Risk Assessment of RYR Food Supplements: Perception vs. Reality

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Thirty-seven red yeast rice (RYR) food supplements were screened for their mycotoxin and natural statin content. Products included pure RYR capsules and multi-ingredient formulations with standardized amounts of monacolin K (MK), marketed both online and retail in the European Union. In terms of mycotoxins, citrinin (CIT) was found in all the monitored products. As CIT content ranged from 100 to 25100 µg/kg, only four products were compliant with maximum EU levels in force until April 2020, while a single product was compliant with the limit of 100 µg/kg introduced after that date. Four contaminated products were labeled as “citrinin free”. In terms of natural statins, nine products had a lower content vs. label statements (from −30 to −83%), while for 24 a larger MK amount (from 10 to 266%) was noticed. Three products had a negligible MK content and only 19 offered a daily dosage exceeding 10 mg as dictated by the health claim granted by EFSA in the EU. No sample had label values compliant with pharmaceutical Good Manufacturing Practices requirements (95–105% content of active constituent). Variable, but small amounts of simvastatin (0.1–7.5 µg per daily dose) were found in 30 samples. These results suggest that limited efficacy and reported safety issues may stem from an under-regulated and undercontrolled market, weakening both effectiveness and risk assessment evaluations.

Keywords: dietary supplement, botanical, mycotoxin, Monascus, quality control

INTRODUCTION

Global sales of food supplements are growing steadily, sustained by consumers perceiving them as a safe and healthy option, mostly according to a supposed pharmaceutical quality in validation, manufacturing and control (1, 2). Quality control of these products is a multifaceted process, given the manifold traits and the variety of substances involved, whose complexity is often overlooked by the narrative surrounding them. For instance, in both the European Union and the United States, standards comparable to pharmaceutical Good Manufacturing Practices are not in force for the food supplement industry (3). Likely as a consequence, a variety of contamination and standardization concerns emerged during the last decade. Many supplements are in fact reportedly suffering from unreliable content of active principles, adulterations with foreign (and sometimes illicit or synthetic) substances, substitution with cheaper ingredients and pesticide or mycotoxin contamination (4–7). While these issues may emerge separately, they may also occur at the same time, altering trustworthiness of research data, safety of use and efficacy. To some extent, both conflicting results in clinical trials, difficult translations of scientific evidence into actual effectiveness, emergence of adverse effects and unreliability of most risk-benefit or risk assessment evaluations, may be a relapse of erratic quality of food supplements (3).
Multiple reports have repeatedly highlighted the lack of consistency between label and actual content in food supplement based on bilberry, blueberry, cranberry and ginkgo, to name a few (8–10). Furthermore, the issues summarized by Lockwood et al. in 2011 have not been corrected according to the subsequent literature, with an adulteration rate in herbal products exceeding 25% (11, 12).

Such scenario has prompted researchers to advocate for a closer attention to quality, for more widespread controls and for a stricter legal framework, to close the gap between reality and consumers perception in terms of quality and safety (13, 14).

Within this context, RYR represents a paradigmatic case. Following the demonstration that MK produced by Monascus purpureus grown on polished rice competitively inhibit human 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase, a key enzyme in the biosynthesis of endogenous cholesterol, several studies confirmed the efficacy of RYR as a lipid-lowering agent. Its usefulness was demonstrated multiple times and is further reinforced by the structural equivalence between MK and Lovastatin, the first statin made commercially available in the pharmaceutical market (15). After an extensive review, the European Food Safety Authority (EFSA) approved a specific health claim applicable to any RYR food supplement, stating that “Monacolin K from RYR contributes to the maintenance of normal blood cholesterol concentrations”. In order to obtain the claimed effect, 10 mg of MK from fermented RYR should be consumed daily. At the same time, despite the large commercial success, no specifications for RYR are available in European, Japanese, Canadian, US Pharmacopeias or in the WHO monographs.

The use of RYR is widespread, especially among the general population, in form of over-the-counter supplements produced by multiple brands and suggested as an alternative to prescription drugs. Besides the issue related to the homology between MK and Lovastatin, some criticality related to quality and safety emerged (16). Most relevant ones involve the erratic MK content, the emergence of side effects and the presence of citrinin, a polyketide mycotoxin with mutagenic, genotoxic and carcinogenic properties (17–20).

Citrinin is a nephrotoxic substance produced by few Aspergillus and Penicillium species, but is often detected also in Monascus purpureus cultures. It disrupts renal function leading to necrosis of the distal tubular epithelium, ultimately impairing renal function. It can be found in a wide variety of cereals, seeds and fruits, but its presence is rather common in food supplements based on RYR, to the point that a recent survey on the occurrence of citrinin in food commissioned by EFSA concluded that these supplements are the samples most prone to contamination with citrinin (21, 22).

EFSA also concluded that, “the composition and content of minor monacolins in food supplements containing fermented red rice is variable and the data does not allow to evaluate their contribution to the cholesterol-lowering efficacy” (23). As a consequence, the European Commission was recently persuaded to introduce a stricter limit for the presence of citrinin, whose permitted maximum amount in food supplements containing RYR was reduced in April 2020 to 00 µg/kg (24, 25).

This study was therefore designed to estimate if RYR supplements available in the marketplace when new EU regulation became effective are compliant with the new limits. Further aims concerned the evaluation of concurrent quality issues, the level of compliance between declared and actual MK content and a detailed evaluation of the actual exposure to citrinin and monacolin K in terms of risk assessment.

**MATERIALS AND METHODS**

**Chemicals and Reagents**

Citrinin (solution in acetonitrile 100 µg mL⁻¹), ochratoxin A (solution in acetonitrile 100 µg mL⁻¹) and aflatoxin mix containing AFBI and AFBJ (2 µg mL⁻¹ in acetonitrile), AFG1 and AFG2 (0.5 µg mL⁻¹ in acetonitrile) were obtained from Romer Labs (Tulln, Austria). Monacolin K (25 mg), monacolin K acid (5 mg) and simvastatin (10 mg) were purchased from Vinci Biochem S.R.L. (Florence, Italy).

HPLC-grade ethanol, and acetonitrile were purchased from Sigma-Aldrich (Taufkirchen, Germany); bidistilled water was obtained using Milli-Q System (Millipore, Bedford, MA, USA). MS-grade formic acid from Fisher Chemical (Thermo Fisher Scientific Inc., San Jose, CA, USA) was also used.

**Red Yeast Rice Supplements: Sampling Plan**

Thirty-seven red yeast rice food supplements, including pure RYR (8) and multi-ingredient formulations (26) with standardized amounts of monacolin K, were bought mainly on internet web sites (24) but also from local pharmacies in Italy (13) between September 2020 and February 2021. RYR products were formulated as capsules (18), tablets (15) and soft capsule (4). Four products were labeled as “citrinin free” (see “Note” column in Table 1). Detailed information on all samples with their form, suppliers, RYR daily dose and MKA content are listed in Table 1. All the samples were analyzed before their expiration date with UHPLC-MS/MS technique.

**Sample Preparation for Mycotoxins Analysis**

Tablets were weighed and then pulverized with a mortar and pestle. For capsules, five samples were weighed, opened and the contents mixed and triturated in a mortar and pestle. Soft capsules were punctured, and the liquid content was carefully squeezed into a beaker and weighed.

Sample preparation was performed following the protocol previously optimized by Wang et al., 2014 (27). To put it briefly, one gram of each sample was extracted with 30 ml ethanol/water (70:30) by shaking on a rotary shaker at 200 strokes/min for 30 min, and it was treated with ultrasonication at 40 °C for 30 min, and then shaken on a rotary shaker at 200 rpm for 1.5 h (27). The extract was centrifuged for 10 min at 1,4000 rpm.
TABLE 1 | List of supplements containing red yeast rice.

| No. | Formulation | Daily dose (Cps) | RYR per daily dose (mg) | Monacolin K declared on label (mg) | Supplier | Form | Notes |
|-----|-------------|-----------------|-------------------------|-----------------------------------|----------|------|-------|
| 1   | Multicomponent | 1 | 200 | 10 | Pharmacy | Tablet |       |
| 2   | Multicomponent | 2 | 337.3 | 10 | Pharmacy | Capsule |       |
| 3   | Pure RYR | 2 | N.D. | 10 | On-line pharmacy | Capsule |       |
| 4   | Pure RYR | 2 | 200 | 10 | On-line store | Capsule |       |
| 5   | Pure RYR | 1 | 600 | 4.8 | On-line store | Capsule |       |
| 6   | Multicomponent | 1 | 250 | 10 | On-line store | Capsule | Citrinin free |
| 7   | Pure RYR | 1 | 300 | 12 | On-line pharmacy | Capsule | Citrinin free |
| 8   | Multicomponent | 1 | 210 | 10.5 | On-line pharmacy | Capsule |       |
| 9   | Multicomponent | 1 | 600 | N.D. | On-line pharmacy | Capsule | Citrinin free |
| 10  | Pure RYR | 1 | 150 | 4.5 | On-line store | Tablet |       |
| 11  | Multicomponent | 1 | 200 | 10 | On-line store | Tablet |       |
| 12  | Multicomponent | 2 | 1,200 | N.D. | On-line store | Capsule |       |
| 13  | Multicomponent | 1 | 100 | 3 | Pharmacy | Tablet |       |
| 14  | Multicomponent | 1 | 200 | 10 | On-line store | Capsule |       |
| 15  | Multicomponent | 1 | 200 | 10 | On-line store | Capsule |       |
| 16  | Multicomponent | 1 | 100 | 3 | On-line store | Capsule |       |
| 17  | Multicomponent | 1 | 200 | 10 | On-line store | Capsule |       |
| 18  | Pure RYR | 1 | 200 | 10 | Pharmacy | Tablet |       |
| 19  | Pure RYR | 2 | 1,200 | N.D. | On-line store | Capsule |       |
| 20  | Multicomponent | 2 | 200 | 10 | On-line store | Soft capsules |       |
| 21  | Multicomponent | 1 | 200 | 10 | Pharmacy | Tablet |       |
| 22  | Multicomponent | 2 | 666.7 | 10 | On-line store | Capsule |       |
| 23  | Multicomponent | 1 | 200 | 10 | On-line store | Tablet | Citrinin free |
| 24  | Multicomponent | 1 | 200 | 10 | On-line store | Tablet |       |
| 25  | Multicomponent | 1 | 333.4 | 10 | On-line store | Tablet |       |
| 26  | Multicomponent | 1 | 333 | 10 | On-line store | Capsule |       |
| 27  | Multicomponent | 1 | 333 | 10 | On-line store | Capsule |       |
| 28  | Multicomponent | 1 | 200 | 3 | Pharmacy | Tablet |       |
| 29  | Multicomponent | 1 | 200 | 10 | Pharmacy | Tablet |       |
| 30  | Multicomponent | 1 | 200 | 10 | Pharmacy | Soft capsules |       |
| 31  | Multicomponent | 1 | 370.4 | 10 | Pharmacy | Tablet |       |
| 32  | Multicomponent | 1 | 400 | 5 | On-line store | Soft capsules |       |
| 33  | Multicomponent | 1 | 167 | 5 | On-line store | Tablet |       |
| 34  | Multicomponent | 1 | 333 | 10 | Pharmacy | Tablet |       |
| 35  | Pure RYR | 2 | 338.4 | 10 | Pharmacy | Capsule |       |
| 36  | Multicomponent | 1 | 220 | 3.3 | On-line store | Tablet |       |
| 37  | Multicomponent | 1 | 417 | 10 | On-line store | Soft capsules |       |

N.D., not declared on label. *Statement made on label by the producer.

at room temperature and then supernatants were quantitatively analyzed using LC-MS/MS.

**Targeted UHPLC-MS/MS Analysis of Mycotoxins and Monacolin K**

UHPLC Dionex Ultimate 3000 separation system coupled to a triple quadrupole mass spectrometer (TSQ Vantage; Thermo Fisher Scientific Inc., San Jose, CA, USA) equipped with an electrospray source (ESI) was employed.

For the chromatographic separation, a reversed-phase C18 Kinetex EVO column (Phenomenex, Torrance, CA, USA) with 2.10 × 100 mm and a particle size of 2.6 µm heated to 40 °C was used. 3 µl of sample extract was injected into the system; the flow rate was 0.35 ml/min.

Gradient elution was performed by using water (eluent A) and methanol (eluent B) both acidified with 0.1% formic acid. Initial conditions were set at 30% B followed by a linear change to 90% B in 5.5 min. 100% B was reached in 2.5 min and kept for 2 min, followed by a reconditioning step for 5 min using initial composition of mobile phases. The total run time was 16 min.

MS parameters: the ESI source was operated in positive ionization mode (ESI+); spray voltage 3,000 V, capillary
temperature at 270 °C, vaporizer temperature was kept at 200 °C, sheath gas flow was set at 40 units and the auxiliary gas flow at 5 units. S-Lens RF amplitude value and collision energies (CE) were optimized during infusion of analyte standard solutions (1000 ng mL⁻¹, in methanol) employing an automatic function of X-calibur software (Thermo Fisher Scientific, San Jose, CA, USA). Detection was performed using multiple reaction monitoring (MRM) mode. The following optimized transitions were used for the quantification: CIT m/z 251 → 233 (CE 18); OTA m/z 404 → 238 (CE = 21 eV); AFB1 m/z 317 → 285 (CE = 42 eV); AFB2 m/z 315 → 287 (CE = 25 eV); AFG1 m/z 329 → 243 (CE = 25 eV); AFG2 m/z 331 → 313 (CE = 30 eV); AFM1 m/z 329 → 273 (CE = 25 eV); MK m/z 427 → 335 (CE = 15 eV); MKA m/z 445 → 343 (CE = 20 eV); simvastatin m/z 441 → 325 (CE = 24 eV). Quantification of target analytes was performed using calibration standards (range 0.001–2 ppm for CIT, 1–50 ppm for MKA, 0.0125–1.25 ppm for simvastatin, and 5–100 ppm for MK). Further analytical details are summarized in Supplementary Table S1. Quantification of targeted analytics was performed employing Thermo Xcalibur 2.2.SP1 QuanBrowser software.
**Risk Dietary Exposure**

For each RYR sample, the daily citrinin intake was established based on the suggested dosage and the mean capsule weight (measured in triplicate) and expressed as µg/kg b.w. considering a man of 60 kg. The Margin of Exposure (MOE) was calculated for each supplement, according to the guidelines previously reported (28).

**RESULTS AND DISCUSSION**

The aim of this study was to estimate whether RYR supplements comply with pharmaceutical quality, and whether their composition is consistent with label statements and with normative modifications related to maximum citrinin content introduced in the EU in April 2020. After setting up a dedicated extraction and LC-MS/MS detection protocol, 37 different food supplements were evaluated. The pool comprised both capsules of pure RYR or multi-ingredient formulations with standardized supplements were evaluated. The pool comprised both capsules of pure RYR or multi-ingredient formulations with standardized amounts of MK and other phytochemicals (see Table 1). Multiple endpoints were evaluated, namely monacolin content, potential adulteration, presence of synthetic statins and citrinin content. Data are reported in Tables 2, 3.

**Compliance Between Label and Actual MK Content**

A first evaluation concerned whether some pharmaceutical quality criteria can be applied to food supplements. At present, no standardization is required in the EU, while EFSA dictate that a daily intake of 10 mg of MK is mandatory to obtain the claimed effect, according to EFSA 10 mg of monacolin K per 10 mg of MK is mandatory to obtain the health claim awarded in 2011. During the fermentation process, the form responsible for the inhibition of cholesterol synthesis (MKA) prevails, favored by water solubility and slightly acidic pH, with a content of approximately 80% on average. The final, drying step in RYR production induces a partial cyclization of MKA. Therefore, in the finished product, the acid: lactone ratio usually varies from 6:4 to 4:6. The lactone form (MK) can be hydroylized into MKA in vivo by the action of hydroxyl esterase (see Figure 1).

To provide an unequivocal determination, calculations were therefore made according to total monacolin K content, i.e. considering both lactone (MK) and acidic form (MKA). Data reported in Table 2 confirm that MK is not standardized in food supplements at present available in both retail and online marketplaces, with major relapses in terms of efficacy, safety and exposure. Nine products provided a lower content as compared to values declared on labels (from −30 to −83%), 24 had a larger MK amount (from 10 to 266%), and three products had negligible MK contents. The actual MK dosage was highly variable, exceeding a 30-fold difference from 1.2 to 36 mg/day and confirming a trend emerged during a recent survey on 26 brands of RYR from large retail chains in the United States (30). No sample had label values compliant with pharmaceutical Good Manufacturing Practices (GMP) requirements (95–105% content of active constituent). In 2011 a health claim has been awarded to RYR by EFSA, stating that “Monacolin K from RYR contributes to the maintenance of normal blood cholesterol concentrations”. Such claim boosted sales of the ingredient and is supported by a large scientific evidence in multiple clinical trials. In order to obtain the claimed effect, according to EFSA 10 mg of monacolin K from fermented RYR preparations should be consumed daily and therefore most producers state to provide such amount. According to our data, however, only five brands provide a content in an arbitrary range comprised between 8 and 12 mg/die.

Monacolin K biosynthesis is reportedly variable according to multiple factors, including strain, temperature, fermentation.

**TABLE 3 | Citrinin content and MOE determination for 37 RYR food supplements.**

| Sample | µg/g | µg/daily dose | ug/kg bw* | MOE |
|--------|------|---------------|-----------|-----|
| 1      | 4.7 ± 0.2 | 1.97 | 0.033 | 608 |
| 2      | 8.8 ± 2.0 | 3.27 | 0.138 | 145 |
| 3      | 2.3 ± 0.02 | 0.97 | 0.035 | 580 |
| 4      | 1.9 ± 0.1 | 0.73 | 0.027 | 734 |
| 5      | 3.2 ± 0.1 | 1.23 | 0.041 | 493 |
| 6      | 4.1 ± 0.2 | 1.23 | 0.034 | 585 |
| 7      | 2.5 ± 0.2 | 1.35 | 0.023 | 889 |
| 8      | 3.2 ± 0.05 | 0.84 | 0.038 | 528 |
| 9      | 2.6 ± 0.3 | 1.30 | 0.022 | 923 |
| 10     | 8.7 ± 0.2 | 4.79 | 0.080 | 251 |
| 11     | 3.0 ± 0.1 | 1.98 | 0.083 | 241 |
| 12     | 3.4 ± 0.01 | 1.87 | 0.083 | 624 |
| 13     | 12.3 ± 0.7 | 5.66 | 0.094 | 212 |
| 14     | 4.3 ± 0.3 | 2.28 | 0.038 | 527 |
| 15     | 2.1 ± 0.04 | 1.18 | 0.020 | 1,020 |
| 16     | 0.1 ± 0.1 | 0.03 | 0.001 | 36,364 |
| 17     | 4.1 ± 0.1 | 1.66 | 0.103 | 195 |
| 18     | 5.4 ± 0.2 | 2.81 | 0.137 | 146 |
| 19     | 4.5 ± 0.1 | 2.43 | 0.041 | 494 |
| 20     | 25.1 ± 1.8 | 23.09 | 0.385 | 52 |
| 21     | 9.2 ± 0.2 | 4.97 | 0.083 | 242 |
| 22     | 7.3 ± 0.1 | 3.80 | 0.063 | 316 |
| 23     | 1.1 ± 0.04 | 1.23 | 0.021 | 974 |
| 24     | 8.2 ± 0.3 | 4.10 | 0.068 | 293 |
| 25     | 4.8 ± 0.001 | 4.80 | 0.080 | 250 |
| 26     | 10.7 ± 0.5 | 8.35 | 0.139 | 144 |
| 27     | 8.8 ± 0.2 | 6.86 | 0.114 | 175 |
| 28     | 8.0 ± 0.4 | 6.48 | 0.108 | 185 |
| 29     | 0.9 ± 0.1 | 3.47 | 0.058 | 345 |
| 30     | 3.2 ± 0.7 | 4.29 | 0.071 | 280 |
| 31     | 8.7 ± 0.07 | 5.66 | 0.094 | 212 |
| 32     | 4.6 ± 2.2 | 4.22 | 0.070 | 284 |
| 33     | 2.9 ± 0.2 | 2.90 | 0.048 | 414 |
| 34     | 5.7 ± 0.6 | 4.79 | 0.080 | 251 |
| 35     | 13 ± 0.2 | 5.98 | 0.100 | 201 |
| 36     | 7.2 ± 0.6 | 3.60 | 0.060 | 333 |

* Considering a reference body weight for human of 60 kg [see ref. #(28)].
techniques and parameters that may lead to an unsatisfactory content in *Monascus* fermentations (26, 31). Precisely for this reason, standardization and post-production control should be extremely accurate, in order to guarantee the same dosage to any consumer and limit the risk of side effects due to excessive administration. The lack of standardization reported represents instead a major shortcoming, outlining a difficult translation from evidence-based science to commercial reality, as clinical and nutritional trials carried out with pure and carefully weighed compounds may not be replicated at post-marketing level. The same critical point concerns the way experimental protocols are set up in those clinical trials using actual supplements instead of formulation prepared (and dosed) on purpose (32). On this regard, a strong commitment to careful analytical determination of MK should be reiterated for any trial intended to test the efficacy of commercial food supplements. As other authors have suggested, results from trials in which an accurate standardization of RYR supplements are not performed may result in unreliable and contradictory data (18, 30). The same applies when exposure is determined to estimate adverse effects and their incidence in the general population: as lipid lowering effects, also most common side effects of RYR are related to MK content and a higher or lower incidence or the emergence of case reports may be ultimately the result of inappropriate dosage stemming from poor standardization, as the 10 products exceeding 90% of declared MK seem to suggest. The trend toward excess is particularly relevant especially considering that the MK contained in the RYR would have a higher bioavailability than the drug to the point that, according to a study (33), 5–6 mg would be equivalent to 20–40 mg of pure, pharmaceutical-grade lovastatin.

**Potential Adulteration**

Nannoni et al. found that monacolin K and its acidic form have a proportional relationship, which can be used to determine whether commercial lovastatin has been added to the RYR (34). By applying the same formula to our data (Table 2), we found that the ratio between the MK / MK + MKA areas is lower than 0.3 in 29 samples, meaning that 80% of the total may be adulterated by the addition of MK from external sources, likely in the form of lovastatin produced by *Aspergillus terreus* or by other means (18, 35). Variable, but limited, amounts of simvastatin (0.1–7.5 µg per daily dose) were also found in 30 samples (Table 2). While these amounts are of no concern, such evidence may open interesting perspectives, as simvastatin is reportedly a synthetic statin. Further investigations may disclose its origin, whether these trace amounts are a consequence of some kind of adulteration (e.g. an impurity from the addition of foreign material) or the result of a previously unreported synthetic capability of *Monascus purpureus*. In both cases the results would be noteworthy, as simvastatin is a derivative of lovastatin whose production is a multistep process. A biocatalytic process starting from monacolin J and a fermentative process for biotransformation of lovastatin into simvastatin, based on *Aspergillus terreus* ATCC20542 fed with 2,2- dimethylbutyrate are available (36).

**Mycotoxin Content**

Since 2017 EFSA clearly identified RYR as the food product with the highest incidence of citrinin contamination (22). Multiple investigations reported frequent and sometimes alarming amounts of this mycotoxin in RYR and our data confirm this evidence. For instance, these data are agreement with previous surveys, in which citrinin was quantified in RYR ranging from 10 to 44240 µg/kg (21, 37, 38). By comparing these literature data with the citrinin levels collected by our study, it can be confirmed that citrinin is almost always present in red rice supplement samples, with values often higher than the legal limit. All tested samples had in fact a citrinin content above LOQ, with values ranging from 100 to 25100 µg/kg, translated in daily dosages comprised between 0.05 and 23.1 µg (Table 3). Four products were compliant with maximum citrinin levels in force in the EU until April 2020, while only a single product was compliant with the limit of 100 µg/kg introduced after that date. Four contaminated products were labeled as “citrinin free”, posing worries of the relevance of this unregulated, self-declared claim which, besides its actual unreliability, seems also to deceptively suggest that products otherwise labeled are of inferior quality.

No ochratoxins nor aflatoxin B1, B2, G1, G2 or M1 were detected in neither sample. However, in 29 samples, an isomeric form of ochratoxin A (OTA) was found. The latter and OTA shown identical fragmentation patterns (see Supplementary Figure S1, Supplemental files), making a mass spectrometric differentiation impossible. On the other hands, the later chromatographic elution (9.7 min) of such isomeric form from the C18 column compared with OTA standard (7.4 min) is consistent with the putative annotation as 2′R-ochratoxin.
A (2′R-OTA) (see Supplementary Figure S1, Supplemental files). Cramer and co-authors (39) previously reported the identification of 2′R-OTA as the thermal degradation product of OTA in coffee. Its presence, if confirmed with the analytical standard, suggests that OTA may be produced by Monascus purpureus or more likely inherited by rice during fermentation after and then subject to degradation during the drying step. This modified form of OTA has a more than seven-fold higher biological half-life in human blood compared to parent compound and while amounts are negligible from toxicological standpoint, its presence suggest that careful, wide-spectrum mycotoxin monitoring could be advisable for RYR.

EFSA has set a level of no concern for citrinin-caused nephrotoxicity at 0.2 µg/kg body weight per day, but it has acknowledged the need for more reliable data on the citrinin level in food and feed before it can undertake exposure studies and risk assessment (40).

Our survey suggests that such reliability, given the wide range of contamination detected, is difficult to obtain.

**Exposure Assessment to Citrinin**

Due to the use of RYR as a natural alternative to statins in lipid-lowering control, it is usually consumed on a regular basis for a long period of time. Therefore, the exposure to possible contaminants should be regarded as chronic.

Based on the scientific evidence, EFSA has classified citrinin as a genotoxic compound. Accordingly, the assessment of possible safety concerns arising from its presence in food and feed, should follow a margin of exposure (MOE) approach. The MOE is the ratio of no-observed-adverse-effect level (NOAEL) obtained from animal toxicology studies to the estimated human exposure level. Under this approach, a MOE ≥ 10,000 is considered of no concern for genotoxic and carcinogenic compounds (28). Although usually for genotoxic compounds a benchmark dose (BMD) approach is usually preferred for the derivation of health-based guidance value, in the case of citrinin only a level of no concern of 0.2 µg/kg b.w. per day is available so far (41). This approximation might lead to an underestimation of the associated risk, being the NOAEL less conservative than the benchmark dose.

The exposure assessment was calculated for all the considered samples, based on a reference body weight for human of 60 kg (29), and reported in Table 3. It can be noticed that in 34 out of 37 samples, the daily intake exceeds the NOAEL, with a maximum of 0.385 µg/kg b.w. for sample #21. When it comes to the MOE, the values are far from the safety threshold of 10,000 for all the batches, with the only exception of sample #17.

Although largely cautionary, the MOE approach clearly shows that the exposure to citrinin from RYR supplements may pose a risk for the consumers. Besides arguments related to public health and regulatory measures, our outcome is even more relevant in terms of communication, when considering that a “citrinin-free” claim was reported on the label of four batches samples in this study. The halo of naturality of RYR supplements together with the associated “claims”, may prompt the consumers to subjectively increase the daily dosage without any perception of possible risk. Therefore, such unsolicited claims should be strongly mistrusted and carefully regulated.

It must be noticed that, compared to food the intake of RYR supplements is less affected by variability ad a consequence of their pre-dosed form. Users, often prompted by health-related issues, take supplements on a regular base according to the suggested dosage, and often choose the same brand over time. While the exposure assessment to contaminants may vary a lot between mean and high-consumers of a certain food, in the case of RYR supplements the variability – and therefore the associated uncertainty – is strongly reduced. As some recent investigations have noticed variability the urinary excretion of citrinin as a marker to assess the exposure, it would be advisable if similar research in the future could track RYR users within the screened population (42).

Considering the toxicological relevance of citrinin, our data poses a serious health concern, and once again underline the need of quality standard and control plan for food supplements.

Overall, our investigation confirms that quality of RYR food supplements is dubious and strict quality control should be implemented. Most commercial RYR products do not meet the maximum level of citrinin allowed in the EU starting April 2020. To date, (accessed 9 January 2021) RASFF database reports only 9 notifications regarding fermented red rice supplements, starting since 2010. Three were related to citrinin content and one to an excess of monacolin; given the occurrence reported in our and in other recently published papers for both these substances, post marketing controls vastly underestimate the scale of the problem. Therefore, pharmaceutical quality criteria should not be taken for granted by both researchers, clinicians, regulators, producers and consumers dealing with RYR. More rigorous standardization and monitoring for mycotoxin content must be encouraged to extrapolate data for risk assessment with respect to the prevalence of citrinin. Given the widespread use, the equivalence between MK and lovastatin and the occurrence of side effects, for RYR supplements the enforcement of pharmaceutical GMP could be a response in order to mitigate the present situation.

**DATA AVAILABILITY STATEMENT**

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

**AUTHOR CONTRIBUTIONS**

LR, RB, and CD’A conceived the study. LR was responsible for the sampling plan, planned and carried out the LC-MS analysis, and the data elaboration. RB and CD’A contributed to the interpretation of the analytical results. RB supervised the project. All authors contributed to the writing process and critically revised the final manuscript.
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REFERENCES

1. Egan B, Hodgkins C, Shepherd R, Timotijevic L, Raats M. An overview of consumer attitudes and beliefs about plant food supplements. Food Funct. (2011) 2:747–52. doi: 10.1039/c1fo10109a
2. Sirico F, Miressi S, Castaldo C, Spera N, Montagnani S, Di Meglio F, et al. Habits and beliefs related to food supplements: Results of a survey among Italian students of different education fields and levels. PLoS ONE. (2018) 13:e0191424. doi: 10.1371/journal.pone.0191424
3. Fibigá J, Šatínský D, Solich P. Current trends in the analysis and quality control of food supplements based on plant extracts. Anal Chim Acta. (2018) 1036:1–15. doi: 10.1016/j.aca.2018.08.017
4. Pickova D, Ostry V, Toman J, Malíř F. Presence of mycotoxins in milk thistle (silybum marianum) food supplements: a review. Toxins. (2020) 12:1–21. doi: 10.3390/toxins120120782
5. Vaclavik L, Kryntskyi AJ, Rader JL. Mass spectrometric analysis of pharmaceutical adulterants in products labeled as botanical dietary supplements or herbal remedies: a review. Anal Bioanal Chem. (2014) 406:6767–90. doi: 10.1007/s00216-014-8159-2
6. Costa JG, Vidovic B, Saraiva N, do Ceu Costa M, Del Favero G, Marko D, et al. Contaminants: a side of dark food supplements? Free Radic Res. (2019) 53:1113–35. doi: 10.1080/10715762.2019.1636045
7. Martínez-Sanz J, Sospedra I, Ortiz E, Baladía E, Gil-Izquierdo A, Ortiz- Moncada R. Intended or unintended doping? A review of the presence of doping substances in dietary supplements used in sports. Nutrients. (2017) 9:1093. doi: 10.3390/nu9101093
8. Czigle S, Tóth J, Jedlinszki N, Házsnagy-Radnai E, Csupor D, Tekeleva D. Presence of mycotoxins in milk thistle (silybum marianum) food supplements: a review. Oxid Med Cell Longev. (2014) 2014:976237. doi: 10.1155/2014/976237

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnut.2021.792529/full#supplementary-material

17. Avula B, Cohen PA, Wang YH, Sagi S, Feng W, Wang M, et al. Chemical profiling and quantification of monacolins and citrinin in red yeast rice commercial raw materials and dietary supplements using liquid chromatography-accurate QToF mass spectrometry: chemometrics application. Public Health Nutr. (2014) 100:243–53. doi: 10.1016/j.jph.2014.07.039
18. Song J, Luo J, Ma Z, Sun Q, Wu C, Li X. Quality and authenticity control of functional red yeast rice—a review. Molecules. (2019) 24:1944. doi: 10.3390/molecules24101944
19. Gordon Ry, Cooperman T, Obermeyer W, Becker Dj. Marked variability of monacolin levels in commercial red yeast rice products: Buyer beware! Arch Intern Med. (2010) 170:1722–7. doi: 10.1001/archinternmed.2010.382
20. Heber D, Lembertas A, Lu QY, Bowerman S, Go VlV. An analysis of nine proprietary chinese red yeast rice dietary supplements: Implications of variability in chemical profile and contents. J Altern Complement Med. (2001) 7:133–9. doi: 10.1089/107555301750164181
21. Silva LJG, Pereira AMPT, Pena A, Lino CM. Citrinin in foods and supplements: a review of occurrence and analytical methodologies. Foods. (2020) 10:14. doi: 10.3390/foods10010014
22. López Sánchez P, de Nijsa M, Spanjerb M, Pietric A, Bertuzzi T, Staraki A, et al. Generation of occurrence data on citrinin in food. EFSA Support Publ. (2017) 14:1177E. doi: 10.2903/sp.efsa.2017.EN-1177
23. Younes M, Aggett P, Aguilar F, Crebelli R, Dusemund B, Filippí M, et al. Scientific opinion on the safety of monacolins in red yeast rice. EFSA J. (2018) 16:05368. doi: 10.2903/j.efsa.2018.5368
24. Commission of the European Union. COMMISSION REGULATION (EC) No 1881/2006. Off Eur Union. (2006) 364–5.
25. Commission of the European Union. (2019). Commission Regulation (EU) No 1881/2006 amending Regulation (EC) No 1881/2006 as regards maximum levels of citrinin in food supplements based on rice fermented with red yeast Monascus purpureus. EFSA Support Publ. (2019) 12:44081–2. doi:10.2903/sp.efsa.2019.EN-124081
26. Huang HN, Huu YY, Bao GR, Xie LH. The quantification of monacolin K in some red yeast rice from Fujian province and the comparison of the other product. Chem Pharm Bull. (2006) 54:687–9. doi: 10.1248/cpb.54.687
27. Wang W, Chen Q, Zhang X, Zhang H, Huang Q, Li D, et al. Comparison of extraction methods for analysis of citrinin in red fermented rice. Food Chem. (2014) 157:408–12. doi: 10.1016/j.foodchem.2014.02.060
28. EFSA. Statement on the applicability of the Margin of Exposure approach for the safety assessment of impurities which are both genotoxic and carcinogenic in substances added to food/feed. EFSA J. (2012) 10:2578. doi: 10.2903/j.efsa.2012.2578
29. Center for Drug Evaluation and Research. Guidance for industry: estimating the maximum safe starting dose in initial clinical trials for therapeutics in adult healthy volunteers. US Dep Heal Hum Serv. (2005) 22:883-9. doi: 10.1186/1477-7525-2-49
30. Cohen PA, Avula B, Khan IA. Variability in strength of red yeast rice supplements purchased from mainstream retailers. Eur J Prev Cardiol. (2017) 24:1431–4. doi: 10.1177/2047487317715714
31. Huang CF, Shen SM, Chen WT, Chen CC. The effects of mutation and citrinin in red yeast rice and CYP450 enzymes/P-glycoprotein and its implication for the metabolism and carcinogenic in substances added to food/feed. EFSA J. (2019) 16:05368. doi: 10.2903/j.efsa.2019.5368
32. Chen C-H, Uang Y-S, Wang S-T, Yang J-C, Lin C-J. Interaction between Red Yeast Rice and CYP450 Enzymes/P-glycoprotein and its implication for the...
clinical pharmacokinetics of lovastatin. *Evid Based Compl Altern Med.* (2012) 2012:1–10. doi: 10.1155/2012/127043

34. Nannoni G, Ali A, Di Pierro F. Development of a new highly standardized and granulated extract from Monascus purpureus with a high content of monacolin K and KA and free of inactive secondary monacolins and citrinin. *Nutrafoods.* (2015) 14:197–205. doi: 10.1007/s13749-015-0047-4

35. Perini M, Carbone G, Camin F. Stable isotope ratio analysis for authentication of red yeast rice. *Talanta.* (2017) 174:228–33. doi: 10.1016/j.talanta.2017.05.057

36. Xie X, Tang Y. Efficient synthesis of simvastatin by use of whole-cell biocatalysis. *Appl Environ Microbiol.* (2007) 73:2054–60. doi: 10.1128/AEM.02820-06

37. Xu BJ, Jia XQ, Gu LJ, Sung CK. Review on the qualitative and quantitative analysis of the mycotoxin citrinin. *Food Control.* (2006) 17:271–85. doi: 10.1016/j.foodcont.2004.10.012

38. Ali N, Degen GH. Citrinin biomarkers: a review of recent data and application to human exposure assessment. *Arch Toxicol.* (2019) 93:3057–66. doi: 10.1007/s00204-019-02570-y

39. Cramer B, Osteresch B, Muñoz KA, Hillmann H, Sibewski W, Humpf HU. Biomonitoring using dried blood spots: Detection of ochratoxin A and its degradation product 2’R-ochratoxin A in blood from coffee drinkers*. *Mol Nutr Food Res.* (2015) 59:1837–43. doi: 10.1002/mnfr.201500220

40. Marley E, Brown P, Leeman D, Donnelly C. Analysis of citrinin in cereals, red yeast rice dietary supplement, and animal feed by immunoaffinity column cleanup and lc with fluorescence detection. *J AOAC Int.* (2016) 99:1025–31. doi: 10.5740/jaocint.16-0060

41. EFSA. Scientific Opinion on the risks for public and animal health related to the presence of citrinin in food and feed. *EFSA J.* (2012) 10:2605. doi: 10.2903/j.efsa.2012.2605

42. Narváez A, Izzo L, Rodríguez-Carrasco Y, Ritieni A. Citrinin dietary exposure assessment approach through human biomonitoring high-resolution mass spectrometry-based data. *J Agric Food Chem.* (2021) 69:6330–8. doi: 10.1021/acs.jafc.1c01776

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