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Published in:
Scientific Reports

DOI:
10.1038/srep29638

Publication date:
2016

Document version
Publisher's PDF, also known as Version of record

Citation for published version (APA):
Jensen, K., Budvytyte, R., Thomas, R. A., Wang, T., Fuchs, A. M., Balabas, M. V., Vasilakis, G., Mosgaard, L. D., Stærkind, H. C., Müller, J. H., Heimburg, T., Olesen, S-P., & Polzik, E. S. (2016). Non-invasive detection of animal nerve impulses with an atomic magnetometer operating near quantum limited sensitivity. Scientific Reports, 6, [29638]. https://doi.org/10.1038/srep29638

Download date: 14. sep., 2024
Non-invasive detection of animal nerve impulses with an atomic magnetometer operating near quantum limited sensitivity

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Magnetic fields generated by human and animal organs, such as the heart, brain and nervous system carry information useful for biological and medical purposes. These magnetic fields are most commonly detected using cryogenically-cooled superconducting magnetometers. Here we present the first detection of action potentials from an animal nerve using an optical atomic magnetometer. Using an optimal design we are able to achieve the sensitivity dominated by the quantum shot noise of light and quantum projection noise of atomic spins. Such sensitivity allows us to measure the nerve impulse with a miniature room-temperature sensor which is a critical advantage for biomedical applications. Positioning the sensor at a distance of a few millimeters from the nerve, corresponding to the distance between the skin and nerves in biological studies, we detect the magnetic field generated by an action potential of a frog sciatic nerve. From the magnetic field measurements we determine the activity of the nerve and the temporal shape of the nerve impulse. This work opens new ways towards implementing optical magnetometers as practical devices for medical diagnostics.

The magnetic field generated around a signaling nerve fiber is of key interest both from a basic scientific and a clinical point of view. The transmembrane potentials have been extensively measured with electrophysiological techniques. Magnetic field measurements are insensitive to the transmembrane currents as the fields from the opposite currents in and out of the membrane cancel. Instead, magnetic field measurements allow for a true measurement of the axon's axial net current, which is the depolarizing wavefront driving the action potential. Magnetic field recordings also allow for non-invasive measurements of the conduction velocity of peripheral nerves¹ which is necessary for diagnostics of multiple sclerosis, myotonia and intoxication in patients.

The magnetic field of a nerve impulse was first measured by Wikswo et al.² using a combination of a superconducting SQUID magnetometer and a toroidal pick-up coil through which the nerve had to be pulled. This method is not compatible with in vivo diagnostics and yields the magnetic field values which are much higher than that in an animal because the return currents in the surrounding tissue are not measured. Here we are able to detect the nerve impulse with the sensor placed beside the nerve, several millimeters away, the setting compatible with in vivo studies. Detection of nerve impulses with a magnetometer based on Nitrogen-Vacancy centers in diamond has recently been reported³. Such magnetometers seem promising for magnetic field microscopy applications where the magnetometer can be placed at micrometer distance or closer to the biological object.

Sensitivity of atomic magnetometers⁴ improves with the number of atoms sensing the field, which for vapor magnetometers is defined by volume and temperature. For example, femtoTesla sensitivity has been achieved with magnetometers operating at a temperature of several hundred of °C in the so-called SERF regime⁵ used also for medical applications⁶–⁸. Similarly high sensitivity has been achieved at room temperature using much fewer atoms by means of quantum state engineering⁹ leading to operation beyond standard quantum limits of

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if the magnetic pulse duration \( \tau \) which is longer than a typical nerve impulse duration in the human body. The close proximity of the sensor to the source of magnetic field is a big advantage as the magnetic field rapidly decreases with the distance from the source. Room-temperature cesium magnetometer has been used for medical applications, however, it operated far above quantum limits of sensitivity.

**Results**

Here we use the approach of ref. 9 for nerve impulse measurements. The sensitive element of the magnetometer is cesium atomic vapour. Cesium has a high vapor pressure such that high sensitivity can be reached at room- or human body temperature. The magnetic moment (spin) of atoms \( \mathbf{J} = (J_x, J_y, J_z) \) is prepared by optical pumping in the \( x \)-direction, along the direction of a bias field \( B_x \) [see Fig. 1(a)]. The magnetic field of the nerve \( B_{\text{nerve}} \) will create a transverse spin component \( J_y = (J_x, J_z) \) which afterwards will rotate in the \( y-z \) plane at the Larmor frequency \( \Omega = B_x / \gamma \) [see Fig. 1(b)], where \( \gamma = 2.2 \times 10^6 \text{rad} / (\text{s} \cdot \text{T}) \) is the cesium gyromagnetic ratio. The \( J_y \) spin component is detected optically by measuring the polarization rotation of the probe light. The magnetic field from the nerve is detected in two modalities, a continuous mode where the magnetic field as a function of time \( B(t) \) is detected, and a pulsed mode where the Fourier component \( |B(\Omega)| \) is detected. In the continuous mode the pump and probe light is continuously on. In the pulsed mode [see Fig. 1(c)], a pulse of pump light is followed by the pulse of magnetic field, and finally the spins are detected with a pulse of probe light.

Optical magnetometers are fundamentally limited by quantum noise consisting of the spin-projection noise (PN) shown as the fuzzy circle in Fig. 1(b), quantum shot noise of the probe light and quantum back action of the probe on the spin\(^7\). For an optimal measurement these noise sources add up to the standard quantum limit (SQL)\(^1\). This limit has been reached for magnetic fields oscillating at hundreds of kHz\(^9\). Here we approach the SQL for the magnetic field measurement of nerve impulses whose frequency is much lower using the techniques described in the Supplementary Information. For continuous measurements, the SQL magnetic field uncertainty \( \Delta B_{\text{SQL}} \) normalized by the total measurement time \( T_{\text{tot}} \) yields the sensitivity \( \Delta B_{\text{SQL}} / \sqrt{T_{\text{tot}}} \approx 1.5 / (\gamma / T J_z / 2) \) in units of \( T / \sqrt{\text{Hz}} \). \( T_s \) is the spin coherence time and \( J_z = 4N_A \) is the total atomic spin for \( N_A \) cesium atoms. At room temperature of 22 °C, the cesium atomic density is 3.1 × 10\(^4\) m\(^{-3}\), the highest of all elements appropriate for atomic magnetometry. The pulsed measurement has the SQL magnetic field uncertainty of \( \Delta |B_{\text{SQL}}(\Omega)| \approx 1.5 / (\gamma / 2T_s) \) if the magnetic pulse duration \( \tau \approx T_s \) (see Supplementary Information).

A long spin-coherence time \( T_s \) is crucial for a high sensitivity. In this work we utilize a vapor cell with the inside surface coated with alkane\(^{12,13}\). The coating protects atomic spin states from decoherence over many thousands of wall collisions and provides \( T_s \approx 2 \text{ ms} \) which is longer than a typical nerve impulse duration \( \tau \approx 2 \text{ ms} \), as required for the ultimate sensitivity. The cesium vapor is contained in a 1.0 mm × 1.0 mm × 7.7 mm channel inside a rectangular glass chip of dimensions 2.3 mm × 8.4 mm × 7.7 mm. This small rectangular vapor cell allows us to have atoms at an average distance of a few mm from the nerve, which is close to a typical distance for many medical applications.

A frog sciatic nerve contains a few nerve bundles each with several thousand axons inside (see Methods section). The nerve is placed inside a plastic chamber where it can be kept alive in a saline solution for more than 5 hours. The nerve is electrically stimulated from one end with a pair of gold electrodes [see Fig. 1(b)]. The stimulus triggers an action potential (a nerve impulse) propagating along the nerve. As a reference measurement we perform an electrical recording of the impulse with another pair of electrodes. Figure 2(a) shows the electrically recorded signals for different stimulation voltages. Figure 2(b) shows the frequency spectra of the nerve signals.
The circumferal magnetic field $B_x$ shows the spectrum and Fig. 2(f) shows the magnetic field Fourier component at the Larmor frequency of 400 Hz. Therefore create a transverse spin component [see Fig. 1(b)]. Figure 2(d) shows the magnetometer signal, Fig. 2(e) the magnetic field from the nerve as $|B_{\text{nerve}}(\Omega)|$ and Fig. 2(c) shows the amplitude of the 400 Hz Fourier component. The nerve is stimulated at $t = 6.0$ ms. The signature of the nerve signal is its non-linear behavior with stimulation voltage, with a firing threshold at around 0.4 V. For voltages above the threshold a nerve impulse is measured with the recording electrodes within the time interval $t = 7.0 \pm 10.0$ ms. We also observe a stimulation artifact at $t = 6.0$ ms [see inset in Fig. 2(a)] which is proportional to the stimulation voltage.

In parallel to the reference electrical recording the nerve signal is detected optically using the pulsed magnetometer mode. The magnetometer is positioned near the middle part of the nerve separated only by a thin microscope cover slip. As the nerve is bent in a U-shape, we mainly detect the field from the 10 mm section of the nerve closest to the magnetometer [see Fig. 1(b)]. The axial ionic current in this 10 mm section can be considered constant, as the action potential has a duration $\approx 3$ ms, the velocity $\approx 30$ m/s, and therefore an extent $\approx 9$ cm $> 10$ mm. The circumferal magnetic field $B_{\text{nerve}}$ from the nerve is on average transverse to the initial spin direction $J_z$ and will therefore create a transverse spin component [see Fig. 1(b)]. Figure 2(d) shows the magnetometer signal, Fig. 2(e) shows the spectrum and Fig. 2(f) shows the magnetic field Fourier component at the Larmor frequency of 400 Hz. A clear threshold for the nerve firing is observed confirming that the magnetometer is capable of detecting the nerve impulse. From calibration measurements (see Methods section) we determine the Fourier component of the magnetic field from the nerve as $|B_{\text{nerve}}(\Omega)| = 9.1$ pT·ms. From Fig. 2(f) we infer that the nerve impulse can be detected optically with a signal to noise ratio $SNR \approx 37$ using 1000 averages. As the SNR scales as $1/\sqrt{N_{\text{avg}}}$ we find the $SNR \approx 1.2$ for a single shot, i.e., it should be possible to detect a nerve impulse in a single shot.

As a control experiment, we make the nerve inexcitable by replacing the saline solution in the plastic chamber with a solution with high potassium concentration. As expected, we clearly observe from both electrical [Fig. 2(c)] and optical [Fig. 2(f)] measurements that the nerve signal is blocked by this solution. Note that the stimulation artifact which was observed in the electrical recording (inset to Fig. 2(a)) should be small in the optical measurements, as the stimulation occurs during the optical pumping [see Fig. 1(c)] where the response to magnetic fields is strongly damped. When the nerve is inexcitable, we do see such a small stimulation artifact (squares in Fig. 2(f)) for the higher stimulation voltages.

The high spatial resolution of our magnetometer (the channel containing the cesium atoms is 1.0 mm across) and the good signal to noise ratio allows us to characterize how the magnetic field decays with distance from the nerve. The results are shown in Fig. 3 and one sees that we can detect the nerve impulse more than 5 mm away. The actual distance dependence may be complicated as the frog sciatic nerve contains 1000’s of axons organized in several bundles (see Supplementary Fig. 1). However, we expect a power law dependence $B(x) \propto 1/(x + x_0)^n$ with $n$ between 1–3. $n = 1$ corresponds to the magnetic field from an infinitely long wire and $n = 3$ corresponds to the field from a magnetic dipole. $x_0$ is the position of the nerve relative to our estimate of 1.9(5) mm based on the size of the vapor cell and dimensions of the nerve. Fitting the data to a power law dependence yields $x_0 = 0.2(8)$ mm and $n = 1.5(4)$ which are within the expected ranges.

The magnetometer can also be operated in the continuous mode which allows for determination of the temporal shape of the magnetic field generated by the nerve, $B_{\text{nerve}}(t)$. The magnetometer response was optimized by matching its frequency response (a Lorentzian centered at the Larmor frequency with a full width at half

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**Figure 2.** Electrical and optical measurements of the nerve impulse for different stimulation voltages. The optical measurements were done in the pulsed mode using 1000 averages. The figures show the signals in time, the square root of the power spectral density $PSD$ and the 400 Hz frequency component. The plotted electrical signals are after 10 times amplification. The uncertainties on the data points in (c) are small to be visible in the figure. The uncertainties on the points in (f) can be estimated from the points without stimulation (0 V) which were measured 9 times and resulted in a 0.25(10) pT·ms signal. By dividing the nerve signal (9.1 pT·ms) by the noise floor obtained without stimulation (0.25 pT·ms) we find the signal to noise ratio, $SNR \approx 37$. and Fig. 2(c) shows the amplitude of the 400 Hz Fourier component. The nerve is stimulated at $t = 6.0$ ms. The signature of the nerve signal is its non-linear behavior with stimulation voltage, with a firing threshold at around 0.4 V. For voltages above the threshold a nerve impulse is measured with the recording electrodes within the time interval $t = 7.0 \pm 10.0$ ms. We also observe a stimulation artifact at $t = 6.0$ ms [see inset in Fig. 2(a)] which is proportional to the stimulation voltage.

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Figure 4(c) shows a comparison of the electrical signal for 0.8 V stimulation and the detected magnetic field signal, as it lasts for several ms and only appears above the threshold for the nerve firing (here 0.8 V or greater). Feature B is due to the nerve and its electrical excitation. Figure 4(a,b) show the temporal profiles of the electrical signal and the magnetic field level, which shows some similarity, as both show the action potential at the time of stimulation and increases linearly with the stimulation voltage. Feature A is due to the stimulation as it starts at the time of stimulation and increases linearly with the stimulation voltage. Feature B is due to the nerve signal, as it last for several ms and only appears above the threshold for the nerve firing (here 0.8 V or greater).

In conclusion, we have demonstrated non-invasive detection of nerve impulses from the frog sciatic nerve by measuring the magnetic field generated by the nerve with a room-temperature sensor with near quantum limited sensitivity. A mm-sized sensor which is sensitive enough to detect sub-picoTesla fields at a distance of a few millimeters from biological objects makes the magnetometer perfect for medical diagnostics. We envision that additional compensation of the ambient magnetic fields and classical fluctuations of the atomic spins could allow for sensitivity beyond the PN limit as well as providing additional compensation of the ambient magnetic fields and classical fluctuations of the atomic spins.

Methods

Nerve preparation. This study was conducted in accordance with the University of Copenhagen Animal Ethics Policy and the frogs were used by the animal facility at the Department of Experimental Medicine at the University of Copenhagen which is accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International (AAALAC). The experiments presented here with the frog sciatic nerve do not require a license.

Sciatic nerves were isolated from green frogs (Rana Esculenta). The frogs were decapitated and the sciatic nerves were isolated from spine and down to the knee. The nerves are 7–8 cm long with a diameter of 1.3 mm. The experiments were conducted in accordance with the AAALAC guidelines and the Ethics Policy of the University of Copenhagen.

Discussion

Projection noise dominated sensitivity can be reached by relatively straightforward steps, such as using multipass vapor cells, by modest heating (increasing the temperature from room-temperature 22 °C to the human body temperature 37 °C will increase the sensitivity by a factor of two) or by employing a low finesse optical cavity. Gradiometry with two cells with oppositely oriented spins allows for generation of nonclassical entangled states leading to sensitivity beyond the PN limit as well as providing additional compensation of the ambient magnetic fields and classical fluctuations of the atomic spins.

In conclusion, we have performed non-invasive detection of nerve impulses from the frog sciatic nerve by measuring the magnetic field generated by the nerve with a room-temperature sensor with near quantum limited sensitivity. A mm-sized sensor which is sensitive enough to detect sub-picoTesla fields at a distance of a few millimeters from biological objects makes the magnetometer perfect for medical diagnostics. We envision that practical, low-cost, and room-temperature atomic magnetometers could be an alternative to the current technology based on cryogenically-cooled SQUID magnetometers which are widely used for magnetoencephalography (MEG) and neuroscience studies. Promising applications of atomic magnetometers include MEG, detection of epilepsy, detection of the synaptic responses in the retina, and cardiography of fetuses.
Figure 4. Electrical and optical measurements of the nerve impulse for different stimulation voltages. The magnetometer was operated in the continuous mode and the signals were averaged 5000 times. (a) Electrical and (b) optical measurements. (c) Electrical signal for 0.8 V stimulation and magnetic field calculated by deconvolution. For these specific measurements the Larmor frequency was 510 Hz and the coherence time 0.37 ms.

Nerve chamber and electrical recording. The nerves are kept in a 3D-printed plastic chamber during the experiments. The chamber is a 47 × 18 mm block of 8.5 mm height that contains a longitudinal U-shaped channel with a diameter of 2 mm in which the nerve is placed. The channel allows maintenance of a saturated water-vapor atmosphere in order to keep the nerve moist. The front part of the chamber is placed close to magnetometer and it is covered by a microscope glass cover slip of 0.13 mm thickness.

The chamber has 6 circular gold electrodes on each side. The electrodes have an outer diameter of 6 mm with the hole inside of 1.5 mm, which fits into the channel. On each side, the distance between electrodes is 5 mm.

The nerve was externally stimulated in the proximal end by applying a short 50 μs square voltage pulse between two spatially separated electrodes surrounding the nerve. The nerve signal was measured in the distal end as a potential difference between a pair of two electrodes. The electrical signal was amplified 10 times and filtered (using a 3 kHz low pass and a 10 Hz high pass filter) and recorded simultaneously with the magnetic field recordings.

Operation of the magnetometer. The magnetometer is based on optical read-out of spin-polarized atomic vapour. The cesium atoms are prepared by an optical pumping with a pulse of circular polarized light such that the total spin vector \( \mathbf{J} = (J_x, J_y, J_z) \) points in the x-direction, which is also the direction of a bias field \( B_x \) [see Fig. 1(a)]. Any magnetic field perpendicular to x-direction (such as the magnetic field from the nerve) will create a transverse spin component \( J_y \), which afterwards will precess around the bias magnetic field at the Larmor frequency \( \Omega = B_x / \gamma \) [see Fig. 1(b)], where \( \gamma = 2.20 \cdot 10^{10} \text{rad/(s·T)} \) is the cesium gyromagnetic ratio. The transverse spin component is detected optically by measuring the polarization rotation of linearly polarized probe light passing through the vapor cell using a balanced polarimeter. The magnetic field from a nerve can be detected in two modalities, a pulsed or a continuous one. In the continuous mode the pulse and probe light is continuously on. In this case, the magnetometer signal \( S(t) \) is proportional to the convolution of the magnetic field with the magnetometer response function \( S(t) = \int_{-\infty}^{\infty} \left[ e^{-\Gamma(t-t')} \cos(\Omega(t-t')) \right] B_x(t') dt' \), where we assumed the transverse field is along the y-direction. The relaxation rate \( \Gamma = 1/T_2 \), which is the inverse of the spin-coherence time, increases linearly with laser power and is in the limit of low power denoted \( \Gamma_{\text{dark}} = 1/T_2^{\text{dark}} \). In the pulsed mode [see Fig. 1(c)], a pulse of pump light first initializes the atomic spins along the x-direction, then the pulse of magnetic field creates a transverse spin component, and finally the spins are detected with a pulse of probe light. In this case, the magnetometer signal is a free induction decay \( S(t) = A \sin(\Omega t + \theta) e^{-\tau T_1} \), where \( \theta \) is a phase and the amplitude \( A \) is proportional to the Fourier component of the magnetic field at the Larmor frequency: \( |B(\Omega)| \sim \int_0^\infty B_x(t) e^{-\tau T_1} dt \), where \( \tau \) is the duration of the magnetic field pulse. For both pulsed and continuous modes, the proportionality constant can be found by applying a known calibration magnetic field.

Calibration of the pulsed magnetometer. The magnetometer is calibrated by applying a known magnetic field. This calibration field is produced by a coil positioned inside the magnetic shield; the field points in the y-direction and has the temporal shape of a single sinusoidal oscillation \( B_{\text{cal}}(t) = B_{\text{cal}} \sin(\Omega_{\text{cal}} t) \) with amplitude \( B_{\text{cal}} = 2 \text{nT} \), frequency \( \Omega_{\text{cal}} = 2\pi \cdot 400 \text{Hz} \) and Fourier component \( |B_{\text{cal}}(\Omega_{\text{cal}})| = \pi B_{\text{cal}} / \Omega_{\text{cal}} = 2.5 \text{nT·ms} \).
In the pulsed mode, the calibration field is applied in between the pump and probe pulses. The recorded magnetometer signal (the free induction decay) is shown in Supplementary Fig. 2(a). The spectrum (calculated as the square-root of the power spectral density $\sqrt{PSD}$) is peaked at the Larmor frequency of the atoms $\Omega = 2\pi \cdot 400$ Hz [Supplementary Fig. 2(b)]. The peak amplitude is proportional to the magnetic field Fourier component: $\sqrt{PSD} \propto |B(t)|$ The proportionality constant can be calculated from the data in Supplementary Fig. 2(b) and the known Fourier component of the calibration field. With this calibration we can calculate the Fourier component $|B_{\text{meas}}(t)|$ of the nerve magnetic field [see Fig. 2(f)] from the measured peak values $\sqrt{PSD} \propto |B(t)|$ [see Fig. 2(e)].

Supplementary Fig. 2(a,b) also show the magnetometer signal without the applied calibration field. The signal was averaged 1000 times before recorded. The noise at the Larmor frequency is 10200 times smaller than the signal obtained with the calibration field, i.e., the calibration field is detected with a SNR $= 10200$ corresponding to a minimal detectable magnetic field Fourier component 2.5 nT/ms/10200 = 0.25 pT·ms using 1000 averages. The single shot minimal detectable Fourier component is then 7.7 pT·ms.

**Calibration of the continuous magnetometer.** In the continuous mode, the lasers are on during the applied calibration field. In this case the magnetometer signal is proportional to the convolution of the magnetic field with the magnetometer response function. Supplementary Fig. 2(c) top shows the detected signal together with a fit to the function $S(t) \propto \int_0^t [\exp(t-t') \cos(\Omega (t-t'))] B(t) dt'$ from which we can determine the Larmor frequency and the coherence time. Using the fitted parameters we can perform numerical deconvolution and by scaling with the amplitude of the calibration field we can obtain the magnetic field as a function of time $B(t)$ [Supplementary Fig. 2(c) bottom]. We see that the deconvolution procedure works well as the deconvolved signal resembles a single sinusoidal oscillation.

Supplementary Fig. 2(d) shows the calculated noise spectrum of the deconvoluted signal. It also shows the noise spectrum when the calibration field was off. In this case the noise at the Larmor frequency is $23 \text{ fT}/\sqrt{\text{Hz}}$ when 100 averages are used. The single shot magnetic field sensitivity is then $230 \text{ fT}/\sqrt{\text{Hz}}$.

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**Acknowledgements**

We would like to thank Bing Chen for help during the initial stages of the experiments, and Hannelouise Kissow for for taking the electron micrograph image of the nerve. This work was supported by the ERC grant INTERFACE and by the DARPA project QUASAR. K. J. was supported by the Carlsberg Foundation. R. A. T. was supported by CNPq/Brazil.

**Author Contributions**

K.J., R.B., R.A.T. and H.C.S. performed the experiments. A.M.F. and S.-P.O. prepared the frog nerves. M.V.B. fabricated the vapor cell. G.V. and J.H.M. contributed with expertise on magnetometry and experimental
techniques and L.D.M. contributed with expertise on the nerves' electric and magnetic responses. K.J., G.V., E.S.P. and S.-P.O. wrote the manuscript. T.H., S.-P.O. and E.S.P supervised the research. All authors discussed the results and commented on the manuscript.

Additional Information
Supplementary information accompanies this paper at http://www.nature.com/srep

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Jensen, K. et al. Non-invasive detection of animal nerve impulses with an atomic magnetometer operating near quantum limited sensitivity. Sci. Rep. 6, 29638; doi: 10.1038/srep29638 (2016).

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