Introduction

*C. burnetii* is a Gram-negative, obligate intracellular bacterium which is the causative agent of Q fever, a zoonotic disease with worldwide prevalence except for New Zealand.\(^1\) Humans get infected by inhalation of infectious material transmitted from domestic livestock. Infection by a single bacterium can result in disease.\(^7\) Most frequently, Q-fever remains asymptomatic or causes only a mild flu-like illness. However, the infection can also lead to interstitial pneumonia, hepatitis or severe chronic disease, which usually presents in form of an endocarditis.\(^3\) Due to the increase in worldwide Q-fever cases over the last decade *C. burnetii* is considered as an emerging pathogen.\(^3\)

Once *C. burnetii* has entered the human body by inhalation it is taken up by mononuclear phagocytes into a phagosome.\(^4\) The *C. burnetii*-containing vacuole (CCV) matures to an acidic, phagolysosomal-like parasitophorous vacuole that is permissive for bacterial replication.\(^5-7\) *C. burnetii* requires a functional Dot/Icm type IV secretion system (T4SS) for the establishment of the replicative CCV.\(^8-10\) The T4SS translocate bacterial virulence factors, termed effector proteins, into the host cell cytoplasm.\(^11\) Importantly, it was shown that *C. burnetii* requires the T4SS to prevent host cell death.\(^9\) To date, more than 130 putative *C. burnetii* effector proteins have been identified,\(^12\) some of which have anti-apoptotic activity,\(^13-15\) whereas the majority still awaits functional characterization.

Apoptosis, a form of programmed cell death, is part of the intrinsic immune defense.\(^16\) It allows removal of damaged or infected cells in the absence of inflammation\(^17\) and is a central mechanism of peripheral immune tolerance.\(^18\) Two main pathways lead to apoptosis induction: The extrinsic and the intrinsic apoptosis pathway. The extrinsic pathway is triggered by ligand binding to
death receptors and adaptor proteins, which activates caspases (cysteinyl aspartate proteases). The intrinsic pathway involves activation of Bax and Bak, which is regulated by the Bcl-2 protein family.\(^{19,20}\) This protein family comprises both positive (BH3-only) and negative (Bcl-2-like) regulators of apoptosis. The ratio of positive and negative regulators expressed in a cell determines whether Bax and Bak are activated. Once activated, Bax and Bak oligomerize and permeabilize the mitochondrial membrane, resulting in the release of small molecules like cytochrome c and activation of caspase 9 through assembly of the apoptosome.\(^{21}\) Activated caspase 9 leads to cleavage and, thus, activation of the key downstream executors of apoptosis: caspase 3 and caspase 7.\(^{22}\) As proteolysis is irreversible, activation of caspases must be tightly regulated. One known mechanism deployed to protect the cell from death receptor and mitochondrial apoptosis is the inhibition of activated caspases by inhibitor of apoptosis proteins (IAPs). In humans the IAPs comprise 8 family members: NAIP, cIAP1, cIAP2, XIAP, survivin, Bruce, ML-IAP and ILP2.\(^{23}\) While it was first believed that all of the IAPs can bind caspases, we now know that in vivo only XIAP functions as a physiological inhibitor of caspases.\(^{24,25}\) The other IAPs require the interaction with cooperative partners to inhibit caspases and thereby apoptosis.\(^{24}\)

Several pathogens have evolved mechanisms to modulate host cell apoptosis, which has emerged as a crucial determinant of virulence. Some bacteria actively induce host cell death to escape from an microbicidal, phagosomal environment or to overcome barriers.\(^{26,27}\) In contrast, obligate intracellular pathogens frequently inhibit apoptosis (e.g. by use of T3SS or T4SS effector proteins) to prevent premature host cell death and to generate a habitat for replication.\(^{28,29}\) \textit{C. burnetii} is no exception as its T4SS to inhibit host cell apoptosis.\(^{9}\) So far, 3 anti-apoptotic effector proteins (AnkG, CaeA and CaeB) have been identified.\(^{14,15,30}\) However, their precise molecular activity has still to be unraveled.

The \textit{C. burnetii} effector protein CaeA localizes to the nucleus when produced in mammalian cells and displays anti-apoptotic activity.\(^{14}\) In this study we found that CaeA does not interfere with the activation of caspase 9 but prevents the cleavage of caspase 7, indicating that CaeA interferes with the apoptotic cascade between the initiator caspase 9 and the executioner caspase 7. Interestingly, CaeA expression resulted in an upregulation of survivin, an IAP known to inhibit activated caspases. However, siRNA experiments revealed that the anti-apoptotic activity of \textit{C. burnetii} was not mechanistically linked to the increased level of survivin. By comparing the sequence of CaeA from 25 \textit{C. burnetii} isolates we identified an EK repetition motif, consisting of alternating glutamic acid and lysine residues, as a site of genetic variability. The EK motif of CaeA was required for anti-apoptotic activity, but not for the intracellular localization of CaeA.

**Results**

**CaeA expression prevents apoptosis at the executioner caspase level**

An increase in the anti-apoptotic Bcl-2-like steady-state protein level results in cell survival. Therefore, we analyzed whether the expression of CaeA (derived from the \textit{C. burnetii} Nine Mile [NM] II strain) results in an altered protein level of anti-apoptotic Bcl-2-family members. Thus, HEK293 cells stably expressing GFP or GFP-CaeA were treated with different UV-light intensities. UV-light induces DNA damage and subsequently intrinsic apoptosis.\(^{31}\) Immunoblot analysis revealed a similar relative abundance of the anti-apoptotic Bcl-2-like proteins Bcl-2 and Bcl-x\textsubscript{L} under all conditions tested (Fig. 1A). In contrast, we detected decreased Mcl-1 protein levels after UV-light treatment, which, however was also true for both GFP-CaeA expressing and GFP control cells. Next, we analyzed whether CaeA inhibits activation of initiator and executioner caspases after UV-light treatment. Apoptosis-induction was measured by assaying the presence of cleaved poly ADP-ribose polymerase (PARP). Proteolytic cleavage of nuclear PARP inactivates DNA repair activity and is a marker for the terminal stages of apoptosis.\(^{32}\) As shown in Figure 1B, GFP-expressing cells treated with UV-light displayed caspase 9 and caspase 7 cleavage as well as PARP cleavage. In contrast, cells expressing GFP-CaeA were refractory to UV-induced proteolytic cleavage of caspase 7 and PARP, but not to proteolytic cleavage of caspase 9. This result suggests that CaeA interferes with the apoptotic cascade downstream of caspase 9 activation and upstream of caspase 7 activation.

**CaeA also inhibits extrinsic apoptosis**

As our data suggested that CaeA inhibits apoptosis at the executioner caspase level, we assumed that CaeA not only inhibits intrinsic apoptosis but also extrinsic apoptosis. Fas-ligand and tumor necrosis factor (TNF) in combination with cycloheximide (CHX) are known inducers of the extrinsic apoptosis pathway.\(^{33}\) Thus, we induced extrinsic apoptosis with TNF and CHX in HeLa cells ectopically producing GFP or GFP-CaeA and measured nuclear fragmentation visualized by DAPI staining. This assay was used as it is a fast and robust assay. Importantly, we used the TUNEL assay to validate our nuclear fragmentation assay. As shown
in Figure 1C the expression of GFP-CaeA protected the cells significantly from TNF plus CHX-induced apoptosis compared to cells expressing GFP alone. To determine whether CaeA expression also inhibits Fas-ligand induced apoptosis, HeLa cells stably expressing the Fas receptor (HeLa-Fas) and transiently expressing GFP or GFP-CaeA were treated with Fas-ligand. As expected, CaeA expression significantly inhibited Fas-ligand-induced extrinsic apoptosis visualized by DAPI staining (Fig. 1D). Importantly, expression of GFP or GFP-CaeA without cell death induction did not influence viability of HeLa and HeLa-Fas cells (data not shown). Together these data indicate that CaeA protects cells from both intrinsic and extrinsic apoptosis.

**CaeA expression results in an up-regulation of survivin**

Apoptotic cell death can be prevented by inhibitor of apoptosis proteins (IAPs). Therefore, we assessed the steady state level of the IAPs XIAP and survivin. XIAP can bind directly to caspase 3, 7 and 9, protecting them from proteolytic cleavage. Survivin is the smallest member of the IAP family and regulates several cellular processes, including apoptosis. We also aimed to analyze the expression level of the other members of the IAP family. However, probably due to low expression level we were unable to detect these IAP at protein and mRNA level.

Thus, HEK293 cells stably expressing GFP or GFP-CaeA were treated with different UV-light intensities. Immunoblot analysis revealed an upregulation of survivin in GFP-CaeA expressing cells as compared to GFP expressing cells, while XIAP was not altered (Fig. 2A). This effect of CaeA was detected during steady-state conditions as well as after UV exposure of the cells (Fig. 2A and 2B).

**CaeA-induced inhibition of apoptosis is not dependent on survivin up-regulation**

As CaeA expression caused inhibition of apoptosis and upregulation of survivin, we investigated whether the
increased protein expression of survivin accounts for the anti-apoptotic effect of CaeA. To this end, we transfected HEK293 cells stably expressing GFP or GFP-CaeA were exposed to UV-light (200 J/m² or 800 J/m²) and incubated for 6 h. (a) Proteins were separated by SDS-PAGE, transferred to a PVDF membrane and probed with antibodies against survivin, XIAP and actin as loading control. The result of one representative experiment out of 3 independent experiments with similar results is shown. (b) Densitometric analysis of the survivin/actin ratio was performed using ImageJ. The data shown represent average values ± SD from at least 3 independent experiments. *P < 0.05 ***P < 0.001

**Figure 2.** CaeA leads to survivin up-regulation. HEK293 cells stably expressing GFP or GFP-CaeA were exposed to UV-light (200 J/m² or 800 J/m²) and incubated for 6 h. (a) Proteins were separated by SDS-PAGE, transferred to a PVDF membrane and probed with antibodies against survivin, XIAP and actin as loading control. The result of one representative experiment out of 3 independent experiments with similar results is shown. (b) Densitometric analysis of the survivin/actin ratio was performed using ImageJ. The data shown represent average values ± SD from at least 3 independent experiments. *P < 0.05 ***P < 0.001

Sequence analysis of C. burnetii strains reveals a genetic variability of CaeA

CaeA has a molecular weight of 25.1 kDa and contains a predicted coiled-coil region that spans from aa 25 to 117 and 2 nuclear localization signals (NLS). Comparison of genomes from different C. burnetii isolates pointed to a significant plasticity in the repertoire of T4SS effector proteins. With respect to single effector proteins, differences in the DNA sequence might help to elucidate their function. Therefore, we analyzed the sequence of caeA from 25 different strains of C. burnetii (Table 1). As shown in Figure 4, the sequence analysis revealed a striking similarity of all strains except for a 5’ gene locus starting from base pair 148 that contains GAA AAG/A repeats. This segment, which is part of the coiled-coil region, encodes alternating glutamic acid (E) and lysine (K) residues. It is present in all strains analyzed, but the number of EK repeats varied between 3 and 13. In addition, 2 single base deletions occurred. Twenty of the sequenced strains showed a single base deletion that

**Figure 3.** Survivin knock-down does not affect CaeA-induced apoptosis-inhibition. HEK293 cells stably expressing GFP or GFP-CaeA were transfected with 50nM non-targeting siRNA (−) or human survivin siRNA (+). (a) 48 hours post-transfection total isolated RNAs were reverse transcribed using SuperScript II reverse transcriptase according to the manufacturer’s protocol and a qRT-PCR was performed with oligonucleotides specific for survivin and hpbgd as a house keeping gene. The ΔΔCt values were calculated for the fold difference of survivin siRNA-treated cells to non-targeting siRNA-treated cells using the 2−ΔΔCt method. (b) 48 hours post-transfection cells were treated with or without UV light (800 J/m²) and incubated for 6h at 37°C in 5% CO₂. Proteins were separated by SDS-PAGE, transferred to a PVDF membrane and probed with antibodies against cleaved PARP, survivin and actin as loading control. The result of one representative experiment out of 3 independent experiments with similar results is shown.
provoked a frame shift and the introduction of a premature ochre stop codon at base pairs 61-63. An additional second single base deletion, downstream of this stop codon, was present in 8 of these isolates. Consequently, \textit{caeA} represents a presumptive pseudogene in 20 of the 25 isolates analyzed in this study. Based on these \textit{in silico} analyses only the \textit{C. burnetii} strain NM I, which contains 6 EK repeats, and 4 further isolates, which all contain 3 EK repeats, are predicted to express a functional CaeA molecule. These five isolates originated from different regions and host species. While strain F-3 was obtained in France from a patient with endocarditis and therefore appears to be associated with chronic Q-fever, strain F-4 was isolated in France from an acute Q-fever patient and the NM strain originated from a tick in Montana, USA. The strain Namibia was obtained from an infected goat in Namibia and the strain Z3574-1/92 was derived from milk of an infected ewe in Germany; in both cases there was no information available on the clinical status of the host animals.

Taken together, the \textit{caeA} gene of \textit{C. burnetii} shows a considerable genetic polymorphism in a region which encodes an EK-repetition motif.

\textbf{The EK repetition motif is required for the anti-apoptotic activity of CaeA}

To assess the functional role of the EK motif, a HEK293 cell line that stably expressed GFP-CaeA lacking the EK repeat (GFP-CaeA\_D\_EK) was established. This cell line was analyzed by flow cytometry together with the already established HEK293 cells stably expressing GFP or GFP-CaeA. As shown in Figure 5A, around 90% of the cells in all 3 stable cell lines expressed GFP. Next, the anti-apoptotic activity of CaeA lacking the EK repeats was tested. While HEK293-GFP cells showed caspase 7 and PARP cleavage after UV-light exposure, HEK293-GFP-CaeA cells were protected from caspase 7 and PARP cleavage (Fig. 5B and 5C), confirming the results in Figure 1B. In contrast, HEK293-GFP-CaeA\_D\_EK showed increased levels of caspase 7 and PARP cleavage after UV-light treatment, suggesting that CaeA lacking the EK repeats was unable to protect cells from apoptosis. Importantly, ectopically expressed GFP-CaeA\_D\_EK still exhibited a proper intracellular localization under steady-state conditions and after apoptosis induction. As depicted in Figure 6, both GFP-CaeA and GFP-CaeA\_D\_EK showed nuclear speckle-like localization with and without apoptosis-induction. Thus, the EK repetition motif enables CaeA to interfere with the apoptosis cascade, but is not required for the intracellular localization of CaeA.

\textbf{CaeA-induced apoptosis inhibition varies with the number of EK repeats}

Sequence analyses of 25 different isolates of \textit{C. burnetii} showed that CaeA contains between 3 and 13 EK repeats. As stated above, only CaeA molecules containing either 3 or 6 EK repeats are expressed as functional effector proteins. To determine how many
repetitions of the EK motif in CaeA are minimally required for efficient inhibition of apoptosis, we tested CHO cells ectopically expressing CaeA \(_D^{\text{EK}}\) with 2 \((\text{CaeA}^{\text{D}}^{\text{EK}})^{C^{2^{\text{EK}}}}\) or 4 EK \((\text{CaeA}^{\text{D}}^{\text{EK}})^{C^{4^{\text{EK}}}}\) repeats. CHO cells expressing wild-type NM II-derived CaeA were significantly protected from apoptosis, whereas cells expressing CaeA\(_{\text{D}}^{\text{EK}}\) were not (Fig. 7), confirming the results obtained in HEK293 cells (Fig. 5B and 5C). Importantly, expression of CaeA\(_{\text{D}}^{\text{EK}}+^{2^{\text{EK}}}\) also inhibited apoptosis, but significantly less than CaeA, while expression of CaeA\(_{\text{D}}^{\text{EK}}+^{4^{\text{EK}}}\) protected the cells as efficiently as wild-type NM II CaeA, which contains 6 EK repeats. From these findings we conclude that at least 4 EK repetitions are required for full anti-apoptotic activity of CaeA.

Discussion

Apoptosis of infected host cells has been identified as an important element of the innate immune response contributes to the clearance of microorganisms cleared from the body. Especially obligate intracellular pathogens have evolved mechanisms to impede the process of host cell apoptosis, but the underlying molecular mechanisms are still incompletely understood. This is also the case for \(C.\ burnetii\), for which inhibition of host cell apoptosis was first described in 2007.\(^{38,39}\) The anti-apoptotic function of the \(C.\ burnetii\) effector proteins AnkG, CaeA and CaeB \(^{14,15}\) suggests that proteins transported by the T4SS are important regulators of host cell survival.
Due to a re-annotation of *C. burnetii* Nine Mile phase I (RSA493) CaeA and CaeB are now annotated as pseudogenes. CaeA (former CBU1524) is now part of CBU1523 and CaeB (former CBU1532) is now part of CBU1531. However, RT-PCR analysis confirmed transcription of CaeA and CaeB and the presence of a functional T4SS translocation signal at its C-terminus. Whether CaeA is only a fragment of CBU1523 or

**Figure 5.** The EK repetition motif is required for apoptosis inhibition. (a) HEK293 cells and HEK293 cells stably expressing GFP, GFP-CaeA and GFP-CaeAΔEK were analyzed by flow cytometry. One representative experiment out of at least 3 independent experiments with similar results is depicted. (b and c) HEK293 cells stably expressing GFP, GFP-CaeA or GFP-CaeAΔEK were exposed to the UV-light intensities indicated and incubated for 6 h at 37 °C in 5% CO2. Proteins were separated by SDS-PAGE, transferred to a PVDF membrane and probed with antibodies against (b) cleaved caspase 7, cleaved PARP (c) and actin as loading control. The result of one representative experiment out of at least 3 independent experiments with similar results is shown.

**Figure 6.** Deletion of the EK repetition motif does not alter intracellular localization of CaeA. Representative immunofluorescence micrographs show CHO-FcR cells transiently transfected with GFP, GFP-CaeA and GFP-CaeAΔEK. The cells were treated with or without 1 μM staurosporine for 6 h at 37 °C in 5% CO2 followed by fixation, permeabilization and staining of the nuclei with DAPI (blue).
whether CBU1523 and CaeA (CBU1524) are 2 independent proteins is unknown and has not been experimentally tested. However, we hypothesize that CaeA is not a fragment of CBU1523 but rather a discrete protein, as the CBU1524 gene contains a potential Shine Dalgarno sequence upstream of the start codon, and an 114bp intergenic stretch, which separates the stop codon of CBU1523 and the start codon of CBU1524. This intergenic stretch is enough sequence space to contain transcriptional and translational start sites for CBU1524.

In this study, we investigated the determinants of the anti-apoptotic function of the C. burnetii effector protein CaeA. CaeA was originally described as an inhibitor of the intrinsic apoptosis pathway when endogenously expressed in HEK293 or CHO cells.14 Our present results suggest that CaeA also protects from the induction of extrinsically triggered apoptosis (Fig. 1C and D). This indicates that CaeA might act at the intersection of both apoptosis pathways. In agreement with this assumption, cleavage of the effector caspase 7 was completely inhibited in presence of CaeA whereas cleavage of the upstream initiator caspase 9 was not (Fig. 1B). IAPs are known to inhibit activated caspases and to interfere at this late step of the apoptotic pathway.24 Some pathogens exploit the anti-apoptotic function of IAPs in order to manipulate the host apoptotic pathway and to promote host cell survival. Importantly, CaeA expression resulted in up-regulation of the steady-state protein level of the IAP survivin (Fig. 2). Several intracellular pathogens have been described to elicit an upregulation of survivin which subsequently improved host cell survival.40-42 For instance, the intracellular protozoan Cryptosporidium parvum caused an increased expression of cIAP1, cIAP2, XIAP as well as survivin in HCT-8 epithelial colon cells, and survivin was shown to be important for apoptosis inhibition by the pathogen.41 These observations contrast with our findings. While CaeA expression led to higher levels of survivin protein in the host cell, CaeA-mediated inhibition of apoptosis was independent of survivin (Fig. 3).

Sequence analysis of caeA from 25 different C. burnetii isolates revealed a noticeable difference in a region consisting of a variable number of GAA AAG/A (EK) codons (Fig. 4). The EK repetition motif of CaeA represents a short tandem repeat (STR) with the 6 bases ‘GAA AAG/A’. In general, STRs or 'microsatellites' are widely distributed in the genomes of eukaryotes and prokaryotes and are characterized by a sequentially repeated unit of one to 6 bp reaching up to 100 nt. Tandem repeats are hot spots for mutational events contributing to gene diversity and the evolutionary process. The high mutation rate and variability of tandem repeats is mainly explained by slipped-strand mispairing and a malfunctioning repair system during replication.43 Consistently, the presence of diverse numbers of EK repeats among the analyzed strains is most likely due to events of slipped-strand mispairing during the replication of the hexanucleotide GAA AAG/A. Besides the variability in the number of GAA AAG/A repetitions, 2 single base deletions were found to contribute to the sequence polymorphism of caeA in the different strains. In isolates encoding 5, 8, 10, 11 or 13 GAA AAG/A repetitions a single base deletion occurred leading to a frameshift and premature stop. In these strains caeA presumably represents a pseudogene. Interestingly, in addition to this first single base deletion, only in isolates encoding a higher number of repetitions, i.e. 10 to 13 GAA AAG/A units, a second single base deletion was found. In the absence of the first single base deletion the second base deletion would result in a frameshift altering the EK motif to an irregular succession of lysine and arginine residues and provoking a premature stop codon directly downstream. Thus, the second base deletion would similarly ensure the expression of a truncated protein. However, every isolate harboring the second base deletion, already showed the first base deletion at position ~55 which per se is sufficient to cause the truncation of CaeA. Remarkably,
in addition to NM only 4 of all 25 analyzed isolates are also capable to express a non-truncated protein. All these isolates of C. burnetii contain 3 EK repeat units and include 2 human, one goat and one sheep isolate. Thus, one might speculate the presence of a low number of EK repeats is a prerequisite for the expression of a non-truncated CaeA.

In prokaryotic genomes STRs exhibit a high mutation rate of up to $10^{-1}$ which contributes to phenotypic variability and fast adaptation to environmental or host conditions as has been shown especially for pathogenic bacteria. Indeed, human pathogens such as Neisseria gonorrhoeae or Haemophilus influenzae have been shown to undergo phase variation and to regulate protein expression similar to a switch on and off mechanism by alteration of the number of 5 and 4 base pair repetition units, respectively. Thus, the expression of H. influenzae lica, a gene that is participates in the synthesis of lipoteichoic acid (LTA), undergoes a phase variation that is mediated by the varying number of a 4 base pair repetition unit. In contrast, variations of the number of tri- or hexanucleotides (such as the CaeA EK repetition motif) do not influence the frame of a sequence and alter gene expression per se. Tri- or hexanucleotide tandem repeats seem to rather influence the function of a protein as variations of the repeat number may alter structural characteristics of a protein. For instance, the activity of the Salmonella enterica serovar Typhimurium and Escherichia coli DNA mismatch repair protein MutL is lost by alteration of the number of the 3 hexanucleotide repeat units encoded in the ATP binding pocket of MutL. As shown by Shaver et al. the deletion as well as the addition of one unit of the hexanucleotide repeat presumably interferes with ATP binding resulting in the inhibition of the ATP-dependent MutL function and an increased mutation rate.

Regarding CaeA, the EK repetition lies within a predicted coiled-coil region, a domain that is organized in $\alpha$-helical bundles facilitating protein-protein interactions. The negatively and positively charged amino acids of the EK motif account for a charged region with a hydrophilic character and presumably lead to the exposition of the domain to the protein surface. Thus, the EK motif might provide a protein interaction site within the coiled-coil region which accounts for the interaction with specific protein partners. Similar to the aforementioned MutL mutation, variation of the EK motif might influence the interaction and the function of the protein. In accordance with this hypothesis the EK repetition motif was shown to be essential for CaeA-mediated inhibition of apoptosis (Fig. 5B and 5C).

Taken together, we have demonstrated that the effector protein CaeA of C. burnetii is an efficient inhibitor of the extrinsic pathway of apoptosis. Furthermore, we defined the EK repetition motif of CaeA as a structural component that is crucial for the anti-apoptotic function of CaeA. Further studies will aim to identify the exact interaction partners of CaeA in the host cell.

Materials and methods

Reagents, cell lines and bacterial strains

Unless otherwise noted, chemicals were purchased from Sigma Aldrich. Complete Protease inhibitor cocktail mixture and X-tremeGENE 9 Transfection Reagent were from Roche. Staurosporine was from Cell Signaling. Cell lines were cultured at 37°C in 5% CO₂ in media containing 5% heat-inactivated fetal bovine serum (Biochrom). Chinese hamster ovary (CHO) fibroblasts were grown in minimal essential medium α medium (Invitrogen) and stable human embryonic kidney (HEK293) cells were maintained in Dulbeccos modified Eagle medium (Invitrogen) supplemented with 1.5 mg/ml G418 (Roth). To construct HEK293 cells stably expressing GFP-CaeAΔEK the corresponding plasmid was transfected into HEK293 cells using X-tremeGENE 9 and selected by culturing in media supplemented with 1.5 mg/ml G418 (Roth) for 7 days. Single GFP-positive cells were sorted and grown for additional 10 to 14 days. C. burnetii strains used in this study are listed in Table 1. To generate plasmids encoding CaeA fusion proteins, genes were amplified by PCR, using C. burnetii Nine Mile phase II clone 4 as template and primers designed according to an earlier annotation of RSA493 (C. burnetii Nine Mile phase I).

siRNA knock-down

The protocol used for siRNA transfection was adapted from Dharmacon’s HeLa cell transfection protocol. One volume of siRNA buffer containing 50nM of non-targeting or survivin siRNA (Dharmacon) was incubated with one volume of serum-free DMEM high glucose containing 5 µl/ml DharmaFECT-1 transfection reagent for 20 min at room temperature. Two volumes of DMEM high glucose supplemented with 20 % FBS containing 5 $\times$ 10⁴ HeLa cells were added. The cells were seeded in a 12-well plates and incubated at 37°C with 5% CO₂. The transfection mix was replaced with 500 µl of fresh media (DMEM high glucose supplemented with 10% FBS) 24 h post transfection. 3 days post transfection, the cells were washed with PBS and exposed to UV light (800 J/m²) in a transilluminator box (Stratagene). After
adding fresh media, the cells were incubated for 6 h at
37°C in 5% CO₂. The samples were separated by SDS-
PAGE and transferred to a PVDF membrane (Millipore).
The membranes were probed with antibodies against
survivin (2808, Cell Signaling), cleaved PARP (611038,
BD Biosciences) and actin (A2066, Sigma-Aldrich). The
proteins were visualized by using horseradish peroxi-
dase-conjugated secondary antibodies (Dianova) and a
chemiluminescence detection system (Thermo Scientific
or Millipore).

**Plasmids and primers**

Plasmid and primers used in this study are listed in
Tables 2–4

**Plasmid construction**

pEGFP-CaeΔEK was generated by mutagenic PCR
with primers 1 and 2 (Table 3) amplified from pCMV-
HA-CaeA. Ligation of PCR product resulted in pCMV-
HA-CaeΔEK. This vector was restricted with BglII and
KpnI, followed by ligation with like-wise restricted
pEGFP-C2.

pEGFP-CaeΔEK+2EK and pEGFP-CaeΔEK+4EK
were generated by mutagenic PCR with primers 1 and 3
or 1 and 4 (Table 3) amplified from pCMV-HA-CaeA-
ΔEK. Ligation of the PCR product resulted in pEGFP-
CaeΔEK+2EK and pEGFP-CaeΔEK+4EK. The vec-
tors were restricted with BglII and KpnI, followed by
ligation with like-wise restricted pEGFP-C2.

**Apoptosis assays**

**Nuclear fragmentation assay**

CHO cells were plated on coverslips in 24-well dishes at
a density of 2.5 × 10⁵ cells/well. After an overnight incu-
bation, cells were transfected with the plasmids indi-
cated. 18 h post-transfection, the cells were incubated
with staurosporine (1 µg/ml) for 6 h at 37°C in 5% CO₂.
The cells were fixed with 4 % paraformaldehyde (Alfa
Aeser) in PBS (Biochrom) for 20min at room tempera-
ture, permeabilized with ice-cold methanol for 30sec and
quenched with 50 mM NH₄Cl (Roth) in PBS for 15min
at room temperature. The cells were mounted using Pro-
Long Gold with DAPI (Invitrogen) to visualize the
nucleus.

**Immunoblotting**

HEK293 cells stably expressing GFP, GFP-CaeA or GFP-
CaeΔEK were seeded in a 12-well plate at a density of
3 × 10⁵ cells/well. The cells were washed with PBS before
exposure with the indicated UV-light intensities (Strata-
gene). After 6h incubation at 37°C in 5%CO₂ samples
were separated by SDS-PAGE and transferred to a
PVDF membrane (Millipore). The membranes were
probed with antibodies directed against Bcl-2 (2870),
Bcl-xL (2764), Mcl-1 (4572), cleaved caspase 7 (9491),
cleaved caspase 9 (9501), XIAP (2045) and survivin
(2808) all from Cell Signaling, cleaved PARP (611038,
BD Biosciences) and actin (A2066, Sigma-Aldrich). The
proteins were visualized by using horseradish peroxi-
dase-conjugated secondary antibodies (Dianova) and a
chemiluminescence detection system (Thermo Scientific
or Millipore).

**TNF + CHX–induced extrinsic apoptosis**

HeLa cells were plated on coverslips in 24-well dishes at
a density of 3.5 × 10⁴ cells/well and were transfected with
the plasmids indicated. 40 h post-transfection, the cells
were incubated with 20 ng/ml TNF (Calbiochem) and 6
µg/ml cycloheximide (Sigma) for 5 h at 37°C in 5% CO₂.
The cells were fixed with 4 % paraformaldehyde (Alfa

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**Table 2. Plasmids used in this study.**

| no | Plasmid              | Reference                  |
|----|----------------------|----------------------------|
| 1  | pCMV-HA-CaeA         | Klingenbeck et al.         |
| 2  | pCMV-HA-CaeAΔEK      | this study                 |
| 3  | pEGFP-CaeΔEK         | this study                 |
| 4  | pCMV-HA-CaeΔEK+2EK   | this study                 |
| 5  | pCMV-HA-CaeΔEK+4EK   | this study                 |
| 6  | pEGFP-CaeΔEK+2EK     | this study                 |
| 7  | pEGFP-CaeΔEK+4EK     | this study                 |

**Table 3. PCR Primers used in this study.**

| no | Oligonucleotide | Sequence 5’-3’ |
|----|----------------|----------------|
| 1  | caeΔEK_fwd     | TTAACACAGCAACTCACCGAAGAACAGGAGCGTTC |
| 2  | caeΔEK_rev     | AAGGTCTGTGCTTTTTTCTTCGAGTCGCTGATAGTCATTG |
| 3  | caeΔEK+2_rev   | TTCTTTTTCTTTTTTTTCTTTTTTTTTTTTTTTTTT |
| 4  | caeΔEK+4_rev   | TTCTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT |
| 5  | caeA_3_fwd     | AAATTCCGCAACCATCTCAAGTGAAGGCCATCAAGG |
| 6  | caeA_4_rev     | GAAGAAAAGCGGCGATCCACTCTCAAT |
Aeser) in PBS (Biochrom) for 20 min at room temperature, permeabilized with ice-cold methanol for 30 sec and quenched with 50 mM NH₄Cl (Roth) in PBS for 15 min at room temperature. The cells were mounted using Pro-Long Gold with DAPI (Invitrogen) to visualize the nucleus.

**Fas-ligand-induced extrinsic apoptosis**

HeLa-Fas cells 50 were plated on coverslips in 24-well dishes at a density of 4 × 10⁴ cells/well. After an overnight incubation, cells were transfected with the plasmids indicated. 18 h post-transfection, the cells were incubated with 0.125 μg/ml anti-Fas IgG (Millipore) for 5 h at 37°C in 5% CO₂. The cells were fixed with 4% paraformaldehyde (Alfa Aeser) in PBS (Biochrom) for 20 min at room temperature, permeabilized with ice-cold methanol for 30 sec and quenched with 50 mM NH₄Cl (Roth) in PBS for 15 min at room temperature. The cells were mounted using ProLong Gold with DAPI (Invitrogen) to visualize the nucleus.

**Confocal microscopy**

2 × 10⁴ CHO cells were plated on coverslips and were transfected with the plasmids indicated. The cells were fixed with 4% paraformaldehyde (Alfa Aeser) in PBS (Biochrom), permeabilized with ice-cold methanol and quenched with 50 mM NH₄Cl (Roth) in PBS for 15 min at room temperature. The cells were mounted using ProLong Gold with DAPI (Invitrogen) to visualize the nucleus. Fluorescence was detected using a Carl Zeiss LSM 700 Laser Scan Confocal microscope and a 64x objective. Images were analyzed using the ZEN2009 software (Jena, Germany).

**FACS analysis**

Protein expression levels of HEK293 cells stably expressing GFP, GFP-CaeA and GFP-CaeAΔEK were analyzed by flow cytometry.

**PCR, sequencing and sequence analysis**

PCR primers 5 and 6 (Table 3) for caeA were designed with Primer Select software implemented in Lasergene 9.0 (DNASTAR, Madison, WI). caeA was amplified using 1x OptiBuffer (Bioline, London, United Kingdom), 1.4 mM MgCl₂, 200μM deoxynucleoside triphosphates (Bioline), 25pmol of each primer, 1.5 U Bio-X-Act Short DNA polymerase (Bioline), and 5 μl of DNA, in a final volume of 25 μl. The PCR was run with the following thermocycling profile: 5 min at 95°C; 1 min at 58°C; 35 cycles, each consisting of 1 min and 15 s at 72°C, 30 s at 95°C, and 30 s at 58°C; and a final elongation step of 10 min at 72°C. Cycle sequencing was performed using BigDye v1.1 chemistry according to the manufacturer’s instructions (Applied Biosystems) (see Sequencing primers in Table 4), purified with DyeEx 96 plates (Qiagen) and electrophoresed on a 3130XL Genetic Analyzer (Applied Biosystems). Sequence analysis and polymorphism evaluation was performed using Lasergene 9.0 (DNASTAR, Madison, WI) and MEGA5 (Tamura et al., 2011).

The sequences of caeA gene are available at GenBank under the accession numbers KT828680-KT828703.

**Isolation of genomic Coxiella burnetii DNA**

Genomic DNAs of 25 paraformaldehyde-fixed or heat-killed C. burnetii isolates was prepared according to the manufacturer’s protocol with the illustra bacteria genomic Prep Mini Spin Kit (GE Health Care), with 3×10 min incubation at 55°C in the beginning and a final elution with 30 μl H₂O.

**Statistical analysis**

An unpaired Student’s t-test or Chi-square test was used for statistical analysis.

**Disclosure of potential conflicts of interest**

No potential conflicts of interest were disclosed.

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