Identification and fine-mapping of a major QTL (PH1.1) conferring plant height in broomcorn millet (Panicum miliaceum)

Tianpeng Liu1,2, Xueying Liu2, Jihong He1, Kongjun Dong1, Wanxiang Pan3, Lei Zhang1, Ruiyu Ren1, Zhengsheng Zhang2* and Tianyu Yang1*

1Crop Research Institute, Gansu Academy of Agricultural Sciences, Lanzhou, China, 2College of Agronomy and Biotechnology, Southwest University, Chongqing, China, 3College of Life Science and Technology, Gansu Agricultural University, Lanzhou, China

The plant height of broomcorn millet (Panicum miliaceum) is a significant agronomic trait that is closely related to its plant architecture, lodging resistance, and final yield. However, the genes underlying the regulation of plant height in broomcorn millet are rarely reported. Here, an F2 population derived from a cross between a normal variety, “Longmi12,” and a dwarf mutant, “Zhang778,” was constructed. Genetic analysis for the F2 and F2:3 populations revealed that the plant height was controlled by more than one locus. A major quantitative trait locus (QTL), PH1.1, was preliminarily identified in chromosome 1 using bulked segregant analysis sequencing (BSA-seq). PH1.1 was fine-mapped to a 109-kb genomic region with 15 genes using a high-density map. Among them, longmi011482 and longmi011489, containing nonsynonymous variations in their coding regions, and longmi011496, covering multiple insertion/deletion sequences in the promoter regions, may be possible candidate genes for PH1.1. Three diagnostic markers closely linked to PH1.1 were developed to validate the PH1.1 region in broomcorn millet germplasm. These findings laid the foundation for further understanding of the molecular mechanism of plant height regulation in broomcorn millet and are also beneficial to the breeding program for developing new varieties with optimal height.

KEYWORDS
broomcorn millet, plant height, BSA-seq, fine-mapping, AP2
Introduction

Broomcorn millet (*Panicum miliaceum*), an allotetraploid species (*2n = 4x = 36*) with two subgenomes originating from *Panicum capillare* and *Panicum repens* (*Hunt et al., 2014*), has been reported as probably one of the earliest domesticated crops around the world (*Lu et al., 2009; Li et al., 2021*). Phytolith evidence suggested that the domestication of broomcorn millet in northern China could be dated to approx. 10,300 years before the present (cal yr BP) (*Yang et al., 2012*). Due to its desirable characteristics including a short growing season, high water use efficiency, and drought tolerance, broomcorn millet has always been an important crop in semiarid regions. The grains of broomcorn millet are rich in minerals, essential amino acids, vitamins, and fatty acid. Especially, the protein content in broomcorn millet is higher than that in rice, maize, wheat, and most other cereals (*Das et al., 2019*). In addition, broomcorn millet is gluten-free and has a low glycemic index, which makes it an ideal nutritious food for a healthy diet (*Das et al., 2019; Zou et al., 2019*). However, the yield and the annual production of broomcorn millet are quite lower than those of other cereals. Therefore, the breeding of broomcorn millet with desirable agronomical traits and high production will be of great significance.

Plant height, determined by the length of the internode and the number of nodes, is a vital agronomical trait of crops and an important factor influencing crop yield. Cereal plants with a high culm are usually susceptible to lodging, thus decreasing final yield. For the breeding programs of main cereal crops such as rice and wheat, the dwarf trait has been exploited in the first “green revolution” and led to a rapidly increasing production. Genes in hormone pathways, such as *sd1*, *d35*, and *ddf1* involved in the gibberellin (GA) signaling pathway (*Sasaki et al., 2002; Itoh et al., 2004; Magome et al., 2004*), *brd1* involved in the brassinosteroid signaling pathway (*Makarevitch et al., 2012*), and *tdl1* involved in the auxin signaling pathway (*Suzuka et al., 2009*), have been reported to play roles in the morphogenesis of plant height. However, most of these genes have been identified from rice, wheat, and maize, whereas no candidate gene has been reported in broomcorn millet.

The rapid development of molecular biology and sequencing technologies provided various tools for modern crop breeding. Map-based cloning of genes involving significant agronomical traits and marker-assisted selection (MAS) have been widely applied in crops (*Chen et al., 2021; Koppolu et al., 2022*). Bulked segregant analysis with whole-genome resequencing (BAS-seq) is an effective approach for the identification of candidate genes and has been successfully utilized (*Lee et al., 2020; Wu et al., 2022; Zhang et al., 2022*). However, genetic and genomic studies of broomcorn millet were more hysteretic than those of other crops. The release of the assembled genome of broomcorn millet in 2019 made it possible to make full use of modern technologies, including BAS-seq and molecular marker development (*Shi et al., 2019; Zou et al., 2019*).

In the present study, the broomcorn millet dwarf mutant Zhang778 was crossed with the regular variety Longmi12 to explore the candidate quantitative trait loci (QTLs) and genes responsible for plant height trait. One major QTL, *PH1.1*, was identified and fine-mapped into a 109-kb interval using BSA-seq and linkage mapping analysis. Fifteen genes were located in the candidate region, and three candidate genes were the most likely candidate genes for *PH1.1*. Three markers linked with *PH1.1* were identified through association analysis, which validated the mapping results and could be utilized as diagnostic markers of plant height for broomcorn millet breeding. Our results laid a foundation for understanding the molecular mechanism of plant height morphogenesis and contribute to breeding varieties of broomcorn millet with optimal plant architecture.

Materials and methods

Plant materials and phenotypic analysis

Longmi12 and Zhang778 were chosen as the parental lines to establish an F2 population in order to analyze the plant height trait of broomcorn millet. Longmi12 is a high-yield variety bred by the Institute of Crop Research in Gansu Academy of Agricultural Sciences, China. The height of mature Longmi12 plants is about 160–180 cm. Zhang778, bred by Zhangjiakou Academy of Agricultural Sciences, China, is a dwarf line created through ethyl methanesulfonate (EMS) mutagenesis, with a final height of about 60–70 cm. Longmi12 was crossed with Zhang778 in Huining in 2018, the F1 plants were planted also in Huining in 2019, and the F2 population with 939 individuals was planted in the summer of 2020 in Huining, Gansu Province. The progenies of each F2 individual were planted in a line to develop the F2.3 population in the summer of 2021 in Huining, Gansu Province.

The plant height of broomcorn millet was measured using a ruler with an accuracy of 0.1 cm. For each F2.3 line, the average height of all plants per line was taken as a representative value. Moreover, the internode length of adjacent nodes and the panicle length in Longmi12 and Zhang778 were measured.

The R package “SEA V2.0” ([https://cran.r-project.org/web/packages/SEA/index.html](https://cran.r-project.org/web/packages/SEA/index.html)) was utilized to analyze the potential inheritance models of plant height in the F2 population. The optimal inheritance model was selected following the instructions in SEA v2.0.

To verify the accuracy of QTL *PH1.1*, a total of 512 broomcorn millet accessions collected from all over the world were used to carry out an association analysis (Supplementary Table S1). These accessions were planted and measured in Huining in 2020 and 2021.
BSA-Seq and resequencing analysis

The DNAs of Longmi12, Zhang778, and the F2 individuals were extracted from young leaves using a modified cetyltrimethyl ammonium bromide (CTAB) method (Zhang et al., 2005). Fifty F2 individuals with height over 160 cm were collected for the “high bulk,” (HB), while 50 individuals shorter than 100 cm were collected for the “dwarf bulk” (DB). The DNA libraries of the parents and the two bulks were sequenced on an Illumina HiSeq 2000 platform. Clean reads were aligned to the reference genome assembled by Zou et al. (2019) using BWA software with default parameters (Li and Durbin, 2009). Single nucleotide polymorphisms (SNPs) and insertions–deletions (Indels) were identified using GATK software. The ΔSNP and ΔIndel index values were calculated to obtain the preliminary intervals associated with the plant height of broomcorn millet.

Marker development, linkage map construction, and fine-mapping

To narrow down the preliminary region obtained from the BSA-seq results, Indels in the candidate region were selected according to the comparative genomic resequencing data between Zhang778 and Longmi12. Indel primers were developed based on the flanking sequence of the target Indels. These primers were first used to screen the parents in order to confirm the polymorphism and then to detect the genotype of the F2 individuals.

The software JoinMap 4.0 (Van Ooijen, 2006) was applied to construct the genetic linkage map, while the Kosambi map function was utilized to convert the recombination values into genetic distances (in centimorgan). Fine-mapping analysis of the QTL and the evaluation of its effect were carried out using MapQTL 6.0 (Van Ooijen, 2009) in the F2 and F2:3 populations independently. For the input trait values, the plant height of each F2 individual and the average value of each F2:3 line were used. MapChart 2.2 (Voorrips, 2002) was applied to represent the genetic map and QTL information graphically.

Prediction of the candidate genes

The genome information and the functional annotation of the genes in the candidate region were obtained from the reference genome of broomcorn millet (Zou et al., 2019). The genome resequencing data of Zhang778 and Longmi12 were used to compare the sequence variance between the two parents. Genes with ASNP/ΔIndel index values equal to 1 or −1 and are harboring sequence variance in the coding regions or potential promoter regions were predicted to be candidate genes.

Regional association analysis

Association analysis of the candidate region was conducted using a total of 512 broomcorn millet accessions. DNA extraction was performed for all accessions using the CTAB method modified by Zhang et al. (2005). The Indel markers in the candidate region were utilized to obtain the corresponding genotype of these accessions. According to the method used by Liu et al. (2017), the genotypes of each Indel in all accessions were obtained with PCR amplification and polyacrylamide gels. Thereafter, the accessions were separated into different haplotypes for each locus, and comparisons of the plant height between haplotypes were conducted using the Student’s t-test. A significance threshold (p < 0.05) was used to determine the significant associations between haplotypes and plant height.

Results

Phenotypic characterization and inheritance of plant height

The plant height of cereal crops is usually determined by the elongation and/or the number of nodes. Here, the dwarf mutant Zhang778 and the normal variety Longmi12, as well as their F2 and F2:3 progenies, were planted and phenotyped in the same location across 2 years. Compared with the height of Longmi12 that ranged from 163.1 to 188.5 cm, that of mature Zhang778 plants was only about 58.3–72.1 cm (Figure 1B). The number of nodes in mature plants was the same in two accessions, suggesting that the dwarf trait in Zhang778 was caused only by the decreased length of the internodes. A comparison of the plant height and the internode length between the two parental lines is shown in Figure 1. Besides plant height, the panicle length of Zhang778 was also shorter than that of Longmi12 (Figures 1A, C).

The plant heights of the F2 and F2:3 progenies were continuous and approximately normally distributed and showed no transgressive segregation (Supplementary Figure S1). These results suggested that there might be more than one underlying gene and that all favorable alleles were contributed by one parent. To understand the genetic architecture of plant height, we used the major gene plus polygene mixed inheritance model and analyzed the F2 population with the R package “SEA V2.0.” The results indicated that 2MG-EA (two major genes with additive effects) was the optimal model, and the heritability of the two major genes was about 88.5% (Supplementary Table S2). In addition, the height distribution ranges of the two generations were slightly different (Supplementary Figure S1), which was identical to the parental performance, suggesting that
Environment factors also had an effect on the final height of broomcorn millet.

Initial mapping of plant height by bulked segregant analysis

BSA-seq was carried out to rapidly identify the genomic regions of plant height. A total of 125 M and 348 M paired reads were generated from the two parents and two bulks, respectively. The average sequencing depth of the parents was 22.4×, while that of the bulks was 61.3× (Supplementary Table S3). The clean reads were aligned to the reference genome of broomcorn millet (Zou et al., 2019), which revealed that about 97% and 93% of the whole genome in the bulks and parents, respectively, were covered by more than 10× reads. A total of 734,746 SNPs and 84,609 small Indels were identified between the two parents, while 220,363 SNPs and 27,565 small Indels were identified between the two bulks (Supplementary Table S3). The ΔSNP index was calculated, and a major region located on Chr1: 1,259,618–6,443,508 was identified with a 99% confidence value (Figure 2; Supplementary Table S4). The ΔIndel index was also calculated, and an interval on Chr1, which overlapped with the ΔSNP index interval, was identified (Supplementary Figure S2; Table S4). Overall, the results of BSA-seq revealed a major candidate region of broomcorn millet plant height on Chr1 (hereafter referred to as PH1.1), with a physical distance of about 5.18 Mb.
Fine-Mapping of PH1.1

To narrow down the mapping region of PH1.1, a total of 68 Indel primers were developed based on the genome resequencing data. The primers were first used to screen for polymorphism between parents. Twenty-eight co-dominant primer pairs with clear bands were then utilized to identify the genotypes of 939 F2 individuals (Supplementary Table S5) and to construct a high-density genetic map around the region of PH1.1 (Figure 3). The sequences of these primers are listed in Supplementary Table S6. QTL analysis was further carried out using the plant height phenotypes from the F2 and F2:3 populations. The results indicated that the region of PH1.1 was narrowed down between markers Indel3.506 and Indel3.719 with a phenotypic variance explanation (PVE) value of 36% and an additive effect of 16.68 cm in the F2 generation and from marker Indel3.506 to Indel3.614 with a PVE value of 45.2% and an additive effect of 14.59 cm in the F2:3 generation (Figure 3; Supplementary Table S7). The corresponding physical distance between Indel3.506 and Indel3.615 was 109 kb (109,478 bp).

Candidate gene prediction analysis for PH1.1

A total of 15 genes (longmi011482–longmi011496) were contained in the PH1.1 interval. Associated genes with DSNP or DIndel index value equal to 1 in the PH1.1 interval included longmi011482, longmi011483, and longmi011489, and SNP variants of longmi011482 and longmi011489 occurred in the intron and exon, respectively (Table 1; Supplementary Table S8). Among these genes, 14 were annotated with functions (Figure 4; Table 2). The sequence alignment of the 15 genes between the parental lines revealed that two nonsynonymous SNPs (Chr1: 3,504,266 and Chr1: 3,564,638) were present in longmi011482 and longmi011489 (Figure 4). No nonsynonymous variation was detected in the coding regions of the other genes. Furthermore, longmi011482 encodes a trichome birefringence-like protein, and longmi011489 encodes the floral homeotic protein APETALA 2 (AP2). Moreover, a few SNPs or Indels were present in the potential promoter regions of longmi011484, longmi011488, longmi011489, and longmi011496 (Supplementary Table S8).

Association test of PH1.1 among the natural population

To verify the fine-mapped region of PH1.1, a total of 512 broomcorn millet accessions (Supplementary Table S1) with various plant heights were genotyped using the four Indel primers in the candidate region (i.e., Indel3.506, Indel3.561, Indel3.614, and Indel3.615). The association test indicated that accessions carrying Longmi12 alleles at both the Indel3.506 and Indel3.614 loci were significantly higher than accessions carrying Zhang778 alleles. In particular, 23 out of the 63 wild

![FIGURE 3](https://example.com/fig3.png)

**FIGURE 3**
Fine-mapping of PH1.1 by linkage analysis. The left panel shows the linkage map, in which the red segment is the fine positioning interval of PH1.1. The right panel displays the log of odds (LOD) value of each locus in the F2 and F2:3 generations.
germplasm showed genotypes different from those of the two parents at Indel3.615. The plant heights with these genotypes were significantly lower than those of the two parents (Figure 5; Supplementary Table S1), indicating that Indel3.615 is a key marker that can distinguish different types of broomcorn millet germplasm. Furthermore, both Indel3.614 and Indel3.615 are located upstream of the longmi011496 gene (Figure 4), suggesting that longmi011496 is also a promising candidate gene. These results confirmed the candidate region of the major QTL PH1.1 and suggested that PH1.1 might be a domesticated QTL of broomcorn millet.

Discussion

Effective strategy to identify and fine-map major QTLs

With the development of molecular biotechnology in recent years, identifying the QTLs/gene of agronomical traits through forward genetic methods has become a significant part of crop breeding (Song et al., 2007; Ying et al., 2019). Detecting the major QTLs for important traits will not only be helpful to understanding the underlying molecular mechanism but also be favorable to
### TABLE 2 Annotated function of the genes located in the interval of PH1.1.

| Gene ID    | Pfam domain                                                                 | Function annotation                                                                 |
|------------|------------------------------------------------------------------------------|-------------------------------------------------------------------------------------|
| longmi011482 | PMR5 N-terminal domain                                                       | Protein trichome birefringence-like 33                                               |
| longmi011483 | PMR5 N-terminal domain                                                       | Protein trichome birefringence-like 33                                               |
| longmi011484 | –                                                                             | Protein FLX-like 1                                                                   |
| longmi011485 | Histidine phosphatase superfamily (branch 2)                                  | Multiple inositol polyphosphate phospatase 1                                           |
| longmi011486 | NAD-dependent epimerase/dehydratase family, 3-beta hydroxysteroid dehydrogenase/isomerase family, GDP-mannose 4,6-dehydratase | Cinnamoyl-CoA reductase 1                                                             |
| longmi011487 | Alfin, PHD finger                                                             | PHD finger protein ALFIN-LIKE 3                                                       |
| longmi011488 | Ribosomal protein S12/S23                                                     | 40S ribosomal protein S23                                                             |
| longmi011489 | AP2 domain                                                                   | Floral homeotic protein APETALA 2                                                     |
| longmi011490 | F-box-like                                                                   | –                                                                                    |
| longmi011491 | Peptidase M1 N-terminal domain                                                | Leukotriene A-4 hydrolase homolog                                                     |
| longmi011492 | –                                                                             | Glycine-rich protein A3                                                                |
| longmi011493 | Heavy metal-associated domain                                                 | Protein SODIUM POTASSIUM ROOT DEFECTIVE 2                                              |
| longmi011494 | Chalcone–flavonone isomerase                                                  | Chalcone–flavonone isomerase                                                          |
| longmi011495 | Domain of unknown function (DUF4378)                                         | –                                                                                    |

**FIGURE 5**

Association test of plant height in the natural population using insertion–deletion (Indel) markers in the PH1.1 interval. (A–H) Association tests of 512 broomcorn millet accessions in 2020 and (A–D) and 2021 (E–H). *, ** represent significant differences at P < 0.05 and 0.001, respectively. NS indicates no significant difference.
The domestication of broomcorn millet from wild species to modern cultivated varieties has greatly increased the size of the panicle and boosted production. The continuous artificial selection of high yield resulted in the tall plant architecture of cultivated broomcorn millet varieties since plant height is usually positively correlated with panicle size in cereal plants (Zhang et al., 2006; Liu et al., 2013; Chen et al., 2014). Here, a major QTL, PH1.1, was identified across two environments and explained about 45% of the variation in the plant height of broomcorn millet. This result indicated that PH1.1 is a stable locus with a large contribution. Through genotyping with the three Indel markers in the PH1.1 region, broomcorn millet accessions with different alleles were significantly segregated according to plant height. In particular, Indel3.615, an insertion/deletion marker in the promoter region, was closely related to its plant height and can be used in the detection of other QTLs in broomcorn millet.  

**PH1.1 is a major domesticated QTL**

In the confidence interval of PH1.1, five of the 15 genes showed sequence variation in the coding region or the potential promoter region. Among them, longmi011489 encodes an AP2 transcriptional factor harboring variations in both the coding and promoter regions. A few mutants of the AP2 genes have been reported to be candidate genes of dwarf plants, such as ddfl in Arabidopsis (Magome et al., 2004), ndl1 in rice (Kusnandar et al., 2022), and dill in maize (Jiang et al., 2012). AP2 genes have been reported to play a significant role in GA signaling pathways, which are involved in the growth and development of plants (Magome et al., 2004; Xue et al., 2022). In a previous study, Zhang778 was proven to be caused by a mutation of the genes related to GA synthesis (Zhang et al., 2020), which is consistent with the function of AP2 genes. In addition, another previous study detected 21 genes involved in the domestication of broomcorn millet, 19 of which were annotated into the AP2 gene family (Li et al., 2021). As mentioned above, PH1.1 was probably a domesticated QTL consistent with the roles of the AP2 gene family involved in the domestication of broomcorn millet. Another candidate gene, longmi011482, encodes a trichome birefringence-like protein related to cell wall polysaccharide O-acetylation in Arabidopsis (Schultink et al., 2015) and fiber development in upland cotton (Yu and Gervers, 2019). Thus, it may participate in the development of the cell wall to affect the elongation of the broomcorn millet internode and then regulate plant height. Furthermore, longmi011496 is an unknown function gene; however, in the association analysis carried out in this study, the base insertion or deletion in its promoter region was closely related to its plant height and can significantly distinguish wild accessions based on the height of the dwarf plant, indicating that the gene may be a specific gene regulating plant height in broomcorn millet. Overall, their functions still need further verification.

**Diagnostic markers of PH1.1 could be applied in MAS**

Diagnostic markers are molecular markers that are significantly linked with one or more traits, which help in the early and rapid identification of a target phenotype (Chakdar et al., 2019). Diagnostic markers have been successfully employed in many traits of various plants, such as the rust resistance trait in wheat (Liu et al., 2021) and common wheat...
(Wu et al., 2018), the testa color in cultivated peanut (Chen et al., 2021), and the bacterial wilt trait in tomato (Abebe et al., 2020).

In the present study, three Indel markers located in the PH1.1 region were validated to be closely linked with the plant height of broomcorn millet using a total of 512 accessions. As do other cereal crops, high broomcorn millet varieties usually suffer from lodging; hence, breeding new broomcorn millet varieties with optimal height will be of great importance. With the three diagnostic markers, particularly Indel3.615, the plant height of new broomcorn varieties could be distinguished at the seedling stages. Therefore, the diagnostic markers in the present study laid a foundation for the molecular breeding of broomcorn millet and could be directly applied in its MAS breeding.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary Material.

Author contributions

TL and XL performed data analysis and drafted the manuscript. ZZ and TY designed the experiment and revised the manuscript. TL and JH constructed the F2 and F2:3 populations. KD, LZ, and RR collected the phenotype. WP completed the collection of population genotypes. All authors contributed to the article and approved the submitted version.

Funding

This study was financially supported by the China Agriculture Research System (CARS-06-14.5-A8); Special Project of Agricultural Science and Technology innovation of GAAS (2021GAAS02); and the Top-Notch Talent Project in Gansu Province (2021).

Acknowledgments

The authors would like to thank Prof. Ping Lu and Dr. Minxuan Liu (Chinese Academy of Agricultural Sciences) for providing seeds and basic information on the 512 broomcorn millet accessions for this study.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2022.1010057/full#supplementary-material

References

Abebe, A. M., Choi, J., Kim, Y., Oh, C. S., Yeam, I., Nou, I. S., et al. (2020). Development of diagnostic molecular markers for marker-assisted breeding against bacterial wilt in tomato. Breed Sci. 70, 462–473. doi: 10.1270/jsbbs.200217

Chakdar, H., Goswami, S. K., Singh, E., Choudhary, P., Yadav, I., Kashyap, P. L., et al. (2019). noxB-based marker for alternaria spp.: A new diagnostic marker for specific and early detection in crop plants. Biotech 9, 249. doi: 10.1007/s13205-019-1779-4

Chen, H., Chen, X., Xu, R., Liu, W., Liu, N., Huang, L., et al. (2021). Fine-mapping and gene candidate analysis for AhRt1, a major dominant locus responsible for testa color in cultivated peanut. Theor. Appl. Genet. 134, 3721–3730. doi: 10.1007/s00122-021-03924-w

Chen, S., Gan, R., Wang, H., Wen, M., Xiao, J., Bao, N., et al. (2014). Characterization of a novel reduced height gene (Rh23) regulating panicle morphology and plant architecture in bread wheat. Euphytica 203, 583–594. doi: 10.1007/s10681-014-1275-1

Das, S., Khound, R., Santra, M., and Santra, D. K. (2019). Beyond bird feed: Proso millet for human health and environment. Agricult. MDPI 9 (3), 1–19. doi: 10.3390/agriculture9030064

Hunt, H. V., Badakshi, F., Romanova, O., Howe, C. J., Jones, M. K., and Hedgpeth, J. S. P. (2014). Reticulate evolution in Panicum (Poaceae): The origin of tetraploid broomcorn millet, p. miliaceum. J. Exp. Bot. 65, 3165–3175. doi: 10.1093/jxb/eru161

Itoh, H., Tatsumi, T., Sakamoto, T., Otomo, K., Toyomasu, T., Kitano, H., et al. (2004). A rice semi-dwarf gene, tan-ginbozu (D35), encodes the gibberellin biosynthesis enzyme, ent-kaurene oxidase. Plant Mol. Biol. 54, 533–547. doi: 10.1023/B:PLAN.0000038261.21060.47

Jiang, F., Guo, M., Yang, F., Duncan, K., Jackson, D., Rafalski, A., et al. (2012). Mutations in an AP2 transcription factor-like gene affect internode length and leaf shape in maize. PLoS One 7, e37040. doi: 10.1371/journal.pone.0037040
Koppolu, R., Jiang, G., Miller, S. G., Muqaddas, Q. H., Rutten, T., Himmelbach, A., et al. (2022). The barley mutant multiformis2 reveals quantitative genetic variation for new spikelet architecture. *Theor. Appl. Genet.* 135, 571–590. doi: 10.1007/s00122-021-03986-w

Kuswandar, A. S., Itoh, J. I., Sato, Y., Honda, E., Hibara, K. I., Kyozuka, J., et al. (2022). Narrow and dwarf leaf 1, the ortholog of arabidopsis enhancer of shoot Regeneration1/Dornroschen, mediates leaf development and maintenance of the shoot apical meristem in oryza sativa l. *Plant Cell Physiol.* 63, 265–278. doi: 10.1093/pcp/peab169

Lee, S. B., Kim, J. E., Kim, H. T., Lee, G. M., Kim, B. S., and Lee, J. M. (2020). Genetic mapping of the c1 locus by GBS-based BSA-seq revealed pseudo-response regulator 2 as a candidate gene controlling pepper fruit color. *Theor. Appl. Genet.* 133, 1897–1910. doi: 10.1007/s00122-020-03565-5

Li, H., and Durbin, R. (2009). Fast and accurate short read alignment with burrows-wheeler transform. *Bioinformatics* 25, 1754–1760. doi: 10.1093/bioinformatics/btp324

Li, C., Liu, M., Sun, F., Zhao, X., He, M., Li, T., et al. (2021). Genetic divergence and population structure in weedy and cultivated broomcorn millets (Panicum miliaceum l.) revealed by specific loci amplified fragment sequencing (SLAF-seq). *Front. Plant Sci.* 10. doi: 10.3389/fpls.2022.1010057

Liu, X., Teng, Z., Wang, J., Wu, T., Zhang, Z., Deng, X., et al. (2017). Validation and characterization of Ghd7.1, a major quantitative trait locus with pleiotropic effects on spikelets per panicle, plant height, and heading date in rice (Oryza sativa l.). *Natl. Acad. Sci. U.S.A.* 114, 729-1357. doi: 10.1073/pnas.0900158106

Magome, H., Yamaguschi, S., Hanada, A., Kamiya, Y., and Oda, K. (2004). Dwarf and delayed-flowering 1, a novel arabidopsis mutant defective in gibberellin biosynthesis because of overexpression of a putative AP2 transcription factor. *Plant J.* 37, 720–729. doi: 10.1111/j.1365-313X.2003.01998.x

Makarevitch, I., Thompson, A., Muehlbauer, G. J., and Springer, N. M. (2012). Narrow and dwarf leaf 1, the ortholog of arabidopsis enhancer of shoot Regeneration1/Dornroschen, mediates leaf development and maintenance of the shoot apical meristem in oryza sativa l. *Plant Cell Physiol.* 63, 265–278. doi: 10.1093/pcp/peab169

Sazuka, T., Kamiya, N., Nishimura, T., Ohmae, K., Sato, Y., Imamura, K., et al. (2004). Dwarf and delayed-flowering 1, the ortholog of arabidopsis enhancer of shoot Regeneration1/Dornroschen, mediates leaf development and maintenance of the shoot apical meristem in oryza sativa l. *Plant Cell Physiol.* 63, 265–278. doi: 10.1093/pcp/peab169

Song, X., Huang, W., Shi, M., Zhu, M., and Lin, H. (2007). A QTL for rice grain width and weight encodes a previously unknown RING-type E3 ubiquitin ligase. *Nat. Genet.* 39, 623–630. doi: 10.1038/ng102014

Van Ooijen, J. W. (2006). JoinMap 4.0: software for the calculation of genetic linkage maps in experimental populations. *Wageningen: Kyazma BV.*

Van Ooijen, J. (2009). MapQTL 6, software for the mapping of quantitative trait loci in experimental populations of diploid species (Wageningen: Kyazma BV).