Effect of the IL-1 Receptor Antagonist Kineret<sup>®</sup> on Disease Phenotype in *mdx* Mice

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Abstract

Duchenne muscular dystrophy (DMD) is an X-linked muscle disease caused by mutations in the dystrophin gene. The pathology of DMD manifests in patients with progressive muscle weakness, loss of ambulation and ultimately death. One of the characteristics of DMD is muscle inflammation, and dystrophin-deficient skeletal muscles produce higher levels of the pro-inflammatory cytokine interleukin 1β (IL-1β) in response to toll like receptor (TLR) stimulation compared to controls; therefore, blocking the IL-1β pathway could improve the disease phenotype in *mdx* mice, a mouse model of DMD. Kineret<sup>®</sup> or IL-1Ra is a recombinant IL-1 receptor antagonist approved by the FDA for treating rheumatoid arthritis. To determine the efficacy of IL-1Ra in a DMD model, we administered subcutaneous injections of saline control or IL-1Ra (25 mg/kg/day) to *mdx* mice daily for 45 days beginning at 5 weeks of age. Functional and histological parameters were measured at the conclusion of the study. IL-1Ra only partially inhibited this signaling pathway in this study; however, there were still interesting observations to be noted. For example, although not significantly changed, splenocytes from the IL-1Ra-treated group secreted less IL-1β after LPS stimulation compared to control mice indicating a blunted response and incomplete inhibition of the pathway (37% decrease). In addition, normalized forelimb grip strength was significantly increased in IL-1Ra-treated mice. There were no changes in EDL muscle-specific force measurements, histological parameters, or motor coordination assessments in the dystrophic mice after IL-1Ra treatment. There was a significant 27% decrease in the movement time and total distance traveled by the IL-1Ra treated mice, correlating with previous studies examining effects of IL-1 on behavior. Our studies indicate partial blocking of IL-1β with IL-1Ra significantly altered only a few behavioral and strength related disease parameters; however, treatment with inhibitors that completely block IL-1β, pathways upstream of IL-1β production or combining various inhibitors may produce more favorable outcomes.
Introduction

Duchenne muscular dystrophy (DMD) is an X-linked muscle disease characterized by inflammation and fibrosis in the skeletal muscles which results from constant cycles of muscle degeneration and regeneration [1–3]. DMD is a progressive muscle weakness disease which causes loss of ambulation by the teenage years and mortality by the third decade of life usually due to cardiovascular complications. Despite the severity of this disease, there are only limited treatment options for DMD patients with the current standard-of-care regimen being glucocorticoids (GCs) [4–6]. These drugs have been demonstrated to delay the onset of symptoms associated with DMD; however, GCs have many side effects in patients, highlighting the need for safer and more effective alternative therapies. The mdx mouse model, identified via a spontaneous mutation in the dystrophin gene, exhibits some of the hallmark pathologies of DMD [3,7,8]. In this model, inflammation develops in both the limbs and diaphragm at 3 weeks of age, with a peak at about age 8–10 weeks, before diminishing in the limbs but not the diaphragm [9]. Pre-clinical testing has demonstrated that anti-inflammatory drugs improve the mdx muscle phenotype and therefore have the potential to alleviate inflammatory pathways in DMD patients [10–13].

Inflammatory cytokines play a major role in the DMD phenotype and these include factors like tumor necrosis factor alpha (TNFα) and interleukin 1 beta (IL-1β). Expression of TNFα in mdx mice has been well characterized and shown to be increased with age in the diaphragm muscle where inflammation is usually high in this model [14]. Many studies have been performed to block this signaling at various levels to improve the dystrophic phenotype by reducing necrosis, degeneration and contraction-induced injury [14–22]. Because TNFα had already been examined extensively, another cytokine of particular interest to target in DMD is IL-1β. It has previously been shown that IL-1β plays a role in the initiation and perpetuation of muscle pathology in both DMD and limb girdle muscular dystrophy 2B (LGMD2B) patients [23]. In addition, IL-1β mRNA levels are higher in mdx mice than in controls, and reducing both the expression and activity of IL-1β could potentially treat muscle inflammation [23]. IL-1β is secreted as a precursor protein and becomes biologically active after undergoing proteolytic cleavage by caspase-1 [24]. IL-1β mediates signaling via the interleukin 1 receptor (IL-1R) and downstream activation of the nuclear factor kappa B (NFκB) pathway. Interestingly, NFκB activity has been previously shown to be elevated in the muscle of mdx mice [23]. Conversely, blocking NFκB activity has been shown to reduce the inflammatory response and IL-1β levels in both DMD patients and mdx mice [11,12,25,26]. These effects are similar to those seen in patients on GCs and can potentially be used in a combinatorial manner to reduce muscle inflammation even further [5,6,27–31].

The interleukin 1 receptor antagonist (IL-1Ra) is a naturally occurring cytokine that inhibits the binding of IL-1β to IL-1R. IL-1Ra lacks the binding domain necessary for recruiting the IL-1R accessory protein to the receptor complex; therefore, preventing downstream pro-inflammatory signaling. A synthetic form of IL-1Ra, anakinra (Kineret®), is a recombinant and nonglycosylated form of human IL-1Ra that has been granted approval for use in arthritis by the Food and Drug Administration (FDA). Anakinra exerts its physiological effects in a similar manner to the naturally occurring antagonist, by competitively binding to the IL-1R and neutralizing the effects of IL-1β. The protective role of anakinra in many diseases, including those affected by inflammation, makes this compound attractive for the treatment of inflammatory diseases of the muscle, such as myositis and DMD. Since inflammation plays a detrimental role in DMD, the high level of IL-1β in the muscles of DMD patients and mdx mice make this pathway an attractive target for reducing the muscle pathology in these affected individuals. We
hypothesized that the inflammatory effects of DMD would be reduced by treatment with anakinra and that, in vivo, mdx mice would have improved muscle function.

Materials and Methods

Animal care

All animal work was conducted in accordance with guidelines for the care and use of laboratory animals provided by the National institutes of health and protocols were approved by the Children’s National Institutional Animal Care and Use Committee (IACUC) (Protocol #304-13-04). For surgery, all animals were anesthetized using ketamine/xylazine and euthanasia was performed while under anesthesia using cervical dislocation. Four-week-old female C57BL/10ScSn-Dmd\(^{mdx}\)/J (mdx) mice weighing 10–18 g were purchased from The Jackson Laboratory (Bar Harbor, ME). Mice were housed in individually ventilated cage system with a 12-h light-dark cycle and received standard mouse chow and water ad libitum. Mice were rested at least 7 days before treatment and treatment began when the mice were 5 weeks old. All functional measures were acquired in a blinded manner.

Study design

IL-1Ra (anakinra, trade name Kineret\(^{\text{TM}}\)) was a gift from Amgen Inc. (Thousand Oaks, CA). Our study involved daily subcutaneous injections of two groups of animals: (a) a control mdx group, dosed with 0.9% NaCl (n = 6), and (b) a drug-treated mdx group, which received IL-1Ra at 25 mg/kg/day in a 50-μL volume of 0.9% NaCl (n = 10). Mice were randomized on the basis of body mass and were treated for 45 days, beginning at 5 weeks of age ending when they were 12 weeks old. IL-1Ra and the vehicle were injected subcutaneously, 7 days a week.

Enzyme-linked immunosorbent assay (ELISA)

Primary splenocytes were isolated at necropsy from mdx mice that had been treated with IL-1Ra or saline; the cells were then maintained for 24 h in vitro. The medium was changed, and the splenocytes were treated for 24 h with lipopolysaccharide (LPS) or LPS-free medium. IL-1 was measured in the medium from the splenocytes. IL-1 was quantified using a Quantikine IL-1 ELISA kit (R&D Systems, Minneapolis, MN) according to the manufacturer’s instructions. N = 4 for each group analyzed.

Rotarod test

Rotarod tests were performed as described previously [32,33]. In brief, mice were trained for 2 days prior to being tested twice a day for 3 consecutive days according to the following parameters: 10 rpm for 60s (the stabilization period), followed by acceleration from 10 rpm to 40 rpm (reached within the first 25s), totaling 240 s. Latency to fall (in seconds, s) was recorded, and six scores were averaged for each mouse. N = 6 saline, n = 10 IL-1Ra treated.

Grip strength testing

Grip strength was assessed using a grip strength meter as previously described [26,33]. Five successful hindlimb and forelimb strength measurements within 2 min were recorded. The maximum values for each day over a 5-day period were used for subsequent analysis, and the data were normalized to body mass and expressed as kilogram force (KGF). N = 6 saline, n = 10 IL-1Ra treated.
In vitro force measurements

Maximal force (mN) and twitch force (mN) generated by the extensor digitorum longus (EDL) muscle from the right hind limb was measured using a force transducer (Aurora Scientific, Aurora, Ontario, Canada) as previously described [11]. Specific force was calculated by dividing the maximal force generated by the EDL muscle by the cross-sectional area of the muscle (kN/m²). N = 4 saline, n = 6 IL-1Ra treated.

Histological evaluations

Mice were anesthetized using ketamine followed by cervical dislocation in order to prevent animal suffering. Muscles were harvested and a portion of each dissected muscle (e.g., gastrocnemius, diaphragm, EDL, or heart) was placed in formalin for paraffin embedding. These tissues were later sectioned and stained with hematoxylin and eosin (H&E). The remaining portion of each tissue was embedded in Tissue-Tek optimal cutting temperature (O.C.T.) compound, (Sakura Finetek USA, Torrance, CA) and frozen in liquid nitrogen chilled isopentane for cryosectioning. Tissues were imaged under a light microscope with a 20X objective, and a digital image was obtained using computer software (Olympus C.A.S.T. Stereology System, Olympus America Inc., Center Valley, PA). The digital images were loaded into Image J (NIH) with additional plug-ins to count the cells. The total number of cells, centralized nuclei, peripheral nuclei, and cells with centralized nuclei were counted and analyzed for comparison between treatment groups. Fibers showing degeneration (as defined by a loss of striations and a homogeneous appearance of the fiber contents), regeneration (as defined by a basophilic cytoplasm and large peripheral or central nuclei with prominent nucleoli), and inflammatory foci per field were assessed in a blinded fashion as described previously [33].

Behavioral activity measurement

Open-field activity was measured using an open-field Digiscan apparatus (Omnitech Electronics, Columbus, OH) as described previously [33]. In brief, all mice were acclimated for 60 min daily in the week prior to data collection. Data were collected every 10 min over a 1-h period each day for 4 consecutive days. Results were calculated as mean ± standard error of the mean (SEM) of all recordings. N = 6 saline, n = 10 IL-1Ra treated.

Statistical analysis

Statistical analyses comparing two groups at a time were performed using parametric, unpaired, two-tailed, t-tests (Graph Pad, Prism software). P-values ≤ 0.05 were considered statistically significant. Values in the graphs in the figures represent Values in the graphs represent mean ± standard error of the mean (SEM).

Results

IL-1Ra blunted the effect of LPS on IL-1 secretion in splenocytes from mdx mice

To test whether IL-1Ra is effective in vivo in inhibiting IL-1 secretion, splenocytes from mdx mice treated with IL-1Ra or saline (control) were stimulated with lipopolysaccharide (LPS). The IL-1 secretion was comparable in the unstimulated splenocytes from the saline-treated mice and the IL-1Ra-treated mice, indicating that IL-1Ra treatment alone did not alter the basal level of IL-1 secretion in the mdx mice (Fig 1). As expected, LPS stimulation significantly increased IL-1 secretion in saline-treated mdx mice. There was also a significant increase in IL-
1Ra-treated splenocytes challenged with LPS. Although it was not statistically significant, it is important to note there was 37% decrease in IL-1 secretion from LPS-stimulated splenocytes of IL-1Ra-treated \textit{mdx} mice when compared to those from saline-treated control \textit{mdx} mice (Fig 1). Therefore, LPS-induced IL-1 secretion was blunted in the splenocytes from mice treated with IL-1Ra when compared to those receiving saline indicating IL-1Ra only partially inhibited this pathway.

**IL-1Ra treatment improves limb muscle strength in \textit{mdx} mice**

Grip strength measurements, performed to measure muscle strength, demonstrated that IL-1Ra-treated \textit{mdx} mice showed a significant 11% increase in maximum forelimb strength measurements and a 7% increase in maximum hindlimb strength where statistical significance was not attained likely due to one outlier (Fig 2A and 2B). Mice treated with IL-1Ra exhibited significantly improved normalized forelimb grip strength (Fig 2C) and improvements in normalized hindlimb strength when compared to control animals (Fig 2D). The normalized forelimb and hindlimb grip strengths increased by approximately 11% (p<0.05) and 7% (p = 0.056), respectively, in the mice treated with IL-1Ra when compared to the saline-injected controls (Fig 2C and 2D). Latency to fall measurements are an indicator of motor coordination, learning, and balance; however, there were no significant differences in these measurements between IL-1Ra-treated and control \textit{mdx} mice (Fig 2E).

**IL-1Ra treatment does not alter \textit{in vitro} force generation or histological parameters in \textit{mdx} mice**

To further examine muscle strength, \textit{in vitro} force measurements were performed. The \textit{in vitro} twitch force (mN) and absolute force (mN) of the EDL muscle did not differ between IL-1Ra-
treated and saline-treated mdx mice (Fig 3A–3C). The specific force (kN/m²) and EDL mass (mg) was not significantly changed but there was a 12% reduction in specific force comparing the mean values of the IL-1Ra-treated animals which could partially be explained by the slight (not statistically significant) differences in mass of the EDL from the IL-1Ra-treated mice versus saline (9.67 mg versus 9.36 mg, Table 1). There were no significant differences in the absolute body mass, skeletal muscle (gastrocnemius, soleus) mass, or heart mass between the saline-treated and IL-1Ra-treated mice (Table 1). From histology, no differences were observed in the number of degenerating or regenerating fibers, inflammatory cells, or centralized or peripheral nuclei in the gastrocnemius muscles from the IL-1Ra-treated and control mdx mice (Fig 4A and 4B).

**IL-1Ra treatment decreases activity in mdx mice**

Behavioral measurements of horizontal and vertical activity were performed using an open-field Digiscan apparatus as previously described [33]. IL-1Ra treatment did not have a significant effect on either horizontal or vertical activity in IL-1Ra-treated mdx mice when compared to saline-treated mice (Fig 2). Maximal strength (KGF) was measured using a grip-strength meter and normalized to body mass in kg to determine the force per kg of animal mass (KGF/KG). Animals treated with IL-1Ra (n = 10) had significantly higher (A) maximal (p = 0.047, 11% increase) and (C) normalized forelimb grip-strength (p = 0.028, 11% increase) than did control saline-treated (n = 6) animals. Hindlimb strength was measured in the same way as forelimb, and there was an increase in (B) maximal hindlimb grip strength (p = 0.100, 7% increase) and (D) normalized hindlimb strength when the means of the IL-1Ra treated (10.56 KGF/KG) and saline-injected mice (9.87 KGF/KG) were compared; this difference almost reached statistical significance (p = 0.058). There were also no significant differences in hindlimb grip strength without normalization, although a similar trend remained. (E) There were no significant differences in the latency to fall in control- (95.44s) and IL-1Ra- (99.15s) treated mdx mice, although the mean values showed a longer latency to fall for the IL-1Ra-treated mice. Values in the graphs represent mean ± SEM. Statistically significant differences were determined by parametric, unpaired, two-tailed t-tests with a p < 0.05 being significant.

**Fig 2. Grip strength is increased in mdx mice treated with IL-1Ra.** Maximal strength (KGF) was measured using a grip-strength meter and normalized to body mass in kg to determine the force per kg of animal mass (KGF/KG). Animals treated with IL-1Ra (n = 10) had significantly higher (A) maximal (p = 0.047, 11% increase) and (C) normalized forelimb grip-strength (p = 0.028, 11% increase) than did control saline-treated (n = 6) animals. Hindlimb strength was measured in the same way as forelimb, and there was an increase in (B) maximal hindlimb grip strength (p = 0.100, 7% increase) and (D) normalized hindlimb strength when the means of the IL-1Ra treated (10.56 KGF/KG) and saline-injected mice (9.87 KGF/KG) were compared; this difference almost reached statistical significance (p = 0.058). There were also no significant differences in hindlimb grip strength without normalization, although a similar trend remained. (E) There were no significant differences in the latency to fall in control- (95.44s) and IL-1Ra- (99.15s) treated mdx mice, although the mean values showed a longer latency to fall for the IL-1Ra-treated mice. Values in the graphs represent mean ± SEM. Statistically significant differences were determined by parametric, unpaired, two-tailed t-tests with a p < 0.05 being significant.

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**Fig 3. IL-1Ra treatment does not alter EDL muscle force.** Muscle force in the extensor digitorum longus (EDL) muscle did not differ in saline-treated and IL-1Ra-treated mdx mice. There was a decrease in the strength capacity of the mice treated with IL-1Ra when compared to the controls for all measurements recorded: (A) twitch force, mN (5% decrease), (B) muscle force, mN (4% decrease) and (C) specific force, kN/m² (12% decrease); however, none of these differences was statistically significant. Values in the graphs represent mean ± SEM. Statistical significance was determined by parametric, unpaired, two-tailed t-tests (n = 4 saline and n = 6 IL-1Ra).

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to saline-injected mice (Fig 5; data not shown). Movement time and total distance traveled were significantly decreased (by 27%) in the IL-1Ra-treated mice when compared to the saline-treated mice (Fig 5B and 5C).

### Discussion

During inflammatory processes, IL-1β is produced by several cell types in response to activation of a variety of innate immune receptors, including the toll-like receptor (TLR) superfamily. Stimulation of IL-1β and subsequent binding to the IL-1R activates the pro-inflammatory NFκB pathway. IL-1β signaling is controlled by IL-1Ra, a natural antagonist of IL-1R, and overexpression of IL-1β has been implicated in the pathology of a wide variety of human

| Parameters     | Saline-treated | IL-1Ra-treated | p-value |
|----------------|----------------|----------------|---------|
| Body mass, g   | 20.93 (0.42)   | 21.85 (0.31)   | 0.12    |
| Gastrocnemius, mg | 109.28 (5.06) | 114.67 (3.00) | 0.36    |
| Soleus, mg     | 7.53 (0.42)    | 7.81 (0.29)    | 0.60    |
| EDL, mg        | 9.36 (0.59)    | 9.67 (0.35)    | 0.66    |
| Heart, mg      | 100.55 (5.73)  | 100.29 (2.68)  | 0.96    |

There were no statistically significant differences in the overall body mass or the masses of the gastrocnemius, soleus, extensor digitorum longus (EDL), or heart between the IL-1Ra-treated and control saline-treated mice. Muscle mass was expressed as the average of the left and right muscle, where applicable, and also as a percentage of the total body weight. Values are expressed as mean, with SEM in parentheses. No significant differences were determined as calculated by parametric, unpaired, two-tailed, t-tests; p-values are indicated.

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Fig 4. IL-1Ra treatment does not alleviate muscle pathology in *mdx* mice. Degenerating (A) and regenerating (B) muscle fibers were quantified in gastrocnemius muscle. Inflammatory cells were also quantified in sections from saline- and IL-1Ra-treated mice (C). Nuclei from sections were scored and quantified as either centralized (D) or peripheral (E). The number of fibers with centralized nuclei (F) was quantified by treatment group Values in the graphs represent mean ± SEM. Statistical significance was determined by parametric, unpaired, two-tailed, t-tests (N = 5, saline; N = 9, IL-1Ra).

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diseases associated with chronic inflammation. Development of inhibitors such as anakinra have been shown to be effective in many diseases with an inflammatory component including: inhibiting apoptotic events during experimental acute myocardial infarction [34], improving symptoms of type 2 diabetes [35–38] and reducing the symptoms of rheumatoid arthritis (RA) in clinical trials [39,40]. Improvements in auto-inflammatory diseases, including Muckle-Wells syndrome (MWS) [41] and gout [42] have also been described after using anakinra. Since inflammation is a major player in DMD, the IL-1β pathway is expected to play a role in the initiation and perpetuation of the muscle pathology in DMD. Muscle inflammation and necrosis is also evident in the mdx mouse model; therefore, IL-1Ra treatment in pre-clinical studies could prevent the inflammatory effects of systemic IL-1β secretion on IL-1R activation potentially limiting cellular inflammation. In addition, it is known that in the mdx mouse model of DMD, there is an increase in the expression of TNF-α and IL-1β prior to disease onset [23]. IL-1Ra is already FDA-approved making this drug a good candidate to try to inhibit IL-1β upregulation and prevent the associated downstream signaling.

To date, many preclinical studies have focused on pharmacological treatments to inhibit TNF-α as a potential treatment for DMD. Several of the drugs tested have been shown to ameliorate the muscle pathologies associated with the mdx phenotype [14–22]. In the present study, inhibiting IL-1β in the mdx model was tested using a synthetic inhibitor of IL-1R, IL-1Ra, treating mdx mice for 45 days. The dosing regimen selected here, 25mg/kg daily, was based on a previous study that demonstrated efficacy in a model of alcoholic steatohepatitis in mice [43]. To assess whether IL-1Ra had engaged the target, we performed an in vitro stimulation assay using LPS to activate splenocytes collected from treated and untreated animals. Here, we saw a blunted effect of IL-1Ra on splenocytes treated with LPS (27% less than control LPS-stimulated splenocytes), indicating IL-1Ra incompletely inhibited IL-1 under our conditions, thus allowing signaling to persist even after treatment (Fig 1).

In addition to examining signaling effects of IL-1Ra, strength testing was also performed and revealed significantly improved forelimb grip strength and a trend for hindlimb grip strength, though not significant, toward increasing (Fig 2). No changes were detected in latency to fall (Fig 2E), or muscle force (twitch, maximal, or specific force, Fig 3). Open-field activity measures were mainly down-regulated in the mice treated with IL-1Ra (Fig 5). One possible explanation for the 27% decrease in movement in the IL-1Ra-treated mice could be that the mice are nauseous and therefore less likely to move about the cage. This is certainly reasonable to consider since this is one of the potential side effects of IL-1Ra in humans (Fig 5). Increased
grip strength and decreased open-field activity measures could have been caused by differential effects of IL-1 signaling. For example, it is known that immobilization stress induces IL-1β in rats [44,45], IL-1 administration induces stress [46], exposure to acute stress induces IL-1 expression in rodents [47]. IL-1R-null mice show decreased anxiety, and conversely IL-1Ra-null mice show increased anxiety as they age [48,49]. Additionally, mice with IL-1β overexpression specifically in the hippocampus (IL-1βXAT) demonstrate increased locomotion, the converse of what was seen in these studies inhibiting that pathway [50]. IL-1 could be released to mediate a stress response, and when the natural inhibitor of IL-1, IL-1Ra, is genetically removed, the animals elicit an increased stress response [49]. For this study, the mice were handled prior to the grip strength measurement, and although they had become acclimated during the week before the measurements were collected, grip strength could have been improved because of increases in IL-1β leading to behavioral stress response. True muscle force is measured by the in vitro force measurements, because these measurements are not confounded by volition. In vitro force was not changed in our study after IL-1Ra treatment, indicating that IL-1Ra did not improve muscle strength possibly due to an incomplete inhibition of this pathway.

The skeletal muscles from the mice treated with IL-1Ra had no differences in histological measurements including inflammatory foci, muscle fibers with centralized nuclei, regenerating fibers, or degenerating fibers when compared to controls. This can also be explained by IL-1Ra being a weak inhibitor of the pathway (Fig 4). There are several other factors that could provide an explanation for this including: lack of potency, dosage, duration of treatment, and the role of the IL-1 pathway in muscle disease progression at the time of treatment. A longer-term study could reveal improved cellular patterning in IL-1Ra-treated mdx mice and indicate that IL-1Ra can indeed diminish the secretion of IL-1; however, this possibility still needs to be tested. For this study, treatment was administered after the onset of skeletal muscle necrosis in mdx mice and others have shown that inhibiting another inflammatory cytokine, TNFα, using Remicade® (an anti-TNFα antibody) at seven days of age delayed onset of acute necrosis at 21 days in mdx mice [22]. Treating mice at earlier time points with IL-1Ra could be considered for future experiments to examine the full potential of this drug at all stages of the disease phenotype in the mdx mice. Interestingly, at 12 weeks there were no significant pathological changes found after Remicade® treatment as we have shown after IL-1Ra treatment [22]. To overcome this challenge with the stabilized mdx muscle phenotype, older mdx mice could be treated with IL-1Ra and challenged using a treadmill to exacerbate the disease phenotype at this stage and examine additional effects from this treatment on dystrophic muscles similarly to what was done with TNFα inhibition [20]. Lastly, it is possible that inhibition of TNFα could be more efficacious than IL-1Ra treatment in the mdx mice; however, considerations have to be taken for examining cardiac function as long-term TNFα inhibition has been demonstrated to have a negative impact on heart function [51].

Overall, the dose of IL-1Ra used here may have been enough to elicit a behavioral response but insufficient to completely block the inflammatory signaling cascade and associated phenotype in the skeletal muscles of the mdx mice. In order to further examine the inhibition of IL-1 and the benefits of down-regulating the secretion of this inflammatory factor, other IL-1R inhibitors could also be examined in preclinical trials. They could be compared with IL-1Ra to determine whether or not they are more potent inhibitors and, if so, what histological and functional outcomes might improve from their use. Others have examined the possibility of decreasing inflammation by using exon-skipping technology targeting IL-1RAcP [52]. Targeting IL-1R can have broad applications for many inflammatory diseases, and the mechanisms of its inhibition are important to understand in order to decrease inflammation associated with the disease. The goal of the ongoing studies in our laboratory is to broaden the therapeutic options for DMD patients and their families beyond prolonged steroid regimens. In this short-
term study of mdx mice, which have a milder phenotype than DMD patients. IL-1Ra treatment displayed an improvement in some functional parameters for muscle strength when compared to saline-treated mice; however, these changes could have been a result of the behavioral effects of the treatment. Optimization of the dosage and timing of the IL-1Ra treatment is clearly needed to clarify whether a greater inhibition of the IL-1β signaling pathway (and resulting clinical improvement) can be achieved in DMD.

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Author Contributions

Conceived and designed the experiments: AS SR JV KN. Performed the experiments: AS SR JV. Analyzed the data: AS SR MBK KN. Wrote the paper: AS SR MBK KN.

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