Prospective study of human herpesvirus 8 oral shedding, viremia, and serological status among human immunodeficiency virus seropositive and seronegative individuals in Sao Paulo, Brazil

Paulo H. Braz-Silva, Tania R. Tozetto-mendoza, Laura M. Sumita, Wilton Freire, Michelle Palmieri, Alan M. do Canto, Vivian I. Avelino-Silva, Marina Gallottini, Philippe Mayaud, and Claudio S. Pannuti

*Instituto de Medicina Tropical de Sao Paulo—IMTSP, Laboratorio de Virologia—UIM 52; *Faculdade de Odontologia—FOUSP, Departamento de Estomatologia, Disciplina de Patologia Geral; *Faculdade de Medicina—FMUSP, Departamento de Molestias Infecciosas e Parasitarias; *Departamento de Estomatologia, Disciplina de Patologia Oral e Maxilofacial; Universidade de Sao Paulo, Sao Paulo, Brazil; *Department of Clinical Research, Faculty of Infectious and Tropical Diseases, London School of Hygiene and Tropical Medicine, London, United Kingdom

ABSTRACT

Human herpesvirus 8 (HHV-8) is a gamma-herpesvirus and etiological agent of all forms of Kaposi sarcoma (KS). Saliva may play an important role in HHV-8 transmission in specific populations. Little is known about HHV-8 oral shedding pattern and the possible correlation with the HHV-8 serological profile and viremia. A prospective study was conducted of HHV-8 salivary excretion among human immunodeficiency virus HIV-seronegative (n = 47) and -seropositive (n = 44) homosexual men and HIV-seropositive women (n = 32) over a 6-month period with monthly HHV-8 serologies (immunofluorescence assays to identify antibodies to latent and lytic HHV-8 viral proteins, and a whole-virus HHV-8 enzyme-linked immunosorbent assay [ELISA]), monthly HHV-8 DNA serum/plasma detection, and daily self-collected oral rinses for HHV-8-DNA detection using real-time polymerase chain reaction.

HHV-8 seropositivity was 51.1%, 63.6%, and 37.5%, in the three studied groups. There was no case of HHV-8 DNA detection in serum/plasma. Intermittent detection of oral HHV-8 DNA was observed during 5.1% (110/2,160) of visits among 28% (18/64) of HHV-8-seropositive individuals, all of whom were males and HHV-8 ELISA seropositive. In immunologically controlled populations of Brazil, HHV-8 oral shedding was limited to HHV-8-seropositive men, occurred infrequently and intermittently, and was not linked to HHV-8 viremia, suggesting a limited potential for oral or blood transmission.

Introduction

Transmission of human herpesvirus 8 (HHV-8), the agent of Kaposi sarcoma (KS) and other human immunodeficiency virus (HIV)-associated rare conditions, has been the subject of conflicting evidence. Oral samples have been consistently shown to have the highest rates of HHV-8 viral detection compared to semen, urine, urethral, vaginal, and anal samples, suggesting that saliva may play an important role in HHV-8 transmission. On the other hand, transmission through blood and blood transfusion remains a controversial issue with no conclusive evidence. Studies among blood donors outside KS endemic areas have shown very rare detection of HHV-8 DNA in blood samples, even among HHV-8-seropositive individuals.

Associations between HHV-8 serological status and HHV-8 DNA oral shedding are also not clear, with some studies even showing HHV-8 oral shedding in HHV-8-seronegative individuals. The lack of a gold standard for HHV-8 serological status further compounds the limited ability to determine ‘true infection’ status.

This study aimed to assess prospectively the frequency of HHV-8 detection in saliva, serum, and plasma and their association with HHV-8 serological status among HIV-seronegative and -seropositive men who have sex with men (MSM) and HIV-seropositive women—groups that have a high probability of being infected with HHV-8.

Methods

Study participants and sample collection

A convenience sample of individuals were included from three groups: HIV-seronegative MSM, HIV-seropositive MSM, and HIV-seropositive women. Volunteer patients either self-presenting or referred by the STD Training Center/AIDS-SP in Sao Paulo for dental care were consecutively recruited at the Dental Special Care Center of the School of Dentistry, University of Sao Paulo, Brazil, where
the purpose and procedures of the study were explained. The patients had no history of KS or other lesions associated with HHV-8. All patients signing informed consent were invited to a prospective follow-up over a 6-month period consisting of two sequential 3-month periods separated by 60 days, with biweekly appointments at the dental clinic where patients would bring their daily (Monday to Friday) self-collected mouthwashes. Self-collection commercial oral rinse kits (Listerine®, Johnson & Johnson, Sao Paulo, Brazil) were distributed to each patient, consisting of 50 mL plastic tubes (pre-labeled for the collection day) and a bottle of Listerine®. All participants were instructed to use 5 mL of the mouthwash solution for 1 min at night and to collect all the contents of the oral rinse in the tube. Samples were stored in patients’ homes in a refrigerator at 4°C until the next dental appointment. The aim was to collect a total of 120 oral rinses from each patient. If a patient had not submitted this number, the collection period was extended up to 4 weeks (2 weeks in each period of 3 months) to attain that number. In addition, blood samples were drawn at the study entry and at monthly appointments. All samples were stored at −80°C until testing.

Serological assays

Six serum samples from each patient were tested without knowledge of other results in a random patient and samples order for HHV-8 serologies at the Laboratory of Virology, Institute of Tropical Medicine, University of Sao Paulo, using in-house immunofluorescence assays (IFA) to identify antibodies to latent and lytic HHV-8 viral proteins, and a whole-virus HHV-8 enzyme-linked immunosorbent assay (ELISA) for immunoglobulin G detection, as described previously [14].

HHV-8 molecular assays

DNA extraction

DNA was extracted from oral rinse, serum, and plasma samples (200 μL) using a NucliSSENS® easyMAG® (bioMerieux, Durham, NC), an IVD-labeled automated system for total nucleic acid extraction based on magnetic silica technology, according to the manufacturer’s instructions. DNA concentration was measured by spectrophotometry at 260 nM in a NanoDrop™ (Thermo Fisher Scientific, Waltham, MA). Beta-actin DNA was also detected in all samples by using real-time polymerase chain reaction (PCR), according to the manufacturer’s protocol (Taq Man DNA Template Reagents Kit; Applied Biosystems, Foster City, CA), in order to evaluate the viabiliy of the DNA isolated from each sample.

ORF 26 real-time PCR.

Oral rinses, serum, and plasma samples were analysed in triplicate for HHV-8 DNA detection and quantification using real-time PCR, as described [11]. Quantities of HHV-8 DNA were log10 transformed to normalize values for analysis. Samples with ≥5 copies/mL (0.7 log10) were considered positive.

Statistical analyses

The proportions of patients and days with detectable HHV-8 DNA in oral rinse, serum, and plasma samples were compared across the groups and by serological assay using chi-square statistics. For the quantitative analysis of viral shedding, the median values of HHV-8 DNA were compared among samples with detectable HHV-8 using the Kruskal–Wallis and Wilcoxon rank-sum tests. Inter-assay (serology) concordance (overall, positive, and negative agreements) was calculated using simple proportions. Performance of serological assays to detect HHV-8 shedding was calculated using standard 2 × 2 tables, and results are given with their 95% confidence intervals (CI). Data were analysed using Stata v14 (Stata Corp., College Station, TX).

Results

Overall, 123 individuals (47 HIV-seronegative MSM, 44 HIV-seropositive MSM, and 32 HIV-seropositive women) were enrolled in this study from March to May 2013, and they were followed up for a 6-month period. Participants’ characteristics are shown in Table 1. All HIV-seropositive individuals were taking highly active antiretroviral therapy (HAART), and around 80% had undetectable HIV plasma viral loads (threshold for detection: 50 copies/mL) and high CD4 + T-lymphocyte counts (68.1% of men and 75.0% of women with CD4+ count ≥500 cells/mm3) at enrolment.

At the end of the study period, all patients had produced 120 self-collected oral samples, six serum samples, and six plasma samples. Forty-two (34.1%) patients needed an extended period to collect the oral rinses (ranging from 3 to 20 samples for a range of 1 to 4 weeks).

Overall, 24/47 (51.1%) HIV-seronegative MSM, 28/44 (63.6%) HIV-seropositive MSM, and 12/32 (37.5%) HIV-seropositive women were positive by at least one HHV-8 serological assay over six time points (Table 1). Considering each assay separately, there was a higher positivity by ELISA (48/123; 39.0%), followed by IFA-LANA (33/123; 26.8%), and IFA-lytic (22/123; 17.9%).

The overall intra-assay concordance in 738 serum samples was 100% (738/738) for ELISA, 99.2% (732/738) for IFA-LANA, and 98.9% (730/738) for IFA-lytic,
Table 1. Participants’ characteristics and HHV-8 antibody and DNA detection among HIV-positive and HIV-negative patients in Sao Paulo, Brazil.

| Characteristics | Male HIV negative, N = 47 | Male HIV positive, N = 44 | Female HIV positive, N = 32 | p-Value |
|-----------------|--------------------------|--------------------------|---------------------------|---------|
| Age (years), median (interquartile range) | 30 (26–37) | 40 (35–45) | 42 (39–52) | <0.01* |
| Proportion with CD4+ >500 cells/mm³ (%) | n/a | 30 (68.1) | 24 (75.0) | 0.52 |
| Proportion with undetectable HIV PVL (%) | n/a | 35 (79.5) | 27 (84.3) | 0.59 |
| HHV-8 serology positivity | ELSA (%) | 18 (38.3) | 24 (54.6) | 6 (18.8) | <0.01 |
| IFA-LANA (%) | 14 (29.8) | 13 (29.6) | 6 (18.8) | 0.49 |
| IFA-lytic (%) | 9 (17.0) | 10 (22.7) | 4 (12.5) | 0.51 |
| Any assay (%) | 24 (51.0) | 28 (63.6) | 12 (37.5) | 0.08 |

HHV-8 DNA detection among all patients

| Serum (%) | 0 | 0 | 0 | — |
| Plasma (%) | 0 | 0 | 0 | — |

Saliva (per person analysis)

| Positive person/tested (%) | 7/47 (14.9) | 11/44 (25.0) | 0 | 0.23 |

Saliva (per sample analysis)

| Positive sample/tested (%) | 40/5,640 (0.7) | 70/5,280 (1.3) | 0 | <0.01 |

HHV-8 DNA saliva detection among HHV-8-seropositive patients only

| Positive individual/tested (%) | 7/24 (29.2) | 11/28 (39.3) | 0 | 0.71 |
| Positive sample/tested (%) | 40/2,880 (1.4) | 70/3,360 (2.1) | 0 | 0.04 |
| Positive samples among shedders samples (%) | 40/840 (4.8) | 70/1,320 (5.3) | 0 | 0.58 |

HHV-8 DNA log₁₀ copies/mL

| 2.8 (2.4–3.2) | 3.7 (3.3–4.5) | 0 | <0.01 |

Continuous variables are presented as medians and interquartile ranges; P-values are presented for comparison across three groups

* Male HIV-negative group was significantly different from male HIV-positive and female HIV-positive groups in pairwise comparison; difference between male HIV-positive and female HIV-positive groups was not significant.

† Threshold for detection 50 copies/mL.

‡ In pairwise comparison, ELISA positivity was significantly different between male HIV-positive and female HIV-positive groups only.

§ Per sample analysis: six serum, six plasma, and 120 saliva samples per patient.

¶ Pairwise comparison for male HIV-negative and male HIV-positive groups.

HHV-8, human herpesvirus 8; HIV, human immunodeficiency virus; PVL, plasma viral load; ELISA, enzyme-linked immunosorbent assay; IFA, immunofluorescence assays.

showing excellent reproducibility for all tests. Patients with discordant intra-assay results were arbitrarily considered positive because the majority (4/6) of their serum samples were positive. The intra-assay concordance for negative results was 100% for each assay (Table 2).

HHV-8 DNA was not detected in any of the 738 serum or plasma samples after triplicate PCR testing. HHV-8 DNA was not detected in any oral samples of the 59 HHV-8-seronegative individuals (total 7,080 samples tested), but it was detected in 18/64 (28%) HHV-8-seropositive individuals: 7/24 (29.2%) HIV-seronegative MSM and 11/28 (39.3%) HIV-seropositive MSM (Table 1). The frequency of HHV-8 DNA detection in samples from shedders was 5.1% (110/2160) overall, 4.8% (40/840) in samples from HHV-8-seronegative MSM, and 5.3% (70/1320) in samples from HHV-8-seropositive MSM (p = 0.58), with a range of positive samples between 3/120 (2.5%) and 11/120 (9.2%) per patient. The median quantity of HHV-8 DNA detected among shedders was significantly higher among the HIV-seropositive compared to HIV-seronegative MSM (3.7 vs. 2.8 log₁₀ copies/mL, p < 0.01; Table 1).

The HHV-8 serological profile and the performance of individual and combination serological assays to detect HHV-8 shedders among the 91 men is shown in Table 3. The evaluation did not include women, since none had any demonstrable shedding. The whole-virus ELISA had the highest sensitivity (100.0%; 95% CI 81.5–100.0), lowest specificity (67.1%; 95% CI 55.1–77.7), but high negative predictive value (NPV; 100.0%; 95% CI: 92.7–100.0). The best combination of assays was ELISA plus IFA-LANA for its high specificity (91.8%; 95% CI 83.0–96.9) without much loss of sensitivity (83.3%; 95% CI 58.6–96.4) and high positive predictive value (PPV; 71.4%; 95% CI 47.8–88.7; Table 3).

Discussion

In these high-risk populations, 29% of HHV-8/HIV dually seropositive MSM, as well as 39% of HHV-8-seropositive/HIV-seronegative MSM, were shedding HHV-8 DNA at least once over a 6-month period, while none of the HHV-8/HIV dually seropositive women shed HHV-8. Among MSM, the frequency of oral shedding was low (around 5% of days), irrespective of HIV serostatus, despite the large number of samples (120) tested per individual. However, the median quantity of oral HHV-8 DNA during episodes was significantly higher among HIV-seropositive shedders. Compared to other serological assays, ELISA would be the best test to identify HHV-8 shedders, as it combines the highest NPV (no shedding detected among ELISA-seronegative individuals) and the highest sensitivity. The pattern of HHV-8
oral shedding was sporadic for all 18 shedding individuals, with shedding days ranging from 2.5% to 17% and very few shedding episodes occurring on consecutive days.

The present findings differ from other studies that have reported a high frequency (32–68%) of HHV-8 shedding and high viral loads among HIV-seropositive women in Africa [3,4]. These studies were carried out in KS endemic areas (Kenya) among very specific populations (sex workers who did not use HAART) [3,4]. The small number of HHV-8-seropositive women in the present study, the different participant characteristics, and the fact that Brazilian patients were immunologically well controlled on HAART may explain these different results. However, it should be considered that some studies on gender susceptibility to HHV-8 and KS report that women are more resistant than men to infection by HHV-8 and development of the disease, which suggests that female hormones may provide protection to women [15,16].

Few studies have systematically analyzed the frequency and dynamics of HHV-8 oral shedding in KS non-endemic areas [2,5–7,17,18]. In the United States, Pauk et al. analyzed 1,134 oropharyngeal samples obtained from daily oral collection among 23 HHV-8-seropositive MSM (including 14 HIV-seropositive men) for a period of approximately 50 days. They found oral HHV-8 DNA detected at least once for 13 participants (57%), with about 50% of those men having frequent shedding (defined as ≥35% positive samples) [2]. Another study from the same group showed frequent and intermittent HHV-8 oral detection: 44 MSM collected oropharyngeal samples for periods of between 25 and 135 days, and 27 (61%) showed at least one positive sample. However, the authors also described HHV-8 oral shedding as intermittent and sporadic [6]. A recent study compared HHV-8 oral shedding between different populations (United States, Peru, Cameroon, Uganda, and Kenya), and the authors did not find a regular pattern of HHV-8 oral shedding, with a great variability observed between participants [17].

The present study found a high prevalence of HHV-8 antibodies in all three groups, although this varied according to the assay used (ranging from 18.7% with IFA-lytic to 56.8% for ELISA). The lack of a gold standard for the serological diagnosis of HHV-8 remains an obstacle for comparing the sensitivity and specificity of each assay, making it difficult to establish what is the most reliable assay to discriminate true HHV-8-infected individuals from those who are not infected [14]. By testing six sequential serum samples blindly from each patient, the present study was able to show excellent intra-assay concordance for all three techniques. Taking into account all 738 samples from the 123 individuals participating in the study, the intra-assay concordance was ≥99% for all assays.

One objective of this study was to determine the performance of serological assays to identify HHV-8 oral shedders. The HHV-8 whole-virus ELISA assay had the highest sensitivity, confirming previous reports [9,11,14], a moderate specificity, and a high NPV. The combination with IFA-LANA assay increased specificity without losing much on sensitivity and had the highest PPV to detect shedders.

It was found that none of the 738 blood samples, including those from the 18 oral shedders, were positive for HHV-8 DNA, despite triplicate testing, confirming earlier findings that in KS non-endemic areas, HHV-8 detection in peripheral blood is very rare, even among HHV-8-seropositive individuals such as blood-bank donors [10,11]. Blood donation by MSM has raised concerns about the possibility of HHV-8 transmission, and in some settings, it has been proposed to impose a lifetime donation deferral for this group [12]. This recommendation would not be supported by the present findings, at least for HHV-8.

### Table 2. Intra-assay concordance for six serum samples obtained from 123 patients in Sao Paulo, Brazil.

| Assay                | Overall concordance, % (n/N) | Concordance among positive results, % (n/N) | Concordance among negative results, % (n/N) | Kappa (95% CI) for overall | p-Value         |
|----------------------|-----------------------------|-----------------------------------------------|---------------------------------------------|---------------------------|----------------|
| ELISA                | 100% (738/738)              | 100% (288/288)                                | 100% (450/450)                              | 99% (90.3–99.9)           | <0.0001        |
| IF-LANA              | 99.2% (732/738)             | 96.9% (192/198)                               | 100% (540/540)                              | 99% (90.3–99.9)           | <0.0001        |
| IF-lytic             | 98.9% (730/738)             | 94.2% (130/138)                               | 100% (600/600)                              | 99% (90.3–99.9)           | <0.0001        |

### Table 3. Performance of serological assays for the detection of salivary HHV-8 DNA shedders among 91 men (47 HIV-seropositive and 44 HIV-seronegative MSM).

| Serologic assay | Positivity, n (%) | Sensitivity % (95% CI) | Specificity % (95% CI) | Positive predictive value % (95% CI) | Negative predictive value % (95% CI) |
|-----------------|------------------|------------------------|------------------------|-------------------------------------|-------------------------------------|
| ELISA           | 18 (100.0)       | 100.0 (81.5–100.0)     | 67.1 (55.1–77.7)       | 42.9 (27.7–59.0)                    | 100.0 (92.7–100.0)                   |
| IFA-LANA        | 15 (83.3)        | 83.3 (58.6–96.4)       | 83.6 (73.0–91.2)       | 55.6 (35.3–74.5)                    | 95.3 (86.9–99.0)                     |
| IFA-lytic       | 6 (33.3)         | 33.3 (13.3–59.0)       | 83.6 (73.0–91.2)       | 33.3 (13.3–59.0)                    | 83.6 (73.0–91.2)                     |
| ELISA + IFA-LANA| 15 (83.3)        | 83.3 (58.6–96.4)       | 91.8 (83.0–96.9)       | 71.4 (47.8–88.7)                    | 95.7 (88.0–99.1)                     |
| ELISA + IFA-lytic| 6 (33.3)         | 33.3 (13.3–59.0)       | 93.2 (84.7–97.7)       | 54.5 (23.4–83.3)                    | 85.0 (75.3–92.0)                     |
| ELISA + IFA-LANA + IFA-lytic | 6 (33.3) | 33.3 (13.3–59.0) | 98.6 (92.6–100.0) | 85.7 (42.1–99.6) | 85.7 (76.4–92.4) |
| IFA-LANA + IFA-lytic | 6 (33.3) | 33.3 (13.3–59.0) | 94.5 (86.6–98.5) | 60.0 (26.2–87.8) | 85.2 (75.6–92.1) |
In conclusion, in immunologically controlled populations of Brazil, HHV-8 oral shedding was limited to HHV-8-seropositive men, occurred infrequently and intermittently, and was not linked to HHV-8 viremia, suggesting a limited potential for oral or blood transmission.

Acknowledgments

We thank the volunteer participants for their time and dedication to this study.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

Supported by FAPESP—Fundação de Amparo a Pesquisa do Estado de São Paulo Grant numbers: 2012/04223-1 and 2012/04303-5.

Notes on contributors

Paulo Henrique Braz-Silva is Assistant Professor of Pathology, Department of Stomatology, University of São Paulo School of Dentistry and Research Associate at the Department of Virology, University of São Paulo Institute of Tropical Medicine since 2014. He earned his Doctor of Dental Surgery (DDS) degree in 2003, and a Master degree in Oral and Maxillofacial Pathology in 2005 from University of São Paulo School of Dentistry. PhD in Oral and Maxillofacial Pathology from University of São Paulo School of Dentistry, and in Cellular and Molecular Biology from University of Nice Sophia Antipolis, France, in 2009. He has conducted researchers in oral shedding of viruses in immunocompromised patients. He has over 30 publications in international and national peer-reviewed journals.

Tania Regina Tozetto-Mendoza is Research Associate at the Department of Virology, University of São Paulo Institute of Tropical Medicine, since 2000. She received her Master (1997) and PhD (2013) degrees in Tropical Disease and International Health from University of São Paulo. She is supervisor of the Program of Specialists in Laboratorial Techniques in Virology and Tropical Hematology of Clinics Hospital of the University of São Paulo School of Medicine. She publishes articles in the field of molecular, cellular and serological diagnosis and her main research interest is in human herpesviruses.

Laura Massami Sumita is Research Associate at Department of Virology, University of São Paulo Institute of Tropical Medicine. She received her Master degree in Medical Sciences from the University of São Paulo Medical School in 2009. She participated in 36 publications in international and national scientific peer-reviewed journals.

Wilton Freire is graduated in Chemistry and works as laboratory specialist at the Department of Virology, University of São Paulo Institute of Tropical Medicine, since 1993.

Michelle Palmieri graduated in Dentistry, from University of São Paulo School of Dentistry (2003). She is Specialist in Oral and Maxillofacial Surgery and assistant professor at Oral and Maxillofacial Surgery Section at FUNDECTO - USP, since 2006. She earned her Master degree in Oral and Maxillofacial Pathology from University of São Paulo School of Dentistry in 2016. She is studying for her PhD degree in Oral and Maxillofacial Pathology from University of São Paulo School of Dentistry. She publishes articles in international and national journals, in the field of Oral and Maxillofacial Surgery and Oral Pathology.

Alan Motta do Canto is assistant surgeon at Unit of Oral and Maxillofacial Surgery, Santa Casa de São Paulo School of Medical Sciences, Brazil; DDS from Universidade Estadual Paulista at Araraquara/São Paulo in 2007. MsC in Oral Diagnosis from University of São Paulo School of Dentistry (2017). Specialist in Oral and Maxillofacial Surgery from Santa Casa de São Paulo School of Medical Sciences, Brazil. He publishes articles in the field of Oral Pathology and Maxillofacial Traumatology.

Vivian Avelino-Silva is a post-doctoral fellow and staff researcher at Department of Infectious and Parasitic Diseases, University of São Paulo Medical School, and Hospital Sirio-Libanes, São Paulo, Brazil. PhD in 2015 and MD in 2004 from University of São Paulo. Author of 24 publications in international and national scientific peer-reviewed journals.

Marina Gallottini graduated in Dentistry from University of São Paulo School of Dentistry in 1986. She earned her Master degree in Oral and Maxillofacial Pathology from the same University in 1989, and a PhD in Oral Pathology from USP in 1994. She is full Professor of Oral and Maxillofacial Pathology, Department of Stomatology, School of Dentistry, University of São Paulo, and Head of the Special Care Dentistry Center at USP. She has experience in the area of Dentistry, with emphasis in Oral Pathology, Oral Medicine and Special Care Needs Patients, working mainly on the following topics: clinical management of systemically compromised patients, HIV/AIDS, developmental disorders, oral opportunistic diseases and salivary markers. She has over 140 peer reviewed journal articles.

Philippe Mayaud is full Professor of Infectious Diseases and Reproductive Health in the Clinical Research Department of the London School of Hygiene & Tropical Medicine (LSHTM), UK, since 2011. He has completed a Medical Doctorate (MD) and training in infectious and tropical diseases at the University of Aix-Marseille, France, and holds a Masters Degree in tropical medicine from the LSHTM, University of London, UK. He is a Visiting Professor at the Faculty of Medicine of the University of São Paulo, Brazil, through a CAPES grant since 2015. He has conducted research on sexually transmitted infections (STIs) and HIV in Africa, and research on human herpes virus type 8 (HHV8) in Brazil. He has over 170 international scientific articles/book chapters; most publications deal with STI/HIV, including randomised interventions trials.

Claudio Sergio Pannuti, PhD and MD, is Associated Professor at the Virology Section of the Institute of Tropical Medicine of São Paulo, University of São Paulo,
and at the Laboratory of Medical Investigation on Virology (LIMHC-FMUSP) of the Clinics Hospital of the University of São Paulo Medical School. He is former Director of the Institute of Tropical Medicine of São Paulo and Head of the Virology Laboratory. He has over 140 publications in international and national peer-reviewed journals.

**ORCID**

Paulo H. Braz-Silva  
http://orcid.org/0000-0002-1842-9521

**References**

[1] Vitale F, Viviano E, Perna AM, et al. Serological and virological evidence of non-sexual transmission of human herpesvirus type 8 (HHV8). Epidemiol Infect. 2000;125:671–675.

[2] Pauk J, Huang ML, Brodie SJ, et al. Mucosal shedding of human herpesvirus 8 in men. N Engl J Med. 2000;343(19):1369–1377.

[3] Taylor MM, Chohan B, Lavreys L, et al. Shedding of human herpesvirus 8 in oral and genital secretions from HIV-1-seropositive and –seronegative Kenyan women. J Infect Dis. 2004;190:484–488.

[4] Phipps W, Saracino M, Selke S, et al. Oral HHV-8 replication among women in Mombasa, Kenya. J Med Virol. 2014;86(10):1759–1765.

[5] Widmer IC, Erb P, Grob H, et al. Human herpesvirus 8 oral shedding in HIV-infected men with and without Kaposi Sarcoma. J Acquir Immune Defic Syndr. 2006;42:420–425.

[6] Casper C, Krantz E, Selke S, et al. Frequent and asymptomatic oropharyngeal shedding of human herpesvirus 8 among immunocompetent men. J Infect Dis. 2007;195:30–36.

[7] Casper C, Redman M, Huang ML, et al. HIV infection and human herpesvirus-8 oral shedding among men who have sex with men. J Acquir Immune Defic Syndr. 2004;35:233–238.

[8] Del Mistro A, Baboci L, Frayle-Salamanca H, et al. Oral human papillomavirus and human herpesvirus-8 infections among human immunodeficiency virus type 1-infected men and women in Italy. Sex Transm Dis. 2012;39(1):894–898.

[9] Souza VAUF, Sumita LM, Nascimento MC, et al. Human herpesvirus-8 infection and oral shedding in Amerindian and non-Amerindian populations in the Brazilian Amazon region. J Infect Dis. 2007;196:844–852.

[10] Qu L, Jenkins F, Triulzi DJ. Human herpesvirus 8 genomes and seroprevalence in United States blood donors. Transfusion. 2010;50:1050–1056.

[11] Levi JE, Nascimento MC, Sumita LM, et al. Non-detection of human herpesvirus 8 (HHV-8) DNA in HHV-8-seropositive blood donors from three Brazilian regions. PLoS One. 2011;6(8):e23546.

[12] Vamvakas EC. Relative risk of reducing the lifetime blood donation deferral for men who have had sex with men versus currently tolerated transfusion risks. Transfus Med Rev. 2011;25(1):47–60.

[13] Johnston C, Orem J, Okuku F, et al. Impact of HIV infection and Kaposi sarcoma on human herpesvirus-8 mucosal replication and dissemination in Uganda. PLoS One. 2009;4(1):e4222.

[14] Nascimento MC, De Souza VA, Sumita LM, et al. Comparative study of Kaposi’s sarcoma-associated herpesvirus serological assays using clinically and serologically defined reference standards and latent class analysis. J Clin Microbiol. 2007;45(3):715–720.

[15] Souza V, Sumita LM, Freire W, et al. Prevalence of antibodies to human herpesvirus-8 in populations with and without risk for infection in Sao Paulo State. Braz J Med Biol Res. 2004;37:123–127.

[16] Pierrotti LC, Etzel A, Sumita LM, et al. Human herpesvirus 8 (HHV-8) infection in HIV/AIDS patients from Santos, Brazil: seroprevalence and associated factors. Sex Transm Dis. 2005;32:57–63.

[17] Bender Ignacio RA, Goldman JD, Magaret AS, et al. Patterns of human herpesvirus-8 oral shedding among diverse cohorts of human herpesvirus-8 seropositive persons. Infect Agent Cancer. 2016;11:7.

[18] Tozetto-Mendoza TR, Sumita LM, Palmieri M, et al. No detectable human herpesvirus-8 oral shedding in seronegative-healthy, immunocompetent individuals from non-endemic regions for Kaposi’s sarcoma: A pilot study. J Invest Clin Dent. 2017;12278.
