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Seroprevalence of *Toxoplasma gondii* in domestic pigs, sheep, cattle, wild boars, and moose in the Nordic-Baltic region: A systematic review and meta-analysis

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**A B S T R A C T**

Background: *Toxoplasma gondii* is an important foodborne zoonotic parasite. Meat of infected animals is presumed to constitute a major source of human infection and may be a driver of geographical variation in the prevalence of anti-*T. gondii* antibodies in humans, which is substantial in the Nordic-Baltic region in northern Europe. However, data on seroprevalence of *T. gondii* in different animal species used for human consumption are scattered.

Methods: We conducted a systematic review of seroprevalence studies and meta-analysis to estimate the seroprevalence of *T. gondii* in five animal species that are raised or hunted for human consumption in the Nordic-Baltic region: domestic pigs (*Sus scrofa domesticus*), sheep (*Ovis aries*), cattle (*Bos taurus*), wild boars (*Sus scrofa*), and moose (*Alces alces*). We searched for studies that were conducted between January 1990 and June 2018, and reported in articles, theses, conference abstracts and proceedings, and manuscripts. Subgroup analyses were performed to identify variables influencing the seroprevalence.

Findings: From a total of 271 studies identified in the systematic review, 32 were included in the meta-analysis. These comprised of 13 studies on domestic pigs, six on sheep, three on cattle, six on wild boars, and four on moose. The estimated pooled seroprevalence of *T. gondii* was 6% in domestic pigs (CI₉₅: 3–10%), 23% in sheep (CI₉₅: 12–36%), 7% in cattle (CI₉₅: 1–21%), 33% in wild boars (CI₉₅: 26–41%), and 16% in moose (CI₉₅: 10–23%). High heterogeneity was observed in the seroprevalence data within each species. In all host species except wild boars, the pooled seroprevalence estimates were significantly higher in animals >1 year of age than in younger animals. Not all studies provided information on animal age, sensitivity...
Toxoplasma gondii is a zoonotic protozoan parasite distributed worldwide and globally ranked fourth among food-borne parasites that pose a threat to public health (FAO/WHO, Food and Agriculture Organization/World Health Organization, 2014). Toxoplasma gondii has a complex life cycle where felids are the only known definitive hosts (Dubey, 2009a; Dubey, 2010). Infected felids can shed millions of oocysts in their feces for a limited time. After sporulation in the environment, the oocysts can infect a wide range of hosts through contaminated soil, water, feed, and food (Dubey, 2010; Jones and Dubey, 2012). In the infected hosts, the parasite undergoes asexual multiplication and can form tissue cysts. Toxoplasma gondii tissue cysts in the meat of animals raised or hunted for human consumption pose a risk to humans if consumed raw or without thorough cooking (Jones and Dubey, 2012). Among the many possible routes of T. gondii infection, ingestion of viable parasites in the tissue cysts of meat originating from infected animals is considered important in humans (Cook et al., 2000; Tenter, 2009).

In most animal host species, the majority of T. gondii infections are subclinical. In farm animals, abortions are considered the most relevant clinical manifestation, especially in sheep, and may lead to economic losses (Dubey, 2009b; Dubey, 2009c; Dubey, 2010). In humans, the infection is often asymptomatic or causes mild symptoms. However, the infection may result in ocular toxoplasmosis; in pregnant women, the infection may result in congenital toxoplasmosis; and especially in immunosuppressed individuals, the infection may be fatal (Montoya and Liesenfeld, 2004). Additionally, recent studies have reported associations between T. gondii infection and psychiatric disorders (Sutterland et al., 2015). The disease burden caused by T. gondii has recently gained more attention (Torgerson and Mastroiacovo, 2013; Mangen et al., 2015; Nissen et al., 2017).

Toxoplasma gondii cannot be detected by routine meat inspection, and there has been relatively little emphasis on the prevention of T. gondii infection in the food chain. Seroepidemiological studies have shown that both farm animals raised for human consumption and game animals farmed or hunted for human consumption are commonly exposed to T. gondii worldwide (Dubey, 2010; Opsteegh et al., 2016).

In the Nordic-Baltic region, the T. gondii seroprevalence in humans varies markedly between the countries. It has been reported as 9% in pregnant women in Norway (Findal et al., 2015), 10% in individuals aged 20–44 years in Iceland (Birgisdóttir et al., 2006), 15% in veterinarians in Finland (Siponen et al., 2019), 20% in individuals aged ≥30 years in Finland (Suvisaari et al., 2017), 20% in pregnant women in Finland (Lappalainen et al., 1992), 23% in individuals aged 20–44 years in Sweden (Birgisdóttir et al., 2006), 28% in pregnant women in Denmark (Lebech et al., 1994), 38% in children aged 14–18 years in Estonia (Lassen et al., 2016), 55% in individuals aged 20–44 years in Estonia (Birgisdóttir et al., 2006), 56% in the general adult population in Estonia (Lassen et al., 2016), and 62% in individuals tested at a clinic in Estonia (Pehk, 1994). These geographical differences in T. gondii seroprevalence may reflect differences in food consumption habits between countries, geographic variation...
in *Toxoplasma gondii* prevalence in animals raised or hunted locally for human consumption, or different levels of oocyst contamination of the environment across the region. Quantification of the seroprevalence of *Toxoplasma gondii* in animals used for human consumption can help estimate the infection risk for humans from different types of meat; however, only sporadic data are available. Therefore, we performed a systematic review and meta-analysis to estimate the seroprevalence of *Toxoplasma gondii* in selected animals raised or hunted for human consumption across the Nordic-Baltic region.

2. Materials and methods

We performed a systematic review of seroprevalence studies on *Toxoplasma gondii* in five animal species in the Nordic-Baltic region. The study was carried out following the recommendations given in “Preferred Reporting Items for Systematic review and Meta-Analysis” (PRISMA) (Liberati et al., 2009). Seroprevalence was defined as the proportion of animals that tested positive out of the total number of animals tested.

2.1. Search strategy and data sources

We set out to identify all studies reporting *Toxoplasma gondii* seroprevalence in domestic pigs (*Sus scrofa domesticus*), sheep (*Ovis aries*), cattle (*Bos taurus*), wild boars (*Sus scrofa*), and moose (*Alces alces*) from the region comprising the Nordic countries (Denmark including the Faroe Islands and Greenland, Finland including the Åland Islands, Iceland, Norway, and Sweden) and the Baltic countries (Estonia, Latvia, and Lithuania). Studies published from beginning of January 1990 to the end of June 2018, and manuscripts identified by the end of June 2018, were considered. Studies reported in peer-reviewed articles, theses, conference abstracts and proceedings, and scientific article manuscripts, in all languages, were considered eligible.

Two of the authors (AO, RB) independently searched for peer-reviewed articles from the CAB Abstract and the MedLine database, using relevant keywords with Boolean operators ‘OR’ and ‘AND’ (Appendix A, Table A.1). Additionally, one author (AO) searched the ProQuest database (Appendix A, Table A.2), the Danish national research database (www.forskningsdatabasen.dk), and the Bulletin of Scandinavian-Baltic Society for Parasitology (http://sbsp.eu/index.php/bulletin). The last search was done on the 30th of June 2018.

The corresponding authors of the identified eligible studies were contacted to identify further studies (Appendix A, Table A.3). Furthermore, the reference lists of the identified eligible studies were screened to identify further studies (particularly grey literature).

2.2. Study selection

Search results from the two databases and other sources were combined. Eligible studies were selected using inclusion and exclusion criteria (Table 1). Two of the authors (AO, RB) used these criteria to independently screen the title and the abstract of each study. Additionally, they identified duplicates and recorded the reason for exclusion of any study during the screening process. The data representing the eligible studies for data extraction were merged and managed in Microsoft Excel 2010 (Appendix C).

2.3. Data extraction

A data extraction sheet was created in Microsoft Excel 2010 for the collection of the following information from the full-text publications: first author, year of publication, country, sample type (plasma, serum, meat juice), sampling period, host species, total number of animals sampled (per host species and per country), number of animals testing seropositive for *Toxoplasma gondii*, total number of animals by age group (young, if ≤1 year; old, if >1 year), total number of seropositive animals by age group, the serological test used, the reported cut-off for classifying a sample as positive or negative, and the reported sensitivity and specificity of the serological test employed. Moreover, for domestic pigs, we collected the total number of pigs and total number of seropositive pigs by production system (indoor, outdoor). The data extraction sheet was pilot-tested by three of the authors (AO, MS, PJ) to assess the feasibility of filling it in with data for each host species and revised before the data extraction process. A data extraction group comprising of five of the authors was formed for each

| Table 1 |
| --- |
| Inclusion and exclusion criteria for the selection of eligible studies for a systematic review on *Toxoplasma gondii* seroprevalence in domestic pigs, sheep, cattle, wild boars and moose in the Nordic-Baltic region. |

| Inclusion criteria | Exclusion criteria |
| --- | --- |
| All studies (journal articles, theses, reports, scientific article manuscripts) on *Toxoplasma gondii* seroprevalence published between 1990 and 2018 (except manuscripts, unpublished at inclusion) | Studies on *Toxoplasma gondii* seroprevalence published before 1990 |
| Samples (blood and meat juice) collected between 1990 and 2018 | Samples collected before 1990 |
| All languages | Countries outside the Nordic-Baltic region |
| Countries from the Nordic and Baltic region i.e., Denmark including the Faroe Islands and Greenland, Finland including the Åland Islands, Iceland, Norway, and Sweden, Estonia, Latvia, and Lithuania | All other host species |
| Domestic pigs (*Sus scrofa domesticus*), sheep (*Ovis aries*), cattle (*Bos taurus*), wild boars (*Sus scrofa*) and moose (*Alces alces*) | Herd-level seroprevalence studies |
| Seroprevalence studies conducted at animal-level | Experimental studies |
| Cross-sectional, cohort and unstructured study designs to estimate *Toxoplasma gondii* seroprevalence in apparently healthy animals | Access to the full text of the study for data extraction |
host species. These groups received the data extraction sheet along with a help file (Appendices D and E) and the assigned articles for reading. Each article was read by at least two authors. The extracted data were collated and compared, any disagreements in the results were resolved by discussion, and the final data were checked by three of the authors (AO, LA, PJ).

2.4. Data handling

When proportions approach zero or one, the variance of the proportions moves towards zero. As a result, studies with either high or low prevalence are given relatively high weight in the meta-analysis of prevalence (Barendregt et al., 2013). This is because the weight is calculated from the inverse of the variance of the prevalence estimate. Therefore, to avoid this, we transformed the prevalence data using the double arcsine method. Meta-analysis was then performed on the double arcsine transformed seroprevalence estimates. For reporting and interpretation of the results, the final pooled seroprevalence estimates and their 95% confidence intervals (CIs) were back-transformed to proportions to ease interpretation (Barendregt et al., 2013).

2.5. Evaluation of heterogeneity and pooled estimates

Meta-analysis was done for each host species separately. For each study, the seroprevalence and its 95% CI were calculated. Individual seroprevalence estimates were pooled using restricted maximum likelihood (REML) method with a random effects model. The individual seroprevalence estimates and the pooled seroprevalence estimates were first visually examined for heterogeneity using a forest plot. The studies were regarded homogenous, when the 95% CIs of all the seroprevalence estimates were overlapping.

| Host species | Database [A] | Other [B] | Total [C] | Duplicates [D] | Excluded [E] | Total [F] | Excluded [G] | Included [H] |
|--------------|--------------|-----------|-----------|----------------|--------------|-----------|--------------|--------------|
| Domestic pig | 95           | 6         | 101       | 50             | 36           | 15        | 4            | 13           |
| Sheep        | 53           | 7         | 60        | 30             | 19           | 11        | 6            | 6            |
| Cattle       | 45           | 4         | 49        | 23             | 20           | 6         | 3            | 3            |
| Wild boar    | 44           | 0         | 44        | 11             | 27           | 6         | 0            | 6            |
| Moose        | 17           | 0         | 17        | 10             | 3            | 4         | 0            | 4            |
| Total        | 254          | 17        | 271       | 124            | 105          | 42        | 13           | 32           |

* a = Number of studies increased during the data extraction process. Lind et al. (1994) reported two studies on domestic pigs. b = Number of studies increased during the data extraction process. Eglite and Keidans (2000) reported two studies on domestic pigs and two studies on sheep.
overlapped (Ried, 2006). Moreover, we used the inverse variance index ($I^2$) to quantify heterogeneity, where $I^2$ values of 25%, 50%, and 75% indicate low, moderate, and high heterogeneity, respectively (Higgins and Thompson, 2002).

To determine the effect of any influential studies on the overall pooled seroprevalence estimates for each host species, we conducted leave-one-out analyses (Appendix B, Figs. B.1 to B.5). A study was considered influential if the pooled seroprevalence estimate calculated without the study was not within the 95% CIs of the overall pooled seroprevalence estimate (Ding et al., 2017).
### 2.6. Subgroup analyses

The number of studies was insufficient for a multivariable regression analysis. Therefore, we performed subgroup analyses to identify possible sources of heterogeneity. The subgroups investigated were the two age groups, and for the studies on domestic pigs also the two production systems. The studies for which information about the subgroup was lacking were omitted from the analysis. Additionally, if a single study had data for both subgroups, then that study was considered as two separate studies in the subgroup analyses. The subgroup analyses were done for those host species for which at least three studies were available with data for the subgroup. Heterogeneity was explored by subgroups within each host species and leave-one-out analysis was conducted in each subgroup within each host species to detect influential studies (Appendix B, Figs. B.6 to B.15).

Subgroup analyses were performed using a mixed effect model, where the random effects model was used to pool the individual seroprevalence estimates within each subgroup and a fixed-effect of the variable of interest was used to test for significant differences between the subgroups (Borenstein et al., 2009; Wang, 2018). When computing the pooled seroprevalence estimates for each subgroup, we assumed the between-study variance to be the same for all subgroups, because the sample size within subgroups or one of the two subgroups was small (<5 studies).

The meta-analysis was performed in R studio (1.1.4), following the R script of Wang (2018).

### 3. Results

#### 3.1. Search results and eligible studies

Fig. 1 outlines the PRISMA process followed for the systematic selection and removal of the studies for each of the five host species. In the literature search, a total of 271 studies were identified: 254 from the databases and 17 from other sources (Table 2). Altogether 11 of the 14 corresponding authors we contacted responded, yielding a response rate of 79% (Appendix

### Table 3C
Characteristics of the three eligible studies on cattle included in the meta-analysis for estimating seroprevalence of *Toxoplasma gondii* in the Nordic-Baltic region.

| Authors                  | Year of publication | Country | Study period | Sample type | Serological method | Sea | Spb | Total no. of cattle ≤1 year of age (no. of seropositives) | Total no. of cattle >1 year of age (no. of seropositives) | Total no. of cattle unknown age (no. of seropositives) | Apparent seroprevalence (%) |
|--------------------------|---------------------|---------|--------------|--------------|-------------------|-----|-----|----------------------------------------------------------|----------------------------------------------------------|--------------------------------------------------------|-------------------------------|
| Egilte and Keidans, 2000 | 2000                | Latvia  | 1993–1998    | Serum        | CF                 | n.r. | n.r. | 254 (2)                                                 | 15 (0)                                                   | 7.5                                      |                               |
| Allén, 2016              | 2016                | Finland | 2013–2014    | Meat juice   | ELISA             | n.r. | n.r. | 185 (15)                                                | 15 (0)                                                   | 7.5                                      |                               |
| Jokelainen et al., 2017 | 2017                | Estonia | 2012–2013    | Serum        | DAT               | n.r. | n.r. | 3679 (707)                                               | 312 (36)                                                 | 18.6                                     |                               |

n.r. = not reported. a = Unspecified commercial complement fixation test (CF). b = Commercial ELISA, ID Screen Toxoplasmosis Indirect Multi-species IDvet Innovative Diagnostics Montpellier France [Cut-off, S/P = 50%]. c = Commercial modified direct agglutination test (DAT). Toxo-Screen DA bioMérieux, Marcy-l’Étoile France [Cut-off, dilution of 1:100]. d = Se, sensitivity. e = Sp, specificity.

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### Table 3D
Characteristics of the six eligible studies on wild boars included in the meta-analysis for estimating seroprevalence of *Toxoplasma gondii* in the Nordic-Baltic region.

| Authors                  | Year of publication | Country | Study period | Sample type | Serological method | Sea | Spb | Total no. of wild boars ≤1 year of age (no. of seropositives) | Total no. of wild boars >1 year of age (no. of seropositives) | Total no. of wild boars of unknown age (no. of seropositives) | Apparent seroprevalence (%) |
|--------------------------|---------------------|---------|--------------|--------------|-------------------|-----|-----|-------------------------------------------------------------|-------------------------------------------------------------|-------------------------------------------------------------|-------------------------------|
| Jokelainen et al., 2012  | 2012                | Finland | 2007–2008    | Serum        | DAT               | n.r. | n.r. | 24 (7)                                                     | 166 (54)                                                   | 7 (4)                                                      | 33.0                          |
| Deksnie and Kirjušina, 2013 | 2013            | Latvia  | 2010–2011    | Meat juice   | ELISA            | n.r. | n.r. | 156 (35)                                                   | 185 (51)                                                   | 130 (27)                                                  | 33.3                          |
| Jokelainen et al., 2015  | 2015                | Estonia | 2012–2013    | Meat juice   | DAT               | n.r. | n.r. | 275 (94)                                                   | 205 (113)                                                  | 847 (450)                                                 | 49.5                          |
| Wallander et al., 2015   | 2015                | Sweden  | 2005–2011    | Serum        | ELISA            | 0.79 | 0.85 | 276 (80)                                                   | 23 (0)                                                     | 29.0                         |                               |
| Malmsten et al., 2018    | 2018                | Sweden  | 2013–2015    | Serum        | ELISA            | n.r. | n.r. | 38 (6)                                                     | 61 (24)                                                    | 2 (0)                                                      | 29.7                          |
| Laforet et al., 2019     | 2019                | Denmark | 2016–2018    | Serum        | ELISA, DAT       | n.r. | n.r. | 156 (35)                                                   | 205 (113)                                                  | 847 (450)                                                 | 49.5                          |

n.r. = not reported. a = Commercial modified direct agglutination test (DAT). Toxo-Screen DA bioMérieux, Marcy-l’Étoile France [Cut-off, S/P = 50%]. b = In-house Enzyme linked immunosorbert assay (ELISA). c = Commercial ELISA, ID Screen Toxoplasmosis Indirect Multi-species IDvet Innovative Diagnostics Montpellier France [Cut-off, S/P = 50%]. d = Se, sensitivity. e = Sp, specificity.
From the 271 studies, 124 studies were duplicates and 105 studies did not meet the inclusion criteria of the screening process (Table 1). The remaining 42 studies were read in full, and 13 of them were further excluded as they did not meet the inclusion criteria. During the data extraction, the number of included studies increased with three because one of the studies on domestic pigs reported seroprevalence data also from a separate, unpublished study (Lind et al., 1994) and another study reported seroprevalence data from two studies on domestic pigs and two studies on sheep (Eglite and Keidans, 2000). Therefore, a total of 32 studies (13 on domestic pigs, six on sheep, six on wild boars, four on moose, and three on cattle) were included in the meta-analysis.

The studies included are summarized in Tables 3A to 3E. They were published between 1994 and 2018 and originated from six of the nine Nordic-Baltic countries. The number of studies per host species from each country ranged from zero to four (Fig. 2). The host species covered by the highest and lowest number of studies were domestic pigs (n = 13) and cattle (n = 3), respectively. The total number of animals tested was highest for domestic pigs (12,727), followed by sheep (6645), cattle (4445), wild boars (4237), and moose (2978).

Different serological methods were used in the studies (Tables 3A to 3E). For example, an enzyme linked immunosorbent assay (ELISA, in-house or commercial) was used in most studies on domestic pigs (n = 10), whereas a modified direct agglutination test (DAT, commercial) was used in all studies on moose (n = 4). Eight studies reported both the sensitivity and the specificity of the serological test used; seven of these studies were on domestic pigs.

### Table 3E

| Authors               | Year of publication | Country | Study period | Sample type | Serological method | Se (%) | Sp (%) | Total no. of moose ≤ 1 year of age (no. of seropositives) | Total no. of moose > 1 year of age (no. of seropositives) | Total no. of moose of unknown age (no. of seropositives) | Apparent seroprevalence (%) |
|-----------------------|---------------------|---------|--------------|-------------|-------------------|--------|--------|----------------------------------------------------------|----------------------------------------------------------|----------------------------------------------------------------|-----------------------------|
| Vikøren et al., 2004  | 2004                | Norway  | 1992–2000    | Serum       | DAT<sup>a</sup>   | n.r.   | n.r.   | 607 (27)                                                | 1468 (233)                                                | 67 (10)                                                          | 12.6                        |
| Jokelainen et al., 2010 | 2010              | Finland | 2008–2009    | Serum       | DAT<sup>a</sup>   | n.r.   | n.r.   | 454 (24)                                                | 729 (90)                                                  | 32 (2)                                                            | 9.5                         |
| Malmsten et al., 2011 | 2011                | Sweden  | 2000–2005    | Serum       | DAT<sup>a</sup>   | n.r.   | n.r.   | 122 (17)                                                | 295 (68)                                                  | 0                                                                  | 20.4                        |
| Remes et al., 2018    | 2018                | Estonia | 2015–2015    | Serum or plasma | DAT<sup>a</sup>   | n.r.   | n.r.   | 143 (18)                                                | 316 (91)                                                  | 4 (2)                                                             | 24.0                        |

<sup>a</sup> Commercial modified direct agglutination test (DAT), Toxo-Screen DA bioMérieux, Marcy-l’Étoile France [Cut-off, dilution of 1:40]. n.r. = not reported.

<sup>b</sup> Se, sensitivity. <sup>c</sup> Sp, specificity.

Fig. 2. Number of Toxoplasma gondii seroprevalence studies in domestic pigs (*Sus scrofa domesticus*), sheep (*Ovis aries*), cattle (*Bos taurus*), wild boars (*Sus scrofa*), and moose (*Alces alces*), by country in the Nordic-Baltic region, 1990–2018.
Toxoplasma gondii

Pooled seroprevalence estimates for five host species in the Nordic-Baltic region, 2018.

| Host species          | No. of studies | No. seropositive animals | Total no. of animals | Pooled seroprevalence (%) (95% CI) | Heterogeneity |
|----------------------|----------------|--------------------------|----------------------|------------------------------------|---------------|
| Domestic pig         | 13             | 568                      | 12,727               | 6.3 (3.5–10.0)                     | 278.8         |
| Sheep                | 6              | 1653                     | 6645                 | 22.7 (12.0–36.0)                   | 367.9         |
| Cattle               | 3              | 760                      | 4445                 | 7.4 (1.0–21.0)                     | 131.5         |
| Wild boar            | 6              | 1146                     | 2792                 | 33.1 (26.0–41.0)                   | 136.3         |
| Moose                | 4              | 582                      | 4237                 | 16.1 (10.0–23.2)                   | 68.7          |

$I^2 =$ Inverse variance index; Q = Cochran’s Q test for heterogeneity; Q-P = probability value of Cochran’s Q test for heterogeneity.

3.2. Heterogeneity in the seroprevalence estimates and subgroup analyses

The T. gondii seroprevalence estimates varied markedly among the included studies: the crude range was 1–30% in domestic pigs, 6–45% in sheep, 1–19% in cattle, 24–50% in wild boars, and 10–24% in moose. For all host species, there was significant heterogeneity among the seroprevalence estimates from the included studies (Fig. 3A to E, Table 4). The highest pooled seroprevalence estimate (PSP) was observed for wild boars (PSP = 33%, CI95%: 26–41%) followed by sheep (PSP = 23%, CI95%: 12–36%), moose (PSP = 16%, CI95%: 10–23%), cattle (PSP = 7%, CI95%: 1–21%), and domestic pigs (PSP = 6%, CI95%: 3–10%). For all host species, the results from the leave-one-out analyses showed that the pooled seroprevalence estimates were not significantly influenced by any of the included studies (Appendix B, Figs. B.1 to B.5).

In all host species except for wild boars, the pooled seroprevalence estimate of T. gondii was significantly higher in old than in young animals (Table 5). The lowest pooled seroprevalence estimates in both the age groups were recorded in domestic pigs (PSPyoung = 4%, CI95%: 2–6% and PSPold = 18%, CI95%: 12–25%, respectively), and the highest were recorded in wild boars (PSPyoung = 26%, CI95%: 16–37% and PSPold = 38%, CI95%: 28–49%, respectively). Heterogeneity in the seroprevalence estimates between studies in the two age groups varied from moderate (young sheep, $I^2$ = 49% and young wild boars, $I^2$ = 71%) to high (young pigs, $I^2$ = 90%; old pigs, $I^2$ = 91%; old sheep, $I^2$ = 99%; old wild boars, $I^2$ = 91%; young moose, $I^2$ = 85%, and old moose, $I^2$ = 94%).

The pooled seroprevalence estimate was 3% (CI95%: 0.3–7%, $I^2$ = 96%) in indoor pigs and 8% (CI95%: 2–16, $I^2$ = 98%) in outdoor pigs. The difference between the pooled seroprevalence estimates in domestic pigs from these two production systems was not statistically significant ($P = 0.11$).

4. Discussion

This systematic literature review and meta-analysis estimated T. gondii seroprevalence in domestic pigs, sheep, cattle, wild boars, and moose in the Nordic-Baltic region, and identified important data gaps. The results of this study summarize the extent of reported exposure to T. gondii in different animal species in the Nordic-Baltic region, which is of importance in the context of food safety. Particularly from the Baltic states, limited data have been previously available, whereas several studies have been conducted since the year 2013 and were included in this systematic review.

The host species covered in this study included both farm animals and game animals. Domestic pigs and wild boars are omnivores and may acquire T. gondii infection by eating sporulated oocysts, shed in unsporulated form by infected felids, or by eating tissues of infected animals. Sheep, cattle, and moose as herbivores likely acquire the infection from sporulated oocysts. All included studies reported seropositive animals, indicating exposure to T. gondii is widespread in the region.

A wide range of T. gondii seroprevalence estimates was observed across the different studies. Because of the high level of heterogeneity between the studies, the pooled seroprevalence estimates presented in this study should be interpreted together with the 95% CIs.

Among the farm animals, domestic pigs had the lowest pooled seroprevalence estimate (PSP = 6%, CI95%: 3–10%), which was on the lower end of the wide range for seroprevalence estimates for pigs from Europe (0.4–64%) (Dubey, 2009b). The highest pooled seroprevalence estimate among farm animals was observed in sheep (PSP = 23%, CI95%: 12–36%) and the estimate was...
close to the mid-range of the seroprevalence estimates for sheep reported in Europe (4–66%) (Dubey, 2009a). From an interpretative perspective, the pooled seroprevalence estimate in cattle (PSP = 7%, CI95%: 1–21%) may be of limited importance due to the low number of studies and high heterogeneity in seroprevalence between them. The relevance of serology for screening of T. gondii in cattle is debatable due to reports of poor correlation between seropositivity and direct detection of the parasites (Opsteegh et al., 2011; Opsteegh et al., 2016).

Among the host species included in our study, wild boars had the highest pooled seroprevalence estimate (PSP = 33%, CI95%: 26–41%), which was in the mid-range of the seroprevalence estimates for wild boars in Europe (5–57%) (Rostami et al., 2017). Toxoplasma gondii is common in wild boar populations throughout the world; the pooled seroprevalence in Europe has been estimated to be 26% (CI: 21–30%) (Rostami et al., 2017). For moose, the pooled seroprevalence estimate in this study (PSP = 16%, CI95%: 10–23%) was also in line with the seroprevalence estimates from other parts of the world (USA 10%; Canada 15%) (Siepierski et al., 1990; Verma et al., 2016).

A higher seroprevalence was observed in animals >1 year of age than in younger animals in all host species covered in our study except wild boars, and similar results have been reported in other studies (Opsteegh et al., 2016). In general, higher seroprevalence in older animals likely reflects a longer period of exposure, which increases the probability of acquiring the infection. Dutch studies have reported higher seroprevalences (5% and 6%) in pigs from outdoor farms compared with pigs from intensive indoor farms (0 and 0.4%) (Kijlstra et al., 2004; van der Giessen et al., 2007). We did not find a statistically significant difference in the pooled seroprevalence estimates between pigs from indoor (PSP = 3%; CI95%: 0.3–7%; I2 = 96%) and outdoor production systems (PSP = 8%; CI95%: 2–16; I2 = 98%) in our study, possibly because of large variation between the limited number of studies. The pooled seroprevalence estimate for pigs from outdoor farms in our study was higher than the estimates from the two Dutch studies.

The studies included in the meta-analysis used different serological methods, sample material, and cut-offs for seropositivity. The performance of serological tests varies between the type of samples (Hill et al., 2006) and the selected cut-offs (Felin et al., 2017). When the serological test is imperfect (sensitivity <100%; specificity <100%), the prevalence estimate obtained from the test is biased; however, this bias can be corrected if the sensitivity and specificity of the tests are known (Diggle, 2011). There is no ‘reference standard’ serological test with 100% sensitivity and 100% specificity for anti-T. gondii antibodies; therefore, it may be useful to apply a latent class approach to estimate the accuracy of a test (Gardner et al., 2010). In this study, we decided to work on apparent seroprevalence estimates from all the studies, without pooling the studies based on the serological test type, due to the lack of data on sensitivity and specificity of the tests needed for adjustment of the seroprevalence estimate. Hence, it is likely that the differences in the serological methods may have contributed to the heterogeneity seen in seroprevalence between the studies.

However, five studies on domestic pigs (Table 3A: (Lind et al., 1994; Lundén et al., 2002; Felin et al., 2015; Koföed et al., 2017; Santoro et al., 2017)), two studies on sheep (Table 3B: (Deksne et al., 2017; Tagel et al., 2019)), three studies on wild boars (Table 3D: (Jokelainen et al., 2015; Wallander et al., 2015; Laforet et al., 2019)), and four studies on moose (Table 3E: (Vikøren et al., 2004; Jokelainen et al., 2010; Malmsten et al., 2011; Remes et al., 2018)) tested young and old animals using the same
test and the same cut-off, and found the seroprevalence to be higher in old animals than in young animals as confirmed by the subgroup analysis (Table 5). Hence, the observed difference in seroprevalence by age group appears to be a direct effect of age and cannot be simply attributed to the choice of diagnostic test.

In meta-analysis, it is recommended to quantify and adjust for publication bias using statistical methods (Møller and Jennions, 2001). This is done to correct for missing studies to avoid overestimation of the true effect size. However, currently available methods such as Egger’s regression test and the trim-and-fill method are not considered useful in studies on proportions (Murad et al., 2018). Additionally, the statistical power of the tests is affected by the presence of high heterogeneity and the limited number of studies (Lau et al., 2006). Therefore, we decided not to look for evidence of publication bias in our study.

Based on the included studies, a substantial proportion of animals investigated were seropositive, which indicates that animals raised or hunted for human consumption in the Nordic-Baltic region were commonly exposed to *T. gondii*. There was a large variation in seroprevalence estimates between the studies in the region. If the observed variations in animal seroprevalence estimates between the studies in the region represent spatial variations in prevalence, then this might partly explain the geographical variation in the reported seroprevalence in human estimates in this region (Lappalainen et al., 1992; Lebech et al., 1994; Pehk, 1994; Birgisdóttir et al., 2006; Findal et al., 2015; Lassen et al., 2016; Suvisaari et al., 2017; Siponen et al., 2019). However, the number of studies available was too low to identify spatial patterns at the country-level, and other differences, such as the age of the tested animals and the serological tests applied may also have affected the estimates. To clarify the sources of heterogeneity, more studies and more data on risk factors that are relevant to each host species are needed. Furthermore, it is important for future studies to report the age of the animals tested, mention the type of production system the animals raised for food are reared in and report the sensitivity, specificity, and cut-off of the serological test used, whenever possible.

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**Conflict of interest statement**

The authors declare that they have no conflict of interest.

**Appendix A. Supplementary data**

Supplementary data to this article can be found online at https://doi.org/10.1016/j.parepi.2019.e00100.

**References**

Allén, E., 2016. E. Toxoplasma gondii –vasta-aineiden esiintyvyys suomalaisilla naudoilla. Retrieved from https://helda.helsinki.fi/bitstream/handle/10138/165893/Lisensiaatinvaihto_EllaAllen_12.4.2016.pdf.

Barendregt, J.J., Doi, S.A., Lee, Y.Y., Norman, R.E., Ves, T., 2013. Meta-analysis of prevalence. J. Epidemiol. Community Health 67 (11), 974–978.

Birgisdóttir, A., Asbjörnsdottir, H., Cook, E., Gislason, D., Jansson, C., Olfsson, L., et al., 2006. Seroprevalence of Toxoplasma gondii in Sweden, Estonia and Iceland. Scand. J. Infect. Dis. 38 (8), 625–631.

Borenstein, M., Hedges, L.V., Higgins, J.P.T., Rothstein, H.R., 2009. Introduction to Meta-Analysis. John Wiley & Sons, Hoboken (Chapter 19).

Cook, A.J., Gilbert, R.E., Buffolano, W., Zufferey, J., Petersen, E., Jenum, P.A., et al., 2000. Sources of toxoplasma infection in pregnant women: European multicentre case-control study. BMJ. 321 (7254), 142–147.

Denske, G., 2010. Occurrence of Toxoplasma gondii in swine in Latvia – preliminary results. Acta Biol. Univ. Daugavp. 10 (2), 143–145.

Denske, G., Kirsiuša, M., 2013. Seroprevalence of Toxoplasma gondii in domestic pigs (Sus scrofa domestica) and wild boars (Sus scrofa) in Latvia. J. Parasitol. 99 (1), 44–47.

Denske, G., Ligere, B., Šneideere, A., Jokelainien, P., 2017. Seroprevalence and factors associated with Toxoplasma gondii infections in sheep in Latvia: Latvian dark headed sheep breed associated with higher seroprevalence. Vector Borne Zoonotic Dis. 17 (7), 478–482.

Diggle, P.J., 2011. Estimating prevalence using an imperfect test. Epidemiol. Res. Int. 608719.

Ding, H., Gao, Y.M., Deng, Y., Lamberton, P.H., Lu, D.B., 2017. A systematic review and meta-analysis of the seroprevalence of Toxoplasma gondii in cats in mainland China. Parasit. Vectors 10 (1), 27. https://doi.org/10.1186/s13071-017-1970-6.

Dubey, J.P., 2009a. History of the discovery of the life cycle of Toxoplasma gondii. Int. J. Parasitol. 39 (8), 877–882.

Dubey, J.P., 2009b. Toxoplasmosis in pigs—the last 20 years. Vet. Parasitol. 164 (2–4), 89–103.

Dubey, J.P., 2009c. Toxoplasmosis in sheep—the last 20 years. Vet. Parasitol. 163 (1–2), 1–14.

Dubey, J.P., 2010. Toxoplasmosis of Animals and Humans. second ed. CRC Press, Boca Raton.

Eglite, L, Keidans, P., 2000. Majdzinieku toksoplasmoze Latvija (in Latvian). In Proceedings of International Conference – Current Issues in Veterinary Medicine, Jelgava, Latvia 45–49.

FAO/WHO, Food and Agriculture Organization/World Health Organization, 2014. Multicriteria-based ranking for risk management of food-borne parasites. Microbiol. Risk Assess. 23, 16.
van der Giessen, J., Fonville, M., Bouwknegt, M., Langelaar, M., Vollema, A., 2007. Seroprevalence of *Trichinella spiralis* and *Toxoplasma gondii* in pigs from different housing systems in the Netherlands. *Vet. Parasitol.* 148 (3–4), 371–374.

Verma, S.K., Carstensen, M., Calero-Bernal, R., Moore, S.A., Jiang, T., Su, C., Dubey, J.P., 2016. Seroprevalence, isolation, first genetic characterization of *Toxoplasma gondii*, and possible congenital transmission in wild moose from Minnesota, USA. *Parasitol. Res.* 115 (2), 687–690.

Vikøren, T., Tharaldsen, J., Fredriksen, B., Handeland, K., 2004. Prevalence of *Toxoplasma gondii* antibodies in wild red deer, roe deer, moose, and reindeer from Norway. *Vet. Parasitol.* 120 (3), 159–169.

Wallander, C., Frössling, J., Vågsholm, I., Uggla, A., Lundén, A., 2015. *Toxoplasma gondii* seroprevalence in wild boars (*Sus scrofa*) in Sweden and evaluation of ELISA test performance. *Epidemiol. Infect.* 143 (9), 1913–1921.

Wallander, C., Frössling, J., Dórea, F.C., Uggla, A., Vågsholm, I., Lundén, A., 2016. Pasture is a risk factor for *Toxoplasma gondii* infection in fattening pigs. *Vet. Parasitol.* 224, 27–32.

Wang, N., 2018. How to conduct a meta-analysis of proportions in R: A Comprehensive Tutorial. https://www.researchgate.net/publication/325486099_How_to_Conduct_a_Meta-Analysis_of_Proportions_in_R_A_Comprehensive_Tutorial (Last access date: 21st January 2019).