It is currently thought that the dementia of Alzheimer’s disease is due to the neurotoxicity of the deposits or aggregates of amyloid-β (Aβ) in the extracellular space of the cerebral cortex. This model has been widely criticized because there is a poor correlation between deposits and dementia. Others have questioned whether Aβ is truly neurotoxic. Yet, in spite of these concerns, the search for therapeutic agents has been based on the development of mouse models transfected with mutant genes associated in humans with early onset Alzheimer’s disease. A major limitation of these models is that although they exhibit many of the pathological and clinical manifestation of the human disease, the bulk of individuals who develop the dementia of Alzheimer’s disease have none of these mutant genes. Furthermore, nine clinical trials of drugs that were effective in transgenic mice failed to show any benefit in patients. Finally, a major unresolved issue with the Aβ model is that since Aβ is produced in everyone, why are deposits only seen in the elderly? This issue must be resolved if we are to understand the etiology of the disease and develop test systems for both diagnosis and drug discovery. Published studies from my laboratory demonstrate that in human cerebrospinal fluid immunoreactive Aβ is only present as a complex with two chaperones, ERP57 and calreticulin and is N-glycosylated. This complex formation is catalyzed by the posttranslational protein processing system of the endoplasmic reticulum (ER). Others have reported that in plaque Aβ is present only as the naked peptide. Together these results suggest that both plaque and dementia are secondary to an age related decline in the capacity of the ER to catalyze protein, posttranslational processing. Since the synaptic membrane proteins necessary for a functioning memory are also processed in the ER, these findings would suggest that the loss of cognition is due to a decline in the capacity of the neuron to produce and maintain functioning synapses. Work from my laboratory and from others further indicate that the components of the ER, posttranslational, protein processing pathway do dramatically decline with age. These data suggest that this decline may be found in all cells and could account not only for the dementia of Alzheimer’s disease, but also for many of the other manifestations of the aging process. These observations also suggest that declining ER function has a role in two well-recognized phenomena associated with aging: a loss of mitochondrial function and a decrease in myelin. Finally, based on this paradigm I propose new cellular and animal models for high-throughput screening for drug discovery.

Keywords: amyloid-β, protein processing, chaperones, N-glycosylation, dementia
late onset disease are found in the apoE and clusterin genes. Furthermore, another difference between the late onset and familial forms is that in human studies Pottier et al. (2012) reported that the mRNA for APP was not elevated in late onset Alzheimer's disease, but was significantly higher in individuals with the various forms of the familial disease and Down's syndrome. Although western blotting of APP would be more definitive, when increases are seen in mRNAs they are commonly accepted as a measure of the total protein content. Hence, these data might suggest that one possible underlying biochemical defect seen in the familial forms of the disease may result in part from elevated levels of APP. Studies on the role of APP in cellular metabolism may give a clue as to how such elevations might lead to the early onset disease.

Knockout studies of APP have reported that this growth factor is necessary for normal neuronal development (Dawson et al., 1999). Furthermore, studies by Nikolaev et al. (2009) have suggested that APP may play an important role at one particular stage in normal embryonic development. As the brain develops the embryo accumulates an excess of neurons and axons. These investigators reported that APP activates death receptor 6 which leads to axon pruning and apoptosis of the excess neurons. These data might suggest that the loss of cognition seen early in the familial forms of the disease before there is overt neuropathology could result from excessive axon pruning due to the high levels of APP; while later in the progression of the disease the high levels of APP could lead to increased neuronal apoptosis with the development of overt neurohistopathology.

Of greater concern there have been nine phase III clinical trials of agents that were effective in transgenic mice but failed to show any efficacy in patients with late onset disease (Table 1).

The probability of nine failures in nine trials for a presumed association is $2^9$ or 1 in 512 which gives a $p = 0.0195$.

Many have ascribed these failures to the initiation of therapy only after the subjects have exhibited significant cognitive decline. This was not true in two of these trials: the GEM (Ginkgo Evaluation of Memory) trial of G. biloba (DeKosky et al., 2008) and the Women's Health Initiative Trial of estrogen alone (Espeland et al., 2004). In the GEM trial of G. biloba the subjects were followed for 7 years (DeKosky et al., 2008). The investigators observed no difference in the incidence of dementia between the placebo and the Ginkgo treated subjects even though this agent was effective in transgenic mice (Figure 2; Stackman et al., 2003).

Similarly, in the estrogen only arm of the Women's Health Initiative Trial the subjects were also followed for 7 years (Figure 3; Espeland et al., 2004). After an initial improvement in cognitive scores which was probably due to a training effect, there were similar declines in the scores of the placebo and estrogen groups.

Furthermore, permanent damage to neurons, such as apoptosis and tangle deposits, is thought to only occur late in the disease. Hence, the late administration of effective agents should at least slow the cognitive decline, if not reverse the disease process. In fact in many of the transgenic mouse studies, elderly animals showed cognitive improvement when given some of these agents (Imbimbo et al., 2012).

In all of these trials only solanezumab gave even the faintest hope of slowing this decline. But even this agent failed to meet the primary end points, and the noted benefit was only seen after combining the results of subjects with early disease in the two trials of the drug, suggesting that the effect may not have been sufficiently robust to be of clinical significance (Doody, 2012). The Food and Drug Administration (FDA) has refused to accept these findings as part of a future New Drug Application (NDA).

In spite of these failures the general thrust of drug development continues to seek new agents which will decrease the production of $\alpha$- or enhance its clearance. These agents include new antibodies, vaccines, and $\gamma$-secretase inhibitors. Also a number of firms have developed inhibitors of the $\beta$-secretase (BACE1). This enzyme cleaves APP at the site which yields the $\alpha$-peptide and the $\beta$-APP (Figure 1). This would seem to be a somewhat hazardous approach for the long term treatment of patients with cognitive impairment, since knocking out BACE1 has been shown to lead to

Table 1 | Failed phase III trials of agents which were effective in transgenic mouse models.

| Drug                          | Mechanism                  | Reference                  | Animal reference |
|-------------------------------|----------------------------|----------------------------|------------------|
| Abeta vaccine                 | Clear abeta                | Holmes et al. (2008)       | Janus et al. (2008) and Morgan et al. (2008) |
| Tarenflurbil                  | $\gamma$-Secretase inhibitor | Green et al. (2009)        | Kukar et al. (2007) and Abramowski et al. (2008) |
| Semagacestat                  | $\gamma$-Secretase inhibitor | Schor (2011)               | Vellas et al. (2012) |
| MK-677                        | IGF-1 secretagogue         | Saviny et al. (2008)       | Carro et al. (2002) |
| Ginkgo biloba                 | Antioxidant                | DeKosky et al. (2008)      | Stackman et al. (2003) |
| Estrogen                      | Hormone replacement        | Espeland et al. (2004)     | Levin-Alhendy et al. (2002) and Carroll et al. (2007) |
| Docosahexaenoic acid          | Omega-3 fatty acid         | Quinn et al. (2010)        | Calon et al. (2004) |
| Bapineuzumab                  | Monoclonal antibody        | Scheitans et al. (2012)    | Imbimbo et al. (2012) |
| Solanezumab                   | Monoclonal antibody        | Doody (2012)               | Imbimbo et al. (2012) |
significant deficits in brain function (Laird et al., 2005; Hu et al., 2010; Rajapaksha et al., 2011). The most serious of these problems is that these animals have been reported to develop a seizure disorder associated with an increased density of surface Nax 1.2 channels and increased intrinsic firing of isolated neurons and in hippocampal brain slices (Hu et al., 2010). In a short term phase I trial in normal volunteers the Merck BACE1 inhibitor, showed no adverse effects. But considering that these adverse effects may require long term administration, since it may dependent upon inducing the Nax 1.2 channels, it may take months to observe this toxicity. Furthermore decreasing the level of Aβ in the brain may require higher doses than those used in the phase I trial. Hence, at higher doses for longer treatment periods there could be an increased risk for observing the toxic side effects.

The only agent which has actually shown any promise of a viable treatment for Alzheimer’s disease is methylene blue (Wischik, 2012). This drug has been used as a urinary tract disinfectant and analgesic since the 1930s. Basic studies have demonstrated that its administration can lead to the loss of tangles, another pathological manifestation of Alzheimer’s disease. Tangles result from cellular stress which leads to hyperphosphorylation of a neuronal, cytoskeletal protein, tau (P-tau). The P-tau then aggregates to form tangles. In a small phase II trial patients receiving 138 mg per day of the drug showed a markedly reduced cognitive decline during a 1 year study period. Yet, those receiving twice the dose showed a parallel decline to the placebo. A chemical modification of this agent is currently going into a phase III clinical trial. It will be interesting to see whether they can replicate their initial findings.

**A NEW PARADIGM FOR THE ETIOLOGY OF THE DEMENTIA OF ALZHEIMER’S DISEASE**

In light of these failures it would seem reasonable to examine new paradigms for the etiology of this dreaded condition. A number of years ago we asked the question: Since Aβ is produced in everyone why is plaque only seen in the brains of the elderly? We postulated that normally Aβ is bound to proteins which keep it in solution and thereby prevent plaque formation. And indeed this turned out to be the case! When we ran western blots of fresh or fresh frozen cerebrospinal fluid (CSF) on denaturing gels, we observed that there was only a single Aβ-immunoreactive band which was consistently found at around 62 kDa (Figure 4; Erickson et al., 2005). And was associated with an endoplasmic reticulum (ER) chaperone, ERp57.

Furthermore, on immunopurification we were able to isolate the complex with antibodies to either Aβ or ERp57 (Figure 5). A large number of studies on the biochemistry of the ER, posttranslational, protein processing pathway have reported that ERp57 binds to nascent proteins at N-glycosylation sites in association with another ER chaperone, calreticulin. In line with these observations, when the CSF was run on a native gel, the band moved to 118 kDa and was associated with both ERp57 and calreticulin (Figure 6).

The fact that the Aβ and the two chaperones all shifted to this higher molecular weight along with the immunopurification data strongly supports our claim that the Aβ band is not either of the two sAPP’s which have similar molecular weights.

Finally, since the Aβ found in plaque is only present as the naked peptide, one possibility is that the complex was originally in the plaque but broke down when it was solubilized by the usual treatment with either concentrated formic or trifluoroacetic acid. This is not likely to be the case since treatment of CSF under these same conditions had no effect on the complex (channels 3 and 4) (Figure 7).

Chaotropic agents such as urea and guanidine isothiocyanate also had no effect on the complex (channel 5 and 6). In fact the only procedure which did affect the complex was storage overnight at 4 °C in a pH 9.0 buffer (channel 9). The other interesting aspect of this study was that the complex bound to a bonate column, a synthetic lectin, suggesting that the Aβ is glycosylated (channel 8).
FIGURE 2 | The effect of Ginkgo biloba on Cognitive Function in the GEM Trial (taken from DeKosky et al., 2008).

The binding of ERp57 to APP is not unique. Two plasma membrane receptors, the vasopressin receptor (Aiyar et al., 1989) and the 1,25 dihydroxy vitamin D plasma membrane receptor (Nemere et al., 2004; Nemere, 2005), have also been shown to have bound ERp57. The latter receptor is present in the plasma membrane of enterocytes and modulates Ca\(^{++}\) uptake from the intestines. The ERp57 would appear to have a role in signal transduction since knocking it down or inhibiting it with antibody to its terminal carboxy end inhibited 1,25 dihydroxy vitamin D stimulated Ca\(^{++}\) uptake (Nemere et al., 2004; Nemere, 2005). It should also be noted that the title of the publication by Aiyar et al. (1989) demonstrating the binding of ERp57 to the vasopressin receptor is incorrect. When these workers originally cloned and sequenced ERp57, they erroneously identified it as a phosphatidylinositol specific phospholipase C (Bennett et al., 1988). We found that the ERp57 purified from rat liver (Srivastava et al., 1991) and rERp57 generated using the authors’ plasmid had no phospholipase activity (Srivastava et al., 1993) and instead it is a thiol protein disulfide oxidoreductase: a family of enzymes that catalyzes both the oxidation and reduction of protein sulfur groups and is usually referred to as a Protein Disulfide Isomerase (PDI).

**THE EFFECT OF AGE ON THE COMPONENTS OF THE ER, POSTTRANSLATIONAL, PROTEIN PROCESSING PATHWAY**

In light of these findings we hypothesized that the dementia seen in late onset Alzheimer’s disease is due to a decline in the capacity of the neurons to catalyze the ER, posttranslational, protein processing of the synaptic membrane proteins that are necessary for a functioning memory (Erksson et al., 2005). In support of this hypothesis we found that ERp57 does decline with age (Figure 8; Erickson et al., 2006).

Several other ER chaperones showed similar declines (Table 2). Finally, data from other laboratories would suggest that there is also a decline in the N-glycosylation pathway with age. Firstly, there is a reduced content of N-glycosylated proteins in the tissues of the elderly (Kousvelari et al., 1988). Yet, this decline could be due to decreases in gene transcription. On the other hand, a number of studies have suggested that it may also be due to changes in the activity of the N-glycosylation pathway. This concept is based on the effect of age on the content of dolichol in various tissues, a necessary cofactor for N-glycosylation.

The initial step in the N-glycosylation pathway is the synthesis of an oligosaccharide bound to dolichol phosphate (Yan and Lennarz, 1999; Helenius and Aebi, 2002; Kelleher and Gilmore, 2006; Figure 9). Dolichol is a high molecular weight terpene which serves as a cofactor for the cytosolic synthesis of the oligosaccharide. Once the complex is fully synthesized, it is transferred to the lumen of the ER and attached to an \(\xi\)-amino group of asparagine by an oligosaccharide transferase (OST; Kelleher and Gilmore, 2006). The addition of the individual sugars to the carbohydrate complex is catalyzed by a family of specific monosaccharide transferases (MST; Yan and Lennarz, 1999; Helenius and Kelleher et al., 1999).
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FIGURE 3 | The effect of estrogen alone on Cognitive Function in the Women’s Health Initiative (taken from Espeland et al., 2004).

Aebi, 2002; Kelleher and Gilmore, 2006). The first sugars added to the dolichol phosphate are a pair of $N$-acetylglucosamines, followed by nine mannoses and three glucoses. A large number of studies have shown that as animals age there is a 5–10-fold increase in the cellular content of both dolichol and dolichol phosphate (for example see Marino et al., 1998). Since there is no increase in the synthesis of dolichol, this finding is consistent with a blockage of the addition of the first $N$-acetylglucosamine. And indeed it has been reported that the activity of ALG7, the MST that adds the first N-acetylglucosamine to the dolichol phosphate, does decline with age (Mota et al., 1994).

OUR STUDIES INDICATE THAT
1. The $A\beta$-chaperone complex is formed during the normal processing of APP in the lumen of the ER.
2. If this complex is not produced, then the naked peptide is secreted and precipitates on its release from the cell to form plaque.
3. On western blotting of a native gel we found that the complex contains two ER chaperones, ERP57 and calreticulin.
4. In the CSF $A\beta$ is probably also N-glycosylated.
5. These data indicate that plaque formation is due to a decline in the capacity of the ER to catalyze the posttranslational processing of APP. It may also account for many other manifestations of aging.

6. Forty percentage of the proteins synthesized in the cell undergo this same posttranslational processing in the ER, including all the synaptic membrane proteins that are necessary for a functioning memory.
7. With age there is a decrease in the components of the ER, protein, post-translational processing pathway.

THE ROLE OF DECREASED ER, PROTEIN, POSTTRANSLATIONAL PROCESSING IN THE DECLINE WITH AGE IN MITOCHONDRIAL FUNCTION AND MYELIN CONTENT
There are numerous phenomena associated with the aging process. Two of the most fundamental are the decline with age in mitochondrial function and myelin content. One of the major problems with the $A\beta$ model is that it does not address what the relationship of either of these two well-documented processes has to do with plaque deposition. I feel that both of these processes may be related to the decline in ER function.

THE ROLE OF THE LOSS OF ER FUNCTION IN THE DECLINE IN MITOCHONDRIAL FUNCTION
It is well-known that as mitochondria age they acquire structural defects (Batlevi and La Spada, 2011). The damaged mitochondria are cleared from the cell by autophagy. Many of the proteins which make up the autophagic vacuoles are processed in the ER and golgi
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FIGURE 4 | Western blots of human, CSF with antibodies to ERp57 and Aβ. (Militello and Colombo, 2011). Hence, with a decline in ER function there is also a decline in autophagy. Such declines have been associated with the onset of some age related neurodegenerative diseases (Pan et al., 2008).

Second, the mitochondria are directly bound to the ER through the mitochondrial associated membrane (MAM; Merkwirth and Langer, 2008; Hayashi et al., 2009; Schon and Przedborski, 2011). This structure serves three, major functions. The first is to transport critical phospholipids from the ER, where they are synthesized, into the mitochondria where they serve vital roles in the maintenance of the structure and function of the mitochondria. Further, it regulates Ca\(^{2+}\) homeostasis in the mitochondria and thereby oxidative phosphorylation.

A final role for the ER in maintaining mitochondrial structure and function is the control of mitochondrial fission and fusion. As defective mitochondria are cleared by autophagy, they are replaced through a process involving both mitochondrial fusion and fission. Recent studies have indicated that several proteins in the MAM play major roles in both of these processes (Merkwirth and Langer, 2008; Friedman et al., 2011). In particular two critical proteins which tether the ER to the mitochondrial outer membrane are mitofusin 1 and 2 (mfn1 and 2; Merkwirth and Langer, 2008; Figure 10). These transmembrane proteins are located in both the ER membrane and the outer membrane of the mitochondria. Both mfn1 and 2 undergo posttranslational processing in the ER. Hence, a decline in the capacity of the ER to catalyze their posttranslational processing would be expected to have a profound effect on mitochondrial structure and function. Such declines in ER function could compromise the ability of the cell to maintain metabolically active mitochondria.

THE ROLE OF THE LOSS OF ER FUNCTION IN THE DECREASE IN MYELIN WITH AGE

Finally, the myelin sheaths are the plasma membranes of oligodendrocytes and Schwann cells (Figure 11). This structure is also

FIGURE 5 | Immunoopurification of the Aβ-ERp57 Complex from Human CSF. Aβ and ERp57 were isolated by immunoprecipitation. The samples were then purified by western blotting on sodium dodecyl sulfate polyacrylamide gel electrophoresis. The bands were identified by antibodies to ERp57 (A) and Aβ (B). (Taken from Erickson et al., 2005)

FIGURE 6 | Western blot of the Aβ–ERp57–calreticulin complex from human CSF after separation on a native gel (taken from Erickson et al., 2005).
processed in the ER. The observed decline seen with age in the capacity of the ER to catalyze this processing could explain the loss of white matter consistently seen in the elderly and the slowing of thought processes as we age.

**PROPOSED CELLULAR AND ANIMAL MODELS FOR DRUG DISCOVERY**

These findings would suggest that we should be seeking agents for the treatment of Alzheimer’s disease that enhance the components of the ER, posttranslational, protein processing pathway. Classically the usual procedure for such a search has been to construct test systems in which the promoter region for the target protein is linked to a reporter system, such as luciferase, a beta galactosidase or a fluorescent protein. Yet, we now know that non-coding regions of the DNA and histone modifications play major roles in controlling both transcription and translation of proteins. Hence, potentially the most fruitful approach for high-throughput drug screening would be to develop cell models in which compact fluorescent proteins, such as green fluorescent protein (GFP), are embedded in an exon of the otherwise intact target gene. This should maximize the number of potential target sites for small molecule, drug discovery.

Such transgenic models are created in embryonic or induced pluripotent stem cells by homologous recombination using a variety of site specific endonuclease systems. These models are usually created with either mouse or human embryonic or induced

| Chaperone | Peak concentration /μg/mg protein | Concentration @ 874 days /μg/mg protein | Constitutive decline % | Show cyclic variation | Cyclic variation % decline |
|-----------|-----------------------------------|----------------------------------------|------------------------|----------------------|--------------------------|
| BiP       | 88.0                              | 48.5                                   | 39                     | Yes                  | 50                       |
| Calnexin  | 57.4                              | 40.5                                   | 29                     | No                   | –                        |
| Calreticun| 76.4                              | 4.8                                    | 8                      | No                   | –                        |
| ERP55     | 34.8                              | 13.1                                   | 51                     | Yes                  | 73                       |
| ERP57     | 15.4                              | 8.2                                    | 32                     | Yes                  | 71                       |
| ERP72     | 141                               | 100                                    | 30                     | No                   | –                        |
| Total     | 336.2                             | 215.1                                  | 37                     |                      |                          |

**Table 2 | Effect of age on the content of rat liver ER chaperones (taken from Erickson et al., 2006).**

![FIGURE 7](image1.png)

**FIGURE 7 | Immunoblot of Aβ in CSF after treatments to dissociate the Aβ-ERp57 complex. Channel 1 – molecular weight markers; channel 2 – untreated CSF; channel 3 – CSF treated with 70% formic acid; channel 4 – CSF treated with 80% trifluoroacetic acid; channel 5 – CSF treated with 8 M guanidine isothiocyanate; channel 6 – CSF treated with 8 M urea; channel 7 – CSF proteins which did not bind to a boronate column; channel 9 – treatment of the complex with glycine buffer, pH 9.0 (taken from Erickson et al., 2006).**

![FIGURE 8](image2.png)

**FIGURE 8 | The effect of age on the ERp57 content of rat, hepatic microsomes (taken from Erickson et al., 2006).**
pluripotent stem cells. In the case of the mouse lines it would also be possible to clone the cells and create transgenic, adult animals. This is an important step since it is necessary to demonstrate that the insertion of the fluorescent protein does not lead to an animal with compromised metabolism. Usually the indicator system is inserted in either the first or last exon of the native protein. If these turn out to produce metabolically compromised cells, then it would be necessary to insert the indicator in an exon that does not compromise metabolic function. The stem cells can then be transformed into a variety of tissue types by incubating with the appropriate growth factors. With the construction of the labeled cell lines, they can be used to automatically screen large libraries of compounds in microtiter plate readers.

Similarly intact animal models could be developed in which the relevant proteins have been knocked down. It is necessary to only knock them down because knockouts of the chaperones and components of the N-glycosylation pathway are lethal mutants. The possible approaches could include a lox-cre system in which the specific tissue cell type, such as neuronal cells have a haploid knock out of the critical gene. Alternatively, it is possible to develop models with a global knock down of the target gene(s) with synthetic, antisense oligonucleotides.

In an unrelated study of the toxicity of methoxychlor, we observed that feeding this insecticide to rats for 3 weeks specifically increased the levels of ERp57 in hepatic microsomes (ER) (Figure 12).

**CONCLUSION**

1. Nine phase III clinical trials of agents that were effective in transgenic mice were not effective in patients with late onset dementia.
2. The reason that plaque does not form in the young is because Aβ is normally N-glycosylated and bound to two ER chaperones, ERP57 and calreticulin, which keep it in solution. 3. These components decline with age.

4. These findings suggest that the dementia seen in late onset Alzheimer’s disease is due to a decline in the ER, posttranslational processing of the synaptic, membrane proteins that are necessary for a functioning memory. 5. It also suggests that the loss of mitochondrial function and white matter in the elderly is due to this decline in ER function. 6. This paradigm suggests new cellular models for drug discovery in which fluorescent proteins are inserted into an exon of the various components of the ER, posttranslational, protein processing pathway. 7. Animal models could be constructed in which the critical proteins are knocked down by standard procedures and then candidate drugs are administered to test whether they improve cognition.

IMPLICATIONS FOR FUTURE DRUG DISCOVERY TO TREAT ALZHEIMER’S DISEASE AND OTHER DISEASES ASSOCIATED WITH AGING

Finally, over the past two decades the pharmaceutical industry and funding agencies and foundations have made and are continuing to make massive investments in the search for disease modifying treatments for Alzheimer’s disease and other diseases of the elderly. Although these efforts have taken a variety of approaches, the bulk have sought treatments based on the Aβ hypothesis that seek to improve cognitive function by decreasing the brain content of Aβ. Considering the number of these agents currently in the pipeline, it is clear that this approach will continue to dominate drug development for the foreseeable future. Yet, since many of these new agents are merely variations on earlier failed investigational drugs, there is a significant probability that they too will not prove to be efficacious in phase III clinical trials.

In view of the long, lag period between the initial identification of new targets and the final regulatory approval of agents developed on the basis of new paradigms, it would seem prudent at this point to consider other models for drug discovery in case the current batch of investigational drugs fail to fulfill their early promise. At the present time there is little chance that this lag period can be shortened by evaluating therapeutic efficacy based on surrogate measures, because changes in the levels of such measures have not been demonstrated to correlate with clinical efficacy. For example, the two most widely accepted biomarkers in the CSF of disease status are low levels of Aβ and high levels of P-tau. Yet in the recent failed bapineuzumab trial, P-tau levels did decline, but the patients still showed no clinical benefit (Blennow et al., 2012).

Many have cited the experience in the treatment of hypertension in which demonstrating that decreasing blood pressure was sufficient grounds for drug approval. But this surrogate measure was based on clinical trial data that had shown that a decrease in blood pressure led to a pronounced decrease in morbidity events (Veterans Administration Cooperative Study Group on Antihypertensive Agents, 1967, 1970). In fact the decrease was so pronounced that in the present era of drug trials, a Data and Safety Monitoring Board would certainly have ended the 1970 trial no later than 3 years instead of continuing it for the full 5 years laid out in the original protocol. Until such robust surrogate measures have been identified or some agent does appear to show efficacy in
REFERENCES

Abramowski, D., Vassiljev, K. H., Fenton, U., Jansen, A. L., Neumann, Schneidereit, A., Ramirez, J. M., et al. (2008). Dynamics of alpha-tau deposition and deposition in different beta-amyloid precursor protein transgenic mouse models following gamma-secretase inhibition. J. Pharmacol. Exp. Ther. 327, 411–424.

Amit, N., Bennett, F. N., Valenti, P., Angeli, M., Munch, M., et al. (1998). Solubilization of rat liver vacuolar proteins receptors as a complex with a guano-nucleotide-binding protein and phospholipid-specific phospholipase C. Biochem. J. 326, 65–78.

Bennett, C. F., Balsamo, J. M., Varricchio, A., and Crock, S. T. (1988). Molecular cloning and complete amino acid sequence of form 1 phospholipid-specific phospholipase C. Nature 334, 268–274.

Batist, V. and La Spada, A. R. (2011). Mitochondrial amyotrophy in neural function, neurodegenerative disease, neuron cell death, and aging. Neurobiol. Dis. 43, 46–61.

Blinn, R., Zetterberg, H., Rinne, J. O., Salloway, S., Wei, J., Black, B., et al. (2008). Effect of immunotherapy for Alzheimer’s disease: results of phase I. J. Neuro Immunol. 176, 93–102.

Cavallini, G., Masini, M., Gori, Z., Cavalli, R., and Vezoli, G. (2011). ER tubules mark sites for mitochondrial division. Cell Death Differ. 18, 81–88.

Casadesús, A., Vassiljev, K. H., Neumann, Schneidereit, A., and Ramirez, J. M. (2008). Golgi Athers for secretion of homing cells to a randomised controlled trial. JAMA 300, 2253–2262.

De Strooper, B., Vanpraet, R., and Gold, B. (2010). The secretases: enzymes with therapeutic potential in Alzheimer’s disease. Nat. Rev. Neurosci. 11, 96–107.

Dawson, B. R. (2012). Safety and efficacy of solanesol in patients with mild to moderate Alzheimer’s disease. Acta Neurol. Scand. 126, 10–30.

Dawson, B. R., Drum, L. M., Olson, D. A., Cohen, D. A., Davis, A. T., Wood, W. G., et al. (2005). In cerebrospinal fluid ER phospholipase C epsilon 5 and calreticulin bind B-amyloid. Biochem. Biophys. Res. Comm. 332, 50–57.

Erickson, R. R., Drum, L. M., and Holtzman, J. L. (2006). The effect of aging on the chaperone concentrations in the hepatic, endoplasmic reticulum of male rats: the possible role of protein misfolding due to the loss of chaperones in the decline in physiological function seen with age. J. Gerontol. A Biol. Sci. Med. Sci. 61, 435–441.

Ernster, M., Rapp, S. B., Shumatink, S. A., Brumans, R., Mannon, J., Sherwood, B. B., et al. (2004). Congenital deficiency in a beta-lipoprotein-conjugated chaperone function in porcine mutant newborns. Proc. Natl. Acad. Sci. U.S.A. 101, 12502–12507.

Gross, K. C., Schmaier, S. L., Amass, D. A., Boulin, A. F., Willcock, G., Swab, E. A., et al. (2009). Effect of curcumin on cognitive decline and activities of daily living in patients with mild Alzheimer’s disease: a randomised controlled trial. JAMA 302, 2577–2584.

Hayashi, T., Rizzuto, R., Hajnoczky, G., Helenius, J., and Aebi, M. (2002). Trans-Golgi network and phospholipid-specific phospholipase C. Curr. Mol. Med. 2, 561–566.

Holtzman, J. L. (2000). The effect of vaccination prevents memory loss in an animal model of Alzheimer’s disease. Nature 408, 982–985.

Holtzman, J. L., Fuchs, J. A., and Holmson, J. L. (2000). The effect of methylene blue administration to rats on hepatic, microsomal cytochrome P450 and phospholipid uptake in intestinal cells. Proc. Natl. Acad. Sci. U.S.A. 97, 7382–7387.

Ikeda, A., MacLaughlin, T., O’Doherty, D. M. M., and Tousse-Laugnou, M. (2009). ARF binds to N-cadherin from neurons in an in situ and normal human and Alzheimer’s disease. Nature 497, 981–984.

Jensen, P. L., Franssena, D. A., and Vorbrodt, H. (2008). The role of autophagy in the treatment of mild-to-moderate Alzheimer’s disease. Expert Rev. Clin. Pharmacol. 1, 13–20.

Koster, J. E., Burreo, D. K., Murphy, L., and Baum, J. B. (1988). N-linked protein glycosylation in the rat peripheral nervous system. J. Neurocytol. 17, 475–482.

Kozma, S. R., Vavrin, T., and Su, T.-P. (2009). MAM: more than just a housekeeper. Mol. Mem. Biol. 26, 545–553.

LeRoth, D., and Torres-Aleman, L. (2011). ER tubules mark sites for mitochondrial division. Cell Death Differ. 18, 81–88.

Morrell, S. L., Fuchs, J. A., and Holmson, J. L. (2000). The effect of methylene blue administration to rats on hepatic, microsomal cytochrome P450 and phospholipid uptake in intestinal cells. Proc. Natl. Acad. Sci. U.S.A. 97, 7382–7387.

Nemer, J. (2005). The 1,25(OH)2D3-MARKS protein: contribution to survival stimulated calcium uptake in cholie and rat. J. Neurobiol. 60, 453–457.

Nemer, J., Fratila-Carson, M. C., Beluc, S., Sherrf, T. M., Norman, A. W., Boyan, B. D., et al. (2004). Bystander knockdown function to links a 1,25(OH)2D3 membrane binding protein (1,25D3-MARKS) and phospholipid uptake in intestinal cells. Proc. Natl. Acad. Sci. U.S.A. 101, 17902–17907.

Parrish, L., Keen, L., and Jankowski, J. (2008). The role of autophagosome lysosome pathway in neurodegeneration associated with Parkinson’s disease. Br. J. Pharmacol. 154, 521–531.

Pettit, C., Wolloen, D., Leonor, A. R., Maitere, D., Besomme, X., Juxhi, S., et al. (2012). Amyloid-beta protein precursor gene expression in animal models of Alzheimer’s Disease and other conditions. J. Alzheimer’s Dis. 26, 563–566.

Quinn, J. F., Ramus, R., Thomas, B. G., Yorke-Mauro, R., Nelson, E. B., Van Deck, C., et al. (2012). Drosophila heannin acid supplementation and
Holtzman Drug discovery models in AD cognitive decline in Alzheimer’s disease. JAMA 341, 1903–1911.
Rajapaksha, T., Eimer, W. A., Beza, T. C., and Vassar, R. (2011). The Alzheimer’s β-secretase enzyme BACE1 is required for accurate axon guidance of olfactory sensory neurons and normal glomerular formation in the olfactory bulb. Mol. Neurodegener. 6, 88.
Schulte, P., Speeling, R., Saliseower, S., and Fos, N. (2012). Reptinotum IV phase 3 results. J. Nutr. Health Aging (abstr.) 16, 797.
Schon, E. A., and Przedborski, S. (2011). Mitochondria: the next (neurode) generation. Neuron 70, 1033–1053.
Schor, N. F. (2011). What the halted phase III γ-secretase inhibitor trial may (or may not) be telling us. Ann. Neurol. 69, 237–239.
Sevigny, J. J., Ryan, J. M., van Dyek, C. H., Peng, Y., Lines, C. R., and Nessly, M. I. (2008). Growth hormone secretagogue MK-677; No clinical effect on AD progression in a randomized trial. Neurology 71, 1702–1708.
Srivastava, S. P., Chen, N.-Q., Liu, Y.-X., and Holtzman, J. L. (1991). Purification and characterization of a new isozyme of thiol protein disulfide oxidoreductase from rat hepatic microsomes: relationship of this isozyme to cytosolic, phospholipid dependent specific phospholipase C from rat kidney. J. Biol. Chem. 266, 20337–20344.
Srivastava, S. P., Fuchs, J. A., and Holtzman, J. L. (1993). The reported cDNA sequence for phospholipase Ca1 encodes protein disulfide isomerase, isozyme Q-2 and not phospholipase C. Biochem. Biophys. Res. Commun. 193, 971–979.
Stackman, R. W., Eckenstein, F., Frei, B., Kulhanek, D., Nowlin, J., and Quinn, J. F. (2003). Prevention of age-related spatial memory deficits in a transgenic mouse model of Alzheimer’s disease by chronic Ginkgo biloba treatment. Exp. Neurol. 184, 510–520.
Vellas, B., Coley, N., Ousset, P.-J., Berrut, G., Dartigues, J.-F., Dubois, B., et al. (2012). Long-term use of standardized Ginkgo biloba extract for the prevention of Alzheimer’s disease (GinkPage): a randomised placebo-controlled trial. Lancet Neurol. 11, 851–859.
Veterans Administration Cooperative Study Group on Antihypertensive Agents. (1967). Effects of treatment on morbidity in hypertension: results in patients with diastolic blood pressures averaging 115 through 129 mm Hg. JAMA 202, 116–122.
Veterans Administration Cooperative Study Group on Antihypertensive Agents. (1970). Effects of treatment on morbidity in hypertension II. Results in patients with diastolic blood pressures averaging 90 through 114 mm Hg. JAMA 213, 1145–1152.
Wischik, C. M. (2012). Limitations of MTC (methylene blue) and replacement LMTX™ in TauRx phase 3 program. J. Nutr. Health Aging (abstr.) 16, 842.
Yan, Q., and Lennarz, W. J. (1999). Oligosaccharyltransferase: a complex multisubunit enzyme of the endoplasmic reticulum. Biochem. Biophys. Res. Commun. 266, 684–689.
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