Phylogenomic and biogeographic reconstruction of the *Trichinella* complex

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Trichinellosis is a globally important food-borne parasitic disease of humans caused by roundworms of the *Trichinella* complex. Extensive biological diversity is reflected in substantial ecological and genetic variability within and among *Trichinella* taxa, and major controversy surrounds the systematics of this complex. Here we report the sequencing and assembly of 16 draft genomes representing all 12 recognized *Trichinella* species and genotypes, define protein-coding gene sets and assess genetic differences among these taxa. Using thousands of shared single-copy orthologous gene sequences, we fully reconstruct, for the first time, a phylogeny and biogeography for the *Trichinella* complex, and show that encapsulated and non-encapsulated *Trichinella* taxa diverged from their most recent common ancestor ~21 million years ago (mya), with taxon diversifications commencing ~10 – 7 mya.

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Parasitic diseases cause substantial morbidity and mortality in billions of animals and humans worldwide, and also major losses to the global food production annually. Parasitic roundworms (nematodes) of the genus *Trichinella* cause disease (trichinellosis) in humans, which can lead to serious morbidity and mortality1–3. Although trichinellosis is endemic in many regions of the world, the predominant impact of human disease relates principally to acute outbreaks following consumption of infected, raw meat products2, with examples in Argentina, China, Laos, Papua New Guinea, Romania and Vietnam3–9.

*Trichinella* is a complex of at least 12 species and genotypes, with a broad geographic range, including Europe, Africa, Asia, the Americas and Australasia10,11. Although only morphologically distinguishable groups (that is, encapsulated and non-encapsulated clad) of *Trichinella* are recognized, based on the appearance of larvae in muscle cells in infected hosts, molecular studies have defined nine species and three genotypes that display extensive biological diversity10–13. Currently, based on genetic studies have defined nine species and three genotypes that display extensive biological diversity10–13. Currently, based on genetic

**Results**

**Genomes and transcriptomes.** We sequenced and assembled 16 draft genomes and 15 transcriptomes from all 12 currently recognized *Trichinella* taxa, including five distinct geographic isolates of *T. pseudospiralis* (Tables 1 and 2 and Supplementary Tables 1–4). The draft assemblies of these genomes (available publicly; see ‘Accession codes’ section) ranged from 46.1 to 51.5 Mb (mean: 49.0 Mb), with average scaffold N50s of 106–294 kb (mean: 196 kb) and GC contents of 32.5–33.7% (33.2%; Tables 1 and 2). For all assemblies, 96.4% (95.6–97.2%) of all 248 core essential genes were identified (Tables 1 and 2), indicating that the gene sets represent substantial proportions of individual genomes. The repeat contents of individual draft genomes were estimated at 6.7–21.8% (mean: 17.7%) of their total nucleotide compositions (Supplementary Tables 5 and 6). On average, the assemblies contain 2.4% (range: 1.1–3.7%) retrotransposons, 2.9% (0.7–5.5%) DNA transposons, 7.7% (0.04–10.6%) unclassified dispersed elements and 4.8% (4.1–5.8%) simple repeats (Supplementary Tables 5 and 6). The repeat content of the genome of genotype T9 (6.7%) is exceptionally low compared with those of all other *Trichinella* taxa (mean: 17.7%). A comparison of the present draft genome for *T. spiralis* (ISS3) with that published previously for same species (ISS195)19 revealed essentially the same percentage of core essential genes (96%), and repeat and GC contents, but a size difference (14 Mb), likely to relate to differing sequencing and assembly methodologies or a genetic distinctiveness between the two geographic isolates.

**Protein-encoding gene sets.** From the 16 genomes representing all *Trichinella* taxa, we predicted 11,006–16,067 (mean: 13,912) protein-encoding genes that were 2,071–3,169 bp in length (mean: 2,632; with introns), exons that were 170–223 bp in length (mean: 203) and introns that were 218–284 bp in length (mean: 259), with 5.7–6.9 exons per gene using complementary, *de novo* and homology-based approaches (Tables 1 and 2). These genes and introns are shorter than those of *Trichuris suis* (mean: 3,812 and 510 bp), *Haemonchus contortus* (6,167 and 832 bp), *Ascaris suum* (mean: 6,636 and 1,081 bp) and *Toxocara canis* (8,416 and 1,133 bp), but are similar to those of *Brugia malayi* (3,106 and 311 bp) and *Caenorhabditis elegans* (3,680 and 320 bp)22–28. In total, 7,659–11,931 (mean: 9,697; 69.9%) of the protein-encoding genes had homologous sequences (BLASTp E value: 10−5) in the NCBI nr protein database (7,655–11,930; mean: 9,694; 69.7%), UniProtKB/SwissProt database (5,143–5,912; mean: 5,603; 40.3%) and WormBase (4,891–5,415; mean: 5,205; 37.4%). Genes with domain and motif matches in InterProScan (n = 5,289–6,372; mean: 5,847; 42.0%) contained 4,626–5,362 (5,035; 36.2%) hits in Pfam, 5,018–5,879 (5,489; 39.5%) in PANTHER, 929–1,037 (992; 7.1%) in PRINTS and 234–266 (252; 1.8%) in PIRSF databases (Supplementary Tables 7 and 8). In total, 7,691–12,000 (mean: 9,730) orthologous genes (E value ≤ 10−5) were associated with biological (KEGG) pathways. The secretomes of individual *Trichinella* taxa were predicted to comprise 314–414 (mean: 363) excretory/secretory (ES) proteins, 238 of which are encoded by single-copy orthologous genes (SCOs; Supplementary Data 1) and include cysteine, metallo- and serine proteases, peptidase inhibitors and orphan (uncharacterized) proteins identified previously29–32.

**Phylogeny.** We reconstructed the phylogeny of all 12 currently recognized *Trichinella* taxa (cf. Tables 1 and 2). First, we conducted an analysis of a complete protein sequence data set representing all 1,284 SCOs shared by all taxa, including five distinct geographic isolates of *T. pseudospiralis*, and two outgroup species (that is, *Trichuris suis* and *A. suum*) using Bayesian inference, maximum likelihood and maximum parsimony to establish the phylogenetic relationships of all encapsulated and non-encapsulated *Trichinella* taxa. Here all three trees constructed had the same topology, with strong support values (1.0; 87–100%) at all nodes, irrespective of the algorithm used (Fig. 1a). Second, employing the same algorithms, we independently assessed the relationships of all 16 *Trichinella* gene sets (without an outgroup) using protein sequences encoded by 2,855 SCOs shared by all taxa. Again, all three resultant trees had the same topology, consistently achieving very strong nodal support (1.0; 94–100%), with the exception of the pair *T. spiralis* + *T. nelsoni* in the maximum parsimony analysis (support: 64%). Third, to address this discrepancy, we studied separately the relationships of all encapsulated taxa using protein sequences.
encoded by 4,090 SCOs, and achieved strong support at all nodes for the three algorithms (1.00–99.100). Then, to assess the root relationship of sister species T. spiralis + T. nelsoni to non-encapsulated clades, we employed the same approach to resolve the relationships of three encapsulated (T. spiralis, T. nelsoni and T. patagoniensis) and one non-encapsulated taxon (T. papuae) using protein sequences encoded by 4,300 SCOs, and achieved the same outcome (that is, absolute support at all nodes). The final consensus tree constructed (Fig. 1a) represents the results from the first and second analyses (for all three algorithms). As expected, the dendrogram constructed from syntenic correlation values (Supplementary Data 2) was consistent in topology with this consensus tree and supported the encapsulated and non-encapsulated clades (Fig. 1a).

For the encapsulated taxa, the consensus tree shows that a lineage with T. spiralis + T. nelsoni is the sister to other taxa, which hierarchically include T. patagoniensis, and pairs of sister species (T. nativa + T6, T. murrelli + T9 and T. britovi + T8; Fig. 1a). These relationships differ from those inferred in previous studies using few ribosomal or mitochondrial DNA sequences, although some previously recognized sister species associations are concordant, and T. spiralis + T. nelsoni
and *T. patagoniensis* are relatively basal. At the time of initial phylogenetic assessment of *Trichinella*\textsuperscript{14}, sequence data were not available for all five *T. pseudospiralis* isolates (T4.1–T4.5) or *T. patagoniensis*. Here, for non-encapsulated taxa, the pair *T. papuae* + *T. zimbabwensis* is basal to all representatives of *T. pseudospiralis* investigated (that is, T4.1–T4.5); T4.4 from North America is basal to T4.5 from Tasmania and to all three isolates from Eurasia (T4.1–T4.3; Fig. 1a). Using the amino-acid sequences encoded by shared SCOs (n = 2,855), tree topology and nodal support values were similar to those achieved using whole-mitochondrial data sets\textsuperscript{21}, although these values were consistently high compared with previous analyses using small gene sets\textsuperscript{14,15}. 

Figure 1 | The evolution and biogeography of *Trichinella* taxa. (a) The phylogeny of all 12 currently recognized taxa of *Trichinella* based on analyses of amino-acid sequence data from shared SCOs (n = 1,284) employing Bayesian inference, ML and MP methods, with *Trichuris suis* and *A. suum* as outgroups; 1,042 and 747 are the numbers of orthologous gene groups, which are unique to encapsulated (red) and non-encapsulated (blue) *Trichinella* taxa, respectively. The topology of the trees constructed using each of these methods was the same; all nodes have absolute statistical support (1.00 or 100%), except for one node (**) in the ML analysis, where it was 99%. The grey bars on the nodes represent 95% confidence intervals for the estimate of taxa, respectively. The topology of the trees constructed using each of these methods was the same; all nodes have absolute statistical support (1.00 or 100%), except for one node (*) in the ML analysis, where it was 99%. The grey bars on the nodes represent 95% confidence intervals for the estimate of species branching time. *T. spiralis* (ISS195)\textsuperscript{19} shares the same phylogenetic position (**) as *T. spiralis* (ISS3). Host animals: suids (*Sus scrofa*) represent both the sylvatic and domestic porcine hosts (left); the reproductive potential of a particular *Trichinella* taxon in *S. scrofa*\textsuperscript{12} is indicated by the colour scale: white: not assessed; light grey; low; dark grey: medium; black: high. Other animals (right) represent examples of carnivorous sylvatic hosts in different geographic regions, including fox, lion, mountain lion, marsupial, crocodile and bird of prey, and the accidental human host. (b) Representation of genome-wide synteny among *Trichinella* taxa (same order as listed on the right in a). Genomic scaffolds (black) sharing at least 10 SCOs between *Trichinella* taxa are displayed. A purple line indicates a single SCO and a red line an inverted SCO. The numbers on the right indicate the genomic length in megabases (Mb). (c) Biogeography of *Trichinella* taxa proposed on the basis of known global (climate, extinctions and/or tectonic) events and diversification times (mya) for *Trichinella* taxa, estimated using a molecular clock approach. Encapsulated taxa: *T. spiralis*, T1; *T. nativa*, T2; *T. britovi*, T3; *T. murrelli*, T5; *T. nelsoni*, T7; *T. patagoniensis*, T12; and *Trichinella* genotypes T6, T8 and T9. Non-encapsulated taxa (infecting mammals, reptiles and/or birds): *T. pseudospiralis*, T4; *T. papuae*, T10; *T. zimbabwensis*, T11 (ref. 12). Geographic distributions of *Trichinella* taxa were reported by Pozio and Zarlanga\textsuperscript{11}. The embedded public domain world map image (https://commons.wikimedia.org/wiki/File:BlankMap-World6.svg) has been modified using the programs World map tool v.1.16 (http://law.nagoya-u.ac.jp/en/appendix/software/worldmap) and GIMP v.2.8 (https://www.gimp.org).
Here, for non-encapsulated taxa, the pair *T. papuae* + *T. zimbabwensis* is basal to all representatives of *T. pseudospiralis* investigated (that is, T4.1–T4.5); T4.4 from North America is basal to T4.5 from Tasmania and to all three isolates from Eurasia (T4.1–T4.3; Fig. 1a). Therefore, we have now been able to establish, for the first time, the phylogenetic relationships of all currently recognized *Trichinella* taxa, including five *T. pseudospiralis* representatives (that is, T4.1–T4.5) and the more recently discovered *T. patagoniensis*.

Divergence and biogeography. Divergence time analysis based on the nematode diversification estimate of 532–382 million years ago (mya)33 (1,000 SCOS) implied that *Trichinella* and *Trichuris suis* had a most recent common ancestor (MRCA) ~281 mya (95% credible interval: 384–204), and that the encapsulated and non-encapsulated *Trichinella* taxa shared an MRCA ~21 (28–15) mya (Fig. 1a), coinciding with the transition from Oligocene to Miocene34. Relative to a deep age for the *Trichinella* lineage, origin of a specific adaptation for encapsulation, associated with the radiation of nine taxa, occurred later in the evolutionary history of these nematodes. Subsequent diversification leading to extant species or species groups within the encapsulated and non-encapsulated clades is temporally circumscribed in the upper and uppermost Miocene during the Tortonian and Messinian periods35,36 commencing ~7 (9–5) and 10 (14–7) mya (Fig. 1a) and continuing into the Pliocene and Pleistocene34. Despite major methodological differences, our estimates are very consistent with those of Zarlanga et al.14 in postulating a geographic distribution for the MRCA of all *Trichinella* taxa in Eurasia. Further concordance is seen in estimates for initial divergence of respective clades for encapsulated and non-encapsulated forms during the mid-Miocene, coincidental with perturbations in temperate ecosystems before diversification of extant species (Fig. 1a,c); this scenario might relate to an early Miocene glaciation37, with low sea levels38 allowing a regional interchange of faunas linking Africa, Eurasia and North America39. The diversification of non-encapsulated *T. pseudospiralis* from the common ancestor of *T. papuae* and *T. zimbabwensis* (in poikilothersms; Fig. 1a,c) coincides with the Tibetan plateau uplift and climate change around 10–8 mya40 and the divergence between *T. papuae* and *T. zimbabwensis* (4.9–2.3 mya), possibly contemporaneously with climate change and the Plio-Pleistocene extinction of some crocodylomorph reptiles41. An avian host might explain the occurrence of *T. pseudospiralis* south of the Tibetan plateau.

Radiation of encapsulated *Trichinella* involves Eurasia, Africa, North America and South America. The occurrence of *T. nelsoni* (7.8–4.1 mya) and *T. britovi* + T8 (3.2–1.7 mya) on the African continent (Fig. 1c) follows temporally discrete and independent expansion events around 7.5 mya (Miocene), 4.5–4 and 3.5 mya (Pliocene) and 2.0 mya (Pleistocene)42. A separation of *T. nelsoni* + *T. spiralis* near the Miocene–Pleistocene boundary further establishes the basis for an independent association with hominins and humans based on ecology and later domestication of primary suid hosts for the latter species43. We hypothesize that the diversification of *T. britovi* and T8 took place in Africa (Fig. 1c) as a consequence of biogeographic barriers and changing environmental conditions, which is in accord with a previous suggestion by Pozio et al.44 Alternatively, the isolation and divergence of an ancestral population across Eurasia and Africa, leading to *T. britovi* and T8, with later secondary expansion, would account for the extensive geographic range of the former species10,14. We suggest that the loss of forests, formation of grasslands and the food scarcity for herbivores during the Miocene45,46 across Eurasia and Africa resulted in a massive expansion of an entirely new guild of predators/hunters that were able to follow scarce prey over vast distances in the later Miocene, Pliocene and Pleistocene47. Encapsulated *Trichinella* taxa initially expanded into North America across Beringia during a time frame deeper than 5 mya, when the landmass was a permanent geographic feature linking Eurasia and the Nearctic, and before the inception of Northern Hemisphere glaciations48. The distribution of *T. patagoniensis* is consistent with the initial emergence of the Panamanian Isthmus (~10 mya), as recently established49-51. New chronologies are also consistent with the complex nature of faunal assembly in the Neotropical region, providing a mechanism for geographic colonization by small cats, followed by extensive radiation in South America after 8 mya, before the Great American Interchange52. Interestingly, there is no current evidence for geographic colonization of South America after 3 mya by nearctic species of *Trichinella* (for example, *T. murrelli* or T6), coincidental with large felids, procyonids, canids and mustelids (Fig. 1c). Inception of Northern Hemisphere glaciation cycles and periodic emergence of the Bering land bridge after 2.5 and 2.0 mya53 led to independent episodes of geographic colonization and host-switching, driving patterns of isolation and genetic divergence (radiation) of *Trichinella*, linking Eurasia and the Nearctic.48 Diversification of Eurasian/Nearctic sister species, including *T. nativa* + T6, *T. murrelli* + T9 and *T. pseudospiralis* in Northern America, reflects an intricate history in response to climate variation and habitat change, facilitating independent events of biotic expansion among carnivorans and other mammalian assemblages, including ursids, canids and mustelids primarily from Eurasia into North America (Fig. 1c). In addition, we hypothesize that T9 diverged from a common ancestor with *T. murrelli* before geographic colonization of the New World, or following isolation across Bering Strait during a glacial maximum (Fig. 1c). In the absence of a fossil record for *Trichinella*, it would be useful to determine the sequence of at least one *Trichinella* taxon from an extinct, infected vertebrate (for example, carnivorans) as a reference in time to assist future studies. The chronological and spatial history of *Trichinella* is described by episodic or cyclical patterns of independent geographic and host colonization on global and regional scales, the extensive development of mosaic faunal assemblages consistent with an integrated history for taxon pulses and ecological fitting mediated by climate and habitat variation over the late Tertiary48,54,55.

Parasite-host adaptation. The significantly higher GC content in the genomes and coding regions of encapsulated compared with non-encapsulated taxa (Kolmogorov–Smirnov (KS) test: *P* values = 3.801 × 10⁻⁴ and 7.466 × 10⁻⁴, respectively; effect size = 1) suggests that these two *Trichinella* clades may have adapted differently to varying environmental stresses (for example, temperature), host immune attacks and/or body temperatures (reptiles versus birds and mammals). We also propose that the intracellular lifestyle and associated bottleneck have resulted, throughout evolution, in a considerable reduction of genome size (49 Mb) and lower GC content (33%) in both encapsulated and non-encapsulated *Trichinella* taxa compared with extracellular relatives such as *Trichurus suis* (78.5 Mb; 44%)22, which accords with observations for selected pathogens56,57. Considering the morphological differences between encapsulated and non-encapsulated *Trichinella* taxa in host muscle cells12, we focused on exploring ES molecules likely to be associated with parasite–host interactions or host cell modification. Although more than half (52.9%) of genes encoding putative ES proteins could not be annotated (Supplementary Data 1), we identified seven SCOS (groups (GRPs) 2,426, 4,153,
Interestingly, the average GC content of differentially transcribed genes (KS test: P \text{value} = 3.059 \times 10^{-7}; \text{effect size} = 1) than that of all genes of all Trichinella taxa. In addition, the significantly lower AT content of differentially transcribed genes (KS test: P \text{value} = 4.076 \times 10^{-2}; \text{effect size} = 0.67) in non-encapsulated taxa suggests a distinct adaptation of genes to the muscle cell compared with encapsulated taxa.

Interestingly, we found expansions in two orthologous groups in non-encapsulated taxa. The first (GRP 149) represents a serine protease precursor with two trypsin-like domains (TssERP and AF331156), which is expanded in all non-encapsulated taxa (Supplementary Data 3) and appears to play a vital role in larval feeding and/or moulting. The second group represents a multicystein-like domain protein (MCD-1) encoded by six gene copies in T. papuae and two copies in T. pseudospiralis (T4.5) from Australia and T. zimbabwensis compared with one copy in all other Trichinella taxa. This protein contains three repeated domains, each with similarity to family 2 cystatins, but lacking critical consensus sites for cysteine protease inhibition. In this respect, the domain organization is similar to that of mammalian kininogens and a known six-domain cystatin from F. hepatica. We propose that MCD-1 might function as a cytokine antagonist by binding to transforming growth factors (TGF)-\beta1 and \beta2 in a manner similar to fetuin\(^{88}\), a proposal that is consistent with previous findings for T. spiralis and evidence of low-level expression of TGF-\beta in the epithelium of the jejunum of infected mice\(^{69}\).

Certainly, family 2 cystatins secreted by other parasitic nematodes have known roles in immune evasion, including the inhibition of proteases involved in antigen presentation and modulation of cytokine responses\(^{70}\). Although MCD-1 is unlikely to function as a typical cystatin, it is expressed at the parasite–host interface and might modulate host immune responses or enable immune evasion. An expanded set of MCD-1s might allow some non-encapsulated Trichinella taxa to suppress immune responses better than encapsulated taxa, possibly facilitating dissemination into a wider range of vertebrate hosts\(^{12}\).

Discussion
Using Illumina-based sequencing and bioinformatics, we assembled draft nuclear genomes representing all 12 recognized taxa of Trichinella, including five geographic isolates of T. pseudospiralis. Using extensive amino-acid sequence data sets derived from all shared SCOs from these nuclear genomes and/or outgroup taxa, we were able to reconstruct the phylogeny of Trichinella. In a previous study, Zarlanga et al.\(^{14}\) had proposed the phylogenetic relationships of Trichinella species and genotypes using small DNA sequence data sets. At the time, these authors provided comprehensive interpretations of the findings and concluded that post-Miocene expansion, colonization and host-switching drove speciation among extant members of the genus Trichinella; although their study was very informative and interesting, the resolution of some clades and the positions of some of the taxa, such as T. nativa and T6 as well as T. murrelli and T9, did not always appear to be well supported statistically by the data presented, when outgroups (mermithids and Trichuris spp.) were included in the analyses. The reason for the limited resolution of some relationships was likely because of the use of sequence data set representing only selected genetic loci (nuclear small subunit and second internal transcribed spacer; mitochondrial large subunit and cytochrome c oxidase subunit I)\(^{14}\). In addition, it appears that some of the species selected as outgroups might have been too distant to achieve a robust phylogeny using all of the concatenated sequence data in the analyses.

Here we utilized an extensive amino-acid sequence data set representing all SCOs (concatenated alignment over 597,495 amino-acid positions) originating from all 16 genomes representing all members of the Trichinella complex, including all known geographic isolates of T. pseudospiralis and T. patagoniensis, not previously available\(^{14,15}\), as well as Trichuris suis and A. suum (outgroups) in the analyses. By contrast, in a previous study\(^{21}\) we could not use Trichuris or Ascaris as outgroups because of excessive sequence divergence in predicted mitochondrial proteins (for example, NAD4 and NAD6) encoded by some genes, such that an unambiguous alignment of homologous characters was impossible; these outgroups therefore had to be excluded from the analyses of mitochondrial data sets, as their inclusion would have led to erroneous results and interpretation. In the present study, the definition of 1,284–2,855 SCOs among all Trichinella taxa (with or without outgroups) allowed, employing an iterative, stepwise approach, the reconstruction of a robust phylogeny utilizing three independent tree-building methods. In accordance with previous investigations\(^{14,21,68,69}\), the present analyses showed that the encapsulated taxa grouped separately from non-encapsulated taxa. Importantly, we were also able to resolve, for the first time, the phylogenetic positions of T. spiralis + T. nelsoni and T. murrelli + T9, five representatives of T. pseudospiralis and T. patagoniensis in relation to all other taxa, with strong (99–100%) statistical support. The diversification times of Trichinella matched well with known historical global events and allowed the biogeography of all taxa to be reconstructed. This biogeography clearly supports the notion that encapsulated and non-encapsulated taxa frequently occur in sympatry and in the context of faunal assembly, driven strongly by events of geographic and host colonization, involving complex spatial and chronological mosaics linking evolutionary
and ecological time. Nonetheless, in the future, extensive sampling of Trichinella taxa from around the world will be required to explore, in depth, population genetic structures to reassess the present phylogenetic and biogeographic reconstruction, and to attempt to identify the host origin of ancestral forms of T. spiralis, the principal causative agent of human trichinellosis. In conclusion, although the present study focused sharply on fundamental phylogenetic and biogeographic aspects, the genomic and transcriptomic resources created here will substantially accelerate many fundamental and applied areas of Trichinella/trichinellosis research. In addition, the genome-wide approach that we employed should have applicability to a wide range of parasites and other metazoon organisms.

**Methods**

**Production and procurement of Trichinella taxa.** Sixteen samples (isolates) of first-stage larvae (L1s) representing all 12 recognized species and genotypes of Trichinella were produced at the International Trichinellosis Reference Center (http://www.iss.it/site/Trichinella/), Istituto Superiore di Sanità (ISS), Rome, Italy. The 16 samples included the following: one sample of each T. spiralis (code: ISS3), T. nativa (ISS10), T. britovi (ISS120), T. murrelli (ISS147), T. nelsoni (ISS37), T. patagoniensis (ISS2496) as well as Trichinella genotypes T6 (ISS334), T8 (ISS272) and T9 (ISS409); five samples representing distinct geographic isolates of T. pseudospiralis (ISS13, ISS588, ISS176 and ISS141 for T4.1, T4.2, T4.3, T4.4 and T4.5, respectively); and one sample of each T. pacuense (ISS1980) and T. zimbabwensis (ISS1029; cf. Tables 1 and 2). These samples were individually produced in female CD1 mice. L1s were recovered from host muscles by pepsin digestion at 40 °C for 30 min, sedimented, washed extensively in physiological saline and suspended in 90% ethanol for storage. Each sample, which represented a packed volume of 50 μl of L1s, was snap-frozen in liquid nitrogen and kept at −80 °C until nucleic acid isolation.

**RNA-seq and transcriptome assembly.** Total RNA was isolated separately from L1s of each isolate of Trichinella employing the TriPure isolation reagent (Roche Molecular Biochemicals). Packed volumes of 20–50 μl were used, equating to thousands of larvae. RNA yields were estimated spectrophotometrically (NanoDrop 1000), and the integrity of RNA was verified using the BioAnalyzer. Total RNA was isolated separately employing the TriPure isolation reagent (Roche Molecular Biochemicals). Packed volumes of 20–50 μl were used, equating to thousands of larvae. RNA yields were estimated spectrophotometrically (NanoDrop 1000), and the integrity of RNA was verified using the BioAnalyzer. RNA-seq was conducted using an established method and relevant data summarized (cf. Supplementary Tables 3 and 4). De novo assemblies were performed using a software pipeline, incorporating the programme Trimmomatic v.0.27 (http://www.usadellab.org/cms/?page=trimmomatic) for read quality-filtering, Kmer v.1.1 (ref. 73) for the reduction of high and low coverage reads, and Velvet v.1.2.07 (ref. 74) and Oases v.0.2.08 (ref. 75) for sequence assembly.

**Other methods.** Genomic sequencing and assembly, prediction of repetitive elements, deep sequencing of protein-coding genes, functional annotation, phylogenetic and divergence time analyses, synteny, GC content and differential transcription analyses are described in Supplementary Methods.

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Acknowledgements

Funding from the Australian Research Council, the National Health and Medical Research Council (NHMRC) of Australia, Yourgene Bioscience and Melbourne Water Corporation is gratefully acknowledged (R.B.G. et al.). This project was funded in part through DG SANCO of the European Commission and Instituto Superiore di Sanita`, Rome (E.P.). This project was also supported by a Victorian Life Sciences Computation Initiative (grant number VR0007) on its Peak Computing Facility at the University of Melbourne, an initiative of the Victorian Government (R.B.G.). N.D.Y. holds an NHMRC Early Career Research Fellowship. P.W.S. thanks the Howard Hughes Medical Institute (HHMI) and the National Institutes of Health (NIH). We thank the staff of Yourgene Bioscience for their contributions. We would specifically like to acknowledge all scientists who developed the programmes used in this study. Given restrictions on the number of articles that could be cited, we were unable to include all original articles in the Methods section. Instead, we have included links to respective websites. P.K.K. is the recipient of a scholarship (STRAPA) from the University of Melbourne.

Author contributions

P.K.K., N.D.Y. and R.B.G. wrote the paper with inputs from all other co-authors including G.L.R. and E.P., and colleagues who produced parasite material. G.L.R. prepared all nucleic acids and undertook laboratory-based evaluations. P.K.K. undertook the genome assembly and all bioinformatic analyses, with support from N.D.Y. and R.B.G. A.V.K. provided initial support with phylogeny. A.H. provided support with structural modelling. E.P.H. explored aspects of historical biogeography and faunal assembly. With support from G.L.R. and E.P., R.B.G. led the project.

Additional information

Accession codes: This Whole Genome Shotgun project has been deposited in the NCBI BioProject database with accession code PRJNA257433. The project includes: all the genome assemblies under accession codes JYDH00000000 to JYDW00000000; raw Illumina read sets for the genomic DNA; raw reads for RNA-seq under accession codes SRS672137, SRS698754, SRS900495, SRS906998, SRS906999 to SRS906994, SRS906996, SRS906998 to SRS907002, SRS908054 and SRS908055; and assembled transcriptomes for the 12 Trichinella taxa under accession codes GEBM0000000 to GEBP000000, GEBR000000000 to GEBZ00000000, GECIA0000000 and GECIC0000000. The versions described in this paper are JYDH10000000 to JYDW10000000 for the genomes, and GEBM010000000 to GEBP010000000, GEBR010000000 to GEBZ010000000, GECIA0100000 and GECIC0100000 for the transcriptomes.

Supplementary Information accompanies this paper at http://www.nature.com/naturecommunications

Competing financial interests: The authors declare no competing financial interests.

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How to cite this article: Korshonen, P. K. et al. Phylogenomic and biogeographic reconstruction of the Trichinella complex. Nat. Commun. 7:10513 doi: 10.1038/ncomms10513 (2016).

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