Association Between Dietary Intakes and Plaque Vulnerability Measured by Optical Coherence Tomography in Patients With Coronary Heart Disease: A Mediation Analysis of Inflammatory Factors

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Although studies have proven that diet has a critical role in preventing or delaying atherosclerosis and is far simpler to adjust and adhere to than other risk factors, the underlying mechanisms behind this effect remain not well comprehended. The purpose of this investigation was to determine the impact of inflammatory factors on the connection between dietary ingestion and coronary plaque fragility as measured via optical coherence tomography (OCT) in patients with coronary heart disease (CHD). This research eventually comprised 194 participants with CHD who met the inclusion and exclusion criteria. Semi-quantitative food frequency questionnaire (SQFFQ) was utilized to investigate dietary consumption status, serum levels of inflammatory biomarkers were analyzed using enzyme-linked immunosorbent assay, and OCT was employed to identify the plaque susceptibility of causative lesions in the body. Following correction for statistically meaningful possible confounders in univariate analysis, quartiles of soy and nuts, fruits and vitamin C were negatively associated with coronary plaque vulnerability. Conversely, the upper quartile group of sodium intake had 2.98 times the risk of developing vulnerable plaques compared with the most minimal quartile group. Meanwhile, we observed an inverse dose–response connection between vitamin C consumption and inflammatory biomarkers as well as plaque vulnerability. More importantly, tumor necrosis factor-α (TNF-α) and interleukin-6 (IL-6) were significant mediators of the connection between vitamin C and plaque vulnerability, suggesting that vitamin C may inhibit the atherosclerotic inflammatory process by decreasing the expression of IL-6 and TNF-α, thereby reducing the risk of vulnerable plaques. These new findings provide crucial clues to identify anti-inflammatory dietary components as effective therapeutic approaches in the management of CHD, while also providing some insights into their mechanisms of action.

Keywords: coronary heart disease, plaque vulnerability, coronary atherosclerosis, optical coherence tomography, diet, nutrient, inflammation, inflammatory cytokine
INTRODUCTION

For despite improvements in interventional and pharmacological therapy for CHD, it is still the primary reason of mortality and disability-adjusted life years (DALYs) worldwide (1). Obstructive atherosclerotic plaque produces a restriction in blood supply to the myocardium, causing CHD (2). It is indicated by robust evidence that CHD evolvement has resulted by the pivotal mechanism of coronary plaque vulnerability, and vulnerable plaque which is distinguished by a large lipid necrotic core and thin fibrous cap is prone to rupture and lead to coronary thrombosis. OCT is a catheter-based imaging technique that uses near-infrared light to obtain cross-sectional images of the coronary arteries. OCT is enabled to evaluate many morphological characteristics of coronary atherosclerosis in vivo due to its unusually high resolution (10–20 m) (3). Clinical findings revealed that OCT-detected vulnerable plaque was inextricably linked to a higher chance of developing major adverse cardiac events (MACE) (4–6). Therefore, effective measures to control and reduce plaque vulnerability are essential to reduce the poor prognosis of CHD.

Among various identified risk factors for CHD, food and eating habits plays a critical influence and are preventable factors (7). Many research have increased our knowledge about the association between nutritional intake and heart health during the last few decades. Increased consumption of vegetables and fruits, nuts, antioxidant vitamins, folate, and dietary fiber may be preventive factors for the incidence and development of CHD, according to prospective studies and meta-analyses (8–13). On the other side, increased consumption of cholesterol, eggs, red meat, processed meat and sugar-sweetened drinks may raise the chance and development of CHD (14, 15). A good diet has also been linked to a decreased incidence of MACE in big populations (16). In patients with CHD treated with percutaneous coronary intervention (PCI), poor consumption of fruits (OR = 0.30, 95% CI = 0.12–0.68) and vegetables (OR = 0.48, 95% CI = 0.27–0.91) are both independent risk factors for in-stent restenosis (ISR) (17). Further research using OCT to evaluate plaque vulnerability, we have found that salt and sodium are dietary components that contribute to plaque vulnerability, while vegetables, fruits, dietary fiber, folate and vitamin C have been revealed to be dietary preventive factors (18). However, the mechanism between dietary intake and plaque vulnerability is unclear.

In fact, atherosclerosis is an inflammatory process, with inflammatory pathways present throughout the entire process, from the onset and progression of atherosclerosis to plaque rupture, erosion, and thrombus formation (19). Inflammatory variables such as high-sensitivity C-reactive protein (hs-CRP), TNF-α, and IL-6 are closely connected to plaque lipid core enlargement and fibrous cap breakdown, suggesting that inflammation and immune cell activation play a key role in defining plaque vulnerability (20–22). At the same time, earlier study has established a clear correlation between inflammation and the overall diet and specific dietary components (23, 24). Fruits and vegetables have been demonstrated to lower inflammation, whereas red meat intake has been shown to promote it (25, 26). Consuming more healthful and anti-inflammatory food ingredients like fruits and green leafy vegetables while reducing consumption of pro-inflammatory foods like processed meat and sugar-sweetened beverages, according to a recent meta-analysis, may be crucial in lowering the risk of CHD and associated mortality (27).

Following the Mediterranean diet (MeDiet) decreases plasma levels of inflammatory cytokines linked with various phases of atheroma plaque formation in old age patients with a high propensity to CHD, according to a five-year multicenter randomized controlled experiment in Spain (28). Miller et al. reported that increasing dietary fiber consumption resulted in considerably reduced plasma indicators of inflammatory state (29). A considerably larger study from the United Kingdom discovered a substantial inverse linear relationship between fiber consumption and hs-CRP (30). Despite this, few studies have been done to see if inflammatory variables play a role in the relationship between dietary intake and coronary plaque vulnerability.

Consequently, our objectives were to: (i) investigate the link between dietary intakes and plaque vulnerability; and (ii) identify inflammatory mediators and their impact on the relationship between food nutrients and coronary plaque vulnerability.

MATERIALS AND METHODS

Study Participants

This study was a cross-sectional design, and used non-probabilistic sampling. From March 2020 to September 2021, 220 CHD patients were recruited to undergo OCT tomography at Harbin Medical University’s Second Hospital Affiliated. The participants were between the ages of 20 and 75. The research excluded patients with current infection, obvious cognitive deficits, mental comorbidity, long-term anti-inflammatory medication usage, left ventricular ejection fraction (LVEF) < 40% and individuals with allergy, rheumatoid disease, or malignancy. In total, 26 participants were excluded based on the exclusion criteria. Thus, 194 individuals with CHD were eventually enrolled throughout this research (Figure 1).

Study Procedures

After informed consent was obtained, these participants were invited to fill questionnaires prior to conducting OCT evaluations. Highly qualified investigators offered in-depth explanations of the surveys to the subjects, and provided necessary help for participants to complete the questionnaire smoothly. During the OCT examination, blood samples were taken. Meantime, sociodemographic and medical information was extracted from their clinical documents and, if required, verified by subjects. Prior to actually participating in the research, every individuals provided written informed permission for participation. The Harbin Medical University (China) Human Ethics Review Committee approved the study (protocol code
ky2020-058, approval date March 20, 2020), which was carried out in accordance with the Helsinki Declaration.

Assessment of Demographic and Clinical Data

Among the information gathered were: gender, age, body mass index (BMI), diagnosis of acute myocardial infarction, familial CHD, hypertension (systolic pressure ≥ 140 mmHg or diastolic pressure ≥ 90 mmHg on two separate occasions, or whether the patient was using anti-hypertensive medications) (31), hyperlipidemia (blood total cholesterol level ≥ 5.7 mmol/L and/or triglycerides concentration ≥ 1.7 mmol/L) (32), diabetes mellitus (DM; fasting plasma glucose level ≥ 7.0 mmol/L, a random venous plasma glucose level ≥ 11.1 mmol/L, 2-h plasma glucose concentration ≥ 11.1 mmol/L following an oral glucose tolerance test, or any combination of these) (33), smoking status (1 cigarette each day or more than 100 cigarettes consumed in an entire life) (34) and drinking condition (a daily alcohol intake of ≥ 24 g) (35).

Fasting plasma glucose (FPG), total cholesterol (TC), triglyceride (TG), low-density lipoprotein cholesterol (LDL-C), and high-density lipoprotein cholesterol (HDL-C) were among the clinical lab indicators examined and collected. The location of the assay was the Department of Clinical Laboratory, Harbin Medical University's Second Hospital Affiliated.

Assessment of Dietary Intakes

Prior conducting this inquiry, every personnel participating inside this investigation received training in the relevant research procedures. The SQFFQ, which was created and verified for the Chinese, was used to assess food consumption and consumption frequency of the study samples. The SQFFQ included 3 parts: food checklist, frequency of food consumption, and weight for every item consumed. Grain and potatoes, legumes and nuts,
vegetables, fruits, domestic animals and poultry, milk, eggs, fish and shrimp, oils and salt were the ten categories in this survey. Subjects recorded their average intake frequency per food over the course of a year on an eight-point scale ranging from “almost never” to “3 times one day or more” (visualized utilizing prevalent container and weight units, for instance, bowls, cups, and spoons). Their mean quotidiam consumption for every foodstuff was obtained via computing weighted frequency everyday and consumption amount per unit, then normalized to gram/day. The Chinese version of the SQFFQ has been validated and can be used to classify study subjects according to the food or nutrient intake during one year (36). The Food Nutrition Calculator V2.7.3 from the China CDC Nutrition and Food Safety Institute was used to calculate daily dietary nutrient consumption.

**Markers of Inflammation Cytokines Measurement**

At the time of OCT checking, fresh venous circulating blood sample (5 mL) was taken from each patient in order to test the serum levels of inflammatory cytokines. Afterward, a 15-min centrifugation at 3,000 rpm and 4°C was performed on these test samples, aliquots were then kept in reserve at −80°C until further analysis was necessary. TNF-α, IL-6, and hs-CRP concentrations were measured using an enzyme-linked immunosorbent assay, with analytic sensitivities of 4.69, 4.69, and 8.00 pg/ml, respectively. Blind method analysis of a single lot was performed. The aforementioned three inflammatory biomarkers had less than 10% inter- and intra-assay coefficients of variation.

**Optical Coherence Tomography Image Acquisition and Analysis**

The intracoronary frequency- or time-domain OCT system (C7XR system, Saint Jude Medical, Westford, MA, United States) was used in this study to do OCT imaging (37). Offline software was utilized to capture and evaluate pictures of coronary arteries bearing culprit plaques. Two seasoned professionals completed the review of all OCT photographs respectively following the standards of OCT Clinical Expert Consensus Statement, and they were not privy to the participant’s electronic healthcare data. Once the conclusions of the two evaluators were inconsistent, the arbitration and compromise was conducted by one additional researcher. The following OCT imaging markers (37) were assessed: Lipid plaques (low signal region with diffuse boundary) and fibrous plaques (homogeneous high-backscattering region) were separated. Lipid arcs of lipid-rich plaques (maximum lipid arc > 2 quadrants) were recorded at 1-mm intervals throughout the lesion, with an average value of the thinnest region tested three times utilized as the fibrous cap thickness parameter. As shown in Figure 2, a lipid plaque with a lipid arc of ≥ 90° and a fibrous cap thickness of < 65 μm was designated as thin cap fibroatheroma (TCFA) in this study as the specific indication utilized to detect plaque vulnerability (38). Fibrous cap discontinuities and plaque cavities were signs of plaque rupture (39). The thrombus presented with an irregular lump extending into an arterial lumen of at least 250 μm in diameter; Macrophage accumulation was characterized by a rise in signal strength inside of plaque, together with the presence of heterogeneous backward shadows; Microchannel showed as speckled black rounded areas with diameter between 50 and 300 μm inside the plaque, and appeared in more than three consecutive frames; Calcification was described as an area with low backscattering signal and a sharp border. Cholesterol crystals were defined as linear, highly backscattering structures within the plaque (40).

**Statistical Analysis**

IBM SPSS Statistics was used to conduct statistical analysis (V.26.0, IBM Corp, Armonk, NY, United States). A statistically significant two-sided p value of 0.05 was used. Before analysis, variables that were not normally distributed were natural-logarithmically converted to achieve near normality, as established by the Kolmogorov-Smirnov test. When applicable, continuous data is provided as the mean (standard deviation), or median (25th, 75th) and evaluated using Student’s t-test or non-parametric test (Mann–Whitney U test). Categorical variables are reported as a percentage, and chi-square tests were used to compare groups. To correspond to a normal distribution, data for inflammatory biomarkers were standardized using log transformations.

The correlations between food and nutrient intakes and all inflammatory biomarkers and atherosclerotic plaque features factors were investigated using the Spearman correlation analysis. The connection of food and nutrient intakes with atherosclerotic plaque features measured by OCT was investigated using logistic regression analysis. The dependent variable was susceptible plaque characteristics, the independent variables were quartiles of food and nutrient intakes that gave a p value < 0.1 in univariate analysis, and any confounding variables that were statistically significant in univariate analysis were adjusted for. For food and nutrient intake categories, multivariable logistic regression models were used to estimate the odds ratio (OR) and 95% confidence interval (CI) of plaque vulnerability, with the lowest intake grouping as the reference. The quartiles median value was used as a quasi-continuous variable in the model to calculate p values for trends.

This study evaluated the mediating effects of inflammatory biomarkers on the link between food and nutrient intakes and plaque vulnerability using Hayes’ (41) PROCESS tool of model 4 (42). As shown in Figure 3, bootstrap mediation analyses were performed. In the absence of mediating variables, the bootstrap analyses made it easier to assess the direct association between food and nutrient intakes and plaque vulnerability (path c; simple relationship in Figure 3A). Following that, a link was found between food and nutrient intakes and each of the suggested mediators (path a), as well as each mediator and plaque vulnerability (path b). The effect of food and nutrient intakes on plaque vulnerability by TNF-α, IL-6 was defined as an indirect effect (path a*b) in the mediation analysis. After correcting for inflammatory indicators, a direct effect (path c’) was defined as the effect of food and nutrient intakes on plaque vulnerability (Mediated relationship in Figure 3B). The mediation analysis was carried out, using 5,000 bootstrap samples. The mediating impact
**FIGURE 2** | Vulnerable plaque and related characteristics in OCT images. (A) Lipid plaque; (B) Thin cap fibroatheroma; (C) Plaque rupture; (D) Thrombus; (E) Macrophage accumulation; (F) Microchannel; (G) Calcification; (H) Cholesterol crystals.

**FIGURE 3** | Simple relationship and Mediated relationship. Non-Mediated pathways (A) and Mediated pathways (B) between food and nutrients and plaque vulnerability. Path $c'$ represents the direct effect of food and nutrients on plaque vulnerability with the mediator included in the model. The indirect effect is the product of path $a$ and path $b$ (path $a \cdot b'$). Each mediator was considered in a separate statistical model.
RESULTS

Patient Characteristics and Laboratory Results

A total of 194 patients were enrolled in the project, with an average age of 56.08 ± 10.99 years. Patients were split into two groups based on whether or not TCFA occurred: the vulnerable plaque group (98, 50.5%) and the non-vulnerable plaque group (96, 49.5%). Table 1 shows that a higher prevalence of myocardial infarction was found (p = 0.015) in the vulnerable plaque group, and the TG, TC and LDL-C levels were significantly higher (p = 0.030, p < 0.001, and p < 0.001, respectively) than the non-vulnerable plaque group. The vulnerable plaque group had greater levels of tumor necrosis factor-α (TNF-α) and interleukin-6 (IL-6) in the laboratory than the non-vulnerable plaque group (both of p = 0.001).

Optical Coherence Tomography Findings

The OCT findings of the two groups were shown in Table 2. When compared to the non-vulnerable plaque group, the vulnerable plaque group had a longer lipid core, thinner fibrous cap thickness, and bigger lipid arc (all p < 0.001). Meanwhile, patients with susceptible plaques had a higher rate of plaque rupture, macrophage infiltration, thrombus, and cholesterol crystal formation than those with non-vulnerable plaques (all p < 0.001). Furthermore, susceptible plaques exhibited a greater rate of calcification and microchannels (p = 0.006 and p = 0.022, respectively).

Multivariate Regression Analysis for the Correlations of Dietary Intakes With Plaque Vulnerability

The baseline nutritional intake of the study participants was summarized in Table 3. The study participants were divided into four groups based on their quartile levels of daily food and nutrient intake, which generated a p value < 0.1 in univariate analysis (Table 3). To find relationships with susceptible plaque, researchers used univariate and multivariable logistic regression models with the lowest intake group as a reference. A backward stepwise multivariable logistic regression model with admission criteria of p < 0.05 and removal criteria of p > 0.1 was used to include factors identified significant in univariate analysis and important variables for professional consideration. After adjusting for gender, age, body mass index, history of myocardial infarction, TC, TG and LDL-C, Table 4 revealed that rising quartiles of dietary consumption of soy and nuts, fruits, and vitamins C remained independently linked with lower risk of susceptible plaque (p for trend with adjustment: 0.028, 0.002, and 0.003, respectively).

The OR between the greatest and lowest levels of soy and nut intake was 0.33 (95% CI = 0.13–0.83). Fruits were associated with 0.26 times (95% CI = 0.10–0.70) decreased risk of incident vulnerable plaques when comparing moderate (Q3 level) to low-consumers, this association was slightly weakened in the highest consumers (Q4 level, OR = 0.37, 95% CI = 0.14–0.99). Vitamin C intake was found to be inversely associated with the development of susceptible plaques, with OR of 0.10 (95% CI = 0.03–0.38) and 0.23 (95% CI = 0.07–0.74) in the highest (Q4) and moderate (Q3) levels, respectively, compared to the lowest level. When compared to low-consumers of sodium, high-consumers had a considerably higher risk of susceptible plaques (OR = 2.98, 95% CI = 1.06–8.37; all p < 0.05).

Correlation Analysis Among the Variables

As shown in Figure 4, vitamin C had negative correlations with TNF-α (r = −0.274, p < 0.001), IL-6 (r = −0.172, p = 0.016) and TCFA (r = −0.270, p < 0.001), rupture (r = −0.216, p = 0.002), lipid plaque (r = −0.217, p = 0.002), thrombus (r = −0.205, p = 0.004), macrophage infiltration (r = −0.173, p = 0.016), cholesterol crystal (r = −0.168, 0.020). Soy and nuts were negatively correlated with TCFA (r = −0.149, p = 0.037) and cholesterol crystal (r = −0.202, p = 0.005), and fruits were associated with rupture (r = −0.286, p < 0.001) and cholesterol crystals (r = −0.161, p = 0.024). Furthermore, TNF-α and IL-6 were positively correlated with TCFA (r = 0.250, p < 0.001 and r = 0.202, p = 0.005, respectively). However, there was no significant correlation between soy and nuts, fruits and TNF-α, IL-6.

Mediation of the Relationship Between Vitamin C Consumption and TCFA by Inflammation Biomarkers

To see if inflammation biomarkers (TNF-α and IL-6) mediated the effect of vitamin C on TCFA, we used SPSS Macro mediatiing analysis (Figure 5). In the mediation analysis, age, gender, BMI, history of myocardial infarction, TG, TC, and LDL-C were all used as covariates. The indirect effects (path a × b) of TNF-α significantly mediated the effects of vitamin C intake Q3 level (95% CI = −0.742, −0.072) and Q4 level (95% CI = −0.734, −0.064) on TCFA when the lowest consumer level (Q1) of vitamin C was used as the reference (Table 5). The 95% Bootstrap confidence interval of Q3 (a2 = −3.923, b = 0.085, a2b = −0.335) and Q4 (a3 = −3.748, b = 0.085, a3b = −0.320) levels relative to Q1 level excluded zero, indicating significant relative mediating effect (Figure 5A and Table 5). That is, the Q3 vitamin C consuming group produced 3.923 (a2 = −3.923, p < 0.01) less TNF-α than the Q1 level group, so the incidence of TCFA was correspondingly reduced (b = 0.085, p = 0.020). The vitamin C consumption group with Q4 level reduced TNF-α by 3.748 (a3 = −3.748, p < 0.01) compared with Q1 level group, resulting in lower TCFA (b = 0.085, p = 0.020). The relative direct effect was significant (95% CI = −2.029, −0.266), indicating that the risk of TCFA decreased by 1.148 (c′ = −1.148, p = 0.011) in the group with Q4 level of vitamin C intake, after excluding the mediating effect (Table 5). And the same is true for IL-6. Taking the lowest intake group as a reference, IL-6 played a relative indirect
TABLE 1 | Characteristics of vulnerable plaque and non-vulnerable plaque individuals (N = 194).

| Variables                        | Total sample (n = 194) | Vulnerable plaque (n = 98) | Non-vulnerable plaque (n = 96) | Test value  | p-value |
|----------------------------------|------------------------|----------------------------|-------------------------------|-------------|---------|
| Age [years, M(SD)]              | 56.08 (10.99)          | 56.26 (11.45)              | 55.90 (10.55)                | t = 0.227   | 0.821   |
| BMI [kg/m², M(SD)]              | 24.91 (3.89)           | 24.91 (3.77)               | 24.91 (4.02)                 | t = -0.001  | 0.999   |
| Male (n, %)                     | 114 (68.8%)            | 63 (64.3%)                 | 51 (53.1%)                   | χ² = 2.493  | 0.114   |
| Smoking history (n, %)          | 115 (59.3%)            | 58 (59.2%)                 | 57 (59.4%)                   | χ² = 0.001  | 0.978   |
| Drinking history (n, %)         | 88 (45.4%)             | 48 (49.0%)                 | 40 (41.7%)                   | χ² = 1.046  | 0.306   |
| Hypertension (n, %)             | 79 (40.7%)             | 39 (39.8%)                 | 40 (41.7%)                   | χ² = 0.070  | 0.791   |
| Diabetes mellitus (n, %)        | 30 (15.5%)             | 15 (15.5%)                 | 15 (15.6%)                   | χ² = 0.004  | 0.961   |
| Hyperlipidemia (n, %)           | 29 (14.9%)             | 12 (12.2%)                 | 17 (17.7%)                   | χ² = 1.139  | 0.286   |
| Family history of CHD (n, %)    | 50 (25.8%)             | 27 (27.6%)                 | 23 (24.0%)                   | χ² = 0.327  | 0.567   |
| Myocardial infarction (n, %)    | 182 (93.8%)            | 96 (98.0%)                 | 86 (89.6%)                   | χ² = 5.863  | 0.015   |
| Laboratory findings             |                        |                            |                               |             |         |
| Blood glucose [mmol/L, M(IQR)]  | 6.64 (5.65, 8.06)      | 6.58 (5.69, 7.99)          | 6.78 (5.54, 8.16)            | z = 0.104   | 0.918   |
| Log TG [mmol/L, M(SD)]          | 0.14 (0.25)            | 0.17 (0.27)                | 0.10 (0.23)                  | t = -2.187  | 0.030   |
| Log TC [mmol/L, M(SD)]          | 0.65 (0.12)            | 0.69 (0.13)                | 0.62 (0.09)                  | t = 4.841   | <0.001  |
| Log LDL-C [mmol/L, M(SD)]       | 0.08 (0.12)            | 0.09 (0.12)                | 0.07 (0.12)                  | t = 0.909   | 0.364   |
| Log hs-CRP [mg/L, M(SD)]        | 0.45 (0.14)            | 0.50 (0.12)                | 0.39 (0.13)                  | t = 6.157   | <0.001  |
| Inflammatory biomarkers          |                        |                            |                               |             |         |
| Log TNF-α [ng/L, M(SD)]         | 1.05 (0.15)            | 1.09 (0.16)                | 1.02 (0.13)                  | t = 3.495   | 0.001   |
| Log IL-6 [pg/mL, M(SD)]         | 1.30 (0.27)            | 1.37 (0.28)                | 1.24 (0.26)                  | t = 3.509   | 0.001   |
| Log hs-CRP [mg/L, M(SD)]        | 0.62 (0.42)            | 0.64 (0.41)                | 0.61 (0.44)                  | t = 0.582   | 0.561   |

M(SD), mean (standard deviation); M(IQR), median (interquartile range); BMI, body mass index; CHD, coronary heart disease; TC, total cholesterol; TG, triglyceride; LDL-C, low density lipoprotein cholesterol; HDL-C, high-density lipoprotein cholesterol; TNF-α, tumor necrosis factor-α; IL-6, interleukin-6; hs-CRP, high sensitivity C-reactive protein; Log, logarithmical.

TABLE 2 | OCT characteristics of vulnerable plaque and non-vulnerable plaque individuals (N = 194).

| Characteristics                        | Total sample (n = 194) | Vulnerable plaque (n = 98) | Non-vulnerable plaque (n = 96) | Test value  | p-value |
|----------------------------------------|------------------------|----------------------------|-------------------------------|-------------|---------|
| Quantitative variable                  |                        |                            |                               |             |         |
| Plaque length [mm, M(IQR)]            | 9.50 (3.18, 15.58)     | 12.35 (7.88, 19.28)        | 5.45 (1.40, 10.80)            | z = 5.938   | <0.001  |
| Fibrous cap thickness [μm, M(IQR)]     | 56.67 (40.00, 70.00)   | 43.33 (33.33, 53.33)       | 70.00 (64.91, 93.33)          | z = -10.497 | <0.001  |
| Lipid arc [deg, M(SD)]                 | 137.34 (83.73)         | 194.68 (55.38)             | 78.81 (65.25)                 | t = 13.346  | <0.001  |
| Qualitative variable                  |                        |                            |                               |             |         |
| Rupture (n, %)                         | 122 (62.9%)            | 89 (90.8%)                 | 33 (34.4%)                   | χ² = 66.191 | <0.001  |
| Lipid plaque (n, %)                    | 128 (66.0%)            | 93 (94.9%)                 | 35 (36.5%)                   | χ² = 73.784 | <0.001  |
| Macrophage infiltration (n, %)        | 144 (74.2%)            | 84 (85.7%)                 | 60 (62.5%)                   | χ² = 13.661 | <0.001  |
| Thrombus (n, %)                        | 164 (84.5%)            | 94 (95.9%)                 | 70 (72.9%)                   | χ² = 19.627 | <0.001  |
| Calcification (n, %)                   | 108 (54.6%)            | 63 (64.3%)                 | 43 (44.8%)                   | χ² = 7.436  | 0.006   |
| Microchannel (n, %)                    | 101 (52.1%)            | 59 (60.2%)                 | 42 (43.8%)                   | χ² = 5.261  | 0.022   |
| Cholesterol crystal (n, %)             | 127 (65.5%)            | 83 (84.7%)                 | 44 (45.8%)                   | χ² = 32.382 | <0.001  |

M(IQR), median (interquartile range); M(SD), mean (standard deviation).

mediating role in the effects of Q2 (95% CI = −0.788, −0.010), Q3 (95% CI = −0.875, −0.030) and Q4 (95% CI = −0.825, −0.010) levels of vitamin C consumption on TCFA (Table 5). Increased vitamin C intake with Q2 (α₁ = −10.117, b = 0.023, α₁b = −0.236), Q3 (α₂ = −11.862, b = 0.023, α₂b = −0.276) and Q4 (α₃ = −11.113, b = 0.023, α₃b = −0.259) level was associated with a decrease in serum levels of inflammatory factors, and furthermore, serum inflammatory factors levels mediated the beneficial effects of vitamin C on vulnerable plaques (Figure 5B and Table 5). In the meantime, the vitamin C consumption with Q4 level had relative direct effects on TCFA after controlling for IL-6 (c’₃ = −1.297, p = 0.004, 95% CI = −2.168, −0.426; Table 5).

DISCUSSION

Our analysis indicated that soy and nuts, fruits, and vitamin C were revealed to be dietary protective factors for plaque vulnerability, whereas sodium was found to be a dietary risk factor for plaque vulnerability. Vitamin C intake was found to be negatively related to IL-6 and TNF-α, whereas serum IL-6 and TNF-α were found to be favorably connected to vulnerable plaque. More importantly, TNF-α and IL-6 mediated the effects of vitamin C on vulnerable plaque, which was the first study to our knowledge to examine the mediating effects of inflammatory variables on the link between vitamin C and coronary plaque.
susceptibility using OCT. The findings of this study reveal that an elevated inflammatory response may explain the link between diet and plaque vulnerability in CHD patients, implying that a vitamin C-rich diet may help to reduce vulnerable plaque progression by lowering the inflammatory response.

Major advancements in coronary artery disease prevention have required early diagnosis of susceptible plaques in recent decades. A vulnerable plaque is a coronary atherosclerotic plaque that is prone to rupture, and TCFA is the most common phenotype of vulnerable plaque today. Large necrotic lipid core with an overlying thin fibrous cap (<65 µm) infiltrated by macrophages characterizes TCFA as the antecedent lesion of plaque rupture (43). TCFA and vulnerable plaques related features can be identified and evaluated by OCT, this permits in vivo detection of macrophages, intimal vasculature, and structures suggestive of calcium accumulation within atherosclerotic plaques, as well as plaque rupture, fibrous cap thickness, and intraluminal thrombus (44). This study by OCT exhibited that vulnerable plaque characteristics, including thinner fibrous cap, more prominent lipid arc, longer lipid core length and TCFA, were seen more frequently compared with non-vulnerable plaque (all p < 0.01). Meanwhile, the incidence of rupture, lipid plaque, macrophage infiltration, thrombus, calcification and cholesterol crystal in vulnerable plaques was fundamentally higher than that in non-vulnerable plaques (all p < 0.01), and huge contrast was also found in the incidence of microchannel (p = 0.022; Table 2). The natural history of coronary atherosclerosis has been shown to be a dynamic process ranging from early lesion development to more advanced plaques complicated by acute rupture and thrombosis; all of these indicators play key roles in the mechanisms of vulnerable plaque formation and rupture, according to studies. However, in cardiovascular illness, the production and disruption of susceptible plaque, followed by thrombosis, is regarded to be a crucial event that leads to acute coronary syndrome (ACS) and sudden cardiac death (43). As a result, physicians would be able to identify individuals at high risk for ACS and catastrophic CHD by detecting susceptible plaque in vivo.

Diet and eating habits have been demonstrated to have a significant impact on the occurrence and progression of heart disease, the world’s leading cause of mortality. Healthy eating habits play a crucial influence in the primary and secondary

### TABLE 3 | Dietary intake status of vulnerable plaque and non-vulnerable plaque patients (N = 194).

| Variables                  | Total Sample (N = 194) | Vulnerable plaque (N = 98) | Non-vulnerable plaque (N = 96) | Test value | p-value |
|---------------------------|------------------------|---------------------------|-------------------------------|------------|---------|
| Cereal and potatoes [g/d, M(SD)] | 504.79 (151.79) | 519.04 (162.03) | 490.23 (139.92) | t = 1.324 | 0.187 |
| Soy and nuts [g/d, M(IQR)] | 9.99 (3.33, 24.58) | 6.67 (2.92, 21.19) | 13.33 (3.33, 31.71) | z = -2.077 | 0.038 |
| Vegetables [g/d, M(IQR)] | 336.96 (256,94, 472.08) | 312.54 (230.63, 425.06) | 361.52 (273.87, 513.89) | z = -2.678 | 0.007 |
| Fruits [g/d, M(IQR)] | 161.32 (69.95, 283.54) | 132.53 (55.06, 229.78) | 216.50 (111.70, 330.04) | z = -3.777 | <0.001 |
| Livestock and poultry [g/d, M(IQR)] | 63.34 (28.04, 120.13) | 59.99 (27.68, 120.13) | 67.26 (28.70, 119.47) | z = -0.645 | 0.519 |
| Milk [g/d, M(IQR)] | 0.00 (0.00, 64.28) | 0.00 (0.00, 52.68) | 0.00 (0.00, 89.29) | z = -0.630 | 0.529 |
| Eggs [g/d, M(IQR)] | 36.26 (18.86, 52.80) | 35.16 (18.52, 52.80) | 37.56 (18.86, 52.80) | z = -0.466 | 0.641 |
| Fish and shrimp [g/d, M(IQR)] | 18.93 (8.33, 44.20) | 17.86 (8.33, 36.54) | 21.43 (8.33, 44.64) | z = -1.009 | 0.313 |
| Oils [g/d, M(IQR)] | 44.44 (33.33, 50.52) | 45.83 (34.58, 54.52) | 41.67 (33.33, 50.00) | z = 0.680 | 0.497 |
| Salt [g/d, M(IQR)] | 6.67 (5.33, 8.33) | 6.67 (5.33, 8.33) | 6.46 (5.00, 8.33) | z = 1.868 | 0.062 |
| Energy [kcal/d, M(SD)] | 92.78 (27.39) | 91.29 (28.12) | 94.31 (26.67) | t = -0.704 | 0.482 |
| Fat [g/d, M(IQR)] | 94.70 (74.00, 120.60) | 91.85 (72.10, 116.85) | 95.35 (74.90, 127.60) | z = -0.957 | 0.339 |
| Carbohydrate [g/d, M(SD)] | 424.67 (117.18) | 424.18 (122.97) | 425.16 (111.61) | t = -0.058 | 0.954 |
| Log Dietary fiber [g/d, M(SD)] | 1.20 (0.17) | 1.17 (0.17) | 1.23 (0.16) | t = -2.769 | 0.006 |
| Cholesterol [mg/d, M(IQR)] | 359.00 (224.75, 485.25) | 347.50 (215.50, 484.50) | 369.00 (242.50, 490.00) | z = -0.533 | 0.594 |
| Log Vitamin A [µgRE/d, M(SD)] | 2.82 (0.23) | 2.79 (0.24) | 2.84 (0.22) | t = -1.739 | 0.084 |
| Thiamine [mg/d, M(IQR)] | 1.42 (0.39) | 1.38 (0.37) | 1.46 (0.41) | t = -1.367 | 0.173 |
| Log Riboflavin [mg/d, M(SD)] | 0.05 (0.16) | 0.02 (0.16) | 0.07 (0.16) | t = -2.059 | 0.041 |
| Niacin [mg/d, M(SD)] | 19.91 (5.90) | 19.59 (6.07) | 20.24 (5.75) | t = -0.768 | 0.443 |
| Vitamin B6 [mg/d, M(IQR)] | 0.41 (0.32, 0.52) | 0.42 (0.31, 0.51) | 0.41 (0.33, 0.54) | z = -0.719 | 0.472 |
| Folate [µg/d, M(IQR)] | 84.65 (59.05, 118.20) | 78.45 (52.48, 96.90) | 94.45 (71.13, 133.30) | z = -3.009 | 0.003 |
| Log Vitamin C [mg/d, M(SD)] | 2.10 (0.20) | 2.05 (0.19) | 2.15 (0.20) | t = -3.646 | <0.001 |
| Log Vitamin D [µg/d, M(IQR)] | 1.50 (0.60, 3.28) | 1.95 (0.68, 3.20) | 1.25 (0.53, 3.30) | z = 0.668 | 0.504 |
| Log Vitamin E [mg/d, M(SD)] | 1.80 (0.13) | 1.80 (0.13) | 1.81 (0.13) | t = -0.880 | 0.380 |
| Log Calcium [mg/d, M(SD)] | 2.71 (0.19) | 2.69 (0.19) | 2.74 (0.19) | t = -1.857 | 0.065 |
| Phosphorus [mg/d, M(SD)] | 1431.01 (414.88) | 1396.46 (403.20) | 1466.29 (425.69) | t = -1.173 | 0.242 |
| Log Potassium [mg/d, M(SD)] | 3.39 (0.13) | 3.37 (0.13) | 3.41 (0.14) | t = -1.921 | 0.056 |
| Log Sodium [mg/d, M(SD)] | 3.58 (0.11) | 3.60 (0.11) | 3.57 (0.11) | t = 1.493 | 0.137 |

M(SD), mean (standard deviation); M(IQR), median (interquartile range); Log, logarithmical. Natural-logarithmically transformed was performed on the values of dietary fiber, vitamin A, riboflavin, vitamin C, vitamin E, calcium, potassium and sodium.
TABLE 4 | Logistic regression analysis on the associations between quartiles of food and nutrients and vulnerable plaque (N = 194).

| Variables                      | Univariate | Multivariable |
|--------------------------------|------------|--------------|
|                                |            |              |
|                                | Adjusted OR (95% CI) | p-value | p for trend | Adjusted OR (95% CI) | p-value | p for trend |
| **Model 1: Food intakes**      |            |              |
| Soy and nuts (g/d)             |            |              |
| Q1 (0 [≤3.33])                 | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (7.14 [3.34–9.99])         | 0.58 (0.23, 1.48) | 0.253 |          | 0.45 (0.14, 1.43) | 0.177 |
| Q3 (15.47 [10.00–24.52])      | 0.53 (0.26, 1.11) | 0.093 |          | 0.48 (0.20, 1.17) | 0.105 |
| Q4 (50.00 [≥24.53])           | 0.41 (0.20, 0.87) | 0.02 | 0.03 | 0.33 (0.13, 0.83) | 0.019 | 0.028 |
| **Vegetables (g/d)**           |            |              |
| Q1 (194.17 ≤257.71)           | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (298.63 [257.72–336.96])   | 0.75 (0.33, 1.69) | 0.482 |          |                   |      |
| Q3 (395.25 [336.97–471.20])   | 0.51 (0.23, 1.15) | 0.106 |          |                   |      |
| Q4 (548.64 [≥471.21])         | 0.32 (0.14, 0.73) | 0.007 | 0.004 |                   |      |
| **Fruits (g/d)**               |            |              |
| Q1 (27.33 [≤70.35])           | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (116.15 [70.36–161.32])    | 1.17 (0.50, 2.73) | 0.718 |          | 1.40 (0.52, 3.76) | 0.505 |
| Q3 (219.10 [161.33–280.20])   | 0.28 (0.12, 0.65) | 0.003 |          | 0.26 (0.10, 0.70) | 0.007 |
| Q4 (345.37 [≥280.21])         | 0.27 (0.12, 0.62) | 0.002 | <0.001 | 0.37 (0.14, 0.99) | 0.047 | 0.002 |
| **Salt (g/d)**                 |            |              |
| Q1 (4.50 [≤5.33])             | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (6.25 [5.34–6.67])         | 2.96 (1.37, 6.38) | 0.006 |          |                   |      |
| Q3 (8.33 [≥8.34])             | 2.64 (1.22, 5.70) | 0.013 |          |                   |      |
| Q4 (10.00 [≥8.34])            | 2.98 (0.82, 10.83) | 0.098 | 0.027 |                   |      |
| **Model 2: Nutrient intakes**  |            |              |
| Dietary fiber (g/d)            |            |              |
| Q1 (10.20 [≤12.1])            | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (14.50 [12.2–16.4])        | 0.66 (0.29, 1.47) | 0.307 |          |                   |      |
| Q3 (17.95 [16.5–20.7])        | 0.53 (0.24, 1.20) | 0.13 |          |                   |      |
| Q4 (24.96 [≥20.8])            | 0.35 (0.15, 0.79) | 0.012 | 0.011 |                   |      |
| Vitamin A (µgRE/d)             |            |              |
| Q1 (366.00 [≤445.0])          | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (562.00 [446.0–681.5])     | 1.05 (0.47, 2.35) | 0.906 |          |                   |      |
| Q3 (779.00 [681.6–886.0])     | 0.61 (0.28, 1.36) | 0.227 |          |                   |      |
| Q4 (1222.00 [≥887.0])         | 0.54 (0.24, 1.20) | 0.129 | 0.070 |                   |      |
| Riboflavin (mg/d)              |            |              |
| Q1 (0.73 [≤0.85])             | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (0.99 [0.86–1.12])         | 0.59 (0.26, 1.33) | 0.207 |          | 0.36 (0.10, 1.33) | 0.127 |
| Q3 (1.29 [1.13–1.50])         | 0.43 (0.19, 0.96) | 0.038 |          | 0.75 (0.17, 3.28) | 0.700 |
| Q4 (1.75 [≥1.51])             | 0.52 (0.23, 1.17) | 0.115 | 0.111 | 3.11 (0.48, 20.28) | 0.236 | 0.074 |
| Folate (µg/d)                  |            |              |
| Q1 (41.90 [≤59.3])            | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (76.00 [59.4–84.7])        | 1.25 (0.55, 2.82) | 0.591 |          |                   |      |
| Q3 (96.45 [84.8–118.2])       | 0.81 (0.37, 1.79) | 0.606 |          |                   |      |
| Q4 (148.10 [≥118.3])          | 0.32 (0.14, 0.74) | 0.006 | 0.003 |                   |      |
| Vitamin C (mg/d)               |            |              |
| Q1 (71.60 [≤99.8])            | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (112.35 [99.9–130.4])      | 0.81 (0.36, 1.85) | 0.618 |          | 0.35 (0.12, 1.02) | 0.054 |
| Q3 (149.40 [130.5–182.1])     | 0.43 (0.19, 0.98) | 0.044 |          | 0.23 (0.07, 0.74) | 0.014 |
| Q4 (211.75 [≥182.2])          | 0.24 (0.10, 0.56) | 0.001 | <0.001 | 0.10 (0.03, 0.38) | 0.001 | 0.003 |
| Calcium (mg/d)                 |            |              |
| Q1 (313.00 [≤387.0])          | 1.00 (Reference) | 1.00 (Reference) |
| Q2 (453.50 [388.0–524.5])     | 0.89 (0.40, 1.99) | 0.77 |          | 2.52 (0.66, 9.62) | 0.176 |
| Q3 (590.00 [524.6–696.0])     | 0.61 (0.27, 1.36) | 0.226 |          | 1.29 (0.26, 6.36) | 0.753 |
| Q4 (849.00 [≥697.0])          | 0.45 (0.20, 1.02) | 0.055 | 0.036 | 0.41 (0.06, 2.59) | 0.342 | 0.082 |

(Continued)
TABLE 4 (Continued)

| Variables | Univariate | Multivariable |
|-----------|------------|--------------|
|          | Adjusted OR (95% CI) | p-value | p for trend | Adjusted OR (95% CI) | p-value | p for trend |
| Sodium (mg/d) | | | | | | |
| Q1 (2851.00 ≤2321.00) | 1.00 (Reference) | | | 1.00 (Reference) | | |
| Q2 (3455.50 >2321.00) | 1.33 (0.60, 2.96) | 0.479 | | 1.16 (0.42, 3.20) | 0.769 | |
| Q3 (4225.00 >3231.00) | 1.00 (0.45, 2.22) | 1.00 (Reference) | 0.127 | 0.127 | 0.198 | 0.107 |
| Q4 (5233.50 >4623.00) | 1.87 (0.84, 4.20) | 0.127 | 0.198 | 2.96 (1.06, 8.37) | 0.038 | 0.111 |

Backward elimination method was applied to avoid multicollinearity. OR (95% CI) are presented with correction. The above model was adjusted for gender, age, body mass index, history of myocardial infarction, low-density lipoprotein cholesterol (LDL-C), total cholesterol (TC) and triglyceride (TG). OR, odds ratio; CI, confidence interval; Q, quartile.

Prevention of cardiovascular disease, according to meta-analysis studies (45). RCTs and observational studies both revealed an inverse relationship between adherence to good dietary patterns, particularly the Mediterranean diet, and cardiovascular risk, including major cardiovascular events (myocardial infarction, stroke, or death from cardiovascular causes) (16, 46). Diets low in whole grains, vegetables, fruits, and nuts but high in sodium were the leading dietary patterns for mortality, according to the latest systematic study of dietary patterns across 195 countries (47). Our findings back up the idea that a low intake of fruits and soy/nuts in the diet is linked to susceptible plaque, and that increasing the intake of cardioprotective food groups (soy/nuts, fruit) in real life is linked to improvements in plaque features in CHD patients. When compared to the lowest-consumers, consumption of ≥ 24.53 g/d (median value 50.00 g/d) of legumes and nuts from daily diet was related with a significantly lower incidence of susceptible plaque (OR = 0.33, 95% CI = 0.13–0.83, p for trend = 0.028). When fruits consumption climbed to 219.10 (161.33–280.20) g/d in CHD patients, a substantial protective impact against susceptible plaques was seen (OR = 0.26, 95% CI = 0.10–0.70, p = 0.007), and this positive effect exhibited a linear trend with the rise of fruits intake (p for trend = 0.002). Our statistically significant findings corroborated previous research, bolstering previous conclusions.

When compared to a low-fat diet, the PREDIMED research found that a Mediterranean diet rich in nuts could reduce CHD end points by about 30% (hazard ratio: 0.72, 95% CI: 0.54, 0.95) (46). The large levels of unsaturated fats, soluble fiber, plant protein, vitamins, minerals, and phytochemicals in nuts are likely to be responsible for their health benefits. Nuts high in long-chain n-3 polyunsaturated fatty acids (PUFAs) and omega-3 PUFAs may reduce plasma TGs and platelet aggregation, as well as have anti-inflammatory properties, which may reduce CHD risk and prevent coronary plaque progression. Nuts would also have provided phytosterols (which are known to inhibit cholesterol absorption and enhance fecal cholesterol excretion) and polyphenols (which are known antioxidants) (48). The high fiber and antioxidant content of legumes may explain their cardiovascular advantages. After 2 months of ingestion, a combination of soluble fiber and soy protein has been shown to drastically lower LDL-C by 20% (49). Increases in legume intake (25 g/d) were connected to increases in paraoxonase-1 activity (PON1, a crucial HDL-bound antioxidant enzyme) and decreases in cholesteryl ester transfer protein (CETP, pro-atherogenic when excessively active) activity, according to Hernández et al. (13). These mechanisms could explain why increasing consumption of dietary legumes and nuts was linked to a lower risk of susceptible plaque. This is the first time, to our knowledge, that the effect of increasing legume and nut consumption on coronary atherosclerotic susceptible plaque risk in humans has been reported.

The evidence to date strongly suggests that increasing fruit consumption may lower the risk of CHD incidence and mortality. According to a meta-analysis, increasing fruit intake by 200 g per day reduces CHD risk by 10%. Globally, 710,000 CHD deaths were attributed to a fruit and vegetable consumption of less than 500 g per day, and this number grew to 1.34 million CHD deaths when 800 g per day was used as the recommended intake (10). The current study also found an inverse relationship between fruit intake and plaque vulnerability in CHD patients, implying that fruits were dietary preventive factors against plaque vulnerability. According to extant research, fruits and vegetables’ preventive mechanisms may include not only some of the recognized bioactive nutrient benefits based on antioxidant, anti-inflammatory, and electrolyte capabilities, but also their functional features, such as low glycemic load and energy density (50). As a result, it’s critical to concentrate on the impact of fruits and vegetables, as well as probable mechanisms of action and dietary recommendations for lowering the risk of vulnerable plaque. Despite the fact that various reviews and meta-analyses of fruit and vegetable diets have found an inverse relationship between intake and CHD risk, epidemiological evidence for causality is mixed. Indeed, differences in the types of fruits and vegetables ingested by research populations, as
well as whether cooked or raw veggies are preferred, may introduce heterogeneity. As a result, stratified analysis of fruits and vegetables separately could help to reduce heterogeneity. This study analysis for fruits and vegetables separately, which were limited to fresh vegetables and fruits, not fruit juices, dry fruits, frozen and canned. But results from this study did not find a significant benefit of increasing vegetables intake for plaque vulnerability prevention. This result may be related to the habit of eating raw vegetables with high-salt sauces in the selected subject samples. Heat processing, on the other hand, can increase the availability of some bioactive substances (such as lycopene from tomatoes and carotenoids from carrots) and convert folate polyglutamate in vegetables into monoglutamate, which is more bioavailable (51). In addition, the research center locations in the cold northeast of China may limit the variety of vegetables available. Fruits and vegetables should be consumed as part of a healthy diet. We should not only consider the daily intake of fruits and vegetables in future dietary guidelines, but also their nutrient content, as well as how they are prepared (storage) and consumed (cooking).

Many elements, including water soluble vitamins, fiber, sterols, and phytochemicals, are credited with the benefits of a diet rich in fruits and vegetables. Dietary vitamins have received the most attention in the literature. These vitamins are hypothesized to defend against reactive oxygen species (ROS), which contribute to atherogenesis by producing superoxide, and so present a potential method by which they may prevent CHD. Fruits (especially citrus fruits like guavas and oranges) are the primary source of these anti-oxidant vitamins, particularly vitamin C (52, 53). Low dietary vitamin C consumption has been linked to an elevated risk of CHD in women in studies (54). Vitamin C intake was found to be inversely related to all-cause

**FIGURE 4** | Correlations between soy and nuts, fruits and Vitamin C nutrients, Inflammatory factors and Plaque characteristics. *p < 0.05; **p < 0.01. TNF-α, tumor necrosis factor-α; IL-6, interleukin-6; TCFA, thin cap fibroatheroma.
and cardiovascular mortality in a prospective cohort analysis (9, 55). Antioxidant vitamin C intake lowered the incidence of poor cardiovascular events, according to a 22-year follow-up research. Low vitamin C consumption from fruits and vegetables was also found to predict intracoronary in-stent restenosis in Type D patients in a prior study (17). The current study also
found an inverse relationship between food-derived vitamin C and plaque vulnerability in CHD patients. And the relationship showed a strongly linear, dose-response trend. We observed a 77% lower risk of vulnerable plaque in the Q3 level versus lowest quartile of vitamin C intake. The protective effect became more significant when intake increased to the Q4 level (median value 211.75 mg/d). It is worth mentioning that the median intake of vitamin C in Q2 to Q4 levels, is greater than the current Recommended Dietary Allowance (RDA) for adults in China (100 mg/d), the median intake of vitamin C in Q4 level is still greater than a proposed intake (PI) for Chinese adults (200 mg/d) which established for the purpose of preventing or reducing the risk of non-communicable diseases, so it would be necessary to consider a larger dose to get the protector effect in high-risk groups of CHD. From this, a diet of more than five daily servings of fresh fruits and vegetables is recommended to achieve an intake of 200 mg/d for vitamin C. Overall, these results support our previous research findings (18). In animal models and in vitro cell culture systems, the link between dietary vitamin C and atheroma formation has been investigated (57). The antioxidant hypothesis has affirmed the protective effects of vitamin C. Meanwhile, the lipid oxidation theory claims that dietary vitamin C can slow the progression of atherosclerosis by lowering the creation of oxidized LDL cholesterol (50). However, the PREDIMED trial found that eating vitamin C-rich fruits and vegetables lowers plasma levels of inflammatory biomarkers such hs-CRP, IL-6, and TNF-α, which are linked to distinct stages of atheroma plaque development (28). As a result, we believe that vitamin C derived mainly from antioxidants diet (fruits and vegetables) is a key mechanism for preventing the formation of vulnerable plaque.

Vitamin C consumption was found to be negatively related to IL-6, TNF-α and vulnerable plaque in this study, but TNF-α and IL-6 were positively connected to vulnerable plaque. To put it another way, we found an inverse dose-response relationship between vitamin C intake with inflammation and plaque vulnerability. Further mediation analysis revealed that IL-6 and TNF-α, were wholly or partially mediating these correlations between vitamin C and vulnerable plaque, implying that the effect of increasing dietary vitamin C intake on vulnerable plaque risk was related to lower levels of serum inflammatory markers. These findings show that the inflammatory pathway may play a role in vitamin C’s preventive actions in the development of vulnerable plaques. Inflammation is hypothesized to aid in the onset and progression of atherosclerosis, as well as the development of acute thrombotic problems (49). The infiltration and retention of LDL cholesterol in the artery wall, in particular, is a crucial initiating event that activates an inflammatory response and promotes the progression of atherosclerosis. Monocyte recruitment is aided by higher amounts of adhesion molecules and chemotactic factors, as well as the promotion of activation, differentiation, and proliferation processes, as well as macrophage immobilization in the inflammatory plaque. The macrophages are eventually converted into foam cells. M-CSF promotes the production of TNF-α and release of proinflammatory cytokines including IL-6 (58), which leads to tissue damage. Activated macrophages then produce inflammatory cytokines that can destabilize lesions and promote thrombus formation, all of which can eventually lead to plaque activation and rupture (59). We believe that vitamin C’s anti-inflammatory and immunomodulating effects are mediated by downregulation of leukocyte adhesion molecules, resulting in a decrease in proinflammatory interleukins IL-6, TNF-α, and their receptors. In fact, inflammatory biomarkers appeared to be inversely associated to vegetable and fruit-based patterns and a priori healthy dietary patterns, as evidenced by intervention studies exploring the Mediterranean diet (28). Studies on the relationship between the dietary inflammatory index (DII, which measures the overall inflammatory impact of food) and cardiovascular diseases have shown that eating a better anti-inflammatory diet can help reduce the risk of CHD and related death (27, 60, 61). Previous research has also found a link between dietary inflammation and plaque susceptibility (60, 62, 63). All of these evidences back up the study’s findings.

Furthermore, this study discovered a favorable link between dietary sodium intake and plaque susceptibility. Subjects in the highest quartile progressed susceptible plaque 2.98 times greater than those in the lowest quartile. Previous research has found that as sodium intake rose, the prevalence of carotid atherosclerotic plaques increased (66 vs 90%, Tertile I vs Tertile III, p = 0.02) (64). According to the most recent meta-analysis, higher sodium consumption was linked to a higher risk of CHD (65). The current study also showed that salt intake has a negative impact on the formation of coronary atherosclerotic plaques. Dietary salt may affect endothelial function, as seen by increased matrix metalloproteinase-9 activity and serum CRP levels (64, 66, 67).

Study Limitations
This study has certain limitations that should be acknowledged. To begin with, this was a single-center, cross-sectional mediation study with a limited sample size, which could result in selection bias. Second, because this was not a follow-up interventional trial, we are unable to establish causal correlations between variables. Third, while we included confounding variables that have been used in other CHD research, our analysis is confined to the common confounding variables that are typically addressed when investigating diet-CHD relationships. As a result, residual confounding from unmeasured variables cannot be ruled out. Finally, although it is convenient to collect data using SQFFQ, there may be recall bias in reporting dietary consumption based on subjects’ eating habits over the previous 12 months, including omission of food items and inaccurate nutrient consumption. In any case, large-scale multicenter prospective studies are needed to investigate the causal association between diet, inflammatory biomarkers, and susceptible plaque progression/regression, as well as to confirm the results of this investigation.

CONCLUSION
In conclusion, the findings of this study add to the growing body of evidence supporting dietary prevention of poor CHD prognosis, suggesting that favoring cardioprotective food groups
and nutrients (including soy and nuts, fruits and vitamin C) and limiting sodium intake help to reduce the likelihood of vulnerable plaque. More importantly, we also have demonstrated for the first time that inflammatory biomarkers IL-6 and TNF-α may mediate the relations between vitamin C and vulnerable plaque. This supports the hypothesis that vitamin C consumption could affect vulnerable plaque development through the inflammatory pathway. These new findings add to the underlying processes by which dietary categories and nutrients may influence CHD outcomes by adding another layer of regulation. CHD patients with suboptimal diet especially those in low vitamin C intake, may be potential beneficiaries of anti-inflammatory therapy. Our findings highlight the relevance of a high-quality food-based diet in general, and we provide a suggestion for a better diet to support the complementary therapeutic therapy of CHD patients at high risk for vulnerable plaques.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Human Ethics Review Committee at Harbin Medical University (China). The patients/participants provided their written informed consent to participate in this study.

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AUTHOR CONTRIBUTIONS

LL and ZZ have conducted investigation and written the original draft of the manuscript. YW and GL were involved in data curation and analysis. XG and BY participated in the OCT analysis and writing – review and editing. PL was responsible for project conception and project supervision. All authors have read and agreed to the published version of the manuscript.
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