Comparison of mannitol and methacholine to predict exercise-induced bronchoconstriction and a clinical diagnosis of asthma

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Abstract

Background: Asthma can be difficult to diagnose, but bronchial provocation with methacholine, exercise or mannitol is helpful when used to identify bronchial hyperresponsiveness (BHR), a key feature of the disease. The utility of these tests in subjects with signs and symptoms of asthma but without a clear diagnosis has not been investigated. We investigated the sensitivity and specificity of mannitol to identify exercise-induced bronchoconstriction (EIB) as a manifestation of BHR; compared this with methacholine; and compared the sensitivity and specificity of mannitol and methacholine for a clinician diagnosis of asthma.

Methods: 509 people (6–50 yr) were enrolled, 78% were atopic, median FEV1 92.5% predicted, and a low NAEPPII asthma score of 1.2. Subjects with symptoms of seasonal allergy were excluded. BHR to exercise was defined as a ≥10% fall in FEV1 on at least one of two tests, to methacholine a PC20 ≤16 mg/ml and to mannitol a 15% fall in FEV1 at ≤635 mg or a 10% fall between doses. The clinician diagnosis of asthma was made on examination, history, skin tests, questionnaire and response to exercise but they were blind to the mannitol and methacholine results.

Results: Mannitol and methacholine were therapeutically equivalent to identify EIB, a clinician diagnosis of asthma, and prevalence of BHR. The sensitivity/specificity of mannitol to identify EIB was 59%/65% and for methacholine it was 56%/69%. The BHR was mild. Mean EIB % fall in FEV1 in subjects positive to exercise was 19%, (SD 9.2), mannitol PD15 158 (CI:129,193) mg, and methacholine PC20 2.1 (CI:1.7, 2.6)mg/ml. The prevalence of BHR was the same: for exercise (43.5%), mannitol (44.8%), and methacholine (41.6%) with a test agreement between 62 & 69%. The sensitivity and specificity for a clinician diagnosis of asthma was 56%/73% for mannitol and 51%/75% for methacholine. The sensitivity increased to 73% and 72% for mannitol and methacholine when two exercise tests were positive.

Conclusion: In this group with normal FEV1, mild symptoms, and mild BHR, the sensitivity and specificity for both mannitol and methacholine to identify EIB and a clinician diagnosis of asthma were equivalent, but lower than previously documented in well-defined populations.

Trial registration: This was a multi-center trial comprising 25 sites across the United States of America. (NCT0025229).
Background
Asthma can be difficult to diagnose and no single symptom or test should be used in isolation to make the diagnosis. A correct diagnosis is important in order for patients to receive appropriate therapy [1]. Because bronchial hyperresponsiveness (BHR) is a hallmark of asthma, bronchial provocation challenge tests (BPTs) with a variety of agents have been used to assist in its diagnosis [2-4].

Methacholine is a commonly used agent, delivered as a wet aerosol. Methacholine acts directly on acetylcholine receptors on smooth muscle causing contraction and airway narrowing. Methacholine has been reported to have a high sensitivity to identify BHR and a negative test is often used to exclude asthma [5,6].

Provocation tests that use indirect stimuli (e.g. exercise and mannitol) have a high specificity for asthma [7] causing smooth muscle contraction by release of endogenous mediators including prostaglandins, leukotrienes, and histamine [8,9]. Evaporative water loss occurs in conditioning the inspired air and causes exercise induced bronchoconstriction (EIB) by inflammatory mediators of mast cell origin [10,11]. Exercise is generally recognised as having a low sensitivity to identify BHR; EIB is consistent with a diagnosis of asthma [12] and responds to chronic treatment with inhaled corticosteroids (ICS) and other drugs used in the treatment of asthma [13-15].

A dry powder of mannitol has been developed as an indirect BPT [16] and is available as a standardized test kit. The test kit contains pre-filled mannitol capsules in escalating doses and a hand-held dry powder inhaler device. Safety and efficacy of mannitol as a BPT were established in a large Phase III clinical trial in patients with asthma and in healthy subjects [7].

The usefulness of mannitol as a BPT in subjects with signs and symptoms of asthma but no clear diagnosis has not been investigated previously. The aim was to study a large population of subjects to compare the sensitivity and specificity of mannitol with methacholine to detect EIB as a manifestation of BHR and to identify a clinician diagnosis of asthma.

Subjects: inclusion criteria
Subjects aged 6–50 years (BMI < 35) with signs and symptoms suggestive of asthma according to the National Institute of Health (NIH) Questionnaire [17] but without a firm diagnosis of asthma or an exclusion of the diagnosis of asthma (e.g. had an equivocal diagnosis of asthma or had been referred for further investigation of asthma-type symptoms) were included. Subjects had at least Step 1 symptoms according to the NAEPPII asthma severity grading (symptoms ≤ 2 times per week; asymptomatic between exacerbations; exacerbations of only a few hours to a few days; and night time symptoms of ≤ 2 times per month). They were required to have an FEV₁ ≥ 70% of the predicted value at the Screening Visit [18,19].

Subjects were excluded from participating in this study if they: had any known other pulmonary disease; had smoked more than 1 cigarette per week within the past year or had a ≥ 10 pack year smoking history; had a respiratory tract infection within the previous 4 weeks; had been skin test positive to aeroallergens that were present in the environment during the time of enrolment and reported worsening of symptoms when exposed to these aeroallergens during the study; had been diagnosed at Screening Visit as definitively having asthma (95 to 100% likelihood) or not having asthma (0 to < 5% likelihood); had clinically significantly abnormal chest x-ray or ECG; or had failed to observe washout of medications that would interfere with BPT (including, but not limited to, no use of corticosteroids within 4 weeks of the Screening Visit).

Methods
The protocol was approved by a central institutional review board. Each subject or parent gave informed consent in writing. The study consisted of 5 visits to the clinic. On the Screening Visit the following were assessed: eligibility; demographic data; medical history; medications; spirometry with reversibility (following 360 mcg of albuterol); allergy skin test reactivity to common allergens (positive test taken as 3 mm wheal). The NIH NAEPPII questionnaire was answered and a score was assigned [20]. Vital signs including blood pressure, heart rate, and respiratory rate were measured in the sitting position and an ECG performed. Based on this information, a respiratory physician assigned one of 6 diagnoses at this visit on the basis of the likelihood of asthma as follows: asthma is extremely likely or definite (95%–100% likelihood); asthma is very likely (72.5 to < 95%); asthma is probable (50 to < 72.5%); asthma is possible (27.5 to < 50%); asthma is unlikely but cannot be excluded (5 to 27.5%); and asthma is very unlikely (0–<5%). Those with 5 to 95% likelihood were included in the study.

Visit 2 occurred 1–4 days after Visit 1 and within 2 hrs of the time of Screening. Adverse events, medications, and withholding times were reviewed (Table 1), and spirometry measured to confirm values on the screening day. This was followed by a brief physical examination. Exercise was performed with vital signs being measured before and after exercise. At Visit 3, the procedures were the same as Visit 2 and occurred within 1–4 days. At Visit 4, adverse events and medications were reviewed, withholding times were checked, and spirometry was performed to confirm FEV₁ was within 15% of Visit 1. The challenge agent was
either mannitol or methacholine, and the choice was randomly determined. The time of the test was documented for each challenge. Vital signs were measured in the sitting position before and after the challenge test. Cough and pulse oximetry were recorded during mannitol challenges. Full spirometry was measured before and at 15 minutes after completion of the mannitol challenge with FEV₁ only being performed after each dose. ECG was performed before and after mannitol challenge. Visit 5 was a repeat of the procedures of Visit 4 with the reciprocal challenge being administered (either mannitol or methacholine). A respiratory physician then assigned one of the diagnoses of likelihood of asthma evaluated at the Screening Visit, above. The NAEPII asthma severity grading score was also re-evaluated at Visit 5 but not necessarily by the same physician.

### Bronchial provocation tests

#### Exercise challenge

Exercise was performed by running on a motorised treadmill whilst breathing medical grade dry air (20–25°C) from a Douglas Bag [14]. Briefly exercise was ramped up

| Table 1: Medications and other factors that may decrease bronchial hyperresponsiveness and their required withholding periods |
|----------------------------------------------------------|
| **FACTOR** | **Withholding Period** |
| Inhaled agents | Short acting bronchodilators (isoproterenol, isetharine, metaproterenol, albuterol, levalbuterol, terbutaline) (e.g. Proventil® or Ventolin®) | 8 hr |
| | Inhaled anticholinergics or combination products (e.g. Atrovent® or Combivent®) | 1 week |
| | Medium acting bronchodilators (ipratropium) | 1 week |
| | Long acting inhaled bronchodilators (salmeterol, formoterol) (e.g. Serevent® or Foradil®) | 2 weeks |
| | Inhaled corticosteroid/long acting inhaled bronchodilator combination (e.g. Advair®) | 4 weeks |
| Oral bronchodilators | Theophylline | 24 hr |
| | Intermediate theophylline | 48 hr |
| | Long acting theophylline | 48 hr |
| | Standard β-agonist tablets | 24 hr |
| | Long acting β-agonist tablets | 48 hr |
| Corticosteroids | There is no washout for topical steroids applied to skin unless they are high potency steroids | 4 weeks |
| Other medications | Hydroxyzine, cetirizine (and other antihistamines) | 72 hr |
| | Tiotropium bromide | 72 hr |
| | Nasals steroids | 1 week |
| | β-blockers | 1 week |
| | Cromolyn sodium | 2 weeks |
| | Nedocromil | 2 weeks |
| | Leukotriene modifiers | 6 weeks |
| Foods | Coffee, tea, cola drinks, chocolate (caffeinated foods) | 12 hr |
| Strenuous exercise or exposure to cold air to a level that would be expected to interfere with challenges | 12 hr |
| Tobacco | 6 hr |
Mannitol and methacholine responses were log transformed for calculation of the geometric means. The analysis plan specified a test of non-inferiority to be achieved if the lower 95% credible limit for the adverse-event-free-ratio \((1 - \text{AER}_{\text{mannitol}})/(1 - \text{AER}_{\text{methacholine}})\) exceeded 0.80.

Analysis

The sensitivity and specificity of a mannitol positive test (defined as ≥ 15% fall in FEV\(_1\) ≤ 635 mg or a ≥ 10% fall in FEV\(_1\) between consecutive doses) and a methacholine positive test (defined as PC\(_{20}\)FEV\(_1\) ≤ 16 mg/ml) with respect to EIB (defined as ≥ 10%, 15% or 20% fall in FEV\(_1\) after a standardized treadmill run) and with respect to a clinician's diagnosis of asthma at Visit 5 were calculated. Additional analyses were performed excluding those with a mannitol test taking longer than 35 minutes because the osmotic gradient will not progressively increase if the time between doses is prolonged.

Safety data

Vital signs (including blood pressure, heart rate, and respiratory rate) using the intent-to-treat population (ITTTP, n = 391) and their changes during challenge tests are described. The adverse events were tabulated following each challenge test. The per-protocol population (PPP, n = 375) included all subjects with no major protocol violations who completed all of the required challenge tests, including methacholine and mannitol challenges.

In the protocol cough was identified as an adverse event if it prevented the challenge from being completed in which case it was documented as severe at the time of testing.

Blinding

A respiratory physician was to make the Clinician diagnosis at the final visit (Visit 5) with access to the data on the exercise challenges, history, examination, skin tests, and FEV\(_1\) reversibility but not the mannitol and methacholine challenge test result. Site staff members performing mannitol and methacholine challenges were not provided with other diagnostic information about the subject to assure that there was no bias in the performance of these tests. Mannitol and methacholine challenges were given in a restricted randomization scheme that produced equal numbers of each order.

To accomplish proper blinding, the mannitol and methacholine challenge team did not have access to the electronic case report form (eCRF) or to other physiological data. Similarly, other site personnel did not have access to the mannitol and methacholine challenge data. Mannitol and methacholine challenge data were able to be reviewed by the Data Manager at CompeWare Inc but they were only provided to the Sponsor (Pharmaxis Ltd) at the end of the study.
Results

The disposition of the subjects is given in Figure 1. There were 16 people in the ITTP who were not included in the PPP. The data are given for the PPP unless otherwise stated because it represents the results for all the subjects who performed all the tests. The age distribution for 375 subjects in the PPP are presented in Table 2. The characteristics of the subjects are summarised in Table 3. There were 96 children and 279 adults (> 18 years); 51.3% female; 76.3% Caucasian, 8.3% Hispanic and 8.5% Black; with near-normal baseline spirometry; with low NAEPPI asthma score of 1.2 (SD 0.5); 78% atopic; and 7.5% responding positively to a bronchodilator. Of the 375, 145 were considered by the respiratory physician to have a 50% or more likelihood of having asthma at Visit 1. There were 163 (43.5%) subjects who had a positive response to exercise (Exc+) with a ≥ 10% fall in FEV1 on at least one test; 119 recording this on the first exercise test with 66 of these 119 recording a ≥ 15% fall in FEV1. There were 168 (44.8%) subjects with a positive test to mannitol (Mann+). Of these, 109 achieved a 15% fall in 635 mg and the remaining achieved a positive test by a 10% fall in FEV1 between consecutive doses. Seventy three percent achieved this response within the first 6 doses of mannitol (≤ 315 mg). There were 156 (41.6%) with a positive test to methacholine (Mech+) with a PC20 ≤ 16 mg/ml, and 26% achieved this response within the first 6 concentrations (≤ 1 mg/ml) being delivered. The percentile values and median data for the positive responses are given in Table 4.

Sensitivity and specificity of mannitol and methacholine to identify a subject with different levels of severity of EIB is given in Table 5. There was no significant difference between mannitol and methacholine to identify EIB; however, these agents did not necessarily identify the same people and the agreement between the mannitol and methacholine test results was 69%. Agreement for mannitol and exercise was 62% and for methacholine and exercise was 63%. The relationship between the reactivity to mannitol expressed as log RDR and the reactivity to exercise expressed as the maximum % fall in FEV1 was significant but not strong (r = 0.256, p < 0.001, n = 312). Maximum % fall in FEV1 to mannitol in relation to methacholine (r = 0.41 p < 0.0001) in those positive and negative to exercise is illustrated in Figure 2. Forty-six (29%) people had a fall ≥ 30% in FEV1 after methacholine, 2 (1.2%) after mannitol and 25 (15.3%) after exercise. The mean maximum % fall (SD) in FEV1 after those with a positive methacholine challenge was 27.7% ± (8.2) and after a positive mannitol challenge was 16.1% ± (5.6). The fall after mannitol was 19.2% ± (4) excluding those who were positive due to a 10% fall between doses, and this was the same as the fall after exercise19.1% (9.4). Sensitivity and specificity of mannitol and methacholine to identify Exc+ 10% in relation to symptoms score is summarised in Table 6.

Mannitol had a sensitivity of 67% to identify a Mech+ PC20 of ≤ 16, 68% for ≤ 12, and 68% for ≤ 8 mg/ml and a sensitivity of 77% to identify ≤ 4 mg/ml and 83% to identify ≤ 2 mg/ml. Methacholine had a sensitivity of 62% for identifying a Mann+ response. An Exc+10% on the first test had a sensitivity of 62% to identify Exc+10% on the second test. The receiver operator curves for mannitol and methacholine in relation to identifying EIB is illustrated in Figure 3 and are almost identical for mannitol and methacholine. The negative and positive predictive values for mannitol and methacholine were close generally differing by 0.01 or less. Similarly the negative (0.64 and 0.65) and positive likelihood ratios (1.71 and 1.79) were similar for mannitol and methacholine respectively.

Sensitivity and specificity for mannitol and methacholine to identify a 10% fall after exercise in children and a clinical diagnosis at Visit 5 is given in Table 7. Of the 375 PPP, there were 240 (64%) identified as having a clinical diagnosis of asthma at Visit 5 (ClinDx5+) and 48% of these had a likelihood of asthma (see definitions above) greater than 50% assigned at Visit 1. Fifteen percent of the subjects who received a clinical diagnosis of asthma were negative to all three challenges. Of the 135 who did not receive a clinical diagnosis of asthma at Visit 5 (ClinDx5_), 78% had a likelihood of asthma of less than 50% assigned at Visit 1. Sensitivity and specificity of mannitol to predict ClinDx5+ was 55% and 73%. The sensitivity for mannitol rose to 73% when ClinDx5+ was associated with two Exc+ 10% tests. The comparable sensitivity and specificity for methacholine (PC20 ≤ 16 mg/ml) was 51% and 75% and sensitivity rose to 72% when ClinDx5+ was associated with two Exc+10% tests.

The positive and negative predictive values for mannitol for a ClinDx5+ were 79% and 48% and for methacholine they were 78% and 46%. There were 106 subjects negative to mannitol, 118 negative to methacholine and 92 negative to exercise who were given a clinician diagnosis of asthma at Visit 5. When the subjects who took longer than 35 min for a mannitol challenge were excluded, the sensitivity of mannitol to identify a 95% risk of having asthma as judged by the clinician was 76%. For methacholine this equivalent value was 67%.
Figure 1
Subject disposition.
The time to perform a positive mannitol test was significantly shorter than a positive methacholine test by approximately 25 min (19.9 min [SE 0.9] vs 44.5 min [SE 1.4]). In those Mech+ only it was 48.3 min (SE 1.7) and for those Mann+ only it was 19.2 min (SE 1.3).

Time for recovery of FEV₁ to 95% of baseline was similar for all tests. Mannitol was 21.6 min (SD 9.0) vs. methacholine 21.06 min (SD 6.96); the maximum time for recovery on the two exercise tests was 22.9 min (SD 13.7). Median time for recovery after mannitol and methacholine was 19 min (interquartile range 17–24).

Sensitivity and specificity for methacholine to identify EIB were considerably less consistent across the centres than for mannitol. The between centre standard deviation for the log odds ratio for methacholine and mannitol was respectively 1.26 vs. 0.32 for sensitivity (p < 0.02) and 0.68 vs 0.47 (p = NS) for specificity. Thus there was significantly less variation in sensitivity of mannitol to identify EIB between centres compared with methacholine.

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The skin test results for the wide variety of allergies are summarised in Table 8 for the per protocol population. The percentage of subjects positive to one or more skin tests for those Exc+ was 42.6%, for Mann+ it was 47.1%.
and for Mech+ it was 41.8%. It was higher in those achieving a ClinDx5+ 63.0%.

During mannitol challenges, frequency of cough was monitored in 419 subjects. Of these 391 had cough (93%) with 204 having occasional cough (49%) that did not interfere with the challenge, 178 frequent cough (42%) that delayed the administration of the next dose and 9 severe cough (2.1%) that caused the challenge to be stopped and an adverse event to be recorded.

Vital signs changed as expected, with increased in systolic and diastolic blood pressure, heart rate, and respiratory rate associated with both exercise challenges. Trivial increases in blood pressure were seen associated with mannitol and methacholine challenges. Oxygen saturation did not change appreciably with mannitol challenge and only 3 subjects had > 3% reduction. Heart rate increased slightly after mannitol challenge and decreased slightly after methacholine challenge. Respiratory rate was also largely unchanged with both mannitol and methacholine challenge.

### Adverse events

All adverse events which commenced after the challenges were reported by MedDRA System Organ Class and Preferred Term and tabulated (Table 9) There were more adverse events on mannitol compared with methacholine 130 vs 89. The distribution across range of severity of events was the same. There were 62 moderate adverse events for any of the challenge tests. Mannitol was non-inferior to methacholine in the sense that the proportion of subjects who did not experience adverse events in the mannitol challenge was at least 80% of the proportion who did not experience adverse events in the methacholine challenge (92.9%) as per the statistical analysis plan.

### Discussion

In these subjects with very mild symptoms suggestive of asthma and good lung function, sensitivity and specificity were equivalent for mannitol and methacholine to identify EIB and a clinical diagnosis of asthma. There were no serious adverse events associated with the tests and both were generally well tolerated.

Mannitol sensitivity to identify EIB was lower than that previously reported in subjects with a definite diagnosis of asthma [21,22]. The lower sensitivity of mannitol to identify EIB reported here is consistent with the mild EIB (50% of subjects had falls in FEV₁ ≤ 15%) documented in this group who did not have a definite diagnosis of asthma. The % fall in FEV₁ in the asthmatics reported by Brannan [21] was 40 ± 19% (SD) and by Munoz [22] it was 37% ± 16% (SD). In the study of Holzer [23] a PD₁₅ to mannitol had a sensitivity of 83% to identify EIB in a group of athletes with a 25.4% ± 15% (SD) fall in FEV₁ after eucapnic voluntary hyperpnea, a surrogate challenge for EIB.
The sensitivity of mannitol to identify a Exc+ 20% fall in FEV₁ was > 80% when those with a prolonged mannitol test time were excluded. Sensitivity of the mannitol to detect a Exc+10% fall was also > 80% in those with an asthma symptom severity score of 2 rather than 1, although the numbers with this score were smaller.

Sensitivity and specificity of methacholine and mannitol for a clinical diagnosis of asthma was equivalent in this study, but the values were lower than those reported in well-defined populations of asthmatic and healthy subjects [7] and similar to those reported for epidemiological studies [24-26]. In the children less than 18 yrs, however, mannitol had a 18.5% higher specificity (81.4% vs 62.9%) for identifying a clinical diagnosis of asthma and a 10% lower sensitivity (60.1% vs 70%) to identify EIB compared with methacholine.

This lower than expected sensitivity for mannitol and methacholine to identify EIB or a clinical diagnosis may be explained by the relatively unusual nature of this group of subjects who would not have qualified either for a clinical study on asthmatic subjects or a study on healthy subjects. The exclusion criteria required that the subjects not be symptomatic to the allergens to which they demonstrated atopy at the time of the study. This too is unusual and may affect mast cell number and IgE status. There was a lack of agreement between the clinical diagnosis and response to the challenge tests in 27% of the subjects; 15% of subjects given a diagnosis of asthma were negative to all three challenge tests and 12% with two or more positive challenge tests were deemed not to have an asthma diagnosis at Visit 5. The dose of mannitol (PD₁₅) or concentration of methacholine (PC₂₀) in those who were positive in this study was consistent with the values reported previously in clinically recognised asthmatics not taking inhaled corticosteroids [27,28].

The prevalence of BHR identified by the three tests differed by only 3.2% (from 41.6% to 44.8%). This suggests that the cut-off points used to define BHR were appropriate. Further, the range of severity of BHR was similar for all the tests with 50% being in the mild range for mannitol and exercise and 75% in the mild range for methacholine [2,29]. However it was not always the same subject responsive to all the BPTs and only 17.9% of subjects were positive to all three challenge tests. This probably reflects the natural variability of the test responses in people with mild BHR and intermittent symptoms.

Some long-held beliefs about methacholine were not upheld by the results of this study. Methacholine did not

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**Table 5: Sensitivity and specificity of challenge at different cut points for a positive test.**

| Exercise Positive Cut-Points – % fall from baseline | 10% | 15% | 20% |
|---------------------------------------------------|-----|-----|-----|
| Mannitol n = 372 | Sensitivity | 58.6 | 69.4 | 78.6 |
| | Specificity | 65.2 | 62.0 | 60.8 |
| Excluding those with challenge > 35 min n = 319 | Sensitivity | 64.1 | 75.3 | 82.7 |
| | Specificity | 59.9 | 57.0 | 55.4 |
| Methacholine 16 mg/ml n = 375 | Sensitivity | 55.2 | 67.4 | 80.3 |
| | Specificity | 68.9 | 66.1 | 65.2 |

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**Table 6: Sensitivity and specificity to identify exercise-induced bronchoconstriction (EIB) in relation to the asthma severity score.**

| NAEPP II | Visit 1 | Visit 5 |
|-------------------|---------|---------|
| Asthma Severity Score | 0 | 1 | 2 | 3 | 0 | 1 | 2 | 3 |
| N mann = 0 | 308 | 47 | 17 | 13 | 294 | 48 | 15 |
| N mech = 0 | 310 | 48 | 17 | 14 | 296 | 48 | 15 |
| Mannitol 635 mg | Sensitivity | 56.6 | 75.0 | 60.0 | 50.0 | 55.1 | 78.3 | 66.7 |
| | Specificity | 69.2 | 48.4 | 42.9 | 72.7 | 67.7 | 56.0 | 16.7 |
| Methacholine 16 mg/ml | Sensitivity | 54.7 | 62.5 | 50.0 | 0.0 | 56.3 | 56.5 | 44.4 |
| | Specificity | 72.8 | 46.8 | 71.4 | 66.7 | 70.8 | 56.0 | 66.7 |
have a higher sensitivity than exercise and mannitol to identify BHR in this population, and the prevalence of BHR was similar for all tests. Methacholine did not have a high negative predictive value for a clinical diagnosis of asthma. There were 118 subjects negative to methacholine who were given a diagnosis of asthma at Visit 5. It was also not uncommon for methacholine to miss EIB, and 73 of the 163 subjects (45%) positive to exercise were negative to methacholine, confirming previous findings in young people with good lung function [30]. The sensitivity of a positive response to the first exercise test to predict a positive response to the second test (62%) was the same as the sensitivity of mannitol and methacholine to predict at least one positive test to exercise. This low sensitivity probably relates to the variability in the mild response to exercise, particularly in those without symptoms of allergy at the time of testing. By performing a second test, an extra 44 people were identified as having EIB. Care was taken in this study to minimise the potential for variability, and there was good overall agreement between the two exercise test results; however, this was primarily due to the number of negative exercise tests.

Only one exercise test and one time point of ≥ 10% was required to be the 'gold' standard used for BHR. This criterion may be interpreted by some investigators as not strict enough because of the variability of the response to exercise. However the value for sensitivity of mannitol to identify EIB was relatively unchanged (59.8% vs 58.3%) when only those subjects with two time points on one exercise challenge were ≥ 10%, or one time point was greater than ≥ 15%, were included in the analysis. The value for sensitivity for methacholine however rose from 55.3% to 63.1% applying the same criteria.

There were 7.5% of subjects with a positive response to bronchodilator at baseline who were not excluded at entry on the basis of having a > 95% chance of having asthma, presumably as the other evidence was not supportive. This group of 28 showed a higher value for sensitivity of mannitol (68.4% vs 59%) and methacholine (73.7% vs 56%) to identify EIB when compared with the entire population.

Cockcroft [31] has found that methacholine sensitivity to identify BHR of 4 mg/ml or more is lower using the dosimeter method compared with the tidal breathing method. The dosimeter method used here is one recommended in the American Thoracic Society guidelines that categorize BHR between 4–16 mg/ml as borderline [2], is in common use, and delivery of the aerosol by this method involves a similar inspired breathing pattern as the mannitol with which it was being compared. We challenged to 16 mg/ml and, on the basis of the Cockcroft

Table 8: The frequency of positive skin tests (> 3 mm wheal) to a variety of allergens in the per protocol population (n = 375).

| Positive | Cat | Cockroach | Dog | Grass | House Dust Mites | Mold | *Cedar | Rag weed | Other Weeds | Trees |
|----------|-----|-----------|-----|-------|------------------|------|--------|----------|------------|-------|
| Tests done | 363 | 299 | 363 | 336 | 347 | 363 | 145 | 298 | 323 | 328 |
| % Positive | 42.1 | 16.1 | 20.7 | 53.6 | 50.4 | 27.3 | 38.6 | 35.6 | 39.0 | 44.5 |

*Mountain Cedar
observation, this may be equivalent to 8 mg/ml on the tidal breathing method. A PC20 of > 8 mg/ml is the value for normality illustrated in the 2007 GINA guidelines [32] and stated by the British Thoracic Society [33].

This study included steroid naïve subjects who were considered to have a greater than 5% and less than 95% probability of having asthma at entry based on spirometry, response to bronchodilator, symptoms, history and skin tests. The clinical value in comparing different diagnostic tools, such as the challenge tests used here, cannot really be judged because there is no gold standard test for the diagnosis of asthma. The strategy used here for a ‘gold standard’ for the diagnosis of asthma was to have a respi-

| System Organ Class | Preferred Term | 1st Exc | 2nd Exc | Mann | Meth | NCR |
|-------------------|----------------|---------|---------|------|------|-----|
| n                 |                | 435     | 429     | 419  | 420  | 436 |
| 1.1 All           |                | 25 (8%) | 21 (7%) | 130  | 89   | 49  |

| Cardiac Disorders | Class Totals | 1 (5%) | 0 | 6 (27%) | 13 (59%) | 2 (9%) |
|-------------------|--------------|-------|---|---------|---------|-------|
| Dizziness         |              | 1     | 5 | 13      |         | 0     |

| Gastrointestinal disorders | Class Totals | 1 (5%) | 1 (5%) | 14 (70%) | 0 | 4 (20%) |
|----------------------------|--------------|-------|--------|----------|---|-------|
| Nausea                     |              | 1     | 5      | 4        |   | 1     |

| Retching                  |              | 1     | 1      | 1        |   | 0     |

| General disorders and administration site concerns | Class Totals | 1 (4%) | 0 | 24 (86%) | 3 (11%) | 0 |
|-----------------------------------------------------|--------------|-------|---|----------|--------|---|
| Burning sensation mucosal                            |              |       |   |          |        |   |
| Feeling jitteral                                      |              |       |   |          |        |   |

| Nervous system disorders | Class Totals | 4 (20%) | 2 (10%) | 6 (30%) | 4 (20%) | 4 (20%) |
|-------------------------|--------------|---------|---------|---------|---------|---------|
| Headache                |              | 2       | 1       | 5       | 4       | 4       |

| Respiratory, thoracic and mediastinal disorders       | Class Totals | 12 (6%) | 11 (6%) | 73 (39%) | 66 (36%) | 23 (12%) |
|-------------------------------------------------------|--------------|---------|---------|----------|----------|----------|
| Chest discomfort                                      |              | 3       | 1       | 13       | 18       | 1        |
| Cough                                                 |              | 2       | 1       | 9        | 8        | 4        |
| Dyspnoea                                              |              | 4       | 3       | 12       | 22       | 1        |
| Nasal congestion                                      |              | 1       | 4       | 3        |          |          |
| Pharyngolaryngeal pain                                |              | 1       | 11      | 1        | 1        |          |
| Reversible airways obstruction                        |              | 1       | 3       | 2        |          |          |
| Rhinitis                                              |              |         |         |          | 3       | 1        |
| Rhinorrhea                                            |              | 2       | 8       |          | 1        |          |
| Wheezing                                              |              | 1       | 6       | 7        | 1        |          |

#Exc = Exercise; Mann = Mannitol; Meth = Methacholine; NCR = not challenge related
In the Phase 3 study on mannitol, in subjects with known asthma and healthy subjects without asthma, the specificity and sensitivity of mannitol to identify a clinical diagnosis of asthma was 59.8% and 95.2% and it was 88.7% and 95% when those with a negative mannitol test being treated with inhaled corticosteroids were excluded [7]. In this study in subjects with symptoms but without a definite diagnosis of asthma at entry mannitol had a sensitivity and specificity of 56% and 73% and this was no different to methacholine 51% and 75%.

In conclusion, mannitol and methacholine provided therapeutic equivalence to identify BHR, EIB, and a clinical diagnosis of asthma in a group of subjects suspected of having asthma but without a clear diagnosis. While mannitol has potential to replace other challenges for BPT, any of these tests can provide specific information and insight regarding mechanism in a particular patient.

Competing interests

SDA is the inventor of the mannitol test however the intellectual property is owned by her employer, the Sydney South West Area Health Service. SDA has not received any fees personally from Pharmaxis. SDA has undertaken research studies that were funded by Pharmaxis. She is a shareholder in Pharmaxis but holds no options. She is likely to benefit from royalties in the future.

BC is an employee and a shareholder of Pharmaxis Ltd.

JW is the President of, and is a shareholder in, CompleWare Corporation. CompleWare received a fee from Pharmaxis Ltd. for services in carrying out the clinical trial.

SN is the statistician employed by CompleWare and carried out the statistical analysis.

SS has received an honorarium from the sponsor of the trial (Pharmaxis Ltd) for chairing a pre-meeting seminar at the AAAAI conference and received a research grant for study participation from Pharmaxis.

DSP participated in the study through Colorado Allergy and Asthma Centers PC, which received a research grant for study participation from Pharmaxis Ltd.

There are no other competing interests or conflicts of interest.

Authors’ contributions

BC, JW, and SDA designed the study (after consultation with the FDA). CompleWare (which employs JW and SN) ran the clinical trial and the data acquisition, and wrote the report on which this paper is based. SN was the statistician for CompleWare. SS and DP were principal investigators who presented abstracts of the work at academic society meetings. All authors contributed suggestions and have approved the final version of the paper. SDA wrote the paper and prepared it for publication and made the decision to submit it to Respiratory Research.

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References

1. Expert Panel Report: Update on Selected Topics 2002: National Asthma Education and Prevention Program. National Institutes of Health, National Heart, Lung and Blood Institute, 2003. NIH publication no. 02-5074. 2003.
2. Crapo RO, Casaburi R, Coates AL, Enright PL, Hankinson JL, Irwin CG, Macintyre NR, McKay RT, Wanger JS, Anderson SD, Cockcroft DW, Fish JE, Stork PJ. Guidelines for methacholine and exercise challenge testing – 1999. Am J Respir Crit Care Med 2000, 161:309-329.
3. Sterk PJ, Fabbri LM, Quanjer PH, Cockcroft DW, O’Byrne PM, Anderson SD, Juniper EF, Malo J-L: Airway responsiveness: Standardized challenge testing with pharmacological and physical and sensitizing stimuli in adults. *Eur Respir J Suppl* 1999, 16:53-83.

4. Van Schoor J, Pauwels R, Joos G: Indirect bronchial hyper responsiveness: the coming of age of a specific group of bronchial challenges. *Clin Exp Allergy* 2005, 35(3):250-261.

5. Cockcroft DW: How best to measure airway responsiveness. *Am J Respir Crit Care Med* 2001, 163:1514-1515.

6. O’Byrne PM, Zamel N: Airway challenges with inhaled constrictor mediators. In *Provocation Testing in Clinical Practice Volume 5*. Edited by: Spector SL. New York: Marcel Dekker; 1995:311-324.

7. Brannan JD, Anderson SD, Perry CP, Freed-Martens R, Lassig AR, Charlton B: The safety and efficacy of inhaled dry powder mannitol as a bronchial provocation test for airway hyperresponsiveness: a phase 3 comparison study with hypertonic saline. *Respir Res* 2005, 6(144):144.

8. Brannan JD, Gulliksson M, Anderson SD, Chew N, Kumpin M: Evidence of mast cell activation and leukotriene release after mannitol inhalation. *Eur Respir J* 2003, 22(3):491-496.

9. Brannan JD, Gulliksson M, Anderson SD, Chew N, Seale JP, Kumpin M: Inhibition of mast cell PGD2 release protects against mannitol-induced airway narrowing. *Eur Respir J* 2006, 27:944-950.

10. O’Sullivan S, Roquet A, Dahlén S-E, Lassig AR, Charlton B, Anderson SD, Brannan J, Spalding N, Rodwell LT, Chan K, Inhibition of mast cell PGD2 release protects against mannitol-induced airway narrowing. *Eur Respir J* 1998, 12(1):330-335.

11. Reiss TF, Hill JB, Harman E, Zang J, Tanaka WK, Bronsky E, Guerreiro D, Hendeles L: Increased urinary excretion of LTE4 after exercise and attenuation of exercise-induced bronchospasm by montelukast, a cysteinyl leukotriene receptor antagonist. *Thorax* 1997, 52(12):1030-1035.

12. Avital A, Springer C, Bar-Yishay E, Godfrey S, Adenosine, methacholine, and exercise challenges in children with asthma or paediatric chronic obstructive pulmonary disease. *Thorax* 1995, 50:516-518.

13. Tan RA, Spector SL: Exercise-induced asthma: diagnosis and management. *Ann Allergy Asthma Immunol* 2002, 89(3):226-235.

14. Weiler JM, Nathan RA, Rupp NT, Kalberg CJ, Emmett A, Dorinsky PM: Effect of fluticasone/salmeterol administered via a single device on exercise-induced bronchoconstriction in patients with persistent asthma. *Ann Allergy Asthma Immunol* 2005, 94:65-72.

15. Pearlman DS, van Adelsberg J, Philip G, Tilles SA, Busse W, Hendeles L, Loeys T, Dass SB, Reiss TF: Onset and duration of protection against exercise-induced bronchoconstriction by a single oral dose of montelukast. *Ann Allergy Asthma Immunol* 2006, 97(1):98-104.

16. Anderson SD, Brannan J, Spring J, Spalding N, Rodwell LT, Chan K, Gonda I, Walsh A, Clark AR: A new method for bronchial-provocation testing in asthmatic subjects using a dry powder of mannitol. *Am J Respir Crit Care Med* 2009, 180(1):769-775.

17. National Heart Lung and Blood Institute: Guidelines for the Diagnosis and Management of Asthma NIH. 1997.

18. Polgar G, Promadhat V: Pulmonary Function Testing in Children: Techniques and Standards. Philadelphia: W.B. Saunders; 1971.

19. Crapo RO, Morris AH, Gardner RM: Reference spirometric values using techniques and equipment that meet ATS recommendations. *Am Rev Respir Dis* 1981, 123(6):659-664.

20. National Institute of Health: Guidelines for the Diagnosis and Management of Asthma. Institutes of Health, National Heart, Lung and Blood Institute Bethesda MD NHLBI/WHO workshop report Publication No 08-4051; 2007.

21. Brannan JD, Koskela H, Anderson SD, Chew N: Responsiveness to mannitol in asthmatic subjects with exercise- and hyperventilation-induced asthma. *Am J Respir Crit Care Med* 1998, 158(4):1120-1126.

22. Munoz PA, Gomez FP, Manrique HA, Roca J, Barbera JA, Young IH, Anderson SD, Rodriguez-Roisin R: Pulmonary gas exchange response to exercise- and mannitol- induced bronchoconstriction in mild asthma. *J Appl Physiol* 2008, 105(5):1477-1485.

23. Holzer K, Anderson SD, Chan H-K, Douglas J: Mannitol as a challenge test to identify exercise-induced bronchoconstriction in elite athletes. *Am J Respir Crit Care Med* 2003, 167(4):534-547.