The genetic causes of infertility in patients with oligozoospermia and azoospermia in Turkish population

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The study was approved by the Ethics Committee of Bakırkoy Dr.Sadi Konuk Training and Research Hospital (Approval number: 2018-01-05) (Date: 2018 Jan 8). All research was performed in accordance with relevant guidelines/regulations, and informed consent was obtained from all participants.
INTRODUCTION

Infertility is considered if the fertilization does not occur in spite of unprotected regular sexual intercourse for one year (1). Infertility is a health problem, which affects 15% of the couples, and almost one-half of infertility caused male-related factors or in combination with female related fertility disorders (1, 2). Azoospermia is the absence of the sperms in the semen analysis and oligozoospermia (<5x10^6 spermatozoa/ml) is defined as the low sperm count in the ejaculate. Infertility in male patients can be occurred by a variety of factors, such as varicosel, antisperm antibodies, infection, spermatid duct obstruction, cryptorchidism, endocrine disorders, testicular trauma, esticular cancer, systemic diseases, retrograde ejaculation, etc (2). Except those cases, in 30–40% of male infertile patients that are presented to as idiopathic, genetic abnormality can be considered (3). The most common genetic cause of male infertility is the chromosomal anomalies and is encountered in 5% of all cases. This rate increases up to 15% in azoospermic males (4). The second most common genetic cause of male infertility is Y chromosome microdeletions, which can’t be detected with cytogenetic methods (5). In this study, we evaluated the genetic causes in patients with nonobstructive azoospermia and severe oligozoospermia in infertile males.

MATERIAL AND METHODS

In this cross-sectional study, after obtaining the ethical approval (2018/01), a total of 350 patients, who admitted to the division of the Andrology from Urology department of University of Health Sciences Bakirkoy Dr. Sadi Konuk Training and Research Hospital due to infertility between January 2015 and January 2017. All participants signed an informed consent form for the use of their blood sample in the study and all records were required to be kept confidential. Patients, who had a semen volume less than 2 cc in the spermiogram analysis, underwent transrectal ultrasound examination for obstructive azoospermia and those with obstructive azoospermia were excluded from the study. The patients had non-obstructive azoospermia (NOA) and severe oligozoospermia in their semen analyses, were analyzed retrospectively. Patients were divided into groups according to sperm count following detailed physical examination and hormonal tests which was performed by a specialist. Semen analyses (motility, sperm concentration and morphology) were performed according to the World Health Organization Guidelines (6). Following the examinations, patients, who had idiopathic azoospermia and severe oligozoospermia (≤ 5 million sperm/ml) with an age between 20 and 45 years were determined. Light microscopic evaluation of sperm motility, concentration, morphology and viability and was performed (6). Specimens originally considered as azoospermic were centrifuged (1000 g for 20 min) and the pellets were examined for spermatozoa, before confirming azoospermia (7). The diagnosis of azospermia and oligozoospermia was confirmed by two spermiograms examined at least 15 days apart following 3 days of sexual abstinence. Blood samples of the patients were taken into tubes containing ethylenediaminetetraacetic acid (EDTA). DNA was isolated from the peripheral blood samples with the DNA isolation device (MagNA Pure Compact System). Karyotype analysis was performed on lymphocyte cultures stimulated by phytohaemagglutinin adapting methodology (8).

For the Y chromosome microdeletion analysis, the amplification of the relevant regions was carried out with the multiplex PCR method using the SRY and ZFY regions and STS sequence-tagged site (STS) primers. Following the multiplex PCR process, the microdeletions in AZFa, AZFb and AZFc regions and SRY and ZFY regions were investigated with the agarose gel electrophoresis.

Statistical Analysis

Data analysis was performed using the SPSS version 16.0 software package. The continuous variables are presented as mean±standard deviation. The frequencies of categorical variables were compared using Pearson Chi-Square test. Statistical significance was considered when p value was <0.05.

RESULTS

A total of 350 infertile men were evaluated, 225 (64.2%) of whom were azoospermic males and 125
(35.7%) oligozoospermic males. The mean age of the NOA and oligozoospermic patients were calculated as 33.4±4.3 years and 34.6±4.8 years, respectively. The overall incidence of cytogenetic abnormalities was 16%. Chromosomal abnormalities were detected in 32 (9.1%) of 350 cases. Numerical and structural chromosomal abnormalities are summarized in Table 1. Four patients in the oligozoospermia group had Kleinfelter syndrome (47,XXY; KS) syndrome (3.2%). In NOA group, twenty-four (11.5%) patients had KS; two (0.88%) patients had 46,XY/47,XXY; one (0.44%) patient had 46,XY/45XO and one (0.44%) patient had 46,XX. In oligozoospermia group, 121 had normal karyotype (96.8%), 4 patients had abnormal karyotype (3.2%). Among NOA group, 197 had normal karyotype (96.8%), 4 patients had abnormal karyotype (3.2%). While the mean age of KS patients in the oligozoospermia group was 28±4 years, in NOA group it was 27±6 years. As semen parameters deteriorated, the chromosomal abnormality frequency increased and statistically significant difference was found (p=0.004; Table 1). In all patients groups, the incidence of KS (47,XXY) was 8% (n=28) and it was the most frequent abnormality. In oligozoospermia and NOA groups, KS frequency were 3.2% and 11.5% respectively (n=4 vs n=24, respectively). The overall incidence of Y chromosome microdeletion was 6.8%. Three (%2.4) patients in oligozoospermia group and twenty-one (9.3%) patients in NOA group, Y chromosome microdeletions were detected. It was statistically significant difference between oligozoospermia and NOA groups (p=0.007). The most frequent region with deletions was the AZFc region. Based on molecular screening of the AZF region, a total of 24 AZF microdeletions were found: 1 in AZFa+b+c, 2 in AZFa, 2 in AZFb, 3 in AZFb+c, and 16 in AZFc (Table 2).

Table 1. Distribution of normal and abnormal karyotypes in the patients included in the study

| Karyotype | NOA (n, %) | Oligozoospermia (n, %) | Total |
|-----------|------------|------------------------|-------|
| Normal (46,XY) | 197 (61.9 %) | 121 (38.1 %) | 318 |
| Abnormal | | | |
| KS (47,XXY) | 24 (85.7 %) | 4 (14.3 %) | 28 |
| Mozaic KS | 2 (100 %) | 2 |
| Other sex chromosome mosaicism 45,X0 (n=1) and 46,XX (n=1) | 2(100 %) | 2 |

KS: Kleinfelter syndrome; NOA: Non-obstructive azoospermia;

Table 2. Y chromosome microdeletions in infertile cohort

| Y chromosome | NOA (n;%) | Oligozoospermia (n;%) | Total |
|--------------|-----------|------------------------|-------|
| Normal | 204 (90,6 %) | 122 (97,6 %) | 326 |
| Deleted | 21 (9,3 %) | 3 (2,4 %) | 24 |
| AZFa | 2 (0,8 %) | 0 | 2 |
| AZFb | 2 (0,8 %) | 0 | 2 |
| AZFc | 13 (5,7 %) | 3 (2,4 %) | 16 |
| AZFbc | 3 (1,3 %) | 0 | 3 |
| AZFabc | 1 (0,4 %) | 0 | 1 |

NOA: Non-obstructive azoospermia;
DISCUSSION

Among numerous etiologic factors, genetic disorders play a primary key role in male infertility with abnormal semen parameters. Spermatogenesis is regulated by a number of genes on the Y chromosome and by autosomes that act at different stages of germ cell development (9). In the present study, the frequency of chromosomal abnormality was founded as 9.1%. These findings are very similar to those reported in earlier studies (10). In the literature there are many studies about chromosomal abnormality rate from different countries which were reported as 6.2-12.6% (11, 12). Consistent with other clinical studies, KS was found to be the most common anomaly in this study (11, 13). In the current study, 26 of the 30 KS patients were azospermic (24 patients with 47, XXY and 2 patients with mosaicism). While the mean age of KS patients in the oligozoospermia group was 18±4 years and it was 27±6 years in NOA group. This finding is consistent with germ cell degeneration and early-onset progressive testicular insufficiency is KS patients (1, 2, 14, 15).

Excessive X chromosome is paternal in 50-60% of cases and maternal in 40-50% of cases (16). Epidemiological studies have shown that the incidence of KS is gradually rising and it is thought to be related with increasing father age (17). However, only 25% of all cases with KS are diagnosed and followed up clinically. However, in recent years, patients with KS have significant health problems not only with infertility but also with high morbidity (70%) and mortality (50%) (17, 18). It is important that the patients are diagnosed early in puberty and the management of the disease is considered as soon as possible. In KS patients, secondary sex characteristics become apparent with testosterone replacement therapy. Depressive mood of the patient disappears and self-confidence increases. In general, patients with KS are accepted as infertile, however assisted reproductive techniques may provide fertilization. These patients may also have children by means of sperm cryopreservation, testicular sperm extraction (TESE), micro-TESE and intracytoplasmic sperm injection (ICSI). Freezing of spermatogonial stem cells or storage of testicular tissue are yet experimental treatments (19).

Y chromosome microdeletions are the second most common cause of male infertility (20). Ethnic and regional factors have been considered as the main parameters affecting diversity and prevalence of these microdeletions (21). An international analysis of a large dataset revealed that the prevalence of Yq microdeletions was between 7% and 10% (21). The incidence of the Y chromosome microdeletion in the Turkish population was investigated by several authors (1.3-9.6%) (4, 22-24). In our study, the frequency of Y chromosome microdeletion was calculated as 6.8%. These frequency variations may be due to the genetic variation in different populations especially in Y chromosome specific haplotypes, genetic background, environmental factors, and different types of primers for AZF-related microdeletions. Furthermore, the population size, patient selection based on the etiology and severity of spermatogenesis defects, and the regional differences can be considered as the possible reasons for these frequency variations. Mirfakhraie et al. reported a higher AZFb (66.67%) microdeletion frequency compared to the AZFc (41.67%) region in general Iranian population (25). It was stated that although AZFc deletion affects the spermatogenesis, it does not always cause infertility. Sperm may be obtained with TESE and fertilization may occur in azoospermic males with AZFc deletion. Germ cells are totally lost in cases with complete deletions of AZFa and AZFb regions. TESE and ICSI will fail in these cases. It was reported that the probability of sperm detection with TESE in partial AZFb deletion was 50% (26). In cases with Yq microdeletion, who underwent assisted reproductive techniques, couples should be informed that the microdeletion will be transferred to the male children and consequently these children may also have spermatogenesis disorders.

The retrospective character of the study and the lack of knowledge of treatment methods applied to the patients and paternity rates during follow-up are the limiting factors.

CONCLUSION

Genetic analysis and consultation should be considered before the implementation of the assisted reproductive techniques, when chromosomal anomalies and Y chromosome microdeletions were detected in infertile males.
Conflict of Interest
The authors have no conflicts of interest to declare.

Financial Disclosure
The authors declared that this study has received no financial support.

Ethical Approval
The study was approved by the Ethics Committee of Bakırkoy Dr.Sadi Konuk Training and Research Hospital (Approval number: 2018-01-05) (Date: 2018, Jan 8). The study protocol conformed to the ethical guidelines of the Helsinki Declaration.

REFERENCES
1. Mundy AR, Andrich DE. Urethral strictures. BJU Int. 2011; 107(1):6-26.
2. Santucci RA, Joyce GF, Wise M. Male urethral stricture disease. J Urol. 2007; 177(5):1667-1674.
3. Anger JT, Buckley JC, Santucci RA, Elliott SP, Saigal CS, Urologic Diseases in America P. Trends in stricture management among male Medicare beneficiaries: underuse of urethroplasty? Urology. 2011; 77(2):481-485.
4. Rassweiler J, Teber D, Kuntz R, Hofmann R. Complications of transurethral resection of the prostate (TURP)–incidence, management, and prevention. Eur Urol. 2006; 50(5):969-979.
5. Lumen N, Hoebeke P, Willemsen P, et al. Etiology of urethral stricture disease in the 21st century. J Urol. 2009; 182(3):983-987.
6. Jarosek SL, Virnig BA, Chu H, Elliott SP. Propensity-weighted long-term risk of urinary adverse events after prostate cancer surgery, radiation, or both. Eur Urol. 2015; 67(2):273-280.
7. Heyns CF, Steenkamp JW, De Kock ML, Whitaker P. Treatment of male urethral strictures: is repeated dilation or internal urethrotomy useful? J Urol. 1998; 160(2):356-358.
8. Launonen E, Sairanen J, Ruutu M, Taskinen S. Role of visual internal urethrotomy in pediatric urethral strictures. J Pediatr Urol. 2014; 10(3):545-549.
9. Blaschko SD, Harris CR, Zaid UB, et al. Trends, utilization, and immediate perioperative complications of urethroplasty in the United States: data from the national inpatient sample 2000-2010. Urology. 2015; 85(5):1190-1194.
10. Kinnaird AS, Levine MA, Ambati D, Zorn JD, Rourke KF. Stricture length and etiology as preoperative independent predictors of recurrence after urethroplasty: A multivariate analysis of 604 urethroplasties. Can Urol Assoc J. 2014; 8(5-6):296-300.
11. Wessells H, Angermeier KW, Elliott S, et al. Male Urethral Stricture: American Urological Association Guideline. J Urol. 2017; 197(1):182-190.
12. Andrich DE, Mundy AR. Non-transecting anastomotic bulbar urethroplasty: a preliminary report. BJU Int. 2012; 109(7):1090-1094.
13. Levy M, Gor RA, Vanni AJ, et al. The Impact of Age on Urethroplasty Success. Urology. 2017; 107:232-238.
14. Santucci RA, McAninch JW, Mario LA, et al. Urethroplasty in patients older than 65 years: indications, results, outcomes and suggested treatment modifications. J Urol. 2004; 172(1):201-203.
15. Barbagli G, Morgia G, Lazzeri M. Dorsal onlay skin graft bulbar urethroplasty: long-term follow-up. Eur Urol. 2008; 53(3):628-633.
16. Lacy JM, Cavallini M, Bylund JR, Strup SE, Preston DM. Trends in the management of male urethral stricture disease in the veteran population. Urology. 2014; 84(6):1506-1509.
17. Arias E, Heron M, Xu J. United States Life Tables, 2013. Natl Vital Stat Rep. 2017; 66(3):1-64.
18. Santucci RA, Mario LA, McAninch JW. Anastomotic urethroplasty for bulbar urethral stricture: analysis of 168 patients. J Urol. 2002; 167(4):1715-1719.
19. Elliott SP, Metro MJ, McAninch JW. Long-term followup of the ventrally placed buccal mucosa onlay graft in bulbar urethral reconstruction. J Urol. 2003; 169(5):1754-7.
20. Barbagli G, Palminteri E, Rizzo M. Dorsal onlay graft urethroplasty using penile skin or buccal mucosa in adult bulbar urethral strictures. J Urol. 1998; 160(4):1307-1309.
21. Blair SL, Schwarz RE. Advanced age does not contribute to increased risks or poor outcome after major abdominal operations. Am Surg. 2001; 67(12):1123-1127.
22. Breyer BN, McAninch JW, Whitson JM, et al. Multivariate analysis of risk factors for long-term urethroplasty outcome. J Urol. 2010; 183(2):613-617.
23. Viers BR, Pagliara TJ, Rew CA, et al. Urethral Reconstruction in Aging Male Patients. Urology. 2018; 113:209-214.
24. Chapman D, Kinnaird A, Rourke K. Independent Predictors of Stricture Recurrence Following Urethroplasty for Isolated Bulbar Urethral strictures. J Urol. 2017; 198(5):1107-1112.

25. Viers BR, Pagliara TJ, Shakir NA, et al. Delayed Reconstruction of Bulbar Urethral strictures is Associated with Multiple Interventions, Longer Strictures and More Complex Repairs. J Urol. 2018; 199(2):515-521.