Pharmacogenetics of Antiplatelet Drugs

Ronan Curtin* and Desmond J. Fitzgerald

Department of Clinical Pharmacology, Royal College of Surgeons in Ireland, 123 St. Stephen’s Green, Dublin 2, Ireland

Received December 7, 2001; Accepted February 1, 2002; Published March 21, 2002

Pharmacogenetics refers to the genetic factors that influence the response to a drug, often involving genetic variations in drug metabolizing enzymes. The pharmacogenetics of antiplatelet agents is in its infancy and largely reflects variations in drug targets or related genes. One particular gene variant, the PlA2 polymorphism of the glycoprotein (GP) IIb/IIIa receptor, is now emerging as a probable determinant of the response to antiplatelet agents including GPIIb/IIIa antagonists. This variant may in part explain the heterogeneity in the response to GPIIb/IIIa antagonists. The PlA2 genotype appears to be associated with an adverse outcome in patients treated with an oral GPIIb/IIIa antagonist and may be a factor in the observed failure of these agents in unselected populations. However, there are preliminary indications that other antiplatelet agents may have an enhanced effect in PlA2 subjects. Further clinical trials in particular are required to definitively characterize the pharmacogenetic effect of PlA2. Other polymorphisms are also likely to contribute to the pharmacogenetics of antiplatelet agents, but these await investigation.

KEY WORDS: pharmacogenetics, PlA2, GPIIb/IIIa receptor, aspirin, dipyridamole, clopidogrel, GPIIb/IIIa antagonists

DOMAINS: thrombosis, pharmacogenomics

INTRODUCTION

Age, concurrent medications, and renal and liver dysfunction are well-characterized factors that can affect the efficacy or toxicity of drug treatment. Inherited factors are also emerging as determinants of the pharmacokinetics and pharmacodynamics of many medications. Pharmacogenetics is that field of science concerned with this hereditary component of drug response. Variations in genes that encode proteins involved in drug metabolism can be important determinants of a patient’s response to therapy. Genetic variations in many drug metabolizing enzymes and their phenotypes have been characterized[1]. However, genetic variation in drug transporters and drug targets, as well as genetic heterogeneity underlying the disease being treated, can also modulate response to medications.
Initially, identification of inherited differences in drug metabolism was based on observations in patients who suffered adverse effects despite being treated with standard doses of a drug[2]. However the focus of research in pharmacogenetics, triggered in part by the human genome project, has shifted to identification of single nucleotide polymorphisms (SNPs) in target genes. Drug trials or disease cohorts provide an opportunity to determine whether such SNPs modify drug response. For example, a variant of the $\beta_2$ adrenergic receptor enhances the response to $\beta$-agonist therapy in asthmatics, as the receptor fails to desensitize[3]. Variants in genes that may not be targets for drugs but are involved in the disease may also influence drug response. For example, recent evidence suggests that the angiotensin converting enzyme (ACE) DD genotype modifies the response to $\beta$-blocker therapy in heart failure[4]. Of particular importance in platelet biology and pharmacology are its surface receptors. Some of these receptors, including glycoprotein (GP) IIb/IIIa are highly polymorphic[5]. It is likely that genetic variation in these receptors is the major determinant of response to antiplatelet agents. This review will focus on the pharmacogenetics of the PI$^{A2}$ polymorphism of GPIIIa. The potential influence of other polymorphisms on antiplatelet therapy awaits investigation.

**THE PI$^{A2}$ POLYMORPHISM**

The GPIIb/IIIa receptor ($\alpha_2\beta_3$) is critical to platelet aggregation and thrombus formation (see Fig. 1). Following platelet activation by agonists such as collagen, ADP, thrombin, thromboxane A$_2$ and epinephrine, the GPIIb/IIIa receptor undergoes a conformational change that allows it to bind fibrinogen and form platelet aggregates[6]. The Platelet Antigen (PI$^{A}$) polymorphism results in a leucine (PI$^{A1}$) to proline (PI$^{A2}$) substitution at position 33 in the extracellular portion of the IIIa subunit of GPIIb/IIIa[7]. It is responsible for most cases of post-transfusion purpura or neonatal alloimmune thrombocytopenic purpura in the Caucasian population, the amino acid substitution causing a conformational change in IIIa and exposure of novel epitopes[5]. The PI$^{A2}$ polymorphism is common in non-Asians with about 20% heterozygous and 2% homozygous for the mutation[8].

Given the importance of the GPIIb/IIIa receptor in platelet aggregation, it was postulated that the PI$^{A2}$ polymorphism might also affect thrombotic risk. Weiss et al.[9] in a small case-control study, showed that the PI$^{A2}$ polymorphism was associated with a 2.8-fold (95%CI 1.2–6.4) increased risk of myocardial infarction or angina. In patients less than 60 years of age, the association appeared stronger with a relative risk (RR) of 6.2 (95%CI 1.8–22.4). More than 30 case-control studies have been published since and most have not confirmed the PI$^{A2}$ genotype as a risk factor for coronary thrombosis. There have been two recent meta-analyses of these studies, the most recent of which suggests a weak association between the PI$^{A2}$ polymorphism and cardiovascular disease (RR 1.10, 95%CI 1.03–1.18)[10,11]. This is not surprising, as any single variant is unlikely to contribute in a major way to the risk of coronary artery thrombosis. Indeed, there is evidence of interactions between the PI$^{A2}$ genotype and environmental factors[12] and other gene variants[13].

Attempts to define the effect of the PI$^{A2}$ polymorphism on platelet function have yielded apparently conflicting results. Although one study has shown an increase in fibrinogen binding in response to ADP[14], others have failed to show an increased affinity of the PI$^{A2}$ receptor for soluble fibrinogen[15,16,17]. Vijayan et al.[16] have shown increased binding by two cell lines (Chinese Hamster Ovary and Human Embryonal Kidney 293 cells) expressing the PI$^{A2}$ isoform of GPIIb/IIIa to immobilized but not soluble fibrinogen. However, Bennett et al.[17] did not reproduce this finding in a B-lymphocyte cell line.

PI$^{A2}$ need not affect affinity of GPIIb/IIIa for its ligand to influence thrombosis. The variant may instead affect thrombus formation by modifying “outside-in” signaling by the receptor.
Outside-in signaling describes the process whereby, after ligand binding to GPIIb/IIIa, information is transmitted into the platelet that is required for aggregation[18]. Vijayan et al.[16] have shown increased peripheral F actin content, cell spreading, and fibrin clot retraction and a modest increase in pp125<sup>FAK</sup> phosphorylation, events characteristic of outside-in signaling, in two cell lines expressing Pl A<sup>2</sup> after binding to immobilized fibrinogen. Platelet α-granule release is another consequence of ligand binding to the GPIIb/IIIa receptor. Michelson et al.[19] have shown that Pl A<sup>1/A2</sup> and Pl A<sup>2/A2</sup> platelets express greater amounts of an α-granule constituent, P-selectin, in a gene-dose manner both at rest and following stimulation with a range of concentrations of ADP.

In <em>vitro</em> measurement of platelet aggregation by turbidometric assay is the standard in determining the efficacy of antiplatelet drugs. However, this method detects only larger platelet aggregates and may be relatively insensitive for detecting platelet hyperfunction[20]. Studies assessing the effect of Pl A<sup>2</sup> on platelet aggregation have been most inconsistent. The largest of these, a cohort of 1,422 participants in the Framingham Offspring study, concluded that Pl A<sup>2</sup> was associated with increased platelet reactivity as evidenced by a lower aggregation threshold concentration to epinephrine and ADP[21]. In addition, Andrioli et al.[22] found increased sensitivity of Pl A<sup>2</sup> platelets to thromboxane A<sub>2</sub>. Although one study has reproduced the finding of increased sensitivity to epinephrine in those homozygous for Pl A<sup>2</sup>[19], others have yielded apparently conflicting results. Lasne et al.[23] showed reduced sensitivity to ADP and TRAP in 8
and 14 PlA2 carriers respectively, and Bennett et al.[17] failed to show any effect of PlA2 on TRAP-induced aggregation over a range of concentrations of the agonist.

ASPIRIN

Aspirin is an effective antiplatelet agent. It causes irreversible inhibition of cyclooxygenase (COX)-1 in platelets, thereby preventing formation of prostaglandin endoperoxides and thromboxane A2, a platelet agonist. Aspirin has been conclusively shown to reduce the risk of further thromboembolic events in patients with a history of cardiovascular, cerebrovascular, and peripheral vascular disease[24]. Yet some patients continue to suffer thromboembolic events despite treatment with aspirin. This may reflect the fact that aspirin is a relatively weak antiplatelet agent. However it has been suggested that some patients are resistant to the antiplatelet effects of aspirin[25]. The term aspirin nonresponder has been used to describe those patients who show a reduced response to aspirin as measured by platelet function assays. In particular, up to 40% of patients on aspirin exhibit little or no prolongation of time to clotting in the bleeding-time test and more recently the Platelet Function Analyser (PFA-100) despite adequate suppression of platelet thromboxane A2 production[26,27]. Whether these definitions of aspirin nonresponse have any impact on clinical outcome has not been established[28,29].

There is evidence to suggest that Pl A2 may modulate response to therapy with aspirin as measured by these assays. In one study of healthy volunteers a shorter baseline bleeding time was reported in 26 PlA2 carriers vs. 54 PlA1A1 controls[30]. Following ingestion of 300 mg aspirin, the bleeding time prolonged by on average 46.5 sec in carriers vs. 112.5 sec in noncarriers, exaggerating a difference found at baseline. Bleeding time was shortened in 7 of the 26 carriers vs. 1 of the 54 controls (p = 0.001). The data suggest that PlA2 accounts in part for aspirin nonresponders.

Platelets are an important contributor to thrombin generation[31]. In addition to bleeding time, Szczeklik et al.[30] analyzed the levels of peripheral blood prothrombin fragment 1.2 (F1.2), a marker of thrombin generation, in carriers and controls before and after aspirin ingestion. Lower levels of F1.2 were found at baseline in PlA2 carriers vs. controls (p = 0.02). After treatment with aspirin, F1.2 levels showed a mild tendency to increase in carriers and decrease in controls, attenuating the baseline differences between the groups. A previous report from this group also showed reduced suppression of thrombin generation in PlA2 carriers on aspirin when measured at the site of vascular injury. Levels of F1.2 were similar at baseline in bleeding time blood in both groups, but after ingestion of aspirin only 9 of 15 PlA2 carriers vs. 23 of 25 PlA1A1 subjects showed reduced thrombin generation. The relative risk of aspirin nonresponse was 6.43 (95%CI 1.38–30.02) in carriers of the Pl A2 allele. More recently this group has further characterized the effect of PlA2 on thrombin generation in the bleeding-time blood of healthy volunteers before and after aspirin ingestion. By measuring prothrombin and a number of thrombin-mediated events, such as thrombin-antithrombin III production, thrombin B-chain formation, fibrinogen consumption, and factor Va generation, they showed evidence of enhanced and more rapid thrombin generation in PlA2 carriers. Aspirin ingestion resulted in a significant reduction in the velocity of all these processes in PlA1A1 subjects but not in PlA2 carriers[32].

Paradoxically, three separate studies have suggested greater sensitivity to aspirin in those with the PlA2 polymorphism when measured by platelet aggregation in vitro. Cooke et al. have shown increased inhibition of epinephrine induced platelet aggregation with aspirin in a study of 11 PlA1A2 subjects and 15 matched PlA1A1 controls[33]. Aggregation in the absence of aspirin was similar in both groups, but there was a tenfold reduction of the IC50 for aspirin with PlA1A2 platelets (p = 0.005). A subsequent study from the same group confirmed increased inhibition of epinephrine induced platelet aggregation by aspirin in platelets from PlA1A2 subjects, but there was a trend towards an opposite effect in platelets from PlA2A2 subjects[19]. In an analysis of 17
PI$^{A1/A1}$, 13 PI$^{A1/A2}$, and 14 PI$^{A2/A2}$ subjects, the IC$_{50}$ for aspirin was lower in heterozygous vs. wildtype subjects (7.4 µmol/l vs. 13.1 µmol/l, p = 0.06). However, the IC$_{50}$ in PI$^{A2/A2}$ subjects was slightly higher (14.0 µmol/l) than in wildtype and significantly higher than in patients heterozygous for PI$^{A2}$ (p = 0.015).

The third study by Andrioli et al.[22] looked at the effect of acetylsalicylic acid on platelet aggregation in washed platelets after stimulation with arachidonic acid. Due to the small number of PI$^{A2/A2}$ samples (n = 2) they were analyzed together with the PI$^{A1/A2}$ samples (n = 12) and compared to PI$^{A1/A1}$ controls (n = 49). PI$^{A2}$ carriers were significantly more sensitive to inhibition by aspirin, with an IC$_{50}$ of $2.7 \pm 0.6$ µmol/l and $23.4 \pm 3.3$ µmol/l for PI$^{A2}$ and PI$^{A1/A1}$ platelets, respectively (p < 0.005).

These apparently conflicting results using different assays of platelet function make it difficult to deduce what effect aspirin may have on platelet inhibition in those with the PI$^{A2}$ polymorphism. The bleeding-time test suffers from poor reproducibility and is sensitive to a host of factors not related to platelet function[34]. In vitro platelet aggregation is more sensitive and specific in detecting changes in platelet function; for example, it will detect changes in platelet reactivity in response to GPIIb/IIIa antagonists at much lower drug concentrations[35]. Prothrombin binds to both activated and resting GPIIb/IIIa. It has been suggested that the reduced number of occupied GPIIb/IIIa receptors resulting from enhanced inhibition of platelet aggregation by aspirin may allow greater prothrombin binding to the platelet surface and subsequent thrombin generation[36]. This theory could explain the contradictory outcome from bleeding-time tests and in vitro aggregation studies in PI$^{A2}$ carriers on aspirin. However, only a prospective study with clinical endpoints will determine whether PI$^{A2}$ has an effect on the response to aspirin.

**DIPYRIDAMOLE**

Dipyridamole is an antiplatelet agent that has been shown to be particularly effective in combination with aspirin in the secondary prevention of stroke[37]. Its mechanism of action includes inhibition of phosphodiesterase and inhibition of adenosine reuptake in platelets[38]. In vitro, dipyridamole reduces platelet adhesion, aggregation, and platelet factor IV availability with plasma concentrations of greater than 3.5 µmol/l[39]. The plasma concentration of dipyridamole may vary as much as tenfold between individuals[40], but no predictors of this variation have been described and no pharmacogenetic studies targeting dipyridamole have been conducted.

**THE THIENOPYRIDINES**

Ticlopidine and clopidogrel inhibit platelet activation by ADP by acting as antagonists of the recently cloned P2Y$_{12}$ purinergic receptor[41]. Clopidogrel is at least as effective as aspirin in reducing recurrent vascular events in high-risk patients[42]. Recently published results of the CURE study show that following an episode of unstable angina or a non-Q-wave-myocardial infarction (MI), combination therapy with aspirin and clopidogrel is superior to aspirin alone[43]. Whether all patients or only certain high-risk subgroups require combination therapy remains to be answered. Substudies from CURE may answer this question[44]. There is unpublished data indicating that some patients do not show platelet inhibition despite treatment with clopidogrel. However there are no known genetic predictors of response to clopidogrel or ticlopidine. Goldschmidt-Clermont et al.[45] studied the effect of clopidogrel in an unspecified number of PI$^{A1/A1}$ and PI$^{A1/A2}$ healthy volunteers and found no significant difference in platelet aggregation to a range of concentrations of epinephrine. Another potential target in pharmacogenetic studies is cytochrome P450-1A (CYP-1A), which is responsible for conversion of clopidogrel to its active metabolite[46]. Although polymorphisms have been described in this enzyme, their functional
significance remains uncertain[1]. In addition, genetic variation in the drug target (the recently identified P2Y12 receptor) could also modify the response to clopidogrel and ticlopidine.

GLYCOPROTEIN IIb/IIIa ANTAGONISTS

The GPIIb/IIIa antagonists are effective antiplatelet agents in the setting of percutaneous coronary intervention and acute coronary syndromes when administered as intravenous infusions[47]. However, attempts to provide prolonged GPIIb/IIIa blockade with oral antagonists has met with failure and may even increase the risk of thrombotic events[48]. Abciximab, the original GPIIb/IIIa antagonist, is a chimeric murine antibody directed against GPIIb/IIIa. Subsequent intravenous and oral compounds are synthetic peptide and peptidomimetic compounds modeled on the Arg-Gly-Asp (RGD) or Lys-Gly-Asp (KGD) recognition sequences of the fibrinogen α and γ chains, respectively[49].

The PiA2 polymorphism does not appear to alter the affinity of the receptor for small-molecule GPIIb/IIIa antagonists. The K, for RGDS has been shown to be similar in PiA1/A1 and PiA1/A2 platelets[17]. Similarly a peptidomimetic GPIIb/IIIa antagonist RWJ 53308, modeled on the KQAGD sequence of the carboxyl terminus of the fibrinogen γ chain, shows similar binding to the receptor, regardless of PiA2 allotype. In addition, RWJ 53308 showed similar inhibition of TRAP-stimulated platelet aggregation at various concentrations of the antagonist in PiA1/A1, PiA1/A2, and PiA2/A2 platelets[17]. However, in a separate study of 23 PiA2 carriers (20 PiA1/A2, 3 PiA2/A2) and 24 PiA1/A1 controls another GPIIb/IIIa antagonist modeled on KGD, eptifibatide, appeared to be less effective in inhibiting platelet aggregation in those with the polymorphism[50]. Eptifibatide inhibited both ADP- (60 and 100 nM) and collagen- (40 nM) induced aggregation less strongly in PiA2 carriers with IC50 values of 115 vs. 67 nM (p < 0.05) and 141 vs. 110 nM (p < 0.05), respectively.

On the other hand, abciximab has been reported to be more effective in inhibiting ADP-induced platelet aggregation in carriers[19]. In an analysis of 20 PiA1/A1, 20PiA1/A2, and 15PiA2/A2 patients, there was an enhanced inhibitory response to abciximab at therapeutic concentrations in PiA1/A2 compared to PiA1/A1 and PiA2/A2 patients (p = 0.099). The inhibitory effect was similar in PiA1/A1 and PiA2/A2 subjects. These preliminary results would appear to indicate that PiA2 differentially influences the response to at least some GPIIb/IIIa antagonists.

Two substudies of oral GPIIb/IIIa antagonist trials suggest that PiA2 may interfere with the response to these agents. In a substudy of SYMPHONY (TIMI 12), 98 patients on sibrafiban were analyzed for an effect of PiA genotype on platelet aggregability and events[51]. Although there was no significant difference in platelet inhibition as measured by ADP-induced aggregation, recurrent MI occurred more commonly in the PiA2 group (7.4 vs. 0%, p = 0.02).

In OPUS, a clinical trial of the oral GPIIb/IIIa antagonist orbofiban, 10,288 patients from 29 countries were recruited within 72 h of an acute coronary syndrome and randomized to one of three groups: (1) orbofiban 50 mg twice daily for 30 days followed by 30 mg twice daily; (2) orbofiban 50 mg twice daily; or (3) placebo[52]. A subgroup of 1,014 patients, the majority of whom were from the U.S. and Canada, donated a genetic sample at a mean of 157 days. These patients were genotyped for the PiA2 polymorphism and its effect on response to orbofiban was examined[53]. The rate of MI was significantly higher among all PiA2 carriers (RR = 2.71, 95% CI 1.37–5.38, p = 0.004). When PiA2 carriers were broken down by treatment, the relative risk of a recurrent MI in those on orbofiban and placebo were 4.27 and 1.04, respectively (p < 0.001) (see Table 1). Of those randomized to treatment with orbofiban, PiA2 carriers had a more than fourfold increased risk of MI vs. noncarriers (2.46 vs. 0.59, p = 0.08). In addition, while there was an expected dose-dependent excess of risk of bleeding events in the noncarriers on treatment, PiA2 carriers showed no increased bleeding risk on treatment. The relative risk of a bleeding event on
TABLE 1
Relative Risk (95% Confidence Interval) for Primary End-Point, Myocardial Infarction, and Bleeding in PlA2 Carriers According to Treatment

| Event       | Placebo          | Treated                  |
|-------------|------------------|--------------------------|
| Primary endpoint$^{1,2}$ | 0.87 (0.49, 1.55) | 1.55 (1.03, 2.34)$^3$   |
| MI$^2$      | 1.04 (0.28, 3.92) | 4.27 (1.82, 10.03)$^4$  |
| Bleeding$^5$| 1.22 (0.67, 2.22) | 0.59 (0.39, 0.92)$^3$  |

*Note:* Risks are relative to PlA2 noncarriers.

$^1$A composite of death, MI, recurrent ischemia at rest leading to rehospitalization, or urgent revascularization or stroke.

$^2$Results of a proportional hazards analysis adjusted for age and sex.

$^3p < 0.05$.

$^4p < 0.001$.

$^5$Results of a proportional hazards analysis adjusted for age, sex, and country.

orbofiban was 1.87 (95%CI 1.29, 2.71) in PlA2 noncarriers vs. 0.87 (95%CI 0.46, 1.64) in PlA2 carriers ($p = 0.05$). The results of this substudy indicate a reduced response to orbofiban or even a negative interaction between drug and PlA2 genotype. A possible explanation for these results is that PlA2 carriers may have a greater propensity to partial agonism when treated with GPIIb/IIIa antagonists[54]. In the setting of coronary artery disease where there is enhanced platelet activation, GPIIb/IIIa antagonists may trigger outside-in signaling and so cause further platelet activation[55]. If plasma drug levels are too low to prevent fibrinogen binding (which occurs with the oral administration of these agents), this enhanced platelet activation may ultimately lead to thrombus formation and increased clinical events.

CONCLUSION

One single nucleotide polymorphism, the PlA2 variant of GPIIIa, appears to modulate the response to antiplatelet agents and GPIIb/IIIa antagonists, although studies report conflicting results. This is not surprising given the millions of single nucleotide polymorphisms in the human genome. It is very likely that many variants are involved in determining the risk of a polygenic disease such as coronary thrombosis and the response to treatment, with each variant contributing modestly to the heterogeneity in drug response. Indeed, given the number of potential variants and resulting haplotypes, it is surprising that studies have detected any effect of the PlA2 variant. Thus, conflicting results may reflect differences in the presence of other gene variants and gene–gene interactions, as well as environmental factors. It is also possible that PlA2 is in linkage with functional polymorphisms close to the GPIIIa gene and is simply a marker of another variant. Larger studies are necessary to explore this issue and to define the role of variants in other genes. Only then will it be possible to profile genes that determine the response to antithrombotic drugs and ultimately provide a means of tailoring therapy for individual patients.

ACKNOWLEDGEMENTS

Ronan Curtin is in receipt of a European Society of Cardiology postdoctoral research fellowship in coronary thrombosis and has also received an Aventis Grant-in-Aid for Cardiovascular trainees.
REFERENCES

1. Evans, W.E. and Relling, M.V. (1999) Pharmacogenomics: translating functional genomics into rational therapeutics. Science 286, 487–491.
2. Gonzalez, F.J., Skoda, R.C., Kimura, S., Umeno, M., Zanger, U.M., Gelboin, H.V., Hardwick, J.P., and Meyer, U.A. (1988) Characterization of the common genetic defect in humans deficient in debrisoquine metabolism. Nature 331, 442–446.
3. Martinez, F.D., Graves, P.E., Baldini, M., Solomon, S., and Erickson, R. (1997) Association between genetic polymorphisms of the β2-adrenoceptor and response to albuterol in children with and without a history of wheezing. J. Clin. Invest. 100, 3184–3188.
4. McNamara, D.M., Holubkov, R., Janosko, K., Palmer, A., Wang, J.J., MacGowan, G.A., Murali, S., Rosenblum, W.D., London, B., and Feldman, A.M. (2001) Pharmacogenetic interactions between beta-blocker therapy and the angiotensin-converting enzyme deletion polymorphism in patients with congestive heart failure. Circulation 103, 1644–1648.
5. Newman, P.J. (1991) Platelet GPIIb-IIIa: molecular variations and alloantigens. Thromb. Haemost. 66, 111–118.
6. Coller, B.S. (1986) Activation affects access to the platelet receptor for adhesive glycoproteins. J. Cell Biol. 103, 451–456.
7. Newman, P.J., Derbes, R.S., and Aster, R.H. (1989) The human platelet alloantigens, Pl A1 and Pl A2, are associated with a leucine33/proline33 amino acid polymorphism in membrane glycoprotein IIIa, and are distinguishable by DNA typing. J. Clin. Invest. 83, 1778–1881.
8. Kim, H.O., Jin, Y., Kickler, T.S., Blakemore, K., Kwon, O.H., and Bray, P.F. (1995) Gene frequencies of the five major human platelet antigens in African American, white, and Korean populations. Transfusion 35, 863–867.
9. Weiss, E.J., Bray, P.F., Tayback, M., Schulman, S.P., Kickler, T.S., Becker, L.C., Weiss, J.L., Gerstenblith, G., and Goldschmidt-Clermont, P.J. (1996) A polymorphism of a platelet glycoprotein receptor as an inherited risk factor for coronary thrombosis. N. Engl. J. Med. 334, 1090–1094.
10. Zhu, M.M., Weedon, J., and Clark, L.T. (2000) Meta-analysis of the association of platelet glycoprotein IIIa PlA1/PlA2 polymorphism with myocardial infarction. Am. J. Cardiol. 86, 1000–1005.
11. Di Castelnuovo, A., de Gaetano, G., Donati, M.B., and Iacoviello, L. (2001) Platelet glycoprotein receptor IIIa polymorphism PlA1/PlA2 and coronary risk: a meta-analysis. Thromb. Haemost. 85, 626–633.
12. Ardissino, D., Mannucci, P.M., Merlino, P.A., Duca, F., Fetiveau, R., Tagliabue, L., Tubaro, M., Galvani, M., Ottani, F., Ferrario, M., Corral, J., and Margaglione, M. (2000) Prothrombotic genetic risk factors in young survivors of myocardial infarction. Blood 94, 46–51.
13. Pastinen, T., Perola, M., Niinii, P., Terwilliger, J., Salomaa, V., Vartiainen, E., Peltonen, L., and Syvanen, A. (1998) Array-based multiplex analysis of case candidate genes reveals two independent and additive genetic risk factors for myocardial infarction in the Finnish population. Hum. Mol. Genet. 7, 1453–1462.
14. Goodall, A.H., Curzen, N., Panes, M., Hurd, C., Knight, C.J., Ouwehand, W.H., and Fox, K.M. (1999) Increased binding of fibrinogen to glycoprotein IIIa-Proline33 (HPA-1b, Pl A2, Zw b) positive platelets in patients with cardiovascular disease. Eur. Heart J. 20, 742–747.
15. Meiklejohn, D.J., Urbiakai, S.J., and Greaves, M. (1999) Platelet glycoprotein IIIa polymorphism HPA 1b (Pl1b): no association with platelet fibrinogen binding. Br. J. Haematol. 105, 664–666.
16. Vijayan, K.V., Goldschmidt-Clermont, P.J., Roos, C., Bray, P.F. (2000) The Pl1b polymorphism of integrin β3 enhances outside-in signaling and adhesive functions. J. Clin. Invest. 105, 793–802.
17. Bennett, J.S., Catella-Lawson, F., Rut, A.R., Vilaire, G., Qi, W., Kapoor, S.C., Murphy, S., and FitzGerald, G.A. (2001) Effect of the Pl1b alloantigen on the function of the β3 integrins in platelets. Blood 97, 3093–3099.
18. Law, D.A., DeGuzman, F.R., Heiser, P., Ministri-Madrid, K., Killeen, N., and Phillips, D.R. (1999) Integrin cytoplasmic tyrosine motif is required for outside-in αIIbβ3 signalling and platelet function. Nature 401, 808–811.
19. Michelson, A.D., Furman, M.I., Goldschmidt-Clermont, P., Mascelli, M.A., Hendrix, C., Coleman, L., Hamlington, J., Barnard, M.R., Kickler, T., Christie, D.J., Kundu, S., and Bray, P.F. (2000) Platelet GP IIIa Pl1b polymorphisms display different sensitivities to agonists. Circulation 101, 1013–1018.
20. Gear, A.R. and Lambrecht, J.K. (1981) Reduction in single platelets during primary and secondary aggregation. Thromb. Haemost. 45, 298.
21. Feng, D., Lindpaintner, K., Larson, M.G., Rao, V.S., O’Donnell, C.J., Lipinska, I., Schmitz, C., Sutherland, P.A., Silberschatz, H., D’Agostino, R.B., Muller, J.E., Myers, R.H., Levy, D., and Toft, G.H. (1999) Increased platelet aggregability associated with platelet GPIIbα Pl1b polymorphism: the Framingham Offspring Study. Arterioscler. Thromb. Vasc. Biol. 19, 1142–1147.
22. Andrioli, G., Minuz, P., Solero, P., Pincelli, S., Ortolani, R., Lussignoli, S., and Bellavite, P. (2000) Defective platelet response to arachidonic acid and thromboxane A2 in subjects with Pl1b polymorphism of β3 subunit (glycoprotein IIIa). Br. J. Haematol. 110, 911–918.
23. Lasne, D., Krenn, M., Pingault, V., Arnaud, E., Fiesinger, J.N., Aiach, M., and Rendu, F. (1997) Interdonor variability of platelet response to thrombin receptor activation: influence of PI(A2) polymorphism. Br. J. Haematol. 99, 801–807.
24. The Antiplatelet Trialists' Collaboration. (1994) Collaborative overview of randomized trials of antiplatelet therapy - I. Prevention of death, myocardial infarction, and stroke by prolonged antiplatelet therapy in various categories of patients. Br. Med. J. 308, 81–106.
25. Doutremepuich, C., de Seze, O., Le Roy, D., Lalanne, M.C., and Anne, M.C. (1990) Aspirin at very ultra low dosage in healthy volunteers: effects on bleeding time, platelet aggregation, and coagulation. Hemostasis 20, 99–105.
26. Buchanan, M.R. and Brister, S.J. (1995) Individual variation in the effects of ASA on platelet function: implications for the use of ASA clinically. Can. J. Cardiol. 11, 221–227.
27. Gum, P.A., Kottke-Marchant, K., Poggio, E.D., Gurm, H., Welsh, P.A., Brooks, L., Sapp, S.K., and Topol, E.J. (2001) Profile and prevalence of aspirin resistance in patients with cardiovascular disease. Am. J. Cardiol. 88, 230–235.
28. Grotemeyer, K.H., Scharafinski, H.W., and Husstedt, I.W. (1993) Two-year follow-up of aspirin responder and aspirin nonresponder. A pilot study including 180 post-stroke patients. Thromb. Res. 71, 397–403.
29. Buchanan, M.R., Schwartz, L., Bourassa, M., Brister, S.J., Peniston, C.M., and the BRAT Investigators. (2000) Results of the BRAT study-a pilot study investigating the possible significance of ASA nonresponsiveness on the benefits and risks of ASA on thrombosis in patients undergoing coronary artery bypass surgery. Can. J. Cardiol. 16, 1385–1390.
30. Szczeklik, A., Undas, A., Sanak, M., Frolow, M., and Wegrzyn, W. (2000) Relationship between bleeding time, aspirin, and the Pl(A1/A2) polymorphism of platelet glycoprotein IIIa. Br. J. Haematol. 110, 965–967.
31. Crook, M. (1990) Platelet prothrombinase in health and disease. Blood Coagul. Fibrinolysis 1, 167–174.
32. Undas, A., Brummel, K., Musial, J., Mann, K.G., and Szczeklik, A. (2001) PI(A2) polymorphism of β3 integrins is associated with enhanced thrombin generation and impaired antithrombotic action of aspirin at the site of microvascular injury. Circulation 104, 2666–2672.
33. Cooke, G.E., Bray, P.F., Hamlington, J.D., Pham, D.M., and Goldschmidt-Clermont, P.J. (1998) PI(A2) polymorphism and efficacy of aspirin. Lancet 351, 1253.
34. Rodgers, R.P. and Levin, J. (1990) A critical reappraisal of the bleeding time. Semin. Thromb. Hemost. 16, 1–20.
35. Cook, J.J., Sitko, G.R., Holohan, M.A., Stranieri, M.T., Glass, J.D., Askew, B.C., McIntyre, C.J., Claremon, D.A., Baldwin, J.J., Hartman, G.D., Gould, R.J., and Lynch, J.J. (1997) Nonpeptide glycoprotein IIb/IIIa inhibitors. 15. Antithrombotic efficacy of L-738,167, a long-acting GPIIb/IIIa antagonist, correlates with inhibition of adenosine diphosphate-induced platelet aggregation but not with bleeding time prolongation. J. Pharmacol. Exp. Ther. 281, 677–689.
36. Bray, P.F., Goldschmidt-Clermont, P.J., Furman, M.I., Michelson, A.D., Barnard, M.R., Mascelli, M.A., Hendrix, C., Coleman, L., Hamlington, J., Kickler, T., Christie, D.J., and Kundu, S. (2001) Platelet glycoprotein IIIa Pl(A) polymorphism and effects of aspirin on thrombin generation. Circulation 103, E33–E34.
37. Diener, H.C., Cunha, L., Forbes, C., Sivenius, J., Smets, P., and Lowenthal, A. (1996) European Stroke Prevention Study 2. Dipyridamole and acetylsalicylic acid in the secondary prevention of stroke. J. Neurol. Sci. 143, 1–13.
38. Hervey, P.S. and Goa, K.L. (1999) Extended-release dipyridamole/aspirin. Drugs 58, 469–475.
39. Rajah, S.M., Crow, M.J., Penny, A.F., Ahmad, R., and Watson, D.A. (1977) The effect of dipyridamole on platelet function: correlation with blood levels in man. Br. J. Clin. Pharmacol. 4, 129–133.
40. Bjornsson, T.D. and Mahony, C. (1983) Clinical pharmacokinetics of dipyridamole. Thromb. Res. 4 (Suppl.), 93–104.
41. Holloper, G., Jantzen, H.M., Vincent, D., Lalanne, M.C., and Anne, M.C. (1990) Aspirin at very ultra low dosage in healthy volunteers: effects on bleeding time, platelet aggregation, and coagulation. Hemostasis 20, 99–105.
42. The CAPRIE Steering Committee. (1996) A randomized, blinded, trial of clopidogrel versus aspirin in patients at risk of ischaemic events (CAPRIE). Lancet 348, 1329–1339.
43. The CURE Investigators. (2001) Effects of clopidogrel in addition to aspirin in patients with acute coronary syndromes without ST-segment elevation. N. Engl. J. Med. 345, 494–502.
44. Mehta, S.R., Yusuf, S., and the CURE Study Investigators. (2000) The Clopidogrel in Unstable angina to prevent Recurrent Events (CURE) trial programme; rationale, design, and baseline characteristics including a meta-analysis of the effects of thienopyridines in vascular disease. Eur. Heart J. 21, 2033–2041.
45. Goldschmidt-Clermont, P.J., Roos, C.M., and Cooke, G.E. (1999) Platelet PI(A2) polymorphism and thromboembolic events: from inherited risk to pharmacogenetics. J. Thromb. Thrombolysis 8, 89–103.
46. Savi, P., Combalt, J., Gaich, C., Rouchon, M.C., Mafrand, J.P., Berger, Y., and Herbert, J.M. (1994) The antiaggregating activity of clopidogrel is due to a metabolic activation by the hepatic cytochrome P450–1A. Thromb. Haemost. 72, 313–317.
47. Kong, D.F., Califf, R.M., Miller, D.P., Moliterno, D.J., White, H.D., Harrington, R.A., Tcheng, J.E., Linehoff, A.M., Hasselblad, V., and Topol, E.J. (1998) Clinical outcomes of therapeutic agents that block the platelet glycoprotein IIb/IIIa integrin in ischemic heart disease. *Circulation* 98, 2829–2835.

48. Curtin, R. and Fitzgerald, D.J. (2000) A cold start for oral glycoprotein IIb/IIIa antagonists. *Eur. Heart J.* 21, 1992–1994.

49. Leffkovits, J., Plow, E.F., and Topol, E.J. (1995) Platelet glycoprotein IIb/IIIa receptor in cardiovascular medicine. *N. Engl. J. Med.* 332, 1553–1559.

50. Rozalski, M., Nowak, A., and Watala, C. (2001) PtA1/A2 polymorphism determines platelet sensitivity to GPIIb/IIIa antagonists [Abstract]. *Thromb. Haemost.*, 86, P2669.

51. Cannon, C.P., Bray, P.F., Ault, K.A., Rizzo, M.J., and Braunwald, E. (1998) PtA2 polymorphism of the platelet IIb/IIIa receptor: increased risk of early recurrent ischaemic events in acute coronary syndromes, increased platelet aggregability, and effective platelet inhibition by an oral IIb/IIIa inhibitor: results from TIMI 12. *Circulation* 98 (Suppl. I), 1–171.

52. Cannon, C.P., McCabe, C.H., Wilcox, R.G., Langer, A., Caspi, A., Berink, P., Lopez-Sendon, J., Toman, J., Charlesworth, A., Anders, R.J., Alexander, J.C., Skene, A., and Braunwald, E. (2000) Oral glycoprotein IIb/IIIa inhibition with orbofiban in patients with unstable coronary syndromes (OPUS-TIMI 16) trial. *Circulation* 102, 149–156.

53. O’Connor, F.F., Shields, D.C., Fitzgerald, A., Cannon, C.P., Braunwald, E., and Fitzgerald, D.J. (2001) Genetic variation in glycoprotein IIb/IIIa (GPIIb/IIIa) as a determinant of the responses to an oral GPIIb/IIIa antagonist in patients with unstable coronary syndromes. *Blood* 98, 3256–3260.

54. Cox, D., Smith, R., Quinn, M., Theroux, P., Crean, P., and Fitzgerald, D.J. (2000) Evidence of platelet activation during treatment with a GPIIb/IIIa antagonist in patients presenting with acute coronary syndromes. *J. Am. Coll. Cardiol.* 36, 1514–1519.

55. Ault, K.A., Cannon, C.P., Mitchell, J., McCahan, J., Tracy, R.P., Novotny, W.F., Reimann, J.D., and Braunwald, E. (1999) Platelet activation in patients after an acute coronary syndrome: results from the TIMI-12 trial. Thrombolysis in myocardial infarction. *J. Am. Coll. Cardiol.* 33, 634–639.

This article should be referenced as follows:

Curtin, R. and Fitzgerald, D.J. (2002) Pharmacogenetics of antiplatelet drugs. *TheScientificWorldJOURNAL* 2, 791–800.

Handling Editor:

Garrett A. FitzGerald, Principal Editor for Thrombosis — a domain of *TheScientificWorldJOURNAL*.

BIOSKETCH

**Dr. Ronan Curtin** is a Cardiology Research Fellow in the Department of Clinical Pharmacology, Royal College of Surgeons in Ireland. He holds MB, BCh, BAO (medical), and MSc (cardiology) degrees. His research interests include monitoring the efficacy of antiplatelet drugs as well as the impact of the PtA2 polymorphism of glycoprotein IIb/IIIa on platelet function and antiplatelet drug response.

**Professor Des Fitzgerald** is Chairman of the Department of Clinical Pharmacology, Royal College of Surgeons in Ireland. He holds an MD and is a Fellow of the European Society of Cardiology. His research interests include platelet adhesion receptors, the regulation of eicosanoid formation, proteomics and genomics.