Auto/paracrine nicotinergic peptides participate in cutaneous stress response to wounding

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Abbreviations: ACh, acetylcholine; AGKOS, agarose gel keratinocyte outgrowth system; GM, growth medium; KCs, keratinocytes; MPO, myeloperoxidase; nAChR, nicotinic acetylcholine receptor; NIgG, normal mouse IgG; SLURP, secreted mammalian Ly-6/urokinase-type plasminogen activator-related protein

Restoration of the epidermal barrier (epithelialization), is a major component of cutaneous response to stress imposed by wounding. Learning physiologic regulation of epithelialization may lead to novel treatments of chronic wounds. The non-canonical ligands of nicotinic acetylcholine receptors SLURP (secreted mammalian Ly-6/urokinase-type plasminogen activator-related proteins)-1 and -2 are produced by keratinocytes (KCs) and inflammatory cells to augment physiologic responses to non-neuronal acetylcholine, suggesting that they can affect wound epithelialization and inflammation. In this study, recombinant (r)SLURP-1 and -2 exhibited dose dependent effects on migration of cultured KCs, and monoclonal antibodies inactivating auto/paracrine SLURPs in mouse skin delayed wound epithelialization. While effects of rSLURPs on migration were opposite, with rSLURP-1 inhibiting and rSLURP-2 stimulating migration of KCs, each anti-SLURP antibody produced a negative effect on epithelialization in vivo, suggesting their more extensive than regulation of keratinocyte migration involvement in wound repair. Since inflammation plays an important role in stress response to wounding, we measured inflammation biomarkers in wounds treated with anti-SLURP antibodies. Both anti-SLURP-1 and -2 antibodies, or their mixture, caused significant elevation of wound myeloperoxidase, IL-1β, IL-6 and TNFα. Taken together, results of this study demonstrated that SLURP-1 slows crawling locomotion of KCs, and exhibits a strong anti-inflammatory activity in wound tissue. In contrast, SLURP-2 facilitates lateral migration of KCs, but shows a lesser anti-inflammatory capacity. Thus, combined biologic activities of both SLURPs may be required for normal stress response to skin wounding, which favors clinical trial of rSLURP-1 and -2 in wounds that fail to heal.

Introduction

Restoration of the epidermal barrier, i.e., epithelialization, is a major component of the natural cutaneous response to a stress imposed by wounding. Epithelialization is just one step of a highly orchestrated process of wound repair that involves a multitude of cells and events. Initial stages (1–3 d post injury) involve the formation of a blood clot, crawling locomotion of epidermal keratinocytes (KCs) over the provisional matrix, and early inflammatory response mediated mainly by neutrophils. At 4–7 d, lymphocytes and macrophages are present in abundance. Formation of new epidermis over the dermal gap of < 2 cm is usually completed by days 10 to 12, concomitant to granulation tissue formation. This phase is accompanied by attenuation of the inflammatory response. Subsequent healing stages are characterized by matrix remodeling, differentiation of the newly formed epidermis and increased elastic fiber content.

The main objective of our research is to identify novel approaches to facilitate epithelialization of wounds that fail to heal. Burn injuries, deep erosions, chronic ulcers, diabetic wounds, graft donor sites are but a few of the clinical situations that could benefit from development of novel approaches of pharmacologic control of epithelialization. We and others have generated an overwhelming evidence of the regulatory function of the auto/paracrine cutaneous acetylcholine (ACh) axis in KCs and inflammatory cells, which connects with the clinical arenas of wound repair. Already completed studies have demonstrated abundance of auto/paracrine of ACh and its receptors in epidermis and illustrated feasibility of treating mucocutaneous erosions and ulcers by a novel pharmacologic approach utilizing cholinergic drugs. Pyridostigmine bromide, which produces cholinomimetic effects due to both inhibition of ACh degradation and activation of nicotinic ACh receptors (nAChRs), healed skin erosion in patients with pemphigus. This justified the use of the solution of the cholinomimetic pilocarpine for topical treatment of pemphigus erosions. The nicotinic agonist nicotine has shown its efficacy in local treatment of cutaneous and oral ulcers. However, nicotine toxicity due to unavoidable off target effects in the neural system precludes its wide use and urges a search for non-toxic nicotinergic drugs.
that can mimic healing effects of nicotine in mucocutaneous wounds.

In a search for a non-toxic nicotinic agonist, we have identified rapid and profound effects on keratinocyte vital functions of the novel non-canonical ligands of nAChRs secreted mammalian Ly-6/urokinase-type plasminogen activator receptor-related protein (SLURP)-1 and -2 that are produced by skin cells to augment the physiologic effects of ACh at nAChRs. We cloned both SLURPs, produced recombinant proteins and generated monoclonal antibodies that visualized SLURP-1 and -2 in human epidermis and oral mucosa. In the intact epidermis, expression patterns of SLURP-1 and -2 are somewhat different, with SLURP-1 being predominant in the upper epithelial layers and SLURP-2—in the lower layers. As expected based on homology between human and murine SLURP molecules, the antibodies raised against human SLURP-1 and -2 visualized their murine counterparts in the epithelializing murine cutaneous wounds. Interestingly, KCs comprising epithelial tongues covering the epithelializing wounds abundantly expressed SLURP-2, but very little amounts of SLURP-1. The opposite was observed in perilesional epidermis, consistent with observations that SLURP-2 is important for keratinocyte survival, whereas SLURP-1—for keratinocyte maturation and terminal differentiation.

The cell function and gene expression studies have shown that SLURPs play important roles in regulating vital functions of KCs and inflammatory cells, suggesting that during wound repair they may control both epithelialization and inflammation. Indeed, pilot studies showed feasibility to accelerate wound epithelialization due to synergistic regulation by recombinant (r) SLURP-1 and -2 of the integrin expression in KCs. rSLURP-1 and rSLURP-2 that of the “migratory” integrins αβ and αβ, and rSLURP-2 that of the “sedentary” integrins αβ and αβ. The effect of rSLURP-1 was predominantly mediated by keratinocyte α7 and α9, nAChR and that of SLURP-2 by non-α7, such as α3 and α9, nAChRs. Since these same receptors modulate expression of the SLURP-1 and -2 genes, it appears that cutaneous stress response to wounding is mediated, in part, by a reciprocally arranged auto/paracrine cholinergic network.

In this study, we ultimately demonstrated the pharmacologic mechanism of action of rSLURP-1 and -2 on the crawling locomotion of human epidermal KCs in vitro, and identified a novel mechanism of protective action of the auto/paracrine SLURPs during cutaneous stress response to wounding mediated by their anti-inflammatory action. Taken together, these findings should justifiably be a trial of rSLURP-1 and -2 in the treatment of chronic wounds.

**Results**

**Dose dependent effects of rSLURP-1 and -2 on keratinocyte migration under agarose.** To determine direct effects of the auto/paracrine nicotinergic peptides SLURP-1 and -2 on lateral migration of KCs, we used the agarose gel keratinocyte outgrowth system (AGKOS), that proved to be a sensitive and reliable in vitro system for assessing pharmacologic effects of test compounds on keratinocyte activities mediating wound epithelialization. To ascertain that the observed changes in the migration distance of KCs under agarose are mediated by the specific, i.e., pharmacologic, effects of rSLURPs, each recombinant protein was used in the increasing concentrations ranging from 0.01 to 0.5 μg/ml. While rSLURP-1 decreased, rSLURP-2 increased the keratinocyte outgrowth rate in the dose-dependent fashion (Fig. 1). The differences from control became significant (p < 0.05) at the 0.1 μg/ml concentration of each rSLURP protein.

These results suggested that auto/paracrine SLURP-1 and -2 exhibit reciprocal regulation of lateral migration of KCs during cutaneous stress response to wounding.

**Delayed epithelialization of cutaneous wounds treated with anti-SLURP antibodies.** Next, we tested the hypothesis auto/paracrine SLURP proteins would exhibit opposite effects on wound epithelialization in vivo. We measured changes in the rate of epithelialization of excisional cutaneous wounds in BALB/c mice treated daily with microinjections of the antibodies to SLURP-1 and/or -2 that recognize their respective target SLURP proteins in murine skin. While the control mice had their wounds fully epithelialized by the 8th day post wounding, the experimental mice treated with anti-SLURP antibodies showed delay in epidermal closure (Fig. 2). The observed alterations in the epithelialization rate were specific to inactivation of endogenous SLURPs, because microinjections of normal mouse IgG (NlgG) did not cause any delays at same time points.

During the first 4 d post wounding, wounds treated with anti-SLRUP-1 antibody healed as fast as control wounds, but exhibited a significant (p < 0.05) delay on the 8th day. In contrast, significantly (p < 0.05) inhibited epithelialization rate of the wounds treated with anti-SLURP-2 antibody was observed on both the 4th and the 8th days post wounding. A combination of both antibodies dramatically deteriorated epithelialization. The wounds treated with a mixture of anti-SLURP-1 and -2 antibodies failed to heal by the time points the wounds treated with each antibody separately had been completely covered by new epidermis (Fig. 2).

These observations indicated that despite their opposite effects on lateral migration of KCs, auto/paracrine functions of SLURP-1 and -2 during the cutaneous stress response to wounding extend beyond regulation of keratinocyte motility.

**Increased inflammation of cutaneous wounds treated with anti-SLURP antibodies.** Since inflammation plays an important role in the cutaneous stress response to wounding, we measured inflammation biomarkers in wounds treated with anti-SLURP antibodies. The intensity of inflammation was judged from the amounts of myeloperoxidase (MPO), IL-1β, IL-6 and TNFα in the homogenates of excised wound tissues at 1, 4 and 12 d post wounding. Microinjections of anti-SLURP-1 or -2 antibody or their mixture caused significant (p < 0.05) elevation of all biomarkers on the 4th day post wounding (Fig. 3). By the 12th day, the amounts of IL-6 and TNFα in the wounds treated with anti-SLURP-2 antibodies decreased to the normal range and those of MPO and IL-1β were close to, yet significantly (p < 0.05) different from norm. Wounds treated with anti-SLURP-1 antibody or a combination of both anti-SLURP antibodies featured significantly (p < 0.05) increased levels of inflammation.
throughout the entire observation period, with the highest degree of inflammation observed in wounds treated with a mixture of both anti-SLURP antibodies (Fig. 3).

These results indicated that auto/paracrine SLURPs control the inflammatory component of cutaneous stress response to wounding, and that SLURP-1 exhibits a stronger anti-inflammatory activity, compared with SLURP-2.

**Discussion**

In this study, we demonstrated for the first time that both nicotinergic peptides SLURP-1 and -2 produce in a complementary fashion a protective action against cutaneous stress caused by wounding, with each protein playing a distinctive biologic role. While SLURP-1 slows down crawling locomotion of KCs, it exhibits a strong anti-inflammatory activity in wound tissue. On the other hand, SLURP-2 facilitates lateral migration of KCs, but has a lesser anti-inflammatory capacity. Thus, combined biologic activities of both SLURPs appear to be required for normal epidermal closure.

Results of our previously completed studies suggested that auto/paracrine SLURP-1 and -2 exhibit reciprocal effects on the migratory function of KCs during wound epithelialization. Since accelerated migration of the KCs re-epithelializing a wound plays a pivotal role in wound repair, we determined in this study the dose-dependence of keratinocyte response to rSLURP-1 and -2 in vitro. The obtained results demonstrated the specificity of rSLURP actions, and suggested that anti-SLURP-2, but not anti-SLURP-1, antibody should delay epithelialization of skin wounds. However, despite opposite effects of rSLURP-1 and -2 on lateral migration of KCs in the AGKOS assay, simultaneous inhibition of auto/paracrine functions of SLURP-1 and -2 in skin wounds with a mixture of anti-SLURP-1 and -2 antibodies delayed epithelialization even more than the anti-SLURP-2 antibody given alone. This observation indicated that during the cutaneous stress response to wounding, the physiologic actions of auto/paracrine SLURPs extend beyond regulation of crawling locomotion of KCs. Hence, we hypothesized that SLURP-1 and -2 facilitate epidermal closure also by controlling inflammation.

In addition to extracellular matrix proteins and auto/paracrine growth factors, keratinocyte migration in wound bed is controlled by a large variety of cytokines and chemokines released by inflammatory cells, which can either stimulate or inhibit epithelialization. The role of inflammation in wound healing remains controversial. Although inflammation is an essential step of normal wound repair, the extent of wound inflammation appears to inversely correlate with the epithelialization rate. For instance, skin wounds in diabetic mice feature robust inflammation in association with overabundance of proliferating epidermis at the wound edge and insufficient migration of KCs, resulting in delayed epithelialization. Noteworthy, KCs themselves produce cytokines and chemokines that can attract inflammatory cells. Thus, the two processes—epithelialization and

**Figure 1.** rSLURP-1 and -2 exhibit opposite effects on lateral migration of human KCs under agarose. Normal human KCs were seeded in the 3 mm wells of the AGKOS plates measuring random cell migration, exposed to increasing concentrations of rSLURP-1 (-●-) or -2 (-■-) dissolved in growth medium, and incubated for 10 d with daily refreshing the media. Migration distance was measured as described in Materials and Methods. Migration distance of control KCs incubated in growth medium without rSLURPs was taken as 100%, and the results expressed as percent of intact controls. Asterisks = p < 0.05 compared with control.
inflammation—are mutually interdependent and inseparable in wound repair. Therefore, to adequately evaluate the therapeutic potential of rSLURPs in chronic wounds, we measured the effect of inactivation of auto/paracrine SLURPs with specific antibodies on the degree of wound inflammation.

Treatments of cutaneous wounds with the antibodies inhibiting biologic activities of auto/paracrine SLURPs increased the amounts of inflammatory molecules in the wound specimens, indicating that these nicotinergic peptides indeed control an inflammatory arm of the cutaneous stress response to wounding. This observation is not surprising, because previous studies have documented that nicotinergic signaling inhibits cutaneous neutrophil infiltration, and pro-inflammatory functions of macrophages, and T- and B-cells. Functional inactivation of auto/paracrine SLURP-1 has a greater impact on the cutaneous immunoregulatory activity, as indicated by a report that patients with genetic SLURP-1 mutation have defective T-cell function. Indeed, both SLURP-1 and SLURP-2 are expressed in various immune cells and organs, and appear to be involved in regulating lymphocyte function via the nAChR-mediated pathways. rSLURPs could mimic both a stimulatory effect of the nAChR agonist nicotine on wound epithelialization and its inhibitory effect on wound inflammation, perhaps, because both SLURP peptides, just like nicotine, bind to and activate the same nAChR subtypes. However, in contrast to nicotine that activates all nAChR subtypes, the specific effects of each SLURP differ, because they preferentially activate distinct nAChRs that are selectively coupled to unique regulation of the epithelial and inflammatory cell functions.

In conclusion, the results of this study established auto/paracrine SLURPs as physiologic regulators of cutaneous stress response to wounding and suggested that rSLURPs may become a prototype drugs for pharmacologic control of wounds that fail to heal. Therapeutic use of rSLURPs should avoid the systemic toxicity of the canonical agonists of nAChRs, such as nicotine, because SLURPs are cytokine-like physiologic substances normally present in blood, saliva, skin, sweat and urine.

Materials and Methods

Chemicals. The full length rSLURP-1 and rSLURP-2 were manufactured at Virusys Corporation, as detailed by us before. Agarose type A was obtained from Accurate Chemical and Scientific Corporation. rSLURP-1 and -2 monoclonal antibodies 336H12–1A3 and 341F10–1F12, respectively, were from Research and Diagnostic Antibodies. N1gG was obtained from Santa Cruz Biotechnology, Inc. The ELISA kit for measuring mouse MPO was purchased from Hycult Biotech, IL-1β and IL-6 from Abcam, Inc. and TNFα from Thermo Fisher Scientific.

Keratinocyte migration assay. Human KCs were purchased from Invitrogen Life Technologies and grown in the serum-free keratinocyte growth medium containing 5 ng/ml EGF and 50 μg/ml bovine pituitary extract (GIBCO-BRL) in accordance to the protocol provided by the vendor. KCs were used in experiments between passages 2 and 4, grown in growth medium (GM) to approximately 80% confluence. The rate of epithelialization in vitro was measured by the random migration distance in AGKOS, as detailed by us elsewhere. Briefly, a confluent keratinocyte monolayer was formed by loading KCs at a high density (4 × 10⁴ cells/10 μl) into 3 mm well in an agarose gel and incubating in GM to allow the cells to adhere to the dish bottom and form intercellular junctions. The cultures were incubated for 10 d in a humid CO₂ incubator with daily changes of GM containing test concentrations of rSLURP-1 or -2 versus GM given alone (control). To standardize results obtained in different experiments, the mean values of the migration distances were converted into the percentage of control. The control value in each experiment was determined by measuring the baseline migration distance (in m) and taken as 100%.

In vivo wounding and morphometric and biochemical analyses of cutaneous wounds. This study was approved by the
The wounds in control animals were similarly treated with plain saline solution alone or containing 1 μg of NlG. Both experimental and control mice were euthanized on days 1, 4, 8 and 12 post wounding, and the wounds with 1 mm margin of surrounding skin were excised. One wound specimen from each animal was fixed, stained and used for the morphometric assay of epithelialization rate, whereas another one was freshly frozen and stored at −80°C until use in biochemical assays. The epithelialization rate was estimated by measuring the lengths of the tongues of new epithelium extending from either side of the central portion of the wound. The percentage epidermal closure was calculated as a ratio of epithelialized wound area (i.e., covered by new epithelium) to initial wound area that was identified by an abrupt change in the epidermal and dermal morphology at the incision site.62-64 For the ELISA assays of MPO, IL-1β, IL-6 and TNFα, University California-Irvine Animal Care and Use Committee. Assay of the skin wound epithelialization rate was performed by histomorphometric analysis in accordance to our previously published protocol.25,60 Briefly, using a 5 mm in diameter Healthlink Biopsy Punch (MedPlusUSA Medical Supplies), full thickness excisions through the panniculus carnosus were made in the anesthetized skin of 6–7 weeks old BALB/c mice (The Jackson Laboratory), in whom the hair cycle had been synchronized by the anagen induction technique.61 Each animal received 2 wounds at the symmetric sites of the central back, 0.5 cm off the vertebral line. Wounds were dressed with the Tegaderm™ (3M) and wounded animals were individually housed under aseptic conditions. Starting 1 h after wounding, we initiated daily treatments of experimental wounds by microinjections of 100 μl saline containing 1 μg of anti-SLURP-1 and/or -2 antibody. The wounds in control animals were similarly treated with plain saline solution alone or containing 1 μg of NlG. Both experimental and control mice were euthanized on days 1, 4, 8 and 12 post wounding, and the wounds with 1 mm margin of surrounding skin were excised. One wound specimen from each animal was fixed, stained and used for the morphometric assay of epithelialization rate, whereas another one was freshly frozen and stored at −80°C until use in biochemical assays. The epithelialization rate was estimated by measuring the lengths of the tongues of new epithelium extending from either side of the central portion of the wound. The percentage epidermal closure was calculated as a ratio of epithelialized wound area (i.e., covered by new epithelium) to initial wound area that was identified by an abrupt change in the epidermal and dermal morphology at the incision site.62-64 For the ELISA assays of MPO, IL-1β, IL-6 and TNFα,
the wound tissue was homogenized using TissueRuptor (Qiagen Inc.) and processed in accordance to the manufacturers’ protocols. The results were expressed as absolute values normalized to the total amount of protein determined by the Quick Start™ Bradford protein assay (BioRad Laboratories). At least 3 animals per time point were used in each experiment.

Statistical analysis. Statistical significance was determined using the Student’s t-test. The differences were deemed as significant when the calculated p value was < 0.05.

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Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

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