Androgenic response of \textit{Triticum durum–Dasypyrum villosum} amphidiploids and their parental forms

H. Stoyanov, I. Belchev

Dobrudzha Agricultural Institute, General Toshevo, Bulgaria

hpstoyanov@abv.bg

Abstract. Wide hybridization in cereal crops is one of the most efficient tools for the enrichment of genetic variability and addressing a number of breeding problems related to resistance and tolerance to biotic and abiotic stresses. Therefore, a large number of amphidiploids between species possessing different morphological, genetic and physiological properties have been developed. One of the most valuable species with regard to the possibilities for introducing valuable traits and properties into wheat species is the wild \textit{Dasypyrum villosum}. With the aim to study the androgenic response of the \textit{Triticum durum–D. villosum} amphidiploids, two accessions and their parental forms – the durum wheat cultivars Gergana and Argonavt and a landrace of the \textit{D. villosum} – were studied. The following parameters were determined: callus induction, plant regeneration, yield of albino and green regenerants. It was found that the callus induction of the two studied amphidiploids differed significantly from that of the parental forms (2.1–7.2 \%), being significantly higher, 30.7 and 16.5 \%, respectively. Regardless of the difference in callus induction, the amphidiploids did not significantly differ from the parental forms in their regeneration ability. The yield of albino plants exceeded the yield of green regenerants and followed the tendency observed in callus induction. Green plants were found only in the amphidiploid Gergana-\textit{D. villosum} and in the parental form durum wheat Gergana. Plants were regenerated from the species \textit{D. villosum}, although they were only albinos, showing its good responsiveness to anther culture. The established characteristics of the amphidiploids and their parental forms make their practical use highly valuable for the improvement of different types of cereal crops.

Key words: anther culture; androgenic response; amphidiploid; \textit{Dasypyrum villosum}; parental forms.

For citation: Stoyanov H., Belchev I. Androgenic response of \textit{Triticum durum–Dasypyrum villosum} amphidiploids and their parental forms. Vavilovskii Zhurnal Genetiki i Selektsi = Vavilov Journal of Genetics and Breeding. 2022;26(2):139-145. DOI 10.18699/VJGB-22-17

Андрогенетическая реакция амфидиплоидов \textit{Triticum durum–Dasypyrum villosum} и их родительских форм

Х. Стоянов, И. Белчев

Добруджанский сельскохозяйственный институт, г. Генерал Тошево, Болгария

hpstoyanov@abv.bg

Аннотация. Отдаленная гибридизация злаков является одним из самых эффективных способов обогащения генетического разнообразия и решения множества селекционных задач в отношении устойчивости и толерантности к биотическому и абиотическому стрессу. Поэтому создано большое количество амфидиплоидов между отдельными видами, которые наделены разными морфологическими, генетическими и физиологическими характеристиками. Дикий вид \textit{Dasypyrum villosum} – один из самых ценных видов с точки зрения возможности интродуктирования ценных качеств и свойств в разные сорта пшеницы. Для изучения реакции амфидиплоидов \textit{Triticum durum–D. villosum} исследованы два образца, их родительские формы – сорта твердой пшеницы Гергана и Аргона, а также местная популяция вида \textit{D. villosum}. Установлены следующие параметры: индукция каллусов, частота всех проростков к 100 эмбриоподобным структурам, частота альбиносных проростков и частота зеленых проростков. По полученным результатам выявлено, что индукция каллусов двух амфидиплоидов статистически достоверно отличается от родительских форм (2.1–7.2 \%), будучи значительно выше, 30.7 и 16.5 \% соответственно. Несмотря на разницу в индукции каллусов, амфидиплоиды практически не отличаются от родительских форм своей регенерирующей способностью. Частота альбиносных проростков значительно превышает частоту зеленых проростков, следуя тенденции, наблюдаемой в индукции каллусов. Зеленые проростки зарегистрированы только у амфидиплоида Гергана-\textit{D. villosum} и у родительской формы твердой пшеницы Гергана. Растения вида \textit{D. villosum} были регенерированы, и то, что были только альбиносные проростки, показывает хорошую отзывчивость вида на культивирование пыльников. Установленные характеристики испытанных амфидиплоидов и их родительских форм делают практическое использование этих амфидиплоидов особенно ценным для селекции различных видов злаков.

Ключевые слова: пыльниковая культура; андрогенетическая реакция; амфидиплоид; \textit{Dasypyrum villosum}; родительские формы.

© Stoyanov H., Belchev I., 2022
This work is licensed under a Creative Commons Attribution 4.0 License
The aim of this study was to determine the reaction of the amphidiploid *T. durum-D. villosum* to anther cultivation in comparison to its parental forms.

**Materials and methods**

**Plant material.** Two accessions of the amphidiploid *T. durum-D. villosum* (1dv (Gergana-*D. villosum*) and 2dv (Argonavt-*D. villosum*)), a part of the collection of Dobrudzha Agricultural Institute were used, as well the durum wheat parental forms (*T. durum* cv. Gergana and cv. Argonavt) and the wild species *D. villosum*.

The accession of *D. villosum* (2n = 2x = 14 (VV); family Poaceae, tribe Triticeae, subtribe Triticeinae, genus *Dasypryrum*) was collected in Dobrich region in 2011.

Crosses Gergana × *D. villosum* and Argonavt×*D. villosum* were made conventionally, without embryo rescue in 2012; the obtained seeds (Gergana × *D. villosum* – 3 seeds and Argonavt×*D. villosum* – 8 seeds) were germinated and at tillering stage the plants (Gergana × *D. villosum* – 1 plant and Argonavt×*D. villosum* – 3 plants) were treated with colchicine in 2013. The seeds from the two obtained primary amphidiploids were multiplied several times.

**Anther culture.** The experiment was carried out during 2016/2017. Anther donor plants were grown under greenhouse conditions. The seeds from the accessions were germinated in Petri dishes and then planted in plastic pots. Fifteen plants from each accession were grown in three pots, using 10 plants per genotype. Primary, seedling were vernalized at 4 °C (3000 lx, 16 h day/8 h night) for 45 days. After this period, the plants were transferred to a cold greenhouse (5–15 °C) for about three months, and the temperature was later increased to 15–20 (25 °C). Tillers bearing spikes containing anthers with microspores at mid- to late uninucleate stage were cut, put in a vessel with water and pretreated at 4 °C for 8–9 days. Ten spikes from each genotype were collected. Cold pretreated spikes were surface sterilized with 70 % ethanol and aseptic conditions. Sixty anthers from each spike were placed in test tubes with 20 ml P2 induction medium (Chuang et al., 1978). The anthers were cultured at 28 °C in darkness for about 60 days. After the 30th day, they were periodically checked for induction of embryogenic structures (calli and embryoids), which were transferred to test tubes with 10 ml regeneration medium (Zhuang, Jia, 1983) and cultured at 25 °C (3000 lx, 16 h day/8 h night). Green and albino regenerants were counted after 30 days.

The androgenic response was estimated by the following traits: callus induction (CI) (number of embryogenic structures induced per 100 cultured anthers, %), plant regeneration (PR) (number of regenerated green and albino plants per 100 embryogenic structures, %), frequency (yield) of green plants (YGR) (number of regenerated green plants per 100 cultured anthers, %) and frequency (yield) of albino plants (YAR) (number of regenerated albino plants per 100 cultured anthers, %).

**Statistics.** The obtained results were summarized over genotypes and parameters. One way ANOVA was carried out with the aim of determining the effect of the genotype on the studied parameters to estimate their androgenic response. Significant differences between the amphidiploids and their
parental forms were calculated based on the Duncan test. To process the data, software MS Office Excel 2003 was used, and to perform ANOVA and the Duncan test – IBM SPSS Statistics v.19.

Results
The results on the androgenic potential of the investigated amphidiploids and their parental forms (Table 1) showed that accession 1dv had the highest callus induction (30.7 %), and durum wheat Argonavt – the lowest (2.1 %). Between the parental forms, there were no significant differences (both between the durum wheat cultivars and between the species durum wheat and D. villosum). The two amphidiploids differed significantly by their callus induction, which was probably related to the effect of the maternal component.

The yield of green plants, averaged for the entire investigated set, was extremely low. In the entire experiment, only 5 green regenerants were produced, one of them being from the durum wheat cultivar Gergana, and the other 4 – from the amphidiploid 1dv (Gergana-Dv). No green regenerants were obtained from cultivar Argonavt and from the amphidiploid 2dv. Also, no green plants were produced from the wild species D. villosum. Although there was a rather small number of the obtained plants for formulating a general tendency for the effect of the parental forms, the presence of green plants in cultivar Gergana and the amphidiploid, in which it was involved, was probably due to genotypic specificity.

The albino plants considerably exceeded the green regenerants. In practice, they were predominant with regard to the total number of regenerants. The amphidiploid 1dv again had the highest yield of albino plants (10.3 %), and the lowest values were observed in cultivar Argonavt (0.4 %). The tendency in yield of obtained albinos largely followed the tendency of callus induction. The two amphidiploids significantly differed from the parental forms by their values, as well as between themselves (10.3 and 5.8 %, respectively). Meanwhile, significant differences between the two durum wheat cultivars and between the durum wheat and the wild species were not registered. The higher yield values of the albino plants in the amphidiploid 1dv may be related to the higher responsiveness of cultivar Gergana, which was the maternal component of this amphidiploid, although the difference between Gergana and Argonavt was not significant.

On the whole, plant regeneration, expressed as a number of regenerants per 100 embryogenic structures, was comparatively low. The highest values were read in the two investigated amphidiploids (35.9 and 35.4 %, respectively), and the lowest – in the wild species D. villosum (13.0 %). This parameter did not follow the tendency observed in the values of callus induction and yield of green and albino regenerants. There were no significant differences between any of the studied accessions. However, higher plant regeneration was registered in the amphidiploids, in comparison to cultivar Argonavt and the wild species, and the difference with cultivar Gergana was considerably lower. The differences not being significant was an indication that the regeneration potential of all studied accessions was practically identical, and the differences formed were entirely random. The total number of regenerants, however, expressly followed the tendency of callus induction and yield of albino plants. The higher responsiveness to anther culture of the two investigated amphidiploids in comparison to either of the parental forms could be clearly observed in this parameter.

The results from the analysis of the variance of the studied parameters (Table 2) showed that the genotype had a significant effect on the parameters callus induction and yield of albino regenerants. This allows supposing that the separate accessions gave specific responses and that there are significant differences between them, as determined by the Duncan test that was carried out. At the same time, the effect of the parental forms was calculated based on the Duncan test. To process the data, software MS Office Excel 2003 was used, and to perform ANOVA and the Duncan test – IBM SPSS Statistics v.19.

Table 1. Androgenic response of parental forms and durum wheat-D. villosum amphidiploids

| Genotype       | NCA | NOC | CI, % | NRC | PR, % | NAR | YAR, % | NGR | YGR, % |
|----------------|-----|-----|-------|-----|-------|-----|--------|-----|--------|
| Gergana        | 600 | 43  | 7.2ab | 13  | 30.2a | 12  | 2ab    | 1   | 0.2ab  |
| Argonavt       | 480 | 10  | 2.1a  | 2   | 20.0a | 2   | 0.4a   | 0   | 0.0a   |
| D. villosum (Dv)| 540 | 23  | 4.3ab | 3   | 13.0a | 3   | 0.6a   | 0   | 0.0a   |
| 1dv (Gergana-Dv)| 600 | 184 | 30.7c | 66  | 35.9a | 62  | 10.3c  | 4   | 0.7b   |
| 2dv (Argonavt-Dv)| 480 | 79  | 16.5b | 28  | 35.4a | 28  | 5.8b   | 0   | 0.0a   |

Note. NCA – number of cultivated anthers; NOC – number of obtained calli; CI – callus induction; NRC – number of regenerative calli; PR – plant regeneration; NAR – number of albino regenerants; YAR – yield of albino regenerants; NGR – number of green regenerants; YGR – yield of green regenerants.
Table 2. ANOVA according to factor “accession” of the studied accessions

| Parameters | Sum of squares | df  | Mean square | F    | Significance |
|------------|---------------|-----|-------------|------|--------------|
| CI         |               |     |             |      |              |
| Between groups | 0.51892 | 4   | 0.12973     | 8.693 | 0.000        |
| Within groups   | 0.59697 | 40  | 0.01492     |       |              |
| Total          | 1.11589 | 44  |             |      |              |
| YAR          |               |     |             |      |              |
| Between groups | 0.06774 | 4   | 0.01694     | 7.799 | 0.000        |
| Within groups   | 0.08686 | 40  | 0.00217     |       |              |
| Total          | 0.15460 | 44  |             |      |              |
| YGR          |               |     |             |      |              |
| Between groups | 0.00032 | 4   | 0.00008     | 2.159 | 0.091        |
| Within groups   | 0.00147 | 40  | 0.00004     |       |              |
| Total          | 0.00179 | 44  |             |      |              |
| PR           |               |     |             |      |              |
| Between groups | 0.36055 | 4   | 0.09014     | 0.899 | 0.474        |
| Within groups   | 4.01134 | 40  | 0.10028     |       |              |
| Total          | 4.37188 | 44  |             |      |              |

Note. CI – callus induction; YAR – yield of albino regenerants; YGR – yield of green regenerants; PR – plant regeneration.

Table 3. Androgenic response of parental forms and durum wheat-D. villosum amphidiploids

| Species         | NCA  | NOC | CI  | NRC | PR  | NAR | YAR, % | NGR | YGR, % |
|-----------------|------|-----|-----|-----|-----|-----|--------|-----|--------|
| T. durum (Td)   | 1080 | 53  | 4.9a | 15  | 28.3a | 14  | 1.3a   | 1   | 0.1a   |
| D. villosum (Dv) | 540  | 23  | 4.3a | 3   | 13.0a | 3   | 0.6a   | 0   | 0.0a   |
| Td-Dv           | 1080 | 263 | 24.4b| 94  | 35.7a | 90  | 8.3b   | 4   | 0.4a   |

Note. NCA – number of cultivated anthers; NOC – number of obtained calli; CI – callus induction; NRC – number of regenerative calli; PR – plant regeneration; NAR – number of albino regenerants; YAR – yield of albino regenerants; NGR – number of green regenerants; YGR – yield of green regenerants.

Table 4. ANOVA according to factor “species” of the studied accessions

| Parameters | Sum of squares | df  | Mean square | F    | Significance |
|------------|---------------|-----|-------------|------|--------------|
| CI         |               |     |             |      |              |
| Between groups | 0.41771 | 2   | 0.20885     | 12.564 | 0.000        |
| Within groups   | 0.69818 | 42  | 0.01662     |       |              |
| Total          | 1.11589 | 44  |             |      |              |
| YAR          |               |     |             |      |              |
| Between groups | 0.05763 | 2   | 0.02881     | 12.480 | 0.000        |
| Within groups   | 0.09698 | 42  | 0.00231     |       |              |
| Total          | 0.15460 | 44  |             |      |              |
| YGR          |               |     |             |      |              |
| Between groups | 0.00011 | 2   | 0.000005    | 1.349 | 0.271        |
| Within groups   | 0.00168 | 42  | 0.00004     |       |              |
| Total          | 0.00179 | 44  |             |      |              |
| PR           |               |     |             |      |              |
| Between groups | 0.31393 | 2   | 0.15696     | 1.625 | 0.209        |
| Within groups   | 4.05795 | 42  | 0.09662     |       |              |
| Total          | 4.37188 | 44  |             |      |              |

Note. CI – callus induction; YAR – yield of albino regenerants; YGR – yield of green regenerants; PR – plant regeneration.

both genotype and species, did not differ as a tendency. The observed differences were not significant (see Tables 3 and 4), which indicated that the studied amphidiploid did not differ from the parental forms by its regeneration capacity.

Discussion
Concerning the results obtained on the androgenic response of the used accessions, it should be emphasized, that no source was found in world literature that would present data on the amphidiploid T. durum-D. villosum or the species D. villosum. An exception was the research of X. Chen et al. (1996), who suggested applying the anther culture method on hybrids (not amphidiploids) of the F₁ (T. durum × D. villosum). These authors reported successful production of amphidiploids, regenerated from colchicine-treated calli. At the same time, there are researches on the use of tissue cultures on three-
component hybrids *T. aestivum × (T. durum-D. villosum)*. H. Li et al. (2005) reported lines with high powdery mildew resistance obtained from such hybrids through the method of embryo rescue and subsequent anther culture.

D. Plamenov et al. (2009), when investigating the androgenic response of accessions from the amphidiploid *T. durum-T. monococcum* ssp. *aegilopoides*, came up with results different from ours. The reported callus induction was 3.3–11.7 % for the two studied accessions, the plant regeneration was considerably higher, 33.8–68.4 %, respectively, and the albino regenerants yield was 1.9–3.2 %. At the same time, the yield of green plants (0.4–0.8 %) was a little higher than the data we obtained in our experiment (0.0–0.7 %). These authors reported a total of seven regenerated plants from both accessions, this parameter being significant, unlike the results we obtained. Using anther culture in the amphidiploid *Ae. variabilis-S. cereale*, and P2 medium, A. Ponitka et al. (2002) observed 1.4–15.7 % of callus induction, and on C17 medium – 20.0–65.2 %. Subsequently, the authors reported 0.1–13.4 % yield of green regenerants using 190-2 regeneration medium. It was found out that the androgenic response was strongly dependent on the genotype, similar to the results of the experiment we conducted. Successful regeneration of green plants through the method of anther culture has also been reported for an aneupolyhaploid of *Thynropyrum ponticum* (Wang et al., 1991), for the amphidiploid *Festuca pratensis-Lolium multiflorum* (Lesniewska et al., 2001; Zwierzykowski et al., 2001; Rapacz et al., 2005) and the amphidiploid *Cyclamen persicum-C. purpurascens* (Ishizaka, 1998).

In contrast to these results, the parental forms were characterized with much lower androgenic response. This was confirmed by the absence of callus induction in *Ae. variabilis* and rye, reported by A. Ponitka et al. (2002), and also in the species *T. monococcum* ssp. *aegilopoides* in the research of D. Plamenov et al. (2009). Durum wheat is also characterized by weak androgenic response, in general. M. Dogramaci-Altunetepe et al. (2001), using 10 durum wheat genotypes, obtained only 248 green regenerants from 86,400 anthers (0.29 %). F. J’Aití et al. (1999), investigating 15 durum wheat genotypes and 7500 cultivated anthers, obtained just three albino regenerants and one green plant.

L. Cistúe et al. (2006), on the other hand, reported significantly higher production of green plants, but including 6-benzylaminopurine or 6-furfurilaminopurine in the induction medium (C17). In more recent researches, the production of haploids, even by the method of isolated microspores, has been of extremely low efficiency in durum wheat (Slama-Ayed et al., 2019). These results entirely corresponded to the data we obtained with regard to the two cultivars Argonavt and Gergana. Clear genotypic specificity was observed in the better response of Gergana to anther culture as compared to Argonavt, although the difference was not statistically significant. It is probable that this tendency is the reason for the amphidiploid Gergana-*D. villosum* having better responsiveness to anther culture. In this respect, the amphidiploid *T. durum-D. villosum* we investigated, and the amphidiploids reported by A. Ponitka et al. (2002) and D. Plamenov et al. (2009) were closer by their androgenic response to the response of triticale (which is a typical amphidiploid crop) than to the response of the parental forms. J. Pauk et al. (2000), K. Marciniak et al. (2003), C. Lantos et al. (2014) and H. Stoyanov et al. (2019) demonstrated that in triticale the albino regenerants are often predominant, similar to the amphidiploid we studied. The values of the green regenerants in triticale also varied (from 0.9 to 27.9 %, but more often within 3–6 %), according to data from various researches (Gonzales, Jouve, 2000; Marciniak et al., 2003; Banaszak, 2011; Lantos et al., 2014).

In contrast to the above responses of the parental forms *Ae. variabilis, S. cereale* and *T. monococcum* ssp. *aegilopoides*, our study, although limited in volume, demonstrated the comparatively good responsiveness of the species *D. villosum* to anther culture. This is the first time when results on regenerants from this species (although only albinos) are being reported. At the same time, it should be emphasized that until this moment results from testing of the reaction of *D. villosum* to the anther culture method have never been reported. This is highly significant for the breeding of the wheat species since it would allow transferring genes from the wild species through the methods of wide hybridization and anther culture more easily, quickly and efficiently. X. Chen et al. (1996) and C. Li et al. (2000) reported common wheat lines resistant to powdery mildew, which were obtained by crossing common wheat to the amphidiploid *T. durum-D. villosum*, followed by embryo rescue and anther culture. Such results showed that the combination of wide hybridization with the method of anther culture is an efficient tool that can be used in the breeding of different cereal crops.

**Conclusion**

Based on the presented results, the following conclusions could be made:

1. For the first time, results on the androgenic response (callus induction, plant regeneration, yield of albino plants, yield of green plants) of the amphidiploid *T. durum-D. villosum* and of the parental component *D. villosum* are being reported.

2. The callus induction of the two studied amphidiploids differed significantly from that of the parental forms (2.1–7.2 %), being considerably higher – 30.7 and 16.5 %, respectively.

3. The plant regeneration of the investigated accessions varied within a certain range (13.0–35.9 %), the differences not being statistically significant. This indicated that in spite of the differences in the callus induction, the amphidiploids did not practically differ from the parental forms by their regeneration capacity.

4. Although plant regeneration was observed in all studied accessions, the yield of albino plants considerably exceeded the yield of green regenerants and followed the tendency observed in callus induction – the two amphidiploids had significantly higher values. At the same time, green plants were registered only in the amphidiploid Gergana-*D. villosum* and in the parental form durum wheat Gergana. Such results emphasized the genotypic specificity of the response to anther culture.

5. Plants were regenerated from the species *D. villosum*, although only albinos, which indicated its good responsiveness to anther culture. This, together with the good response of the amphidiploids with the participation of this species, makes their practical use, in combination with the anther culture method, highly valuable for improving the cereals.
Androgenic response of Triticum durum-Dasypyrum villosum amphidiploids and their parental forms

References

Ando K., Krishna N., Rynesnor S., Rouse M.N., Danilova T., Friebe D., and Pumphey M.O. Introgression of a novel Ug99-effective stem rust resistance gene into wheat and development of Dasyypyrum villosum chromosome-specific markers by genotyping-by-sequencing (GBS). Plant Dis. 2019;103:1068-1074. DOI 10.1099/PI-501-0-18031-RE.

Babaiants O.V., Babaiants L.T., Horash A.F., Vasil'ev A.A., Trakoveskaia V.A., Paliashii V.A. Genetics determination of wheat resistance to Puccinia graminis f.sp. tritici deriving from Aegilops cylindrica, Triticum eureni and amphidiploid 4. Tistol. Genet. 2012; 46(1):10-17. (in Ukrainian)

Banaszak Z. Breeding of triticale in DANKO. In: Tagungsband der 61. Jahrestagung der Vereinigung der Pflanzenzüchter und Saatgutkaufleute. p. 23–25 November 2010. Raumberg-Gumpenstein. Ibidn. 2011;65-68.

Barceló P., Cabrera A., Hégel C., Lötz H. Production of doubled-haploid plants from tritوردum anther culture. Theor. Appl. Genet. 1994; 87:741-745.

Belchev I. Studies on Anther Culture of Common Winter Wheat (Triticum aestivum L.) and Application of Double Haploid Lines in Breeding. General Toshevo, 2003. (in Bulgarian)

Chahal G.S., Gosal S.S. Principles and Procedures of Plant Breeding: Biotechnological and Conventional Approaches. New York: CRC Press, 2000.

Chen X., Xu H.J., Du L.P., Shang L.M., Han B., Shi A., Xiao S. Transfer of gene resistant to powdery mildew from H. villosa to common wheat by tissue culture. Sci. Agric. Sin. 1996;29:1-8.

Chuang C.C., Ouyang T.W., Chia H., Chou S.M., Ching C.K. A set of potato media for wheat anther culture. In: Proc. of Symposium on Plant Tissue Culture 1978. Peking: Science Press, 1978;51-56.

Cistué L., Soriano M., Castillo A.M., Valls M.P., Sanz J.M., Echávarri B. Production of doubled haploids in durum wheat (Triticum turgidum L.) through isolated microspore culture. Plant Cell Rep. 2006;25(4):257-264. DOI 10.1007/s00299-005-0047-8.

Dagüstü N. Diallel analysis of anther culture response in wheat (Triticum aestivum L.). Afr. J. Adv. Biotechnol. 2008;7(9):3419-3423.

Dai S., Li Z., Xue X., Jia Y., Liu D., Pu Z., Zheng Y., Yan Z. Analysis of high-molecular-weight gluten subunits in five amphidiploids and their parental diploid species Aegilops umbellulata and Aegilops uniaristata. Plant Genet. Resour. 2015;13(2):186-189. DOI 10.1074/S1749262114000719.

De Pace C., Snidaro D., Ciaffi M., Vittori D., Ciofo A., Cenci A., Tanzarella O.A., Qualset C.O., Mugnouza G.T.S. Production of doubled haploids in durum wheat and their cytological characterization. Plant Cell. Mol. Biol. Lett. 2011;70963.

De Pace C., Vaccino P., Cionini P.G., Pasquini M., Bizzarri M., Qualset C.O. Ditye Austere 23–25 November 2010. Raumberg-Gumpenstein. Ibidn. 2011;65-68.

Grassy P., Eskridge K., Cassman K. Distinguishing between yield advances and yield plateaus in historical crop production trends. Nat. Commun. 2013;4:2918. DOI 10.1038/ncomms3918.

Ishizaka H. Production of microspore-derived plants by anther culture of an interspecific F1 hybrid between Cyclamen persicium and C. purpurascens. Plant Cell Tissue Organ Cult. 1998;54:21-28.

J’Aiti F., Benhabib O., Sharma H.C., El Jaafari S., El Hadrami I. Geneotypic variation in anther culture and effect of ovary coculture in durum wheat. Plant Cell Tissue Organ Cult. 1999;59:71-76.

Kiani R., Arzani A., Meiby S.A.M.M., Rahimmalek M., Razavi K. Morpho-physiological and gene expression responses of wheat by Aegilops cylindrica amphidiploids to salt stress. bioRxiv. 2021. DOI 10.1101/2020.06.07.139220.

Klimushina M.V., Kroupin P.Y., Bazhenov М.S., Karlov G.I., Divashek M.G. Waxy gene-orthologs in wheat × Thinopyrum amphidiploids. Agronomy. 2020;10(7):963. DOI 10.3390/agronomy10070963.

Lantos C. In vitro antherogenesis in wheat (Triticum aestivum L.). Triticale (×Triticosecale Wittmack), spice pepper (Capsicum annuum L.) and integration of the results into breeding. Thesis of the Ph.D. dissertation. Szent Istvan University, 2009.

Lantos C., Bona L., Boda K., Pauk J. Comparative analysis of in vitro anther- and isolated microspore culture in hexaploid Triticale (×Triticosecale Wittmack) for androgenic parameters. Euphytica. 2014; 197(1):27-37.

Lesniewska A., Ponikta A., Slusarkiewicz-Januzcza A., Zwierzyskowska E., Zwierzyskowzki J., James A.R., Thomas H., Humphreys M.W. Androgenesis from Festuca pratensis × Lolium multiflorum amphidiploid cultivars in order to select and stabilize rare gene combinations for grass breeding. Heredity. 2001;86:167-176.

Li C. Breeding crops by design for future agriculture. J. Zhejiang Univ. Sci. B. 2020;21(6):423-425. DOI 10.1631/jzus.B2010001.

Li H., Chen X., Xin Z.Y., Ma Y.Z., Xu H.J., Chen Y.X., Jia X. Development and identification of wheat–Haynaldia villosa T6D.L.6VS chromosome translocation lines conferring resistance to powdery mildew. Plant Breed. 2005;124:203-205. DOI 10.1111/j.1439-0523.2004.01062.x.

Li H.J., Li Y.W., Zhang Y.M., Li H., Guo B.H., Wang Z.N., Wen Z.Y., Liu Z.Y., Zhu Z.Q., Jia X. Tissue culture induced translocation conferring powdery mildew resistance between wheat and Dasyypyrum villosum and its marker-assisted selection. Yi Chuan Xue Bao. 2000;22(7):608-613.

Liu D., Zhang H., Zhang L., Yuan Z., Mao H., Zheng Y. Distinct hybridization: an interspecific for target manipulation of chromosomes. In: Prapat A., Kumar J. (Eds.). Alien Gene Transfer in Crop Plants. Vol. 1. New York: Springer, 2014. DOI 10.1007/978-1-4614-8585-8. 2. Marciniak K., Kazcmarkz Z., Adamski T., Skumprev S., Surnma M. The anther-culture response of triticale line × tester progenies. Cell. Mol. Biol. Lett. 2003;8:343-351.

Ming D., Xin Liu, Fu Q., Zhao Q., Zhao F., Wang Y. Cytogenetical analysis of hybrid F1 from common wheat and Aegilops ventricosa × Aegilops cylindrica amphidiploid. Agric. Sci. Technol. Hunan. 2011;12(9):1298-1302.

Nemeth C., Yang C.-y., Kasprzak P., Hubbert S., Scholefield D., Mehra S., Skipper E., King L., King J. Generation of amphidiploids from hybrids of wheat and related species from the genera Aegilops, Secale, Thinopyrum, and Triticum as a source of genetic variation for wheat improvement. Genome. 2015;58(2):71-79. DOI 10.1139/gen-2015-0002.

Okada A., Arndell U., Bishoj S., Rama S., Watson-Haigh N.S., Tucker E.J., Baumann U., Langridge P., Whitford R. CRISPR/Cas9-mediated knockout of Ms1 enables the rapid generation of male-sterile hexaploid wheat lines for use in hybrid seed production. Plant Biotechnol. J. 2019;17:1905-1913. DOI 10.1111/pbi.13106.

Pauk J., Poulmalet M., Toth K.L., Monostori T. In vitro androgenesis of triticale in isolated microspore culture. Plant Cell Tissue Organ Cult. 2000;61(3):221-229. DOI 10.1023/A:1006416116366.
Андрогенетическая реакция амфидиплоидов Triticum durum-Dasypyrum villosum и их родительских форм
Х. Стоянов
И. Белчев
2022
26 • 2

Conflict of interest. The authors declare no conflict of interest.