Elevated Peripheral Brain-Derived Neurotrophic Factor Level Associated With Decreasing Insulin Secretion May Forecast Memory Dysfunction in Patients With Long-Term Type 2 Diabetes

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Background: With the progressive course of diabetes and the decline in islet function, the cognitive dysfunction of patients aggravated.

Objective: We aimed to investigate the roles of brain-derived neurotrophic factor (BDNF) and the Val66Met polymorphism in mild cognitive impairment (MCI) in patients with type 2 diabetes mellitus (T2DM).

Methods: A total of 169 Chinese patients with T2DM were involved and divided into long-term (diabetes duration > 10 years) and short-term (diabetes duration ≤ 10 years) diabetes, and in each group, the patients were separated as MCI and the control. Demographic characteristics, clinical variables, and cognitive performances were assessed. The plasma BDNF level was measured via enzyme-linked immunosorbent assay. The Val66Met polymorphisms were analyzed.

Results: Long-term T2DM have lower 2 h postprandial C-peptide (p < 0.05). The BDNF level was slightly higher in patients with MCI than in the controls in each duration group without statistical significance. The relationship of BDNF to Montreal Cognitive Assessment was not proven either. However, in the long-term diabetes group, BDNF concentration remained as an independent factor of logical memory test (β = −0.27; p < 0.05), and they were negatively correlated (r = −0.267; p = 0.022); BDNF was also negatively correlated with fasting C-peptide (r = −0.260; p = 0.022), 2 h postprandial C-peptide (r = −0.251; p = 0.028), and homeostasis model assessment of insulin resistance (r = −0.312; p = 0.006). In genotypic groups, BDNF Val/Val performed better in logical memory test than Met/Met and Val/Met.

Conclusion: Elevated peripheral BDNF level associated with declined islet function, when combined with its Val66Met polymorphism, may forecast memory dysfunction in patients with long-term T2DM.

Keywords: brain-derived neurotrophic factor, mild cognition impairment, type 2 diabetes, islet, insulin
INTRODUCTION

Brain-derived neurotrophic factor (BDNF) belongs to a family of neurotrophins that play a crucial role in the survival and differentiation of the neuronal population (Kowianski et al., 2018; von Bohlen Und Halbach and von Bohlen Und Halbach, 2018). BDNF is abundantly expressed in the central and peripheral nervous systems (Karczewska-Kupczewska et al., 2011), and it may play a non-substitutable role in the hippocampus in supporting the electrophysiological function of memory circuitry (Nagahara and Tuszynski, 2011). Evidence shows that BDNF can cross the blood-brain barrier in both directions (Pan et al., 1998; Karczewska-Kupczewska et al., 2011; Jiang et al., 2018); thus, the correlation between circulating BDNF and brain levels and brain function has been discussed in animal and human studies (Hwang et al., 2015; Weinstein et al., 2017; Ng et al., 2019).

Novel BDNF-focused investigations are being developed for both metabolism and neurological disorders. In neurodegenerative disorders (Weis et al., 2003; Blesch, 2006), including Alzheimer's disease (AD; Lima Giacobbo et al., 2019; Ng et al., 2019) and mild cognitive impairment (MCI; Nagata et al., 2014), the levels of BDNF is one of the most common physiological changes. BDNF plays a critical role in cognition and memory (Kowianski et al., 2018; von Bohlen Und Halbach and von Bohlen Und Halbach, 2018; Miranda et al., 2019). Meanwhile, peripheral BDNF is observed in various metabolic conditions, including high cholesterol, insulin insensitivity, and type 2 diabetes mellitus (T2DM; Hölscher, 2011; Weinstein et al., 2017; Movassat et al., 2019). BDNF increases insulin sensitivity in the brain and periphery (Marosi and Mattson, 2014). Besides, the discovery of brain-specific insulin signaling deficiencies in the early stages of AD pathogenesis has led to the designation of AD as "type 3 diabetes" (de la Monte, 2012; Kandimalla et al., 2017). MCI is considered an intermediate stage between the expected cognitive decline in normal aging and AD (Eshkoor et al., 2015), where memory loss is the main symptom. Thus, the relationship between BDNF and insulin may be a novel research focus in investigating the underlying mechanisms of T2DM and MCI. To our knowledge, in long-term T2DM, islet function decreases, and cognitive dysfunction emerges simultaneously (Kandimalla et al., 2017). It is reported that insulin secretion decreases during the first 10 years after the diagnosis of T2DM (Mauvais-Jarvis et al., 2004). Taking different T2DM duration into account, we must determine whether peripheral BDNF interacts with insulin to influence the cognition decline in MCI patients with T2DM.

Although evidence suggests that BDNF plays a key role in the process of AD and T2DM, the role and association between BDNF polymorphism with these two diseases have been extensively investigated. Among them, a naturally occurring polymorphism in the human BDNF gene, Val66Met, was discovered by a member of the research staff. Physical measurement and demographic characteristics and medical history were collected by a member of the research staff. Physical measurement was observed in a standard way. Body mass index was calculated as body weight divided by the square of the height [body weight (kg)/body height (m$^2$)]. Blood samples were drawn from the subjects the following morning after their admission to the hospital. C-peptide, triglyceride (TG), total cholesterol (TC), high-density lipoprotein (HDL-C), uric acid (UA), and thyroid-stimulating hormone (TSH) were tested using an automatic biochemical analyzer (Roche Group, Basel, Switzerland). Low-density lipoprotein (LDL-C), apolipoprotein A1 (ApoA1), apolipoprotein B (ApoB), and (lipoprotein a) Lpa were measured using an automatic biochemical analyzer (Ningbo Ruiyuan Biotechnology Co., Ltd., Ningbo, China). Glycosylated hemoglobin (HbA1c) was tested using an analyzer (Bio-Rad Laboratories, Inc., United States). Insulin resistance
was calculated according to homeostasis model assessment of insulin resistance (HOMA-IR) using the formula for C-peptide: FPG (mmol/L) × FCP (nmol/L)/22.5 (Kim et al., 2016). The preceding biochemical measurements were performed in the central laboratory of the Affiliated Zhongda Hospital, which implemented internal and external quality control procedures directed by the Chinese Laboratory Quality Control.

**Neuropsychological Function Evaluation**
Participants completed neuropsychological testing comprising the following.

**Montreal Cognitive Assessment**
Montreal Cognitive Assessment is a rapid sensitive assessment tool for screening mild cognition dysfunction, and the total score was 30 points (Nasreddine et al., 2005).

**Mini-Mental State Examination**
The examination was composed of 30 items, in which for each item, a correct answer is awarded one point, with a total of 30 points (Folstein et al., 1975).

**Digit Span Test**
The subjects were instructed to match a symbol with a number within 90 s according to the association between the number and the symbol.

**Verbal Fluency Test**
Verbal fluency test (VFT) assesses the correct number of animals, vegetables, fruits, and common grocery items identified within 1 min.

**Clock Drawing Test**
The participants were asked to draw the face of a clock.

**Auditory Verbal Learning Test**
Auditory verbal learning test (AVLT) evaluated a wide range of auditory and verbal learning functions. Immediate recall, short and long-delayed recall were recorded.

**Logical Memory Test**
Logical memory test (LMT) assessed immediate recall and long-delayed recall were recorded.

**Stroop Color-Word Test**
Stroop Color-Word Test (SCWT) assessed the time taken to correctly identify the number of instances in which the color of the written word in identified (A time and A number), color of the symbol (B time and B number), and the color of the ink rather than the color the word spells (C time and C number).

**Trail Making Test-A and Trail Making Test-B**
Trail Making Test-A and Trail Making Test-B indicate the mean times taken to complete part A (visual conceptual) and part B (visuomotor tracking, which involves motor speed and attention functions).

Montreal Cognitive Assessment and MMSE scores were used to assess global cognitive abilities. SCWT, DST, VFT, and TMTB were conducted to evaluate the executive functions of the patients. CDT was used to analyze visual space function. TMTA test was performed to assess the information processing speed function. LMT and AVLT were used to analyze episodic memory.

Hachinski’s ischemic score, clinical dementia rating, the activity of daily living scale, and self-rating depression scale were obtained simultaneously. An experienced neurologist from the Department of Neurology in the Affiliated Zhongda Hospital of Southeast University performed all procedures, and the participants were blinded to the study design. The duration of the whole examination was approximately 45 min.

**Measurement of Plasma Brain-Derived Neurotrophic Factor Level**
Blood samples were collected via venipuncture between 6:30 a.m. and 7:00 a.m. following an overnight fast. The samples were placed in anticoagulant tubes and centrifuged at 100 × g for 15 min to separate the plasma, which was stored at −80°C until analysis. Plasma BDNF concentrations were measured on the same day using enzyme-linked immunosorbent assay kits (Nanjing Jin Yibai Biological Technology Co., Ltd., Nanjing, China) per the instructions of the manufacturer. Intra- and inter-assay coefficients of variance were less than 9 and 10%, respectively, and the sensitivity limit of the assay was 0.5 µg/L.

**DNA Extraction and Single-Nucleotide Polymorphism Genotyping**
Genomic DNA was extracted from the stored EDTA anticoagulated-venous blood using a DNA purification kit (Puregene, Gentra Systems, Minneapolis, MN, United States). Then, the genotype of BDNF rs6265 was detected using the Sequenom method (Beijing Cnkingbio Biotechnology Corporation, Beijing, China). The following forward and reverse primers were used: 5′-ACGTTGGATGTTGTTTTCTTCAATGGCG-3′ and 5′-ACGTTGGATGGCCTTGCATCATTTGGCTGAC-3′, respectively. The target region was amplified using PCR Accessory Set (Sequenom, 11327). The PCR products were processed using iPLEX Gold reagents and purified using a SpectroCHIP Resin kit (Sequenom, 10117-2). MassARRAY® Analyzer (Agena, 260000) was used to acquire data from the purified samples. No inconsistency was found when 10% of the samples were randomly selected and genotyped again.

**Statistical Analysis**
Data were presented as mean ± standard error of the mean (SEM), median (interquartile range), or percentage. Demographic, clinical characteristics, and neuropsychological test scores of the MCI and control patients were compared using Student’s t-test for normally distributed variables, the non-parametric Mann–Whitney U-test for asymmetrically distributed variables, and chi-squared (χ²) test for the qualitative variables. The test was also conducted to evaluate the distribution of genotypes and allele frequencies and to determine the deviations from the Hardy–Weinberg equilibrium (Santiago Rodriguez, Tom R. Gaunt, and Ian N. M. Day, Hardy–Weinberg
Equilibrium Testing of Biological Ascertainment for Mendelian Randomization Studies). Analysis of variance was used to compare the plasma level of BDNF, glycosylated hemoglobin, and cognitive domains between different genotypes, and the least significant difference method was used for comparisons within each simple genotype group. The partial correlation analysis was performed to explore the relationships between BDNF and cognitive performances. Three hierarchical multiple regression models were used to estimate the predictor variables for LMT in patients with T2DM. In model 1, unique associations of the demographic factors to LMT were tested. Relational factors were added in model 2, and unique associations between health status covariates with LMT beyond effects of the demographic factors were tested. Finally, BDNF was added in model 3, and the unique associations between BDNF with LMT beyond effects of the demographic factors and health status covariates were tested. The linear correlation analysis was conducted to determine the relationship between BDNF and C-peptide, HOMA-IR. Statistical analyses were conducted using SPSS version 19.0 (SPSS Inc., Chicago, IL, United States).

RESULTS

Demographic Characteristics, Clinical Characteristics, and Cognitive Performance
A total of 51 patients with MCI and 118 patients with healthy cognition were involved and divided into two groups according to the duration of T2DM. The demographic characteristics, clinical characteristics, and neuropsychological test scores of the participants are listed in Supplementary Table 1. Patients with T2DM for more than 10 years were older, have a longer hypertension duration, and lower 2 h postprandial C-peptide (2hCP) than those with T2DM for not more than 10 years (all \( p < 0.05 \)). A comparison of the MCI group and the control in each diabetes duration group is shown in Table 1. The plasma BDNF level of MCI was slightly higher than that of the control, but the difference was not significant.

Relationship Between Brain-Derived Neurotrophic Factor and Cognitive Performances
The respective correlations between plasma BDNF level and cognitive performances were analyzed. When sex, age, TSH, and education levels were controlled, the partial correlation indicated that in the patients with T2DM for more than 10 years, BDNF was negatively related to LMT \( (r = -0.267; p = 0.022) \) (Table 2). The relationship of BDNF to other cognition performances was not proven.

We performed the hierarchical multiple regression analysis in patients with T2DM for not more than 10 years, with LMT score as the dependent variable. The demographic factors, sex, age, and education levels, tested in model 1, did not explain meaningful variance in LMT scores. Health status covariates, total cholesterol, and high-density lipoprotein (HDL), which are tested in model 2, explain 11% of the variance in the LMT score beyond the effects of the demographic factors. In step 3, BDNF accounted for a significant proportion of the LMT score controlling beyond the effects of the demographic and health status covariates \( (\Delta R^2 = 0.07; p < 0.05) \). Additionally, in this final model, BDNF \( (\beta = -0.27; p < 0.05) \) was independently related to the LMT score, and the final model explained 23% of the variance in the LMT score (Table 3).

Relationship Between Brain-Derived Neurotrophic Factor to Insulin Secretion and Insulin Resistance
The linear correlation analysis was conducted between BDNF and indicators of islet secretion in subjects with diabetes for more than 10 years, which presented a negative relationship [FCP \( (r = -0.260; p = 0.022) \), 2hCP \( (r = -0.251; p = 0.028) \), HOMA-IR \( (r = -0.312, p = 0.006) \)]. In the group with T2DM for not more than 10 years, the correlation between BDNF and insulin secretion was not proven (Table 4).

Distributions of Brain-Derived Neurotrophic Factor Genotype and Allele Frequencies Between Groups
Distributions of BDNF genotype and allele frequencies of the MCI and the control subjects are presented in Table 5, which are consistent with the Hardy–Weinberg equilibrium in the MCI group \( (\chi^2 = 2.497, df = 2, p > 0.05) \) and control groups \( (\chi^2 = 0.717, df = 2, p > 0.05) \). No significant differences were observed in the distributions between the two groups.

Comparison of Plasma Brain-Derived Neurotrophic Factor Level, Logical Memory Test, Insulin Secretion, and Resistance Between Genotypic Subgroups
The differences in plasma BDNF level, insulin secretion, and resistance of BDNF rs6265 polymorphism groups are not significant as summarized in Table 6. However, the effect of genotype on the LMT scores was significant \( (F = 4.435, p = 0.013) \), while that of the Val/Val group is more remarkable than those of the Met/Met group \( (\text{mean difference} = 2.621, p = 0.008) \) and Val/Met \( (\text{mean difference} = 2.253, p = 0.009) \) (Table 6).

DISCUSSION
In this observational study, we first compared the plasma level of 2hCP in patients with >10 years and <10 years of T2DM and found it was obviously lower in the group of long-term duration. Second, this study compared the plasma levels of BDNF in patients with T2DM between groups with or without MCI in the different diabetes duration groups. In each group, compared with control, the BDNF level of the MCI group was slightly higher, but the difference was not significant. Third, we conducted a correlation analysis between
### TABLE 1 | Baseline characteristics of T2DM patients with different duration.

| Characteristic                | Diabetes duration >10 years (n = 77) | Diabetes duration ≤10 years (n = 92) |
|------------------------------|--------------------------------------|--------------------------------------|
| Age (yr)                     | 64.2 ± 8.24                         | 61.58 ± 8.39                         |
| Male sex, n (%)              | 12.00 (41.40)                        | 29.00 (60.40)                        |
| Fatty liver, n (%)           | 16.00 (55.20)                        | 20.00 (41.70)                        |
| Education level (yr)         | 10.00 (9.00, 12.00)                  | 11.00 (9.00, 12.00)                  |
| Smoking, n (%)               | 8.00 (27.60)                         | 17.00 (35.40)                        |
| Drinking, n (%)              | 3.00 (10.30)                         | 11.00 (22.90)                        |
| Hypertension, n (%)          | 20.00 (69.00)                        | 27.00 (56.30)                        |
| Hypertension duration (yr)   | 8.00 (0.00, 15.50)                   | 9.57 (0.00, 16.50)                   |
| Insulin use, n (%)           | 13.00 (44.80%)                       | 33.00 (68.80)                        |
| Metformin use, n (%)         | 17.00 (58.60)                        | 39.00 (81.30)                        |
| BMI (Kg/m²)                  | 24.62 ± 3.17                         | 25.00 ± 3.34                         |
| Weight (kg)                  | 65.00 (58.00, 72.00)                 | 70.00 (60.25, 76.00)                 |
| Height (cm)                  | 163.00 (155.00, 169.00)*             | 168.00 (160.00, 172.00)*             |
| Systolic BP (mmHg)           | 132.00 (120.00, 150.00)              | 140.00 (122.25, 150.75)              |
| Diastolic BP (mmHg)          | 80.00 (73.00, 87.00)                 | 80.00 (74.00, 90.00)                 |
| HbA1c (%)                    | 9.80 (7.65, 10.85)                   | 8.20 (7.70, 9.38)                    |
| FCP (ng/mL)                  | 0.47 (0.26, 0.64)                    | 0.58 (0.32, 1.02)                    |
| zHCP (ng/mL)                 | 1.63 (1.17, 2.35)                    | 2.11 (1.21, 3.09)                    |
| HOmA-IR                      | 0.06 (0.04, 0.09)                    | 0.07 (0.03, 0.12)                    |
| TG (mmol/L)                  | 1.49 (0.88, 1.94)                    | 1.23 (0.87, 1.85)                    |
| TC (mmol/L)                  | 4.69 ± 0.83                         | 4.36 ± 1.20                         |
| LDL-C (mmol/L)               | 1.14 ± 0.13                         | 1.08 ± 0.89                         |
| ApoA1 (g/L)                  | 1.16 (0.98, 1.37)                    | 1.15 (0.92, 1.44)                    |
| ApoB (g/L)                   | 0.81 (0.69, 1.00)                    | 0.76 (0.67, 0.98)                    |
| HDL-C/LDL-C                  | 0.40 (0.33, 0.59)                    | 0.44 (0.35, 0.52)                    |
| LPa (mmol/L)                 | 144.00 (71.50, 284.00)               | 143.50 (74.25, 240.75)               |
| UA (µmol/L)                  | 283.00 (246.50, 340.00)              | 292.50 (253.25, 337.75)              |
| TSH (µU/ml)                  | 2.76 (1.93, 3.57)*                   | 2.08 (1.22, 2.68)                    |
| MOCO                        | 24.00 (22.00, 24.00)*                | 28.00 (27.00, 29.00)                 |
| MMSE                        | 27.00 (26.00, 28.50)*                | 29.00 (29.00, 30.00)                 |
| CDT                         | 3.00 (2.50, 4.00)*                   | 4.00 (3.25, 4.00)                    |
| DST                         | 11.00 (10.00, 12.00)*                | 12.00 (11.00, 13.00)                 |
| VFT                         | 15.00 (13.00, 17.00)*                | 18.00 (15.00, 21.00)                 |
| TMTA                        | 75.00 (57.00, 84.50)*                | 56.00 (48.00, 64.75)                 |
| SCWT T time                  | 189.00 (141.00, 265.50)*             | 131.50 (106.25, 172.00)              |
| SCWT A time                 | 32.00 (25.50, 43.00)                 | 28.50 (24.30, 35.75)                 |
| SCWT A number               | 50.00 (50.00, 50.00)                 | 50.00 (50.00, 50.00)                 |
| SCWT B time                 | 59.00 (43.50, 67.50)*                | 44.30 (37.50, 55.50)                 |
| SCWT B number               | 48.00 (47.00, 50.00)*                | 49.50 (48.00, 50.00)                 |
| SCWT C time                 | 100.00 (77.50, 125.00)               | 85.00 (71.75, 117.75)                |
| SCWT C number               | 44.00 (42.50, 48.00)*                | 48.00 (46.00, 49.00)                 |
| AVLT immediate              | 16.03 ± 5.00*                       | 19.23 ± 5.54                         |
| AVLT delayed                | 4.66 ± 2.60*                        | 6.56 ± 2.53                         |
| LMT                         | 6.93 ± 3.63*                        | 11.13 ± 4.35                         |
| BDNF (µg/L)                 | 5.85 (4.88, 9.92)                    | 5.53 (4.27, 8.30)                    |

*Significance, p < 0.05; **Significance, p < 0.01.

Data are presented as n (%), mean ± SD, or median (interquartile range) as appropriate. 1 Student’s t-test for the comparison of normally distributed quantitative variables between the MCI group and the control group in diabetic duration more than 10 years. 2 Student’s t-test for the comparison of normally distributed quantitative variables between the MCI group and control group in diabetic duration no more than 10 years. 3 Mann–Whitney U-test for the comparison of asymmetrically distributed quantitative variables between the MCI group and control group in diabetic duration more than 10 years. 4 Mann–Whitney U-test for the comparison of asymmetrically distributed quantitative variables between the MCI group and control group in diabetic duration no more than 10 years. 5 Chi-square (χ²) test was used to compare qualitative variables between the MCI group and control group in diabetic duration no more than 10 years. 6 Chi-square (χ²) test was used to compare qualitative variables between the MCI group and control group in diabetic duration more than 10 years. 7 Chi-square (χ²) test was used to compare qualitative variables between the MCI group and control group in diabetic duration more than 10 years. MCI, mild cognitive impairment; HbA1c, glycated hemoglobin; FPG, fasting plasma glucose; 2hPG, 2 h plasma glucose; FCP fasting C-peptide; 2hCP 2 h postprandial C-peptide; BMI, body mass index; WC, waist circumference; HC, hip circumference; WHR, waist-hip ratio; HOmA-IR, Homeostasis Model Assessment-Insulin Resistance; TG, triglyceride; TC, total cholesterol; LDL-C, low-density lipoprotein; HDL-C, high-density lipoprotein-C; ApoA1, apolipoprotein A1; ApoB, apolipoprotein B; MCOA, Montreal Cognitive Assessment; MMSE, Mini-Mental State Examination; CDT, Clock Drawing Test; DST, Digit Span Test; VFT, Verbal Fluency Test; TMTA, Trail Making Test-A; TMTB, Trail Making Test-B; AVLT, Auditory Verbal Learning Test; LMT, Logical Memory Test; SCWT, Stroop Color-Word Test; BDNF, brain-derived neurotrophic factor.
TABLE 2 | Partial correlation analysis between BDNF, clinical indicators, and LMT score of T2DM patients with different durations.

| LMT | Diabetes duration >10 years | Diabetes duration ≤10 years |
|-----|-----------------------------|-----------------------------|
|     | r  | p     | r  | p |
| BDNF (µg/L) | −0.267 | 0.022 | −0.014 | 0.896 |
| Metformin use | 0.311 | 0.007 | 0.060 | 0.579 |
| TC (mmol/L) | −0.301 | 0.009 | 0.029 | 0.792 |
| HDL-C (mmol/L) | −0.286 | 0.014 | −0.061 | 0.575 |

TABLE 3 | Hierarchical multiple regression analysis of factors associated with LMT score in T2DM more than 10 years.

| Variables | Model 1 | | Model 2 | | Model 3 |
|-----------|---------|-------------------------|---------|-------------------------|---------|
| | Standardized β | 95% CI | | Standardized β | 95% CI | | Standardized β | 95% CI |
| Age (year) | −0.10 | −0.18 to 0.07 | | −0.09 | −0.17 to 0.07 | | −0.08 | −0.16 to 0.07 |
| Sex | −0.17 | −3.61 to 0.53 | | −0.20 | −3.77 to −0.20 | | 0.18 | −3.58 to 0.25 |
| Education level | 0.07 | −0.23 to 0.42 | | 0.13 | −0.14 to 0.50 | | 0.15 | −0.09 to 0.53 |
| TC (mmol/L) | −0.21 | −1.99 to 0.21 | | −0.18 | −5.68 to 1.06 | | −0.25 | −2.14 to 0.01 |
| HDL-C (mmol/L) | −0.27* | −0.87 to −0.10 |
| ΔR² | 0.05 | | 0.11 | | 0.07 | |
| ΔF | 1.19 | | 4.56* | | 6.26* | |

*p < 0.05.

TABLE 4 | Relationship between BDNF and clinical indicators in T2DM patients with different durations.

| BDNF | DM duration >10 years | | DM duration ≤10 years |
|------|------------------------|------------------------|
|     | r  | p     | r  | p |
| FCP (ng/mL) | −0.260 | 0.022 | −0.156 | 0.137 |
| 2hCP (ng/mL) | −0.251 | 0.028 | −0.072 | 0.497 |
| HOMA-IR | −0.312 | 0.006 | −0.160 | 0.127 |

TABLE 5 | Distributions of BDNF genotype and allele frequencies between groups.

| Genotype, n (%) | Allele, n (%) |
|----------------|---------------|
| | Met/Met | Val/Val | Val/Met | p-value | Met | Val | p-value |
| MCI | 18 (35.3) | 11 (21.6) | 22 (43.1) | 0.111 | 58 (56.9) | 44 (43.1) | 0.149 |
| Control | 24 (20.3) | 28 (23.7) | 66 (55.9) | | 114 (48.3) | 122 (51.7) | |

Data are presented as n (%).

TABLE 6 | Comparison of plasma BDNF level, LMT, insulin secretion, and resistance between genotypic subgroups.

| Met/Met (n = 42) | Val/Val (n = 39) | Val/Met (n = 88) | p |
|----------------|----------------|----------------|---|
| FCP (ng/mL) | 0.69 (0.40, 1.03) | 0.51 (0.29, 0.87) | 0.56 (0.37, 0.84) | 0.83 |
| 2hCP (ng/mL) | 2.37 ± 1.21 | 2.23 ± 1.75 | 2.41 ± 1.30 | 0.93 |
| HOMA-IR | 0.07 (0.05, 1.12) | 0.05 (0.04, 1.01) | 0.07 (0.04, 0.11) | 0.87 |
| BDNF (µg/L) | 5.97 (4.78, 9.57) | 6.47 (4.78, 9.45) | 5.37 (4.42, 7.84) | 0.138 |
| LMT | 10.00 (5.00, 15.00) | 12.00 (9.00, 15.00) | 10.00 (7.00, 13.00) | 0.013 |

Data are presented as mean ± SEM, or median (interquartile range) as appropriate.

BDNF and cognitive performance (e.g., MMSE, MOCA, LMT, and other neuropsychological tests) and only found a significant relationship between LMT and BDNF. Only in the individuals with T2DM for more than 10 years, the plasma BDNF levels were obviously negatively correlated with the LMT scores, which was iterated via hierarchical multiple regression. Fourth, the plasma BDNF was negatively associated with FCP, 2hCP, and HOMA-IR only in the longer duration group. Finally, in terms
of polymorphism, the Val homozygote genotype performed best in the LMT test.

In this study, long-term T2DM has a lower 2hCP. This result corresponds with several studies, which reports that insulin secretion decreases in long-term T2DM either (Zangeneh et al., 2010). The difference in BDNF levels between the MCI and control groups was not significant, which was consistent with a meta-analysis of 2019 on the serum BDNF level in MCI (Ng et al., 2019). Higher, lower, or unchanged results have been described in patients with AD compared with healthy controls, but the reasons for such different findings are unclear (Balietti et al., 2018). According to the classification of neuropsychological assessment proposed by van den Berg et al. (2009), the cognitive domains that should be evaluated include memory, intelligence, processing speed, and attention. BDNF promotes synaptic plasticity and is crucial for hippocampus-dependent learning and memory (Binder and Scharfman, 2004). Thus, single cognition domains, such as LMT and testing logic memory, should be discussed specifically. In our examination, the negative correlation between peripheral BDNF level and LMT score was presented among patients with T2DM who were diagnosed for more than 10 years, which is a new supplement about the circulation BDNF levels of patients with AD. We hypothesize that on early cognition dysfunction, damaged logic memory might be the trigger of the compensatory increase in BDNF. This result was found only in the group of patients with long-term T2DM. The different insulin secretion between long and short diabetes duration might be the reason, which was observed in our study either.

Meanwhile, the correlation between BDNF and insulin in different diabetes duration was analyzed. To date, increased (Suwa et al., 2006; Boyuk et al., 2014) and decreased (Krabbe et al., 2007) BDNF were reported in the blood of patients with DM. Our results indicated that, especially in the group with long-term diabetes, the serum BDNF level showed a negative correlation with FCP, 2hCP, and HOMA-IR, and this result is consistent with that of a study of Chinese patients with T2DM (Li et al., 2016). Those results demonstrated that the plasma BDNF level can be regulated only when insulin secretion was decreasing and might explain the ability of BDNF to compensate for cognition decline in long-term diabetes duration.

In terms of genetic factors, to date, the effect of BDNF Val66Met polymorphism on cognition decline remains controversial. The Val allele may be neuroprotective and related to higher cognition, and Met is related to impaired cognitive function (Azeredo et al., 2017; Barha et al., 2019). A recent meta-analysis indicated that performance on memory-based tasks was better in Val/Val carriers (Toh et al., 2018), which is partly consistent with our results. Moreover, it was observed that Met was associated with greater memory decline by using an episodic memory composite score over 126 months, while the effect of BDNF on memory decline was proved to be greatest in preclinical AD. However, in the MCI group, there was no effect of Met66 on memory decline or disease progression to AD over 126 months (Lim et al., 2021). Similarly, in our study, no significant differences were noted between genotypic subgroups in either the MCI or control group. But LMT scores of Val/Val carriers were significantly higher than other patients, which was used to assess the episodic memory performance of patients. Besides, the role of the BDNF Val66Met SNP in episodic memory was also reported in other studies (Cathomas et al., 2010; Kennedy et al., 2015). These results support our hypothesis that on early cognition dysfunction, damaged memory might be correlated with BDNF. In our results, the BDNF Val66Met SNP did not affect plasma BDNF levels, which is consistent with other findings. It did not affect the FCP, 2hCP, and HOMA-IR of T2DM, although Val/Val is prone to developing T2DM (Daily and Park, 2017).

However, several limitations should be considered in our study. First, the relatively small sample size limited the interpretation of our results. Second, the course of disease development was not studied, and thus, a prospective study should be conducted. However, this study is one of the first to investigate the relationship between plasma BDNF, insulin, cognition domain in different duration of T2DM, and the BDNF rs6265 polymorphism. In summary, we suggest that the plasma BDNF in long diabetic duration, correlated with insulin secretion, might contribute to the compensatory rise in memory deficit. BDNF might be the early screening indicator of memory deficit in long-term T2DM before AD was defined. Our results also support a role for the Val66Met BDNF polymorphism in episodic memory. Given that either AD or MCI is a complex disorder, the overall susceptibility cannot be fully explained by diabetes. The specificity of the correlation between Val66Met polymorphisms and the respective cognition domains should be explored further as it may provide insights into the development and progression of various subtypes of AD and MCI.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

AUTHOR CONTRIBUTIONS

SW: study concept, design, and interpretation. XH: writing, analysis, and interpretation. ZX and CW: acquisition of data and study supervision. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphys.2021.686838/full#supplementary-material
REFERENCES

Akbarian, S. A., Salehi-Abargouei, A., Pourmasoumi, M., Kelishadi, R., Nikpour, P., and Heidari-Beni, M. (2018). Association of brain-derived neurotrophic factor gene polymorphisms with body mass index: a systematic review and meta-analysis. Adv. Med. Sci. 63, 43–56. doi: 10.1016/j.admsc.2017.07.002

Alberti, K. G., and Zimet, P. Z. (1998). Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. Diabet. Med. 15, 539–553. doi: 10.1002/(SICI)1096-9166(19980715)15:7<539::AID-DIA1668>3.0.CO;2-S

Azeredo, L. A., De Nardi, T., and Grassi-Oliveira, R. (2017). BDNF Val66Met polymorphism and memory performance in older adults: the Met carrier effect is more complex than previously thought: authors’ reply. Braz. J. Psychopharmacol. 39, 276–277. doi: 10.1590/1516-4446-2017-3904

Arietti, M., Giuli, C., and Conti, F. (2018). Peripheral blood brain-derived neurotrophic factor as a biomarker of Alzheimer’s disease: are there methodological biases? Mol. Neurobiol. 55, 6661–6672. doi: 10.1007/s12055-017-0866-y

Barha, C. K., Liu-Ambrose, T., Best, J. R., Yaffe, K., Rosano, C., Health, A., et al. (2019). Sex-dependent effect of the BDNF Val66Met polymorphism on executive functioning and processing speed in older adults: evidence from the health ABC study. Neurobiol. Aging 74, 161–170. doi: 10.1016/j.neurobiolaging.2018.10.021

Binder, D. K., and Scharfman, H. E. (2004). Brain-derived neurotrophic factor. Mol. Neurobiol. 276–277. doi: 10.1590/1516-4446-2017-0866-y

Blesch, A. (2006). Neurotrophic factors in neurodegeneration. J. Pathol. 16, 295–303. doi: 10.1111/j.1756-6659.2006.00006.x

Boyuk, B., Degirmencioglu, S., Atalay, H., Guzel, S., Acar, A., Celebi, A., et al. (2014). Relationship between levels of brain-derived neurotrophic factor and metabolic parameters in patients with type 2 diabetes mellitus. J. Diabetes Res. 2014:978143. doi: 10.1155/2014/978143

Cathomas, F., Vogler, C., Euler-Sigmund, J. C., De Quervain, D. J., and Papassotiropoulos, A. (2016). Fine-mapping of the brain-derived neurotrophic factor (BDNF) gene supports an association of the Val66Met polymorphism with episodic memory. Int. J. Neuropsychopharmacol. 13, 975–980. doi: 10.1017/S1461145710000519

Daily, J. W., and Park, S. (2017). Interaction of BDNF rs6265 variants and energy metabolism parameters in patients with type 2 diabetes mellitus. Mol. Neurobiol. 53, 439–449. doi: 10.1007/s12055-015-9461-2

Lim, Y. Y., Laws, S. M., Perin, S., Pietrzak, R. H., Fowler, C., Masters, C. L., et al. (2021). BDNF Val66Met polymorphism and memory decline across the spectrum of Alzheimer’s disease. Genes Brain Behav. 20, e12724. doi: 10.1111/gbb.12724

Lima Gibascio, B., Dooodrin, J., Klein, H. C., Dierckx, R. A. J. O., Bromberg, E., and de Vries, E. F. J. (2019). Brain-Derived Neurotrophic Factor in Brain Disorders: focus on neuroinflammation. Mol. Neurobiol. 56, 3295–3312.

Marosi, K., and Mattson, M. P. (2014). BDNF mediates adaptive brain and body responses to energetic challenges. Trends Endocrinol. Metab. 25, 89–98. doi: 10.1016/j.tem.2013.10.006

Mauvais-Jarvis, F., Sobngwi, E., Porcher, R., Rivelino, J. P., Kevorkian, J. P., Vasse, C., et al. (2004). Ketosis-prone type 2 diabetes in patients of sub-Saharan African origin: clinical pathophysiology and natural history of beta-cell dysfunction and insulin resistance. Diabetes 53, 645–653. doi: 10.2337/diabetes.53.3.645

Miranda, M., Morici, J. F., Sanzoni, M. B., and Bekinschtein, P. (2019). Brain-Derived Neurotrophic Factor: a Key Molecule for Memory in the Healthy and the Pathological Brain. Front. Cell Neurosci. 13:363. doi: 10.3389/fncel.2019.00363

Moversatt, J., Delangre, E., Liu, J., Gu, Y., and Janel, N. (2019). Hypothesis and Theory: circulating Alzheimer’s-Related Biomarkers in Type 2 Diabetes. Insight From the Goto-Kakizaki Rat. Front. Neurol. 10:649. doi: 10.3389/fneur.2019.00649

Nagahara, A. H., and Tuszyński, M. H. (2011). Potential therapeutic uses of BDNF in neurological and psychiatric disorders. Nat. Rev. Drug Discov. 10, 209–219. doi: 10.1038/nrd3366

Nagata, T., Kobayashi, N., Shimagawa, S., Yamada, H., Kondo, K., and Nakayama, K. (2014). Plasma BDNF levels are correlated with aggressiveness in patients with amnestic mild cognitive impairment or Alzheimer disease. J. Neural Transm. 121, 433–441. doi: 10.1007/s00702-013-1121-y

Nasreddine, Z. S., Phillips, N. A., Bedirian, V., Charbonneau, S., Whitehead, V., Collin, L., et al. (2005). The Montreal Cognitive Assessment, MoCA: a brief screening tool for mild cognitive impairment. J. Am. Geriatr. Soc. 53, 695–699. doi: 10.1111/j.1532-5415.2005.53221.x

Ng, T. K. S., Ho, C. S. H., Tam, W. W. S., Kua, E. H., and Ho, R. C. (2019). Decreased Serum Brain-Derived Neurotrophic Factor (BDNF) Levels in Patients with Alzheimer’s Disease (AD): a Systematic Review and Meta-Analysis. Int. J. Mol. Sci. 20:257. doi: 10.3390/ijms20020257

Pan, W., Banks, W. A., Fasold, M. B., Bluth, J., and Kastin, A. J. (1998). Transport of brain-derived neurotrophic factor across the blood-brain barrier. Neuropharmacology 37, 1553–1561. doi: 10.1016/0028-3908(98)00141-5

Portet, E., Hassane, P. J., Visser, P. J., Friess, G. B., Nohilly, F., Scheltens, P., et al. (2006). Mild cognitive impairment (MCI) in medical practice: a critical review of the concept and new diagnostic procedure. Report of the MCI Working Group of the European Consortium on Alzheimer’s Disease. J. Neuro. Neurosurg. Psychiatry 77, 714–718. doi: 10.1136/jnnp.2005.085332
Shen, T., You, Y., Joseph, C., Mirzaei, M., Klistorner, A., Graham, S. L., et al. (2018). BDNF Polymorphism: a Review of Its Diagnostic and Clinical Relevance in Neurodegenerative Disorders. *Aging Dis.* 9, 523–536. doi: 10.14336/AD.2017.0717

Suwa, M., Kishimoto, H., Nofuji, Y., Nakano, H., Sasaki, H., Radak, Z., et al. (2006). Serum brain-derived neurotrophic factor level is increased and associated with obesity in newly diagnosed female patients with type 2 diabetes mellitus. *Metabolism* 55, 852–857. doi: 10.1016/j.metabol.2006.02.012

Toh, Y. L., Ng, T., Tan, M., Tan, A., and Chan, A. (2018). Impact of brain-derived neurotrophic factor genetic polymorphism on cognition: a systematic review. *Brain Behav.* 8:e01009. doi: 10.1002/brb3.1009

van den Berg, E., Kloppenborg, R. P., Kessels, R. P., Kappelle, L. J., and Biessels, G. J. (2009). Type 2 diabetes mellitus, hypertension, dyslipidemia and obesity: a systematic comparison of their impact on cognition. *Biochim. Biophys. Acta* 1792, 470–481. doi: 10.1016/j.bbadis.2008.09.004

von Bohlen Und Halbach, O., and von Bohlen Und Halbach, V. (2018). BDNF effects on dendritic spine morphology and hippocampal function. *Cell Tissue Res.* 373, 729–741. doi: 10.1007/s00441-017-2782-x

Weinstein, G., Preis, S. R., Beiser, A. S., Kaess, B., Chen, T. C., Satizabal, C., et al. (2017). Clinical and Environmental Correlates of Serum BDNF: a Descriptive Study with Plausible Implications for AD Research. *Curr. Alzheimer Res.* 14, 722–730. doi: 10.2174/1567205014666170203094520

Weis, J., Saxena, S., Evangelopoulos, M. E., and Kruttgen, A. (2003). Trophic factors in neurodegenerative disorders. *IUBMB Life* 55, 353–357. doi: 10.1080/1521654031000153021

Zangeneh, F., Arora, P. S., Dyck, P. J., Bekris, L., Lernmark, A., Achenbach, S. J., et al. (2006). Effects of duration of type 2 diabetes mellitus on insulin secretion. *Endocr. Pract.* 12, 388–393. doi: 10.4158/EP.12.4.388

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