The complete chloroplast genome of *Mimosa pigra* L. (Fabaceae), a notorious invasive plant

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**ABSTRACT**

*Mimosa pigra* L., also called the giant sensitive tree, is native to tropical America and has invaded Africa, Asia, and Australia. Here, we report the complete chloroplast genome of *M. pigra*, which was 165,996 bp in length and composed of a large single-copy region (LSC; 93,299 bp), a small single-copy region (SSC; 17,989 bp) and two inverted repeat regions (IRs; 27,354 bp). The complete *M. pigra* chloroplast genome included 83 protein-coding genes, 37 tRNAs and 8 rRNAs. Phylogenetic analysis using the maximum likelihood method revealed the monophyly of *M. pigra* and related taxa of the subfamily Caesalpinioideae. In comparison, the members of Papilionoideae were paraphyletic.

*Mimosa* L. 1753 is a genus in Fabaceae that includes 612 accepted species (POWO 2021). *Mimosa pigra* L. 1755, also called the giant sensitive plant, is native to America, but has become a notorious invasive plant in various countries in Asia, Africa and Australia (Shanungu 2009; Mansor and Crawley 2011; Rijal and Cochard 2016; Huynh et al. 2020; Witt et al. 2020; POWO 2021). Because it is harmful to agricultural crops, *M. pigra* has been surveyed and controlled in Cambodia, Malaysia, Vietnam, Zambia and Australia (Shanungu 2009; Mansor and Crawley 2011; Rijal and Cochard 2016; Huynh et al. 2020; Witt et al. 2020). Different phytochemicals have been identified in *M. pigra* extracts (Koodkaew et al. 2018); one extract was shown to protect against cardiovascular diseases (Rakotomalala et al. 2013). Additionally, compounds from a *M. pigra* extract inhibited the growth of other plants, including barnyard grass (Do et al. 2019, 2020). *Mimosa pigra* can also restore polluted soil (Elemike et al. 2019; Pérez-Hernández et al. 2020). These studies revealed both the benefits and disadvantages of *M. pigra*; however, genomic and proteomic researches are required. Therefore, in this study, we sequenced the complete *M. pigra* chloroplast genome using the MiSeq platform to provide the genomic data for future studies of *Mimosa* in particular and Fabaceae in general.

Fresh *M. pigra* leaves were collected in Can Tho, Vietnam (10°02'06.1"N 105°46'04.0"E) and then stored in liquid nitrogen. No specific permission was required because this species is considered an invasive plant in Vietnam. A specimen was identified by Dr. Nguyen Pham Anh Thi and Dr. Khang Do Tan and deposited at the Biotechnology Research and Development Institute (for free access to the sample, contact Dr. Nguyen Pham Anh Thi; email: npathi@ctu.edu.vn) under voucher number BRDI-THI 20200531-001. Total DNA was isolated using the modified CTAB method (Doyle and Doyle 1987). The DNA extract was used to prepare a sequencing library with a TrueSeq Nano DNA Sample Preparation Kit for the Illumina MiSeq platform. The 300 bp paired-end raw reads were imported to Geneious Prime 2021.1 (Kearse et al. 2012) to assemble the chloroplast genome sequence, with *Mimosa pudica* L., 1753 (GenBank acc. no. MH671330) as reference genome. The obtained chloroplast genome was annotated using Geneious Prime 2021.1 and deposited in the NCBI under accession number OL889924.

*Mimosa pigra* had a 165,996 bp in size, typical quadripartite chloroplast genome that includes large single-copy region (LSC; 93,299 bp) and small single-copy region (SSC; 17,989 bp) regions separated by two inverted repeat regions (IR; 27,354 bp). The genome sequence had a 35.4% GC content and contained 83 protein-coding genes, 37 tRNAs, and 8 rRNAs, of which 16 sequences were duplicated in the IR regions: *rpl2, rpl23, rps12, ndhB, ycf2, trnL-CAU, trnM-CAA, trnV-GAC, trnI-GAU, trnR-ACG, trnN-GUU, trnA-UGC, rmn4.5, rm5, rm16, and rm23. The newly sequenced chloroplast genome showed 83.9% similarity to the chloroplast genome of *M. pudica* in NCBI database (GenBank acc. no. MH671330). The junction between the LSC and IR regions of *M. pigra* was located in the rps19 coding region (CDS), which is also similar in *M. pudica* cpDNA. Similarly, the junction between the LSC and SSC regions was in the CDS of *ycf1* in both *Mimosa* species. Although the junctions among the LSC/SSC/IR regions are similar in *M. pigra*, the junctions among the LSC/SSC/IR regions are different in *M. pudica*. The maximum likelihood method revealed the monophyly of *M. pigra* and related taxa of the subfamily Caesalpinioideae. In comparison, the members of Papilionoideae were paraphyletic.
pigra and M. pudica, there are more than 600 species in Mimosa. Therefore, more Mimosa species should be examined to explore the diversity of junctions between the LSC/SSC/IR regions not only in Mimosa but also in Fabaceae.

To conduct a phylogenetic analysis, 78 protein-coding regions in the chloroplast genomes of 18 Fabaceae species were downloaded from the NCBI, of which Bauhinia blakeana S.T.Dunn was used as the outgroup. The sequences were then aligned using MUSCLE (Edgar 2004) embedded in Geneious Prime (Kearse et al. 2012) and concatenated. jModeltest 2.0 determined that the best model for the data matrix was the transversion model (TVM) + proportion of invariable sites (I) + gamma distribution (G) model (Darriba et al. 2012). The IQ-TREE package was used to construct a phylogenetic tree using the maximum likelihood method with 1,000 bootstrap replicates (Minh et al. 2020). The phylogenetic tree was illustrated using Figtree (http://tree.bio.ed.ac.uk/software/figtree/). The phylogenetic analysis showed the monophyly of M. pigra and related species in the sub-family Caesalpinioideae with high support (Figure 1). In Caesalpinioideae, M. pigra and M. pudica formed a clade that is sister to the group of Piptadenia communis Benth., 1841 and Stryphnodendron adstringens (Mart.) Coville, 1910. The complete chloroplast genome provides important information for additional studies on the population genetics of M. pigra and possible strategies for controlling its invasiveness.

**Figure 1.** The maximum likelihood tree inferred from 78 protein-coding regions of 18 chloroplast genomes of Mimosa and related taxa. The numbers are the bootstrap values. CAE: Caesalpinioideae; PAP: Papilionoideae; DIA: Dialioideae; CER: Cercidoideae.

**Author’s contribution**

Le Van Minh and Hoang Dang Khoa Do conceived the conception and design; Nguyen Pham Anh Thi and Khang Do Tan collected and determined the samples; Nguyen Pham Anh Thi, Khang Do Tan and Nguyen Thi Khoa conducted the experiments, analyzed the data, and wrote the draft manuscript; Le Van Minh and Hoang Dang Khoa Do revised the draft manuscript. All authors agreed to the final form of this manuscript.

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**Disclosure statement**

The authors report no conflicts of interest. The collection of M. pigra does not require specific permissions or licenses from the government and local governors. The authors alone are responsible for the content and writing of this article.

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**Data availability statement**

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at [https://www.ncbi.nlm.nih.gov/](https://www.ncbi.nlm.nih.gov/) under the accession no. OL889924. The associated BioProject, SRA, and Bio-Sample numbers are PRJNA788179, SRP350381, and SAMN23929642, respectively.

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