A Missense Mutation in CD38 Associated with Autism Spectrum Disorder in Three Pedigrees

Haruhiro Higashida1,2, Toshio Munesue2, Shigeru Yokoyama1, Minako Hashii1, Keita Koizumi2 and Akihiro Matsushima3
1Department of Biophysical Genetics, Kanazawa University Graduate School of Medicine, Kanazawa, 2Research Center for Child Mental Development, Kanazawa University, Kanazawa, 3Nanao National Hospital, Nanao, Japan

1. Introduction

Autism spectrum disorder (ASD) or pervasive developmental disorder (PDD) is a neurodevelopmental disease, beginning in childhood but extending through to adulthood. ASD is characterised by impairments in reciprocal social interaction and communication, and by restricted or stereotyped patterns of interests and activities. This disorder has received much scientific and social attention1,2. ASD is more common than previously supposed with a frequency of 0.6-3 out of 100 births2-6 and occurs either sporadically or in a familial pattern, and far more commonly in males7-9. The etiology remains largely unknown10. Previously we demonstrated that CD38 acts as a ‘niceness’ protein for mouse social behavior, by regulating release of oxytocin (OT)11, which seems to be essential for mutual recognition and trust12,13. Therefore, here, we describe our results on single nucleotide polymorphisms (SNPs) of CD38 in ASD patients and control subjects14. In addition, we report our experience of treatment of one ASD patient with a CD38 SNP by nasal OT administration.

2. Results

Figures 1 and 2 show human CD38 expression in the frontal cortex, cerebellum, hypothalamus and amygdala, by RT-PCR with human brain RNA samples which were used for synthesizing cDNAs. CD38 mRNA was highly expressed in the hypothalamus in the human brain, suggesting that CD38 has an important role on human social behavior, as in the mouse11.

Armed with this new information about CD38 in the human brain, we set out to examine the human CD38 gene. The mRNA for CD38 is transcribed from human chromosome 4p15,16. The CD38 gene consists of 8 exons, spanning a genomic stretch of 70.51 kb (mRNA: 1227 bases) (http://www.broad.mit.edu/mpg/haploview; Figure 3a). SNP screening in 8 exons and their flanking introns was performed by direct sequencing in 29 unrelated
Fig. 1. Semi-quantitative RT-PCR confesses human CD38 expression in the frontal cortex (F. Cort), the cerebellum (Cbl), the hypothalamus (Hyp) and the amygdala (Amyg). Human brain RNA samples provided commercially (Ambion) were used for synthesizing cDNAs. Relative intensity was calculated by comparing with β-actin expression as a control.
A Missense Mutation in CD38 Associated with Autism Spectrum Disorder in Three Pedigrees

Fig. 2. mRNA expression levels of CD38 gene in various human brain tissues. (a) The relative expression level of human CD38 gene was determined in RNA samples from frontal cortex, cerebellum, hypothalamus and amygdala using real-time quantitative PCR. Housekeeping normalized units (threshold cycle) for each gene obtained in the PCR analysis were used to determine the fold-change among samples. Bars represent fold-changes of the mRNA level of CD38 when comparing cerebellum with other tissues. Data are expressed as the mean ± S.D., performed in duplicate and repeated 3 times. *p<0.01, **p<0.01, Significantly increased from cerebellum value (P<0.01 and P<0.001). (b) Each plot represents the baseline-subtracted fluorescence intensity (ΔRn) that reflects mRNA levels of CD38 or β-actin genes. Horizontal lines indicate threshold lines set in the exponentially increasing area calculated by using SDS software.

Subjects (the sample set A in Table 1) fulfilling Diagnostic and Statistical Manual of Mental Disorders, 4th edition (DSM-IV, American Psychiatric Association, 1994), criteria for ASD, and in 201 non-clinical control subjects (sample set E), in the Kanazawa area in Japan. We detected 12 SNPs that had already been reported, plus 3 novel mutations (Figure 3a). Allelic and genotypic frequencies in these samples are summarized in Tables 2 and 3. Among them, as shown in Figure 3b, we detected the C4693T mutation in exon 3 (SNP13; rs1800561) that leads to an arginine (R)-to-tryptophan (W) substitution at amino acid 140, R140W.

In the following experiments, we focused mainly on this mutation, because of functional abnormality in R140W-substituted-CD38: (1) The R140 is relatively well conserved among multiple species except for the rodent (Figure 3c). R140 is located in the flexible loop (137-141) at the midpoint of the N- and C-terminus domains between two helical domains (αa4
Fig. 3. Genome and molecular structures and mutations of CD38.  
(a) Genomic structure of CD38 and locations of SNPs in introns (upper) and in exons (lower). Exons are indicated by boxes, with translated regions in closed boxes, and untranslated regions in open boxes. Mutations at amino acids at positions of 47, 116, 140, 168 and 264 of CD38 are indicated. Numbering of the nucleotides starts at the A of ATG encoding the initial methionine and refers to Genbank accession number D84284.  
(b) Sequence trace was derived from a blood DNA sample of 4693C/T heterozygote.  
(c) Conservation of R at the 140th amino acid among different species, except for the rodent. Sequences were obtained through the accession numbers of NM001775, AY555148, NM175798, AF117714, AF272974, NM013127, NM007646, D30048, and M85206/M37644 for the indicated species, respectively. It is noted that mutant structure has more open-form conformation than wild-type and its variation is slightly larger.
and αα5) and is the pivot of the hinge region connecting two regions of L-shaped molecule\(^\text{17}\). Therefore, the mutation (W140) causes severe perturbations of the predicted protein structure, if compared with the human (R140), rabbit (K140) or mouse (G140) CD38 (see Figure 7 in ref.\(^\text{14}\)). (2) Indeed, the mutant W140-CD38 showed one third of ADP-ribosyl cyclase activity of wild-type CD38 expressed in the CHO cells\(^\text{15}\). (3) Social amnesia was not rescued by local expression of W140-CD38 in the hypothalamus in \(\text{Cd38}\) knockout mice\(^\text{11}\).

Table 1. Sample sets in this experiment

| Sample set | number | Male/Female | Age range | Description | Country | W140 allele frequency |
|------------|--------|-------------|-----------|-------------|---------|-----------------------|
| A          | 29     | (23/6)      | 12 to 44  | Unrelated ASD | Japan* | 0.052                 |
| B          | 3      | (3/0)       | 21 to 44  | 3 probands in A | Japan* | 1                     |
| C          | 25     | (15/10)     | 21 to 84  | 3 families in B | Japan* | 0.32                  |
| D          | 252    | (252/0)     | 22 to 64  | ASD          | USA** | 0 20, 21              |
| E          | 201    | (106/95)    | 22 to 64  | Unscreened control | Japan* | 0.007                 |

*In the Kanazawa area  **AGRE samples

Table 2. Allelic frequency of SNPs in control and ASD subjects

| SNP* | Control | N | Allele counts | Frequency | ASD | N | Allele counts | Frequency |
|------|---------|---|---------------|-----------|-----|---|---------------|-----------|
| SNP01 | rs3796878 G>A | 400 | 4 | 0.01 | 58 | 0 | 0 |
| SNP02 | rs3796875 A>G | 398 | 100 | 0.25 | 56 | 18 | 0.321 |
| SNP03 | rs6449197 C>T | 392 | 81 | 0.207 | 58 | 16 | 0.275 |
| SNP04 | rs11574927 A>G | 402 | 78 | 0.195 | 58 | 6 | 0.103 |
| SNP05 | rs10805347 A>G | 400 | 180 | 0.452 | 58 | 22 | 0.379 |
| SNP06 | rs3796863 C>A | 398 | 148 | 0.371 | 58 | 20 | 0.345 |
| SNP07 | rs1130169 C>T | 398 | 36 | 0.341 | 56 | 15 | 0.268 |
| SNP08 | rs13137313 A>G | 398 | 176 | 0.442 | 58 | 28 | 0.483 |
| SNP09 | rs17476066 T>C | 392 | 46 | 0.117 | 58 | 6 | 0.103 |
| SNP10 | rs3733593 C>T | 398 | 128 | 0.32 | 56 | 17 | 0.305 |
| SNP11 | C>G | 402 | 1 | 0.002 | 58 | 0 | 0 |
| SNP12 | C>T | 402 | 0 | 0 | 58 | 1 | 0.017 |
| SNP13 | rs1800561 C>T | 402 | 3 | 0.008 | 58 | 3 | 0.052 |
| SNP14 | rs1800051 A>C | 402 | 59 | 0.146 | 58 | 13 | 0.22 |
| SNP15 | C>T | 402 | 6 | 0.015 | 58 | 0 | 0 |

*SNPs are denoted as major allele>minor allele
Fig. 4. Pedigrees for three families of ASD probands carrying the W140. Affected males (proband) are indicated with red arrows. Affected or carrier males or females are indicated by filled squares or circles, respectively. Empty symbols denote individuals with no mutation. Gray symbols indicate undetermined with no DNA available for analysis. Sexes are hidden by diamond symbols upon request. The subjects are indicated by progressive Arabic numbers according to the three generations. The current study was approved by the ethical committee of Kanazawa University Graduate School of Medicine. Subjects marked with symbols are: **, denotes ASD; *, ASD traits; P, PDD-NOS; A, Asperger disorder.
Fig. 5. Expression of the minor allele of rs1800561
(A) cDNA with 4693C has MspAI site. (B) RT-PCR products from blood RNA samples were digested by MspAI. An RT-PCT product from a homozygous 4693C/C subject gives two bands, while a heterozygous 4693C/T subject giving three bands. (C) Sequencing of RT-PCR products confirms the SNP.
Family #3 (3-I-3), in an apparently autosomal dominant fashion. We found a total of 18 carriers in 28 family members cooperative (prevalence=64%). In all cases the mutation was heterozygous (allelic frequency=0.32). The mutant allele was indeed transcribed in the subjects tested (Figure 5).

The kindred were clinically evaluated by interviewing. The probands’ young (1-III-2 in the Family #1) and old (3-III-1 in the family #3) brothers showed clinical features conforming to PDD-NOS (PDD not otherwise specified) or Asperger. Two fathers (1-II-2 and 3-II-1) in their 50s and another father (2-I-1) in his 70s were all diagnosed as having with ASD traits. Most other adults over 50 years old in these pedigrees had not been clinically diagnosed with ASD or other psychiatric diseases, though some showed personal traits such as eccentricity, resulting in 8 ASD subjects out of 13 male carriers (62%). Interestingly, four young female cousins with (1-III-3 and 1-III-4 in the family #1) and without (3-III-3 and 3-III-4 in the family #3) the mutation, had no clinical ASD phenotype.

We also evaluated them from the score of the Autism-Spectrum Quotient (AQ)18,19, in which older subjects esteemed themself by recalling behaviours at their life period of 20s. AQ scores in two young male carriers in the family #1 (1-III-1 and -2) fulfilled the criteria (cut-off point of 28) of ASD, though this score was not obtained from two other ASD probands (in the families #2 and #3), because of low intelligence (Figure 6). Some carriers’ scores were above the standard deviation of average values in noncarrier family members who showed normal control scores, indicating that such carriers may be considered to manifest ASD traits, even though not affected at the clinical level (Figure 6a). Statistically there is no difference between three different age groups (young, middle and old generations), but the males’ score was significantly higher than that of females (p<0.05; Figure 6b). These clinical and self-describing evaluations suggest that this gene polymorphism is important to determine the ASD or ASD trait phenotype.

Given these results, we obtained serum samples from the kindreds to further study the connection between the human CD38 mutation and plasma OT or arginine vasopressin (AVP) levels, since we previously showed that a null mutation of Cd38 resulted in the selective decrease of plasma OT levels in mice11, and low levels of OT have been reported in autistic children20. The plasma OT levels in the carriers (161.3 ± 26.5 pg/ml, n=12) were lower than those of kindred non-carriers (345.8 ± 61.3 pg/ml, n=10; p<0.01), as shown in Figure 7. The differences seem to be found in the younger generation but not so in older subjects (Figure 7c). The OT levels of three probands and two young carriers were compared with ASD patients without the W140 mutation (in the sample set A): the levels of five W140 carriers (79.2 ± 16.6 pg/ml; n=5) were lower than those without the mutation (147.7 ± 15.0 pg/ml; p<0.01, n=26).

Furthermore, the OT level of the carrier ASD probands was significantly lower when compared with that in 101 adult control (198.2 ± 24.7 pg/ml; p<0.01). As expected, there is no difference in AVP levels between CD38-mutation-carriers and –noncarriers in the pedigrees (Figure 7b and d). Low plasma hormone levels were frequently observed in subjects with high AQ scores in carriers in the pedigrees (Figure 7e and f).

We also analysed the R140W mutation in 252 ASD subjects (excluding Hispanic and Asian peoples) recruited to the Autism Genetic Resource Exchange (AGRE; http://www.agre.org)21 in USA (sample set D22). No mutation was found, suggesting ethnicity-dependent frequency differences. Finally, from 201 healthy unscreened control subjects (sample set E), 2 females and 1 male were positive for the mutation, representing allelic frequency of 0.007 (Tables 2 and 3). This frequency is 7.4-fold lower than those (0.052) in ASD patient group in the same residential area (p<0.028; Table 3).
Fig. 6. **AQ score in family members in the three pedigrees.** (a) Assessment groups were: W140: n=14 family members with the monoallelic R140W mutation; R140: n=7 persons without the mutation in the families. Horizontal dashed bar indicates the critical score of 28 obtained from clinical ASD group in a separate experiment\(^\text{18}\). The mean and standard deviation range of AQ scores were illustrated. Note that 5 individuals show the intermediate score above the control range but below the ASD score. The score with R140W is higher than that without (one-way ANOVA, \(p<0.05\)). (b) A plot of AQ scores of each individual of family members with or without the R140W mutation according to age. No significance was found between three generations (20<age<40, 40<age<60 and 60<age) by two-way ANOVA. The scores of males are significantly higher than those of females (\(p<0.05\)). Circles indicate female, and squares, male.
Fig. 7. **Plasma oxytocin and vasopressin levels in family members.** Plasma concentrations of OT (a) and AVP (b) levels in family members with (W140; n=12, red or orange bar) or without (R140; n=10, green or blue bar) heterozygous R140W allele. Mean ± s.e.m. **, p<0.01 (one-way ANOVA). OT (c) or AVP (d) levels in the three kindred according to age. OT (e) or AVP (f) levels as a function of AQ scores. Red and orange symbols or green and blue symbols indicate levels from persons with or without the R140W mutation, respectively. Circles denote female and diamonds or squares, male.
A Missense Mutation in CD38 Associated with Autism Spectrum Disorder in Three Pedigrees

Table 3. Statistical analysis of SNPs in 29 ASD and 201 control subjects in the Kanazawa area

One proband receiving intranasal OT for 6 months showed improvements in the area of social approach, eye-contact and communication behavior without serious adverse effects. These results suggest that the CD38 W140 allele could be a possible risk factor for one form of ASD by abrogating OT function and the carriers become candidates for the OT treatment14.

3. Discussion

ASD is heterogeneous and forms a continuum, and thus is likely to involve many genes7-10,23-26. De novo mutations related to ASD are rarely inherited7, but some ASD also can be inherited from pre-existing genetic variants in parents, for both of which a unified mechanism was proposed9. Our results shed light on genetic mutations of CD38 in relatively mature ASD patients and the inheritable ASD subgroups of three independent families in which the CD38-R140W mutation showed relative co-segregation with diverse phenotypes of ASD. In such multiplex families, the risk of autism or ASD traits in young male offsprings is nearly ~100%, representing dominant transmission of a mutation with high penetrance. However, we cannot completely exclude as yet unidentified contributing factors.

The heterozygous 4693C/T carriers were identified in Japanese and Italian general populations16,27. The presence of this mutation in Japanese has already been reported in the HapMap site (http://www.hapmap.org/) and by Yagui et al. in 199815. Surprisingly, the carriers were also identified in females. Therefore, it is highly possible that even clinically-affected female carriers with this mutant allele could in turn transmit the mutation to their offspring with high penetrance in males. Currently, we are intensively collecting blood samples for mutation analysis from many ASD and control subjects in different cities and countries to support our finding, which so far has been carried out in a relatively small group in a restricted area.

www.intechopen.com
A noteworthy role for OT in social recognition has been shown in rodent and human studies\textsuperscript{12,13}. Recently, a 1.1-Mb deletion of 20p13 including the OT gene (copy number decrease) has been detected in a child with Asperger disorder\textsuperscript{8}. An association of one or two intronic SNPs in the OT receptor gene with autism has also been reported\textsuperscript{25,28,29}, suggesting that defects in OT signaling confer genetic vulnerability to ASD. Though the R140W mutation was not found in 252 American AGRE samples, the association study with tagSNPs showed one SNP (SNP06; rs3796863) that is positively related ($p<0.004$) with American high functioning autism\textsuperscript{14}.

In conclusion, \textit{CD38} mutations provide one genetic basis for those instances of ASD that arises from disruption of the OT signaling. Thus, our finding provides the first theoretical background for the evidence-based treatment by OT infusion for a subgroup of ASD patients with the lower plasma OT level\textsuperscript{14}, which has already been tried for non-selected ASD subjects\textsuperscript{30,31}.

4. Methods summary

Clinical and genetic studies were carried out according to institutional guidelines after ethical approval of participating institutions and informed consent was obtained from all participating patients. A total of 29 unrelated affected individuals out of 96 in the Kanazawa area in Japan (Sample set A) were admitted the Kanazawa University Hospital diagnosed with DSM-IV in accordance with clinical criteria. Blood and platelet biochemical analyses were performed in 29 ASD probands and their parents and family members who agreed to supply. Genotyping for the association study and mutation screening were performed by direct sequencing or TaqMan technology. PCR products were sequenced with the BigDye Terminator Cycle Sequencing Kit (V3.1, Applied Biosystems, Foster City, CA, USA). Samples were then subjected to electrophoresis, using an ABI PRISM genetic analyzer (Applied Biosystems). Absence of genotyping errors was controlled by sequencing the PCR product with the opposite primer in a subset of patients.

**Statistics**: Data are expressed as mean ± s.d. or s.e.m. Statistical analysis was performed using one-way or two-way ANOVA. The criterion for significance in all cases was $p<0.05$.

5. References

[1] Singh J, Hallmayer J & Illes J. Interacting and paradoxical forces in neuroscience and society. \textit{Nature Rev. Neurosci.} 8, 153-60 (2007).

[2] Editorial. New guidance on autism. \textit{Lancet} 370, 1590 (2007).

[3] Williams J G, Higgins J P & Brayne C E. Systematic review of prevalence studies of autism spectrum disorders. \textit{Arch. Dis. Child.} 91, 8-15 (2006).

[4] Baird, G. \textit{et al}. Prevalence of disorders of the autism spectrum in a population cohort of children in South Thames: the Special Needs and Autism Project (SNAP). \textit{Lancet} 368, 210-215 (2006).

[5] Honda H, Shimizu Y, Imai M & Nitto Y. Cumulative incidence of childhood autism: a total population study of better accuracy and precision. \textit{Dev. Med. Child. Neurol.} 47, 10-18 (2005).

[6] Sumi S, Tanai H, Miyachi T & Tanemura M. Sibling risk of pervasive developmental disorder estimated by means of an epidemiologic survey in Nagoya. \textit{J. Hum. Genet.} 51, 518-522 (2006).
A Missense Mutation in CD38 Associated with Autism Spectrum Disorder in Three Pedigrees

[7] Beaudet AI. Autism: highly heritable but not inherited. *Nature Med.* 13, 534-536 (2007).
[8] Sebat J, Lakshmi B, Malhotra D, Troge J, Lese-Martin C, Walsh T, Yamrom B, Yoon S, Krasnitz A, Kendall J, Leotta A, Pai D, Zhang R, Lee YH, Hicks J, Spence SJ, Lee AT, Puura K, Lehtimäki T, Ledbetter D, Gregersen PK, Bregman J, Sutcliffe JS, Jobanputra V, Chung W, Warburton D, King MC, Skuse D, Geschwind DH, Gilliam TC, Ye K & Wigler M. Strong association of de novo copy number mutations with autism. *Science* 316, 445-449 (2007).
[9] Zhao X, Leotta A, Kustanovich V, LaJonchere C, Geschwind DH, Law K, Law P, Qiu S, Lord C, Sebat J, Ye K & Wigler M. A unified genetic theory for sporadic and inherited autism. *Proc. Natl. Acad. Sci. USA* 104, 12831-12836 (2007).
[10] Happé F, Ronald A & Plomin R. Time to give up on a single explanation for autism. *Nature Neurosci.* 9, 1218-1220 (2006).
[11] Jin D, Liu HX, Hirai H, Torashima T, Nagai T, Lopatina O, Shnayder NA, Yamada K, Noda M, Seike T, Fujita K, Takasawa S, Yokoyama S, Koizumi K, Shiraishi Y, Tanaka S, Hashii M, Yoshihara T, Higashida K, Islam MS, Yamada N, Hayashi K, Naguchi N, Kato I, Okamoto H, Matsushima A, Salmina A, Munesue T, Shimizu N, Mochida S, Asano M & Higashida H. CD38 is critical for social behaviour by regulating oxytocin secretion. *Nature* 446, 41-45 (2007).
[12] Insel TR. The challenge of translation in social neuroscience: a review of oxytocin, vasopressin, and affiliative behavior. *Neuron* 65, 768-779 (2010).
[13] Kosfeld M, Heinrichs M, Zak P J, Fischbacher U & Fehr E. Oxytocin increases trust in humans. *Nature* 435, 673-676 (2005).
[14] Munesue T, Yokoyama S, Nakamura K, Anitha A, Yamada K, Hayashi K, Asaka T, Liu HX, Jin D, Koizumi K, Islam MS, Huang JJ, Ma WJ, Kim UH, Kim SJ, Park K, Kim D, Kikuchi M, Ono Y, Nakatani H, Suda S, Miyachi T, Hirai H, Salmina A, Pichugina YA, Soumarokov AA, Takei N, Mori N, Tsuji M, Sugiyama T, Yagi K, Yamagishi M, Sasaki T, Yamasue H, Kato N, Hashimoto R, Taniike M, Hayashi Y, Hamada J, Suzuki S, Ooi A, Noda M, Kamiyama Y, Kido MA, Lopatina O, Hashii M, Amina S, Malavasi F, Huang EJ, Zhang J, Shimizu N, Yoshikawa T, Matsushima A, Minabe Y & Higashida H. Neurosci. Res. 67, 181-91 (2010).
[15] Yagui K, Shimada F, Mimura M, Hashimoto N, Suzuki Y, Tokuyama Y, Nata K, Tohgo A, Ikehata F, Takasawa S, Okamoto H, Makino H, Saito Y & Kanatsuka A. A missense mutation in the CD38 gene, a novel factor for insulin secretion: association with Type II diabetes mellitus in Japanese subjects and evidence of abnormal function when expressed in vitro. *Diabetologia* 41, 1024-1028 (1998).
[16] Malavasi F, Deaglio S, Funaro A, Ferrero E, Horenstein AL, Ortolan E, Vaisitti T & Aydin S. Evolution and function of the ADP ribosyl cyclase/CD38 gene family in physiology and pathology. *Physiol. Rev.* 88, 841-886 (2008).
[17] Liu Q, Kriksunov IA, Graeff R, Lee H C & Hao Q. Structural basis for formation and hydrolysis of the calcium messenger cyclic ADP-ribose by human CD38. *J. Biol. Chem.* 282, 5853-5861 (2007).
[18] Baron-Cohen S, Hoekstra RH, Knickmeyer R & Wheelwright S. The Autism-Spectrum Quotient (AQ)-adolescent version. *J. Autism Dev. Disord.* 36, 343-350 (2006).
[19] Munesue T, Ono Y, Mutoh K, Shimoda K, Nakatani H & Koshino Y. Autonomic and motor responses to Utena’s simple psychiatric function test in autism spectrum disorder (in Japanese). *Seishin Igaku* 49, 599-606 (2007).
[20] Modahl C, Green L, Fein D, Morris M, Waterhouse L, Feinstein C & Levin H. Plasma oxytocin levels in autistic children. *Biol. Psychiatry* 43, 270-277 (1998).

[21] Geschwind DH, Sowinski J, Lord C, Iversen P, Shestack J, Jones P, Ducat L, Spence SJ; AGRE Steering Committee. The autism genetic resource exchange: a resource for the study of autism and related neuropsychiatric conditions. *Am. J. Hum. Genet.* 69, 463-466 (2001).

[22] Toyoda T, Nakamura K, Yamada K, Thanseem I, Anitha A, Suda S, Tsujii M, Iwayama Y, Hattori E, Toyota T, Miyachi T, Iwata Y, Suzuki K, Matsuzaki H, Kawai M, Sekine Y, Tsuchiya K, Sugihara G, Ouchi Y, Sugiyama T, Takei N, Yoshikawa T & Mori N. SNP analyses of growth factor genes EGF, TGFbeta-1, and HGF reveal haplotypic association of EGF with autism. *Biochem. Biophys. Res. Commun.* 360, 715-720 (2007).

[23] Pinto D et al. Functional impact of global rare copy number variation in autism spectrum disorders. *Nature* 466, 368-372 (2010).

[24] Szatmari, P. et al. Mapping autism risk loci using genetic linkage and chromosomal rearrangements. *Nature Genet.* 39, 319-328 (2007).

[25] Weiss LA, Arking DE. Gene Discovery Project of Johns Hopkins & the Autism Consortium, Daly MJ, Chakravarti A. A genome-wide linkage and association scan reveals novel loci for autism. *Nature* 461, 802-808 (2009).

[26] Skuse, GH. Rethinking the nature of genetic vulnerability to autistic spectrum disorders. *Trends Genet.* 23, 387-395 (2007).

[27] Mallone R, Ortolan E, Baj G, Funaro A, Giunti S, Lillaz E, Saccucci F, Cassader M, Cavallo-Perin P & Malavasi F. Autoantibody response to CD38 in Caucasian patients with type 1 and type 2 diabetes: immunological and genetic characterization. *Diabetes* 50, 752-762 (2001).

[28] Wu S, Jia M, Ruan Y, Liu J, Guo Y, Shuang M, Gong X, Zhang Y, Yang X & Zhang D. Positive association of the oxytocin receptor gene (OXTR) with autism in the Chinese Han population. *Biol. Psychiatry* 58, 74-77 (2005).

[29] Wermter AK, Kamp-Becker I, Hesse P, Schlute-Körne G, Strauch K & Remschmidt H. Evidence for the involvement of genetic variation in the oxytocin receptor gene (OXTR) in the etiology of autistic disorders on high-functioning level. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 153B, 629-39 (2010).

[30] Hollander E, Novotny S, Hanratty M, Yaffe R, deCaria C & Aronowitz B & Mosovich S. Oxytocin infusion reduces repetitive behaviors in adults with autistic and Asperger's disorders. *Neuropsychopharmacol.* 28, 193-198 (2003).

[31] Hollander E, Bartz J, Chaplin W, Phillips A, Sumner J, Soorya L, Anagnostou E & Wasserman S. Oxytocin increases retention of social cognition in autism. *Biol. Psychiatry* 61, 498-503 (2007).
The book covers some of the key research developments in autism and brings together the current state of evidence on the neurobiologic understanding of this intriguing disorder. The pathogenetic mechanisms are explored by contributors from diverse perspectives including genetics, neuroimaging, neuroanatomy, neurophysiology, neurochemistry, neuroimmunology, neuroendocrinology, functional organization of the brain and clinical applications from the role of diet to vaccines. It is hoped that understanding these interconnected neurobiological systems, the programming of which is genetically modulated during neurodevelopment and mediated through a range of neuropeptides and interacting neurotransmitter systems, would no doubt assist in developing interventions that accommodate the way the brains of individuals with autism function. In keeping with the multimodal and diverse origins of the disorder, a wide range of topics is covered and these include genetic underpinnings and environmental modulation leading to epigenetic changes in the aetiology; neural substrates, potential biomarkers and endophenotypes that underlie clinical characteristics; as well as neurochemical pathways and pathophysiological mechanisms that pave the way for therapeutic interventions.

How to reference
In order to correctly reference this scholarly work, feel free to copy and paste the following:

Haruhiro Higashida, Toshio Munesue, Shigeru Yokoyama, Minako Hashii, Keita Koizumi and Akihiro Matsushima (2011). A Missense Mutation in CD38 Associated with Autism Spectrum Disorder in Three Pedigrees, Autism - A Neurodevelopmental Journey from Genes to Behaviour, Dr. Valsamma Eapen (Ed.), ISBN: 978-953-307-493-1, InTech, Available from: http://www.intechopen.com/books/autism-a-neurodevelopmental-journey-from-genes-to-behaviour/a-missense-mutation-in-cd38-associated-with-autism-spectrum-disorder-in-three-pedigrees