INTRODUCTION

Angiotensin receptor blockers (ARBs) are antagonists to angiotensin II as it binds to angiotensin II receptor type I and blocks vasoconstriction, retention of sodium and water, cell proliferation, aldosterone release, and sympathetic nerve activation [1,2]. ARBs are safe approved drugs since the 1990s widely used to treat atherosclerosis, cardiovascular diseases, and blood pressure and have been included in commonly prescribed drug for the treatment of hypertension [3,4]. Losartan, candesartan, telmisartan, valsartan, and irbesartan are few of the approved ARBs. Telmisartan and irbesartan are metabosartans classified under the second-generation ARBs with a protective effect on diabetic nephropathy [5]. Adding on telmisartan has also substantially proven neuroprotective effects by possessing anti-inflammatory properties, in stroke patients [6].

There were conflicting reports on the association of ARBs with cancer. Candesartan in heart failure assessment of reduction in mortality and morbidity study showed that there were more fatal cancers in patients administered candesartan than placebo [7]. Meta-analysis using randomized controlled trials performed by Sipahi et al. [8] reported association of unassuring increased risk of new cancer diagnosis and ARBs. Later, a cohort study observed an apparently protective association between the use of ARBs and lung cancer [9]. Another meta-analysis study reported decreased lung cancer risk with ARBs [10].

Lung cancer prevalence among newly diagnosed cases of cancer and cancer deaths constitutes 13% and 19%, respectively, worldwide [11]; 6.9% and 9.3%, respectively, in India [12]. Its prevalence and mortality are due to lack of early detection of the illness, which risks the patient’s life [13]. Due to the association of ARBs to lung cancer, the present study aims to demonstrate the anticancer activity of the two drugs telmisartan and irbesartan using methylthiazolyldiphenyl-tetrazolium bromide (MTT) assay on cell lines. A lung cancer cell line human lung alveolar epithelial cells derived carcinoma cell line A549 was used for analysis. Telmisartan and irbesartan were added to A549 cells at different concentration to determine the effect of these drugs on cancer cell viability in a dose-dependent manner.

METHODS

Cell line and culture

A549 cell line was provided by the National Centre for Cell Sciences, Pune. The cells were cultured and maintained in cell culture media, minimal essential medium (MEM) composed with antibiotics such streptomycin (100 μg/ml) and penicillin (100 U/ml), 10% fetal bovine serum (FBS) in a sui 2. The MTT assay for determining the tumor cell cytotoxicity of the two drugs was performed according to the method of Mosmann [14]. Briefly, cells (1×105/well) were plated in 24-well plates and incubated at 37°C with 5% CO₂ atmosphere. On reaching confluence, the cells were washed in phosphate-buffered saline (PBS) and the medium was...
changed. Increasing concentrations of both the drugs obtained through serial dilution were added to the different wells and were marked, respectively. One of the wells was treated only with the diluent which served as the control. Cell control and drug control were included in each assay. The culture plates were kept under incubation for 24 h at 37°C with 5% CO₂ atmosphere. The medium was separated from wells after incubation, then PBS with pH 7.4 was used for washing the cells. 100 µl/well of 0.5% MTT (5 mg/ml) was added to each well and the cell plate was kept under incubation for 4 h. After incubation, 1 ml of DMSO was added in all the wells. This helps in dissolving the insoluble crystalline formazan product for effective absorbance measurement. UV spectrophotometry was used to measure the absorbance at 570 nm; taking DMSO as blank, and the results were tabulated using the formula as follows:

\[
% \text{Cell viability} = \frac{\text{Absorbance of Treated}}{\text{Absorbance of Control}} \times 100
\]

RESULTS
The absorbance results of MTT assay and the resultant cell viability percentage are tabulated in Table 1 for telmisartan and Table 2 for irbesartan.

Half maximal inhibitory concentration (IC₅₀) level of irbesartan was 31.2 µg and that of telmisartan was 15.6 µg, respectively. This suggests that telmisartan is more cytotoxic than irbesartan. MTT cytotoxicity assay reveals an increase in cell death corresponding to an increase in the concentration of both irbesartan and telmisartan.

Effect of the drug telmisartan and irbesartan on the A549 cells was observed in a phase contrast microscope, and the results are presented in Fig. 1 (telmisartan) and Fig. 2 (irbesartan).

DISCUSSION
Our study proves the anticancer activity of these antihypertensive drugs. Both the drugs were able to establish a good cytotoxicity at lower concentrations indicating that more studies on this field will potentially establish an effective anticancer drug for treating lung cancers. The dot plots of the MTT assay reveal a steep decrease in the cell viability with increasing concentrations of both telmisartan and irbesartan. On extrapolating the IC₅₀ values from the graph, it is shown that telmisartan is more effective than irbesartan. Almost half the concentration of telmisartan (15.6 µg/ml) was able to establish the IC₅₀ when compared to irbesartan (31.2 µg/ml). This suggests that telmisartan will be a better drug of choice for future study. Microscopic observation showed a clear decrease in the live cells when compared to control at the IC₅₀ concentration. Dead cells were seen with distorted and rounded morphology. There were few live elongated cells with normal morphology in Panel D and Panel C while there were no live cells at all in Panel B in comparison with Panel A. This was true for both telmisartan and irbesartan. This clearly shows that the drugs can induce cell death in these lung cancer cell models and thereby can stand as an effective anticancer drug.

Previous studies on telmisartan and irbesartan have demonstrated their ability to block cell proliferation in a dose-dependent manner. This property of these drugs was found to be due to its ability to block extracellular signal-regulated kinase (ERK) activation in aortic vascular smooth muscle [15]. ERK is one of the major mitogen-activated protein kinase (MAPK) signals that the cancer cells use for cell proliferation [16]. This could probably be the reason for the anticancer property noted in our study. More study in this line is required to demonstrate the involvement of MAPK signals in telmisartan- and irbesartan-induced anticancer activity. Further, the effect of irbesartan in blocking hypoxia-induced angiogenesis has also been reported [17].

### Table 1: Effect of various concentrations of telmisartan on the cell viability of A549 cell line

| S. No. | Concentration (µg/ml) | Dilutions | Absorbance (O.D) | Cell viability (%) |
|-------|-----------------------|-----------|------------------|-------------------|
| 1     | 1000                  | Neat      | 0.12             | 16.66             |
| 2     | 500                   | 1:1       | 0.17             | 23.61             |
| 3     | 250                   | 1:2       | 0.21             | 29.16             |
| 4     | 125                   | 1:4       | 0.26             | 36.11             |
| 5     | 62.5                  | 1:8       | 0.30             | 41.66             |
| 6     | 31.2                  | 1:16      | 0.33             | 45.83             |
| 7     | 15.6                  | 1:32      | 0.36             | 50.00             |
| 8     | 7.8                   | 1:64      | 0.44             | 61.11             |
| 9     | Cell control          | -         | 0.72             | 100               |

0.D: Optical density

### Table 2: Effect of various concentrations of irbesartan on the cell viability of A549 cell line

| S. No. | Concentration (µg/ml) | Dilutions | Absorbance (O.D) | Cell viability (%) |
|-------|-----------------------|-----------|------------------|-------------------|
| 1     | 1000                  | Neat      | 0.09             | 12.50             |
| 2     | 500                   | 1:1       | 0.15             | 20.83             |
| 3     | 250                   | 1:2       | 0.24             | 33.33             |
| 4     | 125                   | 1:4       | 0.27             | 37.50             |
| 5     | 62.5                  | 1:8       | 0.32             | 44.44             |
| 6     | 31.2                  | 1:16      | 0.37             | 51.38             |
| 7     | 15.6                  | 1:32      | 0.40             | 55.55             |
| 8     | 7.8                   | 1:64      | 0.42             | 58.33             |
| 9     | Cell control          | -         | 0.72             | 100               |

0.D: Optical density

Fig. 1: Microscopic observation of the telmisartan-treated A549 cell line. Panel A represents the control untreated cells, the panel control shows more viable elongated slender healthy cells; Panel B represents treatment with 1000 µg/ml of the drug, this being the maximal drug concentration shows no viable cells in the field; Panel C represents treatment with 31.2 µg/ml of the drug, this shows very few cells on the whole in the field with more distorted and apoptotic morphology; and Panel D represents 15.6 µg/ml of the drug, being the half maximal inhibitory concentration, it shows the presence of few dead cells and few live cells.

The main reasons for cancer cell survival and proliferation are increased local angiogenesis [18,19], growth suppressor evasion, cell death inhibition, and increased activation of MAPK survival pathway which keeps the cells always in the proliferative phase.
thus promoting the cancerous property [20]. This uncontrolled cell growth, activation of invasion and metastasis, and host immunity avoidance are few of the other properties that retain the cancerous nature of these cells [21]. Nowadays, surgery, chemotherapy, radiation, hormones, and immunotherapy are the main approaches for the cancer treatment. Several problems are associated with the existing treatment methods which include limited efficacy, severe toxicity, and multi-drug resistance [22]. This work is the first of its kind in demonstrating the anticancer activity of telmisartan and irbesartan in a dose-dependent manner. These antihypertensive drugs have been found to have inhibitory roles on the two key processes of cancer, i.e., angiogenesis and MAPK pathway and thus can stand as an effective anticancer drug. This will pave the way for generating novel anticancer drugs with the drugs existing in the market which are relatively less toxic and more effective. The in vitro screening of drugs for anticancer activity is technically simple, quick, cost-effective, and reproducible and provides information about the targets of anticancer action. However, this method has its own limitations as it is error prone and only few molecules reach the human clinical trials. Hence, we need to further extend the study in various in vitro methods and in vivo experimentation in animal models of cancer [23].

CONCLUSION

The study drugs ARBs telmisartan and irbesartan show anticancer activity on the A549 lung carcinoma cell lines. These study results establish a good anticancer activity at lower concentrations of these drugs indicating that more studies on this field are further needed to establish their molecular mechanism of action and to substitute them as effective adjuvant anticancer drugs for lung carcinoma and other cancers which might be safe, tolerable, and effective.

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AUTHORS’ CONTRIBUTION

Dr. K. Punnagai (first author): Conceived the idea, planned the experiments, procured the required cell lines, and analyzed the data and graphs.

Dr. K. Vijayababu (second author and corresponding author): Planned the experiments and conducted the experiments and helped in the calculation of IC50 values and communication with the publication.

Dr. I. Glory Josephine (third author): Added valuable contribution to the conduction of experiments and writing of the manuscript along with the coauthors.

Dr. D. Darling Challathai (fourth author): Supervised the experiments with necessary laboratory requirements and sample preparation and manuscript editing.

CONFLICTS OF INTEREST

The authors declared that they have no conflicts of interest.

REFERENCES

1. Barreras A, Gurk-Turner C. Angiotensin II receptor blockers. Proc (Baylor Univ Med Cent) 2003;16:123-6.
2. de Gasparo M, Catt KJ, Inagami T, Wright JW, Unger T. International union of pharmacology. XXIII. The angiotensin II receptors. Pharmacol Rev 2000;52:415-72.
3. Sleight P, Jakobsen A, Heroy J, Ralph A, Rees T, Shaw M, et al. No HOPE without proof? Do ARBs meet the standard for cardiovascular protection? Medscape J Med 2008;10 Suppl: S6.
4. Soubra L, Nurollin H, Omar A, Salich M. Factors associated with hypertension prevalence and control among Lebanese Type 2 diabetic patients. Int J Pharm Pharm Sci 2018;18:153-9.
5. Nakagami H, Kiomy Osako M, Nakagami F, Shimosato T, Minobe N, Moritani T, et al. Prevention and regression of non-alcoholic steatohepatitis (NASH) in a rat model by metabololast and telmisartan. Int J Mol Med 2010;26:477-81.
6. Jose A, Wilson D, George M, Reshma K Thomas R, Justin A. Comparative study on the beneficial effects of telmisartan and another antihypertensive agents in stroke patients. Int J Pharm Pharm Sci 2017;9:99-102.
7. Pfeffer MA, Swedberg K, Granger CB, Held P, McMurray JJ, Michelson EL, et al. Effects of candesartan on mortality and morbidity in patients with chronic heart failure: The CHARM overall programme. Lancet 2003;362:759-66.
8. Sipahi I, Debanne SM, Rowland DY, Simon DI, Fang JC. Angiotensin II receptor blockade and risk of cancer: Meta-analysis of randomised controlled trials. Lancet Oncol 2010;11:627-36.
9. Blaskaran K, Douglas I, Evans S, van Staa T, Smeeth L. Angiotensin receptor blockers and risk of cancer: Cohort study among people receiving antihypertensive drugs in UK general practice research database. BMJ 2012;344:e2697.
10. Zhang J, Liu J, Chen J, Li X, Wu Y, Chen H, et al. Angiotensin receptor blockers (ARBs) reduce the risk of lung cancer: A systematic review and meta-analysis. Int J Clin Exp Med 2015;8:12656-60.
11. Ferlay J, Soerjomataram I, Ervik M, Dikshit R, Eser S, Mathers C, et al. International Agency for Research on Cancer; 2015. GLOBOCAN. Vol 1 0 Cancer Incidence and Mortality Worldwide. Lyon, France: IARC Cancer Base; 2012.
12. Indian Council of Medical Research; 2013. National Cancer Registry Programme. Three Year Report of Population Based Cancer Registries; 2009-2011.
13. Ford DW, Koch KA, Ray DE, Selecky PA. Palliative and end-of-life care in lung cancer: Diagnosis and management of lung cancer, 3rd ed: American college of chest physicians evidence-based clinical practice guidelines. Chest 2013;143:e498S-e512S.
14. Mosmann T. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. J Immunol Methods 1983;65:55-63.
15. Yamamoto T, Ohashi M, Ho C, Kurtz TW, Rakugi H. TELMISARTAN-induced inhibition of vascular cell proliferation beyond angiotensin receptor blockade and pereoxisome proliferator-activated receptor-gamma activation. Hypertension 2009;54:1353-9.
16. Ruggiero M, Amoroso R. Integrating the MAP kinase signal into the G1 phase cell cycle machinery. Bioessays 2000;22:818-26.
17. Rakusan K, Chvojkova Z, Oliviero P, Ostadalova I, Kolar F, Chassagne C, et al. ANG II type I receptor antagonist irbesartan inhibits coronary angiogenesis stimulated by chronic intermittent hypoxia in neonatal rats. Am J Physiol Heart Circ Physiol 2007;292:H1237-44.
18. Holmgren L, O’Reilly MS, Folkman J. Dormancy of micrometastases;
Balanced proliferation and apoptosis in the presence of angiogenesis suppression. Nat Med 1995;1:149-53.

19. Parangi S, O’Reilly M, Christofori G, Holmgren L, Grosfeld J, Folkman J, et al. Antiangiogenic therapy of transgenic mice impairs de novo tumor growth. Proc Natl Acad Sci U S A 1996;93:2002-7.

20. De Luca A, Maiello MR, D’Alessio A, Pergameno M, Normanno N. The RAS/RAF/MEK/ERK and the PI3K/AKT signalling pathways: Role in cancer pathogenesis and implications for therapeutic approaches. Expert Opin Ther Targets 2012;16 Suppl 2:S17-27.

21. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. Cell 2011;144:646-74.

22. Tan W, Lu J, Huang M, Li Y, Chen M, Wu G, et al. Anti-cancer natural products isolated from Chinese medicinal herbs. Chin Med 2011;6:27.

23. Ahmed SN, Das B, Chakraborty J. Prospective and retrospective animal model used in the pharmacological screening of anti-cancer drug. Int J Curr Pharm Res 2018;10:13-8.