Biomarkers and clinical scores to aid the identification of disease severity and intensive care requirement following activation of an in-hospital sepsis code

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Abstract
Background: Few validated biomarker or clinical score combinations exist which can discriminate between cases of infection and other non-infectious conditions following activation of an in-hospital sepsis code, as well as provide an accurate severity assessment of the corresponding host response. This study aimed to identify suitable blood biomarker (MR-proADM, PCT, CRP and lactate) or clinical score (SOFA and APACHE II) combinations to address this unmet clinical need.

Methods: A prospective, observational study of patients activating the Vall d’Hebron University Hospital sepsis code (ISC) within the emergency department (ED), hospital wards and intensive care unit (ICU). Area under the receiver operating characteristic (AUROC) curves, logistic and Cox regression analysis were used to assess performance.

Results: 148 patients fulfilled the Vall d’Hebron ISC criteria, of which 130 (87.8%) were retrospectively found to have a confirmed diagnosis of infection. Both PCT and MR-proADM had a moderate-to-high performance in discriminating between infected and non-infected patients following ISC activation, although the optimal PCT cut-off varied significantly across departments. Similarly, MR-proADM and SOFA performed well in predicting 28- and 90-day mortality within the total infected patient population, as well as within patients presenting with a community-acquired infection or following a medical emergency or prior surgical procedure. Importantly, MR-proADM also showed a high association with the requirement for ICU admission after ED presentation [OR (95% CI) 8.18 (1.75–28.33)] or during treatment on the ward [OR (95% CI) 3.64 (1.43–9.29)], although the predictive performance of all biomarkers and clinical scores diminished between both settings.

Conclusions: Results suggest that the individual use of PCT and MR-proADM might help to accurately identify patients with infection and assess the overall severity of the host response, respectively. In addition, the use of MR-proADM could accurately identify patients requiring admission onto the ICU, irrespective of whether patients...
Introduction
An early diagnosis of sepsis, irrespective of hospitalisation setting, is crucial in order to provide a rapid and appropriate therapeutic response. This, however, may be complicated by the complex and heterogeneous host response to infection, as well as the high level of patient heterogeneity between departments [1].

A standardised set of defined criteria to aid in the early identification of sepsis therefore remains elusive, despite the established use of specific parameters in conditions such as myocardial infarction and pulmonary embolism [2]. Furthermore, the poor acceptance of previous [3] and current [4, 5] definitions has led to significant debate concerning the most appropriate clinical criteria required to make an accurate diagnosis. Differential presenting signs, symptoms and severities at each stage of hospitalisation, as well as prior surgical history and the setting of the original infectious insult, significantly exacerbate this already complex diagnosis.

The construction and validation of new parameters to aid in the early diagnosis and severity assessment of sepsis patients is therefore crucial. The recent development of tools such as the quick Sequential Organ Failure Assessment (qSOFA) score to screen patients at risk of a poor outcome has resulted in additional parameters becoming available to the treating clinician [6]; however, significant inherent and kinetical limitations make its implementation problematic [7–9]. The use of a panel of blood biomarkers, however, may provide a more elegant solution. Whilst measurement of individual biomarkers to concurrently discriminate between infectious and non-infectious patients and provide an accurate assessment of disease severity is highly desirable [10], no such biomarker appears to have a high enough accuracy to fulfil both clinical requirements. Accordingly, both requirements should be considered separately.

The use of procalcitonin (PCT) and C-reactive protein (CRP) to discriminate between infectious and non-infectious cases is well documented [11–15], despite neither biomarker being well-established in the field of emergency medicine or on the medical ward. Furthermore, neither biomarkers exhibit a robust performance in terms of mortality prediction or in the identification of patients requiring intensive care (ICU) admission. Conversely, the novel blood biomarker, mid-regional proadrenomedullin (MR-proADM) has been shown to be elevated in the early stages of infectious disease progression in the emergency department (ED) [16], after severe burn injury on a medical ward [17], and in critically ill sepsis patients with decreasing PCT concentrations [1] who may require renal replacement therapy (RRT) [18], or are likely to progress towards multiple organ failure [19].

This study therefore aimed to investigate the performance of each biomarker (MR-proADM, PCT, CRP and lactate) and clinical score (SOFA and APACHE II) in patients fulfilling the criteria for the Vall d’Hebron University in-hospital sepsis code (ISC) in order to: (1) discriminate between infected and non-infected patients; (2) assess infection severity according to 28- and 90-day mortality prediction, and (3) identify patients presenting to the ED or undergoing treatment on a medical/surgical ward who require subsequent ICU admission.

Methods
Study design and ethical approval
This single-centre, observational study prospectively enrolled consecutive patients across all hospital departments (ED, ward and ICU) who met the criteria for the activation of the Vall d’Hebron University Hospital in-hospital sepsis code (ISC) [20] between November 2016 and August 2017. The study protocol was approved by the Clinical Research Ethics Committee of Vall d’Hebron University Hospital (PR(AG)333/2016) without the need for informed consent, and was performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments.

Inclusion and exclusion criteria
Inclusion criteria comprised adult patients ≥ 18 years of age presenting with either a suspected or documented infection with the presence of at least one of the following sets of variables, as outlined by the Vall d’Hebron University Hospital in-hospital sepsis code (ISC) [20]: (1) an acute alteration in the level of consciousness not explained by other clinical conditions, or (2) the presence of hyperthermia (axillary temperature > 38.3 °C) or hypothermia (axillary temperature < 36.0 °C), and/or tachycardia (> 110 beats per minute), tachypnoea (> 30 breaths per minute) or desaturation (SpO₂ < 90%), as well as arterial hypotension (systolic arterial pressure < 90 mmHg or mean arterial pressure < 65 mmHg, or a decrease of > 40 mmHg of baseline systolic arterial pressure). Exclusion criteria included non-adult patients, pregnancy, or patients where no blood sample could be obtained.
Study endpoints and analytical aims
Study endpoints and analytical aims were defined as follows. *Infection identification*: either proven infection resulting from a positive blood culture or microbial identification, or a final clinical diagnosis of infection. *28- and 90-day mortality*: all-cause mortality within either 28 or 90 days following enrolment. *Community-acquired infection*: infectious insult developed outside the hospital within the community. *Hospital-acquired infection*: infectious insult developed whilst a patient was hospitalised for a previously non-infectious complaint. *ICU admission*: admission onto either a medical or surgical intensive care unit within 28 days of sepsis code activation.

Data collection and biomarker measurements
Patient comorbidities and demographics were subsequently noted upon sepsis code activation, as well as data concerning triage, routine laboratory values, microbiology testing and final clinical diagnosis. Clinical scores (APACHE II and SOFA) were retrospectively calculated whenever possible upon enrolment. The following biomarker measurements were performed as standard: PCT using a chemiluminescent immunoassay (CLIA), CRP using an immuno-turbidimetric test, and L-lactate using an enzymatic colour test. MR-proADM was retrospectively analysed following blood sampling through a central catheter, and stored at −80 °C until batch analysis using TRACE technology (time-resolved amplified cryptate emission, KRYPTOR platform, Thermo Fisher). Results were unavailable to the treatment physician throughout patient enrolment and hospitalisation.

Statistical analysis
Data were either reported using the mean (standard deviation) for symmetrically distributed variables, or the median [first quartile – third quartile] for variables showing a skewed distribution. Demographic and clinical characteristics between surviving and non-surviving patients up to 28 days following sepsis code activation were assessed using the Chi-square ($\chi^2$) test for categorical variables, Student’s $t$ test for age, and the Mann–Whitney $U$ test for all other continuous variables. Area under the receiver operating characteristic (AUROC) curves were used to identify the biomarker or clinical score with the greatest predictive value for each endpoint, with 95% confidence intervals (95% CI) compared to determine significance. Youden’s criterion established optimal cut-off values with corresponding sensitivity and specificity values. Univariate and multivariate Cox regression models assessed the association with survival time (censored at 28 days following sepsis code activation), with potential confounding variables selected based on a univariate analysis ($p$-value < 0.005 after applying a Bonferroni correction), and subsequently included in a multivariate analysis. Results were either presented as the hazard (HR) or odds ratio (OR) per 1 interquartile-range increase for mortality prediction and ICU admission, respectively, with corresponding 95% confidence intervals. Previously established [16] biomarker cut-offs for 28-day mortality prediction were further analysed within this dataset, and mortality rates in patient subgroups with high biomarker values compared to those with low values. All data were analysed using the statistics software R (version 3.1.2).

Results
Patient characteristics
A total of 150 patients were identified and enrolled after activation of the sepsis code (sepsis code group), comprising 43 (29.1%) presenting to the emergency department (ED), 70 (47.3%) already undergoing treatment on a medical or surgical ward, and 35 (23.6%) already present on the intensive care unit (ICU). 2 patients were removed from the analysis due to a lack of blood sample.

Infection identification following sepsis code activation
Based on final clinical diagnosis, a total of 130 (87.8%) infected (sepsis code infected group) and 18 (12.2%) non-infected patients (sepsis code non-infected group) could be retrospectively identified following initial sepsis code activation. Patient characteristics of the infected population are presented in Table 1. PCT was found to have a high discriminatory performance (Fig. 1a), with an optimal cut-off of 2.02 ng/mL (sensitivity and specificity: 0.73 and 0.78). This cut-off, however, varied considerably depending on the location of sepsis code activation, with an optimal cut-off of 0.37 ng/mL (sensitivity and specificity: 0.86 and 0.75) in the ED, 1.91 ng/mL (sensitivity and specificity: 0.77 and 0.86) on the ward and 2.02 mg/mL (sensitivity and specificity: 0.79 and 0.71) on the ICU.

In the sepsis code infected group, a positive blood culture rate was obtained in 54 (42.2%) cases, with a greater prevalence of Gram-positive as opposed to Gram-negative infections ($N = 34$; 26.2% vs. $N = 17$; 13.1%, $p < 0.01$). A comparison of biomarker and clinical score performance found that PCT had a moderate performance in predicting the likelihood of a positive culture (Fig. 1b). The addition of further biomarkers or clinical scores to initial PCT measurements resulted in no significant increase in discriminatory (infected vs. non-infected patients) or predictive (positive vs. negative blood culture) performance.
### Table 1: Clinical patient characteristics upon activation of the sepsis code with respect to the total infected patient population and subsequent 28-day mortality

|                                    | Patient population ($N=130$) | Survivors ($N=102$) | Non-survivors ($N=28$) | $p$-value |
|------------------------------------|------------------------------|---------------------|------------------------|-----------|
| **Age (years) (mean, S.D.)**       | 63.1 (15.2)                  | 62.3 (15.2)         | 66.1 (15.0)            | 0.205     |
| **Male gender ($N$, %)**           | 83 (63.8%)                   | 61 (59.8%)          | 22 (78.6%)             | 0.348     |
| **Infection characterisation**     |                              |                     |                        |           |
| Infection no SIRS ($N$, %)         | 7 (5.4%)                     | 6 (5.9%)            | 1 (3.6%)               | 0.623     |
| Severe sepsis ($N$, %)             | 41 (31.8%)                   | 37 (36.3%)          | 4 (14.3%)              | 0.028     |
| Septic shock ($N$, %)              | 82 (63.1%)                   | 59 (57.8%)          | 23 (82.1%)             | 0.031     |
| Sepsis-2 ($N$, %)                  | 122 (93.8%)                  | 95 (93.1%)          | 27 (96.4%)             | 0.814     |
| Sepsis-3 ($N$, %)                  | 116 (89.2%)                  | 88 (86.3%)          | 28 (100.0%)            | 0.157     |
| **Location of sepsis code activation** |                              |                     |                        |           |
| Emergency department ($N$, %)      | 39 (30.0%)                   | 33 (32.4%)          | 6 (21.4%)              | 0.568     |
| Ward ($N$, %)                      | 63 (48.5%)                   | 47 (46.1%)          | 16 (57.1%)             | 0.729     |
| ICU ($N$, %)                       | 28 (21.5%)                   | 22 (21.6%)          | 6 (21.4%)              | 0.839     |
| Surgical admissions ($N$, %)       | 47 (36.2%)                   | 37 (36.3%)          | 10 (35.7%)             | 0.487     |
| Medical admissions ($N$, %)        | 83 (63.8%)                   | 65 (63.7%)          | 18 (64.3%)             | 0.487     |
| ICU length of stay (days) (median, IQR) | 6 [2–15]                   | 5 [2–11]            | 12 [3–24]              | 0.093     |
| Hospital length of stay (days) (median, IQR) | 16.5 [8–31] | 17 [8–33]        | 16 [5.5–27]            | 0.298     |
| **Life supporting and intensive care therapies** |                              |                     |                        |           |
| Vasopressors ($N$, %)              | 67 (51.5%)                   | 52 (40.0%)          | 15 (53.6%)             | 0.106     |
| Renal replacement therapy ($N$, %) | 15 (11.5%)                   | 8 (6.2%)            | 7 (25.0%)              | 0.009     |
| Mechanical ventilation ($N$, %)    | 44 (33.8%)                   | 30 (23.1%)          | 14 (50.0%)             | 0.011     |
| Mechanical ventilation duration (days) | 7 [3–16.25]             | 5 [3–17.5]          | 10 [4.5–14.75]         | 0.081     |
| High-flow nasal cannula use ($N$, %) | 19 (14.6%)                     | 16 (15.7%)          | 3 (10.7%)              | 0.386     |
| **Pre-existing comorbidities**     |                              |                     |                        |           |
| Cardiopathy ($N$, %)               | 24 (18.5%)                   | 16 (15.7%)          | 8 (28.6%)              | 0.120     |
| Chronic kidney disease ($N$, %)    | 19 (14.6%)                   | 14 (13.7%)          | 5 (17.6%)              | 0.584     |
| COPD ($N$, %)                      | 19 (14.6%)                   | 10 (9.8%)           | 9 (32.1%)              | 0.003     |
| Immunosuppression ($N$, %)         | 63 (48.5%)                   | 44 (33.4%)          | 19 (67.9%)             | 0.020     |
| Liver cirrhosis ($N$, %)           | 3 (2.3%)                     | 1 (1.0%)            | 2 (7.1%)               | 0.054     |
| **Microbiology**                   |                              |                     |                        |           |
| Positive blood culture ($N$, %)    | 54 (41.5%)                   | 40 (39.2%)          | 14 (50.0%)             | 0.238     |
| Gram positive ($N$, %)             | 34 (26.2%)                   | 24 (23.5%)          | 10 (35.7%)             | 0.895     |
| Gram negative ($N$, %)             | 17 (13.1%)                   | 13 (12.7%)          | 4 (14.3%)              | 0.797     |
| Fungal ($N$, %)                    | 2 (1.5%)                     | 1 (1.0%)            | 2 (7.1%)               | 0.529     |
| **Origin of infection**            |                              |                     |                        |           |
| Abdominal ($N$, %)                 | 41 (31.5%)                   | 35 (34.3%)          | 6 (21.4%)              | 0.194     |
| Bacteria—primary ($N$, %)          | 4 (3.1%)                     | 3 (2.9%)            | 1 (3.6%)               | 0.864     |
| Catheter-related ($N$, %)          | 8 (6.2%)                     | 5 (4.9%)            | 3 (10.7%)              | 0.257     |
| Central nervous system ($N$, %)    | 2 (1.5%)                     | 1 (1.0%)            | 1 (3.6%)               | 0.324     |
| Respiratory ($N$, %)               | 34 (26.2%)                   | 25 (24.5%)          | 9 (32.1%)              | 0.416     |
| Soft-tissue ($N$, %)               | 4 (3.1%)                     | 4 (3.9%)            | 0 (0.0%)               | 0.287     |
| Urinary ($N$, %)                   | 30 (23.1%)                   | 25 (24.5%)          | 5 (17.9%)              | 0.459     |
| Unknown ($N$, %)                   | 3 (2.3%)                     | 1 (1.0%)            | 2 (7.1%)               | 0.054     |
| Other ($N$, %)                     | 4 (3.1%)                     | 3 (2.9%)            | 1 (3.6%)               | 0.864     |
| **Source control**                 |                              |                     |                        |           |
| Focus cleaning ($N$, %)            | 34 (26.2%)                   | 28 (27.5%)          | 6 (21.4%)              | 0.521     |
| Drainage ($N$, %)                  | 8 (6.2%)                     | 6 (5.9%)            | 2 (7.1%)               | 0.806     |
| Surgery ($N$, %)                   | 23 (17.7%)                   | 20 (19.6%)          | 3 (10.7%)              | 0.275     |
| **Biomarker and severity scores**  |                              |                     |                        |           |
| MR-proADM (nmol/L) (median, IQR)   | 3.54 [1.89–6.69]             | 3.18 [1.73–5.64]    | 5.69 [3.98–13.43]      | < 0.001   |
28- and 90-day mortality prediction

The 28-day all-cause mortality rate within the sepsis code infected group was 21.5% (N = 28), with a higher mortality rate in patients enrolled whilst on the ward (N = 16; 25.4%) and ICU (N = 6; 21.4%) as opposed to in the ED (N = 6; 15.4%; Table 1). The 90-day all-cause mortality rate was 24.6% (N = 32). In both cases, MR-proADM, lactate and SOFA were all significantly elevated in non-surviving as opposed to surviving patients (p < 0.01), whereas no significant differences were found in PCT, CRP or APACHE II values.
AUROC analysis found that MR-proADM performed well in predicting 28-day mortality, followed by SOFA and lactate, with an optimal cut-off of 4.28 nmol/L (sensitivity and specificity: 0.76 and 0.65). No significant association was observed with either PCT, CRP or APACHE II (Fig. 2). Similar findings were observed in both the univariate Cox regression analysis and after adjusting for the influence of existing chronic obstructive pulmonary disease (COPD), which was the only pre-existing variable found to significantly influence mortality prediction. The addition of PCT to MR-proADM measurements further improved predictive performance [HR (95% CI) 7.5 (3.7–15.2)], greater than any other biomarker or clinical score combination. Similar results were also obtained for 90-day mortality prediction, with a moderate performance found using MR-proADM, followed by SOFA and lactate. The influence of existing COPD had little impact on predictive performance, whilst the addition of PCT to MR-proADM resulted in the greatest increase in predictive performance [HR (95% CI) 5.7 (3.0–10.9)]. Interestingly, no increase in performance was found for the combination of PCT with either SOFA or lactate measurements [HR (95% CI) 2.9 (1.9–4.6) and 2.1 (1.3–3.2)], respectively.

Using an MR-proADM cut-off of 1.54 nmol/L within the infected patient population, 108 (83.1%) patients were found to have values equal to or exceeding this cut-off, with a corresponding mortality rate of 24.1% (N=26). In comparison, patients with MR-proADM values <1.54 nmol/L (N=22, 16.9%) had a significantly lower mortality rate of 7.3% (N=22, 16.9%).

| Biomarker or clinical score | Patients (N) | Events (N) | AUROC | LR $\chi^2$ | HR IQR [95% CI] | LR $\chi^2$ | HR IQR [95% CI] |
|-----------------------------|--------------|------------|--------|-------------|-----------------|-------------|-----------------|
| **28 day mortality**        |              |            |        |             |                 |             |                 |
| MR-proADM                   | 130          | 28         | 0.75 [0.64 - 0.86] | 23.61 | 4.37 [2.37 - 8.06] | 23.70 | 4.34 [2.34 - 8.05] |
| PCT                         | 127          | 28         | 0.52 [0.40 - 0.64] | 0.00 | 1.01 [0.56 - 1.81] | 1.48 | 0.97 [0.54 - 1.74] |
| CRP                         | 129          | 28         | 0.55 [0.43 - 0.68] | 1.01 | 1.24 [0.79 - 1.92] | 2.42 | 1.24 [0.79 - 1.95] |
| Lactate                     | 116          | 26         | 0.67 [0.54 - 0.80] | 9.64 | 2.17 [1.35 - 3.50] | 10.39 | 2.17 [1.33 - 3.53] |
| APACHE II                   | 64           | 14         | 0.55 [0.43 - 0.68] | 0.13 | 1.14 [0.55 - 2.37] | 0.13 | 1.14 [0.55 - 2.37] |
| SOFA                        | 130          | 28         | 0.73 [0.62 - 0.84] | 21.00 | 3.06 [1.87 - 5.01] | 22.42 | 3.07 [1.88 - 5.02] |
| **90 day mortality**        |              |            |        |             |                 |             |                 |
| MR-proADM                   | 130          | 32         | 0.70 [0.58 - 0.81] | 18.39 | 3.36 [1.92 - 5.88] | 18.56 | 3.31 [1.87 - 5.84] |
| PCT                         | 127          | 32         | 0.54 [0.42 - 0.65] | 0.06 | 0.94 [0.54 - 1.62] | 1.85 | 0.90 [0.52 - 1.55] |
| CRP                         | 129          | 32         | 0.58 [0.46 - 0.69] | 1.91 | 1.33 [0.86 - 2.06] | 3.54 | 1.33 [0.85 - 2.09] |
| Lactate                     | 116          | 30         | 0.65 [0.53 - 0.78] | 8.69 | 2.00 [1.27 - 3.13] | 9.59 | 1.99 [1.26 - 3.15] |
| APACHE II                   | 64           | 18         | 0.53 [0.37 - 0.69] | 0.02 | 1.04 [0.55 - 1.96] | 0.11 | 1.04 [0.55 - 1.96] |
| SOFA                        | 130          | 32         | 0.71 [0.61 - 0.82] | 19.97 | 2.77 [1.75 - 4.36] | 21.59 | 2.78 [1.75 - 4.39] |

**Fig. 2** AUROC, univariate and bivariate analysis for the prediction of 28-day **a** and 90-day **b** mortality for each biomarker and clinical score. APACHE II, Acute Physiological and Chronic Health Evaluation II score; AUROC area under the receiver operating characteristic curve; CI confidence interval; CRP C-reactive protein; HR hazard ratio; IQR interquartile range; LR likelihood ratio; MR-proADM mid-regional proadrenomedullin; N number; PCT procalcitonin; SOFA Sequential Organ Failure Assessment score.
(p < 0.01) lower mortality rate of 13.6% (N = 3). No patient died with an MR-proADM value of < 0.88 nmol/L. Only 6 (4.6%) patients were found to have PCT concentrations < 0.25 μg/mL, of which one died within 28 days.

Mortality prediction based on location of infection development and reason for patient admission

Subgroup analysis found that a total of 67 (51.5%) patients developed an infection in the community and were subsequently hospitalised, compared to 48 (36.9%) patients where the infection was acquired during hospitalisation despite being admitted for a non-infectious related condition. The 28-day mortality rates of both groups were, however, similar (N = 15; 22.4% vs. N = 12; 25.0%). Likewise, 47 (36.2%) patients were identified using the sepsis code following a surgical procedure, as opposed to 83 (63.8%) patients presenting with a medical complaint, again with similar 28-day mortality rates between groups (N = 10; 21.3% vs. N = 18; 21.7%).

AUROC and Cox regression analysis found that MR-proADM had a high predictive value in patients with community-acquired infections, followed by SOFA and lactate (Table 2), which could be further increased after the addition of PCT [HR (95% CI) 10.0 (3.6–27.5)]. The optimal MR-proADM cut-off was 3.67 nmol/L, with a sensitivity and specificity of 0.93 and 0.52, respectively. Conversely, the SOFA score had high predictive performance in patients with hospital-acquired infections (Table 2). Interestingly, the combination of MR-proADM with APACHE II resulted in the largest increase in predictive value [HR (95% CI) 7.9 (1.5–41.1)], whereas the addition of PCT to MR-proADM only resulted in a small overall additive effect. Similarly, AUROC and Cox regression analysis found that MR-proADM had a moderate-to-high predictive value within both medical and surgical patient subgroups, followed by SOFA and lactate (Table 3). The combination of SOFA and APACHE II scores resulted in the greatest predictive performance in the surgical cohort [HR (95% CI) 9.36 (2.13–4.17)], compared to the combination of MR-proADM and PCT in the medical cohort [HR (95% CI) 9.30 (3.89–22.25)].

Intensive care unit admission after sepsis code activation

Finally, biomarker and clinical score performance was compared in order to predict admission to both the surgical and medical intensive care units. Activation of the sepsis code in both the ED and wards resulted in a high rate of admission on the same or subsequent day, with a total of 16 (41.0%) patients transferred from the ED and 28 (45.9%) patients transferred from the ward. MR-proADM showed a moderate-to-high predictive value in both settings, with similar results also found after univariate logistic regression (Table 4).

Discussion

This study compared the performance of a number of clinical severity scores, conventional and novel biomarkers following activation of an in-hospital sepsis code, in order to identify cases of infection and assess the severity of corresponding host response. Results found that the use of PCT to discriminate between sepsis code infected group and sepsis code non-infected group, and MR-proADM to identify disease severity irrespective of the location of infectious insult, had the greatest accuracy in

Table 2 28-day mortality prediction in patients with either community or hospital-acquired infections

| Biomarker or clinical score | Patients (N) | Events (N) | AUROC | Univariate | Bivariate |
|----------------------------|--------------|------------|-------|------------|-----------|
|                            |              |            |       | LR \(\chi^2\) | HR IQR [95% CI] | LR \(\chi^2\) | HR IQR [95% CI] |
| Community                  |              |            |       |            |           |            |              |
| MR-proADM                  | 67           | 15         | 0.76  | 0.64–0.89  | 13.08     | 4.60 [1.95–10.83] | 14.52 | 4.88 [1.96–12.14] |
| PCT                        | 65           | 15         | 0.54  | 0.36–0.71  | 0.09      | 0.89 [0.43–1.85]  | 1.64  | 0.89 [0.43–1.85]  |
| CRP                        | 66           | 15         | 0.57  | 0.40–0.74  | 0.48      | 1.28 [0.62–2.63]  | 2.30  | 1.32 [0.62–2.80]  |
| Lactate                    | 60           | 14         | 0.71  | 0.53–0.89  | 10.03     | 3.44 [1.59–7.45]  | 10.61 | 3.09 [1.40–6.83]  |
| APACHE II                  | 33           | 8          | 0.70  | 0.50–0.90  | 2.10      | 2.07 [0.77–5.55]  | 2.79  | 2.14 [0.76–6.03]  |
| SOFA                       | 67           | 15         | 0.73  | 0.57–0.88  | 8.93      | 2.85 [1.39–5.85]  | 11.16 | 3.01 [1.44–6.28]  |
| Hospital                   |              |            |       |            |           |            |              |
| MR-proADM                  | 48           | 11         | 0.66  | 0.45–0.88  | 5.59      | 3.15 [1.19–8.37]  | 5.70  | 3.32 [1.19–9.25]  |
| PCT                        | 48           | 11         | 0.46  | 0.27–0.65  | 0.03      | 0.91 [0.30–2.77]  | 0.19  | 0.88 [0.29–2.70]  |
| CRP                        | 48           | 11         | 0.53  | 0.32–0.73  | 0.52      | 1.20 [0.70–2.07]  | 0.61  | 1.19 [0.69–2.06]  |
| Lactate                    | 43           | 10         | 0.58  | 0.35–0.81  | 0.87      | 1.40 [0.70–2.77]  | 0.87  | 1.40 [0.70–2.77]  |
| APACHE II                  | 25           | 6          | 0.63  | 0.37–0.89  | 0.85      | 0.61 [0.21–1.74]  | 1.18  | 0.58 [0.19–1.75]  |
| SOFA                       | 48           | 11         | 0.78  | 0.61–1.05  | 12.12     | 3.60 [1.70–7.59]  | 12.18 | 3.58 [1.70–7.55]  |

APACHE II Acute Physiological and Chronic Health Evaluation II score, AUROC area under the receiver operating characteristic curve, CI confidence interval, CRP C-reactive protein, HR hazard ratio, IQR interquartile range, LR likelihood ratio of the \(\chi^2\) test, MR-proADM mid-regional proadrenomedullin, N number, PCT procalcitonin, SOFA Sequential Organ Failure Assessment score.
fulfilling both criteria. Interestingly, MR-proADM was also the most accurate parameter in identifying patients presenting to the ED or receiving treatment on a medical ward who subsequently required immediate ICU admission.

The use of clinical or laboratory parameters which are significantly increased in the early stages of the infectious cascade may allow more appropriate treatment strategies to be initiated upon initial suspicion of infection, thus minimising the risk of patients being either under- or over-treated—both of which may result in undesirable consequences [21]. To this extent, the use of blood biomarkers may provide an easy and rapid source of clinical information with which to inform and help develop clinical strategies. Of these biomarkers, PCT has been studied extensively in both the ED and ICU [12–15, 22–24], with routine use in the ICU especially widespread. Conversely, novel biomarkers such as MR-proADM remain poorly understood, despite a number of recent studies [16, 18, 19, 25]. Furthermore, biomarker performance may vary extensively depending on whether the infection had developed in the community or during hospitalisation. Hence a key aim of this study focused on the comparison of biomarker and clinical score performance

### Table 3 28-day mortality prediction in patients enrolled following either surgical or medical complaints

| Biomarker or clinical score | Patients (N) | Events (N) | AUROC | Univariate | Bivariate |
|-----------------------------|--------------|------------|-------|------------|-----------|
|                             |              |            |       |            |           |
| Surgical                    |              |            |       |            |           |
| MR-proADM                   | 47           | 10         | 0.80  | [0.66–0.94]| 9.03      |
| PCT                         | 47           | 10         | 0.47  | [0.26–0.68]| 0.34      |
| CRP                         | 47           | 10         | 0.59  | [0.38–0.79]| 0.37      |
| Lactate                     | 44           | 8          | 0.73  | [0.49–0.97]| 3.97      |
| APACHE II                   | 29           | 7          | 0.53  | [0.29–0.77]| 0.01      |
| SOFA                        | 47           | 10         | 0.74  | [0.56–0.92]| 10.86     |
| Medical                     |              |            |       |            |           |
| MR-proADM                   | 83           | 18         | 0.72  | [0.57–0.87]| 15.05     |
| PCT                         | 80           | 18         | 0.55  | [0.40–0.71]| 0.10      |
| CRP                         | 82           | 18         | 0.46  | [0.30–0.61]| 0.63      |
| Lactate                     | 72           | 18         | 0.66  | [0.51–0.82]| 6.43      |
| APACHE II                   | 35           | 7          | 0.59  | [0.34–0.82]| 0.20      |
| SOFA                        | 83           | 18         | 0.71  | [0.57–0.86]| 16.32     |

### Table 4 Intensive (ICU) and surgical intensive (ICU) care unit admission from the emergency department and ward

| Biomarker or clinical score | Patients (N) | Events (N) | AUROC | Univariate |
|-----------------------------|--------------|------------|-------|------------|
|                             |              |            |       |            |
| Emergency department        |              |            |       |            |
| MR-proADM                   | 39           | 16         | 0.80  | [0.66–0.94]| 10.55     |
| PCT                         | 37           | 16         | 0.74  | [0.57–0.90]| 7.18      |
| CRP                         | 38           | 16         | 0.64  | [0.46–0.82]| 2.83      |
| Lactate                     | 33           | 14         | 0.45  | [0.24–0.65]| 0.72      |
| APACHE II                   | 16           | 14         | 0.63  | [0.33–0.92]| 0.21      |
| SOFA                        | 39           | 16         | 0.78  | [0.64–0.93]| 10.21     |
| Ward admission              |              |            |       |            |
| MR-proADM                   | 63           | 28         | 0.72  | [0.64–0.79]| 10.6      |
| PCT                         | 63           | 28         | 0.59  | [0.46–0.72]| 3.5       |
| CRP                         | 63           | 28         | 0.57  | [0.41–0.65]| 0.8      |
| Lactate                     | 63           | 28         | 0.57  | [0.44–0.70]| 1.7      |
| APACHE II                   | 20           | 16         | 0.69  | [0.40–0.77]| 1.3      |
| SOFA                        | 63           | 28         | 0.64  | [0.49–0.75]| 3.8      |
according to the location of initial infection development, following activation of an in-hospital sepsis code.

Based on recent publications, the use of MR-proADM was of especial interest within our investigation, with previous studies showing elevated concentrations resulting from increased levels of microcirculatory and endothelial damage, resulting in the early stages of subsequent organ dysfunction [26–29]. The use of a biomarker to identify developing microcirculatory damage may contribute significant information following the activation of a local, in-hospital sepsis code.

Accordingly, results from this investigation potentially indicate two clinically important uses of MR-proADM following sepsis code activation. Firstly, MR-proADM may be used as a tool to aid in the early identification of disease severity, irrespective of medical setting or prior surgical history. Secondly, it may also be used to aid clinical decision-making in assessing ICU requirement within the ED or on a medical ward.

This study found that MR-proADM had a high accuracy in identifying both 28-day and 90-day mortality, compared to all other biomarkers and clinical scores, although many differences were not found to be significant. Performance was maintained irrespective of whether the infection was developed in the community, culminating in a visit to the ED, or following a surgical procedure. Conversely, the SOFA score appeared to more accurately predict mortality in patients following a hospital-acquired infection, although similarly, differences were not found to be significant. In each case, however, the small sample size and relatively low event rate result in few significant differences between biomarkers and scores, as highlighted when comparing 95% confidence intervals. Nevertheless, similar findings have been shown elsewhere. Elke et al. [25] previously found differences in performance depending on whether the infecting agent consisted solely of Gram positive vs. negative bacteria, as well as highlighting the influence of prior surgical procedures in patients directly admitted onto the ICU. A further ICU study by Andaluz-Ojeda et al. [2] in 326 patients with severe sepsis or septic shock, as well as that of Enguix-Armada [30] in 388 patients with septic shock, also found MR-proADM to have the highest predictive value compared to conventional biomarkers such as PCT, CRP and lactate, although interestingly, the greatest performance was found in patients with a lower degree of organ failure. Nevertheless, no subgroup analysis was performed in either study relating to location of infection development or prior surgical history. It is, however, interesting to note that similar results to those high severity, critical care investigations could be observed in our less severe cohort across three different hospital departments. Accordingly, a significant body of evidence now appears to have been formed across all hospital departments, highlighting the use of MR-proADM in identifying the patients at risk of mid- and long-term mortality.

In contrast, few studies have assessed MR-proADM performance in identifying patients requiring ICU admission. An early recognition of such patients is crucial, since it has previously been shown that inappropriate ED discharge can result in higher mortality rates in patients initially classified as suitable for outpatient treatment, but later directly admitted onto the ICU. Similar findings were also reported after initial triage onto a medical ward [31]. The relatively high performance of MR-proADM in identifying these patients after presentation to the ED, with a modest performance during treatment on a medical ward, may provide clinicians with an additional tool in assessing suitability for ICU admission. Nevertheless, further observational and interventional studies are required to confirm these initial findings, and determine the actual clinical value compared to standard routine practice.

We note several limitations and strengths of this study that deserve greater discussion. This was a single-centre pilot study with a relatively low number of enrolled patients, resulting in many subgroups being underpowered for their respective endpoints. Results must therefore be treated with caution. Nevertheless, similar results were found in comparison to those previously discussed in the literature, despite the patient population enrolled in this study being preselected on the basis of fulfilling an in-house sepsis criteria, and can therefore be viewed as additional collaborative evidence. Similarly, the number of patients where the in-hospital sepsis code was activated, yet no infection was subsequently found, only amounted to 18 (12.2%) patients. A greater degree of confidence in the discriminatory results between infected and non-infected patients could be achieved with a larger sample size. Finally, biomarker incorporation within a sepsis code requires a short turnaround time to result, which may be achieved using point-of-care (POC) technology. At this point in time, however, no such platform exists for the measurement of MR-proADM, making routine incorporation of this biomarker in a clinical setting problematic.

**Conclusions**

This study found that the use of PCT to discriminate between infected and non-infected cases following activation of an in-hospital sepsis code, followed by measurement of MR-proADM to determine infection severity and assess the requirement for ICU admission, may help guide initial treatment decisions and intensify therapy in high-risk patients. Interventional studies, specifically focussing on MR-proADM, to potentially optimise early patient
treatment strategies are now crucial before implementation in routine clinical use.

Abbreviations
AUROC: area under the receiver operating characteristic curve; CI: confidence interval; CRB-65: severity scores for community-acquired pneumonia; CRP: C-reactive protein; ED: emergency department; HR: hazard ratio; IQR: inter-quartile range; MR-proADM: mid-regional proadrenomedullin; N: number; OR: odds ratio; PCT: procalcitonin; qSOFA: quick Sequential Organ Failure Assessment; RRT: renal replacement therapy; SIRS: Systemic Inflammatory Response Syndrome; SOFA: Sequential Organ Failure Assessment.

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Author contributions
JB, JCRR and RF conceived and designed the study. JB was the primary author, editor of the manuscript and the lead clinical investigator. All authors apart from DCW contributed to the enrolment of patients and sample collection. DCW provided statistical advice and analysed the data. All authors critically reviewed and approved the final manuscript. RF and JCR take responsibility for the paper as a whole. All authors read and approved the final manuscript.

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Availability of data and materials
The datasets used and/or analysed during the present study are available from the corresponding author upon reasonable request.

Ethics approval and consent to participate
The study protocol was approved by the ethics board of the Vall d’Hebron University Hospital (PPIAG333/2016), with no written informed consent required from patients or legal representatives.

Consent for publication
No individual participant data is reported that would require consent to publish from the participant (or legal parent or guardian for children).

Competing interests
All authors have provided information on potential conflicts of interests directly or indirectly related to the work submitted in the journal’s disclosure forms. The authors declare that they have no competing interests.

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