Introduction

Mesothelioma is a cancer arising from the transformation of mesothelial cells lining the thoracic (pleura) or the abdominal (peritoneum) cavities, and more rarely from transformed mesothelial cells of pericardium or tunica vaginalis testis. The development of mesothelioma has been related to the exposure to carcinogenic mineral fibers, mainly asbestos (1). The large and extensive utilization of asbestos since the mid-20th century, because of its insulating properties and low cost-effectiveness, was followed by a substantial increase in the age-standardized incidence of mesothelioma and related mortality rates until the developed countries strictly regulated (United States) or banned (Europe, Australia) the use of this material, after toxicological studies in vitro and in rodents demonstrated that asbestos fibers were carcinogenic (2).

The national regulatory documents use the generic term asbestos referring to six minerals that were commercially exploited, five amphiboles (crocidolite, actinolite, tremolite, anthophyllite, and amosite) and one serpentine (chrysotile). However, in the natural environment approximately further 400 minerals with similar physical and chemical features remain there, have not been regulated, and can be used unrestrictedly. The fibers of at least some of these minerals have been shown carcinogenic but can be airborne dispersed and affect the local human communities, drawing attention to the inadequacy of the current terminology (3). As an example, the residents of some Cappadocian villages in Turkey and in North Dakota (US) are exposed to naturally occurring erionite fibers, which are more carcinogenic than those of regulated asbestos, but have been used as...
construction or road paving materials (4,5).

We review here the current mechanisms of mesothelioma tumorigenesis, focusing on the impact of carcinogenic mineral fibers on mesothelial cells, the related molecular responses, and the relevance of gene-environment (GxE) interactions.

**Physicochemical features of the fibers govern their carcinogenic potential**

It has been shown by long time that dimensions, durability, and dose (“three D’s”) and physical properties are critical to determine the carcinogenic potency of certain types of mineral fibers (6-8). Fiber dimensions are related to durability and dose because influence bioavailability after inhalation. Long and thin fibers are associated with a higher potency regarding cytotoxicity and mutagenesis. A meta-analysis found that individuals exposed to fibers longer than 10 μm and even 20 μm have a significantly higher risk for asbestos-related disease (9), because longer fibers cannot be efficiently engulfed and cleared by macrophages leading to repeated failed attempts of phagocytosis. The resulting “frustrated phagocytosis” induces the inflammatory cells surrounding the fibers to release free radicals like reactive oxygen species (ROS) and reactive nitrogen species (RNS), exerting mutagenic activity (7,10). According to the WHO (World Health Organization) asbestos fibers can be operationally distinguished in short asbestos fibers (SAF) with length <5 μm and long asbestos fibers (LAF), having length >5 μm, diameter <3 μm and length/diameter ratio >3, which are targeted by the current regulatory rules (11). Moreover, differences in fiber biopersistence after exposure influence tumorigenesis, as serpentine chrysotile characterized by shorter biopersistence (12) compared with amphiboles and erionite, displays a lower carcinogenic potential. However, when the exposure to chrysotile fibers is prolonged, mesothelial cells are equally transformed (13). On the other hand, palygorskite (a very abundant fiber in southern Nevada) with reduced cytotoxicity and biopersistence in vitro, failed to induce carcinogenesis in vivo because it elicits much reduced inflammation compared to carcinogenic fibers (14).

**Fiber-related cell transformation**

The mechanisms of asbestos carcinogenesis have been elusive for long time and the initial hypothesis of the fiber mechanical interference with cell division has been definitely ruled out (2). However, some studies suggested that fiber chemical structure may play a role and iron in particular, as impurity or even as a chemical component of the fiber structure, has been taken into consideration to explain the carcinogenic process induced by asbestos. The deposition of asbestos fibers in tissues offers a surface where iron-reach macromolecular aggregates (asbestos bodies) favoring the development of chronic inflammation (15). Moreover, it has been shown that asbestos-activated macrophages release ROS that, in turn, may induce DNA damage indirectly via formation of 8-hydroxy-2’-deoxyguanosine (8-OHdG) adducts (16). Recent data on iron-catalyzed ROS production suggests that ferroptosis, a non-apoptotic, iron-dependent cell death, may be involved in asbestos-related carcinogenesis (17). Moreover, a role in asbestos-induced carcinogenesis has been postulated for hepatocyte growth factor (HGF), via activation of the PI3K/MEK5/Fra-1 axis (18).

On the other hand, the exposure to asbestos fibers causes death of human mesothelial cell (HM), a cell type particularly susceptible to fiber cytotoxicity that was initially ascribed to apoptosis (19). Afterwards, asbestos pathogenesis was clearly associated with tumor necrosis factor-alpha (TNF-α), a mediator of inflammation (20).

**Chronic inflammation and mesothelioma**

Chronic inflammation plays a major role in the pathogenesis and tumorigenesis induced by asbestos and other carcinogenic mineral fibers. The pro-inflammatory microenvironment established at the site of fiber deposition with the contribution of both HM and macrophages, combined with the biopersistence of many mineral fibers, allow some HM to avoid cell death and eventually go through oncogenic transformation (1).

It has been shown that the largest fraction of HM exposed to crocidolite (21) and chrysotile asbestos (13), as well as to erionite fibers (4), undergo cell death via programmed cell necrosis (21). This is a regulated form of necrosis characterized by the passive release of high mobility group box 1 (HMGB1) by necrotic HM at the site of fiber deposits. HMGB1 is a damage-associated molecular protein (DAMP) that promotes the recruitment of macrophages sustaining the chronic inflammatory process (21,22). HMGB1 binds to RAGE and other HMGB1 receptors of macrophages priming for inflammasome activation, combined with other stimuli, like endogenous ROS formed after asbestos exposure, through the assembly
of NLRP3 inflammasome via oligomerization of inactive NLRP3, apoptosis-associated speck-like protein (ASC) and procaspase-1. NLRP3 inflammasome induces the release of IL-1β, IL-18, IL-1α, and HMGB1, establishing an autocrine chronic inflammation process (23). During this process also TNF-α is secreted and activates NF-κB, promoting survival of HM upon asbestos exposure. The surviving HM will continue to proliferate and accumulate genetic mutations, leading to development of mesothelioma (1). The role of HMGB1 and related chronic inflammation is supported by the report on the preventative role of aspirin for mesothelioma, targeting HMGB1 activities and inflammation, and on the antitumor activity of aspirin in mesothelioma xenograft models (24). Moreover, mesothelioma cell growth was inhibited both in vitro and in vivo by ethyl pyruvate that has been characterized as an effective inhibitor of HMGB1 and suppressor of the expression of the RAGE receptor. Both activities contribute to reducing mesothelioma malignancy (25).

The anti-tumor activity of these widely used anti-inflammatory drugs is explained by the high levels of HMGB1 expression and secretion in the extracellular milieu found in mesothelioma cells, compared with HM and by the findings that competitive inhibitors of HMGB1 delay growth of mesothelioma xenografts (26). HMGB1 is localized mainly in the nucleus of HM, while in mesothelioma HMGB1 was found in both nucleus and cytosol (26). The subcellular localization of HMGB1 is determined by the balance between histone acetyltransferase (HAT) and histone deacetylase (HDAC), controlling the HMGB1 acetylation status (27,28), which is also regulated by poly(ADP-ribose) polymerase-1 (PARP-1) (29). In mesothelioma HMGB1 is actively secreted into the extracellular space (30), where it establishes an autocrine mechanism with RAGE and TLR receptors that promotes proliferation, motility, and survival, leading to the progression of mesothelioma (26).

The role of genes and environment

Asbestos fibers initiate HM death mainly through necrosis (21) and to a lesser extent by other cell death mechanisms (31), are also studied in parallel. Carcinogenesis is commonly related to somatic gene mutations affecting the DNA repair mechanisms, leading to the accumulation of DNA damage and the consequent increase of the fraction of cells carrying damaged DNA. When these cells acquire mechanisms of survival, as those elicited by the HMGB1 pathway in mesothelioma, cancer may develop. The presence of inherited mutations affecting DNA repair and other genes may further contribute to the process of carcinogenesis, by increasing the susceptibility to environmental carcinogens (32). The current approach adopted in the field of carcinogens is to combine genetics and environmental studies to study GxE interactions (2).

The catastrophic event of chromothripsis has been recently attributed to the increase of the mutational level of the cancer cell genome. Chromothripsis develops upon the shattering of a segregated single chromosome that is randomly reassembled, leading to incorrect rearrangements or deletions of DNA sequences. Therefore, even after a short number of cell divisions, massive genome alterations may occur following a single chromothripsis event. In turn, this high mutational status favors oncogene activations or loss of tumor suppressor functions, eventually promoting tumorigenesis (33). Notably, genomic studies of mesothelioma cells and specimens identified non-contiguous biallelic genome alterations with the distinctive pattern of chromothripsis (34,35), and associated with potential neontigen expression, with possible and intriguing implications in mesothelioma immunogenicity (36).

Multiple tumor suppressors involved in the cell cycle control and in apoptosis were found mutated in human mesothelioma. One of the common genetic alterations in mesothelioma is the homozygous deletion on locus 9p21 (37), which affects the transcription of two tumor suppressors: p16INK4a and p14ARF. P16INK4a blocks cell division via binding to CDK4 and CDK6, and p14 promotes apoptosis by inhibiting p53 ubiquitylation. Cytogenetic studies showed that p16 was missing in up to 80% primary pleural mesotheliomas (37), while p16 inactivation suggests the association with poor clinical outcome (38). Transgenic p14 (+/-) mice were more susceptible to asbestos-induced carcinogenesis and harvested primary mice tumors exhibited loss of heterogeneity for p14 (39).

Intermediates in the Hippo signaling pathway are also highly mutated in mesothelioma. Neurofibromatosis type 2 (NF2)/Merlin, an upstream initiator of Hippo, is inactivated in about 40% of malignant mesothelioma (40). Notably, NF2 is the second most frequent mutated gene in mesothelioma after BRCA1 associated protein-1 (BAP1). Heterozygous NF2 (+/-) mice were more sensitive to asbestos exposure and demonstrated an accelerated tumorigenesis compared to wildtype controls (41). Non-functional NF2 leads to nuclear accumulation of yes-associated protein (YAP) and WW Domain-containing...
transcription regulator (WWTR1 or TAZ) in the Hippo pathway. One of the consequence of the pro-inflammatory environment provoked by exposure to asbestos fibers is the enhanced formation in the nucleus of the YAP/TAZ complex, which in turn promotes the expression of multiple proto-oncogenes, supporting cancer cell survival (42).

**Inherited BAP1 mutations and mesothelioma**

Epidemiological studies have provided evidence that only about 5% of miners and shipyard or manufacturing workers with prolonged exposure to asbestos developed mesothelioma (2). Studies on a mesothelioma epidemics among villagers in Cappadocia, Turkey, heavily exposed to erionite fibers and with an unusually high incidence of mesothelioma discovered the transmission of the susceptibility to mesothelioma in Mendelian autosomal dominant inheritance (5,43).

The model of GxE interactions predicts that carriers of germline mutations have greater susceptibility to fiber-induced carcinogenesis and to the development of mesothelioma. This prompted the search of the gene(s) possibly involved. In two unrelated US families with no occupational exposure to asbestos and with high incidence of mesothelioma, array-comparative genomic hybridization (aCGH) and linkage analysis allowed the identification of possible frequent alterations at chromosome 3p21. Subsequent sequencing identified germline BAP1 mutations associated with autosomal dominant transmission of mesothelioma and uveal melanoma (44). In a parallel paper, germline mutations of BAP1 were linked to dominant inheritance of melanocytic tumors (45). The individuals with germline mutated BAP1 were also susceptible to other types of cancer like renal cell carcinoma and squamous cell carcinoma, leading to the identification of the BAP1 cancer syndrome (46).

Animal studies performed using Bap1<sup>+/−</sup> heterozygous mice demonstrated that animals developed mesothelioma when exposed to ten-time lower doses of asbestos fibers that barely caused any mesothelioma in wild type Bap1 mice (47).

BAP1 was initially characterized as a nuclear protein with deubiquitylase activity, which is part of multiprotein transcriptional regulators of genes involved in metabolism, mitochondrial function, and cell proliferation (48). Nuclear BAP1 is also associated in chromatin remodeling (49), DNA double-strand repair (50,51), and auto-deubiquitylation promoting its own nuclear localization (52). Recent studies demonstrated that BAP1 is endowed with a dual activity, both in the nucleus and in the cytoplasm, which cooperate to cause tumor suppression. Cytoplasmic BAP1 is prevalently localized in the endoplasmic reticulum (ER) fraction, where it deubiquitylates and stabilizes the type 3 inositol-1,4,5-trisphosphate receptor (IP3R3). The function of IP3R3 is the release of Ca<sup>2+</sup> from ER into the mitochondrial space through voltage-dependent anion channels (VDACs) of the outer membrane and the mitochondrial uniporter channel (MUC) of the inner mitochondrial membrane. The increase of Ca<sup>2+</sup> concentration in the mitochondria induces the release of cytochrome c activating apoptosis. In heterozygous BAP1<sup>+/−</sup> conditions, as in the individuals of the families with the BAP1 cancer syndrome, the reduced BAP1 dosage impairs both the DNA repair, accumulating DNA damage, and the apoptotic response. This dual effect positively selects cells carrying oncogenic mutations and promotes tumorigenesis (53). Moreover, the reduced BAP1 dosage in the cytoplasm has another consequence that was ascertained by metabolomics analysis. In non-tumoral cells, reduced BAP1 levels induced the Warburg effect, a shift of cell metabolism from oxidative phosphorylation (Krebs cycle) to aerobic glycolysis, originally identified as typical of cancer cells. However, the increase of aerobic glycolysis and lactate production in normal fibroblasts carrying heterozygous BAP1<sup>+/−</sup> indicates a new role for the Warburg effect in anticipating cancer onset by accelerating tumorigenesis (54).

Germline BAP1 mutations were associated with the increased risk of other tumors, like clear cell renal cell carcinoma (55,56), cutaneous melanoma, ocular melanoma (57), atypical Spitz tumors (46,58), basal cell carcinoma (59,60), peritoneal mesothelioma (61-63), cholangiocarcinoma (64), and meningioma (65,66). A worldwide genome analysis of BAP1 germline mutations combined with the survey of the clinical features of the BAP1 cancer syndrome, showed the presence of the core syndrome tumors in a significant fraction of the families investigated by the study (67-69).

Further studies performed a genotype analysis in cancer patients, including those at high risk for familial inheritance, to identify germline alterations in additional genes possibly involved in the predisposition to mesothelioma and other cancers associated with GxE interactions (62,70,71).

**Conclusions**

The cause of mesothelioma was uniquely attributed to asbestos exposure for a long time, however not all
individuals exposed to asbestos and the other unregulated, naturally occurring carcinogenic mineral fibers like erionite, develop mesothelioma. The picture was further clarified when it was demonstrated that the genetic factors play critical roles in susceptibility to mesothelioma (5,43). A progress in the knowledge on the causes of mesothelioma and other cancers related to the model of GxE interaction was the identification of \textit{BAP1} as a predisposition gene for the development of familial mesothelioma (44), leading to the discovery of the BAP1 cancer syndrome (46,48).

The mechanisms of cellular transformation following the exposure of HM to carcinogenic mineral fibers was elucidated by the discovery of the role of chronic inflammation mediated by HMGB1 and the inflammasome (21,23). Moreover, the identification of BAP1 as a main controller of cell death and metabolism contributed to the definition of the complex array of molecular events mediated by asbestos carcinogenesis (53,54).

Further studies will be required to identify the complete picture of the genes predisposing to mesothelioma and their contribution to the molecular mechanisms of asbestos carcinogenesis discovered so far, including chronic inflammation and altered metabolism.

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**Footnote**

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