Abstract: A series of fourteen 2-aryl-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-ones was prepared at room temperature by T3P-mediated cyclization of N-phenyl-C-aryl imines with thionicotinic acid, two difficult substrates. The reactions were operationally simple, did not require specialized equipment or anhydrous solvents, could be performed as either two or three component reactions, and gave moderate–good yields as high as 63%. This provides ready access to N-phenyl compounds in this family, which have been generally difficult to prepare. As part of the study, the first crystal structure of neutral thionicotinic acid is also reported, and showed the molecule to be in the form of the thione tautomer. Additionally, the synthesized compounds were tested against the kinetoplastid parasite Trypanosoma brucei, the causative agent of Human African Sleeping Sickness. Screening at 50 \( \mu \text{M} \) concentration showed that five of the compounds strongly inhibited growth and killed parasites.

Keywords: pyrido(thiazinone); T3P; imine; N-aryl; thionicotinic acid; parasites; \( T. \) brucei

1. Introduction

The 2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one scaffold (Figure 1) features a pyridine ring fused to a thiazine ring. Compounds with this scaffold have previously shown...
anticancer [1], antibacterial [2], and glycosidase inhibitory bioactivity [3]. A compound previously reported by us, 2,3-diphenyl-2,3-dihydro-4H-pyrido[3,2-\(e\)]1,3thiazin-4-one 1j (Scheme 1, Table 1, \(R = \text{H}\)) [4,5], inhibited growth of two kinetoplastid parasites, *Trypanosoma brucei* and *Crithidia fasciculata* [6]. The effect was particularly striking for *T. brucei*, which causes disease in humans and domesticated livestock. Parasite cell growth was inhibited at a specific stage of the cell cycle, indicating that the completion of cytokinesis was delayed or prevented in treated cells. *T. brucei* parasites treated with this compound also had delayed mitochondrial division and showed signs of an endocytosis defect, both of which could disrupt cytokinesis. Although the IC50s were in the micromolar range, precluding further investigation of 1j as an antikinetoplastid therapeutic, these parasites represent a powerful system with which to discover the biological target(s) of this and related compounds, having a great deal of conservation with other eukaryotes and a variety of tools for genetic manipulation and screening. In addition, these compounds may serve as a method to probe the highly orchestrated cell cycle and cell biology of these important pathogens.

![Figure 1](image_url). The 2,3-dihydro-4H-pyrido[3,2-\(e\)]1,3thiazin-4-one scaffold.

![Scheme 1](image_url). Preparation of 2-aryl-3-phenyl-2,3-dihydro-4H-pyrido[3,2-\(e\)]1,3thiazin-4-ones 1.

Compound 1j has also been shown to have activity against two human pathogenic fungi, *Cryptococcus neoformans* and *Lomentospora prolificans* [7], although this work is still ongoing. For potential use as a drug, analogs of lead compound 1j may show enhanced activity and lower IC50s. We are thus interested in producing derivatives of 1j to establish structure–activity relationships and improve the efficacy.

There are only a modest number of 2,3-dihydro-4H-pyrido[3,2-\(e\)]1,3thiazin-4-ones reported in the literature, and the number grows quite small when there is an aryl group on the nitrogen of the 1,3-thiazin-4-one ring [1,2,8–10]. A common method used for preparation of a variety of 2,3-dihydro-4H-1,3-thiazin-4-ones is the cyclization of an imine with a 3-thiocarboxylic acid [11]. However, it is known that \(N\)-aryl imines are less reactive in the preparation of similar 2,3-dihydro-4H-1,3-thiazin-4-ones than \(N\)-alkyl imines [12–14]. Furthermore, Dandia et al. reported in 2004 that reaction of \(N\)-aryl imines with thionicotinic acid (2-mercaptopyridine-3-carboxylic acid) 3 by thermal methods with various catalysts and solvents failed, which they attributed to the “low reactivity” of 3 [9]. Dandia et al.
succeeded in their reactions by using microwaves at 132–145 °C [9]. Arya et al. have reported success with ZSM-5-([MIM]+BF₄⁻), a Brønsted acid ionic liquid catalyst, plus ultrasonication at 95 °C [1,8].

Table 1. Compounds prepared as shown in Scheme 1.

| Compound | R       | Yield of 1 from 2 (%) |
|----------|---------|-----------------------|
| 1a [15]  | p-NO₂   | 45% a                 |
| 1b       | m-NO₂   | 52% a                 |
| 1c       | o-NO₂   | 22% a                 |
| 1d       | p-CF₃   | 63% a                 |
| 1e       | m-CF₃   | 54% a                 |
| 1f       | p-Br    | 40% a                 |
| 1g       | m-Br    | 42% a,b               |
| 1h [15]  | p-F     | 50% a                 |
| 1i       | m-F     | 43% a                 |
| 1j [4,5] | H       | 48% a                 |
| 1k       | p-Me    | 53% c                 |
| 1l       | m-Me    | 35% a,b               |
| 1m       | p-OMe   | 55% a                 |
| 1n       | m-OMe   | 44% a,b               |

a Isolated by chromatography and subsequent recrystallization. b Performed as a three-component coupling rather than premaking the imine. Yield is from aniline. c Isolated by recrystallization.

Where both of these methods involved specialized equipment (microwave oven, ultrasonic processor) and high temperature, we have previously reported simple, room temperature synthesis of various 2,3-diaryl-2,3-dihydro-4H-1,3-thiazin-4-ones [5,12–14], including a few 2-aryl-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-ones [4,5,15], using amide coupling agent 2,4,6-tripropyl-1,3,5,2,4,6-trioxatriphosphorinane-2,4,6-trioxide (T3P). T3P has several advantages over other amide-forming reagents, including that the reagent is safe, the by-products are water soluble and easily removed, and in combination with pyridine epimerization at the carbon alpha to the carboxylic acid is minimized [16]. Taylor’s group has also used T3P to prepare 2,3-dihydro-4H-1,3-thiazin-4-ones [17–19], but have not reported preparation of any 2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-ones.

Herein we report the synthesis of a full series of 2-aryl-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-ones 1, with a variety of substituents in the meta and para positions on the C-aryl ring, as well as one in the ortho position (Scheme 1). Both electron-withdrawing and electron-donating substituents were used. Additionally, initial screening of the compounds against T. brucei is disclosed. The first X-ray crystallographic structure of neutral reagent 3 is also reported.

2. Results and Discussion

The compounds 1a–1n were prepared and the yields are shown in Table 1. The preparations of the imines 2a–2n used have been previously reported [14], but an improved procedure for m-F imine 2i is included here. In cases where the imine was a liquid (2g, 2l, and 2n), the reactions were performed as three-component couplings, forming the imine in situ from an aldehyde and aniline rather than preparing it separately. As we have reported previously in the similar reaction of 3-mercaptopropionic acid [12], yields were expected to be a little lower for the three-component reaction than for the two-component reaction. Equimolar amounts of imine 2 (or aldehyde and aniline) and thionicotinic acid 3 were used, with excesses of T3P and pyridine. Reactions were allowed to proceed at least overnight and followed by TLC. Compounds 1a [15], 1h [15], and 1j [4,5] have been previously reported, but here updated procedures and yields are provided. All of the reactions attempted gave desired products 1a–n. In general, these compounds were readily crystallized. Yields ranged from 22% (ortho-nitro 1c) to 63% and averaged 46%. In the three-
component reactions (1g, 1l, and 1n) the average yield was 40%, ranging from 35–44%. The two-component reactions (1a–1f, 1h–1k, 1m) ranged from 22–63% and averaged 48% yield.

Arya et al. reported yields of 80–94% in reactions of N-aryl imines with 3 using ultrasonication and a Bronsted acid ionic liquid at 95 °C [1,8]. The same group reported similar reactions in 85–92% yield using microwaves at 132–145 °C [9]. Thus, while the yields were high, the reactions required specialized equipment and high temperatures. The method reported here is run at room temperature without special equipment, is simple to perform, and does not require dry solvents.

Reaction of the same meta- and para-substituted imines 2a, 2b, 2d–2n with thiosalicylic acid (2-mercaptobenzoic acid), an aromatic thioacid, had ranged from 12–43% with an average yield of 30% [14]. The more reactive aliphatic 3-mercaptopropionic acid gave yields of 33–75%, averaging 58% [12]. The yields for 3 (35–63%, average 48%) thus fall in between these two—better than thiosalicylic acid but not as good as 3-mercaptopropionic acid. Compound 3 is more complex than it might appear. 2-Mercaptopyridine is known to tautomerize to a nonaromatic thione (Scheme 2) [20]. However, Saleh studied this further, two crystal studies we re undertaken. Growth of crystals of 3T was oxidized to the disulfide [23,24]. This left open several possibilities for precisely how 3 is behaving in the reaction reported herein. To study this further, two crystal studies were undertaken. Growth of crystals of 3 from methanol gave crystals in which 3 was in the form of the thione tautomer 3T (Scheme 3). This appears to be the first time that the structure of the neutral molecule has been determined and shows that the thiol is in fact deprotonated before the carboxylic acid. There is O1-H—S1 hydrogen bonding within the molecule, and N1-H—O2 hydrogen bonding between molecules (Figure 2). Growth of 3 from pyridine, which is part of the reaction medium, produced the disulfide as a pyridine solvate (Figure 3). Half of the molecular species in the crystal have the proton on the carboxylic acid O and the other half have it on the pyridine (solvent) N, suggesting a proton exchange equilibrium in solution. Disulfide formation presumably occurred by O2 oxidation [24], as the crystals were grown by slow evaporation to the atmosphere. The reaction reported here has been run under N2, although air has not been rigorously excluded. Thus, it would appear that the reaction may have an equilibrium between the aromatic 3 and the nonaromatic 3T (Scheme 3), which could account for the yields being between those of aromatic thiosalicylic acid and nonaromatic 3-mercaptopropionic acid.

![Scheme 2](image1.png)

**Scheme 2.** Tautomerization of 2-mercaptopyridine [20].

![Scheme 3](image2.png)

**Scheme 3.** Tautomerization of aromatic 3 to nonaromatic 3T.
Based on these results and mechanisms previously proposed by us [12,14] and Unsworth et al. [17], one possible pathway for the reaction is shown in Scheme 4. In the proposed mechanism, the carboxylate of 3T attacks a phosphorus of T3P, opening the ring and forming a phosphate ester 4. The nitrogen of imine 2 then attacks the carbonyl of 4, with the phosphate group 6 leaving, to yield the iminium ion 5. Finally, ring closure by nucleophilic attack by sulfur on the strongly electrophilic carbon of the iminium ion 5 along with deprotonation by pyridine produces 1. Considering this last step, it would be expected that electron-withdrawing groups on the C-aryl ring would increase the electrophilicity of the iminium ion carbon. However, no pattern was discernible in the yields with regard to electron-withdrawing or electron-donating. This may be because...
the slow step in the process is likely to be the amide bond formation, and the subsequent ring closure is rapid. The low yield of 1c can be attributed to steric hindrance by the ortho substituent.

Scheme 4. Possible reaction pathway.

All fourteen compounds 1a–1n were tested at 50 µM concentration against the kinetoplastid parasite T. brucei brucei. Cultured long slender bloodstream form T. brucei were used for these experiments, as this form most closely represents the stage present in the mammalian host. Of the fourteen compounds analyzed, including 1j that was previously shown to inhibit T. brucei at higher concentrations [6], compounds with trifluoromethyl substitution (1d, 1e), bromo substitution (1f, 1g), and para-methyl substitution (1k) strongly inhibited growth of trypanosomes within the first 18 h (Figure 4; a table of the raw data is in the Supplementary Material). In fact, these compounds not only inhibited growth but killed parasites, with the para-bromo substitution (1f) killing parasites the quickest. Interestingly, the para-methyl substitution (1k) strongly inhibited parasite growth, while the meta-methyl substitution (1l) only showed a subtle effect on parasite growth. This may indicate that the substituent in the para-position may be able to bind more favorably than the meta-position, although more studies would be needed to confirm this. In summary, compounds 1d, 1e, 1f, 1g, and 1k strongly inhibited parasite growth at a concentration of 50 µM and are good candidates for further study.

X-ray crystallographic structures of compounds 1a, 1h, and 1j have been previously reported [4,15], as have physical and spectroscopic properties of 1j [5]. Here, full spectral data and physical properties of 1a–1l and 1k–1n are provided in the Materials and Methods Section and in the Supplementary Material.
Figure 4. Compounds 1a–1n were tested against long slender bloodstream form *Trypanosoma brucei*. Treated cells (50 µM) were compared to a vehicle control (DMSO) over the course of three days. The average of three replicates was plotted for treated cells with standard deviation shown as error bars. DMSO (vehicle) was plotted as the average of four replicates. Cell densities below the level of detection of the hemacytometer (10⁴) are represented by 0, and are not shown on the graph. For example, 1f was not detectable at 18 h, 1d and 1g (overlapping on the graph) were not detectable at 42 h, and 1e was not detectable at 66 h. A table of the raw data is in the Supplementary Material.

3. Materials and Methods

**General:** Thionicotinic acid (2-mercaptopyridine-3-carboxylic acid) 3 was obtained from Oakwood and also purchased from Sigma-Aldrich. 2-Methyltetrahydrofuran and silica gel for flash chromatography were purchased from Sigma-Aldrich. Pyridine was purchased from AlfaAesar. 2,4,6-Tripropyl-1,3,5,2,4,6-trioxatriphosphorinane-2,4,6-trioxide (T3P) in 2-methyltetrahydrofuran (50 weight %) was obtained from Curia. TLC plates (silica gel GF, 250-micron, 10 × 20 cm, cat. No. 21521) were purchased from Analtech. TLCs were visualized under short wave UV, and then with I₂, and then by spraying with ceric ammonium nitrate/sulfuric acid and heating. Infrared spectra of 1e, 1f, 1g, 1h, and 1n were run on a Thermo-Fisher NICOLET iS50 FT-IR using a diamond-ATR attachment for direct powder analysis (Coppin State University). Infrared spectra of 1a, 1b, 1c, 1d, 1i, 1k, and 1m were run on another Thermo-Fisher NICOLET iS50 FT-IR using a diamond-ATR attachment for direct powder analysis (Penn State Schuylkill). 1H, 13C NMR, and 19F NMR experiments (Penn State’s shared NMR facility, University Park) were carried out on a Bruker Avance-III-HD 500.20-MHz (1H frequency) instrument using a 5 mm Prodigy (liquid nitrogen cooled) BBO BB-1H/19F/D Z-GRD cryoprobe. Samples were dissolved in CDCl₃ and analyzed at RT. Typical conditions for ¹H acquisition were 2 s relaxation delay, acquisition time of 4.089 s, spectral width of 8 kHz, 16 scans. Spectra were zero-filled to 128k points, and multiplied by exponential multiplication (EM with LB = 0.3 Hz) prior to FT. For 13C experiments, data were acquired with power-gated ¹H decoupling using a 2 s relaxation delay, with acquisition time of 1.1 s, spectral width of 29.8 kHz, and 256 scans. Spectra were zero-filled once, and multiplied by EM with LB = 2 Hz prior to FT. ¹⁹F NMR data were acquired with acquisition time of 0.58 s, spectral width of 240 ppm, 32 scans, and relaxation delay of 2 s. Exact mass of synthesized compounds was determined LC-MS (Villanova University). Exact mass was measured on a SCIEX Exion LC with a SCIEX 5600+ TripleTOF MS. Separation was achieved on an Agilent Infinity LabPoroshell 120 EC-C18 column maintained at 40 °C with a gradient of 90/10 (water/acetonitrile with 0.1% formic acid) ramped from 5/95 over 6 min at a flow rate of 0.5 mL/min. The TOF-MS was scanned over 100–500 Da and calibrated with the SCIEX APCI positive calibrant solution prior to accurate mass analysis. Compound exact mass was measured in positive ESI.
mode with a DP = 100 V, CE = 10, GAS1 = GAS2 = 60 psi, CUR = 30 psi, ISV = 5500 V, and source temperature of 450 °C. Melting points were performed on an Arthur H. Thomas Co. Thomas Howard Capillary Melting Point Apparatus or on a Vernier Melt Station (Penn State Schuykill). Single crystals of C₆H₅NO₂S and C₁₂H₁₂N₂O₃S₂•2(C₆H₅·N) were grown by slow evaporation of methanol and pyridine solutions, respectively. Suitable crystals were selected and sequentially mounted using a nylon loop and a dab of paratone oil on a Rigaku Oxford diffraction, Synergy Custom system, HyPix-Arc 150 diffractometer (Penn State University Park). The crystals were at 173(2) K during data collection. Using Olex2 [25], the structures were solved with the SHELXT [26] structure solution program using Intrinsic Phasing and refined with the SHELXL [27] refinement package using least-squares minimization.

**X-ray Crystallographic Data**

**Thionicotinic acid grown in methanol:** Crystal Data for C₆H₅NO₂S (M = 155.17 g/mol): orthorhombic, space group Pna2₁ (no. 33), a = 8.2935(4) Å, b = 13.2553(6) Å, c = 5.8995(3) Å, V = 648.55(5) Å³, Z = 4, T = 173(2) K, µ(Cu Kα) = 3.882 mm⁻¹, D_{calc} = 1.589 g/cm³, 1840 reflections measured (12.59 °, I > 2σ(I)), 988 unique (R_{int} = 0.0168, R_{sigma} = 0.0162) which were used in all calculations. The final R1 was 0.0544 (I > 2σ(I)) and wR2 was 0.1385 (all data).

**Thionicotinic acid grown in pyridine:** Crystal Data for C₁₂H₇N₂O₃S₂•2(C₆H₅·N) (M = 466.52 g/mol): monoclinic, space group C2/c (no. 15), a = 7.9786(2) Å, b = 12.3830(3) Å, c = 21.5361(5) Å, β = 96.019(2)°, V = 2116.02(9) Å³, Z = 4, T = 173(2) K, µ(Cu Kα) = 2.616 mm⁻¹, D_{calc} = 1.464 g/cm³, 5474 reflections measured (8.256 ° ≤ 2θ ≤ 149.326°), 2034 unique (R_{int} = 0.0159, R_{sigma} = 0.0133) which were used in all calculations. The final R1 was 0.0310 (I > 2σ(I)) and wR2 was 0.0819 (all data). (Note that in the cif, the molecular formula and weight listed (C₁₁H₉N₂SO₂ and 233.26) is half of that for the moiety, corresponding to the contents in the asymmetric unit.)

**Imines:** Imines 2 were prepared as previously reported [14]. One purification was updated as reported below.

**N-[[3-(Trifluoromethyl)phenyl]methylene]aniline (2e):** Recrystallization from hexanes provided light yellow crystals (9.6692 g, 78%). mp: 45.6–47.8 °C (lit. 47–47.5 °C [28]).

**General Procedure for Three-Component Coupling to Prepare 2-Aryl-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-ones:** A two-necked 25 mL round-bottom flask was oven-dried, cooled under N₂, and charged with a stir bar. Aniline (0.559 g, 6 mmol) and a substituted benzaldehyde (6 mmol) were added. 2-Methyltetrahydrofuran (2.3 mL) was added and the solution was stirred. Thionicotinic acid (0.931 g, 6 mmol) was added. Pyridine (2.9 mL, 36 mmol) was added. Finally, 2,4,6-tripropyl-1,3,5,2,4,6-trioxatriphosphorinane-2,4,6-trioxide (T₃P) in 2-methyltetrahydrofuran (50 weight percent; 11 mL, 18 mmol) was added. The reaction was stirred at room temperature and followed by TLC, then poured into a separatory funnel with dichloromethane (20 mL). The mixture was washed with water (10 mL). The aqueous was then extracted twice with dichloromethane (10 mL each). The organics were combined and washed with saturated sodium bicarbonate (10 mL) and then saturated sodium chloride (10 mL). The organic was dried over sodium sulfate and concentrated under vacuum to give a crude mixture. Further purification was carried out as indicated below for each compound.

**2-(3-Bromophenyl)-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1g):** After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from ethyl acetate and hexanes gave off-white crystals (1.0001 g, 42% yield). mp: 142.6–143.5 °C. ¹H NMR (CDCl₃): δ(ppm): δ 8.53 (dd, J<sub>HH</sub> 4.8, J<sub>H</sub> 1.9 Hz, 1H, para to pyridine N), 8.48 (dd, J<sub>HH</sub> 3.9, J<sub>H</sub> 1.9 Hz, 1H, ortho to pyridine N), 7.93 (s, 1H, Ar), 7.47–7.37 (m, 4H, Ar), 7.34 (d, J<sub>HH</sub> 9.0 Hz, 3H, Ar), 7.24 (dd, J<sub>HH</sub> 7.9, 4.8 Hz, 1H, meta to pyridine N), 7.18 (t, J<sub>HH</sub> 7.9 Hz, 1H, Ar), 6.12 (s, 1H, C2-H). ¹³C NMR (CDCl₃): δ(ppm): δ 163.1 (C=O), 156.2 (Ar), 153.0 (Ar), 141.8 (Ar), 141.5 (Ar), 138.1 (Ar), 131.8 (Ar), 130.1
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(2-aryl-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-ones (1a): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from ethyl acetate and hexanes gave an off-white powder (0.9726 g, 45% yield). mp: 190.0–191.3 °C. 1H NMR (CDCl3): δ (ppm): 8.53 (dd, JHH 4.8, JHH 1.0 Hz, 1H, para to pyridine N), 7.98 (d, JHH 7.9 Hz, 1H, ortho to pyridine N), 7.43 (t, JHH 7.9 Hz, 2H, Ar), 7.35 (d, JHH 8.0 Hz, 2H, Ar), 7.25–6.7 (m, 1H, C2-H), 6.23 (s, 1H, C2-H). 13C NMR (CDCl3): δ (ppm): 163.0 (C=O), 155.8 (pyridine ring), 153.1 (pyridine ring), 147.9 (Ar), 144.6 (Ar), 141.6 (Ar), 138.3 (Ar), 129.6 (Ar), 127.9 (Ar), 127.5 (Ar), 125.9 (Ar), 125.5 (Ar), 124.0 (Ar), 121.8 (Ar), 64.3 (C2). HRMS (m/z): [C19H13BrN3O5S+H]+ of 364.0756 is consistent with calculated [M+H]+ of 364.0756. IR (neat, cm⁻¹): 1652 (C=O). Rf (50% EtOAc/hexanes) = 0.50.

2-(3-Methylphenyl)-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1b): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from a mixture of ethyl acetate and hexanes gave a white solid (1.128 g, 52% yield). mp: 190.0–191.3 °C. 1H NMR (CDCl3): δ (ppm): 8.53 (dd, JHH 4.8, JHH 1.0 Hz, 1H, para to pyridine N), 7.98 (d, JHH 7.9 Hz, 1H, ortho to pyridine N), 7.43 (t, JHH 7.9 Hz, 2H, Ar), 7.35 (d, JHH 8.0 Hz, 2H, Ar), 7.25–6.7 (m, 1H, C2-H), 6.23 (s, 1H, C2-H). 13C NMR (CDCl3): δ (ppm): 163.0 (C=O), 155.8 (pyridine ring), 153.1 (pyridine ring), 147.9 (Ar), 144.6 (Ar), 141.6 (Ar), 138.3 (Ar), 129.6 (Ar), 127.9 (Ar), 127.5 (Ar), 125.9 (Ar), 125.5 (Ar), 124.0 (Ar), 121.8 (Ar), 64.3 (C2). HRMS (m/z): [C19H13BrN3O5S+H]+ of 364.0756 is consistent with calculated [M+H]+ of 364.0756. IR (neat, cm⁻¹): 1652 (C=O). Rf (50% EtOAc/hexanes) = 0.50.

2-(3-Methylphenyl)-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1b): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from a mixture of ethyl acetate and hexanes gave a white solid (1.128 g, 52% yield). mp: 190.0–191.3 °C. 1H NMR (CDCl3): δ (ppm): 8.53 (dd, JHH 4.8, JHH 1.0 Hz, 1H, para to pyridine N), 7.98 (d, JHH 7.9 Hz, 1H, ortho to pyridine N), 7.43 (t, JHH 7.9 Hz, 2H, Ar), 7.35 (d, JHH 8.0 Hz, 2H, Ar), 7.25–6.7 (m, 1H, C2-H), 6.23 (s, 1H, C2-H). 13C NMR (CDCl3): δ (ppm): 163.0 (C=O), 155.8 (pyridine ring), 153.1 (pyridine ring), 147.9 (Ar), 144.6 (Ar), 141.6 (Ar), 138.3 (Ar), 129.6 (Ar), 127.9 (Ar), 127.5 (Ar), 125.9 (Ar), 125.5 (Ar), 124.0 (Ar), 121.8 (Ar), 64.3 (C2). HRMS (m/z): [C19H13BrN3O5S+H]+ of 364.0756 is consistent with calculated [M+H]+ of 364.0756. IR (neat, cm⁻¹): 1652 (C=O). Rf (50% EtOAc/hexanes) = 0.50.
yield). mp: 149.7–149.9 °C. 1H NMR (CDCl3): δ (ppm): δ 8.55–8.52 (m, 1H, para to pyridine N), 8.50 (dd, 3JHH 7.8, 4JHH 1.6 Hz, 1H, ortho to pyridine N), 8.37 (s, 1H, Ar), 8.15 (d, 3JHH 8.3 Hz, 1H, Ar), 7.82 (d, 3JHH 8.9 Hz, 1H, Ar), 7.51 (t, 3JHH 8.0 Hz, 1H, Ar), 7.45 (t, 3JHH 7.9 Hz, 2H, Ar), 7.39–7.32 (m, 3H, Ar), 7.26 (dd, 3JHH 7.8, 4.8 Hz, 1H, meta to pyridine N), 6.25 (s, 1H, C2-H). 13C NMR (CDCl3): δ (ppm): 162.9 (C=O), 155.8 (Ar), 153.1 (Ar), 148.4 (Ar), 141.7 (Ar), 141.6 (Ar), 138.3 (Ar), 132.1 (Ar), 129.8 (Ar), 129.7 (Ar), 129.5 (Ar), 125.7 (Ar), 123.7 (Ar), 121.8 (Ar), 121.6 (Ar), 64.2 (C2). HRMS (m/z): [C19H13NO3S+H]+ of 364.0754 is consistent with calculated [M+H]+ of 364.0756. IR (neat, cm−1): 1654 (C=O). Rf (50% EtOAc/hexanes) = 0.30.

2-(2-Nitrophenyl)-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1c): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from a mixture of ethyl acetate and hexanes gave light yellow crystals (0.4811 g, 22% yield), mp: 195.7–198.5 °C. 1H NMR (CDCl3): δ (ppm): 8.52 (q, 3JHH 1.9 Hz, 1H, para to pyridine N), 8.51 (s, 1H, Ar), 8.18 (dd, 3JHH 8.1, 4JHH 1.5 Hz, 1H, Ar), 7.64 (d, 3JHH 7.0 Hz, 1H, Ar), 7.57 (td, 3JHH 7.6, 4JHH 1.5 Hz, 1H, Ar), 7.53–7.46 (m, 2H, Ar), 7.43–7.30 (m, 3H, Ar), 7.26–7.21 (m, 1H, Ar), 7.15 (s, 1H, C2-H). 13C NMR (CDCl3): δ (ppm): 163.9 (C=O), 156.4 (Ar), 153.2 (Ar), 146.2 (Ar), 142.0 (Ar), 138.1 (Ar), 135.4 (Ar), 133.4 (Ar), 129.7 (Ar), 129.6 (Ar), 128.0 (Ar), 127.5 (Ar), 127.1 (Ar), 125.8 (Ar), 124.9 (Ar), 121.4 (Ar), 60.6 (C2). HRMS (m/z): [C19H13NO3S+H]+ of 364.0750 is consistent with calculated [M+H]+ of 364.0756. IR (neat, cm−1): 1663 (C=O). Rf (50% EtOAc/hexanes) = 0.49.

3-Phenyl-2-[4-(trifluoromethyl)phenyl]-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1d): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from ethyl acetate and hexanes gave slightly off-white crystals (1.4097 g, 63% yield), mp: 185.2–186.1 °C. 1H NMR (CDCl3): δ (ppm): 8.52 (dd, 3JHH 4.8, 4JHH 1.8 Hz, 1H, para to pyridine N), 8.48 (dd, 3JHH 7.9, 4JHH 1.9 Hz, 1H, ortho to pyridine N), 7.69–7.54 (m, 4H, Ar), 7.49–7.39 (m, 2H, Ar), 7.39–7.31 (m, 3H, Ar), 7.24 (dd, 3JHH 7.9, 4.8 Hz, 1H, meta to pyridine N), 6.20 (s, 1H, C2-H). 13C NMR (CDCl3): δ (ppm): 163.0 (C=O), 156.0 (Ar), 152.8 (Ar), 143.2 (Ar), 141.7 (Ar), 138.4 (Ar), 131.3, 131.0, 130.7, 130.5, q, 3JCF 32.6 Hz, Ar C connected to CF3, 129.6 (Ar), 127.8 (Ar), 126.9 (Ar), 126.0 (Ar), 125.8, 125.8, 125.7, 125.7, q, 3JCF 3.7 Hz, Ar ortho to CF3), 125.5 (Ar), δ (ppm): 6.6–6.8. 387.0775 is consistent with calculated [M+H]+ of 387.0778. IR (neat, cm−1): 1652 (C=O). Rf (50% EtOAc/hexanes) = 0.54.

3-Phenyl-2-[3-(trifluoromethyl)phenyl]-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1e): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from ethyl acetate and hexanes gave a white powder (1.2574 g, 54% yield), mp: 119.2–120.2 °C. 1H NMR (CDCl3): δ (ppm): 8.52 (dd, 3JHH 4.8, 4JHH 1.9 Hz, 1H, para to pyridine N), 8.48 (dd, 3JHH 7.9, 4JHH 1.9 Hz, 1H, ortho to pyridine N), 7.73 (s, 1H, Ar), 7.67 (d, 3JHH 7.8 Hz, 1H, Ar), 7.65 (d, 3JHH 7.9 Hz, 1H, Ar), 7.44 (td, 3JHH 8.2, 5.2 Hz, 3H, Ar), 7.37–7.30 (m, 3H, Ar), 7.24 (dd, 3JHH 7.9, 4.8 Hz, 1H, meta to pyridine N), 6.21 (s, 1H, C2-H). 13C NMR (CDCl3): δ (ppm): 163.1 (C=O), 156.1 (Ar), 153.0 (Ar), 141.7 (Ar), 140.4 (Ar), 138.1 (Ar), 136.1, 131.4, 131.1, 130.9, q, 3JCF 32.7 Hz, Ar C connected to CF3), 129.6 (Ar), 129.5 (Ar), 129.2 (Ar), 127.8 (Ar), 126.9, 124.7, 122.5, 120.4, q, 3JCF 272.5 Hz, CF3, 126.0 (Ar), 125.7 (Ar), 125.6, 125.5, 125.5, q, 3JCF 3.9 Hz, Ar C ortho to CF3), 123.5, 123.5, 123.4, 123.4, q, 3JCF 3.8 Hz, Ar C ortho to CF3), 121.6 (Ar), 64.5 (C2). 19F NMR (CDCl3): δ (ppm): -62.8. HRMS (m/z): [C20H13F3N2O2S+H]+ of 387.0773 is consistent with calculated [M+H]+ of 387.0778. IR (neat, cm−1): 1639 (C=O). Rf (50% EtOAc/hexanes) = 0.59.

2-(4-Bromophenyl)-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1f): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from ethyl acetate and hexanes gave off-white crystals (0.9455 g, 40% yield), mp: 171.6–173.6 °C. 1H NMR (CDCl3): δ (ppm): 8.59–8.39 (m, 2H, ortho and para to pyridine N), 8.18 (d, 3JHH 8.8 Hz, 2H, Ar), 7.68 (d, 3JHH 8.9 Hz, 2H, Ar), 7.44 (t, 3JHH 7.8 Hz, 2H, Ar), 7.39–7.31 (m, 3H, Ar), 7.26 (dd, 3JHH 7.9, 4.8 Hz, 2H, Ar), 6.23 (s, 1H, C2-H). 13C NMR
(CDCl₃): δ (ppm): δ 162.3 (C=O), 156.4 (Ar), 152.9 (Ar), 141.8 (Ar), 138.3 (Ar), 138.1 (Ar), 131.8 (Ar), 129.5 (Ar), 128.2 (Ar), 127.6 (Ar), 126.0 (Ar), 125.6 (Ar), 122.8 (Ar), 121.5 (Ar), 64.5 (C2). HRMS (m/z): [C₁₉H₁₃BrN₂O₅·H⁺] of 397.0009 is consistent with calculated [M+H]⁺ of 397.0005. IR (neat, cm⁻¹): 1652 (C=O). Rᵣ (50% EtOAc/hexanes) = 0.59.

2-(4-Fluorophenyl)-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1h): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from ethyl acetate and hexanes gave white crystals (1.0067 g, 50% yield). mp: 127.8–128.4 °C. ¹H NMR (CDCl₃): δ (ppm): δ 8.52 (dd, ³J_{HF} 4.8, ⁴J_{HF} 2.2 Hz, 1H, para to pyridine N), 8.46 (dd, ³J_{HF} 7.9, ⁴J_{HF} 1.9 Hz, 1H, ortho to pyridine N), 7.46–7.40 (m, 1H, Ar), 7.40–7.26 (m, 6H, Ar), 7.24 (dd, ³J_{HF} 7.9, 4.8 Hz, 1H, meta to pyridine N), 7.20 (dd, ³J_{HF} 10.3, ⁴J_{HF} 2.4 Hz, 1H), 7.01–6.95 (m, 1H, Ar), 6.15 (s, 1H, C2-H). ¹³C NMR (CDCl₃): δ (ppm): δ [163.8, 161.8, 1H, C=O, 156.3 (Ar), 152.9 (Ar), 141.9 (Ar), 141.9, 1H, para to pyridine N), 8.46 (dd, ³J_{HF} 7.9, ⁴J_{HF} 1.9 Hz, 1H, ortho to pyridine N), 7.46–7.40 (m, 1H, Ar), 7.40–7.26 (m, 6H, Ar), 7.24 (dd, ³J_{HF} 7.9, 4.8 Hz, 1H, meta to pyridine N), 7.20 (dd, ³J_{HF} 10.3, ⁴J_{HF} 2.4 Hz, 1H), 7.01–6.95 (m, 1H, Ar), 6.15 (s, 1H, C2-H). HRMS (m/z): [C₁₉H₁₃FN₂O₅·H⁺] of 337.0812 is consistent with calculated [M+H]⁺ of 337.0805. IR (neat, cm⁻¹): 1652 (C=O). Rᵣ (50% EtOAc/hexanes) = 0.48.

2-(3-Fluorophenyl)-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1j): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from ethyl acetate and hexanes gave an orange–yellow solid (1.0572 g, 53% yield). mp: 128–130 °C. ¹H NMR (CDCl₃): δ (ppm): δ 8.51 (dd, ³J_{HF} 4.7, ⁴J_{HF} 1.8 Hz, 1H, para to pyridine N), 8.46 (dd, ³J_{HF} 7.9, ⁴J_{HF} 1.9 Hz, 1H, ortho to pyridine N), 7.43–7.30 (m, 7H, Ar), 7.21 (dd, ³J_{HF} 7.9, 4.8 Hz, 1H, meta to pyridine N), 6.81 (d, ³J_{HF} 8.9 Hz, 2H, Ar), 6.15 (s, 1H, C2-H), 2.37 (s, 3H, OCH₃). ¹³C NMR (CDCl₃): δ (ppm): δ 163.4 (C=O), 159.7 (Ar), 156.9 (Ar), 152.8 (Ar), 141.9 (Ar), 137.9 (Ar), 130.9 (Ar), 129.3 (Ar), 127.9 (Ar), 127.4 (Ar), 126.1 (Ar), 125.7 (Ar), 121.2 (Ar), 114.0 (Ar), 64.7 (C2), 55.3 (OCH₃). HRMS (m/z): [C₁₉H₁₆N₂O₃·H⁺] of 349.1005 is consistent with calculated [M+H]⁺ of 349.1005. IR (neat, cm⁻¹): 1644 (C=O). Rᵣ (50% EtOAc/hexanes) = 0.48.

2,3-Diphenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1l): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from 2-propanol gave an ivory solid (1.0914 g, 48% yield). Data previously reported [5].

2-(4-Methoxophenyl)-3-phenyl-2,3-dihydro-4H-pyrido[3,2-e][1,3]thiazin-4-one (1m): After chromatography on 30 g silica gel with mixtures of ethyl acetate and hexanes, recrystallization from 2-propanol gave a yellow solid (1.1419 g, 55% yield). mp: 142.5–143.0 °C. ¹H NMR (CDCl₃): δ (ppm): δ 8.51 (dd, ³J_{HF} 4.7, ⁴J_{HF} 1.8 Hz, 1H, para to pyridine N), 8.46 (dd, ³J_{HF} 7.9, ⁴J_{HF} 1.9 Hz, 1H, ortho to pyridine N), 7.43–7.30 (m, 7H, Ar), 7.21 (dd, ³J_{HF} 7.9, 4.8 Hz, 1H, meta to pyridine N), 6.81 (d, ³J_{HF} 8.9 Hz, 2H, Ar), 6.15 (s, 1H, C2-H), 3.77 (s, 3H, OCH₃). ¹³C NMR (CDCl₃): δ (ppm): δ 163.4 (C=O), 159.7 (Ar), 156.9 (Ar), 152.8 (Ar), 141.9 (Ar), 137.9 (Ar), 130.9 (Ar), 129.3 (Ar), 127.9 (Ar), 127.4 (Ar), 126.1 (Ar), 125.7 (Ar), 121.2 (Ar), 114.0 (Ar), 64.7 (C2), 55.3 (OCH₃). HRMS (m/z): [C₁₉H₁₆N₂O₃·H⁺] of 349.1006 is consistent with calculated [M+H]⁺ of 349.1005. IR (neat, cm⁻¹): 1644 (C=O). Rᵣ (50% EtOAc/hexanes) = 0.48.
Cell Culture: Bloodstream form T. brucei brucei strain 90–13 were cultured at 37 °C in 5% CO₂ in HMI-9 supplemented with 15% fetal bovine serum [29]. Cells were maintained in 5 µg/mL hygromycin and 1 µg/mL neomycin (G418). Parasites from a mid-log culture were split back to 2 × 10⁵ cells/mL in 1 mL, treated with 50 µM of each compound in triplicate, and counted over 3 days using a hemacytometer. The average of 4 replicates for the DMSO vehicle control and 3 replicates for treated samples was plotted with standard deviation shown as error bars (Figure 4).

4. Conclusions

A new, simple preparation of 2-aryl-3-phenyl-2,3-dihydro-4'H-pyrido[3,2-e][1,3]thiazin-4-ones 1 from two difficult substrates (2 and 3) has been demonstrated. The reactions run at room temperature, are simple to perform, and do not require dry solvents or special equipment such as a microwave oven or an ultrasonicator. A series of compounds with various electron-donating and electron-withdrawing groups on the C2-aryl ring was prepared in moderate–good yields. The reactions can be run as either a two- or three-component reaction, although yields were somewhat lower for the three-component reactions.

Some of these compounds strongly inhibited growth of the parasite T. brucei, an important animal pathogen closely related to other species and subspecies that cause disease in humans. These compounds are good candidates for further study as pharmacological agents.

X-ray crystallographic studies of 3 yielded the first crystal structure of the neutral species and showed it to be in the thione form 3T.

Supplementary Materials: The following are available online. ¹H NMR, ¹³C NMR, ¹⁹F NMR, FTIR, Mass spectra, and data for the biological study are available online. CCDC deposition number 2108518–2108519 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via http://www.ccdc.cam.ac.uk/conts/retrieving.html, accessed on 7 August 2021 (or from the CCDC, 12 Union Road, Cambridge CB2 1EZ, UK; Fax: +44-1223-336033; E-mail: deposit@ccdc.cam.ac.uk).

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