Evaluating the role of common risk variation in the recurrence risk of schizophrenia in multiplex schizophrenia families

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INTRODUCTION

Schizophrenia (SCZ) is a severe, clinically heterogeneous psychiatric disorder with a population prevalence of ~1% [1]. Twin, family, and adoption studies consistently show a strong genetic component, with heritability estimates of around 0.75–0.80 [2–6], and family history (FH) remains the strongest risk factor for developing SCZ [7]. Despite high heritability, ~2/3 of SCZ cases report no FH of psychotic illness, and most subjects with a positive FH (FH+) report only a single affected relative [8, 9], concordant with the rates of 31% FH+ and 69% family history negative (FH−) observed in the sample of sporadic SCZ cases analyzed in this study [10].

The Irish Study of High-Density Schizophrenia Families sample (ISHDSF) [11–14] consists of 257 multiplex SCZ families with genotype data, ascertained to have two or more first-degree relatives meeting the Diagnostic and Statistical Manual of Mental Disorders (DSM-III-R) criteria for SCZ or poor-outcome schizoaffective disorder. Such multiplex families, display substantially higher recurrence risk of SCZ than reported in sporadic cases [8, 9], and this discrepancy in recurrence risk suggests that there may be important differences in the genetic architecture between familial and sporadic SCZ cases that warrant further investigation.

One explanation of this difference is that familial SCZ cases may carry a higher burden of common SCZ risk variation as measured by a higher SCZ polygenic risk score (PRS), than ancestry matched sporadic cases. Another explanation is that the increased recurrence risk in multiplex families may be attributable to segregation of rarer, higher risk variation, identifiable through exome or whole-genome sequencing likely in combination with common risk variation. Sequencing studies suggest that rare, deleterious variation in the genome is involved in the genetic etiology of SCZ and other psychiatric disorders [15–22], but the extent to which rare variation contributes to SCZ risk in multiplex families is currently unknown. A third hypothesis, not addressed here, is that familial cases may have increased exposure to environmental risks unique to the families that may explain the higher recurrence risk in multiplex families.

Mega-analyses of SCZ genome-wide association study (GWAS) data by the Psychiatric Genomics Consortium Schizophrenia Working Group (PGC-SCZ) have identified 287 loci associated...
with SCZ [23–25]. GWAS data from such studies are frequently used to construct PRS to index an individual's common genetic variant risk for a disorder. Although current PRS currently lack power to predict SCZ in the general population, they have been shown to index meaningful differences in SCZ liability between individuals. For example, in the European PGC3-SCZ sample, the highest PRS centile has an OR of 44 (95% CI = 31–63) for SCZ compared to the lowest centile of PRS, and OR of 7 (95% CI = 5.8–8.3) when the top centile is compared with the remaining 99% of the individuals in the sample [25].

Common risk variation analyses in multiplex family samples smaller than ISHDSF have been performed [26–28], and we have previously used the summary statistics from the first wave of PGC-SCZ mega-analysis [23] to investigate whether the concept of the genetically influenced psychosis spectrum is supported by empirical data in multiplex SCZ families [29]. Here, we extend our previous work by using PRS profiling in multiplex SCZ families, sporadic SCZ cases and population controls, all from the population of the island of Ireland, to directly test whether common SCZ risk variation in the genome may explain the increased recurrence risk of SCZ in multiplex families. Identifying the source of the increased familial recurrence risk of SCZ is important for future research into the genetic etiology of familial SCZ, and potentially for both diagnosis and treatment of SCZ with different familial backgrounds, as it will determine the relative focus on environmental exposures, as well as common and rare genetic variation in case-control and family studies of SCZ.

**METHODS**

**Sample description**

**Irish study of high-density schizophrenia families (ISHDSF).** Fieldwork for the ISHDSF sample were carried out between 1987 and 1992, with probands ascertained from public psychiatric hospitals in the Republic of Ireland and Northern Ireland, with approval from local ethics committees [30]. Inclusion criteria were two or more first-degree relatives meeting DSM-III-R criteria for SCZ or poor-outcome schizoaffective disorder (PO-SAD), with all four grandparents being born in Ireland or the United Kingdom. Relatives of probands suspected of having psychotic illness were interviewed by trained psychiatrists, and trained social worked interviewed other relatives. Hospital and out-patient records were obtained and abstracted in >98% of cases with SCZ or PO-SAD diagnoses. To avoid bias and detect possible mistakes in diagnosis, an independent review of all diagnostic information such as interview, family history reports, and hospital information was made blind to family assignments by two trained psychiatrists, with each psychiatrists making up to three best estimate DSM-III-R diagnoses, with high agreement between the two psychiatrists (weighted k = 0.94 ± 0.05).

The concentric diagnostic schema of the ISHDSF shown in Table 1 and Supplementary Fig. 4, includes four case definitions: narrow spectrum (SCZ, PO-SAD and simple SCZ), intermediate spectrum (adding schizotypal personality disorder, schizoaffective disorder, and delusional disorder, psychosis not otherwise specified, and good-outcome schizoaffective disorder), broad spectrum (adding psychotic affective illness, paranoid, avoidant and schizoid personality disorders, and other disorders that significantly aggregate in relatives of probands based on previous epidemiological work in Ireland [12] and very broad spectrum (adding any other psychiatric illness in the families). The ISHDSF sample also includes unaffected family members with no diagnosis of any psychiatric illness. The ISHDSF diagnostic schema is described extensively elsewhere [31].

**Irish schizophrenia genomics consortium case/control sample (ISGC).** The ISGC sample was assembled for a GWAS of SCZ in Ireland. Details of recruitment, screening and quality control (QC) methods used for the ISGC sample have been previously described in detail elsewhere [32]. Briefly, the case sample was recruited through community mental health service and inpatient units in the Republic of Ireland and Northern Ireland following protocols with local ethics approval. All participants were interviewed using a structured clinical interview for DSM-III-R and DSM-IV, were over 18 years of age and reported all four grandparents born either in Ireland or the United Kingdom. Cases were screened to exclude substance-induced psychotic disorder or psychosis due to a general medical condition. A subset of sporadic cases sampled by Virginia Commonwealth University (N = 745) have genotypic data and FH information available [10] from completion of the family history research diagnostic criteria (FH-RDC) interview [33]. This includes 233 (~31%) FH+ cases and 512 (~69%) FH– cases, in close concordance with the other large meta-analyses [8, 9]. Controls from the Irish Biobank used in ISGC were blood donors from the Irish Blood Transfusion Service recruited in the Republic of Ireland. Inclusion criteria were all four grandparents born in Ireland or the United Kingdom and no reported history of psychotic illness. Due to the relatively low lifetime prevalence of SCZ, misclassification of controls should have minimal impact on power [34].

**Genotyping and QC**

Samples were genotyped using three different arrays (Supplementary Table 2). 830 individuals representing 237 families from the ISHDSF sample were genotyped on the Illumina 610-Quad Array. An additional 175 ISHDSF individuals from 52 families were later genotyped on the Infinium PsychArray V.1.13 Array. For the case-control sample, 1627 sporadic cases and 1730 controls were successfully genotyped using the Affymetrix V6.0 Array, either at the Broad Institute or by Affymetrix. An additional 487 sporadic cases and 475 controls were later genotyped on the PsychArray along with the additional ISHDSF individuals described above. The same QC protocols were applied to all three datasets and full details are described elsewhere for ISHDSF [31] and the case-control sample [32]. Exclusion criteria for samples were a call rate of <95%, more than one Mendelian error in the ISHDF sample, and difference between reported and genotypic sex. Exclusion criteria for SNPs were MAF < 1%, call rate <98%, and p < 0.0001 for deviation from Hardy-Weinberg expectation. The final ISHDSF sample included 1005 individuals from 257 pedigrees, and the final case-control sample included 4319 individuals (2114 sporadic cases and 2205 controls), whose SNP data passed all QC filters.

**Imputation**

Genotypes passing QC were phased using Eagle V.2.4 [35] and phased genotypes were then imputed to the Haplotype Reference Consortium (HRC) reference panel [36] on the Michigan Imputation Server using Minimac4 [37]. The HRC reference panel includes 64,975 samples from 20 different studies that are predominantly of European ancestry, making it suitable for imputation of the samples studied here. Each of the genotype sets were imputed and the imputed genotype probabilities were extracted and used for PRS construction and downstream analyses. As part of the post-imputation QC, variants with MAF < 1% and imputation quality score of <0.3 [38] were excluded for the initial merging (Supplementary Materials and Supplementary Table 1–3). After imputation and all QC steps, 9,298,121 SNPs in the Affymetrix Array and 11,081,999 SNPs in the PsychArray remained for analysis. In total, 9,008,825 SNPs were shared across all three arrays and were used for PRS construction and all downstream analyses. The mean imputation quality for the SNPs used for PRS construction and downstream analyses on each array was high (mean for all ≥0.96). Detailed information on imputation quality for the SNPs used for PRS construction is provided in Supplementary Materials and Supplementary Table 1.

**Construction of polygenic risk scores**

The ISGC and ISHDSF cohorts are part of the PGC3-SCZ GWAS. To avoid upward bias in PRS estimations, we acquired leave-N-out SCZ summary statistics from the PGC by excluding all cohorts containing any Irish subjects included in the current study. The leave-N-out GWAS summary statistics for PGC3-SCZ (N = 156,509) were first QC’d by excluding variants with MAF < 1% and imputation quality score of <0.9, as well as removing strand ambiguous variants and insertion deletion polymorphisms. We then constructed PRS for all subjects using a Bayesian regression framework by placing a continuous shrinkage prior on SNP effect sizes using PRS-CS with phi value of 1e-2 [39]. PRS-CS uses linkage disequilibrium (LD) information from 1000 Genomes European Phase 3 European sample [40] to estimate the posterior effect sizes for each SNP. Although p-value thresholding method have been previously used frequently [41], PRS-CS has shown substantial improvement in predictive power compared to those methods [42]. Similar to LD Score regression [43], PRS-CS limits the SNPs for PRS construction to approximately 1.2 million variants from HapMap3. By restricting the variants to HapMap3, the partitioning provides ~500 SNPs restricting the variants to HapMap3, the partitioning provides ~500 SNPs

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Table 1. Diagnostic categories present in the ISHDSF sample.

| Category          | Description                                    | Narrow (N=469) | Intermediate (N=112) | Broad (N=52) | Very Broad (N=140) | Other (N=140) |
|-------------------|------------------------------------------------|----------------|----------------------|--------------|--------------------|---------------|
| Schizophrenia     | Narrow Schizophrenia spectrum                 | 409            | 30                   | 9            | 4                  | 48            |
|                   | Intermediate Schizophrenia spectrum           |                | 34                   | 34           | 80                 | 0             |
|                   | Disorders significantly aggregating in the relatives of probands |                | 10                   | 17           | 27                 | 2             |
|                   | Any other psychiatric disorders in the relatives of probands |                | 7                    | 22           | 27                 | 2             |
|                   | Unaffected relatives of probands              |                | 31                   | 21           | 21                 | 6             |
|                   |                                                                                           |                | 2                   | 102           | 27                 | 2             |

The concentric diagnostic hierarchy of the ISHDSF contains four case definitions: narrow, Intermediate, Broad and Very Broad spectrums. These case definitions in the ISHDSF reflect core and periphery of the psychosis spectrum based on previous genetic epidemiology work referenced in the methods section. A visual representation is provided in Supplementary Fig. 4.

1Poor-outcome and good-outcome schizoaffective cases are represented in narrow and intermediate diagnostic category respectively.

2There are four individuals with intellectual disability in the unaffected relatives category.

3Other diagnoses include Anorexia Nervosa (1) and Cyclothymia (1) in the very-broad diagnostic category.

**Note:** Numbers in the table represent the count of cases falling under each diagnostic category.
To show the specificity of the PRS constructed from PGC3-SCZ, an additional PRS for low density lipoprotein (LDL, N = 87,048) from the ENGAGE Consortium [44] was also constructed using the same protocols described above. Genetic correlation and Mendelian Randomization studies by PGC3-SCZ show that there is no genetic correlation or causal relationship between SCZ and LDL, making LDL an appropriate comparison phenotype in which no inflation of SCZ PRS would be expected [45, 46].

Genomic relationship matrix, principal component and statistical analyses
Statistical analyses were carried out using a mixed effects logistic regressions using GMMAT package [47] in R [48]. To account for the high degree of relatedness among individuals, we used glmm.wald() function, fitted by maximum likelihood using Nelder-Mead optimization. Family structure was modeled as a random effect with genetic relationship matrix (GRM) calculated using LDAT [49] in all family members as well as sporadic cases and population controls. Principal component analysis (PCA) of the full sample is consistent with all individuals in the sample having European ancestry (Supplementary Materials, and Supplementary Figs. 5–7). However, to account for fine-scale structure within the Irish population (Supplementary Fig. 8), the top 10 principal components (PC) were also included as covariates in the analyses. While none of the PCs showed association with genetyping arrays or sites, in order to account for other possible batch effects due to genotyping carried out on different arrays or at different sites, we included platform and site as covariates in the model (Supplementary Materials). The final regression models included GRM as a random effect covariate, with the top 10 PCs, genotyping platform, site, and sex as fixed effect covariates. The final results were adjusted for multiple testing using the Holm method in R.

RESULTS
The mean PRS across the diagnostic categories for SCZ are displayed in Fig. 1. No significant differences in LDL PRS were observed between any of the diagnostic categories compared to population controls (Supplementary Fig. 9), indicating the specificity of PGC3-SCZ PRS in this study.

PGC3-SCZ PRS results show that the Narrow spectrum category in the families, which includes familial cases of SCZ, had the highest mean PRS (Z = 1.13, SE = 0.09) followed by sporadic cases (Z = 1.06, SE = 0.09), intermediate spectrum familial cases (Z = 0.81, SE = 0.10), broad familial spectrum cases (Z = 0.67, SE = 0.11), very-broad spectrum cases (Z = 0.53, SE = 0.098), unaffected family members (Z = 0.36, SE = 0.10) and population controls (Z = 0.004, SE = 0.07).

No significant difference between familial and sporadic cases of SCZ
We observe no significant difference in PRS between familial SCZ cases and all sporadic SCZ cases, (p = 0.90), nor between familial SCZ cases and either FH+ (p = 0.88) or FH− (p = 0.82) sporadic SCZ cases. These results suggest that an increased burden of common SCZ risk variation is unlikely to account for the higher recurrence risk of SCZ in multiplex families (Fig. 1). Additionally, we show that there is no significant difference in SCZ PRS between FH+ and FH− sporadic SCZ cases (p = 0.92), suggesting that the inclusion of all sporadic cases in the comparison is unlikely to cause an upward bias in the mean PRS for the full cohort of sporadic cases, and further supporting the hypothesis that increased PRS is unlikely to account for FH of SCZ in the cohort studied here.

All family members carry a high burden of common SCZ risk variants
Familial and sporadic SCZ cases show a significantly higher mean SCZ PRS compared to all other diagnostic categories in the ISHDSF sample and ancestry-matched population controls (Figs. 1 and 2, Supplementary Table 3), underlining the important role of common risk variation in the genetic architecture of both familial and sporadic SCZ cases. All other ISHDSF diagnostic categories also show a significantly higher SCZ PRS compared to the population controls (Figs. 1 and 2). PRS comparison within the ISHDSF sample (Supplementary Table 4) shows no significant difference between mean PRS for intermediate and broad categories, indicating that individuals in both categories have a similar burden of common SCZ risk variants despite the presence of a range of diagnoses on the psychosis spectrum such as atypical psychosis and delusional disorder in the intermediate category, and disorders such as major depressive disorder with psychotic features, and bipolar disorder in the broad category. We observed no significant difference in SCZ PRS loading between the broad category and the very-broad category, which includes any other psychiatric disorder in the ISHDSF sample. The mean SCZ PRS in the very broad category is not significantly different from the unaffected members of the families, indicating a similar burden of common SCZ risk variation in these two distinct diagnostic categories. Finally, we observe a significantly higher PRS in unaffected family members compared to the population controls (p = 4.13 × 10−3), indicating a high baseline risk for SCZ in all members of multiplex families compared to population controls, regardless of their diagnostic status. This observation is consistent with SCZ transmission through some unaffected family members observed in the ISHDSF and other family samples.

DISCUSSION
Multiplex SCZ families represent the upper bounds of the distribution of recurrence risk for SCZ, and this study aimed to investigate the source of this increased recurrence risk. Since sporadic cases are considered to be the norm for most complex diseases including SCZ [50], this makes sporadic SCZ cases a good
obtaining odds ratios. The plots show odds ratios (OR, analyses follow the hypothesis that ISHDSF members and sporadic cases do not have a significant increase in recurrence risk in familial cases. We observed that comparison group to assess whether elevated PRS can account for the increase in recurrence risk in familial cases. We observed that familial SCZ cases do not have a significantly increased PRS compared to sporadic SCZ cases in our modestly sized sample. We further show that this observation holds true regardless of the FH status of sporadic cases. Therefore, our finding provides empirical evidence that increased recurrence risk of SCZ in the ISHDSF sample is unlikely to be attributable to an increased burden of common SCZ risk variation as identified from genome-wide association studies. Therefore, the hypothesis that high familial recurrence risk of SCZ in multiplex families may be attributable to excess rare variation in the genome specific to SCZ, warrants further investigation. Furthermore, these results validate the concept of a genetically influenced psychosis spectrum in multiplex SCZ families as shown by a continuous increase of common SCZ risk variation burden across all members of the ISHDSF, from unaffected family members, to narrow category in the ISHDSF sample.

This analysis reveals potentially important differences in the genetic architecture of familial SCZ cases compared to familial bipolar disorder (BIP) cases. Analysis conducted by Andlauer et al [27] on BIP multiplex families have shown that unlike familial SCZ cases studied here, familial BIP cases have a significantly higher BIP PRS compared to ancestry matched, sporadic cases. These results, in addition to sparse evidence for the involvement of rare risk variation in the genetic architecture of BIP [22], demonstrates the importance of common risk variation in familial BIP, whereas whole exome sequencing studies of SCZ in both family and case–control samples have demonstrated that in addition to common variation, rare variation also plays an important role in the genetic architecture of SCZ [21, 51–53].

Although sequencing studies are only now reaching sample sizes sufficiently powered to detect individually associated rare variation and rare variant enriched genes associated with SCZ [21], earlier sequencing and rare variation studies observe consistent enrichment of rare variation in certain gene-sets and functional categories related to SCZ [51]. In addition, SNP signals from PGC3-SCZ GWAS are shown to be highly enriched in noncoding functional sequences in the genome [25], further underscoring the importance of conducting large scale whole-genome sequencing to identify rare variation in non-coding regions of the genome linked to SCZ. Results from the 1000 Genomes Project demonstrates that rare functional variation is frequent in the genome [54] and shows strong population specificity [55]. For example, using GWAS probe intensity data in the Irish case–control sample used in this study, we have previously detected a rare, novel 149 kb duplication overlapping the protein activated kinase 7 (PAK7) gene only found in the Irish population [36]. This duplication is associated with SCZ in the ISGC (p = 0.0007), and a replication sample of Irish and UK case–controls with 22 carriers in 11,707 cases and 10 carriers in 21,204 controls (p = 0.0004, OR = 11.3). This duplication in PAK7 gene is in strong LD with local haplotypes (p = 2.5 x 10^{-21}), indicating a single ancestral event and inheritance identical by descent in carriers.

We note that the liability that is captured by PRS constructed from PGC3-SCZ is currently insufficient for predicting a diagnosis of SCZ (AUC = 0.71) [25], meaning that PRS alone cannot be used as a diagnostic tool. The results of our study further suggest that current PRS alone is unlikely to be predictive of SCZ recurrence risk in the families of index probands. To address both of these predictive limitations of SCZ PRS, additional components of genetic risk must be identified and included in order to improve both identification of future cases and recurrence risk prediction in the relatives of probands.

The results presented in this study should be interpreted in the context of some limitations. First, current PGC3-SCZ PRS accounts for ~2.6% of the total variance in SCZ liability [25], and genetic risks from rare and structural variation are not represented in the PRS. As a result, some known genetic risk factors for SCZ such as the 22q11 deletion [57] are not included in PRS construction, and such genetic risk factors are best measured through direct assessment of structural variation or whole genome sequencing studies. Despite these limitations, PRS provide the most reliable measurement of common risk variation in the genome and are suitable for indexing an individual’s risk for SCZ in this study. Second, the various diagnostic categories in the ISHDSF sample contain different number of subjects [30]. For example, the lower number of individuals satisfying broad and very broad diagnostic schema in the families, means that the power of analysis in those subgroups is lower. However, the narrow category which includes familial SCZ cases in the ISHDSF sample, has the highest number of individuals across all the diagnostic categories in the ISHDSF, making the sample suitable for the main hypothesis being tested in this study. Third, FH information is only available for a subset of sporadic cases as described in the methods. However, the ratio of FH+ (~31%) and FH− (~69%) sporadic cases studied here is in close agreement with FH data from large meta-analyses samples [8, 9], suggesting the subset of sporadic FH+ and FH− cases available are representative. Fourth, this analysis did not assess the common risk variant burden of each family separately, and the degree to which common risk variation may impact each family could vary between different families. Fifth, since the environmental factors unique to the families have also not been systematically assessed here, integrating rare genetic variation from whole sequencing studies with environmental influences in future analyses could further elucidate the role of rare variation and environmental influences on the recurrence risk of SCZ in multiplex families. Finally, as more samples from under-represented populations are collected, it is essential to replicate and show the generalizability of these findings in more diverse populations.

In conclusion, in this study, we show that differences in common risk variation as indexed by current PRS, is unlikely to account for the increased recurrence risk of SCZ in our cohort of
multiplex SCZ families and ancestry matched sporadic cases. Therefore, our results suggest that both common and rare SCZ risk variation needs to be indexed to potentially improve diagnostic and familial recurrence prediction of SCZ.

DATA AVAILABILITY
GWAS summary statistics for PGC3-SCZ GWAS is publicly available on the PGC website https://www.med.unc.edu/pgc/download-results/
Leave-N-out GWAS summary statistics for PGC3-SCZ GWAS was acquired from the PGC by following the appropriate guidelines and will be shared upon reasonable request.

GWAS summary statistics for LDL is publicly available on the ENGAGE Consortium website http://diagram-consortium.org/2015_ENGAGE_1KG/
We made use of various freely available software tools in this study: PRS-CS: https://github.com/getian107/PRSC
PLINK: https://www.cog-genomics.org/plink/2.0/
GMMAT: https://github.com/hanchenphd/GMMAT
LDAK: http://dougspeed.com/ldak/
PLINK2: https://www.cog-genomics.org/plink/2.0/
The custom scripts used in this study are available upon request.

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AUTHOR CONTRIBUTIONS
MA helped with the conceptualization, carried out the analysis and wrote the original manuscript. AEG, THN, RK, and BCV provided advice on the methodology. SAB, BTW, and MA helped with the conceptualization, carried out the analysis and wrote the original manuscript. ISGC provided access to the full case control sample. All authors contributed to the interpretation of the results and provided critical revisions of the manuscript.

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