Statin drugs to reduce breast cancer recurrence and mortality

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Abstract
Epidemiologic studies have, variably, shown the concomitant use of statin drugs to be beneficial to cancer outcomes. Statin drugs have been FDA approved for three decades for the treatment of high cholesterol and atherosclerotic coronary artery disease and are widely used. This has engendered studies as to their influence on concomitant diseases, including cancers. In this context, statin use has been correlated, variably, with a decrease in deaths from breast cancer. However, there is no extant model for this effect, and the extent of efficacy is open to question. The overarching goal of this article is to communicate to the reader of the potential of statins to reduce breast cancer progression and mortality. This is the use as a secondary prevention measure, and not as a therapy to directly counter active cancer. First, salient aspects of statin pharmacology, as relates to cardiovascular disease, will be discussed. Second, the basic and clinical research studies that investigate statin usage in breast cancer will be presented. Additionally, statin effects in other cancer types will be included for context. Finally, proposals for future basic and clinical research studies to determine the role of statins in breast cancer management will be presented.

Keywords: Breast cancer, Statins, Metastasis, Lipophilicity, Prenylation, Secondary prevention

Background
Statins reduce blood cholesterol by inhibiting the rate-limiting step in the mevalonate pathway: HMG-CoA reductase (HMGCR). While this decreases the cholesterol carried in the blood, the mechanistic aspects also affect cell signaling, in a manner that could directly impact cell proliferation, and thereby cancer cells. Mevalonate pathway inhibition reduces the abundance of farnesyl pyrophosphate (FPP) and geranylgeranyl pyrophosphate (GGPP), isoprenyl groups that critically modify small signaling G proteins involved in cell proliferation, migration, and survival pathways. The influence of the statin drugs on protein prenylation is not limited to just influencing cardiovascular pathologies but can be applied in the context of other diseases, particularly cancer in which progression depends on increased migration, survival, and ultimately proliferation.

Relevant to this review, epidemiologic studies have, variably, shown the concomitant use of statin drugs to be beneficial to cancer outcomes. The aims of this article, based on literature findings both in vivo and in vitro, are to review recent studies related to statin effects on breast cancer from both a clinical and basic aspect, so as to: 1) form a consensus from the existing literature regarding the mechanisms by which statins influence cancer; and 2) establish a model of how these mechanisms influence cancer incidence, recurrence, and mortality. Existing studies suggest that statins can reduce tumor cell growth and survival, which translates to a clinical benefit of statins on reducing cancer recurrence and mortality, with limited effects on incidence of primary cancer.

Statin pharmacology and cardiovascular usage
The first statins were isolated as potent competitive inhibitors of HMG-CoA reductase (HMGCR) by Akira Endo [1], motivated by previous studies that suggested HMGCR was an important node for the regulation of de novo cholesterol synthesis (Fig. 1) [2]. After success in clinical trials, lovastatin became the first FDA-approved statin in 1987 [1, 3]. Subsequently, six other statins have garnered and maintained FDA approval: atorvastatin, fluvastatin, pitavastatin, pravastatin, rosuvastatin, and simvastatin (Additional file 1) [1]. While all statins share the same pharmacophore, their structural characteristics...
guide their binding affinity to HMGCR [4] (Additional file 2). The most biologically relevant molecules affected by inhibition of HMGCR (Fig. 1) are downstream of mevalonate—cholesterol, farnesyl pyrophosphate (FPP), and geranylgeranyl pyrophosphate (GGPP).

The seven FDA-approved statins have similar, but not identical, pharmacokinetics (Additional file 3). Statins are classified as lipophilic or hydrophilic depending on their partition coefficients, with hydrophilic statins exhibiting Log D values less than zero and lipophilic statins exhibiting Log D values greater than zero [5]. Statin uptake into cells occurs through carrier-mediated transport, most commonly by OATP1B1 with lesser contributions from OATP1B3, 1A2, and 2B1, or for lipophilic statins through passive transport across the membrane [6, 7]. Importantly, except for hepatocytes, most cell types do not express these transporters at meaningful levels [6]. This means lipophilic statins can distribute into the cells in extrahepatic tissues by diffusion across the cell membrane and will affect the cholesterol synthesis pathway in most cells, whereas hydrophilic statins are basically inert to most cells [8]. The implications for this review is that only the lipophilic statins would be expected to affect breast cancer cells if the effects on the cancer cells are direct, rather than indirect via cholesterol levels.

The primary action of statins is to reduce circulating lipids, most specifically cholesterol packaged in low density lipoprotein (LDL) particles [9]. Statins accomplish this by acting mainly in hepatocytes, the primary site of cholesterol biosynthesis [10]. Statin binding to HMGCR prevents it from converting HMG-CoA into mevalonate, which reduces the intracellular cholesterol concentration [11]. Cholesterol starvation in hepatocytes increases expression of LDLR, in turn facilitating uptake of LDL particles, which reduces blood cholesterol [12]. Thus, statins both reduce new cholesterol synthesis and enhance hepatic uptake of existing, circulating cholesterol. This aspect would represent an indirect effect on cancer cells via slightly decreasing the levels of circulating nutrient.

In addition to modifying lipid homeostasis, statins exert pleotropic effects on other aspects of cardiovascular pathobiology. Cell-mediated effects stem from reductions in cellular signaling from small G proteins through reducing the availability of FPP and GGPP [9]. For example, in endothelial cells, decreased activation of Rho enhances expression of endothelial nitric oxide synthase (eNOS), which protects endothelial function and impedes cardiovascular disease progression [13]. Thus, if the statin could gain entry (as in lipophilic statins) tumor cells would be affected.

Statin usage in the US has increased dramatically over the past several decades. Between 2003 and 2012, the percentage of adults in the US taking statins increased from 18 to 26% [14]. The clinical potency of statins is determined by the measurement of blood LDL cholesterol (LDL-C) in mg/dL [15]. Statin regimens that achieve <30%, 30–50%, or >50% reductions in LDL-C are labeled low-, moderate-, and high-intensity statin therapy, respectively [16] (Additional file 4). Statins are
the first line of treatment for hyperlipidemia and athero-
sclerotic cardiovascular disease (ASCVD) and are also
used originally as secondary and now as primary pre-
ventative therapy for patients at a high risk for develop-
ing cardiovascular disease.

**Statin usage in cancer**
This background sets the stage for the unexpected effects
statins may have on cancer. As their use is widespread,
there is a large overlap between people on chronic or
long-term statin therapy and those with cancer. Of rele-
vance to the present review are the basic and clinical
research studies that have investigated the roles of statin
drugs in cancer incidence, progression, and mortality.

**Clinical research studies**
While few prospective clinical trials have been conducted
to investigate statin therapy in cancer patients, multiple
retrospective cohort studies and meta-analyses have been
published. Several hypotheses motivate these studies. First,
since statins lower LDL-C, this may influence carcinogen-
esis and tumor progression, as rapidly dividing cells require
increased cholesterol for cell membrane synthesis [17]. In
addition, the influence of statins on reducing prenylation
may reduce signaling through pathways involved in
carcinogenesis and cancer progression [18]. Finally, statins
reduce the secretion of pro-inflammatory cytokines, which
have been shown to drive carcinogenesis and metastasis
[19]. Since these three factors influence both early (car-
cinogenesis) and late (metastasis) stages of cancer progres-
sion, clinical studies have queried the influence of statins
on cancer incidence, recurrence, and mortality (Table 1).
While data on many cancer types will be presented, the
primary cancer type discussed will be breast cancer.

**Statin influence on cancer incidence**
Many studies have investigated the influence of statin
therapy on the risk for developing cancer, with conflict-
ing results. The CARE trial in 1996 published one of the
first associations between statins and cancer incidence,
in which pravastatin users demonstrated a statistically
significant increase in breast cancer risk [20]. Similarly,
the PROSPER trial in 2002 showed a significant increase
in new cancer diagnoses in patients receiving pravastatin
compared to those on placebo [21]. In contrast, some
studies have demonstrated a decreased risk of develop-
ing cancer while taking statins. Shi and colleagues dem-
strated a reduction in the risk of liver cancer among
users of any statin drug, possibly explained by the rela-
tively higher local concentrations of statins in the liver
[22]. In addition, as both lipophilic and hydrophilic
statins gain access to hepatocellular carcinoma cells, this
may represent a special case.

The largest studies have focused on colorectal, lung,
prostate, and breast cancer. In colorectal cancer, large
retrospective cohort studies demonstrated no influence
of statins on disease incidence [23, 24]. In lung cancer, the
largest cohort and meta-analysis studies have shown no
effect of statins on cancer incidence [25, 26]. In prostate
cancer, the majority of studies that have investigated the
influence of statins on incidence find no association, espe-
cially when correcting for the rate of prostate-specific
antigen (PSA) screening [27, 28]. These studies argue for
statins not being contributory to carcinogenesis.

In breast cancer, no association between statin usage
and increased cancer risk has been reported in the litera-
ture [29]. Meta-analyses conducted by Undela et al. [30]
and Bonovas et al. [31] show no significantly different inci-
dence of invasive breast cancer in statin users across mul-
tiple studies. Other meta-analyses that investigated the
incidence of a multitude of cancers among statin users
also show no difference in breast cancer incidence with
statin therapy [32, 33]. Similarly, in the Nurses’ Health
Study, no differences in invasive breast cancer incidence
were observed between statin never users, former users,
or current users [34]. Finally, the Women’s Health Initia-
tive demonstrated no difference in incidence of breast
cancer between statin users and non-users; however, a

| Cancer type       | The effect of statin therapy on cancer incidence | Recurrence | Mortality |
|-------------------|-----------------------------------------------|------------|-----------|
| Breast            | ↔ (Hydrophilic statins)                        | ↓ (All or lipophilic statins) | ↔ (Hydrophilic statins) |
| Prostate          | ↔ (Radical prostatectomy)                     | ↓ (Radiotherapy) | ↓ (All, lipophilic, or hydrophilic statins) |
| Lung              | ↔ (Pravastatin)                               | ↓ (All statins) | ↓ (All statins) |
| Colorectal        | ↔ (Pravastatin)                               | ↓ (All statins) | ↓ (All statins) |
| Primary liver     | ↓ (All Statins)                               | ↓ (All statins or pravastatin) | ↓ (All statins or pravastatin) |

The influence of statins on cancer incidence, recurrence, and mortality for breast, prostate, lung, colorectal, and primary liver cancer. ↔ = no effect, ↓ = reduction.
All statins, hydrophilic statins, and lipophilic statins refer to studies that were conducted examining patients taking any statin, any hydrophilic statin, or any lipophilic statin, respectively.
decrease in the diagnosis of advanced stage breast cancer was seen in users of lipophilic statins [35]. These analyses echo other carcinomas in finding no effect on mammary carcinogenesis, though the decrease in advanced breast cancers may argue for reduction of tumor progression.

Statin influence on breast cancer recurrence and mortality
Fewer studies have investigated the influence of statins on cancer recurrence. While a consistent decrement in recurrence occurs in liver cancer [36], the findings for most other cancers are equivocal. In breast cancer, however, the majority of clinical evidence supports a protective effect of statins on reducing recurrence [29]. The benefits of statins on reducing breast cancer recurrence appears to be strongest in younger patients, suggesting a longitudinal influence of statin therapy [37]. Manthravadi and colleagues demonstrated a 36% reduction in the risk of breast cancer recurrence in patients taking lipophilic statins [38]. The most convincing evidence in the literature was a population study of all stage I–III breast cancer patients in Denmark conducted by Ahern and colleagues [39]. They found a 10% reduction in breast cancer recurrence among women who were prescribed a lipophilic statin (most commonly simvastatin) but a risk identical to those not on statins among women who were prescribed a hydrophilic statin [39]. These data in aggregate suggest lipophilic statins, and not hydrophilic statins, reduce the risk of breast cancer recurrence, which may indicate a role for statins as agents to impede this mortal stage of tumor progression and secondarily implicate a non-cholesterol effect such as direct tumor cell reduction of prenylation or an indirect effect via reduced inflammatory cytokine release.

Further supporting the contention that statins impact tumor progression, the majority of published data suggest that statins have a mortality benefit in most cancer types. In primary liver cancer, pravastatin and any statin usage reduces cancer-specific mortality [40–42]. Similarly, in colorectal cancer, statin usage reduces cancer-specific mortality, in particular when used either prior to diagnosis or prior to recurrence [43, 44]. In lung cancer, retrospective studies have shown that statins reduce cancer-specific mortality [45, 46]. LUNGSTAR, a phase III randomized clinical trial, found no influence of pravastatin on the survival of patients with small-cell lung carcinoma [47]. This result is partly expected if the mechanism is tumor-cell intrinsic, as pravastatin is hydrophilic, meaning its uptake by tumor cells is limited. In prostate cancer, statin usage significantly reduces cancer-specific and all-cause mortality, particularly when used after diagnosis [48, 49].

In breast cancer, the majority of studies suggest statin usage significantly reduces breast cancer-specific mortality (BCSM). A meta-analysis by Liu et al. demonstrated a 43% reduction in BCSM in women with breast cancer, which was confined to those taking lipophilic statins [50]. A nationwide cohort study in Finland by Murtola and colleagues demonstrated a 46% and 54% reduction in BCSM for pre- and post-diagnosis statin users, respectively [51]. A nationwide cohort study in the UK by Cardwell et al. demonstrated a 16% reduction in BCSM considering all statins, but a greater reduction in patients taking simvastatin, a lipophilic statin [52]. Finally, a nationwide cohort study in Scotland by McMenamin et al. demonstrated a 15% reduction in BCSM in pre-diagnosis statin users [53].

Two additional studies provide convincing data for an overall benefit of statins on all site cancer-specific mortality (ASCSM). First, a meta-analysis by Zhong et al. showed a 31% and 23% reduction in ASCSM in pre- and post-diagnosis statin users, respectively [54]. Second, a nationwide cohort study in Denmark demonstrated a 13–17% reduction in ASCSM in pre-diagnosis statin users compared to never-users [55]. These data suggest statin usage can reduce cancer-specific mortality. Coupling this with the foregoing data suggesting statins can decrease breast cancer recurrence leads to the conclusion that statins can reduce the percentage of patients who experience relapse after therapy and thereby increase breast cancer survival.

Summary
Many clinical studies have demonstrated that statins can influence carcinogenesis and cancer progression. While statins do not affect the incidence of most cancers, they do exert significant benefits on recurrence and survival in many cancer types, including breast cancer. Importantly, statin recurrence and mortality benefits are most strongly seen with lipophilic statins. These clinical studies motivate further investigation of statins in basic research studies and targeted clinical trials to evaluate potential additive or synergistic effects of statins when used with standard therapies.

Basic research studies
The clinical studies discussed above have been complemented by basic research studies that focus on the mechanisms by which statins exert anti-tumor effects. Statins may affect tumor cells directly in four main ways: 1) growth suppression, 2) apoptosis induction, 3) anti-invasive and anti-metastatic effects, and 4) anti-angiogenic effects. These would be in addition to the indirect effects of lowering available cholesterol. However, the findings that most beneficial effects of statins accrue to those on lipophilic statins suggest direct effects on the tumor cells. Interestingly, the two exceptions wherein even hydrophilic statins present benefit, liver and prostate cancers, further support the direct effect hypothesis as these cancers possess the transporters that allow cellular entry of the hydrophilic statins.
Statin suppression of cancer cell growth
Previous in vitro studies have demonstrated that statins can halt cancer cell proliferation by inducing G0/G1 or G2/M arrest. Many pathways have been described that contribute to this anti-proliferative effect. First, simvastatin has been shown to induce G1 cell cycle arrest through reduction of CDK4/6 and Cyclin D1 [56]. Additionally, BRCA1 overexpression in breast cancer cells has been shown to sensitize cells to lovastatin treatment through regulation of CDK4/Cyclin D1 [57]. Second, simvastatin, fluvastatin, and lovastatin have been shown to block the CDK2/Cyclin E-mediated G1/S transition [58]. Third, statins block the DNA-binding activity of NF-κB and Activator protein-1 (AP-1), which results in decreased transcription of genes that regulate cell proliferation [4]. Fourth, statins have been shown to inhibit proliferation of breast cancer cells by suppressing FPP and GGPP modification and activation of Ras, Rac, and Rho small GTPases [59]. Rho GTPases normally play an important role in p27 degradation, meaning inactivity results in the accumulation of p27 and cell cycle arrest [60]. Similarly, statin-mediated decreases in Ras activity result in decreased downstream signaling through Akt and diminished cell proliferation [61]. Finally, statins have been shown to inhibit DNA methyltransferases, altering gene transcription and reducing cancer cell proliferation [62]. In summary, statins act to suppress cancer proliferation by inducing cell cycle arrest, blocking proliferation signals, and influencing gene transcription.

Statin induction of cancer cell apoptosis
In addition to suppressing growth of tumor cells, some literature reports that statins can induce apoptosis. Several pathways have been described that contribute to induction of apoptosis. First, statins have been shown to decrease the protein levels of anti-apoptotic proteins such as Bcl-2 and Bcl-xL [63, 64]. Second, statins also have been shown to upregulate the activation of pro-apoptotic molecules such as Bax, Bad, and Caspases 3, 8, and 9 [63–66]. Additionally, statins have been shown to decrease phosphorylation and degradation of Bim in breast cancer cells [67], promoting apoptosis. Third, statins can induce reactive oxygen species (ROS) generation, resulting in an increase in p38 MAPK and activation of apoptotic pathways [68]. Similarly, iNOS-mediated generation of nitric oxide was shown to contribute to simvastatin- and fluvastatin-induced apoptosis in breast cancer cells [66]. Fourth, statins can activate JNK-mediated apoptotic pathways that are independent of p53 activation [69]. Finally, statins can induce calcium-dependent apoptosis through increased mitochondrial uptake of calcium through L-type calcium channels, resulting in the release of cytochrome C [70]. It is suggested that these pro-apoptotic effects are mainly mediated by a reduction in GGPP- rather than FPP-modified proteins [4].

However, other studies demonstrate no influence of statins on apoptosis. For example, simvastatin was shown to decrease proliferation of bladder cancer cells without inducing apoptosis [56]. Moreover, simvastatin at physiologically relevant concentrations was shown to protect osteosarcoma cells from oxidative stress-induced apoptosis by upregulation of Bcl-2 [71]. While atorvastatin treatment could reduce Bcl-2 expression in MDA-MB-231 breast cancer cells, two weeks of neoadjuvant atorvastatin at 80 mg/kg in human breast cancer patients was not found to alter Bax or Bcl-2 expression [69]. Moreover, other studies report minimal effects of statins on inducing cancer apoptosis [72] or a significant influence on apoptosis at concentrations much higher than currently used clinically [4]. Thus, the precise mechanism and magnitude of statin-induced apoptosis of cancer cells remains unclear.

Statin suppression of angiogenesis
Statins exert biphasic effects on angiogenesis—pro-angiogenic effects at low (nanomolar) concentrations, anti-angiogenic effects at higher (micromolar) concentrations [73]. At low concentrations, statins can induce mobilization and differentiation of endothelial progenitor cells (EPCs) through stimulation of the PI3K-Akt pathway [74]. Similarly, statin treatment can induce proliferation in EPCs through upregulation of cyclin genes and downregulation of the CDK2 inhibitor p27 [75]. Moreover, low statin concentrations can enhance endothelial cell migration and tube formation in a PI3K-Akt and endothelial nitric oxide synthase (eNOS)-dependent manner [76]. The PI3K-Akt pathways seem to be required for statin-mediated angiogenesis, as blocking PI3K eliminates pro-angiogenic effects of statins [77].

In contrast, at higher concentrations, statins negatively impact angiogenesis through multiple mechanisms. First, statins have been shown to reduce membrane localization of RhoA by blocking its geranylgeranylation, which reduced the tube-forming ability of human endothelial cells in vitro and was reversed with GGPP supplementation [78]. This inhibition of RhoA subsequently suppresses signaling through pathways that require RhoA activity, such as VEGFR, FAK, and Akt [78]. Second, statins decrease the abundance of caveolin-1 in endothelial cells, which reduces VEGFR2-mediated angiogenic signaling in a cholesterol-independent manner [79]. Third, endothelial cells treated with relatively higher doses of statins show a decrease in hypoxia-stimulated secretion of VEGF and expression of VEGFR2 [73]. Finally, simvastatin has been shown to upregulate vascular epithelial cadherin (VE-cadherin), which limits endothelial cell proliferation, migration, and tube forming ability.

In the context of cancer, statins can reduce tumor angiogenesis through several different mechanisms. First, statins can directly affect the endothelial cells to reduce...
tumor angiogenesis. For example, simvastatin treatment was shown to decrease tumor vascularization in mice at high statin doses [73]. Second, statins can act indirectly on the endothelium by reducing pro-angiogenic or increasing anti-angiogenic protein secretion by tumor cells [80]. Atorvastatin has been shown to reduce circulating VEGF concentrations in human patients with coronary artery disease [81]. Finally, simvastatin and atorvastatin can reduce matrix metalloproteinase (MMP)-9 expression in endothelial cells, which reduces their invasive capability [82]. Moreover, MMP-9 has been shown to be important for releasing the VEGF that is sequestered in breast cancer [83]. Thus, a reduction in MMP-9 impacts both the stimulation for and competency of endothelial cells to initiate angiogenesis.

**Statin suppression of cancer cell invasion and metastasis**

Many in vitro studies have suggested statins reduce the invasiveness and metastatic potential of cancer cells through multiple different mechanisms. First, statin treatment destabilizes the cytoskeletal structure of tumor cells in a RhoA/RhoC-dependent manner. Upon statin treatment, Rho delocalizes from the membrane, which causes breakdown of the actin cytoskeleton, loss of actin stress fibers and focal adhesion sites, and cell rounding [84, 85]. Moreover, statin treatment is sufficient to block EGF-mediated RhoA membrane localization and formation of actin stress fibers [86]. Second, statins block adhesion of cancer cells to both ECM proteins by reducing integrin binding activity [87]. Similarly, statins downregulate E-selectin on tumor endothelial cells, which reduces tumor cell adhesion and invasion through an endothelial barrier [88]. Third, statins reduce expression and activity of the pro-migratory proteases MMP-2, MMP-9, and urokinase by inhibiting Ras and Rho activity [84]. Fourth, statin treatment can downregulate the cancer stem cell marker CD44 in breast cancer cells, which reduces cell migration and invasion [89]. Fifth, statins can reduce expression of the transferrin receptor in breast cancer cells, which causes iron starvation and a reduction in tumor invasiveness [90]. Finally, we have shown that E-cadherin can induce tumor cell resistance to statins [91]. This suggests that statins specifically target the cells primed to undergo the epithelial–mesenchymal transition (EMT), invade, and metastasize.

Statins also impact tumor invasion and metastasis in ex vivo and in vivo models. Statins were shown to reduce prostate cancer migration towards human bone marrow stroma without impacting ECM adhesion [92]. Also, lovastatin, atorvastatin, and simvastatin have been shown to reduce metastatic seeding of melanoma and breast cancer cells in the lung and bone [85, 89, 93]. In breast cancer patients receiving 40 mg simvastatin per day for 4–6 weeks before mastectomy, statin treatment was able to decrease post-explant tumor migration [94]. Moreover, simvastatin treatment reduced the activity of Rho associated protein kinase (ROCK) and expression of RhoC, chemokine receptor type 4 (CXCR4), and CD44 [94]. In a sophisticated ex vivo microphysiological model of breast cancer metastasis to the liver, breast cancer outgrowth in response to an inflammatory lipopolysaccharide/EGF stimulus could be suppressed with atorvastatin treatment [95]. Finally, we have shown in mouse models of spontaneous breast cancer metastasis to liver and lung that atorvastatin is able to suppress metastatic proliferation but not the proliferation of the primary tumor [95]. These data suggest that statins preferentially target mesenchymal, metastatic cells.

**Summary**

Many basic research studies have shown that statins can reduce tumor cell growth and proliferation by inducing cell cycle arrest. Additionally, some, but not all, studies suggest that statins can induce apoptosis in cancer cells. In addition to affecting tumor cell growth and apoptosis, statins can negatively impact the tumor vasculature through suppression of angiogenesis. While statins exhibit a biphasic effect on angiogenesis, their anti-angiogenic effects are observed at similar doses to those used in studies that demonstrate anti-tumor effects. While statins have been shown to reduce cancer cell migration and invasion in vitro, few studies have demonstrated a reduction in metastasis with statin treatment in vivo. These previous studies motivate the investigation of statins in the context of breast cancer metastasis, in particular dormancy and emergence, which is the main cause of morbidity and mortality in this patient population. We propose a model in which statins block the secondary EMT and outgrowth of tumor cells by preventing Ras, Rac, and RhoA prenylation (Fig. 2).

**Toxicity**

An important consideration for repurposing statin drugs for breast cancer treatment is toxicity. Breast cancer patients already receive chemotherapeutic compounds that exhibit varying degrees of toxicities. Fortunately, statins are typically well-tolerated drugs. The most common adverse events associated with statin usage involve muscle tissues, which range in severity from myalgia to fatal rhabdomyolysis [8]. Muscle pain (myalgia) is relatively more common among statin users, with an incidence of approximately 5% [96]. Myopathy, or myositis, only occurs in 0.1% of patients taking statins and involves pain, weakness, and mobility restrictions that often are accompanied by an elevation in serum creatine kinase (CK) to ten times the upper limit of normal [96]. Rarely, myopathy can progress to rhabdomyolysis, which involves CK elevations above 50 times the upper limit of normal and myoglobin accumulation in the blood (myoglobinemia) and urine.
myoglobinuria), which can induce renal failure [96]. The incidence of rhabdomyolysis is extremely rare, with an incidence of 3.4 per 100,000 person years and mortality rate of approximately 10% [97]. Rhabdomyolysis incidence is higher in patients taking atorvastatin, lovastatin, and simvastatin, which is due to their extensive metabolism by CYP3A4 and potential for drug–drug interactions [98]. The less common adverse events observed with statin usage involve the liver, pancreas, kidney, and peripheral nerves.

The key concern for breast cancer patients is that the toxicity of statins leads to generalized inflammation. The initiation of inflammatory cascades can awaken dormant breast cancer cells at the site of micrometastasis to prevent their emergence to form clinically evident metastases. Moreover, patient-derived cells should be tested to validate the cell line data.

Conclusions and future directions
Previous studies have reported mechanistic and pharmacologic insight into the use of statins as anti-tumor agents, but additional questions remain regarding their use in breast cancer. First, the efficacy of statins against breast cancer cells with different molecular subtypes is not yet well understood. The susceptibility of many breast cancer cell lines—hormone receptor-positive, HER2-positive, and triple-negative—should be investigated in order to determine whether molecular subtype plays a role in statin sensitivity. Moreover, patient-derived cells should be tested to validate the cell line data.

Second, the difference in efficacy of statins against primary and metastatic tumor cells should be further investigated. Since the clinical data demonstrate statins selectively reduce breast cancer mortality and recurrence without affecting cancer incidence, this would suggest statins more selectively target metastatic cells. This may result from the evolution of tumor cells between the primary and metastatic tumor or be related to primary tumor drug penetrance. In order to better investigate the differential potency of statins against primary and metastatic tumor cells, paired biopsy samples from clinical patients should be used in in vitro, ex vivo, and in vivo models of breast cancer metastasis. This may reveal the relative importance of intrinsic differences in statin susceptibility of the primary and metastatic cells and the differences in the primary and metastatic microenvironments. Moreover, there may be a difference in statin susceptibility with the site of metastasis.

**Fig. 2 Proposed model for statin action in breast cancer.**

A) Statins Reduce Emergence of Dormant Metastatic Breast Cancer

1. Growth
2. Invasion (EMT)
3. Intravasation
4. Vascular Transit
5. Extravasation
6. Metastatic Seeding (EMT)
7. Dormancy
8. Outgrowth (2nd EMT)

**B) Statins Reduce Signaling Protein Prenylation**

HMGCR

Statin

Pre-prenylation groups (P)

Ras

Soluble Ras

EMT

Outgrowth

Erk

Mex

Raf

Pr

Farnesyl pyrophosphate and geranylgeranyl pyrophosphate, available for prenylating small G proteins, such as Ras (shown), Rac, and RhoA. Decreased prenylation reduces membrane tethering of these G proteins, which reduces downstream proliferative and pro-EMT signaling. Drawing made using images from Servier Medical Art [101].
Finally, it is critical to understand how statins interact with currently used breast cancer therapies, as statin use may either enhance or diminish their efficacy. For example, since statins work by inducing growth arrest and decreasing proliferation, the efficacy of chemotherapeutic agents may be compromised since they are only effective against cycling cells. If chemotherapeutic efficacy is compromised by statins, patients should stop taking their statin during chemotherapy cycles. In contrast, statins have been shown to induce oxidative stress in cancer cells, which may compromise the health of tumor cells and make them more susceptible to cytotoxic drugs. If these findings hold true, patients who don’t normally take statins should supplement their chemotherapy regimen with a lipophilic statin. Finally, since most statins are metabolized by CYP450 isozymes, they may interfere with the metabolism of chemotherapeutic drugs and possibly induce additional adverse events. Examining statins in combination with other chemotherapeutic drugs in ex vivo liver culture systems and in vivo will suggest which drug combinations may result in higher clinical efficacy or toxicity.

Prospective clinical trials should investigate statin use as adjuvant therapy in breast cancer. Since the studies that demonstrate a cancer mortality benefit with statin use are quite large, prospective clinical trials would need to be conducted in a targeted patient population. For example, investigating adjuvant statin therapy in triple-negative or HER2-positive patients would be more feasible than a trial of hormone receptor-positive patients since the former two groups typically relapse earlier. Moreover, patients with nodal metastases or locally invasive primary tumors would be prime candidates for these trials, as they are at the highest risk for metastatic relapse. Patients would be randomized to a lipophilic statin (likely atorvastatin) or placebo and followed longitudinally. These trials would contain defined interim analysis periods to allow patients to crossover from placebo to statin should the statin recurrence or mortality benefits prove significant.

Future basic research studies and clinical trials that build upon the results of past studies have the potential to establish statins as effective and safe agents to delay breast cancer recurrence and mortality. In the short term, studies should address whether current statin users for cardiovascular indications with a history of breast cancer should be uniformly prescribed lipophilic statins. In the long term, clinical trials should address whether statins play a role in adjuvant management of breast cancer in women who do not otherwise require a statin. It is hoped that accomplishing both of these short- and long-term goals will improve the quality of life and survival rates of women with breast cancer.

Additional files

**Additional file 1:** Chemical structures of HMG-CoA, compactin, and FDA-approved statins. The chemical structures of HMG-CoA, compactin, the first derived statin, and the seven FDA-approved statin drugs. Dates in parentheses indicate the initial approval date by the FDA, according to their website [102]. Drug structures were drawn using ChemDoodle Sketcher [103]. (PPTX 6215 kb)

**Additional file 2:** Affinity of HMG-CoA and statins for HMGCR. The affinity (Kd) or mean inhibitory concentration (Ki) of HMG-CoA and statins, respectively, for HMGCR. The statin prefixes are used instead of the whole name (e.g., Atorva = Atorvastatin). Affinity and mean inhibitory concentration values are reported in nanomoles (nM) [104–106]. (PPTX 45 kb)

**Additional file 3:** Pharmacokinetic properties of the seven FDA-approved statins. Absorption, distribution, metabolism, and excretion parameters for each statin are listed. The statin prefixes are used instead of the whole name (e.g., Atorva = Atorvastatin). Times are represented in hours (h). Log D = distribution coefficient at pH 7.4 (higher value means more lipophilic); T1/2, time after oral dose to maximum serum concentration; CYP450 cytochrome P450; T1/2, half-life. Data were acquired from Schachter [8], McKenney et al. [5], and Davidson et al. [107]. (PPTX 57 kb)

**Additional file 4:** High-, moderate-, and low-intensity statin therapy. Statin dosages that achieve plasma reductions in LDL-C of < 30% (low-intensity), between 30 and 50% (moderate-intensity), and > 50% (high-intensity), as tested in randomized control trials (RCTs) and reported in the 2013 American College of Cardiology (ACC) and American Heart Association (AHA) 2013 guidelines [16]. Statin dosages reported in italics are FDA-approved but have not been tested in RCTs. BID twice daily, XL extended release. (PPTX 51 kb)

**Abbreviations**

ASCVD: Atherosclerotic cardiovascular disease; BCSM: Breast cancer-specific mortality; ECM: Extracellular matrix; EMP: Epithelial-Mesenchymal Plasticity; FPP: Farnesyl pyrophosphate; GGPP: Geranylgeranyl pyrophosphate; HMGCR: HMG-CoA reductase; LDL: Low density lipoprotein; LDL-C: LDL cholesterol; LDL-R: Low density lipoprotein receptor; MIF: Matrix metalloproteinase; PCSK9: Proprotein convertase subtiliskin/Kexin type 9

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**Availability of data and materials**

Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

**Authors’ contributions**

CHB drafted the manuscript and all authors reviewed and revised the manuscript. All authors read and approved the final manuscript.

**Ethics approval and consent to participate**

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**Competing interests**

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References
1. Endo A. A historical perspective on the discovery of statins. Proc Japan Acad Ser B. 2010;86(5):484–93. https://doi.org/10.2183/pjpb.86.484.
2. Siperein MD, Fagan VM. Feedback control of mevalonate synthesis by dietary cholesterol. J Biol Chem. 1966;241(3):602–9 http://www.jbc.org/content/241/3/602.long.
3. Tjebbe JA. Lovastatin and beyond: the history of the HMG-CoA reductase inhibitors. Nat Rev Drug Discov. 2003;2(7):517–26. https://doi.org/10.1038/nrd1112.
4. Gazzero P, Proto MC, Gangemi G, et al. Pharmacological actions of statins: a critical appraisal in the management of cancer. Pharmacol Rev. 2012;64(1):102–46. https://doi.org/10.1123/pr.2011.004994.
5. McKinney JM, Ganz P, Wiggins BS, Saseen JS. Statins. In: Ballantyne CM, editor. Clinical lipopidology: a companion to Braunwald’s heart disease: Elsevier; 2009. p. 253–80. https://doi.org/10.1016/S0978-14160546-9.60026-3.
6. Kallikoski A, Niemi M. Impact of OATP transporters on pharmacokinetics. Br J Pharm. 2009;158(3):695–705. https://doi.org/10.1111/j.1476-5381.2009.04040.x.
7. Hu M, Cheung BMY, Tolminson B. Safety of statins: an update. Ther Adv Drug Saf. 2012;3(3):133–44. https://doi.org/10.1177/2042090912439884.
8. Schachter M. Chemical, pharmacokinetic and pharmacodynamic properties of statins: an update. Fundam Clin Pharmaco. 2005;19(1):117–25. https://doi.org/10.1002/fcp.271.
9. Stancu C, Sima A. Statins: mechanism of action and effects. J Cell Mol Med. 1997;32(5):403–20. https://doi.org/10.1111/j.1873-2880.1997.tb00095.x.
10. Charlton-Menys V, Durrington PN. Human cholesterol metabolism and cholesterol transport in carcinogenesis: a review of scientific findings, critical appraisal in the management of cancer. Pharmacol Rev. 2012;64(1):80–101. https://doi.org/10.1123/pr.2011.005399.
11. Undelka K, Srikant V, Bansal D. Statin use and risk of breast cancer: a meta-analysis of observational studies. Breast Cancer Res Treat. 2012;135(1):261–9. https://doi.org/10.1007/s10549-012-1514-x.
12. Bonovas S, Filoussi K, Tsavsis N, Sitaras NM. Statins and cancer risk: a literature-based meta-analysis and meta-regression analysis of 35 randomized controlled trials. J Clin Oncol. 2006;24(30):4808–17. https://doi.org/10.1200/JCO.2006.35560.
13. Dale KM, Coleman CI, Henny JB, et al. Statins and cancer risk. JAMA. 2006;295(1):74–8. https://doi.org/10.1001/jama.295.1.74.
14. Koopmans J, Lamminpaa A, Pulkka E. Statins and cancer: A systematic review and meta-analysis. Eur J Cancer. 2008;44(15):2122–32. https://doi.org/10.1016/j.ejca.2008.06.025.
15. Borquist S, Tamimi RM, Chen WY, et al. Statin use and breast cancer risk in the Nurses’ Health Study. Cancer Epidemiol Biomark Prev. 2016;25(1):120–1. https://doi.org/10.1158/1055-9966.EPI-15-0599.
16. Desai P, Lehman A, Chlebowski RT, et al. Statins and breast cancer stage and mortality in the Women’s Health Initiative. Cancer Causes Control. 2015;26(4):529–39. https://doi.org/10.1007/s10552-015-0530-7.
17. Wu C-Y, Chen Y-L, Ho HJ, et al. Association between nucleoside analogues and risk of hepatitis B virus-related hepatocellular carcinoma recurrence following liver resection. JAMA. 2012;308(18):1906–14 http://www.ncbi.nlm.nih.gov/pubmed/23162861.
18. Sakellakis M, Akinosoglou K, Kostaki A, et al. Statins and risk of breast cancer recurrence. Breast Cancer (Dove Med Press). 2016;8:199–205. https://doi.org/10.2147/BCTT.S101107.
19. Bravata DM, Pignone MP, O’Connor MG, et al. The role of statin therapy for the primary and secondary prevention of cardiovascular disease. Ann Intern Med. 2002;136(11):847–60. https://doi.org/10.7326/0003-4819-136-11-200204160-00007.
20. Stone NJ, Robinson JG, Lichtenstein AH, et al. 2013 ACC/AHA guideline on the treatment of blood cholesterol to reduce atherosclerotic cardiovascular risk in adults. J Am Coll Cardiol. 2014;63(25):2898–938. https://doi.org/10.1016/j.jacc.2013.11.002.
21. Cruz PM, Mo H, McConathy WJ, et al. The role of cholesterol metabolism and cholesterol transport in carcinogenesis: a review of scientific findings, relevant to future cancer therapeutics. Front Pharmacol. 2013;4 https://doi.org/10.3389/fphar.2013.00119.
22. Zhou Q, Liao JK. Statins and cardiovascular diseases: from cholesterol lowering to pleiotropy. Curr Pharm Des. 2009;15(5):467–78 http://www.ncbi.nlm.nih.gov/pubmed/19199975.
23. Landskron G, De la Fuente M, Thuvajit P, et al. Chronic inflammation and cytokines in the tumor microenvironment. J Immunol Res. 2014;2014:1–19. https://doi.org/10.1155/2014/479185.
24. Sacks FM, Pfeffer MA, Moye LA, et al. The effect of pravastatin on coronary events after myocardial infarction in patients with average cholesterol levels. N Engl J Med. 1996;335(14):1001–9. https://doi.org/10.1056/NEJM199610033351401.
systematic review and meta-analysis. Cancer Epidemiol. 2016;45:71–81. https://doi.org/10.1016/j.canep.2016.10.004.

45. Lin JJ, Ezer N, Sigel K, et al. The effect of statins on survival in patients with stage IV lung cancer. Lung Cancer. 2016;99:137–42. https://doi.org/10.1016/j.lungcan.2016.07.006.

46. Hung M-S, Chen I-C, Lee C-P, et al. Statin improves survival in patients with EGFR(T)K lung cancer: A nationwide population-based study. Sougakos J, ed. PLoS One. 2017;12(12):e0171137. https://doi.org/10.1371/journal.pone.0171137.

47. Seckl MJ, Ottensmeier CH, Cullen M, et al. Multicenter, phase III, randomized, double-blind, placebo-controlled trial of pravastatin added to first-line standard chemotherapy in small-cell lung cancer (LUNGSTAR). J Clin Oncol. 2017;35(14):1506–14. https://doi.org/10.1200/JCO.2016.69.7991.

48. Tan P, Wei S, Yang L, et al. The effect of statins on prostate cancer recurrence and mortality after definitive therapy: a systematic review and meta-analysis. Sci Rep. 2016;6(1):29106. https://doi.org/10.1038/srep29106.

49. Larsen SB, Dehliendorf C, Skriver C, et al. Postdiagnosis statin use and mortality in Danish patients with prostate cancer. J Clin Oncol. 2017;35(29):3290–7. https://doi.org/10.1200/JCO.2017.71.8891.

50. Liu B, Yi Z, Guan X, et al. The relationship between statins and breast cancer prognosis varies by statin type and exposure time and type of analysis. Breast Cancer Res Treat. 2017;164(1):1–11. https://doi.org/10.1007/s10549-017-4246-0.

51. Murtola TJ, Visvanathan K, Antama M, et al. Statin use and breast cancer survival: a nationwide cohort study from Finland. PLoS One. 2014;9(10):e110231. https://doi.org/10.1371/journal.pone.0110231.

52. Cardwell CR, Hicks BM, Hughes C, Murray LJ. Statin use after diagnosis of breast cancer and breast cancer survival: a population-based cohort study. Epidemiology. 2015;26(2):168–78. https://doi.org/10.1097/EDE.0000000000000185.

53. Mc Menamin ÚC, Murray LJ, Hughes CM, Cardwell CR. Statin use and breast cancer survival: a nationwide cohort study in Scotland. BMC Cancer. 2016;16(1):600. https://doi.org/10.1186/s12885-016-1665-0.

54. Zhong S, Zhang X, Chen L, et al. Statin use and mortality in cancer patients: Systematic review and meta-analysis of observational studies. Cancer Treat Rev. 2015;41(6):554–67. https://doi.org/10.1016/j.ctrv.2015.04.005.

55. Nielsen SF, Nordestgaard B, Bojesen SE. Statin use and reduced cancer-related mortality. N Engl J Med. 2012;367(19):1792–802. https://doi.org/10.1056/NEJMoa1217355.

56. Wang G, Cao R, Wang Y, et al. Simvastatin induces cell cycle arrest and inhibits proliferation of bladder cancer cells via PPARY signalling pathway. Sci Rep. 2016;6(1):15782. https://doi.org/10.1038/srep15782.

57. Yu X, Luo Y, Zhou Y, et al. BRCA1 overexpression sensitizes cancer cells to lovastatin via regulation of cyclin D1-CDK4-p27KIP1/CDK1/CDK2 pathway: analyses using a breast cancer cell line and tumoral xenograft model. Int J Oncol. 2008;33(3):555–63. http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1895886.

58. Sivaprasad U, Abbas T, Dutta A. Differential efficacy of 3-hydroxy-3-methylglutaryl CoA reductase inhibitors on the cell cycle of prostate cancer cells. Mol Cancer Ther. 2006;5(9):2310–6. https://doi.org/10.1158/1535-7163.MCT-05-0175.

59. Gbelcová H, Rimpelová S, Ruml T, et al. Variability in statin-induced changes following statin treatment in breast cancer. Cancer Res Treat. 2017;164(1):1–7. https://doi.org/10.1007/s10597-016-0786-8.

60. Weis M, Hesse M, Glassford AJ, Cooke JP. Statins have biphasic effects on angiogenesis. Circulation. 2002;105(6):739–45. http://www.ncbi.nlm.nih.gov/pubmed/11839631.

61. Beckwitt CH, Shiraha K, Wells A. Lipophilic statins limit cancer cell growth by inducing apoptosis correlated with activation of p53 and down-regulation of BCL-2 gene expression. Int J Oncol. 2012;40(4):935–41. https://doi.org/10.3892/ijo.2011.1273.

62. Banarait B, Suwananloet W, Naowaboot J. Simvastatin potentiates doxorubicin activity against MCC-7 breast cancer cells. Oncol Lett. 2017;14(5):6243–50. https://doi.org/10.3892/ol.2017.7683.
by 3-hydroxy-3-methylglutaryl-coenzyme A reductase inhibitors. Cancer Res. 2001;61(12):4885–91. http://www.ncbi.nlm.nih.gov/pubmed/11406567.

87. Glynn SA, O’Sullivan D, Eustace AJ, et al. The 3-hydroxy-3-methylglutaryl-coenzyme A reductase inhibitors, simvastatin, lovastatin and mevastatin inhibit proliferation and invasion of melanoma cells. BMC Cancer. 2008;8(1):9. https://doi.org/10.1186/1471-2407-8-9.

88. Nübel T, Dippold W, Kleinert H, et al. Lovastatin inhibits Rho-regulated expression of E-selectin by TNFα and attenuates tumor cell adhesion. FASEB J. 2004;18(1):140–2. https://doi.org/10.1096/fj.03-0261fje.

89. Mandal CC, Ghosh-Choudhury N, Yonedo T, et al. Simvastatin prevents skeletal metastasis of breast cancer by an antagonistic interplay between p53 and CD44. J Biol Chem. 2011;286(13):11314–27. https://doi.org/10.1074/jbc.M110.193714.

90. Kanegane AK, Gollavilli PN, Vasamsetti SB, et al. Statin-induced inhibition of breast cancer proliferation and invasion involves attenuation of iron transport: intermediacy of nitric oxide and antioxidant defence mechanisms. Febs J. 2014;281(16):3719–38. https://doi.org/10.1111/febs.12893.

91. Beckwitt CH, Clark AM, Ma B, et al. Statin-induced mevalonate pathway inhibition attenuates the growth of mesenchymal-like cancer cells that lack functional E-cadherin mediated cell cohesion. Sci Rep. 2014;4:7593. https://doi.org/10.1038/srep07593.

92. Brown M, Hart C, Tawadros T, et al. The differential effects of statins on the metastatic behaviour of prostate cancer. Br J Cancer. 2012;106(10):1689–96. https://doi.org/10.1038/bjc.2012.138.

93. Collison EA, Kleer C, Wu M, et al. Atorvastatin prevents Rhoc isoprenylation, invasion, and metastasis in human melanoma cells. Mol Cancer Ther. 2003;2(10):941–8. http://www.ncbi.nlm.nih.gov/pubmed/14578459.

94. Yulian ED, Bustami A, Dosan R, et al. Simvastatin inhibits human breast cancer migration through Rho/Rho-associated coiled-coil-containing protein kinase signaling. Adv Sci Lett. 2017;23(7):6789–94. https://doi.org/10.1166/asl.2017.9398.

95. Beckwitt CH, Clark AM, Ma B, et al. Statins attenuate outgrowth of breast cancer metastases. Br J Cancer. 2018;119:1094–1105.

96. Jacobson TA. Toward “pain-free” statin prescribing: clinical algorithm for diagnosis and management of myalgia. Mayo Clin Proc. 2008;83(6):687–700. https://doi.org/10.4065/83.6.687.

97. Law M, Rudnicka AR. Statin safety: a systematic review. Am J Cardiol. 2006;97(8):552–60. https://doi.org/10.1016/j.amjcard.2005.12.010.

98. Maje D, Shahid S, Solanki D, Gaurav K. Safety of statins. Indian J Endocrinol Metab. 2013;17(4):536. https://doi.org/10.4103/2230-8210.113754.

99. Khazali AS, Clark AM, Wells A. Inflammatory cytokine IL-8/CXCL8 promotes tumour escape from hepatocyte-induced dormancy. Br J Cancer. 2018;118(4):566–76. https://doi.org/10.1038/bjc.2017.414.

100. Taylor DP, Clark A, Wheeler S, Wells A. Hepatic nonparenchymal cells drive metastatic breast cancer outgrowth and partial epithelial to mesenchymal transition. Breast Cancer Res Treat. 2014;144(3):551–60. https://doi.org/10.1007/s10549-014-2875-0.

101. Beckwitt et al. Breast Cancer Research (2018) 20:144

102. FDA Drug Approval Dates. https://www.fda.gov/Drugs/InformationOnDrugs/ucm079874.htm. Accessed 25 Sept 2018.

103. iChemLabs LLC. ChemDoodle Sketcher. https://web.chemdoodle.com/demos/sketcher/. Accessed 25 Sept 2018.

104. Mukhtar RYA, Reid J, Reckless JPD. Pitavastatin. Int J Clin Pract. 2005;59(2):239–52. https://doi.org/10.1111/j.1742-1241.2005.00461.x.

105. McKenney JM. Pharmacologic characteristics of statins. Clin Cardiol. 2003;26(13):832–8. http://www.ncbi.nlm.nih.gov/pubmed/12708637.

106. Holdgate GA, Ward WH, McTaggart F. Molecular mechanism for inhibition of 3-hydroxy-3-methylglutaryl CoA (HMG-CoA) reductase by rosuvastatin. Biochem Soc Trans. 2003;31(Pt 3):3528–31. https://doi.org/10.1042/bst0310528.

107. Davidson MH, Robinson JG. Lipid-lowering effects of statins: a comparative review. Expert Opin Pharmacother. 2006;7(13):1701–14. https://doi.org/10.1517/14656566.7.13.1701.