Lipid Regulation Effects of Raw and Processed Notoginseng Radix Et Rhizome on Steatotic Hepatocyte L02 Cell

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Received 5 March 2016; Revised 25 May 2016; Accepted 16 June 2016

Academic Editor: Vanessa Souza-Mello

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Introduction. Raw and processed Notoginseng Radix Et Rhizome (NRR) have been widely used in treatment of metabolic syndromes and related disease, including nonalcoholic fatty liver disease (NAFLD). This study was designed to investigate lipid regulation effects of raw and processed NRR in steatotic L02 cell. Materials and Methods. Steatotic L02 cells were obtained after being cultured with 5% fat emulsion-10% FBS-RPMI 1640 medium for 48h. Contents of total cholesterol (TC), triglyceride (TG), free fatty acid (FFA), high-density lipoprotein cholesterol (HDL-C), and low-density lipoprotein cholesterol (LDL-C) in steatotic L02 cells were evaluated after treatment. Furthermore, the lipid metabolism regulation mechanism of Panax notoginseng saponins (PNS) and its monomers were evaluated by detecting the expressions of hydroxymethyl glutaric acyl coenzyme A reductase (HMG-CoAR), sterol regulating element binding protein-2 (SREBP-2), and cholesterol 7α-hydroxylase (CYP7α).

Results. TG and TC contents were doubled in model group compared to those in normal L02 cell group. Raw NRR and NRR heated with sand (NRR-B) showed much remarkable lipid-lowering effects in steatotic L02 cells. PNS, notoginsenoside R1, ginsenoside Rg1, and ginsenoside Rb1 displayed the best TG and TC regulation activity, which could significantly reduce contents of SREBP-2 and HMG-CoAR and increase the content of CYP7α.

Conclusions. Our results may support the fact that both raw NRR and NRR-B might have more satisfactory effects in the treatment of NAFLD.

1. Introduction

Nonalcoholic fatty liver disease (NAFLD) is a common health problem worldwide. As the data collected from all over the world shows, about one in four adults suffers from NAFLD. In western and Asian countries, prevalence of NAFLD is up to approximately 30% in the general population. The detection rate of NAFLD increased with age and body mass index (BMI) [1]. The prevalence of NAFLD was increased from 35.47% in 2006 to 46.46% in 2014 among 1948 subjects in China [2]. NAFLD, highly related to metabolic syndrome, is defined as the complex lipid metabolism dysfunction in hepatic cells. Hypertriglyceridemia (HTG), which may or may not be associated with hypercholesterolemia, is usually presented in NAFLD. Insulin resistance and inflammation are believed to play main roles in the pathogenesis and progression of NAFLD. Metabolic syndromes, which include obesity, type 2 diabetes mellitus, and hyperlipidemia, are risk factors of NAFLD [3].

Notoginseng Radix Et Rhizome (NRR), known as a valuable traditional Chinese medicine, is the dried roots of Panax notoginseng (Burk.) F. H. Chen in Araliaceae family. NRR can be used with its both raw and processed products. Several processing methods are recorded and applied in clinic use of NRR. NRR can be steamed solely [4, 5] or heated with sand or sesame oil [6, 7]. Different processed products of NRR have obvious differences in clinical practice application. Panax notoginseng has got a high evaluation in the treatment of hematric disease, such as coronary artery disease, angina, peptic ulcer, hyperlipidemia, and hypertension [8, 9]. Raw NRR could promote blood circulation for removing blood stasis, remove edema, and relieve pain [10]. It is considered as sovereign remedies for traumatism and first-line choice for hemostasis in traditional Chinese medicine. The steamed NRR (NRR-A) and NRR heated with sand (NRR-B) have better pharmacological effects in nourishing blood and improving the microcirculation [11]. NRR fried with sesame oil (NRR-C) plays a major role in enhancing memory.
retention and the immunity of the body [12]. In recent decades, raw and processed NRR are all used in the treatment of NAFLD and metabolic syndromes related disease [13]. The contents of Panax notoginseng saponin (PNS, which is one of the most important compounds extracted from roots of the herb Panax notoginseng) and its monomers, such as notoginsenoside R1, ginsenoside Rg1, ginsenoside Rb1, ginsenoside Re, and ginsenoside Rd, reduced in different degrees after processing [14, 15]. However, little is known about the activities and possible differences between raw and processed NRR on lipid regulation.

In this study, steatotic L02 hepatocyte of normal human was used to explore the lipid metabolism regulation effects of raw NRR and its processed products. Moreover, potential activities of PNS, notoginsenoside R1, ginsenoside Rg1, ginsenoside Rb1, ginsenoside Rd, and ginsenoside Re were also investigated.

2. Materials and Methods

2.1. Main Reagents. PNS (the contents of notoginsenoside R1, ginsenoside Rg1, ginsenoside Rb1, ginsenoside Rd, and ginsenoside Re are 6.9%, 28.0%, 29.7%, 73.%, and 3.8%, resp.), notoginsenoside R1, ginsenoside Rg1, ginsenoside Rb1, ginsenoside Rd, and ginsenoside Re were purchased from the National Institute for the Control of Pharmaceutical and Biological Products, China. Lovastatin (Jiangsu Ocean Pharmaceutical Co., Ltd., China) and fenofibrate (Laboratoires Fournier S.A., France) were used as positive control for lowering cholesterol and triglyceride, respectively. Fetal bovine serum (FBS) and penicillin-streptomycin were purchased from Fisher Scientific International Inc., USA. Fat emulsion was purchased from Sichuan Koren Pharmaceutical Co., Ltd., China.

2.2. Processing Procedures of NRR. The Notoginseng Radix Et Rhizome was collected in Wenshan County of Yunnan Province by the authors in September 2012 and identified as the root and rhizome of P. notoginseng (Burk.) F. H. Chen by Professor Xiaojian Mao, Yunnan University of Traditional Chinese Medicine. Voucher specimens were deposited in the Herbarium of Pharmacognosy, Yunnan University of Traditional Chinese Medicine. NRR-A was steamed from the root and rhizome of Panax notoginseng of Chinese Academy of Sciences. The cell was maintained in an incubator (MCO-20AIC CO2 Incubator, Japan’s Sanyo Electric Co., Ltd.) with 5% carbon dioxide, 95% air, at 37°C in RPMI-1640 medium (Gibco Invitrogen Corporation, USA) supplemented with 10% fetal bovine serum (FBS, Hyclone). 0.25% trypsin (1–2 mL) (Amresco, USA) was used to passage cells at 80–90% confluence.

2.3. Determination of Notoginsenoside R1, Ginsenoside Rg1, Ginsenoside Rb1, Ginsenoside Rd, and Ginsenoside Re Contents in the Raw and Processed NRR by HPLC. In order to illustrate the chemical profiles of raw and processed NRR, concentrations of notoginsenoside R1, ginsenoside Rg1, ginsenoside Rb1, ginsenoside Re, and ginsenoside Rd were detected by HPLC. Sample solutions were prepared following the procedures developed in our previous studies [14]. 0.6 g powder of NRR, NRR-A, NRR-B, and NRR-C was precisely weighed and refluxed with 50 mL 70% methanol for 2 h. These solutions were filtered through 0.45 μm membrane and 10 μL was injected into the HPLC system, respectively.

All experiments were performed with Agilent Zorbax SB-C18 (4.6 mm × 250 mm, 5 μm). The mobile phase consisted of acetonitrile (A) and water (B) as gradient elution for separation. The following gradient was used: 0–21 min, 20% A; 21–30 min, 20–25% A; 30–41 min, 25–41% A; 41–51 min, 41–45% A; 51–56 min, 45–20% A; and 56–66 min, 20% A. The column temperature was maintained at 20°C. The analysis was conducted at a flow rate of 1.0 mL/min and detected at 203 nm.

2.4. Preparation of Extractions of Raw and Processed NRR. 10 g powdered samples of raw and processed NRR were refluxed with 100 mL 50% ethanol for three times, 1 hour for each time, separately. The extracts were combined, condensed, and lyophilized. The extraction rates of NRR, NRR-A, NRR-B, and NRR-C were 18.0%, 16.5%, 19.0%, and 16.0% crude drug, respectively.

2.5. Culture Conditions and Treatment of Cells [17]. The whole research was carried out by human normal hepatocyte cell line L02, which was purchased from Kunming Institute of Zoology, Chinese Academy of Sciences. The cell was maintained in an incubator (MCO-20AIC CO2 Incubator, Japan’s Sanyo Electric Co., Ltd.) with 5% carbon dioxide, 95% air, at 37°C in RPMI-1640 medium (Gibco Invitrogen Corporation, USA) supplemented with 10% fetal bovine serum (FBS, Hyclone). 0.25% trypsin (1–2 mL) (Amresco, USA) was used to passage cells at 80–90% confluence.

L02 cells were inoculated into 6-well plates at a cell density of 3 × 10^4 and then incubated for 48 h till 80–90% confluences with 10% FBS-RPMI 1640 medium. Cells were incubated in G0 medium (0.2% FBS-RPMI 1640 medium) for 24 h to induce the cell cycle synchronous before the test. In the subsequent experiments, the control group was incubated with 10% FBS-RPMI 1640 medium alone, while the model group was incubated with 5% fat emulsion-10% FBS-RPMI 1640 medium for 48 h, respectively. After 48 h, cell morphological changes were investigated by inversion fluorescence microscope (ECLIPSE TS100, Nikon Corporation, Japan).

2.6. Preparation of Treatment Solutions. The stock solutions of raw and processed NRR extracts were achieved by dissolving them in dimethylsulfoxide (DMSO, Sigma, USA) at a concentration of 10 mg/mL. Then they were further diluted with G0 medium (0.2% FBS-RPMI 1640 medium) to grade concentrations of 10 μg/mL, 20 μg/mL, 40 μg/mL, 80 μg/mL, and 100 μg/mL, respectively.

Similarly, the Panax notoginseng saponins were diluted with G0 medium to grade concentrations of 10 μg/mL,
20 μg/mL, and 40 μg/mL, respectively. Meanwhile, lovastatin and fenofibrate were diluted with G0 medium to make stock solutions at a concentration of 10 μg/mL.

2.7. Lipid Metabolism Regulation Effects. Steatotic hepatocyte L02 cells were treated with the extracts of NRR, NRR-A, NRR-B, and NRR-C at the concentrations of 10, 20, 40, 80, and 100 μg/mL for another 24 h to evaluate their lipid metabolism regulation activity. 24 h after exposure, the cells were washed with PBS for 2 times. Then, the cells were trypsinized, collected, and resuspended with PBS and centrifuged to separate PBS from cells. Cells debris were lysed by 0.01% Triton X-100 (Sigma, USA) solution and then centrifuged at 5,000 rpm (Allegra X-22 Centrifuge) (Beckman Coulter, USA) for 10 minutes. TG and TC contents in the supernatant were examined using assay kits purchased from the Biosino Bio-technology & Science Inc. The contents of HDL-C and LDL-C in the supernatant were tested by assay kits purchased from the Changchun Sunostik Medical Technology Co., Ltd., China. FFA contents in the supernatant were tested by assay kits purchased from the Nanjing technology Co., Ltd., China. All these assays were carried out by DNM-9602G enzyme microplate reader purchased from the Beijing Pulang New Technology Co., Ltd., China.

Cells were lysed and collected under the above conditions. The contents of TG and TC in PNS, Rg1, Rb1, Rd, Re, and R1 treated groups at the concentrations of 10, 20, and 40 μg/mL were measured by DNM-9602G enzyme microplate reader.

In order to explore the mechanism of Panax notoginseng on lipid metabolism, the contents of cholesterol 7 alpha hydroxylase, HMG-CoAR, and SREBP-2 in PNS, R1, Rg1, and Rb1 treated groups were tested by ELISA assay kits purchased from Wuhan biological engineering Co., Ltd., China.

2.8. Statistical Analysis. Statistical analysis was performed using SPSS version 17. The quantitative data were given as the mean ± SD. The data of multiple groups were compared with one-way analysis of variance. p values < 0.05 were considered significant.

3. Results

3.1. The Contents of Triterpenoid Saponins in Raw and Processed NRR. The percentages of notoginsenoside R1, ginsenoside Rg1, ginsenoside Rb1, ginsenoside Re, and ginsenoside Rd in raw and processed NRR were listed in Table 1. Most of the saponins contents reduced after processing. The contents of ginsenoside Rg1, ginsenoside Rb1, ginsenoside Re, and ginsenoside Rd in raw and processed NRR were listed in Table 1. However, percentage of notoginsenoside R1 was increased in NRR-C.

3.2. Fat Emulsion Incubation Induced Cellular Steatotic. A model of steatotic hepatocyte was successfully established by the incubation with 5% fat emulsion-10% FBS-RPMI 1640 medium in human liver L02 cell line. Morphological changes indicated that lipid droplets (LDs) were obviously accumulated in the model group. Cell swelling, characteristic of cell
Table 1: The percentage of notoginsenoside R₁, ginsenoside Rg₁, ginsenoside Re, ginsenoside Rb₁, and ginsenoside Rd in Notoginseng Radix Et Rhizome and its processed products.

| Sample | Notoginsenoside R₁ | Ginsenoside Rg₁ | Ginsenoside Re | Ginsenoside Rb₁ | Ginsenoside Rd |
|--------|-------------------|----------------|---------------|----------------|----------------|
| NRR    | 0.98              | 3.27           | 0.38          | 2.72           | 0.66           |
| NRR-A  | 0.79              | 2.69           | 0.33          | 2.49           | 0.61           |
| NRR-B  | 0.30              | 1.15           | 0.11          | 1.00           | 0.24           |
| NRR-C  | 1.09              | 2.56           | 0.18          | 2.44           | 0.57           |

The contents of notoginsenoside R₁, ginsenoside Rg₁, ginsenoside Re, ginsenoside Rb₁, and ginsenoside Rd were analysed by HPLC as described in the text. NRR, Notoginseng Radix Et Rhizome; NRR-A, steamed NRR; NRR-B, NRR heated with sand; NRR-C, NRR fried with sesame oil.

Table 2: Contents of TG in raw and processed NRR treated steatotic L02 cells.

| Sample       | 10 µg/mL | 20 µg/mL | 40 µg/mL | 80 µg/mL | 100 µg/mL |
|--------------|----------|----------|----------|----------|-----------|
| NRR          | 0.40 ± 0.03### | 0.41 ± 0.06### | 0.48 ± 0.05### | 0.51 ± 0.00### | 0.36 ± 0.03### |
| NRR-A        | 0.42 ± 0.06### | 0.38 ± 0.01### | 0.45 ± 0.02### | 0.44 ± 0.01### | 0.50 ± 0.03### |
| NRR-B        | 0.44 ± 0.01### | 0.41 ± 0.01### | 0.39 ± 0.05### | 0.39 ± 0.03### | 0.38 ± 0.05### |
| NRR-C        | 0.44 ± 0.01### | 0.48 ± 0.07### | 0.42 ± 0.04### | 0.55 ± 0.01### | 0.50 ± 0.02### |
| Control group| 0.25 ± 0.01##  |          |          |          |           |
| Model group  | 0.53 ± 0.01*** |          |          |          |           |
| Lovastatin   | 0.31 ± 0.01### |          |          |          |           |
| Fenofibrate  | 0.30 ± 0.01### |          |          |          |           |

Triglyceride (TG) contents were assayed by assay kits as described in the text. Values were mean ± SD (n = 3) and expressed in mmol/L. NRR, Notoginseng Radix Et Rhizome; NRR-A, steamed NRR; NRR-B, NRR heated with sand; NRR-C, NRR fried with sesame oil. NRR and its processed products were dissolved in 50% ethanol. * indicates a significant difference compared with control group cells, and # indicates a significant difference compared with model group cells. ### p < 0.001. ## p < 0.01. ### p < 0.001.

Death and cell gap grew bigger than that of the control group. As the positive control, only slight dead cells were observed, with no significant accumulation of LDs. The intracellular TG contents were increased from 0.25 ± 0.01 mmol/L to 0.53 ± 0.01 mmol/L (p < 0.001) in steatotic L02 cells, while the intracellular contents of TC were increased from 0.13 ± 0.01 mmol/L to 0.23 ± 0.01 mmol/L (p < 0.001), which indicated the full and successful simulation of human NAFLD status.

3.3. TG, TC, HDL-C, LDL-C, and FFA Regulation Activity of Raw and Processed NRR. The TG, TC, HDL-C, LDL-C, and FFA contents in raw and processed NRR were shown in Tables 2, 3, 4, 5, and 6. Morphological observations showed that raw and processed NRR could reduce the accumulation of LDs at different levels, cells arranged closely, and intercellular space is small. The morphological changes of NRR-B were similar to the positive control groups. 50% ethanol extract of NRR and NRR-B showed better TG and TC regulation activity than NRR-A and NRR-C. Nevertheless, the effects of them were weaker than those of lovastatin and fenofibrate. 50% ethanol extract of NRR could reduce TC content by about 43% (p < 0.001), and 50% ethanol extract of NRR-B reduced TG content by about 24% (p < 0.001). HDL-C content was increased from 0.22 ± 0.03 mmol/L to 0.41 ± 0.02 mmol/L in 10 µg/mL NRR-B group (p < 0.01), which was similar to that of control group. 50% ethanol extract of NRR and NRR-B could reduce LDC-L content by 47% and 39% (p < 0.001), respectively. 50% ethanol extract of NRR showed more remarkable FFA regulation effects compared to its processed procedures, and the FFA content in 100 µg/mL NRR group was reduced from 762.89 ± 52.31 mmol/L to 386.59 ± 15.46 mmol/L in steatotic L02 cells (p < 0.001).

3.4. TG and TC Regulation Activities of Triterpenoid Saponins. PNS, including but not limited to notoginsenoside R₁, ginsenoside Rg₁, ginsenoside Rb₁, ginsenoside Re, and ginsenoside Rd, represented the main active ingredients of P. notoginseng. Changes in cell morphology indicated that PNS and notoginsenoside R₁ could significantly reduce the formation of LDs, and the arrangement of basal epithelial cells was tight. The TG-lowering and TC-lowering effects of PNS and related triterpenoid saponins were displayed in Figures 2(a) and 2(b). PNS, notoginsenoside R₁ and ginsenoside Rg₁, showed better TG-lowering activities than those of ginsenoside Rb₁, ginsenoside Re, and ginsenoside Rd. 40 µg/mL ginsenoside Rg₁ group could reduce the TG content from 0.53 ± 0.01 to 0.31 ± 0.01 mmol/L (p < 0.001), which was similar to that of lovastatin. PNS and notoginsenoside R₁ could reduce about 35% of TC content (p < 0.001), while ginsenoside Rg₁ and ginsenoside Rb₁ could reduce about 26% of TC content (p < 0.001).

3.5. The Mechanism of Panax notoginseng on Lipid Metabolism. Lipid-regulating effects are associated with the increasing of CYP7A and decreasing of SREBP-2 content, suggesting a possible antistatic mechanism of PNS, notoginsenoside R₁, ginsenoside Rg₁, and ginsenoside Rb₁. The
relative contents of CYP7α, HMG-CoAR, and SREBP-2 after treatments were shown in Figures 3(a), 3(b), and 3(c). Notoginsenoside R₁ at the concentration of 10 μg/mL showed great CYP7α increasing effect from 13.87 ± 0.77 to 78.73 ± 1.18 mmol/L (p < 0.001). PNS and notoginsenoside R₁ could relieve the steatotic state of liver by lowering the HMG-CoAR activity from 21.88 ± 0.23 to 12.91 ± 0.09 and 13.19 ± 0.33 mmol/L at the concentration of 10 μg/mL (p < 0.001), respectively. The SREBP-2 regulation activities of high concentration group of PNS and low and medium concentration groups of notoginsenoside R₁ showed a better effect than that of the positive group. They reduced nearly 65% of SREBP-2 content (p < 0.001). SREBP-2 lowering effects of ginsenoside Rg₁ and ginsenoside Rb₁ were weaker than those of other groups.

4. Discussion

TG and TC contents in hepatocytes are the defining characteristic of NAFLD. In addition, FFA has a direct effect on NAFLD when it esterified to TG [18]. LDL is in charge of transporting most of the lipid synthesized in the liver to the whole body via circulation system. On the contrary, HDL is in charge of transporting lipid back to the liver for biodegradation. Thus, it is important to point out that the levels of TG, TC, HDL-C, LDL-C, and FFA were the key factors for hyperlipidemia. All of them are considered as typical indicators in the diagnosis of NAFLD, hypertriglyceridemia, and hypercholesterolemia.

CYP7α and HMG-CoAR are key enzymes of biosynthesis and biodegradation of TC, while these procedures are regulated by SREBP-2. HMG-CoAR is the rate-limiting enzyme for cholesterol synthesis. Inhibiting its activity could reduce the LDL-C synthesis. CYP7α is the key rate-limiting enzyme for cholesterol transformation into bile acid in the liver, therefore promoting the excretion of cholesterol [19]. SREBP-2 is an important transcription factor, which could regulate fatty acid and cholesterol synthesis and impact the transcription of several genes involved in fatty acid and triglyceride metabolism [20].

In this research, L02 hepatocyte of normal human was exposed to 10% FBS and 5% fat emulsion for 48 hours as study objects. Steatotic human liver L02 cell was considered as a rapid and sensitive in vitro model to investigate the lipid regulation effects of raw and processed NRR. The contents of TG and TC in model group increased drastically compared to the control group (p < 0.001). The model can provide a reliable model for experimental study of pathogenesis of NAFLD and screening drugs for NAFLD treatment. Therefore, this steatotic model was frequently used in published research.
Table 5: Contents of LDL-C in raw and processed NRR treated steatotic L02 cells.

| Sample    | 10 μg/mL | 20 μg/mL | 40 μg/mL | 80 μg/mL | 100 μg/mL |
|-----------|----------|----------|----------|----------|-----------|
| NRR       | 1.48 ± 0.16*** | 1.43 ± 0.19*** | 1.51 ± 0.14*** | 1.33 ± 0.08*** | 1.56 ± 0.08*** |
| NRR-A     | 1.63 ± 0.12*** | 1.87 ± 0.02*** | 1.47 ± 0.22*** | 1.47 ± 0.13*** | 1.73 ± 0.13*** |
| NRR-B     | 1.39 ± 0.11*** | 1.27 ± 0.08*** | 1.32 ± 0.12*** | 1.40 ± 0.21*** | 1.21 ± 0.09*** |
| NRR-C     | 1.70 ± 0.11*** | 1.75 ± 0.19*** | 1.81 ± 0.19*** | 1.83 ± 0.20*** | 2.32 ± 0.06*** |
| Control group | 2.40 ± 0.12*** |            |          |          |           |
| Model group | 1.23 ± 0.10*** |            |          |          |           |
| Lovastatin | 1.73 ± 0.12*** |            |          |          |           |
| Fenofibrate| 1.54 ± 0.22*** |            |          |          |           |

Low-density lipoprotein cholesterol (LDL-C) contents were assayed by assay kits as described in the text. Values were mean ± SD (n = 3) and expressed in mmol/L. NRR, Notoginseng Radix Et Rhizome; NRR-A, steamed NRR; NRR-B, NRR heated with sand; NRR-C, NRR fried with sesame oil. NRR and its processed products were dissolved in 50% ethanol. * indicates a significant difference compared with control group cells, and # indicates a significant difference compared with model group cells.

**P < 0.01. ***P < 0.001. ****P < 0.01. **P < 0.01. ***P < 0.001.

Table 6: Contents of FFA in raw and processed NRR treated steatotic L02 cells.

| Sample    | 10 μg/mL | 20 μg/mL | 40 μg/mL | 80 μg/mL | 100 μg/L |
|-----------|----------|----------|----------|----------|----------|
| NRR       | 713.91 ± 69.25*** | 484.53 ± 25.77*** | 465.63 ± 41.60*** | 398.62 ± 47.67*** | 386.59 ± 15.46*** |
| NRR-A     | 577.32 ± 32.19** | 493.13 ± 82.53** | 513.75 ± 47.89** | 487.11 ± 27.57** | 505.15 ± 14.58** |
| NRR-B     | 539.52 ± 24.36*** | 613.4 ± 62.96*** | 637.46 ± 61.06*** | 525.77 ± 5.15*** | 436.43 ± 31.5*** |
| NRR-C     | 582.47 ± 96.57*** | 652.92 ± 12.97*** | 673.54 ± 39.03*** | 656.36 ± 75.99*** | 817.87 ± 56.78*** |
| Control group | 386.60 ± 5.15*** |            |          |          |           |
| Model group | 762.89 ± 52.31*** |            |          |          |           |
| Lovastatin | 680.41 ± 8.93*** |            |          |          |           |
| Fenofibrate| 639.18 ± 94.42** |            |          |          |           |

Free fatty acid (FFA) contents were assayed by assay kits as described in the text. Values were mean ± SD (n = 3) and expressed in mmol/L. NRR, Notoginseng Radix Et Rhizome; NRR-A, steamed NRR; NRR-B, NRR heated with sand; NRR-C, NRR fried with sesame oil. NRR and its processed products were dissolved in 50% ethanol. * indicates a significant difference compared with control group cells, and # indicates a significant difference compared with model group cells.

* P < 0.05. ** P < 0.01. *** P < 0.001. **** P < 0.01. ** P < 0.01. *** P < 0.001.

Figure 2: TG and TC contents in the steatotic L02 cells after treatment of triterpenoid saponins. Triglyceride (TG) (a) and total cholesterol (TC) (b) contents were assayed by assay kits as described in the text. Values were mean ± SD (n = 3) and expressed in mmol/L. NRR, Notoginseng Radix Et Rhizome; PNS, Panax notoginsenosides. * indicates a significant difference compared with control group cells, and # indicates a significant difference compared with model group cells. **P < 0.01. ***P < 0.001. ****P < 0.01. ***P < 0.001.
investigations by our research group [21] and other scientists [22].

Processing procedure changed both chemical constituents and pharmacological activities of NRR. Data suggest that 50% ethanol extracts of NRR and NRR-B displayed distinctive lipid-lowering effects in steatotic L02 cells. NRR-A showed ordinary lipid-lowering effects, while NRR-C fared the worst.

Saponins, dencichine, volatile oil, and polysaccharides are the main phytochemical constituents in NRR. Among them, saponins are considered to be one of the most important active ingredients. Notoginsenoside R₁, ginsenoside Rg₁,
and ginsenoside Rb₁ were high adaptive compositions
[23] among all saponins with distinctive TG-lowering and
TC-lowering activities. On the whole, notoginsenoside R₁,
ginsenoside Rg₁, and ginsenoside Rb₁ showed the best TG-
lowering activities similar to that of PNS in the concentration
of 40 μg/mL in steatotic L02 cells compared with other
triterpenoid saponins.

The contents of triterpenoid saponins decreased after
processing, possibly due to the high temperature during
processing procedure. Contents of triterpenoid saponins
decreased most significantly in NRR-B; however, NRR-B
showed the best activity on alleviating the fat denaturation,
and that is probably related to the generation of other new
constituents in the processing procedure [14]. Therefore, PNS,
R₁, Rg₁, and Rb₁, but not limited to them, were considered as
material foundations of *Panax notoginseng*.

Abnormal cholesterol level in steatotic L02 cells could be
associated with the cholesterol biosynthesis pathway and the
transformation-related protein expression. PNS, Rg₁, Rb₁,
and R₁ could inhibit cholesterol synthesis and reduce the
content of SREBP-2. PNS, Rg₁, Rb₁, and R₁ showed better
lipid-lowering activities by increasing the content of CYP7α
downregulated the TC content in steatotic L02 cells. The study of
*Panax notoginseng* on lipid-lowering and its mechanisms can provide new ideas for the further studies on
NAFLD.

In conclusion, raw NRR and NRR heated with sand show
better lipid regulation effects than other processing proce-
dures in the treatment of hyperlipidemia, NAFLD, and its
related diseases. PNS, notoginsenoside R₁, ginsenoside Rg₁,
ginsenoside Rb₁, and ginsenoside Rd may be the main active
ingredients to reduce TG and TC. The mechanisms of lipid-
lowering effect of PNS, notoginsenoside R₁, ginsenoside Rg₁,
and ginsenoside Rb₁ may be related to the downregulation of
HMG-CoAR and SREBP-2 and upregulation of CYP7α. Future studies focusing on animal experiments are still being
carried out by our research group in order to elucidate the
lipid regulation activities of raw and processed NRR in vivo.

Competing Interests

The authors declare that there are no competing interests
regarding the publication of this paper.

Acknowledgments

This research was financially supported by the National
Natural Science Foundation of China (Grant no. 81460623),
the Natural Science Foundation of Yunnan Province (Grant
no. 2014FA035), and the Southern Medicine Collaborative
And Innovation Center (30270100500).

References

[1] C. H. Lee and K. S. Lam, “Managing non-alcoholic fatty liver
disease in diabetes: challenges and opportunities,” *Journal of
Diabetes Investigation*, 2016.

[2] Z.-Y. Lu, Z. Shao, Y.-L. Li et al., “Prevalence of and risk factors
for non-alcoholic fatty liver disease in a Chinese population: an
8-year follow-up study,” *World Journal of Gastroenterology*, vol.
22, no. 13, pp. 3663–3669, 2016.

[3] Z. H. Li, Y. W. Chen, L. M. Xu et al., “The overview of
biological effects of galanin on major risk factors and pathology
of NAFLD,” *Chinese Journal of Gastroenterology and Hepatology*,
vol. 20, no. 11, pp. 1069–1072, 2011.

[4] X. Gai and B. Liu, “Improve the preparation of notoginseng,” *Qilu
Pharmaceutical Affairs*, vol. 24, no. 5, pp. 304–305, 2005.

[5] W. T. Xu and X. G. Wang, “Optimum processing technology
of Notoginseng Radix et Rhizoma was obtained by orthogonal
experiment,” *Shandong Journal of Traditional Chinese Medicine*,
vol. 31, no. 8, pp. 601–602, 2012.

[6] D. J. Ye and S. C. Zhang, *The Science of Processing Chinese
Materia Medica*, People’s Medical Publishing House, Beijing,
China, 1999.

[7] D. G. Tang, “The processing methods of frying in oil of *Panax
Notoginseng*,” *Chinese Traditional Patent Medicine*, vol. 13, no. 7,
pp. 18–19, 1991.

[8] Y. S. Liu, “The clinical application of Notoginseng Radix et
Rhizoma,” *Research and Practice of Chinese Medicines*, vol. 26,
no. 2, pp. 88–89, 2012.

[9] X. J. Liu, “The modern clinical application of Notoginseng
Radix et Rhizoma,” *Journal of Traditional Chinese Medicine*,
vol. 26, no. 11, pp. 2355–2356, 2011.

[10] Commission of Chinese Pharmacopoeia, *Chinese Pharma-
copia*, vol. 1, China Medico-Pharmaceutical Science & Technol-
ology Publishing House, Beijing, China, 2015.

[11] H. Y. Tian, X. Y. Qu, and P. H. Xiong, *Collection of Processing
Procedure of Ethnomedicines in China*, TCM Ancient Books
Publishing House, Beijing, China, 2000.

[12] B. Chen, H.-L. Koh, and X.-B. Jia, “Research progress and
thought on processing of *Panax notoginseng*,” *Chinese Tradi-
tional and Herbal Drugs*, vol. 44, no. 4, pp. 482–487, 2013.

[13] I. W. Joo, J. H. Ryu, and H. J. Oh, “The influence of Sam-Chil-Geun (*Panax notoginseng*) on the serum lipid levels and
inflammations of rats with hyperlipidemia induced by poloxamer-407,” *Yonsei Medical Journal*, vol. 51, no. 4, pp. 504–
510, 2010.

[14] X. H. Zhou, R. H. Zhao, R. P. Zhang et al., “Determination of
five saponins in different heat processed Notoginseng Radix et
Rhizoma,” *Journal of Yunnan University of Traditional Chinese
Medicine*, vol. 36, no. 6, pp. 11–14, 2013.

[15] J. P. Qin, G. Z. Zhang, B. B. Zhang et al., “Determining the
contents of saponins in Notoginseng Radix et Rhizome of
different processed products,” *Chinese Traditional and Herbal
Drug*, vol. 37, no. 8, pp. 1175–1177, 2006.

[16] Y. Jiang and Q. M. Huang, *Modern Research on Processing of
Traditional Chinese Medicine*, Sciences Press, Beijing, China,
2010.

[17] M. J. Wang, P. Lin, Y. F. Li et al., “Lipid regulation effects of raw
and processed Polygoni Multiflori Radix on steatotic human
liver cell line L02,” *World Phytomedicines*, vol. 28, no. 4, pp. 499–
499, 2013.

[18] D. Cupp, J. P. Kemp, and A. M. Kleinfeld, “Fatty acid-albumin
complexes and the determination of the transport of long chain
free fatty acids across membranes,” *Biochemistry*, vol. 43, no. 15,
pp. 4473–4481, 2004.

[19] R. D. Jones, A. M. Lopez, E. Y. Tong et al., “Impact of
physiological levels of chenodeoxycholic acid supplementation
on intestinal and hepatic bile acid and cholesterol metabolism
in *Cyp7a1*-deficient mice,” *Steroids*, vol. 93, pp. 87–95, 2015.
[20] L. J. Zhang, B. Yang, and B. P. Yu, “Paeoniflorin protects against nonalcoholic fatty liver disease induced by a high-fat diet in mice,” *Biological and Pharmaceutical Bulletin*, vol. 38, no. 7, pp. 1005–1011, 2015.

[21] M. Wang, R. Zhao, W. Wang, X. Mao, and J. Yu, “Lipid regulation effects of Polygoni Multiflori Radix, its processed products and its major substances on steatosis human liver cell line L02,” *Journal of Ethnopharmacology*, vol. 139, no. 1, pp. 287–293, 2012.

[22] J.-W. Wang, X.-Y. Wan, H.-T. Zhu et al., “Lipotoxic effect of p21 on free fatty acid-induced steatosis in L02 cells,” *PLoS ONE*, vol. 9, no. 4, Article ID e96124, 2014.

[23] Y. Wang, Y. Chu, W. Li, X.-H. Ma, and Z.-P. Wei, “Advances in study on saponins in *Panax notoginseng* and their pharmacological activities,” *Chinese Traditional and Herbal Drugs*, vol. 46, no. 9, pp. 1381–1392, 2015.