Incidence of infections due to third generation cephalosporin-resistant Enterobacteriaceae - a prospective multicentre cohort study in six German university hospitals

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Abstract

Background: Infections caused by third generation cephalosporin-resistant Enterobacteriaceae (3GCREB) are an increasing healthcare problem. We aim to describe the 3GCREB infection incidence and compare it to prevalence upon admission. In addition, we aim to describe infections caused by 3GCREB, which are also carbapenem resistant (CRE).

Methods: In 2014–2015, we performed prospective 3GCREB surveillance in clinically relevant patient specimens (screening specimens excluded). Infections counted as hospital-acquired (HAI) when the 3GCREB was detected after the third day following admission, otherwise as community-acquired infection (CAI).

Results: Of 578,420 hospitalized patients under surveillance, 3367 had a 3GCREB infection (0.58%). We observed a similar 3GCREB CAI and HAI incidence (0.28 and 0.31 per 100 patients, respectively). The most frequent pathogen was 3GCR E. coli, in CAI and HAI (0.15 and 0.12 per 100 patients). We observed a CRE CAI incidence of 0.006 and a HAI incidence of 0.008 per 100 patients (0.014 per 1000 patient days).

Conclusions: Comparing the known 3GCREB admission prevalence of the participating hospitals (9.5%) with the percentage of patients with a 3GCREB infection (0.58%), we conclude the prevalence of 3GCREB in university hospitals to be about 16 times higher than suggested when only patients with 3GCREB infections are considered. Moreover, we find the HAI and CAI incidence caused by CRE in Germany to be relatively low.

Keywords: Fluoroquinolone, Carbapenem, Gram-negative, ESBL, Enterobacter spp., Klebsiella spp., E. coli, CRE, Hospital-acquired infections, Community-acquired infections
Background
Emerging multidrug-resistant Gram-negative bacteria are a global health concern, especially those harbouring extended-spectrum beta-lactamases (ESBL), which render Enterobacteriaceae resistant to third generation cephalosporins (3GC) and extended-spectrum penicillins [1]. Third generation cephalosporin resistant Enterobacteriaceae (3GCREB) infections are a special threat to patient safety, as resistance may cause a delay in effective antimicrobial therapy and thereby lead to worsening patient outcomes [2]. EU surveillance data shows that the 3GC resistance rate of E. coli in blood and cerebrospinal fluid samples has increased in many EU countries (EU mean 2012: 11.9%, 2015: 13.1%) [3]. The incidence density of 3GCREB in clinical specimens in German intensive care units (ICUs) rose from 2001 to 2015 (E. coli: 0.16 to 3.83/1000 patient days, K. pneumoniae: 0.25 to 1.41/1000 patient days) [4]. The percentage of hospital-acquired infections (HAI) caused by ESBL-producing Enterobacteriaceae in German ICUs and surgical departments increased as well (2007: 10.9% to 2012: 15.5%) [5].

The ATHOS (Antibiotic Therapy Optimization Study) project aimed at assessing the 3GCREB admission prevalence and 3GCREB incidence of community-acquired and hospital-acquired infections (CAI, HAI) in six German university hospitals in 2014 and 2015. The prevalence data was published previously [6]. Here, we describe the incidence of 3GCREB infections in the same hospitals and relate the data to the 3GCREB admission prevalence. Furthermore, we analyse the distribution of additional resistance phenotype patterns in those 3GCREB that caused infections.

Methods
Study design and data sources
The ATHOS project was a prospective observational cohort study that monitored hospitalized patients in general wards and ICUs for their first 3GCREB detection in clinical specimens (active surveillance). Each microbiology finding was followed by checking the health record or by contacting the wards and clinicians directly. The study was performed in six German university hospitals from January 1, 2014 to December 31, 2015. Patients hospitalized in the departments of dermatology, gynaecology/obstetrics, ophthalmology, otorhinolaryngology, paediatrics and psychiatry were excluded from surveillance.

Microbiological analysis
Gram-negative bacteria were identified down to species level using either MALDI-TOF MS or VITEK®2 (bioMérieux, Nürtingen, Germany). Antimicrobial susceptibility testing was performed using VITEK®2. Enterobacteriaceae were classified as susceptible or resistant based on minimal inhibitory concentrations according to EUCAST breakpoints [7]. Non-susceptibility was regarded as resistance. Indicator antimicrobials for third generation cephalosporin resistance were cefotaxime and ceftazidime, indicators for carbapenem resistance were imipenem and meropenem.

Definitions
A 3GCREB case was defined as the first 3GCREB isolate detected in clinical specimens (e.g. urine, wound swab, blood culture, tracheobronchial secretion or other clinical specimens) in a patient during a single hospital stay. Re-admission followed by another 3GCREB detection created a new case. Cases were distinguished in colonisations and cases with infections. Infections were defined as the detection of 3GCREB in a clinical specimen with additional signs and symptoms of infection as determined by a clinician followed by adequate antimicrobial therapy. A single case could present several 3GCREB species, each with several infections (though counting only by infection type). Acquisition was defined as follows: detection on day 1–3 (admission day = day 1) counted as community-acquired (CA), later detections counted as hospital-acquired (HA) [8]. We stratified for the following infection types: urinary tract infection (UTI), lower respiratory tract infection (LRTI), surgical site infection (SSI) and bloodstream infection (BSI). Other infection types were pooled in the category "other infections". We analysed HAI caused by HA-3GCREB and CAI caused by CA-3GCREB. Cases with an ambiguous acquisition (CA-3GCREB with HAI and HA-3GCREB on top of existing CAI) were discarded.

Statistical analysis
Infection incidence was calculated as infections per 100 patients, incidence density as infections per 1000 patient days. Both were stratified by species, resistance phenotype and infection type. Then 95% confidence intervals were calculated. The species distribution over infection types was tested with $X^2$ test ($R \times C$ table). The comparison of resistance phenotypes among 3GCREB responsible for different infection types was tested with Fisher's exact test ($2 \times 2$ table, carbapenem-resistant versus -susceptible). $P$-values < 0.05 were considered significant. Statistical analysis was performed with SAS 9.4 (SAS Institute, Cary, NC, USA) and OpenEpi (Open Source Epidemiologic Statistics for Public Health, V3.01 http://www.openepi.com).

Ethics and data protection
3GCREB surveillance was performed in accordance with the German Infection Protection Act [9]. The ethics committee at Charité, University Medicine Berlin, Germany, approved this study (EA/018/14). Data from the six hospitals was entered into an online accessible database approved by the data protection commissioner.
Results
The ATHOS project was conducted at six German university hospitals comprising a total of 283 wards and 4957 beds in the surveillance area. The majority were general wards with surgical specialty \( (n = 104) \) followed by medical specialty \( (n = 83) \), intensive care units (ICU, \( n = 49 \) ), haematology/oncology \( (n = 35) \) and intermediate care wards \( (n = 12) \).

In the period 2014–2015, 578,420 patient admissions with 3,385,112 patient days were under surveillance. After excluding invalid data and cases of colonization \( (n = 2262) \), 3367 clinical cases with one or more 3GCREB infections \( (0.58\% \text{ of the patients}) \) were analysed (Fig. 1a). The median age was 69 (IQR 58–77 years) and 55\% of the cases were male. Of the cases, 92\% had one infection, 7\% had two infections and 1\% had three or four infections.

To compare different infection types, cases were broken down into single infections and those with ambiguous acquisition were discarded, yielding 3370 single infections. The majority of infections were hospital-acquired (3GCREB HAI, Fig. 1b). We observed a difference in infection incidence among the university hospitals, e.g. the 3GCREB HAI incidence ranged from 0.17–0.42 per 100 patients. Therefore, the data of the individual hospitals was pooled for further analysis. The absolute numbers of infections stratified by species, resistance phenotype, and infection type are shown in Table 1, the infection incidences and incidence densities in Table 2. The 3GCREB CAI incidence was 0.28 per 100 patients, and that of HAI 0.31 per 100 patients. The majority of CAI were caused by 3GCREB which were also fluoroquinolone resistant (FQR, 0.17 per 100 patients), in HAI that incidence was lower (0.14 per 100 patients).

Fig. 1 Flow chart of third generation cephalosporin resistant Enterobacteriaceae (3GCREB) cases (a) and infections (b). a) Readmission followed by 3GCREB detection created a second case. b) Each case could represent several infections but only one per infection type.
Table 1: Distribution of third generation cephalosporin resistant *Enterobacteriaceae* (3GCREB) infections in the ATHOS project, 2014–2015, Germany

| Parameter               | Category | 3GCREB infections n (%) | 3GCREB CAI n (%) | 3GCREB HAI n (%) |
|-------------------------|----------|-------------------------|------------------|------------------|
|                         |          | Covers **total** cases. |                  |                  |
|                         |          | 3370 (100%)             | 1596 (100%)      | 1774 (100%)      |
| Species                 |          | E. coli                |                  |                  |
|                         |          | 1624 (48%)             | 910 (57%)        | 714 (40%)        |
|                         |          | Enterobacter spp.      |                  |                  |
|                         |          | 702 (21%)              | 221 (14%)        | 481 (27%)        |
|                         |          | Klebsiella spp.        |                  |                  |
|                         |          | 562 (17%)              | 264 (17%)        | 298 (17%)        |
|                         |          | Citrobacter spp.       |                  |                  |
|                         |          | 209 (6%)               | 76 (5%)          | 133 (7%)         |
|                         |          | Other species a        |                  |                  |
|                         |          | 273 (8%)               | 125 (8%)         | 148 (8%)         |
| Resistance              |          | all 3GCREB             |                  |                  |
|                         |          | covers **species**.    |                  |                  |
|                         |          | Only 3GCR              |                  |                  |
|                         |          | 1488 (44%)             | 580 (36%)        | 908 (51%)        |
|                         |          | + FQR                  |                  |                  |
|                         |          | 1796 (53%)             | 979 (61%)        | 817 (46%)        |
|                         |          | + CR                   |                  |                  |
|                         |          | 86 (3%)                | 37 (2%)          | 49 (3%)          |
|                         |          | E. coli                |                  |                  |
|                         |          | 1624 (100%)            | 910 (100%)       | 714 (100%)       |
|                         |          | Only 3GCR              |                  |                  |
|                         |          | 409 (25%)              | 209 (23%)        | 200 (28%)        |
|                         |          | + FQR                  |                  |                  |
|                         |          | 1203 (74%)             | 698 (77%)        | 505 (71%)        |
|                         |          | + CR                   |                  |                  |
|                         |          | 12 (1%)                | 3 (0%)           | 9 (1%)           |
|                         |          | Enterobacter spp.      |                  |                  |
|                         |          | 702 (100%)             | 221 (100%)       | 481 (100%)       |
|                         |          | Only 3GCR              |                  |                  |
|                         |          | 553 (79%)              | 168 (76%)        | 385 (80%)        |
|                         |          | + FQR                  |                  |                  |
|                         |          | 122 (17%)              | 45 (20%)         | 77 (16%)         |
|                         |          | + CR                   |                  |                  |
|                         |          | 27 (4%)                | 8 (4%)           | 19 (4%)          |
|                         |          | Klebsiella spp.        |                  |                  |
|                         |          | 562 (100%)             | 264 (100%)       | 298 (100%)       |
|                         |          | Only 3GCR              |                  |                  |
|                         |          | 157 (28%)              | 59 (22%)         | 98 (33%)         |
|                         |          | + FQR                  |                  |                  |
|                         |          | 362 (64%)              | 181 (69%)        | 181 (61%)        |
|                         |          | + CR                   |                  |                  |
|                         |          | 43 (8%)                | 24 (9%)          | 19 (6%)          |
| Infections              |          | SSI                   |                  |                  |
|                         |          | 401 (12%)              | 99 (6%)          | 302 (17%)        |
|                         |          | UTI                   |                  |                  |
|                         |          | 1528 (45%)             | 872 (55%)        | 656 (37%)        |
|                         |          | LRTI                  |                  |                  |
|                         |          | 571 (17%)              | 158 (10%)        | 413 (23%)        |
|                         |          | BSI                   |                  |                  |
|                         |          | 459 (14%)              | 220 (14%)        | 239 (13%)        |
|                         |          | Other infections b     |                  |                  |
|                         |          | 411 (12%)              | 247 (15%)        | 164 (9%)         |
| Year                    |          | 2014                  |                  |                  |
|                         |          | 1784 (53%)             | 847 (53%)        | 937 (53%)        |
|                         |          | 2015                  |                  |                  |
|                         |          | 1586 (47%)             | 749 (47%)        | 837 (47%)        |
| Ward type               |          | General ward           |                  |                  |
|                         |          | 2174 (65%)             | 1102 (69%)       | 1072 (60%)       |
|                         |          | ICU/interm. Care       |                  |                  |
|                         |          | 1196 (35%)             | 494 (31%)        | 702 (40%)        |
| Specialty               |          | Surgical              |                  |                  |
|                         |          | 1407 (42%)             | 522 (33%)        | 885 (50%)        |
|                         |          | Non-surgical           |                  |                  |
|                         |          | 1963 (58%)             | 1074 (67%)       | 889 (50%)        |

3GCREB infections among 578,420 patient admissions and 3,385,112 patient days in six German university hospitals. * Other *Enterobacteriaceae species* include Cedecea, Hafnia, Morganella, Pantoea, Proteus, Providencia, Raoultella, and Xerobacter species. * Other infections* includes all other infection types. 3GCREB = third generation cephalosporin-resistant *Enterobacteriaceae*, 3GCR third generation cephalosporin resistance, FQR fluoroquinolone resistance, CR carbapenem resistance, CA community-acquired, HA hospital-acquired, SSI surgical site infection, UTI urinary tract infection, LRTI lower respiratory tract infection, BSI bloodstream infection, ICU intensive care unit, interm. Care intermediate care. Column percentages were calculated for each parameter with respect to “3GCREB total.”

patients). 3GCREB that were also carbapenem resistant caused 0.006 CAI and 0.008 HAI per 100 patients.

Among 3GCREB species, *E. coli* caused the highest incidence of CAI (0.15 per 100 patients) and HAI (0.12 per 100 patients). The most frequent infections were UTIs irrespective of the acquisition; the incidence of 3GCREB CA-UTI exceeded that of 3GCREB HA-UTI (0.15 vs. 0.11 per 100 patients). For SSI and LRTI, the incidence of 3GCREB HAI exceeded that of CAI (Table 2).

Figure 2 shows the distribution of 3GCREB infections stratified by CAI/HAI and by species. Species distribution differed significantly in UTI, LRTI and BSI: HA-UTI, -LRTI and -BSI were caused more frequently
Table 2  Incidence (densities) of infections with third generation cephalosporin-resistant Enterobacteriaceae (3GCREB), ATHOS project, 2014–2015, Germany

| Parameter | Category | 3GCREB CAI incidence per 100,000 admissions (95% CI) | 3GCREB CAI incidence per 100 admissions (95% CI) | 3GCREB HAI incidence per 1000 admissions (95% CI) | 3GCREB HAI incidence density per 1000 patient days (95% CI) |
|-----------|----------|---------------------------------------------------|-----------------------------------------------|-----------------------------------------------|--------------------------------------------------|
| 3GCREB    | total    | 276 (263–290)                                     | 0.28 (0.26–0.29)                               | 0.31 (0.29–0.32)                               | 0.52 (0.50–0.55)                                  |
| Species   | E. coli  | 157 (147–168)                                     | 0.16 (0.15–0.17)                               | 0.12 (0.12–0.13)                               | 0.21 (0.20–0.23)                                  |
|           | Enterobacter spp. | 38 (33–44)                                  | 0.04 (0.03–0.04)                               | 0.08 (0.08–0.09)                               | 0.14 (0.13–0.16)                                  |
|           | Klebsiella spp. | 45 (40–52)                                  | 0.05 (0.04–0.05)                               | 0.05 (0.05–0.06)                               | 0.09 (0.08–0.10)                                  |
| Other species | 22 (18–26)                        | 0.02 (0.02–0.03)                               | 0.03 (0.02–0.03)                               | 0.04 (0.04–0.05)                                 |
| Resistance | all 3GCREB |                                                  |                                               |                                               |                                                  |
|           | only 3GCR | 100 (92–109)                                    | 0.10 (0.09–0.11)                               | 0.16 (0.15–0.17)                               | 0.27 (0.25–0.29)                                  |
|           | + FQR     | 169 (159–180)                                    | 0.17 (0.16–0.18)                               | 0.14 (0.13–0.15)                               | 0.24 (0.23–0.26)                                  |
|           | + CR      | 6 (5–9)                                           | 0.006 (0.005–0.009)                            | 0.008 (0.006–0.011)                            | 0.014 (0.011–0.019)                               |
| E. coli   | only 3GCR | 36 (31–41)                                       | 0.04 (0.03–0.04)                               | 0.04 (0.03–0.04)                               | 0.06 (0.05–0.07)                                  |
|           | + FQR     | 121 (112–130)                                    | 0.12 (0.11–0.13)                               | 0.09 (0.08–0.10)                               | 0.15 (0.14–0.16)                                  |
|           | + CR      | 1 (0–2)                                          | 0.001 (0.000–0.002)                            | 0.002 (0.001–0.003)                            | 0.003 (0.001–0.005)                               |
| Enterobacter spp. | only 3GCR | 29 (25–34)                                       | 0.03 (0.03–0.03)                               | 0.07 (0.06–0.07)                               | 0.11 (0.10–0.13)                                  |
|           | + FQR     | 8 (6–10)                                         | 0.01 (0.01–0.01)                               | 0.01 (0.01–0.02)                               | 0.02 (0.02–0.03)                                  |
|           | + CR      | 1 (1–3)                                          | 0.001 (0.001–0.003)                            | 0.003 (0.002–0.005)                            | 0.006 (0.003–0.009)                               |
| Klebsiella spp. | only 3GCR | 10 (8–13)                                        | 0.01 (0.01–0.01)                               | 0.02 (0.01–0.02)                               | 0.03 (0.02–0.04)                                  |
|           | + FQR     | 31 (27–36)                                       | 0.03 (0.03–0.04)                               | 0.03 (0.03–0.04)                               | 0.05 (0.05–0.06)                                  |
|           | + CR      | 4 (3–6)                                          | 0.004 (0.003–0.006)                            | 0.003 (0.002–0.005)                            | 0.006 (0.003–0.009)                               |
| Infections | SSI       | 17 (14–21)                                       | 0.02 (0.01–0.02)                               | 0.05 (0.05–0.06)                               | 0.09 (0.08–0.10)                                  |
|           | UTI       | 151 (141–161)                                    | 0.15 (0.14–0.16)                               | 0.11 (0.11–0.12)                               | 0.19 (0.18–0.21)                                  |
|           | LRTI      | 27 (23–32)                                       | 0.03 (0.02–0.03)                               | 0.07 (0.07–0.08)                               | 0.12 (0.11–0.13)                                  |
|           | BSI       | 38 (33–43)                                       | 0.04 (0.03–0.04)                               | 0.04 (0.04–0.05)                               | 0.07 (0.06–0.08)                                  |
| Other infections | 43 (38–48)                       | 0.04 (0.04–0.05)                               | 0.03 (0.02–0.03)                               | 0.05 (0.04–0.06)                                 |

a “Other Enterobacteriaceae species” include Cedecea, Hafnia, Morganella, Pantoaea, Proteus, Providencia, Raoultella, and Serratia species. b “Other infections” includes all other infection types. 3GCREB = third generation cephalosporin-resistant Enterobacteriaceae, 3GCR third generation cephalosporin resistance, FQR fluoroquinolone resistance, CR carbapenem resistance, 3GCR + CR carbapenem resistance Enterobacteriaceae (CRE), CAI community-acquired infection, HAI hospital-acquired infection, SSI surgical site infection, UTI urinary tract infection, LRTI lower respiratory tract infection, BSI bloodstream infection. 95% CI 95% confidence interval.

Discussion

The incidence of 3GCREB infections among patients admitted to German university hospitals in 2014/15 was < 1%. Additional fluoroquinolone resistance was frequent in particular in CAI, while additional carbapenem resistance was rare, both in CAI and HAI (0.006 and 0.008 per 100 patients; HAI incidence density 0.014 per 1000 patient days). An interesting finding was that among CA-LRTI caused by 3GCR Klebsiella spp., the percentage of additional carbapenem resistance (28%) was substantial and significantly higher than in 3GCR Klebsiella spp. causing HA-LRTI (6%, \( p = 0.015 \)).

In an admission prevalence study performed in parallel, we screened a minimum of 500 patients per hospital for...
rectal 3GCREB carriage on admission (day 1–3, admission day = day 1). This study showed a 3GCREB colonization prevalence of 9.5% [6]. Therefore, we conclude that the colonization rate of patients in university hospitals is about 16 times higher (=9.5%/0.58% patients with 3GCREB infection) than suggested when only patients with 3GCREB infections are considered. The reason for this high colonization prevalence and for a CAI incidence comparable to HAI is most likely the case mix in tertiary care university hospitals due to their position at the end of the treatment chain. On admission, many patients most likely have received previous antimicrobial treatment and have higher co-morbidity scores than patients in other hospital types.

The incidence of CAI causing 3GCREB that were also resistant to fluoroquinolones exceeded that of HAI (Table 2). One reason for this may be the high antibiotic use reported by patients admitted to the participating hospitals (34% of the 3GCREB-negative and 53% of the 3GCREB-positive patients) [6]. Another reason may be an enhanced use of fluoroquinolones in outpatient care [10]. This excess of fluoroquinolone use might be caused (among other reasons) by over prescription and non-adherence to antibiotic prescription guidelines in outpatient care [11–13]. A reduction of fluoroquinolone prescriptions would be desirable.

In 2013/14, the EU mean of carbapenemase-producing (CP) *E. coli* and *K. pneumoniae* was found to be 0.025 per 1000 patient days and 0.006 per 1000 patient days were reported for Germany (EuSCAPE study) [14]. We observed a CRE HAI incidence density for *Klebsiella* spp. of 0.006 (95% CI 0.000–0.008) and for *E. coli* of 0.003 per 1000 patient days (95% CI 0.001–0.005). The EuSCAPE study showed that among CRE, 70% of the *K. pneumoniae* and 30% of the *E. coli* produced carbapenemases [14]. Combining these percentages of CPE among CRE with our CRE data yields an estimated CPE incidence density for *Klebsiella* spp. and *E. coli* of 0.005 per 1000 patient days (*Klebsiella* spp. 0.004 and *E. coli* 0.001 per 1000 patient days). Thus, our data is comparable to the EuSCAPE data for Germany [14].

In contrast to other *Enterobacteriaceae* species, 3GCR *Klebsiella* spp. showed a high percentage of carbapenem resistance among CA infections, especially in LRTI and BSI. In a UK study, the prevalence of carbapenem resistance in clinically relevant *K. pneumoniae* specimens was also due primarily to community-acquired isolates (70%) [15]. The EU mean of carbapenem resistance in invasive *K. pneumoniae* isolates increased from 2012 to 2015 to 8%. In two European countries, carbapenem resistance was observed in over 25% of *K. pneumoniae* (Italy 34% and Greece 62%). In isolates from Germany, carbapenem resistance was rare (0.1%) [3]. Surveillance in German ICUs showed that the carbapenem resistance rate of *K. pneumoniae* in clinically relevant specimens increased from 2001 to 2015 to 1.5% [4]. A large admission prevalence study found a low CRE admission prevalence in Germany (comparable to UK, both 0.1%) [6, 16]. In light of this data, we conclude that even with increasing trends Germany currently still is a low CRE prevalence region.

The ATHOS project was a prospective observational study. One major limitation is the lack of patient-based information on previous healthcare contacts. Therefore, a classification into the important “healthcare-associated”
category was not possible. Instead, the somewhat arbitrary
day 3 limit for classification into CA and HA was applied,
as it is commonly used for surveillance of HAI [8]. None-
theless, we have some insight into the healthcare contacts
of our patient mix from our patients in the admission
prevalence study sample, 9% of whom had stayed in a re-
habilitation centre, 5% in a long-term care facility and 26%
in another hospital in the 6 months prior to admission [6].
Some CAI may, in fact, be healthcare-associated and thus
the incidence of CAI may be overestimated in our anal-
ysis. This CA/HA classification was also used for SSI.
We cannot exclude that some CA-SSI cases may be re-
admitted cases. An advantage of the study is the inclusion
of general wards. We can describe the incidence of
3GCREB infections in German university hospitals com-
prehensively and are not limited to ICU data. However,
due to the hospital-wide surveillance, we lack ward-
specific denominator data and are not able to calculate
department-specific incidences.

One strength of the study is the inclusion of most
Enterobacteriaceae species, since other studies are often
restricted to E. coli or K. pneumoniae. Such studies are
likely to underestimate the real incidence of CRE. We
found a major part of CRE-HAI caused by Enterobacter
group (0.006 per 1000 patient days, 95% CI 0.000–0.008).
Imipenem resistance of E. cloacae complex has increased
over the last years in German ICUs [4]. In addition,
long-term surveillance by the US Veterans Health
Administration of CRE in patients also showed a steady increase of carbapenem resistance in *E. cloacae* [17]. However, we have noticed that Vitek² frequently over-reported imipenem resistance. For future CRE studies, we suggest including *Enterobacter* spp. and using additional, more reliable diagnostic methods (e.g. disk diffusion test or agar gradient diffusion) to determine if the high frequency of Vitek² imipenem resistance represents the true epidemiology. Another drawback of using routine diagnostic methods is that we were unable to compare sequence types of 3GCREB causing CAI and HAI.

As a result of the 3GCREB admission prevalence study performed in parallel [6] and the surveillance data for infections, we can for the first time estimate the 3GCREB prevalence to be about 16-times higher than indicated by the 3GCREB infection incidence (0.58%). This will enable other university hospitals with a similar patient mix to estimate roughly the dimension of colonization prevalence present in their patients. Furthermore, we conclude that very few 3GCREB carriers are identified using clinically indicated diagnostic procedures. Therefore, we believe the majority of hospitals underestimate the extent of 3GCREB prevalence. This study gives a comprehensive description of the incidence of CRE found in German university hospitals.

**Conclusion**

Overall, our analysis showed that German university hospitals have a low 3GCREB infection incidence when compared to the admission prevalence. As we observed a comparable incidence of 3GCREB CAI and HAI, it is important that clinicians consider cephalosporin resistance in their empirical treatment decisions, irrespective of the acquisition type (CA vs. HA) of the infection.

**Abbreviations**

3GC (3GCR): 3rd generation cephalosporin (–resistant); 3GCREB: 3rd generation cephalosporin-resistant *Enterobacteriaceae*; ATHOS: Antibiotic therapy optimisation study; BSI: Bloodstream infection; CA (CAI): Community-acquired (infection); CP (CPE): Carbapenemase-producing (*Enterobacteriaceae*); CR (CRE): Carbapenemase-resistant (*Enterobacteriaceae*); *E. cloacae*: *Enterobacter cloacae*; E. coli: *Escherichia coli*; ESBL: extended spectrum β-lactamase; EU: European Union; EUCAST: European committee on antimicrobial susceptibility testing; HA (HAI): Hospital-acquired (infection); ICU: Intensive care unit; IQR: Interquartile range; *K. pneumoniae*: * Klebsiella pneumoniae*; LRTI: Lower respiratory tract infection; MALDI-TOF MS: Matrix-assisted laser desorption/ionization-time of flight mass spectrometry; spp.: species; SSA: Surgical site infection; UK: United Kingdom; US: United States; UTI: Urinary tract infection.

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**Availability of data and materials**

The data that underlie this analysis will be available from the corresponding author upon founded request in statistical analysis software format (SAS).

**Authors’ contributions**

AMR local site coordinator, local data collection, supervision of data collection in partner sites, data analysis, drafting and revising manuscript. JZ study design, surveillance protocol, local site coordinator. MWP study design, surveillance protocol, local site coordinator. FS study design, data analysis. MB database setup, supervision of data collection in partner sites. AK microbiological analysis. BO local data collection. JKMK local principal investigator, microbiological analysis. SF microbiological analysis, local data collection. CQ local data collection. FG local site coordinator. AM microbiological analysis. Vl local data collection. WS local site coordinator, local data collection. SA local data collection. SP microbiological analysis. ET local principal investigator. AH microbiological analysis. HS principal investigator of the study, study design, surveillance protocol. MJGTV local site coordination, local data collection. WPK principal investigator of the project, study design, surveillance protocol. PG principal investigator of the study, study design, surveillance protocol. All authors contributed to writing the manuscript through commenting and approved the final document.

**Ethics approval and consent to participate**

The ethics committee at Charité, University Medicine Berlin, Germany, approved this study (EA/018/14). Surveillance was performed in accordance with the German Infection Protection Act and did not require patient consent.(9).

**Consent for publication**

Not applicable.

**Competing interests**

No competing interests unless stated.

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References

1. World Health Organization: Antimicrobial resistance: global report on surveillance. In. WHO Library Cataloguing-in-Publication Data: World Health Organisation 2014.

2. Rottier WC, Ammerlaan HS, Bonten MJ. Effects of confounders and intermediates on the association of bacteraemia caused by extended-spectrum beta-lactamase-producing Enterobacteriaceae and patient outcome: a meta-analysis. J Antimicrob Chemother. 2012;67(6):1311–20.

3. European Centre for Disease Prevention and Control: Antimicrobial resistance surveillance in Europe 2015. Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). In. Stockholm: ECDC; 2017.

4. Remschmidt C, Schneider S, Meyer E, Schroener-Boersch B, Gastmeier P, Schwab F. Surveillance of antibiotic use and resistance in intensive care units (SARI). Dtsch Arztebl Int. 2017;114(50):858–65.

5. Leistner R, Schoder C, Geffers C, Breier AC, Gastmeier P, Behnke M. Regional distribution of nosocomial infections due to ESBL-positive Enterobacteriaceae in Germany: data from the German National Reference Center for the surveillance of nosocomial infections (KISS). Clin Microbiol Infect. 2015;21(3):255.e251–5.

6. Hamprecht A, Rohde AM, Behnke M, Fehl S, Gastmeier P, Gebhardt F, Kern WV, Knobloch J, Mischlak A, Obermann B, et al. Colonization with third-generation cephalosporin-resistant Enterobacteriaceae on hospital admission: prevalence and risk factors. J Antimicrob Chemother. 2016;71(10):2957–63. https://doi.org/10.1093/jac/dkw216.

7. Breakpoint tables for interpretation of MICs and zone diameters. Version 4.0. http://www.eucast.org.

8. Multidrug-Resistant Organism & Clostridium difficile Infection (MDRO/CDI) Module. https://www.cdc.gov/nhsn/pdfs/pscmanual/12pscmdro_cdadcurrent.pdf.

9. Federal Ministry of Justice and Consumer Protection: German Infection Protection Act, §23. In; 2001.

10. Dingle KE, Didelot X, Quan TP, Eyre DW, Stoesser N, Golubchik T, Harding RM, Wilson DJ, Griffiths D, Vaughan A, et al. Effects of control interventions on Clostridium difficile infection in England: an observational study. Lancet Infect Dis. 2017;17(4):411–21.

11. Povuens KB, Dolk FCK, Smith DRM, Robotham JV, Smieszek T. Actual versus ‘ideal’ antibiotic prescribing for common conditions in English primary care. J Antimicrob Chemother. 2018;73(suppl_2):19–26.

12. Smieszek T, Pouwels KB, Dolk FCK, Smith DRM, Hopkins S, Sharland M, Hay AD, Moore MV, Robotham JV. Potential for reducing inappropriate antibiotic prescribing in English primary care. J Antimicrob Chemother. 2018;73(suppl_2):i36–43.

13. Zweigner J, Meyer E, Gastmeier P, Schwab F. Rate of antibiotic prescriptions in German outpatient care - are the guidelines followed or are they still exceeded? GMS Hyg Infect Control. 2018;13 Doc04.

14. Grundmann H, Glasner C, Albiger B, Aanensen DM, Tomlinson CT, Andrae AS, Grant RP, Tan Z, Gregory AW, Giske CG, et al. Occurrence of carbapenemase-producing Klebsiella pneumoniae and Escherichia coli in the European survey of carbapenemase-producing Enterobacteriaceae (EuSCAPE): a prospective, multinational study. Lancet Infect Dis. 2017;17(2):153–63.

15. Trepanier P, Mallard K, Meunier D, Pike R, Brown D, Ashby JP, Donaldson H, Awad-El-Karrim FM, Balakrishnan L, Cubbon M et al. Carbapenemase-producing Enterobacteriaceae in the UK: a national study (EuSCAPE-UK) on prevalence, incidence, laboratory detection methods and infection control measures. J Antimicrob Chemother. 2017;72(2):596–603.

16. Otter JA, Dyakovs KN, Queral-Rubiera A, Patul A, Ahanonu C, Tosas August D, Edgeworth JD, Goldenberg SD. Universal hospital admission screening for carbapenemase-producing organisms in a low-prevalence setting. J Antimicrob Chemother. 2016;71(12):3556–61.

17. Kaase M, Schlimanski S, Schiller R, Beyer L, Thurner A, Steinmann J, Kempf VA, Hess C, Sobottka L, Femmer L, et al. Multicentre investigation of carbapenemase-producing Escherichia coli and Klebsiella pneumoniae in German hospitals. Int J Med Microbiol. 2016.