Title
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Permalink
https://escholarship.org/uc/item/9vh713g4

Journal
Inflammation research : official journal of the European Histamine Research Society ... [et al.], 60(1)

ISSN
1023-3830

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Publication Date
2011

DOI
10.1007/s00011-010-0238-9

Peer reviewed
Human skin penetration of a copper tripeptide in vitro as a function of skin layer

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Received: 24 September 2009 / Accepted: 16 July 2010 / Published online: 20 August 2010 © The Author(s) 2010. This article is published with open access at Springerlink.com

Abstract

Objective and design Skin retention and penetration by copper applied as glycyl-L-histidyl-L-lysine cuprate diacetate was evaluated in vitro in order to assess its potential for its transdermal delivery as an anti-inflammatory agent.

Materials and methods Flow-through diffusion cells with 1 cm² exposure area were used under infinite dose conditions. 0.68% aq. copper tripeptide as permeant was applied on isolated stratum corneum, heat-separated epidermis and dermatomed skin and receptor fluid collected over 48 h in 4 h intervals using inductively coupled plasma mass spectrometry to analyze for copper in tissues and receptor fluid.

Results The permeability coefficient of the compound through dermatomed skin was $2.43 \pm 0.51 \times 10^{-4}$ cm/h; 136.2 $\pm$ 17.5 µg/cm² copper permeated 1 cm² of that tissue over 48 h, while 97 $\pm$ 6.6 µg/cm² were retained as depot.

Conclusions Copper as tripeptide was delivered in potentially therapeutically effective amounts for inflammatory disease.

Keywords Absorption · Arthritis · Bioavailability · Copper complex · Depot · Inflammation

Introduction

Acute and chronic inflammation are characterized by changes in the metabolism of copper and by a pronounced responsiveness to therapy with copper compounds [1, 2]. The copper content and ceruloplasmin activity of serum are significantly elevated above normal values in inflammatory diseases in man and laboratory animals, and copper in widely different chemical forms is used as therapeutic agent in therapy for chronic and acute inflammation [2]. The rise in total serum copper measured in inflammation is held to represent the natural anti-inflammatory (AI) response of the organism itself. The responsiveness of inflammatory disorders to copper supplementation noted suggests that the control exerted by endogenous copper on inflammation is susceptible to enhancement by exogenous sources. When copper complexes were administered subcutaneously, the reduction in inflammation was proportional to total amount of copper injected [3].

The present research seeks a viable transdermal alternative to what so far has been a therapy for inflammatory disorders relying on intra-articular or intramuscular injection of drugs which can cause patient discomfort and untoward side effects. Data on human skin penetration by copper compounds is rare. Pirot et al. [4] determined the Skin permeability (Kp) for copper sulfate and chloride at $3.2 \times 10^{-6}$ cm/h, respectively, for the pyrrolidone carboxylate at $5.7 \times 10^{-5}$ cm/h. Using a model membrane made of stratum corneum lipids, Mazurowska suggested that copper peptides, and GHK-Cu in particular, might permeate skin [5–7]. Experiments using skin models generally underestimate absorption since the composition of model membranes does not reflect the stratum corneum, a heterogeneous, multicomponent membrane [8–10]. Orth [11] and Finkey [12] measured uptake of copper into...
superficial facial skin strata in vivo when applying glycyl-
L-histidyl-L-lysine copper cuprate, diacetate (GHK-Cu).
However, to date, bioavailability of copper was not studied
quantitatively using human skin ex vivo. Therefore, evi-
dence that copper does penetrate the skin in any form is so
far based on its observed effects only. Anti-inflammatory
(AI) activity by topical application of copper complexes
was shown in animals by Brown [1], and in man by
Sorenson [2], and has been extensively reviewed by
Milanino [13].

Chemical: Glycyl-L-histidyl-L-lysine copper cuprate, diacetate GHK-
Cu(Ac)_2, Formula: C_{18}H_{29}CuN_{6}O_{8}

Criteria for optimal diffusivity of cupriphores are hard to
define due to the complexity of equilibria that zwitterions
are subject to in the variable systemic environments they
encounter upon diffusion. Earlier investigations, however,
help in the design of complexes of adequate pharma-
ecological activity. Those include:

- Copper derivatives of well-known antiarthritic drugs,
  including steroidal and nonsteroidal AI agents. The
copper complexes of those drugs have been found to be
active or more effective AI agents than the parent drugs
[14].
- The AI effect of copper is non-specific with respect to
inflammatory pathologies and is enhanced by exoge-
nous sources [13].
- AI activity of exogenous copper sources is independent
of ligand [15]. On topical administration, copper
complexes are subject to dynamic ligand exchange in
the process of diffusion through skin [16]. Thereby,
copper oxidation state can change from monovalent to
divalent to the elemental state, and the original complex
administered topically is likely to differ from the
complex which subsequently forms with endogenous
amino acids to act as an AI agent [17].

The function of copper ligands thus is to promote skin
penetration only, and this study evaluates skin penetration
of a nature-identical copper complex [18]. If therapeuti-
cally significant, that is, if transdermal AI therapy can
be achieved, it may present an alternative to current
approaches involving intra-articular or intramuscular
injection, which can cause patient discomfort, particularly
in arthritic patients where long term therapy is employed.

Materials and methods

Materials

The permeant was GHK-Cu received from the ProCyte
Corp. (Kirkland, WA). It was not analyzed for stoichiom-
metric purity since, in several studies, copper complexes in a
physiological environment were subject to de-complexation
and rearrangement, depending on pH and complex stability
constants [11, 19–21]. Those investigations indicated
the presence of multiple species for GHK-Cu (e.g., tri-and
hexa-peptide) in solutions, together with copper-free
oligopeptide. Copper as a transition metal of variable oxi-
dation states has the ability to form complexes with
different geometries and stability. Here, the copper tripep-
tide is summarily referred to as GHK-Cu. Concentrations
are reported in terms of copper per ml of solution.

Experimental design

Copper penetration was studied using split thickness skin,
isolated stratum corneum and entire epidermis.

Tissue preparation

Human cadaver skin was obtained from the Northern
California Transplant Bank (San Rafael, CA) and stored in
a freezer (−20°C) until use. All experiments were per-
formed within 2 months after freezing. All subjects were
Caucasian, aged 21–53 years, and both genders were
represented.

Split thickness skin (entire epidermis with a portion of
the dermis) was obtained by use of a dermatome targeted to
500 μm and stored (−20°C) until use.

Entire epidermis, (isolated. epidermis incl. stratum cor-
neum), was obtained by heat separation (60°C) and stored
(−20°C) until use.

Isolated stratum corneum was obtained from isolated
epidermis which was placed on Whatman paper soaked in a
0.05% w/w trypsin Type III (from bovine pancreas, Sigma
Chemical, St. Louis, USA) solution in 0.15 M phosphate
buffered saline (PBS, pH 7.4, Sigma) for 2 h at 37°C.
Remaining trypsin activity was blocked by submerging the
stratum corneum sheet in 0.05% w/w trypsin inhibitor
Typed II (from soybean, Sigma Chemicals, St. Louis,
USA), then the stratum corneum sheet was washed twice in
purified water, dried and stored above silica gel at room
temperature.
Separation of epidermis (incl. stratum corneum) from dermis following split thickness skin diffusion experiment C (sep. epidermis): recovered split thickness skin was placed on polyethylene film, covered with polyethylene film and placed under an aluminum block heated to 65°C for 90 s, allowing separated epidermis to be lifted from the dermis.

Diffusion experiment

Replicates were run on tissue from the same donor. Glass low-volume flow cells with 1 cm² exposure area were kept at 37°C. Receptor fluid was collected over 48 h in 4 h intervals. Cells were mounted in the wells of an aluminum cell holder (Stratacor, Richmond, CA), whose internal channels were perfused with water at 37°C from a circulating water bath, and holders were mounted on a fraction collector. Water containing 1 ml/l of gentamycin sulfate solution (100 mg/ml) was pumped through the cells at approximately 2 ml/h. Tissues were cut into circles with a biopsy punch (1 cm²), mounted on the diffusion cells and the donor chambers clamped in place.

To test for tissue integrity, prior to the experiment with tissue discs clamped in place, the peristaltic pump was run at 10 rpm, generating back pressure in the receptor cells (five times the normal pressure) for 2 h to check for leakers, which were eliminated.

About 1 ml of test solution was placed in the donor chambers, closed with ground glass stoppers and sealed with Parafilm to prevent donor solution evaporation.

At completion, the content of donor cells were collected with a pipette, the skin surface washed (3 × 9 ml) with distilled water and the rinses saved in LSC vials with polyethylene capliners (Perkin-Elmer, Shelton, CT).

Analytical methodology

For original copper levels, portions of isolated stratum corneum, isolated epidermis, as well as split thickness dermis were analyzed to determine the increment retained there during diffusion. Following diffusion experiment C, split thickness skin was also separated into epidermis and dermis (entire separated epidermis and separated dermis) for analysis of tissue depot formation in the experiment.

Experimental details and results are reported in terms of copper, analyzed by inductively coupled plasma mass spectrometry (ICP-MS).

Analyses were performed by DuPont Analytical Solutions, Wilmington, DE. There, samples were prepared by diluting 1:10 (volume/volume), using 2% HNO₃ immediately prior to analysis by ICP-MS. The ICP-MS equipment used was an Agilent 7500c, operated under helium collision gas mode. The instrumental detection limit was <100 pg/ml copper. Standard solutions were prepared by diluting a 1,000-µg/ml Cu stock solution with 2% HNO₃. Two separate (low concentration and high concentration) six-point external calibration curves (r² > 0.999) were used for quantitation of both copper isotopes (63 and 65 Cu) with 89Y used as an internal standard. The low concentration calibration curve was constructed from 1, 2, 5, 10, and 20 ng/ml Cu standards and the high concentration calibration curve was from 20, 100, 1,000, 5,000, and 10,000 ng/ml Cu standards. The instrument was calibrated using the five standards above, and the samples bracketed between the standards without extrapolation either below the calibration curve or above it.

Data analysis

Kp (cm/h) was calculated from steady-state flux Jss (µg/cm²/h) of copper. Tissue copper levels reported from diffusion experiments were corrected for endogenous levels determined prior to the experiments.

Standard deviations (SD) were calculated by the XL program.

Results

Skin permeability coefficients (Kps) for the copper tripeptide ranged from 3 × 10⁻⁷ cm/h through isolated epidermis (Table 1, Exp. B) to 5.54 × 10⁻⁴ cm/h through isolated stratum corneum (Table 1, Exp. A).

Lag times measured from 2 h on isolated stratum corneum and split thickness skin (Table 1, Exp. A, C), to 7 h on isolated epidermis (Table 1, Exp. B).

Copper collected in the receptor fluid in 48 h ranged from 0.60 ± 0.21 µg/cm² or 0.006 ± 0.001% of the dose through isolated epidermis (Exp. B; Table 2), to 136.2 ± 17.5 µg/cm² or 2.0 ± 0.31% through split thickness skin (Exp. C, Table 2).

Amounts of membrane retention in stratum corneum tissue were 290 ± 69 µg/cm², a 438-fold increase over baseline levels (Tables 2, 3, Exp. A), and a total 96.9 µg/cm² in the complete dermatomed skin (Table 2, Exp. C) based on analysis of the separated strata after diffusion through split thickness skin.

Discussion

This study evaluated absorption and skin retention of a copper tripeptide on separate skin compartments: the stratum corneum, epidermis and split thickness skin.

The amount of copper deposited from the tripeptide in the stratum corneum as well as in the entire epidermis and
C Split thickness

B Isolated epidermis

Accordingly, at least 80% of the amount in the stratum corneum will become available if the lag time for the turnover time for the stratum corneum [23]. Retained drug may be lost to natural desquamation, limiting its eventual systemic availability, to be sure. On closer analysis, however, it was also found that the fraction of chemical that sloughs off versus the part that becomes systemically available depends on the ratio of lag time to the turnover time for the stratum corneum [23]. Accordingly, at least 80% of the amount in the stratum corneum will become available if the lag time for penetration through the stratum corneum is less than about 16 h, which corresponds to 5% of the stratum corneum turnover time of typically 14 days. Consequently, based on our results where lag times ranged from 2 to 7 h, the copper retained in the stratum corneum may become largely systemically available. As a result of the experiment with dermatomed skin, by topical application of GHK-Cu under the present conditions 200–250 μg/cm² copper would become systemically available. Since the experiment was conducted ex vivo, it is possible that even larger amounts will be retained in the deeper dermal tissue there, followed by penetration in vivo.

The permeation rate of GHK-Cu through stratum corneum seems high, the highest of all three skin strata studied. It seems consistent with a model of porous transport, as appendageal penetration is considered to be one, if not the predominant pathway for the diffusion of multipolar ions [24–26]. In addition, the high penetration level through stratum corneum alone can be explained by the fact that there was no tissue retention from underlying skin that may have limited penetration.

dermis appears remarkable. For instance, an about 400-fold increase over baseline was found in the stratum corneum alone. Such high tissue retention of copper may be attributed to intradermal decomplexation and re-binding to various endogenous amino acids with higher nucleophilic donor capacity than the original ligand.

Whether or not skin levels retained in the stratum corneum and epidermis/dermis in the diffusion process of drugs should be included in the overall estimation of absorbed material, i.e., assessing whether permeant retained in skin tissues eventually will become available systemically, is an issue of ongoing discussions. Reservoir formation in the course of in vitro studies, in particular, has been addressed [22]. Retained drug may be lost to natural desquamation, limiting its eventual systemic availability, to be sure. On closer analysis, however, it was also found that the fraction of chemical that sloughs off versus the part that becomes systemically available depends on the ratio of lag time to the turnover time for the stratum corneum [23].

Table 1 Copper skin penetration as function of skin strata from aq. GHK-Cu solution

| Exp. | Skin strata                        | Kp × 10² (cm/h) ± SD | Flux (μg/cm²/h) ± SD | Lag time (h) |
|------|-----------------------------------|----------------------|---------------------|--------------|
| A    | Isolated stratum corneum (n = 3)  | 55.9 ± 13.2          | 38 ± 9              | 2            |
| B    | Isolated epidermis (n = 3)        | 0.003 ± 0.0006       | 0.002 ± 0.0004      | 7            |
| C    | Dermatomed skin (n = 8)           | 2.5 ± 0.51           | 1.7 ± .35           | 2            |

Table 2 Human skin penetration and retention of copper in function of skin strata after 48 h (μg/cm²; percent of applied dose)

| Exp. | Skin strata                        | Donor recovered | Stratum corneum | Total epidermis | Dermis | Receptor fluid | Total recovered |
|------|-----------------------------------|-----------------|-----------------|-----------------|--------|----------------|----------------|
| A    | Isolated stratum corneum (n = 3)  | (79 ± 13)       | 290.3 ± 69 (4.0 ± 0.15) | NA | NA | 2,110 ± 589 (19.85 ± 3.9) | (98 ± 27) |
| B    | Isolated epidermis (n = 3)        | (94 ± 12)       | ND              | 59.7 ± 13.1 (1.0 ± 0.3) | NA | 0.60 ± 0.21 (0.006 ± 0.001) | (95 ± 12) |
| C    | Split thickness (n = 8)           | 91 ± 14         | ND              | 1.0² ± 0.4 (66.0 ± 6.6) | 0.09 ± 0.01 (30.9² ± 6.8) | 136.2 ± 17.5 (2.0 ± 0.31) | (94 ± 7) |

NA not applicable; ND not determined

a Separated following split thickness diffusion

Table 3 Copper reservoir in skin layers versus original levels from 0.68% aq. GHK-Cu at 48 h

| Skin compartment | Cu at 0 h (μg/cm²) | Cu at 48 h (μg/cm²) | Increase vs. 0 h |
|------------------|--------------------|---------------------|------------------|
| Isolated SC (Exp. A) | 0.68 ± 0.28; n = 3 | 290 ± 69; n = 3 | 438×             |
| Isolated Epi (Exp. B) | 0.37 ± 0.17; n = 3 | 61.1 ± 7.0; n = 3 | 165×             |
| Dermatomed skin (Exp. C) | 1.72 ± 0.47; n = 8 | 53 ± 8.1; n = 8 | 31×              |
| Separated epidermisᵃ | 0.37 ± 0.7; n = 8 | 66.7 ± 7.89; n = 8 | 180×             |
| Separated dermisᵇ | 1.72 ± 0.27; n = 8 | 27.63 ± 8.83; n = 8 | 16×              |

ᵃ Separated following split thickness diffusion; total epidermis
ᵇ Dermis separated from split thickness skin following the diffusion experiment
The high hydrophilicity of the copper tripeptide (pseudo log \( P_{ow} \approx -4.5 \), quotient sat. n-octanol/sat. aqueous solution),\(^1\) is reconcilable with shunt penetration typical for highly hydrophilic solutes such as salts. Significant penetration through appendages can occur within minutes following exposure, in contrast to non-appendageal pathways [27–30].

However, the Kps of GHK-Cu through epidermis and dermatomed skin were lower at about one order of magnitude. Restriction of the pore area may cause the discrepancy in the diffusion constant. This suggests, in fact, that the diffusion pathway for copper penetration is restricted as compared to stratum corneum alone.

Skin retention and permeation by metal complexes is not often investigated, and our observations on the subject of a copper tripeptide merit more detailed discussion.

Copper is an essential trace element, critical for a variety of biological processes; for hemoglobin synthesis, enzyme activation, (i.e., superoxide dismutase), or more generally as a key component of mitochondrial, cytoplasmic and nuclear enzyme systems [31].

It is a key factor in the production of collagen and elastin [11, 12, 18], and the AI activity of numerous copper complexes is well documented [13, 32]. Such activity was greater than either treatment with inorganic copper salts or the complexing agents alone: while many among the parent compounds or ligands were inactive in test models of inflammation even at larger screening doses, in coordination with copper they were active. The compounds thus investigated include copper complexes of well-known antiarthritis drugs, including steroidal and nonsteroidal anti-inflammatory drugs: niflumic acid, D-penicillamine, hydrocortisone, dexamethasone, dimethylsulfoxide, clopirac, ketoprofen, (+)-naproxen, indomethacin, mefenamic acid, diclofenac, ibuprofen among a number of others. The copper complexes of those drugs have been found to be more effective anti-inflammatory agents than the parent drugs [33, 34].

Acute or chronic inflammation induces a net increase of total copper in blood, synovial fluid and inflamed areas, mobilized from endogenous stores [13]. Chronic inflammation may lead to depletion of copper stores, implying the need for exogenous copper supply in order to effectively cope with the inflammatory pathologies. While copper complexes are commonly administered by injection, the AI activity of subcutaneous injection of copper aspirinate in rats without adverse effects, while it proved effective in treating experimental subplantar edema even at a dose of 10 mg/kg.

When administering copper to the organism, its toxicity should be addressed.

There is ample evidence that the copper ions released will be subject to the highly efficient homeostatic control in eukaryotic organisms that prevents its circulation at undetermined levels. Levels of free ionic Cu, a relatively toxic metal, are moderated to the minimum levels sufficient for physiologic needs, reported in the range of \( 10^{-11} \) to \( 10^{-19} \) mol estimated in plasma, by binding to ceruloplasmin and metallothionein [35, 36]. The dynamic equilibrium between ceruloplasmin and metallothionein in stored in liver tissue prevents toxic accumulation of free copper ion in mammals [37]. That hepatic reservoir is released slowly back into the blood as ceruloplasmin and copper metallothionein in response to inflammation due to injury or arthritis [38]. The average level of bound copper in human control plasma is 1.12 \( \mu g/ml \) [39]; in arthritic patients it is 1.40 \( \mu g/ml \) [40]. A therapeutically effective copper level in inflammation cannot be assessed in definite terms, however, because copper status induced in response to inflammatory diseases in humans varies depending on target organ [13].

In contrast to injection, in topical application the skin forms a natural barrier which controls drug release over an extended period, under approximately steady conditions over a period of days. Flux, and not elimination from the skin surface seems to be the limiting step, and repeated dosage may result only in a build-up on the surface, possibly without significantly affecting the kinetics.

Both historical and more recent investigative data illustrate the safe exposure to exogenous copper as metal, as its salts, or in complex form. Records on human exposure to copper and its benefits date back to prehistoric times [41, 42]. As an example, in current medical practice copper histidinate is administered intravenously to children suffering from Menke’s disease (a.k.a. kinky hair syndrome), a genetically transmitted inability to absorb the metalloelement. In the following, three examples for the efficacy of topical copper compounds are provided:

1. Based on the observation that topical applications of copper salicylate produced anti-inflammatory effects in humans volunteers, in certain countries (e.g., Australia), ethanolic Cu(II) salicylate-containing preparations (Alcusal) are available over the counter for

\(^1\) RCC, 2002. Determination of the partition coefficient (n-octanol/water) of copper tripeptide-1 (PC 1044). RCC Ltd, Environmental Chemistry & Pharmaceutical Division; CH-4452 Ittingen/Switzerland.
human use to alleviate arthritic pain [43], an example of synergism between Cu (II) and an AI drug.

2. A chlorophyllin copper complex (CCC), derived from chlorophyll by replacing the chelated magnesium with copper, has anti-inflammatory and antimicrobial properties, as well as a marked stimulating effect on epithelial cell growth rates and cell regeneration. First established in tissue culture studies, these findings were confirmed clinically through wound healing and deodorizing characteristics observed in animals and humans [44]. Administered orally, CCC is classified as a safe and effective internal deodorant by the US FDA [45].

3. In order to document the tolerability of metallic copper which must be oxidized in contact with the skin in order to penetrate, Gorter examined absorption and tolerance of metallic copper-containing ointments on human volunteers over a period of several weeks. Copper analysis in serum showed that levels had increased significantly above control levels, without overt untoward effects, either in the skin or systemically, following application of ointments containing up to 20% of the metal in its elemental form [46]. It is well established that the Kp of organic chemicals is determined by their lipophilicity and size [47]. This is also likely to apply to (transition) metals complexed with ligands subject to changes in pH and other parameters in the skin with depth [48], which determine ionization as well as availability of ligands and lipophilicity. Copper, a transition metal of variable oxidation state, has the ability to form complexes with different geometries and stability.

Similar to most transition metals, the mechanism by which copper complexes permeate skin is little understood. While copper distribution in the tissues analyzed resulted in remarkably high stratum corneum retention, through that barrier it also reached the highest values in the receptor fluid. Under present study conditions, significant passive transport of the copper species across separate skin strata of differing barrier properties seems to indicate the various potential routes of passage from the surface of the skin into the subepidermal or dermal tissue: the appendageal, the inter- and the intra-cellular pathways. Further, an additional factor appeared to determine the results obtained: intrinsic complex rearrangement in the various skin compartments leading to retention and diffusion of copper ion as a function of ligand. An undetermined part of copper might transit through the skin as the undissociated original tripeptide, and another might diffuse bound to endogenous amino acids of higher complex stability constants such as the histidinate [21], the predominant copper complex in plasma, with cystinate as the secondary ligand.

Studies with labeled ligands (i.e., radio-labeled) may help with further understanding the complex skin penetration behavior of copper complexes. In the present study, only diffusion followed by copper analysis on isolated or separated tissue strata allowed a measure of insight.

Conclusions

Quantitative assessment of skin penetration and specifying the actual penetration mechanism for a copper peptide is difficult due to possible ligand exchanges during diffusion. The results described here, however, show that a topical organic copper complex such as GHK-Cu may be an alternative to copper delivery by injection, also potentially providing a skin reservoir likely to become available over time. This effect warrants further development towards transdermal delivery of the AI agent, especially when considering GHK, a nature-identical carrier tripeptide [18]. Several laboratories in the past had demonstrated local and systemic AI activity of copper compounds by oral, ip, iv, and particularly subcutaneous administration. Having shown permeation of the skin barrier by GHK-Cu in a therapeutically effective dose here, application of the principal to transdermal dosing now may become a realistic alternative to current, traditional routes and agents, however. Taking our results a step further, controlled release of copper for AI therapy using patch technology could be optimized with well-designed formulations. The fact that a deficit in that essential element may be significantly corrected by choosing appropriate cupriphores is a significant outcome of our endeavours towards alleviating rheumatoid arthritis and similar musculoskeletal conditions often afflicting the general population.

Acknowledgments We express our appreciation for the guidance received from the late Roberto Milanino, MD, of the University of Verona, who brought insight into the biological relevance of our research and to whose memory we dedicate this manuscript. We thank Len Patt of the ProCyte Corporation (Redmond, WA) for making a complimentary sample of the copper tripeptide available for our investigation. We thank Larry Ryan, DuPont Analytical Solutions (Wilmington, DE) for supervising inductively coupled plasma mass spectroscopy analyses, and Jeannette Woodward for their execution.

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