Autologous Bone-Marrow vs. Peripheral Blood Mononuclear Cells Therapy for Peripheral Artery Disease in Diabetic Patients

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Introduction

Peripheral artery disease (PAD) is a part of non-coronary artery syndrome caused by changes in the lower extremities’ arterial structure and function, which atherosclerotic lesion and thrombo-embolism are generally considered as the underlying pathophysiology (1). Worldwide, more than 200 million people were affected by this disease in 2010 (2). Many studies have shown a strong association between diabetes mellitus (DM) and an increased prevalence of PAD (3, 4). Indeed, PAD is an established manifestation of macrovascular complications in diabetes other
than coronary artery disease and stroke. Approximately 20% of DM patients older than 40 years were affected by PAD. Meanwhile, 20–30% of PAD patients have DM (3). Our data in Dr. Cipto Mangunkusumo National General Hospital, Jakarta, Indonesia, showed that between 2008 and 2012, there were 54 patients with PAD underwent endovascular revascularization and almost 80% of them had type 2 diabetes (T2D) (5).

As evident, the presence of diabetes greatly accelerates PAD disease progression, with a more severe below-the-knee lesion, and significantly increases the risk of ischemic ulcer, gangrene, and major amputation rates, compared to those without DM (6). Around 10–20% of patients with lower limb PAD will progress to critical limb ischemia (CLI), a severe form of PAD characterized by pain at rest, ulceration, and necrosis (7, 8). CLI is associated with unfavorable prognosis, recurrent hospitalization, lower quality of life, higher risk for cardiovascular death, and 5-year survival rates of 50% or even less (9). In addition to optimum medical therapy and risk factors modification, the current gold standard management for severe PAD and CLI is revascularization through endovascular intervention or surgical bypass (10, 11). Unfortunately, up to 40% of these patients were not good candidates for revascularization procedures due to excessive operative risk or unfavorable vascular involvement (12). Therefore, a new approach is in great demand for these ‘no option’ patients to reduce the amputation and mortality rates. In the past two decades, autologous stem cell therapy, especially mononuclear cells (MNC), has been used extensively for ‘no option’ CLI patients with promising results. However, it is still not available for routine clinical practice. This review will compare the two primary sources of MNC, peripheral blood and bone marrow (BM), for its use in CLI management in diabetes patients.

**Stem Cell Therapy**

Stem cell therapy has emerged as a new therapeutic strategy for many diseases. Stem cells are cells that have the capacity to renew themselves and to produce differentiated cells. Based on the extent to which they can differentiate into different cell types, stem cells can be classified as totipotent, pluripotent, multipotent, oligopotent, or unipotent (13). Additionally, based on their origin, stem cells can be classified into four broad types: embryonic, fetus, infant, and adult stem cells (13, 14). Embryonic stem cells, which have pluripotent properties, can differentiate into any type of cell in the body. Meanwhile, the adult stem cells with multipotent properties pose a higher specificity than embryonic stem cells. They could produce different types of cells only for specific organs and tissues (14). In contrast to embryonic stem cells, the ethical aspects of adult stem cells are relatively not controversial because they originate from tissue samples of adult humans, not human embryos. Although, safety issues especially for long-term follow-up, are still becoming a concern (15).

In normal tissue experiencing an ischemic injury, there will be a revascularization compensatory response consisting of angiogenesis, vasculogenesis, and arteriogenesis that plays an essential role in repairing damaged blood vessels and forming new blood vessels or neovascularization (16). These very intricate processes involved inflammatory cells, numerous chemokines and cytokines, various growth factors, and stem cells (16–18). Unfortunately, these mechanisms are defective in diabetes and CLI patients (19, 20). The tremendous potential inherited by stem cells to differentiate and evolve into differentiated cell types holds the prospect to be used as one modality for PAD or CLI management in T2D patients. The stem cells gradually get stimulated by the surrounding cellular environment (damaged hypoxic tissues), which will lead to the formation of specialized cells identical to those they come in contact and grow with (21, 22). Paracrine effects induced by the stem cells were also responsible for supporting these reparative processes (23).

**Bone Marrow Mononuclear Cells (BMMNC)**

As one primary source of adult stem cells, bone marrow has been explored extensively for its use in stem cell therapy in CLI. Type of stem cells from the bone marrow that has been widely used as therapy include bone marrow mononuclear cells (BMMNC), or its subsets, which consist of endothelial progenitor cells (EPC) and mesenchymal stem/stromal cells (MSC).

The term BMMNC is used to describe cells with unilobed or round nuclei and lack of granules in the cytoplasm that are found in the bone marrow. This broad definition results in a diverse cell population that comprises hematopoietic stem and progenitor cells (HSC/HPC), lymphoid cells, monocyte, and mesenchymal stem cells (MSCs). These characteristics differ from those of myeloid and red blood cell progenitor, thereby facilitating the separation of BMMNC by physical means (24). Further efforts have been made to characterize the angiogenic stem and progenitor population in the BMMNC fraction. Asahara et al. (22) have managed to identify a class of circulating mononuclear cells expressing CD34 with the ability to differentiate into endothelial cells in vitro and con-
tribute to vasculogenesis in adult animal models, thus defined as putative EPC. Although, until now, the concept of EPC has become a subject of debate and controversies, as many studies from different research groups have used different isolation methods and identification to define EPC-like cells (25-28). Moreover, many markers that were previously used to describe putative EPC, such as CD34, CD117, and CD133, have also been known to be shared by myeloid cells of various stages of differentiation (29, 30). Thus, make the issues more complicated.

Another imperative fraction of BMMNC is the MSC. Despite their small number in bone marrow (~1 in every ten thousand bone marrow cells), these cells can be multiplied ex-vivo into larger quantities to achieve required doses (31). In addition, there are also alternative sources of MSCs, including adipose tissues (32, 33), dental pulp (34), periodontal ligament (35) peripheral blood (36), cord blood (37), umbilical cord (38, 39), and placenta (40). International Society for Cell Therapy (ISCT) proposed the minimum criteria to describe multipotent MSCs, which are plastic adherence in ordinary culture conditions; expression of cell surface CD73, CD90, and CD105; lack of expression of cell-surface CD34, CD45, HLA-DR, CD14, CD79a, or CD19; and multilineage differentiation capacity (41). Despite these proposition, it is prominent that MSCs isolated from different tissue and/or under different conditions will likely result in diverse or more differentiated stem cells, albeit still defined as multipotent progenitor cells (42, 43). Due to these variations, it should be kept in mind that MSCs between studies are less likely to signify identical cell types since details such as expansion technique and isolation method may affect clinical results.

BMMNC used for therapeutic purposes in clinical trials usually obtained by bone-marrow aspiration from ileum under local or general anesthesia depending on the withdrawn volume (44, 45). Next, the aspirate will undergo a purification and concentration step using a density gradient centrifugation, either manually or preferably by an automated cell separator (46). The application of an automated cell separator allowed for shorter processing time and reduced risk of microbial contamination with higher mononuclear cells recovery and preserved or even better functional capacity (47, 48).

Peripheral Blood Mononuclear Cells (PBMNC)

A peripheral blood mononuclear cell (PBMNC) is any peripheral blood cell having a round nucleus, consisting of lymphocytes (T cells, B cells, NK cells) and monocytes (49). The EPCs could also be identified in isolated PBMNC, although only as a small fraction (50). As the mononuclear cells in the peripheral blood originally come from bone marrow, these cells could be stimulated or mobilized by giving a systemic injection of granulocyte colony-stimulating factor (G-CSF) or granulocyte-macrophage colony-stimulating factor (GM-CSF) to increase the number in the peripheral blood without affecting its function and capacity (51-53). Thus, the quantities or dosage of this CD34+ mobilized PBMNC will be sufficient for therapeutic angiogenesis. The receptors of G-CSF are expressed primarily on neutrophils and bone marrow precursor cells. Whereas, for GM-CSF, the receptors are more widely expressed and present on neutrophils, monocytes, eosinophils, dendritic cells, basophils, and B cells (54). Both growth factors decreased the expression of adhesion molecules on the CD34+ cells, such as very late antigen-1 (VLA-1), L-selectin, vascular cell adhesion molecule-1 (VCAM-1), and CXCR4 – a receptor for stromal-cell derived factor-1 (SDF-1) – thus facilitating the mobilization of these CD34+ cells to peripheral blood (55).

Several trials have shown that G-CSF and GM-CSF administration leads to a dose-dependent increase of EPC in peripheral blood (56, 57). In addition, G-CSF is known to augment differentiation of marrow cells into vasculature endothelial cells, resulting in early blood flow recovery in ischemic tissues (58). However, G-CSF is considered preferable than GM-CSF due to the higher yield of CD34+ cells and less or shorter duration of apheresis (59-61). This might explain the scarcity of study using GM-CSF mobilized PBMNC for critical limb ischemia. Important drawbacks of research using stem cell mobilization are their efficacy is significantly influenced by race, ethnics, and disease status (62). Therefore, it is essential to weight patient characteristics when comparing each different PBMNC study. After mobilization with G-CSF or GM-CSF, the PBMNCs are then harvested by means of cell apheresis, either alone or in conjunction with an additional blood-cell separator system (57, 63-65).

Potential Mechanism of BMMNC and PBMNC in PAD

The mechanism on how BMMNC or PBMNC can promote neovascularization is still considered as an ongoing investigation. A study by Asahara et al. (22) has first demonstrated that peripheral blood CD34+ mononuclear cells can differentiate into endothelial cells and several other studies support this finding (66-68). However, this concept still becomes a controversy since other studies did not ob-
serve similar result. Studies by Göthert et al. (69) and Purhonen et al. (70) that incorporates fluorescent protein-labeled BM-derived cells failed to discover these cells in the vascular walls after transplantation.

Another proposed mechanism for the role of stem cells in PAD is the stabilization of neovascularization through pericyte differentiation. Pericytes are branched cells that present at intervals along the walls of capillary blood vessels (and postcapillary venules), embedded within the microvascular basement membrane and wrapping endothelial cells (71). Pericytes are communicating with endothelial cells by direct physical contact and paracrine signaling pathways. They play an essential role in stabilizing the vessel wall, regulating microvascular blood flow, and endothelial cell proliferation (72, 73). By utilizing green fluorescent protein (GFP)-chimeric mice, Rajantie et al. (74) were unable to find evidence of BM-derived endothelial cells at sites of tumor or vascular endothelial growth factor (VEGF)-induced angiogenesis. Instead, they consistently observed BM-derived periendothelial cells at the sites, and these cells expressed NG2 proteoglycan, a marker for developing pericytes. These findings are supported by Ziegelhoeffer et al. (75), who also used GFP-chimeric mice that underwent unilateral femoral artery occlusions as a model of hind-limb ischemia. They failed to colocalize GFP signals with endothelial or smooth muscle cells marker in the growing collateral arteries. Nevertheless, accumulations of GFP-positive cells were detected in the perivascular space, appearing like pericytes and fibroblasts based on their shape and distribution. These suggest that in the adult organism, BM-derived cells do not promote vascular growth by incorporating into vessel walls but may function as supporting cells.

Besides direct incorporation into the vascular structure, BMMNC might play an essential role in neovascularization in PAD via its potent paracrine signaling. In the site of vascular injury, BMMNC and PBMNC have been found to secrete various pro-angiogenic factors, such as basic fibroblast growth factor (bFGF), VEGF, hepatocyte growth factor (HGF), and angiopoietin-1 (76, 77). VEGF is one of the main regulators in angiogenesis. It acts as a chemotactic marker for macrophage and granulocyte and stimulates endothelial cells migration, matrix metalloproteinase activity, and forming of the blood vessel lumens (78). Additionally, it can also promote vessel enlargement, increase vessel branching, and maintaining vascular homeostasis (79). Meanwhile, bFGF exerts its role in angiogenesis by stimulating endothelial cells migration, invasion, and production of plasminogen activator, as well as influencing other growth factors and chemokines secretion (80, 81). HGF has similar properties to bFGF and can enhance the angiogenesis potency of VEGF (82). These growth factors have been used in several clinical trials as a therapy for critical limb ischemia, although with mixed results (83).

**Clinical Trials of Stem Cell Therapy for PAD in Diabetes Patients**

Animal trials using rabbit, rat, and mouse in hind-limb ischemia (HLI) models have shown promising result in which BMMNC, PBMNC, or MSC transplantation could improve capillary density, neovascularization and collateral growth, blood perfusion, upregulation of b-FGF and VEGF levels, as well as decreases in auto amputation rate and muscle atrophy (84).

These encouraging results from animal studies prompt Tateishi-Yuyama et al. (85) to conduct the Therapeutic Angiogenesis using Cell Transplantation (TACT) study, the earliest human clinical trial using intramuscular injection of autologous BMMNC or unstimulated PBMNC in the ischemic limb of PAD patients with CLI. This pilot study demonstrated significant improvement in ankle-brachial index (ABI), transcutaneous oxygen pressure (TCPO2), resting pain, and pain-free walking time, that persists after 24 weeks in BMMNC compared to the unstimulated PBMNC.

Clinical studies of BMMNC and PBMNC therapy for limb ischemia have significantly improved from uncontrolled phase I trial to ongoing randomized, placebo-controlled, clinical phase III trial. Huang et al. (56) first described the benefit of autologous mobilized PBMC transplantation in diabetic CLI in terms of lower limb amputation, limb ulcer healing, ischemic pain, blood perfusion, and ABI. This study utilized five days of G-CSF therapy to mobilize the MNCs to peripheral blood, followed by apheresis which was then used for autologous injection. Despite Tateishi-Yuyama et al. (85) demonstrated the superiority of BMMNC compared to unstimulated PBMNC, it is important to note that Tateishi-Yuyama et al. did not mobilize the MNCs using G-CSF. This might result in a very low number of MNCs or CD34+ cells in the PBMC-treated subjects, thus potentially affecting the outcome. However, a recent study by Persiani et al. (86) utilizing unstimulated PBMC showed a significant improvement of ABI and ischemic pain in diabetic CLI patients after transplantation. Ozturk et al. (57) reported better improvement of Fontaine score, ABI, TcPO2, and pain score in diabetic CLI patients treated with G-CSF mobilized PBMC compared to control subjects treated with stand-
ard of care. Additionally, a more recent larger-scale randomized clinical trial by Horie et al. (87) using G-CSF mobilized PBMNC transplantation showed that PBMNC therapy was able to significantly improve the progression-free survival of mild-to-moderate PAD.

Several trials conducted to compare the efficacy between BMMNC and PBMNC as CLI therapy has shown mixed results. A study by Dubsky et al. (65) diabetic CLI patients, showed no differences in the improvement of TcPO2 between BMMNC and PBMNC group, which complied with the finding from Huang et al. (64). However, the latter study reported significantly better improvement in ABI, skin temperature, and resting pain after 12 weeks in PBMNC than BMMNC group. In addition, a meta-analysis by Liew et al. (88) demonstrated a better reduction of major amputation rate, improvement of ulcer healing, and ABI in PBMNC but not BMMNC when compared to the control group. Therefore, PBMNC might not be inferior to BMMNC, especially with the added practicality of avoiding invasive and painful procedure, its adverse risk, and cost. In the future, PBMNC treatment can even be commenced as an outpatient procedure to reduce patient burden.

There has been a significant number of meta-analysis assessing the effect of stem cells therapy in PAD. Mostly, these meta-analyses demonstrated a substantial improvement of surrogate endpoints such as ABI, TcPO2, ischemic pain, ulcer healing, and pain-free walking distance (PFWD) after stem cells transplantation. However, in terms of primary outcomes such as amputation rate, overall survival or mortality, and amputation-free survival (AFS), mixed results have been observed (89-93). Ai et al. (90) and Rigato et al. (91) showed a reduced major amputation rate and an increased rate of AFS in cell-based therapy. Xie et al. (92) and Gao et al. (94) also revealed a reduction in total amputation rate after stem cell therapy but failed to show improvement in major limb salvage. Meanwhile, meta-analyses that only include trials with randomized placebo-controlled intervention or with low risk of bias consistently did not show a significant improvement in total or major amputation rate, mortality, and AFS in the treatment group compared to placebo, after stem-cells transplantation (88, 93, 95).

It appears that the earlier relatively small and non-placebo controlled trials demonstrated more evident advantages of cell therapy, whereas the larger placebo-controlled studies showed less convincing results. It is important to note that there is a high risk of bias in the type of trials without randomization and placebo-controlled, whether in the intervention given or outcome evaluation. Many clinical trials have failed to show the advantages of stem cell therapy for PAD in the major amputation rate and overall survival. We could speculate that the improvement of neovascularization as shown by the improved surrogate outcome is not sufficient to the extent of preventing major amputation. Moreover, diabetic patients with CLI might already have many complications in the other organs and the possibility of diffuse atherosclerotic lesions in the other vascular beds, including cardiovascular and cerebrovascular system (6). These conditions will eventually increase the risk of mortality, which cannot be corrected by merely administering stem cell therapy.

**Pitfalls of Stem Cell Therapy for PAD in Diabetes**

Preclinical studies initially raise some concerns regarding the efficacy of autologous MNC in diabetic individuals. Hyperglycemia condition in diabetes is associated with decreased number and function of EPC. Animal studies have shown that diabetic EPCs exhibit decreases adhesion and migration activities; decreased fibronectin, collagen I, vitronectin, and laminin expression; impaired expression of eNOS and VEGF; and reduced capability to incorporate into the vascular structure (19, 96). Many studies have revealed a significantly decreased number of circulating CD34+ cells in diabetic patients, especially with foot ulcers and PAD (97-99). Additionally, this reduced quantity of circulating EPC is more prevalent in complicated diabetes rather than uncomplicated diabetes (96). Despite all these drawbacks, treatment of autologous mobilized PBMNC could still improve neovascularization, albeit to a lesser extent than nondiabetics (100). Interestingly, a recent meta-regression analysis by Rigato et al. (91) and meta-analysis by Gao et al. (94) implied that the benefit of cell therapy on amputation rate was higher in trials with a majority of patients having diabetes mellitus.

With regards to optimization studies, discussion about optimal dose and delivery route remains open. A meta-analysis by Ai et al. (90) did not find a significant correlation between cell dosages and the therapeutic effects. Additionally, a study by Losordo et al. (101) showed the lowest beneficial dose for CD34+ cells for PAD is $1 \times 10^5$ cells/kg body weight and increasing the number of injected cells would not result in significant improvement of the outcomes. However, a meta-analysis by Pan et al. (93) showed significant results in major amputation, total amputation, and complete ulcer healing rate in the high CD34+ cell dosage ($1 \times 10^6$ cells/kg body weight), which were not observed in low-dose studies. Hence, more studies comparing different cell dosages with a sufficient num-
ber of subjects and randomized placebo-controlled designs are needed to confirm the optimal dosage for MNC therapy in PAD.

The intramuscular approach injects a high concentration of stem cells in the ischemic areas that require angiogenesis. Despite documented low engraftment rate (0.44 – 10% after 96 hours) (84), the justification behind intramuscular injection is to deposit cell reservoir near the ischemic area (Fig. 1). On the other hand, the intraarterial method aims to direct stem cells to margin areas with adequate supply of oxygen and nutrients to promote new cell activities. However, it is not yet known how many cells are able to reach the ischemic areas from the blood vessels. Studies that directly compare intramuscular and intra-arterial injection of BMMNC in CLI patients have shown no significant differences (102, 103).

The safety of BMMNC and PBMNC therapy for PAD has generally been established. Although bone marrow aspiration as a procedure to obtain the BMMNC is considered more invasive compared to apheresis for obtaining PBMNC, it is still categorized as low-risk procedure. Some potential adverse events associated with bone marrow aspiration include pain, low-grade fever, bleeding, and infection (104). In addition, bone marrow aspiration, especially with large volume aspirate, has also been described to decrease mean hematocrit by 2.6% compared to control. However, this condition is tolerable in most patients (102). Nevertheless, this should be considered in at-risk patients such as anemic or coronary artery disease patients. Meanwhile, the administration of G-CSF for mobilization of PBMNC before apheresis carries some potential risks. Mostly, the adverse events are mild such as bone pain, headache, fatigue, and flu-like symptoms (103). However, there are some case reports on very rare cases of serious adverse events associated with G-CSF injection in healthy subjects, like spleen rupture (106), acute lung injury (107),

![Fig. 1. A schematic diagram of angiogenesis process following stem cell injection of the affected area of PAD. EC: Endothelial Cells. Adapted from: Lawall H, et al. Thromb Haemost; 2010.](image)

### Table 1. Advantages and drawbacks of cell products used for peripheral artery disease

| Cell therapy types          | Advantages                                                                 | Drawbacks                                                |
|-----------------------------|---------------------------------------------------------------------------|----------------------------------------------------------|
| Bone-Marrow derived         | Most widely researched cell product for PAD.                               | Risk and complications during bone marrow aspiration.    |
| Mononuclear Cells (BMMNC)   | - Several meta-analyses showed improvement in ABI, TcPO2, pain scores, and ulcer healing in BMMNC treated patients (90–92). |                                                          |
|                             | - Availability of long term safety result (3 years) (109).                 |                                                          |
| Peripheral Blood derived    | Head-to-head comparison with BMMNC showed similar benefits (65).          | Risk and complications during G-CSF/GM-CSF administration (105). |
| Mononuclear Cells (PBMNC)   | - Recent meta-analysis showed PBMNC, but not BMMNC are able to reduce risk of major amputation and improve ulcer healing (88). | Efficacy of stem cells mobilizations are significantly influenced by race, ethnics, and disease status (62). |
| Mesenchymal Stem Cells (MSC) | - Promising result from preclinical and clinical studies.                  | - Risk and complications during bone marrow aspiration (in BMMSC). |
|                             | - Able to be isolated and expanded to achieve consistent and desirable dose. | Reports of potential role of MSC in tumor development.    |
|                             | - Allow larger-scale manufacturing with good quality control to produce an ‘off the-shelf’ cellular products. | - High variability of cell products between protocols and research facility. |
|                             | - Possibility of allogeneic therapy due to its non or low-antigenic properties. | - Relatively more expensive.                             |
and transient state of hypercoagulability that could give rise to thrombotic complications (108). No musclar damage, blood vessel malformation, or acute kidney injury has been identified with any intramuscular injection of MNCs (88). Additionally, there is also no report of unexpected long-term adverse effects (109, 110). The advantages and drawbacks of cell products used for PAD are shown in Table 1.

Conclusions

Extensive phase I and II trials have established the safety of BMMNC and PBMNC as well as its efficacy in improving ischemic pain, reducing ulcer size, pain-free walking distance, ABI, and TcPO2. Nonetheless, the capacity to establish the efficacy to reduce major amputation rates, AFS, and all-cause mortality is still lacking due to the limited number of phase III studies. In addition, an occurring trend is observed in which small, non-placebo-controlled study seems to demonstrate a seemingly larger benefit compared to large, placebo-controlled study. This phenomenon raises some concern, especially because several important endpoints are susceptible to bias. Future larger placebo-controlled RCTs are needed before further translation of MNC therapy to standard care. Additionally, current clinical trials using MNC therapy in PAD are only performed in end-stage disease PAD when the patients are unable to undergo revascularization treatment. It might be important to conduct clinical study in mild to moderate PAD to not miss the potential benefit of this novel therapy as the more advanced disease is related to the decreased regenerative capabilities of the vasculature and the functionality of the autologous stem cells itself.

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Potential Conflict of Interest

The authors have no conflicting financial interest.

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