Overexpressed p-S6 associates with lymph node metastasis and predicts poor prognosis in non-small cell lung cancer

Yaoxiang Tang1, Jiadi Luo1, Ying Zhou1, Hongjing Zang1, Yang Yang1, Sile Liu1, Hongmei Zheng1, Jian Ma2, Songqing Fan1 and Qiuyuan Wen1*

Abstract

Background: Ribosomal protein S6 (S6), a downstream effect media of the AKT/mTOR pathway, not only is a part of 40S small subunit of eukaryotic ribosome, but also involves in protein synthesis and cell proliferation during cancer development.

Methods: In present study, we explore the association between phosphorylated S6 (p-S6) protein expression and clinicopathological features as well as prognostic implications in NSCLC. P-S6 was detected in tissue microarrays (TMAs) containing 350 NSCLC, 53 non-cancerous lung tissues (Non-CLT), and 88 cases of matched metastatic lymph node lesions via immunohistochemistry (IHC). Transwell assays and wound healing assay were used to assess the effects of p-S6 inhibition on NSCLC cell metastasis.

Results: The p-S6 expression in NSCLC was more evident than that in Non-CLT (p < 0.05). Compared to NSCLC patients who have no lymph node metastasis (LNM), those with LNM had higher p-S6 expression (p = 0.001). Regardless of lung squamous cell carcinoma (SCC) or adenocarcinoma (ADC), p-S6 was increased obviously in metastatic lymph nodes compared with matched primary cancers (p = 0.001, p = 0.022, respectively). Inhibition of p-S6 decreased the metastasis ability of NSCLC cells. In addition, p-S6 was an independent predicted marker for LNM in patients with NSCLC (p < 0.001). According to survival analysis, patients with highly expressed p-S6 had a lower survival rate compared with that with lower expression (p = 0.013). P-S6 is an unfavorable independent prognostic factor for NSCLC patients (p = 0.011).

Conclusion: Increased expression of p-S6 is not only a novel predictive biomarker of LNM but also poor prognosis in NSCLC.

Keywords: P-S6, Lymph node metastasis, Prognosis, Non-small cell lung cancer
of the NSCLC patients are diagnosed in advanced clinical stages, often accompanied by LNM and distant metastasis, which are important reasons for the poor prognosis [3, 4]. Therefore, it is an urgent need to find new biomarker for early detection of LNM of NSCLC and predict the prognosis.

Recently, the protein kinase B (AKT)/mammalian target of rapamycin (mTOR) signal pathway is over-activated in numerous tumors, including NSCLC, and associated with tumor angiogenesis, invasion, metastasis and so on [5–7]. Ribosomal protein S6 (S6), a part of 40S subunit of eukaryotic ribosome, is a famous downstream effect media of the AKT/mTOR pathway [8, 9]. As the first ribosomal protein proved to undergo inducible phosphorylation, S6 is mostly induced by activated 70kDaS6 Kinases (S6K) at five phosphorylation sites (Ser235, Ser236, Ser240, Ser244, Ser247), mainly Ser236 [9–12]. Phosphorylated S6 (p-S6) plays a crucial part in protein synthesis, cell size control and cell proliferation as increasing the affinity of the ribosome and mRNAs and improving the efficiency of protein translation [13]. In addition, p-S6 is also the key effector of mTOR in regulating cell size, whose decreased expression results in smaller cell size and reflects the growth defect [14, 15]. Overexpression of p-S6 is found in various solid tumors, such as gastric cancer, glioblastomas, and renal cell carcinomas (RCCs), and associated with poor prognosis [16–18]. S6 phosphorylation was even associated with malignant potential and glucose metabolism of intra-ducal papillary mucinous neoplasm of the pancreas [19]. In addition, the phosphorylation of S6 is considered to contribute to acquired resistance to MAPK pathway inhibitors in cancers, suggesting that p-S6 plays an essential role in the mechanism of anti-cancer drugs [20, 21].

However, the prognostic implications of p-S6 are poorly understood in NSCLC. In this present research, we have measured the level of p-S6 expression in tissue microarrays (TMAs) of non-cancerous lung tissues (Non-CLT) and NSCLC by immunohistochemistry (IHC) and decreased p-S6 expression in NSCLC cell lines with mTOR inhibitor (RAD001), to estimate not only the association between p-S6 and clinicopathological/prognostic features of NSCLC patients, but also the alteration of NSCLC cell migration and invasion ability.

**Material and Methods**

**Patients and tissue samples**

All tissue specimens which were collected by surgical resection acquired from the Thoracic Surgery Department of the Second Xiangya Hospital of Central South University (CSU) between 2003 and 2013. Approval of all protocols were obtained from the Institutional Human Experiment and Ethics Committee of the Second Xiangya Hospital of CSU (Changsha, China) (approval No. S039/2011), and approval of all protocols were obtained. 350 cases of NSCLC, 53 cases of Non-CLT, and 88 cases of matched metastatic lymph node lesions were included in this research. Patients in the study had definite histological diagnosis on the basis of 2015 WHO Classification of lung cancer [22], and were comprehensively staged as per the Eighth Edition Lung Cancer Stage Classification [23]. Complete clinicopathological data obtained from medical records were available for all patients (Supplementary Table).

**Cells lines and reagents**

Two human NSCLC cell lines, A549 and SPC-A1, were from the Cell Bank, Chinese Academy of Sciences (Shanghai, China) and those were grown in 37°C, 5% CO2 incubator. Both cell lines were culture in RPMI-1640 (BI, Israel) medium comprising 10% fetal bovine serum (BI, Israel). The mTOR inhibitor Everolimus (RAD001) was procured from Selleckchem, USA. Rabbit antibodies against p-S6 (S235/236) (#4857) were obtained from Cell Signaling Technology, USA. Mouse antibodies against S6 (cat no. 66886-1-Ig) and β-Actin (cat no. 66009-1-Ig) were purchased from proteintech, USA. Peroxidase-conjugated Affinipure Goat anti-Rabbit IgG (cat no. HG-SAR00002a) and anti-Mouse IgG (cat no. YG-SAM00001a) were purchased from HonorGene, China.

**Construction of the tissue microarrays**

As previously described, tissue microarrays (TMAs) were constructed using paraffin-embedded tissue of NSCLC and Non-CLT [24]. The perforation diameter of each sample was 0.6 mm. For TMAs of NSCLC, each case included two tumor cores, of which 88 cases each also contained two matched metastatic lymph node cores. For TMAs of Non-CLT, two cores per case was included. The average score of the two cores was regarded as the final score for each case.

**Immunohistochemistry and scores**

IHC for p-S6 in TMAs was carried out and the conditions of antibody staining were adjusted as described previously [25]. Briefly, primary antibody for p-S6 (1:800 of Ser235/236) was applied to detect its expression level. The positive control slide and negative control slide were included in each experiment. The IgG isotype-matched antibody was applied as negative contrast to confirm the antibody specificity.

Immunoreactivity was assessed semi-quantitatively by two independent pathologists. The staining intensity score was multiplied by percentage score of positive expressed tumor cells as the total score. Specifically,
staining intensity was scored as 0 to 3 (0 for negative; 1 for mild expression; 2 for moderate expression; 3 for strong expression). The percentage score was identified as 0 to 4 in view of positive cytoplasmic staining: 0 = 0%; 1 = 1-25%, 2 = 26-50%, 3 = 51-75%, 4 = 76-100%. The range of total score was from 0 to 7. Cutoff level which determined according to overall survival (OS) of NSCLC patients was 2. Expression of p-S6 in tumor cells of which total score is more than 2 was considered high expression and others were regarded as low expression. Agreement between the two assessors is 96%, borderline and ambiguous cases are resolved through discussion.

Western blotting
As previously described [5, 26], SPC-A1 and A549 cells were seeded and then treated with DMSO or 5 nmol/L RAD001 for 24 h, preparation of protein lysates and western blotting analysis were performed. The blots were cut prior to hybridisation with antibodies. The antibodies were used as follows: 1:1000 of Ser235/236 for anti-p-S6 and 1:5000 for anti-β-Actin.

Wound healing assay
1 × 10⁵ SPC-A1 and A549 cells were seeded and then treated with DMSO or 5 nmol/L RAD001 at about 60% confluence and cultured until 90%. Serum free RPMI-1640 was used to replace the culture medium and wounds were scratched with the tip of 10 μl pipette in each well. The size of wounds was observed using inverted microscope (Leica, Germany) and images were captured at 0 h, 24 h and 48 h. The data presented were repeated in triplicate. Wound healing percentage (%) = (1 – scratch area at t / scratch area at 0 h) × 100% (t means 24 h or 48 h).

Transwell assays
24-well chambers (Costar 3422; Corning, USA) were used to perform transwell migration assays. A549 and SPC-A1 cells were treated with DMSO or 5 nmol/L RAD001 for 24 h at 37°C. Then 5 × 10⁴ cells were extracted from each group and suspended in 200 μl of serum-free RPMI-1640 to the upper chamber, while 10% FBS RPMI-1640 was added to the lower room. The cells could migrate through the membrane for 12 h to 48 h. The membrane was fixed for 20 min in 4% paraformaldehyde solution and dyed with crystal violet for 20 min at room temperature. The count of cells under the cell membrane was calculated. The implementation of invasion assay protocol be analogous to that of migration assay, except that Matrigel was coated on the upper cavity and the incubation time was 1 h (356,234, Corning, USA).

Statistical analysis
The difference expression pattern of p-S6 between Non-CLT and NSCLC, as well as the association between p-S6 and clinicopathological features were analyzed by Chi-square (χ²) test. The difference between the metastatic lymph nodes and matched primary tumors was also analyzed by Chi-square (χ²) test. Multi-logistic regression method was performed to identify whether p-S6 is an independent marker for LNM of NSCLC. Survival rate curve was evaluated by Kaplan-Meier analysis, and comparisons were analyzed via log-rank test. Furthermore, cox proportional hazards model was performed for determining independent prognostic markers. The deviation between control and RAD001 groups was analyzed using one way t-test. All these statistical analyses were put into effect with SPSS Statistics software (version 24). Numerical data were presented as mean ± SEM. P < 0.05 (Two-sided) indicates that the result is statistically significant.

Results
P-S6 was significantly overexpressed in NSCLC
Firstly, we detected the intracellular localization and the level of p-S6 expression by IHC in a total of 403 samples, of which 350 were NSCLC. The results indicated that p-S6 was mainly observed in the cell cytoplasm of lung SCC, ADC and Non-CLT (Fig. 1). Quantification of p-S6 expression showed that its percentage of high expression was 30.5% (47/154) in lung SCC, 62.2% (122/196) in lung ADC and 15.1% (8/53) in Non-CLT, respectively. As shown in Supplementary Fig. 1, compared with Non-CLT, the level of p-S6 protein was obviously increased both in lung SCC and ADC (p = 0.031, p < 0.001, respectively).

Association between p-S6 and clinicopathological features
As data showed in Table 1, p-S6 in female was higher than that in male (p < 0.001). Patients with lung ADC had obviously higher p-S6 expression level than patients with SCC (p < 0.001). Overexpressed p-S6 was significantly correlated with LNM (p = 0.001). Moreover, patients with high p-S6 expression level suffered a lower overall survival (OS) rate than that with low expression (p = 0.005). However, there was not significantly association between p-S6 expression and other features, such as clinical stages, pathological grade and age (all p > 0.05).

Impact of p-S6 expression on LNM in NSCLC
Among the 350 NSCLC patients, 210 cases had LNM and 140 cases were free from LNM. NSCLC patients with LNM had obviously higher p-S6 expression compared with those without LNM (p = 0.001, Table 1). We also evaluated the level of p-S6 in primary NSCLC tissues and its matched LNM lesions. As shown in Fig. 2, regardless
of lung SCC or ADC, the percentage of high p-S6 expression is significantly higher in metastatic lymph node lesions with compared with the matched primary cancer \((p = 0.001, p = 0.022, \) respectively). To determine whether p-S6 expression was the independent predicted parameter for LNM in NSCLC, a multivariate logistic regression analysis was carried out. As mentioned in Table 2, highly expressed p-S6 \((p = 0.001), \) clinical stages \((p < 0.001), \) and age \((p = 0.010)\) were notably correlated with LNM status of NSCLC patients. These results indicated that overexpressed p-S6 is an independent factor for LNM of NSCLC regardless of other parameters.

### Inhibition of p-S6 reduced the cell migration and invasion of NSCLC

The alteration of cell migration and invasion ability of NSCLC were assessed after inhibition of p-S6 expression level in NSCLC cell lines (A549 and SPC-A1). Firstly, we evaluated the inhibition efficiency by Western blotting after the cell lines were treated with RAD001. According to our previous research [5, 26], NSCLC cells was treated with RAD001 at 5 nmol/L for 24 h. Figure 3 showed that p-S6 expression was significantly reduced after treatment with RAD001. Secondly, to detect the effect of p-S6 on the cell migration, wound healing and transwell migration experiments were conducted. Figure 4 showed that compared with the control group, the wound healing percentage of RAD001 group was decreased by 10.19 and 20.44% at 24 h and 48 h in A549 cells \((p = 0.018, p = 0.001, \) respectively), by 5.90 and 7.55% in SPC-A1 cells \((p = 0.028, p = 0.048, \) respectively), respectively. Figure 5A revealed that the inhibition of p-S6 expression in A549 and SPC-A1 cell lines decreased the migration ability by transwell migration assay (both \(p < 0.001\)). Moreover, the cell invasion assay demonstrated that p-S6 inhibition in NSCLC cell lines reduced the number of invaded cells (both \(p < 0.001\) (Fig. 5 B). Together, these results revealed that inhibition of p-S6 expression could reduce the cell migration and invasion of NSCLC.

### Impact of p-S6 expression on patients’ prognosis

In univariate survival analysis, survival rate was estimated by Kaplan-Meier method in 350 NSCLC patients,
Fig. 2 Comparison of p-S6 expression in metastatic NSCLC and matched primary cancer. A: P-S6 was strongly positive expressed in cell cytoplasm in primary lung SCC (A-a), primary lung ADC (A-b) and mild positive staining in its matched LNM lesions (A-c, A-d). B: Whether it is lung SCC or lung ADC, the expression level of p-S6 in metastatic cancer foci is significantly higher than that in primary cancer foci ($p = 0.001$, $p = 0.022$, respectively).

Table 2 Multivariate logistic analysis of LNM factors in NSCLC patients

| Variables              | B     | S.E. | Wald  | P-value | Exp(B) | 95.0% CI for Exp(B) |
|------------------------|-------|------|-------|---------|--------|---------------------|
| p-S6 expression        | 0.921 | 0.270| 11.644| 0.001*  | 2.512  | 1.480 - 4.262       |
| Clinical stages        | 1.957 | 0.264| 54.968| 0.000*  | 7.068  | 4.220 - 11.880      |
| Histological type      | -0.026| 0.276| 0.009 | 0.925   | 0.974  | 0.567 - 1.674       |
| Pathological grade     | -0.051| 0.256| 0.039 | 0.843   | 0.951  | 0.575 - 1.571       |
| Age                    | -0.759| 0.293| 6.688 | 0.010*  | 0.468  | 0.263 - 0.832       |
| Gender                 | 0.607 | 0.324| 3.516 | 0.061   | 1.835  | 0.973 - 3.462       |

Abbreviations: S.E Standard error, Exp(B) Odds ratio, CI Confidence interval, LNM Lymph node metastasis. * $P < 0.05$

Fig. 3 Effect of RAD001 on expression of p-S6 protein in NSCLC cell lines (A549 and SPC-A1). NSCLC Cells were treated with DMSO or RAD001 (5 nmol/L) for 24 h. In A549 and SPC-A1, compared with Control group, p-S6 protein in RAD001 groups were significantly decreased. Original blots are presented in Supplementary Fig. 2.
and comparison was performed via log-rank test. NSCLC patients with highly expressed p-S6 had a poor prognosis compared to those with lowly expressed p-S6 \((p = 0.013)\) (Fig. 6 A & Table 3). The OS rate of NSCLC patients with advanced clinical stage (stage III) was lower relative to those with early stages (stage I-II) \((p < 0.001)\) (Fig. 6 B & Table 3). Of note, patients without LNM have higher OS than those with LNM \((p < 0.001)\) (Fig. 6 C & Table 3). Meanwhile, compared with NSCLC patients with well and moderated pathological grade, lower OS could be seen in those with poor differentiation \((p = 0.003)\) (Fig. 6 D & Table 3).

To further investigate whether the overexpressed p-S6 was an independent prognostic index for NSCLC patients, Cox regression analysis was carried out. The results shown in Table 3 indicated that up-regulated p-S6 might be a poor prognostic marker for patients with NSCLC \((p = 0.011)\), as are clinical stage \((p < 0.001)\), LNM status \((p = 0.015)\) and pathological grade \((p = 0.013)\). Besides, age, gender, histological type had no significantly effect on the prognosis of patients with NSCLC.

**Discussion**

In this study, we demonstrated that the level of p-S6 protein expression in NSCLC was significantly higher than that in Non-CLT. The percentage of high p-S6 expression was obviously increased in patients with LNM than that without LNM. Inhibition of p-S6 expression reduced the invasion and migration ability of NSCLC cells. Our analysis indicated that NSCLC patients with increased p-S6 expression had a lower rate of survival than that with low expression ones. Taking together, our results imply that p-S6 may play a significant part in the progression of NSCLC and aberrant expression of p-S6 might be a novel prognostic marker for NSCLC. Increasing evidences have shown that mTOR/S6K/S6 pathway plays a crucial role in p53-mediated tumor inhibition \([27, 28]\). P-S6, the key downstream effector of mTOR/S6K pathway, is involved in the occurrence and development of many cancers \([9]\). Its expression is ascendant in numerous tumors, such as RCCs, pancreatic cancer, and esophageal squamous cell carcinoma \([18, 27, 29]\). The phosphorylation of S6 can attenuate the autophagy.
induced by damage-regulated autophagy modulator 1 (DRAM1) and p-S6 is a requirement for AKT-driven malignant transformation of pancreatic islet β cells [28, 30]. Due to the oncogenic role of p-S6, descending its expression could potentially provide a clue to find new idea for the targeted treatment cancers. For example, the suppression of p-S6 can block the further inhibition of the therapeutic mTOR inhibitor everolimus on protein synthesis and proliferation of RCCs cells [18]. Knockdown of S6K1 gene can obviously decrease the expression of cyclin D, leading to the decline of survival ability of esophageal cancer cells [29].

Activating invasion and metastasis is one of biological capabilities of cancers [31]. Our study showed that NSCLC patients with LNM had higher p-S6 expression. Furthermore, the expression level of p-S6 in metastatic lymph node lesion was higher than that in matched primary lesion. Multivariate analysis indicated that the high p-S6 expression could be an independent predicted marker for LNM in patients with NSCLC. Ribosomal protein S6, as a famous effector of mTOR signal pathway, mostly involved in the regulation of cell size and proliferation [9]. However, there is rarely report about the effect of p-S6 on metastasis of NSCLC. To further clarify the metastatic ability of p-S6 in NSCLC, we inhibited p-S6 expression in A549 and SPC-A1 cell lines and found that down regulation of p-S6 weakened the migration and invasion ability of NSCLC cells. Taking together, these data suggest that p-S6 might play a major part in promoting invasion and metastasis of NSCLC. Similar to our discoveries, previous studies have reported that overexpressed p-S6 is positively related to LNM in RCCs, colorectal cancer, and epithelial ovarian cancer [18, 32, 33]. On the other hand, inhibition of p-S6 expression or S6 gene knockdown can significantly suppress the cell invasion and migration of several kinds of human cancers, such as esophageal cancer and colorectal cancer cells [29, 34]. All the mentioned above indicate p-S6 positively affects cell invasion and migration. Despite the number of studies describing the aberrant expression of p-S6 in tumors activating invasion and metastasis, the concrete

Fig. 5 Effects of p-S6 inhibition on migration and invasion of NSCLC cell lines (A549 and SPC-A1). A549 and SPC-A1 cells were treated with 5 nmol/L RAD001 and were subjected to transwell assay to detect cell migration and invasion activity. A Results showed that migration of cell lines, A549 and SPC-A1, decreased following treatment with RAD001. B Inhibition of p-S6 decreased the invasion ability of cell lines A549 and SPC-A1. ***p < 0.001, Control vs. RAD001. (200 ×)
biological mechanism is still unclear. One study reported that the phosphorylation defect of S6 inhibits the phosphorylation of paxillin, a focal adhesion protein, leading to inhibit the formation of local adhesion [29]. In this current study, we have little knowledge of how the aberrant expression of p-S6 involved in the LNM of NSCLC, further explorations for the intrinsic mechanism are required in the future.

Our results indicated that the OS rate of NSCLC patients with highly expressed p-S6 was obviously lower than that of those with low level of p-S6. Multivariate analysis showed that high expressed p-S6 was an independently prognostic indicator in NSCLC patients, which seem to favor the oncogenic role of p-S6. Previous studies on RCCs, gastric cancer, and glioblastomas also identified p-S6 as a novel poor prognosis biomarker [16–18]. Up to now, there are many inhibitors that can prevent S6 phosphorylation. Studies have reported that inhibition of p-S6 can significantly reduce tumor growth, which is important for effective response to treatment of triple negative breast cancer [35, 36]. In addition, mTOR inhibitor everolimus can effectively inhibit the level of p-S6, thereby reversing the resistance of HER2-mutant cancers to neratinib and exerting anti-tumor effects [37]. These further suggest that p-S6 may be a powerful biomarker in tumors and may provide novel strategy in targeted therapy of NSCLC.

Conclusions
In summary, a significant overexpression of p-S6 was found in NSCLC. Inhibition of p-S6 expression could weakened the migration and invasion ability of NSCLC cells and aberrant expression level of p-S6 might be an independent predictor for LNM of NSCLC patients. In addition, overexpressed p-S6 may be a novel poor prognostic biomarker for NSCLC patients.
### Table 3: Summary of univariate/multivariate analysis for overall survival in patients with NSCLC (n= 350)

| Variables          | Univariate analysis | Multivariate analysis |
|--------------------|---------------------|-----------------------|
|                    | Average survival    |                      | Exp (B)     | 95.0%CI     | P-value    |
|                    | (SE) 95%CI          | P-value               |                |            |            |
| **p-S6**           |                     |                       |                |            |            |
| High expression    | 44.561 (3.050)      | 38.583-50.540         | 0.013*       | 1.549      | 1.105-2.173| 0.011*     |
| Low expression     | 67.500 (5.336)      | 57.041-77.958         |               |            |            |
| **Clinical stages**|                     |                       |                |            |            |
| Stage I-II         | 68.200 (4.195)      | 59.977-76.423         | 0.000*       | 1.980      | 1.356-2.893| 0.000*     |
| Stage III          | 47.862 (3.834)      | 40.347-55.378         |               |            |            |
| **LN status**      |                     |                       |                |            |            |
| LNM                | 44.969 (3.201)      | 38.695-51.243         | 0.000*       | 1.609      | 1.098-2.360| 0.015*     |
| No LNM             | 74.932 (5.919)      | 63.330-86.533         |               |            |            |
| **Histological type**|                   |                       |                |            |            |
| SCC                | 63.464 (5.825)      | 52.046-74.882         | 0.240        | 1.137      | 0.806-1.603| 0.464      |
| ADC                | 50.588 (3.367)      | 43.988-57.188         |               |            |            |
| **Pathological grade**|                 |                       |                |            |            |
| Well/moderated     | 58.662 (3.509)      | 51.784-65.541         | 0.003*       | 1.517      | 1.094-2.104| 0.013*     |
| Poor               | 54.226 (4.164)      | 46.066-62.387         |               |            |            |
| **Age**            |                     |                       |                |            |            |
| ≤ 50               | 45.395 (3.633)      | 38.274-52.515         | 0.454        | 1.014      | 0.718-1.430| 0.939      |
| >50                | 61.199 (4.064)      | 53.234-69.165         |               |            |            |
| **Gender**         |                     |                       |                |            |            |
| Female             | 54.566 (4.467)      | 45.810-63.321         | 0.210        | 0.718      | 0.484-1.064| 0.099      |
| Male               | 57.363 (3.886)      | 49.747-64.978         |               |            |            |

Abbreviations: CI Confidence interval, Exp(B) Odds ratio, SE Standard error, LNM Lymph node metastasis; *P < 0.05

### Abbreviations

NSCLC: Non-small cell lung cancer; ADC: Adenocarcinoma; SCC: Squamous cell carcinoma; LNM: Lymph node metastasis; AKT: The protein kinase B; mTOR: Mammalian target of rapamycin; S6: Ribosomal protein S6; S6K: 70 kDa S6 Kinases; P-S6: Phosphorylated S6; RCCs: Renal cell carcinomas; TMAs: Tissue microarrays; Non-CLT: Non-cancerous lung tissues.

### Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s12885-022-09664-4.

Additional file 1.
Additional file 2.

### Acknowledgements

Not applicable.

### Authors’ contributions

Qiuyuan Wen and Songqing Fan conceived and designed the study. Yaoxiang Tang and Jiadi Luo performed the experiments. Yaoxiang Tang, Yin Zhou, and Hongjing Zang wrote the paper. Yang Yang and Sile Liu analyzed the data. Qiuyuan Wen, Hongmei Zheng, and Jian Ma reviewed and edited the manuscript. All authors read and approved the manuscript and agree to take responsibility and be accountable for contents of the article.

### Funding

The work was supported by the Natural Science Foundation of Hunan Province (grant No. 2017JJ3457), Hunan Provincial Innovation Foundation For Postgraduate (grant No. CK20210373), the Fundamental Research Funds for the Central Universities of Central South University (grant No. 2021zzts1045) and the National Natural Science Foundation of China (grant No. 81703009, 81773218, and 81972838).

### Availability of data and materials

All data generated or analyzed during this study are included in this published article (and its supplementary information files). The datasets generated and analysed during the current study are available in the figshare repository, https://doi.org/10.6084/m9.figshare.16901161.v4.

### Declarations

#### Ethics approval and consent to participate

In this study, all procedures performed on research samples were confirmed by the ethical standards of the Institute Ethics Committee of the Second Xiangya Hospital of Central South University (approval No. S039/2011) and in accordance with the 1964 Helsinki declaration and its later amendments of comparable ethical standards. Informed consent was obtained from all subjects or their legal guardian.

#### Consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no conflicts of interest.

### Author details

1 Department of Pathology, The Second Xiangya Hospital, Central South University, Changsha 410011, Hunan, China. 2 Cancer Research Institute of Central
South University, Hunan Key Laboratory of Nonesolving Inflammation and Cancer, Central South University, Changsha-410011, Hunan, China.

Received: 30 December 2021 Accepted: 10 May 2022

Published online: 20 May 2022

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394–424. https://doi.org/10.3322/caac.21492.

2. Duma N, Santana-Davila R, Molina JR. Non-Small Cell Lung Cancer: Epidemiology, Screening, Diagnosis, and Treatment. Mayo Clin Proc. 2019;94(8):1623–40. https://doi.org/10.1016/j.mayocp.2019.01.013.

3. Miller KD, Nogueira L, Mariotto AB, Rowland JH, Yabroff KR, Alfano CM, et al. Cancer treatment and survivorship statistics, 2019. CA Cancer J Clin. 2019;69(5):363–85. https://doi.org/10.3322/caac.21565.

4. Woodard GA, Jones KD, Jablons DM. Lung Cancer Staging and Prognosis. Cancer Treat Rev. 2016;70:174–75. https://doi.org/10.1016/j.ctrv.2017.01.006.

5. Wen Q, Wang W, Luo J, Chen L, Xie G, et al. CGP57380 enhances efficacy of p-Akt, p-mTOR and p-eIF4E proteins associates with metastasis and unfavorable prognosis in non-small cell lung cancer. PLoS One. 2012;7(5):e35168. https://doi.org/10.1371/journal.pone.0035168.

6. Lu J, Zhang H, Zheng J, Zhan Y, Yang Y, Zhang Y, et al. Overexpression of p-Akt, p-mTOR and p-eIF4E proteins associates with metastasis and unfavorable prognosis in non-small cell lung cancer. PLoS One. 2012;7(5):e35168. https://doi.org/10.1371/journal.pone.0035168.

7. Aoki M, Fujishita T. Oncogenic Roles of the PI3K/AKT/mTOR Axis. Curr Proteomics. 2006;3(2):127–49. https://doi.org/10.2174/156802606776710058.

8. Wool IG, Chan YL, Gluck A. Structure and evolution of mammalian ribosomal protein S6 phosphorylation sites in Swiss mouse 3T3 fibroblasts. J Biol Chem. 1974;249(21):6917–25. https://doi.org/10.1016/j.tibs.2006.04.003.

9. Ruvinsky I, Meyuhas O. Ribosomal protein S6 phosphorylation: from protein synthesis to cell size. Trends Biochem Sci. 2006;31(6):342–8. https://doi.org/10.1016/j.tibs.2006.04.003.

10. Gressner AM, Wool IG. The phosphorylation of liver ribosomal proteins in vivo. Evidence that only a single small subunit protein (S6) is phosphorylated. J Biol Chem. 1974;249(21):6917–25.

11. Bandi HR, Ferrari S, Krieg J, Meyer HE, Jbrotov KR, Alfano CM, et al. Cancer treatment and survivorship statistics, 2019. CA Cancer J Clin. 2019;69(5):363–85. https://doi.org/10.3322/caac.21565.

12. Tanimura Y, Furukawa A, Shimizu M, Kuroki T, Itoh H, Itoh A, et al. Ribosomal protein S6 phosphorylation is a determinant of cell size and glucose homeostasis. Genes Dev. 2005;19(18):2199–211. https://doi.org/10.1101/gad.351605.

13. Ruvinsky I, Katz M, Dreazen A, Gielchinsky Y, Saada A, Freedman N, et al. Mouse deficient in ribosomal protein S6 phosphorylation suffer from muscular weakness that reflects a growth defect and energy deficit. PLoS One. 2009;4(5):e5618. https://doi.org/10.1371/journal.pone.0005618.

14. Ruvinsky I, Sharon N, Lerner T, Cohen H, Stolovich-Rain M, Nir T, et al. Ribosomal protein S6 phosphorylation is a determinant of cell size and glucose homeostasis. Genes Dev. 2005;19(18):2199–211. https://doi.org/10.1101/gad.351605.

15. Ruvinsky I, Katz M, Dreazen A, Gielchinsky Y, Saada A, Freedman N, et al. Mouse deficient in ribosomal protein S6 phosphorylation suffer from muscle weakness that reflects a growth defect and energy deficit. PLoS One. 2009;4(5):e5618. https://doi.org/10.1371/journal.pone.0005618.

16. Zheng Z, Zheng Z, Zhang M, Wang J, Gu G, Wang W, et al. Reciprocal expression of p-AMPKa and p-S6 is strongly associated with the prognosis of gastric cancer. J Biomed Biotechnol. 2011;144(5):646–74. https://doi.org/10.1186/1472-6750-144-5-646.

17. Nakamura H, Goto K, Sugiyama T, Kodama T, Suwa Y, Kurokawa K, et al. Ribosomal protein S6 phosphorylation attenuates DNA damage and cell proliferation via inhibition of the phosphoinositide 3-kinase-Akt-mTOR-ribosomal protein S6 pathway. Cell Commun Signal. 2019;17(1):28. https://doi.org/10.1186/s12964-019-0341-7.

18. Kim SH, Yang TH, Chau GC, Pyo S, Um SH. Prognostic significance and function of phosphorylated ribosomal protein S6 in esophageal squamous cell carcinoma. Mod Pathol. 2013;26(3):327–35. https://doi.org/10.1038/modpathol.2012.161.

19. Wittenberg AD, Azar S, Klockendler A, Stolovich-Rain M, Avraham S, Birnbaum L, et al. Phosphorylated Ribosomal Protein S6 Is Required for Akt-Driven Hyperplasia and Malignant Transformation, but Not for Hyper trophy, Aneuploidy and Hyperfunction of Pancreatic beta-Cells. PLoS One. 2016;11(2):e0149905. https://doi.org/10.1371/journal.pone.0149905.

20. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646–74. https://doi.org/10.1016/j.cell.2011.02.013.

21. Lai ET, Chen ZG, Zhou X, Xiao Y, Zhang Y, et al. DEPTOR expression is strongly associated with serous ovarian cancer. Mod Pathol. 2011;144(5):646–74. https://doi.org/10.1016/j.modpathol.2011.02.013.

22. Tang et al. BMC Cancer (2022) 22:564 Page 10 of 11

23. Seshadri N, Turraa M, Li J, Lee MW, Kwon S, Lee KS, et al. 5-Hydroxytryptamine 4 receptor antagonism inhibits proteasomal degradation of oestrogen receptor-β in breast cancer cells. Curr Med Chem. 2018;25(46):5685–95. https://doi.org/10.2174/0929867325870838.

24. Knoll M, Macher-Goeppinger S, Kopitz J, Duensing S, Rahemt S, Hohenfellner M, et al. The ribosomal protein S6 in renal cell carcinoma functional relevance and potential as biomarker. Oncotarget. 2016;7(1):418–32. https://doi.org/10.18632/oncotarget.6225.
37. Sudhan DR, Guerrero-Zotano A, Won H, Ericsson PG, Servetto A, Huerta-Rosario M, et al. Hyperactivation of TORC1 Drives Resistance to the Pan-HER Tyrosine Kinase Inhibitor Neratinib in HER2-Mutant Cancers. Cancer Cell. 2020;37(2):258–9. https://doi.org/10.1016/j.ccell.2020.01.010.

**Publisher’s Note**
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.