Postexercise urinary alpha-1 acid glycoprotein is not dependent on hypoxia

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Abstract

Proteinuria is a transient physiological phenomenon that occurs with a range of physical activities and during ascent to altitude. Exercise intensity appears to dictate the magnitude of postexercise proteinuria; however, evidence also indicates the possible contributions from exercise-induced hypoxemia or reoxygenation. Using an environmental hypoxic chamber, this crossover-designed study aimed to evaluate urinary alpha-1 acid glycoprotein (α1-AGP) excretion pre/postexercise performed in hypoxia (HYP) and normoxia (NOR). Sixteen individuals underwent experimental sessions in normoxia (NOR, 20.9% O2) and hypoxia (HYP, 12.0% O2). Sessions began with a 2-h priming period before completing a graded maximal exercise test (GXT) on a cycle ergometer, which was followed by continuation of exposure for an additional 2 h. Physiological responses (i.e., blood pressure, heart rate, and peripheral oxygenation), Lake Louise Scores (LLSs), and urine specimens (analyzed for albumin and α1-AGP) were collected pre- and postexercise (after 30, 60, and 120 min). Peak power output was significantly reduced in HYP (193 ± 45 W) compared with NOR (249 ± 59 W, P < 0.01). Postexercise urinary α1-AGP was greater in NOR (20.04 ± 14.84 μg·min−1) than in HYP (15.08 ± 13.46 μg·min−1), albeit the difference was not significant (P > 0.05). Changes in urinary α1-AGP from pre- to post-30 min were not related to physiological responses or performance outcomes observed during GXT in NOR or HYP. Despite profound systemic hypoxemia with maximal exercise in hypoxia, postexercise α1-AGP excretion was not elevated above the levels observed following normoxic exercise.

NEW & NOTEWORTHY By superimposing hypoxic exposure and maximal exercise, we were able to investigate the impact of hypoxia on postexercise proteinuria. Urinalysis for α1-AGP (via particle-enhanced immunoturbidimetry) in specimens collected pre-/postexercise enabled the sensitive detection of altered glomerular permeability. Data indicated that exercise intensity, rather than the degree of exercise-induced hypoxemia, determines postexercise proteinuria.

alpha-1 acid glycoprotein; exercise; hypoxia; orosomucoid; proteinuria

INTRODUCTION

Glomerular proteinuria is a transient physiological phenomenon that occurs in healthy individuals following exercise (1) and during ascent to altitude (2, 3). Exhibited across a range of physical activities [e.g., swimming (4), running (5), and rowing (6)], it can be characterized by increases in urinary albumin or alpha-1 acid glycoprotein (α1-AGP; 1, 7), the latter being a potentially more sensitive marker of proteinuria (3). Exercise intensity ultimately dictates the degree of the postexercise increases (8, 9), although the mechanism(s) for this are not well understood. Intermittent exercise has been shown to elicit greater increases in proteinuria compared with continuous exercise (10), implicating a possible contribution from exercise-induced hypoxemia (or reoxygenation; 11). This is supported by findings from altitude studies that have shown relationships between urinary α1-AGP and blood oxygenation (12). Furthermore, hypoxia (HYP) is known to potentiate a variety of factors that have been implicated for postexercise proteinuria such as increased oxidative stress (13), acid-base balance disruption (8, 14), and increased blood pressure via changes in catecholamines (15) or elements of the renin-angiotensin-aldosterone-system (16, 17).

Exercising at altitude presents a unique way to investigate the involvement of hypoxemia in the development of postexercise glomerular proteinuria. Hypoxic ventilatory response (HVR) tests would support such investigation given that HVR relates to the degree of hypoxemia exhibited with exercise at altitude (18). Therefore, it was hypothesized that: 1) exercise under hypoxic conditions would elicit greater postexercise α1-AGP excretion compared with a sea-level equivalent exercise bout and 2) a greater HVR may attenuate exercise-induced hypoxemia and thereby limit postexercise elevations in proteinuria.

To evaluate the impact of hypoxemia on postexercise glomerular proteinuria, the objective of this study was to measure α1-AGP excretion following maximal exercise in an
environmental chamber. This study aimed to examine: 1) the time course and degree of urinary α1-AGP excretion surrounding exercise in normoxia (NOR) and hypoxia (HYP); 2) physiological responses to exercise in NOR and HYP in relation to postexercise urinary α1-AGP excretion; and 3) whether any associations existed between HVR test outcomes and the degree of postexercise urinary α1-AGP (exhibited in NOR vs. HYP).

**METHODS**

**Participants and Design**

Ethical approval was granted by the University of Birmingham (ERN_18-1270). Written informed consent was obtained before participation. Consented participants completed a general health history questionnaire to screen for signs, symptoms, or history of cardiovascular, pulmonary, renal, or metabolic disease with only healthy individuals included. Individuals who reported any of the following were also excluded: 1) active pharmacotherapy, 2) actively being under the care of a GP (for any reason), or 3) history of smoking. If a participant exhibited any of the following (after repeated measurements) at rest during any session, they were withdrawn: 1) systolic blood pressure (SBP) ≥ 140 mmHg, 2) diastolic blood pressure (DBP) ≥ 90 mmHg, or 3) heart rate (HR) > 100 beats·min⁻¹.

All participants first completed a familiarization session that included a graded incremental maximal exercise test (GXT) in ambient conditions (see *Graded Incremental Maximal Exercise Tests*). A randomized crossover design was then adopted with all participants completing two experimental sessions (NOR and HYP; see *Experimental Sessions*), with each consisting of physiological response measurements, a GXT, and timed urine collections (see *Urine Experiments*). Resting control trials and HVR tests (see *Hypoxic Ventilatory Response Tests*) were also completed in a subset of participants. For all sessions, participants were asked to refrain from 1) partaking in strenuous exercise or consuming alcoholic beverages 24 h prior and 2) consuming caffeine 4 h prior. All sessions were separated by no less than 48 h.

**Experimental Sessions**

Experimental sessions in NOR (20.9% O₂) and HYP (~12.0% O₂, equivalent to 4,000 m) were conducted in an environmental chamber (TIS Services, Hampshire, UK) as outlined in Fig. 1. Participants were blinded to condition wherever possible [e.g., using sham hypoxia for NOR sessions (19)]. Sessions consisted of a preexercise 2-h priming exposure (upright seated), during which participants were encouraged to consume 500 mL of water, before completing a GXT that lasted ~15 min. The GXT was followed by a continuation of exposure for an additional 2 h (total session duration, 4.5 h). Resting control sessions were identical to experimental sessions except GXT, which was substituted with a time-controlled resting period (familiarization GXT duration used). If at any point a participant felt too unwell to continue, presented with moderate or severe symptoms of

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**Figure 1.** Experimental session timeline. Detailed schematic for the conduct of resting control, normoxic (NOR), and hypoxic (HYP) sessions. Resting control sessions were identical to NOR and HYP sessions other than the graded incremental maximal exercise (GXT), which was substituted with a resting period (length of familiarization GXT). Each session proceeded as follows: 1) participant arrived and underwent 10–15 min of seated rest; 2) “baseline” (sea-level) physiological responses (heart rate, systolic and diastolic blood pressures, and peripheral oxygenation) and Lake Louise Scores were recorded; 3) participant emptied their bladder, which started the clock for timed urine collections; 4) participant entered the chamber; 5) participant was encouraged to drink at least 500 mL of water in the 2 h leading up to GXT during which physiological response outcomes were recorded at 30-min intervals; 6) urine specimens collected immediately preexercise; 7) GXT was administered with staged increases adjusted for condition (30 W vs. 20 W/2 min; NOR and HYP, respectively; to standardize test duration between conditions) and physiological response outcomes recorded throughout (every 1–2 min; refer to *Graded Incremental Maximal Exercise Tests*); 8) postexercise physiological response outcomes and timed urine specimens collected at 30, 60, and 120 min following GXT; and 9) participant exited the environmental chamber and was monitored for approximately 15 min (and up to 1 h) before departure.
acute mountain sickness [i.e., Lake Louise Score, LLS, >5 (20)], or requested to leave the chamber for any reason, they were removed immediately.

Peripheral oxygenation (SpO₂; via finger pulse oximetry; WristOx₂, Model 3i50, Nonin Medical Inc., Plymouth, MN), heart rate (HR; via 3-lead electrocardiogram, ECG), SBP and DBP (via automated sphygmomanometry of the brachial artery; Tango M2 Stress Monitor, SunTech Medical, Morrisville, NC), and LLS were measured: upon arrival, at 30-min intervals throughout experimental sessions, and for at least 15 min (and up to 1 h) following each session. Mean arterial pressure (MAP, mmHg) at 30-min intervals was calculated from resting SBP, DBP, and HR using a weighted formula (21). Urine specimens were also collected throughout these sessions as outlined in Specimen collection and handling.

Graded Incremental Maximal Exercise Tests

Graded exercise tests were conducted on an upright cycle ergometer (Velotron, Quarq Technology, Spearfish, SD) and began with a 3-min warm-up, which was performed at 50 W and had a target rating of perceived exertion (RPE) of 14 [Borg Scale (22)]. Following the warm-up, a 2-min stage was performed at 60 W. To control for total exercise duration between conditions, wattage was increased every 2 min by 20 W or 30 W (in HYP and NOR, respectively) until volitional fatigue (23) or an RPE of 20 was achieved. To ensure maximal effort in both, NOR and HYP, the anticipated maximal power output (Wₘₐₓ) for each condition was estimated from familiarization W_max (i.e., NOR = 100% of familiarization W_max and HYP = 70%-80% of familiarization W_max). After W_max was achieved an unloaded, self-paced cool-down was performed for 3–5 min.

SpO₂ and HR were recorded each minute throughout GXT, whereas power output (in watts), SBP, DBP, and RPE were recorded at the end of each 2-min stage. Performance outcomes (maximal power output, W_max) and physiological response outcomes (SBP_max, DBP_max, HR_max, and SpO₂_max) were also recorded at the time maximal effort was achieved. MAP_max was calculated using the previously mentioned weighted formula by inputting SBP_max, DBP_max and HR_max measurements. Blood lactate (BLA) was analyzed (Lactate Plus, Nova Biomedical, Waltham, MA) before and after GXT from blood obtained by finger prick. Delta BLA (Δ BLA) was estimated from these measurements.

Urine Experiments

Specimen collection and handling.

To mark the start of timed urine collections, participants emptied their bladder (not collected) within 15–20 min of arrival and before entering the environmental chamber. All urine produced thereafter was collected into 3,000 mL, UV-protected, polyethylene containers designed for human urine collection (SARSTEDT, Nümbrecht, Germany). Separate containers were used for each timed collection with specimens collected immediately preexercise (arrival to preexercise; ~2 h) and at the following timepoints following exercise: post-30 min (preexercise to post-30 min; GXT duration + 30 min), post-60 min (post-30 to post-60 min), and post-120 min (post-60 to post-120 min; 60 min; see Fig. 1). Specimen volume (mL), weight (g), and collection time (h:min) were immediately recorded upon collection with a sample (20 mL), then aliquoted into a conical centrifuge tube and temporarily stored on ice (0°C) until the end of the session, when all samples were centrifuged (5,400 rpm at 21°C) for 10 min. Following centrifugation, supernatants were further aliquoted into 2 mL cryovials (4 × replicates) and frozen at ~80°C until urinalysis.

Urine specimens were thawed in a warming cabinet (37°C) for 1 h before urinalysis. Particle-enhanced immunoturbidimetry for low-concentration α₁-AGP (measuring range: 0.08–148.20 mg·L⁻¹) and albumin (measuring range: 11–66,500 mg·L⁻¹) was performed using the Optilite autoanalyzer (The Binding Site, Ltd., Birmingham, UK). Triplicate (α₁-AGP) results were used in conjunction with specimen volume (or weight) and collection duration to estimate α₁-AGP excretion rate (µg·min⁻¹). Triplicate excretion rates for each specimen were then averaged with this result used for statistical analysis. Change (Δ) in α₁-AGP was estimated as the difference between preexercise and post-30-min excretion.

Hypoxic Ventilatory Response Tests

An additional visit was undertaken among a subset of participants (n = 6) to assess their HVR. HVR tests were administered via a dynamic end-tidal forcing (DEF) system (v1, BreathDP, University of Oxford) using a stepped, isocapnic-hypoxia protocol (24). Tests began with participants seated at rest while breathing ambient room air for 5 min via a mouthpiece connected to the DEF, which allowed ventilation to stabilize. During this resting period, “baseline” partial pressure of end-tidal CO₂ (PETCO₂) was recorded. Next, a series of 3-min stepped decreases in the partial pressure of end-tidal oxygen (PETO₂) were administered (at 100 mmHg, 65 mmHg, and 55 mmHg) with PETCO₂ clamped ~1 mmHg above “baseline” throughout. Three-minute steps were followed by a final 5-min step at 100 mmHg PETO₂ with total test duration being ~20 min. Measures of ventilation (tidal volume; respiratory rate; and minute ventilation, VE), SpO₂, SpO₂, and HR (via 3-lead ECG) were recorded continuously throughout.

Thirty-second averages were estimated for VE, SpO₂, and VE/SpO₂ (L·min⁻¹·%⁻¹), and plotted as a function of time for the three isocapnic-hypoxic steps. VE/SpO₂ represented the direct relationship between VE and SpO₂ at each 30-s interval, whereas VE/SpO₂ slope (or ΔVE/ΔSpO₂) represented the slope of the best-fit line plotted (within individuals) from the linear regression (VE, y-axis; and SpO₂, x-axis) across all steps (25–27). Peak VE (L·min⁻¹), minimum SpO₂ (%), and mean values for VE, SpO₂, and VE/SpO₂ were reported for each step, whereas VE/SpO₂ slope (ΔVE/ΔSpO₂) was reported for the entire HVR test (accounting for all three steps). By convention, VE/SpO₂ slopes were presented as absolute values with actual values used for statistical analysis. Void of missing data for all steps, repeated-measures ANOVA with Tukey’s post hoc test was used to compare HVR outcomes between isocapnic-hypoxic steps.

Statistical Analysis

Normality of distribution was assessed using Shapiro-Wilk test for each outcome measure with data log-
results likely attributable to similarities in the physiological responses following exercise (i.e., change from +30 to +120) in NOR and HYP. No significant main effect of time was observed for $S\text{PO}_2\,$ ($P = 0.368$) or SBP ($P = 0.411$). Similarly, no interaction effect between time and condition was observed for $S\text{PO}_2\,$ ($P = 0.161$), HR ($P = 0.054$), DBP ($P = 0.242$), SBP ($P = 0.083$), or MAP ($P = 0.127$).

### Maximal Exercise Tests

Results from paired t tests (or Wilcoxon tests) for GXT performance and physiological response outcomes are presented in Table 1. As expected, $S\text{PO}_{2\text{max}}$ and $W_{\text{max}}$ were significantly lower in HYP than in NOR (both, $P < 0.001$), whereas $SBP_{\text{max}}$ and $MAP_{\text{max}}$ were significantly greater in HYP than in NOR (both, $P < 0.05$). $DBP_{\text{max}}$, $HR_{\text{max}}$, RPE, $\Delta$ BL, and ambient CO were no different between NOR and HYP.

### Urinalysis

A grand total of 136 urine specimens were successfully collected from the 156 possible time points for NOR, HYP, and control sessions. In these specimens, urinalysis for $\alpha$-AGP was 95.6% successful, whereas urinalysis for albumin was only 23.7%. Urinary $\alpha$-AGP excretion rates (means ± SD, $\mu g \cdot min^{-1}$) from resting control, NOR, and HYP sessions are presented in Fig. 3 along with the corresponding plots and comparisons of log-transformed values. Mixed-effects analysis comparing log($\alpha$-AGP) values from resting control, NOR, and HYP sessions demonstrated the impact of exercise, as evidenced in the significant differences observed at post-30 min (as shown in Fig. 3B), as well as the significant main effects of condition [$F(2,09, 31.34) = 10.82, P < 0.001$] and time [$F(140, 20.94) = 24.83, P < 0.001$]. However, no interaction effect was apparent [$F(21.5, 10.40) = 3.35, P = 0.073$]. When log($\alpha$-AGP) values were compared between NOR and HYP sessions independent of the resting control, no significant main effects were observed for condition [$F(110.00, 15.00) = 3.88, P = 0.068$] or interaction [$F(25.0, 21.38) = 0.733, P = 0.519$], however, the significant main effect of time remained apparent [$F(27.40, 70.76) = 20.89, P < 0.001$]. Consistent with the former, $\Delta$-AGP was no different between NOR (15.09 ± 14.77 $\mu g \cdot min^{-1}$) and HYP (11.22 ± 14.71 $\mu g \cdot min^{-1}$, $P = 0.233$).

### HVR Tests

Mean and individual data for 30-s averages of $V\text{E}$, $S\text{PO}_2$, and $V\text{E}/S\text{PO}_2$, are presented in Supplemental Fig. S2 (see https://doi.org/10.6084/m9.figshare.14870154). As expected, a significant effect of $PETO_2$ on peak $V\text{E}$ ($P = 0.007$) and mean $V\text{E}$ ($P = 0.006$) was observed, with both increasing as $PETO_2$ was decreased. Similarly, $S\text{PO}_2$ was lower at each step, reflected in the significant main effect of $PETO_2$ on minimum and mean $S\text{PO}_2$ (both $P < 0.001$). Direct estimations of $V\text{E}/S\text{PO}_2$ (L·min$^{-1}$·%$^{-1}$) for each step also demonstrated a significant effect of $PETO_2$ on $V\text{E}/S\text{PO}_2$ ($P = 0.008$). By contrast, $V\text{E}$/$S\text{PO}_2$ slope (or $\Delta V\text{E}/\Delta S\text{PO}_2$), which was representative of the dynamic relationship between $V\text{E}$ (y-axis) and $S\text{PO}_2$ (x-axis) for HVR tests, was plotted from data from all steps, albeit with linear regressions applied to individual 30-s interval data (as shown in Fig. 4).
Results from correlation analyses are presented in Table 2. Significant positive relationships were observed between log (Δ α1-AGP) and SBPmax (P = 0.045; R² = 0.317) and MAPmax (P = 0.040; R² = 0.331) in HYP only. No other performance, physiological response, or HVR test outcomes were related to α1-AGP for either condition (Table 2). Of note, however, was the participant exhibiting the uncharacteristically elevated HVR ("ID 05" shown in red in Fig. 4). This participant was the only participant to exhibit greater post-30-min α1-AGP excretion in HYP (absolute, 33.70 μg·min⁻¹; Δ 31.56 μg·min⁻¹) than in NOR (absolute, 27.74 μg·min⁻¹; Δ 26.26 μg·min⁻¹), despite a reduction in Wmax (33% reduction) that was greater than that of the group (23 ± 7% reduction). This participant also exhibited the second highest MAPmax (163 mmHg) out of the group in HYP.

**DISCUSSION**

The present study assessed the contribution(s) of hypoxia to postexercise proteinuria by evaluating urinary α1-AGP excretion surrounding maximal exercise tests in NOR and...
HVR test outcomes (refer to Table 2). Nevertheless, excretion and the degree of desaturation during exercise orersh rerelationships were observed between postexercise tion (refer to Fig. 3). In line with this, no signifi
cance was exhibited by concomitant increases in postexercise HYP exercise, marked reductions in SpO2 were not accompa-
ned by concomitant increases in postexercise 1-AGP excretion (and changes observed in this study support earlier cycling-based studies that have demonstrated similar reductions (7) or unchanged postexercise excretion in hypoxia (28). The present observations were attributed to concomitant reductions in exercise performance known to accompany hypoxic exercise (29), which have, in part, been attributed to reductions in blood flow to the working limbs (30). It is suggested that in response to vigorous hypoxia (i.e., maximal exercise in HYP), an alteration in the redistribution of blood flow occurs, which promotes a balance between glomerular blood flow and pressure (31) that does not favor proteinuria to the same extent as NOR exercise (or lower exercise intensities (32)). This implicates the likely role of renal hemodynamics in the progression of postexercise glomerular proteinuria (33). Development of significant relationships between

Table 1. Performance and physiological response outcomes from maximal graded exercise tests

|                | Normoxia | Hypoxia |
|----------------|----------|---------|
| **Means ± SD** |          |         |
| Wmax, W        | 249 ± 59 | 193 ± 45 |
| SBPmax, mmHg   | 175 ± 36 | 198 ± 25 |
| DBPmax, mmHg   | 78 ± 12  | 80 ± 16  |
| MAPmax, mmHg   | 126 ± 19 | 139 ± 17 |
| HRmax, beats·min−1 | 186 ± 19 | 186 ± 20 |
| SpO2max, %    | 95 ± 3   | 81 ± 7   |
| RPE*          | 19 ± 2   | 19 ± 2   |
| ΔBLA, mmol·L−1·k−1 | 9.6 ± 12.3 | 7.3 ± 7.3 |
| CO2max, ppm   | 855 ± 386 | 674 ± 301 |

GXT outcomes are presented as means ± SD or median ± interquartile range (IQR), where marked by asterisk, *paired samples t tests (and t statistics) were used to compare between conditions (normoxia vs. hypoxia) for normally distributed data, whereas Wilcoxon matched-pairs tests (and z-scores) were used when distribution was abnormal (outcomes indicated by *). Similarly, mean differences (95% confidence intervals, CIs) were presented for normally distributed data and median differences (95% CIs) for data with an abnormal distribution. **Significant difference between normoxia vs. hypoxia. Subscript (max) refers to measurements obtained at the time maximal effort was achieved. BLA, blood lactate; CO2, ambient carbon dioxide; DBP, diastolic blood pressure; GXT, graded maximal exercise test; HR, heart rate; MAP, mean arterial pressure; RPE, rating of perceived exertion; SBP, systolic blood pressure; SpO2, peripheral oxygenation; Wmax, maximal power output.

Figure 3. Urinary alpha-1 acid glycoprotein (1-AGP) excretion rates. A: 1-AGP excretion (means ± SD, µg·min−1) during resting control sessions (among a subset of participants, n = 7) and surrounding exercise tests performed in normoxia and hypoxia (among the entire cohort, n = 16). B: corresponding box and whisker plots for log-transformed 1-AGP excretion rates (median ± 25% and 75% quartiles range; whiskers: 5th–95th percentiles). A mixed-effects analysis with Tukey’s test was used to compare log(1-AGP) values between sessions (resting control vs. normoxia vs. hypoxia) at the 30-min intervals surrounding maximal exercise tests. Compared with resting control, log(1-AGP) was significantly greater 30 min following exercise in both normoxia (P = 0.002) and hypoxia (P = 0.004). **All statistical tests were two-tailed with significance set to α < 0.05.
postexercise α1-AGP and SBP_max as well as MAP_max in HYP, and despite attenuated α1-AGP excretion in HYP, may indicate underlying pathophysiology with, but not limited to, glomerular involvement (e.g., diabetes; 37). Taken together with α1-AGP urinalysis being more sensitive than albumin urinalysis (refer to Urinalysis), it is reasonable to conclude that α1-AGP urinalyses will afford identification of subclinical changes in glomerular responses, translating to earlier disease detection or improved status monitoring.

Limitations and Future Directions

The absence of a prospective a priori power analysis for postexercise α1-AGP excretion may be considered a limitation of this study. Unfortunately, pre-/postexercise α1-AGP data were not available from the literature, making the effect size required to detect a difference between NOR and HYP unclear, which in turn prevented sample size the estimation. That said, biological relevance does not always coincide with indexed effect sizes (38). Thus, a contextualized approach, able to weigh various aspects of the experiment against the importance of reducing error, was considered to be most appropriate (39, 40). Nevertheless, results from this study should be interpreted in the context of the cohort studied.

Table 2. Correlation results for postexercise alpha-1 acid glycoprotein excretion

| Exercise Tests | Log(α1-AGP_post-30) | Log(α1-AGP_) | Log(α1-AGP_post-30) | Log(α1-AGP_) |
|---------------|---------------------|--------------|---------------------|--------------|
| W_max         | 0.347               | 0.450        | 0.234               | 0.129        |
| SpO2_max      | 0.520†              | 0.508†       | 0.184†              | 0.398†       |
| HR_max        | 0.816†              | 0.573†       | 0.892†              | 0.867†       |
| SBP_max       | 0.674               | 0.726        | 0.072               | 0.045**      |
| DBP_max       | 0.458               | 0.477        | 0.554               | 0.501        |
| MAP_max       | 0.816               | 0.815        | 0.070               | 0.040**      |
| HRV tests     | Peak Ve             | 0.564†       | 0.497†              | 0.714†       |
| Minimum SpO2  | 0.803†              | 0.919†       | 0.497†              | 0.497†       |
| Ve/SpO2 slope | 0.497†              | 0.419†       | 0.564†              | 0.564†       |

Correlation analyses were performed between log-transformed α1-AGP excretion rates [i.e., post-30 min or change (Δ) from pre- to post-30 min] and performance and physiological response outcomes from exercise tests, as well as hypoxic ventilatory responses (HVR) test outcomes using Pearson r (or Spearman rho, ℓ). Linear regression was applied to data demonstrating a significant relationship. Significant relationships were evident between log(Δ) and SBP_max (r = 0.563, R² = 0.317), as well as MAP (r = 0.564; R² = 0.319) for hypoxia only. All statistical tests were two-tailed with significance set at α < 0.05. Significant results are indicated by double asterisk, **. Ve/SpO2 slope (or ΔVe/ΔSpO2) represented the slope of the best-fit line plotted for the simple linear regression of data from all three isocapnic-hypoxic steps. α1-AGP, alpha-1 acid glycoprotein; DBP, diastolic blood pressure; HR_max, maximal heart rate; MAP, mean arterial pressure; SBP, systolic blood pressure; SpO2, peripheral oxygenation; Ve, minute ventilation; W_max, maximal power output.
Inability to report performance outcomes (e.g., exercise intensity) in relative measures (e.g., W·kg⁻¹ or VO₂max in mL·kg⁻¹·min⁻¹) is also an obvious limitation and likely the cause for the absence of any significant relationship between power output and postexercise α1-AGP. Nevertheless, the purpose of this study was not to evaluate the effect of intensity but rather the effect of hypoxemia by delivering a potent hypoxic stimulus (i.e., maximal exercise in HYP). Moreover, the proteinuric effect of such hypoxic exercise was expected to transcend that of intensity.

Arguments against the expression of ventilatory responsiveness as a function of SpO₂ can be made. However, in this instance, such expression adequately shows the change in ventilation and also demonstrates its effectiveness at increasing SpO₂, notwithstanding the limitations of SpO₂ measurements. Further, although it is acknowledged that the relationship between PO₂ and ventilation is hyperbolic (41), interpretation of ventilatory responsiveness using linear regression (applied to ventilation and SpO₂) was considered more appropriate in the context of the three isocapnic-hypoxic steps, which are representative of only a fragment of the hyperbolic response curve.

Building upon this study, and to improve understanding related to the role of renal hemodynamics in postexercise proteinuria, future researchers may consider: 1) evaluating a larger cohort; 2) analyzing physiological markers of sympathetic nerve activation (e.g., plasma epinephrine or norepinephrine), glomerular endothelial permeability (e.g., urinary glycosaminoglycans), or renal oxidative stress; and 3) measuring renal blood flow (via nonradioactive microsphere) or perfusion (via near-infrared spectroscopy, NIRS).

Conclusions

Findings demonstrate the utility of immunoturbidimetric analysis for urinary α1-AGP and examinations of postexercise proteinuria. Despite profound systemic hypoxemia, exercise-induced increases in urinary α1-AGP excretion appear to be dictated by exercise intensity.

SUPPLEMENTAL DATA

Supplemental Fig. S1: https://doi.org/10.6084/m9.14870136.
Supplemental Fig. S2: https://doi.org/10.6084/m9.14870154.

GRANTS

This research was supported by the JABBS Foundation.

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

K.E.J., G.M.B., A.R.B., and S.J.E.L. conceived and designed research; K.E.J., G.M.B., C.B., A.F., and S.J.E.L. performed experiments; K.E.J. and C.B. analyzed data; K.E.J. interpreted results of experiments; K.E.J. prepared figures; K.E.J. and S.J.E.L. drafted manuscript; K.E.J., G.M.B., A.R.B., and S.J.E.L. edited and revised manuscript; K.E.J., G.M.B., C.B., A.F., A.R.B., and S.J.E.L. approved final version of manuscript.

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