Research Article

Selected Parameters of Nutritional and Pro-Health Value in the Common Carp (Cyprinus carpio L.) Muscle Tissue

J. Klobukowski,1 K. Skibniewska,2 K. Janowicz,1 F. Klobukowski,3 E. Siemianowska,2 E. Terech-Majewska,4 and J. Szarek5

1Department of Human Nutrition, University of Warmia and Mazury in Olsztyn, Olsztyn, Poland
2Department of Foundations of Safety, University of Warmia and Mazury in Olsztyn, Olsztyn, Poland
3Department of Food Commodity Science, Medical University of Gdańsk, Gdańsk, Poland
4Department of Epizootiology, University of Warmia and Mazury in Olsztyn, Olsztyn, Poland
5Department of Pathophysiology, Forensic Veterinary Medicine and Administration, University of Warmia and Mazury in Olsztyn, Olsztyn, Poland

Correspondence should be addressed to K. Janowicz; katarzyna.janowicz@uwm.edu.pl

Received 12 December 2017; Revised 10 February 2018; Accepted 27 February 2018; Published 26 April 2018

1. Introduction

Consumption of fish in Central and Eastern Europe is still insufficient, which is a consequence of consumers’ unsatisfactory knowledge and prices. Greater demand for this type of food is observed among people with university education and in circles where eating fish is a family tradition [1]. However, it is beyond doubt that increasing attention is being paid to food quality, both in Poland and elsewhere. The quality of fish muscle tissue, in terms of both the qualitative and the quantitative characteristics, including its nutritional value and chemical composition of muscles, can be modified by changing breeding conditions and, more specifically, feeding technologies.

Considerations of the pro-health effect of food must take into account the important role of animal origin fat, including polyunsaturated fatty acids and cholesterol in human nutrition. Fish, which is a food unmodified over the centuries and which can be regarded as one of humans’ prime foods, is a valuable source of those nutrients [2, 3]. The spectrum of health-promoting effects of n-3 PUFAs on the cardiovascular system is very broad. Numerous studies have demonstrated that the consumption of a large number of foods providing a source of these acids, inter alia the application of a Mediterranean diet that is rich in fish, contributes to a reduction in the risk of the incidence of coronary heart disease and cardiac insufficiency. Moreover, the effectiveness of PUFAs in the course of coronary heart...
The results of laboratory tests carried out after four weeks subjected to testing. In the diet of the first one, 200 g of a fillet of common carp fed with a mixture with increased n-3 PUFA content (439 ± 146 mg/100 g of the fillet) was incorporated. The total energy consumption in both groups was the same. The results of laboratory tests carried out after four weeks of the experiment clearly indicated the positive effect of the consumption of common carp in the event of CVDs. Plasma lipid levels were significantly improved in the group of patients who had fish incorporated into their diet compared to the control group. A reduction in the total cholesterol level by 27% was demonstrated, with a decrease by 2% in the control group (p < 0.001). LDL cholesterol level decreased by 26% as compared to 4% (p < 0.001); the TG value in the plasma decreased by 26% as compared to 3% (p < 0.001). An increase in HDL cholesterol level by 30% in relation to 10% was observed (p < 0.001). Based on a POLKARD–SPOK study, Filipiak and Opolski [13] indicate the occurrence of disturbances in lipid metabolism in 78% of the population of patients at high risk of death due to CVDs. The data suggests that it is beneficial to enrich the diet with polyunsaturated fatty acids to improve the values of blood serum lipidogram through an increase in the consumption of common carp, particularly in Central European countries.

2. Material and Methods

Material from five fish farms in different locations in Poland was used in the study; the production technology applied in them was intensive (I) in 2 farms, low-intensive (NI) in 1 farm, and semiextensive (SE) in 2 farms. In order to avoid the effect of climate on the growth of common carps, which are poikilotherms, samples were taken from ponds situated in various parts of the country. Most farms were characterised by single-season production. Only one of the farms, with a semiextensive technological level, carried out multiseason production. Of the mentioned types of farms, three types of ponds were noted: commercial and fattening fish bins (I), only commercial (NI), and commercial I recreational and angling (SE). The stocking density in the farm applying the intensive production technology was 1,600 fish/ha, in the ponds with the low-intensive technological level was 1,315 fish/ha, and for the semiextensive production was 600 fish/ha. Particular farms were distinguished by production at a level of 1,002.5 kg/ha, 993 kg/ha, and 659 kg/ha, respectively, for the intensive, low-intensive, and semiextensive technological levels. On average, the weight of common carps harvested in the farm with intensive production technology was 1,159.5 g per fish; common carps harvested from the ponds with the low-intensive production technology had an average weight of 1,523 g per fish, whereas those from ponds characterised by semiextensive production technology had an average weight of 1,424 g per fish. The following types of feed were used in different farms:

(i) Cereal mixture—wheat, barley, and rye in the ratio of 3:1:1 or only fishing bait in semiextensive farms.

(ii) Cereal mixture (wheat, barley, and rye in the ratio of 2:1:1) or a cereal mixture (maize + wheat and barley in the ratio of 1:3) in low-intensive production technology.

(iii) Granulate Aller-Aqua, which comprises dried distiller grain, sunflower protein concentrate, soybean protein concentrate, fish meal, poultry meal, blood meal, feather meal, rapeseed oil, wheat, triticale, rape, soybean, vitamins, minerals, and amino acids, in intensive production technology.
 Altogether, 75 fish were taken for the study, 15 consumer common carps of the S grade (0.8–1 kg) and D grade (1-2 kg). The fish were then killed, boned, and stored at −18°C.

In order to conduct a chemical analysis, a 5 cm wide boneless and skinless section was taken from a frozen fillet from the ventral to the dorsal side. The next stage involved grinding and homogenisation of a sample in a homogeniser (Type PRO350 BIOEKO, time: 40 minutes, speed: 11,000 RPM).

The dry weight was determined by drying samples to a constant mass, in accordance with the guidelines of AOAC [14]. Total ash was determined by drying, carbonising, and incineration [14]. Total protein content (nitrogen ×6.25) was determined by the Kjeldahl method [15].

Fat was extracted in accordance with the procedure proposed by Folch et al. [16]. Two-gram homogenised samples of muscle tissue were homogenised with 20 ml of methanol for 1 minute and with 40 ml of chloroform, also for 1 minute. The homogenate was filtered through a degreased filter. The remainder was washed with 60 ml of mixture of methanol and chloroform at a ratio of 2:1. The filtrate was then washed with 40 ml of chloroform and 20 ml of methanol and combined with 0.88% of sodium chloride at 25% of the filtrate volume. The mixture was shaken and left overnight for the layers to separate. The upper layer was collected with a vacuum pump and rejected. The lower layer was a lipid extract. A mixture of water and methanol in a ratio of 1:1 was added to it, in two replicates, in an amount of 1/4 of the amount of filtrate, and its top layer was each time discarded. The lipid extract was filtered through calcined sodium sulphate (VI) on a degreased filter and the solvent was then distilled off. The remainder was weighed in accordance with AOAC 2002 guidelines [17].

Methyl esters of fatty acids were prepared by the modified method developed by Peisker [18]. A 0.5 g sample of fat was put into an ampoule, 2 cm² of methylating mixture (methanol: chloroform: concentrated sulphuric acid, at a ratio of 100:100:1 (v/v/v)) was added, and the ampoule was sealed. The ampoules were heated in a water bath for two hours at 100°C. The fatty acids were analysed by gas chromatography on a 6890N Agilent Technologies chromatograph under the following conditions: capillary column with internal diameter of 0.32 mm, length 30 m (Supelcowax 10 liquid phase, film thickness of 0.25 µm), injector (split 50:1) (flow rate of 10 ml/min), injector temperature of 225°C, detector temperature of 250°C, and column temperature of 180°C. Fatty acids were identified through a comparison of retention times of the standards (a mixture of 37 acids) and peaks in the test sample.

The gross calorific value (\( W_g \) kcal/100 g) of breeding common carp muscle tissue was calculated using physical energy equivalents from the following formula: \( W_g = 5.65 \times B + 9.45 \times T + 4.15 \times W \) [kcal/g] [19], where \( B \) is the protein content in a sample, g/100 g; \( T \) is the fat content in a sample, g/100 g; \( W \) is the carbohydrates content in a sample, g/100 g; 5.65, 9.45, and 4.15 are physical energy equivalents for proteins, fats, and carbohydrates, respectively, kcal/g.

The net calorific value (\( W_n \) kcal/100 g) of the carcasses was calculated using Atwater net equivalents, from the following formula: \( W_N = 4 \times B + 9 \times T + 4 \times W \) [kcal/g] [19], where \( B \) is the protein content in a sample, g/100 g; \( T \) is the fat content in a sample, g/100 g; \( W \) is the carbohydrates content in a sample, g/100 g; 4, 9, and 4 are Atwater net equivalents for proteins, fats, and carbohydrates, respectively, kcal/g. When the calorific values \( W_g \) and \( W_N \) were expressed in kJ/100 g, a conversion factor of 1 kcal = 4.19 kJ was applied. Since common carp muscle tissue contains only trace amounts of carbohydrates; this energy component was left out of the calculations [19].

The dietary atherogenic index (AI) and thrombogenic index (TI) were estimated with the formulae developed by Ulbricht and Southgate [20]. The atherogenic index was calculated from the formula \( AI = C12:0 + 4 \times C14:0 + C16:0/\sum PUFA \times n-6 + \sum PUFA \times n-3 + \sum MUFA \), where \( C12:0 \), \( C14:0 \), and \( C16:0 \) are the content of saturated acids: lauric, myristic, and palmitic (%); \( \sum PUFA \times n-6 \) are the total polyunsaturated fatty acids n-6 (%); \( \sum PUFA \times n-3 \) are the total polyunsaturated fatty acids n-3 (%); \( \sum MUFA \) are the total monounsaturated fatty acids (%). The thrombogenic index (TI) was calculated from the formula \( TI = C14:0 + C16:0 + C18:0/(0.5 \times C18:1) + 0.5 \times (MUFA-C18:1) + (0.5 \times PUFA-n-6) + 3 \times PUFA \times n-3 + (PUFA \times n-3/PUFA \times n-6) \), where: \( C14:0 \), \( C16:0 \), and \( C18:0 \) are the content of saturated acids: myristic, palmitic, and stearic (%); MUFA is monounsaturated fatty acid; PUFA n-6 are polyunsaturated fatty acids n-6 (%); PUFA n-3 are polyunsaturated fatty acids n-3 (%).

The statistical analysis was performed using Statistica 12 software, with the t-test for samples independent from groups at \( p \leq 0.05 \).

3. Results

An analysis of the composition of common carp muscle tissue from fish farms of three types has shown a distinct effect of the method of feeding on the nutritional and energy value of the meat. Common carp muscle tissue from a farm which applied the intensive breeding technology had the highest calorific value (gross: 182 kcal; net: 102 kcal) followed by the material from farms where semiextensive production technology was applied (116 and 91 kcal, resp.). The lowest calorific value was recorded for the muscle tissue of common carp bred by the low-intensive technology; the gross calorific value of the product was 109 kcal and net calorific value was 88 kcal (Table 1). The net calorific value is an especially important parameter for living organisms; it is the amount of energy actually used by an organism for life processes. The muscles of fish fed by the intensive method also contained the highest percentage of dry matter (24.6%), protein (18.74%), and fat (15.8%) in the carcass, which obviously has its effect on the calorific value. Moreover, the content of dry matter is closely connected with the amount of fat in muscle tissue, as it is a component whose form does not change significantly during drying. Muscle tissue of fish from low-intensive breeding contained the lowest level of fat of all the samples: 0.56%. The dry matter content was 22.31% and the total protein content was 18.32% (Table 1). Despite the similarity of the feed used in low-intensive and semiextensive farms (cereal mixtures), the higher protein content in fish muscle tissue from the
first type of breeding could be an effect of an addition of maize in the feed. This cereal contains similar levels of protein to other species, but it was an additional component of the feed, thereby increasing its consumption by fish. Regardless of the feed or production technology, monounsaturated fatty acids dominated in common carp muscle tissue (Table 2). For example, the values for C18:1 n-9 were not statistically significant differences between the two extreme types of fish farms: intensive and semiextensive. Only for C18:2 n-6 or C20:1 n-9 acids were no statistically significant differences found. Only two cases with no statistically significant differences were identified in an analysis of common carp muscle tissue by the low-intensive and semiextensive method. Those were the content levels of fatty acids C20:2 n-9 and C22:6 n-3 (Table 2). It can be concluded from the results of the statistical analysis of the fatty acid profile of fish muscle tissue from three types of fish farms that the type of feed has a great effect on its composition. The contents of various profile components vary from one fish type to another. There are some other cases with no differences between them, but only on the farms where intensive and low-intensive production technology was applied. This arises from the similarity of feed used in these fish ponds and the small number of results with no statistically significant differences between farms: I-SE, LE-SE, which clearly shows the differences in the nutritional value of the feed and translates into the material composition. Regardless of the type of common carp production technology, the fish fat contained saturated and unsaturated fatty acids at a beneficial ratio (Figure 1). The ratio was 0.84 in common carp caught on intensive farms, 0.74 in the low-intensive ones, and 0.87 in semiextensive ones. An examination of the fatty acid profile in common carp muscles has shown that they are not only a source of PUFA n-3 and PUFA n-6; the values of ω3 and ω6 are considerably lower than for other animal fats (Figure 2). For example, the atherogenic index for cow milk is ca. 3−4. The values for fish muscle tissue are 0.43, 0.54, and 0.35 for intensive, low-intensive, and semiextensive technology, respectively. For the samples of muscles under study, the values were 0.46, 0.65, and 0.84 for intensive, low-intensive, and semiextensive breeding, respectively.

| Item                     | Intensive (n = 30) | Production technology | Low-intensive (n = 15) | Semiextensive (n = 30) |
|--------------------------|-------------------|-----------------------|------------------------|------------------------|
| Gross calorific value (kJ/100 g) | x 182            |                       | 109                    | 116                    |
| Gross calorific value (kJ/100 g) | x 764            |                       | 458                    | 487                    |
| Net calorific value (kJ/100 g)      | x 102            |                       | 88                     | 91                     |
| Net calorific value (kJ/100 g)      | x 428            |                       | 370                    | 382                    |
| Dry matter (%)            | x 24.6a          |                       | 22.3b                  | 21.98b                 |
| Total protein (%)         | x 18.74a         |                       | 18.32a                 | 17.94a                 |
| Total fat (%)             | x 1.58a          |                       | 0.56b                  | 1.49a                  |
| Total ash (%)             | x 1.01a          |                       | 1.11a                  | 1.02a                  |

**Table 1:** Calorific value and basic chemical composition in muscle tissue of common carp samples under study.

The same letters in a row denote the absence of significant statistical differences between mean values at $p \leq 0.05$. Different letters in a row denote the presence of significant statistical differences between mean values at $p \leq 0.05$. 

fatty acids: C18:4 n-3, C20:1 n-9, C20:1 n-7, C20:3 n-3, C20:4 n-3, and C20:5 n-6. Not many significant differences were found between the fatty acid contents in common carp muscle tissue from the two extreme types of fish farms: intensive and semiextensive. Only for C18:2 n-6 or C20:1 n-9 acids were no statistically significant differences found. Only two cases with no statistically significant differences were identified in an analysis of common carp muscle tissue by the low-intensive and semiextensive method. Those were the content levels of fatty acids C20:2 n-9 and C22:6 n-3 (Table 2). It can be concluded from the results of the statistical analysis of the fatty acid profile of fish muscle tissue from three types of fish farms that the type of feed has a great effect on its composition. The contents of various profile components vary from one fish type to another. There are some other cases with no differences between them, but only on the farms where intensive and low-intensive production technology was applied. This arises from the similarity of feed used in these fish ponds and the small number of results with no statistically significant differences between farms: I-SE, LE-SE, which clearly shows the differences in the nutritional value of the feed and translates into the material composition. Regardless of the type of common carp production technology, the fish fat contained saturated and unsaturated fatty acids at a beneficial ratio (Figure 1). The ratio was 0.84 in common carp caught on intensive farms, 0.74 in the low-intensive ones, and 0.87 in semiextensive ones. An examination of the fatty acid profile in common carp muscles has shown that they are not only a source of PUFA n-3 and PUFA n-6; the values of ω3 and ω6 are considerably lower than for other animal fats (Figure 2). For example, the atherogenic index for cow milk is ca. 3−4. The values for fish muscle tissue are 0.43, 0.54, and 0.35 for intensive, low-intensive, and semiextensive technology, respectively. For the samples of muscles under study, the values were 0.46, 0.65, and 0.84 for intensive, low-intensive, and semiextensive breeding, respectively.
Table 2: Fatty acid profile in muscle tissue of common carp under study (%).

| Fatty acids | Intensive (n = 30) | Low-intensive (n = 15) | Semiextensive (n = 30) |
|-------------|--------------------|------------------------|------------------------|
| Saturated fatty acids |                     |                        |                        |
| C14:0       | 1.59<sup>a</sup>   | 1.44<sup>b</sup>      | 0.84<sup>c</sup>      |
| C15:0       | 0.3<sup>a</sup>     | 0.22<sup>b</sup>      | 0.1<sup>b</sup>       |
| C16:0       | 19.02<sup>a</sup>   | 21.65<sup>b</sup>     | 18.3<sup>c</sup>      |
| C17:0       | 0.28<sup>a</sup>    | 0.19<sup>b</sup>      | 0.13<sup>c</sup>      |
| C18:0       | 5.05<sup>a</sup>    | 5.58<sup>b</sup>      | 6.40<sup>c</sup>      |
| C20:0       | 0.11<sup>a</sup>    | 0.08<sup>b</sup>      | 0.11<sup>a</sup>      |
| C22:0       | 0.19<sup>a</sup>    | 0.03<sup>b</sup>      | 0<sup>c</sup>         |
| Total ratio UFA | 26.54            | 29.19                  | 25.88                  |
| Monounsaturated fatty acids |                 |                        |                        |
| C14:1       | 0.09<sup>a</sup>   | 0.08<sup>b</sup>      | 0.04<sup>c</sup>      |
| C16:1 n-7   | 8.65<sup>a</sup>   | 10.29<sup>b</sup>     | 8.17<sup>c</sup>      |
| C17:1       | 0.37<sup>a</sup>   | 0.25<sup>b</sup>      | 0.15<sup>c</sup>      |
| C18:1 n-9   | 33.87<sup>a</sup>  | 39.99<sup>b</sup>     | 45.61<sup>c</sup>     |
| C18:1 n-7   | 3.55<sup>a</sup>   | 2.96<sup>b</sup>      | 2.52<sup>c</sup>      |
| C20:1 n-11  | 0.38<sup>a</sup>   | 0.32<sup>b</sup>      | 0.16<sup>b</sup>      |
| C20:1 n-9   | 2.28<sup>a</sup>   | 2.1<sup>a</sup>       | 2.24<sup>b</sup>      |
| C20:1 n-7   | 0.10<sup>a</sup>   | 0.09<sup>b</sup>      | 0.05<sup>b</sup>      |
| C22:1 n-11  | 0.37<sup>a</sup>   | 0.09<sup>b</sup>      | 0.06<sup>c</sup>      |
| C22:1 n-9   | 0.14<sup>a</sup>   | 0.07<sup>b</sup>      | 0.05<sup>c</sup>      |
| Total ratio MUFA | 49.8             | 56.24                  | 59.05                  |
| Polyunsaturated fatty acids |                |                        |                        |
| C18:3 n-3   | 3.03<sup>a</sup>   | 2.06<sup>b</sup>      | 0.71<sup>c</sup>      |
| C18:4 n-3   | 0.73<sup>a</sup>   | 0.52<sup>a</sup>      | 0.14<sup>b</sup>      |
| C20:3 n-3   | 0.15<sup>a</sup>   | 0.14<sup>a</sup>      | 0.07<sup>b</sup>      |
| C20:4 n-3   | 0.36<sup>a</sup>   | 0.28<sup>a</sup>      | 0.14<sup>b</sup>      |
| C20:5 n-3   | 2.59<sup>a</sup>   | 1.34<sup>b</sup>      | 0.73<sup>c</sup>      |
| C22:5 n-3   | 0.85<sup>a</sup>   | 0.38<sup>b</sup>      | 0.27<sup>c</sup>      |
| C22:6 n-3   | 2.20<sup>a</sup>   | 0.96<sup>b</sup>      | 0.84<sup>b</sup>      |
| Total ratio n-3 | 9.91             | 5.68                   | 2.9                   |
| C18:2 n-6   | 10.59<sup>a</sup>  | 7.13<sup>b</sup>      | 9.92<sup>a</sup>      |
| C18:3 n-6   | 0.32<sup>a</sup>   | 0.24<sup>a</sup>      | 0.37<sup>c</sup>      |
| C20:3 n-6   | 0.44<sup>a</sup>   | 0.25<sup>b</sup>      | 0.33<sup>c</sup>      |
| C20:4 n-6   | 1.55<sup>a</sup>   | 0.85<sup>b</sup>      | 1.06<sup>c</sup>      |
| C22:5 n-6   | 0.14<sup>a</sup>   | 0.14<sup>a</sup>      | 0.22<sup>b</sup>      |
| Total ratio n-6 | 13.04            | 8.61                   | 11.09                 |
| Ratio n-3/n-6 | 0.74             | 0.66                   | 0.26                  |
| C20:2 n-9   | 0.41<sup>a</sup>   | 0.29<sup>b</sup>      | 0.28<sup>b</sup>      |
| Total ratio PUFA | 23.36           | 14.58                  | 15.08                 |

<sup>ab</sup>The same letters in a row denote the absence of significant statistical differences between mean values at \( p \leq 0.05 \). <sup>abc</sup>Different letters in a row denote the presence of significant statistical differences between mean values at \( p \leq 0.05 \).

4. Discussion

The muscle tissue of common carp fed exclusively with cereal mixtures or fishing bait (i.e., food of a simple composition, without being enriched with animal protein or fat, therefore having a lower nutritional value) contained the smallest percentage of dry matter and protein. The percentage of protein in all carcasses did not deviate from its mean content in the muscle tissue of other fish species (13% to 25%) [21]. It is noteworthy that the protein of common carp, regardless of the production technology, contains essential amino acids in amounts much higher than in the standard protein [22]. This protein contains a higher percentage of amino acids such as phenylalanine, leucine, isoleucine, threonine, methionine, cystine, and valine, compared with beef, pork, or mutton [21, 23]. From a nutritional point of view, the protein of
this fish species is regarded as complete. The composition of rainbow trout muscle tissue (another freshwater fish) is similar. A deficit of essential amino acids in the human diet has been shown to result in many adverse changes in the body. A sufficient supply of valine helps to maintain the right coordination of movements, the right body weight, function of dendritic cells, and a feeling of hunger. A deficit of leucine can lead to neurological disorders and a deficit of methionine impairs body immunity [24].

It has been shown in a number of studies that the type of feed given to fish affects the chemical composition of their muscle tissue. The fatty acid profile, affected by the type of feed, is very important in human nutrition. Unlike the meat of slaughter animals, fish fat is also a rich source of vitamins A and D [24]. Like marine fish, freshwater fish can be a rich source of essential fatty acids; when included in the diet, they supply the body with cardioprotective, hypotensive, and antitumour substances [11, 25]. The availability of omega-3 fatty acids for different human tissues depends on the diet and is of great importance both for their correct development and for prevention and treatment of chronic diseases [26]. Similar observations were included in the paper by Grela et al. [27], who analysed the composition of marine and freshwater fish muscle tissue. Their muscle tissue was found to contain the highest percentage of C16:0 palmitic (ca. 20%) and C18:0 stearic acid, although the content of the latter in the samples was much lower. Similarly, the highest percentage of palmitic acid in the fatty acid profile was found in muscle tissue of other freshwater fish of South Asia, rohu (Labeo rohita) of the Cyprinidae family and Mozambique tilapia (Oreochromis mossambicus) [28], as well as muscle tissue of common carp from Lake Beyşehir in Turkey and rainbow trout (15–18%) bred in Poland [24], which indicates that this fatty acid dominates the MUFA profile in fat of freshwater fish regardless of the climate and season [29]. Monounsaturated fatty acids reduce the level of “bad” cholesterol (LDL) and increase the level of “good” cholesterol (HDL) in blood [30, 31]. The same results were obtained in a study of the common carp muscle tissue composition conducted by Guler et al. [29]. They also demonstrated seasonal variability of the content of total fat in fish carcasses, with the highest content observed in winter (4.45%).

The preparation given to the fish contained 56% of protein and 11% of lipids. It is important to be able to model the lipid profile of food because fats are the main high-energy nutrient. It has been shown in numerous studies that high consumption of fat and its improper composition can stimulate the development of civilizational diseases. The fatty acid profile, especially the proportion of saturated to mono- and polyunsaturated acids, is of particular importance. Polyunsaturated essential omega-3 and omega-6 fatty acids are not synthesised in the human body and they must be supplied with food [32, 33]. The group’s main representative ones include α-linolenic acid (C18:3) of the n-3 family; it is a precursor of C20:5 n-3 eicosapentaenoic (EPA) and 22:6 n-3 docosahexaenoic acid (DHA) and linoleic acid (C18:2 n-6), a precursor of C20:4 n-6 arachidonic acid (AA) [34]. Modelling of PUFA content of the muscle tissue of freshwater fish is of interest to numerous researchers. A similar experiment was conducted in the Czech Republic by Mraz and Pickova (2011) [35] who also analysed three systems of nutrition. In the first one, the fish had access to plankton; in the second one, the fish had access to plankton with the addition of cereals and rapeseed cake granules. The fatty acid profile in the white muscles of fish was then analysed. The muscles of fish which were not provided with supplements were characterised by a high content of n-3 PUFAs (in particular EPA and DHA). The supplementation with rapeseed cake granules resulted in the PUFA n-3 content being at a moderate level. The addition of cereals led to a high
content of oleic acid and a low content of n-3 polyunsaturated fatty acids.

Similar literature reports have also pointed to freshwater fish as a valuable source of omega-3 fatty acids and emphasised their significant role in the daily diet [36, 37]. Another method for modifying the composition of fatty acids is the application of the finishing feeding strategy. Since the composition of fish muscles is highly variable, it would be valuable if producers could produce raw material with a high and repeatable level of n-3 polyunsaturated fatty acid content. The difficulties primarily encountered by fish breeders who intend to enrich the feed with components providing n-3 PUFAs include the increasingly rare use of fish oil as well as a high price and low availability of algae and microorganisms. For this reason, the possibility is being considered of the application of finishing feeding developed with the following in mind: species of carnivorous fish and those with a medium fat content, such as the turbot (Psetta maxima), fatty fish such as the Atlantic salmon (Salmo salar), and lean fish such as the Atlantic cod (Gadus morhua) and the Murray cod (Maccullochella peeli peeli) [38]. Supplementation serves a significant role in nutrition. The content of n-3 HUFA s (highly unsaturated fatty acids) can be increased with the supplementation of ALA, taking advantage of the common carp's capacity for bioconversion of ALA into n-3 HUFA s [35]. An effect of feed modification on the fatty acid profile was studied by Menoyo et al. [39], who examined how an addition or total substitution of feed with linseed oil will influence the quality of muscle tissue of Atlantic salmon. It has been shown that feed can be fully substituted with linseed oil, with no effect on the productivity or sensitivity of muscles to oxidation of lipids. Linseed oil affected the metabolism of fatty acids in the liver, increasing the activity of glucose-6-phosphate dehydrogenase and accumulation of C20:4 n-6 eicosatetraenoic acid. This was accompanied by a decrease in the concentrations of C20:5 n-3 eicosapentaenoic and C22:6 n-3 docosahexaenoic acid in fish muscle tissue. According to the analysis of factors affecting the distribution of fatty acids within the muscle tissue of the common carp, carried out by Mrzaz and Pickova (2011) [35], a fish fillet is not uniform, and thus the distribution of fatty acids in the muscles varies. The highest lipid content was noted in the abdominal wall (approx. 30%) and in the red muscles (16-17%), while the lowest was in the white dorsal muscles (approx. 1-2%). The role of genetic factors must not be omitted. It was demonstrated that the fatty acid content of the muscle tissue is hereditary; moreover, there is a positive correlation between the body size (body length and weight) and the lipid content [40]. Consideration of the effect of external factors on the quality of fish muscle tissue should also take into account the climate. Çelik et al. [41] examined the composition of muscle tissue of zander (Sander lucioperca) and showed that a higher percentage of omega-3 fatty acids were present in muscle tissue of fish bred in a colder climate. Furthermore, Çağlak and Karşı [42] showed seasonal influences not only on the lipid index of the edible portion of zander, but also on higher content of aspartic acid, glutamic acid, and lysine in muscle tissue of these fish in autumn than in spring. Similar results were presented in a study conducted by Guler et al. [29], where a significantly higher omega-3 to omega-6 acid ratio was shown during a spawning period as well as in spring and in autumn. Pleadin et al. [43] investigated the impact of seasonal changes and the location of breeding farms in the Adriatic Sea on the chemical composition of sea brass (Dicentrarchus labrax) and sea bream (Sparus aurata). The study was conducted in October 2012 and January 2013. They showed a significant influence of seasonality on moisture content and fat content in fish muscle tissue, while the location of farms did not significantly influence the indicators. The value of these relations is similar to other species of fish such as S. scholl or T. lineatus, where PUFA/SFA ratios are 0.4 and 1.7, respectively. The lower values of the ratio of polyunsaturated to saturated fatty acids are characterised by muscle tissue of species such as L. niloticus, B. bajad, and O. niloticus [44].

There are two dietary indexes associated with the lipid profile: the atherogenic index and the thrombogenic index. Atherogenesis denotes the development of atherosclerotic changes in blood vessels, which result in the development of ischaemic heart disease. A negative lipid ratio in blood speeds up atherogenesis. Thrombogenic components denote particles which facilitate the formation of blood clots. Edible fats are classified into two types: atherogenic (i.e., those that favour the development of atherosclerotic changes) and antiatherogenic (which have an antiatherosclerotic effect). Most animal fats have an atherogenic effect due to the high content of saturated fatty acids and cholesterol. The other group includes mainly vegetable oils [45]. This means that the fat has much higher antiatherosclerotic and cardioprotective properties than milk fat and emphasises its pro-health value. The thrombogenic index, which for milk ranges from 3.75 to 4.71, is another parameter which indicates the beneficial properties of common carp fat [46]. The nutritional value of the common carps tested, given indicators such as the thrombogenicity index and the atherogenic index, was very favourable, as was the case with commercially important species of marine fish such as bream (Spa rus aurata), sea bass (Dicentrarchus labrax), dentex (Dentex dentex), and turbot (Scophthalmus maximus) [47].

The findings suggest that the muscle tissue of common carp and other freshwater fish can be a valuable replacement for marine fish in the diet. It is a rich source of PUFA in desired proportions. Moreover, there is a distinct, beneficial effect of intensive technology of fish breeding on selected parameters of lipid profile, the amount of components with the cardioprotective action, total amount of fat, and the associated content of fat-soluble vitamins. The differences in results for samples obtained from various types of farms arise from the type of feed given to the animals. The feed given in the semintensive technology was a cereal mixture: wheat, barley, and rye in the ratio of 3:1:1, or only fishing bait; in the low-intensive technology it was a cereal mixture—wheat, barley, and rye in the ratio of 2:1:1—or a cereal mixture—maize + wheat and barley in the ratio of 1:3. The fish were given industrial feeds in the intensive production technology; these were made of the following ingredients: yeast, wheat gluten, fish meal, krill meal, fish oil, wheat, vitamins, minerals, and amino acids.
Their composition was much more diverse; it contained both complete animal protein and plant protein. An addition of fish oil in the right proportions, especially saturated fatty acids, allows for modification of the composition of fatty acids in the bodies of fed animals. Yeasts are used in animal feeding as a source of vitamin B, some bioelements, enzymes, and digestible protein. They have a beneficial effect on intestinal flora and stimulate the growth and development of young animals and their health and productivity [48].

5. Conclusions

(1) The type of production technology, especially intensive breeding technique, has a significant effect on dry weight and total fat in the common carp muscle tissue under study.

(2) The type of feed used in intensive, low-intensive, and semiextensive common carp breeding has a highly significant effect on the fatty acid profile. Regardless of the technology of common carp breeding, fish fat has a beneficial proportion of unsaturated (UFA) to saturated (SFA) fatty acids and is a rich source of polyunsaturated fatty acids (PUFAs).

(3) Low values of atherogenic index (AI) and thrombogenic index (TI) of common carp fat indicate its antiatherosclerotic and cardioprotective properties, which are much stronger than in milk fat. This confirms the pro-health properties of fat in common carp muscle tissue.

(4) Owing to its nutritional and pro-health value, muscle tissue of common carp, especially from intensive breeding, can be a complete substitute for marine fish in the human diet.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Acknowledgments

The authors would like to express their gratitude to Dr. Janusz Zakrzewski for help in conducting the experiment and obtaining the results necessary to write this publication.

References

[1] P. Skalecki, M. Florek, A. Litwinczuk et al., “Wartość użytkowa i skład chemiczny mięsa karpi (Cyprinus common carpio L.) i pstrągów tczewnych (Onchorhynchus mykiss Walb.) pozyskanych z gospodarstw rybacych regionu lubelskiego,” Roczniki Naukowe Towarzystwa Zootchnicznego, vol. 9, no. 2, pp. 57–62, 2013.

[2] K. Achemrowicz and K. Szary-Sworst, “Wielonienasycone kwasyltuszczowe czynniaki poprawy stanu zdrowia człowieka,” Żywnosc. Nauka. Technologia. Jakość, vol. 3.44, pp. 23–35, 2005.

[3] K. Bieniarz, F. Borowiec, and Z. Okoniewski, “Zawartość tłuszczu, kwasów tłuszczowych i cholesterolu w mięsie karpi (Cyprinus common carpio L.) chowanych w różnych warunkach pokarmowych,” Roczniki Naukowe Zootechniki, Suplement, vol. 12, pp. 129–135, 2001.

[4] W. Steffens, “Aquaculture produces wholesome food: cultured fish as a valuable source of n-3 fatty acids,” Aquaculture International, vol. 24, no. 3, pp. 787–802, 2016.

[5] J. Ciborska, “Lipidy w żywności, żywieniu i zdrowiu człowieka Cz. Il. Aspekty żywieniowe i zdrowotne,” PRZEMYSŁ SPOŻYWACZY, vol. 1, no. 9, pp. 46–51, 2017.

[6] K. A. Skibniewska and J. Zakrzewski, “Technologia produkcji rybackiej ajakosć karpia. Wpływ rodzaju technologii produkcji rybacyjnej i jakości środowiska wodnego na wybrane wskazniki hodowlane i patomorfologiczne karpia konsumpcyjnego (Cyprinus common carpio L.),” Olsztyn, Wydanie w ramach projektu Sektorowego Programu Operacyjnego „Rybołówstwo i Przetwórstwo Ryb 2004–2006” współfinansowanego przez Unię Europejską, pp. 65–72, 2008 (Portuguese).

[7] A. Sánchez-Villegas, L. Verberne, J. De Iraola et al., “Dietary Fat Intake and the Risk of Depression: The SUN Project,” PLoS ONE, vol. 6, no. 1, p. e16268, 2011.

[8] A. P. DeFilippis, M. J. Blaha, and T. A. Jacobson, “Omega-3 fatty acids for cardiovascular disease prevention,” Current Treatment Options in Cardiovascular Medicine, vol. 12, no. 4, pp. 356–380, 2010.

[9] J. Tkaczewska and W. Migdał, “Porównanie wydajności rzeźnej, zawartości podstawowych składników odżywczych oraz poziomu metali ciężkich w mięśniach karpi (Cyprinus common carpio L.) pochodzących z różnych rejonów Polski,” Żywność. Nauka. Technologia. Jakość, vol. 6, no. 85, pp. 180–189, 2012.

[10] R. Puchala, M. Pilarczyk, and R. Puchala, “Wpływ żywienia na skład chemiczny mięsa karpia,” Inżynieria Rolnicza, vol. 5, no. 93, pp. 363–368, 2007.

[11] W. Steffens and M. Wirth, “Influence of nutrition on the lipid quality of pond fish: Common carp (Cyprinus carpio) and tench (Tinca tinca),” Aquaculture International, vol. 15, no. 3-4, pp. 313–319, 2007.

[12] V. Adamkova, J. Mraz, P. Kacer, and P. Suchanek, “The consumption of common carp meat and plasma lipids in secondary prevention in the heart ischemic disease patients,” Neurope-docinol. Lett., vol. Lett. 32, no. Suppl. 2, pp. 101–104, 2011.

[13] K. J. Filipiak and G. Opolski, Epidemiologiczne aspekty zaburzeń lipidowych oraz terapii tych zaburzeń w Polsce. W Zaburzenia lipidowe, 51-64. Poznań: Termidea Wydawnictwa Medyczne, 51-64. Poznań, Termidea Wydawnictwa Medyczne, 2010.

[14] AOAC, Official Methods of Analysis, Association of Official Analytical Chemists, Washington, DC, USA, 18th edition, 2005.

[15] AOAC, Official Methods of Analysis, vol. Gaithersburg, MD, USA., Gaithersburg, MD, USA., 17th ed edition, 2000.

[16] J. Folch, M. Lees, and G. H. Sloane Stanley, “A simple method for the isolation and purification of total lipides from animal tissues,” The Journal of Biological Chemistry, vol. 226, no. 1, pp. 497–509, 1957.

[17] AOAC, Official Methods of Analysis., method 996.06; Fat (Total, Saturated, Unsaturated, and Monounsaturated), VA, USA, 17th edition, 2002.

[18] Z. Zegarska, J. Jaworski, Z. Borejszo, and Z. Zegarska, “Ocena zmodyfikowanej metody Peiskera otrzymywania estrow metyloowych kwasów tłuszczowych," Acta Academiae Agriculturae ac Technicae Olstenensis, Technologia Alimentorum, 1991.

[19] J. Gawęcki, “Żywienie człowieka. Podstawy nauki o żywieniu,” Wydawnictwo Naukowe PWN,” 2016.

[20] T. L. V. Ulbricht and D. A. T. Southgate, “Coronary heart disease: seven dietary factors,” The Lancet, vol. 338, no. 8773, pp. 985–992, 1991.

[21] V. V. Vladau, I. Bud, and S. Reka, “Nutritive value of fish meat comparative to some animals meat,” in Bulletin of the University
of Agricultural Sciences & Veterinary Medicine Cluj-Napoca, vol. 65, pp. 301–305, Animal Science & Biotechnologies, 2008.

[22] WHO/FAO/UNU, “Protein and amino acid requirements in human nutrition,” Report of joint WHO/FAO/ UNU expert consultation, WHO, Geneva, Switzerland, 2002, WHO Technical Report, Series 935.

[23] K. A. Skibniewska, J. Zakrzewski, J. Klobukowski et al., “Nutritional value of the protein of consumer carp cyprinus carpio L.,” Czech Journal of Food Science, vol. 31, no. 4, pp. 313–317, 2013.

[24] J. Szares, K. A. Skibniewska, J. Zakrzewski, and J. Gaziur, The quality of rainbow trout (Oncorhynchus mykissWalbaum 1972) from technologies applied in Poland. Testing the trout production technologies applied in Poland in the light of the Commission Regulation (WE) 710/2009, ElSet, Olsztyn, Poland, 2013.

[25] A. Philibert, C. Vanier, N. Abdelouahab, H. M. Chan, and K. A. Skibniewska, J. Zakrzewski, J. Kłobukowskiet al., “Nutritional quality of common commercial Nile fish of different species farmed in the Adriatic Sea,” International Journal of Fishes and Aquaculture, vol. 3, no. 6, pp. 99–104, 2011.

[26] G. O. Guler, B. Kiztanir, A. Aktumsek, O. B. Citil, and H. E. R. Grela, R. K. Pisarski, E. Kowalczuk-Vasilev, and A. Craciunescu, “A comparison of dietary fatty acids and carbohydrates on the ratio of serum total to HDL cholesterol and on serum lipids and apolipoproteins: a meta-analysis of 60 controlled trials,” American Journal of Clinical Nutrition, vol. 77, no. 5, pp. 1146–1155, 2003.

[27] R. P. Mensink, P. L. Zock, A. D. M. Kester, and M. B. Katan, “Effects of dietary fatty acids and carbohydrates on the ratio of serum total to HDL cholesterol and on serum lipids and apolipoproteins: a meta-analysis of 60 controlled trials,” American Journal of Clinical Nutrition, vol. 77, no. 5, pp. 1146–1155, 2003.

[28] C. M. Williams, “Dietary fatty acids and human health,” Annales de Zootechnie, vol. 49, no. 3, pp. 165–180, 2000.

[29] R. J. Nicolosi, “Dietary fat saturation effects on low-density-lipoprotein concentrations and metabolism in various animal models,” American Journal of Clinical Nutrition, vol. 65, no. 5, 1997.

[30] S. Yu, J. Derr, T. D. Etherton, and P. M. Kris-Etherton, “Plasma cholesterol-predictive equations demonstrate that stearic acid is neutral and monounsaturated fatty acids are hypocholesterolemic,” American Journal of Clinical Nutrition, vol. 61, no. 5, pp. 1129–1139, 1995.

[31] D. Ljubojević, V. Radosavljević, N. Puvača et al., “Interactive effects of dietary protein level and oil source on proximate composition and fatty acid composition in common carp (Cyprinus carpio L.),” Journal of Food Composition and Analysis, vol. 37, pp. 44–50, 2015.

[32] J. Mráz and J. Pickova, “Factors influencing fatty acid composition of common carp (Cyprinus carpio) muscle,” Neuroendocrinology, vol. 32, no. 2, pp. 3–8, 2011.

[33] S. Bulut, “Fatty acid composition and ω6/ω3 ratio of the pike (Esox lucius) muscle living in Eber Lake, Turkey,” Scientific Research and Essays, vol. 5, no. 23, pp. 3776–3780, 2010.
