Regulation of glutamate receptor subunit availability by microRNAs

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The efficacy of synaptic transmission depends, to a large extent, on postsynaptic receptor abundance. The molecular mechanisms controlling receptor abundance are poorly understood. We tested whether abundance of postsynaptic glutamate receptors (Glus) in Drosophila neuromuscular junctions is controlled by microRNAs, and provide evidence that it is. We show here that postsynaptic knockdown of dicer-1, the endoribonuclease necessary for microRNA synthesis, leads to large increases in postsynaptic GluR subunit messenger RNA and protein. Specifically, we measured increases in GluRIIA and GluRIIB but not GluRIIC. Further, knockout of MiR-284, a microRNA predicted to bind to GluRIIA and GluRIIB but not GluRIIC, increases expression of GluRIIA and GluRIIB but not GluRIIC proportional to the number of predicted binding sites in each transcript. Most of the de-repressed GluR protein, however, does not appear to be incorporated into functional receptors, and only minor changes in synaptic strength are observed, which suggests that microRNAs primarily regulate Drosophila receptor subunit composition rather than overall receptor abundance or synaptic strength.

Introduction

Most synaptic transmission in mammalian brains occurs via glutamate receptors (Glus). Synthesis and localization of Glus is essential for brain development, and Glu changes underlie learning, memory, and many different neuropathologies. Several lines of evidence suggest that Glu synthesis may be regulated by microRNAs, which suppress gene expression in a variety of organisms and tissue types (Lee et al., 2007; Baek et al., 2008; Selbach et al., 2008). First, microRNAs are particularly abundant in brains, and have recently been shown to play important roles in several aspects of nervous system development (Cao et al., 2006; Bicker and Schratt, 2008; Fiore et al., 2008). Second, some postsynaptic proteins, possibly including GluR subunits, seem to be locally translated near postsynaptic densities (Sigrist et al., 2000; Job and Eberwine, 2001; Miller et al., 2002; Sutton et al., 2004; Sutton and Schuman, 2005; Martin and Zukin, 2006; Pfeiffer and Huber, 2006; Schuman et al., 2006; Bramham, 2008), whereas the microRNA synthesis protein dicer and many microRNAs themselves may be enriched at synapses (Lugli et al., 2005, 2008). Finally, acetylcholine receptor abundance in Caenorhabditis elegans neuromuscular junctions (NMJs) has recently been shown to be regulated by microRNAs (Simon et al., 2008).

Here, we test whether GluR synthesis in Drosophila melanogaster is regulated by microRNAs. Drosophila NMJs are glutamatergic synapses that are individually identifiable at the cellular level and experimentally accessible throughout development in vivo (Keshishian et al., 1996; Featherstone and Broadie, 2004; Ruiz-Canada and Budnik, 2006). Drosophila NMJs contain two biophysically, pharmacologically, and spatially distinct subtypes of ionotropic Glus (DiAntonio et al., 1999; Marrus et al., 2004; Chen et al., 2005; Featherstone et al., 2005; Qin et al., 2005), called “A type” and “B type.” A-type receptors contain the subunit GluRIIA but not GluRIIB, plus GluRIIC, GluRIID, and GluRIIE. B-type receptors contain GluRIIB but not GluRIIA, plus GluRIIC, GluRIID, and GluRIIE.

Our results show that microRNAs strongly regulate the abundance of GluRIIA and GluRIIB Glur subunit mRNAs, and that this has dramatic effects on relative subunit protein abundance. In the absence of microRNA-mediated subunit repression,
GluRIIA and GluRIIB proteins accumulate in the cell; in the absence of a particular microRNA—\textit{MiR-284}—GluR subunits accumulate such that receptor subtype ratios in the synapse are dramatically changed.

**Results**

Because microRNAs bind to target transcripts via base pairing, a variety of computational methods have been developed that compare the sequence of putative microRNAs with predicted transcripts (Rajewsky, 2006; Watanabe et al., 2007; Bartel, 2009). If appropriate complementarity is detected, an interaction between the microRNA and transcript is presumed to occur in vivo. To determine whether \textit{Drosophila} NMJ GluR transcripts might be regulated by microRNAs and identify candidate microRNAs for each subunit transcript, we bioinformatically screened the entire sequence of \textit{GluRIIA}, \textit{GluRIIB}, \textit{GluRIIC}, \textit{GluRIID}, and \textit{GluRIIE} transcripts for potential microRNA-binding sites. We screened the entire transcript because although it was initially thought that microRNAs selectively associated with untranslated regions (UTRs) to inhibit target translation, microRNAs might also trigger target transcript degradation by binding to complementary sites wherever they might be found in a transcript.

Unfortunately, although we tried several different tools representing several different approaches (Enright et al., 2003; Stark et al., 2003; Brennecke et al., 2005; Krek et al., 2005; Rusinov et al., 2005; Sethupathy et al., 2006), our bioinformatics experiments did not converge toward any usable predictions. To circumvent this problem, we considered a different approach. MicroRNA synthesis requires cleavage of stem-loop precursor RNAs by a protein complex containing the type III ribonuclease “dicer” (Bernstein et al., 2001; Hutvagner et al., 2001). Indeed, the property of being processed by dicer is one of the classical definitions of a microRNA (Berezikov et al., 2006). Therefore, if GluR transcripts are regulated by microRNAs, then they must also regulated (indirectly) by dicer. Specifically, eliminating dicer function should increase GluR protein abundance if GluR transcripts are regulated by microRNAs. Elimination of dicer should also cause an increase in GluR mRNA abundance if microRNAs regulate GluR transcript stability (as opposed to translation).

The \textit{Drosophila} genome encodes two dicer paralogues: dicer-1 and dicer-2. Dicer-1 and dicer-2 interact with different sets of proteins and have functionally distinct roles in vivo: dicer-1 preferentially cleaves precursor microRNAs after they are exported from the nucleus, whereas dicer-2 preferentially cleaves exogenous double-stranded RNA, presumably as a defense against viral infection (Lee et al., 2004). Elimination of dicer-1 function should therefore prevent synthesis of \textit{Drosophila} microRNAs, allowing us to determine whether NMJ GluRs are regulated by microRNAs. Unfortunately, consistent with the fact that dicer is required for early development in every complex organism examined to date (Giraldez et al., 2005; Murchison et al., 2005), \textit{Drosophila} dicer-1 mutants (\textit{Dcr-1[d102]} and \textit{Dcr-1[1147X]}) died in embryogenesis before proper NMJ formation and GluR expression. We also examined “semieviable” \textit{ago1[04845]} mutants. Argonaute 1 (AGO1), like dicer-1, is thought to be preferentially required for microRNA synthesis (Okamura et al., 2004). However, homozygous \textit{ago1[d04845]} mutants, like dicer mutants, also died before normal NMJ formation and GluR expression.

As a result, we were unable to use dicer-1 (or AGO1) mutants to test whether NMJ GluRs are regulated by microRNAs. Such an analysis would have been, in any case, complicated by possible presynaptic microRNA-dependent changes affecting transsynaptic induction of receptor expression (Broadie and Bate, 1993). However, the previously described segregation of dicer-1 and dicer-2 function in \textit{Drosophila} (Lee et al., 2004) suggested another possible approach. If dicer-1 is preferentially required for microRNA synthesis, but dicer-2 is preferentially required for processing of exogenous double-stranded RNA (Lee et al., 2004), then it should be possible to knock down dicer-1 function via RNAi (Fig. 1 A). RNAi knockdown of dicer-1 might be more permissive for development (and thus study of GluR expression) than mutants for two reasons: (1) RNAi typically reduces, but does not eliminate, target function; and (2) RNAi transgene expression in \textit{Drosophila} can be temporally and spatially restricted using the \textit{Gal4–UAS} system (Brand and Perrimon, 1993), allowing us to knock down dicer-1 only in postsynaptic muscle cells while leaving all other tissues as wild type.

To test this approach, we generated a \textit{UAS–dicer-1 RNAi} transgene. Global expression of this transgene using a tubulin Gal4 driver (\textit{TubGal4/UAS-dcr1.RNAi}; Lee and Luo, 1999) caused early embryonic lethality, similar to that observed in dicer-1 and AGO1 mutants. In contrast, expression of dicer-1 RNAi specifically in postsynaptic body wall muscles (\textit{24BGal4/UAS-dcr1.RNAi}) allowed animals to hatch, feed, and develop through third instar larval stages. To confirm that body wall muscle dicer-1 was knocked down in these animals, and to quantify that knockdown, we examined dicer-1 antibody immunoreactivity in third instar larval body wall tissues (Fig. 1, B and C). In control animals, dicer-1 immunoreactivity was prominent as distinct puncta scattered throughout body wall muscle cells (Fig. 1 B). Presumably, these puncta represent micro ribonucleoprotein complexes, with which dicer proteins are thought to associate. However, we also observed dicer-1 immunoreactivity as a faint “haze” distributed throughout the muscle cells (Fig. 1 B). Body wall muscles expressing dicer-1 RNAi showed a dramatic decrease in both haze intensity and number of dicer-1 immunoreactive puncta, which is consistent with elimination of approximately two thirds of muscle dicer-1 (Fig. 1, B and D).

MicroRNAs can repress gene expression by decreasing target transcript abundance or by suppressing translation. To test whether NMJ GluR transcript abundance is dependent on dicer-1 (and thus regulated by microRNAs), we measured GluRIIA and GluRIIB transcript abundance (using quantitative real-time RT-PCR and FISH) in control animals and in animals expressing \textit{dicer-1 RNAi} specifically in postsynaptic body wall muscles (\textit{24BGal4/UAS-dcr1.RNAi}). We focused on these two transcripts because GluRIIA and GluRIIB are thought to be rate-limiting for formation of A- and B-type GluRs, respectively;
and GluRIIA and GluRIIB are expressed only in body wall muscle (DiAntonio et al., 1999). Muscle-specific knockdown of dicer-1 resulted in large statistically significant increases in GluRIIA and GluRIIB transcript abundance, as measured by real-time RT-PCR, which is consistent with the idea that stability of both transcripts is regulated by microRNAs (Fig. 2 A).

To confirm these results, we used FISH to visualize GluRIIA and GluRIIB mRNA in body wall muscles of control larvae and larvae expressing dicer-1 RNAi (Fig. 2 C). Consistent with our real-time RT-PCR results, FISH showed a large increase in GluRIIA and GluRIIB mRNA abundance after muscle-specific expression of dicer-1 RNAi (Fig. 2 B). We conclude, based on these results, that GluRIIA and GluRIIB transcripts are targets for microRNA-mediated repression, and that the microRNAs that repress GluRIIA and GluRIIB do so, at least in part, by reducing target transcript abundance. Presumably, this also affects GluRIIA and GluRIIB protein abundance (Fig. 2 D).

To test whether microRNAs regulate GluRIIA and GluRIIB protein abundance, we stained larval NMJs using antibodies against GluRIIA and GluRIIB (Fig. 3). In control animals, most GluRIIA and GluRIIB protein is tightly localized at synapses, which is consistent with the role of A- and B-type receptors in NMJ transmission. In contrast, very little GluRIIA and GluRIIB immunoreactivity is detectable in wild-type non-synaptic muscle cell regions. This small amount of immunoreactivity is consistent with a low density of functional non-synaptic
dissected 24BGal4/UAS-dcr1.RNAi larvae were briefly incubated with the fixable nonpermeant membrane dye FM1-43FX, in order to label plasma membranes. These preparations were then fixed and stained with GluRIIA and GluRIIB antibodies to label GluRIIA and GluRIIB protein. To determine the spatial relationship of receptor subunit proteins and the muscle plasma membranes, we imaged labeled preparations by confocal microscopy and generated 3D reconstructions of the neuromusculature (Fig. 4). As expected, we observed a high concentration of GluRIIA and GluRIIB immunoreactivity at NMJs (Fig. 4 A, arrows), which likely represents functional synaptic A- and B-type GluRs. There was also a relatively low density of GluRIIA and GluRIIB associated with extrasynaptic plasma membrane, GluRs measured in previous studies (Nishikawa and Kidokoro, 1995; Featherstone et al., 2000). In 24BGal4/UAS-dcr1.RNAi larvae, the density of GluRIIA and GluRIIB immunoreactivity at synapses was not different from controls, which was consistent with our electrophysiological results. However, we observed dramatically high levels of nonsynaptic GluRIIA and GluRIIB immunoreactivity in 24BGal4/UAS-dcr1.RNAi larvae (Fig. 3). Most of this nonsynaptic immunoreactivity was punctate, similar in size and shape to that of synaptic GluR clusters.

To test whether the nonsynaptic GluRIIA and GluRIIB protein puncta observed after expression of dicer-1 RNAi represented (possibly functional) surface receptors, we used immunocytochemistry and confocal microscopy (Fig. 4). Manually
Intracellular GluRIIA and GluRIIB could represent assembled GluRs unable to traffic to plasma membranes, or they could represent subunit protein that is not assembled into a receptor complex. The former suggests that GluRs are assembled early and then distributed throughout postsynaptic cells, whereas the latter suggests that unassembled GluR protein might be widely distributed before being assembled into multimeric receptors. To determine whether extrasynaptic intracellular GluRIIA and GluRIIB immunoreactivity might represent assembled but intracellular GluRs, we examined immunoreactivity for the essential receptor subunit GluRIIC (also known as GluRIII; Marrus et al., 2004). GluRIIC is a required subunit in which is consistent with the presence of some functional extrasynaptic receptors. However, the vast majority of GluRIIA and GluRIIB immunoreactivity in 24BGal4/UAS-dcr1.RNAi larval muscles was distributed inside the thin layer of FM1-43 fluorescence representing the muscle plasma membrane (Fig. 4, A, B, D, and E), which suggests that the ectopic GluRIIA and GluRIIB protein observed in this genotype is mainly in the interior of muscle cells. We conclude, based on this evidence, that although dicer-1 knockdown dramatically increases abundance of GluRIIA and GluRIIB protein, most of the de-repressed excess GluRIIA and GluRIIB is nonfunctional and distributed throughout the interior of the muscle cells.
However, 24BGal4/UAS-dcr1.RNAi larvae showed a small but significant increase in sEJC amplitude, which suggests a slight increase in the number of postsynaptic GluRs (Fig. 6, C and D). To specifically check for changes in the number of functional nonsynaptic receptors, we pressure-ejected 1 mM glutamate onto extrasynaptic muscle membrane in control and 24BGal4/UAS-dcr1.RNAi larvae. The amplitude of nonsynaptic GluR currents was not increased in 24BGal4/UAS-dcr1.RNAi larvae, compared with controls (Fig. 7).

We conclude, based on our electrophysiology, that dicer-1 knockdown has very little effect on total numbers of functional postsynaptic GluRs, and no effect on the number of functional nonsynaptic receptors. These results are consistent with our immunocytochemical results (Figs. 2–5).

The data described above suggest that: (1) Drosophila GluRs are indeed regulated by microRNAs; (2) this regulation occurs, at least in part, via degradation of GluRIIA and GluRIIB subunit transcripts; and (3) microRNA-mediated regulation of GluRIIA and GluRIIB may be most important for control of receptor assembly and/or subunit composition rather than synaptic strength. However, these data do not address the possibility that regulation of GluRIIA and GluRIIB occurs indirectly (via regulation of an unknown transcript that in turn regulates GluR transcript abundance and/or receptor assembly). Nor do the data reconcile the apparent discrepancy between bioinformatic predictions and the both A- and B-type GluRs (Marrus et al., 2004). Therefore, if GluRIIC immunoreactivity in 24BGal4/UAS-dcr1.RNAi larvae is not associated with nonsynaptic GluRIIA or GluRIIB immunoreactivity, then the GluRIIA and GluRIIB immunoreactivity cannot represent assembled receptors. As shown in Fig. 5, GluRIIC immunoreactivity in 24BGal4/UAS-dcr1.RNAi larvae matched that observed in controls. We therefore conclude that the abundant nonsynaptic intracellular GluRIIA and GluRIIB protein observed in 24BGal4/UAS-dcr1.RNAi larvae (Figs. 3 and 4) represents GluR subunits unable to multimerize, perhaps because of the relatively restricted availability of other required subunits.

To confirm our immunocytochemical results, we turned to voltage-clamp electrophysiology (Fig. 6). Electrophysiology is extremely sensitive to changes in functional GluR abundance and distribution; in the Drosophila larval NMJ, differences of only a few dozen individual postsynaptic receptors can be reliably detected and statistically differentiated using this method. First, we measured total synaptic strength by stimulating the presynaptic motor nerve and measuring the amplitude of the resulting postsynaptic excitatory junction currents (EJCs). EJCs in 24BGal4/UAS-dcr1.RNAi larvae were slightly, but not significantly, larger than those in controls (Fig. 6 A). The frequency of spontaneous EJCs (sEJCs) was not significantly altered in 24BGal4/UAS-dcr1.RNAi larvae compared with controls (Fig. 6 B).
Interestingly, none of the predicted microRNA-binding sites in each genotype. Error bars represent SEM. n.s., not significant.

phenotypes observed in 24BGal4/UAS-dcr1.RNAi larvae. To address these problems, we returned to bioinformatics.

The lack of the GluRIIC phenotype in 24BGal4/UAS-dcr1.RNAi larvae indicated that GluRIIC transcripts are not regulated by microRNAs in vivo. Given this, we reasoned that it might be possible to fine-tune bioinformatic search parameters such that the algorithm could simultaneously predict microRNA-bindings sites in GluRIIA and GluRIIB, which is consistent with our experimental results, but no (or relatively few) microRNA-binding sites in GluRIIC, which is also consistent with our experimental results. Many published algorithms predict no microRNA-binding sites in GluRIIA, GluRIIB, or GluRIIC transcripts, which suggests that these algorithms are too restrictive. Several of these algorithms assume highly complementary microRNA–target binding, which has subsequently been shown to not be absolutely required for microRNA-mediated transcript inhibition (Rajewsky, 2006). In contrast, the “MicroInspector” algorithm of Rusinov et al. (2005) is designed to identify all possible sites of microRNA-target interaction, which can then be sorted according to predicted strength. Using default/recommended parameters (25°C hybridization, −26 kcal/mol free energy cutoff), MicroInspector (version 1.0) predicted 106 binding sites for 43 different Drosophila microRNAs in the GluRIIA transcript, 91 binding sites for 43 different microRNAs in the GluRIIB transcript, and 132 binding sites for 45 different microRNAs in the GluRIIC transcript (see Tables S1–S3). The predicted binding sites in each case were scattered throughout the entire transcript (Tables S1–S3).

To restrict the parameters such that MicroInspector’s predictions were more consistent with our experimental results, we sorted the entire set of predicted interactions according to the calculated free energy of binding, and selected parameters that would predict microRNA-binding sites in GluRIIA and GluRIIB but not GluRIIC. Specifically, we found that 20°C hybridization and a more restrictive free energy cutoff of −37.1 kcal/mol predicted two binding sites (for MiR-284 and MiR-306) in GluRIIA, two binding sites (both for MiR-284) in GluRIIB, and zero binding sites in GluRIIC (Fig. 8 A). Interestingly, none of the predicted binding sites in GluRIIA or GluRIIB were in UTRs. However, there is no a priori reason to assume that microRNA-binding sites must be in UTRs, as the dicer-dependent loss of GluRIIA and GluRIIB transcript that we observed could theoretically be triggered by microRNA binding to any region of the transcript, including protein-coding sequences.

If MiR-284 regulates GluRIIA and GluRIIB, then MiR-284 loss-of-function mutants should show large increases in GluRIIA and GluRIIB similar to or (more likely) larger than those observed after dicer-1 knockdown. To test this, we examined GluRIIA and GluRIIB protein distribution in homozygous Df(3R)kar-Sz12 mutant larvae, in which the MiR-284 transcript, which is consistent with our experimental predictions (Fig. 8 A). Specifically, nonsynaptic GluRIIA and GluRIIB similar to or (more likely) larger than those observed after dicer-1 knockdown. To test this, we examined GluRIIA and GluRIIB protein distribution in homozygous Df(3R)kar-Sz12 mutant larvae, in which the MiR-284 gene is deleted. Homozygous Df(3R)kar-Sz12 mutant larvae were 100% prepupal lethal, but a small fraction (<1%) of animals survived to the third instar larval stage. The behavior and gross morphology of Df(3R)kar-Sz12 animals appeared normal by light microscopy, except for a tendency toward slightly swollen bodies and increased “bursting” when cuticles were initially penetrated for dissection, which suggests increased hemolymph pressure or weaker body walls. Importantly, light microscopic examination of dissected Df(3R)kar-Sz12 larvae showed relatively normal neuromuscular anatomy.

As shown in Fig. 8 B, homozygous Df(3R)kar-Sz12 mutant larvae showed dramatic increases in nonsynaptic GluRIIA and GluRIIB protein, which is consistent with the revised bioinformatic predictions (Fig. 8 A). Specifically, nonsynaptic GluRIIA was increased ~500%, and nonsynaptic GluRIIB was increased ~2,800% (Fig. 8 C). As with knockdown of dicer-1 by RNAi, synaptic GluRIIA was not increased in Df(3R)kar-Sz12 larvae.

Figure 5. Knockdown of dicer-1 does not alter abundance or distribution of GluRIIC protein. GluRIIC protein distribution is normal after muscle-specific knockdown of dicer-1; all GluRIIC remains tightly localized to NMJs in both control and 24BGal4/UAS–dicer-1.RNAi larvae. (bottom) Quantification from preparations like those shown in the micrographs (n = 10–15 for each genotype). Error bars represent SEM. n.s., not significant.
As shown in Fig. 9 (A–C), transgenic expression of MiR-284 in both types of MiR-284 deletion mutant backgrounds rescued GluRIIA and GluRIIB overexpression phenotypes to near control levels. Expression of the MiR-284 transgene also rescued both homozygous Df(3R)kar-Sz12 mutants and Df(3R)kar-Sz12/Df(3R)exel7317 mutants to adulthood, which suggests that loss of MiR-284 was by itself responsible for a significant portion of the lethality in these genotypes. Both homozygous Df(3R)kar-Sz12 mutants and Df(3R)kar-Sz12/Df(3R)exel7317 mutants showed 100% preadult lethality in the absence of the MiR-284 transgene.

Significantly, the relatively large increase in GluRIIB abundance relative to GluRIIA abundance that we observed in synaptic receptors after loss of MiR-284 (Figs. 8 C and 9 B) was also completely rescued, which suggests that MiR-284 controls postsynaptic GluR subtype ratios. To evaluate this, we computed postsynaptic GluR subtype ratios from unnormalized immunocytochemical data and used them to create pie charts representing relative proportions of A-type (green) and B-type (red) receptors at the NMJ. As shown (Fig. 9 D), wild-type NMJs are slightly enriched for A-type receptors. But after MiR-284 deletion (mean of Df(3R)kar-Sz12 and Df(3R)kar-Sz12/Df(3R)exel7317 phenotypes), the NMJ becomes enriched for Synaptic GluRIIB immunoreactivity, however, was increased ∼250% in Df(3R)kar-Sz12 larvae (Fig. 8 C). In contrast to GluRIIA and GluRIIB, the abundance and distribution of GluRIIC immunoreactivity in Df(3R)kar-Sz12 larvae matched that of wild-type controls (Fig. 8 D), which is consistent with the conclusion that GluRIIC mRNA is not regulated by MiR-284. Interestingly, microRNA-mediated repression of GluRIIA and GluRIIB seemed to be dose dependent, in that GluRIIB, with two predicted MiR-284 binding sites, was much more strongly de-repressed after knockout of MiR-284 than was GluRIIA, which has only one predicted MiR-284 binding site.

To confirm that the Df(3R)kar-Sz12 mutant phenotype shown in Fig. 8 was specifically caused by loss of MiR-284, we performed two types of experiments. First, we performed a complementation test using another MiR-284 deletion, Df(3R)exel7317. As expected, Df(3R)kar-Sz12/Df(3R)exel7317 larvae showed dramatically increased nonsynaptic GluRIIA and GluRIIB immunoreactivity (Fig. 9 A, left), similar to that observed in homozygous Df(3R)kar-Sz12 mutants (Fig. 8) and after dicer-1 knockdown (Fig. 3). Second, we cloned a small genomic fragment containing only the MiR-284 gene, and expressed this transgene in homozygous Df(3R)kar-Sz12 mutant larvae, as well as in Df(3R)kar-Sz12/Df(3R)exel7317 mutant larvae (Fig. 9, A–C). As shown in Fig. 9 (A–C), transgenic expression of MiR-284 in both types of MiR-284 deletion mutant backgrounds rescued GluRIIA and GluRIIB overexpression phenotypes to near control levels. Expression of the MiR-284 transgene also rescued both homozygous Df(3R)kar-Sz12 mutants and Df(3R)kar-Sz12/Df(3R)exel7317 mutants to adulthood, which suggests that loss of MiR-284 was by itself responsible for a significant portion of the lethality in these genotypes. Both homozygous Df(3R)kar-Sz12 mutants and Df(3R)kar-Sz12/Df(3R)exel7317 mutants showed 100% preadult lethality in the absence of the MiR-284 transgene.

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Figure 6. NMJ synaptic transmission is largely unaltered after muscle-specific knockdown of dicer-1 RNAi. Synaptic transmission was assayed by voltage-clamping postsynaptic ventral longitudinal muscle 6 (at −60 mV) and recording either evoked (EJC) or spontaneous (sEJC) synaptic transmission at NMJs. (A) EJC amplitude is not significantly changed after muscle-specific knockdown of dicer-1 (n = 8–10 for each genotype). (B) Knockdown of muscle dicer-1 also does not significantly change sEJC ("mini") frequency (n = 10–20 animals of each genotype; thousands of sEJCs). (C and D) Muscle-specific knockdown of dicer-1 leads to a slight increase in sEJC amplitude, which suggests a slight increase in the number of functional postsynaptic GluRs (n = 10–20 animals of each genotype; thousands of sEJCs). Error bars represent SEM. n.s., not significant. *, P < 0.01.

Figure 7. Knockdown of muscle dicer does not increase the number of functional nonsynaptic GluRs. Glutamate-gated currents from pressure ejection of 1 mM glutamate onto extrasynaptic muscle plasma membrane are not changed after muscle-specific knockdown of dicer-1 (n = 11–19 of each genotype). Error bars represent SEM. n.s., not significant.
Discussion

Our data demonstrate that microRNAs, including but not limited to MiR-284, control Glu subunit abundance and synaptic receptor subtype ratios in the Drosophila larval NMJ. Specifically, we describe regulation of GluRIIA and GluRIIB. Our conclusion that microRNAs suppress GluRIIA and GluRIIB abundance is based on several pieces of evidence: (1) quantitative real-time PCR showing an increase in GluRIIA and GluRIIB transcript abundance after dicer-1 knockdown; (2) FISH showing an increase in microscopically visible GluRIIA and GluRIIB mRNA after dicer-1 knockdown; (3) immunohistochemistry showing a dramatic increase in GluRIIA and GluRIIB protein after dicer-1 knockdown; (4) electrophysiology showing an increase in sEJC amplitude after dicer-1 knockdown; and (5) immunohistochemistry showing an increase in GluRIIA and GluRIIB protein after genetic deletion of MiR-284. To our knowledge, this is the first evidence for regulation of GluRs by small noncoding RNAs in any organism.

Because GluRIIA and GluRIIB are rate-limiting for GluR formation, and because MiR-284 regulated GluRIIB more strongly compared with GluRIIA, changes in MiR-284 activity lead to changes in the postsynaptic A-type/B-type receptor ratio. A- and B-type GluRs are biophysically and pharmacologically distinct (DiAntonio et al., 1999; Marrus et al., 2004; Schmid et al., 2008), and the A-type/B-type receptor ratio varies developmentally (Schmid et al., 2008) and along the larval ventral–dorsal axis (Lee et al., 2008). MicroRNA-mediated regulation of GluR subunit transcripts, and MiR-284 in particular, may explain how these developmental and spatial differences are achieved.

We also showed that RNAi can effectively knock down Drosophila dicer-1, which is in agreement with the conclusion that dicer-1 and dicer-2 play largely independent roles for processing and/or synthesis of microRNA and RNAi (Lee et al., 2004), and demonstrated that dicer-1 is predominantly localized to distinct puncta, which is consistent with the conclusion that micro ribonucleoprotein complexes aggregate in vivo. However, dicer-1 puncta were scattered throughout postsynaptic muscle cells, and not obviously associated with any particular organelle, the perinuclear region, or postsynaptic densities, as previously described for mouse brain (Lugli et al., 2005).

Interestingly, MiR-284 regulated GluR protein abundance in proportion to the number of predicted binding sites in each target transcript: GluRIIB had two predicted MiR-284 binding sites, and was most strongly affected by loss of MiR-284; GluRIIA, a deletion (Df[3R]kar-Sz12) that removes MiR-284 show abundant mis-localized GluRIIA and GluRIIB in body wall muscles. (C) Quantification from images like that shown in B. GluRIIB protein is dramatically increased both in synapses and in nonsynaptic intracellular areas, whereas GluRIIA is increased only nonsynthetically (n = 10–12). ***, P < 0.0001. [D] GluRIIC is not changed in MiR-284 deletion mutants (Df[3R]kar-Sz12; n = 5). Error bars represent SEM, n.s., not significant.
with one predicted binding site, was less affected; and GluRIIC, with no predicted binding sites, was not affected at all. This suggests that microRNA-mediated regulation of any particular transcript may depend on the number and type of individual microRNA-binding sites, where more potential binding sites lead to stronger repression. Also, interestingly, none of the predicted MiR-284 binding sites in either GluRIIA or GluRIIB were in UTRs.

However, as noted, we were unable to obtain consistent a priori bioinformatic predictions regarding the number and type of microRNA-binding sites in *Drosophila* GluR transcripts. Even the experimentally supported prediction that MiR-284 binds to GluR mRNAs was not generally replicable using most bioinformatics tools, including, importantly, later versions of the program (MicroInspector) that made the original prediction. These sorts of bioinformatics inconsistencies have been commented upon by others (Bartel, 2009), and demonstrate that the molecular mechanisms by which microRNAs interact with their targets in vivo are still imperfectly understood or not yet consistently implemented in publicly accessible computational form. However, it is important to note that we were eventually able to successfully predict MiR-284 regulation of GluRIIA and GluRIIB using a combination of bioinformatic “tuning” and preliminary biological data. We therefore think that computational tools for prediction of microRNA–target interactions remain an important goal, and deserve high priority given the obvious usefulness that accurate algorithms will have.

Figure 9. A Mir-284 transgene rescues GluR phenotypes caused by deletion of the native Mir-284 gene. [A] Larvae hemizygous for each of two different Mir-284 deletions [Df(3R)kar-Sz12/Df(3R)Exel7317] show abundant mislocalized GluRIIA and GluRIIB in body wall muscles (left), and this phenotype is largely rescued with a single copy of a Mir-284 transgene (right). [B and C] Quantification of synaptic (B) and nonsynaptic (C) GluRIIA and GluRIIB immunoreactivity (green and red, respectively) for homozygous Df(3R)kar-Sz12 mutants, mutants of the genotype Df(3R)kar-Sz12/Df(3R)Exel7317, and each of these two mutants expressing a Mir-284 transgene. Error bars represent SEM. n.s., not significant. ***, P < 0.0001. [D] Pie charts showing relative amounts of GluRIIA and GluRIIB immunoreactivity in control NMJs, in NMJs after deletion of Mir-284, and in Mir-284 deletion mutants rescued with a Mir-284 transgene.
Materials and methods

UAS-dicer-1 transgene construction

The UAS-dicer-1 IR (UAS-dicer RNAi) transgene was created using previously described methods and reagents (Lee and Carthew, 2003). In brief, a 513-bp dicer-1–specific PCR product was amplified from genomic DNA using primers 5′-XbaDcr1 (5′-TTTGCAGGGATCTTACGACATCT-CAAGCATGAC-3′) and 3′-XbaDcr1 (5′-TTTGGATCTAAGATGAGGATTGAGAATCT-AGAGTAACCAGCATC-3′). This PCR product was digested with XbaI and ligated “tail-to-tail” into the AvrII site of the pWIZ vector (Lee and Carthew, 2003). This construct was then used to transform yeast (6c23) flies according to standard methods (Spradling and Rubin, 1982). Two independent transgene insertion lines were obtained and mapped to chromosomes 2 and 3. We expressed UAS-dicer RNAi transgenes using two different Gal4 drivers: TubGal4, which drives in all tissues (Lee and Luo, 1999), and 24BGal4, which drives exclusively in mesoderm (muscle; Brand and Perrimon, 1993).

UAS-Mir-284 transgene construction

The “rescue” transgene containing the MiR-284 genomic region was created using the Gateway cloning system (Invitrogen). To clone the MiR-284 genomic region, we PCR-amplified the 21-bp MiR-284 gene (5′-TGAAGCTGAGACCATCTGATCCAGCAATTG-3′) plus an additional 1 kb in the upstream direction, and 300 bp in the downstream direction. Primers were designed with attB sites added according to the Gateway cloning protocol. The primers were: MiR-284 L3 (5′-GAATCTCCGGGTAATCAGCAGAATG-3′) and MiR-284 R3 (5′-TTAGTCTAGAATGAGGTAATCATAG-3′). The pUAST Gateway destination vector was designed by B. Ackley (University of Kansas, Lawrence, KS) using the Gateway Vector Conversion System (Invitrogen). The pUAST-MiR-284 transgene was then sequenced and transformed using standard methods (Spradling and Rubin, 1982).

All rescue animals were individually genotyped using PCR to confirm that the native MiR-284 gene was deleted and that they also carried the MiR-284 transgene, as follows. First, we confirmed that the native MiR-284 gene was deleted using primers 5′-CCAAAATCGCAGAGGACTGACCAT-3′ and 5′-CTCGGATCTAGCAGCATTG-3′. These primers produce a 762-bp band when the wild-type genomic region is present; a negative PCR result indicated the larva was homozygous for the MiR-284 deletion. Actin 5C was used a positive control for the PCR reaction, using primers 5′-GGAACACCACTCTTCACAAGC-3′ and 5′-TACAAGCGAGAGACACAGGG-3′, which amplify a 171-bp region of the actin 5C gene.

Real-time RT-PCR and quantification of GluR mRNA abundance

For real-time RTPCR, RNA isolation, reverse transcription, and PCR was performed as described previously (Featherstone et al., 2005; Liebl et al., 2005; Liebl et al., 2006), with slight modifications for more precise quantification. In brief, Drosophila total RNA was isolated from several animals of each genotype using standard TRIzol extraction (Roberts, 1998), and then reverse transcribed using poly dT primers. Real-time PCR was then performed on mRNAs of interest (GluRIIA, GluRIIB, and the control/standard actin 5C) using a thermocycler (Opticon 2; MJ Research) with SYBR green for amplification detection and quantitation. Quantification of starting mRNA was computed according to Harz et al. (2004). In brief, the cycle threshold (Ct) values for the target (GluRIIA or GluRIIB) and an actin 5C control were determined for each sample, and the target was normalized to the control using the following calculation: ΔCt(sample) = Ct(target) – Ct(Actin 5C). Normalized sample Ct values were then referenced to a wild-type control sample (the “calibrator”) to determine the relative amount of target mRNA in the test sample. The formula used for determining relative target levels was: ΔΔCt(sample) = ΔCt(sample) – ΔCt(calibrator). The amount of target for each sample is reported as a ratio relative to the calibrator using 2-ΔΔCt. Essentially, actin 5C serves as a control for RNA isolation and amplification in each sample, and the wild type provides a standard quantity of target transcript to which all samples are referenced. Other aspects of the calculations correct for the fact that PCR product abundance is logarithmically (rather than linearly) related to the starting template.

FISH

RNA probes were synthesized in vitro from full-length target cDNAs, using standard methods appropriate to the cDNA vector (e.g., SP6 RNA polymerase for vectors containing an SP6 promoter). cDNAs for GluRIIA and GluRIIB were gifts from S. Sigrist (Freie Universität Berlin, Berlin, Germany); GluRIIC cDNA was obtained from the Drosophila Genomics Resource Center. After synthesis, probes were shortened using alkaline hydrolysis for better penetration of tissues (Cox et al., 1984), and precipitated using standard methods. In most cases, the probe was labeled with digoxigenin-UTP (DIG-UTP; Roche) during transcription and then eventually visualized using anti-DIG antibodies. In some cases, the synthesis reaction included an amine-modified UTP that could be subsequently directly labeled with a fluorophore (“FISH tag” kit; Invitrogen). In situ hybridization was then performed using standard methods similar to those described previously (Braissant and Wahl, 1998; Sigrist et al., 2000; Tomancak et al., 2002). In brief, manually filled olorless third instar larvae were fixed for 1 h in 4% paraformaldehyde in phosphate-buffered saline, and then permeabilized using TritonX-100. After fixation, larvae were incubated with probes at 55°C overnight, then stained with anti-HRP antibodies to visualize NMJs, and with either anti-DIG antibodies or fluorophores with affinity to amine-modified UTP for visualization of mRNA using confocal fluorescent microscopy.

Immunocytochemistry and confocal microscopy

For immunocytochemistry and confocal microscopy, animals were manually dissected and fixed in Bouin’s fixative, as described previously (Chen and Featherstone, 2005; Chen et al., 2005). Drosophila dicer-1 mouse polyclonal antibody (A1031a; used at 1:1,000) was generated against the dicer-1–specific peptide TRHLYDPRQHSGAQDTR by the CIM antibody core at Arizona State University, but is no longer available from this source due to loss of funding. Mouse monoclonal anti-GlurIIA, from a cell line made by C. Schuster (University of Heidelberg, Heidelberg, Germany) and available from University of Iowa’s Developmental Studies Hybridoma Bank, was used at 1:100–1:200 (Featherstone et al., 2002). Rabbit polyclonal anti-GlurIIA and rabbit polyclonal anti-GlurIIB, first described in Liebl et al. (2004) and subsequently replicated in our own laboratory (Liebl et al., 2005), were used at 1:2,000. Fluorescently conjugated anti-HRP was used at 1:100–1:200. Immunoreactivity was visualized using FITC, TRITC, or CY5-conjugated goat anti–mouse/rabbit secondary antibodies (Jackson Immunoresearch Laboratories) at 1:400. For plasma membrane staining using the fixable membrane dye FM-4-64FX (Invitrogen), dissected larval preparations were exposed to 5 µg/ml FM-4-64FX in ica-cold phosphate-buffered saline for 1 min, then fixed immediately in Bouin’s fixative. For imaging, preparations were mounted on slides in Vectashield mounting medium (Vector Laboratories). All images of NMJs were from ventral longitudinal muscles 6 and 7 in abdominal segments 3 and 4. Control and experimental preparations were always stained and imaged in parallel.

Images were captured using a laser scanning confocal system (Fluoview FV500; Olympus) running Fluoview software 4.3 with a 40×/1.3 NA Plan-Fluar or a 60×/1.4 NA Plan-Apochromat objective lens (Olym pus). All imaging was performed at room temperature (20–23°C). Quantification of staining intensity was performed as described previously (Featherstone et al., 2001). In brief, fluorescence intensity of relevant structures from unaltered, unconfocal maxima intensity projections was measured by manually selecting the region of interest in ImageJ and measuring the mean pixel intensity of that region. To control for differences in individual preparation immunoreactivity, excitation, fluorescence attenuation, and detection, we then subtracted the mean pixel intensity of a similarly sized region of unstained “background” in the same fluorescence wavelength channel in the same image. 3D isosurface reconstructions were performed using Amira 3.1 (Mercury Computer Systems). Photoshop CS2 was then used to crop images and adjust contrast for printing.

Microscopic puncta (dicer-1 immunoreactivity or Glur mRNA aggregates detected by H3F; Figs. 1 C and 2 B) were counted manually or using Particle Counting macros in ImageJ 1.42 (http://rsb.info.nih.gov/ij/). Both methods yield similar results. Spots that were 1–2 pixels in total size were in both cases assumed to be noise, and therefore not counted.

Electrophysiology

All electrophysiological recordings were obtained from larval ventral longitudinal muscle 6 (A2–4) using a two-electrode voltage clamp (−60 mV) as described previously (Featherstone et al., 2000, 2005; Liebl et al., 2005). Note that this is the same NMJ that was analyzed for all other experiments in this study. All dissections and electrophysiology were performed under standard Drosophila saline (135 mM NaCl, 5 mM KCl, 4 mM MgCl2, 1.8 mM CaCl2, 5 mM TES [Tris, EDTA, and NaCl], and 72 mM sucrose).

Statistics

Statistical significance was determined using unpaired Student’s t test (for normally distributed data) or nonparametric Mann-Whitney tests (when posthoc F-tests determined that variances were significantly different). SEIC amplitude distributions were compared using Kolmogorov-Smirnov tests. In all figures, asterisks indicate statistical significance (*, P < 0.05; **, P < 0.01; ***, P < 0.001; ****, P < 0.0001). “a.s.” indicates a lack of statistical significance (i.e., not significantly different), and error bars represent SEM.
Bioinformatics
Online supplemental material includes three Microsoft Excel files converted from comma-delimited .csv data files, which represent output from MicroInspector (version 1.0; Rusinov et al., 2005). Each of these files was generated using MicroInspector 1.0 default/recommended parameters (Hybridization temperature 25°C, ~26 kcal/mole free energy cutoff). Rows are sorted by free energy, where the most likely interactions are listed first. As described in the text, our experimental results support only the most likely two interactions for GluRIIA and GluRIIB, and none of the interactions for GluRIIC. Note that algorithms in subsequent versions of MicroInspector have been modified, and results from the current online version may not be identical to those described here.

Online supplemental material
Table S1 is the MicroInspector output after the Flybase 5.2 GluRIIA transcript sequence was used as input. Table S2 is the MicroInspector output after the Flybase 5.2 GluRIIB transcript sequence was used as input. Table S3 is the MicroInspector output after the Flybase 5.2 GluRIIC transcript sequence was used as input. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200902062/DC1.

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