The RGD motif in fibronectin is essential for development but dispensable for fibril assembly

Seiichiro Takahashi,1 Michael Leiss,1 Markus Moser,1 Tomoo Ohashi,2 Tomoe Kitao,3 Dominik Heckmann,4 Alexander Pfeifer,5 Horst Kessler,4 Junichi Takagi,3 Harold P. Erickson,2 and Reinhard Fässler1

1Max Planck Institute of Biochemistry, Department of Molecular Medicine, 82152 Martinsried, Germany
2Department of Cell Biology, Duke University Medical Center, Durham, NC
3Laboratory of Protein Synthesis and Expression, Institute for Protein Research, Osaka University, Suita, Osaka, Japan
4Center of Integrated Protein Science at Department Chemie, Technical University Munich, 85747 Garching, Germany
5Institute of Pharmacology and Toxicology, University of Bonn, 53113 Bonn, Germany

Fibronectin (FN) is secreted as a disulfide-bonded FN dimer. Each subunit contains three types of repeating modules: FN-I, FN-II, and FN-III. The interactions of α5β1 or αv integrins with the RGD motif of FN-III repeat 10 (FN-III10) are considered an essential step in the assembly of FN fibrils. To test this hypothesis in vivo, we replaced the RGD motif with the inactive RGE in mice. FN-RGE homozygous embryos die at embryonic day 10 with shortened posterior trunk, absent tail bud–derived somites, and severe vascular defects resembling the phenotype of α5 integrin–deficient mice. Surprisingly, the absence of a functional RGD motif in FN did not compromise assembly of an FN matrix in mutant embryos or on mutant cells. Matrix assembly assays and solid-phase binding assays reveal that αvβ3 integrin assembles FN-RGE by binding an isoDGR motif in FN-I5, which is generated by the nonenzymatic rearrangement of asparagines (N) into an iso-aspartate (iso-D). Our findings demonstrate that FN contains a novel motif for integrin binding and fibril formation whose activity is controlled by amino acid modification.

Introduction

Fibronectin (FN) is a ubiquitous and abundant ECM protein, which is secreted as a disulfide-bonded dimer consisting primarily of three types of repeating modules (I, II, and III; Hynes, 1992; Kornblith et al., 1996). Distinct FN modules bind cell surface receptors such as integrins and syndecans (Plow et al., 2000) and ECM proteins such as collagens, fibrin, and FN itself (Pankov and Yamada, 2002). FN plays an essential role during development, in physiology (tissue repair), and in disease (cancer progression and invasion, inflammation, atherosclerosis, etc.). The functions of FN are critically dependent on the assembly of the secreted FN dimers into a fibrillar network. FN assembly is a cell-driven process that occurs in a stepwise manner: the dimeric FN protein is secreted in a compact or inactive conformation; integrin binding converts FN into an active and extended dimer; the dimeric state of FN induces integrin clustering and accumulation of integrin-bound FN, which in turn allows FN–FN interactions at different sites along the FN molecule and the formation of fibrils (Wierzbiacka-Patynowski and Schwarzbauer, 2003).

The activation of FN is most efficiently induced by α5β1 integrin binding to the RGD motif in the type III-10 module along with the synergy sequence located in the adjacent type III-9 module (Pierschbacher and Ruoslahti, 1984; McDonald et al., 1987; Nagai et al., 1991; Sechler et al., 1997). Additional RGD binding integrins, including αvβ3 and αIIbβ3, can substitute for α5β1 in FN matrix assembly but are less able to form a dense and delicate fibrillar network in vitro (Wu et al., 1995, 1996; Wennerberg et al., 1996). Furthermore, it has also been reported that FN can be assembled in an RGD-independent manner through binding of α4β1 integrin to the connecting segment-1 (CS1) in the alternatively spliced V region near the C terminus of FN (Sechler et al., 2000). It is not clear, however, whether the CS1-dependent FN assembly occurs and is of significance in vivo.

Gene ablation studies in mice confirmed that the two RGD-dependent FN assembly mechanisms induced by either α5β1 integrin or αv integrins exist in vivo and that each of them can compensate for the absence of the other. Mice lacking the α5 integrin gene, as well as mice lacking the αv integrin gene,
exhibit normal assembly of FN matrix (Yang et al., 1993; Bader et al., 1998). In contrast, mice carrying deletions of both the α5 and the αv integrin genes display severely compromised FN matrix formation (Yang et al., 1999).

Antibody and peptide inhibition studies in amphibians and chicks (Darribère et al., 1988; Harrisson et al., 1993), chemical mutagenesis in zebras (Julich et al., 2005; Koshida et al., 2005), and gene targeting in mice (Bouvard et al., 2001) identified a crucial role for the FN matrix in the development of mesoderm and mesoderm-derived structures. The mesoderm is one of the three germ layers that forms during gastrulation and is essential for morphogenesis and organ development. FN-deficient mice commence gastrulation and die at embryonic day (E) 8–8.5 with several mesodermal defects, including shortened anterior–posterior axis, disorganized notochord, absence of heart and somites, and abnormal vasculogenesis (George et al., 1993). Mice lacking the expression of both the α5 and the αv integrins (and hence all FN-RGD binding integrins) also display arrested development at the beginning of gastrulation (Yang et al., 1999). α5/αv double-null mice are arrested at an earlier stage than the FN-deficient mice. The α5/αv double mutants completely lack the mesodermal germ layer and contain only a few mesodermal cells in the developing head fold region and allantois, indicating that the two integrins must perform functions in addition to FN matrix assembly. Interestingly, mice lacking only the α5 integrin die ~E9.5–10, and despite their ability to develop a normal FN matrix, they also have mesodermal defects. These are restricted to the posterior region of the embryo and include an arrest in posterior axis elongation, the absence of posterior somites, and vascular defects (Yang et al., 1993). These observations indicate that the α5β1–FN interaction conveys essential signals in the posterior trunk mesoderm that are not compensated by αv integrins or other FN binding proteins. It is also possible, however, that the α5β1 integrin requires interaction with ECM proteins other than FN to extend the posterior trunk and develop posterior somites. In contrast, mice without αv integrin display no mesodermal abnormalities and die either of placenta defects or perinatally as a result of severe intracerebral hemorrhage, indicating that the role of the αv integrins for mesoderm formation is fully compensated by α5β1 (Bader et al., 1998).

To test how FN binding receptors compensate for each other during development and FN fibril formation, it would be necessary to generate double, triple, or even higher order combinations of integrin and syndecan knockout mice. However, the necessary intercross combinations cannot be performed because most integrin-null mice are recessive lethal (α4, α5, α8, and αv). Moreover, integrins and syndecans interact with multiple ECM proteins, and ECM proteins interact with multiple integrins, which would further complicate the interpretation of the phenotypes.

Therefore, we decided to specifically test the role of FN-RGD binding integrins in vivo by generating mice in which the aspartic acid (D) residue of the RGD motif was replaced with a glutamate (E). Many previous studies have shown that this replacement completely inactivates this site for cell adhesion. We found that the FN(RGE/RGE) mice died early in embryogenesis from multiple defects, but surprisingly they assembled an apparently normal FN matrix. Furthermore, our studies revealed a novel integrin binding site in the fifth N-terminal type I module of FN, which was essential for the assembly of the FN-RGE matrix.

### Table I. Genotypes of progeny from heterozygous intercrosses

| Stage | Total | n | %  | n  | %  | n  | %  |
|-------|-------|---|----|----|----|----|----|
| E7.5  | 16    | 5 | 31.2| 8  | 50.0| 3  | 18.8|
| E8.5  | 170   | 46| 27.1| 81 | 47.6| 43 | 25.3|
| E9.5  | 95    | 26| 27.3| 46 | 48.4| 23 | 24.2|
| E10.5 | 46    | 12| 26.1| 21 | 45.7| 13 | 28.3|
| E11.5 | 26    | 11| 42.3| 15 | 57.7| 3  | 11.5|
| E12.5 | 15    | 4 | 26.6| 11 | 73.3| 0  | 0.0 |
| E13.5 | 4     | 2 | 50.0| 2  | 50.0| 0  | 0.0 |
| P21   | 157   | 55| 35.0| 102| 65.0| 0  | 0.0 |

*Embryos without a beating heart.

Results

#### Generation of FN(RGE/RGE) knockin mice

To test the role of the RGD motif in FN-III10 in vivo, we cloned part of the mouse FN gene, changed the GAC codon in exon 30 to GAG by site-directed mutagenesis, and inserted a loxP flanked neo cassette into intron 30 (Fig. S1, A and B, available at http://www.jcb.org/cgi/content/full/jcb.200703021/DC1). The C>G nucleotide change resulted in a replacement of the aspartate (D) residue of the RGD motif with a glutamate (E) and created a novel Hinfl restriction site in the mutant FN gene (Fig. S1, A and B). The DNA vector was used to transfer the mutation via embryonic stem cells into the germline of mice. The neo cassette was removed by breeding to a deleter-Cre strain. Mice were genotyped by PCR analysis of tail or yolk sac biopsies, and the knockin mutation was confirmed by sequencing of the PCR amplicon (Fig. S1, B and C). Mice heterozygous for the mutant FN allele (FN(RGE+/−)) were normal.

Heterozygous mice were intercrossed to obtain homozygosity (FN(RGE/RGE)). Out of 157 viable offspring, we obtained 65% heterozygous and 35% wild type but no homozygous mutant mice (Table I). This indicates that the RGE mutation in FN represents a recessive embryonic lethal trait.

To determine the exact time point of lethality, we dissected embryos derived from heterozygous intercrosses and determined their genotypes by genomic PCR using DNA extracted from either the yolk sac or embryos. We found that genotypes...
showed a normal Mendelian distribution until E10.5, whereas beyond E10.5, no homozygous mutant embryos with beating hearts were recovered (Table 1). At ~E11.5, FN\textsuperscript{RGE/RGE} embryos started to disintegrate or were already partially resorbed.

**FN\textsuperscript{RGE/RGE} embryos display multiple abnormalities**

To search for defects and identify the cause for the lethality, we isolated whole-mount embryos from heterozygous intercrosses at different time points of development. At E8.5, the FN\textsuperscript{RGE/RGE} embryos had slightly shorter anterior–posterior axes but were otherwise indistinguishable from their wild-type or heterozygous littermates (Fig. 1, A and B). Both the FN\textsuperscript{RGE/RGE} embryos and wild-type littermates contained three to four somite pairs and had a well-developed head fold, neural tube, and beating heart. At E9.5, wild-type embryos were turned into the fetal position and their posterior trunks extended into a C-like shape (Fig. 1 C). In contrast, FN\textsuperscript{RGE/RGE} embryos were only partially turned and displayed a kinked neural tube and severely shortened posterior trunk without somites (Fig. 1 D). E9.5 FN\textsuperscript{RGE/RGE} embryos contained never more than 12 or 13 somite pairs, whereas wild-type littermates had ~21 somite pairs (Fig. 1, C and D). The neural tubes were noticeably kinked, the limb buds were absent, and the embryonic and yolk sac vasculature was frequently dilated and sometimes ruptured, leading to blood cell leakage into the amniotic cavity (unpublished data). At E10.5, the head structures, heart, and fore limb buds of FN\textsuperscript{RGE/RGE} embryos were further developed, whereas the number of somites had not increased further (Fig. 1, E and F).

To test whether proliferation or survival was affected by the FN-RGE mutation, we analyzed Ki67, TUNEL, and activated caspase-3 signals. Proliferation as tested by Ki67 staining was similar in control and FN\textsuperscript{RGE/RGE} embryos (unpublished data). In sharp contrast, apoptotic cell death of tail bud–derived mesodermal cells was dramatically elevated, as shown in a double staining for TUNEL (Fig. 1, G and H) and activated caspase-3 signals (Fig. 1, I and J).

**FN-RGE is normally distributed and assembled in vivo**

It has become generally accepted that the RGD motif in FN is essential for FN matrix assembly (Nagai et al., 1991; Schwarzauer, 1991; Sechler et al., 1997). Consequently, we expected mutation of the RGD motif to abolish fibril assembly and hamper fibril stability or function. To evaluate FN-RGE matrix assembly, we performed immunostaining and Western blot experiments. Immunostaining of parasagittal, paraffin-embedded sections from PFA-fixed E9.5 embryos revealed that FN-RGE was widely expressed. Closer inspection of different areas within the FN\textsuperscript{RGE/RGE} embryos revealed strong FN-RGE matrix surrounding the nervous tissue, somites, and large vessels and a fine, network-like distribution in the heart, developing facial, and trunk mesenchyme, within the somites and the tail bud mesoderm (Fig. 2 A). Neither the distribution nor the intensity of the FN immunosignal differed between wild-type and FN\textsuperscript{RGE/RGE} littermates (Fig. 2 A). Western blot assays of whole embryo lysates confirmed the normal expression levels of FN-RGE in FN\textsuperscript{RGE/RGE} embryos (Fig. 2 B).

![Figure 1](image.png)

**Figure 1. Whole-mount pictures of wild-type and FN\textsuperscript{RGE/RGE} embryos.**

(A and B) Side view of an E8.5 wild-type embryo with four pairs of somites (A) and an FN\textsuperscript{RGE/RGE} embryo (B) with four pairs of somites and a well-developed allantois (al). The FN\textsuperscript{RGE/RGE} embryo is slightly smaller. (C and D) Side view of an E9.5 wild-type (C) and FN\textsuperscript{RGE/RGE} (D) embryo. The wild-type embryo has turned into the fetal position and contains 21 pairs of somites (C). The E9.5 FN\textsuperscript{RGE/RGE} embryo shows incomplete turning, a kinked neural tube, and a truncation of the posterior trunk region with 13 pairs of somites in the anterior region (D). Inset shows anterior somites at higher magnifications. The head region of the embryo is normally developed. (E and F) Side view of an E10.5 wild-type embryo with 29 somite pairs (E) and an E10.5 FN\textsuperscript{RGE/RGE} embryo with 13 somite pairs (F). The first branchial arch and fore limb buds (fl) are developed, whereas the second branchial arch is missing (f). (G and H) Apoptotic cell death in FN\textsuperscript{RGE/RGE} embryos. TUNEL and cleaved form of caspase-3 staining reveal massive cell death in posterior trunk regions of E9.5 FN\textsuperscript{RGE/RGE} embryo in comparison with very few positive cells in the wild-type embryo. Note the colocalization (yellow) between TUNEL (green) and activated cleaved form of caspase-3 (red) staining. h, heart; s, somites; nt, neural tube. Bars: (B, D, and F) 500 μm; (J) 100 μm.

The apparently normal FN-RGE fibrils in FN\textsuperscript{RGE/RGE} embryos was unexpected and could be due to a cross-linking artifact induced by the PFA fixation or, alternatively, to α4β1 integrin–dependent assembly. To test whether either was the case, we immunostained unfixed cryosections derived from
Importantly, to rule out the possibility that the observed differences between control and FNRGE/RGE embryos are caused by any differences in the sera they were maintained in, we grew FNRGE/RGE embryos in serum-free media. To avoid serum-derived ECM, we used serum-free media. To avoid serum-derived FN, we grew FNRGE/RGE embryos in serum-free media.

When labeled wild-type plasma FN (pFN) was added to the FNRGE/RGE culture, FNRGE/RGE cell clones assembled it into a normal fibril network, with 𝛼v integrin colocalizing into typical fibril adhesions (Fig. 3 B). These data suggest that 𝛼v integrins are involved in the assembly of FN-RGE and are not responsible for the differences observed between control and FNRGE/RGE cells.

Next, we tested whether FNRGE/RGE cells are able to assemble their endogenously produced mutant FN-RGE. Assembly of self-produced FN critically depends on the absence of exogenous serum-derived FN, which is a widely used supplement of cell culture media. To avoid serum-derived FN, we grew control and FNRGE/RGE cells in serum replacement medium and, because they lacked the serum-derived ECM components, seeded them on defined substrates.

Control and FNRGE/RGE cells plated on LM111 adhered via their laminin binding 𝛼6𝛽1 integrin, leaving the 𝛼5𝛽1 integrin, as well as the 𝛼v integrins, free to contribute to the assembly of the secreted FN. In contrast to the previous notion that the RGD sequence of FN is essential for the initiation of matrix assembly, FNRGE/RGE cells initiated and completed the assembly of FN-RGE into fibrils on their cell surface (Fig. 3 A). Similar to mouse embryos, FNRGE/RGE cells were able to assemble a dense fibrillar network (Fig. 3 A). Although the kinetic of fibril formation was similar between control and FNRGE/RGE cells, the size and the distribution of fibrils differed between the two cell types. Control cells produced an elaborate network consisting of thin and long FN fibrils (Fig. 3 A). In contrast, FN fibrils formed by FNRGE/RGE cells appeared short and thick.

To determine which integrin, 𝛼v and/or 𝛼5, mediates assembly of FN-RGE, we plated cells on VN. On VN, control and FNRGE/RGE cells adhered via their 𝛼v integrins. A previous study showed that this depletes 𝛼vB3 integrin from the cell surface but leaves 𝛼5B1 integrin diffusely distributed and presumably free to participate in matrix assembly (Fath et al., 1989). Control cells developed a regular FN network when plated on VN, whereas FNRGE/RGE cells were unable to form FN-RGE fibrils (Fig. 3 B). These data suggest that 𝛼v, but not 𝛼5B1, integrins mediate the assembly of FN-RGE into matrix fibrils.

αv integrins can trigger an RGD-independent FN assembly pathway

To further investigate whether 𝛼v and not 𝛼5B1 integrins executed FN-RGE fibril formation, we immunostained FNRGE/RGE cells growing on LM111 with 𝛼5 integrin and FN antibodies. As expected, control cells showed coalignment of the 𝛼5 integrin with the thin and elaborate FN network (Fig. 4 A). In sharp contrast, LM111-growing FNRGE/RGE cells failed to align 𝛼5 integrin with FN-RGE-containing fibrils. Instead, the 𝛼5 integrin signal was diffusely distributed over the entire cell surface.
further indicating that it is not engaged in FN-RGE binding (Fig. 4 A). Unfortunately, because of the poor quality of the \( \alpha_v \) integrin antibodies that were available to us, we could not directly demonstrate a colocalization of \( \alpha_v \) integrins with FN-RGE fibrils.

To confirm that \( \alpha_v \) integrins were indeed assembling FN-RGE fibrils, we depleted \( \alpha_v \) integrins in FN\(^{RGE/RGE} \) cells by viral transduction of \( \alpha_v \)-specific siRNAs and assayed for FN assembly. FN\(^{RGE/RGE} \) cells expressed high levels of \( \alpha_v \) integrins (Fig. S3 A and Fig. S4, available at http://www.jcb.org/cgi/content/full/jcb.200703021/DC1), which were reduced by \( \sim80\% \) after retroviral expression of siRNAs (Fig. S4). The \( \alpha_v \) integrin–depleted FN\(^{RGE/RGE} \) cells were cultured in serum replacement medium and seeded on LM111. Despite the availability of free \( \alpha_5 \beta_1 \) integrins, the \( \alpha_v \)-depleted FN\(^{RGE/RGE} \) cells were unable to form FN-RGE fibrils (Fig. 4 B). Occasionally, we could observe a few thick fibrils after longer culture periods, which likely developed as a result of the remaining low levels of \( \alpha_v \) integrins on FN\(^{RGE/RGE} \) cells (Fig. 4 B).

To address the question of whether assembly of FN-RGE is accomplished via the RGD binding pocket of \( \alpha_v \) integrins, we tested FN-RGE fibrillogenesis in the presence of linear Gly-Arg-Gly-Ser-Pro peptides (linRGD) or cyclic cyclo(-Arg-Gly-Asp-D-Phe-Val-) peptides (cycRGD). The more constrained cycRGD was reported to preferentially bind \( \alpha_v \beta_3 \) integrin (Pfaff et al., 1994). Incubation of control and FN\(^{RGE/RGE} \) cells with the linRGD peptide considerably inhibited assembly of wild-type as well as FN-RGE fibrils (Fig. 5). Treatment with the cycRGD peptide inhibited assembly of FN-RGE much more strongly but showed only partial inhibition of assembly of wild-type FN (Fig. 5). Treatment of control and FN\(^{RGE/RGE} \) cells with peptides, in which the RGD binding motif is mutated (linear Gly-Arg-Gly-Glu-Ser-Pro peptide [linRGE] or cyclic cyclo(-Arg-Ala-Asp-D-Phe-Val-) peptides [cycRAD]), had no effect on assembly of wild-type FN or FN-RGE (Fig. 5). Similarly, heparin treatment, which blocks the binding of FN to cell surface proteoglycans such as syndecans, partially inhibited assembly of wild-type FN but had almost no effect on FN-RGE fibrils. These data demonstrate that the RGD binding pocket of \( \alpha_v \) integrins is essential for the assembly of the FN-RGE matrix.
The FN-I1-9 domains bind αvβ3 integrin with high affinity

The data so far indicate that αv integrins bind and assemble FN-RGE into fibrils via a novel integrin binding site on FN. To localize the novel αv binding/assembly sites, we cultured FN-RGE cells on LM111 and treated them with recombinant FN fragments spanning almost the entire FN molecule (Fig. 6). Although FN-I1-9, FN-III7-10, and FN-III10-12 strongly inhibited the formation of both wild-type and FN-RGE fibrils, FN-III1-5, FN-III4-7, FN-III7-10RGE, and FN-III13-15 had no effect on either (Fig. 6 and Fig. S5, available at http://www.jcb.org/cgi/content/full/jcb.200703021/DC1).

FN-I1-9 is generally recognized as a potent inhibitor of matrix assembly and is thought to participate in binding to receptors on the cell surface or to specific FN-III domains of another FN molecule (McKeown-Longo and Mosher, 1984; Aguirre et al., 1994; Hocking et al., 1994; Bultmann et al., 1998). Because assembly of the FN-RGE matrix depended on αvβ3 integrin, we wondered if this integrin might bind directly to FN-I1-9.

We therefore examined whether αvβ3 integrin is capable of binding to the recombinant FN-I1-9 fragment. To this end, we performed direct and competitive solid-phase binding assays, in which the recombinant FN fragments were adsorbed to microtiter plates and then incubated with serially diluted biotinylated αvβ3 integrin. Direct binding assays showed comparable binding of αvβ3 integrins to FN-I1-9, FN-III7-10, and FN-III10-12 but no binding to the other fragments tested (Fig. 7 A and Fig. S5). Interestingly, α5β1 was also able to bind FN-I1-9, although α5β1 was incapable of assembling FN-RGE fibrils (Fig. 7 C).

Competitive binding assays showed that incubation with the linRGD peptide inhibited binding of both αvβ3 integrins to FN-I1-9, FN-III10, and FN-III10 but no binding to the other fragments tested (Fig. 7 A and Fig. S5). Interestingly, α5β1 was also able to bind FN-I1-9, although α5β1 was incapable of assembling FN-RGE fibrils (Fig. 7 C).
The GNGRG motif in FN-I$_5$ represents a novel $\alpha\beta_3$ binding and assembly site for FN

Primary and tertiary structure analyses showed that murine FN contains several GNGRG loops within the FN-I$_{1-9}$ that are well conserved in human, bovine, rat, amphibian, and fish (Di Matteo et al., 2006). While we were preparing this manuscript, Curnis et al. (2006) reported that a single GNGRG loop in FN-I$_1$ can convert to GisoDGRG (where isoD is isoa aspartate) and that the GisoDGRG provided a novel binding site for $\alpha\beta_3$ integrin. To explore this observation, we synthesized the (*Cys-Asn-Gly-Arg-Cys*) peptide (NGR-2C) as well as the (*Cys-isoAsp-Gly-Arg-Cys*) peptide (isoDGRG-2C), which adopts a conformation similar to the deamidated, rearranged GNGRG loop of FN-I$_1$ (Di Matteo et al., 2006) and used them in our competitive solid-phase binding and FN in vitro assembly assays. As shown in Fig. 7, isoDGRG-2C strongly inhibited assembly of the DGRG motif. The rearrangement of NGR-2C into GisoDGRG and GDGRG is a well-known reaction occurring spontaneously under physiological conditions (Robinson et al., 2004). Because NGR-2C inhibited FN-RGE formation occurring spontaneously under physiological conditions compared with the GisoDGRG, as well as isoDGRG-2C (Fig. 8 A), we tested the kinetics of the rearrangement of the NGR-2C into isoDGRG-2C. HPLC assays revealed that $\sim$10% of NGR-2C was converted into isoDGRG-2C after an incubation of 16 h at 37°C at pH 7.2 (Fig. 8 B). The experiment also shows that before incubation at 37°C, the peptide was nearly 100% NGR-2C. Therefore, when testing for inhibition of FN matrix assembly, only a small amount of isoDGRG-2C would have accumulated in the later time of the 14-h assay. The inhibition observed is likely due to unmodified NGR-2C interacting with $\alpha\beta_3$. Altogether, these data indicate that the GNGRG motif in FN-I$_1$ represents a novel and selective $\alpha\beta_3$ binding that can function for FN fibril assembly in vitro and in vivo.

Discussion

The RGD binding motif in FN is thought to play a central role in FN matrix assembly and RGD-mediated integrin signaling in vivo. In the present study, we assessed these roles by creating a nonfunctional RGE site in the FN gene of mice. The phenotype of FN$^{RGE/RGE}$ mice is identical in type and quality to the defects reported for $\alpha\beta_1$ integrin–deficient mice (Yang et al., 1993) but much less severe than the phenotype of FN-null or $\alpha\alpha/\alpha\varepsilon$ double-null mice (Yang et al., 1999). This finding allows us to draw several important conclusions. First, the mesodermal defects caused by absent $\alpha\beta_1$ integrin expression are solely based on the disruption of $\alpha\beta_1$ binding to the RGD motif of FN and do not involve binding to other $\alpha\beta_1$ ligands (collagen XVIII, ADAMs, etc.). Second, the development of a less severe phenotype in FN$^{RGE/RGE}$ mice than in $\alpha\varepsilon/\alpha\varepsilon$ double-null mice indicates that $\alpha\varepsilon$ integrin–FN interactions either are not required for early development or occur in an RGD-independent manner. Third, the direct observation of FN matrix in FN$^{RGE/RGE}$ mice shows that the RGD motif in FN must be dispensable for the formation of a functional fibrillar FN network.

![Figure 7](image-url)

**Figure 7.** Direct solid-phase FN and integrin binding assay. (A) Titration curve of $\alpha\varepsilon$3 integrin bound to different recombinant FN fragments. After blocking of nonspecific binding sites, $\alpha\varepsilon$3 was allowed to bind in TBS buffer containing 1 mM MnCl$_2$. Mean ± SD ($n = 3$). Note that $\alpha\varepsilon$3 integrin bound with high affinity to FN-I$_{1-9}$. (B) FN-I$_{1-9}$ was immobilized at 0.25 $\mu$g/ml heparin (triangles), 750 $\mu$M isoDGRG-2C (squares), or 850 $\mu$M linRGD peptide (circles). The values plotted are normalized to the maximal binding value to give the yield of saturation. Maximal binding of $\alpha\beta_3$ to FN-I$_{1-9}$ corresponds to an OD (405 nm) value of 1.08. Note that $\alpha\varepsilon$3 binding to FN-I$_{1-9}$ is inhibited by both linRGD and isoDGRG-2C peptides. (C) The same experimental procedure was performed as described in B, with the exception that $\alpha\beta_3$ integrin was used for titration. Maximal binding of $\alpha\beta_1$ to FN-I$_{1-9}$ corresponds to an OD (405 nm) value of 1.04. Note that $\alpha\beta_1$ integrin binding to FN-I$_{1-9}$ is inhibited by linRGD peptides but not by isoDGRG-2C peptides. In all binding assays, the absorbance measured in BSA-coated wells was <15% relative to maximum absorbance values and was routinely subtracted. All binding was determined in duplicate.
Based on this mouse work and previous FN assembly studies performed in vitro, one would have predicted that a non-functional RGD motif in FN would abrogate binding to both α5β1 and αv integrins and hence render fibrillogenesis equally impossible as in α5αv double-null mice. However, this was not the case. We found neither quantitative nor qualitative differences in the FN network of FNRGE/RGE mice, irrespective of whether the embryo sections were treated with fixative or examined in native cryosections.

One reason for this unexpected finding could have been that the D>E mutation was too conservative and hence would still permit binding to integrins. Such a possibility, however, was ruled out experimentally: the assembly of FN-RGE by FN^{RGD/RGE} cells grown on LM111 was inhibited by the addition of neither linear RGE peptides nor recombinant FN-III_{1,10} fragment, in which the RGD was converted into an RGE. Moreover, the same recombinant FN-III_{1,10}RGE fragment was unable to bind integrins in a highly sensitive solid-phase binding assay.

**FN-RGE is assembled by αv integrins**

FN assembly via the RGD motif can occur through either α5β1 or αv integrins. The αv integrin–mediated assembly, however, leads to a less dense network with shorter and thicker fibrils in vitro (Wennerberg et al., 1996), which is thought to be due to the different subcellular localization and associated shape of αv versus α5β1 integrin clusters (α5β1 form thin fibrillar adhesions and αv integrins arrowhead-like focal adhesions) and/or to the differential activation of small GTPases such as RhoA, which occurs through FN binding to α5β1 but not to αv integrins (Danen et al., 2002).

The FN-RGE fibrils of FN^{RGD/RGE} cells displayed appearance and behavior typical of αv-mediated fibrils: they were short and thick, whereas fibrils of wild-type cells were thin, long, and ramified. Furthermore, the assembly of FN-RGE could be efficiently inhibited by cycRGD, which was shown to bind with higher affinity to αvβ3 than to α5β1 integrins (Pfaff et al., 1994). The interpretation that αv integrins mediate FN-RGE assembly was further supported by siRNA-mediated depletion of the αv integrin protein in FN^{RGD/RGE} cells, which showed that αv integrin–depleted cells were unable to form a fibrillar FN-RGE network.

The role of αv integrins was further confirmed by direct binding assays, with recombinant fragments spanning almost the entire FN protein, which revealed a novel high-affinity binding interaction between αvβ3 integrin and the N-terminal FN-I_{1,9} fragment. Moreover, the assembly of FN-RGE fibrils by FN^{RGD/RGE} cells was efficiently inhibited by FN-I_{1,9} and by synthetic peptides comprising the novel αvβ3 integrin binding site.

RGD-independent mechanisms for FN assembly have been observed with different cell culture systems. One report showed that binding of Mn^{2+}-activated α4β1 integrin to the CS1 site can induce assembly of FN in vitro (Sechler et al., 2000). It is not clear, however, whether the α4β1-mediated mechanism also operates in vivo and, if so, whether its role is substantial during embryogenesis. In the E9.5 control and FN^{RGD/RGE} embryos, we observed a very restricted expression of α4β1 on a few cells in the cranial region and the developing epicard. Thus, the abundant FN-RGE fibrils in sites without α4β1 expressing
cells must have assembled using an α4β1-independent mechanism. This conclusion was further confirmed with FN\textsuperscript{RG/GE} cell lines that lacked α4 integrin but still efficiently assembled FN-RGE fibrils.

A second RGD-independent FN assembly mechanism has been described for cells isolated from α5 integrin-deficient mice (Sottile et al., 2000). Incubation of FN-null cells growing in serum replacement medium and on VN with recombinant FN lacking the RGD motif (FN\textsubscript{ΔRGD}) assembled short stitch-like aggregates on the cell surface. A potential explanation for the development of dot-like FN\textsubscript{ΔRGD} aggregates could be that a few αvβ3 integrins failed to bind the VN substrate and were available to recruit FN\textsubscript{ΔRGD} to the cell surface by binding to the site in FN-11-9. Alternatively, the recruitment and aggregation of FN\textsubscript{ΔRGD} may have been accomplished by syndecans, which also bind FN. Potential syndecan–FN interaction would also explain why heparin treatment inhibited the formation of the stitch-like aggregates on FN-null cells. We also tested a portion of FN\textsubscript{Δ} with heparin. Interestingly, heparin had no effect on FN-I\textsubscript{1-9} aggregates on the cell surface. A potential explanation for the heparin inhibition suggests that the native NGR sequence itself may already be able to act as a high-affinity binding site for αvβ3 and function for matrix assembly of FN-RGE. It is more likely, however, that the NGR motif was sufficiently modified in culture to inhibit FN-RGE assembly. Supporting this hypothesis is the fact that NGR peptides failed to block αvβ3 binding to the FN-I\textsubscript{5} module (Curnis et al., 2006). Moreover, assembly of the FN-RGE matrix in both embryos and cell culture requires only a small fraction of isoDGR-modified FN molecules to bind cell surface integrins, whereas the remaining fraction becomes integrated into fibrils via FN–FN interactions, which proceeds independent of the isoDGR modification.

Interestingly, α5β1 also bound the FN-I\textsubscript{5} peptide, but the isoDGR-2C peptide failed to inhibit this binding or assembly of wild-type FN. A likely explanation is that the interaction of α5β1 with FN-I\textsubscript{5} is weaker than the interaction of αvβ3 with the isoDGR motif in FN-I\textsubscript{5} and fails to provide the necessary binding strength to allow assembly of FN-RGE fibrils. Conversely, our findings also indicate that FN-I\textsubscript{1-9} inhibits FN assembly by affecting not only FN–FN but also FN–αvβ3 interactions, which has profound consequences for assembly of FN-RGE. Collectively, these results revealed that the GNGRG loop in FN-I\textsubscript{5} represents a novel functional αvβ3 binding and FN fibril assembly site. Future experiments will be necessary to test whether additional motifs in FN-I\textsubscript{1-9} can also provide functional integrin binding and FN assembly sites (Shapiro et al., 2005).

**The GNKRG motif in FN-I\textsubscript{5} is a novel αvβ3 binding site that can function for FN matrix assembly**

The FN-I\textsubscript{1-9} fragment (also called 70-kD N-terminal fragment) inhibits FN assembly with efficiency similar to RGD peptides. It is generally believed that the inhibitory effect of FN-I\textsubscript{1-9} is caused by high-affinity binding sites for FN, which block the FN–FN interactions required to align and cross-link FNs into fibrils (McKeown-Longo and Mosher, 1985; McDonald et al., 1987; Schwarzbauser, 1991; Aguirre et al., 1994; Hocking et al., 1994; Sechler and Schwarzbauser, 1998). The fact that we could inhibit FN-RGE assembly with FN-I\textsubscript{1-9} pointed to the possibility that FN-I\textsubscript{1-9} may, in addition to its FN binding activity, contain an αv integrin binding site. We confirmed this in direct binding assays and found that FN-I\textsubscript{1-9} bound αvβ3. Curnis et al. (2006) reported that the Asn-Gly-Arg (NGR) sequence of FN-I\textsubscript{5} could be converted to a high-affinity binding site for αvβ3 integrin through the deamidation and rearrangement of the asparagine residue and the creation of an isoDGR sequence. Isoaspartate formation is a well-known, nonenzymatic process that can occur during aging of ECM proteins (David et al., 1998; Lanthier and Desroisiers, 2004) and during isolation and storage of proteins (Paranandi et al., 1994). The properties of the novel αvβ3 integrin binding sites in FN agreed with our direct binding assay, which showed that FN-I\textsubscript{1-9} bound αvβ3 and offered a potential explanation for the αv integrin–dependent and FN-I\textsubscript{1-9}–sensitive assembly of FN-RGE. Indeed, we found that an isoDGR peptide efficiently inhibited the binding of αvβ3 to FN-I\textsubscript{1-9} in our direct binding assay, and it blocked the assembly of FN-RGE by FN\textsuperscript{RG/GE} cells.

We found that assembly of the FN-RGE matrix was inhibited equally by peptides NGR-2C and isoDGR-2C. This suggests that the native NGR sequence itself may already be able to

The integrity of the RGD motif of FN is essential for development

The FN\textsuperscript{RG/GE} mice survive substantially longer than FN-null mice, indicating that FN-RGE fibrils are biologically active. FN\textsuperscript{RG/GE} embryos display defects that are strikingly similar to those observed in α5-null embryos (Yang et al., 1993), suggesting that high-affinity interactions of α5β1 with FN triggers unique functions during early development that cannot be compensated by αv integrin–RGD, αv integrin–NGR, or α5β1–FN\textsubscript{I1-9} interactions. In both mouse mutants, the posterior trunk is shortened, underdeveloped, and lacks somites. TUNEL assays and immunostaining for activated caspase-3 revealed diminished survival of posterior trunk mesoderm in FN\textsuperscript{RG/GE} mice, suggesting that RGD-triggered α5β1 integrin signals inhibit caspase-3 activation and anoikis.

A consistent feature of all FN\textsuperscript{RG/GE} embryos analyzed between E9.5 and E10.5 was that their somite number was limited to 12–13 somites. A plausible explanation for this finding could be that somites 12–13 represent the transition from the gastrulation- to tail bud–derived mesoderm (Wilson and Beddington, 1996). This would indicate that the integrin-triggered survival signals are mainly required for the posterior trunk- and tail bud–derived mesoderm. Such an α5β1 integrin independence of the anterior, streak-derived mesoderm could be explained by expression and compensation of αv integrins. Unfortunately, we were unable to test whether αv is high in the anterior and low or absent in the posterior mesoderm because all anti-αv antibodies that were available to us produced only background staining.

Zebrafish also require α5 integrin for somite development along the body axis. In contrast to the mouse, α5 integrin display anterior somite defects, suggesting
that α5 is indispensable for anterior somitogenesis, whereas the posterior somites developed normally (Julich et al., 2005; Koshida et al., 2005). Interestingly, knockdown of fibronectin1 in α5 integrin mutant zebrafish, as well as knockdown of fibronectin1 and fibronectin3, led to a more severe phenotype, suggesting that the restriction of defects in zebrafish to the anterior somites is probably due to functional redundancy (Julich et al., 2005; Koshida et al., 2005). A further difference from our findings is the absence of apoptosis in the anterior somites of α5 integrin−deficient zebrafish. It was even shown that the anterior somites develop perfectly well, but their boundaries cannot be maintained because of the loss of FN fibrils that would normally surround somite boundaries. In about half of our FN<sup>RGEO</sup> embryos, we observed normal amounts of FN-RGE fibrils at anterior somite boundaries, whereas in the remaining half, the levels were reduced (unpublished data).

In conclusion, we found that FN-RGE can be assembled into fibrils in vivo and in vitro via αvβ3 binding to the isoDGR motif in FN-I. The isoDGR site in FN-I<sub>5</sub> is generated by NGR deamidation (Curnis et al., 2006) and, as previously demonstrated, can readily be reversed by the enzymatic action of the protein l-isoaspartyl methyltransferase (Reissner and Aswad, 2003; Lanthier and Desroisiers, 2004). The ability of tissues to swiftly activate and deactivate binding of αvβ3 and likely other αv integrins to FN may represent a novel strategy to spatially and temporally trigger FN-mediated αv integrin signaling during development and disease.

**Materials and methods**

**Generation of knockin mice**

A 129/Sv mouse PAC (P1-derived artificial chromosome) clone was used to construct the FN-RGE targeting vector (Fig. S1 A). A 3.3-kb EcoRV–Stul fragment containing exons 28–30 was subcloned into BlueScript (Stratagene). A 0.8-kb SacI–Stul fragment containing exon 30 was subcloned and used for site-directed mutagenesis. The nucleotide C1238T of the GAC codon encoding the aspartic acid residue of the ArgGly-Asp (RGD) motif in exon 30 was converted with the QuickChange Mutagenesis kit (Stratagene) into a G, resulting in a GAG codon encoding glutamic acid (Carninci et al., 2006) and, as previously demonstrated, can readily be reversed by the enzymatic action of the protein l-isoaspartyl methyltransferase (Reissner and Aswad, 2003; Lanthier and Desroisiers, 2004). The ability of tissues to swiftly activate and deactivate binding of αvβ3 and likely other αv integrins to FN may represent a novel strategy to spatially and temporally trigger FN-mediated αv integrin signaling during development and disease.

Histology and immunofluorescence

To test FN expression, E9.5 embryos were dissected and either immediately embedded in optimal cutting temperature compound (OCT; Thermo Scientific) to obtain unfixed sections or fixed in 4% PFA and embedded in paraffin (Paraplast X-tra; Sigma-Aldrich) to obtain paraffin sections. Cryosections of 10-µm thickness and paraffin sections of 6-µm thickness were cut and used for histchemistry and immunofluorescence as described previously (Fässler and Meyer, 1995). TUNEL staining was performed according to the manufacturer's instructions (Roche). Antibodies against the following proteins were used: anti-FN (Chemicon), anti-α4 integrin (Biolegend), and anti–cleaved caspase-3 (New England Biolabs, Inc.). Cy3- or FITC-conjugated secondary antibodies were purchased from Sigma-Aldrich or Jackson ImmunoResearch Laboratories.

Cells were seeded on glass coverslips (Lab-Tek Chamber Slides; Nunc) precoated with 5 µg/ml VN (Sigma-Aldrich) or 10 µg/ml LM111 (a gift from R. Deutzmann, Regensburg University, Regensburg, Germany) for 2 h at 37°C, incubated at 37°C for 16–96 h, fixed with 4% PFA, and incubated with primary and secondary antibodies, respectively. In some cases, cells were permeabilized with 0.5% Triton X-100 after PFA fixation.

The following antibodies were used: anti-FN (Chemicon); anti-α5 (BD Biosciences); and Cy2. Cy3- and Cy5-conjugated secondary antibodies were used for detection (Jackson ImmunoResearch Laboratories). Nuclei were stained with DAPI (Roche). Fluorescent images were taken at room temperature with a microscope (DMRA2; Leica) equipped with NPLAN 40/0.65 NA objective and a camera (DMRA2, Leica), or with a confocal microscope (DMIRE2; Leica) equipped with a 100× 1.4 NA oil objective, using Leica Confocal Software (version 2.5, Build 1227) for data acquisition. Images were collected as TIF files and analyzed using Photoshop (version 9.0; Adobe).

**Western blotting**

E8.5 embryos were solubilized in RIPA buffer, briefly sonicated, and boiled for 2 min in nonreducing sample buffer. The samples were separated on 6% SDS polyacrylamide gels, blotted onto Immobilon-P polyvinylidene difluoride membranes (Millipore), and hybridized with polyclonal antibodies specific for FN (Chemicon) and tubulin (Sigma-Aldrich). Bound antibodies were hybridized with horseradish peroxidase–conjugated secondary antibodies and detected using the ECL kit (GE Healthcare).

**Generation of FN<sup>RGEO</sup> cell lines**

To establish cell lines, FN<sup>RGEO</sup> mice were intercrossed and embryos harvested at E9. To release cells, embryos were trypsinized for 5 min at 37°C and disrupted by pipetting up and down. Cells were cultured in DME supplemented with 10% FBS. Cells were genotyped by PCR, immortalized by retroviral transduction of the SV40 large T antigen, and cloned. Control (FN<sup>+/+</sup>) and FN<sup>RGEO</sup> cell lines were adopted to grow in serum replacement medium (47.5;47.5;5.5.1 ratio of DME/AIM-V Medium [Invitrogen]/ RPM1640/nonessential amino acids) and then used for FN assembly.

**Cell adhesion assays**

Cell adhesion assays were performed as described previously (Czuchra et al., 2006).

**Flow cytometry**

Flow cytometry was performed as described by Czuchra et al. (2006). The following antibodies were used: anti–β1, –β3, –α2, –α4, –α5, –α6, and –αv integrins (all obtained from BD Biosciences), which were FITC conjugated (anti–β1, –α2, and –α6 integrins), biotinylated (anti–β3, –α5, and –αv integrins), or PE conjugated (anti–α4 integrin). Biotinylated antibodies were detected with streptavidin-Cy5 (Jackson ImmunoResearch Laboratory). To assess autofluorescence and nonspecific staining, cells were stained with IgM isotype–FITC (IgM) and IgG2a isotype–FITC (IgG2a; BioLegend), and IgG2a isotype–PE (for α4 integrin), or streptavidin-Cy5 (β3, α5, and αv; all obtained from BD Biosciences).

**Generation of αv integrin-depleted cell lines**

The sequence of the siRNA constructs was chosen according to the protocol of Ui-Tei et al. (2004) and Naito et al. (2004). Full-length mRNA sequences of the target gene were checked using the online software siDirect for 19-bp stem-loop structure. The sequence obtained was synthesized as a 5′-terminally phosphorylated primer with a 5′-HindIII and 3′-BglII recognition site (S‘-CATCCCCGGTACGTTAAGTTTGATCTCGAAGAGAATCC-3‘). Subsequently, primers were annealed and inserted into HindIII-BamHI linearized pSuper.retro.puro (OligoEngine). The primer

**Generation of αv integrin-depleted cell lines**

The sequence of the siRNA constructs was chosen according to the protocol of Ui-Tei et al. (2004) and Naito et al. (2004). Full-length mRNA sequences of the target gene were checked using the online software siDirect for 19-bp stem-loop structure. The sequence obtained was synthesized as a 5′-terminally phosphorylated primer with a 5′-HindIII and 3′-BglII recognition site (S‘-CATCCCCGGTACGTTAAGTTTGATCTCGAAGAGAATCC-3‘). Subsequently, primers were annealed and inserted into HindIII-BamHI linearized pSuper.retro.puro (OligoEngine). The primer

**Generation of αv integrin-depleted cell lines**

The sequence of the siRNA constructs was chosen according to the protocol of Ui-Tei et al. (2004) and Naito et al. (2004). Full-length mRNA sequences of the target gene were checked using the online software siDirect for 19-bp stem-loop structure. The sequence obtained was synthesized as a 5′-terminally phosphorylated primer with a 5′-HindIII and 3′-BglII recognition site (S‘-CATCCCCGGTACGTTAAGTTTGATCTCGAAGAGAATCC-3‘). Subsequently, primers were annealed and inserted into HindIII-BamHI linearized pSuper.retro.puro (OligoEngine). The primer
Expression of FN fragments

The human FN-III7-10 fragment was subcloned into pET15b plasmid (Invitrogen) and mutated into the pET15b-FN-FN11-12-GRG by using the QuikChange II Site-Directed Mutagenesis kit. The following primers were used to introduce the mutation: RGD-＞RGE forward, 5′-CAC-TGGCGCTGAGAGAGCCCCCAAAGCAG3′; RGD-＞RGE reverse, 5′-CTG-CCTGGGGGCTCTCCTAGCCGCGCACTGA-3′; RGD-＞RGRG forward, 5′-GTGATGGCTGCAGCGAGGCAGGGACAGAAGCAGC-3′; and RGD-＞MRGD reverse, 5′-GCTTGGCCTGGGGGCCTGAGGAGTGCAGACA-GCATACAG-3′. The plasmids were transformed into BL21 (DE3)plysS Escherichia coli cells. Protein expression and purification of these and other fragments were performed as described previously (Ohashi and Erickson, 2005).

Inhibition assay

All experiments were performed in serum replacement medium. Control and FN-GRG/RGE cells were seeded on 10 μg/ml ML111-coated 8-well Lab-Tek Chamber Slides (80,000 cells/well), allowed to spread for 2 h, and incubated with inhibitory peptides dissolved in PBS for another 14 h at 37°C and 5% CO2. The assay was stopped by fixation with 4% PFA in PBS and immunostained.

The final concentrations of the used peptides were as follows: linRGD and linRGE, 0.5 mg/ml (900 μM; BIOMOL Research Laboratories, Inc.); cycRGD and cycRGRG, 0.25 mg/ml (410 μM; BIOMOL Research Laboratories, Inc.); isocycRGD-2C and NGR-2C, 800 μM (for synthesis, see Peptide synthesis on TCP resin using the Fmoc strategy. Fmoc-Asp-Oiso methylpyrrolidinone for 20 min. The linear peptide was cleaved by using the QuikChange II XL Site-Directed Mutagenesis kit. The following primers were used Pbf protected. To avoid positive charge at the N terminus and to facilitate HPLC purification by enhancing the UV absorption, the terminal was used as RGD-＞MRGD.

Direct solid-phase binding assay using purified recombinant integrins and recombinant FN fragments

Direct solid-phase assay for determining the relative binding of biotinylated αvβ3 integrins to various FN fragments was performed as described previously (Eble et al., 1998). In brief, 0.25 μM solutions of recombinant FN fragments in TBS were used to coat 96-well polyvinylchloride microtiter plates (Maxisorp; Nunc) by an overnight incubation at 4°C. Coating with 0.25 μg/ml BSA was used to determine the background values °C. In the fol-

References

Aguirre, K.M., R.J. McCormick, and J.E. Schwarzbauer. 1994. Fibronectin self-association is mediated by complementary sites within the amino-terminal one-third of the molecule. J. Biol. Chem. 269:27863–27868.

Bader, B.L., H. Rayburn, D. Crowley, and R.O. Hynes. 1998. Extensive vasculogenesis, angiogenesis, and organogenesis precede lethality in mice lacking all αv integrins. Cell. 95:507–519.

Bouvard, D., C. Brakebusch, E. Gustafsson, A. Aszodi, T. Bengtsson, A. Berna, and R. Fassler. 2001. Functional consequences of integrin gene mutations affecting α5β1 and α5β3. J. Cell. Biol. 155:211–221.

Bultmann, H., A.J. Santos, and D.M. Peters. 1998. Fibronectin fibrillogenesis involves the heparin II binding domain of fibronectin. J. Biol. Chem. 273:2601–2609.

Curnis, F., R. Longhi, L. Crippa, A. Cattaneo, E. Dondossola, A. Bachi, and A. Corti. 2006. Spontaneous formation of L-isoaspartate and gain of function in fibroblasts. J. Biol. Chem. 281:36466–36476.

Cruzhia, A., H. Meyer, K.R. Legate, C. Brakebusch, and R. Fassler. 2006. Genetic analysis of β1 integrin activation motifs” in mice. J. Cell Biol. 174:889–899.

Dane, E.H., P. Sonneveld, C. Brakebusch, R. Fassler, and A. Sonnenberg. 2002. The fibronectin-binding integrins α5β1 and αvβ3 differentially modulate RhoA-GTP loading, organization of cell matrix adhesions, and fibronectin fibrillogenesis. J. Cell Biol. 159:1071–1086.

Darrbire, T., K.M. Yamada, K.E. Johnson, and J.C. Boucaut. 1988. The 140-kDa fibronectin receptor complex is required for mesodermal cell adhesion during gastrulation in the amphibian Pleurodeles waltlisi. Dev. Biol. 126:182–194.

David, C.L., I. Orpiszewski, X.C. Zhu, K.J. Reissner, and D.W. Aswad. 1998. Isoaspartate in chondroitin sulfate proteoglycans of mammalian brain. J. Biol. Chem. 273:32063–32070.

Di Matteo, P., F. Curnis, R. Longhi, G. Colombo, A. Sacchi, L. Crippa, M.P. Pottti, M. Ponzoni, S. Toma, and A. Corti. 2006. Immunogenic and structural properties of the Asn-Gly-Arg (NGR) tumor neovasculature-homing motif. Mol. Immunol. 43:1509–1518.

Eble, J.A., K.W. Wucherpfennig, L. Gauthier, P. Dersch, E. Krukonis, R.R. Isberg, and M.E. Hemler. 1998. Recombinant soluble human α3β1 integrin: purification, processing, regulation, and specific binding to laminin-5 and invasion in a mutually exclusive manner. Biochemistry. 37:10945–10955.

Fassler, R., and M. Meyer. 1995. Consequences of lack of β1 integrin gene expression in mice. Genes Dev. 9:1906–1908.
Scheller, J.L., S.A. Corbett, and J.E. Schwarzbauer. 1997. Modulatory roles for integrin activation and the synergy site of fibronectin during matrix assembly. Mol. Biol. Cell. 8:2563–2573.

Scheller, J.L., A.M. Cimiskey, D.M. Gazzola, and J.E. Schwarzbauer. 2000. A novel RGD-independent fibronectin assembly pathway initiated by αβ1 integrin binding to the alternatively spliced V region. J. Cell Sci. 113:1491–1498.

Shapiro, N., I.R. Ellis, T.J. Dines, A.M. Schor, S.L. Schor, D.G. Norman, and R. Marquez. 2005. Synthesis of an IGD peptidomimetic with motogenic activity. Mol. Biolyst. 1:318–320.

Sottile, J., D.C. Hocking, and K.J. Langenbach. 2000. Fibronectin polymerization stimulates cell growth by RGD-dependent and -independent mechanisms. J. Cell Sci. 113:4287–4299.

Takagi, J., B.M. Petre, T. Walz, and T.A. Springer. 2002. Global conformational rearrangements in integrin extracellular domains in outside-in and inside-out signaling. Cell. 110:599–611.

UeI-Tei, K., Y. Naito, F. Takahashi, T. Haraguchi, H. Ohki-Hamazaki, A. Juni, R. Ueda, and K. Saigo. 2004. Guidelines for the selection of highly effective siRNA sequences for mammalian and chick RNA interference. Nucleic Acids Res. 32:936–948.

Wennerberg, K., L. Lohikangas, D. Gullberg, M. Pfaff, S. Johansson, and R. Fassler. 1996. β1 integrin-dependent and -independent polymerization of fibronectin. J. Cell Biol. 132:227–238.

Wierzbicka-Patynowski, I., and J.E. Schwarzbauer. 2003. The ins and outs of fibronectin matrix assembly. J. Cell Sci. 116:3269–3276.

Wilson, V., and R.S. Beddington. 1996. Cell fate and morphogenetic movement in the late mouse primitive streak. Mech. Dev. 55:79–89.

Wu, C., V.M. Keivens, T.E. O’Toole, J.A. McDonald, and M.H. Ginsberg. 1995. Integrin activation and cytokineskeletal interaction are essential for the assembly of a fibronectin matrix. Cell. 83:715–724.

Wu, C., P.E. Hughes, M.H. Ginsberg, and J.A. McDonald. 1996. Identification of a new biological function for the integrin αvβ3: initiation of fibronectin matrix assembly. Cell Adhes. Commun. 4:149–158.

Yang, J.T., H. Rayburn, and R.O. Hynes. 1993. Embryonic mesodermal defects in α5 integrin-deficient mice. Development. 119:1093–1105.

Yang, J.T., B.L. Bader, J.A. Keisberg, M. Ullman-Cullere, J.E. Trevithick, and R.O. Hynes. 1999. Overlapping and independent functions of fibronectin receptor integrins in early mesodermal development. Dev. Biol. 215:264–277.

Fath, K.R., C.I. Edgell, and K. Burridge. 1989. The distribution of distinct integrins in focal contacts is determined by the substratum composition. J. Cell Sci. 92:67–75.

George, E.L., E.N. Georges-Labouesse, R.S. Patel-King, H. Rayburn, and R.O. Hynes. 1993. Defects in mesoderm, neural tube and vascular development in mouse embryos lacking fibronectin. Development. 119:1079–1091.

Harrison, F., L. Van Nassauw, J. Van Hoof, and J.M. Fvidart. 1993. Microinjection of anti-fibronectin antibodies in the chicken blastoderm: inhibition of mesoblast cell migration but not of cell ingestion at the primitive streak. Anat. Rec. 236:685–696.

Hocking, D.C., J. Sottile, and P.J. McKeown-Longo. 1994. Fibronectin’s III-1 module contains a formation-dependent binding site for the amino-terminal region of fibronectin. J. Biol. Chem. 269:19183–19187.

Hynes, R.O. 1992. Integrins: versatility, modulation, and signaling in cell adhesion. Cell. 69:11–25.

Julich, D., R. Geisler, and S.A. Holly. 2005. Integrin α5 and delta/notch signaling have complementary spatiotemporal requirements during zebrafish somitogenesis. Dev. Cell. 8:575–586.

Kombliitt, A.R., C.G. Pesce, C.R. Alonso, P. Cramer, A. Srebow, S. Werbjah, and A.F. Muro. 1996. The fibronectin gene as a model for splicing and transcription studies. FASEB J. 10:248–257.

Koshida, S., Y. Kishimoto, H. Ustumi, T. Shimizu, M. Furutani-Seiki, H. Kondoh, and S. Takada. 2005. Integrin α5-dependent fibronectin accumulation for maintenance of somite boundaries in zebrafish embryos. Dev. Cell. 8:587–598.

Lanthier, J., and R.R. Desrosiers. 2004. Protein L-isooaspartyl methyltransferase repairs abnormal aspartyl residues accumulated in vivo in type-I collagen and restores cell migration. Exp. Cell Res. 293:96–105.

McDonald, J.A., H.J. Quade, T.J. Broekelmann, R. LaChance, K. Forsman, E. Haegawa, and S. Akiyama. 1987. Fibronectin’s cell-adhesive domain and an amino-terminal matrix assembly domain participate in its assembly into fibroblast pericellular matrix. J. Biol. Chem. 262:2957–2967.

McKeown-Longo, P.J., and D.F. Mosher. 1984. Mechanism of formation of disulfide-bonded multimers of plasma fibronectin in cell layers of cultured human fibroblasts. J. Biol. Chem. 259:12210–12215.

McKeown-Longo, P.J., and D.F. Mosher. 1985. Interaction of the 70,000-mol-wt amino-terminal fragment of fibronectin with the matrix-assembly receptor of fibroblasts. J. Cell Biol. 100:364–374.

Nagai, T., N. Yamakawa, S. Aota, S.S. Yamada, S.K. Akiyama, K. Olden, and K.M. Yamada. 1991. Monoclonal antibody characterization of two distant sites required for function of the central cell-binding domain of fibronectin in cell adhesion, cell migration, and matrix assembly. J. Cell Biol. 114:1295–1305.

Naito, Y., T. Yamada, K. Ui-Tei, S. Morishita, and K. Saigo. 2004. siDirect: highly effective, target-specific siRNA design software for mammalian RNA interference. Nucleic Acids Res. 32:W124–W129.

Ohashi, T., and H.P. Erickson. 2005. Domain unfolding plays a role in superfibronection formation. J. Biol. Chem. 280:39143–39151.

Pankov, R., and K.M. Yamada. 2002. Fibronectin at a glance. J. Cell Sci. 115:3861–3863.

Paranandi, M.V., A.W. Guzzetta, W.S. Hancock, and D.W. Aswad. 1994. Deamidation and isoaspartate formation during in vitro aging of recombinant tissue plasminogen activator. J. Biol. Chem. 269:243–253.

Pfaff, M., K. Tangemann, B. Muller, M. Gurrath, G. Muller, H. Kessler, R. Timpl, and J. Engel. 1994. Selective recognition of cyclic RGD peptides of NMR defined conformation by αβ1β3, αvβ3, and α5β1 integrins. J. Biol. Chem. 269:20233–20238.

Pfeifer, A., T. Kessler, S. Siletto, D.A. Cheres, and J.M. Verma. 2000. Suppression of angiogenesis by lentiviral delivery of PEX, a noncatalytic fragment of matrix metalloproteinase 2. Proc. Natl. Acad. Sci. USA. 97:12227–12232.

Pierschbacher, M.D., and E. Ruoslahti. 1984. Cell attachment activity of fibronectin can be duplicated by small synthetic fragments of the molecule. Nature. 309:30–33.

Plow, E.F., T.A. Haas, L. Zhang, J. Lofus, and J.W. Smith. 2000. Ligand binding to integrins. J. Biol. Chem. 275:21785–21788.

Reissner, K.J., and D.W. Aswad. 2003. Deamidation and isoaspartate formation in proteins: unwanted alterations or surreptitious signals? Cell. Mol. Life Sci. 60:1281–1295.

Robinson, N.E., Z.W. Robinson, B.R. Robinson, A.L. Robinson, J.A. Robinson, M.L. Robinson, and A.B. Robinson. 2004. Structure-dependent nomenclature deamidation of glutaminyl and asparaginyl pentapeptides. J. Pept. Res. 63:426–436.

Schwarzbauer, J.E. 1991. Identification of the fibronectin sequences required for assembly of a fibrillar matrix. J. Cell Biol. 113:1463–1473.

Sechler, J.L., and J.E. Schwarzbauer. 1998. Control of cell cycle progression by fibronectin matrix architecture. J. Biol. Chem. 273:25533–25536.