Nomogram for Prediction of Bronchial Mucus Plugs in Children with *Mycoplasma pneumoniae* Pneumonia

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The presence of bronchial mucus plugs (BMP) in children with *Mycoplasma pneumoniae pneumonia* (MPP) results in delayed clinical and radiographic resolution and long-standing pulmonary sequelae. The predictive factors associated with BMP formation remains poorly defined. Nomograms to predict BMP presence in children with MPP were proposed using a cohort of patients who underwent bronchoscopy intervention at Children's Hospital in Eastern China. Patients with MPP in an earlier period formed the training cohort (n = 872) for nomogram development, and those thereafter formed the validation cohort (n = 399) to confirmed model’s performance. BMP in children with MPP were found in 196 (22.5%) and 91 (22.8%) patients in the training and validation cohorts, respectively. The independent risk factors associated with BMP were age > 5 years (OR 2.06; 95% CI 1.43 to 2.98), higher IL-10 level (> 10 ng/L, 2.19; 95% CI 1.46 to 3.28), higher IFN-γ level (> 30 ng/L, 1.69; 95% CI 1.13 to 2.54), and presence of complication (3.43; 95% CI 1.45 to 8.09). Incorporating these 4 factors, the nomogram achieved good concordance indexes of 0.771 (95% CI, 0.734–0.808) and 0.796 (95% CI, 0.744–0.848) in predicting BMP in the training and validation cohorts, respectively. The nomogram achieved an optimal prediction of BMP in children with MPP. Using this model, the risk of BMP formation would be determined, contributing to a rational therapeutic choice.
marker for early prediction of refractory MPP\textsuperscript{18}. Additionally, there is evidence that increased serum concentrations of tumor necrosis factor (TNF) alpha, interferon (IFN) gamma, and interleukin 18 (IL-18) were closely associated with refractory MPP\textsuperscript{19–21}. Although atelectasis is a stronger indicator for bronchoscopy intervention, many MPP children with BMP had no significant atelectasis signs in chest imaging. Thus it is especially essential to predict BMP presence in children with MPP using some key clinical variables. Therefore, the aim of the present study was to define clinical factors associated with BMP formation in MPP children using a cohort of patients. In particular, we sought to create an internally validated nomogram to predict the individual risk of BMP in MPP children.

**Methods**

**Patients and data collection.** A retrospective study was conducted on a primary cohort of patients who had MPP and underwent flexible bronchoscopy between January 2016 and December 2018 at the respiratory department of Children's hospital, Zhejiang University School of Medicine (Hangzhou, China). Diagnosis of MPP was based on diagnosis of CAP and etiology using our previous study\textsuperscript{22}. Those patients with tuberculosis or HIV positive were excluded from the study, and those with chronic lung diseases, congenital heart disease, cerebral palsy or tumor were also excluded. Eligible consecutive patients with MPP between January 2016 and December 2017 were included into the training cohort for development of the nomogram to predict risk of BMP, and those between January and December 2018 were enter into the validation cohort.

Demographic and clinical data were collected, including imaging examination (suggestive of pneumonia or pulmonary consolidation, atelectasis and pleural effusions), ultrasonography (suggestive of pleural effusions), and bronchoscopy results. All laboratory tests for collection were performed before bronchoscopy. Based on our
previous study, the cutoff values of IL-10 and IFN-γ were determined as 10 ng/L and 30 ng/L, respectively22. The definition of complications referred to the presence of pulmonary atelectasis and/or pleural effusion. The primary outcomes of interest were risk of BMP formation in children with MPP.

**Ethical considerations.** This study was approved by the Ethic Review Board of Children's Hospital, Zhejiang University School of Medicine. Written informed consents were obtained from a parent and/or legal guardian of each participant and all research was performed in accordance with the relevant guidelines and regulations.

### Table 1. Clinical characteristics of patients in the training and validation cohort.

| Variable                  | Training cohort (n = 872) | Validation cohort (n = 399) | P value |
|---------------------------|---------------------------|----------------------------|---------|
| Plugs (Y/N)               | 196/676                   | 91/308                     | 0.90    |
| Gender (M/F)              | 481/351                   | 209/190                    | 0.36    |
| Age (year)                | 5.20 ± 2.93               | 4.99 ± 2.86                | 0.20    |
| Age (> 5 y, n)            | 387                       | 165                       | 0.31    |
| Birthweight               | 3.30 ± 0.49               | 3.32 ± 0.51                | 0.307   |
| Fever (day)               | 8.64 ± 4.41               | 8.77 ± 5.32                | 0.85    |
| WBC counts (x10^9/L)      | 8.21 ± 3.65               | 8.31 ± 3.80                | 0.65    |
| Neutrophil (%)            | 60.08 ± 16.66             | 60.89 ± 15.57              | 0.41    |
| Hgb (g/L)                 | 121.94 ± 10.38            | 122.79 ± 9.74              | 0.17    |
| PLT (x10^9/L)             | 312.62 ± 108.12           | 306.35 ± 114.13            | 0.35    |
| CRP (mg/L)                | 26.63 ± 33.75             | 23.60 ± 26.47              | 0.08    |
| PCT (ng/L)                | 0.35 ± 2.07               | 0.27 ± 0.52                | 0.54    |
| ALT (U/L)                 | 31.90 ± 112.67            | 25.62 ± 52.57              | 0.29    |
| AST (U/L)                 | 51.34 ± 151.55            | 45.87 ± 61.16              | 0.49    |
| IgG (g/L)                 | 9.51 ± 2.94               | 9.20 ± 2.71                | 0.09    |
| IgA (g/L)                 | 1.03 ± 0.64               | 1.19 ± 0.66                | 0.001** |
| IgE (U/L)                 | 223.70 ± 285.99           | 207.38 ± 280.46            | 0.38    |
| IL-2 (1.1–9.8 ng/L)       | 3.88 ± 2.88               | 3.19 ± 3.31                |         |
| IL-4 (0.1–3.0 ng/L)       | 3.74 ± 2.77               | 3.38 ± 3.86                | 0.06    |
| IL-6 (1.7–16.6 ng/L)      | 36.69 ± 69.10             | 49.97 ± 90.46              | 0.009** |
| IL-10 (2.6–4.9 ng/L)      | 7.88 ± 10.19              | 9.27 ± 13.91               | 0.09    |
| TNF-α (0.1–5.2 ng/L)      | 2.67 ± 1.93               | 2.55 ± 2.63                |         |
| IFN-γ (1.6–17.3 ng/L)     | 23.83 ± 36.51             | 25.28 ± 48.26              | 0.55    |
| Pleural effusion (n)      | 244                       | 112                       | 0.53    |
| Atelectasis (n)           | 124                       | 63                        | 0.93    |
| Complications (n)         | 298                       | 141                       | 0.97    |

**Figure 2.** Distribution of clinical characteristics between the training and validation cohorts. (a, b, c, d, e, and f) represented white blood cell (WBC) counts, neutrophil percentage (N%), hemoglobin (Hgb) concentration, platelet (PLT) counts, C reactive protein (CRP) and alanine aminotransferase (ALT) levels, respectively.
Statistical analysis. Continuous variables were expressed as mean and standard deviation (SD), and compared using an unpaired, 2-tailed t test or Mann-Whitney test. Categorical variables were compared using X^2 test or Fisher exact test. Clinical variables associated with increased risk of BMP formation were assessed based on clinical importance, scientific knowledge, and predictors identified in previously published articles. The associations of relevant clinical variables with BMP formation in MPP children were assessed using logistic regression models. Stepwise analysis with the Akaike information criterion (AIC) was used to identify variables for the multivariable regression models. Odds ratios (OR) were reported with their 95% CIs. Selected variables were incorporated in the nomogram to predict the probability of BMP formation in MPP children using statistical software (rms package of R, version 3.4.3; http://www.r-project.org). The nomogram was based on regression coefficient in multivariate logistic regression to 0 to 100-point scale.

The model performance was evaluated by the concordance index (C statistics) and calibration. The C statistic estimates the probability of concordance between predicted and observed outcomes and is equivalent to the area under the receiver operating characteristic curve (ROC). ROC analysis was used to calculate the optimal cutoff value, determined by maximizing the Youden index. Accuracy of cutoff value was assessed by the sensitivity, specificity, and predictive values. Calibration was evaluated using a calibration plot, a graphic representation of the relationship between the observed outcome and the predicted probabilities. The model was validated using bootstrapped resampling to decrease the overfitting bias. R statistical software packages were used to perform all the statistical analysis and graphics (R code, see supplementary information), P < 0.05 was considered statistically significant.

Results

Clinical characteristics. Four hundred children undergoing bronchoscopy were confirmed to have bronchial mucus plugs (BMP). Patients who had no M. pneumoniae infections and those with missing values on relevant predictors were not included in the study. Therefore, 287 children with MPP and BMP were included in the analytic cohort. Totally, 1271 children with MPP met the inclusion criteria. 872 and 399 children with MPP were divided into the training and validation cohorts, respectively (Fig. 1). The clinical characteristics of patients are seen Table 1. The baseline clinical data were similar between the training and validation cohorts, except serum IgA and IL-6 levels (Figs. 2 and 3). Bronchoscopy identified BMP in children with MPP were found in 196 (22.5%) and 91 (22.8%) patients in the two cohorts, respectively.

Predictors of BMP formation in children with MPP. In the training cohort, MPP children with BMP had longer duration of fever (7.18d vs 6.12d), higher neutrophil percentage (64.65% vs 58.75%), higher CRP level (36.64 vs 23.73 ng/L), higher IL-6 (61.25 vs 29.57 ng/L), IL-10 (11.28 vs 7.02 ng/L), and IFN-γ (40.58 vs 18.97 ng/L) levels compared with those without BMP (Table 2). A univariate logistic regression showed that age older than 5 years (OR, 2.95; 95% CI 2.11 to 4.10), longer fever time (OR, 1.06; 95% CI 1.02 to 1.09), higher neutrophil percentage (OR, 1.02; 95% CI 1.01 to 1.03), higher CRP level (OR, 1.01; 95% CI 1.006 to 1.014), higher ALT (>40 vs ≤40 u/L; OR, 3.04; 95% CI 1.99 to 4.63), higher IL-6 (OR, 1.006; 95% CI 1.003 to 1.008), IL-10 (OR, 1.04; 95% CI 1.02 to 1.05), and IFN-γ (OR, 1.015; 95% CI 1.01 to 1.02) levels, presences of pleural effusion (OR, 4.73; 95% CI 3.37 to 6.63) and atelectasis (OR, 3.57; 95% CI 2.40 to 5.33) were independently associated with BMP formation in MPP children, especially higher IL-10 (>10 vs ≤10 ng/L; OR, 2.84; 95% CI 1.97 to 4.05), higher IFN-γ (>30 vs ≤30 ng/L; OR,
2.83; 95% CI 1.99 to 4.03), and presence of complication (OR, 5.60; 95% CI 3.98 to 7.88) (Fig. 4). On multivariate regression analysis, age >5 years (OR 2.06; 95% CI 1.43 to 2.98), higher IL-10 level (>10 ng/L; OR, 2.19; 95% CI 1.46 to 3.28), higher IFN-γ level (>30 vs ≤30 ng/L; OR, 1.69; 95% CI 1.13 to 2.54), and presence of complication (OR, 3.43; 95% CI 1.45 to 8.09) were each independently associated with BMP formation (Table 3).

**Development and validation of a BMP-predicted nomogram.** Nomogram to predict BMP formation in children with MPP from the training cohort was shown in Fig. 5. The nomogram to predict BMP was created based on the following 4 independent predictive factors: age >5 years, higher IL-10 level (>10 vs ≤10 ng/L), higher IFN-γ level (>30 vs ≤30 ng/L), and presence of complication. Higher total points based on the sum of the assigned number of points for each factor in this nomogram were associated with the risk of BMP. The model demonstrated a good discriminative ability in estimating the risk of BMP, with a C statistics of 0.771 (95% CI, 0.734–0.808). Furthermore, calibration plot graphically showed a good agreement on BMP formation in children with MPP via bootstrap resampling (Fig. 6a).

In the validation cohort, the nomogram also showed a good discriminative ability for the estimation of BMP formation with C statistics of 0.796 (95% CI, 0.744–0.848). There was also a good calibration curve for the risk estimation of BMP in children with MPP (Fig. 6b). The optimal cutoff value of the total nomogram scores was determined as 80. The sensitivity and specificity when used in discriminating the presence of BMP from MPP children were 73.0% (95% CI, 70.0–75.9%) and 70.6% (95% CI, 67.6–73.6%) in the training cohort, and 84.6% (95% CI, 81.1–88.1%) and 60.7% (95% CI, 58.3–63.1%) in the validation cohort, respectively.

**Discussion**

The presence of BMP results in the poor clinical prognosis in children with MPP. Our study suggests that clinical variables, including age >5 years, higher IL-10 level (>10 ng/L), higher IFN-γ level (>30 ng/L), and presence of complication, are significantly associated with presence of BMP in children with MPP. Particularly, the presence of complications, including pleural effusion and atelectasis, strongly predicted the BMP formation in children with MPP.

| Variable | No plug (n = 676) | Plugs (n = 196) | P value |
|----------|-----------------|---------------|---------|
| Birthweight | 3.28 ± 0.50 | 3.35 ± 0.45 | 0.09 |
| Fever (d) | 6.12 ± 4.24 | 7.18 ± 4.25 | 0.002** |
| Gender (M/F) | 378/298 | 103/93 | 0.40 |
| Age (>5 y, n) | 260 | 127 | <0.01** |
| WBC counts (×10^9/L) | 8.30 ± 3.73 | 7.90 ± 3.37 | 0.17 |
| Neutrophil (%) | 58.75 ± 16.60 | 64.65 ± 16.07 | <0.01** |
| Hgb (g/L) | 121.90 ± 10.45 | 122.07 ± 10.20 | 0.84 |
| PLT (×10^9/L) | 313.97 ± 106.11 | 308.01 ± 114.86 | 0.50 |
| CRP (mg/L) | 23.73 ± 27.25 | 36.64 ± 48.85 | <0.01** |
| PCT (ng/L) | 0.30 ± 2.14 | 0.50 ± 1.83 | 0.27 |
| ALT (U/L) | 30.01 ± 121.05 | 38.39 ± 76.98 | 0.36 |
| AST (U/L) | 49.44 ± 168.51 | 57.86 ± 65.22 | 0.50 |
| LDH (U/L) | 296.79 ± 124.84 | 311.92 ± 107.87 | 0.124 |
| IgG (g/L) | 9.50 ± 2.84 | 9.58 ± 3.27 | 0.76 |
| IgA (g/L) | 0.95 ± 0.61 | 1.29 ± 0.66 | <0.01** |
| IgE (U/L) | 231.01 ± 292.34 | 197.57 ± 261.25 | 0.17 |
| IL-2 (ng/L) | 4.08 ± 3.18 | 3.19 ± 1.23 | <0.01** |
| IL-4 (ng/L) | 3.79 ± 2.61 | 3.58 ± 3.25 | 0.41 |
| IL-6 (ng/L) | 29.57 ± 53.67 | 61.25 ± 102.84 | <0.01** |
| IL-10 (ng/L) | 7.02 ± 8.24 | 11.28 ± 14.65 | <0.01** |
| TNF-α (ng/L) | 2.73 ± 2.11 | 2.43 ± 1.12 | 0.05 |
| IFN-γ (ng/L) | 18.97 ± 26.81 | 40.58 ± 55.70 | <0.01** |
| Pleural effusion (n) | 137 | 107 | <0.01** |
| Atelectasis (n) | 68 | 56 | <0.01** |
| Complications | 170 | 128 | <0.01** |
| ALT (>40 U/L) | 62 | 46 | <0.01** |
| IL10 (>10 ng/L) | 105 | 67 | <0.01** |
| IFN-γ (>30 ng/L) | 113 | 71 | <0.01** |

**Table 2.** Clinical characteristics of patients in the training cohort. ALT = alanine aminotransferase; AST = aspartate aminotransferase; CRP = C reactive protein; ESR = erythrocyte sedimentation rate; Hgb = hemoglobin; IFN = interferon; IgA = Immunoglobulin A; IgG = Immunoglobulin G; IL = interleukin; LDH = lactate dehydrogenase; PCT = procalcitonin; PLT = blood platelet; TNF = tumor necrosis factor; WBC = white blood cells. *P < 0.05, **P < 0.01.
In the present study, a univariate logistic regression showed that fever, neutrophil percentage, serum CRP, IL-2, IL-4, IL-6, and TNF-α levels were significantly associated with BMP formation in children with MPP. However, their OR values were relatively small, and had only a low statistical power. Therefore, these clinical variables were not included in further analysis. Additionally, multivariate regression model demonstrated that elevated serum ALT, the presence of pleural effusion or atelectasis had no significant correlation with BMP formation. Eventually, only age > 5 years, higher IL-10 level, higher IFN-γ level, and presence of complication entered the predictive model. Among the currently available predictive tools, a nomogram has high accuracy and good discrimination abilities in predicting outcomes with its convenience. The present nomogram performed well as supported by the C index values of 0.771 and 0.796 in the training and validation cohorts, respectively, and the optimal calibration curves demonstrating the agreements between prediction and actual observation.

A previous study showed that serum IL-18 and lactate dehydrogenase (LDH) levels can be used as parameters to determine which patients are candidates for corticosteroid therapy. Serum LDH level might be a useful...
marker for the evaluation of therapeutic efficacy in refractory MPP. A combination of age and serum levels of LDH, ESR ≥ 25 mm/h, and CRP ≥ 15 mg/L could be a predictive panel marker for early prediction of refractory MPP. The serum concentrations of TNF-α and IFN-γ were significantly higher in children with refractory MPP. Xu and colleagues demonstrated that age, fever duration, CRP and LDH were independent risk factors for BMP from patients with refractory MPP. In our study, although serum CRP in MPP children with BMP was higher than those without BMP, its statistical power was relatively small and not included in our nomogram. Different from the previous studies, our study did not show a significant difference in serum LDH levels between the children with and without BMP. This result might be more convincing because of a much larger population in our study. On the other hand, due to the limited availability of the commercial cytometric bead array kit, our institute only detected IL-2, 4, 6, 10, TNF-α, and IFN-γ, other cytokines were not included in the present nomogram. Of course, combined detection of cytokines would have a greater confidence to predict the presence of BMP in children with MPP.

In our BMP risk estimation nomogram, age >5 years, higher IL-10 level, higher IFN-γ level, and presence of complication demonstrated significant predictive value. For clinical use of the model, we assessed the risk of BMP in children with MPP using 80 as the cutoff value. MPP children with a score of 80 or more are high-risk of BMP formation. For example, an MPP child with >5 years plus serum IFN-γ level >30 ng/L would have a total of 82 points (50 points for age >5 years, and 32 points for serum IFN-γ level), strongly indicating the BMP presence.

In view of the prediction, the nomogram could serve as a tool to select patients for evaluating the necessity of flexible bronchoscopy. In general, the presence of persistent atelectasis in chest imaging is a strong indicator for flexible bronchoscopy intervention. Thus early bronchoscopic examination may be necessary for children with MPP presented with clinical sign of atelectasis, regardless of the presence of BMP. Considering a strong drive of atelectasis for bronchoscopy intervention, atelectasis had the top score and the highest weight in our prediction model. Notably, another complication of pleural effusion also showed similar value in the present predictive model. However, there was no relevant report about the relationship between pleural effusion and BMP formation in children with MPP. Their potential mechanism needs further investigation.

The use of the nomogram in estimating the risk of BMP in children with MPP to perform flexible bronchoscopy is a new concept. Our model depended on demographic data, thoracic imaging and serum cytokine levels to predict the risk of BMP in children with MPP. In addition, we demonstrated no improvement in model performance with the addition of variables originated from clinical manifestations (fever duration) and other laboratory markers (such as CRP level, neutrophil percentage, and serum IL-6 level). Although these variables are clearly important for diagnosis and management of BMP especially refractory MPP, the lack of improvement in model performance might further indicate complexity of MPP conditions.

### Table 3. Multivariate logistic regression analysis of bronchial mucus plugs based on the training cohort.

| Variable               | Odds Ratio (95%CI) | P value |
|------------------------|--------------------|---------|
| Age (>5 y vs ≤5 y)    | 2.06 (1.43–2.98)   | <0.001**|
| ALT (>40 vs ≤40 U/L)  | 1.56 (0.97–2.57)   | 0.07    |
| IL-10 (>10 vs ≤10 ng/L)| 2.19 (1.46–3.28)   | <0.001**|
| IFN-γ (>30 vs ≤30 ng/L)| 1.69 (1.13–2.54)   | 0.011*  |
| Pleural effusion (yes vs no) | 0.97 (0.45–2.08) | 0.93    |
| Atelectasis (yes vs no) | 1.41 (0.78–2.53)   | 0.25    |
| Complications         | 3.43 (1.45–8.09)   | 0.005** |

**Table 3.** Multivariate logistic regression analysis of bronchial mucus plugs based on the training cohort. ALT = alanine aminotransferase; IL = interleukin; IFN = interferon. *P < 0.05, **P < 0.01.
The present study had several limitations. First, our nomogram was internally validated using bootstrap methods, further studies are needed to externally validate the proposed nomogram. Moreover, a prospective study is required to further confirm the reliability of the nomogram. Second, because cytokine detections could not be available in all the patient or hospital, it would probably limit the popularity of this model. However, considering a higher weight of clinical variables for predicting BMP formation, clinicians should comprehensively assess a patient's condition before making a decision. Also, data collection was limited to those children with 1 to 15 years, so it is not clear whether the model would apply to younger children (<1 year) or older children (>15 years).

Third, in the present respective cohort, bronchoscopies were performed only in children with refractory MPP. Those children who got better after admission did not undergo bronchoscopy. The children could be considered to have no BMP. Therefore, it would inevitably lead to selective bias. Finally, because the nomogram was based on our institutional clinical data, other specific markers (such as IL-18) to estimate BMP risk might further improve the accuracy.

In conclusion, this study presents a nomogram demonstrating independent risk factors associated with BMP formation in children with MPP. The nomogram has potential to be used for early identification of BMP in children with MPP, contributing to a rational therapeutic choice.

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References
1. Ning, G. et al. The etiology of community-acquired pneumonia among children under 5 years of age in mainland China, 2001-2015: A systematic review. Hum. vaccines immunotherapeutics 13, 2742–2750 (2017).
2. Principi, N. & Esposito, S. Management of severe community-acquired pneumonia of children in developing and developed countries. Thorax 66, 815–822 (2011).
3. Wallihan, R. & Ramilo, O. Community-acquired pneumonia in children: current challenges and future directions. J. Infect. 69(Suppl 1), S87–90 (2014).
4. Sinaniotis, C. A. & Sinaniotis, A. C. Community-acquired pneumonia in children. Curr. Opin. Palm. Med. 11, 218–225 (2005).
5. Xu, Q. et al. Prediction of Bronchial Mucus Plugs Formation in Patients with Refractory/Mycoplasma PneumoniaePneumonia. Journal of Tropical Pediatrics, Iwm064 (2016).
6. Zhou, M. et al. Cause-specific mortality for 240 causes in China during 1990–2013: a systematic subnational analysis for the Global Burden of Disease Study 2013. Lancet 387, 251–272 (2016).
7. Waits, K. B. New concepts of Mycoplasma pneumoniae infections in children. Pediatr. Pulmonol. 36, 267–278 (2003).
8. Griffiths, U. K. et al. Pneumonia Mortality among Children under 5 in China from 1996 to 2013: An Analysis from National Surveillance System. PLoS one 10, e0133620 (2015).
9. Huang, L. et al. Independent predictors for longer radiographic resolution in patients with refractory Mycoplasma pneumoniae pneumonia: a prospective cohort study. BMJ open. 8, e023719 (2018).
10. Akashi, Y. et al. Clinical features and seasonal variations in the prevalence of macrolide-resistant Mycoplasma pneumoniae. J. Gen. family Med. 19, 191–197 (2018).
11. Moynihan, K. M. et al. Severe Mycoplasma Pneumoniae Infection in Children Admitted to Pediatric Intensive Care. Pediatr. Infect. Dis. J. 37, e336–e338 (2018).
12. Waits, K. B. & Tallington, D. F. Mycoplasma pneumoniae and its role as a human pathogen. Clin Microbiol Rev 17, 697–728, table of contents (2004).
13. Zhang, Y. et al. Effects of bronchoalveolar lavage on refractory Mycoplasma pneumoniae pneumonia. Respiratory care 59, 1433–1439 (2014).
14. Smalley, N., MacLaren, G., Best, D., Paul, E. & Butt, W. Outcomes in children with refractory pneumonia supported with extracorporeal membrane oxygenation. Intensive care Med. 38, 1001–1007 (2012).
15. Efrati, O. et al. Flexible bronchoscopy and bronchoalveolar lavage in pediatric patients with lung disease. Pediatric Crit. Care Med. 10, 80–84 (2009).
16. Ding, Y. et al. High expression of HMGB1 in children with refractory Mycoplasma pneumoniae pneumonia. BMC Infect. Dis. 18, 439 (2018).
17. Dunican, E. M. et al. Mucus plugs in patients with asthma linked to eosinophilia and airflow obstruction. J. Clin. investigation 128, 997–1009 (2018).
18. Wang, Z., Li, Y.-C., Zhou, X.-J. & Wu, J.-Y. Prediction of Refractory Mycoplasma Pneumoniae Pneumonia in Pediatric Patients. Pediatric Allergy, Immunology, Pulmonol. 30, 92–96 (2017).
19. Inamura, N. et al. Management of refractory Mycoplasma pneumoniae pneumonia: utility of measuring serum lactate dehydrogenase level. J. Infect. chemotherapy: Off J. Jpn. Soc. Chemotherapy 20, 270–273 (2014).
20. Wang, M. et al. Clinical and laboratory profiles of refractory Mycoplasma pneumoniae pneumonia in children. Int. J. Infect. diseases: IJD: Off. Publ. Int. Soc. Infect. Dis. 29, 18–23 (2014).
21. Wang, Y., Zhang, Y., Lu, W. & Wang, L. Serum Tumor Necrosis Factor-alpha and Interferon-gamma Levels in Pediatric Mycoplasma pneumoniae Pneumonia: A Systematic Review and Meta-Analysis. Can. respiratory J. 2018, 8354892 (2018).
22. Xu, X. E., Li, X. J., Liu, J. J., Wu, L. & Chen, Z. M. Serum cytokine profile contributes to discriminating M. pneumoniae pneumonia in children. Cytokine 86, 73–78 (2016).
23. Kim, Y. et al. Nomograms to Predict Recurrence-Free and Overall Survival After Curative Resection of Adrenocortical Carcinoma. JAMA Surg. 151, 365–373 (2016).
24. Lei, Z. et al. Nomogram for Preoperative Estimation of Microvascular Invasion Risk in Hepatitis B Virus–Related Hepatocellular Carcinoma Within the Milan Criteria. JAMA Surg. 151, 356 (2016).
25. Steyerberg, E. W. & Vergouwe, Y. Towards better clinical prediction models: seven steps for development and an ABCD for validation. Eur. heart J. 35, 1925–1931 (2014).
26. Moons, K. G. M. et al. Transparent Reporting of a multivariable prediction model for Individual Prognosis Or Diagnosis (TRIPOD): Explanation and Elaboration. Ann. Intern. Med. 162, W1 (2015).
27. Smeijsters, K. M. G. et al. Effect of Bronchoscopy on Gas Exchange and Respiratory Mechanics in Critically Ill Patients With Atelectasis: An Observational Cohort Study. Front. Med. 5, 301 (2018).
28. Bhat, J. J. et al. Flexible Bronchoscopy in Non-resolving Pneumonia. Indian. J. pediatrics 84, 681–684 (2017).
29. Yang, M. et al. Interleukin 17A as a good predictor of the severity of Mycoplasma pneumoniae pneumonia in children. Sci. Rep. 7, 12934 (2017).
30. Zhang, Y. et al. Cytokines as the good predictors of refractory Mycoplasma pneumoniae pneumonia in school-aged children. Sci. Rep. 6, 37037 (2016).
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Author contributions
All authors contributed to the development of the selection criteria, the risk of bias assessment strategy and data extraction criteria. Z.M.C. is the guarantor. X.F.X., H.W.L. and Y.J.S. drafted the manuscript. X.F.X., L.W. and Z.M.C. developed the search strategy. D.L.W., L.Y.L. and Y.T. provided statistical expertise. All authors read, provided feedback and approved the final manuscript.

Competing interests
The authors declare no competing interests.

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