Utility of Magnetic Resonance Imaging and Positron Emission Tomography in Rheumatic Diseases

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The use of magnetic resonance imaging (MRI) and positron emission tomography (PET) in rheumatology allows a better understanding of the pathophysiology of the disease as well as early diagnosis and appropriate evaluation of the disease status and treatment responses. Despite the limited availability because of the high cost, there is growing evidence for the advantages and clinical application of these modalities to various rheumatic diseases. This review discusses the basic mechanisms and clinical applications of MRI and PET in certain rheumatic diseases, including rheumatoid arthritis, axial spondyloarthritis, idiopathic inflammatory myopathies, large vessel vasculitis, central nervous system disease associated with rheumatic diseases, polymyalgia rheumatica, adult-onset Still’s disease, and immunoglobulin G4-related disease. (J Rheum Dis 2020;27:136-151)

Key Words. Magnetic resonance imaging, Positron emission tomography, Rheumatic diseases

INTRODUCTION

Recently, novel imaging technologies, such as magnetic resonance imaging (MRI) and positron emission tomography (PET), have emerged in the field of rheumatology. Despite their limited availability owing to their high cost, they provide unique information on patients with rheumatic diseases. They may allow a better understanding of the pathophysiology of the disease as well as an early diagnosis and appropriate evaluation of the disease status and treatment response. This paper reviews the basic mechanisms and clinical applications of MRI and PET in certain rheumatic diseases, including rheumatoid arthritis (RA), axial spondyloarthritis (axSpA), idiopathic inflammatory myopathies (IIM), large vessel vasculitis (LVV), central nervous system (CNS) disease associated with rheumatic diseases, polymyalgia rheumatica (PMR), adult-onset Still’s disease (AOSD), and immunoglobulin G4-related disease (IgG4-RD).

MAIN SUBJECTS

MRI

1) Principles of MRI

MRI uses powerful magnets and radio frequency (RF) to produce three-dimensional images [1]. The application of a strong, external magnetic field aligns protons within the tissue, which is then disturbed by an external RF energy. Tissue can be characterized by two different relaxation times, T1 and T2, which are distinguished according to the time from the excitation of protons to relaxation [1,2]. Water with long T1 values appears dark (low signal), whereas fat with short T1 values appears bright (high signal) in the T1-weighted images. On the other
hand, T2-weighted images depict both fat and water with high signal intensity [2]. Muscle shows intermediate signal intensity that is slightly higher than water but lower than fat in the T1-weighted images, and a much lower signal intensity than both fat and water in the T2-weighted images [3].

Inflammatory lesions generally produce high-intensity signals in the T2-weighted images caused by edema (water); the fat also constitutes a significant proportion of the high-intensity signals. Intentional suppression of the fat signal augments true inflammatory lesions from the T2-weighted images [4-6] and can be applied to both T1- and T2-weighted sequences. Currently, fat saturation and short tau inversion recovery (STIR) are common methods for fat suppression [7]. Contrast agents, such as gadolinium, enhance highly vascularized lesions and are generally recommended when assessing the peripheral joints, particularly in inflammatory settings [8,9]. Occasionally, post-contrast T1 sequences are fat-suppressed to enhance the contrast resolution and improve the visibility of lesions [4,10].

MRI has excellent soft tissue resolution and can differentiate between fat, water, and muscle, and better characterize different joint structures, such as ligaments, hyaline, and fibrocartilage. Early bone involvement and inflammatory soft tissue changes to synovitis and tenosynovitis, which cannot be detected using conventional radiographic methods, can be visualized and assessed in detail by MRI [11]. Moreover, MRI is currently the only imaging modality available for assessing inflammation within the bone marrow (i.e., osteitis), which has been reported to be highly predictive of subsequent bone erosions [11-15]. Although computed tomography (CT) is considered the gold standard for the detection of bony destruction, MRI is another good method for monitoring the progression of erosions [16-18].

Because MRI does not involve ionizing radiation exposure, it can be applied in vulnerable populations, such as pregnant women and pediatric patients. On the other hand, MRI has several limitations, including relatively high cost and patient inconveniences, such as an enclosed claustrophobic space, long examination time, and loud noises. Moreover, MRI is sensitive to metal, thereby requiring specific safety measures in patients with metallic/electronic implants or metallic foreign bodies. The technique has a relatively low spatial resolution, resulting in thicker slices compared to CT [10,19].

2) Clinical applications of MRI in rheumatic diseases
The utility of MRI for non-inflammatory arthritis, such as rheumatoid arthritis, is also important. MRI can clearly demonstrate bone erosions, subchondral bone cysts, and subchondral bone lesions, which are often missed by conventional radiographs [20]. MRI can also provide valuable information on the extent and severity of joint lesions, including cartilage damage and bone edema, which are crucial for the management of rheumatic diseases [21].

Reference:
Kim et al. J Korean Rheum Assoc 1995;2:9-18 [20].
as osteoarthritis, lies in its excellent anatomical resolution of soft tissues, of which the virtue is often missing from CT. The cartilage loss and degenerative changes in the supporting tissues, such as tendons and ligaments, can be identified by MRI. MRI has been the topic of interest in terms of its role in the diagnosis, monitoring, and prognosis prediction of inflammatory rheumatic diseases because of its ability to detect inflammatory changes as well as anatomic resolution.

1. MRI in RA

MRI in the diagnosis of RA: Inflammatory arthritis is characterized by synovitis, bone marrow edema (BME), and adjacent tenosynovitis. The inflamed and hypertrophied synovium is depicted by the high-intensity signal in T2-weighted images, which would be more prominent in the fat suppressed T2-weighted images [9,20] (Figure 1). BME lesions, which are some of the earliest findings of inflammatory arthritis, are detectable on MRI as early as nine weeks from the onset of symptoms [21,22].

Longitudinal studies of undifferentiated arthritis (UA) reported MRI to be an independent predictor of a subsequent diagnosis of RA [23,24]. In 116 patients with UA, a prediction model, including clinical hand arthritis, morning stiffness of more than one hour, rheumatoid factor positivity, and the MRI BME score in the metatarsophalangeal and wrist joints, correctly identified the development of RA and non-RA in 82% of patients (sensitivity 81%, specificity 82%) [24].

In another study of UA, an assessment of MRI-detected tenosynovitis showed the best accuracy in predicting a future RA diagnosis, particularly among UA patients with oligoarthritis (odds ratio = 7.5, 95% confidence interval [95% CI] 2.4–23). The technique also had a high negative predictive value of 95% (95% CI 89%–98%) [25].

According to the European League against Rheumatism (EULAR) recommendations, MRI can be used to improve the certainty of diagnosis above the clinical criteria alone when a diagnosis of RA is in doubt. The presence of inflammation on MRI can predict the progression of UA to clinical RA [26].

MRI in assessing the disease activity and structural damage in RA: MRI is valuable for assessing disease activity and structural damage in RA. Synovitis on MRI was shown to be associated significantly with histologic synovial inflammation, clinical disease activity, and patient-reported outcomes in RA [27-29]. When evaluating the responsiveness of MRI, ultrasonography, conventional radiography, and the core measures of the disease activity in RA patients during the first year of treatment with tumor necrosis factor (TNF) inhibitors, the best measure of inflammation was a composite measure comprising MRI synovitis, tenosynovitis, and BME [30].

MRI was superior to conventional radiography in detecting the early effects of different treatments on structural damage to bone and cartilage [31-33]. The level of agreement for the detection of bone erosions between MRI and CT was high (concordance at 77%–90% of sites) in the RA wrist and metacarpophalangeal joints (MCPs) [16,17,34]. Based on CT as the reference method, the reported sensitivities and specificities for MCP erosions were 68% and 92% for MRI, 44% and 95% for US, and 26% and 98% for radiography [18].

The RA MRI scoring system (RAMRIS) developed by the Outcomes Measures in Rheumatology (OMERACT) is a validated scoring system that quantifies both inflammation (synovitis, tenosynovitis, and BME) and joint damage (bone erosion and joint space narrowing) in RA hands and wrists [35]. The system has demonstrated construct validity, good inter- and intra-reader reliability, excellent correlation with traditional radiographic methods, and high sensitivity to change [36-38]. Assessments of inflammation and joint damage on MRI using the RAMRIS method have been used widely to evaluate the treatment responses in RA patients [30,31,39,40].

Predictive value of MRI for radiographic progression in RA: MRI assessments of inflammation and joint damage in the wrist or MCPs have predictive value for subsequent radiographic progression in the bilateral hands, wrists, and feet [12-15,32,41]. In particular, BME was reported to be the strongest predictor of future radiographic progression in RA [12-15]. Regression analyses in the three- and five-year follow-up in two cohorts and clinical trials showed that BME on MRI had independently predicted the short-term and long-term (3, 5, and 11 years) radiographic progression in RA [13,41-43]. These studies showed that an MRI prediction for radiographic progression was better than the clinical measure of the disease activity alone. Subclinical inflammation observed on MRI in RA patients in clinical remission or low disease activity was also reported to be predictive of subsequent radiographic progression [44,45].

2. MRI in axSpA

MRI in the diagnosis and classification of axSpA: MRI of the sacroiliac joints (SI) is essential for the diagnosis and classification of axSpA. The 2009 Assessment of
SpondyloArthritis international Society (ASAS) classification criteria led to differentiation between ankylosing spondylitis and non-radiographic axSpA (nr-axSpA) based on the presence of definite radiographic sacroiliitis and sacroiliitis in MRI [46]. To define nr-axSpA, MRI evidence of SI inflammation, which is radiographically occult, was included. The ASAS published definitions for a positive MRI of the SI. Active sacroiliitis is indicated by the clear presence of BME in the subchondral bone of the SI on STIR, fat saturated T2-weighted, or gadolinium-enhanced T1-weighted images [5,47] (Figure 2). The sole presence of other inflammatory lesions, such as synovitis, enthesitis, or capsulitis without concomitant BME is not sufficient, and structural lesions, including fat metaplasia, sclerosis, erosion or ankylosis, may be observed but are not required to meet the definition of active sacroiliitis on MRI [5].

The ASAS-defined MRI changes based on BME lack specificity. Positive MRI findings, according to the ASAS definition, were also observed in patients with non-specific back pain [48], postpartum women [49], military recruits [50], runners [49], athletes [51], as well as in the general population [52]. This suggests that the imaging arm of the ASAS classification criteria may identify false-positive classification and diagnosis. On the other hand, deep extensive BME lesions (a homogeneous, unequivocal increase in signal ≥ 1 cm from the articular surface) were reported to be highly specific for axSpA-associated sacroiliitis [49].

In addition to the inflammatory MRI changes, structural changes in the SI have value for the classification and diagnosis of axSpA [53,54]. The SIMACT (sacroiliac joint MRI and CT study Sacroiliac MAgetic resonance Computed Tomography) study reported that using low-dose CT as the standard of reference, MRI T1-weighted images had better sensitivity with significantly more correct imaging findings than conventional radiographs for erosions (79% vs. 42%), joint space changes (75% vs. 41%), and overall positivity (85% vs. 48%). At the same time, there were no differences regarding the specificity between radiographs and MRI T1-weighted sequences (> 80% for all scores) [53]. MRI T1-weighted images were inferior to radiographs only for sclerosis (sensitivity 30% vs. 70%, respectively), but the differences were not significant [53].

Fat metaplasia, which manifests as increased periarticular signal intensity on the T1-weighted MR images, is typical for sacroiliitis [55,56]. On the other hand, MRI-detected fat metaplasia in SI was reported to be prevalent in subjects without axSpA subjects, ranging from 50.6% in patients aged < 45 years to 94.4% in those ≥ 75 years, whereas MRI-detected erosions were uncommon, less than 3% of the entire study population [57]. Some authors proposed a positive MRI for structural lesions of the SI for classifying axSpA as follows: fatty lesions ≥ 3, erosions ≥ 3, or fatty lesions and/or erosions ≥ 5 [54,58].

MRI in assessment of the treatment efficacy in axSpA: Owing to its high performance on soft tissue and BM abnormalities, MRI has been used to evaluate the efficacy of treatments in axSpA. Treatment with biological agents, such as TNF inhibitors and interleukin-17 inhibitors, reduced the MRI-detected inflammation sig-

Figure 2. Active inflammatory lesions of the sacroiliac joints in non-radiographic axial spondyloarthritis. (A) There is no evidence of sacroiliitis on plain radiography. (B and C) Magnetic resonance imaging short tau inversion recovery (B) and fat suppressed gadolinium-enhanced T1-weighted images (C) show bone marrow edema with high-intensity signal (arrows). Adopted from Lee. J Rheum Dis 2014;21:110-21 [47].
nificantly in the spine and SI [59-64], which is not measured by CT or plain radiography.

In addition to these inflammatory changes, the structure-modifying effects of TNF inhibitors have also been documented using MRI in patients with axSpA [62,65]. The mean SPARCC (the Spondyloarthritis Research Consortium of Canada) SI erosion score decreased (-0.6), and the SPARCC SI backfill score increased (+0.8) after a 12-week treatment with adalimumab [62]. Significant reductions in erosions (-0.57) and increases in backfill (+0.36) in the SI compared to placebo (-0.08 and 0.06, respectively) were also observed in patients with nr-axSpA following 12-week etanercept therapy [65].

3) MRI in IIM

MRI is an essential imaging tool for evaluating muscle involvement in the IIM [66]. MRI assists in the diagnosis of IIM by providing important pathological changes, including muscle edema (inflammation), fascitis, subcutaneous inflammation, muscle atrophy, and fatty replacement. Because inflammatory lesions are often patchy, MRI plays a role in selecting the optimal site for a muscle biopsy and reducing the rate of false-negative results [67,68]. MRI also helps define the extent of muscle involvement, disease activity (water signal changes), and the chronicity of the disease (fatty replacement and muscle atrophy) [66,69]. Therefore, MRI is considered an appropriate diagnostic and monitoring tool that can reflect both inflammation and damage to the affected muscle.

No standardized and validated scoring system for evaluating the muscle MRI has been developed [70]. Various semi-quantitative scoring systems and quantitative methods for assessing different muscle MRI signal parameters have been suggested. Muscle edema has been evaluated semi-quantitatively not only in the individual muscles [71,72] but also in whole muscle compartments [73]. A Norwegian study proposed a scoring system for evaluating the MRI changes associated with the disease activity (edema in muscles and fascia; score 0~42) and damage (fatty replacement and muscle volume reduction; score 0~36) with a total MRI score of 0~78 [73]. Because quantitative evaluation systems are based on computer analysis of the pixel intensity values, they provide a more precise evaluation of the muscle changes with statistically well-defined outcomes. These quantitative methods showed a good correlation with semi-quantitative evaluation systems [74,75].

Thigh MRI patterns varied by the clinical subsets of IIM have been described. Polymyositis (PM) is likely to involve the proximal thigh muscles, either globally or predominantly in the posterior muscle group [76]. The muscle edema in PM was diffuse due to the immune reaction in the myofibers. On the other hand, muscle edema in dermatomyositis (DM) involved the anterior muscle groups predominantly and was rather focal and patchy [77]. Fascial edema appeared to be characteristic of DM [78,79] because of the underlying pathophysiology of DM, such as perivascular and perifascicular inflammation, and possible ischemic damage [80]. Immune-mediated necrotizing myopathies (IMNM), particularly in patients with the anti-signal recognition particle, had significantly more widespread muscle edema, atrophy and fatty replacement than those with PM and DM, using semi-quantitative scoring systems [79]. Patients with inclusion body myositis showed more severe muscle atrophy and fatty replacement than those with PM, DM, and anti-hydroxy-3-methylglutaryl-coenzyme A reductase-positive IMNM [79].

4) MRI/MRA in LVV

LVV is the most common form of primary vasculitis comprising Takayasu arteritis (TA) and giant cell arteritis (GCA). MRI and magnetic resonance angiography (MRA) are valuable methods for evaluating LVV. MRA is a specialized MRI technique to focus on blood vessels and flow, and is an alternative to or used in conjunction with MRI. The advantages of MRI/MRA are the absence of radiation and the high resolution of both the walls and lumens of the heart, entire aorta, and its branches. Wall edema (water content) on the T2-weighted images and diffuse circumferential wall thickness or mural contrast enhancement on the T1-weighted images are early signs of vessel inflammation [81]. EULAR has recommended MRI/MRA for diagnosing and evaluating LVV [82].

MRI/MRA in TA: TA primarily affects the aorta and its main branches. When contrast-enhanced MRI/MRA was compared with conventional angiography in 30 suspected patients, MRI/MRA was highly sensitive to the aorta and its branches (100%); only 50% of pulmonary artery lesions were detected [83]. Therefore, the diagnostic accuracy can differ according to the arteries involved. According to meta-analysis, MRA showed a higher sensitivity of 92% compared to ultrasound (81%), even though both had a similar specificity of > 90% for a TA diagnosis [84].

MRI/MRA has also been used to monitor the disease activity of TA. Evidence of wall edema of at least one vessel segment by MRI/MRA decreased from 94% (17/18) of
patients with clinically active disease to 56% (24/43) of clinically remitted patients [85]. Although MRI could provide more accurate information on the inflammatory activity than the clinical criteria, the presence of vessel wall edema did not correlate with the subsequent vascular anatomy changes. This has raised concerns regarding assessments of the disease activity relying on MRI vessel wall edema solely.

**MRI/MRA in GCA:** GCA also involves the aorta and its major branches, with a predilection for the branches of the carotid, including the superficial temporal artery. The MRI findings of GCA correlated well with the histological findings of temporal arteries and with the fulfillment of the American College of Rheumatology clinical criteria [86,87]. High-resolution MRI allowed detailed visualization of the temporal arteries and occipital arteries bilaterally with mural thickening and contrast enhancement as the vasculitic findings. In particular, MRI/MRA may be useful in those with non-temporal cranial vessel involvement. According to a meta-analysis of five prospective studies, the sensitivity and specificity of MRI were 68% ~ 89% and 73% ~ 97%, respectively, for the diagnosis of GCA [88]. Unlike diagnostic purposes, MRI provides only limited evidence for monitoring the disease activity in GCA. On the other hand, some studies showed that mural enhancement improved significantly after treatment [86,89].

**(5) MRI in CNS lesions of rheumatic diseases**

The organic CNS involvement of rheumatic diseases is diverse, including parenchymal inflammatory lesions, ischemic events, demyelinating lesions, and meningitis. Parenchymal inflammatory lesions consist of focal lesions, such as vasculitis or diffuse lesions, such as encephalitis. MRI is considered the gold standard of CNS imaging based on its superior sensitivity and accuracy than other modalities for the morphological detection of these lesions [90]. In particular, MRI offers better visualization of the posterior cranial fossa, containing the brainstem and the cerebellum, as well as good contrast between the grey and white matter. The detailed anatomical structures of CNS are well contrasted by T1- and T2-weighted images with or without enhancement, which are the most widely used sequences for inflammatory CNS lesions with a similar manner of interpretation for musculoskeletal images [91].

MRI is the most useful neuroimaging study in patients with neuropsychiatric systemic lupus erythematosus (NPSLE). The technique may reveal abnormalities that reflect focal NPSLE. From a large series (n=108) of NPSLE, abnormal MRI findings of the brain were found in 60%. Small-vessel diseases, such as white-matter hyperintensities, atrophy, microbleeds, lacunae, and recent small subcortical infarct, were the most common (57%), followed in order by large-vessel diseases (13%) and inflammatory-like lesions (6.5%). Cerebrovascular syndrome correlated with large-vessel diseases and microbleeds, whereas cognitive dysfunction correlated with white-matter hyperintensities [92]. White matter lesions or periventricular hyperintensities can be observed in patients with SLE who do not have neuropsychiatric symptoms [93].

MRI should be performed in all patients suspected of having primary CNS vasculitis on account of sensitivity in detecting abnormalities in CNS vasculitis [94]. MRI of the brain commonly shows multiple infarcts in multiple vascular territories, but it is not specific for CNS vasculitis. Infarcts can often be found in areas of the brain not affected by the more common causes of stroke (such as the corpus callosum) and rarely tumor-like lesions [95]. MRA, which is the most reliable for assessing the large proximal arteries, is inadequate for primary CNS vasculitis because most cases of primary CNS vasculitis have medium and small intracranial arteries affected rather than the large proximal arteries. Involvement of the large proximal intracranial arteries indicates an alternative diagnosis in the appropriate clinical context.

Several different imaging sequences other than conventional T1- and T2-weighted images have also been used. For ischemic lesions, diffusion-weighted (DW) sequences, in which the diffusion of water molecules generates contrast in MRI, are useful. DW images allow the early detection of ischemic lesions before overt diffuse damage in CNS, which is evident in conventional T1- or T2-weighted images by tracing the reduced water contents associated with reduced blood flow [96]. Fluid attenuation inversion recovery images are often used to evaluate white matter plaque near the ventricles or to identify demyelination.

MRI is also effective for evaluating the longitudinal changes of the above lesions after treatment. Overall, MRI is an indispensable tool for evaluating diverse CNS pathologies associated with rheumatic diseases, providing information for quick and critical decision making for diagnosis and treatment.
PET

1) Principles of PET

PET is an imaging technique that uses radioactive substances to visualize and measure various metabolic processes in the body [97]. \(^{18}\)F-fluorodeoxyglucose (FDG) is a radiopharmaceutical that is used to evaluate the glucose metabolism and is currently the most widely used PET tracer [98] to detect enhanced glucose uptake from the high glycolytic activity of cancer cells or inflammatory cells [99,100]. CT and PET have been combined (PET/CT) to evaluate the anatomy and function because accurate anatomical localization of various functional abnormalities is difficult from PET alone [101].

In addition to the diagnosis of cancer [99], PET/CT is used under a range of conditions, including inflammatory, infectious, cardiovascular, and neurological diseases [102]. In particular, its indication has been widening to rheumatic diseases, such as LVV, PMR, IIM, AOSD, and IgG4-RD.

2) Clinical applications of PET in rheumatic diseases

(1) PET in LVV

Determination of the PET activity in LVV: A visual grading scale (vascular to liver uptake) is recommended when interpreting the arterial wall FDG uptake in PET. A standardized 0~3 grading system is used for a visual grading scale: 0=no uptake (< mediastinum); 1=low-grade uptake (< liver); 2=intermediate-grade uptake (=liver); 3=high-grade uptake (> liver). Grade 2 is possibly indicative, and grade 3 is considered positive for active LVV [103,104]. A total vascular score (TVS; 0~21) can be calculated from a summation of each score from seven vascular regions (thoracic and abdominal aorta, and sub-clavian, carotid, axillary, iliac, and femoral arteries).

Quantitative methods, such as simple standardized uptake value (SUV) metrics and target-to-background ratios (TBR), have also been proposed. Erroneous signal quantification can occur when using the injected radiotracer dose, patient weight, and imaging time point. Therefore, TBR is recommended over SUV because the use of the ratio between two measurements can minimize the effects on signal quantification of errors [105]. TBR methods using lung [106], liver [107], or blood pool [108,109] as a reference have been proposed. A target-to-blood pool method was applied successfully in LVV [109,110], and is currently recommended by the cardiovascular committee of the European Association of Nuclear Medicine for the assessment of vascular wall inflammation [105].

PET in the diagnosis of LVV: Previous studies have reported the utility of PET in diagnosing LVV [104,111-114] (Figure 3). Meta-analysis of eight studies, including 170 LVV patients with GCA or TA and 230 controls, reported that the pooled sensitivity and specificity of PET or PET/CT were 75.9% and 93.0%, respectively, with a high positive likelihood ratio of 7.267 [111]. PET showed higher diagnostic performance for the identification of GCA than TA (87% vs. 58%, respectively; p<0.0001) [104,111]. In a study that evaluated the concordance of PET/CT and aortic CT angiography (CTA) in the diagnosis of GCA, both PET/CT and CTA could detect large-vessel involvement with comparable results in a per-patient analysis. On the other hand, PET/CT showed higher performance in a per-segment analysis, particularly in the detection of inflammation in branches of the aorta [115]. PET/CT was also useful for assessing local inflammatory and vascular remodeling events independent
of systemic inflammation during the follow-up, even in lesions where the arterial wall was less than 4 mm thick [116]. Recent EULAR recommendations for the imaging modalities in LVV included PET to diagnose LVV [82].

**PET in monitoring efficacy of treatment in LVV:** Patients with LVV can develop new arterial lesions during periods of apparent clinical remission, and the PET activity during clinical remission was shown to predict future clinical relapse and angiographic progression of the disease [112,117]. Treatment of patients with LVV showed a decrease in arterial FDG uptake with corresponding improvement in the clinical and serological outcomes [118-125] (Figure 4). Most of these studies used only small patient cohorts with retrospective study designs, as well as relatively short follow-up intervals between imaging assessments. In a prospective study of 35 GCA patients with repetitive whole-body PET/CT acquired at the baseline and after three and six months of glucocorticoid therapy, the mean TVS declined significantly on repeated PET at three months (from 7.9±5.5 to 2.4±3.5), but no further decrease was observed at six months [120].

Another prospective study in 12 active TA patients revealed a significant decrease in the qualitative PET Vascular Activity Score (PETVAS), and quantitative PET parameters on the follow-up PET/CT at week 30 after infliximab biosimilar therapy [123]. A recent six-month prospective study of 52 LVV patients using PETVAS reported that the treatment intensification led to a significant decrease in disease activity by imaging, clinical, and inflammatory markers. On the other hand, weakened treatment significantly worsened the PET activity without changing the clinical and serologic activity [124]. Based on the available evidence, PET is a potential tool for assessing the disease activity and treatment responses in LVV [126] (Figure 4). Nevertheless, further studies with a longer duration of repeated PET will be needed to confirm whether the PET uptake continues to improve with treatment.

(2) **PET in PMR**

PMR is an inflammatory rheumatic condition characterized by aching and morning stiffness of the shoulders, hip girdle, and neck, which is sometimes associated with...
GCA. A prospective study of 35 PMR patients, using the same methodology as that for GCA [120], reported increased FDG uptake in the shoulder, hips, and spinous processes of the vertebrae [127]. Further studies confirmed that PET/CT is useful for the diagnosis of PMR [128,129]. A high FDG uptake at two or more sites among ischial tuberosities, greater trochanters, and lumbar spinous processes showed high sensitivity (85.7%) and specificity (88.2%) for the diagnosis of PMR [129]. Bilateral diffuse lower limb, pelvic girdle, or inflammatory low back pain was reported to be predictors of a positive PET/CT scan for LVV in PMR [130]. Positive PET scans in patients with PMR did not correlate with their risk of relapse [127].

Regarding an assessment of the treatment response by PET/CT in PMR, the FDG uptake may reflect the disease activity. The FDG uptake was shown to decrease but was still present after three and six months of glucocorticoid therapy in patients with PMR [127]. A multicenter clinical trial assessing tocilizumab therapy in PMR patients with PMR did not correlate with their risk of relapse [127].

PET/CT may be used to detect inflammation in active IIM [132,133]. On the other hand, increased FDG muscle uptake is non-specific for myositis because it may simply reflect the intensity of metabolic activity within the affected muscles [132]. Increased FDG uptake can be observed even in normal muscles after physical exercise. PET/CT can also be used as a reliable screening tool of cancer in IIM, where an exceptionally high risk of malignancy exists [134].

(4) PET in AOSD
AOSD is a rare systemic inflammatory disease, of which fever, arthritis, and salmon-colored rash are the main characteristics. The patients may have pharyngitis or lymphadenopathy. Leukocytosis and elevated levels of serum ferritin can be noted. The patients usually present with fever of unknown origin (FUO). The diagnosis of AOSD is one of the challenges because infections, malignant tumors, and other autoimmune diseases that may have similar symptoms should be excluded ahead [135].

PET/CT can be applied to detect or exclude malignant tumors and infections [136-138]. In addition, it can also be used for a differential diagnosis between AOSD and other connective tissue diseases. In the case of AOSD, PET/CT shows characteristic FDG accumulation in the bone marrow, spleen, lymph nodes, joints, and salivary glands [139,140]. This helps select the appropriate biopsy location [141]. Because the visual grade and SUV intensity of FDG from lymph nodes, spleen, and bone marrow were co-related with the AOSD disease activity, it can be a tool for assessing the disease activity [142]. Compared to other connective tissue diseases, patients with AOSD show more frequent and stronger FDG uptake by the spleen and bone marrow, and lymphadenopathy is characterized by symmetrical involvement, mainly in the neck and axilla. In some cases of lymphoma, it is difficult to distinguish from AOSD, but they appear to show enlarged lymph nodes with morphological characteristics different from reactive hyperplasia [143].

From a large prospective study on the patients with FUO or inflammation of an unknown origin, of which a vast majority were finally identified as having rheumatic diseases, such as AOSD, PET/CT led to a substantial increase in the diagnosis of the patients (up to 57% of them). In particular, 80% of afebrile elderly patients with high CRP were predicted to have their underlying disease identified [144].

(5) PET in IgG4-RD
IgG4-RD is an immune-mediated fibro-inflammatory syndrome that shares certain pathological, serological, and clinical features but with an unknown etiology [145]. Patients often present with tumors in the affected organs. This is common among middle-aged and elderly men. The main pathological features of IgG4-RD are the tissue infiltration of IgG4 positive plasma cells and small lymphocytes [146]. In general, a patient has multiple symptoms that can be classified into four categories: pancreatic-hepatic-biliary disease, retroperitoneal fibrosis and aortitis, head and neck disease, and classic Mikulitz syndrome with systemic involvement [147].

A prospective cohort study on the patients with IgG4-RD found that PET/CT was more sensitive in detecting organ involvement than other techniques [148]. The study reported characteristic findings, such as increased diffuse uptake in the pancreas and salivary glands, patch lesions in the retroperitoneal region and vascular wall, and multi-organ involvement that was less likely to be a metastasis. PET/CT was also useful in a biopsy, treatment decisions, and evaluations of the treatment response [148]. In a study of patients with clinically suspected IgG4-RD, PET/CT of the patients diagnosed as having IgG4-RD showed more likely multi-organ involvement, lower maximum SUV (SUV_{max}) of the main involved organ and higher SUV_{max} of the submandibular
glands, compared to those with other diseases [149]. Using these parameters as the diagnosis criteria, the sensitivity and specificity for a diagnosis of IgG4-RD were up to 86% and 66%, respectively. Subsequent studies demonstrated PET/CT were helpful for diagnosis, identification of the involved organs, biopsy guidance, and treatment response evaluation of IgG4-RD, including sclerosing sialadenitis, autoimmune pancreatitis, and retroperitoneal fibrosis [150-153].

**CONCLUSION**

The growing evidence of MRI and PET justify their use in research and the routine care of patients with various rheumatic diseases. MRI and PET can play an important role in early diagnosis and monitoring the disease activity and response to therapy in patients with rheumatic diseases.

**CONFLICT OF INTEREST**

No potential conflict of interest relevant to this article was reported.

**AUTHOR CONTRIBUTIONS**

E.H.P. contributed to the study design, literature review and analysis, and to writing the manuscript. C.H.Y. contributed to the study conception and design, and to reviewing the manuscript. E.H.K. and H.J.B. equally contributed to the study conception and design, literature review and analysis, and to writing and completing the manuscript.

**REFERENCES**

1. Westbrook C, Roth CK, Talbot J. MRI in practice. 4th ed. Chichester, Wiley-Blackwell, 2011.
2. Grover VP, Tognarelli JM, Crosse MM, Cox IJ, Taylor-Robinson SD, McPhail MJ. Magnetic resonance imaging: principles and techniques: lessons for clinicians. J Clin Exp Hepatol 2015;5:246-55.
3. May DA, Disler DG, Jones EA, Balkissoon AA, Manaster BJ. Abnormal signal intensity in skeletal muscle at MR imaging: patterns, pearls, and pitfalls. Radiographics 2000;20:S295-315.
4. Østergaard M, Duer A, Møller U, Ejbjerg B. Magnetic resonance imaging of peripheral joints in rheumatic diseases. Best Pract Res Clin Rheumatol 2004;18:861-79.
5. Lambert RG, Bakker PA, van der Heijde D, Weber U, Rudwaleit M, Hermann KG, et al. Defining active sacroiliitis on MRI for classification of axial spondyloarthritis: update by the ASAS MRI working group. Ann Rheum Dis 2016;75:1958-63.
6. Sung S, Kim HS, Kwon JW. MRI assessment of sacroiliitis for the diagnosis of axial spondyloarthropathy: comparison of fat-saturated T2, STIR and contrast-enhanced sequences. Br J Radiol 2017;90:20170090.
7. Defaut EM, Beltran J, Johnson G, Rousseau J, Marchandise X, Cotten A. Fat suppression in MR imaging: techniques and pitfalls. Radiographics 1999;19:373-82.
8. Østergaard M, Conaghan PG, O’Connor P, Szkułdarek M, Klaurlund M, Emery P, et al. Reducing invasiveness, duration, and cost of magnetic resonance imaging in rheumatoid arthritis by omitting intravenous contrast injection -- does it change the assessment of inflammatory and destructive joint changes by the OMERACT RAMRIS? J Rheumatol 2009;36:1806-10.
9. Sudoł-Szołpiszka I, Jurik AG, Eshed I, Lennart J, Grainger A, Østergaard M, et al. Recommendations of the ESSR Arthritis Subcommittee for the use of magnetic resonance imaging in musculoskeletal rheumatic diseases. Semin Musculoskelet Radiol 2015;19:396-411.
10. Eshed I, Hermann KGA. MRI in imaging of rheumatic diseases: an overview for clinicians. Clin Exp Rheumatol 2018;36 Suppl 114:10-5.
11. Østergaard M, Pedersen SJ, Døhn UM. Imaging in rheumatoid arthritis--status and recent advances for magnetic resonance imaging, ultrasonography, computed tomography and conventional radiography. Best Pract Res Clin Rheumatol 2008;22:1019-44.
12. Hetland ML, Ejbørg BJ, Hørslev-Petersen K, Jacobsen S, Vestergaard A, Jurik AG, et al.; CIMESTRA study group. MRI bone oedema is the strongest predictor of subsequent radiographic progression in early rheumatoid arthritis. Results from a 2-year randomised controlled trial (CIMESTRA). Ann Rheum Dis 2009;68:384-90.
13. Hetland ML, Stengaard-Pedersen K, Junker P, Østergaard M, Ejbørg BJ, Jacobsen S, et al.; CIMESTRA study group. Radiographic progression and remission rates in early rheumatoid arthritis - MRI bone oedema and anti-CCP predicted radiographic progression in the 5-year extension of the double-blind randomised CIMESTRA trial. Ann Rheum Dis 2010;69:1789-95.
14. Haavardsholm EA, Boesene P, Østergaard M, Schildvold A, Kvien TK. Magnetic resonance imaging findings in 84 patients with early rheumatoid arthritis: bone marrow oedema predicts erosive progression. Ann Rheum Dis 2008;67:794-800.
15. Nieuwenhuis WP, van Steenbergen HW, Stomp W, Stijnen T, Huizinga TW, Bloem JL, et al. The course of bone marrow edema in early undifferentiated arthritis and rheumatoid arthritis: a longitudinal magnetic resonance imaging study at bone level. Arthritis Rheumatol 2016;68:1080-8.
16. Døhn UM, Ejbørg BJ, Hasselquist M, Narvestad E, Møller J, Thomsen HS, et al. Detection of bone erosions in rheumatoid arthritis wrist joints with magnetic resonance imaging, computed tomography and radiography. Arthritis Res Ther 2008;10:R25.
17. Døhn UM, Ejbørg BJ, Court-Payen M, Hasselquist M, Narvestad E, Szkułdarek M, et al. Are bone erosions de-
ected by magnetic resonance imaging and ultrasonography true erosions? A comparison with computed tomography in rheumatoid arthritis metacarpophalangeal joints. Arthritis Res Ther 2006;8:R110.

18. Defun UM, Ejbjerg B, Boonen A, Hetland ML, Hansen MS, Knudsen LS, et al. No overall progression and occasional repair of erosions despite persistent inflammation in adalimumab-treated rheumatoid arthritis patients: results from a longitudinal comparative MRI, ultrasonography, CT and radiography study. Ann Rheum Dis 2011;70:252-8.

19. Oldendorf W. Advantages and disadvantages of MRI. In: Oldendorf W, Oldendorf W Jr, eds. Basics of magnetic resonance imaging. Boston, Springer, 1988.

20. Kim HY, Kim JA, Lee SK, Hong CS, Suh JS. The utility of wrist magnetic resonance imaging in rheumatoid arthritis for disease activity measurement. J Korean Rheum Assoc 1995;2:9-18.

21. Ostendorf B, Scherer A, Mödder U, Schneider M. Diagnostic value of magnetic resonance imaging of the forefoot in early rheumatoid arthritis when findings on imaging of the metacarpophalangeal joints of the hands remain normal. Arthritis Rheum 2004;50:2094-102.

22. McQueen FM, Gao A, Ostergaard M, King A, Shalley G, Robinson E, et al. High-grade MRI bone oedema is common within the surgical field in rheumatoid arthritis patients undergoing joint replacement and is associated with oestis in subchondral bone. Ann Rheum Dis 2007;66:1581-7.

23. Tamai M, Kawakami A, Uetani M, Takao S, Arima K, Iwamoto N, et al. A prediction rule for disease outcome in patients with undifferentiated arthritis using magnetic resonance imaging of the wrists and finger joints and serologic autoantibodies. Arthritis Rheum 2009;61:772-8.

24. Duer-Jensen A, Hørslev-Petersen K, Hetland ML, Bak L, Ejbjerg BJ, Hansen MS, et al. Bone edema on magnetic resonance imaging is an independent predictor of rheumatoid arthritis development in patients with early undifferentiated arthritis. Arthritis Rheum 2011;63:2192-202.

25. Nieuwenhuis WP, van Steenbergen HW, Magnlus N, Newsum EC, Bloem JL, Huizinga TWJ, et al. Evaluation of the diagnostic accuracy of hand and foot MRI for early Rheumatoid Arthritis. Rheumatology (Oxford) 2017;56:1367-77.

26. Colebatch AN, Edwards CJ, Østergaard M, van der Heijde D, Balint PV, D’Agostino MA, et al. EULAR recommendations for the use of imaging of the joints in the clinical management of rheumatoid arthritis. Ann Rheum Dis 2013;72:804-14.

27. Østergaard M, Stoltenberg M, Lovgren-Nielsen P, Volck B, Jensen CH, Lorenzen I. Magnetic resonance imaging-determined synovial membrane and joint effusion volumes in rheumatoid arthritis and osteoarthritis: comparison with the macroscopic and microscopic appearance of the synovium. Arthritis Rheum 1997;40:1856-67.

28. Humby F, Mahto A, Ahmed M, Barr A, Kelly S, Buch M, et al. The relationship between synovial pathobiology and magnetic resonance imaging abnormalities in rheumatoid arthritis: a systematic review. J Rheumatol 2017;44:1311-24.

29. Baker JF, Conaghan PG, Emery P, Baker DG, Østergaard M. Relationship of patient-reported outcomes with MRI measures in rheumatoid arthritis. Ann Rheum Dis 2017;76:486-90.

30. Haavardsholm EA, Østergaard M, Hammer HB, Bøyesen P, Boonen A, van der Heijde D, et al. Monitoring anti-TNFalpha treatment in rheumatoid arthritis: responsiveness of magnetic resonance imaging and ultrasonography of the dominant wrist joint compared with conventional measures of disease activity and structural damage. Ann Rheum Dis 2009;68:1572-9.

31. Peterfy C, Emery P, Tak PP, Østergaard M, DiCarlo J, Otsa K, et al. MRI assessment of suppression of structural damage in patients with rheumatoid arthritis receiving rituximab: results from the randomised, placebo-controlled, double-blind RA-SCORE study. Ann Rheum Dis 2016;75:170-7.

32. Peterfy C, Strand V, Tian L, Østergaard M, Lu Y, DiCarlo J, et al. Short-term changes on MRI predict long-term changes on radiography in rheumatoid arthritis: an analysis by an OMERACT Task Force of pooled data from four randomised controlled trials. Ann Rheum Dis 2017;76:992-7.

33. Østergaard M, Emery P, Conaghan PG, Fleischmann R, Hsia EC, Xu W, et al. Significant improvement in synovitis, ostitis, and bone erosion following golimumab and methotrexate combination therapy as compared with methotrexate alone: a magnetic resonance imaging study of 318 methotrexate-naive rheumatoid arthritis patients. Arthritis Rheum 2011;63:3712-22.

34. Perry D, Stewart N, Benton N, Robinson E, Yeoman S, Crabbe J, et al. Detection of erosions in the rheumatoid hand; a comparative study of multidetector computerized tomography versus magnetic resonance scanning. J Rheumatol 2005;32:256-67.

35. McQueen F, Lassere M, Edmonds J, Conaghan P, Peterfy C, Bird P, et al. OMERACT rheumatoid arthritis magnetic resonance imaging studies. Summary of OMERACT 6 MR imaging module. J Rheumatol 2003;30:1387-92.

36. Ginat S, Lillegraven S, Haavardsholm EA, Eshed I, Conaghan PG, Peterfy C, et al. Validation of the OMERACT magnetic resonance imaging joint space narrowing score for the wrist in a multireader longitudinal trial. J Rheumatol 2015;42:2480-5.

37. Østergaard M, Bøyesen P, Eshed I, Gandjbakhch F, Lillegraven S, Bird P, et al. Development and preliminary validation of a magnetic resonance imaging joint space narrowing score for use in rheumatoid arthritis: potential adjunct to the OMERACT RA MRI scoring system. J Rheumatol 2011;38:2045-50.

38. Haavardsholm EA, Østergaard M, Ejbjerg BJ, Kvan NP, Uhlig TA, Lillefs G, et al. Reliability and sensitivity to change of the OMERACT rheumatoid arthritis magnetic resonance imaging score in a multireader, longitudinal setting. Arthritis Rheum 2005;52:3860-7.

39. Axelsen MB, Eshed I, Hørslev-Petersen K, Stengaard-Pedersen K, Hetland ML, Møller J, et al.; OPERA study group. A treat-to-target strategy with methotrexate and intra-articular triamcinolone with or without adalimumab effectively reduces MRI synovitis, ostitis and tenosynovitis and halts structural damage progression in early rheumatoid arthritis: results from the OPERA randomised controlled trial. Ann Rheum Dis 2015;74:867-75.

40. Conaghan PG, Durez P, Alten RE, Burmester GR, Tak PP,
Klarseskg L, et al. Impact of intravenous abatacept on synovitis, osteitis and structural damage in patients with rheumatoid arthritis and an inadequate response to methotrexate: the ASSET randomised controlled trial. Ann Rheum Dis 2013;72:1287-94.

41. Baker JF, Ostergaard M, Emery P, Hsia EC, Lu J, Baker DG, et al. Early MRI measures independently predict 1-year and 2-year radiographic progression in rheumatoid arthritis: secondary analysis from a large clinical trial. Ann Rheum Dis 2014;73:1968-74.

42. Bøyesen P, Haavardsholm EA, Ostergaard M, van der Heijde D, Sesseng S, Kvien TK. MRI in early rheumatoid arthritis: synovitis and bone marrow oedema are independent predictors of subsequent radiographic progression. Ann Rheum Dis 2011;70:428-33.

43. Hetland ML, Ostergaard M, Stengaard-Pedersen K, Junker P, Ejbjerg B, Jacobsen S, et al.; The CIMESTRA Study Group. Anti-cyclic citrullinated peptide antibodies, 28-joint Disease Activity Score, and magnetic resonance imaging bone oedema at baseline predict 11 years’ functional and radiographic outcome in early rheumatoid arthritis. Scand J Rheumatol 2019;48:1-8.

44. Brown AK, Conaghan PG, Karim Z, Quinn MA, Ikeda K, Peterfy CG, et al. An explanation for the apparent discrepancy between clinical remission and continued structural deterioration in rheumatoid arthritis. Arthritis Rheum 2008;58:2958-67.

45. Gandjbakhch F, Conaghan PG, Ejbjerg B, Haavardsholm EA, Foltz V, Brown AK, et al. Synovitis and osteitis are very frequent in rheumatoid arthritis clinical remission: results from an MRI study of 294 patients in clinical remission or low disease activity state. J Rheumatol 2011;38:2039-44.

46. Rudwaleit M, van der Heijde D, Landewé R, Listing J, Akkoc N, Brandt J, et al. The development of Assessment of SpondyloArthritis International Society criteria for axial spondyloarthritis (part II): validation and criteria for the Assessment of Spondylarthritis International Society classification criteria for axial spondyloarthritis (part II): validation and final selection. Ann Rheum Dis 2009;68:777-83.

47. Lee S. MRI features of axial spondyloarthritis and differential diagnosis: focusing on the spine and sacroiliac joint. J Rheum Dis 2014;21:110-21.

48. Arnbak G, Grethe Jurik A, Horslev-Petersen K, Hendricks O, Hermansen LT, Loft AG, et al. Associations between spondyloarthritis features and magnetic resonance imaging findings: a cross-sectional analysis of 1,020 patients with persistent low back pain. Arthritis Rheumatol 2016;68:892-900.

49. de Winter J, de Hooge M, van de Sande M, de Jong H, van Hoven L, de Koning A, et al. Magnetic resonance imaging of the sacroiliac joints indicating sacroiliitis according to the Assessment of SpondyloArthritis International Society definition in healthy individuals, runners, and women with postpartum back pain. Arthritis Rheumatol 2018;70:1042-8.

50. Varkas G, de Hooge M, Renson T, De Mts S, Carron P, Jacques P, et al. Effect of mechanical stress on magnetic resonance imaging of the sacroiliac joints: assessment of military recruits by magnetic resonance imaging study. Rheumatology (Oxford) 2018;57:508-13.

51. Weber U, Jurik AG, Zejden A, Larsen E, Jørgensen SH, Ruffbach K, et al. Frequency and anatomic distribution of magnetic resonance imaging features in the sacroiliac joints of young athletes: exploring “background noise” toward a data-driven definition of sacroiliitis in early spondyloarthritis. Arthritis Rheumatol 2018;70:736-45.

52. Baraliakos X, Richter A, Feldmann D, Ott A, Buelow R, Schmidt CO, et al. Frequency of MRI changes suggestive of axial spondyloarthritis in the axial skeleton in a large population-based cohort of individuals aged <45 years. Ann Rheum Dis 2020;79:186-92.

53. Diekhoff T, Hermann KG, Greese J, Schwenke C, Poddubnyny D, Hamb M, et al. Comparison of MRI with radiography for detecting structural lesions of the sacroiliac joint using CT as standard of reference: results from the SIMACT study. Ann Rheum Dis 2017;76:1502-8.

54. Bakker PA, van den Berg R, Hoeve M, van Lunteren M, Ez-Zaitouni Z, Fagerli KM, et al. Impact of replacing radiographic sacroiliitis by magnetic resonance imaging structural lesions on the classification of patients with axial spondyloarthritis. Rheumatology (Oxford) 2018;57:1186-93.

55. Maksymowycz WP, Wichuk S, Chioveranwisawakit P, Lambert RG, Pedersen SJ. Fat metaplasia and backfill are key intermediaries in the development of sacroiliac joint ankylosis in patients with ankylosing spondylitis. Arthritis Rheumatol 2014;66:2958-67.

56. Song IH, Hermann KG, Haibel H, Althoff CE, Poddubnyny D, Listing J, et al. Relationship between active inflammatory lesions in the spine and sacroiliac joints and new development of chronic lesions on whole-body MRI in early axial spondyloarthritis: results of the ESTHER trial at week 48. Ann Rheum Dis 2011;70:1257-63.

57. Ziegeler K, Eskhal K, Schorr C, Sieper J, Diekhoff T, Makowsky MR, et al. Age- and sex-dependent frequency of fat metaplasia and other structural changes of the sacroiliac joints in patients without axial spondyloarthritis: a retrospective, cross-sectional MRI study. J Rheumatol 2018;45:915-21.

58. de Hooge M, van den Berg R, Navarro-Compán V, Reijnierse M, van Gaalen F, Fagerli K, et al. Patients with chronic back pain of short duration from the SPACE cohort: which MRI structural lesions in the spine are most specific for axial spondylarthritis? Ann Rheum Dis 2016;75:1308-14.

59. Lambert RG, Salonen D, Rahman P, Inman RD, Wong RL, Einstein SG, et al. Adalimumab in patients with non-radiographic axial spondyloarthritis: synovitis and bone marrow oedema are in-...
the sacroiliac joints of patients in the randomized, double-blind, placebo-controlled danish multicenter study of adalimumab in spondyloarthritis, as assessed by the Berlin and Spondyloarthritis Research Consortium of Canada methods. Arthritis Rheumatol 2016;68:418-29.

63. Braun J, Baraliakos X, Hermann KG, Landewé R, Machado PM, Maksymowych WP, et al. Effect of certolizumab pegol over 96 weeks of treatment on inflammation of the spine and sacroiliac joints, as measured by MRI, and the association between clinical and MRI outcomes in patients with axial spondyloarthritis. RMD Open 2017;3:e000430.

64. Baraliakos X, Borah B, Braun J, Baeten D, Laurent D, Sieper J, et al. Long-term effects of secukinumab on MRI findings in relation to clinical efficacy in subjects with active ankylosing spondylitis: an observational study. Ann Rheum Dis 2016;75:408-12.

65. Maksymowych WP, Wichuk S, Dougados M, Jones HE, Pedersen R, Szumski A, et al. Modification of structural lesions on MRI of the sacroiliac joints by etanercept in the EMBARK trial: a 12-week randomised placebo-controlled trial in patients with non-radiographic axial spondyloarthritis. Ann Rheum Dis 2018;77:78-84.

66. Pipitone N. Value of MRI in diagnostics and evaluation of myositis. Curr Opin Rheumatol 2016;28:625-30.

67. Lampa J, Nennesmo I, Einarsdottir H, Lundberg I. MRI guided muscle biopsy confirmed polymyositis diagnosis in a patient with interstitial lung disease. Ann Rheum Dis 2001;60:423-6.

68. Tomášová Studynková J, Čharvát F, Jarosová K, Vencovský J. The role of MRI in the assessment of polymyositis and dermatomyositis. Rheumatology (Oxford) 2007;46:1174-9.

69. Theodorou DJ, Theodorou SJ, Kakitsubata Y. The role of MRI in the assessment of polymyositis and dermatomyositis. Br J Radiol 2012;85:e1298-308.

70. Kubinová K, Mann H, Vencovský J. MRI scoring methods used in evaluation of muscle involvement in patients with idiopathic inflammatory myopathies. Curr Opin Rheumatol 2017;29:623-31.

71. Pipitone N, Notarnicola A, Levrini G, Spaggiari L, Scardapane A, Iannone F, et al. Do dermatomyositis and polymyositis affect similar thigh muscles? A comparative MRI-based study. Clin Exp Rheumatol 2016;34:1098-100.

72. Barsotti S, Zampa V, Talarico R, Minichilli F, Ortolini S, Iacopetti V, et al. Thigh magnetic resonance imaging for the evaluation of disease activity in patients with idiopathic inflammatory myopathies followed in a single center. Muscle Nerve 2016;54:666-72.

73. Andersson H, Kirkhus E, Garen T, Walle-Hansen R, Pedersen B, Zampà V, Talrico R, Mincichielli F, Ortolini S, Iacopetti V, et al. Thigh magnetic resonance imaging for the evaluation of disease activity in patients with idiopathic inflammatory myopathies followed in a single center. Arthritis Res Ther 2017;19:17.

74. Yao L, Yip AL, Sharde JA, Mesdaginia S, Volochayev R, Jansen AV, et al. Magnetic resonance measurement of muscle T2, fat-corrected T2 and fat fraction in the assessment of idiopathic inflammatory myopathies. Rheumatology (Oxford) 2016;55:441-9.

75. Elessawy SS, Abd El-Ghaffar Borg M, Abd El-Salam Mohamed M, Elhawy GE, Abd El-Salam EM. The role of MRI in the evaluation of muscle diseases. Egypt J Radiol Nucl Med 2013;44:607-15.
Radue EW, Weigel M, Wiest R, Urbach H. Introduction to magnetic resonance imaging for neurologists. Continuum (Minneap Minn) 2016;22:1379-98.

Sarbu N, Aloebedi F, Toledano P, Espinosa G, Giles I, Rahman A, et al. Brain abnormalities in newly diagnosed neuropsychiatric lupus: systemic MRI approach and correlation with clinical and laboratory data in a large multicenter cohort. Autoimmun Rev 2015;14:153-9.

Kozora E, West SG, Kotzin BL, Julian L, Porter S, Bigler E. Magnetic resonance imaging abnormalities and cognitive deficits in systemic lupus erythematosus patients without overt central nervous system disease. Arthritis Rheum 1998;41:41-7.

Salvarani C, Brown RD Jr, Calamia KT, Christianson TJ, Weigand SD, Miller DV, et al. Primary central nervous system vasculitis: analysis of 101 patients. Ann Neurol 2007;62:442-51.

Melloy ES, Singhal AB, Calabrese LH. Tumour-like mass lesion: an under-recognised presentation of primary angiitis of the central nervous system. Ann Rheum Dis 2008;67:1732-5.

Kucharczyk J, Mintorovitch J, Asgari HS, Mosesy L. Diffusion/perfusion MR imaging of acute cerebral ischemia. Magn Reson Med 1991;19:311-5.

Bailey DL, Townsend DW, Valk PE, Maisey MN. Positron emission tomography: basic sciences. Secaucus, NJ, Springer-Verlag, 2005.

Martínez MJ, Ziegler SI, Beyer T. PET and PET/CT: basic principles and instrumentation. In: Dresel S, ed. PET in oncology. Berlin, Heidelberg, Springer, 2008.

Fletcher JW, Djulbegovic B, Soares HP, Siegel LA, Lowe VJ, Lyman GH, et al. Recommendations on the use of 18F-FDG PET in oncology. J Nucl Med 2008;49:480-508.

Kubota R, Yamada S, Kubota K, Ishiwata K, Tamahashi N, Ido T. Intratumoral distribution of fluorine-18-fluorodeoxyglucose in vivo: high accumulation in macrophages and granulation tissues studied by microautoradiography. J Nucl Med 1992;33:1972-80.

Keidar Z, Haim N, Guralnik L, Wollner M, Bar-Shalom R, Ben-Nun A, et al. PET/CT using 18F-FDG in suspected lung cancer recurrence: diagnostic value and impact on patient management. J Nucl Med 2004;45:1640-6.

Zhuang H, Codreanu I. Growing applications of FDG PET/CT imaging in non-oncologic conditions. J Biomed Res 2015;29:189-202.

Lensen KD, Comans EF, Voskuyl AE, van der Laken CJ, Brouwer E, Zwijnenburg AT, et al. Large- vessel vasculitis: interobserver agreement and diagnostic accuracy of 18F-FDG-PET/CT. Biomed Res Int 2015;2015:914692.

Soussan M, Nicolas P, Schramm C, Katssahan S, Pop G, Fain O, et al. Management of large-vessel vasculitis with FDG-PET: a systematic literature review and meta-analysis. Medicine (Baltimore) 2015;94:e6522.

Bucurieus J, Hyafil F, Verberne HJ, Slart RH, Lindner O, Sciagra R, et al. Position paper of the Cardiovascular Committee of the European Association of Nuclear Medicine (EANM) on PET imaging of atherosclerosis. Eur J Nucl Med Mol Imaging 2016;43:780-92.

Moosig F, Czech N, Mehl C, Henze E, Zeuner RA, Kneba M, et al. Correlation between 18-fluorodeoxyglucose accumulation in large vessels and serological markers of inflammation in polymyalgia rheumatica: a quantitative PET study. Ann Rheum Dis 2004;63:870-3.

Hautzel H, Sander O, Heinzel M, Schneider M, Müller HW. Assessment of large-vessel involvement in giant cell arteritis with 18F-FDG PET: introducing an ROC-analysis-based cutoff ratio. J Nucl Med 2008;49:1107-13.

Tetzka D, Haraguchi T, Ishihara T, Ohigashi H, Inagaki H, Suzuki J, et al. Role of FDG PET-CT in Takayasu arteritis: sensitive detection of recurrences. JACC Cardiovasc Imaging 2012;5:422-9.

Besson FL, de Boysson H, Parienti JJ, Bouvard G, Bienvenu B, Agostini D. Towards an optimal semiquantitative approach in giant cell arteritis: an 18F-FDG PET/CT case-control study. Eur J Nucl Med Mol Imaging 2014;41:155-66.

Stellingwerff MD, Brouwer E, Lensen KJ, Rutgers A, Arends S, van der Geest KS, et al. Different scoring methods of FDG PET/CT in giant cell arteritis: need for standardization. Medicine (Baltimore) 2015;94:e1542.

Lee YH, Choi SJ, Ji JD, Song GC. Diagnostic accuracy of 18F-FDG PET or PET/CT for large vessel vasculitis: a meta-analysis. Z Rheumatol 2016;75:924-31.

Grayson PC, Alehashemi S, Bagheri AA, Vivelek AC, Cupps TR, Kaplan MJ, et al. 18F-fluorodeoxyglucose-positron emission tomography as an imaging biomarker in a prospective, longitudinal cohort of patients with large vessel vasculitis. Arthritis Rheumatol 2018;70:439-49.

Kim J, Oh MD. FDG PET-CT in the diagnosis of Takayasu arteritis presenting as fever of unknown origin: a case report. Infect Chemother 2015;47:190-3.

Yoo SJ, Lee JC, Kim Y, Yoo IS, Shim SC, Kim KH, et al. Clinical Implication of 18F-FDG PET in diagnosing and monitoring disease activity in a case of subclinical stage of giant cell arteritis. J Rheum Dis 2015;22:382-6.

de Boysson H, Dumont A, Liozon E, Lambert M, Boutemy J, Maingé G, et al. Giant-cell arteritis: concordance study between aortic CT angiography and FDG-PET/CT in detection of large-vessel involvement. Eur J Nucl Med Mol Imaging 2017;44:2274-9.

Incerti E, Bombetti E, Fallanca F, Baldissera EM, Alongi P, Tombolini E, et al. 18F-FDG PET reveals unique features of large vessel inflammation in patients with Takayasu’s arteritis. Eur J Nucl Med Mol Imaging 2017;44:1109-18.

Dellavedova L, Carletto M, Faggioni P, Sciascera A, De Sole A, MaZZone A, et al. The prognostic value of baseline and clinical factors of 18F-FDG PET/CT in steroid-naïve large-vessel vasculitis: introduction of volume-based parameters. Eur J Nucl Med Mol Imaging 2016;43:340-8.

de Leeuw K, Bijl M, Jager PL. Additional value of positron emission tomography in diagnosis and follow-up of patients with large vessel vasculitides. Clin Exp Rheumatol 2004;22(6 Suppl 36):S21-6.

Stenová E, Mistec S, Povinec P. FDG-PET/CT in large-vessel vasculitis: its diagnostic and follow-up role. Rheumatol Int 2010;30:1111-4.

Blockmans D, de Ceuninck L, Vanderschueren S, Knockaert D, Mortelmans L, Bobbaers H. Repetitive 18F-fluorodeoxyglucose positron emission tomography in giant cell arteritis: a prospective study of 35 patients. Arthritis Rheum 2006;55:131-7.
121. Muto G, Yamashita H, Takahashi Y, Miyata Y, Morooka M, Minamimoto R, et al. Large vessel vasculitis in elderly patients: early diagnosis and steroid-response evaluation with FDG-PET/CT and contrast-enhanced CT. Rheumatol Int 2014;34:1545-54.

122. Karunanithi S, Sharma P, Bal C, Kumar R. 18F-FDG PET/CT for diagnosis and treatment response evaluation in large vessel vasculitis. Eur J Nucl Med Mol Imaging 2014;41:586-7.

123. Park EH, Lee EY, Lee YJ, Ha YJ, Yoo WH, Choi BY, et al. Infliximab biosimilar CT-P13 therapy in patients with Takayasu arteritis with low dose of glucocorticoids: a prospective single-arm study. Rheumatol Int 2018;38:2233-42.

124. Banerjee S, Quinn KA, Gribbons KB, Rosenblum JS, Civelek AC, Novakovich E, et al. Effect of treatment on imaging, clinical, and serologic assessments of disease activity in large-vessel vasculitis. J Rheumatol 2020;47:99-107.

125. Choi YW, Jung S, Jung TY, Kim YH, Han DS, Bang SY. Fluorine-18-fluorodeoxyglucose positron emission tomography/computed tomography in diagnosis and assessment of takayasu arteritis and ulcerative colitis. J Rheum Dis 2017;24:55-9.

126. Pelletier-Galarneau M, Ruddy TD. PET/CT for diagnosis and management of large-vessel vasculitis. Curr Cardiol Rep 2019;21:34.

127. Blockmans D, De Ceuninck L, Vanderschueren S, Blockmans D, De Ceuninck L, Vanderschueren S, et al. Value of 18F-fluorodeoxyglucose positron emission tomography/computed tomography in patients with active polymyalgia rheumatica. Mod Rheumatol 2011;21:334-6.

128. Yamashita H, Kubota K, Takahashi Y, Minamimoto R, Morooka M, Kaneko H, et al. Clinical value of 18F-fluorodeoxyglucose positron emission tomography/computed tomodiagnosis of first onset Still’s disease: a seven-case series and review of the literature. Mod Rheumatol 2014;24:645-50.

129. Gerfaud-Valentin M, Maucott-Boulch D, Hot A, Iwaz J, Ninet J, Durieu I, et al. Adult-onset Still’s disease: manifestations, treatment, outcome, and prognostic factors in 57 patients. Medicine (Baltimore) 2014;93:91-9.

130. Prieto-Peña D, Martínez-Rodríguez I, Loricera J, Banzo I, Prieto-Peña D, Martínez-Rodríguez I, Loricera J, et al. Whole-body fluorodeoxyglucose positron emission tomography in diagnosis of Takayasu arteritis and ulcerative colitis. Semin Arthritis Rheum 2014;43:1624-34.

131. Schönau V, Vogel K, Englbrecht M, Wacker J, Schmidt D, Manger B, et al. The value of 18F-fluorodeoxyglucose positron emission tomography in the assessment of disease activity of adult-onset Still’s disease. Eur J Nucl Med Mol Imaging 2012;39:586-95.

132. Wallace ZS, Zhang Y, Perugino CA, Naden R, Choi HK, Stone JH, et al. Clinical phenotypes of IgG4-related disease: an analysis of two international cross-sectional cohorts. Ann Rheum Dis 2019;78:406-12.

133. Zhang J, Chen H, Ma Y, Xiao Y, Niu N, Lin W, et al. Characterizing IgG4-related disease with 18F-FDG PET/CT: a prospective cohort study. Eur J Nucl Med Mol Imaging 2014;41:1624-34.

134. Lee J, Hyun SH, Kim S, Kim DK, Lee JK, Moon SH, et al. Utility of FDG PET/CT for differential diagnosis of patients clinically suspected of IgG4-related disease. Clin Nucl Med 2016;41:237-43.

135. Takano K, Yajima R, Kubota K, Yamamoto M, Takahashi H, Yama N, et al. Clinical utility of 18F-fluorodeoxyglucose/positron emission tomography in diagnosis of immunoglobulin G4-related sclerosing sialadenitis.
151. Cheng MF, Guo YL, Yen RF, Chen YC, Ko CL, Tien YW, et al. Clinical utility of FDG PET/CT in patients with autoimmune pancreatitis: a case-control study. Sci Rep 2018; 8:3651.

152. Wang Y, Guan Z, Gao D, Luo G, Li K, Zhao Y, et al. The value of $^{18}$F-FDG PET/CT in the distinction between retroperitoneal fibrosis and its malignant mimics. Semin Arthritis Rheum 2018;47:593-600.

153. Zhao Z, Wang Y, Guan Z, Jin J, Huang F, Zhu J. Utility of FDG-PET/CT in the diagnosis of IgG4-related diseases. Clin Exp Rheumatol 2016;34:119-25.