Pharmacogenetics Role of Genetic Variants in Immune-Related Factors: A Systematic Review Focusing on mCRC

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Abstract: Pharmacogenetics plays a key role in personalized cancer treatment. Currently, the clinically available pharmacogenetic markers for metastatic colorectal cancer (mCRC) are in genes related to drug metabolism, such as DPYD for fluoropyrimidines and UGT1A1 for irinotecan. Recently, the impact of host variability in inflammatory and immune-response genes on treatment response has gained considerable attention, opening innovative perspectives for optimizing tailored mCRC therapy. A literature review was performed on the predictive role of immune-related germline genetic biomarkers on pharmacological outcomes in patients with mCRC. Particularly, that for efficacy and toxicity was reported and the potential role for clinical management of patients was discussed. Most of the available data regard therapy effectiveness, while the impact on toxicity remains limited. Several studies focused on the effects of polymorphisms in genes related to antibody-dependent cellular cytotoxicity (FCGR2A, FCGR3A) and yielded promising but inconclusive results on cetuximab efficacy. The remaining published data are sparse and mainly hypothesis-generating but suggest potentially interesting topics for future pharmacogenetic studies, including innovative gene–drug interactions in a clinical context. Besides the tumor immune escape pathway, genetic markers belonging to cytokines/interleukins (IL-8 and its receptors) and angiogenic mediators (IGF1) seem to be the best investigated and hopefully most promising to be translated into clinical practice after validation.

Keywords: colorectal cancer; mCRC; biomarkers; genetic susceptibility factors; immunotherapy; pharmacogenetics; precision medicine; personalized medicine; immune system; clinical implementation

1. Introduction

Colorectal cancer (CRC) is the third most frequent cancer and the second leading cause of death in the world [1]. Further, 20% of the cases have a metastatic CRC (mCRC) at diagnosis and, among the remaining non-metastatic patients, it was observed a 30% probability to develop metastasis during the treatment [2]. 5-fluorouracil (5-FU) in combination with leucovorin (LV) remains the cornerstone of most chemotherapeutic schedules used to treat advanced CRC. The combination of 5-FU/LV with either oxaliplatin (e.g., FOLFOX) or irinotecan (e.g., FOLFIRI) represents a well-established standard first-line treatment [3,4]. Capecitabine could be efficaciously used in place of 5-FU/LV association in several regimens, as XELOX (capecitabine and oxaliplatin) and XELIRI (capecitabine and irinotecan) [4]. A quartet combination FOLFOXIRI (5-FU, LV, oxaliplatin, irinotecan) was also positively tested. In the last decade, the treatment of mCRC has further achieved great advances with the development of biological agents targeting the vascular endothelial growth factor (VEGF, i.e., bevacizumab, aflibercept, ramucirumab) and epidermal growth factor receptor (EGFR, i.e., cetuximab, panitumumab) cascade or leading to a multiple-kinase inhibition (regorafenib) [4]. In the recent period, immunotherapies have also been considered in specific mCRC molecular subtypes [5]. Particularly, regulatory agencies (the Food and Drug Administration (FDA) and the European Medicines Agency (EMA))
approved pembrolizumab for the treatment of metastatic and unresectable microsatellite instability-high (MSI-H)/mismatch repair deficiency (dMMR) CRC, as well as nivolumab plus ipilimumab for the treatment of adult and pediatric patients aged 12 years and older with MSI-H/dMMR mCRC after progression following the use of fluoropyrimidine, oxaliplatin, and irinotecan. The introduction of the immunocheckpoint inhibitors (ICI) has revolutionized the management and survival of many mCRC patients with highly immunogenic characteristics such as MSI-H features. Immunotherapy could be adopted to boost the immune response in the body and/or suggest to immune cells how to identify and destroy cancer cells in the tumor microenvironment (TME), ameliorating the clinical outcome of these patients.

Chronic inflammation and host immune system dysfunction are well-recognized important factors that contribute to CRC development, progression, and prognosis [6–9]. Based on molecular features including immune-related aspects, four different consensus subtypes (CMSs) with different clinical implications have been determined for CRC: the highly immunogenic known as “immune” (CSM1), the inflamed immune-suppressive known as “mesenchymal” (CSM4), and other two poorly immunogenic subtypes called “canonical” (CSM2) and “metabolic” (CSM3) [10]. The cancer-related inflammatory and immune response has been also reported to play a crucial role in the modulation of the efficacy of mCRC treatment, not only concerning ICI but also to standard chemotherapy and targeted agents (Figure 1).

Figure 1. The interplay between immune-related factors and pharmacological strategies to fight mCRC.

Inflammation and the immune system were indicated to impact chemotherapy effectiveness by multiple different mechanisms, including the modulation of chemotherapy-mediated tumor cell death and regulation of inflammatory-related transcriptional factors that in turn affect the expression of absorption, distribution, metabolism, and excretion (ADME) genes [11–13]. Moreover, the same chemotherapeutics, including irinotecan, 5-FU, and oxaliplatin, were indicated to display an immune-modulator effect, influencing the overall antitumor response and disease outcome [14–18]. The immune response has been reported to be important also in determining the cytotoxicity of some targeted agents such as cetuximab. Another mechanism of action of the anti-EGFR drug, beyond the EGFR blockade, is antibody-dependent cellular cytotoxicity (ADCC), also referred to as antibody-
dependent cell-mediated cytotoxicity [18–20]. The cetuximab structure exhibits not only the antigen-binding but also a crystalline fragment (Fc fragment), enabling it to bind to the fragment Cy receptor (FcγR) of immune cells, including the natural killer (NK). The binding of cetuximab to Fc fragment triggers the ADCC pathway.

Despite favorable therapeutic results in recent years, a significant inter-individual heterogeneity in therapy outcome still constitutes a critical problem in mCRC management. Moreover, with the increasing number of effective drugs for mCRC patients often used in combination, the selection of the more appropriate first-line therapeutic options becomes a complex issue influencing the course of therapy [21]. Therefore, the definition of molecular markers that predict which patients will benefit from a specific treatment option could significantly impact the clinical decision-making and therapeutic planning. Some molecular characteristics of tumors (i.e., KRAS, NRAS, and BRAF mutations; MSI-H/dMMR status) [21–24], as well as clinical features (i.e., right- and left-sided mCRC) [21,25], have been identified as predictors of therapy outcome and patient prognosis. However, a significant variability in the response to treatment is still present and additional markers should be defined. On this ground, the evaluation of the host genetic profile could contribute to predict the chemo-responsiveness and to better stratify patients who undergo therapy for mCRC based on the treatment outcome [26]. The emerging role of inflammation and the immune system in mCRC treatment opens a novel field for pharmacogenetic studies aimed at defining the role of host variability in inflammation and immune-related genes in predicting treatment response and patient outcome. In particular, the study of genetic variants in biomarkers, involved in the acute phase, innate, and adaptive immune responses, provides a better understanding of the role of immune regulation in cancer and thus its influence not only on the efficacy and toxicity of chemotherapy, but also on the mechanisms of resistance to therapy. The aim of this review is to critically report and discuss the current literature on the effect of inflammation and immune-related germline variants as predictive markers of mCRC systemic therapy outcome and how they can help stratify patients according to the toxicity risk as well as the likelihood to benefit from the administration of specific anti-tumor agents.

2. Methods

This work used a systematic review methodology and adheres to the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) guidelines. The selected papers regard the predictive role in terms of efficacy and/or toxicity of genetic variants in immune-related biomarkers in patients with mCRC. The systematic literature search was conducted on 21 October 2021 using PubMed database. Boolean operators AND/OR were used to combine search terms. The search strategy included key terms as follows: ((("colorectal cancer" OR CRC) AND (patient OR patients) AND (immun*) AND (therapy OR chemotherapy OR treatment* OR leucovorin OR oxaliplatin OR irinotecan OR fluoropyrimidine OR 5-FU OR *fluorouracil* OR 5FU OR capecitabine OR cetuximab OR bevacizumab OR aflibercept OR ramucirumab OR regorafenib OR panitumumab OR ipilimumab OR nivolumab OR pembrolizumab OR trifluridine OR tipiracil OR encorafenib OR TAS-102 OR vemurafenib OR pertuzuzumab OR trastuzumab OR lapatinib OR deruxtecan OR sotorasib OR adagrasib) AND (polymorphism* OR pharmacogenetic* OR pharmacogenomic*)) NOT ((review[Publication Type]) OR (“systematic review”[Publication Type]) OR (“review”[Publication Type]))). Only manuscripts published in English were considered, and systematic reviews, reviews, short communications, abstracts and case reports were excluded after a systematic screening. Only studies with metastatic patients and immune-related biomarkers, defined as acute-phase cytokines and enzymes, angiogenic mediators, nuclear receptors, toll-like receptors, cytokines and chemokines activators in the early phase of immune response, tumor immune escape factors, cytokines and chemokines activators in the late phase of immune response, cell death regulation factors were considered eligible. Studies with ADME-related, DNA repair, drug-transporters, miRNA, long non-coding RNA (lncRNA), RNA-binding, cell cycle, VEGF-related genes were excluded. In particular,
all the inherited genetic variants with a predictive role in terms of efficacy measured as overall survival (OS), progression-free survival (PFS), overall response rate (ORR), response rate (RR), disease control rate (DCR) and/or toxicity measured as any toxicities, hematotoxicity, diarrhea, gastrointestinal, skin rash toxicity were included in the systematic review based on immune system factors acting in the acute-phase/innate, and adaptive immune response (Figure 2), and their possible role in clinical management was discussed. This systematic research was performed independently by two authors (A.B. and L.S.).

Figure 2. Studies selection flow diagram.

3. Results

3.1. Studies Selection

A total of 506 records were found running the search strategy on 21 October 2022. No duplicate was found. After the first screening evaluating the language and the type of records, 30 records were excluded. All the remaining 476 papers were screened by title, abstract, and full text to identify the eligible studies. 432 records were excluded because not fit with the eligibility criteria. A total of 44 manuscripts were considered and discussed in the systematic review (Figure 2).

3.2. Summary of Evidences

All the genes and their inherited genetic variants with a predictive role in terms of efficacy and/or toxicity retrieved from the 44 studies highlighted by the systematic review are summarized in Table 1. These results are subdivided in eight groups: (1) acute-phase cytokines and enzymes, (2) angiogenic mediators, (3) nuclear receptors, (4) toll-like receptors, (5) cytokines and chemokines activators in the early phase of immune response, (6) tumor immune escape factors, (7) cytokines and chemokines activators in the late phase of immune response, (8) cell death regulation factors. Moreover, to highlight the risk of bias of the results, four levels of evidences (high, moderate, low, very low) were adopted. First of all, the study design was evaluated, attributing “high” level of evidence to the meta-analysis and randomized controlled studies, a “moderate” level to training/replication/control studies, a “low” level to observational studies. Then, other aspects including low sample size, heterogeneity in patients’ ethnicity, in treatment used, and in results of different studies were considered to downgrade the level of evidence.
Table 1. Summary of the efficacy and toxicity evidence for immune-related genes divided in 8 groups belonging to the acute-phase/innate and adaptive immune response.

| Genes                                      | rs Code/ Alias | Therapy (N Patients in Cohorts)                  | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect                                                                                          | Ref.     |
|--------------------------------------------|----------------|-------------------------------------------------|----------|-----------|-------------------|-------------------|------------------------------------------------------------------------------------------------------|----------|
| **Acute-phase cytokines and enzymes**      |                |                                                 |          |           |                   |                   |                                                                                                      |          |
| IL6                                        | rs2069837      | FOLFIRI+BV (223 training, 228 validation),      | 775      | Caucasian | Efficacy         | Moderate          | rs2069837-G allele: lower mPFS (training, validation). rs2069837-AA genotype: higher tumor response (training, validation). No significant association with OS. | [27]     |
|                                            |                | FOLFIRI+CTX (264 control)                       |          |           |                   |                   |                                                                                                      |          |
|                                            | rs1800795      | FOLFIRI+BV (223 training, 228 validation), FOLFIRI+CTX (264 control) | 775      | Caucasian | Efficacy         | Moderate          | No significant association with RR, PFS, OS.                                                        | [27]     |
|                                            |                | BV+FOLFOX/XELOX 132                            | 132      | Caucasian | Efficacy         | Low               | rs1800795-G allele: higher OS.                                                                      | [28]     |
| STAT3                                      | rs744166; rs4796793 | FOLFIRI+BV (223 training, 228 validation), FOLFIRI+CTX (264 control) | 775      | Caucasian | Efficacy         | Moderate          | No significant association with RR, PFS, OS.                                                        | [27]     |
|                                            |                | BV+FOLFOX/XELOX 132                            | 132      | Caucasian | Efficacy         | Low               | rs1053004-C allele: lower risk of grade 3–4 GI toxicity.                                             | [29]     |
| **IL1RN (IL-1RA)**                         | rs4251961; rs579543 | FOLFOX/FOLFIRI 180                             | 180      | Not found | Efficacy         | Low               | rs4251961-TT or rs579543-TT genotypes: higher survival.                                              | [30]     |
| **PTGS2 (COX-2)**                          | rs5275         | CTX 130                                        | 130      | Mainly Caucasian | Efficacy/Toxicity | Low               | rs5275-T allele: better PFS. No associations with OS, ORR, skin rash toxicity.                        | [31]     |
|                                            |                | XELOX                                           | 76       | Asian     | Efficacy         | Very low          | rs5275-TT genotype: better PFS and OS. No significant association with ORR, skin rash toxicity.      | [32]     |
|                                            |                | mFOLFOX/XELOX ± CTX 815                        | 815      | Caucasian | Efficacy/Toxicity | Moderate          | No significant association with RR, skin rash toxicity.                                             | [33]     |
|                                            |                | CTX 130                                        | 130      | Mainly Caucasian | Efficacy/Toxicity | Low               | rs20417-GG: shorter mPFS. No significant association with RR, skin rash toxicity.                    | [31]     |
| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|---------------|--------------------------------|----------|-----------|------------------|------------------|-------------|------|
| **STING1**<sup>(STING)</sup> | rs7380824 | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 control cohort 1, 215 control cohort 2) | 451 | Caucasian | Efficacy | Moderate | rs7380824-any T allele: lower ORR in CTX cohort. | [34] |
| | rs1131769 | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 control cohort 1, 215 control cohort 2) | 451 | Caucasian | Efficacy | Moderate | rs1131769-any T allele: shorter OS in CTX cohort. | [34] |
| **CGAS**<sup>(cCAS)</sup> | rs610913; rs311678; rs6907936 | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 control cohort 1, 215 control cohort 2) | 451 | Caucasian | Efficacy | Moderate | No significant association with ORR, PFS, OS. | [34] |
| **IFNB1** | rs1051922 | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 control cohort 1, 215 control cohort 2) | 451 | Caucasian | Efficacy | Moderate | rs1051922-G/A and A/A genotype: shorter PFS in CTX cohort. | [34] |
| | rs10964831 | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 control cohort 1, 215 control cohort 2) | 451 | Caucasian | Efficacy | Moderate | No significant association with ORR, PFS, OS. | [34] |
| **Angiogenic mediators** | | | | | | | | |
| **CXCR4** | rs2228014 | FOLFOX/XELOX+BV (144 training), FOLFIRI+BV (424 validation A), FOLFIRI+BV (229 validation B), FOLFIRI+BV (77 control) | 874 | Asian and Caucasian | Efficacy | Moderate | rs2228014-T allele: shorter mPFS and trend of shorter mOS. | [35] |
| **CXCL12** | rs1801157; rs3740085 | FOLFOX/XELOX+BV (144 training), FOLFIRI+BV (424 validation A), FOLFIRI+BV (229 validation B), FOLFIRI+BV (77 control) | 874 | Asian and Caucasian | Efficacy | Moderate | No significant association with PFS, OS, ORR. | [35] |
| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|----------------|-------------------------------|----------|-----------|-------------------|------------------|-------------|------|
| CCL2  | rs4586         | FOLFIRI+BV (228 discovery, 248 validation), FOLFIRI+CTX (248 control) | 724      | Caucasian | Efficacy          | Moderate         | rs4586-C allele: better PFS in KRAS mutant of discovery cohort. No significant association with OS, RR. | [36] |
| CCR2  | rs3092964      | FOLFIRI+BV (228 discovery, 248 validation), FOLFIRI+CTX (248 control) | 724      | Caucasian | Efficacy          | Moderate         | No significant association with PFS, OS, ORR. | [36] |
| CCL18 | rs14304        | FOLFIRI+BV (228 discovery, 248 validation), FOLFIRI+CTX (248 control) | 724      | Caucasian | Efficacy          | Moderate         | rs14304-T allele: longer PFS (not validated in FIRE-BV cohort). No significant association with OS, RR. | [36] |
| TBK1  | rs7486100; rs12313449 | FOLFIRI+BV (228 discovery, 248 validation), FOLFIRI+CTX (248 control); FOLFIRI+BV (486) | 486–724  | Caucasian | Efficacy          | Moderate         | rs7486100-T allele: worse PFS in BV cohorts, worse OS only in KRAS wild-type. No significant association with OS, RR in all population. | [36,37] |
| IRF3  | rs2304205; rs10415576 | FOLFIRI+BV (228 discovery, 248 validation), FOLFIRI+CTX (248 control) | 724      | Caucasian | Efficacy          | Moderate         | rs2304205-C allele: better PFS in KRAS mutant of discovery cohort. No significant association with OS, RR. | [36] |
| IGF1  | rs6220         | BV+FOLFOX/XELOX                | 132      | Caucasian | Efficacy          | Low              | rs6220-G allele: increased OS. No significant association with RR and PFS. | [28] |
|       |                | FOLFIRI+BV (295), FOLFIRI+CTX (305) | 614      | Caucasian | Efficacy          | Moderate         | No significant association with PFS, OS. | [38] |
|       |                | CTX                          | 130      | Caucasian | Efficacy          | Low              | No significant association with ORR, PFS, OS. | [39] |
|       | rs6214         | CTX                          | 130      | Caucasian | Efficacy          | Low              | rs6214-TT genotype: worse PFS and OS. | [39] |
|       |                | FOLFIRI+BV (295), FOLFIRI+CTX (305) | 614      | Caucasian | Efficacy          | Moderate         | No significant association with PFS, OS. | [38] |
|       | rs2946834      | CTX                          | 130      | Caucasian | Efficacy          | Low              | rs2946834-AA genotype: better PFS in all population and better PFS, ORR in RAS wild-type subgroup. | [39] |
|       |                | FOLFIRI+BV (295), FOLFIRI+CTX (305) | 614      | Caucasian | Efficacy          | Moderate         | rs2946834-A allele: better PFS in all population and better PFS, OS in RAS wild-type subgroup. | [38] |
|       | rs7136446      | CTX                          | 130      | Caucasian | Efficacy          | Low              | rs7136446-A allele: worse PFS in all patients. No significant association with OS, ORR. | [39] |
Table 1. Cont.

| Genes   | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect                                                                 | Ref. |
|---------|----------------|---------------------------------|----------|-----------|-------------------|-------------------|-------------------------------------------------------------------------------|------|
| IGF1R   | rs2016347      | CTX                             | 130      | Caucasian | Efficacy          | Low               | rs2016347-G allele: worse OS in all population and in wild-type KRAS subgroup. No significant association with PFS, ORR. | [39] |
|         |                | FOLFIRI+BV (295), FOLFIRI+CTX (305) | 614      | Caucasian | Efficacy          | Moderate          | No significant association with PFS, OS.                                      | [38] |
|         | rs2272037      | CTX                             | 130      | Caucasian | Efficacy          | Low               | rs2272037-C allele: worse OS. No significant association with ORR, PFS.        | [39] |
|         | rs2229765      | CTX                             | 130      | Caucasian | Efficacy          | Low               | No significant association with ORR, PFS, OS.                                 | [39] |
| IRS1    | rs1801123      | FOLFIRI+BV (295), FOLFIRI+CTX (305) | 614      | Caucasian | Efficacy          | Moderate          | rs1801123-C allele: worse OS in all population and in RAS wild-type. No significant association with PFS, OS. | [38] |
|         | rs1801278      | FOLFIRI+BV (295), FOLFIRI+CTX (305) | 614      | Caucasian | Efficacy          | Moderate          | No significant association with PFS, OS.                                     | [38] |
| IRS2    | rs2289046; rs1805097 | FOLFIRI+BV (295), FOLFIRI+CTX (305) | 614      | Caucasian | Efficacy          | Moderate          | No significant association with PFS, OS.                                     | [38] |
|         | rs2070744      | BV+FOLFOX/FOLFIRI (114 study), FOLFIRI (123 control) | 237      | Caucasian | Efficacy          | Low               | ENOS Haplo1/Haplo1: longer mPFS; combining ENOS Haplo1/Haplo1 and ENOS Haplo 2/Haplo 2: longer PFS, OS. No significant association with RR. | [40] |
|         |                | BV+FOLFOX6 (120 study), FOLFIRI (112 control) | 232      | Caucasian | Efficacy / Toxicity | Low               | No significant association with ORR, PFS, OS, toxicity.                       | [41] |
| NOS3/ENOS (eNOS) | rs1799983 | BV+FOLFOX/FOLFIRI (114 study), FOLFIRI (123 control) | 237      | Caucasian | Efficacy          | Low               | rs1799983-GT: worse ORR, PFS, OS. ENOS Haplo1/Haplo1: longer mPFS; combining ENOS Haplo1/Haplo1 and ENOS Haplo 2/Haplo 2: longer PFS, OS. | [40] |
|         |                | BV+FOLFOX6 (120 study), FOLFIRI (112 control) | 232      | Caucasian | Efficacy / Toxicity | Low               | rs1799983-TT genotype: higher severe toxicity in BV-based group. No significant association with ORR, PFS, OS. | [41] |
| VNTR    | rs4a/b 27pb    | BV+FOLFOX/FOLFIRI (114 study), FOLFIRI (123 control) | 237      | Caucasian | Efficacy          | Low               | VNTR-4bb: longer PFS, OS. ENOS Haplo1/Haplo1: longer mPFS; combining ENOS Haplo1/Haplo1 and ENOS Haplo 2/Haplo 2: longer PFS, OS. No significant association with ORR. | [40] |
Table 1. Cont.

| Genes     | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect                                                                 | Ref. |
|-----------|----------------|---------------------------------|----------|-----------|-------------------|-------------------|------------------------------------------------------------------------------|------|
| NOS2/NOS  | rs27779248     | BV+FOLFIRI/FOLFOXIRI (227+231 evaluation cohorts), BV+FOLFIRI/FOLFOXIRI (301+187 validation cohorts) | 946       | Caucasian | Efficacy          | Moderate          | No significant association with PFS, OS, RR.                                 | [42] |
|           |                | CCTTT repeat                    |          |           |                   |                   |                                                                              |      |
| HIF1A     | rs11549465     | BV+FOLFIRI/XELOX                | 132      | Caucasian | Efficacy          | Low               | rs11549465-T allele: increased PFS only in univariate analysis. However, in the construction of the decision tree was the most important factor that determines PFS. | [28] |
| TXN       | rs2301242      | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 BV cohort) | 236      | Caucasian | Efficacy          | Low               | No significant association with PFS, tumor response neither in FOLFIRI+CTX or in FOLFIRI+BV. | [43] |
| TXN2      | rs4821494      | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 BV cohort) | 236      | Caucasian | Efficacy          | Low               | rs4821494-any G allele: worse OS in FOLFIRI+CTX. No significant association with PFS, tumor response in FOLFIRI+CTX, neither in FOLFIRI+BV. | [43] |
|           | rs9619611; rs59841625 | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 BV cohort) | 236      | Caucasian | Efficacy          | Low               | No significant association with PFS, tumor response neither in FOLFIRI+CTX or in FOLFIRI+BV. | [43] |
| CAT       | rs7943316; rs564250; rs11604331; rs1001179; rs769217 | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 BV cohort) | 236      | Caucasian | Efficacy          | Low               | No significant association with PFS, tumor response neither in FOLFIRI+CTX or in FOLFIRI+BV. | [43] |
| GPX4      | rs4807542      | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 BV cohort) | 236      | Caucasian | Efficacy          | Low               | rs4807542-any A allele: worse OS in FOLFIRI+CTX. No significant association in FOLFIRI+BV. | [43] |
|           | rs757229; rs713041 | FOLFIRI+CTX (129 CTX cohort), FOLFIRI+BV (107 BV cohort) | 236      | Caucasian | Efficacy          | Low               | No significant association with PFS, tumor response neither in FOLFIRI+CTX or in FOLFIRI+BV. | [43] |
### Table 1. Cont.

| Genes      | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect                                                                 | Ref. |
|------------|----------------|--------------------------------|----------|-----------|-------------------|-------------------|-----------------------------------------------------------------------------|------|
| Nuclear receptors |                |                                |          |           |                   |                   |                                                                             |      |
| NR1I2 (PXR) | rs10934498     | FOLFIRI/FOLFIRINOX ± BV/CTX | 109      | Caucasian (French) | Toxicity          | Low               | rs10934498-A allele: decrease risk of grade 3-4 hematotoxicity.             | [44] |
|            | rs1523127      | FOLFIRI/FOLFIRINOX ± BV/CTX | 109      | Caucasian (French) | Toxicity          | Low               | rs1523127-C allele: increased risk of grade 3-4 hematotoxicity.             | [44] |
|            | rs2472677      | FOLFIRI/FOLFIRINOX ± BV/CTX | 109      | Caucasian (French) | Toxicity          | Low               | rs2472677-G allele: increased risk of all type of grade 3-4 toxicity and of-grade 3-4 hematotoxicity. | [44] |
|            | rs3814055      | FOLFIRI/FOLFIRINOX ± BV/CTX | 109      | Caucasian (French) | Toxicity          | Low               | rs3814055-T allele: increased risk of grade 3-4 hematotoxicity.             | [44] |
| NR1I3 (CAR) | rs2307418; rs2307424; rs2501873; rs2502815; rs3003596; rs4073054; rs6686001 | FOLFIRI/FOLFIRINOX ± BV/CTX | 337; 335 | Caucasian | Efficacy          | Moderate           | No significant association with toxicity.                                   | [44] |
| NR1I1 (VDR) | rs7299460      | FOLFIRI (247 Italian discovery, 90 Canadian replication); FOLFIRI (250 Italian discovery, 92 Canadian replication) | 337; 335 | Caucasian | Efficacy          | Moderate           | rs7299460-T: longer OS. Highly predictive genetic score when combined with IL15RA-rs7910212-TC/CC, SMAD3-rs7179840-TT, VDR-rs7299460-CC. | [45,46] |
| Toll-like receptors |                |                                |          |           |                   |                   |                                                                             |      |
| TLR1       | rs5743618      | FOLFIRI+BV (228 discovery, 297 validation) | 525      | Caucasian | Efficacy          | Moderate           | rs5743618-T allele: worse RR (validated), worse PFS and OS only in the discovery cohort (not validated). | [47] |
|            | rs5743565      | FOLFIRI+BV (228 discovery, 297 validation) | 525      | Caucasian | Efficacy          | Moderate           | No significant association with RR, PFS, OS.                               | [47] |
| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|----------------|---------------------------------|----------|-----------|-------------------|-------------------|-------------|------|
| **TLR2** | rs3804099 | FOLFIRI+BV (228 discovery, 297 validation) | 525 | Caucasian | Efficacy | Moderate | rs3804099-C allele: better PFS only in univariate analyses (not validated). No significant association with RR, OS. | [47] |
| | rs4696480 | FOLFIRI+BV (228 discovery, 297 validation) | 525 | Caucasian | Efficacy | Moderate | rs4696480-T allele: worse PFS only in univariate analyses (not validated). No significant association with RR, OS. | [47] |
| **TLR3** | rs3775291 | CTX+OXA-based | 1948 | Caucasian | Efficacy | Moderate | No significant association with DFS, OS. | [48] |
| **TLR4** | rs4986790 | IRI/FOLFIRI | 46 | Brazilian | Toxicity | Very low | rs4986790-(AG+GG) genotypes: more likely to experience diarrhea. | [49] |
| | rs4986790 | CTX+OXA-based | 1948 | Caucasian | Efficacy | Moderate | No significant association with DFS or OS. | [48] |
| | rs4986791 | IRI/FOLFIRI | 46 | Brazilians | Toxicity | Very low | TLR4-rs4986791-(CT+TT) genotypes: more likely to experience diarrhea. | [49] |
| | rs4986791 | CTX+OXA-based | 1948 | Caucasian | Efficacy | Moderate | No significant association with DFS, OS. | [48] |
| **TLR6** | rs3821985 | FOLFIRI+BV (228 discovery, 297 validation) | 525 | Caucasian | Efficacy | Moderate | No significant association with ORR, PFS, OS. | [47] |
| | rs5743818 | FOLFIRI+BV (228 discovery, 297 validation) | 525 | Caucasian | Efficacy | Moderate | rs5743818-AA genotype: lower RR (not validated), lower PFS in discovery and validation cohorts. No significant association with PFS, OS. | [47] |
| **TLR7** | rs3853839 | FOLFIRI+CTX (244 discovery), FOLFIRI+BV (246 control), FOLFOX/SOX+CTX (76 validation) | 566 | Caucasian | Efficacy | Moderate | rs3853839-GG genotype: longer PFS (validated for CTX-based) and OS (not validated). This preliminary association with PFS was not observed in BV-based cohort. No significant association with RR. | [50] |
| | rs187084 | FOLFIRI+CTX (244 discovery), FOLFIRI+BV (246 control), FOLFOX/SOX+CTX (76 validation) | 566 | Caucasian | Efficacy | Moderate | rs187084-C allele: better PFS (not validated). No significant association with RR, OS. | [50] |
| Genes       | rs Code/ Alias   | Therapy (N Patients in Cohorts)                                                                 | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect                                                                 | Ref. |
|-------------|------------------|-----------------------------------------------------------------------------------------------|----------|-----------|-------------------|-------------------|-----------------------------------------------------------------------------|------|
| FPR1        | rs867228         | CTX+OXA-based 1948                                                                             | 1948     | Caucasian | Efficacy          | Moderate          | No significant association with DFS, OS.                                   | [48] |
| MMP2        | rs243865         | FOLFIRI+BV 486                                                                                   | 486      | Caucasian | Efficacy          | Moderate          | rs243865-any T: better OS in KRAS mutant patients.                          | [37] |
| MAP3K7      | rs1145727        | FOLFIRI + BV (228 discovery, 297 validation)                                                   | 525      | Caucasian | Efficacy          | Moderate          | rs1145727- AA: shorter OS (not validated). No significant association with RR, PFS. | [47] |
| MAP3K7      | rs157688         | FOLFIRI+BV (228 discovery, 297 validation)                                                     | 525      | Caucasian | Efficacy          | Moderate          | rs157688- CC genotype: longer PFS and OS only in univariate analyses (not validated). No significant association with RR. | [47] |
|             | rs157432         | FOLFIRI+BV (228 discovery, 297 validation)                                                     | 525      | Caucasian | Efficacy          | Moderate          | No significant association with RR, PFS, OS.                               | [47] |
| Cytokines and chemokines in the early phase of immune response |          |                                                                                              |          |           |                   |                  |                                                                           |      |
| CXCL8       | rs4073           | BV+FOLFOX6 (120 discovery Caucasian), FOLFOX6 (112 control Caucasian); BV-based (125 Asian); BV+FOLFOXIRI (180 Caucasian) | 125–232  | Mainly Caucasian | Efficacy / Toxicity | Moderate / Low | rs4073-A variant: shorter PFS, OS, ORR, and higher IL-8 levels only in BV-based group. | [40,51,52] |
|            |                  | CTX (130); 5-FU/OXA (105); Regorafenib (47); FOLFIRI+CTX (30)                                 | 30–130   | Mainly Caucasian | Efficacy / Toxicity | Low / Very low | No significant association with PFS, OS, ORR, skin rash toxicity.         | [31,53–55] |
| CXCR1/ IL8RA | rs2234671       | BV+FOLFOX/XELOX 132                                                                           | 132      | Caucasian | Efficacy          | Low               | rs2234671-GG genotype: higher tumor RR. No significant association with PFS, OS. The combinations of CXCR1 variants may improve the prediction success for PFS and OS, and rs2234671 was the main split criteria in the decision tree for RR. | [28] |
|            |                  | 5-FU/OXA 105                                                                                   | 105      | Mainly Caucasian | Efficacy          | Low               | rs2234671-GC genotype (vs GG): worse TTP.                                  | [53] |
|            |                  | CTX ± mFOLFOX/XELOX 815                                                                         | 815      | Caucasian | Efficacy / Toxicity | Moderate          | No significant association with RR, skin rash toxicity.                   | [33] |
| Genes       | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect                                                                                                                                                                                                 | Ref. |
|-------------|----------------|--------------------------------|----------|-----------|-------------------|-------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------|
| CXCR2/IL8RB | rs2230054      | BV+FOLFOX/XELOX 132 Caucasian | Efficacy | Low       | rs2230054-TT: lower tumor RR. An ethnicity effect was also reported: lower RR in Caucasians but not in Asians and Hispanics. No significant association with PFS, OS.                                            | [28] |
|             |                | BV-based 125 Asian             | Efficacy | Low       | No significant association with ORR, PFS, OS.                                                                                                                                     | [51] |
|             |                | Regorafenib 47 Caucasian       | Efficacy / Toxicity | Very low | No significant association with RR, PFS, toxicity.                                                                                                                                | [54] |
|             |                | 5-FU/OXA 105 Mainly Caucasian | Efficacy | Low       | No significant association with OS, RR, TTP.                                                                                                                                       | [53] |
| IL10        | rs1800896      | BV-based 125 Asian             | Efficacy | Low       | No significant association with ORR, PFS, OS.                                                                                                                                     | [51] |
| IL17A       | rs2275913      | BV-based 122 Caucasian         | Efficacy | Low       | No significant association with OS, PFS, serum cytokine levels.                                                                                                                  | [56] |
| IL17F       | rs763780       | BV-based 122 Caucasian         | Efficacy | Low       | No significant association with OS, PFS, serum cytokine levels.                                                                                                                  | [56] |
| Tumor immune escape factors |                |                                |          |           |                   |                   |                                                                                                                                                                                                           |      |
| CD24        | rs52812045     | CTX ± IRI (105 training),     | 924      | Caucasian | Efficacy          | Moderate          | rs52812045-AA genotype or A allele: shorter mPFS and OS in CTX-based groups and no relation in BV-based groups.                                                                                           | [57] |
|             |                | CTX+FOLFIRI (225)/OXA (74) (validation cohorts), BV+FOLFIRI (520 control) |          |           |                   |                   |                                                                                                                                                                                                           |      |
| CTLA4       | rs231777       | CTX ± IRI (105 training),     | 924      | Caucasian | Efficacy          | Moderate          | rs231777-T allele and CT: higher risk of progression, and worse PFS in CTX-based group, respectively.                                                                                                   | [57] |
|             |                | CTX+FOLFIRI (225)/OXA (74) (validation cohorts), BV+FOLFIRI (520 control) |          |           |                   |                   |                                                                                                                                                                                                           |      |
| PDCD1 (PD1) | rs2227981; rs7421861 | CTX ± IRI (105 training),     | 924      | Caucasian | Efficacy          | Moderate          | No significant association with RR, PFS, OS.                                                                                                                                                               | [57] |
|             |                | CTX+FOLFIRI (225)/OXA (74) (validation cohorts), BV+FOLFIRI (520 control) |          |           |                   |                   |                                                                                                                                                                                                           |      |
### Table 1. Cont.

| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|----------------|--------------------------------|----------|-----------|-------------------|-------------------|-------------|------|
| CD274 (PDL1) | rs2297137 | CTX ± IRI (105 training), CTX+FOLFIRI (225)/OXA (74) (validation cohorts), BV+FOLFIRI (520 control) | 924 | Caucasian | Efficacy | Moderate | rs2297137-G allele: worse tumor response in CTX-based group. rs2297137-A allele had a prolonged OS in BV-based group. [57] |
| CD274 (PDL1) | rs2297136 | CTX ± IRI (105 training), CTX+FOLFIRI (225)/OXA (74) (validation cohorts), BV+FOLFIRI (520 control); OXA-based+BV (76), IRI-based+BV (65), OXA- or IRI-based + BV (152) | 924; 141–152 | Caucasian; Chinese | Efficacy | Low | No significant association with RR, PFS, OS in Caucasian. CD274-rs2297136-AA genotype: better PFS and OS in Chinese. In KRAS mutant Chinese subgroup, CD274-rs2297136-AA: longer PFS, trend longer OS. [57–59] |
| rs10122089 | | CTX ± IRI (105 training), CTX+FOLFIRI (225)/OXA (74) (validation cohorts), BV+FOLFIRI (520 control) | 924 | Caucasian | Efficacy | Moderate | No significant association with RR, PFS, OS. [57] |
| KIRs/HLAs | KIR haplotype combination | FOLFIRI | 224 | Caucasian | Efficacy | Low | Presence of haplotype combination of KIR2DL5A, 2DS5, 2DS1, 3DS1, and KIR3DS1/HLA-Bw4-I80 and absence of KIR2DS4 and 3DL1: increased CR rates. Absence of KIR2DS5 and presence of KIR3DL1/HLA-Bw4-I80: better OS. [58] |
| 16 KIRs | | CTX-based | 70 | Caucasian | Efficacy | Very low | No significant association with OS, PFS. [60] |
| ADORA2A (A2AR) | rs5751876 | FOLFIRI+BV (107 discovery), FOLFIRI+BV (215 validation), FOLFIRI+CTX (129 control) | 451 | Caucasian | Efficacy | Moderate | No significant association with OS, PFS, RR. [61] |
| ADORA2B (A2BR) | rs2015353 | FOLFIRI+BV (107 discovery), FOLFIRI+BV (215 validation), FOLFIRI+CTX (129 control) | 451 | Caucasian | Efficacy | Moderate | rs2015353-TT: longer OS in FOLFIRI+BV group. [61] |
| CD39/ ENTPD1 | rs11188513 | FOLFIRI+BV (107 discovery), FOLFIRI+BV (215 validation), FOLFIRI+CTX (129 control) | 451 | Caucasian | Efficacy | Moderate | rs11188513-C allele: shorter PFS (not validated), OS in FOLFIRI-BV group (validated). [61] |
| | rs2226163 | FOLFIRI+BV (107 discovery), FOLFIRI+BV (215 validation), FOLFIRI+CTX (129 control) | 451 | Caucasian | Efficacy | Moderate | rs2226163-GG: longer OS only in FOLFIRI+BV group. [61] |
| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|----------------|---------------------------------|----------|-----------|-------------------|-------------------|-------------|------|
| **CD73/ NT5E** | rs2229523 | FOLFIRI+BV (107 discovery), FOLFIRI+BV (215 validation), FOLFIRI+CTX (129 control) | 451 | Caucasian | Efficacy | Moderate | rs2229523-A allele: longer OS in FOLFIRI+BV group. | [61] |
| | rs9657182 | CTX ± IRI (105 training), CTX+FOLFIRI/OXA (225 Caucasian/ 74 Japanese validation), BV+FOLFIRI (520 control) | 924 | Mainly Caucasian | Efficacy | Moderate | rs9657182-CT: longer OS in CTX-based group and shortest mOS in the Japanese validation cohort. Ethnicity effect for OS. rs9657182-T allele: shorter PFS in FOLFIRI+BV group. | [57] |
| **IDO1** | rs3739319 | CTX ± IRI (105 training), CTX+FOLFIRI/OXA (225 Caucasian/ 74 Japanese validation), BV+FOLFIRI (520 control) | 924 | Mainly Caucasian | Efficacy | Moderate | rs3739319-GG: longer mOS in CTX-based group. rs3739319-A allele: longer PFS in CTX-based group. | [57] |
| | rs10108662 | CTX ± IRI (105 training), CTX+FOLFIRI/OXA (225 Caucasian/ 74 Japanese validation), BV+FOLFIRI (520 control) | 924 | Mainly Caucasian | Efficacy | Moderate | No significant association with RR, PFS, OS. | [57] |
| **HIF1A** | rs2057482 | FOLFIRI+BV (107 discovery), FOLFIRI+BV (215 validation), FOLFIRI+CTX (129 control) | 451 | Caucasian | Efficacy | Moderate | No significant association with OS, PFS, RR. | [61] |
| | rs11549465 | FOLFIRI+BV (107 discovery), FOLFIRI+BV (215 validation), FOLFIRI+CTX (129 control) | 451 | Caucasian | Efficacy | Moderate | No significant association with OS, PFS, RR. | [61] |
| **HNF1A** | rs2244608 | FOLFIRI ± BV/other (167 Canadian study), FOLFIRI (250 Italian validation) | 417 | Caucasian | Efficacy | Low | rs2244608-G: improved PFS, enhanced blood exposure to SN-38, 41% increased biliary index, 24% decreased glucuronidation ratio. | [62] |

**Cytokines in the late phase of immune response**

| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|----------------|---------------------------------|----------|-----------|-------------------|-------------------|-------------|------|
| **IFNG** | rs2069718; rs1861493 | OXA-based+BV (76), IRI-based+BV (65); OXA-based or IRI-based+BV (152) | 141–152 | Chinese | Efficacy | Low | No significant association with OS, PFS. | [58,59] |
| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|---------------|---------------------------------|----------|-----------|-------------------|------------------|-------------|------|
| **IFNGR1** | | | | | | | | |
| rs2234711 | OXA-based+BV (76), IRI-based+BV (65); OXA-based or IRI-based+BV (152) | 141–152 | Chinese | Efficacy | Low | rs2234711-G allele: longer OS, only in univariate analysis. [58,59] | | |
| rs9376267 | OXA-based+BV (76), IRI-based+BV (65); OXA-based or IRI-based+BV (152) | 141–152 | Chinese | Efficacy | Low | rs9376267-T allele: longer OS, only in univariate analysis. [58,59] | | |
| **IFNGR2** | | | | | | | | |
| rs9608753; rs1059293 | OXA-based+BV (76), IRI-based+BV (65); OXA-based or IRI-based+BV (152) | 141–152 | Chinese | Efficacy | Low | No significant association with OS, PFS. [58,59] | | |
| **JAK1** | | | | | | | | |
| rs112395617 | OXA-based+BV (76), IRI-based+BV (65); OXA-based or IRI-based+BV (152) | 141–152 | Chinese | Efficacy | Low | No significant association with OS, PFS. [58,59] | | |
| **JAK2** | | | | | | | | |
| rs1887429; rs1887428 | OXA-based+BV (76), IRI-based+BV (65); OXA-based or IRI-based+BV (152) | 141–152 | Chinese | Efficacy | Low | No significant association with OS, PFS. [58,59] | | |
| **STAT1** | | | | | | | | |
| rs3088307; rs41430444; rs6745710 | OXA-based+BV (76), IRI-based+BV (65); OXA-based or IRI-based+BV (152) | 141–152 | Chinese | Efficacy | Low | No significant association with OS, PFS. [58,59] | | |
| **STAT2** | | | | | | | | |
| rs2020854 | OXA-based+BV (76), IRI-based+BV (65); OXA-based or IRI-based+BV (152) | 141–152 | Chinese | Efficacy | Low | No significant association with OS, PFS. [58,59] | | |
| **IL15RA** | rs7910212 | FOLFIRI (250 Italian discovery and 92 Canadian validation) | 335 | Caucasian | Efficacy | Moderate | rs7910212-C allele: worse OS. Highly predictive genetic score when combined with IL15RA-rs7910212-TC/CC, SMAD3-rs1779840-TT, NR1I2-rs1054190-TT, VDR-rs7299460-CC. [46] | | |
| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|---------------|--------------------------------|----------|-----------|-------------------|-------------------|-------------|------|
| SMAD3 | rs7179840     | FOLFIRI (250 Italian discovery and 92 Canadian validation) | 335      | Caucasian | Efficacy          | Moderate          | rs7179840-C allele: better OS. Highly predictive genetic score when combined with IL15RA-rs7910212-TC/CC, SMAD3-rs7179840-TT, NR1I2-rs1054190-CC. | [46] |
|       |               | Regorafenib (79 Japan discovery, 150 Italian validation) | 229      | Japan and Italian | Efficacy | Low | rs1130371-A allele: shorter PFS (in Japan and Italian cohort) and OS (only in validation Italian cohort). Ethnicity effect for OS. rs1130371-GG variant: higher CCL5 serum level changes between baseline and day 21 but did not correlate with CCL3 levels. | [63] |
| CCL3  | rs1130371     | FOLFOX+BV (61 evaluation), FOLFOX/XELOX+BV (71 validation), FOLFOX (84 control) | 216      | Asian     | Efficacy         | Low | No significant association with PFS, OS, RR. | [64] |
|       |               | Regorafenib (79 Japan discovery, 150 Italian validation) | 229      | Japan and Italian | Efficacy (serum level) | Low | rs1130371-GG genotype: increased CCL3 levels at day 21. | [65] |
| CCL4  | rs1634517     | Regorafenib (79 Japan discovery, 150 Italian validation) | 229      | Japan and Italian | Efficacy         | Low | rs1634517-A allele: shorter PFS (in Japan and Italian cohort), OS (only in Italian validation cohort), rs1634517-CC variant: higher CCL5 serum level changes at baseline and day 21 but did not correlate with CCL4 levels. | [63] |
|       |               | FOLFOX+BV (61 evaluation), FOLFOX/XELOX+BV (71 validation), FOLFOX (84 control) | 216      | Asian     | Efficacy          | Low | rs1634517-A allele: shorter OS (only in control group treated without BV). | [64] |
|       |               | Regorafenib (79 Japan discovery, 150 Italian validation) | 229      | Japan and Italian | Efficacy (serum level) | Low | rs1634517-CC variant: higher CCR5 changes between baseline and day 21. Pattern of decreased CCL4 levels at day 21 had a trend toward longer PFS. | [65] |
Table 1. Cont.

| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|----------------|-------------------------------|----------|-----------|-------------------|-------------------|------------|------|
| CCL5  | rs2280789      | Regorafenib (79 Japan discovery, 150 Italian validation) | 229      | Japan and Italian | Efficacy / Toxicity | Low   | rs2280789-GG: longer OS (only in Japan discovery cohort), higher incidence of grade>3 hand–foot skin reaction, lower CCL5 level at baseline and day 21, lower VEGF-A level at day 21. rs2280789 and rs3817655 showed high LD. | [63] |
|       |                | FOLFOX+BV (61 evaluation), FOLFOX/XELOX+BV (71 validation), FOLFOX (84 control) | 216      | Asian     | Efficacy          | Low   | rs2280789-GG: shorter OS (in control group treated without BV). rs2280789-G allele: longer PFS, OS, RR in FOLFOX+BV compared FOLFOX. rs2280789-G allele: higher VEGF-A level at baseline, greater decrease of VEGF-A levels at day 14 and 56. | [64] |
|       |                | FOLFIRI+CTX (244 evaluation), FOLFIRI+BV (247 control) | 491      | KRAS wt  Caucasian | Efficacy | Moderate | rs2280789-G allele: shorter OS. Combined with tumor location (left/right) a better stratification emerged: left- rs2280789-AA better mOS, right-rs2280789-G allele worse mOS. | [66] |
|       |                | Regorafenib (79 Japan discovery, 150 Italian validation) | 229      | Japan and Italian | Efficacy (serum level) | Low   | rs2280789-G allele: higher CCL3 level between baseline and day 21, but CCL4 decreased. | [65] |
|       | rs3817655      | Regorafenib (79 Japan discovery, 150 Italian validation) | 229      | Japan and Italian | Efficacy / Toxicity | Low   | rs3817655-TT: longer OS (only in Japan discovery cohort), higher incidence of grade>3 hand–foot skin reaction, lower CCL5 level at baseline and day 21, lower VEGF-A level at day 21. rs2280789 and rs3817655 showed high LD. | [63] |
|       |                | Regorafenib (79 Japan discovery, 150 Italian validation) | 229      | Japan and Italian | Efficacy (serum level) | Low   | rs3817655-TT: higher CCL3 level between baseline and day 21, but CCL4 decreased. Increased CCL3 level at PD was associated with longer OS. | [65] |
### Table 1. Cont.

| Genes | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|-------|----------------|---------------------------------|----------|-----------|-------------------|-------------------|-------------|------|
| CCR5  | rs1799988      | Regorafenib (79 Japan discovery, 150 Italian validation) | 229 Japan and Italian | Efficacy / Toxicity | Low | rs1799988-TT variant: higher risk of no DC (PD), higher CCL5 serum level changes at baseline and day 21. No significant association with OS, PFS. | [63] |
|       |                | FOLFOX+BV (61 evaluation), FOLFOX/XELOX+BV (71 validation), FOLFOX (84 control) | 216 Asian | Efficacy | Low | rs1799988-T allele: shorter OS (only in control group without BV). CCR5 rs1799988-T allele: shorter OS. | [64] |
|       |                | FOLFIRI+CTX (244 evaluation), FOLFIRI+BV (247 control) | 491 KRAS wt Caucasian | Efficacy | Moderate | rs1799988-T allele: lower RR (trend), shorter PFS (only in evaluation). Opposite effects between right- and left-sided tumors: TT variant: favorable in right-sided tumors, while T allele was unfavorable in left-sided tumors for tumor response, PFS, and OS. | [66] |
|       |                | Regorafenib (79 Japan discovery, 150 Italian validation) | 229 Japan and Italian | Efficacy (serum level) | Low | Patients with TS had lower mean changes in serum CCR5 levels between baseline and day 21. No significant differences in DC and TS at baseline, higher in TS at day 21. | [65] |
| CDX2  | rs3812863      | FOLFOX ± BV (146 Japan discovery), FOLFOXIRI+BV (230 Caucasian validation), FOLFIRI+BV (228 Caucasian control) | 604 Japan and Caucasian | Efficacy | Moderate | rs3812863-GG genotype: higher ORR, trend of better OS, PFS. | [67] |
| MS4A12 | rs4939378     | FOLFOX ± BV (146 Japan discovery), FOLFOXIRI+BV (230 Caucasian validation), FOLFIRI+BV (228 Caucasian control) | 604 Japan and Caucasian | Efficacy | Moderate | rs4939378-GG genotype: longer OS, PFS. | [67] |
| HIF1A | rs12434438     | Regorafenib (79 Japan discovery, 150 Italian validation) | 229 Japan and Italian | Efficacy / Toxicity | Low | No significant association with PFS, OS, toxicity. | [63] |
Table 1. Cont.

| Genes                   | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect                                      | Ref.      |
|-------------------------|----------------|---------------------------------|----------|-----------|-------------------|-------------------|-------------------------------------------------|-----------|
| Regulated cell death factors |                |                                 |          |           |                   |                   |                                                 |           |
| FCGR2A                  | rs1801274      | CTX-based                        | 39–1123  | Mainly Caucasian | Efficacy          | High              | Meta-analysis: rs1801274-His (A allele): generally, no benefit for mOS, mORR, mDCR in CTX-based regimen. | [31,60,68–86] |
| FCGR3A                  | rs396991       | CTX-based                        | 39–2831  | Mainly Caucasian | Efficacy / Toxicity | High              | Meta-analysis: rs396991-Phe/Phe (AA genotype): longer OS, PFS, and PFS in KRAS wt. | [31,60,68–86] |
| ANXA1                   | rs1050305      | BV+FOLFOX (161 discovery), BV+FOLFOXIRI (109 validation), BV+FOLFIRI (378 control) | 648      | Caucasian | Efficacy          | Moderate          | rs1050305-G allele: worse OS (validated), PFS, RR only in BV+FOLFOXIRI group. No significant association with PFS, OS, ORR in BV+FOLFIRI group. rs1010222-A allele: better PFS (not validated). No significant association with PFS, OS, ORR in BV+FOLFIRI group. | [87] |
|                          | rs1010222      | BV+FOLFOX (161 discovery), BV+FOLFOXIRI (109 validation), BV+FOLFIRI (378 control) | 648      | Caucasian | Efficacy          | Moderate          |                                                 |           |
| CALR                    | rs1049481      | BV+FOLFOX (161 discovery), BV+FOLFOXIRI (109 validation), BV+FOLFIRI (378 control) | 648      | Caucasian | Efficacy          | Moderate          | No significant association with PFS, OS, ORR. | [87] |
| ATG4B                   | rs35271226; rs1130910; rs7421; rs34691302 | OXA-based (188), IRI-based (137) | 325      | Chinese   | Efficacy          | Low               | No significant association with OS, PFS, DCR. | [88] |
| ATG16L1                 | rs6758317; rs2241878; rs7595748 | OXA-based (188), IRI-based (137) | 325      | Chinese   | Efficacy          | Low               | No significant association with OS, PFS, DCR. | [88] |
|                          | rs17094017     | OXA-based (188), IRI-based (137) | 325      | Chinese   | Efficacy          | Low               | rs17094017-T allele (additive model): Increased OS, PFS, DCR in overall population. rs17094017-T allele (additive model): Increased OS, PFS, DCR in OXA-based not in IRI-based group (stratifying for chemotherapy). | [88] |
| ATG2B                   | rs8019013      | OXA-based (188), IRI-based (137) | 325      | Chinese   | Efficacy          | Low               | rs8019013-T allele (additive model): Shorter PFS, prolonged DCR in univariate, not confirmed after FDR correction. | [88] |
|                          | rs12432561; rs10134160 | OXA-based (188), IRI-based (137) | 325      | Chinese   | Efficacy          | Low               | No significant association with OS, PFS, DCR. | [88] |
### Table 1. Cont.

| Genes   | rs Code/ Alias | Therapy (N Patients in Cohorts) | Patients | Ethnicity | Clinical Outcomes | Level of Evidence | Main Effect | Ref. |
|---------|----------------|---------------------------------|----------|-----------|-------------------|-------------------|-------------|------|
| ATG13   | rs13448        | FOLFIRI+BV or FOLFIRI+CTX       | 657      | Chinese   | Toxicity          | Moderate          | rs13448-any C: lower rate of grade 2–3 hypertension | [89] |
| FIP200  | rs1129660      | FOLFIRI+BV or FOLFIRI+CTX       | 657      | Chinese   | Toxicity          | Moderate          | rs1129660-any G: lower rate of grade 2 or 3 hypertension | [89] |
| ULK1    | rs9481         | FOLFIRI+BV or FOLFIRI+CTX       | 657      | Chinese   | Toxicity          | Moderate          | rs9481-A allele: lower rate of grade 2 or 3 hypertension | [89] |
| GABARAPL2 | rs11149841; rs6564267 | OXA-based (188), IRI-based (137) | 325      | Chinese   | Efficacy          | Low               | No significant association with OS, PFS, DCR. | [88] |
| WIP1    | rs11658979; rs11077558; rs2011143; rs2909207; rs883622; rs883620; rs35271226 | OXA-based (188), IRI-based (137) | 325      | Chinese   | Efficacy          | Low               | No significant association with OS, PFS, DCR. | [88] |

**Abbreviations:** CT: chemotherapy, BV: bevacizumab, CTX: cetuximab, PANI: panitumumab, FL: fluoropyrimidines, OXA: oxaliplatin, Cape: capecitabine; 5-FU: 5-fluoropyrimidine, IRI: irinotecan, SOX: oxaliplatin + 5-FU regimen, (m)OS: (median) overall survival; (m)PFS: (median) progression-free survival; (O)RR: (overall) response rate; ADCC: antibody-dependent cellular cytotoxicity; DCR: disease control rate; PD: progression; TS: tumor shrinkage; CR: complete response; LD: linkage disequilibrium; N: number of patients; Ref.: references. 3. Acute-phase cytokines and enzymes.
4. Discussion

4.1. Acute-phase Cytokines and Enzymes

Interleukin-1β (IL-1β), interleukin-6 (IL-6), and tumor necrosis factor-α (TNF-α) are pyrogenic cytokines with a role in the acute-phase response. Under hypoxic states, IL-6 promotes hypoxia-inducible factor 1α (HIF-1α, HIF1A) and signal transducer and activator of transcription-3 (STAT-3) transcription, which stimulates VEGF expression, blood vessel formation, and tumor growth through defective angiogenesis. In a total of 775 mCRC patients, 511 patients were treated with first-line FOLFIRI plus bevacizumab (223 in training and 288 in validation cohorts) and 264 with FOLFIRI plus cetuximab (control cohort), polymorphisms in the IL-6/STAT-3 pathway were investigated in the development, invasion, and spreading of mCRC due to their role in facilitating immune tolerance within the TME [27]. Patients carrying the IL6-rs2069837-G allele had shorter median progression-free survival (PFS) compared with the AA genotype both in the training and validation cohorts (Hazard Ratio (HR): 1.50; \( p = 0.033 \); HR: 1.34, \( p = 0.047 \), respectively) [27]. Moreover, patients with the IL6-rs2069837-AA genotype had higher tumor response (67% AA vs. 52% any G, \( p = 0.026 \)) only in the validation cohort. The IL6-rs2069837-G allele leads to increase IL-6 expression and was supposed that it serves as a surrogate for resistance towards anti-VEGF therapy in mCRC patients [27,90].

The inflammatory-related transcription factor, STAT-3, is hyper-activated in several human cancers, leading to proliferation, apoptosis, division, and differentiation of tumor cells. Usually, prolonged activation of the IL-6/STAT-3 signal may result in poor therapy outcomes as well as drug-related side effects via alterations of drug bioavailability [91]. Altered expression of STAT-3 deriving from polymorphic variants may predispose patients administered irinotecan-based chemotherapy to gastrointestinal (GI) epithelium damage and consequent side effects (i.e., mucositis, diarrhea). STAT3-rs1053004 polymorphism is located in the 3′UTR region and regulates the protein expression by altering its transcriptional activity [92]. In this regard, a discovery/replication study in 400 mCRC patients treated with first-line FOLFIRI, highlighted the protective role of the STAT3-rs1053004-C allele against grade 3-4 GI toxicity (OR: 0.51, \( p = 0.045 \); OR: 0.39, \( p = 0.043 \), respectively, in the discovery and replication groups) [29].

The prolonged inflammatory effects of IL-1β are counterbalanced by the action of the interleukin-1 receptor antagonist (IL-1RA) cytokine that competitively inhibits the binding of circulating IL-1β during an inflammatory event to quench acute inflammation and avoid tumor angiogenesis/metastasis. Two polymorphisms in the IL-1RA gene (rs4251961 and rs579543) influencing IL-1RA circulating levels may alter this mechanism. The highest IL-1RA production was reported in carriers of the homozygous IL-1RA-rs4251961-TT and IL-1RA-rs579543-TT genotypes. In 180 mCRC patients treated with second-line irinotecan or oxaliplatin-based regimens ± cetuximab, carriers of at least one T allele for these two polymorphisms had a favorable survival profile (risk ratio: 0.64, \( p = 0.018 \)) [93].

Prostaglandin-endoperoxide synthase 2 (PTGS2, also known as cyclooxygenase 2 [COX-2]) is the key enzyme in inflammatory prostaglandin biosynthesis in many cells, including colonic epithelial cells, and it is induced in response to inflammatory stimuli. Pro-inflammatory prostaglandin E2 (PGE2) is a potent inflammatory lipid mediator that is generated by COX-2 conversion of arachidonic acid, representing the main prostaglandin in CRC [94]. The alterations in multiple pathways, including PTGS2, EGFR, as well as RAS, are significantly involved in CRC development and progression. The cross-talk between pathways of inflammatory response/prostaglandins biosynthesis and EGFR signaling plays a key role in enhancing the growth and spread of tumor cells [94,95]. Particularly, during the inflammatory response, PTGS2 is involved in the processes of vasoconstriction, vasodilatation, platelet aggregation, and immunomodulation. Prostaglandins and other lipid mediators of inflammation are produced quickly by the macrophages and then their actions are followed by those of cytokines such as IL-6 and interleukin-10 (IL-10) with the consequent activation of the EGFR signal, promoting the proliferation of tumor cells [94,96]. Concerning response to anti-EGFR agents, PTGS2-rs5275 and PTGS2-rs20417
polymorphisms have raised the interest of researchers as potential predictor biomarkers of efficacy [31,33]. The rs20417 polymorphism is located in the 3′UTR region of the PTGS2 gene, and the minor C allele has been associated with a decrease in promoter activity, whereas the minor C allele of the rs5275 polymorphism in exon 10 decreases mRNA stability [95]. In 76 mCRC patients treated with XELOX chemotherapy, the PTGS2-rs5275-TT genotype was correlated with better PFS and OS (HR:0.47, \( p = 0.046 \), and HR:0.16, \( p = 0.013 \), respectively) [32]. Contrariwise, in the study by Lurje et al. involving 130 mCRC patients treated with cetuximab, PTGS2-rs20417-CC and PTGS2-rs5275-CC genotypes were associated with longer PFS in univariate analysis (relative risk (RR):0.31, \( p = 0.032 \) and RR:0.67, \( p = 0.0003 \), respectively). PTGS2-rs5275 maintained significance also in multivariate analysis (RR:0.53; \( p = 0.013 \)) [31]. Conflicting data also come from a large study of 815 Caucasian mCRC patients treated with cetuximab-based therapy in which the PTGS2-rs5275 polymorphism did not emerge as a predictive marker of efficacy for the therapy [33]. Further studies are certainly needed to clarify the actual role of these polymorphisms in the response to anti-EGFR treatment. The impact that such polymorphisms have on both gene expression and COX-2 activity may help to better explain the resistance to targeted therapies that sometimes occur in a TME in which the immune response to cancer cells appears to be impaired.

In addition, genes involved in the cyclic GMP–AMP synthase (cGAS)–stimulator of the interferon genes (STING) pathway have an important role in the early phase of immune response by inducing type-I interferon (IFN) [97]. In a case/control study of 451 Caucasian mCRC patients, 129 treated with FOLFIRI+cetuximab and the others with FOLFIRI+bevacizumab as controls, the genes in the cGAS/STING pathway were evaluated. In the FOLFIRI+cetuximab group, patients with STING-rs7380824-any T allele showed a lower ORR (any T vs. CC: 60.7% vs. 81.5%, \( p = 0.04 \)), while patients with STING-rs11311769-any T allele had a shorter OS (HR:2.98; \( p = 0.0085 \)) [34]. These polymorphisms are missense variants (rs7380824: R293Q; rs11311769: R232H) and their variants can determine a reduction in the level of IFN production and then in the innate immune response [98]. In the FOLFIRI-cetuximab group, another variant related to INF emerged as significant: the IFNB1-rs1051922 polymorphism that affects the IFN-β production due to a premature stop codon. The presence of IFNB1-rs1051922-GA and AA genotypes was correlated with shorter PFS (HR:2; \( p = 0.02 \); HR:2.19, \( p = 0.02 \), respectively) [34]. In the FOLFIRI+bevacizumab group, the cGAS-rs6907936 polymorphism emerged as significant; the presence of any G allele showed higher ORR (81.5% vs. 58.4%, \( p = 0.04 \)) [34]. The mechanism of action of the antiangiogenic drug bevacizumab seems less likely to directly impact the immune system by ADCC; however, an indirect effect on tumor vasculatures due to the STING/cGAS pathway could interfere with the bevacizumab antitumor activity. Nevertheless, more studies are needed to confirm this hypothesis.

### 4.2. Angiogenic Mediators

Inflammation can cause endothelium changes, known as endothelial cell activation, which allow it to participate in the inflammatory response [99,100]. The immune-related core changes of endothelial cell activation include cytokine production, expression of leukocyte adhesion molecules, and up-regulation of human leukocyte antigen (HLA) molecules, then determining neutrophil recruitment [99].

The C-X-C motif chemokine ligand 12 (CXCL12)/C-X-C chemokine receptor type 4 (CXCR4) axis, mainly activated by VEGF and hypoxic stress typical of inflammation, contributes to the vessels formation, promoting angiogenesis and metastasis [101]. Blocking CXCR4 in the signal cascade leads to a reduction in tumor endothelium with a mechanism that is VEGF-independent [102]. Tumor-associated macrophages (TAMs) with the M2-like phenotype are related to tumor cell growth and spreading. In CRC, CXCL12/CXCR4 axis activation stimulates the up-regulation of miRNAs on TAMs, leading to enhanced immunosuppression, epithelial to mesenchymal transition (EMT), as well as angiogenesis via VEGF and EGF secretion [103].
colleagues, including a total of 874 Caucasian and Asian mCRC patients receiving first-line chemotherapy with bevacizumab, found that the CXCR4-rs2228014-CC genotype showed a longer PFS compared with CT/TT genotypes in multivariate analysis (training cohort, HR:1.77, \( p = 0.029 \); validation cohort B, HR:1.87, \( p = 0.009 \)). These results remained significant after adjusting by primary tumor location for patients with left-sided mCRC [35].

TAMs-related functions are also regulated by several genes, including C-C motif chemokine ligand 2 (CCL2), C-C motif chemokine ligand 18 (CCL18), TANK-binding kinase 1 (TBK1), and IFN regulatory factor 3 (IRF3), whose signaling pathway is activated during tumor angiogenesis. In particular, TBK1 is triggered under hypoxic conditions and in turn activates IRF3 and NF-kB-dependent factors associated with inflammation [104]. TBK1 and IRF3 polymorphisms were investigated for their effects on tumor growth and vascularization and thus on response to bevacizumab therapy. The discovery-validation study by Sunakawa et al. on mCRC patients evidenced that carriers of the TBK1-rs7486100-T allele had shorter PFS than those with the AA genotype (HR:1.46, adjusted \( p = 0.028 \)) in the wild-type KRAS validation set. Moreover, carriers of the IRF3-rs2304205-C allele experienced better RR compared to the AA genotype (79% vs. 54%, adjusted \( p = 0.0056 \)) limited to the wild-type KRAS discovery cohort. The authors also highlighted that in the mutant KRAS discovery set, mCRC patients carrying CCL2-rs4586-C and IRF3-rs2304205-C alleles had longer PFS (HR:0.51, adjusted-\( p = 0.026 \), and HR:0.56, adjusted-\( p = 0.039 \) in the dominant model, respectively) compared to other genotypes in multivariable analysis. The CCL18-rs14304- TT genotype was associated with worse PFS compared to the CC genotype (HR:1.24, adjusted-\( p = 0.007 \)) [36].

Reduced oxygen tension is crucial for tumor vascularization and regulates VEGF production. The modulation of both VEGF expression and tumor angiogenesis is also driven by molecules belonging to HIF1α and insulin-like growth factor 1 (IGF1)/insulin-like growth factor 1 receptor (IGF1R) signaling [105,106]. The role of polymorphisms in IGF1R was here reported because it was suggested that a decreased IGF1R expression may also affect CD4+ Th cell lineage commitment [107]. The response to bevacizumab-based therapy may be thus influenced by variants in HIF1α, IGF1, and IGF1R genes. A study conducted on 132 mCRC patients administered bevacizumab in association with fluoropyrimidines and oxaliplatin, evidenced that patients carrying the HIF1α-rs11549465-T allele had longer PFS (HR:0.55, \( p = 0.038 \)) compared to the CC genotype in univariate analysis [28]. Moreover, carriers of a combination of HIF1α-rs11549465-CC, VEGF-rs699947-A, and EGF-rs2227983-GG experienced shorter PFS (HR:2.66, \( p < 0.001 \)). The IGF1-rs6220-G allele was associated with better overall survival (OS) compared to the AA genotype in multivariate analysis (HR:0.60, \( p = 0.046 \)) [28]. In the FIRE 3 trial, 614 mCRC Caucasian patients receiving fluoropyrimidines and irinotecan-based chemotherapy plus bevacizumab or cetuximab were assessed for IGF and insulin receptor substrate (IRS) 1 and 2 isoforms. From multivariate analysis emerged that carriers of the IGF1-rs2946834-T allele had longer PFS (HR:0.78, \( p = 0.010 \)) and OS (HR:0.65, \( p = 0.003 \)) compared to carriers of the CC genotype. In the subgroup of FOLFIRI plus bevacizumab patients, IGF1-rs2946834-T allele carriers showed only a trend of longer PFS in the multivariable model (HR:0.77, \( p = 0.074 \)) [38]. In the whole population analysis, the IRS1-rs1801123-C allele was correlated independent from KRAS status with shorter OS (HR:1.28, \( p = 0.054 \)) than the TT genotype in multivariate analysis, resulting more a prognostic than a predictive biomarker of therapy response [38].

In 132 mCRC patients treated with first-line FOLFOX or XELOX and bevacizumab, the IGF1-rs6214-G allele, associated with increased VEGF mRNA expression, showed increased OS (HR:0.51, \( p = 0.005 \)) and remained significant in multivariate analysis (HR:0.60, adjusted-\( p = 0.046 \)) [28]. Several studies have also suggested that the IGF1 pathway is a key mediator of resistance to cytotoxic chemotherapy and anti-EGFR treatment [108–111]. An analysis of 130 formalin-fixed, paraffin-embedded (FFPE) samples from patients with mCRC treated with a cetuximab-based regimen showed that polymorphisms in the IGF pathway were significantly associated with PFS and/or OS. In particular, IGF1-rs6214-TT was associated to worse PFS (HR:2.239, \( p = 0.008 \)) and OS (HR:2.282, \( p = 0.011 \) in multi-
variate analysis), the \( IGF1R\text{-rs2016347-G} \) allele was associated with shorter OS (HR:1.734, \( p = 0.033 \) in multivariate analysis), while the \( IGF1\text{-rs2946834-G} \) allele was associated to shorter PFS (HR:2.943, \( p = 0.002 \) in multivariate analysis) [39].

Endothelial NOS (eNOS, also known as NOS3), constitutively expressed at the level of the endothelium, plays a key role in endothelial health during angiogenesis, as well as in the regulation of immune activation. ENOS is a gene involved in the production of nitric oxide (NO) and is therefore essential for hemodynamic maintenance of blood flow due to its ability to regulate blood vessel permeability. Production of NO by macrophages, dendritic cells, and NK increases levels of VEGF, which phosphorylates eNOS by binding to endothelial cells, stimulating cell production and thus the growth of new blood vessels [112]. Two immune pathways are involved in this process: HIF-1\( \alpha \) induction and PGE2 production. HIF-1\( \alpha \) is regulated by NO, but also by cytokines and growth factors. HIF-1\( \alpha \), as a cellular hypoxia sensor, induces genes that promote vasodilation, vascular permeability, and angiogenesis. Regulation of NO is also essential for tissue repair. Specifically, NO activates metalloproteinases (MMPs) that remodel the cellular matrix for neovascularization through proteolytic activity [112,113]. Variants in ENOS gene reduce the enzyme activity and/or expression and thus the NO levels may enhance the response to anti-angiogenic-based treatments. Ulivi et al. assessed 237 Caucasian mCRC patients and found that heterozygous carriers of \( ENOS\text{-rs1799983-GT} \) variant had shorter PFS (HR:1.70, \( p = 0.013 \)), OS (HR:1.80, \( p = 0.014 \)), and overall response rate (ORR) (42.5\% vs. 63.1\%, \( p = 0.030 \)) compared to GG and TT carriers [40]. Moreover, patients with the \( ENOS\text{-rs2070744-TT} \) genotype had both shorter PFS (HR:3.63, \( p = 0.036 \)) and OS (HR:5.48, \( p = 0.007 \)) than CT/CC carriers. Another polymorphism reducing the protein expression is the intronic variable number tandem repeat (VNTR) of 27 nucleotides in the \( ENOS \) gene. Patients carrying \( ENOS\text{-VNTR-4bb} \) (homozygous for five repeats) showed both longer PFS (HR:0.63, \( p = 0.034 \)) and OS (HR:0.54, \( p = 0.015 \)) than patients carrying at least four repeats (\( VNTR\text{-4ba/4aa} \)). Combining haplotype (i.e., Haplo 1: \( VNTR\text{-b/rs1799983-G} \) and Haplo 2: \( VNTR\text{-b/rs1799983-T} \) ENOS Haplo1/Haplo1 and Haplo2/Haplo2 showed an improved PFS (15.0 vs. 9.1 months, \( p = 0.001 \)), OS (34.5 vs. 20.5 months, \( p = 0.002 \)), and ORR (71\% vs. 45.9\%, \( p = 0.013 \)) compared to other ENOS haplotype combinations [40].

\( ENOS\text{-rs1799983} \) was investigated in 232 mutant KRAS Caucasian mCRC patients administered chemotherapy alone (n = 112) or chemotherapy plus bevacizumab (n = 120). In the bevacizumab group, patients carrying the \( ENOS\text{-rs1799983-TT} \) genotype, leading to a reduced NO production, had higher severe toxicity, particular grade 3-4 hypertension and proteinuria (GG+GT: 8\% vs. TT: 50\%; \( p = 0.0002 \)) [41].

Another gene involved in the production of NO is \( NOS2 \) coding for iNOS (also known as nitric-oxide synthase 2 [NOS2]), an NADPH-dependent enzyme known as a surrogate marker of M1 macrophage activation. In 227 mCRC patients receiving FOLFIRI plus bevacizumab, patients with CCTTT any >13 repeats showed improved median PFS compared with patients carrying the \( \leq 13/\leq 13 \) repeats variant (HR:0.64, \( p = 0.010 \)). Similar results were shown adopting the \( >26\text{repeats}/\leq 26 \) repeats (HR:0.56, \( p = 0.005 \)). These data were partially confirmed in 231 patients receiving FOLFOXIRI plus bevacizumab: a better median PFS was observed in patients with >26 vs. \( \leq 26 \) repeats. However, these data were not confirmed in the two validation cohorts of 301 and 187 patients [42]. These results seem to suggest that the \( ENOS \) gene has a more important role in maintaining endothelial cell functional integrity.

The reactive oxygen species (ROS) could be directly scavenged by different antioxidant molecules, including superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), and cytosolic and mitochondrial thioredoxins (TXN1 and TXN2). A study of 236 mCRC Caucasians, 129 patients treated with first-line FOLFIRI+cetuximab and 107 with FOLFIRI+bevacizumab, analyzed the effects of polymorphisms in genes involved in the antioxidant pathway [43]. In the FOLFIRI+cetuximab group, they found that the patients’ carrier \( TXN2\text{-rs4821494-G} \) allele and \( GPX4\text{-rs4807542-A} \) allele had worse OS, confirmed by multivariate analysis (HR:2.47, \( p = 0.03 \); HR:2.24, \( p = 0.05 \)).
polymorphism is located in the 5′UTR and the T mutant allele was related to a lower TXN2 expression in sigmoid and transverse colon tissue. The role of ROS is pleiotropic and it includes the inflammasome activation critical for an efficient immune response [114].

4.3. Nuclear Receptors

Nuclear receptors (NRs) are key sensors in regulating gene expression of proteins involved in drug transport and metabolism and in orchestrating the activation of the immune system triggered by xenobiotics themselves [11,12,115,116]. Particularly, the pregnane X receptor (PXR, NR1I2) is a major regulator of both xenobiotic detoxification and innate immunity to xenobiotics. PXR ligands stimulate the expression of NLR family pyrin domain containing 3 (NLRP3), nucleotide-binding oligomerization domain 1 (NOD1), and Toll-like receptor (TLR)-2, TLR-4, and TLR-9 in endothelial cells by causing activation of the inflammasome and simultaneous activation of PXR and innate immunity [117]. Activation of PXR, in turn, stimulates several phase 1 (e.g., cytochromes [CYPs]) and phase 2 enzymes (e.g., glutathione S-transferase [GSTs], sulfotransferase [SULTs], UDP-glucuronosyltransferase [UGT1As]), as well as the membrane ATP-binding cassette (ABC)/solute carrier (SLC) transporters that have a crucial role in the modulation of ADME of chemotherapeutics [11,12,115]. Overexpression of the PXR in vivo and in vitro models has been correlated with irinotecan resistance due to UGT1A induction [118]. By contrast, the active metabolite of irinotecan (SN-38) has been associated with PXR induction, improving irinotecan metabolism [119]. A study involving 109 advanced CRC patients receiving 180 mg/m² irinotecan treatment highlighted a correlation between severe hematological toxicity and NR1I2-rs10934498-A (OR:0.17, \( p = 0.009 \)), NR1I2-rs2472677-G (OR:41.55, \( p = 0.003 \)), NR1I2-rs3814055-T (OR:9.25, \( p = 0.009 \)), and NR1I2-rs1523127-C (OR:7.23, \( p = 0.009 \)) alleles. In addition, NR1I2-rs2472677-G carriers experienced a higher grade of any type of toxicity than other genotypes (OR:6.78, \( p = 0.031 \)) [44]. In a discovery/replication study involving about 400 mCRC patients treated with irinotecan-based therapy, those carrying the NR1I2-rs1054190-TT genotype had significant short OS (HR = 6.78, \( p = 0.0021 \), and HR = 3.56, \( p = 0.0414 \) in the discovery and replication sets, respectively) compared to carriers of the C ancestral allele [45].

The vitamin D receptor (VDR), another member of NRs, is a mediator of the genomic actions of 1,25-dihydroxycholecalciferol and, more recently, emerged as an immunomodulatory player. Polymorphisms of VDR and vitamin D3 (VitD3) serum levels have been correlated to (auto)immune-diseases development since VDR is expressed in several immune cell types [120]. The VitD3/VDR axis shows regulatory activity in the adaptive immune response, suggesting a possible influence of the nuclear receptor and its polymorphisms on tumor cell growth, inflammation, invasion, metastasis, and angiogenesis [121]. In addition, this protein regulates calcium and phosphate homeostasis as well as the transcription of genes affecting the metabolism of drugs (i.e., CYPs, UGT1As, ABC/SLC transporters) [45]. The study by De Mattia et al. highlighted an association of the VDR-rs7299460-T allele with longer OS (HR:0.61, \( p = 0.0076 \); HR:0.57, \( p = 0.0478 \) in discovery and replication sets, respectively). Additionally, a study involving 250 mCRC patients suggests that VDR-rs11574077 polymorphism is correlated with irinotecan-related toxicity. The pharmacokinetic analysis evidenced that the VDR-rs11574077-G minor allele affects the efficacy of irinotecan glucuronidation and detoxification. VDR-rs11574077-G carriers concordantly experienced higher grade gastrointestinal toxicity although limited to the discovery set (OR:4.46, \( p = 0.010 \)) [29].

4.4. Toll-Like Receptors (TLRs)

The TME and immune responses not only play a key role in tumor growth and progression but could also influence the efficacy and toxicity of cancer therapies. TLRs, together with NOD-like receptors (NLRs), belong to pattern recognition receptors (PRRs) and can recognize a variety of pathogen-associated molecular patterns (PAMPs). In particular, TLRs expressed on numerous immune cells (e.g., macrophages, dendritic cells, NK cells, and T and B lymphocytes) and non-immune cells (e.g., endothelial cells, fi-
broblasts) are sensors of the innate immune response, and some members induce the production of type I interferons (IFN-α and IFN-β), which further stimulates the immune response [122]. TLRs are membrane-receptors located on the cell surface or endosomes with different specificities [123,124].

TLR-4, a bacterial sensor whose activation stimulates NF-κB signaling and inflammatory cytokine production, has been associated with the risk of developing gastrointestinal toxicity during irinotecan treatment, likely related to intestinal mucosal damage and associated infections [125]. In addition, irinotecan promotes activation of the innate immune response through direct binding to the TLR-4/MD-2 complex and stimulates the inflammatory response [126]. This mechanism appears to be confirmed in a preliminary, albeit small, study of patients with advanced CRC treated with irinotecan. Analysis showed that carriers of TLR-4 rs4986790-G and TLR-4 rs4986791-T minor alleles had increased IL-6 plasma levels and a more frequent occurrence of severe and late-onset diarrhea [49].

A study of 5000 patients with CRC, in which 40% had advanced disease, investigated the polymorphisms FPR1 rs867228, TLR-3 rs3775291, and TLR-4 rs4986790, and in other proteins of the PRRs family, without finding any significant association [48].

TLR-7 polymorphisms have been investigated in cancer immunotherapy due to the ability of this PRR to induce a robust release of anti-cancer cytokines such as IL-12. A study by Okazaki et al. found TLR-7 rs3853839 to be a potential independent biomarker of PFS in patients treated with anti-EGFR therapy. Particularly, the multivariable model showed that patients carrying the TLR-7 rs3853839-GG genotype had longer PFS compared to those carrying GC/CC genotypes (HR: 2.02, \( p = 0.015 \)) [50].

As abovementioned, TLRs are also expressed on non-immune cells and, therefore, they also play a role in modulating the functions of endothelial cells via mitogen-activated protein kinase kinase kinase 7 (MAP3K7 or TAK1) signaling that controls cell viability and the inflammation response [127,128]. TLR-1 and TLR-6, both of which dimerize with TLR-2, play critical roles in the intestinal mucosal immune response. Genetic variants in these PRRs and their common downstream signaling molecule TAK1 could lead to interindividual differences in drug response in mCRC patients [129]. In a study of 535 mCRC patients treated with FOLFIRI and bevacizumab, patients with the TLR-1 rs5743618-TT genotype had a worse RR than GT/GG genotypes (discovery set: 43% vs. 62%, \( p = 0.025 \); validation set: 46% vs. 65%, \( p = 0.021 \)). Patients with the TLR-6 rs5743818-AA genotype had a lower response rate (RR) and shorter PFS than AC/CC carriers, limited to univariate analysis. In the discovery arm, patients with the TLR-1 rs5743618-TT genotype had shorter PFS and OS compared with GT/TT genotypes (HR: 1.50, \( p = 0.046 \); HR: 1.53, \( p = 0.025 \), respectively). In the same group, patients with the TAK1 rs1145727-AA genotype had a shorter OS than patients with GA/GG genotypes (23 vs. 26.8 months, \( p = 0.008 \)) [47]. The TLR-1 rs5743618 polymorphism is a missense (Ser602Ile) variation in which the presence of the mutant G allele is associated with a decrease in IL-6 and TNF-α levels and therefore with a reduction in TLR-1-mediated immunity [130]. This may explain the altered pro-tumorigenic activity linked to the worse response of patients carrying the TLR-1 rs5743618-T allele.

### 4.5. Cytokines and Chemokines in the Early Phase of Immune Response

Other pro-inflammatory cytokines, such as interleukin-8 (IL-8) and interleukin-12 (IL-12), play important roles as chemo-attractants for innate immune cells such as neutrophils, basophils, dendritic cells, eosinophils, Langerhans cells, mast cells, monocytes, macrophages, neutrophils, NK and T cells at the site of inflammation. IL-8, encoded by CXCL8, plays a role not only in altering vascular permeability by promoting neutrophil recruitment and migration but also in their activation, triggering the respiratory burst, thereby ROS and NO. Moreover, IL-8 has a pivotal role in the angiogenesis, proliferation, and migration of tumor cells as well as TME modulation [131,132]. Polymorphisms in CXCL8 may have a role in the modulation of the abovementioned functions, possibly influencing the response to chemotherapy in the mCRC setting. Mutant RAS mCRC patients carrying the CXCL8 rs4073-TT genotype showed better PFS (HR: 0.53, \( p = 0.002 \)) and OS.
(HR:0.64, p = 0.03) after bevacizumab-based therapy compared to those carrying TA/AA genotypes but only PFS retained significance in multivariate analysis (HR:1.8, p = 0.0006). This interindividual variability in response to treatment was supported by different IL-8 serum levels: CXCL8-rs4073-TT genotype carriers had lower cytokine serum levels [41]. Another study involving 125 metastatic or relapsed CRC patients treated with first-line cytotoxic chemotherapy combined with bevacizumab correlated the CXCL8-rs4073-T allele to a tendency for longer PFS, whereas it was significantly associated with better ORR compared to the AA genotype (OR:0.394, p = 0.067) [51]. In another study with 108 KRAS wild-type patients treated with bevacizumab-based therapy, the presence of the CXCL8-rs4073-Any A allele (HR:1.51, p = 0.037) and the TBK1-rs7486100-TT genotype (HR:1.94, p = 0.037) were associated with worse PFS [52]. The prognostic effect of the CXCL8-rs4073 variant is independent to the KRAS status but seems related to the interaction with bevacizumab-containing regimens. The role of this variant was also investigated in another three studies in patients with mCRC treated with cetuximab-based or oxaliplatin-based therapies or regorafenib; however, no significant associations with response, survival, and skin rash toxicity were found [31,53,54].

A study on 132 mCRC Caucasian patients administered first-line bevacizumab/oxaliplatin-based therapy investigated IL-8 receptors, CXCR1 and CXCR2. From the analysis emerged an association between the CXCR1-rs2234671-GG genotype and good RR (71% GG vs. 37% GC vs. 17% CC, p < 0.001). The CXCR2-rs2230054-CC genotype showed worse RR (62% TT vs. 44% TC vs. 21% CC, p = 0.008), retaining significance after multiple testing [28]. In Asian patients, the CXCR2-rs2230054 variant did not show any significant relation with bevacizumab-related efficacy outcome as well as the IL10-rs1800896, which was another biomarker with a role in the angiogenic pathway [51]. In 105 mCRC patients mainly Caucasians treated with 5-FU or oxaliplatin-based regimens without bevacizumab, the presence of CXCR1-rs2234671-GC compared to the GG genotype showed worse TTP while no association was reported for CXCR2-rs2230054 [53]. The CXCR1-rs2234671 was also included in a comprehensive pharmacogenetic profiling of biomarkers belonging to the EGFR pathway and its relation with response and toxicity in patients treated with cetuximab-based regimens was evaluated without finding significant association [33].

Finally, other pro-inflammatory cytokines, the IL-17A and IL-17F, are mainly produced by Th17 (T-helper cell type 17) cells in adaptive immunity, but they are also secreted by other immune cells including NK, NKT, neutrophils, and intestinal Paneth cells, highlighting their role also in the innate response [133]. Moreover, IL-17 was investigated for its involvement in several other pathways [134]. For example, a dysregulated IL-17A and IL-17F production can impair myeloid cell recruitment and determine an excessive pro-inflammatory cytokine expression, and consequently chronic inflammation, which leads to cancer development. A study in 122 Caucasians mCRC treated with bevacizumab-based therapy evaluated the predictive role of IL17A-rs2275913 and IL17F-rs763780 variants, however, no significant association with efficacy outcomes was found. Only the baseline serum IL-17A concentration was significantly associated with the response to bevacizumab [56].

4.6. Tumor Immune Escape Factors

Antigen presentation is a crucial step in the cell-mediated mechanisms essential for the activation of the immune system. Tumor cells could up-regulate the immune checkpoint and their ligands, inhibiting activity or inducing apoptosis of the immune cells (tumor immune escape mechanism). Programmed cell death ligand 1 (PD-L1, CD274), cytotoxic T lymphocyte-associated protein 4 (CTLA-4), Cluster of Differentiation 47 (CD47), and some other innate immune checkpoints have been investigated by Volz et al. in a total of 924 mCRC patients [57].

In the training cohort (cetuximab-based), the CD274-rs2297137-G allele was associated with worse tumor response (56% AA vs. 19% GA vs. 16% GG; p = 0.029), the CTLA4-rs231777-CT genotype with shorter PFS (multivariate HR: 1.76; p = 0.019), and the CD24-rs52812045-AA genotype with shorter median PFS and OS (multivariate PFS HR:3.18,
The association of the CD24-rs52812045-AA genotype with shorter PFS was validated in 74 patients treated with oxaliplatin plus cetuximab (multivariate HR:2.12; *p* = 0.018). The CD274-rs2297137-A allele was also associated with prolonged OS in multivariate analysis in the additional control cohort 2 (FOLFIRI plus bevacizumab) (HR:0.65; *p* = 0.021) [57]. Moreover, in 141 mCRC Chinese patients treated with first-line bevacizumab-based regimens, the CD274-rs2297136-AA genotype was associated with better PFS (HR:1.68; *p* = 0.018) and OS (HR:1.88; *p* = 0.012) in the whole population and also in the KRAS mutant subgroup [58]. The binding of CD24, present on tumor cells, to Siglec-10, expressed on immune cells, causes inhibition of the immune response mediated by the tyrosine phosphatases SHP-1 and SHP-2. SHP-1/2 phosphorylate the immunoreceptor tyrosine-based inhibitory motif (ITIM) at the cytoplasmic end of Siglec-10, blocking TLR-mediated inflammation and immune response and promoting tumor cell escape from the immune system [135]. HIF1-α appears to modulate human leukocyte antigen-G (HLA-G) expression, a tolerogenic molecule proposed as an immune checkpoint [136,137], and the HIF1A-rs2244608 polymorphism has been associated with the efficacy of irinotecan-based CRC therapy. In particular, patients carrying the HNF1A-rs2244608-G allele had better PFS than patients with the AA genotype (HR:0.72; *p* = 0.002) [62]. HLA-G has been studied as an independent marker of efficacy in CRC [138–140], and its polymorphisms deserve further investigation in mCRC [140].

The regulation of the killing function of NK and T cells is mediated by a family of type I transmembrane glycoproteins called killer-cell immunoglobulin-like receptors (KIRs) that interact with MHC class I. NK cells are able to discriminate MHC class I in transformed and virus-infected cells, ignoring potential targets, expressing normal levels of autologous MHC class I, and leading to inhibition of antiviral and cytotoxic activities [141]. Genetic variants in the KIRs and their ligands, HLA class I (HLA-A, -B, -C), were investigated in 224 mCRC patients receiving FOLFIRI [58]. KIR3DL1/HLA-Bw4-I80 combination emerged as a positive predictive marker of survival (HR for T80:2.8, *p* < 0.001 and HR for no functional KIR/HLA interaction:1.8, *p* = 0.007) and the presence of the KIR2DL5A, 2DS5, 2DS1, 3DS1, and KIR3DS1/HLA-Bw4-I80 combination was associated with an increased rate of complete response (ORs ranging from 2.1 to 4.3) [58]. Most KIRs have an inhibitory effect on immune cells, while some, such as KIR2DS4, can activate immune cells. KIR2DS4 encodes both a full-length protein (KIR2DS4f) and a truncated protein (KIR2DS4d) due to a deletion of 22 base pairs in exon 5. This truncated protein cannot attach to the cytoplasmic membrane and is therefore considered a non-functional receptor. In a small group of 70 mCRC with mutant KRAS and FCGR2A-His131 patients treated with cetuximab, a significantly longer OS was observed in patients with the KIR2DS4d/4d genotype compared with individuals carrying at least a functional receptor (KIR2DS4f) (HR:2.27, *p* = 0.026) [60].

Adenosine, which plays an immunosuppressive and angiogenic modulator role in the TME, is produced by the ectodinucleotidase enzymes, CD39 and CD73, which are expressed in the surface membrane of tumor cells, but also in B cells or regulatory T cells [142,143]. Bevacizumab-induced hypoxia stimulates the up-regulation of HIF1-α and consequently of CD39 and CD73 in cancer cells, leading to suppression of cytotoxic T lymphocytes (CTLs) and NK cells. Recently, a large study of 322 Caucasian patients with mCRC treated with FOLFIRI plus bevacizumab demonstrated that in the discovery cohort, patients with both CD39-s11188513-C allele and CD73-rs2229523 GG genotype had a shorter OS in multivariable analysis (HR:2.10, *p* = 0.031; HR:0.49, *p* = 0.026, respectively), and was also confirmed in the validation cohort (HR:1.53, *p* = 0.013; HR:0.62, *p* = 0.024, respectively) [61]. Interestingly, these results were not replicated in the control arm in which patients received cetuximab-based chemotherapy, suggesting that the adenosine pathway may be a promising marker of specific bevacizumab response [61]. Moreover, extracellular adenosine exerts its immunosuppressive effect on tumor cells via A2BR, one of its four receptors, promoting tumor growth and metastasis. Carriers of the A2BR-rs2015353-TT genotype had a favorable OS compared with patients with at least one minor C allele in the multivariable model (HR: 0.24, *p* = 0.004) [61].
4.7. Cytokines in the Late Phase of Immune Response

The main cytokines produced by T cells are known as interleukins (ILs). IL-15, a type I cytokine together with IL-2, IL-4, IL-7, IL-9, and IL-21, promotes survival of T, B, and NK cells by binding to IL-15\(\alpha\), encoded by \(\text{IL15RA}\), negatively regulating both carcinogenesis and tumor growth [144]. IL-2, IL-4, IL-7, IL-9, and IL-15 receptors have a common gamma (\(\gamma\)c) chain, also referred to as CD132. Besides CD132, IL-15\(\alpha\) shares a \(\beta\) chain (IL-2/15R\(\beta\) or CD122) with IL-2, a cytokine structurally related to IL-15. The binding of IL-15 to CD122 stimulates the phosphorylation of Jak3 and subsequent activation of STAT5. The Jak3/STAT5 pathway induces cell survival of T, NK, and NK-T lymphocytes and promotes the differentiation of NK lymphocytes by increasing the expression of anti-apoptotic proteins such as Bcl2 and Bcl-x [145]. The overproduction of soluble IL-15/IL-15R\(\alpha\) could represent a novel mechanism of immune escape [146]. Furthermore, IL-15, through the balance between IL-2 and IL-15, stimulates survival for CD8 memory T cells [147]. The main cytokine produced by the CD8 T cells is the IFN\(\gamma\), which mainly increases the activation of MHC class I and II on macrophages. Variants in the INFG receptor gene (\(\text{INFGR1}\)) have been investigated for their predictive role in 141 Chinese mCRC treated patients with bevacizumab-based regimens. In particular, patients with the \(\text{INFGR1}-\text{rs}2234711\)-G allele and \(\text{INFGR1}-\text{rs}9376267\)-T allele showed longer OS only in univariate analysis (\(p=0.041\); \(p=0.0312\)) [58].

The TGF-\(\beta\)/SMAD-3 pathway is involved in chemoresistance modulating the TME: TGF-\(\beta\) through SMAD3 inhibits CD4 T cell proliferation and effectors function, silencing the IL-2 expression [148]. In a recent study involving 335 mCRC patients who received first-line FOLFIRI, the \(\text{IL15RA-}\text{rs}7910212\)-C allele was associated with short OS (HR:1.57, \(p=0.0327\); HR:1.71, \(p=0.0411\) in discovery and replication sets, respectively) according to the dominant model. In the same analysis groups, the \(\text{SMAD3-}\text{rs}7179840\)-C allele was instead associated with longer OS (HR:0.65, \(p=0.0202\); HR:0.61, \(p=0.0216\) in discovery and replication sets, respectively) in the dominant model [46].

Other pathways involved in the immune system are C-C motif chemokine ligand 5 (CCL5)/C-C chemokine receptor type 5 (CCR5) axis, C-C motif chemokine ligand 3 (CCL3), and C-C motif chemokine ligand 4 (CCL4), which stimulate the dendritic cells to produce IL-12, induce migration of endothelial progenitor cells, leading to increased VEGFA expression. A study comprised two independent sets of mCRC patients (79 Asian and 150 Caucasian patients for discovery and validation set, respectively) treated with regorafenib as monotherapy [63], indicated that patients carrying the \(\text{CCL4-}\text{rs}1634517\)-A allele have significantly shorter PFS than those carrying the CC genotype (discovery set: HR:1.58, adjusted-\(p=0.058\); validation set: HR:1.59, adjusted-\(p=0.012\)). In addition, patients carrying the CCL3-\text{rs}1130371-A allele were suggested to have a shorter PFS than those carrying GG (discovery set: HR:1.48, adjusted-\(p=0.064\); validation set: HR:1.50, adjusted-\(p=0.027\)). The \(\text{CCL4-}\text{rs}1634517\)-A and CCL3-\text{rs}1130371-A alleles resulted also in predictors of shorter OS in the validation set (HR:1.46, adjusted-\(p=0.041\), and HR:1.44, adjusted-\(p=0.047\), respectively) [63]. Interestingly, in a later analysis of the same cohorts, Suenaga et al. correlated polymorphisms in the CCL5/CCR5 pathway with serum levels of the CCR5 circulating protein and its ligands. From the study emerged that patients carrying the \(\text{CCL5-}\text{rs}2280789\)-G allele had higher CCL3 but lower CCL4 serum levels compared to AA carriers (72.7% vs. 23.1%, \(p=0.006\) and 31.8% vs. 69.2%, \(p=0.043\), respectively) [65]. Considering that the levels of these markers (increased CCL3 and decreased CCL4) are related to a good response to regorafenib, their association with \(\text{CCL5-}\text{rs}2280789\) polymorphism suggests a mechanism of action in the CCR5 network beyond the CCL5/VEGFA pathway.

The role of polymorphism in two enterocyte-specific genes, \(\text{MS4A12}\) and its transcriptional activator \(\text{CDX2}\), in predicting the efficacy of oxaliplatin was also investigated in a total of 604 mCRC patients who were divided into a discovery cohort (146, FOLFOX +/− bevacizumab), a validation cohort (230, FOLFOXIRI plus bevacizumab), and a control cohort (228, FOLFIRI plus bevacizumab) [67]. Enterocytes play a role in the absorption of small molecules from the lumen and in intestinal immunity by secreting antimicro-
bial proteins and cytokines and acting as antigen-presenting cells (APCs) for intestinal T lymphocytes. The *MS4A12*-rs4939378-G allele was correlated to longer PFS than the AA genotype in multivariate analysis (HR:0.65, \( p = 0.035 \)) in patients expressing the wild-type KRAS. Only in the mutant KRAS subgroup, patients with the *CDX2*-rs3812863-GG variant showed longer PFS than those with any A allele in univariate analysis (HR:0.39, \( p = 0.004 \)), pointing out the prognostic role of these biomarkers [67].

4.8. Regulated Cell Death Factors

Regulated cell death is a form of cell death due to signal transduction that can be pharmacologically or genetically modulated. The cytotoxic T cells induce the apoptosis of their targets both through the release of cytotoxic granules and also contribute to the immune homeostasis maintenance through the Fas/FasL interaction that leads to activation-induced cell death. This last form of apoptosis is induced by repeated T cell receptor (TCR) stimulation, responsible for the peripheral deletion of activated T cells through the caspases [149].

Alterations in genes involved in homeostasis maintenance through the apoptosis mechanism are investigated in mCRC. However, in 76 mCRC patients treated with XELOX chemotherapy, no association with response and survival were found for polymorphisms in *Caspase 3*, *TP53*, *BCL2L*, *TNFRSF10B*, *AKT1*, *BID*, *RIPK1*, *FAS*, and *FASL* genes [32]. Moreover, the autophagy-related gene *ATG2B* was investigated for its prognostic role in 325 Chinese patients treated with oxaliplatin-based or irinotecan-based chemotherapy. The *ATG2B*-rs17094017-T allele was associated with an increased OS (HR:0.65, \( p = 0.002 \)), PFS (HR:0.76, \( p = 0.007 \)), and DCR (OR:0.60, \( p = 0.03 \)) in the overall population, though stratifying by the chemotherapy used, these effects were confirmed only for patients treated with oxaliplatin-based regimen (OS, HR:0.64, \( p = 0.0219 \); PFS, HR:0.72, \( p = 0.0215 \); DCR, OR:0.39, \( p = 0.126 \)) [88]. Further investigation should be done to better identify the functional role of this polymorphism on the ATG2B expression and the exact mechanism underlying its influence on the immune system.

Some chemotherapeutic agents, including oxaliplatin, are capable of triggering a process known as immunogenic cell death (ICD) [150]. Oxaliplatin-based treatments lead to the production of DAMPs in ICD by stimulating the release of calreticulin (CALR), annexin A1 (ANXA1), and high-mobility group box 1 (HMGB1). These factors are recognized by PRRs, including LDL protein-related receptor 1 (LRP1) and purinergic receptor P2X 7 (P2RX7) [150]. In a study of 648 mCRC patients with discovery/validation design treated with an oxaliplatin-containing regimen (161 FOLFOX/bevacizumab in the discovery cohort, 109 FOLFOXIRI/bevacizumab in validation cohort; and 378 FOLFIRI/bevacizumab in two control cohorts), both the *ANXA1*-rs1050305-AA and *LRP1*-rs1799986-CC genotypes correlated with better OS [87]. Particularly, the *ANXA1*-rs1050305-AA genotype showed longer OS compared to any G allele both in the discovery and validation cohorts (HR:1.87, \( p = 0.03 \); HR:2.69, \( p < 0.001 \), respectively) and *LRP1*-rs1799986-CC had better OS than any T allele (HR:1.69, \( p = 0.03 \)) although not confirmed in the validation cohort [87]. In the discovery cohort, the *CALR*-rs1010222-A allele had a better PFS in univariate analysis (HR:0.61, \( p = 0.008 \)) compared to the GG genotype, with only a concordant trend in multivariate analysis. This association was not confirmed in the validation cohort.

 Destruction of tumor cells mediated by the binding of the antibodies (including cetuximab) through the crystalline fragment (Fc) receptors of NK cells is termed ADCC [18,20]. ADCC represents a mechanism by which an effector cell without antigenic specificity can mediate antigen-specific functions by recognizing the Fc portions of antibodies used to bind to the tumor cell. Activated NKs lyse tumor cells and stimulates the immune response via cross-talk with dendritic cells (DCs), and macrophages, via IFN\( _\gamma \), chemokines, and cytokines. Tumor cell lyses, by lytic granules, further stimulate the release of tumor antigens, which triggers additional cytotoxic activity via presentation to cytotoxic T cells by DCs [20,83]. The Fc\( _\gamma \)Rs are encoded by *FCGR1* (CD64), *FCGR2* (CD32), and *FCGR3* (CD16) in chromosome 1, and polymorphisms in the Fc receptor of immune cells, altering the affin-
ity for IgG Fc fragments, may influence the response to cetuximab proving to be a promising host genetic biomarker. Currently, a couple of SNPs have been associated with different IgG affinities: FCGR2A-rs1801274 (535 A > G, Arg131His) and FCGR3A-rs396991 (818 A > C, Val158Phe). The FCGR2A-rs1801274-His131 allotype displays a greater affinity for IgG1 compared to Arg131, whereas the FCGR3A-rs396991-Val158 allotype showed an increased affinity for IgG1, IgG2, and IgG3 compared to Phe158, improving the immune response through ADCC activation [151,152]. However, investigations on FCGR polymorphisms in mCRC patients are not yet conclusive as to their hypothetical significance at the clinical level. Despite the correlation between these polymorphisms and IgG binding affinity, some studies in mCRC patients showed no significant results assessing FCGR2A-rs1801274 and/or FCGR3A-rs396991 genotypes and cetuximab efficacy in terms of PFS, OS, and RR, irrespective of KRAS status [31,60,68–71,83]. However, in the study by Geva et al., an exploratory analysis in the subgroup of mutant KRAS mCRC patients, FCGR3A-rs396991-Phe/Phe was correlated to increased disease control rate (DCR) and OS compared to other genotypes (Pearson χ² p = 0.042 Phe/Phe vs. non-Phe/Phe and Log-Rank p = 0.030 Phe/Phe vs. non-Phe/Phe, respectively) [68]. In the NCIC CTG CO.17 trial, the subgroup of wild-type KRAS patients carrying FCGR2A-rs1801274-His/His had both improved OS and PFS, demonstrating respectively a 5.5 months benefit (HR:0.36, p = 0.003) and a 3.7 months benefit (HR:0.19, p = 0.02) compared to carriers of the Arg allele [80]. Shepshelovich and colleagues highlighted similar results by assessing 572 Australian wild-types KRAS mCRC patients. Those harboring FCGR2A-rs1801274-His/His showed longer OS (HR:0.66, p < 0.001) compared to any Arg. Despite FCGR3A-rs396991 genotypes not being associated with clinical outcomes, an advantage in terms of OS was found combining FCGR2A-rs1801274-His/His and FCGR3A-rs396991-Phe/Phe (HR:0.33, p = 0.003) compared to FCGR2A-rs1801274-Arg/Arg and FCGR3A-rs396991-Val/Val [81]. The role of FCGR3A-rs396991 polymorphism was evaluated in the meta-analysis by Ying and colleagues including 16 trials and a total of 2831 mCRC patients administered cetuximab-based chemotherapy. The analysis evidenced an improved PFS for patients carrying FCGR3A-rs366991-Phe/Phe in the dominant model considering both the overall population (MSR:0.680, p = 0.027) and the wild-type KRAS subgroup (MSR:0.728, p = 0.12) [72]. A study involving 96 mCRC Caucasian patients highlighted a significant correlation between the FCGR3A-rs366991-Val allele and longer PFS compared to Phe/Phe (10.8 vs. 5.1 months respectively, p = 0.05) [73]. Contrasting results were evidenced in a previous small study in which mCRC patients carrying FCGR3A-rs396991-Val/Val had shorter PFS (Phe/Phe: 2.3 vs. Val/Val: 2.4 vs. Val/Phe: 2.5 months; p = 0.055). However, the combination of FCGR2A-rs1801274-Arg/Arg and FCGR3A-rs396991-Val/Val highlighted a shorter median PFS compared to other combinations (1.1 vs. 3.7 months, p = 0.001) [74]. A later combination analysis evidenced that mCRC patients carrying FCGR2A-rs1801274-His/His and/or FCGR3A-rs396991-Val/Val genotypes had longer PFS than carriers of Arg and Phe, respectively (5.5 vs. 3.0 months; p = 0.005) [75]. Other latest studies of comparable size attempted to define the role of FCGR3A-rs1396991 and FCGR2A-rs1801274 polymorphisms about cetuximab efficacy generating, however, contrasting results [76–78,82,86]. Because the main toxicity of cetuximab is skin rash and the high degree of toxicity has very often been correlated with a good response to the drug, some studies have attempted to evaluate these polymorphisms as markers of toxicity, but insignificant results have been obtained [31,70,78]. Overall, FCGR2A and FCGR3A polymorphisms emerge as promising biomarkers for predicting the efficacy of cetuximab. However, further prospective studies are needed to conclusively define the role of the rs1801274 and rs396991 polymorphisms concerning cetuximab efficacy, including consideration of KRAS status.

5. Conclusions

In recent years, the growing number of studies on metastatic colorectal cancer has highlighted how the different mechanisms of tumor onset, as well as the pathways and mechanisms of evasion of cancer cells, are strictly interconnected with the immune system,
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making further investigations necessary. At present, the inflammation and immune-related germline genetic markers emerge as promising predictors of the therapy outcome in mCRC patients, representing a further potential tool for personalizing and optimizing treatments. Most of the available data regard the impact of these markers on the therapy effectiveness while their effect on the risk to develop severe toxicity remains limited (Table 2).

Several studies focused on the impact of polymorphisms in genes belonging to ADCC (FCGR2A, FCGR3A) and yielded promising, but not conclusive, results on cetuximab efficacy. The remaining published data are sparse and have mainly hypothesis-generating value, suggesting, however, potentially interesting topics for future pharmacogenetic studies. Particularly, in addition to the tumor immune escape pathway, genetic markers belonging to the cytokines/interleukins, including CXCL8, CCL5, CCR5 and angiogenic mediators including IGF1, IRF3, NOS2, appear to be the most promising (Figure 3).

Table 2. Final summary with high-level evidence for efficacy and toxicity focusing on therapy.

| Therapy | Genes | rs Code/Alias | Allele/Genotype | Clinical Outcome |
|---------|-------|---------------|-----------------|-----------------|
| CETUXIMAB | STING1 | rs7380824 | T allele | Worse ORR |
| | | rs1131769 | T allele | Worse OS |
| | IFNBI | rs1051922 | GA, AA genotypes | Worse PFS |
| | TLR7 | rs3853839 | GG genotype | Better PFS |
| | CD24 | rs52812045 | A allele | Worse PFS, OS |
| | CTLA-4 | rs231777 | T allele | Worse PFS |
| | CD274 | rs2297137 | G allele | Worse ORR |
| | IDO1 | rs9657182 | CT genotype | Better OS in Caucasian |
| | | rs3739319 | GG genotype | Better OS in Japanese |
| | CCL5 | rs2280789 | G allele | Worse OS in KRAS wild-type |
| | CCR5 | rs1799988 | T allele | Worse PFS, RR |
| | FCGR3A | rs396991 | C allele | Better OS, PFS, and PFS in KRAS wild-type |
| IL6 | rs2069837 | G allele | Worse PFS, ORR |
| CXCR4 | rs2228014 | T allele | Worse PFS, OS |
| CCL2 | rs4586 | C allele | Better PFS in KRAS mutant |
| CXCL8 | rs4073 | A allele | Worse PFS, OS, ORR |
| TBK1 | rs7486100 | T allele | Worse PFS in KRAS wild-type |
| IRF3 | rs2304205 | C allele | Better PFS in KRAS wild-type |
| CD274 | rs2297137 | G allele | Better OS |
| ADORA2B | rs2015353 | TT genotype | Better PFS |
| CD39/ENTPD1 | rs11188513 | C allele | Worse OS |
| | rs2226163 | GG genotype | Better OS |
| CD73/NT5S | rs2229523 | A allele | Better OS |
Table 2. Cont.

| Therapy                             | Genes | rs Code/Alias | Allele/Genotype | Clinical Outcome       |
|-------------------------------------|-------|--------------|-----------------|------------------------|
| FOLFIRI + BEVACIZUMAB               | TLR1  | rs5743618    | TT genotype     | Worse RR               |
|                                     | TLR6  | rs5743818    | AA genotype     | Worse PFS              |
|                                     | IDO1  | rs9657182    | T allele        | Worse PFS              |
|                                     | NOS2  | CCTTT repeat | >13 repeats     | Better PFS             |
|                                     |       | rs1053004    | C allele        | Lower toxicity         |
|                                     | IGF1  | rs2946834    | A allele        | Better PFS; PFS, OS in RAS wild-type |
|                                     | IRS1  | rs1801123    | C allele        | Worse OS; OS in RAS wild-type |
|                                     | NR1L2/PXR | rs1054190  | TT genotype     | Worse OS               |
|                                     | NR1L1/VDR | rs7299460  | T allele        | Better OS              |
|                                     | IL15RA | rs7910212    | C allele        | Worse OS               |
|                                     | SMAD3  | rs5719840    | C allele        | Better OS              |
| OXALIPLATIN                         | CDX2  | rs3812863    | GG genotype     | Better ORR, OS, PFS    |
|                                     | MS4A12 | rs4939378    | GG genotype     | Better OS, PFS         |
|                                     | ANXA1  | rs1050305    | G allele        | Worse OS               |

Several studies focused on the impact of polymorphisms in genes belonging to ADCC (FCGR2A, FCGR3A) and yielded promising, but not conclusive, results on cetuximab efficacy. The remaining published data are sparse and have mainly hypothesis-generating value, suggesting, however, potentially interesting topics for future pharmaco-genetic studies.

Particularly, in addition to the tumor immune escape pathway, genetic markers belonging to the cytokines/interleukins, including CXCL8, CCL5, CCR5 and angiogenic mediators including IGF1, IRF3, NOS2, appear to be the most promising (Figure 3).

Figure 3. Promising genes and related pathways influenced by polymorphisms. Abbreviations: VEGF: Vascular Endothelial Growth Factor; MMP: Matrix metalloproteinases; EMT: Epithelial-mesenchymal transition; ADME: absorption, distribution, metabolism, and excretion; TME: tumor microenvironment; Nk: Natural Killer; DCs: Dendritic Cells.
The data published to date on the predictive role of genetic variants of the acute phase, innate, and adaptive immune responses may also lead to a better understanding of the involvement of immune regulation in modulating treatment outcomes and mechanisms of treatment resistance. In particular, genetic variants belonging to angiogenic mediators (CXCR4, CCL2, CXCL8, TBK1, IRF3) and tumor immune escape molecules (CD24, CTLA-4, CD274, IDO1) are mainly investigated in anti-VEGF and anti-EGFR therapies, respectively, generating promising, although preliminary data, suggesting a potential translation, after validation, into clinical practice. Furthermore, these data may provide information regarding possible novel biological targets for putative immunotherapies and vaccine strategies. Finally, the discovery of new immune/pharmacogenetics markers could ameliorate the current CMS classification in CRC, identifying further cancer subtypes that may be helpful in further personalizing treatments. In fact, alteration in genes with an immunosuppressive role could compromise the immune system homeostasis, causing an increase in its activity and a prolonged state of chronic inflammation, and then favoring tumor progression and therapy ineffectiveness.

In addition to the host genetic variants, other inflammation and immune-related molecular markers have been indicated to be helpful in predicting the outcome of mCRC treatment. Somatic biomarkers, such as Immunoscore, immunoprofiling, or tumor mutational burden (TMB), have been proposed to have a predictive or prognostic value [153,154], although their implementation in clinical practice is still tricky, mainly for the limited availability of tumor tissues. However, noninvasive approaches based on the body fluids of patients could overcome this limit. In particular, liquid biopsy approaches, such as noninvasive genetic tests, isolating tumor-derived entities could monitor the tumor and its development by an analysis of immune-related genomic and proteomic data [155]. In addition, both the germline variant profile of patients and immune system biomarkers monitored during chemotherapy can help predict survival, response to treatment, or development of toxicity.

Last but certainly not least, the increasing knowledge about the composition of the gut microbiota has made the interaction between the microbiota and the immune system a new field of interest. The composition of the microbiota, polymorphisms that alter immune-related genes, as well as genes involved in the production of reactive oxygen species and reactive nitrogen species, could adversely affect immune homeostasis, which is a key factor in preventing the development of cancer and influencing the effects of pharmacological treatments.

In conclusion, according to the literature data discussed in this review, the study of inflammatory and immune-related genetic markers is certainly an important and essential topic that should be the goal of future studies aimed at personalizing the treatment of mCRC.

**Author Contributions:** L.S.: Conceptualization, Investigation, Data curation, Writing—original draft; Funding acquisition; A.B.: Investigation, Data curation, Writing—original draft; E.D.M.: Validation, Writing—review and editing; E.C.: Writing—review and editing, Supervision; G.T.: Funding acquisition, Supervision. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the Italian Ministry of Health (Ricerca Corrente), and 5 × 1000 “Ricerca Sanitaria”.

**Institutional Review Board Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.
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