Gender-specific relationship between thigh muscle and fat mass and brain amyloid-β positivity

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Abstract

Background: The relationship of specific body composition in the thighs and brain amyloid-beta (Aβ) deposition remained unclear, although there were growing evidence that higher muscle and fat mass in thighs had a protective effect against cardiometabolic syndromes. To determine whether muscle mass and fat mass in the thighs affected amyloid-beta (Aβ) positivity differently in relation to gender, we investigated the association of muscle mass and fat mass with Aβ positivity using positron emission tomography (PET) in individuals without dementia.

Methods: We recruited 240 participants (134 [55.8%] males, 106 [44.2%] females) without dementia ≥45 years of age who underwent Aβ PET, bioelectrical impedance analysis (BIA) and dual-energy X-ray absorptiometry (DEXA) scans of the hip in the health promotion center at Samsung Medical Center in Seoul, Korea. Lower extremity skeletal muscle mass index (LASMI) was measured using BIA, and gluteofemoral fat percentage (GFFP) was estimated using DEXA scans of the hip. We investigated the associations of LASMI and GFFP with Aβ positivity using logistic regression analyses after controlling for age, APOE4 genotype, and cognitive stage.

Results: Higher muscle mass in the thighs, measured as LASMI (odds ratio [OR]=0.27, 95% confidence interval [CI] 0.08 to 0.84, \(p=0.031\)) was associated with a lesser risk of Aβ positivity in only females. Higher fat mass in the thighs, measured as GFFP (OR=0.84, 95% CI 0.73 to 0.95, \(p=0.008\)) was associated with a lesser risk of Aβ positivity in only males. However, the association between LAMSI (\(p\) for interaction= 0.810), GFFP (\(p\) for interaction= 0.075) and Aβ positivity did not significantly differ by gender. Furthermore, LAMSI only negatively correlated with centiloid (CL) values in females (\(r=-0.205, p=0.037\)), and GFFP only negatively correlated with CL values only in males (\(r=-0.253, p=0.004\)).

Conclusions: Our findings highlight the importance of recognizing that gender differences exist with respect to the specific body composition to potentially protect against Aβ deposition. Therefore, our results may help in designing gender-specific strategies for controlling body composition to prevent Aβ deposition.

Keywords: Amyloid-β (Aβ), Muscle, Fat, Thigh, Gender

Background

Deposition of amyloid-beta (Aβ) is the most prominent pathological change in Alzheimer’s disease (AD). According to many studies on in-vivo Aβ biomarkers, brain Aβ deposition precedes brain atrophy and cognitive impairment. Cognitively normal participants with Aβ biomarkers are reported to be more likely to develop...
mild cognitive impairment (MCI) or dementia than those without Aβ biomarkers [1–3]. Thus, Aβ positivity is an important predictor of AD dementia prognosis.

Growing body of evidence shows that weight loss is associated with a higher risk of AD [4, 5]. Weight loss is also reported to be the early sign of AD [6]. Furthermore, there were several studies investigating the associations of BMI or weight loss with Aβ deposition [7, 8]. However, since BMI reflects nonspecific measures of body composition, previous studies using BMI were not able to investigate which body components (ie, muscle mass and fat mass) might be associated with a higher risk of Aβ deposition.

Previous studies have suggested that thigh circumference may be an important determinant of cardiometabolic syndrome [9] and mortality [10]. Both increased fat and muscle mass in the thighs, which reflect for a large composition of human body [11, 12], have protective effects against cardiometabolic syndrome [13, 14]. Specifically, higher fat mass in the thighs was found to be associated with a better metabolic marker status including higher adiponectin levels, lower insulin resistance, lower cholesterol levels, and lower C-reactive protein levels [15–18]. Higher thigh fat mass also prevents cardiometabolic syndrome, including hypertension, diabetes, and hyperlipidemia [19, 20]. In contrast, progressive loss of muscle mass in the thighs with aging, which is known as sarcopenia [21], is associated with a greater risk of cardiovascular disease [22, 23]. In addition, there is increasing evidence that lower muscle mass is related to metabolic impairment and poor health outcomes, including increased morbidity and mortality rates [24, 25]. Considering that cardiometabolic syndromes are closely associated with the development of AD, it would be reasonable to expect that decreased fat and muscle mass of the thighs might be associated with increased risk of Aβ deposition in individuals without dementia. Furthermore, previous studies have revealed differences in body composition between males and females. In fact, it was found that fat mass was lower and muscle mass was higher in males than in females [14]. Therefore, it is possible that Aβ deposition in males might be vulnerable to decreased fat mass, while Aβ deposition in females might be vulnerable to decreased muscle mass.

The objective of our study was to investigate the association of muscle mass in the thighs, as measured using bioelectrical impedance analysis (BIA), and fat mass in the thighs, as measured using dual-energy X-ray absorptiometry (DXA) scans of the lower body, with Aβ positivity using positron emission tomography (PET) in individuals without dementia. Specifically, we determined whether fat and muscle mass in the thighs affected Aβ positivity differently between males and females.

**Methods**

**Study Participants**

We enrolled 240 participants without dementia ≥45 years of age who underwent a full health screening examination, including BIA and DXA scans of the hip in the health promotion center at Samsung Medical Center (Seoul, Korea) between August 2015 and October 2020. All patients also underwent standardized neuropsychological test battery using the Seoul Neuropsychological Screening Battery 2nd edition (SNSB-II) [26, 27], blood tests, brain magnetic resonance imaging (MRI), and Aβ PET. Participants without dementia were participants with normal cognition (NC) and those with MCI. All participants with NC met the following criteria: (1) no medical history that was likely to affect cognitive function based on Christensen’s health screening criteria [28]; (2) no objective cognitive impairment in any cognitive domain on a comprehensive neuropsychological test battery (above at least -1.0 SD of age-adjusted norms on any cognitive test); and (3) independence in activities of daily living. All patients with MCI met the criteria for MCI with the following modifications [29, 30]: (1) subjective cognitive complaints by the participants or caregivers; (2) objective memory impairment below -1.0 SD on verbal or visual memory tests, (3) no significant impairment in activities of daily living, and (4) non-demented status.

We excluded participants with significant whiter matter hyperintensities (cap or band >10 mm and longest diameter of deep white matter lesion >25 mm), structural lesions including cerebral infarction, intracranial hemorrhage, brain tumors, and hydrocephalus on MRI, and abnormal laboratory results on complete blood count, electrolyte, vitamin B12 and folate levels, syphilis serology, and liver/kidney/thyroid function tests.

All participants underwent BIA and DXA scans of the hip on the same day, and these measurements were performed within three years before or after the Aβ PET. The median time interval between the measures was 4 months (interquartile range, 2–7 months). Two hundreds and twelve (88.3%) out of 240 participants had less than 12-month interval between BIA and DXA scans and Aβ PET.

The institutional review board of the Samsung Medical Center approved this study. Written informed consent was obtained from all participants.

**Aβ PET acquisition**

All participants underwent Aβ PET (18F-florbetaben PET and 18F-flutemetamol PET scans using a Discovery StE PET/CT scanner (GE Medical Systems, Milwaukee, WI, USA). For 18F-florbetaben PET or 18F-flutemetamol PET, a 20-minute emission PET scan in dynamic mode (consisting of 4 × 5 min frames) was performed 90 min after
an injection of a mean dose of 311.5 MBq \(^{18}\text{F-florbetaben}\) or 197.7 MBq \(^{18}\text{F-flutemetamol}\), respectively. Three-dimensional PET images were reconstructed in a \(128 \times 128 \times 48\) matrix with \(2 \text{ mm} \times 2 \text{ mm} \times 3.27 \text{ mm}\) voxel size using the ordered-subsets expectation maximization algorithm \((^{18}\text{F-florbetaben, iteration} = 4 \text{ and subset} = 20; \ ^{18}\text{F-flutemetamol, iteration} = 4 \text{ and subset} = 20)\).

A\(\beta\) PET quantification using centiloid values
We used a centiloid (CL) method previously developed by our group [31] to standardize the quantification of A\(\beta\) PET images obtained using different ligands. The CL method for FBB and FMM PET enables the transformation of the standardized uptake value ratio (SUVR) of FBB and FMM PETs to CL values directly without conversion to the \(^{11}\text{C-labeled Pittsburgh compound SUVR}.

There are three steps to obtain CL values [31]: (1) pre-processing of PET images, (2) determination of CL global cortical target volume of interest (CTX VOI), and (3) conversion of SUVR to CL values. First, to pre-process the A\(\beta\) PET images, PET images were co-registered to each participant’s MR image and then normalized to a T1-weighted MNI-152 template using the SPM8 unified segmentation method. We used T1-weighted MR image correction with the N3 algorithm only for intensity non-uniformities, without applying corrections to the PET images for brain atrophy or partial volume effects. Second, we used the FBB-FMM CTX VOI defined as areas of AD-specific brain A\(\beta\) deposition in our previous study [31]. Briefly, to exclude areas of aging-related brain A\(\beta\) deposition, the FBB-FMM CTX VOI was generated by comparing SUVR parametric images (with the whole cerebellum as a reference area) between 20 typical patients with Alzheimer’s disease-related cognitive impairment (AD-CTX) and 16 healthy elderly participants (EH-CTX) who underwent both FBB and FMM PET scans. To generate the FBB-FMM CTX VOI, the average EH-CTX image was subtracted from the average AD-CTX image. We then defined the FBB-FMM CTX VOI as areas of AD-related brain A\(\beta\) accumulation common to both FBB and FMM PET. Finally, the SUVR values of the FBB-FMM CTX VOI were converted to CL units using the CL conversion equation. The CL equation was derived from the FBB-FMM CTX VOI separately for FBB and FMM PET and applied to the FBB and FMM SUVR.

To determine the participants’ CL cut-off-based A\(\beta\) positivity, we applied the optimal cut-off value derived using a k-means cluster analysis in 527 independent samples of participants with normal cognition. The cut-off value was set at 27.08, representing the 95th percentile of the lower cluster [32], and the whole cerebellum was used as a reference region.

Thigh muscle mass measurement
To obtain body mass index (BMI) and appendicular skeletal muscle mass (ASM) of the bilateral lower limbs, all participants underwent BIA using a multifrequency BIA device (InBody 720; InBody Co., Ltd., Seoul, Korea) according to the manufacturer’s guidelines [33]. The appendicular skeletal muscle mass index in the lower extremity (LASMI) was calculated by dividing the sum of the ASM in the bilateral lower limbs by the square of the height (ASM in bilateral lower extremity/[height]²). According to previous results that LASMI was highly correlated with thigh muscle volume in MRI scans of thigh [34], we considered LASMI as a proxy marker of muscle mass in the thigh.

Standardized neuropsychological test battery
All participants underwent the SNSB-II [26, 27], which includes standardized and validated tests of various cognitive functions. The SNSB-II evaluates lots of cognitive factors including verbal and visual memory, visuospatial function, language, praxis, components of Gerstmann syndrome (acalculia, agraphia, right/left disorientation, finger agnosia), and frontal/executive functions. We chose to use six cognitive measures, which were representative and important neuropsychological tests to evaluate the cognitive function in five cognitive domains as follows: (1) Memory: the Seoul Verbal Learning Test SVLT delayed recall and Rey-Osterrieth Complex Figure Test (RCFT) delayed recall; (2) Language: Korean version of the Boston Naming Test; (3) Visuospatial function: RCFT copying Test; (4) Frontal executive function: the Stroop Test color reading; and (5) Attention: Digit Span Test backward.
**Statistical analyses**

All statistical analyses were performed separately in males and females. We used independent t-tests and chi-squared tests to compare the demographic and clinical characteristics of Aβ-positive (Aβ+) and Aβ-negative (Aβ−) groups. To explore the association between BMI and Aβ positivity, we performed a logistic regression analysis with BMI as predictor after controlling for age, APOE4 genotype, and cognitive stage (NC and MCI). To check the correlation among BMI, LASMI, and GFFP, we used Pearson's correlation with BMI as a dependent variable and LASMI or GFFP as an independent variable. To investigate the association between body composition (muscle and fat mass) in the thighs and Aβ positivity, we performed a logistic regression analysis with LASMI and GFFP together as predictors after controlling for age, APOE4 genotype, and cognitive stage (NC and MCI). To further validate the relationship between Aβ burden and body composition (muscle and fat mass) in the thighs, we explored the relationship using quantified CL values rather than cut-off-based categorization. In this analysis, we used partial correlation with CL values as dependent variables and LASMI or GFFP as independent variables after controlling for age, APOE4 genotype, and cognitive stage (NC and MCI). Finally, to evaluate the effect modification of gender and body composition in the thighs on Aβ positivity, we performed multivariable logistic regression analysis with LASMI, GFFP, and gender together as main effects and LASMI*gender and GFFP*gender as interaction effects after controlling for age and APOE4 genotype in all participants (males and females).

All reported p-values were two-sided, and the significance level was set at 0.05. All analyses were performed using R version 4.3.0 (Institute for Statistics and Mathematics, Vienna, Austria; www.R-project.org).

**Results**

**Clinical characteristics of participants**

Among the 240 participants, there were 134 males with a mean age of 71.3±6.7 years and 106 females with a mean age of 69.9±8.1 years (Table 1). There were no differences between males and females in the frequency of Aβ+ (32.8% and 38.7%, p=0.421) (Table 1) and in the frequency of MCI stage (53.7% and 42.5%, p=0.108) (Additional file 1: Table S1). Males (13.7±4.0) had a significantly higher mean years of education than females (12.0±4.8, p=0.002). However, rates of hypertension, diabetes, and APOE4 genotype were not different between males and females. Males (5.57±0.47) had a higher mean LASMI than females (4.64±0.44, p<0.001) and females (25.19±4.54) had a higher mean GFFP than males (18.72±4.05, p<0.001), although mean BMI was not different between gender (p=0.166).

**Effect of BMI on Aβ uptakes**

Among males, the Aβ+ group had (22.85±2.41) lower BMI than Aβ− (24.56±2.57, p<0.001). Among females, the Aβ+ group also had (22.69±2.84) lower BMI than Aβ− (24.01±2.91, p=0.024). BMI was also associated with Aβ positivity in both males (OR=0.76, 95% CI 0.61 to 0.92) and females (OR=0.82, 95% CI 0.68 to 0.96).

Regarding the relationship of BMI with LASMI and GFFP, among males, BMI was positively correlated with LASMI (r=0.581, p<0.001) and GFFP (r=0.422, p<0.001). Among females, BMI was also correlated with LASMI (r=0.533, p<0.001) and GFFP (r=0.349, p=0.001).

**Effects of body composition on Aβ uptakes**

As illustrated in Fig. 1, among males, Aβ+ group (17.15±3.66) had lower GFFP than Aβ− (19.48±4.02, p=0.001), while mean LASMI was not different between Aβ+ and Aβ− group (p=0.111). Among females, Aβ+ group (4.72±0.42) had lower LASMI than Aβ− (4.50±0.44, p=0.011), while mean GFFP was not different between Aβ+ and Aβ− group (p=0.803). Among males, GFFP (OR=0.84, 95% CI 0.73 to 0.95) was independently associated with Aβ positivity, while LASMI (OR=0.49, 95% CI 0.16 to 1.40) was not associated with Aβ positivity (Table 2). In contrast, among females, a higher LASMI (OR=0.27, 95% CI 0.08 to 0.84) was independently associated with lesser Aβ positivity, while GFFP (OR=0.99, 95% CI 0.89 to 1.10) was not associated with Aβ positivity (Table 2). There was no interaction effect of gender with LASMI (gender*LASMI, p=0.875) and GFFP (gender*GFFP, p=0.092) on Aβ positivity in all participants (Table 2).

Figure 2 shows the correlation between LASMI and CL values. Among males, GFFP (r=−0.253, p=0.004) negatively correlated with CL values, while LASMI (r=−0.042, p=0.636) did not correlate with CL values. In contrast, among females, LASMI (r=−0.205, p=0.037) negatively correlated with CL values, while GFFP (r=0.022, p=0.827) did not correlate with CL values.

**Discussion**

In the present study, we systematically investigated the effects of muscle and fat mass in the thighs on brain Aβ deposition between males and females in a large cohort. We found that higher muscle mass, as reflected by the LASMI, was associated with a lesser risk of Aβ positivity in females. Higher fat mass, as reflected by the GFFP, was associated with a lesser risk of Aβ positivity in males. Thus, our findings suggest that there may be gender differences in the effects of body composition on Aβ deposition. Furthermore, our results may help in designing strategies for controlling body composition to prevent Aβ deposition.
Our first major finding was that higher muscle mass in the thighs was associated with a lesser risk of Aβ positivity in females. Previous studies have shown that sarcopenia is associated with incident probable AD dementia and cognitive impairment in older adults [35, 36]. However, information on the association between sarcopenia and AD biomarkers is lacking. Several possible mechanisms have been proposed to explain the relationship between muscle mass and brain Aβ positivity. Specifically, this relationship may be explained by the fact that Aβ-mediated neurodegeneration in the hypothalamus is involved in regulating dietary intake and energy expenditure [37], and systemic pro-inflammatory changes [38, 39]. However, we did not observe this relationship in males. To our knowledge, our findings are the first to report the gender-specific deleterious effects of sarcopenia on Aβ deposition. One study showed that sarcopenia only increased all-cause mortality in females [40]. It is important to identify the reason why females are more vulnerable to sarcopenia. Hormonal and socio-behavioral effects may contribute to gender differences. In particular, testosterone and estrogen may play crucial roles. Testosterone possesses anabolic properties, and estrogen modulates and prevents inflammation [41]. A decrease in testosterone level in elderly males and estrogen deficiency in elderly females triggers the discontinuance of anabolic reactions in males and increases inflammatory reactions in females. These characteristics are known to be distinct precipitating factors for sarcopenia in relation to gender [41]. In fact, the Framingham Heart Study found that higher levels of inflammatory proteins, such as interleukin, were associated with sarcopenia in males [42]. Hence, it was expected that sarcopenia in females would be closely associated with inflammation, which might lead to increased Aβ deposition.

Our second major finding was that higher fat mass in the thighs was associated with a lesser risk of Aβ positivity in males. The association of fat mass with Aβ might lead to increased Aβ deposition. A previous study showed that sarcopenia only increased all-cause mortality in females [40].

**Table 1** Demographic variables and body composition profiles of study participants

| Variables                  | Males                        | Females                      |
|----------------------------|------------------------------|------------------------------|
|                            | Total (n = 134) | Aβ (−) (n = 90) | Aβ (+) (n = 44) | Total (n = 106) | Aβ (−) (n = 65) | Aβ (+) (n = 41) |
| Cognitive stage             |                             |                              |                  |                |              |                |
| NC                         | 62 (46.3%)                  | 49 (54.4%)*                  | 13 (29.5%)*      | 61 (57.5%)     | 43 (66.2%)*    | 18 (43.9%)*    |
| MCI                        | 72 (53.7%)                  | 41 (45.6%)*                  | 31 (70.5%)*      | 45 (42.5%)     | 22 (33.8%)*    | 23 (56.1%)*    |
| Demographics               |                             |                              |                  |                |              |                |
| Age, years                 | 71.3±6.7                    | 70.5±7.2*                    | 72.8±5.3*        | 69.9±8.1       | 68.9±8.3      | 71.5±7.5       |
| Education, years           | 13.7±4.0                    | 14.2±3.5                     | 12.8±4.6         | 12.0±4.8       | 12.4±4.7      | 11.2±4.9       |
| APOE, e4 carrier           | 47 (35.1%)                  | 19 (21.1%)*                  | 28 (63.6%)*      | 33 (31.1%)     | 10 (15.4%)     | 23 (56.1%)*    |
| Hypertension               | 64 (47.8%)                  | 46 (51.1%)                   | 18 (40.9%)       | 49 (46.2%)     | 29 (44.6%)    | 20 (48.8%)     |
| Diabetes                   | 32 (23.9%)                  | 25 (25.6%)                   | 9 (20.5%)        | 17 (16.0%)     | 11 (16.9%)    | 6 (14.6%)      |
| Body composition           |                             |                              |                  |                |              |                |
| BMI, kg/m²                 | 24.0±2.6                    | 24.56±2.57*                  | 22.85±2.41*      | 23.5±2.9       | 24.01±2.91*   | 22.69±2.84*    |
| LASMI, kg/m²               | 5.57±0.47                   | 5.62±0.50                    | 5.48±0.39        | 4.64±0.44      | 4.72±0.42*    | 4.50±0.44*     |
| GFFP, %                    | 18.72±4.05                  | 19.48±4.02*                  | 17.15±3.66*      | 25.19±4.54     | 25.28±4.08    | 25.05±5.24     |
| Aβ deposition              |                             |                              |                  |                |              |                |
| Centiloid                  | 270±42.1                    | 1.3±6.7*                    | 79.6±34.2*       | 27.1±37.1      | 2.2±6.3*      | 64.4±30.9*     |
| Cognitive tests            |                             |                              |                  |                |              |                |
| SVLT recall                | 4.4±2.9                     | 5.2±2.9*                    | 2.9±2.3*         | 5.2±3.4       | 6.1±3.1*      | 3.5±3.2*       |
| RCFT recall                | 13.2±6.9                    | 15.0±7.0*                   | 9.7±5.1*         | 11.0±6.9      | 13.0±6.7*     | 7.8±6.1*       |
| K-BNT                      | 46.7±7.8                    | 47.7±7.3*                   | 44.7±8.5*        | 43.3±9.7      | 45.1±8.6*     | 40.3±10.8*     |
| RCFT copying               | 32.2±4.5                    | 32.9±3.1*                   | 30.7±6.2*        | 30.4±7.2      | 31.7±5.7*     | 28.3±8.8*      |
| DSB                        | 3.9±1.0                     | 3.9±1.0                     | 3.9±1.0          | 3.9±1.4       | 4.1±1.5       | 3.7±1.3        |
| Stroop CR                  | 73.0±24.9                   | 76.1±23.0*                  | 66.8±27.6*       | 80.5±28.5     | 88.0±25.7*    | 68.0±28.7*     |

Values are presented as mean ± standard deviation

Abbreviations: Aβ (−) amyloid negative, Aβ (+) amyloid positive, n number of patients whose data were available for analysis, BMI body mass index, CR color reading, DSB Digit Span Test backward, LASMI lower extremity appendicular skeletal muscle mass index, GFFP gluteofemoral fat percentage, K-BNT Korean version of the Boston Naming Test, RCFT Rey-Osterrieth Complex Figure Test, SVLT Seoul Verbal Learning Test delayed recall

* Significant difference at p<0.05 between Aβ (−) and Aβ (+) in the same gender
suggested that males have more abdominal fat than females, whereas females have more fat mass in their thighs than males [14]. Unlike abdominal fat, fat mass in the thighs is known to have beneficial effects on metabolic health, which is closely related to a higher level of adiponectin known as a fat tissue-specific hormone [15]. In fact, it has been reported that there are lower levels of adiponectin in males than in age-and BMI-matched females [43]. Growing evidence indicates that adiponectin possesses anti-inflammatory and insulin-sensitizing properties [15] and it has a protective role in the development of neurodegenerative diseases [44, 45]. In particular, a recent study revealed that higher levels of adiponectin were associated with higher A\(\beta\) levels in the cerebrospinal fluid, which suggested possible neuroprotective effects of adiponectin against A\(\beta\) positivity [46]. Thus, it was expected that a reduction in adiponectin levels might be a possible explanation for male-specific vulnerability following a reduction of fat mass in the thighs.

**Limitations**
The strength of the present study is that we systematically investigated the effects of body composition on A\(\beta\) positivity with measurements of muscle and fat mass in the thighs in a large-sized cohort. However, our study has several limitations that should be addressed. First, since this
was a cross-sectional study, it was difficult to determine the temporal relationships between the changes in body composition and Aβ deposition. Second, we were not able to assess repeated measurements over time, and then we did not cover the change or variability of muscle and fat mass. Third, we lacked more precise data of thigh muscle and fat mass, because we used LASMI and GFFP as proxy measures for muscle and fat mass in the thighs. However, existing findings show that GFFP highly correlates with actual values of thigh fat mass measured using DXA scans of the whole body [47], and LAMSI correlates with thigh muscle volume measured using MRI scans of the thigh [34]. Fourth, we used muscle and fat mass measured within 3 years before or after the Aβ PET. However, this limitation was alleviated to some extent by previous findings that the annual increase in Aβ is very low [48]. Finally, our participants were recruited from a population undergoing a comprehensive preventive health exam that was not covered by national medical insurance. The cohort was also restricted to participants undergoing DXA testing, which might have differed from the cohort of patients who did not undergo such testing. This study may have thus resulted in the enrollment of a healthier or more “health-seeking” population, which may also limit the generalizability of this study to other populations.

Conclusions
In the present study, we highlight the importance of recognizing that gender differences may exist with respect to how specific body compositions may potentially protect against Aβ deposition. Furthermore, our findings suggest that the detection of changes in thigh muscle and fat mass in relation to gender are needed for early diagnosis and prevention of AD.

Supplementary Information
The online version contains supplementary material available at https://doi.org/10.1186/s13195-022-01086-5.

Additional file 1: Table S1. Demographic variables of study participants stratified by cognitive stage.

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Authors' contributions
S.H. Kang, S.W. Seo, and M. Kang contributed to the study conception and design. S.H. Kang, Y. Chang, S.W. Seo, and M. Kang contributed data collection. S.H. Kang performed the statistical analysis and wrote the first draft of the manuscript. S.H. Kang, Y. Chang, S.W. Seo, and M. Kang contributed interpretation of data and manuscript revision. All authors read and approved the final manuscript.

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Availability of data and materials
Anonymized data for our analyses presented in this report are available upon request from the corresponding authors.
Declarations

Ethics approval and consent to participate
Approval was obtained from the ethics committee of Samsung Medical Centre. The procedures used in this study adhere to the tenets of the Declaration of Helsinki. Written informed consent was obtained from all patients and caregivers.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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References

1. Rowe CC, Bourget P, Ellis KA, Brown B, Lim YY, Mulligan R, et al. Predicting Alzheimer disease with beta-amyloid imaging: results from the Australian imaging, biomarkers, and lifestyle study of ageing. Ann Neurol. 2013;74:905–13.
2. Villedame VG, Pike KE, Chetelat G, Ellis KA, Mulligan RS, Bourget P, et al. Longitudinal assessment of Abeta and cognition in aging and Alzheimer disease. Ann Neurol. 2011;69:181–92.
3. Ye BS, Kim HJ, Kim YJ, Jung NY, Lee JS, Lee J, et al. Longitudinal outcomes of amyloid positive versus negative amnestic mild cognitive impairments: a three-year longitudinal study. Sci Rep. 2018;8:5557.
4. Tolpanen AM, Ngandu T, Kärhölt I, Laatikainen T, Rusanen M, Soninen H, et al. Midlife and late-life body mass index and late-life dementia: results from a prospective population-based cohort. J Alzheimers Dis. 2014;38:201–9.
5. Bell SP, Liu D, Samuels LR, Shah AS, Gifford KA, Hohman TJ, et al. Late-Life Body Mass Index, Rapid Weight Loss, Apolipoprotein E ε4 and the Risk of Cognitive Decline and Incident Dementia. J Nutr Health Aging. 2017;21:1259–67.
6. Johnson DK, Wilkins CH, Morris JC. Accelerated weight loss may precede diagnosis in Alzheimer disease. Arch Neurol. 2006;63:1312–7.
7. Vidoni ED, Townley RA, Honea RA, Burns JM. Alzheimer disease biomarkers are associated with body mass index. Neurology. 2011;77:1913–20.
8. Kang SH, Kim JH, Chang Y, Cheon BK, Choe YS, Jang H, et al. Independent effect of body mass index variation on amyloid-β positivity. Front Aging Neurosci. 2022;14:924550.
9. Kim YH, So WM. Relative lower body circumferences are associated with the prevalence of metabolic syndrome and arterial stiffness. Technol Health Care. 2017;25:211–9.
10. Chen CL, Liu L, Huang JY, Yu YL, Shen G, Lo K, et al. Thigh Circumference and Risk of All-Cause, Cardiovascular and Cerebrovascular Mortality: A Cohort Study. Risk Manag Healthc Policy. 2020;13:1977–87.
11. Patel P, Abate N. Body fat distribution and insulin resistance. Nutrients. 2013;5:2019–27.
12. Abe T, Kawai K, Kondo M, Fukunaga T. Comparison of ultrasound-measured age-related, site-specific muscle loss between healthy Japanese and German men. Clin Physiol Funct Imaging. 2011;31:320–5.
13. Yim JE, Heshka S, Albu JB, Heymsfield S, Gallagher D. Femoral-gluteal subcutaneous and intermuscular adipose tissues have independent and opposing relationships with CVD risk. J Appl Physiol. 1985;60:700–7.
14. Schorr M, Dichtel LE, Gerweck AV, Valera RD, Torriani M, Miller KK, et al. Sex differences in body composition and association with cardiometabolic risk. Biol Sex Differ. 2018;9:28.
15. Turer AT, Khera A, Ayers CR, Turer CB, Grundy SM, Vega GL, et al. Adipose tissue mass and location affect circulating adiponectin levels. Diabetologia. 2011;54:2515–24.
16. Vega GL, Adams-Huet B, Peshock R, Willett D, Shah B, Grundy SM. Influence of body fat content and distribution on variation in metabolic risk. J Clin Endocrinol Metab. 2006;91:4459–66.
17. Grundy SM, Adams-Huet B, Vega GL. Variable contributions of fat content and distribution to metabolic syndrome risk factors. Metab Syndr Relat Disord. 2008;6:281–8.
18. Khera A, Vega GL, Das SR, Ayers C, McGuire DK, Grundy SM, et al. Sex differences in the relationship between C-reactive protein and body fat. J Clin Endocrinol Metab. 2009;94:3251–8.
19. Okura T, Nakata Y, Yamabuki K, Tanaka K. Regional body composition changes exhibit opposing effects on coronary heart disease risk factors. Atherosclerosis. 2004;179:923–9.
20. Snijder MB, Zimmet PZ, Wser M, Dekker JM, Seidell JC, Shaw JE. Independent and opposite associations of waist and hip circumferences with diabetes, hypertension and dyslipidemia: the AusDiab Study. Int J Obes Relat Metab Disord. 2004;28:402–9.
21. Morley JE, Baumgartner RN, Roubenoff R, Mayer J, Naar KS. Sarcopenia. J Lab Clin Med. 2001;137:231–3.
22. Tyrovolas S, Panagiotakos D, Georgousepolou E, Chrysouhou C, Tousoulis D, Haro JM, et al. Skeletal muscle mass in relation to 10 year cardiovascular disease incidence among middle aged and older adults: the ATTICA study. J Epidemiol Community Health. 2020;74:26–31.
23. Celis-Morales CA, Walsh P, Lyall DM, Steel L, Petermann F, Anderson J, et al. Associations of grip strength with cardiovascular, respiratory, and cancer outcomes and all cause mortality: prospective cohort study of half a million UK Biobank participants. BMJ. 2018;361:k1651.
24. Abellan van Kan G. Epidemiology and consequences of sarcopenia. J Nutr Health Aging. 2009;13:708–12.
25. Petersen SJ, Braunschweig CA. Prevalence of sarcopenia and associated outcomes in the Clinical Setting. Nutr Clin Pract. 2016;31:40–8.
26. Kang SH, Park YH, Lee D, Kim JP, Chin J, Ahn Y, et al. The Cortical Neuroanatomy Related to Specific Neuropsychological Deficits in Alzheimer’s Continuum. Dement Neurocogn Disord. 2019;18:77–95.
27. Kang Y, Jahng S, Na DL. Seoul Neuropsychological Screening Battery. 2nd ed. Seoul: Human Brain Research & Consulting Co.; 2012.
28. Christensen KJ, Multhaupt KS, Nordstrom S, Voss K. A cognitive battery for dementia: Development and measurement characteristics. Psychol Assess J Consul Clin Psychol. 1991;3:168–74.
29. Petersen RC. Clinical practice. Mild cognitive impairment. N Engl J Med. 2011;364:2227–34.
30. Jeong HJ, Lee H, Lee S-Y, Seo S, Park KH, Lee YB, et al. [18F]THK5351 PET Imaging in Patients with Mild Cognitive Impairment. J Nucl Med. 2020;61:202–14.
31. Cho SH, Choe YS, Kim HJ, Jang H, Kim Y, Kim SE, et al. A new Centiloid method for [18F]-flortetaben and [18F]-flutemetamol PET without conversion to PiB. Eur J Nucl Med Mol Imaging. 2020;47:1938–48.
32. Villeneuve S, Rabinovici GD, Cohn-Sheehy BI, Madison C, Ayakta N, Ghosh PM, et al. Existing Pittsburgh Compound-B postmortem emission tomography thresholds are too high: statistical and pathological evaluation. Brain. 2015;138:2030–33.
33. Kang JH, Choi SH, Lim S, Kim KW, Lim JY, Cho NH, et al. Assessment of appendicular skeletal muscle mass by bioimpedance in older community-dwelling Korean adults. Arch Gerontol Geriatr. 2014;58:303–7.
34. Hoshino O, Yoshikawa N, Shirumizu N, Kiryu S, Uehara M, Kobayashi H, et al. Quantitative analysis of skeletal muscle mass in patients with rheumatic diseases under glucocorticoid therapy—comparison among biocellulose impedance analysis, computed tomography, and magnetic resonance imaging. Mod Rheumatol. 2015;25:257–63.
35. Burns JM, Johnson DK, Watts A, Swerdlow RH, Brooks WM. Reduced lean mass in early Alzheimer disease and its association with brain atrophy. Arch Neurol. 2010;67:428–33.

36. Boyle PA, Buchman AS, Wilson RS, Leurgans SE, Bennett DA. Association of muscle strength with the risk of Alzheimer disease and the rate of cognitive decline in community-dwelling older persons. Arch Neurol. 2009;66:1339–44.

37. Loskutova N, Honea RA, Brooks WM, Burns JM. Reduced limbic and hypothalamic volumes correlate with bone density in early Alzheimer’s disease. J Alzheimers Dis. 2010;20:313–22.

38. Yaffe K, Kanaya A, Lindquist K, Simonsick EM, Harris T, Shorr RI, et al. The metabolic syndrome, inflammation, and risk of cognitive decline. JAMA. 2004;292:2237–42.

39. Krabbe KS, Pedersen M, Bruunsgaard H. Inflammatory mediators in the elderly. Exp Gerontol. 2004;39:687–99.

40. Batsis JA, Mackenzie TA, Barre LK, Lopez-Jimenez F, Bartels SJ. Sarcopenia, sarcopenic obesity and mortality in older adults: results from the National Health and Nutrition Examination Survey III. Eur J Clin Nutr. 2014;68:1001–7.

41. Anderson LJ, Liu H, Garcia JM. Sex Differences in Muscle Wasting. Adv Exp Med Biol. 2017;1043:153–97.

42. Payette H, Roubenoff R, Jacques PF, Dinarello CA, Wilson PW, Abad LW, et al. Insulin-like growth factor-1 and interleukin 6 predict sarcopenia in very old community-living men and women: the Framingham Heart Study. J Am Geriatr Soc. 2003;51:1237–43.

43. Hausman DB, DiGirolamo M, Bartness TJ, Hausman GJ, Martin RI. The biology of white adipocyte proliferation. Obes Rev. 2001;2:239–54.

44. Kamogawa K, Kohara K, Tabara Y, Uetani E, Nagai T, Yamamoto M, et al. Abdominal fat, adipose-derived hormones and mild cognitive impairment: the J-SHIPP study. Dement Geriatr Cogn Disord. 2010;30:432–9.

45. Gorska-Ciebiada M, Saryusz-Wolska M, Borkowska A, Ciebiada M, Loba J. Adiponectin, leptin and IL-1 beta in elderly diabetic patients with mild cognitive impairment. Metab Brain Dis. 2016;31:257–66.

46. Waragai M, Adame A, Trinh I, Sekiyama K, Takamatsu Y, Ume K, et al. Possible Involvement of Adiponectin, the Anti-Diabetes Molecule, in the Pathogenesis of Alzheimer’s Disease. J Alzheimers Dis. 2016;52:1453–9.

47. Leslie WD. Prediction of body composition from spine and hip bone densitometry. J Clin Densitom. 2009;12:428–33.

48. Kemppainen NM, Schenin NM, Koivunen J, Johansson J, Toivonen JT, Någren K, et al. Five-year follow-up of 11C-PiB uptake in Alzheimer’s disease and MCI. Eur J Nucl Med Mol Imaging. 2014;41:283–9.

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