Extramedullary Myeloid Leukemia in the Setting of a Myeloproliferative Neoplasm

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Abstract

Extramedullary acute myeloid leukemia (EML), also known as myeloid sarcoma (MS), is an extramedullary solid mass derived from the proliferation of myeloblasts outside of the bone marrow. EML can present independently or concurrently with intramedullary acute myeloid leukemia (iAML). It can happen de novo or secondary to iAML, myeloproliferative neoplasm (MPN), chronic myelomonocytic leukemia (CMML), or myelodysplastic syndrome (MDS). We present a 57-year-old female with a history of Janus kinase 2 (JAK-2)-positive essential thrombocythemia (ET) evolving into EML in the setting of a persistent TP53 mutation. We discuss the essential diagnostic studies including tissue biopsy and fluorodeoxyglucose positron emission tomography/computed tomography (F-FDG PET/CT) imaging. We also investigate the significance of cytogenetics and next-generation sequencing (NGS) along with the unique pathogenesis, treatment and prognostic implications.

Keywords: Myeloid sarcoma; JAK-2-positive essential thrombocythemia; TP53; Venetoclax; 5-azacitidine; Eprenetapopt; CD33 CAR T-cell therapy

Introduction

Extramedullary acute myeloid leukemia (EML) is a solid constituent formed by the migration and proliferation of myeloblasts outside of the intramedullary tissue [1]. Even though its incidence is not well reported [2], EML is estimated to occur in about 2-8% of the adult population with intramedullary acute myeloid leukemia (iAML) [3]. Myeloid sarcoma (MS) most commonly presents with concomitant iAML, but it can also present as a recurrence of it or de novo [2]. EML can also occur secondary to other hematological conditions such as myeloproliferative neoplasm (MPN), myelodysplastic syndrome (MDS), and chronic myelomonocytic leukemia (CMML) [1]. EML has a myriad of presentations, most commonly found in bone, lymphoid tissue, gastric mucosa, breast, skin, and in rare cases in the central nervous system (CNS) [4-6]. In the pediatric population, leukemia cutis is the most common presentation [7, 8]. MS resembles solid cancers in histology, immunohistochemistry (IHC), and RNA sequencing [9]. Its morphology can be described as a single-filing architectural pattern, identical to that of invasive lobular breast carcinoma and with dense desmoplastic keloid-like fibrosis, similar to colon, gallbladder, and pancreatic carcinomas [9].

Cytogenetic and molecular studies have identified similar mutation patterns as those found in AML [10, 11]. Prognostic cytogenetic and molecular abnormalities have not been determined yet in EML; however, reported cases show that the presence of MS confers a worse prognosis [12]. In recent clinical practice, the treatment modalities for EML incorporate surgery and radiation therapy that can be utilized in localized disease as well as in the palliative setting [13]. The definitive treatment approach almost always requires the addition of systemic therapy usually in the form of AML induction chemotherapy, as well as consolidation with allogeneic stem cell transplantation (allo-SCT) in the first line setting [13]. New targeted therapies and chimeric antigen receptor (CAR) T-cell therapy comprise a potential therapeutic breakthrough for the future. In this paper, we present a case of extramedullary bone and epidural involvement of AML arising from an underlying MPN.

Case Report

Investigations

We present a case of a 57-year-old female with history of cerebral aneurysm, transient ischemic attack and JAK-2-positive essential thrombocythemia (ET). The patient was initially diagnosed in 1987 and progressed to spent-phase myelofibrosis in 2011. The bone marrow pathology at the time of spent phase diagnosis revealed myelofibrosis with no increase in blast count and a decision was made to treat the patient with hydroxyurea. Six years later, she presented to the hospital...
with debilitating low back pain and pancytopenia.

**Diagnosis**

On presentation, a magnetic resonance imaging (MRI) with contrast of the lumbar spine showed multiple lesions throughout the lumbar spine and a mass centered at the S1 vertebrae extending into the epidural space as well as into the S1-S2 neural foramen (Fig. 1). The surgical sacral mass biopsy was indicative of a small blue cell neoplasm favoring MS. The tissue stained positive for CD34, CD117 (not shown), CD99 (not shown), CD33, CD11c, and equivocal for CD 68-KPI (Fig. 2). Negative stains included CD43 (weak), CD45, CD56, CD13, TdT, and myeloperoxidase (MPO). The bone marrow biopsy indicated presence of AML, arising within a chronic MPN (Fig. 3). Approximately a third of the marrow space was involved by a dense proliferation of primitive cells. The remainder of the marrow demonstrated marked megakaryocytic expansion with abnormal forms associated with scattered erythroid and myeloid precursors. The reticulin stain revealed marked reticulin fibrosis in much of the marrow. The bone marrow aspirate was insufficient for evaluation. Cytogenetic studies revealed a normal karyotype. The fluorescence in situ hybridization (FISH) study was normal and the next-generation sequencing (NGS) panel identified a TP53 Arg273His mutation with 17% allele frequency.

**Treatment**

The patient underwent a decompressive laminectomy for a right L5-S1 epidural tumor. Next, she received one fraction of radiation therapy to the S1 joint which resulted in the improvement of her lower back pain. She was subsequently induced with cytarabine and daunorubicin 7 + 3 protocol.

**Follow-up and outcomes**

A repeat bone marrow biopsy at day 14 after induction chemotherapy revealed a hypocellular marrow with less than 5% cellularity. The stain for CD34 was largely negative. There were mostly areas of dense fibrosis with few lymphoid cells coinciding with myelofibrosis. There was no evidence of AML on flow cytometric evaluation. The bone marrow aspirate was insufficient for evaluation. A bone marrow biopsy on day 28 was positive for an MPN (post-ET myelofibrosis). There was no morphological or immunophenotypic evidence of AML. The marrow had a cellularity of 95-100% with 1% blasts present. The repeat NGS panel identified a persistent TP53 Arg273His mutation with unchanged allele frequency at 17%.

Two months after, a repeat MRI with contrast of the lumbar spine redemonstrated marrow signal abnormality identified throughout the lumbosacral spine concerning for interval progression of prior EML. The epidural mass described previously, at the level of S1, appeared less prominent (Fig. 4). A positron emission tomography/computed tomography (PET/CT) identified focal areas of fluorodeoxyglucose (FDG) activity within the posterior left iliac bone as well as in the sacral region (Fig. 5). A bone biopsy of the left iliac bone showed bone remodeling fibrotic changes. A repeat biopsy of the prior soft tissue area near the right sacroiliac (SI) joint showed fatty tissue with necrosis. Based on the biopsy results, we concluded that the patient had regenerative bone changes as opposed to relapsed leukemia. A repeat bone marrow biopsy showed stro-
mal changes including loose fibrosis and maturing trilineage hematopoiesis. The marrow had 30-45% cellularity and 1% blasts. The flow cytometry was negative. The bone marrow biopsy was compatible with an underlying MPN, and showed no evidence of AML. The TP53 Arg273His mutation persisted in the bone marrow with 25% allele frequency, which had increased since the prior biopsy. The patient received four cycles of 5-azacytidine and venetoclax with goals to bridge to allo-SCT and suppress the TP53 mutation. She received full intensity fludarabine and busulfan and then proceeded with allo-SCT from a 10/10 matched unrelated donor. A repeat bone marrow biopsy 60 days after transplant did not show any evidence of iAML. The cytogenetics and FISH analysis were normal, but the TP53 mutation testing was not done at the time. Shortly after, the patient was admitted with altered mental status and was found to have leukemic meningitis. At this point,

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**Figure 2.** Staining from the sacral mass biopsy: (a) CD68-positive cells; (b) CD11c-positive cells; (c) CD33-positive cells; (d) CD34-positive cells.

**Figure 3.** (a, b) Bone marrow pathology sample showing increasing mononuclear cells.
the patient decided to pursue hospice care. The time from her diagnosis of EML to her death was 9 months.

Discussion

The diagnosis of MS involves a multi-modality approach. Imaging is one of the first diagnostic tools that can detect a suspicious mass early and it can also be used to evaluate treatment response [13, 14]. PET/CT scans should be considered in diagnosing new-onset EML, as well as relapsed EML, because it can help detect small lesions that are difficult to visualize with other imaging modalities [15]. PET/CT scans aid in facilitating diagnosis as well as identifying obscure foci of relapsed disease [16]. A retrospective analysis showed that an F-FDG-PET/CT is more accurate than an F-FDG-PET or CT alone [17]; however, in both cases, bone marrow and tissue biopsies are essential for diagnosis [13]. The differential of EML includes Ewing’s sarcoma, peripheral neuroendocrine tumors, neuroblastomas, lymphoblastic lymphomas, Wilms’ tumors, sarcomas, and Burkitt’s lymphoma. Often MS resembles small, round, blue-cell tumors that are embryonic in appearance [18, 19]. Morphologically, EML is characterized by infiltrative myeloblasts derived from adnexal destruction [20]. In regards

Figure 4. T1-weighted MRI with contrast sagittal (a) and axial (b) views showing marrow signal abnormality identified throughout the lumbosacral spine. The S1 mass described previously appears less prominent. MRI: magnetic resonance imaging.

Figure 5. (a) PET-CT identifying focal areas of SUV activity. There are intense areas of activity identified within the sacrum, in the left sacral alar with an average SUV of 1.9 and a maximum SUV of 4.3 as well as in the mid sacrum S1 level with an average SUV of 2.1 and a maximum SUV of 4.2. (b) There is an average SUV of 1.6 with maximum of 3.0 in the posterior left iliac bone adjacent to the sacroiliac joint as well. Discrete lesions in these areas are not identified. PET: positron emission tomography; CT: computed tomography; SUV: standard uptake value.
to IHC, CD68-KP1 is the most expressed marker followed by MPO [13, 20, 21]. Other markers include CD34, CD43, CD45, CD68, CD56, CD117, CD11c, CD13, and CD33 [13, 19, 20]. Cytogenetic aberrations correlate closely with those found in AML, MPN, or MDS. The most common cytogenetic abnormalities are t(8;21)(q22;q22) and inv(16)(p13;q22) which are also associated with AML subclassifications with the most favorable prognosis [20, 22-24]. Even though cytogenetics can characterize MS, their prognostic value is uncertain in EML. In our patient's case, the NGS panel only revealed a TP53 mutation. In other reports, FLT3 and NPMI mutations have shown to be the most common mutations found in EML case reports [20, 22-24]. Gene encoding tyrosine kinases (FLT3, KIT, and KRAS), tumor suppressors genes (WTI and TP53), spliceosome proteins (SF3B1 and SRSF2), transcription factors (RUNX1), as well as epigenetic modifiers (TET2 and ASXL1) are commonly found in EML [11, 24].

The incidence of the TP53 mutation is only 12.7% in AML; however, it reaches an incidence up to 40-50% in solid malignancies [25-27]. In AML, the TP53 mutation is most commonly associated with the use of prior chemotherapy or with the presence of a complex karyotype. TP53 mutations have also been found in secondary AML associated with polycythemia vera (PV) and ET [25, 28]. We identified five cases of MS associated with TP53 mutations published in literature [29]. One of these cases reported a medistinal MS preceding AML with a PICALM-MLLT10 fusion gene mutation in addition to the TP53 mutation [29]. Two other cases reported de novo MS and two additional cases had underlying MDS before developing MS [1, 11]. Preclinical studies have shown that inactivation of TP53 leads to uncontrolled cell proliferation and increased oncogenesis in hematopoietic stem cells as well as enhanced engraftment in various tissues [27]. In mice, TP53-deficient cells were shown to have an engraftment advantage over the TP53 wild-type stem cells which could potentially aid in the engraftment of myeloblasts to the extramedullary tissues [30-34]. This could relate to our patient's case, where a TP53 mutation may have helped the development of MS. TP53 is often seen as a later finding in hematological malignancies, as it is known to confer poor prognosis.

The pathogenesis of EML has yet to be fully elucidated. For a long time, CD56, the blast neural cell adhesion molecule, was a protein of high interest in the mechanism of homing of myeloid blasts to various extramedullary sites that expressed high CD56 [10, 20, 35]. High CD56 expression is most commonly associated with t(8;21) and 11q23 aberrations [35, 36]; however, most patients with MS do not express CD 56 in their leukemic cells [12, 37, 38]. Another important cell adhesion protein is CD11b (surface β2-integrin member macrophage-1 antigen) which is derived from mononuclear cells and is found to be highly expressed in AML with monoblastic/myelomonocytic differentiation, where the risk of developing MS is higher [38, 39]. Studies have shown the expression of CD11b in AML associated with MS to be higher than in AML alone [37]. The RAS-MAPK/ERK pathway is another potential pathway in the development of EML as metastasis-suppressor RAF kinase inhibitor protein (RKIP) loss has been reported in 50% of patients with AML associated with MS, versus only 14% in patients with AML alone [40].

Several factors need to be taken into consideration while choosing therapy for the management of MS, such as the timing of initial diagnosis versus relapse, the presence of concurrent AML, and the presence of local symptoms [20]. Local therapies such as surgical decompression or radiation therapy have been used in isolated MS cases, as well as in cases of severe symptoms with palliative intent [1]. In our patient with bone involvement, surgical decompression followed by radiation therapy played a major role in her treatment approach, as well as in the relief of her symptoms. The risk for impending cord compression remains a real risk for patients with epidural/bone involvement. Even though our patient did not have cord compression, her symptoms required surgical intervention. Systemic therapy, however, should almost always be considered in both local and metastatic disease. Clinical data have shown that 75-90% of patients with isolated MS who do not receive systemic therapy progress to AML within 4 months [12, 41].

AML induction chemotherapy has found usefulness in the treatment of EML since this is preceded or followed by iAML within a short period of time in most cases [42]. The current suggested approach is utilizing anthracycline-based induction regimens, and allogeneic transplantation in the initial setting [13, 43]. The presence of EML itself confers a poor prognosis in patients with iAML [12]. A series of case reports have demonstrated that addition of systemic therapy to surgery and radiation can achieve a 65% remission rate and an overall survival (OS) of 40 months in patients with EML [13, 44]. Retrospective data suggest that allo-SCT should be offered in the first line setting, after induction chemotherapy in patients with isolated EML as well as in patients with concurrent iAML [13, 43, 45]. Patients who benefit the most from allogeneic transplant are those who achieve complete response (CR) prior to it and second transplants are sometimes indicated as salvage therapy in patients with relapsed disease [12, 46-49].

While induction chemotherapy with cytarabine and anthracycline combination is the preferred initial treatment in fit AML patients, this is not the case in TP53-mutated patients due to the discouraging data in this group [25]. More recently, the combination of venetoclax and the hypomethylating agent (HMA) 5-azacytidine has gained great interest in the treatment of TP53-mutated patients. In a recent randomized prospective trial of older AML patients, the doublet was associated with a higher incidence of composite remission in the TP53-mutated group, but this was not translated into OS benefit [50]. The mechanism of apoptosis mediated by venetoclax appears to be TP53-independent, while HMA agents have shown up to 47% response rates in TP53-mutated patients [51, 52]. Currently, there are several clinical trials investigating novel therapies in this group of patients. The mutant p53 reactivator, eprenetapopt (APR-246), has shown some promise. Phase I/II data show the combination of APR-246 with 5-azacitidine in patients with TP53-mutant MDS and AML has achieved a CR of 44% [53]. TP53 mutation predicted higher CR when this regimen was given as maintenance therapy after allo-SCT [52]. An anti-CD47 IgG4 monoclonal antibody that stimulates cellular phagocytosis and T cell-mediated cytotoxic effect on leukemic cells called magrolimab is also being investigated in combination with azacytidine in the untreated AML as well as relapsed setting. In the TP53-mutant cohort, this combination has shown a CR of 45%.
with a median duration of response of 7.6 months [52]. Other targeted therapies that are being investigated are eprenetapopt in combination with venetoclax and azacytidine and bispecific dual affinity retargeting antibody (DART) against CD3 and CD123, flotetuzumab in the early relapse setting [52].

Non-randomized data that support venetoclax combination with HMA in AML patients also exist in the salvage setting. This is supported by a small retrospective analysis of AML patients that reported a CR of 58% with venetoclax and 5-azacytidine in the salvage/pre-transplant setting [54]. We thought that this combination of therapy would have benefitted our patient as a bridge to allo-SCT, by hopefully inducing CR. In the relapse setting, other potential targeted therapies that have shown activity in relapse EML based on reported patient cases are anti-CD33 monoclonal antibody gemtuzumab ozogamicin in patients with concurrent positive CD33 AML [55] and the tyrosine kinase inhibitor imatinib for patients with FIP1L1-PDGFRα and BCR-ABL mutations [13, 56]. The newest consideration for relapse EML is the PRGN-3006 ultra-CAR-expressing T-cell therapy, which is currently under investigation in a phase 1/1b trial. This is a multigenic, autologous CD33 CAR T-cell therapy trial that is currently open for enrollment for relapsed/refractory AML, including extramedullary disease [57].

**Learning points**

MSs are infrequent and vexing for the clinician when it comes to making a diagnosis and approaching treatment outcomes.

We hypothesize that TP53 mutation may have been the initiating event for developing EML and its persistency may have been a major factor of contribution in this patient’s poor prognosis.

Localized treatment with surgery and radiation in conjunction to AML-based systemic therapy and stem cell transplantation remain the current approach to treating EML, although the outcomes are poor overall.

TP53-mutated AML patients have poor outcomes with standard therapy, therefore targeted therapies and clinical trials should be considered. We can predict similar poor outcomes in EML with TP53 mutations.

Considerations for targeted molecular therapies in relapse MS are anti-CD33 monoclonal antibody gemtuzumab ozogamicin, as well as the tyrosine kinase imatinib.

CD33 CART therapy is currently under investigation for relapse disease, including EML, in the clinical trial setting.

Further large randomized controlled studies are necessary to establish these and other novel agents as the standard of care for the treatment of EML.

In general, extramedullary presentations are unique and not fully understood. Finding the best treatment approach poses a complicated challenge.

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**Conflict of Interest**

We do not have any conflict of interest.

**Informed Consent**

Verbal consent from the patient’s family was obtained.

**Author Contributions**

Dr. Jorgena Kosti gathered the patient’s information and wrote the manuscript. Dr. Timothy Mervak provided the pathology and the pathology slides as well as pictures. Dr. Howard Terебelo edited and supervised the writing of this paper.

**Data Availability**

The authors declare that data supporting the findings of this study are available within the article.

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