LOW SERUM ANGIOPOIETIN-1, HIGH SERUM ANGIOPOIETIN-2, AND HIGH ANG-2/ANG-1 PROTEIN RATIO ARE ASSOCIATED WITH EARLY ONSET SEPSIS IN SURINAMESE NEWBORNS

Rens Zonneveld,‡† Rianne Jongman,§‡ Amadu Juliana,‡ Wilco Zijlmans,‡* Frans Ploetz,§ Grietje Molema,* and Matijs van Meurs‡§

*Department of Pathology and Medical Biology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands; †Academic Pediatric Center Suriname, Paramaribo, Suriname; §Department of Critical Care, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands; ¶Department of Anesthesiology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands; ‡Department of Pediatrics, Diakonessen Hospital, Paramaribo, Suriname; and §Department of Pediatrics, Tergoii Hospitals, Blaricum, The Netherlands

ABSTRACT—Purpose: Vascular inflammation and leakage in sepsis is mediated by Angiopoietin-1 (Ang-1) and Angiopoietin-2 (Ang-2) and their phosphorylation of the endothelial Tie-2 receptor. This study investigates levels of Ang-1 and Ang-2 in newborns to gain insight in the vascular pathophysiology of early onset sepsis (EOS) within 72 h after birth. Methods: A prospective cohort study was performed among 71 Surinamese newborns treated with antibiotics for suspected EOS and 20 control newborns. Newborns with suspected EOS were divided in two groups: blood culture negative and positive EOS. Ang-1 and Ang-2 levels were measured in serum obtained at the start of antibiotic treatment and at re-evaluation after 48 to 72 h. Results: In this cohort 8.5% of newborns had a positive blood culture. At the start of antibiotic treatment Ang-1 serum levels were lower ($P<0.01$), and Ang-2 and Ang-2/Ang-1 serum protein ratios were higher ($P<0.01$ and $P<0.01$, respectively) in newborns with blood culture positive EOS than in controls. These levels were not dependent on timing of first blood draw after birth. After 48 to 72 h levels of Ang-1 further decreased in blood culture positive EOS, while in the other groups no change was observed. Conclusions: Our findings support the hypothesis that a disbalance in the Angiopoietins plays a role in the vascular pathophysiology of EOS.

KEYWORDS—Angiopoietins, early onset sepsis, newborn, Suriname

ABBREVIATIONS—Ang-1—angiopoietin-1; Ang-2—angiopoietin-2; EOS—early onset sepsis; Tie-2—TEK tyrosine kinase endothelial-Tie-2 receptor

INTRODUCTION

Sepsis is a syndrome with physiologic, pathological, and biochemical changes induced by an infection, and occurs in all age groups (1). Early onset sepsis (EOS) in newborns, defined as onset of sepsis within 72 h after birth, remains a clinical diagnostic and therapeutic challenge due to its nonspecific clinical presentation. This is associated with late discovery and undertreatment of septic newborns or overtreatment with antibiotics of uninfected ones (2–4). These diagnostic and therapeutic problems arise because the pathophysiology of EOS is not completely understood (3–5).

One of the pathological changes in septic patients is microvascular dysfunction leading to increased vascular inflammation and leakage (6). Vascular endothelial cells control these changes through the Angiopoietin/Receptor Tyrosine Kinase (Tie)-2 endothelial receptor system, which is severely disturbed in sepsis (6–9). The system consists of the ligands Angiopoietin-1 (Ang-1) and Angiopoietin-2 (Ang-2) (9). In health, Ang-1-Tie2 binding promotes intracellular Tie-2 phosphorylation, which prevents the occurrence of vascular inflammation and vascular leakage (10). During sepsis, Ang-2 dose dependently competes with Ang-1, which inhibits Tie-2 phosphorylation and induces destabilizing vascular inflammation and leakage (11, 12). In sepsis in children and adults, higher Ang-2 levels and Ang-2/Ang-1 ratios in blood are associated with presence, severity, and outcome of sepsis, while changes in Ang-1 levels are less uniformly present (13–17). To date, there is insufficient knowledge if disturbances in the Angiopoietin/Tie2 system also reflect the activation state of the endothelium during EOS in newborns (18). Furthermore, no data exists on the Angiopoietins during EOS from non-Western countries, such as Suriname.

Therefore, we studied the levels and behavior of Ang-1 and Ang-2 at the start of antibiotic treatment and at re-evaluation...
between 48 to 72 h in Surinamese newborns with suspected EOS. We hypothesized that lower Ang-1 and higher Ang-2 and Ang-2/Ang-1 protein ratio were associated with blood culture positive EOS.

PATIENTS AND METHODS

Study design and subjects

A prospective observational cohort study was performed at the neonatal care facility of the Academic Pediatric Center Suriname at the Academic Hospital Paramaribo. Patients were included in a 14-month period between April 1, 2015 and May 31, 2016. Newborns with a gestational age equal to or above 34 weeks in whom antibiotics were started within the first 72 h of life for suspected EOS were included. Excluded were neonates of whom no serum was obtained or not enough information was available after the study period to confirm outcomes. Written informed consent was obtained from at least one parent for the use of residual serum and clinical information. The study protocol was approved by the Surinamese Medical-Ethical Board (VG-021-14A) and was made available on clinicaltrials.gov (Trial registration: NCT02486783 registered 27/6/2015).

Clinical protocol

For all newborns, the standard local protocol for the management of suspected EOS was followed. This included the start of antibiotics after blood collection for culture and serial laboratory testing of infectious parameters (t = 0). Intravenous ampicillin (50–75 mg/kg/day) and gentamycin (3 mg/kg/ day) were started based on the presence of maternal risk factors for infection (i.e., positive group B streptococcus culture, (premature) prolonged rupture of membranes, intrapartum fever or intrapartum antibiotics) and/or clinical signs of infection of the newborn. Controls were newborns without signs of infection receiving blood draws for hyperbilirubinemia. In these controls, no antibiotics were started. Newborns in whom antibiotics were started were divided into two groups based on blood culture result: blood culture negative EOS and blood culture positive EOS.

Data collection

For all newborns maternal information (i.e., history, pregnancy complications (i.e., presence of diabetes mellitus, pregnancy-induced hypertension or preeclampsia) and maternal risk factors for infection) was recorded, along with gestational age (if unknown according to Ballard), Apgar scores, birth weight, gender, ethnicity, results from laboratory testing (white blood cell counts and C-reactive protein (CRP) levels), duration of antibiotic treatment, blood culture results, hospital course, and mortality.

Sample collection, preparation, and analysis

Blood samples were collected in serum microtainers using standard blood collection during the insertion of a venous cannula. This time point was labeled t = 0. After 48 to 72 h of treatment with antibiotics a second blood sample was obtained using capillary collection. This time point was labeled t = 48 to 72 h. CRP and hematological parameters were determined routinely at the clinical laboratory of the Academic Hospital Paramaribo. Blood was allowed to clot at room temperature and serum was separated by centrifugation at 2,300 x g for 8 min, the serum was harvested and residual sample was stored at −80 °C until further analysis. Frozen samples were transported on dry ice from Suriname to the Netherlands. For analysis, the samples were thawed on ice and immediately analyzed. Measurement of levels of Ang-1 and Ang-2 was performed using the Human Lumienx Screening Assay LXSAAH (R&D Systems, Minneapolis, Minn) according to the manufacturer’s instructions. We determined interassay coefficients of variation (CV) and accepted a maximum of 20%. Median interassay CV ranged from 7.3% to 10.5% for Ang-1 and 4.6% to 10.3% for Ang-2, respectively.

Statistical analysis

Categorical variables were presented as numbers and percentages with 95% CI and compared with chi-square. Continuous variables were presented as median and interquartile range (IQR) Due to the nonparametric nature of the data a Mann–Whitney or Kruskal–Wallis test with Dunn correction for multiple comparisons was used for the analysis of continuous variables. Spearman rho was used to assess bivariant associations between CRP levels and Ang-1 and Ang-2 levels, respectively. P values < 0.05 were considered statistically significant. All the analyses were performed using Prism version 7.0a (Graphpad Software Inc, San Diego, Calif).

RESULTS

Demographics

Of 101 eligible newborns eight newborns were excluded for incomplete clinical information and two for insufficient serum. For the 91 included newborns demographics are given in Table 1. Birth weight, age at presentation, Apgar score, and clinical course at t = 48 to 72 h were distributed unevenly among the three groups. Six (8.5%; 95% CI 3.9–17.2) newborns receiving antibiotic treatment had a positive blood culture (all gram-negative bacteria, Klebsiella pneumoniae (n = 2), Enterobacter cloacae (n = 2) and Escherichia coli (n = 2)).

Newborns with EOS received respiratory and circulatory support more often than controls (P < 0.001). A total of five newborns with EOS died. White blood cell, neutrophil and trombocyte counts, and CRP levels were not different between groups (Table 2).

Levels of angiopoietins

At t = 0, median levels of Ang-1 were significantly lower in blood culture positive EOS (28.3 (28.0) ng/mL) versus controls (77.4 (65.2) ng/mL), P < 0.01 (Table 2) (Fig. 1A). Median Ang-2 levels were higher in blood culture EOS (21.1 (13.3) ng/mL) versus controls (10.2 (1.9) ng/mL), P < 0.001, respectively (Fig. 1B). The Ang-2/Ang-1 protein ratio was higher in blood culture positive EOS (median (IQR) 0.77 (0.77) versus controls (median (IQR) 0.13 (0.13) (P < 0.01) (Fig. 1C)).

There was no difference in median levels of Ang-1, Ang-2, and Ang-2/Ang-1 protein ratio between blood culture negative EOS and controls. At t = 48 to 72 h, median Ang-1 levels had decreased 21-fold in blood culture positive EOS from levels at t = 0 (P = 0.10), while median Ang-2 levels remained high (P = 0.99) (Table 2) (Fig. 1, A and B). Median levels of Ang-1, Ang-2, and Ang-2/Ang-1 protein ratio were not different when comparing blood culture positive or blood culture negative EOS with controls.

Levels of Ang-1 and Ang-2 were tested for dependency on timing of first blood draw (t = 0) after birth. For controls and EOS (blood culture negative plus blood culture positive EOS) median levels at t = 0 were not different between newborns if t = 0 was before 24 h or between 24 to 72 h after birth (Fig. 2, A and B).

Because CRP levels at t = 48 to 72 h increased from levels at t = 0 in blood culture negative and positive EOS (Table 2), correlation of Ang-1 and Ang-2 with CRP was assessed among 44 newborns with blood culture negative (n = 2) EOS in whom all data had been recorded (Fig. 3, A and B). Lower median Ang-1 (rho = −0.46; 95% CI −0.67 to −0.19), but not higher Ang-2, correlated with higher CRP at t = 48 to 72 h.

DISCUSSION

In this study, we investigated the serum levels of Ang-1 and Ang-2 to better understand the vascular pathophysiology in near-term and term Surinamese newborns treated for EOS.
Lower levels of Ang-1, higher Ang-2, and a higher Ang-2/Ang-1 protein ratio in serum of newborns were associated with blood culture positive EOS at the start of antibiotic treatment. Levels of Ang-1 further decreased over time in newborns with blood culture positive EOS and correlated negatively with higher levels of CRP. These results indicate a role for the Angiopoietins in vascular inflammation during EOS in Suriname. An estimated 5% to 10% of total EOS data is from non-Western countries such as Suriname, while there is strong indication that over 90% of global deaths due to EOS occur in these settings (2, 19, 20). Thus, our data add critical basic and clinical knowledge on the true global impact of EOS.

Our results of Ang-1 levels are in line with other studies that reported reduced Ang-1 levels in children associated with septic shock and death (21, 22). The mechanism for low Ang-1 remains poorly understood. While Ang-1 levels are low, the levels of its soluble ligand sTie-2 are higher in the blood of septic patients. Soluble Tie 2 may act as a decoy.

### TABLE 1. Descriptive statistics of the study group (n = 91)

|                      | Early onset sepsis | Blood culture negative (n = 65) | Blood culture positive (n = 6) | P value |
|----------------------|--------------------|--------------------------------|-------------------------------|---------|
| Pregnancy, n (%)     |                    |                                |                               |         |
| Complications†       |                    |                                |                               |         |
|                     | 3 (15)             | 16 (25)                        | 1 (17)                        | 0.63    |
| Chorioamnionitis‡    | 0                  | 18 (28)                        | 0                             |         |
| Mode of delivery, n (%) |                  |                                |                               |         |
| Vaginal              | 12 (60)            | 46 (75)                        | 4 (67)                        | 0.54    |
| Caesarean            | 8 (40)             | 19 (25)                        | 2 (33)                        |         |
| Sex, n (%)           |                    |                                |                               |         |
| Male                 | 9 (45)             | 29 (45)                        | 5 (83)                        | 0.19    |
| Female               | 11 (55)            | 36 (55)                        | 1 (17)                        |         |
| Ethnicity, n (%)     |                    |                                |                               |         |
| Maroon and Creole    | 12 (60)            | 44 (68)                        | 4 (67)                        |         |
| Hindo-Surinamese     | 3 (15)             | 14 (21)                        | 1 (17)                        | 0.61    |
| Other‡               | 5 (25)             | 7 (11)                         | 1 (17)                        |         |
| Gestational age, n (%) (weeks) |            |                                |                               |         |
| 34–37                | 1 (5)              | 22 (34)                        | 0                             |         |
| 37–40                | 14 (70)            | 30 (46)                        | 4 (67)                        | 0.06    |
| >40                  | 5 (25)             | 13 (20)                        | 2 (33)                        |         |
| Apgar score, n (%)   |                    |                                |                               |         |
| <5                   | 0                  | 5 (8)                          | 2 (33)                        | 0.03    |
| Birth weight, median (IQR) (g) |            | 3,130 (700)                   | 2,840 (835)                   | 3,500 (906) | 0.02 |
| Age at presentation, n (%) (h) |            |                                |                               |         |
| <24                  | 4 (20)             | 43 (66)                        | 2 (33)                        |         |
| 24–48                | 7 (35)             | 13 (20)                        | 1 (17)                        | <0.01   |
| 48–72                | 9 (45)             | 9 (14)                         | 3 (50)                        |         |
| Clinical course (at 48–72 h), n (%) |            |                                |                               |         |
| CPAP                 | 0                  | 9 (14)                         | 0                             |         |
| Mechanical ventilation | 0               | 7 (11)                         | 2 (33)                        |         |
| Cardiotonics         | 0                  | 5 (8)                          | 1 (17)                        | <0.001  |
| Mortality            | 0                  | 3 (5)                          | 2 (33)                        |         |

†Presence of pregnancy-induced hypertension, preeclampsia, or diabetes mellitus.
‡Defined as intrapartum fever or administration of antibiotics.
§Includes: Javanese, Chinese, Caucasian, and Amerindian.

CPAP indicates continuous positive airway pressure; IQR, interquartile range; N/A, not applicable.

### TABLE 2. Infection biomarkers in baseline controls and newborns with suspected and blood culture positive early onset sepsis

| Time point | Controls (n = 20) | Blood culture negative (n = 65) | Blood culture positive (n = 6) | P value |
|------------|------------------|---------------------------------|-------------------------------|---------|
| White blood cells (×10^9/L) |        |                                |                               |         |
| t = 0      | 88 (97)          | 15.3 (8.2)                      | 17.5 (9.7)                    | 21.9 (82.4) | 0.68 |
| Neutrophils (×10^9/L) |        |                                |                               |         |
| t = 0      | 72 (79)          | 7.1 (8.5)                       | 9.2 (7.9)                     | 10.2 (34.1) | 0.57 |
| Platelets (×10^9/L) |        |                                |                               |         |
| t = 0      | 83 (91)          | 235 (82)                        | 239 (60)                      | 74 (164.5) | 0.07 |
| C-reactive protein (mg/dL) |        |                                |                               |         |
| t = 0      | 75 (82)          | <0.5 (0)                        | <0.5 (0.7)                    | 0.7 (4.8) | 0.34 |
| t = 48–72 h | 44 (48)          | N/A                             | 0.7 (1.8)                     | 1.4 (16.3) | 0.81 |
| Angiopeoetine-1 (ng/mL) |        |                                |                               |         |
| t = 0      | 91 (100)         | 77.4 (65.2)                     | 82.2 (45.7)                   | 28.3 (28.0) | <0.01 |
| t = 48–72 h | 49 (54)          | 68.9 (44.5)                     | 73.6 (67.3)                   | 13.6 (16.1) | 0.02 |
| Angiopeoetine-2 (ng/mL) |        |                                |                               |         |
| t = 0      | 91 (100)         | 10.2 (1.9)                      | 11.2 (6.9)                    | 21.1 (13.3) | <0.01 |
| t = 48–72 h | 49 (54)          | 9.9 (1.5)                       | 11.8 (4.7)                    | 19.0 (18.9) | 0.07 |

Data presented as median (IQR).

1Data analyzed with a Kruskal-Wallis test between all groups or with a Mann-Whitney test between blood culture negative and positive groups. EOS indicates early onset sepsis; IQR, interquartile range; N/A, not applicable.
receptor binding Ang-1 with high affinity, thereby decreasing its circulating levels. On the other hand, increasing Ang-1 levels, thereby increasing endothelial Tie-2 receptor phosphorylation, may help to inhibit vascular inflammation and leakage. In a clinically relevant murine model, intravenous recombinant Ang-1 treatment was sufficient to improve sepsis-associated organ dysfunctions and survival time, most likely by preserving endothelial barrier function (23).

Higher levels of Ang-2 may be reflective of vascular inflammation and vascular leakage. Intravenous lipopolysaccharide injection in human volunteers, adult human sepsis, and secondary infection in critically ill patients causes higher levels of circulating Ang-2 and higher Ang-2/Ang-1 ratios (24–28). As intracellular Tie-2 phosphorylation cannot be assessed in patients, an increased Ang-2/Ang-1 ratio might be predictive for reduced endothelial Tie-2 receptor phosphorylation with subsequent vascular inflammation and leakage.

EOS can occur following colonization of the newborn with bacterial pathogens following intrauterine infection or in the birth canal during labor (4). Two studies found higher maternal and amniotic fluid levels of Ang-2 in cases of intrauterine infection in at term and preterm birth (29, 30). To our knowledge, placental Ang-2 crossing has not been described. The presence of intrauterine infection may result in EOS and cause subsequent suppression of neonatal levels of Ang-1 and release of neonatal Ang-2 from endothelial cells. Our finding that levels of Ang-1 and Ang-2 are similar between infected newborns included directly after birth and after 24 h supports this hypothesis.

A remarkable finding in our study was that levels of Ang-1 in newborns were up to a 10-fold higher, specifically in healthy newborns and those with blood culture negative EOS, than in children or adults in earlier studies (13–17, 21, 22). Placental levels of Ang-1 and Ang-2 are high during pregnancy and then quickly drop after birth (31, 32). Only one earlier study compared both antepartum and postcaesarean maternal samples with neonatal umbilical cord blood samples (32). Ang-1 concentrations were significantly higher in umbilical samples, suggesting separate Angiopoietin regulation in the newborn.
Animal models of pregnancy may help elucidate the exact dynamics of Angiopoietins in newborns. These animal models may also be instrumental in detecting endothelial Tie-2 receptor phosphorylation in different microvascular beds.

From a clinical perspective, our findings indicate that serial measurement of Angiopoietins may predict or exclude bacteremia in newborns before blood culture results are known. High serial Ang-1 and low serial Ang2/Ang-1 ratio may be extra arguments to discontinue antibiotics, alongside serial measurement of CRP. A known limitation to CRP is its slow synthesis limiting its utility in early prediction of EOS. To overcome this issue, inflammatory mediators that precede CRP synthesis, such as interleukin (IL)-1ß, IL-6, and IL-8, and tumor-necrosis factor (TNF)-α, have been of interest in EOS research. These mediators have short half-lives, which limits their clinical use and establishment of appropriate cut-off values. In our study, levels of Ang-1 remained high in healthy and low in the sickest newborns at re-evaluation 48 to 72 h after the start of antibiotics, indicating persistent association with severity of disease over time and clinical utility. Additionally, TNF-α has been shown to correlate with Ang-2 levels in adult patients with sepsis. For these reasons it would be interesting to evaluate temporal relations of the Angiopoietins with a panel of inflammatory mediators, such as TNF-α, IL-6, and IL-1ß in EOS.

Our study has several limitations. First, our sample size was relatively small to assess relevance of the Angiopoietins as clinical biomarkers and results may have been confounded by birth weight and asphyxia, which were distributed unevenly among the groups. Second, as levels of the Angiopoietins were determined with a Luminex Screening Assay we were unable to compare levels with results from other studies measured with ELISA, and small sample volumes acquired in newborns precluded assessment of other inflammatory mediators. Future studies will focus on eliminating these limitations to enable us to validate the current observations in newborns in Surinamese newborns.

In summary, our data show changes in the ligands of the Angiopoietin/Tie2 endothelial receptor system Ang-1 and Ang-2 in EOS and support the hypothesis that increased vascular inflammation and increased vascular leakage leads to microvascular dysfunction in the pathophysiology of EOS. The potential impact of intra-uterine-infection deserves attention in future investigations to further elucidate dynamics of Angiopoietins in newborns with and without EOS.

ACKNOWLEDGMENTS

The authors acknowledge the efforts of all employees of the Clinical Laboratory of the Academic Hospital Paramaribo and the Central Laboratory of Suriname, Paramaribo, Suriname, for assistance with sample storage, handling, and transport.

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