Vitellogenin Level in the Plasma of Russian Sturgeon (Acipenser gueldenstaedtii) Northern Israel

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Abstract

In the present study, we examined the vitellogenin (Vg) level of Russian sturgeon maintained under specific and relatively constant aquaculture conditions at water temperature of 18-22°C during growth and maturation.

An increase in Vg in the blood plasma was found in the yellow oocytes stage and the gray oocytes stage. However, no Vg was found in the pre-vitellogenic stage. Based on the present study and previous studies on hormonal control of reproduction and growth, we proposed a quality model that correlated between egg development and the hormones involved in controlling vitellogenesis (VTL).

Keywords Vitellogenin; Russian sturgeon; Vitellogenesis; Oocytes

Introduction

Russian sturgeon (Acipenser gueldenstaedtii) is osteichthyes fish that belong to the Osteichthyes class, Actinopterygii sub-class, Acipenseriformes order and Acipenseridae family [1]. Russian sturgeon (A. gueldenstaedtii) belong to one of the most ancient group of osteichthyes fish found along the coasts of the Atlantic and Pacific Oceans, in the Caspian and Black Seas, and in many rivers, lakes and inland seas [2]. The decline of sturgeon populations in their native habitats, mainly in the Caspian Sea, is due to overfishing for meat and caviar production, the destruction of their spawning grounds and water pollution. This has led to bringing about their introduction into aquaculture.

The adaption of Russian sturgeon to aquaculture conditions [3] was studied, as well as different aspects of hormonal control of growth and reproduction under artificial conditions in order to improve survival rate, growth and reproduction success [4-9]. Nevertheless, information on gonadal development [9] and puberty of Russian sturgeon in northern Israel aquaculture is important for caviar production, although much information about this species has still not been studied [10,11]. During oogenesis, the Vg is a phospholipoglycoprotein precursor contained in the yolk of the fish oviparous, similar to other vertebrates, and is an estrogen-induced female-specific serum protein that is biosynthesized in the liver, secreted into the bloodstream, and involved in the process of yolk formation during oogenesis. This effect on VTL (also known as yolk deposition in oocytes) is very important for processing oocytes transfer in preparation for becoming caviar, and therefore various sturgeon species were studied [10-12].

The few hormones involved in controlling VTL are mainly from the gonadotropin axis hypothalamic gonadotropin-releasing hormone (GnRH), Luteinizing (LH) and Follicle stimulating hormone (FSH) [4,7] and the somatotropin axis growth hormone (GH) and insulin-like growth factor I (IGF-I) [6,8,13], as well as the interaction between them.

The aim of the present study was to measure the level of Vg in Russian sturgeon under specific aquaculture conditions through growth and oogenesis.

Materials and Methods

Fish and sampling procedure

Russian sturgeons were approved for study by the government since they could add to our knowledge about caviar production. Fertilized eggs of Russian sturgeon (A. gueldenstaedtii) originating in the Caspian Sea were brought from Russia and reared in the Dan Fish Farms (Upper Galilee, Israel; 31°30’N, 34°45’E) under aquaculture conditions, as previously described [9] (Figure 1). Fish were maintained in 250 m³ ponds made of concrete in 30kg/m³ at a water temperature of 18-22°C and 12h photoperiod. The oxygen concentration was 6 ppm and was controlled by an oxygenic machine. Ammonia and nitrite were below 0.5 ppm.

The fish were fed twice a day with 1% body weight of dry food [48% protein and 18% fat].

Fish and sampling

Blood samples from Russian sturgeon males and females were taken. Each fish was anesthetized in a clove oil bath (0.25 mg/l) and placed in a V-shaped base for examination, as described previously by Hurvitz et al. [9]. Samples of 1 ml were collected from each fish from the caudal blood vessels. The samples were taken from females of various sizes, ages (1, 2, 3, 4, 5, 6 and 7 years old) and ovary...
development; the stage of ovarian maturity was analyzed by endoscopy (Figure 2).

Serum was then separated by centrifugation at 1,500 RPM for 10 min and stored at -20°C until use. Blood was collected from immature females, with transparent oocytes at a diameter of 0.4 mm, and from females with ovaries in the yellow oocytes stage and the gray oocytes stage at a 2-2.5 diameter. The different states were photographed by endoscopy [9].

Histological analysis

A histological analysis was generally performed according to [5]. Briefly, gonadal samples were fixed in Bouin’s fluid for 48 h and then transferred to 70% ethanol for storage until processing for light microscopy. Paraffin sections of 4 to 7 μm were stained with hematoxylin and eosin. For a description of the gonadal stages, the terminology used by Hurvitz et al. [9] was adopted and the histology of oocytes was described by this study. Mean oocyte diameter was calculated for each fish after measuring five of the largest oocytes present in the fixed tissue before processing. Oocyte diameter was measured using an optical binocular with notches (CETI Belgium) at an accuracy ± 50 μm.

Vitellogenin (Vg) analysis

Vg was measured by gel electrophoresis and Western blot analyses. Electrophoresis and Western blot analysis SDS-PAGE (5–22.5% or 8–
25%) were carried out using electrophoresis on 7.5% polyacrylamide gel electrophoresis (PAGE), as described previously by Laemmli (1970) using a system of PROTEAN II mini (Biorad, USA).

An analysis was made for the protein transfer using a Western blot membrane (Biorad, USA) with TBS (20 mM Tris-HCl, 150 mM NaCl, pH 7.5). To label the marking of the VTG, the membranes were incubated for two hours with mouse anti-Gulf sturgeon VTG (Biosense Laboratories, AS diluted 1:250) at room temperature and washed three times with TBS (20 mM Tris-HCl, 150 mM NaCl, pH 7.5) including 0.05% Tween. For the second incubation, IgG (TBS-T horse radish peroxidase conjugated to goat anti-mouse IgG) was used for 1 hour and then washed again as described previously. The marked bands were identified by ECL (Amersham-Pharmacia-Biotech).

Results

![Figure 4: SDS-PAGE of plasma Vg from white sturgeon. The 250 KD band represents VTG in female sturgeon. Females with ovaries full of gray eggs were used as positive control (PC) in the left panel, and male plasma was used as negative control. Numbers 141-154 refer to individual females sampled, and + or – markings in the table represent gonads stage as determined by endoscopy. The ovaries at the different states were photographed by endoscopy: yo – yellow oocytes stage, and go – gray oocytes stage. All of the oocytes in the ovary are transparent with a diameter of 0.4 mm. + A few yellow oocytes are dispersed in ovaries smaller than 1 mm. + + Many yellow oocytes are dispersed in ovaries with a diameter of 1-2 mm. +++ All of the ovaries contain yellow or gray oocytes with a diameter of 2-2.5 mm.](image)

![Figure 5: Suggested model and hormones involved in the vitellogenin protein that controls vitellogenesis in Russian sturgeon oogenesis. A – [4], B – [5] C [9], D –[6,13] E – [7] and F – present study [10,12](image)

Discussion

The present study contributes not only to providing a description of the level of Vg in the plasma of Russian sturgeon for the first time during the various stages of oocytes development of fish grown in specific aquaculture conditions in which the water is of high quality and at a constant temperature all year round, but also to comparing the results of the hormones involved in controlling reproduction specifically on the gonadotropin [4,5,7,9] and the somatotropin [6,8] axes (Figure 5). The results of the present study are in agreement with a previous study on white sturgeon, which found that plasma Vg concentrations correlated with total plasma calcium and stage of ovarian maturity [10]. However, in white sturgeon, Vg could be measured in the previtellogenic stages.

Vitellogenin (Vg) is a phospholipoglycoprotein precursor contained in the yolk of oviparous vertebrates, and is an estrogen-induced female-specific serum protein biosynthesized in the liver, secreted into the bloodstream, and involved in the process of yolk formation, called VTL. Although Vg has been purified and characterized in many fish species [12], including species belonging to the genus Acipenser [10-12], the biochemical characteristics in Russian sturgeon have not been studied extensively, and we assume that there are no significant differences compared to other species belonging to the Acipenser. VTG in sturgeon grown in the Dan Fish Farms is an example of fish grown under relatively constant conditions, and the growth rate correlates to VTG. This environmental condition effect during hormone control reproduction on VTG was described in other fish under experimental conditions [14]. In the present and previous research studies carried out on Russian sturgeon grown in the Dan Fish Farms under constant conditions, the growth rate was linear. Vg might be correlated to

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growth, but a more specific study on Russian sturgeon under these conditions is required.

In summary, for the relationship between eggs development and hormones involved in controlling VTG based on previous studies, we suggested the quality model described in Figure 5.

References

1. Vlasenko AD, Pavlov AV, Sokolovm LI, Vasilev VP (1989) The freshwater fishes of europe. Acipenseriformes. Acipenser gueldenstaedtii brandt 1.
2. Dettlaff TA, Ginsburg AS, Schmalhausen OI (1993) Sturgeon fishes: Developmental biology and aquaculture. Springer Verlag.
3. Doroshevt SI, Moberg GP, Van Eenennaam VP (1997) Observations on the reproductive cycle of cultured white sturgeon, acipenser transmontanus. Environ Biol Fisches 48: 265-278.
4. Hurvitz A, Degani G, Goldberg D, Yom Din S, Jackson K, et al.(2004) Cloning of fshβ, lhbβ and glycoprotein α subunits from the russian sturgeon (acipenser gueldenstaedtii), β subunits mrna expression, gonad development and steroid levels in immature fish. Gen Comp Endocrinol 140: 61-73.
5. Jackson K, Hurvitz A, Yom Din S, Goldberg D, Perelson O, et al. (2006) Anatomical, hormonal and histological descriptions of captive russian sturgeon (acipenser gueldenstaedtii) with intersex gonads. Gen Comp Endocrinol 148: 359-367.
6. Yom Din S, Hurvitz A, Goldberg D, Jackson K, Levavi-Sivan B, et al. (2008) Cloning of russian sturgeon (acipenser gueldenstaedtii) growth hormone and insulin-like growth factor 1 and their expression in male and female fish during the first period of growth. Journal of Endocrinological Investigation 31: 201-210.
7. Yom-Din S, Hollander-Cohen JA, Boehm BI, Shpilman M, Golan M, et al. (2016) Gonadotropins in the russian sturgeon: Their role in steroid secretion and the effect of hormonal treatment on their secretion. PLOS ONE 13: 1-23.
8. Degani G, Yom-Din S, Hurvitz A (2017) Transcription of insulin-like growth factor receptor in russian sturgeon (acipenser gueldenstaedtii) ovary during oogenesis. Universal Journal Agricultural Research 5: 119-124.
9. Hurvitz A, Jackson K, Degani G, Levavi-Sivan B (2007) Use of endoscopy for gender and ovarian-stage determinations in russian sturgeon (acipenser gueldenstaedtii) grown in aquaculture. Aquaculture 270: 158-166.
10. Linares-Casenave J, Kroll KJ, Van Eenennaam JP, Doroshevt SI (2003) Effect of ovarian stage on plasma vitellogenin and calcium in cultured white sturgeon. Aquaculture, 221: 645-656.
11. Van Eenennaam JP, Doroshevt SI (1998) Effect of age and body size on gonadal development of atlantic sturgeon. J Fish Biol 53: 624-637.
12. Zhang Y, Qu Q, Sun D, Liu X, Suo L, et al.(2011) Vitellogenin in amur sturgeon (acipenser schrenckii): Induction, purification and changes during the reproductive cycle. J Appl Ichthyol 27: 660-665.
13. Wuertz S, Nitsche A, Jastroch M, Gessner J, Klingenspor M, et al. (2007) The role of the igf-1 system for vitellogenesis in maturing female starlet, acipenser ruthenus linaeus. Gen Comp Endocrinol 150: 140-150.
14. Levy G, David D, Degani G (2011) Effect of environmental temperature on growth- and reproduction-related hormone gene expression in the female blue gourami (trichogaster trichopterus). Comparative Biochemistry and Physiology 160: 381-389.