG. Effects of sevoflurane and isoflurane on cardiac and coronary dynamics in chronically instrumented dogs. *J Am Soc Anesthesiol.* (1990) 72:659–62. doi: 10.1097/00005422-199004000-00014
16. Chaudhry S, Hussain R, Rajasundaram R, Corless D. Gangrenous cholecystitis in an asymptomatic patient found during an elective laparoscopic cholecystectomy: a case report. *J Med Case Rep.* (2011) 5:1–3. doi: 10.1186/1752-1947-5-199
17. Ohno H, Ishida H, Komatsuda T, Matsuda A, Watanabe T. Asymptomatic emphysematous cholecystitis. *J Med Ultrasound.* (2009) 36:153–6. doi: 10.1111/vru.12601
18. Bertolini G. *Body MDCT in Small Animals: Basic Principles, Technology, and Clinical Applications.* Cham: Springer (2017). p. 134–5. doi: 10.1007/978-3-319-46904-1
19. Baan M, Kidder AC, Johnson SE, Sherding RG. Rhinoscopic diagnosis of Equus caballus boehmi infection in a dog. *J Am Anim Hosp Assoc.* (2011) 47:60–3. doi: 10.5326/JAAHA-MS-5707
20. Winston JA, Piperisova I, Neel J, Gookin JL. Cynicoliomycetes guttulatus infection in dogs: 19 cases (2006-2013). *J Am Anim Hosp Assoc.* (2016) 52:42–51. doi: 10.5326/JAAHA-MS-6307
21. Day MJ, Mackin A, Littlewood JD. *BSAVA Manual of Canine and Feline Haematology and Transfusion Medicine.* Chedegley: British Small Animal Veterinary Association (2000). p. 117–9.
22. Brugge WR, Brand DL, Atkins HL, Lane BP, Abel WG. Gallbladder dyskinesia in chronic acalculous cholecystitis. *Dig Dis Sci.* (1986) 31:461–7. doi: 10.1007/BF01320308
23. Zachary JF, McGavin MD. *Pathologic Basis of Veterinary Disease.* 5th ed. Amsterdam: Elsevier Health Sciences (2012). p. 449–50.

**ETHICS STATEMENT**

Ethical review and approval was not required for the animal study because, this study is a case report of examinations, surgeries, and biopsies performed for the purpose of treatment of patients, and no action contrary to treatment was performed. Written informed consent was obtained from the owners for the participation of their animals in this study.

**AUTHOR CONTRIBUTIONS**

JL and HY: conception and design and drafting the article. JL, JK, SH, and HY: acquisition of data. JL, JK, SH, KL, and HY: analysis and interpretation of data, revising article for intellectual content, and final approval of the completed article. All authors contributed to the article and approved the submitted version.

**FUNDING**

The research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (2019R1A6A1A03033084).
24. Sepesy LM, Center SA, Randolph JE, Warner KL, Erb HN. Vacuolar hepatopathy in dogs: 336 cases (1993-2005). *J Am Vet Med Assoc.* (2006) 229:246–52. doi: 10.2460/javma.229.2.246

25. Peyron C, Lecoindre P, Chevallier M, Guerret S. Vacuolar hepatopathy in 43 French Scottish Terriers: a morphological study. *Rev Méd Vét.* (2015) 166:176–84. doi: 10.13140/RG.2.1.1167.9445

26. Ritchie CJ, Hsieh J, Gard MF, Godwin JD, Kim Y, Crawford CR. Predictive respiratory gating: a new method to reduce motion artifacts on CT scans. *Radiology.* (1994) 190:847–52. doi: 10.1148/radiology.190.3.8115638

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher’s Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Lee, Kang, Heo, Lee and Yoon. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.