Effect of Organic Loading Rate on the Biological Performance of the Thermophilic Anaerobic Membrane Bioreactor Treating Pulp and Paper Primary Sludge

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Research Article

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Abstract

Waste-to-energy or value-added products have been increasingly considered in many pulp and paper mills (PPMs) worldwide. However, developing appropriate conversion technologies is a major challenge in transforming PPMs wastes into biofuels or value-added biomaterials. In the present study, a long-term (320 d) anaerobic digestion of primary sludge of a thermomechanical pulp mill (TMP) was carried out for the first time in a thermophilic anaerobic membrane bioreactor (ThAnMBR). Effect of organic loading rate (OLR) in the range of 2.5–6.8 kg-COD/m$^3$ d and hydraulic retention times (HRT) of 3–8 d on the process performance was investigated. Under various OLRs, stable biogas productions were obtained, and the best results were achieved with lower OLR (2.5 kg-COD/m$^3$ d) and higher HRT (8 d), at biogas yields of 189 L biogas/kg MLSS fed. However, it was found that biogas production and sludge biomass degradation decrease when the organic loading rate increases. The proportion of sludge reduction ranged from 28.9 to 46.7% depending on the applied OLRs. Despite varying OLRs, stable membrane performance was obtained, where the required membrane flux was easily maintained during the reactor operation. In this study, also the properties of digestate and membrane permeates were studied under different operating conditions, and they fluctuated to some extent with OLR. ThAnMBR is a promising new technology for pulp and paper mill primary sludge treatment.

1. Introduction

Pulp and paper mill sludge (PPMS) represents a large part of the industrial waste and contains a high portion of the organic matter (Elliott and Mahmood, 2012). Sludge waste is produced from both virgin and recycled paper production processes (Simão et al., 2018). However, this substrate is usually undesirable and needs to be treated or disposed of in an environmentally acceptable manner. The common handling strategies of the waste sludge are landfilling, incineration, and recycling in the paper making process. The first two practices place a significant financial burden on the forestry industry because of the high capital expenses and high energy required to dry large amounts of water before burning the waste sludge (Meyer et al., 2018). Reportedly, sludge disposal takes a large part of the wastewater treatment facility budget ($\approx$ $30 per wet ton), which may reach approximately 60% of the total cost (Elliott and Mahmood, 2012; Park et al., 2012). In addition to the high expenses, these traditional handling approaches have been characterized by several problems such as air (incineration) and water (landfill leachate) pollution. Conversely, PPM sludge contains valuable molecules (25–75% carbohydrate). For example, Kraft PPM primary sludge is composed of woody materials such as cellulose (58 wt%), hemicelluloses (12 wt%), and lignin (20 wt%), along with inorganic materials that have been used in the pulping process (Bayr and Rintala, 2012). While, the secondary sludge is composed of microbial biomass, cellulose (36–50 wt%), and non-degradable lignin's compounds (19–44 wt%) (Likon and Trebše, 2012; Kinnunen et al., 2015). The quantity of sludge production varies depending on the pulping processes, but primary sludge represents the majority of the total solids compared to secondary sludge (Bajpai, 2015). The application of traditional sludge treatment processes leads to the loss of these
precious resources accompanied by environmental pollution (Bayr et al., 2013; Hazarika and Khwairakpam, 2018).

Several technologies, including direct combustion (Manwatkar et al., 2012; Pio et al., 2020a), pyrolysis (Yin et al., 2021), gasification (Pio et al., 2020b), and hydrothermal liquefaction (Zhang et al., 2021) processes have evolved to convert sludge waste into fuels. In addition to power generation, these technologies were characterized by reducing the volume of sludge, destroying harmful pathogens, and stabilizing heavy metals (Liew et al., 2012). However, the high initial investment cost and requirements for the pre-dewatering process hamper their wide application (Liew et al., 2012). Among the treatment alternatives, one strategy that has been recognized worldwide as a feasible option to improve the energy efficiency of PPM sludge is anaerobic digestion (AD). Anaerobic digestion (AD) offers a promising alternative relative to the above-mentioned options due to its reduced environmental footprint, small reactor size, and requires no sludge dewatering, where sludge dewatering represents a considerable economic burden. Also, the residual organic matter and nutrients that are retained in effluents (digestate) can be returned and reused for different applications (Veluchamy and Kalamdhad, 2017a).

The substrate's content of sugar, fat, and protein controls its anaerobic digestion potential and energy production; accordingly, PPM sludge appears an appropriate substrate for AD due to its high carbohydrate and high-water content. Also, one of the attractive features of PPM sludge is that the raw material cost is zero, no excessive pre-treatment is required due to previously processed biomass, and the possibility of using the existing pulp or paper mill equipment (Gurram et al., 2015). AD has been applied extensively and for a wide range of organic substrates. Nonetheless, the literature shows very limited studies treating PPM sludge as an anaerobic digestion substrate. Yet, studies on PPM sludge digestion are in their infancy and no industrial application has been reported to date. This may be due to long residence times (20–60 d), and low yield of bio-methane and bio-hydrogen due to its low degradability (30–50%) (Lin et al., 2009; Kinnunen et al., 2015), plus the biomass separation problems of the traditional anaerobic digestion processes (Dereli et al., 2012). Additionally, effluent quality from anaerobic treatment is usually poorer than that from aerobic treatment and needs further polishing. This defect might limit the application of anaerobic technology, especially in places that require wastewater reclamation and reuse, such as an integrated forest biorefinery (IFB), where the resulting wastes and by-products are recycled and utilized as a resource.

Compared to traditional anaerobic digestion, the anaerobic membrane bioreactor (AnMBR), a combination of anaerobic digestion and membrane filtration, is a relatively new technology for the treatment of municipal and industrial wastes but has demonstrated its superiority over conventional anaerobic biological processes in terms of higher effluent quality for reuse/reclamation, recovery of most of the potential energy in biodegradable waste streams, and decoupling of solids retention time (SRT) from hydraulic retention time (HRT) (Bokhary et al., 2020). By integrating the membrane process for microbial biomass retention, anaerobic microorganisms can proliferate without being washed out of the system. This leads to higher biomass concentration in AnMBR, which may enhance biogas yield compared to conventional processes. Also, decoupling the HRT and SRT in AnMBRs can achieve a
significant reduction in reactor volumes, thereby reducing capital costs. Thus, AnMBR could be a promising option for primary sludge digestion. However, to the authors’ knowledge, no study has ever been reported on the treatment of pulp and paper primary sludge using thermophilic anaerobic membrane bioreactor (ThAnMBR) technology. Of the few experimental studies involving conventional anaerobic digestion of PPM sludge, only two studies tested PPM primary sludge (Jokela et al., 1997; Bayr and Rintala, 2012), while other studies focused on the treatment of secondary sludge, or sludge mixtures (Karlsson et al., 2011; Lin et al., 2011; Saha et al., 2011; Bayr and Rintala, 2012). Most PPM sludge studies have been conducted at mesophilic temperatures using batch assays, but few have been conducted in thermophilic conditions (Teghammar et al., 2012; Lopes et al., 2018). Bayr and Rintala (2012) examined AD of kraft mill primary sludge under thermophilic conditions using a semi-continuous bioreactor (CSTR) but at relatively long HRTs and low OLRs. OLR is an essential parameter and limited information is available about the steady-state performance of the ThAnMBR for biogas production under high OLRs.

Taking into account all the advantages of AnMBR mentioned above, the current research program focuses on the application of a relatively new ThAnMBR technology for biogas production from primary sludge of thermomechanical pulp (TMP) aiming for improving biogas yield and solids reduction and reducing HRT without the risk of washing out the microbial population. This study aimed to investigate the possibility of performing AD of TMP primary sludge at low HRT and high OLRs. Of interest is elucidating an optimal organic loading rate that maximizes bio-methane production and solids reduction, minimizes membrane fouling propensity, and produces permeate with suitable quality for reuse in a pulp and paper biorefinery.

2. Materials And Methods

2.1. Feed and inocula

PPM sludge was obtained from a local Canadian thermomechanical pulp mill. Sludge received from this mill was dewatered sludge from a primary clarifier. The mill uses a mixture of hardwood and softwood as raw materials for pulp production. After delivered to the laboratory, the collected PPMS was stored in the refrigerator at 4°C before being prepared and used as a substrate. Primary sludge was mixed with tap water to create a slurry with a certain concentration as a feeding substrate. As primary sludges are relatively low in nitrogen and phosphorus (Hagelqvist 2013), ammonium chloride and dipotassium phosphate were added in COD: N: P of 100:2.6:1 ratio to improve its nutrients content. Table 1 shows the characteristics of the primary sludge and inoculum used in this study. The inoculum was collected from a thermophilic anaerobic membrane bioreactor operated for one year by a member of our research group (Jiang, 2018), and the system was treating secondary sludge from the local pulp and paper mill.

2.2. Laboratory scale submerged AnMBR setup and operation
The submerged AnMBR setup consists of a CSTR reactor with an active volume of 6.5 L and flat sheet microfiltration (MF) membrane module with an active surface area of 0.03 m$^2$ (10 cm × 15 cm on each side of the module) and pore size of 0.1µm. The membrane material was a polyvinylidene fluoride (PVDF) (DAFU Membrane Technology Co., P.R. China). The bioreactor temperature was maintained at 50°C by circulating the warm water from an adjustable water bath through the water jacket of the reactor. The pH of the bioreactor was controlled at 7.2 ± 0.1 using a NaOH solution. For reactor feeding, the sludge was pumped into the bottom of the bioreactor by a feeding pump (Iwaki Magnet Pump, Iwaki co., LTD. Tokyo Japan). The pump was controlled by a water level sensor (Madison Co., USA), and a controller (Flowline, USA) to maintain a constant liquid level in the bioreactor. Figure 1 shows a schematic diagram of the ThAnMBR setup used in this study. After feeding the reactor with the required amount of inoculum and primary sludge, it was flushed with nitrogen gas for 5 min to create anaerobic conditions. Permeate was withdrawn from the system using a peristaltic pump (Masterflex C/L®, Cole-Parmer, Montreal, Canada), and collected in its tank for further analysis. The peristaltic pump was controlled by a timer that operated in three minutes on and two minutes off mode. Transmembrane pressure was monitored with a pressure gauge (Omega, model 656201BA4CD3ACD1. Korea). To sustain the required SRT, reactor sludge was taken out from CSTR once a day during the daily measurement of the biogas production rate, reactor pH, and TSS of digestate.

For fouling control, the membrane was sparged by biogas. This was accomplished using a stainless-steel pipe diffuser placed below the membrane unit on each side, and the biogas was recirculated by two gas recycling pumps (Masterflex Console Drive, Model 7520-40, Thermo Fisher Scientific, USA). The biogas sparging rate was fixed at 3.76 ± 0.08 L/min during the experiments by adjusting the digital pump speeds. At the bottom of the reactor, a gentle mixing was applied using a magnetic stirrer blade (Barnstead Thermolyne, Cimarec Plate Stirrer, Dubuque, IA, USA) to keep the reactor in suspension. For biogas composition measurement, the biogas samples were taken from the top reactor’s port, while the biogas produced was collected in a graduated cylinder using a water displacement method. The reactor was operated at OLRs between 2.5 and 6.8 kg-COD/m$^3$ d and hydraulic retention times (HRTs) in the 3–8 d range.

2.3. Analytical methods

2.3.1. Primary sludge samples, reactor feeds, and inoculum

Primary sludge was characterized for total solids (TS), total suspended solids (TSS), and volatile solids (VS) according to standard methods (Baird et al., 2017). TSS of the reactor feed and digestate was analyzed three times a week, while pH was monitored daily. Total COD of the permeate and feed were analyzed every alternate week, using HR COD test vials (K-7365, 0–1500 ppm) and a COD reactor (Hanna Instruments), followed by absorbance measurement using a colorimeter (Hach DR 2800 Spectrophotometer). Ash content in the sludge sample was measured after dry oxidation of PS at 575°C using a muffle furnace, according to standard methods for biomass analysis (Cai et al., 2017). The pH was measured using an Oakton pH 700 benchtop meter. Elemental composition (C, N, H, S, and O) was
measured by an elemental analyzer (Elementar Vario EL). The inoculum was analyzed for TS, TSS, VS, COD, ash, pH, and elemental content as per the methods mentioned previously. Table 1 shows the results of the primary sludge, feed, and inoculum characterization. Air dry primary sludge contains 96.32 ± 0.28 % total solids, 93.93 ± 0.24% volatile solids, and 6.07 ± 0.244 % fixed solids. The reduction of TSS and VSS is usually used as an indicator of the biodegradation efficiency of sludge. There is a linear relationship between SCOD solubilization and VSS reduction (Zhang et al., 2013), here the higher the solubilization of COD, the higher is the reduction of VSS.

Table 1
Characteristics of the primary sludge and thermophilic inoculum used in this study.

| Item     | Unit       | Primary sludge       | Mesophilic inoculum |
|----------|------------|----------------------|---------------------|
| TSS      | (g/L)      | 11.54 ± 0.42         | 36.63 ± 0.31        |
| TS       | (g/L)      | 11.80 ± 0.65         | 39.62 ± 0.43        |
| VS       | (%TS)      | 93.93 ± 0.24         | 83.48 ± 0.37        |
| C        | (%TS)      | 41.69 ± 8.29         | 43.66 ± 0.60        |
| H        | (%TS)      | 6.19 ± 0.60          | 5.82 ± 0.29         |
| O        | (%TS)      | 42.16 ± 2.4          | -                   |
| N        | (%TS)      | 0.15 ± 0.07          | 3.25 ± 0.10         |
| S        | (%TS)      | 0.05 ± 0.03          | 1.02 ± 0.11         |
| C/N      |            | 285.60               | 13.43               |
| TCOD     | (g/L)      | 20.078 ± 1.865       | 47.66 ± 1.04        |
| Soluble COD | (g/L)     | 0.194 ± 0.010        | 1.150 ± 0.100       |
| Ash      | (%)        | 3.61 ± 0.37          | 16.52 ± 0.38        |
| pH       |            | 7.2 ± 1              | 6.99 ± 0.030        |

= indicates not measured values

2.3.2. Biogas production and composition

The biogas production rate was measured daily throughout the experimental period using the water displacement method and graduated cylinder, and then the biogas yields were calculated as weekly averages. Biogas composition (methane, nitrogen, and carbon dioxide) was measured by a gas chromatography system (Shimadzu, GC-2014, Kyoto Japan). The biogas samples were taken from the headspace of the bioreactor using a 5 ml syringe and injected in the GC system equipped with a thermal conductivity detector (room temperature +10°C to 400°C), a silica gel packed column, and injectors. Helium was used as an equipment carrier gas at a flow rate of 30 mL min\(^{-1}\) (digital setting by electronic
flow controller (AFC)). Samples were injected at room temperature via a single packed injector to
determine the composition. The composition was measured every two days throughout the experimental
period.

### 2.3.3. Permeate quality and digestate properties

COD of the permeate was measured according to the standard protocols (APHA, 2005) every other week.
MLSS was measured three times a week and monitored throughout the experimental period. Elemental
composition (C, N, H, S, O) of digestates was analyzed by an Elementar Vario EL (GmbH, Hanau,
Germany). Chemical element concentrations of the permeate were measured by the ICP-AES (Inductively
coupled plasma atomic emission spectroscopy) elemental analyzer. NanoBrook Zeta PALS Analyzer (Brookhaven Instruments Corp, USA) was used to analyze the zeta potential of digestates suspension.
Structural changes and degradation of primary sludge components after anaerobic digestion were
studied using a Bruker Tensor 37 Fourier Transform Infrared (FT-IR) Spectrophotometer. Digestates
dewaterability was measured by Capillary Suction Time (CST) (Triton Electronics Ltd., Bigods Hall,
Dunmow, UK), using CST filter paper and a 1 cm sludge funnel. Some measurements were performed
throughout the experimental period, others were made in triplicate, and results were reported as means ±
standard deviations.

### 2.3.4. Particle size distributions

Mastersizer 2000 Particle Size Analyzer (Malvern Instruments Ltd. Worcestershire, UK) was used for the
measurement of the particle size distributions of the mixed liquor and feed samples. It consists of an
optical bench (to collect raw data), dispersion units (for sample preparation/delivery), and a standalone
computer unit that runs Malvern software. This instrument uses laser diffraction or low light angle
scattering and can measure particle sizes in the range of 0.02 to 2000 microns using a single optical
measurement path. The mastersizer detects the scattered light, which is a combination of two light
sources, by a detector that is used to measure across the particle size range and converts a signal to size.
This device measures each sample three times automatically and calculates the average.

### 3. Results And Discussion

#### Effect of OLR on biological performance and stability

The biological performance was determined in this study based on biogas productivity/composition,
permeate quality/digestate properties, and solids reduction. Table 2 shows the operating conditions of
ThAnMBR. The experiment was divided into three phases as shown in Table 2. Phase I was operated at
HRT of 5 d and OLR of 3.7 kg-COD/m³ d, while phases II and III were operated at HRTs of 3 and 8 d and
OLRs of 6.8 and 2.59 kg-COD/m³ d, respectively. This study aims to test the feasibility of the primary
sludge digestion under relatively higher organic loading rates and lower HRTs. Regarding the primary
sludge studies, Bayr and Rintala (2012) studied primary sludge digestion under hydraulic retention times
(HRT) in the range of 16–32 d, which is relatively high compared to the current study. Likewise, Veluchamy and Kalamdhad (2017b) investigated pulp and paper mill sludge digestion under HRT of 45 d.

All phases of this study were run at a constant solid retention time of 32 d. In all phases, the primary sludge was introduced as a sole substrate to examine its biogas potential and solids reduction. At the end of each phase, the reactor was shut down, and the membrane was taken out for characterization. To initiate each phase, the reactor was mixed, and a new membrane module was installed, then, the required HRT and OLR were set up. Each phase lasted for more than 3 months, once the reactor biogas production and MLSS concentration were stable, the reactor was shifted into the next operating conditions, as shown in Fig. 2 (a–c). The system was operated in three different organic loading environments by changing the HRT (flow rate) while maintaining a constant SRT for the process.

| Phase | Duration (d) | Temperature (°C) | Average permeate flux L/m².hr. | Feed COD (g/L) | Feed TS (g/L) | SRT (d) | HRT (d) | OLR (kg-COD/m³ d) |
|-------|--------------|------------------|--------------------------------|----------------|--------------|---------|---------|------------------|
| I     | 1–94         | 50               | 2.43 ± 0.20                    | 19.39 ± 1.22   | 11.57 ± 0.74 | 32      | 5       | 3.73 ± 0.35      |
| II    | 99–206       | 50               | 4.59 ± 0.28                    | 20.61 ± 1.49   | 11.59 ± 0.30 | 32      | 3       | 6.84 ± 0.89      |
| III   | 212–323      | 50               | 1.48 ± 0.11                    | 19.76 ± 1.01   | 11.48 ± 0.14 | 32      | 8       | 2.55 ± 0.19      |

*The COD was the solids COD from primary sludge but not soluble COD.

### 3.1. Biogas production and composition

Reportedly, optimal biogas yield is a result of appropriate reactor design and enhanced growth of methanogens forming bacteria, which in turn depends on the favorable operating environment (e.g., the optimal amount of feedstock (OLR), temperature, and HRT). Biogas production and percentage of biogas composition of different operating conditions studied are presented in Fig. 2 (a–c). Reactor performance was studied at different OLR ranges from 2.5 to 6.8 kg-COD/m³ d. The best results were obtained at higher HRTs and lower organic loading rates. At the beginning of each phase, a slight decrease in biogas yield was observed but the system soon recovered and adapted to new conditions over time. The average biogas production rate of the reactor for loading rates of 3.7 ± 0.4, 6.8 ± 0.9, and 2.5 ± 0.1 kg COD/ m³ d was observed as 0.96, 1.14, and 1.6 L/d, respectively (Fig. 2a). Best biogas productivity was achieved at OLR 2.5 kg COD/ m³ d and HRT 8 d. The biogas production at 2.5 kg COD/ m³ d OLR was about two times higher than that at 3.7 kg COD/m³ d OLR. The system showed a decrease in biogas production with an increased loading rate. The increased solids content can be outside the capacity of microorganisms to handle it due to the recalcitrant nature of the primary sludge. A decrease in biogas
production with an increase in OLR is observed in many previous findings. For example, Gou et al. (2014) observed an approximately 43% decrease in CH$_4$ yield when the OLR increased from 1 to 6 g VS/L/day, during the co-digestion of waste activated sludge with food waste in a thermophilic system. Hassan et al. (2015) reported a similar result when anaerobically treated recycled paper mill effluent by a hybrid baffled reactor. The biomass degradation was also decreased as OLR increased. Several studies have shown similar findings. For example, Mel et al. (2015) reported a reduced COD degradation as the organic loading increased. This can be attributed to the recalcitrant property of the primary sludge, which requires a longer residence time, and the overloading of the system. These results illustrate that it is not feasible to operate the system under the high solids rate of the primary sludge, and OLR 2.5 kg COD/m$^3$/d and HRT 8 d may be consider optimum and can be suggested as design criteria for PS treatment. Different optimal OLRs are reported for anaerobic digestion of organic wastes, but this result is similar to results obtained by Liu et al. (2017) when they treated food waste under a thermophilic condition and an optimum OLR of 2.5 was reported. Figure 2b shows biogas yield based on the amount of the feed suspended solids added. The average biogas production obtained during phase I at OLR of 3.7 kg COD/m$^3$/d was approximately 53 L biogas/kg MLSS fed, however, when the OLR increased to 6.8 kg COD/m$^3$/d in phase II this resulted in a decreased biogas production of 45 L biogas/kg MLSS fed. However, when OLR was reduced to 2.5 kg COD/m$^3$/d in phase III this resulted in an improved biogas production of 155 L biogas/kg MLSS fed. The biogas production at phase II is corresponding to biogas production of 45 m$^3$/t VS$_{removed}$, which is reported by Jokela et al. (1997) for primary sludge from the TMP mill using the Biochemical Methane Potential (BMP) test. For comparison, the reported biogas yield for Kraft pulp mill primary sludge under the same operating temperature (thermophilic conditions) was higher than the biogas yield from the thermomechanical pulp mill reported in this study. For example, Bayr and Rintala (2012) reported 190–230 m$^3$CH$_4$/tVS$_{fed}$ methane yields for primary sludge from bleached Kraft mill under OLR of 1–1.4 kgVS/m$^3$/d and HRT of 16–32 d using CSTR. The OLR used in this study showing an excellent biogas production rate was 2-2.5 times of the OLR used in the study of Bayr and Rintala (2012), demonstrating the advantages of the THAnMBR, as compared to conventional CRTR. Differences in biogas production can be attributed to the different pulping conditions between the Kraft and TMP plants, as well as the resulting sludge. In the TMP process, the fibers are produced by thermal treatment followed by mechanical refining, while the kraft process involves treating the wood chips with a hot mixture containing water, NaOH, and Na$_2$S, known as white liquor. Hence, kraft fiber is flexible and easily degradable fiber compared to TMP fiber. Also, the surface of the kraft fibers contains less amounts of lignin and extractives than the TMP fibers, and the wood extractives are well-known inhibitory compounds for biological processes.

Under the steady-state, the methane content of the biogas was around 55.5 ± 5.78%, while, carbon dioxide and nitrogen content were 31.3 ± 6.3 and 13.2 ± 5.6, respectively (Fig. 2c). No significant change in the methane content was observed with the change in the solid loading rate, but the profile of the biogas production rate was different. The methane content achieved in this study was close to the results of most lignocellulosic treatment studies. It has been reported that the percentage of methane and carbon
dioxide in biogas is largely dependent on the feedstock processed as well as the duration and extent of bio-methanation over the retention period (Kavuma, 2013). The relatively high CO₂ content in the produced biogas of this study can be a result of the presence of acidifying microorganisms in methanogens, which causes the accumulation of volatile fatty acids in the process (Wijekoon et al., 2011; Franke-Whittle et al., 2014). Usually, a high VFAs concentration reduces the pH value in the reactor and prevents methane formation due to the inhibition of the methanogenesis process.

### 3.2. Permeate quality

The effect of OLRs on effluent (permeate) characteristics was evaluated in this study. The effluent properties were analyzed in terms of COD concentration, pH, and chemical element concentrations. Effluent COD and chemical element concentrations have fluctuated to some extent with OLRs in the experiment. In the first phase, the average effluent COD was 1.67 ± 0.46 g/L at OLR of 3.7 kg-COD/m³ d while the average concentration of the COD in the second and third phases were 0.70 ± 0.34 and 0.32 ± 0.11 g/L at OLRs of 6.8 and 2.59 kg-COD/m³ d, respectively. As can be seen from Fig. 3, the COD of the effluent fluctuated slightly in the first phase, which could be due to the beginning of the acclimatization of microorganisms to the new environment and the addition of the inoculation substance (inoculum). Thereafter, the COD concentration stabilized and varied between 0.16 g/L and 0.6 g/L throughout the experimental period. This trend has also been reported in other studies, for example, Lin et al. (2011) reported a high COD value (800 mg/L) in the initial start-up period and a low COD value (425 mg/L) during the stable-state process of the submerged anaerobic membrane reactor (SAnMBR). A similar pattern was observed by Hafuka et al. (2019), they reported a gradual decrease in the effluent COD concentration in the range of 840–240 mg/L. Sato et al. (2016) reported COD values in the effluent in the range of 510 – 251 mg/L for a membrane bioreactor treating sludge biomass. Hafuka et al. (2019) attributed the high COD concentration of the membrane filtrate in early operating time to the high concentration of soluble-COD of the seed sludge. The soluble COD of inoculum (seed sludge) was 1.2 g/L in this study, which could also be linked to the high COD concentration in the first phase. COD concentration decreased with increasing operating time can also be attributed to an increase in biological activity in the reactor (high degradation rate).

On the other hand, the OLR has shown some influence on the effluent COD, where the lower OLR was associated with a lower COD concentration. This may indicate that the high solid loading rate is not fully digested by the reactor microorganisms. It is worth noting that the COD of the effluent at the lower OLR was to a certain extent stable compared to the higher OLRs. Higher effluent COD with higher OLRs was also reported in other research works. For example, Boonyungyuen et al. (2014) reported 132.0 ± 4.3 mg/L effluent COD concentration for OLR of 0.52 kg/m³ d compared to 84.4 ± 10.2 mg/L for OLR of 0.13 kg/m³ d, when they treated textile wastewater by MBR. Wijekoon et al. (2011) attributed the higher permeate concentration to the increased generation of volatile fatty acid (VFA) on the reactor with increased OLR, and they achieved VFA removal efficiency in descending order of 96%, 94%, and 82% for OLRs of 5, 8, and 12 kg COD/m³ d, respectively. In this study, the pH of the effluent ranged from 7.2 to 7.4, which is in the methanogenic range indicating the stability of the system. Concentrations of chemical elements differed
slightly between the tested conditions (Table 3), where the permeates of the shorter HRTs had more concentrations of phosphorus, copper, sodium, and potassium, compared to the longer HRT permeate. However, the remaining elements did not differ greatly between the varied operating conditions.
### Table 3
**Effluent characteristics under different SRTs**

| Description | Minimum detection limit (MDL) | Permeate properties at different HRTs | 5d | 3d | 8d |
|-------------|-------------------------------|--------------------------------------|-----|----|----|
| **Macronutrients (mg/L)** | | | | | |
| Phosphorus | 0.05 | | | | |
| Sulfur | 0.3 | | | | |
| **Micronutrients (mg/L)** | | | | | |
| Iron | 0.005 | 0.039 | 0.044 | 0.084 | |
| Cobalt | 0.002 | < 0.002 | < 0.002 | < 0.002 | |
| Nickel | 0.03 | < 0.03 | < 0.03 | < 0.03 | |
| Zinc | 0.001 | 0.0023 | 0.0023 | < 0.001 | |
| Copper | 0.002 | 0.0073 | 0.0043 | < 0.0027 | |
| Manganese | 0.001 | 0.034 | 0.040 | 0.057 | |
| Molybdenum | 0.05 | < 0.05 | < 0.05 | < 0.05 | |
| Selenium | 0.05 | < 0.05 | < 0.05 | < 0.05 | |
| **Common cations (mg/L)** | | | | | |
| Sodium | 0.03 | 505 | 403.33 | 346.67 | |
| Potassium | 0.5 | 167 | 145 | 117.33 | |
| Calcium | 0.01 | 11.99 | 9.10 | 12.23 | |
| Magnesium | 0.01 | 3.16 | 2.92 | 3.49 | |
| Lead | 0.03 | < 0.03 | < 0.03 | < 0.03 | |
| Aluminum | 0.03 | < 0.03 | < 0.03 | < 0.03 | |
| Barium | 0.04 | < 0.04 | < 0.04 | < 0.04 | |
| Chromium | 0.002 | 0.0053 | < 0.002 | < 0.003 | |
| Tin | 0.05 | < 0.05 | < 0.05 | < 0.05 | |
| Strontium | 0.01 | 0.023 | < 0.01 | 0.017 | |
| COD (mg/L) | | | | | |
| 1.7 ± 0.46 | 0.70 ± 0.34 | 0.32 ± 0.11 | | |
| pH | | | | | |
| 7.3 ± 0.3 | 7.2 ± 0.1 | 7.3 ± 0.1 | | |

**3.3. Digestates properties**
The performance of the system was assessed also by taking into consideration the digestate properties under different operating conditions of ThAnMBR. Figure 4 shows the concentrations of MLSS (mixed liquor suspended solids), FSS (Feed suspended solids), and reactor pH upon changing of different OLRs in the reactor. The MLSS concentrations for the reactor ranged from 16 to 28 g/L. The first phase showed fluctuation in MLSS concentration, which could be due to microorganisms’ acclimatization (Fig. 4). In the second phase, the MLSS concentration increased from 20 to 24 g/L due to the increased organic loading rate (from 3.7 to 6.8 kg COD/m$^3$ d) and stabilized at about 26 g/L throughout the phase time. However, when the OLR was reduced to 2.5 kg COD/m$^3$ d in the third phase, the MLSS concentration remained stable and high. The reason for this was the accumulation of sludge on the wall of the reactor at the liquid and biogas interface in the second phase, and the falling of the accumulated headspace sludge on the bioreactor at the end of the second phase resulted in a higher MLSS concentration in the following phase (third phase). The sludge accumulated in the headspace of the reactor in the second phase due to the high OLR coupled with reactor sparging by biogas for fouling control. The latter blew out the sludge into the reactor headspace and then the sludge accumulated on the headspace and between the fittings. Although a drop in MLSS concentrations was expected due to reduced OLR, the MLSS concentration in the third phase ranged between 25 and 26.5 g/L because of the aforesaid reasons, and no major fluctuation was seen, as shown in Fig. 4.

In this study, the total solids of the treated sludge were 11.80 ± 0.65, and NH$_4$Cl and K$_2$HPO$_4$ were added as nutrients to the feed to meet the COD:N:P ratio of 100:2.6:1. At this concentration, the bioreactor can be easily fed with the sludge substrate using a magnetic drive centrifugal pump (Iwaki magnetic pump). It has been reported that there is an optimal concentration of TS content in raw materials and usually ranges between 1–10% and that the volume of the produced biogas diminishes by reducing and increasing the TS concentration portion below and above the optimum value (Yavini et al., 2014; Dhar et al., 2016). The pH profile of the reactor during the experimental time is shown in Fig. 4. The pH of the system ranged between 7.2 and 7.5 in this study. Reportedly, the pH values and temperature regime have a direct impact on biogas production. Methanogens have been reported to grow better in the pH range 6.8–7.5, which is favorable for biogas production. However, the anaerobic digester process can tolerate a range of 6.5 to 8.0 (Cioabla et al., 2012). On the other hand, lower pH levels may lead to a complete reactor failure by inhibiting the CH$_4$-forming bacteria.

Table 4 shows the properties of digestate under different operating conditions. The average soluble COD for digestates was 0.188, 0.326, and 0.153 g/L in phase I, II, and III, respectively. The soluble COD of the digestate was higher in the third phase compared to the first (HRT, 5 d) and second (HRT, 8 d) phases. This may be due to the short HRT (3 d), which corresponds to the highest OLR (6.8 kgCOD/m$^3$ d). Elemental composition (C, N, H, and S) was analyzed by an elemental analyzer. As can be seen from Table 4, there is no significant difference in C, N, H, and S concentrations between the tested OLRs. However, nitrogen concentration decreased slightly with increasing digestion time. It can be concluded from Table 4 that digestates contain high carbon sources and low nutrient sources. Thereby, for the further valorization of this digestate via bioconversion processes, the value of these two crucial elements
needs to be optimized for improved microorganisms' growth and degradation rate. As indicated in Table 4, the weekly average of the biogas yield ranged between 0.061 and 0.095 L biogas/g MLSS removed in the first phase and ranged between 0.045 and 0.059 in the second phase. Whereas the biogas yield in the third phase was in the range of 0.316–0.339 L biogas/g MLSS removed, indicating the effect of OLR on the biogas yield and a lower OLR led to an increased biogas yield.

Solid's reduction ratio is an important parameter in the anaerobic digestion of sludge. The reduction of the sludge biomass ranged from 28.9 to 46.7 % in this study, depending on the applied OLRs. Sludge biomass degradation efficiency decreased with increasing OLR. Sludge biomass reduction was 45.5% at the OLR of 3.7 kg COD/m$^3$ d but was markedly reduced to 28.9 % when the OLR increased to 6.8 kg COD/m$^3$ d. While the solids reduction in the reactor varied between 46.13 and 46.7% for the OLR of 2.5 kg COD/m$^3$ d. This result is similar to the previously reported results for MBRs. For example, Sato et al. (2016) achieved about 47.4% sludge biomass reduction when the OLR was reduced from 0.45 to 0. 225 kg COD/m$^3$ d, using pilot-scale MBR. Using conventional bioreactors, Lin et al. (2009) reported cellulose degradation efficiency in the range of 21.8 to 65% for pulp and paper sludge, but no pronounced change was observed for lignin degradation even after NaOH pretreatment, indicating the low degradability of the lignin. Bayr and Rintala (2012) achieved cellulose removal efficiency in the range of 70–73% after anaerobic digestion of pulp and paper primary sludge with overall volatile solids (VS) removal between 25 and 40% (similar to the results from this study), but hemicellulose did not degrade much compared to cellulose with a degradation efficacy between 7 and 27%. Kinnunen et al. (2015) reported VS removal between 11 and 26% for pulp and paper industry bio-sludge, which is similar to the finding (9–23% VS removal) reported by Saha et al. (2011) for pulp and paper mill secondary sludge. Thus, for improved organic matter degradation, pretreatment of the sludge substrate could be recommended.
### Table 4
Digestates properties, biogas yield, and solids reduction ratio under different operating conditions of anaerobic digestion of pulp and paper mill primary sludge in thermophilic AnMBR.

| Component                          | Digestate characteristics at different phases |
|------------------------------------|----------------------------------------------|
|                                    | Phase I                                      | Phase II                                    | Phase III                                   |
| Days                               | 1–94                                        | 99–206                                      | 212–323                                     |
| OLR (kgCOD/m$^3$d)                 | 3.7                                          | 6.8                                         | 2.5                                         |
| HRT (d)                            | 5                                            | 3                                            | 8                                           |
| SRT (d)                            | 32.5                                         | 32.5                                         | 32.5                                        |
| Nitrogen addition $^a$             | yes                                          | yes                                         | yes                                         |
| Phosphorus addition $^b$           | yes                                          | yes                                         | yes                                         |
| VS (%)                             | 90.18 ± 0.39                                 | 92.83 ± 1.2                                 | 92.35 ± 0.64                                |
| Fixed solids (%)                   | 9.82 ± 0.38                                  | 7.17 ± 1.2                                  | 7.65 ± 0.65                                 |
| SCOD (g/l)                         | 0.188 ± 0.007                                | 0.326 ± 0.009                               | 0.153 ± 0.058                               |
| Biogas yield (L biogas/g MLSS$_{removed}$) | 0.095                                      | 0.059                                       | 0.334                                       |
| Average methane conc. (%)          | 55                                           | 58.7                                        | 56                                          |
| Solid reduction (%)                | 45.5 ± 0.74                                  | 28.9 ± 0.02                                 | 46.13 ± 0.95                                |
| C (%)                              | 45.22 ± 1.26                                 | 45.59 ± 0.70                                | 45.07 ± 1.20                                |
| H (%)                              | 5.98 ± 0.50                                  | 5.94 ± 0.15                                 | 6.01 ± 0.48                                 |
| N (%)                              | 0.86 ± 0.18                                  | 0.79 ± 0.24                                 | 0.75 ± 0.15                                 |
| S (%)                              | 0.29 ± 0.08                                  | 0.25 ± 0.06                                 | 0.24 ± 0.07                                 |
| C/N                                | 52.58                                        | 57.71                                       | 60.09                                       |
| pH                                 | 7.2 ± 0.10                                   | 7.2 ± 0.05                                  | 7.2 ± 0.09                                  |

$^a$NH$_4$Cl  
$^b$KH$_2$PO$_4$

### 3.4. Particle size distribution

The particle size distribution (PSD) measurement of the digestates is shown in Fig. 5. The PSD of the digestates did not vary greatly with different operating conditions (OLRs), as they ranged between 1 and 1000 microns. The second phase contained larger particles (peaked at about 500 microns) than the other...
phases, possibly due to the higher loading rate (OLR, 6.8 kg COD/m³ d). The other phases had OLRs in the range of 2.5–3.7 kg COD/m³ d. All phases showed low amounts of small particles, which peaked at about 0.8 µm. The average size of the digestates of the different phases was around 30 µm. On the other hand, the size of the digestates molecules did not appear to have any clear effect on the membrane pores blocking, and this may be due to their large sizes. The employed membrane has a pore size of 0.1µm, and it is well known that when the pore size is relatively small, fouling may result mainly from the foulants adsorption. It has been reported that when foulants are similar or smaller than the pores of the membrane, adsorption and pore-blocking mechanisms occur; however, when the foulants are greater than the pores of the membrane, a cake layer will likely form on the surface of the membrane (Bokhary et al., 2018).

4. Conclusion

In this study, thermophilic anaerobic digestion of primary sludge of a thermomechanical pulp mill in a relatively new type of bioreactor (ThAnMBR) was studied for the first time. In particular, the variations of OLRs with operational parameters and their interrelations were investigated. The results of this study showed that the primary sludge of the TMP mill can be satisfactorily treated with a ThAnMBR. The reactor showed stable performance and biogas production in the range of 0.059–0.334 L biogas/g MLSS removed was achieved. Lower OLRs and higher HRTs have been associated with higher biogas production compared to higher OLR and shorter HRT, and OLR of 2.5 kg COD/m³ d and HRT 8 d could be considered optimum and can be suggested as design criteria for PS treatment. The reduction of the sludge biomass ranged from 28.9 to 46.7 % depending on the applied OLRs. Sludge biomass degradation efficiency decreased with increasing OLR. Also, changing the OLRS has shown some influence on the effluent COD, where the lower OLR was associated with a lower COD concentration. Similarly, digestate properties have fluctuated to some extent with OLRs. Regardless of OLRs, membrane performance was stable, and the required membrane flux was easily maintained during the operation. The results demonstrated that ThAnMBR is a promising new technology for pulp and paper mill primary sludge treatment and has many advantages as compared to conventional bioreactors.

Declarations

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Figures
Figure 1

a schematic diagram of the ThAnMBR setup
Figure 2

(a) Biogas production rate (b) biogas yield based on the amount of the feed suspended solids added (c) percentage of biogas composition at different organic loading rates over digestion time.
Figure 3

Permeate COD and pH under the tested conditions over digestion time.

Figure 4

Mixed liquor suspended solids (MLSS) and pH of the digester and the suspended solids of the feeding substrate (FSS).
Figure 5

Particle size distribution of the mixed liquor suspended solids in the three tested conditions.

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