Research Article

Coordination Dynamics and Coordination Mechanism of a New Type of Anticoagulant Diethyl Citrate with Ca\(^{2+}\) Ions

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1. Introduction

An anticoagulant must be added to dialysates to prevent blood solidification \textit{in vitro} (in a dialysis machine). Sodium citrate (Na\(_3\)Cit) is an important anticoagulant used in clinical settings [1–3]. However, using Na\(_3\)Cit as an anticoagulant easily causes hypocalcemia and hypercalcemia [4, 5] because of the strong chelating ability of Na\(_3\)Cit with Ca\(^{2+}\) ions. Given this ability, the dissociation metabolism of the formed chelate CaCit in \textit{vivo} takes 30 min. Using Na\(_3\)Cit also negatively affects the maintenance of coagulation stability of high-risk hemorrhage patients \textit{in vivo}, which easily causes complications such as hypocalcemia during or after dialysis.

Our group has previously synthesized a new anticoagulant [6], namely, diethyl citrate (Et\(_2\)Cit). The anticoagulant mechanism of Et\(_2\)Cit is based on the formation of Ca\(^{2+}\) with Et\(_2\)Cit. This formation decreases the Ca\(^{2+}\) concentration in blood and inhibits prothrombin conversion into thrombin, thereby influencing the anticoagulant effect. The large steric effect of Et\(_2\)Cit weakens the coordination of Ca\(^{2+}\) ion compared with that of Na\(_3\)Cit. Therefore, hypocalcemia and hypercalcemia can be avoided using Et\(_2\)Cit as anticoagulant [7]. The frequency of blood gas analyses can also be lessened by repeatedly taking the venous blood of patients to monitor serum calcium levels, which can help relieve the pain of patients and the workload of nurses.

The stability of the complex of Et\(_2\)Cit with Ca\(^{2+}\) (CaEt\(_3\)Cit) is reportedly weaker than that of CaCit [8]. At pH 7.4 and 37°C, the stability constants (\(K'_s\)) are 1988 for CaCit and 231 for CaEt\(_3\)Cit. However, several problems remain unsolved when Et\(_2\)Cit is used as an anticoagulant. These problems include the reaction kinetics of Et\(_2\)Cit with Ca\(^{2+}\) and coordination reaction mechanisms, as well as the composition and characterization of the complex. Accordingly, the coordination dynamics of Et\(_2\)Cit and Na\(_3\)Cit with Ca\(^{2+}\), as well as the influencing factors, were studied. The underlying coordination principle was also proposed.
2. Materials and Methods

2.1. Instruments and Reagents. The instruments used were as follows: CHN-O– rapid type element analyzer (Foss-Heraeus Company), Bruker AM 500 nuclear magnetic resonance (NMR) spectrometer (with CDCl₃ as solvent and TMS as internal standard), Nicolet-170 SX type FT-IR spectrometer, D/max 2400 (Rigaku) X-ray diffractometer, inductively coupled plasma emission spectrometry (ICP) system (PE Company, USA), PHS-3C pH meter (Shanghai Precision & Scientific Instrument Co., Ltd., China), and sodium chloride injection system (Wuhan Binhu Double-Crane Pharmaceutical Co., Ltd., China).

All chemical reagents used were of analytical grade. Et₂Cit was prepared in our laboratory (99.3% purity) [6].

2.2. Experimental Methods

2.2.1. Reaction Rate Constants of Et₂Cit and Na₃Cit with Ca²⁺. CaCl₂ and Et₂Cit solutions (2.0 mmol/L) were prepared and mixed. A calcium-ion-selective electrode was used to determine the change in electrode potential of the mixed solution with reaction time at pH 7.4 and 37 °C under stirring. The result was then compared with that of Na₃Cit.

The linear regression equation of the calcium ion-selective electrode was \( y = 29x + 69 \) (where \( y \) is the electrode potential and \( x \) is \( -p(Ca^{2+}) \)). The concentration of \( Ca^{2+} [c(Ca^{2+})] \) at \( t \) time was also calculated. Given that CaCl₂ was mixed with Na₃Cit or Et₂Cit (1:1) and that the reaction of \( Ca^{2+} \) with Na₃Cit or Et₂Cit was equal in solution [7], the following reaction rate equation can be established using \( r \) to represent the reaction rate:

\[
 r = k c^n, \tag{1}
\]

where \( k \) is the reaction rate constant and \( n \) is the reaction order. Assuming that \( x \) is the amount of \( Ca^{2+} \) substance concentration that disappeared at \( t \) time, that is, \( x = a - c \) (\( Ca^{2+} \)), the following can be obtained by arranging formula (1):

\[
 r = -\frac{dc}{dt} = -\frac{d(a-x)}{dt} = \frac{dx}{dt} = k(a-x)^n. \tag{2}
\]

After logarithm on both sides we get

\[
 \log r = \log \left( -\frac{d(a-x)}{dt} \right) = \log k + n \log(a-x) = \log k + n \log c. \tag{3}
\]

From the plot of \( x \) versus \( t \), we can calculate the tangent slope of the curve \( dx/dt \), which is the reaction rate of various points. Formula (3) shows a linear relationship between \( \log r \) and \( \log c(Ca^{2+}) \). In the diagram of \( \log r \) on \( \log c(Ca^{2+}) \), the slope of the straight line is the reaction order \( n \), whereas the intercept is \( \log k \).

2.2.2. Effect of pH on Reaction Rate. The pH of the system was adjusted to 6.0, 7.4, and 8.0. Then, the effect of pH on \( k \) and \( n \) was determined.

2.2.3. Synthesis of Diethyl Citrate Calcium Complex Crystal. About 1.665 g (15 mmol) of anhydrous CaCl₂ was completely dissolved in water. Then, 1.241 g (5 mmol) of Et₂Cit was slowly trickled under stirring. The pH was adjusted to 7.0 after obtaining a colorless and transparent solution. The solution was sealed with a plastic wrap having holes and then placed in an oven at 37 °C for slow volatilization and crystallization. The precipitated colorless, needle-like crystals were filtered, washed with anhydrous ethanol, dried, and characterized. The methods of characterization included elemental analysis, X-ray powder diffraction (XRD), Fourier-transform infrared spectroscopy (FT-IR), ¹H NMR, and ICP.

3. Results and Discussion

3.1. Reaction Rate Equation of Et₂Cit and Na₃Cit with Ca²⁺. The change in concentration of free Ca²⁺ ion \([c(Ca^{2+})]\) with \( t \) in reaction system of Et₂Cit and Na₃Cit with CaCl₂ is shown in Figure 1. A rapid decrease in \( c(Ca^{2+}) \) was observed with prolonged \( t \) from 0 s to 30 s. This finding indicated that Et₂Cit or Na₃Cit was rapidly coordinated with \( Ca^{2+} \). At \( t = 30 \) s, \( c(Ca^{2+}) \) decreased from 1.0 mmol/L to 0.49 mmol/L in the Na₃Cit system and from 1.0 mmol/L to 0.87 mmol/L in the Et₂Cit system. \( c(Ca^{2+}) \) slowly decreased when \( t > 120 \) s, indicating that the system was in a dynamic equilibrium of complexation dissociation.

The tangent slope \( (dx/dt) \) of points on the curve, that is, the reaction rate \( r \) of each point formula (2), can be obtained according to Figure 1. In the diagram of \( \log r \) versus \( \log c(Ca^{2+}) \) (Figure 2), the slope of the line was the reaction order \( n \) (formula (3)). The intercept of the line was \( \log k \) in Figure 2, as shown in the following:

\[
\text{Et}_2\text{Cit-Ca}_3: \ n = 2.46; \ \log k = 2.06, \ \text{so} \ k = 120; \\
\text{Na}_3\text{Cit-Ca}_3: \ n = 2.44; \ \log k = 2.46, \ \text{so} \ k = 289. \tag{4}
\]
Figure 2: Plots of log $r$ vs. log $c$(Ca\(^{2+}\)) in different systems: (a) Et\(_2\)Cit-Ca\(^{3+}\) system and (b) Na\(_2\)Cit-Ca\(^{3+}\) system.

The reaction rate equations of Et\(_2\)Cit and Na\(_2\)Cit with Ca\(^{2+}\) were as follows:

Et\(_2\)Cit-Ca system: $r = k_a 2.46 = 120 a^{2.46}$,

Na\(_2\)Cit-Ca system: $r = k_a 2.44 = 289 a^{2.44}$.

(5)

Given that $k$ can directly reflect the reaction rate, (5) shown that the complexation rate of Na\(_2\)Cit with Ca\(^{2+}\) was faster than that of Et\(_2\)Cit.

The anticoagulant mechanism of Na\(_2\)Cit and Et\(_2\)Cit was based on the combination of calcium ion (Ca\(^{2+}\)) in serum, as well as the reduced concentration of free Ca\(^{2+}\) in plasma that disturbed the blood clotting process from reaching the anticoagulation effect in vitro [9–11]. However, the strong coordination ability of Na\(_2\)Cit, particularly as an anticoagulant, can coordinate a large number of Ca\(^{2+}\) ions in the blood. This phenomenon can lead to the low serum concentration of calcium in patients, as well as to hypocalcemia and all kinds of complications [12–15]. Therefore, calcium is needed to be replenished in the anticoagulation process of Na\(_2\)Cit [16]. Meanwhile, calcium citrate (CaCit) can dissociate during the metabolism and release Ca\(^{2+}\) after entering the body in the dialysis process. Additionally, hypercalcemia easily ensued in patients with presupplementary Ca\(^{2+}\). Therefore, the incidence of hypocalcemia and hypercalcemia can be reduced if we can reduce the coordination ability of anticoagulant.

The reaction rate was equal to the inverse reaction rate when the reaction reached equilibrium, as shown in the following:

$$k(a − x)^n = k_{re} x^n.$$

(6)

The above equation can be written as follows [17]:

$$\frac{x^n}{(a − x)^n} = \frac{k}{k_{re}} = K_n,$$

(7)

where $k$ is the reaction rate constant, $k_{re}$ is the inverse reaction rate constant, and $K_n$ is the complex stability constant.

In a previous article [8], the $K_n$ values of CaEt\(_2\)Cit and CaCit were 231 and 1988 at pH 7.4 and 37°C, respectively, and the $k$ values in the coordination reaction of Et\(_2\)Cit and Na\(_2\)Cit with Ca\(^{2+}\) were 120 and 289 L mol\(^{-1}\) s\(^{-1}\), respectively. According to (7), $k_{re}$ of Et\(_2\)Cit and Na\(_2\)Cit with Ca\(^{2+}\) in the coordination reaction were 0.52 and 0.15 L mol\(^{-1}\) s\(^{-1}\), respectively. Thus, the rate of decomposition and release of Ca\(^{2+}\) was faster for CaEt\(_2\)Cit than for CaCit. The above results indicated that Et\(_2\)Cit can complex with Ca\(^{2+}\) and reduce the free Ca\(^{2+}\) concentration during anticoagulation; thus, anticoagulation can be achieved. Meanwhile, the complexing ability of Et\(_2\)Cit with Ca\(^{2+}\) was weaker than that of Na\(_2\)Cit. After Et\(_2\)Cit coordinated with Ca\(^{3+}\), the Ca\(^{2+}\) releasing rate of CaEt\(_2\)Cit was faster than that of CaCit. Therefore, the occurrence of hypocalcemia in patients can be avoided. Moreover, only a small amount of calcium or none at all was needed using Et\(_2\)Cit as anticoagulant during dialysis unlike using Na\(_2\)Cit. Thus, the occurrence of hypercalcemia can be avoided using Et\(_2\)Cit as an anticoagulant.

3.2. Effect of pH on Reaction Rate. At present, the main dialy- sates in clinical practice are bicarbonate and acetic acid liquid. The pH of acetate dialysate is generally controlled to remain at 6.0 to 7.2 [18]. In [19], the pH range of the dialysate is 5.3–8.2. At the entrance of the dialysis machine, the pH of a patient’s whole blood was between 7.15 and 7.4, whereas the pH of the exports of the dialysis machine was between 6.2 and 7.4.

In the dialysis process, the pH values of different dialysis sates varied. The acidities of different anticoagulants also dif- fered. Therefore, the pH of blood in the dialysis process also changed. Considering that Na\(_2\)Cit was a strong base-weak acid salt, 1 mol of Na\(_2\)Cit contained 3 mol of carboxylate (COO\(^-\)), wherein Na\(_2\)Cit was alkaline. Therefore, when Na\(_2\)Cit was used as an anticoagulant, the blood pH decreased and metabolic alkalosis likely ensued.

Considering that one Et\(_2\)Cit molecule only had one –COO\(^-\), the possibility of causing alkalis was signifi- cantly reduced when Et\(_2\)Cit was used as anticoagulant. With increased pH from 6.0 to 8.0, free c(Ca\(^{2+}\)) decreased faster in the system (Figure 3) because increased pH benefited the ionization of –OH and –COOH of Et\(_2\)Cit or Na\(_2\)Cit, which in turn benefited the coordination with Ca\(^{2+}\).

Table 1 shows the reaction rate constants $k$ of Et\(_2\)Cit and Na\(_2\)Cit with CaCl\(_2\), as well as the complex dissociation rate $k_{re}$ when the pH values of the system were 6.0, 7.4, and 8.0.

The reaction rate and dissociation rate of the complex were found to accelerate with increased pH. The reaction rate of Et\(_2\)Cit and Na\(_2\)Cit with CaCl\(_2\) was influenced by pH because H\(^+\) inhibits the ionization of the active H of –COOH in Et\(_2\)Cit molecule, as well as changing the course of coordination reaction. Thus, the reaction rate constant and reaction order changed.

Within pH 6.0–8.0, the pH increase accelerated the dissociation rate of the complex. With increased pH from 6.0 to 8.0, $k_{re}$ of the Et\(_2\)Cit-CaCl\(_2\) system increased from 0.04 to 19.8, whereas $k_{re}$ of the Na\(_2\)Cit-CaCl\(_2\) system increased from 0.03 to 6.79. The dissociation rate of the complex coordination of Et\(_2\)Cit and Na\(_2\)Cit with calcium under an alkaline
3.0
3.1
3.2
3.3
3.4
0 50 100 150 200 250 300
Time (s)
\[ \log(c(\text{Ca}^{2+})) \]

**Figure 3:** Plots of concentration change of free Ca\(^{2+}\) ions with reaction time under different pH conditions: (a) Et\(_2\)Cit and (b) Na\(_3\)Cit.

| pH   | 6.0 | 7.4 | 8.0 |
|------|-----|-----|-----|
| Et\(_2\)Cit-CaCl\(_2\) system |     |     |     |
| reaction order (n)       | 2.03| 2.46| 2.73|
| stability constants (K\(_s\)) | \(10^{3.93}\) | \(10^{2.06}\) | \(10^{3.06}\) |
| rate constant (k)/L-mol\(^{-1}\)-s\(^{-1}\) | 9   | 120 | 4571 |
| reverse reaction constant (k\(_r\))/L-mol\(^{-1}\)-s\(^{-1}\) | 0.04| 0.52| 19.80 |
| Na\(_3\)Cit-CaCl\(_2\) system |     |     |     |
| reaction order (n)       | 2.16| 2.44| 3.0 |
| \(K_j\) | \(10^{1.98}\) | \(10^{2.46}\) | \(10^{1.83}\) |
| \(k/L\)-mol\(^{-1}\)-s\(^{-1}\) | 60  | 289 | 13489 |
| \(k_r/L\)-mol\(^{-1}\)-s\(^{-1}\) | 0.03| 0.15| 6.79 |

**Table 1:** Reaction rate constant and reaction order of Et\(_2\)Cit and Na\(_3\)Cit with Ca\(^{2+}\) ions.

3.3. Research on Et\(_2\)Cit and Ca Complexes

3.3.1. Elemental Analysis and Ca Content as Determined by ICP. To further study the coordination of Et\(_2\)Cit with Ca\(^{2+}\), the complex of Et\(_2\)Cit with Ca\(^{2+}\) was synthesized. Its composition was analyzed using elemental analysis and ICP, and the results are shown in Table 2. Et\(_2\)Cit was found to form the complex of CaEt\(_2\)Cit with Ca\(^{2+}\) in 1:1 coordination ratio.

**Table 2:** Elemental analysis data and Ca content measured by the ICP of complex CaEt\(_2\)Cit.

|     | C% | H% | Ca% |
|-----|----|----|-----|
| EA results | 41.55 (41.64)* | 5.73 (5.55) | — |
| ICP result | — | — | 13.68 (13.93) |

*The value in bracket was theoretical value, which is calculated according to the formula of complex CaEt\(_2\)Cit.

Therefore, the experimental value was consistent with the theoretical value.

3.3.2. XRD Analysis. Figure 4 is the XRD pattern of CaCl\(_2\) and CaEt\(_2\)Cit crystals. The diffraction peaks of CaCl\(_2\) appeared at \(d = 5.97, 2.78, 3.03, 4.28,\) and \(2.90\) Å (Figure 4(a)),

condition was faster than that under an acidic condition. Therefore, the pH increase of anticoagulants such as Et\(_2\)Cit and Na\(_3\)Cit and dialysis under alkaline conditions achieved the purpose of anticoagulation and avoided the occurrence of dialysis acidosis, thereby improving the survival rate and quality of life.

**Figure 3:** Plots of concentration change of free Ca\(^{2+}\) ions with reaction time under different pH conditions: (a) Et\(_2\)Cit and (b) Na\(_3\)Cit.

Therefore, the experimental value was consistent with the theoretical value.
Table 3: Wavenumber of the main absorption peaks of FT-IR spectra of Et$_2$Cit and its complex CaEt$_2$Cit.

|          | Et$_2$Cit/cm$^{-1}$ | CaEt$_2$Cit/cm$^{-1}$ |
|----------|---------------------|------------------------|
|          | 3480 (OH)$^*$        | 3430 (OH)              |
| $\nu$(OH)| 2986 (CH$_2$, CH$_3$)| 2982 (CH$_2$, –CH$_3$) |
|          | 1732 (O–C=O)         | 1709, 1624 (O–C=O)     |
|          | 1100 (C–O–C)         | 1081, 1041 (C–O–C)     |

Table 4: Absorption peak section and its assignment of the $^1$H NMR spectra of Et$_2$Cit and CaEt$_2$Cit.

|          | Et$_2$Cit/ppm | CaEt$_2$Cit/ppm |
|----------|---------------|-----------------|
| $\delta$ | 1.24–1.31 (–CH$_3$) | 1.22–1.28 (–CH$_3$) |
|          | 2.80–2.97 (–CH$_2$CO) | 2.77–2.98 (–CH$_2$CO) |
|          | 4.13–4.30 (–OCH$_2$) | 4.12–4.17 (–OCH$_2$) |
|          | 7.26, 6.28 (–OH) |                  |

Figure 4: XRD patterns of CaEt$_2$Cit and CaCl$_2$: (a) CaEt$_2$Cit and (b) CaCl$_2$.

Figure 5: FT-IR spectra of Et$_2$Cit and its complex CaEt$_2$Cit: (a) Et$_2$Cit and (b) CaEt$_2$Cit.

whereas the diffraction peaks of the complex appeared at $d = 6.99$ and 3.02 Å.

3.3.3. FT-IR Analysis. The FT-IR spectra of Et$_2$Cit and CaEt$_2$Cit complex are shown in Figure 5. The wavenumbers of the main absorption peaks are shown in Table 3 [20].

(1) The peak at 3430 cm$^{-1}$ was due to the stretching vibration of the hydroxyl group in the CaEt$_2$Cit complex, which red shifted by approximately 50 cm$^{-1}$ more than that of Et$_2$Cit (3480 cm$^{-1}$), indicating a hydrogen bond.

(2) The carbonyl absorption peak (C=O) of CaEt$_2$Cit split into two peaks, which were 1709 and 1624 cm$^{-1}$, respectively, indicating two different coordination environments in carbonyl. The position of both peaks red-shifted by approximately more than 30 and 110 cm$^{-1}$ compared with the carbonyl absorption peaks of Et$_2$Cit at 1736 cm$^{-1}$. This finding indicated that the carbonyl of Et$_2$Cit was coordinated with the calcium ions and was consistent with the change in the carbonyl characteristic absorption peak before and after coordination, as reported in [20].

(3) The absorption peak of the symmetric stretching vibrations of (C–O–C) in C–O–C of Et$_2$Cit was at 1100 cm$^{-1}$. However, the peak split into two in the complex, that is, at 1081 and 1041 cm$^{-1}$, respectively. This phenomenon was ascribed to one of the three C–O–C groups of the Et$_2$Cit molecular complex with Ca$^{2+}$, in which C–O–C absorption was bimodal and red shifted.

(4) The peak at 2982 cm$^{-1}$ was the absorption peak of the methyl hydrocarbon of CaEt$_2$Cit. It did not significantly change compared with the absorption peak of Et$_2$Cit methyl hydrocarbon (2986 cm$^{-1}$).

3.3.4. $^1$H NMR. The $^1$H NMR spectra of Et$_2$Cit and CaEt$_2$Cit were studied using CDCl$_3$ as a solvent, and the results are shown in Figure 6. The absorption peaks of $^1$H NMR are shown in Table 4.

(1) The proton peaks of the ligand at $\delta = 7.26$ and 6.28 ppm disappeared, indicating that –COOH participated in the coordination reaction. Meanwhile, the hydrogen in –OH group is very active; it can be easily dissociated and be partially or entirely substituted by deuterium in CDCl$_3$ solution.

(2) At 2.70 ppm to 3.0 ppm, the two groups of Et$_2$Cit quartets were –CH$_2$C=O (Figure 6(b)). –CH$_3$C=O groups occurred in different chemical environments, that is, 1,3-Et$_2$Cit and 1,5-Et$_2$Cit. The physical and chemical properties of the two isomers were very similar, so the two peaks did not significantly differ. After CaEt$_2$Cit was generated, the chemical environment of Et$_2$Cit changed and resulted in obvious dispersion.
and specificity of the two peaks of 2.70 ppm from 3.0 ppm. This result indicated that after 1,3-Et₂Cit and 1,5-Et₂Cit coordinated with calcium ions, the property difference of the two formed complexes increased compared with those of the original two ligands.

3) At 2.70 ppm to 3.0 ppm, the H peaks of –CH₂– shifted from 2.85 ppm to 2.91 ppm, and then to 2.81 ppm to 2.94 ppm after Et₂Cit coordinated with calcium. This finding was due to the O in –OCH₂ that coordinated with Ca, consistent with the IR spectra.

4) The peak at δ = 4.0 ppm was assigned to –OCH₂ of –COOCH₂CH₃ (Figure 6(c)). Compared with Et₂Cit (δ = 4.13 ppm to 4.20 ppm), this peak of the complex (δ = 4.12 ppm to 4.17 ppm) shifted to a high field. This finding indicated the weakening of the induction effect of attracting electrons of O in –OCH₂ from H after the O atom in –OCH₂ coordinated with Ca. Thus, the total electron density of H increased, and the absorption peaks moved to a high field.

Elemental analysis, ICP, XRD, FT-IR, and ¹H NMR revealed that Et₂Cit formed a 1:1 complex with Ca²⁺, that is, CaEt₂Cit.

3.3.5. Coordination Mechanism. The above results showed that Ca²⁺ was coordinated with Et₂Cit. O in –COO and C–O–C of Et₂Cit was coordinated with Ca²⁺ in bidentate ligand. Two kinds of –OCH₂CH₃ had different chemical environments in the crystals, that is, 1,3-CaEt₂Cit and 1,5-CaEt₂Cit. However, their proportions were still difficult to ascertain because of their similar physical and chemical properties. Based on the above characterization results, two kinds of coordination of Et₂Cit with Ca²⁺ are shown in Figure 7.

We rule out the possible coordination of hydroxyl group of Et₂Cit based on the reason that the FT-IR (Figure 5) and ¹H NMR spectra (Figure 6) have confirmed that one of the carboxyl of Et₂Cit was coordinated with the calcium ion. When one –COOH and one –COOCH₂CH₃ in Et₂Cit were coordinated with calcium ion, the –OH group and

![Figure 6: ¹H NMR spectra of Et₂Cit and its complex CaEt₂Cit. (a₁, b₁, and c₁) are Et₂Cit; (a₂, b₂, and c₂) are CaEt₂Cit. (a) Total spectra, (b) δ = 2.70–3.00 section, (c) δ = 4.10–4.21 section.](image-url)
the coordinated Ca ion were separated on opposite sides of the center C atom of Et$_2$Cit (Figure 7); thus –OH cannot coordinate with calcium ion because of the space steric hindrance.

4. Conclusion

The coordination dynamics and effect of Et$_2$Cit and Na$_3$Cit pH on Ca$^{2+}$ in saline water were studied. In 37°C saline water, the coordination dynamics equations of Et$_2$Cit and Na$_3$Cit with Ca$^{2+}$ were $r = 120a^{2.46}$ and $r = 289a^{2.44}$, respectively. The reverse reaction rate constants ($k_{\text{rs}}$) of coordination with CaCl$_2$ were 0.52 and 0.15 L·mol$^{-1}$·s$^{-1}$ for Et$_2$Cit and Na$_3$Cit, respectively. The dissociation rate of Ca$^{2+}$ of CaEt$_2$Cit was faster than that of CaCit. The increased pH accelerated the dissociation of the complex. With increased pH from 6.0 to 8.0, $k_{\text{rs}}$ of Et$_2$Cit-CaCl$_2$ increased from 0.04 to 19.80, which was beneficial in improving the anticoagulant effect. Et$_2$Cit and Ca$^{2+}$ were coordinated to form a 1:1 complex, and O atoms in –COOH and C–O–C of Et$_2$Cit were coordinated with Ca$^{2+}$ in bidentate ligand. Et$_2$Cit was able to coordinate with Ca$^{2+}$, and its release capacity of Ca$^{2+}$ was stronger than that of Et$_2$Cit. Thus, it did not require an intravenous infusion of calcium when used as an anticoagulant, thereby avoiding hypocalcemia and hypercalcemia that can be caused by Na$_3$Cit. Overall, Et$_2$Cit was a better anticoagulant than Na$_3$Cit.

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