Assumption-free estimation of the genetic contribution to refractive error across childhood

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Purpose: Studies in relatives have generally yielded high heritability estimates for refractive error: twins 75–90%, families 15–70%. However, because related individuals often share a common environment, these estimates are inflated (via misallocation of unique/common environment variance). We calculated a lower-bound heritability estimate for refractive error free from such bias.

Methods: Between the ages 7 and 15 years, participants in the Avon Longitudinal Study of Parents and Children (ALSPAC) underwent non-cycloplegic autorefraction at regular research clinics. At each age, an estimate of the variance in refractive error explained by single nucleotide polymorphism (SNP) genetic variants was calculated using genome-wide complex trait analysis (GCTA) using high-density genome-wide SNP genotype information (minimum N at each age = 3,404).

Results: The variance in refractive error explained by the SNPs (“SNP heritability”) was stable over childhood: Across age 7–15 years, SNP heritability averaged 0.28 (SE = 0.08, p < 0.001). The genetic correlation for refractive error between visits varied from 0.77 to 1.00 (all p < 0.001) demonstrating that a common set of SNPs was responsible for the genetic contribution to refractive error across this period of childhood. Simulations suggested lack of cycloplegia during autorefraction led to a small underestimation of SNP heritability (adjusted SNP heritability = 0.35; SE = 0.09). To put these results in context, the variance in refractive error explained (or predicted) by the time participants spent outdoors was < 0.005 and by the time spent reading was < 0.01, based on a parental questionnaire completed when the child was aged 8–9 years old.

Conclusions: Genetic variation captured by common SNPs explained approximately 35% of the variation in refractive error between unrelated subjects. This value sets an upper limit for predicting refractive error using existing SNP genotyping arrays, although higher-density genotyping in larger samples and inclusion of interaction effects is expected to raise this figure toward twin- and family-based heritability estimates. The same SNPs influenced refractive error across much of childhood. Notwithstanding the strong evidence of association between time outdoors and myopia, and time reading and myopia, less than 1% of the variance in myopia at age 15 was explained by crude measures of these two risk factors, indicating that their effects may be limited, at least when averaged over the whole population.

Myopia is the most common eye disorder worldwide [1,2] and is a risk factor for several important ocular pathologies, including glaucoma, retinal detachment, and chorioretinal degeneration [3–6]. The etiology of myopia is poorly understood, although genetic and environmental influences are known to be involved [for recent reviews, see 1,2,6–15].

The extent to which a quantitative trait is determined by genetic factors is termed “heritability.” Heritability in the “broad sense” (h²), as calculated in twin studies, refers to the proportion of the phenotypic variation in the trait due to additive, dominant, and gene–gene interaction effects [16] while heritability in the “narrow sense” (h²), as calculated in family studies, refers to the proportion of phenotypic variance explained by additive genetic effects alone [17]. Since genes and environment never operate in isolation, the utility of a heritability estimate may appear questionable [16,18]. However, the narrow sense heritability indicates the potential of genetic testing for identifying individuals at high risk of a specific disease [19]. Refractive error has been shown to be highly heritable [1,2,6–15,20–24]: In a recent review [25], the broad sense heritability ranged from 0.75 to 0.90 and the narrow sense heritability from 0.15 to 0.70.

The availability of techniques to accurately and cost-effectively survey genetic variation across the genome now makes it feasible to quantify the contribution of causal variants tagged by genotyped or imputed single nucleotide polymorphisms (SNPs) to differences in metric traits or disease susceptibility between individuals [26,27]. The inter-subject
variance in a trait that can be explained by all the SNPs has been termed SNP heritability \( (h_{SNP}^2) \). For an infinitely large population in which every polymorphism was genotyped without error, SNP heritability \( h_{SNP}^2 \) would equal heritability in the narrow sense \( h^2 \). For finite-sized samples in which only common SNPs can be genotyped or imputed accurately, \( h_{SNP}^2 \) therefore corresponds to a lower-bound estimate of \( h^2 \). Theoretical and empirical work suggests that \( h_{SNP}^2 \) provides a valuable guide to, first, the potential for predicting at-risk individuals using currently available genotyping platforms and, second, the potential for identifying specific disease-associated genetic variants in a genome-wide association study (GWAS) using existing genotyping and imputation methods but applied to larger samples.

An important, fundamental difference between SNP-heritability estimation methods and prior methods of estimating heritability is that \( h_{SNP}^2 \) is computed using a sample of essentially unrelated individuals. Thus, \( h_{SNP}^2 \) estimates have the attractive feature of not being biased by the confounding effects of shared family environment. Furthermore, they are not reliant on other assumptions required to obtain comparable estimates in twins [26,27]. As with conventional \( h^2 \) estimates, the genetic correlation between two traits (i.e., the degree to which the two traits are determined by a shared set of additive genetic variants) can also be calculated using SNP data.

Prior work suggests that at least 40–50% of the between-subject variance in height [27,28], axial length [29], and corneal curvature [29] can be explained by common SNPs. We applied this approach to estimate the SNP heritability of refractive error for children participating in a birth cohort study. We also calculate genetic correlations between refractive error measured at different ages, to infer whether the same or different genetic variants influence susceptibility to refractive error across childhood.

**METHODS**

**Subjects and ethical approval:** The Avon Longitudinal Study of Parents and Children (ALSPAC) recruited 14,541 pregnant women resident in Avon, England, with expected dates of delivery 1 April 1991 to 31 December 1992. When the oldest children were approximately 7 years of age, an attempt was made to bolster the initial sample with eligible cases who had failed to join the study originally, resulting in an additional 713 children being enrolled. The total number of children who were alive at 1 year of age was 14,701. The enrollment is described in more detail in a cohort profile article [30]. This research adhered to the tenets of the Declaration of Helsinki. Ethical approval for the study was obtained from the ALSPAC Ethics and Law Committee and the Local Research Ethics Committees (LRECs). Separate consent was given for the collection of refractive data from the participant’s own optometrist. The ALSPAC website contains details of all the data available through a fully searchable data dictionary.

**Phenotypes:** ALSPAC participants were invited to a research clinic when they were aged approximately \( 7\frac{1}{2}, 10\frac{1}{2}, 11\frac{1}{2}, 12\frac{1}{2}, \) and \( 15\frac{1}{2} \) years old. At each visit, refractive error was estimated with non-cycloplegic autorefraction (Canon R50 instrument, Canon USA Inc., Lake Success, NY) as described and validated previously [31-33]. After the outlier readings were removed, the mean spherical equivalent (SE) refractive error was calculated as the autorefraction sphere power plus half of the cylinder power and averaged between the two eyes. For one set of analyses, refractive errors were transformed to normal scores [34] to test whether the non-normal distribution of the trait biased the results. As discussed below, and in the Appendix, autorefraction in the absence of cycloplegia is known to lead to inaccuracy, with the degree of measurement error varying with the age of the child, his or her level of refractive error, the type of autorefractor used, and other subject-specific and random factors [33,35-38].

When the study children were aged 8–9 years, a questionnaire was completed by their mother that included the following questions: (1) “On a weekend day, how much time on average does your child spend each day out of doors in summer?” (2) “On normal days in school holidays, how much time on average does your child spend each day reading books for pleasure?” The response options for these questions were “None at all,” “1 hour,” “1–2 hours,” and “3 or more hours.”

**Genetic and statistical analyses:** DNA samples were genotyped using Illumina HumanHap550-Quad bead arrays, as described [39,40]. Blood sampling and DNA extraction were carried out as described [30]. Samples of known non-European ancestry, with missingness >3%, minimal or excessive autosomal heterozygosity, cryptic relatedness >10% identity by descent (IBD), or with a sex mismatch were excluded. EIGENSTRAT analysis and multidimensional scaling analysis seeded with individuals from HapMap phase 2 revealed no additional outliers. SNPs with call rate <95%, minor allele frequency (MAF) <1%, or Hardy–Weinberg equilibrium (HWE) p value ≤5×10^{-7} were excluded. After phasing haplotypes with ShapeIT v2.r644 [41], IMPUTE2 (V2.2.2) [42] was used to impute unobserved marker genotypes for the 8,237 participants whose samples passed the quality control criteria (imputation was performed simultaneously with a sample of 8,196 mothers of ALSPAC participants [43], who were included to improve phasing and imputation accuracy) with the Dec 2013 release of the 1000 Genomes Project.
haplotypes (Version 1, Phase 3) as the reference set. Data for seven participants were excluded from further analysis after the subjects withdrew their consent to participate. Genomic control lambda ($\lambda_{GC}$) values computed after GWAS analyses for a range of traits using the ALSPAC children data set showed minimal evidence of population stratification ($\lambda_{GC}<1.06$).

The statistical approach used for estimating SNP heritability contrasts fundamentally with that of GWAS gene discovery analyses, in which millions of SNPs are tested one by one, resulting in millions of statistical tests. A SNP-heritability analysis requires only one statistical test to be performed (a linear regression analysis). Specifically, the difference in phenotype between pairs of individuals is used as the dependent variable, and genetic relatedness between the same pairs of individuals is used as the primary explanatory variable (modeled as a random effect using restricted maximum likelihood; REML). Other explanatory variables such as age and sex can be included in the model as fixed effects. PLINK 1.90 [44] and GCTA v1.24.4 [45] were used to compute the pairwise genetic relationship matrix (for individual chromosomes and all chromosomes, respectively). There has been debate in the literature about the advantages and disadvantages of excluding SNPs in linkage disequilibrium when computing the genetic relationship matrix for SNP-heritability analysis [46-48]. In light of this debate, the genetic relationship matrix was calculated as described by Zhu et al. [49]. Thus, SNPs with an imputation reliability score <0.6, MAF <0.01, or HWE test p value <10⁻⁶ were excluded, followed by extraction of SNPs present in HapMap3, which left 1,192,778 markers for calculating genetic relatedness in a sample of 8,230 individuals. One subject from each of the 1,028 pairs within the cohort with an estimated genetic relationship >0.025 (the approximate degree of sharing expected between second or third cousins) was excluded, leaving 7,202 “unrelated” individuals. SNP-heritability analyses were run either with the raw refractive error readings or after transforming refractive error measurements to normal scores [34], to determine whether the non-normal distribution of the trait influenced the SNP-heritability estimate. Bivariate analyses were run to estimate the genetic correlation between all possible pairs of traits [29]. Univariate and bivariate modeling was performed with GCTA. Sex was included as a covariate; however, for the results reported age was not included since its omission had no effect on the parameter estimates.

Simulations were performed to assess the impact of refractive error measurement error due to the lack of cycloplegia. Briefly, this involved simulating a hypothetical trait such that it had a predetermined SNP-heritability level (ranging from 0.2 to 0.8 in steps of 0.1) and then assessing the SNP heritability before and after measurement error “noise” was added to the trait. The choice of the noise level is discussed in Appendix 1 and Figure 1 and Figure 2. In practice, the genotype data and genetic relatedness information for the 3,404 subjects in the age 15 years analysis were used to simulate phenotypic trait values at a given SNP-heritability level (0.2, 0.3, …, 0.8) by randomly selecting 1,000 SNPs (MAF range 0.01 to 0.50) from the genome as “causal variants,” with the effect size for each SNP drawn from a normal distribution, using LDAT [46]. In R, the standard deviation of the simulated trait was converted to 1.29 D to match the standard deviation of refractive error in 15-year-old ALSPAC participants (Table 1) and a randomly selected measurement error noise value was added, drawn from a normal distribution with a mean zero and a standard deviation of 0.50 D. GCTA was used to calculate the SNP heritability for the trait before and after noise was added. Fifty replicates were simulated at each heritability level.

The variance in refractive error at ages 10, 11, 12, and 15 predicted (or explained) by time spent outdoors was assessed using a general linear model, using the information reported by the mother when the child was aged 8–9 (see the Phenotypes section, above). Sex was included as a covariate. The variance predicted (explained) by time spent reading was gauged using the same method.

**RESULTS**

At each ALSPAC research clinic, non-cycloplegic autorefraction measurements were obtained for between 3,404 and 5,103 participants who also had genotype data available and were less related to each other than second to third cousins (Table 1). Univariate GCTA modeling suggested the SNP heritability for refractive error was 0.19 to 0.27 (weighted mean $h^2_{SNP}=0.23$, SE≈0.08, p<0.001) when estimated using untransformed trait values and 0.20 to 0.34 (weighted mean $h^2_{SNP}=0.28$, SE≈0.08, p<0.001) after the trait was transformed to a normal distribution (Table 2). These estimates therefore represent the proportion of the inter-subject variance in refractive error that can be explained by common SNPs in linkage disequilibrium (LD) with causal genetic variants that influence refractive error. Accordingly, a lower bound estimate for the heritability of refractive error—free from any bias due to sharing a common home environment—is approximately 25%.

Between 2,997 and 4,238 unrelated genotyped participants underwent autorefraction at two different ages (Table 3). The genetic correlation between refractive error assessed at each age varied between 0.77 and 1.00 (SE=0.07)
for untransformed trait values, and after refractive error readings were transformed into normal distributions. Estimates from closely-spaced clinic visits were generally higher than those from more widely-spaced visits, as would be expected, and genetic correlation estimates for refractive error assessed at or after the age of 10 were all ≥0.94 (Table 3). Thus, at each of the ages examined the same set of genetic variants explained about 25% of the variance in refractive error, instead of one set of genetic variants acting early in childhood and a different set of variants acting later in childhood.

To assess the impact of measurement error during autorefraction due to the lack of cycloplegia, we simulated cycloplegic and non-cycloplegic autorefraction traits for the 3,404 children included in the age 15 years analysis (Table 1). In each simulation run, 1,000 SNPs were randomly selected as “causal variants” and assigned a phenotypic effect such that the simulated trait had a predetermined $h^2_{SNP}$ value. Then the $h^2_{SNP}$ was calculated using GCTA before and after phenotypic measurement error noise was added at the level expected to occur because of the lack of cycloplegia (Appendix 1; Figure 1 and Figure 2). As shown in Figure 3, there was a close correspondence between the simulated and observed $h^2_{SNP}$ before the measurement noise was added (“cyclo” autorefraction simulation in Figure 3; slope=1.059). However, the observed $h^2_{SNP}$ was underestimated after measurement error noise was added to the trait (“non-cyclo” autorefraction simulation in Figure 3; slope=0.851). Applying a conversion factor of 1.244 (1.059/0.851=1.244) to account for the level of underestimation due to the lack of cycloplegia yielded an adjusted weighted mean $h^2_{SNP}$ of 0.35 (SE=0.09). Since the measurement error caused by the lack of cycloplegia would be at least as bad at younger ages, and since any non-systematic source of measurement error such as hyperopes accommodating more than myopes would cause $h^2_{SNP}$ to be further underestimated, this adjustment is likely to be conservative.

To help give context to the genetics findings, the variance explained (or predicted) by time spent reading and by time spent outdoors was also estimated (Table 4). Time spent

![Figure 1](http://www.molvis.org/molvis/v21/621)

**Figure 1.** Comparison of subjective refraction recorded by the subject’s own optometrist and non-cycloplegic autorefraction obtained at the 15-year ALSPAC research clinic. Whether the Avon Longitudinal Study of Parents and Children (ALSPAC) clinic record indicated that the subject was not wearing contact lenses during the autorefraction test is indicated by the symbol color. Note that the single outlying data point for the subjects confirmed as not wearing contact lenses corresponds to a myopic participant recorded as perfectly emmetropic by the optometrist, suggesting an error in data entry or retrieval instead of a highly inaccurate autorefraction measurement.
outdoors assessed using a questionnaire completed by the mother when the participants were aged 8–9 years predicted or explained <0.5% of the variance in refractive error at older ages, while the corresponding figure for time spent reading for pleasure assessed at a similar age was <1%.

**DISCUSSION**

The SNPheritability estimate of approximately 35% obtained here gives by far the most compelling support to date for a polygenic inheritance component for refractive error. Unlike prior heritability studies, the present estimate is free from assumptions about, and confounded by, shared environment effects, for example, parents and their children, or siblings, sharing the same home environment.

**Figure 2.** Difference between non-cycloplegic autorefraction obtained at the 15-year ALSPAC research clinic and subjective refraction recorded by the subject’s own optometrist. Two frequency distributions are plotted according to whether the Avon Longitudinal Study of Parents and Children (ALSPAC) clinic record indicated that the subject was or was not wearing contact lenses during the autorefraction test. Note the reduction in outliers and better conformity to a normal distribution when contact lens wear was excluded.
What is the relationship between SNP heritability ($h^2_{\text{SNP}}$), broad sense heritability ($H^2$), and narrow sense heritability ($h^2$)? Simulation studies and GWAS-GCTA analyses with large sample sizes for traits such as height [27,50] have helped clarify this issue. $h^2_{\text{SNP}}$ gives a lower-bound estimate of $h^2$. Thus, taking a figure of 54% (95% confidence interval [CI] 36 to 72%) for the narrow sense heritability of refractive error (calculated for a sample of 890 ALSPAC participants who had refraction information available for a sibling [51]) the gap between our $h^2_{\text{SNP}}$ estimate of 35% and the $h^2$ figure of 54% is likely due, first, to upward bias in the estimation of $h^2$ from shared family environment effects and, second, to two sources of downward bias in $h^2_{\text{SNP}}$: namely, the limited ability of genetic relatedness indices to capture the additive genetic effects of rare variants, and statistical “noise” associated with a modest sample size. For height, studies in larger samples and employing higher-density genotyping platforms have shown that $h^2_{\text{SNP}}$ steadily increases toward $h^2$ [50].

### Table 2. SNP-heritability (estimates of variance explained by SNPs) for refractive error, at a range of ages.

| Research clinic visit | N       | Untransformed trait | Transformed trait |
|-----------------------|---------|---------------------|------------------|
|                       |         | $h^2_{\text{SNP}}$ | SE     | p-value | $h^2_{\text{SNP}}$ | SE     | p-value |
| 7                     | 5103    | 0.250               | 0.067  | 9.7e-05 | 0.342               | 0.068  | 1.6e-07 |
| 10                    | 4862    | 0.198               | 0.069  | 1.5e-03 | 0.204               | 0.069  | 1.1e-03 |
| 11                    | 4433    | 0.194               | 0.075  | 4.0e-03 | 0.260               | 0.076  | 2.0e-04 |
| 12                    | 4438    | 0.270               | 0.076  | 1.4e-04 | 0.323               | 0.076  | 5.1e-06 |
| 15                    | 3404    | 0.259               | 0.097  | 2.9e-03 | 0.270               | 0.096  | 1.8e-03 |
| Mean                  |         | 0.233               | ~0.08  | <0.001  | 0.281               | ~0.08  | <0.001  |

Analyses included sex as a covariate. Estimates are shown for refractive error in Diopters (“untransformed trait”) or after refractive error readings were transformed to normal scores (“transformed trait”). P values correspond to a test of whether the heritability estimate is different from zero. The mean was calculated after weighting by sample size (N), using the formula, $\Sigma(h^2 N)/\Sigma N$.

### Table 3. Genetic correlations between refractive error assessed at two different ages.

| Age    | Age 7   | Age 10  | Age 11  | Age 12  | Age 15  |
|--------|---------|---------|---------|---------|---------|
| Age 7  | x       | (0.94)  | (0.98)  | (0.91)  | (0.77)  |
|        | n=4238  | n=3837  | n=3862  | n=3007  |
|        | 0.87    | 1.00    | 1.00    | 1.00    |
| Age 10 | (0.083) | x       | (0.035) | (0.037) | (0.083) |
|        | n=4238  | n=4060  | n=4062  | n=3146  |
|        | 0.89    | 1.00    | 0.99    | 0.96    |
| Age 11 | (0.089) | (0.077) | x       | (0.032) | (0.088) |
|        | n=3837  | n=4060  | n=3878  | n=2997  |
|        | 0.78    | 1.00    | 0.99    | 1.00    |
| Age 12 | (0.089) | (0.067) | (0.058) | x       | (0.055) |
|        | n=3862  | n=4062  | n=3878  | n=3127  |
|        | 0.77    | 1.00    | 0.96    | 0.94    |
| Age 15 | (0.124) | (0.107) | (0.099) | (0.070) | X       |
|        | n=3007  | n=3146  | n=2997  | n=3127  |

The standard error of the genetic correlation is shown in brackets. N refers to the sample size. Analyses included sex as a covariate. Estimates are shown for refractive error in Dioptres above the diagonal, and after refractive error was transformed to a normal score below the diagonal.
The gap between $h^2$ and $H^2$ is generally attributed to non-additive sources of genetic variation, which are included in $H^2$ but absent from $h^2$. Unlike $h^2_{SNP}$, estimates of $h^2$ and $H^2$ are likely to be inflated due to misallocation of unique or common environment variance. Although our results do not allow us to address the extent of this upward bias (only giving a lower bound estimate for $h^2$ and $H^2$), the large gap between the narrow sense heritability and the broad sense heritability estimates for refractive error (e.g., 15–70% and 75–90%, respectively [25]) implies that dominance and/or interaction

### Table 4. Variance in refractive error explained by time spent reading and time spent outdoors.

| Research clinic visit | N     | Untransformed trait |       |       | Transformed trait |       |
|-----------------------|-------|---------------------|-------|-------|-------------------|-------|
|                       |       | $r^2$               | P-value |       | $r^2$            | P-value |
| **Time spent reading at age 8–9 years-old** |       |                     |       |       |                   |       |
| 10                    | 4616  | 0.003               | 7.7e-04 |       | 0.003            | 3.7e-04 |
| 11                    | 4203  | 0.009               | 6.7e-09 |       | 0.007            | 4.1e-07 |
| 12                    | 4209  | 0.008               | 1.4e-07 |       | 0.007            | 3.0e-07 |
| 15                    | 3298  | 0.008               | 2.1e-06 |       | 0.005            | 1.9e-04 |
| **Time spent outdoors at age 8–9 years-old** |       |                     |       |       |                   |       |
| 10                    | 4626  | 0.001               | 3.7e-01 |       | 0.001            | 9.6e-02 |
| 11                    | 4215  | 0.001               | 4.4e-01 |       | 0.001            | 2.2e-01 |
| 12                    | 4225  | 0.003               | 4.6e-03 |       | 0.002            | 1.4e-02 |
| 15                    | 3298  | 0.002               | 2.1e-02 |       | 0.001            | 3.3e-02 |

Analyses included sex as a covariate. Estimates are shown for refractive error in Diopters (“untransformed trait”) or after refractive error readings were transformed to normal scores (“transformed trait”).

Figure 3. Comparison of SNP-heritability estimation for a simulated refractive error trait before and after the addition of measurement error “noise.” Traits were simulated using a model designed to yield a specified SNP-heritability level (x-axis). The simulated trait was either analyzed directly (“cyclo”) or after the addition of noise designed to mimic the measurement error of non-cycloplegic autorefraction (“non-cyclo”). Measurement error led to the progressive under-estimation of the true SNP heritability.
The SNP heritability for refractive error was about 35%, providing an approximate upper limit for the capacity to predict the future refractive error of premyopic children using existing genotype platforms and imputation methods alone. Technical advances and larger sample sizes would be expected to raise $h^2_{\text{SNP}}$ and improve the ability to predict children at risk of myopia. Inclusion of gene–gene and/or gene–environment interaction effects would be expected to further improve predictive capacity. Notably, recent studies employing Bayesian or mixed models, similar in spirit to GCTA, have demonstrated the potential of genetic prediction from genome-wide common SNPs, which means that identifying individual risk SNPs through GWAS is not a prerequisite for effectively predicting disease [52].

Our $h^2_{\text{SNP}}$ estimate of about 35% provides an approximate upper limit for the capacity to predict the future refractive error of premyopic children using existing genotype platforms and imputation methods alone. Technical advances and larger sample sizes would be expected to raise $h^2_{\text{SNP}}$ and improve the ability to predict children at risk of myopia. Inclusion of gene–gene and/or gene–environment interaction effects would be expected to further improve predictive capacity. Notably, recent studies employing Bayesian or mixed models, similar in spirit to GCTA, have demonstrated the potential of genetic prediction from genome-wide common SNPs, which means that identifying individual risk SNPs through GWAS is not a prerequisite for effectively predicting disease [52].

Crude collection of information on two key environmental risk factors for myopia, time spent outdoors and time reading for pleasure, had limited capacity to predict or explain future refractive error, consistent with previous reports [53, 54]. However, our finding that these exposures explained <1% of the variance in refractive error likely underestimates the true effects for several reasons. First, the parental questionnaire used to collect the information had a categorical scoring scale rather than a continuous scale, which would have weakened statistical power to predict fine-scale differences. Second, exposure at only one particular age was examined, which may not have been representative of the true exposure. Third, for some participants the level of exposure may have been reported inaccurately. Finally, there may be nonlinear relationships between the time children engage in these activities and refractive error development, such as threshold effects, which would not have been taken into account when we assessed the variance they explained.

Our study had two main limitations. First, the assessment of refractive error at ALSPAC research clinics was done without cycloplegia, which is known to produce a subject-specific measurement error that can either underestimate or overestimate true refractive error. Lack of cycloplegia will therefore have reduced phenotypic accuracy, especially at younger ages and in hyperopes, and made our SNP-heritability estimates overly conservative (Appendix 1). Note that any systematic “over minus” component of the measurement error due to lack of cycloplegia during autorefraction would have had no impact, since heritability is a measure of explained variance, i.e., between-subject differences, whereas a systematic effect relates to the mean phenotype in the population. Second, the latest age at which ALSPAC participants’ refractive error was assessed was 15-years-old, which is before the age at which myopia typically stabilizes [55]. Therefore, further work will be needed to discover whether the set of genetic variants found here to influence refractive development over the 7- to 15-year-old age range continues to exert an effect into adulthood. Such information will be valuable for genetic prediction and for understanding the extent to which genetically-determined causal mechanisms are shared between early-onset and later-onset myopia.

Conclusion: The SNP heritability for refractive error was estimated to be approximately 35%. This value—calculated using only information about refractive error and an index of genetic similarity between each pair of individuals in the sample—provides the strongest evidence to date that refractive error has an important genetic component. In comparison, the two main environmental risk factors identified to date for myopia—time spent reading and time spent outdoors—together explained or predicted about 1% of the variance in refractive error. Nevertheless, the SNP-heritability figure is lower than that for axial length and corneal curvature ($h^2_{\text{SNP}}$ about 40–50%) calculated in the same population [29] suggesting that environmental effects exert more influence in determining refractive error than they do for the individual ocular components. The same set of genetic variants was found to influence refractive error at the various ages examined, rather than different sets of genetic variants acting at specific ages.

APPENDIX 1. THE LEVEL OF MEASUREMENT ERROR NOISE DUE TO LACK OF CYCLOPLEGIA DURING AUTOREFRACTION.

Autorefraction without cycloplegia is inaccurate in children. The level of inaccuracy depends on the type of instrument (e.g., open-field versus closed-field), the age and refractive error of the subjects, other subject-specific factors, and random sources of variation [33, 35-38]. In general, the degree of inaccuracy is worse in younger compared to older children, and typically causes hyperopia to be underestimated and myopia to be overestimated. A convenient and widely-used, albeit simplistic, method to quantify the inaccuracy of non-cycloplegic autorefraction is to assess the mean ± standard deviation of the difference between the non-cycloplegic measurement and a gold standard measurement (e.g., cycloplegic autorefraction or subjective refraction). This approach assumes that there is a systematic measurement error coupled with a random measurement error: it therefore ignores the effects of the child’s age and refractive error. Reports of non-cycloplegic autorefraction measurement...
error in population-representative samples vary widely, e.g., −0.36±0.41 D for 13-year-old children in the BATS & TEST cohorts [35] and −1.23±0.97 D for children aged 7–18 years-old in the Shunyi district RESC study [38], reflecting the age range and refractive error distribution of the respective samples. As reported previously [32] we compared non-cycloplegic autorefraction measurements obtained at ALSPAC research clinics against subjective refraction findings recorded by the participant’s own optometrist, for optometrist eye examinations performed within 6 months of the 15-year ALSPAC research clinic visit (n=346). The mean difference between the two measures was −0.22±0.84 D. Although this sample is not representative of the ALSPAC cohort, it does at least provide an indication of the degree of measurement error. New information provided by ALSPAC allowed us to exclude results for 13 of the above subjects whose clinic record indicated that an “error” occurred during autorefrac-
tion, and also to examine separately the findings for subjects who were confirmed as not wearing contact lenses during the autorefractive measurement (Figure 1 and Figure 2); typically, this ALSPAC record either confirmed that contact lenses were not worn, or was left incomplete by the clinical assessor. The measurement error of non-cycloplegic autorefrac-
tion better approximated a normal distribution in subjects confirmed as not wearing contact lenses (contact lens wear included: standard deviation of difference=0.60 D, n=108; contact lens wear not included: standard deviation of difference=0.95 D, n=225) suggesting that outlier autorefraction readings – likely due to a handful of subjects not removing their contact lenses before autorefration and thus errone-
uously appearing to be emmetropic by autorefration – had inflated the standard deviation of the subjective refraction versus non-cycloplegic autorefration comparison (Figure 2). The greater the degree of measurement error noise, the greater the underestimation of SNP-heritability (Figure 3), therefore we chose to use a normal distribution with a stan-
dard deviation of 0.50 D from which to simulate measurement error noise, to provide a conservative adjusted estimate of SNP-heritability. To access the data, click or select the words “Appendix 1.”

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REFERENCES

1. Hornbeak DM, Young TL. Myopia genetics: a review of current research and emerging trends. Curr Opin Ophthalmol 2009; 20:356–62. [PMID: 19587595].
2. Pan CW, Ramamurthy D, Saw SM. Worldwide prevalence and risk factors for myopia. Ophthalmic Physiol Opt 2012; 32:3-16. [PMID: 22150586].
3. Mitchell P, Hourihan F, Sandbach J, Wang JJ. The relationship between glaucoma and myopia - The Blue Mountains Eye Study. Ophthalmol 1999; 106:2010-5. [PMID: 10519600].
4. Wong TY, Klein BE, Klein R, Knudtson M, Lee KE. Refrac-
tive errors, intraocular pressure, and glaucoma in a white population. Ophthalmol 2003; 110:211-7. [PMID: 12511368].
5. Mitry D, Charteris DG, Fleck BW, Campbell H, Singh J. The epidemiology of rhegmatogenous retinal detachment - Geographic variation and clinical associations. Br J Ophthalmol 2010, 94:678-84. [PMID: 19515646].
6. Flitcroft DI. The complex interactions of retinal, optical and environmental factors in myopia aetiology. Prog Retin Eye Res 2012; 31:622-60. [PMID: 22772022].
7. Charman N. Myopia: its prevalence, origins and control. Ophthalmic Physiol Opt 2011; 31:3-6. [PMID: 21158881].
8. Feldkaemper M, Schaeffel F. An updated view on the role of dopamine in myopia. Exp Eye Res 2013; 114:106-19. [PMID: 23434455].
9. Flitcroft DI. Is myopia a failure of homeostasis? Exp Eye Res 2013; 114:16-24. [PMID: 23454097].
10. Hawthorne FA, Young TL. Genetic contributions to myopic refractive error: Insights from human studies and supporting evidence from animal models. Exp Eye Res 2013; 114:141-9. [PMID: 23379998].
11. Morgan IG, Ohno-Matsui K, Saw SM. Myopia. Lancet 2012; 379:1739-48. [PMID: 22559900].
12. Nickla DL. Ocular diurnal rhythms and eye growth regulation: Where we are 50 years after Lauber. Exp Eye Res 2013; 114:25-34. [PMID: 23298452].
13. Sivak J. The cause(s) of myopia and the efforts that have been made to prevent it. Clin Exp Optom 2012; 95:572-82. [PMID: 22845416].
14. Stone RA, Pardue MT, Iuvone PM, Khurana TS. Pharmacology of myopia and potential role for intrinsic retinal circadian rhythms. Exp Eye Res 2013; 114:35-47. [PMID: 23313151].
Molecular Vision 2015; 21:621-632 <http://www.molvis.org/molvis/v21/621>

15. Wojciechowski R. Nature and nurture: the complex genetics of myopia and refractive error. Clin Genet 2011; 79:301-20. [PMID: 21155761].

16. Tenesa A, Haley CS. The heritability of human disease: estimation, uses and abuses. Nat Rev Genet 2013; 14:139-49. [PMID: 23329114].

17. Falconer DS. Heritability. Introduction to quantitative genetics. 3rd ed. Harlow, UK: Longman; 1989.

18. Hemani G, Knott S, Haley C. An evolutionary perspective on epistasis and the missing heritability. PLoS Genet 2013; 9:e1003295-[PMID: 23509438].

19. Dudbridge F. Power and predictive accuracy of polygenic risk scores. PLoS Genet 2013; 9:e1003348-[PMID: 23555274].

20. Lopes MC, Andrew T, Carbonarof F, Spector T, Hammond CJ. Estimating heritability and shared environmental effects for refractive error in twin and family studies. Invest Ophthalmol Vis Sci 2009; 50:126-31. [PMID: 18757006].

21. Dirani M, Chamberlain M, Shekar SN, Islam AF, Garoufalis P, Chen CY, Guymer RH, Baird PN. Heritability of refractive error and ocular biometrics: The genes in myopia (GEM) twin study. Invest Ophthalmol Vis Sci 2006; 47:4756-61. [PMID: 17065484].

22. Lyhne N, Stolje AK, Kyvik KO, Green A. The importance of genes and environment for ocular refraction and its determiners: a population study based on twins 20–45 year old twins. Br J Ophthalmol 2001; 85:1470-6. [PMID: 11734523].

23. Klein AP, Sukittipat B, Duggal P, Lee KE, Klein R, Bailey-Wilson JE, Klein BE. Heritability analysis of spherical equivalent, axial length, corneal curvature, and anterior chamber depth in the Beaver Dam Eye Study. Arch Ophthalmol 2009; 127:649-55. [PMID: 19433716].

24. Chen CY, Seurr J, Stankovich J, Garoufalis P, Dirani M, Pertile KK, Richardson AJ, Mitchell P, Baird PN. Heritability and shared environment estimates for myopia and associated ocular biometric traits: the Genes in Myopia (GEM) family study. Hum Genet 2007; 121:511-20. [PMID: 17205325].

25. Sanfilippo PG, Hewitt AW, Hammond CJ, Mackey DA. The heritability of ocular traits. Surv Ophthalmol 2010; 55:561-83. [PMID: 20851442].

26. Visscher PM, Medland SE, Ferreira MA, Morley KL, Zhu G, Cornes BK, Montgomery GW, Martin NG. Assumption-free estimation of heritability from genome-wide identity-by-descent sharing between full siblings. PLoS Genet 2006; 2:e41-25. [PMID: 16565746].

27. Yang J, Benyamin B, McEvoy BP, Gordon S, Henders AK, Nyholt DR, Madden PA, Heath AC, Martin NG, Montgomery GW, Goddard V, Visscher PM. Common SNPs explain a large proportion of the heritability for human height. Nat Genet 2010; 42:565-9. [PMID: 20562875].

28. Yang J, Manolio TA, Pasquale LR, Boerwinkle E, Caporaso N, Cunningham JM, de Andrade M, Feenstra B, Feingold E, Hayes MG, Hill WG, Landi MT, Alonso A, Lettge G, Lin P, Ling H, Lowe W, Mathias RA, Melbye M, Pugh E, Cornelis MC, Weir BS, Goddard ME, Visscher PM. Genome partitioning of genetic variation for complex traits using common SNPs. Nat Genet 2011; 43:519-25. [PMID: 21552263].

29. Guggenheim JA, Zhou X, Evans DM, Timpson NJ, McMahon G, Kemp JP, St Pourcain B, Northstone K, Ring SM, Fan Q, Wong T-Y, Cheng CY, Khor CC, Aung T, Saw SM, Williams C. Coordinated genetic scaling of the human eye: Shared determination of axial eye length and corneal curvature. Invest Ophthalmol Vis Sci 2013; 54:1715-21. [PMID: 23385790].

30. Boyd A, Golding J, Macleod J, Lawlor DA, Fraser A, Henderson J, Molloy L, Ness A, Ring S, Davey Smith G. Cohort Profile: The ‘Children of the 90s’–the index offspring of the Avon Longitudinal Study of Parents and Children. Int J Epidemiol 2013; 42:111-27. [PMID: 22507743].

31. Guggenheim JA, Northstone K, McMahon G, Ness AR, Deere K, Mattocks C, St Pourcain B, Williams C. Time outdoors and physical activity as predictors of incident myopia in childhood: A prospective cohort study. Invest Ophthalmol Vis Sci 2012; 53:2856-65. [PMID: 22491403].

32. Northstone K, Guggenheim JA, Howe LD, Tilling K, Paternoster L, Kemp JP, McMahon G, Williams C. Body stature growth trajectory during childhood and the development of myopia. Ophthalmol 2013; 120:1064-73. .

33. Williams C, Lumb R, Harvey I, Sparrow JM. Screening for refractive errors with the Topcon PR2000 Pediatric Refractometer. Invest Ophthalmol Vis Sci 2000; 41:1031-7. [PMID: 10752938].

34. Bartlett MS. The use of transformations. Biometrics 1947; 3:39-52. [PMID: 20240416].

35. Sanfilippo PG, Chu BS, Bigault O, Kearns LS, Boon MY, Young TL, Hammond CJ, Hewitt AW, Mackey DA. What is the appropriate age cut-off for cycloplegia in refraction? Acta Ophthalmol (Copenh) 2014; 92:e458-62. [PMID: 24641244].

36. Suryakumar R, Bobier WR. The Manifestation of Noncycloplegic Refractive State in Pre-School Children is Dependent on Autorefractor Design. Optom Vis Sci 2003; 80:578-86. [PMID: 12917577].

37. Williams C, Miller L, Northstone K, Sparrow JM. The use of non-cycloplegic autorefract data in general studies of children's development. Br J Ophthalmol 2008; 92:723-4. [PMID: 18441189].

38. Zhao J, Mao J, Luo R, Li F, Pokharel GP, Ellwein LB. Accuracy of noncycloplegic autorefraction in school-age children in China. Optom Vis Sci 2004; 81:49-55. [PMID: 14747761].

39. Cheng C-Y, Schache M, Ikrak MK, Young Terri L, Guggenheim Jeremy A, Vitart V, MacGregor S, Verhoeven Virginie JM, Barathh Veluchamy A, Liao J, Hysi Pirro G, Bailey-Wilson Joan E, St. Pourcain B, Kemp John P, McMahon G, Timpson Nicolos J, Evans David M, Montgomery Grant W, Mishra A, Wang Ya X, Wang Jie J, Rochtchina E, Polasek O, Wright Alan F, Amin N, van Leeuwen Elisabeth M, Wilson James F, Penrell Craig E, van Duijn Cornelia M, de Jong Paulus T, Vingerling Johannes R, Zhou X, Chen P, Li R, Tay W-T, Zheng Y, Chew M, Burdon KP, Craig JE,
40. Li Q, Wojciechowski R, Simpson C, Hysi P, Verhoeven VM, Ilkram M, Höhn R, Vitari V, Hewitt A, Oxley K, Mäkelä K-M, MacGregor S, Pirastu M, Fan Q, Cheng C-Y, St Pourcain B, McMahon G, Kemp J, Northstone K, Rahi J, Cumberland P, Martin N, Sanfilippo P, Lu Y, Wang Y, Yang C, Polasek O, Campbell H, Bencic G, Wright A, Wedenoja J, Zeller T, Schillert A, Mirshahi A, Lackner K, Ye P, Yap MH, Ried J, Gieger C, Murgia F, Wilson J, Fleck B, Yasz E, Vinger B, Hofman A, Uitterlinden A, van Duijn C, Aung T, van Duijn C, Mitchell PA, Oosterud BA, Teo YY, Hammond CJ, Stambolian D, Mackay DA, Klaver CCW, Wong T-Y, Saw S-M, Baird PN. Nine loci for ocular axial length identified through genome-wide association studies, including shared loci with refractive error. Am J Hum Genet 2013; 93:264-77. [PMID: 24144296].

41. Williams AL, Patterson N, Glessner J, Hakonarson H, Reich D. Phasing of many thousands of genotyped samples. Am J Hum Genet 2012; 91:238-51. [PMID: 22883141].

42. Howie B, Fuchsberger C, Stephens M, Marchini J, Abecasis GR. Fast and accurate genotype imputation in genome-wide association studies through pre-phasing. Nat Genet 2012; 44:955-9. [PMID: 22820512].

43. Fraser A, Macdonald-Wallis C, Tilling K, Boyd A, Golding J, Davey Smith G, Henderson J, Macleod J, Molloy L, Ness A, Ring S, Nemoto SM, Lawlor DA. Cohort Profile: The Avon Longitudinal Study of Parents and Children: ALSPAC mothers cohort. Int J Epidemiol 2013; 42:97-110. [PMID: 22507742].

44. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. Gigascience 2015; 4:7-[PMID: 25722852].

45. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: A tool for genome-wide complex trait analysis. Am J Hum Genet 2011; 88:76-82. [PMID: 2167468].

46. Speed D, Hennam G, Johnson Michael R, Balding David J. Improved heritability estimation from genome-wide SNPs. Am J Hum Genet 2012; 91:1011-21. [PMID: 23217325].

47. Speed D, Hennam G, Johnson MR, Balding DJ. Response to Lee et al.: SNP-based heritability analysis with dense data. Am J Hum Genet 2013; 93:1155-7. [PMID: 24314551].

48. Lee SH, Yang J, Chen GB, Ripke S, Stahl EA, Hultman CM, Sklar P, Visscher PM, Sullivan PF, Goddard ME, Wray NR. Estimation of SNP heritability from dense genotype data. Am J Hum Genet 2013; 93:1151-5. [PMID: 24314550].

49. Zhu Z, Bakshi A, Vinkhuyzen AA, Hennam G, Lee SH, Nolte IM, van Vliet-Ostaptchouk JV, Snieder H, Esko T, Milani L, Magni R, Metspalu A, Hill WG, Weir BS, Goddard ME, Visscher PM, Yang J. Dominance genetic variation contributes little to the missing heritability for human complex traits. Am J Hum Genet 2015; 96:377-85. [PMID: 25683123].

50. Wood AR, Esko T, Yang J, Vedantam S, Pers TH, Gustafsson S, Chu AY, Estrada K, Luan J, Kutalik Z, Amin N, Buchkovich ML, Croteau-Chonka DC, Day FR, Duan Y, Fall T, Fehrman R, Ferreira T, Jackson AU, Karjalainen J, Lo KS, Locke AE, Magi R, Mihaylov I, Porcu E, Randall JC, Scherag A, Vinkhuyzen AAE, Westra H-J, Winkler TW, Workalemahu T, Zhao JH, Absher D, Albricht E, Anderson D, Baron J, Beekman M, Demirkiran A, Ehret GB, Feenstra B, Feitosa MF, Fischer K, Fraser RM, Goel A, Gong J, Justice AE, Kanoni S, Kleber ME, Kristiansson K, Lim U, Lotay V, Lui JC, Mangino M, Leach IM, Medina-Gomez C, Nalls MA, Nyholt DR, Palmer CD, Pasko D, Peclihavis S, Prokopenko I, Ried JS, Ripke S, Shungin D, Stancakova A, Strawbridge RJ, Sung YJ, Tanaka T, Teumer A, Trompet S, van der Laan SW, van Setten J, van Vliet-Ostaptchouk JV, Wang Z, Yengo L, Zhang W, Afzal U, Arbnow J, Arscott GM, Bandinelli S, Barrett A, Bellis C, Bennett AJ, Berne C, Blusher M, Bolton JL, Bottcher Y, Boyd HA, Bruijnenberg M, Buckley BM, Buyskes S, Caspersen IH, Chines PS, Clarke R, Claudi-Boehm S, Cooper M, Daw EW, De Jong PA, Deelen J, Delgado G, Denny JC, Domnukse-Rutten R, Dimitriou M, Dooley ASF, Dorr M, Eklund N, Eury E, Folkesens L, Garcia ME, Geller F, Giedraitis V, Go AS, Graff H, Grammer TB, Graszler J, Gronberg H, de Groot LCPGM, Groves CJ, Haessler J, Hall P, Haller T, Hallmans G, Hannemann A, Hartman CA, Hassinen M, Hayward C, Heard-Costa NL, Helmer Q, Hennam G, Henders AKH, Hillege HL, Hlatky MA, Hoffmann W, Hoffmann P, Holmen O, Huijvink-Duistermaat JJ, Illig T, Isaacs A, James AL, Jeff J, Johansen B, Johansson A, Julies J, Juliusdottir T, Junttila J, Kho AN, Kinnunen L, Klopp N, Kocher T, Kratzer W, Lichtner P, Lind L, Lindstrom R, Lobbens S, Lorentzon M, Lu Y, Lyssenko V, Magnusson BE, Hessman GR, Mahajan A, Maillard M, McArdle WL, McKenzie CA, McLauchlan S, McLaren PJ, Menni C, Merger S, Milani L, Moayyeri A, Monda KL, Morken MA, Muller G, Muller-Nurasyid M, Musk AW, Nairi N, Nauck M, Nolte IM, Nothen MM, Oozerse B, Padic L, Rayner WN, Renstrom F, Robertson NR, Rose LM, Roussel R, Sanna S, Scharnagl H,
Molecular Vision 2015; 21:621-632 <http://www.molvis.org/molvis/v21/621>

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Scholtens S, Schumacher FR, Schunkert H, Scott RA, Sehmi J, Seufferlein T, Shi J, Silventoinen K, Smit JH, Smith AV, Smolonska J, Stanton AV, Stirrups K, Stott DJ, Stringham HM, Sundstrom J, Swertz MA, Syvanen A-C, Tayo BO, Thorleifsson G, Tyrer JP, van Dijk S, van Schoor NM, van der Velde N, van Heemst D, van Oort FVA, Vermeulen SH, Verweij N, Vonk JM, Waite LL, Waldenberger MG, Wannenaar R, Wilkens LR, Willenborg C, Wilsaard T, Wojczynski MK, Wong A, Wright AF, Zhang Q, Arveiler D, Bakker SJL, Beilby J, Bergman RN, Bergmann S, Biffar R, Blangero J, Boomsma DI, Bornstein SR, Bovet P, Brambilla P, Brown MJ, Campbell H, Caulfield MJ, Chakravarti A, Collins R, Collins FS, Crawford FD, Cupples LA, Danesh J, de Faire U, den Ruijter HM, Erbel R, Erdmann J, Eriksson JG, Farrall M, Ferrarini E, Ferrieres J, Ford I, Forouhi NG, Forrester TJ, Gaedkevoort DI, Gieger C, Golay A, Gottesman O, Gudnason V, Gysemans C, Haas DW, Hall AS, Harris TB, Hattersley AT, Heath AC, Hicks AA, Hindorff LA, Hingorani AD, Hofman A, Hovingh GK, Humphries SE, Hunt SC, Hypponen E, Jacobs KB, Jarvelin MR, Jousilahti P, Kaprio J, Kastelein JJP, Kayser M, Kee F, Keinanen-Kiukaanniemi SM, Kiekenbaum LA, Kooperberg C, Koskinen S, Kovacs P, Kraja AT, Kruisjistederck L, Laukka TA, Lu, Laidenberg C, Le Marchand L, Lettmann R, Lupoli S, Madden PA, Mannisto S, Manunta P, Marme CA, Matisse TC, McKnight B, Meitinger T, Moll FL, Montgomery GW, Morris AD, Morris AP, Murray JC, Nelson S, Ochsod L, Oldenhake J, Ong KK, Ouwehand WH, Pasterkamp G, Peters A, Pratt P, Price JF, Qiu L, Raitakari OT, Rankinen T, Rao DC, Rice TK, Ritchie M, Rudan I, Salomon V, Samani NJ, Sarakis J, Sarzynski MA, Schwarz PEH, Sebert S, Sever P, Shejner AR, Sinisalo J, Steinthorsdottir V, Stok RP, Tartif J-C, Tonjes A, Tremblay RT, Tremoli E, Virtamo J, Vohl M-C, The Electronic Medical R, Genomics C, The MC, The PC, The LifeLines Cohort S, Amouelj P, Asselbergs FW, Assimes TL, Bouchard M, Boehm BO, Boerwinkle E, Bottiger EP, Bouchard C, Cauchi S, Chambers JC, Chanock SJ, Cooper RS, de Bakker PIW, Deleusse G, Ferrucci L, Franks PW, Fregul P, Groop LC, Haiman CA, Hamsten A, Hayes MG, Hui J, Hunter DJ, Hveem K, Jukema JW, Kaplan RC, Kivimaki M, Kuh D, Laakso M, Liu Y, Martin NG, Marz W, Melbye M, Moebus S, Munroe PB, Njolstad I, Oostra BA, Palmer CNA, Pedersen NL, Perola M, Perussi L, Peters U, Powell JE, Power C, Quertermous T, Rauramaa R, Reinmaa E, Rickard PM, Rivadeneira F, Rotter JI, Saaristo TE, Saleheen D, Schlessinger D, Slagboom PE, Snijder E, Spector TD, Strachuk K, Stumvoll M, Tuomilehto J, Uutisluoto M, van der Harst P, Volzke H, Walker M, wareham NH, Watkins H, Wichmann HE, Wilson JT, Zanen P, Deloukas P, Heid IM, Lindgren CM, Mohlke KL, Speliotes EK, Thorsteinsdottir U, Barroso I, Fox CS, North KE, Strachan PD, Beckmann JS, Berndt SI, Boehnke M, Borecki IB, McCarthy MI, Metspalu A, Stefansson K, Uitterlinden AG, van Duijn CM, Franke L, Will CJ, Price AL, Lettre G, Loos RJF, Weedon MN, Ingelsson E, O’Connell JR, Abecasis GR, Chasan DI, Goddard ME, Visscher PM, Hirschhorn JN, Frayling TM. Defining the role of common variation in the genomic and biological architecture of adult human height. Nat Genet 2014; 46:1173-86. [PMID: 25282103].

51. McMahon G. PhD thesis: The genetics and epidemiology of myopia in the ALSPAC cohort. Cardiff: University Press; 2010.

52. Golan D, Rosset S. Effective genetic-risk prediction using mixed models. Am J Hum Genet 2014; 95:383-93. [PMID: 25279982].

53. Jones-Jordan LA, Sinnott LT, Mann RE, Cotter S, Kleinsein RN, Mutti DO, Twelker D, Zadnik K. Early childhood refractive error and parental history of myopia as predictors of myopia. Invest Ophthalmol Vis Sci 2010; 51:115-21. [PMID: 19737876].

54. Jones-Jordan LA, Sinnott LT, Graham ND, Cotter SA, Kleinsein RN, Mann RE, Mutti DO, Twelker JD, Zadnik K. CSG. The contributions of near work and outdoor activity to the correlation between siblings in the Collaborative Longitudinal Evaluation of Ethnicity and Refractive Error (CLEERE) Study. Invest Ophthalmol Vis Sci 2014; 55:6333-9. [PMID: 25205866].

55. The Comet Group. Myopia stabilization and associated factors in Correction of Myopia Evaluation Trial (COMET) participants. Invest Ophthalmol Vis Sci 2013; 54:7871-7884. [PMID: 24159085].