Arch Clin Infect Dis. 2020 April; 15(2):e90449. 
doi: 10.5812/archcid.90449.

Research Article

In Silico Design of Multi-Epitope ESAT-6:Ag85b:Fcγ2a Fusion Protein as a Novel Candidate for Tuberculosis Vaccine

Masoud Keikha1, Mohsen Karbalaei2 and Kiarash Ghazvini1,*

1Department of Microbiology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran
2Department of Microbiology, School of Medicine, Jiroft University of Medical Sciences, Jiroft, Iran

*Corresponding author: Department of Microbiology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran. Email: ghazvinik@mums.ac.ir

Received 2019 February 08; Revised 2020 April 05; Accepted 2020 March 12.

Abstract

Mycobacterium tuberculosis (MTB), which is the causative agent of tuberculosis (TB), is among the most important infectious bacteria with high morbidity and mortality rates worldwide. Bacilli Calmette-Guerin (BCG) vaccine has been discovered for about a century, and it is considered as a major vaccine for humans. However, some factors, such as its attenuated nature and its inefficacy against the latent form of the disease, have led to the use of alternative vaccines. Multi-epitope subunit vaccines are new-generation vaccines that are being developed in clinical trial phases. For the production of a subunit vaccine, the selection of immunodominant antigens and targeted delivery systems to antigen presenting cells (APCs) are considered as basic parameters. In the present study, we designed the novel multi-epitope ESAT-6:Ag85b:Fcγ2a, which was evaluated completely by various online tools as an optimum vaccine against TB. The early secreted antigenic target of 6 kDa (ESAT-6) and antigen 85B (Ag85B) are two immunodominant antigens, and Fcγ2a is a targeted delivery system. This vaccine candidate can be used for future preclinical studies.

Keywords: ESAT-6, Ag85B, Mycobacterium tuberculosis, Vaccine, Tuberculosis

1. Background

Tuberculosis (TB) is a re-emerging disease that is still a major global health concern (1, 2). According to the World Health Organization (WHO), a worldwide incidence of 10 million new TB cases and 1.5 million deaths in 2018 have been reported (2).

Tuberculosis is an infectious disease and vaccination is one of the best schemes in preventing infectious diseases. Since the 1920s, Bacilli Calmette-Guerin (BCG) has been the only licensed TB vaccine used for the prevention of TB among children and adults. However, according to numerous clinical trials, this vaccine has some disadvantages, including lack of efficient long-term immunity against TB, various efficacy in adult pulmonary TB (0% - 80%), and side-effects such as severe cutaneous reaction or even disseminated BCG infection, and it is not recommended for HIV-infected individuals. Therefore, more research is required to introduce new-generation TB vaccines (1, 3-5).

M. tuberculosis is a facultative intracellular bacterium which could survive and cause persistent infection within alveolar macrophages using about 1,500 various proteins such as ESAT-6, CFP-10, MTP64, hspX, and Ag85 complex (6).

ESAT-6 (Rv3875) and fibronectin binding protein b (Ag85Bs)(Rv1886c) are known as major virulence factors expressed and secreted in early phases of infection with M. tuberculosis (MTB). These antigens induce strong immune responses both in animals (e.g., IFN-γ or IL-2) and humans (6-8). Several favorable vaccine categories have been designed using ESAT-6 and Ag85Bs proteins, which could induce a high level of immunity against the early stage of MTB infection with valid and impressive results both in animal studies and I, IIa, and IIb clinical trials (5, 6). Recently, epitope-based vaccines have become more popular than classical DNA vaccines. The epitope-based vaccine technology has provided several advantages such as the insertion of multi-immunodominant epitopes from several proteins in the form of one macromolecule, reduction of potential allergenicity, and decreasing cloning and expression problems (9, 10).

Surprisingly, clinical experiments have shown that binding these antigens to the Fc domain of immunoglobulin (Ig) can lead to their selective uptake. The Fc delivery system plays a pivotal role in improving antigen delivery to macrophages and DC cells (11). Consequently, the use of this system leads to efficient uptake of antigens and processing by antigen presenting cells (APCs) and creates great immune response by Th1 and cytotoxic T-cells (CTLs) (11, 12). In addition, subunit vaccines connected to the Fc
Keikha M et al.

fragment of IgG increase serum half-life, solubility, stability, and purification of these subunit antigens (11-13).

In recent decades, vaccine design has focused on producing multi-epitope vaccines as a new approach, and research in bioinformatics can help design effective subunit vaccine candidates against TB. Computational tools, which are cost-effective and time-saving, play an important role in detecting appropriate epitopes (14).

2. Objectives

In the current study, based on the early secretory protein of MTB infection, we designed a novel multi-epitope subunit vaccine by focusing on the Fc fragment of IgG using computational tools.

3. Methods

3.1. Selection of Appropriate Targets

There are numerous antigen candidates to be selected in early replication (e.g., EsxA, EsxB, Ag85 complex, and Mpt64) and dormancy (e.g., DosR families and HspX) phases. In this study, we attempted to design a multi-epitope vaccine based on the early phase of MTB infection. These antigens should be immunodominant in pathogenesis and secreted in the early stages of MTB replication.

3.2. Primary Sequence Alignment and Homology

ESAT-6 and Ag85B gene sequences were retrieved from NCBI (www.ncbi.nlm.nih.gov) for in silico analysis and were then compared with other related sequences using BLAST software to select their conserved regions. In addition, the mouse FC fragment of IgG2a (Fcγ2a) domain (coding regions of hinge-CH2-CH3) was added to the final fusion construct to increase the selective uptake process (11, 13).

3.3. Evaluating Physicochemical Features

Online servers, including Uniprot, PDB, and ProtParam were employed to evaluate Ag85B and ESAT-6 antigens. By the use of online servers, some properties (e.g., molecular weight, isoelectric pH, negatively and positively charged residues, half-life and instability index) of these antigens were evaluated (11).

3.4. Primary Constructed Synthetic Sequence

Protein sequences of the Ag85B, ESAT-6, and Fcγ2a domains were joined to one another by GGSSGG and AAY linkers as a synthetic fusion construct, named Ag85b:ESAT-6:Fcγ2a (11).

3.5. Immune-Informatics Analysis

3.5.1. Prediction of MHC class I and II Binding Epitopes

We used IEDB, nHLAPred, and MHC2Pred online tools in order to predict MHC class I and II epitope-restricted binding sequences (15).

3.5.2. Prediction of B Cell Binding Epitopes

Prediction of B cell binding-epitopes was confirmed using the BepiPred online server from the IEDB online tool according to the Kolaskar and Tongaonkar antigenicity scale (15).

3.5.3. Evaluating Epitopes for HLA Binding

Binding sites of human HLAs, that is, MHC class I and II-restricted epitopes were evaluated using the IDEB online server.

3.6. Fusion Construct Design and Analysis

3.6.1. Primary Synthetic Multi-Epitopes Construct Design

A new mouse construct was designed based on combining T cell and B cell high immune-modulatory of ESAT-6 and Ag85B selected epitopes. These epitopes bind with each other with appropriate linkers, and subsequently, the fusion construct binds to mFcγ2a (mouse IgG2a) for the enhancement of selective uptake by APCs.

3.6.2. Codon Optimization

The multi-epitopes construct was codon optimized using the JCat tool (http://www.jcat.de) online server.

3.6.3. Construct Sequence Analysis

ProtParam (expasy.org/tools/protparam.html) was employed for the evaluation of amino acid composition and physicochemical properties.

3.6.4. Prediction of Dimer Conservation in Protein Modeling Process

The synthetic construct was developed in dimeric form, and protein modeling was performed using the MODELER tool and protein structure prediction servers such as the (PS)2-v2 and I-TASSER online servers. Then, all the models were assessed according to their scores.

3.6.5. Prediction of mRNA Structure

The secondary structure of mRNA was predicted using the Secondary Structure Web Server (16).
3.6.6. Post-Translational Modification (PTMs) of Multi-Epitopes Construct

Post-translational modifications such as glycosylation, phosphorylation, methylation, and acylation are specifically for eukaryotic cells, while such modifications do not exist in prokaryotic cells. Modification of the new construct may affect the structure of constructs. Therefore, modified structures were predicted by several online servers, which are listed in Table 1 (15, 16).

3.6.7. Prediction of Antigenicity Properties of Designed Construct

VaxiJen was employed to evaluate the antigenicity parameters of our multi-epitopes construct.

3.6.8. Prediction of Allergenicity of Designed Construct

AlgPred and PREAL online servers were applied to predict the allergenicity of our construct.

3.6.9. Docking Stimulation

Docking was carried out by the PatchDock and Xex 8.0 software programs to evaluate potential binding groove of MHC I (PDB ID: 3VCL, 4XXC, and 4O2C) and MHC II molecules (PDB ID: 1BX2, 1AQD, 5JLZ) (17).

4. Results

4.1. Primary Sequence Alignment and Homology

We selected ESAT-6 (NC_000962.3) and Ag85B (AY207396.1) gene sequences and compared them with other related gene sequences of the MTB complex. Using BLAST software, 99% similarity between gene sequences was shown for each of the MTB complex genes. Finally, conserved regions were selected for future evaluation. In addition, the mRNA sequence of IgG2a (accession number: V00798.1) was downloaded from NCBI in order to select coding regions of hinge region and Fc domain of mouse IgG2a.

4.2. Evaluating Physicochemical Features

ESAT-6 (PBD: 1WA8) and Ag85B (UniProt: Q847N4) protein sequences were downloaded from PBD and UniProt and evaluated using ProtParam as an initial screening for determining the first suitable confirmation for the vaccine candidate. These proteins were considered appropriate to be selected as a TB vaccine candidate.

4.3. Primary Constructed Synthetic Sequence

The primary construct was synthesized using the combination of Ag85B-GGSGG-ESAT-6-AAYAAY-Fcγ2a sequences (Figure 1A-C).

4.4. Immune-Informatics Analysis

4.4.1. Prediction of MHC Class I and II Binding Epitopes

Appropriate immune epitopes were predicted according to BALB/c mouse H-2 class I alleles, including H2-Kd, H2-Db, and H2-Ld. Three epitopes were selected based on high score values (including LSNAVPTGS, MTDSVRKI-RAWGRRLMIGTAAAVVLPLGLAGGAATAGAFSRPL, and FS-RPGPLVEYLQVPSMG with the total scores of 0.83, 0.81 and 0.71, respectively). Moreover, the protein sequence was submitted on IDEB/MHC II for evaluating potential binding with BALB/c mouse H-2 Class II alleles consisting of H2-IAd and H2-IEd. Subsequently, three epitopes were selected based on the highest score results via online tools, including QGNVTSHISL, EAGQAMASTEGRNMFA, or VQFQSGGNNSPA with the cut-off scores of 0.72, 0.68, and 0.62, respectively.

4.4.2. Prediction of B Cell Binding Epitopes

Predicted B cell linear epitopes were AAAAVLPGLVGLA, GLPVEYLQVPS, and AVYLLD domains with an appropriate threshold value higher than 0.5. In addition, Kolaskar and Tongaonkar plot was conducted with a high rate of 1.227 (Figure 2).

4.4.3. Evaluating Epitopes for HLAs Binding

The selected MHC I and II restricted epitopes were analyzed to assess their potential ability in binding to human HLA alleles by the IDEB online tool. All the selected epitopes were confirmed and were able to bind to human HLA alleles with acceptable population coverage higher than 50%, such as AKMKAAGG, GLIGGS, or DKKVEPKS with the total rates of 98.5, 98.0 and 97.5, respectively.

4.5. Fusion Construct Design and Analysis

4.5.1. Primary Synthetic Multi-Epitopes Construct Design

The final construct was designed based on MHC class I, II, and linear B-cell epitopes that were joined using the GGSGG linker and were also connected to the Fcγ2a fragment of IgG (PDB: 1IGT) to increase selective uptake delivery system.

4.5.2. Codon Optimization

The synthetic gene was a codon adapted according to a famous eukaryotic expressing system as Saccharomyces cerevisiae for appropriate post-translational modification process.
4.5.3. Construct Sequences Analysis

Physicochemical parameters of the new construct were evaluated by the ProtParam online tool. The properties of this construct included the number of amino acids: 293, molecular weight: 30 kDa, pH: isoelectric 8.29, negative charge: 20, positive charge: 22, half-life > 30 hours, aliphatic index: 78.57 and gravy: -0.01.

4.5.4. Prediction of Dimer Conservation in the Protein Modeling Process

The synthetic gene was converted to dimeric forms, and all the predicted models were evaluated based on a high score rate (GDT_TS score > 60%, ProQ score 1.6, Bit score 120.3 with E value 0.014). Finally, construct stability was confirmed using ERRAT (http://services.mbi.ucla.edu/ERRAT/) and Ramachandran software programs to be 96% and 90%, respectively.
4.5.5. Prediction of mRNA Structure

Twenty mRNA structures were predicted, and all the structures were checked to select the best optimal structure according to $\Delta G$ rate. Our mRNA model had $\Delta G$: $-225$ kcal/mol and did not have leucine zipper or coiled coil, and the hairpin loop and stem loop domain was lower than 20%.

4.5.6. Post-Translational Modification (PTMs) of Multi-Epitope Construct

We employed numerous online servers for PTMs prediction including Glycosciences, NetOGlyc, NetPhos, NetPhosK, Myristoylator, NMT, CELLO, and BaCelLo (Table 1).

4.5.7. Prediction of Antigenicity Properties of Construct Design

The synthetic gene construct was determined as an antigen according to the Vaxijen online software (cut-off: 0.7).

4.5.8. Allergenicity Evaluation of Construct Design

Both AlgPred and PREAL online tools predicted that the synthetic construct ESAT-6:Ag85B:Fc;2a was not an allergen (reliability: 90%).

4.5.9. Docking Results

Several epitopes such as SVTVPSSLGTQTY, FS-RPGLPEYLQPSPMGR, VQFQSGGNNSPA and DGLGGSV were evaluated for docking with several MHC class I and II alleles. Among them, SVTVPSSLGTQTY and FS-RPGLPEYLQPSPMGR were bound closely with MHC (4O2C and 1AQD) with the high scores of 11910 and 10196, respectively (Figure 4).

5. Discussion

Nowadays, in addition to therapeutic protocols for the treatment of TB infections, the role of other strategies such as autophagy, antithetical effects of microRNA molecules and the obvious role of T regulatory (T reg) cells in the pathogenesis of TB have been known to researchers (18-21). However, it can be stated that prescription of effective preventive vaccines is considered as an important strategy against TB infection. BCG is the only available TB vaccine approved by the WHO from 1921. However, this vaccine has serious drawbacks and side effects, including its contraindication among HIV patients, its inefficacy in adult...
pulmonary TB and its inability to control reactivation of the latent form (3, 21). According to the literature, BCG vaccine is not able to induce the production of long-life T cells, which are continuously expressed for pro-inflammatory cytokines. There are numerous cases that have received attenuated BCG vaccine in childhood, but they have been affected by TB after puberty (5). Recently, subunit vaccines have been used as BCG prime boost and are also considered proper candidates for the new generation of TB vaccines. These can provoke CD4+ T cells’ memory, which, in turn, produce a protective response against TB that could remain more than one year (22).

Nowadays, subunit vaccines are applied for several purposes, including prophylaxis (protection), post-exposure (prevention of TB reactivation using dormant antigens) and therapeutic targets (even treatment of MDR-TB) (3, 4). Subunit vaccines such as H1/IC31 can also be recommended for HIV-infected individuals or immunocompromised patients (23). The problems associated with subunit vaccines are high production costs, inefficient
bioavailability, low antigen stability during formulation or hydrolysis, and inactivation process (6). Therefore, computational analysis is the first choice for rational design and is considered as the best economical option to predict and process synthetic subunit vaccine properties (16).

Multi-epitope subunit vaccines are new strategies which are developed to produce highly efficient vaccines. The multi-epitope-pulsed dendritic cell vaccine and multi-epitope TARP peptides are famous examples of epitopes which have entered clinical trials (24). Online bioinformatics databases have provided the possibility to predict the antigenic region of proteins. Each of the efficient vaccines should be screened and evaluated by bioinformatics tools before production (14). On the other hand, PCR amplification of high GC-rich genes such as fbpB is challenging and not easily setup. Consequently, construction of fusion epitopes is appropriate and very efficient for triggering cellular immunity response (25). In addition, Ag85B and ESAT-6 are considered as important early expressed antigens of MTB, and subunit vaccine ESAT-6:Ag85B has entered phase IIa of the clinical trial (7).

In the present study, we applied highly immunogenic epitopes of ESAT-6 and Ag85B connected to Fcγ2a and as-

| Online Server URL Address | Function | Modification Types | Residue |
|---------------------------|----------|--------------------|--------|
| http://www.glycosciences.de/ | Prediction of O-linked glycosylation sites in eukaryotes | O-linked glycosylation | T2 |
| www.cbs.dtu.dk/services/NetNGlyc | Prediction of N-linked glycosylation sites in eukaryotes | O-linked glycosylation | T13 |
| http://montana.informatics.indiana.edu/ModPred/index.html; http://montana.informatics.indiana.edu/ModPred/index.html | Prediction of di-sulfide bond sites | Disulfide linkage | C14 |
| www.cbs.dtu.dk/services/NetPhos | Prediction of phosphorylation sites in eukaryote | Phosphorylation | No |
| www.cbs.dtu.dk/services/NetPhosK | Prediction of kinase sites in eukaryote | No | No |
| mendel.imp.ac.at/myristate/SUPL predictor.html | Prediction of N-terminal myristylation | Lipid | No |
| mendel.imp.ac.at/myristate/SUPL predictor.html | Prediction of N-terminal N-myristylation | No | No |
| www.cbs.dtu.dk/services/SignalP | Prediction of signal peptide (SPase) sites in protein sequence | Signal peptide | E29 |
| gpcr.biocomp.unibo.it/bacello/ | Prediction of extracellular localization of proteins | Extracellular | Extracellular |
| cello.life.nctu.edu.tw/ | Prediction of extracellular localization of proteins | Extracellular | Extracellular |
| https://wolfpsort.hgc.jp/ | Prediction of extracellular localization of proteins | Secreted pathway | Secreted pathway |

Table 1. Post-Translation Modification and Related Online Servers

Keikha M et al.

Arch Clin Infect Dis. 2020; 15(2):e910449.
sessed those using certifiable online servers. According to the literature, MTB as a facultative intracellular pathogen is responsible for the activities of the cellular immune system such as Th1 and CTLs control and elimination of TB bacilli from the human body (26). Thus, our construct was designed based on MHC class I and II, T cell, HLA alleles and B cell restricted high score epitopes. Subsequently, fusion-epitopes were combined with the constant region of the Fcγ2a fragment. The Fcγ2a fragment is necessary for enhancing selective uptake and stability and promoting the aliphatic index of the synthetic construct. In order to increase antigenic capacity, multi-epitopes subunit vaccine ESAT-6:Ag85B:Fcγ2a was converted to dimer form, and the 3D model of the dimeric peptide was calculated by I-TASSERS with reliable C-score, ERRAT-score, and acceptable Ramachandran plot.

mRNA stability plays a significant role in the amount of protein expression and total protein stability. Computational analysis showed that the predicted model of our final synthetic construct did not have a hairpin loop or leucine zipper, which in turn, could lead to mRNA stability or prevention of mRNA aggregation, and subsequently, hydrolysis. Moreover, ΔG index is a reliable marker for the prediction of mRNA stability structure, and the low amount of ΔG suggests an increase in mRNA stability (15).

Post-translation modifications cause substantial changes in proteins, which could lead to protein maturation in eukaryotic systems after the completion of the translation phase (27). According to bioinformatics databases, our fusion protein was glycosylated and had disulfide bonds and signal peptides and was expressed as an extracellular protein. According to the literature, glycosylation can improve the immunogenic properties of proteins, and disulfide bonds stabilize the tertiary or quaternary protein structures and fix dimeric peptides together (15, 28). Also, the signal peptide sequence provokes the secretion of proteins outside of the cell.

Docking analysis was evaluated as a reliable test to determine the possibility of interaction of antigens with HLA molecules. The computational docking analysis is reported by energy scores, where low energy scores reveal a high possibility of receptor-ligand interaction.

5.1. Conclusions

In summary, after several screening tests, we concluded that our synthetic multi-epitopes ESAT6:Ag85B:Fcγ2a construct can be considered as a potential vaccine. The reliability of our new construct is established based on various reasons including (1) using ESAT6 and Ag85B as two main secretory antigens in the active phase of MTB infection; (2) using Fc fragment as a natural biological targeted delivery system to APCs; and (3) using these three components in the form of a multi-epitope fusion protein for targeted and strong stimulation of cellular immune system. In addition, basic bioinformatic studies can significantly impact the development of efficient vaccines and reduce production costs. Overall, we hope that this synthetic multi-epitope fusion candidate will be applied for in-vitro and in-vivo studies illin the near future.

Footnotes

Authors’ Contribution: GK did study design. KG and KM did data collection and analysis. KM and KM wrote the manuscript.

Conflict of Interests: The authors have no conflicts of interest.

Funding/Support: None to declare.

References

1. Keikha M, Esfahani BN. The relationship between tuberculosis and lung cancer. Advanced biomedical research. 2018;7. [PubMed: 29657943].
2. World Health Organization. Global status report on alcohol and health 2018. Geneva: World Health Organization; 2019.
3. Babaki MKZ, Soleimanzad S, Rezaee SA. Antigen 85 complex as a powerful Mycobacterium tuberculosis immunogen: Biology, immunopathogenicity, applications in diagnostics, and vaccine design. Microbial pathogenesis. 2017;112:20–9. [PubMed: 28942772].
4. Keikha M, Karbalaei M. Can Multi-Stage Recombinant Fusion Proteins be Considered as Reliable Vaccines Against Tuberculosis? A Letter to the Editor. Modern Care Journal. 2019;16(2). doi: 10.5812/modercnj.9493.
5. Khoshnood S, Heidary M, Haeili M, Drancourt M, Darban-Sarokhalil D, Nasiri MJ, et al. Novel vaccine candidates against Mycobacterium tuberculosis. International journal of biological macromolecules. 2018;120:180–8. [PubMed: 30098365].
6. Khademi F, Derakhshan M, Youssefi-Avarvand A, Tafaghodi M, Soleimanzad S. Multi-stage subunit vaccines against Mycobacterium tuberculosis: an alternative to the BCG vaccine or a BCG-prime boost? Expert review of vaccines. 2018;17(1):31–44. [PubMed: 29148853].
7. van Dessel JT, Arend SM, Prins C, Bang P, Tingskov PN, Lingnau K, et al. Ag85B:ESAT-6 adjuvanted with IC31® promotes strong and long-lived Mycobacterium tuberculosis specific T cell responses in naive human volunteers. Vaccine. 2010;28(20):3571–81. [PubMed: 20226890].
8. Navid S, Keikha M. Can Fusion Proteins Be Considered a New Candidate for Tuberculosis Vaccine? Gene, Cell and Tissue. 2018;3(3). doi: 10.5812/gct.79943.
9. Sharbier A. Multi-epitope DNA vaccines. Immunology and cell biology. 1997;75(4):402–8. [PubMed: 9154885].
10. Negahdaripour M, Nezafat N, Esiami M, Ghoshoon MB, Shoiliain E, Najafipour S, et al. Structural vaccinology considerations for in silico design of a multi-epitope vaccine. Infection, Genetics and Evolution. 2018;58:96–109. [PubMed: 29253673].
11. Farsani H, Mosavat A, Soleimanzad S, Sadeghian H, Eydgahi MRA, Ghazvini K, et al. Fc-based delivery system enhances immunogenicity of a tuberculosis subunit vaccine candidate consisting of the ESAT-6: CFP10 complex. Molecular BioSystems. 2016;12(7):2189–201. [PubMed: 27138226].

Keikha M et al.

Arch Clin Infect Dis. 2020;15(2):e90449.
12. Soleimanpour S, Farsiani H, Mosavat A, Ghazvini K, Eydghaï MRA, Sankian M, et al. APC targeting enhances immunogenicity of a novel multistage Fc-fusion tuberculosis vaccine in mice. *Applied microbiology and biotechnology*. 2015;99(24):10467–80. [PubMed: 26373723].

13. Babaki MKZ, Taghiabadi M, Soleimanpour S, Moghadam MS, Mosavat A, Amini AA, et al. Mycobacterium tuberculosis Ag85B: hfc/γ1 recombinant fusion protein as a selective receptor-dependent delivery system for antigen presentation. *Microbial pathogenesis*. 2019;129:68–73. [PubMed: 30751546].

14. Phuphanich S, Wheeler CJ, Rudnick JD, Mazer M, Wang H, Nuno MA, et al. Phase I trial of a multi-epitope-pulsed dendritic cell vaccine for patients with newly diagnosed glioblastoma. *Cancer Immunology, Immunotherapy*. 2013;62(1):125–35. [PubMed: 22847020].

15. Jahangiri A, Rasooli I, Gargari SEM, Owlia P, Rahbar MR, Amani J, et al. An in silico DNA vaccine against Listeria monocytogenes. *Vaccine*. 2011;29(40):6948–58.

16. Moradi J, Tabrizi M, Izad M, Mosavari N, Feizabadi MM. Designing a novel multi-epitope DNA-based vaccine against tuberculosis: in silico approach. *Jundishapur Journal of Microbiology*. 2017;10(3). doi: 10.5812/jjm.43950.

17. Khatoon N, Pandey RK, Prajapati VK. Exploring Leishmania secretory proteins to design B and T cell multi-epitope subunit vaccine using immunoinformatics approach. *Scientific reports*. 2017;7(1):3-12. [PubMed: 28816060].

18. Yousefi M, Eslami M, Karbalaei M, Keikha M, Ghazvini K. Autophagy as one of the most important strategies for the treatment of tuberculosis; Mini-review. *Reviews in Clinical Medicine*. 2019;6(4):135-9.

19. Keikha M, Shabani M, Navid S, Sadegh EB, Karbalaei ZBM. What is the role of “T reg Cells” in tuberculosis pathogenesis? *The Indian journal of tuberculosi*. 2018;65(4):360. [PubMed: 30522628].

20. Keikha M, Karbalaei M. Antithetical Effects of MicroRNA Molecules in Tuberculosis Pathogenesis. *Advanced biomedical research*. 2009;8. [PubMed: 30775345].

21. Keikha M, Soleimanpour S, Eslami M, Yousefi B, Karbalaei M. The mystery of tuberculosis pathogenesis from the perspective of T regulatory cells. *Meta Gene*. 2019;10:6632. doi: 10.1016/j.mgene.2019.100662.

22. Aagaard C, Hoang T, Dietrich J, Cardona P, Izzo A, Dolganov G, et al. A multistage tuberculosis vaccine that confers efficient protection before and after exposure. *Nature medicine*. 2011;17(2):189. [PubMed: 21258338].

23. Andersen P. Vaccine strategies against latent tuberculosis infection. *Trends in microbiology*. 2007;15(1):7-13. [PubMed: 17141504].

24. Wood LV, Fojo A, Roberson BD, Hughes MS, Dahut W, Gulley JL, et al. TARP vaccination is associated with slowing in PSA velocity and decreasing tumor growth rates in patients with Stage D0 prostate cancer. *Oncoimmunology*. 2016;5(8). e1197459. [PubMed: 27822067].

25. Hanke T, Neumann VC, Blanchard TJ, Sweeney P, Hill AV, Smith GL, et al. Effective induction of HIV-specific CTL by multi-epitope using gene gun in a combined vaccination regime. *Vaccine*. 1999;17(6):589-96. doi: 10.1016/S0264-410X(98)00238-2.

26. Raja A. Immunology of tuberculosis. *Indian Journal of Medical Research*. 2004;120(Oct):213-22.

27. Anderson P. Post-transcriptional regulons coordinate the initiation and resolution of inflammation. *Nature Reviews Immunology*. 2009;10(1):31-42. [PubMed: 19808419].

28. Mohammad N, Karsabet MT, Amani J, Ardjmand A, Zadeh MR, Gholi MK, et al. In silico design of a chimeric protein containing antigenic fragments of Helicobacter pylori; a bioinformatic approach. *The open microbiology journal*. 2016;10:97. [PubMed: 27335622].