Detection of multiple respiratory pathogens during primary respiratory infection: nasal swab versus nasopharyngeal aspirate using real-time polymerase chain reaction

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Abstract In this study, we present the multiple detection of respiratory viruses in infants during primary respiratory illness, investigate the sensitivity of nasal swabs and nasopharyngeal aspirates, and assess whether patient characteristics and viral load played a role in the sensitivity. Healthy infants were included at signs of first respiratory tract infection. Paired nasopharyngeal aspirates and nasal swabs were collected. Real-time polymerase chain reaction (PCR) was carried out for 11 respiratory pathogens. Paired nasopharyngeal aspirates and nasal swabs were collected in 98 infants. Rhinovirus (n=67) and respiratory syncytial virus (n=39) were the most frequently detected. Co-infection occurred in 48% (n=45) of the infants. The sensitivity of the nasal swab was lower than the nasopharyngeal aspirate, in particular, for respiratory syncytial virus (51% vs. 100%) and rhinovirus (75% vs. 97%). The sensitivity of the nasal swab was strongly determined by the cycle threshold (CT) value (p<0.001). The sensitivity of the swab for respiratory syncytial virus, but not rhinovirus, was 100% in children with severe symptoms (score ≥11). It is concluded that, for community-based studies and surveillance purposes, the nasal swab can be used, though the sensitivity is lower than the aspirate, in particular, for the detection of mild cases of respiratory syncytial virus (RSV) infection.

Abbreviations
NS Nasal swab
NPA Nasopharyngeal aspirate
PCR Polymerase chain reaction
hMPV Human metapneumovirus
RSV Respiratory syncytial virus

Background
Respiratory viruses are a common cause of illness in children, in particular during their first years of life, and may lead to more severe morbidity and hospitalisation [1–4]. Different types of specimen are available for viral diagnosis. The nasopharyngeal aspirate (NPA) has been considered to be the best sampling technique, but is more invasive and results in significantly more distress of the infant than a nasal swab (NS) [5]. A number of studies have compared the sensitivity of the NPA with nasopharyngeal swabs [6], nose–throat swabs [7] and NS [5, 8–10]. Generally, conventional techniques such as viral culture and antigen detection methods were used. The use of real-time polymerase chain reaction (PCR) may overcome differences in sensitivity for respiratory viruses as a result of specimen type [7].

Limited data are available on the comparison of these sampling methods combined with real-time PCR. It was
observed that nose–throat swabs are a less invasive diagnostic technique, with adequate sensitivity for use in outpatient and large community-based settings in children [7]. However, no tests were performed for rhinovirus, even though this virus commonly infects infants [11]. The aim of this study was to present the detection of common respiratory pathogens in infants during primary respiratory illness, to investigate the sensitivity of the NS and NPA, and assess the role of patient characteristics and viral load in the sensitivity of either sampling method.

Study design

Study cohort

The study is part of the Netherlands Amnion Fluid Study of the Utrecht University Medical Centre (UUMC), the Netherlands [12]. Healthy infants were included at birth and were at risk for primary respiratory infection until the age of one year. The data collection and episode sampling stopped one year after birth. From April 2006 to April 2008, including two winter seasons, paired NPA and NS specimens were obtained from 98 infants. Parents were instructed to notify the clinical staff within 24 h after the onset of symptoms. Clinical staff visited the child within 36 h and the history of illness was taken by a standardised questionnaire. Symptoms were scored, according to Gern et al. [13], with points presented in parentheses: fever (>38°C) (1); cough, mild (1), moderate (2), severe (3); rhinorrhoea, mild (1), moderate to severe (2); hoarseness (1); duration of illness >4 days (1); apnoea (3); wheezing (5); retractions (5); tachypnoea (5); cyanosis (5). Mild, moderate and severe infection were defined as sum scores 0–4, 5–10 and 11 and higher, respectively. Specially trained clinical staff obtained paired NS and NPA.

Collection of specimens

The NPA was obtained by the use of an infant mucus extractor (VYGON). Both nostrils were suctioned. In addition, an NS was collected; samples were collected from one nostril and one from the hard palate using separate cotton-tipped swabs (Infant Mucus Extractor, Vygon Pharmaceutiques, Ecouen, France). The two swabs were then inserted into one vial containing 2 ml of virus transport medium (Gly medium).

Real-time PCR

Semi-quantitative real-time PCR was conducted on both the NS and NPA for respiratory syncytial virus (RSV), rhinovirus, human metapneumovirus, adenovirus, corona-virus, influenza, parainfluenza virus (type 1, 3), parainfluenza (type 2, 4), bocavirus, Mycoplasma pneumoniae and Chlamydia pneumoniae. Nucleic acids were extracted using the QIAamp DSP virus kit (QIAGEN, Valencia, CA, USA). Each sample was eluted in 200 μl buffer. cDNA was synthesised by using MultiScribe reverse transcriptase (RT) and random hexamers (both from Applied Biosystems, Foster City, CA, USA) [14]. Each 100-μl reaction mixture contained 60 μl cDNA mix and 40 μl of eluted RNA. After incubation for 10 min at 25°C, RT was carried out for 30 min at 48°C, followed by RT inactivation for 5 min at 95°C [14]. Samples were assayed in a 50-μl reaction mixture containing 20 μl (c)DNA and a 30-μl mix of the forward and reverse primers and probes. All samples had been spiked before extraction with an internal control virus (murine encephalomyocarditis virus [RNA] and phocine herpes virus [DNA]). The amplification and detection were performed by the use of an ABI Prism 7700 sequence-detection system; 2 min at 50°C to acquire optimal AmpErase UNG activity and 10 min at 95°C to activate AmpliTaq Gold DNA polymerase, followed by 45 cycles of 15 s at 95°C and 1 min at 60°C [15]. Primers and probes for the real-time PCR detection of RSV, influenza virus, parainfluenza virus and adenovirus are available from van de Pol et al. [16] and real-time PCR assays were performed as described previously [17–19].

Statistical analysis

Similar to previous studies, a consensus standard was used to assess the sensitivity of each testing method: a positive result in either the NPA or NS was considered as the gold standard for the presence of a pathogen and was used to calculate the sensitivity of the NPA and NS for the detection of the respiratory pathogens. The Chi-square test was used and a logistic regression analysis was performed. The outcome variable was defined as the sample being positive. Variables used in the model were age, gender, symptom score and multiple virus detection. Statistical significance was concluded if the p-value was <0.05. The statistical analyses were performed in STATA 10.0 (StataCorp LP, College Station, TX, USA).

Results

A total of 163 respiratory pathogens were identified in 94 children. The majority of children (73.5%) were ill for less than 4 days at the time of sampling. The median age at primary infection was 104 days (range 33–269) and the median score of symptoms was 3, indicating a mild illness (Table 1). In all children with an illness of 5 days or longer, one or more pathogens were detected.
Multiple pathogens in half of the children

In 49 children (50%), one pathogen was detected: 29 rhinovirus, 11 RSV, four coronavirus, two hMPV, two parainfluenza virus type 2 and 4, and one bocavirus. In 25 children, two pathogens were detected, of which 24 (96%) were rhinovirus with RSV. In 20 children, more than two pathogens were detected during the first episode of respiratory symptoms (three pathogens: \( n=17 \); four pathogens: \( n=2 \); five pathogens: \( n=1 \)). Co-infection rates by pathogen were: rhinovirus (57%), hMPV (60%), RSV (72%), coronavirus (71%) and bocavirus (91%).

Sensitivity of nasal swab is lower than the aspirate for RSV and rhinovirus

Rhinovirus was found the most frequently (\( n=67 \)), followed by RSV (\( n=39 \)) and coronavirus (\( n=14 \)) (Table 2).

No influenza viruses or parainfluenza type 1 and 3 viruses were detected. The sensitivity for detecting any pathogen of the NPA was 92% (CI\(_{95\%}\) 86.7–95.7), whereas the sensitivity of the NS was lower at 67% (CI\(_{95\%}\) 59.1–74.0). For the detection of RSV and rhinovirus, the sensitivity of the NS was lower than the NPA (Table 2).

Sensitivity of nasal swab depends on viral load

The sensitivity values of the NPA and NS were investigated in more detail for rhinovirus and RSV (Table 3). For children with a low symptom score, the sensitivity of the NS was lower than the NPA. The NS had a lower sensitivity than the NPA for the 30–40 cycle threshold (CT) values. To assess whether the sensitivity of the NS differed by age group, gender, multiple pathogens, symptom score and CT value, Chi-squared tests were performed. The sensitivity of the NS for the detection of RSV was related to the symptom score (\( p=0.001 \)) and the sensitivity

| Characteristics                  | All infants (\( N=98 \)) | Rhinovirus detected (\( n=67 \)) | RSV detected (\( n=39 \)) |
|----------------------------------|--------------------------|----------------------------------|---------------------------|
| Median age in days (range)       | 104 (33–269)             | 99 (33–269)                      | 115 (51–269)              |
| Median days of illness at time sampling (range) | 3.5 (2–31)\(^c\) | 4 (2–16)                          | 4 (2–12)                  |
| % male                           | 58%                      | 60%                              | 62%                       |

The values represent percentages, unless indicated otherwise

\(^a\) Reported by parents

\(^b\) \( N=97 \)

\(^c\) Duration of illness: IQR=3–5
of the NS was related to the CT values for both RSV and rhinovirus ($p<0.001$).

In the logistic regression analysis, age and gender did not significantly predict the detection of RSV or rhinovirus. The symptom score predicted RSV detection in both the NPA (odds ratio [OR]: 1.21; CI95% 1.07–1.39) and the NS (OR: 1.28; CI95% 1.12–1.48), while an inverse relationship was observed for symptom score and rhinovirus detection in the two samples (OR: 0.87; CI95% 0.78–0.98). The presence of more than one pathogen predicted RSV (OR: 8.98; CI95% 3.03–26.7) or rhinovirus detection (OR: 3.66 CI95% 1.33–10.08) in the NPA. When the same analysis was performed as a backwards regression with $p<0.2$, the results did not change.

**Discussion**

This study illustrates that the proportion of infants where a respiratory pathogen is detected was high (96%) and co-infections were common. In 20 children, more than two pathogens were detected during the first episode of respiratory symptoms. Co-infections were observed frequently for RSV (72%), coronavirus (71%) and bocavirus (91%) in particular.

High rates of co-infection in young children have been described recently for childhood pneumonia, in particular, in children aged less than 12 months [20] and in children hospitalised with acute respiratory tract infection [21, 22]. The most frequently detected virus was RSV, followed by human bocavirus and rhinovirus [21, 23]. A common combination has been reported to be RSV and bocavirus [21]. Even though a high occurrence of co-infections has been reported, ranging from 14–16% [21, 22] to 27% [20], our study presents an even higher rate of co-infection. A possible explanation for this high co-infection rate may be related to the sampling of both nostrils for the NPA. Human bocavirus is a newly identified virus and has been detected in respiratory tract secretions in patients with acute respiratory symptoms in 2 to 19% of the samples [24]. Co-infection with another virus has been observed in 40% of bocavirus-positive children [25]. The frequent associations of bocavirus with other respiratory viruses might be explained by the persistence of bocavirus in the respiratory tract [25]. Furthermore, we investigated the sensitivity of the NPA and NS tested by a real-time PCR method. The sensitivity of the NPA was 92%, while for the NS, it was 67%. In particular for the detection of rhinovirus and RSV, the NS had a lower sensitivity (75% and 51%, respectively) compared to the NPA (97% and 100%, respectively). The sensitivity of the NS for RSV was 100% for children with high symptom scores. For both RSV and rhinovirus, viral load, indicated by the CT value, was the major determinant of the sensitivity of the NS in a dose-dependent fashion. The symptom score predicted RSV detection in both the NPA and the NS, while an inverse relationship was observed for symptom score and rhinovirus detection in the two samples.

The use of a swab has been considered as a suitable replacement in community-based research or epidemiological studies. The major advantage of a swab is that collection is less painful and more convenient than an aspirate, as no additional devices are needed [5]. These factors may outweigh some reduction in sensitivity. The advantage of molecular methods in the detection of respiratory viruses has been reported [26, 27] and Lambert et al. reported that using these methods seemed to overcome the previously observed sensitivity reduction when less invasive specimens were combined with the conventional laboratory methods [7]. With the recently

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**Table 2** Detection of respiratory pathogens and the sensitivity by sampling method

| Respiratory pathogen          | NPA ($n$) | NS ($n$) | Total | NPA Sensitivity | 95% CI* | NS Sensitivity | 95% CI* |
|------------------------------|-----------|----------|-------|----------------|--------|---------------|--------|
| Rhinovirus                   | 65        | 50       | 67    | 97%            | 89.6–99.6 | 75%            | 62.5–84.4 |
| RSV                          | 39        | 20       | 39    | 100%           | 91.0–100 | 51%            | 34.8–67.6 |
| Coronavirus                   | 13        | 10       | 14    | 93%            | 66.1–99.8 | 71%            | 41.9–91.6 |
| Bocavirus                    | 8         | 7        | 11    | 73%            | 39.0–94.0 | 64%            | 30.8–89.1 |
| Adenovirus                   | 9         | 6        | 11    | 82%            | 48.2–97.7 | 55%            | 23.4–83.3 |
| Parainfluenza type 2 and 4   | 9         | 9        | 11    | 82%            | 48.2–97.7 | 82%            | 48.2–97.7 |
| hMPV                         | 4         | 3        | 5     | 80%            | 28.4–99.5 | 60%            | 14.7–94.7 |
| *Mycoplasma pneumoniae*      | 2         | 2        | 4     | 67%            | 9.4–99.2  | 67%            | 9.4–99.2  |
| *Chlamydia pneumoniae*       | 1         | 2        | 3     | 50%            | 1.2–98.7  | 100%           | 15.8–100  |
| Total                        | 150       | 109      | 163   | 92%            | 86.7–95.7 | 67%            | 59.1–74.0 |

NPA: nasopharyngeal aspirate; NS: nasal swab; RSV: respiratory syncytial virus; hMPV: human metapneumovirus; CI: confidence interval

*A one-sided 97.5% confidence interval was used in case the sensitivity was 100%
developed flocked swabs, the sensitivity is even further improved and the flocked swabs have the advantage of being rapid and less traumatic for paediatric patients [28]. However, the sensitivity of the flocked swab in outpatients with respiratory tract infections may be lower than in hospitalised patients. Further studies are required considering different types of swabs and patient populations, and should test for a broad spectrum of respiratory pathogens.

Our findings demonstrated a lower sensitivity of the NS, in particular for RSV. Similar results were reported in other studies where conventional, non-amplification-based methods were used [8, 9]. Lambert et al. did not test for rhinovirus and this was the most frequently detected virus in our study and elsewhere [11, 29]. No influenza detections were found in our study. This is not explained by sampling bias, because most swabs were taken during

Table 3 Sensitivity of the NPA and NS for the detection of rhinovirus and RSV presented by age group, gender, symptom score, presence of multiple pathogens and CT-value of the NPA

| Respiratory pathogen | subgroups | NPA (n) | NTS (n) | Total (n) | NPA Sensitivity | 95% CI | NS Sensitivity | 95% CI |
|----------------------|-----------|---------|---------|-----------|----------------|-------|---------------|-------|
| **Rhinovirus**       | Age       | 1–3 months | 28 | 24 | 28 | 100% | 87.7–100 | 86% | 67.3–96.0 |
|                      |           | 3–6 months | 30 | 22 | 32 | 94% | 79.2–99.2 | 69% | 50.0–83.9 |
|                      |           | 6–12 months | 7 | 4 | 7 | 100% | 59.0–100 | 57% | 18.4–90.1 |
|                      | Gender    | Boy | 38 | 32 | 40 | 95% | 83.1–99.4 | 80% | 64.4–90.0 |
|                      |           | Girl | 27 | 18 | 27 | 100% | 82.1–100 | 67% | 46.0–83.5 |
|                      | Symptom score | 0 to 4 | 46 | 36 | 48 | 96% | 85.7–99.5 | 75% | 60.4–86.4 |
|                      |           | 5 to 10 | 15 | 11 | 15 | 100% | 78.2–100 | 73% | 44.9–92.2 |
|                      |           | over 11 | 4 | 3 | 4 | 100% | 39.8–100 | 75% | 19.4–99.4 |
|                      | Multiple pathogen | No | 29 | 25 | 29 | 100% | 88.1–100 | 86% | 68.3–96.1 |
|                      |           | Yes | 36 | 25 | 38 | 95% | 82.3–99.4 | 66% | 48.7–80.4 |
|                      | CT NPA | 0–20 | 3 | 3 | 3 | 100% | 29.2–100 | 100% | 29.2–100 |
|                      |           | 20–25 | 22 | 22 | 22 | 100% | 84.6–100 | 100% | 84.6–100 |
|                      |           | 25–30 | 17 | 15 | 17 | 100% | 80.4–100 | 88% | 63.6–98.5 |
|                      |           | 30–35 | 10 | 6 | 10 | 100% | 69.2–100 | 60% | 26.2–87.8 |
|                      |           | 35–40 | 10 | 2 | 10 | 100% | 69.2–100 | 20% | 2.5–55.6 |
|                      |           | 40–45 | 3 | 0 | 3 | 100% | 2.9–100 | 0% | 0–70.8 |
| **RSV**              | Age       | 1–3 months | 12 | 7 | 12 | 100% | 73.5–100 | 58% | 27.7–84.8 |
|                      |           | 3–6 months | 20 | 11 | 20 | 100% | 82.3–100 | 55% | 31.5–77.0 |
|                      |           | 6–12 months | 7 | 2 | 7 | 100% | 59.0–100 | 29% | 7.6–64.8 |
|                      | Gender    | Boy | 24 | 11 | 24 | 100% | 85.8–100 | 46% | 25.6–67.2 |
|                      |           | Girl | 15 | 9 | 15 | 100% | 78.2–100 | 60% | 32.3–83.4 |
|                      | Symptom score | 0 to 4 | 22 | 8 | 22 | 100% | 84.6–100 | 36% | 17.2–59.3 |
|                      |           | 5 to 10 | 10 | 5 | 10 | 100% | 69.2–100 | 50% | 18.7–81.3 |
|                      |           | over 11 | 7 | 7 | 7 | 100% | 59.0–100 | 100% | 59.0–100 |
|                      | Multiple pathogen | No | 11 | 10 | 11 | 100% | 71.5–100 | 91% | 58.7–99.8 |
|                      |           | Yes | 28 | 10 | 28 | 100% | 87.7–100 | 36% | 18.6–56.0 |
|                      | CT NPA | 0–20 | 4 | 4 | 4 | 100% | 39.8–100 | 100% | 39.8–100 |
|                      |           | 20–25 | 11 | 10 | 11 | 100% | 71.5–100 | 91% | 58.7–99.8 |
|                      |           | 25–30 | 4 | 4 | 4 | 100% | 39.8–100 | 100% | 39.8–100 |
|                      |           | 30–35 | 7 | 1 | 7 | 100% | 59.0–100 | 14% | 0.4–57.9 |
|                      |           | 35–40 | 10 | 0 | 10 | 100% | 69.2–100 | 0% | 0–30.8 |
|                      |           | 40–45 | 3 | 1 | 3 | 100% | 29.2–100 | 33% | 0.8–90.6 |

NPA nasopharyngeal aspirate; NS nasal swab; RSV respiratory syncytial virus; CT Cycle threshold value

*a One-sided 97.5% confidence interval was used in case sensitivity was 100%

*b The CT-value of the NPA was used as a reference to compare with NS, therefore the sensitivity of the NPA is 100% for all categories
the winter season, during which both RSV and influenza had their peak incidence. A possible reason may be related to the patient population and the small population size. Another study showed similar results, with rhinovirus and RSV being the most frequently detected [29].

There were a number of limitations of this study. Firstly, one limitation was the timing of sampling. For five cases, sampling occurred 10 days after the onset of illness. Since the viral shedding of RSV is highest between days 0 and 6, sampling should preferably occur in this period [30]. The high proportion of positive samples, however, indicates that this effect was not a major drawback of this study. Secondly, in this study, pain and discomfort of the collection of the samples was not assessed, but other studies provided reference to this [5]. Finally, it is unknown whether the order of obtaining the specimens may have resulted in a lower detection rate in the NS. It is possible that, by suctioning both nostrils for the NPA, the secretions with virus or viral nucleic acids were reduced. This corresponds with the finding that few mild cases were detected with the NS, as the sensitivity of the NS dropped with lower symptoms score and higher CT values. Because the order and nature of the sample collection was slightly different from the study performed by Lambert et al., the sensitivity of the NS may be an underestimate, and caution needs to be taken when interpreting the NS sensitivity.

RSV and rhinovirus were commonly detected in infants during primary respiratory infection, and co-infections occurred in about half of the children. The sensitivity of the NPA was higher than the NS, in particular for the detection of RSV and rhinovirus. Although the sensitivity of a method is important, one must also take into account the advantages that different sampling methods offer. The great advantage of the NS is that this method can be performed in outpatient settings without needing special equipment. The medical ethical committee of the Utrecht University Medical Centre (UUMC) approved the study protocol and written informed consent was obtained from the parents of the participating children.

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The medical ethical committee of the Utrecht University Medical Centre (UUMC) approved the study protocol and written informed consent was obtained from the parents of the participating children.

Conflicts of interest The authors declare that they have no conflicting interests in publishing this paper.

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