INTRODUCTION

The oral route is the most convenient and extensively used route for drug administration in the body. It is likely that at least 90% of all the drugs administered by the oral route [1]. The oral route is the most preferred route, and it received more interest in the pharmacy sector because it provides more flexibility in designing of the dosage form as compared to another path. There are different drug deliveries to administer the drug by the oral route. The two difficulties of drug delivery systems (DDS) are, less gastric retention time and less gastric emptying (GE) time. Decrease response of dose due to incomplete drug release from the dosage form in the absorption zone [2, 3]. Due to physiological property the drug releases in unsatisfactory and highly fluctuating among and between individuals and generally affected by the gastrointestinal transit of the form, specifically its residence time in gastric, which seems to be one of the major causes of the overall transit time variability [4]. In the delivery of drugs with narrow absorption windows in the small intestinal region, the gastric retention will provide advantages [4]. Various approaches have been hypothesized to control the residence of DDS in the upper part of the gastrointestinal tract (GIT) such as the incorporation of passage delaying food agents, ion exchange resins, raft system, high-density DDS, floating drug delivery systems (FDDS), swelling or expandable DDS and mucoadhesive DDS [5, 6, 22]. The oral controlled DDS main objective is to attain more expected and enhanced bioavailability [7]. The common property of conventional controlled release (CR) technologies is that a large part of the drug load is released in the colon, where the dosage form stays for a relatively long period. This delivery approach, while desirable for many molecules, was found to be inappropriate for drugs that are poorly absorbed from the lower portion of the GIT. Under certain circumstances extending the gastric retention of a DDS is necessary for achieving the bigger therapeutic advantage of the drug. The benefit of gastric retention is for those drugs that are absorbed in the proximal portion of the GIT, and the drugs that are having a solubility less or a dip in the alkaline pH may advantage of gastric retention [8]. Drug delivery to the proximal small intestine and local and sustained drug delivery to the abdominal to treat certain diseases, extending gastric retention of the therapeutic substance may offer several benefits including enhancement of therapeutic efficacy and possible reduction of the dose size, improved bioavailability [9]. Fundus and body is a component part of the proximal stomach, which functions as a reservoir for swallowing materials, whereas the major site of mixing motions is the distal region (pylorus), conveying as a pump to push gastric contents for GE [10, 22]. In, fasting as well as fed states the GE occurs. As a consequence of gastric contractions, the GE occurs and nature depends upon the contents of the stomach. The appropriate classification of GE can be digestible solids, indigestible solids and GE of liquid. Due to the generation of intragastric pressure because of the gentle muscular contractions happening mainly from the proximal stomach, i.e. from the upper body of the stomach the liquids can be removed [11]. Compare to all the DDS the FDDS have a bulk density is lesser than gastric fluids and so longer buoyant in the stomach without disturbing GE rate for an extended period of time [22].

MATERIALS AND METHODS

Materials

Losartan potassium was procured from Farma glow Ltd, Mumbai, India and guar gum were procured from Oxford Chemicals, Delhi. Gum karaya and HPMC k15m are obtained from NR chemicals limited, Mumbai. Talc and magnesium stearate were procured from SD Fine Chem. Ltd, Mumbai, India, and sodium bicarbonate were obtained from Rankem Ltd., Delhi. The residual reagents which were used having an analytical grade [16, 17].

Methods

Preparation of losartan potassium floating tablets

All the required ingredients sufficient for 20 tablets were weighed accurately and thoroughly mixed after passing through the sieve no.
Vt for determining powder flow is Carr’s index. The powder arch shape and cohesiveness of particles, particle size distribution, was assessed using the compressibility index, which is defined as:

\[ \rho_t = \frac{M}{V_t} \]

Where,
- \( \rho_t \) = tapped density (kg/m\(^3\)),
- \( M \) = mass of powder (kg),
- \( V_t \) = tapped volume of powder (m\(^3\)).

Compressibility index

From the values of bulk and tapped densities a secondary method for determining powder flow is Carr’s index. The powder arch potency or bridge strength and stability of the powder were directly measured by the percentage compressibility. Each formulation’s Carr’s index was calculated by the below-given equation [15, 18].

\[ \text{Carr's index} = \left( \frac{\rho_t - \rho_b}{\rho_b} \right) \times 100 \]

Where,
- \( \rho_t \) = tapped density (kg/m\(^3\)),
- \( \rho_b \) = bulk density (kg/m\(^3\))

Hausner’s ratio

Hausner’s ratio is calculated by tapped density versus bulk density [18].

\[ \text{Hausner’s ratio} = \left( \frac{\text{Tapped Density}}{\text{Bulk Density}} \right) \]

Post-compression parameters

Tablet thickness and diameter

Thickness, as well as the diameter of the tablets, was determined by using a vernier caliper. By picking three tablets from each formulation individually and randomly the tablet’s thickness was measured [14, 16, 17].

Hardness

The hardness test is used to check the binding strength of a tablet. Due to tablets, insufficient hardness may undergo chipping or breakage during transportation or breakage during handling. Randomly five tablets were selected and with the help of Monsanto hardness tester hardness of each tablet was calculated. The tablet hardness is commonly measured in kg/cm\(^2\)[16, 21].

Friability

By taking randomly 20 tablets into Roche friabilitator, the friability of tablets was determined. It is expressed in percentage. Primarily 20 tablets were weighed and transferred into Roche friabilitator. At 25 rpm for 4 min the friabilitator drum was rotated, or the drum was rotated total 100 revolutions. After completion of 100 revolutions, the 20 tablets were removed and again weighted. The weight loss percentage was then measured by the below-given equation [15, 16, 17].

\[ f = \left( \frac{W_e - W_f}{W_e} \right) \times 100 \]

Where,
- \( f \) = friability,
- \( W_e \) = initial weight,
- \( W_f \) = final weight.

Uniformity of weight

The weight variation arises due to the non-uniform size of granules, poor flow property, and mechanical problems. If the granular size is large, non-uniformly the dies will be filled. And due to the formation of non-uniform size tablets, the drug content also became non-uniform.

Method: Uncoated tablets comply with this test.

The 20 tablets average weight is determined and compared with individual weight. From the average weight, not more than two tablets deviate from a percentage larger than that given. If more than two tablet weight is outside the average percentage weight then (according to IP) the test will be failed [14, 15, 17].

Uniformity of drug content

In the formulation, the drug content was checked the dose uniformity. Randomly 10 tablets were selected individually and powered. The randomly selected powdered individual tablets were placed in a 100 ml volumetric flask containing pH 1.2 buffer solution and left undisturbed overnight. The drug content was determined after suitable dilutions by U. V-spectrophotometer at 234 nm against blank [14, 19, 21].
Buoyancy studies

To evaluate the floating lag time and extent of floating the *in vitro* buoyancy was conducted. The glass beaker containing 0.1 M HC1 250 ml buffer pH 1.2 into it, the tablets were placed. Then the floating lag time (tablet to reach from the water inside to surface) and entire floating time (tablet's floating duration) were measured [14, 19, 20].

Drug excipients compatibility studies

In all pharmaceutical dosage forms, the excipients are found almost as integral components. The careful selection of the excipients is the successful formulation of a stable and effective solid dosage form. The role of excipients to simplify administration, promote the bioavailability and the constant release of the drug and also protect it from deterioration. One of the strongest analytical techniques to detect functional groups of a drug is infrared spectroscopy. Fourier transform infrared spectroscopy (FTIR) studies the interaction between a pure drug and its formulations were evaluated. In the current study, the employed method was a potassium bromide pellet method. The dry powdered potassium bromide was thoroughly mixed with the sample. The mixture was then compacted by using dies to form a disc. In the spectrophotometer, the spectrum was recorded after placing the disc [16, 17, 20].

In *in vitro* dissolution studies using pH 1.2 buffer

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### Table 1: Formulation composition of losartan potassium

| Ingredients                              | F1       | F2       | F3       | F4       | F5       | F6       | F7       | F8       | F9       |
|------------------------------------------|----------|----------|----------|----------|----------|----------|----------|----------|----------|
| Losartan potassium (% w/w)              | 138.5    | 138.5    | 138.5    | 138.5    | 138.5    | 138.5    | 138.5    | 138.5    | 138.5    |
| HPMC-K4M (% w/w)                        | -        | 103.875  | -        | -        | -        | -        | 34.625   | 34.625   | -        |
| Guar gum (% w/w)                        | -        | -        | -        | 103.875  | -        | -        | -        | -        | 34.625   |
| Gum karaya (% w/w)                      | -        | -        | -        | -        | 103.875  | 34.625   | -        | -        | 34.625   |
| Mannitol (% w/w)                        | 30.25    | 30.25    | 30.25    | 45.625   | 45.625   | 45.625   | 30.25    | 30.25    | 30.25    |
| Sodium bicarbonate (% w/w)              | 50       | 50       | 50       | 50       | 50       | 50       | 50       | 50       | 50       |
| Talc (% w/w)                             | 6        | 6        | 6        | 6        | 6        | 6        | 6        | 6        | 6        |
| Magnesium Stearate (% w/w)              | 6        | 6        | 6        | 6        | 6        | 6        | 6        | 6        | 6        |
| Total Weight (mg)                        | 300      | 300      | 300      | 350      | 350      | 350      | 300      | 300      | 300      |
| HPMC-Hydroxy propyl methyl cellulose    |          |          |          |          |          |          |          |          |          |

**Table 2: Pre-compression parameters**

| Formulation | Bulk density (kg/m^3)* | Tapped density (kg/m^3)* | Carr’s index (%)* | Hauser’s ratio* | Angle of repose (°)* |
|-------------|-----------------------|--------------------------|-------------------|-----------------|---------------------|
| F1          | 0.359±0.06            | 0.423±0.05               | 15.13±0.17        | 1.17±0.03       | 20.14±0.41          |
| F2          | 0.375±0.25            | 0.441±0.12               | 14.97±0.14        | 1.17±0.01       | 25.11±1.27          |
| F3          | 0.355±0.15            | 0.412±0.31               | 13.83±0.19        | 1.16±0.03       | 20.14±0.01          |
| F4          | 0.326±0.17            | 0.377±0.15               | 13.53±0.24        | 1.15±0.01       | 29.2±0.94           |
| F5          | 0.361±0.18            | 0.423±0.18               | 14.66±0.16        | 1.16±0.03       | 28.5±1.44           |
| F6          | 0.326±0.16            | 0.377±0.19               | 13.53±0.18        | 1.15±0.01       | 29.2±0.41           |
| F7          | 0.392±0.17            | 0.443±0.17               | 11.51±0.24        | 1.13±0.04       | 25.27±0.59          |
| F8          | 0.363±0.21            | 0.423±0.05               | 14.18±0.21        | 1.18±0.03       | 22.8±0.93           |
| F9          | 0.365±0.16            | 0.416±0.06               | 12.26±0.04        | 1.14±0.02       | 25.23±0.32          |

*Mean±standard deviation (SD). n=3

**Table 3: Post-compression parameters**

| Formulation | Thickness (mm)* | Hardness (kg/cm)* | Friability (%)* | Uniformity of weight (mg)* | Assay (%)* |
|-------------|-----------------|-------------------|-----------------|---------------------------|-----------|
| F1          | 4.02            | 5.02              | 0.46            | 300.4         | 99.47     |
| F2          | 3.94            | 5.94              | 0.35            | 279.39        | 98.25     |
| F3          | 4.13            | 5.30              | 0.26            | 298.69        | 98.29     |
| F4          | 3.93            | 5.42              | 0.45            | 301.12        | 98.53     |
| F5          | 4.24            | 5.24              | 0.31            | 296.45        | 99.49     |
| F6          | 4.06            | 5.06              | 0.52            | 297.26        | 97.27     |
| F7          | 4.06            | 5.21              | 0.23            | 297.59        | 98.68     |
| F8          | 3.95            | 5.38              | 0.15            | 299.35        | 99.24     |
| F9          | 4.16            | 5.12              | 0.14            | 298.95        | 99.39     |

*Mean±standard deviation (SD). n=3

**Drug release kinetics**

To inspect the drug release mechanism and kinetics, the cumulative percentage of drug release data were fitted to models presenting zero-order (cumulative percentage of drug release versus time), first-order (log cumulative percentage of drug remaining versus time), Higuchi (cumulative percentage drug release versus square root of time), Hixon-Crowell (cube root of drug percentage remaining versus time in 'h') and Korsmeyer-Peppas (log cumulative percentage drug release versus log time) correspondingly [14, 17, 20].

**RESULTS AND DISCUSSION**

Pre-compression parameters of losartan potassium granules

The losartan potassium formulations F1 to F9 the Carr’s index and Hausner’s ratio data between 11.51 (F7) to 15.13 (F1) and 1.18 (F8). The losartan potassium prepared tablets bulk density and tapped density was found in between 0.326 (F6) to 0.392 (F7) and 0.377 (F4) to 0.443 (F7) respectively. The angle of repose values range is in between 20.14 (F1) to 29. 2 (F6) indicate good flow property about the granules.

Post-compression parameters

In the formulations, the drug content values vary from 98.25% (F2) to 99.49% (F5). The hardness and friability of the prepared gastro retentive drug delivery system (GRDDS) of losartan potassium were found in between the range of 5.02 (F1) to 5.94 (F2) kg/cm² and 0.14% (F9) to 0.52% (F6). The thickness and weight variation data of the GRDDS of losartan potassium tablets were found in between 3.93 mm (F4) to 4.24 mm (F5) and 296.45 mg (F5) to 301.12 mg (F4), were presented in table 3.
Table 4: In vitro buoyancy properties of losartan potassium

| Formulation | Floating lag time (s)* | Total floating time (h)* |
|-------------|------------------------|--------------------------|
| F1          | 63                     | >12                      |
| F2          | 75                     | >12                      |
| F3          | 95                     | >12                      |
| F4          | 70                     | >12                      |
| F5          | 85                     | >12                      |
| F6          | 92                     | >12                      |
| F7          | 87                     | >12                      |
| F8          | 91                     | >12                      |
| F9          | 98                     | >12                      |

*mean±standard deviation (SD), n=3

Floating property

The all GRDDS of losartan potassium floating lag time values vary in between 63 and 98 s and total floating time was found more than 12 h.

FTIR studies

In the current study, Fourier transforms infrared spectroscopy (FTIR) data of the best formulation were matched with the standard spectrum of pure drug losartan potassium over the range 400-4000 cm⁻¹ analyzed. The spectrum of pure losartan potassium shows prominent and strong absorption bands at wave numbers of 1256.53 cm⁻¹, 2870.26 cm⁻¹, 3172.30 cm⁻¹ and 762.24 cm⁻¹ corresponding to cyclic amines, C-H stretches, O-H bending and Chlorine, respectively. The FTIR spectrum of the optimized formulation displayed the characteristic bands of both drug and excipient without any significant spectral shift. This suggested there was no potential chemical interaction between the components of the formulations.

Fig. 1: FTIR spectrum of pure losartan potassium

Fig. 2: FTIR spectrum of pure losartan potassium with excipients
**Fig. 3:** Percentage drug release of losartan potassium from formulation F1 to F9, all values were calculated mean±standard deviation; n=3

Table 5: *In vitro* drug release kinetic studies of different formulations

| Formulation | Zero-order (R²) | First order | Higuchi | Hixon Crowell | Release exponent (n) |
|-------------|----------------|-------------|---------|---------------|---------------------|
| F1          | 0.975          | 0.629       | 0.926   | 0.802         | 0.868               |
| F2          | 0.962          | 0.71        | 0.958   | 0.854         | 0.669               |
| F3          | 0.93           | 0.613       | 0.863   | 0.754         | 0.760               |
| F4          | 0.949          | 0.976       | 0.978   | 0.969         | 0.987               |
| F5          | 0.981          | 0.985       | 0.986   | 0.989         | 0.759               |
| F6          | 0.99           | 0.988       | 0.988   | 0.993         | 0.780               |
| F7          | 0.989          | 0.715       | 0.959   | 0.872         | 0.913               |
| F8          | 0.976          | 0.842       | 0.965   | 0.919         | 0.515               |
| F9          | 0.951          | 0.668       | 0.894   | 0.813         | 0.496               |

**In vitro drug release profile**

*In vitro* dissolution studies were performed for all the batches of GRDDS of losartan potassium using USP XXIII dissolution test apparatus-II at 50 rpm, 900 ml of 0.1N HCl (pH 1.2) used as dissolution media. The *in vitro* drug release data was given in fig. 3. All the tablet formulations showed more than 8% drug release within 1 h, except F4, but formulation F8 showed maximum 23.17% drug release within 1 h. After the ideal study, drug release for formulations F1, and F4 (with HPMC k4m) were found to be 98.64%, after 10 h and 60.28% after 12 h, respectively. Formulations F2 and F5 have guar gum the drug releases 97.56%, after 10 h and 64.83% after 12 h, respectively, and same like Formulations F3 and F6 having gum karaya the drug releases 98.65%, after 10 h and 65.24% after 12 h, respectively. As the drug release could only be observed up to 10 h in F1-F3 with the sustained release polymer concentration of 23%, a further high concentration of polymer was employed in F4-F6 to sustain the drug release up to 12 h, as the drug release was found to be a function of polymer concentration. As these results were far behind the satisfactory release rate, a combination of polymers in 1:1 ratio the concentration of 23% was employed to negotiate the slow release characteristics of F7 to F9 formulations, and the drug release was found to be optimum in F9 (97.77%) drug release having two natural gums, one is water-soluble (guar gum) and the other is water-insoluble (gum karaya) compared to F7 and F8. Hence, F9 was considered to be the optimized formulation. The drug release data were then subjected to Korsmeyer-Peppas equation for determination of release mechanism and the release exponent ‘n’ varied from 0.496-0.987 that indicates F9 having Fickian diffusion and remaining F1 to F8 followed non-Fickian diffusion [13].

**CONCLUSION**

This study discourses the formulation and evaluation of gastroretentive tablets of losartan potassium. The effervescent based floating drug delivery was a hopeful approach to achieve *in vitro* buoyancy. The addition of polymer HPMC k4m, natural polymers’ guar gum, gum karaya and gas generating agent sodium bicarbonate was important to achieve *in vitro* buoyancy. Formulation F9 showed a preferred drug release profile up to 12 h following zero-order release kinetics and formulations F1 to F8 followed non-Fickian diffusion except F9 was having Fickian diffusion. Thus, the conclusion of this research work clearly points out, a promising potential of this losartan potassium floating prolong release dosage form is as a substitute to the conventional dosage form for the management of hypertension.

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**AUTHORS CONTRIBUTIONS**

All the authors have contributed equally

**CONFLICTS OF INTERESTS**

Declared none
REFERENCES

1. Streubel A, Siepmann J, Bodmeier R. Drug delivery to the upper small intestine window using gastro-retentive technologies. Curr Opinion Pharmacol 2006;6:501-8.
2. Bardonnet PL, Faivre V, Pugh WJ, Piffaretti JC, Falson F. Gastroretentive dosage forms: overview and the special case of Helicobacter pylori. J Controlled Release 2006;111:1-18.
3. Timmermans J, Moees AJ. How well do floating dosage forms float? Int J Pharm 1999;6:207-16.
4. Streubel A, Siepmann J, Bodmeier R. Floating matrix tablets based on low-density foam powder: effects of formulation and processing parameters on drug release. Eur J Pharm Sci 2003;18:37-45.
5. Nayak AK, Malakar J, Sen KK. Gastroretentive drug delivery technologies: current approaches and future potential. J Pharm Educ Res 2010;1:1-10.
6. Kotthari Abhaykumar H, Manish J. Gastroretentive drug delivery system and its approaches: a review. Int J Pharm Res Development 2013;4:7-16.
7. Rouge N, Buri P, Doelker E. Drug absorption sites in the gastrointestinal tract and dosage form for site-specific delivery. Int J Pharm 1996;136:117-39.
8. Fell JT, Whitehead L, Collet H. Prolonged gastric retention using floating dosage forms. Pharm Technol 2000;24:82-90.
9. Streubel A, Siepmann J, Bodmeier R. Gastroretentive drug delivery system. Expert Opin Drug Delivery 2006;3:217-33.
10. Bramhankar DM, Jaiswal SB. Controlled release medication. Jain MK editor. Design of controlled drug delivery systems. 1st ed. Delhi: Vallabh Prakashan; 2002. p. 335-7.
11. Minami H, McCallum RW. The physiology and pathophysiology of gastric emptying in humans. Gastroenterology 1984; 86:1592-610.

12. Patel VF, Patel NM, Yeole PG. Studies on the formulation and evaluation of ranitidine floating tablets. Indian J Pharm Sci 2005;67:703-9.
13. Banker GS, Rhodes CT. Target-oriented drug delivery systems and packaging of pharmaceutical dosage forms. Banker GS, Rhodes CT editors. 3rd ed. New York: Marcel Dekker; 1996. p. 678-721.
14. Samyuktha M, Vasanth PM, Suresh K, Ramesh T, Ramesh M. Formulation and evaluation of gastroretentive floating tablets of losartan potassium. Int J Biopharm 2013;4:18-26.
15. Sandhyarani S, Ramesh A, Krishna KVM. Formulation and evaluation of stable floating tablet of losartan potassium for oral controlled drug delivery system. J Pharm Sci Res 2015;7:946-51.
16. Audumbar DM, Ritesh SB. Development and evaluation of gastroretentive floating tablets of a quinapril HCL by direct compression technique. Int J Pharm Sci 2017;9:35-46.
17. Ahmed AA, Wedad KA, Al-saady FA. Formulation and evaluation of prochlorperazine maleate sustained floating release tablet. Int J Pharm Sci 2017;9:89-98.
18. Manojkumar SP, Vidyasagar G, Patil VB. Formulation, optimization and evaluation of floating tablets clarithromycin. Int J Pharm Sci 2015;7:320-6.
19. Radhika PR, Nishala N, Kiruthika M, Iswarya S. Design and evaluation of intragastric buoyant tablets of venlafaxine hydrochloride. Asian J Pharm Clin Res 2017;10:166-70.
20. Sreejan M, Jayasri K, Kancherla RA, Lakshmi KK. Alginate-based gastro-retentive raft forming tablets for enhanced bioavailability of tinidazole. Int J Appl Pharm 2017;9:16-21.
21. Bharat WT, Umesh TJ, Shruti GP, Vijay RP. Formulation and in vitro evaluation of floating tablets of cepodoxime proxetil. Int J Curr Pharm Res 2017;9:18-22.
22. Shah HP, Prajapati ST, Patel CN. Gastroretentive drug delivery systems: from conception to commercial success. J Crit Rev 2017;4:10-21.