Association between Enterovirus Infection and Clinical Type 1 Diabetes Mellitus: Systematic Review and Meta-analysis of Observational Studies

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Summary: Numerous animal models and epidemiological and observational studies have demonstrated that enterovirus(EV) infection could be involved in the development of clinical type 1 diabetes mellitus(T1DM), but its etiology is not fully understood. Therefore, we reviewed the association between EV infection and clinical T1DM. We searched PubMed and Embase from inception to April 2021 and reference lists of included studies without any language restrictions in only human studies. The correlation between EV infection and clinical T1DM was calculated as the pooled odds ratio(OR) and 95% confidence intervals(CI), analysed using random effects models. Subgroup and sensitivity analyses were performed to evaluate the robustness of the associations. A total of 25 articles(22 case-control studies and 3 nested case-control studies) met the inclusion criterion including 4854 participants(2948 cases and 1906 controls) with a high level of statistical heterogeneity($I^2=80\%, P<0.001$) mainly attributable to methods of EV detection, study...
type, age distribution, source of EV sample and control subjects. Meta-analysis showed a significant association between EV infection and clinical T1DM ($OR=5.75$, $95\%CI=3.61-9.61$). There is a clinically significant association between clinical T1DM and EV infection.

**Key words:** Clinical T1DM; Enterovirus Infection; Observational Studies; Meta-analysis

**Introduction**

Type 1 diabetes mellitus (T1DM) is generally believed to be a chronic autoimmune disease characterized by the destruction of insulin-producing β-cells that results from a complex interaction between genetic susceptibility, the immunological factors and environmental agents[1]. It has been reported that the present global number of individuals with diabetes was estimated at 415 million, but has reached as much as 642 million by 2040. The estimated incidence rates of T1DM increased annually by 1.4% during 2002-12 in America[2], 1.01% during 2010-15 in China respectively[3, 4]. The rapid increase in incidence, especially in children under the age of 5 years[5], cannot be fully attributed to genetic factors. Reports have linked viral infections[6], obesity[7], socioeconomic status[8], vitamin D deficiency[9, 10], diet, immunisation, seasonal variation[11] to an increased risk of T1DM. However more recent evidence regarding a putative role for enterovirus (EV) infection in the development of clinical T1DM comes from case-control studies that have shown a significant temporal association after enterovirus epidemics, and the detection of EV RNA or EV capsid protein in pancreatic biopsies of patients with current onset T1DM[6]. On the other
hand, evidence from diabetic animal models and cell studies suggests that EVs are likely to destroy the pancreas via immunological cross-reaction (molecular mimicry) because of the sequence homology between the coxsackievirus P2 protein and glutamic acid decarboxylase 65 (GAD65) or is directed to destroy insulin producing islet cells via T lymphocytes (bystander damage) [1]. T1DM may also contribute to the children’s psychological and mental problems [12], such as depression and anxiety, since a strong correlation between diabetes and the status of mind or quality of life in children has been reported. Besides, diabetes could increase the economic burden to families and societies in low- and middle-income countries because of lifelong treatment and management of illness. Furthermore, identification of these risk factors could lead to a better understanding of T1DM and contribute to developing strategies to prevent T1DM so as to reduce the economic burden of diabetes and improve the quality of life.

In 2004, a systematic review of coxsackie B virus serology did not indicate a relation with T1DM [13], but there was another study showing that EV infection, confirmed only by reverse transcription polymerase chain reaction, did show a clinically significant association with T1DM in 2011 [14]. Moreover, these case-control or cohort studies did not increase the statistical power and provided precise estimates because of the relatively small sample size for each individual study. However, the correlation between EV and clinical T1DM remains unclear due to the source of EV samples, different methods to confirm EV infection and study type and
so on. Based on these facts, we conducted a systematic review and meta-analysis to clarify the relationship between EV infection and the risk of clinical T1DM.

**Methods**

This study was performed in accordance with the Meta-analysis of Observational Studies in Epidemiology (MOOSE)[15] and was registered on PROSPERO (registration number: CRD42021236044).

**Search strategy** Two reviewers (SY and XL D) independently performed a systematic search for observational studies of enterovirus (EV) infection and clinical type 1 diabetes mellitus (T1DM) according to Medical Subject Headings (MeSH) or Emtree combined with free-text terms on PubMed and Embase, from inception to April, 2021. The search terms used were “type 1 diabetes”, “enterovirus infection”, “echovirus infection” and “coxsackievirus infection”. The search strategy is reported in detail in the Supplementary Materials. The search was confined to original articles including humans in any language and was conducted by manually searching the reference lists of the eligible studies and by direct contact with authors.

**Study Selection Criteria** Studies were eligible for inclusion in line with the below criteria: (1) a case-control or cohort study design; (2) assessment of the relation between EV infection and clinical T1DM; (3) established or newly diagnosed T1DM without HIV or hepatitis viruses; (4) evidence of EV infection via measuring virus RNA or specific antibodies in blood, stool or tissue of patients or laboratory investigations or other biopsies; (5) available data on odds ratio (OR) with 95% confidence interval (CI) or numbers of events/total in both case and control;
(6) non-human studies were excluded. Disagreements between the two reviewers were resolved via discussion with a third arbitrary investigator (LW C), when necessary.

**Data Extraction**  The information of each study was extracted using a standardised form as follows: first author’s name, date of publication, design of study, location, age distribution, number of cases and controls (matching criteria), methods to confirm EV infection, virus species or serotypes, assessment of diabetes and islet autoantibodies. One investigator (ZZ) extracted above the data checked by another investigator (YL Z).

**Quality Assessment**  Two investigators (SY and BY Z) independently evaluated the included study quality independently using the Newcastle-Ottawa scale (NOS) for case-control studies or cohort studies, as recommended by the Cochrane Collaboration, and different opinions were resolved through consensus. This tool assessed three areas—selection, comparability, exposure or outcome—total score of 9 stars, with 5 stars or more deemed as good methods. In the comparability category, we highlighted the evaluation of controls matched for age and sampling time, as these two factors are most likely to affect the incidence of EV infection. The possibility of publication bias was evaluated by visual inspection of the funnel plot.

**Statistical Analysis**  Review Manager software (version 5.3) was used to calculate pooled ORs with 95% CI and P value for EVs infection in patients with diabetes versus no diabetes from the published data in studies using the Mantel-Haenszel method. We analysed the association between EV infection and clinical T1DM using both fixed and random-effects models. However, only combined ORs from the
random-effects models are presented because of a high level of heterogeneity in the study design. A forest plot summarised the results of all eligible studies. Statistical heterogeneity was explored using Cochran’s Q and \( I^2 \) statistics, indicating the proportion of variance in outcomes between studies. Statistical significance was defined as \( P<0.05 \) was considered statistically significant heterogeneity, while \( I^2 \) less than 25%, 50% and 75% were regarded as low, moderate and high heterogeneity respectively. Subgroup and sensitivity analyses were conducted for age distribution, the initial time of clinical T1DM, methods to confirm EV infection, source of EV sample and control subjects, virus species or serotypes, study type, and summary ORs were calculated. We also performed sensitivity analyses by individual study, study size, study location and NOS score.

**Results**

*Study Selection*  Our search returned all 706 publications after the removal of 272 duplicate articles. We identified and included 53 potential original articles by screening titles and abstracts. Furthermore, 25 relevant studies were evaluated by reviewing the full text and finally included in the systematic review and meta-analysis. Four studies were excluded owing to the same case-control subjects for different research aims. Figure 1 shows the search flowchart for eligible studies.

*Characteristics of Included Studies*  Demographic characteristics of the included 25 studies is presented in Table 1. All studies were case-control designs, and of these samples of 3 nested case-control studies were collected from diabetic cohort studies. The majority of included study subjects were from Europe and a few participants
were from non-European countries. Twenty-five studies included 2150 patients with T1DM and 2704 controls, aged range between 0 and 70 years, but who were mostly children, adolescents and young adults. EV infection was confirmed by real transcription polymerase chain reaction (RT-PCR) and in situ hybridization (ISH) to detect EV-RNA in 15 studies, specific IgM antibodies using neutralisation test (NT), ELISA, immunofluorescence assay (IFA), and radioimmunoassay (RIA) to identify antibodies against EV or coxsackievirus for seven studies, and immunohistochemistry (IHC) to investigate enteroviral capsid protein vp1 for three studies. Of the 25 studies, 5 studies simultaneously used two methods of EV detection. Most articles did not report data on EV species or serotypes, but only 5 studies provided data on IgM antibodies against coxsackievirus B serotypes and of which one only examined echovirus and coxsackievirus A.

**Meta-analysis results** The results suggested that EV infection was significantly related to clinical T1DM mellitus as compared with no T1DM, but with evidence of high heterogeneity between the 25 studies ($P<0.001$, $I^2=80\%$) (Figure 2). ORs ranged from 0.14 to 426.19, with a combined OR of 5.75 (95% CI = 3.61–9.16) (figure 2).

**Subgroup and sensitivity analyses** We also performed subgroup analyses with respect to methods to confirm EV infection, source of EV sample, virus species or serotypes, initial time of clinical T1DM, age distribution, source of control subjects and study type (Table 2). The combined ORs for NT, RIA, ISH, IHC and RT-PCR were 1.58 (95% CI = 0.76–3.30), 3.02 (95% CI = 0.17–56.64), 5.21 (95% CI = 2.31–11.79), 7.29 (95% CI = 1.42–37.58), 7.48 (95% CI = 4.20–13.32) respectively. There was no
heterogeneity ($I^2 = 0\%, P = 0.35$) across the two studies that measured EV-RNA in the intestinal mucosa by ISH, but the other subgroups by NT, RIA, IHC and RT-PCR showed significant heterogeneity (Table 2).

When we analysed the source of EV sample, the summary ORs for serum, intestinal mucosa, plasma, peripheral blood mononuclear cells (PBMCs) and pancreatic tissue were 3.90 ($95\% CI = 3.24-4.70$), 4.36 ($95\% CI = 2.24-8.49$), 4.49 ($95\% CI = 3.12-6.46$), 11.42 ($95\% CI = 4.27-30.58$), 27.60 ($95\% CI = 8.48-89.78$) respectively, with significantly statistically heterogeneity ($I^2 = 72.30\%, P = 0.006$) across the 5 groups, while there were no heterogeneity in PBMCs group ($I^2 = 0\%, P = 0.52$) and pancreatic tissue group ($I^2 = 0\%, P = 0.67$), moderate heterogeneous in intestinal mucosa group ($I^2 = 50\%, P = 0.14$), high heterogeneous in serum group ($I^2 = 83\% , P < 0.001$) and plasma group ($I^2 = 77\% , P = 0.002$) respectively (Table 2).

For only six studies that examined specific IgM antibodies to coxsackievirus B (CB) serotypes, the pooled ORs for CB1, CB2, CB3, CB5, CB6 were 1.76 ($95\% CI = 1.18-2.63$), 0.90 ($95\% CI = 0.50-1.64$), 0.95 ($95\% CI = 0.68-1.33$), 0.91 ($95\% CI = 0.37-2.21$), 0.88 ($95\% CI = 0.50-1.53$) respectively, while the pooled OR for CB4 was comparatively higher [2.03 ($95\% CI = 0.87-4.75$)] for the other CB1, CB2, CB3, CB5, CB6 serotypes, and with moderate heterogeneity ($I^2 = 60\% , P = 0.04$) versus no or mild heterogeneity respectively ($I^2 = 0\% , P = 0.67$; $I^2 = 25\% , P = 0.26$; $I^2 = 0\% , P = 0.51$; $I^2 = 43\% , P = 0.15$; $I^2 = 0\% , P = 0.87$) (Table 2).

The combined ORs for newly diagnosed clinical T1DM and previously diagnosed clinical T1DM were 4.76 ($95\% CI = 2.84-7.98$), 4.91 ($95\% CI = 2.49-9.67$),
with high heterogeneity ($I^2=82\%, P<0.001$) versus mild heterogeneity ($I^2=32\%, P=0.22$) respectively (Table 2). For the summary OR of age 0–9 years group [33.82 (95% CI = 1.87–612.91)] was significantly higher in the 0–20 years group [4.89 (95% CI = 2.51–9.51)] and age 0–71 years group [7.53 (95% CI = 3.61–15.72)], probably because of the high rates of EV infection in children [18, 21, 34]. It is generally believed to children with immature immune system in whom they have greater risk of infection in comparison with adults who have mature immune system. The rates background refers to this. Although there was no appropriate data on HLA genotypes in all cases and controls to perform subgroup analysis, we investigated the relationship between EV infection and clinical T1DM affected by HLA typing based on different sources of control subjects, The relative controls are selected from the siblings, while the normal subjects are collected from the unrelated individuals, so that we are able to elucidate the effect of genetic factor on the result. The combined OR of relative controls [3.19 (95% CI = 1.65–6.14)] was significantly higher for the normal (Table 2) subjects [56.41 (95% CI = 3.54–899.40)], indirectly demonstrating that EV infection could increase the risk of clinical T1DM in genetically susceptible individuals (table 2).

When we analysed the results from the only three nested case-control studies, the summary OR [2.16 (95% CI = 0.95–4.92)] was lower than that of the 22 case-control studies [7.49 (95% CI = 4.20–13.36)], probably because of the variance in study design (Table 2). In summary, subgroup analyses indicate that none of subset significantly affected the stability of overall results, in addition to group B coxsackievirus that obviously decreased the combined OR of 5.75 (95% CI = 3.61–9.16)
for an relation between EV infection and clinical T1DM to that of 1.14(95%CI=0.92-1.41) for an association between CB infection and clinical T1DM.

We also carried out sensitivity analysis by study size, study location and NOS score to examine the robustness of the correlation. The summary ORs for more than 100 participants and less than 100 participants were 5.88(95%CI=3.46-9.99), 6.00(95%CI=2.21-16.26) respectively, with no heterogeneity between the groups. The pooled ORs for European area and non-European area were 5.72(95%CI=2.94-11.94), 6.02(95%CI=3.02-12.01) respectively, with no heterogeneity between both groups. Sensitivity analysis by study quality was classified into 3 groups(8~9 score group, 7 score group, 5~6 score group) because all studies scored more than 5 on the NOS(Table 2). Although most studies did not report diagnostic criteria in detail for clinical T1DM ,and clinical presentation and laboratory investigations were poorly described, insulin therapy was started in patients with T1DM after diagnosis. Finally, sensitivity analysis by individual studies did not significantly affect the summary effect estimates.

Quality Assessment. Newcastle Ottawa scores ranged from 5 to 9 stars, with all studies scoring 5 stars or more, suggesting good methodological quality overall, with no studies reporting a non-response rate (Table 1). The funnel plot showed reasonable symmetry, with no evidence of publication bias(Figure 3). However there was great variability across the studies, with significant statistical heterogeneity; therefore, the meta-analysis results should be interpreted with caution when extended to the general population.
Discussion

To our knowledge, this meta-analysis and systematic review is the first to report the relationship between EV infection and the risk of clinical T1DM by systematically reviewing molecular and serological observational studies. This study, which included 25 articles, suggest that EV infection had a more than about six times the risk of clinical T1DM, approximate 34 times the risk in children when compared with the control. Those who have clinical T1DM with positivity for islet autoantibodies are not at greater risk of EV infection than those with negative results for islet autoantibodies; therefore, so enterovirus infection might be a risk factor for clinical type 1 diabetes. Our results suggest that the pathogenesis of clinical T1DM triggered by EV may not be one mechanism. Subgroup analysis demonstrated that age group, methods of EV detection, source of EV sample, study design and genetic factors may have a tremendous influence on the results.

To date, a great many epidemiological and observational studies have investigated the relationship between EV infection and the risk of T1DM[6, 10, 11, 17, 20, 22-29, 31-33, 35-37]. However their findings have been inconsistent because the prevalence and incidence of T1DM differ greatly for geographic areas, methods of EV detection, age and source of EV samples, and genetically predisposed individuals.

Since J. E. Banatvala et al. first reported evidence of the association between Coxsackie B1-5 viruses and children under the age of 5 years with clinical T1DM in Austria, England and Australia[38], subsequently most studies have been conducted in Europe. Overall, it is recognised by most investigators that EV infection could
accelerate the progression of T1DM or transiently emerge autoantibodies associated with T1DM in genetically susceptible populations, however, we perform an analysis of the islet autoantibody-positive individuals due to lack of sufficient data that provided unreliable results. We included children and adults with clinical T1DM, decreasing the high rates background of bacterial and viral infections in children. Global studies were included to decrease the risk of geographical bias associated with infection rates. Most studies, however, were from European countries[6, 10, 16-26, 28, 30-33] where the incidence of diabetes is higher than that in Asian and African countries. Given the heterogeneity of study populations’ heterogeneity, complex pathogenesis, and multiple environmental agents, we used random effects models due to high heterogeneity across individual studies, providing more conservative and reliable effect estimates. However, our results should be carefully interpreted due to significant statistical differences among all studies, when particular, as extended to the external population.

Although the initial factors of the anti-islet cell autoimmune response are not understood, a few possible mechanisms for the relationship between EV infection and risk of T1DM have been inferred. First, patients with clinical T1DM are more likely to be infected with a variety of pathogens, such as bacteria, viruses and fungi, compared to individuals without T1DM. Viruses can promote diabetes either by directly infecting and destroying islet beta cells or by triggering an autoimmune attack on islet cells[1]. In addition, seasonal variation plays an important role in the pathogenesis of T1DM[11]. Furthermore, diet and perinatal factors are more likely to
increase the risk of developing T1DM[39]. Secondly, there was one possible fact that molecular mimicry due to homology between Glutamic acid decarboxylase 65 (GAD65) and a causative agent such as Coxsackie B4 virus. A search has investigated several autoantigens (IAA, ICA, IA-2A, and ZnT8) within pancreatic beta cells play significant roles in the initiation or progression of autoimmune pancreatic injury. Nonetheless, children or adults with other autoimmunity, most commonly autoimmune thyroiditis and celiac disease, are at increased risk for catching T1DM; however, there was an extreme lack of available data in the eligible studies, so that we could not perform the related analysis. In the long term, clinical T1DM is an autoimmunity that gives rise to a complex interaction between genetically susceptible population and environmental factors[1].

In the future, larger multicentre international prospective or birth cohort studies could investigate the relationships between clinical T1DM and age distribution, genetic factors, enterovirus various, and EV serotypes. Moreover, clinical trials are needed to develop useful and feasible strategies, as vaccines against EV species or serotypes, to lessen the prevalence and incidence of T1DM.

Limitations

We performed a set of standard and comprehensive literature searches, and made no any language restrictions to limit our ability to evaluate the association between EVs infection and the risk of T1DM. However, there were several limitations in our meta-analysis. First, the included studies were confined to case-control studies with inherent factors, such as different data collection, various detection methods of viral
infections (RT-PCR, specific antiviral neutralising antibodies and hybridization, and samples from different sites. Second, it was true that we performed subgroup and sensitivity analyses to reduce potential confounding factors, but all eligible studies also had a high level of heterogeneity. Third, other environmental agents might alter the risk of T1DM, such as maternal virus infection [40], cow’s milk and vitamin D [41], moreover, it is impossible to improve all of these potential confounders in retrospective studies. Fourth, the results of this meta-analysis cannot prove that EV infection has a cause-and-effect role in the development of clinical T1DM. Despite these limitations, this meta-analysis has increased the statistical power by pooling the findings of a single case-control study with overall good methodological quality, to some extent, which was sufficient evidence to draw this conclusion.

In summary, we demonstrated that EV infection may be a dependent risk factor for clinical T1DM. Further studies are needed to explore the potential pathways and focus on whether the virus vaccine could decrease the risk of clinical T1DM or not.

Author contributions

SY and BY Z were involved in searching databases and screening articles and drafted the first manuscript. BY Z and XL D were engaged in data extraction data and reviewing the quality of included studies with the aid of YL Z. Statistical analysis was carried by SY and Z Z. All authors reviewed this article and contributed the revisions. LW C is the guarantor for this article. SY and BY Z contributed to the work equally and should be regarded as co-first authors.

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Conflict of interest

All of authors have no conflict of interest.

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Data availability statement. The data that support the findings of this study are available as Supplementary Materials.

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Fig. 1. Flow diagram of the literature selection process.
Fig. 2. Forrest plot of the association between clinical T1DM and EV infection

Fig. 3. Funnel plot of the association between clinical T1DM and EV infection
Table 1 summary of individual study investigating type 1 diabetes and enterovirus infection

| Study          | country | cases/controls | age(years) | details of cases | method of detection | details of controls | details of methods to confirm viral infections | Islet Autoantibody |
|----------------|---------|----------------|------------|------------------|--------------------|--------------------|-----------------------------------------------|--------------------|
| Antonio, 1985[16] | Italy   | 22/46          | 0–16       | T1DM onset; Samples were collected from 1981 to 1982 | CB1–6 neutralizing antibodies in serum | NT                | age,sample time-matched healthy children in the same season | ICA,IAA,GAD,IA-2 |
| Gun, 1985[17]   | Sweden  | 24/48          | 2–15       | T1DM onset; Samples were continuously collected from 1982 to 1984 | CB-virus-specific IgM in serum | RI                 | 48 age,sample time-matched nondiabetic children in 24 of which have non-EV infections; the other 24 for planned surgical procedure | 7                  |
| Donn, 1992[18]  | America | 225/163        | 0–29       | T1DM onset; HLA-DR3 typing; Samples were collected from 1984 to 1987 | CB-virus-specific IgM in serum | NT                 | age,sex,sample time and socioeconomic status-matched friends | 8                  |
| Frisk, 1992[19] | Sweden  | 35/47          | case:0–15; control:3–18 | T1DM onset; Samples were collected from 1983–8 | CB-virus-specific IgM in serum | RIA                | siblings | 7 |
| Nairn, 1999[20] | British | 110/182        | 1–17       | T1DM onset; Samples were collected from 1991–7 | EV-RA in serum | RT-PCR             | age,place,sample time-matched nondiabetic children | 8                  |
| Lonnrot, 2000[21] | Finland | 47/34          | 2.5–20     | T1DM onset; Samples were collected from Finnland DiMe study | EV-RA in serum | RT-PCR             | siblings | 7 |
| Miroslav, 2000[11] | Slovak | 336/707 children | T1DM onset; samples were collected from 1978-91; cases and controls were taken from the Slovak National Register of Childhood Diabetes | age-matched healthy children were collected from 1985-98 | NT | CB antibody in serum | 5 |
| Wassim, 2000[22] | Sweden | 56/24 | 12 newly T1DM children and 20 newly T1DM adults; 13 previously T1DM children and 13 previously T1DM adults; samples were collected from 1997 to 1998 | 17 children and 20 adults | RT-PCR | EV-RNA in serum | 8 |
| Maria, 2003[23] | Australia | 206/160 | T1DM onset, HLA-DRB/DQB typing; samples were collected from 1997-9 | children from the community | RT-PCR | EV-RNA in either plasma or stool | ICA, IAA, GAD-65, IA-2 | 9 |
| Moya, 2005[24] | Germany | 47/100 | T1DM onset; autoantibody-positive and negative children for 50,50 respectively | IHC | - | EV-RNA and CB-virus-specific IgM/IgG in serum | GAD-65, ICA, IA-2 | 7 |
| Francesco, 2007[6] | Italy | 6/26 | T1DM onset; 5 multigorgan donors and 1 whole pancreas graft | normal multigorgan donors | IHC | enteroviral VP-1 in pancreatic tissues | 6 |
| Author(s)             | Year | Country | Sample Size | Sample Collection Details | T1DM Onset | Sample Type | Controls | Methodology Details |
|----------------------|------|---------|-------------|---------------------------|------------|-------------|----------|---------------------|
| Elfaitouri et al.     | 2007 | Sweden  | 33/51       | 0~17                      | 24 siblings and 27 healthy children | RT-PCR     | 32 ICA positive relative controls; 31 ICA negative relative controls; 19 healthy subjects, age, sex, place, sample time-matched | GAD-65 7 |
| Liu et al.            | 2007 | Cuba    | 34/25       | case:1~15; control:1~47   | 0~50 duration of T1DM; Samples were collected from 1995-2000 | RT-PCR     | 0~50 duration of T1DM; Samples were collected from 1995-2000 enterovirus in intestinal mucosa | 6 |
| Oikarinen et al.      | 2008 | Finland | 12/10       | case:18~53; control:23~71 | 11 normal neonate; 39 normal children; 69 normal adults | ISH/IHC    | 11 normal neonate; 39 normal children; 69 normal adults enteroviral capsid protein VP1 staining in pancreatic tissue | 7 |
| Richardion et al.     | 2009 | Britain | 72/11       | case:1~42; control:23~71  | 11~42 duration of T1DM; Samples were collected from 2005-2010 | IHC        | 11~42 duration of T1DM; Samples were collected from 2005-2010 | 7 |
| Barbara et al.        | 2010 | The Netherlands | 10/20 | case:5~14; control:6~17 | T1DM duration: 1~57; Samples were collected from 2003-4 | RT-PCR     | 1~57 hospitalized children without endocrine disorders; HLA typing | 5 |
| Mercalli et al.       | 2012 | Italy   | 25/48       | case:3~77; control:1~67   | T1DM duration: 0~38; HLA typing; Samples were collected from 2005-6 | ISH/R T-PCR/IHC | T1DM duration: 0~38; HLA typing; Samples were collected from 2005-6 EV-RNA/V-P1 in intestinal mucosa | GADA, I-A/2A, IAA, TGG, TGA, TMA 7 |
| Maarit, Finland       | 2012 | Finland | 39/81       | case:18~63; control:18~76 | T1DM duration: 1~57; Samples were collected from 2005-6 | ISH/R T-PCR/IHC | T1DM duration: 1~57; Samples were collected from 2005-6 EV-RNA and VP1 protein in intestinal mucosa | 7 |
| Author(s) | Year | Country | Case:Control | Sample Collection | Sample Type | EV-R NA in plasma | GADA,IAA-2,IAA,ZnT8 | RT-PCR EV-R NA in plasma |
|-----------|------|---------|--------------|-------------------|-------------|-------------------|---------------------|------------------------|
| Salvatore, 2013 | 2013 | Italy | 24/11 6 | case:6~13, control:4.9~46 | T1DM onset; Samples were collected from 2010-2015 | 20 siblings, 41 parents, 29 non-diabetic children and 26 healthy adults; age, place, sample time-matched | | a |
| Sami, 2014 | 2014 | European | 249/249 | case:1.1~22.7, control:1.0~25.5 | T1DM onset; HLA-DR3/DR4 genotype; cases and controls were taken from European VirDiab Study | age, sex, place, sampling time-matched children | | |
| Cekin, 2014 | 2014 | Turkish | 86/10 0 | 9.9±2.3 | 24 T1DM onset; 62 previously T1DM | age, sex, mat ched children | | |
| Imen, 2017 | 2017 | Tunisian | 95/14 1 | 1~48 | T1DM duration: 0~30, 41 children, 54 adults | 57 children and 84 adults | | |
| Waled, 2018 | 2018 | Egyptian | 382/100 | case:2~16, control:3~14 | T1DM; Samples were collected from 2013-2014 | sex, age, mat ched children | | |
| Giovanni, 2018 | 2018 | Italian | 82/11 7 | 2.1~18 | T1DM | sex, age, place, sample time-matched short stature or minor trauma children | | |
| Murat, 2018 | 2018 | Turkish | 40/30 1~16 | T1DM onset | sample time and place-matched healthy children | | | |

T1DM = type 1 diabetes mellitus; RT-PCR = reverse transcription polymerase chain reaction; GADA = antiautoantibodies to GAD; EV = enterovirus; ICA = antiautoantibodies to ICA; IAA = antiautoantibodies to IAA; ZnT8 = antiautoantibodies to ZnT8; EV-R NA = enterovirus neutralization assay; IFA = immunofluorescence assay; IHC = immunohistochemistry; CB = group B coxsackievirus; IgM = IgM antiautoantibodies to EV; RT-PCR EV-R NA = reverse transcription polymerase chain reaction for EV-R NA.

Table 2 Summary Odds Ratios and Heterogeneity for association of EV Infection and Clinical T1DM in subgroup and sensitivity analyses.
| Variables                        | No. of Studies | OR(95%CI)         | Weight(%) | I² (%) | P for heterogeneity |
|---------------------------------|----------------|-------------------|-----------|--------|---------------------|
| Geographical areas              |                |                   |           |        |                     |
| Europe                          | 18             | 5.72(2.94–11.14)  | 70.4      | 83     | <0.001              |
| Non- Europe                     | 7              | 6.02(3.02–12.01)  | 29.6      | 75     | <0.001              |
| Age distribution(years)         |                |                   |           |        |                     |
| 0–9                             | 2              | 33.82(1.87–612.91)| 6.4       | 74     | <0.001              |
| 0–20                            | 14             | 4.89(2.51–9.51)   | 52.9      | 83     | <0.001              |
| 0–71                            | 11             | 7.53(3.61–15.72)  | 40.7      | 81     | <0.001              |
| The initial time of clinical    |                |                   |           |        |                     |
| T1DM                            |                |                   |           |        |                     |
| Newly clinical T1DM             | 18             | 4.76(2.84–7.98)   | 81.9      | 82     | <0.001              |
| Previously clinical T1DM        | 4              | 4.91(2.49–9.67)   | 18.1      | 32     | 0.22                |
| Methods of EV detection         |                |                   |           |        |                     |
| NT                              | 5              | 1.58(0.76–3.30)   | 25.1      | 89     | <0.001              |
| RIA                             | 2              | 3.02(0.17–54.64)  | 8.4       | 91     | <0.001              |
| ISH                             | 2              | 5.21(2.31–11.79)  | 6.5       | 0      | 0.350               |
| IHC                             | 5              | 7.29(1.42–37.58)  | 16        | 81     | <0.001              |
| RT-PCR                          | 13             | 7.48(4.20–13.32)  | 44        | 63     | 0.002               |
| Source of EV sample             |                |                   |           |        |                     |
| Serum                           | 14             | 3.90(3.24–4.70)   | 73.3      | 83     | <0.001              |
| Intestinal Musoca               | 3              | 4.36(2.24–8.49)   | 5.2       | 50     | 0.14                |
| Plasma                          | 5              | 4.49(3.12–6.46)   | 19.1      | 77     | 0.002               |
| PBMC                            | 2              | 11.42(4.27–30.58) | 1.5       | 0      | 0.52                |
| Pancretic tissue                | 2              | 27.60(8.48–89.78) | 0.9       | 0      | 0.67                |
| Type of controls                |                |                   |           |        |                     |
| Relatives                       | 3              | 3.19(1.65–6.14)   | 31.6      | 0      | 0.75                |
| Normal Subjects                 | 3              | 56.41(3.54–899.49)| 27.5      | 87     | <0.001              |
| autoantibody positive individuals | 2          | 1.07(0.43–2.67)   | 21.5      | 40     | 0.20                |
| autoantibody negative           | 2              | 2.29(0.15–35.84)  | 19.4      | 83     | 0.01                |
| Coxsackievirus Serotypes |   |          |   |          |   |
|-------------------------|---|----------|---|----------|---|
| CB1                     | 4 | 1.76(1.18--2.63) | 14.8 | 0 | 0.67 |
| CB2                     | 4 | 0.90(0.50--1.64)  | 19 | 25 | 0.26 |
| CB3                     | 4 | 0.95(0.68--1.33)  | 18.5 | 0 | 0.51 |
| CB4                     | 5 | 2.03(0.87--4.75)  | 20.9 | 60 | 0.04 |
| CB5                     | 4 | 0.91(0.37--2.21)  | 17.4 | 43 | 0.15 |
| CB6                     | 2 | 0.88(0.50--1.53)  | 9.3 | 0 | 0.87 |

Sample size

|   |          |          |  |          | <0.001 |
|---|----------|----------|---|----------|--------|
| <100 | 12 | 5.88(3.46--9.99) | 57.7 | 83 |        |
| >100 | 13 | 6.00(2.21--16.26) | 42.3 | 80 | <0.001 |

NOS score

|   |          |          |  |          | <0.001 |
|---|----------|----------|---|----------|--------|
| 9--10 | 6 | 5.89(2.42--14.38) | 27.4 | 81 | <0.001 |
| 8 | 13 | 5.12(2.10--12.49) | 50.6 | 55 | <0.001 |
| 6--7 | 6 | 7.93(3.75--16.78) | 22.0 | 61 | 0.02  |

Study type

|   |          |          |  |          | <0.001 |
|---|----------|----------|---|----------|--------|
| CC | 22 | 7.49(4.20--13.36) | 85.9 | 80 | <0.001 |
| NCC | 3 | 2.16(0.95--4.92) | 14.1 | 83 | <0.001 |

T1DM=type 1 diabetes mellitus; NT=neutralization test; RIA=radioimmunoassay; IFA=immunofluorescence assay; IHC=immunohistochemistry; RT-PCR=reverse transcription polymerase chain reaction; CB=group B coxsackievirus; CC=case-control study; NCC=nested case-control study