Plastics in robots: a degradation study of a humanoid skin mask made of soft urethane elastomer

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Abstract
Understanding the degradation of plastic materials is a big challenge for curators, conservators and conservation scientists in museums worldwide aiming to preserve their collections due to the variety of formulations of synthetic polymers and pigments. The conservation of polyurethane (PUR) based objects is challenging because they can suffer from extensive degradation. Particularly PUR elastomers can degrade shortly after their production, as occurred to the mask of the Japanese robot SAYA, which within 8 years suffered from two large tears, discoloration and stickiness. This research aims at studying the degradation phenomena of the androids’ synthetic skin. Better knowledge of the chemical composition of the mask and the chemical and physical decay will contribute to planning a suitable stabilization treatment. Within a multi-analytical approach, colorimetric and microscopic investigations highlighted discolored areas, which showed further color changes within a five months monitoring campaign, confirming the instability of the material likely due to ongoing degradation. Raman microscopy allowed the identification of Pigment White 6 (titanium dioxide TiO₂) in the anatase form, known to promote the photosensitivity of PUR substrates towards ultraviolet (UV) light. Attenuated total reflectance Fourier transform infrared (ATR-FTIR) spectroscopy identified the PUR composition of the mask, the presence of phthalates as plasticizers and suggested the formation of quinone chromophores in the polymer structure as a result of photo-oxidation, possibly responsible for the mask yellowing. Evolved gas analysis–mass spectrometry (EGA-MS) and double-shot-gas chromatography/mass spectrometry (TD/Py–GC/MS) analyses support the characterization of the formulation of the mask as being made of methylene diphenyl diisocyanate (MDI) PUR ether elastomer. Plasticizers in high concentration, mainly diisononyl phthalate (DINP) and bis(2-ethylhexyl)phthalate (DEHP), and the UV stabilizer Tinuvin 328 were also detected. In addition, the presence of styrene-acrylonitrile (SAN) could also contribute to the mask’s chemical instability. More amount of UV stabilizer and phthalates were detected at the surface (contributing to its stickiness) than in the inner core. The degradation of the mask results from the light susceptibility of MDI PUR ether and SAN, as well as the higher photochemical activity of anatase. The mask was transferred on to a mannequin and placed in the storage area to prevent light exposure and photo-oxidation. As loose edges had to be stabilized, tests were conducted and adhesive stripes glued with a PUR dispersion were selected for keeping the head’s shape. The novelty of this study is the implementation of conservation science on the study of androids with PUR elastomeric components in robotic collections, which are becoming increasingly popular in technical museums, however still seldomly studied.

Keywords: Humanoid robot, Urethane elastomer (PUR), Natural aging, Yellowing, Stickiness, Raman microscopy, ATR-FTIR, EGA-MS, TD/Py–GC/MS, Preventive conservation

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Introduction

Since the 1990s, technology research has led to the development of robots increasingly similar to humans. The studies of this type of robot imply investigating the possible mechanisms, intelligence, and/or functions required for an effective interaction with humans, such as facial expression. Therefore, several robots with an anthropomorphic face capable of mimicking emotions, e.g. surprise, fear, disgust, anger, happiness and sadness, were developed [1–6]. Investigations of the biomechanics of the human face permitted the reproduction of basic facial movements, controlled by several actuators with mechanical drives or smart systems [6]. The first robotic faces were composed of: a mechanical structure with actuators, a skull frame often made of fibre-reinforced plastic (FRP) and facial skin [2–4]. The similarity of robot’s facial expressions with human beings depends on the number and locations of control points on the facial skin for deformation. Therefore, the artificial mask needs to be deformable, soft and similar to the human skin texture. Soft polyurethane (PUR) or synthetic silicone rubber [2, 4, 6–8] were usually used to reproduce the humanoid masks created by mould technology. The production, often used in art and for the reproduction of special cinematic effects, follows a series of moulding processes, which consist of: (i) production of a human face mould; (ii) injection of an elastomer in the mould; (iii) release of the skin and finish the manufacture by trimming, painting and making up [4, 5]. PUR elastomers are formed by polymerization of diisocyanate (either aromatic or aliphatic), polyester or polyether polyols (macrodils) and chain extenders (typically small-molecule diol or a diamine), which are characterized by an alternation of hard (diisocyanate-chain extender sequence) and soft (macerdiol sequence) segments [9, 10]. Their casting procedure consists of mixing two low viscous liquid components under a specific ratio, generally expressed by the supplier, which cures at room temperature. The two components are often a polyol mixture and an isocyanate. Based on the required softness, a certain quantity of plasticizer can be also added [9, 11]. Other additives, such as UV absorbers, antioxidants and pigments, are introduced into the polyol part during the beginning of the moulding [11].

Objects made of PUR, together with those made of cellulose nitrate, cellulose acetate and polyvinylchloride (PVC), tend to be more susceptible to degradation than other plastic objects in museum collections [12–14]. Several studies were conducted to understand the aging of PUR foams [12–18] or PUR coatings [19, 20], which alterations depend mainly on the ether or ester composition of the polymer. PUR ether polymers are more sensitive to oxidation, which causes discoloration and loss of mechanical properties, while PUR ester polymers degrade mainly by hydrolysis, caused by exposure to moisture, heat and light, and lead to chain scission of the polymer and its crumbling [10, 12, 21]. However, only few studies were dedicated to objects made of PUR elastomer and their degradation. An example is the design object Soft Vase (1994) of the Dutch designer Hella Jongerius made by PUR ester elastomer internally plasticized with 1,4-butaneoldiol (chain extenders), which presented only mechanical damages [9]. Other investigated objects were the works of art of the series Stage Evidence of the Italian artist Loris Cecchini. Capanna et al. [22] have reported mechanical damages, loss of elasticity, weeping of liquids and stickiness for Stage Evidence 2002 (fotocopistrice), while de Groot et al. [9] and Lazzari et al. [23] have reported also yellowing for Finestre II (2000) and Stage Evidence (1999), respectively. The objects were identified as PUR ester elastomer [9, 22] and PUR ether elastomer [23] with a high content of phthalates used as plasticizers. A similar degradation phenomenon found on the objects of Loris Cecchini was observed on the facial mask of the robot SAYA. The android was developed and built by Professor Hiroshi Kobayashi and his team from the Tokyo University of Science in 2009, including the facial skin made of soft PUR elastomer [2]. The prototype was tested as a telecommunication interface with humans and served initially as a receptionist and, later on, as an English teacher in Japanese elementary schools [24]. SAYA was loaned to the Deutsches Museum (Munich, Germany) in 2017 to be part of the future permanent robotics’ exhibition. However, already in 2017 the facial mask was severely degraded. Besides the allover stickiness, a localized discoloration of the PUR elastomer was visible, changing from a human-looking skin to spotty colored patterns. Moreover, two large tears located around the neck and close to the left shoulder compromised the mask’s integrity (Additional file 1: Fig. S1). Their development is likely associated with tensile stresses, deriving from the head movements during function and/or tilted back resting position.

The investigation aimed to characterize the composition of the PUR mask; understand its degradation phenomena, and suggest measures for minimizing its degradation.

Within this study colorimetry, microscopy, scanning electron microscopy–energy dispersive X-ray (SEM–EDX), Raman microscopy (μ-Raman), attenuated total

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1 The name Saya has the following two meanings: 1) “SAYA” is originally written in Chinese character (KANJI) meaning “bloom in the field”; 2) Saya means “me” in Malayan. The Japanese researcher team used it to mean that the robot has an ego (Prof. Takuya Hashimoto, personal communication, November 2, 2020).
reflectance Fourier transform infrared spectroscopy (ATR-FTIR), evolved gas analysis-mass spectrometry (EGA-MS) followed by double-shot-gas chromatography/mass spectrometry (TD/Py–GC/MS) were employed. Based on the material analysis, a new support (mannequin) was chosen, and the effectiveness of gluing stripes for stabilizing the tears was assessed. Thanks to the integration of the information gathered from the multi-technique approach, new insights into the chemical composition of the PUR elastomeric mask and its degradation were achieved, helping to suggest preventive and active conservation measures for its long-term preservation at the Deutsches Museum.

Materials and methods
The SAYA mask
The humanoid robot SAYA’s mask is made of a soft PUR elastomer, HITOHADA GEL® by EXSEAL CO., LTD (Prof. Takuya Hashimoto, personal communication, November 5, 2019). The elastomeric skin was severely degraded when it arrived at the Deutsches Museum in November 2017 (Fig. 1).

To prepare the future permanent exhibition, several meetings with the robotic curators started in March 2019. Unfortunately, due to the poor condition state of the mask, it was decided to replace it. A diagnostic campaign started on SAYA’s mask to understand its decay and protect it in the storage area. Therefore, the skin was transferred onto a new inert mannequin made of polypropylene (PP) with a similar shape to the robot’s head (Fig. 2). While removing the mask, it was observed that the actuators located inside the skull could control the facial movements through metal connections between themselves and the inner surface of the mask. Mask and skull were secured via velcros (e.g. cheeks, top of the head and ears).

Furthermore, the external surface of the mask displayed an inhomogenous coloration: a pale pink color around the eyes, ears, band on the top of the head, zip in the back, neck and a part of the chest; and yellow on the rest (Additional file 1: Fig. S1). The location of those color regions on the different parts of the mask could not be associated with a specific degradation trend.

Two distinct color profiles were observed on the mask’s edges (Fig. 3): yellow | pale pink core | yellow (section a); pale pink | glue (section b) corresponding to regions exposed to oxygen from the outside and with a glue in the inside. Samples from the yellow surface and the pale pink core were taken for investigation.

The glue (Fig. 3, section b) helped fix the metallic connections between the mask and the skull, as well as the adhesion between the mask and a velcro tape. Microsamples of the glue and of the skull were taken for analysis.

Colorimetry
The color measurement on the mask’s surface was conducted with the portable Konica Minolta CM-700d Spectrophotometer (Minolta Co. Ltd., Japan) in May 2019. The measurement campaign was repeated 5 months later (October 2019) to monitor possible changes or color shifts. The following parameters were used for the
measurements: spectral range 400–700 nm with a wavelength pitch of 10 nm, d/8° geometry, SCI mode, 10° colorimetric observer (CIE 1964), D65 standard illuminant and measurement area of 3 mm in diameter. In addition, measurement points of the mask’s external surface were selected in both the pale pink and yellowed regions, recording two replicas for each point. The color difference was calculated as the coefficient ΔE using the of CIEL*a*b* color space (L* as a parameter for brightness between 100 for white and 0 for black, while a* and b* are the chromaticity coordinates) and the mathematical formula of CIEDE 1976 \( \Delta E_{a,b} \) \[25\]:

\[
\Delta E_{a,b} = (L_2^* - L_1^*)^2 + (a_2^* - a_1^*)^2 + (b_2^* - b_1^*)^2 / 0.5
\]

This formula was applied to calculate both the color change between non degraded (pale pink, subscript 1) and degraded (yellow, subscript 2) areas and possible color variations in correspondence of these two areas over the monitoring period (May 2019, subscript 1; October 2019, subscript 2). A positive difference of Δa* and Δb* describes red and yellow shifts, while a negative difference of Δa* and Δb* describes green and blue shifts \[19, 26\].

Microscopic and scanning electron microscopy—energy dispersive X-ray spectrometry (SEM–EDX) investigations

Visual differences between the yellowed surface and pale pink core were examined under a digital microscope (Keyence VHX-1000) equipped with a 5–50× objective lens and a scanning electron microscope Phenom ProX, version G3 (Phenomworld, now ThermoFisher), low vacuum, Ce-hexahybrid cathode. EDX-analysis were performed with 15 kV, point beam current and 90 s,
standardless ZAF quantification. The sample was analyzed without any preliminary preparation.

**Raman microscopy (μ-Raman)**

For the identification of pigment(s), the PUR sample was analyzed superficially and in cross-section by μ-Raman spectroscopy. The sample was placed onto glass slides on the stage of an Olympus BXFM-ILHS microscope coupled to a Jobin Yvon LabRam 300 confocal Raman spectrometer. The μ-Raman study was carried out using 785 nm line of diode laser (100-mW), for the Raman excitation. The apparatus included an air-cooled multi-channel CCD detector. Gratings of 600 groves/mm were used to collect spectra between 100 and 1000 cm⁻¹. The system was calibrated by using the 520.7 cm⁻¹ band of a standard silicon (1 0 0) wafer. The confocal pinhole was maintained at a minimum aperture (200 μm) to reduce the thickness of the focal plane. Integration time and accumulations were noted to achieve an acceptable s/n ratio. The laser beam was focused on the sample by a ×100 objective lens to give a spot size of approximately 1 μm. The laser power at the surface of the samples was controlled using neutral density filters (optical densities 0.3, 0.6, 1 and 2). Spectra were recorded as an extended scan. Raman data analysis was performed using LabSpec 5 software (Horiba Jobin Yvon).

**Attenuated total reflectance Fourier transform infrared spectroscopy (ATR-FTIR)**

Infrared analyses of the yellow surface (Fig. 3a), the pale pink core and the glue (Fig. 3b) found on the reverse of the mask were performed with an Alpha FT-IR Spectrometer (Bruker) equipped with a diamond crystal. Infrared spectra were collected after 64 scans from 4000 to 400 cm⁻¹ with a resolution of 4 cm⁻¹, recording two replicas for each point. The interpretation of spectra was performed with the OPUS 7.2 software (Bruker). Bis(2-ethylhexyl)phthalate (DEHP, Sigma Aldrich) was used as plasticizer reference material to perform a spectral comparison.

**Evolved gas analysis-mass spectrometry (EGA-MS)**

Evolved gas analyses of the yellow surface and the pale pink core were carried out using a Multi-shot Pyrolyzer EGA/PY-3030D (Frontier Lab.). Around 200 μg of the sample were added directly into a stainless steel Eco-cup sample holder (Frontier Lab.). The pyrolyzer temperature program varied from 100 to 700 °C (hold for 5 min), increasing 20 °C/min. The evolved gas was transferred directly into the 5977B MSD mass spectrometer (Agilent) through a deactivated and uncoated stainless-steel transfer Frontier Ultra Alloy® EGA tube (UADTM 2.5N–2.5m-l.D. 0.15 mm, O.D. 0.47 mm, Frontier Lab.) maintained at 300 °C in the 7890B GC system oven (Agilent) with Helium as the carrier gas, a flow rate of 1.2 mL/min and a split ratio of 25:1. Ion detection was carried out in the m/z range 25–550. The thermogram interpretation and the volatile compound identification were performed using a mass spectrum comparison and a selective ion curve extraction with the software and database F-Search 3.5.0 (Frontier Lab.) as well as literature data [17, 27].

**Double-shot-gas chromatography/mass spectrometry (TD-Py/GCMS)**

In order to collect more information about the volatile organic compounds and the polymeric composition, double-shot-GCMS (see instrumental details in the section above) was applied to measure ca. 200 μg of the yellow surface and ca. 200 μg of the pale pink core. The first step, thermal desorption (TD-GC/MS), was performed from 50 °C (hold for 2 min) to 320 °C (hold for 3 min) with an increasing ratio of 20 °C/min. At the beginning of the separation column, the focalization of the volatile compounds was assured by a cryo-trap maintained at −180 °C with liquid nitrogen [28]. The second step, flash pyrolysis (Py-GC/MS), was conducted at 600 °C for 6 s. GC separation was performed using a Frontier UA5 capillary column (30 m-0.25F, 30 m × 250 μm × 0.25 μm, Frontier Lab.), Helium as the carrier gas with a flow rate of 1.2 mL/min and a split of 30:1. The injector temperature was set to 300 °C. The column temperature program was set from 40 °C (hold for 2 min) with a heating rate of 20 °C/min until 280 °C (hold for 5 min). The ionization was EI positive mode at 70 eV, source temperature at 230 °C, quadrupole at 150 °C and mass scanning range of 15 to 550 m/z. Compound identification was performed using NIST MS Search 2.2 and F-Search 3.5.0 (Frontier Lab.) databases and literature data [17, 27, 29].

**Testing the effectiveness of gluing stripes for stabilizing the tears on the new mannequin**

In order to stabilize and minimize further deformations, an active conservation treatment was planned. It aimed to find suitable gluing stripes that can restabilize adhesion between the tears and connect the mask outward bent edges with the mannequin for keeping its original shape.

The gluing stripes consist of Japanese paper, which were tested with several adhesives. The ripped long-fibred Japanese paper (100% Kozo fibres, 12 g/m², Defner & Johann GmbH), a material widely used in the conservation field [30–32], should allow a smoother transition between edges and provide sufficient strength due to the Kozo fibres length. The adhesives should be soluble in water to avoid the exposure of the PUR to harsh solvents and guarantee a long-term flexibility and elasticity
therefore, cellulose-based adhesives were tested: (i) Methocel A 15 LV (Merck KGaA), (ii) Tylose MH 300 (Kremer Pigmente GmbH & Co. KG), (iii) Tylose MH 1000 (Kremer Pigmente GmbH & Co. KG), iv) wheat starch paste (Kremer Pigmente GmbH & Co. KG). A synthetic polymer-based adhesive, the aqueous PUR dispersion with no free isocyanate groups PU 52 (Kremer Pigmente GmbH & Co. KG) [35, 36], was also considered in order to investigate the effectiveness of a compatible product with the PUR mask. Van Oosten [14] reported that a similar product with another aliphatic PUR ester dispersion (Impranil® DLV) was already tested to stabilize PUR foam material with promising results. Characteristics of the adhesives selected for investigation are presented in Table 1.

### Preparation of the adhesives and test strips
Distilled aqueous solutions of 5% of Methocel A 15 LV, Tylose MH 300 and Tylose MH 1000 were prepared under continuous magnetic stirring at 20 °C. 20% of wheat starch paste was heated for 30 min under constant stirring in distilled water. Extra 20 mL of distilled water were added to compensate for moisture lost during the cooking. Then, the wheat starch paste was sieved two times and consequently diluted with water to get a concentration of 10%. The polyurethane dispersion PU 52 was used without any further preparation.

Before testing the adhesives on the original surfaces, the proper adhesion between the Japanese paper and the testing adhesives was assessed, as well as their resulting flexibility. For this a set of Japanese paper strips were embedded with the adhesive solutions (Table 1) and left to dry at 20 °C and 50% RH. Since the adhesives did not delaminate, the strips retained their original shape and demonstrated a good flexibility when pulled manually, they were all selected for further tests on the intended PUR elastomer and PP mannequin.

The PUR and PP surfaces were cleaned using a mildly damp fine-pored polyurethane sponge before being treated. The adhesives were applied in drops onto the paper strips and then gently pressed on the testing surface using a flat short-haired brush. Afterwards, the paper strips were left dry without further pressure at 20 °C and 50% RH (Fig. 4). After 7 days, the gluing treatments were assessed visually and manually by pulling the strips to determine their adhesion to the substrates. In addition, the flexibility of the paper embedded with adhesives was determined after 7 days of drying by controlling the occurrence of cracks due to shrinkage and/or failure after a manual tension test.

### Results and discussion

#### Colorimetry

The colorimetric analysis performed in May 2019 (after twenty-two months from the loan) highlighted two different colors of the humanoid mask. As reported in Fig. 5, two clusters were recognized for the pale pink and yellow regions (for sampling spots and coordinate values, see Additional file 1: Fig. S2 and Table S1), which showed ΔE* values of 1.6 and 2.5 respectively, after 5 months of monitoring. These variations indicate an ongoing color changing progress meanwhile the mask was stored in...
laboratory conditions (ca. 22 $^\circ$C, 40–60% RH, mostly kept in the dark and for short times exposed to 250 lx). PUR elastomer, especially polyether- and aromatic isocyanates-based, is known to be very sensitive to light [19, 37–39]. In the SAYA mask case, the points uncovered by the cloths and wig yellowed with time. A clear color change trend was not observed for the other spots, highlighting the inherent complexity of the PUR elastomer degradation and its related discoloration over time.

**Microscopic and SEM–EDX investigations**

The appearance of the pale pink core differed from the yellow surface of the mask. In Fig. 6a, the pinkish part appears smooth, homogeneous and translucent, with only a few fibers attached on it. Instead, the yellowish surface (Fig. 6b) appears dirty with several soiling particles. The stickiness, a degradation phenomenon also observed in plastics like PVC, is likely caused by the migration of plasticizers to the surface, mainly in the liquid form such as bis(2-ethylhexyl)phthalate [39, 40].

The observation of the sample under SEM emphasized the two different morphologies between the consistent pale pink core (P) and heterogeneous yellow surface (Y) (Fig. 6c). The EDX analysis identified mainly carbon and oxygen for the pale pink area. Whereas the yellowed surface is characterized by further elements as magnesium (Mg), aluminium (Al) and silicon (Si), indicating the likely presence of aluminosilicates adhering to the sticky surface.

**Raman microscopy (μ-Raman)**

Only whitish particles were observed within the PUR elastomer under the coupled microscope, identified as Pigment White (PW) 6 (titanium dioxide, TiO$_2$, C.I. n. 77891) as rutile and anatase crystalline form by μ-Raman. As depicted in Fig. 7, overlapping signals from the two crystalline Raman signatures were detected. In the Raman spectrum, three bands of rutile (R) at 226 (multi-photon process), 441 (E$_g$) and 614 (A$_{1g}$) cm$^{-1}$ are observed, while only the most intense Raman-active optical phonon modes at 141 (E$_g$) cm$^{-1}$ characterize the presence of anatase (A) [41–44]. Pigments based on TiO$_2$ are the preferred white pigments used for polymers, including PUR [45]. The detection of both TiO$_2$ crystalline forms reveals the use of this polymorphic mixture for the mask’s pigmentation. The photochemistry of TiO$_2$ has been subject of detailed research [46]. Studies have demonstrated how its presence could influence the photo-stability of several polymers [47–51]. In coloring plastics, the different photo-active properties of the two crystalline forms are well known [52]. Chen et al. [53] demonstrated the strong photo-sensitizing effect of anatase and, on the contrary, the photo-stabilizing properties of rutile against photo-oxidation of PUR under UV irradiation. The presence of the crystalline form anatase may impart a greater susceptibility of the elastomer to photo-oxidation.

No other pigments were identified in the PUR sample. A strong fluorescent background characterized the degraded polymeric matrix, limiting the μ-Raman investigation and probably detecting the red component of the pink color. However, a possible composition of the pink pigment could be a mixture of TiO$_2$ with iron oxides, Pigment Red (PR) 101 and Pigment Yellow (PY) 42 (and traces of Pigment Black (PBk) 7, carbon black), as reported in the safety data sheet of the pink pigment produced by the Japanese manufacturer of the PUR elastomer [54]. These pigments are known in the literature for the coloring of PUR [45]. The inorganic pigments used for the pink coloring could have been dispersed in small quantities and are known for having a small size (10–30 µm) [45], which made their identification in polymeric matrix a challenge. No differences between the pale pink core and the yellow surface were observed.

**ATR-FTIR**

Figure 8 shows the spectra of the pale pink core and the yellow surface of the mask. The band assignments are reported in Table 2. Similar absorption bands characterize the two samples, with the characteristic stretching vibration C–O–C at 1100 cm$^{-1}$ of the polyether-based PUR [18, 39, 55]. Other typical features are: (1) the bands at 2966 cm$^{-1}$, 2928 cm$^{-1}$ and 2870 cm$^{-1}$ of CH$_3$ and CH$_2$ stretching; (2) 1727 cm$^{-1}$ band of the urethane C=O group; (3) 1530 cm$^{-1}$ band of the N–H bend vibration with C–N stretch vibration in the –C–NH group (amide...
II band); 4) 1273 cm\(^{-1}\) band of the C–O stretch [19, 38, 52–58]. However, the typical band between 3340 and 3320 cm\(^{-1}\) of the stretching vibration of N–H group is weak. This could indicate the loss of the urethane structures due to UV exposure [19, 59] or the low amount of urethane bonds in the plastic formulation [11].

Besides the characteristic IR bands of the PUR elastomer, slight differences were recognized between the pale pink core and yellowed surface, as highlighted by the negative and positive intensities in the spectrum (Fig. 8). Especially for the yellowed surface, the band at 3674 cm\(^{-1}\) of hydroxyl stretching band and the peaks at 1039 cm\(^{-1}\) and 1015 cm\(^{-1}\) ascribed to in-plane Si–O vibrations, could indicate the presence of aluminum silicates (e.g. kaolinite) on the surface as attached dust particle [23], which confirms the EDX analysis (dust presence).

Moreover, when compared with the bands of the pale pink core, the spectrum of yellowed sample denotes a slight decrease of intensity of the following bands: (1) C–H stretch vibrations in the 3000–2800 cm\(^{-1}\) region attributed to the vibrations of the methylene groups; (2) the urethane C=O stretch free at 1727 cm\(^{-1}\); (3) the band at 1100 cm\(^{-1}\) of the C–O–C stretching vibrations of the ether group (Additional file 1: Fig. S3) [19, 37]. These changes can be correlated with the beginning of the production of quinone structures, which led to yellowing [19]. It is known that the aromatic structures of PUR are oxidized by UV radiation in the central methylene group, resulting in conjugated quinones and, therefore in the formation of colored products and loss of physical and mechanical properties [10, 19, 37].
Both spectra (pale pink core and yellowed surface) present characteristic absorption bands of phthalate. As shown for the reference spectrum of di(2-ethylhexyl) phthalate (DEHP) (Fig. 8, blue line), the main absorptions are: 1727 cm$^{-1}$ (ν C=O of O=C–O), 1600 cm$^{-1}$ and 1580 cm$^{-1}$ (ν C=C of benzene ring), 1464 cm$^{-1}$ (ν C–H), 1273 cm$^{-1}$ (ν C–O of O=C–O), the shoulder at 1077 cm$^{-1}$ (ν C–O of O–CH$_2$) and 744 cm$^{-1}$ (ν C–H) [15, 39, 60, 61]. The band at 744 cm$^{-1}$, used to discriminate the presence of phthalate from the polymeric matrix, resulted slightly increased in the yellowed surface compared to the pale pink core, a signal of the plasticizer migration to the surface (Additional file 1: Fig. S4). No FTIR signals correlated with the presence of pigments were observed in PUR samples, probably because of the low pigment concentration, well below the detection limit of the technique (around <3%) [62].

The spectrum of the glue found on the reverse side of the mask is also shown in Fig. 8. The absorbance bands are comparable to those reported for the mask samples, which determine its composition as a PUR ether-based adhesive. Further bands are at 3294 cm$^{-1}$ (stretching vibration of N–H), 1534 cm$^{-1}$ (amide II band) and at 1224 cm$^{-1}$ (amide III) [19, 37, 63]. Moreover, the main absorption bands for phthalate were also identified at the glue’s surface. The Japanese team probably decided to use a PUR glue chemically compatible with the PUR elastomer. PUR adhesives are extensively used in several applications for their good adhesion on many substrates, good toughness, excellent flexibility, durability, resistance to water, and a broad range of chemicals [64–66]. PUR is known to form hydrogen bonds and covalent bonds when active hydrogen is present [20]. As described in the previous section (see The SAYA mask), the areas with an internal layer of glue have a pale pink color similar to the original one. This fact could suggest the positive effect of the glue to act as a barrier against oxygen, slowing down the yellowing process that characterizes the other areas of the mask. Further research is needed to investigate this aspect.

The spectrum of the sample from the skull revealed to be made of glass fiber reinforced polyester, as presented and discussed in Additional file 1: Fig. S5 and Table S2.

EGA-MS
Figure 9 (top) shows the EGA-MS thermograms of the pale pink core and the yellow surface. For both samples, two thermal decomposition zones are identified: zone 1 from 160 to 330 °C and zone 2 from 330 to 500 °C. The first band describes the evolved gas emitted by additives, with the most abundant ions in the mass spectrum of $m/z$ 43, 57, 71, 85, 127, 149, 167, 279 and 293 (Fig. 9a, c). These ions suggested the presence of phthalates used as plasticizers, in particular diononyl phthalate (DINP, quantitative ion $m/z$ 293, confirmation ions $m/z$ 149, 267) and DEHP (quantitative ion $m/z$ 279, confirmation ions $m/z$ 149, 167) [67, 68]. The second band is related to the pyrolysis of the PUR,

\[ \text{2 The presence of DINP and DEHP in the PUR elastomer were confirmed by TD-GC/MS analysis (see the paragraph Double-shot – GC/MS).} \]
characterized by the ions \( m/z \) 43, 59, 73, 91, 104 and 117 (Fig. 9b, d). La Nasa et al. [17] and Neumann et al. [69] reported that these fragments are attributable to the soft segments of polyether polyols of PUR. The presence of the hard segment of PUR, whose volatilization presumably occurred between 250 and 300 °C [70], was not identified probably due to the too intense signal of the additives.

Focusing on the mass spectra, the thermal zones of the two samples are similar (Fig. 9a–d). However, their EGA profiles present differences. While the temperature of the second step of the polymer did not change with a peak centered at 404 °C, the temperature of first step related to the evolution of additives slightly shifted from 268 °C for the pale pink core to 273 °C for the yellowed surface. This shift could be due to the two main phthalates present in different ratios in the bulk and at the surface (i.e. their different speed of migration towards the surface or their different evaporation rates). Moreover, the ratio between the intensities of the bands changed: the pale pink inner core sample is characterized by a less intense band of additives than that of the polymer; the yellowed surface sample, instead, presents a more intense additive band and a reduction of the band related to the polymer. These variations could be due to the mask aging, especially regarding the migration of the plasticizer and, its consequent enrichment to the surface of the mask, causing the stickiness phenomenon [71]. Plasticizers in the formulation were necessary to create soft and elastic PUR elastomer material to reproduce the human skin.

### Table 2 Vibrational band assignments for the ATR-FTIR spectra of the pale pink core and yellow surface of the SAYA mask

| Wavenumber (cm\(^{-1}\)) | Assignment | References |
|-------------------------|------------|------------|
| 3674                    | \( v \) O–H free | [23]        |
| 3350–3250               | \( v \) NH | [13, 19, 39, 59] |
| 2966                    | \( v \)a C–H\(_3\) | [13, 15, 17–19, 38, 60] |
| 2928                    | \( v \)a C–H\(_2\) | [13, 15, 17–19, 38, 60] |
| 2870                    | \( v \)s C–H\(_3\) | [13, 15, 17–19, 38, 60] |
| 1727                    | \( v \) C=O | [13, 38, 56, 61] |
| 1649                    | \( v \) C=C in alkanes | [21, 62] |
| 1600                    | \( v \) C=C (benzene ring) | [13, 15, 17–19, 39, 56, 60, 61] |
| 1580                    | \( v \) C=C (benzene ring) | [13, 17, 18, 39, 56, 60, 61] |
| 1530                    | \( \delta \) O=\(C\)–N–H, \( v \) O=\(C\)–N–H | [17–19, 39] |
| 1454                    | \( \delta \)a C–H\(_3\) | [13, 17, 18, 38] |
| 1373                    | \( \delta \)s C–H\(_3\) | [13, 17, 18, 38] |
| 1345                    | \( \omega \) C–H\(_2\) | [13, 17, 18] |
| 1273                    | \( v \) C–O | [60, 61] |
| 1100                    | \( va \) C–O–C Polyether-based polyol | [15, 18, 38, 39, 55] |
| 1077                    | \( v \) C–O | [38, 39, 60, 61] |
| 1039                    | Si–O | [23] |
| 1015                    | Si–O | [23] |
| 933                     | \( p \) C–H\(_3\) | [17, 18, 38] |
| 744                     | \( v \) C–H | [38, 39, 60, 61] |
| 669                     | Amide \( V \) urethane seg | [37] |

### Double-shot-GC/MS

Figure 10 shows the volatile organic compounds detected with the first step of the analysis (TD-GC/MS) of the pale pink core and yellow surface. The peak identification is reported in Table 3. The presence of the phthalates DEHP (n° 19) and DINP (n° 21) as plasticizers was confirmed and traces of diethyl phthalate (DEP, n° 12) and dibutyl phthalate (DBP, n° 16) were also identified. Moreover, the UV stabilizer 2-(2′,3,5-di-tert-amylphenyl) benzotriazole (Tinuvin 328, n° 20) was detected, as well as the principal components of the polymer, e.g. propylenglycol (n° 3), disopropyl ether (n° 4) and methylene diphenyl diisocyanate (MDI) (n° 17). The identification of MDI supports the finding of FT-IR analysis and corroborates the photo-oxidation of the PUR mask and, therefore, its visible yellowing [19]. Besides PUR, specific fragments of another polymer were recognized: small peaks, which became more intense in the pyrograms, related to styrene-acrylonitrile (SAN, n° 6, 10,
Fig. 9  EGA-thermograms of the pale pink core (red line) and the yellow surface (black line) of PUR mask. The mass spectra obtained for zone 1 (160–330 °C) and zone 2 (330–500 °C) are also reported.
11, 13–15, 18) were identified. The presence of SAN is discussed later in the text.

Considering the results of the pale pink core and the yellowed surface, a relative increase of the abundance of the main additives DEHP (n° 19), Tinuvin 328 (n° 20) and DINP (n° 21) was detected on the yellow surface (see Additional file 1: Table S3). These results confirm those obtained from the EGA-MS analysis regarding the additive migrations to the mask’s surface. This degradation process is already well known for plastics with a high amount of additives, such as PVC [39, 72, 73].

Figure 11 shows the pyrograms of the mask obtained in the second step (Py-GC/MS analysis) at 600 °C. The relative peaks are reported in the Table 3. Diisopropyl ether (n° 26) and 2-propanone, 1-(1-methylethoxy)-(n° 27) were the main compounds from the PUR polymer. All the other fragments, instead, belonged to the pyrolysis of the SAN amount, in particular: acrylonitrile (n° 23), styrene (n° 29), α-methylstyrene (n° 31), 4-phenylbutyronitrile (n° 33), SA: 4-phenylpent-4-enenitrile (n° 34), 2-methylene-4-phenethylpentadinitrile (n° 35), 2-methylene-4-phenylheptanedinitrile (n° 36), 2-(2-phenylallyl) pentanedinitrile (n° 37) and 2-phenethyl-4-phenylpent-4-enenitrile (n° 38) [27]. The presence of SAN in PUR museum objects was already observed by Izzo et al. [29], as an additive capable of improving the mechanical properties of PUR polymers [74]. Its presence, however, could have also influenced the photo-stability of the PUR elastomer negatively. SAN is known to turn yellow in color when exposed to light and oxygen due to photo-oxidative degradation of the polystyrene component [75, 76].

When comparing the two sample’s compositions, no significant differences were detected. This could mean that the degradation process of PUR mask is still at an initial stage and has not yet caused evident chemical alterations in the polymeric matrix composition.

**Assessment of the adhesive treatment effectiveness**

The results of the adhesion tests are summarized in Table 4. The cellulose-based adhesives gave good results...
on SAYA PUR elastomer, with the exception of Methocel A 15 VL, while not good adhesion was found for the PP mannequin. The best results were obtained with the wheat starch paste and PU 52 in both supports. The PUR dispersion was the unique adhesive with good flexibility. Because of its good adhesion properties and chemical compatibility with the mask’s composition, a complete treatment of the tears was suggested to the museum staff as adequate.

### Table 3

Major peak identification in the TD-step (from 50 to 320 °C) and in the Py-step (600 °C) of the pale pink core and yellow surface of PUR elastomer

| Peak number | Peak identification | Main ions (m/z) |
|-------------|---------------------|-----------------|
| TD-step     |                     |                 |
| 1           | Acetaldehyde        | 40, 41, 42, 43, 44 |
| 2           | Propylene oxide     | 29, 43, 58      |
| 3           | Propyleneglycol     | 43, 45, 76      |
| 4           | Diisopropyl ether   | 44, 45, 87, 102 |
| 5           | 2-Ethyl-1-hexene    | 41, 42, 55, 70, 112 |
| 6           | Styrene             | 51, 77, 78, 104 |
| 7           | 2-Ethylhexanol      | 41, 55, 57, 70, 83, 98, 130 |
| 8           | 1-Nonanol           | 29, 41, 56, 70, 83, 97, 126 |
| 9           | Phthalic anhydride  | 50, 76, 104, 148 |
| 10          | 4-Phenylbutyronitrile | 51, 65, 77, 91, 104, 115, 145 |
| 11          | 4-Phenylpent-4-enenitrile (hybrid dimer) | 51, 63, 77, 91, 103, 115, 117, 130, 142, 157 |
| 12          | Diethyl phthalate (DEP) | 50, 65, 76, 105, 149, 177, 222 |
| 13          | 2-Methylene-4-phenyl heptanedinitrile (hybrid trimer) | 51, 65, 77, 91, 105, 156, 209, 210 |
| 14          | 2-Methylene-4-phenethyl pentanedinitrile (hybrid trimer) | 51, 65, 77, 91, 104, 117, 128, 144, 210 |
| 15          | 2-(2-Phenylallyl)pentadinitrile (hybrid trimer) | 51, 65, 77, 91, 104, 118, 129, 144, 156, 170, 198, 210 |
| 16          | Diethyl phthalate (DBP) | 76, 104, 149, 204, 278 |
| 17          | Methylenediphenyl disocyanate (MDI) | 77, 132, 165, 180, 208, 222, 250 |
| 18          | 2-Phenethyl-4-phenylpent-4-enenitrile (hybrid trimer) | 51, 65, 77, 91, 105, 115, 118, 129, 142, 156, 170, 261 |
| 19          | Bis(2-ethylhexyl)phthalate (DEHP) | 43, 57, 71, 104, 149, 167, 279, 390 |
| 20          | 2-(2′-Hydroxy,3′,5′-di-tert-amylphenyl)benzotriazole (Tinuvin 328) | 43, 71, 92, 149, 252, 322, 351 |
| 21          | Diisononyl phthalate (DINP) | 43, 57, 71, 85, 97, 127, 149, 167, 293, 418 |
| Py-step     |                     |                 |
| 22          | Propylene           | 38, 39, 40, 41, 42 |
| 23          | Acrylonitrile       | 50, 51, 52, 53 |
| 24          | Metacrylonitrile    | 39, 41, 52, 67 |
| 25          | Pentenenitrile      | 39, 41, 54, 66, 81 |
| 26          | Diisopropyl ether   | 41, 43, 45, 59, 87, 102 |
| 27          | 2-Propanone, 1-(1-methylethoxy)- | 41, 43, 58, 73, 116 |
| 28          | Ethylbenzene        | 51, 65, 77, 91, 106 |
| 29          | Styrene             | 51, 63, 78, 104 |
| 30          | Isopropylbenzene    | 51, 77, 91, 105, 120 |
| 31          | α-Methylstylene     | 51, 63, 78, 91, 103, 118 |
| 32          | 2-Methylenepentanedinitrile | 52, 66, 79, 106 |
| 33          | 4-Phenylbutyronitrile | 51, 65, 77, 91, 104, 115, 145 |
| 34          | 4-Phenylpent-4-enenitrile (hybrid dimer) | 51, 63, 77, 91, 103, 115, 130, 142, 157 |
| 35          | 2-Methylene-4-phenethyl pentanedinitrile (hybrid trimer) | 51, 65, 77, 91, 105, 156, 209, 210 |
| 36          | 2-Methylene-4-phenyl heptanedinitrile (hybrid trimer) | 51, 65, 77, 91, 104, 117, 144, 210 |
| 37          | 2-(2-Phenylallyl)pentadinitrile (hybrid trimer) | 51, 57, 65, 77, 91, 104, 118, 129, 144, 156, 170, 198, 210 |
| 38          | 2-Phenethyl-4-phenylpent-4-enenitrile (hybrid trimer) | 51, 65, 77, 91, 105, 115, 118, 129, 142, 156, 170, 261 |

Characteristic ions in mass spectra: molecular weight (M_w) bold, base peak underlined
Conclusions

The multi-analytical approach applied in this study allowed a comprehensive characterization of the material composition of SAYA’s facial/skin mask and its degradation. This study confirms how the joint application of in situ ATR-FTIR and µ-Raman, with µ-destructive EGA-MS and TD/Py–GC/MS analyses is a successful approach for addressing unknown complex plastic formulation at any level of degradation to be applied also in future studies.

This paper highlights the inherent complexity and chemical instability of PUR elastomer-based artifacts, and the color change detected within 5 months of monitoring pointed out a still ongoing degradation. After only 10 years from the production, this alteration extent is likely a result of the chemical composition of the mask, which makes it deeply susceptible to changes.

Based on the results of this study and literature discussion, two main degradation processes might be assumed: (a) photo-oxidation of the MDI-PUR elastomer, responsible for the yellowing and loss of physical and mechanical properties, enhanced by the presence of anatase form and SAN in the formulation; (b) migration of UV stabilizer and phthalates to the surface, which, especially for phthalates (DINP and DEHP), caused a widespread stickiness and the consequent attachment of dust particles. The role of UV-stabilizer in the color change needs still clarification as literature suggests decay leading to possible yellowing [77, 78].

Table 4  Results of the adhesion and flexibility tests

| Adhesive                    | Adhesion on PUR | Adhesion on PP | Flexibility |
|-----------------------------|-----------------|---------------|-------------|
| Methocel A 15 LV (5%)       | 0               | -             | -           |
| Tylose MH 300 (5%)          | +               | 0             | 0           |
| Tylose MH 1000 (5%)         | +               | 0             | -           |
| Wheat starch paste (10%)    | +               | +             | 0           |
| PUR dispersion PU 52        | +               | +             | +           |

+ : good; 0 : mediocre; -- : poor
The specific knowledge of SAYA’s mask composition and degradation supported interdisciplinary discussions with the team of curators and conservators of the Deutsches Museum. Even though a possible adhesion treatment could be suggested to minimize further degradation, additional research is necessary, including the long-term stability of the treatment through accelerated aging experiments.

Abbreviations
BSD: Backscatter electron detector; CIE: Commission internationale de l’éclairage; DBP: Dibutyl phthalate; DEHP: Bis(2-ethylhexyl)phthalate; DEP: Diethyl phthalate; DINP: Disononyl phthalate; EGA-MS: Evolved gas analysis-mass spectrometry; FRP: Fiber reinforced plastic; FTIR-ATR: Fourier transform infrared spectroscopy in attenuated total reflection; MDI: Methylene diphenyl diisocyanate; Mₜ: Molecular weight; PP: Polypropylene; PUR: Polyurethane; PVC: Polyvinylchloride; SAN: Styrene acrylonitrile; SEM–EDX: Scanning electron microscope–energy dispersive X-ray spectroscopy; TD/Py–GC/MS: Thermal desorption/pyrolysis–gas chromatography/mass spectrometry; UV: Ultraviolet.

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Authors’ contributions
MP and AM defined the research project and designed the experiments. AM, MP and EMS performed the analyses, the data analysis and interpretation. JS performed the conservation treatment and gave recommendations for preventive conservation and storage. AM wrote the first draft manuscript and all co-authors discussed the results and commented on the manuscript. All authors reviewed, edited the final manuscript. All authors read and approved the final manuscript.

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Availability of data and materials
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Declarations
Competing interests
The authors declare that they have no competing interests.

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