A narrative review of the last decade’s literature on the diagnostic accuracy of septic arthritis of the native joint

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Abstract
While septic arthritis can be a straightforward diagnosis, there are many cases when the diagnosis is difficult to make. The aim of this study was to review the last decade’s literature on the diagnosis of septic arthritis of the native joint in adults and summarize that data in an easy to follow algorithm to clarify how the last decade’s data may be applied to the diagnosis of septic arthritis. A search of PubMed and CINAHL databases was performed to identify studies that compared results diagnostic tests for septic arthritis. We cross referenced this search with searches of additional databases (including Cochrane Library and Scopus) to confirm similar search results. The Quality Assessment of Diagnostic Accuracy Studies (QUADAS) tool was used by two independent reviewers to determine study quality and risk of bias. After applying inclusion and exclusion criteria to the initial search, 15 papers total were included for analysis. All 15 papers were of high quality methodology as determined by the QUADAS tool. There were 26 different diagnostics tests used across the 15 papers included for review. Three of those diagnostic tests had specificity and sensitivity greater than 80%. Eight tests had a positive likelihood ratio of ≥10. Three tests had a negative likelihood ratio < 0.1, indicating that they may help to rule out septic arthritis. A flowchart was created to summarize the findings of our review, so that physicians may reference this visual in making the appropriate diagnosis when the commonly held standards of cell count, gram stain, and culture aren’t enough to make the diagnosis.

Keywords: Septic arthritis, Septic joint, Joint aspiration

Introduction
The diagnosis of septic arthritis primarily relies on patient clinical presentation and synovial fluid analysis of the affected joint. The differential diagnosis of septic arthritis is broad and may include gout, pseudogout, trauma, hemarthrosis, rheumatic fever, rheumatoid arthritis, spondyloarthropathies, osteomyelitis, viral arthropathies, septic bursitis, and lyme disease [7]. While synovial fluid analysis is obtained as standard of care, it is well-documented that synovial fluid findings can be highly variable and there can be significant overlap in patients with the underlying diagnosis of gout, pseudogout, or rheumatoid arthritis [9, 30]. A synovial white blood cell (WBC) count of 50,000 has typically been used as an appropriate cut-off for the diagnosis of septic arthritis, though gout and pseudogout are known to result in similar WBC counts [23]. Conversely, some patients with septic arthritis may have WBC counts < 50,000, and the immunosuppressed patient may mount little to no leukocytic response at all. Septic arthritis can also coexist with crystalline arthropathy, thus further confounding the diagnosis of septic arthritis [7, 22]. Septic arthritis, gout, and pseudogout are known to have elevated serum inflammatory markers such as erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP), rendering them of little use in differentiating the diagnosis of septic arthritis from systemic
disease. Similarly, serum glucose and protein have failed to show diagnostic utility [9]. Recent analysis has shown some promise for the use of other serum markers such as procalcitonin levels, the delta neutrophil index, calprotectin levels, and the lactate/glucose ratio [3, 5, 21]. Culture and gram stain can help to confirm septic arthritis, though these can be falsely positive due to contamination from skin flora. Colony counts and sensitivities can help to confirm actual infection vs. contamination. Alternatively, a negative culture can occur in septic arthritis due to initiation of antibiotics prior to synovial fluid sampling, inadequate fluid sampling volume, or inadequate plating and growth requirements. All of this points to the difficulty that can arise in properly diagnosing septic arthritis and the need for a synopsis of recent literature on the topic of the diagnostic accuracy of septic arthritis.

The aim of this study was to review the last decade’s literature on the diagnosis of septic arthritis of the native joint in adults and summarize that data in an algorithm in order to clarify how the last decade’s literature may be applied to the diagnosis of septic arthritis.

Methods
Study design
This narrative review was performed according to the criteria of the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) recommendations [14].

Search strategy
A systematic search of relevant literature was conducted using PubMed and CINAHL, with cross referencing total manuscript counts in the Cochrane Library and Scopus, as well as hand-searching reference lists of included articles. Search terms used were "septic arthritis", "septic joint", "diagnosis", "approach", and "synovial fluid". We subsequently re-ran the search with additional terms such as "infection" "infectious" and confirmed that our results were similar. Results were then narrowed to include only the adult population, native joint infections, and literature from the last 10 years.

Study selection
A total of 933 articles were reviewed by the first author to identify studies related to the diagnostic accuracy of clinical tests in septic arthritis of the native joint in the adult population. Titles and abstracts were initially reviewed for a primary screen. Full text articles were retrieved during further screening for inclusion (Fig. 1).

Eligibility criteria
Diagnostic studies were eligible if they included 1) a description of the clinical approach to diagnosing septic arthritis; 2) an assessment of the accuracy of their diagnostic test (e.g. sensitivity, specificity, positive and negative likelihood ratios, or odds ratios) 3) an acceptable reference standard for comparison; 4) were written in the English language; and 5) conducted their study on the adult population with native joints. Studies that included pediatric patient populations were excluded as more formalized diagnostic criteria already exist for some cases of pediatric septic arthritis, such as the Kocher criteria. Patients with prosthetic joints were not included in this review. Studies were excluded if they did not provide an adequate reference standard or a report of diagnostic accuracy.

Data extraction
Data extraction was performed by the first author. Data extracted included: study population, types of arthritides included, studied diagnostic test, diagnostic reference standard, diagnoses made by the authors, sensitivity, specificity, positive likelihood ratios, and negative likelihood ratios, when available. In studies where odds ratios were provided, a quadratic formula described by Simel et.al. was used to back-calculate sensitivity, specificity, and likelihood ratios if positive and negative test result data was available [27]. Due to heterogeneity amongst outcome measures and diagnostic methods, outcome measures and data could not be combined into a summary set of measures for meta-analysis to create overall diagnostic outcome effects.

Sensitivity, defined as the ability of a test to correctly classify a person with the disease of interest as having that disease, was calculated as (true positives/(true positives + false negatives)). Specificity, defined as the ability to correctly classify a person without the disease of interest as not have that disease, was calculated as (true negatives/(true negatives + false positives)). A likelihood ratio, defined as the likelihood of a given test result in a person with the disease of interest compared with the likelihood of the same result in a person without the disease of interest, was also calculated. A positive likelihood ratio was calculated as (sensitivity/(1-specificity)). A negative likelihood ratio was calculated as ((1-sensitivity)/specificity). Sensitivity and specificity were considered to be sufficient if they were both >= 90%. A likelihood ratio greater than 10 indicated that a positive test was good at ruling in a diagnosis, while a likelihood ratio less than 0.1 indicated that a negative result was good at ruling out a diagnosis [8, 13].

Quality assessment
Articles included in the study were assessed for potential bias and quality using the Quality Assessment of Diagnostic Accuracy Studies (QUADAS) tool. This is a 14-question tool with each question scoring 1 point for a "yes" answer and 0 points for a "no" or "unclear" answer. Papers that received a score of 10 or above were
considered high quality with less risk of design bias, while those which received a score below 10 were considered low quality with higher risk of design bias. Two reviewers (EHGT and MHL) independently assessed each paper using the QUADAS tool. Disagreements between reviewers were discussed and resolved.

Level of evidence
The level of evidence was assessed using the Knee Surgery, Sports Traumatology, Arthroscopy level of evidence table provided to authors at: https://www.kssta.org/authors-homepage/level-of-evidence/.

Source of funding
None.

Results
Systematic search results and study selection
A total of 933 articles were retrieved after the initial searches (Search 1: 706 articles, Search 2: 227 articles). The initial 933 were reviewed by title and summary and
narrowed to 46. These 46 papers were reviewed via full text and narrowed to 15 after removing studies that included children, periprosthetic joint infections, tests that would not be readily available to aid in the rapid diagnosis of septic arthritis in the hospital setting, did not have the diagnosis of septic arthritis as the primary diagnosis, or did not have sensitivity, specificity, or positive and negative result data needed to compare the diagnostic accuracy of their results. Figure 1 provides a flow diagram describing study selection.

Characteristics of included studies
Table 1 provides details on study characteristics. Diagnostic tests investigated, participants, diagnostic reference tests used, and diagnoses made by investigators were included. There were 26 different diagnostic tests used across 15 papers included in this review. These tests assessed a total of 1997 patients.

The most common reference standard used for a diagnosis of septic arthritis was a positive synovial fluid culture. Many studies also used Newman’s criteria as their reference standard [16]. Some studies used either Newman’s criteria or a positive synovial fluid culture or a combination of clinical presentation, synovial fluid leukocyte counts, and the exclusion of other noninfectious causes. One study used either positive synovial fluid cultures or a diagnosis of septic arthritis by an orthopedic surgeon with surgical intervention and intravenous antibiotics given during the hospital stay.

Sizes of study populations between papers ranged from 46 patients to 458.

The different diagnostic tests included synovial lactate, synovial glucose, synovial lactate/glucose, PCR, synovial culture, synovial white blood cell count (WBC) with and without differential, C-reactive protein, procalcitonin, synovial calprotectin, microcalorimetry, urate, synovial total protein, synovial erythrocytes, synovial total polymorphic cells, synovial total mononuclear cells, hemoglobin, platelet counts, absolute neutrophil counts, neutrophil differential, crystals, interleukin-6, leukocyte esterase and glucose test strips, lactate dehydrogenase, and tumor necrosis factor alpha.

Diagnostic accuracy
The diagnostic accuracy of the clinical tests studied, calculated by sensitivity, specificity, and positive and negative likelihood ratios are provided in Table 2. Sensitivity ranged from 23% - 100% and specificity ranged from 3.5% - 100%. Positive likelihood ratios ranged from 0 to 111.88 and negative likelihood ratios ranged from 0 to 2.23. Three diagnostic tests had both sensitivity and specificity >90%; these were synovial leukocytes > 50,000 or PMNs > 90% (94%, 100%), leukocyte esterase ++ or +++ and glucose – (89.5% and 99.2%), and PMNs > 75% (100%, 94%). Eight tests had a positive likelihood ratio of ≥10. These were lactate-glucose ratio > 5 (LR+ 27), synovial lactate > 10 (LR+ 41.6), synovial glucose < 1.0 (LR+ 33.3), calprotectin > 150 mg/L (LR+ 12.2), neutrophils > 95% in the absence of crystals (LR+ 11.36), leukocytes > 50,000 in the absence of crystals (LR+ 10.94), leukocyte esterase ++ or +++ and glucose – (LR+ 111.88), and PMNs > 75% (LR+ 16.67).

Quality scores
The quality scores of the manuscripts as assessed by the QUADAS tool ranged from 10 to 13, indicating that all of the papers were high quality with low risk of design bias (Table 3). There was 93% initial agreement using QUADAS scores and 100% after further discussion.

Level of evidence
The level of evidence as assessed using the Knee Surgery, Sports, Traumatology, Arthroscopy table provided to authors indicated that all the papers were of either level II (60%) or level III (40%) evidence, in the Diagnostic category (Table 3).

Discussion
While septic arthritis is sometimes easily diagnosed, there are many situations in which the diagnosis can be confounded by underlying or concomitant disease processes or pathologies. Our goal was to review the last decade’s literature on the diagnosis of septic arthritis of the native joint in adults and provide an algorithm summarizing the findings of the last decade into an easy to follow workflow. The typical patient presentation consists of an acute onset of joint pain with erythema, warmth, limited joint range of motion, and possible effusion within the joint [7]. The first step in diagnosing any suspected septic arthritis is to perform a joint aspiration for gram stain, culture, and cell count of the aspirated synovial fluid, prior to the administration of antibiotics. Successful aspiration of various joints requires knowledge of the anatomical structures that could preclude access to the joint space. When aspirating the ankle, the needle should be inserted 2.5 cm proximally and 1.3 cm anteriorly to the tip of the lateral malleolus, just lateral to the peroneus tertius tendon. The knee can be aspirated on the lateral side, at the superior aspect of the patella. The needle must be advanced through the lateral retinaculum to enter the joint. The hip can be aspirated from either a lateral, medial, or anterior approach. It may be necessary to use advanced imaging to aid in
| Author                  | Test Evaluated                        | Subjects (N, Gender, Mean age) | Reference Standard                                                                                                                                 |
|-------------------------|---------------------------------------|-------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------|
| Berthoud (2020) [3]     | Lactate, Glucose, Lactate/Glucose Ratio | 233                           | 151 males mean age 61.2 Newmann’s Criteria and 1) pathogen isolated from synovial fluid 2) pathogen isolated from blood culture with typical clinical presentation for arthritis or 3) arthrocentesis revealed purulent synovial fluid in combination with presence of typical clinical presentation for septic arthritis, absence of crystals, and absence of other suitable diagnosis |
| Sigmund (2019) [26]     | mPCR, synovial culture                 | 72                            | 39 males mean age 64 1) Newmann’s Criteria or 2) Pathologic features of septic arthritis and leukocyte count > 50,000 or PMNs > 90% in synovial fluid        |
| Chouk (2019) [5]        | WBC, CRP, Procalcitonin               | 98                            | 51 males mean age 65.2 Newman’s Criteria                                                                                                           |
| Shu (2019) [25]         | Synovial lactate                      | 39                            | 28 males mean age 51 1) Positive synovial fluid culture or 2) Septic arthritis diagnosed by institution orthopedists with surgical intervention and IV antibiotics given during hospital stay |
| Baillet (2019) [1]      | Calprotectin                           | 74                            | 38 males mean age 70.4 Bacteria in synovial culture or blood culture without crystals present                                                                 |
| Morgenstern (2018) [15] | SF leuk w/ diff, PCR, and microcalorimetry | 57                            | 31 males mean age 62 Positive synovial fluid culture or 1) local clinical signs and symptoms, 2) increased SF leukocyte count and 3) exclusion of noninfectious causes |
| Pavic (2018) [20]       | Highest recorded temp w/ 24 h, hemoglobin, WBC, platelet, CRP, ESR, Urate, Syn total leuk, Syn total eryth, syn tot polymorphos, syn tot mononuc, gender, symptom duration, features of sepsis, presence of sweat, presence of chills/rigors, joint swelling, joint erythema, joint warmth, joint tenderness to palpation, joint ROM restriction | 165                           | 119 males mean age 59.35 Positive synovial fluid culture                                                                                           |
| Ferreyra (2017) [6]     | ALC, ANC, neutrophil differential, crystals | 208                           | 125 males mean age 59.6 Pyogenic organism in joint fluid or blood samples                                                                              |
| Borzio (2016) [4]       | ESR, serum WBC, syn WBC, neutrophils, lymphocytes, temp | 458                           | (gender, age not given) Positive synovial fluid culture                                                                                             |
| Paosong (2015) [18]     | Procalcitonin                         | 75                            | no gender, age given Newman’s Criteria                                                                                                               |
| Lenski (2014) [19]      | IL6, Synovial Lactate                 | 119                           | 54 males mean age 69.9 Positive synovial fluid culture                                                                                              |
| Omar (2014) [17]        | Leukocyte Esterase and Glucose Strips | 146                           | 64 males mean age 59 1) A pathogen was isolated from the synovial fluid 2) A pathogen was isolated from a                                             |
needle placement within the hip capsule. The shoulder is most easily aspirated anteriorly, where the bony landmarks are readily palpable. The needle should be inserted at half the distance between the coracoid process and the anterolateral edge of the acromion, with the needle directed posteriorly so as to avoid the neurovascular bundle of the brachial plexus. The elbow is typically aspirated via a posterior approach, with the needle inserted just lateral to the olecranon [11] (Table 4). Previous literature has shown that the commonly held threshold of synovial WBC > 50,000 is not sensitive enough to effectively rule out septic arthritis [10, 12]. It is also well described that other arthritides such as gout, pseudogout, or rheumatoid arthritis can cause a high synovial WBC count as well [23]. Notably, other studies have argued that synovial leukocytes > 50,000 is not sufficiently sensitive to rule out a diagnosis of septic arthritis. Other tests had excellent likelihood ratios of > 10, indicating that they could aid in the diagnosis as a confirmatory “rule in” test, after synovial cell counts were already collected. The tests with positive likelihood ratios > 10 included lactate:glucose ratio > 5, synovial lactate ≥ 10, synovial glucose < 1.0, calprotectin > 150 mg/L, neutrophils > 95% in the absence of crystals, leukocytes > 50,000 in the absence of crystals, leukocyte esterase ++ or +++ and glucose –, and PMNs > 75% [1, 3, 6, 17]. Of these tests presented, serum lactate and glucose are easily drawn labs, readily available in the vast majority of hospital settings. A dipstick for synovial leukocyte esterase and glucose is also an easily available test, with an excellent positive likelihood ratio. While Ferreyra et al. detailed the ways in which the absence of crystals and a high leukocyte or neutrophil count can help to confirm the diagnosis of septic arthritis, the presence of crystals cannot effectively rule out septic arthritis, as described earlier [6, 29].

To that end, several tests had good negative likelihood ratios (LR- ≤ 0.1) and may help to more quickly rule out septic arthritis. Baillet et al. demonstrated that a synovial

| Author                  | Test Evaluated | Subjects (N, Gender, Mean age) | Reference Standard |
|-------------------------|----------------|-------------------------------|--------------------|
| Baran (2014) [2]        | WBC, %PMNs     | 96                            | Positive synovial fluid culture |
| Lenski (2014) [9]       | Serum markers (WBC, CRP, UA), synovial markers (lactate, glucose, UA, LDH, WBC, tot prot, IL 6) | 82                | Positive synovial fluid culture |
| Talebi-Taher (2013) [28]| serum/synovial procal, serum IL-6, TNF-a, CRP, ESR, synovial WBC and PMN % | 75, 41 males mean age 52.2 | 1) Purulent material in the joint space with isolation of a bacterial pathogen from the joint fluid or 2) Positive gram stain |
### Table 2  Diagnostic Accuracy of Included Tests

| Test                                | Sensitivity | Specificity | NPV   | PPV   | LR+   | LR-   |
|-------------------------------------|-------------|-------------|-------|-------|-------|-------|
| Berthoud Lactate/Glucose Ratio > 5  | 52.0%       | 98.10%      | 27    | 0.49  |       |       |
| Synovial Lactate >=10               | 40.0%       | 99.00%      | 41.6  | 0.61  |       |       |
| Synovial Glucose < 1.0              | 32.0%       | 99.00%      | 33.3  | 0.69  |       |       |
| Sigmund SF mPCR                     | 38.0%       | 100.00%     | 100   | 53.6  | 0     | 0.62  |
| Sf Culture                          | 29.0%       | 100.00%     | 100   | 50    | 0     | 0.71  |
| Combined culture + mPCR             | 43.0%       | 100.00%     | 100   | 55.6  | 0     | 0.57  |
| Tissue Culture                      | 40.0%       | 100.00%     | 0     | 0.6   |       |       |
| Chouk PCT > 0.5 ng/ml               | 65.0%       | 91.00%      | 91    | 7.20  | 0.40  |       |
| Chouk PCT > 0.2 ng/ml               | 80.0%       | 74.40%      | 44.4  | 93.5  | 3.10  | 0.30  |
| Shu Synovial Lactate >=10           | 27.0%       | 97.00%      | 7.90  | 0.80  |       |       |
| Shu Synovial Lactate >=5            | 55.0%       | 76.00%      | 2.30  | 0.60  |       |       |
| WBC >= 50,000                       | 27.0%       | 97.00%      | 7.90  | 0.80  |       |       |
| WBC >=100,000                       | 18.0%       | 100.00%     | 10.00 | 0.80  |       |       |
| Baillet Calprotectin > 150 mg/L     | 73.0%       | 94.00%      | 84    | 88    | 11.64 | 0.29  |
| Baillet Calprotectin < 52 mg/L      | 96.0%       | 44.00%      | 47    | 95    | 1.71  | 0.09  |
| Morgenstern Serum CRP > 10          | 78.0%       | 40.00%      | 48    | 71    | 1.30  | 0.55  |
| Morgenstern Serum WBC > 10          | 62.0%       | 74.00%      | 65    | 71    | 2.38  | 0.51  |
| SF Leaks > 50,000 or PMNs > 90%     | 94.0%       | 100.00%     | 100   | 97    | 0.00  | 0.06  |
| Culture                             | 46.0%       | 100.00%     | 100   | 74    | 0.00  | 0.54  |
| PCR                                 | 23.0%       | 91.00%      | 63    | 65    | 2.56  | 0.77  |
| Microcalorimetry                    | 46.0%       | 94.00%      | 83    | 73    | 7.67  | 0.57  |
| Pavic Features of Sepsis (RR > 25 bpm, HR > 120 bpm, SBP < 100, temp > 38.5 w/1 24 h of review) | 75.0% | 72.10% | 24.7 | 93.5 | 1.93 | 0.41 |
| Ferreyra Neutrophil count > 50,000  | 50.0%       | 94.40%      | 893   | 0.53  |       |       |
| Ferreyra Neutrophil count < 15,000  | 92.3%       | 77.00%      | 4.013 | 0.10  |       |       |
| Leukocyte count > 50,000             | 53.6%       | 91.70%      | 5.76  | 0.51  |       |       |
| Leukocyte count > 70,000             | 39.3%       | 95.60%      | 8.93  | 0.63  |       |       |
| Leukocyte count < 20,000             | 92.3%       | 70.60%      | 3.14  | 0.11  |       |       |
| %Neutrophils > 90                   | 71.4%       | 79.70%      | 3.52  | 0.36  |       |       |
| %Neutrophils > 95                   | 50.0%       | 89.00%      | 4.55  | 0.56  |       |       |
| %Neutrophils < 80                   | 96.2%       | 56.00%      | 2.19  | 0.07  |       |       |
| %Neutrophils > 90 + no crystals     | 67.9%       | 92.80%      | 8.28  | 0.35  |       |       |
| %Neutrophils > 95 + no crystals     | 50.0%       | 95.60%      | 11.36 | 0.52  |       |       |
| > 50,000 leukocytes + no crystals   | 53.6%       | 95.10%      | 10.94 | 0.49  |       |       |
| > 50,000 neutrophils + no crystals  | 35.7%       | 96.70%      | 10.82 | 0.66  |       |       |
| Borzio Synovial fluid WBC > 64,000  | 40.0%       | 90.00%      | 4     | 0.25  |       |       |
| Kocher criteria + synovial WBC > 64,000 | 0.0% | 98.60% | 0 | 90.8 | 0 | 1.0149878 |
| Paosong Procalcitonin >= 0.66       | 59.0%       | 86.00%      | 69.9  | 79.6  | 4.21  | 0.48  |
| Lenski1 Synovial tot prot 4.3       | 55.6%       | 75.00%      | 2.22  | 0.59  |       |       |
| Syn Gluc 40                         | 56.6%       | 83.00%      | 3.33  | 0.52  |       |       |
| Synovial Lactate 6.2                | 74.5%       | 87.20%      | 5.81  | 0.29  |       |       |
| Syn WBC 14.4                        | 71.2%       | 84.90%      | 4.71  | 0.34  |       |       |
| Synovial IL6 7000                   | 92.5%       | 64.10%      | 2.58  | 0.12  |       |       |
| Omar LE ++ or +++                   | 94.7%       | 73.20%      | 34.6  | 98.9  | 3.54  | 0.08  |
### Table 2 Diagnostic Accuracy of Included Tests (Continued)

| Test Description                          | Sensitivity | Specificity | NPV  | PPV  | LR+  | LR-  |
|-------------------------------------------|-------------|-------------|------|------|------|------|
| LE ++ or +++ and GLC -                   | 89.5%       | 99.20%      | 94.4 | 98.4 | 111.88 | 0.11 |
| WBC > 50,000                             | 72.7%       | 92.30%      | 9.09 | 0.30 |      |      |
| % PMNs > 90                              | 81.8%       | 67.30%      | 2.50 | 0.27 |      |      |
| %PMNs > 85                               | 88.6%       | 57.70%      | 2.05 | 0.23 |      |      |
| %PMNs > 80                               | 93.2%       | 53.80%      | 2.32 | 0.11 |      |      |
| Synovial WBC > 50,000                    | 72.7%       | 92.30%      | 9.09 | 0.30 |      |      |
| % PMNs > 90                              | 81.8%       | 67.30%      | 2.50 | 0.27 |      |      |
| %PMNs > 85                               | 88.6%       | 57.70%      | 2.05 | 0.23 |      |      |
| %PMNs > 80                               | 93.2%       | 53.80%      | 2.32 | 0.11 |      |      |
| Synovial Glucose < 5.1                   | 65.9%       | 92.00%      | 8.24 | 0.37 |      |      |
| (Gout) Synovial uric acid of 7.0         | 78.1%       | 82.80%      | 4.53 | 0.27 |      |      |
| (Gout) Serum uric acid of 7.2            | 70.00%      | 85.20%      | 4.73 | 0.35 |      |      |
| Synovial lactate >/= 4.3                 | 89.5%       | 77.30%      | 3.04 | 0.14 |      |      |
| Synovial Glucose < 5.1                   | 65.9%       | 92.00%      | 8.24 | 0.37 |      |      |
| (Gout) Synovial uric acid of 7.0         | 78.1%       | 82.80%      | 4.53 | 0.27 |      |      |
| (Gout) Serum uric acid of 7.2            | 70.00%      | 85.20%      | 4.73 | 0.35 |      |      |
| Synovial LDH >/= 1900                    | 68.9%       | 88.90%      | 6.2  | 0.35 |      |      |
| Synovial WBC >/= 38.0                    | 58.2%       | 86.20%      | 4.22 | 0.49 |      |      |
| Synovial total prot of >/= 4.4           | 48.90%      | 75.00%      | 1.96 | 0.68 |      |      |
| Synovial IL-6 of >/= 7000                | 93.90%      | 13.60%      | 1.09 | 0.44 |      |      |
| Serum CRP of >/= 0.5                     | 92.30%      | 3.50%       | 0.96 | 2.23 |      |      |
| Peripheral WBC >/= 10.0                  | 55.80%      | 41.40%      | 0.95 | 1.07 |      |      |
| WBC > 50,000                             | 100%        | 66%         | 59.52| 100  | 2.94  | 0.00 |
| PMN > 75%                                | 100.00%     | 94.00%      | 89.29| 100  | 16.67 | 0.00 |
| CRP > 18 mg/L                            | 92.00%      | 30%         | 76.47| 79.31| 1.31  | 0.27 |
| ESR (> 17 for men, > 25 for women)       | 100.00%     | 26.00%      | 40.32| 100  | 1.35  | 0.00 |
| TNF-alpha > 10                           | 96.00%      | 62%         | 55.81| 96.88| 2.58  | 0.06 |
| IL6 > 20                                 | 12.00%      | 92%         | 42.86| 67.65| 1.50  | 0.96 |
| Serum PCT > 0.5 ng/ml                    | 68.00%      | 80.00%      | 62.96| 83.33| 3.40  | 0.40 |
| Synovial fluid PCT > 0.5%/ng/ml          | 24.00%      | 96%         | 75   | 71.64| 6.00  | 0.79 |

### Table 3 QUADAS Tool Scoring and Level of Evidence (LOE)

| Author                        | Q1 | Q2 | Q3 | Q4 | Q5 | Q6 | Q7 | Q8 | Q9 | Q10 | Q11 | Q12 | Q13 | Q14 | TOTAL | LOE   |
|-------------------------------|----|----|----|----|----|----|----|----|----|-----|-----|-----|-----|-----|-------|-------|
| Berthoud (2020)               | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 0   | 1   | 1   | 1   | 12   | Diagnostic II |
| Morgenstern (2018)            | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 0   | 1   | 1   | 0   | 10   | Diagnostic II |
| Pavic (2018)                  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 0   | 1   | 0   | 1   | 11   | Diagnostic II |
| Ferreyra (2017)               | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 0   | 1   | 0   | 1   | 11   | Diagnostic II |
| Baran (2014)                  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 0   | 1   | 0   | 1   | 11   | Diagnostic II |
| Lenski (2014)                 | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 0   | 1   | 0   | 1   | 11   | Diagnostic II |
| Talebi-Taher (2013)           | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 1  | 0  | 0   | 1   | 0   | 0   | 10   | Diagnostic II |
calprotectin of < 52 mg/L was a highly sensitive test (96%) with a LR – of 0.09 and Ferreyra et al. found that a synovial neutrophil count of < 15,000 had a sensitivity of 92% with a LR of 0.10 [1, 6]. Lastly, Talebi-Taher et al. described serum TNF-alpha as an excellent discriminator between septic and inflammatory arthritis with a sensitivity of 96%, a negative predictive value of 96.88%, and a negative likelihood ratio of 0.06. These additional studies may help to rule out septic arthritis in more difficult to diagnose patients with synovial WBC counts less than 50,000.

From the information synthesized above we have created an evidence-based algorithm summarizing our findings that practitioners may use to more rapidly narrow the diagnosis of septic arthritis while waiting for gram stain and culture to return (Fig. 2). The literature review confirms that for any suspected septic arthritis joint aspiration with synovial fluid analysis consisting of cell count, gram stain, and culture be performed. While awaiting culture results, additional tests based on the algorithm in Fig. 2 may be utilized to aid in “ruling in” or “ruling out” the diagnosis of septic arthritis, depending on the synovial cell counts.

Limitations
The main limitation in this review is the absence of a universally accepted gold standard for the diagnosis of septic arthritis. While many papers cited a positive synovial fluid culture as their reference standard, others referred to clinical criteria or a clinical course consisting of surgical intervention and antibiotics. The heterogeneity of diagnostic tests as well as the varying population sizes included in the studies makes it difficult to create uniformly generalizable conclusions about the diagnosis of septic arthritis. Additionally, laboratories at various institutions may have different ‘normal ranges’ and therefore the numbers presented may not be universal measurements, but may need to be converted into differing institutional ranges. This review is further limited by the small number of articles published about the potential diagnostic accuracy of tests in septic arthritis. The narrow field may have introduced bias into our review. Language limitations of the reviewers narrowed the acceptable studies to those published in English. Lastly, while an attempt was made to assess the potential for bias and quality using the QUADAS tool, this tool does have its own limitations. A paper may score poorly on the QUADAS tool if certain methodological steps are not included in the manuscript, even if they were part of the analysis. This field would further benefit from high quality methodological research to further narrow the diagnostic potential of some of the referenced biomarkers in the approach to septic arthritis.

Conclusions
This narrative review aggregates and synthesizes the last decade of published literature on the approach to diagnosing septic arthritis in the adult native joint, and we have provided a visual algorithm summarizing our review. The initial step in diagnosis is a joint aspiration with culture and gram stain. Notably, culture and gram stain are still the gold standard when it comes to accurately diagnosing septic arthritis of the native joint, however, while culture and gram stain are pending, there are additional tests that can help to either rule in or rule out septic arthritis if the preliminary cytology is not overly convincing. Test findings such as TNF-alpha < 10, percent neutrophils < 80, blood procalcitonin < 0.39, synovial calprotectin < 52 mg/L, and neutrophil count < 15,000 all have a negative likelihood ratio < 0.1 and can help to rule out the diagnosis of septic arthritis. In order to help rule in the diagnosis of septic arthritis test findings such as synovial lactate/glucose ratio > 5, synovial lactate > 10, synovial glucose < 1.0, calprotectin > 150 mg/L, synovial leukocyte esterase ++ or +++ with glucose on
Fig. 2 Algorithm summarizing findings of this review

*In all scenarios, antibiotics should be held until joint aspiration is complete.
*ROM = Range of Motion
*LR = Negative Likelihood Ratio
49 *LR+ = Positive Likelihood Ratio
50 *WBC = White Blood Cell
51 *TNF-α = Tumor Necrosis Factor Alpha

Patient with painful joint

Joint pain Swelling
Erythema
Restricted ROM

*Joint aspiration <50K cells ± crystals, culture pending
Consider additional “rule out” tests (LR_0 <0.1)

TNF-α <10
%Neutrophils < 80
Blood Procalcitonin <0.39
Synovial Calprotectin <52mg/L
Neutrophil Count <15,000
If one or more of these tests is in the “rule out” range, consider other causes of arthritis

*Joint aspiration >50k – 100k cells ± crystals, culture pending
Consider additional “rule in” tests (LR+ ≥ 10)

Synovial Lactate/Glucose Ratio >5
Synovial Lactate ≥ 10
Synovial Glucose < 1.0
Calprotectin >150 mg/L
%Neutrophils >95, crystals absent
>50,000 leukocytes, crystals absent
>50,000 neutrophils, crystals absent
Leukocyte esterase ++ or +++ with – glucose on testing strip
If one or more of these tests is in the “rule in” range, consider proceeding to irrigation, debridement, and antibiotics

*Joint aspiration >100K cells ± crystals, culture pending
Joint aspiration with WBC ≥ 100,000 has 100% specificity for septic arthritis (Shu)
Irrigation, debridement, and antibiotics
dipstick, percent neutrophils > 95, > 50,000 leukocytes, or > 50,000 neutrophils, all in the absence of crystals have likelihood ratios > 10.

Acknowledgements
None.

Authors’ contributions
All authors contributed to the study conception and design. Material preparation, data collection, and analysis were performed by EHGT and MHL. The first draft of the manuscript was written by EHGT. All authors read and approved of the final draft of the manuscript.

Funding
None.

Availability of data and materials
Data is available in tables and figures within the manuscript.

Ethics approval and consent to participate
Not applicable.

Consent for publication
Not applicable.

Competing interests
AMS is a paid consultant for Stryker. EHGT and MHL have no disclosures.

Received: 1 October 2020 Accepted: 25 November 2020

Published online: 09 January 2021

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