Research Paper

Association between Sumoylation-Related Gene rs77447679 Polymorphism and Risk of Gastric Cancer (GC) in a Chinese Population

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Abstract

Purpose: Sumoylation plays a critical role in gene regulation and tumorigenesis, and is hypothesized to correlate with the development of various cancers. So far, there has been no reported association between sumoylation-related genes and the risk of gastric cancer (GC).

Methods: A total of 17 tagging single-nucleotide polymorphisms (tag-SNPs) in 5 sumoylation-related genes were selected and genotyped by SNaPshot in a case-control study, including 1021 GC patients and 1304 controls. Odds ratio (OR) and 95% confidential interval (CI) were computed to evaluate the genetic association of the onset of GC.

Results: We demonstrated that CBX4 rs77447679 polymorphism was significantly associated with GC risk ($P = 0.017$; adjusted OR: 1.71; 95% CI: 1.10-2.66). The patients with CC genotype had a lower risk of GC (CC vs. CA+AA, $P = 0.017$; adjusted OR: 1.24; 95% CI: 1.04-1.49).

Conclusion: This study revealed that CBX4 rs77447679 polymorphism was positively associated with GC, and individuals with CC genotype had less risk of GC. The risky effects and functional effect of this polymorphism in GC require further investigation.

Key words: sumoylation, haploview, polymorphism, gastric cancer, susceptibility.

Introduction

Gastric cancer (GC) is the fourth most common cancer diagnosed in men and the fifth in women globally [1]. Multiple pathogenic factors, such as unhealthy lifestyle, and Helicobacter pylori infection, contribute to the etiology of GC [2]. Consistent with multifactorial pathogenesis, clinical outcomes and GC prevalence might result from environmental factors. Besides, the influence of host-specific factors, especially host genetic variation, could impose a remarkable effect on the susceptibility of GC in a population with high incidence rate [3].

Post-translational modifications (PTMs) include enzymatic changes such as addition of chemical group adducts, including several ubiquitin, small ubiquitin-like modifiers (SUMO) and phosphor. These changes play an essential role in gene regulation, cellular function, tissue development, and metabolism, and they are closely relevant to cancer occurrence [4, 5]. Understanding the influence of
sumoylation on gene regulation and protein homeostasis could provide valuable insights of early diagnosis and treatment for cancer [6]. Sumoylation refers to the post-translational process that a SUMO protein moiety is enzymatically conjugated to a substrate protein [7, 8]. Sumoylation requires the following series of enzymatic events: A. SUMO-activation by enzyme E1, including SUMO-activation enzyme E1 (SAE1) and ubiquitin-activating enzyme 2 (UBA2); B. SUMO-conjugation by enzyme E2, such as UBC9 and ubiquitin-conjugating enzyme 2I (UBE2I); C. SUMO-ligation by enzyme E3, such as ran-binding protein 2 (RANBP2) and polycomb chromobox homolog 4 (CBX4) [4, 9-12]. The sumoylation pathway, one of the most important PTMs, was closely related with pathogenesis of human diseases through regulation of genome stability, gene expression, nuclear functions, and protein-protein interactions [13-15]. Besides, sumoylation is also relevant in various cancers, including prostate cancer, bladder cancer, brain cancer, breast cancer, and cervical carcinoma, through the mediation of some key oncogenes and tumor suppressors [9, 16-19]. Recently, some current GC-association studies for sumoylation have existed [20, 21], but knowledge of the detailed mechanism of GC carcinogenesis is still unclear. So far, there is no study focusing on the influence of sumoylation-related genes on the risk of GC.

This study focuses on the single nucleotide polymorphisms (SNPs) located in the functional region of sumoylation-related genes, and we aim to provide a new perspective on the mechanism of GC. We performed a case-control study of tag-SNPs in 5 potential sumoylation-related genes (RANBP2, SAE1, UBE2I, UBA2 and CBX4) in a Chinese population, and CBX4 rs77447679 has been identified significantly associated with GC.

Methods and Materials

Study participants
A total of 1021 histopathologically confirmed GC patients and 1304 GC-free participants without a family history of GC (as control group) were enrolled in this study. All participants were genetically unrelated Han Chinese. These GC patients had been referred to the Department of Oncology in the Second Affiliated Hospital of Nanjing Medical University, Shanghai Tenth People's Hospital, and Tongji University School of Medicine in China, between 2011 and 2014. The disease-free controls were enrolled from those sought routinely physical examination in the outpatient department of these hospitals. A standardized questionnaire was developed and used to obtain basic demographic information and clinical characteristics of the participants. This study was approved by each of the participating institutes' ethics committee, in accordance with the principles of the Helsinki Declaration (Approval Number: SHSY-IEC-pap-16-15), and written informed consent was obtained from all participants in this study (both patients and control).

SNP selection
The potential SNPs were screened and selected from unrelated Han Chinese genotype data from the 1000 Genome Project database. Identification of the tag-SNPs was performed in the Haploview 4.2 software (BROAD institute, Cambridge, MA, UK) with pairwise option, using \( r^2 = 0.8 \) as the threshold. Besides, we also scanned these tag-SNPs' corresponding minor allele frequencies (MAFs) in the NCBI SNP database, and excluded those without reports in Chinese population or MAF < 0.05. The identification number of the final candidate tag-SNPs, their relative positions in the genome, and the linkage disequilibrium (LD) of these tag-SNPs are shown in Fig 1.

DNA extraction and genotyping
Approximately 10 ml whole blood from each participant was extracted and transferred into an ACD tube. Genomic DNA was extracted from peripheral blood samples by protease K (Roche Applied Science, IN, USA) digestion and phenol-chloroform extraction. DNA extraction was performed in the laminar flow hood using sterile techniques. Genotypes of the 33 tag-SNPs were determined by SNaPshot (Applied Biosystems, CA, USA). The primers to amplify different fragments, including each tag-SNP, were designed with the Primer 5 software (PREMIER Biosoft, Palo Alto, CA, USA). Amplifications were performed in the ABI3130 genetic analyzer (Applied Biosystems, CA, USA), and allelic discrimination was completed using the GeneMapper™ software (Applied Biosystems, CA, USA), using manufacturers’ instruction and protocols. Positive controls through sequencing and negative controls without DNA samples were contained in each plate to ensure the accuracy of genotyping. The genotyping procedure in this study was conducted in a double-blinded way. Approximately 10% of the total samples were randomly selected for duplicated assays, and the results were 100% concordant.

http://www.jcancer.org
Figure 1. Hapmaps of sumoylation-related genes: A. Hapmap of RANBP2; B. Hapmap of UBE2I; C. Hapmap of CBX4; D. Hapmap of UBA2 and E. Hapmap of SAE.
Statistical analysis

χ² test and Student’s t-test were performed to quantify the differences of demographic characteristics, selected variables, genotype frequencies and allele distributions between the case (GC-patient) and control groups. Association between genetic variation and the risk of GC was estimated by odds ratio (OR) and corresponding 95% confidence intervals (CI) from an unconditional logistic regression, adjusted for age and sex. Hardy-Weinberg equilibrium (HWE) for each SNP in the case and control groups were checked using χ² test. P < 0.05 was considered as statistically significant in this study. Statistical analyses were performed in SPSS 23.0 software (SPSS, Inc., IL, USA).

Results

Characteristics of the study cases and controls

1021 patients (case group) with histopathologically confirmed GC (806 patients, comprising 79% of all cases were non-cardia, and the remaining 21% were cardia) and 1304 healthy participants (control group) were enrolled in this study. There was no significant difference of age or sex between the case and control groups. 470 patients (46%) had clinical stage I/II GC, and the other 551 (54%) were in stage III/IV GC. The demographic information and clinical characteristics of the participants are shown in Table 1.

Association between SUMO gene polymorphisms and the risk of GC

19 potential SNPs were selected from RANBP2, UBE21, CBX4, UBA2 and SAE1 genes as the candidate sumoylation-related tag-SNPs. The characteristics of these 19 tag-SNPs, as well as the allele frequencies of the polymorphisms among the case and control groups are reported in Table 2. After SNP genotyping, 5 SNPs (UBE21 rs8062881, rs75998799 and rs12925270, CBX4 rs190846371, and SAE1 rs6509314) were excluded because of their low MAF (P < 0.05). The observed genotype frequencies of the SUMO genes were consistent with the HWE. Of the remaining 14 SNPs, there was significant difference between the case and control group for rs77447679 in CBX4 gene, and consequently strong association between SNP rs77447679 and the risk of GC (P = 0.017; adjusted OR: 1.24; 95% CI: 1.07-1.44).

Table 1. Demographic and clinical characteristics of GC patient (cases) and control groups.

| Variables            | Cases N = 1021 | Control N = 1304 | P    |
|----------------------|---------------|------------------|------|
| Tumor site           |               |                  |      |
| Non-cardia           | 806 (0.79)    | 1021 (0.80)      |      |
| Cardia               | 214 (0.21)    | 283 (0.20)       |      |
|          |  |                  |      |
| TNM stage            |               |                  |      |
| I/II                 | 470 (0.46)    | 1111 (0.86)      |      |
| III/IV               | 551 (0.54)    | 293 (0.22)       |      |
|          |  |                  |      |
| age                  |               |                  |      |
| ≤60                  | 459 (0.45)    | 612 (0.47)       |      |
| >60                  | 562 (0.55)    | 692 (0.53)       |      |
| sex                  |               |                  |      |
| male                 | 715 (0.70)    | 887 (0.68)       |      |
| female               | 306 (0.30)    | 417 (0.32)       |      |

Table 2. Tag-SNPs results for GC in sumoylation-related genes.

| SNP       | Gene   | CHR position | Major allele | Minor allele | MAF in control | OR (95% CI) | P       |
|-----------|--------|--------------|--------------|--------------|----------------|-------------|---------|
| rs12614691| RANBP2 | 2            | 109362565    | A            | G              | 0.092       | 1.08 (0.88-1.33) | 0.465 |
| rs8062881 | UBE21  | 16           | 1361651      | C            | G              | 0.036       | 0.98 (0.71-1.34) | 0.885 |
| rs12920764| UBE21  | 16           | 1362187      | T            | A              | 0.020       | 1.14 (0.72-1.79) | 0.576 |
| rs75998799| UBE21  | 16           | 1366079      | G            | A              | 0.106       | 0.92 (0.72-1.16) | 0.473 |
| rs7610599 | UBE21  | 16           | 1374524      | G            | A              | 0.049       | 0.86 (0.61-1.21) | 0.372 |
| rs12925270| UBE21  | 16           | 137937     | C            | C              | 0.343       | 1.10 (0.97-1.25) | 0.153 |
| rs1285251 | CBX4   | 17           | 77809823    | C            | C              | 0.156       | 1.03 (0.87-1.21) | 0.756 |
| rs1285249 | CBX4   | 17           | 77810143    | G            | C              | 0.289       | 0.97 (0.85-1.10) | 0.606 |
| rs73422123| CBX4   | 17           | 77810391    | T            | A              | 0.166       | 1.24 (1.04-1.49) | 0.017 |
| rs77447679| CBX4   | 17           | 77810474    | C            | G              | 0.488       | 1.06 (0.94-1.19) | 0.316 |
| rs2289728 | CBX4   | 17           | 77811854    | A            | G              | 0.177       | 0.96 (0.82-1.13) | 0.641 |
| rs1285243 | CBX4   | 17           | 77814106    | T            | A              | 0.126       | 0.98 (0.82-1.18) | 0.851 |
| rs12980229| UBA2   | 19           | 34914144    | A            | G              | 0.403       | 1.01 (0.90-1.14) | 0.848 |
| rs177918  | SAE1   | 19           | 47634313    | G            | T              | 0.187       | 0.92 (0.79-1.07) | 0.272 |
| rs309184  | SAE1   | 19           | 47636827    | T            | G              | 0.080       | 0.90 (0.72-1.12) | 0.357 |
| rs5580126 | SAE1   | 19           | 47650489    | C            | A              | 0.171       | 1.02 (0.87-1.21) | 0.786 |
| rs1261339 | SAE1   | 19           | 47684908    | G            | G              | 0.275       | 0.89 (0.78-1.02) | 0.091 |
| rs3745621 | SAE1   | 19           | 47689921    | C            | C              | 0.042       | 1.00 (0.73-1.35) | 0.983 |
| rs5590314 | SAE1   | 19           | 47696626    | T            |                |             |         |

* adjusted for age and sex in logistic regression
Detailed allele distributions of rs77447679 in the case and control groups are shown in Table 3. Individuals with the CA genotype had significantly higher risk of GC, (1.71-fold compared to the baseline genotype CC, \( P = 0.018 \); adjusted OR: 1.71; 95% CI: 1.10-2.66); besides, recessive model also had a significantly increased risk of GC (CC vs. CA+AA, \( P = 0.017 \); adjusted OR: 1.24; 95% CI: 1.04-1.49). However, there was no statistically significant difference of AA genotype between the patient and control groups (\( P = 0.065 \); adjusted OR: 1.19; 95% CI: 0.99-1.44).

**Table 3.** Genotype frequencies of CBX4 rs77447679 polymorphism in GC patients and controls.

| rs77447679 genotype | Controls | OR (95% CI)* | P* |
|---------------------|----------|--------------|----|
| C>A                 |          |              |    |
| CC                  | 648      | 1.00 (Reference) |    |
| CA                  | 296      | 1.71 (1.10-2.66) | 0.018 |
| AA                  | 46       | 1.19 (0.99-1.44) | 0.065 |
| CA+AA               | 342      | 1.24 (1.04-1.49) | 0.017 |
| CC+CA               | 944      | 1.00 (Reference) |    |
| AA                  | 46       | 1.63 (1.05-2.52) | 0.030 |

* Adjustment for age and sex in logistic regression

**Discussion**

In this study, we have identified 17 tag-SNPs in 5 sumoylation-related genes (RANBP2, SAE1, UBE2I, UBA2 and CBX4) and quantified their association with GC susceptibility in a Chinese population. We demonstrated that CBX4 rs77447679 was significantly associated with the risk of GC, and individuals with CA genotype had a significantly higher risk of GC, comparing to the baseline CC genotype. This is a pioneer investigation to quantify the relationship between the sumoylation-related gene polymorphism and the risk of GC in Chinese population.

First reported in 1996, sumoylation modifies various eukaryotic proteins [7]. Sumoylation is governed by a conserved cascade consisting an E1-activating enzyme complex, an E2-conjugating enzyme and an E3 ligase [22]. The 5 sumoylation-related genes involved in this study (RANBP2, SAE1, UBE2I, UBA2, and CBX4) are responsible for gene regulation of the E1 enzyme complex, E2 or multiple E3 ligases. In this study, we have demonstrated that CBX4 polymorphism had strong association with GC development. CBX4 gene is a critical component of the polycomb repressive complex 1 (PRC1), which synergizes with PRC2 to silence gene expression by specifically modifying nucleosomal histones [23-25]. Additionally, CBX4 is a SUMO E3 ligase, which could upregulate the sumoylation of a more limited repertoire of substrate involved in tumorigenesis [26, 27]. Several recent studies have reported that CBX4 is closely related with the proliferation and differentiation of cloned embryos, thymic epithelial cells, and transarterial chemoembolization in hepatocellular carcinoma (HCC) [28-31]. Furthermore, CBX4 expression has been reported to upregulate tumor tissues, and higher CBX4 expression is correlated with the α-fetoprotein (AFP) level in serum, tumor size, pathological differentiation, and the TNM (tumor, node, metastasis) stages in HCC [32]. In addition, Jiao HK et al. demonstrated that CBX4 could promote angiogenesis and metastasis in HCC by the sumoylation pathway dependent on the hypoxia-inducible factor-1α protein and could be an independent prognostic factor for HCC patients [30]. The amplification and biological roles of the CBX4 gene have not yet been reported in GC. Regarding its well-known role in HCC, we hypothesize that CBX4 gene is involved in the pathogenesis of GC. However, subsequent studies on different populations are needed to confirm our finding.

This study has suggested that SNPs in gene’s functional regions could influence gene expression and functions [33-35]. Thus, 7 selected tag-SNPs (rs1285251, rs1285249, rs73422123, rs77447679, rs2289728, rs190846371, and rs1285243) of the CBX4 genes, located in the functional regions of CBX4, were examined for their potential association with GC. CBX4 rs77447679 variant could be involved in the development of GC and leading to abnormal expression of CBX4 proteins; moreover, patients with CC genotype had a lower risk of GC, which suggested that the C to A substitution of CBX4 rs77447679 could increase the transcription and expression of CBX4, and contribute to the development of GC (\( P = 0.018 \); adjusted OR: 1.71; 95% CI: 1.10-2.66). Thus, CBX4 rs77447679 polymorphism is a biologically plausible factor in GC. The association between CBX4 gene polymorphism and cancer risk has been reported in previous studies [36]. In a large-scale study of esophageal squamous cell carcinoma (ESCC), it was demonstrated that CBX4 gene was substantially amplified and upregulated in a Chinese population and could act as oncogenes and potential therapeutic targets in ESCC. Recurrent focal CNAs, including amplified chromosomal segments containing CBX4, were validated in ESCC, which implied that CBX4 amplification and the corresponding protein upregulation contributed to the development of ESCC [36]. Our findings on CBX4 rs77447679 polymorphism were consistent with previous studies of ESCC, which together could provide a novel interventional target for GC patients and people with certain susceptible genotypes (e.g., CC comparing to CA) and thus with a higher GC risk. We expect a better understanding of
sumoylation in the occurrence and development of GC.

There were, however, some limitations in this study. First, multiple factors, such as age, sex, smoking status, drinking status, and dietary habits, might influence the development of GC, and stratification would be recommended. Besides, the expression of CBX4 proteins in patients’ tumor tissues and controls, as well as the underlying mechanisms of this polymorphism in GC-related genes require further investigation. Finally, this case-control study was based on hospital participants, so there was inherent possibility of selection bias.

Conclusion

We have identified that the CBX4 rs77447679 polymorphism was statistically significantly associated with GC, and individuals with CC genotype had lower GC risk. More comprehensive investigation is necessary to evaluate the interactions between the genetic and environmental factors, as well as the underlying mechanisms of the association between rs77447679 polymorphism and the risk of GC.

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Competing Interests

The authors have declared that no competing interest exists.

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