Impact of percentage and particle size of sugarcane biochar on the sorption behavior of clomazone in Red Latosol

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ABSTRACT

Biochar is a carbonaceous material that has excellent potential as a fertilizer and soil conditioner. However, there is a lack of information concerning the effects of the amount and particle size of this pyrogenic material on the soil sorption capacity. In this work, evaluation was made of changes in clomazone (CMZ) sorption in a Red Latosol following soil conditioning using different percentages (0.25, 0.5, and 1% w/w) of sugarcane biochar in three particle sizes (<106, 106-508, and 508-610 µm). The conditioned soils presented apparent sorption coefficients (K_d) up to 1300 times higher than that of pure soil, besides changes in the behavior of CMZ sorption. The biochar particle size and percentage influenced sorption of the herbicide as well as its retention in the amended soil during desorption processes. Both sorption and desorption Freundlich constants were linearly correlated with the external surface area of the biochar present in the soil.

Key words: Apparent sorption coefficient, batch experiment, external surface area, factorial design, Freundlich isotherm, herbicide retention.

INTRODUCTION

The Terras Pretas de Índio (TPI) are highly fertile soils that occur in small areas of the Amazon region (Glaser and Birk 2012, Rittl et al. 2015, Schmidt et al. 2014). The presence of organic matter from pyrogenic origins is one of the main factors responsible for the advantageous physical characteristics, structure, and fertility of these soils (Benites et al. 2009). The use of biochar to mimic the TPI has shown satisfactory results in terms of the retention and availability of nutrients in the soil, since these materials have large surface areas and high ion exchange capacities, and also provide an additional source of nutrients (Glaser et al. 2002, Petter and Madari 2012, Wu et al. 2017, Zhao et al. 2017).

The feasibility of using biochar as a soil conditioner is of considerable interest to Brazilian agroindustry, which generates a large quantity of organic waste that often has no suitable destination. The sugar and alcohol industry is especially
notable in this respect, since it produces around 200 million tons of organic waste annually. These wastes are often burned in furnaces at the industrial installations, in order to avoid transportation costs (IPEA 2012). The use of the pyrolyzed solid waste in sugarcane crop production would make the production process more sustainable, as well as help to reduce environmental contamination by pesticides. The use of pesticides in the production of this crop accounts for approximately 7.5% of the 914,200 tons of these substances consumed in Brazilian plantations (2014), with the great majority being herbicides (Scorza Júnior 2009, SINDIVEG 2015). This makes sugarcane cultivation the second largest consumer of these agricultural inputs (SINDIVEG 2015).

Clomazone (CMZ, Table I), a selective herbicide that belongs to the isoxazolidinone chemical group, is one of the active ingredients applied for weed control in sugarcane crops. The compound is absorbed by the roots and is transported through the plants in the xylem, and shows higher efficiency when applied pre-emergence (MAPA 2013). Its activity is related to the inhibition of carotenoid production, which leads to the loss of chlorophyll, since in the absence of carotenoids, chlorophyll is more readily degraded by photooxidation processes (Silva et al. 2007).

The high solubility of CMZ in water (1102 mg L⁻¹ at 20 °C) can lead to potential negative impacts on groundwater. Its half-life has been found to vary between 79 and 124 days, depending on the composition of the soil, so its moderate persistence can affect successive crops in the same area. In addition, CMZ is a non-dissociable compound that presents high volatility (19.2 mPa at 25 °C) (IUPAC 2007, Van Scoy and Tjeerdema 2014).

The addition of biomass charcoal to soil alters the sorptive capacity, leading to greater retention of pesticides and potentially reducing environmental contamination. Conditioning of soil with bamboo biochar was found to result in greater sorption of the herbicide pentachlorophenol in the soil, hence reducing the contamination of soil and groundwater through leaching of the herbicide (Xu et al. 2012). The incorporation of wheat straw biochar in soil increased hexachlorobenzene sorption 42-fold, compared to soil without addition of biochar, and reduced dispersion of the herbicide (Song et al. 2012).

Although it is known that biochar can assist in mitigating pesticide contamination of soil, greater understanding is required of its properties in terms of sorptive capacity and the mechanisms of sorption of chemicals in the soil. The objective of this study was to evaluate the influence of the addition of sugarcane biomass biochar, in different percentages and particle sizes, on the retention of CMZ in soil. Investigation was made of the behavior of CMZ sorption, as well as in the changes of physicochemical characteristics of the soil amended with biochar.

**EXPERIMENTAL**

**CHEMICALS**

The chemicals used were clomazone (98.1% w/w) and HPLC grade acetonitrile (Sigma-Aldrich, Seelze, Germany), dihydrated calcium chloride and sodium azide (Vetec, Rio de Janeiro, Brazil), and deionized water purified using a Millipore system.

A stock 1000 mg L⁻¹ standard solution of CMZ was prepared by dilution in acetonitrile. The working solutions used in the sorption tests were prepared by diluting the stock solution with a previously prepared solution containing CaCl₂ (1.11 mg L⁻¹, employed to simulate the ionic strength of soil solutions) and NaN₃ (250 mg L⁻¹, used to inhibit microbial activity).

**SOIL COLLECTION AND BIOCHAR PRODUCTION**

A Red Latosol with low organic matter content was collected at 0-20 cm depth, in an area without recent history of herbicide application, in the municipality...
of Tangará da Serra in Mato Grosso State (latitude 14° 39’ 4.01” S, longitude 57° 26’ 0.64” W). The soil was air dried and sieved through a 2 mm mesh, and was subsequently characterized (EMBRAPA 1997). The physicochemical characteristics of the soil are presented in Table II.

The sugarcane biomass (with 60% moisture content) was dried for 48 h at 60 °C in a forced air circulation drying oven. The dried material was chopped in a disintegrator and then pyrolyzed at 400 °C for 6 h under a CO₂ atmosphere in a laboratory muffle furnace. The resulting biochar was milled, dried at 60 °C for 12 h, and sieved through different meshes, yielding three fractions with particle sizes of 610-508, 508-106, and <106 µm, which were used to investigate the influence of particle size on the CMZ sorption processes in the soil. The biochars were mixed with soil and homogenized to obtain amended soils with biochar in different particle sizes and percentage. The physicochemical characteristics of the soil amended using 1% (w/w) biochar with particle size <106 µm are shown in Table II.

HPLC ANALYSES

The chromatographic analyses were performed on an HPLC system (Shimadzu, Tokyo, Japan) equipped with an LC-20AT pump, a SIL-10AF autosampler, a CTO-10ASVP oven, and an SPD-20A UV/Vis detector set at 210 nm.

The chromatographic separation was performed using a Shimadzu C18 column (VP-ODS, 150 x 4.6 mm, 4.6 ± 0.3 µm) maintained at 30 °C. Isocratic elution was performed with a mobile phase composed of acetonitrile and deionized water (40:60, v/v), at a flow rate of 1.2 mL min⁻¹. The injected sample volume was 20 µL and the retention time was 12.4 min. Quantification of CMZ was performed by the external standards method. The Limit of detection (LOD) and quantification (LOQ) were determined using blank soil solution (solution containing 1.11 mg L⁻¹ CaCl₂ and 250 mg L⁻¹ NaN₃ stirred with a soil sample for 24 h). They were estimated based on a signal-to-noise ratio (S/N) of 3 and 10 for LOD and LOQ, respectively. The accuracy and precision were evaluated using blank solutions fortified with 0.25, 5.0 and 15 mg L⁻¹ (in triplicate) through a recovery test and the relative standard deviation (RSD), respectively, as suggested by ANVISA (2017).

Effect of particle size and biochar percentage on sorption/desorption of CMZ in the soil

A 2² factorial design with 3 replications at the central point was employed to investigate the combined effects of biochar particle size and biochar/soil percentage (% w/w) on the retention and behavior of CMZ in the soil. Table III shows the particle sizes and biochar/soil percentages evaluated, together with the coded values. These percentages are equivalent to 1.25, 2.5 and 5 tons biochar ha⁻¹, considering the application of biochar in the first 5 cm-layer of soil.

Sorption tests were used to obtain the apparent sorption coefficient ($K_{f,app}$), adopted as the response in the factorial design.

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**TABLE I**

| Clomazone | Physicochemical properties |
|-----------|----------------------------|
| Molecular formula: | C₁₂H₁₄ClNO₂ |
| Molar mass: | 239.7 g mol⁻¹ |
| Vapour pressure at 25 °C: | 1.44×10⁻⁴ mmHg |
| $K_w$: | 347 |
| Solubility in water at 20° C: | 1102 mg L⁻¹ |
| Non-ionizable compound |
CLOMAZONE SORPTION/DESORPTION TESTS IN SOIL AMENDED WITH BIOCHAR

Sorption equilibrium time

The assays were carried out based on Procedure 106 (Batch Equilibrium) of the Organization for Economic Co-operation and Development (OECD 2000).

Firstly, it was necessary to determine the time required for CMZ to reach sorption equilibrium in the soil. This was achieved using soil samples amended with 1% (w/w) of biochar with particle size <106 µm, and in the absence of biochar. The soil/solution ratio was 1:5 (w/v), and CMZ solutions were prepared in the presence of calcium chloride (1.11 mg L⁻¹) and sodium azide (250 mg L⁻¹) to inhibit microbiological activity.

The time required for equilibrium sorption of CMZ was determined by suspending 2.00 g portions of soil in 10.0 mL volumes of a solution containing CaCl₂ (1.11 mg L⁻¹), NaN₃ (250 mg L⁻¹), and CMZ (10.0 mg L⁻¹), in glass centrifuge tubes. The suspensions were maintained under horizontal stirring (180 rpm) for periods of up to 32 h, at constant temperature (21 °C), followed by centrifugation at 1300 x g for 5 min. Aliquots of the supernatants were filtered using syringe filters containing 0.45 µm PTFE membranes, and were then analyzed using HPLC.

The concentrations of CMZ in the soil (\( C_s^{\text{Sor}} \), mg kg⁻¹) after different stirring times were calculated from the difference between the initial (\( C_0 \)) and final (\( C_\text{aq} \)) concentrations in the aqueous solution (mg L⁻¹):

\[
C_s^{\text{Sor}} = \frac{V_0}{m_{\text{soil}}} \left[ C_0 - C_\text{aq}^{\text{Sor}} \right]
\]  

where \( m_{\text{soil}} \) (kg) is the mass of soil (or soil amended with biochar) and \( V_0 \) (L) is the volume of CMZ solution added to the soil.

The data were analyzed using one-way analysis of variance (ANOVA) and the post-hoc Tukey test, adopting a 95% confidence level. The \( C_s^{\text{Sor}} \) values were plotted against time (h) and the equilibrium time was defined as the point at which the sorption values reached a plateau (OECD 2000).

The pseudo-first order, pseudo-second order, and Elovich models were applied in order to

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**TABLE II**

Physicochemical characteristics of soil (S) and soil amended by 1% of biochar in particle size <106 µm (S + B).

| Soil   | S          | S + B       | B (estimated) |
|--------|------------|-------------|---------------|
| pH (in H₂O) | 5.59       | 6.00        |               |
| P (mg kg⁻¹) | 0.1        | 0.9         | 80.1          |
| K⁺ (mg kg⁻¹) | 11         | 116         | 10511         |
| Mg²⁺ (mmol kg⁻¹) | 0.9        | 1.7         | 80.9          |
| CEC⁺ (mmol kg⁻¹) | 52.0       | 55.6        | 412           |
| OM (%) | 2.37       | 2.50        |               |
| Texture (%) |            |             |               |
| Sand (> 0.053 mm) | 66.5       | 66.1        |               |
| Silt (0.053-0.002 mm) | 10.3       | 9.0         |               |
| Clay (<0.002 mm) | 23.2       | 22.9        |               |

a CEC – Cationic Exchange Capacity at pH 7.
b OM – Organic Matter.

Analyses carried out in the Laboratório de Análise de Solo, Tecido Vegetal e Fertilizante from Universidade Federal de Viçosa, according to the methodology of Empresa Brasileira de Pesquisa Agropecuária –EMBRAPA.
evaluate the effect of biochar addition on the kinetics of CMZ sorption in the soil. The pseudo-first order model is given by:

\[ q_t = q_e \left(1 - e^{-k_1 t}\right) \]  \hspace{1cm} (2)

where \( q_e \) (mg kg\(^{-1}\)) and \( q_t \) (mg kg\(^{-1}\)) are the amounts absorbed at equilibrium and at a given time \( t \), respectively, and \( k_1 \) (h\(^{-1}\)) is the rate constant (Janos et al. 2007, Plazinski et al. 2009, Qiu et al. 2009).

The pseudo-second order model is described by the following linear equation (Janos et al. 2007):

\[ \frac{t}{q_t} = \frac{1}{k_2q_e^2} + \frac{t}{q_e} \]  \hspace{1cm} (3)

where the rate constant \( (k_2, \text{ mg kg}\(^{-1}\) h\(^{-1}\)) \) and \( q_e \) (mg kg\(^{-1}\)) are obtained from the slope and intercept, respectively, of a plot of \( t/q_t \) against \( t \) (h) (Peruchi et al. 2015, Plazinski et al. 2009, Qiu et al. 2009).

The Elovich model (Eq. 4), which was originally proposed for describing the adsorption of gases in solid systems, can also be used to describe the removal of pollutants from aqueous solutions (Qiu et al. 2009). Use of this model is restricted to the beginning of the sorption process, since it neglects the simultaneous occurrence of desorption, when the sorption process is at equilibrium (Plazinski et al. 2009). In this model, the intercept \( (1/Y) \ln(\chi Y) \), mg kg\(^{-1}\) represents the amount sorbed during the initial stage of the sorption process, corresponding to the fast adsorption step, due to movement of the adsorbate to the most accessible part of the sorbent, while the slope \( (1/Y) \) (mg kg\(^{-1}\) h\(^{-1}\)) is related to the second slow sorption stage and to the duration of this process \((\ln t)\) (Fernández-Bayo et al. 2008, Peruchi et al. 2015).

\[ q_t = \frac{1}{Y} \ln(\chi Y) + \frac{1}{Y} \ln t \]  \hspace{1cm} (4)

**Sorption assays**

The sorption tests used to investigate the effects of particle size and biochar percentage on the retention and behavior of CMZ in the soil were also based on the Batch Equilibrium procedure (OECD 2000).

In the experiments based on the factorial design, the apparent sorption coefficients \( (K_{app}) \) were determined using a 10.0 mg L\(^{-1}\) CMZ solution containing CaCl\(_2\) (1.11 mg L\(^{-1}\)) and NaNO\(_3\) (250 mg L\(^{-1}\)). Volumes of 10.0 mL of this solution were added to glass centrifuge tubes containing 2.00 g of the soil samples (as indicated in the factorial design), and the suspensions were stirred for a period corresponding to the previously established equilibrium time. After this period, the tubes were centrifuged and the supernatants were filtered and analyzed by HPLC. The same procedure was performed (in triplicate) using the soil without
biochar, for use as a reference in comparison of the results. The apparent sorption coefficients were calculated using Eq. 5:

$$K_d = \frac{C_s}{C_{aq}}$$  \tag{5}

where $C_s$ (mg$_{CMZ}$ kg$_{soil}^{-1}$) is the concentration of the sorbed compound and $C_{aq}$ (mg$_{CMZ}$ L$_{aqueous solution}^{-1}$) is the concentration of herbicide present in solution, at equilibrium (MacKay and Vasudevan 2012). The data analysis was performed with Statistica v. 8 software (StatSoft, Tulsa, USA).

Sorption isotherms were obtained for the factorial design experiments and also for soil without biochar. Five different CMZ concentrations were added (10, 12.5, 15, 17.5, and 20 mg L$^{-1}$) in 2.00 g portions of the soil samples described in the factorial design, as well as made to determine the values.

The results were fitted using the Freundlich model (Eq. 6), which describes the relation between the amount of the compound present in solution ($C_{aq}$, in mg L$^{-1}$) and the amount sorbed in the soil ($C_s$, in mg kg$^{-1}$), at equilibrium.

$$C_{aq} = K_{f}^{orr} C_{aq}^{orr}(eq)^{\frac{1}{n}} $$ \tag{6}

The Freundlich constant ($K_{f}^{orr}$, in mg$^{-1/n}$ L$^{1/n}$ kg$^{-1}$) is a parameter that characterizes the soil sorption capacity for a given compound (Wu et al. 2011).

Desorption assays

The desorption tests were performed as a continuation of the sorption assays. In this procedure, all the remaining supernatant in the samples from the sorption test was removed. Subsequently, the volume of solution removed was replaced by an equal volume of solution containing CaCl$_2$ and NaN$_3$ (without CMZ). The samples were then subjected to the same stirring, centrifugation, and filtration procedure used in the sorption assays, and aliquots of the final supernatants were analyzed by HPLC. The data obtained were used to determine $K_{d}^{des}$ and $K_{f}^{des}$, provided by the Freundlich desorption isotherms.

Estimation of biochar external surface area

The external surface areas of environmental solid particles can be estimated based on their geometric shape, density ($\rho$, g cm$^{-3}$), and diameter (d, μm). Assuming spherical shape of the biochar particles, their external areas ($A_{ext}$, m$^2$) could be calculated according to Eq. 7 (Papelis et al. 2003, Schmukat et al. 2012). The total area of the biochar ($A_{tot, biochar}$, m$^2$), could then be obtained by multiplying $A_{ext}$ by the mass of the adsorbent ($m_{biochar}$, g) employed in each experiment of the factorial design, as shown in Eq. 8.

$$A_{ext} = \frac{6}{d \times \rho} $$ \tag{7}

$$A_{tot, biochar} = \frac{6 \times m_{biochar}}{d \times \rho} $$ \tag{8}

The value of $m_{biochar}$ was obtained by multiplying the percentage of biochar in the amended soil (% biochar, % w/w) by its mass ($m_{soil+biochar}$, g), as shown in Eq. 9.

$$m_{biochar} = \frac{\% biochar}{100} \times m_{soil+biochar} $$ \tag{9}

Substituting $m_{biochar}$ in Eq. 8 resulted in Eq. 10. Since the biochars used in the experiments had the same density (constant $\rho$), given that they originated from a single sample, $A_{tot, biochar}$ was dependent on the ratio between % biochar and the particle diameter d, as shown in Eq. 11.

$$A_{tot, biochar} = \frac{6 \times m_{biochar}}{d \times \rho} \frac{6 \times \% biochar \times m_{soil+biochar} \times 100}{d \times \rho} $$ \tag{10}

As $A_{tot, biochar}$ was proportional to the ratio % biochar/d (μm$^{-1}$), determination was made of the correlations between % biochar/d and $K_{f}^{orr}$ and
The sorption of Clomazone in soil amended with biochar was investigated by comparing the retention time of the analyte in a fortified extract with the retention time of a standard solution of the herbicide prepared in acetonitrile. The selectivity was confirmed by comparing the chromatograms for an extract obtained by stirring soil (amended using 1% biochar with particle size <106 µm) with a 20 mg L⁻¹ CMZ solution and an extract obtained using the same procedure, but without CMZ (Figure 1).

CMZ was quantified by the external standards method. The LOD and LOQ were found to be 0.006 and 0.019 mg L⁻¹. The analytical curve was obtained by analyzing solutions containing the herbicide at concentrations between 0.025 and 20 mg L⁻¹, together with 1.11 mg L⁻¹ CaCl₂ and 250 mg L⁻¹ NaN₃. The curve showed a correlation coefficient (r) of 0.9998, indicative of low dispersion of the experimental data and low uncertainty of the regression equation, confirming that the analytical technique enabled reliable quantification of CMZ (ANVISA 2003). The precision of the method (expressed as the RSD) was 1.6%, 5.2% and 0.5%, and the accuracy (expressed as the average recoveries) was 111%, 103% and 99% at concentration levels 0.25, 5.0 and 15 mg L⁻¹, respectively.

EQUILIBRIUM TIME

The soils with addition of biochar (at 1%, using the biochar with particle size <106 µm) and without biochar addition were stirred with a 10 mg L⁻¹ CMZ solution for different periods up to 32 h (Figure 2). It was found that after 2 h of stirring, the concentrations of CMZ in the soil showed no significant differences (Tukey post-hoc test, 95% confidence level). For both soils, slight increases in sorption were observed until the equilibrium plateau was reached after 8 h of stirring (apparent equilibrium).

The equilibrium time was shorter than reported by Li et al. (2004) for four Chinese soils originating from rice crop fields, where it was found that the CMZ sorption processes reached equilibrium after 24 h. The results obtained for the kinetics of CMZ sorption (Table IV) showed that the pseudo-second order kinetic model presented the best fit to the experimental data, based on the coefficient of determination (R² > 0.99). This indicated that the sorption kinetics was entirely controlled by sorption/desorption processes and that it was governed by the availability of sorption sites on the soil (or the soil amended with biochar) surface, rather than by the concentration of CMZ in the solution (Liu 2008, Plazinski et al. 2009). The qₑ values provided by the kinetic model revealed a notable
increase of the soil sorption capacity following the addition of biochar. According to Wang and Xing (2007), pyrolysis results in changes in the surface chemical groups, as well as the formation of new micropores in the biomass. Consequently, the addition of biochar to soils results in increased sorption capacity, due to its high surface area, high porosity, and the presence of aromatic components in its matrix.

EFFECTS OF BIOCHAR PARTICLE SIZE AND PERCENTAGE ON CMZ SORPTION IN SOIL

The effects of biochar particle size and biochar/soil percentage on CMZ sorption in soil were evaluated using a 2² factorial design with a central point. The results, in terms of the apparent sorption coefficient \( K_{d}^{\text{app}} \), are shown in Table V.

The effects of the factors on the CMZ sorption on soil (Figure 3) showed that both particle size (negative effect) and percentage (positive effect) of the biochar were significant (at the 95% confidence level). In other words, a smaller particle size and higher biochar percentage helped increase sorption of the herbicide in the soil. The curvature and the interaction between the factors were also significant.

The biochar particle size and percentage used in experiment E3 provided the highest sorption of CMZ in soil, based on the \( K_{d}^{\text{app}} \) values shown in Table V (\( K_{d}^{\text{app}} = 616 \)). It could therefore be concluded that the conditions that favored greater sorption, in the range of particle sizes and percentages studied, were the highest percentage (1%) and the smallest particle size (<106 μm).

In the absence of biochar (\( K_{d}^{\text{app}} = 0.47 \)), the partitioning of CMZ between the aqueous solution and the soil was lower than in all the experiments with biochar addition (Table V). Comparison of the sorption capacities of the unmodified (without addition of biochar) and amended soils showed that addition of the pyrogenic material increased the \( K_{d}^{\text{app}} \) value by up to 1300-fold (experiment E3). The relative contributions of the biochar to the increase sorption were 95.3, 70.6, 99.9, 88.3, and 88.9% for experiments E1, E2, E3, E4, and E5, respectively, showing that the biochar provided more effective sorption, compared to the other soil fractions. Similar results were obtained by Yu et al. (2011), who showed that the addition of 0.5% eucalyptus biochar increased the sorption of acetamiprid insecticide in three different soils, and that the relative contribution of biochar to the
Sorption varied according to the quantity of organic matter present in the soil.

In the work of Zheng et al. (2010), it was shown that smaller particle sizes of biochar resulted in lower availability of the herbicide atrazine in aqueous solutions, as well as shorter times to reach equilibrium sorption of the herbicide by the biochar. Similar features were observed here for CMZ (Table V), with greater sorption of the herbicide by the smaller particle size biochar, and enhancement of the effect with increasing percentage of biochar in the soil.

The results of the desorption assays showed that the soils modified with biochars presented lower extractions (using aqueous CaCl$_2$ and NaN$_3$ solution) of CMZ initially retained in the soil, compared to the unmodified soil, which showed complete desorption of CMZ by the aqueous solution. It was observed that the particle size factor also contributed to lower extraction. In experiments E2, E4, and E5, with larger particle size biochars, the herbicide concentrations in the soils were 4.0, 15.0, and 12.9 times higher (based on $K^{d}_{des}$ values), respectively, than the concentrations of herbicide

### Table IV

| Soil      | Pseudo-first order | Pseudo-second order | Elovich |
|-----------|--------------------|---------------------|---------|
|           | $q_e$ (mg kg$^{-1}$) $k_1$ (h$^{-1}$) $R^2$ | $q_e$ (mg kg$^{-1}$) $k_2$ (kg mg$^{-1}$ h$^{-1}$) $R^2$ | $1/Y \ln(xy)$ (mg kg$^{-1}$) $1/Y$ (kg mg$^{-1}$ h$^{-1}$) $R^2$ |
| S         | 7.611              | 2.091               | 0.450   | 8.693                  | 0.163               | 0.998               | 5.675               | 0.883               | 0.941               |
| S + B     | 47.992             | 15.434              | 0.259   | 48.333                  | 2.540               | 0.999               | 47.488              | 0.312               | 0.802               |

### Table V

| Experiments | Particle Size (1) | Percentage (2) | $K^{sor}_d$ (L kg$^{-1}$) | $K^{des}_d$ (L kg$^{-1}$) | Percentage Desorbed (l%) |
|-------------|-------------------|----------------|---------------------------|---------------------------|--------------------------|
| E1          | < 106 µm          | 0.25           | 10.1                      | 31.5                      | 13.7                     |
| E2          | 610-508 µm        | 0.25           | 1.6                       | 4.0                       | 55.4                     |
| E3          | < 106 µm          | 1              | 616.3                     | *                         | *                        |
| E4          | 610-508 µm        | 1              | 4.0                       | 15.0                      | 25.0                     |
| E5          | 508-106 µm        | 0.5            | 4.22±0.24                 | 12.9±1.2                  | 28.0±1.9                 |
| SOIL        |                   |                | 0.47±0.02                 | **                        | 100**                    |

E1 - Soil amended with 0.25% of biochar in particle size < 106 µm.
E2 - Soil amended with 0.25% of biochar in particle size between 508-610 µm.
E3 - Soil amended with 1% of biochar in particle size < 106 µm.
E4 - Soil amended with 1% of biochar in particle size between 508-610 µm.
E5 - Soil amended with 0.5% of biochar in particle size between 106-508 µm.
SOIL - Soil without addition of biochar.

$K^{sor}_d$ – Apparent sorption coefficient, after contact of the soil (amended or without biochar) sample, with a CMZ solution (10 mg L$^{-1}$) and under stirring for 8 h.

$K^{des}_d$ – Apparent desorption coefficient, after contact of the soil samples (obtained at the end the sorption assay) with a CaCl$_2$ and NaN$_3$ solution (free of CMZ) and under stirring for 8 h.

* In this experiment it was not possible to quantify the CMZ in aqueous solution due to its concentration was below the analytical concentration range assessed in this work.

** In this experiment all CMZ was desorbed.
in the aqueous solutions. In experiment E1, using biochar with smaller granulometry, the retention was even higher and the CMZ concentration in the solid phase was 31.5 times greater than in solution, confirming the important contribution of surface area in determining the sorption capacity of the biochar.

An increase in the biochar percentage also led to a greater amount of herbicide retained in the soil, as shown by comparison of the \( K_{\text{des}} \) values obtained in experiments E2 and E4. The same effect could be seen for comparison of experiments E1 and E3, although the amount of herbicide extracted by the aqueous solution in experiment E3 was so small that its quantification was not possible. This could be explained by the high relative contribution of the biochar to CMZ sorption in this experiment, providing further evidence of the strength of the interaction between the herbicide and the biochar.

**SORPTION AND DESORPTION ISOTHERMS**

In order to improve understanding of the CMZ sorption mechanisms, sorption and desorption isotherms were obtained for the factorial design experiments and the assay using unmodified soil. The CMZ concentration range employed in the isotherms was between 10 and 20 mg L\(^{-1}\). This high concentration range was selected because at lower CMZ concentrations (in preliminary tests), conditioning of the soil using the smaller particle size biochar resulted in sorption of all the CMZ present in the aqueous solution, making its quantification by HPLC impossible.

The sorption and desorption isotherms obtained are shown in Figure 4, together with the curve obtained using the Freundlich equation to fit the data. The Freundlich equation provided statistically significant explanations of the sorption and desorption processes in all the experiments (from the factorial design and using unmodified soil), with p-values <0.01. The parameters obtained for each model are shown in Table VI.

The sorption isotherms obtained for the SOIL (unmodified soil) and E2 (soil amended using 0.25% of biochar with particle size of 508-610 µm) experiments presented the highest 1/n values, which were close to unity, so they were classified as type C. This type of isotherm indicated a constant affinity between the CMZ molecules and the sorption sites present in the adsorbent (Giles et al. 1974, McBride 1994).

A decrease in biochar particle size and/or an increase in the biochar percentage led to a decrease of the 1/n parameter and alteration of the sorption behavior. In experiments E1 (soil amended using 0.25% of biochar with particle size <106 µm), E4 (soil amended using 1% of biochar with particle size 508-610 µm), and E5 (soil amended using 0.5% of biochar with particle size 106-508 µm), the curves obtained could be classified as type L. These isotherms suggested high affinity (early during the sorption process) between the herbicide and the soil amended with biochar. The sorption isotherm for experiment E3 (obtained for soil amended using 1% of biochar with particle size <106 µm) represented an extreme case of this classification, with the lowest 1/n value, and could be considered a type H isotherm. This isotherm type was indicative of stronger interactions between the CMZ molecules and the soil amended with biochar (McBride 1994).

The sorption behavior of CMZ in the soil was altered following the addition of biochar, and was also influenced by the characteristics of the biochar (particle size and percentage). This was demonstrated by the changes in the 1/n parameter (Table VI) and the type of isotherm when the biochar particle size was decreased and the biochar percentage was increased.

According to MacKay and Vasudevan (2012), another way of predicting the sorption behavior of polyfunctional organic compounds in environmental solids is by evaluating the types of
SORPTION OF CLOMAZONE IN SOIL AMENDED WITH BIOCHAR

Figure 4 - Isotherms obtained in the CMZ sorption and desorption assays in a Red Latosol amended with different particle sizes and percentages of biochar (E1: soil amended with 0.25% of biochar in particle size < 106 µm; E2: soil amended with 0.25% of biochar in particle size between 508-610 µm; E3: soil amended with 1% of biochar in particle size < 106 µm; E4: soil amended with 1% of biochar in particle size between 508-610 µm and E5: soil amended with 0.5% of biochar in particle size between 106-508 µm) and without addition of the sorbent (SOIL).

TABLE VI
Parameters of Freundlich isotherms (Kf and 1/n) and its coefficients of determination.

| Experiment | Phenomenon studied | Freundlich |
|------------|--------------------|------------|
|            | Kf (µg L⁻¹ (mg L⁻¹)⁻¹ (g⁻¹) | 1/n | R² | p-value |
| E1         | Sor*               | 23.0219    | 0.2806 | 0.8934 | 1.7238x10⁻⁶ |
|            | Des**              | 28.2427    | 0.1893 | 0.5530 | 4.9229x10⁻⁶ |
| E2         | Sor*               | 2.4292     | 0.7934 | 0.7575 | 1.0115x10⁻⁴ |
|            | Des**              | 4.8916     | 0.4319 | -0.1497 | 0.00656 |
| E3         | Sor*               | 83.4806    | 0.2126 | 0.9606 | 2.3583x10⁻⁶ |
|            | Des**              | 112.8474   | 0.1618 | 0.9635 | 1.3409x10⁻⁵ |
| E4         | Sor*               | 8.4023     | 0.5900 | 0.9733 | 4.9290x10⁻⁷ |
|            | Des**              | 16.7875    | 0.5753 | 0.8587 | 8.1121x10⁻⁶ |
| E5         | Sor*               | 13.6423    | 0.3216 | 0.7547 | 1.1102x10⁻⁶ |
|            | Des**              | 17.1957    | 0.0583 | -0.0472 | 1.2845x10⁻¹³ |
| SOIL       | Sor*               | 0.7657     | 0.7302 | 0.7881 | 2.0872x10⁻¹⁴ |

*Sorption and **Desorption.
interactions between these compounds and the sites present on the surfaces of solids. These interactions were classified as type I (hydrophobic partitioning and electron donor-acceptor interactions) and type II (interactions involving ion exchange, surface complexation, and cation bridges).

CMZ is a non-ionic polyfunctional compound whose structure includes regions with high electron density, such as the aromatic part and the isoxazolidinone group. These enable electron donor interactions with neutral species (with low electron density) such as metal oxides and surface aluminosilicate groups. Due to the presence of nitrogen and oxygen atoms in the isoxazolidinone group, hydrogen bonds can also be formed with the hydroxyl groups present in organic matter, and complexes can be formed with surface aluminum and iron atoms (MacKay and Vasudevan 2012).

Like graphite, the structure of biochars includes disorderly stacked polyaromatic sheets, which form highly sorptive aromatic π-systems. These aromatic structures have the ability to perform π-donor and π-acceptor interactions, depending on the substituent groups attached to aromatic rings of compounds to be sorbed. In the case of sorption of π-acceptor compounds (which have deactivating groups on the aromatic ring), both hydrophobic van der Waals interactions and hydrogen bonding are possible mechanisms that can provide strong sorption. In the case of π-donor compounds (which have activating groups present on the aromatic ring), such as CMZ, van der Waals and π-π interactions are possible mechanisms of adsorption, with their intensities increasing according to the number of aromatic rings present and the planarity of the molecule (Keiluweit and Kleber 2009). These features suggest that the sorption of CMZ in the soil (and in soil amended with biochar) followed a type I mechanism governed by hydrophobic partitioning mechanisms and electron donor-acceptor interactions (MacKay and Vasudevan 2012).

**CORRELATION BETWEEN \( A_{\text{tot.biochar}} \) AND PARAMETERS \( K_f \) AND \( 1/n \)**

The Freundlich isotherm \( K_f \) and \( 1/n \) parameters obtained in the sorption and desorption assays were correlated with the % biochar/d ratio, which is proportional to the total external surface area of the biochar \( (A_{\text{tot.biochar}}) \). The results of these correlations and the parameters obtained in the linear regressions are shown in Figure 5 and Table VII, respectively.

The values of \( K_f^{\text{Sor}} \) and \( K_f^{\text{Des}} \) were highly correlated with the % biochar/d ratio \((R > 0.99, p < 0.0001)\), showing that the processes of sorption and desorption of CMZ on the amended soil were directly influenced by the total external surface area of the biochar present in it. An increase in biochar surface area implies an increase in the number of sorption sites that enable hydrophobic partitioning, electron donor-acceptor interactions, and hydrogen bonding (Wang et al. 2016), providing higher sorption (higher \( K_f^{\text{Sor}} \)) and lower desorption (higher \( K_f^{\text{Des}} \)) of CMZ in soil. The \( 1/n_{\text{ads}} \) and \( 1/n_{\text{des}} \) parameters showed poor correlations with the % biochar/d ratio \((R < 0.62, p > 0.05)\), although there was a downward trend of these parameters with increasing % biochar/d ratio. This inverse behavior could be attributed to increased aromatic domains in the soil, due to a greater total surface area of the biochar, contributing to nonlinearity of the isotherms and change in the sorption mechanisms (Wang et al. 2016, Zhu et al. 2004).

**EFFECT OF BIOCHAR ADDITION ON SOIL PHYSICOCHEMICAL PROPERTIES**

Assessment of changes in the properties of the soil was carried out by comparing the physicochemical characteristics of unmodified soil and the soil amended with biochar that provided the greatest CMZ sorption, determined in the factorial design experiments (Table II).

The results showed that the addition of sugarcane biochar to soil increased the
Figure 5 - Evaluation of the influence of the total external surface area of biochar on the sorption and desorption of CMZ by the correlation of ratio % biochar/d with the parameters of the Freundlich equation $K_f^{\text{sor}}$ (a), $K_f^{\text{des}}$ (b), $1/n_{\text{sor}}$ (c) and $1/n_{\text{des}}$ (d).

Table VII
Parameter values obtained from the correlation between the ratio % biochar/d and parameters of Freundlich equation $K_f^{\text{sor}}$, $K_f^{\text{des}}$, $1/n_{\text{sor}}$ and $1/n_{\text{des}}$.

| Correlation | R       |
|-------------|---------|
| % Biochar/d vs $K_f^{\text{sor}}$ | 8976.40 (% Biochar/d) - 1.46731 | 0.9930 |
| % Biochar/d vs $K_f^{\text{des}}$ | 12171.80 (% Biochar/d) - 2.1224 | 0.9985 |
| % Biochar/d vs $1/n_{\text{sor}}$ | -50.87748 (% Biochar/d) + 0.62083 | 0.6132 |
| % Biochar/d vs $1/n_{\text{des}}$ | -22.2132 (% Biochar/d) + 0.35288 | 0.3745 |

concentrations of P, K, and Mg in the soil by 0.8 mg kg$^{-1}$, 105 mg kg$^{-1}$, and 0.8 mmol kg$^{-1}$, respectively. These results indicate that the material has potential to act as a fertilizer. In addition, biochar improved the values for CEC, OM, and pH, which are important factors affecting the sorption processes of nutrients and organic compounds in soil. These findings were in agreement with the review provided by Glaser et al. (2002), who reported that the addition of biochar to soil had the combined effects of increasing the organic matter content, improving the retention and availability of nutrients, increasing the cation exchange capacity (CEC), and altering the pH.
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