Evaluation of oxidative/nitrative stress and uterine artery pulsatility index in early pregnancy

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ABSTRACT

Introduction: Increased oxidative/nitrative stress is characteristic not only in pathologic, but also in healthy pregnancy. High uterine artery pulsatility index (UtAPI) at the end of the first trimester is associated with altered placentation and elevated risk for adverse pregnancy outcomes. We aimed to examine the relationship of systemic oxidative/nitrative stress and uterine artery pulsatility index in the first trimester and their correlation to pregnancy outcomes. Material and methods: Healthy pregnant women were recruited at 12–13th gestational week ultrasound examination; UtAPI was determined by color Doppler ultrasound. Patients were divided into high (UtAPI ≥ 2.3) (n = 30) and low (n = 31) resistance groups, and pregnancies were followed until labor. Systemic oxidative/nitrative stress was estimated by measuring total peroxide level, total antioxidant capacity and nitrotyrosine level. Results: Plasma total peroxide level was significantly lower (2,510 ± 39 mM vs. 2,285 ± 59 mM), total antioxidant capacity was higher (781 ± 16 mM CRE vs. 822 ± 13 mM CRE) in the high UtAPI group, which were accompanied by lower birth weight (3,317 ± 64 g vs. 3,517 ± 77 g, P < 0.05). Plasma total peroxide level showed a negative correlation (by Pearson) to UtAPI (P < 0.01) and positive correlation to birth weight (P < 0.05). Conclusions: According to our results, lower systemic oxidative stress showed correlation with high UtAPI measured between the 12–13th weeks of gestation. We also found significant...
differences in the birth weight of healthy newborns; therefore it is worth examining this relationship in pathological pregnancies.

**KEYWORDS**

uterine artery pulsatility index (UtAPI), pregnancy, birth weight, oxidative stress, nitrative stress

**INTRODUCTION**

The decrease in uterine artery pulsatility index (UtAPI) on the border of the first and second trimester is a well-established clinical marker of physiological placentation. This decline is due to the development of the specialized uteroplacental vasculature: expanded, high-flow, low-resistance vessels to satisfy the growing nutritional and oxygen requirements of the fetus [1]. However, delay in this process might contribute to altered placentation and can be a warning sign of later placental dysfunction.

Examination of the uterine artery’s blood flow by ultrasound Doppler method at the end of the first trimester with the determination of pulsatility index and resistance is widely accepted in obstetrical practice. Its predictive value was proposed in intrauterine growth restriction (IUGR) and preeclampsia. Over 95 percentile might be pathognostic to preeclampsia [2–5]. The simultaneous measurement of other early markers, like maternal plasma pregnancy related plasma protein A (PAPP-A), placental growth factor (PlGF), soluble fms-like tyrosine kinase-1 (sFlt-1), etc. may increase the sensitivity and specificity of the prognosis [6–8].

In healthy pregnancy and placentation, in the first trimester, during the period of trophoblast invasion, concentrations of reactive oxygen species (ROS) rise. At this time, placental oxygen tension is relatively low, ca. 20 mmHg. This low oxygen tension is necessary for producing the required level of angiogenic factors such as VEGF, PlGF and HIF resulting in placental angiogenesis and normal cell and tissue proliferation. When the maternal side of placental circulation has developed, both oxygen tension and ROS level get tripled. The placenta adapts to elevated oxygen tension and free radical concentration by modulating HIF-1α and increasing cellular levels of antioxidants. In physiological adaptation this process is advantageous in respect of fetal development and leads to a decrease in vascular resistance. In the high resistance group this process is partially flawed and results in damaged maternal vascular network of the placenta, which may be the consequence of lower oxygen tension and the elevation of ROS concentration. Oxygen- and nitrogen-derived reactive species are involved in biological pathways, where they act as mediators; however, their overproduction leads to oxidative/nitrative stress that takes part in the pathogenesis of numerous diseases [9, 10]. Oxidative stress increases during the course of physiological pregnancy; however, in pathological complications, like gestational diabetes mellitus (GDM), preeclampsia and IUGR this elevation is more pronounced both in the placenta and in the blood plasma [11, 12]. Nitrative stress rises by the third trimester in physiological pregnancy. In pathological conditions mentioned above, this elevation might appear earlier and can reach higher levels [13, 14].

Our aim was to examine the relationship between systemic oxidative/nitrative stress and the uterine artery pulsatility index (UtAPI) on the 12–13th week of healthy pregnancy, and their correlation to pregnancy outcome and neonatal characteristics.
MATERIALS AND METHODS

Patients

A prospective observational study was performed at the Department of Obstetrics and Gynecology and the Department of Physiology, Faculty of Medicine, Semmelweis University, Budapest, Hungary between 27 May 2016 and 12 December 2017. The study was approved by the Scientific and Research Ethics Committee of the Hungarian Medical Research Council (43102-2/2014/EKU (425/2014) 49768-1/2015/EKU (392/2015)). Written informed consent was obtained from all subjects.

Healthy pregnant women were invited randomly during the 12th week ultrasound screening (transabdominal sonography) to participate in the study (12+0–12+6 week of gestation). Method: A transducer should be positioned longitudinally and placed medially. The vessel can be identified with colored Doppler. The best place for measuring uterine artery PI and RI is 1 cm after the branching from the iliac artery. Exclusion criteria were age below 18 and over 40 years, malignant tumors, hypertension, diabetes mellitus, obesity (BMI over 30), twin pregnancy and chronic inflammatory diseases. During transabdominal ultrasound examination by GE Volusion E8 (Boston, MA, USA), flowmetric measurement by color Doppler method was performed on the uterine artery 1 cm from the branching of the iliac artery; the pulsatility index was determined according to the equation below. Pregnant women were divided into two groups based on the mean of the right and left artery pulsatility indices; normal UtAPIgroup (termed in the text as low UtAPI < 2.3), (n = 31) and high UtAPIgroup (n = 30).

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PI = \frac{\text{peak systolic velocity (PSV) } - \text{end diastolic velocity (EDV)}}{\text{time averaged velocity (TAV)}}
\]

Inclusion/admission method was: when recruiting a high UtAPI patient, we also recruited an age- and BMI-fitted low UtAPI patient. Cut off point was set to 90% percentile of the average of the European population [15]. The study size was estimated according to earlier measurements of plasma total peroxide level in healthy women. With a pooled standard deviation of 269 μM, a 200-μM difference by unpaired Student’s t-test may be detected with \( \alpha = 0.05 \) and a power of 0.8 by a sample size of 29 per group [16].

Study protocol

At the 12th week, following the ultrasound examination, participants filled in a questionnaire about their family, internal medicines and obstetric anamnesis, and blood samples were taken. From these samples routine laboratory tests (total blood cell count, CRP, hepatic and renal function) were performed; furthermore serum, plasma and mononuclear leukocyte fractions were isolated for measuring oxidative/nitrative stress laboratory parameters. Circulating mononuclear cells were isolated from blood samples by gradient centrifugation using Histopaque-1077 according to the users’ manual (Sigma-Aldrich, St. Louis, MO, USA). Methanol-fixed smears were made from the cell suspension and stored at 4 °C until further processing. Pregnancies were followed until birth in order to note pregnancy complications, labor circumstances, and neonates’ anthropometric data. We were not able to register labor circumstances in six cases (low UtAPIgroup: 4, high UtAPIgroup: 2).
**Determination of oxidative/nitrative stress related parameters**

Total plasma peroxide (PRX) concentration, reflecting systemic oxidative stress, was determined from plasma samples by colorimetric method using OxyStat assay (Biomedica, Wien, Austria). Plasma total antioxidant capacity was measured from plasma samples by a commercially available assay kit (OxiSelect™ Total Antioxidant Capacity (TAC) Assay Kit, Cell Biolabs Inc., San Diego, CA, USA). Nitrative stress was characterized by measuring serum levels of 3-nitrotyrosine (NT) using competitive ELISA based on HRP-conjugated anti-3NT antibody (OxiSelect™ Nitrotyrosine Elisa Kit, Cell Biolabs Inc., San Diego, CA, USA). To examine intracellular nitrative stress, immunohistochemistry was performed on methanol-fixed leukocyte smears with anti-nitrotyrosine rabbit polyclonal antibody (Abcam, Cambridge, UK) (1:80, 4 °C, overnight). Aspecific labeling was avoided by incubating the smears in 15% normal horse serum for 1 h at room temperature. Secondary labeling was achieved by horseradish peroxidase-conjugated horse anti-rabbit immunoglobulin (Vector Laboratories, Burlingame, CA, USA) (30 min, room temperature). Brown colored diaminobenzidine (6 min, room temperature, brown color) was used for visualizing the labeling (Vector Laboratories). Smears were counter-stained with blue colored hematoxylin (Vector Laboratories). Images of five representative microscopic fields of each smear were taken with 200x magnification (Zeiss-Imager. A1 light microscope, 20x/0.45 objective, AxioCam MRc5 camera, AxioVision – Rel. 4.8 software; Carl Zeiss Microscopy GmbH, Jena, Germany). The ratio of the positively stained cellular area compared to the total cellular area was determined by a blinded experimenter using ImageJ software (MBFImageJ, NIH, Bethesda, MA, USA); at least 300 cells were evaluated on each smear. Lower number of data in measured parameters was due to samples inadequate for analysis (hemolyzed plasma, serum or low cell count on leukocyte smears).

In order to examine the possible additive value of PRX in the correlation between UtAPI and birth weight, a new parameter was calculated, namely the quotient of UtAPI and plasma PRX: PRX:PIP\textsubscript{X} = UtAPI/PRX*100.

**Statistical analysis**

Normal distribution data are presented as mean ± SEM. In the case of non-Gaussian distribution, logarithmic transformation was implemented (TAC) and data are presented as median [IQR]. Statistical significance between the two study groups was determined by the two-tailed unpaired Student’s t-test. Nominal variables were tested by Chi-square test. Correlations between variables were estimated using Pearson’s correlation. Missing data were treated as missing. Probability values of \( P < 0.05 \) were considered significant. For the statistical analysis SPSS 22.0 and Graphpad Prism 6.0 softwares were used.

**RESULTS**

**Correlation between uterine artery PI and demographic and historical parameters**

Pulsatility index of the uterine artery as our grouping variable was significantly higher in the high resistance group (Fig. 1A).

Age and BMI did not differ in the study groups. On the other hand, gravidity, parity, and the number of miscarriages were significantly lower among women with higher UtAPI (Table 1).
Clinical laboratory parameters including plasma creatinine, liver enzymes, hemoglobin, CRP and glucose levels were similar in our study groups; only LDH was significantly lower in women with higher UtAPI (Table 1).

**Correlation between uterine artery pulsatility index and pregnancy outcome**

The rate of Cesarean section was also similar in the two groups. All newborns were in the normal body parameter range; however, newborns of the high UtAPI group had significantly lower birth weight and chest circumference than those of the low UtAPI group. There was no difference in gestational weeks at labor; therefore these differences were not due to earlier delivery (Table 1).

**Correlation between uterine artery PI and oxidative/nitrative stress**

Plasma total peroxide level, which refers to systemic oxidative stress, was significantly lower in the high UtAPI group than in the low UtAPI group (Fig. 1B), whereas total antioxidant capacity was significantly higher in the high UtAPI group (Fig. 1C). PIPX calculated from UtAPI and PRX (PIPX: UtAPI/PRX × 100) was significantly higher in the high UtAPI group (Fig. 1D). On the other hand, serum and mononuclear cell NT levels were similar in the low and high UtAPI groups (Fig. 2A and B).
Table 1. Clinical and anthropometric data of women and their offspring. The two study groups were matched according to their age and BMI. On the other hand, gravidity, parity and the number of previous miscarriages were lower in the high UtAPI group. There were no significant difference between the two groups in their major clinical laboratory parameters, however LDH level was significantly lower in the high UtAPI group. Development of preeclampsia was not observed among the study subjects. The occurrence of GDM and Cesarean section was similar in the two groups. The newborns of the high UtAPI group had smaller birth weight and chest circumference despite the gestational age of the mothers at labor being similar. Data are presented as mean ± SEM, where not indicated otherwise.

| Variable                      | low UtAPI (n = 31) | high UtAPI (n = 30) | Significance |
|-------------------------------|-------------------|---------------------|--------------|
| **Anamnestic data**           |                   |                     |              |
| Age (years)                   | 32.00 ± 0.66      | 31.00 ± 0.73        | ns           |
| (min–max) (26–39)             | (22–38)           |                     |              |
| BMI (kg/m²)                   | 21.60 ± 0.71      | 22.60 ± 0.39        | ns           |
| (min–max) (0–6)               | (0–4)             |                     |              |
| Gravidity                     | 1.60 ± 0.27       | 0.80 ± 0.19         | P < 0.05     |
| (min–max) (0–6)               | (0–4)             |                     |              |
| Parity                        | 1.00 ± 0.19       | 0.40 ± 0.15         | P < 0.05     |
| (min–max) (0–4)               | (0–4)             |                     |              |
| Miscarriages                  | 0.50 ± 0.15       | 0.10 ± 0.07         | P < 0.05     |
| **Clinical parameters**       |                   |                     |              |
| LDH (U/L)                     | 152.66 ± 2.75     | 144.70 ± 2.47       | P < 0.05     |
| Hemoglobin (g/L)              | 128.73 ± 1.53     | 126.92 ± 1.56       | ns           |
| Hematocrite (L/L)             | 0.38 ± 0.00       | 0.37 ± 0.00         | ns           |
| Glucose (mmol/L)              | 4.60 ± 0.12       | 4.63 ± 0.14         | ns           |
| Triglycerides (mmol/L)        | 1.40 ± 0.10       | 1.45 ± 0.73         | ns           |
| HDL (mmol/L)                  | 1.87 ± 0.05       | 1.77 ± 0.05         | ns           |
| LDL (mmol/L)                  | 3.31 ± 0.13       | 3.30 ± 0.10         | ns           |
| Bilirubin (µmol/L)            | 8.34 ± 0.79       | 7.80 ± 0.42         | ns           |
| Creatinine (µmol/L)           | 47.77 ± 1.55      | 47.43 ± 1.25        | ns           |
| AST (U/L)                     | 18.45 ± 0.53      | 18.70 ± 0.83        | ns           |
| ALT (U/L)                     | 14.58 ± 0.88      | 15.77 ± 1.62        | ns           |
| γGT (U/L)                     | 13.32 ± 0.78      | 13.17 ± 0.96        | ns           |
| CRP (U/L)                     | 6.27 ± 0.70       | 5.99 ± 0.60         | ns           |
| ALP (U/L)                     | 56.81 ± 2.89      | 53.60 ± 2.25        | ns           |
| **Pregnancies’ outcome**      |                   |                     |              |
| Cesarean section (N)          | 13                | 11                  | ns           |
| out of emergent fetal/maternal indication/total (N) | 1/1/2 | 3/2/5 | ns |
| GDM (N)                       | 3                 | 3                   | ns           |
| PE (N)                        | 0                 | 0                   | ns           |
| IUGR (N)                      | 0                 | 0                   | ns           |
| **Newborns’ anthropometry**   |                   |                     |              |
| Bodyweight (g)                | 3,517.41 ± 77.02  | 3,316.79 ± 63.76    | P < 0.05     |
| Gestational week              | 39.80 ± 0.19      | 39.30 ± 0.20        | ns           |
| Chest circumference (cm)      | 34.41 ± 0.29      | 33.57 ± 0.26        | P < 0.05     |
| Head circumference (cm)       | 35.00 ± 0.26      | 34.39 ± 0.30        | ns           |
| Apgar 1’                      | 9.44 ± 0.11       | 9.61 ± 0.09         | ns           |
| Apgar 5’                      | 10.00 ± 0.00      | 10.00 ± 0.00        | ns           |
Correlations between oxidative/nitrative stress, UtAPI and clinical data in the total study cohort

In the total study cohort, plasma total peroxide level showed negative correlation to UtAPI and positive correlation to the birth weight of the newborns (Table 2). On the other hand, TAC showed negative correlation to the number of previous pregnancies and positive correlation to serum bilirubin and creatinine levels (Table 2). Intracellular NT immunohistochemical staining intensity showed positive correlation to γGT levels. It also positively correlated to the occurrence of Cesarean section.

We also found that the negative correlation of UtAPI to birth weight in our cohort can be improved by combining UtAPI measurements with plasma PRX level by calculating their ratio (PIPX) (Table 3).

Fig. 2. Pulsatility index and nitritative stress markers. Panel A and B. Serum and mononuclear cell 3-nitrotyrosine levels were similar in the study groups. (N = 21; 28 and N = 16; 14). Data are presented as mean ± SEM, or Median [IQR]in case of TAC. *: P < 0.05, **: P < 0.01, ***: P < 0.001

Table 2. Clinical parameters correlating to systemic oxidative/nitrative stress markers measured on the 12–13th weeks of gestation. Plasma total peroxide level positively correlated to birth weight. Plasma total antioxidant capacity negatively correlated to the number of previous healthy pregnancies and positively to serum bilirubin and creatinine levels. Intracellular tyrosine nitration of circulating mononuclear cells positively correlated to γGT levels and the occurrence of Cesarean section.

| Dependent variable | R     | P     | N  |
|--------------------|-------|-------|----|
| **Independent variable: Plasma Total Peroxide** |       |       |    |
| Birth weight       | 0.342 | 0.031 | 40 |
| UtAPI              | −0.428| 0.004 | 44 |
| **Independent variable: Log (Plasma TAC)**     |       |       |    |
| Gravidity          | −0.323| 0.015 | 56 |
| Bilirubin          | 0.268 | 0.045 | 56 |
| Creatinine         | 0.357 | 0.007 | 56 |
| **Independent variable: Leukocyte NT**          |       |       |    |
| γGT                | 0.407 | 0.026 | 30 |
| Cesarean section   | 0.391 | 0.048 | 26 |
Results only for primiparas

All statistical analysis was also repeated with primiparas only, and the results were similar to those of the entire study group.

**DISCUSSION**

Several research groups have examined the relationship between the first and second trimesters’ elevated UtAPI or resistance index and lower birth weight, mainly because of the risk for intrauterine growth restriction (IUGR) [17–20]. In our healthy study cohort, the newborns of high-UtAPI mothers were born with significantly smaller weight and chest circumference than the newborns of the low-UtAPI group, although there was no difference in the gestational week at birth. In the total study cohort, UtAPI negatively correlated to the birth weight of the neonates that were within the normal range. These results may suggest that UtAPI measured in this period may indicate later placental function and fetal development also in physiological pregnancy.

Plasma PRX indicating oxidative stress was significantly lower in the high UtAPI group, whereas total antioxidant capacity was significantly higher compared to the low UtAPI group. Low oxygen tension in the first trimester having an important role in the initial normal placentation triples by the end of the first trimester, along with the levels of reactive oxygen species, a change that significantly contributes to further placental development. The placenta adapts to the elevated oxidative stress by modulating the levels of cellular antioxidants [21]. The low level of oxidative stress in the high UtAPI group may reflect the impairment or delay of this oxidative burst. This hypothesis may be supported by the negative correlation of oxidative stress to UtAPI and its positive correlation to birth weight of the neonates in the total study cohort even in physiological pregnancies – therefore further examination of oxidative stress markers might be also useful in pathological pregnancies. That is the reason why we try to develop more sensitive diagnostic indicators, like PIPX, involving classical and oxidative stress parameters.

As both UtAPI and plasma PRX showed a correlation to lower birth weight, we generated a variable that reflects their simultaneous change defined as PIPX:UtAPI/PRX*100. PIPX showed a stronger correlation to birth weight than UtAPI alone, suggesting that measuring oxidative stress may improve the predictive value of UtAPI on fetal development.

At this time, together with the oxidative burst, the secondary invasion of the trophoblast cells occurs in the placenta [22] that is associated with cellular degradation. Tissue degradation usually leads to the elevation of serum LDH concentration. In our study, serum LDH

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Table 3. Correlation of birth weight of the newborns to UtAPI, plasma total peroxide level and their combined variable PIPX (UtAPI/PRX*100). All analyzed parameters showed correlation to birth weight; however, the strongest correlation could be found in case of PIPX

| Dependent variable | R     | P     | N   |
|-------------------|-------|-------|-----|
| **Independent variable: UtAPI** |       |       |     |
| Birth weight      | −0.347| 0.009 | 55  |
| **Independent variable: PIPX** |       |       |     |
| Birth weight      | −0.450| 0.004 | 40  |
concentration was lower in the high UtAPI group at the end of the first trimester. We hypothesize that this lower LDH reflects the decreased trophoblast invasion.

Additionally, it has been previously shown by our research group that even after a healthy pregnancy, the degree of oxidative stress is elevated after three years [16]. It is also conceivable that low-resistance pregnant women had higher oxidative stress levels because of their higher number of previous pregnancies. Our observation that plasma total antioxidant capacity negatively correlates to the number of previous pregnancies may also support this hypothesis.

Gravidity and parity were significantly higher in our low UtAPI group. This observation may be related to findings showing that multiparity protects against the development of high uterine artery pulsatility index. The significantly higher parity and gravidity in our low UtAPI group may reflect that multiparous women’s body had previously practiced the steps of placentation, leading to a higher chance of success in trophoblast invasion and circulatory adaptation [23].

Despite the decreased oxidative stress in the high UtAPI group, we found no difference in nitrative stress between the two study groups. Nitrotyrosine is mainly produced by the reaction of protein tyrosine residues and peroxynitrite, whereas peroxynitrite is produced by the reaction of nitric oxide and superoxide anion [24]. According to this reaction pathway, the change in nitrotyrosine levels might be expected to follow the changes in PRX concentration. On the other hand, previous studies also failed to confirm to this relationship, especially in chronic disease and healthy conditions [16, 25].

Intracellular nitrative stress showed a positive correlation to serum γGT value. Several prior studies suggested that γGT could be used as a clinical marker of inflammation and oxidative stress. Simona Bo et al. studied the relationship between serum NT and serum γGT. Based on their results, these two markers showed a positive correlation [26]. An interesting finding of our study is that the level of intracellular NT also correlated to Cesarean section outcome, however further investigations are required to clarify its mechanism and importance.

The limitations of the study were the relatively small number of cases and the high variability of oxidative/nitrative stress markers among pregnant women. Due to these, we were not able to define distinct cutoff values of oxidative/nitrative stress markers for pregnancy outcomes. Combined parameters, like the variable calculated from UtAPI and PRX, could be expected to improve the predictive value of oxidative/nitrative stress parameters. On the other hand, additional studies are required to identify a more clinically relevant oxidative/nitrative stress parameter.

**CONCLUSION**

In conclusion, according to our results high UtAPI without other risk factors results in smaller newborns in the normal range. On the other hand, plasma oxidative stress and LDH were decreased in the high UtAPI group of our cohort that may be in connection with the disturbance of placentation. This hypothesis is further supported by our finding that combining UtAPI with an oxidative stress parameter (PRX) strengthens its correlation to birth weight. Measurement of oxidative stress markers together with UtAPI at the 12–13th weeks of gestation may provide additional information about placentation. We found significant differences also in the birth weight of healthy newborns; therefore it is worth examining this relationship in pathological pregnancies.
Data availability: According to the ethical approval, the detailed datasets of the present study cannot be shared with a third party without permission. Permission may be asked upon individual request.

Conflict of interest: The authors declare that no competing financial interests exist.

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ABBREVIATIONS

γGT Gamma-glutamyl transferase
GDM Gestational diabetes mellitus
HRP Horse raddish peroxidase
IUGR Intrauterine growth restriction
LDH Lactate dehydrogenase
NT Serum nitrotyrosine
PIGF Placental growth factor
PAPP-A Plasma pregnancy related plasma protein A
PE Praeclampsia
PIPX The quotient of UtAPI and plasma PRX, UtAPI/PRX*100
PRX Plasma total peroxide
sFlt-1 Soluble fms-like tyrosine kinase-1
TAC Plasma total antioxidant capacity
UtAPI Uterine artery pulsatility index

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