Effects of Nrf2 Deficiency on Bone Microarchitecture in an Experimental Model of Osteoporosis

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Received 3 February 2014; Accepted 20 June 2014; Published 7 July 2014

Academic Editor: Jean-Claude Lavoie

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Objective. Redox imbalance contributes to bone fragility. We have evaluated the in vivo role of nuclear factor erythroid derived 2-related factor-2 (Nrf2), an important regulator of cellular responses to oxidative stress, in bone metabolism using a model of postmenopausal osteoporosis.

Methods. Ovariectomy was performed in both wild-type and mice deficient in Nrf2 (Nrf2−/−). Bone microarchitecture was analyzed by μCT. Serum markers of bone metabolism were also measured. Reactive oxygen species production was determined using dihydrorhodamine 123.

Results. Sham-operated or ovariectomized Nrf2−/− mice exhibit a loss in trabecular bone mineral density in femur, accompanied by a reduction in cortical area in vertebrae. Nrf2 deficiency tended to increase osteoblastic markers and significantly enhanced osteoclastic markers in sham-operated animals indicating an increased bone turnover with a main effect on bone resorption. We have also shown an increased production of oxidative stress in bone marrow-derived cells from sham-operated or ovariectomized Nrf2−/− mice and a higher responsiveness of bone marrow-derived cells to osteoclastogenic stimuli in vitro.

Conclusion. We have demonstrated in vivo a key role of Nrf2 in the maintenance of bone microarchitecture.

1. Introduction

Multiple pathogenetic mechanisms are responsible for loss of bone mass and skeletal microarchitectural deterioration leading to osteoporosis which represents a major challenge to our health systems. Excessive bone resorption or an inadequate bone formation in response to increased resorption during bone remodeling results in skeletal fragility [1]. Epidemiological studies have revealed that the majority of the affected individuals are postmenopausal women. It is estimated that in developed countries around 50 percent of women aged 50 and older will sustain an osteoporotic fracture during their lifetime. Osteoporotic fractures are associated with chronic pain and greatly impact the quality of life. Experimental findings support the concept that estrogen deficiency leads to increased bone turnover and plays a key role in osteoporosis [2].

Recent studies have demonstrated an important contribution of redox imbalance to bone fragility [3] and biochemical markers of oxidative stress are significantly increased in osteoporosis [4, 5]. In addition, menopause-related estrogen withdrawal might contribute to make bone more vulnerable to oxidative injury. Loss of estrogens decreases defensive mechanisms against oxidative stress and accelerates the effects of aging on bone [6]. Therefore, reduced antioxidant levels would enhance bone resorption [7] whereas the reduction in oxidative stress may provide protection against osteoporosis in aged and ovariectomized rats [8].

Nuclear factor erythroid derived 2-related factor-2 (Nrf2) plays an important role in the cellular defense against...
avitro studies have shown a role for this transcription factor oxygen species (ROS) signaling [16, 17]. In contrast, some induced osteoclastogenesis through modulation of reactive susceptibility to radiation-induced bone loss [15].

Ovaries of Valencia and Autonomous University of Madrid, Spain). The C57BL/6J Nrf2 knockout mice were kindly provided by Dr. Masayuki Yamamoto (University of Tsukuba, Japan). Genotyping of wild-type (Nrf2+/+) C57BL/6j mice and Nrf2-deficient (Nrf2−/−) littermates was done as reported previously [22]. Female Nrf2−/− and wild-type C57BL/6j mice between 6 and 9 months of age were included in this study. Animals of each strain were randomly allocated to ovariotomy (OVX) (n = 20) or Sham groups (n = 20). Surgery was performed after the mice were anesthetized with isoflurane and treated with butorfanol (2 mg/kg, s.c.). Ovaries of the mice in OVX group were removed through a midline incision of the skin and flank incisions of the peritoneum. The skin incision was then closed with metallic clips. Mice in the sham-operated group were anesthetized and sutured without removal of the ovaries. All mice were maintained in cages with a 12-hour light/dark cycle and free access to standard diet and water. Mice were housed and cared for by the veterinary staff in accredited facilities and were routinely screened for health status. On day 30 after surgery, mice were anesthetized, blood samples were taken by retro-orbital puncture, and animals were killed by cervical dislocation. Hind paws and L3–L5 vertebrae from randomly selected animals in each group were isolated and frozen for μCT and histomorphometric analysis.

2.2. Histomorphometric Analysis. Lumbar spines were dissected (L3–L5) and fixed in 10% formalin. After decalcification in 10% EDTA, the specimens were processed for paraffin embedding. Tissue sections (7 μm) were stained with hematoxylin and eosin to measure trabecular and cortical bone area, and osteoblast number relative to bone perimeter (N.Ob/BPm, mm−1) or with TRAP and counterstained with hematoxylin and aniline blue to determine the percentage of bone surface covered by osteoclasts (Oc.S/BS, %).

2.3. Measurement of Bone Markers. Enzyme-linked immunosorbent assay (ELISA) kits were used to determine serum levels of tartrate-resistant acid phosphatase (TRAP)-5b, with sensitivity of 0.1 U/L, and C-terminal telopeptides of type I collagen (CTX-I), with sensitivity of 2.0 ng/mL (IDS, Paris, France). Serum levels of osteocalcin, osteoprotegerin (OPG), and RANKL were determined by LumineX, with sensitivity of 70, 2.3, and 3.3 pg/mL, respectively (Millipore Corporation, Billerica, MA, USA). Alkaline phosphatase (ALP) levels in serum were determined as previously described [21].

2.4. X-Ray Microcomputed Tomography (μCT). Hind paws were kept wrapped in gauzes soaked in 0.9% NaCl at −20°C until their analysis by μCT. Samples were analyzed with a SkyScan 1172 μCT equipment (Bruker microCT NV, Kontich, Belgium). Three-dimensional trabecular microarchitecture was analyzed at the distal metaphysis of the femur in a region of 1.5 mm. Samples were imaged with an X-ray tube voltage of 50 kV, current of 200 μA, at a scanning voxel size of 5.5 μm, and with the use of an aluminum filter (0.5 mm in thickness). The scanning total angular rotation was 185° with an angular increment of 0.4°. Datasets were reconstructed using a modified Feldkamp algorithm [23] and segmented into binary images using adaptive local thresholding previous to analysis.

2.5. Isolation of Bone Marrow Cells. Bone marrows were flushed out of femur and tibia with DMEM (Sigma-Aldrich, St Louis, MO, USA) containing penicillin (100 U/mL) and streptomycin (100 mg/mL), supplemented with 10% fetal bovine serum (Sigma-Aldrich) (complete DMEM) using a 25-gauge needle. Cells were treated with red blood cells lysis buffer (0.15 M NH₄Cl, 1 mM KHCO₃, 0.1 mM Na₂EDTA, pH 7.4), washed twice with complete DMEM, and seeded in 96-well tissue culture plates at 5 × 10⁵ cells per well (oxidative stress quantification) or 3 × 10⁵ cells per well (osteoclast differentiation).

2.6. Oxidative Stress Quantification. ROS production in bone marrow-derived cells was determined using dihydrodorozamide 123 (Molecular Probes, Invitrogen S.A., Barcelona, Spain), which is oxidized to fluorescent rhodamine 123 (485 nm (excitation)/534 nm (emission)). Cells were washed with DMEM (Sigma-Aldrich), incubated with dihydrodorozamide 123 (5 μM) in DMEM for 10 min at 37°C and then 12-O-tetradecanoylphorbol-13 acetate (TPA, 200 ng/mL) was added and cells were incubated for 30 min at 37°C. Cells were washed, resuspended in PBS, and analyzed by flow cytometry.
Figure 1: Structure of trabecular bone of femur. (a) Three-dimensional μCT analysis. vBMD: volumetric bone mineral density; BV/TV: bone volume fraction; BS/TV: bone surface density; Tb.Sp: trabecular spacing; Tb.N: trabecular number; Tb.Th: trabecular thickness; OVX: ovariectomy. Results are expressed as mean ± SD (n = 3). ∗P < 0.05, ∗∗P < 0.01. (b) Representative μCT images of trabecular bone of femur are shown.

2.7. Osteoclast Differentiation. Bone marrow-derived cells were cultured in complete DMEM medium containing macrophage-colony stimulating factor (M-CSF) (30 ng/mL) and RANKL (50 ng/mL). Medium was changed every 2 days. TRAP staining was performed after 8 days of culture. Cells were fixed with 4% paraformaldehyde in PBS for 5 min, washed, and incubated in 0.2 M acetate buffer (0.2 M sodium acetate, 0.05 M tartaric acid, and water; pH 5.0) for 20 min at 37°C. Cells were then washed and incubated in a solution containing distilled water, 0.5 mg/mL naphthol AS-MX phosphate (Sigma-Aldrich), and 1.1 mg/mL fast red violet LB salt (Sigma-Aldrich) for 3 h at 37°C. Finally, cells were washed and counterstained with hematoxylin/aniline blue, and TRAP+ multinucleated cells were counted.

2.8. Statistical Analysis. The results are presented as mean ± SD. The level of statistical significance was determined by one-way ANOVA followed by Bonferroni’s post test.

3. Results

3.1. Effects on Bone Architecture in Femur. The μCT analysis indicated that Nrf2 deficiency in either sham-operated or ovariectomized mice affected bone architecture in femur. As shown in Figures I(a) and I(b), Nrf2 deficiency significantly reduced volumetric bone mineral density (vBMD) in trabecular bone and a tendency was observed towards a reduction in bone volume fraction (BV/TV) and bone surface density (BS/TV). In addition, Nrf2 deficient mice also tended to show lower values of trabecular number (Tb.N), accompanied with a higher trabecular spacing (Tb.Sp) in ovariectomized mice.

(FACSCanto, Becton Dickinson, Franklin Lake, NJ, USA). Each experiment was done in triplicate.
3.2. Effects on Vertebral Bone. Histomorphometric analysis of vertebral bone showed a significant reduction in the ratio cortical area/total area accompanied with a tendency to a lower trabecular area/total area ratio in sham-operated or ovariectomized Nrf2-deficient mice compared with wild-type controls (Figure 2). In addition, a tendency to higher osteoblast numbers was observed in the presence of Nrf2 deficiency in either sham-operated or ovariectomized animals. Interestingly, osteoclast surface was significantly higher in Nrf2−/− mice relative to their corresponding wild-type controls. Ovariectomy tended to reduce osteoblast numbers and significantly increased osteoclast surface.

3.3. Effects on Serum Markers of Bone Turnover. No significant effects were obtained for serum ALP and osteocalcin (early and late markers of osteoblast differentiation, resp.) levels (Figure 3) although a tendency to higher values was observed in the presence of Nrf2 deficiency. CTX-1 and TRAP-5b levels indicate osteoclast activity and osteoclast number, respectively, and the ratio CTX-1/TRAP-5b is a sensitive resorption index [24]. CTX-1/TRAP-5b as well as the ratio RANKL/OPG, an index of osteoclastic activation, increased in Nrf2−/− mice. In addition, ovariectomy augmented CTX-1/TRAP-5b and RANKL/OPG.

3.4. Osteoclast Differentiation. Increased serum levels of TRAP-5b/CTX-1 and RANKL/OPG as well as osteoclast surface in vertebrae sections were observed in Nrf2−/− mice. In order to confirm the response to relevant osteoclastogenic stimuli [16, 17] in our experimental model, we used bone marrow-derived cells of mice from the different experimental groups treated with M-CSF and RANKL. Figure 4 shows that deficiency in Nrf2 enhanced in vitro osteoclast differentiation in either sham-operated or ovariectomized animals.

3.5. Oxidative Stress. The possible effects on ROS production were investigated in our experimental model. For this purpose, we assessed oxidative stress in bone marrow-derived
cells by quantifying intracellular ROS accumulation. As expected, Nrf2 deficiency resulted in a higher oxidative stress either in sham-operated or ovariectomized mice (Figure 5). In addition, ovariectomy tended to increase ROS production.

4. Discussion

A wide range of evidence indicates that Nrf2 regulates the expression of genes involved in cellular detoxification and maintains cellular redox homeostasis [25, 26]. In this work, we focused on the possible role of Nrf2 in bone metabolism because oxidative stress is an important mediator of bone loss in estrogen deficiency. Analysis of bone parameters in Nrf2−/− mice and their wild-type littermates either in the absence or presence of ovariectomy showed that Nrf2 is required for the maintenance of bone quality. We demonstrate that mice deficient in Nrf2 exhibit a loss in trabecular bone mineral density in femora and a reduction in cortical bone area in vertebrae. These changes induced by Nrf2 deficiency may result in bone fragility and fracture risk.

Nevertheless, enhanced bone formation activity has been reported in 9-week-old Nrf2−/− mice [27]. Further studies are needed to determine the influence of age on Nrf2 regulatory effects in bone.

This study has demonstrated an increased production of ROS by bone marrow cells from either sham-operated or ovariectomized Nrf2-deficient mice which is consistent with the constitutive lower levels of antioxidants reported in cells from these animals [9]. A wide range of evidence indicates that ROS are intermediaries in osteoclast metabolism leading to osteoclast differentiation or activation [7, 28]. Therefore, ROS are mediators of RANKL signaling in osteoclasts [29] although they do not seem to be directly involved in the bone resorption process [30]. We have shown that Nrf2 deficiency increases osteoclast numbers in vivo which would be the consequence of a higher osteoclastogenenic stimulation. Previous in vitro studies have shown higher levels of osteoclast differentiation induced by RANKL and bone resorption using cells from Nrf2-deficient mice [17, 27]. In addition to a higher responsiveness of bone marrow-derived cells.
to osteoclastogenic stimuli in vitro, we have demonstrated the consequences of Nrf2 deficiency on relevant in vivo biomarkers such as the ratio RANKL/OPG in serum which was significantly increased leading to enhanced osteoclast differentiation in vivo. Serum levels of resorption markers and the μCT and histomorphometric analyses confirmed that the increased osteoclast number reflects an increased bone resorption in vivo. We have also shown a high production of oxidative stress in bone marrow-derived cells after ovariectomy. The accumulation of ROS in these cells would activate T cells through CD80 upregulation on dendritic cells [31]. In addition, the lowering of antioxidants by estrogen deficiency may sensitize osteoclasts to osteoclastogenic signals promoting bone resorption [7]. Nevertheless, our results in this experimental model suggest that the maximal osteoclastogenic response to oxidative stress would be elicited by Nrf2 deficiency as ovariectomy did not further increase it.

On the other hand, an increased oxidative stress may contribute to the inhibition of osteoblast differentiation [32] and proliferation [33] or the induction of cell death [34]. Previous studies have suggested the involvement of Nrf2 in bone mineralization. Stable overexpression of Nrf2 inhibits
Figure 5: Production of oxidative stress by bone marrow-derived cells. Oxidative stress was measured in bone marrow-derived cells stimulated with TPA by flow cytometry using dihydrorhodamine 123 (DHR). RFI: relative fluorescence intensity; OVX: ovariectomy. Data represent mean ± SD (n = 4). *P < 0.05.

The results of our study thus support a role for Nrf2 in bone metabolism in vivo.

Development of therapies for improving bone formation and bone repair has considerable impact. We found that genetic deletion of Nrf2 enhances bone turnover and results in deterioration of bone structure either in sham-operated or ovariectomized mice. Our data indicate that this transcription factor is indispensable for normal bone microarchitecture and suggest that Nrf2 may be a pharmacological target to maintain bone integrity in pathological situations.
Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This work was supported by grants SAF2010-22048, SAF2010-17822, RD12/0043/0013 (Ministerio de Economía y Competitividad, ISCIII, FEDER), and Prometeo2010-047 (Generalitat Valenciana). Lidia Ibáñez thanks Spanish Ministerio de Economía y Competitividad for a fellowship (FPI).

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