Epigenetic Alterations Associated With Early Prenatal Dexamethasone Treatment

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Prenatal treatment with dexamethasone (DEX) reduces virilization in girls with congenital adrenal hyperplasia (CAH). It has potential short- and long-term risks and has been shown to affect cognitive functions. Here, we investigate whether epigenetic modification of DNA during early developmental stages may be a key mediating mechanism by which prenatal DEX treatment could result in poor outcomes in the offspring. We analyzed genome-wide CD4+ T cell DNA methylation, assessed using the Infinium HumanMethylation450 BeadChip array in 29 individuals (mean age = 16.4 ± 5.9 years) at risk for CAH and treated with DEX during the first trimester and 37 population controls (mean age = 17.0 years, SD = 6.1 years). We identified 9672 differentially methylated probes (DMPs) associated with DEX treatment and 7393 DMPs associated with a DEX × sex interaction. DMPs were enriched in intergenic regions located near epigenetic markers for active enhancers. Functional enrichment of DMPs was mostly associated with immune functioning and inflammation but also with nonimmune-related functions. DEX-associated DMPs enriched near single nucleotide polymorphisms (SNPs) associated with inflammatory bowel disease, and DEX × sex-associated DMPs enriched near SNPs associated with asthma. DMPs in genes involved in the regulation and maintenance of methylation and steroidogenesis were identified as well. Methylation in the BDNF, FKBP5, and NR3C1 genes were associated with the performance on several Wechsler Adult Intelligence Scale–Fourth Edition subscales. In conclusion, this study indicates that DNA methylation is altered after prenatal DEX treatment. This finding may have implications for the future health of the exposed individual.

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Freeform/Key Words: prenatal treatment, dexamethasone, methylation, epigenetics, fetal programming, congenital adrenal hyperplasia

Abbreviations: CAH, congenital adrenal hyperplasia; DEX, dexamethasone; DMP, differentially methylated probe; FDR, false-discovery rate; GAT, Genetic Association Test; GO, gene ontology; GREAT, Genomic Regions Enrichment of Annotations Tool; GWAS, genome-wide association studies; IBD, inflammatory bowel disease; PNMS, prenatal maternal stress; SNP, single nucleotide polymorphism; WAIS-IV, Wechsler Adult Intelligence Scale–Fourth Edition.
The synthetic glucocorticoid dexamethasone (DEX) has been used for several decades to ameliorate prenatal virilization of girls with classic congenital adrenal hyperplasia (CAH), an inborn error of cortisol synthesis. Girls with CAH are treated during the entire gestational period from week 7, whereas treatment in boys with CAH and in healthy children is stopped when the genotype of the fetus is known (gestational weeks 12 to 14) [1]. We have previously identified substantial negative effects of DEX on verbal working memory in children not having CAH and who were treated during the first trimester of fetal life [2, 3]. These effects seem to be sex dimorphic, as cognitive deficits were more pronounced in treated girls [3]. Furthermore, women with CAH and treated during the entire gestational period perform worse on several cognitive tasks than women with CAH not treated prenatally [4].

Early-life glucocorticoid treatment or stressful events are good examples of fetal programming factors, as they have been associated with short- and long-term adverse effects on human health [5]. For example, antenatal glucocorticoid treatment in preterm infants resulted in increased aortic arch stiffness and altered glucose metabolism in early adulthood [6], as well as altered stress reactivity [7]. Another consequence is cortical thinning in general and in particular, of the rostral anterior cingulate cortex [8]. Thickness of the left rostral anterior cingulate cortex was further associated with more affective problems [8].

Furthermore, studies on children subjected to prenatal maternal stress (PNMS) during a natural disaster identified prenatal stress as a programming factor that may lead to long-lasting consequences for immunity [9]. Additionally, exposure to PNMS was positively associated with higher insulin levels and higher body mass index during adolescence [10]. At 5½ years of age, children exposed to high levels of objective stress had deficits in cognitive abilities [11]. Finally, PNMS was associated with functionally organized changes in the T cell methylome with specific changes in gene programming of the immune system itself [12].

Here, we investigate whether DNA methylation is altered after prenatal glucocorticoid treatment. We hypothesize that epigenetic modification of DNA during early developmental stages may be a key mediating mechanism by which prenatal DEX treatment could result in poor outcomes in the offspring. We investigated whether prenatal DEX treatment generates long-lasting effects on CpG methylation profiles. To this end, we isolated CD4+ T cells from children and adults at risk for CAH, but not having CAH, and exposed to DEX during the first trimester of fetal life. Data on whole-genome DNA methylation in DEX-exposed cases were compared with data from untreated population controls.

1. Patients and Methods

A. Subjects

The study is part of a longitudinal project evaluating prenatal treatment of CAH in a total of 237 individuals in Sweden (for details, see Wallensteen et al. [3]). Here, we focus on the effects of first-trimester DEX treatment on whole-genome methylation in prenatally treated subjects without CAH compared with healthy, untreated controls from the Swedish general population. Written, informed consent was obtained from all participants, and the study was approved by the Regional Ethics Committee of Stockholm. In total, 66 subjects [29 DEX-treated subjects (mean age = 16.4 years, SD = 5.9 years), 12 young women and 17 young men, and 37 population controls (mean age = 17.0 years, SD = 6.1 years), 18 young women and 19 young men] were included.

B. Isolation of T Cells and Flow Cytometry

Peripheral blood mononuclear cells were separated by density centrifugation from 50 ml of whole blood per subject. CD4+ T cells were isolated from peripheral blood mononuclear cells using magnetic-activated cell sorting (Miltenyi Biotec) and stored at −80°C.

The purity of CD4+ cell populations was evaluated by immunophenotyping using two-color antibody panels. Data were acquired and analyzed using the Cyan ADP Analyzer (Summit...
4.3; Beckman Coulter). T Cell population purity was 94.9% (SD 3.1). For a more detailed description of T cell isolation and flow cytometry, see Reinius et al. [13].

C. DNA Extraction, Bisulfite Treatment, and DNA Methylation Measurements Using the 450K BeadChip Array

Genomic DNA was isolated from T cell pellets using the QiAmp DNA Mini Kit (Qiagen), according to the manufacturer’s instructions. The DNA concentration was measured using the Qubit 2.0 concentration (Thermo Fisher Scientific). Bisulfite treatment was performed using the EZ-96 DNA Methylation Kit (Zymo Research), and DNA methylation measurements were performed using the Infinium HumanMethylation450 BeadChip array (Illumina).

D. Differential Methylation Analysis

The data analysis was performed in R, and preprocessing was conducted using the lumi package [14, 15]. Five samples, three controls, and two treated subjects were dropped during the quality-control phase as a result of poor genome-wide correlation with other samples and an aberrant distribution of β values. In addition, the following probes were excluded from the analysis: (i) probes located on the Y and X chromosomes; (ii) probes with a single nucleotide polymorphism (SNP) located within 3 bp of the interrogated CpG site to exclude false-positive probes caused by genetic variations; and (iii) CpG probes with detection values of $P > 0.01$ [16].

After filtering, 395,462 probes were included in the differential analysis. Estimations of β values for the probes were performed using a previously described, three-step pipeline [17]. Batch effects were identified using a principal component analysis and subsequently corrected using the Combat function from the Surrogate Variable Analysis (sva) Bioconductor package [18]. Differential DNA methylation was computed by transforming β values into M values and using the Linear Models for Microarray Data (limma) package to define a linear model, including the following variables: DEX, age, sex, and the interaction of the treatment (DEX) with the sex of the subject (DEX × sex) [15, 19]. The interaction between treatment and sex was studied as a result of previous results, indicating a sex-dimorphic effect of DEX treatment [3]. For the DEX-derived differential methylation analysis, we identified three sets of relevant CpG sites: probes with (i) $p_{uncorrected} < 0.01$, (ii) $p_{uncorrected} < 0.01$ and a group difference in methylation of 5%, and (iii) $p_{uncorrected} < 0.01$ and a group difference in methylation of 10%. Similar lists were computed for the DEX × sex interaction.

D-1. Association between DNA methylation and cognitive performance

We also sought to investigate whether there would be associations between methylation (β values) in a subset of candidate genes with biological relevance for brain function and cognitive outcome. To this end, we related methylation in differentially methylated probes (DMPs) from the candidate genes $BDNF$, $NR3C1$, $NR3C2$, and $FKBP5$ to the cognitive performance of the participants. For a detailed description of the procedures and tests used for assessment of cognitive performance, see Karlsson et al. [20]. Here, the test scores (of subjects 16 years of age or older) that estimate general intelligence (Wechsler Adult Intelligence Scale—Fourth Edition (WAIS-IV), “Matrices” and “Vocabulary”), executive functions (WAIS-IV, “Coding,” “Digit Span,” “Span Board,” and the Stroop test), and learning and memory (WAIS-IV, “List learning”) were analyzed using multiple linear regression with β values from DMPs in the $BDNF$, $NR3C1$, $NR3C2$, and $FKBP5$ genes as predictors, together with age, sex, and $β × sex$.

E. Functional Enrichment

E-1. GREAT analysis

To investigate the functional relevance of DEX-associated methylation changes, the Genomic Regions Enrichment of Annotations Tool (GREAT; version 3.0.0; http://bejerano.stanford.
edu/great) was applied [21]. Functional enrichment of DMPs was performed for DEX and DEX \times sex-associated DMPs and running separate analyses for the three lists of differential methylated probes described in Fig. 1A. Gene sets with a false-discovery rate (FDR) <0.05 were considered significant and selected. To avoid threshold-driven results, enriched gene sets from all analyses were subsequently overlapped, and a gene ontology (GO) term was considered to be enriched if it appeared to be significant in at least two gene-set enrichment analyses.

E-2. Enrichment analysis of disease susceptibility loci

With the purpose of evaluating whether altered states of DNA methylation increase susceptibility to disease, we investigated if DMPs (P < 0.01) were enriched at disease-associated loci. The enrichment was evaluated using gene-set enrichment analysis (GSEA) with the Gene Set Enrichment Analysis (GSEA) software (version 4.1.0). Gene sets were obtained from the Molecular Signatures Database (MSigDB) [22]. The FDR was set at 0.05, and the overlap of gene sets from all analyses was used to determine significant enrichment.

A) 

| Differential Methylation [DEX] | Differential Methylation [DEX \times sex] | Overlap [DEX + DEX \times sex] |
|-------------------------------|----------------------------------------|-------------------------------|
| p < 0.01: 9672 CpG sites; 5220 genes | p < 0.01: 7393 CpG sites; 4421 genes | Overlap: 2882 CpG sites |
| Hypermethylated: 3482 CpG sites | Hypomethylated: 6190 CpG sites | Hypermethylated: 3129 CpG sites | Hypomethylated: 4264 CpG sites |

| p < 0.01 + \beta-value > 5%: 2234 CpG sites; 1422 genes | p < 0.01 \beta-value > 5%: 2786 CpG sites; 1749 genes | Overlap: 776 CpG sites |
| Hypermethylated: 519 CpG sites | Hypomethylated: 1715 CpG sites | Hypermethylated: 1613 CpG sites | Hypomethylated: 1173 CpG sites |

| p < 0.01 \beta-value > 10%: 42 CpG sites; 24 genes | p < 0.01 \beta-value > 10%: 200 CpG sites; 159 genes | Overlap: 11 CpG sites |
| Hypermethylated: 19 CpG sites | Hypomethylated: 23 CpG sites | Hypermethylated: 89 CpG sites | Hypomethylated: 111 CpG sites |

B) 

Figure 1. Overview of the differential methylation analysis. (A) Overview of the differential methylation analysis showing results for the effect of DEX and DEX \times sex interaction. Number of DMPs in total, hyperhypomethylated, and unique genes is shown for the three levels of analysis. (B) Scatter plots showing $-\log_{10}(p)$ and $\beta$ values for all analyzed CpG sites in the 450K array for the effect of DEX (left) and for the interaction between DEX and sex (right).
SNPs identified in genome-wide association studies (GWAS; https://www.ebi.ac.uk/gwas/). We focused on a set of inflammatory and autoimmune disorders, where a programming effect for altered disease susceptibility as a result of DEX treatment could be plausible. These were asthma, pulmonary function, inflammatory bowel disease (IBD), ulcerative colitis, and rheumatoid arthritis. We also included a set of the following negative-control SNPs associated with terms unlikely to be affected by DEX: colorectal cancer, migraine, major depressive disorder, age-related macular degeneration, mean platelet volume, and iron status biomarkers. For each one of these 11 sets, we also computed a negative control set consisting of common SNPs acquired from the online University of California Santa Cruz SNP database (dbSNP; v.147; https://genome.ucsc.edu/). These sets were selected by matching each SNP with the CpG probe density of the SNP from the GWAS sets and thereby controlling for the number of SNPs included and for CpG probe density. Enrichment was computed using the Genetic Association Test (GAT) in four genomic bins: 1, 2, 5, and 10 kb around DMPs and SNPs [22]. Enrichments with an FDR <0.05 were considered significant.

F. Validation by Bisulfite Pyrosequencing

Bisulfite pyrosequencing was used to validate the 450K array. The selected CpGs were cg16280132 and cg26348243, located in the LTA gene and cg02322400 in the MAML2 gene. Methods and primers are available upon request.

2. Results

A. Differential Methylation Analysis

A-1. Genome-wide methylation analysis

In this study, we aimed to identify alterations in DNA methylation to find evidence for fetal epigenetic reprogramming after prenatal glucocorticoid treatment. Three levels of differential methylation associated with the DEX treatment and the DEX × sex interaction were investigated. We conducted the analysis in this manner on the basis of the following assumptions: (i) most differences in methylation between DEX-treated subjects and controls would be mild; (ii) we investigated a large number of probes that required multiple testing otherwise; (iii) our aim was to determine the biological impact of DEX on methylation level. An overview of the results from the differential methylation analyses is given in Fig. 1. The majority of the group differences, for the DEX and the DEX × sex-associated DMPs, was between 0% and 10% in magnitude (Fig. 1A). DEX × sex-associated DMPs had more hypomethylated probes (53.5%) than hypermethylated probes (46.5%) when investigating all analyzed sites (Fig. 1B). The percentages for DEX alone were 50.3% for hypermethylated probes and 49.7% for hypomethylated probes.

After correction for multiple comparisons, the distribution of the DMPs revealed an enrichment of them in gene bodies and intergenic regions in relation to the nearest gene (all values, \( P < 0.05 \)) and in open seas in relation to the nearest CpG island (all values, \( P < 0.05 \); Fig. 2). In addition, DEX × sex-associated DMPs enriched in south shelves <2 kb, flanking outward from a CpG shore (\( P < 0.05 \); Fig. 2). The pattern of probe distributions was similar for all probe outputs [23].

A-2. Overlap with epigenetic features

As enrichment of DMPs was identified both in gene bodies and in intergenic regions, we aimed to identify a mechanism by which these sites could potentially affect gene regulation. Therefore, we overlapped the probes that have a substantial change in methylation of at least 10% and that were located within intergenic regions or gene bodies with genomic regions enriched for the histone modifications H3K4me1 and H3K27ac (enriched at active enhancers)
or H3K36me (enriched in actively transcribed gene bodies). In addition, all probes, regardless of the location, were overlapped with DNase 1 hypersensitive sites, i.e., sites of uncondensed chromatin. DMPs were analyzed compared with the proportion of overlapping probes included in the 450K array using Fisher’s exact test. The proportion of probes in intergenic regions and associated with the DEX x sex interaction was significantly higher than expected for H3K4me1 sites (odds ratio = 2.84, \( P = 0.0004 \)) and for H3K27ac sites (odds ratio = 2.54, \( P = 0.002 \)). Genomic locations for markers were acquired from the Roadmap Epigenomics Project (http://egg2.wustl.edu/roadmap/web_portal/).

A-3. Targeted analyses of candidate genes

Furthermore, we wanted to investigate subsets of targeted genes from previous DNA methylation studies, targeting similar or close systems, to see whether we could replicate their results. We analyzed a subset of genes specifically reported to be differentially methylated in individuals exposed to high glucocorticoid/stress levels [12, 24–28] (Table 1). We also investigated other clinically/biologically relevant genes involved in the regulation and maintenance of DNA methylation, per se (Table 2), as well as genes involved in steroid action, regulation, and production (Table 3). All genes were analyzed independently on the 450K array, and CpG probes located within the gene or 1500 bp, up- or downstream, were evaluated. We identified DMPs in the majority of the targeted gene sets; results are presented in Tables 1–3.

A-4. Association between methylation and cognitive performance

We investigated whether methylation in DMPs in the \( BDNF, NR3C1, NR3C2 \), and the \( FKBP5 \) genes was associated with cognitive performance, assessed with the WAIS-IV and the Stroop tests [29]. These genes were chosen for this analysis as a result of their biological relevance for brain function and the fact that they all contained DMPs. In the \( BDNF \) gene, \( \beta \) values of two CpGs (cg10558494 and cg23947039) and their interaction with sex were associated with performance during the working memory task Digit Span and the learning and memory task List learning (immediate recall; all values, \( P < 0.05 \)). Furthermore, \( \beta \) values in cg06025631 and their interaction with sex were associated with performance during the...
processing speed task Coding (both \( P < 0.05 \)). In the \( FKBP5 \) gene, \( \beta \) values and their interaction with sex for cg14642437 were associated with performance on the task measuring fluid intelligence Matrices (both \( P < 0.05 \)). In the \( NR3C1 \) gene, \( \beta \) values in cg18068240 associated with performance in Matrices, as well as in the task-assessing ability to inhibit an overlearned response, “The Stroop color and word test” (both \( P < 0.05 \)). The interaction between \( \beta \) values and sex was also significantly associated with Matrices (\( P = 0.047 \)).

### B. Functional Enrichment

#### B-1. GREAT analysis

To add functional relevance, an enrichment analysis of DMPs was performed for GO terms in “Biological processes,” “Molecular functions,” and “Cellular components,” using the annotation tool GREAT. In total, 31 different GO terms were significantly enriched for either an association with DEX or for an association with DEX \( \times \) sex (FDR < 0.05) [29]. Enriched ontologies are summarized in Fig. 3. The majority of the enriched GO terms relates to immune functioning and inflammation, e.g., “interleukin 1 production and secretion” and “T cell receptor complex.” However, there are enriched terms related to biological functions unrelated to the functionality of the T cell, indicating an effect on multiple biological systems.

#### B-2. Enrichment analysis of disease susceptibility loci

GAT was used to investigate whether the DMPs enrich with disease risk SNPs from the GWAS catalog. Results for all enrichments are described [23]. Overall, the significance of the
colocalization increased with an increase in the size of the genomic bin. This is expected, as an increased bin size results in more overlaps between DMPs and SNPs. Here, we focus on the results from enrichment at 2 kb, as it has been shown that most CpGs are influenced by SNPs within a 2-kb range [30]. In the 2-kb bin, after FDR correction, DMPs associated with DEX show enrichment around IBD-associated SNPs (q = 0.022) and DEX × sex-associated probes enriched with asthma-associated SNPs (q = 0.022; Fig 4). The enrichment for IBD and asthma was substantial in all genomic bins analyzed (q < 0.05) [23]. No controls were enriched in the 2-kb bin (all q > 0.05).

C. Validation With Bisulfite Pyrosequencing

Results for five CpGs, located in the LTA and MAML2 genes, were replicated using bisulfite pyrosequencing. The directions of the changes in methylation corresponded to the changes seen with the Illumina 450K array for the CD4+ T cells (Table 4).

3. Discussion

To find evidence of long-lasting fetal programming effects after prenatal glucocorticoid treatment, we investigated DNA methylation in children at risk for, but not having, CAH,
who were treated with DEX during the first trimester of fetal life. The prenatal treatment of CAH presents a dilemma, as the majority of the treated cases is subjected to a potentially harmful therapy without any benefit. This type of therapy therefore requires an adequate risk-benefit analysis and a long-term follow-up of treated cases.

We choose the T cell as a model system, as it is easily accessible, and the first trimester DEX treatment affects the fetus during a period of gestation (gestation weeks 7 to 12) when the fetal thymus is formed, and the first lymphocyte gene rearrangements occur [31]. The T cell can be used to study the effects on this cell type per se but also as a model system for studying mechanisms or events that may occur in other cell types during embryogenesis after glucocorticoid exposure [32]. The sample size of our study is relatively small with a risk of detecting false positives. The study describes the epigenetic alterations from prenatal glucocorticoid treatment in humans. To evaluate how these alterations affect the health of treated individuals requires studies on larger cohorts, in addition to functional studies.

The genome-wide analysis revealed substantial differential methylation associated with prenatal DEX treatment, and in addition, the DEX effects were modified by sex. DEX × sex-associated probes also included more hypomethylated probes than hypermethylated ones. DMPs were enriched in gene bodies and intergenic regions. DMPs, located in intergenic regions and associated with the DEX × sex interaction, enrich with marks enriched for the specific histone modifications H3K4me1 and H3K27ac, i.e., markers of active enhancers. This suggests that changes in methylation associated with prenatal DEX treatment may affect the gene regulation through alterations of the chromatin state and thereby change the accessibility of regulatory elements in the genome. This is reinforced by the results from the functional enrichment analysis with GREAT, where DEX-associated probes are enriched for the GO term, “histone displacement” (Fig. 3). These data are in line with data from studies in rats, where prenatal DEX treatment decreased the production of proinflammatory cytokines, along with decreased signs of active chromatin (acetylation of histone 3 lysines, H3K4me1/3, and H3K36me3) in the promoter region of the TNF-α gene [33]. TNF-α contained DMPs in our study (Table 1).

| Gene    | Probe ID          | Location          | P[DEX]  | β[DEX] | P[DEX × Sex] | β[DEX × Sex] |
|---------|-------------------|-------------------|---------|--------|--------------|--------------|
| CRH     | cg19035496        | TSS200            | 0.020   | -0.04  | 0.047        | 0.05         |
| CRH     | cg08215831        | TSS200            | 0.072   | -0.03  | 0.018        | 0.05         |
| CRHR1   | cg27410679        | Body              | 0.008   | -0.05  | 0.031        | 0.06         |
| CYP11A1 | cg24482024        | TSS1500           | 0.009   | -0.03  | 0.039        | 0.03         |
| CYP11A1 | cg18068537        | TSS200            | 0.044   | -0.03  | 0.065        | 0.04         |
| CYP11A1 | cg06285340        | First exon; TSS1500 | 0.010   | -0.05  | 0.023        | 0.06         |
| CYP11A1 | cg03449379        | 3'UTR             | 0.010   | -0.04  | 0.196        | 0.03         |
| CYP11A1 | cg22186216        | Body; 5'UTR       | 0.031   | -0.03  | 0.065        | 0.04         |
| CYP11A1 | cg24578679        | Body; TSS200      | 0.012   | 0.01   | 0.040        | -0.01        |
| CYP11A1 | cg02916102        | First exon; body; 5'UTR | 0.118   | 0.01   | 0.029        | -0.02        |
| CYP11A1 | cg16332610        | First exon; body; 5'UTR | 0.213   | 0.01   | 0.036        | -0.02        |
| CYP11B1 | cg20073007        | Body              | 0.011   | -0.03  | 0.007        | 0.05         |
| CYP11B1 | cg05416055        | Body              | 0.044   | -0.03  | 0.045        | 0.05         |
| CYP11B1 | cg17015094        | TSS200            | 0.002   | -0.05  | 0.005        | 0.06         |
| CYP11B1 | cg21288978        | TSS200            | 0.007   | -0.04  | 0.029        | 0.05         |
| CYP11B2 | cg11324259        | TSS200            | 0.004   | -0.04  | 0.033        | 0.05         |
| CYP11B2 | cg07954324        | TSS200            | 0.019   | -0.03  | 0.108        | 0.03         |
| CYP19A1 | cg01916429        | 5'UTR             | 0.010   | -0.02  | 0.006        | 0.03         |
| CYP19A1 | cg19946085        | 5'UTR             | 0.022   | 0.01   | 0.005        | -0.01        |
| CYP21A2 | cg04771084        | TSS200            | 0.560   | -0.01  | 0.026        | 0.07         |
| SRD5A2  | cg04900872        | Body              | 0.002   | -0.05  | 0.012        | 0.05         |

The location and the direction of change in methylation are shown for probes importantly associated with prenatal DEX treatment or with the DEX × sex interaction. Probes were analyzed in a linear model, including age and sex as covariates. Significance was defined as an uncorrected P < 0.05. Significant results are marked in boldface.
Functional enrichment of the DMPs further revealed that the changes in DNA methylation in the T cell were functionally organized, especially regarding immune functioning and inflammation. We were able to show that DMPs were enriched near disease-related SNPs for IBD and asthma. This raises the possibility that when DEX alters DNA methylation in cis, it could contribute to the development of these disorders by altering gene expression. A consequence of prenatal DEX exposure may therefore be that the treatment creates a long-lasting program for altered immunity, which could potentially lead to development of immune-mediated inflammatory disease.

The results presented here are in line with the aforementioned Canadian study investigating long-term effects associated with the DEX treatment (white) and DEX × sex interaction (gray) or both (black). GO terms from the same biological pathway and that had overlapping probes are clustered together in a single bar with the lowest –log10(q value) available for the specific cluster. The bold line indicates significance threshold after FDR correction. AIM2, absent in melanoma 2; RES, retention and splicing.

Figure 3. Summary of enriched GO terms. Summary of enriched GO terms significantly associated with the DEX treatment (white) and DEX × sex interaction (gray) or both (black). GO terms from the same biological pathway and that had overlapping probes are clustered together in a single bar with the lowest –log10(q value) available for the specific cluster. The bold line indicates significance threshold after FDR correction. AIM2, absent in melanoma 2; RES, retention and splicing.

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The results presented here are in line with the aforementioned Canadian study investigating long-term effects associated with PNMS. T Cell DNA methylation was found to correlate with the levels of cytokines in the blood of the offspring [34]. The altered cytokine levels were subsequently attributed to a shift in the levels of T helper 1 cells toward T helper 2 cells [9, 34]. Moreover, higher levels of PNMS predicted a greater lifetime risk of wheezing and asthma. These effects were only observed in girls and suggest that maternal distress/glucocorticoid treatment during pregnancy affects fetal programming of the immune system and that these effects may be sex dimorphic [35].

In our targeted analysis, we identified DMPs in genes involved in the regulation and maintenance of DNA methylation (Table 2). This suggests that DEX treatment interferes with certain parts of the epigenetic regulatory system. Through this mechanism, it may have more widespread and long-lasting effects on gene regulation. These results are in line with studies in guinea pigs, where prenatal treatment with betamethasone was associated with altered expression of genes involved in the regulation and maintenance of DNA methylation per se [36].

We further identified several DMPs located upstream of the transcriptional start site region (TSS200, promotor regions) in genes involved in adrenal steroidogenesis (Table 3).
This may reflect an adaptation of the hypothalamic-pituitary-adrenal axis on the genomic level as a response to prenatal DEX treatment.

With regard to genes identified in the literature and being affected by glucocorticoids, stress, and traumatic events, all investigated genes contained DMPs (Table 1). Furthermore, in genes relevant for the central nervous system, we found important associations between DNA methylation in BDNF, NR3C1, and FKBP5 and performance in cognitive tasks [29]. We have previously identified deficits in executive functions in individuals exposed to DEX during the first trimester [2, 3]. The mechanism for these DEX-induced deficits is not known, but a link between epigenetic alterations in specific neurons and in cells involved in the function of the hypothalamic-pituitary-adrenal axis may be part of the answer.

### Table 4. Methylation Data for Validated CpG Sites

|                | MAML2               | LTA               |
|----------------|---------------------|-------------------|
|                | Pos. 1 (cg02322400) | Pos. 1 (cg16290132) | Pos. 2 (cg26348243) | Pos. 3 | Pos. 4 |
| 450K Array (T cells) |                     |                   |                   |        |        |
| DEX (%)        | 47.4 (9.9)          | 20.9 (5.5)        | 13.1 (7.0)        | –      | –      |
| Control (%)    | 42.1 (8.8)          | 17.2 (7.0)        | 7.0 (7.1)         | –      | –      |
| \(P_{\text{DEX}}\) | 3.33e-07            | 0.007             | 0.011             | –      | –      |
| Pyrosequencing (T cells) |                     |                   |                   |        |        |
| DEX (%)        | 32.9 (6.7)          | 16.5 (9.7)        | 16.4 (7.8)        | 12.1 (7.4) | 18.4 (8.7) |
| Control (%)    | 28.1 (7.5)          | 8.8 (11.2)        | 9.6 (9.4)         | 6.0 (9.1) | 10.2 (11.5) |
| \(P_{\text{DEX}}\) | 1.39E-05            | 0.023             | 0.042             | 0.066  | 0.013  |

Results from the 450K array and from the c nucleotide. Methylation at interrogated CpG sites in T cells showed a similar state in the 450K array as in the bisulfite pyrosequencing.
4. Conclusion

Here, we show associations between DNA methylation and first-trimester DEX treatment in treated subjects. The effects are moderated by sex and may change gene regulation by alterations in the state of chromatin. The effect of DEX on T cell DNA methylation is mostly related to immune functioning and inflammation and may therefore potentially program long-lasting effects on immunity and subsequently, lead to development of immune-related disorders.

How the identified alterations affect the health of the individual require studies on larger cohorts, in addition to functional studies.

In conclusion, this study indicates that prenatal DEX treatment affects fetal epigenetic programming, and together with the added cognitive risks, we recommend that the protocol for prenatal treatment of CAH should be revised to avoid treatment of healthy fetuses.

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