Antecedent infections in Fisher syndrome: sources of variation in clinical characteristics

Michiaki Koga1 · Masahiko Kishi2,3 · Toshihiro Fukusako4 · Naomi Ikuta5 · Masayuki Kato6 · Takashi Kanda1

Received: 7 March 2019 / Revised: 1 April 2019 / Accepted: 1 April 2019 / Published online: 6 April 2019
© The Author(s) 2019

Abstract
The clinical features of Guillain–Barré syndrome (GBS) are highly variable, according to the type of antecedent infection. Although a major GBS phenotype, Fisher syndrome (FS), has been shown to be preceded by infections similar to those preceding GBS, whether or not the clinical features in FS also vary according to antecedent infection remains unclarified. Frequent antecedent infections among this study of 70 FS patients included Haemophilus influenzae \( [n = 15 \ (21\%)] \), Campylobacter jejuni \( [n = 10 \ (14\%)] \), and cytomegalovirus (CMV) \( [n = 6 \ (8.6\%)] \). Compared with other FS patients, H. influenzae-seropositive FS patients more frequently had a history of prior upper respiratory tract infection; double vision as the initial symptom; and, except for oculomotor disturbance, more rarely showed cranial nerve involvement. C. jejuni-related FS occurred predominantly in younger male patients and characteristically presented with blurred vision. According to GBS disability scale, CMV-related FS tended to be more severe, although every patient received immunotherapy. Serum anti-GQ1b IgG antibodies were detected in most cases, regardless of antecedent infection type. At the nadir of illness, the most frequent diagnosis in H. influenzae-related cases was “pure FS” without limb weakness or central nervous system involvement (71%), in C. jejuni-related cases “incomplete FS” such as acute ophthalmoparesis with or without ataxia (60%), and in CMV-related cases (50%) advanced conditions such as GBS overlap and Bickerstaff brainstem encephalitis. These findings indicate that the type of preceding infection determined the neurological features of FS. CMV-related FS appeared to be similar to H. influenzae- and C. jejuni-related FS regarding anti-GQ1b antibody-mediated pathogenesis, as opposed to CMV-related GBS.

Keywords Fisher syndrome · Antecedent infection · Haemophilus influenzae · Campylobacter jejuni · Cytomegalovirus · Intercellular adhesion molecule 1

Introduction
Guillain–Barré syndrome (GBS) is a postinfectious autoimmune-mediated neuropathy, the clinical manifestations of which greatly vary among individual cases [1]. An at least partial spectrum of antecedent infections has been determined, which indicates that Campylobacter jejuni is the most frequent causal agent of GBS [2]. To clarify the pathogenesis of GBS, classifying GBS into subgroups based on the type of antecedent infection, such as “C. jejuni-associated GBS” and “cytomegalovirus (CMV)-associated GBS” could be important, because antecedent infections are closely associated with neurological phenotypes and specific autoantibodies [3]. For example, GBS patients with CMV infection showed higher blood levels of soluble intercellular adhesion molecule 1 (sICAM-1) than the levels in other GBS patients, which suggests that a cytotoxic mechanism is involved in the development of CMV-associated GBS [4].
Fisher syndrome (FS) is a major clinical phenotype of GBS. It is characterized neurologically by the triad of ophthalmoplegia, ataxia, and areflexia and immunologically by the frequent presence of anti-GQ1b IgG autoantibodies [5]. Our previous case-controlled serological study showed that *C. jejuni* and *Haemophilus influenzae* were the major identified agents of antecedent infection in FS [6]. However, unlike for GBS, close associations of clinical features with types of antecedent infections are yet to be clarified for FS, perhaps because of stereotypical consideration with regard to a relatively uniform clinical picture and presence of specific autoantibodies (anti-GQ1b antibodies), and also of limitations of our previous study which did not include cases of incomplete forms of FS, such as acute ophthalmoplegia without ataxia, and of advanced FS that included Bickerstaff brainstem encephalitis (BBE) [6]. The aim of this study was to clarify whether or not the clinical and laboratory features of FS vary as they do in GBS, according to antecedent infections.

### Methods

#### Study participants

This study consisted of a cohort of 70 patients with FS, all of whom had undergone a medical examination or were referred to Yamaguchi University Hospital for assessment of the presence of serum anti-glycolipid antibodies between 2001 and 2017. The clinical criteria for enrollment have been reported previously [7]. However, unlike our previous studies [6, 7], this study included patients with FS in the broad sense to recruit a wide range of patients with FS-related conditions as follows: without ataxia or areflexia or both (namely “incomplete FS”) and those who initially presented with the triad of manifestations characteristic of FS and then developed generalized muscle weakness (namely, FS/GBS overlap) or disturbance in consciousness (final diagnosis being BBE) or both. To collect clinical data from cases not our own, we requested information on other relevant patients from their primary physicians, which included neurological findings and GBS disability scale (GDS) [8] at the nadir of illness. For the analysis of sICAM-1 level, serum samples from 31 patients with GBS were also used in addition to those from FS patients. The study protocol was approved by the Institutional Ethics Committee at Yamaguchi University Hospital.

#### Serological analysis

Serum IgM and IgG antibodies against isolated gangliosides (GM2, GM1, GD1a, GalNAc-GD1a, GD1b, GT1a, GT1b, or GQ1b; 10 pmol/well) and ganglioside complexes (GM1/GD1a and GM1/GT1a) were measured by enzyme-linked immunosorbent assay (ELISA) as described elsewhere [9]. IgG subclasses of anti-GQ1b and anti-GT1a antibodies were examined as reported previously [10]. Antecedent *C. jejuni* and *H. influenzae* infections were investigated by an in-house ELISA method, as reported elsewhere [6]. CMV, Epstein–Barr virus, and *Mycoplasma pneumoniae* infections were serologically identified using commercially available kits, as described elsewhere [6]. Serum level of sICAM-1 was measured using a commercially available ELISA kit (R&D Systems, Inc., Minneapolis, USA).

### Data analysis

Differences in frequencies between groups were analyzed using the Fisher exact test. Differences in medians were examined by the Mann–Whitney *U* test. Differences were considered significant for *P* < 0.05 in two-sided tests. Statistical calculations were performed by SPSS 19 software (IBM Japan, Tokyo, Japan).

### Results

#### Serology and symptoms of antecedent infections

Among 70 patients with FS, serological evidence of recent infection with *H. influenzae*, *C. jejuni*, CMV, and *M. pneumoniae* was seen in 15 (21%), 10 (14%), 6 (8.6%), and 1 (1.4%) patient, respectively. One patient showed seropositivity for both *H. influenzae* and CMV. No agent of antecedent infection was identified in 39 (56%) patients. Patients with *H. influenzae* or *C. jejuni* infections were younger [median age, 43 year (*P* = 0.06) and 41 year (*P* = 0.02), respectively] than the other patients. The male–female ratio of patients with *C. jejuni* infections was disproportionally male (8:2) compared to the gender ratio of the other patients (27:33, *P* = 0.04) (Table 1).

Any symptom of infection preceded the onset of neurological manifestations in 66 (94%) of 70 patients. Respiratory tract symptoms were the most frequent in patients with *H. influenzae* infection [15 of 15 (100%)], which was a significantly higher proportion of patients than the proportion of other FS patients with respiratory tract symptoms (62%, *P* = 0.002). In contrast, gastrointestinal symptoms were the most frequent in patients with *C. jejuni* infection [8 of 10 (80%)], which was a significantly higher proportion of patients than the proportion of other FS patients with gastrointestinal symptoms (20%, *P* = 0.0004). Among the six patients with CMV seropositivity, four (67%) had a history of prior respiratory tract symptoms, one (17%) had gastrointestinal symptoms, and one (17%) did not have any preceding symptom of infection.
The most frequent initial neurological symptom was double vision, regardless of antecedent infection, but this manifestation was pronounced in patients with H. influenzae (87%) compared to patients with C. jejuni (40%) and CMV (50%) infections. It is noteworthy that three (30%) of ten patients with C. jejuni infections, in whom obvious mydriasis was observed at admission, initially presented with blurred vision without double vision or ptosis. Neurological findings at admission (or first visit to the neurology department by patients who had not been hospitalized) are shown in Table 1. Lack of cranial nerve palsy except for ophthalmoplegia was characteristic for patients with H. influenzae infection. Frequent occurrence of sensory disturbance was characteristic for patients with CMV infection, although the difference was not statistically significant (compared to the others: \( P = 0.09 \)). The differences between GDS of the patients with each type of antecedent infection at admission were not remarkable.

### Immunotherapy, final diagnosis, and disability scores at nadir (Table 2)

As immunotherapy, intravenous immunoglobulin therapy was administered to most patients. The median times from the onset to the nadir of illness for patients with H. influenzae or C. jejuni infections were both 6 days, whereas it tended to be longer for those with CMV infection (9 days). The neurological findings at the nadir varied, depending on the type of antecedent infection: patients with H. influenzae infection presented with the FS triad only, and 50% of patients with CMV infection had bulbar palsy. Variation in neurological disturbances was well reflected by the final diagnoses: “pure FS” (FS triad without limb weakness or disturbance of the central nervous system disturbance) in 71% of patients with H. influenzae infection, “incomplete FS” (acute ophthalmoparesis without ataxia and/or a-/hyporeflexia) in 60% of C. jejuni patients, and FS/GBS overlap or BBE in 50% of CMV patients.

Regarding GDS and duration of hospitalization, the patients with H. influenzae and C. jejuni infections were

---

**Table 1 Neurological findings at admission**

|                | Total | H. influenzae | C. jejuni | CMV | None identified |
|----------------|-------|---------------|-----------|-----|-----------------|
| Number         | 70    | 15*           | 10        | 6*  | 39              |
| Age, median (range) | 55 (2–76) | 43 (2–69) | 41 (12–58)* | 49 (23–65) | 60 (9–76)** |
| Gender, male/female | 35/35 | 7/8          | 8/2**** | 2/4 | 19/20          |
| Admission, days after onseta | 4 (1–28) | 3 (1–11) | 4 (2–11) | 3 (1–11) | 4 (1–28) |
| Neurological findings at admissionb | | | | |
| Abducens-dominancy in OP | 32/52 (62%) | 9/13 (69%) | 4/7 (57%) | 3/4 (75%) | 17/29 (59%) |
| Internal OP | 35/69 (51%) | 7 (47%) | 7 (70%) | 5 (83%) | 16/39 (41%) |
| Facial palsy | 14/66 (21%) | 0/13**** | 3 (30%) | 1 (17%) | 9/38 (24%) |
| Bulbar palsy | 15/68 (22%) | 1/13 (7.7%) | 2 (20%) | 3 (50%) | 8 (21%) |
| Cranial nerve palsy except for OP | 22/66 (33%) | 1/13 (7.7%)* | 4 (67%) | 3 (50%) | 13/37 (35%) |
| Neck weakness | 6/59 (10%) | 0/13 | 2/8 (25%) | 1 (17%) | 3/33 (9.1%) |
| Limb weakness | 7 (10%) | 0 | 1 (10%) | 1 (17%) | 4 (10%) |
| Ataxia | 58/69 (84%) | 14 (93%) | 6 (60%) | 6 (100%) | 33/38 (87%) |
| Sensory disturbance | 22/65 (34%) | 5/14 (36%) | 3 (30%) | 4 (67%) | 9/35 (26%) |
| Autonomic disturbance | 4/60 (6.7%) | 1/12 (8.3%) | 0/8 | 0 | 3/34 (8.8%) |
| GBS disability scale, median (range) | 2 (1–5) | 2 (1–4) | 2 (1–4) | 3 (2–4) | 2 (1–4) |

CMV cytomegalovirus, OP ophthalmoplegia, GBS Guillain–Barré syndrome

*Lower than others (\( P = 0.02 \))
**Higher than others (\( P = 0.0004 \))
****Larger proportion of males than others (\( P = 0.04 \))
*****Rarer than others (\( P = 0.03 \))
******Rarer than others (\( P = 0.03 \))

*a Including one patient who was double positive for H. influenzae and CMV

*b First visit to neurology department in patients who had not been hospitalized
similar (median GDS, both 3; median hospitalization duration, 15 and 17 days, respectively). These data were in contrast with those with CMV infection, in whom GDS tended to be higher (4 in all cases) and the duration of hospitalization longer (median 24 days: more than 2 weeks in all). Patients without an identified antecedent infection were positioned between the patients with *H. influenzae* / *C. jejuni* infections and the patients with CMV infection with respect to time from onset to nadir of illness, neurological findings at the nadir, and duration of hospitalization.

**Laboratory data**

IgG antibodies against GQ1b and GT1a were detected in most cases, regardless of antecedent infection, and the pattern of other anti-ganglioside IgG antibodies did not considerably differ between the groups (Table 3). However, the IgG subclasses of anti-GQ1b antibodies were unevenly distributed, as follows: patients with *H. influenzae* and *C. jejuni* infections most often showed IgG1 anti-GQ1b seropositivity, whereas those with CMV infection only showed IgG3 anti-GQ1b seropositivity. Anti-GM2 IgM antibodies were negative in all patients. Differences in the results of cerebrospinal fluid analysis between the groups were not significant.

As in the previous study [4], the serum concentration of sICAM-1 tended to be higher in patients with CMV-associated GBS than those with other antecedent infection-associated GBS (Fig. 1). In contrast, no apparent increase of sICAM-1 concentration was seen in CMV-associated FS.

**Discussion**

Because of the frequent detection of anti-GQ1b antibodies, FS tends to be considered as a relatively uniform clinical entity, and there have been few studies regarding variability in the clinical picture of FS. This study has clearly shown that FS has a markedly variable clinical picture, and that three antecedent infections (*H. influenzae*, *C. jejuni*, and CMV) are closely associated not only with antecedent symptoms of infection, but also with neurological features and disability scores, as follows: (1) *H. influenzae*-related FS

---

**Table 2** Immunotherapies and clinical features at nadir

|                     | Total          | *H. influenzae* | *C. jejuni* | CMV | None identified |
|---------------------|----------------|-----------------|-------------|-----|----------------|
| **Number**          | 70             | 15              | 10          | 6   | 39             |
| **Immunotherapies** |                |                 |             |     |                |
| Intravenous         | 52/69 (75%)    | 10/14 (71%)     | 8 (80%)     | 6 (100%) | 28 (72%) |
| Steroid pulse       | 2/69 (2.9%)    | 0/14            | 0           | 0   | 2 (5.1%)     |
| Duration from onset | 6 (2–30)       | 6 (3–11)        | 6 (3–11)    | 9 (3–14) | 8 (2–30) |
| > 7 days            | 29/68 (43%)    | 2/13 (15%)*     | 2 (20%)     | 4 (67%) | 20 (51%)     |
| **Neurological**    |                |                 |             |     |                |
| findings at nadir   |                |                 |             |     |                |
| Conscious disturbance| 2/66 (3.0%)    | 0/14            | 1/9 (11%)   | 1 (17%) | 0/38         |
| Bulbar palsy        | 15/66 (23%)    | 1/14 (7.1%)     | 3/9 (33%)   | 3 (50%) | 8/38 (21%) |
| Limb weakness       | 13/66 (20%)    | 2/14 (14%)      | 1/9 (11%)   | 2 (33%) | 8/38 (21%) |
| Artificially         | 2/67 (3.0%)    | 0/14            | 0/9         | 0   | 1/38 (2.6%) |
| **Diagnosis at nadir** |             |                 |             |     |                |
| “Pure FS”**         | 39/69 (57%)    | 10/14 (71%)     | 2 (20%)**   | 3 (50%) | 25 (64%)     |
| “Incomplete FS” (AO ± ataxia) | 12/69 (17%) | 2/14 (14%) | 6 (60%)*** | 0 | 4 (10%) |
| FS/GBS overlap or BBE | 18/69 (26%)  | 2/14 (14%)      | 2 (20%)     | 3 (50%) | 10 (26%)    |
| GDS at nadir, median | 3 (1–5)       | 3 (1–4)         | 3 (1–4)     | 4 (3–4) | 2 (1–5)     |
| GDS ≥ 3             | 36/67 (54%)    | 7/13 (54%)      | 5 (50%)     | 6 (100%)**** | 18/38 (47%) |
| Duration of hospitalization, median (range) | 20 (2–57) | 15 (5–36) | 17 (6–48) | 24 (15–36) | 20 (2–57) |

CMV cytomegalovirus, FS Fisher syndrome, AO acute ophthalmoparesis, GBS Guillain–Barré syndrome, BBE Bickerstaff brainstem encephalitis, GDS GBS disability scale

* Rarer than others (*P* = 0.02)
** Rarer than others (*P* = 0.01)
*** More frequent than others (*P* = 0.001)
**** More frequent than others (*P* = 0.02)

*Having the triad of ophthalmoplegia, ataxia, and a/hyporeflexia without limb weakness and central nervous system disturbance*
occurred after upper respiratory tract symptoms, patients presented with double vision as an initial symptom, showed “pure FS” at the nadir of illness without cranial nerve involvement other than oculomotor disturbance and limb weakness, and showed mild disability according to the GDS; (2) C. jejuni-related FS occurred predominantly in younger male patients after gastrointestinal symptoms, remained “incomplete FS” with manifestations such as acute ophthalmoparesis with mild disability, and sometimes was characterized by an exceptional initial symptom of blurred vision; and (3) CMV-related FS tended to manifest bulbar palsy and sensory disturbance, and to advance to FS/GBS overlap or BBE, with severe disability at the nadir of illness despite immunotherapy.
A large cohort study that included 194 patients with anti-GQ1b IgG antibodies showed that various phenotypes, including acute ophthalmoparesis, FS, BBE, and their manifestations overlapping with GBS, formed a distinct disease entity, namely “anti-GQ1b IgG antibody syndrome” [11]. However, that study did not clarify why such clinical variation occurs, except for limb weakness, which was partially associated with common existence of anti-GM1 or anti-GD1a antibodies in addition to anti-GQ1b antibodies. To the best of our knowledge, our study is the first to identify an antecedent infection as a causative factor in determining the subtypes of “IgG anti-GQ1b antibody syndrome”.

It remains unclear why neurological variation occurs in FS despite the common occurrence of anti-GQ1b IgG antibodies. A possible explanation might be the cross-reactivity of anti-GQ1b antibodies with ganglioside complexes. Depending on antibody specificities, anti-GQ1b antibodies are divided into three types: GQ1b-specific, GM1/GQ1b-reactive, and GD1a/GQ1b-reactive [12], and such diverse specificities might lead to clinical differences between FS patients [13]. In our simple analysis that only assessed two anti-ganglioside-complex antibodies (anti-GM1/GT1a and anti-GM1/GD1a), we failed to find differences in antibody specificities according to the antecedent infection. Additional detailed analyses of a large number of anti-ganglioside complex antibodies are needed to evaluate the theory that diverse antibody specificities might lead to clinical differences between FS patients.

Among the three identified subtypes of FS, H. influenzae- and C. jejuni-related FS showed several similar characteristics, including onset of illness in younger patients, short period of time between onset to the nadir of illness (median 6 days), neurological deficits limited to the triad of FS with mild disability at the nadir, and IgG1 subclass predominance of anti-GQ1b antibodies. In contrast, CMV-related FS manifested different characteristics, including longer period of time between onset to nadir of illness (median 9 days); neurological deficits beyond the triad of FS, including sensory disturbance and limb weakness; increased severity in disability at the nadir of illness, and IgG3 subclass predominance of anti-GQ1b antibodies. These clinical features of CMV-related FS are similar to those of CMV-related GBS, which was characterized by longer period of time from onset to nadir of illness (median 10 days) compared to C. jejuni-related GBS (median 7 days), frequent cranial nerve palsy and sensory disturbance, and severe disability [14–16]. Although anti-GM2 IgM antibodies and anti-moesin IgG antibodies have been proposed to be the autoantibodies associated with CMV-related GBS [17–19], findings casting doubt on their contributory roles in the pathogenesis of GBS have also been reported [20, 21], and therefore a role for pathogenic autoantibodies remains unclear in CMV-related GBS. This issue, as well as the finding of elevated concentrations of serum sICAM-1 in patients with CMV-related GBS [4], indicate the possible role of a cytotoxic mechanism rather than a humoral mechanism in the development of CMV-related GBS. Therefore, we had at first hypothesized that anti-GQ1b IgG antibodies would not be present in CMV-related FS. However, this study found that anti-GQ1b antibodies were as frequent in patients with CMV-related FS as they were in patients with FS preceded by other infections, and that serum sICAM-1 concentration was not elevated in patients with CMV-related FS in contrast to patients with CMV-related GBS. We concluded that a humoral mechanism could be a causative factor in CMV-related FS as well.

We previously reported that the presence of IgG1 subclass of antibodies against GM1 and other motor gangliosides was closely associated with poor recovery in GBS patients [10, 22]. In the current study, information on disability scale during recovery phase was not available, and therefore it remains unclear whether IgG1 subclass of anti-GQ1b antibodies is also associated with poor recovery in FS. We instead compared the duration of hospitalization between the groups and found that patients with IgG1 anti-GQ1b antibodies tended to need rather shorter hospitalization [median 17 (5–55) days] than those without IgG1 antibodies [20 (8–57) days: \( P = 0.11 \)]. This suggests that IgG1 subclass of anti-GQ1b antibodies is not an indicator for poor recovery in FS, but further investigation should be necessary regarding this matter.

This study has limitations. First, this study included a relatively small number of FS patients \( (N = 70) \), only six patients with CMV-related FS were analyzed, and statistical significance could not be confirmed for several situations. Second, inaccurate clinical data and selection bias cannot be completely denied, because the data were derived from a multicenter cohort. To reduce the possible problems, we contacted the involved primary physicians, and without divulging information on the serology on antecedent infections, asked them to answer a questionnaire as well as to provide a summary on their patients. Finally, no antecedent infectious agent was identified in more than a half of FS patients in our study, and a wider range of possible antecedent infections should be investigated. Two case–control studies from Bangladesh and The Netherlands reported a significant association for hepatitis E virus (HEV) infection with GBS [23, 24]. One FS patient positive for anti-HEV IgM antibodies has also been identified among six Japanese FS patients [25]; however, our preliminary study did not find a patient with serological evidence of recent HEV infection among 65 Japanese patients with FS (Koga, unpublished data).

In conclusion, our data strongly suggest that the type of preceding infection determines the neurological features of FS. In clinical practice, cases of CMV-FS especially should...
be actively treated, because they tended to progress to severe disability. Moreover, CMV-related FS appeared to be similar to *H. influenzae-* and *C. jejuni-*related FS regarding anti-GQ1b antibody-mediated pathogenesis as opposed to CMV-related GBS.

**Acknowledgements** We acknowledge Dr. Ohori (Japanese Red Cross Yamaguchi Hospital, Japan), Dr. Tanaka and Dr. Sano (NHO Kanmon Medical Center, Japan), Dr. Abe (JCHO Tokuyama Central Hospital, Japan), and Dr. Kanai (NHO Takasaki Medical Center, Japan) for providing us with their patients’ clinical data. Dr. Koga received a Grant-in-Aid for Scientific Research (C) (KAKENHI 18K07115) from the Ministry of Education, Culture, Sports, Science and Technology of Japan. Prof. Kanda received a Research Grant for Neuroimmunological Diseases from the Ministry of Health, Labour and Welfare of Japan.

**Author contributions** MKo: design of study, collection, analysis and interpretation of the data, drafting of original manuscript. MKi: collection and interpretation of the data, revision of the manuscript. TF: collection of the data, revision of the manuscript. NE: collection of the data, revision of the manuscript. NKa: collection of the data, revision of the manuscript. TK: supervision of the study, interpretation of the data, revision of the manuscript.

**Compliance with ethical standards**

**Conflicts of interest** None of the authors reports any disclosures.

**Ethical standards** This study was approved by the Institutional Review Board of Yamaguchi University Hospital.

**Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

**References**

1. Yuki N, Hartung HP (2012) Guillain–Barré syndrome. N Engl J Med 366:2294–2304
2. Jacobs BC, Rothbarth PH, van der Meché FG, Hadden RD, Karch H, Hartung HP, Zielasek J, Weissbrich B, Cornblath DR, Swan AV, Hughes RA, Schubert J, Weishaupt A, Toyka KV, Plasma Exchange/Sandoglobulin Guillain–Barré Syndrome Trial Group (2001) Preceding infections, immune factors, and outcome in Guillain–Barré syndrome. Neurology 56:758–765
3. Chiba A (2016) Miller Fisher syndrome. In Willison HJ, Goodfellow JA (eds) GBS100: celebrating a century of progress in Guillain–Barré syndrome and new serological markers. J Neurol 259:1366–1374
4. Hughes RA, Newsom-Davis JM, Perkin GD, Pierce JM (1978) Controlled trial prednisolone in acute polyneuropathy. Lancet 2(8093):750–753
5. Koga M, Takahashi M, Yokoyama K, Kanda T (2015) Ambiguous value of anti-ganglioside IgM autoantibodies in Guillain–Barré syndrome and its variants. J Neurol 262:1954–1960
6. Koga M, Yuki N, Hirata K, Morimatsu M, Mori M, Kusunoki S (2003) Anti-GM1 antibody IgG subclass: a clinical recovery predictor in Guillain–Barré syndrome. Neurology 60:1514–1518
7. Koga M, Gilbert M, Takahashi M, Li J, Hirata K, Kanda T, Yuki N (2012) GQ1b-seronegative Fisher syndrome: clinical features and new serological markers. J Neurol 259:1366–1374
8. Hughes RA, Newsom-Davis JM, Perkin GD, Pierce JM (1978) Controlled trial prednisolone in acute polyneuropathy. Lancet 2(8093):750–753
9. Koga M, Takahashi M, Yokoyama K, Kanda T (2015) Ambiguous value of anti-ganglioside IgM autoantibodies in Guillain–Barré syndrome and its variants. J Neurol 262:1954–1960
10. Koga M, Yuki N, Hirata K, Morimatsu M, Mori M, Kusunoki S (2003) Anti-GM1 antibody IgG subclass: a clinical recovery predictor in Guillain–Barré syndrome. Neurology 60:1514–1518
11. Okada M, Yuki N, Hirata K (2001) Anti-GQ1b IgG antibody syndrome: clinical and immunological range. J Neurol Neurosurg Psychiatry 70:50–55
12. Kaida K, Kanzaki M, Morita D, Kamakura M, Motoyoshi K, Hirakawa M, Kusunoki S (2006) Anti-ganglioside complex antibodies in Miller Fisher syndrome. J Neurol Neurosurg Psychiatry 77:1043–1146
13. Kanzaki M, Kaida K, Ueda M, Morita D, Hirakawa M, Motoyoshi K, Kamakura K, Kusunoki S (2008) Ganglioside complexes containing GQ1b as targets in Miller Fisher and Guillain–Barré syndromes. J Neurol Neurosurg Psychiatry 79:1148–1152
14. Visser LH, van der Meché FG, Meulstee J, Rothbarth PP, Jacobs BC, Schmitz PI, van Doorn PA (1996) Cytomegalovirus infection and Guillain–Barré syndrome: the clinical, electrophysiologic, and prognostic features. Dutch Guillain–Barré Study Group. Neurology 47:668–673
15. Orlikowski D, Porcher R, Sivadon-Tardy V, Quincampoix JC, Raphaël JC, Durand MC, Sharrar T, Roussi J, Caudie A, Annane D, Rozenberg F, Leruez-Ville M, Gaillard JL, Gault E (2011) Guillain–Barré syndrome following primary cytomegalovirus infection: a prospective cohort study. Clin Infect Dis 52:837–844
16. Caudie A, Quitard Pinon A, Taravel D, Sivadon-Tardy V, Orlikowski D, Rozenberg F, Sharrar T, Raphaël JC, Gaillard JL (2011) Preceding infections and anti-ganglioside antibody profiles assessed by a dot immunoassay in 306 French Guillain–Barré syndrome patients. J Neurol 258:1958–1964
17. Irie S, Saito T, Nakamura K, Kanazawa N, Ogino M, Nakazawa T, Ito H, Tanai Y, Kowa H (1996) Association of anti-GM2 antibodies in Guillain–Barré syndrome with acute cytomegalovirus infection. J Neuromuscular Musculoskelet Disord 68:19–26
18. Jacobs BC, van Doorn PA, Groeneveld JH, Tio-Gillen AP, van der Meché FG (1997) Cytomegalovirus infections and anti-GM2 antibodies in Guillain–Barré syndrome. J Neurol Neurosurg Psychiatry 62:641–643
19. Sawai S, Sato M, Mori M, Misawa S, Sogawa K, Kazami T, Ishibashi M, Beppu M, Shibuya K, Ishitake T, Sekiguchi Y, Nomura K, Sato K, Matsuhashita K, Kodera Y, Nomura F, Kawai N (2014) Moesin is a possible target molecule for cytomegalovirus-related Guillain–Barré syndrome. Neurology 83:113–117
20. Yuki N, Tagawa Y (1998) Acute cytomegalovirus infection and IgM anti-GM2 antibody. J Neurol Sci 154:14–17
21. Miyaji K, Devaux J, Yuki N (2014) Moesin is a possible target molecule for cytomegalovirus-related Guillain–Barré syndrome. J Neurol 261:1946–1951
22. Jacobs BC, Koga M, van Rijs W, Geleijns J, van Doorn PA, Willison HJ, Yuki N (2008) Subclass IgG to motor gangliosides related to infection and clinical course in Guillain–Barré syndrome. J Neuromuscular Disord 19:181–190
23. Geurtvanskessel CH, Islam Z, Mohammad QD, Jacobs BC, Endtz HP, Osterhaus AD (2013) Hepatitis E and Guillain–Barré syndrome. Clin Infect Dis 57:1369–1370
24. van den Berg B, van der Eijk AA, Pas SD, Hunter JG, Madden RG, Tio-Gillen AP, Dalton HR, Jacobs BC (2014) Guillain–Barré syndrome associated with preceding hepatitis E virus infection. Neurology 82:491–497

25. Fukae J, Tsugawa J, Ouma S, Umez T, Kusunoki S, Tsuboi Y (2016) Guillain–Barré and Miller Fisher syndromes in patients with anti-hepatitis E virus antibody: a hospital-based survey in Japan. Neurol Sci 37:1849–1851.