Polymorphisms within the Promoter Region of the Gamma Interferon (IFN-γ) Receptor1 Gene are Associated with the Susceptibility to Chronic HBV Infection in an Iranian Population

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ARTICLE INFO

Article type: Research Article

Article history:
Received: 17 Jul 2012
Revised: 15 Aug 2012
Accepted: 15 Sep 2012

Keywords:
Hepatitis B Virus
Single Nucleotide Polymorphism
Interferon Gamma Receptor Polymorphism, Restriction Fragment Length

ABSTRACT

Background: chronic hepatitis B virus (HBV) infection is a multifactorial disease that can result in serious clinical complications. Host genetic background especially the genes that encode immunologic factors like INF-γ and its receptor (IFN-γ R) are critical in the pathogenesis of infection.

Objectives: The current study aimed to investigate the association between two single nucleotide polymorphisms (SNPs) at positions -611 and -56 within the promoter region of gamma interferon receptor1 gene (IFN-γ R1) and chronic HBV infection.

Materials and Methods: Genomic DNA from peripheral blood samples of 200 chronically HBV infected patients and 200 healthy blood donors, as controls, were collected and genomic DNA was extracted by phenol-chloroform method and DNA analysis genotype identification was performed by PCR-RFLP.

Results: The results indicated that both SNP’s frequency had a significant difference in the patient and control groups. At position -56, TT genotype was associated with patient group and P value was 0.002 and at position -611, GG genotype was further observed in control group and P value was 0.006.

Conclusions: Presence of G allele at position -611 within promoter of IFN-γ R1 gene in the enrolled population for the study was related to lower risk of disease, and presence of T allele at position -56 was also related to susceptibility to chronic HBV infection. Men had higher frequency of chronic HBV infection, which might be the result of high risk behavior.

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DOI: 10.5812/hepatmon.7283
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1. Background

Chronic HBV infection is one of the most common viral infections worldwide, with about 400 million chronic carriers living in the world (1). More than 90% of HBV infections in adults are self-limited within six months with or without clinical symptoms (acute infection), whereas 5 to 10% of infections progress towards chronic infection in which the virus is not cleared from the body (2). Chronic infection can lead to serious clinical consequences such as liver cirrhosis and hepatocellular carcinoma (HCC) (3). The host genetic background can play a critical role in the pathogenesis of disease and single nucleotide polymorphism (SNPs) may be effective on the clinical heterogeneity of chronic infection (4). The immune system’s response to primary HBV infection determines the clinical course of HBV infection. The persistent HBV infection is considered a multifactorial disorder, which various factors such as the host genetic background, viral and environmental factors are involved in its pathogenesis (5). IFN-γ is a cytokine that plays a crucial role in defense against intracellular pathogens (especially virus) and the stimulation of an immune mediated inflammatory response (6). The importance of IFN-γ in the clearance of HBV has been recently demonstrated. During an acute self-limited HBV infection, a high level of IFN-γ is secreted by T lymphocytes, which have significant roles on HBV clearance (7). IFN-γ can inhibit the replication of HBV and directly reduces the viral load (8-10). Studies conducted on laboratory animals especially in the transgenic mice have shown that the expression of cytokines can cause decrease in HBV replication, without lethal effect on hepatocytes (7). Nucleotide variations in the regulatory regions of coding cytokine genes and their receptors can lead to modulation in the cytokine expression, a large number of evidences indicate that SNPs within the binding site of transcription factors may cause decrease in the promoter function of a cytokine/receptor complex and thus can affect the symptoms of a disease (7, 8). The host genetic backgrounds, especially (SNPs) can be a marker of the clinical heterogeneity of HBV infection (17, 19). IFN-γ has a heterodimeric receptor that is made up of two chains (IFN-γR1 and IFN-γR2), that each mediates a specific function; IFN-γR1 is involved in binding to the ligand (IFN-γ) and IFN-γR2 plays a role in the signaling pathway (6). The binding of homodimeric IFN-γ to IFN-γR1 causes oligomerization of the two receptor subunits. Each receptor subunit is associated with a distinct Janus kinase (Jak), which is activated by reciprocal transphosphorylation upon oligomerization of IFN-γR1 and IFN-γR2. Then, Jak activation causes phosphorylation of IFN-γR1. A latent transcription factor, Stat1, binds to the phosphorylated IFN-γR1, undergoes tyrosine phosphorylation, and forms homodimers that translocate to the nucleus and regulate transcription of inducible genes by IFN-γ (14). Previous studies have shown that SNPs in the IFN-γR and its receptor can affect the pathogenesis of chronic HBV infection.

2. Objectives

The current study aimed to investigate the association between two single nucleotide polymorphisms (SNPs) at positions -611 and -56 within the promoter region of gamma interferon receptor1 gene (IFN-γR1) and chronic HBV infection.

3. Materials and Methods

3.1. Study Groups

The study included 200 chronic patients positive for HBsAg, who were from different provinces referring to Taleghani hospital during 2010-2011, with symptoms of impaired liver function tests (transaminases twice the level of normal for at least a 6 month period), detectable IgG anti-HBc and/or sonographic, and clinical results compatible with chronic liver disease. 200 blood donors, who were negative for both anti-Hbc and HBsAg, were studied as control subjects (Table 1). All individuals who enrolled for the study were from different regions of Iran, that researchers obtained their ethical approvals. Serum HBsAg, HBeAg, anti-HBs, anti-HBe and anti-HBc were assayed by ELISA kits (DRG International Inc., USA), according to the manufacturer’s instruction. None of patients had a history of intravenous drug abuse or extreme alcohol consumption, and all were negative for anti-HCV and anti-HIV. Patients with other liver diseases (hemochromatosis, Wilson’s disease, α-1 antitrypsin deficiency)
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or history of receiving anti-viral or immunosuppressive drugs were excluded. The blood samples were obtained from each participant and DNA was extracted from buffy coat using the phenol-chloroform method (18).

3.2. Detection of IFN-γR1 Gene Polymorphism

The regions of IFN-γR gene containing two single nucleotide polymorphisms (SNPs) at positions -56 and -611 were amplified by PCR. The PCR products for analysis were digested by the Hpy188I restriction enzyme (New England Biolab) for -611 SNP and the AfeI restriction enzyme (New England Biolab). The PCR conditions were optimized for -56 SNP as follows: 95°C for 5 min; 33 cycles of 95°C for 30 s, 59°C for 30 s, and 72°C for 30 s; and a final extension at 72°C for 10 min; and for -611 SNP as follows: 94°C for 5 min; 35 cycles of 95°C for 35 s, 61.7°C for 35 s, and 72°C for 35 s; and a final extension at 72°C for 10 min. The PCR products were run on a 1% agarose gel and stained with ethidium bromide for visualization under UV light. The RFLP products for -56 promoter SNPs were visualized on 2% agarose gel. The products for -611 promoter SNPs were visualized on 18% polyacrylamide gel electrophoresis. To confirm the RFLP genotyping results, 10% of samples were randomly sequenced via direct sequencing by ABI genetic analyzer 3130xl system.

3.3. Statistical Methods

The data were analyzed using chi-square test according to the Hardy–Weinberg equilibrium (HWE) and independent sample t test, at P < 0.05 significance level. SPSS software version 16.0 was employed to calculate Odds Ratios (ORs) and 95% confidence intervals (95% CI) by binary logistic regression.

4. Results

The polymorphisms of IFN-γR1 promoter were identified in subjects under study (Figure). They were C to T transition at -56 and G to A at -611 relative to promoter site. There were no significant relationships between age and gender of case and control groups and the status of disease (P = 0.380). Mean age of patients and controls were 45.71 ± 15.37 and 43.99 ± 16.17 respectively. As indicated in Table 3, irrespective of age, and gender factors, a case-control analysis of the population revealed a significant difference in both of the site polymorphisms. Logistic regression analyses revealed that genotype frequency of -56 TT was significantly higher in cases than in controls with (P < 0.002, OR = 0.469, 95% CI = 0.292-0.754) and genotype frequency of -611GG was notably higher in controls than in cases with (P < 0.006, OR = 0.164, 95% CI = 0.046-0.580). Therefore a significant association be-

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Table 1. Clinical Data of Control and Patient Groups

|                | CHP       | CHS       | P-value | OR (95% CI) | Adj OR (95% CI) | Total, No. |
|----------------|-----------|-----------|---------|-------------|----------------|------------|
| Male/female, No. | 130/70    | 108/92    |         |             |                |            |
| HBsAg + −       |           |           |         |             |                |            |
| Anti-HBsAg − +  |           |           |         |             |                |            |
| HBeAg + −       |           |           |         |             |                |            |
| Anti-HBeAg − −  |           |           |         |             |                |            |
| Anti-HBcAg + −  |           |           |         |             |                |            |
| ALT 55.8 ± 62.2 | 24.6 ± 10.4 |          |         |             |                |            |

Abbreviations: ALT, alanine aminotransferase; CHP, chronic hepatitis B patient; HBsAg, hepatitis B surface antigen; HBeAg, hepatitis B antigen; HBcAg, hepatitis B core antigen; HCS, healthy controls.

Table 2. Primers and Restriction Enzymes for Genotyping

| SNP, position | PCR primer sequence (5′- 3′) | Restriction Enzyme | Allele Phenotype |
|---------------|------------------------------|--------------------|-----------------|
| -611A/G(p)    | F:CTCTTCATGAGAGGCTGTCT R:TAACCTTGGAGTTCACCTGG | Hpy188I | A: 260bp G:240 + 20bp |
| -56T/C(P)     | F:TGCATGACAAGGGGTAGGAG R:CAACCAGGTGAAGTCCAAGAG | AfeI | T: 430bp C:339 + 91bp |

Table 3. Genotype Distributions of SNPs in Study Participants

| Polymorphism | Genotype | CHP, No. (%) | HCS, No. (%) | P-value | OR (95% CI) | Adj OR (95% CI) | Total, No. |
|--------------|----------|--------------|--------------|---------|-------------|----------------|------------|
| 56           | TT       | 74 (37)      | 42 (20.9)    | 1.00    | 1.00        | 1.00           | 116        |
|               | TC       | 86 (43)      | 103 (51.7)   | 0.469   | 0.292-0.754 | 0.486          | 190        |
|               | CC       | 40 (20)      | 55 (27.4)    | 0.413   | 0.237-0.720 | 0.421          | 95         |
| 611          | AA       | 82 (41)      | 76 (37.8)    | 1.00    | 1.00        | 1.00           | 158        |
|               | AG       | 115 (57.5)   | 117 (53.7)   | 0.981   | 0.656-1.484 | 1.00           | 233        |
|               | GG       | 3 (1.5)      | 17 (8.5)     | 1.00    | 0.046-0.580 | 0.177          | 20         |

Abbreviations: CHP, chronic hepatitis B patients; HCS, healthy controls; OR, odd’s ratio.
The digestion pattern of Hpy188I restriction enzyme on 18% poly-
donors. Two polymorphisms of IFN-γ were identified, including two SNPs at position -56 and -611. The genetic variations within promoter of IFN-γ may affect the clinical course of HBV infection. The results of the study suggest that the mutant allele T is a functional SNP and is associated with susceptibility to chronic HBV infection (similar to the current study population at -56 T/C SNP) but no polymorphism have been found at position -611 (19). Genetic polymorphism in the IFN-γ R1 promoter was linked to atopic cataracts (13). The genetic variations within promoter of IFN-γ among different ethnicities and various cell lines have been studied, a deletion/insertion polymorphism at promoter locus -470, found in Africans, is absent in Europeans and Asians (19). The functional effects of -56T/C SNP have also been investigated. The presence of C allele exhibited a decreased transcriptional activity in reporter gene expression in the human B cell line and myeloid cell lines whereas at -56T/C SNP mutant allele T in the hHepG2 cell line illustrated a reduction in transcription and wild allele C showed a high level of transcription (15).

It is plausible that chronic HBV infection is a multifactorial disease. The host immunological factors that can be effective in viral persistence are not well understood. Rapid advances in human genomics offer great opportunities to associate genetic variations with risks of diseases. In conclusion, the existence of two polymorphisms within the IFN-γ R1 promoter in an Iranian population were detected. According to the results of the current study, especially at position -56 SNP, it seems that allele T can be considered as a risk factor towards susceptibility to chronic infection in the population under study; and genetic variations in promoter of IFN-γ R1gene may affect the clinical course of HBV Infection.

Acknowledgments
The authors would like to thank all colleagues from the liver group of the RCGLD, especially Azar Sanati, Shohreh Almasi, Parvaneh Mohammadi, Mahsa Khanyaghma and Hanieh Mirtalebi. All patients and healthy blood donors for providing blood samples. The authors are also grateful to Mr. M. Mahlouji for reading the manuscript critically. This work was supported by a grant from Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran and Research Center of Gastroenterology and Liver Diseases, Taleghani Hospital, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

Authors’ Contribution
Sayyad Khanizadeh conducted experimental studies and drafted manuscript as his MSc thesis. Mehrdad Ravanshad (supervisor) and Seyed Reza Mohebbi (advisor) designed the study concepts and prepared the manuscript. Hamed Naghoosi, Mohamad Abrahim Tahaei, Seyed Dawood Mousavi Nasab, Sara Romani, Pedram Azimzadeh, Azar Sanati and Mohammad Reza Zali were involved in sample and data collection and analysis.
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Financial Disclosure
None declared.

Funding/Support
None declared.

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