Identification of Optically Active Pyrimidine Derivatives as Selective 5-HT\textsubscript{2C} Modulators

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Abstract: A series of pyrimidine derivatives 4a–i were synthesized and evaluated for their binding affinities towards 5-HT\textsubscript{2C} receptors. With regard to designed molecules 4a–i, the influence of the size of alkyl ether and the absolute configuration of a stereogenic center on the 5-HT\textsubscript{2C} binding affinity and selectivity was studied. The most promising diastereomeric mixtures 4d and 4e were selected in the initial radioligand binding assay and they were further synthesized as optically active forms starting from optically active alcohols 5d and 5e, prepared by an enzymatic kinetic resolution. Pyrimidine analogue (R,R)-4e displayed an excellent 5-HT\textsubscript{2C} binding affinity with good selectivity values against a broad range of other 5-HT receptor subtypes.

Keywords: pyrimidine; optically active; enzymatic kinetic resolution; 5-HT\textsubscript{2C} receptor; binding affinity; selectivity

1. Introduction

Serotonin receptors, also known as 5-hydroxytryptamine (5-HT) receptors, are mainly located in the central nervous system (CNS) and play an important role in mediating both excitatory and inhibitory neurotransmission. There are 14 different 5-HT receptors classified into seven major subfamilies (5-HT\textsubscript{1–7}). They have been known to regulate various physiological functions such as mood, depressive behavior, appetite, biorhythm, and feeding [1,2]. According to recent studies, the 5-HT\textsubscript{2C} receptor (5-HT\textsubscript{2C}R) is expected to be a potential drug target for the diagnosis and treatment of a number of CNS disorders including schizophrenia, depression, substance abuse, and Parkinson diseases, as well as obesity and urinary incontinence [3–8]. In particular, 5-HT\textsubscript{2C}-specific modulators may have few undesired side effects on peripheral tissues because this receptor is exclusively expressed in the CNS [9,10]. However, the 5-HT\textsubscript{2C} receptor belongs to the 5-HT\textsubscript{2} receptor family, together with 5-HT\textsubscript{2A} and 5-HT\textsubscript{2B}, which have a high similarity in terms of their amino acid sequences [11]. It has been reported that the activation of 5-HT\textsubscript{2A} and 5-HT\textsubscript{2B} is strongly implicated in hallucinations and valvular heart disease [12,13]. Thus, the discovery of 5-HT\textsubscript{2C} agonists with a high specificity and subtype selectivity for 5-HT\textsubscript{2A} and 5-HT\textsubscript{2B} receptors is important for avoiding side effects.
To date, several compounds, including lorcaserin 1 and vabicaserin 2, have been identified as 5-HT_{2C} agonists with significant selectivity against 5-HT_{2A} and 5-HT_{2B} (Figure 1) [14–18]. Lorcaserin (ADP-356) was developed for the treatment of obesity and was approved for clinical use by FDA in 2012 [19]. Vabicaserin is a drug developed for the treatment of acute schizophrenia or appetite suppressants, but has not been shown to be effective in clinical trial studies [20,21]. In addition, other small molecule 5-HT_{2C} agonists have been reported to be in preclinical or early clinical development [15–18].

![Figure 1. The chemical structures of representative 5-HT_{2C} receptor ligands and their binding affinities for 5-HT_{2} receptor subtypes [19,21].](image)

During our efforts toward the development of 5-HT receptor modulators, we have initiated a program to discover 5-HT_{2C} agonists as potential therapeutic and diagnostic agents for CNS diseases. Recently, pyrimidine analogue 3 with good pharmacological and pharmacokinetic properties has been reported as a potential 5-HT_{2C} agonist [22]. Our in vitro study of this molecule proved that it has a high potency against the 5-HT_{2C} receptor and relative good selectivity against the 5-HT_{2A} receptor [23]. However, the binding affinity of 3 to the 5-HT_{2B} receptor is still too high, which prompted us to investigate this compound for the development of 5-HT_{2C} selective agonists. Based on the report of the activation selectivity of 5-HT_{2C} over 5-HT_{2B} [11], we postulated that altering the chain length between the phenyl group and pyrimidine may induce a subtle structural change in the molecule, which can make a large difference in its interaction with 5-HT_{2C} and 5-HT_{2B} because the binding site of 5-HT_{2C} is slightly deeper than that of 5-HT_{2B} [11]. Thus, pyrimidine derivatives 4 with hydrocarbon chains shorter or longer than that of the parent molecule 3 were designed to examine the effect of the structural modification of compound 3 on the target selectivity toward 5-HT_{2C} over 5-HT_{2B} (Figure 2). In this paper, we report our progress in the synthesis and biological evaluation of pyrimidine derivatives 4 as selective 5-HT_{2C} agonists.

![Figure 2. The structures of reported 5-HT_{2C} agonist 3 and designed compounds 4.](image)

2. Results and Discussion

2.1. Synthesis of Pyrimidine Derivatives

The synthesis of compounds 4a–i is described in Scheme 1. Following the literature procedure [22,24], a series of pyrimidine derivatives 4a–i were synthesized starting from either primary or secondary alcohols 5a–i. The alcohols 5a–e were obtained from commercial sources, whereas the others 5f–i were easily prepared via the reduction/oxidation of phenyl propanoic acids 8 and 9. Thus, alcohols 5a–i were first reacted with 2,4-dichloro-5-fluoropyrimidine in the presence of NaO/Bu to
afford 4-alkoxypyrimidines 6a–i. A nucleophilic aromatic substitution reaction (SNAr) of 6a–i with (R)-(+)-1-Boc-3-methylpiperazine gave 2-amino-4-alkoxypiperazine 7a–i. Finally, the BOC group in 7a–i was removed in the presence of trifluoroacetic acid or 4 M hydrochloric acid to furnish the corresponding pyrimidine derivatives 4a–i.

Scheme 1. Synthesis of the pyrimidine derivatives 4a–i. Reagents and conditions: (a) 2,4-dichloro-5-fluoropyrimidine, NaOEt, toluene, 0 °C, 70–91%; (b) (R)-(+)-1-Boc-3-methylpiperazine, DIPEA, CH3CN, 110 °C, 14–53%; (c) TFA, CH2Cl2, r.t., 52–72%; (d) 4 M HCl, dioxane, 0 °C, 53%; (e) BH3-Me2S, THF, r.t., 84–94%; (f) PCC, CH2Cl2, r.t., 73–77%; (g) MeMgBr, THF, 0 °C, 72–85%.

2.2. Biological Evaluation

The serotonin receptor binding affinity of our synthesized compounds 4a–i was examined by a radioligand binding assay in transfected CHO-K1 cell lines using [3H]mesulergine as a radioligand. Practically, a displacement of radioligand with compounds 4a–i was first evaluated at a concentration of 10 μM, and then their Ki values were determined on the basis of the dose-response curves. As summarized in Table 1, compounds 4d and 4e showed the highest binding affinities to 5-HT2C and good selectivity values for 5-HT2A. However, the binding affinities of 4d and 4e to 5-HT2B were comparable to the value of 3. At this moment, we assumed that both diastereomeric mixtures 4d and 4e might have a negative influence on the selectivity for 5-HT2B. Thus, we planned to synthesize each diastereomer derived from 4d and 4e as an optically pure form to test its in vitro activity against 5-HT2 receptors.

Table 1. Binding affinities of pyrimidine analogues 4a–i against 5-HT2 receptor subtypes.

| Entry | Comp. | n | F | % Binding | 5-HT2A K_i (nM) | 5-HT2B K_i (nM) | 5-HT2C K_i (nM) |
|-------|-------|---|---|-----------|----------------|----------------|-----------------|
| 1     | 4a    | 0 | 2-F | 93.1        | 80.0           | 96.9           | 56.0            | 97.8           | 31.0           |
| 2     | 4b    | 0 | 3-F | 97.4        | 43.0           | 98.5           | 14.0            | 99.0           | 5.1            |
| 3     | 4c    | 0 | 4-F | 94.6        | 51.0           | 97.6           | 14.0            | 98.8           | 22.0           |
| 4     | 4d    | 0 | 3-F | 91.9        | 213.0          | 98.9           | 4.8             | 99.3           | 2.7            |
| 5     | 4e    | 0 | 4-F | 97.6        | 138.0          | 95.6           | 24.0            | 97.8           | 20.0           |
| 6     | 4f    | 2 | 3-F | 96.5        | 90.0           | 97.7           | 22.0            | 97.9           | 22.0           |
| 7     | 4g    | 2 | 4-F | 96.5        | 170.0          | 98.7           | 16.0            | 98.6           | 14.0           |
| 8     | 4h    | 2 | 3-F | 94.3        | 138.0          | 97.6           | 22.0            | 95.0           | 12.0           |
| 9     | 4i    | 2 | 4-F | 96.9        | 173.0          | 97.6           | 22.0            | 95.0           | 12.0           |
| 10    | 4j    | 1 | 3-F | 95.6        | 128.0          | 97.4           | 7.9             | 98.2           | 0.7            |
2.3. Synthesis and In Vitro Evaluation of Optically Active Pyrimidines

In order to synthesize 4d and 4e as optically active diastereomers, optically active secondary alcohols 5d and 5e should be prepared. For this purpose, we initially attempted the separation of diastereomers 10 and 11, which were synthesized by the reaction of 5d and 5e with (R)-(-)-O-acetylmadelic acid using EDCI as a coupling reagent. However, diastereomeric mixtures 10 and 11 were not completely separated by column chromatography on silica gel. Alternatively, the enzymatic kinetic resolution was applied to separate racemic 5d and 5e, as shown in Scheme 2. It has been reported that CAL-B lipase can selectively acetylate the (R)-form of secondary benzyl alcohols using vinyl acetate as an acyl transfer reagent (Scheme 2) [25,26]. According to the literature procedure, the selective acetylation reactions of racemic secondary alcohols 5d and 5e were performed using 0.5 equivalent of vinyl acetate, pyridine, and the CAL-B enzyme in hexane. Enantiomeric pure acetate was obtained by further acetylation of the remaining alcohols with an excess of vinyl acetate and CAL-B enzyme followed by chromatographic separation.

Scheme 2. The synthesis of (R)/(S)-5d and 5e using enzymatic kinetic resolution. Reagents and conditions: (a) vinyl acetate (0.5 eq), CAL-B, pyridine, hexane, r.t.; (b) 1 M NaOH, MeOH, r.t., 17–32% (two steps); (c) vinyl acetate (1 eq), CAL-B, pyridine, hexane, r.t., then separation, 44–46%; (d) (R)-(−)-O-acetylmadelic acid, DCC, DMAP, CH2Cl2, r.t., 68–86%.

For determining the optical purity of (R)/(S)-5d and 5e, they were converted to the corresponding mandelic ester (R)/(S)-10 and 11, respectively. A 1H-NMR analysis of diastereomeric protons in (R)/(S)-10 and 11 indicated that the ee’s of (R)/(S)-5d and 5e were greater than 93%. We also confirmed that the optical rotation values of our compounds (R)/(S)-5d and 5e are almost identical to those of the compounds reported in the literature.
With each enantiomer (R)- or (S)-5d and 5e in hand, the optically active pyrimidine derivatives 4d–4e were synthesized following the same reaction sequences (Scheme 3). Finally, compounds 4d and 4e were assessed for their binding affinity to 5-HT2 receptor subtypes by radioligand binding assays. The in vitro assay results are demonstrated in Table 2. In general, (R,R)-4d and 4e prepared from secondary alcohols (R)-5d and (R)-5e showed excellent binding affinities to 5-HT2C, whereas (S,R)-forms of 4d and 4e exhibited low potencies for 5-HT2A and 5-HT2B. A further evaluation of these compounds for other 5-HT receptor subtypes was also performed and is provided in the supplementary data (Table S2). These results combined with the in vitro data in Table 1 suggested that pyrimidine derivatives 4 with a short alkyl chain could maintain their binding affinity to 5-HT2C comparable to that of 3 and the binding affinity for other 5-HT subtypes could be significantly influenced by an absolute configuration of the stereogenic methyl group in 4. Considering the potencies and selectivities of 4, we can conclude that (R,R)-4e would be a viable candidate for a selective 5-HT2C modulator.

![Scheme 3. Synthesis of the optically active pyrimidine derivatives 4d–4e. Reagents and conditions: (a) 2,4-dichloro-5-fluoropyrimidine, NaOtBu, toluene, 0 °C, 55–72%; (b) (R)-(+)1-Boc-3-methylpiperazine, toluene, 150 °C, 44–57%; (c) TFA, CH2Cl2, r.t., 67–71%; (d) HCl, dioxane, 0 °C, 81–82%.

![Table 2. Binding affinities of optically active pyrimidines 4d and 4e against 5-HT2 receptor subtypes](attachment:image.png)

| Entry | Comp. | n | F | R | 5-HT2A % Binding | Kᵢ (nM) | 5-HT2B % Binding | Kᵢ (nM) | 5-HT2C % Binding | Kᵢ (nM) |
|-------|-------|---|---|---|------------------|--------|------------------|--------|------------------|--------|
| 1     | (R,R)-4d | 0  | 3-F | Me (R) | 93.3 | 222.0 | 100.2 | 2.6 | 98.5 | 1.2 |
| 2     | (S,R)-4d | 0  | 3-F | Me (S) | 82.4 | 475.0 | 95.2 | 67.0 | 97.7 | 14.0 |
| 3     | (R,R)-4e | 0  | 3-F | Me (R) | 0.9 | 19.0 | 99.6 | 19.0 | 94.2 | 4.0 |
| 4     | (S,R)-4e | 0  | 4-F | Me (S) | 64.8 | 1024.0 | 95.3 | 128.0 | 97.8 | 23.0 |

3. Materials and Methods

3.1. General Methods

All reactions were conducted under oven-dried glassware under an atmosphere of nitrogen. All commercially available reagents were purchased and used without further purification. Solvents and gases were dried according to standard procedures. Organic solvents were evaporated with reduced pressure using a rotary evaporator. Reactions were followed by analytical thin layer chromatography (TLC) analysis using glass plates precoated with silica gel (0.25 mm). TLC plates were visualized by exposure to UV light (UV), and were then visualized with a KMnO₄ or p-anisaldehyde stain followed by brief heating on a hot plate. Flash column chromatography was performed using silica gel 60 (230–400 mesh, Merck, Darmstadt, Germany) with the indicated solvents. ¹H-NMR spectra were measured with 400MHz and ¹³C-NMR spectra were measured with 100MHz using CDCl₃ and MeOD. ¹H-NMR spectra are represented as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), integration, and coupling constant (J) in Hertz (Hz). ¹H-NMR chemical shifts are reported relative to CDCl₃ (7.26 ppm). ¹³C NMR was recorded relative to the central line of CDCl₃ (77.0 ppm). High resolution mass spectra (HR-MS) were obtained using...
positive electrospray ionization and mass/charge (m/z) ratios that are reported as values in atomic mass units.

3.2. Synthesis of Pyrimidines 4a–i

3.2.1. General Procedure for Preparing Compounds 5f and 5g

To a solution of 3-(3 or 4-fluorophenyl)propanoic acid 8 or 9 (1.78 mmol) in THF (8.90 mL), borane-dimethyl sulfide (3.57 mmol) was added dropwise. The reaction mixture was allowed to stir at room temperature for 1 h. After completion of the reaction (monitored by TLC), the mixture was slowly quenched with MeOH until bubbling ceased, extracted with EtOAc. The organic layers were dried over anhydrous MgSO\(_4\) and concentrated in vacuo. The resulting residue was purified by flash chromatography on silica gel (EtOAc:n-hexane = 1:4) to afford propanol 5f or 5g.

3-(3-Fluorophenyl)propan-1-ol (5f): Yield: 84%; \(^1\)H-NMR (400 MHz, CDCl\(_3\)) \(\delta\) 7.26–7.21 (m, 1H), 6.97 (d, \(J = 7.6\) Hz, 1H), 6.92–6.86 (m, 2H), 3.68 (q, \(J = 5.8\) Hz, 2H), 2.72 (t, \(J = 7.7\) Hz, 2H), 1.92–1.85 (m, 2H). \(^{13}\)C-NMR (100 MHz, CDCl\(_3\)) \(\delta\) 162.9 (d, \(J = 243\) Hz), 144.4 (d, \(J = 7\) Hz), 129.8 (d, \(J = 8\) Hz), 124.1 (d, \(J = 3\) Hz), 115.2 (d, \(J = 21\) Hz), 112.7 (d, \(J = 21\) Hz), 62.0, 33.9, 31.7 (d, \(J = 2\) Hz).

3-(4-Fluorophenyl)propan-1-ol (5g): Yield: 94%; \(^1\)H-NMR (400 MHz, CDCl\(_3\)) \(\delta\) 7.17–7.14 (m, 2H), 7.00–6.95 (m, 2H), 3.67 (t, \(J = 6.4\) Hz, 2H), 2.69 (t, \(J = 7.8\) Hz, 2H), 3.67 (t, \(J = 6.4\) Hz, 2H), 1.91–1.84 (m, 2H). \(^{13}\)C-NMR (100 MHz, CDCl\(_3\)) \(\delta\) 161.6 (d, \(J = 241.6\) Hz), 137.7 (d, \(J = 3.1\) Hz), 130.0 (d, \(J = 7.6\) Hz), 115.4 (d, \(J = 21\) Hz), 62.3, 34.6, 31.5.

3.2.2. General Procedure for Preparing Compounds 5f' and 5g'

To a solution of propanol 5f or 5g (0.482 mmol) in CH\(_2\)Cl\(_2\) (4.80 mL), PCC (0.964 mmol) was added at 0 \(^\circ\)C. The reaction mixture was allowed to stir at room temperature for 1 h. After completion of the reaction (monitored by TLC), the mixture was filtered with silica gel and a celite pad, and extracted with ether. The organic layers were dried over anhydrous MgSO\(_4\) and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (EtOAc:n-hexane = 1:4) to afford propanal 5f' or 5g'.

3-(3-Fluorophenyl)propanal (5f'): Yield: 77%; \(^1\)H-NMR (400 MHz, CDCl\(_3\)) \(\delta\) 9.82 (t, \(J = 1.2\) Hz, 1H), 7.27–7.22 (m, 1H), 6.97 (d, \(J = 7.8\) Hz, 1H), 6.91–6.88 (m, 2H), 2.95 (t, \(J = 7.5\) Hz, 2H), 2.80–2.77 (m, 2H). \(^{13}\)C-NMR (100 MHz, CDCl\(_3\)) \(\delta\) 201.0 (d, \(J = 243\) Hz), 142.9 (d, \(J = 8\) Hz), 130.4 (d, \(J = 8\) Hz), 124.0 (d, \(J = 2\) Hz), 115.2 (d, \(J = 21\) Hz), 113.2 (d, \(J = 21\) Hz), 44.9, 27.8 (d, \(J = 2\) Hz).

3-(4-Fluorophenyl)propanal (5g'): Yield: 73%; \(^1\)H-NMR (400 MHz, CDCl\(_3\)) \(\delta\) 9.77 (t, \(J = 1.2\) Hz, 1H), 7.13–7.09 (m, 2H), 6.96–6.91 (m, 2H), 2.91–2.87 (m, 2H), 2.72 (t, \(J = 7.5\) Hz, 2H). \(^{13}\)C-NMR (100 MHz, CDCl\(_3\)) \(\delta\) 201.2, 161.5 (d, \(J = 243\) Hz), 136.0 (d, \(J = 3\) Hz), 129.7 (d, \(J = 8\) Hz), 115.3 (d, \(J = 21\) Hz), 45.4, 27.3.

3.2.3. General Procedure for Preparing Compounds 5h and 5i

To a solution of propanal 5f' or 5g' (0.204 mmol) in THF (2.00 mL), MeMgBr (3.0 M in ether: 0.245 mmol) was added dropwise at 0 \(^\circ\)C. The reaction mixture was allowed to stir at room temperature for 1 h. After completion of the reaction (monitored by TLC), it was quenched with saturated aqueous NH\(_4\)Cl, extracted with EtOAc and washed with brine. The organic layers were dried over anhydrous MgSO\(_4\) and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (EtOAc:n-hexane = 1:2) to afford butan-2-ol 5h or 5i.

4-(3-Fluorophenyl)butan-2-ol (5h): Yield: 85%; \(^1\)H-NMR (400 MHz, CDCl\(_3\)) \(\delta\) 7.26–7.20 (m, 1H), 6.97 (d, \(J = 7.7\) Hz, 1H), 6.92–6.85 (m, 2H), 3.82 (d, \(J = 2.3\) Hz, 1H), 2.80–2.63 (m, 2H), 1.79–1.73 (m, 2H), 1.24 (d, \(J = 6.2\) Hz, 3H).
4-(4-Fluorophenyl)butan-2-ol (5i): Yield: 72%; 1H-NMR (400 MHz, CDCl₃) δ 7.12 (dd, J = 5.6, 8.2 Hz, 2H), 6.93 (t, J = 8.7 Hz, 2H), 3.82–3.75 (m, 1H), 2.74–2.58 (m, 2H), 1.75–1.68 (m, 2H), 1.20 (d, J = 6.2 Hz, 3H). 13C-NMR (100 MHz, CDCl₃) δ 161.3 (d, 1J = 241 Hz), 137.7 (d, 2J = 3 Hz), 129.7 (d, 3J = 8 Hz), 115.1 (d, 2J = 21 Hz), 67.4, 40.9, 31.3, 23.7.

3.2.4. General Procedure for Preparing Compounds 6a–i

A solution of sodium tert-butoxide (2.72 mmol) in toluene (18.2 mL) was treated with primary or secondary alcohol (1.82 mmol) dropwise at 0 °C. After 5 min, 2,4-dichloro-5-fluoropyrimidine (2.18 mmol) was added to the mixture. The reaction mixture was allowed to stir at room temperature for 1 h. After completion of the reaction (monitored by TLC), it was quenched with saturated aqueous NH₄Cl, extracted with EtOAc, and washed with brine. The organic layers were dried over anhydrous MgSO₄ and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (EtOAc:n-hexane = 1:8) to afford pyrimidine 6.

2-Chloro-5-fluoro-4-(2-fluorobenzyl)oxy)pyrimidine (6a): Yield: 91%; 1H-NMR (400 MHz, CDCl₃) δ 8.21 (d, J = 2.2 Hz, 1H), 7.53–7.49 (m, 1H), 7.49–7.34 (m, 1H), 7.20–7.09 (m, 2H), 5.57 (s, 2H). 13C-NMR (100 MHz, CDCl₃) δ 161.2 (d, 1J = 247 Hz), 159.0 (d, 2J = 12 Hz), 153.2 (d, 3J = 5 Hz), 145.9 (d, 1J = 263 Hz), 144.3 (d, 2J = 20 Hz), 131.1 (d, 3J = 4 Hz), 131.0 (d, 3J = 8 Hz), 124.3 (d, 4J = 4 Hz), 121.8 (d, 2J = 14 Hz), 115.7 (d, 2J = 20 Hz), 63.9 (d, 3J = 4 Hz).

2-Chloro-5-fluoro-4-((3-fluorobenzyl)oxy)pyrimidine (6b): Yield: 90%; 1H-NMR (400 MHz, CDCl₃) δ 8.23 (d, J = 2.2 Hz, 1H), 7.40–7.34 (m, 1H), 7.25 (d, J = 7.26 Hz, 1H), 7.19 (d, J = 9.4 Hz, 1H), 7.09–7.04 (m, 1H), 5.50 (s, 2H). 13C-NMR (100 MHz, CDCl₃) δ 162.9 (d, 1J = 245 Hz), 158.9 (d, 2J = 11 Hz), 153.2 (d, 3J = 5 Hz), 145.8 (d, 1J = 245 Hz), 144.4 (d, 2J = 20 Hz), 136.9 (d, 3J = 8 Hz), 130.3 (d, 3J = 8 Hz), 123.9 (d, 4J = 3 Hz), 115.7 (d, 2J = 21 Hz), 115.3 (d, 2J = 22 Hz), 68.9 (d, 3J = 2 Hz).

2-Chloro-5-fluoro-4-((4-fluorobenzyl)oxy)pyrimidine (6c): Yield: 75%; 1H-NMR (400 MHz, CDCl₃) δ 8.20 (d, J = 2.2 Hz, 1H), 7.47 (dd, J = 5.4 Hz, 8.7 Hz, 2H), 7.08 (t, J = 8.7 Hz, 2H), 5.47 (s, 2H). 13C-NMR (100 MHz, CDCl₃) δ 163 (d, 1J = 246 Hz), 159.03 (d, 2J = 12 Hz), 153.2 (d, 3J = 5 Hz), 145 (d, 1J = 263 Hz), 144.3 (d, 2J = 20 Hz), 130.8 (d, 3J = 9 Hz), 130.3 (d, 3J = 4 Hz), 115.2 (d, 2J = 21 Hz), 69.2.

2-Chloro-5-fluoro-4-(1-(3-fluorophenylethoxy)pyrimidine (6d): Yield: 79%; 1H-NMR (400 MHz, CDCl₃) δ 8.17 (d, J = 2.2 Hz, 1H), 7.35–7.30 (m, 1H), 7.22 (d, J = 7.7 Hz, 1H), 7.17–7.14 (m, 1H), 7.02–6.97 (m, 1H), 6.31–6.27 (m, 1H), 1.70 (d, J = 6.6 Hz, 3H). 13C-NMR (100 MHz, CDCl₃) δ 162.9 (d, 1J = 245 Hz), 158.6 (d, 2J = 11 Hz), 153.2 (d, 3J = 5 Hz), 146.0 (d, 1J = 263 Hz), 144.4 (d, 2J = 20 Hz), 142.9 (d, 3J = 7 Hz), 130.2 (d, 4J = 8 Hz), 122.0 (d, 4J = 2 Hz), 115.3 (d, 2J = 21 Hz), 113.3 (d, 2J = 22 Hz), 75.7 (d, 4J = 2 Hz), 22.2.

2-Chloro-5-fluoro-4-(1-(4-fluorophenylethoxy)pyrimidine (6e): Yield: 89%; 1H-NMR (400 MHz, CDCl₃) δ 8.35 (d, J = 2.2 Hz, 1H), 7.66 (dd, J = 5.3, 8.7 Hz, 2H), 7.29–7.25 (m, 2H), 6.54–6.49 (m, 1H), 1.92 (d, J = 6.6 Hz, 3H). 13C-NMR (100 MHz, CDCl₃) δ 162.6 (d, 1J = 245 Hz), 158.6 (d, 2J = 11 Hz), 153.1 (d, 3J = 5 Hz), 146.0 (d, 1J = 262 Hz), 144.3 (d, 2J = 20 Hz), 136.1 (d, 4J = 3 Hz), 128.4 (d, 3J = 8 Hz), 115.6 (d, 2J = 22 Hz), 75.9, 22.1.

2-Chloro-5-fluoro-4-(3-(3-fluorophenyl)propoxy)pyrimidine (6f): Yield: 86%; 1H-NMR (400 MHz, CDCl₃) δ 8.19 (d, J = 2.2 Hz, 1H), 7.28–7.22 (m, 1H), 6.98 (d, J = 7.7 Hz, 1H), 6.92–6.87 (m, 2H), 4.48 (t, J = 6.4 Hz, 2H), 2.80 (t, J = 7.6 Hz, 2H), 2.19–2.12 (m, 2H). 13C-NMR (100 MHz, CDCl₃) δ 163.0 (d, 1J = 244 Hz), 159.4 (d, 2J = 11 Hz), 153.3 (d, 3J = 5 Hz), 145.9 (d, 1J = 263 Hz), 144.0 (d, 2J = 20 Hz), 143.3 (d, 3J = 7 Hz), 123.0 (d, 4J = 8 Hz), 124.1 (d, 4J = 3 Hz), 115.3 (d, 2J = 21 Hz), 113.1 (d, 2J = 21 Hz), 67.6, 31.6 (d, 4J = 1 Hz), 29.6.

2-Chloro-5-fluoro-4-(3-(4-fluorophenyl)propoxy)pyrimidine (6g): Yield: 93%; 1H-NMR (400 MHz, CDCl₃) δ 8.17 (d, J = 4.2 Hz, 1H), 7.16–7.12 (m, 2H), 6.99–6.93 (m, 2H), 4.45 (t, J = 6.4 Hz, 2H), 2.75 (t, J = 7.6 Hz, 2H), 2.14–2.10 (m, 2H). 13C-NMR (100 MHz, CDCl₃) δ 161.4 (d, 1J = 246 Hz), 159.4 (d, 2J = 11 Hz), 153.3
(R)-tert-Butyl-4-(5-fluoro-4-(1-(3-fluorophenyl)ethoxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate

2-Chloro-5-fluoro-4-(((2-fluorobenzyl)oxy)pyrimidine (6h): Yield: 70%; 1H-NMR (400 MHz, CDCl3) δ 8.17 (d, J = 2.3 Hz, 1H), 7.26–7.20 (m, 1H), 6.95–6.87 (m, 3H), 5.40–5.35 (m, 1H), 2.79–2.69 (m, 2H), 2.18–2.11 (m, 1H), 2.02–1.96 (m, 1H), 1.43 (d, J = 6.2 Hz, 3H). 13C-NMR (100 MHz, CDCl3) δ 163.0 (d, J = 5 Hz), 159.2 (d, J = 11 Hz), 153.3 (d, J = 5 Hz), 146.0 (d, J = 262 Hz), 144.1 (d, J = 20 Hz), 143.6 (d, J = 7 Hz), 129.9 (d, J = 8 Hz), 124.0 (d, J = 3 Hz), 115.2 (d, J = 21 Hz), 113.0 (d, J = 21 Hz), 75.1, 37.0, 31.4 (d, J = 1 Hz), 19.6.

2-Chloro-5-fluoro-4-(((4-fluorophenyl)butan-2-yl)oxy)pyrimidine (6i): Yield: 79%; 1H-NMR (400 MHz, CDCl3) δ 8.17 (d, J = 2.3 Hz, 1H), 7.14–7.11 (m, 2H), 6.99–6.94 (m, 2H), 5.40–5.35 (m, 1H), 2.78–2.65 (m, 2H), 2.17–2.09 (m, 1H), 2.00–1.92 (m, 1H), 1.42 (d, J = 6.2 Hz, 3H). 13C-NMR (100 MHz, CDCl3) δ 161.4 (d, J = 242 Hz), 159.2 (d, J = 11 Hz), 153.3 (d, J = 5 Hz), 146.0 (d, J = 262 Hz), 144.0 (d, J = 21 Hz), 136.6 (d, J = 3 Hz), 129.7 (d, J = 8 Hz), 115.2 (d, J = 21 Hz), 75.2, 37.4, 30.8, 19.6.

3.2.5. General Procedure for Preparing Compounds 7a–i

To a solution of pyrimidine 6 (0.569 mmol) in CH3CN (2.90 mL), (R)-(−)-1-Boc-3-methylpiperazine (1.14 mmol) and N,N-diisopropylethylamine (1.14 mmol) were added. The reaction mixture was allowed to stir at 110 °C for 14 h. After completion of the reaction (monitored by TLC), it was quenched with saturated aqueous NH4Cl, extracted with EtOAc, and washed with brine. The organic layers were dried over anhydrous MgSO4 and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (EtOAc/CHCl3/n-hexane = 1:4:8) to afford methyl piperazine carboxylate 7.

(R)-tert-Butyl-4-(5-fluoro-4-((2-fluorobenzyl)oxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7a): Yield: 14%; 1H-NMR (400 MHz, CDCl3) δ 7.98 (d, J = 2.8 Hz, 1H), 7.49–7.45 (m, 1H), 7.35–7.29 (m, 1H), 7.17–7.13 (m, 1H), 7.11–7.06 (m, 1H), 5.53–5.44 (m, 2H), 4.73 (bs, 1H), 4.32 (d, J = 13.3, 1H), 4.14–3.90 (m, 2H), 3.17–2.89 (m, 3H), 1.49 (s, 9H), 1.12 (d, J = 6.7 Hz, 3H). 13C-NMR (100 MHz, CDCl3) δ 160.7 (d, J = 246 Hz), 157.6 (d, J = 11 Hz), 156.8 (d, J = 2 Hz), 155.2, 143.5 (d, J = 19 Hz), 139.9 (d, J = 247 Hz), 130.8 (d, J = 4 Hz), 130.05 (d, J = 9 Hz), 124.2 (d, J = 3 Hz), 123.4 (d, J = 14 Hz), 115.4 (d, J = 21 Hz), 79.8, 67.6 (d, J = 5 Hz), 48.5, 47.2, 44.0, 42.9, 38.7, 28.4, 14.1.

(R)-tert-Butyl-4-(5-fluoro-4-((3-fluorobenzyl)oxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7b): Yield: 27%; 1H-NMR (400 MHz, CDCl3) δ 7.99 (d, J = 2.8 Hz, 1H), 7.36–7.30 (m, 1H), 7.19 (d, J = 7.7 Hz, 1H), 7.15–7.09 (m, 1H), 7.03–6.98 (m, 1H), 5.43–5.36 (m, 2H), 4.69 (bs, 1H), 4.29 (d, J = 13.0 Hz, 1H), 4.14–3.84 (m, 2H), 3.16–2.87 (m, 3H), 1.48 (s, 9H), 1.11 (d, J = 6.6 Hz, 3H). 13C-NMR (100 MHz, CDCl3) δ 162.9 (d, J = 245 Hz), 157.5 (d, J = 10 Hz), 156.8 (d, J = 2 Hz), 155.2, 143.6 (d, J = 19 Hz), 139.9 (d, J = 240 Hz), 138.7 (d, J = 7 Hz), 130.1 (d, J = 8 Hz), 123.2 (d, J = 3 Hz), 115.1 (d, J = 21 Hz), 114.6 (d, J = 22 Hz), 79.9, 67.1 (d, J = 1 Hz), 48.5, 47.2, 44.0, 42.9, 38.8, 28.4, 14.1.

(R)-tert-Butyl-4-(5-fluoro-4-((4-fluorobenzyl)oxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7c): Yield: 21%; 1H-NMR (400 MHz, CDCl3) δ 7.98 (d, J = 2.8 Hz, 1H), 7.43–7.39 (m, 2H), 7.07–7.03 (m, 2H), 5.40–5.34 (m, 2H), 4.72 (bs, 1H), 4.30 (d, J = 12.8 Hz, 1H), 4.15–3.78 (m, 2H), 3.17–2.87 (m, 3H), 1.48 (s, 9H), 1.13 (d, J = 6.7 Hz, 3H). 13C-NMR (100 MHz, CDCl3) δ 162.7 (d, J = 245 Hz), 157.7 (d, J = 11 Hz), 156.8 (d, J = 2 Hz), 155.2, 143.5 (d, J = 20 Hz), 139.9 (d, J = 247 Hz), 131.9 (d, J = 3 Hz), 129.9 (d, J = 8 Hz), 115.5 (d, J = 21 Hz), 79.9, 67.2, 48.5, 47.2, 44.0, 42.9, 38.8, 28.4, 14.1.

(R)-tert-Butyl-4-(5-fluoro-4-((1-(3-fluorophenyl)ethoxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7d): Yield: 44%; 1H-NMR (400 MHz, CDCl3) δ 7.94–7.93 (m, 1H, 1H′), 7.32–7.25 (m, 1H, 1H′), 7.16–7.06 (m, 2H, 2H′), 6.97–6.90 (m, 1H, 1H′), 6.09–6.00 (m, 1H, 1H′), 4.57 (bs, 1H, 1H′), 4.22–3.85 (m, 3H, 3H′), 3.08–2.74 (m, 3H, 3H′), 1.65 (d, J = 6.6 Hz, 3H, 3H′), 1.46 (d, J = 2.4 Hz, 9H, 9H′), 1.13–0.90 (m, 3H, 3H′). 13C-NMR (100 MHz, CDCl3, Diastereomeric mixture) δ 162.9 (d, J = 244 Hz),
157.14 (d, J = 11 Hz), 157.11 (d, J = 11 Hz), 156.7 (d, J = 3 Hz), 155.20, 155.18, 145.0 (d, J = 23 Hz), 144.91 (d, J = 23 Hz), 143.47 (d, J = 20 Hz), 143.44 (d, J = 20 Hz), 140.0 (d, J = 245 Hz), 130.11 (d, J = 8 Hz), 130.07 (d, J = 9 Hz), 121.42 (d, J = 3 Hz), 121.17 (d, J = 3 Hz), 114.6 (d, J = 21 Hz), 114.5 (d, J = 21 Hz), 112.8 (d, J = 22 Hz), 112.6 (d, J = 22 Hz), 79.8, 79.8, 73.92, 73.90, 73.80, 73.79, 48.4, 47.1, 44.0, 42.8, 38.7, 28.4, 23.0, 22.8, 13.9.

(R)-tert-Butyl-4-(5-fluoro-4-(1-(4-fluorophenyl)ethoxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7g): Yield: 40%; 1H-NMR (400 MHz, CDCl₃, Diastereomeric mixture) δ 7.95–7.94 (m, 1H, 1H'), 7.94–7.33 (m, 2H, 2H'), 7.06–6.98 (m, 2H, 2H'), 6.12–6.05 (m, 1H, 1H'), 4.60 (bs, 1H, 1H'), 4.25–3.88 (m, 3H, 3H'), 3.10–2.87 (m, 3H, 3H'), 1.66 (d, J = 6.6 Hz, 3H, 3H'), 1.48 (d, J = 2.0 Hz, 9H, 9H'), 1.15–0.96 (m, 3H, 3H'). 13C-NMR (100 MHz, CDCl₃, Diastereomeric mixture) δ 162.3 (d, J = 245 Hz), 162.2 (d, J = 245 Hz), 157.3 (d, J = 11 Hz), 157.2 (d, J = 11 Hz), 156.7, 155.2, 143.4 (d, J = 20 Hz), 143.3 (d, J = 19 Hz), 140.0 (d, J = 247 Hz), 138.1 (d, J = 3 Hz), 137.9 (d, J = 3 Hz), 127.7 (d, J = 8 Hz), 127.4 (d, J = 8 Hz), 115.5 (d, J = 22 Hz), 115.4 (d, J = 21 Hz), 79.9, 73.9, 73.8, 48.5, 47.2, 43.8, 42.9, 38.7, 38.6, 28.4, 23.0, 22.8, 14.0.

(R)-tert-Butyl-4-(5-fluoro-4-(3-(3-fluoropropoxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7h): Yield: 53%; 1H-NMR (400 MHz, CDCl₃) δ 7.94 (d, J = 3.0 Hz, 1H, 1H'), 7.26–7.20 (m, 1H), 6.96 (d, J = 7.7 Hz, 1H), 6.90–6.85 (m, 2H), 4.65 (bs, 1H), 4.35 (t, J = 6.5 Hz, 2H), 4.25–4.21 (m, 1H), 4.18–3.88 (m, 2H), 3.13–2.87 (m, 3H), 2.77 (t, J = 7.6 Hz, 2H), 2.13–2.06 (m, 2H), 1.47 (s, 9H), 1.12 (d, J = 6.7 Hz, 3H). 13C-NMR (100 MHz, CDCl₃, Diastereomeric mixture) δ 163.6 (d, J = 244 Hz), 158.7 (d, J = 11 Hz), 157.5 (d, J = 2 Hz), 155.9, 144.4 (d, J = 7 Hz), 143.8 (d, J = 20 Hz), 140.6 (d, J = 247 Hz), 130.5 (d, J = 9 Hz), 124.8 (d, J = 3 Hz), 115.9 (d, J = 21 Hz), 113.6 (d, J = 21 Hz), 80.4, 66.0, 49.1, 47.8, 44.6, 43.5, 39.4, 32.5, 30.6, 29.1, 14.6.

(R)-tert-Butyl-4-(5-fluoro-4-(3-(3-fluoropropoxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7i): Yield: 45%; 1H-NMR (400 MHz, CDCl₃) δ 7.96 (d, J = 2.9 Hz, 1H), 7.16–7.13 (m, 2H), 6.99–6.94 (m, 2H), 4.66 (bs, 1H), 4.39–4.33 (m, 2H), 4.25–3.89 (m, 3H), 3.20–3.06 (m, 2H), 3.02–2.98 (m, 1H), 2.88–2.73 (m, 2H), 2.12–2.05 (m, 2H), 1.48 (s, 9H), 1.12 (d, J = 6.6 Hz, 3H). 13C-NMR (100 MHz, CDCl₃) δ 161.4 (d, J = 242 Hz), 158.1 (d, J = 11 Hz), 156.9 (d, J = 3 Hz), 155.2, 143.1 (d, J = 20 Hz), 140.0 (d, J = 247 Hz), 136.8 (d, J = 8 Hz), 115.2 (d, J = 21 Hz), 79.8, 65.4, 48.5, 47.1, 44.0, 42.9, 38.7, 31.3, 30.4, 28.4, 14.0.

(R)-tert-Butyl-4-(5-fluoro-4-(4-(3-fluorobutyl)oxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7j): Yield: 40%; 1H-NMR (400 MHz, CDCl₃, Diastereomeric mixture) δ 7.96–7.95 (m, 1H, 1H'), 7.12–7.07 (m, 2H, 2H'), 6.96–6.91 (m, 2H, 2H'), 5.26–5.18 (m, 1H, 1H'), 4.64 (bs, 1H, 1H'), 4.22–3.89 (m, 3H, 3H'), 3.14–3.05 (m, 2H, 2H'), 2.89–2.62 (m, 3H, 3H'), 2.13–2.05 (m, 1H, 1H'), 1.94–1.85 (m, 1H, 1H'), 1.48 (d, J = 0.6 Hz, 9H, 9H'), 1.38 (d, J = 6.2 Hz, 3H, 3H'), 1.13–1.11 (m, 3H, 3H'). 13C-NMR (100 MHz, CDCl₃, Diastereomeric mixture) δ 161.3 (d, J = 242 Hz), 157.9 (d, J = 10 Hz), 156.9, 155.2, 143.1 (d, J = 20 Hz), 140.2 (d, J = 246 Hz), 137.07 (d, J = 3 Hz), 137.06 (d, J = 2 Hz), 129.76 (d, J = 7 Hz), 129.72 (d, J = 7 Hz), 115.14 (d, J = 21 Hz), 115.12 (d, J = 21 Hz), 79.8, 72.10, 72.17, 48.5, 47.2, 44.0, 42.9, 38.7, 37.7, 30.99, 30.96, 28.4, 19.9, 19.8, 13.9.
3.2.6. General Procedure for Preparing Compounds 4a–i

Methods A: To a solution of methyl piperazine carboxylate 7 (0.161 mmol) in CH$_2$Cl$_2$ (1.60 mL), trifluoro acetic acid (4.01 mmol) was added at 0 °C. The reaction mixture was allowed to stir at the same temperature for 1 h. After completion of the reaction (monitored by TLC), the mixture was diluted with saturated aqueous NaHCO$_3$, and extracted with EtOAc. The organic layers were dried over anhydrous MgSO$_4$ and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (DCM/MeOH = 10:1) to afford methyl piperazine 4.

Methods B: To a solution of methyl piperazine carboxylate 7 (0.144 mmol) in dioxane (1.45 mL), 1 M HCl in ether (1.44 mmol) was added at 0 °C. The reaction mixture was allowed to stir at the same temperature for 2.5 h. After completion of the reaction (monitored by TLC), the mixture was diluted with saturated aqueous NaHCO$_3$, and extracted with EtOAc. The organic layers were dried over anhydrous MgSO$_4$ and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (DCM/MeOH = 10:1) to afford methyl piperazine 4.

(R)-5-Fluoro-4-(((2-fluorobenzyl)oxy)-2-(2-methylpiperazin-1-yl)pyrimidine (4a): Methods A: Yield: 52%;

H-NMR (400 MHz, MeOD) δ 8.16 (d, J = 3.1 Hz, 1H), 7.67–7.63 (m, 1H), 7.56–7.50 (m, 1H), 7.37–7.27 (m, 2H), 5.71–5.61 (m, 2H), 4.86–4.83 (m, 1H), 4.49–4.45 (m, 1H), 3.24–3.17 (m, 2H), 3.09–3.01 (m, 2H), 2.87–2.80 (m, 1H), 1.35 (d, J = 6.8 Hz, 3H).

C-NMR (100 MHz, MeOD) δ 160.9 (d, J = 245 Hz), 157.6 (d, J = 11 Hz), 156.9 (d, J = 2 Hz), 143.0 (d, J = 12 Hz), 139.6 (d, J = 246 Hz), 130.1 (d, J = 3 Hz), 130.0 (d, J = 8 Hz), 124.1 (d, J = 3 Hz), 123.4 (d, J = 14 Hz), 115.0 (d, J = 21 Hz), 61.5 (d, J = 5 Hz), 49.1, 46.3, 44.9, 38.8, 12.4; HRMS-ESI (m/z): [M + H]$^+$ calcd. for C$_{16}$H$_{19}$F$_2$N$_4$O: 321.1521; found: 321.1525; HPLC purity, 6.7 min, 98.8%.

(R)-5-Fluoro-4-(((2-fluorobenzyl)oxy)-2-(2-methylpiperazin-1-yl)pyrimidine (4b): Methods A: Yield: 72%;

H-NMR (400 MHz, MeOD) δ 8.18 (d, J = 2.9 Hz, 1H), 7.67–7.54 (m, 1H), 7.43 (d, J = 7.6 Hz, 1H), 7.36 (d, J = 9.6 Hz, 1H), 7.31–7.21 (m, 1H), 5.66–5.58 (m, 2H), 4.83 (bs, 1H), 4.47 (d, J = 12.6 Hz, 1H), 3.25–3.18 (m, 2H), 3.10–3.02 (m, 2H), 2.88–2.81 (m, 1H), 1.35 (d, J = 6.8 Hz, 3H).

C-NMR (100 MHz, MeOD) δ 162.9 (d, J = 243 Hz), 157.6 (d, J = 11 Hz), 156.9 (d, J = 2 Hz), 143.1 (d, J = 20 Hz), 139.6 (d, J = 245 Hz), 139.3 (d, J = 8 Hz), 130.0 (d, J = 8 Hz), 123.0 (d, J = 3 Hz), 114.4 (d, J = 21 Hz), 114.0 (d, J = 22 Hz), 66.9 (d, J = 1 Hz), 49.2, 46.4, 44.8, 38.8, 12.4; HRMS-ESI (m/z): [M + H]$^+$ calcd. for C$_{16}$H$_{19}$F$_2$N$_4$O: 321.1521; found: 321.1523; HPLC purity, 6.9 min, 97.1%.

(R)-5-Fluoro-4-(((3-fluorobenzyl)oxy)-2-(2-methylpiperazin-1-yl)pyrimidine (4c): Methods A: Yield: 57%;

H-NMR (400 MHz, MeOD) δ 8.19 (d, J = 3.1 Hz, 1H), 7.68–7.65 (m, 2H), 7.30 (t, J = 8.8 Hz, 2H), 5.65–5.57 (m, 2H), 4.89–4.86 (m, 1H), 4.53–4.48 (m, 1H), 3.28–3.21 (m, 2H), 3.14–3.06 (m, 2H), 2.92–2.84 (m, 1H), 1.38 (d, J = 6.9 Hz, 3H).

C-NMR (100 MHz, MeOD) δ 162.6 (d, J = 243 Hz), 157.7 (d, J = 11 Hz), 156.9 (d, J = 2 Hz), 143.0 (d, J = 20 Hz), 132.5 (d, J = 4 Hz), 129.8 (d, J = 9 Hz), 114.9 (d, J = 22 Hz), 67.0, 49.1, 46.3, 44.8, 38.7, 12.4; HRMS-ESI (m/z): [M + H]$^+$ calcd. for C$_{16}$H$_{19}$F$_2$N$_4$O: 321.1521; found: 321.1524; HPLC purity, 7.4 min, 97.4%.

5-Fluoro-4-(((1-(3-fluorophenyl)ethoxy)-2-((R)-2-methylpiperazin-1-yl)pyrimidine (4d): Methods A: Yield: 71%;

H-NMR (400 MHz, MeOD, Diastereomeric mixture) δ 8.13 (s, 1H, 1H$^1$), 7.52–7.49 (m, 1H, 1H$^3$), 7.39–7.36 (m, 1H, 1H$^1$), 7.32–7.29 (m, 1H, 1H$^3$), 7.16–7.14 (m, 2H), 6.31–6.22 (m, 1H, 1H$^1$), 4.71 (bs, 1H, 1H$^3$), 4.43–4.33 (m, 1H, 1H$^1$), 3.18–2.99 (m, 4H, 4H$^1$), 2.86–2.72 (m, 1H, 1H$^3$), 1.83–1.81 (m, 3H, 3H$^3$), 1.39–1.32 (m, 3H, 3H$^1$).

C-NMR (100 MHz, MeOD, Diastereomeric mixture) δ 162.93 (d, J = 245 Hz), 162.91 (d, J = 245 Hz), 157.13 (d, J = 11 Hz), 157.10 (d, J = 11 Hz), 156.8, 145.6 (d, J = 7 Hz), 145.3 (d, J = 7 Hz), 143.1 (d, J = 20 Hz), 143.0 (d, J = 19 Hz), 139.7 (d, J = 246 Hz), 130.1 (d, J = 8 Hz), 130.0 (d, J = 8 Hz), 121.14 (d, J = 27 Hz), 121.11 (d, J = 27 Hz), 114.0 (d, J = 22 Hz), 113.9 (d, J = 21 Hz), 74.05 (d, J = 2 Hz), 73.97 (d, J = 1 Hz), 49.1, 49.0, 46.3, 44.8, 44.7, 38.72, 38.68, 22.0, 21.8, 12.5, 12.4; HRMS-ESI (m/z): [M + H]$^+$ calcd. for C$_{17}$H$_{23}$F$_2$N$_4$O: 335.1678; found: 335.1680; HPLC purity, 7.5 min, 98.1%.

5-Fluoro-4-(((4-fluorophenyl)ethoxy)-2-((R)-2-methylpiperazin-1-yl)pyrimidine (4e): Methods B: Yield: 53%;

H-NMR (400 MHz, MeOD, Diastereomeric mixture) δ 8.16–8.14 (m, 1H, 1H$^1$), 7.64–7.59 (m, 2H, 2H$^1$),...
Molecules 2017, 22, 1416

7.28–7.23 (m, 2H, 2H′), 6.33–6.28 (m, 1H, 1H′), 4.78–4.76 (m, 1H, 1H′), 4.45–4.43 (m, 1H, 1H′), 3.21–3.10 (m, 2H, 2H′), 3.08–2.99 (m, 2H, 2H′), 2.89–2.75 (m, 1H, 1H′), 1.84–1.82 (m, 3H, 3H′), 1.41–1.17 (m, 3H, 3H′).

(3)-5-Fluoro-4-((3-fluorophenyl)propoxy)-2-(2-methylpiperazin-1-yl)pyrimidine (1f): Methods A: Yield: 62%; 1H-NMR (400 MHz, CDCl3) δ 7.99 (d, J = 3.0 Hz, 1H), 7.30–7.24 (m, 1H), 7.00 (d, J = 7.6 Hz, 1H), 6.95–6.89 (m, 2H), 4.67–4.61 (m, 1H), 4.42–4.36 (m, 2H), 4.29–4.25 (m, 1H), 3.09–2.98 (m, 3H), 2.90 (d, J = 12.2 Hz, 1H), 2.83–2.77 (m, 3H), 2.18–2.11 (m, 2H), 1.24 (d, J = 6.8 Hz, 3H). 13C-NMR (100 MHz, MeOD) δ 163.0 (d, J′ = 21 Hz), 158.3 (d, J′ = 11 Hz), 156.2, 144.1 (d, J′ = 8 Hz), 142.9 (d, J′ = 21 Hz), 140.3 (d, J′ = 247 Hz), 129.7 (d, J′ = 8 Hz), 121.4 (d, J′ = 4 Hz), 114.8 (d, J′ = 21 Hz), 112.3 (d, J′ = 21 Hz), 65.5, 46.6, 43.3, 35.8, 31.3, 29.7, 12.3; HRMS-ESI (m/z): [M + H]+ calc. for C19H25F2N4O: 349.1834; found: 347.1837; HPLC purity, 71.9 min, 98.7%.

(3)-5-Fluoro-4-((3-fluorophenyl)propoxy)-2-(2-methylpiperazin-1-yl)pyrimidine (4f): Methods A: Yield: 63%; 1H-NMR (400 MHz, CDCl3) δ 7.93 (d, J = 3.0 Hz, 1H), 7.14–7.11 (m, 2H), 6.96–6.92 (m, 2H), 4.58 (t, J = 5.0 Hz, 1H), 4.35–4.31 (m, 2H), 4.23–4.19 (m, 1H), 3.03–2.92 (m, 3H), 2.84 (d, J = 12.3 Hz, 1H), 2.75–2.71 (m, 3H), 2.10–2.03 (m, 2H), 1.18 (d, J = 6.8 Hz, 3H). 13C-NMR (100 MHz, MeOD) δ 161.4 (d, J′ = 241 Hz), 158.4 (d, J′ = 11 Hz), 156.0, 142.8 (d, J′ = 20 Hz), 140.5 (d, J′ = 247 Hz), 137.1, 129.8 (d, J′ = 8 Hz), 114.6 (d, J′ = 21 Hz), 65.7, 46.9, 44.3, 43.1, 35.3, 30.7, 30.1, 12.4; HRMS-ESI (m/z): [M + H]+ calc. for C18H23F2N4O: 349.1834; found: 347.1837; HPLC purity, 5.2 min, 95.5%.

5-Fluoro-4-((3-fluorophenyl)butan-2-ylxoxy)-2-((R)-2-methylpiperazin-1-yl)pyrimidine (4h): Methods A: Yield: 68%; 1H-NMR (400 MHz, MeOD, Diastereomeric mixture) δ 8.16–8.15 (m, 1H, 1H′), 7.46–7.40 (m, 1H, 1H′), 7.16–7.15 (m, 1H, 1H′), 7.09–7.05 (m, 2H, 2H′), 5.48–5.38 (m, 1H, 1H′), 4.80–4.72 (m, 1H, 1H′), 4.42–4.37 (m, 1H, 1H′), 3.24–3.15 (m, 2H, 2H′), 3.12–3.03 (m, 1H, 1H′), 2.99–2.83 (m, 3H, 3H′), 2.31–2.24 (m, 1H, 1H′), 2.18–2.09 (m, 1H, 1H′), 1.57–1.56 (m, 3H, 3H′), 1.39–1.35 (m, 3H, 3H′). 13C-NMR (100 MHz, MeOD, Diastereomeric mixture) δ 162.9 (d, J′ = 242 Hz), 157.86 (d, J′ = 11 Hz), 157.84 (d, J′ = 20 Hz), 156.9, 144.3 (d, J′ = 7 Hz), 142.7 (d, J′ = 245 Hz), 129.68 (d, J′ = 9 Hz), 129.66 (d, J′ = 9 Hz), 124.02 (d, J′ = 3 Hz), 123.98 (d, J′ = 3 Hz), 114.8 (d, J′ = 21 Hz), 114.7 (d, J′ = 21 Hz), 112.3 (d, J′ = 22 Hz), 112.2 (d, J′ = 22 Hz), 72.01, 71.92, 49.12, 49.10, 46.24, 46.22, 44.82, 44.80, 38.70, 38.68, 36.9, 31.0, 18.66, 18.64, 12.4, 12.3; HRMS-ESI (m/z): [M + H]+ calc. for C19H25F2N4O: 363.1991; found: 363.1992; HPLC purity, 8.0 min, 98.4%.

5-Fluoro-4-((3-fluorophenyl)butan-2-ylxoxy)-2-((R)-2-methylpiperazin-1-yl)pyrimidine (4i): Methods A: Yield: 53%; 1H-NMR (400 MHz, MeOD, Diastereomeric mixture) δ 8.12 (d, J = 2.4 Hz, 1H, 1H′), 7.32–7.29 (m, 2H, 2H′), 7.14–7.08 (m, 2H, 2H′), 5.43–5.34 (m, 1H, 1H′), 4.74–4.67 (m, 1H, 1H′), 4.35–4.31 (m, 1H, 1H′), 3.18–3.11 (m, 2H, 2H′), 3.08–2.99 (m, 2H, 2H′), 2.93–2.79 (m, 1H, 1H′), 2.28–2.19 (m, 1H, 1H′), 2.13–2.03 (m, 1H, 1H′), 1.54–1.51 (m, 3H, 3H′), 1.35–1.31 (m, 3H, 3H′). 13C-NMR (100 MHz, MeOD, Diastereomeric mixture) δ 161.3 (d, J′ = 241 Hz), 157.87 (d, J′ = 11 Hz), 157.85 (d, J′ = 11 Hz), 156.9, 142.7 (d, J′ = 20 Hz), 139.9 (d, J′ = 246 Hz), 137.3 (d, J′ = 4 Hz), 129.72 (d, J′ = 7 Hz), 129.67 (d, J′ = 7 Hz), 114.57 (d, J′ = 21 Hz), 114.55 (d, J′ = 21 Hz), 72.00, 71.95, 49.13, 49.09, 46.24, 46.22, 44.83, 44.79, 38.71, 38.68, 37.3, 30.5, 30.4, 18.66, 18.62, 12.4, 12.3; HRMS-ESI (m/z): [M + H]+ calc. for C19H25F2N4O: 363.1991; found: 363.1993; HPLC purity, 8.2 min, 96.9%.
3.3. Synthesis of Optically Active Pyrimidines 4d and 4e

3.3.1. General Procedure for Preparing Compound (R)-(−)-5d and (R)-(−)-5e

To a solution of 1-(3 or 4-fluorophenyl)ethanol (6.67 mmol) in n-hexane (22.2 mL), CAL-B (147 mg), vinyl acetate (3.34 mmol), and triethylamine (0.667 mmol) were added. The reaction mixture was allowed to stir at room temperature for 1 h. After completion of the reaction (monitored by TLC), the mixture was filtered and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (EtOAc:n-hexane = 1:8) to afford acetate intermediate (315 mg) as a colorless oil. To a solution of acetate (1.73 mmol) in MeOH (3.45 mL), 1 M NaOH (2.59 mmol) was added. The reaction mixture was allowed to stir at room temperature for 1 h. After completion of the reaction (monitored by TLC), it was quenched with distilled water and extracted with EtOAc. The organic layers were dried over anhydrous MgSO₄ and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (EtOAc:n-hexane = 1:8) to afford alcohol (R)-(−)-5d and (R)-(−)-5e.

(R)-1-(3-Fluorophenyl)ethan-1-ol (R)-(−)-5d: Yield: 17%; ¹H-NMR (400 MHz, CDCl₃) δ 7.32–7.26 (m, 1H), 7.12–7.07 (m, 2H), 6.97–6.92 (m, 1H), 4.87 (q, J = 6.4 Hz, 1H), 2.18 (s, 1H), 1.47 (d, J = 6.4 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ 163.0 (d, 3J = 244 Hz), 148.5 (d, 3J = 6 Hz), 130.0 (d, 3J = 8 Hz), 121.0 (d, 3J = 3 Hz), 114.2 (d, 2J = 21 Hz), 112.3 (d, 2J = 21 Hz), 69.8, 25.2; Optical rotation for (R)-(−)-5d: [α]₂⁰ +43.7° (c 0.7, CHCl₃).

(R)-1-(4-Fluorophenyl)ethan-1-ol (R)-(−)-5e: Yield: 32%; ¹H-NMR (400 MHz, CDCl₃) δ 7.32–7.26 (m, 2H), 7.03–6.97 (m, 2H), 4.84 (q, J = 6.1 Hz, 1H), 2.34 (s, 1H), 1.44 (d, J = 6.4 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ 162.1 (d, 3J = 243 Hz), 141.5 (d, 3J = 22 Hz), 112.1 (d, 3J = 8 Hz), 115.2 (d, 2J = 21 Hz), 69.7, 25.3; Optical rotation for (R)-(−)-5e: [α]₂⁰ +51.9° (c 0.5, CHCl₃).

3.3.2. General Procedure for Preparing Compound (S)-(−)-5d and (S)-(−)-5e

To a solution of 1-(3 or 4-fluorophenyl)ethanol (6.67 mmol) in n-hexane (22.2 mL), CAL-B (147 mg), vinyl acetate (13.3 mmol), and triethylamine (0.667 mmol) were added. The reaction mixture was allowed to stir at room temperature for 12 h. After completion of the reaction (monitored by TLC), the mixture was filtered and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (EtOAc:n-hexane = 1:8) to afford alcohol (S)-(−)-5d and (S)-(−)-5e.

(S)-1-(3-Fluorophenyl)ethan-1-ol (S)-(−)-5d: Yield: 44%; ¹H-NMR (400 MHz, CDCl₃) δ 7.31–7.26 (m, 1H), 7.11–7.06 (m, 2H), 6.96–6.91 (m, 1H), 4.85 (td, J = 5.5, 7.5 Hz, 1H), 2.31 (s, 1H), 1.46 (d, J = 6.5 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ 163.0 (d, 3J = 245 Hz), 148.6 (d, 3J = 6 Hz), 130.0 (d, 3J = 8 Hz), 121.0 (d, 3J = 3 Hz), 114.2 (d, 2J = 21 Hz), 112.3 (d, 2J = 22 Hz), 69.8 (d, 4J = 2 Hz), 25.2; Optical rotation for (S)-(−)-5d: [α]₂⁰ −46.9° (c 0.4, CHCl₃).

(S)-1-(4-Fluorophenyl)ethan-1-ol (S)-(−)-5e: Yield: 46%; ¹H-NMR (400 MHz, CDCl₃) δ 7.33–7.30 (m, 2H), 7.04–6.98 (m, 2H), 4.85 (q, J = 6.4 Hz, 1H), 2.16 (s, 1H), 1.45 (d, J = 6.4 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃) δ 162.1 (d, 3J = 243 Hz), 141.5 (d, 4J = 3 Hz), 127.1 (d, 3J = 8 Hz), 115.2 (d, 2J = 21 Hz), 69.8, 25.3; Optical rotation for (S)-(−)-5e: [α]₂⁰ −49.7° (c 0.6, CHCl₃).

3.3.3. General Procedure for Preparing Compounds (R)/(S)-10 and 11

To a solution of (R) or (S)-secondary alcohol (0.0749 mmol) in CH₂Cl₂ (0.400 mL), (R)-2-acetoxy-2-phenylacetic acid (0.112 mmol), EDCI (0.112 mmol), and DMAP (0.112 mmol) were added. The reaction mixture was allowed to stir at room temperature for 12 h. After completion of the reaction (monitored by TLC), the mixture was filtered, extracted with CH₂Cl₂, and washed with brine. The organic layers were dried over anhydrous MgSO₄ and concentrated in vacuo. The resulting residue was purified by flash column chromatography on silica gel (EtOAc:n-hexane = 1:8) to afford mandelate (R)/(S)-10 and 11.
(R)-1-(3-Fluorophenyl)ethyl (R)-2-acetoxy-2-phenylacetate (6d): Yield: 72%; 1H-NMR (400 MHz, CDCl3) δ 8.18 (d, J = 2.2 Hz, 1H), 7.37–7.31 (m, 1H), 7.23 (d, J = 7.7 Hz, 1H), 7.18–7.15 (m, 1H), 7.03–6.99 (m, 1H), 6.30 (q, J = 6.5 Hz, 1H), 1.71 (d, J = 6.6 Hz, 3H). 13C-NMR (100 MHz, CDCl3) δ 156.8 (d2, J = 11 Hz), 153.2 (d4, J = 5 Hz), 146.0 (d1, J = 263 Hz), 144.4 (d2, J = 20 Hz), 142.9 (d3, J = 7 Hz), 130.3 (d2, J = 8 Hz), 122.0 (d, J = 3 Hz), 115.3 (d2, J = 21 Hz), 113.3 (d2, J = 22 Hz), 75.6 (d, J = 2 Hz), 22.2; Optical rotation for (R)-6d: [α]27D +178.3° (c 0.7, CHCl3).

(S)-1-(3-Fluorophenyl)ethyl (R)-2-acetoxy-2-phenylacetate (6e): Yield: 65%; 1H-NMR (400 MHz, CDCl3) δ 8.16 (d, J = 2.2 Hz, 1H), 7.47–7.42 (m, 2H), 7.08–7.02 (m, 2H), 6.30 (q, J = 6.6 Hz, 1H), 1.71 (d, J = 6.6 Hz, 3H). 13C-NMR (100 MHz, CDCl3) δ 156.8 (d2, J = 11 Hz), 153.2 (d4, J = 4 Hz), 146.0 (d1, J = 262 Hz), 144.4 (d2, J = 20 Hz), 142.9 (d3, J = 7 Hz), 130.3 (d2, J = 8 Hz), 122.0 (d, J = 3 Hz), 115.3 (d2, J = 21 Hz), 113.3 (d2, J = 22 Hz), 75.7 (d, J = 1 Hz), 22.2; Optical rotation for (S)-6e: [α]27D +197.3° (c 0.8, CHCl3).

tert-Butyl-(R)-4-(5-fluoro-4-((R)-1-(3-fluorophenyl)ethoxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate (7d): Yield: 50%; 1H-NMR (400 MHz, CDCl3) δ 7.94 (d, J = 2.8 Hz, 1H), 7.31–7.26 (m, 1H), 7.14 (d, J = 7.7 Hz, 1H), 7.07 (d, J = 9.6 Hz, 1H), 6.96–6.91 (m, 1H), 6.03 (q, J = 6.6 Hz, 1H), 4.58 (bs, 1H), 4.18–3.86 (m, 3H), 3.08–3.01 (m, 2H), 2.84–2.74 (m, 1H), 1.66 (d, J = 6.6 Hz, 3H), 1.47 (s, 9H), 0.91 (d, J = 6.5 Hz, 3H). 13C-NMR (100 MHz, CDCl3) δ 162.9 (d, J = 245 Hz), 157.1 (d2, J = 11 Hz), 156.7 (d, J = 2 Hz), 155.2, 145.1 (d, J = 7 Hz), 143.4 (d, J = 20 Hz), 140.0 (d, J = 246 Hz), 130.1 (d, J = 8 Hz), 121.2 (d, J = 3 Hz), 114.5 (d, J = 21 Hz), 112.7 (d2, J = 22 Hz), 79.8, 73.9, 48.4, 47.1, 43.9, 42.8, 38.7, 28.4, 23.0, 13.9; Optical rotation for (R)-7d: [α]27D +105.0° (c 0.4, CHCl3).
tert-Butyl-(R)-4-(5-fluoro-4-((S)-1-(3-fluorophenyl)ethoxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate ((S,R)-7d): Yield: 57%; 1H-NMR (400 MHz, CDCl₃) δ 7.95 (d, J = 2.9 Hz, 1H), 7.32–7.26 (m, 1H), 7.16 (d, J = 7.7 Hz, 1H), 7.11–7.09 (m, 1H), 6.97–6.93 (m, 1H), 6.06 (q, J = 6.6 Hz, 1H), 4.55 (bs, 1H), 4.22–3.84 (m, 3H), 3.07–2.87 (m, 3H), 1.66 (d, J = 6.6 Hz, 3H), 1.47 (s, 9H), 1.13 (d, J = 6.6 Hz, 3H). 13C-NMR (100 MHz, CDCl₃) δ 163.0 (d, J₁ = 245 Hz), 157.1 (d, J₂ = 11 Hz), 156.7 (d, J₄ = 2 Hz), 155.2, 144.9 (d, J₃ = 7 Hz), 143.5 (d, J₂ = 19 Hz), 140.0 (d, J₁ = 247 Hz), 130.0 (d, J₃ = 8 Hz), 121.4 (d, J₁ = 3 Hz), 114.6 (d, J₂ = 21 Hz), 112.8 (d, J₂ = 22 Hz), 79.8, 73.8, 48.4, 47.1, 43.9, 42.9, 38.7, 28.4, 22.8, 14.0; Optical rotation for (S,R)-7d: [α]²⁸D +215.3° (c 0.7, CHCl₃).

tert-Butyl-(R)-5-(4-fluorophenyl)ethoxy)pyrimidin-2-yl)-3-methylpiperazine-1-carboxylate ((R,R)-7e): Yield: 44%; 1H-NMR (400 MHz, CDCl₃) δ 7.93 (d, J = 2.8 Hz, 1H), 7.39–7.33 (m, 2H), 7.04–6.98 (m, 2H), 6.07 (d, J = 6.6 Hz, 1H), 4.60 (bs, 1H), 4.23–3.86 (m, 3H), 3.09–2.77 (m, 3H), 1.65 (d, J = 6.6 Hz, 3H), 1.46 (s, 9H), 0.95 (d, J = 6.6 Hz, 3H). 13C-NMR (100 MHz, CDCl₃) δ 162.6 (d, J₁ = 245 Hz), 157.2 (d, J₂ = 11 Hz), 156.7 (d, J₄ = 2 Hz), 155.2, 143.3 (d, J₂ = 19 Hz), 140.0 (d, J₁ = 247 Hz), 138.1 (d, J₃ = 3 Hz), 127.4 (d, J₃ = 8 Hz), 115.4 (d, J₂ = 21 Hz), 79.8, 73.9, 73.9, 48.4, 47.1, 43.7, 42.8, 38.7, 28.4, 23.0, 14.0; Optical rotation for (R,R)-7e: [α]²⁸D +77.9° (c 0.4, CHCl₃).

5-Fluoro-4-((1-(3-fluorophenyl)ethoxy)-2-((R)-2-methylpiperazin-1-yl)pyrimidine ((R,R)-4d): Methods A: Yield: 67%; 1H-NMR (400 MHz, CDCl₃) δ 7.99 (d, J = 2.4 Hz, 1H), 7.32–7.26 (m, 1H), 7.13 (d, J = 7.6 Hz, 1H), 7.06 (d, J = 9.5 Hz, 1H), 6.97–6.93 (m, 1H), 5.99 (q, J = 6.5 Hz, 1H), 4.88 (bs, 1H), 4.47 (d, J = 13.7 Hz, 1H), 3.41–3.14 (m, 4H), 2.84 (bs, 1H), 1.68 (d, J = 6.6 Hz, 3H), 1.12 (d, J = 6.9 Hz, 3H). 13C-NMR (100 MHz, CDCl₃) δ 163.0 (d, J₁ = 245 Hz), 157.6 (d, J₂ = 11 Hz), 145.2 (d, J₄ = 2 Hz), 144.8 (d, J₃ = 7 Hz), 143.1 (d, J₂ = 20 Hz), 140.6 (d, J₁ = 246 Hz), 130.3 (d, J₃ = 8 Hz), 121.0 (d, J₁ = 3 Hz), 114.7 (d, J₂ = 21 Hz), 112.5 (d, J₂ = 23 Hz), 74.7, 47.2, 44.4, 43.1, 35.5, 23.0, 13.2; HRMS-ESI (m/z): [M + H]⁺ calcd for C₁₇H₂₁F₂N₄O: 335.1678; found: 335.1681; HPLC purity, 9.4 min, 98.4%; Optical rotation for (R,R)-4d: [α]²⁸D +118.2° (c 0.3, MeOH).

5-Fluoro-4-((R)-1-(3-fluorophenyl)ethoxy)-2-((R)-2-methylpiperazin-1-yl)pyrimidine ((S,R)-4d): Methods A: Yield: 71%; 1H-NMR (400 MHz, CDCl₃) δ 7.99 (d, J = 2.4 Hz, 1H), 7.34–7.26 (m, 1H), 7.15 (d, J = 7.7 Hz, 1H), 7.09 (d, J = 9.6 Hz, 1H), 7.00–6.95 (m, 1H), 6.03 (q, J = 6.5 Hz, 1H), 4.88–4.85 (m, 1H), 4.53 (d, J = 13.6 Hz, 1H), 3.43 (d, J = 12.1 Hz, 1H), 3.34–3.27 (m, 4H), 1.68 (d, J = 6.6 Hz, 3H), 1.36 (d, J = 7.1 Hz, 3H). 13C-NMR (100 MHz, CDCl₃) δ 163.0 (d, J₁ = 245 Hz), 157.7 (d, J₂ = 11 Hz), 155.5 (d, J₄ = 3 Hz), 144.5 (d, J₃ = 7 Hz), 143.1 (d, J₂ = 21 Hz), 140.5 (d, J₁ = 249 Hz), 130.2 (d, J₃ = 8 Hz), 121.2 (d, J₁ = 3 Hz), 114.8 (d, J₂ = 21 Hz), 112.7 (d, J₂ = 22 Hz), 74.6, 47.0, 44.4, 43.2, 35.7, 22.3, 13.5; HRMS-ESI (m/z): [M + H⁺] calcd for C₁₇H₂₁F₂N₄O: 335.1678; found: 335.1682; HPLC purity, 7.3 min, 97.4%; Optical rotation for (S,R)-4d: [α]²⁸D –228.6° (c 0.4, MeOH).

5-Fluoro-4-((R)-1-(4-fluorophenyl)ethoxy)-2-((R)-2-methylpiperazin-1-yl)pyrimidine ((R,R)-4e): Methods B: Yield: 82%; 1H-NMR (400 MHz, CDCl₃) δ 7.94 (d, J = 2.8 Hz, 1H), 7.40–7.34 (m, 2H), 7.04–6.99 (m, 2H), 6.08 (q, J = 6.6 Hz, 1H), 4.60–4.55 (m, 1H), 4.18 (d, J = 3.5 Hz, 1H), 3.06–2.86 (m, 4H), 2.72–2.65 (m, 1H), 2.54 (bs, 1H), 1.65 (d, J = 6.6 Hz, 1H), 1.04 (d, J = 6.8 Hz, 1H). 13C-NMR (100 MHz, CDCl₃) δ 162.2 (d, J₁ = 244 Hz), 157.2 (d, J₂ = 11 Hz), 156.9 (d, J₄ = 2 Hz), 143.3 (d, J₂ = 19 Hz), 139.9 (d, J₁ = 246 Hz), 138.2 (d, J₄ = 3 Hz), 127.5 (d, J₃ = 8 Hz), 115.4 (d, J₂ = 21 Hz), 73.8, 50.3, 16.6, 45.8, 39.5, 23.0, 13.6; HRMS-ESI (m/z): [M + H⁺] calcd for C₁₇H₂₁F₂N₄O: 335.1678; found: 335.1680; HPLC purity, 7.5 min, 95.7%; Optical rotation for (R,R)-4e: [α]²⁸D +132.0° (c 0.3, MeOH).
5-Fluoro-4-((S)-1-(4-fluorophenyl)ethoxy)-2-((R)-2-methylpiperazin-1-yl)pyrimidine (S,R)-4e: Methods B: Yield: 81%; 1H-NMR (400 MHz, CDCl₃) δ 7.95 (d, J = 2.9 Hz, 1H), 7.40–7.37 (m, 2H), 7.05–7.00 (m, 2H), 6.11 (q, J = 6.6 Hz, 1H), 4.61–4.57 (m, 1H), 4.27–4.23 (m, 1H), 3.09–3.86 (m, 4H), 2.78–2.71 (m, 1H), 2.59 (bs, 1H), 1.66 (d, J = 6.6 Hz, 1H), 1.23 (d, J = 6.8 Hz, 1H). 13C-NMR (100 MHz, CDCl₃) δ 162.3 (d, 1J = 244 Hz), 157.2 (d, 2J = 11 Hz), 156.9 (d, 3J = 2 Hz), 143.4 (d, 4J = 20 Hz), 140.0 (d, 5J = 246 Hz), 137.9 (d, 6J = 3 Hz), 127.7 (d, 7J = 8 Hz), 115.4 (d, 8J = 21 Hz), 73.7, 50.2, 46.5, 45.8, 39.5, 22.7, 13.6; HRMS-ESI (m/z): [M + H]+ calcd for C₁₇H₁₇F₂N₄O: 335.1678; found: 335.1678; HPLC purity, 7.4 min, 97.7%; Optical rotation for (S,R)-4e: [α]D₂⁰ = –226.5° (c 0.5, MeOH).

3.4. Serotonin Receptor Binding Affinity Assays

Eleven dilutions (5 × assay concentration) of the test and reference compounds (Table S1) were prepared in standard binding buffer (50 mM tris(hydroxymethyl)-aminomethane-HCl (Tris-HCl), 10 mM MgCl₂, 1 mM ethylenediaminetetraacetate (EDTA), pH 7.4) by serial dilution: 0.05 nM, 0.5 nM, 1.5 nM, 5 nM, 15 nM, 50 nM, 150 nM, 500 nM, 1.5 µM, 5 µM, and 50 µM. The radioligand (Table S3) was diluted to five times the assay concentration in standard binding buffer. Aliquots (50 µL) of the radioligand were dispensed into the wells of a 96-well plate containing 100 µL of standard binding buffer. Triplicate aliquots (50 µL) of the test and reference compound dilutions were then added. Finally, crude membrane fractions (50 µL) of cells (HEK293 or CHO) expressing human recombinant receptors were dispensed into each well. A total of 250 µL of the reaction mixtures was incubated at room temperature and shielded from light for 1.5 h, and was then harvested by rapid filtration onto Whatman GF/B glass fiber filters presoaked with 0.3% polyethyleneimine, by using a 96-well Brandel harvester (Gaithersburg, MD, USA).

Four rapid washes were performed with chilled standard binding buffer (500 µL) to decrease nonspecific binding. Filters were placed in 6 mL scintillation tubes and allowed to dry overnight. The next day, 4-mL of EcoScint scintillation cocktail (National Diagnostics) was added to each tube. The tubes were capped, labeled, and counted by liquid scintillation counting. The filter mats were dried, and the scintillant was melted onto the filters, then the radioactivity retained on the filters was counted in a Microbeta scintillation counter. The IC₅₀ values were obtained by using the Prism 4.0 program (GraphPad Software, La Jolla, CA, USA) and were converted into Ki values. Each compound was tested at least in triplicate.

4. Conclusions

In summary, we have synthesized a series of pyrimidine derivatives 4a–i and evaluated their binding affinities towards 5-HT₂C receptors. Our initial biological study indicated that 2-amino-4-alkoxy pyrimidines 4b, 4d, and 4e, possessing a short carbon chain between the phenyl group and pyrimidine, have excellent 5-HT₂C binding affinities, which are comparable to that of the reported pyrimidine analogue 3. In order to improve the selectivity for other 5-HT₂ receptor subtypes, the most potent compounds 4d and 4e were selected and their diastereomeric isomers were synthesized as optically pure forms. For this purpose, optically active secondary alcohols 5d and 5e were also prepared by an enzymatic kinetic resolution. (R,R)-4d and 4e displayed excellent 5-HT₂C binding affinities with less selectivity towards 5-HT₂A and 5-HT₂B, whereas (S,R)-4d and 4e exhibited low potencies for 5-HT₂A and 5-HT₂B with a slight loss of the 5-HT₂C binding affinity. These results suggest that the pyrimidine analogue (R,R)-4e is a potential lead compound for identifying a 5-HT₂C selective modulator.

Supplementary Materials: The supplementary materials are available online.

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References

1. Cheng, J.; Giguere, P.M.; Lv, W.; Roth, B.L.; Kozikowski, A.P. Design and synthesis of (2-(5-chloro-2,2-dimethyl-2,3-dihydrobenzofuran-7-yl)cyclopropyl)methanamine as a selective serotonin 2C agonist. Tetrahedron Lett. 2015, 56, 3420–3422. [CrossRef] [PubMed]

2. Berg, K.A.; Clarke, W.P.; Cunningham, K.A.; Spampinato, U. Fine-tuning serotonin$\text{ }_{2C}$ receptor function in the brain: Molecular and functional implications. Neuropharmacology 2008, 55, 969–976. [CrossRef] [PubMed]

3. Sargent, B.J.; Henderson, A.J. Targeting 5-HT receptors for the treatment of obesity. Curr. Opin. Pharmacol. 2011, 11, 52–58. [CrossRef] [PubMed]

4. Ahmad, S.; Ngui, K.; Miller, K.J.; Wu, G.; Hung, C.-P.; Malmstrom, S.; Zhang, G.; O’Tanyi, E.; Keim, W.J.; Cullen, M.J.; et al. Tricyclic dihydroquinazolinones as novel 5-HT$\text{ }_{2C}$ selective and orally efficacious anti-obesity agents. Bioorg. Med. Chem. Lett. 2010, 20, 1128–1133. [CrossRef] [PubMed]

5. Berger, M.; Gray, J.A.; Roth, B.L. The expanded biology of serotonin. Annu. Rev. Med. 2009, 60, 355–366. [CrossRef] [PubMed]

6. Liu, K.K.-C.; Cornelius, P.; Patterson, T.A.; Zeng, Y.; Santucci, S.; Tomlinson, E.; Gibbons, C.; Maurer, T.S.; Marala, R.; Brown, J.; et al. Design and synthesis of orally-active and selective azaindane 5HT$\text{ }_{2C}$ agonist for the treatment of obesity. Bioorg. Med. Chem. Lett. 2010, 20, 266–271. [CrossRef] [PubMed]

7. Andrews, M.D.; Fish, P.V.; Blagg, J.; Brabham, T.K.; Brennan, P.E.; Bridgeland, A.; Brown, A.D.; Bungay, P.J.; Conlon, K.M.; Edmunds, N.J.; et al. Pyrimido[4,5-j]azepines as potent and selective 5-HT$\text{ }_{2C}$ receptor agonists: Design, synthesis, and evaluation of PF-3246799 as a treatment for urinary incontinence. Bioorg. Med. Chem. Lett. 2011, 21, 2715–2720. [CrossRef] [PubMed]

8. Kozikowski, A.P.; Cho, S.J.; Jensen, N.H.; Allen, J.A.; Svennebring, A.M.; Roth, B.L. HTS and Rational Drug Design [RDD] to generate a class of 5-HT$\text{ }_{2C}$-selective ligands for possible use in schizophrenia. Chem. Med. Chem. 2010, 5, 1221–1225. [CrossRef] [PubMed]

9. Meltzer, H.Y.; Roth, B.L. Lorcaserin and pimavanserin: Emerging selectivity of serotonin receptor subtype-targeted drugs. J. Clin. Investig. 2013, 123, 4986–4991. [CrossRef] [PubMed]

10. Holmes, A. Genetic variation in cortico-amygdala serotonin function and risk for stress-related disease. Neurosci. Biobehav. Rev. 2008, 32, 1293–1314. [CrossRef] [PubMed]

11. Heifetz, A.; Storer, R.I.; McMurray, G.; James, T.; Morao, I.; Aldeghi, M.; Bodkin, M.J.; Biggin, P.C. Application of an integrated GPCR SAR-modeling platform to explain the activation selectivity of human 5-HT$\text{ }_{2C}$ over 5-HT$\text{ }_{2B}$. ACS Chem. Biol. 2016, 11, 1372–1382. [CrossRef] [PubMed]

12. Kaumann, A.J.; Levy, F.O. 5-Hydroxytryptamine receptors in the human cardiovascular system. Pharmacol. Ther. 2006, 111, 674–677. [CrossRef] [PubMed]

13. Nichols, D.E. Hallucinogens. Pharmacol. Ther. 2004, 101, 131–181. [CrossRef] [PubMed]

14. Green, M.P.; McMurray, G.; Storer, R.I. Selective 5-HT$\text{ }_{2C}$ receptor agonists: Design and synthesis of pyridazine-fused azepines. Bioorg. Med. Chem. Lett. 2016, 26, 4117–4121. [CrossRef] [PubMed]

15. Cheng, J.; Giguere, P.M.; Schmerberg, C.M.; Pogorelov, V.M.; Rodriguiz, R.M.; Huang, X.-P.; Zhu, H.; McCorvy, J.D.; Wetsel, W.C.; Roth, B.L.; et al. Further advances in optimizing (2-Phenylcyclopropyl) methanamines as Novel Serotonin 2C Agonist: Effects on hyperlocomotion, prepulse inhibition, and cognition models. J. Med. Chem. 2016, 59, 578–591. [CrossRef] [PubMed]

16. Siuciak, J.A.; Chapin, D.S.; McCarthy, S.A.; Guanowsky, V.; Brown, J.; Chiang, P.; Marala, R.; Patterson, T.; Seymour, P.A.; Swick, A.; et al. CP-809,101, a selective 5-HT$\text{ }_{2C}$ agonist, shows activity in animal models of antipsychotic activity. Neuropharmacology 2007, 52, 279–290. [CrossRef] [PubMed]

17. Andrews, M.D.; Green, M.P.; Allerton, C.M.N.; Batchelor, D.V.; Blagg, J.; Brown, A.D.; Gordon, D.W.; McMurray, G.; Milns, D.J.; Nichols, C.L.; et al. Design and synthesis of piperazinyl pyrimidinones as novel selective 5-HT$\text{ }_{2C}$ agonists. Bioorg. Med. Chem. Lett. 2009, 19, 5346–5350. [CrossRef] [PubMed]
18. Neelamegam, R.; Hellenbrand, T.; Schroeder, F.A.; Wang, C.; Hooker, J.M. Imaging evaluation of 5HT₂C agonists, [¹¹C]WAY-163909 and [¹¹C]Vabicaserin, formed by pictet-spengler cyclization. *J. Med. Chem.* 2014, *57*, 1488–1494. [CrossRef] [PubMed]

19. Thomsen, W.J.; Grottick, A.J.; Menzaghi, F.; Reyes-Saldana, H.; Espitia, S.; Yuskin, D.; Whelan, K.; Martin, M.; Morgan, M.; Chen, W.; et al. Lorcaserin, a novel selective human 5-hydroxytryptamine 2C agonist: In vitro and in vivo pharmacological characterization. *J. Pharmacol. Exp. Ther.* 2008, *325*, 577–587. [CrossRef] [PubMed]

20. Shen, J.H.; Zhao, Y.; Rosenzweig-Lipson, S.; Popp, D.; Williams, J.B.; Giller, E.; Detke, M.J.; Kane, J.M. A 6-week randomized, double-blind, placebo-controlled, comparator referenced trial of vabicaserin in acute schizophrenia. *J. Psychiatr. Res.* 2014, *53*, 14–22. [CrossRef] [PubMed]

21. Dunlop, J.; Watts, S.W.; Barrett, J.E.; Coupet, J.; Harrison, B.; Mazandarani, H.; Nawoschik, S.; Pangalos, M.N.; Ramamoorthy, S.; Schechter, L.; et al. Characterization of vabicaserin (SCA-136), a selective 5-hydroxytryptamine 2C receptor agonist. *J. Pharmacol. Exp. Ther.* 2011, *337*, 673–680. [CrossRef] [PubMed]

22. Kalgutkar, A.S.; Bauman, J.N.; McClure, K.F.; Aubrecht, J.; Cortina, S.R.; Paralkar, J. Biochemical basis for differences in metabolism-dependent genotoxicity by two diazinylpiperazine-based 5-HT₂C receptor agonists. *Bioorg. Med. Chem. Lett.* 2009, *19*, 1559–1563. [CrossRef] [PubMed]

23. Kim, J.; Moon, B.S.; Lee, B.C.; Lee, H.Y.; Kim, H.-J.; Choo, H.; Pae, A.N.; Cho, Y.S.; Min, S.-J. A potential PET radiotracer for the 5-HT₂C receptor: Synthesis and in vivo evaluation of 4-(3-[¹⁸F]fluorophenethoxy) pyrimidine. *ACS Chem. Neurosci.* 2017, *8*, 996–1003. [CrossRef] [PubMed]

24. Kim, J.; Cho, Y.S.; Min, S.-J. Facile synthesis of 2-amino-4-alkoxypyrimidines via consecutive nucleophilic aromatic substitution (SₖAr) reactions. *Bull. Korean Chem. Soc.* 2016, *37*, 1998–2008. [CrossRef]

25. Faraldos, J.A.; Giner, J.; Smith, D.H.; Wilson, M.; Ronhovde, L.; Wilson, E.; Clevette, D.; Holmes, A.E.; Rouhier, K. Enzymatic resolution of 1-phenylethanol and formation of a diastereomer: An undergraduate ¹H-NMR experiment to introduce chiral chemistry. *J. Chem. Educ.* 2011, *88*, 334–336. [CrossRef] [PubMed]

26. Wu, X.-M.; Xina, J.-Y.; Sun, W.; Xia, C.-G. Environmentally friendly, efficient resolution of racemic secondary alcohols by lipase-catalyzed enantioselective transesterification in ionic liquids in the presence of organic bases. *Chem. Biodivers.* 2007, *4*, 183–188. [CrossRef] [PubMed]

**Sample Availability:** Samples of the compounds 4a–4i are available from the authors.

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