The stromal cell–derived factor-1α/CXCR4 ligand–receptor axis is critical for progenitor survival and migration in the pancreas

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The SDF-1α/CXCR4 ligand/chemokine receptor pair is required for appropriate patterning during ontogeny and stimulates the growth and differentiation of critical cell types. Here, we demonstrate SDF-1α and CXCR4 expression in fetal pancreas. We have found that SDF-1α and its receptor CXCR4 are expressed in islets, also CXCR4 is expressed in and around the proliferating duct epithelium of the regenerating pancreas of the interferon (IFN) γ–nonobese diabetic mouse. We show that SDF-1α stimulates the phosphorylation of Akt, mitogen-activated protein kinase, and Src in pancreatic duct cells. Furthermore, migration assays indicate a stimulatory effect of SDF-1α on ductal cell migration. Importantly, blocking the SDF-1α/CXCR4 axis in IFNγ–nonobese diabetic mice resulted in diminished proliferation and increased apoptosis in the pancreatic ductal cells. Together, these data indicate that the SDF-1α–CXCR4 ligand receptor axis is an obligatory component in the maintenance of duct cell survival, proliferation, and migration during pancreatic regeneration.

Introduction
Chemokines are a superfamily of small secreted proteins known initially for their role in leukocyte trafficking (Luster, 1998; Gale and McColl, 1999; Rossi and Zlotnik, 2000). Chemokines have received much attention for their involvement in the regulation of HIV infection, proliferation, and mobilization of hematopoietic stem cells, fetal development, and regulation of angiogenesis (Kim and Broxmeyer, 1999).

Although most chemokines and receptors have overlapping binding specificity with other family members, SDF-1α and its receptor CXCR4 bind only each other (Kim and Broxmeyer, 1999). CXCR4 has also been the focus of numerous papers because it is a coreceptor for the entry of HIV into T cells (Feng et al., 1996). SDF-1α is involved in the migration of hematopoietic cells to the marrow, and hematopoietic precursors from the bone marrow via the circulation into peripheral tissues (Aiuti et al., 1997; D’Apuzzo et al., 1997). A comprehensive study of the expression of SDF-1α and CXCR4 from gastrulation to organogenesis in the mouse embryo provides evidence of the continuous involvement of the SDF-1α/CXCR4 axis during embryogenesis (McGrath et al., 1999). Furthermore, disruption of the genes for SDF-1α or its receptor results in late embryonic lethality. Importantly, the SDF-1α–deficient mouse and the corresponding CXCR4 mutants are the only known chemokine/chemokine receptor mutants that display embryonic lethality (Murphy et al., 2000). Genetically deficient embryos display severe defects in their gastrointestinal vasculature, cerebellar neuron migration, cardiac ventricular septal closure, B cell development, and hematopoietic bone marrow colonization (Nagasawa et al., 1996; Ma et al., 1998; Tachibana et al., 1998; Zou et al., 1998). The extensive consequences observed in different organ systems in the SDF-1α and CXCR4 knockout mice indicate that the SDF-1α–CXCR4 axis is an essential component of differentiation of numerous tissues.

The terminal differentiation of the pancreatic endocrine cells from epithelial progenitor cells occurs during their mi-

Abbreviations used in this paper: C-10, small inducible cytokine A6; Eotaxin, small inducible chemokine A 11; IP-10, IFN-γ–inducible protein 10 kD; MCP, monocyte chemoattractant protein; MIG, monokine induced by γ IFN; MIP, macrophage inflammatory protein; NOD, nonobese diabetic; PDX1, pancreatic duodenal homeobox 1; RANTES, regulated on activation normal T-cell expressed and secreted; TCA-4, thymus-derived chemotactic agent 4.
migration from the duct wall into primitive islet-like structures (Slack, 1995). The process involves the remodeling of the cell surface and a change in the adhesive properties of these cells as they migrate (Cirulli et al., 2000). The signals governing this migration are not fully defined. Based on the demonstrated involvement of the SDF-1α-CXCR4 pair in the migration and proliferation of hematopoietic stem cells, we hypothesized that this ligand–receptor pair may be important during pancreatic endocrine cell development. However, because the genetically deficient mice suffer from widespread midgestational defects, it is difficult to address the requirement for CXCR4/SDF-1α during embryonic development. Therefore, we sought to determine the role of CXCR4 ligation during pancreatic islet regeneration.

In transgenic mice in which the cytokine IFN-γ is expressed under the control of the insulin promoter, the pancreas displays remarkable ductal hyperplasia and regeneration of new islets (Sarvetnick et al., 1988; Gu and Sarvetnick, 1993, 1994). Previous work suggests the pancreatic islet regeneration proceeds through the same intermediates as does islet formation during ontogeny (Gu and Sarvetnick, 1993, 1994; Kritzik et al., 1999, 2000). We have found that spontaneous islet regeneration in the IFN-γ transgenic mouse recapitulates the pancreatic developmental program in adults (Kritzik et al., 1999, 2000).

In this work, we tested the hypothesis that CXCR4 ligation is required for the differentiation of pancreatic islets during regeneration. Our results strongly support an essential role for the SDF-1α–CXCR4 pair during IFN-γ induced pancreatic regeneration.

Results

Chemokine expression in the IFN-γ-nonobese diabetic (NOD) pancreas

The cytokine IFN-γ has been shown to induce the expression of a number of chemokines, specifically IFN-γ-inducible protein 10 kD (IP-10), monokine induced by γ IFN (MIG), and IFN-inducible T cell α chemoattractant in different tissues (Luster et al., 1985; Kaplan et al., 1987; Luster and Ravetch, 1987; Gottlieb et al., 1988; Ransohoff et al., 1993; Cassatella et al., 1997; Sauty et al., 1999). We hypothesized that IFN-γ-induced chemokines may have an effect on the migration of pancreatic progenitors to their appropriate niche where they could differentiate into the endocrine cells of the pancreas. To investigate the induction of chemokines by IFN-γ expression in the pancreas, we used RNase protection assays. The chemokines that were induced by the IFN-γ transgene compared with nontransgenic NOD mice were: small inducible cytokine A6 (C-10); IP-10; macrophage inflammatory protein (MIP)-2; regulated on activation normal T-cell expressed and secreted (RANTES; Fig. 1 A); MIG; monocyte chemoattractant protein (MCP)-5; SDF-1α; thymus-derived chemotactic agent 4 (TCA-4); and small inducible chemokine A11 (Eotaxin; Fig. 1 B). In the NOD pancreas, five of the chemokines were detected: C-10,
Localization of SDF-1α and CXCR4 expression in the pancreas

Of the chemokines and chemokine receptors that we found to be induced by IFNγ, the SDF-1α/CXCR4 pair has been shown to be an important regulator of cell migration, proliferation, and embryonic development. Next, we performed histological studies to determine whether the immunolocalization of SDF-1α and CXCR4 was consistent with their involvement in epithelial migration in the transgenic pancreas.

In the NOD mouse pancreas, SDF-1α and CXCR4 expression was detected in islets by immunofluorescence (Fig. 3 A). In addition, SDF-1α expression was confirmed by Western blot analysis of proteins expressed by isolated pancreatic islets (unpublished data). The expression of SDF-1α in NOD mice appeared constitutive because in vitro treatment of the islets by IFNγ (1,000 U/ml) for 24 h did not augment SDF-1α expression, as determined by Western blotting (n = 2; unpublished data). In the IFNγNOD mouse, SDF-1α staining was localized to cells within the islet mass (Fig. 3 B and C). In the duct epithelium, frequent cells, which express only CXCR4 (Fig. 3 E, green), and occasional cells, which coexpress CXCR4 and SDF-1α (Fig. 3 E, yellow), were observed. Thus, the majority of duct cells express CXCR4, suggesting that these cells may be migrating toward the SDF-1α-expressing newly forming islets.

Figure 2. Expression of chemokine receptors in adult regenerating pancreas. (A) Expression of CCR group of chemokine receptors in whole pancreas from IFNγNOD transgenic mice (lane a) and NOD (lane b) controls. The RPA probe for CCR is shown in lane c. (B) Expression of CXCR group of chemokine receptors in IFNγNOD (lane a) and NOD (lane b) pancreas was determined as described in A. The RPA probe for CXCR is shown in lane c. As with the chemokine expression experiments a second experiment yielded similar findings. Gel positions of chemokine receptors are indicated with arrows.

Chemokine receptor expression in the IFNγNOD pancreas

Chemokines induce their effects by binding G protein–coupled receptors. We used probes for chemokine receptors to determine their expression in NOD and IFNγNOD mouse pancreas. The chemokine receptors induced in the IFNγNOD mouse pancreas were CCR6, CCR7, CCR1, CCR3, CCR5, CXCR5, DARC, CX3CR1, and CXCR4 (Fig. 2, A and B). In the NOD only CXCR4, CXCR5, and DARC were detected. The relative expression of the chemokine receptors in the transgenic pancreas compared with the NOD pancreas is shown in Table I. As determined by densitometry, C-10 expression was elevated 12-fold in the IFNγ transgenic pancreas compared with the NOD pancreas. The expression of MIG was enhanced approximately sixfold in the transgenic pancreas. SDF-1α expression was elevated threefold, and TCA-4 and Eotaxin were expressed at five- and fourfold higher levels than in the NOD pancreas, respectively. Of the CHK2 chemokines expressed, IP10, C-10, and RANTES expression were modest compared with the NOD pancreas.

Table I. Chemokine and chemokine receptor induction by the IFNγ transgene in the NOD mouse pancreas

| Chemokines | NOD | IFNγNOD |
|------------|-----|---------|
| C-10       | 1   | 12X     |
| MIG        | 1   | 6X      |
| SDF-1α     | 1   | 3X      |
| TCA-4      | 1   | 5X      |
| Eotaxin    | 1   | 4X      |
| Chemokine receptors | | |
| CXCR4      | 1   | 4X      |
| CXCR5      | 1   | 2.5X    |
| DARC       | 1   | 2.5X    |
| MIG        | 1   | 6X      |
| SDF-1α     | 1   | 12X     |
| TCA-4      | 1   | 5X      |
| Eotaxin    | 1   | 4X      |

MIG, SDF-1α, TCA-4, and Eotaxin. The relative expression of chemokines in the transgenic mouse pancreas compared with the NOD pancreas is shown in Table I. As determined by densitometry, C-10 expression was elevated 12-fold in the IFNγ transgenic pancreas compared with the NOD pancreas. The expression of MIG was enhanced approximately sixfold in the transgenic pancreas. SDF-1α expression was elevated threefold, and TCA-4 and Eotaxin were expressed at five- and fourfold higher levels than in the NOD pancreas, respectively. Of the CHK2 chemokines expressed, IP10, C-10, and RANTES expression were modest compared with the NOD pancreas. The expression of MIG was enhanced approximately sixfold in the transgenic pancreas. SDF-1α expression was elevated threefold, and TCA-4 and Eotaxin were expressed at five- and fourfold higher levels than in the NOD pancreas, respectively. Of the CHK2 chemokines expressed, IP10, C-10, and RANTES expression were modest compared with the NOD pancreas.
play CXCR4 (Fig. 4 B) expression. In the embryonic pancreas, both SDF-1α (Fig. 4 C, green) and CXCR4 (Fig. 4 D, green) were found to be frequently coexpressed with insulin (red). CXCR4 staining was often adjacent to insulin expressing cell clusters (Fig. 4 D). Double immunofluorescent staining of CXCR4 and SDF-1α (Fig. 4, E and F) revealed that some cells in the primitive islet structures express SDF-1α (red) and duct cells expressed CXCR4 (Fig. 4, D and E, green). However, the two were often colocalized, suggesting that this chemokine may be involved in the recruitment of cells from ducts into the developing islet cell clusters.

**In vitro stimulation of cell migration by SDF-1α**

To test the hypothesis that SDF-1α can stimulate the migration of ductal progenitor cells of the regenerating pancreas, we performed in vitro migration assays using cells isolated from the pancreatic ductal network of regenerating IFNγ-NOD.scid mice. We measured the effect of SDF-1α on the migration of those cells using native or type 1 collagen–coated membrane inserts. Interestingly, the stimulatory effect of SDF-1α was most pronounced in the absence of collagen. 100 and 300 ng/ml of SDF-1α enhanced duct cell migration by 52 and 93% (P < 0.02), respectively. In the presence of collagen, chemotaxis was increased by ~38% at both 100 and 300 ng/ml concentrations of SDF-1α (P < 0.001; Fig. 5 A). Interestingly, collagen coating of the inserts greatly enhanced the apparent basal and SDF-1α induced migration of the ductal cells (Fig. 5, D and E). This could be because the freshly isolated cells exhibit increased survival in the presence of the matrix coating. Alternatively, collagen may increase the adhesion of these cells, accounting for the reduced ability of SDF-1α to induce chemotaxis. Together, the data demonstrate that ductal cells from the regenerating pancreas migrate in response to SDF-1α and their migration in vitro is clearly modulated by the presence of ECM.

**Effect of CXCR4 neutralization on ductal cell proliferation in the IFNγ-NOD pancreas**

We have hypothesized that cell migration is critical for the regenerative process to occur. Because we observed the CXCR4 receptor induced in the expanding epithelium of the transgenic pancreas, and because in vitro studies demonstrated the ability of SDF-1α to induce transgenic ductal cell migration, we asked whether this receptor–ligand interaction participated in the regenerative process. Therefore, we determined the outcome of blocking SDF-1α binding by treating the IFNγ transgenic mice with a CXCR4 neutralizing antibody (Gonzalo et al., 2000). For this analysis, two groups of 12-wk-old IFNγ-NOD mice were intravenously treated with 20 µg/mouse CXCR4 blocking antibody (n = 8) or rabbit IgG (n = 7) every third day for 2 wk. Normally, 12-wk-old transgenic mice display strong proliferative activity, as evidenced by BrdU incorporation, with the development of elaborate ducts and formation of new islets in the pancreas (Gu and Sarvetnick, 1993). We stained sections from different levels of the pancreas of each mouse and counted the BrdU-positive duct cells, as well as the total number of duct cells. The CXCR4 antibody treatment resulted in a 42% de-
crease in the ratio of BrdU labeled duct cells to the total number of duct cells in the IFN\textgamma{}NOD pancreas compared with IgG-treated control mice (CXCR4 = 19.8%; IgG controls = 11.41%; Fig. 6). This significant (P < 0.005) decrease in the proportion of BrdU-positive duct cells indicates that the SDF-1\textalpha{}/CXCR4 axis has a critical role in the net expansion of the regenerating duct epithelium.

The effect of CXCR4 neutralization on survival in IFN\textgamma{}NOD pancreata
Remodeling and directed movement of tissue in development involves regulation of both cell proliferation and programmed cell death (Zakeri and Lockshin, 2002). Apoptotic cell death has also been demonstrated to be involved in the homeostatic regulation of hematopoiesis (Peters et al., 1998). Indeed, the

Figure 4. SDF-1\textalpha{} and CXCR4 expression in embryonic NOD pancreas. Panel A illustrates SDF-1\textalpha{} expression by DAB staining in primitive islet structures in the fetal pancreas. Ductal areas are clear of SDF-1\textalpha{} staining. Panel B depicts CXCR4 expression in primitive islets and also in some ductal cells (d, duct; i, islet). (C) Representative double immunofluorescent images of SDF-1\textalpha{} (green) and insulin (red) reveal extensive colocalization (yellow) in the E18 pancreas with a population of cells expressing SDF-1\textalpha{} alone. (D) Insulin (red) and CXCR4 (green) immunofluorescent staining demonstrating that some cells display coexpression of CXCR4 and insulin (yellow), with a significant number of cells staining only for CXCR4. (E) Double immunofluorescent staining of CXCR4 (green) and SDF1-\textalpha{} (red) demonstrates that contiguous cells in the primitive islet clusters can express the ligand, the receptor, or both (yellow). (F) A ductal region surrounded by developing islet clusters magnified from E. Arrows point to duct cells. Bars, 25 \mu m.

Figure 5. SDF-1\textalpha{} stimulates in vitro migration of pancreatic ductal cells. (A) Cell migration was measured in the presence and absence of collagen coating. Each bar represents either basal migration or fold stimulation from basal in a total of six membranes from three experiments (mean ± SEM); P < 0.02 for migration on native membranes and P < 0.001 on collagen-treated membranes by analysis of variance. B and C are two representative fields of (B) basal and (C) SDF-1\textalpha{}-stimulated (300 ng/ml each) ductal cells on uncoated membranes. D and E depict two fields of (D) basal and (E) SDF-1\textalpha{}-stimulated ductal cells migrating on collagen I-coated membranes.
decreased incorporation of BrdU in the CXCR4-treated mice could reflect either decreased duct cell replication or increased turnover of the duct cell population. Therefore, we decided to assess cell turnover by measuring apoptosis in the pancreatic epithelial duct cells, quantitating fragmented DNA using the TUNEL method in the CXCR4 antibody–treated and control IgG–treated mice. Treatment of IFNγ/NOD transgenic mice with the CXCR4 blocking antibody resulted in a fourfold increase in the number of pancreatic ductal nuclei displaying fragmentation (2.0%; \(P < 0.003\); Fig. 7). The number of duct cells undergoing apoptosis was 0.5% in control IFNγ/NOD mice treated with rabbit IgG. Our results demonstrate a destabilizing effect of blocking CXCR4–SDF-1 interactions on the survival of duct cells in vivo.

**The effect of CXCR4 neutralization on the proportion of pancreatic progenitors present in ductal cells**

We postulated that the decline in duct cell replication, and augmentation in apoptosis in response to CXCR4 neutralizing antibody could affect the proportion of pancreatic progenitor cells in the regenerating pancreas. The pdx1 gene is a key regulator of pancreatic development and insulin expression on β cells (Jonsson et al., 1994; Offield et al., 1996). We have demonstrated previously that the proliferating ducts in the IFNγ transgenic mouse pancreas express pancreatic duodenal homeobox 1 (PDX1; Kritzik et al., 1999).

Treatment with the CXCR4 neutralizing antibody \((n = 4)\) led to a 50% decrease in the number of ductal cells expressing PDX1 compared with IgG-treated controls \((n = 4; P < 0.03;\) Fig. 8). These results suggest that the SDF-1α–CXCR4 interaction may be affecting the size of the population of ductal cells that could differentiate into insulin-producing cells. Interestingly, the inhibition of the SDF-1α–CXCR4 axis impacted the average diameter of ducts. Although the actual number of duct cells in the treatment and control groups were not statistically significantly different (not depicted), the number of duct cells residing in smaller ducts were increased significantly in the CXCR4-treated group \((n = 7)\) compared with IgG-treated controls \((n = 7; P < 0.001;\) Fig. 9). These data are consistent with a role for SDF-1α and CXCR4 in the initiation of expansion and recruitment to facilitate islet differentiation.

**Stimulation of freshly isolated ductal cells with SDF-1α induces tyrosine phosphorylation of Src, MAPK, and Akt (protein kinase B)**

SDF-1α has been shown to transduce its actions by binding its G protein–coupled seven transmembrane receptor CXCR4 and activating p44/42 MAPK, PI3-Kinase, and Akt (Ganju et al., 1998; Majka et al., 2000; Vlahakis et al., 2002;
Floridi et al., 2003). The Src family of tyrosine kinases have been reported to be involved in the activation of migration in different cell types (Datta et al., 1997; Coffer et al., 1998; Lowell and Berton, 1998; O’Laughlin-Bunner et al., 2001; Inngejerdingen et al., 2002). Therefore, we wanted to determine if SDF-1α could stimulate the phosphorylation of MAPK, Akt, or Src, signaling proteins that are potentially associated with the proliferation, survival, or migration of ductal cells. Cells isolated from the pancreatic ducts of IFNγ-NOD mice were serum-starved overnight, and stimulated with 100 or 300 ng/ml SDF-1α or 10 ng/ml EGF for 5 min. Fig. 10 A shows SDF-1α and EGF-stimulated phosphorylation of Src, MAPK, and Akt in vitro. EGF-stimulated MAPK and Akt phosphorylation in these cells. SDF-1α-stimulated Src phosphorylation about twofold at 300 ng/ml, where we observed a striking effect of the chemokine on migration. Our findings suggest a potential role for Src in the regulation of migration by SDF-1α in the duct progenitor cells of the regenerating pancreas.

The time courses of stimulation of the phosphorylation of signaling proteins vary depending on the specific ligand. Therefore, we examined the time courses of SDF-1α stimulation of MAPK and Akt phosphorylation in freshly isolated ductal cells. Cells were isolated and serum-starved overnight as indicated in the previous paragraph, and stimulated with 300 ng/ml SDF-1α for 0, 2, 5, 10, 30 or 60 min. Fig. 10 B shows the time course of the phosphorylation of MAPK (p42, p44 ERK1, and ERK2) and Akt. SDF-1α-stimulated ERK1 and ERK2 phosphorylation was first seen at 2 min and reached a peak at ~10 min. The stimulation of Akt phosphorylation by SDF-1α was not evident until 10 min, at which point it was very potent (~30-fold by densitometry). The dramatic phosphorylation of Akt and to a lesser extent MAPK and Src, in response to SDF-1α in duct cells of the IFNγNOD transgenic mice, indicate a potential involvement of these signaling proteins in the regulation of migration, proliferation, and survival of the pancreatic progenitor population.

**Discussion**

Our focus was to identify chemokines that are involved in cell migration within the regenerating pancreas. SDF-1α and CXCR4 expression, as determined by RNase protection
assays, were greatly enhanced in the IFNγ/NOD mouse pancreas. Our experiments revealed expression of this chemokine–chemokine receptor pair in the regenerating IFNγ/NOD pancreas and in the developing islet clusters of NOD embryos. In vivo blocking experiments demonstrated that the CXCR4 receptor is an obligatory component of pancreatic regeneration. A concomitant augmentation in apoptosis in response to CXCR4 antibody treatment indicated that an antiapoptotic effect of CXCR4 ligation in the ductal epithelial cells is a component of the enhanced ductal expansion in the regenerating pancreas. Furthermore, we report a direct stimulatory effect of SDF-1α on the migration of cells isolated from the ducts of the regenerating adult pancreas. Therefore, our data suggests that the IFNγ induced elevation in the expression of SDF-1α in the duct epithelium of the pancreas results in the chemotactic migration of CXCR4 expressing cells, in response to the SDF-1α gradient.

Importantly, neutralization with CXCR4 antibody elicited a fourfold increase in the proportion of apoptotic ductal cells. The role of CXCR4 in the regulation of apoptosis is context dependent. CXCR4 involvement in the stimulation of apoptosis by HIV envelope proteins in CD4+ T cells has been an area of extensive study (Herbein et al., 1998; Biard-Piechaud et al., 1999; Colamussi et al., 2001; Yao et al., 2001; Arthos et al., 2002). Similarly, CXCR4 regulation of the apoptotic effect of HIV coat proteins on neurons of the neocortex has also been reported (Corasaniti et al., 2001). In contrast, SDF-1α has been reported to promote survival by inhibiting apoptosis in hematopoietic progenitor cells (Lataillade et al., 2002). Furthermore, in fetal thymus organ culture, SDF-1α enhanced viability of serum-depleted cells in culture by down-regulating the pro-apoptotic bax protein and up-regulating the antiapoptotic bcl-2 protein (Hernandez-Lopez et al., 2002). Interestingly, transgenic mice expressing SDF-1α under a Rous sarcoma virus promoter display enhanced spleen and bone marrow myelopoiesis in vivo (Broxmeyer et al., 2003). Myeloid progenitors from the SDF-1α transgenic mice also exhibit prolonged survival in the absence of growth factors in vitro compared with progenitors from wild-type mice. In our pancreatic regeneration model, it appears that blocking the CXCR4 receptor resulted in an augmentation of apoptosis, indicating a role for the CXCR4 receptor in promoting survival of the ductal cell precursor pool in the regenerating ducts, similar to its role in hematopoietic progenitor cells. Our observation of diminished numbers of ductal cells expressing PD1, a critical pancreatic progenitor marker, in mice treated with CXCR4 neutralizing antibody is consistent with the concurrent enhanced programmed cell death.

The endocrine progenitor cells in the ductal epithelium of the IFNγ/NOD pancreas display primitive cell markers (unpublished data), and may be migrating in response to local changes in SDF-1α concentration and variations in CXCR4 expression in the cells in response to the cytokine IFNγ. Once recruited to a niche where growth factors stimulate their proliferation and differentiation, these progenitors assume an endocrine cell lineage. The stimulatory effect of SDF-1α on the migration of CD34+ hematopoietic progenitor cells was established relatively early (Aiuti et al., 1997; Kim and Broxmeyer, 1998; Mohle et al., 1998). Of the many chemokines assayed SDF-1α was the first to have been shown to affect directed movement of myeloid progenitor cells (Broxmeyer et al., 1999). Wright et al. (2002) have reported recently that a purified population of hematopoietic stem cells expresses CXCR4 and migrates in response to SDF-1α in vitro. The mobilization of stem cells in and out of the bone marrow is important in therapeutic transplant procedures. However, what is more intriguing is whether these hematopoietic stem cells can migrate to sites of inflammation and differentiate into other tissue cell types (Krause et al., 2001). In vivo, SDF-1α is produced by bone marrow (Bleul et al., 1996) and the epithelial cells in many organs such as lung (Aiuti et al., 1997). Two recent papers provide evidence of CXCR4 expression in epithelial colon cells (Dwinell et al., 1999; Jordan et al., 1999). Interestingly, another class of cells of epithelial origin that express CXCR4 receptors are breast cancer cells, both primary and metastatic (Muller et al., 2001). Furthermore, using neutralizing antibodies for CXCR4 resulted in a significant reduction in metastatic ability indicating a clear effect of CXCR4 in the migration of breast cancer cells.

We present evidence that Src, MAPK, or Akt phosphorylation may potentially be involved in the stimulation of migration of the ductal cells by SDF-1α. In vitro stimulation of the freshly isolated ductal cells with SDF-1α resulted in the phosphorylation of MAPK and Akt. The robust SDF-1α effect on Akt phosphorylation coupled with the in vivo results of an augmentation of apoptosis with CXCR4 neutralization suggests an involvement of Akt in the promotion of survival of duct cells. Therefore, it is possible for SDF-1α to have a direct effect on proliferation and survival. Alternatively, CXCR4 expression might induce migration of progenitor cells to niches where they can be stimulated to proliferate, and subsequently, differentiate into endocrine cells. Importantly, we have shown previously that inhibition of the infiltration of macrophages did not play a role in the observed proliferation and islet regeneration in the IFNγ transgenic mice (Gu et al., 1995).

In the current paper, we report the limited expression of the chemokines C-10, MIG, TCA-4, Eotaxin, and SDF-1α in the 8-wk-old NOD pancreas. We had reported previously low level C-10 expression in the 10-wk-old NOD mouse pancreas (Bradley et al., 1999). In this earlier paper, Th1 cells harvested from primary cultures expressed high levels of lymphotactin, MIP-1α, MIP-1β, and MCP-1, and low levels of IP-10 and RANTES, when stimulated with anti-CD3. In addition, Chen et al. (2001) have reported the expression of MCP-1 in pancreatic islets isolated from NOD mice at the peak of insulitis (8–10 wk) using RT-PCR; and Cameron et al. (2000) observed a progressive increase in MIP-1α production by the NOD pancreas peaking at 5 wk, and MCP-1 expression starting to rise by 10 wk. In the latter paper, chemokine expression was measured by ELISA. The discrepancies in the findings by different investigators may reflect differences in the time course of insulitis progression in the NOD mice, methods used to measure expression of the chemokines, and the genders of the NOD mice. Furthermore, as the analysis of whole pancreas RNA clearly masks important local differences in chemokine and chemokine receptor expression, the localization and potential significance of chemokines both in the pathogenesis of diabetes and islet regeneration still remain challenging fields of study.
In conclusion, this paper provides in vivo evidence for a role of the SDF-1α–CXCR4 chemotaxis axis in a model of tissue regeneration. Importantly, elucidating the molecular mechanisms involved in the stimulation of migration and proliferation and the diminution of apoptosis in the epithelial precursor cells in the regenerating pancreas will help devise effective means for islet replacement in the future.

Materials and methods

Mice

The transgenic mice expressing IFN in the pancreatic β cells have been described previously (Gu and Sarvetnick, 1993, 1994; Sarvetnick et al., 1988, 1990). The IFNα transgenic mice used in the present work were on the NOD and NOD.scid background (Maikino et al., 1980; Prochna et al., 1992). The NOD background provides an excellent model for type 1 diabetes.

RNase protection assays

Total RNA was extracted from pancreata from 8- to 12-wk-old male IFNγ−/− and NOD mice using an RNeasy Kit (Qiagen) according to the manufacturer’s instructions. Pancreatic RNA was pooled from three IFNγ−/− transgenic mice and three NOD mice for the RNase protection assays. This process was repeated two independent times. In-house chemokine and chemokine receptor probes were used to perform the RNase protection assays as described previously (Asensio and Campbell, 1997; Boztug et al., 2002). The CHK1 chemokine probe detects lymphotactin, C-10, MIP-2, MCP-3, MIP-1β, T-cell activation gene 3, MCP-1, IP-10, MIP-1α, and RANTES. The CHK2 probe detects MIG, MCP-5, SDF-1α, TCA-4, growth-regulated oncogene-α, Eotaxin, lipopolysaccharide-induced CXC chemokine, IFN-inducible T-cell alpha chemoattractant, and Fractalkine. The probe for the CCR group of chemokine receptors included CCR6, CCR7, CCR8, CCR9, CCR4, CCR3, CCR1, and CCR5. The probe for the CXCR group of receptors included CXCR4, CXCR2, CXCR5, DARC, CXCR1, CXCR3, and CXCR1. Autoradiographs were analyzed by densitometry using NIH Image 1.63 for quantitation.

Immunohistochemistry and immunofluorescence

Pancreata from IFNγ−/− transgenic, NOD mice, and E18 NOD embryos were fixed in 10% neutral buffered formalin and embedded in paraffin. Paraffin embedded tissue was cut into 4-μm sections and stained using rabbit polyclonal anti–SDF-1α antibody against mouse SDF-1α (Cell Sciences) or rabbit polyclonal anti–CXCR4 antibody raised against the N-terminal extracellular domain of mouse CXCR4 receptor (Capralogics Inc.), or guinea pig antibody against insulin (DakoCytomation). Sections were counterstained with hematoxylin. For double immunofluorescent detection, biotinylated secondary goat, rabbit, and guinea pig antibodies (Vector Laboratories) were used, followed by streptavidin-conjugated Alexa Fluor 488 and 568 (Molecular Probes Inc.). The first streptavidin-conjugated Alexa Fluor 488 (488) incubation was followed by avidin and biotin blocking (Vector Laboratories). After the Alexa Fluor (568) incubation, the sections were placed in mounting medium from The Slowfade Light Antifade Kit (Molecular Probes Inc.). Nuclei were visualized with TOPRO3 (Molecular Probes Inc.). Sections were analyzed on a scanning confocal microscope (model MRC 1024; BioRad Laboratories), mounted on an Axiovert TV-100 with 40 or 63x objectives (Carl Zeiss MicroImaging, Inc.).

Pancreatic ductal cell purification and islet isolation

Cells of the pancreatic ductal network were purified from 10- to 12-wk-old IFNγ−/−, scid mice for the migration experiments and IFNα−/− mice for the assessment of in vitro phosphorylation of signaling proteins. The pancreata were collagenase (1 mg/ml; Roche) digested for 45 min. The digest was filtered through a 200-μm mesh and the ductal network above the mesh was treated with 0.05% trypsin, 0.33 mM EDTA. The resulting cell suspension was filtered through a 70-μm cell strainer. The cells in the filtrate were resuspended in RPMI medium. Approximately one million duct cells were derived from each pancreas preparation. The viability of these duct cell ranges from 75 to 95%. Pancreatic islets were isolated and cultured as described previously (Flodstrom et al., 2002).

Migration assays

Chemotaxis was assessed using uncoated or collagen-coated culture plate inserts (12-μm pore; 12-mm diam; Millipore) placed into 24-well plates. Half the inserts were coated with 1.48 mg/ml collagen type I (BD Biosciences), diluted threefold with 95% ethanol, 2 h at RT, and blocked with 1% BSA. Freshly isolated ductal network cells resuspended in serum-free RPMI 1640 medium (200,000 cells in 20 μl) were added to the upper chambers. 600 μl of RPMI 1640 medium with 0, 100, or 300 μg/ml of SDF-1α (PeproTech) was added to the bottom chambers and the cells were allowed to migrate for 24 h at 37°C. At the end of the assay, the cells from the upper chamber were aspirated and the membranes were fixed in 10% neutral buffered formalin, stained with hematoxylin and eosin, and mounted on slides upside down. Images of eight fields were captured from each membrane (at a magnification of 20) and the number of cells in or associated with the pores was counted.

Experimental protocol for the CXCR4 antibody neutralization study

12-wk-old IFNγ−/−NOD mice were divided into two groups of eight. Rabbit IgG or a rabbit polyclonal CXCR4 neutralizing antibody (20 μg/mouse; Gonzalo et al., 2000; Millennium Pharmaceuticals, Inc.) was injected intravenously every third day for a period of 2 wk. The total number of injections per mouse was five. On day 13 of treatment, 100 μg/g BrdU (Sigma-Aldrich) was administered intraperitoneally. 15 h after the BrdU injection, pancreatic tissue was fixed in Bouin’s. Monoclonal rat anti-BrdU antibody (Accurate Chemical) was used to assess proliferation.

In pancreata from a subgroup of mice (n = 2 per group) treated with CXCR4 neutralizing antibody or control IgG, TUNEL staining was performed using the in situ cell death detection POD kit (Roche) according to the manufacturer’s instructions. TUNEL staining was repeated on sections from two different levels of the pancreas for each animal.

Pancreatic islets from mice treated with IgG or CXCR4 neutralizing antibody (n = 4 per group) were evaluated for PDX1 staining using rabbit polyclonal anti-PDX1 antibody (CHEMICON International, Inc.). Images were captured from at least six ductal areas from each mouse and the PDX1-positive duct number and the total duct cell number in these areas were quantified. In seven of the mice from each group, the percentage of small ducts (defined as ducts comprising <15 cells) and large ducts were quantified by counting duct cell foci at least. Images captured the overlay of images captured from one hematoxylin stained section from each mouse.

Assessment of SDF-1α-stimulated Src, MAPK, and Akt phosphorylation

Freshly isolated cells from the pancreatic ductal network from IFNγ−/− transgenic mice were serum-starved overnight, and stimulated with 100 or 300 mg/ml SDF-1α or 10 ng/ml EGF for 5 min at 37°C. Cells were lysed with RIPA buffer containing 0.5% Nonidet P-40, 1% sodium deoxycholate, 1% sodium dodecyl sulfate, 10 mM EDTA, and 0.5% sodium azide (Pierce). Cell lysates were prepared for Western blot analysis. Rabbit polyclonal antibodies to phospho-Src (Tyr 416), dually phosphorylated phospho-MAPK (Thr202/Tyr204), and phospho-Akt (Ser473) (Cell Signaling Technology) were used for immunodetection (Cell Signaling Technology). In a second stimulation experiment, the cells were isolated and serum starved as above and treated with 300 mg/ml SDF-1α for 0, 2, 5, 10, 30, and 60 min. Pancreatic islets were precultured for 5–6 d before a 24-h exposure to 1,000 U/ml IFNγ (BD Biosciences) or vehicle. After exposure, the islets were homogenized in RIPA buffer, and lysates prepared for Western blot analysis and were immunoblotted with SDF-1α antibody. All membranes were stripped and reblotted with a mouse mAb to actin to confirm equal protein loading (ICN Biomedicals).

Statistical analysis

Analysis of variance was used to analyze data in Fig. 5. The t test was used to analyze data in Figs. 6–9.

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