Comparison of antifungal and cytotoxicity activities of titanium dioxide and zinc oxide nanoparticles with amphotericin B against different Candida species: In vitro evaluation

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Abstract

Background: Candida species are known to cause serious fungal infections that produce cutaneous, mucosal, and systemic infections. Nowadays, mortality and morbidity candidiasis in immunocompromised patients have increased. Nanotechnology is a new world-known technology and includes particles ranging from about 1 to 100 nanometers. The purpose of this study was to evaluate the antifungal and cytotoxicity activities of titanium dioxide nanoparticles (TiO2-NPs) and zinc oxide nanoparticles (ZnO-NPs) compared to amphotericin B (AmB) on different Candida spp in in vitro conditions.

Methods: In the present study, susceptibility of different Candida species to TiO2-NPs and ZnO-NPs compared to AmB was determined by broth microdilution (BMD) and agar well diffusion methods. Cytotoxicity of TiO2-NPs and ZnO-NPs and amphotericin B was measured by MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide) assay.

Results: The results indicated that the TiO2-NPs and ZnO-NPs showed antifungal activities against pathogenic Candida spp. The minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) of TiO2-NP ranges against Candida spp. were 128-256 µg/mL and 256-512 µg/mL, respectively. The MIC and MFC values of ZnO-NPs were 64-128 µg/mL and 256-512 µg/mL, respectively. However, MICs and MFCs of AmB were 8-16 µg/mL and 16-32 µg/mL, respectively. The MTT assay results showed that the CC50% belonged to ZnO-NPs 706.2 µg/mL, for TiO2-NPs 862.1 µg/mL, and for AmB 70.19 µg/mL, respectively.

Conclusion: Our findings showed that TiO2-NPs and ZnO-NPs had antifungal effects against all Candida species, yet the antifungal properties of TiO2-NPs and ZnO-NPs were significantly less than those of AmB. The CC50% of AmB was significantly lower than ZnO-NPs and TiO2-NPs.
1 | INTRODUCTION

Candidiasis is an opportunistic fungal disease with a great variety of clinical symptoms that can affect various body parts including the skin, nails, oral mucosa, vagina, and internal organs. These infections can be fatal depending on the host’s immune system. This infection is known as one of the major important causes of death, particularly among immunocompromised hosts. Predisposing factors of Candida species include avitaminosis, obesity, pregnancy, alcohol consumption, broad-spectrum antibiotics and corticosteroids, physiological changes, and age. Several species of Candida such as C. albicans and non-albicans Candida (NAC) species including C. tropicalis, C. kefyr, C. krusei, C. glabrata, and C. parapsilosis are known as the common cause of candidiasis. Different species of Candida are able to convert to invasive pathogens when the immune system is weakened or the microbial balance of the normal flora of the body is disturbed. Topical antifungal drugs such as nystatin, clotrimazole, and miconazole are used to treat superficial candidiasis. For systemic or disseminated Candidiasis, oral ketoconazole, oral and intravenous fluconazole (FLC), and intravenous amphotericin B (AmB) are recommended.

A major concern of the increasing consumption of chemical drugs like antifungal agents is their side effects, which might in some cases be even more dangerous than the disease itself. Headache, nausea, vomiting, hepatotoxicity, and anaphylaxis are some side effects of these drugs. Other side effects of antifungal drugs include the development of resistant fungal species and therapeutic failures. For these reasons, many studies have been conducted to find new compounds with antifungal effects. They are one of the compounds of nanoparticles. The nanoparticles have been studied both individually and in combination with antifungal drugs to achieve better methods for treating various diseases. Furthermore, the nanoparticles are easily available, more affordable, and more effective with similar effects, which makes them a good alternative to chemical drugs.

Titanium dioxide (TiO2) is a nanoparticle, which has been investigated more than any other substance due to its very high photocatalytic activity. It exists in three crystalline phases including anatase, rutile, and brookite the most stable of which, at normal pressure and temperature, is the anatase structure and the other two phases are semi-stable. Some properties of this material that make it preferable to other particles include its high chemical resistance, non-toxicity, endurance, availability, and low production cost. In recent decades, the using of zinc oxide (ZnO-NPs) has increased due to large energy band gaps, chemical-thermal stability, high oxidation dependence (60 mV), and non-toxicity.

MTT assay is used to determine the rate of cell proliferation and cell viability, and its mechanism depends on the decrease of insoluble crystals of MTT formazan by the enzyme succinate dehydrogenase in the mitochondria of the cell. Dimethyl sulfoxide (DMSO) solution was used to dissolve these crystals. Then, the amount of light absorption was measured in terms of the intensity of the blue color of formazan at a wavelength of 540 nm. If the cell is alive and reproducing, the rate of dye production and the amount of absorption read are higher, while if more cells are dead and inactive, the rate of light absorption is lower. The aim of this study was to evaluate the antifungal and cytotoxicity activities of titanium dioxide nanoparticles and zinc oxide nanoparticles compared to amphotericin B.

2 | MATERIALS AND METHODS

2.1 | Preparation and determination of Characterization of TiO2-NPs and ZnO-NPs

TiO2-NP and ZnO-NP powders were purchased from Iranian Nanomaterial Pioneer Company (Mashhad, Iran). Characterization of TiO2-NPs and ZnO-NPs was determined by scanning electron microscope (SEM), UV spectroscopy, and X-ray diffraction (XRD). The SEM micrographs demonstrated the shapes and sizes of the TiO2-NPs and ZnO-NPs. The crystalline structure of these nanoparticles was measured using X-ray diffraction (XRD) (Panalytical, Almelo, Netherlands).

2.2 | Candida spp. and growth condition

This study was carried out on five different Candida species isolated from patients with different types of candidiasis. These species were previously identified by real-time PCR High Resolution Melting Analysis and sequencing methods. The species included C. tropicalis, C. parapsilosis, C. krusei, C. albicans, and C. lusitaniae. First, the Candida spp was subcultured onto Sabouraud dextrose agar (SDA) (Liofilchem Company, Italy).

2.3 | In vitro antifungal susceptibility testing

2.3.1 | Broth microdilution (BMD) method

To determine MIC and MFC values of TiO2-NPs and ZnO-NPs compared to AmB on five different Candida species, the guideline of Clinical and Laboratory Standard Institute (CLSI, M27-ED4 document) was followed. MIC was determined by broth microdilution and agar well diffusion methods. For broth microdilution method, serial dilutions of 4096-8 μg/mL for TiO2-NPs and ZnO-NPs and 128-0.125 μg/mL for AmB were prepared in a 96-well microplate containing RPMI-1640 medium (Sigma-Aldrich, USA). Then,
a suspension containing $1.5 \times 10^3$ cells/mL of each Candida species was added to all wells. The microplates were incubated in shaker incubator at 35°C for 24 hours. After incubation, each well was compared with the controls and the results as MIC were recorded. MIC values were the lowest concentrations of the TiO2-NPs, ZnO-NPs, and AmB, which inhibited the growth of Candida by 90% compared to the growth of control. Here, AmB was used as positive control. To measure MFC, 10 µL from wells with no turbidity were cultured on SDA. The plates were incubated at 35°C for 24 hours. After incubation, the lowest concentrations of TiO2-NPs, ZnO-NPs, and AmB, with three or less Candida colonies, were reported as MFC values. All experiments were carried out in triplicate.

### 2.3.2 | Agar well diffusion method

The inhibitory effects of various concentrations of TiO2-NPs, ZnO-NPs (8192, 4096, 2048, 1024, 512, 256, 128, 64, 32, 16, and 8 µg/mL), and AmB (128, 64, 32, 16, 8, 4, 2, 1, 0.5, 0.25, and 0.125 µg/mL) were evaluated on Candida species. A suspension containing $1.5 \times 10^3$ cells/mL of each Candida species was cultured on Sabouraud dextrose agar medium. Then, 6-mm wells were created inside the Sabouraud dextrose agar medium by a special punch. Various concentrations of TiO2-NPs, ZnO-NPs, and AmB were added to the wells. A well containing distilled water (DD) was considered as a negative control. The plates were placed in an incubator at 35°C for 24 hours. After incubation, the diameters of zones of inhibition of TiO2-NPs, ZnO-NPs, and AmB for each Candida spp were measured. The tests were performed three times, and the average of the diameters of zones of inhibition of TiO2-NPs, ZnO-NPs, and AmB was recorded.

### 2.4 | Measuring cytotoxicity of TiO2-NPs and ZnO-NPs compared to AmB by MTT assay

MTT assay was used to determine cell proliferation and viability of macrophage J/774. $1 \times 10^5$ mouse macrophages J/774 were cultured in a 96-well plate. After 24 hours, the culture medium was altered. Then, 10 µL of TiO2-NPs and ZnO-NPs at concentrations of 81920-320 µg/mL and AmB at concentrations of 1280-2.5 µg/mL was added to mouse macrophages J/774. Then, mouse macrophages J/774 were incubated at 37°C, 5% CO2 and 95% air. After 72 hours of incubation, the conditioned medium was discarded, and 25 µL of MTT solution (Sigma Chemical Co., Germany) was added to each well and incubated in the dark for 3 hours. After this time, the medium containing MTT was carefully removed and 100 µL of DMSO (Merck, Germany) was added to each well of the plate. Finally, the optical absorption was calculated based on the intensity of blue-colored formazan using a 96-well microplate reader (BioTek, USA) at 540 nm. If the mouse macrophages J/774 are viable and proliferating, the color is consequently more intense and higher absorbance readings are obtained. The experiment was repeated three times.

### 3 | RESULTS

#### 3.1 | Characterizations of TiO2-NPs and ZnO-NPs

As shown in Figure 1, TiO2-NPs have anatase form of 99% purity. The size of many nanoparticles is in the range of 10-25 nm. The SEM image shows a uniform distribution of the nanoparticles. The TiO2-NPs were spherical or rod-shaped and composed of 80% anatase and 20% rutile (Figure 1). Figure 2 shows the ZnO-NPs with a spherical structure of 99.8% purity. The size of most ZnO-NPs is in the range of 10-30 nm with an average of 20 nm. The SEM image shows a uniform distribution of the nanoparticles.
3.2 | X-ray diffraction patterns of TiO₂-NPs and ZnO-NPs

Figure 3 shows the X-ray diffraction pattern of TiO₂-NPs in the anatase phase. Several sharp peaks are observed in the range 2θ within 20-80°. The identified peaks in the range 2θ are 25.12°, 38.25°, 48.10°, 54°, 55.1°, 63°, 69.0°, 70.50°, and 75.10°. In the X-ray diffraction pattern of ZnO-NPs, several sharp peaks are observed in the range 2θ in the range 20-120° (see Figure 4). These peaks are identified in the range 2θ: 28.36°, 32.25°, 34.32°, 48.0°, 58.91°, 63.0°, 70.0°, and 71.20°.

3.3 | Antifungal effects of TiO₂-NPs and ZnO-NPs compared to AmB on different Candida species

Susceptibility of different Candida species to TiO₂-NPs and ZnO-NPs compared to AmB is shown in Figures 5 and 6. As seen in Figures 5 and 6, MIC and MFC values of TiO₂-NPs against Candida spp. were 128-256 µg/mL and 256-512 µg/mL, respectively. The range of MIC and MFC of ZnO-NPs was 64-128 µg/mL and 256-512 µg/mL, respectively. MIC and MFC values of AmB were 8-16 µg/mL and 16-32 µg/mL, respectively.

As shown in Figures 5 and 6, the lowest MIC of TiO₂-NPs in C albicans, C krusei, and C parapsilosis was 128 µg/mL and the highest MIC of TiO₂-NPs in C lusitaniae and C tropicalis was 256 µg/mL. Moreover, the results indicated that the highest MFC of TiO₂-NPs for C tropicalis was 512 µg/mL and the lowest MFC of TiO₂-NPs in C parapsilosis, C lusitaniae, C krusei, and C albicans was 256 µg/mL. The minimum amount of ZnO-NPs required for the growth inhibition of C parapsilosis and C krusei was 64 µg/mL and for C lusitaniae, C albicans, and C tropicalis was 128 µg/mL. The lowest MIC of ZnO-NPs was observed for C parapsilosis and C krusei, and the highest MIC values were obtained for C lusitaniae, C albicans, and C tropicalis.

The lowest MIC value of AmB in C lusitaniae, C albicans, and C tropicalis was 8 µg/mL, and the highest MIC value of AmB in C parapsilosis, and C krusei was 16 µg/mL. The results revealed that the highest MFC of AmB in C tropicalis and C albicans was 32 µg/mL and the lowest MFC of AmB in C parapsilosis, C lusitaniae, and C krusei was 16 µg/mL.

In the present study, using the agar well diffusion method, the diameters of zones of inhibition of five different Candida spp in confronting various concentrations of TiO₂-NPs, ZnO-NPs and AmB were measured. The highest zone of inhibition of the TiO₂-NPs at concentrations 8192 and 4096 µg/mL on all tested Candida spp was ≥60 mm. No growth inhibition in Candida species was observed in concentrations <512 µg/mL of TiO₂-NPs.

The largest inhibition zone of ZnO-NPs at concentrations of 8192 µg/mL on C parapsilosis was obtained by the size equal to 20 mm. No growth inhibition was observed at concentrations 2048 µg/mL of ZnO-NPs for C parapsilosis, C tropicalis, and C krusei, and at concentrations <2048 µg/mL of ZnO-NPs for all tested Candida spp. The maximum zones of inhibition on C albicans, followed by C krusei and C parapsilosis produced by concentrations of 128 µg/mL of AmB. Totally, it suggested that the antifungal activities of TiO₂-NPs, ZnO-NPs, and AmB increased with increase in concentrations. The diameters of zones of inhibition of TiO₂-NPs, ZnO-NPs, and AmB at various concentrations against different Candida species (mm) are shown in Tables 1-3, respectively.

3.4 | MTT assay results

Survival rate of mouse macrophages J/774 exposed to TiO₂-NPs and ZnO-NPs compared to AmB is shown in Figure 7. The MTT results showed that the CC50% were for ZnO-NPs 706.2 µg/mL, for TiO₂-NPs 862.1 µg/mL, and for AmB 70.19 µg/mL, respectively. Our finding indicated that the rate of cytotoxicity increased with increasing concentration. The CC50% of AmB was significantly lower than ZnO-NPs and TiO₂-NPs (P < .05). The cytotoxicity ratio ranking on mouse macrophages J/774 was TiO₂-NPs > ZnO-NPs > AmB.

4 | DISCUSSION

In the recent decades, the prevalence of opportunistic diseases such as fungal infections in immunocompromised patients has been increased. Candidiasis is the major common invasive fungal infection in human. Due to the side effects of antifungal drugs and the
development of resistant fungal species and therapeutic failures, attention has been drawn to the use of novel antifungal compounds with fewer side effects. One of these novel compounds is nanoparticles. The nanoparticles are very small substances <100 nm in size that have various physical and chemical properties, act on living cells at the nanoscale and initiate different effects. Of all nano-sized materials, metal oxide nanoparticles are the most attractive. Some of these metal nanoparticles are considered in medicine.
and pharmaceutical sciences due to their unique properties. ZnO-NPs, magnesium oxide, and TiO2-NPs are examples of these nanoparticles.

The primary aim of this study was to evaluate the antifungal effects of TiO2-NPs and ZnO-NPs compared to AmB on different Candida spp. Our findings showed that the TiO2-NPs and ZnO-NPs had antifungal potentials against pathogenic Candida spp. and could stop the growth of all tasted Candida spp at different concentrations, although their inhibitory effect is less than AmB. The results of Karimiyan et al study were consistent with those of the present study. They showed that ZnO-NPs and copper oxide (CuO) nanoparticles exerted their inhibitory effect at a higher concentration compared to AmB; also, magnesium and silicon nanoparticles had no inhibitory effect on C albicans.

Moreover, Memarian et al demonstrated that gold nanoparticles exerted their inhibitory effect on resistant strains of C albicans at a higher concentration as compared with FLC and conjugated FLC. Dananjaya et al concluded that significant changes occurred in the external morphology of C albicans after treatment with both types of ZnO-NPs due to the damage to the fungal cell membrane. In addition, the cytotoxicity of both nanoparticles was compatible with the increase in the concentration of Hep2 cells and C albicans. In another study, chemically synthesized TiO2-NPs exhibited higher anti-candidal property compared with FLC.

In the present study, the inhibitory effect of the ZnO-NPs occurred at MIC = 128 μg/mL for C albican, while MIC of ZnO-NPs and ZnO-chitosan nanocomposites on C albican was 200 μg/mL and 75 μg/mL, respectively. In the study concluded by Grabchev et al, copper complex molecules and benzofuran-cyclam had a greater antibacterial and antifungal effect than copper free ligand.

The secondary aim of the present study was to compare the cytotoxic effect of TiO2-NPs and ZnO-NPs with AmB. Our findings indicated that the rate of cytotoxicity increased with increasing concentration. The CC50% of AmB was significantly lower than ZnO-NPs and TiO2-NPs.

Previous studies have shown the cytotoxic activity of different nanoparticles against various cell lines. Ghadimi et al reported that the toxicity of the nanoparticles was dose-dependent at 24 hours and the highest cytotoxic effects were observed at concentrations of 50 and 100 μg/mL and IC50 equal to 7.14 μg/mL. In the current study, the CC50% of TiO2-NPs was significantly higher on mouse macrophages J/774, than those reported in the study performed by Ankita Chatterjee et al, who reported that the TiO2-NPs at a concentration of 200 μg/mL exhibited cytotoxic activity against the Mg 63 osteosarcoma cell lines. The cytotoxic effects of ZnO-NPs on lung cancer cell line A549 occurred at 33-37 μg/mL. Different nanoparticles have completely different antifungal and cytotoxic effects, which can be explained by the type, size, concentration of nanoparticles, and other factors.
and synthesis mode of the nanoparticles as well as the studied *Candida* species.

## 5 | CONCLUSION

Our finding showed that the TiO$_2$-NPs and ZnO-NPs had antifungal potentials against pathogenic *Candida* spp. and could inhibit the growth of all tested *Candida* spp. However, its antifungal properties were significantly less than those of AmB. MTT assay results revealed that the rate of cytotoxicity increased with increasing concentrations. Finally, the CC$_{50}$% of AmB was significantly lower than ZnO-NPs and TiO$_2$-NPs.

## CONFLICT OF INTEREST

The authors declare no competing interests.

## AUTHORS CONTRIBUTIONS

SS and PGHA developed the study concept and design. SHA collected the data. SS analyzed and interpreted the data. SS wrote the article. PGHA and SS revised and edited the article. All authors read and approved the final article.

## ETHICAL APPROVAL

The study was evaluated and approved by the Ethics Committee of the Kerman Medical University and Kerman Research Council (IR. KMU.REC.1397.542).

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## REFERENCES

1. Zahabi ZF, Sharififar F, Ghasemi Nejad Almani P, Salari S. Antifungal activity of different fractions of *Salvia rhytidea* Benth as a valuable medicinal plant against various species of *Candida* in Kerman Province, southeast Iran. *Gene Rep.* 2020;19:1-7.
2. Khan SN, Khan S, Misba L, Sharief M, Hashmi A, Khan AU. Synergistic fungicidal activity with low doses of eugenol and amphotericin B against *Candida albicans*. *Biochem Biophys Res Commun.* 2019;518(3):459-464.
3. Cheng M-F, Yang Y-L, Yao T-J, et al. Risk factors for fatal candidemia caused by *Candida albicans* and non-albicans *Candida* species. *BMC Infect Dis.* 2005;5(1):1-5.
4. Barati M, Mirkalantari S, Ansari S, Salari S, Fattahi A. Determination of antimicrobial susceptibility pattern of *Candida* species isolated from patients with symptomatic candiduria. *Res J Med Sci.* 2019;24:1-3.
5. McManus BA, Coleman DC. Molecular epidemiology, phylogeny and evolution of *Candida albicans*. *Infect Genet Evol.* 2014;21:166-178.
6. Eghtedar Nejad E, Ghasemi Nejad Almani P, Mohammad MA, Salari S. Molecular identification of *Candida* isolates by Real-time PCR-high-resolution melting analysis and investigation of the genetic diversity of *Candida* species. *J Clin Lab Anal.* 2020;34(7):1-8.
7. Hasanvand S, Azadegan Qomi H, Kord M, Didehdar M. Molecular epidemiology and in vitro antifungal susceptibility of *candida* isolates from women with vulvovaginal candidiasis in northern cities of Khuzestan province, Iran. *Jundishapur J Microbiol.* 2017;10(8):1-6.
8. Salari S, Almani Ghasemi Nejad P. Antifungal effects of Lactobacillus acidophilus and *Lactobacillus plantarum* against different oral *Candida* species isolated from HIV/AIDS patients: an in vitro study. *J Oral Microbiol.* 2020;12(1):1-10.
9. d’Enfert C, Hube B. *Candida: Comparative and Functional Genomics*. Vol 10. UK: Caister Academic Press; 2007.
10. Seddighi NS, Salari S, Izadi AR. Evaluation of antifungal effect of iron-oxide nanoparticles against different *Candida* species. *IET Nanobiotechnol.* 2017;11(7):883-888.
11. Jeena V, Robinson RS. Green oxidations: titanium dioxide induced tandem oxidation coupling reactions. *Beilstein J Org Chem.* 2009:5(1):1-4.
12. Augugliaro V, Loddo V, López-Muñoz MJ, et al. Home-prepared anatase, rutile, and brookite TiO$_2$ for selective photocatalytic oxidation of 4-methoxybenzyl alcohol in water: reactivity and ATR-FTR study. *Photochem Photobiol Sci.* 2009;8(5):663-669.
13. Enyashin AN, Seifert G. Structure, stability and electronic properties of TiO$_2$ nanostructures. *Phys Status Solidi B.* 2005;242(7):1361-1370.
14. Mirzaei H, Darroudi M. Zinc oxide nanoparticles: biological synthesis and biomedical applications. *Ceram Int.* 2017;43(1):907-914.
15. Yazdi-Rouholami E, Motamed N, Tahmasb M, Omidfar K. Silibinin cytotoxic effect on MCF-7 cell line. *J Sabzevar Univ Med Sci.* 2016;23(3):386-391.
16. CLSI. *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts: 4th informational Supplement CLSI M27-S.* Wayne, PA: CLSI; 2017.
17. Salari S, Seddighi NS, Almani PGN. Evaluation of biofilm formation ability in different *Candida* strains and anti-biofilm effects of Fe$_3$O$_4$-NPs compared with Fluconazole: an in vitro study. *J Mycol Med.* 2018;28(1):23-28.
18. Arastefar A, Badiee P. A Review of antifungals and their mono-and combination-therapy in the treatment of invasive fungal infections. *J Kerman Univ Med Sci.* 2016;23(6):829-842.
19. Medina C, Santos-Martinez M, Radomski A, Corrigan O, Radomski M. Nanoparticles: pharmacological and toxicological significance. *Br J Pharmacol.* 2007;150(5):552-558.
20. Niederberger M, Pinna N. *Metal Oxide Nanoparticles in Organic Solvents: Synthesis, Formation, Assembly and Application*. Germany: Springer Science & Business Media; 2009.
21. Zou C, Gao W. Fabrication, optoelectronic and photocatalytic properties of some composite oxide nanomaterials. *Trans Electr Electron Mater.* 2010;11(1):1-10.
22. Karimiyan A, Najafzadeh H, Ghorbannour M, Hekmati-Moghadam SH. Antifungal effect of magnesium oxide, zinc oxide, silicon oxide and copper oxide nanoparticles against *Candida albicans*. *Zahedan J Res Med Sci.* 2015;17(10):19-23.
23. Memarian M, Javadi A, Fateh R. Antifungal effects of gold nanoparticles conjugated fluconazole against fluconazole resistant strains of *candida albicans* isolated from patients with chronic vulvovaginitis. *Qom Univ Med Sci J.* 2016;10(7):10-19.
24. Dananjaya S, Kumar RS, Yang M, Nikapitiya C, Lee J, De Zoyas M. Synthesis, characterization of ZnO-chitosan nanocomposites and evaluation of its antifungal activity against pathogenic *Candida albicans*. *Int J Biol Macromol.* 2018;108:1281-1288.
25. Haghighi F, Roudbar M, Mohammd P, Eskandari M. Comparative evaluation of the effects of TiO$_2$ nanoparticles and its photocatalytic form on the formation of fungal biofilms. *Arak Univ Med Sci.* 2012;15(1):27-34.
26. Grabchev I, Yordanova S, Vasileva-Tonkova E, et al. A novel benzo-furazan-cyclam conjugate and its Cu(II) complex: Synthesis, characterization and in vitro cytotoxicity and antimicrobial activity. *Dyes Pigm.* 2016;129:71-79.
27. Ghadimi F, Mirzae A, Arasteh J. Antibacterial and cytotoxicity of synthesized silver nanoparticles using Erica carnea extract on breast cancer cell line (MCF-7) and analysis of its apoptotic effects. *Razi J Med Sci*. 2019;26(6):84-94.

28. Chatterjee A, Nishanthini D, Sandhiya N, Abraham J. Biosynthesis of titanium dioxide nanoparticles using Vigna radiata. *Asian J Pharm Clin Res*. 2016;9(4):85-88.

29. Reddy A, Srividya L. Evaluation of in vitro cytotoxicity of zinc oxide (ZnO) nanoparticles using human cell lines. *J Toxicol Risk Assess*. 2018;4(1):1-3.

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