Magnetic Resonance of Myelin Water: An \textit{in vivo} Marker for Myelin

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Abstract. Myelin is critical for healthy brain function. An accurate \textit{in vivo} measure of myelin content has important implications for understanding brain plasticity and neurodegenerative diseases. Myelin water imaging is a magnetic resonance imaging method which can be used to visualize myelination in the brain and spinal cord \textit{in vivo}. This review presents an overview of myelin water imaging data acquisition and analysis, post-mortem validation work, findings in both animal and human studies and a brief discussion about other MR techniques purported to provide \textit{in vivo} myelin content. Multi-echo $T_2$ relaxation approaches continue to undergo development and whole-brain imaging time now takes less than 10 minutes; the standard analysis method for this type of data acquisition is a non-negative least squares approach. Alternate methods including the multi-flip angle gradient echo mcDESPOT are also being used for myelin water imaging. Histological validation studies in animal and human brain and spinal cord tissue demonstrate high specificity of myelin water imaging for myelin. Potential confounding factors for \textit{in vivo} myelin water fraction measurement include the presence of myelin debris and magnetization exchange processes. Myelin water imaging has successfully been used to study animal models of injury, applied in healthy human controls and can be used to assess damage and injury in conditions such as multiple sclerosis, neuromyelitis optica, phenylketonuria, neurofibromatosis, niemann pick’s disease, stroke and concussion. Other quantitative magnetic resonance approaches that are sensitive to, but not specific for, myelin exist including magnetization transfer, diffusion tensor imaging and $T_1$ weighted imaging.

Keywords: Myelin, myelin water, brain, white matter, demyelination

INTRODUCTION

Myelin is critical for healthy brain function. It plays a fundamental role in determining the speed of action potentials; when myelin integrity is compromised, brain function is affected. Alternatively, when new brain functions are learned or compensatory pathways are formed following neuronal damage, one expects to see increased myelination in the involved nerve tracts. Therefore, an accurate \textit{in vivo} measure of myelin content has important implications for understanding brain plasticity and neurodegenerative diseases.

Magnetic resonance (MR) imaging provides exquisite contrast and detail in images of the brain. While conventional MR is a highly sensitive but notoriously unspecific technique for imaging brain, more advanced MR techniques have the promise to provide us with much more specific information. In particular in recent years many MR techniques have been developed and implemented for imaging...
myelination. In this review we shall focus primarily on one of these techniques - myelin water imaging.

It is an interesting time to review myelin water imaging because, while considerable progress has been made over the last 2 decades, the field is still not mature and fundamental questions remain to be explored. There are several reviews on measuring myelin with MR which we encourage the reader to also peruse, for example [1, 2], and on myelin water imaging [3, 4]. This review will cover the basic principles of the myelin water imaging technique and its validation, applications to humans and animal models. We will also briefly review other magnetic resonance approaches developed for monitoring myelination in vivo. Our goal is to present an up-to-date overview of myelin water imaging that includes a comprehensive summary of how this technique has been applied to study development, healthy controls and disease.

Central Nervous System (CNS) tissue

From the point of view of an MR imaging scientist, CNS tissue contains two fundamentally different kinds of cells (myelinated and unmyelinated) and four different locations for water (intracellular, extracellular, between myelin bilayers and cerebrospinal fluid). While the non-aqueous components of CNS tissue, i.e. lipids, proteins and nucleic acids, have complex dynamic structures, 70–85% of brain mass is simply H2O. Unmyelinated neurons and glial cells have single bilayer membranes while myelinated neurons contain multi-bilayer membranes for which approximately 40% of the mass is compartmentalized water (Fig. 1). Note that this highly simplified description of CNS tissue ignores microstructure geometry and blood, both of which have been studied extensively by MRI and other techniques.

MR of CNS tissue

The most common type of MR is known as ‘proton’ MR which is sensitive to signal from all of the hydrogen atoms or protons in CNS tissue. Practically speaking, the majority of the signal measured by MR of brain emanates from hydrogen in water molecules and the reason for this is as follows. The signals from larger, slowly tumbling, molecules such as lipids and proteins decay rapidly to zero because they possess a very wide distribution of MR frequencies. This broad range of frequencies arises from the dipolar fields that hydrogen’s produce at neighbouring hydrogen sites. However, when molecules undergo isotropic tumbling at rapid rates, the aforementioned broad range of signal frequencies is replaced by a much narrower frequency distribution about the average proton MR frequency. This phenomenon, which is known as motional narrowing, occurs for small molecules in CNS tissue. Water makes up 100.0% of such small molecules in CNS tissue [5]. Conventional MR techniques cannot access the rapidly decaying signal from the larger non-aqueous molecules; hence MR images from CNS tissue arise from water. Also, it has been shown that all of the water in CNS tissue has a MR signal which can be detected by MRI [5, 6].

Conventional MR of CNS tissue

There are three fundamental contrast mechanisms in MR: proton density, T1 relaxation and T2 relaxation. Proton density is proportional to water content [7]. T1 relaxation, often called spin-lattice relaxation, is influenced by water content as well as the presence of other tissue constituents such as iron and myelin, and factors including field strength, temperature and MR magnetization exchange processes [8–11]. T2 relaxation, often called spin-spin relaxation, is also related to water content, the nature of the tissue microstructure, iron, pH and MR magnetization exchange. While the detailed mechanisms of T1 and T2 relaxation in CNS tissue are not well understood, these processes provide qualitative contrast weightings that yield superb in vivo visualization of CNS tissue and have proved invaluable as a diagnostic and patient management tool in medicine.

More quantitative MR measurements in CNS tissue

While conventional MR techniques play a crucial role in medical MRI, more quantitative approaches have focused on measuring specific tissue properties [7]. Highly successful examples include measurements of blood flow [12] and of tissue water diffusion [13, 14]. Another type of MR imaging, which has profound applications in the study of brain function and plasticity, is functional MRI [15], which measures the haemodynamic response, or oxygenated blood flow which occurs when brain regions are activated. A number of MR techniques have been developed for in vivo measurement of myelin content. This review will focus mainly on one of these techniques - myelin water imaging.
MYELIN WATER IMAGING: DATA ACQUISITION AND ANALYSIS

The myelin water fraction

It is commonplace to assume that an MRI voxel (i.e. volume element of an image, typically 1–5 mm$^3$) consists of homogeneous tissue. This is an oversimplification and particularly not true when imaging myelinated CNS tissue. While the entire MR signal is from protons on water molecules, as mentioned earlier, individual water molecules can experience very different microscopic environments. For a homogeneous sample (for example a bottle of pure water or CSF fluid), the $T_2$ decay curve, a plot of MR signal vs time from excitation (TE), gives rise to an exponentially decaying signal with rate constant $1/T_2$. The original myelin water imaging measurements [16, 17] took advantage of the fact that the $T_2$ time of water is determined by its local environment. Myelin water has a $T_2$ time between 10 and 20 ms, while intra-and extra-cellular water has a $T_2$ time longer than 60 ms. If the imaging voxel contains water in several different non-exchanging environments, the resulting $T_2$ decay curve from that voxel is the sum of exponential decays with amplitudes proportional to the relative amounts of water in each environment. In myelin water imaging, the $T_2$ decay curve is separated into its exponential components and expressed as a $T_2$ distribution [18], which is a plot of signal amplitude vs $T_2$ time (Fig. 2). The myelin water fraction (MWF) is generally defined as the ratio of the area in the $T_2$ distribution arising from myelin water (between 10 and 40 ms for humans in vivo) to the area of the entire $T_2$ distribution. MWF can be visually presented as a myelin water image (Fig. 3).

Variants of myelin water imaging

The original in vivo myelin water measurements implemented in the mid 1990s were slow to acquire and produced only a single brain slice in 25 minutes; today it is possible to collect whole brain MWF images in less than 10 minutes. At least four markedly different approaches to myelin water imaging have been explored.

Carr-Purcell-Meiboom-Gill (CPMG) $T_2$

The most common approach, known as the Carr-Purcell-Meiboom-Gill (CPMG) or spin echo method, (e.g. [16, 19, 20] relies on acquisition of the $T_2$ decay curve by application of a $90^\circ$ pulse followed

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Fig. 1. Electron Micrograph of myelinated CNS tissue (Figs. 4–7 in Basic Neurochemistry: Molecular, Cellular and Medical Aspects. 6th edition. Siegel GJ, Agranoff BW, Albers RW, et al., editors. Philadelphia: Lippincott-Raven; 1999. ISBN 0-397-51820-X).
by a train of $180^\circ$ pulses. The CPMG approach typically collects images from 32 echoes with spacing between the echoes of 10 ms; i.e. images at 10 ms, 20 ms, ..., 320 ms. The CPMG approach can be sped up by a factor of 3 by collecting three images at each echo time [20]. At least two groups, [21–23], implemented a more rapid acquisition by collecting brain volumes for a limited number of echo times using a fast imaging approach (spiral acquisitions) rather than collecting images for each echo time. This approach has been shown to collect full brain myelin water imaging results in as little as 4 minutes [24].

As explained above, for these multiple TE time approaches to MWI, the analysis involves extracting the myelin water signal with the shorter $T_2$ time from the total $T_2$ distribution. Traditionally, this has involved non-negative least squares (NNLS) analysis [18] incorporating regularization by minimizing the square of the solution. For data acquired at 3T, transmitter radio frequency ($B_1^+$) inhomogeneity has mandated inclusion of a correction for stimulated echo artifacts due to sub-optimal refocusing pulses [25]. The NNLS analysis makes minimal a priori assumptions about the nature of the $T_2$ distribution. Another approach has been to simplify the analysis by decomposing the $T_2$ decay curves into three log-Gaussian components which are defined to be myelin water, intra/extra cellular water and CSF [26, 27]. While the latter approach may be more robust, problems could arise if the tissue actually contains more than three $T_2$ components. Other approaches to analysis include a multivoxel Bayesian algorithm using spatial prior information which takes into account the expectation that volume fractions and $T_2$ relaxation times of tissue compartments change smoothly within coherent brain regions [28], or a two-Gaussian model to approximate the $T_2$ distribution followed by an expectation-maximization framework with an edge-preserving prior which imposes spatial consistency and smoothness constraints [29]. A mixture of three Wald distributions with unknown mixture weights, mean and shape parameters to represent the distribution of the relative amount of water in between myelin.
sheets, tissue water, and cerebrospinal fluid has also been employed [30]. As mentioned above, models which underestimate, or overestimate, the number of T2 components may give misleading results. A novel post processing method with an efficient multi-slice acquisition scheme, called T2 spectrum analysis uses a weighted regularized NNLS algorithm and nonlocal mean filter (T2SPARC) [31].

Gradient echo decay curve T2*
While T2 data is acquired from an echo train produced by 180° degree refocusing pulses, T2* decay arises from an echo train derived by magnetic field gradient reversals. T2* decay curves are steeper than T2 decay curves because they are susceptible to inhomogeneous magnetic fields and to local differences in magnetic resonance frequency. Several groups have obtained myelin water images from T2* decay curve measurement [32–35]. These myelin water images are substantially improved when the analysis includes individual resonance frequencies for each of the water pools [33, 34] and when signal loss due to inherent magnetic field gradients is recovered [36]. Another approach uses weighted combinations of measured decay signals in a spatially independent neighborhood to combine tissues with similar relaxation parameters. To determine optimal weighting factors, the authors define a spatially independent neighborhood for each pixel and a distance with respect to decay rates that effectively includes pixels with similar decay characteristics, and which therefore have similar relaxation parameters [37].

T1 Acquisitions
It has been found that measurement of T1 relaxation in CNS tissue yields two components with T1’s of 150–200 ms and 1.0 to 1.5 s [38]. Just as for T2, the shorter T1 component has been associated with myelin water. By making an image of only the shorter T1 components in CNS tissue, images which look much like myelin water images have been created [39]. A disadvantage of T1 approaches for measuring myelin water is that they are vulnerable to magnetization exchange effects [40].

mcDESPOT
The mcDESPOT technique [41] is fundamentally different from myelin water imaging techniques which are derived from T2 or T2* decay curves. mcDESPOT makes use of the flip angle dependence of two fast gradient echo imaging sequences (a spoiled gradient sequence and a balanced free precession sequence) to enable extraction the myelin water and intra/extracellular water components in CNS tissue. An advantage of mcDESPOT is that it enables whole brain myelin water fraction imaging with high spatial resolution in a relatively short time.

VALIDATION OF MYELIN WATER IMAGING

Animal studies

Peripheral nervous system
The first observations of a short T2 component in myelinated tissue were reported in frog sciatic nerve in the 1960’s and 70’s [42, 43]. Several decades later the effect of peripheral nerve injury was examined in a number of rat models. In a study of injured rat sciatic nerve, Webb et al. demonstrated that MWF reflected myelin loss and remyelination, due to Wallerian degeneration and regeneration [44]. Using tumor necrosis factor-alpha to induce inflammation with little demyelination and axonal loss in rat sciatic nerve, Stanisz et al. found that multicomponent T2 was the best at distinguishing between inflammation and demyelination [45], while other MR methods (magnetization transfer) were more likely to be influenced by both changes in myelin and pH. Pun et al. and Odrobina et al. examined the effect of demyelination induced by tellurium on the sciatic nerve of rats, with both MR and histology [46, 47]. Tellurium treated animals showed a decrease in healthy myelin in the sciatic nerve and a decrease in the area of the short T2 component; a strong correlation was observed between the degree of myelin staining and the size of the short T2 component (R2 = 0.59). In comparison to other MR methods (T1 relaxation) also used in these studies, the MWF was the best measure of demyelination. Finally, demyelination mediated by dithiocarbamate revealed a decreased contribution from the shortest T2 component in multicomponent T2 spectra, consistent with demyelination [48].

Central nervous system
Myelin water in CNS white matter was first observed in a cat model in 1991[49]. Shortly there afterwards, guinea pigs with experimental autoimmune encephalomyelitis (EAE) demonstrated reduced MWF consistent with histologically con-
firmed myelin loss in both the brain and spinal cord [6, 50]. More recently, a strong correlation between MWF in rat spinal cord and histological staining for myelin was observed [51], and a remyelinating mouse model found that MWF increased with remyelination [52]. Longitudinal assessment of MWF in vivo correlates with myelin staining to accurately characterize white matter damage in a dorsal column transection model of rat spinal cord injury [53].

Human studies

Several studies using human tissue have examined the relationship between MWF and histological staining for myelin. Evidence to date suggests that there is minimal change of the myelin water signal post mortem, both shortly after death in-situ and upon tissue fixation with formalin [54], making such correlative studies possible. The T2 distribution of formalin fixed CNS tissue is qualitatively similar in shape to that from in vivo, although the T2's are shifted to shorter times. A good qualitative correspondence was first observed between MWF in formalin fixed brain and the anatomical distribution of myelin by Moore et al. [55] as indicated by the myelin phospholipid stain Luxol Fast Blue (LFB) [56–59]. Follow-up brain sample studies demonstrated an excellent quantitative correlation between MWF and LFB optical density at both 1.5T and 7T (Fig. 4) [54, 60]. Similarly, comparisons between MWF and myelin staining in human spinal cord also show excellent correspondence between the MR and histology markers for myelin [61].

**MYELIN WATER IMAGING IN HUMANS**

**Development**

Myelination in humans begins in the fifth fetal month. Rapid myelin growth occurs during the first two years and continues throughout life until the 6th decade [62, 63]. The onset of myelination is marked by a rapid increase in brain lipid and protein content and a corresponding decrease in brain water content [64]. Conventional T1 and T2 weighted imaging show changes in contrast in both white and grey matter due to these rapid changes in brain tissue and water content [65]. However, conventional imaging is not specific to myelination.

Myelin water imaging can provide more quantitative information about myelin content in the developing brain. Applying this method in pre-term infants is feasible, and demonstrates variation in brain MWF for different brain structures [66, 67]. Using the mcDESPOT technique, several studies have reported the pattern of normal myelin development from 2.5 months to 5.5 years of age (Fig. 5) [68–71]. These studies demonstrate a spatial temporal pattern of myelination beginning in the cerebellum, pons, and internal capsule; proceeding caudocranially from the splenium of the corpus callosum and optic radiations (3–4 months) to the occipital and parietal lobes (4–6 months) and then to the genu of the corpus callosum and frontal and temporal lobes (6 – 8 months). This pattern of myelination is consistent with reports of myelination beginning in posterior regions before anterior regions of the brain [72]. Distinct male and female developmental patterns were observed, and
significant relationships between myelin content and measures of cognitive function were demonstrated [70]. Examining slightly older children with a multi-echo approach, Whitaker et al. [73] found that the number of highly myelinated voxels in the corpus callosum was strongly correlated with verbal IQ tests in a study of preadolescent children (9–12 yrs). However, the number of highly myelinated voxels did not correlate with age. Based on myelination trajectory observations in healthy infants, toddlers and young children, Deoni et al. developed a model that may be useful for identifying developmental abnormalities [69].

One factor which may influence development is breastfeeding. The prevailing consensus suggests that breastfeeding is associated with higher IQ and improved cognitive functioning. Using mcDEPOSIT myelin water imaging, it was demonstrated that breastfed children exhibited increased white matter development in later maturing frontal and association brain regions [74]. Positive relationships between white matter microstructure and breastfeeding duration were also observed in several brain regions anatomically consistent with improvements in cognitive and behavioural performance measures [74]. A commentary on this study raises the possibility that the premature introduction of cow’s milk in the second year of life may also negatively influence cognitive development [75].

A second factor which may be linked to cognitive performance is the presence of the apolipoprotein E (APOE) ε4 allele, a major susceptibility gene for late-onset Alzheimer’s disease. The presence of this gene may influence brain development already in infancy, as demonstrated by a recent myelin water MRI study where it was found that infant ε4 carriers had a lower myelin water fraction than noncarriers in areas of the brain preferentially affected in Alzheimer’s disease [76].

**Healthy adults**

**Brain**

In healthy adults, white matter has substantially larger MWF than grey matter. Myelin water in the brain varies between different white matter areas by more than a factor of 2 [17, 22, 77–80]. Reproducibility and reliability of MWF in healthy controls, both at a single site and multiple sites is very good [81–84]. Regional MWF differences are believed to reflect differing regional myelin concentrations (Fig. 3). One study found that MWF in males was significantly greater than females for the rostral body ($p < 0.05$) and posterior midbody ($p < 0.005$) while splenium MWF in males was significantly less than that in females ($p < 0.05$) [85]. Frontal lobe MWF is correlated with age, as well as years of education, in normal adults (Fig. 6) [86], and a second study by the same authors found significant positive relationships between regional MWF and age, North American Adult Reading Test (NAART) IQ, and years of completed education in healthy volunteers [87]. Recent
Fig. 6. Plots of frontal myelin water fraction in relation to age and education in healthy comparison subjects (control) and patients with schizophrenia. While statistically significant relations between frontal myelin water fraction and both age ($r = 0.47$, $p = 0.01$) and education ($r = 0.51$, $p = 0.006$) were observed in healthy subjects, no statistically significant relations were seen in patients with schizophrenia (Fig. 4, Flynn SW, Lang DJ, Mackay AL, Goghari V, Vavasour IM, Whittall KE et al. Abnormalities of myelination in schizophrenia detected in vivo with MRI, and post-mortem with analysis of oligodendrocyte proteins. Mol Psychiatry. 2003;8(9):811-20).

work using mcDESPOT found a positive correlation between activity level and MWF in the right parahippocampal cingula [88].

**Spinal Cord**

Myelin water techniques applied in the brain can also be used to study spinal cord myelination. However, imaging the spinal cord is difficult for a number of reasons including magnetic field inhomogeneities, the presence of flow from CSF and the small diameter of the cord. Nevertheless, several $T_2$ studies have shown the feasibility of measuring MWF in spinal cord in vivo. MWF has been found to be approximately 50% higher in spinal cord than normal brain white matter and is reported to vary along the length of the cord [19, 89–92]. Younger adults (20–30 years old) are reported to have a lower cervical cord MWF compared to older (50–75 yr) study participants [91]. An indepth review of myelin water in the cord can be found elsewhere [93].

**Multiple sclerosis**

Multiple sclerosis (MS) is an autoimmune disease of the CNS that is characterized by edema, inflammation, demyelination and axonal loss [94]. Given that myelin damage is a key component of MS pathogenesis, much of the pioneering work in myelin water imaging began in the field of MS.

**Lesions**, or focal plaques, which appear bright on proton density and $T_2$ weighted imaging were the initial focus for imaging myelin in MS (Fig. 7).
MWF has been shown to be variably decreased in MS lesions [16, 21, 77, 78, 84, 95–99]. On average, lesions show \( \sim 50\% \) less MWF than normal appearing white matter (NAWM) [77], however, myelin loss in MS lesions is extremely heterogeneous with some lesions having no myelin, and others having normal, or near normal, white matter levels [77, 95, 99]. Lesion variability likely reflects different lesion pathologies, as MWF does differ between T2 weighted lesions, contrast enhancing lesions and T1 black holes [99, 100]. Using discriminant function analysis, Vavasour et al. found that myelin water was most important in separating lesions based on their age, clearly delineating lesions less than one year old from lesions older than one year. This suggests there is a time dependence to demyelination in new lesions; on average, lesions less than a year old had only slightly reduced myelin water, whereas lesions at least one year old had a larger reduction in myelin water [100]. However, whether lesions less than a year old have a higher myelin water because of an absence of demyelination or due to the presence of a cycle of demyelination and remyelination is not clear.

Longitudinal assessment of MWF in MS can provide insight into the processes of demyelination and remyelination. Several studies have demonstrated reductions and subsequent increases in MWF as lesions develop and resolve over the time scale of months [84, 101, 102]. A serial study examining larger lesions showed low myelin water in the core of the lesion upon first appearance, which suggests demyelination may be present in the early stages of lesion development [101]. Several months later, one lesion showed a reduction in low MWF volume, reflecting remyelination, whereas the volume of low myelin water in the second lesion remained low and constant, suggestive of a failure to repair.

Normal appearing white matter (NAWM) are areas of brain and spinal cord which appear ‘normal’ on standard clinical imaging. Much effort has also been put into studying NAWM in MS, which is known from post-mortem histological studies to demonstrate myelin abnormalities demyelination [103–107]. MWF of NAWM was found to be diffusely reduced in both brain (by 6–37\%) [21, 77, 96, 99, 102] and spinal cord (by 11–25\%) [19, 90] when compared to healthy controls. NAWM MWF can distinguish between different subtypes of MS [108] and reduced brain MWF is also related to increased disability [96, 108], suggested that more global disease process can influence clinical status.

Longitudinal changes in MWF can also be measured in NAWM, for example examining the cervical cord in primary progressive multiple sclerosis found a 10\% decrease in MWF over 2 years, suggesting ongoing demyelination may be contributing to the disease process in this subgroup of patients [90]. Of course, such diffuse myelin loss could be due to concomitant...
loss of axons in Wallerian degeneration as reported in NAWM [109–111].

**Diffusely abnormal white matter (DAWM)** is a third subtype of white matter observed in MS brain [112]. DAWM is typically periventricular and consists of regions of mild hyperintensity with a poorly defined boundary on proton density and T2 weighted imaging. Since DAWM is only now becoming more recognized, it is possible some of the previous studies on NAWM may have included DAWM in their assessments. MWF is reduced in areas of DAWM by approximately 23% *in vivo* and 30% in post mortem tissue relative to adjacent NAWM, and appears to be a very sensitive indicator of pathology [113, 114]. Post mortem work correlating MWF with histological staining for different components of the myelin bilayer finds a selective loss of myelin lipids with relative preservation of myelin proteins [115].

Because MS is a characterized by a variety of pathological processes, it is important to consider other possible sources for changes in MR markers claiming to be specific for myelin. Changes in water content (WC) arising from inflammation or edema can influence MWF as this measure is a ratio of the myelin water signal to the total signal from all water, hence decreases in MWF could arise from increases in WC. However, modeling shows that the observed increases in WC in MS of about 2% should only result in about a 2% decrease in MWF, while the observed MWF decrease is about 15%. If the MWF decreases observed in lesions and NAWM were to have occurred purely due to increases in WC, the volume increases would have to have been larger than 15% making this scenario unrealistic [77]. Nevertheless, it is important to keep in mind that small changes in MWF can certainly, at least in part, arise from an increase in water content.

**Neuromyelitis Optica (NMO)**

Neuromyelitis Optica is a rare relapsing autoimmune disorder that preferentially causes damage to the optic nerve and spinal. Several studies have examined differences in MWF between MS and NMO. One report indicates that myelin water fraction in MS lesions is more greatly reduced [98], thereby suggesting that myelin loss is more severe in MS than in NMO. In contrast, in a combined transcranial magnetic stimulation / myelin water fraction study, the degree of damage was found to be greater in NMO than MS: NMO showed larger reductions in corticospinal tract recruitment curves and lower MWF than MS and controls [116]. The differing results between the Jeong and Manogaran studies may reflect differences in patient populations, especially considering that MS is such a heterogeneous disease.

**Schizophrenia and first episode psychosis**

Schizophrenia is characterised by disorganized thinking, hallucinations, delusions, changes in behaviour and changes in emotions. Schizophrenia often develops during the late teens or early 20’s, during the time of myelination in frontal brain regions. The cause of schizophrenia is unknown, but there is growing evidence of neuronal and oligodendrocyte-related abnormalities, including myelin and fatty-acid biosynthesis dysfunction based on post-mortem brain studies [117], abnormal expression of myelin-related genes [118] and damage to the myelin sheath due to the formation of concentric lamellar bodies and decreased volume density of mitochondria in oligodendrocytes [119]. Myelin water imaging studies support evidence of myelin-related abnormalities in the white matter of people with schizophrenia. Flynn et al. found a 12% decrease in global white matter MWF and a 36% reduction in left genu MWF in schizophrenic subjects when compared to controls [86]. Furthermore, the relationship between white matter MWF with both age and years of education observed in healthy controls is absent in schizophrenia (Fig. 6) [86]. A second, more recent study, found altered patterns of association between age, years of education, and MWF in people with first episodes of psychosis, suggesting that subtle disturbances in myelination may be present early in the course of their disease [87].

**Phenylketonuria**

Phenylketonuria (PKU) is a metabolic disorder whereby people lack the ability to metabolize the amino acid phenylalanine (PHE). Most affected individuals who are not treated with a diet restricted in PHE develop severe mental retardation. Several studies have described myelin abnormalities in the brain and spinal cord of people with PKU [120–124]. Using myelin water imaging, Sirrs et al. found the MWF to be reduced in PKU by up to 56% in normal appearing white matter; a reduction in MWF was also reported in diffuse white matter lesions [125].
Fig. 8. Group differences in myelin water fraction (MWF) between healthy controls and participants with chronic stroke. MWF was significantly reduced in whole-cerebrum normal appearing white matter (NAWM) in the stroke group compared to whole-cerebrum NAWM in the healthy group (A), in the ipsilesional posterior limb of internal capsule (PLIC) and non-dominant PLIC MWF in the healthy group (B) and in the contralesional PLIC for the stroke group versus dominant PLIC in the healthy group (C) (*p < 0.05). Error bars represent standard deviation (Fig. 5, Borich MR, Mackay AL, Vavasour IM, Rauscher A, Boyd LA. Evaluation of white matter myelin water fraction in chronic stroke. Neuroimage Clin. 2013;2:569-80).

**Autism**

Autism continues to increase in prevalence and some evidence suggests abnormal white matter anatomy and impaired brain connectivity may play an important role in this disease’s symptomology. To study the role of myelin in autism Deoni et al. employed the mcDESPOT method to compare 14 individuals with autism and 14 age- and IQ-matched controls [126]. Individuals with autism showed widespread MWF differences compared to controls. In particular, worse current social interaction skills were related to reduced MWF.

**Stroke**

At least one study has examined myelin water in ischemic stroke [83]. Borich et al. examined 20 individuals with ischemic stroke and 12 matched healthy controls. Reduced MWF was observed in whole-cerebrum white matter (p < 0.001) and in the ipsilesional (p = 0.017) and contralesional (p = 0.037) posterior limb of internal capsule (PLIC) after stroke compared to whole-cerebrum and bilateral PLIC MWF in healthy controls (Fig. 8). These observations provide novel insight into myelin specific tissue changes that occur in stroke, and may have important consequences for understanding neuropathology of stroke.

**Neurofibromatosis**

Unidentified bright objects (UBOs) are often observed in neurofibromatosis-1 (NF1), a genetic disease which causes tumors along the nervous system. The histopathological basis of these UBOs is not clear. Billiet et al. examined 17 patients with NF1 and found no significant alterations in MWF (Fig. 9), which suggests demyelination is unlikely to be present in UBOs which appear to be mainly caused by a shift towards a higher T2-value of the intra- and extracellular water pool [127]. The authors suggest the appearance of UBOs may arise from altered microstructural compartmentalization, and an increase in ‘extracellular-like’, intracellular water, possibly due to intramyelinic edema.

**Niemann-Pick disease**

Niemann-Pick disease is a rare progressive genetic disorder characterized by an inability of the body to transport cholesterol and other fatty substances (lipids) inside of cells. This leads to the abnormal accumulation of these substances within various tissues of the body, including white matter. In a small case study, Davies-Thompson et al. used myelin water imaging to examine demyelination in 2 patients with Niemann-Pick disease type C [128]. The results suggest that this technique may be useful for identifying regional changes in myelination in this condition.
Primary lateral sclerosis (PLS) and Amyotrophic lateral sclerosis (ALS)

Both PLS and ALS are progressive demyelinating diseases which target the motor neurons of the CNS. Diagnosis of PLS and ALS is clinical, and it may be difficult to distinguish the two diseases at the early stages. Kolind et al. studied 23 ALS, 7 PLS and 12 controls using mcDESPOT and found that PLS patients were distinguished by widespread cerebral MWF reductions, independent of disease duration and clinical upper motor neuron burden, while ALS patients showed a significant increase in intra- and extra-cellular water, indirectly linked to neuroinflammatory activity [129]. Furthermore, cognitive impairment in the ALS group was associated with myelin changes within the anterior corpus callosum and frontal lobe projections, and longitudinal changes in MWF were only significant in the PLS group. Given their observations, the authors conclude that myelin water imaging may have the potential to distinguish PLS from ALS, and may have value as a marker of extramotor involvement.

Concussion

Traumatic brain injury and concussion are a major public health concern but continue to be poorly understood. Evidence for myelin damage in mild traumatic brain injury (TBI) comes from one prospective study that examined two varsity hockey teams (45 players) over one season of athletic competition [130]. 11 players sustained a concussion, and were scanned at 72 hours, 2 weeks, and 2 months post-injury. At 2 weeks post-injury, MWF was reduced in several brain areas relative to preseason; values recovered to pre-season values by 2 months post-injury (Figs. 10, 11). These results could indicate a transient myelin disruption following a single mTBI, with subsequent remyelination of affected neurons.

LIMITATIONS OF MYELIN WATER IMAGING

While MWF has shown strong qualitative and quantitative correspondence with histological markers for myelin (as described earlier), it is important to be aware of several potential confounding factors that may influence in vivo measurement of myelin water.

Exchange

T₂ decay curve approaches to measurement of myelin water fraction assume that water molecules stay in the myelin bilayers for times long compared to the decay curve measurement time. Evidence from studies of rodent spine [131, 132] suggested that movement of water from myelin during the measurement can occur at rates sufficiently fast to cause the measured myelin water fraction to be artificially low, an observation which has also been characterized using a 4 pool model of white matter [133]. However, other animal studies involving bovine brain and optic nerve suggest exchange does not play a large role in MWF measurements [134, 135]. The effect of exchange in human brain has not yet been accurately characterized; however, it seems likely that measured myelin water fractions are slight underestimates of the true myelin water fraction [136].

Magnetization exchange with non-aqueous tissue

It has been demonstrated that when measuring T₁ in CNS tissue, magnetization exchange between non-aqueous tissue and water in CNS tissue can give rise to relatively short T₁ components [40, 137–139]. It may
be difficult to separate these components from myelin water T₁ components, which are themselves altered by exchange. Hence exchange between the multiple components of CNS tissue makes it challenging to extract an accurate myelin water fraction from T₁ measurements.

**Myelin debris**

Damage to myelin, whether from an acute event (i.e. trauma) or chronic condition (i.e. multiple sclerosis) will result in myelin debris. Eventually macrophages will clear debris from the injury site, but until that occurs the presence of myelin debris may influence the measured myelin water fraction. Several animal studies have investigated how myelin debris impacts MWF. Using a rat sciatic nerve cut/crush injury model it was shown that myelin water was unable to distinguish between intact myelin, degenerating myelin and myelin debris [44]. In a longitudinal study of rat spinal cord injury via dorsal column transection, MWF increased 3 weeks post injury due to reduced extracellular space filled with myelin debris [140]. Finally, a lyssolecithin induced injury in mouse spinal cord found that demyelination shown by myelin water lagged behind the histological evidence of demyelination by 1 week [52]. Although no such work has been completed in human tissue, given these observations it is possible that presence of myelin debris would impact MWF measurement of recent demyelination in humans in vivo.

**mcDESPOT**

The mcDESPOT approach for measurement of MWF offers many advantages over the decay curve approaches, including accommodation for exchange effects; however, it also brings several challenges. The analysis involves a two component model (plus CSF) which may not be applicable to all situations in CNS tissue, in particular pathology which frequently contains pools of water (e.g. edema) with intermediate T₂ times [141, 142]. The analysis also requires estimation of a large number of parameters from a limited amount of relatively noisy data points. Consequently, the accuracy of mcDESPOT derived myelin water fraction is presently an active area of research [143–148]. Finally, it has been clearly demonstrated that the mcDESPOT curves are affected appreciably by magnetization transfer [149, 150]; original analy-
OTHER MR TECHNIQUES SENSITIVE TO MYELINATION

A large literature exists on the imaging of myelin using magnetic resonance. It is possible to subdivide this literature into three areas: 1) imaging myelin in subjects under 2 years old, 2) imaging myelin in white matter in subjects more than 2 years old and 3) imaging myelin in the cortex. Perhaps surprisingly, these three applications have very little overlap in the literature.

Imaging myelin in subjects under 2 years old

Because the microstructure of human brain changes so extensively over the first two years, it has, so far, been impossible to define a specific MR technique for measuring myelin content in this subject group. In the context of radiological imaging, it has been noted that 1) myelin causes signal increase on T1 weighting [65], 2) myelin causes signal decrease on T2 weighting [65] and 3) myelin causes an increase on diffusion anisotropy [151]. However the extent of these effects change with the age of the child [151]. Therefore, it may be quite difficult to quantitatively characterize plasticity in childhood brain. A number of studies are in the literature that use the mcDESPOT approach to myelin water imaging, however, interpretation of findings from these studies should also keep in mind the potential confounding factor of the large water content changes that occur in the brain from 0–2 years.

Imaging myelin in white matter in subjects more than 2 years old

In adults it is much easier to image myelin in white matter than in grey matter because the concentration of myelin is much higher in white matter. The largest literature on myelin imaging comes from the white matter field and the largest application has been the characterization of neuronal degeneration in diseases like MS.

Magnetization transfer (MT)

MT techniques measure decreases in MR signal following off resonance excitations. The premise of the MT approach is that (1) compared to hydrogens attached to water, hydrogens attached to non-aqueous molecules (i.e. all lipids and proteins in myelin and glial cells) have a much broader range of MR frequencies and that (2) hydrogens within myelin preferentially exchange magnetization with water molecules [134, 152]. There is an extensive literature on the use of MT to characterize myelination in MS [2]. Data collection and analysis is straightforward and readily available on MR scanners and the method is sensitive to small differences between groups. Several studies [153, 154] have showed correlation between MT parameters and histological measures of myelin. However, not all studies support the use of MT as a surrogate for myelin [52, 155]. A distinct advantage of MT techniques is their relative robustness compared to many other putative myelin sensitive techniques. A disadvantage of the use of MT to monitor myelin is that while a change in myelin will cause a change in MT, a change in MT is not necessarily due to a change in myelin; for example, changes in water content which could be caused by inflammation or edema will result in changes in MT [156]. Examining the magnetization transfer ratio (MTR) [157] and T2 relaxation measures in an EAE guinea pig model, Gareau et al. showed both measures were reduced in the NAWM. However, MTR and myelin water appeared to be influenced by different aspects of EAE, as modulating the inflammation strongly influenced MTR but had minimal effect on myelin water. This result suggests that the short T2 component is specific for myelin while pathological features other than myelin content may also be important in the interpretation of MTR [158]. McCreaey et al. examined a murine model of spinal cord damage induced by lyssolecithin whereby demyelination is followed by spontaneous remyelination and found that MWF increased with remyelination while MTR did not [52].

An exciting new approach to MT and myelin is the inhomogeneous MT technique [159, 160]. Initial work on this technique suggests that it is preferentially sensitive to the signal from hydrogens on lipids in CNS tissue. Given that myelin is 70–80% lipid, more research is warranted on this variant of MT.

Diffusion

Diffusion MR takes advantage of the fact that the anisotropic nature of cellular microstructure influences the thermally induced random motions of water molecules in brain and spinal cord. Particularly in
white matter, tissue water travels preferentially along the local axonal orientations. In diffusion tractography, local diffusion directions are combined to produce maps of nerve tracts [13, 14]. In diffusion tensor imaging (DTI), measures of diffusion and anisotropy of diffusion are acquired at each imaging voxel. DTI has been used to show evidence on plasticity in both preclinical [161, 162] and human learning tasks [163]. While there is a growing literature on MR diffusion techniques and plasticity, caution is warranted and one should be careful about interpreting diffusion anisotropy changes as myelin changes [164]. The perpendicular component of the diffusion tensor (often called radial diffusivity or lambda perp) has been demonstrated to be inversely related to myelination in animal models [165, 166]. This latter approach to measuring myelination is complicated by the fact that simplistic DTI measures of the perpendicular component of the diffusion tensor are confounded by water content changes [166].

**Ultrashort Echo Time (UTE)**

Water protons in brain have T2s greater than 10 ms and can be easily imaged by conventional MR techniques with conventional echo times. However, CNS tissue also contains non-aqueous sources of protons, including, but not limited to, the lipids and proteins that make up myelin. The T2 of non-aqueous protons is very short (≈50 μs) [167] and cannot be directly measured with conventional MRI. To access the dipolar broadened, non-aqueous signal from CNS tissue, ultrashort TE (UTE) techniques must be used [168]. Visualization of UTE signal is improved by using long-T2 suppression RF pulses that improve the contrast of short-T species by removal of contaminating signal from tissue with longer T2 values [169]. A number of studies have used these methods to study brain tissue [170–174]. Two of the studies which used UTE for myelin measurement [167, 171] employed smaller bore magnets and short excitation pulses which yielded signals from non-aqueous brain with T2 times which were in agreement with previous NMR spectrometer measurements of brain samples [135]. In vivo, white matter demonstrates short-T2 components [170, 172, 173] and reduced signal in areas of pathology [170, 174]. However, in vivo UTE characterizations of brain using whole body MR systems have reported unrealistically long myelin T2 times, suggesting that it may be very difficult to use UTE techniques on whole body MRs to measure myelin content in humans in vivo. UTE methods are available on most newer MR systems [173, 174] and it is expected this area of research will continue to expand.

**Imaging myelin in the cortex**

Myelin concentrations are substantially less in the cortex than in white matter; hence it is much more difficult, and perhaps even impossible, to accurately
measure cortical myelin content. Thus far cortical myelin measurement has been largely qualitative. Several groups [9, 175] have made the assumption that changes in T<sub>1</sub> relaxation time are proportional to changes in myelin. Then, a T<sub>1</sub> map is presumed to be a qualitative cortical myelin map. A problem with these T<sub>1</sub> approaches is that in CNS tissue T<sub>1</sub> is also correlated with water content and with iron content [9, 10]. Yet another qualitative approach to measuring myelin content in brain is to assume that the ratio of T<sub>1</sub> weighted signal intensity to T<sub>2</sub> weighted signal intensity is related to cortical myelin content [176, 177]. Remarkably, these two approaches result in images which look like what one might expect for a myelin image in cortex. However, the lack of a solid theoretical background for these contrasts as myelin measures and the fact that, the assumption of myelin specificity is likely to break down in abnormal brain, make it unlikely that these approaches could be generally specific for myelin. Unfortunately, due to the low concentrations of myelin in grey matter, more quantitative myelin measurement techniques fail; hence such qualitative approaches might be the only means available to monitor myelination in grey matter.

CONCLUDING REMARKS

Imaging myelin in vivo is a challenging research area and, as described above, many approaches are being explored. Based upon theoretical specificity and strong correspondence with histology, this review makes the case for myelin water imaging as the technique of choice for monitoring myelination in white matter. It is an exciting time to be in this field, since it is clear that obtaining a better understanding of brain plasticity through myelination changes will have profound influences on medicine and neuroscience.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

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