Research Article

Correlation of Serum C-Peptide, Soluble Intercellular Adhesion Molecule-1, and NLRP3 Inflammasome-Related Inflammatory Factor Interleukin-1β after Brain Magnetic Resonance Imaging Examination with Cerebral Small Vessel Disease

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Objective. To explore the correlation of serum c-peptide, soluble intercellular adhesion molecule-1 (sICAM-1), and NLRP3 inflammasome-related inflammatory factor interleukin-1β (IL-1β) after brain magnetic resonance imaging (MRI) examination with cerebral small vessel disease (CSVD). Methods. A total of 72 CSVD patients treated in our hospital from December 2018 to December 2019 were selected as the case group and another 72 patients who presented cerebrovascular risk factors but obtained normal brain MRI examination result in the same period were selected as the control group. The serum specimen of patients in the two groups were collected, their serum c-peptide levels were measured by radio immunoassay, and their serum sICAM-1 and NLRP3 inflammasome-related inflammatory factor IL-1β were measured by enzyme-linked immunosorbent assay (ELISA), so as to analyze the correlation between these indicators and CSVD. Results. Compared with the control group, the level values of serum c-peptide, sICAM-1, and IL-1β were significantly higher in the case group (P < 0.001), with CSVD being the dependent variable, and age, smoking, uric acid, history of stroke, serum c-peptide, sICAM-1, and IL-1β being the independent variables. A logistic regression analysis was conducted, and the result showed that age, smoking, serum c-peptide, sICAM-1, and IL-1β were the risk factors for CSVD, and by drawing the ROC curves, it could be concluded that the area under sICAM-1 curve was larger than that of other single indicator. Conclusion. Elevation of level values of serum c-peptide, sICAM-1, and NLRP3 inflammasome-related inflammatory factor IL-1β is correlative with CSVD, and age, smoking, serum c-peptide, sICAM-1, and IL-1β are the independent risk factors for CSVD.

1. Introduction

Cerebral small vessel disease (CSVD) refers to the disease caused by small vessel lesion in the brain, which involves a vessel diameter of 30–800 μm [1, 2]. It is a brain wide dynamic disease, the exact pathogenesis of which has not been fully elucidated, and it is currently believed to be mainly related to hypoperfusion, microembolic embolism, ischemic damage, structural changes in small vessels, and some metabolic encephalopathies, such as homocysteinemia. With the acceleration of population aging and high frequency of risk factors for cerebrovascular disease, the prevalence of CSVD is increasing year by year. Recent advances in science and technology have deepened the understanding of CSVD [3], and studies have shown that CSVD increases patients’ risk of developing cerebral infarction and death. Meanwhile, further studies have confirmed that the impairment of CSVD on cognitive function increases patients’ probability of developing dementia, undoubtedly illustrating the immense danger of the disease [4, 5]. CSVD has a complex cause of disease with
variable and more insidious clinical symptoms, and its diagnosis mainly relies on imaging examinations, so early detection and implementation of targeted intervention is the key to improving the prognosis of the disease [6]. At present, age, hypertension, etc., are recognized risk factors for CSVD, but there is a lack of specific serological markers. Previous studies have confirmed that serum c-peptide can indirectly reflect the concentration of insulin, and insulin resistance is related to the abnormal increase of insulin content, which is an independent risk factor for the occurrence of CSVD [7].

NLRP3 inflammasome is an important pattern recognition receptor in the cytoplasm, which can sense pathogenic microorganisms and metabolites in cells and start the assembly of inflammatory complex. As an inflammatory factor of NLRP3, IL-1β is one of the key messengers involved in the interaction between the CNS and the immune system, and it also involves in immune regulation in the CNS and promotes neuronal growth and development under physiological conditions, but its high concentrations will inhibit the excitability and synaptic function of neurons in the hippocampal region, which in turn will lead to cognitive dysfunction [8]. Soluble intercellular adhesion molecule-1 (sICAM-1), as a biomarker of endothelial cell injury, participates in inflammatory response. Some studies [9] have found that a higher level of sICAM-1 may be a risk factor for leukoaraiosis. Leukoaraiosis refers to the hypersoftening of cerebral white matter, which is a normal aging process, but will lead to leukoaraiosis when softening is too fast. Currently, there are no reports pointing out the correlation of serum c-peptide, NLRP3 inflammasome-related inflammatory factor IL-1β, and sICAM-1 after brain MRI examination with CSVD. To prove the aforesaid opinions, the study was carried out, and the results are reported as follows.

2. Materials and Methods

2.1. General Data. A total of 72 CSVD patients treated in our hospital from December 2018 to December 2019 were selected as the case group, and 72 patients who presented cerebrovascular risk factors but obtained normal brain MRI examination result in the same period were selected as the control group. The study met the World Medical Association Declaration of Helsinki (2013) [10]. Within 24 after admission, neurologists collected the clinical data and information of the study subjects who met the enrollment conditions, including their gender, age, smoking history, history of hypertension, and total cholesterol (TC) level.

2.2. Inclusion and Exclusion Criteria. Inclusion criteria: (1) the patients met the Diagnostic Criteria for CSVD Revised by National Conference on Cerebrovascular Diseases [11] and were aged between 18 and 80 years; (2) the patients had focal neurological signs; (3) the patients had an acute or subacute onset; and (4) the patients did not have history of serious brain injury.

Exclusion criteria: (1) the patients had stroke with other specific causes; (2) the patients were unconscious or were unable to cooperate with the study; (3) the patients were complicated with severe dysfunction of kidney, liver, lung, etc.; and (4) the patients presented obvious intracranial artery atherosclerosis.

2.3. Methods. Patients in the two groups received the brain MRI examination for measurement of their expression levels of serum c-peptide, NLRP3 inflammasome-related inflammatory factor IL-1β, and sICAM-1, with the following specific methods.

Brain MRI examination: Philip 1.5T MRI system (manufactured: Koninklijke Philips N.V.) was used for brain scan in the conventional axial position and sagittal positions T1WI (TR 550 ms, TE 15 ms, slice thickness 4 mm, slice gap 0.4 mm, and 20 slices), T2WI (TR 2.7 ms, TE 150 ms, slice thickness 4 mm, slice gap 0.4 mm, and 20 slices), and DWI (TR 2.7 ms, TE 50 ms, b value 1,000 s/mm², slice thickness 4 mm, slice gap 0.4 mm, and 20 slices); after image collection, two senior imaging physicians analyzed the images by the double blind method to observe and record situations such as leukoaraiosis and cerebral microhemorrhage.

Serum detection: the patients in the two groups were checked and told the objective and cooperation method of blood collection. 5 ml of fasting venous blood was drawn from their suitable vein in the morning and placed for 10–20 min under room temperature and then centrifuged under 3,000 r/min for 15 min to collect the supernatant and put it in an ultralow temperature freezer. In case of precipitation occurred during preservation, the blood samples should be centrifuged again for supernatant collection. The serum c-peptide level was measured by the radioimmunoassay with the kits purchased from Nanjing Xinfan Biotechnology Co., Ltd. The IL-1β and sICAM-1 levels were measured by the ELISA method with the experiment steps in strict accordance with the specification on the kits (manufactured: Wuhan Adanti Biotechnology Co., Ltd.). The kits were required to be preserved under the environment of 2–8°C before use; if use, the kits should be taken out from the cold storage environment and then placed under the room temperature for 15–30 min before operation.

2.4. Statistical Methods. In this study, the data were processed with the professional statistical software SPSS23.0, and the picture drawing software was GraphPad Prism 7 (GraphPad Software, San Diego, USA). The related risk factors for the occurrence of CSVD were analyzed by multifactor logistic regression, and differences were considered statistically significant at $P < 0.05$.

3. Results

3.1. Comparison of Clinical Data between the Case Group and the Control Group. Other than the mean age, smoking history, uric acid, and history of stroke, other clinical indicators were not significantly different between the two groups ($P > 0.05$). See Table 1.

3.2. Between-Group Comparison of Level Values of Serum C-Peptide, sICAM-1, and IL-1β. Compared with the control group, the level values of serum c-peptide, sICAM-1, and IL-1β were significantly higher in the case group ($P < 0.001$). See Table 2.
3.3. Correlation of Serum C-peptide, sICAM-1, and IL-1β with CSVD. With CSVD being the dependent variable, and age, smoking, uric acid, history of stroke, serum c-peptide, sICAM-1, and IL-1β being the independent variables, a logistic regression analysis was conducted, and the result showed that age, smoking, serum c-peptide, sICAM-1, and IL-1β were the risk factors for CSVD. See Table 3.

3.4. ROC Curve Analysis on Serum C-Peptide, sICAM-1, IL-1β, and CSVD. Figure 1 shows the ROC curve analysis on serum c-peptide, sICAM-1, IL-1β, and CSVD.

3.5. Comparison of Areas under the Curves among Various Indicators. The area under the curve of sICAM-1 was larger than that under other single indicator. See Table 4.

4. Discussion

As the population aging deepens in recent years, the prevalence of senile dementia is increasing, which not only puts great pressure on society and families but also seriously affects the physical and mental health of the elderly population, making it an important cause of death in them [12]. CSVD is an intracranial small vessel lesion in sites such as arterioles, small veins, and capillaries caused by a combination of factors, which damages the white matter and gray matter in depth of brain and finally leads to a series of pathological and neuroimaging changes, thereby resulting in cognitive impairment and other clinical syndromes in patients [13]. Studies have confirmed [14] that CSVD is not only related to vascular cognitive dysfunction but also to the pathogenesis of neurodegenerative diseases such as Alzheimer’s disease (AD), and as these two diseases are overlapped to some extent, most AD patients have signs of CSVD, including cognitive dementia, progressive cognitive decline, bladder sphincter dysfunction, and mental disorders [15]. Currently, there is no consensus on CSVD pathogenesis, but it may be due to the fact that chronic progressive subclinical ischemia in the brain increases vascular

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### Table 1: Between-group comparison of clinical data.

| Item                              | Case group | Control group | X²/t  | P     |
|-----------------------------------|------------|---------------|-------|-------|
| Gender                            |            |               |       |       |
| Male/female                       | 37/35      | 40/32         | 0.251 | 0.616 |
| BMI (mean ± SD, kg/m²)            |            |               |       |       |
| 21.23 ± 1.72                      | 21.63 ± 1.94| 1.309         | 0.193 |
| Mean age (mean ± SD, years)       |            |               |       |       |
| 64.68 ± 8.23                      | 54.60 ± 5.73| 8.529         | <0.001|
| Smoking history                   |            |               |       |       |
| Yes                               | 53 (73.61%)| 21 (29.17%)   | 28.466| <0.001|
| No                                | 19 (26.39%)| 51 (70.83%)   |       |       |
| Drinking history                  |            |               |       |       |
| Yes                               | 40 (55.56%)| 42 (58.33%)   | 0.113 | 0.736 |
| No                                | 32 (44.44%)| 30 (41.67%)   |       |       |
| History of hypertension           |            |               |       |       |
| Yes                               | 40 (55.56%)| 43 (59.72%)   | 0.256 | 0.613 |
| No                                | 32 (44.44%)| 29 (40.28%)   |       |       |
| History of coronary heart disease |            |               |       |       |
| Yes                               | 40 (55.56%)| 39 (54.17%)   | 0.028 | 0.867 |
| No                                | 32 (44.44%)| 33 (45.83%)   |       |       |
| Fasting blood glucose value (mean ± SD, mmol/L) | 5.42 ± 1.09 | 5.28 ± 1.17 | 0.743 | 0.459 |
| TC (mean ± SD, mmol/L)            |            |               |       |       |
| 4.28 ± 0.76                       | 4.05 ± 0.83 | 1.734         | 0.085 |
| Triacylglycerol (mean ± SD, mmol/L)| 1.16 ± 0.41| 1.19 ± 0.38 | 0.455 | 0.650 |
| Uric acid (mean ± SD, μmol/L)     |            |               |       |       |
| 324.64 ± 19.69                    | 243.51 ± 12.51| 29.510       | <0.001|
| History of stroke                 |            |               |       |       |
| Yes                               | 20 (27.78%)| 2 (2.78%)     | 17.383| <0.001|
| No                                | 52 (72.22%)| 70 (97.22%)   |       |       |

### Table 2: Between-group comparison of level values of serum c-peptide, sICAM-1, and IL-1β (mean ± SD).

| Group               | n  | Serum c-peptide (pmol/L) | sICAM-1 (ng/ml) | IL-1β (ng·L⁻¹) |
|---------------------|----|--------------------------|-----------------|----------------|
| Case group          | 72 | 1.54 ± 0.09               | 153.96 ± 4.90   | 21.16 ± 0.75   |
| Control group       | 72 | 0.77 ± 0.14               | 101.37 ± 1.56   | 15.36 ± 1.29   |
| t                   |    | 39.257                    | 87.778          | 33.323         |
| P                   |    | <0.001                    | <0.001          | <0.001         |

### Table 3: Multi-factor logistic regression analysis on CSVD.

| Item                              | B    | SE    | Wald  | P     | OR   |
|-----------------------------------|------|-------|-------|-------|------|
| Age                               | 0.168| 0.073 | 0.016 | 0.006 | 1.253|
| Smoking                           | 4.235| 1.376 | 7.263 | 0.012 | 0.202|
| Uric acid                         | 0.527| 0.326 | 5.273 | 0.427 | 1.836|
| History of stroke                 | 16.278| 4.721| 0.263 | 0.724 | 2.367|
| Serum c-peptide                   | 0.753| 0.352 | 5.261 | 0.025 | 2.031|
| sICAM-1                           | 0.867| 0.386 | 6.261 | 0.031 | 2.638|
| IL-1β                             | 0.536| 0.137 | 8.025 | 0.006 | 1.362|
permeability, causing extravasation of intravascular substances, disruption of the blood-brain barrier, local inflammatory response, and then resulting in vascular and perivascular tissue damage [16].

Brain MRI has become a common imaging modality at present, and it applies radiofrequency pulses of specific frequency to the human body in the static magnetic field, so that the hydrogen protons in the human body are stimulated and then cause magnetic resonance; after stopping the pulse, the protons produce MR signal during relaxation, and with the continuous promotion of MRI imaging sequence and image, brain MRI provides a good basis for the diagnosis of brain lesions [17]. Moreover, MRI is effective in detecting cerebral small vessel injury because it is noninvasive and easy to operate. However, applying MRI for early screening of CSVD is more costly and time-consuming, so serologic markers have become a new research direction.

A foreign meta-analysis found that [18] compared with nonstroke patients, CSVD patients have significantly increased serum c-reactive protein (CRP), tumor necrosis factor-α (TNF-α), and interleukin-6 (IL-6), indicating that the inflammatory process participates in the pathomechanism of CSVD. The activation and amplification of inflammatory response is an important factor in the development and progression of inflammatory diseases, and the immune inflammasome is the initiator of inflammation. Current studies have confirmed that [19] the increased expression level of NLRP3 inflammasome is closely related to the occurrence and progression of chronic obstructive pulmonary disease, malignancy, and chronic viral hepatitis. As a macromolecular multiprotein complex, NLRP3 inflammasome functions to regulate the secretion and maturation of downstream proinflammatory cytokines such as IL-8 and IL-1β. These proinflammatory cytokines produced by the activated NLRP3 inflammasome can mediate the activation of the NF-kB signaling pathway and induce the occurrence of brain neuroinflammation. When cerebral ischemia occurs, IL-1β and other inflammatory mediators can activate local endothelial cells and leukocytes and induce significant upregulation of the number of adhesion molecules on the cell surface, which leads to massive adhesion between leukocytes and endothelial cells, causing microvascular obstruction and aggravating brain tissue damage [20, 21]. High insulin level is a well-established risk factor for atherosclerosis, whereas microvascular endothelial cells are vulnerable to insulin metabolism and growth promoting effects. Serum c-peptide can indirectly reflect the concentration of insulin, and abnormally increased insulin content together with insulin resistance is an independent risk factor contributing to the development of CSVD [22]. SICAM-1 is an inflammatory factor secreted by vascular endothelial cells that mediate adhesion of platelets, leukocytes, and vascular endothelium and is involved in blood-brain barrier (BBB) disruption and brain tissue damage induced by chronic cerebral ischemia, and high levels of SICAM-1 may also increase the risk of future recurrent stroke and vascular dementia [23].

The study found that compared with the control group, the level values of serum c-peptide, SICAM-1, IL-1β, and CSVD.

**Figure 1:** ROC curve analysis on serum c-peptide, SICAM-1, IL-1β, and CSVD.

**Table 4:** Comparison of areas under the curves among various indicators.

| Test result variable | Area   | SE   | Asymp. sig. | Asymp. 95% CI | Lower limit | Upper limit |
|----------------------|--------|------|-------------|----------------|-------------|-------------|
| Serum c-peptide      | 0.819  | 0.037| 0.000       | 0.747 0.892   |             |             |
| SICAM-1              | 0.840  | 0.035| 0.000       | 0.771 0.910   |             |             |
| IL-1β                | 0.813  | 0.038| 0.000       | 0.739 0.886   |             |             |
two subgroups (with or without acute cerebral infarction) for intercomparison.

**Data Availability**

Data to support the findings of this study are available on reasonable request from the corresponding author.

**Conflicts of Interest**

The authors have no conflicts of interest to declare.

**References**

[1] J. Chung, S. Marini, J. Pera et al., “Genome-wide association study of cerebral small vessel disease reveals established and novel loci,” *Brain*, vol. 142, pp. 3176–3189, 2019.

[2] Y. Ma, A. Song, V. Anand et al., “Blood pressure variability and cerebral small vessel disease: a systematic review and meta-analysis of population-based cohorts,” *Stroke*, vol. 51, pp. 82–89, 2020.

[3] C. A. J. L. Rutten and S. N. Rost, “Emerging insights from the genetics of cerebral small-vessel disease,” *Annals of the New York Academy of Sciences*, vol. 1471, pp. 5–17, 2020.

[4] X. Liu, L. Tan, S. Diao et al., “The global burden of cerebral small vessel disease related to neurological deficit severity and clinical outcomes of acute ischemic stroke after IV rt-PA treatment,” *Neurological Sciences*, vol. 40, pp. 1157–1166, 2019.

[5] Y. Chen, H. Yu, J. Zhu et al., “Low carotid endothelial shear stress associated with cerebral small vessel disease in an older population: a subgroup analysis of a population-based prospective cohort study,” *Atherosclerosis*, vol. 288, pp. 42–50, 2019.

[6] S. C. Elizabeth, R. D. Jorgensen, J. P. Gianaros, M. Joseph, and R. Caterina, “Cerebrovascular disease: neuroimaging of cerebral small vessel disease,” *Prog Mol Biol Transl Sci*, vol. 165, pp. 225–255, 2019.

[7] M. V. S. Uniken, M. Sandro, K. U. Lena et al., “Impact of cerebral small vessel disease on functional recovery after intracerebral hemorrhage,” *Stroke*, vol. 50, pp. 2722–2728, 2019.

[8] S. Mahdiye, K. Ahmad, P. Fereshteh, A. Ataie, M. Monif, and H. R. Nouri, “Involvement of NLRP4 inflammasome through caspase-1 and IL-1β augments neuroinflammation and contributes to memory impairment in an experimental model of Alzheimer’s like disease,” *Brain Research Bulletin*, vol. 154, pp. 81–90, 2020.

[9] J. B. Jiménez, R. L. Iolanda, J. Pizarro et al., “Kidney function changes and their relation with the progression of cerebral small vessel disease and cognitive decline,” *Journal of the Neurological Sciences*, vol. 409, Article ID 116635, 2020.

[10] World Medical Association, “World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects,” *JAMA*, vol. 310, no. 20, pp. 2191–2194, 2013.

[11] S. R. Natalia and E. Mark, “Cerebral small vessel disease,” *Continuum*, vol. 26, pp. 332–352, 2020.

[12] T. Katie, G. Cem, S. Thierry et al., “Levels of CXCL13 and sICAM-1 correlate with disease activity score in patients with rheumatoid arthritis treated with tocilizumab,” *Adv Rheumatol*, vol. 59, 2019.

[13] F. Bieuzen, C. Hausswirth, and B. Dugué, “Circulating soluble intercellular adhesion molecule-1 (sICAM-1) after exercise-induced muscular damage: does the use of whole-body cryostimulation influence its concentration in blood?” *Cryobiology*, vol. 87, pp. 120–122, 2019.

[14] M. Li, J. Wang, X. Wang et al., “Clinical efficacy of aspirin combined with clopidogrel in treating cerebral infarction and its effect on serum hs-CRP, sICAM-1 and TNF-a,” *Experimental and Therapeutic Medicine*, vol. 19, pp. 939–944, 2020.

[15] X. Chen, H. Feng, H. Lin, X. Xianrong, W. Jianchang, and J. Zhanguo, “Carotid imaging changes and serum IL-1β, sICAM-1, and sVAP-1 levels in benign paroxysmal positional vertigo,” *Scientific Reports*, vol. 10, Article ID 21494, 2020.

[16] K. Strzelczyk, C. Piotr, B. Benjamin et al., “The levels of sICAM-1, sELAM-1, TNFα and sTNFR1 proteins in patients with colorectal adenocarcinoma in tumor and corresponding normal mucosa,” *Acta Biochimica Polonica*, vol. 67, pp. 579–585, 2020.

[17] X. Li, Y. Zhou, R. Zou et al., “β Associations of serological biomarkers of sICAM-1, IL-1, MIF, and su-PAR with 3-month mortality in acute exacerbation of idiopathic pulmonary fibrosis,” *Mediators of Inflammation*, vol. 2020, Article ID 4534272, 9 pages, 2020.

[18] P. Eudoxie, I. Ioanna, P. Despina, A. Pantopoulou, G. Agroianiss, and I. Vrotsos, “The effect of experimental periodontitis, experimental diabetes and their combination on the serum levels of adiponectin, leptin, IL-6, IL-18, MCP-1, RANTES and sICAM-1 in rats,” *Journal of the International Academy of Periodontology*, vol. 22, pp. 1–10, 2020.

[19] M. Jakob, C. A. Andrea, O. J. Lamontagne et al., “Distinct effects of complement and of NLRP3- and non-NLRP3 inflammasomes for choroidal neovascularization,” *Elife*, vol. 9, 2020.

[20] L. H. Chung, Y. J. Chen, W. Y. Huei et al., “Cbl negatively regulates NLRP3 inflammasome activation through GLUT1-dependent glycolysis inhibition,” *International Journal of Molecular Sciences*, vol. 21, 2020.

[21] X. Fang, Y. Wang, Y. Zhang, L. Yelin, K. K. Joanne, and W. Li, “NLRP3 inflammasome and its critical role in gynecological disorders and obstetrical complications,” *Frontiers in Immunology*, vol. 11, Article ID 555826, 2020.

[22] D. C. Candela, F. M. Reiner, R. N. Bonetti et al., “AP-1 (activated protein-1) transcription factor JunD regulates ischæmia/reperfusion brain damage via IL-1β (Interleukin-1β),” *Stroke*, vol. 50, pp. 469–477, 2019.

[23] A. M. Gregory, M. A. Leticia, S. G. Sánchez et al., “Soluble intercellular adhesion molecule-1 (sICAM-1) as a biomarker of vascular cognitive impairment in older adults,” *Dementia and Geriatric Cognitive Disorders*, vol. 47, pp. 243–253, 2019.

[24] Z. Zhou, H. Li, S. Tian et al., “Critical roles of NLRP3 inflammasomes in IL-1β secretion induced by Corynebacterium pseudotuberculosis in vitro,” *Molecular Immunology*, vol. 116, pp. 11–17, 2019.