**In vitro** Antioxidant and Anti-obesity Activities of Freeze-dried *Canarium sp.*, *Averrhoa bilimbi* L. and *Malus domestica*

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Received date: Jun 25, 2019; Revised date: Sep 5, 2019; Accepted date: Sep 9, 2019

**BACKGROUND:** The number of obesity cases is still increasing worldwide and has reached an epidemic scale. Plants are known to have a protection role in the development of obesity, however their antioxidant and anti-obesity activities have not widely known. This study was conducted to assess the *in vitro* antioxidant and anti-obesity activities of different types of freeze-dried fruits cultivated in Indonesia, especially *Canarium sp.*, *Averrhoa bilimbi* L. and *Malus domestica*.

**METHODS:** Total phenolic content of freeze-dried fruits was identified by Folin-Ciocalteu method, while the total flavonoid content was measured by aluminium chloride colorimetric assay. To assessed the antioxidant activity, 2,2-diphenyl 1-picrylhydrazyl (DPPH) scavenging activity assay and 2,2’-azino-bis (3-ethylbenothiazoline-6-sulphonic acid) (ABTS) reducing activity assay were performed. The α-amylase and lipase inhibitory activity assay were performed to assess the anti-obesity activity. For comparison, hydroxycitric acid (HCA) compound was also assessed with DPPH, ABTS, α-amylase and lipase assays.

**RESULTS:** *A. bilimbi* had the highest total phenol content (6.35 µg GAE/mg), meanwhile *M. domestica* had the highest total flavonoid content (2.06 µg QE/mg). *A. bilimbi* also showed the highest antioxidant activity both in DPPH and ABTS assay, with inhibitory concentration (IC$_{50}$)=279.99 µg/mL and 631.78 µg/mL, respectively. The freeze-dried *M. domestica* had the highest anti-α-amylase activity (IC$_{50}$=258.85 µg/mL), while *Canarium sp.* had the highest anti-lipase activity (IC$_{50}$=118.66 µg/mL).

**CONCLUSION:** Freeze-dried fruits demonstrate *in vitro* benefits toward obesity. *A. bilimbi* has potent antioxidant activity and is beneficial against obesity-related adverse health effect by relieving oxidative stress. *M. domestica* and *Canarium sp.* hamper the fat accumulation by reducing the carbohydrate absorption and dietary lipid.

**KEYWORDS:** antioxidant, anti-obesity, *Canarium sp.*, *Averrhoa bilimbi* L., *Malus domestica*, hydroxycitric acid

Indones Biomed J. 2019; 11(3): 320-6

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Indones Biomed J. 2019; 11(3): 320-6

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**Introduction**

Obesity is defined as excessive body fat accumulation, which increase the risk of adverse health effect. As described by World Health Organization (WHO), people with body mass index (BMI) over 30 kg/m² are diagnosed as obese.(1) In Indonesia, the BMI of 22.9 kg/m² is considered to be the normal weight.(2)

In recent years, the number of obesity cases is still increasing worldwide up to 650 million in adults (3), and has reached the epidemic scale, especially in high-income
countries (4). Obesity is the main preventable cause of death. An obese person has a higher risk of developing various adverse health effects, including cardiovascular diseases and diabetes.(5) There are multiple risk factors ranging from an unhealthy lifestyle to genetics that contribute to the pathogenesis of obesity.(6) Oxidative stress, a condition caused by excessive production of reactive oxygen species (ROS), plays an important role in the development of obesity and its adverse health effect.(7,8)

Plants are the source of multitude phytochemical compounds that are useful in various biological activities. Previous study found that several plant extracts were able to prevent fat absorption through the inhibition of pancreatic lipase.(9) Other study demonstrated that plant extract could protect an animal model from hyperglycemia.(10) Nowadays, the use of fruits continues to be developed, considering its less side effects and toxicity.

Despite its many benefits, fruits are easily spoiled. Thus, diverse methods are found to preserve fruits, one of which is the freeze-drying technique.(11) Freeze-drying technique is known to have many advantages to maintain fruits and its phytochemicals quality. By using the technique, the colour stability and anthocyanin content in fruit are able to be preserved.(12)

Unfortunately, the antioxidant and anti-obesity activities of several common fruits found in Indonesia, such as Canarium sp. (galip nut), Averrhoa bilimbi L. (star fruit) and Malus domestica (apple), especially when preserved samples were washed and milled, before being freeze-dried using the freeze drier (Nanbei, Henan, China), which resulted in 28.22 gram freeze-dried Canarium sp., 9.98 gram freeze-dried A. bilimbi and 57.6 gram freeze-dried M. domestica.

For comparison, hydroxycitric acid (HCA) compound (Cat #59847-5G, Sigma Aldrich, Missouri, USA), a derivative of citric acid which is found in some tropical plants, was also prepared and further analysed.

The Identification of Total Phenolic Content
The total phenolic content in freeze-dried fruit samples was evaluated using the modified colorimetric method. The method was marked by the reduction of Folin-Ciocalteu reagent (Cat #1.09001.0500, Merck, Darmstadt, Germany) by phenolic compounds with a concomitant formation of a blue complex. Each freeze-dried fruit samples and 100 µL of gallic acid standard was mixed with 75 µL of 10% Follin-Ciocalteau reagent and 60 µL of 7.5% Na₂CO₃ in the microplate, and then was incubated at the temperature of 45-50°C for 10 minutes. The absorbance was measured in 750 nm of wavelengths using the microplate reader.(13,14)

The Identification of Total Flavonoid Content
The total flavonoid content of freeze-dried fruits was measured by aluminium chloride colorimetric assay. Briefly, each freeze-dried fruit samples and 250 µL of standard was mixed with 75 µL of 5% NaNO₂. After 6 minutes, 150 µL of 10% AlCl₃·H₂O solution was added. After another 5 minutes, 0.5 mL of 1M NaOH solution was added to the mixture. The final mixture was measured using the microplate reader absorbance at 510 nm of wavelength.(15)

The 2,2-diphenyl-1-picrylhydrazyl (DPPH) Scavenging Activity Assay
To evaluate the antioxidant activity, the DPPH assay was conducted. Fifty µL of freeze-dried fruit samples and HCA at various concentrations were added to the 96-well microplate. Two-hundred µL of DPPH solution (Cat #D913-2, Sigma Aldrich) was then added into each well, and was incubated at 20-25°C in the darkroom for 30 minutes. The absorbance was measured at 517 nm of wavelengths using Multiskan™ GO Microplate Spectrophotometer (Thermo Scientific, Massachusetts, USA). A 250 µL of DPPH solution was used as the negative control, while 250 µL of methanol was used as the blank control. The scavenging activity was measured based on the sample DPPH absorption rate in comparison to negative control.(14,16)

The 2,2’-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid (ABTS)-reducing Activity Assay
Two µL of each freeze-dried fruit samples and HCA were added to the sample well, then 198 µL of ABTS solution (Cat #A1888-2G, Sigma Aldrich) was added to each well,
and was incubated at 30°C for 6 minutes. The absorbance was measured every 1 minute at 420 nm of wavelength using the Multiskan™ GO Microplate Spectrophotometer reader (Thermo Scientific) for 8 minutes. This method was done to evaluate the antioxidant activity.

**The α-amylase Inhibitory Activity Assay**

To assess the anti-obesity activity of each freeze-dried fruits and HCA, α-amylase inhibitory activity assay were performed. Thirty µL of each freeze-dried fruit samples and HCA were added into the sample and negative control well, while 30 µL of dimethyl sulfoxide (DMSO) was served as the blank control. Ten µL of α-amylase enzyme (Cat #7595-50ML, Sigma Aldrich) was added into each well, except for the blank control well. After incubated at 37°C for 10 minutes, 40 µL starch solution was added to each well except for the negative control well. Forty µL of phosphate buffer was then added to the negative control well, and was incubated at 37°C for 15 minutes. The enzymatic reaction was terminated by the addition of 100 µL of acidic iodine solution in each well. The absorbance was measured at 565 nm of wavelength using the Multiskan™ GO Microplate Spectrophotometer (Thermo Scientific).

**The Lipase Inhibitory Activity Assay**

Lipase inhibitory activity assay to evaluate the anti-obesity activity was performed based on following procedures. Twenty mL of each freeze-dried fruit samples and HCA, 20 mL of porcine pancreas lipase (PPL) enzyme (Cat #L3126-100G, Sigma Aldrich), and 100 mL of potassium phosphate buffer were added to the 96-well microplate, and then was incubated at 37°C for 1 hour. Furthermore, 10 mL of nitrophenyl butyrate (NPB) (Cat #N9876-1G, Sigma Aldrich) substrate was added, and was incubated at 37°C for 5 minutes. The absorbance was measured at 405 nm of wavelength.

**Results**

**Total Phenolic Content**

Total phenolic content of each freeze-dried fruits was evaluated. The result was expressed in µg gallic acid equivalent (GAE)/mg. Freeze-dried *A. bilimbi* had the highest level of phenolic content compared to the other two freeze-dried fruits, with the concentration of 6.35±0.95 µg GAE/mg. Meanwhile, the lowest concentration of phenol was found in the freeze-dried *M. domestica* 2.46±0.93 µg GAE/mg (Table 1).

**Total Flavonoid Content**

The flavonoid content of freeze-dried fruits was also evaluated. The result was expressed in µg quercetin equivalent (QE)/mg. Freeze-dried *M. domestica* had the highest level of flavonoid content compared to the other freeze-dried fruits, with concentration of 2.06±0.46 µg QE/mg, while the lowest was the freeze-dried *Canarium* sp., with the concentration of 0.04±0.32 µg QE/mg (Table 1).

| Freeze-dried Fruit Sample     | Total Phenolic Content (µg GAE/mg) | Total Flavonoid Content (µg QE/mg) |
|-------------------------------|-----------------------------------|-----------------------------------|
| *Canarium* sp.                | 6.07±0.50                         | 0.04±0.32                         |
| *Averrhoa bilimbi* L.         | 6.35±0.95                         | 0.29±0.48                         |
| *Malus domestica*             | 2.46±0.93                         | 2.06±0.46                         |

*The data was presented as mean±standard deviation.

**DPPH Scavenging Activity**

The antioxidant activity of each freeze-dried fruits and HCA based on DPPH scavenging assay were evaluated. Based on the result, freeze-dried *A. bilimbi* had the highest DPPH scavenging activity (49.73%) when compared to the freeze-dried *Canarium* sp., freeze-dried *M. domestica* and HCA (Figure 1A). Based on the inhibitory concentration (IC$_{50}$) values, freeze-dried *A. bilimbi* also showed the highest scavenging activity (279.99 µg/mL), compared to the other three compounds (Table 2).

**ABTS-reducing Activity**

The antioxidant activities of each freeze-dried fruits and HCA were also evaluated based on the ABTS assay. The freeze-dried *A. bilimbi* had the highest percentage in ABTS-reducing activity (12.46%) compared to freeze-dried *Canarium* sp., freeze-dried *M. domestica* and HCA (Figure 1B). The IC$_{50}$ value was calculated, HCA had the highest ABTS-reducing activity (166.23 µg/mL), and followed by freeze-dried *A. bilimbi* (634.37 µg/mL) which had the highest value among the three freeze-dried fruits, whereas the freeze-dried *M. domestica* had the lowest ABTS-reducing activity (3183.41 µg/mL) (Table 2).

**α-amylase Inhibitory Activity**

To assess the anti-obesity of each freeze-dried fruits and HCA, the α-amylase inhibitory activity was evaluated. Figure 2A showed that freeze-dried *M. domestica* had the...
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Indones Biomed J. 2019; 11(3): 320-6
DOI: 10.18585/inabj.v11i3.728

Figure 1. Concentration of freeze-dried Canarium sp., Averrhoa bilimbi L., Malus domestica, and HCA based on DPPH scavenging activity assay (A) and ABTS-reducing activity assay (B).

Table 2. The IC$_{50}$ value of freeze-dried Canarium sp., Averrhoa bilimbi L., Malus domestica, and HCA based on DPPH scavenging activity assay and ABTS-reducing activity assay.

| Freeze-dried Fruit Sample | IC$_{50}$ (µg/mL) |
|---------------------------|--------------------|
|                           | DPPH              | ABTS               |
| Canarium sp.              | 370.84            | 1210.93            |
| Averrhoa bilimbi L.       | 279.99            | 631.78             |
| Malus domestica           | 805.11            | 3183.41            |
| Hydroxycitric acid        | 349               | 166.23             |

Discussion

Phytochemical has various biological activities underlying the utilization of plant extract as traditional medicine.(22) Some phytochemical contents, especially the phenolic and polyphenolic compound, are well known for its antioxidant capacity, since its structures are able to turn into its less-reactive form.(23,24) The consumption of high phenolic content beverages, such as wine and tea, is also found to increase the serum antioxidant capacity as measured with antioxidant assay.(25,26) The phenolic and polyphenolic content in fruits are able to be metabolized and secreted by the body over a 24 h period after ingestion, thus further increase the safety of its consumption.(27,28)

Freeze-drying is one solution to keep nutritional value, antioxidant activity and phytochemical content of fruits from changing.(12,29) In present study, the phenolic and polyphenolic content of freeze-dried fruits were measured, and it was found that freeze-dried A. bilimbi had the highest phenolic content compare to the other freeze-dried fruits. However, the result of this study showed that the freeze-dried A. bilimbi and freeze-dried Canarium sp. had lower phenolic content compared to previous studies.(30,31) Meanwhile, the freeze-dried Malus domestica, which contained the least phenolic content, showed similar amount of phenolic content (±3 µg/mg) compared to another study.(32)

Total flavonoid assay suggested that freeze-dried M. domestica had the highest flavonoid content, while freeze-dried Canarium sp. had the lowest. Flavonoid content of freeze-dried M. domestica found in this study was slightly

highest α-amylase inhibitory activity (18.18%), compared to freeze-dried Canarium sp., freeze-dried A. bilimbi and HCA. While based on the IC$_{50}$ values, HCA had the highest α-amylase inhibitory activity (196.16 µg/mL), followed by freeze-dried M. domestica (258.85 µg/mL). While freeze-dried Canarium sp. had the lowest α-amylase inhibitory activity (16496.93 µg/mL) (Table 3).

Lipase Inhibitory Activity

The lipase inhibitory activity of freeze-dried fruits and HCA were evaluated to assess the anti-obesity activity. Freeze-dried Canarium sp. had the highest lipase inhibitory activity (77.89%) at the highest concentration tested, while A. bilimbi had the lowest among the three freeze-dried fruits (Figure 2B). Based on the calculated IC$_{50}$ values, HCA had the best value for inhibiting the enzyme lipase (31.22 µg/mL), followed by freeze-dried Canarium sp. (118.66 µg/mL).
lower compared to previous research.(33) The difference on both phenolic and flavonoid content exhibited in present study may be caused by numerous factors, including different cultivars, solvents polarity, and the extraction method used. (31) In addition, the fruit phytochemical content is also affected by different environmental conditions, ranging from the nutritional intake to the climate changes.(34)

Obesity pathogenesis was a complex process triggered by excessive fat consumption and hypoxic condition of the adipose tissue. When combined, both conditions trigger the adipocyte to produce ROS and further cause a condition known as oxidative stress. Oxidative stress stimulates the fat accumulation in adipose tissue and the differentiation of the adipocyte.(35,36) Antioxidant attenuated obesity-related health effect on mice.(37) Based on both DPPH and ABTS assay, the results of this study showed that freeze-dried *A. bilimbi* had the highest antioxidant activity, which is likely due to the high phenolic content found in freeze-dried *A. bilimbi*. Thus, freeze-dried *A. bilimbi* has a good potential to counteract the free radicals. Antioxidant plant extract was also able to protect obese animal model from obesity related adverse health effect.(38) As evidence suggest, *A. bilimbi* may either prevent obesity and/or its adverse health effects through antioxidant activity by relieving the oxidative stress.

In addition to oxidative stress, inhibition of carbohydrate absorption and dietary lipid are also known to be beneficial in treating obesity. After consumption of food rich in carbohydrate, glucose concentration in plasma increased rapidly. The increased blood-sugar induces the body to store monosaccharide either as glycogen in the liver or fat in adipose tissue. Recent clinical trials conclude that the inhibition of α-amylase promoted weight loss.(39) Previous research reported that food rich in polyphenol had potential inhibitory properties against α-amylase and α-glucosidase.(40) As carbohydrate digestion and obesity were linked, inhibition of enzyme may provide benefits for treating this condition. The present study found that freeze-dried *M. domestica* had potential α-amylase inhibition. This activity may be related to the high polyphenolic content found in freeze-dried *M. domestica*, in this study showed by the high flavonoid content in freeze-dried *M. domestica*. This result also supported by previous research stating that *M. domestica* consumption may also limit the absorption of carbohydrate, preventing obesity.

Up until now, the inhibition of pancreatic lipase is the most widely studied mechanism for the evaluation of anti-obesity agents since inhibition of this enzyme hampers the absorption of fatty acid by the gut.(41,42) Polyphenols extracted from black tea were able to inhibit the fat absorption in the gut, which occurs due to the inhibition of the enzyme lipase. (43) The present study found that freeze-dried *M. domestica* has potential lipase inhibitory activity. This activity may be related to the high polyphenolic content found in freeze-dried *M. domestica*, in this study showed by the high flavonoid content in freeze-dried *M. domestica*. This result also supported by previous research stating that *M. domestica* consumption may also limit the absorption of carbohydrate, preventing obesity.

![Figure 2](image-url)  
**Figure 2.** Concentration of freeze-dried *Canarium* sp., *Averrhoa bilimbi* L., *Malus domestica*, and HCA based on α-amylase inhibitory activity assay (A) and lipase inhibitory activity assay (B).

| Freeze-dried Fruit Sample | IC₅₀ (µg/mL) | α-Amylase | Lipase |
|---------------------------|-------------|-----------|-------|
| *Canarium* sp.            | 16496.93    | 118.66    |
| *Averrhoa bilimbi* L.     | 4984.57     | 165.33    |
| *Malus domestica*         | 258.85      | 161.01    |
| Hydroxycitric acid        | 196.16      | 31.22     |
of lipase enzyme. (43) The result of present study showed that the freeze-dried Canarium sp. was able to inhibit lipase activity, as having the highest lipase inhibitory activity. Freeze-dried Canarium sp. may help to prevent obesity by reducing gut fat absorption.

This study showed that freeze-dried fruit demonstrates in vitro benefit toward obesity while extending its quality and storage capability. However, further in vivo study is required to conclude the amount of freeze-dried fruit required, since the sole purpose of this study is to investigate the possible antioxidant and anti-obesity potential of freeze-dried fruit, not to determine the dosage.

**Conclusion**

In summary, freeze-dried fruits demonstrate in vitro benefits toward obesity. Freeze-dried A. bilimbi has potent antioxidant activity and may be beneficial against obesity-related adverse health effect by relieving oxidative stress. While freeze-dried M. domestica and freeze-dried Canarium sp. hamper the fat accumulation by reducing the carbohydrate absorption and dietary lipid in the digestive tract.

**Acknowledgements**

We are grateful to Ministry of Research, Technology and Higher Education of the Republic of Indonesia for the Research Grant of Applied Product Research (2017) for their financial support. We also gratefully acknowledge Universitas YARSI and Aretha Medika Utama Biomolecular and Biomedical Research Center for the methodology and facilities support.

**References**

1. World Health Organization. Obesity: preventing and managing the global epidemic. Report of a WHO consultation. World Health Organ Tech Rep Ser. 2000; 894: 1-253.
2. World Health Organization. Global Status Report on Noncommunicable Diseases 2014. Geneva: World Health Organization; 2015.
3. World Health Organization [Internet]. Obesity and Overweight [updated 2018 Feb 16; cited 2018 Dec 10]. Available from: https://www.who.int/news-room/fact-sheets/detail/obesity-and-overweight.
4. Hruby A, Hu FB. The epidemiology of obesity: a big picture. Pharmacoeconomics. 2015; 33: 673–89.
5. Walls HL, Backholer K, Proietto J, McNeil JJ. Obesity and trends in life expectancy. J Obes. 2012; 2012: 107989.
6. Wyatt SB, Winters KP, Dubbert PM. Overweight and obesity: prevalence, consequences, and causes of a growing public health problem. Am J Med Sci. 2006; 331: 166–74.
7. Marsegilia L, Manti S, D’Angelo G, Nicotera A, Parisi E, Di Rosa G, et al. Oxidative stress in obesity: a critical component in human diseases. Int J Mol Sci. 2014; 16: 378–400.
8. Savini I, Catani M, Evangelista D, Gasperi V, Avigliano L. Obesity-associated oxidative stress: strategies finalized to improve redox state. Int J Mol Sci. 2013; 14: 10497–538.
9. Slanc P, Doljak B, Kreft S, Lunder M, Janeš D, Strukelj B. Screening of selected food and medicinal plant extracts for pancreatic lipase inhibition. Phyther Res. 2009; 23: 874–7.
10. Boqué N, Campión J, de la Iglesia R, de la Garza AL, Milagro FI, San Román B, et al. Screening of polyphenolic plant extracts for anti-obesity properties in Wistar rats. J Sci Food Agric. 2013; 93: 1226–32.
11. Sagar VR, Suresh Kumar P. Recent advances in drying and dehydration of fruits and vegetables: a review. J Food Sci Technol. 2010; 47: 15–26.
12. Sanchez V, Baeza R, Chirife J. Comparison of monomeric anthocyanins and colour stability of fresh, concentrated and freeze-dried encapsulated cherry juice stored at 38°C. J Berry Res. 2015; 5: 243–51.
13. Ivanova D, Gerova D, Chervenkov T, Yankova T. Polyphenols and antioxidant capacity of Bulgarian medicinal plants. J Ethnopharmacol. 2005: 96: 145–50.
14. Widowati W, Rani AP, Hamzah RA, Arumwardana S, Affiah E, Kusuma HSW, et al. Antioxidant and antiaging assays of Hibiscus sabdariffa extract and its compounds. Nat Prod Sci. 2017; 23: 192–200.
15. Sudha G, Priya MS, Shree RI, Vadivukkarasi S. In Vitro Free Radical Scavenging Activity of Raw Pepino Fruit (Solanum muricatum Aiton.). Int J Curr Pharm Res. 2011; 3: 137–40.
16. Widowati W, Wargasetia TL, Affiah E, Mozef T, Kusuma HSW, Nufus H, et al. Antioxidant and antidiabetic potential of Carcuma longa and its compounds. Asian J Agri Biol. 2018; 6: 149–161.
17. Etoundi C, Kuaté D, Ngondi J, Oben J. Anti-amilase, anti-lipase and antioxidant effects of aqueous extracts of some Cameroonian spices. J Nat Prod. 2010; 3: 165–71.
18. Widowati W, Fauziah N, Herdiman H, Afni M, Afifah E, Kusuma HSW, et al. Antioxidant and anti aging assays of oryz sativa extracts, vanillin and coumaric acid. J Nat Remedies. 2016; 16: 88.
19. Adnyana IK, Abuzaid AS, Kurniati NF, Kusuma HSW, et al. Antioxidant and antiobesity properties in Wistar rats. J Sci Food Agric. 2013; 93: 1226–32.
20. Bhutkar MA. In vitro studies on alpha amylase inhibitory activity of some indigenous plants. Mod Appl Pharm Pharmacol. 2018; 2018: 1-5.
21. Liu S, Li D, Huang B, Chen Y, Lu X, Wang Y. Inhibition of pancreatic lipase, α-glucosidase, α-amylase, and hypolipidemic effects of the total flavonoids from Nelumbo nucifera leaves. J Ethnopharmacol. 2013; 149: 263–9.
22. Zheng W, Wang SY. Antioxidant activity and phenolic compounds in selected herbs. J Agric Food Chem. 2001; 49: 5165–70.
23. Sulaiman CT, Sadashiva CT, George S, Gopakrishnan VK, Balachandran I. Chromatographic studies and in vitro screening for acetyl cholinesterase inhibition and antioxidant activity of three acacia species from South India. Anal Chem Lett. 2013; 3: 111–8.
24. Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. J Nutr Sci. 2016; 5: e47.
25. Tsang C, Higgins S, Duthie GG, Duthie SJ, Howie M, Mullen W, et al. The influence of moderate red wine consumption on antioxidant
status and indices of oxidative stress associated with CHD in healthy volunteers. Br J Nutr. 2005; 93: 233–40.
26. Benzie IFF, Szeto YT, Strain JJ, Tomlinson B. Consumption of green tea causes rapid increase in plasma antioxidant power in humans. Nutr Cancer. 1999; 34: 83–7.
27. Stalmach A, Steiling H, Williamson G, Crozier A. Bioavailability of chlorogenic acids following acute ingestion of coffee by humans with an ileostomy. Arch Biochem Biophys. 2010; 501: 98–105.
28. Ottaviani JI, Borges G, Momma TY, Spencer JPE, Keen CL, Crozier A, et al. The metabolome of [2-14C](-)epicatechin in humans: implications for the assessment of efficacy, safety and mechanisms of action of polyphenolic bioactives. Sci Rep. 2016; 6: 29034.
29. Orak H, Aktas T, Yagar H, İsbilir SS, Ekinci N, Sahin FH. Effects of hot air and freeze drying methods on antioxidant activity, colour and some nutritional characteristics of strawberry tree (Arbutus unedo L) fruit. Food Sci Technol Int. 2012; 18: 391–402.
30. Hasanuzzaman M, Ali MR, Hossain M, Kuri S, Islam MS. Evaluation of total phenolic content, free radical scavenging activity and phytochemical screening of different extracts of Averrhoa bilimbi (fruits). Int Curr Pharm J. 2013; 2: 92–6.
31. Mannan A, Rahman MM, Habib MR, Hasan MA, Al Amin M, Saha A. Comparative assessment on in vitro antioxidant activities of ethanol extracts of Averrhoa bilimbi, Gymnema sylvestre and Capsicum frutescens (fruits). Int Curr Pharm J. 2013; 2: 92–6.
32. Navarro M, Moreira I, Arnaez E, Quesada S, Azofeifa G, Vargas F, et al. Polyphenolic characterization and antioxidant activity of Malus domestica and Prunus domestica cultivars from Costa Rica. Foods. 2018; 7: 15.
33. Kschonsek J, Wolfram T, Stöckl A, Böhm V. Polyphenolic compounds analysis of old and new apple cultivars and contribution of polyphenolic profile to the in vitro antioxidant capacity. Antioxidants. 2018; 7: 20.
34. Papetti A, Daglia M, Gazzani G. Anti- and pro-oxidant water soluble activity of Cichorium genus vegetables and effect of thermal treatment. J Agric Food Chem. 2002; 50: 4696–704.
35. Higuchi M, Dusting GJ, Peshavariya H, Jiang F, Hsiao STF, Chan EC, et al. Differentiation of human adipose-derived stem cells into fat involves reactive oxygen species and forkhead Box O1 mediated upregulation of antioxidant enzymes. Stem Cells Dev. 2013; 22: 878–88.
36. Lee H, Lee YJ, Choi H, Ko EH, Kim J. Reactive oxygen species facilitate adipocyte differentiation by accelerating mitotic clonal expansion. J Biol Chem. 2009; 284: 10601–9.
37. Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, et al. Increased oxidative stress in obesity and its impact on metabolic syndrome. J Clin Invest. 2004; 114: 1752–61.
38. Hogan S, Canning C, Sun S, Sun X, Zhou K. Effects of grape pomace antioxidant extract on oxidative stress and inflammation in diet induced obese mice. J Agric Food Chem. 2010; 58: 11250–6.
39. Barrett ML, Udani JK. A proprietary alpha-amylase inhibitor from white bean (Phaseolus vulgaris): A review of clinical studies on weight loss and glycemic control. Nutr J. 2011; 10: 24.
40. Kwon YI, Apostolidis E, Shetty K. Inhibitory potential of wine and tea against α-amylase and α-glucosidase for management of hyperglycemia linked to type 2 diabetes. J Food Biochem. 2008; 32: 15–31.
41. Foster-Schubert KE, Cummings DE. Emerging therapeutic strategies for obesity. Endocr Rev. 2006; 27: 779–93.
42. Yun JW. Possible anti-obesity therapeutics from nature – A review. Phytochemistry. 2010; 71: 1625–41.
43. Yuda N, Tanaka M, Suzuki M, Asano Y, Ochi H, Iwatsuki K. Polyphenols extracted from black tea (Camellia sinensis) residue by hot-compressed water and their inhibitory effect on pancreatic lipase in vitro. J Food Sci 2012; 77: H254–61.