A unique mineralization mode of hypermineralized pleromin in the tooth plate of *Chimaera phantasma* contributes to its microhardness

Mayumi Iijima¹,³* & Mikio Ishiyama²,³

Tooth plates of the chimaeroids, holocephalian fishes, are unique dental hard tissues. Unlike the teeth of other animals, the tooth plates are located on the roof of the mouth and in the lower jaw. Their tooth plates consist, to a large extent, of lightly mineralized tissue (osteodentin) and hypermineralized tissue (pleromin). Notably, the mineral phase of pleromin is whitlockite, while that of other animals is apatite. Dietary habits of chimaeroids and wearing features of their tooth plates suggest an extreme hardness of pleromin, but this has never been investigated. We examined the microhardness of the tooth plate of *Chimaera phantasma* and found that pleromin in the biting region was extremely hard, comparable with the hardness of mature tooth enamel, whereas the hardness of immature pleromin was lower than that of bovine dentin. The hardness of osteodentin, on the other hand, was equivalent to that of bovine dentin and almost the same throughout the tooth plate. Immature pleromin was sparsely packed with oval crystals of whitlockite and, as pleromin matures, the space between crystals was filled with small intercrystalline materials. The maturing process of pleromin could partly contribute to its remarkable hardness and have some implications for designing novel biomaterials.

The chimaeroid fishes (Holocephali), which appeared about 420 Myr ago¹ in the Devonian period, belong to a subclass of cartilaginous fish (Chondrichthyes) and is a sister group of the other subclass, Elasmobranchii. The chimaeroid fishes are distinguished among the chondrichthians by the possession of a unique dental organ, i.e., three pairs of tooth plates, which are not shed or replaced, but evergrowing appositionally (statodont dentition) from caudal to rostral²-⁴. On the other hand, the dentition of elasmobranchs, as represented by shark, that comprises rows of multiple teeth are shed and replaced throughout life (lyodont dentition). Thus, the dentition of chimaeroids contrasts well with that of the Elasmobranchii. Although each of the six tooth plate of chimaeroid was reported to have developed from an individual tooth plate primordium⁵, embryological studies of chimaeroid tooth plates have shown an implication of a compound nature that represents the fusion of adjustment tooth-forming territories⁶. Furthermore, some early holocephalians, e.g., *Helodus*, was found to have a dentition consisting partially of tooth files, that represents a transitional form between tooth files and tooth plates⁶-⁸.

Tooth plates of chimaeroids are composed of three types of hard tissues: (1) a lightly mineralized tissue that occupies the bulk of the tooth plate called osteodentin⁹, which is also called trabecular dentin¹⁰, and sclerotic osteodentin¹¹; (2) a hypermineralized tissue, named pleromin¹² which has been also referred to as Kosmin¹³, tubular dentin¹⁴, orthotabeculine¹⁵, and whitlockin¹¹. The distribution and morphology of the hypermineralized tissue are different by taxa¹⁶,¹⁷; and (3) a thin veneer called vitrodentin¹³ or the outer dentin¹¹ that overlays the tooth plate and quickly wears out on exposed surfaces¹¹. Terms for these hard tissues are sometimes changed by various workers with new data¹⁸. In our study, we refer to the lightly mineralized bulky tissue as osteodentin, the hypermineralized pads and tritoral pad as vascular pleromin, and hypermineralized rods and ovoid as compact pleromin, following our previous papers²,¹⁹.

¹Department of Applied Biological Chemistry, Graduated School of Agricultural and Life Sciences, The University of Tokyo, 1-1-1 Yayoi, Bunkyo-ku, Tokyo 113-8657, Japan. ²Department of Histology, The Nippon Dental University School of Life Dentistry at Niigata, 1-8 Hamaura-cho, Chuo-ku, Niigata 951-8580, Japan. ³These authors contributed equally: Mayumi Iijima and Mikio Ishiyama. *email: mayumi01@mx5.canvas.ne.jp
Two types of pleromin develop embedded in osteodentin\textsuperscript{11,20}; the vascular pleromin is a single large pad that encloses vascular canals, while the compact pleromin is constituent of a series of small bead-like rods and lacks vascular canals. The differentiation of these pleromin is of developmental interest. Similar to tooth enamel, pleromin is a hypermineralized hard tissue. However, pleromin differs from enamel but is more like bone, because pleromin is formed by mesenchyme-derived cells, not by epithelial-derived cells\textsuperscript{2}. Furthermore, unlike enamel or enameloid that covers the surface of dentin, pleromin does not cover the entire biting area.

As Ishiyama et al.\textsuperscript{2} reported previously, the mineral phase of hypermineralized pleromin is whitlockite. Whitlockite is assigned the ideal formula \(\text{Ca}_9\text{Mg}(\text{HPO}_4)(\text{PO}_4)_6\)\textsuperscript{21}. The hard tissue composed of whitlockite is of primary importance in terms of biomimicry, because, to our knowledge, the mineral components of the hard tissue in all other vertebrates belong to apatite-group \(\text{Ca}_9(\text{PO}_4)_6(\text{OH},\text{F},\text{CO}_3)\), whose composition differs by species\textsuperscript{22}. Extant lungfish (\textit{Lepidosiren paradoxa} and \textit{Protopus} sp.) also have tooth plates with a constituent of hypermineralized petrodentin\textsuperscript{23–25}. However, the petrodentin in the tooth plates consists of hydroxyapatite\textsuperscript{26}. Whitlockite is known as the major constituent of the human dental calculus and is involved in the pathological calcification\textsuperscript{27,28}, but it is not a constituent of normal dental hard tissues\textsuperscript{29}. Thus, pleromin is currently the only example of whitlockitic hard tissue that forms in physiological conditions. Because of this rare mineral phase of pleromin, a recent study proposed to refer to this hypermineralized tissue as “whitlockin”\textsuperscript{30}.

Whitlockite gives an X-ray diffraction (XRD) profile similar to that of \(\beta\text{-Ca}_3(\text{PO}_4)_2\) (\(\beta\text{-TCP}\)) and Mg-contain- ing \(\beta\text{-TCP} (\beta\text{-TCa,MgP})\). Since both whitlockite and \(\beta\text{-TCa,MgP}\) contain Mg and exhibit XRD profiles similar to that of \(\beta\text{-TCP}\), it is difficult to distinguish them based on their XRD profiles and Mg content\textsuperscript{29}. Thus, a little ambiguity regarding the mineral phase of pleromin remained needs to be solved. Since \(\text{HPO}_4\) is a key to assign- ing \(\text{KH}_3 \text{PO}_4\) (IP-3, 4 in Fig. 4a) was higher than that of the PP (PP-1, 2 in Fig. 4a); in the immature VP, the KH of the PP was 0.1–0.5 GPa, whereas the KH of the IP was 0.86–1.1 GPa. The KH increased gradually as the distance from VC appeared at the central region of IP, regardless of its maturity. The KH of immature IP (0.86–1.1 GPa, IP-3, 4 in Fig. 4a) was 1.7 times increase.

The wear resistance and dietary habit of the chimaeroids, which involves crushing of shellfish\textsuperscript{4,11}, implies that the hypermineralized pleromin is extremely hard. To date, however, there has been no study to determine mechanical properties of tooth plates. Here, we report for the first time the hardness of tooth plates in \textit{Chimaera phantasmas} (Holocephali) and discuss the hardness in relation to its microstructure.

**Results**

**General morphology of the palatine tooth plate.** A contact microradiogram (CMR) (Fig. 2a) and optical microscopic images (Fig. 2b) of a longitudinal section cut along the white line illustrated in Fig. 1 show the distribution of hypermineralized pleromin in the palatine tooth plate. The CMR (Fig. 2a) shows the pleromin as a radiopaque white region and less mineralized osteodentin (OD) (Fig. 2a,b) as radiolucent gray and black regions. The two types of pleromin, vascular pleromin (VP) and compact pleromin (CP), exhibiting different morphologies, are shown in Fig. 2a and b. VP is a large pad of pleromin that contains vascular canals (VC) and occupies an occlusal part of the tooth plate. On the other hand, CP lacks VC and forms rows in the interior region that was named “pearlstrings”\textsuperscript{13}. In the posterior region, both VP and CP are radiolucent (Fig. 2a) and opaque (Fig. 2b,b1), indicating that these tissues are immature. As their maturity increases toward the anterior region, pleromin becomes radiopaque (Fig. 2a) and transparent (Fig. 2b,b2). In the biting area (arrows in Fig. 2b), VP is exposed on the occlusal surface by wear. The interior structure of immature and mature VP is shown by Fig. 2b1 and b2, respectively. The enlarged images of VP show that pleromin formed around VC (peritubular pleromin (IP)) is relatively opaque, while pleromin surrounding PP (interglobular pleromin (IP)) is relatively transparent. In contrast, CP is uniformly and densely packed with mineral components (Fig. 2b3).

Figures 3a and b show the distribution maps of Ca, P, and Mg in the immature and the mature VP, respectively. In the immature VP (Fig. 3a), it is evident that Ca and P localize more in IP than in PP. Whereas in the mature VP, the difference between PP and IP is unclear (Fig. 3b), presumably because of an increase in the calcification level in mature VP (Table 1). In the CP, these ions distributed evenly (Supplementary Fig. S2 online). An energy dispersive X-ray spectrometer (EDS) analysis showed that the Mg content of immature pleromin and mature pleromin was higher than that of osteodentin (Table 1). In VP, the content of Ca, P, and Mg increases as this tissue matures; however, the ratio of Mg substitution for Ca, represented by the Mg/(Mg + Ca) atomic ratio of VP, was approximately 0.044, which indicates that 4.4% of Mg substitutes for Ca in VP, regardless of its maturity. On the other hand, about 1.5% of Mg substitutes for Ca in osteodentin.

**Microhardness of the tooth plate.** Microhardness of vascular pleromin. The Knoop hardness (KH) of VP in immature and mature regions are shown in Fig. 4a. In both immature and mature VPs, the hardness of IP (IP-3, 4 in Fig. 4a) was higher than that of the PP (PP-1, 2 in Fig. 4a); in the immature VP, the KH of the PP was 0.1–0.5 GPa, whereas the KH of the IP was 0.86–1.1 GPa. The KH increased gradually as the distance from VC increased (Fig. 4a). In the mature VP, the KH of the PP was 0.3–1 GPa (PP-1,2 in Fig. 4a) and that of the IP was 3–5.9 GPa (PP-3,4 in Fig. 4a). The KH increased drastically as the distance from VC increased. The increase of the KH from PP (1 GPa) to IP (3 GPa) in the mature VP was three times higher than that in immature VP (from 0.5 to 0.86 GPa, 1.7 times increase).

In the PP, the hardness increased from the junction of PP-IP toward the center of IP. The maximum KH appeared at the central region of IP, regardless of its maturity. The KH of immature IP (0.86–1.1 GPa, IP-3, 4 in Fig. 4a, Supplementary Fig. S1 online) was as low as that of bovine dentin (0.88 ± 0.01 GPa, Supplementary Table S1 online). In contrast, the KH of central region of IP in the biting region was about 5.86 ± 0.13 GPa (IP-4 of mature pleromin in Fig. 4a, about 600 µm below the surface, Supplementary Fig. S1 online). In two other central regions of IP, the KH was 5.96 ± 0.27 GPa and 5.97 ± 0.13 GPa (both of them located at 300 µm below the surface). These three values show no statistically significant difference (P = 0.900, ANOVA/Tukey) and are
larger than the KH of the outer enamel (about 200 µm below the surface) of mature bovine tooth, 4.53 ± 0.14 GPa (Supplementary Table S1 online, Fig. 4a).

Microhardness of compact pleromin. The KH of three CPs at different maturity levels (A–C in Supplementary Fig. S1 online) are shown in Fig. 4b. In each CP, the hardness increased from outer regions (1 in Fig. 4b) toward the center (4 in Fig. 4b). The central region was the hardest, regardless of the maturity. In the most mature CP-C, the hardness increased from 2.9 GPa in the outermost region (C-1 in Fig. 4b) to 4.9 GPa in the central region (C-4 in Fig. 4b) (1.7 times increase). In CP-B, the hardness was 0.9 GPa in the outermost region (B-1 in Fig. 4b), while 3.7 GPa in the central region (B-4 in Fig. 4b) (4 times increase). In the most immature CP-A, a large increase was observed, from 39–59 MPa in the outermost region (A-1 in Fig. 4b) to 784 MPa in the central region (A-4 in Fig. 4b; 16 times increase). The CPs that were immature than CP-A were too soft to indent.

Microhardness of osteodentin. The KH of OD, 618 ± 43 MPa, that is the average of six different points from posterior to anterior (for indented positions, see Supplementary Fig. S1 online), is indicated in Fig. 4a. The KH of OD was lowest in the most posterior region (point No. 2 in Supplementary Fig. S1 online), 538 ± 5 MPa, while that in the biting region (point No. 6 in Supplementary Fig. S1 online) was relatively high, 659 ± 4 MPa. In the outer OD of the tooth plate covering the CPs (point No. 1 in Supplementary Fig. S1 online), the KH was 624 ± 105 MPa. The KH at point No. 2–No. 6 did not show statistical difference (P > 0.6, ANOVA/Tukey). The average KH of OD was smaller than that of bovine dentin (880 ± 10 MPa) (Fig. 4a).

Mineral characterization. The SEM images of VP at different maturity levels are shown in Fig. 5. Three observed positions a, b, and c are indicated in Supplementary Fig. S1 online. Immature pleromin is roughly packed with oval-shaped crystals approximately 300–400 nm in longitudinal direction (Fig. 5a). These crystals are located around fibers and arranged randomly. As pleromin matures, the space between the crystals is filled with small intercrystalline materials (Fig. 5b), and both fibers and oval-shaped crystals become unrecognizable.
(Fig. 5c). As a result, pleromin grows into a dense tissue. The mineralization process of pleromin undergoes (1) a growth of whitlockite into oval-shaped crystals and (2) a subsequent deposition of intercrystalline materials between the whitlockite crystals.

The powder XRD profile of pleromin is shown in Fig. 6a1, and the peaks for standard β-TCP (JCPDS 09-0169) are indicated in Fig. 6a2. The diffraction peaks of pleromin are extremely sharp, and its profile is similar to the profile of β-TCP. The diffraction angles (2θ) are, however, slightly higher than those of β-TCP. The refined lattice dimensions are $a = 10.408(1)$ Å and $c = 37.244(4)$ Å. These are smaller than the values reported for β-TCP ($a = 10.439(1)$ Å and $c = 37.375(6)$ Å)\(^\text{30}\). The smaller lattice dimension is consistent with the peak shift to a higher angle.

Figure 6b and c show the micro-beam X-ray photographs of immature and mature pleromin, and Fig. 6e shows their densitometric traces. The XRD peak positions of immature pleromin and those of mature pleromin
are equivalent, which indicates that the mineral phase of the immature pleromin is the same as that of mature pleromin. Debye–Scherrer rings in the micro-beam X-ray photographs indicate that the crystallites in pleromin have no preferred orientation, which is consistent with our SEM observation of pleromin (Fig. 5). In mature pleromin, four unidentifiable peaks at 12.5°, 15.3°, 23.5°, and 25.3° in 2θ (* in Fig. 6e) were observed, while none of these peaks were detected in immature pleromin. On the other hand, our micro-beam X-ray photograph of osteodentin (Fig. 6d) revealed that the mineral component of osteodentin was apatite.

Figure 7 shows the microscopic ATR FT-IR spectra of pleromin at different maturation levels and osteodentin. Immature pleromin-A is located in the most posterior region, and immature pleromin-B is located between immature pleromin-A and mature pleromin. In immature pleromin-B and in the mature pleromin, bands derived from phosphatic components are prominent. The major absorption bands in the range of 800–1200 cm⁻¹ for both immature pleromin-B and mature pleromin overlap with bands derived from whitlockite (red arrows in Fig. 7). The band at 862 cm⁻¹ is assigned to HPO₄ stretching and the bands at 990 and 1135 cm⁻¹ are assigned to υ₃PO₄ (Fig. 7). It is notable that this band that can be ascribed to whitlockite is also observed in immature pleromin-A, because this suggests that the mineral phase in the early stage is whitlockite. In the spectrum of immature pleromin-A, amide I (1655 cm⁻¹) and amide II (1535 cm⁻¹) are prominent (blue arrows in Fig. 7). We ascribe other bands in the range of 1100–1800 cm⁻¹ to the resin (* in Fig. 7, Supplementary Fig. S3 online).

Table 1. Elemental analysis of vascular pleromin (VP) and osteodentin. Atomic % of Ca, P, and Mg obtained by EDS analysis and rate of Mg substitution for Ca, Mg/(Mg + Ca) are indicated. The maturity of pleromin increases in the order of immature VP-A < immature VP-B < mature VP. Numbers in parenthesis show 2σ of each data.

| Position          | Ca/atm%  | P/atm%  | Mg/atm% | Mg/(Mg + Ca) |
|-------------------|----------|---------|---------|--------------|
| Immature VP-A     | 4.00 (4) | 4.00 (3)| 0.18 (1)| 0.043        |
| Immature VP-B     | 8.32 (5) | 7.75 (4)| 0.39 (1)| 0.045        |
| Mature VP         | 24.6 (1) | 22.8 (1)| 1.17 (2)| 0.045        |
| Osteodentin       | 3.93 (4) | 3.59 (3)| 0.06 (1)| 0.015        |

Figure 3. Elemental analysis of vascular pleromin (VP). Distribution maps of Ca, P, and Mg in (a) immature VP and (b) mature VP. (a) and (b) correspond, respectively, to areas (b1) and (b2) in Fig. 2.
The spectrum of osteodentin is similar to that of apatite, i.e., that of bovine dentin (Supplementary Fig. S4 online). The absorption bands specific to PO$_4$ of apatite are indicated in Fig. 7. The bands at 875, 1418, and 1456 cm$^{-1}$ are ascribed to CO$_3^{2-}$ ions in apatite lattice$^{34}$. These bands are also observed in the ATR FT-IR spectra of bovine tooth enamel (Supplementary Fig. S4 online). Thus, the mineral phase of osteodentin was identified as CO$_3$-containing apatite. The bands of amide I (1535 cm$^{-1}$) and amide II (1655 cm$^{-1}$) are also observed in the spectrum of osteodentin (blue arrows in Fig. 7).

**Discussion**

Our measurement of the microhardness of the tooth plate of *Chimaera phantasma* revealed a complex hardness distribution in the tooth plate. Although overall hardness of pleromin in the immature region was lower than or equivalent to that of osteodentin, the hardness increased from posterior to anterior regions. It is notable that
pleromin in the functioning areas is the hardest of all, approximately 6 GPa. The KH of pleromin was larger than that of the outer enamel (about 200 µm below the surface) of the mature bovine tooth measured in this study (Fig. 4a) and that of the tooth enamel studied previously (2.6–4.3 GPa)35–38. In contrast, the overall hardness of osteodentin was approximately 600 MPa. The hardness of pleromin and osteodentin well explains the observation that pleromin and osteodentin show different wear rates, and pleromin becomes exposed on the biting surface upon wear4,11. The KH of VP in a posterior immature region was slightly lower than that of osteodentin, probably due to the sparse packing of crystals in this region (Fig. 5a). In the case of CP, immature four CPs younger than CP-A (Supplementary Fig. S1, online) were too soft to indent. This could be ascribed to the empty structure of the earliest CP11 (ovoid in11). In CP, the KH increased from margin to the center regardless of the maturity of CP (Fig. 4b). This hardness change is consistent with the mineralization process observed by Smith et al.11. That is, as the mineralization proceeds, the empty center was filled with granules, indicating that mineralization began in the center of CP11. The central part of immature CP contained crystal bundles and was dense, while mineralized periphery of CP was composed with network of tubules, which contained small mineral granules within11. This very early stage mineralization pattern could be taken over in the later stage, because the distribution pattern of the microhardness in more matured CP was similar to that of immature CP (Fig. 4b).

Nanoindentation study of tooth enamel39 showed that the hardest region (~ 6 GPa) is located within 100 µm of the occlusal surface, and the hardness decreases in regions deeper from the surface and closer to

![Figure 6. XRD profiles of pleromin and osteodentin. (a1) Powder XRD profile of pleromin. (a2) XRD peaks for β-TCP (JCPDS 09-0169). Micro-beam X-ray photographs of (b) immature pleromin, (c) mature pleromin, and (d) osteodentin. Note that the Debye–Scherrer rings indicate that crystallites in pleromin (b, c) and in osteodentin (d) have no preferred orientation. (e) Densitometric traces of (b–d). *Indicates peak other than β-TCP.](https://www.nature.com/scientificreports/)
the enamel-dentin junction. This tendency was also observed in bovine tooth enamel examined in this study (Fig. 4a and Supplementary Table S2 online). It is reported that the hardness of tooth enamel shows only a weak correlation with the degree of the prism alignment, but a strong correlation with the chemical composition\(^3\). In sea urchin, the microhardness of teeth increases from peripheral regions to the center, which is correlated with an increase in the Mg content of calcite\(^4\). The maximum hardness of the working zone was reported to be 3.54 GPa, which is much harder than the hardness of other calcitic materials (1.5 GPa for pure calcite)\(^5\). In the tooth plate of *Chimaera phantasma*, hypermineralized pleromin is composed of Mg-containing whitlockite and pleromin drastically increases in KH with maturation (Fig. 4), as the content of Ca, P, and Mg increases. In fact, the Mg content of mature pleromin was significantly higher than that of the immature pleromin. However, the Mg/(Mg + Ca) ratio remains the same in immature pleromin and mature pleromin (Table 1), indicating that the Mg content of whitlockite in pleromin was largely unchanged during the maturation process of pleromin. This is also supported by our XRD analysis, which shows that the individual peak position of the immature and mature pleromin was unchanged (Fig. 6). Similar relationship between the Mg content and hardness of the hard tissue can be seen in the immature and mature tooth enamel.

The chemical formula of porcine tooth enamel was determined using immature and erupted tooth\(^2\). That is, Ca\(_{4.155}\)Mg\(_{0.034}\)Na\(_{0.191}\)K\(_{0.004}\)(HPO\(_4\))\(_{0.35}\)(CO\(_3\))\(_{0.357}\)(PO\(_4\))\(_{2.384}\)(OH)\(_{0.007}\) for immature enamel and Ca\(_{4.568}\)Mg\(_{0.032}\)Na\(_{0.11}\)K\(_{0.002}\)(HPO\(_4\))\(_{0.143}\)(CO\(_3\))\(_{0.256}\)(PO\(_4\))\(_{2.712}\)(OH)\(_{0.378}\) for erupted one. According to the chemical formula, 0.8% and 0.7% Mg substitute for Ca in immature and erupted enamel, respectively. These values are much lower than the acceptable quantity of Mg in synthesized Mg-substituted hydroxyapatite (2.1 wt%)\(^6\). It should be noted that although the erupted tooth enamel is much harder than immature enamel, Mg content was almost the same between them. Therefore, hardening of these hypermineralized tissues can be attributed not principally to Mg, but to some other factors.

In the early stage of pleromin formation, initial crystals deposited in tubular saccules, which is the major constituents of the immature pleromin matrix\(^1\). The mineral deposition in tubular matrix is also detected by optic observation\(^1\). These deposits grew into oval-shaped crystals (Fig. 5a). In this stage, the KH of pleromin was much lower than that of osteodentin and bovine dentin (Fig. 4a). In the later maturation stage of pleromin,
the space between the oval-shaped crystals was filled with small intercrystalline materials (Fig. 5b,c), majority of which are identical with the oval crystals in terms of mineral phase (Fig. 6) and composition (Table 1). It should be noted that in the very early mineralization stage of CP (ovoid in Fig. 5), existence of a labile non-crystalline mineral of high Mg content was observed using different optical methods13. Furthermore, the non-crystalline mineral transformed rapidly into hypermineralized pleromin11 (HD in Fig. 5). In the process of the mineralization, intercrystalline space was filled with irregular shaped mineral deposits (Fig. 5b,c). If the formation of whitlockite starts with Mg-containing amorphous calcium phosphate (Mg-ACP), these deposits could be Mg-ACP, the intermediate transition material from Mg-ACP to whitlockite, and/or whitlockite. It is plausible that unidentified XRD peaks in Fig. 6e were caused by such an intermediate phase. In any case, regardless of the type of mineral phase of the secondary precipitate, an increment of the pleromin density by the secondary precipitation of the intercrystalline materials is a most plausible factor to harden pleromin, rather than the presence of Mg in the mineral. Generally, the space-filling method is effective to make the powdery substance dense. Pleromin could acquire a remarkable hardness through this mineralization mode. This process is distinct from the formation of tooth enamel. In the maturation stage of mammalian tooth enamel formation, the space between the thin ribbon-like immature crystals is filled by the lateral growth of the mother crystals. Enamel crystals thus increase the width and thickness until the crystals are finally in contact with each other14.

Extant lungfish also use tooth plates to crush hard prey. The crushing action is reported to occur between powerful jaws, equipped with palatal and lingual tooth plates, and has been observed in the three genera16. The tooth plates are composed of osteodentin and petrodentin24,25. Petrodentin is formed as a special and unique hypermineralized portion11,45. The crushing action implies that the petrodentin is very hard. This has been proved by the hardness measurement. The Vickers hardness of osteodentin is reported to be ~ 430 MPa and that of petrodentin is ~ 3.12 GPa in the plan along the long axis and ~ 2.48 GPa in the plan transverse to the long axis of apatite crystals26. Thus, the hardness of the hypermineralized petrodentin was almost comparable to the reported hardness of tooth enamel and shown to be sufficiently high to take the crushing action. In the case of the pleromin of Chimaera phantasma, it was not possible to measure the microhardness perpendicular to the biting surface due to its surface condition (see Materials and Methods). However, in contrast to the anisotropic petrodentin, which is composed of the needle shaped hydroxyapatite crystals26, pleromin is supposed to be more isotropic than petrodentin: Pleromin is composed of oval-shaped and irregular shaped fine whitlockite crystals without preferred orientation (Fig. 5). Micro-beam XRD profiles (Fig. 6e) indicate that there is no preferred orientation in pleromin constituting crystals. Therefore, mechanical property is supposed to be relatively isotropic. Although chimaera and lungfish belong to different class and the mineral phase of the hypermineralized tissue in the tooth plate is different, both tooth plates have a common nature; that is, a hard and hypermineralized tissue develops in a soft and less mineralized matrix. There might be a common mechanism to form hypermineralized tissue in the osteodentin matrix between them.

The uniqueness of pleromin in Chimaera phantasma may be highlighted by comparison with the dental tissues of other fishes. Sharks (Elasmobranchii) also belong to cartilaginous fish (Chondrichthyes) and are the closest relative of tissues of other fishes. Sharks (Elasmobranchii) also belong to cartilaginous fish (Chondrichthyes) and are the closest relative of tissues of other fishes. Nevertheless, the dental tissues of sharks differ considerably from those of Chimaera. Notably, sharks have a series of teeth on the jaws, and the hypermineralized part of teeth (enameloid) entirely covers dentin. Furthermore, the mineral phase of shark enameloid is fluorapatite. Unlike sharks, lungfish have tooth plates, but the hypermineralized tissue in tooth plates (petrodentin) is composed of hydroxyapatite26. Thus, chimaeroid pleromin is distinct among biominerals in vertebrates, which are composed of calcium phosphates. Our findings of the relationship between the mineralization mode and the hardness would help creating novel biomaterials.

Conclusion
The hypermineralized portion of the tooth plate of Chimaera phantasma is composed of whitlockite that shows a remarkable hardness, comparable to the hardness of tooth enamel. The hardness of immature pleromin increases strikingly, as it grows toward its functioning region, and as it undergoes hypermineralization. The mineral phase and the rate of Mg substitution for Ca of the mature pleromin are equivarent to those of the immature pleromin. We hypothesize that a striking increase in hardness could be correlated with the mineralization process of pleromin, which involves (1) the growth of whitlockite into oval-shaped crystals, and (2) the subsequent deposition of intercrystalline materials between the whitlockite crystals. A drastic increment of the hardness of pleromin in the biting region of the tooth plate should be the key for chimaeroids to preying upon hard-bodied diet.

Methods
Sample preparation. Tooth plates of Chimaera phantasma (adult, length ~ 80 cm, Shimoda, Japan) (Fig. 1a) was used. Palatine tooth plates (arrowed in Fig. 1b) was removed from the upper jaw and subsequently soaked in 10% formalin with neutral pH for 48 h, dehydrated by immersing the tooth plate in 10–100% ethanol progressively for 60 min in each concentration, and embedded in polyester resin (Rigolac). The resin block was cut longitudinally along the line in Fig. 1b into two half with a diamond blade (Buehler Ltd.) at a low speed. One of the halves of the resin block was used for the measurement of microhardness. The cut surface was polished with waterproof SiC-abrasive papers (#1000–#2000) and Al2O3-polishing papers with Al2O3 particle size of 3 μm (#4000), 1 μm (#8000), and finally 0.3 μm (#15000). The polished block was cleaned using an ultrasonic cleaning apparatus in distilled water for 3 s.

The other half of the resin block was cut into longitudinal cross sections (thickness of ~ 200 μm), using a saw microtome (Leitz-1600) and used for characterization of the mineral phase of pleromin and osteodentin. Since the biting surface of the tooth plates is uneven and have lots of concavities, it was not possible to make a mirror
polishing surface for hardness measurement. Therefore, hardness measurement was conducted normal to the biting direction using the longitudinally cut surface.

**Observation of general morphology of the tooth plate.** To determine the position to analyze, the morphology and distribution of hypermineralized pleromin in the longitudinal cross section was examined using an optical microscope and a contact microradiography (CMR) (Sofron, SRO-M50, 4 kV, 4 mA). CMR was performed using a polished section with a thickness about 70 µm.

**Measurement of microhardness.** The Knoop hardness (KH) was measured for (1) immature vascular pleromin (VP) in posterior regions and mature VP in anterior regions (b1 and b2 in Supplementary Fig. S1 online), (2) three compact pleromin (CP) with different maturity levels (A, B, and C in Supplementary Fig. S1 online), and (3) six points in osteodentin (OD) (1–6 in Supplementary Fig. S1 online), using a Knoop microindenter (Shimadzu, Microindentor HMV-2000). Indentation of VP was conducted in arrays directed from an inner part of the PP towards the center of the IP (Supplementary Fig. S1 online). Three indented CPs are indicated in Supplementary Fig. S1 online as A, B, and C. Maturity of the CPs increases in the order of A, B, and C. In each CP, four points from peripheral regions to the center were indented (Supplementary Fig. S1 online). CPs that are more immature than A, were too soft and could not be indented. Indentation of the OD was performed at six different points in the regions beginning from a posterior region to more anterior regions (Supplementary Fig. S1 online).

Mature VP and CP were indented with a load 200 or 100 gf for 30 s. A small load was applied to soft regions; that is, immature VP and CP in posterior regions and osteodentin, were loaded at 50 or 10 gf for 30 s. Immediately after indentation, length of the long indentation diagonal was measured three times. Knoop hardness number (KHN) (kgf/mm²) was obtained for each scale reading. Then, mean KHN and its standard deviation (SD) (N = 3) were calculated for one indentation point. To represent Knoop hardness (KH), KHN was converted into GPa. For comparison, the hardness of bovine molar tooth, which was embedded and prepared equally, was measured. For enamel, outer surface, middle regions, and inner regions near the enamel-dentin junction were indented. For dentin, the central region between the enamel-dentin junction and dental pulp was indented. The KH of enamel and dentin of a bovine tooth were obtained by measuring the KHN of three closely spaced positions. The Knoop microindentor was adjusted using a standard stainless steel block with hardness of 9.4–10.2 GPa before measurement of the sample. The standard stainless steel block gave KH of 10.20 ± 0.02 GPa. All the measurements were performed at room temperature.

Multiple comparison tests of the mean hardness of various points were performed using KaleidaGraph (ver. 4.0). The data were statistically analyzed by a one-way classification analysis of variance (ANOVA) with the Tukey posterior multiple comparison.

**Characterization of the mineral phase.** To identify the mineral phase in a specific position of pleromin and osteodentin, a micro-Laue camera (Rigaku) was used. Longitudinal cross sections of a tooth plate (Fig. 2b) were fixed onto a sample-holder, and a selected area of immature and mature vascular pleromin (a, c in Supplementary Fig. S1 online) and a central part of osteodentin (2 in Supplementary Fig. S1 online) were irradiated with a focused X-ray beam using a collimator of 100 µm diameter. A micro-Laue photograph of each position was taken with an exposure time of 20 h (56 kV, 200 mA, CuKα). The lattice dimension of whitlockite was refined by means of a powder X-ray diffractometer (Rigaku, Rint2500, 56 kV, 200 mA, CuKα) using 2θ peaks. Diffraction angle (2θ) of each peak was corrected using Si powders.

To analyze a specific position of pleromin and osteodentin, microscopic Fourier Transformed Infra-red (FT-IR) analysis was performed using the attenuated total reflection (ATR) method (Shimadzu, FT-IR 8800). For measurement, a longitudinally sectioned tooth plate (Fig. 2b) was used. Three positions of VP with different maturity levels (a, b, c in Supplementary Fig. S1 online) and a central portion of osteodentin (3 in Supplementary Fig. S1 online) were examined. Resin that surround sample was also measured (Supplementary Fig. S3 online). Each point was lightly pressed with a Ge-prism and an IR spectrum from a small area (100 × 100 µm²) was obtained. For comparison, the spectra of bovine tooth enamel and dentin were obtained using a thin section of the tooth and the same ATR FT-IR instrument (Supplementary Fig. S4 online).

After a longitudinal cross section was coated with a thin layer of carbon using a carbon evaporator, the tooth plate was observed by a scanning electron microscope (SEM) (HITACHI, S4500) and atomic analysis of the same area was conducted by an energy dispersive X-ray spectrometer (EDS) (HORIBA, EMAX-7000), equipped with SEM. The content and distribution of calcium (Ca), phosphorus (P), and magnesium (Mg) of these areas were analysed under a 20 kV of operating voltage, 0.44 nA of probe current, and acquisition time of 3000 s. Three positions of vascular pleromin (VP) at different maturity levels (a–c in Supplementary Fig. S1 online) were examined.

**Ethical approval.** All methods were carried out in accordance with relevant guidelines and regulations. All experimental protocols were approved by a named institutional and/or licensing committee. This study was approved by Animal Experiment Committee of the Nippon Dental University School of Life Dentistry at Niigata.

Received: 15 June 2020; Accepted: 14 October 2020
Published online: 29 October 2020
References

1. Inoue, J. G. et al. Evolutionary origin and phylogeny of the modern holoccephalans (Chondrichthyes: Chimaeriformes): mitogenomic perspective. *Mol. Biol. Evol.* **27**, 2576–2586 (2010).

2. Ishiyama, M., Sasagawa, I. & Akai, J. The inorganic content of pleromin in tooth plates of the living Holoccephalan consists of a crystalline calcium phosphate known as β-Ca3(PO4)2 (whitlockite). *Arch. Hist. Jap.* **47**, 89–94 (1984).

3. Patterson, C. Interpretation of the toothplates of chimaeroid fishes. *Zool. J. Linn. Soc.* **106**, 33–61 (1992).

4. Didier, D. A., Stahl, B. J. & Zangerl, R. Development and Growth of compound tooth plates in *Callorhinus milii* (Chondrichthyes, Holoccephali). *J. Morphol.* **222**, 73–83 (1994).

5. Kemp, A. A comparison of the developing dentition of *Neoceratodus forsteri* and *Callorhinus milii*. *Proc. Linn. Soc. NSW* **107**, 245–262 (1984).

6. Patterson, C. The phylogeny of the chimaeroids. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **249**, 101–219 (1965).

7. Finarelli, J. A. & Coates, M. I. First tooth-set outside the jaws in a vertebrate. *Proc. R. Soc. B* **279**, 775–779 (2011).

8. Itano, W. M. & Lambert, L. L. A new cochliodont anterior tooth plate from the Mississippian of Alabama (USA) having implications for the origin of tooth plates from tooth files. *Zool. Lett.* **4**, 12–21 (2018).

9. Örvig, T. Phylogeny of tooth tissue: evolution of some calcified tissues in early vertebrates. In *Structural and Chemical Organization of Teeth* 1st edn (eds Miles, A. E. W.) 45–110 (Academic Press, London, 1967).

10. Peyer, B. Comparative Odontology (University of Chicago Press, Chicago, 1968).

11. Smith, M. M., Underwood, C., Goral, T., Healy, G. & Johanson, Z. Growth and mineralogy in dental plates of the holoccephalan *Harriotta raleighana* (Chondrichthyes): novel dentin and conserved patterning combine to create a unique chondrichthyan dentin. *Zool. Lett.* https://doi.org/10.1186/s40123-019-0125-z (2019).

12. Örvig, T. Palaeohistological notes3 The interpretation of pleromin (pleromic hard tissue) in the dermal skeleton of psammosteid heterostracans. *Zool. Scr.* **5**, 35–47 (1976).

13. Bargmann, W. Die Zahnplatten von *Chimaera monstrosa*. *Z. Zellforsch Mikrosch.* **19**, 37–561 (1933).

14. Mov-Thomas, J. A. The early evolution and relationships of elasmobranchs. *Biol. Rev.* **14**, 1–26 (1939).

15. Zangerl, R., Winter, H. F. & Hansen, M. C. Comparative microscopic dental anatomy in the Petalodontida (Chondrichthyes, Elasmobranchii). *Fieldiana Geol. Ser.* **26**, 1–43 (1993).

16. Didier, D. A. Phylogenetic systematics of extant chimaeroid fishes (Holoccephali, Chimaeroidae). *Am. Mus. Novit.* **3111**, 1–86 (1995).

17. Ishiyama, M. & Teraki, Y. The fine structure and formation of hypermineralized petrodentine in tooth plate of extant lungfish *Neoceratodus forsteri* (Holocephali). *J. Morphol.* **2052**, 2052–2059 (2003).

18. Smith, M. M. Petrodentine in extant and fossil dipnoan dentitions: microstructure, histogenesis and growth. *Calcif. Tissue Int.* **54**, 356–364 (1990).

19. Ishiyama, M., Yoshie, S., Teraki, Y. & Cooper, E. W. T. Ultrastructure of pleromin, a highly mineralized tissue comprising crystalline calcium phosphate known as whitlockite. In *Mechanisms and Phylogeny of Mineralization in Biological Systems* (eds Suga, S. & Nakahara, H.) 453–457 (Springer, New York, 1991).

20. Örvig, T. Histologic studies of ostracoderms, placoderms and fossil elasmobranchs. 4. Ptyctodontid tooth plates and their bearing on holoccephalan ancestry: the condition of *Ctenurus* and *Ptyctodus*. *Zool. Scr.* **9**, 219–259 (1980).

21. Keppler, U. Zum whitlockite-problem. *Neues Jahrb. Miner. Monatsh.* **4**, 12–21 (1974).

22. Lowenstam, H. A. & Weiner, S. *On Bionitralization* 8–15 (Oxford Univ. Press, Oxford, 1989).

23. Lison, L. The structure of teeth of lungfish. *C. R. Seances Soc. Biol. Filiales.* **135**, 431–433 (1941).

24. Smith, M. M. Petrodentine in extant and fossil dipnoan dentitions: microstructure, histogenesis and growth. *Proc. Linn. Soc. NSW.* **20**, 367–407 (1984).

25. Reisz, R. R. & Smith, M. M. Lungfish dental pattern conserved for 360 Myr. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **362**, 121–139 (2017).

26. Ishiyama, M. & Teraki, Y. The fine structure and formation of hypermineralized petrodentine in tooth plate of extant lungfish (Lepidosiren paradoxa and *Protopterus* sp.). *Acta Odont. Scand.* **121–139 (1957).

27. Chevron, Y., Daculsi, G., Kiejkowska, R. & Kerebel, B. The effect of magnesium on the formation of apatites and whitlockites. In *Magnesium in Health and Disease* (eds Itokawa, Y. & Durlach, J.) 11–19 (John Libbey & Co., Ltd., New York, 1989).

28. Tovborg, A. J. & Rowles, S. L. Magnesium whitlockite, a major constituent of human dental calculus. *Acta Odont. Scand.* **15**, 121–139 (1957).

29. Elliott, J. C. The crystallographic structure of dental enamel and related apatites. *Ph.D Thesis. (University of London, 1964).*

30. Currey, J. D. & Abeysekera, R. M. The microhardness and fracture surface of the petrodentine of Lepidosiren (Dipnoi), and of other mineralized tissues. *Archs. Oral Biol.* **48**, 439–444 (2003).

31. Fong, H. et al. Enamel structure properties controlled by engineering proteins in transgenic mice. *J. Bone Miner. Res.* **18**(11), 2052–2059 (2003).

32. Atkin, T. et al. Correlation of microhardness and wear in differently eroded bovine dental enamel. *Archs. Oral Biol.* **42**(5), 243–250 (1997).

33. Meredith, N., Sherriff, M., Setchell, D. J. & Swanson, S. A. V. Measurement of the microhardness and Young's modulus of human dental enamel and dentine using an indentation technique. *Archs. Oral Biol.* **41**(6), 539–545 (1996).

34. Curr, J. et al. Nanoindentation mapping of the mechanical properties of human molar tooth enamel. *Archs. Oral Biol.* **47**, 281–291 (2002).

35. Wang, R., Addadi, L. & Weiner, S. Design strategies of sea urchin teeth: structure, composition and micromechanical relations to function. *Phil. Trans. R. Soc. Lond.* **352**, 469–480 (1997).

36. Wang, R. Fracture toughness and interfacial design of a biological fiber-matrix ceramic composition in sea urchin teeth. *J. Am. Ceram. Soc.* **81**, 1037–1040 (1998).

37. Aoba, T. & Moreno, E. C. Changes in the nature and composition of enamel mineral during porcine amelogenesis. *Calcif. Tiss. Int.* **47**, 356–364 (1990).

38. Stippiece, L. et al. Characterization of Mg-substituted hydroxyapatite synthesized by wet chemical method. *Ceram. Int.* **40**, 3261–3276 (2014).

39. Stivel, K. A. & Halse, A. Crystal growth in rat incisor enamel. *Anat. Rec.* **173**, 453–468 (1972).

40. Smith, M. M. & Johanson, Z. The dipnoan dentition: A unique adaptation with a longstanding evolutionary record. In *The biology of Lungfish* (eds Jorgensen, J. M. & Joss, J.) 219–242 (CRC Press, Taylor and Francis, Cambridge, 2011).
Acknowledgements
The authors are grateful to Shimadzu Corporation for performing the micro ATR FT-IR spectroscopy. We would thank to Dr. Kazuhiko Kawasaki (The Pennsylvania State University) for providing constructive comments.

Author contributions
M.I. and M.I. coordinated and conducted this study and equally contributed.

Competing interests
The authors declare no competing interest.

Additional information
Supplementary information is available for this paper at https://doi.org/10.1038/s41598-020-75545-0.

Correspondence and requests for materials should be addressed to M.I.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2020