Exploring the Role of Novel Medical Therapies for Aggressive Pituitary Tumors: A Review of the Literature—“Are We There Yet?”

Lydia S. Lamb 1,2, Hao-Wen Sim 2,3,4 and Ann I. McCormack 1,2,3,*

1 Department of Endocrinology, St Vincent’s Hospital, Sydney 2010, New South Wales, Australia; l.lamb@garvan.org.au
2 Garvan Institute of Medical Research, Sydney 2010, New South Wales, Australia; hao-wen.sim@svha.org.au
3 St Vincent’s Clinical School, University of New South Wales, Sydney 2010, New South Wales, Australia
4 Kinghorn Cancer Centre, Darlinghurst 2010, New South Wales, Australia

* Correspondence: a.mccormack@garvan.org.au, Tel.: +61-2-922-958-489

Received: 7 January 2020; Accepted: 22 January 2020; Published: 28 January 2020

Abstract: Aggressive pituitary tumors account for up to 10% of pituitary tumors and are characterized by resistance to medical treatment and multiple recurrences despite standard therapies, including surgery, radiotherapy, and chemotherapy. They are associated with increased morbidity and mortality, particularly pituitary carcinomas, which have mortality rates of up to 66% at 1 year after diagnosis. Novel targeted therapies under investigation include mammalian target of rapamycin (mTOR), tyrosine kinase, and vascular endothelial growth factor (VEGF) inhibitors. More recently, immune checkpoint inhibitors have been proposed as a potential treatment option for pituitary tumors. An increased understanding of the molecular pathogenesis of aggressive pituitary tumors is required to identify potential biomarkers and therapeutic targets. This review discusses novel approaches to the management of aggressive pituitary tumors and the role of molecular profiling.

Keywords: aggressive pituitary tumors; pituitary carcinoma; molecular profiling; targeted therapy; immunotherapy

1. Introduction

Pituitary tumors account for approximately 10% to 15% of all intracranial tumors [1], with a prevalence of clinically significant pituitary tumors reported as 80 to 100 cases per 100,000 [2]. The majority are benign and slow growing; however, up to 10% have a more aggressive clinical course [3]. The definition of aggressive pituitary tumors (APT)s has been controversial. The previous edition of the World Health Organisation (WHO) Classification of Tumors of the Pituitary Gland (2004) classified three types of pituitary tumors, including typical adenoma, atypical adenoma, and pituitary carcinoma. Atypical adenomas were defined as tumors with histological features suggestive of aggressive clinical behavior, including an elevated mitotic index, proliferation marker Ki-67 labeling index >3%, and overexpression of tumour protein p53 by immunohistochemistry [4]. In the most recent WHO classification in 2017, the term atypical adenoma is no longer recommended due to a lack of evidence for its prognostic value [5–7]. It has been more recently proposed that APT be defined clinically as invasive tumors displaying an unusually rapid tumor growth rate or tumor growth despite optimal standard therapies, including surgery, radiotherapy, and chemotherapy [1–3]. The new WHO classification describes pituitary tumor subtypes along with histological markers,
such as Ki67 > 3%, that may suggest a potential for aggressive clinical behavior [5]. Pituitary carcinomas (PCs), defined by the presence of craniospinal or systemic metastases, are very rare, comprising just 0.2% of all pituitary tumors [1,5]. The mortality rates of PCs are known to be high, historically up to 66% at 1 year following diagnosis [8,9]. Recently, it has been demonstrated that the mortality associated with APTs approaches that of PCs, with mortality rates for APT of 28% and PC 42.5% at a median duration of 11 years following pituitary tumor diagnosis [3]. Temozolomide is the first chemotherapeutic agent to show activity against APTs and PCs and is now recommended as first-line chemotherapy [2,10]. Overall, temozolomide treatment is associated with a 37% radiological response rate in APTs and PCs, with demonstrated increased survival rates among responders [3,11]. However, 30% of patients demonstrate progression despite temozolomide treatment, and of those that respond, subsequent tumor progression is common after cessation of temozolomide [3]. Thus, there is an urgent need to identify alternative effective treatments to help manage this challenging condition.

Modern day medical oncology has seen a transition towards precision medicine and the use of targeted therapies in the management of cancer. Advances in tumor molecular profiling and increased use of next generation sequencing technology in clinical practice have facilitated the discovery of molecular drivers of cancer and development of treatments to target specific molecular signatures [12,13]. In the management of APTs and PCs, the use of novel targeted therapies has emerged somewhat incidentally on the back of successful application in other cancer types. In addition, there may be a role for immune checkpoint inhibitor therapies, which have revolutionized oncology practice and are now used across multiple cancer types. However, further understanding of the molecular pathogenesis of pituitary tumors will help inform the applicability of such novel therapies.

Herein, we will review the role of novel targeted therapies and initial experience with immunotherapy in the management of APT and PC and propose a focus on the development of molecular profiling of these tumors to help rationalize treatment modalities.

2. Targeted Therapies

Growth factors and their receptors, intracellular signaling pathways, and proteins involved in cell cycle regulation are all known to have important roles in pituitary tumorigenesis and thus are potential targets for therapy (Figure 1, Tables 1 and 2).
Figure 1. Growth factors, such as vascular endothelial growth factor (VEGF), epidermal growth factor (EGF), and fibroblast growth factor (FGF), bind receptor tyrosine kinases (RTKs) on the cell surface. This initiates intracellular signaling via the P13K/Akt/mTOR and Raf/Mek/ERK pathways, which promotes cell cycle progression within the cell nucleus. Cell cycle progression is activated by cyclins binding to cyclin-dependent kinases and inhibited by a number of proteins, including p16, p21, p27, p57, as well as p53, which acts by upregulating p21. Pituitary tumour transforming gene (PTTG) is an oncogene involved in multiple cellular processes, including induction of growth factors VEGF and FGF; downregulation of cell cycle inhibitors, such as p21; and modulation of cell cycle progression. Targeted therapies have been developed, which interrupt the cellular signaling pathway at different points and act to downregulate cellular differentiation and proliferation in cancer.
Table 1. Published cases of targeted therapies for aggressive pituitary tumors.

| Author                  | Age | Gender | Subtype                | Tumor molecular targets                      | Previous chemotherapy | Drug                  | Duration of treatment (months) | Concurrent treatment | Outcome                             |
|-------------------------|-----|--------|------------------------|----------------------------------------------|------------------------|-----------------------|------------------------------|------------------------|-------------------------------------|
| Ortiz et al. 2012       | 44  | Male   | Silent ACTH Carcinoma | VEGF immunoreactivity in tumor cell cytoplasm | Temozolomide           | Bevacizumab           | 26                          | None                   | SD over 26 months                   |
| O’Riordan et al. 2017   | 25  | Female | ACTH Carcinoma         | None                                         | Temozolomide           | Bevacizumab           | 6                           | Pasireotide            | Reduction in ACTH SD               |
| Rotman et al. 2019      | 51  | Male   | ACTH                   | NR                                           | None                   | Bevacizumab           | 26                          | Radiotherapy            | Biochemical cure SD >8 years        |
| Touma et al. 2017       | 63  | Male   | ACTH                   | NR                                           | None                   | Bevacizumab           | 2                           | Radiotherapy            | CR 5 years                         |
| Kurowska et al. 2015    | 56  | Female | ACTH                   | NR                                           | Temozolomide           | Bevacizumab           | NR                          | None                   | Died from neurosurgical complications |
| McCormack et al. 2018   | 9   | Male   | Immuno-negative        | NR                                           | Temozolomide           | Bevacizumab           | 4                           | Temozolomide           | PR at 4 months                    |
| McCormack et al. 2018   | 46  | Male   | ACTH                   | NR                                           | Temozolomide           | Bevacizumab           | 9                           | None                   | PR at 9 months                    |
| McCormack et al. 2018   | 20  | Male   | PRL                    | NR                                           | Temozolomide           | Bevacizumab           | 9                           | Temozolomide           | PD                                  |
| McCormack et al. 2018   | 55  | Male   | Immuno-negative        | NR                                           | Temozolomide           | Bevacizumab           | 5                           | None                   | SD at 5 months                    |
| McCormack et al. 2018   | 52  | Male   | ACTH                   | NR                                           | Temozolomide           | Bevacizumab           | 2                           | None                   | PD                                  |
| McCormack et al. 2018   | 4   | Male   | GH                     | NR                                           | None                   | Bevacizumab           | 21                          | Temozolomide           | PR at 21 months                   |
| Wang et al. 2019        | 41  | Female | GH                     | VEGFR-2                                      | None                   | Apatinib              | 12                          | Temozolomide           | CR                                  |
| McCormack et al. 2018   | NR  | NR     | NR                     | NR                                           | NR                     | Sunitinib             | NR                          | NR                     | PD                                  |
| Study, Year | Gender | Tumor Type | Mutation(s) | salvage therapy | Duration | Response |
|-------------|--------|------------|-------------|-----------------|----------|----------|
| Cooper et al. 2019 | NR Female | PRL | NR | Lapatanib | 6 | None |
| Cooper et al. 2019 | NR Female | PRL | NR | Lapatanib | 6 | None |
| Cooper et al. 2019 | NR Female | PRL | NR | Lapatanib | 6 | None |
| Cooper et al. 2019 | NR Female | PRL | NR | Lapatanib | 6 | None |
| Cooper et al. 2019 | NR Male | PRL | NR | Lapatanib | 6 | None |
| Cooper et al. 2019 | NR Male | PRL | NR | Lapatanib | 6 | None |
| McCormack et al. 2018 | NR | NR | NR | Lapatanib | NR | NR |
| McCormack et al. 2018 | NR | NR | NR | Lapatanib | NR | NR |
| McCormack et al. 2018 | NR | NR | NR | Erlotinib | NR | NR |
| Zhang et al. 2019 | 68 Male | PRL | Increased p-AKT, p-4EBP1 and p-S6 | Everolimus | 16 | Cabergoline |
| Donovan et al. 2016 | 46 Female | ACTH Carcinoma | STK11 (F298L), NOTCH1 (R1672H), FGFR2(P443A) and PDGRFRB (A713T) mutation | Everolimus | 7 | Capecitabine |
| Jouanneau et al. 2011 | 45 Male | ACTH Carcinoma | Activation of AKT1, inactivation of PI3K, overexpression CCND1 | Temozolomide | Everolimus | 3 | Octreotide |
| McCormack et al. 2018 | NR | NR | NR | Everolimus | NR | NR |
| McCormack et al. 2018 | NR | NR | NR | Everolimus | NR | NR |
| Target | Drugs | Outcomes in other cancers | Results of pre-clinical studies | In vitro therapeutic studies | In vivo therapeutic studies |
|--------|-------|---------------------------|---------------------------------|-----------------------------|-----------------------------|
| VEGF targeted therapy | Bevacizumab (VEGFA inhibitor) | Prolongs PFS in metastatic CRC, cervical cancer, non-small cell lung cancer, ovarian cancer, mesothelioma, and metastatic RCC. Improved PFS and OS with apatinib in gastric, breast, and lung cancer. | • Increased VEGF expression in APT compared with non-aggressive pituitary tumors  
• Octreotide reduces VEGF expression and secretion in GH-secreting adenomas | N/A | Reduced tumor size, prolactin secretion, vascularity and intra-tumoral hemorrhage in prolactin-secreting tumors. |
| EGFR targeted therapy | mAbs against EGFR | Improved survival in non-small cell lung cancer, metastatic colorectal cancer, head and neck, pancreatic, and breast cancer. | • Increased EGFR expression in APT/PCs  
• Increased EGFR expression in corticotroph adenomas | Gefitinib suppresses POMC and ACTH in murine and canine corticotroph adenoma cell cultures  
Gefitinib suppresses expression POMC in human corticotroph adenoma cell cultures  
• Lapatinib suppresses prolactin mRNA and secretion and reduces tumor volume in vitro in human prolactinoma cell lines and in vivo in rat models. | Gefitinib attenuates growth and ACTH secretion |

Table 2. Overview of targeted therapy for aggressive pituitary tumours and pituitary carcinomas. Therapeutic targets, established drugs, outcomes in other cancers, evidence from pre-clinical studies relating to therapeutic targets in pituitary tumors, and in vitro and in vivo studies of targeted therapy in pituitary tumors.
| FGF targeted therapy | FGFR TKIs in development | Ongoing investigation in pre-clinical and clinical trials | N/A | N/A |
|----------------------|--------------------------|---------------------------------------------------------|------|------|
| • 60% of pituitary adenomas express FGFR-4 isoform, which is not identified in normal pituitary | • Increased FGFR-4 in macroadenomas and correlation with Ki67 and tumor invasiveness | • Increased expression of FGF2 and FGFR-1 in PAs and significantly increased FGFR-1 in invasive tumors | • Downregulation of FGFR-2 in PAs |

| Raf/Mek/ERK pathways | BRAF inhibitors (e.g., Vemurafenib, dabrafenib) | Improved PFS and OS in melanoma | • B-Raf mRNA expression and pMEK1/2 and pERK1/2 are increased in pituitary adenomas compared to normal pituitaries | • Treatment of murine corticotroph pituitary tumor cells with vemurafenib resulted in a greater reduction of ACTH in the cells expressing the V600E BRAF mutation compared to tumor cells with wild-type BRAF |
|----------------------|-------------------------------------------------|---------------------------------|---------------------------------|----------------------------------------------------------|
| • Akt expression, pAkt, and Akt activity are increased in pituitary tumors compared with normal pituitary tissue | • PI3KCA gene amplifications in pituitary tumors | • PI3KCA gene mutations are associated with invasive tumors and tumor recurrence | • In rat GH3 cell lines, Rapamycin, Everolimus, and PI3K inhibitors decrease cell viability and proliferation. | • The pan-PI3K inhibitor Buparlisib reduces tumor volume, prolactin secretion and mitotic index in rat models of prolactin tumors. |
| PI3K/Akt/mTOR | mTOR inhibitors prolong PFS in renal cell carcinoma, neuroendocrine tumors and advanced breast cancer | mTOR inhibitors | • Akt expression, pAkt, and Akt activity are increased in pituitary tumors compared with normal pituitary tissue | • Dual mTOR/P13K inhibitor reduces cell proliferation and promotes cell death in GH3 cells and prolactin secreting cell cultures | • The combination of everolimus and cabergoline inhibits GH3 cell proliferation and prolactin levels. |
| mTOR inhibitors (e.g., Rapamycin and everolimus) | | | • Akt expression, pAkt, and Akt activity are increased in pituitary tumors compared with normal pituitary tissue | • Dual mTOR/P13K inhibitor reduces cell proliferation and promotes cell death in rat NFPAs | • The combination of dual PI3K/mTOR inhibition and temozolomide synergistically inhibits tumor growth and reduces GH/PRL levels in pituitary adenoma cell lines and in a mouse GH3 tumor model. |
### Notch Signaling Pathway

| Agents targeting Notch are in development | Notch 3 receptor and its ligand Jagged1 are increased in NFPAs and PRLs compared with normal pituitary |
|------------------------------------------|---------------------------------------------------------------------------------------------------|

- Response demonstrate in Phase 1 and 2 clinical trials in CRC, breast, lung, ovarian and papillary thyroid cancer, anaplastic astrocytoma, sarcoma, glioblastoma multiforme, and melanoma.

### Hedgehog Signaling Pathway

| Vismodegib | Increased OS in metastatic BCC |
|-------------|--------------------------------|

- In PA cell cultures exogenous SHH increased secretion of GH, PRL, and ACTH from their respective tumors
- Administration of SHH in corticotroph cell lines exerted anti-proliferative effects
- Administration of SHH inhibitor increased proliferation in GH3 cell lines

### Cell Cycle-Targeted Therapy

| CDK 4/6 inhibitors | Prolong PFS in estrogen receptor positive breast cancer. |
|---------------------|---------------------------------------------------------|

- Reductions in pRb and p16 or increased expression of cyclin D1 are observed in up to 80% tumors
- Cyclin D1 is over expressed in aggressive NFPAs
- Cyclin E is over expressed and p27 reduced in Cushing’s disease
- Mutations to p53 are demonstrated in corticotroph adenomas
- R-roscovitine (cyclin E/CDK2 inhibitor) reduces cell number, induces cell cycle arrest, induces senescence and reduces ACTH expression and secretion in mouse ACTH-secreting pituitary cells.
- R-roscovitine demonstrated a reduction in tumor size and serum and tumor ACTH expression in mice with corticotroph tumors.

### PTTG

| N/A | N/A |

- PTTG is overexpressed in approximately 90% PAs compared with low or no expression in normal pituitary tissue
- PTTG correlates with Ki67 and tumor invasiveness and aggression
- Overexpression of c-terminal truncated PTTG in rat prolactin- and GH-secreting pituitary tumor GH3 cells suppressed prolactin promotor activity, prolactin mRNA expression and hormonal levels.
- Injecting rats with c-terminal-truncated PTTG-transfected GH3 cells resulted in smaller tumors.
### Pituitary Tumor Epigenetics

| Zebularine (DNMT) | Trichostatin A (HDAC) | N/A |
|-------------------|-----------------------|-----|
| • Multiple epimutations have been identified in pituitary adenomas | • Reversal of epigenetic changes and re-expression of EFEMP1 gene with zebularine and TSA in AtT-20 and GH3 cell lines | N/A |
| • Reversal of epigenetic changes and restored expression of BMP-4 with zebularine and TSA in AtT-20 and GH3 cell lines. | • Reversal of epigenetic changes and re-expression of HMGA with zebularine and TSA in GH3 cell lines. | N/A |

### ICI therapy

| Anti PD-1, anti PD-L1, Anti CTLA4 antibodies | Effective and approved for use in the treatment of melanoma, lung cancer, RCC, head and neck SCC, lymphoma, and urothelial carcinoma | Pituitary tumors express PD-L1 and CD8+ tumor infiltrating lymphocytes with higher PD-L1 expression in functioning adenomas and a correlation between PD-L1 expression and hormonal levels and Ki67 |
|--------------------------------------------|-------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------|

Aggressive pituitary tumor (APT), pituitary carcinoma (PC), vascular endothelial growth factor (VEGF), vascular endothelial growth factor receptor (VEGFR) progression free survival (PFS), overall survival (OS), colorectal cancer (CRC), renal cell carcinoma (RCC), growth hormone (GH), epidermal growth factor receptor (EGFR), monoclonal antibodies (mABs), tyrosine kinase inhibitors (TKIs), fibroblast growth factor (FGF), fibroblast growth factor receptor (FGFR), pituitary adenoma (PA), non-functioning pituitary adenomas (NFPAs), basal cell carcinoma (BCC), sonic hedgehog (SHH), pituitary tumor transforming gene (PTTG), DNA methyltransferase (DNMT), histone deacetylase (HDAC), EGF containing fibulin-like extracellular matrix protein (EFEMP1), high mobility group A (HMGA), immune checkpoint inhibitor (ICI), programmed cell death protein 1 (PD-1), programmed death ligand 1 (PD-L1), cytotoxic T lymphocyte associated protein 4 (CTLA4).
3. Growth Factor-Targeted Therapies

3.1. Inhibition of Vascular Endothelial Growth Factor (VEGF)

Vascular endothelial growth factor (VEGF) plays a key role in tumorigenesis by promoting angiogenesis and vascular permeability as well as modulating the tumor immune microenvironment [14,15]. VEGF contributes to the pro-tumor immunosuppressive microenvironment by inhibiting dendritic cell maturation, accumulating myeloid-derived suppressor cells, and inducing T regulatory cells (T reg), as well as increasing the expression of inhibitory receptors on cytotoxic CD8+ T cells, including programmed cell death protein 1 (PD-1), cytotoxic T lymphocyte associated protein 4 (CTLA-4), T cell immunoglobulin and mucin domain containing protein 3 (TIM-3), and lymphocyte activation gene 3 (LAG-3) [16,17]. VEGF is upregulated in many human tumors [15] by common genetic events, which lead to malignant transformation, such as a loss of tumor suppressor genes and activation of oncogenes. Hypoxia, a common neoplastic characteristic, is also a driver of VEGF expression and upregulation of VEGF tyrosine kinase receptors (VEGFRs) VEGFR-1 and VEGFR-2 [18]. In turn, VEGFR-1 and to a greater extent VEGFR-2 are major transducers of important oncogenic downstream signaling pathways, such as phosphoinositide 3-kinase (PI3K)/Akt and mitogen activated protein kinase (MAPK) pathways [14].

Targeted therapy with antibodies directed at VEGF and VEGFR are now used in the treatment of many cancers [14]. Bevacizumab, a recombinant monoclonal antibody given intravenously, blocks VEGFA and inhibits the interaction between VEGF and VEGFR. It prolongs progression-free survival in metastatic colorectal cancer, cervical cancers, non-small cell lung, ovarian cancer, mesothelioma, and metastatic renal cell carcinoma [19]. However, in some cancers, bevacizumab has not been demonstrated to increase survival and the explanation for this lack of improvement remains unknown [19]. Apatinib, an oral therapy, inhibits VEGFR2 predominantly but has mild inhibitory effects on other tyrosine kinase receptors, such as c-Kit and c-Src [20]. Clinical trials have demonstrated improved progression-free and overall survival with apatinib treatment in gastric, breast, and lung cancer [20].

The mechanism of action of anti-VEGF therapy is multifactorial and includes inhibition of angiogenesis, induction of epithelial cell apoptosis, direct effect on tumor cells expressing VEGF receptors, modulation of the immune system with enhanced cytotoxic T cell and dendritic cell activity, and reduced activity and accumulation of immunosuppressive T reg cells, tumor-associated macrophages, and myeloid-derived suppressor cells [17,21].

Markers of tumor angiogenesis, such as VEGF expression, have been found to be higher in APT compared with non-aggressive pituitary adenomas [22–25]. VEGF expression and secretion is reduced by octreotide in growth hormone (GH)-secreting adenomas and pasireotide in non-functioning (NF) adenomas in vitro, suggesting that somatostatin analogues may inhibit pituitary tumor cell viability by inhibiting the action of VEGF [26,27]. In mice with and without multiple endocrine neoplasia 1 (MEN1), anti-VEGF therapy reduces tumor size, prolactin secretion, and vascularity in prolactin-secreting adenomas [28–30] as well as normalizing blood vessels and reducing intra-tumoral hemorrhage [30].

Thirteen cases have been described of APT or PC targeted with anti-VEGF or anti-VEGFR therapy. Nine cases have been reported to respond to treatment with the VEGF antibody bevacizumab. Four of these patients were treated concurrently with temozolomide and bevacizumab with or without radiotherapy, including one receiving a second course of temozolomide and five cases commenced bevacizumab following progressive disease on temozolomide, including one case in combination with pasireotide [3,31–34]. One case has been reported of an aggressive GH-secreting adenoma responding to treatment with a combination of temozolomide and the VEGFR-2 inhibitor apatinib, with normalization of GH and a >90% reduction in the tumor size [35]. Two cases have been reported of progressive disease on bevacizumab following previous chemotherapy. In addition, one case with progressive disease on the VEGFR inhibitor sunitinib is reported [3]. Based on these outcomes, VEGF-targeted therapy is a promising therapeutic option; however, further investigation is required to identify predictors of response to treatment and guide patient selection.
3.2. Inhibition of Epidermal Growth Factor Receptors (EGFRs)

The epidermal growth factor receptor (EGFR) family comprises EGFR (ErbB1/HER1), ErbB2 (ErbB2/HER2/Neu), Erb B3 (ErbB3/HER3), and Erb B4 (ErbB4/HER4), a group of membrane receptor tyrosine kinases that promote cell proliferation and suppress apoptosis [36]. Mutations to and overexpression of EGFR commonly occurs in cancer and leads to the upregulation of the downstream PI3K/Akt/mammalian target of rapamycin (mTOR) and Raf/mitogen-activated protein kinase (Mek)/extracellular signal-regulated kinase (ERK) pathways, which promote cancer cell proliferation [36–38]. Increased EGFR expression has been associated with poorer clinical outcome in a number of malignancies, including bladder, breast, lung, and head and neck cancers [38].

Given the role of EGFR and its signaling pathways in oncogenesis, two types of EGFR-targeted therapies have been developed. Humanized monoclonal antibodies (mAbs) against the EGFR extracellular domain block ligand binding and mediate downregulation of EGFR while tyrosine kinase inhibitors (TKIs) bind to the receptor pocket, excluding ATP and thus preventing signal transduction [36,38]. These therapies have been shown to improve survival in EGFR-positive cancers and have been approved for use in non-small-cell lung cancer, metastatic colorectal cancer, head and neck cancer, pancreatic cancer, and breast cancer [36,39]. Predictors of response to EGFR-targeted therapy have been examined. In patients with non-small cell lung cancers, high levels of EGFR protein expression or increased EGFR copy numbers derive greater therapeutic benefit from EGFR-directed mAbs [40,41]. Mutations in exon 18-21 of EGFR cause variable activation of EGFR in the absence of ligand binding, driving tumorigenesis. The majority of these mutations confer sensitivity to EGFR blockade, with rare activating mutations conferring resistance [41]. Tumor analysis for EGFR mutations is therefore routine in clinical practice to predict response to therapy in lung adenocarcinoma [42].

In pituitary tissue, EGFR is expressed at variable levels in normal pituitary as well as functioning and non-functioning pituitary adenomas, with higher expression demonstrated in invasive adenomas and carcinomas [37,43,44]. EGFR overexpression in corticotroph adenomas is associated with downstream activation of the ERK pathway. The expression levels of EGFR were found to correlate with adrenocorticotropic hormone (ACTH) and cortisol levels as well as tumor recurrence [45]. Increased EGFR expression in corticotroph adenomas has been attributed to recently identified somatic mutations in ubiquitin-specific peptidase 8 (USP8), a deubiquitinase enzyme that protects EGFR from degradation. USP8 mutations are located in the 14-3-3 protein binding motif and enhance the proteolytic cleavage and catalytic activity of USP8, resulting in increased EGFR signaling. USP8-mutated tumors have demonstrated higher levels of proopiomelanocortin (POMC), the precursor to ACTH [46,47]. Interestingly, surgical remission rates are higher in patients with USP8-mutated tumors, which may reflect these tumors are smaller and thus may suggest USP8 mutations are not a critical factor in the development of aggressive tumor behavior. There are likely to be other, currently unidentified, factors that may also contribute to EGFR signaling in pituitary tumors. Nevertheless, these findings raise the possibility of EGFR-directed therapy, particularly in corticotroph adenomas.

Gefitinib, a TKI-targeting EGFR, suppresses expression of POMC and ACTH secretion in murine and canine corticotroph adenoma cell cultures, suppresses expression of POMC in human corticotroph adenoma cell cultures, as well as attenuating the growth and ACTH secretion of EGFR-expressing tumors in vivo in mice [48]. In GH3 rat lactosomatotroph pituitary tumor cells transfected with EGFR2, lapatinib, a dual EGFR/ErbB1 and HER2/ErbB2 TKI, has been shown to suppress prolactin mRNA expression and secretion, and reduce tumor volume to a greater effect than gefitinib in both in vivo rat models and in vitro human prolactinoma cell lines [49].

To date, seven cases have been reported of lapatinib therapy for aggressive prolactinomas, with variable results. Tumor size decreased by 22% in one patient, remained stable in three, and increased in three. Prolactin level decreased in three patients, with a median decrease of 42%, but increased in the other three cases [3,50]. Lapatinib has also been used unsuccessfully in one case of an aggressive silent GH tumor and an aggressive ACTH-secreting adenoma with disease progression [3].
3.3. Inhibition of Fibroblast Growth Factor

Fibroblast growth factor (FGF) and their receptors are involved in the regulation of cellular proliferation, survival, migration, and differentiation, and play a key role in angiogenesis. Dysregulation of FGF signaling contributes to oncogenesis by driving cellular proliferation and promoting angiogenesis [51,52]. FGF receptor (FGFR) mutations and amplifications have been implicated in a number of cancers, including multiple myeloma, bladder, cervical, prostate, breast, and gastric cancer [51]. Paradoxically, FGF-2 has been demonstrated to inhibit cellular proliferation and promote apoptosis in certain human breast cancer cell lines as well as being associated with good prognostic indicators when over expressed in some breast cancers [52]. In addition, FGFR-2 expression is associated with decreased tumor progression in some tumors, such as astrocytomas, bladder, prostate, and thyroid carcinomas [52].

In normal pituitary tissue, FGF has been reported to be produced by the folliculostellate cells and regulates GH, prolactin, and TSH secretion [53]. There are four distinct FGFRs (FGFR 1, 2, 3, and 4); however, normal pituitary tissue expresses the mRNA for only FGFR-1, FGFR-2, and FGFR-3 along with the secretable immunoglobulin-like domain of FGFR-4 [54,55]. In around 60% of pituitary adenomas, an N-terminally truncated FGFR-4 isoform has been identified, which includes a transmembrane and kinase domain not identified in normal pituitary tissue. The expression of pituitary tumor-derived FGFR-4 is increased in macroadenomas compared with microadenomas and correlates with Ki67 and tumor invasiveness [55]. In pituitary adenomas, higher expression of FGF2 and FGFR-1 has been demonstrated in comparison with normal pituitary, with significantly more enhanced expression of FGFR-1 in invasive tumors [56]. In contrast, FGFR-2 has been shown to be downregulated in pituitary adenomas due to FGFR-2 promoter methylation; however, this has not been clinically assessed in relation to tumor invasiveness or aggressiveness [57].

Therapies targeting FGF and their receptors have been proposed in the management of cancer, and FGFR tyrosine kinase inhibitors are currently being investigated in preclinical and clinical trials [58]. The complex involvement of FGF and their receptors in the pathogenesis of pituitary tumors suggests there may be a role for therapies targeting this pathway in the treatment of pituitary tumors, although this has not yet been investigated.

4. Targets of Intracellular Signaling Pathways

4.1. PI3K/Akt/mTOR and Raf/Mek/ERK Pathways

Under normal circumstances, cellular regulation is under the control of complex intracellular signaling pathways, including the PI3K/Akt/mTOR and Raf/Mek/ERK pathways, which regulate cell cycle proliferation and differentiation, cell survival, protein synthesis, and cellular metabolism [59,60] (Figure 1). Signal transduction is initiated by growth factors activating receptor tyrosine kinases (RTKs). In the PI3K/Akt/mTOR pathway, RTKs activate the signal transduction component lipid kinase, PI3K. This then activates Akt, which phosphorylates several downstream targets, leading to cell proliferation [59–61]. In the Raf/Mek/ERK pathway, Ras GTPase stimulation results in Raf phosphorylation and activation of MEK1 and MEK2, which in turn phosphorylate and activate ERK1 and ERK2. ERK signaling targets multiple kinases, phosphatases, transcription factors, and cytoskeletal proteins, resulting in the modulation of genes associated with cell proliferation [60,61]. A major downstream target of both the PI3K/Akt/mTOR and Raf/Mek/ERK pathway is mTOR, which stimulates cell growth. Mutations in these pathways, in mTOR itself and in the mTOR complexes mTORC1 and mTORC2, account for up to 30% of all human cancers [60,62].

Therapies that inhibit mTOR prolong progression-free survival and have been approved for use in multiple cancers, including renal cell carcinoma, neuroendocrine tumors, and advanced breast cancer, with trials ongoing in other malignancies [63–66]. mTOR inhibitors seem to demonstrate better efficacy in renal cell carcinoma and breast cancer than in other malignancies, which is interesting as they share similar genetic alterations in the mTOR signaling pathway, including mutations to PI3KCA and mutations and deletions of phosphatase and tensin homolog (PTEN). This may suggest a role for genetic and molecular analysis to predict response to treatment [67]. Potential
biomarkers of response to mTOR inhibition, including loss of PTEN function, AKT phosphorylation, and PI3K, mTOR, tuberous sclerosis complex (TSC) 1 or 2 mutations [40,63,68]; and those that predict resistance, including overexpression of the apoptosis-inhibitory protein Bcl-2 and KRAS, BRAF, and TSC mutations [40,63]. Validation of these biomarkers in clinical trials is required.

Both the PI3K/Akt/mTOR and Raf/Mek/ERK pathways are upregulated in pituitary tumors and implicated in pituitary tumorigenesis [61]. The \textit{PIK3CA} gene encodes PIK3CA of class IA PI3K and \textit{PIK3CA} gene mutations and amplifications cause increased enzymatic activity of PIK3CA, activation of AKT signaling, and growth factor-independent growth, invasion, and metastasis of cancer cells [69]. Amplifications of the \textit{PIK3CA} gene are observed in invasive and non-invasive pituitary tumors, with a prevalence of 20% to 40% [69,70]. \textit{PIK3CA} mutations are present in pituitary tumors [70] and are particularly associated with invasive tumors, with one study demonstrating somatic mutations in 9% of invasive pituitary tumors but none among the cohort of non-invasive tumors. Furthermore, the presence of a \textit{PIK3CA} mutation was associated with increased recurrence rates [69]. Akt mRNA expression, phosphorylated Akt, and Akt activity are increased in pituitary tumors compared with normal pituitary tissue [61,71,72]. The expression of B-Raf mRNA and phosphorylated MEK1/2 and ERK1/2 are also significantly higher in pituitary adenomas compared to normal pituitaries [61,73]. Interestingly, \textit{H-ras} point mutations have been reported in distant metastatic pituitary tumor deposits of three patients but not in their respective primary pituitary tumors, suggesting that \textit{H-ras} mutations may have a unique role in the development of pituitary metastases [74]. The oncogenic \textit{V600E BRAF} mutation has been identified in 16.5% of corticotroph adenomas but not in other types of pituitary adenomas. In a murine corticotroph cell line, increased phosphorylation of ERK1/2 was demonstrated in cells expressing the \textit{BRAF} mutation, consistent with increased MAPK activity. These cells also exhibited increased levels of \textit{POMC} mRNA [75].

Given the important role of the PI3K/Akt/mTOR and Raf/Mek/ERK pathways in pituitary tumor pathogenesis, a number of agents targeting these pathways have been investigated in the treatment of pituitary adenomas; however, most evidence of activity comes from preclinical studies. Treatment of murine corticotroph pituitary tumor cells with the \textit{BRAF} inhibitor vemurafenib resulted in a greater reduction of ACTH in the cells expressing the \textit{V600E BRAF} mutation compared to tumor cells with wild-type \textit{BRAF}, suggesting a potential role for \textit{BRAF} inhibitor therapy in patients with \textit{BRAF}-mutated tumors [75]. Rapamycin is a lipophilic macroline that acts to inhibit mTOR, leading to a reduction in cell cycle progression and cell proliferation [62,76]. In rat GH-secreting pituitary tumor GH3 cell lines, rapamycin, its bioavailable analog RAD001 (Everolimus), and PI3K inhibitors have been shown to decrease cell viability and proliferation through inhibition of mTOR activity and interruption of normal cell cycle function [77,78]. These effects appear to occur without elimination of the p70S6K negative feedback loop, which leads to treatment resistance in other cancers. However, in human GH-secreting pituitary adenoma cell lines, everolimus has produced a reduction in cell viability in one study but not in another [77,78]. The dual mTOR/PI3K inhibitor NVP-BEZ235 reduces cell proliferation and promotes cell death in rat non-functioning pituitary adenomas in vitro and in vivo [79], in growth hormone-secreting GH3 cell lines in vitro, and in primary cell cultures of human prolactinomas [80]. The pan-PI3K inhibitor NVP-BKM120 (Buparlisib) has demonstrated no significant effect on GH3 cells in vitro but did reduce the tumor volume, prolactin secretion, and mitotic index in rat models of prolactin tumors in vivo [80]. Synergies may also be effective. The combination of everolimus and cabergoline was shown to inhibit mammosomatotroph tumor GH3 cell proliferation and prolactin levels in vitro [72]. The combination of dual PI3K/mTOR inhibition and temozolomide has been demonstrated to synergistically inhibit tumor growth and reduce GH/PRL levels in pituitary adenoma cell lines and in a mouse GH3 tumor model [81]. In other cancers, including glioblastoma, melanoma, and renal cell carcinoma, this combination has been effective in overcoming temozolomide resistance [81].

Clinical experience with everolimus in pituitary tumors is limited to case reports with variable outcomes. Two cases have been described with biochemical and radiological response to everolimus [72,82]. Both cases had abnormalities demonstrated in the mTOR signaling pathway, including an STK11 mutation in an ACTH-secreting PC and increased p-Akt expression in an aggressive
prolactinoma [72,82]. In contrast, in another case of a recurrent ACTH-secreting PC, there was no response to the combination of everolimus and octreotide [83]. This may have been due to the development of rapamycin resistance due to the feedback of rapamycin on the phosphorylation of Akt, a mechanism that seems to be absent in GH- and prolactin-secreting tumors [83]. Three other cases reported had progressive disease despite treatment with everolimus [3].

4.2. Notch and Hedgehog Signaling Pathways

The Notch signaling pathway regulates normal cell proliferation and differentiation and is of relevance in cancer biology, affecting multiple tumor cell types, including cancer stem cells, immune, endothelial, and tumor cells [84]. Agents targeting the Notch signaling pathway are under investigation in the treatment of cancer and response has been demonstrated in phase 1 and 2 clinical trials in colorectal, breast, lung, ovarian and papillary thyroid cancer, anaplastic astrocytoma, sarcoma, glioblastoma multiforme, and melanoma [84,85].

The hedgehog signaling pathway is implicated in the maintenance and homeostasis of stem cells and regulates cell proliferation, survival, and angiogenesis [84]. Aberrations in the hedgehog pathway have been implicated in many cancers, including pancreatic, colon, gastric, lung, breast and prostate cancer, basal cell carcinoma, medulloblastoma rhabdosarcoma, leukemia, and multiple myeloma [86]. Vismodegib, which is an antagonist of the smoothened (SMO) receptor in the hedgehog pathway, increases overall survival in metastatic basal cell carcinoma (BCC) and has been approved for use in patients with metastatic or locally advanced BCC who are not candidates for surgery or radiotherapy. Clinical trials are ongoing to examine the role of Vismodegib and another SMO inhibitor Saridegib in other cancers; however, no benefit has yet been demonstrated in gastric, lung, colorectal, ovarian, pancreatic cancer, myelofibrosis, or chondrosarcoma [84].

The Notch and hedgehog pathway are involved in normal pituitary development and there is evidence for their role in pituitary tumor pathogenesis [87]. The Notch 3 receptor and its ligand Jagged1 are increased in non-functioning pituitary adenomas and prolactinomas compared with normal pituitary [88,89]. The expression of the signaling protein sonic hedgehog (SHH) is downregulated in pituitary adenomas and absent in corticotroph adenomas compared with normal pituitary. Furthermore, administration of SHH in AtT20 corticotroph cells exerted antiproliferative effects whereas administration of a SHH inhibitor increased proliferation in GH3 mammosomatotropinomas [90]. These findings suggest that downregulation of the hedgehog pathway may play a role in the pathogenesis of pituitary adenomas [87]. On the other hand, in primary cell cultures established from pituitary adenomas, exogenous SHH increased secretion of GH, prolactin, and ACTH from somatotropinomas, lactotropinomas, and Cushing’s tumors [90]. Further research is required to understand the role of the Notch and hedgehog signaling pathways in pituitary pathogenesis as they may represent novel targets for therapy in the management of APT and PC, although this has not yet been investigated.

5. Cell Cycle-Targeted Therapy

Normal cellular proliferation relies on progression through the cell cycle under the control of cyclin dependent kinases (CDKs), which are modulated by activators (cyclins) and inhibitors (Ink4, and Cip and Kip inhibitors), composed of p21, p27, and p57 [91]. In normal functioning cells, CDK 4 and 6 form a complex with cyclin D1 to initiate phosphorylation of retinoblastoma tumor suppressor protein (Rb), which releases suppression of E2F transcription factors. Activated E2F results in the transcription of genes, including cyclin E, which binds CDK2, forming a complex that promotes cell cycle progression from G1 to S. This process is regulated by p16, a CDK inhibitor, which when activated, inhibits the CDK4/6 and cyclin D1 complex, leading to hypophosphorylated Rb and halting cell cycle progression [92,93]. The tumor suppressor gene p53 also plays an important role in cell cycle regulation by upregulating the expression of the CDK inhibitor p21 [91]. Cell cycle disruption with unregulated cell cycle progression is a common feature of human cancer [91–94].

Therapies targeting the cell cycle have recently been developed for use in human cancers. In particular, CDK4/6 inhibitors prolong progression-free survival and are now approved for use in
estrogen receptor-positive breast cancer [95,96]. Cyclin D1 deficiency has been shown to impair mammary epithelial proliferation [97] while cyclin D1 and CDK4 are necessary for breast cancer development in mice [98] and cyclin D1 is overexpressed in the majority of human breast cancers [99].

In pituitary tumors, reductions in pRb and p16 or increased expression of cyclin D1 are observed in up to 80% of tumors [61,100,101]. In particular, cyclin D1 is overexpressed in aggressive, non-functioning tumors [102] while cyclin E is overexpressed and p27 reduced in Cushing’s disease [102,103]. Mutations in the tumor suppressor gene TP53 have been identified rarely in corticotroph adenomas and may provide a therapeutic target in the future [104–106]. Clinical trials into the restoration of wild-type p53 in other cancer types are ongoing, but this has not been explored in APT. R-roscovitine is a cell cycle inhibitor that acts on cyclin E/CDK2 to cause cell cycle arrest. In mouse ACTH-secreting pituitary cells, treatment with R-roscovitine reduced the cell number and induced cell cycle arrest, demonstrated by decreased cyclin E, increased p27Kip1, p57Kip2, and p21Cip1 expression, as well as reduced Thr821 phosphorylation of Rb [107]. R-roscovitine treatment also induced senescence as evidenced by increased β-gal expression, and caused a reduction in ACTH expression and secretion. In vivo, mice with an induction of corticotroph tumors treated with R-roscovitine demonstrated a reduction in tumor size as well as serum and tumor ACTH expression [107]. Ander et al. reported an interesting case that supports a potential role for cell cycle inhibition in the treatment of pituitary tumors. A 71-year-old female with an incidental pituitary macroadenoma under observation was treated with the CDK4/6 inhibitor palbociclib for metastatic breast cancer, which resulted in significant regression of the pituitary tumor [94].

6. Pituitary Tumor-Transforming Gene (PTTG)

Pituitary tumor-transforming gene (PTTG) is an oncogene involved in multiple cellular processes, including cellular proliferation, DNA repair, angiogenesis, and induction of genetic instability [108]. PTTG interferes with normal cell cycle progression via complex mechanisms, including inhibition of sister chromatid separation in anaphase, interaction with the transcription factor Sp1 to mediate S1/G phase transition, interaction with the p53 pathway, and suppression of the activity of CDK inhibitor p21. PTTG is also involved in angiogenesis via induction of FGF and VEGF [108]. Overexpression of PTTG contributes to cellular transformation and tumor development and has been implicated in endocrine and non-endocrine tumors, including those of the pituitary, thyroid, ovary, breast, prostate, lung, esophagus, colon, and central nervous system [108,109]. PTTG is overexpressed in approximately 90% of pituitary adenomas compared with low or no expression in normal pituitary tissue and PTTG has been demonstrated to correlate with Ki67 and tumor invasiveness and aggression [109–111].

Targeting PTTG has been explored in prolactinoma, glioma, follicular thyroid, and cervical cancer cell lines as well as in vivo mouse models of lung cancer and hepatoma and demonstrate the therapeutic promise of PTTG inhibition [108]. In rat prolactin- and GH-secreting pituitary tumor cell lines, overexpression of c-terminal-truncated PTTG suppressed prolactin promoter activity, mRNA expression, and hormone levels. In addition, injecting rats with GH3 cells transfected with truncated PTTG resulted in smaller tumors compared with those injected with the control vector [112]. These findings demonstrate how the interruption of PTTG action may have therapeutic potential in the management of prolactinomas.

7. Pituitary Tumor Epigenetics

Epigenetics is defined as a process that heritably influences gene expression without genetic change to the underlying gene sequence and occurs via modifications, including methylation of cytosine bases in DNA, modification of histone proteins, and expression of microRNA [113,114]. A number of drugs have been developed for use in various tumors types that reverse DNA methylation and histone modifications through the inhibition of the enzymes responsible for these changes, including DNA methyltransferases (DNMTs) and histone deacetylases (HDACs) [115].
In pituitary tumors, a number of epimutations have been identified that affect the transcription of hormone and growth factor receptors, signal transduction pathway molecules, transcription factors, and cell cycle regulators. Usually, these include changes to the methylation or modification of histones [116].

Research into targeting epigenetic changes in the management of pituitary tumors is limited. The majority of pituitary adenomas have reduced expression of the EGF-containing fibulin-like extracellular matrix protein (EFEMP1) gene due to epigenetic silencing, which is reversed in AtT-20 and GH3 cell lines incubated with the DNMT inhibitor zebularine and the HDAC inhibitor trichostatin A (TSA). There was, however, no effect on cell proliferation or apoptotic end points [117]. Bone morphogenetic protein-4 (BMP-4) has been proposed as both an oncogene and a tumor suppressor gene in pituitary adenomas. Reduced BMP-4 expression is associated with epigenetic changes, which are reversed in AtT-20 and GH3 cell lines incubated with the epiprep drugs zebularine and TSA [118]. Similarly, overexpression of high mobility group A (HMGA) genes, a frequent finding in pituitary adenomas, results from epigenetic changes that downregulate microRNA targeting HMGA genes. Epiprep drugs also revert these modifications [119]. The clinical implications of these findings remain to be determined.

8. Immune Checkpoint Inhibitor Therapies

The significant advances in cancer therapies over the recent years have been based on an increased understanding of the tumor immune microenvironment and development of immunotherapies to target and modulate the immune response. Immune checkpoint inhibitors include anti-CTLA4, anti-PD-1 and anti-programmed death ligand 1 (PD-L1) antibodies, which target CTLA4 and PD-1 receptors on the surface of T cells, and PD-L1 expressed on the surface of tumor cells, blocking the subsequent inhibitory signal, and resulting in increased T cell activation and anti-tumor immune response (Figure 2). These agents are effective and have been approved for use in the treatment of multiple malignancies, including melanoma, lung cancer, renal cell carcinoma, squamous cell head and neck cancer, lymphoma, and urothelial carcinoma [120].

**Figure 2.** T cell activation requires presentation of tumor antigen to the T cell receptor (TCR) by major histocompatibility complex (MHC) on the antigen-presenting cell (APC) and also co-stimulation via B7 on the APC binding to CD28 on the T cell. Cytotoxic T lymphocyte associated protein 4 (CTLA4) is an inhibitory molecule that binds B7 with a higher infinity than CD28 and downregulates the T cell. In addition, PD1 on the T cell binds PD-L1 expressed by tumor cells, which also acts to downregulate the T cell. Immune checkpoint inhibitors, including anti-CTLA4, anti-PD1, and anti-PD-L1
monoclonal antibodies, block the interaction of B7 with CTLA4 and PD1 with PD-L1, eliminating the inhibitory signal and upregulating the T cell.

Pituitary tumors have been demonstrated to express PD-L1 and CD8+ tumor-infiltrating lymphocytes with higher PD-L1 expression in functioning adenomas and a correlation between PD-L1 expression, hormone levels, and Ki67 [121–123]. These findings provide a rationale for exploring the role of immunotherapy in the management of APTs. However, just one case has been reported to date in the literature of a patient with a pituitary carcinoma responding to treatment with the combination checkpoint inhibitor therapy of ipilimumab and nivolumab [124]. The authors presented a case of a 35-year-old female with an aggressive ACTH-secreting pituitary adenoma that initially responded to temozolomide and capecitabine prior to presentation with a liver metastasis. She subsequently received ipilimumab and nivolumab, resulting in a 92% reduction in the volume of the liver metastasis, 59% reduction in the intracranial tumor volume, and normalization of ACTH levels. Interestingly, subsequent genetic analysis of the hepatic metastasis demonstrated development of a hypermutated phenotype (5275 mutations, or 93 mutations/Mb) classic for temozolomide exposure, including an MSH6 mutation. It was postulated that a temozolomide-induced hypermutated tumor may be more sensitive to immune checkpoint inhibitor therapy. It is worth noting that the liver metastasis had low (<1%) PD-L1 expression by immunohistochemistry [124].

This case suggests there may be a role for immune checkpoint inhibitor therapy in the management of APT and PC. The immune microenvironment of pituitary tumors as well as predictors of response in pituitary tumors remain poorly understood and represent new avenues for research.

9. Active Clinical Trials

There are four registered active clinical trials examining treatments for patients with APTs. Three trials examining the use of immune checkpoint inhibitor therapies are recruiting, including a phase II trial of Nivolumab Plus Ipilimumab in People with Aggressive Pituitary Tumors (Memorial Sloane Kettering Cancer Centre, United States), Capecitabine and Temozolomide for Treatment of Recurrent Pituitary Adenomas (Weill Cornell Medical College, United States), and Nivolumab and Ipilimumab in Treating Patients with Rare Tumors (University of Alabama et al., United States) [125–127]. One study entitled Targeted Therapy with Lapatanib in Patients with Recurrent Pituitary Tumors Resistant to Standard Therapy (Cedars-Sinai Medical Center et al., United States) examined EGFR-targeted therapy and has been completed, although as yet, no results are available [128].

10. Conclusions

There is certainly a healthy preclinical research base that provides rationale for exploring a range of targeted therapies, as well as immunotherapy, in the management of APTs. The role of these therapies in APT, however, remains unclear in clinical practice. Unfortunately, rare tumors, such as APTs, are significantly underserved with regard to clinical trial opportunities. Advancement is limited while case reports and occasional case series provide for a slow progression in this field. Routine molecular profiling of APTs may both guide treatment decisions and provide justification for access to targeted therapies used routinely in other cancer types. The future may look bright, but for APTs, we are definitely not there yet.

Author Contributions: All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.
1. Dworakowska, D.; Grossman, A.B. Aggressive and malignant pituitary tumours: State-of-the-art. *Endocr. Relat. Cancer* 2018, 25, R559–R575.

2. Raverot, G.; Burman, P.; McCormack, A.; Heaney, A.; Petersenn, S.; Popovic, V.; Trouillas, J.; Dekkers, O.M.; Endocrinology, T.E.S.O. European Society of Endocrinology Clinical Practice Guidelines for the management of aggressive pituitary tumours and carcinomas. *Eur. J. Endocrinol.* 2018, 178, G1–G24.

3. McCormack, A.; Dekkers, O.M.; Petersenn, S.; Popovic, V.; Trouillas, J.; Raverot, G.; Burman, P.; Hubalewska-Dydejezky, A.; Assie, G.; Bach, L.; et al. Treatment of aggressive pituitary tumours and carcinomas: Results of a European Society of Endocrinology (ESE) survey 2016. *Eur. J. Endocrinol.* 2018, 178, 265–276.

4. IARC; World Health Organization; IA. *Pathology and Genetics of Tumours of Endocrine Organs*; IARC: Lyon, France, 2004; p. 324.

5. Lloyd, R.; Osamura, R.; Kloppel, G.; Rosai, J. *WHO Classification of Tumours of Endocrine Organs*, 4th ed.; IARC: Lyon, France, 2017; Volume 1.

6. Inoshita, N.; Nishioka, H. The 2017 WHO classification of pituitary adenoma: Overview and comments. *Brain Tumor Pathol.* 2018, 35, 51–56.

7. Mete, O.; Lopes, M.B. Overview of the 2017 WHO Classification of Pituitary Tumors. *Endocr. Pathol.* 2017, 28, 228–243.

8. Yoo, F.; Kuan, E.C.; Heaney, A.P.; Bergsneider, M.; Wang, M.B. Corticotrophic pituitary carcinoma with cervical metastases: Case series and literature review. *Pituitary* 2018, 21, 290–301.

9. Pernicone, P.J.; Scheithauer, B.W.; Sebo, T.J.; Kovacs, K.T.; Horvath, E.; Young, W.F.; Lloyd, R.V.; Davis, D.H.; Guthrie, B.L.; Schoene, W.C. Pituitary carcinoma: A clinicopathologic study of 15 cases. *Cancer* 1997, 79, 804–812.

10. McCormack, A.I.; Wass, J.A.H.; Grossman, A.B. Aggressive pituitary tumours: The role of temozolomide and the assessment of MGMT status. *Eur. J. Clin. Investig.* 2011, 41, 1133–1148.

11. Lasolle, H.; Cortet, C.; Castinetti, F.; Cloix, L.; Caron, P.; Delember, B.; Desailloud, R.; Jublanc, C.; Lebrun-Frenay, C.; Sadoul, J.-L.; et al. Temozolomide treatment can improve overall survival in aggressive pituitary tumors and pituitary carcinomas. *Eur. J. Endocrinol.* 2017, 176, 769–777.

12. Chae, Y.K.; Pan, A.P.; Davis, A.A.; Patel, S.P.; Carneiro, B.A.; Kurzrock, R.; Giles, F.J. Path toward Precision Oncology: Review of Targeted Therapy Studies and Tools to Aid in Defining “Actionability” of a Molecular Lesion and Patient Management Support. *Mol. Cancer Ther.* 2017, 16, 2645–2655.

13. Re, A.; Nardella, C.; Quattrone, A.; Lunardi, A. Editorial: Precision Medicine in Oncology. *Front. Oncol.* 2018, 8, 479.

14. Goel, H.L.; Mercurio, A.M. VEGF targets the tumour cell. *Nat. Rev. Cancer* 2013, 13, 871–882.

15. Ferrara, N.; Gerber, H.-P.; LeCouter, J. The biology of VEGF and its receptors. *Nat. Med.* 2003, 9, 669–676.

16. Voron, T.; Colussi, O.; Marcheteanu, E.; Pernot, S.; Nizard, M.; Pointet, A.L.; Latreche, S.; Bergaya, S.; Benhamouda, N.; Tanoch, C.; et al. VEGF-A modulates expression of inhibitory checkpoints on CD8+ T cells in tumors. *J. Exp. Med.* 2015, 212, 139–148.

17. Yang, J.; Yan, J.; Liu, B. Targeting VEGF/VEGFR to Modulate Antitumor Immunity. *Front. Immunol.* 2018, 9, 978.

18. Ferrara, N. Vascular endothelial growth factor as a target for anticancer therapy. *Oncologist* 2004, 9, 2–10.

19. Zirlik, K.; Duyster, J. Anti-Angiogenics: Current Situation and Future Perspectives. *Oncol. Res. Treat.* 2018, 41, 166–171.

20. Zhang, H. Apatinib for molecular targeted therapy in tumor. *Drug Des. Devel. Ther.* 2015, 9, 6075–6081.

21. Ellis, L.M.; Hicklin, D.J. VEGF-targeted therapy: Mechanisms of anti-tumour activity. *Nat. Rev. Cancer* 2008, 8, 579–591.

22. Lloyd, R.V.; Scheithauer, B.W.; Kuroki, T.; Vidal, S.; Kovacs, K.; Stefaneanu, L. Vascular Endothelial Growth Factor (VEGF) Expression in Human Pituitary Adenomas and Carcinomas. *Endocr. Pathol.* 1999, 10, 229–235.

23. Wang, Y.; Li, J.; Tohti, M.; Hu, Y.; Wang, S.; Li, W.; Lu, Z.; Ma, C. The expression profile of Dopamine D2 receptor, MGMT and VEGF in different histological subtypes of pituitary adenomas: A study of 197 cases and indications for the medical therapy. *J. Exp. Clin. Cancer Res.* 2014, 33, 56.
24. Sánchez-Ortiga, R.; Sánchez-Tejada, L.; Moreno-Perez, O.; Riesgo, P.; Niveiro, M.; Picó Alfonso, A.M. Over-expression of vascular endothelial growth factor in pituitary adenomas is associated with extrasellar growth and recurrence. *Pituitary* 2013, 16, 370–377.

25. Cristina, C.; Perez-Millan, M.I.; Luque, G.; Dulce, R.A.; Sevlever, G.; Berner, S.I.; Becu-Villalobos, D. VEGF and CD31 Association in Pituitary Adenomas. *Endocr. Pathol.* 2010, 21, 154–160.

26. Kurosaki, M.; Saeger, W.; Abe, T.; Lüdecke, D.K. Expression of vascular endothelial growth factor in growth hormone-secreting pituitary adenomas: Special reference to the octreotide treatment. *Neurol. Res.* 2008, 30, 518–522.

27. Zatelli, M.C.; Piccin, D.; Vignali, C.; Tagliati, F.; Ambrosio, M.R.; Bondanelli, M.; Cimino, V.; Bianchi, A.; Schmid, H.A.; Scarnarini, M.; et al. Pasireotide, a multiple somatostatin receptor subtypes ligand, reduces cell viability in non-functioning pituitary adenomas by inhibiting vascular endothelial growth factor secretion. *Endocr. Relat. Cancer* 2007, 14, 91–102.

28. Luque, G.M.; Perez-Millan, M.I.; Ornstein, A.M.; Cristina, C.; Becu-Villalobos, D. Inhibitory Effects of Antivascular Endothelial Growth Factor Strategies in Experimental Dopamine-Resistant Prolactinomas. *J. Pharmacol. Exp. Ther.* 2011, 337, 766–774.

29. Korsisaari, N.; Ross, J.; Wu, X.; Kowanetz, M.; Pal, N.; Hall, L.; Eastham-Anderson, J.; Forrest, W.F.; Van Bruggen, N.; Peale, F.V.; et al. Blocking Vascular Endothelial Growth Factor-A Inhibits the Growth of Pituitary Adenomas and Lowers Serum Prolactin Level in a Mouse Model of Multiple Endocrine Neoplasia Type 1. *Clin. Cancer Res.* 2008, 14, 249–258.

30. Chauvet, N.; Romanò, N.; Lafont, C.; Guilhou, A.; Galibert, E.; Bonnemont, P.; Fedele, M.; Fusco, A.; Mollard, P.; et al. Complementary actions of dopamine D2 receptor agonist and anti-vegf therapy on tumoral vessel normalization in a transgenic mouse model. *Int. J. Cancer* 2017, 140, 2150–2161.

31. Rotman, L.E.; Vaughan, T.B.; Hackney, J.R.; Riley, K.O. Long-Term Survival After Transformation of an Adrenocorticotropic Hormone–Secreting Pituitary Macroadenoma to a Silent Corticotroph Pituitary Carcinoma. *World Neurosurg.* 2019, 122, 417–423.

32. Touma, W.; Hoostal, S.; Peterson, R.A.; Wiernik, A.; Santacruz, K.S.; Lou, E. Successful treatment of pituitary carcinoma with concurrent radiation, temozolomide, and bevacizumab after resection. *J. Clin. Neurosci.* 2017, 41, 75–77.

33. O’Riordan, L.M.; Greally, M.; Coleman, N.; Breathnach, O.S.; Hennessy, B.; Thompson, C.J.; Grogan, W. Metastatic ACTH-producing pituitary carcinoma managed with combination pasireotide and bevacizumab following failure of temozolomide therapy: A case report. *JCO* 2013, 31, e13022.

34. Ortiz, I.D.; Syro, L.V.; Scheithauer, B.W.; Ensen, A.; Uribé, H.; Fadul, C.E.; Rotondo, F.; Horvath, E.; Kovacs, K. Anti-VEGF therapy in pituitary carcinoma. *Pituitary* 2011, 15, 445–449.

35. Wang, Y.; He, Q.; Meng, X.; Zhou, S.; Zhu, Y.; Xu, J.; Tao, R. Apatinib (YN968D1) and Temozolomide in Recurrent Invasive Pituitary Adenoma: Case Report and Literature Review. *World Neurosurg.* 2019, 124, 319–322.

36. Wee, P.; Wang, Z. Epidermal Growth Factor Receptor Cell Proliferation Signaling Pathways. *Cancers* 2017, 9, 52.

37. Cooper, O.; Vlotides, G.; Fukuoka, H.; Greene, M.I.; Melmed, S. Expression and function of ErbB receptors and ligands in the pituitary. *Endocr. Relat. Cancer* 2011, 18, R197–R211.

38. Mendelsohn, J.; Baselga, J. Status of Epidermal Growth Factor Receptor Antagonists in the Biology and Treatment of Cancer. *J. Clin. Oncol.* 2003, 21, 2787–2799.

39. Harandi, A.; Zaidi, A.S.; Stocker, A.M.; Laber, D.A. Clinical Efficacy and Toxicity of Anti-EGFR Therapy in Common Cancers. *J. Oncol.* 2009, 2009, 1–14.

40. Bonomi, P.; Gandara, D.; Hirsch, F.; Kerr, K.; Obasaju, C.; Paz-Ares, L.; Bellomo, C.; Bradley, J.; Jr, P.B.; Culligan, M.; et al. Predictive biomarkers for response to EGFR-directed monoclonal antibodies for advanced squamous cell lung cancer. *Ann. Oncol.* 2018, 29, 1701–1709.

41. Prudkin, L.; Wistuba, I.I. Epidermal growth factor receptor abnormalities in lung cancer. Pathogenetic and clinical implications. *Ann. Diagn. Pathol.* 2006, 10, 306–315.

42. Lindeman, N.I.; Cagle, P.T.; Aisner, D.L.; Arcila, M.E.; Beasley, M.B.; Bernicker, E.H.; Colasacco, C.; Dacic, S.; Hirsch, F.R.; Kerr, K.; et al. Updated Molecular Testing Guideline for the Selection of Lung Cancer Patients for Treatment with Targeted Tyrosine Kinase Inhibitors: Guideline from the College of American Pathologists, the International Association for the Study of Lung Cancer, and the Association for Molecular Pathology. *Arch. Pathol. Lab. Med.* 2018, 142, 321–346.
43. Cooper, O.; Mamelak, A.; Bannykh, S.; Carmichael, J.; Bonert, V.; Lim, S.; Cook-Wiens, G.; Ben-Shlomo, A. Prolactinoma ErbB receptor expression and targeted therapy for aggressive tumors. *Endocrine* 2013, 46, 318–327.

44. Onguru, O.; Scheithauer, B.W.; Kovacs, K.; Vidal, S.; Jin, L.; Zhang, S.; Ruebel, K.H.; Lloyd, R.V. Analysis of epidermal growth factor receptor and activated epidermal growth factor receptor expression in pituitary adenomas and carcinomas. *Mod. Pathol.* 2004, 17, 772–780.

45. Liu, X.; Feng, M.; Dai, C.; Bao, X.; Deng, K.; Yao, Y.; Wang, R. Expression of EGFR in Pituitary Corticotroph Adenomas and Its Relationship with Tumor Behavior. *Front. Endocrinol.* 2019, 10, 785.

46. Ma, Z.Y.; Song, Z.J.; Chen, J.H.; Wang, Y.F.; Li, S.Q.; Zhou, L.F.; Mao, Y.; Li, Y.M.; Hu, R.G.; Zhang, Z.Y.; et al. Recurrent gain-of-function USP8 mutations in Cushing’s disease. *Cell Res.* 2015, 25, 306–317.

47. Reincke, M.; Sbiera, S.; Hayakawa, A.; Theodoropoulou, M.; Oswald, A.; Beuschlein, F.; Meitinger, T.; Mizuno-Yamasaki, E.; Kawaguchi, K.; Saeki, Y.; et al. Mutations in the deubiquitinase gene USP8 cause Cushing’s disease. *Nat. Genet.* 2015, 47, 31–38.

48. Fukuoka, H.; Cooper, O.; Ben-Shlomo, A.; Mamelak, A.; Ren, S.-G.; Bruyette, D.; Melmed, S. EGFR as a therapeutic target for human, canine, and mouse ACTH-secreting pituitary adenomas. *J. Clin. Investig.* 2011, 121, 4712–4721.

49. Fukuoka, H.; Cooper, O.; Mizutani, J.; Tong, Y.; Ren, S.G.; Bannykh, S.; Melmed, S. HER2/ErbB2 Receptor Signaling in Rat and Human Prolactinoma Cells: Strategy for Targeted Prolactinoma Therapy. *Mol. Endocrinol.* 2011, 25, 92–103.

50. Cooper, O.; Bonert, V.; Rudnick, J.; Pressman, B.; Melmed, S. SUN-442 EGFR/ErbB2 Targeted Therapy for Aggressive Prolactinomas. *J. Endocr. Soc.* 2019, 3, 442.

51. Turner, N.; Grose, R. Fibroblast growth factor signalling: From development to cancer. *Nat. Rev. Cancer.* 2010, 10, 116–129.

52. Korc, M.; Friesel, R.E. The role of fibroblast growth factors in tumor growth. *Curr. Cancer Drug Targets* 2009, 9, 639–651.

53. Ezzat, S.; Smyth, H.S.; Ramyar, L.; Asa, S.L. Heterogenous in vivo and in vitro expression of basic fibroblast growth factor by human pituitary adenomas. *J. Clin. Endocrinol. Metab.* 1995, 80, 878–884.

54. Abbass, S.A.A.; Asa, S.L.; Ezzat, S. Altered Expression of Fibroblast Growth Factor Receptors in Human Pituitary Adenomas. *J. Clin. Endocrinol. Metab.* 1997, 82, 7.

55. Qian, Z.R.; Sano, T.; Asa, S.L.; Yamada, S.; Horiguchi, H.; Tashiro, T.; Li, C.C.; Hirokawa, M.; Kovacs, K.; Ezzat, S. Cytoplasmic Expression of Fibroblast Growth Factor Receptor-4 in Human Pituitary Adenomas: Relation to Tumor Type, Size, Proliferation, and Invasiveness. *J. Clin. Endocrinol. Metab.* 2004, 89, 1904–1911.

56. McCabe, C.J.; Khaira, J.S.; Boelaert, K.; Heaney, A.P.; Tannahill, L.A.; Hussain, S.; Mitchell, R.; Olliff, J.; Sheppard, M.C.; Franklyn, J.A.; et al. Expression of pituitary tumour transforming gene (PTTG) and fibroblast growth factor-2 (FGF-2) in human pituitary adenomas: Relationships to clinical tumour behaviour. *Clin. Endocrinol.* 2003, 58, 141–150.

57. Zhu, X.; Lee, K.; Asa, S.L.; Ezzat, S. Epigenetic Silencing through DNA and Histone Methylation of Fibroblast Growth Factor Receptor 2 in Neoplastic Pituitary Cells. *Am. J. Pathol.* 2007, 170, 1618–1628.

58. Loriot, Y.; Necchi, A.; García-Donas, J.; Huddart, R.; Burgess, E.; Fleming, M.; Rezazadeh, A.; Mellado, B.; Varlamov, S.; et al. Erdafitinib in Locally Advanced or Metastatic Urothelial Carcinoma. *J. Hematol. Oncol.* 2019, 318–318.
64. Baselga, J.; Campone, M.; Piccart, M.; Burris, H.A.; III; Rugo, H.S.; Sahmoud, T.; Noguchi, S.; Gnant, M.; Pritchard, K.I.; Lebrun, F.; et al. Everolimus in Postmenopausal Hormone-Receptor–Positive Advanced Breast Cancer. *N. Engl. J. Med.* 2012, 366, 520–529.

65. Motzer, R.J.; Escudier, B.; Oudard, S.; Hutson, T.E.; Porta, C.; Bracarda, S.; Grünwald, V.; Thompson, J.A.; Figlin, R.A.; Hollaender, N.; et al. Efficacy of everolimus in advanced renal cell carcinoma: A double-blind, randomised, placebo-controlled phase III trial. *Lancet* 2008, 372, 449–456.

66. Hudes, G.; Carducci, M.; Tomczak, P.; Dutcher, J.; Figlin, R.; Kapoor, A.; Staroslaw ska, E.; Sosman, J.; McDermott, D.; Bodrogi, I.; et al. Temsirolimus, Interferon Alfa, or Both for Advanced Renal-Cell Carcinoma. *N. Engl. J. Med.* 2007, 356, 2271–2281.

67. Tian, T.; Li, X.; Zhang, J. mTOR Signaling in Cancer and mTOR Inhibitors in Solid Tumor Targeting Therapy. *Int. J. Mol. Sci.* 2019, 20, 755.

68. Kwiatkowski, D.J.; Choueiri, T.K.; Fay, A.P.; Rini, B.I.; Thorner, A.R.; De Velasco, G.; Tyburnczy, M.E.; Hamieh, L.; Albigès, L.; Agarwal, N.; et al. Mutations in TSC1, TSC2, and MTOR Are Associated with Response to Rapalogs in Patients with Metastatic Renal Cell Carcinoma. *Clin. Cancer Res.* 2016, 22, 2445–2452.

69. Lin, Y.; Jiang, X.; Shen, Y.; Li, M.; Ma, H.; Xing, M.; Lu, Y. Frequent mutations and amplifications of the PIK3CA gene in pituitary tumors. *Endocrinol. Relat. Cancer* 2009, 16, 301–310.

70. Murat, C.; Braga, P.; Fortes, M.; Bronstein, M.; Correa-Giannella, M.; Giorgi, R. Mutation and genomic amplification of the PIK3CA proto-oncogene in pituitary adenomas. *Braz. J. Med. Biol. Res.* 2012, 45, 851–855.

71. Muşat, M.; Korbonits, M.; Kola, B.; Borboli, N.; Hanson, M.R.; Nanzer, A.M.; Grigson, J.; Jordan, S.; Morris, D.G.; Gueorguiev, M.; et al. Enhanced protein kinase B/Akt signalling in pituitary tumours. *Endocr. Relat. Cancer* 2005, 12, 423–433.

72. Zhang, N.; Way, J.S.; Zhang, X.; Sergey, M.; Bergsneider, M.; Wang, M.B.; Yong, W.H.; Heaney, A.P. Effect of Everolimus in Treatment of Aggressive Prolactin-Secreting Pituitary Adenomas. *J. Clin. Endocrinol. Metab.* 2019, 104, 1929–1936.

73. Ewing, J.; Pedder-Smith, S.; Franchi, G.; Ruscica, M.; Emery, M.; Vax, V.; Garcia, E.; Czirják, S.; Hanzely, Z.; Kola, B.; et al. A mutation and expression analysis of the oncogene BRAF in pituitary adenomas. *Clin. Endocrinol.* 2007, 66, 348–352.

74. Pei, L.; Melmed, S.; Schethauer, B.; Kovacs, K.; Prager, D. H-ras mutations in human pituitary carcinoma metastases. *J. Clin. Endocrinol. Metab.* 1994, 78, 842–846.

75. Chen, J.; Jian, X.; Deng, S.; Ma, Z.; Shou, X.; Shen, Y.; Zhang, Q.; Song, Z.; Li, Z.; Peng, H.; et al. Identification of recurrent USP48 and BRAF mutations in Cushing’s disease. *Nat. Commun.* 2018, 9, 3171.

76. Benjamin, D.; Colombi, M.; Moroni, C.; Hall, M.N. Rapamycin passes the torch: A new generation of mTOR inhibitors. *Nat. Rev. Drug Discov.* 2011, 10, 868–880.

77. Gorshtein, A.; Rubinfeld, H.; Kendler, E.; Theodoropoulou, M.; Cerovac, V.; Stalla, G.K.; Cohen, Z.R.; Hadani, M.; Shimon, I. Mammalian target of rapamycin inhibitors rapamycin and RAD001 (everolimus) induce anti-proliferative effects in GH-secreting pituitary tumor cells in vitro. *Endocr. Relat. Cancer* 2009, 16, 1017–1027.

78. Pivonello, C.; Fatalano, R.; Solari, D.; Auriemma, R.S.; Frio, F.; Vitulli, F.; Grasso, L.F.S.; Di Cera, M.; De Martino, M.C.; Cavallo, L.M.; et al. Effect of combined treatment with a pan-PI3K inhibitor or an isoform-specific PI3K inhibitor and everolimus on cell proliferation in GH-secreting pituitary tumour in an experimental setting. *Endocrine* 2018, 62, 663–680.

79. Lee, M.; Wiedemann, T.; Gross, C.; Leinhauser, I.; Roncaroli, F.; Braren, R.; Pellegrata, N.S. Targeting PI3K/mTOR Signaling Displays Potent Antitumor Efficacy against Nonfunctioning Pituitary Adenomas. *Clin. Cancer Res.* 2015, 21, 3204–3215.

80. Chanal, M.; Chevallier, P.; Raverot, V.; Fonteneau, G.; Lucia, K.; Garcia, J.L.M.; Rachwan, A.; Jouanneau, E.; Trouillas, J.; Honnorat, J.; et al. Differential Effects of PI3K and Dual PI3K/mTOR Inhibition in Rat Prolactin-Secreting Pituitary Tumors. *Mol. Cancer Ther.* 2016, 15, 1261–1270.

81. Dai, C.; Zhang, B.; Liu, X.; Ma, S.; Yang, Y.; Yao, Y.; Feng, M.; Bao, X.; Li, G.; Wang, J.; et al. Inhibition of PI3K/AKT/mTOR Pathway Enhances Temozolomide-Induced Cytotoxicity in Pituitary Adenoma Cell Lines in Vitro and Xenografted Pituitary Adenoma in Female Nude Mice. *Endocrinology* 2013, 154, 1247–1259.
Yu, Q.; Geng, Y.; Sicinski, P. Specific protection against breast cancers by cyclin D1 ablation.

Direct Role in Hormone Secretion and Cell Proliferation. Expression and Function of Sonic Hedgehog Pathway Components in Pituitary Adenomas: Evidence for a proteomic analyses. Molecular pathogenesis of human prolactinomas identified by gene expression profiling, RT-qPCR, and Evans, C.-O.; Moreno, C.S.; Zhan, X.; McCabe, M.T.; Vertino, P.M.; Desiderio, M.M.; Oyesiku, N.M. Requirement for CDK4 kinase function in breast cancer.

Breast. Yu, Q.; Sicinska, E.; Geng, Y.; Ahnström, M.; Zagozdzon, A.; Kong, Y.; Gardner, H.; Kiyokawa, H.; Harris, S.J.; Weinberg, R.A. Cyclin D1 Provides a Link between Development and Oncogenesis in the Retina and

Sicinski, P.; Donaher, J.L.; Parker, S.B.; Li, T.; Fazeli, A.; Gardner, H.; Haslam, S.Z.; Bronson, R.T.; Elledge, S.J.; Weinberg, R.A. Cyclin D1 Provides a Link between Development and Oncogenesis in the Retina and

Hormones and Wnt pathways in cancer stem cells: Clinical update. Nat. Rev. Clin. Oncol. 2015, 12, 445–464.

Venkatesh, V.; Nataraj, R.; Thangaraj, G.S.; Karthikeyan, M.; Gnanasekaran, A.; Kaginelli, S.B.; Kuppanna, G.; Kalappa, C.G.; Basalingappa, K.M. Targeting Notch signalling pathway of cancer stem cells. Stem Cell Invest. 2018, 5, 5.

Amakye, D.; Jagani, Z.; Dorsch, M. Unraveling the therapeutic potential of the Hedgehog pathway in cancer. Nat. Med. 2013, 19, 1410–1422.

Yavropoulou, M.P.; Maladaki, A.; Yovos, J.G. The role of Notch and Hedgehog signaling pathways in pituitary development and pathogenesis of pituitary adenomas. Hormones 2015, 14, 5–18.

Lu, R.; Gao, H.; Wang, H.; Cao, L.; Bai, J.; Zhang, Y. Overexpression of the Notch3 receptor and its ligand Jagged1 in human clinically non-functioning pituitary adenomas. Oncol. Lett. 2013, 5, 845–851.

Evans, C.-O.; Moreno, C.S.; Zhan, X.; McCabe, M.T.; Vertino, P.M.; Desiderio, M.M.; Oyesiku, N.M. Molecular pathogenesis of human prostates identified by gene expression profiling, RT-qPCR, and proteomic analyses. Pituitary 2008, 11, 231–245.

Vila, G.; Theodoropoulou, M.; Stalla, J.; Torn, J.C.; Løsa, M.; Renner, U.; Stalla, G.K.; Paez-Pereda, M. Expression and Function of Sonic Hedgehog Pathway Components in Pituitary Adenomas: Evidence for a Direct Role in Hormone Secretion and Cell Proliferation. J. Clin. Endocrinol. Metab. 2005, 90, 6687–6694.

Malumbres, M.; Barbacid, M. Cell cycle, CDKs and cancer: A changing paradigm. Nat. Rev. Cancer 2009, 9, 153–166.

Dowdy, S.F.; Hinds, P.W.; Louie, K.; Reed, S.I.; Arnold, A.; Weinberg, R.A. Physical interaction of the retinoblastoma protein with human D cyclins. Cell 1993, 73, 301–312.

Sellers, W.R.; Kaelin, W.G. Role of the retinoblastoma protein in the pathogenesis of human cancer. J. Clin. Oncol. 1995, 13, 3001–3312.

Anderson, E.; Heller, R.S.; Lechan, R.M.; Heilman, C.B. Regression of a nonfunctioning pituitary macroadenoma on the CDK4/6 inhibitor palbociclib: Case report. Neurosurg. Focus 2018, 44, E9.

Turner, N.C.; Ro, J.; André, F.; Loi, S.; Verma, S.; Iwata, H.; Harbeck, N.; Loibl, S.; Huang Bartlett, C.; Zhang, K.; et al. Palbociclib in Hormone-Receptor–Positive Advanced Breast Cancer. N. Engl. J. Med. 2015, 373, 209–219.

Hortobagyi, G.N.; Stemmer, S.M.; Burris, H.A.; Yap, Y.-S.; Sonke, G.S.; Paluch-Shimon, S.; Campone, M.; Blackwell, K.L.; André, F.; Winer, E.P.; et al. Ribociclib as First-Line Therapy for HR-Positive, Advanced Breast Cancer. N. Engl. J. Med. 2016, 375, 1738–1748.

Sicinski, P.; Donaher, J.L.; Parker, S.B.; Li, T.; Fazeli, A.; Gardner, H.; Haslam, S.Z.; Bronson, R.T.; Elledge, S.J.; Weinberg, R.A. Cyclin D1 Provides a Link between Development and Oncogenesis in the Retina and

Breast. Cell 1995, 82, 621–630.

Yu, Q.; Sicinska, E.; Geng, Y.; Ahnström, M.; Zagrozdzon, A.; Kong, Y.; Gardner, H.; Kiyokawa, H.; Harris, L.N.; Stål, O.; et al. Requirement for CDK4 kinase function in breast cancer. Cancer Cell 2006, 9, 23–32.

Yu, Q.; Geng, Y.; Sicinski, P. Specific protection against breast cancers by cyclin D1 ablation. Nature 2001, 411, 1017–1021.

Simpson, D. Aberrant expression of G1/S regulators is a frequent event in sporadic pituitary adenomas. Carcinogenesis 2001, 22, 1149–1154.

Hibberts, N.A.; Simpson, D.J.; Bicknell, J.E.; Broome, J.C.; Hoban, P.R.; Clayton, R.N.; E Farrell, W. Analysis of cyclin D1 (CCND1) allelic imbalance and overexpression in sporadic human pituitary tumors. Clin. Cancer Res. 1999, 5, 2133–2139.

Jordan, S.; Lidhar, K.; Korbonits, M.; Lowe, D.G.; Grossman, A.B. Cyclin D and cyclin E expression in normal and adenomatous pituitary. Eur. J. Endocrinol. 2000, 143, R1–R6.

Roussel-Gervais, A.; Bilodeau, S.; Vallette, S.; Berthelet, F.; Lacroix, A.; Figarella-Branger, M.; Brue, T.; Drouin, J. Cooperation between Cyclin E and p27Kip1 in Pituitary Tumorigenesis. Mol. Endocrinol. 2010, 24, 1835–1845.
104. Sbiera, S.; Perez-Rivas, L.G.; Taranets, L.; Weigand, I.; Flitsch, J.; Graf, E.; Monoranu, C.M.; Saeger, W.; Hagel, C.; Honegger, J.; et al. Driver mutations in USP8 wild-type Cushing’s disease. Neuro Oncol. 2019, 21, 1273–1283.

105. Guo, F.; Wang, G.; Wang, F.; Xu, D.; Liu, X. Identification of Novel Genes Involved in the Pathogenesis of an ACTH-Secreting Pituitary Carcinoma: A Case Report and Literature Review. Front. Oncol. 2018, 8, 510.

106. Kawashima, S.T.; Usui, T.; Sano, T.; Iogawa, H.; Hagiwara, H.; Tamanaha, T.; Tagami, T.; Naruse, M.; Hojo, M.; Takahashi, J.A.; et al. P53 gene mutation in an atypical corticotroph adenoma with Cushing’s disease. Clin. Endocrinol. 2009, 70, 656–657.

107. Liu, N.-A.; Jiang, H.; Ben-Shlomo, A.; Wawrowsky, K.; Fan, X.-M.; Lin, S.; Melmed, S. Targeting zebrafish and murine pituitary corticotroph tumors with a cyclin-dependent kinase (CDK) inhibitor. Proc. Natl. Acad. Sci. USA 2011, 108, 8414–8419.

108. Salehi, F.; Kovacs, K.; Scheithauer, B.W.; Lloyd, R.V.; Cusimano, M. Pituitary tumor-transforming gene in endocrine and other neoplasms: A review and update. Endocr. Relat. Cancer 2008, 15, 721–743.

109. Trott, G.; Ongaratti, B.R.; Silva, C.B.D.O.; Abech, G.D.; Haag, T.; Rech, C.G.S.L.; Ferreira, N.P.; Oliveira, M.D.C.; Pereira-Lima, J.F.S. PTTG overexpression in non-functioning pituitary adenomas: Correlation with invasiveness, female gender and younger age. Am. J. Clin. Pathol. 2019, 41, 83–89.

110. Filippella, M.; Galland, F.; Kujas, M.; Young, J.; Faggiano, A.; Lombardi, G.; Colao, A.; Meduri, G.; Chanson, P. Pituitary tumour transforming gene (PTTG) expression correlates with the proliferative activity and recurrence status of pituitary adenomas: A clinical and immunohistochemical study. Clin. Endocrinol. 2006, 65, 536–543.

111. Zhang, X.; Horwitz, G.A.; Heaney, A.P.; Nakashima, M.; Prezant, T.R.; Bronstein, M.D.; Melmed, S. Pituitary Tumor Transforming Gene (PTTG) Expression in Pituitary Adenomas. None 1999, 84, 761–767.

112. Horwitz, G.A.; Miklovsky, I.; Heaney, A.P.; Ren, S.-G.; Melmed, S. Human Pituitary Tumor-Transforming Gene (PTTG1) Motif Suppresses Prolactin Expression. Mol. Endocrinol. 2003, 17, 600–609.

113. Eccleston, A.; DeWitt, N.; Gunter, C.; Marte, B.; Nath, D. Epigenetics. Nature 2007, 447, 395.

114. Farrell, W.E. Epigenetics of pituitary tumours: An update. Curr. Opin. Endocrinol. Diabetes Obes. 2014, 21, 299.

115. Sharma, S.; Kelly, T.K.; Jones, P.A. Epigenetics in cancer. Carcinogenesis 2009, 31, 27–36.

116. Yacqub-Uzman, K.; Richardson, A.; Duong, C.V.; Clayton, R.N.; Farrell, W.E. The pituitary tumour epigenome: Aberrations and prospects for targeted therapy. Nat. Rev. Endocrinol. 2012, 8, 486–494.

117. Duong, C.V.; Yacqub-Uzman, K.; Emes, R.; Clayton, R.N.; Farrell, W.E. The EFEMP1 Gene: A Frequent Target for Epigenetic Silencing in Multiple Human Pituitary Adenoma Subtypes. NEN 2013, 98, 200–211.

118. Yacqub-Uzman, K.; Duong, C.V.; Clayton, R.N.; Farrell, W.E. Epigenomic Silencing of the BMP-4 Gene in Pituitary Adenomas: A Potential Target for Epidrug-Induced Re-expression. Endocrinology 2012, 153, 3603–3612.

119. Kitchen, M.O.; Yacqub-Uzman, K.; Emes, R.D.; Richardson, A.; Clayton, R.N.; Farrell, W.E. Epidrug mediated re-expression of miRNA targeting the HMGA transcripts in pituitary cells. Pituitary 2015, 18, 674–684.

120. Havel, J.J.; Chowell, D.; Chan, T.A. The evolving landscape of biomarkers for checkpoint inhibitor immunotherapy. Nat. Rev. Cancer 2019, 19, 133–150.

121. Mei, Y.; Bi, W.L.; Greenwald, N.F.; Du, Z.; Agar, N.Y.R.; Kaiser, U.B.; Woodmansee, W.W.; Reardon, D.A.; Freeman, G.J.; Fecic, P.E.; et al. Increased expression of programmed death ligand 1 (PD-L1) in human pituitary tumors. Oncotarget 2016, 7, 76565–76576.

122. Wang, P.F.; Wang, T.J.; Yang, Y.K.; Yao, K.; Li, Z.; Li, Y.M.; Yan, C.X. The expression profile of PD-L1 and CD8+ lymphocyte in pituitary adenomas indicating for immunotherapy. J. Neuroendocrinol. 2018, 139, 89–95.

123. Wang, P.; Wang, T.; Yang, Y.; Yu, C.; Liu, N.; Yan, C. Detection of programmed death ligand 1 protein and CD8+ lymphocyte infiltration in plurihormonal pituitary adenomas: A case report and review of the literatures. Medicine 2017, 96, e9056.

124. Lin, A.L.; Jonsson, P.; Tabar, V.; Yang, T.J.; Cuaron, J.; Beal, K.; Cohen, M.; Postow, M.; Rosenblum, M.; Shia, J.; et al. Marked Response of a Hypermutated ACTH-Secreting Pituitary Carcinoma to Ipilimumab and Nivolumab. J. Clin. Endocrinol. Metab. 2018, 103, 3925–3930.

125. Nivolumab and Ipilimumab in People with Aggressive Pituitary Tumors—Full Text View—ClinicalTrials.gov. Available online: https://clinicaltrials.gov/ct2/show/NCT04042753 (accessed on 17 December 2019).
126. Nivolumab and Ipilimumab in Treating Patients with Rare Tumors—Full Text View—ClinicalTrials.gov. Available online: https://clinicaltrials.gov/ct2/show/NCT02834013 (accessed on 17 December 2019).

127. Capecitabine and Temozolomide for Treatment of Recurrent Pituitary Adenomas—Full Text View—ClinicalTrials.gov. Available online: https://clinicaltrials.gov/ct2/show/NCT03930771 (accessed on 17 December 2019).

128. Targeted Therapy with Lapatinib in Patients with Recurrent Pituitary Tumors Resistant to Standard Therapy—Full Text View—ClinicalTrials.gov. Available online: https://clinicaltrials.gov/ct2/show/NCT00939523 (accessed on 17 December 2019).

© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).