Leaky severe combined immunodeficiency in mice lacking non-homologous end joining factors XLF and Mrı

Sergio Castañeda-Zegarra\textsuperscript{1,2,†}, Qindong Zhang\textsuperscript{1,2,†}, Amin Alirezaylavasani\textsuperscript{1,2}, Valentyn Oksenych\textsuperscript{1,2,3,*}

\textsuperscript{1}Department of Clinical and Molecular Medicine (I KOM), Norwegian University of Science and technology, 7491 Trondheim, Norway;
\textsuperscript{2}St. Olavs Hospital, Trondheim University Hospital, Clinic of Medicine, Postboks 3250 Sluppen, 7006 Trondheim;
\textsuperscript{3}Department of Biosciences and Nutrition (BioNut), Karolinska Institutet, 14183 Huddinge, Sweden

\textsuperscript{†}These authors contributed equally

\textsuperscript{*}Correspondence: valentyn.oksenych@ntnu.no;

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Abstract

Non-homologous end joining (NHEJ) is a DNA repair pathway required to detect, process and ligate DNA double-stranded breaks (DSBs) throughout the cell cycle. The NHEJ pathway is required for the V(D)J recombination in developing B and T lymphocytes. During the NHEJ, the core factors Ku70 and Ku80 form a heterodimer called Ku, which recognizes DSBs and promotes the recruitment of accessory factors (e.g., PAXX, Mri, Dna-pkcs, Artemis) and downstream core factors XLF, XRCC4, and Lig4. Mutations in NHEJ genes result in immunodeficiency. Deletion of individual NHEJ genes Mri, Paxx or Xlf in mice resulted in no or modest phenotype, while combined inactivation of Xlf and Mri, or Xlf and Paxx, led to late embryonic lethality. Here, we demonstrated that the deletion of pro-apoptotic factor Trp53 rescued the embryonic lethality of mice with combined deficiencies in Xlf and Mri. The Xlf−/− Mri−/− Trp53+/− mice possessed reduced body weight, severely reduced B and T cell lymphocyte counts in spleen and thymus, and accumulation of progenitor B cells in the bone marrow. To the contrary, combined inactivation of Mri and Paxx resulted in live-born mice with phenotype not distinguishable from Mri-deficient mice. We concluded that Mri is functionally redundant with XLF during the B and T lymphocyte development in vivo.
1. Introduction

The non-homologous end-joining (NHEJ) is the DNA repair pathway that recognizes, processes and ligates DNA double-stranded breaks (DSB) throughout the cell cycle. The NHEJ is required for lymphocyte development, in particular, to repair DSBs induced by the recombination activating genes 1 and 2 (RAG) in developing B and T lymphocytes, and by activation-induced cytidine deaminase (AID) in mature B cells [1]. The NHEJ is initiated when core subunits Ku70 and Ku80 (Ku) are recruited to the DSB sites. Ku then mediates the recruitment of accessory NHEJ factors, including the DNA-dependent protein kinase catalytic subunit (DNA-PKcs) and nuclease Artemis. Additional core factors DNA ligase 4 (Lig4), X-ray repair cross-complementing protein 4 (XRCC4) and XRCC4-like factor (XLF) mediate DNA end ligation. Accessory factors parologue of XRCC4 and XLF (PAXX) and modulator of retroviral infection (Mri) stabilize the NHEJ complex [2, 3].

Inactivation of Ku70, Ku80, Dna-pkcs or Artemis results in severe combined immunodeficiency (SCID) characterized by a lack of mature B and T lymphocytes [4-8]. Deletion of Xrcc4 [9] or Lig4 [10] results in late embryonic lethality in mice that correlates with apoptosis in the central nervous system. Inactivation of the Xlf (Cernunnos) gene results in only modest immunodeficiency in mice [11, 12], while mice lacking Paxx [13-16] or Mri [2, 17] displayed no overt immunodeficiency phenotype.

Mild phenotype observed in mice lacking XLF was explained by functional redundancy between XLF and multiple DNA repair factors, including Ataxia telangiectasia mutated (ATM), histone H2AX [18], Mediator of DNA Damage Checkpoint 1 (MDC1) [19], p53-binding protein 1 (53BP1) [16, 20], RAG2 [21], DNA-PKcs [19, 22, 23], PAXX [3, 13, 14, 19, 24-26] and Mri [2]. Moreover, combined inactivation of Xlf and Paxx, as well as Xlf and Mri, resulted in late embryonic lethality in mice, which challenged studies of B and T lymphocyte development in vivo.

In this study, we rescued synthetic lethality between Xlf and Mri by inactivating one or two alleles of pro-apoptotic gene Trp53. Resulting Xlf−/−Mri−/−Trp53+/− mice possessed leaky SCID phenotype with nearly no B and T lymphocytes detected in spleens, nearly no mature T cells detected in thymi, and
accumulated progenitor B cells in the bone marrow. The $Xlf^{--}Paxx^{--}Trp53^{++}$ mice possessed a more severe phenotype with even lower levels of mature B cells detected in the spleen. Therefore, we concluded that both PAXX and Mri function in B and T lymphocyte development \textit{in vivo}, and their roles are compensated by XLF.

2. Materials and Methods

2.1. Mice

All experiments involving mice were performed according to the protocols approved by the Comparative Medicine Core Facility (CoMed) at the Norwegian University of Science and Technology (NTNU, Trondheim, Norway). $Xlf^{+/-}$ [11] and $Dna-pkcs^{+/-}$ [4] mice were imported from the laboratory of Professor Frederick W. Alt at Harvard Medical School. $Trp53^{++}$ mice [27] were imported from Jackson Laboratories. $Paxx^{+/-}$ [15] and $Mri^{+/-}$ [17] mice were generated on request for the Oksenych group and described previously.

2.2. Lymphocyte development

Lymphocyte populations were analyzed by flow cytometry as described earlier [15, 17, 18, 20]. Briefly, spleens, thymi, and femora were isolated from 5-7-week-old mice, the cells were lysed using red blood cell lysis buffer Hybri-Max™ (Sigma Aldrich, St. Louis, MO, USA; #R7757), resuspended either in PBS (Thermo Scientific, Basingstoke, UK; #BR0014G) or in 95% PBS-5% Fetal bovine serum, FCS (Sigma Life Science, St. Louis, Missouri, United States; Cat#F7524), and counted using Countess™ II Automated Cell Counter (Invitrogen, Carlsbad, CA, United States; #A27977). The cell suspension was spun down and diluted with PBS to get a final cell concentration of $2.5 \times 10^7$/mL. The cells were then incubated with fluorochrome-conjugated antibodies and analyzed by flow cytometry.

2.3. Antibodies
For the flow cytometry analyses, the following antibodies were used: rat anti-CD4-PE-Cy7 (BD Pharmingen™, Binningerstrasse, Allschwil, Switzerland, #552775, 1:100); rat anti-CD8-PE-Cy5 (BD Pharmingen™, Binningerstrasse, Allschwil, Switzerland, #553034, 1:100); anti-CD19-PE-Cy7 (Biolegend, San Diego, CA, USA, #115520, 1:100); hamster anti-mouse anti-CD3-FITC (BD Pharmingen™, Binningerstrasse, Allschwil, Switzerland, #561827, 1:100); rat anti-mouse anti-CD43-FITC (BD Pharmingen™, Binningerstrasse, Allschwil, Switzerland, #561856, 1:100); rat anti-mouse anti-CD45R/B220-APC (BD Pharmingen™, Binningerstrasse, Allschwil, Switzerland; #553092, 1:100); rat anti-mouse anti-IgM-PE-Cy7 (BD Pharmingen™, Binningerstrasse, Allschwil, Switzerland, #552867, 1:100).

2.4. Statistics

Statistical analyses were performed using one-way ANOVA, GraphPad Prism 8.0.1. 244 (San Diego, CA, USA). In all statistical tests, $p<0.05$ were taken to be significant (*, $p<0.05$; **, $p<0.01$; ***, $p<0.001$; ****, $p<0.0001$).

3. Results

3.1. Inactivation of the Trp53 gene rescued embryonic lethality in mice lacking XLF and Mri

Combined inactivation of Xlf and Mri resulted in synthetic lethality [2]. To rescue embryonic lethality in Xlf$^{−/−}$Mri$^{+/−}$ mice, we inactivated one allele of Trp53. For this, we intercrossed Mri$^{−/−}$ [17] and Xlf$^{−/−}$Trp53$^{−/−}$ [19] mice. In the next step, we selected and intercrossed triple heterozygous mice, Xlf$^{−/−}$Mri$^{−/−}$Trp53$^{+/−}$. By PCR screening, we identified Xlf$^{−/−}$Mri$^{−/−}$Trp53$^{+/−}$ (n=6), Xlf$^{−/−}$Mri$^{−/−}$Trp53$^{−/−}$ (n=2) and one Xlf$^{−/−}$Mri$^{−/−}$ mouse (Fig. 1). The mice lacking both XLF and Mri factors possessed reduced weight (12 g on average, $p<0.0001$) when compared with gender- and age-matched WT (19 g), Xlf$^{−/−}$ (19 g) and Mri$^{−/−}$ (20 g) controls. We used these mice to characterize the development of B and T lymphocytes (Fig. 1-4).
3.2. Leaky SCID in Xlf⁻/⁻Mri⁻/⁻Trp53⁺/⁺ mice

To determine the roles of Mri in lymphocyte development in vivo, we isolated thymi, spleens and femora from Xlf⁻/⁻Mri⁻/⁻Trp53⁺/⁺ mice, as well as from the Xlf⁻/⁺, Mri⁻/⁺ and WT controls. Combined deficiency for XLF and Mri resulted in 3-fold reduced sizes of thymi (32 mg, on average, p<0.0001) and 9-fold reduced counts of thymocytes (1.9x10⁷, p<0.0001), when compared with single deficient or WT controls (Fig. 1). The average weight of spleens (22 mg, p<0.0001), as well as a count of splenocytes (2x10⁷, p<0.0001) were about 4-5-fold reduced in 6-week-old Xlf⁻/⁻Mri⁻/⁻Trp53⁺/⁺ mice when compared with WT and single deficient controls (Fig. 1). The reduced number of splenocytes was due to decreased populations of B and T lymphocytes (Fig. 2). In particular, the reduction of CD3⁺ T cells count was 4 folds (p<0.0001), while the count of CD19⁺ B cells was 20 folds reduced (p<0.0001) in Xlf⁻/⁻Mri⁻/⁻Trp53⁺/⁺ mice when compared with single deficient and WT controls (Fig. 2). Counts of all CD4⁺, CD8⁺ and CD4⁺CD8⁺ T cells in thymi, as well as counts of CD4⁺ and CD8⁺ T cells in spleens, were dramatically reduced in Xlf⁻/⁻Mri⁻/⁻Trp53⁺/⁺ mice when compared with single deficient and wild type controls (about 4 folds, p<0.0001; Fig. 2). We concluded that XLF and Mri are functionally redundant during the B and T lymphocytes development in mice.

3.3. Leaky SCID in mice lacking XLF and PAXX

Combined inactivation of XLF and PAXX resulted in embryonic lethality in mice [3, 13, 14, 19]. To determine the impact of XLF and PAXX on B and T cell development in vivo, we rescued the synthetic lethality by inactivating one allele of Trp53 as described earlier [19]. Resulting Xlf⁻/⁻Paxx⁻/⁻ Trp53⁺/⁺ mice possessed 30-40 folds reduced counts of thymocytes (4x10⁶, p<0.0001) when compared with WT (1.31x10⁸), Xlf⁻/⁻ (1.36x10⁸) and Paxx⁻/⁻ (1.66x10⁸) mice (Fig. 2, 3) due to decreased levels of both double-positive CD4⁺CD8⁺ and single-positive CD4⁺ and CD8⁺ T cells (Fig. 2,3). Development of spleens in 6-week old mice lacking both XLF and PAXX was dramatically affected when compared with WT and single deficient controls due to the lack of B cells and decreased counts of T cells (Fig. 3). In particular, when compared with the WT and single knockout controls, in
Xlf<sup>−/−</sup>Paxx<sup>−/−</sup>Trp53<sup>−/+</sup> mice the counts of CD19+ B splenocytes were reduced 400-600 folds (0.1x10<sup>6</sup>, p<0.0001); and the counts of CD3+ T splenocytes were reduced 70-90 folds (to 0.3x10<sup>6</sup>). We concluded that XLF and PAXX are functionally redundant during the B and T lymphocyte development in mice.

3.4. Early B cell development is abrogated in mice lacking XLF and Mri, or XLF and PAXX

Reduced counts and proportions of mature B lymphocytes in Xlf<sup>−/−</sup>Mri<sup>−/−</sup>Trp53<sup>−/+</sup> mice suggested a block in B cell development in the bone marrow. We isolated the bone marrow cells from the femora of mice lacking XLF, Mri or both XLF/Mri and analyzed proportions of B220+CD43+IgM- progenitor B cells, B220+CD43+IgM- pre-B cells and B220+CD43+IgM+ immature and mature B cells. We detected only background levels of B220+CD43+IgM+ B cells in bone marrows isolated from Xlf<sup>−/−</sup>Mri<sup>−/−</sup>Trp53<sup>−/+</sup> mice (Fig. 4A,B). To the contrary, these mice possessed 2-3 fold increased proportions of pro-B cells when compared with WT, Mri<sup>−/−</sup> and Xlf<sup>−/−</sup> controls (Fig. 4A, C). Moreover, Xlf<sup>−/−</sup>Paxx<sup>−/−</sup>Trp53<sup>−/+</sup> mice possessed background levels of IgM+ B cells, and 3-4 folds increased proportions of pro-B cells when compared with WT, Paxx<sup>−/−</sup> and Xlf<sup>−/−</sup> controls (p<0.0001; Fig. 4A,C). We concluded that B cell development is blocked at pro-B and pre-B cell stages of both Xlf<sup>−/−</sup>Mri<sup>−/−</sup>Trp53<sup>−/+</sup> and Xlf<sup>−/−</sup>Paxx<sup>−/−</sup>Trp53<sup>−/+</sup> mice.

3.5. Normal development of Paxx<sup>−/−</sup>Mri<sup>−/−</sup> mice

Both PAXX and Mri are accessory NHEJ factors that are functionally redundant with XLF in mouse development. Combined inactivation of Paxx and Xlf [3, 13, 14, 19], or Mri and Xlf [2]; this study) genes resulted in synthetic lethality in mice, as well as in abrogated V(D)J recombination in vAbI pre-B cells [2, 3, 13, 14, 25]. To determine if Paxx genetically interacts with Mri, we intercrossed mice heterozygous or null for both genes (e.g., Paxx<sup>−/−</sup>Mri<sup>−/+</sup>). We found that resulting Paxx<sup>−/−</sup>Mri<sup>−/+</sup> double knockout mice are of nearly WT size (17g, p>0.9999), fertile, and possess normal development of B and T cells, indistinguishable from wild type and single deficient Paxx<sup>−/−</sup> and Mri<sup>−/+</sup> controls. In particular, Paxx<sup>−/−</sup>Mri<sup>−/+</sup> mice possessed normal counts of thymocytes and splenocytes,
including the CD4+, CD8+, and CD4+CD8+ T cells; and CD19+ B cells (not shown). We concluded that there is no genetic interaction between Paxx and Mri genes in vivo.

4. Discussion

Genetic inactivation of Xlf [11], Paxx [3, 13-15], or Mri [2, 28] in mice resulted in modest or no detectable phenotype. However, inactivation of other NHEJ factors resulted in the blockage of B and T cell development in mice, e.g., observed in Ku70−/− [5], Ku80−/− [6], Artemis−/− [7], Dna-pkcs−/− [4]; or embryonic lethality, e.g., Xrcc4−/− [9] and Lig4−/− [10]. Moreover, combined inactivation of Xlf and Mri (Xlf−/−Mri−/−) [2] or Xlf and Paxx (Xlf−/−Paxx−/−) [3, 13, 14], also resulted in embryonic lethality in mice, which correlated with increased levels of neuronal apoptosis in the central nervous system. Inactivation of one or two alleles of Trp53 rescued the embryonic lethality of the Xrcc4−/− [9, 29], Lig4−/− [10, 30], Xlf−/− Dna-pkcs−/− and Xlf−/− Paxx−/− [19; 23]. Findings during the last years suggest that Mri forms heterogeneous complexes containing PAXX or XLF, which function during DNA DSB repair by the NHEJ [2]. Combined inactivation of Xlf and Mri in vAbl pre-B cells resulted in a severe block in V(DJ) recombination and accumulation of unrepaired DNA double-strand breaks in vitro [2]. However, it was unclear whether the combined inactivation of Xlf and Mri will result in the lack of B and T lymphocytes in adult mice. Similarly, the double deficient vAbl pre-B cells lacking Xlf and Paxx were unable to maintain the V(DJ) recombination. However, it was not known whether the double-deficient mice lacking XLF and PAXX will possess any mature B cells. Importantly, the lack of progenitor T cell model system left the question on T cell development in Xlf−/−Mri−/− and Xlf−/−Paxx−/− mice completely unexplored.

Here, we demonstrated that mice lacking XLF, Mri and p53, although liveborn, possessed a leaky severe combined immunodeficiency phenotype. The Xlf−/−Mri−/−Trp53−/− mice, however, had a clear fraction of mature B cells in the spleen (CD19+) and bone marrow (B220+CD43−IgM+) (Fig. 2, 4), as well as clear fractions of double and single-positive T cells in the thymus (CD4+CD8+, CD4+, CD8+) and single-positive T cells in the spleen (CD4+ and CD8+) (Fig. 2). Similarly, the triple deficient mouse
model lacking XLF, PAXX, and p53 was liveborn and possessed only minor proportions of mature B cells in spleen and bone marrow, as well as only minor fractions of mature single positive T cells in the thymus and spleen (Fig. 3,4). Due to the presence of small but clear fractions of mature B and T cells in Xlf⁻/⁻Mri⁻/⁻Trp53⁺⁻ and Xlf⁻/⁻Paxx⁻/⁻Trp53⁺⁻ mice, we refer to the observed immunodeficient phenotypes as “leaky SCID”. Previously, leaky SCID was described in mice lacking other NHEJ factors, e.g. Ku70⁻/⁻ [5], Artemis⁻/⁻ [7], Lig4⁻/⁻Trp53⁺⁻, Xrc4⁻/⁻Trp53⁺⁻ [9, 29], Xlf⁻/⁻Atm⁻/⁻ [18] and Xlf⁻/⁻Rag2⁻/⁻ [21].

The Inactivation of Trp53 is not always sufficient to rescue the embryonic lethality in mice. For example, the mice deficient for the PLK1-interacting checkpoint helicase (PLIC) possess developmental defects in the presence or absence of p53 [31].

Moreover, we found that the mice with combined inactivation of Paxx and Mri (Paxx⁻⁻/⁻Mri⁻⁻) were live-born, fertile, and possessed normal B and T cell development. These data are further highlighting the previous observations using an in vitro model system. In particular, the vAbl pre-B cells with combined inactivation of Paxx and Mri genes showed as efficient V(D)J recombination as the single deficient Mri⁻⁻ and Paxx⁻⁻ and wild type controls [2]. Furthermore, combined inactivation of Paxx and Ku80 (Paxx⁻⁻/⁻Ku80⁻⁻), Atm (Paxx⁻⁻/⁻Atm⁻⁻) [14], and Dna-pkcs (Paxx⁻⁻/⁻Dna-pkcs⁻⁻) [19] showed a similar phenotype to the single deficient mice, Ku80⁻⁻, Atm⁻⁻ and Dna-pkcs⁻⁻, correspondently. Thus, both Paxx and Mri interact genetically with Xlf, but not with each other or different DNA repair factors tested so far.

In conclusion, we have developed two complex genetic models, Xlf⁻⁻/⁻Mri⁻⁻Trp53⁺⁻ and Xlf⁻⁻/⁻Paxx⁻⁻/⁻Trp53⁺⁻ and found that the B and T lymphocyte development is severely blocked in these mice at the progenitor stages.

Author contributions
All the authors designed the study, analyzed and interpreted the results. SCZ, QZ, and AL performed most of the experiments. VO wrote the paper with the help of SCZ and QZ.

**Conflict of interest statement**

The authors declare no conflict of interest.

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**Figure legends**

**Fig. 1. Growth and immune system development in Xlf^−/− Mri^−/− Trp53^+/− mice.** (A) The comparison of body size, thymus and spleen size between a Xlf^−/− Mri^−/− Trp53^+/− mouse and an Xlf^−/− mouse of the same age. (B) The weight of WT, Xlf^+/+, Mri^+/+, Xlf^−/− Mri^−/− Trp53^+/− mice. (C, D) The number (×10^6) of thymocytes (C) and splenocytes (D) in WT, Xlf^−/−, Mri^−/−, Xlf^−/− Mri^−/− Trp53^+/− mice. Comparisons between every two groups were made using one-way ANOVA, GraphPad Prism 8.0.1. *XMT=Xlf^−/− Mri^−/− Trp53^+/−

**Fig. 2. T and B cell development in Xlf^−/− Mri^−/− Trp53^+/− mice.** (A) Flow cytometric analysis of thymic and splenic T cell subsets and splenic CD19+ B cells. (B; C; D) The number (×10^6) of splenic CD19+ B cells (B), splenic CD3+ T cells (C) and thymic CD4+CD8+ double positive (DP) T cells in WT, Xlf^−/−, Mri^−/−, Xlf^−/− Mri^−/− Trp53^+/− mice. Comparisons between every two groups were made using one-way ANOVA, GraphPad Prism 8.0.1. *XMT=Xlf^−/− Mri^−/− Trp53^+/−; *XPT=Xlf^−/− Paxx^-/- Trp53^+/−; *DPK=Dna-pkcs^-/-
**Fig. 3.** T and B cell development in \(\text{Xlf}^{-/-}\ P\text{axx}^{-/-}\ \text{Trp53}^{+/+}\) mice. Examples of flow cytometric analysis of thymic and splenic T cell subsets and splenic CD19+ B cells.

**Fig. 4.** B cell development is abrogated in bone marrow of \(\text{Xlf}^{-/-}\ M\text{ri}^{-/-}\ \text{Trp53}^{+/+}\) and \(\text{Xlf}^{-/-}\ P\text{axx}^{-/-}\ \text{Trp53}^{+/+}\) mice. (A) Flow cytometric analysis of developing B cells. The upper left box marks progenitor B cell population, the lower left box marks the pre-B cells, and the lower right box marks the IgM+ immature and mature B cells. (B, C) The frequencies (%) of IgM+ B cells (B) and progenitor B cells (C) in WT, \(\text{Xlf}^{-/-}\), \(\text{Mri}^{-/-}\), \(\text{Xlf}^{-/-}\ \text{Mri}^{+/+}\ \text{Trp53}^{+/+}\), \(\text{Paxx}^{-/-}\), and \(\text{Xlf}^{+/+}\ \text{Paxx}^{-/-}\ \text{Trp53}^{+/+}\) mice. Comparisons between every two groups were made using one-way ANOVA, GraphPad Prism 8.0.1. *XMT=\(\text{Xlf}^{-/-}\ \text{Mri}^{+/+}\ \text{Trp53}^{+/+}\); *XPT=\(\text{Xlf}^{+/+}\ \text{Paxx}^{-/-}\ \text{Trp53}^{+/+}\).

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Figure 1

A

Mice

Xlf<sup>-/-</sup>/Mri<sup>+/+</sup>

5 cm

1 cm

1 cm

B

Mouse weight, g

***p<0.0001

WT Xlf<sup>−/−</sup> Mri<sup>+/+</sup> *XMT

C

Thymocyte count, 10<sup>6</sup>

***p<0.0001

WT Xlf<sup>−/−</sup> Mri<sup>+/+</sup> *XMT

D

Splenocyte count, 10<sup>6</sup>

***p<0.0001

WT Xlf<sup>−/−</sup> Mri<sup>+/+</sup> *XMT

*XMT=Xlf<sup>−/−</sup>/Mri<sup>+/+</sup>/Trp53<sup>−/−</sup>
Figure 2

A  

| Thymus   | Spleen     |
|----------|------------|
| WT       |            |
| 10       | 17         |
| 87       | 60         |
| CD19+ splenocytes,10^6 |
| Xlf^-/  |            |
| 8        | 20         |
| 88       | 39         |
| CD19+ splenocytes,10^6 |
| Mri^-/   |            |
| 9        | 15         |
| 87       | 62         |
| CD19+ splenocytes,10^6 |
| Xlf^-/ Mri^-/ Trp53^-/- | |
| 3        | 6          |
| 92       | 12         |
| 6        | 22         |
| CD19+ splenocytes,10^6 |

CD4  
CD8  
CD19  
CD3  

B  

| CD19+ splenocytes,10^6 |
|------------------------|
| WT                     |
| Xlf^-/                 |
| Mri^-/                 |
| Xlf^-/ Mri^-/ Trp53^-/- |

****p<0.0001  
****p<0.0001  
ns, p>0.9999  

C  

| CD3+ splenocytes,10^6 |
|------------------------|
| WT                     |
| Xlf^-/                 |
| Mri^-/                 |
| Xlf^-/ Mri^-/ Trp53^-/- |

****p<0.0001  
****p<0.0001  
ns, p>0.9999  

D  

| DP thymocytes,10^6 |
|--------------------|
| WT                 |
| Xlf^-/             |
| Mri^-/             |
| Xlf^-/ Mri^-/ Trp53^-/- |

****p<0.0001  
****p<0.0001  
ns, p>0.9999  

*XMT=Xlf^-/ Mri^-/ Trp53^-/-
*XPT=Xlf^-/ Paxx^-/ Trp53^-/-
*DPK=Dna-pkcs^-/-

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Figure 3

Thymus

WT

Xlf

Paxx

Xlf/Paxx

Trp53

Spleen

8

6

15

12

53

28

9

80

18

53

0.88

8

80

12

57

0.73

1

85

4

2

1

0.15

CD4

CD8

CD4

CD8

CD19

CD3
Figure 4

A

CD43

IgM

WT
Xlf/-
Mri/-
Xlf/-Mri/-Trp53+/

Paxx/-
Xlf/-Paxx/-Trp53+/

B

C

IgM+ B cells, %

B220+CD43+IgM- pro-B cells, %

****p<0.0001

****p<0.0001

****p<0.0001

****p<0.0001

*XMT=Xlf/-Mri/-Trp53+/
*XPT=Xlf/-Paxx/-Trp53+/

(Indian waxbill, Tricholaema megalops)