Vitamin A in resistance to and recovery from infection: relevance to SARS-CoV2

C. B. Stephensen1* and G. Lietz2*

1Immunity and Disease Prevention Research Unit, USDA Western Human Nutrition Research Center, and Nutrition Department, University of California, Davis, CA, USA
2Human Nutrition Research Centre, Population Health Sciences Institute, Newcastle University, Newcastle upon Tyne NE2 4HH, UK

(Submitted 12 September 2020 – Final revision received 30 December 2020 – Accepted 13 January 2021 – First published online 20 January 2021)

Abstract

SARS-CoV2 infects respiratory epithelial cells via its cellular receptor angiotensin-converting enzyme 2, causing a viral pneumonia with pronounced inflammation resulting in significant damage to the lungs and other organ systems, including the kidneys, though symptoms and disease severity are quite variable depending on the intensity of exposure and presence of underlying conditions that may affect the immune response. The resulting disease, coronavirus disease 2019 (COVID-19), can cause multi-organ system dysfunction in patients requiring hospitalisation and intensive care treatment. Serious infections like COVID-19 often negatively affect nutritional status, and the resulting nutritional deficiencies may increase disease severity and impair recovery. One example is the viral infection measles, where associated vitamin A (VA) deficiency increases disease severity and appropriately timed supplementation during recovery reduces mortality and hastens recovery. VA may play a similar role in COVID-19. First, VA is important in maintaining innate and adaptive immunity to promote clearance of a primary infection as well as minimise risks from secondary infections. Second, VA plays a unique role in the respiratory tract, minimising damaging inflammation, supporting repair of respiratory epithelium and preventing fibrosis. Third, VA deficiency may develop during COVID-19 due to specific effects on lung and liver stores caused by inflammation and impaired kidney function, suggesting that supplements may be needed to restore adequate status. Fourth, VA supplementation may counteract adverse effects of SARS-CoV2 on the angiotensin system as well as minimises adverse effects of some COVID-19 therapies. Evaluating interactions of SARS-CoV2 infection with VA metabolism may thus provide improved COVID-19 therapy.

Key words: SARS-CoV2: COVID-19: Vitamin A: Retinoic acid: Immunity: Angiotensin-converting enzyme 2 (ACE2): Lipofibroblasts

The Coronaviridae family of single-stranded RNA viruses includes four genera: alpha-, beta-, gamma- and delta-coronavirus[1]. Seven coronaviruses are known to infect humans. Two (229E and NL63) from the alpha genus and two (OC43 and HKU-1) from the beta genus cause mild upper respiratory tract infections and symptoms of the common cold. The remaining three human coronaviruses are also in the beta genus but cause severe lower respiratory tract infections, respiratory failure and death. These viruses are MERS-CoV, which causes the Middle East respiratory syndrome; SARS-CoV1, which causes severe acute respiratory syndrome, and SARS-CoV2, which is causing the current pandemic of coronavirus disease 2019 (COVID-19)[1,2]. SARS-CoV2 spreads readily from person-to-person due to the long incubation period of 2–14 d following infection[3] and, according to data available to the WHO, has infected 75 million people and caused 1.7 million deaths worldwide between December 2019 and December 2020[4].

SARS-CoV2 infection causes both mild (in approximately 80% of cases) and severe disease. Symptoms during mild disease include fever, dry cough, dyspnoea, sore throat, malaise, myalgia and, less frequently, gastrointestinal symptoms such as anorexia, nausea and diarrhoea. Mild cases are typically treated at home with supportive care, but no specific anti-viral therapy is currently recommended[5]. Some patients (approximately 20% of all infections) progress from mild to more severe disease, the diagnosis of which involves a positive test for SARS-CoV2, tachypnoea and hypoxaemia and specific findings of decreased arterial oxygen concentration or a chest X-ray showing pneumonia, indicating extensive lung inflammation. Some patients with severe infection (approximately 5% of all infections) develop renal failure and intravascular coagulation, require prolonged...
involving VA supplementation, as well as other appropriate
COVID-19 disease might be improved by nutritional support
recovery of normal lung function, then recovery from severe
and VA deficiency impairs host resistance to infection and
develop during this period of severe disease. If this is the case,
severe COVID-19 cases often have prolonged hospitalisation
of phagocytes, binding to melanin and blocking organic anion
transporter family member 1A2 OATP1A2(10). The anti-
inflammatory corticosteroid dexamethasone was shown to
reduce the death of SARS-CoV2-ventilated patients in the
RECOVERY study by one-third(5), but not in patients experienc-
ing a milder disease(8).

The purpose of this review is to examine how vitamin A (VA)
status might affect host resistance to, or recovery from, COVID-
19 disease. VA is of particular interest in this regard for at least
three reasons. First, VA deficiency impairs host resistance to
infection. VA was termed ‘the anti-infective’ vitamin approxi-
mately 90 years ago because of the increased risk of infec-
tions, particularly of lung infections, in VA-deficient
experimental animals(11), indicating that some aspect of host resis-
tance to, or recovery from, infection was impaired. Research in
the past 30 years has demonstrated that VA (retinol) is con-
verted to its active metabolite retinoic acid by cells of the
immune system and that retinoic acid then acts as an autocrine
or paracrine factor to regulate many aspects of immune func-
tion, as has been reviewed recently(12,13). Second, the action of
VA is crucial in normal lung development and in repair of lung
tissue subsequent to injury(14); thus, VA status may be particu-
larly important during recovery from COVID-19. Third, severe
infections are known to have a significant, negative impact on
VA status by several mechanisms, including reduced absorp-
tion(15), decreased hepatic mobilisation(16) and urinary loss of
VA due to glomerular and/or tubular dysfunction(17). Since
severe COVID-19 cases often have prolonged hospitalisation
with renal involvement, it is plausible that VA deficiency could
develop during this period of severe disease. If this is the case,
and VA deficiency impairs host resistance to infection and
recovery of normal lung function, then recovery from severe
COVID-19 disease might be improved by nutritional support
involving VA supplementation, as well as other appropriate
nutritional support. Fourth, medication used to reduce the
impact of severe COVID-19 infection could alter the metabo-
ism of VA in affected tissues and therefore induce tissue-spe-
cific VA deficiency(18), which could reduce the ability to repair
tissues(19).

SARS-CoV2 pathogenesis and effects of vitamin A on the
immune system

SARS-CoV2 infection of the respiratory epithelium

Similarly to SARS-CoV1, the cellular infection mechanism of
SARS-CoV2 is mainly mediated by the cell-surface receptor
angiotensin-converting enzyme 2 (ACE2)(19). SARS-CoV2 uses
ACE2 for receptor-mediated cell entry and serine protease trans-
membrane serine protease 2 (TMPRSS2) for S protein priming(20).
Cell entry of SARS-CoV2 can be partially blocked by the clinically
proven serine protease inhibitor camostat mesylate, which is
active against serine protease transmembrane serine protease
2 (TMPRSS2)(20). Although very low levels of ACE2 protein have
been observed in the normal respiratory system(21), the presence
of ACE2 protein levels and viral RNA has been confirmed in cli-
culated epithelial cells, indicating that low-level protein expression
in upper airway epithelial cells facilitates infection of ciliated
cells, followed by a rapid interferon-induced increase of ACE2
expression in lower airway ciliated and type-2 alveolar epithelial
cells, potentially allowing SARS-CoV2 to spread across the respi-
atory mucosa(22). The cell-surface ACE2 receptor internalises on
binding to the SARS-CoV spike protein, leading to ACE2 receptor
down-regulation(23,24). The SARS-CoV spike protein-mediated
ACE2 down-regulation could contribute to the severity of lung
pathologies such as pulmonary fibrosis and acute respiratory dis-
tress syndrome, since the counterbalance of ACE2 on angioten-
sin II production in the renin–angiotensin system is deregulated,
affecting its ability to modulate the innate immune system and to
regulate inflammation(25,26). ACE2 deficiency is associated with up-regulation of putative mediators of atherogenesis, such as cytokines and adhesion mol-
ecules, and reduced ACE expression is observed in established
atherosclerotic plaques(27). Finally, the observed difference in
SARS-CoV2 severity between children and adults could in part
be attributed to the lower expression of the SARS-CoV2-targeted
receptor ACE2 and serine protease serine protease transmem-
brane serine protease 2 (TMPRSS2) in nasal and bronchial air-
ways in children compared with adults(20).

Vitamin A and epithelial barriers

VA is needed to maintain healthy respiratory and intestinal
epithelial barriers(17). VA deficiency may not produce obvious
changes in healthy epithelial surfaces, but following viral
infection in the intestine, for example, pathological changes
to infected epithelial surfaces were found to be much greater
in VA-deficient than in control mice(29). In the respiratory tract,
VA deficiency also increases epithelial damage and impairs
recovery, sometimes leading to squamous metaplasia in
alveoli and airways, following noxious exposures such as
Vitamin A and COVID-19 1665

Effects of vitamin A on innate immunity

Many cells of the innate immune system, including innate lymphoid cells, macrophages and granulocytes, are affected by VA. Type 1 innate lymphoid cells, including natural killer (NK) cells, play a role in responses to viral infection and their activity and number in peripheral blood is decreased by VA deficiency(54), while retinoic acid treatment can also dampen activity(55). Neutrophil and macrophage function is also directly affected by VA. Phagocytic and bacterial-killing activity of these cells is impaired by VA deficiency, leading to decreased resistance to some infections, though production of the CD4+ T-helper type 1 (Th1)-promoting cytokine IL-12 can be increased, causing increased type 1 inflammation(57). Ex vivo and animal studies show that treatment of macrophages with retinoic acid can have anti-inflammatory effects, dampening production of IL-12 and pro-inflammatory cytokines such as TNF-α, as well as increasing production of the regulatory cytokine IL-10(55). Other effects on macrophage phenotype are also caused by retinoic acid. For example, in Leishmania infection of mice, retinoic acid treatment diminishes the development of M1 macrophages, a component of protective type 1 immunity in this model system, and increases the development of M2 macrophages and type 2 immunity, which is not protective. Thus, retinoic acid intervention in this system decreases resistance to infection(50). Conversely, in Schistosoma mansoni infection of mice, VA deficiency exacerbates disease due to excessive inflammation and tissue damage, while retinoic acid treatment increases conversion of macrophages into a tissue-resident phenotype that dampens inflammation and allows control of infection and enhancement of tissue repair(57). While many of the effects of retinoic acid on innate immune cells can be seen as anti-inflammatory, a lesson from these and other studies is that intervention with retinoic acid during infections can have unpredictable effects on the course of a particular infection depending on the specifics of the immune response that are responsible for pathogen clearance. This topic has been discussed in a recent review(58).

Adaptive immune response to SARS-CoV2 infection

Dendritic cells link the initial anti-viral response of the innate immune system to development of adaptive immunity, involving
B and T lymphocytes, that will help clear infections that cannot be resolved by innate immunity alone. Adaptive immunity also has a memory component that, in the case of most acute viral infections, provides a rapid and robust response upon subsequent exposure to the same virus, typically providing protection against symptomatic infection. Such responses develop for other coronaviruses \(^{59}\), and hopefully, this will also be true of SARS-CoV2.

To initiate adaptive immunity, dendritic cells at the site of infection are also activated by pattern-recognition receptor-mediated recognition of viruses, as well as by local cytokine production, and deliver viral antigen from the site of infection to a regional lymph node to initiate development of virus-specific effector and memory T cells, as well as memory B cells that results in development of antibody-producing plasma cells. Immunologists recognise three basic types of protective immunity against infectious diseases, types 1, 2 and 3 \(^{59}\). Type 1 immunity is typically induced in response to viruses and other intracellular pathogens and involves development of Th1 cells producing the signature cytokine IFN-γ and CD8 \(^+\) cytotoxic T-lymphocyte which can directly kill virus-infected cells, and a robust serum IgG antibody response, often targeting the epitope of the viral glycoprotein involved in binding to its cellular receptor (the spike [S] protein in the case of SARS-CoV2), thus neutralising the ability of the virus to infect cells. Macrophages activated by Th1 cells at sites of infection are also a key component of type 1 immunity. Type 2 immunity provides protection against intestinal parasites, involves development of CD4 \(^+\) Th2 cells producing IL-4, -5 and -13, induction of an antibody response involving IgE and activation of eosinophils and basophils at mucosal surfaces, near the sites of pathogen exposure. Type 3 immunity targets extracellular pathogens, involves development of CD4 \(^+\) Th17 cells producing IL-17 and IL-22, development of appropriate antibody responses and engagement of mucosal epithelial cells and neutrophils to provide barrier protection. CD4 \(^+\) follicular-helper T cells develop during all of these responses to provide help for development of antigen-specific memory B cells and a robust antibody response. CD4 \(^+\) regulatory (Treg) cells also develop during most adaptive responses to provide a pathogen-specific cell type capable of dampening pro-inflammatory immunity by direct cell-to-cell interaction or production of the cytokines IL-10 and TGF-β.

Research on the adaptive immune response to SARS-CoV2 is in an early state, but two recent reviews indicate that patients recovering from COVID-19 have the Th1, CD8 \(^+\) cytotoxic T-lymphocyte and neutralising antibody responses expected of a typical type 1 response against a viral infection \(^{52,60}\). Briefly, Th1 responses predominate over Th2 and Th17, as expected, and viral epitopes from all major structural proteins, including spike (S), matrix (M) and nucleocapsid (N), are seen in recovering patients. CD8 T-cell responses are also seen against the same proteins. The magnitude of the response is somewhat greater in patients with more severe disease, which could be due to greater antigenic stimulation. Similar patterns of response were seen against SARS-CoV1 in recovered SARS patients \(^{63}\), and protection against death by CD8 cells was demonstrated in a mouse model \(^{62}\). Similar CD4 and CD8 responses were seen against MERS-CoV in MERS survivors 0-5-2 years after infection \(^{63}\). These findings suggest that patients recovering from SARS-CoV2 infection may have protective T-cell memory responses. In contrast to the potential benefit of such Th1 responses, some evidence from SARS patients \(^{60}\) as well as preliminary work from COVID-19 patients cited in a recent review \(^{60}\) suggests that Th2 responses may be associated with poor outcomes to infection. B-cell responses develop in response to SARS-CoV2, as demonstrated by the serum IgM and IgG antibody responses that are seen within 2 weeks of infection, as is also seen with MERS-CoV and SARS-CoV1 \(^{65}\).

**Antibodies** are directed primarily against the S and N proteins, and the ACE2-binding site is quite immunogenic, resulting in robust neutralising antibody responses \(^{52}\). 

### Effects of vitamin A on adaptive immunity

As discussed above, dendritic cells are a key bridge from innate to adaptive immunity during infection and immunisation. Dendritic cells provide the signals necessary to stimulate development of pathogen-appropriate adaptive immune responses. These signals include peptide antigen to provide pathogen-specificity (i.e. recognition of a specific strain of the flu virus by memory T cells), co-stimulation to insure continued proliferation of developing T cells and specific cytokines to steer CD4 T-cell development towards the pathogen-appropriate Th1, Th2, Th17, Treg and CD4 \(^+\) follicular-helper T phenotypes. In addition, a fourth signal that is often provided by dendritic cells, particularly in the intestinal and other mucosal-associated immune tissues, is retinoic acid \(^{66}\), the production of which is diminished during VA deficiency. Retinoic acid affects several aspects of T-cell development, including modification of T-cell phenotype development and mucosal homing of lymphocytes. These effects have recently been reviewed \(^{12}\). They are pleiotropic and depend on modifying factors (e.g. the local cytokine environment), but several main points are clear.

Perhaps, the most consistent and biologically important effect of retinoic acid produced by dendritic cells, and a principal reason for retinoic acid production being constitutive in mucosal dendritic cells, is that retinoic acid induces transcription of two mucosal-homing molecules on lymphocytes, chemokine receptor 9 and α4β7 integrin. Thus, memory T cells, B cells and antibody-producing plasma cells which develop in the intestine or respiratory tract (both are initiating sites of the common mucosal immune response) will preferentially recirculate to these sites, whereas T cells developing from systemic lymph nodes are less likely to do so \(^{60}\). Mucosal infections often trigger secretory IgA responses, and the plasma cells responsible for secreting IgA are found at submucosal sites due to the imprinting of chemokine receptor 9 and α4β7 integrin expression. This mechanism may explain why VA deficiency sharply reduces the secretory IgA response to influenza A virus infection in the nasal mucosa of mice \(^{67}\). Furthermore, impaired IgA antibody and infrequent virus-specific CD8 \(^+\) T cell response following respiratory viral infection induced a cytokine storm in the upper respiratory tract of VA-deficient mice \(^{69}\). Retinoic acid also enhances intestinal homing of other cell types, including Treg cells \(^{69}\).
Second, VA deficiency has contrasting effects on Th1/Th2 development, with deficiency not affecting or sometimes enhancing Th1 development (via increased IL-12 production, as discussed above) but dampening Th2 development, particularly during the early stages of phenotype commitment immediately after antigenic stimulation. In contrast, retinoic acid, or high-level dietary VA, can enhance type 2 immunity(70). However, retinoic acid, at a somewhat later stage in development of Th1 memory cells, can help stabilise phenotype commitment to the Th1 lineage(73). Similarly, effects of VA on CD8+ cytotoxic T-lymphocyte development, a key component of type 1, anti-viral immunity are mixed(12). Using model viral infections to explore the effects of VA deficiency has shown no impairment of clearance of acute influenza A virus infection in mice, a slight enhancement of some aspects of the type 1 response squamous metaplasia during recovery(31). Using chronic lymphocytic choriomeningitis virus infection in mice also showed an elevated type 1 response, more severe pathological changes, higher virus load and increased mortality, effects which were reversed with retinoic acid(72).

Third, VA regulates the balance between Th17 and Treg development. Both cell types often develop in the intestinal lymphoid tissue (as well as at other sites) with Treg cells being predominant when inflammation is minimal and constitutive TGF-β and retinoic acid production bias memory T-cell development towards a Treg phenotype which is appropriated for antigens derived from food and commensal bacteria. However, when inflammation develops, IL-6 is produced and this shifts the balance towards Th17. During VA deficiency, Treg development is diminished because retinoic acid production is reduced(55).

Fourth, retinoic acid affects B cell development, and VA deficiency decreases the antibody response to T-cell-dependent antigens, in particular(12). While VA deficiency in mice slightly enhances type 1-related IgG2a antibody responses, the IgG1 response is diminished and, in particular, the IgA response is specifically diminished because retinoic acid can enhance class-switching to IgA(73,74).

**Contribution of the immune system to COVID-19 pathogenesis**

Approximately 5% of COVID-19 patients, many with underlying conditions such as CVD, diabetes mellitus and obesity, develop severe disease with a high risk of death. Male sex and age above 65 years are also risk factors(69). The pathogenesis of severe disease involves excessive activation of the immune system that can result in immunopathology that increases disease severity, as has been reviewed recently(92,93,77). During the initial innate immune response, a robust IFN type I/III response appears to predict a milder course of disease, presumably by controlling initial virus replication and allowing development of the adaptive response. However, patients with severe disease are more likely to have a lower initial IFN type I/II response (though it may be elevated later) which may result in a failure to control initial viral replication. Prolonged viral replication then appears to result in high-level production of pro-inflammatory cytokines (including TNF-α, IL-1, IL-6 and many others), likely produced by activated macrophages but perhaps by other innate cells such as neutrophils that may be attracted to the lungs. This local hyper-inflammation may dampen development of adaptive immunity and in patients with severe disease lymphopenia, including low levels of T cells, is pronounced. During this ‘cytokine storm’ with activation of innate immune cells, a systemic response develops that includes elevation of acute phase proteins, particularly CRP, and activation of the complement system which may result in vascular leakage as well as diffuse intravascular coagulation which can cause organ damage thus increasing disease severity. Developing therapeutic interventions to dampen this immune-mediated pathology is a high priority of current research. Potential drug targets include the NF-κB pathway to attenuate TNF-α and IL-6 expression, the JAK/STAT signalling pathway and the sphingosine-1-phosphate receptor 1 pathway(78). The JAK/STAT and sphingosine-1-phosphate receptor 1 signalling pathways are of particular interest since SARS-CoV2 potential down-regulation of ACE2 expression could result in over-production of angiotensin II via the related ACE enzyme, leading to enhanced IL-6 production. JAK/STAT inhibitors such as Baricitinib or sphingosine-1-phosphate receptor 1 receptor agonists such as Fingolimod could attenuate the cytokine storm(78).

**Possible implications of vitamin A deficiency for immune response to SARS-CoV2**

The effects of VA in the immune system discussed above have several possible implications for individuals with VA deficiency infected by SARS-CoV2. First, VA deficiency is not likely to impair a type 1 anti-viral response to SARS-CoV2, but could increase the severity of type 1 inflammation and tissue damage in the lungs following viral infection. Second, after clearance of SARS-CoV2 infection, VA deficiency could impair repair of damaged alveolar pneumocytes and airway epithelium. Third, protective immunity to SARS-CoV2 could be impaired by VA deficiency, particularly the mucosal IgA response, which could be important in resistance to reinfection, though development of memory Th1 and CD8+ cytotoxic T-lymphocyte response might also be affected.

**Effect of infections on vitamin A metabolism and status**

VA is absorbed in the intestine with high efficiency in healthy individuals (>80%) and mainly stored in the liver (>90%), with smaller amounts stored in intestine, kidney, adipose tissue and the lung(79). Release of serum VA bound to retinol-binding protein (RBP4) is homoeostatically controlled and does not change over a wide range of liver reserves(79). During infection, the acute protein response increases the synthesis of hepatic inflammatory cytokines, but at the same time reduces the release of negative acute phase proteins such as RBP4. The subsequent reduction of holo-RBP4 (retinol bound to apo-RBP4) causes a decline in circulating VA even before the acute-phase proteins CRP and AGP have reached peak concentrations(80). Reduced serum retinol and RBP4 concentrations could potentially overestimate the prevalence of VA deficiency in populations with high levels of inflammation(80). Severe infections have also shown to reduce food intake(81), impair absorption by 20–30%(82), increase
metabolic requirements or catabolic losses\(^{(80)}\) and result in substantial urinary losses\(^{(17,82)}\). Importantly, respiratory infections combined with low dietary VA intake could deplete liver VA stores in COVID-19 patients suffering from pneumonia to levels associated with VA deficiency, since urinary losses of >1000 μg retinol/d have been observed in 24 % of ICU patients with pneumonia and sepsis, representing a higher amount than the RDA for VA\(^{(82)}\). We estimate that this level of urinary loss, combined with a postulated decrease in intake of approximately 50 % of the RDA for women hospitalised with COVID-19, could lead to a loss of approximately 1350 μg/d from liver stores. Such losses would lead to deficient liver stores (<20 μg/g) in approximately 3 weeks for a 61 kg woman with relatively low pre-existing liver stores (40 μg/g; estimated liver weight of 2-4 % of body weight). Using data on estimated liver stores among US women of 129 (so 89) μg/g who consumed high levels of VA (a mean intake of 175 (so 111) % of the RDA of 700 μg retinol\(^{(83)}\)), we estimate that approximately 16 % of women (those at least one so below the mean) in a population with similarly high intake would have reserves of 40 μg/g or less.

In populations consuming lower levels of VA, the percentage of individuals at risk for systemic VA deficiency would be higher. It is important to note however that these systemic effects of VA are different to tissue-specific VA deficiency that could develop following COVID-19 infection (Fig. 1). First, pulmonary fibrosis induced by SARS-CoV2 is characterised by the conversion of lipofibroblasts to activated myofibroblasts that excessively deposit extracellular matrix proteins and lose their retinoid stores\(^{(45,48)}\).

Second, inflammation induced reduction of retinoic acid uptake and metabolism to polar metabolites\(^{(80)}\) could affect the ability of the lung to regenerate from pulmonary fibrosis, since lipofibroblasts rely on retinoids to initiate, coordinate and regulate alveolar septal eruption and alveolo-genesis\(^{(45)}\). Third, the ability to deliver retinoids to the lung is compromised during severe infection due to the decline in circulating VA\(^{(80)}\). Nutritionists have long recognised that acute infection exacerbates the risk of malnutrition, as there is potential to fall into a ‘vicious circle’ of acute illness, deficiency and susceptibility to secondary infection.

However, the timing of VA supplementation during SARS-CoV2 infection and the recovery period may be critical, as early intervention during the infectious period may be contraindicated due to its impact on the ACE2 receptor\(^{(37,86)}\), but intervention during the recovery period may be critical to address the potentially developing tissue-specific VA deficiency and its effects on the respiratory immune response. Importantly, VA supplementation may not be beneficial depending on age and co-morbidity, since VA supplementation can increase the risk of acute respiratory tract infections in children with adequate VA status\(^{(84)}\) and can delay recovery from community-acquired pneumonia in children\(^{(45)}\) and respiratory syncytial virus infection in infants\(^{(86)}\). However, VA supplementation was associated with improved recovery from Ebola virus infection in adults\(^{(87)}\) and can have beneficial effects in obesity by correcting tissue VA deficiency, altering lung immune cell composition, improving vaccine responses and assisting in respiratory virus clearance\(^{(88)}\). In particular, high-dose VA supplementation during recovery from measles has been shown to decrease mortality and improve recovery, including decreasing the severity of metabolic requirements or catabolic losses\(^{(80)}\) and result in substantial urinary losses\(^{(17,82)}\). Importantly, respiratory infections combined with low dietary VA intake could deplete liver VA stores in COVID-19 patients suffering from pneumonia to levels associated with VA deficiency, since urinary losses of >1000 μg retinol/d have been observed in 24 % of ICU patients with pneumonia and sepsis, representing a higher amount than the RDA for VA\(^{(82)}\). We estimate that this level of urinary loss, combined with a postulated decrease in intake of approximately 50 % of the RDA for women hospitalised with COVID-19, could lead to a loss of approximately 1350 μg/d from liver stores. Such losses would lead to deficient liver stores (<20 μg/g) in approximately 3 weeks for a 61 kg woman with relatively low pre-existing liver stores (40 μg/g; estimated liver weight of 2-4 % of body weight). Using data on estimated liver stores among US women of 129 (so 89) μg/g who consumed high levels of VA (a mean intake of 175 (so 111) % of the RDA of 700 μg retinol\(^{(83)}\)), we estimate that approximately 16 % of women (those at least one so below the mean) in a population with similarly high intake would have reserves of 40 μg/g or less.

In populations consuming lower levels of VA, the percentage of individuals at risk for systemic VA deficiency would be higher. It is important to note however that these systemic effects of VA are different to tissue-specific VA deficiency that could develop following COVID-19 infection (Fig. 1). First, pulmonary fibrosis induced by SARS-CoV2 is characterised by the conversion of lipofibroblasts to activated myofibroblasts that excessively deposit extracellular matrix proteins and lose their retinoid stores\(^{(45,48)}\).

Second, inflammation induced reduction of retinoic acid uptake and metabolism to polar metabolites\(^{(80)}\) could affect the ability of the lung to regenerate from pulmonary fibrosis, since lipofibroblasts rely on retinoids to initiate, coordinate and regulate alveolar septal eruption and alveolo-genesis\(^{(45)}\). Third, the ability to deliver retinoids to the lung is compromised during severe infection due to the decline in circulating VA\(^{(80)}\). Nutritionists have long recognised that acute infection exacerbates the risk of malnutrition, as there is potential to fall into a ‘vicious circle’ of acute illness, deficiency and susceptibility to secondary infection.

However, the timing of VA supplementation during SARS-CoV2 infection and the recovery period may be critical, as early intervention during the infectious period may be contraindicated due to its impact on the ACE2 receptor\(^{(37,86)}\), but intervention during the recovery period may be critical to address the potentially developing tissue-specific VA deficiency and its effects on the respiratory immune response. Importantly, VA supplementation may not be beneficial depending on age and co-morbidity, since VA supplementation can increase the risk of acute respiratory tract infections in children with adequate VA status\(^{(84)}\) and can delay recovery from community-acquired pneumonia in children\(^{(45)}\) and respiratory syncytial virus infection in infants\(^{(86)}\). However, VA supplementation was associated with improved recovery from Ebola virus infection in adults\(^{(87)}\) and can have beneficial effects in obesity by correcting tissue VA deficiency, altering lung immune cell composition, improving vaccine responses and assisting in respiratory virus clearance\(^{(88)}\). In particular, high-dose VA supplementation during recovery from measles has been shown to decrease mortality and improve recovery, including decreasing the severity of metabolic requirements or catabolic losses\(^{(80)}\) and result in substantial urinary losses\(^{(17,82)}\). Importantly, respiratory infections combined with low dietary VA intake could deplete liver VA stores in COVID-19 patients suffering from pneumonia to levels associated with VA deficiency, since urinary losses of >1000 μg retinol/d have been observed in 24 % of ICU patients with pneumonia and sepsis, representing a higher amount than the RDA for VA\(^{(82)}\). We estimate that this level of urinary loss, combined with a postulated decrease in intake of approximately 50 % of the RDA for women hospitalised with COVID-19, could lead to a loss of approximately 1350 μg/d from liver stores. Such losses would lead to deficient liver stores (<20 μg/g) in approximately 3 weeks for a 61 kg woman with relatively low pre-existing liver stores (40 μg/g; estimated liver weight of 2-4 % of body weight). Using data on estimated liver stores among US women of 129 (so 89) μg/g who consumed high levels of VA (a mean intake of 175 (so 111) % of the RDA of 700 μg retinol\(^{(83)}\)), we estimate that approximately 16 % of women (those at least one so below the mean) in a population with similarly high intake would have reserves of 40 μg/g or less.

In populations consuming lower levels of VA, the percentage of individuals at risk for systemic VA deficiency would be higher. It is important to note however that these systemic effects of VA are different to tissue-specific VA deficiency that could develop following COVID-19 infection (Fig. 1). First, pulmonary fibrosis induced by SARS-CoV2 is characterised by the conversion of lipofibroblasts to activated myofibroblasts that excessively deposit extracellular matrix proteins and lose their retinoid stores\(^{(45,48)}\).

Second, inflammation induced reduction of retinoic acid uptake and metabolism to polar metabolites\(^{(80)}\) could affect the ability of the lung to regenerate from pulmonary fibrosis, since lipofibroblasts rely on retinoids to initiate, coordinate and regulate alveolar septal eruption and alveolo-genesis\(^{(45)}\). Third, the ability to deliver retinoids to the lung is compromised during severe infection due to the decline in circulating VA\(^{(80)}\). Nutritionists have long recognised that acute infection exacerbates the risk of malnutrition, as there is potential to fall into a ‘vicious circle’ of acute illness, deficiency and susceptibility to secondary infection.

However, the timing of VA supplementation during SARS-CoV2 infection and the recovery period may be critical, as early intervention during the infectious period may be contraindicated due to its impact on the ACE2 receptor\(^{(37,86)}\), but intervention during the recovery period may be critical to address the potentially developing tissue-specific VA deficiency and its effects on the respiratory immune response. Importantly, VA supplementation may not be beneficial depending on age and co-morbidity, since VA supplementation can increase the risk of acute respiratory tract infections in children with adequate VA status\(^{(84)}\) and can delay recovery from community-acquired pneumonia in children\(^{(45)}\) and respiratory syncytial virus infection in infants\(^{(86)}\). However, VA supplementation was associated with improved recovery from Ebola virus infection in adults\(^{(87)}\) and can have beneficial effects in obesity by correcting tissue VA deficiency, altering lung immune cell composition, improving vaccine responses and assisting in respiratory virus clearance\(^{(88)}\). In particular, high-dose VA supplementation during recovery from measles has been shown to decrease mortality and improve recovery, including decreasing the severity of
secondary bacterial pneumonia\(^\text{99}\). Delivering VA through inhalation may be more efficient to address the immunological lesions in VA-deficient individuals\(^\text{90,91}\) particularly since inflammation also reduces retinoic acid uptake and metabolism to polar metabolites\(^\text{80}\).

**Interaction of vitamin A with olfactory function**

SARS-CoV2 causes olfactory and/or gustatory dysfunctions in COVID-19 patients\(^\text{92-95}\). A recent meta-analysis of eight studies confirmed that patients with COVID-19 had significantly higher risks of developing olfactory and/or gustatory dysfunctions compared with normal subjects \(\text{OR of 65-9}\) or patients with acute respiratory infection without detectable virus \(\text{OR of 11-3}\)\(^\text{98}\). On average, 50 % of COVID-19 patients suffered from olfactory and/or gustatory dysfunctions\(^\text{84}\), although olfactory impairment was reported to vary between 46-8 % in China, 60 % in the USA and 85-6 % in Europe\(^\text{98}\). The mean time from olfactory loss to recovery onset was reported as 12 d, with complete olfactory recovery in only 51 % of patients\(^\text{93}\). Viral infections are a common cause of olfactory dysfunction\(^\text{94}\), although the pathogenesis of sensory loss after viral infections is not well characterised\(^\text{95,96}\). It is assumed that the nasal epithelium and olfactory bulb could be critical in the pathological development of COVID-19, with mild outcomes of COVID-19 leading to localised effects with mild effects on olfactory function. Interestingly, the effects of SARS-CoV2 on chronic olfactory impairment increase with age, affecting up to 50 % of people aged \(\geq 65\) years and >80 % of people aged \(\geq 80\) years\(^\text{92}\).

VA supplementation may play an important role in the regeneration of olfactory receptor neurons, particularly since retinoic acid signalling is involved in neuronal regeneration\(^\text{97}\). Indeed, a recent retrospective cohort analysis in patients with post-infectious olfactory dysfunction indicated that topically applied VA at a dose of 10 000 IU/d for 8 weeks improved clinical outcome\(^\text{88}\). This beneficial effect of VA to improve olfactory dysfunction is supposedly due to increased local concentration of VA at the olfactory neuroepithelium\(^\text{98}\). Thus, similarly for treating VA deficiency in the lung, topical administration of VA may increase bioavailability and reduce toxic side effects of high-dose systemic VA therapy needed to overcome localised VA deficiency\(^\text{94,96,98}\). However, since no studies have investigated the benefit of topical application of VA in COVID-19 patients suffering from olfactory dysfunction, prospective, double-blind and placebo-controlled trials are needed to confirm if VA therapy would be of benefit to those patients that struggle to recover from olfactory impairment.

**Interaction of vitamin A with current SARS-CoV2 medication**

The RECOVERY trial results demonstrated that low-dose dexamethasone reduced the death rate of patients that require respiratory support\(^\text{31}\). However, although corticosteroids have anti-inflammatory, antioxidant, pulmonary vasodilator and anti-edematous effects\(^\text{100}\), their impact on limiting immune responses might counteract viral clearance\(^\text{101}\).\n
Previous studies associate the use of corticosteroid in SARS, MERS and influenza with a higher risk of a delayed viral clearance and long-term complications in survivors\(^\text{102}\). Furthermore, dexamethasone decreases alveolar numbers, and this effect on alveolar architecture can be long term or permanent\(^\text{101}\). The drug also reduces retinoid-binding proteins and receptors in mice\(^\text{101}\), which potentially could affect the metabolism of VA in affected tissues. Since chronic nutritional VA deficiency can result in pulmonary fibrosis\(^\text{114}\), long-term complications in survivors after dexamethasone treatment could include lung tissue scarring due to medication-induced VA deficiency. On the other hand, supplementation with a nutrient-metabolite combination containing VA and retinoic acid in a 10:1 molar ratio (VARA) was effective in increasing the concentration of VA to a level that was not antagonised by dexamethasone, probably since the VARA-induced lung tissue VA accumulation compensated the VA loss caused by dexamethasone\(^\text{103}\). Importantly, the delivery of retinoids to the lung tissue determines whether or not the negative side effects of dexamethasone on lung VA levels can be overcome. Treatment with retinoic acid was shown to reverse pulmonary emphysema\(^\text{104}\), but caused significant toxic side effects\(^\text{105}\). To obtain biologically effective levels in the lung, inhalation of retinoids rather than oral administration may be a better approach\(^\text{91}\), particularly since this approach induces cellular retinol-binding protein 1 protein expression which was shown to be reduced following dexamethasone treatment\(^\text{106}\). However, effectiveness of either VARA or retinoic acid inhalation to optimise lung VA stores would need to be tested in COVID-19-affected individuals who have been treated with dexamethasone.

**Summary**

Traditionally, VA plays a very important role in the response of the immune system to infection, and this role may be particularly important in the development of a protective immune response after a SARS-CoV2 infection has occurred. This stresses the need for an adequate VA status prior to infection in order to support the immune system in its ability to deal with the inflammatory response due to SARS-CoV2 via a range of mechanisms that have been summarised in this review. However, in spite of an adequate \textit{a priori} VA reserve, SARS-CoV2 infection itself may diminish VA stores. Such an infection-induced VA deficiency may impair the ability of the lung to repair damaged epithelial surfaces, potentially leading to long-lasting scarring, lung fibrosis and reduced pulmonary capacity. Thus, VA supplementation during recovery may be needed even though VA supplementation early in infection may not have been indicated due to a lack of VA deficiency prior to infection. In addition, earlier VA supplementation may have unpredictable effects for the following reasons: First, it is plausible that VA supplementation at the time of infection early in infection may not have been indicated due to a lack of VA deficiency prior to infection. In addition, earlier VA supplementation may have unpredictable effects for the following reasons:

- First, it is plausible that VA supplementation at the time of infection early in infection may not have been indicated due to a lack of VA deficiency prior to infection.
- Second, however, activation of ACE2 in infected patients could reduce the risk of sympathetic
over-activation seen in severe SARS-CoV2 infection, and in particular in obese and diabetic COVID-19 patients. Topical application of VA may be beneficial to patients suffering from post-infectious olfactory dysfunction, since VA supplementation may play an important role in the regeneration of olfactory receptor neurons. Finally, there could be a beneficial use of VA as a co-medication in severe SARS-CoV2-affected patients who are treated with corticosteroids, particularly since dexamethasone was shown to negatively affect VA metabolism in lung tissues. It is plausible to assume that VA deficiency in at risk population groups would increase the risk of severe COVID-19 infection, but data to confirm this are currently missing. Importantly, COVID-19 has doubled the number of people in lower- and middle-income countries facing acute food insecurity and is predicted to reduce the coverage of VA supplementation programmes to VAD-affected populations between 15 and 50% (10). Since the risk of SARS-CoV2 infection is low in under 5-year-olds, and the benefit of VA supplementation outweighs the risk of developing a severe COVID-19 infection due to increased VA intake, VA supplementation programmes should continue for this age group.

Acknowledgements
We acknowledge the help of Kieran Finney and Chloe Elliott for the literature review and Catherine Meplan for her help in designing the figure.

Funding for C. B. S. was provided by USDA-ARS project number: 2032-51500-026-00-D.

The manuscript was written by C. B. S. and G. L.

Both authors have no conflicts of interest to declare.

References
1. Fung TS & Liu DX (2019) Human coronavirus-host-pathogen interaction. *Annu Rev Microbiol* **73**, 529–557.
2. Chen Y, Liu Q & Guo D (2020) Emerging coronaviruses: genome structure, replication, and pathogenesis. *J Med Virol* **92**, 418–423.
3. Florindo HF, Kleiner R, Vaskovich-Koubi D, et al. (2020) Immune-mediated approaches against COVID-19. *Nat Nanotechnol* **15**, 630–645.
4. WHO Coronavirus Disease (COVID-19) dashboard. https://covid19.who.int/ (accessed December 2020).
5. Gandhi RT, Lynch JB & Del Rio C (2020) Mild or moderate Covid-19. *N Engl J Med* **383**, 1757–1766.
6. Berlin DA, Gulick RM & Martinez FJ (2020) Severe Covid-19. *N Engl J Med* **383**, 2451–2460.
7. Yadav R, Aggarwal S & Singh A (2020) SARS-CoV-2 host dynamics: increased risk of adverse outcomes of COVID-19 in obesity. *Diabetes Metab Syndr* **14**, 1355–1360.
8. Johnson RM & Vinetz JM (2020) Dexamethasone in the management of Covid-19. *Bmj* **370**, m2648.
9. Hu TY, Lee JZ & Asrivatham SJ (2020) Cardiovascular considerations in coronavirus disease 2019 with a special focus on arrhythmia. *J Innovations Card Rhythm Manag* **11**, 4191–4198.
10. Paniri A, Hosseini MM, Rasoulinejad A, et al. (2020) Molecular effects and retinopathy induced by hydroxychloroquine during SARS-CoV-2 therapy: role of CYP450 isoforms and epigenetic modulations. *Eur J Pharmacol* **886**, 1753–1754.
11. Green HN & Mellanby E (1928) Vitamin A as an anti-infective agent. *Br Med J* **2**, 691–696.
12. Larange A & Cheroutre H (2016) Retinoic acid and retinoid acid receptors as pleiotropic modulators of the immune system. *Annu Rev Immunol* **34**, 369–394.
13. Guo Y, Brown C, Ortiz C, et al. (2015) Leukocyte homing, fate, and function are controlled by retinoic acid. *Physiol Rev* **95**, 125–148.
14. Timonedo J, Rodríguez-Fernández L, Zaragozá R, et al. (2018) Vitamin A deficiency and the lung. *Nutrients* **10**, 1132.
15. Akdamat EK, Mulenga M, Dueker SR, et al. (2010) Accelerator mass spectrometry can be used to assess vitamin A metabolism quantitatively in boys in a community setting. *J Nutr* **140**, 1588–1594.
16. Gieng SH, Green MH, Green JB, et al. (2007) Model-based compartmental analysis indicates a reduced mobilization of hepatic vitamin A during inflammation in rats. *J Lipid Res* **48**, 904–913.
17. Stephensen CB (2001) Vitamin A, infection, and immune function. *Annu Rev Nutr* **21**, 167–192.
18. Hind M & Maden M (2004) Retinoic acid induces alveolar regeneration in the adult mouse lung. *Eur Respir J* **23**, 20–27.
19. Kuba K, Imay R, Rao SA, et al. (2005) A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. *Nat Med* **11**, 875–879.
20. Hoffmann M, Begen M, Lafon Y, et al. (2020) Influence of gle-nohumeral joint muscle insertion on moment arms using a finite element model. *Comput Methods Biomech Biomed Engin* **23**, 1117–1126.
21. Hikmet F, Meir L, Edvinsson A, et al. (2020) The protein expression profile of ACE2 in human tissues. *Mol Syst Biol* **16**, e9610.
22. Nawijn MC & Timens W (2020) Can ACE2 expression explain SARS-CoV-2 infection of the respiratory epithelia in COVID-19? *Mol Syst Biol* **16**, e9841.
23. Hamming I, Cooper ME, Haagmans BL, et al. (2007) The emerging role of ACE2 in physiology and disease. *J Pathol* **212**, 1–11.
24. Gagliardi MC, Tieri P, Ortona E, et al. (2020) ACE2 expression and sex disparity in COVID-19. *Cell Death Discovery* **6**, 37.
25. Kuba K, Imai Y & Penninger JM (2006) Angiotensin-converting enzyme 2 in lung diseases. *Curr Opin Pharmacol* **6**, 271–276.
26. Sodhi CP, Jenny N, Yamaguchi Y, et al. (2019) A dynamic variation of pulmonary ACE2 is required to modulate neutrophilic inflammation in response to pseudomonas aeruginosa lung infection in mice. *J Immunol* **203**, 3000–3012.
27. Tikellis C & Thomas MC (2012) Angiotensin-converting enzyme 2 (ACE2) is a key modulator of the renin angiotensin system in health and disease. *Int J Pept* **2012**, 256294.
28. Saheb Sharif-Askari N, Saheb Sharif-Askari F, Alabed M, et al. (2020) Airways expression of SARS-CoV-2 receptor, ACE2, and TMPRSS2 is lower in children than adults and increases with smoking and COPD. *Mol Tber Methods Clin Dev* **18**, 1–6.
29. Ahmed F, Jones DB & Jackson AA (1990) The interaction of vitamin A deficiency and rotavirus infection in the mouse. *Br J Nutr* **63**, 563–573.
30. Paquette NC, Zhang LY, Ellis WA, et al. (1996) Vitamin A deficiency enhances ozone-induced lung injury. *Am J Physiol* **270**, L475–L482.
31. Stephensen CB, Blount SR, Schoeb TR, et al. (1993) Vitamin A deficiency impairs some aspects of the host response to influenza A virus infection in BALB/c mice. *J Nutr* **123**, 823–833.
