Molecular neurobiological clues to the pathogenesis of bipolar disorder
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Bipolar disorder is a serious psychiatric disorder, with a high heritability and unknown pathogenesis. Recent genome-wide association studies have identified the first loci, implicating genes such as CACNA1C and ANK3. The genes highlight several pathways, notably calcium signalling, as being of importance. Molecular studies suggest that the risk variants impact on gene regulation and expression. Preliminary studies using reprogrammed patient-derived cells report alterations in the transcriptome and in cellular adhesion and differentiation. Mouse models show that genes involved in circadian biology, acting via dopaminergic effects, reproduce aspects of the bipolar phenotype. These findings together represent significant advances in identification of the genetic and molecular basis of bipolar disorder, yet we are still far from an integrated, evidence-based understanding of its aetiology.

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Genomics of bipolar disorder
As with other psychiatric disorders, genome-wide association studies (GWAS) of single nucleotide polymorphisms (SNPs) indicate that the heritability of BD is attributable largely to multiple loci of small effect. Table 1 summarises the current genome-wide significant loci. See [5,7] for recent reviews of BD genetics. Table 1 also notes the gene(s) implicated at each locus, but it is worth emphasising that the ‘causal’ gene, and true risk SNP or haplotype, at each locus remains unknown, hindering the interpretation of the GWAS signals, as is the case for all psychiatric disorders (see [8] for discussion; see below for discussion of some of the BD-implicated genes).

Other studies have begun to investigate genotype–phenotype relationships within BD, and between BD and other disorders. Within BD, genetic loci have been related to variables such as presence of psychotic symptoms, suicidality, and body mass index (e.g. [9]). Some of these analyses have revealed genome-wide associations, but the samples are inevitably smaller than for the overall BD-control comparisons; there is also the problem that detailed phenotyping is often not available. Reflecting the clinical and familial overlaps between BD and other psychiatric disorders, cross-disorder analyses have been carried out. These show substantial sharing of risk loci between BD and schizophrenia [10], and also significant commonalities with major depression, but little with autism or attention-deficit hyperactivity disorder [10,11].
Shedding new genetic light on an old debate, these data support the view that BD and schizophrenia are on a continuum, rather than being discrete disorders, as they have conventionally been classified [12].

Despite the progress, several major limitations and unknowns deserve mention, beyond the need to identify unambiguously the affected gene(s) at each locus. First, the causal genetic variant being tagged by the GWAS SNPs has not been identified. In the absence of known coding variants in linkage disequilibrium, the likelihood is that the risk SNPs alter gene regulation and expression [13]. There is preliminary evidence for this in terms of expression quantitative trait loci (eQTLs) [11*,14*,15] and for some of the individual genes, as discussed below. Second, the current BD GWAS signals explain only a fraction of the heritability. The source of the remainder is unknown, but likely includes many more independent loci, as well as gene–gene interactions (epistasis), gene–environment interactions, and a role for copy number variants (CNVs), although at present the data suggest that the latter are less important in BD than in schizophrenia or autism [12]. Other rare variants may also be involved, including those encoding calcium channel and GABA_A receptor subunits [16,17*].

### Bipolar disorder genes and pathways

Two main approaches are being taken to understand the neurobiological consequences of the genes implicated to date in BD: bioinformatic identification of gene networks and pathways, and empirical studies of the expression and function of individual genes. The latter studies are also addressing the molecular basis of the genetic associations.

Pathway analyses using gene ontology and related methods are being carried out in various ways. A prominent category, for which there is convergent evidence, is calcium signalling, particularly voltage-gated calcium channels, identified in several datasets and using differing approaches. It is notable that calcium signalling was already hypothesised to be important in BD and its treatment [18,19]; it was perhaps less expected that this enhancement of calcium channel genes applies similarly to schizophrenia, and also extends to other disorders [11*].

A range of other pathways are also highlighted in BD by studies which have combined GWAS data with gene expression data; these include hormonal regulation, second messenger systems and glutamatergic signalling [20*] as well as histone and immune pathways [21]. Each of these findings should be seen as provisional and in need of replication, nor least given the inadequacies of gene ontology categories, and the incomplete information being fed into the analyses. Nevertheless, they do provide some of the first steps to a more meaningful integrative molecular classification and understanding of BD.

Table 1

| Genome-wide significant loci for bipolar disorder. |
|-----------------------------------------------|
| Locus | Implicated gene(s) and symbol(s) |
|---|---|
| **Genome-wide significant in BD** | |
| 10q21.2 | Ankyrin 3 (ANK3) |
| 12p13.3 | Calcium channel, voltage-dependent, L-type, alpha 1C subunit (CACNA1C) |
| 11q14.1 | Teneurin transmembrane protein 4 (TENM4, formerly known as ODD4) |
| 19p12 | Neurocan (NCAN) |
| 6q25.2 | Spectrin repeat containing, nuclear envelope 1 (SYNE1) |
| 3p22.2 | Tetratricopeptide repeat and ankyrin repeat containing 1 (TRANK1) |
| 5p15.31 | Adenylate cyclase 2 (ADCY2) |
| 6q16.1 | MicroRNA 2113 (MR2113); POU class 3 homeobox 2 (POU3F2; formerly known as OTF7) |
| 10q24.33 | Arsenite methyltransferase (AS3MT) |
| **Genome-wide significant in BD + schizophrenia (combined)** | |
| 2q32.1 | Zinc finger protein 804A (ZNF804A) |
| 3p21.1 | Inter-alpha-trypsin inhibitor heavy chain 3 (ITIH3); inter-alpha-trypsin inhibitor heavy chain 4 (ITIH4) |
| 16p11.2 | Mitogen-activated protein kinase 3 (MAPK3) |
| **Genome-wide significant in BD + unipolar depression (combined)** | |
| 3p21 | Polynucleotide 1 (PBRM1) |

Adapted from [5,7,49].

Complementing these approaches, the leading BD risk genes are being investigated individually, with the goal of understanding the neurobiology of the gene, the molecular basis of the disease association, and the functional impact of the putative risk variant. Three examples are given here.

*CACNA1C* encodes the L-type calcium channel Cav1.2 subunit and is arguably the best supported BD gene, especially if the various lines of prior evidence for altered calcium signalling in the disorder are taken into account [18,19]. In addition to many studies reporting neuroimaging, cognitive and neurophysiological correlates of *CACNA1C* genotype, molecular studies have sought to show the proximal effect of the BD-associated SNPs. The risk haplotype, including the main SNP, rs1006737, resides within the large (300 kb) third intron, and is hence non-coding. Any functionality is likely to be via an effect on expression or splicing of *CACNA1C* – though, as with all such genetic associations, other explanations are possible (e.g. effects on antisense transcripts, non-coding RNAs, or distant genes [13]). The size and complexity of the gene (∼6.5 Mb, with at least 55 exons, and an unknown repertoire of transcript and protein isoforms) makes this a daunting task – especially since gene regulation is often different between tissues (as well as cell types and developmental stages), mandating the use of post mortem brain tissue as part of this approach [22]. In the first such study, in dorsolateral prefrontal cortex of over 250 individuals, Bigos et al. [23] showed that genotype affects *CACNA1C* mRNA abundance with risk
homozygotes having highest expression. Results in induced neurons derived from fibroblasts show the same profile (see below). However, Gershon et al. [16] found the opposite result in cerebellum, and no genotype effect in parietal cortex, in separate brain series; these authors examined a larger range of CACNA1C SNPs, but had considerably smaller sample sizes, than [23]. Roussos et al. [15] also provide some evidence that the risk allele is associated with reduced CACNA1C expression in brain tissue, and in cell lines, and reveal a more complex relationship between CACNA1C gene structure and regulation beyond that conferred by rs1006737. Further studies are clearly required, taking into account the possibility of temporal and spatial specificity of effects, and genetic influences on splicing of the gene — to date only the overall CACNA1C transcript abundance has been assayed.

ANK3 encodes ankyrin G, a scaffold protein involved in many cellular processes [24]. Like CACNA1C, ANK3 is a large and complex gene (700 kb, with at least 44 exons and multiple protein isoforms). With regard to BD, perhaps the best known feature of ANK3 is that it is located in the axon initial segment and at nodes of Ranvier, wherein it couples voltage-gated sodium channels to the cytoskeleton [25]. However, a recent study shows that ANK3 is dispensable for this function [26], whilst Chang et al. [27] show that ANK3 is expressed by oligodendrocytes, with ankyrin G abundant on the glial rather than axonal side of the nodes. These unexpected findings are complemented by reports showing that ankyrin G may have important additional localisations and functions. For example, Durak et al. [28] show that ANK3 regulates the β-catenin/Wnt signalling pathway, already implicated in BD. Smith et al. [29] identified a novel short ANK3 isoform which is located in dendritic spines and regulates NMDA receptor-dependent plasticity. Interestingly, ANK3 becomes enriched in this sub-synaptic fraction after chronic lithium treatment [30]. In BD, ANK3 mRNA is increased in blood [31], but its expression has not been reported in brain in the disorder. The likelihood of isoform-selective functions of ANK3 is complemented by preliminary evidence for differential expression of ANK3 isoforms in human brain and their regulation by BD associated SNPs [32,33]. The latter study [33] suggests that the risk allele at rs9804190 is associated with decreased ANK3 mRNA, but other ANK3 SNPs did not show significant effects on expression.

ZNF804A (zinc finger protein 804A) was originally identified as a schizophrenia risk gene but then found to be associated with a broader psychosis phenotype including BD. It is thought to encode a transcription factor, and regulates expression of various genes in vitro [34]. The best-established psychosis risk allele (rs1344706), located in the third intron, influences ZNF804A mRNA in foetal but not adult human brain [35], with the risk allele being associated with lower expression. Tao et al. [36**] replicated this observation in prefrontal cortex of a separate, large sample (n = 697) and showed that genotype impacts not on the full-length transcript, but on a newly identified, shorter ZNF804A mRNA isoform. The effect was again limited to foetal brain, although they also found a genotype-by-diagnosis interaction in their BD sample, with lower expression in patients homozygous for the risk allele. Complicating interpretation of the data, translation of the protein encoded by the short ZNF804A transcript has not been unambiguously demonstrated in human brain; moreover, its predicted sequence lacks the zinc finger domain of ZNF804A, and hence nothing is known about its potential function nor how it may be involved in BD pathogenesis.

In summary, these examples illustrate the depth and breadth of research which will be needed to move from statistical association of BD with a genomic locus to (a) identifying the affected gene(s), (b) clarifying its role in the pathogenesis of BD, and (c) determining the pathophysiological consequences of the risk SNP or haplotype.

Animal models of bipolar disorder

All animal models of psychiatric disorders have inherent limitations, but BD has suffered particularly from the lack of a model which recapitulates its core feature, viz. spontaneous oscillations between manic-, euthymic-, and depressive-like phenotypes. However, there are several genetic mouse models which show manic-like behaviour and which have been used to explore its molecular and neural basis.

SHANK3 point mutations and deletions are linked with autism, but not with BD. However, Shank3-overexpressing mice show manic-like behaviour, altered circadian rhythms, and a synaptic excitation/inhibition imbalance, via a pathway involving the Arp2/3 pathway and increases in F-actin [37]. Their behavioural phenotype could be normalised by one commonly used mood stabiliser, sodium valproate, but not by lithium.

The Clock Δ19 mouse has a mutation in the Clock gene, a core regulator of the circadian system. The mouse was noted several years ago to have a diurnal manic-like behaviour, being hyperactive and with more reward-related and reduced depressive and anxiety behaviours during the light phase, but normal behaviour at night [38]. This manic-like daytime profile coincides with increased firing of dopamine neurons in the ventral tegmental area. Sidor et al. [39], using a novel chronic optogenetic stimulation paradigm, provide convincing evidence that enhanced dopaminergic activity is causally related to the behaviour. They also showed that the mechanism likely involves Clock acting as a transcriptional repressor of tyrosine hydroxylase, a gene involved in dopamine synthesis, suggesting that the Clock Δ19 mutant
may be less efficient in this respect. Interestingly, another mouse model has recently been reported which also involves a circadian gene (the nuclear receptor REV-ERBa) regulating dopamine regulation via tyrosine hydroxylase repression, and with similar mood-related behaviour [40*]. These two studies together argue for a renewed focus on circadian biology, and dopamine, in the core pathophysiology — and perhaps treatment — of BD and related psychiatric disorders.

The Shank3 transgenic, Clock Δ19 and REV-ERBa mice illustrate how genetic models can be used to identify synaptic, cellular and synaptic mechanisms underlying BD-relevant phenotypes. Animal models are also being used to investigate the function of BD-associated genes. For example, Leuiss et al. [41**] studied AnK3 using heterozygous Ank3+/− mice, and RNA interference to knock down Ank3 in the dentate gyrus. Both manipulations led to decreased anxiety and increased motivation for reward, which were ameliorated by chronic lithium administration. Moreover the Ank3+/− mice were more susceptible to depression-like behaviours after chronic stress, and had higher corticosterone levels, suggestive of enhanced stress reactivity.

Modelling bipolar disorder in reprogrammed cells
Notwithstanding these developments, the lack of established animal models of bipolar disorder has encouraged the early adoption of induced pluripotent stem cell (iPSC) research into the field.

Chen et al. [42] generated iPSCs from fibroblasts of three BD patients and three controls, and differentiated them into cells with a forebrain neuronal phenotype. They reported alterations in the transcriptome of the BD-derived neurons (but not the iPSCs), including up-regulation of some receptor and ion channel transcripts and those specifying a ventral rather than dorsal telencephalic fate.

Wang et al. [43] took a different technical approach, directly trans-differentiating fibroblasts to neuron-like cells (induced neurons), from 12 BD patients (6 lithium responders, 6 non-responders) and 6 healthy controls. Using a label-free optical imaging method, they measured parameters related to cell growth, size, and adhesion. The main finding was that lithium responsive patients’ neurons showed greater adhesion compared to those of lithium non-responders. Cells from control subjects were intermediate between these two groups, and the BD group overall did not show differences from the controls. The study highlights that research using iPSCs, and derived neuronal-related cells may have value for understanding drug actions as well as disease pathophysiology.

Madison et al. [44] used a family-based paradigm, generating iPSC lines and then neural progenitor cells from two brothers with BD and their two unaffected parents. No differences were seen for the iPSCs between affected and unaffected members, but the neural progenitors showed several differences, including alterations in neurogenesis and in the expression of neuroplasticity-related genes including Wnt and ion channels.

Another experimental design is to compare cells derived from subjects on the basis of genotype, rather than by diagnosis, to examine genetic mechanisms. Yoshimizu et al. [45**] took fibroblasts to make induced neurons from 24 subjects genotyped for rs1006737, the risk-associated SNP in CACNA1C, and examined the expression and function of calcium channels. They found elevated CACNA1C mRNA, and greater calcium current density, in the neurons derived from risk homozygotes compared to heterozygotes and non-risk homozygotes. These data support the view that the risk variants of CACNA1C may involve a gain rather than a loss of function, and strengthen the rationale for L-type calcium channel antagonists as potential therapeutic agents in BD.

In summary, studies using reprogrammed cells are providing intriguing clues about the pathophysiology of neuronal-like cells derived from patients with BD, or related to its genetic risk. However, no clear pattern or convergence of findings has yet emerged, and much remains to be done in terms of detailed phenotyping, and particularly in the scaling up of experiments to provide robust and generalisable results.

Conclusions
Recent findings have made inroads into revealing the genetic and molecular basis of BD, and have provided support for the long-suspected (but still poorly characterised) role of calcium signalling in the disorder, as well as implicating pathways and processes which were less anticipated. There is a long road ahead in terms of identifying the pathophysiological implications of the risk genes, and understanding how their functioning is impacted by the risk variants. At present, the effects appear to be mediated through altered gene expression and thence function, with some evidence that specific isoforms in each gene may be important. The challenges and opportunities ahead are both empirical and bioinformatic, and both will expand substantially with the forthcoming GWAS from the Psychiatric Genomics Consortium involving 20,000 BD cases and 28,000 controls [7] which will no doubt reveal many more loci. In tandem with the ongoing efforts to understand the molecular mechanism of action of lithium and other mood stabilisers [46-48], there are grounds for cautious optimism that transformative advances in therapies for BD will soon be possible. These are long overdue.

Conflicts of interest
No relevant conflicts of interest.
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Neurobiology of disease

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