Immunodetection of FGF1-2 and FGFR1-2 indicates that these proteins disappear in the wound epidermis and blastema of the scarring limb in lizard

Abstract
In lizards while the tail broadly regenerates after amputation, the limb generally forms short scarring outgrowths. FGFs are important growth factors that are present in regenerating tissues and appear to stimulate tail regeneration in lizards. In the limb, immunohistochemistry at 15-16days post-amputation indicates that FGFs and FGFR1-2 are present with different intensity in the wound (regenerating) epidermis and sparsely in connective, muscle and cartilaginous cells of the healing tissues of the stump. However the immunofluorescence study indicate that FGF and FGF1-2 receptors are little or no immunodetectable at 25-26days post-amputation, especially in the wound epidermis, when the limb blastema is turning into a scarring connective tissue of the short limb outgrowth. Western blot analysis shows no low MW bands for FGF1-2 and altered or degraded bands of FGFR1-2 in the limb at 25-26days post-amputation when scarring is taking place. The present observations indicate that the disappearance of these proteins is related to loss of cell proliferation that leads to the failure of limb regeneration in lizards. The study further stress the essential role of FGFs and their receptors in the stimulation of organ regeneration in lizards and likely in other amniotes.

Keywords: lizard, limb, regeneration, scarring, FGFs, FGFR1. immunohistochemistry

Introduction
Among amniotes only lizards are capable of regenerating the tail after amputation but not the limb or digits that instead forms a scar or tail-like outgrowths.1-4 The initial phase of limb healing determines the re-epithelialization of the stump and the accumulation of a bulk of connective cells forming a small soft mound termed limb blastema. The microscopic studies of the blastema have shown the presence of few mesenchymal cells accumulating over the injured tissues and instead a massive infiltration of granulocytes and macrophages during the first 3 weeks post-amputation. These cells determine an intense inflammatory reaction that is retained as the main cause of the lack of limb regeneration, as it has been recently indicated also by a transcriptome analysis.8 One of the consequences of the intense inflammation is the stimulation of the differentiation of a dense, irregular connective tissue while the amputated bone a cartilaginous cup seals the damage in the bone tissue. The final result is therefore the rapid transition from a soft blastema to a denser, harder, scaled, and flat or short scarring outgrowths.3,6-8

In particular it is believed that inflammation impedes the formation/maintenance of an Apical Epidermal Peg (AEP) at the tip of the wound epidermis of the limb, while keratinocytes rapidly differentiate into a mature epidermis.6-9 The AEP is retained essential to maintain tail regeneration, and is linked to the production of Wntless protein (Wnt) and Fibroblast Growth Factors (FGFs). Previous studies have indicated that FGF1-2 tend to disappear in the epidermis of the scarring limb blastema,10 suggesting that inflammation depresses the FGF signaling pathway and the maintenance of an AEP that is needed for maintaining tail regeneration. The use of FGFR1 antagonists also reduces or eliminates regeneration in lizards if administered before or at the blastema stage.11-13 Finally, recent transcriptome studies have also shown the up-regulation of FGFs and their receptors in the regenerating tail6 while these are absent in the scarring limb.9 The present study extends these initial findings on FGF1-2 to the detection and localization of FGF receptors 1 and 2 (FGFR1-2), the specific membrane proteins that bind to FGFs.11,16 The present study stresses the importance of the FGF signaling pathways for organ regeneration in lizards, and amniotes in general.

Materials and methods
In the present study we have utilized fixed tissues originally prepared in previous studies from adults of the lizard Podarcis muralis.17 The experiments were conducted following the Italian guidelines for animal care and handling (art. 5, DL 116/92). The lizards were kept in a terrarium at 25–33°C with a photoperiod of 12-14hours of light, and they were fed with a diet of mealworms, fruit-flies and fly larvae. The lizards were maintained at 4-6°C for 30 minutes to produce hypothermia and numbness, and hindlimb amputation was done in the distal part of the tight close to the knee, using a sharp razor blade, after anesthetizing the lizards with ethyl ether. Stump samples with their healing blastema from 15-16days (n=3), and 25-26days (n=3) post-amputation, were selected among others for the present study, and were fixed in 4% paraformaldehyde in 0.1M Phosphate buffer at pH 7.4 at 0-4°C for 5hours, rinsed in buffer for about 30minutes, dehydrated in ethanol (70%, 80%, 95%), infiltrated in xylene, and embedded in wax. The tissues were sectioned at 6-8-8m in the longitudinal plane using a microtome, and were collected and dried on glass slides. Sample sections were stained with Hematoxylin-Eosin (H & E) for the general histological study.

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Results

In the selected limb blastemas presented in this study, the regenerating limb at 16 days after amputation appeared as a small outgrowth covered by a shiny dark epidermis tissue here termed a regenerating blastema for analogy with the blastema of the regenerating tail details in. 6,7,17 However, differently from the tail blastema, the limb blastema is generally destined to form a scarring outgrowth in the following days, and it grows to about 1 mm before turning into a scaled outgrowth. Among the proximal tissues included in the limb stump at 16 days after the amputation, some fragmented and repairing muscles were present, and the sectioned femur showed a compact lamellar bone in the diaphysis while its proximal, untouched epiphysis showed in the cavity of the secondary ossification center the presence of some cartilaginous cells (Figure 1). At 26 days the analyzed blastema was still covered by a thick wound epidermis with a thicker corneous layer and contained a dense and irregular connective tissue, most composed by elongated fibroblasts in the superficial, apical region, and an even denser connective tissues formed by spindle-shaped fibroblasts in proximal regions, in contact or penetrating among the original tissues of the stump (Figure 1C-G). Numerous blood vessels were sparsely distributed among fibroblasts, and mitotic figures of dividing fibroblasts were rare to absent.

Figure 1 Histological examples of regenerating limb at 16 days (A, B) and 26 days (C-G) post-amputation. A, at 16 days the amputated limb appears partially covered by a scab (arrowheads in the inset, Bar 0.5mm). Histological section shows a thick wound epidermis covering a blastema made of a loose connective tissue. Bar, 10µm. B, detail on the femur remnant contacting the blastema, and showing the degenerated cartilage in the epiphysis. Bar, 50µm. C, limb stump covered by a roundish blastema (arrowhead). Bar, 0.5mm. D, histology of the blastema showing a loose connective tissue with sparse blood vessels. Bar, 50µm. E, detail on the thick wound epidermis with a multilayered corneous layer (arrow). Bar, 10µm. F, a detail of the superficial blastema shows numerous fibroblasts. Bar, 10µm. G, a more proximal region of the blastema, near the stump tissues, shows elongated fibroblasts forming a dense connective tissue. Bar, 10µm. Legends: bl, blastema; bm, bone marrow; bo, femur compact bone (diaphysis); ep, epiphysis; gp, growth plate (metaphysis); mu, stump muscles; sc, scale; v, blood vessels; we, wound (regenerating) epidermis.

Western blotting observations from the analyzed healing limbs of 25-26 days post-amputation showed main immunopositive bands at 48-50, 67-70 and 75-77kDa for FGF1 and FGF2, the latter showing...
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After separation of the regenerating epidermis from the underlying connective tissues indicated for simplification as “regenerated dermis” (Figure 2), western blot immunodetection for receptors showed in the regenerating (wound) epidermis bands at 55 and 58-60kDa for FGFR1 (Figure 2, fourth lane) and a band around 120kDa and 18kDa in the regenerating dermis for FGFR1 (Figure 2, fifth lane). The regenerating epidermis showed a main band around 80kDa, and weaker at 85 and 57-60kDa for FGFR2 (Figure 2 sixth lane), while almost no bands or a very weak band at 47kDa for FGFR2 was seen in the regenerating dermis (Figure 2, seventh lane).

Immunolabeling for FGF1 and FGF2 showed a more intense immunofluorescence in the wound epidermis and less in the blastema with both FGF1 and FGF2 at 16 days post-amputation, although FGF1 showed a higher labeling also in blastema cells (Figure 3A) (Figure 3B). Periosteal cells covering the lamellar bone of the femur and chondroblasts of its repairing distal end, originally transected, also showed some immunolabeling for FGF1 and FGF2 at 16 days (Figure 3C). Immunofluorescence for both FGFs lowered in the healing tissues of the limb blastema at 26 days post-amputation, especially in the wound epidermis covering the outgrowth (Figure 3D) (Figure 3E). An un-even immunofluorescence remained in the repairing or regenerating muscles, in the periosteum and in chondrocytes of the healing femur at its distal, previously transected ends (data not shown). No labeling was seen in control sections, aside a non-specific fluorescence in blood vessels and in some auto fluorescence in the corneous layer of the epidermis (Figure 3F).

The immunolocalization at 16 days post-amputation for FGFR1 showed an immune labeled wound epidermis while sparse and weakly labeled cells were observed in cells located in the connective tissue located underneath the wound epidermis, here, indicated as blastema (Figure 4A) (Figure 4B). The FGFR1 labeling was instead present among the repairing muscles and in the cartilage cells and periosteum of the distal, healing region of the initially transected bone of the femur (Figure 4C) (Figure 4D). Interestingly, sparse immune labeled cells were also detected in the cartilaginous cells present in the proximal epiphysis of the femur, a region untouched by the amputation (Figures 1B) (Figure 4E). Control sections appeared immunonegative (Figure 4F). Differently from 16 days, almost no immune labeling for FGFR1 was seen in the blastema at 26 days post-amputation, in both the wound epidermis and the underlying blastema (Figure 5A).

However, in the cartilage of the regenerating femur and in that of its proximal epiphyses, the latter untouched by the amputation, sparse cartilaginous labeled cells for FGFR1 were still seen even at 26 days post-amputation (Figure 5B) (Figure 5C). A similar observation was also made for FGFR2, that appeared present in the apical wound epidermis at 16 days but was low to absent in the blastema (Figure 5D). Sparse labeled cells were however detected in the proximal epiphyses of the femur (not hit by the transaction) at 16 and 26 days post-injection also for FGFR2 (Figure 5E). Immunofluorescence for FGFR2 lowered or disappeared at 26 days in both wound epidermis and blastema cells, despite the non-specific auto fluorescence of blood vessels (Figure 5F).

Figure 2 Western blotting for FGF1, FGF2, FGFR1 and FGFR2 in the regenerating limb (RL), in the regenerating limb epidermis (RLE) and regenerated limb dermis (+inner tissues, indicated as RLD). The MW is indicated in the ordinate in kilo Daltons (kDa).

Figure 3 TRITC immunofluorescence for FGF1 (A, C, D) and FGF2 (B, E) in regenerating limbs at 16 and 26 days post-amputation. Bars indicate 10 μm in all figures. A, at 16 days both the wound epidermis and sparse cells in the blastema are immunofluorescent (v indicate non-specific fluorescence in blood vessels). B, thick and immunofluorescent wound epidermis at 16 days. C, sparse immunofluorescence in both the wound epidermis and underlying blastema connective and the end of the transected femur (outlined by dashes) showing labelled cells (chondroblasts and periosteal cells). D, lowering of immunofluorescence in the repairing connective and muscles of the blastema, that almost disappears in the wound epidermis aside non-specific labeling of fragments of the corneous layer. E, lowering to disappearing of immunofluorescence in the wound epidermis and underlying connective blastema. F, immunonegative control (CO). Legends: bl, blastema; bo, bone tissue (femur); c, corneous layer; ca, cartilage tissue; mu, periosteal cells on the transected femur; we, wound epidermis (filled with auto-fluorescent erythrocytes); we, wound epidermis. The dashes underline the wound epidermis in A, D, E, F, and they outline the cartilaginous apex of the transected femur in C and D.

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In the cases here analyzed, FGF1-2 and FGFR1-2 appear present where cell proliferation is still active at 15-16 days post-injury, especially in the regenerating epidermis, in sparse blastema cells, and mainly in the repairing muscle cells or in the periosteum and cartilaginous epiphyses of the femur. FGF immune labeling tends to disappear later on when the proliferation in the limb decreases and only few tissues such a muscle and cartilage are still repairing. Cell proliferation and FGF immune labeling is also abolished in scarring tails after cauteronization, an intervention that drives the tail to form a scar instead of regenerate into a new tail. The importance of FGFs and FGFRs for lizard regeneration confirms previous observations, and the numerous data available for amniotes such as amphibians and fish, two groups of vertebrates where tissue and organ regeneration is even higher than in lizards. The researches on amphibians and fish have indeed shown that FGFs and their receptors are mitogenic growth factors for the regeneration of the amphibian limb or the fish fin, mimicking most of the activity of the Neurotrophic Factor. FGFs are released from nerves and the spinal cord, and maintain cell proliferation in the blastema, especially FGF1 while FGF2 is present and release from the Apical Epidermal Cap and maintains cells proliferation in concert with other signaling proteins.

Discussion

The regeneration of limb outgrowths of different length in lizards depends from some conditions, including trimming the exposed femur after the shrinkage of the stump connective and muscle tissues during the first days post-trauma, and keeping the temperature above 25°C. In our selected cases, we analyzed the best conditions when the stump was fully re-epithelialized already at 16 days after the injury and formed a blastema-like outgrowth at 26 days that was accumulating fibroblasts and formed ascolling connective tissues rich in irregular extracellular fibrils.

In the cases where analyzed, FGF1-2 and FGFR1-2 appear present where cell proliferation is still active at 15-16 days post-injury, especially in the regenerating epidermis, in sparse blastema cells, and mainly in the repairing muscle cells or in the periosteum and cartilaginous epiphyses of the femur. FGF immune labeling tends to disappear later on when the proliferation in the limb decreases and only few tissues such a muscle and cartilage are still repairing. Cell proliferation and FGF immune labeling is also abolished in scarring tails after cauteronization, an intervention that drives the tail to form a scar instead of regenerate into a new tail.

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The initial presence of FGF and their receptors in the first 2 weeks of limb healing in lizards likely stimulates the wound epidermis to proliferate in order to cover the exposed tissues of the stump. FGF2 also stimulated angiogenesis and the rate of blastema growth in the tail. However, the intense and persisting inflammation in the connective tissues, bones and muscles of the limb due to a massive accumulation of immune cells, macrophages and lymphocytes, in the stump and initial blastema, leads to scarring. The detected immunopositive bands for FGFs and their receptors at 50-80kDa, a range outside that expected for FGF (16 to 27kDa), indicate polymerization of lower bands or degradation of the High MW precursors in our material. The lack of the active protein bands at 18-22kDa for FGF1 and FGF2 and of higher MW bands for their receptors in the limb, further confirms the decrease or complete disappearing of these signaling proteins during these later stages of limb recovery. FGFs are instead present in the regenerating tail where FGF appears stimulating tissue regeneration. Another indication of the important role of FGFs for tail regeneration in lizards derives from experiments where the administration of FGF and/or FGFs inhibitors before and during blastema formation determines failure of tail regeneration. In conclusion FGF and FGFR1-2 are part of the essential signaling pathways that sustain the growth of the blastema and regeneration of cartilage, muscle, and the new epidermis in lizards and vertebrates in general.

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None.

Conflict of interest
The author declares no conflict of interest.

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