LncRNA SNHG20 promotes cell proliferation and invasion by suppressing miR-217 in ovarian cancer

Xuefeng Xing¹ · Ming An¹ · Tonghua Chen¹

Abstract
Background Ovarian cancer is the most common female gynecological malignancy. SNHG20, as a long non-coding RNA, has been proven to be an important regulator in the occurrence and development of various tumors. However, the potential mechanism of SNHG20 in ovarian cancer is unclear.

Objective The present study was aimed to investigate the functions and mechanisms of SNHG20 in ovarian cancer.

Methods The expression of SNHG20 and miR-217 in ovarian cancer tissues and cell lines was detected by qRT-PCR. CCK-8 assay was used to measure cell proliferation in transfected cells. The transwell assay was used to detect the relative invasion rate of transfected cells. The putative binding sites between SNHG20 and miR-217 were predicted by software LncBase v.2, and the interaction between SNHG20 and miR-217 was confirmed by dual-luciferase reporter assays and RIP assay. The rescue experiments were used to illustrate potential mechanisms.

Results SNHG20 was upregulated in ovarian cancer tissues and cell lines. Overexpression of SNHG20 promoted ovarian cancer cell proliferation and invasion. MiR-217 was downregulated in ovarian cancer tissues and cells, and was negatively regulated by SNHG20. Moreover, miR-217 overexpression inhibited ovarian cancer cell proliferation and invasion. Furthermore, miR-217 mimic reversed the inhibitory effect of SNHG20 overexpression on the biological behavior of ovarian cancer cells.

Conclusions SNHG20 promoted cell proliferation and invasion by sponging miR-217 in ovarian cancer. These results suggested that SNHG20 and miR-217 might provide new targets for therapeutic application in ovarian cancer.

Keywords Ovarian cancer · LncRNA SNHG20 · MiR-217 · Proliferation · Invasion

Introduction
Ovarian cancer is one of the most common female gynecologic malignancy and is the fifth leading cause of cancer-related death in women (Webb et al. 2017; Siegel et al. 2017). The overall prognosis of patients with ovarian cancer has improved due to dramatic improvements in diagnosis and therapeutic approaches (Holmes 2015). However, most patients with ovarian cancer are already in the advanced stage when they are diagnosed (Chien and Poole 2017). Advanced ovarian cancer is prone to drug resistance, recurrence, and metastasis, which leads to a 5-year survival rate of less than 40% (Allemani et al. 2015; Matz et al. 2017; Sankaranarayanan and Ferlay 2006). Thus, it is necessary to further explore the molecular mechanisms of the occurrence and development of ovarian cancer and to develop new treatment strategies.

A growing number of studies suggested that non-coding protein sequences in the genome were involved in the occurrence and metastasis of cancer, and were expected to become cancer therapeutic targets (Panoutsopoulou et al. 2018; Yoshida and Kimura 2017). Long non-coding RNA (lncRNA) is a type of non-coding RNA longer than 200 nucleotides, which plays an important regulatory role in the occurrence and development of tumors (Bill et al. 2019; Huarte 2015). However, the functions of lncRNA in tumors are complex, and many of them are unknown. Currently, the lncRNA-microRNAs (miRNAs) interaction network is often used as a research direction (Wang et al. 2015). Previous researches have indicated that some lncRNAs were dysregulated in ovarian cancer and played vital roles...
in tumorigenesis and progression, such as MAIT (Zhou et al. 2020), PVT1 (Chen et al. 2018), HOTAIR (Chang et al. 2018), and SNHG17 (Pan et al. 2020). The long non-coding RNA small nucleolar RNA host gene 20 (SNHG20) is located on 17q25.2, which has been reported to act as an oncogene in various tumors (Zhao et al. 2019), such as prostate cancer (Wu et al. 2019), gastric cancer (Yu et al. 2019), bladder cancer (Zhao et al. 2018), and lung cancer (Chen et al. 2017). Further researches have shown that SNHG20 knockdown inhibited ovarian cancer cell proliferation, migration, invasion, epithelial-mesenchymal transition and promoted cell apoptosis (He et al. 2018; Wang et al. 2019). However, the potential molecular mechanism of SNHG20 in ovarian cancer is unclear.

MicroRNA (miRNA) is a short non-coding RNA with 18–22 nucleotides in length (Tristán-Ramos et al. 2020), which always regulates gene expression through targeting mRNAs. Mounting evidence suggested that MiR-217 functions as a tumor suppressor. It has been reported that miR-217 could suppress cell proliferation, migration, and invasion in non-small cell lung cancer by regulating SIRT1 and P53/KAI1 signaling (Jiang et al. 2020). In gastric cancer, miR-217 inhibited epithelial-to-mesenchymal transition through targeting PTPN14 (Chen et al. 2020). A study revealed that miR-217 suppressed the migration and invasion of HeLa cells through modulating MAPK1 (Zhu et al. 2019). In ovarian cancer, miR-217 acted as a tumor suppressor role by targeting IGF1R (Li et al. 2016). However, further studies on the role of miR-217 in ovarian cancer are needed, and the link between SNHG20 and miR-217 is still unclear.

LncRNA can interact with miRNA and act as a competitive endogenous RNA (ceRNA) to regulate the expression of target genes, thus playing an important role in the occurrence and development of tumors (Bartel 2009). In this study, we aimed to explore the expression pattern, role, and potential functional mechanism of SNHG20 in ovarian cancer. Then, we further studied the relationship between SNHG20 and miR-217 in the proliferation and invasion of ovarian cancer cells. We also hope to find novel theoretical targets for the treatment of ovarian cancer.

Materials and methods

Clinical sample selection

A total of 30 surgically resected ovarian cancer tissues and adjacent normal tissues were collected at the Sanya People’s Hospital from 2017 to 2019. The characteristics of 30 patients with ovarian cancer in this study are presented in Table S1. All clinical samples were immediately snap-frozen in liquid nitrogen and then stored at 80 °C until RNA isolation. Besides, the research was approved by the Ethics Committee of the Sanya People’s Hospital (Approval No. 2017. 138).

Cell culture

Ovarian cancer cell lines (SKOV3 and A2780) and normal human ovarian surface epithelial cell line (HOSEpiC) were obtained from American Type Culture Collection (ATCC, Manassas, VA, USA). All cells were cultured in RPMI-1640 medium (Gibco, Thermo Fisher Scientific) supplemented with 10% fetal bovine serum (FBS, Gibco; Thermo Fisher Scientific), and incubated in a humidified atmosphere at 37 °C with 5% CO2.

Cell transfection

SNHG20 small interference RNA (si-SNHG20) or its negative control (si-NC), miR-217 mimics and inhibitor (miR-217 and anti-miR-217) or its control (miR-NC and anti-miR-217) were synthesized by GenePharma Co., Ltd (Shanghai, China). The overexpression vector pcDNA 3.1 was acquired from Thermo Fisher Scientific. The above oligonucleotides and SNHG20 overexpression plasmid (pcDNA-SNHG20) or its negative control (pcDNA) were transfected into ovarian cancer cell lines (SKOV3 and A2780) using Lipofectamine™ 3000 transfection reagent (Invitrogen, Carlsbad, CA, USA), according to the manufacturer’s suggested protocols.

Total RNA extraction and quantitative real-time polymerase chain reaction (qRT-PCR)

Total RNA was extracted from human ovarian cancer tissues and cell lines using Trizol reagent (Invitrogen, Thermo Fisher Scientific, Inc.). Reverse transcription was performed to acquire complementary DNA using the PrimeScript RT reagent Kit (Takara, Tokyo, Japan). Real-time PCR was performed using SYBR® Premix Ex Taq™II Kit (Takara, Tokyo, Japan) by an ABI 7500 real-time PCR system (Applied Biosystem, MA, USA). All primers were designed and synthesized by Sangon Biotech Co., Ltd. (Sangon, Shanghai, China). GAPDH (glyceraldehyde-3-phosphate dehydrogenase) and U6 were used as internal controls for SNHG20 and miR-217, respectively. The primer sequences were listed as follows: SNHG20 forward, 5′-ATGGCTTATA AATAGATACGCC-3′ and reverse, 5′-GGTCAAAACAGG GAGGGA-3′; miR-217 forward, 5′-CGCCTACTGCAT CAGGAACGT-3′ and reverse, 5′-CGGCCAGTGTTCA CACT-3′; GAPDH forward, 5′-GACCAGTCCATGC CATCAC-3′ and reverse, 5′-ACGCTCTTCACACTAC CTT-3′; U6 forward, 5′-CGCTTCCCGCAGCACATA TAC-3′ and reverse, 5′-ATGGAACGCTTCCAGATTT GC-3′.
The 2−ΔΔCt method was used to analyze the expression of SNHG20 or miR-217.

**Cell counting kit-8 (CCK-8) assay**

The proliferation of ovarian cancer cells was measured by cell counting kit-8 (CCK-8, Beyotime Institute of Biotechnology, Shanghai, China). SKOV3 or A2780 cells (4 × 10^5 per well) with different transfection were cultured in 96-well plants. After 0, 24, 48, or 72 h of incubation, 10 µl of CCK-8 solution was added into each well, then the cells were incubated at 37 °C for 2 h. The absorbance at 450 nm of transfected cells was measured with a microplate reader (Thermo Fisher, USA).

**Transwell assay**

Cell invasion ability was assessed by transwell chambers (10 μm pore size; BD Biosciences, San Jose, CA, USA). SKOV3 or A2780 cells (1 × 10^5 cells per well) were cultured in the upper chamber, and the upper chamber was pre-coated with 100 µl of Matrigel gel (BD, Franklin Lakes, NJ, USA). At the same time, RPMI-1640 medium supplemented with 10% FBS was added to the lower chamber. After incubation for 24 h, the cells on the lower surface were fixed with methanol. At last, the wells were stained with 0.5% crystal violet solution and observed by a microscope.

**Dual-luciferase reporter assay**

The SNHG20 sequences containing the binding sites (SNHG20-WT) and non-binding sites (SNHG20-MUT) for miR-217 were synthesized and inserted into a psiCHECK2 reporter vector (Promega, WI, USA). The SNHG20-WT or SNHG20-MUT reporter vector was co-transfected into SKOV3 and A2780 cells with anti-miR-217 (miR-217 inhibitor) or anti-NC (inhibitor NC), then the cells were seeded into 24-well plates. After transfection for 48 h, the luciferase activity was assessed by the Dual-Luciferase Reporter Assay Kit (Promega, WI, USA).

**RNA immunoprecipitation (RIP) assay**

The Magna RIP RNA Binding Protein Immunoprecipitation Kits (Millipore, Billerica, MA, USA) were used for the RIP assays. The operation steps of RIP analysis were consistent with the previously mentioned description (Ma et al. 2020).

**Statistical analysis**

GraphPad Prism 8. (GraphPad Software, Inc.) and SPSS 22.0 (SPSS Inc., Chicago, IL, USA) were used for data analysis. All experiments were carried out in three independent biological replicates, and the data were expressed as the mean ± SD (standard deviation). The difference between the two groups was compared by the paired or unpaired Student's t-test. The relationship between SNHG20 expression and miR-217 was analyzed by Spearman’s correlation coefficient. *P < 0.05* was considered statistically significant.

**Results**

**SNHG20 was up-regulated in ovarian cancer tissues and cell lines**

First, the expression of SNHG20 in 30 ovarian cancer tissues and their adjacent normal tissues was measured by qRT-PCR assay. The result showed that SNHG20 was overexpressed in ovarian cancer compared with that in adjacent normal tissues (Fig. 1a). In addition, we investigated whether there were some relationships between SNHG20 expression and ovarian cancer stages. The qRT-PCR data suggested that there were no significant differences in SNHG20 expression associated with different tumor stages (Fig. S1). Then, SNHG20 expression in ovarian cancer cell lines (SKOV3 and A2780) and human ovarian surface epithelial cell line (HOSEpiC) were also determined by qRT-PCR assay. The qRT-PCR data showed that SNHG20 expression was significantly higher in ovarian cancer cell lines (SKOV3 and A2780) than in HOSEpiC cell line (Fig. 1b). These results indicated that dysregulated SNHG20 might involve in the occurrence and development of ovarian cancer.

**SNHG20 positively promoted ovarian cancer cell proliferation and invasion in vitro**

To explore the functional role of SNHG20 in ovarian cancer, SKOV3 and A2780 cells were transfected with pcDNA, SNHG20, si-NC, or si-SNHG20. First, the endogenous expression level of SNHG20 was measured. The qRT-PCR results showed that SNHG20 expression was remarkably increased in SKOV3 and A2780 cells transfected with SNHG20, and the expression of SNHG20 was significantly decreased after transfection of si-SNHG20 (Fig. 2a). Next, the CCK-8 assay was performed to measure the ability of cell proliferation. The data indicated that SNHG20 significantly promoted cell proliferation and SNHG20 knockdown inhibited cell proliferation (Fig. 2b). Besides, transwell assay concluded that the number of invasive cells was collectively enhanced in SKOV3 and A2780 cells transfected with SNHG20, and knockdown of SNHG20 weakened...
Fig. 1 SNHG20 was significantly overexpressed in ovarian cancer tissues and cell lines. a QRT-PCR analysis of SNHG20 expression in ovarian cancer tissues (n = 30) and adjacent normal tissues (n = 30).

b The SNHG20 expression in ovarian cancer cell lines and normal human ovarian surface epithelial cell line (HOSePiC). **P < 0.01

Fig. 2 Effects of lncRNA SNHG20 on proliferation and invasion of ovarian cancer cells. SKOV3 and A2780 cells were transfected with pcDNA, SNHG20, si-NC, or si-SNHG20, respectively. pcDNA and si-NC were used as controls. a Expression of SNHG20 in transfected cells was measured by qRT-PCR. b Proliferation ability of transfected cells was assessed by CCK-8 assay. c The transwell assay was used to assess the relative invasion rate of transfected cells. **P < 0.01, ***P < 0.001
cell invasion (Fig. 2c). In short, these data indicated that SNHG20 regulated ovarian cancer cell proliferation and invasion.

miR-217 was a target of SNHG20

To further study the molecular mechanism of SNHG20 in ovarian cancer, the potential targets of SNHG20 were predicted using the online software LncBase Predicted v.2. The results showed that miR-217 was a target gene of SNHG20, and the binding sites between SNHG20 and miR-217 were shown in Fig. 3a. Then, to confirm whether there was direct interaction, the luciferase reporter vector was co-transfected into SKOV3 and A2780 cells with miR-217 or miR-NC. The luciferase activity of SKOV3 and A2780 cells in the SNHG20-WT group was markedly reduced by introducing miR-217, whereas the mutant vector was unaffected (Fig. 3b). Furthermore, the RIP assay indicated that SNHG20 and miR-217 were significantly enriched in human Ago2 antibodies compared with the IgG antibody control (Fig. 3c). All the above data implied that miR-217 was binding SNHG20.

Fig. 3 SNHG20 directly interacted with miR-217. a The putative binding sites between SNHG20 and miR-217 were analyzed by software LncBase v.2. b The relationship between SNHG20 and miR-217 was confirmed by dual-luciferase reporter assay. c The interaction between SNHG20 and miR-217 was further confirmed by RIP assay. ***P < 0.001

Next, the expression of miR-217 in ovarian cancer tissues and SKOV3 and A2780 cells was measured by qRT-PCR assay. The qRT-PCR data suggested that the expression of miR-217 in ovarian cancer and SKOV3 and A2780 cells were lower than in normal tissues and cells (Fig. 4a, b). The correlation between SNHG20 and miR-217 expression was performed by Pearson’s correlation analysis. The results indicated that the expression of miR-217 was negatively correlated with SNHG20 in ovarian cancer tissues (Fig. 4c). Furthermore, the SKOV3 and A2780 cells were transfected with pcDNA, SNHG20, si-NC, or si-SNHG20, and the expression of miR-217 in these cells was examined. The results determined that miR-217 expression was significantly reduced in SKOV3 and A2780 cells transfected with SNHG20 and were remarkably increased with a low SNHG20 expression (Fig. 4d). These results demonstrated that miR-217 might act as a potential role in ovarian cancer, and the expression of miR-217 was negatively regulated by SNHG20.
Overexpression of miR-217 suppressed ovarian cancer cell proliferation and invasion

To explore the effect of miR-217 on ovarian cancer cell growth and invasion, SKOV3 and A2780 cells were transfected with miR-NC or miR-217 mimic, respectively. Compared with the control group, the miR-217 expression was remarkably increased in ovarian cancer cells SKOV3 and A2780 transfected with miR-217 (Fig. 5a), while there was no significant difference in SNHG20 expression (Fig. S2). Moreover, the CCK-8 assay indicated that miR-217 overexpression suppressed cell proliferation (Fig. 5b). Furthermore, the transwell assay confirmed that overexpression of miR-217 reduced the ability of cell invasion (Fig. 5c). These results suggested that miR-217 might play a negative role in the proliferation and invasion of ovarian cancer cells.

SNHG20 promoted cell proliferation and invasion by inhibiting miR-217 expression

To clarify the relationship between SNHG20 and miR-217 clearly, some rescue experiments were designed in vitro. The qRT-PCR data exhibited that miR-217 mimic reversed the inhibition of endogenous miR-217 expression in SKOV3 and A2780 cells transfected with SNHG20 (Fig. 6a), and miR-217 inhibitor (anti-miR-217) reversed the promotion of miR-217 expression in SKOV3 and A2780 cells transfected with si-SNHG20 (Fig. 6b). Additionally, miR-217 suppressed cell proliferation in SKOV3 and A2780 cells transfected with SNHG20 (Fig. 6c), and anti-miR-217 promoted cell proliferation in ovarian cancer cells transfected with si-SNHG20 (Fig. 6d). Moreover, the ability of cell invasion was decreased in SKOV3 and A2780 cells co-transfected with SNHG20 + miR-217 (Fig. 6e), and si-SNHG20 + anti-miR-217 co-transfection was increased (Fig. 6f). These results implied that SNHG20 regulated cell proliferation...
and invasion via suppressing miR-217 expression in ovarian cancer.

**Discussion**

Ovarian cancer is a common gynecological cancer. Most patients with ovarian cancer are diagnosed at an advanced stage due to the lack of specific clinical manifestations of early ovarian cancer (Chien and Poole 2017). The low 5-year survival rate was also responsible for the higher mortality rate of ovarian cancer (Li et al. 2018). Recently, increasing studies on the molecular mechanism of ovarian cancer carcinogenesis have been reported, and many molecular mechanisms are still unknown. Hence, more efforts are needed to reveal the underlying molecular mechanism of ovarian cancer. Moreover, accumulated studies have shown that IncRNAs played an important role in tumorigenesis, and acted as potential treatment targets in cancer. In this study, we clarified the functions and molecular mechanism of IncRNA SNHG20 in ovarian cancer.

SNHG20, a member of small nucleolar RNA host genes, has been verified to make a big difference in tumorigenesis. A previous study has shown that SNHG20 was markedly elevated in glioma cells, and inhibition of SNHG20 expression could increase the apoptosis of glioma cells (Guo et al. 2019). Jin et al. reported that SNHG20 was overexpressed in non-small cell lung cancer, and knockdown of SNHG20 inhibited proliferation, migration and invasion of non-small cell lung cancer and increased apoptosis (Jin et al. 2019). It was proved that SHNG20 expression was significantly increased in prostate cancer, and SNHG20 overexpression promoted prostate cancer cell proliferation and invasion and decreased cell apoptosis (Wu et al. 2019). In ovarian cancer, SNHG20 promoted ovarian cancer progression via Wnt/β-catenin signaling (He et al. 2018) or by regulating the proliferation regulators and epithelial-mesenchymal transition (EMT)-related proteins (Wang et al. 2019). Consistent with previous studies, we verified that the expression of SNHG20 was upregulated in ovarian cancer tissues and SKOV3 and A2780 cells. Knockdown of SNHG20 inhibited the proliferation and invasion of ovarian cancer cells, whereas SNHG20 overexpression promoted cell growth and invasion. Taken
together, SNHG20 might play an important role in the cell proliferation and invasion of ovarian cancer.

In recent years, it has been confirmed that miRNAs regulated cell growth, invasion, metastasis, and apoptosis in cancer (Cao et al. 2020; Yang et al. 2020). Many miRNAs have been determined to be involved in the development of ovarian cancer. MiR-134-3p inhibited the progression of ovarian cancer via targeting flap structure-specific endonuclease 1 in vitro (Zhao et al. 2020). MiR-195 regulated tumor growth and MICU1 expression in ovarian. MiR-6086 suppressed ovarian cancer angiogenesis via regulating the OC2/VEGFA/EGFL6 axis (Wu et al. 2020). Besides, overexpression of miR-217 in epithelial ovarian cancer cells inhibited cell proliferation, migration, and invasion (Li et al. 2016). In the present study, our findings indicated that miR-217 was decreased in ovarian cancer tissues and cell lines, and miR-217 overexpression inhibited the proliferation and invasion in vitro. The above results suggested that miR-217 was involved in the proliferation and invasion of ovarian cancer as a tumor suppressor.

Previous studies have confirmed that lncRNAs are widely involved in the network of ceRNA regulation and can act as ceRNA and further function as miRNA sponge to regulate the transcription level of miRNA (Xin et al. 2018). To further investigate the potential mechanism of SNHG20 in ovarian cancer, the online software LncBase v.2 was used to predict the target genes of SNHG20. The results showed that miR-217 was a target gene of SNHG20. Then, the Pearson’s correlation analysis indicated that the expression of miR-217 was negatively correlated with SNHG20 in ovarian cancer tissues. Besides, the direct interaction of SNHG20 with miR-217 was confirmed by dual-luciferase reporter gene and RIP analysis, and miR-217 expression was regulated by SNHG20 in vitro. Furthermore, overexpression of miR-217 suppressed the proliferation and invasion and reversed the effect of SNHG20 in ovarian cancer cells.

![Fig. 6](image-url) SNHG20 promoted cell proliferation and invasion by regulating miR-217 in ovarian cancer. SKOV3 and A2780 cells were transfected with pcDNA, SNHG20, SNHG20+miR-NC, SNHG20+miR-217, si-NC, si-SNHG20, si-SNHG20+anti-miR-NC, and si-SNHG20+anti-miR-217, respectively. a, b Expression of miR-217 in transfected cells was determined by qRT-PCR assay. c, d The CCK-8 assay was used to assess the proliferation ability of transfected cells. e, f The transwell assay was used to detect the relative invasion rate of transfected cells.
All these results proved that SNHG20 promoted ovarian cancer cell proliferation and invasion by inhibiting miR-217 expression. The previous study has shown that miR-217 played an inhibitory role in epithelial ovarian cancer by suppressing IGFR1 expression (Li et al. 2016). However, it was unknown whether SNHG20/miR-217 regulated the progression of ovarian cancer by regulating IGFR1 gene or other target genes. Therefore, exploring the downstream genes of SNHG20/miR-217 in ovarian cancer is the focus of our future work. In addition, future investigations are needed to determine more specific functions of SNHG20/miR-217 in ovarian cancer cells, such as their role in cell apoptosis.

In conclusion, lncRNA SNHG20 was up-regulated in ovarian cancer tissues and cell lines. Up-regulated SNHG20 promoted growth and invasion of ovarian cancer cells, while knockdown of SNHG20 inhibited cell proliferation and invasion. Software predictive analysis indicated that miR-217 was a target gene of SNHG20. Further research showed that miR-217 expression was negatively regulated by SNHG20. Besides, miR-217 could reverse the effect of SNHG20 overexpression. Together, SNHG20 promoted the proliferation and invasion of ovarian cancer via suppressing miR-217.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s13258-021-01138-4.

Acknowledgements All authors would like to thank the Sanya People’s Hospital.

Author contributions MA designed the study, analyzed the experiments, and wrote the paper. XX and TC carried out the data collection and data analysis and revised the paper. All authors read and approved the final version of the manuscript.

Data availability The data in this research is available from the corresponding author on reasonable request.

Declarations

Conflict of interest Xuefeng Xing, Ming An, and Tonghua Chen declare that they have no conflict of interest.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

Allemanni C, Weir H, Carreira H, Harewood R, Spika D, Wang X, Bill M, Papaioannou D, Karunasis M, Kohlschmidt J, Pepe F, Walker C, Walker A, Brannan G, Pathmanathan A, Zhang X et al (2015) Global surveillance of cancer survival 1995–2009: analysis of individual data for 25,676,887 patients from 279 population-based registries in 67 countries (CONCORD-2). Lancet 385:977–1010

Bartel D (2009) MicroRNAs: target recognition and regulatory functions. Cell 136:215–233

Bill M, Papaioannou D, Karunasis M, Kohlschmidt J, Pepe F, Walker C, Walker A, Brannan G, Pathmanathan A, Zhang X et al (2019) Expression and functional relevance of long non-coding RNAs in acute myeloid leukemia stem cells. Leukemia 33:2169–2182

Cao Y, Xiong J, Zhang G, Liu Y, Jie Z, Li Z (2020) Long noncoding RNA UCA1 regulates PRL-3 expression by sponging microRNA-495 to promote the progression of gastric cancer. Mol Ther Nucleic Acids 19:853–864

Chang L, Guo K, Yuan Z, Shi H, Zhang D (2018) LncRNA HOTAIR regulates CCND1 and CCND2 expression by sponging miR-206 in ovarian cancer. Cell Physiol Biochem 49:1289–1303

Chen Z, Chen X, Chen P, Yu S, Nie F, Lu B, Zhang T, Zhou Y, Chen Q, Wei C et al (2017) Long non-coding RNA SNHG20 promotes non-small cell lung cancer cell proliferation and migration by epigenetically silencing of P21 expression. Cell Death Dis 8:e3092

Chen Y, Du H, Bao L, Liu W (2018) LncRNA PVT1 promotes ovarian cancer progression by silencing miR-214. Cancer Biol Med 15:238–250

Chen G, Yang Z, Feng M, Wang Z (2020) microRNA-217 suppressed epithelial-to-mesenchymal transition through targeting PTPN14 in gastric cancer. Biosci Rep 40:BSR20193176

Chien J, Poole E (2017) Ovarian cancer prevention, screening, and early detection: report from the 11th biennial ovarian cancer research symposium. Int J Gynecol Cancer 27:S20–S22

Guo L, Zhang Z, Li R, Li H, Cui Y (2019) Influences of LncRNA SNHG20 on proliferation and apoptosis of glioma cells through regulating the PTEN/PI3K/AKT signaling pathway. Eur Rev Med Pharmacol Sci 23:253–261

He S, Zhao Y, Wang X, Deng Y, Wan Z, Yao S, Shen H (2018) Up-regulation of long non-coding RNA SNHG20 promotes ovarian cancer progression via Wnt/β-catenin signaling. Biosci Rep 38:BSR20170681

Holmes D (2015) Ovarian cancer: beyond resistance. Nature 527:232–242

Huarte M (2015) The emerging role of lncRNAs in cancer. Nat Med 21:1253–1261

Jiang W, Hou L, Wei J, Du Y, Zhao Y, Deng X, Lin X (2020) Hsa-miR-217 inhibits the proliferation, migration, and invasion in non-small cell lung cancer cells via targeting SIRT1 and P53/KAI1 signaling. Balk Med J 37:208–214

Jin L, Jiang X, He G, Shi J, Su F, Zhu H (2019) SNHG20 knockdown suppresses proliferation, migration and invasion, and promotes apoptosis in non-small cell lung cancer through acting as a microRNA sponge. Biomed Pharmacother 112:108648

Li J, Li D, Zhang W (2016) Tumor suppressor role of miR-217 in human epithelial ovarian cancer by targeting IGFR1. Oncol Rep 35:1671–1679

Li N, Zhan X, Zhan X (2018) The lncRNA SNHG3 regulates energy metabolism of ovarian cancer by an analysis of mitochondrial proteomes. Gynecol Oncol 150:343–345

Ma Q, Qi X, Lin X, Li L, Chen L, Hu W (2020) LncRNA SNHG3 promotes cell proliferation and invasion through the miR-384/hepatoma-derived growth factor axis in breast cancer. Hum Cell 33:232–242

Matz M, Coleman M, Carreira H, Salmerón D, Chirlaque M, Allemanni C (2017) Worldwide comparison of ovarian cancer survival:
histological group and stage at diagnosis (CONCORD-2).

Pan X, Guo Z, Chen Y, Zheng S, Peng M, Yang Y, Wang Z (2020) STAT3-induced IncRNA SNHG17 exerts oncogenic effects on ovarian cancer through regulating CDK6. Mol Ther Nucleic Acids 22:38–49

Panoutsopoulou K, Avgeris M, Scorilas A (2018) miRNA and long non-coding RNA: molecular function and clinical value in breast and ovarian cancers. Expert Rev Mol Diagn 18:963–979

Sankaranarayanan R, Ferlay J (2006) Worldwide burden of gynecological cancer: the size of the problem. Best Pract Res Clin Obstet Gynaecol 20:207–225

Siegel RL, Miller KD, Jemal A (2017) Cancer statistics, 2017. CA Cancer J Clin 67:7–30

Tristán-Ramos P, Rubio-Roldan A, Peris G, Sánchez L, Amador-Cubero S, Viollet S, Cristofari G, Heras S (2020) The tumor suppressor microRNA let-7 inhibits human LINE-1 retrotransposition. Nat Commun 11:5712

Wang P, Ning S, Zhang Y, Li R, Ye J, Zhao Z, Zhi H, Wang T, Guo Z, Li X (2015) Identification of IncRNA-associated competing triplets reveals global patterns and prognostic markers for cancer. Nucleic Acids Res 43:3478–3489

Wang D, Dai J, Hou S, Qian Y (2019) IncRNA SNHG20 predicts a poor prognosis and promotes cell progression in epithelial ovarian cancer. Biosci Rep 39:BSR20182186

Webb PM, Gynaecological Cancers Group, Jordan SJ, Cancer Causes and Care Group (2017) Epidemiology of epithelial ovarian cancer. Best Pract Res Clin Obstet Gynaecol 41:3–14

Wu X, Xiaoy, Zhou Y, Zhou Z, Yan W (2019) IncRNA SNHG20 promotes prostate cancer migration and invasion via targeting the miR-6516-5p/SCGB2A1 axis. Am J Transl Res 11:5162–5169

Wu B, Zhang L, Wu Y, Lu T, Zhang Y, Zhu W, Song Q, Lv C, Guo J, Tian Y et al (2020) miR-6086 inhibits ovarian cancer angiogenesis by downregulating the OC2/VEGFA/EGFL6 axis. Cell Death Dis 11:345

Xin X, Wu M, Meng Q, Wang C, Lu Y, Yang Y, Li X, Zheng Q, Pu H, Gui X et al (2018) Long noncoding RNA HULC accelerates liver cancer by inhibiting PTEN via autophagy cooperation to miR15a. Mol Cancer 17:94

Yang C, Zheng X, Ye K, Sun Y, Lu Y, Fan Q, Ge H (2020) miR-135a inhibits the invasion and migration of esophageal cancer stem cells through the hedgehog signaling pathway by targeting Ssm. Mol Ther Nucleic Acids 19:841–852

Yoshida N, Kimura T (2017) Pathogen-associated regulatory non-coding RNAs and oncogenesis. Front Biosci 22:1599–1621

Yu J, Shen J, Qiao X, Cao L, Yang Z, Ye H, Xi C, Zhou Q, Wang P, Gong Z (2019) SNHG20/miR-140-5p/NDRG3 axis contributes to 5-fluorouracil resistance in gastric cancer. Oncol Lett 18:1337–1343

Zhao Q, Gao S, Du Q, Liu Y (2018) Long non-coding RNA SNHG20 promotes bladder cancer via activating the Wnt/β-catenin signalling pathway. Int J Mol Med 42:2839–2848

Zhao W, Ma X, Liu L, Chen Q, Liu Z, Zhang Z, Ma S, Wang Z, Li H, Wang Z et al (2019) SNHG20: a vital IncRNA in multiple human cancers. J Cell Physiol. https://doi.org/10.1002/jcp.28143

Zhao M, Ji H, Fu Q, Cheng Q, Zhang Y, Yang Y (2020) MicroRNA-134-3p inhibits ovarian cancer progression by targeting flap structure-specific endonuclease 1 in vitro. Oncol Rep 45:119–128

Zhou S, Xu A, Song T, Gao F, Sun H, Kong X (2020) IncRNA MIAT regulates cell growth, migration, and invasion through sponging miR-150-5p in ovarian cancer. Cancer Biother Radiopharm 35:650–660

Zhu L, Yang S, Wang J (2019) miR-217 inhibits the migration and invasion of HeLa cells through modulating MAPK1. Int J Mol Med 44:1824–1832

Publisher’s Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.