Article

Thermodynamic Characterization of Free and Surface Water of Colloidal Unimolecular Polymer (CUP) Particles Utilizing DSC

Peng Geng, Ashish Zore and Michael R. Van De Mark *

Department of Chemistry, Missouri S&T Coatings Institute, Missouri University of Science and Technology, Roll, MO 65409, USA; pgkr4@mst.edu (P.G.); aszbnd@mst.edu (A.Z.)
* Correspondence: mvandema@mst.edu

Received: 28 May 2020; Accepted: 22 June 2020; Published: 24 June 2020

Abstract: Colloidal Unimolecular Polymer (CUP) particles are spheroidal, 3–9 nm with charged groups on the surface and a hydrophobic core, which offer a larger surface water fraction to improve the analysis of its characteristics. Differential scanning calorimetry (DSC) was performed to determine the characteristics of surface water. These properties include the amount of surface water, the layer thickness, density, specific heat of the surface water above and below the freezing point of water, melting point depression of free water, effect of charge density and particle size. The charge density on the CUP surface was varied as well as the molecular weight which controls the particle diameter. The surface water is proportional to the weight fraction of CUP <20%. Analogous to recrystallization the CUP particles were trapped in the ice when rapidly cooled but slow cooling excluded the CUP, causing inter-molecular counterion condensation and less surface water. The density of surface water was calculated to be 1.023 g/mL to 1.056 g/mL depending on the surface charge density. The thickness of surface water increased with surface charge density. The specific heat of surface water was found to be 3.04 to 3.07 J/g·K at 253.15 K and 3.07 to 3.09 J/g·K at 293.15 K. The average area occupied by carboxylate and ester groups on the CUP surface were determined.

Keywords: Colloidal Unimolecular Polymer (CUP); Differential Scanning Calorimetry (DSC); surface water; density; heat of fusion; thickness; cooling rate; specific heat; melting point depression; counterion condensation

1. Introduction

The freezing of water has been an extensively studied thermodynamic property. The freeze thaw of sperm, eggs, cryogenic preservation, crop freezing, food preservation by freezing, paint freeze thaw stability, road and walkway ice and aircraft icing are just a few examples of where ice formation is a critical issue. Surface associated water plays an important role in most of these examples. However, the study of surface water has been difficult due to the low ratio of surface to free water present in a system. Nano particles can offer a significantly enhanced window into surface water due to the high surface water to particle weight ratio.

Colloidal Unimolecular Polymer (CUP) particles are a new class of spheroidal nanoscale polymer particles (3–9 nm) with charged hydrophilic groups on the surface and a hydrophobic backbone [1]. These nano particles can be inexpensively and easily synthesized, particle size can be varied with narrow particle size dispersity by controlling the molecular weight and different surface charge density can be designed with both positive and negative charges, this will provide a very predictable, controllable and reproducible system. Also, CUP particles are spheroidal with charges on the surface but unlike latex particles, CUP solutions are free of surfactant or additives and are zero VOC, making it a promising model
for fundamental scientific studies. The CUP surface has a layer of surface associated or bound water, which does not freeze until a much lower temperature than free water [2–6], shown in Figure 1.

![Figure 1. Colloidal Unimolecular Polymer (CUP) particles with surface water.](image)

Due to the small particle size, CUP’s surface area per gram is ultra-high, giving a high surface area to volume ratio and surface functionality [7]. The effect of surface water can be neglected when the size of the particle is very large. The size of a typical latex particle is about 100 nm and the diameter of water molecule is only 0.28 nm. Assuming that there is one layer of water bounded on the particle surface, the volume ratio of bound water and latex particle is only 1.69%. However, when the particle is as small as 3.90 nm, the ratio is 49.56% [1], which makes the contribution of surface water much more significant as shown in Figure 2.

![Figure 2. Comparison of latex, polyurethane dispersion (PUD) and CUP (25.4k).](image)

These facts make CUP an ideal model for surface water studies, offering a huge advantage over proteins which are limited in size, structure and availability [7]. CUPs can contribute to our understanding of proteins and micelles and also contribute to the study of other materials with water interfaces. Zero VOC CUPs are a very good candidate for future applications including coatings, adhesives, sealants and many others.

The formation of CUP particles is generally accomplished through a water reduction process, according to the Flory-Huggins theory [8]. The process is driven by the polymer-polymer interaction being greater than the polymer-solvent interaction and entropically favored by releasing water analogously to micelle formation. The charged groups repel each other to create the spheroidal shape. Once formed, CUPs are thermodynamically stable in water solution.

CUP solutions are made through a water reduction process, as shown in Figure 3. The brown spheres represent the hydrophobic polymer backbone, each sphere is a methyl methacrylate unit (or hydrophobic monomer). While the gray spheres represent the ionizable carboxylic acid side-chain groups (or methacrylic acid unit). Polymers were dissolved in a low boiling water miscible solvent,
like THF, stirred overnight to ensure all polymer chains are in a random coil configuration, Structure I. Base was slowly added to the solution to pH 8.5 forming carboxylate salts and the chains became more extended due to the charge repulsion, Structure II. As pH adjusted water was gradually added, the sodium carboxylate ions became solvated and separated. The repulsion between adjacent ions increased due to the increasing dielectric caused by the added water and the chain extended toward linearity which increased the viscosity [9], Structure III. At a critical water to THF ratio the Mark-Houwink parameter reaches the highest value and the polymer–polymer interaction became greater than the polymer–solvent interaction, the carboxylate groups oriented into the water phase, organizing to produce maximum separation of charge. Hence, water released from the polymer backbone increases the entropy and, the hydrophobic polymer chain collapses into spheroidal shape, Structure IV. Finally, the low boiling solvent, THF was stripped off under reduced pressure. The presence of ionic groups on the surface is the driving force to prevent the particles from aggregating through charge-charge repulsion [10]. Once formed, these colloidal solutions are thermodynamically stable [11]. The ratio of hydrophobic and hydrophilic monomers on the polymer chain (HLB value) is very critical in the unimolecular collapse of the polymer chain during water reduction process. If the chains are too hydrophobic, the collapsed chain tend to aggregate and if the chains are too hydrophilic, the polymer will dissolve or form a more extended or barbell like structure [1,12,13].

The term “surface water” has been used since 1922 [14]. Ever since that, many techniques have been used to investigate the surface water. Previous studies suggest that water molecules were arranged in several layers adjacent to a solid interface changing the properties of the water [15–25]. The state of water has been widely investigated by various techniques.

Van De Mark et al. correlated the surface water with the rheological behavior of CUP particles [10]. They found that from the intrinsic viscosity of the CUP solution, by combining the density and molecular weight of CUPs, the thickness of water layer was estimated to be 0.57 nm, which is similar to the case of protein [2] and cylindrical nanoropes [26].

Nuclear magnetic resonance (NMR) is another method which has been used to examine surface water. The mobility of surface water molecules is lower than free water molecules, therefore NMR probing of the mobility has been used to verify the existence of structured water around colloidal polyvinyl acetate (PVA) with particle size of 0.13 and 0.8 micron used by Clifford et al. By measuring the spin lattice relaxation time constants of protons, they concluded that the total amount of bound water per unit surface area was larger for larger particles [27–29].
Katayama et al. investigated the states of water in a polyacrylamide gel using spin-lattice relaxation time constant measurements and concluded that the macromolecule must have a hydrophilic substituent to be able to capture surface water and the amount of surface water remarkably depends on the amount and the nature of the substituents [30]. Van De Mark et al. used the same method to determine the surface water thickness of CUP particles, which ranged from 0.19 nm to 0.693 nm and it was found to be both molecular weight and temperature dependent [31].

Quasielastic neutron scattering (QENS) has been used for both organic and inorganic water-containing materials. Mamnontov used neutron scattering to investigate the vibrational dynamics of surface water in ZrO$_2$. The rotational diffusion of surface water molecules was found to be slower by about a factor of 2 when compared with free water and the residence time for translational diffusion of surface water was about 40 times longer than free water. It was proposed that there were about two hydration layers on top of the layer of surface OH groups [32].

Toney et al. measured the water density profile perpendicular to silver (III) at two voltages by using X-ray scattering and found that the first inner layer of surface water has a greater density compared with free water, 1.1 ($-0.23$ V) to 1.8 ($+0.52$ V) water molecules per Ag atom for surface water, while they expected 0.8 water molecules per Ag atom, based on the density of free water. Which indicated that the hydrogen-boding network changed and resulted in very different properties in this layer from those in free water [33,34].

Drost-Hansen et al. studied the water adjacent to silica surfaces by using differential thermal analysis (DTA). They observed thermal property changes in the slope and displacements of the baseline in thermograms and small endothermic peaks in heating curves. These peaks were associated with structural phenomena changes of the water at silica/water interface, indicating the properties of water close to solid interfaces are very different from free water [35].

Velazquez et al. described the interaction of water with hydrophilic materials indicating that a portion of water was firmly bounded to individual sites. By using infrared spectroscopy, the bound water amount on a methylcellulose film was identified. It was reported that the bending vibration mode of bound water shifted toward lower frequency by about 15.5 cm$^{-1}$. When the humidity increased, the bound water content remained at below 5% [36].

Differential scanning calorimetry (DSC) has been widely used to investigate detailed information on the state of water by distinguishing the amount of freezeable water from non-freezeable water for many water absorbed system [37–43], Ross [44], Bushuk et al. [45] and Biswas et al. [46] all concluded that the amount of unfrozen water per gram of solute increased with the increasing dilution, this dependence was more obvious for concentrations smaller than 1 M. Berlin et al. studied whey protein systems by using DSC and determined that 0.5 g of water per gram of whey protein would not freeze at $-40$ °C. By adding lactose and salts, the unfreezeable water increased as the concentration of lactose and salts were increased, which varied between 0.5 and 1.2 g of water per gram of solids [47,48]. Hatakeyema et al. quantitatively calculated the amount of bound water in poly(vinyl alcohol) hydrogel using DSC. It was reported that each hydroxyl group of PVA can associate with 1–1.5 molecules of non–freezeable water and 5–6 molecules of freezeable bound water [49]. Ostrowska-Czubenko et al. analyzed the state of water in chitosan hydrogel membranes and found that freezeable water content increased linearly with the water uptake, while non-freezeable water content remained constant beyond critical water content value. The non-freezeable surface water ranged from 0.47 to 0.65 g per gram of dry membrane [50]. Muffett et al. investigated the amount of unfrozen water in soy proteins by DSC and reported that the unfrozen water amount increased in the range of 0.09–0.14 g of unfrozen water per gram of total water, with the increasing of total water [51]. Garti et al. determined the thickness of bound water layer in the n-dodecane/1-pentanol/C$_{12}$(EO)$_8$ system to be 0.5 nm by using DSC [40]. Kobayashi et al. determined the thickness of fully hydrated dimyristoylphosphatidylcholine to be 3.2–3.4 nm, using DSC, X-ray and densimetry [52].

Many of the previous surface water studies utilizing DSC gives confidence to investigate surface water properties of CUP system. DSC has many advantages over other techniques for evaluating the
surface water behavior. First, DSC is the most common thermal analysis technique to obtain heat content change (enthalpy) and heat capacity with ease and speed. Second, DSC can hold a precise temperature with no drift and it can be used for kinetic studies in a faster and more straightforward way than other methods [53–55]. In this study, using hermetically sealed pans and sample size of 30 mg, accurate water melting and specific heat data should be readily obtainable for even low weight fraction of CUP solutions.

CUP opens a window to investigate the thermodynamics of surface water and free water for particulate systems. This work presents a primary study of the CUP surface water properties utilizing DSC. It includes—synthesis and production of CUP through water reduction; determination of the thickness of surface water of CUP by measuring the heat of fusion and particle size; determine the actual density of surface water; determine the specific heat of surface water; evaluate the effect of cooling rate on the amount of surface water; determine the average surface area of functional groups on the CUP surface by knowing the melting point depression of CUP solutions; study the relationship between CUP surface water and molecular weight and charge density in ions per nm$^2$. This work represents the first comprehensive study of the functionality and size effects on CUP surface water behavior during freeze/thaw conditions.

2. Materials and Methods

2.1. Materials

Methyl methacrylate (MMA), methacrylic acid (MAA), 2,2′-azobis(2-methylpropionitrile) (AIBN) and 1-dodecanethiol were purchased from Aldrich (St Louis, MO, USA). MMA was purified by washing with a 10% (w/w) solution of sodium bicarbonate, followed by rinsing with de-ionized water and then brine and then dried over sodium sulfate and filtered. Copper (I) bromide was added to the MMA as an inhibitor and simple distillation under nitrogen was carried out. MAA was purified by distillation with copper (I) bromide under reduced pressure. THF was dried and distilled under protection of nitrogen. AIBN was re-crystallized from methanol and 1-dodecanethiol was used as received.

2.2. Synthesis of Poly(MMA/MAA) Copolymer

Polymers were synthesized by a free radical polymerization method in tetrahydrofuran (THF) [21]. To a 2 L three neck flask, 10.40 mol of THF was added. Methyl methacrylate (MMA) and methacrylic acid (MAA) were used with various molar ratios. 1-Dodecanethiol was added as a chain transfer agent based on desired molecular weight of polymer. The initiator AIBN was then added, 0.0007 times the total moles of monomers. The reaction was carried out under nitrogen, with condenser and a gas outlet adapter connected to an oil bubbler to allow a positive flow of nitrogen throughout the polymerization. The mixture was heated slowly to reflux under stirring for 24 h. The polymer solutions were then cooled to room temperature and part of the THF was removed by rotovap. Finally, the polymer was precipitated in cold deionized water under high shearing rate and dried in a 323.15 K oven under vacuum for 24 h. Table 1 shows the component amount for the synthesis of Polymers 1–7.

2.3. Water Reduction Method

To a 500 mL Erlenmeyer flask were added 10 g of dry polymer, 40 g of THF and stirred for 12 h to let the polymer chains fully obtain a random coil configuration. Based on the measured acid number, 1 M NaOH solution was slowly added to neutralize the solution to pH 8.5 by peristaltic pump at the rate of 1.24 g/min. Then 90 g of deionized water modified to pH = 8.5 using 1 M NaOH solution was then gradually added by peristaltic pump at the rate of 1.24 g/min. The pH of system was maintained at 8.5 throughout the process of water reduction. Majority of THF was stripped off by rotovap and the sample was left in vacuum to remove the last trace of THF, giving zero VOC CUP solution. The clear solutions were then filtered through a 0.45 micron filter to remove any extraneous trace particulate contaminants, the typical loss on filtering was less than 0.05% of the solids by weight. The CUP weight
percent solids was determined and the CUP solution was either diluted by adding pH modified water or concentrated under vacuum to prepare the different CUP solids fractions needed for the study.

Table 1. Polymer synthesis: the amount of materials used.

| Polymer | Molecular Weight (g/mol) | Monomer 1 | Monomer 2 | Initiator | Chain Transfer Agent | Solvent |
|---------|--------------------------|-----------|-----------|-----------|----------------------|---------|
|         |                          | MMA (mol) | MAA (mol) | AIBN (mol)| 1-Dodecanethiol (mol)| THF (mol)|
| 1       | 28,900                   | 2.25      | 0.25      | $1.75 \times 10^{-3}$ | $7.61 \times 10^{-3}$ | 10.40   |
| 2       | 59,800                   | 2.25      | 0.25      | $1.75 \times 10^{-3}$ | $3.79 \times 10^{-3}$ | 10.40   |
| 3       | 122,500                  | 2.25      | 0.25      | $1.75 \times 10^{-3}$ | $1.87 \times 10^{-3}$ | 10.40   |
| 4       | 25,400                   | 2.22      | 0.33      | $1.78 \times 10^{-3}$ | $8.62 \times 10^{-3}$ | 10.40   |
| 5       | 73,500                   | 2.30      | 0.23      | $1.77 \times 10^{-3}$ | $3.10 \times 10^{-3}$ | 10.40   |
| 6       | 49,700                   | 2.35      | 0.17      | $1.76 \times 10^{-3}$ | $4.51 \times 10^{-3}$ | 10.40   |
| 7       | 22,700                   | 2.39      | 0.13      | $1.76 \times 10^{-3}$ | $9.46 \times 10^{-3}$ | 10.40   |

3. Characterization of Polymers

3.1. Absolute Molecular Weight of Copolymers

Gel permeation chromatography utilized a Viscotek model 305 manufactured by Malvern Corp. system (Westborough, MA, USA). The GPC was equipped with a triple detector array TDA305—refractive index detector, low and right-angle light scattering detector and intrinsic viscosity detector, the column, two PAM-505 from PolyAnalytik (London, Ontario, Canada) with a guard column, the column size is 7.5 mm (ID) $\times$ 300 mm (L). Polymer samples were prepared to 2 mg/cc. Injection volume was 100 µL and THF flow rate was 0.5 ml/min.

3.2. Density of CUP Solution

Densities of CUP solutions were directly measured by density meter (DDM 2911 plus by Rudolph Research Analytical (Hackettstown, NJ, USA), at various weight fractions at 298.15 K. The accuracy is 0.00001 g/cm³.

3.3. Density of Dry CUP Polymer

The solutions of CUP were dried in a vacuum oven, heated at 323.15 K with the presence of solid sodium hydroxide to absorb carbon dioxide. After the clear crystal-like material formed, the sample was then heated to 383.15 K to constant weight. The densities of the dry CUPs were measured by a gas displacement pycnometer, Micrometritics AccuPycII 1340. The temperature was controlled at 299.04 ± 0.04 K. Twenty five readings were made for each sample and the results were reported by average and standard deviation.

3.4. Acid Number (AN)

The acid number of the copolymers were determined by the titration method ASTM D974 and reported in mg of KOH/g of polymer sample. The method was modified by using potassium hydrogen phthalate (KHP) in place of hydrochloric acid and phenolphthalein in place of methyl orange. THF was used as the solvent of the titration.

3.5. Viscosity of CUP Solution

Viscosity of CUP solution was measured based on ASTM D445, ASTM-D446 and ISO 3104, 3105. Ubbelohde capillary viscometer J-340 from Cannon instrument company was used to determine the viscosity of CUP solutions at two different temperatures—298.15 K and 302.15 K. Before each
measurement, CUP solution was transferred to the Ubbelohde capillary viscometer and kept in a constant temperature water bath at 298.15 ± 0.1 K for 20 min with plastic wrap covering on top of the viscometer to prevent potential evaporation and carbon dioxide contamination of the solution. A stopwatch with 0.01 s precision was used to monitor the elution time and each measurement was repeated for at least three times and the error being less than 0.5%. Absolute viscosity was then calculated by Equation (1).

\[ \eta = t \cdot \rho \cdot c, \]  

(1)

where \( \eta \) is the viscosity of CUP solution (cP), \( t \) is the elution time (s), \( \rho \) is the density of CUP solution (g/mL) and \( c \) is the Ubbelohde capillary viscometer constant (0.009749 mm\(^2\)/s).

### 3.6. Particle Size of CUP

Particle size was measured by dynamic light scattering, using Microtrac Nanotrac 250 particle size analyzer from Microtrac with a laser diode of 780 nm wavelength and 180° measuring angle. The viscosity of solution was used, due to high weight fraction of CUP having high viscosity that caused by electronic repulsion [10] and low weight fraction of CUP produce very poor light scattering efficiency and a weak signal, because of small particle size, around 5 nm. The CUP solutions were diluted to 10% weight fraction by pH adjusted Mili-Q ultrapure water with resistance of 18.3 MΩ. The particle size was calculated by Stokes-Einstein Equation (2).

\[ D = \frac{k_B \cdot T}{6\pi \cdot \eta \cdot r}, \]  

(2)

where \( k_B \) is the Boltzman constant, \( T \) is the absolute temperature of solution, \( \eta \) is the viscosity of solution and \( r \) is the radius of the CUP particle.

### 3.7. Differential Scanning Calorimetry

Differential scanning calorimeter from TA instruments Q2000 was used to measure the heat of fusion, specific heat and melting point depression of CUP solutions. About 30 mg of CUP samples were sealed in the Tzero Hermetic pan from DSC Consumables Inc., then cooled to the target temperature at a series of cooling rates, isothermal for 10 min and heated up to 313.15 K at 3 K/min rate. The mass of sealed pan was measured before and after each measurement with the result being considered valid if the mass difference was smaller than 0.001 mg.

### 4. Results and Discussion

#### 4.1. Polymer Synthesis and Characterization

It was found that polymers with molecular weight lower than 13,000 g/mol tended to be unstable, due to insufficient electrostatic stabilization causing the polymer to aggregate. Polymers with very high molecular weight, higher than 200,000 g/mol might have issue during water reduction process due to chain–chain entanglement. At high molecular weight the reduction solution must be very dilute since as the molecular weight increases the chance for chain entanglement increases therefore the solutions must be diluted. Thus, seven polymers with different molecular weights (22,700–122,500 g/mol) and different monomer ratios were synthesized and successfully made into CUP solutions. Polymer 1, 2 and 3 have different molecular weight but the same monomer ratio, 9 mol MMA to 1 mol MAA. In order to investigate the behavior at very low charge density values, Polymer 4 and 5 were designed...
to have different molecular weight but same charge density in ions per \( \text{nm}^2 \) as Polymer 2. The charge density \( \rho_v \) was determined by Equation (3).

\[
\rho_v = \frac{M_W}{4\pi \cdot r^2 \left( n_1 \cdot M_{MMA} + n_2 \cdot M_{MAA} \right)}
\]

(3)

where \( \rho_v \) is the charge density in ions per \( \text{nm}^2 \), \( M_W \) is the molecular weight of CUP polymer, \( r \) is the radius of the CUP particle, \( n_1 \) is the moles of MMA and \( n_2 \) is the moles of MAA used per average repeat unit, \( M_{MAA} \) is the molecular weight of monomer methacrylic acid, \( M_{MMA} \) is the molecular weight of monomer methyl methacrylate. Polymer 6 and Polymer 7 were synthesized to have a much higher monomer ratio which was able to investigate the behavior at very low charge density values.

The molecular weight, acid number, particle size and density of the polymers are listed in Table 2. The densities of the dry polymer increased with the increasing of molecular weight, because when molecular weight is larger, the number of polymer chain end groups decrease, result in less free volume and higher density [56]. All polymers had consistent acid numbers with respect to the monomer feed.

Table 2. Molecular weight, particle size, acid number and density of the polymers.

| Sample ID | \( M_n \) (g/mol) | Monomer Ratio | Particle Size (nm) | AN (mg KOH/g) | Density of Dry CUP, \( \rho_p \) (g/mL) | Charge Density in Ions per \( \text{nm}^2 \) |
|-----------|-------------------|---------------|-------------------|---------------|--------------------------------------|------------------------------------------|
| Polymer 1 | 28,900            | 9:1           | 4.22              | 56.8          | 1.2246 ± 0.0018                      | 0.52                                    |
| Polymer 2 | 59,800            | 9:1           | 5.38              | 57.0          | 1.2311 ± 0.0014                      | 0.66                                    |
| Polymer 3 | 122,500           | 9:1           | 6.83              | 56.9          | 1.2342 ± 0.0018                      | 0.84                                    |
| Polymer 4 | 25,400            | 6.8:1         | 4.04              | 73.2          | 1.2243 ± 0.0018                      | 0.66                                    |
| Polymer 5 | 73,500            | 9.8:1         | 5.76              | 52.6          | 1.2315 ± 0.0018                      | 0.66                                    |
| Polymer 6 | 49,700            | 14:1          | 5.06              | 37.7          | 1.2307 ± 0.0016                      | 0.42                                    |
| Polymer 7 | 22,700            | 19:1          | 3.90              | 28.2          | 1.2241 ± 0.0018                      | 0.24                                    |

The polymers were then reduced, the THF removed and the CUP solutions were concentrated. The samples were then measured for particle size and validated as to single chain particles due to a match in particle size with the calculated size and distribution from the absolute molecular weight [1]. The samples were either concentrated or diluted into different weight fractions for DSC analysis.

4.2. Effect of Cooling Rate and Ice Formation

The DSC measurements were done using about 30 mg of CUP solution in Tzero hermetically sealed pans. Enough head space was allowed to avoid rupture or leaking during the freeze thaw cycle and large enough to give an accurate measurement. DSC samples were cooled to 233.15 K and held for 10 min to ensure all freezable water froze and then heated to 313.15 K to allow all the ice to melt without causing leakage. It was difficult to precisely obtain accurate area of the ice formation peak due to some super-cooling during the temperature lowering scan. Therefore, the data presented here represents the heating, melting process only. The mass of the DSC pan was measured before and after to ensure that no water had been lost during the cycle, only runs with losses of less than 0. 001 mg were used.

To evaluate the time required to freeze the sample as well as to determine if the surface water will slowly leave the outer surface and freeze, samples of a 10.03% 28.9k CUP solution, Polymer 1, were cooled to 233.15 K at 10 K/min, isothermal for 10 min, then heated up to 313.15 K at 3 K/min. The same
sample was also measured following the same protocol, except the isothermal time was changed to 1 h, 2 h, 4 h and 8 h. The endothermic peaks represented the measured heat of fusion of freezable water. The same area and shape indicated that all freezable water was able to completely freeze within the 10 min isothermal period, shown in Figure 4. Thus, 10 min was chosen as the experimental isothermal time.

Ice forms typically near 273 K, however, the endothermic peak is somewhat broad so is the freezing peak making baseline determinations near 273 K ± 10 K difficult. Evaluation of the lowest temperature needed was evaluated by running scans at with 10.40% 28.9k CUP solution cooled to 233.15 K, 243.15 K and 253.15 K at 10 K/min from room temperature, isothermal at each temperature for 10 min. Then the sample was heated up to 313.15 K at the rate of 3 K/min. The heat of fusion was unchanged over this range. Which implied that the 233.15 K isothermal temperature could assure all freezable water froze. When water-based latex resins are exposed to cold temperature conditions, water will start to freeze and form ice crystals excluding the latex. As the ice crystals grow, there is less and less liquid water existing in the solution and therefore the weight fraction of latex particles increases in the remaining liquid and the particles are forced close together [57], shown in Figure 5.

Thus, in theory, CUP solutions should perform in a similar way. If the CUP solution was cooled at an infinity slow rate, the distance between each CUP particles will become a minimum, reaching maximum inter-molecular counterion condensation effect and result in the minimum amount of surface water, regardless of initial weight fraction of CUP particles.
In the literature, different freezing methods result in different supercooling effects [58]. For normal protein freeze-drying process, the fastest cooling rate can be obtained by liquid nitrogen freezing with a small volume of sample, that gives the most supercooling; while the lowest supercooling is for the pre-cooled shelf method [59–61].

In order to understand the effect of the cooling rate on the measurement. Variation in the cooling rate was evaluated analogously for Polymer 1. The DSC of CUP solutions were cooled at 0.1 K/min, 0.2 K/min, 0.5 K/min, 1 K/min, 2 K/min, 3 K/min, 5 K/min, 7 K/min and 10 K/min. Eight weight fractions of Polymer 1 CUP solutions were cooled to 233.15 K at different rates, then isothermal for 10 min and heated to 313.15 K at the rate of 3 K/min. The amount of non-freezable surface water per CUP particle was calculated by Equation (4).

\[
m_{SW} = \left(\frac{\Delta H_W - \Delta H_{FW}}{\Delta H_W} - X_{CUP}\right) \cdot M \cdot N_A \cdot X_{CUP},
\]

where \(m_{SW}\) is the mass of surface water per CUP particle in grams, \(N_A\) is Avogadro constant, \(\Delta H_W\) is the heat of fusion of water (333.5 J/g), \(\Delta H_{FW}\) is the heat of fusion of freezable water in CUP solution, \(M\) is the molecular weight of CUP polymer, \(X_{CUP}\) is the weight fraction of CUP in the solution.

In order to demonstrate the effect of cooling rate on the amount of surface water, the DSC measurement of Polymer 1 solutions was conducted at 10 K/min cooling rate. Results showed that the amount of surface water was constant at concentrations below where Manning condensation occurs, Figure 6. However, as the cooling rate was reduced the amount of surface water measured decreased and showed a small concentration dependency. As the concentration of CUP increased the surface water decreased only slightly. During the freezing process, ice nucleates and the crystals grow. In a supercooling situation, when ice nucleation occurs growth is extremely rapid and traps the CUP particles in the crystal matrix before significant diffusion can occur. However, when slow cooled the ice growth was slower and allowed time for CUP diffusion and subsequent Manning condensation to occur [62]. When this charge condensation occurs the amount of surface water drops. At 0.1 degrees per minute and 5.23% CUP, the amount of surface water is \(4.64 \times 10^{-20}\) g per particle but at 10 degrees per minute it was \(4.94 \times 10^{-20}\) g per CUP particle, the drop in surface water was only 6.07%. All the CUPs in this study were polymerized from methyl methacrylate and methacrylic acid, the surface charged groups originated from neutralization of carboxylic acid by NaOH. The effective charge on the surface is subject to the dissociation equilibrium and there are two counterion condensations that exist in this system, inter-molecular counterion condensation and intra-molecular counterion condensation [63–65]. Inter-molecular counterion condensation occurs between particles when the weight fraction of CUP particles is high and intra-molecular counterion condensation occurs where the charge density of a single particle is high. Slow cooling rates allowed more time for CUP particles to migrate, when CUP particles were forced to become closer, the inter-molecular counterion condensation effect increased which reduced the number of effective charges on CUP’s surface, releasing part of the surface water where it became freezable. The counterion condensation effect was greater for higher concentrations of CUPs. This small difference is likely caused by the increased chance of charge condensation in the more crowded higher concentration.

Solutions with 21.40% and higher weight fraction CUP showed less surface water per particle at 10 K/min, Figure 7. The cause for this decrease was inter-molecular counterion condensation. When the weight fraction of CUP is high, with short distances between CUP particles, inter-molecular counterion condensation existed even before the cooling process. The inter-molecular counterion condensation also reduced the ice formation driven charge condensation which occurs during slower freezing rates. As the concentration increases the effect of slow freezing is reduced and at about 36% the surface water is a constant being independent of the cooling rate. It is important to note that the viscosity of the CUP solution is concentration dependent and rises rapidly due to the onset of Manning condensation [8,66]. The ion-ion repulsion reduces the diffusion rate of the CUP particle and thus makes it harder to move away from the advancing crystals of ice being formed and the CUP becomes an inclusion.
exist in this system, inter-molecular counterion condensation and intra-molecular counterion condensation [63–65]. Inter-molecular counterion condensation occurs between particles when the weight fraction of CUP particles is high and intra-molecular counterion condensation occurs where the charge density of a single particle is high. Slow cooling rates allowed more time for CUP particles to migrate, when CUP particles were forced to become closer, the inter-molecular counterion condensation effect increased which reduced the number of effective charges on CUP's surface, releasing part of the surface water where it became freezable. The counterion condensation effect was greater for higher concentrations of CUPs. This small difference is likely caused by the increased chance of charge condensation in the more crowded higher concentration.

Figure 6. Amount of surface water per CUP particle of Polymer 1 at 5.23%, 10.40% and 14.92% at different cooling rates.

Figure 7. Amount of surface water per CUP particle of Polymer 1 at different weight fraction at different cooling rates.

Direct comparison of the amount of surface water of the solutions with different CUP particle weight fractions are shown in Figure 8. The effect of the cooling rate decreased with the increasing of CUP particle weight fraction and gradually disappeared. The initial concentration independent region was again below the inter-molecular counterion condensation region. At approximately 20% the contribution due to the inter-molecular counterion condensation and subsequent viscosity effect obviously controlled the system. When the gel point is approached the particles cannot significantly move and the surface water reaches a point where the rate of cooling does not affect the result. Figure 8 illustrates the effect of 0.1 K/min versus 10 K/min cooling rates. At low concentration the surface water per cup was a constant until Manning condensation occurs then as the concentration increases the differences reduce to zero at the gel point.
Figure 8. Amount of surface water per CUP particle of Polymer 1 at different weight fraction at 0.1 K/min and 10 K/min cooling rate.

The packing of spheroidal materials is most likely to form either random close packing or hexagonal close packing. The maximum volume fraction 0.634 is for random close packing and for hexagonal close packing it is 0.7405 [67–71]. CUP solution is able to approach hexagonal close packing with the volume fraction of CUP particle being 0.7405 including surface water. Due to the small particle size and relatively low density the Brownian motion of CUP can help to move the particles to a position where the electrostatic repulsion and Brownian motion reach a balanced stable structure. Based on the previous publication by Van De Mark et al. [8], the weight fraction of CUP particle for each packing model can be calculated by Equations (5)–(7).

\[
\phi = \frac{\rho_s \cdot X_{\text{CUP}}}{\rho_p}
\]

\[
\phi_{\text{max}} (1 + \lambda/r)^3 = 0.634
\]

\[
\phi_{\text{max}} (1 + \lambda/r)^3 = 0.7405,
\]

where \(\phi\) is the CUP volume fraction, \(\rho_s\) is the density of CUP solution, \(\rho_p\) is density of CUP particle, \(X_{\text{CUP}}\) is the weight fraction of CUP, \(\phi_{\text{max}}\) is maximum volume fraction, \(\lambda\) is the thickness of surface water, \(r\) is radius of the CUP particle.

For CUP solutions of Polymer 1, the weight fraction of CUP particles was 36.16% when reaching random close packing, which is the gel point and 43% for hexagonal close packing. When the weight fraction of CUP particles was close to 30%, the cooling rate effect on the amount of surface water was almost gone, due to the very high viscosity and close to maximum inter-molecular counterion condensation. The surface water difference between the two cooling rates approached zero when the weight fraction of CUP particles is close to 43%, at this point and the particle is very difficult to move and any free water will be slow to migrate. It is also the point where CUP particles reaches maximum inter-molecular counterion condensation over the concentration range of 20% to 43%, CUP's the surface water amount drops by 47.55% at 10 K/min and 45.07% at 0.1 K/min.

The behavior of CUP solutions were similar to that of latex resins in that as the ice forms the particles are forced together. The higher charge density on the surface and the increased surface area create an increased amount of surface water which does not freeze. The same effects should
be observed in globular proteins and other nanoscale particles in water as well as in vitro and vivo materials when frozen.

4.3. Weight Fraction of Surface Water

In order to minimize the effect of counterion condensation in the remaining portion of this work, all the CUP solutions were cooled at 10 K/min, which is the highest cooling rate the DSC was capable of. The isothermal time was kept at 10 min and heated to 313.15 K at the rate of 3 K/min. Furthermore, since the CUP samples were neutralized by NaOH solution, heat of fusion of NaOH modified water sample was measured. Since there was no difference in the heat of fusion and melting point depression between NaOH modified water and deionized water (333.5 J/g), the deionized water was used as the standard. Different molecular weight polymers were made into CUP solutions and were prepared to various weight fractions. The heat of fusion of each solution was measured by DSC.

Figure 9 shows the DSC endothermic peak of Polymer 1 CUP solutions with three weight fractions. It was obvious that with the increasing of the weight fraction, the endothermic peaks shift to lower temperature, this can be explained by Raoult’s law [72]. CUP solutions contain only CUP particles, free water and surface water. The surface water does not freeze until a very low temperature and the free water which freezes like normal water. The endothermic peak for all the samples exhibited only a single peak. Thus, the observed difference in heat of fusion, between a water sample and a CUP solution, is due to the CUP particles and a fraction of water that does not freeze in the DSC measurement [46]. Therefore it is possible to determine the weight fraction of surface water utilizing the heat of fusion by DSC. For CUP samples, the weight fraction of free water can be determined by dividing the heat of fusion of the CUP solution by that of deionized water, which was used as a standard, Equation (8).

\[
X_{FW} = \frac{\Delta H_{FW}}{\Delta H_W}, \quad (8)
\]

where \(X_{FW}\) is the weight fraction of freezable water, \(\Delta H_{FW}\) is the heat of fusion of freezable water obtained from DSC, \(\Delta H_W\) is the heat of fusion of water, 333.5 J/g. The surface water weight fraction, \(X_{SW}\) can be calculated knowing the weight fraction of CUP, \(X_{CUP}\) and \(X_{FW}\), Equation (9).

\[
X_{SW} = 1 - X_{FW} - X_{CUP}. \quad (9)
\]
Combining Equations (8) and (9) to obtain Equation (10).

\[ X_{SW} = 1 - X_{CUP} - \left( \frac{\Delta H_{FW}}{\Delta H_W} \right) \tag{10} \]

From Equations (8)–(10), the weight fraction of surface water of each CUP solution was determined. Figure 10 shows the weight fraction of surface water vs weight fraction of the CUP particles.

The results indicate that there was a significant amount of non-freezable water in this system. The amount of water was linearly dependent upon the weight fraction of CUP over the range shown. Thus, the amount of water was constant per particle. Comparing Polymer 1, 2 and 3, that have a 9:1 MMA:MAA ratio, the higher molecular weight result in a lower slope, due in part to the lower surface area per gram of polymer. However, there is another variable, the charge density of ions on the CUP surface. As the molecular weight increases at a constant MMA:MAA ratio the surface charge density increases since all the charged groups will attempt to be on the surface. It would be expected that more charge density the more surface water should be observed.

Molecular weight of Polymer 4 is slightly larger than Polymer 1, the weight fraction of surface water should have been similar if the surface water is only dependent upon surface area and not on charge density. However, the charge density of Polymer 4 is much higher than polymer 1 and the surface water was significantly higher. This increase indicates that the surface water is dependent upon charge density. In addition, by comparing Polymer 2 and 5, with similar charge density, Polymer 2 has smaller molecular weight, so that it has more surface area and also more surface water. The conclusion can be made that both molecular weight and charge density play important roles in affecting the weight fraction of surface water. As for the same monomer ratio, larger molecular weight CUP particles will have higher charge density and if the monomer ratio is different, with similar molecular weight, the surface water weight fraction is proportional to the ions per nm² and higher charge density gives a higher surface water weight fraction. When the charge density is the same, higher molecular weight CUP has less surface water weight fraction that is, the surface area defines the surface water weight fraction.

All the CUP solutions discussed in Figure 10 are below the concentration where Manning condensation occurs. Polymer 1 was used to evaluate higher concentrations as is shown in Figure 11.
At concentrations above 20% the weight fraction of water begins to plateau at about 35%. Increasing the concentration causes an increase in the inter-molecular counterion interaction as the distance between ions becomes shorter. This interaction causes Manning condensation and reduces the effective charge on the surface. With less charge, the hydration layer decreases. Looking at Figures 10 and 11, the linear behavior at low concentration can be used to define when Manning condensation becomes significant by noting where the plot becomes non-linear.

4.4. Surface Water Density

Surface water has been found to have a greater density than bulk or free water [10,33,34]. To gain a deeper understanding of CUP surface water, the density of the CUP solutions were measured by high precision density meter at various weight fractions of CUP particles. Values of $1/\rho_s$ were plotted against weight fraction of CUP particles. The reciprocal of density of CUP solution was found to have a linear relationship with the weight fraction of CUP particles at low concentration [8].

Figure 12 shows polymer 1 as an example. It was observed that with the increasing of weight fraction of CUP particles, the reciprocal of density had an excellent linear regression fit up to about 20%. However, at higher concentrations the data deviated from linearity. This deviation can be explained by the increased weight fraction of CUP particles shortening the distance between the charged particles causing inter-molecular counterion condensation. The condensation reduces the amount of bound water per particle and thus alters the observed density. Higher concentrations could not be accurately measured for density due to their high viscosity.

In a given CUP solution, there are three components—free water, surface water and CUP particles. The combination of volume of CUP solid, volume of surface water and volume of free water is equal to the volume of the solution. By knowing the weight fraction of surface water, free water and CUP solids, as well as the density of CUP solid, free water and solution, the density of surface water can be determined by Equation (11).

$$\frac{1}{\rho_s} = \frac{X_{FW}}{\rho_{FW}} + \frac{X_{SW}}{\rho_{SW}} + \frac{X_{CUP}}{\rho_{CUP}}, \quad (11)$$

where $\rho_s$ is the density of CUP solution, $\rho_{FW}$ is the density of surface water, $\rho_{FW}$ is the density of free water, $\rho_{CUP}$ is the density of CUP particle.

![Figure 11. Weight fraction of surface water vs CUP particles of Polymer 1.](image-url)
Polymers 2020, 12, 1417

Figure 12. Dependence of $1/\rho_s$ on weight fraction of Polymer 1 CUP solution.

For each polymer, the surface water density was found to be constant at low weight fraction of CUP particles and when the weight fraction was high enough to cause inter-molecular counterion condensation the density of surface water drops due to the decreased effective charge density as shown in Table 3.

Table 3. Density of surface water for Polymers 1–7.

| CUPs | $X_{\text{CUP}}$ | $\rho_{sw}$ (g/mL) | CUPs | $X_{\text{CUP}}$ | $\rho_{sw}$ (g/mL) | CUPs | $X_{\text{CUP}}$ | $\rho_{sw}$ (g/mL) |
|------|-----------------|-------------------|------|-----------------|-------------------|------|-----------------|-------------------|
| 1    | 5.85% 1.0413    | 9.57% 1.0411      | 3    | 122.5k (0.84)   | 10.12% 1.0560     | 5    | 73.5k (0.66)    | 11.19% 1.0491     |
|      | 13.32% 1.0412   | 16.55% 1.0412     |      |                 | 18.10% 1.0563     |      |                 | 21.72% 1.0512     |
| 2    | 59.8k (0.66)    |                   | 4    | 25.4k (0.66)    | 5.18% 1.0500      | 6    | 49.7k (0.42)    | 4.35% 1.0335      |
|      | 10.35% 1.0508   | 11.35% 1.0509     |      |                 | 13.41% 1.0502     |      |                 | 16.11% 1.0357     |
|      | 18.72% 1.0507   |                   |      |                 | 14.30% 1.0501     |      |                 | 19.32% 1.0360     |
| 7    | 22.7k (0.24)    | 5.00% 1.0231      |      |                 |                   |      |                 |                   |

When comparing the densities of surface water of CUP solutions with different molecular weights, as it is shown in Figure 13, Polymer 2, Polymer 4 and Polymer 5 have same charge density (0.66 charges per nm²) and the densities of surface water are the same. CUPs with higher charge density have higher surface water density. All the CUP polymers except Polymer 3 fit a linear regression relationship of surface water density to surface charge density. Polymer 3 has the largest molecular weight and highest charge density which results in intra-molecular counterion condensation reducing the surface effective charge. This condensation leads to the actual effective charge density being smaller, thus resulting in a slightly smaller surface water density than expected by the linear relationship. When extrapolating the linear function to zero, where there is no charge on the surface, the surface water density is 1.0076 g/mL,
which is slightly larger than free water (0.997043 g/mL) [73]. This zero point should also represent the association with the ester groups, which is the other group capable of hydrogen bonding interaction with water.

The density of the surface water can be related to the hydrogen bonding of water to the carboxylates and the ester groups on the surface. The carboxylate groups form up to six strong hydrogen bonds and the esters four weaker. Therefore, since the carboxylate groups form a stronger interaction, the density would be higher than with the ester groups. As the charge density increases, the number of carboxylates increase as does the density [74].

### 4.5. Surface Water Thickness

To gain a deeper understanding of CUP surface water, it is important to relate our results to a structural model for the calculation of the surface water layer thickness. The total surface area of the CUP particles was calculated based on the measurement of the diameter of the CUP particle. The densities of surface water were determined, by knowing the weight fraction of surface water and number of CUP particles, the thickness of each sample was determined by Equation (12) [8].

\[
\frac{4}{3}\pi(\lambda + \frac{d}{2})^3 - \frac{4}{3}\pi(\frac{d}{2})^3 = \frac{X_{SW}M}{X_{CUP}N_A\rho_{SW}},
\]

where \(\lambda\) is the thickness of surface water, \(d\) is the diameter of CUP particle, \(X_{SW}\) is the weight fraction of surface water, \(X_{CUP}\) is the weight fraction of CUP particle, \(M\) is the molecular weight of CUP, \(N_A\) is Avogadro constant, and \(\rho_{SW}\) is the density of surface water.

Table 4 shows the surface water thickness of each CUP at different weight fractions. The thickness of CUPs ranges from 0.43 to 0.77 nm, which is about two to four water molecules thick. While previous studies have been suggested that surface water is as thick as from a few water molecules to several dozens of water molecules depending on the particles surface properties [75,76]. The thickness is likely to be primarily controlled by the charge density of the CUP particle surface and any hydrogen bonding groups at the surface. Charges will hold the water in a more bound state than water more loosely held at more hydrophobic surface groups. Comparing Polymer 1, 2 and 3, it can be concluded that the larger
particle has the thicker surface water layer due to it having more carboxylate groups at the surface per unit area. A higher charged surface will form a thicker electrical double layer, giving more counterions and associated more water molecules. This result agrees with the findings of Chen et al. [66]. While Polymer 4 has a lower molecular weight but the same ions per nm\(^2\) as Polymer 2, the thickness of surface water are the same. By looking at Polymer 1, the thickness of surface water is 0.636 nm when the weight fraction is small and the thickness decreases to 0.564 nm when the weight fraction is 25% due to the increased inter-molecular counter-ion condensation. This again implies that, the thickness of surface water is proportional to ions per nm\(^2\), which also corresponds to the finding of weight fraction of surface water.

**Table 4. Surface water thickness of each CUP at different concentrations.**

| CUPs | X\(_{\text{CUP}}\) | \(\lambda\) (nm) | CUPs | X\(_{\text{CUP}}\) | \(\lambda\) (nm) | CUPs | X\(_{\text{CUP}}\) | \(\lambda\) (nm) |
|------|-----------------|-----------------|------|-----------------|-----------------|------|-----------------|-----------------|
| 1 28.9k (0.52) | 5.85% | 0.635 | 3 122.5k (0.84) | 4.53% | 0.766 | 5 73.5k (0.66) | 11.19% | 0.734 |
| (0.52) | 9.57% | 0.637 | | 10.12% | 0.766 | | 14.67% | 0.733 |
| | 13.32% | 0.636 | | 13.72% | 0.765 | | 17.62% | 0.734 |
| | 16.55% | 0.636 | | 18.10% | 0.766 | | 21.71% | 0.732 |
| | 25.01% | 0.564 | | | | | |
| 2 59.8k (0.66) | 4.83% | 0.733 | 4 25.4k (0.66) | 5.18% | 0.732 | 6 49.7k (0.42) | 4.35% | 0.556 |
| (0.66) | 10.35% | 0.735 | | 10.15% | 0.735 | | 11.25% | 0.556 |
| | 11.35% | 0.734 | | 13.41% | 0.734 | | 16.11% | 0.557 |
| | 18.72% | 0.735 | | 14.30% | 0.735 | | 19.32% | 0.556 |

Figure 14 shows the plot of surface water thickness and charge density in ions per nm\(^2\). When the charge density increases on a single CUP particle, the intra-molecular counterion condensation occurs on itself, reducing the effective charges on the surface, resulting in a thinner surface water layer. When charge density was smaller than 0.66 nm, surface water thickness increases with the increasing of charge density and when charge density is higher, surface water thickness increases but less than expected due to the inter-molecular counter-ion condensation. Extrapolation of the line to zero shows that when there is no effective charge on CUP surface, there is still a 0.2513 nm surface water layer, which is approximately a monolayer of water [77–79].

### 4.6. Melting Point Depression

The melting point depression of CUP solutions can also offer potential information. DSC was used to accurately measure the heat flow associated with thermal transitions in CUP solutions. As is well known, by adding a non-volatile solute, the melting point of a solvent decreases [80]. If we consider the solution is ideal, the freezing/melting point depression can be described by Equation (13).

\[
\Delta T_F = K_F \cdot b \cdot i, \tag{13}
\]

where \(\Delta T_F\) is the melting point depression in K, \(K_F\) is cryoscopic constant (1.853 K·kg/mol), \(i\) is van’t Hoff constant, \(b\) is the molality of solute in mol/kg.
Since the CUP solutions were modified by NaOH to pH = 8.5, it is important to know if the Na\(^+\) might contribute to the temperature depression, which might affect the van’t Hoff factor in Equation (13). The molality of Na\(^+\) in CUP solutions was calculated by Equation (14).

\[
b_{Na} = \frac{10^{\text{pH} - 14} \cdot V}{m_{\text{solution}} - m_{\text{NaOH}}},
\]

where \(b_{Na}\) is the molality of sodium ions, \(V\) is the volume of solution, \(m_{\text{solution}}\) is the mass of solution, \(m_{\text{NaOH}}\) is the mass of NaOH.

The measured melting point depression of water and NaOH modified water was obtained by DSC, there was no difference between pure and pH modified water. By knowing the molality of Na\(^+\), the melting point depression, contributed by sodium ion in solution, was calculated to be \(1.1755 \times 10^{-5}\) \(K\), obtained from Equation (14), which was negligible.

Knowing the molality \(b = (n_{\text{solute}}/m_{\text{solvent}})\), the molality of CUP particles was calculated from Equation (15).

\[
b_{\text{CUP}} = \frac{X_{\text{CUP}}}{(1 - X_{\text{CUP}}) \cdot M_{W}},
\]

where \(b_{\text{CUP}}\) is the molality of CUP particle, \(M_{W}\) is the molecular weight of CUP, \(X_{\text{CUP}}\) is the weight fraction of CUP.

By using Equation (15), the molality of CUP can be calculated, the relation between the molality of CUP and melting point depression was plotted in Figure 15.

The molality of CUP being equal to 0.001 mol/kg was picked, the corresponding \(\Delta T_{\text{CUP}}\) was determined from Figure 15. Rearranging Equation (13), the van’t Hoff factor can be described as Equation (16).

\[
i = \frac{\Delta T_{\text{CUP}}}{b \cdot K_F}.
\]

In each CUP particle, the number of repeat units \((n_{\text{rep}})\) was calculated by dividing the molecular weight of CUP by the molecular weight of each repeat unit, Equation (17).

\[
n_{\text{rep}} = \frac{M_{W}}{n \cdot M_{\text{MMA}} + M_{\text{MAA}}},
\]

where \(n_{\text{rep}}\) is the number of repeat unit in one CUP particle, \(M_{\text{MMA}}\) is the molecular weight of MMA monomer, \(M_{\text{MAA}}\) is the molecular weight of MAA monomer, \(n\) is the molar ratio of monomer MMA/MAA.

The number of effective groups that contributed to van’t Hoff factor can be expressed as Equation (18).

\[
n_{\text{eff}} = \frac{i}{n_{\text{rep}}},
\]

where \(n_{\text{eff}}\) is the number of effective groups in each repeat unit.

The van’t Hoff factor, which is the number of ions per individual molecules of solute. As it was well known, van’t Hoff factor is equal to 2 for NaCl and 3 for BaCl\(_2\) as an example. In each repeat unit, there is one carboxylate group and several ester groups. Each carboxylate group was neutralized by NaOH, thus sodium ion may or may not be taken into account, shown in Equations (19) and (20).

\[
n_{\text{ester}} = n_{\text{eff}} - n_c
\]

\[
n_{\text{ester}} = n_{\text{eff}} - 2 \cdot n_c
\]

where \(n_c\) is the number of carboxylate group in each repeat unit, which equals to 1, \(n_{\text{ester}}\) is the number of ester group in each repeat unit.
increasing of charge density and when charge density is higher, surface water thickness increases but less than expected due to the inter-molecular counterion condensation. Extrapolation of the line to zero shows that when there is no effective charge on CUP surface, there is still a 0.2513 nm surface water layer, which is approximately a monolayer of water [77–79].

Figure 14. Surface water thickness vs charge density in ions per nm$^2$.

4.6. Melting Point Depression

The melting point depression of CUP solutions can also offer potential information. DSC was used to accurately measure the heat flow associated with thermal transitions in CUP solutions. As is well known, by adding a non-volatile solute, the melting point of a solvent decreases [80]. If we consider the solution is ideal, the freezing/melting point depression can be described by Equation (13).

$$\Delta T_F = \text{ibKT} FF \cdot \cdot = \Delta$$

(13)

where $\Delta T_F$ is the melting point depression in K, $K_F$ is cryoscopic constant (1.853 K·kg/mol), $i$ is van't Hoff constant, $b$ is the molality of solute in mol/kg.

Since the CUP solutions were modified by NaOH to pH = 8.5, it is important to know if the Na$^+$ might contribute to the temperature depression, which might affect the van't Hoff factor in Equation (13). The molality of Na$^+$ in CUP solutions was calculated by Equation (14).

$$\text{NaOH}_{\text{solution}} \cdot \text{pH} \cdot \text{Na} \cdot \text{mm} \cdot Vb \cdot = \cdot - 1410$$

(14)

where $b_{\text{Na}}$ is the molality of sodium ions, $V$ is the volume of solution, $m_{\text{solution}}$ is the mass of solution, $m_{\text{NaOH}}$ is the mass of NaOH.

The measured melting point depression of water and NaOH modified water was obtained by DSC, there was no difference between pure and pH modified water. By knowing the molality of Na$^+$, the melting point depression, contributed by sodium ion in solution, was calculated to be $1.1755 \times 10^{-5}$ K, obtained from Equation (14), which was negligible.

Figure 15. Molality of CUPs vs melting point depression.

The surface area of the CUP particle can be expressed as $A_{\text{CUP}} = 4\pi r^2$, average area of each repeat unit on CUP was determined.

$$A_{\text{rep}} = \frac{A_{\text{CUP}}}{n_{\text{rep}}}$$

(21)
Since the surface area of CUP was taken by carboxylate and ester groups, in each repeat unit, there is one carboxylate group and several ester groups. Using Equation (22), the average area of each ester group and carboxylate group were determined by using different molecular weight CUP particles.

\[
A_{\text{rep}} = A_c + n_{\text{ester}} \cdot A_{\text{ester}},
\]

where \(A_c\) is the average area of one carboxylate group occupied on CUP surface, \(A_{\text{ester}}\) is the average area of one ester group occupied on CUP surface.

The calculation indicated that sodium ion did contribute to the van’t Hoff factor value, the average area of an ester group on CUP surface was 0.374 nm\(^2\), while average area of a carboxylate group is 0.287 nm\(^2\). If assuming each group is a plane circle, the radius of carboxylate group is 0.244 nm, the radius of carboxylic acid group is 0.213 nm. Which is similar to the total of the length of C=O and O–H bond. The number of carboxylate groups varies upon different molecular weight and charge density of CUP particles. Results are shown in Table 5 [81,82].

### Table 5. Average area of each carboxylate and ester group on CUPs.

| CUPs | \(\Delta T_{\text{CUP}}\) (K) | \(b_{\text{CUP}}\) | \(i\) | \(n_{\text{rep}}\) | \(n_{\text{eff}}\) | \(A_{\text{rep}}\) (nm\(^2\)) | \(n_{\text{ester}}\) | \(A_e\) (nm\(^2\)) | \(A_c\) (nm\(^2\)) | \(n^*_{c}/n^*_{\text{ester}}\) per CUP |
|------|-----------------|-----------------|-----|--------|--------|-----------------|--------|-----------------|--------|-----------------|
| 1    | 0.344           | 0.001           | 185.645 | 29.276  | 6.341  | 1.911           | 4.341  | 0.375           | 0.284  | 29.3/126.3      |
| 2    | 0.589           | 0.001           | 317.863 | 60.579  | 5.247  | 1.501           | 3.247  | 0.374           | 0.285  | 60.6/196.7      |
| 3    | 1.010           | 0.001           | 545.062 | 124.096 | 4.392  | 1.181           | 2.392  | 0.374           | 0.285  | 124.1/296.8     |
| 4    | 0.330           | 0.001           | 178.090 | 33.121  | 5.377  | 1.548           | 3.377  | 0.373           | 0.289  | 33.1/111.9      |
| 5    | 0.674           | 0.001           | 363.734 | 68.869  | 5.282  | 1.513           | 3.282  | 0.374           | 0.286  | 68.9/226.0      |
| 6    | 0.462           | 0.001           | 249.325 | 33.406  | 7.463  | 2.408           | 5.463  | 0.372           | 0.290  | 33.4/182.5      |
| 7    | 0.262           | 0.001           | 141.392 | 11.417  | 12.397 | 4.185           | 10.397 | 0.375           | 0.290  | 11.4/118.7      |

\(\Delta T_{\text{CUP}}\): melting point depression in K, determined from Figure 15. \(b_{\text{CUP}}\): molality of CUP particles. \(i\): van’t Hoff factor. \(n_{\text{rep}}\): number of repeat unit in each CUP particle. \(n_{\text{eff}}\): number of effective groups per repeat unit. \(n_{\text{ester}}\): number of ester groups in each repeat unit. \(A_{\text{rep}}\): area of one repeat unit. \(A_c\): average area each carboxylate group takes (in equation \(\Delta T = i \cdot K_F \cdot b\), consider \(i = 2\)). \(A_e\): average area each ester group takes (in equation \(\Delta T = i \cdot K_F \cdot b\), consider \(i = 2\)). \(n^*_c/n^*_\text{ester}\): number of carboxylate and ester groups on one CUP particle surface.

Polymer 1, 2 and 3 have the same monomer ratio, with the increasing of molecular weight of CUP, the number of ester groups per repeat unit decreases. Because higher molecular weight CUP has more of hydrophobic groups on the inside, leaving more hydrophilic charged groups on the surface, giving higher ions per nm\(^2\). Polymer 2, 4 and 5 have similar ions per nm\(^2\) but different molecular weight, the number of ester groups per repeat unit are fairly close. This again indicates that the number of ester groups on the surface is dependent on the charge density with the carboxylates taking surface positions over esters during the formation of CUPs. The implication is that the area of the surface minus the area needed by the carboxylates leaves the remainder to be filled in by the esters with most esters being on the particle interior.

### 4.7. Specific Heat Analysis

Specific heat is also an important thermodynamic component. The specific heat measurement at a given temperature were also gathered from the DSC measurement, however, in order to obtain proper and critical measurement, the zeroline has to be measured separately and subtracted from the measured curve before evaluation [83]. An empty DSC pan was used to determine the heat flow rate...
of the zeroline $q_0$ (T) and a calibration substance of a known behavior, water, was placed into another DSC pan with similar mass, using the same experimental procedure. The precise specific heat $C_S$ can be calculated by simply subtracting the calibrated heat flow rate zeroline and combing with the known substance, shown in Equation (23).

$$C_S = \frac{q_S - q_0}{q_{ref} - q_0} \cdot \frac{m_{ref}}{m_S} \cdot C_{ref},$$

(23)

where $C_S$ is the specific heat of sample, $C_{ref}$ is the specific heat of reference, $q_S$ is the heat flow rate of the sample, $q_0$ is the heat flow rate of empty DSC pan, $q_{ref}$ is the heat flow rate of reference, $m_{ref}$ is the mass of reference, $m_S$ is the mass of sample.

If considering the mass difference of two DSC pans used for zeroline calibration and actual measurement, it is possible to further make routine calculation, Equation (24). However, the DSC pan correction result in an error smaller than 1%, which is negligible.

$$C_S = \frac{q_S - q_0}{q_{ref} - q_0} \cdot \frac{m_{ref}}{m_S} \cdot C_{ref} + \frac{m_{Cr,ref} - m_{Cr,S}}{m_S} \cdot C_{Cr},$$

(24)

where $m_{Cr,ref}$ is the mass of reference DSC pan, $m_{Cr,S}$ is the mass of sample DSC pan.

The dry polymer is a fine powder with low thermal conductivity, in order to have better thermal conductivity, the dry polymer was put in an open cap DSC sample pan and heated to 453.15 K to get rid of the moisture and let the polymer melt to have better contact with the pan. The pan was then sealed, cooled to 233.15 K, isothermal for 10 min, heated to 313.15 K. Two temperatures were picked for measurement, 253.15 K and 293.15 K, because at 253.15 K, the free water is in its ice form, while at 293.15 K, free water is liquid. Also, at these two temperatures, the baseline is strait, there is no overlapping with the endothermic peak and for one sample at a fixed temperature, the specific heat of dry CUP and surface water are constant. The specific heat of CUP solution is the total of the specific heat of free water (ice), dry CUP and surface water. The specific heat of each component at 263.15 K and 293.15 K was obtained from DSC measurement. Knowing the weight fraction, the specific heat of surface water was determined by Equations (25) and (26).

$$C_{P(s)} = aC_{P(ice)} + bC_{P(CUP)} + cC_{P(sw)},$$

(25)

$$C_{P(s)} = aC_{P(Fw)} + bC_{P(CUP)} + cC_{P(sw)},$$

(26)

where $C_{P(s)}$ is the specific heat of CUP solution, $C_{P(ice)}$ is the specific heat of ice, $C_{P(CUP)}$ is the specific heat of CUP particle, $C_{P(sw)}$ is the specific heat of surface water, $C_{P(Fw)}$ is the specific heat of free water, $a$, $b$ and $c$ represent the weight fraction of each component in the CUP solution.

The specific heat of each component in Polymer 1–7 were calculated, surface water specific heat is about 3.07 to 3.09 J/g·K at 293.15 K and 3.04 to 3.07 J/g·K at 253.15 K, which exhibit a small dependency on charge density, Table 6. The values exhibit only a small decrease in going from liquid water temperatures to well below freezing for water. The specific heat of surface water was independent of concentration for the range studied, which were all below the point of Manning condensation. The data for surface water at 293.15 K indicates that the water has more freedom than ice but less than liquid water. The small lowering of the specific heat with a 40 degree drop was similar to many materials not undergoing a phase change or other transition in mobility.

Surface water associated to the carboxylate and ester groups on CUP surface, known the specific heat of surface water at 293.15 K and 253.15 K, as well as the number of each functional groups, the specific heat of water associated to the carboxylate and ester groups could be estimated by Equation (27).

$$C_{P(sw)} = \frac{n_c}{n_c + n_{ester}} \cdot C_{P(swC)} + \frac{n_{ester}}{n_c + n_{ester}} \cdot C_{P(sw)},$$

(27)
where \( C_{P(swc)} \) is the specific heat of water associated to carboxylate groups, \( C_{P(swe)} \) is the specific heat of water associated to ester groups.

Table 7 shows the specific heat due to the ester and the carboxylate groups on the surface. This estimation of the effect of each group gives a relatively consistent average value which could be used to define a new polymer based on a group contribution basis.

**Table 6.** Specific heat of each components in polymer 1–7 (J/g K).

| CUPs | X_{CUP} | CUP Solid (253.15 K) | Surface Water (253.15 K) | Ice (253.15 K) | CUP Solid (293.15 K) | Surface Water (293.15 K) | Free water (293.15 K) |
|------|---------|---------------------|-------------------------|----------------|---------------------|-------------------------|---------------------|
| 1 28.9k (0.52) | 5.85% | 1.235 | 3.055 | 1.943 | 1.321 | 3.081 | 4.182 |
| 2 59.6k (0.66) | 10.35% | 1.230 | 3.051 | 1.943 | 1.315 | 3.076 | 4.182 |
| 3 122.5k (0.84) | 11.23% | 1.228 | 3.048 | 1.943 | 1.311 | 3.074 | 4.182 |
| 4 25.4k (0.66) | 11.25% | 1.231 | 3.058 | 1.943 | 1.316 | 3.087 | 4.182 |
| 5 73.5k (0.66) | 19.32% | 1.231 | 3.059 | 1.943 | 1.314 | 3.086 | 4.182 |
| 6 49.7k (0.42) | 5.00% | 1.237 | 3.066 | 1.943 | 1.324 | 3.093 | 4.182 |

**Table 7.** Specific heat of surface water associated with carboxylate and ester groups (J/g K).

| CUPs | \( C_{P(sw)} \) of Carboxylate (293.15 K) | \( C_{P(sw)} \) of Ester (293.15 K) | \( C_{P(sw)} \) of Carboxylate (253.15 K) | \( C_{P(sw)} \) of Ester (253.15 K) |
|------|--------------------------------|-----------------|--------------------------------|-----------------|
| 1 (0.52) | 2.979 | 3.103 | 2.965 | 3.073 |
| 2 (0.66) | 2.979 | 3.103 | 2.954 | 3.078 |
| 3 (0.84) | 2.984 | 3.102 | 2.953 | 3.078 |
| 4 (0.66) | 2.965 | 3.107 | 2.956 | 3.078 |
| 5 (0.66) | 2.975 | 3.105 | 2.965 | 3.073 |
| 6 (0.42) | 2.960 | 3.109 | 2.954 | 3.078 |
| 7 (0.24) | 2.978 | 3.104 | 2.956 | 3.077 |
5. Conclusions

This work discussed the thermal properties of CUPs with different molecular weight, monomer ratio and CUP surface charge density (ions per nm$^2$), based on the heat of fusion, specific heat and melting point depression aspects. It was found that surface water occupied a significant amount of volume in CUP solutions. Rapid cooling of CUP solutions will result in larger amount of surface water due to more rapid ice crystal growth and less time for CUP particles to migrate and undergo Manning condensation. The effect of cooling rate is less on the higher weight fraction solutions due to charge–charge repulsion which lowers mobility. The density of surface water was calculated and ranged from 1.023 g/mL to 1.056 g/mL depending on the charge density. The thickness of surface water was calculated, showing a trend of increasing with the increasing surface charge density. The molecular weight had no effect on the thickness of the surface water layer. However, as the weight fraction of CUP particles increased above approximately 20%, inter-molecular counterion condensation occurs and decreases surface water layer thickness. The melting point depression was found to linearly dependent upon molality of CUPs with the slope being related to the number of ions on the surface of CUPs. Using the melting point depression data, the average area of carboxylate and ester groups were determined and its results are independent of the molecular weight. The specific heat of surface water was found to be 3.07 to 3.09 J/g·K at 293.15 K and 3.04 to 3.07 J/g·K at 253.15 K, which was between ice and free water and exhibited a small dependency with the surface charge density. CUPs are true nanoscale spheroidal particles with the molecular weight and surface charge being easily designed and synthesized. The findings can be readily extended and utilized in many fields including biochemical and life sciences. Therefore, CUP particles offer an excellent model to investigate the behavior of surface water, which can be of fundamental importance to protein, micelle, hydrogel and material science. In addition, CUP solution is zero VOC and free of surfactant, it has great potentials in coating, adhesive and many other applications. This work represents a comprehensive study of CUP surface water which defines structural components and their effects on surface water in a detailed quantitative manner for the first time.

Author Contributions: Performed all DSC work and co-wrote the paper, P.G.; CUP solutions density measurement, A.Z.; Theory development, co-wrote and supervised the project, M.R.V.D.M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: The authors would like to thank the Department of Chemistry for the use of the DSC and the Missouri S&T Coating Institute for financial support. Also Rachel Keppler for her assistance as an undergraduate researcher and Dave Satterfield for DSC calibration.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Van De Mark, M.R.; Natu, A.; Gade, S.V.; Chen, M.; Hancock, C.; Riddles, C. Molecular Weight(Mn) and Functionality Effects on CUP Formation and Stability. J. Coat. Technol. Res. 2014, 11, 111–122. [CrossRef]
2. Oncley, J.L. Evidence from Physical Chemistry Regarding the Size and Shape of Protein Molecules from Ultra-centrifugation, Diffusion, Viscosity, Dielectric Dispersion, and Double Refraction of Flow. Ann. N. Y. Acad. Sci. 1941, 41, 121–150. [CrossRef]
3. Pammenter, N.W.; Vertucci, C.W.; Berjak, P. Homeohydrous (Recalcitrant) Seeds: Dehydration, the State of Water and Viability Characteristics in Landolphiakirkii. Plant Physiol. 1991, 96, 1093–1098. [CrossRef]
4. Yaghmur, A.; Aserin, A.; Tiunova, I.; Garti, N. Sub-zero temperature behaviour of non-ionic microemulsions in the presence of propylene glycol by DSC. J. Therm. Anal. Calorim. 2002, 69, 163–177. [CrossRef]
5. Jahnert, S.; Vaca, C.F.; Schaumann, G.E.; Schreiber, A.; Schonhoff, M.; Findenegg, G.H. Melting and freezing of water in cylindrical silica nanopores. Phys. Chem. Chem. Phys. 2008, 10, 6039–6051. [CrossRef]
6. Findenegg, G.H.; Jahnert, S.; Akcakayiran, D.; Schreiber, A. Freezing and Melting of Water Confined in Silica Nanopores. Chem. Phys. Chem. 2008, 9, 2651–2659. [CrossRef]
7. Riddles, C.; Zhao, W.; Hu, H.J.; Chen, M.; Van De Mark, M.R. Self-assembly of Water Insoluble Polymers into Colloidal Unimolecular Polymer (CUP) Particles of 3–9 nm. *Polymers* 2013, 55, 48–57. [CrossRef]
8. Chen, M.; Riddles, C.; Van De Mark, M. Gel point behavior of colloidal unimolecular polymer (CUP) particles. *Colloid Polym. Sci.* 2013, 291, 2893–2901. [CrossRef]
9. Vidulich, G.A.; Evans, D.F.; Kay, R.L. The Dielectric Constant of Water and Heavy Water between 0 and 40. degree. *J. Phys. Chem.* 1967, 71, 656–662. [CrossRef]
10. Chen, M.; Van De Mark, M.R. Rheology Studies on Colloidal Unimolecular Polymer (CUP) Particles in Absence of NaCl. *Preprints* 2011, 52, 336–337.
11. Natu, A.; Van De Mark, M.R. Synthesis and characterization of an acid catalyst for acrylic-melamine resin systems based on colloidal unimolecular polymer (CUP) particles of MMA-AMPS. *Prog. Org. Coat.* 2015, 81, 35–46. [CrossRef]
12. Aseyev, V.O.; Tenhu, H.; Klenin, S.I. Contraction of a Polyelectrolyte upon Dilution. Light-Scattering Studies on a Polycation in Saltless Water-Acetone Mixtures. *Macromolecules* 1998, 31, 7717–7722. [CrossRef]
13. Dobrynin, A.V.; Rubinstein, M.; Obukhov, S.P. Cascade of Transitions of Polyelectrolytes in Poor Solvents. *Macromolecules* 1996, 29, 2974–2979. [CrossRef]
14. Newton, R.; Gortner, R.A. A Method for Estimating Hydrophilic Colloid Content of Expressed Plant Tissue Fluids. *Bot. Gaz.* 1922, 74, 442–446. [CrossRef]
15. Bockris, J.O.; Conway, B.E.; Yeager, E. Comprehensive Treatise of Electrochemistry. *Angewandte* 1981, 93, 840.
16. Fawcett, W.R.; Levine, S.; de Nobriga, R.M.; McDonald, A.C. A molecular model for the dielectric properties of the inner layer at the mercury/aqueous solution interface. *J. Electroanal. Chem.* 1980, 111, 163–180. [CrossRef]
17. Lee, C.; McCammon, J.A.; Rossky, P.J. The structure of liquid water at an extended hydrophobic surface. *J. Chem. Phys.* 1984, 80, 4448–4455. [CrossRef]
18. Patey, G.N.; Torrie, G.M. Water and salt water near charged surface: A discussion of some recent theoretical results. *Chem. Scr.* 1989, 29, 39–47.
19. Schmickler, W.; Henderson, D. New model for the structure of the electrochemical interface. *Progr. Surf. Sci.* 1987, 22, 323–420. [CrossRef]
20. Price, D.; Halley, J.W. Electronic structure of metal-electrolyte surfaces: Three-dimensional calculation. *Phys. Rev. B* 1988, 38, 9857–9867. [CrossRef]
21. Glosli, J.N.; Philpott, M.R. Molecular-dynamics simulation of adsorption of ions from aqueous media onto charged electrodes. *J. Chem. Phys.* 1992, 96, 6962–6969. [CrossRef]
22. Spohr, E. Computer simulation of the water/platinum interface. *J. Phys. Chem.* 1989, 93, 6171–6180. [CrossRef]
23. Raghavan, K.; Foster, K.; Berkowitz, M. Comparison of the structure and dynamics of water at the platinum(111) and platinum(100) interfaces: Molecular dynamics study. *Chem. Phys. Lett.* 1991, 177, 426–432. [CrossRef]
24. Nagy, G.; Heinzinger, K. A molecular dynamics study of water monolayers on charged platinum walls. *J. Electroanal. Chem.* 1992, 327, 25–30. [CrossRef]
25. Aloisi, G.; Foresti, M.L.; Guidelli, R. A Monte Carlo simulation of water molecules near a charged wall. *J. Chem. Phys.* 1989, 91, 5592–5596. [CrossRef]
26. Erko, M.; Findenegg, G.H.; Cade, N.; Michette, A.G.; Paris, O. Confinement-induced structural changes of water studied by Raman scattering, *Phys. Rev. B* 2011, 84, 104205. [CrossRef]
27. Johnson, G.A.; Lecchini, S.M.A.; Smith, E.G.; Clifford, J.; Pethica, B.A. Role of water structure in the interpretation of colloid stability. *Discuss. Faraday Soc.* 1966, 42, 120–133. [CrossRef]
28. Clifford, J. Properties of Micellar Solutions. Part 4.—Spin Lattice Relaxation Times of Hydrocarbon Chain Protons in Solutions of Sodium Alkyl Sulphates. *Trans. Faraday Soc.* 1965, 61, 1276–1282. [CrossRef]
29. Clifford, J.; Lecchini, S. State of Liquid Water near Solid Interfaces. *SCI Monogr.* 1967, 25, 174–195.
30. Katayama, S.; Fujiwara, S. NMR study of the spatial effect of polyacrylamide gel upon the water molecules confined in it. *J. Am. Chem. Soc.* 1979, 101, 4485–4488. [CrossRef]
31. Van De Mark, M.R.; Zore, A.; Geng, P.; Zheng, F. Colloidal Unimolecular Polymer Particles: CUP. In *Single-Chain Polymer Nanoparticles*; Pomposo, J.A., Ed.; Wiley-VCH: Weinheim, Germany, 2017; pp. 259–312.
32. Mamontov, E. Dynamics of surface water in ZrO2 studied by quasielastic neutron scattering. *J. Chem. Phys.* 2004, 121, 9087–9097. [CrossRef]
33. Toney, M.F.; Howard, J.N.; Richer, J.; Borges, G.L.; Gordon, J.G.; Melroy, O.R.; Wiesler, D.G.; Yee, D.; Sorensen, L.B. Voltage-dependent ordering of water molecules at an electrode-electrolyte interface. *Nature* 1994, 368, 444–446. [CrossRef]

34. Toney, M.F.; Howard, J.N.; Richer, J.; Borges, G.L.; Gordon, J.G.; Melroy, O.R.; Wiesler, D.G.; Yee, D.; Sorensen, L.B. Distribution of water molecules at Ag(111)/electrolyte interface as studied with surface X-ray scattering. *Surf. Sci.* 1995, 335, 326–332. [CrossRef]

35. Ling, C.S.; Hansen, W.D. DTA Study of Water in Porous Glass. *Adsort. Interface* 1975, 8, 129–156.

36. Velazquez, G.; Herrera-Gomez, A.; Martin-Polo, M.O. Identification of bound water through infrared spectroscopy in methycellulose. *J. Food Eng.* 2003, 59, 79–84. [CrossRef]

37. Lee, K.Y.; Ha, W.S. DSC studies on bound water in silk fibroin/S-carboxymethyl kerateine blend films. *Polymer* 1999, 40, 4131–4134. [CrossRef]

38. Higuchi, A.; Lijima, T. DSC investigation of the states of water in poly (vinyl alcohol) membranes. *Polym. Sci.* 1985, 26, 1207–1211. [CrossRef]

39. Ratto, J.; Hatakeyama, T.; Blumstein, R.B. Differential scanning calorimetry investigation of phase transitions in water/chitosan systems. *Polymer* 1995, 36, 2915–2919. [CrossRef]

40. Garti, N.; Aserin, A.; Ezrati, S.; Tiunova, I.; Berkovic, G. Water Behavior in Nonionic Surfactant Systems I: Subzero Temperature Behaviour of Water in Nonionic Microemulsions Studied by DSC. *J. Colloid Interface Sci.* 1995, 178, 60–68. [CrossRef]

41. Tahmasebi, A.; Yu, J.; Su, H.; Han, Y.; Lucas, H.; Zheng, H.; Wall, T. A differential scanning calorimetric (DSC) study on the characteristics and behaviour of water in low-rank coals. *Fuel* 2014, 135, 243–252. [CrossRef]

42. Monti, D.; Chetoni, P.; Burgalassi, S.; Najarro, M.; Saettone, M.F. Increased corneal hydration induced by potential ocular penetration enhancers: Assessment by differential scanning calorimetry (DSC) and by desiccation. *Int. J. Pharm.* 2002, 232, 139–147. [CrossRef]

43. Garti, N.; Aserin, A.; Tiunova, I.; Fanun, M. A DSC study of water behaviour in water-in-oil microemulsions stabilized by sucrose esters and butanol. *Colloids Surf. A* 2000, 170, 1–18. [CrossRef]

44. Ross, K.D. Differential scanning calorimetry of nonfreezeable water in solute-macromolecule-water system. *J. Food Sci.* 1978, 43, 1812–1815. [CrossRef]

45. Bushuk, W.; Mehrotra, V.K. Studies of water binding by differential thermal analysis. II. Dough studies using the melting mode. *Cereal Chem.* 1977, 54, 320–325.

46. Biswas, A.B.; Kumsah, C.A.; Pass, G.; Philips, G.O. The effect of carbohydrates on the Heat of Fusion of Water. *J. Solut. Chem.* 1975, 4, 581–590. [CrossRef]

47. Berlin, E.; Kliman, P.G.; Anderson, B.A.; Pallansch, M.J. Water binding in whey protein concentrates. *J. Dairy Sci.* 1973, 56, 984–987. [CrossRef]

48. Berlin, E.; Kliman, P.G.; Anderson, B.A.; Pallansch, M.J. Changes in state of water in proteinaceous systems. *J. Colloid Interface Sci.* 1970, 34, 488–494. [CrossRef]

49. Hatakeyama, T.; Yamauchi, A. Studies on bound water in poly (vinyl alcohol) hydrogel by DSC and FT-NMR. *Eur. Polym. J.* 1984, 20, 61–64. [CrossRef]

50. Ostrowska-Czubenko, J.; Pierog, M.; Gierszewska-Druzynska, M. State of water in noncrosslinked and crosslinked hydrogel chitosan membranes-DSC studies. *Prog. Chem. Appl. Chitin Deriv.* 2011, 16, 147–156.

51. Muffett, D.J.; Snyder, H.E. Measurement of unfrozen and free water in soy proteins by differential scanning calorimetry. *J. Agric. Food Chem.* 1980, 28, 1303–1305. [CrossRef]

52. Kobayashi, Y.; Fukada, K. Characterization of swollen lamellar phase of dimyristoylphosphatidylcholine-gramicidin A mixed membranes by DSC, SAXS, and densimetry. *Biophys. Acta* 1998, 1371, 363–370. [CrossRef]

53. ASTM E2161-08, *Standard Terminology Relating to Performance Validation in Thermal Analysis*; ASTM International: West Conshohocken, PA, USA, 2008.

54. ASTM E473-08, *Standard Terminology Relating to Thermal Analysis and Rheology*; ASTM International: West Conshohocken, PA, USA, 2008.

55. *Principles and Applications of Thermal Analysis*; Wiley-Blackwell: Hoboken, NJ, USA, 2007.

56. East, G.C.; Margerison, D.; Pulat, E. Variation of polymer density with molecular weight and consequences in dilatometric studies of addition polymerization. *Trans. Faraday Soc.* 1966, 62, 1301–1307. [CrossRef]

57. Bosen, S.F.; Bowles, W.A.; Ford, E.A.; Person, B.D. *Antifreezes. Ullmann’s Encyclopedia of Industrial Chemistry*, 5th ed.; Wiley-VCH: Weinheim, Germany, 1985; Volume A3.
58. Abdelwahed, W.; Degobert, G.; Stainmesse, S.; Fessi, H. Freeze-drying of nanoparticles: Formulation, process and storage considerations. Adv. Drug Deliv. Rev. 2006, 58, 1688–1713. [CrossRef] [PubMed]
59. Jiang, S.; Nail, S.L. Effect of process conditions on recovery of protein activity after freezing and freeze-drying. Eur. J. Pharm. Biopharm. 1998, 45, 249–257. [CrossRef]
60. Searles, J.A.; Carpenter, J.F.; Randolph, T.W. The ice nucleation temperature determines the primary drying rate of lyophilization for samples frozen on a temperature-controlled shelf. J. Pharm. Sci. 2001, 90, 860–871. [CrossRef]
61. Auveillain, M.; Cave, G.; Fessi, H.; Devissaguet, J.P. Lyophilisation de vecteurs colloïdaux submicroniques. STP Pharma 1989, 5, 738–744.
62. Chang, B.S.; Patro, S.Y. Freeze-drying Process Development for Protein Pharmaceuticals. In Lyophilization of Biopharmaceuticals; Costantino, H.R., Pikal, M.J., Eds.; American Association of Pharmaceutical Scientist: Alington, VA, USA, 2004; pp. 113–138.
63. Manning, G.S. A limiting law for the conductance of the rod model of a salt-free polyelectrolyte solution. J. Phys. Chem. 1975, 79, 262–265. [CrossRef]
64. Ninham, B.W.; Parsegian, V.A. Electrostatic Potential between Surfaces Bearing Ionizable Groups in Ionic Equilibrium with Physiologic Saline Solution. J. Theor. Biol. 1971, 31, 405–428. [CrossRef]
65. Belloni, L.; Interdisciplinary Laboratory of Nanoscale and Supramolecular Organization, DSM/IRAMIS/SIS2M, CEA/Saclay, 91191 Gif-sur-Yvette CEDEX, France. Private Communication, 1971.
66. Chen, M.; Riddles, C.J.; Van De Mark, M.R. Electrorheological contribution to the rheology of colloidal unimolecular polymer (CUP) particles in water. Langmuir 2013, 29, 14034–14043. [CrossRef]
67. de Kruif, C.G.; van Iersel, E.M.F.; Vrij, A. Hard sphere colloidal dispersions: viscosity as a function of shear rate and volume fraction. J. Chem. Phys. 1983, 83, 4717–4725. [CrossRef]
68. van der Werff, J.C.; de Kruif, C.G.; Blom, C.; Mellema, J. Linear viscoelastic behavior of dense hard-sphere dispersions. Phys. Rev. A 1989, 39, 795–807. [CrossRef] [PubMed]
69. van de Werff, J.C.; de Kruif, C.G.; Blom, C.; Mellema, J. Linear viscoelastic behavior of dense hard-sphere dispersions. J. Rheol. 1989, 33, 421–454. [CrossRef]
70. Pishvaei, M.; Grailat, C.; McKenna, T.F.; Cassagnau, P. Rheological behaviour of polystyrene latex near the maximum packing fraction of particles. Polymer 2005, 46, 1235–1244. [CrossRef]
71. Dames, B.; Morrison, B.; Willenbacher, N. An empirical model predicting the viscosity of highly concentrated, bimodal dispersions with colloidal interactions. Rheol. Acta 2001, 40, 434–440. [CrossRef]
72. Villa, S.; Riesco, N.; García de la Fuente, I.; Gonzalez, J.A.; Cobos, J.C. Thermodynamics of mixtures with strongly negative deviations from Raoult’s law: Part 5. Excess molar volumes at 298.15 K for 1-alkanols + dipropylamine systems: Characterization in terms of the ERAS model. Fluid Phase Equilib. 2001, 190, 113–125. [CrossRef]
73. Kotz, J.C.; Treichel, P.M.; Weaver, G.C. Kinetic Theory of Gases. In Chemistry and Chemical Reactivity, 6th ed.; Thomson Brooks Cole, Cengage Learning: Boston, MA, USA, 2005.
74. March, J. Advanced Organic Chemistry, 4th ed.; John and Wiley and Sons: Hoboken, NJ, USA, 1992.
75. Palasantzas, G.; Svetovoy, V.B.; van Zwol, P.J. Influence of ultrathin water layer on the van der Waals/Casimir force between gold surfaces. Phys. Rev. B 2009, 79, 235434. [CrossRef]
76. Schufle, J.A.; Huang, C.T.; Drost-Hansen, W. Temperature dependence of surface conductance and a model of vicinal (interfacial) water. J. Colloid Interface Sci. 1976, 54, 184–202. [CrossRef]
77. Ganta, D.; Dale, E.B.; Rosenberger, A.T. Measuring sub-nm adsorbed water layer thickness and desorption rate using a fused-silica whispering-gallery microresonator. Meas. Sci. Technol. 2014, 25, 055206. [CrossRef]
78. Eisenberg, D.; Kauzmann, W. The Structure and Properties of Water; Oxford University Press: Oxford, UK, 1969.
79. Opitz, A.; Scherge, M.; Ahmed, S.I.-U.; Schaefer, J.A. A comparative investigation of thickness measurements of ultra-thin water films by scanning probe techniques. J. Appl. Phys. 2007, 101, 064310. [CrossRef]
80. Abbott, A.P.; Boothby, D.; Capper, G.; Davies, D.L.; Rasheed, R.K. Deep eutectic formed between choline chloride and carboxylic acids: Versatile alternatives to ionic liquids. J. Am. Chem. Soc. 2004, 126, 9142–9147. [CrossRef]
81. March, J.; Smith, D. Advanced Organic Chemistry, 5th ed.; Wiley: Hoboken, NJ, USA, 2001.
82. Demaison, J.; Herman, M.; Lievin, J. The equilibrium OH bond length. *J. Int. Rev. Phys. Chem.* 2007, 26, 391–420. [CrossRef]

83. Huhne, G.; Hemminger, W.; Flammersheim, H.J. *Differential Scanning Calorimetry*; Springer: Berlin/Heidelberg, Germany, 1996.

© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).