Salidroside Protects Against Influenza A Virus-Induced Acute Lung Injury in Mice

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Abstract
Influenza A virus infections can cause acute lung injury (ALI) in humans; thus, the identification of potent antiviral agents is urgently required. Herein, the effects of salidroside on influenza A virus-induced ALI were investigated in a murine model. BALB/c mice were intranasally inoculated with H1N1 virus and treated with salidroside. The results of this study show that salidroside treatment (30 and 60 mg/kg) significantly attenuated the H1N1 virus-induced histological alterations in the lung and inhibited inflammatory cytokine production. Salidroside also decreased the wet/dry ratio, viral titers, and Toll-like receptor 4 expression in the lungs. Therefore, salidroside may represent a potential therapeutic reagent for the treatment of influenza A virus-induced ALI.

Keywords
salidroside, influenza A virus, acute lung injury, inflammatory cytokines, Toll-like receptor 4

Introduction
As one of the most important human pathogens, influenza A virus infections can result in seasonal and pandemic morbidity and mortality. Influenza A viruses infect millions of people each year, leading to severe respiratory distress and death. Severe influenza virus infection, acute respiratory distress syndrome (ARDS), bilateral pulmonary infiltration, and hypoxemia can occur clinically. Moreover, hypoxic respiratory failure is the main cause of death. Recently, human and avian influenza viruses that continue to infect humans, causing acute lung injury that results in a large number of deaths, have attracted worldwide attention and an urgent need to identify potent antiviral agents.

Rhodiola rosea is a common folk medicine, primarily used to treat symptoms of altitude sickness, altitude hypoxia, and hypoxia. The main effective component of Rhodiola rosea is salidroside (SDS) (Figure 1A), which is a phenylpropanoid glycoside. A large number of studies have shown that SDS has multiple pharmacological effects, including anti-inflammatory, anti-viral, anti-hypoxia, anti-tumor, anti-fibrosis, and anti-aging properties. Moreover, SDS has been shown to exhibit anti-viral activity against dengue virus by inhibiting viral protein synthesis and boosting host immunity. Our previous research demonstrated that the anti-inflammatory effects of SDS against lipopolysaccharide-induced acute lung injury (ALI) in mice are attributed to its ability to inhibit the Toll-like receptor 4 (TLR4)-mediated nuclear factor-kappa B (NF-kB) signaling pathway.

In this study, we examined the protective effects of SDS in a mouse model of influenza virus-induced ALI and clarified the potential anti-inflammatory mechanism. The results of this study may provide a novel method for the clinical treatment of influenza virus-induced ALI.

Materials and Methods
Animal Preparation and Treatment
Six-week-old healthy female BALB/c mice (n = 96) were maintained in a pathogen-free facility and housed in cages...
containing sterilized food and drinking water, as described previously. A/Puerto Rico/8/1934(H1N1) virus (PR8, mouse-adapted virus) was grown and titrated in 10-day-old embryonated chicken eggs. SDS was purchased from the National Institute for the Control of Pharmaceutical and Biological Products, Beijing, China.

The mice were randomly divided into the following groups: 1) control; 2) virus infected; 3) SDS (15, 30, and 60 mg/kg) + virus infected; and 4) oseltamivir + virus infected, as described previously. There are 6 groups in this study, and each group contained 16 mice. BALB/c mice were intranasally inoculated with 10^6 50% tissue culture infective doses (TCID50) per 0.1 ml of PR8 virus, as described previously. The mice were treated with 15, 30, and 60 mg/kg of body weight/day of SDS or 20 mg/kg/day oseltamivir (as a positive control) twice daily via intraperitoneal injection (i.p.) for 5 days. Treatment was initiated at the same time.

On Days 3 and 6 post-viral challenge, the plasma and lung tissues from 5 mice per group were collected and the viral titers and cytokine levels were measured. The survival rate, degree of weight loss, and disease symptoms were observed in the remaining 6 mice for the 14 days following inoculation as described previously. The lungs were collected from the mice for virus titration in embryonated chicken eggs using the Reed and Muench method. All animal experiments were performed in accordance with the recommendations of the Office International des Epizooties (OIE).

**Histopathological Analysis**

Tissue sections were subjected to histopathological analysis. Lung tissues from virus-inoculated mice were fixed in 10% neutral buffered formalin for at least 24 h before processing. The tissues were embedded in paraffin using standard tissue processing procedures. Sections (4-μm thick) were cut and fixed on glass slides. Standard hematoxylin and eosin (HE) staining was performed as previously described.

**Lung Wet Dry Ratio**

The lung wet dry ratio (W/D ratio) was used to evaluate the severity of pulmonary edema. Dry ice was used as narcotic and euthanasia to alleviate the pain of mice. The lungs were removed and the wet weight was measured. The lungs were then placed in an incubator at 60°C for 24 h to obtain the dry weight as previously described.

![Figure 1](image-url)
Cytokine Measurement

After mice were anesthetized with dry ice as previously described, a trachea cannula was inserted into the lungs, through which PBS was injected and aspirated to collect BALF. The fluid recovery rate was greater than 80%. The BALF was collected and centrifuged, and the cytokine levels in the supernatant were subsequently analyzed. The levels of the proinflammatory cytokines IL-1β, IL-6, and TNF-α in the BALF supernatant were evaluated using an ELISA in accordance with the recommended manufacturer instructions. The protein concentrations in the supernatant of the BALF were quantified using a bicinchoninic acid protein assay kit to evaluate the vascular permeability in the airways of mice as previously described.

Western Blot Analysis

The lungs of the mice were removed and immediately frozen in liquid nitrogen until homogenization. Protein was extracted from the lung tissue homogenates and quantified using a bicinchoninic acid protein assay kit, per the manufacturer’s instructions. The same amount of protein was loaded into each well of a sodium dodecyl sulfate polyacrylamide gel. The protein was transferred to polyvinylidene difluoride membrane. The immunoactive proteins were detected by using an enhanced chemiluminescence Western blot detection kit per the manufacturer’s instructions, as described previously.

Statistical Analysis

All data were expressed as the mean ± SD and statistical analysis was performed with SPSS 17.0. The differences were analyzed using a one-way ANOVA (Dunnett’s t-test), and a threshold of \( P < 0.05 \) was considered to be statistically significant.

Results

Clinical Symptoms

On day 3 post-inoculation, the infected mice exhibited similar clinical symptoms (inactivity and wrinkled fur). On day 5, the infected mice developed more severe clinical symptoms, including inactivity, wrinkled fur, hunched posture, loss of appetite, shortness of breath, and loud crackling. Treatment with SDS or oseltamivir relieved these clinical symptoms and virus titer of lung (Figure 1B and 1C). From day 5 post-inoculation, the infected mice began to die, and the body weight of the infected mice was significantly decreased on day 6 post-inoculation compared with the control group (Figure 1D).

Histopathological Analyses

To evaluate the histological changes in the influenza virus-infected mice after SDS treatment, the lung tissues were subjected to HE staining. Histopathological analyses revealed that...
lung tissues from the control mice exhibited a normal structure with no histopathological changes under a light microscope. At 3 days post-inoculation, the lung tissue had a multifocal mild or moderate interstitial inflammatory hyperemia and exudative pathological changes. By 6 days post-inoculation, the lesions in the lung tissue became enlarged and multiple patchy lesions had fused (Figure 2). These influenza virus-induced pathological changes were significantly attenuated by both SDS (30 and 60 mg/kg) and oseltamivir treatment.

**Lung W/D Ratio**

The results showed that influenza virus infection could significantly increase the lung W/D ratio (Figure S1A). However, treatment with SDS (30 and 60 mg/kg) or oseltamivir significantly decreased the lung W/D ratio ($P < 0.05$) compared to those in the virus infected group. The total protein concentration in the BALF was significantly increased in the virus infected group (Figure S1B). In contrast, treatment with SDS (30 and 60 mg/kg) or oseltamivir significantly decreased the total protein concentration ($P < 0.05$) compared to the virus infected group.

**Levels of Proinflammatory Cytokines**

Proinflammatory cytokines appear during the early phase of an inflammatory response, which play a critical role in ALI and contribute to the severity of lung injury. Previous studies have shown that increased levels of proinflammatory cytokines, TNF-$\alpha$, IL-6, and IL-1$\beta$ in the BALF are observed in ARDS patients, and the persistent elevation of proinflammatory cytokines in individuals with ALI has been associated with more severe outcomes. After 6 days post-inoculation, the effect of SDS on IL-1$\beta$, IL-6, and TNF-$\alpha$ production was analyzed by ELISA. The results showed that the concentrations of IL-1$\beta$, IL-6, and TNF-$\alpha$ in the BALF were significantly increased after influenza virus inoculation (Figure S1C-E). However, treatment with SDS (30 and 60 mg/kg) or oseltamivir significantly reduced the level of IL-1$\beta$, IL-6, and TNF-$\alpha$ production compared to those in the virus infected group ($P < 0.05$).

**Discussion and Conclusion**

TLR4 is one of the most well-characterized TLRs, and is located on the plasma membrane. Previous studies have shown that influenza virus infection can lead to activation of various TLR signaling pathways. In particular, inactivated H5N1 influenza virus can induce ALI through the TLR4-NF-$\kappa$B signaling pathway. As an important transcription factor, NF-$\kappa$B plays a critical role in diverse cellular processes. Inhibitor of NF-$\kappa$B (IkBs) and NF-$\kappa$B p65 are important factors involved in the TLR4-NF-$\kappa$B signaling pathway. A Western blot analysis was performed to detect the expression of TLR4, IkB-$\alpha$, and NF-$\kappa$B p65 (Figure 3A). Our results show that influenza virus significantly increased TLR4 expression and IkB-$\alpha$ and NF-$\kappa$B p65 phosphorylation, whereas treatment with SDS (30 and 60 mg/kg) significantly decreased TLR4.
expression, as well as IxB-α and NF-κB p65 phosphorylation (Figure 3B).

ALI is characterized by severe hypoxemia, pulmonary edema, and neutrophil accumulation in the lungs, and there are few effective measures for successful treatment. Therefore, the development of novel therapies for ALI is urgently required.16-19 In this study, an ALI model was established by influenza virus infection in mice, and we observed the effects of SDS on influenza virus-induced ALI. Thus, SDS may represent an effective clinical treatment for influenza virus-induced ALI.

Declaration of Conflicting Interests
The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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References
1. Herold S, Steinmueller M, von Wulffen W, et al. Lung epithelial apoptosis in influenza virus pneumonia: the role of macrophage-expressed TNF-related apoptosis-inducing ligand. J Exp Med. 2008;205(13):3065-3077.
2. Oshansky CM, Gartland AJ, Wong SS, et al. Mucosal immune responses predict clinical outcomes during influenza infection independently of age and viral load. Am J Respir Crit Care Med. 2014;189(4):449-462.
3. Kelly GS. Rhodiola rosea: a possible plant adaptogen. Altern Med Rev. 2001;6(3):293-302.
4. Qian EW, Ge DT, Kong SK. Salidroside protects human erythrocytes against hydrogen peroxide-induced apoptosis. J Nat Prod. 2012;75(4):531-537.
5. Sharma N, Mishra KP, Ganju L. Salidroside exhibits anti-dengue virus activity by upregulating host innate immune factors. Arch Virol. 2016;161(12):3331-3344.
6. Lu R, Wu Y, Guo H, Huang X. Salidroside protects lipopolysaccharide-induced acute lung injury in mice. Dose Response. 2016;14(4):1559325816678492.
7. Wu H, Peng X, Cheng L, et al. Genetic and molecular characterization of H9N2 and H5 avian influenza viruses from live poultry markets in Zhejiang Province, eastern China. Sci Rep. 2015;5:17508.
8. Carty M, Bowie AG. Recent insights into the role of Toll-like receptors in viral infection. Clin Exp Immunol. 2010;161(3):397-406.
9. Reed L, Muench H. A simple method for estimating fifty percent endpoints. Am J Hyg. 1938;27(3):493-497.
10. Hai-bo W, Chao-tian G, Ru-feng L, et al. Characterization of a highly pathogenic H5N1 avian influenza virus isolated from ducks in Eastern China in 2011. Arch Virol. 2012;157(6):1131-1136.
11. Tang Y, Chen Y, Chu Z, et al. Protective effect of cryptotanshinone on lipopolysaccharide-induced acute lung injury in mice. Eur J Pharmacol. 2014;723:494-500.
12. Lin Z, Xu C, Liu B, et al. Analysis of the phylogeny of Chinese H9N2 avian influenza viruses and their pathogenicity in mice. Arch Virol. 2014;159(10):2575-2586.
13. Singla S, Jacobson JR. Statins as a novel therapeutic strategy in acute lung injury. Palm Circ. 2012;2(4):397-406.
14. Imai Y, Kuba K, Neely GG, et al. Identification of oxidative stress and Toll-like receptor 4 signaling as a key pathway of acute lung injury. Cell. 2008;133(2):235-249.
15. Marchant D, Singhera GK, Utokaparch S, et al. Toll-like receptor 4-mediated activation of p38 mitogen-activated protein kinase is a determinant of respiratory virus entry and tropism. J Virol. 2010;84(21):11359-11373.
16. Blank R, Napolitano LM. Epidemiology of ARDS and ALI. Crit Care Clin. 2011;27(3):439-458.
17. Aggarwal NR, King LS, D’Alessio FR. Diverse macrophage populations mediate acute lung inflammation and resolution. Am J Physiol Lung Cell Mol Physiol. 2014;306(8):L709-L725.
18. Lalu MM, Moher D, Marshall J, et al. Efficacy and safety of mesenchymal stromal cells in preclinical models of acute lung injury: a systematic review protocol. Syst Rev. 2014;3:48.
19. Hale BG, Albrecht RA, Garcia-Sastre A. Innate immune evasion strategies of influenza viruses. Future Microbiol. 2010;5(1):23-41.