Impact of intracerebroventricular enzyme replacement therapy in patients with neuronopathic mucopolysaccharidosis type II

Joo-Hyun Seo, Motomichi Kosuga, Takashi Hamazaki, Haruo Shintaku, and Torayuki Okuyama

INTRODUCTION
Mucopolysaccharidosis II (MPS II) (Hunter syndrome, OMIM 309900) is a rare X-linked lysosomal storage disorder caused by a deficiency of iduronate-2-sulfatase (IDS).1 IDS is an essential enzyme for the catabolism of glycosaminoglycans (GAGs), such as heparan sulfate, dermatan sulfate.1 IDS mutations lead to progressive lysosomal accumulation of GAGs in many organs and tissues, resulting in a wide spectrum of symptoms, including psychomotor developmental delay, hepatosplenomegaly, joint contracture, obstructive respiratory disease, and cardiac dysfunction.1 Patients with MPS II exhibit a number of neurological and behavioral problems associated with HS accumulation, including aggression, hyperactivity, sleep disturbances, and progressive decline of language and cognitive ability.2 In patients with MPS II with central nervous system (CNS) involvement, severe cognitive impairment and progressive neurological decline are observed.1 Patients are categorized according to the presence (neuronopathic MPS II) or absence (attenuated MPS II) of CNS involvement, with approximately two-thirds of patients having neuronopathic MPS II.3 Neuronopathic MPS II can be further divided into two groups based on the genetic mutation: group MS is characterized by missense mutations and is presumed to have slight residual enzyme activity, and group NT is considered to have null type mutations, such as deletions, recombination with pseudogene, and nonsense mutations.4 The patients in group NT show a more rapid decline than do those in group MS.5

Enzyme replacement therapy (ERT) with human recombinant idursulfase (Elaprase®) has been used since 2006 for the treatment of MPS II. Idursulfase is administered intravenously once a week and has been used for more than 120 Japanese patients since its approval in Japan in 2007.6 However, because intravenously administered idursulfase cannot penetrate the blood-brain barrier, it cannot reach the cerebral parenchyma. As such, CNS involvement in patients with neuronopathic MPS II, such as psychomotor developmental delay and neurological regressive episode, cannot be improved with current intravenous ERT. To improve the CNS symptoms, including cognitive decline, enzyme preparations that can reach the cerebral parenchyma have been sought. Non-clinical evidence has shown that idursulfase administered intrathecally or intracerebroventricularly (i.c.v.) rapidly reaches the brain.6 Consequently, global clinical studies of intrathecal idursulfase in patients with MPS II have been conducted. A phase 1/2 study of intrathecal idursulfase in children with MPS II conducted in the US and the UK supported the continued development of intrathecal administration as a potential therapy for the cognitive impairment caused by MPS II.7

Idursulfase beta (Hunterase®), developed by GC Pharma for ERT in patients with MPS II, was approved in South Korea for intravenous
administration in 2012 and received orphan drug designation in the US in 2013. Idursulfase and idursulfase beta have identical amino acid sequences but differ slightly in their glycosylation patterns because they are produced in different cell lines using different manufacturing processes. However, the structure, biological activity, and pharmacokinetics of the enzymes are almost the same. A recent non-clinical study in a mouse model of MPS II demonstrated that i.c.v. ERT with idursulfase beta significantly reduced HS concentrations in cerebrospinal fluid (CSF) and produced improvements in biochemical, histological, and memory/learning function parameters. i.c.v. ERT can be an effective and safe therapy for treating CNS symptoms, as demonstrated for cerliponase alfa (Brineura®), an approved treatment for neuronal ceroid lipofuscinosis type 2 disease. However, i.c.v. administration of idursulfase beta has not been evaluated in humans. This article presents the 100-week results of a multicenter, open-label, phase 1/2 clinical study to evaluate the efficacy and safety of i.c.v. idursulfase beta in patients with MPS II.

RESULTS
Six patients with a confirmed diagnosis of severe MPS II having significant developmental delay (group MS or NT) provided informed consent and were enrolled in the study. All patients had tolerated ≥24 weeks of treatment with intravenous idursulfase before the start of this study. During the study, all patients received idursulfase beta for 100 weeks via a CSF reservoir implanted under the patient’s scalp for i.c.v. administration; intravenous administration of idursulfase was also continued throughout the study. One patient initially did not receive i.c.v. idursulfase beta owing to fever observed 1 day before the planned day of reservoir placement; however, this patient later re-consented and re-enrolled in the study. All six patients were included in both the full analysis set and the safety analysis set. All patients were Japanese and male, with a mean age (range) at screening of 42.2 (23–65) months (Table 1).

| Patient no. | Sex | Group by gene mutation | Details of gene mutation | Developmental retardation | Age (months) |
|-------------|-----|------------------------|--------------------------|--------------------------|--------------|
|             |     |                        |                          |                          | Diagnosis    | Screening   | 0 weeks of injection (baseline) | 100 weeks of injection |
| A           | M   | NT                     | c.2_3C>G                 | yes                      | 13           | 23          | 25                        | 48                     |
| B           | M   | NT                     | c.1349_1350insGA (p.D450Efsx12) | yes                      | 26           | 36          | 36                        | 59                     |
| C           | M   | NT                     | c.1349_1350insGA (p.D450Efsx12) | yes                      | 26           | 36          | 36                        | 59                     |
| D           | M   | NT                     | c.1139_1140insA (p.Y380X) | yes                      | 38           | 47          | 48                        | 70                     |
| E           | M   | MS                     | c.419G>T (p.G140V)       | yes                      | 28           | 65          | 65                        | 88                     |
| F           | M   | NT                     | c.814C>T (p.Q274*)       | yes                      | 39           | 47          | 48                        | 70                     |

Mean age 28.3 42.2 43.0 59.0 59.0 65.7

aGroup MS is characterized by missense mutations and is presumed to have slight residual enzyme activity; group NT is considered to have null mutations, such as deletions, recombination with pseudogene, and a nonsense mutation.

The mean overall developmental age (DA), determined by the Kyoto Scale of Psychological Development 2001 (KSPD), increased from the screening period (23.2 months) up to week 76 (28.8 months), then decreased slightly (27.3 months) at week 100. DAs in each patient were compared with 13 Japanese patients with neuronopathic MPS II treated with intravenous idursulfase (n = 13). In the one patient in group MS (patient E), the DA from the screening period up to week 52 was similar to the least-squares means of the historical control group but then increased up to week 100 (Figure 2A). In the patients in group NT (other than patient F), the DA either was similar to or increased above the least-squares means for the historical control group (Figure 2B). Patient F, in group NT, started the i.c.v. treatment at the age of 47 months. He had increases in overall DA from the screening period (36 months) up to week 76 (45 months), followed by a marked decrease at week 100 (23 months). However, because the person who conducted the KSPD commented that the patient was distracted and disconnected during the observation at week 100, the numerical DA value should be interpreted carefully.

The differences in overall DA from the screening period to week 100 in the six patients (patients A–F) receiving i.c.v. idursulfase beta were +16.0, +14.0, +4.0, −1.0, +5.0, and −13.0 months (Figure 3). The corresponding least-squares means of the DA increase in the historical control group (n = 13) were +8.0, −0.5, −0.5, −4.7, −3.2, and +10.0 months.
and ~4.7 months, respectively (Figure 3). The mean difference in the change in DA from the screening period to week 100 between the six patients and the historical control group was 5.1 months. In the comparison with the historical control group, the developmental improvement was confirmed in five of six patients (all except patient F) in the study group that received i.c.v. administration.

Through week 100, adverse events (AEs) occurring from placement of the reservoir for i.c.v. administration of idursulfase beta were observed in three patients (Table 2); all AEs were mild. Two AEs (pyrexia and restlessness) in one patient were suspected as being related to the procedure. AEs occurring after i.c.v. administration of idursulfase beta were observed in all patients. All patients reported mild (n = 6 patients) or moderate (n = 3 patients) AEs. The most common AEs reported were pyrexia, upper respiratory tract infection, and vomiting (n = 6 patients, 100% each). AEs suspected as being related to the study drug were observed in all six patients (vomiting [n = 6, 100%], pyrexia [n = 3, 50.0%], procedural nausea [n = 2, 33.3%], and blood bilirubin increase and urticaria [n = 1 each, 16.7%]). No deaths or discontinuations owing to AEs occurred during the study.

Eleven serious AEs (SAEs), all designated as being because of hospitalization or prolongation of hospitalization, were reported in four patients. One patient experienced six SAEs (three asthma, one vomiting, one respiratory syncytial virus infection, and one inguinal hernia); one patient experienced two SAEs (pyrexia and gastroenteritis norovirus); one patient experienced two SAEs (pyrexia and viral pharyngitis); and one patient had one SAE (caries tooth). Two SAEs (pyrexia and vomiting) were suspected as being related to the study drug. One 4-year-old patient (patient D) experienced vomiting after drug administration at week 68. Another 4-year-old patient (patient F) experienced moderate pyrexia and vomiting 6 h after the start of the first study drug administration. Because CSF examination showed mild inflammation, intravenous antibacterial infusion was administered. The patient’s fever was resolved after 2 days and he was discharged the next day. The pyrexia was reported as an SAE.

From the safety point of view, we measured anti-IDS antibodies in serum and CSF. Anti-IDS antibodies in serum were detected in three patients (A, C, and F) at baseline and in three patients (B, C, and F) at 24 and 52 weeks. These antibodies might be a result of previous intravenous idursulfase treatment, as all patients had been treated with idursulfase before and throughout this study. All patients were negative for anti-IDS antibodies in serum at week 100. Anti-IDS antibodies in CSF were not detected in all patients during this study.

**DISCUSSION**

This study was the first to evaluate the efficacy and safety of i.c.v. administration of idursulfase beta in Japanese patients with severe MPS II. At data cutoff, six patients had received idursulfase beta (maximum 30 mg) once every 4 weeks for 100 weeks. Five patients had a ≥50% decrease from baseline at weeks 52 and 100. CSF, cerebrospinal fluid; HS, heparan sulfate; MPS, mucopolysaccharidosis.
in the HS concentration relative to baseline was 62.6% at week 52 and 71.2% at week 100. In addition, all but one patient had a greater increase in DA at week 100 compared with historical controls, with an average difference for all six patients of +5.1 months. Thus, unlike intravenously administered idursulfase and idursulfase beta, which cannot penetrate the blood-brain barrier, i.c.v. administration of idursulfase beta may be expected to improve CNS manifestations.

The concentration of HS in CSF was selected as the primary endpoint of this study because HS accumulation is considered to be the cause of CNS symptoms, such as psychomotor developmental delay, in patients with MPS II. Impairment in the morphogenesis of nerve cell dendrites and inflammatory nerve cell death owing to the accumulation of the GM2 ganglioside, as well as neurodegeneration-mediated oxidative stress, have been reported as mechanisms underlying CNS symptoms resulting from HS accumulation. It has been suggested that patients with MPS II with cognitive disorders tend to have high HS concentrations in the CSF. Furthermore, the concentrations of HS in CSF correlated with those in brain tissues of the mice treated by i.c.v. administration of idursulfase beta. These results suggest that HS concentrations in CSF reflect HS concentrations in brain tissue. The precise mechanism of the enzyme transfer from CSF to brain parenchyma is still unclear, and short-time injection may have facilitated driving idursulfase beta from cephalad toward the brain parenchyma. HS concentrations in CSF are an informative biomarker for evaluating neurodegeneration in MPS II.

As sufficient data of HS concentrations in CSF of normal children are not available, we only evaluated the decrease of HS concentration in CSF in each treated patient. In this study, monthly i.c.v. administration of idursulfase beta reduced the HS concentration in the CSF at week 52 in all patients, which was maintained through week 100. Recently, it was reported that weekly intravenous administration of JR-141, a new IDS fused with anti-human transferrin receptor antibody, for 3 weeks reduced HS concentrations in the CSF of patients with MPS II; however, the study did not have DA data, and the reduction in the HS concentration in the CSF was lower (25.1% and 31.5% in patients treated with 1 [n = 6] or 2 [n = 5] mg/kg/week, respectively) than in this study. More research is needed to confirm whether HS concentrations could be directly correlated with a reduction in CNS symptoms.

DA was set as the secondary endpoint to evaluate the effect of i.c.v. idursulfase beta on psychomotor developmental delay in pediatric patients. To date, only one other study directly evaluated the improvement in CNS symptoms in patients with MPS II who were administered intrathecal idursulfase. This study showed that monthly i.c.v. administration of idursulfase beta maintained or increased DA in five of six patients compared with the historical control group receiving intravenous idursulfase. At 100 weeks (about 2 years) after starting this study, six patients who received i.c.v. idursulfase beta had a 5.1-month increase in mean DA compared with 13 historical control patients who received only intravenous administration of idursulfase. Comparing patient ages when starting i.c.v. idursulfase beta, three patients who started at age <3 years increased DA by 9.0 months, whereas the other three patients who started at age ≥3 years increased DA by only 1.2 months. i.c.v. idursulfase beta may be expected to improve DA in patients at an early age when DA is still...
increasing; for older patients in whom DA is starting to decrease, i.c.v. idursulfase beta could increase DA slightly or slow the rate of DA decline. This suggests the importance of early initiation of i.c.v. administration because of the irreversible nature of CNS disease. In general, the first step in the diagnosis of MPS II is to identify the clinical manifestations characteristic of the disorder.19 However, children begin to present these characteristic manifestations at 3–4 years of age, by which time intellectual retardation is already advanced. In exceptional cases, children with a family history may be diagnosed before disease onset.20 Recently, neonatal screening for MPS II initiated in some countries and territories has increased the number of children diagnosed before intellectual retardation occurs.21,22 Thus, i.c.v. administration of idursulfase beta may be initiated for children before progression of intellectual retardation, and this will be a new treatment option for patients.

On the basis of the frequency and severity of AEs, including AEs associated with placement of the implantable reservoir, idursulfase beta was well tolerated during the long-term study period. This may be related to short-time (1 min) injection, which may have helped to avoid the risk of bacterial infection. The CSF reservoir we used in this study was a well-practiced device, and we did not have any trouble related to the device. From a safety point of view, short-time injection and the CSF reservoir were good methods and materials.

Our study has some limitations that must be acknowledged. As MPS II is a rare disease, the number of eligible patients was limited. Early treatment affects prognosis and the patient’s subsequent medical condition. Consequently, the study was designed to be open label with no placebo control group, because of an acceptable risk/benefit ratio as with device placement and i.c.v. administration of an inactive comparator, although a historical control group receiving intravenous idursulfase was used for DA comparison. Additionally, we used the KSPD for assessment of DA. This is another limitation of our study because KSPD is only applicable to Japan, although it is strongly correlated with the Bayley III,23 which is applicable to many other countries.

In conclusion, monthly i.c.v. administration of idursulfase beta using the implantable CSF reservoir for 100 weeks in Japanese pediatric patients with severe MPS II reduced CSF HS concentrations, maintained DA, and appeared to be well tolerated. These results suggest that i.c.v. idursulfase beta penetrates the brain and improves CNS manifestations. This ongoing study will further evaluate the long-term efficacy and safety outcomes associated with idursulfase beta; however, the results obtained to date support the continued development of idursulfase beta as a potential therapy for MPS II.

MATERIALS AND METHODS

Study design and participants

This was a phase 1/2, open-label, non-controlled, investigator-initiated clinical study (JMACCT CTR JMA-IIA00350, date of registration June 4, 2018). A placebo control was not considered because of an unacceptable risk/benefit ratio as with device placement and i.c.v. administration of an inactive comparator. The study was initiated at two clinical sites in Japan in July 2016 and is ongoing until March 2021. This analysis includes data collected up to February 2021.
2019 (100-week data cutoff). The study was conducted in compliance with the Declaration of Helsinki and the International Council for Harmonisation Guideline for Good Clinical Practice. The protocol and patient informed consent form were reviewed and approved by the Institutional Review Boards of the National Center for Child Health and Development and the Osaka City University Graduate School of Medicine. All legally acceptable representatives of patients provided signed informed consent at the screening period.

Male patients aged 1.5 to <15 years with a confirmed diagnosis of severe MPS II having significant developmental delay (group MS or NT) were eligible for this study if they had never received hematopoietic stem cell transplantation (HSCT) and tolerated ≥24 weeks of treatment with intravenous idursulfase (administered ≥20 times during the 24 weeks before the start of this study). Confirmed diagnosis of severe MPS II was defined as IDS activity in leukocytes that is low or below the quantitation limit, a urinary uronic acid value that exceeds the reference value, and having genetic variants observed exclusively in neuronopathic MPS II.24–26 Patients were excluded for any of the following: prior HSCT, previous intrathecal administration of idursulfase, urinary uronic acid level ≥50-fold the upper limit of the reference value by age, ventricular/intraperitoneal shunt, end-stage organ dysfunction or other serious diseases, malignant neoplasm, participation in other clinical studies within 6 months before the study start, or history of anaphylactic shock from any component of the study drug.

### Procedures

For enrolled patients, a CSF reservoir was implanted under the patient’s scalp for i.c.v. administration. Idursulfase beta i.c.v. injection (15 mg/mL), provided by GC Pharma (Yongin, South Korea), was used for this study. The appropriate dose of idursulfase beta was diluted with normal saline under sterile conditions, and 2 mL was administered for 1 min. The first dose was administered in a hospital setting to allow detailed observation of the patient. Idursulfase beta was i.c.v. administered once every 4 weeks: 1 mg at weeks 0 (baseline) and 4; 10 mg at weeks 8 and 12; 30 mg at weeks 16, 20, and 24; and 42 administrations from weeks 28 to 100 at the final dose as decided by the data monitoring committee based on changes in the HS concentration in the CSF and the safety evaluation (Figure 4). CSF samples were drawn immediately before administration of each idursulfase beta i.c.v. dose. Intravenous administration of idursulfase (0.5 mg/kg/week) was continued throughout the study; an interval of ≥24 h was set between intravenous idursulfase and i.c.v. idursulfase beta. In order to evaluate the efficacy and safety, HS concentration, urinary uronic acid, biochemistry tests, vital signs, immunogenicity tests (IDS antibody, anti-IDS antibody), a head computed tomography or magnetic resonance imaging scan, AEs, KSPD, and other tests were measured and checked for all patients at regular intervals during the clinical trial period (Table 3).

### Outcomes

The primary endpoint was the HS concentration in the CSF. CSF samples were collected every 4 weeks from baseline and every 8 weeks from week 28 prior to each drug administration, and HS concentrations were measured by Toray Research Center (Kamakura, Japan).27,28 The CSF sample (20 μL) was transferred to a glass tube and evaporated to dryness under nitrogen at 40°C. Methanolic hydrochloric acid (3 M, 200 μL) and 2,2-dimethoxy propane (20 μL) were added, and the solution was mixed, sonicated, heated (at 65°C for 75 min), and evaporated. The residue was reconstituted with 200 μL of internal standard working solution and further mixed and sonicated. The solution was centrifuged at 10,000 × g at 4°C for 3 min, and the resultant filtrate was placed into an autosampler vial and analyzed. A UPLC® system (Waters, Milford, MA, USA) and a triple quadrupole mass spectrometer API5000 (AB SCIEX, Framingham, MA, USA) were used for liquid chromatography with tandem mass spectrometry analysis. Analyst v1.5.1 (AB SCIEX, Framingham, MA, USA) was used for data processing.

The secondary endpoint was DA determined by the KSPD, which is an individualized face-to-face test to assess a child’s development.

Table 2. Summary of AEs

| AE, adverse event; n, number of patients with AE. | Preferred term | 52 weeks n (%) | 100 weeks n (%) |
|-----------------------------------------------|----------------|----------------|-----------------|
| Total N                                       | 6              | 6              |
| AEs associated with placement procedure of implantable reservoir | 3 (50.0)       | 3 (50.0)       |
| constipation                                  | 1 (16.7)       | 1 (16.7)       |
| contusion                                     | 1 (16.7)       | 1 (16.7)       |
| post-procedural haemorrhage                   | 1 (16.7)       | 1 (16.7)       |
| pyrexia                                       | 1 (16.7)       | 1 (16.7)       |
| restlessness                                  | 1 (16.7)       | 1 (16.7)       |
| wheezing                                      | 1 (16.7)       | 1 (16.7)       |
| AEs after study drug administration            | 6 (100.0)      | 6 (100.0)      |
| pyrexia                                       | 5 (83.3)       | 6 (100.0)      |
| upper respiratory tract infection             | 5 (83.3)       | 6 (100.0)      |
| vomiting                                      | 5 (83.3)       | 6 (100.0)      |
| eczema                                        | 3 (50.0)       | 4 (66.7)       |
| gastroenteritis                               | 2 (33.3)       | 4 (66.7)       |
| urticaria                                     | 3 (50.0)       | 3 (50.0)       |
| arthropod sting                               | 0 (0.0)        | 2 (33.3)       |
| blood urine present                           | 2 (33.3)       | 2 (33.3)       |
| dermatitis diaper                             | 1 (16.7)       | 3 (50.0)       |
| eye discharge                                 | 0 (0.0)        | 2 (33.3)       |
| faeces soft                                   | 1 (16.7)       | 2 (33.3)       |
| injection site extravasation                  | 2 (33.3)       | 2 (33.3)       |
| procedural nausea                             | 1 (16.7)       | 2 (33.3)       |

AE, adverse event; n, number of patients with AE.24–26 Patients were excluded for any of the following: prior HSCT, previous intrathecal administration of idursulfase, urinary uronic acid level ≥50-fold the upper limit of the reference value by age, ventricular/intraperitoneal shunt, end-stage organ dysfunction or other serious diseases, malignant neoplasm, participation in other clinical studies within 6 months before the study start, or history of anaphylactic shock from any component of the study drug.
in the following three areas: postural-motor, cognitive-adaptive, and language-social. The KSPD is a standardized tool that is widely used in Japan for developmental assessment in all age groups. The Bayley Scales of Infant Development III (BSID-III), which is used globally to assess the developmental/cognitive function of young children, has not been standardized in Japanese. It has been reported that the developmental quotients of the KSPD are strongly correlated with the corresponding composite score of the BSID-III.

Safety measures included incidence and severity of AEs, vital signs, electrocardiograms, and clinical laboratory tests. AEs were separately collected as: (1) AEs occurring from placement of the reservoir until i.c.v. administration of idursulfase beta, and (2) AEs occurring after i.c.v. administration of idursulfase beta. AEs were classified by system organ class and preferred term using the Medical Dictionary for Regulatory Activities version 18.1. Immunogenicity tests, including anti-idursulfase beta antibodies in serum and CSF, were performed using the BioNote IDS Ab ELISA kit (BioNote, Gyeonggi, South Korea).

**Statistical analysis**
A planned sample size of at least four patients was set in consideration of the number of eligible pediatric patients. The efficacy analysis set was defined as patients who had received idursulfase beta and for whom data were available for at least one time point after the first injection.

### Table 3. Schedule of clinical trial

| Tests                              | Time (weeks) |
|------------------------------------|--------------|
|                                    | –5 to –1    | 0  | 1  | 2  | 4  | 8  | 10 | 12 | 16 | 18 | 20 | 24 | 28 | 32 | 36 | 40 | 44 | 48 | 52 | +4 | 100 |
| Implantation of CSF reservoir      | X            |
| Administration                     | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |
| Injection dose (mg) b              | 1            | 1  | 10 | 10 | 30 | 30 | 30 | 30 | 30 | 30 | 30 | 30 | 30 | 30 | 30 | 30 | 30 | ~ 30 |
| HS concentration in CSF c          | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |
| Urinary uronic acid                | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |
| KSPD                               | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |
| Biochemistry tests                 | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |
| Vital signs                        | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |
| Immunogenicity tests               | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |
| Head CT or MRI scan                | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |
| Adverse events check               | X            | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  | X  |

*aScreening period.

bIdursulfase beta was intracerebroventricularly administered once every 4 weeks: 1 mg at weeks 0 (baseline) and 4; 10 mg at weeks 8 and 12; 30 mg at weeks 16, 20, and 24; and a final dose (all patients received 30 mg) from weeks 28 to 100.

cCSF samples were drawn immediately before administration.
administration of idursulfase beta. The safety analysis set included all patients who had received idursulfase beta. The primary endpoint, HS concentration in CSF, is presented at each time point for each patient and as the mean of all patients. In addition, the HS concentrations relative to baseline were calculated, and the percentage of patients whose HS values at week 100 decreased by >50% compared with baseline were reported. The secondary endpoint, DA in a total of three areas, is presented at each time point for each patient and as the mean of all patients. DAs in each patient were compared with 13 Japanese patients with neuronopathic MPS II treated with intravenous idursulfase, defined as patients with severe MPS II and etiological mutation of the IDS gene who received only intravenous idursulfase and had two or more KSPD evaluations collected before the study. Least-squares means calculated by a linear mixed-effects model in the reference group were used for comparison with patients in this study.

Data availability
The data for this study contain personal information that is not suitable for sharing in its current format. Appropriately de-identified datasets for the current study can be made available by the corresponding author upon reasonable request.

ACKNOWLEDGMENTS
The authors would like to thank all study participants. This research was supported by the Japan Agency for Medical Research and Development (AMED) under grant no. JP17lk0103012 and GC Pharma. The study drug used for this investigator-initiated study was provided by GC Pharma, manufacturer/licensee of idursulfase beta. Medical writing assistance funded by GC Pharma was provided by Hiroko Ebina, BPharm, Ph, MBA, CMPP and Rebecca Lew, PhD, CMPP of ProScribe – Envision Pharma Group. ProScribe’s services complied with international guidelines for Good Publication Practice (GPP3). The funder of the study had no role in the study design, data collection, data analysis, data interpretation, or writing of the report. The corresponding author had full access to all the data in the study and had final responsibility for the decision to submit for publication.

AUTHOR CONTRIBUTIONS
All authors were involved in the investigation and participated in the writing, review, editing, and approval of the final version of the manuscript. J.H.S. was involved in the data curation and formal analysis. T.O. was involved in the conceptualization, funding acquisition, and supervision.

DECLARATION OF INTERESTS
T.H. has received research grants from Amicus Therapeutics, JCR Pharmaceuticals, and Sanofi Genzyme. T.O. has received research grants from GC Pharma, Sanofi, and Sumitomo Dainippon and is the Principal Investigator for enzyme replacement therapy clinical trials for mucopolysaccharidosis II sponsored by GC Pharma and JCR Pharmaceuticals. The remaining authors declare no competing interests.

REFERENCES
1. Wraith, J.E., Scarpa, M., Beck, M., Bodamer, O.A., De Meirleir, L., Guffon, N., Meldgaard Lund, A., Malm, G., Van der Ploeg, A.T., and Zeman, J. (2008). Mucopolysaccharidosis type II (Hunter syndrome): a clinical review and recommendations for treatment in the era of enzyme replacement therapy. Eur. J. Pediatr. 167, 267–277.
2. Shapiro, E.G., Jones, S.A., and Escolar, M.L. (2017). Developmental and behavioral aspects of mucopolysaccharidoses with brain manifestations—neurological signs and symptoms. Mol. Genet. Metab. 122S, 1–7.
3. Stapleton, M., Kubaski, F., Mason, R.W., Yabe, H., Suzuki, Y., Orii, K.E., Orii, T., and Tomatsu, S. (2017). Presentation and treatments for mucopolysaccharidosis type II (MPS II; Hunter syndrome). Expert Opin. Orphan Drugs 5, 295–307.
4. See, J.-H., Okuyama, T., Shapiro, E., Fukushima, Y., and Kosuga, M. (2020). Natural history of cognitive development in neuronopathic mucopolysaccharidosis type II (Hunter syndrome): contribution of genotype to cognitive developmental course. Mol. Genet. Metab. Rep. 24, 100630.
5. Japanese Society for Inherited Metabolic Diseases (2019). Practical Guideline for the Management of Mucopolysaccharidosis (MPS) Type II 2019 [Japanese] (Shindan to Chiryu Sha).
6. Calias, P., Papisov, M., Pan, J., Savioli, N., Belov, V., Huang, Y., Lottherhand, J., Alessandrinii, M., Liu, N., Fischman, A.J., et al. (2012). CNS penetration of intra-thecal-lumbar idursulfase in the monkey, dog and mouse: implications for neurological outcomes of lysosomal storage disorder. PLoS ONE 7, e30341.
7. Muenzer, J., Hendriksz, C.J., Fan, Z., Visjayaraghavan, S., Perry, V., Santra, S., Solanki, G.A., Mascelli, M.A., Pan, L., Wang, N., et al. (2016). A phase II/II study of intrathecal idursulfase-IT in children with severe mucopolysaccharidosis II. Genet. Med. 18, 73–81.
8. Kim, C., Seo, J., Chung, Y., Ji, H.J., Lee, J., Sohn, J., Lee, B., and Jo, E.C. (2017). Comparative study of idursulfase beta and idursulfase in vitro and in vivo. J. Hum. Genet. 62, 167–174.
9. Sohn, Y.B., Ko, A.R., Seong, M.R., Lee, S., Kim, M.R., Cho, S.Y., Kim, J.S., Sakaguchi, M., Nakazawa, T., Kosuga, M., et al. (2018). The efficacy of intracerebroventricular idursulfase-beta enzyme replacement therapy in mucopolysaccharidosis II murine model: heparan sulfate in cerebrospinal fluid as a clinical biomarker of neuropathology. J. Inherit. Metab. Dis. 41, 1235–1246.
10. Schulz, A., Ajayi, T., Specchio, N., de Los Reyes, E., Gissen, P., Ballon, D., Dyke, J.P., Cahan, H., Slator, P., Jacoby, D., and Kohlshütter, A.; CLN2 Study Group (2018). Study of intraventricular cerliponase alfa for CLN2 disease. Neurology 81, 2522–2529.
11. Ikuzawa, M., Iwachidou, S., and Oogami, R. (2001). In The Guide of Kyoto Scale of Psychological Development 2001 [Japanese]., M. Ikuzawa, Y. Matsushita, and A. Tomatsu, eds. (Kyoto Kokusai Shakiakufukusho Center).
12. Ngu, L.H., Ong Peitee, W., Leong, H.Y., and Chew, H.B. (2017). Case report of treatment experience with idursulfase beta (Hunterase) in an adolescent patient with MPS II. Mol. Genet. Metab. Rep. 12, 28–32.
13. Hendriksz, C.J., Muenzer, J., Vandererve, A., Davis, J.M., Burton, B.K., Mendelsohn, N.J., Wang, N., Pan, L., Pato, A., and Barbier, A.J. (2015). Levels of glycosaminoglycans in the cerebrospinal fluid of healthy young adults, surrogate-normal children, and Hunter syndrome patients with and without cognitive impairment. Mol. Genet. Metab. Rep. 5, 103–106.
14. Öhmi, K., Greenberg, D.S., Rajavel, K.S., Ryazantsev, S., Li, H.H., and Neufeld, E.F. (2003). Activated microglia in cortex of mouse models of mucopolysaccharidoses I and IIIB. Proc. Natl. Acad. Sci. USA 100, 1902–1907.
15. Villani, G.R.D., Gargiulo, N., Farao2nio, R., Castaldo, S., Gonzalez Y Reyero, E., and Di Natale, P. (2007). Cytokines, neurotrophins, and oxidative stress in brain disease from mucopolysaccharidosis IIIB. J. Neurosci. Res. 85, 612–622.
16. Wallace, S.U., Siegel, D.A., and Dobrenis, K. (1995). GM2 ganglioside and pyramidal neuron dendritogenesis. Neurochem. Res. 20, 1287–1299.
17. Hendriksz, C.J., Muenzer, J., Burton, B.K., Pan, L., Wang, N., Naimy, H., Pano, A., and Barbier, A.J. (2015). A cerebrospinal fluid collection study in pediatric and adult patients with Hunter syndrome. J. Inborn Errors Metab. Screen. 3, 1–5.
18. Okuyama, T., Eto, Y., Sakai, N., Minami, K., Yamamoto, T., Sonoda, H., Yamaoka, M., Tachibana, K., Hirato, T., and Sato, Y. (2019). Iduronate-2-sulfatase with anti-human transferrin receptor antibody for neuropathic mucopolysaccharidosis II: a phase 1/2 trial. Mol. Ther. 27, 456–464.

19. Scarpa, M., Almássy, Z., Beck, M., Bodamer, O., Bruce, I.A., De Meirleir, L., Guffon, N., Guillin-Navarro, E., Hensman, P., Jones, S., et al.; Hunter Syndrome Europena Expert Council (2011). Mucopolysaccharidosis type II: European recommendations for the diagnosis and multidisciplinary management of a rare disease. Orphanet J. Rare Dis. 6, 72.

20. Tajima, G., Sakura, N., Kosuga, M., Okuyama, T., and Kobayashi, M. (2013). Effects of idursulfase enzyme replacement therapy for mucopolysaccharidosis type II when started in early infancy: comparison in two siblings. Mol. Genet. Metab. 108, 172–177.

21. Chuang, C.K., Lin, H.Y., Wang, T.J., Huang, Y.H., Chan, M.I., Liao, H.C., Lo, Y.T., Wang, L.Y., Tu, R.Y., Fang, Y.Y., et al. (2018). Status of newborn screening and follow up investigations for mucopolysaccharidoses I and II in Taiwan. Orphanet J. Rare Dis. 13, 84.

22. Donati, M.A., Pasquini, E., Spada, M., Polo, G., and Burlina, A. (2018). Newborn screening in mucopolysaccharidoses. Ital. J. Pediatr. 44 (Suppl 2), 126.

23. Kono, Y., Yonemoto, N., Kusuda, S., Hirano, S., Iwata, O., Tanaka, K., and Nakazawa, J. (2016). Developmental assessment of VLBW infants at 18 months of age: A comparison study between KSPD and Bayley III. Brain Dev. 38, 377–385.

24. Kosuga, M., Mashima, R., Hirakiyama, A., Fuji, N., Kumagai, T., Seo, J.H., Nikado, M., Saito, S., Ohno, K., Sakuraba, H., and Okuyama, T. (2016). Molecular diagnosis of 65 families with mucopolysaccharidosis type II (Hunter syndrome) characterized by 16 novel mutations in the IDS gene: genetic, pathological, and structural studies on iduronate-2-sulfatase. Mol. Genet. Metab. 118, 190–197.

25. Sohn, Y.B., Ki, C.-S., Kim, C.-H., Ko, A.-R., Yook, Y.-J., Lee, S.-J., Kim, S.I., Park, S.W., Yeau, S., Kwon, E.-K., et al. (2012). Identification of 11 novel mutations in 49 Korean patients with mucopolysaccharidosis type II. Clin. Genet. 81, 185–190.

26. Vafadaki, E., Cooper, A., Heptinstall, L.E., Hatton, C.E., Thornley, M., and Wraith, J.E. (1998). Mutation analysis in 57 unrelated patients with MPS II (Hunter’s disease). Arch. Dis. Child. 79, 237–241.

27. Auray-Blais, C., Lavoie, P., Zhang, H., Gagnon, R., Clarke, J.T.R., Maranda, B., Young, S.P., An, Y., and Millington, D.S. (2012). An improved method for glycosaminoglycan analysis by LC-MS/MS of urine samples collected on filter paper. Clin. Chim. Acta 413, 771–778.

28. Zhang, H., Young, S.P., Auray-Blais, C., Orchard, P.J., Tolar, J., and Millington, D.S. (2011). Analysis of glycosaminoglycans in cerebrospinal fluid from patients with mucopolysaccharides by isotope-dilution ultra-performance liquid chromatography-tandem mass spectrometry. Clin. Chem. 57, 1005–1012.