Evaluation of the effectiveness of BG-Sentinel and CDC light traps in assessing the abundance, richness, and community composition of mosquitoes in rural and natural areas

André B. B. Wilke1*, Chalmers Vasquez2, Augusto Carvajal2, Maday Moreno2, William D. Petrie2 and John C. Beier1

Abstract

Background: Vector-borne diseases are a major burden to public health. Controlling mosquitoes is considered the most effective way to prevent vector-borne disease transmission. Mosquito surveillance is a core component of integrated vector management, as surveillance programs are often the cornerstone for the development of mosquito control operations. Two traps are the most commonly used for the surveillance of adult mosquitoes: Centers for Disease Control and Prevention miniature light trap (CDC light trap) and BG-Sentinel trap (BioGents, Regensburg, Germany). However, despite the importance of the BG-Sentinel trap in surveillance programs in the United States, especially in the Southern states, its effectiveness in consistently and reliably collecting mosquitoes in rural and natural areas is still unknown. We hypothesized that BG-Sentinel and CDC light traps would be more attractive to specific mosquito species present in rural and natural areas. Therefore, our objective was to compare the relative abundance, species richness, and community composition of mosquitoes collected in natural and rural areas by BG-Sentinel and CDC light traps.

Methods: Mosquitoes were collected from October 2020 to March 2021 using BG-Sentinel and CDC light traps baited with dry ice, totaling 105 trap-nights.

Results: The BG-Sentinel traps collected 195,115 mosquitoes comprising 23 species from eight genera, and the CDC light traps collected 188,594 mosquitoes comprising 23 species from eight genera. The results from the permutational multivariate analysis of variance (PERMANOVA) and generalized estimating equation model for repeated measures indicate the BG-Sentinel and CDC light traps had similar sampling power.

Conclusion: Even though BG-Sentinel traps had a slightly better performance, the difference was not statistically significant indicating that both traps are suitable to be used in mosquito surveillance in rural and natural areas.

Keywords: Mosquitoes, Mosquito surveillance, Vector-borne diseases, Arboviruses, Malaria

*Correspondence: axb1737@med.miami.edu
1 Department of Public Health Sciences, Miller School of Medicine, University of Miami, 1120 Northwest 14th Street, Miami, FL 33136, USA
Full list of author information is available at the end of the article
Background

Vector-borne diseases are a major burden to public health. Currently, half of the world’s population is at risk of vector-borne pathogen infections, resulting in approximately 1 billion infections every year [1–4]. In 2019, approximately 3 million cases of dengue were reported in the Americas [5], and vector-borne pathogen transmission is being reported more frequently not only in endemic areas [6–9], but also in non-endemic countries such as Croatia, France, and Italy [10–12].

The availability of effective drugs and vaccines is limited to a few pathogens and has reduced effectiveness in decreasing the prevalence and incidence of vector-borne pathogens [13–16]. In this context, controlling mosquitoes is widely accepted as the most effective way to prevent vector-borne pathogen transmission to humans and animals [17–19]. However, many mosquito vector species are responsible for transmitting different arboviruses and other pathogens, and the timely and precise detection of mosquito vector species in a given area is key for the development of targeted and effective control strategies [20].

Mosquito surveillance is a core component of integrated vector management, as surveillance programs are often the cornerstone for the development of mosquito control operations [18]. Most mosquito control districts in the United States operate surveillance systems to inform their control operations and guide control efforts including source reduction, chemical interventions, and environmental management [21–23]. However, consistently, accurately, and reliably assessing the presence and relative abundance of mosquito species is not a simple task often relying upon multiple trap types and approaches [23–26]. Furthermore, invasive mosquito species are an increasing threat to public health, and vector species such as *Aedes albopictus* and *Culex coronator* have expanded their range and abundance considerably in the last decade [27–32]. A reliable mosquito surveillance system should be able to early detect invasive species allowing stakeholders to implement control efforts to curb their proliferation and avoid their establishment.

Therefore, a surveillance system should be able to inform stakeholders regarding the relative abundance, species richness, and community composition of mosquitoes. However, different traps have different levels of attractiveness for different mosquito species, and choosing the right trap for collecting adult mosquitoes in different areas is key to achieving reliable and actionable results. In this context, two traps are the most commonly used for the surveillance of adult mosquitoes, Centers for Disease Control and Prevention miniature light trap (CDC light trap) and BG-Sentinel trap (BioGents, Regensburg, Germany). BG-Sentinel traps are the current gold standard for collecting *Aedes stegomyia* mosquitoes [33]. On the other hand, CDC light traps are considered more of a generalist trap that will attract a wider range of mosquito species, including *Anopheles* and *Culex* [23, 34, 35].

However, despite the importance of the BG-Sentinel trap in surveillance programs in the United States, especially in the Southern states, its effectiveness in consistently and reliably collecting mosquitoes in rural and natural areas in the United States is yet to be determined. We hypothesized that BG-Sentinel and CDC light traps have different levels of attractiveness and will be more attractive to specific mosquito species present in rural and natural areas leading to different outcomes of the community composition assessment. Therefore, our objective was to compare the relative abundance, species richness, and community composition of mosquitoes collected in natural and rural areas by BG-Sentinel and CDC light traps.

Methods

Collection of mosquitoes

Mosquitoes were collected from October 2020 to March 2021 using battery-powered BG-Sentinel 2 and CDC light traps, totaling 105 trap-nights. Firstly, we set three CDC and three BG-Sentinel traps at no more than 50 m from each other once a week for 24 h for 7 weeks in a rural area in the southern region of Miami-Dade County, Florida known for having great richness and abundance of mosquitoes (25°24′19.9″N; 80°30′03.9″W). Secondly, to test if the mosquito species richness and relative abundance after 7 weeks of collections would be similar in other areas of the Miami-Dade County, we selected 11 different collection sites in rural and natural areas, in which one BG-Sentinel and one CDC light trap were deployed together for 24 h at no more than 50 m from each other (Fig. 1). An insulated cooler with approximately 2 kg of dry ice was placed next to the traps as bait [36]. The traps were placed under similar environmental conditions hidden in the vegetation in shaded areas to protect the traps from the elements and enhance mosquito collections. CDC traps were set in tree branches at a height of approximately 1 m above ground level and BG-Sentinel traps were set directly on the ground. The collected mosquitoes were transported to the Miami-Dade County Mosquito Control Laboratory and subsequently morphologically identified to species using taxonomic keys [37].

Statistical analyses

Biodiversity analyses were carried out for each trap type based on the Shannon, dominance, and equitability indices. The Shannon index takes into consideration species...
abundance and richness, therefore, less diversity results in lower values, and more diversity results in higher values [38]. Dominance (1-Simpson) index estimates the association between species richness and abundance, values close to 1 indicate the presence of dominant species whereas values closer to 0 imply a more even distribution between species richness and abundance [39]. The equitability index is calculated using the Shannon diversity index divided by the logarithm of the number of species [40]. This measures the evenness with which specimens are divided among the species in a given mosquito community. Analyses were carried out with 10,000 randomizations where each randomization is done without replacement using a 95% confidence interval (CI). To compare the mosquito species composition collected by CDC light traps and BG-Sentinel traps, we performed a permutational multivariate analysis of variance (PERMANOVA) with 9999 permutations based on Bray–Curtis distances [41, 42]. The data were organized into two groups (Group 1 = CDC light traps; and Group 2 = BG-Sentinel traps) to compare the mosquito species composition collected by the two different traps. Then, we used the SIMPER (similarity percentage) method to assess the contribution of each mosquito species to the observed differences between trap types [43]. Analyses were done using PAST v3.2 [44].

We performed a generalized estimating equation (GEE) model for repeated measures to assess differences in the species richness (number of species) and relative abundance of mosquitoes collected by BG-Sentinel and CDC light traps [45]. Species richness and relative abundance were used as dependent variables, trap type (BG-Sentinel and CDC light trap) as units, and collection date as repeated measures (longitudinal model). The model was done in SPSS v.28 software.

**Results**

A total of 26 species from nine genera were collected by both BG-Sentinel and CDC light traps, totaling 383,709 specimens. The BG-Sentinel traps collected 195,115 mosquitoes comprising 23 species from eight genera. The CDC light traps collected 188,594 mosquitoes comprising 23 species from eight genera. *Aedes triseriatus*, *Coquillettidia perturbans*, and *Culex quinquefasciatus* were only collected by the BG-Sentinel traps, whereas *Aedeomyia squamipennis*, *Aedes condolelsens*, and *Wyeomyia mitchelli* were only collected by the CDC light traps. The BG-Sentinel traps collected 6521 more mosquitoes than the CDC light traps. *Culex nigripalpus* was the most abundant species collected by both the BG-Sentinel (131,661) and CDC light traps (131,237), followed by *Culex erraticus* (BG-Sentinel = 16,127; CDC light trap = 13,442) and

---

**Fig. 1** Map showing the location of the collection sites in Miami-Dade, Florida. The first set of 7 weeks of collection sites and trap locations are displayed in green, and the second set of experiments showing the 11 different collection sites in rural and natural areas and trap locations are displayed in red. The figure was produced using ArcGIS 10.2 (Esri, Redlands, CA) using freely available layers from the Miami-Dade County’s Open Data Hub—https://gis-mdc.opendata.arcgis.com/
Anopheles crucians (BG-Sentinel = 15,515; CDC light trap = 13,878). Aedes triseriatus and Ae. condolescens were the least common species, being collected only once by a BG-Sentinel and a CDC light trap, respectively. Mansonia dyari was collected in larger numbers by BG-Sentinel traps (BG-Sentinel = 16,823; CDC light trap = 7984), on the other hand, Culex panocossa was collected in larger numbers by CDC light traps (BG-Sentinel = 2148; CDC light trap = 8387) (Table 1).

The diversity indices yielded similar values for the mosquito community identified by both BG-Sentinel and CDC light traps. The mosquito community estimated by the BG-Sentinel traps yielded a dominance index of 0.47, whereas the CDC light traps yielded a dominance index of 0.5. The Shannon and equitability indices also yielded similar results for both traps 1.2 and 0.38, respectively (Table 2).

The PERMANOVA did not yield significant results for the comparison between the mosquito community comprising the mosquitoes collected by the BG-Sentinel and CDC light traps (F = 1.54; P = 0.11). The subsequent SIMPER analysis of the mosquito community showed that Cx. nigripalpus, An. crucians, and Cx. erraticus contributed the most to the observed differences (Table 3).

The results of the GEE models for repeated measures for species richness and relative abundance of mosquitoes collected by BG-Sentinel and CDC light traps showed no statistically significant differences between traps (Table 4). Even though the comparison of the species richness and relative abundance of the mosquitoes collected by the BG-Sentinel and CDC light traps were not significantly different, the BG-Sentinel traps collected more species per trap-night when compared to CDC light traps. Both the BG-Sentinel and CDC light traps had similar performances estimating the relative abundance of mosquitoes (Fig. 2).

**Discussion**

BG-Sentinel and CDC light traps have been extensively used to successfully assess the relative abundance, species richness, and community composition of vector

### Table 1: Total number of mosquitoes collected by BG-Sentinel and CDC light traps in Miami-Dade County, Florida

| Species                  | BG-Sentinel trap | CDC light trap | Grand total |
|--------------------------|------------------|---------------|-------------|
|                          | Males | Females | Total | Males | Females | Total | Males | Females | Total |
| Aedeomyia squamipennis  |       |         |       |       |         |       | 3     | 39     | 42    |
| Aedes albopictus         | 17    | 17      | 34    | 3     | 11      | 14    |
| Aedes atlanticus         | 10    | 5174    | 5184  | 2978  | 2978    | 5956  |
| Aedes condoleiscens      | 0     | 1       | 1     | 1     | 1       | 2     |
| Aedes infirmatus         | 27    | 27      | 54    | 55    | 55      | 110   |
| Aedes scapularis         | 5     | 5       | 10    | 36    | 36      | 72    |
| Aedes taeniomyrnchus     | 55    | 55      | 110   | 101   | 101     | 202   |
| Aedes tortilis           | 177   | 177     | 354   | 346   | 346     | 692   |
| Aedes triseriatus        | 1     | 1       | 2     | 1     | 1       | 2     |
| Anopheles crucians       | 192   | 15,323  | 15,515| 32    | 13,846  | 13,878| 25,293|
| Anopheles quadrimaculatus| 2     | 3089    | 3091  | 3     | 4299    | 4302  | 4591  |
| Anopheles walkeri        | 51    | 51      | 102   | 97    | 97      | 194   |
| Coquilletidina perturbans| 7     | 7       | 14    | 7     | 7       | 14    |
| Culex coronator          | 124   | 124     | 248   | 64    | 64      | 128   |
| Culex erraticus          | 27    | 16,100  | 16,127| 77    | 13,365  | 13,442| 20,807|
| Culex intergator         | 206   | 206     | 412   | 379   | 379     | 758   |
| Culex nigripalpus        | 62    | 131,599 | 131,661| 176   | 131,061 | 131,237| 262,898|
| Culex panocossa          | 10    | 2138    | 2148  | 117   | 8270    | 8387  | 10,535|
| Culex quinquefasciatus   | 5     | 5       | 10    | 5     | 5       | 10    |
| Mansonia dyari           | 16,823| 16,823  | 33,646| 7984  | 7984    | 15,968| 24,807|
| Mansonia titillans       | 3735  | 3735    | 7470  | 5157  | 5157    | 10314 |
| Psorophora columbiae     | 108   | 108     | 216   | 32    | 32      | 64    |
| Uranotaenia lowii        | 2     | 24      | 26    | 8     | 43      | 51    |
| Uranotaenia sapphirina   | 1     | 20      | 21    | 2     | 8       | 10    |
| Wyeomyia mitchelli       | 0     | 3       | 3     | 3     | 3       | 6     |
| Wyeomyia vanduzei        | 1     | 1       | 1     | 1     | 1       | 2     |
mosquitoes [23, 46, 47], and are essential for mosquito control operations. Our results show that the BG-Sentinel and the CDC light traps performed equally in assessing the species richness and relative abundance of mosquitoes. The number of mosquitoes collected by the BG-Sentinel and the CDC light traps varied less than 4%. Both the BG-Sentinel and the CDC light traps collected 23 species from eight genera from a total of 26 species from nine genera detected in total. The diversity indices yielded virtually identical results and the GEE model for repeated measures showed no significant differences between the performance of the BG-Sentinel and CDC light traps. However, even though the difference was not statistically significant, the BG-Sentinel traps had slightly superior performance; they collected more mosquitoes in total and yielded higher species richness in more trap-nights.

Most species collected by the BG-Sentinel and the CDC light traps during this study were fairly evenly distributed between traps. However, *Ma. dyari* and *Cx. panocossa* were the exceptions. Twice as many *Ma. dyari* were collected by BG-Sentinel traps, and *Cx. panocossa* was collected approximately 4 times more by CDC light traps. Furthermore, BG-Sentinel traps failed to collect *Ad. squamipennis*, and CDC light traps failed to collect *Cq. perturbans*. These results indicate that even though the performance in collecting mosquitoes of the BG-Sentinel and the CDC light traps were not statistically significantly different, some species were more attracted by one trap instead of the other.

The results of the PERMANOVA showed no significant differences in the mosquito community collected by the BG-Sentinel and the CDC light traps, in agreement with the GEE model for repeated measures and the diversity indices. Furthermore, the subsequent SIMPER analysis showed that the most abundant mosquitoes contributed the most to the observed differences indicating that the performance of the traps in collecting mosquitoes was similar. These results indicate that both trap types had similar performances in collecting rare species or failing to collect specific species. Studies done in Europe and China had similar results to the ones obtained in this study, in which BG-Sentinel traps performed equally or were slightly superior to CDC light traps [48, 49]. However, in another study done in South Africa CDC traps had a superior performance in comparison to BG-Sentinel traps [50]. Local environmental and climatic conditions have a major influence on the development and proliferation of mosquitoes and greatly affect their behavior and ecology [51–54]. Therefore, locally assessing the effectiveness of the traps used to investigate the mosquito community composition, species richness and relative abundance in rural and natural areas is essential to improve the reliability and usefulness of mosquito surveillance and early warning systems.

Our results showed the presence of mosquito vector species in the rural and natural areas surveyed in this study. Among them two primary vectors of pathogens were collected in large numbers, *Anopheles quadrimaculatus* (primary vector of malaria in the Americas) and *Cx. nigrpalpus* (primary vector of West Nile virus) [53, 55]. Mosquito surveillance in rural and natural areas
bordering urban areas is key to avoiding vector-borne pathogen transmission to human and animal populations [56–58]. Anthropogenic alterations in the environment such as deforestation and defaunation often lead to habitat fragmentation [59]. Such human-made environmental alterations have a substantial impact on the mosquito community composition, relative abundance, and species richness. The behavior and ecology of mosquito vector species that are anthropophilic but non-synanthropic are greatly affected by anthropogenic alterations in the environment [60–63]. As a consequence, these mosquito vector species will invade urban areas seeking resources and will increase their contact with humans leading to a higher risk of pathogen spillover to human populations [64–66]. Therefore, the correct identification of the relative abundance and species richness in rural and natural areas bordering urban areas is key to determining the risk of vector-borne pathogens to humans [67–69].

Reliable and effective mosquito surveillance systems are key for the early detection of invasive species and to help to prevent their establishment as well as to inform mosquito control operations and guide control efforts [70–72]. Even though BG-Sentinel and the CDC light traps have had similar performances and were able to assess the community composition of mosquitoes in rural and natural areas, other sampling methods should also be considered to improve the effectiveness of surveillance systems. Immature mosquito surveillance systems should also be considered as the information obtained from such surveillance systems is complementary to adult mosquito surveillance systems, providing important information on what aquatic habitats are being used by each species and where the highest relative abundance levels of immature mosquitoes are concentrated [24]. Gravid traps are also an important tool since they use a different approach than traps that mimic a host (e.g., BG-Sentinel and the CDC light traps), and thus potentially complementing the sampling power of the surveillance system [73–75].

Mosquito collections were done between October 2020 and March 2021, and therefore, we were unable to assess all weather and season variations that would have provided further insight into the population dynamics and the mosquito community composition. CDC light traps and BG-Sentinel traps were not rotated in the first set of experiments due to the need to tie CDC light traps to tree branches. For this reason, the traps were set in the same environment at no more than 50 m from each other to avoid biases and inconsistencies in the collections.

### Table 4: Results of the generalized estimating equation models for repeated measures for species richness and relative abundance of mosquitoes collected by BG-Sentinel and CDC light traps

| Dependent variables | Parameters | Parameter estimates | Tests of model effects |
|---------------------|------------|---------------------|------------------------|
|                     |            | Standard error | 95% Wald CI | Wald Chi-square | df | P-value | Wald Chi-square | df | P-value |
| **Species richness** | Intercept  | 1.30 | 5.06 | 10.17 | 34.22 | 1 | >0.001 | 100.40 | 1 | >0.001 |
|                     | Trap type  | 1.49 | −3.22 | 2.63 | 0.03 | 1 | 0.84 | 0.04 | 1 | 0.840 |
| **Relative abundance** | Intercept | 1616.40 | 844.55 | 7180.72 | 6.16 | 1 | 0.013 | 14.11 | 1 | >0.001 |
|                     | Trap type  | 1995.68 | −4439.89 | 3383.01 | 0.07 | 1 | 0.79 | 0.07 | 1 | 0.790 |

**Fig. 2** Comparison of the effectiveness of BG-Sentinel and CDC light traps in assessing species richness and relative abundance of mosquitoes in rural and natural areas of Miami-Dade, Florida
Conclusion

The results of the BG-Sentinel and CDC light traps in assessing the relative abundance and species richness of mosquitoes in rural and natural areas of Miami-Dade indicate that both traps performed equally, yielding similar results in all analyses. Therefore, we were able to reject the hypothesis that BG-Sentinel and CDC light traps would be more attractive to specific mosquito species present in rural and natural areas. Even though BG-Sentinel traps had a slightly better performance, the difference was not statistically significant indicating that both traps are suitable to be used in mosquito surveillance in rural and natural areas.

Acknowledgements

We thank the staff of the Miami-Dade County Mosquito Control Division for their help in the processing and identification of the mosquitoes.

Authors’ contributions

ABBW and CV conceived of and designed the study. ABBW, AC, and CV were responsible for the mosquito collection. MM was responsible for the taxonomic identification. ABBW and CV developed the study methodology and data analysis methodologies. ABBW analyzed the data and prepared the original figures. ABBW wrote the original draft of the paper. All authors contributed to reviewing and editing the paper. CV, WDP, and JCB were responsible for the project administration, funding acquisition, resources, supervision, and validation of this study.

Funding

This research was supported by the Miami-Dade Mosquito Control Division and by the CDC (https://www.cdc.gov/) grant 1U01CK000510-05. Southeastern Regional Center of Excellence in Vector-Borne Diseases: The Gateway Program. CDC had no role in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Declarations

Ethics approval and consent to participate

Since this study posed less than minimal risk to participants and did not involve endangered or protected species, the Institutional Review Board at the University of Miami determined that the study was exempt from institutional review board assessment (IRB Protocol Number: 20161212).

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

1 Department of Public Health Sciences, Miller School of Medicine, University of Miami, 1120 Northwest 14th Street, Miami, FL 33136, USA. 2 Miami-Dade County Mosquito Control Division, Miami, FL, USA.

Received: 22 September 2021   Accepted: 21 January 2022   Published online: 08 February 2022

References

1. Bhatt S, Gething PW, Brady OJ, Messina JP, Farlow AW, Moyes CL, et al. The global distribution and burden of dengue. Nature. 2013;496:504–7.
2. Gaythorpe KAM, Hamlet A, Keusch GT. The intolerable burden of malaria: a new look at the numbers. Am J Trop Med Hyg. 2001;64:iv–vii.
3. World Health Organization—WHO. Vector-borne diseases. https://www.who.int/news-room/fact-sheets/detail/dengue-and-severe-dengue
4. Panamerican Health Organization—PAHO. Reported cases of dengue fever in The Americas, 2019. http://www.paho.org/data/index.php/en/ mnu-topics/indicadores-dengue-en/dengue-nacional-en/252-dengue-pais-ano-en.htm
5. Abdul-Ghani R, Mahdy MAK, Al-Eryani SMA, Fouque F, Lenhart AE, Allkawi A, et al. Impact of population displacement and forced movements on the transmission and outbreaks of Aedes-borne viral diseases: dengue as a model. Acta Trop. 2019;197:105066.
6. Roiz D, Wilson AL, Scott TW, Fonseca DM, Jourdain F, Müller P, et al. Autochthonous dengue fever and chikungunya fever in France: from bad dream to reality! Clin Microbiol Infect. 2010;16:1702–4.
7. Wilke ABB, Vasquez C, Medina J, Carvajal A, Petrie WD, et al. Autochthonous dengue fever in Croatia, August-September 2010. Euro Surveill. 2011;16:1–4.
8. Wilke ABB, Beier JC, Benelli G. Filariasis vector control down-played to reality! Clin Microbiol Infect. 2010;16:1702–4.
9. Poletti P, Messeri G, Ayelli M, Vallarini R, Rizzo C, Merler S. Transmission potential of chikungunya virus and control measures: the case of Italy. PLoS ONE. 2011;6:e18860.
10. Kraemer MUG, Faria NR, Reiner RC, Golding N, Nikolay B, Stasse S, et al. Autochthonous dengue fever and chikungunya virus in South-East Asia: a modelling study. Lancet Infect Dis. 2017;17:330–8.
11. Barrett ADT. Yellow Fever in Angola and Beyond—therapeutic update. N Engl J Med. 2016;375:301–3.
12. Lizzio K, Qualls WA, Brown SC, Beier JC. Expanding integrated vector management to promote healthy environments. Trends Parasitol. 2014;30:394–400.
13. World Health Organization—WHO. Vector-borne diseases: dengue as a model. Acta Trop. 2019;197:105066.
14. Wilke ABB, Beier JC, Benelli G. Filariasis vector control down-played to reality! Clin Microbiol Infect. 2010;16:1702–4.
15. Poletti P, Messeri G, Ayelli M, Vallarini R, Rizzo C, Merler S. Transmission potential of chikungunya virus and control measures: the case of Italy. PLoS ONE. 2011;6:e18860.
16. Kraemer MUG, Faria NR, Reiner RC, Golding N, Nikolay B, Stasse S, et al. Autochthonous dengue fever and chikungunya fever in France: from bad dream to reality! Clin Microbiol Infect. 2010;16:1702–4.
17. Wilke ABB, Beier JC, Benelli G. Filariasis vector control down-played to reality! Clin Microbiol Infect. 2010;16:1702–4.
18. Kraemer MUG, Faria NR, Reiner RC, Golding N, Nikolay B, Stasse S, et al. Autochthonous dengue fever and chikungunya fever in France: from bad dream to reality! Clin Microbiol Infect. 2010;16:1702–4.
19. Roiz D, Wilson AL, Scott TW, Fonseca DM, Jourdain F, Müller P, et al. Autochthonous dengue fever in Croatia, August-September 2010. Euro Surveill. 2011;16:1–4.
20. Wilke ABB, Beier JC, Benelli G. Filariasis vector control down-played to reality! Clin Microbiol Infect. 2010;16:1702–4.
21. Stevens MCAA, Faulkner SC, Wilke ABB, Beier JC, Vasquez C, Petrie WD, et al. Spatially clustered count data provide more efficient search strategies in invasion biology and disease control. Ecol Appl. 2021;31:1–11.
22. Wilke ABB, Vasquez C, Carvajal A, Ramirez M, Cardenas E, Petrie WD, et al. Effectiveness of adulticide and larvicide in controlling high densities of Aedes aegypti in urban environments. PLoS ONE. 2021;16:e0246046.
23. Wilke ABB, Vasquez C, Medina J, Carvajal A, Petrie W, Beier JC. Community composition and year-round abundance of vector species of mosquitoes make Miami-Dade County, Florida a receptive gateway for arbovirus entry to the United States. Sci Rep. 2019;9:8732.
24. Wilke ABB, Davis C, Vasquez C, Carvajal A, Medina J, Petrie WD, et al. Urbanization creates diverse aquatic habitats for immature mosquitoes in urban areas. Sci Rep. 2019;9:15335.
Aedes aegypti (Diptera: Culicidae) in the Continental United States. J Med Entomol. 2020;58:10–25.

27. Wilke ABB, Benelli G, Beier JC. Beyond frontiers: on invasive alien mosquito species in America and Europe. PLoS Negl Trop Dis. 2020;14:e0007864.

28. Wilke ABB, Vásquez C, Cardenas G, Carvalj A, Medina J, Petrie WD, et al. Invasion, establishment, and spread of invasive mosquitoes from the Culex coronator complex in urban areas of Miami-Dade County, Florida. Sci Rep. 2021;11:4620.

29. Benelli G, Wilke ABB, Beier JC. Aedes albopictus (Asian Tiger Mosquito). Trends Parasitol. 2020;36:942–3.

30. Alto BW, Connelly CR, Meara GFO, Hickman D, Karr N. Reproductive biology and susceptibility of Florida Culex coronator to infection with West Nile Virus. Vector Borne Zoonotic Dis. 2014;14:606–14.

31. Roche B, Léger L, Lambert G, Lacour G, Fousadier R, Besnard G, et al. The spread of Aedes albopictus in Metropolitan France: contribution of environmental drivers and human activities and predictions for a near future. PLoS ONE. 2015;10:11.

32. Cebri S. A literature review of host feeding patterns of invasive Aedes mosquitoes in Europe. Insects. 2020;11:848.

33. Maciel-de-Freitas R, Eiras AE, Lourenço-de-Oliveira R. Field evaluation of the effectiveness of the BG-Sentinel, a new trap for capturing adult Aedes aegypti (Diptera: Culicidae). MemInst Oswaldo Cruz. 2006;101:321–5.

34. Medeiros-Sousa AR, Fernandes A, Ceretti-Junior W, Wilke ABB, Marrelli MT. Mosquitoes in urban green spaces: using an island biogeography approach to identify drivers of species richness and composition. Sci Rep. 2017;7:17826.

35. Sudia WD. Battery-operated light trap, an improved model. Mosq News. 1962; p. 265.

36. Wilke ABB, Carvajal A, Medina J, Anderson M, Nieves VJ, Ramirez M, et al. Assessment of the effectiveness of the BG-Sentinel traps baited with CO2 and BG-Lure for the surveillance of vector mosquitoes in Miami-Dade County, Florida. Samy AM, editor. PLoS One 2019;14:e0212688. https://doi.org/10.1371/journal.pone.0212688

37. Darsie FR Jr, Morris CD. Keys to the adult females and fourth-instar larvae of the mosquitoes of Florida (Diptera, Culicidae), vol. 1, 1st edn. Tech Bull Florida Mosq Cont Assoc. 2000.

38. Hutcheson K. A test for comparing diversities based on the Shannon formula. J Theor Biol. 1970;29:151–4.

39. Sheldon AL. Equitability indices: dependence on the species count. Ecology. 1969;50:466–7.

40. Colwell RK. Biodiversity: concepts, patterns, and measurement. Commu- nicities Ecosyst. 2009. p. 257–64.

41. Anderson MJ. Permutational multivariate analysis of variance (PER-MANOVA). Wiley StatsRef: Statistics Reference Online. 1–15 (2017) DOI: https://doi.org/10.1002/9781118445112.stat07841.

42. Amano S, Takahata N, Azuma J, Suwa K, Okada H, Hamada H, et al. The spread of Aedes albopictus in Japan as assessed by genetic analysis. Sci Rep. 2017;7:13711/19

43. Gorsich EE, Beechler BR, van Bodegom PM, Govender D, Guardo MM, Venter M, et al. A comparative assessment of adult mosquito trapping methods to estimate spatial patterns of abundance and community composition in southern Africa. Parasit Vectors. 2019;12:462.

44. Wilke ABB, Vásquez C, Carvalj A, Medina J, Chase C, Cardenas G, et al. Proliferation of Aedes aegypti in urban environments mediated by the availability of key aquatic habitats. Sci Rep. 2020;10:12925.

45. Wilke ABB, Wisinski BF, Benelli G, Vásquez C, Mutebi J, Petrie WD, et al. Local conditions favor dengue transmission in the contiguous United States. Entomol Gen. 2021;41:523–9.

46. Link WS, Collins S, Gaffney D, Armstrong MJ, et al. A global map of dominant malaria vectors. Parasit Vectors. 2012;5:69.

47. Ferraguti M, Martínez-de la Puente J, Roiz D, Ruiz S, Soriguera R, Figuerola J. Effects of landscape anthropization on mosquito community composition and abundance. Sci Rep. 2016;6:29002.

48. Centers for Disease Control and Prevention. Mosquito species in which West Nile virus has been detected. 2017. https://www.cdc.gov/westnile/resources/pdfs/MosquitoSpecies1999-2012.pdf

49. Burkett-Cadena ND, Vittor AV. Deforestation and vector-borne disease: forest conversion favors important mosquito vectors of human pathogens. Basic Appl Ecol. 2018;26:101–10.

50. Dunphy BM, Kovach KB, Gehrke EJ, Field EH, Rowley WA, Bartholomay LC, et al. Long-term surveillance defines spatial and temporal patterns implicating Culex tarsalis as the primary vector of West Nile virus. Sci Rep. 2019;9:1–10.

51. Weaver SC. Urbanization and geographic expansion of zoonotic arboviral diseases: mechanisms and potential strategies for prevention. Trends Microbiol. 2013;21:360–3.

52. Izro R, Young H, Galetti M, Ceballos G, Nick J, Collen B. Defaunation in the Anthropocene. Science. 2014;345:401.

53. Multini LC, Wilke ABB, Marrelli MT. Neotropical Anopheles (Kerteszia) mos- quitoes associated with bromelial-malaria transmission in a changing world. Acta Trop. 2020;205:105413.

54. Souza D, Goes D, Paula D, Road SP, Tropical M, Janeiro D, et al. Genomic surveillance of yellow fever virus epidemic waves in São Paulo, Brazil, 2017–2018. PLOS Pathog. 2020;16:e1008699.

55. Abreu FV, Ribeiro IP, Ferreira-de-Brito A, Santos AA, Miranda RM, Bonelli ID, et al. Haemagogus leucocelaenius and Haemagogus janthinomys are the primary vectors in the major yellow fever outbreak in Brazil, 2016–2018. Emerg Microbes Infect. 2018;28:13–31.

56. Lorenz C, Azevedo TS, Virginio F, Sallum MAM. Impact of environmental factors on neglected emerging arboviral diseases. PLoS Negl Trop Dis. 2017;11:1–9.

57. Multini LC, de Souza AL, Marrelli MT, Wilke AB. The influence of anthropo-genic habitat fragmentation on the genetic structure and diversity of the malaria vector Anopheles cruzii (Diptera: Culicidae). Sci Rep. 2020;10:18018.

58. Laporta GZ, de Prado PIKL, Kraelnik RA, Coutinho RM, Sallum MAM. Biodiversity can help prevent malaria outbreaks in tropical forests. PLoS Negl Trop Dis. 2013;7:e2139.

59. Zohdy S, Schwartz TS, Oaks JR. The coevolution effect as a driver of spillover. Trends Parasitol. 2019;35:399–408.

60. Multini LC, Marrelli MT, Beier JC, Wilke ABB. Increasing complexity threatens the elimination of Extra-Amazonian malaria in Brazil. Trends Parasitol. 2019;35:383–7.

61. Laporta GZ, Burattini MN, Levy D, Fukuya LA, de Oliveira TMP, Maselli LM, et al. Plasmodium falciparum in the southeastern Atlantic forest: a challenge to the bromelial-malaria paradigm? Malar J. 2015;14:181.

62. Laporta GZ, de Prado PIKL, Kraelnik RA, Coutinho RM, Sallum MAM. Biodiversity can help prevent malaria outbreaks in tropical forests. PLoS Negl Trop Dis. 2013;7:e2139.

63. Zohdy S, Schwartz TS, Oaks JR. The coevolution effect as a driver of spillover. Trends Parasitol. 2019;35:399–408.

64. Multini LC, Marrelli MT, Beier JC, Wilke ABB. Increasing complexity threatens the elimination of Extra-Amazonian malaria in Brazil. Trends Parasitol. 2019;35:383–7.
72. Ammar SE, McIntyre M, Swan T, Kasper J, Derraik JGB, Baker MG, et al. Intercepted mosquitoes at New Zealand's ports of entry, 2001 to 2018: current status and future concerns. Trop Med Infect Dis. 2019;4:101.
73. Williams GM, Gingrich JB. Comparison of light traps, gravid traps, and resting boxes for West Nile virus surveillance. J Vector Ecol. 2007;32:285–91.
74. Eiras AE, Buhagiar TS, Ritchie SA. Development of the gravid Aedes trap for the capture of adult female container-exploiting mosquitoes (Diptera: Culicidae). J Med Entomol. 2014;51:200–9.
75. Irisha SR, Moorea SJ, Deurac YA, Brucea J, Camerona MM. Evaluation of gravid traps for the collection of Culex quinquefasciatus, a vector of lymphatic filariasis in Tanzania. Trans R Soc Trop Med Hyg. 2013;107:15–22.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.