Persistence and clearance of viral RNA in 2019 novel coronavirus disease rehabilitation patients

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Abstract

Background: A patient’s infectivity is determined by the presence of the virus in different body fluids, secretions, and excreta. The persistence and clearance of viral RNA from different specimens of patients with 2019 novel coronavirus disease (COVID-19) remain unclear. This study analyzed the clearance time and factors influencing 2019 novel coronavirus (2019-nCoV) RNA in different samples from patients with COVID-19, providing further evidence to improve the management of patients during convalescence.

Methods: The clinical data and laboratory test results of convalescent patients with COVID-19 who were admitted to from January 20, 2020 to February 10, 2020 were collected retrospectively. The reverse transcription polymerase chain reaction (RT-PCR) results for patients’ oropharyngeal swab, stool, urine, and serum samples were collected and analyzed. Convalescent patients refer to recovered non-febrile patients without respiratory symptoms who had two successive (minimum 24 h sampling interval) negative RT-PCR results for patients’ oropharyngeal swab, stool, urine, and serum samples were collected and analyzed. Convalescent patients refer to recovered non-febrile patients without respiratory symptoms who had two successive (minimum 24 h sampling interval) negative RT-PCR results for viral RNA from oropharyngeal swabs. The effects of cluster of differentiation 4 (CD4)+ T lymphocytes, inflammatory indicators, and glucocorticoid treatment on viral nucleic acid clearance were analyzed.

Results: In the 292 confirmed cases, 66 patients recovered after treatment and were included in our study. In total, 28 (42.4%) women and 38 men (57.6%) with a median age of 44.0 (34.0–62.0) years were analyzed. After in-hospital treatment, patients’ inflammatory indicators decreased with improved clinical condition. The median time from the onset of symptoms to first negative RT-PCR results for oropharyngeal swabs in convalescent patients was 9.5 (6.0–11.0) days. By February 10, 2020, 11 convalescent patients (16.7%) still tested positive for viral RNA from stool specimens and the other 55 patients’ stool specimens were negative for 2019-nCoV following a median duration of 11.0 (9.0–16.0) days after symptom onset. Among these 55 patients, 43 had a longer duration until stool specimens were negative for viral RNA than for throat swabs, with a median delay of 2.0 (1.0–4.0) days. Results for only four (6.9%) urine samples were positive for viral nucleic acid out of 58 cases; viral RNA was still present in three patients’ urine specimens after throat swabs were negative. Using a multiple linear regression model ($F = 2.669, P = 0.044$, and adjusted $R^2 = 0.122$), the analysis showed that the CD4+ T lymphocyte count may help predict the duration of viral RNA detection in patients’ stools ($t = 2.699, P = 0.010$). The duration of viral RNA detection from oropharyngeal swabs and fecal samples in the glucocorticoid treatment group was longer than that in the non-glucocorticoid treatment group (15 days vs. 8.0 days, respectively; $t = 2.550, P = 0.013$) and the duration of viral RNA detection in fecal samples in the glucocorticoid treatment group was longer than that in the non-glucocorticoid treatment group (20 days vs. 11 days, respectively; $t = 4.631, P = 0.001$). There was no statistically significant difference in inflammatory indicators between patients with positive fecal viral RNA test results and those with negative results ($P > 0.05$).

Conclusions: In brief, as the clearance of viral RNA in patients’ stools was delayed compared to that in oropharyngeal swabs, it is important to identify viral RNA in feces during convalescence. Because of the delayed clearance of viral RNA in the glucocorticoid treatment group, glucocorticoids are not recommended in the treatment of COVID-19, especially for mild disease. The duration of RNA detection may relate to host cell immunity.

Keywords: COVID-19; 2019-nCoV; Nucleic acid detection; Glucocorticoid
Introduction

In January 2020, a new coronavirus was confirmed as the cause of unexplained pneumonia in a group of patients from Wuhan, Hubei, and was subsequently named the 2019 novel coronavirus (2019-nCoV).[11] Due to increasing numbers of cases reported out of China, the World Health Organization announced on January 30 that the emerging new coronavirus pneumonia epidemic constituted a public health emergency of international concern.[2] Up until February 11, 2020, there were 44,653 confirmed 2019 novel coronavirus disease (COVID-19) cases reported in China and 395 cases in 24 other countries.[3] The transmission capacity of 2019-nCoV was underestimated at first. Initial studies showed its regeneration number, \( R_0 \), was 2.2 to 2.9.[5-7] meaning that each infector could transmit to another 2.2 to 2.9 people. Recently a novel study revealed that the \( R_0 \) of 2019-nCoV is 3.77 based on clinical and epidemiological data from nearly 8866 patients in 30 provinces,[8] which is higher than the \( R_0 \) of severe acute respiratory syndrome coronavirus (SARS-CoV).[9]

The patient’s infectivity is determined by the presence of the virus in different body fluids, secretions, and excreta. All patients with positive viral RNA detection need to be isolated. As mentioned in the “Diagnosis and Treatment Scheme of New Coronavirus Infected Pneumonia” (trial version 5), only after the relief of symptoms and two successive (minimum 24 h sampling interval) negative viral nucleic acid results for respiratory specimens, the isolated cases can be disisolated. However, the persistence and clearance of viral RNA in different specimens of COVID-19 patients remains unclear. In this study, viral RNA detection was performed on throat swabs, and stool, urine, and serum specimens, which were analyzed based on different clinical conditions and lab results, to figure out the clearance time of the virus and factors which may influence this.

Methods

Ethical approval

This retrospective study was approved by the Shanghai Public Health Clinical Center Ethics Committee (No. YJ-2020-S015-01) and was exempted from the need for informed consent from patients.

Subjects

From January 20, 2020 to February 10, 2020, all confirmed patients with COVID-19 in the Shanghai region were admitted to the Shanghai Public Health Clinical Center. The convalescent patients refer to recovered non-febrile patients without respiratory symptoms who had two successive (minimum 24 h sampling interval) negative reverse transcription polymerase chain reaction (RT-PCR) results for viral RNA from oropharyngeal swabs.

Clinical measures

Clinical data and lab results were recorded at admission and last fecal viral RNA test, including sex, age, cluster of differentiation 4 (CD4)+ T lymphocyte counts, erythrocyte sedimentation rate (ESR), C-reactive protein (CRP), and procalcitonin (PCT). The glucocorticoid treatment group included any patients who were ever treated with glucocorticoids, such as prednisolone or dexamethasone. The feces-positive and feces-negative groups were classified based on the detection of viral RNA in patients’ feces. The clinical conditions and lab results of study subjects, together with the viral RNA results from different specimens (oropharyngeal swab, stool, urine, and serum) from each day were collected retrospectively.

Detection of viral RNA in COVID-19

A magnetic bead-method nucleic acid extraction kit was applied in a fully automated nucleic acid extraction instrument (Master Biotechnology, China). The total RNA was extracted from a 200-μL sample and dual fluorescence PCR (Applied Biosystems 251658240 7500 Real-Time PCR Systems, Foster City, CA, USA) was performed according to the manufacturer’s instructions.

Statistical analysis

Statistical analyses were performed using SPSS 20.0 (International Business Machines Corporation, IBM, Armonk, New York, USA). Normally distributed continuous variables are summarized as the mean and standard deviation, and non-normally distributed data are recorded as median and interquartile range (IQR) as appropriate. Categorical variables are expressed as counts and percentages for each category. The \( t \)-tests or Wilcoxon rank-sum tests were applied to test differences between two groups; Fisher exact tests or Chi-square tests were used for categorical variables. Multiple linear regression was applied to determine the relationship between outcomes and the exploratory factor. \( P < 0.05 \) was considered significant.

Results

Demographics and laboratory examination results

From January 20, 2020 to February 10, 2020, 292 patients with COVID-19 were admitted to the Shanghai Public Health Clinical Center. Sixty-six convalescent patients were included in our study. In total, 28 (42.4%) women and 38 men (57.6%) with a median age of 44.0 (34.0–62.0) years were analyzed. The oldest patient was 78 years old and the youngest was 16 years old. There was no difference in sex or age between those with or without glucocorticoid treatment (\( \chi^2 = 0.342, P = 0.599; t = 1.059, P = 0.294 \)). On admission, the average high level of ESR was 70.0 (25.5–90.0) mm/h, high-sensitivity CRP was 8.4 (1.6–20.3) mg/L, and PCT was 0.03 (0.02–0.05) ng/mL, which decreased to 44.0 (29.5–81.3) mm/h, 0.5 (0.5–2.1) mg/L, and 0.02 (0.02–0.02) ng/mL, respectively, upon treatment [Table 1].

Virus RNA detection in different samples from patients with COVID-19

The median time from the onset of symptoms to first negative RT-PCR results for oropharyngeal swabs of
convalescent patients was 9.5 (6.0–11.0) days with improvement in symptoms such as fever, cough, and dyspnea. This time varied greatly between patients, ranging from 2 to 22 days. The RT-PCR for viral RNA was performed using stool, urine, and blood specimens during convalescence. Until the end of the observation period (February 10, 2020), 11 convalescent patients (16.7%) still tested positive for viral RNA in stool specimens. The other 55 patients (83.3%) were negative for viral RNA following a median duration of 11.0 (9.0–16.0) days. Twelve patients (21.8% 12/55) still tested positive for viral RNA in stool samples from patients with COVID-19 and viral nucleic acids were subsequently detected in patients' stool specimens. There was no statistically significant difference in viral RNA detection in patient stool samples between the glucocorticoid treatment group (8.0 days, t = 2.550, P = 0.013) and the non-glucocorticoid treatment group (11 days, t = 1.509, P = 0.294) [Table 1].

Factors related to virus clearance

An analysis of the correlation between the absolute values of CD4+ T lymphocytes, CRP, red blood cell sedimentation rate, PCT, and the time of detoxification for feces during convalescence was performed. Using a multiple linear regression model (F = 2.699, P = 0.044, and adjusted R² = 0.122), the analysis showed that the CD4+ T lymphocyte count may help predict the duration of viral RNA detection in patient stool samples (t = −2.699, P = 0.010) [Table 2].

During hospitalization, five patients received glucocorticoid treatment. The duration of viral RNA detection in throat swabs and fecal samples in the glucocorticoid treatment group was longer than that in the non-glucocorticoid treatment group (8.0 days vs. 11 days, t = 4.631, P < 0.001, respectively). We further analyzed the differences in the results of the last tests for inflammatory indicators upon positive results for viral RNA in fecal samples. There was no statistically significant difference in inflammatory indicators between patients with positive and negative viral fecal RNA test results (P > 0.05) [Table 3].

Discussion

The novel coronavirus was firstly identified in respiratory specimens from patients with COVID-19 and viral nucleic acids were subsequently detected in patients' stool, urine, and gastrointestinal mucosa.[10-12] Recently a neonatal infection was reported, indicating the possibility of fecal-oral and vertical transmission from mother to child, in addition to the currently confirmed droplet transmission and direct contact transmission. In

| Table 1: Clearance time of viral RNA with or without glucocorticoid treatment. |
|------------------|-----------------|-----------------|-------------|-------------|
| Parameters       | All patients    | GC treatment    | No GC treatment | Statistics | P     |
|                  | (n = 66)        | (n = 5)         | (n = 61)       |             |       |
| Gender           |                 |                 |               |             |       |
| Female           | 38              | 4               | 34            | χ² = 0.342  | 0.559 |
| Male             | 28              | 1               | 27            |             |       |
| Age (years)      | 44.0 (34.0–62.0)| 51.0 (44.0–68.8)| 41.0 (34.0–61.3)| t = 1.059  | 0.294 |
| Pharyngeal swab virus nucleic acid negative time (days) | 9.5 (6.0–11.0) | 15.0 (9.8–16.8) | 8.0 (6.0–11.0) | t = 2.550  | 0.013 |
| Fecal virus nucleic acid negative time (days) | 11.0 (9.0–16.0) | 20.0 (17.5–22.5) | 11.0 (9.0–14.0) | t = 4.631  | <0.001|
| Difference between fecal and swab virus nucleic acid negative time (days) | 2.0 (1.0–4.0) | 8.0 (2.25–11.0) | 2.0 (1.0–3.0) | t = 1.983  | 0.115 |

Values are presented as n or median (IQR). Pharyngeal swab virus nucleic acid negative time: the time from when the patient has fever and respiratory symptoms to the first negative nucleic acid test of the throat swab. Fecal virus nucleic acid negative time: the time from the onset of symptoms to when the fecal virus nucleic acid test is first negative. Difference between fecal and swab virus nucleic acid negative time: the time between the first negative nucleic acid test of the throat swab and that of the fecal sample. GC: Glucocorticoid.

| Table 2: Multiple linear regression analysis of immune and inflammatory parameters with respect to virus clearance. |
|------------------|-----------------|-----------------|-------------|-------------|
| Independent variables | Coefficient | St. Error | Beta | t   | P  |
| CD4+ T lymphocyte counts (cell/μL) | −0.009 | 0.003 | −0.445 | −2.699 | 0.010 |
| ESR (mm/h) | 0.019 | 0.019 | 0.153 | 1.006 | 0.320 |
| CRP (mg/L) | −0.027 | 0.045 | −0.102 | −0.589 | 0.559 |
| PCT (ng/mL) | −6.278 | 15.607 | −0.064 | −0.402 | 0.689 |

CD4: Cluster of differentiation 4; ESR: Erythrocyte sedimentation rate; CRP: C-reactive protein, PCT: Procalcitonin.
Glucocorticoids have been widely used in the treatment of severe acute respiratory syndrome and Middle East respiratory syndrome, and are now also used in conjunction with other drugs to treat patients infected with 2019-nCoV. However, in the published clinical management opinions for the COVID-19, the application of glucocorticoids is not recommended unless there are other indications.[13,14] The use of glucocorticoids may delay the clearance of viral nucleic acids in patients and should be avoided during viral replication. Some bias exists in our study because the patients in the glucocorticoid treatment group had more severe disease and lower CD4+ T lymphocytes counts. Our point is that mild patients are not recommended glucocorticoid treatment, which may delay virus clearance. The randomized controlled double-blind experiments with expanded sample sizes will help clarify this issue. T cell immunity may play an important role in 2019-nCoV infection. The absolute values of CD4+ T lymphocytes, CRP, ESR, and PCT measured upon admission were analyzed with respect to virus clearance. The lower the absolute value of CD4+ T lymphocytes before treatment, the longer duration of virus clearance. The relationship between the fecal viral RNA results and inflammatory indicators of patients were analyzed and no statistical difference in ESR, CRP, or PCT during rehabilitation was found.

In brief, as the clearance of viral RNA from patients’ stools was delayed compared to that from oropharyngeal swabs, it is important to detect the viral RNA in feces during convalescence. Because of the delayed clearance of viral RNA in the glucocorticoid treatment group, glucocorticoids are not recommended in the treatment of COVID-19, especially for mild disease. The duration of RNA detection may be related to host cell immunity.

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### Conflicts of interest

None.

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