Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.
Revisiting the sigma-1 receptor as a biological target to treat affective and cognitive disorders

Kinga Sałaciak, Karolina Pytka *
Department of Pharmacodynamics, Faculty of Pharmacy, Jagiellonian University Medical College, Medyczna 9, 30-688 Krakow, Poland

ARTICLE INFO
Keywords:
Sigma-1 receptors
Memory deficits
Depression
Animal models

ABSTRACT
Depression and cognitive disorders are diseases with complex and not-fully understood etiology. Unfortunately, the COVID-19 pandemic dramatically increased the prevalence of both conditions. Since the current treatments are inadequate in many patients, there is a constant need for discovering new compounds, which will be more effective in ameliorating depressive symptoms and treating cognitive decline. Proteins attracting much attention as potential targets for drugs treating these conditions are sigma-1 receptors. Sigma-1 receptors are multifunctional proteins localized in endoplasmic reticulum membranes, which play a crucial role in cellular signal transduction by interacting with receptors, ion channels, lipids, and kinases. Changes in their functions and expression may lead to various diseases, including depression or memory impairments. Thus, sigma-1 receptor modulation might be useful in treating these central nervous system diseases. Importantly, two sigma-1 receptor ligands entered clinical trials, showing that this compound group possesses therapeutic potential. Therefore, based on preclinical studies, this review discusses whether the sigma-1 receptor could be a promising target for drugs treating affective and cognitive disorders.

1. Introduction
Sigma-1 receptors are proteins that attract much attention as potential targets for drugs treating many medical conditions. The substantial therapeutic potential of sigma-1 receptor ligands is due to the involvement of the sigma-1 receptor in the development of many disorders, such as retinal diseases (Cantarella et al., 2007; Ha et al., 2011; Martin et al., 2004; Smith et al., 2008), drug addiction (Hirani et al., 2013; Romieu et al., 2002), alcohol addiction (Skuz, 2013), pain (Bravo-Caparrós et al., 2019; Bruna et al., 2018; Shin et al., 2020), ischemic stroke (Urfer et al., 2014), schizophrenia (Borison et al., 1991; Gewirtz et al., 1994), Alzheimer disease (Maurice et al., 1998), Parkinson disease (Francardo et al., 2019, 2014), amyotrophic lateral sclerosis (Mancuso et al., 2012; Ono et al., 2014), multiple sclerosis (Collina et al., 2017; Lisak et al., 2020), Huntington’s disease (Bol’shakova et al., 2017; Squitieri et al., 2015; Vetel et al., 2021), as well as cognitive and affective disorders (Albayrak and Hashimoto, 2012; Furuse and Hashimoto, 2010; Mandelli et al., 2017; Wang et al., 2019a). The prevalence of the latter two conditions has dramatically increased recently due to the COVID-19 pandemic (Ettman et al., 2020; Hampshire et al., 2021). Studies indicate that even three times as many adults report symptoms of depression or some form of cognitive decline (e.g., brain fog) now in comparison to last year (Ettman et al., 2020; Hampshire et al., 2021). Taking that into account, in this review, based on preclinical studies, we will discuss the therapeutic potential of sigma-1 receptor ligands as drugs improving mood and memory.

2. Basic information on sigma-1 receptor
2.1. Structure, distribution, and localization
Sigma-1 receptors are endoplasmic-reticulum-resident transmembrane proteins, acting as intracellular receptors with chaperone protein function. They are mainly expressed in the central nervous system (hippocampus, hypothalamus, mesencephalon, olfactory bulb). However, they can also be found in the liver, kidney, lungs, muscles, urinary bladder, as well as endocrine, immune and reproductive tissues (Gundlach et al., 1986; Kekuda et al., 1996; Mei and Pasternak, 2001). In the central nervous system, sigma-1 receptors are present on neurons and glial cells (astrocytes, oligodendrocytes, and ependymocytes).

The sigma-1 receptor is a single polypeptide containing 223 amino acids (26 kDa). The crystal structure revealed a trimeric organization.
with a single transmembrane domain for each protomer, and the carboxy-terminal domain included a cupin-like β-barrel with the ligand-binding site buried at its center (Schmidt et al., 2016). Sigma-1 receptors occur on the endoplasmic reticulum (ER)/mitochondrial associated membranes (MAM) and at very low levels in post-synaptic thickenings of the neuronendoplasmic reticulum (Alonso et al., 2000; Su et al., 2010). At MAM, they are sequestered with glucose-related protein 78/binding immunoglobulin protein (BiP), a major ER chaperone protein (Hayashi and Su, 2007). After the activation, the receptors separate from binding immunoglobulin protein and act either within the MAM or translocate to plasma membrane regulating calcium signaling (Hayashi and Su, 2007). Sigma-1 receptor may also transfer from the MAM to the nucleus membrane, interact with various nuclear factors and regulate gene transcription (Tsai et al., 2015).

2.2. Oligomerization

Studies demonstrated that sigma-1 receptors exist as monomers, dimers, and higher order oligomers (Pal et al., 2007). However, the ligand-binding properties occur only in an oligomeric state (Gromek et al., 2014). Interestingly, the agonist’s presence favors the monomeric and dimeric form of sigma-1 receptors, while the antagonist promotes higher order receptor oligomers (Mishra et al., 2015; Yano et al., 2018). It is an exciting feature of the sigma-1 receptor, considering its possible role in receptor functions. Furthermore, a GXXG motif of the sigma-1 receptor takes part in its oligomerization – mutations within this region disrupt this process and result in an increased number of smaller oligomeric states (Gromek et al., 2014).

2.3. Functions of sigma-1 receptors

2.3.1. Interaction with other proteins

Sigma-1 receptors interact with ion channels, G-protein coupled receptors (GPCRs), and cell-signaling molecules (Fig. 1). Therefore, it is not surprising that the sigma-1 receptor participates in diverse physiological and pharmacological processes. Moreover, the sigma-1 receptor regulates the compartmentalization of lipids on the endoplasmic reticulum and their further transport to the plasma membrane and cytosol (Hayashi and Su, 2003).

2.3.1.1. Ion channels. Sigma-1 receptors regulate ion currents interacting with various channels, such as Kv1.2 channels, increasing their surface expression (Abraham et al., 2019; Kourrich et al., 2013). They also inhibit Kv1.3, Kv1.4, and Kv1.5 channels (Aydar et al., 2002; Kinoshita et al., 2012; Zhang and Cuevas, 2005), regulate Kv2.1 channels, inhibit L-type voltage-gated calcium channels (Church and Fletcher, 1995; Mueller et al., 2013; Tchedre et al., 2008; Zhang and Cuevas, 2002), inhibit N-type Ca\(^{2+}\) channels currents (Zhang and Cuevas, 2002; Zan et al., 2017a, b, c), inhibit voltage-gated sodium ion channels: Nav1.2/1.4 (Gao et al., 2012) and Nav 1.5 (Johannesen et al., 2009; Koornneef et al., 2009), and regulate hERG channels (Balasuriya et al., 2014; Crottes et al., 2011). Moreover, sigma-1 receptors inhibit acid-sensing ion channels (ASIC1a) by activating a PTX-sensitive G-protein and stimulating AKAP150 bound calcineurin (Carnally et al., 2010; Mari et al., 2014). They also inhibit store-operated Ca\(^{2+}\) entry by diminishing coupling of stromal interaction molecule 1 (STIM1) to calcium release-activated calcium channel

![Fig. 1. A summary of the interaction of sigma-1 receptors with various proteins.](image-url)
2.3.1.7. Neurotransmitters.

2.3.2. Neurotransmission

Activating sigma-1 receptor can enhance glutamatergic and cholinergic synaptic neurotransmissions. Low doses of sigma-1 receptor agonists potentiated the N-methyl-D-aspartate (NMDA)-induced activation of pyramidal neurons in the CA3 region of the hippocampus and monoamines release (Monnet et al., 1995, 1992). The possible mechanism of current enhancement through NMDA receptors involved preventing small conductance Ca$^{2+}$-activated K$^+$ current (SK) channel activation (Martina et al., 2007), which, in turn, increased Ca$^{2+}$ influx through the NMDA receptors and potentiated receptor-mediated responses, including long-term potentiation (LTP). Interestingly, sigma-1 agonists increased the expression of GluN2A and GluN2B subunits and postsynaptic density protein 95 (PSD-95), proteins required for synaptic plasticity associated with NMDA receptor signaling. They also enhanced the interaction between GluN2 subunits and sigma-1 receptors but attenuated the coupling between GluN2B and PSD-95 (Pabba et al., 2014). Additionally, sigma-1 receptors stimulation increased the NMDA receptors levels on the cell surface (Pabba et al., 2014).

Moreover, the blockade of sigma-1 receptors modulated gamma-aminobutyric acid (GABA) and glutamate uptake, transport-mediated release, and exocytosis (Pozdnyakova et al., 2020). Interestingly, the sigma-1 receptor antagonist decreased the glutamate release but induced a biphasic response for GABA – low doses of NE-100 increased GABA uptake and with increasing dose, the uptake rate was decreasing (Pozdnyakova et al., 2020).

Sigma-1 receptors regulate the cholinergic neurotransmission either indirectly, through modulation of NMDA receptors in the brain, or directly. Stimulating sigma-1 receptors increased the extracellular acetylcholine level (Horan et al., 2002; Junien et al., 1998; Matsuno et al., 2008). Low doses of NE-100 increased GABA uptake and with increasing dose, the uptake rate was decreasing (Pozdnyakova et al., 2020).
2.3.3. Neurite outgrowth
Sigma-1 receptors influence neurite outgrowth. Scientists showed that sertraline inhibited nerve growth factor (NGF)-induced neurite outgrowth in PC12 cells, and this effect was blocked by both agonist and antagonist of sigma-1 receptors (Matsushima et al., 2019). The study suggested that the antidepressant acted as an inverse agonist of the sigma-1 receptor (Matsushima et al., 2019). On the other hand, fluvoxamine improved NGF-induced neurite outgrowth, which was inhibited by dexamethasone, acting via sigma-1 receptors and normalizing also the disrupted phosphorylation of a serine/threonine protein kinase (Akt) (Matsushima et al., 2018).

Similarly, dipentylammonium, a sigma-1 receptor agonist, enhanced neurite outgrowth (Brimson et al., 2018). Moreover, the compound protected from glutamate toxicity and prevented nuclear factor kappa B (NFκB) activation in HT-22 cells as well as dopamine-induced cell death in the APP/Swe expressing Neuro2A cells (Brimson et al., 2018). This stimulation of neurite-outgrowth might have involved increased production of adenosine triphosphate (ATP).

2.3.4. Neuroprotection
Sigma-1 receptors can promote neuroprotection. Studies demonstrated that sigma-1 receptor stimulation caused a neuroprotective effect, resulting from receptor translocation to the endoplasmic reticulum, stabilizing the IP$_3$ receptor, reducing ER stress, and modulating calcium flux. Eventually, the normalization of the calcium flow to mitochondria prevented mitochondrial leakage and stimulated metabolism (Hayashi and Su, 2007). In addition, the sigma-1 receptor agonists are neuroprotective against glutamate-, rotenone-, oligomycin-, and NMDA-induced neurotoxicity in the neuronal cell lines (Brimson et al., 2018; Franchini et al., 2020; Nishikawa et al., 2000; Senda et al., 1998a, b; Shimazu et al., 2000).

N,N-Dimethyltryptamine, a sigma-1 receptor agonist, prevented cell apoptosis in the ischemic injury, suggesting that the compound counteracted ferroptotic cell death linked to oxidative stress (Szabó et al., 2021). Also, dehydroepiandrosterone (DHEA), neurosteroid stimulating sigma-1 receptors, showed anti-amoosic activity when it was administered between 3-48 h after ischemia. However, when given during ischemia or early reperfusion, it demonstrated neurotoxic effects by increasing Ca$^{2+}$ influx through NMDA receptors (Li et al., 2009). Its neuroprotective mechanism may depend on sigma-1 receptor-mediated changes like inhibition of NMDA-induced nitric oxide synthase hyperactivity, reduction of nitric oxide production (Kurata et al., 2004), stimulation of phosphoinositide 3-kinase (PI3) activity (Raval et al., 2003), and antiapoptotic Bel-2 proteins (Charalampopoulos et al., 2004), or upregulation of glutamate transporter-1 (GLT-1) (Sokabe et al., 2007) and extracellular signal-regulated kinase (ERK) phosphorylation (Shi et al., 2021). Interestingly, the optimal time window for the administration of sigma-1 receptor ligands is crucial for retinal neuroprotection – (+)-pentazocine given since post-natal day 14 (P14), but not P18, P21 or P24, to the Pde6jdr10/J (rd10) mice (mouse model of retinitis pigmentosa) preserved the visual acuity (Wang et al., 2021a, b).

This neuroprotective mechanism of sigma-1 receptors may depend on modulation of miR-214-3p in retina, a microRNA involved in the oxidative stress (Wang and Smith, 2019).

Sigma-1 receptors are also a crucial node in pathways determining cellular survival – they enhance the IRE1 stability at MAMs under endoplasmic reticulum stress (Mori et al., 2013). Also, the overexpression of these receptors promotes the protection from ER stress, whilst their decrease leads to apoptosis (Hayashi and Su, 2007). The cytoprotective effect of sigma-1 receptor ligands can be observed in both ischemic hepatocytes and hypertrophic cardiocytes. In ischemic hepatocytes, the sigma-1 receptor-mediated cytoprotective effect results from reducing the leakage of hepatic enzymes, increasing the metabolic capacities, and increasing the ATP synthesis (Klouz et al., 2008). In hypertrophic cardiocytes, the same effect is due to restoring mitochondrial Ca$^{2+}$ mobilization and ATP production (Tagashira et al., 2013b).

Moreover, the sigma-1 receptor also plays a significant role in mitochondria-targeted cell apoptosis – after activation, it increases a protective bcl-2 gene expression (Yang et al., 2007). In cultured cortical neurons co-incubated with A25-35 peptide, sigma-1 receptor agonists prevented the cell death after exposure to β-amyloid toxicity (Marrazzo et al., 2005). Moreover, the sigma receptor antagonist diminished the neuroprotective effects of memantine against Aβ-induced neurotoxicity in SH-SY5Y cells (Keshavarz et al., 2020). Interestingly, an anxiolytic drug, afovale, protected neurons regulating the intracellular calcium overload during ischemia and acidosis via activation of sigma-1 receptors (Cuevas et al., 2011a). Moreover, the compound also prevented the microglia activation and decreased their response to ATP during and after an ischemic episode in vitro (Cuevas et al., 2011b). That suggests the activation of sigma-1 receptors may exert neuroprotective properties modulating the microglia activation and functioning.

2.3.5. Oxidative stress
Sigma-1 receptor knock-out animals have increased oxidative stress levels (Pal et al., 2012). Similarly, in cells with overexpressed sigma-1 receptors, agonists decreased whilst antagonists increased oxidative stress levels (Pal et al., 2012). Interestingly, sigma-1 receptors activated the antioxidant response elements and upregulated NAD(P)H quinone oxidoreductase 1 (NQO1) and superoxide dismutase 1 (SOD1) mRNA expression (Pal et al., 2012). Moreover, reactive oxygen species (ROS) level was elevated in retinal Müller cells lacking sigma-1 receptors (Wang et al., 2015). On the other hand, the activation of sigma-1 receptors in retinal Müller cells attenuated the production of ROS by enhancing the nuclear factor (erythroid-derived 2)-like 2 (Nrf2) signaling pathway (Wang et al., 2015). Furthermore, (−)-pentazocine, a sigma-1 receptor agonist, ameliorated the LPS-induced increase in ROS production (Zhao et al., 2014). Moreover, a novel sigma-1 modulator with acetylcholinesterase inhibiting properties decreased ROS production in SH-SY5Y human neuroblastoma cell lines (Rui et al., 2018). The sigma-1 receptor activation disrupts the complex between PSD-95 and neuronal nitric oxide synthase (nNOS) (Yang et al., 2010), leading to decreased nitric oxide production (Goyagi et al., 2001). On the other hand, in the spinal cord and brain mitochondria, activation of the sigma-1 receptor leads to increased ROS production (Choi et al., 2013; Natavlishvili et al., 2015). In normal conditions, sigma-1 receptor agonists generate moderate oxidative stress in mitochondria through ROS production, involving complex I activity (Goguadze et al., 2019). In pathological conditions, such as Aβ-induced stress, sigma-1 receptor activation restores the mitochondrial physiology – normalizing the complex I and IV activity responsible for increased ROS production (Goguadze et al., 2019).

2.3.6. Neuroinflammation
Sigma-1 receptors, expressed in microglia, modulate microglial activation and attenuate neuroinflammation (for more information please see Liu et al., 2018). In microglial cells co-incubated with LPS, the expression of sigma-1 receptor was significantly decreased as a result of toll-like receptor 4 (TLR4) activation via the TLR4-TAK1-p38 MAPK pathway and the activation of histone deacetylase 6 (HDAC6) (Iwamoto et al., 2015). Another sigma-1 receptor agonist, (+)-pentazocine, reduced the release of tumor necrosis factor α (TNFα), interleukin 10 (IL-10), monocyte chemoattractant protein-1 (MCP-1), and nitric oxide (NO) and inhibited LPS-induced activation of ERK and c-Jun N-terminal kinase (JNK) pathways (Zhao et al., 2014). Moreover, in murine
microglial BV2 cells, the activation of the sigma-1 receptors by SKF83959 prevented M1 microglial activation induced by LPS and decreased the release of inflammatory cytokines like tumor TNF-α, IL-1β, and inducible nitric oxide synthase (iNOS) (Wu et al., 2015). Interestingly, in mouse model of motor neuron degeneration, sigma-1 receptors activation led to increased anti-inflammatory M2 microglial phenotypes (Peviani et al., 2014). The antioxidative and anti-inflammatory effect after the sigma-1 receptor activation was mediated by promoting Nrf2/HO-1 signaling and suppressing NF-kB activity (Wang and Zhao, 2019). SA4503 downregulated the expression of iNOS and TNF-α and upregulated glutathione (GSH) in cultured astrocytes after LPS-induced inflammation (Wang and Zhao, 2019). Moreover, activation of sigma-1 receptors is beneficial in mice after stroke, as it normalizes matrix metalloproteinase-9 overexpression and hyperreactive astrogliosis (Sánchez-Blázquez et al., 2018). Also, dexmedetomidine, a sigma-1 receptor agonist, alleviated the hypoxia/reoxygenation-induced neuroinflammation manifested by an increased level of inflammatory factors like TNF-α, IL-6 or MCP-1 (Zha et al., 2021). What is worth mentioning, the activation of sigma-1 by afobazole regulated the microglial activation preventing the amyloid-induced toxicity (Behensky et al., 2013).

3. Sigma-1 receptor ligands

Even though the sigma-1 receptor is highly expressed in mammalian cells, no endogenous ligand has been clearly identified; however, both dimethyltryptamine (DMT) and neurosteroids activate the receptor and may be considered as putative candidates (Fontanilla et al., 2009; Maurice et al., 1999b). Moreover, the sigma-1 receptor binds various drugs and compounds with diverse chemical structures and different therapeutic and pharmacological profiles (Table 1) (Fallica et al., 2021; Weber and Wünsch, 2017).

Unfortunately, most of the functional assays to identify and classify sigma-1 receptor ligands are not precise. Many studies described various in vitro, in vivo, and ex vivo systems responding in a specific manner to sigma-1 receptor agonists (for more information please see Colabufo et al., 2009). However, most of these methods are not appropriate for high throughput use as they rely on measuring downstream effects of sigma-1 receptor stimulation. For many years, by convention, we classify compounds as sigma-1 receptor agonists if they induce the typical sigma-1 receptor-mediated effects in animal studies or resemble the phenotypes of receptor overexpression, like increased translocation of sigma-1 receptors to the plasma membrane, interaction with various proteins, neuroprotection, but also increased proliferation of cancer cells (Alam et al., 2017; Hall et al., 2009; Hayashi and Su, 2007; Meunier and Hayashi, 2016; Wu et al., 2018). On the contrary, antagonists are compounds that do not exert pharmacological effects on their own but abolish the effects of sigma-1 receptors activation or mimic the phenotype of receptor knock-down.

On the other hand, since the sigma-1 receptors influence calcium signaling, some proposed methods measure calcium levels (Wu and Bowen, 2008). However, these assays lack the quantitative precision and dynamic range required to capture accurate dose-response curves adequately. Quantitative cell-based assays assuming that sigma-1 receptor agonists induce neurite outgrowth also depend on specific downstream effects, but a limited number of compounds demonstrate this effect (Takebayashi et al., 2002). Similar conclusions can be drawn from studies on striatal tissue from guinea pigs, where sigma-1 receptor activation inhibits the dopamine release from striatum – the assay is imperfect as a decrease in dopamine level may occur after stimulating or blocking other receptors (Gonzalez and Werling, 1997). Another proposed method is based on the use of novel receptor fluorescence resonance energy transfer (FRET) based biosensor – the sigma-1 receptor agonists decrease and antagonists increase the FRET signal (Gómez-Soler et al., 2014). This effect results from how sigma-1 receptor ligands affect the oligomerization process – antagonists promote the high order oligomers, while agonists – monomers and dimers (Mishra et al., 2015). However, the agonist-bound receptor crystallizes similarly to antagonist-bound sigma-1 receptors, and overall receptor conformation remains practically unchanged, except for a 1.8 Å shift of helix α4 found when compared the (+)-pentazocine-bound relative to the PD 144418-bound structure (Schmidt et al., 2018). Moreover, we cannot assume that all potential sigma-1 receptor agonists stabilize only the monomeric structure and antagonists – the high-molecular-weight oligomers. Therefore, current structural data are insufficient to differentiate the intrinsic activity of sigma-1 receptor ligands.

The most common and quite relevant method examines the sigma-1 receptor/BiP association in ELISA kits as activation of this receptor relies on BiP dissociation (Fujimoto et al., 2012). Additionally, the positive modulators of these receptors can be identified when they are co-administered with known sigma-1 receptor agonists, and they potentiated the efficacy of the sigma-1 receptor agonists (Martin et al., 2020).

What is worth mentioning is that agonists and antagonists affect the stability of sigma-1 receptors multimers, decreasing or increasing the number of receptor’s multimers, respectively (Hong, 2020). The bioluminescence resonance energy transfer (BRET) assays enable analyses of ligand-induced multimerization of sigma-1 receptors as well as its interaction with BiP (Yano et al., 2018). The antagonists enhanced sigma-1 receptor homomer BRET signals, whereas agonists do not. Moreover, BRET assays also help differentiate the agonists and antagonists based on the interaction between the sigma-1 receptor and BiP (Yano et al., 2018).

Most of the above methods are not ideal and do not give us the certainty of whether a compound is the agonist or antagonist of the

Table 1

| Agonists               | Antagonists            | Modulators         |
|------------------------|------------------------|--------------------|
| (+)-pentazocine        | NE-100                 | SOMCL-668          |
| (+)-SKF10047           | Haloperidol            | OZP002             |
| Fluvoxamine            | Sertraline             | ER1                |
| Igmesine (JO-1784)     | BD1047                 |                    |
| Pregnemonone (PRE)     | BD1063                 |                    |
| Dehydroepiandrosterone | PB212                  |                    |
| Donepezil              | A266                   |                    |
| Cutamesine (SA4503)    | BD1008                 |                    |
| PRE-048                | Dipyrenylamonium       |                    |
| Quetiapine             |                        |                    |
| Ziprasimine            |                        |                    |
| Venlafaxine            |                        |                    |
| Bupropion              |                        |                    |
| Dextromethorphan       |                        |                    |
| PCP                    |                        |                    |
| Desipramine           |                        |                    |
| Fluoxetine             | R278995/CRA0450        |                    |
| Berberine              |                        |                    |
| Ropinirol              |                        |                    |
| UMB62                  |                        |                    |
| UMB82                  |                        |                    |
| Ditylglycanidine (DTG) |                        |                    |
| 17β-estradiol          |                        |                    |
| N-n-propyl-3-(3-hydroxyphenyl)piperidine ((+)-3-PPP) | | |
| LS-1-137               |                        |                    |
| KT-95                  |                        |                    |
| ANAVEXI-41             |                        |                    |
| Dimemorfan             |                        |                    |
| (-)-MB22               |                        |                    |
| AP710B                 |                        |                    |
| Choline                |                        |                    |
Neuroscience and Biobehavioral Reviews 132 (2022) 1114–1136

4. Depressive-like state and sigma-1 receptors

Recent studies have shown that sigma-1 receptors play a significant role in the pathophysiology of neuropsychiatric disorders like depression, and their ligands may constitute a promising option for treating depression (reviewed in Fishback et al., 2010). In addition, we already know that many currently used antidepressants bind to sigma-1 receptors, and their pharmacological action may partially depend on these receptors (Hashimoto, 2009).

To assess the possible antidepressant-like effects of compounds targeting sigma-1 receptors, we can use either tests measuring the animals’ reaction to specific environmental conditions (like forced swim, tail suspension, and sucrose preference tests) (for more information on an amygdala (Zhang et al., 2017a, b, c)). However, the possible factors impaired long-term depression (LTD) induction in the basolateral reaction to specific environmental conditions (like forced swim, tail suspension, and sucrose preference tests) (for more information on an amygdala (Zhang et al., 2017a, b, c)).

The sigma-1 receptor knockout mice show depressive-like behaviors observed in male but not female knockout mice (Chevallier et al., 2011; Di et al., 2017). Only when sigma-1 knockout female mice were ovariectomized, these differences were not observed (Sha et al., 2015). Similarly, the same study reported that 17β-estradiol given to male mice demonstrated neuroprotective and antidepressant-like properties (Sha et al., 2015). The above studies suggest that estrogens can protect from sigma-1 receptors (Hashimoto, 2009). The attenuated GR-mediated feedback phosphorylation (Di et al., 2017) or models mimicking not only behavioral but also cellular and molecular changes observed in depression (like unpredictable chronic mild stress procedure, olfactory bulbectomy, chronic corticosterone administration, maternal separation) (for more information on animal models of depression, please see Wang et al., 2017). In the following subchapters, we review the available literature on the role of the sigma-1 receptor in regulating depressive-like states in rodents.

4.1. Sigma-1 receptor knock-out animals

The sigma-1 receptor knockout mice show depressive-like behaviors observed in the forced swim and tail suspension tests (Akune et al., 2001; Di et al., 2017; Sabino et al., 2009; Zhang et al., 2017a, b, c). Interestingly, the decrease in immobility and impaired neurogenesis was observed in male but not female knockout mice (Chevallier et al., 2011; Di et al., 2017). Only when sigma-1 knockout female mice were ovariectomized, these differences were not observed (Sha et al., 2015). Similarly, the same study reported that 17β-estradiol given to male mice demonstrated neuroprotective and antidepressant-like properties (Sha et al., 2015). The above studies suggest that estrogens can protect from depressive-like behavior and impaired neurogenesis induced by the absence of the sigma-1 receptor.

Moreover, sigma-1 receptor knockout mice had decreased expression of glucocorticoid receptors (GR) via reduced protein kinase C (PKC) phosphorylation (Di et al., 2017). The attenuated GR-mediated feedback inhibition of the hypothalamic–pituitary–adrenal (HPA) axis produced a long-lasting HPA axis hyperactivity and subsequent depressive-like phenotype (Di et al., 2017). Another research showed that the depressive-like phenotype observed in sigma-1 knockout mice developed due to decreased nNOS activity and nitric oxide level with a subsequent decrease in presynaptic glutamate and GABA release and impaired long-term depression (LTD) induction in the basolateral amygdala (Zhang et al., 2017a, b, c). However, the possible factors contributing to the specific change in behavior in sigma-1 knockout mice could be multifactorial.

4.2. The effect of sigma-1 receptor ligands on depressive-like behavior in rodents

Various sigma-1 receptor agonists showed antidepressant-like activity in tests assessing antidepressant-like effect (Kim et al., 2006; Matsumo et al., 1996; Phan et al., 2002; Reddy et al., 1998; Skuza and Rogóz, 2009; Ukai et al., 1998; Urani et al., 2001; Wang et al., 2007a, b).

The ammonium salts are sigma-1 receptor ligands, among which the most promising is dipentylammonium. The compound binds not only to sigma-1 receptors but also to sigma-2 receptors, showing antidepressant-like activity in the forced swim and tail suspension tests (Brimson et al., 2020). Moreover, the administration of BD1047, a sigma-1 receptor antagonist, reversed the decrease in immobility in mice treated with dipentylammonium (Brimson et al., 2020). The results of this study suggest that sigma-1 receptor activation contributes to the antidepressant-like effect. The molecular mechanism responsible for the antidepressant-like activity of sigma-1 receptor agonists might involve Ca2+ mobilization (Urani et al., 2002a).

Not only sigma-1 receptor agonists improve the animals’ performance in the tests assessing depressive-like behaviors. Such properties also show allosteric modulators of the sigma-1 receptor. One of the examples is SOMC1-G668, which decreased the immobility in the forced swim and tail suspension tests, and its effect was blocked after pretreatment with BD1047 (sigma-1 receptor antagonist) (Wang et al., 2016). OZP002, a positive modulator of sigma-1 receptors, showed antidepressant-like properties in the forced swim test and potentiated the effect of 17β-estradiol administered at sub-effective doses (Maurice et al., 2019). Given that the positive modulator led to the same behavioral effect as the classic sigma-1 receptor agonist, and the use of both sigma-1 receptors knock out mice or selective sigma-1 receptor antagonist prevented antidepressant-like effect, may suggest that OZP002 compound could potentiate the endogenous activation of the sigma-1 receptor.

Interestingly, scientists observed a depressive-like state assessed by the tail suspension, forced swim, and sucrose preference tests in mice with induced cardiac dysfunction. The depressive-like state was reversed by either intracerebroventricular infusions of prazosin or administration of sigma-1 receptor agonists – PRE084 and SA4503, respectively (Ito et al., 2012; Shinoda et al., 2016). The observed changes in behavior were most likely due to the decreased sigma-1 receptor expression in the hippocampus caused by elevated corticosterone levels (Moriguchi et al., 2013). Moreover, chronic dexamethasone infusions resulted in behavioral, cellular, and molecular changes in animals – they were more immobile in the forced swim test (Terada et al., 2014). Dexamethasone-treated mice showed decreased sigma-1 receptors and BDNF expressions, and chronic administration of fluvoxamine reversed these changes. Considering that sigma-1 receptor activation increases BDNF expression via cAMP-response element binding protein (CREB) activation (Ji et al., 2017; Xu et al., 2015), the increased sigma-1 receptor expression after treatment with fluvoxamine may be related to the increased BDNF expression.

Importantly, the sigma-1 receptors are involved in the antidepressant-like properties of multimodal compounds (Berkann et al., 2004; Chaki et al., 2004; Dhir and Kulkarni, 2007a, b; Khulbe et al., 2013; Kulkarni and Dhir, 2008; Nguyen et al., 2017; Villard et al., 2011b). For instance, sigma-1 receptors mediate the antidepressant-like activity of many antidepressants or antipsychotics. The quetiapine-dependent decrease in mice immobility in the forced swim test was potentiated by sigma-1 receptor agonists and abolished by pretreatment with sigma antagonists (Kotagale et al., 2013). Also, the anti-immobility effects of venlafaxine, bupropion, or fluvoxamine in the forced swim test may require interaction with sigma-1 receptors (Dhir and Kulkarni, 2007b, 2008c; Terada et al., 2014). Neurosteroids like dehydroepiandrosterone sulfate and pregnenolone sulfate also exhibited antidepressant-like effect in the tail suspension and the forced swim tests in mice after activating sigma-1 receptors (Dhir and Kulkarni, 2008b; Reddy et al., 1998; Urani et al., 2001). The synergic administration of inactive doses of sigma-1 receptor agonists with NMDA antagonist, 5-HT1A receptor agonist or selective serotonin reuptake inhibitors.
Akt/GSK-3β/b-catenin, or ERK. The antidepressant-like effect of sigma-1 agonists is mediated not by one but several signaling pathways, including nNOS-NO-CREB, nNOS-NO-CREB signaling (Zhang et al., 2017a, b, c). In olfactory bulbar neurons, the activation of sigma-1 receptors abolishes the negative behavioral effects of chronic stress (Chen et al., 2020; Liu et al., 2018, 2019). Moreover, only one-week treatment with SOMCL-668 (an allosteric modulator of sigma-1 receptor) was enough to reverse the stress-induced decrease in the sucrose preference (Wang et al., 2016). Furthermore, the sigma-1 allosteric modulator inactivated glycan synthase kinase 3 beta (GSK3β) and normalized the BDNF level in the stressed mice (Wang et al., 2016). Another study demonstrated that the stimulation of sigma-1 receptors by fluvoxamine ameliorated the depressive-like behaviors in streptozocin-induced diabetic rats, and the mechanism of this effect depended on the sigma-1 receptor-BDNF pathway (Lenart et al., 2016). In agreement, other findings indicated that the chronic stimulation of sigma-1 receptors increased the BDNF level in the hippocampus (Kikuchi-Utsumi and Nakaki, 2008). Moreover, several antidepressants increased the neurite outgrowth in PC12 cells via interaction with sigma-1 receptors (Ishima et al., 2014; Takebayashi et al., 2002). In addition, fluvoxamine increased the level of pro-BDNF, mature BDNF, and sigma-1 receptors which eventually led to cell protection in diabetic animals (Lenart et al., 2016). The sigma-1 receptors agonist also alleviated the symptoms of postpartum depression in mice by increasing the nNOS-NO-CREB signaling (Zhang et al., 2017a, b, c). In olfactory bulbectomized mice, the activation of sigma-1 receptors by chronic administration of dehydroepiandrosterone decreased the immobility time of animals in the forced swim test, suggesting antidepressant-like effect (Rogacz and Skuzu, 2006; Skuzu and Rogacz, 2002, 2006, 2007).

Interestingly, we have recently discovered that HBK-15, a multimodal compound showing fast antidepressant-like effect and anti-amnesic properties (Pytka et al., 2015, 2017a, 2017b, 2018; Waszkiewicz et al., 2014), also binds to sigma-1 receptors (unpublished results). It is very likely that the unique pharmacological profile of HBR-15 may depend on sigma-1 receptor-mediated signaling. However, our hypothesis requires confirmation.

The possible molecular mechanism of the antidepressant-like effect of sigma-1 receptor agonists may involve various signaling pathways. Studies showed that sigma-1 receptor stimulation led to molecular, cellular, and biochemical changes, including an increase in NR2A, CREB, NGF, and BDNF (Fukunaga and Moriguchi, 2017; Ji et al., 2017; Matsushima et al., 2018; Yagasaki et al., 2006). Both sigma-1 agonist (PB190) and antagonist (PB212) decreased the corticosterone-induced catalase (CAT) activity, an enzyme involved in the oxidative stress responses (Skuzu et al., 2011). That is an unexpected result, considering the activity of both agonist and antagonist of sigma-1 receptors. However, both compounds also possess a very high affinity towards sigma-2 receptors (Berardi et al., 2005), so these outcomes may be the effect of modulating the sigma-2 receptor’s activity. Fluvoxamine and dehydroepiandrosterone sulfate led to Akt-1 phosphorylation via sigma-1 receptors in PC12 cells, underlying their antidepressant-like activity (Nakano et al., 2010). Sigma-1 receptors also enhanced the glutamate release from presynaptic terminals caused by fluvoxamine in the stressed mice, which may be linked to the potential memory-improving effect of the drug in depressed individuals (Fu et al., 2012).

In animal models of depression, sigma-1 ligands also showed antidepressant-like activity. Rats, after chronic unpredictable mild stress procedure, demonstrated despair (increased immobility in the forced swim test), and anhedonic (decreased sucrose consumption) behaviors (Chen et al., 2020; Liu et al., 2019). Chronic treatment with SA4503 or fluvoxamine, sigma-1 receptor agonists, abolished the negative behavioral effects of chronic stress (Chen et al., 2020; Liu et al., 2018, 2019). Moreover, only one-week treatment with SOMCL-668 (an allosteric modulator of sigma-1 receptor) was enough to reverse the stress-induced decrease in the sucrose preference (Wang et al., 2016). Furthermore, the sigma-1 allosteric modulator inactivated glycan synthase kinase 3 beta (GSK3β) and normalized the BDNF level in the stressed mice (Wang et al., 2016). Another study demonstrated that the stimulation of sigma-1 receptors by fluvoxamine ameliorated the depressive-like behaviors in streptozocin-induced diabetic rats, and the mechanism of this effect depended on the sigma-1 receptor-BDNF pathway (Lenart et al., 2016). In agreement, other findings indicated that the chronic stimulation of sigma-1 receptors increased the BDNF level in the hippocampus (Kikuchi-Utsumi and Nakaki, 2008). Moreover, several antidepressants increased the neurite outgrowth in PC12 cells via interaction with sigma-1 receptors (Ishima et al., 2014; Takebayashi et al., 2002). In addition, fluvoxamine increased the level of pro-BDNF, mature BDNF, and sigma-1 receptors which eventually led to cell protection in diabetic animals (Lenart et al., 2016). The sigma-1 receptors agonist also alleviated the symptoms of postpartum depression in mice by increasing the nNOS-NO-CREB signaling (Zhang et al., 2017a, b, c). In olfactory bulbectomized mice, the activation of sigma-1 receptors by chronic administration of dehydroepiandrosterone decreased the immobility time of animals in the forced swim test (Moriguchi et al., 2013). The authors suspected that this effect occurred after increased neurogenesis in the hippocampus via activation of the Akt/GSK-3β/b-catenin pathway (Moriguchi et al., 2013). All above findings indicate that the antidepressant-like effect of sigma-1 agonists is mediated not by one but by several signaling pathways, including nNOS-NO-CREB, Akt/GSK-3β/b-catenin, or ERK.

In addition to neurons, astrocytes play a vital role in depression pathophysiology and treatment. Sigma-1 receptors modulate the activation of astrocytes by increased phosphorylation of ERK and GSK3β (Wang et al., 2019a, b). The activation of sigma-1 receptors also alleviated depressive-symptoms (in conditioned fear stress and forced swim test) in mice infused with β-amyloid protein (Urani et al., 2002b, 2004), proving that sigma-1 receptor agonists may exert antidepressant-like activity in various disorders often co-existing with depression.

A recent study shed new light on the role of sigma-1 receptors in depression. Plasma exosomes from depressed mice ameliorated depressive-like behaviors, deficiency of BDNF expression, and neuro-inflammation in LPS-treated mice via sigma-1 receptor delivery (Wang et al., 2021a, b). The exosomes without sigma-1 receptors no longer showed the antidepressant-like effect, suggesting that sigma-1 receptors are crucial for antidepressant-like activity.

4.3. Sigma-1 receptor ligands in clinical trials

Up to date, only SA4503 (cutamines) has completed phase II clinical trials for major depressive disorder (ClinicalTrials.gov Identifier: NCT00551109, Table 2) (M’s Science Corporation, 2008). Moreover, the compound was also tested for safety and motor function restoration in patients after acute ischemic stroke (ClinicalTrials.gov Identifier: NCT00639249) (M’s Science Corporation, 2009). Additionally, igmesine, another sigma-1 receptor agonist, was also tested in depressed patients; however, the compound failed to be effective in phase III clinical trials (Pande et al., 1998, 1999).

4.4. Conclusion

To conclude, the lack of sigma-1 receptors contributes to the development of depressive-like phenotype in male rodents. Conversely, the activation of sigma-1 receptors mediates the antidepressant-like response in naïve and depressed animals (Fig. 2), probably via influence on BDNF and NGF expression. The pharmacological effects of sigma-1 receptor ligands in tests assessing antidepressant-like effect are summarized in Table 3.

5. Memory functioning and sigma-1 receptors

Sigma-1 receptor ligands, especially agonists, have been studied in various models of memory impairments in rodents, either assessing emotional/aversive memory (using passive avoidance test (Jarvik and Kopp, 1967) or fear conditioning (Anagnostaras et al., 1999)), spatial working and reference memory (using Morris water maze (Morris, 1984), Y-maze (Dennis and Sollenberger, 1934), or recognition/episodic-like memory (using object recognition test) (for more information on animal memory tests please see Beuzen and Belzung, 1995; Lueptow, 2017; Vorhees and Williams, 2014)). Since compounds targeting sigma-1 receptors control calcium mobilization, neurotransmitters release, including glutamate and acetylcholine, and neurotrophic factors expression, it is not surprising that they often show anti-amnestic properties. Moreover, sigma-1 receptors may play a crucial role in the development of Alzheimer’s disease 2b/3 (Anavex Life Sciences Corp., 2021a, 2021b), FTD (Anavex Life Sciences Corp., 2020), and Rett syndrome (Anavex Life Sciences Corp., 2021c, 2021d).

Table 2: Sigma-1 receptor agonists in clinical studies.

| Sigma-1 receptor agonist | Conditions | Phase | Reference |
|--------------------------|------------|-------|-----------|
| SA4503 (cutamines)       | Major depressive disorder | 2     | (M’s Science Corporation, 2009) |
|                          | Acute ischemic stroke     | 2     | (M’s Science Corporation, 2009) |
| ANAVEX2-73 (blarcamines) | Alzheimer’s disease       | 2b/3  | (Anavex Life Sciences Corp., 2021a, 2021b) |
|                          | Parkinson’s disease dementia | 2    | (Anavex Life Sciences Corp., 2020) |
|                          | Rett syndrome             | 2     | (Anavex Life Sciences Corp., 2021c, 2021d) |

1120

Neuroscience and Biobehavioral Reviews 132 (2022) 1114–1136

K. Satuciak and K. Pytka
role in neurodegenerative diseases like Alzheimer’s disease. Sigma-1 agonists not only reversed memory impairments in rodent Alzheimer’s disease models after a single injection before the test (Maurice et al., 1998) but also showed neuroprotective properties (Antonini et al., 2011; Marrazzo et al., 2005; Maurice et al., 2019).

5.1. Sigma-1 receptor knock-out animals

Sigma-1 knockout mice exhibited impaired NMDA-dependent neurogenesis, the formation of LTP and LTD (Shá et al., 2013; Snyder et al., 2016; Zhang et al., 2017a, b, c). The lack of sigma-1 receptors caused the overproliferation of progenitor cells, but newborn neuronal cells’ survival was decreased in the dentate gyrus (Shá et al., 2013). Moreover, diminished expression of sigma receptors contributed to decreased NMDA NR2B phosphorylation in the basolateral amygdala and dentate gyrus, as well as the nNOS coupling to PSD-95 and NO level (Zhang et al., 2017a, b, c). Such an effect might be beneficial in neurodegenerative diseases because of the decrease in NMDA receptor-mediated neurotoxicity. The receptor knockdown also impaired the dentritic spine formation, and thus synaptic plasticity, crucial for memory and learning processes (Tsai et al., 2009).

The first study on sigma-1 knockout mice showed no significant cognitive function changes (Langa et al., 2003). However, later experiments demonstrated that only female homozygous and heterozygous mice showed spatial and working memory impairments in the water and Y-maze tests, respectively (Chevallier et al., 2011; Xu et al., 2017). Moreover, these deficits were more pronounced with age, and they could be reversed by 17β-estradiol (Chevallier et al., 2011). Interestingly, the emotional memory deficits assessed by the passive avoidance test appeared only in 12-month-old homozygous females (Chevallier et al., 2011). On the other hand, sigma-1 knockout male and female mice had impaired long-term, but not short-term, recognition memory (Xu et al., 2017). Also, the Hamlet test, where animals are exposed to an enriched environment, showed that lack of sigma-1 receptors completely prevented the establishment of topographic memory and behavioral resilience, as well as the hippocampal neurogenesis (Crouzier et al., 2020). These differences between males and females suggest that sex hormones may affect sigma-1 receptors signaling and in consequence memory processes. However, not all types of memory are equally affected in knock-out animals proving that sigma-1 receptors may have a diverse effect on different types of memory and learning processes.

5.2. The role of sigma-1 receptor in the LTP formation

Long-term potentiation (LTP) is a process based on the long-lasting increase in the synaptic strength following a high-frequency stimulation, being a foundation of synaptic plasticity and, thus, learning and memory (for more information please see (Lynch, 2004)). The sigma-1 receptors are a critical node in the NMDA-independent LTP formation. Neurosteroids via sigma receptors transiently enhanced presynaptic function (Chen et al., 2006; Jafari-Sabet et al., 2019). The same effect was observed in the hippocampus of rats treated with alcohol, suggesting the significant role of sigma-1 receptors in synaptic plasticity (Sabeti and Gruol, 2008). Furthermore, impaired LTP formation inolfactory bullectomized mice was improved after a chronic treatment with DHEA (Moriguchi et al., 2013), which increased calcium/calmodulin-dependent protein kinase II (CaMKII), PKC, and ERK activity in the hippocampus (Moriguchi et al., 2011). More specifically, DHEA led to increased CaMKII autophosphorylation and GluR1 phosphorylation in the dentate gyrus (Moriguchi et al., 2013). These molecular changes also affected animal behavior. DHEA reversed the spatial, recognition, and emotional memory deficits (Moriguchi et al., 2011). Moreover, the beneficial effect of DHEA was visible in the ischemic rats. The compound prevented the disruption of the LTP formation in CA1 by attenuating the reduction in tyrosine phosphorylation of NR2B (Li et al., 2006).

5.3. The effect of sigma-1 receptor ligands on various types of memory in rodents

Compounds with an affinity towards sigma receptors showed anti-amnesic properties in numerous studies, in which cognitive impairments were induced by different factors, such as drug (scopolamine, dizocilpine, phencyclidine, neurotransmission, cocaine, nimodipine) or toxin (β-amyloid(25–35)-peptide, CO, trimethyltin) administration, stress (post-traumatic stress disorder, prenatal restraint), or brain damage (brain ischemia, olfactory bullectomy) (Maurice et al., 1994a, 1994b, 1998, 1999a, 2001a; Meunier et al., 2006c). All mentioned animal models cause memory deficits that differ in severity and memory type affected (for more information on animal models of memory deficits please see Gallagher, 1997; Götz and Götz, 2009; More et al., 2016; van der Staay et al., 2011). They may be a result of impaired cholinergic (scopolamine), glutamatergic (dizocilpine, phencyclidine) or serotonergic neurotransmission (para-chlorophenylalanine), neurons death (β-amyloid(25–35)-peptide, 192 IgG-saporin, trimethyltin, ibotenic acid, carbon monoxide (CO), brain ischemia, olfactory bullectomy), oxidative stress (cysteine) or oxidative phosphorylation (prenatal stress) (for more information on animal models of cognitive impairments please see Beraki et al., 2009; Cheng et al., 2012; Deryabina et al., 2020; Li et al., 2020; Muriach et al., 2010; Nakajima et al., 2007; Pepeu, 2004; van der Staay et al., 2011).

Interestingly, these factors induce various changes in sigma-1 receptor density. For example, cholinergic lesion (caused by unilateral NMDA infusion to nucleus basalis) resulted in increased sigma-1 receptor expression in the parietal cortex, contrary to sleep deprivation that caused a decrease in the level of the receptor in the pons and midbrain (Ramakrishnan et al., 2015). Mice during the alcohol withdrawal showed the increased sigma-1 receptor expression in the hippocampus (Meunier et al., 2006a), while phencyclidine administration led to a decrease in the sigma-1 receptor level in the same structure (Kunii et al., 2009). However, the density of sigma-1 receptors remained unchanged throughout life (Phan et al., 2003). These findings suggest that the level of the sigma-1 receptor may be affected by drug or toxin administration, but not age.

Results regarding whether the activation or blockade of sigma-receptors is beneficial for memory are ambiguous. Most studies demonstrated that administration of sigma-1 receptor agonists/antagonists did not affect normal memory function. However, they showed such an effect might be beneficial in neurodegenerative diseases because of the decrease in NMDA receptor-mediated neurotoxicity.
### Table 3
Antidepressant-like activity of sigma-1 receptor ligands.

| Potential modulator                                      | Animal Models/Tests | Reference                  |
|----------------------------------------------------------|---------------------|----------------------------|
| **Sigma-1 receptor agonists**                            |                     |                            |
| SA4503, (+)-pentazocine, ditolylguanidine (DTG), J0-1784 (single administration) | Mice Forced swim test | (Matsuno et al., 1996)     |
| SA4503, (+)-pentazocine, DTG, desipramine, fluoxetine    | Mice Tail suspension test | (Ukai et al., 1998)     |
| UMB23, UMB82                                             | Mice Forced swim test | (Wang et al., 2007a, b)   |
| Igmnesine, (+)-SKF-10047, dehydroepiandrosterone sulfate | Mice Forced swim test | (Urani et al., 2001)     |
| PRE-084                                                  | Mice Forced swim test | (Skura and Rogoz, 2009)   |
| Dehydroepiandrosterone sulfate (DHEAS), pregnenolone sulfate (PS) | Rats Forced swim test | (Reddy et al., 1998)     |
| Dipentylammonium (DPA)                                   | Mice Forced swim test, tail suspension test | (Brimson et al., 2020)   |
| Igmnesine                                                | Mice Forced swim test | (Urani et al., 2002a; Villard et al., 2011b) |
| PRE084                                                   | Mice with accelerate cardiac dysfunction Forced swim test | (Ito et al., 2012) |
| SA4503                                                   | Mice after transverse aortic constriction Forced swim test, tail suspension test, sucrose preference test | (Shinoda et al., 2016) |
| Dehydroepiandrosterone (DHEA)                            | Olfactory bulbectomized mice Forced swim test, tail suspension test | (Moriguchi et al., 2013) |
| 17+/estradiol                                            | Mice Forced swim test, tail suspension test | (Urani et al., 2002a; Villard et al., 2011b) |
| Deuterated (d6)-dextromethorphan                         | Mice Forced swim test, tail suspension test | (Nguyen et al., 2017) |
| R278995/SRAO450                                          | Rats, mice Forced swim test, tail suspension test | (Chaki et al., 2004) |
| Berberine                                                | Naive and reseparinated mice Forced swim test, tail suspension test | (Kulkarni and Dhir, 2008) |
| Hydromethanolic extract of flowers of Tagetes erecta     | Mice Forced swim test | (Kulibe et al., 2013)    |
| Ropinol                                                  | Mice Forced swim test, tail suspension test | (Urani et al., 2002a; Villard et al., 2011b) |
| Quetiapine                                               | Mice Forced swim test | (Kotagale et al., 2013)  |
| Venlafaxine                                              | Mice Forced swim test | (Urani et al., 2002a; Villard et al., 2011b) |
| Bupropion                                                | Mice Forced swim test | (Urani et al., 2002a; Villard et al., 2011b) |

### Table 3 (continued)
Potential modulator Animal Animal Models/Tests Reference

| Potential modulator                                      | Animal Models/Tests | Reference                  |
|----------------------------------------------------------|---------------------|----------------------------|
| Dehydroepiandrosterone sulfate (DHEAS), pregnenolone sulfate (PS) | Rats Forced swim test | (Skura and Rogoz, 2007)   |
| DTG, SA4503                                               | Rats Forced swim test | (Skura and Rogoz, 2007)   |
| SA4503, siramesine                                        | Rats Forced swim test | (Skura and Rogoz, 2006, 2002) |
| Pramipexole + sertraline                                  | Rats Forced swim test | (Boguz and Skura, 2006)   |
| SA4503                                                   | Chronic unpredictable stressed rats Sucrose preference test, forced swim test | (Liu et al., 2015, 2016)   |
| Fluvoxamine                                              | Chronic unpredictable stressed rats Sucrose preference test, forced swim test | (Chen et al., 2020) |
| Fluvoxamine                                              | Diabetic-induced depressed rats Forced swim test | (Lenart et al., 2016) |
| PRE-084                                                  | Ovariectomized mice Forced swim test, tail suspension test | (Zhang et al., 2017a, b, c) |
| Igmnesine, (+)-SKF-10,047, dehydroepiandrosterone sulfate (igne, PRE-084) | A/25-35 peptide-treated mice Conditioned fear stress test | (Urani et al., 2004) |
| Sigma-1 receptor modulators                              |                     |                            |
| SOMCL-668                                                 | Naive mice, chronic unpredictable stressed mice Forced swim test, tail suspension test, sucrose preference test | (Wang et al., 2016) |
| OZP002                                                   | Mice Forced swim test | (Maurice et al., 2019)    |

their pharmacological effects when learning was impaired. Interestingly, one study demonstrated a negative effect of the sigma-1 receptor agonist on emotional memory (assessed by the passive avoidance test) (Freeman and Young, 2001). The research was performed on a day-old chicks, in which sigma-1 receptor agonist administration 5 h after training induced learning and memory deficits (Freeman and Young, 2001). However, this effect occurred when specific dose of (+)-SKF10047 was given to chicks – higher or lower dose did not disrupt memory. This suggests that sigma-1 receptor activation might be crucial for emotional memory consolidation, possibly by modulating the NMDA, GABA, and acetylcholine neurotransmission. However, this study did not profoundly investigate the cause of the amnesic effect of the sigma-1 receptor agonist. Nevertheless, most studies concerning the effects of sigma-1 receptor ligands on memory were performed on mice or rats, not chicks, so these discrepancies could be due to species differences.

### 5.3.1. Emotional/aversive memory
Sigma-1 receptor agonists enhanced disrupted emotional memory in rodents (Earley et al., 1991; Matsuno et al., 1993b; Maurice et al., 1994a, 2000, 2001a). Sigma-1 receptor agonists reversed emotional memory deficits caused by either dizocilpine administration, probably
targets such as the serotonin 5-HT$_2$A receptor (OPC-14523) (Tottori et al., 2002) or muscarinic receptors (ANAVEX2-73) (Espallergues et al., 2007; Villard et al., 2011a). Similarly, sigma-1 receptor modulators (OZP002 or E1R) were effective in scopolamine-induced memory impairments and these compounds also enhanced the anti-amnesic effect of either PRE084 or igmesine (Maurice et al., 2013; Zvejniece et al., 2014).

This suggests that sigma-1 receptor modulators affect the cholinergic neurotransmission, and they are an attractive therapeutic target for drugs treating memory deficits caused by a decrease in acetylcholine level.

Studies showed that sigma-1 receptor agonists might reverse memory deficits induced by sleep deprivation. Such example is cutamezine (SA4503), which prevented emotional memory impairment induced by sleep deprivation in the passive avoidance task (Ramakrishnan et al., 2015b). Interestingly, this anti-amnesic effect observed at higher doses may result from more prolonged exposure and/or drug binding to other receptors or channels. As sleep deprivation-induced memory impairments are caused by the increase in misfolded proteins’ level and expression of inflammatory cytokines (Esumi et al., 2011; Rothman et al., 2013), the fact that sigma-1 receptor agonist reversed the consolidation and memory deficits is promising. Considering that many diseases are associated with more or less pronounced sleep disturbances, which may worsen their clinical course and lead to memory deficits, treatment with sigma-1 receptor agonists might be a promising strategy.

Worth mentioning is that the anti-amnesic effect of memory-enhancing drugs such as donepezil or rivastigmine can be correlated with stimulation of NGF-mediated neurite outgrowth via sigma-1 receptors (Ishima et al., 2008; Terada et al., 2018). In mice with dizocilpine-induced emotional memory deficits, donepezil showed anti-amnesic activity, which was attenuated when the sigma-1 receptors were pharmacologically blocked (Maurice et al., 2006). The above findings suggest that sigma-1 is an essential target for anti-amnesic effect.

In trimethyltin-induced memory disruption, igmesine (JO-1784), PRE-084, and dextromethorphan, a nonselective serotonin reuptake inhibitor and a sigma-1 receptor agonist, improved emotional memory after sigma-1 receptors stimulation (Maurice et al., 1999a; O’Connell et al., 1996; Shin et al., 2007). Moreover, PRE-084 and DTG were effective to ameliorate the memory and learning deficits after chronic CO exposure (Maurice et al., 1994c, 1999a). However, the neuroprotective effect of another sigma-1 receptor agonist, DHEA in CO-induced emotional memory impairments did not depend on sigma-1 receptors as their pharmacological blockade did not abolish memory-enhancing properties of the neurosteroid (Maurice et al., 2000). The basal forebrain lesion, caused by ibotenic acid administration, caused memory deficit in the passive avoidance task and these impairments were reversed after sigma-1 receptor activation (Senda et al., 1996).

In p-chloroamphetamine-induced amnesia sigma-1 receptor agonists showed anti-amnesic effects in the passive avoidance test (Matsumoto et al., 1994). Moreover, stimulating these receptors reversed memory impairments during the encoding, consolidation, and recall phase, thus all stages of learning and memory processes (Matsumoto et al., 1994, 1998; Senda et al., 1997).

The anti-amnesic effect of sigma-1 receptors activation may strongly depend on restoring the disrupted acetylcholinergic transmission. This hypothesis is based on the fact that similar to acetylcholinesterase inhibitors, sigma-1 receptor agonists showed anti-amnesic effect after serotonin depletion in the central nervous system, and sigma-receptor stimulation increased acetylcholine level in the frontal cortex and hippocampus, key structures for memory (Junien et al., 1991b; Matsumoto et al., 1992, 1993a). Also, the memory-enhancing properties disappeared after pretreatment with either scopolamine, a muscarinic receptor antagonist, or hemicholinium-3, a Na$^+$/dependent high-affinity choline uptake inhibitor. However, scopolamine itself causes memory deficits so it is difficult to state whether the anti-amnesic effect of the sigma-1 receptor agonist was antagonized, or the potentiated memory impairments were too potent to be reversed by sigma-1 receptor activation.

On the contrary, in the methamphetamine-induced memory disruption, the blockade of sigma-1 receptors improved emotional memory, assessed by the passive avoidance test (Seminero et al., 2013). This suggests that sigma-1 receptor antagonists improve memory deficits caused by decreased dopamine levels. Moreover, other studies showed that sigma-1 receptor antagonists prevented attenuated methamphetamine-induced apoptosis, necrosis, ROS/RNS generation, and dopamine release both in vitro and in vivo studies (Kaushal et al., 2012; Matsumoto et al., 2008).

Importantly, memory deficits occur most often during senescence. In senescence-accelerated mice, sigma-1 receptor agonists showed anti-amnesic properties by improving emotional memory (Maurice et al., 1996). Moreover, OPC-14523 (a sigma-1 receptor agonist) exhibited anti-amnesic properties in age-associated memory impairments (Tottori et al., 2002). However, as mentioned earlier, the compound also acts as a 5-HT$_2$A receptor agonist. Therefore, its effects might be related to the influence on serotonergic neurotransmission. Interestingly, only a single administration of OPC-14523 was enough to reverse behavioral changes in the scopolamine-induced memory deficits model (Tottori et al., 2002). However, the compound was unable to reverse age-related memory and learning impairments when it was administered once. On the hand, OPC-14523 ameliorated age-induced cognitive impairments, when it was administered chronically (Tottori et al., 2002).

Emotional memory deficits can also be caused by ischemia. Studies demonstrated that in mice after brain ischemia, sigma-1 receptor agonists restored proper memory functioning assessed by the passive avoidance test (Xu et al., 2015, 2017; Yabuki et al., 2015). These memory-enhancing properties might be mediated via NR2A-calcium/calmodulin-dependent protein kinase type IV (CaMKIV) - target of rapamycin complex 1 (TORC1) pathway as the sigma-1 receptor activation normalized the level of BDNF, NR2A, CaMKIV and TORC1 in the hippocampus (Xu et al., 2015). Other studies show that neuroprotective properties of sigma-1 receptor agonists in ischemic mice may be the result of increased ATP production, activation of Akt, CaMKII, and ERK as well as up-regulation of sigma-1 receptor expression in the hippocampus (Yabuki et al., 2015).

The anti-amnesic effect of the sigma-1 receptor agonist was also seen in prenatally stressed juvenile rats. IGF-1 improved spatial and emotional memory by normalizing the impaired glutamatergic, cholinergic, and adrenergic neurotransmission as well as HPA-axis functioning (decreasing the corticosterone release through an anti-CRF effect (Derocz et al., 1995; Eaton et al., 1996; Junien et al., 1991a; Mennier et al., 2004). Interestingly, another sigma-1 receptor agonist, SA-4503, also improved olfactory-bulbectomized rats’ performance in the cued and contextual fear conditioning and this effect was abolished after NMDA blockade (Wang et al., 2007a, b). This is an interesting result considering that memory deficits observed in olfactory bulbectomy arise from the degeneration of the cholinergic system and the decrease in acetylcholine level (Yamamoto et al., 1997; Yoshimura et al., 1974). That confirms that sigma-1 receptors affect cholinergic neurotransmission, and this effect may also depend on NMDA receptors.

Memory impairments also occurred after addictive drug exposure. Both igmesine and DHEA by activating sigma-1 receptors alleviated...
memory impairment observed in the passive avoidance task in prenatally cocaine-exposed mice (Meunier and Maurice, 2004). Studies showed that cocaine exposure led to sigma-1 receptor-dependent upregulation of D-type K⁺ current in the nucleus accumbens with a subsequent decrease in neuronal activity and enhancing behavioral cocaine response (Kourrich et al., 2013). The sigma-1 receptor agonist may modulate neuronal and behavioral response to cocaine since the cocaine-induced complex formation between the sigma-1 receptor and Kv1.2 channels redistributes both proteins from intracellular compartments to the plasma membrane (Kourrich et al., 2013).

5.3.2. Spatial working and reference memory

Various sigma-1 receptor agonists improved impaired spatial working and reference memory (water maze, eight-arm radial maze, Y-maze test) after dizocilpine treatment, and their memory-enhancing effect was abolished after pretreatment with sigma-1 receptor antagonist (Maurice et al., 1994a, 1994b, 1994d, 1997; Ohno and Watanabe, 1995; Zou et al., 1998, 2000) or oligodeoxynucleotide antisense (Maurice et al., 2001a, 2001b). Sigma-1 receptor agonists like (+)-SKF10047 or SA4503, or positive receptor modulators (OZP002 and E1R) reversed scopolamine-induced spatial short-term memory impairments in the Y-maze alternation test and long-term memory deficits in the water maze (Hiramatsu et al., 2002; Malik et al., 2015; Maurice et al., 2001a, 2019; Urani et al., 1998; Wang et al., 2003; Zevnieje et al., 2014).

Interestingly, both isomers of pentazocine exerted anti-amnesic properties in scopolamine-treated animals via stimulating sigma receptors (Hiramatsu and Hoshino, 2005). However, only the (+)-pentazocine reversed the dizocilpine-induced memory deficits in the Y-maze task. Another sigma-1 receptor agonist – OPC-14523 – showed memory-enhancing properties not only in mice with scopolamine-induced memory impairment (Morris water maze), but also improved cognition in age-induced learning and memory deficits (Tottori et al., 2002). Even when the nonselective compound was tested in animals with scopolamine-induced memory decline (KT-95 or U-50, 488H with affinity toward all opioid receptors), its anti-amnesic effect depended mainly on sigma receptors (Hiramatsu et al., 2006; Hiramatsu and Hoshino, 2004). Also, ANAVEX-2-73, sigma-1 and muscarinic receptor agonist, reversed the scopolamine- and dizocilpine-induced spatial memory impairments, and this effect depended on sigma-1 and muscarinic receptors (Espallergues et al., 2007; Villard et al., 2011a). Thus, the above studies suggest that muscarinic receptors are more important for the short-term memory, while sigma receptors are more engaged in the long-term memory formation.

What is interesting, sigma-1 receptors also play a significant role in the memory impairment caused by neurotoxic organometal – trimethylin. Igmese (JO 1784), PRE-084, and dextromethorphan reversed behavioral changes in the spatial memory assessing tasks (Maurice et al., 1999a, O’Connell et al., 1996; Shin et al., 2007). Since the neurotoxicity of trimethylin is the result of overstimulation of NMDA receptors and increased glutamate release, it seems that sigma-1 receptor activation normalizes the disrupted glutamatergic neurotransmission.

Mild or severe memory deficits may also develop after partial or almost complete cholinergic depletion. The sigma-1 receptor agonists (PPCC and SA4503) ameliorated the working and reference memory impairments acting via sigma-1 receptors (Antonini et al., 2009; Senda et al., 1998a, b). Interestingly, the administration of the sigma-1 receptor to animals with normal cognitive functions did not change any parameters (Antonini et al., 2009). It suggests that stimulating sigma-1 receptors exerts memory-enhancing properties only in situations when cognition is impaired. Moreover, when the level of endogenous steroids is diminished, their role in the CA1 activity and spatial learning is more visible, as castration increased the sigma-1 receptor mRNA expression level (Moradpour et al., 2016; Phan et al., 1999).

Diminished NO levels can also cause cognitive impairments. The reduced NO synthesis led to the short-term spatial memory impairment, which was reversed after sigma-1 receptor activation and a subsequent guanylate cyclase enhancing NO/cGMP pathway (Mamiya et al., 2000). In the nimodipine-induced memory impairments, PRE-084 attenuated impaired performance in the Y-maze and water maze tasks (Maurice et al., 1995). The study showed that sigma-1 receptor agonists were effective in various models of memory impairments. Moreover, the study proved that sigma-1 receptor agonists show anti-amnesic properties via facilitating the intracellular Ca²⁺ fluxes, which are involved in the memory processes.

Repeated exposure to carbon monoxide leads to the long-lasting but delayed memory deficits (Piantadosi et al., 1997). Like models of brain ischemia, the etiology of memory and learning impairments involves the neurotoxicity of excitatory amino acids, and subsequently disrupted hippocampal cholinergic system by hypoxic toxicity. As the sigma-1 receptor stimulation attenuated the CO-induced spatial memory and learning deficits (Maurice et al., 1994c, d; Meunier et al., 2006b), we can conclude that the anti-amnesic effect is the result of sigma-1 receptor-mediated neuroprotection and antioxidative properties. Interestingly, similar to emotional memory, sigma-1 receptor activation also ameliorated spatial learning and memory deficits in brain ischemia/r- reperfusion mice via increasing the level of BDNF (Xu et al., 2015, 2017; Yabuki et al., 2015).

Numerous stress models disrupt memory in animals. The memory impairments can be observed in the rat model of post-traumatic stress disorder, and the sigma-1 receptor activation prevented the spatial memory deficits as well as increased the phosphorylation of BDNF, TrkB, and ERK (Ji et al., 2016). Also, the prenatal restraint stress impaired the later cognitive abilities during adulthood (Wu et al., 2007). Rats showed impaired spatial performance in the Y-maze and Morris water maze due to the disrupted neurotransmission and the HPA axis functioning (Meunier et al., 2004). The administration of igmesine normalized animals’ behavior and disrupted the activity of the HPA axis (Meunier et al., 2004). This suggests that sigma-1 receptor activation thanks to neuromodulatory effects can alleviate memory impairments linked to developmental disorders.

Additionally, spatial memory deficits may also occur after drug exposure, especially during the prenatal period. The cocaine-treated dams exhibited the memory impairment, which was reversed after the sigma-1 receptor stimulation by igmesine or dehydroepiandrosterone (Meunier and Maurice, 2004). This neuroprotective effect of both compounds may result from restoring the normal functioning of several receptors, including NMDA, acetylcholine, dopaminergic or norepinephrine receptors and K⁺ channels, and normalizing the activity of the HPA axis and stress hormones’ levels.

One of the possible explanations why sigma-1 receptors play a crucial role in spatial memory may be the fact that in various models of memory deficits we observe decreased expression of sigma-1 receptors in the hippocampus or hypothalamus (Ito et al., 2013). For example, mice with myoccardial infarction showed impaired spatial working memory (decrease in Y-maze spontaneous alternation) (Ito et al., 2013), which was normalized via intracerebroventricular infusion of the sigma-1 receptor agonist (Ito et al., 2013).

What is worth mentioning, the sigma-1 receptor activation is also beneficial for age-related spatial working and reference memory impairments in senescence-accelerated mice as well as normal aging rats (Maurice et al., 1996; Maurice, 2001). Moreover, a recent study showed that sigma-1 receptor agonists were also effective in α-thalassemia X-linked intellectual disability (ATR-X) syndrome. SA4503, sigma-1 receptor agonist, reversed the abnormal axonal development and dendritic spine formation (Yamaguchi et al., 2018). ATR-X model mice spatial learning and memory deficits, which were ameliorated by SA4503 via increasing BDNF level in the medial prefrontal cortex (Yamaguchi et al., 2018).

5.3.3. Recognition/episodic-like memory

The role of the sigma-1 receptor in the recognition memory is ambiguous. Dehydrocorybulbine, a sigma-1 receptor ligand obtained
from Corydalis yanhusuo, attenuated MK-801-induced memory deficits in the object recognition test (Wang et al., 2019a, b). However, it is difficult to unambiguously state whether this memory-enhancing effect was mediated via sigma receptors, since the alkaloid binds to the various receptors (dopaminergic, serotonergic or adrenergic) (Wang et al., 2019a, b).

Among currently used drugs, many show significant affinity toward sigma receptors. The chronic administration of either fluvoxamine or donepezil, drugs with significant sigma-1 receptor affinity, ameliorated phencyclidine-induced impairments of the recognition memory (Hashimoto et al., 2007; Kunitachi et al., 2009). The pretreatment with NE-100, a sigma-1 receptor antagonist, abolished the drugs’ effects (Hashimoto et al., 2007; Kunitachi et al., 2009). Thus, the anti-amnesic activity of fluvoxamine and donepezil depended on sigma-1 receptor stimulation. Moreover, SA4503, a sigma-1 receptor agonist, reversed the recognition memory deficits as well as abnormal dendritic spine formation in the mouse model of ATR-X syndrome (Yamaguchi et al., 2018). Given these results, sigma-1 receptor agonists are a promising therapeutic option for alleviating various symptoms of intellectual disability, like those observed in ATR-X syndrome.

Interestingly, both sigma-1 receptor agonist and antagonist reversed the impaired recognition memory in alcohol-dependent mice during the withdrawal (Meunier et al., 2006a). These surprising results may be explained by the down-regulation of the increased level of sigma-1 receptors. The sigma-1 receptor agonists normalized the receptor over-expression but led to the accumulation of the protein in the terminal sites. On the other hand, sigma-1 receptor antagonists decreased the receptor binding in the microsomal fractions (Meunier et al., 2006a). Moreover, another study showed that the blockade of the sigma-1 receptor by AZ66 improved the recognition memory in methamphetamine-treated mice (Seminero et al., 2013). AZ66 significantly abolished striatal dopamine depletions and dopamine transporter reductions caused by methamphetamine (Seminero et al., 2013). This suggests that stimulating sigma-1 receptor leads to anti-amnesic effect in the case of memory impairments caused by decreased dopamine levels. Considering all reports, in this specific type of memory, a balanced modulation of sigma-1 receptor activity seems to be crucial. Studies suggest that depending on the cause of memory deficit either stimulation or blockade of sigma-1 receptors is responsible for anti-amnesic effect.

5.4. Sigma-1 receptor and Alzheimer’s disease – preclinical studies

Sigma-1 receptor agonists showed their anti-amnesic properties in various behavioral tests assessing different types of memory impaired in Alzheimer’s disease. The first paper linking the sigma-1 receptor with Alzheimer’s disease was a work by Maurice and colleagues (Maurice et al., 1998). In this study, the selective sigma-1 receptor agonists (+)-pentazocine, PRE-084, SA4503 attenuated the learning and memory deficits after Aβ25-35 peptide infusion in the spontaneous alternation performance and passive avoidance tests (Maurice et al., 1998). Subsequent papers confirmed these findings showing that the administration of various sigma-1 receptor agonists like PRE-084, donepezil or (-)-MR22 restored the proper memory functioning in the Y-maze and passive avoidance tasks in the Aβ25-35 peptide-infused mice (Antonini et al., 2011; Maurice, 2016; Meunier et al., 2006c). Their additive properties might have involved enhancing acetylcholine neurotransmission since donepezil acts as both acetylcholinesterase inhibitor and agonist of α7 subtype of nicotinic receptors as well as sigma-1 receptor agonist, increasing the acetylcholine release in the hippocampus and cortex. Moreover, the neuroprotective activity of pregnenolone sulfate (PREGS) might depend not only on the sigma-1 receptor but also on α7 nicotinic receptors. The compound improved the Aβ25-35-treated animals’ performance in the Morris water maze (Yang et al., 2012). ANAVEX2-73 also ameliorated the spatial and emotional learning deficits in mice injected with Aβ25-35 peptide, also showing neuroprotective properties by decreasing the level of lipid peroxidation and blocking tau hyperphosphorylation (Lahmy et al., 2013; Villard et al., 2011a). However, as mentioned earlier, this effect may be mediated not only via sigma-1 receptors, but also muscarinic receptors. The same results were observed for another aminotetrahydrofuran derivative, which stimulates sigma-1 receptors – ANAVEX1-41 (Villard et al., 2009). Moreover, a compound with a similar pharmacological profile, AF710B, improved recognition and spatial memory in aged McGill-R-Thy1-APP rats by reducing amyloid plaques, normalizing the microglia distribution and synaptophysin levels (Hall et al., 2018). Fluoxetine treatment also reversed memory and learning deficits in the active avoidance task in the nucleus-basalis-lesioned rats (experimental model of Alzheimer’s disease) (Ivkovic et al., 2004). Treatment of J20 amyloidogenic mice with another SSRI, fluvoxamine, improved memory in the object recognition task but did not affect other types of memory (assessed in the Y-maze and cheeseboard task) (Kim et al., 2016). Interestingly, the chronic administration of choline, which also binds to sigma receptors and potentiates IP3-evoked Ca2+ release (Bralolotu et al., 2019), improved the spatial memory in the Morris water maze in APP/PS1 mice and, like afobazole, reduced the microglial activation (Velasquez et al., 2019). This anti-amnesic effect of sigma-1 receptor activation may involve the inhibition of β-secretase activity (enzyme responsible for the cleavage of β-amyloid protein precursor) and subsequently reducing the Aβ production (Kim et al., 2018). The sigma-1 receptor positive modulator (OZP002) was also active in two mouse models of Alzheimer’s disease – it prevented the amyloid toxicity and memory impairment (Maurice et al., 2019).

What is worth mentioning, when the activity of sigma-1 receptors was blocked (either genetically or pharmacologically), the memory impairments, lipid peroxidation and Bax expression, caused by Aβ25-35 infusion, were more pronounced compared with mice with normal sigma-1 receptor function (Maurice et al., 2018). Moreover, APPSwelnd/sigma-1 receptor knockout mice exhibited more deficits in learning and memory processes than APPSwelnd mice (Maurice et al., 2018). This suggests that abnormal levels of sigma-1 receptors determine more severe pathology of Alzheimer’s disease mouse models. However, Yin with co-workers showed that heterozygous sigma-1 receptors knockout mice injected with Aβ25-35 peptide did not exert such memory deficits compared with wild-type mice after Aβ25-35-treatment (Yin et al., 2015). Here, the study suggests that the decreased expression of sigma-1 receptors is neuroprotective in Alzheimer disease mouse model against Aβ25-35-induced increased NR2B phosphorylation and subsequent neuronal cell death and spatial memory deficits (Yin et al., 2015). Considering the previously described research, where the stimulation of sigma-1 receptors exerted neuroprotective and anti-amnesic effects in Aβ25-35-treated animals, this difference might lie in time-dependent modulation of sigma-1 receptor activity. Up to 48 h post- Aβ25-35 infusion, the genetic or pharmacological decrease in the activity of the sigma-1 receptors protects neurons from NMDA-dependent excitability. However, after that time, not the blockade but the stimulation of sigma-1 receptors is responsible for the neuroprotective effect, probably via ERK and PI3K-Akt pathways (Yang et al., 2012) or antioxidant properties (Hyrsylyosto et al., 2013; Pal et al., 2012). The neuroprotection and beneficial properties of sigma-1 receptor activation in the Alzheimer disease may also result from the formation and stabilization of mushroom spines (Ryskamp et al., 2019).

5.5. Sigma-1 receptor ligands in clinical trials

Only one sigma-1 receptor agonist has been investigated in clinical trials concerning various memory deficits to the best of our knowledge (Table 2). ANAVEX2-73 (blarcamesine), a mixed sigma-1/muscarinic receptor ligand, is currently in the phase 2b/3 clinical trial for Alzheimer’s disease (ClinicalTrials.gov Identifier: NCT04314934 and ClinicalTrials.gov Identifier: NCT03790709) (Anavex Life Sciences Corp., 2022).
deficits, and sigma-1 receptor antagonists do not affect memory in rodents. Further studies showed that the blockade of these receptors might also mediate the anti-amnesic effect. The anti-amnesic effect in various animal models of memory impairments and subsequent expression of sigma-1 receptors, as well as the administration of different compounds and different doses used. However, given that the vast majority of studies demonstrated that sigma-1 receptor antagonists are ineffective in rodent models of memory deficits, we can conclude that the sigma-1 receptor should be activated to achieve anti-amnestic effects. The anti-amnestic effect of sigma-1 receptor agonists may be the result of normalizing the disrupted glutamatergic and cholinergic neurotransmission and increasing BDNF and ERK levels. Moreover, only the sigma-1 receptor knock-out female mice had impaired memory. However, considering males, the research results were ambiguous, showing both impaired and normal memory functioning. The activity of sigma-1 receptor ligands in tests assessing the anti-amnesic effect is summarized in Table 4.

6. Discussion and conclusions

Sigma-1 receptors are unique and riveting therapeutic targets with exciting features. The last decades brought a new perspective on these proteins, first identified as a type of opioid receptors, and now classified as chaperone proteins. They outstand among other receptors and proteins as:

• their sequence is quite different from other mammalian proteins, suggesting that their role in cells may be irreplaceable,
• they are widely distributed in the organism, not only in the central nervous system but also in the peripheral tissues and organs; thus, they may be a pharmacological target in various diseases with different etiology and phenotypes,
• they bind ligands with very diverse structures implicating that multiple compounds may modulate the activity of sigma-1 receptors, and therefore they can be used in the treatment of various disorders,
• they can translocate from ER to different cell compartments, thus modulating the activity of ion channels, receptors, transporters, kinases, neurotrophic factors, and other proteins, rendering sigma-1 receptors as an important node in the inter-organelle communication in cells.

Moreover, we need to highlight that sigma-1 receptor agonists have a favorable safety profile – animal studies proved that stimulation of these receptors and their modulatory activity occurred only in the pathological conditions but not in the normal physiological state. Therefore, such compounds may have limited side effects, which is beneficial.

However, one of the problems we need to solve is establishing a standardized method to characterize the ligand’s intrinsic activity. Available methods and techniques are very diverse, most of them imperfect, often irreproducible, and do not give certainty whether a ligand is an agonist, antagonist, positive or negative allosteric modulator. So far, the function of sigma-1 receptor ligands is predominantly identified according to the specific in vitro or in vivo pharmacological effects compared to the phenotype of sigma-1 receptor knock-out animals or to the effects after administration of standard sigma-1 receptors ligands.

Therefore, if we want to design novel sigma-1 receptor-targeted molecules with a possible therapeutic use, a standard functional assay is necessary. However, we need to raise the issue that since the sigma-1 receptor is a chaperone and not a classical receptor, it does not possess a specific transduction system. Nevertheless, by convention, we still name sigma-1 protein as a receptor. Therefore, we should consider whether using the terms: agonist or antagonist concerning sigma-1 receptor is appropriate and if we should not use the general description and classify compounds binding to the sigma-1 receptor just as ligands/modulators which affect the downstream pathways/proteins.

On the other hand, given that sigma-1 receptors affect numerous molecular proteins, a careful and detailed investigation of their role in the organism needs to be done as their upstream, and downstream intracellular signal cascades may differ depending on the cell types and organs, or systems. Moreover, scientists need to clarify the exact role of sigma-1 receptors in the pathophysiology of each disease, where the administration of sigma-1 receptor ligands may be beneficial. Importantly, we cannot exclude the possibility that broad possible indications for sigma-1 receptor ligands, reported so far, might be due to the activation or inhibition of other proteins.

As mentioned above, the currently known sigma-1 receptor ligands bind not only to sigma-1 receptors but also interact with other receptors or channels. Given that one of the intriguing features of sigma-1 receptors is their ability to interact with various molecules, which sometimes differ entirely from each other in terms of the chemical structure, it may be challenging to design highly selective sigma-1 receptor ligands. However, it does not necessarily need to be a disadvantage in treating complex CNS diseases such as depression or cognitive impairments. In such cases, multimodal compounds targeting the sigma-1 receptor might be superior to highly selective molecules. Thus, a multi-treatment approach may be optimal and the most beneficial if we want to treat complex neurodegenerative diseases with multidimensional etiologies and phenotypes.

As mentioned in the introduction COVID-19 pandemic increased the prevalence of depression and memory disorders. Interestingly, the sigma-1 receptor agonists may be attractive and putative therapeutic candidates for the treatment of COVID-19, not only considering their
Table 4

| Potential modulator | Model of memory impairments | Animals | Animal Models/Tests | Reference |
|---------------------|----------------------------|---------|--------------------|-----------|
| **Sigma-1 receptor agonists** | | | | |
| (+)-SKF-10047 | Scopolamine | Rats | Passive avoidance | (Matsuno et al., 1995) |
| Igmesine, (+)-3-PPP, DTG | Scopolamine | Rats | Passive avoidance | (Earley et al., 1991) |
| (+)-SKF-10047, (±)-pentazocine | Scopolamine | Mice | Passive avoidance | (Matsuno et al., 1993b) |
| SA4503 | Scopolamine, ibotenic acid forebrain lesion | Rats | Passive avoidance | (Senda et al., 1996) |
| SA4503 | Scopolamine | Rats | Passive avoidance | (Matsuno et al., 1997) |
| (+)-SKF-10047, SA4503 | Scopolamine | Mice | Y-maze, water maze | (Senda et al., 1998a, b) |
| DHEA-S, PREG-S | Scopolamine | Mice | Y-maze, passive avoidance | (Maurice et al., 2001a) |
| PRE-084, SA4503 | Scopolamine | Mice | Y-maze, water maze | (Tottori et al., 2002) |
| OPC-14523 | Scopolamine | Rats | Morris water maze | (Tottori et al., 2002) |
| LS-1–137 | Scopolamine | Mice | Active avoidance, water maze | (Malik et al., 2015) |
| Fluoxetine | Nucleus basalis lesion | Rats | Active avoidance | (Ivoviov et al., 2004) |
| (+)-Pentazocine (+)-SKF-10047 | Scopolamine | Mice | Y-maze | (Hiramatsu et al., 2002; Hiramatsu and Hoshino, 2005, 2004) |
| **Sigma-1 receptor antagonists** | | | | |
| KT-95 | Scopolamine | Mice | Y-maze, passive avoidance | (Hiramatsu et al., 2006) |
| ANAVEX1-41 | Scopolamine | Mice | Y-maze, passive avoidance, water maze | (Espallergues et al., 2007) |
| ANAVEX2-73 | Scopolamine | Mice | Y-maze, passive avoidance | (Villard et al., 2011a) |
| Dimemorfan | Scopolamine | Mice | Passive avoidance, water maze | (Wong et al., 2003) |
| (+)-PPCC | 192G-saporin induced lesions, atropine sulfate | Rats | Morris water maze | (Antonini et al., 2009) |
| (+)-SKF-10047, (+)-pentazocine | I-NAME, 7-nitroindazole | Mice | Y-maze | (Mamiya et al., 2000) |
| (+)-SKF-10047, (+)-pentazocine, DTG | Dizocilpine | Mice | Y-maze, passive avoidance, elevated plus maze | (Maurice et al., 1994a) |
| DHEA-S | Dizocilpine | Mice | Y-maze, passive avoidance | (Maurice et al., 1997) |
| SA4503 | Dizocilpine | Mice | Y-maze, passive avoidance | (Maurice and Privat, 1997) |
| (+)-SKF-10047, SA4503 | Dizocilpine | Rats | Radial arm maze | (Zou et al., 1998) |
| SA4503, DHEA-S, PREG-S | Dizocilpine | Mice | Passive avoidance, water maze | (Noda et al., 2001) |
| DTG, (+)-pentazocine, (+)-SKF-10047 | Dizocilpine | Mice | Y-maze | (Maurice et al., 1994b) |
| PRE-084, SA4503 | Dizocilpine | Mice | Y-maze, passive avoidance | (Maurice et al., 2001a) |
| PRE-084, DHEA-S, PREG-S | Dizocilpine | Mice | Y-maze, passive avoidance | (Maurice et al., 2001b) |
| SA4503, (+)-pentazocine (+)-SKF-10047 | Phenecyclidine, dizocilpine | Mice | One-trial water-finding task | (Noda et al., 2001) |
| Donepezil, igmesine | Dizocilpine | Mice | Y-maze, passive avoidance | (Maurice et al., 2006) |
| ANAVEX2-73 | Dizocilpine | Mice | Y-maze, passive avoidance | (Villard et al., 2011a) |
| PRE-084 | Dizocilpine | Mice | Y-maze, passive avoidance | (Maurice et al., 1994d, Phan et al., 1999) |
| Fluvoxamine, SA4503, DHEA-S | Phenecyclidine | Mice | Object recognition task | (Ishihimoto et al., 2007) |
| Donepezil | Phenecyclidine | Mice | Object recognition task | (Kunitachi et al., 2009) |
| (+)-SKF-10047, (+)-pentazocine | p-Chloroamphetamine | Mice | Passive avoidance | (Matsuno et al., 1993b) |
| (+)-SKF-10047, DTG, (+)-3-PPP | p-Chloroamphetamine | Mice | Passive avoidance, water maze | (Matsuno et al., 1994) |
| PRE-084 | Nimodipine | Mice | Y-maze, passive avoidance, water maze | (Maurice et al., 1996) |
| (+)-SKF-10047, DTG, (+)-3-PPP | CDEP | Mice | Passive avoidance | (Matsuno et al., 1998b) |
| PRE-10,047, DTG | Repeated CO exposure | Mice | Y-maze, passive avoidance | (Maurice et al., 1994a) |
| DHEA | Repeated CO exposure | Mice | Y-maze, passive avoidance | (Maurice et al., 1994c) |
| Donepezil, igmesine | Repeated CO exposure | Mice | Y-maze, passive avoidance | (Maurice et al., 2000) |
| Igmesine | Trimethyltin | Rats | Passive avoidance, radial arm maze | (O’Connell et al., 1996) |
| Pre-084, DTG | Trimethyltin | Mice | Passive avoidance | (Maurice et al., 1999a) |
| Dextromethorphan | Trimethyltin | Rats | Water maze, passive avoidance | (Shin et al., 2007) |
| Igmesine | Prenatal restraint | Rats | Y-maze, T-maze, water maze, passive avoidance | (Meunier et al., 2004) |
| Igmesine, DHEA | Prenatal cocaine treatment | Rats | T-maze, water maze, passive avoidance | (Meunier and Maurice, 2004) |
| Igmesine, PRE084 | Senescence-accelerated mouse | Mice | Y-maze, water maze, passive avoidance, open field | (Maurice et al., 1996) |
| PRE-084 | Normal aging | Rats | Morris water maze | (Maurice, 2001) |
| OPC-14523 | Normal aging | Rats | Morris water maze | (Tottori et al., 2002) |
| PRE-084 | Normal aging | Mice | Morris water maze | (Phan et al., 2003) |
| Cutaneusine | Rapid eye movement sleep deprivation | Rats | Passive avoidance | (Ramakrishnan et al., 2015b) |
| PRE-084 | Post-traumatic stress disorder | Rats | Water maze | (Li et al., 2016) |
| PRE-084 | Myocardial infarction | Mice | Y-maze | (Ito et al., 2013) |
| PRE-084, DTG | Brain ischemia | Mice | Y-maze, object recognition test, water maze | (Xu et al., 2015) |
| PRE-084 | Brain ischemia | Mice | Y-maze, object recognition test, water maze | (Xu et al., 2017) |
| DHEA | Brain ischemia | Mice | Y-maze, object recognition test, passive avoidance | (Tabuki et al., 2015) |
| SA4503 | Olfactory bullectomy | Rats | Cued and contextual fear-conditioning test | (Wang et al., 2007a, b) |
| Igmesine | Alcohol withdrawal | Mice | Object recognition task | (Meunier et al., 2006a) |
| (+)-pentazocine, PRE-084, SA4503, PREG-S, DHEA-S | β-Amyloid(25–35) peptide | Mice | Y-maze, passive avoidance | (Maurice et al., 1998) |

(continued on next page)
antidepressant-like and anti-amnestic properties, but also the role of sigma-1 receptors in the disease progression itself. The newest reports showed that the sigma-1 receptors are important in the replication of SARS-CoV-2 in cells and that the sigma-1 receptor is a promising therapeutic target for COVID-19 infection (reviewed in Hashimoto, 2021)). Nevertheless, growing evidence indicates that sigma-1 ligands can modulate multiple physiological and pathological processes, including excitotoxicity, calcium dysregulation, mitochondrial and endoplasmic reticulum dysfunction, oxidative stress, and inflammation. This feature may be associated with the significant involvement of the receptors in the pathomechanism of many CNS diseases, including affective and cognitive disorders. Thus, considering all published research, the sigma-1 receptor ligands, and predominantly agonists, might be the next generation of psychotherapeutic agents.

### Funding

This study has been conducted as part of a research project financed by the National Science Centre, Poland (grant number 2019/34/E/NZ7/00454) and Jagiellonian University Medical College (grants number N42/DBS/000117, N42/DBS/000140).

### References

Abraham, M.J., Fleming, K.L., Raymond, S., Wong, A.Y.C., Bergeron, R., 2019. The sigma-1 receptor and the hERG voltage-gated K+ channel revealed by atomic force microscopy and homogeneous time-resolved fluorescence (HTRF®). J. Biol. Chem. 289, 32353-32363. https://doi.org/10.1074/jbc.M114.583566.

Beggiato, S., Borelli, A.C., Borroto-Escuela, D., Corbicel, I., Tomasini, M.C., Marti, M., Antonelli, T., Tanganelli, S., Fuxe, K., Ferrari, L., 2017. Cocaine modulates allosteric D2(2)-e(-) receptor-receptor interactions on dopamine and glutamate nerve terminals from rat striatum. Cell. Signal. 40, 116-124. https://doi.org/10.1016/j.cellsig.2017.09.007.

Beheney, A.A., Yancy, I.E., Shuster, A.M., Seredenin, S.B., Petrow, A.V., Cuebas, V., 2013. Stimulation of sigma receptors with afobazole blocks activation of Microglia and reduces toxicity caused by amyloid-β25-35. J. Pharmacol. Exp. Ther. 347, 458-467. https://doi.org/10.1124/jpet.113.208348.

### Table 4 (continued)

| Potential modulator | Model of memory impairments | Animals | Animal Models/Tests | Reference |
|---------------------|----------------------------|---------|---------------------|-----------|
| Donepezil, PRED84 | β-Amyloid(25-35) peptide | Mice | Y-maze, passive avoidance | (Meunier et al., 2006c) |
| ANAVEX-41 | β-Amyloid(25-35) peptide | Mice | Y-maze, passive avoidance, radial arm maze | (Villard et al., 2009) |
| ANAVEX-23 | β-Amyloid(25-35) peptide | Mice | Y-maze, passive avoidance, | (Villard et al., 2011a) |
| Dimemorfan | β-Amyloid(25-35) peptide | Mice | Passive avoidance, water maze | (Wang et al., 2003) |
| Fluvoxamine | J20 amyloidiogen mouse model of Alzheimer’s disease | Mice | Object recognition test | (Kim et al., 2018) |
| Choline | APP (amyloid precursor protein)/PS1 (presenilin1) mice | Mice | Morris water maze | (Velazquez et al., 2019) |
| (−)-MR22 | Cholinergic lesion and amyloid infusion | Rats | Morris water maze, radial arm water maze | (Antonini et al., 2011) |
| ANAVEX-23/73 | Injection of Aβ/25-35 peptide | Mice | spontaneous alternation in the Y-maze, passive avoidance test, object recognition task | (Lahny et al., 2013) |
| AP710B | McGill-R-Thy1-APP transgenic (Tg) rats | Object recognition test, Morris water maze | (Hall et al., 2018) |
| ANAVEX-23/73, PRE-084 (−donepezil) | Injection of Aβ/25-35 peptide | Mice | spontaneous alternation in the Y-maze, passive avoidance test | (Maurice, 2016) |
| PREG-5 | Injection of Aβ/25-35 peptide | Mice | Morris water maze | (Yang et al., 2012) |
| SA4503 | α-thalassemia X-linked intellectual disability | Mice | Barnes maze, Y-maze, object recognition task | (Yamaguchi et al., 2018) |

### Sigma-1 receptor antagonists

| Sigma-1 receptor antagonists | | | | |
|-------------------------------|------------------------|-----------------|-----------------|---|
| BD1047 | Alcohol withdrawal | Mice | Object recognition test | (Meunier et al., 2006a) |
| AZ66 | Methamphetamine | Mice | Object recognition test, passive avoidance task | (Seminero et al., 2013) |

### Sigma-1 receptor modulators

| Sigma-1 receptor modulators | | | | |
|-----------------------------|------------------------|-----------------|-----------------|---|
| 0PZ2002 | Scopolamine, Inhibition of Aβ/25-35 peptide | Mice | T-maze, water-maze learning, passive avoidance, object recognition test | (Maurice et al., 2019) |
| EIR | Scopolamine | Mice | Y-maze, passive avoidance | (Zvejniece et al., 2014) |

Anavex Life Sciences Corp. 2021d. ClinicalTrials.gov Identifier: NCT04575259 [WWW Document]. URL https://clinicaltrials.gov/ct2/show/NCT04575259?

Anavex Life Sciences Corp. 2021b. ClinicalTrials.gov Identifier: NCT04314934 [WWW Document]. URL https://clinicaltrials.gov/ct2/show/NCT04314934?

Anavex Life Sciences Corp. 2020. ClinicalTrials.gov Identifier: NCT04575259 [WWW Document]. URL https://clinicaltrials.gov/ct2/show/NCT04575259?

Anavex Life Sciences Corp. 2021a. ClinicalTrials.gov Identifier: NCT03941444 [WWW Document]. URL https://clinicaltrials.gov/ct2/show/NCT03941444?

This study has been conducted as part of a research project financed by the National Science Centre, Poland (grant number 2019/34/E/NZ7/00454) and Jagiellonian University Medical College (grants number N42/DBS/000117, N42/DBS/000140).
Beraki, S., Diaz-Heijtz, R., Tai, F., Berardi, F., Ferorelli, S., Abate, C., Pedone, M.P., Colabufo, N.A., Contino, M., Chen, L., Dai, X.-N., Sokabe, M., 2006. Chronic administration of Charalampopoulos, I., Tsatsanis, C., Dermitzaki, E., Alexaki, V.-I., Castanas, E., Carnally, S.M., Johannessen, M., Henderson, R.M., Jackson, M.B., Edwardson, J.M., Cantarella, G., Bucolo, C., Di Benedetto, G., Pezzino, S., Lempereur, L., Calvagna, R., Brimson, J.M., Safrany, S.T., Qassam, H., Tencomnao, T., 2018. Dipentylammonium phencyclidine on cognition and gene expression in C57BL/6 mice. Int. J. Neurosci. 118, 1051–1058. https://doi.org/10.1080/00207454.2017.1391418.

Collina, S., Rui, M., Stotani, S., Bignardi, E., Rossi, D., Curti, D., Giordanetto, F., Crott Crouzier, L., Couly, S., Roques, C., Peter, C., Belkhiter, R., Arguel Jacquemin, M., 2007. Protective effects of the σ1 receptor agonist ANAVEX-1801 in a mouse model of neuropharmac.2005.10.015.

Brailoiu, E., Chakraborty, S., Brailoiu, G.C., Unterwald, E.M., Abood, M.E., Taylor, C.W., Dennis, W., Sollenberger, J.R., 1994. Neuronal activity is revealed gender-related anxiety, depressive-like and memory alterations. J. Neurosci. 24, 7351–7359. https://doi.org/10.1083/jneurosci.24.21.7351.

Choi, S.-R., Roh, D.-H., Yoon, S.-Y., Kang, S.-Y., Moon, J.-Y., Kwon, S.-G., Choi, H.-S., Han, H.-J., Beitz, A.J., Oh, S.-B., Lee, J.-H., 2013. Spinal σ1 receptor agonists activate NADPH oxidase 2 leading to the induction of pain hypersensitivity in mice and mechanical allodynia in neuropathic rats. Pharmacol. Res. 74, 56–67. https://doi.org/10.1016/j.phrs.2013.02.025.

Church, J., Fletcher, E.J., 1995. Blockade by sigma site ligands of high voltage-activated Ca2+ channels in rat and mouse cultured hippocampal pyramidal neurons. Br. J. Pharmacol. 116, 2801–2810. https://doi.org/10.1111/j.1368-9462.1995.tb15929.x.

Colabo, N.A., Contino, M., Inglese, C., Niso, M., Perrone, R., Roperto, S., Roperto, F., 2009. In vitro and ex vivo characterization of sigma-1 and σ2 receptors: agonists and antagonists in biological assays. Cent. Nerv. Syst. Agents Med. Chem. 9, 161–171. https://doi.org/10.1080/10611860802590316.

Cortes, D.,rial, S., Rapetti-Maas, L.R., Pisani, D.F., Lordi, C., Pellissier, B., Martin, P., Chevet, E., Borgese, F., Soriani, O., 2011. SIGIRP regulates hERG channel expression through a post-translational mechanism in leukemia cells. J. Biol. Chem. 286, 27947–27958. https://doi.org/10.1074/jbc.M111.227038.

Crouzier, L., Coudy, S., Roques, C., Peter, C., Belkhiter, R., Arguel Jacquemin, M., Bonetto, A., Delprat, B., Maurice, T., 2020. Sigma-1 (σ1) receptor activity is necessary for physiological brain plasticity in mice. Eur. Neuropharmacol. 39, 29–45. https://doi.org/10.1016/j.euroneuro.2020.08.010.
Gundlach, A.L., Largent, B.L., Snyder, S.H., 1986. Autoradiographic localization of sigma receptors in cerebral cortex: a medicinal chemistry perspective. J. Med. Chem. 29, 7962-7966.

Feltmann, K., Borroto-Escuela, D.O., Rüegg, J., Jurnak, F. L., de Oliveira Sergio, T., Gromek, K.A., Suchy, F.P., Meddaugh, H.R., Wrobel, R.L., LaPointe, L.M., Chu, U.B., Freeman, F.M., Young, I.G., 2001. Involvement of the sigma-1 receptor regulator. Science 293, 934-937. https://doi.org/10.1126/science.1066125.

Francardo, V., Bez, F., Wieloch, T., Nissbrandt, H., Ruscher, K., Cenci, M.A., 2014. Pharmacological stimulation of sigma-1 receptors has neuroprotective effects in experimental parkinsonism. Brain 137, 1998-2004. https://doi.org/10.1093/brain/azt113.

Francardo, V., Geva, M., Deniz, Q., Steiner, L., Hayden, M.R., Cenci, M.A., 2019. Sigma-1 receptors suppress multiple aspects of microglial activation. Glia 57, 744-754.

Hall, A.A., Herrera, Y., Ajmo, C.T.J., Cuevas, J., Pennykaker, K.R., 2009. Sigma receptors suppress multiple aspects of microglial activation. Glia 57, 744-754.

Hall, I., Iulita, M.F., Gubert, P., Flores Aguilar, L., Ducatenzeiler, A., Fisher, A., Cuello, A.C., 2018. APT7108, an M1-sigma-1 receptor agonist with long-lasting disease-modifying properties in a transgenic rat model of Alzheimer’s disease. J. Neurochem. 146, 221-234. https://doi.org/10.1111/jnc.14572.

Hampl, A., Trender, W., Chamberlain, S.R., Jolly, A.E., Grant, J.E., Patrick, F., Mazibuko, N., Cr, S., Barnby, J.M., Hellyer, P., 2021. Cognitive de ciites in people who have recovered from COVID-19. EclinicalMedicine 100, 101044. https://doi.org/10.1016/j.eclinm.2021.101044.

Hashimoto, K., 2009. Sigma-1 receptors and selective serotonin reuptake inhibitors: clinical implications of their relationship. Clin. Pharm. Ther. 88, 295-301. https://doi.org/10.1111/j.1365-2141.2009.01067.x.

Hashimoto, K., Tujita, Y., Iyo, M., 2003. Metyrapone-induced cognitive de cites in mice are improved by subsequent subchronic administration of fluvanoxamine: role of sigma-1 receptors. Neuropharmacology 50, 314-521. https://doi.org/10.1016/j.neuropharm.2005.08.007.

Hayashi, T., Tu, P.P., 2001. Regulating ankyrin dynamics: roles of sigma-1 receptors. Proc. Natl. Acad. Sci. U. S. A. 98, 491–496. https://doi.org/10.1073/pnas.98.1.491.

Hayashi, T., Tu, S.P., 2003. 11-receptors (11-binding sites) form raft-like microdomains and target lipid droplets on the endoplasmic reticulum: roles in endoplasmic reticulum lipid compartmentalization and export. J. Biol. Chem. 278, 718-725. https://doi.org/10.1074/jbc.M105107200.

Hayashi, T., Tu, S.P., 2007. Sigma-1 receptor chaperones at the ER: mitochondrial interface regulate Ca2+ signaling and cell survival. Cell 131, 596-610. https://doi.org/10.1016/j.cell.2007.08.036.

Hayashi, T., Tu, S.P., 2008. Sigma-1 receptor chaperones: a new class of protein chaperones that function in the ER. Faseb J. 22, 2014-2022. https://doi.org/10.1096/fj.08-115868.

Hayashi, T., Tu, S.P., 2012. The lifetime of UDP-galactose:ceramide galactosyltransferase is controlled by a distinct endoplasmic reticulum-associated degradation (ERAD) regulated by sigma-1 receptor chaperones. J. Biol. Chem. 287, 43156-43169. https://doi.org/10.1074/jbc.M112.380444.

Heston, J., Oliveira, A.L., Meister, B., Calheme, S., 2003. Large cholinergic nerve terminals on terminals of motoneurons and their relation to muscarinic receptor type 2. J. Comp. Neurol. 460, 476-486. https://doi.org/10.1002/cne.10648.

Hiramatsu, M., Hoshino, T., 2004. Involvement of e-opioid receptors and e receptors in memory function deterioration after head injury. Brain Res. 1030, 241-255. https://doi.org/10.1016/j.brainres.2004.09.009.

Hiramatsu, M., Hoshino, T., 2005. Improvement of memory impairment by (-)- and (+)-pentazocine via sigma, but not kappa opioid receptors. Brain Res. 1057, 72-80. https://doi.org/10.1016/j.brainres.2005.07.028.

Hiramatsu, M., Hoshino, T., Kameyama, T., Nabeshima, T., 2002. Involvement of e-opioid and receptors in short-term memory in mice. Eur. J. Pharmacol. 453, 91-98. https://doi.org/10.1016/s0014-2999(02)02989-9.

Hiramoto, M., Mizuno, N., Kanematsu, K., 2006. Pharmacological characterization of the ameliorating effect on learning and memory impairment and antiinociceptive effect of KT-95 in mice. Behav. Brain Res. 167, 219-225. https://doi.org/10.1016/j.bbr.2005.09.009.

Hiramoto, T., Mereu, M., Soto, P.L., Tanda, G., Katz, J.L., 2013. Self-administration of sigma-1 receptor agonists modulates dopamine transporter conformation and cocaine binding and may thereby potentiate cocaine self-administration in rats. J. Biol. Chem. 292, 11250-11261. https://doi.org/10.1074/jbc.M112.380444.

Horan, B., Gifford, A.N., Matsuoka, K., Mita, A., Ashby, C.R.J., 2002. Effect of SA4503 on e-opioid eceptors in mice. Adv. Exp. Med. Biol. 964, 251–298. https://doi.org/10.1006/abme.2001.6475.

Husi, M., Gino, A., Matsuoka, K., Mita, A., Ashby, C.R.J., 2002. Effect of SA4503 on e-opioid eceptors in mice. Adv. Exp. Med. Biol. 964, 251–298. https://doi.org/10.1006/abme.2001.6475.

Jaffe, H., Raubitschek, A., 1953. A transmitter-blocking agent: a study of the role of monoamine in the central nervous system. Am. J. Ment. Defic. 41, 219-237. https://doi.org/10.1177/0002629253413017.

K. Saatiack and K. Pyka
Neuroscience and Biobehavioral Reviews 132 (2022) 1114-1136

Katz, J.L., 2017. The sigma-1 receptor modulates dopamine transporter function in a cell culture model of Parkinson’s disease. J. Comp. Neurol. 460, 476–486. https://doi.org/10.1002/cne.10648.

Kato, Y., Iyo, M., 2005. Metyrapone-induced cognitive de cites in mice are improved by subsequent subchronic administration of fluvanoxamine: role of sigma-1 receptors. Neuropharmacology 50, 314-521. https://doi.org/10.1016/j.neuropharm.2005.08.007.

Kato, Y., Iyo, M., 2008. Sigma-1 receptor chaperones: a new class of protein chaperones that function in the ER. Faseb J. 22, 2014-2022. https://doi.org/10.1096/fj.08-115868.

Kawada, Y., Kishimoto, K., Ishibashi, T., Kato, Y., Iyo, M., 2011. Interaction of new antidepressants with sigma-1 receptor chaperones and their potential of neurite outgrowth in PC12 cells. Eur. J. Pharmacol. 677, 167-173. https://doi.org/10.1016/j.ejphar.2011.01.064.

Kato, I., Hirosaki, Y., Matsukawa, N., Nakano, M., Sunagawa, K., 2012. Decreased brain sigma-1 receptor contributes to the relationship between heart failure and depression. Cardiovasc. Res. 93, 33-40. https://doi.org/10.1093/cvr/cvr255.
Mammia, T., Noda, Y., Noda, A., Hirotsu, M., 2000. Effects of sigma receptor agonists on the impairment of spontaneous alternation behavior and decrease of cyclic GMP level induced by nitric oxide synthase inhibitors in mice. Neuropsychoendocrinology 29, 2391–2398.

Mancuso, R., Mengoli, S., Rando, A., Casas, C., Otta, R., Navarro, X., 2012. Sigma-1r agonist improves motor function and motoneuron survival in ALS mice. Neuroreport. J. Am. Sci. Exp. Neuroreport 9, 814–826. https://doi.org/10.1016/j.neuro.2011.01.040-y.

Mandelli, L., Wang, S.-M., Han, C., Lee, S.-J., Patkar, A.A., Masand, P.S., Par, C.-U., Serretti, A. 2017. The impact of a single nucleotide polymorphism in SIGHMAR1 on depressive symptoms in major depressive disorder and bipolar disorder. Adv. Ther. 34, 713–724. https://doi.org/10.1007/s12325-017-0482-2.

Mari, Y., Katnik, C., Cuesvas, J., 2014. ζ1-receptor inhibition of ASIC1a channels is dependent on a perturbations-toxin-sensitive G protein and an AKAP150/calcineurin complex. Neurochem. Res. 40, 2055–2067. https://doi.org/10.1007/s11064-014-1324-0.

Marrazzo, A., Caraci, F., Salinaro, E.T., Su, T.P., Copani, A., Ronsisvalle, G., 2005. Mancuso, R., Oliv

Matsuno, K., Matsunaga, K., Senda, T., Mita, S., 1993a. Increase in extracellular acetylcholine level in rat frontal cortex induced by N-acetyltyramine. Brain Res. Mol. Brain Res. 123, 66–75. https://doi.org/10.1016/0169-328X(93)90096-r.

Matsuno, K., Matsunaga, K., Mita, S., 1992. Increase of extracellular acetylcholine level induced in vitro by homocysteine and glutamate. Brain Res. Mol. Brain Res. 123, 311–315. https://doi.org/10.1016/0169-328X(92)90096-r.

Martina, M., Turcotte, M.E.B., Halman, S., Bergeron, R., 2007. The sigma-1 receptor antagonist SA4503, a selective PCP derivative, attenuates MK-801-induced impairment of learning in mice. Pharmacol. Biochem. Behav. 49, 859–869. https://doi.org/10.1016/j.phsol.2006.08.001.

Maurice, T., Su, T.-P., Paradise, D.W., Nabeshima, T., Privat, A., 1994d. Pre-O84, a sigma selective POP derivative, attenuates MK-801-induced impairment of learning in mice. Pharmacol. Biochem. Behav. 49, 859–869. https://doi.org/10.1016/j.phsol.2006.08.001.

Maurice, T., Su, T.-P., Paradise, D.W., Nabeshima, T., 1995. Prevention of nimodipine-induced impairment of learning by the selective sigma ligand Pre-084. J. Neural Transm. Gen. Sect. 102, 1–18. https://doi.org/10.1007/bf01275581.

Maurice, T., Roman, F.J., Su, T.P., Privat, A., 1996. Beneficial effects of sigma agonists on the age-related learning impairment in the senescence-accelerated mouse (SAM). Brain Res. 733, 219–230. https://doi.org/10.1016/S0006-8993(93)90565-3.

Maurice, T., Junien, J.L., Privat, A., 1997. Dehydrodopamineindorene sultate attenuates dixisul-coulned induced learning impairment in mice via σ-receptors. Behav. Brain Res. 83, 159–164. https://doi.org/10.1016/S0166-4328(97)80661-5.

Maurice, T., Su, T.-P., Privat, A., 1998. Sigma1 (σ1) receptor agonists and neurosteroids attenuate B25-35-amyloid peptide-induced amnesia in mice through a common mechanism. Neurosci. Biobehav. Rev. 22, 421–428. https://doi.org/10.1016/S0149-7634(97)00017-8.

Maurice, T., Phan, V.L., Noda, Y., Yamada, K., Privat, A., 1999a. The attenuation of learning impairments induced after exposure to CO or trimethyltin in mice by sigma (σ) receptor ligands involves both σ1 and σ2 sites. Br. J. Pharmacol. 127, 335–342. https://doi.org/10.1039/a00255j.phy.s60.116178.

Maurice, T., Phan, V.L., Urani, A., Kamei, H., Noda, Y., Nabeshima, T., 1999b. Neuroactive neurosteroids as endogenous effectors for the sigma(1) receptor: pharmacological evidence and therapeutic opportunities. Jpn. J. Pharmacol. 81, 125–155. https://doi.org/10.1253/jph.81.125.

Meunier, J., Maurice, T., 2004. Beneficial effects of the sigma1 receptor agonists on the impairment of passive avoidance response in mice: comparison with acetylcholinesterase inhibitors. Psychopharmacology (Berl.) 112, 134–141. https://doi.org/10.1007/s00213-010-1011-0.

Meunier, J., Guiri, D., Pirat, J.L., 2019. Neuroprotection in non-transgenic and transgenic mouse models of Alzheimer’s disease by positive modulation of σ1 receptors. Pharmacol. Res. 144, 315–330. https://doi.org/10.1016/j.phrs.2019.04.026.

Me, J., Pustemak, G.W., 2001. Molecular cloning and pharmacological characterization of the rat sigma receptor. Biochem. Pharmacol. 62, 349–355. https://doi.org/10.1016/s0006-2952(01)00666-9.

Meunier, J., Hayashi, T., 2010. Sigma 1 receptors regulate Bcl-2 expression by reactive oxygen species-dependent regulation of nuclear factor kappa B. J. Pharmacol. Exp. Ther. 332, 388–397. https://doi.org/10.1124/jpet.109.160960.

Meunier, J., Maurice, T., 2004. Beneficial effects of the sigma 1 receptor agonists igmesine and dehydrodopamine on learning impairments in rats prenatally exposed to cocaine. Neurotoxicol. Teratol. 26, 783–797. https://doi.org/10.1016/j.neurotox.2004.07.003.

Meunier, J., Gut, M., Récaens, M., Maurice, T., 2004. Attenuation by a sigma 1 (σ1) receptor agonist of the learning and memory deficits induced by a prenatal restraint stress in juvenile rats. Br. J. Pharmacol. 142, 689–700. https://doi.org/10.1038/sj.bjp.0705835.

Meunier, J., Demeilliers, B., Célier, A., Maurice, T., 2006a. Compensatory effect by soma-1 receptor stimulation during alcohol withdrawal in mice performing an object recognition task. Behav. Brain Res. 166, 166–176. https://doi.org/10.1016/j.bbr.2005.07.019.

Meunier, J., Jeni, J., Maurie, T., 2006b. Antiannexin and neuroprotective effects of donepezil against amyloid-beta25-35 peptide-induced toxicity in mice involve interaction with the sigma receptor. Br. J. Pharmacol. 149, 998–1012. https://doi.org/10.1038/sj.bjp.0706927.
Tagashira, H., Bhuiyan, M.S., Shioda, N., Fukunaga, K., 2014. Fluvoxamine rescues effects of mitochondrial Ca2+ mobilization and ATP production following cardiac hypertrophy. Biochem. Biophys. Acta - Gen. Subj. 1830, 3082–3094. https://doi.org/10.1016/j.bbabio.2013.12.019.

Takeda, K., Izumo, N., Suzuki, B., Karube, Y., Morikawa, T., Ishibashi, Y., Kameyama, T., 1998. Beneficial effects of mitochondrial Ca2+ transport and ATP production through σ1-receptor in hypertrophic cardiac myocytes. Life Sci. 95, 89–100. https://doi.org/10.1016/s0024-3205(98)00093-8.

Toheda, S.T., Hung, R.Q., Dibas, A., Krishnamoorthy, R.R., Dillon, G.H., Yorio, T., Tchedre, K.T., Huang, R.Q., Dibas, A., Krishnamoorthy, R.R., Dillon, G.H., Yorio, T., 2001a. The antidepressant-like activity of the sigma-1 receptor agonist cutamesine (SA4503) for the treatment of Alzheimer’s disease. Neuropharmacology 192, 108612. https://doi.org/10.1016/j.neuropharm.2021.108612.

Urani, A., Roman, F.J., Maurice, T., 2001. The antidepressant-like evaluation of σ1-receptor agonists in rats after chronic intracerebroventricular infusion of β-amyloid (1–40) peptide. Eur. J. Pharmacol. 486, 151–161. https://doi.org/10.1016/s0014-2999(01)01565-5.

Urfer, R., Moebius, H.J., Skoludzik, D., Santamarina, E., Sato, W., Mita, S., Muir, K.W., 2014. Phase II trial of the Sigma-1 receptor agonist cutamesine (SA4503) for recovery enhancement after acute ischemic stroke. Stroke 45, 3304–3310. https://doi.org/10.1161/strokeaha.114.005885.

van der Staay, F.J., Rutten, K., Erb, C., Blokland, A., 2011. Effects of the cognition imparer MK-801 on learning and memory in mice and rats. Behav. Brain Res. 220, 215–229. https://doi.org/10.1016/j.bbr.2011.11.1070.

Velaquez, R., Ferreira, E., Knowles, S., Fux, C., Rodin, A., Winslow, W., Oddo, S., 2019. Lifelong choline supplementation ameliorates Alzheimer’s disease pathology and associated cognitive deficits by attenuating microglia activation. Aging Cell 18, https://doi.org/10.1111/acel.13037.

Vetel, S., Foucault-Bruchard, L., Tronel, C., Buron, F., Vergote, J., Bordal, S., Routier, S., Serrière, S., Chalon, S., 2021. Neuroprotective and anti-inflammatory effects of a therapy combining agonists of nicotinic α7 and σ1 receptors in a rat model of Parkinson’s disease. Neurogen. Res. 16, 1099–1114. https://doi.org/10.1136/nij.2021.001961.

Wang, Y., Jiang, H., Chen, J.W., Cao, L., Li, C.-J., Chen, F.-C., 2003. Anti-amnesic effect of dimorfan in mice. Br. J. Pharmacol. 138, 944–949. https://doi.org/10.1038/sj.tpj.7005117.

Wang, D., Noda, Y., Tsuchekawa, H., Zhou, Y., Itozaki, M., Senzaki, K., Nitta, A., Nabeshima, T., 2007a. Role of N-methyl-D-aspartate receptors in antidepressant-like effects of σ1 receptor agonist 1-(3-acetylamino-4-chlorophenyl)-4-(3-pyridyl)-3,4-dihydro-2-[1H]-quinoline monomethanesulfonate. J. Pharmacol. Exp. Ther. 303, 1227–1234. https://doi.org/10.1124/jpet.102.024370.

Wang, D., Noda, Y., Tsuchekawa, H., Zhou, Y., Itozaki, M., Senzaki, K., Nitta, A., Nabeshima, T., 2007b. Phase II trial of the Sigma-1 receptor agonist cutamesine (SA4503) for the treatment of Alzheimer’s disease. Neuropharmacology 55, 310–322. https://doi.org/10.1016/j.neuropharm.2006.06.002.

Wang, J., Smith, S.B., 2019. A novel mechanism of sigma 1 receptor neuroprotection: modulation of miR-214-3p. Adv. Exp. Med. Biol. 1185, 463–467. https://doi.org/10.1007/978-3-030-27878-1_26.

Wang, H., Lin, Z., Chen, J.W., Chiou, C.-L., Cao, L.-J., Chen, F.-C., 2003. Anti-amnesic effect of dimorfan in mice. Br. J. Pharmacol. 138, 944–949. https://doi.org/10.1038/sj.tpj.7005117.
