INTRODUCTION

In terms of species abundance and biomass, the sub-order Notothenioidei constitute the dominant fish group in the Southern Ocean (Eastman and Mc Cune, 2000). The suborder is thought to have evolved before the formation of the Antarctic Polar Front (APF), around 20 million years ago (Clarke and Johnston, 1996). Almost a hundred nototheniid species have an exclusively Antarctic distribution,
whereas there are 26 non-Antarctic species (Eastman and Eakin, 2000; Montgomery and Clements, 2000). Evidence from molecular phylogenetic analysis suggests that some families, namely Bovichtidae, Pseudaphritidae and Eleginopidae, spread out of Antarctic waters before the establishment of the APF (Stankovic et al., 2002). Similar analyses suggest that the barrier of APF was permeated in some opportunities allowing a successful migration from the Antarctic and colonisation of Subantarctic waters (Bargelloni et al., 2000). The largely non-Antarctic genus Patagonotothen resulted especially successful, giving rise to 14 species (Eastman, 1995).

Information about reproductive features such as maturation and spawning schedule, fecundity or reproductive effort of Subantarctic species of notothenioids is available only for a few species. Therefore, comparative analyses between species with Antarctic and Subantarctic distributions are hampered.

The energetic demands for reproduction and growth are conditioned by food intake and the physiological reproductive strategies (Calow, 1985). Low temperatures and highly seasonal productivity of the Antarctic marine environment modulate the energy budget of Antarctic fish and the seasonality of spawning. Some species of Antarctic Notothenioidei, like Nototthenia rossii, N. coriiceps and a great number of channichthyids, spawn a few thousand large yolky oocytes (more than 3 mm in diameter), usually during autumn, while other species of Nototheniidae spawn a greater number of small oocytes at any time of the year (Ekau, 1991; Kock and Wilhelms, 2003). Pseudochoaenichthys georgianus and Champsocephalus gunnari require about one year to complete the yolk deposition, while Chaenocephalus aceratus and N. coriiceps need a longer time (Everson et al., 1996; Everson, 1994; Kock and Everson, 1997; Kock and Kellermann, 1991). To understand the gonadal cycle and yolk deposition process, a more intense sampling schedule throughout the year would be necessary.

The reproductive characteristics of fish from the Southern Ocean were studied in many species, in some of which histological gonadal descriptions were made (Eastman and DeVries, 2000; Everson et al., 1991; La Mesa et al., 2003; Macchi and Barrera-Oro, 1995; Russo et al., 2000; Shandikov and Faleeva, 1992; Van der Molen and Matallanas, 2003; see Kock and Kellerman, 1991 and references therein). Most studies on reproduction were carried out only in summer–autumn (Kock and Kellermann, 1991; Duhamel et al., 1993; Kock et al., 2000; Kock and Wilhelms, 2003) because that is the season when most of the Antarctic scientific cruises take place.

Different attempts to determine the energy budget of fish species of the Southern Ocean have been made in recent years. Food intake requirements and feeding energetics were analysed by Kock (1992) in several species of notothenioids, by Johnston and Battram (1993) in Nototthenia neglecta and by Chekunova and Naumov (1982) in Nototthenia rossi marmorata. The proximal biochemical compositions of N. r. marmorata and Lepidonotothen (=Nototthenia) gibberifrons were studied by Kozlov (1981; 1982). The lipid compositions of high Antarctic nototheniid fish were studied by Friederich and Hagen (1994), Hagen et al. (2000) and Kamler et al. (2001).

In the present study we analysed the reproductive characteristics of Antarctic Notothenioidei species, comparing fish samples from the northern and southern branches of the Scotia Arc. In addition, we analysed the energetic content in gonads, liver and muscle of Antarctic notothenioids for the first time.

MATERIAL AND METHODS

Fishes were collected during the LAMPOS survey in April 2002 (Fig. 1) and detailed cruise information has been published by Arntz and Brey (2003) for the total number of fishes caught during the cruise and the number considered for the various investigations in this paper (see Tables 1 and 2). Shipboard sampling involved taxonomic identification, labelling and measurement of total length (TL) and standard length (SL) to the nearest cm below. Total weight (TW), sex, and weight of liver, stomach and gonads were determined to the nearest 0.1 g below. Gonado-somatic index (GSI) was calculated as the percentage of gonadal weight per total body weight.

Gonad samples for histological studies were preserved in Bouin fixative, dehydrated and included in Paraplast. Histological sections (5 to 7 µm thick) were stained with haematoxylin-eosin and Periodic Acid-Schiff-haematoxylin (PAS-H).

Absolute fecundity was estimated by extrapolating the number of oocytes of 3 weighed subsamples to the weight of the entire ovary. The number of oocytes produced per gram of total weight (relative fecundity) was also determined. A portion of ovary
was preserved in 10% formaldehyde to determine the frequency distribution of the oocyte diameters.

Samples of axial somatic muscle, liver and gonads were kept frozen (-20°C) for energy studies. Wet mass of each sample was measured, and the material was dried at 60°C to constant weight and ground in a pestle mortar. Pellets (75-200 mg) were burned in a Parr 1425 micro-bomb calorimeter (Lucas, 1996). The values, corrected by ash and acid contents, were expressed as kJ·g⁻¹ ash free dry weight (AFDW).

Total energy content was calculated as the sum of the energy content of gonads, liver, and muscle. Wet and dry weight values were used to calculate wet weight/dry weight conversions for energy determinations. The Friedman test was performed to compare the energy content between organs of each species (STATISTICA software).

Table 1. Oocyte diameter, absolute and relative (Oocytes/ TW) fecundity in Antarctic Notothenioidei (*no ripe oocytes).

| Species          | n  | Oocyte diameter (µm) | Oocyte wet weight (mg) | Oocyte dry weight (mg) | Absolute fecundity | Relative fecundity |
|------------------|----|----------------------|------------------------|------------------------|---------------------|-------------------|
| N. rossii        | 1  | 2150 - 3150          | 8.06                   | 2.67                   | 89793               | 27                |
| C. gunnari       | 4  | 1800 - 3100          | 6.52                   | 2.33                   | 3933                | 14                |
| C. rastrospinosus| 1  | 3900 - 4600          | 30.24                  | 8.41                   | 4073                | 10                |
| G. gibberifrons  | 4  | 500 - 1200*          | 0.51                   | 0.13                   | 84067               | 103               |
| P. georgianus    | 2  | 3800 - 4700          | 26.92                  | 7.54                   | 7317                | 9                 |
| L. kempi         | 1  | 700 - 1200*          | 0.62                   | 0.19                   | 116420              | 175               |

Table 2. Relationship between total body weight (TW) / total body length (TL) and total energy content (TEC) / TL. Mean percentage water content (MWC %). TEC for standardised fish at 40 cm TL (TEC₄₀). Mean energy density (ED ± SD) for each species analysed. n₁ = total number of specimens caught in the LAMPOS survey; n₂ = number of specimens used for energetic determination.

| Species          | n₁ | TW = a·TLᵇ | r² | n₂ | TEC = a·TLᵇ | r² | MWC % | TEC₄₀ | Mean ED (SD)kJ·g⁻¹ wet mass |
|------------------|----|-------------|----|----|-------------|----|-------|-------|---------------------------|
| C. aceratus      | 78 | 0.0010·LT ᵇ | 0.96 | 8  | 0.0043·TL ᵇ | 0.99 | 78.8  | 1944 | 5.38 (0.45)                |
| C. gunnari       | 76 | 0.0005·LT ᵇ | 0.95 | 5  | 0.0433·TL ᵇ | 0.47 | 76.8  | 2217 | 5.21 (0.50)                |
| P. georgianus    | 49 | 0.0008·LT ᵇ | 0.97 | 11 | 0.0003·TL ᵇ | 0.89 | 80.0  | 2238 | 4.55 (0.41)                |
| C. rastrospinosus| 13 | 0.0040·LT ᵇ | 0.97 | 10 | 0.0040·TL ᵇ | 0.93 | 78.8  | 2615 | 4.99 (0.24)                |
| G. gibberifrons  | 92 | 0.0015·LT ᵇ | 0.97 | 14 | 0.0028·TL ᵇ | 0.98 | 76.9  | 3183 | 4.89 (0.17)                |
| D. eleginoides   | 38 | 0.0249·LT ᵇ | 0.95 | 5  | 0.2216·TL ᵇ | 0.88 | 68.7  | 4689 | 7.80 (1.27)                |
| L. kempi         | 158| 0.0042·LT ᵇ | 0.99 | 8  | 0.0279·TL ᵇ | 0.97 | 75.5  | 7253 | 5.40 (0.48)                |
RESULTS

Gonad maturation

Chaenocephalus aceratus

In the ovaries of Chaenocephalus aceratus (off South Georgia and South Orkney Islands) the vitellogenic oocytes showed different degrees of atresia (Fig. 2a). Some oocytes were starting the atretic process, with a fragmented chorion and hypertrophy of granulose cells. Other oocytes were already in an advanced stage of resorption, with a reduced cytoplasm invaded by phagocytic cells distributed between yolk globules.

Champsocephalus gunnari

All the ovaries of Champsocephalus gunnari showed a bimodal oocyte diameter distribution (Fig. 3). The smaller oocytes (< 600 µm) had basophilic cytoplasm and primary yolk vesicles. The bigger oocytes (1600 to 3200 µm) were in an advanced stage of maturation, with homogeneous yolk stained heavily by PAS and eosin (Fig. 2b; GSI = 8). Males had testes with wide central tubules full of sperm.
and the more external tubules containing isolated spermatogoniae placed against the tubule wall (Fig. 4a, GSI = 2.5).

**Pseudochaenichthys georgianus**

Samples of *Pseudochaenichthys georgianus* were composed of both juvenile and sexually mature adult individuals. Ovaries of the former contained only basophilic or previtellogenic oocytes, while the latter had ovaries with mature oocytes (3800 to 4800 µm, Fig. 3; GSI = 18). Sampled males were all mature. Their testes had tubules with numerous spermatogonia and a small amount of cysts containing spermatocytes (Fig. 4b; GSI = 2.02). Abundant spermatozoa, both in cysts and free, were observed in the central ducts. The irregular arrangement of spermatozoa and the phagocytic cells found between them suggested a previous evacuation.

**Chionodraco rastrospinosus**

Two types of ovaries were distinguished in this species. One type had scanty oocytes, with few yolk globules and recent post-ovulatory follicles (POF, Fig. 2c; GSI= 2.54). The POF were large, with a wide central lumen, and consisted of hypertrophied follicular cells arranged in an irregular convoluted structure (Fig. 2d). A monolayer theca with abundant small blood vessels and connective fibres surrounded the POF. The other type contained mature oocytes with homogeneous yolk (Fig. 2e; GSI= 22.49) and a batch of previtellogenic oocytes. This second type of ovary showed a bimodal distribution of oocyte diameters with a batch larger than 700 µm and a batch of mature oocytes between 3900 and 4600 µm (Fig. 3). In all sampled males the central testis tubules contained free spermatozoa while the peripheral tubules contained a small number of residual spermatozoa (Fig. 4c; GSI = 1.33). Occasionally, phagocytic cells were present among the residual spermatozoa. Isolated spermatogonia (about 12 µm) were distributed along the tubule walls and groups of them were close to the blind end of the tubules. The general histology of the testes suggested a recent evacuation of sperm.

**Lepidonotothen larseni**

The ovaries had oocytes with numerous secondary yolk globules. Some of them were starting the process of yolk coalescence (Fig. 2f; GSI = 5.15).

**Lepidonotothen kempi**

Two ovary categories were found in the samples of this species. One type (fish captured especially at station C2, near the South Sandwich Islands), with GSI lower than 2, contained advanced (old) POF, oocytes in primary vitellogenesis and some residual oocytes in the process of resorption (Fig. 2g; GSI = 1.7).
The other group of ovaries (most of the fish captured at station C5) showed GSI between 3 and 10, and contained a batch of oocytes in secondary vitellogenesis (Fig. 2h; GSI = 7.2). The diameter distribution of oocytes in smears showed a bimodal distribution with 50% of basophilic oocytes (300 µm) and 50% of maturing oocytes (500 – 1300 µm, Fig. 3).

Two different degrees of maturation were found in males of *Lepidonotothen kempi*. One group showed testes with active spermatogenesis, containing a large number of cysts of spermatogonia and spermatocytes arranged close to the tubule walls (Fig. 4d). A second group had scarce spermatogoniae, spermatocytes free in the lumina of the ducts and phagocytes between the spermatocytes (Fig. 4e), suggesting a previous sperm evacuation.

**Gobionotothen gibberifrons**

Ovaries of this species contained basophilic, previtellogenic and yolky oocytes (Fig. 2i). The ovaries from the fish sampled off South Georgia (B2 Station, Fig. 1) showed a lower degree of maturation (GSI=1.4) than the ovaries from the fish sampled off Elephant Island (E1 Station, Fig. 1; GSI =5.9). In histological sections the oocytes from fish sampled off South Georgia had a diameter of about 700 µm, while in the ones from off Elephant Island the diameter was around 880 µm.

In smears, a bimodal distribution of oocytes was found, with previtellogenic ones of less than 500 µm diameter and yolky ones between 500 and 1300 µm in diameter (Fig. 3).

Residual mature oocytes, which appeared to be a remainder from a previous spawning, were found in ovaries from fish sampled off Elephant Island (Fig 2j). Testes were in the process of maturation, containing cysts of spermatocytes.

**Dissostichus eleginoides**

The total length of the specimens (53 to 93 cm) was smaller than or close to the first maturation size (Everson and Murray 1999). Ovaries had basophilic
and previtellogenic oocytes filling the ovigerous lamellae (Fig. 2k). Two types of testes were found, one of them containing exclusively spermatogonia covering the tubule wall (Fig. 4f, GSI = 0.02) and the other one containing cysts with spermatogonia and spermatocytes (Fig. 4g, GSI = 3.2). There was a lumen in the centre of each tubule. Spermatozoa were not found in cysts or in tubules.

**Notothenia rossii**

Only one female specimen was captured, with its ovary in maturation. In smears almost 45% of the oocytes were previtellogenic opaque oocytes (300 µm diameter), while small diameter oocytes (400 to 1000 µm.) containing globular yolk reached about 35%. The larger translucent yolky eggs had diameters between 2100 and 3200 µm (Fig. 3).

**Fecundity**

The number of oocytes contained in the ovary was estimated in three species of Channichthyidae and in three species of Nototheniidae (Table 1). The only species with ovaries containing fully mature oocytes were *N. rossii*, *C. rastrospinosus* and *C. gunnari*. In the other species used for the estimation, the larger oocytes were in an advanced stage of vitellogenesis. Absolute and relative fecundity were lower in the Channichthyidae than in the Nototheniidae.

**Energy content**

Due to the small number of specimens studied in each species, the energy content (kJ/g) of both sexes was analysed together. The energy content of muscle and gonads only showed significant differences in *D. eleginoides* and *C. rastrospinosus* (Fig. 5A and B). The former had higher energy content in the muscle than in the gonads. Conversely, the latter had higher energy content in the gonads than in the muscle. The energy content of the liver was higher than the energy content of the gonads in all species (p<0.001), except in *C. gunnari*.

The energy content of the liver of *G. gibberifrons* (Fig. 6) was significantly higher in individuals sampled off South Georgia than in individuals sampled off Elephant Island (stations B2 and E1 respectively; Fig. 1).

The total energy content, size and mass for each species are shown in Table 2. Total energy content
calculated for a standard specimen (40 cm TL) was lower in channichthyids than nototheniids. The highest value was measured in *L. kempi*. Minimum and maximum values of energy density (ED kJ/g wet mass) were found in *C. aceratus* and *D. eleginoides* respectively.

**DISCUSSION**

The scarcity of material sampled from different species at the stations covered does not allow for generalisations about reproductive patterns and energetic characteristics of notothenioids along the Scotia Arc. However, the data are sufficient to perform an analysis in nine fish species caught mainly off South Georgia and in the southern branch of the Scotia Arc.

Biogeographically, our sampling sites are located in the ichthyofaunistic subregion of the Seasonal Pack Ice Zone and northern islands in which the Scotia Arc is included (sensu Kock, 1992). This is dominated by notothenioids of the genera *Notothenia* and *Lepidonotothen*, harpagiferids and channichthyids. Six of the species analysed in this study (Tables 1 and 2) were described as inhabiting exclusively this subregion. The other three species studied show a wider distribution. *Chionodraco rastrosinus* inhabits both low and high Antarctic zones (Kock and Stransky, 2000); *Lepidonotothen kempi* (=*L. macrophtalma + L. squamifrons*, Schneppenheim et al., 1994, Eastman and Eakin, 2000) occurs in the low Antarctic zone and on the Patagonian shelf (Kock and Stransky, 2000); and *Dissostichus eleginoides* has a wide latitudinal distribution related to Subantarctic deep waters (as far as 35ºS, off southern Peru in the Pacific Ocean, and off Uruguay in the Atlantic Ocean; De Witt et al., 1990).

Christiansen et al. (1998) found differences in the reproductive timing between low and high Antarctic species, and considered that they were caused by both geographical gradients and different modes of spawning. In the present study, two principal reproductive groups were recognised in agreement with Christiansen et al. (1998): substrate spawning fish, with low fecundity, demersal oocytes of great diameter with a high energetic content (most of channichthyids), and pelagic spawners with numerous small-diameter oocytes (notothenioids and *C. gunnari*).

The values of relative and absolute fecundity found in this study (Table 1) are in agreement with the range of the values found in previous studies (Christiansen et al., 1998; Kock, 1989; Kock and Kellermann, 1991; Kock and Everson, 1997; Permitin, 1973). Although the mature females were few in number, our results help to understand the reproductive patterns of Antarctic notothenioids. They are characterised by delayed gonadal maturation, prolonged gametogenesis, usually one unique spawning per year and moderately high fecundity (North and White, 1987; Kock, 1992). Accordingly, most of the species analysed in the present study showed synchronous oocyte growth, with only one generation of maturing oocytes indicating that spawning occurred only once in each spawning season.

In all specimens of *C. aceratus* captured near both the South Orkney and South Georgia islands (Stations B2 and D2 respectively, Fig. 1), yolky oocytes of up to 1500 µm diameter were found in atretic condition (Fig. 2a). The presence of atretic oocytes and the small size of healthy oocytes (1 mm) suggest that these fish neither had spawned recently nor were prepared to spawn given the proximity of the spawning season (March – May, c.f. Permitin 1973; Kock, 1989). This generalised atretic phenomenon is considered as a failure in the attainment of final oocyte maturation that could be caused by the poor physical condition of the females, probably due to food shortage (Hunter and Macewicz, 1985; Yoneda et al., 2002). In the case of this species, food should be available all year round because adult specimens are considered opportunistic predators (Everson et al., 2000). The main cause of massive ovarian atresia in all sampled female specimens is not clear. Nevertheless, skipping spawning, when environmental conditions are unsuitable, could be a way to maintain somatic viability (McEvoy and McEvoy, 1992). When one species shows massive ovarian atresia and the reproductive cycle is studied using only macroscopic staging of ovaries without histological support, the risk of confusion between pre-reproductive atresia and the normal maturation process is high because the atretic oocytes can be misidentified as maturing ones (La Mesa et al., 2003).

Ovaries of *C. gunnari* captured off the South Orkneys (Station D2) contained mature oocytes with homogeneous yolk, with a diameter of about 3.2 mm. The advanced stage of maturation of the ovaries and the testes full of mature sperm ready to evacuate found in the present study suggest the proximity of the spawning season in March–May, in agreement with Kock (1989). The ovaries analysed
were healthy and showed no signs of an atretic process. Pre-reproductive regression was described in different opportunities for *C. gunnari* off South Georgia (Everson *et al.*, 1991; Macchi and Barrera-Oro, 1995). These authors established a relation between the failure in ovarian maturation and the interannual variation in krill availability, the main food source for this species. Kock and Kellermann (1991) postulated that part of the population of *C. gunnari* in the Atlantic sector of the Southern Ocean does not spawn every year. The annual variability of individuals participating in spawning emphasises the importance of checking the actual percentage of spawning individuals every year in species under exploitation.

The coexistence of mature males and females of *P. georgianus* with oocytes around 4500 µm in the South Georgia samples indicates the occurrence of spawning during April in agreement with Kock’s and Kellermann’s (1991) findings.

In *C. rastrospinosus* ovaries with either mature oocytes or recent POF (Fig. 2c-e) were found off South Georgia (Station B2). The histology of the testes showed sperm evacuation. These data are in agreement with the data of Kock (1989) and Kock and Kellermann (1991), who described the spawning of this species taking place in April around Elephant Island. These results suggest that spawning occurs simultaneously in an extensive area at both sides of the Antarctic Polar Front.

Female *L. larsoni* (off Elephant Island, Station E1) had ovaries containing oocytes with numerous secondary yolk globules beginning the yolk coalescence process (Fig. 2f). This ovarian histology suggests that this species probably spawns in early winter, in agreement with the results of Kock (1989).

In *L. kempi*, the presence of two kinds of ovaries and testes (spent and maturing) indicates that spawning had occurred previously near the South Sandwich Islands (Fig. 1, Station C2, 254-262 m deep) since a maturing process with heavier ovaries containing yolked oocytes took place in deeper waters (Fig. 1, Station C5, 380-390 m deep). Our samples containing yolked oocytes in April suggest a protracted maturation during winter and spring. The presence of old POF in April (Fig. 2g) indicates either that the POF remained in the ovary as long as four months, or that the spawning off the South Sandwich Island occurred near the end of summer. If this was the case, spawning could occur later than November-December, as was described by Kock and Kellermann (1991) off the South Orkney Islands.

In April ovaries of *G. gibberifrons* were in the process of maturation. The yolky oocytes had not yet reached their maximum diameter (Fig. 3), which Kock (1989) determined at 2.5 mm. The presence of mature residual oocytes in the ovaries collected in April (Fig. 2j) could indicate the occurrence of a slow process of resorption, since the anterior spawning period should have been in the previous winter, according to Kock (1989) for the population at Elephant Island. Testes in our samples contained only spermatocytes and not spermatozoa, suggesting that males were in the early process of maturation as well.

Female *D. eleginoides* were immature, probably because they had not reached the size of first maturity yet (98.2 cm for females, Everson and Murray, 1999). One group of males was smaller than the size of first maturity (78.5 cm, Everson and Murray, 1999), and accordingly the testes were immature. A second group, composed of bigger males, was in the process of maturation (Fig. 4g), suggesting that these specimens could be ready to evacuate during the next spawning season, which Kock and Kellermann (1991) established as July-September.

Comparisons of reproductive features between the Antarctic and Subantarctic notothenioids are difficult because available data concerning the latter group are restricted to a few species, usually inhabiting shallow waters and without closely related species in the Antarctic zone. There are two exceptions: the genera *Harpagifer* and *Champsocephalus*. The first comprises several species in coastal waters of different peri-Antarctic islands (*H. bispinis, H. georgianus* *georgianus, H. georgianus* *palliolatus, H. kerguelenensis, H. spinosus*) and one Antarctic species (*H. antarcticus*) (Fischer and Hureau, 1985). Reproductive characteristics were studied only in *H. antarcticus* (Daniels, 1978; White and Burren, 1992). The genus *Champsocephalus* has the non-Antarctic species *C. esox*, confined to the Magellan Province, whose reproductive biology was studied in a population inhabiting the Beagle Channel (Calvo *et al.*, 1999). Sexually mature males are found from January to September, and females containing 3300 to 8600 mature oocytes or POF are found from February to November.

Individuals of the non-Antarctic species *Patagonotothen tessellata* spawn twice a year, in winter and at the end of summer. Each male guards egg masses spawned by several females in nests under flat rocks in lower levels of the intertidal zone of the Beagle Channel. Female *P. tessellata* spawn...
7600–62,000 oocytes with a diameter of 1000 to 1500 µm (Rae and Calvo, 1995; 1996). P. cornucola has not specifically been studied but preliminary observations indicate a similar timing of spawning (unpubl. observ.).

The non-Antarctic notothenioids show the only verified protandric hermaphroditic species that inhabits the Magellan region, Eleginops maclovinus (Calvo et al., 1992). This species spawns partially in June in central to southern Chile (Panozo, 1996), and shows a protracted spawning season between September and December around the Islas Malvinas/Falkland Islands in waters of 30-100 metres depth. Each batch comprises between 1.1 and 7.3 million oocytes with a diameter of 1 to 1.2 mm (Brickle et al., 2005). Hence, taking into account the extended spawning period and the possible repetitive spawning of the non-Antarctic notothenioids vs. the usual single spawning of Antarctic notothenioids, Subantarctic waters obviously impose less severe constraints on the reproductive effort than Antarctic waters.

The axial swimming muscles of fish represent more than 60% of their total body mass (Johnston, 2001; Sanger and Stoiber, 2001), and their energy content affects the total energy content value of the body. In all the species analysed in this study the energy content (kJ/g) of the axial muscle had similar values except for D. eleginoides, which reached a significantly higher value (Fig. 5A). Gonads of D. eleginoides had the lowest energetic content but differences from other species were not significant. It is noticeable that the average values of the energy content (kJ/g) of gonads (Fig. 5B) were not different between the species studied in spite of the diverse degree of sexual maturation.

Energetic values of liver in G. gibberifrons showed a great dispersion (Fig. 5C) that could probably be explained by the different origin of fishes. G. gibberifrons caught off Elephant Island (Station E1, Fig. 6) had significantly lower values of liver energy content and higher GSI and bigger yolky oocytes than the females caught off South Georgia (Station B2).

D. eleginoides showed the highest values of energy density (7.8 kJ/g wet mass, Table 2), probably due to the high lipid content of this species (Eastman, 1993). The rest of the species studied varied between 4.55 and 5.40 kJ/g.

The energy density (ED) of notothenioids (Table 2) is higher than the values reported for Pleuronectes asper (3.5 to 4.5 kJ/g wet mass; Paul, 1997) but equivalent to or slightly lower than the energy values of sexually mature Clupea pallasi (8-10 kJ/g; Paul et al., 1998). These results stress the important role played by notothenioids in energy transfer in Southern Ocean food webs.

The highest total energy content (TEC) found in this study (Table 2) for a standardised fish of 40 cm TL corresponds to L. kempi (7253 kJ), although the ED (5.4 kJ/g) of this species was not the highest. This high TEC could be explained by the higher TW/TL relationship that L. kempi has in comparison with the other notothenioids, which have TEC values of 2000 to 4600 kJ (Table 2). These values are similar to the values reported for Gadus morhua of 45 cm TL (2400 to 4100 kJ; Lambert and Dutil, 1997).

The estimations of total energy density in the species studied (expressed in relation to TL or TW; Table 2) could be useful to quantify the energy transfer in piscivorous predators such as seabirds or marine mammals (Anthony et al., 2000; Cherel y Ridoux, 1992).

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