Uterine transplantation in transgender women

BP Jones, a,b NJ Williams, c S Saso, a,b M-Y Thum, b,d I Quiroga, e J Yazbek, a,b S Wilkinson, c S Ghaem-Maghami, a,b P Thomas, f JR Smith a,b

a West London Gynaecological Cancer Centre, Hammersmith Hospital, Imperial College NHS Trust, London, UK b Department of Surgery and Cancer, Imperial College London, London, UK c Department of Politics, Philosophy and Religion, Lancaster University, Lancaster, UK d Lister Fertility Clinic, The Lister Hospital, London, UK e The Oxford Transplant Centre, The Churchill Hospital, Oxford University Hospitals NHS Trust, Oxford, UK f Brighton Gender Clinic, Nuffield Health Hospital, Brighton, UK

Correspondence: Mr BP Jones, Department of Surgery and Cancer, Imperial College London, Du Cane Road, London W12 0NN, UK.
Email: Benjamin.jones@nhs.net

Accepted 12 August 2018. Published Online 20 September 2018.

Please cite this paper as: Jones BP, Williams NJ, Saso S, Thum M-Y, Quiroga I, Yazbek J, Wilkinson S, Ghaem-Maghami S, Thomas P, Smith JR. Uterine transplantation in transgender women. BJOG 2019;126:152–156.

Linked article This article is commented on by N Hammond-Browning, p. 157 in this issue. To view this mini commentary visit https://doi.org/10.1111/1471-0528.15482.

Introduction

Gender dysphoria, defined as the persistent discomfort with one’s gender identity or biological sex, affects between 0.5 and 1.4% of adult males. Treatment aims at congruence, to allow those who experience it to find comfort within their gendered self, which optimises psychological wellbeing and self-fulfilment. Although many experiencing gender dysphoria require partial treatment or social transition, others only find comfort following surgical intervention to change their external genitalia and sexual characteristics. Traditionally, infertility has been an unfortunate consequence of the realignment of a transgender person’s body with their gender identity.

Following a successful clinical trial investigating uterine transplantation (UTx) in Sweden, resulting in eight live births so far, UTx appears to be a viable therapeutic option for women with absolute uterine factor infertility (AUFi). More than 42 UTx procedures have now been performed globally, and at least 12 live births have been reported. Following the establishment of the International Society of Uterine Transplantation (ISUTx), and the formation of research teams globally, it is anticipated that UTx will make the transition from research to clinical care in the future.

Anatomical considerations

Uterine transplantation entails the transplantation of the uterus, including the cervix, a cuff of vagina, the surrounding ligamentous and connective tissues, as well as the major blood vessels to the level of the internal iliac vessels. The uterus is then placed orthotopically in the pelvis of the recipient, where it is structurally supported using the uterosacral, round and broad ligaments laterally, the bladder peritoneum anteriorly, and the vagina and paravaginal tissues inferiorly.
Vascular anastomosis
Most UTx procedures performed to date have achieved revascularisation through bilateral internal iliac artery to external iliac end-to-side anastomoses. To determine the plausibility of retrieving a graft from a woman, with subsequent implantation into a natal male pelvis, the intersex differences in pelvic vascular anatomy require consideration. Fätu et al. assessed the morphometry of the internal iliac arteries between sexes, and concluded there was no difference in internal iliac artery length, with a mean length of 49 mm. However, the calibre of the vessels was noted to be 1.6 mm wider in females than in males (6.2 versus 7.8 mm). Although a significant funnelling effect could predispose to thrombosis, this difference could be negated by anastomosing further proximally, where the vessels are similarly sized. With regard to the external iliac arteries, data from lower limb angioplasties show there is no significant difference in external iliac artery calibre between sexes.

Vaginal anastomosis
In all UTx cases performed to date, the recipient’s vagina has been anastomosed to a vaginal cuff, of varying length, which is retrieved as part of the graft. This restores reproductive anatomy and allows physiological excretion of discharge and menstruation, but also allows direct visualisation of the cervix and access to take biopsies, which is the only reliable way to detect rejection following UTx. In the M2F transgender model, it would therefore only be possible following gender reassignment surgery (GRS), which traditionally includes orchidectomy, penectomy, clitoroplasty, and labiaplasty, with the subsequent creation of a neovagina. The inverted penile skin flap is the standard technique for neovagina creation, to line a newly created space between the bladder/prostate and the rectum. However intestine, or pelvic peritoneum, have also been utilised, particularly in cases with penoscrotal hypoplasia, which can be an iatrogenic consequence of feminising hormones. However, the absence of a physiologically functioning vaginal mucosa may be problematic. The vagina is lined by multiple layers of stratified squamous epithelium, the top layer of which removes adherent micro-organisms by desquamation into the vaginal lumen. Vaginal epithelium also facilitates the recognition of pathogens and stimulates production of antimicrobial peptides and pro-inflammatory cytokines. These protective mechanisms contribute to the creation of a commensal microflora, predominantly consisting of lactobacilli, which provides an optimal physiological environment to prevent infection and maintain pregnancy. In M2F transgender women, the pH in penile skin-lined neovaginas is elevated, owing to an inability to support the growth of acidic lactobacilli, with colonisation of bacteria from skin or intestinal microfloras instead. Following M2F transgender UTx, the presence of a skin or intestinal neovagina, in the context of immunosuppression, may increase susceptibility to recurrent neovaginal infections and create a hostile environment that may be incapable of sustaining pregnancy. This was exemplified in the UTx case performed in Turkey in a recipient with an intestinal neovagina. Despite multiple embryo transfers and at least six early pregnancy miscarriages, she has yet to achieve a live birth. Moreover, the only woman in the Swedish series to have not yet given birth following successful UTx, despite suffering at least five miscarriages, has a skin neovagina. Although it appears the absence of a physiologically functioning vagina is detrimental, albeit to a currently unquantified extent, small numbers of live births have been reported in women with skin neovaginas, including two following UTx in the Swedish series, highlighting that successful pregnancy is possible.

To overcome this anatomical hindrance in M2F transgender women, a utero-vaginal transplant could be performed, utilising as much donor vagina as possible, en-bloc with the uterus (Figure 1). This would be achievable using a similar technique to that employed at radical hysterectomy, with
preservation of the vaginal branches of the uterine vessels. This would necessitate retrieval from deceased donors, excluding female living donors. An alternative donor pool may be female to male (F2M) transgender men undergoing hysterectomy and vaginectomy, although the increased radicality of the hysterectomy may not be acceptable. However, recent evolution of the surgical technique, following cases in China and Dallas, has potentially significantly reduced donor risk. The modified technique utilises venous drainage of the graft via the ovarian or utero-ovarian veins, as opposed to the unpredictable and tortuous uterine venous plexus which leads to the internal iliac veins. This negates the need for the complex and time-consuming ureteric dissection away from the uterine veins, reducing surgical risk. Moreover, it reduces operative times from 12 hours to 4–5 hours, which in turn decreases potential venous thromboembolism (VTE) risk. This dissection also favours minimally invasive retrieval techniques, which should enhance recovery and reduce potential morbidity further. As most F2M transgender men will also undergo bilateral oophorectomy, whereas the ovaries it would not be transplanted, it would allow the retrieval of elongated ovarian vascular pedicles to facilitate the implantation.

The graft for implantation, using deceased donors, is displayed in Figure 1. This would be anastomosed to the recipient’s neovagina as shown in Figure 2. Although prostatectomy is not routinely undertaken in GRS, the oestrogenised environment in transgender women causes prostatic atrophy, which should not cause a structural hindrance in UTx.

**Ligamentous support**

The ligamentous and soft tissue support is provided anteriorly by approximating donor and recipient bladder peritoneum. This technique is directly transferable to the M2F transgender model. Postero-laterally, the broad and uterosacral ligaments from the donor are connected to the pelvic side wall and uterosacral remnants, respectively, in the recipient. Although M2F transgender women do not have uterosacral remnants, this could be overcome by a more radical ligamentous retrieval, with subsequent anastomosis to the recipient paraneovaginal region (Figure 2).

**Hormonal factors**

Exogenous estrogen optimises the development of female secondary sexual characteristics, whereas anti-androgens, such as spironolactone or finasteride, minimise male features. Progestogen is not routinely administered, as its role in feminisation remains unclear. However, considering that unopposed estrogen is a significant risk factor for endometrial hyperplasia and endometrial carcinoma in post-menopausal women, progestogen supplementation would become essential in transgender women following UTx.

Continuous hormone replacement therapy (HRT) is the usual regimen prescribed in M2F transgender women, but sequential HRT would be more appropriate following UTx in transgender women. Not only is withdrawal bleeding an important sign of graft function, but it is intrinsically part of being female and therefore contributes to gender identity, which may have psychological benefits.

Graft thrombosis is one of the most common serious complications following organ transplantation. Although oral estrogen was previously implicated in an increased VTE risk in M2F transgender women, this was later attributed to the use of ethinylestradiol, a particularly thrombogenic estrogen that is no longer in routine use. A subsequent study on 2236 M2F transgender individuals reaffirmed this, with no additional risk of VTE seen in those receiving different hormone therapy.

**Fertility considerations**

Fertility preservation should be discussed in all M2F cases prior to the commencement of hormone therapy or contemplation of GRS. M2F transgender women can preserve their...
fertility prior to transition using sperm cryopreservation, with subsequent in vitro fertilisation (IVF) or intratruterine insemination (IUI) in a female partner or surrogate.

Following UTx, embryo transfer should not be attempted until at least 6 months postoperatively, to allow healing and stabilisation of immunosuppression. Achieving pregnancy may be feasible utilising hormone regimens that have been used with success in women with premature ovarian insufficiency or following physiological menopause. Following the withdrawal bleed on sequential combined HRT, estrogen supplementation should be commenced to stimulate the endometrium. Once >7 mm in thickness, progesterone should be supplemented to maintain the endometrial lining for implantation. A single embryo can then be transferred into the uterus. Multiple embryo transfers should be avoided owing to the additional risks associated with multiple gestations. All women should have previously undergone orchidectomy, with resultant low testosterone levels. However, if anti-androgens such as finasteride or spironolactone are being taken, these should also be stopped in advance of fertility treatment, owing to their teratogenic potential.

**Obstetric considerations**

Male pelvices differ from their female counterparts, to an extent that they can be used to determine gender at autopsy. This dimorphism has evolved as a consequence to sex-specific selection pressures. Natal males need a pelvis suitable for bipedal locomotion, whereas the female pelvis must also accommodate a fetus during pregnancy and be adequately capacious for childbirth. Although most skeletal measurements are larger in males than females, the true pelvis of the female has evolved to become larger and broader. This dimorphism is most marked at the anteroposterior diameter of the pelvic inlet, the transverse diameter of the midplane between the ischial spines, and the transverse diameter of the pelvic outlet. Moreover, whereas the pelvic inlet is oval-shaped in females, it is heart-shaped in males. These intersex differences in pelvic morphology would predispose M2F females after UTx to cephalophevic disproportion should labour be attempted. However, as the requisite mode of delivery in women following UTx is caesarean section, owing to concerns regarding the mechanical strain of labour, this should also be the case in M2F transgender women.

Sexual dimorphism arises predominantly due to the outcome of gender-determined autosomal genes, which are regulated by sex-specific hormones and influenced by hormone receptor sensitivity. This dimorphism has been demonstrated in the pelvis, where despite similar growth patterns throughout childhood, it is not until puberty when the growth trajectory increases in females, and not until the late twenties when the pelvis attains the most favourable obstetric dimensions. As such, if M2F transgender women undergo hormone therapy at a young enough age, they may develop similar pelvic morphology to natal females. Although there is no evidence in the context of M2F transgender women, the opposite effect has been demonstrated in female to male (F2M) transgender men, where a biometric analysis of pelvic characteristics after the onset of hormone therapy revealed evidence of ‘masculinisation’.

**Conclusion**

Despite a number of anatomical, hormonal, fertility, and obstetric considerations that require consideration, there is no overwhelming clinical argument against performing UTx as part of GRS. However, the increased radicality associated with the retrieval operation, including a longer vaginal cuff and more extensive ligamentous dissection, potentially necessitates the use of deceased donors. Alternatively, F2M transgender men may offer an alternative donor pool should they accept the increased risk compared with standard hysterectomy. Prior to undertaking UTx in transgender women, further research should be undertaken including cadaveric retrieval and implantations to assess the feasibility of the anatomical considerations discussed herein. Furthermore, it is recommended that animal studies are revisited to identify potential unknown risks and determine whether genetic males can successfully conceive and maintain pregnancy.

The reproductive aspirations of M2F transgender women deserve equal consideration to those assigned female at birth and, subject to feasibility being shown in the suggested areas of research, it may be legally and ethically impermissible not to consider performing UTx in this population.

**Disclosure of interests**

None declared. Completed disclosure of interest forms are available to view online as supporting information.

**Contribution to authorship**

The article was conceived and written by BPJ. NJW helped write the article and reviewed the final draft. SS, MYT, IQ, JY, SW, SGM, PT, and JRS reviewed and contributed to the manuscript.

**Details of ethics approval**

Not applicable.

**Funding**

This work was supported by the Wellcome Trust [097897/Z/11/Z].

© 2018 The Authors. BJOG: An International Journal of Obstetrics and Gynaecology published by John Wiley & Sons Ltd on behalf of Royal College of Obstetricians and Gynaecologists
Acknowledgements
The authors would like to thank Dee McLean for the artwork in Figures 1 and 2.

References

1 American Psychiatric Association. Diagnostic and Statistical Manual of Mental Disorders, 5th edn. Arlington, VA: American Psychiatric Press; 2013.

2 Coleman E, Bockting W, Botzer M, Cohen-Kettenis P, De Cuypere G, Feldman J, et al. Standards of care for the health of transgender, transgender, and gender-nonconforming people version 7. Int J Transgend 2012;13:165–232.

3 Bränström M. Current status and future direction of uterus transplantation. Curr Opin Organ Transplant 2018;https://doi.org/10.1097/MOT.0000000000000568

4 Alghrani A. Uterus transplantation: does procreative liberty encompass a right to gestate?. J Law Biosci 2016;3:636–41.

5 Murphy TF. Assisted gestation and transgender women. Bioethics 2015;29:389–97.

6 Wilkinson S, Williams NJ. Public funding, social change, and uterus transplants. J Med Ethics 2016b;42:572–3.

7 Fätu C, Puigcorró M, Fätu IC. Morphometry of the internal iliac artery in different ethnic groups. Ann Anat 2006;188:541–6.

8 Timaran CH, Stevens SL, Freeman MB, Goldman MH. External iliac and common iliac artery angioplasty and stenting in men and women. J Vasc Surg 2001;34:440–6.

9 Mölne J, Broecker V, Ekberg J, Nilsson O, Dahn-Kåhler P, Bränström M. Monitoring of human uterus transplantation with cervical biopsies: a provisional scoring system for rejection. Am J Transplant 2017;17:1628–36.

10 Horbach SE, Bouman MB, Smit JM, Özer M, Buncamper ME, Mullender MG. Outcome of vaginoplasty in male-to-female transgenders: a systematic review of surgical techniques. J Sex Med 2015;12:1499–512.

11 Bouman MB, van der Sluis WB, Buncamper ME, Özer M, Mullender MG, Meijerink WJ. Primary total laparoscopic sigmoid vaginoplasty in transgender women with penoscrotal hypoplasia: a prospective cohort study of surgical outcomes and follow-up of 42 patients. Plast Reconstr Surg 2016;138:614e–23e.

12 Slater M, Vinaja X, Ally I, Loukas M, Terrell M, Schober J. Neovaginal construction with pelvic peritoneum: reviewing an old approach for a new application. Clin Anot 2018;31:175–80.

13 Patton DL, Thwin SS, Meier A, Hooton TM, Stapleton AR, Eschenbach DA. Epithelial cell layer thickness and immune cell populations in the normal human vagina at different stages of the menstrual cycle. Am J Obstet Gynecol 2000;183:541–6.

14 Anderson DJ, Marathe J, Pudney J. The structure of the human vaginal stratum corneum and its role in immune defense. Am J Reprod Immunol 2014;71:618–23.

15 Weyers S, Verstraeten H, Gerrets J, Monstrey S, Santiago Gidós S, Saaens B, et al. Microflora of the penile skin-lined neovagina of transsexual women. BMC Microbiol 2009;20:102.

16 Akar ME, Ozkan O, Aydinuraz B, Diirican K, Cincik M, Mendiciciglu I, et al. Clinical pregnancy after uterus transplantation. Fertil Steril 2013;100:1358–63.

17 Ozkan O, Dogan NU, Ozkan O, Mendiciciglu I, Dogan S, Aydinuraz B, et al. Uterus transplantation: from animal models through the first heart beating pregnancy to the first human live birth. Womens Health (Lond) 2016;12:442–9.

18 Bränström M. 1st Congress of International Society of Uterine Transplantation (ISUTx); 18–19 September 2017; Gothenberg.

19 Acipn M, Acici M, Quereda F, Santoyo T. Cervicovaginal agenesia: spontaneous gestation at term after previous reimplantation of the uterine corpus in a neovagina: case report. Hum Reprod 2008;23:548–53.

20 Wei L, Xue T, Tao KS, Zhang G, Zhao GY, Yu SQ, et al. Modified human uterus transplantation using ovarian veins for venous drainage: the first report of surgically successful robotic-assisted uterus procurement and follow-up for 12 months. Fertil Steril 2017;108:346–56 e1.

21 Testa G, McKenna GJ, Gunby RT Jr, Anthony T, Koon EC, Warren AM, et al. First live birth after uterus transplantation in the United States. Am J Transplant 2018;18:1270–4.

22 Jin B, Turner L, Walters WA, Handelsman DJ. The effects of chronic high dose androgen or estrogen treatment on the human prostate. J Clin Endocrinol Metab 1996;81:4290–5.

23 Wesp LM, Deutsch MB. Hormonal and surgical treatment options for transgender women and transfeminine spectrum persons. Psychiatr Clin North Am 2017;40:99–111.

24 Furness S, Roberts H, Marjoribanks J, Lethaby A. Hormone therapy in postmenopausal women and risk of endometrial hyperplasia. Cochrane Database Syst Rev 2012;(8):CD000402.

25 Smith DC, Prentice R, Thompson DJ, Herrmann WL. Association of exogenous estrogen and endometrial carcinoma. N Engl J Med 2015;372:1164–7.

26 Branteld IE, Nilvér H, Ahelegen S. Menstruation during a lifespan: a qualitative study of women’s experiences. Health Care Women Int 2014;35:600–16.

27 van Kesteren PJ, Asscheman H, Megens JA, Gooren LJ. Mortality and morbidity in transsexual subjects treated with cross-sex hormones. Clin Endocrinol (Oxf) 1997;47:337–42.

28 Gooren LJ, Giltay EJ, Bunc MC. Long-term treatment of transsexuals with cross-sex hormones: extensive personal experience. J Clin Endocrinol Metab 2008;93:19–25.

29 Ameratunga D, Weston G, Osanlis T, Catt J, Vollenhoven B. In vitro fertilisation (IVF) with donor eggs in post-menopausal women: are there differences in pregnancy outcomes in women with premature ovarian failure (POF) compared with women with physiological age-related menopause? J Assist Reprod Gen. 2009;26:511–4.

30 Keller DL. Risks related to the use of finasteride (propecia) in pregnant women. Am Fam Physician 2010;82:102.

31 Rathnayake D, Sinclair R. Use of spironolactone in dermatology. Skinmed 2010;8:328–32.

32 Iscan MY, Steyn M. The Human Skeleton in Forensic Medicine. Springfield, IL: Charles C Thomas Publisher; 2013.

33 Rosenberg KR. The evolution of modern human childbirth. Yearb Phys Anthropol 1992;35:89–124.

34 Correia H, Balseiro S, De Areia M. Sexual dimorphism in the human pelvis: testing a new hypothesis. Homo 2015;56:153–60.

35 Williams TM, Carroll SB. Genetic and molecular insights into the development and evolution of sexual dimorphism. Nat Rev Genet 2009;10:797–804.

36 Sítik A, Fajkáková M, Žďáňínská E, Antoszewski B. Biometric characteristics of the pelvis in female-to-male transsexuals. Arch Sex Behav 2012;41:1303–13.