T-786C Polymorphism of Endothelial Nitric Oxide Synthase Gene and Serum Level of Nitric Oxide in Nonsmoker and Nondiabetic Patients Suffering from Coronary Artery Disease

Yaghoubi AliReza, Khaki-Khatibi Fatemeh*, Zarghami Nosratallah, Rahbani-Nobar Mohammad and Porhassan Mohammad
Cardiovascular Research Center, Department of Clinical Biochemistry, Faculty of Medicine, Tabriz University of Medical sciences, Tabriz, Iran

Abstract

**Background:** Various mutations on endothelial nitric oxide synthase (eNOS) gene cause reduced production of NO, the endothelial relaxing factor, and may accelerate the process of atherosclerosis. The study designed to investigate the frequency of T-786C polymorphism of the gene in patients suffering from CAD in North West of Iran.

**Material and methods:** One hundred twenty subjects including 60 patients with angiographically diagnosed CAD and 60 age and sex matched CAD-free subjects as control were studied. The levels of Nitric oxide in the samples were measured with the Griess Method. The genotype studies were carried using allele specific PCR.

**Results:** Comparing with the control reduced levels of NO was noticed in the patient group (P<0.05) and significantly high frequency of eNOS -786C genotype was found in CAD patients (P<0.05).

**Conclusions:** The low levels of NO and increased frequency of T-786C polymorphism might be a risk factor in progression of coronary artery disease in the studied subjects.

**Keywords:** Coronary artery disease; Endothelial nitric oxide synthase gene; T-786C polymorphism; Nitric oxide

**Introduction**

Coronary artery disease (CAD) is the major cause of mortality and morbidity in most countries. Among many traditional risk factors for CAD development positive family history are now being considered as significant and novel risk factor. Atherosclerosis, an essential factor for the development of CAD, results from a defective endothelial function, which is ascribed mainly to an altered production of nitric oxide (NO), an important endothelium-derived relaxing factor [1]. NO is synthesized via a reaction that includes the conversion of L-arginine to L-citruline catalyzed by endothelial nitric oxide synthase (eNOS), which is one of the three isoforms of the enzyme [2]. eNOS is the product of eNOS gene, which is 21kb in size and consists of 26 exons [3]. Additionally, promoter region of the eNOS gene harbors several transcription factor binding sites, regulating gene expression [4].

Nitric oxide (NO) is a vasoactive substance and a major mediator of endothelium-dependent vasodilatation, which is synthesized and also released in the vascular endothelium [4]. Nitric oxide diffuses from the endothelium to the vascular smooth muscle cells, where it increases the concentration of cyclic guanosine monophosphate (cGMP) by stimulating soluble guanylyl cyclase, leading to vascular relaxation [5].

Nitric oxide (NO) is a free radical molecule that plays an essential role in numerous physiological actions, including vaso regulation, inhibition of platelet aggregation, and immunological reactions [6]. Endothelial nitric oxide synthase (eNOS), an isoform of NO-producing enzymes that is fairly specific to endothelial cells, has been found to play a prominent role in both angiogenesis and vasculogenesis [7].

Recently, the eNOS gene was studied extensively for genetic polymorphisms to elucidate its genetic role in cardiovascular diseases. As a result, a single-nucleotide polymorphism (SNP) in the promoter region, T-786C, was found to modify the promoter activity in vitro [8].

The eNOS gene is encoded by a gene (NOS3) located on chromosome 7q35-q36 [9]. A polymorphism in the 5’ flanking region of the NOS3 gene (T-786C) has been associated with coronary spasm among Japanese [8,10]. It is believed that these mutations might result in altered NO metabolism and impaired NO release, leading to increased vascular tone and elevation in blood pressure. One of the mutations in the eNOS gene is a result of a thymidine (T) being replaced by a cytosine (C) at nucleotide -786 (T-786C).

The aim of present study was to determine the prevalence of T-786C polymorphism of eNOS in CAD patient and control. Because of significant effect of diabetes and smoking on the NO metabolism the samples obtained from non-diabetic and non-smoking subjects.

**Materials & Methods**

**Subjects**

The CAD group included 30 females and, 30 males with a mean age of 58 years, ranging from 40-78 years. They had various degrees of stenosis in one or more of the main branch of coronary artery documented by coronary angiography. Patients with diabetes mellitus, renal disease, chronic obstructive pulmonary disease, hepatitis and smoker were excluded from the study group. The controls included 30 females, 30 males with a mean age of 57 years, ranging from 40-76 years.

*Corresponding author: Fatemeh Khaki Khatibi PhD, Cardiovascular Research Center, Department of Clinical Biochemistry, Faculty of Medicine, Tabriz University of Medical sciences, Tabriz, Iran, Tel: +98-914-4068385; Fax: +98-411-3364668; E-mail: fatemeh.khakikhatibi@yahoo.com

Received January 09, 2012; Accepted February 09, 2012; Published February 11, 2012

Citation: AliReza Y, Fatemeh KK, Nosratallah Z, Mohammad RN, Mohammad P (2012) T-786C Polymorphism of Endothelial Nitric Oxide Synthase Gene and Serum Level of Nitric Oxide in Nonsmoker and Nondiabetic Patients Suffering from Coronary Artery Disease. J Biotechnol Biomater 2:125. doi:10.4172/2155-952X.1000125

Copyright: © 2012 AliReza Y, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

DOI: 10.4172/2155-952X.1000125
years. The subjects proved to be healthy by health screening and had no obstructions in the coronary artery by angiography.

**Blood sampling**

Blood samples were collected in the morning by venipuncture after an overnight fast and were allowed to clot at room temperature for about 1 hour. Sera were separated from cells by centrifugation at 1500xg for 10 min and kept at -80°C and blood samples were stored at -20°C until analysis.

**Measurement of serum NO**

The levels of NO in serum were determined colorimetrically by Griess method [11]. In this method, NO undergoes a series of reactions with several molecules present in biological fluids including O₂, O₂⁻, and NO₃⁻. The final products of NO in vivo are nitrite (NO₂⁻) and nitrate (NO₃⁻). The relative proportion of NO₂⁻ and NO₃⁻ is variable and cannot be predicated with certainty. Thus the best index of total NO production is the sum of both NO₂⁻ and NO₃⁻. The method used in this study provides an accurate and convenient measurement of nitrate/nitrite concentration in a simple two-step process. The first step is conversion of nitrate to nitrite utilizing nitrate reductase. The second step is the addition of Griess reagent which converts nitrite into deep purple azo compound. Colorimetric measurement of the absorbance due to this azo chromophore accurately determines nitrate concentration.

**Molecular analyses**

DNA was extracted using standard phenol-chloro for procedure. Genotyping of T-786C, a newly developed allele-specific polymerase chain reaction (PCR) was used (Figure 1) [12]. The oligonucleotide primers used in the reaction are listed in (Figure 1). Artificial mismatches were included in the 2684T and 2684C primers as indicated in (Figure 1). Amplification was performed in a total volume of 20 µL containing 100 ng genomic DNA, 5 µM 2684T and 2684C primers, 1.6 µM Tₐ and Cₐ primers and 10 µL Master Mix. After a hot start at 96°C, amplification was achieved by 36 cycles at 94°C for 40 seconds, 54°C for 40 seconds, and 72°C for 45 seconds. The C and T alleles gave a 176bp and a 250bp product, respectively, with a 387bp common product (Figure 2).

**Statistical analysis**

Data were analyzed with t-test, expressed as mean ± SD and χ² test. Data were compared in the groups by using SPSS software version 16. P<0.05 was chosen as the level of significance. Allele frequencies were calculated from the genotype counts. The observed genotype counts were compared with those expected under Hardy-Weinberg equilibrium with χ² test.

**Results**

The characteristics of the case (CAD) as well as control group are summarized in (Table 1). NO differences were noticed between the mean values of age, sex and family history of CAD in patient and control groups. The percent of hypertensive subject in the patient groups was significantly higher than that of control group (p<0.05) (Table 1).

Comparing NO levels in the patient and control groups, significantly low levels of NO were noticed in the patients group (p<0.05) (Table 2).

Genotype frequencies of T-786C among the patient and control groups are shown in (Table 3). In the present study we found a marked difference in the frequency of the T-786C mutation of eNOS gene between patients and control groups. In total patients group, the frequency of T/T, C/T and C/C genotypes was 38.3%, 46.7% and 15% respectively. In the control the frequency of T/T, C/T and C/C genotypes was 38.3%, 46.7% and 15% respectively. The frequency of C allele was significantly higher in the patient group.

**Discussion**

Nitric oxide (NO) plays an essential role in regulating vascular tone and hemodynamic. NO stimulates endothelial proliferation and angiogenesis, thereby playing an important role in wound healing and microcirculation. In addition to NO inhibits the release of endothelin-1 (a vasoconstrictor) [13]. Besides being an endothelium derived vasodilator molecule, NO also has important physiological and pathological effects. It can be synthesized in most tissue and cells. Its most prominent roles in cardiovascular system are blood pressure regulation, inhibition of thrombocyte aggregation, leukocyte adhesion, smooth muscle cell proliferation, and LDL oxidation. The decreases in production and bioavailability are associated with events that accelerate development of atherosclerosis such as vasoconstriction, thrombocyte aggregation, migration of monocytes to the vascular wall, oxidized LDL.
they found a significant association between the T-786C mutation and CAD. Colombo et al. [19] showed a positive association between the polymorphisms and the extent of CAD in the Italian population. T-786C polymorphism was also reported as being related to coronary spasm in the Japanese by Nakayama et al. [20]. In this study high frequency of C/C genotype of T-786C was noticed in the patient group. The frequency of C allele was significantly higher in patient group. The results are in agreement with those reported by others.

According to Nakayama et al. [8], the -786C allele would be associated with a significantly reduced eNOS promoter activity. The reduced endothelial production of NO in the coronary arteries would predispose carriers of the C allele to coronary spasm. The coronary spasm might be more severe and prolonged in CC homoygotes, increasing the risk of CAD [21]. Also according popov et al. [22], report the eNOS T-786C polymorphism could serve as a possibility to differentiate high risk subgroups in heterogeneous population individual with cardiac diseases who need cardiac surgery with CPB.

The T-786C mutation in the promoter region of eNOS resulted in the reduction of eNOS promoter transcription rate [8], leading to the reduced NO production in blood vessels and endothelial dysfunction [23].

Conclusions

It was concluded that presence of the eNOS mutant allele reduces endothelial production of NO and may predispose the patients carrying the mutant allele to coronary spasm, hypertension and coronary artery diseases.

Since the study was limited to nonsmoker and no diabetic subjects the limitation of the study was small sample size.

References

1. Davignon J, Ganz P (2004) Role of endothelial dysfunction in atherosclerosis. Circulation 109: I102–I112.
2. Mayer B, Hemmens B (1997) Biosynthesis and action of nitric oxide in mammalian cells. Trends Biochem Sci 22: 477–481.
3. Marsden PA, Heng HH, Scherer SW (1993) Structure and chromosomal localization of the human constitutive endothelial nitric oxide synthase gene. J Biol Chem 268: 17476–17486.
4. Karantziouls-Fegaras F, Antoniou H, Lai SL (1999) Characterization of the human endothelial nitric-oxide synthase promoter. J Biol Chem 274: 3076–3093.
5. Moncada S, Higgs A (1993) The L-arginine-nitric oxide pathway. N Engl J Med 329: 2002–2012.
6. Gross SS, Wolin MS (1995) Nitric oxide: pathophysiological mechanisms. Annu Rev Physiol 57: 737–769.
7. Kimura H, Esumi H (2003) Reciprocal regulation between nitric oxide and vascular endothelial growth factor in angiogenesis. Acta Biochim Pol 50: 49–59.
8. Nakayama M, Yasue H, Yoshimura M, Shimasaki Y, Kugiyama K, et al. (1999) T-7863C mutation in the 5'-flanking region of endothelial nitric oxide synthase gene is associated with coronary spasm. Circulation 99: 2886–2870.
9. Nadaud S, Bonnardeaux A, Lathrop GM, Soubrier F (1994) Gene structure, polymorphism and mapping of the human endothelial nitric oxide synthase gene. Biochem Biophys Res Commun 198: 1027–1033.
10. Casas JP, Cavalleri GL, Bautista LE (2006) Endothelial nitric oxide synthase gene polymorphisms and cardiovascular disease: a huge review. Am J Epidemiol 164: 921–935.
11. M-Miranda K, G-Espey M, A-Wink D (2001) A rapid, simple spectrophotometric method for simultaneous detection of nitrate and nitrite. Nitric Oxide 5: 62–71.
12. Little S (1995) Amplification-refractory mutation system (ARMS) analysis of point mutation. In: Boyle AL, ed. Current Protocols in Human Genetics. New York, NY: John Wiley & Sons 9:8: 1–1
13. Ignarro LJ, Cirino G, Casini A, Napoli C (1999) Nitric oxide as a signaling molecule in the vascular system: an overview. J Cardiovasc Pharmacol 34: 879–886.

14. Elizalde M, Ryden M, van-Harmelen V (2000) Expression of nitric oxide synthase in subcutaneous adipose tissue of nonobese and obese humans. J Lipid Res 41: 1244–1251.

15. Scribner AW, Loscalzo J, Napoli C (2003) The effect of angiotensin-converting enzyme inhibition on endothelial function and oxidant stress. Eur J Pharmacol 482: 95-99.

16. Hyndman ME, Parsons HG, Verma S, Bridge PJ, Edworthy S, et al. (2002) The T-786 C mutation in endothelial nitric oxide synthase is associated with hypertension. Hypertension 39: 919–922.

17. Rossi GP, Cesari M, Zanchetta M, Colonna S, Maiolino G, et al. (2003) The T-786 C endothelial nitric oxide synthase genotype is a novel risk factor for coronary artery disease in Caucasian patients of the GENICA study. Am J Cardiol 41: 930–937.

18. Nakayama M, Yasue H, Yoshimura M, Shimasaki Y, Ogawa H, et al. (2000) T-786 C mutation in the 5′-flanking region of the endothelial nitric oxide synthase gene is associated with myocardial infarction, especially without coronary organic stenosis. Am J Cardiol 86: 628–634.

19. Colombi MG, Paradossi U, Andreassi MG, Botto N, Manfredi S, et al. (2003) Endothelial nitric oxide synthase gene polymorphisms and risk of coronary artery disease. Clin Chem 49: 3389–3395.

20. Kunnas TA, Ilveskoski E, Niskakangas T, Laippala P, Kajander OA, et al. (2002) Association of the endothelial nitric oxide synthase gene polymorphism with risk of coronary artery disease and myocardial infarction in middle-aged men. J Mol Med 80: 605–609.

21. Jaramillo PC, Lanas C, Lanas F (2006) T-786C polymorphism of the endothelial nitric oxide synthase gene in Chilean subjects with coronary artery disease and controls. Clin Chim Acta 387: 105-108.

22. Popov AF, Henker C, Schmitto JD, Wiese CH, Coskun KO, et al. (2010) Clinical relevance of eNOS T-786C polymorphism for hospital mortality and morbidity in cardiac surgical patients. J Cardiovasc Surg (Torino) 51: 265-72.

23. Kim JJ, Bae J, Lim SW, Cha DH, Cho HJ, et al. (2007) Influence of endothelial nitric oxide synthase gene polymorphisms (T-786C, 4a4b, G894T) in Korean patients with coronary artery disease. Thromb Res 119: 579-585.