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Different environmental gradients affect different measures of snake β-diversity in the Amazon rainforests

Rafael de Fraga Corresp., 1, 2, 3, Miquéias Ferrão 1, Adam J Stow 2, William E Magnusson 4, Albertina P Lima 4

1 Programa de Pós-Graduação em Ecologia, Instituto Nacional de Pesquisas da Amazônia, Manaus, Amazonas, Brazil
2 Department of Biological Sciences, Macquarie University, Sydney, New South Wales, Australia
3 Programa de Pós-graduação em Sociedade, Natureza e Desenvolvimento, Universidade Federal do Oeste do Pará, Santarém, Pará, Brazil
4 Coordenação de Biodiversidade, Instituto Nacional de Pesquisas da Amazônia, Manaus, Amazonas, Brazil

Corresponding Author: Rafael de Fraga
Email address: r.defraga@gmail.com

Mechanisms generating and maintaining biodiversity at regional scales may be evaluated by quantifying β-diversity along environmental gradients. Differences in assemblages result in biotic complementarities and redundancies among sites, which may be quantified through multi-dimensional approaches incorporating taxonomic β-diversity (TBD), functional β-diversity (FBD) and phylogenetic β-diversity (PBD). Here we test the hypothesis that snake TBD, FBD and PBD are influenced by environmental gradients, independently of geographic distance. The gradients tested are expected to affect snake assemblages indirectly, such as clay content in the soil determining primary production and Height Above the Nearest Drainage (HAND) determining prey availability, or directly, such as percentage of tree cover determining availability of resting and nesting sites, and climate (temperature and precipitation) causing physiological filtering. We sampled snakes in 21 sampling plots, each covering 5 km², distributed over 880 km in the central-southern Amazon Basin. We used dissimilarities between sampling sites to quantify TBD, FBD and PBD, which were response variables in multiple-linear-regression and RDA models. We show that patterns of snake community composition based on TBD, FBD and PBD are associated with environmental heterogeneity in the Amazon. Despite positive correlations between all β-diversity measures, TBD responded to different environmental gradients compared to FBD and PBD. Our findings suggest that multi-dimensional approaches are more informative for ecological studies and conservation actions compared to a single diversity measure.
Different environmental gradients affect different measures of snake $\beta$-diversity in the Amazon rainforests

Rafael de Fraga$^{1,2,3,*}$, Miquéias Ferrão$^1$, Adam J. Stow$^3$, William E. Magnusson$^4$, Albertina P. Lima$^4$

1 Programa de Pós-graduação em Ecologia, Instituto Nacional de Pesquisas da Amazônia, Manaus, Amazonas, Brazil
2 Programa de Pós-graduação em Sociedade, Natureza e Desenvolvimento, Universidade Federal do Oeste do Pará, Santarém, Pará, Brazil
3 Department of Biological Sciences, Macquarie University, Sydney, New South Wales, Australia
4 Coordenação de Biodiversidade, Instituto Nacional de Pesquisas da Amazônia, Manaus, Amazonas, Brazil

* corresponding author: Rafael de Fraga r.defraga@gmail.com
Abstract

Mechanisms generating and maintaining biodiversity at regional scales may be evaluated by quantifying β-diversity along environmental gradients. Differences in assemblages result in biotic complementarities and redundancies among sites, which may be quantified through multi-dimensional approaches incorporating taxonomic β-diversity (TBD), functional β-diversity (FBD) and phylogenetic β-diversity (PBD). Here we test the hypothesis that snake TBD, FBD and PBD are influenced by environmental gradients, independently of geographic distance. The gradients tested are expected to affect snake assemblages indirectly, such as clay content in the soil determining primary production and Height Above the Nearest Drainage (HAND) determining prey availability, or directly, such as percentage of tree cover determining availability of resting and nesting sites, and climate (temperature and precipitation) causing physiological filtering. We sampled snakes in 21 sampling plots, each covering 5 km², distributed over 880 km in the central-southern Amazon Basin. We used dissimilarities between sampling sites to quantify TBD, FBD and PBD, which were response variables in multiple-linear-regression and RDA models. We show that patterns of snake community composition based on TBD, FBD and PBD are associated with environmental heterogeneity in the Amazon. Despite positive correlations between all β-diversity measures, TBD responded to different environmental gradients compared to FBD and PBD. Our findings suggest that multi-dimensional approaches are more informative for ecological studies and conservation actions compared to a single diversity measure.
Introduction

Investigating how environmental gradients influence community structure is crucial to understanding mechanisms and processes affecting biodiversity at different scales (Keddy, 1992). Quantifying species-habitat associations across continuous landscapes helps disentangle the mechanisms generating and maintaining patterns of regional and local biodiversity. This has been widely demonstrated in the Amazon rainforests by estimates of assemblage β-diversity associated with environmental gradients at mesoscales (e.g., Drucker, Costa & Magnusson, 2008; Fraga, Lima & Magnusson, 2011; Bueno et al., 2012; Ribeiro-Jr, Lima & Magnusson, 2012; Rojas-Ahumada, Landeiro & Menin, 2012; Moulatlet et al., 2014; Menger et al., 2017). In general, levels of β-diversity across heterogeneous continuous landscapes has been estimated through biotic complementarities and redundancies among sites. Most studies focused on measures of β-diversity based on between-site dissimilarities in quantitative (based on abundance data) and qualitative (presence/absence data) species composition. However, use of multiple dimensions may be more informative, because different diversity measures often carry complementary information (Devictor et al., 2010; Weinstein et al., 2014).

Numerous methods have been developed to quantify β-diversity, and each method potentially gives different insights into the mechanisms driving biodiversity (Dehling et al., 2014). Values of taxonomic β-diversity (TBD) may change across heterogeneous landscapes in response to variation in availability of resources, because of selection for different physiological characteristics, ecological plasticity, intra and interspecific interactions, and dispersal ability (Mariac et al., 2011; Hangartner, Laurila & Räsänen, 2012). In general, patterns of community assembly are caused by different portions of environmental gradients providing suitable
conditions to habitat-specialist species, while generalists cover larger portions of gradients (e.g., Kinupp & Magnusson, 2005).

Estimates of functional β-diversity (FBD) may change across heterogeneous landscapes because function is mediated by phenotypes potentially affecting the species performance and fitness, such as morphological, biochemical, behavioral and phenological traits (Petchey & Gaston, 2002). Functional traits can be environmentally filtered when environmental heterogeneity is sufficient to cause variation in local adaptation to different selection pressures (Weinstein et al., 2014). Additionally, some traits such as foraging mode may determine the ability of snakes to cross different habitat patches in Amazonia (Fraga et al., 2017). High levels of FBD are often related to ecosystem dynamics, stability and productivity (Tilman, 2001).

Estimates of phylogenetic β-diversity (PBD) usually incorporate information on the evolutionary history that is shared among species throughout assemblages (Milcu et al., 2013). High values of PBD suggest convergent adaptation among historically isolated assemblages, but recently connected (Weinstein et al., 2014), which is expected as a result from timescale variation in stability and connectivity among habitat patches (Morlon et al., 2011; Jetz & Fine, 2012). In the Amazon rainforests, a combination between the Andean uplift and climate change are though as major factors driving habitat evolution, and consequently driving species diversification, dispersal and extinction (Hoorn et al., 2010). Alternatively, levels of PBD positively related to FBD suggest assemblages evolutionary structured by niche conservatism (Wiens & Graham, 2005).

Measures of β-diversity based on between-site differences in TBD, FBD and PBD are more effective at identifying factors shaping community structure than measures of α-diversity, such as number of species or functional groups at particular sites (Cadotte et al., 2009; Flynn et
This is because mechanisms influencing community assembly act on biotic complementarities and redundancies among sites, and not on the number of organism-units within sites (Diaz & Cabido, 2001; McGill et al., 2006). In general, α-diversity measures fail to capture the contribution of each species to the regional diversity, because different sites may have equal values of diversity (e.g., number of species), even if the species found in each site are taxonomically, functionally or phylogenetically distinct. Identifying mechanisms underpinning β-diversity has clear implications for conservation management. These include identification of unique characteristics in the regional diversity, which makes a site irreplaceable and therefore a priority for conservation actions (Pressey et al., 1993). This approach has been used to test the efficiency of protected areas in France (Meynard et al., 2011), the effects of forest modification on birds and trees in South Africa (Grass et al., 2015), and the efficiency of environmental law in Brazil (Fraga, Lima & Magnusson, 2011; Bueno et al., 2012).

Although much of the Amazon basin appears relatively homogeneous in satellite images, ecological studies at mesoscales have shown that subtle changes along environmental gradients influence patterns of co-occurrence of frogs (Ribeiro-Jr, Lima & Magnusson, 2012; Rojas-Ahumada, Landeiro & Menin, 2012), understory birds (Bueno et al., 2012; Menger et al., 2017), plants (Costa, Magnusson & Luizão, 2005; Drucker, Costa & Magnusson, 2008) and snakes (Fraga, Lima & Magnusson, 2011). In this study, we sampled a continuous landscape across about 880 km of rainforest, from central to southwest Amazonia. Most species of snakes recorded are widely distributed throughout the Amazon basin and some occur in other ecosystems in South America. However, species do not occupy all sites within their ranges, and different assemblages could be expected even at scales of tens of kilometers (Fraga, Lima & Magnusson, 2011). The wide distributions indicate that assemblage differences are more likely to
result from environmental effects than from historical contingencies, such as dispersal limitation across geographic barriers.

In this study we examine the influence of environmental gradients on TBD, FBD and PBD estimates for snake assemblages in the Amazon rainforests. Investigating multiple assemblage dimensions in the same study system potentially allows accessing broad pictures of factors causing and maintaining biodiversity. Snakes are useful organisms to test for multidimensional changes in assemblages over landscapes in the Amazon because of the exceptionally-high species diversity (e.g., Bernarde et al., 2012), great species-trait diversity (e.g., body size and colors, reproductive and foraging modes), which implies large variation in functional traits (e.g., Martins & Oliveira, 1999), and heterogeneous habitats that potentially affect dispersal and gene flow (Fraga et al., 2017). In addition, snakes have been included in estimates of global reptile decline (Gibbons et al., 2000), which highlights the importance of assessing a broad picture of mechanisms generating diversity. This is particularly important in the area we sampled, because it is under strong anthropogenic pressure due to rapid urban growth (Soares-Filho et al., 2006; Fraga et al., 2013a), deforestation associated with paving the major access road (Fearnside & Graça, 2006; Soares-Filho et al., 2006), and flooding by hydroelectric power plants (Fearnside, 2014). Few studies have attempted to identify multiple factors driving snake community assembly (e.g., Fraga, Lima & Magnusson, 2011). Furthermore, to our knowledge, the standardized sampling effort used over such a large area in this study is unprecedented.

We evaluate the effects of environmental gradients on snake assemblages and investigate spatial structuring causing levels of TBD, FBD and PBD among sites. We test the general hypothesis that patterns of community composition in taxonomic, functional and phylogenetic
spaces result from current environmental heterogeneity. Additionally, we hypothesize that different β-diversity measures should respond to different environmental gradients, because they carry complementary information on snake assemblages.

Materials & Methods

Snake sampling. We sampled snakes in 21 RAPELD sampling sites (Magnusson et al., 2005, 2013), each of which has trails covering 5 km² (5 km separated by 1 km). RAPELD is a Brazilian acronym to accommodate two study scales, rapid assessments (RAP) and long-term ecological research (PELD – in Portuguese, Pesquisas Ecológicas de Longa Duração). RAPELD sampling sites will be mentioned throughout the text as modules.

In each module, we sampled ten 250 m long by 10 m wide plots with center lines following the altitudinal contours. The plots were distributed along two parallel 5-km long trails (five plots per trail) with standardized distance of 1 km between neighboring plots. The modules were distributed almost linearly over 880 km (Fig. 1) from central (Manaus, Amazonas) to southwest Amazonia (Porto Velho, Rondônia). The study area includes Central Amazonia, to the north of the Amazon River, the interfluve between the middle regions of the Madeira and Purus rivers, and the upper Madeira River, in southwestern Amazonia. Three modules were installed at the Ducke Reserve, which is a 100-km² fragment of non-flooded primary rainforest, located on the northern outskirts of Manaus. Eleven modules were installed along the federal highway BR-319 that connects Manaus to western Brazil. The highway was largely abandoned in the 1980s, and the modules were installed mainly to enable multi-taxa impact assessments of the effect of the road on biodiversity. Along this road, modules were placed in areas covered by primary and old-secondary rainforest, with patches of flooded forest and campinarana (forest on white sand).
The southern Madeira River region contains seven modules. The Madeira River was recently
dammed by two large hydroelectric power plants in the Porto Velho region, and the modules
were installed along the banks of the river for monitoring the effects of flooding on biodiversity.
The data used in this study were collected prior to flooding (see details below). The region is
covered by primary and secondary rainforest under increasing threat of human occupation.
We sampled snakes by nocturnal active visual search, with two observers per plot, and
standard searching time of 1 hour per plot. In the Amazon, nocturnal search simultaneously
allows finding foraging nocturnal species, and resting diurnal species (field observation). To
increase the sampling effort and accuracy in species detection, we undertook four non-
consecutive surveys of each plot between 2007 and 2014. We show below that temporal
variation (e.g., caused by seasonal environmental variables) in assemblage structure is unlikely,
because species composition did not differ among the surveys. Snakes were collected under
RAN-ICMBio / IBAMA (Ministry of Environment, Government of Brazil) permanent license nº
13777-2/2008 in the name of Albertina Pimentel Lima (coordinator of the sampling expeditions).
The license includes ethical approval of all the procedures used.
Detection probabilities of snakes are usually very low (Steen, 2010; Fraga et al., 2014),
and they may bias the results by generating statistical artifacts such as the arc effect (Gauch,
1982), mainly because no species occurrence is shared between sampling units. To avoid
statistical artifacts in this study, we used modules as sampling units rather than plots. This results
in loss of degrees of freedom, but it increases the predictive power of the analysis because the
sampling units usually shared more than one species. All the analyses in this study are based on
26 species (Table S1), belonging to four families (Boidae, Colubridae, Dipsadidae and
Viperidae).
**Taxonomic β-diversity (TBD).** We represented TBD as the scores from a PCoA (Principal Coordinates Analysis) ordination of a matrix of Forbes’ similarity index (*Forbes, 1907*) on species presence/absence data between modules. We transformed Forbes’ similarities in dissimilarities between modules by 1 - Forbes. The Forbes index has been indicated as robust in the case of incomplete sampling (*Alroy, 2015*), which is common in studies of snakes in the Amazon due to the low detection probabilities of most species (*Fraga et al., 2014*). The PCoA was undertaken in the vegan package (*Oksanen et al., 2015*) in R (*R Development Core Team, 2015*).

**Functional β-diversity (FBD).** We constructed a trait matrix using ten continuous and discrete traits, measured or observed for adult individuals only. These were maximum total length, tail length proportional to body length, diameter of the eye proportional to head length, maximum size of offspring, discrete habitat (aquatic, arboreal, cryptozoic, fossorial, terrestrial), period of activity (diurnal, nocturnal), foraging mode (ambush, active), diet (birds, bird eggs, centipedes, earthworms, frogs, fish, lizards, mammals, snails, snakes, Squamata eggs, tadpoles), defensive behavior (ball posture, bite, caudal autotomy, cloacal discharge, constriction, enlarged head, flattened body, hidden head, inflated neck, liana imitation, shown mucosa, sound, strike, tail shaking, tail sting, venom, vomit) and reproductive mode (oviparous, viviparous). All the traits used have been described as ecologically relevant for snakes (literature compilation in *Burbrink & Myers, 2015*). Further details on functional traits may be found in Table S2.

The continuous traits were measured, and we used average values per species (see *Petchey & Gaston, 2006*). For the species for which we found less than five individuals, we supplemented our data with published data (*Beebe, 1946*; *Belluomini & Hoge, 1958*; *Duellman, 1978*; *Cunha & Nascimento, 1983*; *Dixon & Soini, 1986*; *Michaud & Dixon, 1989*; *Cunha &
Nascimento, 1993; Starace, 1998; Martins & Oliveira, 1999; Fraga et al., 2013a). We also obtained most of the data for discrete traits per species from the literature, and they were supplemented with field observations. The levels of most discrete traits are not mutually exclusive (e.g. species which feed on a variety of prey), so we coded discrete traits into independent binary traits as suggested by Petchey & Gaston (2007).

We used the trait matrix to estimate FBD using the dFBD function in the FD R-package (Laliberté & Legendre, 2010; Laliberté, Legendre & Shipley, 2014). This function calculates Gower distances between species, which is an index thought to be more appropriate when analyzing mixed continuous and discrete traits, although the results are often strongly correlated with Euclidean distances (Petchey & Gaston, 2007). The dFBD function transforms the Gower distance matrix by calculating square roots. This is important to avoid negative eigenvalues in the PCoA calculated from the distance matrix, which should be set in Euclidean space to avoid biased estimates of FD (Laliberté & Legendre, 2010). PCoA was used to obtain scores representing four different functional indices (Villéger, Mason & Mouillot, 2008), which are functional richness (FRic), functional evenness (FEve), functional divergence (FDiv) and functional dispersion (FDis). In this study, we represented FBD using FDis, because this index estimates functional β-diversity based on average distances to the centroid of multivariate dispersion (Anderson, 2006). FDis has been described as a β-diversity index which is not affected by species richness, it can handle any number and type of traits and it is little biased by outliers (Anderson, Ellingsen & McArdle, 2006).

We visually controlled the robustness of the FBD estimate by constructing a functional tree (Fig. S1) based on Gower pairwise distances between species, which was calculated in the vegan R-package (Oksanen et al., 2015). We undertook a hierarchical cluster analysis on the
Gower dissimilarity matrix to build an UPGMA functional tree, using the hclust function (argument average) in R.

**Phylogenetic β-diversity (PBD).** We estimated PBD based on a well-supported phylogenetic hypothesis proposed by *Pyron, Burbink & Wiens (2013)*. Phylogeny of Squamata reptiles was reconstructed by analyzing 12 concatenated genes (5 mtDNA and 7 nuclear) from more than 4,000 species. Levels of clade support were estimated by non-parametric Shimodaira-Hasegawa-Like implementation of the approximate likelihood-ratio test (further details in *Pyron, Burbink & Wiens, 2013*). We used the ape R-package (*Paradis, 2012*) to obtain a subtree composed of the species sampled in this study (Fig. S2).

We estimated PD using the phylosor function of the picante R-package (*Kembel et al., 2010*), which estimates fractions of branch lengths in a phylogenetic tree that are shared among communities (*Kembel et al., 2010*). The phylosor function returns a pairwise phylogenetic similarity matrix, which was converted to a pairwise distance matrix (1-phylosor matrix) and summarized by PCoA scores.

**Environmental gradients and inferential analysis.** We evaluated the influence of clay content in the soil on the diversity measures because this gradient affects primary production, which influences the overall trophic network (*Cintra et al., 2013*). Clay content was measured in a pooled 5g sample derived from six subsamples per plot, and we used averages per module. Technicians at the Laboratório Temático de Solos e Plantas (LTSP) of the Instituto Nacional de Pesquisas da Amazônia (INPA, Manaus) conducted the physical analyses following standard methods (*Embrapa, 2009*).

The Height Above the Nearest Drainage (HAND) algorithm estimates the depth to the water table, which represents a gradient of relative water potential, in which higher values...
indicate large gravitational potential and lower values may reflect soil waterlogging in the absence of drainage. Here we tested the effects of HAND (log-normalized) on snake diversity because vertical and horizontal distances from drainage are correlated in a micro-watershed scale (Schietti et al., 2013), and horizontal distance from drainage was previously identified as an important factor affecting snake assemblages through availability of physiological optimal or prey availability at assemblage (Fraga, Lima & Magnusson, 2011) and population (Fraga et al., 2013b) levels in Amazonia. We used percentage of tree cover (log-normalized) because this gradient potentially filters species by their adaptability to variation in habitat openness, considering the variation in factors such as availability of resting and nesting sites (Burger & Zappalorti, 1986), availability of prey and protection from predators (Webb & Shine, 1996), and light intensity (Pringle, Webb & Shine, 2003). The gradients HAND and tree cover were obtained from raster surfaces downloaded from the public repository Ambdata (Amaral et al., 2013; www.dpi.inpe.br/Ambdata).

We used temperature of the coldest month and precipitation of the wettest month because climate is often considered as a primary factor determining distribution of biodiversity at numerous spatial and temporal scales. Climate may limit species distribution through physiological filtering, especially in ectothermic animals such as snakes (Blain et al., 2009).

Climate data were obtained from raster surfaces downloaded from the Worldclim database (Fick & Hijmans, 2017). All raster surfaces used in this study have a resolution of 1 km, and values for each gradient were extracted by the raster R-package (Hijmans, 2015) using geographic coordinates per module.

Geographic distance has been found to be an important factor driving β-diversity for several vertebrates, because potentially carries unmeasured environmental variation (Qian &
Ricklefs, 2012). Our study area covers about 4,400 km², and therefore we expect spatial autocorrelation in the environmental data. To reduce the effects of geographic distance on the environmental gradients we calculated Euclidean distances between centroid coordinates per module, and reduced dimensionalities using the first axis from a PCoA. The coordinates were used in linear regressions given by $\text{gradient} = a + b(\text{geographic distance})$. Residuals from these models were used as independent variables in inferential models to quantity proportions of snake diversity that are explained by environmental variation. We used this approach because multiple-linear-regressions using raw environmental data as independent variables returned slight spatial autocorrelation in the residuals for TBD (Moran’s $I = -0.272$, $P = 0.06$), FBD (Moran’s $I = -0.320$, $P = 0.02$) and PBD (Moran’s $I = -0.261$, $P = 0.08$), which was negatively significant ($P < 0.05$) in 20–30% of ten geographic distance classes (Fig. S3).

To test for the effects of gradients on the diversity measures we used multiple-linear-regression models, that were built following the general formula $\text{diversity measure} = a + b_1(\text{clay content residuals}) + b_2(\text{HAND residuals}) + b_3(\text{tree cover residuals}) + b_4(\text{temperature residuals}) + b_5(\text{precipitation residuals})$. The gradients measured are given in different units, so we scaled them using the scale function in R (Becker, Chambers & Wilks, 1988). The residuals representing gradients showed little multicollinearity ($\leq 0.7$ in all cases). We have considered significant relationships at $P \leq 0.05$ after Bonferroni correction. Additional information (e.g., amplitude and average values) on the gradients can be found in the Table S3.

Alternatively, we used redundancy analysis (RDA) to test the effects of environmental gradients on raw distance matrices among sampling units, separately for each diversity measure. This approach was useful to verify the robustness of the results obtained by the multiple linear regressions, considering information loss by dimensionality compression through PCoA. We
constructed the models using taxonomic, functional and phylogenetic distance matrices as dependent variables, and environmental gradients as independent variables, which is equivalent to multiple-multivariate linear regression models (Legendre, Fortin & Borcard, 2015). We used a permutation test of significance (5000 randomizations) to decide between accept or reject null hypothesis.

We also compared the subsets of species that were found per plot in each of the four surveys (2007–2014), to control any effect of temporal variation on the diversity measures. We calculated Forbes pairwise dissimilarities among each survey on each plot, summarized the resulting matrix using PCoA (axis 1), and tested differences among surveys using ANOVA. We set a two-factors model – plot and survey – to decouple assemblage compositions between space and time. We found that differences in assemblage composition are associated with plots (P = 0.03), and plots and surveys interacting with each other (P = 0.01), but not with surveys alone (P = 0.26). This finding demonstrates that the patterns of spatial assemblage structure shown in this study have no bias of assemblages changing over time.

To quantify relationships between different diversity measures we used matrix regression with permutation test of significance (5000 randomizations). The models were set up with the pairwise distance matrices used to summarize TBD, FBD and PBD.

Results

The first axis of PCoA ordination captured 59% of the original variance in the raw data used to estimate TBD, and the second axis captured 39%. However, the variance captured by axis 1 was not related to the environmental variation quantified by the gradients tested (P > 0.25 in all cases). Therefore, we used axis 2 to represent TBD. PCoA axis 1 captured 100% of the
variance for FBD and 30% for PBD and were used as univariate versions of the diversity measures in the inferential models.

The multiple linear regression explained 61 percent (adjusted $R^2$) of the variance in TBD ($F_{5,15} = 7.31, P = 0.001$, residual standard error = 0.29). This finding was particularly associated to the effects of clay content ($P = 0.002$) and tree cover ($P = 0.034$) on $\beta$-diversity among the sampling modules. HAND, temperature and precipitation were not related to TBD ($P > 0.45$ in all cases). About 37 percent of the variance in FBD was explained by the multiple regression ($F_{5,15} = 3.42, P = 0.02$, residual standard error = 0.45), which was mainly due to the effects of HAND ($P = 0.023$) on dissimilarities between modules. Clay content, tree cover, temperature and precipitation were not related to FBD ($P > 0.85$ in all cases). About 32 percent of the variance in PBD was explained by the gradients ($F_{5,15} = 2.87, P = 0.05$, residual standard error = 0.11), but only HAND ($P = 0.034$) contributed to the model. The other gradients tested were not related to PBD ($P = 1$ in all cases). All the multiple regression models returned residuals that did not differ statistically from a normal distribution (Shapiro-Wilk $W > 0.92$, $P > 0.12$ in all cases).

A complete summary of the multiple-regression models can be found in Table 1 and the partial regressions from each model are presented in Fig. 2.

The RDA models significantly captured 32% of the constrained variance to TBD ($F_{5-15} = 1.42, P = 0.05$), 43% to FBD ($F_{5-15} = 2.27, P = 0.04$) and 36% to PBD ($F_{5-15} = 1.71, P = 0.03$). In general, the results were consistent with the multiple regressions (Table S4). However, tree cover did not significantly explain TBD ($P = 0.24$).

All diversity measures were positively related to each other (Fig. 3). However, TBD was more strongly related to PBD ($r^2 = 0.67, P < 0.0001$) compare to the relationships between TBD and FBD ($r^2 = 0.13, P < 0.0001$), and between FBD and PBD ($r^2 = 0.23, P < 0.0001$).
Discussion

Our data indicate that environmental gradients affect snake co-occurrence in central-southern Amazonia, which results in levels of β-diversity identified by taxonomic, functional and phylogenetic dissimilarities among modules. These findings are consistent with temperate snake assemblages, which may be structured by phylogenetic species variability and trait variability (Burbrink & Myers, 2015). Furthermore, our data corroborate a previous study in Brazil, which found variation in phylogenetic and phenotypic compositions of snake assemblages associated with environmental gradients (Cavalheri, Both & Martins, 2015). However, the variation found in that study was primarily due to differences between forested and open biomes, which is expected to be pronounced. In this study, we show that variation in snake assemblages along environmental gradients is consistent even within biomes, where structural differences between sites are often subtle.

Environmental gradients affecting species composition have been found in many groups of organisms in the Amazon, such as frogs (Ribeiro-Jr, Lima & Magnusson, 2012, Rojas-Ahumada, Landeiro & Menin, 2012), understory birds (Bueno et al., 2012; Menger et al., 2017), plants (Drucker, Costa & Magnusson, 2008; Moulatlet et al., 2014) and snakes (Fraga, Lima & Magnusson, 2011). In general, it is expected that species occupy portions of gradients in a way to optimize the balance between dispersal capacity, physiological needs and availability of resources (Laliberté, Legendre & Shipley, 2014). However, our data indicate that TBD is affected by different sets of environmental gradients compared to FBD or PBD, which suggests that patterns of spatial structure in snake assemblages in the Amazon may not be characterized by a single diversity measure. In fact, choosing environmental gradients as predictors in species-
habitat association models is not a trivial task, although it is an effective approach to evaluate conservation issues such as environmental legislation (Fraga et al., 2011; Bueno et al., 2012).

We found that clay content in the soil predicted snake community assembly based on taxonomic dissimilarities among modules. Associations between soil texture and patterns of vertebrate community structure are often assumed as indirectly causal, because soil texture affects many factors driving regional species occurrence, such as vegetation density (Woinarski, Fisher & Milne, 1999) and distance from streams (Bueno et al., 2012). The environmental heterogeneity in the Amazon rainforests include soil texture gradients from poorly-drained, seasonally-flooded sandy soils (Cintra et al., 2013), in which plants find poor substrate for rooting (Quesada et al., 2012), to well-drained soils that support older well-developed forests (Castilho et al., 2006; Emilio et al., 2013). Therefore, variation in soil texture across landscapes generates high β-diversity via suitability of the conditions for dispersal and colonization, which has been found in plant (Costa, Magnusson & Luizão, 2005; Costa et al., 2008), invertebrate (Franklin, Magnusson & Luizão, 2005) and vertebrate (Woinarski, Fisher & Milne, 1999; Bueno et al., 2012; this study) assemblages.

We showed that HAND is related to snake assemblages based on functional traits and phylogeny. Distance above the drainage has been identified as an important factor structuring plant (Drucker, Costa & Magnusson, 2008) and animal (Fraga, Lima & Magnusson, 2011; Bueno et al., 2012; Rojas-Ahumada, Landeiro & Menin, 2012) assemblages in the Amazon, and riparian zones may be biologically distinct from adjacent areas (Sabo et al., 2005). Despite the fact that most Amazonian snakes can cross different habitats, distance from drainage may influences β-diversity (Fraga, Lima & Magnusson, 2011) and structures populations in terms of variation in density and body size (Fraga et al., 2013b). In this study, we found that HAND
affects FBD mainly because lower distances from drainage favor assemblages composed by arboreal (e.g., *Imantodes lentiferus*) and aquatic species (e.g., *Helicops angulatus*). Lower values of HAND are often associated with seasonal surface waterlogging (*Schietti et al., 2013*), which may complicate fossorial and terrestrial lifestyles. Therefore, variation in HAND across the landscape generates regional dispersal corridors that may be more suitable for some species than others, which could affect FBD and PBD.

We found inconsistency between the effects of tree cover on TBD estimated by multiple regression and RDA. Different results may reflect different levels of statistical sensitivity of the models to the data structure, or unidentified sampling bias. Therefore, we assume tree cover in this study as a probable filter to snake regional co-occurrence. Variation in tree cover along continuous landscapes generates mosaics of more and less suitable habitats for different subsets of species, through factors that directly affects species biology. These include availability (*Lindenmayer et al., 1991*) and non-random selection of nesting and resting sites (*Burger & Zappalorti, 1986*; *Webb & Shine, 1996*), thermo-regulatory requirements, availability of prey, scent chemical trails from potential prey (e.g., small mammals), protection from predators (*Webb & Shine, 1996*), variation in light intensity, air and ground temperatures and wind speed (*Pringle, Webb & Shine, 2003*). Furthermore, variation in tree cover at wider spatial scales (e.g., biomes) may define regional subsets of species according to their morphological adaptation to use different plant strata (*Cavalheri, Both & Martins, 2015*). This finding is relevant for conservation, because maintenance of regional assemblages depends on protecting large areas, which contain wide amplitudes of tree-cover gradients.

Changes in phylogenetic (PBD) composition among plots were proportional to changes in the snake taxonomic identities (TBD). Spatial congruence between estimates of phylogenetic and
taxonomic $\beta$-diversity has been suggested as often higher compared to $\alpha$-diversity measures (Devictor et al., 2010; Bernard-Verdier et al., 2013; Arnan, Cerdá & Retana, 2016), especially in cases of niche conservatism (Wiens & Graham, 2005). The tendency of species to retain ancestral characteristics along generations results in local or regional assemblages structured by environmental filtering, despite different environmental gradients may affect different diversity measures (Webb et al., 2002). Additionally, the positive correlations between diversity measures show that TBD and FBD efficiently captured a phylogenetic signal, which suggests assemblages evolutionary structured by Brownian motion, in which species change mainly through genetic drift and natural selection randomly directed (Losos, 2008). Concerning the positive relation between FBD and PBD, we found that environmental gradients drive co-occurrence of species that are simultaneously phylogenetically related and have similar ecological requirements (Keddy, 1992; Myers & Harms, 2009), and one measure may be used as a proxy to the other. Contrarily, TBD was not a good proxy for FBD, because large proportions of data were poorly fitted between both diversity measures. Ultimately, combinations of TBD and FBD should be part of studies on community ecology, because they give integrative approaches that reveal taxonomic, ecological and evolutionary forces acting on community structuring, which is very useful for conservation (Devictor et al., 2010).

From the point of view of conservation, the positive relationships between diversity measures directs the focus of biodiversity monitoring programs and reserve planning to cover higher levels of phylogenetic diversity, because this measure reflects the maintenance of ecosystem processes operating over long timescales (Cadotte, Carscadden & Mirotchnick, 2011). Functional diversity is estimated based on sets of traits that reflect environmental tolerances and requirements, which in turn determine where species can live (Lavorel et al., 1997) and interact with each other in
assemblages (Davies et al., 2007). Therefore, loss of evolutionarily distinct species may result in irreversible loss of functions for ecosystems (Bracken & Low, 2012). However, at larger scales it may be difficult to decide which diversity measure should be prioritized in conservation, and a multi-dimensional approach may be more appropriate (Devictor et al., 2010), despite greater difficulties of interpretation.

Our findings are unlikely to be biased by the spatial distribution of sampling units (see González-Caro et al., 2012), because the RAPELD system provides regular distribution of plots across the landscape regardless of logistical issues (Magnusson et al., 2013). However, snakes usually have low detection probabilities (Steen, 2010; Fraga et al., 2014), which have been estimated at less than 10 percent for multiple surveys of many Amazonian species in RAPELD plots (Fraga et al., 2014). Low detection probabilities often cause false absences of species from plots, and this may generate misinterpretation of how species respond to landscape change (Gu & Swihart, 2003). We are unable to totally discount effects of low detectability on our results. However, the strong relationships between diversity measures and environmental gradients showed that a combination of high sampling effort, different methods used to quantify biodiversity and the use of an appropriate multivariate distance measure may considerably reduce the effect of false absences and return reliable results.

Conclusions

We used an unprecedented standardized sampling effort to show that environmental heterogeneity is associated with β-diversity in Amazonian-forest snakes. Positive correlations between β-diversity measures estimated show that PBD alone may be sufficient to investigate spatial structure in Amazonian snake assemblages under taxonomic, functional and phylogenetic
perspectives. However, TBD response to different environmental gradients suggests that testing
the effects of a set of environmental gradients on at least two β-diversity measures can generate
deeper understanding of factors causing spatial community assembly. This finding highlights the
efficiency of using multi-dimensional approaches to quantify biodiversity in community-level
conservation status assessments and decision-making on natural resources management.

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References

Alroy J. 2015. A new twist on a very old binary similarity coefficient. Ecology 96:575–586.
Amaral S, Costa CB, Arasato LS, Ximenes AC, Rennó CD. 2013. AMBDATA: Variáveis
ambientais para Modelos de Distribuição de Espécies (MDEs). Anais do XVI Simpósio
Brasileiro de Sensoriamento Remoto (SBSR) 16:6930–6937.
Anderson MJ. 2006. Distance-based tests for homogeneity of multivariate dispersions.
Biometrics 62:245–253.
Anderson, MJ, Ellingsen KE, McArdle BH. 2006. Multivariate dispersion as a measure of beta
diversity. Ecology Letters. 9:683–693.
Arnan X, Cerdá X, Retana J. 2017. Relationships among taxonomic, functional, and
phylogenetic ant diversity across the biogeographic regions in Europe. Ecography 40:448–
457.
Becker, RA, Chambers JM, Wilks AR. 1988. The new S language: a programming environment for data analysis and graphics. Wadsworth & Brooks/Cole.

Beebe W. 1946. Field notes on the snakes of Kartabo, British Guiana, and Caripito, Venezuela. Zoologica 31:11–52.

Bellomini, HE, Hoge AR. 1958. Contribuição à biologia de Eunectes murinus (Linnaeus 1758) (Serpentes). Observações sobre hábitos alimentares de “sucuris” em cativeiro. Memórias do Instituto Butantan 28:207–216.

Bernard-Verdier M, Flores O, Navas M-L, Garnier E. 2013. Partitioning phylogenetic and functional diversity into alpha and beta components along an environmental gradient in a Mediterranean rangeland. Journal of Vegetation Science 25(5):877–889.

Bernarde PS, Albuquerque S, Barros TS, Turci LCB. 2012. Serpentes do estado de Rondônia. Biota Neotropica 12:1–29.

Blain H-A, Bailon S, Cuenca-Bescós G, Arsuaga JL, Castro JMB, Carbonell E. 2009. Long-term climate record inferred from early-middle Pleistocene amphibian and squamate reptile assemblages at the Gran Dolina Cave, Atapuerca, Spain. Journal of Human Evolution 56:55–65.

Bracken MES, Low NHN. 2012. Realistic of rare species disproportionately impact higher trophic levels. Ecology Letters 15:461–467.

Bueno AS, Bruno RS, Pimentel TP, Sanaiotti TM, Magnusson WE. 2012. The width of riparian habitats for understory birds in an Amazonian forest. Ecological Applications 22:722–734.

Burbrink FT, Myers EA. 2015. Both traits and phylogenetic history influence community structure in snakes over steep environmental gradients. Ecography 38:001–013.
Burger J, Zappalorti RT. 1986. Nest site selection by Pine Snakes, *Pituophis melanoleucus*, in the New Jersey Pine Barrens. *Copeia* 1986:116–121.

Cadotte MW, Cavenders-Bares J, Tilman D, Oakley TH. 2009. Using phylogenetic, functional and trait diversity to understand patterns of plant community productivity. *PloS ONE* 4:e5695.

Cadotte MW, Carscadden K, Mirotchnick N. 2011. Beyond species: functional diversity and the maintenance of ecological processes and services. *Journal of Applied Ecology* 48:1079–1087.

Castilho CV, Magnusson WE, De-Araújo RNO, Luizão RCC, Luizão FJ, Lima AP, Higuchi N, De-Castilho CV. 2006. Variation in aboveground tree live biomass in a central Amazonian forest: effects of soil and topography. *Forest Ecology and Management* 234:85–96.

Cavalheri H, Both C, Martins M. 2015. Filtering and spatial processes in structuring communities: the case of Neotropical snake communities. *PLoS ONE* 10:e0127959.

Cintra BBL, Schietti J, Emillio T, Martins D, Moulatlet G, Souza P, Levis C, Quesada CA, Schöngart J. 2013. Soil physical restrictions and hydrology regulate stand age and wood biomass turnover rates of Purus-Madeira interfluvial wetlands in Amazonia. *Biogeosciences* 10:7759–7774.

Costa FRC, Magnusson WE, Luizão RC. 2005. Mesoscale distribution of Amazonian understory herbs in relation to topography, soil and watersheds. *Journal of Ecology* 93:863–878.
Costa FRC, Guillaumet JL, Lima AP, Pereira OS. 2008. Gradients within gradients: the mesoscale distribution patterns of palms in a central Amazonian forest. *Journal of Vegetation Science* 20:69–78.

Cunha OR, Nascimento FP. 1983. Ofídios da Amazônia XIX - As espécies de Oxyrhopus Wagler, com uma subspécie nova, e Pseudoboa Schneider, na Amazônia oriental e Maranhão. (Ophidia, Colubridae). *Boletim do Museu Paraense Emílio Goeldi, série Zoologia* 122:1–42.

Cunha OR, Nascimento FP. 1993. Ofídios da Amazônia: As cobras da região leste do Pará. *Boletim do Museu Paraense Emílio Goeldi, série Zoologia* 9:1–191.

Davies TJ, Meiri S, Barraclough TG, Gittleman JL. 2007. Species co-existence and character divergence across carnivores. *Ecology Letters* 10:146–152.

Dehling DM, Fritz SA, Töpfer T, Päckert M, Estler P, Böhning-Gaese K, Schleuning M. 2014. Functional and phylogenetic diversity and assemblage structure of frugivorous birds along an elevational gradient in the tropical Andes. *Ecography* 37:1047–1055.

Devictor V, Mouillot D, Meynard C, Jiguet F, Thuiller W, Mouquet N. 2010. Spatial mismatch and congruence between taxonomic, phylogenetic and functional diversity: the need for integrative conservation strategies in a changing world. *Ecology Letters* 13:1030–1040.

Dias-Terceiro RG, Kaefer IL, Fraga R, Araújo MC, Simões PI, Lima AP. 2015. A matter of scale: historical and environmental factors structure anuran assemblages from the upper Madeira River, Amazonia. *Biotropica* 47:259–266.

Diaz S, Cabido M. 2001. Vive la difference: plant functional diversity matters to ecosystem processes. *Trends in Ecology and Evolution* 16:646–655.
Dixon JR, Soini P. 1986. The reptiles of the upper Amazon Basin, Iquitos region, Peru. Milwaukee: Milwaukee Public Museum.

Drucker DP, Costa F, Magnusson WE. 2008. How wide is the riparian zone of small streams in tropical forests? A test with terrestrial herbs. *Journal of Tropical Ecology* 24:65–74.

Duellman WE. 1978. The biology of an equatorial herpetofauna in Amazonian Ecuador. *Miscellaneous Publications, Museum of Natural History, University of Kansas* 65:1–352.

Emilio T, Quesada CAN, Costa F, Magnusson WE, Schietti J, Feldpausch TR, Brienen RJW, Baker TR, Chave J, Álvarez E, Araújo A, Bánki O, Castilho CV, Honório C, Euridice N, Killeen TJ, Malhi Y, Mendoza EMO, Monteagudo A, Neill D, Parada GA, Peña-Cruz A, Ramirez-Angulo H, Schwarz M, Silveira M, Ter SH, Terborgh JW, Thomas R, Lezama AR, Vilanova E, Phillips OL. 2013. Soil physical conditions limit palm and tree basal area in Amazonian forests. *Plant Ecology & Diversity* 8:229-241.

Empresa Brasileira de Pesquisa Agropecuária (Embrapa). 2009. *Manual de análises químicas de solos, plantas e fertilizantes*. Brasília: Embrapa Informação Tecnológica.

Fearnside PM. 2014. Impacts of Brazil’s Madeira river dams: unlearned lessons for hydroelectric development in Amazonia. *Environmental Science & Policy* 38:164–172.

Fearnside PM, Graça PMLA. 2006. BR-319: Brazil’s Manaus-Porto Velho highway and the potential impact of linking the arc of deforestation to central Amazonia. *Environmental Management* 38:705–16.

Fick SE, Hijmans RJ. 2017. Worldclim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* 37(12):4302–4315.
Flynn DFB, Mirotchnick N, Jain M, Palmer MI, Naeem S. 2011. Functional and phylogenetic diversity as predictors of biodiversity-ecosystem function relationships. *Ecology* 92:1573–1581.

Forbes SA. 1907. On the local distribution of certain Illinois fishes: an essay in statistical ecology. *Bulletin of the Illinois State Laboratory of Natural History* 7:272–303.

Fraga R, Lima AP, Magnusson WE. 2011. Mesoscale spatial ecology in a tropical snake assemblage: the width of riparian corridors in central Amazon. *Herpetological Journal* 21:51–57.

Fraga R, Lima AP, Prudente ALC, Magnusson WE. 2013a. *Guide to the snakes of the Manaus region – central Amazonia*. Manaus: INPA.

Fraga R, Magnusson WE, Abrahão CR, Sanaiotti T, Lima AP. 2013b. Habitat selection by *Bothrops atrox* (Serpentes: Viperidae) in central Amazonia, Brazil. *Copeia* 2013:684–690.

Fraga R, Stow AJ, Magnusson WE, Lima AP. 2014. The costs of evaluating species densities and composition of snakes to assess development impacts in Amazonia. *PloS ONE* 9:e105453.

Fraga R, Lima AP, Magnusson WE, Ferrão M, Stow AJ. 2017. Contrasting patterns of gene flow for Amazonian snakes that actively forage and those that wait in ambush. *Journal of Heredity* 108:524–534.

Franklin E, Magnusson WE, Luizão FJ. 2005. Relative effects of biotic and abiotic factors on the composition of soil invertebrate communities in an Amazonian savanna. *Applied Soil Ecology* 29:259–273.

Gauch HG. 1982. *Multivariate analysis in community ecology*. Cambridge: Cambridge University Press.
Gibbons JW, Scott DE, Ryan TJ, Buhlmann KA, Tuberville TD, Metts BS, Greene JL, Mills T, Leiden Y, Poppy S, Winne CT. 2000. The global decline of reptiles, déjà vu amphibians. *BioScience* 50:653–666.

González-Caro S, Parra JL, Graham CH, McGuire JA, Cadena CD. 2012. Sensitivity of metrics of phylogenetic structure to scale, source of data and species pool of hummingbird assemblages along elevational gradients. *PloS ONE* 7:e35472.

Grass I, Brandl R, Botzat A, Neuschulz EL, Farwig N. 2015. Contrasting taxonomic and phylogenetic diversity responses to forest modifications: comparisons of taxa and successive plant life stages in South African scarp forest. *PloS ONE* 10:e0118722.

Gu W, Swihart RK. 2003. Absent or undetected? Effects of non-detection of species occurrence on wildlife-habitats models. *Biological Conservation* 116:195–203.

Hangartner S, Laurila A, Räsänen K. 2012. Adaptive divergence in moor frog (*Rana arvalis*) populations along an acidification gradient: inferences from QST – FST correlations. *Evolution* 66:867–881.

Hijmans RJ. 2015. raster: Geographic Data Analysis and Modeling. R package version 2.3-40. Available at http://CRAN.R-project.org/package=raster (accessed on 10 April 2018).

Hoorn C, Wesselingh FP, ter Steege H, Bermudez MA, Moura A, Sevink J, Sanmartín I, Sanchez-Meseguer A, Anderson CL, Figueiredo JP, Jaramillo C, Riff D, Negri FR, Hooghiemstra H, Lundberg J, Stadler T, Särkinen T, Antonelli A. 2010. Amazonian through time: Andean uplift, climate change, landscape evolution, and biodiversity. *Science* 330:927–931.

Jetz W, Fine PVA. 2012. Global gradients in vertebrate diversity predicted by historical area-productivity dynamics and contemporary environment. *PLOS Biology* 10(3):e1001292.
Keddy PA. 1992. Assembly and response rules: two goals for predictive community ecology. *Journal of Vegetation Science* 3:157–164.

Kembel SW, Cowan PD, Helmus MR, Cornwell, Morlon H, Ackerly DD, Blomberg SP, Webb CO. 2010. Picante: R tools for integrating phylogenies and ecology. *Bioinformatics* 26:1463–1464.

Kinnup VF, Magnusson WE. 2005. Spatial patterns in the understory shrub genus *Psychotria* in central Amazonia: effects of distance and topography. *Journal of Tropical Ecology* 21:1–12.

Laliberté E, Legendre P. 2010. A distance-based framework for measuring functional diversity from multiple traits. *Ecology* 91:299–305.

Laliberté E, Legendre P, Shipley B. 2014. FD: measuring functional diversity from multiple traits, and other tools for functional ecology. R package version 1.0-12. Available at http://CRAN.R-project.org/web/packages/FD (accessed on 10 April 2018).

Lavorel S, McIntyre S, Landsberg J, Forbes TDA. 1997. Plant functional classifications: from general groups to specific groups based on response to disturbance. *Trends in Ecology and Evolution* 12:474–478.

Legendre P, Fortin M-J, Borcard D. 2015. Should the Mantel test be used in spatial analysis? *Methods in Ecology and Evolution* 6:1239–1247.

Lindenmayer DB, Cunninham RB, Tanton MT, Smith AP. 1991. The conservation of arboreal marsupials in the montane ash forest of the central highlands of Victoria, south-east Australia, III. Models of the habitat requirements of Leadbeater’s possum *Gymnobelideus leadbeateri* and the diversity and abundance of arboreal marsupials. *Biological Conservation* 56:295–315.
Losos, J. 2008. Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. *Ecology Letters* 11:995–1007.

Magnusson WE, Lima AP, Luizão R, Luizão F, Costa FRC, Castilho CV, Kinupp VF. 2005. RAPELD: a modification of the Gentry method for biodiversity surveys in long-term ecological research sites. *Biota Neotropica* 5(2):1–6.

Magnusson WE, Braga-Neto R, Pezzini F, Baccaro F, Bergallo H, Penha J, Rodrigues D, Verdade LM, Lima A, Albernaz AL, Hero J-M, Lawson B, Castilho C, Drucker D, Franklin E, Mendonça F, Costa F, Galdino G, Castley G, Zuanon J, Vale J, Luizão L, Luizão R, Cintra R, Barbosa RI, Lisboa A, Koblitz RV, Cunha CN, Pontes ARM. 2013. *Biodiversity and Integrated Environmental Monitoring*. Manaus: Áttema.

Mariac C, Jehin L, Saïdou AA, Thuillet AC, Couderc M, Sire P, Judgé H, Adam H, Bezançon G, Pham JL, Vigouroux Y. 2011. Genetics basis of pearl millet adaptation along an environmental gradient investigated by a combination of genome scan and association mapping. *Molecular Ecology* 20:80–91.

Martins M, Oliveira ME. 1999. Natural history of snakes in forests of the Manaus region, Central Amazonia, Brazil. *Herpetological Natural History* 6:78–150.

McGill BJ, Enquist BJ, Weiher E, Westoby M. 2006. Rebuilding community ecology from functional traits. *Trends in Ecology and Evolution* 21:178–185.

Menger J, Magnusson WE, Anderson MJ, Schlegel M, Pe’er G, Henle K. 2017. Environmental characteristics drive variation in Amazonian understory bird assemblages. *PLoS ONE* 12:e0171540.
Beyond taxonomic diversity patterns: how do $\alpha$, $\beta$ and $\gamma$ components of bird functional and phylogenetic diversity respond to environmental gradients across France? Global Ecology and Biogeography 20:893–903.

Michaud EJ, and Dixon JR. 1989. Prey items of 20 species of the Neotropical colubrid snake genus Liophis. Herpetological Review 20:39–41.

Milcu A, Allan E, Roscher C, Jenkins T, Meyer ST, Flynn D, Bessler H, Buscot F, Engels C, Gubsch M, König S, Lipowsky A, Loranger J, Renker C, Scherber C, Schmid B, Thébault E, Wubet T, Weisser WW, Scheu S, Eisenhauer N. 2013. Functionally and phylogenetically diverse plant communities key to soil biota. Ecology 94:1878–1885.

Morlon H, Schwilk DW, Bryant JA, Marquet PA, Rebelo AG, Tauss C, Bohannan BJM, Green JL. 2011. Spatial patterns of phylogenetic diversity. Ecology Letters 14(2):141–149.

Moulatlet GM, Costa FRC, Rennó CD, Emílio T, Schietti J. 2014. Local hydrological conditions explain floristic composition in lowland Amazonian forests. Biotropica 46:395–403.

Myers JA, Harms KE. 2009. Seed arrival, ecological filters, and plant species richness: a meta-analysis. Ecology Letters 12:1250–1260.

Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR, O’Hara RB, Simpson GL, Solymos P, Stevens MHH, Szoecs E, Wagner H. 2015. Vegan: Community Ecology Package. Available at http://cran.r-project.org (accessed on 10 April 2018).

Petchey OL, Gaston KJ. 2002. Functional diversity (FD), species richness and community composition. Ecology Letters 5:402–411.
Petchey OL, Gaston KJ. 2006. Functional diversity: back to basics and looking forward. *Ecology Letters* 9:741–758.

Petchey OL, Gaston KJ. 2007. Dendrograms and measuring functional diversity. *Oikos* 116:1422–1426.

Pressey RL, Humphries CJ, Margules CR, Vane-Wright RI, Williams PH. 1993. Beyond opportunism: key principles for systematic reserve selection. *Trends in Ecology and Evolution* 8:124–128.

Pringle RM, Webb JK, Shine R. 2003. Canopy structure, microclimate, and habitat selection by a nocturnal snake, *Hoplocephalus bungaroides*. *Ecology* 84:2668–2679.

Pyron RA, Burbink FT, Wiens JJ. 2013. A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC Evolutionary Biology* 13:1–53.

Qian H, Ricklefs RE. 2012. Disentangling the effects of geographic distance and environmental dissimilarities on global patterns of species turnover. *Global Ecology and Biogeography* 21:341–351.

Quesada CA, Phillips OL, Schwarz M, Czimczik CI, Baker TR, Patiño S, Fyllas NM, Hodnett MG, Herrera R, Almeida S, Alvarez DE, Arneth A, Arroyo L, Chao KJ, Dezzeo N, Erwin T, Di-Fiore A, Higuchi N, Coronado EH, Jimenez EM, Killeen T, Lezama AT, Lloyd G, López-González G, Luizão FJ, Malhi Y, Monteagudo A, Neill DA, Vargas PN, Paiva R, Peacock J, Peñuela MC, Cruz AP, Pitman N, Filho NP, Pietro A, Ramírez H, Rudas A, Salomão R, Santos AJB, Schmerler J, Silva N, Silveira M, Vásquez R, Vieira I, Terborgh J, Lloyd J. 2012. Basin-wide variations in Amazon forest structure and function are mediated by both soils and climate. *Biogeosciences* 9:2203–2246.
R Development Core Team. 2015. R: a language and environment for statistical computing. Vienna: R Foundation for Statistical Computing. Available at http://www.R-project.org (accessed on April 10 2018).

Ribeiro-Jr JW, Lima AP, Magnusson WE. 2012. The effect of riparian zones on species diversity of frogs in Amazonian forests. Copeia 2012:375–381.

Rojas-Ahumada DP, Landeiro VL, Menin M. 2012. Role of environmental and spatial processes in structuring anuran communities across a tropical rainforest. Austral Ecology 37:865–873.

Sabo JL, Sponseller R, Dixon M, Gade K, Harms T, Heffernan J, Jani A, Katz G, Soykan C, Watts J, Welter J. 2005. Riparian zones increase regional species richness by harboring different, not more, species. Ecology 86:56–65.

Schietti J, Emilio T, Rennó CD, Drucker DP, Costa FRC, Nogueira A, Baccaro FB, Figueiredo F, Castilho CV, Kinupp V, Guillaumet JL, Garcia ARM, Lima AP, Magnusson WE. 2013. Vertical distance from drainage drives floristic composition changes in an Amazonian rainforest. Plant Ecology & Diversity 2:241–253.

Soares-Filho BS, Nepstad DC, Curran LM, Cerqueira GC, Garcia RA, Ramos CA, Voll E, McDonald A, Lefebvre P, Schlesinger P. 2006. Modelling conservation in the Amazon basin. Nature 440:520–523.

Starace F. 1998. Guide des Serpents at Amphisbènes de Guiane. Matoury: Ibis Rouge Éditions.

Steen D. 2010. Snakes in the grass: secretive natural histories defy both conventional and progressive statistics. Herpetological Conservation & Biology 5:183–188.

Tilman D. 2001. Functional diversity. In: Levin SA, ed. Encyclopedia of Biodiversity. New York: Academic Press, 109–120.
Venables WN, Ripley BD. 2002. *Modern applied statistics with S*. 4th edition. New York: Springer.

Villéger S, Mason WH, Mouillot D. 2008. New multidimensional functional diversity indices for a multifaceted framework in functional ecology. *Ecology* 89:2290–2301.

Webb JK, Shine R. 1996. Out on a limb: conservation implications of tree-hollow use by a threatened snake species (*Hoplocephalus bungaroides*: Serpentes, Elapidae). *Biological Conservation* 81:21–33.

Weinstein BG, Tinoco B, Parra JL, Brown LM, McGuire JA, Stiles FG, Graham CH. 2014. Taxonomic, phylogenetic, and trait beta diversity in South American Hummingbirds. *The American Naturalist* 184(2):211–224.

Wiens JJ, Graham CH. 2005. Niche conservatism: integrating evolution, ecology, and conservation biology. *Annual Review of Ecology, Evolution, and Systematics* 36:519–539.

Winemiller KO, López-Fernández H, Taphorn DC, Nico LG, Duque AB. 2008. Fish assemblages of the Casiquiare River, a corridor and zoogeographical filter for dispersal between the Orinoco and the Amazon basins. *Journal of Biogeography* 35:1551–1563.

Woinarski JCZ, Fisher A, Milne D. 1999. Distribution patterns of vertebrates in relation to an extensive rainfall gradient and variation in soil texture in the tropical savannas of the Northern Territory, Australia. *Journal of Tropical Ecology* 15:381–398.
Summary of statistical coefficients from multiple linear regressions testing the effects of environmental gradients on estimates of snake β-diversity in Brazilian Amazonia.

TBD = taxonomic β-diversity, FBD = functional β-diversity, PBD = phylogenetic β-diversity. HAND = Height Above the Nearest Drainage. Bolded P-values are statistically significant after Bonferroni correction. $R^2$ values are adjusted to the number of sampling units. SE = standard error.
| Coefficient                      | TBD     | FBD     | PBD     |
|----------------------------------|---------|---------|---------|
| $R^2$                            | 0.612   | 0.377   | 0.319   |
| $P$                              | **0.001** | **0.02** | **0.05** |
| Clay content                     | SE      | $t$     | $P$     |
|                                 | 0.007   | -4.431  | **0.002** |
|                                 | 0.01    | -0.499  | 1       |
|                                 | 0.002   | 1.233   | 1       |
| HAND                             | SE      | $t$     | $P$     |
|                                 | 0.08    | -0.045  | **0.023** |
|                                 | 0.127   | 3.325   | 0.033   |
|                                 | 0.033   | 3.142   |         |
| Tree cover                       | SE      | $t$     | $P$     |
|                                 | 0.16    | 3.929   | **0.007** |
|                                 | 0.247   | -1.435  | 0.859   |
|                                 | 0.064   | 0.661   |         |
| Temperature of the coldest month | SE      | $t$     | $P$     |
|                                 | 0.23    | -0.34   | 1       |
|                                 | 0.035   | -0.044  | 1       |
|                                 | 0.009   | -0.574  |         |
| Precipitation of the wettest month | SE   | $t$     | $P$     |
|                                 | 0.003   | -1.804  | 0.457   |
|                                 | 0.005   | 1.382   | 0.936   |
|                                 | 0.001   | -0.589  | 1       |
Figure 1

Map of the study area in Brazilian Amazonia showing plots where snake assemblages were sampled.

Black circles are 5 km$^2$ RAPELD sampling modules (see definition of RAPELD in the methods). The numbers can be used to check the species found in each module in Table S1. (A) Ducke Reserve, Manaus. (B) Federal highway BR-319, Purus-Madeira interfluve. (C) Upper Madeira River, Rondônia.
Figure 2

Relationships (partials from multiple linear regressions) between different measures of snake β-diversity and environmental gradients in central-southwestern Brazilian Amazonia.

(A) TBD = taxonomic β-diversity, (B) FBD = functional β-diversity, (C) PBD = phylogenetic β-diversity. HAND = Height Above the Nearest Drainage. Environmental gradients are shown as residuals from linear regressions used to reduce the effects of geographic distance on the environmental heterogeneity measured.
Figure 3

Relationships between dissimilarity matrices used to summarize different estimates of snake β-diversity in central-southwestern Brazilian Amazonia.

TBD = taxonomic β-diversity, FBD = functional β-diversity, PBD = phylogenetic β-diversity. (A) Relationship between TBD and FBD. (B) Relationship between TBD and PBD. (C) Relationship between FBD and PBD.