Research Article

Immunoinformatic Analysis to Identify Proteins to Be Used as Potential Targets to Control Bovine Anaplasmosis

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Received 10 April 2020; Revised 6 July 2020; Accepted 3 August 2020; Published 27 August 2020

1. Introduction

Ticks and tick-borne pathogens constitute a major challenge for the cattle industry due to their impact on production losses [1]. Bovine anaplasmosis is a disease caused by Anaplasma marginale, an important tick-transmitted, intraerythrocytic Gram-negative bacterium, that is endemic in Mexico [2]. The disease has a worldwide distribution and causes serious economic losses, particularly in beef cattle, as they are more exposed to A. marginale transmitted by the tick-vector Rhipicephalus microplus [3]. The control of bovine anaplasmosis does not only depend on controlling the pathogen itself, but also the vector which transmits it as they have coevolved with the host [4]. So far, there are no commercial vaccines against bovine anaplasmosis, and those that have been prepared from freed bacteria (initial bodies) have a limited use due to a wide antigenic diversity of the pathogen, while those prepared from live attenuated organisms carry the risk for the cotransmission of other blood-borne pathogens [2, 5].
an integral view of the tick-pathogen relationship should be considered to propose not only effective control measures but also successful diagnostic and vaccination methods as those recently reported [8, 9, 10].

Recently, the Next-Generation Sequencing (NGS) in veterinary medicine has revealed the potential to design primary diagnostic, control, and prevention methods [11]. The genomes of *A. marginale* and *R. microplus* have been sequenced and published [12, 13, 14], and many detailed studies have been performed with the Major Surface Proteins (MSPs) of *A. marginale*, whether as vaccine prospects or diagnostic targets. In the first instance, individual or conglomerate proteins have not been very successful in inducing protective solid immunity; as diagnostic targets, a recombinant Msp5 has been successfully used for molecular detection and the serological ELISA test [15, 16].

In contrast, in *R. microplus*, several proteins have been studied as vaccine candidates [17]. The use of Bm86 in the vaccines TickGard [18] (no longer commercially available) and GAVAC [19] reduced parasitism poorly, and the immunologic memory induced was short-lived [20, 21]. The recent approaches in tick research have facilitated a different design of vaccines based on genomics, proteomics (hemo-lymph) [22], and transcriptomics (sialomes) studies [23].

Despite the amount of data produced for both *A. marginale* and *R. microplus*, a wide repertoire of proteins still remains to be studied, as well as the possibility to find protective antigens to bovine anaplasmosis and its vector *R. microplus*.

In this work, we carried out an immununomic analysis of the genomes of all reported Mexican *A. marginale* strains with the aim to find previously nonreported potential vaccine candidates. Derived from a transcriptomic analysis of *R. microplus*, we found a protein involved in immunological processes whose absence makes ticks susceptible to acaropathogenic organisms used in biological control.

2. Materials and Methods

2.1. Genomic Analysis and Protein Selection. The amino acid sequences for the reported genomes of the seven Mexican strains were searched online for outer membrane and membrane proteins. The sequences were downloaded and grouped in a list, with the exception of the MSPs and the type four secretion system (VirB) sequences.

The sequence of the peroxinectin (*pxn*) gene was searched and retrieved from the transcriptome (unpublished data) of different stages of *R. microplus* (Arthropodology Unit, INIFAP, Mexico). The coding sequence of the *pxn* gene was translated to a protein sequence with the Translate Tool from Expasy.

This study was approved by the Animal Experimentation and Ethics Committee of the National Center for Disciplinary Research in Animal Health and Safety (CENID-SAI, Mexico) which is a branch of the INIFAP. The study took ethical and methodological aspects into considerations in accordance with the Mexican regulations on use, housing, and transportation of experimental animals (NOM-062-ZOO-1999 and NOM-051-ZOO-1995).

2.2. Prediction of Antigenic Proteins. To find the highest antigenic protein, selected protein sequences of *A. marginale* and the protein sequence of the peroxinectin of *R. microplus* were submitted to Vaxijen v2.0 server (http://www.ddgpharmfac.net/vaxijen/Vaxijen/Vaxijen.html) with default parameters.

2.3. Prediction of Subcellular Localization and Stability of the Proteins. Predicted antigenic proteins of *A. marginale* and the peroxinectin protein of *R. microplus* were submitted to different servers to predict their subcellular localization. We used the secondary structure and subcellular prediction server Constrained Consensus TOpology (CCTOP; http://cttop.enzim.ttk.mta.hu). The proteins of interest were also submitted to the CELLO v.2.5 server (http://cello.life.nctu.edu.tw/). To analyze the stability and secondary structure of the target proteins sequence, ProtParam server (http://web.expasy.org/protparam/) and SOPMA server (https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page = /NPSA/npsa_sopma.html) were used with default parameters.

2.4. Linear B-Cell Epitope Prediction. B-cell epitopes can be categorized as linear (continuous) and conformational (discontinuous) based on their spatial structure. We used, at least, three online available tools for the prediction of linear epitopes: ABCpred (http://crdd.osdd.net/raghava/abcpred/) was set at sequences for relevant linear B-cell epitopes at 18-mers with a threshold of 0.85 and overlapping filters “on”, BCEpred (http://crdd.osdd.net/raghava/bcepred/) predicts epitopes with an 58.7% accuracy using flexibility, hydrophilicity, polarity, and surface properties combined at a threshold of 2.38, and BepiPred 2.0 (http://www.cbs.dtu.dk/services/BepiPred/) predicts B-cell epitopes based on epitope and nonepitope amino acids determined from crystal structures. Further analysis was performed with IED Antibody Epitope Prediction (Kolaskar and Tongaonkar, 1990).

2.5. Three-Dimensional Modelling. The three-dimensional (3D) structure of selected proteins of *A. marginale* and the peroxinectin protein of *R. microplus* were predicted using the PHYRE2 server (Kelley et al., 2015). Phyre2 PDB files were visualized with EzMol tool (Reynolds et al., 2018).

3. Results

3.1. Protein Selection. We identified 21 conserved amino acid sequences corresponding to membrane-associated proteins in all of the seven Mexican *A. marginale* strains (Table S1, supplementary material). Three protein sequences (PleD, MurJ, and TolC) of *A. marginale* were selected from this list, and the peroxinectin protein sequence of *R. microplus* was chosen after a transcriptomic analysis. The sequences selected have not been previously studied in *A. marginale* and/or *R. microplus* and have attributed functions that make them attractive as potential vaccine candidates.
3.2. PleD Family of Two-Component System Response Regulators. PleD is present in all seven Mexican published sequenced strains; it is composed of 455 amino acids, with an approximate molecular weight of 51.4 kDa, and a theoretical pI of 4.99 (Table 1). Clustal Omega alignment showed identity percentages between 99.56% and 100% among all Mexican sequences.

The subcellular location explored with CCTOP gave no potential model; thus, the sequence was analyzed with TMHMM, where the proposed model turned out to be an extracellular protein (data not shown), but when analyzed with both the CELLO v.2.5 and the PSORTb servers, they predicted a cytoplasmatic protein, in agreement with the purported function. Functional domain analysis with ScanProsite presents a typical PleD three-domain structure, two of them with the response-regulatory expected domains (4–121 and 158–274 amino acids, respectively) and a GGDEF domain. Vaxijen analysis showed that PleD is a possible antigen at 0.5 threshold. B-cell linear epitope analysis gave several representative sequences. From the analysis with ABCpred at 0.80 threshold and an 18 amino acid-length sequence, there were more than a dozen potential B-cell epitopes, yet only three sequences were recognized by the programs proposed in Materials and Methods. These epitopes were also individually analyzed by Vaxijen, and all three had scores ≥1.0. These sequences are graphically represented in Figure 1 along with the Phyre2 3D model, visualized with the EzMol online tool. The model obtained from Phyre2 analysis was obtained by comparison with the crystal structure of the “Response Regulator PleD” from Caulobacter vibrioides (PDB ID: 5c1w25 B). The 3D model obtained had >90% confidence out of a 41% identity. The three sequences representative of B-cell epitopes are SYDLPFIDLFNPGDGGLRF (amino acids 200–217), KRPNIDTIPHTVGDELLQQ (amino acids 337–354), and NFRSNNTRYTIPILVLDD (amino acids 220–236). These epitopes are also found in all other A. marginale sequences available at NCBI.

3.4. Outer Membrane Protein TolC. In all Mexican strains, TolC is composed of 408 amino acids, but in the strains of Aguaascalientes, Atitalaquia, and Puente de Ixtla, its sequences have a single amino acid change in position 43. The protein has a molecular weight of 45.3 kDa, a theoretical pI of 9.48, and an aliphatic index of 101.25 (Table 1). TMHMM and PSORTb predicted an outer membrane protein, whereas Phyre2 and CCTOP predicted a cytoplasmatic protein. The 3D model obtained from Phyre2, based on the models of its crystallized E. coli homolog (PDB ID: c1tqqC) is basically the same as that obtained with CCTOP, except that these tools consider the fact that the TolC final form is a homotrimer [25]. Thus, we consider TolC as a mostly cytoplasmatic protein, with a transmembrane domain that traverses to both internal and external membranes, and a short extracellular domain. Vaxijen analysis shows that at 0.5 threshold, TolC is not antigenic (Table 1). B-cell linear peptides analysis with ABCpred at 0.75 threshold showed about two dozen representative sequences, and from those, only three were chosen for their antigenicity scores, with two other programs. Epitope sequences INVDKASQRLEVRLRFPV (amino acids 273–290), GFLPRVTYDFVVQKDGRH (amino acids 60–77), and EAIKQEAKLNLKTTDLVL (amino acids 200–217) were all recognized as probable antigen by Vaxijen with scores above 0.8. These three epitopes are shown within the context of a 3D model generated with Phyre2 tool and visualized with EzMol (Figure 3). These three B epitope sequences are present in all of the seven Mexican strains and were also present in all other A. marginale, A. centrale, and A. ovis reported genomes.

3.5. Peroxinectin of R. microplus. In this work, we report, for the first time, a peroxinectin protein from the tick R. microplus.

This protein has a molecular weight of 90.35 kDa and a periplasmatic localization according to both CELLO and TMHMM servers (Table 2).

The prediction of ScanProsite shows a peroxidase domain that is shared with chorion peroxonectin of Ixodes scapularis with an identity of 94% according to Blastp. Although Vaxijen predicted the peroxinectin as a nonantigenic protein at 0.5 threshold, B-cell linear epitopes predicted with ABCpred at 0.75 threshold showed 45 sequences, and from those, only three were selected (SIFTAMHTLMREHNRV, amino acids 465–481; IGNVFAAAAYRYGHTL, amino acids 550–566; and VEQIRKASLARIICDN, amino acids 750–766) which are shown in the 3D model generated with Phyre2 tool and visualized with EzMol (Figure 4). The 3D model obtained from Phyre2 was based on the model of the crystallized human myeloperoxidase (PDB ID: c5fmaA) with a 100% confidence and a 37% identity with the peroxidase family domain.
4. Discussion

Recently, sequencing of genomes and omics approaches for pathogens of veterinary importance and their vectors have provided a significant amount of data. The analysis of these data is an important step in the process of designing new vaccines that can protect against *A. marginale*. The studies of the tick *R. microplus* have contributed to the understanding of the *A. marginale* interactions and the identification of possible targets to control tick infestations [26, 27].

In *A. marginale*, a large number of laboratory studies have focused on the MSPs and the Type Four Secretion

| Protein | PleD | MurJ | TolC |
|---------|------|------|------|
| NCBI accession number | KAA8473141 | KAA8472909 | KAA8472779 |
| Length AA | 455 | 501 | 408 |
| Minimum % identity | 99.56% | 99.60% | 99.75% |
| MW Da | 51479.39 | 54909.97 | 45329.33 |
| Theoretical pl | 4.99 | 9.37 | 9.48 |
| Aliphatic index | 93.01 | 127.49 | 101.25 |
| Domains (domain location) | Two response-regulatory (4–121 aa, 158–274 aa) GGDEF domain, (324–455 aa) | Lipid II flippase MurJ (26–484 aa) | Two outer membrane efflux domains, (25–203 aa and 222–403 aa) |
| Vaxijen threshold | 0.5725 | 0.4482 | 0.4482 |
| Location | Cytoplasmic 9.97 (PSORTb) | Cytoplasmic 10.00 (PSORTb) | Cytoplasmic protein (CCTOP) |

*Strain Mex-15 was used as reference for the calculation of parameters of each protein. Identity in all cases ranges from 99.56% to 100%.*

Figure 1: PleD graphic representation. (a) Phyre2 model. (b) SYDLFIIDLNFGGDGLRF epitope (amino acids 200–217); (c) KRYNDTFGHVGDELLQQ epitope (amino acids 337–354); and (d) NFRSNNNTYTPILVLLD epitope (amino acids 220–237). These sequences are recognized by all three epitope-prediction programs and have a score above 1.0 when analyzed by Vaxijen. The first residue of the chain is indicated with 1.
Figure 2: MurJ graphic representation. (a) Phyre2 model; (b) SRIMMVYLFCSLSSVVC epitope (amino acids 127 to 144); (c) EFKI-PAFFSCISVTNAL epitope (amino acids 373–390); and (d) YLKIHNLSERKSL epitope (amino acids 422–439). These sequences are recognized by all three epitope-prediction programs and, when analyzed by Vaxijen, the scores were above 0.75.

Figure 3: TolC graphic representation. (a) Phyre2 model; (b) INVDKASQRLEVRLRFPV epitope (amino acids 273–290); (c) GFLPRVTYDFVQKDGRH epitope (amino acids 60–77); and (d) EAIKQEAKLNLKTTLDVL epitope (amino acids 354–371). These sequences were recognized by all three epitope-prediction programs and were as well recognized as probable antigens by Vaxijen with scores above 0.8.
System proteins, but none of them have resulted in a vaccine capable of protecting cattle [28, 29, 30]. In the case of *R. microplus*, several recombinant vaccines have been developed; however, the search for new targets continue [6, 8]. In the present work, we have performed an analysis of the complete genome sequences of all Mexican *A. marginale* strains and extracted the sequences of membrane-associated proteins. Our approach was to edit MSPs, TFSS, and OMPs as curated by the NCBI and search for proteins that are highly conserved between all seven strains. From the seven Mexican strains, PleD, MurJ, and TolC were almost identical, with only few amino acid variations, ranging from 99.56% to 99.75% identity (Table 1). These proteins have important functions in Gram-negative bacteria, and two of them (PleD and TolC) have been studied in other Rickettsiaceae. PleD along with PleC form part of one of the several two-component signal transduction systems in Gram-negative bacteria commonly used to coordinate intracellular responses with environmental cues [31]. MurJ is a lipid II type flippase involved in the translocation of peptidoglycan from the cytoplasm to the periplasmic space in Gram-negative bacteria [32]. While there is no evidence of peptidoglycan synthesis in Rickettsiaceae [33], it seemed reasonable to explore MurJ in this context. Finally, TolC belongs to a family of multidrug transporters that provide an essential first-line defense mechanism against antibiotics, and also for expelling toxic compounds from the cell [34].

There is evidence that, at least, the *R. typhi* ankyrin secretion

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**Table 2: Physicochemical characteristics of peroxinectin protein.**

| Characteristic                  | Value       |
|--------------------------------|-------------|
| Length AA                       | 808         |
| MW Da                           | 90352.86    |
| Theoretical pI                   | 6.03        |
| Aliphatic index                  | 78.38       |
| Domains (domain location)        | Animal heme peroxidase (228–805 aa) |
| Vaxijen threshold                | 0.4894      |
| Location                        | Periplasmic protein (CELLO, TMHMM) |

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**Figure 4:** *Rhipicephalus microplus* peroxinectin graphic representation. (a) Phyre2 model; (b) SLTAMHTLMREHNRV epitope (amino acids 465–481); (c) IGNVFAAYRYGHTL epitope (amino acids 550–566); and (d) VEQIRKASLARIICDN, epitope (amino acids 750–766). These sequences were recognized by all three epitope-prediction programs and were visualized with EzMol.
is dependent on TolC [35]. The role of TolC in A. marginale is not known yet, but it has been reported that ankyrin is expressed during the infection of the tick vector, so it is possible that TolC may also play a role during infection of the tick vector. Furthermore, we described, for the first time, the presence of a peroxinectin protein of R. microplus. This protein has an important role in the process of melanization, where melanin acts as a protective barrier as part of a vital mechanism to defense against pathogens [38, 39]. Also, in insects, hemocyte opsonization mediated by peroxinectin facilitates the internalization of bacteria. The activities of peroxinectin reported in insects reinforce the potential of this protein as a probable target to control tick infestations.

The scope of our study was to report potential vaccine candidates under the criterion of conservancy, function, and antigenicity. The three selected proteins of A. marginale and the peroxinectin of R. microplus fill this criterion. The 3D models obtained allow for the localization of the proposed B-cell epitopes in terms of exposition to possible attack from specific antibodies. Then, peptides that contain these epitopes may be synthetized and used as a new alternative in designing diagnostic targets [40] or for vaccine candidates [41], as well as the use of Multiple Antigenic Peptides (MAPs) where one or even two epitope sequences can be included in a tetramer or an octamer to be synthesized and used as vaccine [41]. In our case, the combination of one or more of these peptides with an antigen from the tick vector may be a better alternative as on one hand, the pathogen is targeted and on the other, the vector can be targeted as well, probably minimizing transmission.

5. Conclusions

In this work, we present an alternative to the study of therapeutic targets against pathogens, considering the bioinformatic tools a good strategy for the design of new candidate molecules to control pathogens. Here, we present an alternative collection of immunogenic targets derived from in silico analysis of A. marginale proteins and their transmission vector R. microplus. The B-cell epitopes from the three proteins of A. marginale (PleD, MurJ, and TolC), and ToIC, have not been previously reported as immunogenic targets, and they could be a viable alternative to the known proteins used to design vaccines against bovine anaplasmosis. In regards to the R. microplus tick, we report the Pxn protein, considered a protein with a role in immunological, cellular, and reproductive mechanisms of tick and, therefore, a good candidate for its control.

Data Availability

Data are available from the corresponding author on request.

Conflicts of Interest

The authors declare no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Acknowledgments

This work was supported by INIFAP projects SIGI numbers 1232936014, 1351293413, and 112353134456.

Supplementary Materials

Table S1: proteins present in seven Mexican strains of Anaplasma marginale with immunogenic properties predicted by in silico analysis. (Supplementary Materials)

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