Effects of orthodontic force on root surface damage caused by contact with temporary anchorage devices and on the repair process

Özge Çelik Güler a
Sıddık Malkoç b

Department of Orthodontics, Faculty of Dentistry, Çanakkale Onsekiz Mart University, Çanakkale, Turkey
Private Practice, Istanbul, Turkey

Objective: This study aimed to evaluate the effects of force loading on root damage caused by contact with temporary anchorage devices (TADs) during orthodontic treatment and to examine the repair process 4, 8, and 12 weeks after TAD contact by micro-computed tomography (CT). Methods: We enrolled 42 volunteers who required bilateral upper first premolar extractions. The experimental study design was as follows. For both first premolars, cantilever springs were placed, and then TADs were immediately inserted between the premolars of all volunteers. According to the removal order of the appliances, the participants were divided into the TAD group (Group T: n = 21, only TAD removal) and the spring group (Group S: n = 21, only spring removal). A split-mouth design was adopted in both groups as follows. For each volunteer, the left premolars were extracted 4, 8, or 12 weeks after TAD–root contact. The right premolars were extracted immediately after contact in both groups (Groups T-C and S-C) and used as positive controls. Resorption volumes and numbers of craters were determined by micro-CT. Results: The numbers of resorption craters were higher in Group T than in Group S at 8 and 12 weeks (p < 0.01). Crater volumes were higher in Group T than in Group S at 4 and 12 weeks (p < 0.01, both). Conclusions: Root injury was not completely repaired 12 weeks after root–TAD contact, even when the TADs were removed in cases of continuous force application. [Korean J Orthod 2019;49(2):106-115]

Key words: Root resorption, Root damage, Micro-computed tomography, Force loading

Received August 17, 2018; Revised October 21, 2018; Accepted November 28, 2018.

Corresponding author: Özge Çelik Güler.
Assistant Professor, Department of Orthodontics, Faculty of Dentistry, Çanakkale Onsekiz Mart University, Terzioğlu Kampüsü Mühendislik Fakültesi Ek Bina, Çanakkale 17100, Turkey.
Tel +90-539-3000737 e-mail dt.celikozge@gmail.com

How to cite this article: Çelik Güler Ö, Malkoç S. Effects of orthodontic force on root surface damage caused by contact with temporary anchorage devices and on the repair process. Korean J Orthod 2019;49(2):106-115.

© 2019 The Korean Association of Orthodontists.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.
INTRODUCTION

Temporary anchorage devices (TADs) have been used in clinical practice for anchorage and tooth movement, including intrusion and extrusion, distalization and mesialization, correction of the tooth axis and occlusal cant, en-masse retraction procedures,¹² and to apply orthopedic force.¹ They can be placed at several sites during a single appointment without the need for a complicated surgical procedure.³ However, TADs are also associated with various complications including vessel and sinus injury, TAD fracture during removal, TAD failure during insertion,⁵⁶ and TAD movement during orthodontic loading.⁷ One possible reason for these complications has been suggested to be inadequate interradicular space between the root and TAD. The preferred and most secure site for TAD application was suggested to be the inter-radicular bone between the roots of adjacent teeth. However, this area was also associated with a risk of root damage.⁸⁻¹⁰ In addition to incorrect placement, TADs may also accidentally contact the root during the application of orthodontic force.

It was reported that root contact with TADs resulted in root resorption but that root repair occurred after contact was discontinued; damage was restricted to the cementum or dentin with no pulpal perforation.¹ Studies evaluating the healing period after root damage caused by TADs have reported conflicting results. For example, Kadioglu et al.¹¹ reported that complete healing occurred in 8 weeks, whereas Ahmed et al.¹² suggested that more than 12 weeks were required for complete healing in their clinical studies of patients. There is a lack of knowledge about the management of complications caused by TADs.²

Micro-computed tomography (CT) is a fast, high-resolution method with a low error rate that enables advanced visual and three-dimensional (3D) evaluations.¹³¹⁴ Previously, it was used to monitor and measure resorption craters on the root surfaces of extracted teeth after orthodontic tooth movement.¹⁵ Previous studies examined the accuracy of 3D evaluations of root resorption craters versus two-dimensional histological and radiographic evaluations.¹⁶¹⁷ To the best of our knowledge, there has been no 3D imaging study on the effect of the removal order of TADs or on the force used for root repair 12 weeks after accidental root-TAD contact. Thus, the aim of the current study was to evaluate force loading on root damage after root–TAD contact and on the root repair process over time (at 4, 8, and 12 weeks) after root–TAD contact. An additional aim was to add to the knowledge base on the management of this complication. We hypothesized that orthodontic force would cause more damage to the root surface than a TAD and that healing after damage would be incomplete at 12 weeks.

MATERIALS AND METHODS

This study adopted an experimental clinical design previously described in the literature.⁸ Forty-two volunteers (24 female subjects and 18 male subjects aged between 13 and 18 years; mean, 15.15 ± 1.55 years) who required bilateral upper first premolar extractions who presented for orthodontic treatment between May 2014 and December 2015 in the Department of Orthodontics, Faculty of Dentistry, Inonu University were included. The current study was approved by the clinical research ethics committee of Inonu University (#2014-22). Written and detailed informed consent was obtained from the parents or guardians of all participants.

The exclusion criteria for all participants were a history of orthodontic treatment, poor oral hygiene with gingivitis detected by a periodontal clinical examination, metal allergies, non-cooperation, craniofacial or dentoalveolar anomalies, incomplete tooth apexification confirmed by radiographs, and close proximity between roots and impacted canines.¹⁶

Study groups and design

The randomization was performed at baseline for patient selection. All right sides were selected as the positive control group and all left sides were selected as the experimental group. Participants were divided into the TAD group (Group T: only TAD removal, n = 21) and the spring group (Group S: only spring removal, n = 21) according to the type of the removed appliances. For both groups’ first premolars, cantilever springs were placed, and TADs were then immediately inserted between the first and second premolars. A split-mouth design was used in both groups as follows. In each volunteer, the left premolars were extracted 4, 8, or 12 weeks after TAD–root contact (Group T and Group S). The right premolars were extracted immediately after TAD–root contact and used as positive controls (Groups T-C and S-C) (Figure 1). Resorption crater volumes and numbers of craters were determined by micro-CT.

Clinical procedure

Molar bands (Seamless Bands; 3M Unitek, Monrovia, CA, USA) were cemented to the upper first molars with glass ionomer cement (Ketac; 3M ESPE, Seefeld, Germany) and 0.022-inch self-ligating brackets (Empower; American Orthodontics, Sheboygan, WI, USA) were then applied to both upper first premolars (Transbond XT; 3M Unitek). Titanium molybdenum alloy cantilever springs (BT3, 14 × 0.017 × 0.025 inch; G&H, Franklin, IN, USA) were applied with a 150-g force (Figure 2) and TADs (Aarhus screw; Medicon, Tuttinglen, Germany)
with a 1.5-mm diameter and 8-mm length were placed between the first and second premolars. Resin-modified glass ionomer cement (Multi-Cure Glass Ionomer Orthodontic Band Cement; 3M Unitek) was placed on the surface of the mandibular first molars to avoid occlusal interference (Figure 3). The positions of the TADs were confirmed on periapical radiographs (Figure 4A and 4B). The force was applied until TAD–root contact was obtained, which occurred after 4 weeks in all participants.

In all individuals in Groups T-C and S-C, the right premolars were extracted immediately after TAD–root contact was confirmed by periapical radiographs. Following TAD–root contact, the left premolars were extracted 4, 8, or 12 weeks after TAD removal (Group T) or spring removal (Group S).

The same surgeon performed all extractions under local anesthesia. During the extraction process, care was taken when using extraction forceps to avoid dam-
age to the surfaces of the roots. After the extractions, blood and tissue residues on the teeth were removed by pressureless washing with an isotonic solution without damaging the root surface. For each tooth, root resorption was observed by visual inspection with the aid of a magnifying glass (Figure 5). The extracted teeth were then transferred to sterile tubes. Finally, the teeth were transferred to the laboratory for radiographic analysis.

Radiographic analysis
The numbers and volumes of resorption craters were detected using a micro-CT system (SkyScan 1172; Bruker, Kontich, Belgium). In total, 1,300 to 1,350 sections were obtained with a rotation interval of 0.4° and thickness of about 9.0 µm while rotating the device 360° to scan the teeth. The area of evaluation was limited to the cervicoapical direction of cross-section slides (CTAn 1.15.4.0, SkyScan; Kontich). The total root surface was divided into three parts: cervical, middle, and apical. Each part was further divided into four sub-regions: mesiobuccal, distobuccal, mesiolingual, and distolingual (Figure 6). The resorption area in all three planes was separated from other tissues by using the same program (CTAn 1.15.4.0, SkyScan). In the second step, a binary view page was used to perform thresholding in order to determine the resorption craters. A custom processing page was utilized and the same task list was used as the standard to calculate the volumes and numbers of resorption craters in each sample. Finally, the volumes and numbers of craters in the roots were calculated.

Statistical analysis
To determine the sample size for each group, a power analysis was carried out based on an alpha significance level of 0.05 and beta value of 0.1 to achieve 90% power to detect an average difference of 0.5 mm³ ±
0.20 mm$^3$) in resorption craters on root surfaces from the different groups (version 3.0.10, G*Power; Franz Faul Universität, Kiel, Germany). The power analysis showed that seven patients were required in each group. Thus, to achieve 90% power (significance levels: alpha = 0.05, beta = 0.1), the patients in Group T and Group S were randomly divided into three subgroups according to the observation period (4, 8, or 12 weeks) with seven patients in each group.

Descriptive statistical data (the mean and standard deviation) are presented. Non-parametric statistical tests were conducted to determine the volumes and numbers of resorption craters. The Kruskal–Wallis test, a one-way analysis of variance, and the Mann–Whitney U test with the Bonferroni correction were performed for statistical assessments. The data were analyzed using SPSS software ver. 22.0 (IBM Corp., Armonk, NY, USA).

**RESULTS**

**Demographic data**

There were no significant differences in the ages of patients between Group T (15.81 ± 1.59 years, 14.60 ± 1.42 years, and 15.03 ± 1.39 years) and Group S (15.05 ± 0.93 years, 15.67 ± 1.87 years, and 14.74 ± 2.07 years) at 4, 8, and 12 weeks, respectively. Group T comprised 10 male subjects and 11 female subjects and Group S comprised 8 male subjects and 13 female subjects. The non-equivalence of the female and male subjects in the groups originated from the fact that some volunteers did not complete the study and new volunteers were added immediately.

**Resorption crater volume**

The total crater volume in Group T was higher than in Group S at 4 and 12 weeks after TAD–root contact ($p = 0.006$) but similar at 8 weeks ($p > 0.05$) (Figure 7A, Table 1). In Group T, the total crater volume was higher at 4 weeks compared to those at 8 and 12 weeks ($p = 0.03$). In Group S, the total crater volume was lowest at 12 weeks compared to those at 4 and 8 weeks ($p = 0.01$) (Figure 8A and 8B, Table 2).

**Table 1. Comparisons of crater volumes and numbers in Groups T and S**

| Group | Volume (mm$^3$) | Number |
|-------|-----------------|--------|
| T4    | 0.57 ± 0.24     | 9.8 ± 4.6 |
| S4    | 0.19 ± 0.15     | 10.1 ± 6.5 |
| p-value | 0.006*         | 1.000  |
| T8    | 0.27 ± 0.30     | 13.0 ± 2.7 |
| S8    | 0.23 ± 0.21     | 10.2 ± 8.8 |
| p-value | 0.940           | 0.050*  |
| T12   | 0.25 ± 0.15     | 16.4 ± 5.8 |
| S12   | 0.04 ± 0.06     | 5.7 ± 2.8  |
| p-value | 0.006*         | 0.007*   |

Values are presented as mean ± standard deviation. Group T, Temporary anchorage device experimental group; Group S, spring experimental group; T4, Group T at 4 weeks; S4, Group S at 4 weeks; T8, Group T at 8 weeks; S8, Group S at 8 weeks; T12, Group T at 12 weeks; S12, Group S at 12 weeks. The $p$-values are analyzed by Mann–Whitney $U$-test; *significance level of $p \leq 0.05$. 

Figure 7. A, The crater volumes in Groups T and S. The resorption crater volumes were significantly lower in Group S than in Group T at 4$^*$ and 12$^†$ weeks (both, $p = 0.006$). B, The numbers of craters in Groups T and S. The numbers of resorption craters were significantly higher in Group T than in Group S at 8$^†$ and 12$^* $weeks ($p = 0.05$ and $0.007$, respectively).

Group T, Temporary anchorage device experimental group; Group S, spring experimental group.

**Figure 7.** A, The crater volumes in Groups T and S. The resorption crater volumes were significantly lower in Group S than in Group T at 4* and 12† weeks (both, $p = 0.006$). B, The numbers of craters in Groups T and S. The numbers of resorption craters were significantly higher in Group T than in Group S at 8† and 12* weeks ($p = 0.05$ and 0.007, respectively).
The crater volume in Group T was higher than that in Group T-C at 4 weeks ($p = 0.006$) but similar between Group S and Group S-C at 4, 8, and 12 weeks ($p > 0.05$) (Figure 9A and 9B, Table 3).

**Numbers of resorption craters**

The total number of craters in Group T was significantly higher than that in Group S at 8 ($p = 0.05$) and 12 weeks ($p = 0.007$) but similar at 4 weeks ($p > 0.05$) (Figure 7B, Table 1). However, there were no differences in crater numbers between Group T at 4, 8 and 12 weeks and between Group S at 4, 8 and 12 weeks ($p > 0.05$) (Figure 8C and 8D, Table 2).

Moreover, there were no differences in crater numbers between Groups T and T-C or between Groups S and S-C at 4, 8, or 12 weeks ($p > 0.05$) (Figure 9C and 9D, Table 3).

**DISCUSSION**

Root damage caused by TADs is a common complication seen in clinical practice. There is limited knowledge on the management of this complication, including the repair process of the root surface and the timing of force loading. Thus, the current study investigated the repair process for root damage caused by TADs using micro-CT. The results revealed that a 12-week period may not be adequate for root healing. The results also suggested that when TADs cause root damage, clinicians should remove the loading immediately and should not perform reloading for longer than 12 weeks.

Several studies have evaluated changes to the root surface after TAD-root contact. Kadioglu et al. examined root damage in 10 patients over an 8-week period histologically and reported that collagen fibrils completely covered the damaged surface during the entire period. In the current study, which adopted a similar group design to that used by Kadioglu et al., an 8-week period was not adequate for the root repair process, as evaluated by micro-CT. Instead, longer than 12 weeks was required for the repair process. The discordance be-

**Table 2. Comparisons of crater volumes and numbers in Groups T and S over different periods**

| Group | Volume (mm$^3$) | Number  |
|-------|----------------|---------|
| T4    | 0.570 ± 0.240$^A$ | 9.8 ± 4.6 |
| T8    | 0.270 ± 0.300$^B$ | 13.0 ± 2.7 |
| T12   | 0.250 ± 0.150$^B$ | 16.4 ± 5.8 |
| $p$-value | 0.03$^*$ | 0.64 |
| S4    | 0.190 ± 0.150$^A$ | 10.1 ± 6.5 |
| S8    | 0.230 ± 0.210$^A$ | 10.2 ± 8.8 |
| S12   | 0.040 ± 0.060$^B$ | 5.7 ± 2.8 |
| $p$-value | 0.01$^*$ | 1.0 |

Values are presented as mean ± standard deviation. Group T, Temporary anchorage device experimental group; Group S, spring experimental group; Group T at 4 weeks; T8, Group T at 8 weeks; T12, Group T at 12 weeks; S4, Group S at 4 weeks; S8, Group S at 8 weeks; S12, Group S at 12 weeks. $^A,B$ There is no statistically significant difference between groups with the same letters. The $p$-values are analyzed by Kruskal–Wallis and Mann–Whitney U-test; $^*$ significance level of $p ≤ 0.05$.

**Figure 8.** A, A crater volume comparison in Group T over different periods. The total crater volume was significantly higher at 4 weeks compared to those at 8 and 12 weeks ($p = 0.03$). B, A crater volume comparison in Group S over different periods. The lowest total crater volume was seen at 12 weeks compared to those at 4 and 8 weeks ($p = 0.01$). C, A comparison of crater numbers in Group T over different periods. D, A comparison of crater numbers in Group S over different periods.

Group T, Temporary anchorage device experimental group; Group S, spring experimental group.
the study findings may be attributed to the methodologies used, as a histological analysis is superior to micro-CT in terms of the detection of repaired cementum.  
Kadioglu et al. also evaluated the removal order of the spring or TAD but they did not report a comparison of the removal order. In addition, in the same study, they did not conduct a statistical analysis of the impact of the removal order of springs or TAD on the healing period. In the current study, immediately after TAD–root contact, the first step was removal of the spring, followed by no reloading for longer than 12 weeks. In this study, crater volumes and numbers significantly decreased after removing the spring and leaving the TAD in contact with the root in situ (Group S) compared with removal of the TAD with continuous force (Group T) for 12 weeks. Both groups showed a tendency toward decreased crater volumes after 12 weeks but with no difference in crater numbers. Thus, complete root healing did not appear to have occurred by 12 weeks, as shown by micro-CT.
Ahmed et al. examined cementum repair after TAD–root contact in human upper first premolars and reported that 70% of teeth showed ≥ 90% healing at 12 weeks, with most healing completed by 8 weeks. They concluded that longer than 12 weeks was required for complete healing. However, they did not examine the effects of force loading and did not leave the TAD in situ after TAD–root contact. In the current study, similar to the findings of Ahmed et al., a 12-week period was not sufficient for healing. In addition, leaving the TAD in contact with the root affected the repair process less adversely compared with removal of the TAD with continuous force. TAD–root contact may not be detected immediately in a clinical setting. The results of the present study suggest that it is better to remove the spring at the time of the detection of TAD–root contact rather than leaving it in situ.

**Figure 9.** A, A comparison of crater volumes in Groups T and T-C. The crater volume was higher in Group T than in Group T-C at 4 weeks (*p = 0.006). B, A comparison of crater volumes in Groups S and S-C. Comparisons of the distributions of the total number of craters in Groups T and T-C (C) and in Groups S and S-C (D).

Group T, Temporary anchorage device experimental group; Group S, spring experimental group.
They concluded that when TAD–root contact occurs, the TAD should be removed immediately. They also reported that when the TAD was left in contact with the root in situ, no definitive cementum repair was observed for up to 16 weeks. In the current study, in the group in which the TAD was left in situ without force (Group S), crater volumes were decreased at 12 weeks compared to those at 4 weeks, whereas crater numbers remained similar at all observation time points. The decrease in crater volume was attributed to spring removal, which may cause relapse and thus the TAD–root contact may be not be tight. In the study by Kim and Kim, the authors removed the TAD after TAD–root contact and evaluated root healing over a 16-week period without force loading. They observed partial healing after 4 weeks and increased healing over time, with complete healing observed by 8 to 16 weeks. In the current study, crater volumes in the group that underwent TAD removal with continuous force (Group T) decreased over time, but crater numbers were similar in Group T at all observation time points. The differences in crater volumes were attributed to continued force loading from springs in the TAD removal group (Group T) possibly decreasing healing. Kim and Kim also reported that root damage caused by TADs was irreversible when the TADs penetrated the pulp. In the current study, no pulpal damage was detected. The effects of TAD removal on healing were not compared with the findings of Kim and Kim because of the difference in study designs.

Hembree et al. evaluated the healing process following root damage caused by TAD–root contact over both short (6 weeks) and long (12 weeks) periods in seven beagle dogs. They reported that damage caused by leaving the TAD in situ and in contact with the roots was the same at 6 and 12 weeks compared to that at baseline, contrary to expectations. They also reported that leaving a TAD in contact with the root may not stimulate root resorption without continuous force. Similar to their findings, we found significantly lower crater volumes and numbers at 12 weeks in Group T compared to those in Group S. In the current study, force loading from springs appeared to cause more resorption and less healing than leaving a TAD in contact with the root in situ without force for 12 weeks.

Similar to Hembree et al., Renjen et al. evaluated the effect of TAD–root contact on the root surface in three beagle dogs for 12 weeks and concluded that TAD penetration of the root surface did not influence cementum repair or healing. In the current study, similar to the findings of Renjen et al., leaving the TADs in situ appeared to have less of an effect on healing than a continuous force.

The minimum distance between the root and TAD was suggested to be 1.5 mm. In addition, root resorption was reported when the mini-implant was closer than

Table 3. Comparisons of crater volumes and numbers in the experimental and control groups

| Group | Experimental | Control | p-value |
|-------|--------------|---------|---------|
| Volume (mm³) | | | |
| T4 | 0.57 ± 0.24 | 0.28 ± 0.10 | 0.006* |
| T8 | 0.27 ± 0.30 | 0.24 ± 0.21 | 0.740 |
| T12 | 0.25 ± 0.15 | 0.20 ± 0.12 | 0.560 |
| S4 | 0.19 ± 0.15 | 0.26 ± 0.36 | 0.650 |
| S8 | 0.23 ± 0.21 | 0.19 ± 0.14 | 0.940 |
| S12 | 0.04 ± 0.06 | 0.07 ± 0.04 | 0.080 |
| Number | | | |
| T4 | 9.8 ± 4.6 | 7.0 ± 3.7 | 0.200 |
| T8 | 13.0 ± 2.7 | 8.9 ± 6.0 | 0.060 |
| T12 | 16.4 ± 5.9 | 13.6 ± 5.3 | 0.300 |
| S4 | 10.1 ± 6.6 | 7.0 ± 4.5 | 0.360 |
| S8 | 10.3 ± 8.8 | 9.5 ± 3.3 | 0.560 |
| S12 | 5.7 ± 2.9 | 8.7 ± 2.9 | 0.070 |

Values are presented as mean ± standard deviation.
Group T, Temporary anchorage device experimental group; Group S, spring experimental group; T4, Group T at 4 weeks; T8, Group T at 8 weeks; T12, Group T at 12 weeks; S4, Group S at 4 weeks; S8, Group S at 8 weeks; S12, Group S at 12 weeks.

The p-values are analyzed by Mann–Whitney U-test; *significance level of p ≤ 0.05.

than to remove the TAD and that longer than a 12-week period should elapse before the application of a reloading force.

Maino et al. evaluated the effects of a root notch caused by drills or TAD contact in two male patients and reported that the root notch partially healed with the formation of cellular cementum almost 8 weeks after root drilling. After the root notch occurred and the root was in contact with a TAD for 1 week, followed by a delay of 4 weeks, the defect was only partially healed. They concluded that a force of 200 g from an open-coil spring may disturb root healing following root damage. Similar to their results, in the current study, only partial healing of root damage related to TADs was observed after 4 weeks, and healing was reduced compared to that at 8 and 12 weeks. Total healing was not observed until 12 weeks. Furthermore, in the current study, a loading force of 150 g from cantilever springs prolonged the healing period.

Kim and Kim examined root damage caused by TAD–root contact in four mini-pigs for up to 16 weeks and evaluated the impact of leaving the TAD in contact with the teeth in situ on the healing of damaged roots. They concluded that when TAD–root contact occurs, the
0.6 mm to the root. In the current study, to prevent the possible stimulation of root absorption by TADs, the TADs were placed using a minimally invasive technique and a low torque between the middle of the roots; initial cone-beam computed tomography (CBCT) was not acquired and this was accepted as a limitation. Although we did not measure the distance between the root and TAD at baseline using CBCT, no invasion of the periodontal ligament of the roots occurred, as confirmed by radiographs and a visual inspection.

Previous research reported that root resorption due to iatrogenic trauma may differ from root resorption due to orthodontically induced inflammation. Therefore, the response to healing may vary. In the present study, the experimental model aimed to mimic accidental contact between a TAD and roots during anchorage. Thus, the current results may apply only to orthodontically induced inflammatory root resorption caused by TADs. Further studies on trauma-related root injury and inflammatory root resorption caused by TAD insertion or drilling may be needed.

Clinically, during conventional orthodontic treatment, resorption and healing can occur between appointments. A continuous orthodontic force initiates the resorption process. However, when this force is discontinued or falls below a certain level, the resorption lacuna begins to repair. In cases where root resorption is detected during orthodontic treatment, it is recommended that the application of active force be stopped. In some cases, iatrogenic injuries induced by TADs may contribute to root resorption. As shown in the present experimental model, accidental contact between the root and TAD, even when it is inserted in an ideal position, may cause root resorption. Thus, when TAD-root contact occurs, clinicians should not continue force loading on the damaged root. They should allow time for the healing process, which may require longer than 12 weeks.

Some previous studies used passive retention wires. According to Xu et al., the most serious relapse in tooth movement occurred immediately after removal of the wires, and then the tooth returned completely to its original position. In the current study, a passive wire was not applied after the spring was removed to keep the teeth in position. This was a limitation of the present study. As subsequent relapses may have increased resorption, the use of a passive wire in future studies may prevent tooth movement during retention or recovery.

According to the results of the current study, a 12-week period may not be adequate for total healing of damaged roots, and further studies with longer healing periods would be beneficial to provide more knowledge about the healing process. However, conducting studies of longer durations would likely prove difficult, as they would extend patients’ treatment periods, and the patients would obtain no additional benefit from the treatment other than contributing to science.

**CONCLUSION**

Within the limitations of this study, we conclude that force loading from springs on damaged roots caused by TAD contact during orthodontic treatment may increase damage and reduce healing compared with leaving a TAD in contact with the root in situ without force. In addition, a 12-week healing period may not be adequate for total root healing, as detected by micro-CT, and force reloading on damaged roots should not take place for longer than 12 weeks.

**CONFLICTS OF INTEREST**

No potential conflict of interest relevant to this article was reported.

**ACKNOWLEDGEMENTS**

This study was supported by TUBITAK (The Scientific and Technological Research Council of Turkey) research grant and the project number 114S936.

**REFERENCES**

1. Papadopoulos MA, Tarawneh F. The use of miniscrew implants for temporary skeletal anchorage in orthodontics: a comprehensive review. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2007;103:6-15.
2. Alves M Jr, Baratieri C, Mattos CT, Araújo MT, Maia LC. Root repair after contact with mini-implants: systematic review of the literature. Eur J Orthod 2013;35:491-9.
3. Cakir E, Malkoç S, Kirtay M. Treatment of Class II malocclusion with mandibular skeletal anchorage. Am J Orthod Dentofacial Orthop 2017;151:1169-77.
4. Reyniers RM, Ronchi L, Lalu L, Di Girolamo N, de Lange J, Roberts N, et al. Barriers and facilitators to the implementation of orthodontic mini implants in clinical practice: a systematic review. Syst Rev 2016;5:181.
5. Kim JK, Park YC, Vanarsdall RL. Applications of orthodontic mini implants. Chicago: Quintessence Publishing Company; 2007.
6. Kravitz ND, Kusnoto B. Risks and complications of orthodontic miniscrews. Am J Orthod Dentofacial Orthop 2007;131:43-51.
7. Liou EJ, Pai BC, Lin JC. Do miniscrews remain stationary under orthodontic forces? Am J Orthod Dentofacial Orthop 2004;126:42-7.
8. Kadioglu O, Büyükyilmaz T, Zachrisson BU, Maino BG. Contact damage to root surfaces of premolars touching miniscrews during orthodontic treatment. Am J Orthod Dentofacial Orthop 2008;134:353-60.

9. Maino BG, Weiland F, Attanasi A, Zachrisson BU, Büyükyilmaz T. Root damage and repair after contact with miniscrews. J Clin Orthod 2007;41:762-6; quiz 750.

10. Ahmed V KS, Rooban T, Krishnaswamy NR, Mani K, Kalladka G. Root damage and repair in patients with temporary skeletal anchorage devices. Am J Orthod Dentofacial Orthop 2012;141:547-55.

11. Demir N, Ozturk AN, Malkoc MA. Evaluation of the marginal fit of full ceramic crowns by the micro-computed tomography (micro-CT) technique. Eur J Dent 2014;8:437-44.

12. Malkoç MA, Sevimay M, Tatar İ, Çelik HH. Micro-CT detection and characterization of porosity in luting cements. J Prosthodont 2015;24:553-61.

13. Harris DA, Jones AS, Darendeliler MA. Physical properties of root cementum: part 8. Volumetric analysis of root resorption craters after application of controlled intrusive light and heavy orthodontic forces: a microcomputed tomography scan study. Am J Orthod Dentofacial Orthop 2006;130:639-47.

14. Chan EK, Darendeliler MA, Petocz P, Jones AS. A new method for volumetric measurement of orthodontically induced root resorption craters. Eur J Oral Sci 2004;112:134-9.

15. Weiland F. Constant versus dissipating forces in orthodontics: the effect on initial tooth movement and root resorption. Eur J Orthod 2003;25:335-42.

16. Malek S, Darendeliler MA, Swain MV. Physical properties of root cementum: part I. A new method for 3-dimensional evaluation. Am J Orthod Dentofacial Orthop 2001;120:198-208.

17. Cheng LL, Türk T, Elekdag-Türk S, Jones AS, Petocz P, Darendeliler MA. Physical properties of root cementum: Part 13. Repair of root resorption 4 and 8 weeks after the application of continuous light and heavy forces for 4 weeks: a microcomputed-tomography study. Am J Orthod Dentofacial Orthop 2009;136:320.e1-10; discussion 320-1.

18. Kim H, Kim TW. Histologic evaluation of root-surface healing after root contact or approximation during placement of mini-implants. Am J Orthod Dentofacial Orthop 2011;139:752-60.

19. Hembree M, Buschang PH, Carrillo R, Spears R, Rossouw PE. Effects of intentional damage of the roots and surrounding structures with miniscrew implants. Am J Orthod Dentofacial Orthop 2009;135:280.e1-9; discussion 280-1.

20. Renjen R, Maganzini AL, Rohrer MD, Prasad HS, Kraut RA. Root and pulp response after intentional injury from miniscrew placement. Am J Orthod Dentofacial Orthop 2009;136:708-14.

21. Huang LH, Shotwell JL, Wang HL. Dental implants for orthodontic anchorage. Am J Orthod Dentofacial Orthop 2005;127:713-22.

22. Lee YK, Kim JW, Baek SH, Kim TW, Chang Yl. Root and bone response to the proximity of a mini-implant under orthodontic loading. Angle Orthod 2010;80:452-8.

23. Gonzales C, Hotokezaka H, Darendeliler MA, Yoshida N. Repair of root resorption 2 to 16 weeks after the application of continuous forces on maxillary first molars in rats: a 2- and 3-dimensional quantitative evaluation. Am J Orthod Dentofacial Orthop 2010;137:477-85.

24. Rygh P. Orthodontic root resorption studied by electron microscopy. Angle Orthod 1977;47:1-16.

25. Reitan K. Effects of force magnitude and direction of tooth movement on different alveolar bone types. Angle Orthod 1964;34:244-55.

26. Öwman-Moll P, Kurol J, Lundgren D. Repair of orthodontically induced root resorption in adolescents. Angle Orthod 1995;65:403-8; discussion 409-10.

27. Xu X, Zhou J, Yang F, Wei S, Dai H. Using micro-computed tomography to evaluate the dynamics of orthodontically induced root resorption repair in a rat model. PLoS One 2016;11:e0150135.