3,4-Unsubstituted 2-tert-Butyl-pyrrolidine-1-oxyls with Hydrophilic Functional Groups in the Side Chains

Andrey I. Taratayko 1,*, Yurii I. Glazachev 2, Ilia V. Eltsov 3, Elena I. Chernyak 1 and Igor A. Kirilyuk 1

1 N.N. Vorozhtsov Institute of Organic Chemistry SB RAS, Academician Lavrentiev Ave. 9, 630090 Novosibirsk, Russia; chernyak@nioch.nsc.ru (E.I.C.); kirilyuk@nioch.nsc.ru (I.A.K.)
2 Voevodsky Institute of Chemical Kinetics and Combustion SB RAS, Institutskaya 3, 630090 Novosibirsk, Russia; glaza@kinetics.nsc.ru
3 Department of Natural Sciences, Novosibirsk State University, Pirogova Str. 1, 630090 Novosibirsk, Russia; eiv@fen.nsu.ru
* Correspondence: taratayk@nioch.nsc.ru

Abstract: Pyrrolidine nitroxides with four bulky alkyl substituents adjacent to N–O group are known for their high resistance to bioreduction. The 3,4-unsubstituted 2-tert-butyl-2-ethylpyrrolidine-1-oxyls were prepared from the corresponding 2-tert-butyl-1-pyrroline-1-oxides via either the addition of ethinylmagnesium bromide with subsequent hydrogenation or via treatment with ethyllithium. The new nitroxides showed excellent stability to reduction with ascorbate with no evidence for additional large hyperfine couplings in the EPR spectra.

Keywords: nitroxides; nitrones; organometallic compounds; pyrrolidine; reduction kinetics

1. Introduction

Nitroxides are a broad family of stable free radicals with one unpaired electron localized on the unbonding orbital of the N–O group. These radicals have found broad application in various fields of science and technology [1–3]. The broad majority of commonly used nitroxides have a cyclic structure with four methyl substituents adjacent to the N–O group. Recently, nitroxides with methyl groups replaced with bulkier alkyl substituents have attracted much attention. These so-called “sterically shielded” nitroxides demonstrated much higher stability against chemical reduction to diamagnetic compounds with components of biological systems then their tetramethyl analogs did [4–6]. The advantage of these reduction-resistant radicals over conventional tetramethyl-substituted nitroxides is especially obvious when they are used for EPR measurements inside living cells [7,8], or in vivo for functional imaging using MRI or EPR techniques [9,10]. Pyrrolidine nitroxides with four ethyl groups (or three ethyl and one tert-butyl group) in the nearest environment of the nitroxide moiety demonstrated the highest resistance to reduction [4,5,11–14]. However, nitroxides with multiple bulky alkyl substituents are lipophilic or even insoluble in water, therefore their use in biophysics and structural biology is complicated [14]. Moreover, the EPR spectra of these nitroxides may reveal large additional hyperfine couplings (ca. 0.2 mT) with methylene hydrogens of the ethyl groups [11,13,14]. These couplings are typical for 3-substituted or 3,4-disubstituted five-membered rings sterically shielded nitroxides and were shown to result from interaction of ethyl group with a cis-substituent at the neighboring asymmetric center, which prevents averaging of spin density on methylene hydrogens of the ethyl group [15].

The 3,4-unsubstituted sterically shielded pyrrolidine nitroxides with carboxylic groups in the side chains were described by Lampp et al. [12]. The EPR spectra of these nitroxides showed relatively narrow lines ($\Delta B_{pp}$ (Gaussian) = 0.18–0.21 mT) with no evidence for large additional splittings on $\gamma$-hydrogens. Here, we combined the general approach used by Lampp with the methods suggested in our previous papers [11,13,14] to prepare new water-soluble sterically shielded nitroxides.
2. Results and Discussion

Reductive cyclization of aliphatic $\gamma$-nitroketones is one of the most widely used methods of synthesis of pyrrolidine N-oxides [12,16–18], and reactions of the latter with organometallic reagents are commonly used for the preparation of pyrrolidin nitroxides [12,16]. Bulky substituents and functional groups can be introduced on the first step of this sequence of procedures. Sodium methoxide-catalyzed Michael addition of nitro-compound 1a,b [19,20] to 4,4-dimethylpent-1-en-3-one 2 [21] afforded $\gamma$-nitroketone 3a,b (Scheme 1), which were treated with Zn/NH$_4$Cl without purification. Resulting nitrones 4a,b were isolated as a colorless oils with 57–60% yields for two steps.

![Scheme 1. Synthesis of pyrrolidine N-oxides.](image1)

The addition of ethynylmagnesium bromide to pyrrolidine N-oxides can proceed without affecting the carboxylic group, and the ethynyl moiety can be converted into an ethyl group via hydrogenation [13]. Here, we exploited the same approach. Alkaline hydrolysis of ester groups in 4a,b gave corresponding mono- and dicarboxylic acids 5a,b with nearly quantitative yields (Scheme 2).

![Scheme 2. Alkaline hydrolysis of 4a,b.](image2)

The addition of ethynylmagnesium bromide to 5a proceeded slowly, presumably due to sterical hindrance. Complete conversion was achieved only on the 7th day of treatment with 15-fold excess (0.5–1 M in THF) of the reagent at room temperature. Subsequent treatment analogous to the literature procedure [13] afforded a mixture of two diastereomeric nitroxides 7a and 7a’, which could not be separated by column chromatography or crystallization (Scheme 3). The ratio of isomers 7a and 7a’ was 6.5:1 according to $^1$H NMR spectra recorded after reduction of the sample with Zn in the presence of trifluoroacetic acid in CD$_3$OD (see [5] for detailed procedure of reduction).

Similar to that described for 3-carboxypyrroline nitroxides [13], the hydrogenation of a mixture of 7a,a’ on Pd/C in methanolic solution gives a crystalline precipitate of poorly soluble zwitterionic 1-hydroxy pyrroline carboxylates. As a result, the addition of hydrogen to multiple carbon-carbon bonds occurs slowly, leaving no chance to obtain the desired 2,5-diethyl-substituted nitroxide with a satisfactory yield [13]. To prevent the precipitation of inner salts, mixture 7a,a’ was first treated with diazomethane solution (Scheme 4), and resulting mixture of esters 8a was subjected to hydrogenation at atmospheric pressure on Pd/C (4%) in methanolic solution. Subsequent treatment with alkali in aqueous methanol in aerobic conditions afforded 10a in 92% yield. Pure major isomer 10a was isolated via crystallization from hexane. To determine the relative configuration of
the asymmetric centers in 10a, the nitroxide was reduced to corresponding amine with Zn in CF₃COOH–CD₃OD at 65 °C and NMR spectra were recorded. Analysis of the HSQC, HMBC, COSY, and NOESY spectra unambiguously showed the trans-configuration of the ethyl groups in the pyrrolidine ring (see Supplementary Materials SI and Appendix A).

Thus, the addition of the ethynylmagnesium bromide to 5a predominantly occurs cis to the 2-carboxyethyl group, presumably due to coordination of the organometallic reagent with carboxylate anion.

Scheme 3. Addition of ethynylmagnesium bromide to nitron 5a.

In contrast to 5a, no trace of conversion was observed upon treatment of nitrone 5b with a twenty-fold excess of the ethynylmagnesium bromide, even with an increase in reaction time of up to 3 months. The explanation is most likely the formation of insoluble magnesium salt (Scheme 5).

Scheme 5. Reaction of nitrone 5b with ethynylmagnesium bromide.

Organolithium compounds can be successfully used to prepare highly strained nitroxides from cyclic α-tert-butynitronees [14]. To avoid side reactions and increased reagent consumption, esters were reduced to hydroxymethyl groups and the latter were protected. Treatment of nitrones 4a,b with a threefold excess of lithium aluminum hydride resulted in reduction of both the ester and nitron groups (Scheme 6). To regenerate the nitrone groups, the resulting N-hydroxyamines were oxidized with MnO₂. The hydroxy groups were protected via treatment with dimethoxypropane in the presence of pyridinium tosylate. Pure nitrones 12a,b were isolated using column chromatography as colorless oils.
Organolithium compounds can be successfully used to prepare highly strained nitroxides. Scheme 6 shows the synthesis of the nitrones 12a,b.

Nitrones 12a,b readily reacted with ethyllithium, the resulting hydroxylamines 13a,b were oxidized to nitroxides by air oxygen in presence of methylene blue without isolation, and the protecting groups were removed to give 14a,b (Scheme 7).

Scheme 6. Synthesis of the nitrones 12a,b.

HPLC analysis and 1H NMR spectra after reduction of the sample with Zn in the presence of trifluoroacetic acid in CD$_3$OD showed that 14a is a mixture of diastereomers in equal ratio. Thus, unlike the literature examples [14], addition of ethyllithium to 12a does not exhibit any stereoselectivity. The nitroxide 14b is a racemate.

The EPR spectra of the nitroxides 10a and 14b and diastereomeric mixtures 7a,a′, 8a, and 14a are represented in Figure 1 by broadened triplets with no additional resolved hyperfine structure. The broadening obviously resulted from multiple smaller couplings with neighboring γ and δ-hydrogens, indicating some averaging due to relatively free rotation of the alkyl groups. Parameters of EPR spectra, partition coefficients octanol/water and rate constants of reduction with ascorbate of mixtures 7a,a′ and 8a, and individual nitroxides 10a, 14a, and 14b are listed in the Table 1. In mixtures of 7a,a′ and 8a, the contribution of the minor component did not exceed 15% and produced no effect on the apparent linewidths. Moreover, the spectrum of 14a, which is a mixture of two diastereomers in equal ratio, does not show broader lines than spectra of 10a and 14b. Linewidths in the spectra of 10a and 14a,b were ca. 40% higher than those reported for nitroxides prepared by Lampp et al. [12]; however, they were ca. 40% lower than overall widths of the component of the nitroxide triplet (H$_{pp}$ + αH) in similar 2-tert-butyl-3,4-disubstituted nitroxide [14].

The spectra of nitroxides with ethynyl moiety showed remarkably smaller linewidths than those of similar nitroxides with ethyl groups. All the new nitroxides are lipophilic, but their partition coefficients were significantly lower than those of earlier described β-tert-butyl-substituted nitroxides of pyrrolidine series [14]. The nitroxide with two 3-hydroxypropyl groups 14b showed the lowest partition coefficient in octanol-water mixtures. The 7a,a′ and 10a are weak acids and their distribution octanol–water mixtures may be accompanied by partial ionization of carboxylic groups. Adjustment of pH of the water phase to 9 leads to redistribution of the nitroxides into water solution with $K_p < 0.1$. Similar to the earlier described pattern [14], tert-butyl-substituted nitroxides 10a, 14a and 14b demonstrate very high resistance to reduction. The distant functional groups in the side chains (carboxy or hydroxyl) produce minor effect on the reduction rate. As expected, nitroxides 7a,a′ and 8a were much stronger oxidants due to the electron-withdrawing effect of ethynyl group.
Table 1. Parameters of EPR spectra in water, partition coefficients octanol/water, and reduction rate constants with ascorbate.

| Nitro oxide | g-Factor | Hp-p, mT (Center) | $a_N$, mT | Partition Coefficient | $k_{red}$, M$^{-1}$ s$^{-1}$ |
|-------------|----------|------------------|-----------|-----------------------|--------------------------|
| 7$a,a'$     | 2.00571 ± 2 | 0.17 ± 0.005     | 1.61 ± 0.005 | 150                  | 0.2 ± 0.05               |
| 8a          | 2.00571 ± 2 | 0.17 ± 0.005     | 1.61 ± 0.005 | 600                  | 0.6 ± 0.1                |
| 10a         | 2.00563 ± 2 | 0.28 ± 0.005     | 1.61 ± 0.005 | 240                  | (4 ± 1) × 10$^{-5}$     |
| 14a         | 2.00560 ± 2 | 0.26 ± 0.005     | 1.61 ± 0.005 | 400                  | (3 ± 1) × 10$^{-5}$     |
| 14b         | 2.00562 ± 2 | 0.26 ± 0.005     | 1.61 ± 0.005 | 70                   | (5 ± 1) × 10$^{-5}$     |

3. Materials and Methods

3.1. General Information

The IR spectra were recorded on a Bruker Vector 22 FT-IR spectrometer (Bruker, Billerica, MA, USA) in KBr pellets (1:150 ratio) or in neat samples (see Figures S1–S13). The 1H NMR spectra were recorded on a Bruker AV 300 (300.132 MHz), AV 400 (400.134 MHz) and DRX 500 (500.130 MHz) spectrometers (Bruker, Billerica, MA, USA). 13C NMR spectra were recorded on a Bruker AV 300 (75.467 MHz), AV 400 (100.614 MHz) and DRX 500 (125.758 MHz) spectrometers (see Figures S14–S35). All NMR spectra were acquired for 5–10% solutions in CDCl$_3$, (CD$_3$)$_2$SO or CDCl$_3$–CD$_3$OD mixtures at 300 K using the signal of the solvent as a standard. HRMS analyses were performed with high-resolution mass spectrometer DFS (Thermo Electron, Waltham, MA, USA). NMR spectra of nitroxides for analysis and structure assignment were recorded after reduction with Zn in CD$_3$OD–CF$_3$COOH at 65 °C, as described in [13]. HPLC analysis was performed with an Agilent 1100 liquid chromatography system (Agilent Technologies, Santa Clara, CA, USA) equipped with a quaternary pump, online degasser, autosampler, and diode array detector. Chromatographic separations were carried out on a ZORBAX SB-C18 column (150 mm × 4.6 mm, 5.0 μm) by use of binary solvent system MeCN:H$_2$O (4:1 v/v). Flow rate was 0.7 mL/min. Detection was performed at 238 nm.

Figure 1. EPR spectra of mixtures 7$a,a'$ and 8a, and individual nitroxides 10a, 14a, and 14b (0.2 mM in deoxygenated water). For spectrometer settings, see Section 3.
Reactions were monitored by thin-layer chromatography (TLC) on Merck TLC Silica gel 60 F254 plates. Kieselgel60 (Macherey-Nagel GmbH & Co. KG, Düren, Germany) was utilized as sorbent for the column chromatography.

EPR experiments were performed on an X-band (9.8 GHz) EPR spectrometer ER-200D (Bruker, Billerica, MA, USA). All measurements were performed in a 50 µL glass capillary.

3.1.1. Conditions for Spectral Analysis

Radicals concentrations were 0.2 mM in deoxygenated distilled deionized water. Spectra were fitted with Winsim program to calculate hyperfine constants. g-factor was calculated versus Li:F (g = 2.002293). EPR settings: modulation amplitude, 0.8 G; MW power, 5 mW; time constant, 50 ms; total acquisition time, 3 min.

3.1.2. Conditions for Kinetics Experiments

Radical concentrations were 0.1–0.2 mM in phosphate buffer (5 mM, pH ~7.5). Glutathione, 5 mM; ascorbic acid: 100, 200, 300 mM (for nitroxides 10a, 14a, 14b); 33, 66, 100 mM (for 7a,b and 8a). Kinetics of the decrease in EPR signal of radicals with time was recorded. Total kinetics time was 3 h (nitroxides 10a, 14a, 14b) or 10 min (7a,a’ and 8a). Pseudo first-order reaction rate constants were calculated for each ascorbic acid concentration from initial slope (nitroxides 10a, 14a, 14b) or exponential decay rate (7a,a’ and 8a) of kinetics. Second-order rate constants were calculated from linear fitting of the pseudo first order rate constants versus ascorbic acid concentration. EPR settings: modulation amplitude, 2.5 G; microwave power, 5 mW; time constant, 20 ms.

3.1.3. Partition Coefficient Measurements

A solution of a nitroxide (0.1–0.2 mM) in water (1 mL) was prepared. Aliquots of the solution were taken into 50 µL capillary for measurements of amplitude of low field component of EPR spectrum. Thereafter, this aliquot was returned back, required volume octanol (1–5% of water volume) was added, and the mixture was shaken and then shortly centrifuged to separate aqueous and octanol phase. Then, the aliquot of aqueous phase was carefully taken to measure the EPR signal again. This procedure was repeated at least three times for each sample. The amplitude of the intensity of the low field band of the nitroxide EPR spectrum in octanol fraction and the intensity of the initially recorded spectrum in water were used to calculate the partition coefficient. To measure partition coefficient at pH 9 larger volume of octanol was used (100–300% to that of water). EPR settings: modulation amplitude—2.0–2.5 G; microwave power—5 mW; time constant—50 ms.

Compounds 1a,b and 2 were prepared using literature procedures [19–21].

3.2. Synthesis

3.2.1. Synthesis of tert-Butyl Substituted Nitrones (General Method)

A methyl-4-nitroheptanoate (1a) or dimethyl 4-nitroheptanedioate (1b) (100 mmol) was added dropwise to freshly prepared solution of sodium methylate in methanol (0.35 M, 40 mL) within 15 min. The yellow solution formed was stirred for 20 min at room temperature, then a solution of 4,4-dimethylpent-1-en-3-one 2 (100 mmol) in 40 mL of dry methanol was added dropwise within 30 min. The reaction mixture was allowed to cool down to room temperature, then heated to reflux for 3 h. (TLC control on SiO₂, chloroform, stained with phosphomolybdic acid). The pH was adjusted to neutral by addition of glacial acetic acid (ca. 1.5 mL). and the solvent was distilled off in vacuum. The residue was dissolved in ethyl acetate (150 mL) and successively washed with water (70 mL), saturated solution of sodium bicarbonate (100 mL) and again with water (100 mL). The organic layer was dried with anhydrous Na₂SO₄ and the solvent was evaporated in vacuum. The residue was dissolved in 30 mL of THF and mixed with aqueous solution ammonium chloride (1.7 M, 60 mL). The resulting solution was cooled to 7 °C, and zinc dust (24.0 g, 370 mmol) was added by small portion maintaining the temperature in the range of 7–15 °C. The reaction mixture was stirred for 1 h at 5 °C and then 1 h at room temperature. Inorganic
precipitate was filtered off and washed with ethanol (200 mL). The filtrate was concentrated in vacuum and the residue was dissolved in water (50 mL). Water solution was extracted with diethyl ether (50 mL) and organic layer was discarded. Water layer was saturated with NaCl and extracted with CHCl₃ (3 × 100 mL). The combined organic extracts were dried with anhydrous Na₂SO₄. The solvent was evaporated in vacuum, the crude residue was purified by column chromatography (SiO₂, chloroform-methanol 40:1 mixture as an eluent, detected under UV lamp) to give 4a or 4b.

Methyl 3-(5-tert-butyl-2-ethyl-1-oxido-3,4-dihydro-2H-pyrrol-2-yl)propanoate (4a): yield 15.3 g (60%). Colorless oil. HRMS (EI/DFS) m/z [M⁺] calcd. for (C₁₄H₂₃NO₃)⁺: 255.1829; found: 255.1831. IR (neat) νmax: 1737 (C=O), 1573 (C=N). ¹H NMR (500 MHz; CDCl₃, δ): 0.77 (t, J₁ = 7.4 Hz, 3H), 1.22 (s, 9H, t-Bu), 1.41–1.50 (m, 1H), 1.73–1.82 (m, 2H), 1.83–1.92 (m, 2H), 2.07–2.15 (m, 1H), 2.16–2.28 (m, 2H), 2.51 (dd, J₄₁ = 7.6 Hz, J₄₂ = 7.2 Hz, 2H), 3.58 (s, 3H, OCH₃). ¹³CNMR (125 MHz; CDCl₃, δ): 7.29, 23.98, 25.26, 27.31, 28.23, 30.50, 32.67, 33.33, 51.38, 79.25, 150.67, 173.3.

Dimethyl 3,3’-(5-tert-butyl-1-oxido-3,4-dihydro-2H-pyrrole-2,2-diyl)dimpropanoate (4b): yield 17.8 g (57%). Colorless oil. HRMS (EI/DFS) m/z [M⁺] calcd. for (C₁₆H₂₅NO₄)⁺: 313.1884; found: 313.1880. IR (neat) νmax: 1737 (C=O), 1573 (C=N). ¹H NMR (400 MHz; CDCl₃, δ): 1.15 (s, 9H, t-Bu), 1.71–1.82 (m, 4H), 2.02–2.11 (m, 2H), 2.11–2.19 (m, 4H), 2.45–2.51 (m, 2H), 3.52 (s, 6H, OCH₃). ¹³CNMR (100 MHz; CDCl₃, δ): 24.44, 25.15, 27.13, 28.07, 32.53, 33.35, 51.49, 78.04, 151.06, 173.02.

3.2.2. 3-(5-Tert-Butyl-2-ethyl-1-oxido-3,4-dihydro-2H-pyrrol-2-yl)propanoic Acid (5a)

An aqueous solution of sodium hydroxide (3.2 M, 50 mL) was added to the solution of 4a (10.2 g, 40 mmol) in methanol (50 mL), and the resulting homogeneous mixture was left at room temperature for 24 h. Methanol was distilled off in vacuum, the water solution was shaken with diethyl ether (50 mL), and the organic layer was discarded. Chloroform (70 mL) was added to the water layer and 2 M aqueous solution of H₂SO₄ (40 mL) was added to the mixture upon stirring. The organic layer was separated, and the water layer was extracted with CHCl₃ (2 × 50 mL). Combined organic extracts were dried with anhydrous Na₂SO₄, the solvent was evaporated in vacuum, the crude residue was treated with diethyl ether (30 mL), and and white crystalline precipitate of 5a was filtered off, yield 8.5 g (91%), m.p. 160.6–162.0 °C (ethyl acetate). Found: C, 64.91; H, 9.31; N, 5.93; calcd. For C₁₅H₂₅NO₃: C, 64.70; H, 9.61; N, 5.80%; IR (KBr) νmax: 1726 (C=O), 1594 (C=N). ¹H NMR (500 MHz; CDCl₃, δ): 0.79 (t, J₁ = 7.4 Hz, 3H), 1.26 (s, 9H, t-Bu), 1.46–1.55 (m, 1H), 1.76–1.86 (m, 2H), 1.87–1.96 (m, 2H), 2.09–2.21 (m, 2H), 2.26–2.34 (m, 1H), 2.54–2.66 (m, 2H), 11.16 (br. S, 1H). ¹³CNMR (125 MHz; CDCl₃, δ): 7.00, 24.01, 25.55, 28.03, 28.76, 30.55, 32.10, 33.84, 80.39, 157.62, 174.74.

3.2.3. 3,3’-(5-Tert-Butyl-1-oxido-3,4-dihydro-2H-pyrrole-2,2-diyl)di propioanoic Acid (5b)

An aqueous solution of sodium hydroxide (3.2 M, 5 mL) was added to the solution of 4b (1.25 g, 4 mmol) in methanol (5 mL). The resulting homogeneous solution was allowed to stand at room temperature for 24 h, methanol was distilled off in vacuum pressure and 2M solution of H₂SO₄ (4.5 mL) was added to the residue upon stirring. The formed precipitate of 5b was filtered off and successively washed with iced-cold water (2 mL) and diethyl ether (10 mL) and dried in vacuum, yield 1.0 g (95%). White crystalline solid, m.p. 223.3–224.5 °C (ethanol). Found: C, 59.22; H, 8.29; N, 4.78; calcd. For C₁₄H₂₅NO₃: C, 58.93; H, 8.12; N, 4.91%; IR (KBr) νmax: 1720 (C=O), 1594 (C=N). ¹H NMR (300 MHz; (CD₃)₂SO, δ): 1.20 (s, 9H, t-Bu), 1.62–1.76 (m, 2H), 1.80–2.05 (m, 6H), 2.16–2.32 (m, 2H), 2.54–2.67 (m, 2H). ¹³CNMR (75 MHz; (CD₃)₂SO, δ): 23.66, 24.89, 26.15, 26.78, 28.00, 32.71, 33.07, 77.91, 150.65, 174.17.
3.2.4. 2-tert-Butyl-5-(2-carboxyethyl)-5-ethyl-2-ethynylpyrrolidine-1-oxyl (7a,a′)

A powder of 5a (3.6 g, 15 mmol) was added to a 0.5–1 M solution of ethynyl-magnesium bromide in THF (250 mL) upon stirring. The mixture was allowed to stand at room temperature for 168 h (TLC control on SiO₂, ethyl acetate: methanol: acetic acid 200:20:1, detected under UV lamp), then quenched with water (20 mL) and acidified with saturated aqueous sodium bisulfate solution (150 mL) to pH 3–4. The organic layer was separated, and the water phase was extracted with ethyl acetate (2 × 100 mL). The combined organic layers were dried with anhydrous Na₂SO₄. The solvent was evaporated in vacuum and the crude residue was dissolved in methanol (70 mL) and basified with sodium hydroxide solution (1 M, 20 mL). Methylene blue (6 mg, 0.02 mmol) was added to the mixture and the air was bubbled until the solution turned dark blue. The methanol was distilled off in vacuum, and the remaining aqueous solution was washed with diethyl ether (30 mL), acidified with saturated aqueous sodium bisulfate solution (20 mL) to pH < 4, and extracted with ethyl acetate (3 × 50 mL). The organic phase was dried with Na₂SO₄ and the solvent was evaporated in vacuum. The residue was purified by column chromatography (SiO₂, eluent ethyl acetate: methanol: acetic acid 100:10:1) to give of 7a,a′, yield 2.40 g (60%), yellow crystalline solid, m.p. 109.9–112.3 °C (hexane: ethyl acetate 5:1). Found: C, 67.66; H, 9.04; N, 5.22; calcld. for C₁₅H₂₅NO₅: C, 67.64; H, 9.08; N, 5.26%; IR (KBr) ν max: 3309, 2983, 2969, 2956, 1718, 1214, 619. 1H NMR(400 MHz; CD₃OD/CDCl₃, Zn/CF₃COOH system δ): 0.63 (t, J₁ = 7.5 Hz, 3H), 0.86 (s, 9H, t-Bu), 1.34–1.53 (m, 1H), 1.53–1.67 (m, 2H), 1.71–1.85 (m, 3H), 1.89–2.06 (m, 1H), 2.06–2.16 (m, 1H), 2.23–2.29 (m, 2H), 2.75 (s, 1H). Signals of minor isomer: 0.50 (t, J₁ = 7.5 Hz, 3H), 0.67 (s, 9H, t-Bu), 2.64 (s, 1H).

3.2.5. 2-tert-Butyl-5-ethyl-2-ethynyl-5-(3-methoxy-3-oxopropyl)pyrrolidine-1-oxyl (8a)

A solution of 7a (1.0 g, 3.76 mmol) in diethyl ether (10 mL) was slowly added to the 0.4 M solution (40 mL) of diazomethane in diethyl ether at 0 °C. The reaction mixture was stirred for 1 h at this temperature. The reaction was monitored by TLC (SiO₂, ethyl acetate: methanol: acetic acid 200:20:1 mixture; detected under UV lamp). Then, acetic acid (2 mL) was slowly added to remove excess of diazomethane, and the resulting solution was washed with 10% aqueous sodium bicarbonate (2 × 25 mL) and water (1 × 25 mL). The organic phase was dried with Na₂SO₄ and the solvent was evaporated in vacuum. The residue was purified by column chromatography (SiO₂, ethyl acetate as an eluent, detected under UV lamp) to give 8a, yield 1.0 g (95%), yellow crystalline solid, m.p. 65.4–66.2 °C (hexane). Found: C, 68.79; H, 9.22; N, 4.97; calcld. for C₁₅H₂₇NO₅: C, 68.54; H, 9.35; N, 5.00%; IR (KBr) ν max: 3259 (C=O); 1H NMR (500 MHz; CD₃OD/CDCl₃, Zn/CF₃COOH system δ): 0.97 (t, J₁ = 7.5 Hz, 3H), 1.20 (s, 9H, t-Bu), 1.86–2.26 (m, 6H), 2.26–2.47 (m, 2H), 2.60–2.68 (m, 2H), 3.12 (s, 1H), 3.74 (s, 3H), 7.26–7.42 (m, 5H), 7.42–7.50 (m, 5H), 7.50–7.58 (m, 5H).

3.2.6. 2-tert-Butyl-5-(2-carboxyethyl)-2,5-diethylpyrrolidine-1-oxyl (10a)

A solution of 8a (1.0 g, 3.6 mmol) in methanol (10 mL) was placed in the reaction vessel equipped with magnetic stirrer and connection line to gasometer filled with hydrogen. The catalyst (Pd/C, 4%, 30 mg) was added, and the system was purged with hydrogen and closed. The mixture was vigorously stirred until hydrogen absorption ceased (ca. 7 h, 0.22 L of hydrogen absorbed), then the catalyst was filtered off and washed with methanol. Filtrate was mixed with 2M aqueous solution of sodium hydroxide (10 mL) and allowed to stand at room temperature for 10 h. The solution turned yellow. Methanol was distilled off in vacuum, the remaining water solution was shaken with diethyl ether (15 mL) and organic layer was discarded. Chloroform (20 mL) was added to the mixture was acidified with saturated aqueous sodium bisulfate solution (10 mL) to pH < 4 upon stirring. Organic layer was separated and water layer was extracted with chloroform (20 mL). Combined extract was dried with Na₂SO₄ and the solvent was evaporated in vacuum. The residue was purified by column chromatography (SiO₂, ethyl acetate: methanol: acetic acid 100:10:1 mixture as an eluent) to give 10a, yield 0.90 g (92%), yellow crystalline solid, m.p. 101–102 °C dec. (hexane). Found: C, 66.81; H, 10.49; N, 5.14; calcld. for
Molecules 2022, 27, 1922

C_{15}H_{26}N_3O_3; C, 66.63; H, 10.44; N, 5.18%; IR (KBr) ν_{max}: 1704 (C=O). ^1H NMR (400 MHz; CD_3OD/CDCl_3, Zn/C\textsubscript{5}F\textsubscript{5}COOH system): 0.71 (t, J = 7.4 Hz, 3H), 0.84 (t, J = 7.5 Hz, 3H), 0.88 (s, 9H, t-Bu), 1.43–1.60 (m, 3H), 1.63–1.71 (m, 1H), 1.74–2.03 (m, 6H), 2.29–2.45 (m, 2H).

3.2.7. Synthesis of tert-Butyl Substituted Nitrones, Containing 3-Hydroxypropyl Moiety (General Method)

A solution of corresponding nitrone 4a,b (10 mmol) in anhydrous diethyl ether (10 mL) was added dropwise within 20 min to a suspension of lithium aluminum hydride (1.14 g, 29.9 mL). The reaction mixture was cooled in an ice bath, and excess of lithium aluminum hydride was quenched with water (5 mL). The ether layer was separated by decantation and residue was washed with ether (2 × 50 mL). The combined organic layers were dried with anhydrous Na_2SO_4, the solvent was evaporated in vacuum, and the residue was purified by column chromatography (SiO\textsubscript{2}, chloroform-methanol 15:1 mixture as an eluent, detected under UV lamp).

3-(5-tert-Butyl-2-ethyl-1-oxido-3,4-dihydro-2H-pyrrole-2-yl)-1-propanol (11a): 0.95 g, yield 42%. Colorless oil. HRMS (EI/DFS) m/z [M]+ calcd. for (C_{13}H_{25}NO_3)^+: 227.1880; found:227.1882. IR (neat) ν_{max}: 3346 (O–H), 1583 (C = N). ^1H NMR (400 MHz; CDCl_3, δ): 0.70 (t, J = 7.4 Hz, 3H), 1.17 (s, 9H, t-Bu), 1.22–1.34 (m, 1H), 1.34–1.51 (m, 3H), 1.73–1.89 (m, 4H), 2.45–2.51 (m, 2H), 3.46 (dt, J = 6.2 Hz, J = 1.8 Hz, 2H), 4.21 (br.s, 1H); 13C NMR (100 MHz; CDCl_3, δ): 7.27, 23.63, 25.24, 26.05, 27.56, 30.98, 33.32, 33.53, 61.62, 80.12, 152.49.

3,3'-{5-tert-Butyl-1-oxido-3,4-dihydro-2H-pyrrole-2,2-diyl}di(1-propanol) (11b): 1.64 g, yield 64%. White crystalline solid, m.p. 136.7–137.2 °C (ethyl acetate). Elemental analysis: found: C, 65.33; H, 10.57; N, 5.44; calcd. for C_{14}H_{27}NO_4: C, 65.53; H, 10.78; N, 5.51%; IR (KBr) ν_{max}: 3344 (O–H), 1600 (C=O). ^1H NMR (500 MHz; CDCl_3, δ): 1.20 (s, 9H, t-Bu), 1.23–1.33 (m, 2H), 1.42–1.55 (m, 4H), 1.81–1.91 (m, 4H), 2.52–2.56 (m, 2H), 3.49 (t, J = 6.2 Hz, 4H), 4.38 (br.s, 2H); 13C NMR (125 MHz; CDCl_3, δ): 24.19, 25.35, 26.02, 27.75, 33.50, 34.20, 61.60, 79.87, 154.38.

3.2.8. Reaction of Nitrone 11a and 11b with 2,2-Dimethoxypropane (General Method)

A mixture of 11a or 11b (10 mmol), 2,2-dimethoxypropane (48.9 mL, 400 mmol), PPTS (502 mg, 2 mmol), molecular sieves 4 Å (10 g) and anhydrous chloroform (50 mL) was stirred at room temperature for 3 h. The reaction was monitored by TLC (SiO\textsubscript{2}, chloroform-methanol 10:1 mixture; stained with Dragendorff’s reagent). The reaction mixture was cooled in an ice bath, and excess of lithium aluminum hydride was quenched with water (5 mL). The ether layer was separated by decantation and residue was washed with ether (2 × 50 mL). The combined organic layers were dried with anhydrous Na_2SO_4, the solvent was evaporated in vacuum and the crude residue was stirred with chloroform (50 mL) and manganese dioxide (3.44 g, 40 mmol) at room temperature for 3 h. The reaction was monitored by TLC (SiO\textsubscript{2}, chloroform-methanol 10:1 mixture; stained with Dragendorff’s reagent). Manganese oxides were filtered off, precipitate was washed with chloroform (30 mL) and methanol (30 mL), the filtrate was evaporated in vacuum, and the residue was purified by column chromatography (SiO\textsubscript{2}, chloroform-methanol 15:1 mixture as an eluent, detected under UV lamp).

5-tert-Butyl-2-ethyl-2-[3{(1-methoxy-1-methylethoxy)propyl]-3,4-dihydro-2H-pyrrole 1-oxide (12a): 2.48 g, yield 83%. Colorless oil. HRMS (EI/DFS) m/z [M]+ calcd. for (C_{17}H_{33}NO_3)^+: 299.2455; found: 299.2465. IR (neat) ν_{max}: 1578 (C = N). ^1H NMR (500 MHz; CDCl_3, δ): 0.75 (t, J = 7.4 Hz, 3H), 1.21 (s, 9H, t-Bu), 1.23 (s, 6H), 1.52 (dt, J = 13.2 Hz, J = 4.4 Hz, 1H), 1.77 (dt, J = 13.2 Hz, J = 4.4 Hz, 1H), 1.81–1.88 (m, 3H), 2.48–2.53 (m, 2H), 3.09 (s, 3H), 3.22–3.29 (m, 1H), 3.32–3.37 (m, 1H); 13C NMR (125 MHz; CDCl_3, δ): 7.37, 23.69, 23.90, 24.20, 25.31, 27.48, 30.74, 33.30, 34.63, 48.21, 60.37, 79.89, 99.53, 150.58.

5-tert-Butyl-2,2-bis [3{(1-methoxy-1-methylethoxy)propyl]-3,4-dihydro-2H-pyrrole 1-oxide (12b): 3.2 g, yield 80%. Colorless oil. HRMS (EI/DFS) m/z [M]+ calcd. for (C_{22}H_{43}NO_5)^+:
3.2.9. Reaction of Nitrones 12a and 12b with Ethyllithium (General Method)

A solution of ethyllithium in n-pentane (0.7 M, 70 mL) was slowly added dropwise to a solution of 12a or 12b (6 mmol) in dry benzene (10 mL) upon stirring under argon. The mixture was stirred for 20 h at room temperature, cooled in the ice bath, and quenched with water (20 mL). The organic layer was separated and water layer was extracted with diethyl ether (2 × 30 mL). Combined extracts were dried with Na₂SO₄ and the solvent was evaporated in vacuum. The crude residue was dissolved in methanol (70 mL) and basified with aqueous solution of sodium hydroxide (1 M, 20 mL). Methylene blue (6 mg, 0.02 mmol) was added to the mixture, and the air was bubbled until the solution had turned dark blue. The methanol was distilled off in vacuum, and the residue was extracted with diethyl ether (3 × 30 mL). Combined extract was dried with Na₂SO₄ and the solvent was evaporated in vacuum. The residue was dissolved in the 30 mL of mixture methanol:water (1:1), and PPTS (500 mg) was added. The mixture was stirred for 1 h at room temperature, methanol was distilled off in vacuum and residue was extracted with diethyl ether (3 × 30 mL). Combined extract was dried with Na₂SO₄ and the solvent was evaporated in vacuum. The residue was purified by column chromatography (SiO₂, chloroform-methanol 30:1 or hexane-ethyl acetate:1 mixture as an eluent, detected under UV lamp) to give desired nitroxide.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/molecules27061922/s1, Figures S1–S13: IR spectra of 4a,b, 5a,b, 7a,a’, 7a, 7a,a”, 8a, 10a, 11a,b, 12a,b, 14a,b; Figures S14–S39: NMR spectra of 4a,b, 5a,b, 7a,a”, 8a, 10a_red, 11a,b, 12a,b, 14a_red, 14b_red; Figure S40: HPLC analysis data of 14a.
Author Contributions: Synthesis A.I.T.; EPR experiments Y.I.G.; NMR experiments I.V.E.; HPLC analysis E.I.C.; writing—original draft preparation, A.I.T. and I.A.K.; writing—review and editing, I.A.K.; supervision, I.A.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Russian Scientific Foundation (grant number 21-73-00281).

Data Availability Statement: Data are contained within the article or Supplementary Material.

Acknowledgments: We thank the personnel of the Multi-Access Center of SB RAS for recording IR, NMR, and HRMS spectra.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds 10a, 14a and 14b are available from the authors.

Appendix A

The 1D and 2D NMR spectra of 10a_red were recorded from a mixed CD$_3$OD/CDCl$_3$ solution at 303K on a Bruker Avance III 500 FT-spectrometer with working frequencies 500.13 and 125.76 MHz for $^1$H and $^{13}$C, respectively. $^1$H and $^{13}$C NMR chemical shifts are reported in ppm of the δ scale and referred to signals of the CD$_3$OD (3.31 ppm for residual protons for $^1$H and 49.15 for $^{13}$C NMR spectra). Signals were assigned using 2D (HSQC, HMBC, COSY) NMR techniques.

The presence of two asymmetric centers causes nonequivalent of hydrogen atoms in the methylene groups of the molecule. Two methylene groups of pyrrolidine were identified using $^1$H, $^{13}$C-HSQC, and HMBC techniques. Two of four signals overlapped with other multiplets, but the approximate value of $J_{HH}$-coupling constant can be elicited for them: for the first methylene group: $\delta_C = 28.57$ ppm, $\delta_{H_1} = 1.99$ ($J_1 = 10.4$ Hz, $J_2 = 6.7$ Hz, $J_3 = 3.3$ Hz) and $\delta_{H_2} = 2.17$ ($J_1 = 10.4$ Hz, $J_2 = 7.6$ Hz) ppm; for the second: $\delta_C = 38.01$ ppm, $\delta_{H_1} = 1.90$ ($J_1 = 13$ Hz, $J_2 = 7.6$ Hz, $J_3 = 3.3$ Hz) and $\delta_{H_2} = 2.08$ ($J_1 = 13$ Hz, $J_2 = 6.7$ Hz). The third $J$-constants were not observed for signals at $\delta_{H_1} = 2.17$ and $\delta_{H_2} = 2.08$ ppm, so their values were approximately equal to 0 Hz. So, synclinal orientation for protons pairs with angle close to 90° at 1.90 and 1.99 ppm on the one hand, and 2.08 and 2.17 ppm on the other hand can be suggested.

![Figure A1](image-url)

**Figure A1.** (top) Real and (bottom) simulated (fragment of pyrrolidine ring) $^1$H-NMR spectra.

Weak cross-peaks in the $^1$H,$^1$H–NOESY spectrum showed interaction of the methyl group at 1.09 ppm with protons at 2.17 ppm (methylene group of pyrrolidine ring) and at 2.05 ppm (ethyl group). Some cross-peaks for the other side of molecule were also detected: between signals at 0.93 and 2.58 ppm, 1.72 and 2.08 ppm, 1.79 and 1.90 ppm, 2.03 and 2.18 ppm. So, we can conclude that tert-butyl and ethyl groups were oriented in the same plane with hydrogen atoms at 2.08 and 2.17 ppm of the pyrrolidine cycle.
References

1. Ouari, O.; Gigmes, D. Nitroxides: Synthesis, Properties and Applications; Royal Society of Chemistry: London, UK, 2021; ISBN 978-1-78801-752-7. [CrossRef]

2. Kokorin, A.I. (Ed.) Nitroxides—Theory, Experiment and Applications; IntechOpen: London, UK, 2012; ISBN 978-9535107224.

3. Likhtenshtein, G.I.; Yamauchi, J.; Nakatsuji, S.; Smirnov, A.; Tamura, R. Nitroxides. Applications in Chemistry, Biomedicine and Material Science; Wiley: New York, NY, USA, 2008; ISBN 978-3-527-31889-6. Available online: https://www.intechopen.com/books/2507 (accessed on 17 February 2022).

4. Paletta, J.T.; Pink, M.; Foley, B.; Rajca, S.; Rajca, A. Synthesis and reduction kinetics of sterically shielded pyrrolidine nitroxides. Org. Lett. 2012, 14, 5322–5325. [CrossRef] [PubMed]

5. Jagtap, A.P.; Krstic, I.; Kunjir, N.C.; Hänsel, R.; Prisner, T.F.; Sigurdsson, S.T. Sterically shielded spin labels for in-cell EPR spectroscopy: Analysis of stability in reducing environment. Free Radic. Res. 2015, 49, 78–85. [CrossRef] [PubMed]

6. Ovcherenko, S.S.; Chinak, O.A.; Chechushkov, A.V.; Dobrynin, S.A.; Kirilyuk, I.A.; Krumkacheva, O.A.; Richter, V.A.; Bagryan-skaya, E.G. Uptake of cell-penetrating peptide tfl2 by human lung cancer cells: Monitoring by electron paramagnetic resonance and confocal laser scanning microscopy. Molecules 2021, 26, 5442. [CrossRef] [PubMed]

7. Ketter, S.; Dajka, M.; Rogozhnikova, O.; Dobrynin, S.A.; Tormyshev, V.M.; Bagryanskaya, E.G.; Joseph, B. In situ distance measurements in a membrane transporter using maleimide functionalized orthogonal spin labels and 5-pulse electron-electron double resonance spectroscopy. J. Magn. Reson. Open 2022, 10–11, 100041. [CrossRef] [PubMed]

8. Karthikeyan, G.; Bonucci, A.; Casano, G.; Gerbaud, G.; Abel, S.; Thomé, V.; Kodjabachian, L.; Magalon, A.; Guigliarelli, B.; Belle, V.; et al. A Bioreistant Nitroxide Spin Label for In-Cell EPR Spectroscopy: In Vitro and In Oocytes Protein Structural Dynamics Studies. Angew. Chem. Int. Ed. 2018, 57, 1366–1370. [CrossRef] [PubMed]

9. Soikkeli, M.; Kettunen, M.I.; Niväjärvi, R.; Olsion, V.; Rönkkö, S.; Laakkonen, J.P.; Lehto, V.P.; Kavakka, J.; Heikkinen, S. Assessment of the Relaxation-Enhancing Properties of a Nitroxide-Based Contrast Agent TEEPO-Glc with in Vivo Magnetic Resonance Imaging. Contrast Media Mol. Imaging 2019, 2019, 5629997. [CrossRef] [PubMed]

10. Emoto, M.; Mito, F.; Yamasaki, T.; Yamada, K.I.; Sato-Akaba, H.; Hirata, H.; Fujii, H. A novel ascorbic acid-resistant nitroxide in fat emulsion is an efficient brain imaging probe for in vivo EPR imaging of mouse. Free Radic. Res. 2011, 45, 1325–1332. [CrossRef] [PubMed]

11. Dobrynin, S.A.; Glazachev, Y.I.; Gatilov, Y.V.; Chernyak, E.I.; Salnikov, G.E.; Kirilyuk, I.A. Synthesis of 3,4-Bis(hydroxymethyl)-2,2,5,5-tetraethylpyrrolidin-1-oxyl via 1,3-Dipolar Cycloaddition of Azomethine Ylide to Activated Azalene. J. Org. Chem. 2018, 83, 5392–5397. [CrossRef] [PubMed]

12. Lampp, L.; Morgenstern, U.; Merzweiler, K.; Imming, P.; Seidel, R.W. Synthesis and characterization of sterically and electrostatically shielded pyrrolidine nitroxide radicals. J. Mol. Struct. 2019, 1182, 87–94. [CrossRef]

13. Dobrynin, S.A.; Usatov, M.S.; Zhurko, I.F.; Morozov, D.A.; Polienko, Y.F.; Glazachev, Y.I.; Parkhomenko, D.A.; Tyumentsev, M.A.; Gatilov, Y.V.; Chernyak, E.I.; et al. A Simple Method of Synthesis of 3-Carboxy-2,2,5,5-Tetraethylpyrrolidine-1-oxyl and Preparation of Reduction-Resistant Spin Labels and Probes of Pyrrolidine Series. Molecules 2021, 26, 5761. [CrossRef] [PubMed]

14. Zhurko, I.F.; Dobrynin, S.; Gorodetski, A.A.; Glazachev, Y.I.; Rybalova, T.V.; Chernyak, E.I.; Asanbaeva, N.; Bagryanskaya, E.G.; Kirilyuk, I.A. 2-Butyl-2-tert-butyl-5,5-diethylpyrrolidine-1-oxyls: Synthesis and properties. Molecules 2020, 25, 845. [CrossRef] [PubMed]
15. Bobko, A.A.; Kirilyuk, I.A.; Gritsan, N.P.; Poloyvanenko, D.N.; Grigor’ev, I.A.; Khramtsov, V.V.; Bagryanskaya, E.G. EPR and Quantum Chemical Studies of the pH-sensitive Imidazoline and Imidazolidine Nitroxides with Bulky Substituents. *Appl. Magn. Reson.* **2010**, *39*, 437–451. [CrossRef] [PubMed]

16. Keana, J.F.W.; Pou, S.; Rosen, G.M. Synthesis and properties of some nitroxide α-carboxylate salts. *J. Org. Chem.* **1989**, *54*, 2417–2420. [CrossRef]

17. Black, D.S.C.; Edwards, G.L.; Evans, R.H.; Keller, P.A.; Laaman, S.M. Synthesis and reactivity of 1-pyrroline-5-carboxylate ester 1-oxides. *Tetrahedron* **2000**, *56*, 1889–1897. [CrossRef]

18. Earla, A.; Walter, E.D.; Braslau, R. Synthesis and spin trapping properties of polystyrene supported trifluoromethylated cyclic nitrones. *Free Radic. Res.* **2019**, *53*, 1084–1100. [CrossRef] [PubMed]

19. Ballini, R.; Bosica, G. Conjugated addition reactions of nitroalkanes with electrophilic alkenes in aqueous media. *Eur. J. Org. Chem.* **1998**, *1998*, 355–357. [CrossRef]

20. Pollini, G.; Barco, A.; De Guili, G. Tetramethylguanidine-catalyzed addition of nitromethane to α,β-unsaturated carboxylic acid esters. *Synthesis* **1972**, *1972*, 44–45. [CrossRef]

21. Golod, E.L.; Bagal, L.I. Destructive nitration of polinitrocarnonylic compounds. I. New method of hexanitroethane synthesis. *Russ. J. Org. Chem.* **1994**, *90*, 29–32.