Ischaemia Modified Albumin and Malondialdehyde Level in Subjects Suffering from Hypothyroidism

Jayati Roy Choudhury¹, Amrita Karmakar¹*, Barnita Guha¹, Brahmarshi Das² and Jayanta Kumar Rout¹

¹Department of Biochemistry, R. G. Kar Medical College, Kolkata, India.
²Department of Biochemistry, Midnapore Medical College, Paschim Medinipur, India.

Authors’ contributions

This work was carried out in collaboration between all authors. Authors AKK, JRC and BD designed the study, wrote the protocol and author JKR supervised the work. Authors AKK, JRC and BG carried out all laboratories work and performed the statistical analysis. Authors BD and JKR managed the analyses of the study. Author JRC wrote the first draft of the manuscript. Author AKK managed the literature searches and edited the manuscript. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJBcRR/2015/17638

Editors:
(1) Richard A. Manderville, Departments of Chemistry and Toxicology, University of Guelph, Canada.

Reviewers:
(1) Anonymous, USA.
(2) Sandro Percario, Oxidative Stress Research Lab., Institute of Biological Sciences, Federal University of Para, Brazil and Entomology Branch, Division of Parasitic Diseases and Malaria, US Centers for Disease Control and Prevention, USA.

Complete Peer review History: http://www.sciencedomain.org/review-history.php?id=1038&iid=3&aid=9188

ABSTRACT

Thyroid hormone deficiency (hypothyroidism) leads to many serious conditions in body. Oxidative stress (OS), a state of excess free radicals and reactive metabolites formation which ultimately leads to an imbalance between the production of oxidants and their elimination by antioxidative systems in the body. Researchers all around the globe are in search of the link between this oxidative stress by estimating its different markers and hypothyroidism. In this present study oxidative stress is evaluated in hypothyroidism by estimating serum ischaemia modified albumin (IMA) and malondialdehyde (MDA).

Settings and Design: 56 patients attending different Out-Patient Department (OPD) who have fulfilled inclusion criteria were included as cases and 43 apparently healthy persons were selected as control. Serum fT4 and TSH level measured by immunoassay and serum IMA and MDA level measured by well standardised validated methods.

*Corresponding author: E-mail: amritatua@gmail.com;
1. INTRODUCTION

The thyroid gland produces two related hormones, thyroxine (T4) and triiodothyronine (T3). Acting through nuclear receptors, T3 and T4 play a critical role in cell differentiation during development and help to maintain thermogenic and metabolic homeostasis in the adult. Thyroid hormone deficiency (hypothyroidism) leads to many serious conditions in body like growth failure, atherosclerosis, pleural and pericardial effusion, perceptive deafness, infertility etc. [1].

Oxidative stress (OS) represents an imbalance between the production of oxidants and their elimination by antioxidative systems in the body. Many studies have linked OS to thyroid dysfunction by showing its association with abnormally regulated oxidative or antioxidative molecules [2,3]. Although, the high reactivity and short half life of reactive oxygen and poly unsaturated fatty acid (PUFA) or nitrogen species make difficult their direct determination [3], several methods have been tried. We propose an approach for the determination of oxidative stress in human serum by measuring ischaemia modified albumin (IMA) and malondialdehyde (MDA) which is a familiar member of thiobarbituric acid reacting substances (TBARS).

Ischemia Modified Albumin (IMA) has been developed and found to be very useful for the detection of acute myocardial ischemia and the study of oxidative stress in different disorder. Initially the test was named as Albumin Cobalt Binding (ACB) assay since it is based on the reduced binding affinity of the human serum albumin for metal ions (Cobalt) in patients suffering from oxidative stress. The reduction in the binding affinity of albumin has been attributed to the free radical damage to the N-terminal of albumin molecule in patients [4-8]. In 2012 it was reported that IMA levels are increased in patients with thyroid dysfunction, particularly in overt hypothyroidism [6]. Contradicting to this Ersoy et al. [5] stated that Serum IMA levels did not differ among patients with overt or subclinical hypothyroidism.

Malondialdehyde (MDA), a secondary product of lipid peroxidation (LPO) and is used as an index of tissue damage [9,10], as it is the major reactive aldehyde resulting from peroxidation of biological membrane polyunsaturated fatty acids (PUFA). The peroxidation of poly unsaturated fatty acids of the biological membranes leads to conjugated dienes (CD) formation, followed by the cleavage of the fatty acid chains and subsequent release of the reactive aldehyde products, i.e. malondialdehyde, 4-hydroxy 2,3-transnonenal, 4-hydroxy-2,3-transhexanal etc. often referred to as thiobarbituric acid reacting substances (TBARS) [11,12]. Malondialdehyde is only formed by fatty acids with three or more double bonds and is relatively more stable than others [13].

1.1 Aims and Objectives

1. To find out if there is any correlation between fT4 and TSH with IMA and MDA in our specified groups.
2. To find out if there is any difference of the means of IMA and MDA between our case and control group.

2. MATERIALS AND METHODS

Patients attending different Out-Patient Department (OPD) of R. G. Kar Medical College & Hospital, Kolkata, West Bengal who had serum TSH value >6.5 mIU/L were included in this study. 56 patients were well thought-out as cases. They had attained the age of 18 years and could give informed consent after understanding the objectives of the study. A group of 43 age and sex matched healthy individuals with no history of thyroid disorder served as controls. Control subjects were selected from persons accompanying the patients in OPD who were free from any detectable disorders including hypothyroidism or conditions which may interfere with our study parameter. Patients who are suffering from

Results: By undertaking independent sample’s t test it is found that mean and standard deviation (SD) of IMA (t=4.149, p<0.001) and MDA (t=19.171, p<0.001) are significantly increased in case group than the control group. In case group it was found that serum IMA & MDA are significantly correlated with fT4 {r= -0.835 (IMA) & -0.765(MDA)} & TSH {(r= +0.859(IMA) & +0.672(MDA)).

Keywords: Hypothyroidism; ischaemia modified albumin; malondialdehyde, oxidative stress; free radicals.
diabetes mellitus, neoplasm, neurological diseases were excluded from study. All of the selected patients are free from intake of alcohol or nicotine by any means in any form. As participants attended the hospital OPD from a large rural base, they had approximately similar ethnicity, socioeconomic status and dietary habits. Institutional ethics committee permission was taken prior to start of the work. Informed consent was taken from all participants before inclusion in the study in their convenient language as applicable.

Fasting blood was collected by venipuncture for the determination of different biochemical parameters. The blood samples were subjected to centrifugation at 3,000 rpm for 10 min for separation of serum. Serum thus obtained was analysed for biochemical parameters Serum fT4 and TSH level were measured by ELISA with kit provided by Accubind-Monobind with help of TECAN ELISA (Austria based company) reader and washer. Within assay precision of fT4, TSH was determined by analysis control materials.

IMA estimation [14-17] is based on the premise that hypothyroidism due to oxidative stress causes changes in human serum albumin (HSA) that are demonstrated by reduced exogenous cobalt binding. The concentration of ischemia modified serum albumin can be determined by addition of a known amount of cobalt (II) to a serum specimen and measurement of the unbound cobalt (II) by colorimetric assay using dithiothreitol (DTT). An inverse relationship thus exists between the level of albumin bound cobalt and the intensity of the colour formation. Preparations for the Co (II) albumin binding protocol involved the addition of 200 ml of patient serum to 50 ml of a solution of 1g ml cobalt chloride, followed by vigorous mixing and 10-min incubation. Dithiothreitol (50 ml of a 1.5 g/l solution) was then added and mixed. After 2 min. incubation, 1.0 ml of a 9.0 g/l solution of NaCl was added. The absorbance of the assay mixture was read at 470 nm using a Spectrophotometer. The blank was prepared similarly with the exclusion of DTT. IMA assay was standardized in the Department of Biochemistry R. G. Kar Medical College and a standard curve was prepared in the range 6.0-60.0 µg CoCl2/ml. One IMA unit was defined as “µg of free Co (II)” in the reaction mixture per ml of serum sample”. The assay was found to maintain linearity within this range. The calculated factor from this standard curve is 411.8.

So,

\[
IMA=411.8 \times (\text{Absorbance of unknown}) - \times (\text{Absorbance of blank}) \times 5 \mu g/ml.
\]

For the convenience in the present study we expressed our values as absorbance units (ABSU).

MDA estimation [18-20] is based on that polyunsaturated fatty acids react with molecular oxygen to undergo destructive free radical mediated auto-oxidation. During this process numerous peroxides and aldehyde compounds are formed. These non-volatile compounds decompose to form MDA and other substances during acid heating in laboratory (Prayer et al.). These substances react with thiobarbituric acid in an acidic medium to form a pink colour compound. Then the product is readily extractable into organic solvents such as butanol and absorbance was taken at 532 nm using dual beam spectrophotometer.

2.1 Estimation of MDA in Samples

To 0.5 ml of freshly prepared serum 2.5 ml of trichloroacetic acid (20 mg/dl) was added and the tube was allowed to stand for 10 minutes at room temperature. Then 2.5 ml of sulphuric acid (0.05 M) was added and stirred thoroughly. Then 3.5 ml of TBA reagent (200 mg of TBA was mixed with 100 ml of 2 M sodium sulphate solution) was added to this. The coupling of lipid peroxide with TBA was carried out by heating in boiling water bath for 30 minutes. It was then cooled in water. 4.0 ml of n-butanol was added and chromogen was extracted to organic phase by vigorous shaking and vortexing followed by centrifugation at 3000 rpm for 10 minutes.

The supernatant organic phase was pipetted to a clean test tube. Its absorbance was determined at 532 nm wave length by spectrophotometer. N-Butanol was used as blank to assure zero reading. The optical density was noted and level of MDA was calculated from standard curve in the range of 5-15 nmol/ml. The calculated factor from the standard curve is = 60.

Hence,

\[
\text{Concentration of MDA of a unknown solution} = \frac{\text{Absorbance of unknown}}{60} \times 2 \text{ nmol/ml}.
\]
3. RESULTS

Serum TSH (18.3±8.1 mIU/ml) was significantly raised among case group in comparison to control (5±0.87 mIU/ml). IMA (0.51±0.14 μg/ml) and MDA (9.27±1.27 nmol/ml) also significantly increased among case group in respect to control. Serum fT4 was significantly decreased among case group (vide Table 1). In control group Pearson’s bivariate correlation shows that serum IMA & MDA though correlated with fT4 (-ve) & TSH (+ve) but the level is not statistically significant in controls. In the case group it was found that serum IMA & MDA are significantly correlated with fT4 (r=-0.835 & -0.765) & TSH (r=+0.859 & +0.672) (vide Table 2).

4. DISCUSSION

Oxidative stress represents an imbalance between the production of oxidants and their elimination by antioxidative systems in the body. In this study, serum IMA and malondialdehyde had been determined as oxidative stress marker taking into account the relationship between free radicals and thyroid diseases. In 2012 Erdamar H et al. [2] revealed an increased generation of reactive oxygen species and impairment of the antioxidant system in patients with hypothyroidism. Malondialdehyde, nitrite, vitamin E and myeloperoxidase activity increased in patients with hypothyroidism. Some other studies also showed association of thyroid dysfunction with abnormally regulated oxidative or antioxidative molecules [3]. Contradicting to this Ersoy et al. [5] stated that Serum IMA levels did not differ among patients with overt or subclinical hypothyroidism.

In this study 56 cases and 43 age and sex matched controls had been recruited. After analyzing different parameters and applying appropriate statistical analysis it was found that the mean and standard deviation (SD) of different studied parameters are significantly different in these two groups. After doing independent sample ‘t’ test it was found that all the studied parameters has a statistically significant difference of mean of cases from that of controls group.

In control group Pearson’s bivariate correlation shows that serum IMA & MDA though correlated with fT4 (-ve) & TSH (+ve) but the level is not statistically significant in controls. In cases we found that serum IMA & MDA are significantly correlated with fT4 (r=-0.835 & -0.765) & TSH (r=+0.859 & +0.672).

Hence, from this study it can be concluded that in hypothyroidism there is oxidative stress which is reflected by the existence of significant level of free radical mediated generation of serum MDA and IMA.

Results revealed an increased generation of reactive oxygen species and impairment of the antioxidant system in patients with hypothyroidism.

Table 1. Mean and standard deviation and their comparison by independent t test

| Biochemical parameter | Case (mean±SD) | Control (mean±SD) | 't' value | 'p' value |
|-----------------------|----------------|-------------------|-----------|-----------|
| fT4 (ng/dl)           | 0.93±0.58      | 1.6312±0.15       | -7.750    | <0.001    |
| TSH (m IU/ml)         | 18.31±8.1      | 5.0±0.87          | 10.71     | <0.001    |
| IMA (μg/ml)           | 0.51±0.14      | 0.42±0.047        | 4.14      | <0.001    |
| MDA (nmol/ml)         | 9.27±1.27      | 4.6±1.09          | 19.17     | <0.001    |

Table 2. Pearson’s bivariate correlation of fT4 & TSH with different study parameters of controls and cases are as follows

| Thyroid function tests | Control IMA | Case IMA | Control MDA | Case MDA |
|-----------------------|-------------|----------|-------------|----------|
| fT4                   | -2.69       | -0.835   | -0.225      | -0.765   |
| Sig. (2-tailed)       | >0.05       | <0.001   | >0.05       | <0.001   |
| TSH                   | .296        | .859     | .241        | .672     |
| Sig. (2-tailed)       | >0.05       | <0.001   | >0.05       | <0.001   |

IMA and MDA levels were significantly correlated with fT4 and TSH only among case group.
5. CONCLUSION

Malondialdehyde and IMA concentrations increased in patients with hypothyroidism and also correlated with the severity of the disease as reflected by the levels of FT4 and TSH. These findings indicate that thyroid hormones have a strong impact on oxidative stress and the antioxidant system.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Jameson JL, Weetman AP. Disorders of the thyroid gland. in: Kasper DL, Fauci AS, Longo DL, Braunwald E, Hauser SL, Jameson JL, editors. Harrison’s principles of internal medicine. 16th ed. New York: Mcgraw-hill Medical Publishing Division. 2005;2104-27.
2. Xing M. Oxidative stress: A new risk factor for thyroid cancer. Endocr Relat Cancer. 2012;19(1):C7-11.
3. Metere A, Chiesa C, Di Cosimo C, Fierro G, Giacomelli L, Pietraforte D. A novel approach to study oxidative stress in thyroid diseases: A preliminary study. Eur Rev Med Pharmacol Sci. 2012;16(5):646-52.
4. Nanda N, Bobby Z, Hamide A, Koner BC, Sridhar MG. Association between oxidative stress and coronary lipid risk factors in hypothyroid women is independent of body mass index. Metabolism. 2007;56(10):1350-5.
5. Ersoy K, Anaforoglu I, Algun E. Serum ischemic modified albumin levels might not be a marker of oxidative stress in patients with hypothyroidism. Endocrine. 2013;43(2):430-3.
6. Ma SG, Yang LX, Bai F, Xu W, Hong B. Ischemia-modified albumin in patients with hyperthyroidism and hypothyroidism. Eur J. Intern Med. 2012;23(6):e136-40.
7. Chawla R, Goyal N, Calton R, Goyal S. Ischemia Modified Albumin: A novel marker for acute coronary syndrome. Indian Journal of Clinical Biochemistry. 2006;1(21):77-82.
8. Govender R, De Greef J, Delport R, Becker PJ, Vermaak WJ. Biological variation of ischaemia-modified albumin in healthy subjects. Cardiovasc J. Afr. 2008;19(3):141-4.
9. Vaca CE, Wilhelm J, Harms-Ringdahl M. Interaction of lipid peroxidation products with DNA. A review. Mutat Res. 1988;195(2):137-49.
10. Vaca CE, Wilhelm J, Harms-Ringdahl M. Studies on lipid peroxidation in rat liver nuclei and isolated nuclear membranes. Biochim Biophys Acta. 1988;956(3):375-87.
11. Kehrer JP. Free radicals as mediators of tissue injury and disease. Crit Rev Toxicol. 1993;(23):21-48.
12. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxidation in animal tissues by thiobarbituric acid reaction. Anal Biochem. 1979;(95):351.
13. Hayashi O, Shumizu T, Yag K. Metabolic and functional significance of prostaglandins in lipid peroxide research, in Lipid peroxides in biology and medicine. Academic Press, New York. 1982;(41).
14. Chawla R, Goyal N, Calton R, Goyal S. Ischemia modified albumin: A novel marker for acute coronary syndrome. Indian J. Clin Biochem. 2006;21(1):77-82.
15. Beetham R, Monk C, Keating L, Benger JR, Kendall J. Effects of storage at -20 degrees C on ischaemia-modified albumin results. Ann Clin Biochem. 2006;43(Pt6):500-2.
16. Maguire OC, O’Sullivan J, Ryan J, Cunningham SK. Evaluation of the albumin cobalt binding (ACB) assay for measurement of ischaemia-modified albumin (IMA) on the Beckman Coulter LX-20. Ann Clin Biochem. 2006;43(Pt6):494-9.
17. Keating L, Benger JR, Beetham R, Bateman S, Veysey S, Kendall J, et al. The PRIMA study: Presentation ischaemia-modified albumin in the emergency department. Emerg Med J. 2006;23(10):764-8.
18. Janero DR, Burghardt B. Thiobarbituric acid-reactive malondialdehyde formation during superoxide-dependent, iron-catalyzed lipid peroxidation: Influence of peroxidation conditions. Lipids. 1989;24(2):125-31.
19. Janero DR, Burghardt B. Analysis of cardiac membrane phospholipid peroxidation kinetics as malondialdehyde: Nonspecificity of thiobarbituric acid-reactivity. Lipids. 1988;23(5):452-8.

20. Gutteridge JM, Quinlan GJ. Malondialdehyde formation from lipid peroxides in the thiobarbituric acid test: The role of lipid radicals, iron salts and metal chelators. J. Appl Biochem. 1983; 5(4-5):293-9.

© 2015 Choudhury et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited.