Review Article

MiRNAs in Gestational Diabetes Mellitus: Potential Mechanisms and Clinical Applications

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Gestational diabetes mellitus (GDM) is a common pregnancy complication which is normally diagnosed in the second trimester of gestation. With an increasing incidence, GDM poses a significant threat to maternal and offspring health. Therefore, we need a deeper understanding of GDM pathophysiology and novel investigation on the diagnosis and treatment for GDM. MicroRNAs (miRNAs), a class of endogenic small noncoding RNAs with a length of approximately 19-24 nucleotides, have been reported to exert their function in gene expression by binding to proteins or being enclosed in membranous vesicles, such as exosomes. Studies have investigated the roles of miRNAs in the pathophysiological mechanism of GDM and their potential as noninvasive biological candidates for the management of GDM, including diagnosis and treatment. This review is aimed at summarizing the pathophysiological significance of miRNAs in GDM development and their potential function in GDM clinical diagnosis and therapeutic approach. In this review, we summarized an integrated expressional profile and the pathophysiological significance of placental exosomes and associated miRNAs, as well as other plasma miRNAs such as exo-AT. Furthermore, we also discussed the practical application of exosomes in GDM postpartum outcomes and the potential function of several miRNAs as therapeutic target in the GDM pathological pathway, thus providing a novel clinical insight of these biological signatures into GDM therapeutic approach.

1. Overview of Gestational Diabetes Mellitus

Gestational diabetes mellitus (GDM) is a common maternal complication that occurs or is recognized during pregnancy. Since the pathophysiology of GDM is characterized by chronic insulin resistance in the second half of pregnancy, it is not diagnosed until the late second or early third trimester of gestation. A globally estimated prevalence of 1.8-31% has been reported due to the lack of consistency in GDM diagnostic criteria between countries [1]. GDM exerts various adverse implications for mothers and their offspring. Mothers complicated by GDM have higher rates of preeclampsia and adverse pregnancy outcomes, such as cesarean deliveries and shoulder dystocia [2]. They are more susceptible to developing postpartum type 2 diabetes mellitus (T2DM) compared with normal women [3]. Additionally, their offspring may suffer from long-term metabolic disorders and related health conditions such as obesity, T2DM, and cardiovascular disease (CVD) [4, 5]. According to a new set of diagnostic criteria published by IADPSG, pregnant women should perform an oral glucose tolerance test (OGTT) during 24-28 weeks of gestation [6]. However, compliance with the test may decline because it requires fasting and multiple blood types and may cause discomfort such as vomiting. However, OGTT is not recommended as a routine screening for GDM at an earlier trimester of gestation [6], and hence, treatment cannot be applied promptly for the prevention of GDM. Therefore, finding early predictors of GDM is significant to improve the prognosis of mothers and fetuses.

Moreover, since coronavirus disease 2019 (COVID-19) pneumonia pandemic-induced local lockdown measures have been carried out worldwide, their negative effects on psychosocial states of pregnant women and on the glycemic
balance in GDM patients have been observed. A review indicated negative implications of lockdowns and unhealthy lifestyle for pregnancy [7]. Solitude and mental burden such as anxiety and depression may lead to unhealthy dietary habits and reduced exercise. On the other hand, increased snack consumption and carbohydrate intake were revealed with a high glycemic index; increased total diet intake was found to be associated with a rise in HbA1c levels [8, 9] during the COVID-19 pandemic lockdown. A retrospective study conducted in France reported a lower postprandial glycemic control and a higher use of insulin therapy during quarantine (18 March–7 May 2020). These observations were explained by anxiety, reduced physical activity, and changes in diet [10]. These risk factors, coordinating with self-reported boredom/solitude and enhanced consumption of snacks, unhealthy foods, and sweets, have caused increased weight gain in some obese individuals [11]. A higher rate of GDM was observed in pregnant women during March–April 2020 compared with the same period in 2019 [12].

miRNAs, first discovered from C. elegans by Ambros and Ruvkun, represent small, short noncoding, and single-stranded RNA sequences consisting of approximately 22 nucleotides (nt) in length and act as negative regulators by inhibiting mRNA translation or leading to its degradation [13]. In most cases, miRNAs can also mediate posttranscriptional gene silencing by complementary binding to the target mRNA 3′-untranslated region (3′-UTR) or 5′-UTR or open reading frame (ORF) regions via their seed sequence region [14, 15]. Many animal model systems have been established to detect miRNAs, and their number is primarily associated with the organism’s complexity [16]. miRNAs present potential roles in the regulation of β-cell function and mass, as well as in metabolic processes [17]. The genome-wide analysis has demonstrated over 600 miRNAs expressed in placenta and their essential role in pregnancy and GDM [17–19]. Given the high stability of placental miRNAs in maternal circulation and their accessibility from maternal blood, they may become an early diagnostic biomarker of GDM [19]. Meanwhile, the role of a low glycemic or Mediterranean diet and particularly the favorable impact of plant-derived foods (e.g., vegetables, fibers, and fruits) on oxidative stress by enhancing antioxidant compounds has represented a new aspect in the pathogenesis of GDM [20]. Moreover, the correlation with miRNAs was not fully understood. This review is aimed at reporting updated literature in miRNA regarding to pathogenesis of GDM and the associated potential application.

2. The Biogenesis Pathway for miRNAs

During the process of miRNA biogenesis (shown in Figure 1), miRNAs located in intergenic regions and introns are transcribed by RNA polymerases II and III, from their promoter or cotranscribed with their own host gene or other miRNAs in the initial stage. The primary miRNA (pri-miRNA), an ~1000 nt capped and polyadenylated transcript, is known to contain a stem-loop structure in the nucleus [21]. The microprocessor complex subsequently crops this pri-miRNA to produce a precursor miRNA (pre-miRNA) with a length of 60 nt. The Exportin5-RanGTP system then exports this pre-miRNA to the cytoplasm for further processing. Eventually, the Dicer/TRBP complex cleaves the terminal loop of the pre-miRNA to create a miRNA duplex [21].

The remaining double-stranded RNA is loaded into a multiprotein complex called an RNA-induced silencing complex (RISC) and further unwinds in the center of RISC (an Argonaute protein) [21, 22]. During this process, the guide RNA strand from the miRNA duplex is selected as the mature miRNA, while the other passenger RNA strand is degraded. This guide strand remains in the RISC to form the miRNA-RISC complex as an essential component and serves to regulate gene expression epigenetically [23].

The miRNA-RISC has the capacity to regulate gene expression through base-pairing to the 3′-untranslated region (UTR), 5′-UTR, and protein-coding region of the messenger RNA (mRNA) target [13, 24]. The specific interaction between miRNAs and the target mRNA is primarily directed by the miRNA binding. This binding requires a certain number of nucleotides to match the sequence flanking the seed region [25]. The processes of the regulation in gene translation by miRNA-RISC are divided into two steps [26]: (i) the miRNA-RISC complex obstructs the binding between ribosomes and the mRNA target [27]; (ii) this consequently leads to mRNA degradation characterized by mRNA deadenylation and decapping, leading to accelerated destabilization and decay, thus suppressing translation of the target mRNA ultimately [28].

A single miRNA can target hundreds of mRNAs, and a specific target mRNA is often under the control of several distinct miRNAs. It has been established that miRNAs have potential function in many essential biological activities, such as cell proliferation, differentiation, apoptosis, disease initiation, and development [29–33]. Their dysregulation or dysfunction was revealed in many metabolic researches regarding obesity, T2DM, and cardiovascular disease. In addition, extracellular miRNAs are present in biological fluids such as plasma and are being packed into various carriers such as microvesicles (e.g. exosomes) or lipoproteins, rendering them a potential role as biomarkers or therapeutic targets [34].

3. miRNA Identification and Quantification Techniques

Several specific and sensitive approaches were applied to detect, validate, and quantify miRNAs, including quantitative reverse transcriptase PCR (RT-qPCR) [35, 36], in situ hybridization [37], Northern blot analysis [38, 39], miRNA microarray [40, 41], and next-generation sequencing (NGS) [42]. Deciding on the optimal miRNA profiling and quantification technology depends on the experimental designs, specific types of sample, research objective, and intended therapeutic use.

However, the expressions of several miRNAs in some findings we will review in detail further are not shared across each other. Different source materials such as serum or
plasma used during the detecting process or discrepancy in the analysis platform’s application might contribute to such differences [43]. Therefore, minimizing experimental variations through experimental normalization, data processing, and optimization is also significant for the precise evaluation of the level of miRNA from a specific sample [44].

Currently, quantitative reverse transcriptase PCR (RT-qPCR) is known as the gold-standard approach for miRNA quantification, which serves as the most reproducible and sensitive method [45–47]. Stem-loop RT-based TaqMan miRNA assay is widely used as the main PCR technique in research due to the advantage of high sensitivity and specificity [48]. Besides, direct RT-based and poly(A) tailing-based SYBR miRNA assays are considered as practical alternatives for miRNA detection and quantification [48]. A high-throughput qRT-PCR platform has been established as a more available approach for rapid miRNA profiling of a great quantity of biological samples. Some advances have been achieved in quantification using low amounts of miRNA [49]. TaqMan low density array (TLDA) possesses the advantage of cost-effectiveness and serves as the most widely used qRT-PCR miRNA expression profiling method [50].

Efforts have been made for the possibility of shortening the technique execution time as well as lowering amounts of miRNA used in quantification [51–53]. Microarrays represent a practicable discovery tool used for miRNA
identification on the basis of the principle in hybridization of cDNA to the DNA probe [54]. However, this technique is not quite promising for miRNA profiling, since they are not capable of detecting highly expressed miRNAs or distinguishing between mature and immature miRNAs [55, 56]. Moreover, several limitations related to this technique including low sensitivity, high requirement for RNA input amount (100 ng to 1 μg), background, and cross-hybridization still remain to be solved.

Another two alternatives for miRNA identification are also applied in research. In situ hybridization serves to contrast the level of miRNAs in different cells through utilizing radioactive, fluorescent, or dioxigen in probes [57]. It is noted that ISH also presents several disadvantages, including long processes, strenuous steps, and a higher rate of errors [57]. Additionally, next-generation sequencing technology (NGS) is a highly accurate technique with an advantage over other technologies, as it has the capability to identify novel miRNAs. Nevertheless, NGS is a more laborsome technique compared with qRT-PCR and microarrays and presents a higher requirement for RNA input amount (500 ng to 5 μg). Of note, high costs of this technique may contribute to a limitation of its wider availability [58].

Moreover, the most frequent normalization technique involves strategies using exogenous spike-in miRNA, such as C. elegans miR-39, which is validated to be more reliable compared with endogenous reference genes like miR-16 [59]. However, researchers prefer an application of combining both exogenous and endogenous miRNA reference genes, due to no ideal normalization strategy exists and the application of a single type of reference gene is insufficient for accurate miRNA results [60].

4. miRNA: The Role in Pathophysiology of GDM

4.1. miRNA-Related Maternal Metabolic Adaptation. In the past decade, people have been interested in the link of novel placenta-derived factors such as placenta-derived miRNA to pregnancy. More and more studies have explored the biological functions of placental-derived miRNA and their applicability as biomarkers in some pregnancy complications, such as GDM. Moreover, it is well established that an improper maternal metabolic adaptation to these placental-derived miRNAs has been observed [61, 62]. Therefore, variations in the expression of placental-related miRNA may indicate changes to maternal metabolic adaptation mechanism, thus providing insight into the pathogenesis of GDM pregnancy. Besides, variations in the expression of miRNAs in circulating samples may also indicate their involvement in maternal metabolic adaptation. Several studies have investigated the regulation of placenta-associated miRNA and circulating miRNAs as well as their related metabolic adaptation in GDM (Table 1).

Kokkinopoulou et al. first described a T2DM-specific expression profile of miRNAs that target disease-susceptibility genes, such as CDKN2A, CDK5, IGF2BP2, KCNQ1, and TSPAN8. miR-98-5p, one of miRNAs expressing decreased levels in T2DM patients compared with controls, was reported [63]. Moreover, miR-98 is also known to be implicated in embryo implantation during the initial stage of pregnancy. In 2016, Cao et al. showed a significant upregulation of miR-98 derived from placenta at gestation of 37–40 weeks in GDM patients (n = 193) compared to normal pregnant subjects (n = 202). Additionally, experimental validation in JEG-3 (human choriocarcinoma cell line) provided supportive evidence for its role in the regulation of glucose uptake. Specifically, by regulating MeCP2 and in turn targeting Trpc3, it has subsequent regulative effects on insulin-mediated glucose uptake in GDM [64]. This experimental evidence further confirmed the role of miR-98 in the development of GDM.

Zhao et al. reported a significantly upregulated concentration of miR-518d in the placenta of women affected by GDM compared with the normal subjects at 37-40 weeks of gestation. It is further proven that concentration of miR-518d in term placenta was negatively correlated with the expression of peroxisome proliferator-activated receptor-α (PPARα) [65]. PPAR plays a role in regulating the pathway related to inflammation, accidental formation, oxidative stress, and insulin signaling metabolism [66, 67]. The downregulation of the PPARγ expression in GDM may accelerate glucose intolerance [68]. Reduced expression of PPARα and RXRα were also found in the placenta of women with GDM [69].

In 2011, the same group demonstrated a significant downregulation of miR-132, miR-29a, and miR-222 in serum derived from GDM patients (n = 24) at gestation during the 16th and 19th weeks in comparison with healthy pregnant women (n = 24) [50]. Contrarily, a significant upregulation of miR-222, 1 of 17 differentially expressed miRNAs identified by Shi et al., was found in omental adipose tissues from GDM patients. By conducting a validation study in 10 GDM pregnant women compared with 10 healthy subjects of normal glucose tolerance, they further confirmed that the level of miR-222 was negatively correlated with the protein concentration of transporter glucose transporter 4 (GLUT4) in omental adipose tissue, as well as estrogen receptor-α (ER-α); the implication of the latter was validated in glucose homeostasis and insulin regulation [70–72]. Furthermore, they also validated the involvement of miR-222 in insulin resistance induced by estrogen in GDM through experiments performed on 3T3-L1 adipocytes by using antisense oligonucleotides [55].

Later on, Stirm et al. demonstrated a significant upregulation of miR-340 in whole blood cells (WBC) and lymphocytes from GDM women (n = 8) at 24–32 weeks of gestation, compared to healthy subjects (n = 8) [73]. A significant downregulation of polyadenylate-(poly(A))- binding protein- (PABP-) interacting protein 1 (PAIP1), known as a key promoter of translation that was never described in GDM before [74], was observed only in WBCs in GDM women, in comparison with normal glucose tolerant (NGT) subjects. An inverse correlation between miR-340 and PAIP1 expression in lymphocytes was observed, indicating that miR-340 might negatively regulate PAIP1. They further conducted experiments and observed reduced expression of miR-340 in human lymphocytes cultured in high-glucose medium. After adding insulin to the high-
miRNA | Regulation | Stage of pregnancy | Source | Cell studied | Putative target | Related metabolic adaptation
---|---|---|---|---|---|---
miR-222 [55] | ↑ | 38-39 wk | Omental adipose tissue | 3T3-L1 cells | ER-α | ↑ estrogen induced insulin resistance
miR-98 [64] | ↑ | 37-40 wk | Placenta | JEG-3 cells | Mecp2, Trpc3 | ↓ insulin-mediated glucose uptake
miR-518d [65] | ↑ | 37-40 wk | Placenta | HEK-293 cells | PPARα | ↓ glucose intolerance
miR-340 [73] | ↑ | 24-32 wk | Whole blood cells | Lymphocytes | PAIP1 | ↑ maternal intolerance
miR-130b, miR-148a [75] | ↑ | Newborns | HUVECs | HUVECs & BeWo cells | AMPKα1 | ↓ glucose metabolism

4.2. miRNA-Related Maternal Pancreatic β-Cell Dysfunction.

The development of GDM may be attributed to the dysfunction of maternal pancreatic β-cell during the compensatory mechanism for insulin resistance. Recent studies have established a conceivable link between circulating miRNAs, placental miRNAs, and maternal pancreatic β-cell dysfunction in GDM (Table 2).

Feng et al. assessed the level of miRNAs in peripheral blood samples derived from 12 GDM pregnancies and 12 healthy pregnancies. miR-33a-5p was demonstrated to be significantly upregulated in GDM group with respect to the NGT group. Furthermore, the authors found a positive correlation between miR-33a-5p expression and blood glucose. Notably, overexpression or inhibition of miR-33a-5p performed on INS-1 cells was revealed to significantly inhibit or promote cell growth and insulin production under high glucose condition, respectively. miR-33a-5p was found to directly target its downstream gene ABCA1, and Inc-DANCR exerts as a sponge in the regulation of antagonizing the function of miR-33a-5p [79]. These results confirmed that the Inc-DANCR-miR-33a-5p-ABCA1 signaling pathway exerts a significant role in regulating the biological function of INS-1 cells.

Similarly, Sebastiani et al. evaluated the level of miR-330-3p and found its hyperexpression in the blood sample of 21 GDM pregnancies versus 10 normal pregnancies at 24–33 weeks of pregnancy using a highly standardized approach. Interestingly, circulating miR-330-3p expression was negatively associated with fasting insulin only in GDM patients. Furthermore, two age- and BMI-matched populations were distinguished by differential level of miR-330-3p that divided into high and low groups, respectively [80]. Moreover, overexpression of miR-330-3p was validated to target and downregulate key genes, such as E2F1, known as essential modulators in glucose-stimulated insulin secretion and β-cell maintenance, such as β-cell growth and proliferation [81, 82]. The authors thus postulated that the hyperexpression of miR-330-3p in the blood sample may be harmful for β-cell function and/or proliferation.

Oppositely, He et al. analyzed the expression of miR-494 in the blood sample from 20 pregnancies affected by GDM and 20 normal women [83]. A significant downregulation of miR-494 was found in GDM pregnancies compared to CTRLs and was negatively associated with blood glucose. Furthermore, overexpression of miR-494 enhanced insulin secretion, induced cell proliferation, and inhibited cell apoptosis, whereas miR-494 knockdown achieved the opposite results. miR-494 was revealed to directly target phosphatase and tensin homolog (PTEN), known to exert a crucial role in apoptosis, in pancreatic β-cells. Notably, downregulation of PTEN induced by siRNA rescued the impact brought by miR-494 knockdown on insulin secretion, cell proliferation, and apoptosis of pancreatic β-cells. In conclusion, the results underline implication of miR-494 in β-cell dysfunction of GDM.

Li et al. also reported a significant downregulation of miR-96 in placental tissue from 3 GDM pregnancies compared to 3 healthy pregnancies. In addition, miR-96 expression was also found inversely correlated with blood glucose. It is noted that the knockdown of miR-96 reduced insulin level, lowered cell viability, and increased apoptosis in INS-1 cells under high glucose condition. Interestingly, similar correlation between miRNA and blood glucose was also observed in GDM rats. Zhao et al. analyzed the miRNA-
miR-33a-5p [80] 
miR-330-3p [80] 
miR-494 
miR-96 [85] 
miR-221 [84]

Table 2: Studies investigating the regulation of miRNA and related maternal pancreatic β-cell dysfunction in GDM.

| miRNA     | Regulation | Stage of pregnancy | Source                          | Cell studied | Putative target | Related pancreatic β-cell dysfunction                  |
|-----------|------------|--------------------|---------------------------------|--------------|-----------------|--------------------------------------------------------|
| miR-33a-5p| ↑          | 24–28 wk           | Blood samples                   | INS-1, HEK293T cells | ABCA1           | cell growth, insulin production                         |
| miR-330-3p| ↑          | 24–33 wk           | Plasma samples                  | —            | E2F1, CDC42     | cell proliferation, insulin secretion                   |
| miR-494   | ↓          | —                  | Peripheral blood                | INS-1 cells  | PTEN            | insulin secretion, cell proliferation, cell apoptosis   |
| miR-96    | ↓          | —                  | Placental tissue                | INS-1, HEK293T cells | PAK1           | insulin secretion, cell viability                       |
| miR-221   | ↓          | —                  | Placental tissue of GDM rats    | INS-1 cells  | PAK1            | insulin secretion, cell proliferation, cell apoptosis   |

221 expression in placental tissues of GDM rats by the microarray. A downregulation of miRNA-221 was reported in GDM rats, and a negative correlation between the miRNA-221 level and the blood glucose level was demonstrated. Notably, knockdown of miRNA-221 lowered insulin production and increased apoptosis in INS-1 cells, while opposite results were observed in miRNA-221-overexpressed INS-1 cells. Of note, miRNA-221 and miR-96 were proven to directly target PAK1 in two researches, and these results suggested that the dysfunction of β-cell might be attributed to dysregulation of miRNA-221 and miR-96 with a subsequent effect through targeting PAK1 [84, 85].

5. miRNAs in Placental Function and Fetal Complication

We have reviewed the role of several placenta-associated and circulating miRNAs in maternal metabolic adaptation and pancreatic β-cell dysfunction. In addition, several studies have investigated the role of miRNAs in placental function, as well as GDM-related fetal complication of the next generation.

By using RNA sequence and qRT-PCR validation, Ding et al. confirmed several dysregulated miRNAs in the placenta derived from 8 GDM pregnancies versus 8 healthy subjects. These differentially expressed miRNAs were predicted to be involved in placenta morphology and development. Notably, miR-138-5p was selected for biological functional assay due to its significant overexpression in GDM. Its overexpression inhibited the proliferative and migratory ability of HTR-8/SVneo trophoblast cells. A specific target of miR-138-5p was TBL1X, an oncogene in the activation of the WNT/β-catenin signaling pathway. This pathway crucially participates in placental biological processes, such as proliferation, differentiation, and invasion [86–88]. Moreover, miR-138-5p was validated to target sirtuin 1 (SIRT1) [89]; although limited studies reported the association between SIRT1 and GDM, reliable data confirmed its implication in the inflammation and glucose metabolic pathway in human placenta. Mac-Marcjanek et al. conducted experiments to investigate SIRT1-dependent specific gene alteration in GDM pregnancies and identified four diabetes-relevant genes linked to metabolism, inflammation, and transporting functions in SIRT1-overexpressed leukocytes [90]. SIRT1 was also found increasingly expressed in GDM women exposed to hyperglycemia at one day postpartum [91]. However, other authors observed a reduced level of SIRT1 in fetal endothelial colony-forming cells (EFCFs) and HUVECs in GDM pregnancies [92, 93], suggesting that dysregulation of SIRTs may be related to fetal complication. These evidences suggested that miR-138-5p serves as a potential biomarker in GDM management.

Li et al. identified 29 differentially expressed placenta-derived miRNAs from 15 GDM pregnancies in respect to 15 normoglycemia subjects to investigate the alteration of miRNAs. By conducting a miRNA microarray and RT-qPCR analysis approach, they validated 9 dysregulated miRNAs (miR-508-3p, miR-27a, miR-9, miR-137, miR-92a, miR-33a, miR-30d, miR-362-5p, and miR-502-5p). Furthermore, these miRNAs were predicted to target key genes implicated in the EGFR/phosphatidylinositol 3-kinase (PI3K)/protein kinase B (AKT) signaling pathway [56]. Of note, it is well known that the insulin tyrosine kinase receptor could activate the PI3K/AKT pathway and promote glucose transporting by enhancing the delivery of intracellular GLUT4 to the cell surface. Specifically, miR-508-3p, one of the overexpressed miRNAs, was revealed to directly regulate PI3Kγγ, a reverse modulator of the epidermal growth factor receptor (EGFR). PI3Kγγ exerts an essential role in adequate placental development and fetal growth [94]. The upregulation of miR-508-3p was validated to repress the expression of PI3Kγγ and aberrantly activate the EGFR/PI3K/AKT signaling [56]. Thus, the dysregulation of miR-508-3p may potentially promote the development of macrosomia, a specific fetal complication related to GDM.

Floris et al. reported an upregulated expression of miR-101 in HUVEC cells from GDM (n = 22) compared to healthy subjects (n = 24) and confirmed its crucial role in endothelial function and angiogenesis [95]. Moreover, miR-101 was found to target enhancer of zester homolog 2 (EZH2) [95–100], which exhibited reduced concentration in its isoform and histone H3K27 trimethylation in cultured human umbilical vein endothelial cells (HUVECs) from a GDM-exposed fetus [101]. A negative correlation between miR-101 and EZH2 was reported in a feedback loop of epigenetic regulation, suggesting a decreased functionality in GDM placenta. The dysfunctionality of the GDM placenta...
may contribute to miR-101 upregulation and functional alterations observed in HUVECs, including cell apoptotic activities and angiogenic and migratory capacities [101]. However, the maintenance of the alteration in this pathway and associated adverse impact on generation’s health still remain unclear. Some metabolic disorders such as cardiovascular disease might emerge in their adulthood life.

Notably, miRNAs could also function as a protective mechanism. Diaz-Perez et al. discovered another two differentially expressed miRNAs in GDM placental tissue and revealed their potential role in placental pathophysiology. Specifically, upregulation of miR-221 and miR-222 was reported in human fetoplacental endothelial cells (fpEC) isolated from four GDM placenta during the third trimester compared to four CTRLs [102]. What is more, miR-221 and miR-222 were validated to negatively regulate ICAM1 protein, whose reduced concentration was observed in the fetoplacental endothelium derived from GDM [103, 104]. These miRNAs may lead to the downregulation of ICAM-1 and function as a protective mechanism against inflammation characterized by leucocyte transmigration from blood to placenta due to hyperglycemia during GDM [102].

6. Exosomes and miRNAs in GDM

Exosomes are known specifically as extracellular vesicles (EVs), with the characteristic of a bilayered lipid and ~50-150 nm in diameter, originating from the endosomal compartment and actively secreted by multiple cell types [105]. Recently, exosomal miRNAs and their involvement in gene expression are gaining increasing scientific attention, suggesting their potential role for regenerating new therapies [106]. Exosomal miRs can be derived from different biological fluids, such as saliva, serum, amniotic fluid, urine, and breast milk, and can be released from various cells into the extracellular space [107, 108]; such a characteristic renders them to be potential clinical biomarkers and even novel targets for therapeutic intervention.

Three modes of mechanisms have been reported in the protection of miRNAs from degradation [34, 109–113]. These mechanisms could guarantee intercellular communication of miRNAs and their stability as cargos when delivered to recipient cells, subsequently inducing expressional and functional response. Therefore, similar to the cell-to-cell contact-dependent signaling pattern, the capacity of circulating EVs in conveying information is also considered an essential way for intercellular communication [114].

It is widely acknowledged that placenta is tightly linked to alteration of metabolic status in pregnancy. It is considered that adverse placental condition might be mirrored by the miRNA expression profile in placenta-derived exosomes (PdEs). In this part, we will emphasize PdE’s contribution to the development of GDM and give our viewpoints for their application in GDM management.

6.1. Tissue-Derived Exosomes and Exosomal miRNAs in GDM

Rice et al. performed the pilot study to demonstrate an altered exosomal concentration in GDM pregnancy. They observed a significantly higher exosomal level in the plasma sample of GDM women compared to normal subjects. The results also revealed that a high D-glucose level promotes exosomes released from trophoblast cells during the first-trimester pregnancy, suggesting a correlation between high glucose and exosomal bioactivity, which is of clinical relevance in GDM pathophysiology [115]. Furthermore, these exosomes released from trophoblast cells were confirmed to induce the expression of cytokine mediators such as interleukin-8 (IL-8) and TNF-a by in vitro experiments conducted on human umbilical vein endothelial cells (HUVECs), suggesting that exosomes could regulate immune responses to maternal metabolic adaptation during pregnancy.

Another study conducted by Salomon et al. also investigated the profile of PdEs in plasma during pregnancy. A progressive increase in the amount of these PdEs was observed, and the profile of these PdEs released into peripheral circulation at the 6-week gestation was characterized by gestational age. Furthermore, Salomon et al. confirmed these results in a prospective cohort through comparing the gestational-age PdE profile in GDM maternal plasma to normal subjects [116]. Similarly, they also observed an altered release of proinflammatory cytokines from HUVECs when treated with these PdEs derived from GDM pregnant women [117]. A more recent study conducted by Nardi et al. also reported similar results, indicating such pregnancy-related alterations of circulating EVs might provide a first hint for their role in the regulation of immune response during pregnancy [118].

Nakahara et al. also reported total PdE exosomal alterations in a cohort study and revealed their association with gestational age and pregnancy outcome. They also found a significantly higher PdE level in GDM pregnancies and PE versus normal pregnancies. In addition, several significant risk factors for GDM, including glucose concentration, maternal body mass index (BMI), and fetal body weight, were strongly associated with the PdE concentration during pregnancy, indicating that PdEs may reflect maternal metabolic adaptation and diagnostic utility to predict adverse pregnancy outcomes at an early stage [119]. Similarly, Elfeky et al. revealed a significant correlation between exosome concentration in maternal circulation and maternal BMI. Specifically, maternal BMI was inversely correlated with the contribution of PdEs to the total exosomes across gestation. A stronger effect was observed in exosomes derived from women of higher BMI in respect to lean, suggesting a potential influence of exosomes on the maternal systemic inflammation during gestation [120]. This study established the exosomal variation could be attributed to maternal BMI.

The role of exosomes derived from adipose tissue (exo-AT) is less investigated in the pathogenesis of GDM. Another study indicates that these exo-AT can function as regulators in the placental glucose metabolism through communication with placenta tissues in GDM, making it a potential to become an effective target for therapeutic intervention to prevent consequences complicated by GDM such as fetal overgrowth [121, 122]. Recently, it has been established that exo-AT might promote insulin resistance (IR) and other obesity-related metabolic statuses in obesity. Novel findings provided the evidence for the pivotal role of
the dysregulated release of exo-AT in the onset and development of GDM in obese mothers [122].

Interestingly, PdEs can also function as regulators in the communication with other organs/tissues. Recent studies have identified several exosomal miRs and suggested their potential roles as biomarkers for myogenesis, nutrient metabolism, and muscle mass variation in pathophysiological conditions [123–126]. There may exist a potential link between placenta-specific exosomal miRNAs and skeletal muscle. Nair et al. assessed the concentration of exosomal miRNA in chorionic villi explants derived from 12 pregnancies complicated by GDM compared to 12 normal subjects using next-generation sequencing (NGS) [123]. They further revealed a dysregulated set of 27 placenta-specific exosomal miRNAs and further explored the concentration of several exosomal miRs, including miR-22-3p, miR-125a-3p, miR-197-3p, miR-99b-5p, and miR-224-5p. These specific miRNAs were selected for their differentially expressed patterns between GDM and CTRLs, as well as variation in a consistent pattern in skeletal muscle samples and in GDM maternal circulation. Of note, several differentially expressed miRNAs were predicted to target glucose metabolism-associated genes such as the PI3K/AKT signaling pathway, suggesting their involvement in skeletal muscle insulin sensitivity of GDM. Therefore, placenta-specific exosomal miRNAs might exert a crucial role in the interrelation between gestational tissues and skeletal muscle with subsequent possible effects on peripheral insulin resistance in GDM.

Therefore, such research for the role of exosomes as paracrine vectors might help discover useful research hypotheses and novel knowledge for deciphering GDM pathophysiology and generating valuable and accessible biomarkers for the diagnostics and prediction in GDM. In addition, as regards the dysregulation of miRNA expression which has been linked to the complication of pregnancy, exosomal content including miRNA could be profiled and discovered as biomarkers for GDM. However, their involvement in the pathophysiology of GDM still needs to be further investigated for diagnostic purposes and therapeutic intervention.

6.2. Exosomes and miRNAs in GDM Treatment. Exosomes can be potential candidates for effective and regenerative therapies, thus establishing a new therapeutic area in regard to postpartum outcomes of GDM mothers, such as stress urinary incontinence (SUI), a common pathological state observed in nearly 30% of postpartum women [127]. Likewise, therapies based on MSC-exosomes have also been explored and represent as a promising approach in the improvement of GDM-caused myopathy.

Notably, Ni et al. demonstrated that some functional and histological improvements were achieved in a SUI rodent model when treated with hADSCs-exosomes. Additionally, several proteins contained in hADSCs-exosomes were linked to some crucial pathways such as Wnt, PI3K-Akt, and Jak-STAT signaling pathways, which were potentially implicated in skeletal muscle and nerve regeneration [127].

Similarly, Liu et al. reported the capacity of hADSCs-exosomes in increasing type I collagen content through stimulating collagen synthesis and inhibiting collagen degradation in vaginal fibroblasts from SUI women and established promising evidence in the field of therapeutic strategy for treating SUI [128]. Experimental evidence further confirmed the role of exosomes released from fibroblasts of SUI women in regulating endothelial cell angiogenesis [129].

Importantly, it has been established that miRNAs could be a potential candidate for effective and personalized therapy of GDM due to the discovery that exosomes possess diverse functions, including therapeutic function in the GDM avenues.

Moreover, several studies have reported promising approaches in treating GDM. By using microarray analysis, Chen et al. identified differentially expressed genes and miRNAs involved in the regulation of flotillin2 (FLOT2). The results indicated a negative correlation and a target relationship between miR-351 and FLOT2. Specifically, they treated GDM mice with a series of mimic, inhibitor, and small interfering RNA to investigate the bioactivity of miR-351 in insulin resistance (IR), cell apoptosis in pancreatic tissues, and liver gluconeogenesis [130]. The results showed that an upregulation of miR-351 suppressed the expression of FLOT2 with subsequent effects on liver gluconeogenesis by downregulating the PI3K/AKT pathway in GDM mice. These results indicated that miR-351 serve to prevent GDM development, and miR-351 was identified as a therapeutic target in the intervention of GDM.

Another study conducted by Tang et al. explored the role of miR-335-5p on insulin resistance and pancreatic islet β-cell secretion via activation of the TGFβ signaling pathway by downregulating VASH1 expression in GDM mice. They observed that overexpression of miR-335-5p and inhibition of VASH1 might contribute to the downregulation of insulin and insulin release levels [131]. These findings provided evidence for the role of miR-335-5p in the development of insulin resistance and the inhibition of pancreatic islet β-cell through downregulating VASH1 and subsequently activating the TGF-β pathway in GDM mice, thus providing more clinical insight into the GDM treatment.

7. Discussion

Gestational diabetes mellitus is regarded as one of adverse pregnancy complications, presenting an increasing prevalence throughout the world. It may lead to maternal postpartum metabolic disorders, such as obesity and diabetes, and bring about adverse influence on later development of the offspring. Although GDM is well known as a common pregnancy complication, it could not be diagnosed until the late second trimester [6]. Hence, novel biological signatures for timely diagnosis and therapeutic intervention are of significance. Nowadays, early recognition, diagnostic criteria, and therapeutic targets related to GDM are of great interest and with controversies, for diversity exists in race, region, genetics, environmental factors, and diagnostic criteria for GDM [132–135].

It is demonstrated that lifestyle strategy initiated in the first trimester of pregnancy has been proven effective
[136–141], reinforcing the importance of exploring biomarkers in early pregnancy. More importantly, identifying novel and available biomarkers in an early pregnancy provides clinical value not only for GDM early diagnosis but also for the prevention of obstetric and maternal-fetal complications.

Our team has investigated thyroid hormone in early pregnancy and revealed a negative correlation between its level and GDM. A low FT4 level in early pregnancy was found to increase the risk for developing GDM [142]. More recently, our team has established an advanced ML model for the early prediction of GDM [143]. Through employing machine learning (ML) models of high accuracy, a clinically cost-effective 7-variable logistic regression (LR) model that achieved effective discriminate power (AUC = 0.77) was ultimately investigated. The results demonstrated that low body mass index (BMI) (≤17) was revealed as a risk factor for GDM. Meanwhile, total 3,3',5'-triiodothyronine (T3) and total thyroxin (T4) showed superiority over free T3 and free T4 in predicting GDM, respectively. Besides, a promising predictive value of lipoprotein was also validated (AUC = 0.66).

As a class of short noncoding RNAs, miRNAs have achieved rising attention in GDM pathophysiology and development. Moreover, miRNAs have also induced interest as mediators of tissue cross-talk, such as adipose tissue and skeletal tissues, in the development of GDM. Notably, apart from previous findings related to miRNA in adipose tissue, adipocyte-derived markers also include adiponectin and leptin [144–146]. Likewise, some other placenta-derived markers, such as follistatin-like-3 [147–149] and placental growth factor [150, 151], could also function as biochemical predictors. These evidences may indicate a potential link of miRNAs to these serum biological signatures, suggesting their capacity as regulators of gene expression at the epigenetic level. Theoretically, the capacity of miRNAs in epigenetic modifications from an early pregnant stage holds evidence for their specific use in predicting GDM. Therefore, further investigation on miRNAs’ changes in concentration and corresponding epigenetic alterations in various biological tissues should be carried out.

8. Conclusion

In conclusion, we reviewed miRNAs revealed in placental tissues and investigated their roles in metabolic adaptations (e.g., insulin resistance, pancreatic, and β-cell function), placental function, and fetal complication. We also reviewed plasma exosomes and molecular content involved in GDM etiology; these evidences help in elucidating GDM pathophysiological pathways. However, their clinically diagnostic and predictive value still needs further investigation. Although several miRs were detected in the first trimester of pregnancy, it is noted that sample collection for miRNA analysis in most studies reviewed were restricted to the late second trimester of gestation. There still exists a lack in the evidence for miRNAs. Therefore, further research is needed in the validation of miRNA profiles for the earlier prediction of GDM. We will conduct more research to establish the potentiality of miRNAs for their predicting value in the diagnosis of GDM later on.

Conflicts of Interest

The authors declare no conflict of interest.

Authors’ Contributions

Zhao-Nan Liu was responsible for conceptualization, PubMed search, and manuscript preparation. Ying Jiang was responsible for PubMed search and manuscript preparation. Xuan-Qi Liu was responsible for PubMed search and manuscript preparation. Meng-Meng Yang was responsible for review and editing and supervision and revision of the manuscript. Cheng Chen was responsible for manuscript review and editing. Bai-Hui Zhao was responsible for manuscript supervision and revision. He-Feng Huang was responsible for conceptualization and manuscript review, supervision, and revision. Qiong Luo was responsible for conceptualization and manuscript review, supervision, and revision. Zhao-Nan Liu, Ying Jiang, and Xuan-Qi Liu contributed equally to this work.

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References

[1] H. D. McIntyre, P. Catalano, C. Zhang, G. Desoye, E. R. Mathiesen, and P. Damm, “Gestational diabetes mellitus,” Nature Reviews. Disease Primers, vol. 5, no. 1, p. 48, 2019.
[2] B. E. Metzger, D. R. Coustan, and E. R. Trimble, “Hyperglycemia and adverse pregnancy outcomes,” Clinical Chemistry, vol. 65, no. 7, pp. 937–938, 2019.
[3] J. H. Moon, S. H. Kwak, and H. C. Jang, “Prevention of type 2 diabetes mellitus in women with previous gestational diabetes mellitus,” The Korean Journal of Internal Medicine, vol. 32, no. 1, pp. 26–41, 2017.
[4] T. D. Clausen, E. R. Mathiesen, T. Hansen et al., “Overweight and the metabolic syndrome in adult offspring of women with diet-treated gestational diabetes mellitus or type 1 diabetes,” The Journal of Clinical Endocrinology and Metabolism, vol. 94, no. 7, pp. 2464–2470, 2009.
[5] Y. Yu, O. A. Arah, Z. Liew et al., “Maternal diabetes during pregnancy and early onset of cardiovascular disease in offspring: population based cohort study with 40 years of follow-up,” BMJ, vol. 367, article i6398, 2019.
[6] International Association of Diabetes and Pregnancy Study Groups Consensus Panel, B. E. Metzger, S. G. Gabbe et al., “International Association of Diabetes and Pregnancy Study Groups recommendations on the diagnosis and classification of hyperglycemia in pregnancy,” Diabetes Care, vol. 33, no. 3, pp. 676–682, 2010.
[7] A. L. Fedullo, A. Schiattarella, M. Morlando et al., "Mediterranean diet for the prevention of gestational diabetes in the Covid-19 era: implications of IL-6 in diabetes," *International Journal of Molecular Sciences*, vol. 22, no. 3, p. 1213, 2021.

[8] R. Zufo, F. Castellana, R. Sardone et al., "Preliminary trajectories in dietary behaviors during the COVID-19 pandemic: a public health call to action to face obesity," *International Journal of Environmental Research and Public Health*, vol. 17, no. 19, p. 7073, 2020.

[9] C. Munekawa, Y. Hosomi, Y. Hashimoto et al., "The impact of the COVID-19 virus emergency, gestational diabetes mellitus: a retrospective study," *Diabetes & Metabolism*, vol. 47, no. 2, pp. 201–210, 2021.

[10] L. Ghesquiere, C. Garabedian, E. Drumez et al., "The complexity of miRNA-mediated regulation," *Cell Death and Differentiation*, vol. 22, no. 1, pp. 22–33, 2015.

[11] N. Justman, G. Shahak, O. Gutzeit et al., "MicroRNA inhibition of translation initiation in vitro by targeting the cap-binding complex eIF4E," *Science*, vol. 317, no. 5845, pp. 1764–1767, 2007.

[12] A. Elalouf, E. Hutzinger, T. Nishihara, J. Rehwinkel, M. Fauser, and E. Izaurralde, "Deadenylation is a widespread effect of miRNA regulation," *RNA*, vol. 15, no. 1, pp. 21–32, 2009.

[13] K. J. Png, N. Halberg, M. Yoshida, and S. F. Tavazoie, "A microRNA regulon that mediates endothelial recruitment and metastasis by cancer cells," *Nature*, vol. 481, no. 7380, pp. 190–194, 2011.

[14] H. E. Gee, C. Camps, F. M. Buffa et al., "miR-10b and breast cancer metastasis," *Nature*, vol. 455, no. 7216, pp. E8–E9, 2008.

[15] Y. Tay, J. Zhang, A. M. Thomson, B. Lim, and I. Rigoutsos, "MicroRNAs to Nanog, Oct4 and Sox2 coding regions modulate embryonic stem cell differentiation," *Nature*, vol. 455, no. 7216, pp. 1124–1128, 2008.

[16] J. Kota, R. R. Chivukula, K. A. O'Donnell et al., "Therapeutic microRNA delivery suppresses tumorigenesis in a murine liver cancer model," *Cell*, vol. 137, no. 6, pp. 1005–1017, 2009.

[17] L. Ma, J. Teruya-Feldstein, and R. A. Weinberg, "Tumour invasion and metastasis initiated by microRNA-10b in breast cancer," *Nature*, vol. 449, no. 7163, pp. 682–688, 2007.

[18] J. D. Arroyo, J. R. Chevillet, E. M. Kroh et al., "Argonaute2 complexes carry a population of circulating microRNAs independent of vesicles in human plasma," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 12, pp. 5003–5008, 2011.

[19] J. Wang, J. Chen, P. Chang et al., "MicroRNAs in plasma of pancreatic ductal adenocarcinoma patients as novel blood-based biomarkers of disease," *Cancer Prevention Research*, vol. 2, no. 9, pp. 807–813, 2009.

[20] A. J. Pratt and I. J. MacRae, "The RNA-induced silencing complex: a versatile gene-silencing machine," *The Journal of Biological Chemistry*, vol. 284, no. 27, pp. 17897–17901, 2009.

[21] A. E. Pasquinelli, "MicroRNAs and their targets: recognition, regulation and an emerging reciprocal relationship," *Nature Reviews Genetics*, vol. 13, no. 4, pp. 271–282, 2012.
C. Chen, R. Tan, L. Wong, R. Fekete, and J. Halsey, K. W. Witwer, Y. H. Yuan, B. Z. Chi, S. H. Wen, R. P. Liang, Z. M. Li, and J. D. J. Wang, P. L. Paris, J. Chen et al., H. Wang, R. Peng, J. Wang, Z. Qin, and L. Xue, Y. X. Chen, K. J. Huang, and K. X. Niu, J. M. Thomson, J. Parker, C. M. Perou, and S. M. Hammond, G. S. Pall and A. J. Hamilton, C. Zhao, J. Dong, T. Jiang et al., A. Hrustincova, H. Votavova, and M. Dostalova Merkerova, E. Varallyay, J. Burgyan, and Z. Havelda, Z. Shi, C. Zhao, X. Guo et al., A. Lendvai, M. J. Deutsch, T. Plosch, and R. Ensenauer, S. J. Holdsworth-Carson, R. Lim, A. Mitton et al., A. Garcia-Ocana, M. Javelle and M. C. P. Timmermans, S. Madadi and M. Soleimani, J. Li, L. Song, L. Zhou et al., S. J. Holdsworth-Carson, R. Lim, A. Mitton et al., A. Hrustincova, H. Votavova, and M. Dostalova Merkerova, E. Varallyay, J. Burgyan, and Z. Havelda, Z. Shi, C. Zhao, X. Guo et al., A. Lendvai, M. J. Deutsch, T. Plosch, and R. Ensenauer, S. J. Holdsworth-Carson, R. Lim, A. Mitton et al., A. Hrustincova, H. Votavova, and M. Dostalova Merkerova, E. Varallyay, J. Burgyan, and Z. Havelda, Z. Shi, C. Zhao, X. Guo et al., A. Lendvai, M. J. Deutsch, T. Plosch, and R. Ensenauer, S. J. Holdsworth-Carson, R. Lim, A. Mitton et al., A. Hrustincova, H. Votavova, and M. Dostalova Merkerova, E. Varallyay, J. Burgyan, and Z. Havelda, Z. Shi, C. Zhao, X. Guo et al., A. Lendvai, M. J. Deutsch, T. Plosch, and R. Ensenauer, S. J. Holdsworth-Carson, R. Lim, A. Mitton et al., A. Hrustincova, H. Votavova, and M. Dostalova Merkerova, E. Varallyay, J. Burgyan, and Z. Havelda, Z. Shi, C. Zhao, X. Guo et al., A. Lendvai, M. J. Deutsch, T. Plosch, and R. Ensenauer, S. J. Holdsworth-Carson, R. Lim, A. Mitton et al.
[71] A. Nadal, P. Alonso-Magdalena, S. Soriano, I. Quesada, and A. B. Rooper, “The pancreatic β-cell as a target of estrogens and xenoestrogens: Implications for blood glucose homeostasis and diabetes,” *Molecular and Cellular Endocrinology*, vol. 304, no. 1–2, pp. 63–68, 2009.

[72] P. Alonso-Magdalena, A. B. Rooper, M. P. Carrera et al., “Pancreatic insulin content regulation by the estrogen receptor ER alpha,” *PloS One*, vol. 3, no. 4, article e2069, 2008.

[73] L. Stirn, P. Huypens, S. Sass et al., “Maternal whole blood cell miRNA-340 is elevated in gestational diabetes and inversely regulated by glucose and insulin,” *Scientific Reports*, vol. 8, no. 1, 2018.

[74] Q. R. Wang, A. N. Han, L. Y. Chen et al., “Paip1 overexpression is involved in the progression of gastric cancer and predicts shorter survival of diagnosed patients,” *Oncotargets and Therapy*, vol. Volume 12, pp. 6565–6576, 2019.

[75] J. B. Tryggestad, A. Vishwanath, S. N. Jiang et al., “Influence of gestational diabetes mellitus on human umbilical vein endothelial cell miRNA,” *Clinical Science*, vol. 130, no. 21, pp. 1955–1967, 2016.

[76] D. S. Novikova, A. V. Garabazhiu, G. Melino, N. A. Barlev, and V. G. Tribulovich, “AMP-activated protein kinase: structure, function, and role in pathological processes,” *Biochemistry*, vol. 80, no. 2, pp. 127–144, 2015.

[77] K. E. Boyle, H. Hwang, R. C. Jannsen et al., “Gestational diabetes is characterized by reduced mitochondrial protein expression and altered calcium signaling proteins in skeletal muscle,” *PloS One*, vol. 9, no. 9, article e106872, 2014.

[78] S. Liong and M. Lappas, “Activation of AMPK improves inflammation and insulin resistance in adipose tissue and skeletal muscle from pregnant women,” *Journal of Physiology and Biochemistry*, vol. 71, no. 4, pp. 703–717, 2015.

[79] Y. Feng, X. Qu, Y. Chen et al., “MicroRNA-33a-5p sponges to inhibit pancreatic β-cell function in gestational diabetes mellitus LncRNA DANCR,” *Reproductive Biology and Endocrinology*, vol. 18, no. 1, p. 61, 2020.

[80] G. Sebastiani, E. Guarino, G. E. Greico et al., “Circulating microRNA (mirna) expression profiling in plasma of patients with gestational diabetes mellitus reveals upregulation of miRNA mir-330-3p,” *Frontiers in Endocrinology*, vol. 8, 2017.

[81] J. S. Annicotte, E. Blanchet, C. Chavey et al., “The CDK4-pRB-E2F1 pathway controls insulin secretion,” *Nature Cell Biology*, vol. 11, no. 8, pp. 1017–1023, 2009.

[82] Z. X. Wang, E. J. Oh, and D. C. Thurmond, “Glucose-stimulated Cdc42 Signaling Is Essential for the Second Phase of Insulin Secretion,” *The Journal of Biological Chemistry*, vol. 282, no. 13, pp. 9536–9546, 2007.

[83] Y. F. He, J. Bai, P. Liu et al., “mir-494 protects pancreatic β-cell function by targeting PTEN in gestational diabetes mellitus,” *EXCLI Journal*, vol. 16, pp. 1297–1307, 2017.

[84] H. Zhao and S. Tao, “miRNA-221 protects islet β cell function in gestational diabetes mellitus by targeting PAK1,” *Biochemical and Biophysical Research Communications*, vol. 520, no. 1, pp. 218–224, 2019.

[85] L. Li, S. Wang, H. Y. Li et al., “MicroRNA-96 protects pancreatic β-cell function by targeting PAK1 in gestational diabetes mellitus,” *BioFactors*, vol. 44, no. 6, pp. 539–547, 2018.

[86] R. Ding, F. Guo, Y. Zhang et al., “Integrated transcriptome sequencing analysis reveals role of miR-138-5p/ TBLIX in placenta from gestational diabetes mellitus,” *Cellular Physiology and Biochemistry*, vol. 51, no. 2, pp. 630–646, 2018.

[87] X. C. Zeng, F. Q. Liu, R. Yan et al., “Downregulation of miR-610 promotes proliferation and tumorigenicity and activates Wnt/β-catenin signaling in human hepatocellular carcinoma,” *Molecular Cancer*, vol. 13, no. 1, 2014.

[88] M. Knoller and J. Pollheimer, “Human placental trophoblast invasion and differentiation: a particular focus on Wnt signaling,” *Frontiers in Genetics*, vol. 4, p. 190, 2013.

[89] B. G. Luan and C. X. Sun, “mir-138-5p affects insulin resistance to regulate type 2 diabetes progression through inducing autophagy in HepG2 cells by regulating SIRT1,” *Nutrition Research*, vol. 59, pp. 90–98, 2018.

[90] K. Mac-Marcjanek, A. Zieniaki, M. Zurawski-Klis, K. Cypryk, L. Woziak, and M. Wojcik, “Expression profile of diabetes-related genes associated with leukocyte sirtuin 1 overexpression in gestational diabetes,” *International Journal of Molecular Sciences*, vol. 19, no. 12, p. 3826, 2018.

[91] S. Sultan, N. Alzahrani, and K. Al-Sakkaf, “The postpartum effect of maternal diabetes on the circulating levels of sirtuins and superoxide dismutase,” *FEBS Open Bio*, vol. 8, no. 2, pp. 256–263, 2018.

[92] M. Lappas, “Anti-inflammatory properties of sirtuin 6 in human umbilical vein endothelial cells,” *Mediators of Inflammation*, vol. 2012, 11 pages, 2012.

[93] J. Gui, A. Potthast, A. Rohrbach, K. Borns, A. M. Das, and F. von Versen-Hoynek, “Gestational diabetes induces alterations of sirtuins in fetal endothelial cells,” *Pediatric Research*, vol. 79, no. 5, pp. 788–798, 2016.

[94] E. E. Er, M. C. Mendoza, A. M. Mackey, L. E. Rameh, and J. Bennis, “AKT facilitates EGF trafficking and degradation by phosphorylating and activating PIKfyve,” *Science Signaling*, vol. 6, no. 279, 2013.

[95] M. Smits, S. E. Mir, R. J. A. Nilsson et al., “Down-regulation of miR-101 in endothelial cells promotes blood vessel formation through reduced repression of EZH2,” *PloS One*, vol. 6, no. 1, article e16282, 2011.

[96] S. Varambally, Q. Cao, R. S. Mani et al., “Genomic loss of microRNA-101 leads to overexpression of histone methyltransferase EZH2 in cancer,” *Science*, vol. 322, no. 5908, pp. 1695–1699, 2008.

[97] T. Mitic, A. Caporali, I. Floris et al., “EZH2 Modulates Angiogenesis In Vitro and in a Mouse Model of Limb Ischemia,” *Molecular Therapy*, vol. 23, no. 1, pp. 32–42, 2015.

[98] C. H. Lu, H. D. Han, L. S. Mangala et al., “Regulation of tumor angiogenesis by EZH2,” *Cancer Cell*, vol. 18, no. 2, pp. 185–197, 2010.

[99] F. Kottakis, C. Polytarchou, P. Foltopoulou, I. Sanidas, S. C. Kampranis, and P. N. Tsichlis, “FGF-2 regulates cell proliferation, migration, and angiogenesis through an NDY1/ KDM2B-miR-101-EZH2 pathway,” *Molecular Cell*, vol. 43, no. 2, pp. 285–298, 2011.

[100] H. Dreger, A. Ludwig, A. Weller et al., “Epigenetic regulation of cell adhesion and communication by enhancer of zeste homolog 2 in human endothelial cells,” *Hypertension*, vol. 60, no. 5, pp. 1176–1183, 2012.

[101] I. Floris, B. Descamps, A. Vardeu et al., “Gestational diabetes mellitus impairs fetal endothelial cell functions through a mechanism involving microRNA-101 and histone methyltransferase enhancer of zeste homolog-2,” *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 35, no. 3, pp. 664–674, 2015.
endothelium in GDM,” *Cell Adhesion & Migration*, vol. 10, no. 1-2, pp. 18–27, 2016.

[103] M. Duan, H. H. Yao, G. K. Hu, X. M. Chen, A. K. Lund, and S. Buch, “HIV Tat induces expression of ICAM-1 in HUVECs: implications for miR-221/-222 in HIV-associated cardiomyopathy,” *PLoS One*, vol. 8, no. 3, p. e60170, 2013.

[104] G. K. Hu, A. Y. Gong, J. Liu, R. Zhou, C. S. Deng, and X. M. Chen, “miR-221 suppresses ICAM-1 translation and regulates interferon-γ-induced ICAM-1 expression in human cholangiocytes,” *American Journal of Physiology-Gastrointestinal and Liver Physiology*, vol. 298, no. 4, pp. G542–G550, 2010.

[105] B. Bahkshandeh, M. A. Kamaledinn, and K. Aalishah, “A comprehensive review on exosomes and microvesicles as epigenetic factors,” *Current Stem Cell Research & Therapy*, vol. 12, no. 1, pp. 31–36, 2017.

[106] B. Peng, Y. M. Chen, and K. W. Leong, “MicroRNA delivery for regenerative medicine,” *Advanced Drug Delivery Reviews*, vol. 88, pp. 108–122, 2015.

[107] J. Rak and A. Guha, “Extracellular vesicles - vehicles that spread cancer genes,” *BioEssays*, vol. 34, no. 6, pp. 489–497, 2012.

[108] T. Matsumura, K. Sugimachi, H. Inuma et al., “Exosomal microRNA in serum is a novel biomarker of recurrence in human colorectal cancer,” *British Journal of Cancer*, vol. 113, no. 2, pp. 275–281, 2015.

[109] A. Gallo, M. Tandon, I. Alevizos, and G. G. Illei, “The majority of microRNAs detectable in serum and saliva is concentrated in exosomes,” *PLoS One*, vol. 7, no. 3, article e30679, 2012.

[110] A. Michael, S. D. Bajracharya, P. S. T. Yuen et al., “Exosomes from human saliva as a source of microRNA biomarkers,” *Oral Diseases*, vol. 16, no. 1, pp. 34–38, 2010.

[111] L. L. Lv, Y. H. Cao, D. Liu et al., “Isolation and quantification of microRNAs from urinary exosomes/microvesicles for biomarker discovery,” *International Journal of Biological Sciences*, vol. 9, no. 10, pp. 1021–1031, 2013.

[112] Q. Zhou, M. Z. Li, X. Y. Wang et al., “Immune-related microRNAs are abundant in breast milk exosomes,” *International Journal of Biological Sciences*, vol. 8, no. 1, pp. 118–123, 2012.

[113] Z. B. Hu, X. Chen, Y. Zhao et al., “Serum microRNA signatures identified in a genome-wide serum microRNA expression profiling predict survival of non-small-cell lung cancer,” *Journal of Clinical Oncology*, vol. 28, no. 10, pp. 1721–1726, 2010.

[114] T. Kuroiwa, E. G. Lee, C. L. Danning, G. G. Illei, I. B. McInnes, and D. T. Boumpas, “CD40 ligand-activated human monocytes amplify glomerular inflammatory responses through soluble and cell-to-cell contact-dependent mechanisms,” *Journal of Immunology*, vol. 163, no. 4, pp. 2168–2175, 1999.

[115] G. E. Rice, K. Scholz-Romer, E. Sweeney et al., “The effect of glucose on the release and bioactivity of exosomes from first trimester trophoblast cells,” *The Journal of Clinical Endocrinology and Metabolism*, vol. 100, no. 10, pp. E1280–E1288, 2015.

[116] C. Salomon, M. J. Torres, M. Kobayashi et al., “A gestational profile of placental exosomes in maternal plasma and their effects on endothelial cell migration,” *PLoS One*, vol. 9, no. 6, article e98667, 2014.

[117] C. Salomon, K. Scholz-Romer, S. Sarker et al., “Gestational diabetes mellitus is associated with changes in the concentration and bioactivity of placenta-derived exosomes in maternal circulation across gestation,” *Diabetes*, vol. 65, no. 3, pp. 598–609, 2016.

[118] F. D. Nardi, T. F. Michelon, J. Neumann et al., “High levels of circulating extracellular vesicles with altered expression and function during pregnancy,” *Immunobiology*, vol. 221, no. 7, pp. 753–760, 2016.

[119] A. Nakahara, O. Elfeky, C. Garvey, D. Guanzon, S. A. Longo, and C. Salomon, “Exosome profiles for normal and complicated pregnancies—a longitudinal study [3O],” *Obstetrics & Gynecology*, vol. 133, no. 1, p. 162, 2019.

[120] O. Elfeky, S. Longo, A. Lai, G. E. Rice, and C. Salomon, “Influence of maternal BMI on the exosomal profile during gestation and their role on maternal systemic inflammation,” *Placenta*, vol. 50, pp. 60–69, 2017.

[121] G. Mignot, S. Roux, C. Thery, E. Segura, and L. Zitvogel, “Prospects for exosomes in immunotherapy of cancer,” *Journal of Cellular and Molecular Medicine*, vol. 10, no. 2, pp. 376–388, 2006.

[122] N. Jayabalan, A. Lai, S. Nair et al., “Quantitative proteomics by SWATH-MS suggest an association between circulating exosomes and maternal metabolic changes in gestational diabetes mellitus,” *Proteomics*, vol. 19, no. 1-2, article 1800164, 2018.

[123] S. Nair, N. Jayabalan, D. Guanzon et al., “Human placental exosomes in gestational diabetes mellitus carry a specific set of miRNAs associated with skeletal muscle insulin sensitivity,” *Clinical Science*, vol. 132, no. 22, pp. 2451–2467, 2018.

[124] I. Guller and A. P. Russell, “MicroRNAs in skeletal muscle: their role and regulation in development, disease and function,” *Journal of Physiology*, vol. 588, no. 21, pp. 4075–4087, 2010.

[125] F. Catapano, J. Domingos, M. Perry et al., “Downregulation of miRNA-29, -23 and -21 in urine of Duchenne muscular dystrophy patients,” *Epigenomics*, vol. 10, no. 7, pp. 875–889, 2018.

[126] D. Cacchiarelli, T. Incitti, J. Martone et al., “miR-31 modulates dystrophin expression: new implications for Duchenne muscular dystrophy therapy,” *EMBO Reports*, vol. 12, no. 2, pp. 136–141, 2011.

[127] J. Ni, H. Li, Y. Zhou et al., “Therapeutic potential of human adipose-derived stem cell exosomes in stress urinary incontinence - an in vitro and in vivo study,” *Cellular Physiology and Biochemistry*, vol. 48, no. 4, pp. 1710–1722, 2018.

[128] X. C. Liu, S. W. Wang, S. H. Wu et al., “Exosomes secreted by adipose-derived mesenchymal stem cells regulate type I collagen metabolism in fibroblasts from women with stress urinary incontinence,” *Stem Cell Research & Therapy*, vol. 9, no. 1, p. 159, 2018.

[129] X. C. Liu, S. H. Wu, W. Y. Wang, Q. Hao, Z. D. Guo, and W. Z. Wang, “Regulatory effect of exosomes secreted by vaginal wall fibroblasts on angiogenesis in patients with stress urinary incontinence,” *Zhonghua Yi Xue Za Zhi*, vol. 99, no. 7, pp. 510–514, 2019.

[130] S. H. Chen, X. N. Liu, and Y. Peng, “MicroRNA-351 eases insulin resistance and liver gluconeogenesis via the PI3K/AKT pathway by inhibiting FLOT2 in mice of gestational diabetes mellitus,” *Journal of Cellular and Molecular Medicine*, vol. 23, no. 9, pp. 5895–5906, 2019.

[131] X. W. Tang and Q. X. Qin, “miR-335-5p induces insulin resistance and pancreatic islet β-cell secretion in gestational diabetes mellitus mice through VASH1-mediated TGF-β
signaling pathway,” *Journal of Cellular Physiology*, vol. 234, no. 5, pp. 6654–6666, 2019.

[132] A. Duran, S. Sáenz, M. J. Torrejón et al., “Introduction of IADPSG criteria for the screening and diagnosis of gestational diabetes mellitus results in improved pregnancy outcomes at a lower cost in a large cohort of pregnant women: the St. Carlos Gestational Diabetes Study,” *Diabetes Care*, vol. 37, no. 9, pp. 2442–2450, 2014.

[133] D. R. Coustan, L. P. Lowe, B. E. Metzger, and A. R. Dyer, “The Hyperglycemia and Adverse Pregnancy Outcome (HAPO) study: paving the way for new diagnostic criteria for gestational diabetes mellitus,” *American Journal of Obstetrics and Gynecology*, vol. 202, no. 6, pp. 654.e1–654.e6, 2010.

[134] H. Long and T. Cundy, “Establishing consensus in the diagnosis of gestational diabetes following HAPO: where do we stand?,” *Current Diabetes Reports*, vol. 13, no. 1, pp. 43–50, 2013.

[135] Y. Yang, Q. Li, Q. Wang, and X. Ma, “Thyroid antibodies and gestational diabetes mellitus: a meta-analysis,” *Fertility and Sterility*, vol. 104, no. 3, pp. 665–671.e3, 2015.

[136] S. B. Koivusalo, K. Rönö, M. M. Klemetti et al., “Gestational diabetes mellitus can be prevented by lifestyle intervention: the Finnish Gestational Diabetes Prevention Study (RADIEL) a randomized controlled trial,” *Diabetes Care*, vol. 39, no. 1, pp. 24–30, 2016.

[137] C. Chiswick, R. M. Reynolds, F. Denison et al., “Effect of metformin on maternal and fetal outcomes in obese pregnant women (EMPOWAR): a randomised, double-blind, placebo-controlled trial,” *The Lancet Diabetes and Endocrinology*, vol. 3, no. 10, pp. 778–786, 2015.

[138] C. Song, J. Li, J. Leng, R. C. Ma, and X. Yang, “Lifestyle intervention can reduce the risk of gestational diabetes: a meta-analysis of randomized controlled trials,” *Obesity Reviews*, vol. 17, no. 10, pp. 960–969, 2016.

[139] D. Simmons, M. N. van Poppel, and DALI consortium, “UPBEAT, RADIEL, and DALI: what’s the difference?,” *Lancet Diabetes Endocrinol*, vol. 3, no. 10, p. 761, 2015.

[140] L. Poston, R. Bell, H. Croker et al., “Effect of a behaviourial intervention in obese pregnant women (the UPBEAT study): a multicentre, randomised controlled trial,” *The Lancet Diabetes and Endocrinology*, vol. 3, no. 10, pp. 767–777, 2015.

[141] D. Simmons, R. Devlieger, A. van Assche et al., “Effect of physical activity and/or healthy eating on GDM risk: the DALI lifestyle study,” *The Journal of Clinical Endocrinology and Metabolism*, vol. 102, no. 3, pp. 903–913, 2017.

[142] S. Yang, F. T. Shi, P. C. K. Leung, H. F. Huang, and J. X. Fan, “Low thyroid hormone in early pregnancy is associated with an increased risk of gestational diabetes mellitus,” *The Journal of Clinical Endocrinology and Metabolism*, vol. 101, no. 11, pp. 4237–4243, 2016.

[143] Y. T. Wu, C. J. Zhang, B. W. Mol et al., “Early prediction of gestational diabetes mellitus in the Chinese population via advanced machine learning,” *The Journal of Clinical Endocrinology and Metabolism*, vol. 106, no. 3, pp. e1191–e1205, 2021.

[144] M. Lacroix, M. C. Battista, M. Doyon et al., “Lower adiponectin levels at first trimester of pregnancy are associated with increased insulin resistance and higher risk of developing gestational diabetes mellitus,” *Diabetes Care*, vol. 36, no. 6, pp. 1577–1583, 2013.

[145] T. Ravnsborg, L. L. T. Andersen, N. D. Trabjerg, L. M. Rasmussen, D. M. Jensen, and M. Overgaard, “First-trimester multimarker prediction of gestational diabetes mellitus using targeted mass spectrometry,” *Diabetologia*, vol. 59, no. 5, pp. 970–979, 2016.

[146] M. A. Williams, C. F. Qiu, M. Muy-Rivera, S. Vadakkorkia, T. Song, and D. A. Luthy, “Plasma adiponectin concentrations in early pregnancy and subsequent risk of gestational diabetes mellitus,” *The Journal of Clinical Endocrinology and Metabolism*, vol. 89, no. 5, pp. 2306–2311, 2004.

[147] D. V. Tortoriello, Y. Sidis, D. A. Holtzman, W. E. Holmes, and A. L. Schneyer, “Human follistatin-related protein: a structural homologue of follistatin with nuclear localization,” *Endocrinology*, vol. 142, no. 8, pp. 3426–3434, 2001.

[148] Y. Sidis, A. Mukherjee, H. Keutmann, A. Delbaere, M. Sadatsuki, and A. Schneyer, “Biological activity of follistatin isoforms and follistatin-like-3 is dependent on differential cell surface binding and specificity for activin, myostatin, and bone morphogenetic proteins,” *Endocrinology*, vol. 147, no. 7, pp. 3586–3597, 2006.

[149] R. Thadhani, C. E. Powe, M. L. Tjøa et al., “First-trimester follistatin-like-3 levels in pregnancies complicated by subsequent gestational diabetes mellitus,” *Diabetes Care*, vol. 33, no. 3, pp. 664–669, 2010.

[150] A. Syngelaki, R. Kotecha, A. Pastides, A. Wright, and K. H. Nicolaides, “First-trimester biochemical markers of placentation in screening for gestational diabetes mellitus,” *Metabolism*, vol. 64, no. 11, pp. 1485–1489, 2015.

[151] C. Y. T. Ong, T. T. Lao, K. Spencer, and K. H. Nicolaides, “Maternal serum level of placental growth factor in diabetic pregnancies,” *The Journal of Reproductive Medicine*, vol. 49, no. 6, pp. 477–480, 2004.