Review article

Innate-adaptive immunity interplay and redox regulation in immune response

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ABSTRACT

Innate and adaptive immune cell activation and infiltration is the key characteristic of tissue inflammation. The innate immune system is the first line of host defense in which innate immune cells are activated by danger signals, including pathogen- and danger-associated molecular pattern, and metabolite-associated danger signal. Innate immunity activation can directly contribute to tissue inflammation or immune resolution by phagocytosis and secretion of biologically active molecules, or indirectly via antigen-presenting cell (APC) activation-mediated adaptive immune responses. This review article describes the cellular and molecular interplay of innate-adaptive immune systems. Three major mechanisms are emphasized in this article for their role in facilitating innate-adaptive immune interplay. 1) APC can be formed from classical and conditional innate immune cells to bridge innate-adaptive immune response. 2) Immune checkpoint molecular pairs connect innate and adaptive immune cells to direct one-way and two-way immune checkpoint reactions. 3) Metabolic reprogramming during immune responses leads to excessive cytotoxic and mitochondrial reactive oxygen species (ROS) production. Increased NADPH oxidase-derived extracellular and intracellular ROS are mostly responsible for oxidative stress, which contributes to functional changes in immune cells. Further understanding of innate-adaptive immunity interplay and its underlying molecular basis would lead to the identification of therapeutic targets for immunological and inflammatory disease.

1. Introduction

Innate and adaptive immunity are two branches of the immune system that are essential for the initiation and progression of chronic inflammatory diseases, including atherosclerosis [1,2]. Innate immunity serves as the first line of host defense against danger signals in a quick and non-specific manner. For example, a wide range of exogenous and endogenous danger signals, such as oxidized low density lipoprotein (ox-LDL), could activate endothelial cells (ECs) and classical innate immune cells, such as monocytes, macrophages, and dendritic cells (DCs), by binding to toll-like receptors (TLR) [3]. Activated innate immune cells contribute to tissue inflammation by secreting pro-inflammatory mediators, such as tumor necrosis factor α (TNFα), interleukin-6 (IL-6), and reactive oxygen species (ROS) [4]. Along with the innate immune responses, tissue inflammation is also accompanied by the infiltration of adaptive immune cells which convey antigen-specific immune responses by T cells that differentiate into effector T cells with pro- or anti-inflammatory cytokine production, and B cells that differentiate into plasma cells with specific antibody production. Numerous studies highlighted the important role of T and B cell responses in atherosclerosis [5,6].

Three signals were recognized as the mechanism of innate control of the adaptive immune responses, consisting of signal 1 engagement of the T/B cell receptor (TCR/BCR) by antigen peptide on major histocompatibility complex (MHC), signal 2 ligation of immune checkpoint molecular pairs (co-stimulation and co-inhibition), and signal 3 cytokine stimulation [7–9]. We recently proposed signal 4, metabolite-associated danger signals (MADS) recognition by metabolic sensor (MS), as a novel mechanism to connect innate and adaptive immune responses [7]. This signal 4 theory is mostly based on the discovery of...
homocysteine-mediated CD40+ monocyte activation and CD40L production in chronic kidney disease (CKD) patients [10]. As CD40/CD40L is a co-stimulatory molecular pair, essential for both T and B cell activation, uric acid and Hcy-induced CD40/CD40L production/ligation plays a critical role in activating both innate and adaptive immune cells. These findings sparked the hypothesis that a two-way immune checkpoint may be an important modulating machinery for the crosstalk between innate and adaptive immunity [7]. Numerous evidence support the notion that MADS, such as hyperhomocysteinemia (HHcy), uric acid, hyperglycemia, and hyperlipidemia, stimulate tissue inflammation by activating the innate and adaptive immune system [10–14].

We recently summarized major features of metabolic reprogramming in innate and adaptive immune cells and established models of metabolic interplay in anti- and pro-inflammatory responses [15]. During pro-inflammatory immune responses, most immune cells switch towards enhanced glycolysis and pentose phosphate pathway to prompt an activation and proliferation status and adopt pro-inflammatory effector functions. This metabolic reprogramming is associated with increased cytosolic and mitochondrial ROS production [15]. ROS has been increasingly recognized not only as a by-product of metabolic reprogramming but also play an important coordinated role in mediating cell-cell interaction and promoting immune cell activation and differentiation [16]. However, it is challenging to describe the molecular basis and to define the role of ROS in immune cell activation.

This review provides a comprehensive overview for molecular and redox regulation in innate-adaptive immunity interplay and tissue inflammation.

2. Myeloid cells are the major populations in tissue inflammation

Inflammation is a tissue pathological process with the major purpose of resolving infection and tissue repair. Numerous innate and adaptive immune cells reside in inflammatory tissue where they contribute to immune defense and tissue homeostasis in metabolic diseases [17,18]. Recently, single-cell RNA sequencing is appreciated to be an unbiased strategy to explore cell clusters with shared transcriptional signatures and to identify specific molecular signatures that are unique to different cell population. Data from several single-cell RNA sequencing studies [19,20] of aortic leukocytes (CD45+) from healthy and atherosclerotic mice have been reorganized and presented in Table 1. These information indicated that in the healthy aortic vessel, lymphocytes, especially T cells, were the most abundant leukocyte population in Ldlr-/– and Apoe-/– mice. In the atherosclerotic aorta, myeloid cell clusters, including monocyte, macrophage and dendritic cells, were increased in both mice fed a western/high-fat diet (WD/HFD) for 11–12 weeks. With disease progression, myeloid cells were further increased to 63.8% of total immune cells in advanced atherosclerotic aorta of Ldlr-/– mice fed a HFD for 20 weeks, which is similar to the composition in human carotoid plaque (65%). Myeloid cells have also been reported as the major immune cell populations in aortic lesion in hyperlipidemia and HHcy mouse model (Tg-hCBS ApoE−/–, Cbs−/– and Ldlr-/–, Cbs−/–) [12,21]. Therefore, it is recognized that myeloid cells are the major populations in advanced atherosclerotic lesion.

3. Innate immune response

Conditional innate immune cell and metabolic sensor system — Innate immunity is the first line of host defense, providing initial non-specific defense against danger signals. Professional innate immune cells include DC, monocyte, macrophage, and B cell. Recent studies recognized EC and smooth muscle cell as conditional innate immune cells that share most features of professional innate immune cells in response to extracellular environmental changes [22–24]. As described in Fig. 1A, professional and conditional innate immune cells rely on pattern-recognition receptors (PRRs) or metabolic sensor (MS) to rapidly recognize and respond to danger signals derived from invading pathogens, injured self-cells or metabolic stresses. Pathogen-associated molecular patterns (PAMPs) are danger signals produced by microorganisms which active innate immune cells by PRR (PAMP:PRR recognition), such as TLR, nod-like receptors, and C-lectin receptors, with high affinity to facilitate the downstream signaling cascade to initiate immune responses. It was thought that immune system functioned by making a distinction between self- and non-self-constituents until Dr. Matzinger proposed the “danger model” claiming that “danger/alarm signals” from injured cells could initiate immune response in the 1990s [25]. The danger theory gives us a different view that self-constituents can trigger an immune response if they are dangerous, and non-self-constituents can be tolerated if they are not dangerous. Given these, many kinds of endogenous molecules were identified to be danger signals under pathological conditions. Danger-associated molecular patterns (DAMPs) are molecules released under stress or tissue damage condition and initiate immune response by DAMP:PRR recognition.

However, current knowledge could not support the idea that all pro-inflammatory endogenous metabolites can bind to PRRs. MADS are proposed as a novel category of danger signals comprising small molecular metabolites which trigger immune responses mostly by MADS: MS recognition [7]. Using HHcy as a prototype disease model, we found that the Hcy-methionine (HM) cycle functioned as a MS system determining methylation-regulated pathogenic signaling [12,26]. This hypothesis is proved in CKD patients in which Hcy mediates inflammatory CD40+ monocyte differentiation, a biomarker for sever CKD [10]. The HM cycle determines production of S-adenosylmethionine and S-adenosylhomocysteine, a universal donor and competitive inhibitor of cellular methylation, respectively. Unlike PAMP/DAMP:PRR recognition, MADS:MS recognition is mostly receptor-independent, and largely depends on metabolic reprogramming and metabolic modification on targeted molecules and pathogenic signaling [12,26].

**Effector response and trained immunity** — Immunogenic recognition triggers innate immune cell activation and a series of effector responses (Fig. 1A). One of the effector responses is pathogen elimination by secreting cytokines and chemokines, phagocytosis, and others.

**Table 1**

| Cell type | Aorta (Ldlr−/−) | Aorta (Apoe−/−) | Carotid plaque (Human, deconvolve)** |
|-----------|---------------|----------------|-----------------------------------|
|           | 12W CD19 | 12W WD19 | 20W HFD20 | 20W HFD20 | 12W CD19 | 12W WD19 | 12W WD20 | 12W WD20 |
| Macrophage | 13.6 | 27.0 | 28.9 | 49.6 | 4.9 | 9.6 | 9.6 | 51.1 |
| Monocyte | 6.2 | 21.1 | 12.3 | * | 4.9 | 9.6 | 9.6 | 51.1 |
| MoDC/DC | - | 3.9 | 14.9 | 14.2 | 4.9 | 9.6 | 9.6 | 51.1 |
| NRG | 1.7 | 2.1 | * | * | 2.1 | 3.3 | 3.3 | 5.3 |
| B cell | 24.4 | 4.0 | * | * | 2.4 | 1.6 | 1.9 | 4.17 |
| T cell | 54.1 | 45.8 | 28.3 | 25.3 | 66.0 | 49.0 | 38.2 | 19.3 |
| Other cell | - | - | 15.7 | 11.0 | - | - | 9.6 | - |

*Cells were grouped in “other cells”.*
addition, immune recognition induces phenotypic and functional maturation of innate immune cells into potent antigen-presenting cells (APC) which initiate adaptive immune responses [30,31]. Trained immunity or innate immune memory is a novel feature of innate immune cells and conditional innate immune cells [32]. These cells mount protection against secondary infection and respond in a non-specific sensitized manner by undergoing epigenetic reprogramming and metabolism rewiring [32]. Evidence for trained immunity first emerged in humans showing vaccination-induced protections did not only work against the targeted diseases, but also other pathogens [33]. β-glucan, lipopolysaccharides, ox-LDL, uric acid and some pro-inflammatory cytokines (IL-8 and IL-12) induced trained immunity in monocytes [34–36]. Conditional innate immune cells can also be trained to confer long-lasting immunological memories [37]. For example, lysophosphatidylcholine has been shown to elicit trained immunity in ECs by up-regulating glycolysis, mevalonate, and acetyl-CoA generating pathways [38].

### 4. Adaptive immune response

The adaptive immune response is antigen-specific and encompasses cell-mediated (T cell) and humoral-mediated (B cell) immunity which are both critical to drive tissue inflammation or repair (Fig. 1B). T cell response involves 4 distinct regulatory signals [7,39]. Signal 1 (antigen recognition) determines the specificity of T cell response. Antigen peptide is presented by MHCI/II on APC and engaged with antigen-specific TCRs on naive T cell. Signal 2 (immune checkpoint) is marked by the ligation of stimulatory/inhibitory molecular pairs [7]. Signal 3 (cytokine stimulation) strengthens immune cell activation. Activated APCs produce various cytokines that enhance T cell clonal expansion and differentiation [40,41]. Signal 4 (MADS recognition) is a novel signal we proposed to describe small molecular metabolite-induced MADS/MS recognition [7,10].

B cell response is marked by antibody production that play an important role in both innate and adaptive immunity. There are 2 types of B cell immunity. T cell-dependent B cell response describes B2 cells mount antibody responses with the help of follicular helper T (Tfh) cells. Three signals are involved in T cell-dependent B cell response. Signal 1 (antigen recognition) determines the specificity of B cell response in which B cell receptor recognizes specific antigens. Signal 2 (immune checkpoints) is featured by CD40/CD40L ligation contributing to B cell activation, isotype switching and affinity maturation. Signal 3 (cytokine stimulation) strengthens B cell immune response. T cell-independent B cell response refers to the innate defense of B1 cells against a wide range of PAMP/DAMPs. B1 cells produce both natural antibodies and immunomodulatory molecules, such as IL-10, IL35, and granulocyte-macrophage colony-stimulating factor, to regulate acute and chronic inflammatory diseases. The natural antibodies produced by B1 cells differ from B2 cell adaptive antibodies by their low affinity and polyreactivity. Current evidence supports that the IgM antibody generated by B1 cells are atheroprotective [43,44].
5. Innate-adaptive immunity interplay

APC activation bridges innate-adaptive immunity — APC plays a central role in initiating adaptive immune responses and bridging innate-adaptive immunity through their capacity to present antigen to T cells (Fig. 2A). Upon rapid activation by PAMP/DAMP:PRR or MADS:MS recognition, innate immune cells acquire enhanced capacity to process and present antigen by upregulating MHCII and immune checkpoint molecules, such as CD80, CD86 and CD40, which allow them to form APC and efficiently activate CD4 and CD8 T cells [4, 7, 45–47]. DC is one of the professional APCs which link the innate and adaptive response [48, 49]. In response to ox-LDL, human peripheral monocytes can differentiate into DCs which further induce Th1 and Th17 activation and proliferation [50]. Of note, ECs have also been shown to effectively take up, process, and present antigens, with the expression of MHCII/II, immune checkpoint molecules and cytokines, to prime adaptive immunity [51].

Considering the important roles of T and B cells in chronic inflammatory diseases, identification of specific antigens driving adaptive immunity is critical to advance therapeutic strategies. In Fig. 2B, we summarized a series of antigens which have been associated with the pathogenesis of atherosclerosis. Lipoprotein a, lipoprotein-lipase and advanced glycation end products are established MADS relevant to atherosclerotic plaque development. They can induce the expression of MHCII, in-
mune checkpoint molecules and cytokines, to prime adaptive immunity [51].

Pneumocystis carinii pneumonia (PCP) is a gram-negative bacterium which may be causative for atherosclerosis [57]. Knockout of TLR2/4 attenuated C. pneumoniae- accelerated atherosclerosis and reduced foam cell formation in ApoE-/- mice. Several DAMPs, including heat shock proteins, extracellular matrix components and some cell death associated products, are described in atherogenesis. Heat shock proteins may be engaged in humoral immune response as the level of plasma heat shock proteins and their autoantibodies positively correlated with atherosclerosis in humans [58]. T cells specifically responding to heat shock proteins was also found in atherosclerotic plaques [59].

Immunogenic checkpoint connects innate and adaptive immune cell response — As one of the mechanisms for innate control of adaptive immunity, signal 2 has been well-studied about its function to direct T cell function. Based on the findings over the past few years, we classified signal 2 immune checkpoint into one-way and two-way immune checkpoints (Fig. 3A) [7]. The one-way immune checkpoint involves only forward signaling and directs T cell activation. ICOS/ICOSL is one of the well-described one-way immune checkpoints, which are essential for T cell expansion and differentiation [60]. However, the two-way immune checkpoint is bi-directional involving forward and reverse signaling, which modulates both innate and adaptive immunity by inducing T cell and monocyte/APC differentiation/suppression. CD40/CD40L is the best representative of two-way co-stimulatory immune checkpoints, which modulate effector functions of both T cell [61] and monocytes [62, 63]. Immune checkpoints can be further divided into two groups by their co-stimulatory or co-inhibitory function (Fig. 3B) [63–65]. The stimulatory immune checkpoint molecules deliver positive signals leading to APC/T cell activation, differentiation and immune memory. The inhibitory immune checkpoint molecules deliver negative signals resulting in APC/T cell suppression, apoptosis and immune tolerance. Thirty immune checkpoint molecular pairs, consisting of 16 co-stimulatory and 14 co-inhibitory immune checkpoints are summarized in Fig. 3B. This list is based on our recent publication [63] and included updated information by literature search.

Redox regulation in immune responses — ROS are generated by the partial reduction of oxygen to form chemically reactive species, such as superoxide (O2·-), hydrogen peroxide (H2O2), hydroxyl radical (OH·), peroxynitrite (ONOO·), and hypochlorous acid (HOCl), which are also natural byproducts of metabolic reactions [66, 67]. Recently, we summarized current understanding on immune cell metabolic reprogramming and established models of metabolic switch in the progress of immune cell activation [15]. Briefly, increased glycolysis and impaired oxidative phosphorylation are two major metabolic changes in proinflammatory innate and adaptive immune cells. These metabolic changes are associated with excessive ROS production which play critical roles in physiological and pathological processes. Recent redox biology researches in immune cells have generated important information regarding to redox signaling and regulation in immune responses (Fig. 4A).

Redox signaling and regulation in innate immunity — Innate immune response is initiated by immune recognition through PRR or MS which can induce ROS production. Increased ROS is the main antimicrobial response in phagocytes and has been known as respiratory/oxidative burst [68]. In response to large microbes, neutrophil produced an excess amount of extracellular ROS by plasma membrane NADPH oxidase (NADPH) activation which caused selective oxidation of NF-κB leading to increased IL1β expression and amplified neutrophil recruitment [69]. Moreover, mitochondrial has been shown to contribute to ROS production and antimicrobial responses in macrophages in response to E. coli or TLR1,4 and 6 agonist [70]. Reducing mtROS by using mitochondrial electron transport chain complex II SDH-specific inhibitor (3-nitropropionic acid) or overexpressing mitochondrial catalase resulted in defective bacterial killing in macrophages [70, 71]. ROS also induces dendritic cell differentiation and their antigen presenting functions [72].

It is established that NF-κB activation mediates ROS-induced innate immune response. For example, mtROS-mediated formation of disulfide bond in the NF-κB essential modulator (NEMO) is essential for the activation of ERK1/2 and NF-κB signaling in infected macrophages [73]. Innate immune activation induced by asbestos inhalation was revealed to be mediated by NOX-derived ROS-NLPR3 inflammasome signaling [74]. In response to TLR agonist R848, the increased mtROS production was associated with increased cross-presentation capacity of plasmacytoid DC [75]. However, high ROS is not always associated with NF-κB activation, as LPS-treated ROS-high DC had low NF-κB activity compared with ROS-low DC [76].

Natural killer T (NKT) cells play critical roles in killing infected and malignant cells and has been shown to produce higher levels of NOX1/2-related ROS, especially NKT1/17, compared to CD4+ /CD8+ T cells [77]. However, in myeloid specific NOX2-deficient mice or by using NOX2-inhibitor histamine dihydrochloride, myeloid cell-derived ROS has been demonstrated to facilitate metastasis of melanoma cells by suppressing IFNγ-producing NKT cell function [78]. EC has been proposed as one of the conditional innate immune cells which can be activated to form APC and to initiate adaptive immune responses [22, 79]. Consistently, IFNγ induced the expression of MHCII in EC partially mediated by ROS-mediated signaling [80, 81].

Taken together, NOX and mitochondria are two major sources of increased ROS production in innate immune cells, which play critical roles in regulating innate immunity via modulating mostly ERK1/2, NF-κB, TLR and NLPR3 signalings [74].

Redox signaling and regulation in adaptive immunity — Excessive ROS production is associated with the activation, differentiation and survival in T and B cells. Indeed, moderate ROS production is essential for T cell activation whereas excessive ROS can inhibit NF-κB phosphorylation and T cell activities [82]. NOX2-derived ROS was responsible for CD3/CD28 stimulation-mediated CD8+ T cell activation [83]. MtROS production was also induced after TCR activation and promoted T cell activation by regulating IL2 and IL4 expression [84]. Hepatic oxidative stress induced humoral and cellular immune responses in methionine-choline deficient model of non-alcoholic steatohepatitis [85]. Upon activation, T cells differentiate to distinct subsets with different cytokine production. ROS can modulate both T cell
A. APC activation bridges innate-adaptive immune response

![Diagram of APC activation bridges innate-adaptive immune response]

B. Antigen presented by APC in atherosclerosis

| Antigen                                      | PMID                                      |
|----------------------------------------------|-------------------------------------------|
| Homocysteine                                | 28738836, 27006445, 25009174, 19858416,   |
|                                              | 22628578                                  |
| Oxidized low-density lipoprotein             | 10946219,15081469,16270279, 29588316      |
| β2-glycoprotein-1                            | 10441168, 11023938, 16483919              |
| Lipoprotein-lipase                           | 15529371                                  |
| Lipoprotein A                                | 9344887                                   |
| Advanced glycation end products              | 16100036                                  |

**MADS**

**Bacterial**

- Porphyromonas gingivalis: 16013219, 16299299, 20505314
- Chamydia pneumoniae: 10539855
- Bacteroides forsythus: 16013219
- Streptococcus mutans: 16013219, 16299299, 12135941
- Helicobacter pylori: 11795863
- Echerichia coli: 10096931, 11506459

**Virus**

- Enterovirus: 9843459
- Cytomegalovirus: 17680510, 29409508
- human immunodeficiency viruses: 19996940
- Human papilloma virus: 26664889
- Viperin: 16196199

**PAMP**

**DAMP**

- Heat-shock protein: 10604986, 12818399, 25027488
- Extracellular matrix: 29290139
- Collagen: 17327474
- Nucleotide triphosphate: 27339459
- High mobility group box 1: 18574060

Fig. 2. APC bridges innate-adaptive immune responses. A. APC bridges innate-adaptive immune responses. The initial process of immune response is immune recognition of innate immune cell or innate immune-like cell-mediated by MADS:MS or DAMP/PAMP:PRR recognition. Activated innate immune cells process antigens intracellularly and present them via MHCI molecules. This process is called APC formation and serves as a bridge between innate and adaptive immunity. APC presents antigen to T cell leading to T cell activation. B. Antigen presented by APC in atherosclerosis. Antigens presented by APC in atherogenesis are summarized based on literature searching.
differentiation and cytokine production by polarized T cell subsets. For instance, H$_2$O$_2$ reduced IFN$\gamma$ production of activated Th1 and increased IL4 secretion of activated Th2 in vitro which was associated with bronchial hyper-responsiveness and airway remodeling [86]. NOX-derived ROS was also important for Treg differentiation and function since mice with mutated p45phox or gp91phox displayed hampered Treg induction and T cell suppression [87]. Moreover, ROS was involved in activation-induced cell death via Fas/FasL pathway to maintain T cell homeostasis [88].

Similarly, BCR stimulation induced rapid ROS production in primary resting murine B cells [89]. Two distinct sources of ROS were revealed downstream of the BCR signaling, Nox2 in the early stage of B cell activation and mitochondrial respiration at later stage of B cell activation [89]. However, the role of ROS in B cell subsets differentiation remain elusive. Collectively, ROS produced by NOX and mitochondrial are involved in adaptive immune responses which potentially involve in NF-κB activation.

**Redox regulation in innate-adaptive immunity interplay** — Considering that MHC and immune checkpoint molecules are two key molecular features of innate-adaptive immunity activation, we paid special attention on the role of ROS-modulated MHC and immune checkpoint molecules expression in APC (Fig. 4B).

H$_2$O$_2$-treated DC has been shown to be more efficient in promoting T cell proliferation compared with normal DC due to increased expression of MHC-I, MHC-II, and the co-stimulatory molecules CD40 and CD86 [90]. Moreover, xanthine oxidase-derived O$_2^•−$ induced phenotypic and functional maturation of DC, partly through an NF-κB-dependent mechanism [91]. However, in response to ROS inducer, including glutathione synthesis inhibitor, buthionine sulphoximine, and paclitaxel, human and mouse tumor-associated macrophages acquired immunosuppressive phenotype by upregulating the expression of PD-L1 and immunosuppressive cytokines, including IL-4, IL-10 and IL17 [92]. Oxidatively truncated lipid species formed via oxidative degradation of triacylglyceride and cholesterol ester decreased trafficking of peptide-MHC-I complexes to the cell surface in tumor-associated DCs, consequently impaired their ability to initiate and sustain adequate CD8$^+$ T cell responses to tumor cells [93]. Anti-oxidant α-tocopherol vitamin E abrogated this defect by preventing the generation of per-oxidized lipids [93].

Overall, ROS production can be induced in both activated innate and adaptive immune cells, and in turn regulate their activities as well as innate-adaptive immunity interplay. However, further mechanistic studies are needed to translate this knowledge into therapeutic opportunities for metabolic and inflammatory diseases.

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**Fig. 3. Immune checkpoint molecules in innate-adaptive immunity interplay.** A. Immune checkpoint (signal 2) in regulating innate-adaptive immune response. We defined two types of immune checkpoints, one-way and two-way, based on signal 2 direction. The one-way immune checkpoint only involves forward signaling and directs to T cell activation or suppression. The two-way immune checkpoint involves forward and reverse signaling which modulates both innate and adaptive immunity by inducing TC activation/suppression and innate immune cell/APC activation/suppression. B. Thirty immune checkpoint molecular pairs regulate T cell and APC function. Thirty immune checkpoint molecular pairs and their functions are illustrated. Red color symbol describes the stimulatory molecular pair and green color symbol describes inhibitory molecular pairs. Stimulatory immune checkpoint molecular pair ligation leads to T cell/APC activation and differentiation, whereas inhibitory immune checkpoint molecular pair ligation leads to T cell/APC suppression and apoptosis. (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)
A. Redox regulation and signaling in innate and adaptive immune cells

| ROS Induction                  | Cell Types | Disease Conditions /Stimuli | Molecular/cellular Change                      | Effector Response | PMID       |
|-------------------------------|------------|-----------------------------|------------------------------------------------|-------------------|------------|
| Extracellular (PM NOX-derived)| NP         | Large-size microbes         | ↑IL-1β amplifying Nφ recruitment                | Activation        | 28314592   |
| Intracellular (Mt-derived)    | pDC        | TLR stimulation             | Triggering CD8⁺ T cell                          | Activation        | 29884826   |
| Extracellular (NOX2-derived)  | Myeloid cell| Melanoma                    | ↓IFN-γ secretion, ↓NK cell infiltration         | Suppression        | 28760732   |
| Intracellular (NOX2-derived)  | NKT1/17    | -                           | ↑IFN-γ / IL-17                                  | Activation        | 29021374   |
| Intracellular (Lipid peroxidation) | CD4⁺ T cell | NASH                        | Th-1 activation of CD4⁺ T cell                 | Activation        | 24115128   |
| Intracellular (NOX2-derived)  | CD8⁺ T cell| CD3/CD28 stimulation        | CD39, ↑IFN-γ secretion                          | Activation        | 26549640   |
| Intracellular (NOX2/Mt-derived)| B cell    | BCR stimulation             | IgM and IgG1 antibody titers                    | Activation        | 23024271   |
| Intracellular or extracellular| EC         | IFNγ                        | ↑MHC-I                                        | Transformed to APC| 12006557   |

B. Oxidative stress-related MHC and checkpoint molecular changes in APC

| Oxidative stress in APC   | Cell Types | Disease Conditions /Stimuli | APC Checkpoint Molecular Change | Checkpoint Function | TC Response | PMID       |
|---------------------------|------------|-----------------------------|--------------------------------|---------------------|-------------|------------|
| H₂O₂ treatment           | DC         | Healthy volunteers          | ↑MHC-I, I; ↑CD40 and CD86     | Co-stimulatory      | Activation  | 9890657    |
| ↑XO-derived O₂⁻ in DC    | DC         | Not analyzed                | ↑CD80, CD83, CD86              | Co-stimulatory      | Activation  | 12554628   |
| ↑Non-specific ROS in Mφ   | Mφ         | TNBC                        | ↑PD-L1                         | Co-inhibitory       | CD8⁺ T cell suppression | 30770442 |
| ↑Oxidatively truncated lipids in IDC | IDC | Tumor                   | pMHC-I on cell surface         | Impairing cross-presentation | CD8⁺ suppression | 29242535 |

C. Model of redox regulation in immune response

- **BCR/TCR/CD3/CD28 stimulation** → ↑NOX/Mt → ↑Intracellular ROS → BC/TC/NKT → Proliferation/activation (↑IFN-γ, IL-17, CD39)
- **LPS/TLR stimulation** → ↑NOX/Mt → Intracellular ROS → pDC → Proliferation/activation (↑IL-1β)
- **LPS/INF-γ stimulation** → ↑NOX/Mt → Intracellular ROS → Extracellular ROS → EC → APC-like EC → Conditional APC formation/activation (↑MHC-II)
- **Inflammation** → ↑NOX/NOX → Intracellular ROS → Extracellular ROS → DC → T cell → APC formation/activation → T cell activation (↑CD40, CD80, CD83, CD86, MHC-I, II)
- **Tumor (TNBC/Melanoma)** → ↑NOX/Mt → Intracellular ROS → Extracellular ROS → MC/MMφ/IDC → T cell → APC formation/activation → T cell suppression (↑PD-L1, ↑IFN-γ, pMHC I on cell surface)

D. NOX1/2/4 play major role in intracellular and extracellular ROS induction

- **Cell 1**
  - NOX1/2 → AQP → O₂⁻ → SOD3 → H₂O₂ → Extracellular ROS

- **Cell 2**
  - NOX1/2 → AQP → O₂⁻ → SOD1 → H₂O₂ → Cytosol ROS
  - NOX1/2 → ER/ES → PKC, c-Src, p-P47 → PDK → Mitochondrial ROS

(caption on next page)
6. Model of redox regulation in immune response

In Fig. 4C, we presented a model to characterize redox regulation in different immune cells during immune responses. Various danger signals led to increased intracellular and extracellular ROS production in immune cells. Elevated NOX- and mitochondria-derived intracellular ROS is associated with B and T cell activation and proliferation in response to BCR/TCR/CDS/CD28 stimulation. Under TLR stimulation, the production of NOX- and mitochondria-derived intracellular and extracellular ROS in neutrophils and pDC are increased and can further promote their activation and proliferation. Moreover, in innate immune-like cell EC, pro-inflammatory cytokine IFN-γ/LPS induce NOX- and mitochondria-derived intracellular and extracellular ROS production, which contribute to the conversion and activation of ECs to APC by upregulating MHCII molecules. The increased NOX/xanthine oxidase-derived intracellular and extracellular ROS in innate immune cells under inflammatory danger signal stimulation played a critical role in mediating APC and T cell activation by modulating the expression of MHC and immune checkpoint molecules. Differently, in the tumor microenvironment, excessive intracellular or extracellular ROS production in innate immune cells, including monocyte, macrophage and DC, is associated with APC formation and activation but leads to T cell suppression partially mediated by increased co-inhibitory immune checkpoint molecule PD-L1.

7. Intracellular ROS and ROS compartmentalization

ROS generation within specific subcellular compartments is mostly related with spatial metabolic processes, differential thiol concentration and specific redox enzyme distribution. This compartmentalization provides local control of ROS regulation and signaling between compartments. We modeled and discussed biochemical and molecular basis underlying ROS compartmentalization in our recent publications [15, 67]. For instance, glycolysis and pentose phosphate pathway contribute to cytosolic ROS (cROS) production by generating reducing equivalents NADPH catalyzed by NOX to produce O$_2^•$ or H$_2$O$_2$. As a consequence of electron leakage, the mitochondrial electron transport chain continuously generates mitochondrial ROS (mROS) using NADH and FADH$_2$ as reductants [15]. Mitochondria are a major source of intracellular ROS and especially susceptible to oxidative stress. This is most likely due to high levels of ROS generated from lipid peroxidation, protein oxidation and mitochondrial DNA mutations. However, it was found that mitochondria have higher concentration of glutathione (GSH) which is a major hydrophilic antioxidant and most reactive electron donor [94]. About 10–15% of the total cell GSH content was found in mitochondria which play a critical role in the maintenance of mitochondrial function and cell survival [95].

Moreover, redox enzyme subcellular localization is one of the major mechanisms defining ROS compartmentalization. As indicated in Fig. 4D, ROS clearance enzyme superoxide dismutase (SOD) has 3 family members in mammal, of which SOD1 is located in the cytoplasm, SOD2 in the mitochondria, and SOD3 is extracellular [96]. NOX, a membrane-bound multi-enzyme complex, is the major producer of ROS with various tissue distribution and subcellular localization. NOX family is responsible for transporting electrons across biological membranes and the reduction of oxygen into superoxide. NOX consists of seven members, NOX1, NOX2, NOX3, NOX4, NOX5, DUOX1 and DUOX2 [67]. Localization of NOX play critical roles in regulating ROS production and the pathways they control. NOX1 and NOX2 has been found in plasma membrane, endosome, phagosome, and endoplasmic reticulum (ER). The NOX4 has been identified in focal adhesions, nucleus, ER and mitochondrion intermembrane. The DUOX1 and DUOX2 were localized to the plasma membrane. However, little information is available about the subcellular localization of NOX3 and NOX5. As NOX and NOX-derived ROS have been implicated in regulating immune responses and the pathologies of various diseases, we further illustrated the biochemical processes of how NOX contributes to the generation of intracellular and extracellular ROS in immune cells in Fig. 4D.

NOX1/2/4 play major role in intracellular and extracellular ROS induction (Fig. 4D) — Plasma membrane-located NOX1/2 is the major producer of extracellular ROS which may serve as communication molecules in immune responses. The activity of NOX is dependent on subunit interaction [97]. In response to danger signals, including PAMP, DAMP and MADS, plasma membrane located NOX1/2 are activated [98]. Activated NOX1/2 produce and release O$_2^•$ directly into the extracellular spaces which are dismutated into H$_2$O$_2$ by superoxide dismutase (SOD) 3. Different from O$_2^•-$H$_2$O$_2$ is relatively permeable and may be diffused into cell via aquaporins which are initially defined as membrane-intrinsic proteins water channel and involved in osmotic volume regulation. Recently, increasing studies have demonstrated that some aquaporin isotypes from mammals and plants can transport H$_2$O$_2$ across organelle and plasma membranes and play a role in oxidative stresses [99–102]. However, the efficiency of H$_2$O$_2$ transport varies with different aquaporin homologs. For instance, two plant aquaporins, AP1P2; 4 and SoPPIP2; 1, and a human aquaporin AQP1 were all transporters of both H$_2$O and H$_2$O$_2$, but the plant aquaporins were more permeable to H$_2$O than the human homologue [100]. Controversially, by using improved H$_2$O$_2$ imaging, another study demonstrated that H$_2$O$_2$ produced in HeLa cells upon EGFR activation can not be diffused out across the cytoplasm [103]. Further studies are needed to address how aquaporins regulate H$_2$O$_2$ transport across organelle and plasma membranes, and whether H$_2$O$_2$ molecules produced outside or inside the cells function differently.

NOX located in ER, endosome, and mitochondrial play major roles in intracellular ROS production. The compartmentalization of NOX-derived ROS may contribute to its functional specificity [102]. Excessive cROS in immune cells is mostly produced by NOX1/2 located in ER and endosome, which can be activated via protein kinase C (PKC)-induced P47phox phosphorylation. Intraellular O$_2^•$ can be dismutated by SOD1 to form H$_2$O$_2$. Further, cytosolic H$_2$O$_2$ can activate Mt-KATP via PKC-ε signaling. Increased K$^+$ influx reduces the mitochondrial membrane potential ($\Delta$Vm) leading to electron transport chain-derived O$_2^•$ production. NOX4 is located in the mitochondrial intermembrane, which is the key enzyme facilitating mitochondria-derived O$_2^•$ production. NOX4-derived O$_2^•$ is released into mitochondrial matrix and dismutated by SOD2 to H$_2$O$_2$ which can then be transported into the cytosol via QAQ.
AQP. Therefore, NOX1/2/4-located in cytoplasm and ER/endosome membranes and mitochondrial intermembrane play major roles in intracellular and extracellular ROS induction in immune response.

The general structure of different NOX isotypes is similar, consisting of two transmembrane subunits, three cytosolic subunits, and G protein Rac. The membrane subunits form the catalytic core of NOX, such as gp91phox (also known as NOX2, or its homologs) and p22phox. The cytosolic subunits, including p47phox, p67phox, p40phox, and G protein Rac, are required for assembly and activation of NOX complex [104]. The regulation of NOX activity varies depending on the isoform [105]. Generally, NOX activation requires the translocation of the cytosolic subunits and Rac to the transmembrane subunits to activate the catalytic core of NOX which produces O$_2^•$ by transferring electrons from NADPH to molecular oxygen [104]. Phosphorylation of p47phox, p67phox and Rac by kinases, such as PKC, PKA, mitogen activated protein kinase, CAMP dependent kinase, and p21 activated kinase, plays critical roles in facilitating these process [106–108]. Overall, NOX activity is mostly regulated by the activation and interaction of its subunits.

8. Therapeutic potential of antioxidant

Redox regulation is implicated in modulating innate and adaptive immunity and pathophysiology of many diseases, such as atherosclerosis, diabetes, rheumatoid arthritis, cancer, and neurodegenerative diseases [98,109]. Increasing efforts have been made to study the effects of antioxidants on the prevention and treatment of these diseases [110]. There are two major category of antioxidants, enzymatic and non-enzymatic antioxidants [111]. Enzymatic antioxidants are endogenous enzymes catalyzing the neutralization of ROS and reactive oxygen species (ROS) and reactive nitrogen species (RNS). However, ROS and RNS are effective signaling molecules controlling several cellular functions [112]. The general structure of different NOX isotypes is similar, consisting of two transmembrane subunits, three cytosolic subunits, and G protein Rac. The membrane subunits form the catalytic core of NOX, such as gp91phox (also known as NOX2, or its homologs) and p22phox. The cytosolic subunits, including p47phox, p67phox, p40phox, and G protein Rac, are required for assembly and activation of NOX complex (Fig. 5). The regulation of NOX activity varies depending on the isoform. Generally, NOX activation requires the translocation of the cytosolic subunits and Rac to the transmembrane subunits to activate the catalytic core of NOX which produces O$_2^•$ by transferring electrons from NADPH to molecular oxygen. Phosphorylation of p47phox, p67phox and Rac by kinases, such as PKC, PKA, mitogen activated protein kinase, CAMP dependent kinase, and p21 activated kinase, plays critical roles in facilitating these processes. Overall, NOX activity is mostly regulated by the activation and interaction of its subunits.

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**Fig. 5. Innate-adaptive immunity interplay and redox regulation in immune response.** Danger signals (DAMP/PAMP/MADS) induce immune response firstly via innate immune cell activation which leads to cytokine/chemokine production, trained immunity, pathogen elimination, and APC activation. The activated APC interacts with Naïve CD8$^+$ and Naïve CD4$^+$ T cell and function as a bridge to connect the innate and adaptive immune systems. Immune checkpoint molecule pairs (signal 2) determine the stimulatory or inhibitory immune response on T cell and APC. In the course of immune cell activation, active metabolic reprogramming leads to redox regulation and increased intracellular and extracellular ROS production. This results in immune subset differentiation. Naïve CD8$^+$ T cell differentiates into CTL which lyse infected and tumor cells. Naïve CD4$^+$ T cell differentiates into inflammatory subsets Th1/2/9/17/22, anti-inflammatory subsets Treg, and Thf subset which helps Naïve B cell/B2 to differentiate into a plasma cell and the memory cell. MC/MΦ are classical APC and can be differentiated into anti-inflammatory subsets (not listed) and pro-inflammatory subsets (CD14++,CD16+, CD14’CD40+, CD1b + Ly6C+(m), M1, M4, Mox). Immune cell subsets produce distinguish effector molecules which contribute to tissue inflammation or repair.

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nitrogen species, including SOD, catalase, glutathione peroxidase, and thioredoxin reductase. Whereas non-enzymatic antioxidants are small molecules produced endogenously by metabolism, such as GSH, coenzyme W, and bilirubin, or provided by food or supplements, such as vitamin E, vitamin C, and carotenoids [111]. By neutralizing the excess of free radicals, antioxidant has been shown to be protective in vitro and contribute to disease prevention in several mouse models [112–114]. However, the results from clinical trials are controversial. For instance, although the most well studied antioxidants, vitamin C and E, have been shown to be atheroprotective in mouse [115], randomized controlled trials revealed that neither vitamin C nor vitamin E supplementation reduced the risk of major cardiovascular events in middle-aged and older men after a mean of 8 years of treatment and follow-up [116]. A systematic review and meta-analysis included 68 randomized trials with 232,606 participants revealed that treatment with beta carotene, vitamin A, and vitamin E may increase mortality [117]. In fact, although redox homeostasis is essential to life, the application of antioxidant supplement or therapy should undergo further evaluation. Disease site, cell type and compartment-specific control of ROS by agents with stable antioxidant activities may be potential targets for future investigation and treatment.

9. Summary

Three mechanisms are highlighted in this review to illustrate the innate-adaptive immunity interplay in tissue immune response (Fig. 5). 1) APC activation bridges innate-adaptive immune responses. 2) Immune checkpoint molecular reaction directs innate and adaptive immune cell responses. 3) Metabolic reprogramming leads to excessive intracellular and extracellular ROS production, mostly related to NOX activation in immune cells.

Immune recognition via DMAP/PAMP/PRR or MADS/MS in innate immune cells triggers APC activation, which bridges the innate and adaptive immune systems by interacting with Naïve CD4+ /CD8+ T cells. As mentioned, four signals are initiated by APC to elicit adaptive immune responses. Immune checkpoint molecule pairs (signal 2) play a critical role in connecting innate and adaptive immune cells for stimulatory or inhibitory immune response, and direct immune response towards either cell by one-way or two-way immune checkpoint reactions. Importantly, as detailed in our recent publication, immune cells undergo metabolic reprogramming during immune responses, which is characterized as altered energy production, biosynthesis and epigenetic reprogramming [15]. To fulfill increased energetic and biosynthetic demands for defense response and damage repair, activated immune cells tend to increased glycolysis activity but decrease oxidative phosphorylation for prompt ATP production to adopt proliferative status and pro-inflammatory effector functions. As one of the consequences of cellular metabolism changes, distinct epigenetic reprogramming has also been found in activated immune cells due to altered accessibility of acetyl/methyl group donor and metabolite-modulated activity of epigenetic enzymes [18]. Elevated acetylation but suppressed methylation is often associated with a pro-inflammatory status in immune cells in many cases [15].

In activated immune cells, metabolic reprogramming directly leads to increased ROS production. For example, activated glycolysis and pentose phosphate pathway lead to increased NADPH and cROS production [118]. Elevated levels of mtROS can also be attributed to mitochondrial intermembrane-located NOX4 complex activation. In addition, plasma membrane bounded-NOX1/2 complexes activation results in extracellular ROS production. ER and endosome membrane bound-NOX1/2 complex activation leads to cROS production. Excessive ROS production in immune cells leads to oxidative stress, which causes molecular damage, disrupted biochemical and cellular functions.

The regulation of innate-adaptive immunity interplay by APC activation, immune checkpoint molecular reactions and metabolic reprogramming determines distinct immune cell subsets differentiation and direct the immune responses to tissue inflammation, immune resolution, tissue repair or immune tolerance. In response to danger signals, monocyte can be differentiated into anti-inflammatory subsets (CD14+CD16+ monocyte in human, CD1b+Ly6C+ monocyte in mice, CD14+CD40+ monocyte and M2/Mhem macrophage) and pro-inflammatory subsets (CD14+CD16+ monocyte in human, CD1b+Ly6C+ monocyte in mice, CD14+CD40+ monocyte, M1 and M4 macrophage) [4,119]. The pro-inflammatory monocytes selectively traffic to the sites of inflammation and are most likely to be differentiated to pro-inflammatory macrophages which contribute to tissue inflammation by producing TNFα, IL-1β, ROS and other inflammatory mediators [4]. However, anti-inflammatory MCs are prone to differentiate to anti-inflammatory macrophages and secrete anti-inflammatory cytokines (IL-10), resulting in tissue repair. Naïve CD8+ T cells can be activated to cytotoxic T lymphocyte to destroy infected cells and tumor cells. In contrast, naïve CD4+ T cell can be differentiated into different effector T cells, including the pro-inflammatory subsets Th1/2/9/17/22, anti-inflammatory regulatory T cells and Tfh cell. Here, we refer the details about the functions of different immune cells subsets in tissue inflammation and diseases to another review article we recently published [119].

This article comprehensively illustrated the updated knowledge of innate-adaptive immunity interplay with a focus on the bridging role of APC formation, immune checkpoint, and redox regulation. Further understanding the molecular basis of innate-adaptive immunity interplay and how they contribute to metabolic and inflammatory diseases could lead to the identification of novel therapeutic targets.

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Declaration of competing interest

None.

Nonstandard abbreviations and acronyms.

| Abbreviation | Description |
|--------------|-------------|
| Ag           | Antigen     |
| APC          | Antigen-presenting cell |
| AQP          | Aquaporin   |
| BCR          | B cell receptor |
| CDK          | Chronic kidney disease |
| cROS         | Cytosolic reactive oxygen species |
| DC           | Dendritic cell |
| DAMP         | Danger-associated molecular pattern |
| EC           | Endothelial cell |
| ER           | Endoplasmic reticulum |
| ES           | Endosome |
| HIF1α        | Interferon |
| IL6          | Interleukin6 |
| MADS         | Metabolite-associated danger signal |
| MHC          | Major histocompatibility complex |
| MtROS        | Mitochondrial reactive oxygen species |
| MS           | Metabolic sensor |
| Mφ            | Macrophage |
| NKT          | Natural killer T cell |
| NOX          | NADPH oxidase |
| Nφ            | Neutrophil |
| PAMP         | Pathogen-associated molecular pattern |
| pDC          | Plasmacytoid dendritic cell |
| PKC          | Protein kinase C |
| PRR          | Pattern recognition receptor |
| ROS          | Reactive oxygen species |
| SOD          | Superoxide dismutase |
| TCR          | Toll cell receptor |

(continued on next page)
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