Association of miR-34a-3p/5p, miR-141-3p/5p, and miR-24 in Decidual Natural Killer Cells with Unexplained Recurrent Spontaneous Abortion

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Background: The specific causes of recurrent spontaneous abortion (RSA) remain unknown in 37–79% of affected women. The aim of this study was to explore the expression levels of 6 miRNAs in natural killer (NK) cells from the decidua of patients with unexplained RSA (URSA) and to predict the target genes of 3 miRNAs.

Material/Methods: Two groups were examined: URSA (n=20) and controls (n=20). Flow cytometry analysis was used to identify NK cells isolated from the decidua. Transcriptional levels of miRNA were monitored using quantitative real-time reverse transcription-polymerase chain reaction. Prediction and analysis of mRNA targets of differentially expressed miRNAs were performed using bioinformatics methods.

Results: Five miRNAs [miR-34a (+281%, P<0.001), miR-155 (+396%, P<0.001), miR-141 (+142%, P<0.01), miR-125a (+279%, P<0.001), and miR-125b (+185%, P<0.001)] were up-regulated, while miR-24 was down-regulated (~64%, P<0.01) in the URSA group, compared to the control group. This study identified potential mRNA targets: miR-34a-3p/5p, 58S/1718 (targets of miR-34a-3p/targets of miR-34a-5p), miR-141-3p/5p, 2270/629 (targets of miR-141-3p/targets of miR-141-5p), and miR-24, 2320 target genes. A total of 140 pathways related to target genes were identified including PI3K-Akt, focal adhesion, MAPK, Wnt, regulation of the actin cytoskeleton, T cell receptor, TGF-β, and estrogen signaling pathways.

Conclusions: This study suggests that miR-34a-3p/5p, miR-141-3p/5p, and miR-24 in decidual NK cells could be associated with URSA. These findings might contribute to the panel of diagnostic and prognostic biomarkers with clinical utility, and facilitate the development of new strategies for targeted therapy against URSA.

MeSH Keywords: Abortion, Induced • Killer Cells, Natural • MicroRNAs

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Background

Recurrent spontaneous abortion (RSA) is defined as 3 or more consecutive pregnancy losses before the 22nd gestational week or spontaneous abortion of an embryo/fetus weighing less than 500 g [1]. Diagnosis of RSA requires multiple tests to detect parental chromosomal anomalies and maternal thrombophilic, endocrine, or immunological disorders. However, the specific causes of RSA remain unknown in 37–79% of affected women [1,2]. Over 50% of RSA cases are classified as idiopathic unexplained RSA (URSA) and the mechanisms of URSA are not completely understood [3]. Parental exposure to pheno- l-based pollutants might play a role in URSA [4], as well as polymorphisms in genes such as methylene tetrahydrofolate reductase and methionine synthase reductase [5]. Nevertheless, the immune system is considered one of the main culprits. Indeed, studies have shown that Th17 cells play a major role in rejecting conceptus antigens due to a Th17/Treg imbalance in URSA [6,7]. In addition, the Th1/Th2 ratio also plays a role in URSA [8–10], as well as the aberrant expression of the human leukocyte antigen (HLA) [11], dysfunction of uterine natural killer (NK) cells [12], and the presence of some auto-antibodies [13]. Immunotherapy with mononuclear cells from the father has been shown to bias the Th17/Treg ratio and to be beneficial for pregnancy.

The decidua and the immune cells residing in it play roles in the maintenance of pregnancy. The decidua invades the uterine mucosa during the first trimester of pregnancy and is thought to be critical for controlling the depth of invasion of trophoblasts. The decidua contains a unique complement of immune cells comprising 70% of NK cells [14]. Decidual NK cells differ from blood NK cells, both phenotypically and functionally. A number of studies have suggested that decidual NK cells play a role in mediating trophoblast invasion and vascular remodeling through their ability to secrete an array of regulatory molecules, chemokines, and cytokines [15]. Therefore, these cells might play a critical role in the occurrence of URSA, but their regulation is still poorly known.

MicroRNAs (miRNAs) are a class of small non-coding regulatory RNAs influencing mRNA translation via direct interaction and/or degradation; they are involved in the control of a variety of cellular functions, including proliferation, stem cell maintenance, differentiation, cell death, and metabolism [16]. Recent studies have indicated involvement of aberrant miRNAs and their target genes in URSA. These studies have shown that miRNA-133a and its target HLA-G, miRNA-125 and its target LIFR/ERBB2, and miRNA-17 were dysregulated, which are involved in immune system regulation and cell proliferation [17–19]. A study has shown that 5 miRNAs were upregulated in the decidua of patients with RSA and that these miRNAs are involved in adhesion, apoptosis, and angiogenesis [20]. Higher expression of miR-486-3p and lower expression of miR-3074-5p in placental villa might also play a role in RSA [21]. In parallel, studies have revealed the ability of some miRNAs to regulate cell proliferation and apoptosis [22,23], which are critical processes of normal embryogenesis [24]. Among candidates miRNAs, members of the miRNA-34 family (miR-34a, miR-34b, and miR-34c) are direct transcriptional targets of the tumor suppressor protein p53, with the potential to regulate both apoptosis and cell proliferation [25]. miR-155 has been shown to regulate apoptosis and was proposed to target caspase 3 and NF-κB signaling [26, 27]. miRNA-125b is suggested to control expression of the tumor necrosis factor α (TNF-α) [28], a powerful activator cytokine of NF-κB [29]. miR-24 has potential target sites in the 3’UTR region of IFN-γ mRNA and negatively regulates IFN-γ expression. This miRNA controls a complex network of apoptotic and angiogenic pathways in endothelial cells [30,31]. miR-141, a member of the miR-200 family, is reportedly associated with various human malignancies. For instance, miR-141 is upregulated in ovarian cancer [32], but downregulated in hepatocellular and prostate cancers [33,34].

Therefore, this study aimed to examine the expression of miR-34a, miR-155, miR-141, miR-125a, miR-125b, and miR-24 in URSA. This study identified miRNAs that might be involved in URSA, as well as their target genes. This new knowledge could be used in the design of novel strategies against URSA.

Material and Methods

Patients and samples

All study participants were Chinese and recruited at the Family Planning Department, Beijing Obstetrics and Gynecology Hospital, Capital Medical University (China), between August 2014 and October 2014. The RSA group consisted of 20 childless Chinese women aged 25–35 (31.0±2.87) years with at least 3 or more successive spontaneous abortions at 7–10 (56.4±3.6 days) gestational weeks. Subjects did not suffer from systemic diseases, such as chronic renal failure, heart failure, agglutination disorders, lupus, ovarian polycystic, maternal reproductive anatomical hormonal abnormalities, paternal and maternal chromosomal abnormalities, or infection. RSA was detected using ultrasound. The control group included 20 fertile Chinese women aged 25–35 (29.9±2.0) years at 7–10 (57.9±4.3 days) gestational weeks with 1 or more children and no history of spontaneous abortion. Normal pregnancies were also detected using ultrasound. Decidual tissues were obtained from cases of induced abortion (IA) at about 7 gestational weeks. All patients provided written informed consent, and the study protocol was approved by the Ethics Committee of the Beijing Obstetrics and Gynecology Hospital, Capital Medical University.
Table 1. Primers for real-time PCR.

| microRNA     | Forward primer                  | Reverse primer                  |
|--------------|--------------------------------|---------------------------------|
| miRNA34a     | 5’-CTCGCTTCGGCAGCACA -3’        | 5’-AACGCTTCACGAATTTCGGT-3’      |
| miR-24       | 5’-CTCGCTTCGGCAGCACA -3’        | 5’-AACGCTTCACGAATTTCGGT-3’      |
| miR-155      | 5’-CTCGCTTCGGCAGCACA -3’        | 5’-AACGCTTCACGAATTTCGGT-3’      |
| miR-141      | 5’-CTCGCTTCGGCAGCACA -3’        | 5’-AACGCTTCACGAATTTCGGT-3’      |
| miR-125a/b   | 5’-CTCGCTTCGGCAGCACA -3’        | 5’-AACGCTTCACGAATTTCGGT-3’      |
| U6 snRNA     | 5’-CTCGCTTCGGCAGCACA -3’        | 5’-AACGCTTCACGAATTTCGGT-3’      |

Flow cytometry

Decidual tissues were cut into 1-mm pieces and enzymatically digested for 20 min using vigorous shaking and 1.5 mg of type I DNAse and 24 mg of type IV collagenase in 15 mL of RPMI-1640 medium (HyClone, Thermo Fisher Scientific, Waltham, MA, USA). This procedure was repeated 3 times. After an additional 5 min of incubation at room temperature without shaking, the supernatants were collected and loaded on Ficoll density gradient to purify the lymphocyte population. NK cells were purified using the human NK cell isolation kit (BD Biosciences, Franklin Lake, NJ, USA). The following mouse anti-human monoclonal antibodies (mAbs) conjugated with PCS and PE were used for cell sorting: anti-CD56 and anti-CD3 (all from BD Biosciences, Franklin Lake, NJ, USA), and matching conjugated isotype controls. For staining and cell sorting, cells were washed in PBS supplemented with 2% FCS and incubated with mAbs on ice for 30 min, followed by washing twice. Cell sorting and fluorescence measurements were performed on a MoFlo high-performance cell sorter (Cytomation, Dako, Glostrup, Denmark). Data from single cell events were collected using a standard FACScalibur™ flow cytometer (Immunocytometry systems; BD Biosciences, Franklin Lake, NJ, USA).

RNA extraction and qRT-PCR

Transcriptional levels of miRNAs were monitored using quantitative real-time reverse transcription-polymerase chain reaction (qRT-PCR). Total RNA was extracted using TRIzol total RNA isolation reagent (Invitrogen, Carlsbad, CA, USA), according to the manufacturer’s protocol. The quality of RNA was confirmed using an Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA, USA) [35]. Total RNA (1 μg) was used for cDNA synthesis with the First-Strand cDNA Synthesis Kit (Fermentas, Burlington, Canada). Real-time PCR was performed as follows: 95°C for 15 min for hot-start, followed by 95°C for 30 s, and 60°C for 60 s for 40 cycles, using the SYBR Green Quantitative PCR Master Mix (Applied Biosystems, Foster City, CA, USA) on a Bio-Rad iQ5 instrument (Bio-Rad, Hercules, CA, USA). Primers for amplification of microRNA (RiboBio, Guangzhou, China) are listed in Table 1. Each reaction was performed in triplicate and measured using the comparative Ct (2^ΔΔCt) method with U6 snRNA as the normalization control.

Prediction and analysis of mRNA targets of 3 differentially expressed miRNAs

Prediction of differentially expressed miRNA target genes was performed using the TargetScan 5.0 database (UK). Data on intersecting predicted target genes of differentially expressed miRNAs were collected. The negative regulatory relationships between miRNAs and genes were highlighted for analysis. Similarly, pathway analysis was used to determine the significant pathways negatively correlated with intersection of target genes according to the Kyoto encyclopedia of genes and genomes (KEGG, Japan), Biocarta (Germany), and Reatome (USA) databases [36,37].

Statistical analysis

SPSS 16.0 (IBM, Armonk, NY, USA), was used for data analysis. Data are presented as means ± standard deviation (SD). Statistical significance was evaluated by independent sample t-test. P<0.05 was considered statistically significant.

For pathway analysis, the Fisher’s exact and the chi-square tests were used to obtain the P and P(k) values to select significant pathways. The false discovery rate (FDR) was used to determine the threshold of the P-value in multiple tests and analyses. Significant pathways were considered as being those with FDR <0.05 and P<0.05.

A post hoc power analysis was performed. miR-141 was selected since it was the one with the smallest difference between the 2 groups. For the comparison of relative miR-141 expression levels between controls and patients with URSA, the power was 100% (n=20 and n=20, respectively, α=5%).

Table 1. Primers for real-time PCR.
Results

Identification of decidual NK cells

Figure 1 shows that the purity of CD56^+ CD3^- NK cells in decidua were more than 70%. These cells were sorted using flow cytometric analysis and used for the subsequent experiments.

Expression levels of miRNA in decidual NK cells from patients with URSA

The expression levels of the 6 identified miRNAs were evaluated using qRT-PCR. Five of the miRNAs (miR-34a (+281%, \( P < 0.001 \)), miR-155 (+396%, \( P < 0.001 \)), miR-141 (+142%, \( P < 0.01 \)), miR-125a (+279%, \( P < 0.001 \)), and miR-125b (+185%, \( P < 0.001 \)) were up-regulated, while miR-24 was down-regulated. ** \( P < 0.01 \), *** \( P < 0.001 \).
Target gene prediction and pathway analysis

Three of the above miRNAs (miR-34a, miR-141, and miR-24) were subjected to further target gene prediction and functional analyses. Available databases were used to compile potential targets for all the miRNA genes, leading to the identification of miR-34a-3p/5p, 585/1718 (number of target genes of miR-34a-3p/number of target genes of miR-34a-5p), miR-141-3p/5p, 2270/629 (number of target genes of miR-141-3p/number of target genes of miR-141-5p), and miR-24, 2320 target genes. KEGG analyses were subsequently performed to determine the biological pathways of the target genes. A total of 140 target gene-related pathways were identified (Supplementary Figure 1), among which the 29 most significant pathways are shown in Figure 3, including PI3K-Akt, focal adhesion, MAPK, Wnt, regulation of the actin cytoskeleton, T cell receptor, TGF-beta, and estrogen signaling pathways.

Discussion

The specific causes of RSA remain unknown in 37–79% of affected women. Evidence suggests that the immune system is involved in URSA, but the precise mechanisms are poorly understood. Therefore, this study aimed to explore the expression levels of 6 miRNAs in NK cells from the decidua of patients with URSA and to predict the target genes of 3 miRNAs. Results showed that 5 miRNAs (miR-34a, miR-155, miR-141, miR-125a, and miR-125b) were up-regulated, while miR-24 was down-regulated in the URSA group, compared to the control group. This study identified potential miRNA targets, such as miR-34a-3p/5p, 585/1718, miR-141-3p/5p, 2270/629, and miR-24, 2320, and 140 pathways related to target genes were identified including PI3K-Akt, focal adhesion, MAPK, Wnt, regulation of the actin cytoskeleton, T cell receptor, TGF-beta, and estrogen signaling pathways. These results suggest some pathways could be involved in URSA. These pathways should be explored because related treatments could be possible.

In this study, decidual CD56+ CD3− NK cells were isolated and flow cytometric analysis revealed a purity of more than 70%. This profile of expression is deemed sufficient to correctly isolate NK cells from the decidua [38,39]. Furthermore, the use of cell sorting allowed the use of a relatively pure population of cells.

As previously shown, members of the miR-34 family are direct regulators of p53 and are involved in the regulation of apoptosis and cell proliferation [25]. No previous study is available showing a possible association between miR-34 and RSA. Nevertheless, previous studies revealed that p53 is involved in URSA. Indeed, Shang et al. [40] showed that the p53-CDKN1A and p53-Bax pathways were activated in RSA, while Wei et al. [41] showed that the placental villi from patients with URSA expressed high levels of p53. A previous study in mice revealed that miR-141 might influence cell proliferation and apoptosis in the endometrium by downregulating the expression of PTEN and that miR-141 plays important roles in embryo implantation [42]. miR-24 is aberrantly expressed in polycystic ovary syndrome, which is in itself associated with pregnancy problems [43].
Supplementary Figure 1. A total of 140 target gene-related pathways.
addition, miR-24 downregulates MYC, a key proliferative factor involved in embryo implantation [43]. However, the exact relation between these miRNAs and their targets is still poorly understood. Some of the data presented above suggest possible beneficial effects of their miRNAs on pregnancy, but the present study suggests detrimental effects. Additional studies are necessary to address these issues.

Pathway analysis for predicted target genes of miR-34a, miR-141, and miR-24 revealed association with numerous pathways, including PI3K-Akt, MAPK, Wnt, T cell receptor, and TGF-β signaling, all of which regulate discrete cellular functions and interact as a network. The PI3K-Akt signaling pathway plays a significant role in cell survival. Activation of this pathway inhibits apoptosis, allowing cell proliferation [44]. The mitogen-activated protein kinase (MAPK) cascade is a highly conserved module involved in various cellular functions, including proliferation, differentiation, and migration [45].

This study is not without limitations. First, CD56+CD3– NK cells could comprise more than 1 population of cells. Future studies should aim to better characterize these cells. Second, only a few miRNAs were investigated. Third, the targets of the identified miRNAs were not directly assessed. Future studies should examine these target proteins in URSA. Further comprehensive research, such as genome-wide association analyses, may be required to facilitate the identification of more effective targeted treatments and significant markers in URSA.

Conclusions

This study suggests that miR-34a-3p/5p, miR-141-3p/5p, and miR-24 in decidual NK cells are associated with URSA. These findings might contribute to the panel of diagnostic and prognostic biomarkers with clinical utility, and facilitate the development of new strategies for targeted therapy against URSA.

Competing interests

All authors declare that they have no conflict of interest.

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