HOTAIR Promotes Proliferation, Migration, and Invasion of Ovarian Cancer SKOV3 Cells Through Regulating PIK3R3

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Background: The aim of this study was to determine the effect on proliferation, migration, and invasion after silencing HOTAIR in ovarian cancer SKOV3 cells, and to elucidate the potential mechanism.

Material/Methods: We analyzed the mRNA expression level of HOTAIR and PIK3R3 in ovarian cancer SKOV3, OVCAR3, and A2780 cell lines. We analyzed the mRNA expression level of HOTAIR and PIK3R3 in ovarian cancer SKOV3 after transection with miR-214 or miR-217. We analyzed the mRNA and protein expression level of PIK3R3 when silencing HOTAIR. We analyzed the expression of HOTAIR when silencing PIK3R3. We analyzed the proliferation, migration and invasion in ovarian cancer SKOV3 after silencing HOTAIR or PIK3R3.

Results: The expression of HOTAIR and PIK3R3 in ovarian SKOV3 and OVCAR3 was increased compared with A2780 cells (P<0.05). The mRNA level of HOTAIR and PIK3R3 in ovarian cancer SKOV3 cells was decreased when transected with miR-214 or miR-217 compared to negative control (p<0.05). The mRNA and protein level of PIK3R3 was decreased when HOTAIR was silenced and the mRNA level of HOTAIR was decreased when PIK3R3 was silenced (p<0.05). The proliferation, migration and invasion was decreased in ovarian SKOV3 when HOTAIR or PIK3R3 was silenced (p<0.05).

Conclusions: HOTAIR can promote proliferation, migration, and invasion in ovarian SKOV3 cells as a competing endogenous RNA.

MeSH Keywords: Cell Migration Assays, Leukocyte • MicroRNAs • Ovarian Neoplasms

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Background

Epithelial ovarian cancer (EOC) is among the leading causes of cancer death in women. Because most ovarian cancers become drug-resistant, the survival rates are still low [1]. Thus, it is important to understand the potential molecular mechanisms involved in EOC metastasis. It is crucial to find biomarkers of EOC for diagnosis and predict the clinical outcome of the patient. 

Long non-coding RNAs (LncRNA Long intergenic non-coding RNA) have been reported to regulate gene expression at various levels, including in chromatin modification, transcription, and posttranscriptional processing [2,3]. LncRNA-p21 and GASS act as tumor-suppressor molecules [4]. Expression of lincRNA-p21 is regulated by the p53 and upon transcription, it suppresses expression of the genes transcriptionally regulated through p53 by binding to the hnRNP-K complex [5]. GASS plays an important role in the induction of apoptosis. It suppresses some antiapoptotic genes through binding to the glucocorticoid receptor (GR) and hence prevents GR from binding to the glucocorticoid response elements on the target DNA molecule [6].

Long noncoding RNA HOTAIR (HOX antisense intergenic RNA) has been considered as a pro-oncogene in multiple cancers [7]. HOTAIR expression was obviously increased in leukemic cell lines and primary AML blasts. It modulated c-KIT expression by competitively binding miR-193a [8]. The up-regulation of HOTAIR has been found in both estrogen receptor-positive and triple-negative breast cancer. It has been reported that HOTAIR is regulated by estrogen via ERs in ER-positive breast cancer [9]. HOTAIR expression was increased in oral squamous cell carcinoma compared to non-tumor tissue and is associated with metastasis, stage, and histological differentiation [10]. However, the underlying molecular mechanism of HOTAIR in ovarian cancer remains under unclear.

The PI3K (phosphoinositide-3-kinase, 3)/AKT/mammalian target of rapamycin (mTOR) pathway plays a critical role in the malignant transformation of human tumors and their subsequent growth, proliferation, and metastasis. Preclinical investigations have suggested that the PI3K/AKT/mTOR pathway is frequently activated in ovarian cancer [11,12]. PIK3R3 is regarded as a potential therapeutic target of ovarian cancer [13]. However, there is no report about the relation between PIK3R3 and HOTAIR.

In this study, we investigated the function of HOTAIR in ovarian cancer SKOV3 cells, then elucidated the mechanism.

Material and Methods

Immunohistochemistry (IHC)

The patient and control groups had no significantly difference in terms of age and parity (p>0.05). The tissue samples were fixed in formalin, embedded in paraffin, sectioned (5 μm thick), deparaffinized in xylene, dehydrated in a graded series of ethanol, subjected to antigen retrieval in citrate buffer for 30 min in a steamer, and washed in phosphate-buffered saline (PBS). PBS was used for all subsequent washes and for antiserum dilution. Tissue sections were quenched sequentially in 3% hydrogen peroxide and blocked with PBS containing 10% goat serum (Sigma) for 1 h at 37°C. Next, slides were incubated with the primary antibody overnight at 4°C, which included Rabbit monoclonal (1:100; ab186612; Abcam). Negative controls included omission of primary antibody and use of irrelevant primary antibodies. After several washes (3×3 min) to remove excess antibody, the slides were incubated with diluted (1:300) anti-rabbit biotinylated antibodies (Beijing Biosynthesis Biotechnology CO., LTD, China) for 1 h. All the slides were washed in PBS and were incubated in avidin biotin peroxidase complex (ABC, Beijing Zhongshan Golden Bridge Biotechnology Co. Ltd., Beijing, China) diluted 1:300 in PBS for 30 min in humidified chambers at 37°C. DAB (Beijing Biosynthesis Biotechnology CO., Ltd., China) was used as a chromogen and hematoxylin was used as a nuclear counter-stain.

Slides were evaluated independently by 3 pathologists for distribution and intensity of signal as described by De Falco et al. [14] Intensity was scored from 0 to 3: 0 (absent immunopositivity); 1 (low immunopositivity); 2 (moderate immunopositivity); and 3 (intense immunopositivity). An average of 22 fields was observed for each tissue.

Cell lines and culture conditions

Ovarian cancer cell lines (SKOV3, OVCAR3, and A2780) were purchased from the Institute of Biochemistry and Cell Biology of the Chinese Academy of Sciences (Shanghai, China). Cells were cultured in RPMI 1640 medium supplemented with 10% fetal bovine serum (10% FBS), 100 U/ml penicillin, and 100 mg/ml streptomycin (Invitrogen) in humidified air at 37°C with 5% CO2. The miRNA mimics were purchased from GenePharma Co., Ltd. (Shanghai, China). The negative control was non-specifically transfected siRNA.

Cell proliferation assay

Cell proliferation was determined using the CCK-8 assay, as described previously [15], and EdU assay was performed using the Cell-Light TM EdU imaging detecting kit according to the instructions in the kit (Ruibo Biotechnology, Guangzhou, China).
EdU is a thymidine analog that can be used to label cells undergoing DNA replication [16].

Quantitative real-time polymerase chain reaction

Total RNA was isolated using an RNA pure High-purity Total RNA Rapid Extraction Kit (BioTeke, RP1201, China), as per the instructions provided in the kit. cDNA was synthesized using the iSCRIPT cDNA synthesis kit (Bio-Rad). The primers used for amplifying HOTAIR, PIK3R3, and GAPDH were synthesized by Guangzhou Funeng Co., Ltd. The real-time PCR kit was purchased from Guangzhou Funeng Co., Ltd. PCR conditions were 95°C for 10 s, 60°C for 20 s, and 72°C for 10 s. Each sample was analyzed in triplicate. Relative quantification of mRNA was performed using the comparative threshold cycles (CT) method. This value was used to plot the gene expression using the formula $2^{-\Delta\Delta CT}$.

Western blot analysis

Expression of MAPK1 protein was analyzed by Western blotting, as previously described [17]. The primary antibodies used were polyclonal goat anti-PIK3R3 (1:1000; ab186612; Abcam) and monoclonal rabbit anti-GAPDH (1:5000; ab181602; Abcam). The bands were detected via enhanced chemiluminescence (ECL) reagent (Beyotime Institute of Biotechnology, Jiangsu, China) and quantified by ImageQuant 3.3 software (Molecular Dynamics).
**Statistical evaluation**

All values are expressed as mean ±SEM. The Mann-Whitney U test was used. A p value of <0.05 was considered statistically significant.

**Results**

The expression of PIK3R3 in ovarian cancer tissues

The expression of PIK3R3 was significantly higher in ovarian carcinoma samples than in normal ovarian samples (Figure 1; P<0.05). PIK3R3 was predominantly localized on the plasma membrane and cytoplasm (Figure 1A, 1B).

The expression of HOTAIR and PIK3R3 in ovarian cancer cells

To investigate the expression of HOTAIR and PIK3R3 in ovarian cancer cells, we detected the mRNA expression of HOTAIR and PIK3R3 in SKOV3, OVCAR3, and A2780 cells. The mRNA expression of HOTAIR and PIK3R3 in ovarian cancer SKOV3 and OVCAR3 was increased compared with A2780 cells (Figure 2A) (P<0.05).

The interaction between HOTAIR and PIK3R3 in ovarian cancer SKOV3 cells

We selected SKOV3 cells to determine the high-level expression of HOTAIR and PIK3R3. The protein and mRNA level of PIK3R3 were decreased when silencing HOTAIR. The mRNA level of HOTAIR was decreased when silencing PIK3R3 (P<0.05).
When silencing dicer, the interaction between HOTAIR and PIK3R3 was abolished (Figure 2B–2D).

**HOTAIR and PIK3R3 are targets of miR-214 or miR-217**

We investigated whether HOTAIR and PIK3R3 are targets of miR-214 or miR-217. MiR-214 or miR-217 mimics and inhibitors were transfected into ovarian cancer SKOV3 cells. The expression of HOTAIR and PIK3R3 was decreased when transfected with miR-214 or miR-217 mimics (Figure 3A; P<0.05). In contrast, the expression of HOTAIR and PIK3R3 was increased when transfected with miR-214 or miR-217 inhibitors (Figure 3B) (P<0.05).

**Silencing HOTAIR or PIK3R3 increased the expression of miR-214 or miR-217**

We investigated whether HOTAIR and PIK3R3 regulates miR-214 or miR-217. The expression of miR-214 or miR-217 was increased when HOTAIR or PIK3R3 was silenced (Figure 3C) (P<0.05). The present work provides the first evidence for a positive HOTAIR/PIK3R3 correlation and the crosstalk between miR-214 or miR-217, HOTAIR, and PIK3R3. Luciferase assays confirmed the existence of specific crosstalk between the HOTAIR and PIK3R3 mRNA through competition for miR-214 or miR-217 binding (Figure 3D, 3E).

**Silencing HOTAIR or PIK3R3 inhibited the proliferation, migration and invasion in SKOV3 cell**

We studied the effects of HOTAIR or PIK3R3 on SKOV3 cells. The proliferation, migration, and invasion of SKOV3 was inhibited when HOTAIR or PIK3R3 was silenced (Figure 4 and 5)(P<0.05).

**Discussion**

Long noncoding RNAs are a recently reported class of non-protein coding RNAs, which have now increasingly been shown to be involved in a wide variety of biological processes as regulatory molecules. Recent reports have suggested that IncRNAs could potentially interact with miRNAs and regulated their regulatory role by interactions [18].

In this study, we found that the expression of HOTAIR and PIK3R3 in ovarian SKOV3 and OVCAR3 was increased compared to that in A2780 cells. The repression of HOTAIR and PIK3R3 were strongly correlated. Furthermore, we found that HOTAIR interacted with PIK3R3 through miR-214 or miR-217. The proliferation, migration, and invasion were inhibited in ovarian SKOV3 after silencing HOTAIR or PIK3R3.

Emerging evidence suggests that IncRNAs may participate in this regulatory circuitry. We hypothesized that HOTAIR may also serve as a ceRNA. Bioinformation analysis (http://starbase.sysu.edu.cn/) indicated that HOTAIR is a putative ceRNA of PIK3R3. Firstly, we investigated the expression of PIK3R3 and HOTAIR in ovarian cancer SKOV3, OVCAR3, and A2780. We found that the expression of HOTAIR and PIK3R3 in ovarian SKOV3 and OVCAR3 was increased compared with A2780 cells (P<0.05) suggesting that HOTAIR may be related to PIK3R3. Silencing HOTAIR down-regulated the protein and mRNA level of PIK3R3. Silencing PIK3R3 downregulated the expression of HOTAIR, indicating that HOTAIR may interact with PIK3R3. When silencing dicer, the interaction between HOTAIR and PIK3R3 was abolished. Therefore, we hypothesized that the miRNA may be involved in this process.
Bioinformation analysis predicts that miR-214 and miR-217 are shared by HOTAIR and PIK3R3 [19,20]. Therefore, we determined whether the interaction of HOTAIR and PIK3R3 is regulated by miR-214 and miR-217. As expected, miR-214 and miR-217 mimics can inhibit the expression of HOTAIR and PIK3R3. When silencing HOTAIR, the expression of miR-214 and miR-217 was increased, suggesting that the interaction of HOTAIR and PIK3R3 is mediated through miR-214 and miR-217.

Phosphatidylinositol 3-kinase (PI3K) is involved in a wide range of cancer-associated signaling pathways [21]. The role of PI3K in ovarian cancer will undoubtedly provide new insights for pharmacologic intervention in this malignancy. Molecular cloning of PI3Ks reveals a large and complex family that contains 3 classes of multiple subunits and isoforms [22]. PIK3R3 had increased expression in ovarian cancer and knockdown of PIK3R3 mRNA expression increases cell apoptosis in ovarian cancer in vitro [13]. Our study revealed that silencing PIK3R3 can inhibit the proliferation, migration, and invasion in ovarian cancer SKOV3 cells. This is consistent with a previous report [13] that HOTAIR is overexpressed in males in colorectal cancer, hepatocellular carcinoma, pancreatic cancer, and gastrointestinal stromal tumors, and in females in ovarian cancer [23–37]. Down-regulation of HOTAIR can inhibit the malignant behavior of cancer [28]. In this study, silencing HOTAIR inhibited the proliferation, migration, and invasion of ovarian cancer SKOV3 cells.

Conclusions

Our study determined that HOTAIR interacting with PIK3R3 regulates ovarian cancer skov3 cells proliferation, migration, and invasion through miR-214 and miR-217. It can serve as a therapy target of ovarian cancer. This study gives new insight into ovarian cancer research, pathogenesis, and treatment. In the future, we will study the interaction between HOTAIR and PIK3R3 in vivo.
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