Replication of Type 2 Diabetes Candidate Genes Variations in Three Geographically Unrelated Indian Population Groups

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Abstract

Type 2 diabetes (T2D) is a syndrome of multiple metabolic disorders and is genetically heterogeneous. India comprises one of the largest global populations with the highest number of reported type 2 diabetes cases. However, limited information about T2D associated loci is available for Indian populations. It is, therefore, pertinent to evaluate the previously associated candidates as well as identify novel genetic variations in Indian populations to understand the extent of genetic heterogeneity. We chose to do a cost effective high-throughput mass-array genotyping and studied the candidate gene variations associated with T2D in literature. In this case-control candidate genes association study, 91 SNPs from 55 candidate genes have been analyzed in three geographically independent population groups from India. We report the genetic variants in five candidate genes: TCF7L2, HHEX, ENPP1, IDE and FTO, are significantly associated (after Bonferroni correction, p<5.5E–04) with T2D susceptibility in combined population. Interestingly, SNP rs7903146 of the TCF7L2 gene passed the genome wide significance threshold (combined P value = 2.05E–08) in the studied populations. We also observed the association of rs7903146 with blood glucose (fasting and postprandial) levels, supporting the role of TCF7L2 gene in blood glucose homeostasis. Further, we noted that the moderate risk provided by the independently associated loci in combined population with Odds Ratio (OR) <1.38 increased to OR = 2.44, (95%CI = 1.67–3.59) when the risk providing functional SNPs ofrs7903146 were considered. We also observed the association of rs11618490 of the IDE gene in blood glucose homeostasis.

Introduction

The prevalence of type 2 diabetes (T2D), a complex disorder, is increasing at an alarming rate and becoming a major health problem. The highest incidence of T2D is seen in developing countries where 80% of deaths occur due to diabetes [1]. It has been proposed that the highest number of diabetic patients would be in Asia by the year 2025 [2,3]. The increased prevalence of type 2 diabetes (T2D) is thought to be due to environmental factors, acting on genetically susceptible individuals [4]. The heritability of T2D is one of the best established among common diseases [5,6], and consequently, genetic risk factors for T2D have been the subject of intense research [7]. Linkage studies have reported many T2D-linked chromosomal regions and have identified putative, causative genetic variants in CAPN10, ENPP1, HNF4A, WFS1 and ACDC [8–10]. In parallel, candidate-gene association studies have reported many T2D-associated loci, with coding variants in the nuclear receptor PPARγ (P12A) [11] and the potassium channel KCNJ11/E23K [12] being among the very few that have been replicated in most of the populations. Multiple genome wide association studies identified the genes including TCF7L2, as well as a non-synonymous SNP in the zinc transporter SLC30A8 and variants in HHEX, CDKAL1, IGF2BP2 and CDKN2A/B [13–17]. Study by WTCCC involving a common set of controls for the seven UK wide case cohorts led to the finding of FTO to be associated with T2D through its effect on body mass index (BMI) [18]. The Diabetes and Genetics Replication and Meta-Analysis (DIAGRAM) consortium was formed to carry out meta-analyses of three of the previously published studies; WTCCC, DGI and FUSION [13–15]. This international collaborative effort identified six new loci JAZF1, CDC123-CAMK1D, TSPAN8-LGR5, THADA, ADAMTS9 and NOTCH2 [19]. Few studies from East-Asian ancestry identified additional loci, KCNJ1, PTPRD, SRR, 13q13.1, UBE2E2 and
CDCA4, CDCA4B associated with T2D [20–24]. However, a majority of loci show association in some populations but did not replicate in others, most plausibly a result of genetic heterogeneity in T2D, instead of all being false positive associations. India comprises one of the largest global populations, which had 62.4 million people with type 2 diabetes in year 2011. International Diabetes Federation has predicted it to be 100 million people by year 2030 [25]. It makes it important to replicate and evaluate the previously associated candidates to identify common T2D associated variations/genes, as well as identify novel genetic variations in various Indian population groups to understand the extent of genetic heterogeneity. In the present study, we tried to address it and analyzed 91 SNPs from 55 candidate genes, most of which are previously associated with T2D susceptibility (Table S1) in different world-populations, in three geographically isolated Indian population groups.

Results and Discussion

Type 2 Diabetes (T2D) is a syndrome of multiple metabolic disorders. It includes abnormally high blood glucose levels (hyperglycemia); involving insulin resistance related signaling pathways and defects in insulin-mediated glucose uptake in muscle; impaired insulin secretion due to dysfunction of pancreatic β cells; disruption of secretory function of adipocytes and an impaired insulin action in liver. The etiology of human T2D involves a strong genetic background [26]. Various approaches including the linkage, candidate gene and the recent genome-wide-studies have been successful to identify more than 40 common genetic variants associated with T2D [27,28]. These gene variants are related to different metabolic pathways in the disease [29]. However, the total genetic variants roughly account for 10% of the heritability of T2D, suggesting that much remains to be discovered [30]. There is a need to replicate previously associated loci in multiple populations of the world, specifically in Asia including India, where relatively fewer studies have been carried out to identify the common global T2D associated variations/genes; and simultaneously assess the genetic heterogeneity among different population groups for these loci.

The three different population groups of India (Punjab; Jammu and Kashmir; Orissa) recruited in this study were genotyped for 91 SNPs from 55 candidate genes, including those previously associated with T2D susceptibility (Table S1). IBS (Identity by state) analysis (Table S2) showed no significant difference among the cases and controls, in any of the three studied population sets, suggesting those as homogeneous population groups. The detailed description of SNPs, status of Hardy Weinberg equilibrium, allelic frequencies in cases and controls is provided in Table S3.

| SNP Chr: bp position Gene | Punjab Cases/Controls | Jammu and Kashmir Cases/Controls | Orissa Cases/Controls | P value OR(95% CI) | Combined (Fix effects model) | P value OR(95% CI) | OR(95% CI) |
|---------------------------|----------------------|-------------------------|-------------------|-----------------|-----------------------------|------------------|-------------|
| rs7903146_C/T 10:114758349 TCF7L2 | 0.359/0.303 0.003278 1.28(1.08–1.52) | 0.363/0.277 0.000452 1.46(1.13–1.93) | 0.340/0.263 0.000946 1.48(1.19–1.85) | 2.05E–08 1.3833 | | |
| rs1044498_A/C 6:132172368 ENPP1 | 0.189/0.144 0.003242 1.37(1.11–1.70) | 0.201/0.153 0.01724 1.39(1.05–1.82) | 0.188/0.156 0.0864 1.24(0.96–1.61) | 4.24E–05 1.3411 | | |
| rs9939609_T/A 16:53820527 FTO | 0.351/0.293 0.001944 1.30(1.10–1.54) | 0.369/0.323 0.06208 1.22(0.98–1.52) | 0.369/0.306 0.00673 1.32(1.08–1.62) | 7.38E–06 1.2905 | | |
| rs3751812_G/T 16:53818460 FTO | 0.349/0.292 0.002282 1.30(1.09–1.54) | 0.372/0.325 0.007303 1.30(1.06–1.58) | 0.372/0.325 0.007303 1.30(1.06–1.58) | 1.2786 | | |
Replication of T2D Candidate Genes Variations

[31,32]; and was replicated in genome-wide association and candidate gene studies [16,17]. We wanted to explore whether the association signal is independent of each other or these genes are in linkage disequilibrium (LD). LD analysis of 14 SNPs of TCF7L2, IDE, HHEX along with SIRT1 genes located at chromosome 10q23-25 was performed using Haploview software. Interestingly, the LD analysis showed a very weak or no LD between SNPs from these genes (Figure S1), suggesting an independent risk effect of each locus. These loci remained significant in logistic regression analysis after adjustment with BMI, age and gender as covariates, (Table S5). Along with, the common disease associated variations, shared by all the groups, we observed some variations showing association in population specific manner (p<0.05) and are provided in Table S4. These variations either represent the genetic heterogeneity among the populations or are some false positives, which warrant screening in larger sample sets.

Further, analysis of studied SNPs, using one way Anova and linear correlation, with epidemiological/clinical parameters of diabetes [waist to hip circumference ratio (WHR), BMI, Blood glucose fasting/Postprandial] showed a significant association of SNP, rs7903146 of TCF7L2 gene with blood glucose (fasting as well as postprandial) in combined population (Table S6), probably indicating that Indian diabetic patients commonly fall in the category of being deficient in Insulin secretion rather than having insulin resistance [33,34]. TCF7L2, a transcription factor, is reported to be involved in glucose homeostasis, insulin secretion and biosynthesis through GLP1 and wingless type (wnt) signaling pathway is also involved in developmental and growth regulatory mechanism of cells [35–37].

The other associated SNPs in this study also belong to important genes that play an important role in various metabolic pathways of diabetes pathobiology. HHEX, a hematopoietically expressed homebox protein is another transcription factor that is suggested to reduce the β cell secretion capacity and sensitivity of insulin [38]. IDE, a major enzyme (Zn2⁺- regulated metalloproteinase) expressed ubiquitously including all insulin-responsive tissues responsible for insulin degradation and thereby influencing the extent of the cellular response to insulin [39,40]. The substrate specificity of IDE coincides with peptides capable of amyloid formation, and may prevent accumulation of amyloidogenic peptides. Disruption of this scavenging function might promote aggregation of the islet amyloid, a characteristic of type 2 diabetes [41]. Interestingly, some other studies have shown the risk allele of rs1879722 in association with increased post loading hyper-insulinemia [42]. ENPP1, an ectonucleotide pyrophosphatase phosphodiesterase, has a role in the insulin resistance by directly inhibiting insulin-induced conformational changes of the insulin receptor, thereby affecting its activation and downstream signaling, which resulted in fasting hyper-insulinemia, a strong predictor of the subsequent development of obesity in children [9,43]. FTO gene variant has been strongly associated with predisposition to diabetes through an effect on BMI [18,44]. We wanted to see if these biologically related and significantly associated candidate genes show any interactive effect in association to T2D, hence we evaluated interaction in the significantly associated SNPs. The risks provided by the independent loci in our study were moderate (OR<1.38, in combined population, at all loci) as has been shown in other studies [7]. Interaction analysis of genotype combinations of these SNPs with diabetes susceptibility showed an increased effect in associations. We observed that the pair-wise interaction analyses followed by multiple gene interactions (Table S7) shows an increased risk (p = 4.52E⁻⁶, OR = 2.44, 95%CI = 1.67–3.59) when the risk providing genotypes of TCF7L2, HHEX, ENPP1 and FTO genes were combined (distributed in 7.24% of patients compared to 3.08% of controls). An increased protection (p = 2.68E⁻⁹, OR = 0.28, 95%CI = 0.19–0.43) was also observed for the protection providing genotype combination of IDE, HHEX, ENPP1 and FTO genes (present in 7.63% of controls as compared to 2.12% patients). The observations suggest the importance of identifying not only novel loci in providing disease risk but also understand the role of other mechanisms including the gene-gene and pathway based interaction between multiple functionally important genes, in complex diseases like T2D.

In conclusion, our study suggests TCF7L2, HHEX, IDE, ENPP1 and FTO as commonly associated T2D susceptibility genes in the three Indian populations. Interaction analyses have shown an increased effect in associations suggesting the importance of gene-gene and pathway based interaction between multiple functionally important genes. This study also highlights the importance of multiple populaton groups based studies in identifying common disease causing genes. Genetic heterogeneity and phenocopies are among the vagaries of complex disorders like T2D, which make understanding of such diseases challenging. It is anticipated that sub-categorization of sample sets by clinical parameters as well as by social groupings like religions castes, etc. and studies of larger data sets will help us better understand the genetic heterogeneity, in complex diseases like T2D especially in Indian populations, our perspective of future studies.

Materials and Methods

Ethics Statement

A written informed consent was obtained from all the participants. The data were analysed anonymously, and the study was approved by the ethical committee of Jawaharlal Nehru University.

Subjects

In the present study, a total of 2900 samples, independent from our previous studies [45], including 1583 well characterized diabetes patients and 1317 controls belonging to three geographically independent population groups of India (649 patients and 600 controls from Punjab, 507 patients and 300 controls from Jammu and Kashmir; 427 patients and 417 controls from Orissa), were included. Diagnosis of T2D was made according to the criteria of World Health Organization (Expert Committee 2003). The patients with a history of ketoacidosis/requiring continuous insulin treatment since diagnosis/having exocrine pancreatic disease/or with exceptionally early age of onset (<30 years), were excluded. Patients with severe liver or renal dysfunction were also excluded. Non-diabetic individuals with no known positive family history of diabetes were included in the study. The studied individuals were confirmed of being unrelated for three generations. Anthropometric measurements and other features are summarized in Table S8.

Assessment of the Clinical Parameters

The patient was diagnosed with hypertension when the systolic blood pressure (SBP) was 140 mmHg and the diastolic blood pressure (DBP), 90 mmHg. Overweight and Obese together were clinically characterized by body mass index (BMI) of >24.9 (BMI is defined by ratio of weight in kilograms to square of height in meters) and the increased abdominal fat was measured by waist to hip circumference ratio (WHR) of 0.94.

SNP Selection and Genotyping

SNPs in this study (Table S1) were included from those genes which have been implicated with T2D or diabetes related traits
through genome-wide association studies, mostly in European populations, and further replicated in other populations using candidate gene approach [13,16,17,46–57]. In addition, other gene SNPs that have been studied with T2D susceptibility but not replicated or studied in multiple populations were also included. Genotyping of SNPs was performed using High-throughput genotyping MassArray platform (SEQUENOM) as described earlier [58]. SNP genotyping success rate was >95%. For quality control of SNP genotyping, each 96 well plate contained three or more duplicate samples and a negative control. The concordance rate for genotyping was >99.5%.

**Statistical Analyses**

Statistical analyses were mainly performed using PLINK v. 1.07 (http://www.pmgu.mgh.harvard.edu/plink/). Each SNP was tested for Hardy Weinberg Equilibrium. Pairwise IBS (Identity by state) distances between all individuals have also been calculated with respect to binary phenotype (non-significant SNPs of this study only), to know if there are hints of group differences. IBS analysis is most robust for genome-wide data; however, this analysis provides a preliminary evidence of no population stratification. Significant association of SNPs was tested by 3×2 Chi square test for overall genotype frequency distributions between diabetes patients and controls. Association of SNP with type 2 diabetes was further confirmed by conditional logistic regression analysis with forward conditional method adjusted for possible confounding factors: age, gender and BMI. Odds ratios (ORs) were calculated with respect to risk allele. Meta-analysis was performed by combining summary estimates both under random effect and fixed effect models using PLINK v. 1.07, which also provides Cochran’s Q test statistic, a test of heterogeneity among the studies, P value <5.5×10^-4 (0.05/91) was considered significant after Bonferroni correction. We also explored genotypic interactions of significantly associated SNPs using logistic regression with forward conditional method. These analyses were performed using statistical software SPSS v20.0 (SPSS, Chicago III, IL, USA).

Association of SNPs with quantitative traits was determined using one way ANOVA adjusted for age, sex, population and BMI as appropriate, in control, patients and total population. P value <2.77×10^-4 (0.05/180) was considered significant after Bonferroni correction (10 SNPs×6 parameters in 3 population groups).

Epidemiological parameters [Age, Waist/Hip ratio, BMI, Gender, Systolic Blood pressure, Diastolic Blood pressure] were compared between patients and controls using linear regression analysis. Statistical power of the study was estimated using QUANTO version 1.2 (http://hydra.usc.edu/gxe/). Sample size included in this study had 70–97% power to detect the association with OR of 1.3–1.5 assuming minor allele frequency of 0.20.

### Supporting Information

**Figure S1** Linkage disequilibrium (LD) analysis (r^2 value) of 14 SNPs of TCF7L2, IDE, HHEX and SIRT1 genes located at chromosome 10q23-25. (TIF)

**Table S1** Details of the SNPs selected for present study. (XLS)

**Table S2** Permutation test for: between group (case-control) Identity by State (IBS) difference with respect to binary phenotype for independent analysis of three populations. (DOC)

**Table S3** Characteristic of 91 SNPs from 55 genes studied in three different populations (Punjab; Jammu and Kashmir; and Orissa) of India. (XLS)

**Table S4** Allele frequencies, P values, odds ratios of the studied SNPs in three studied populations of India. (XLSX)

**Table S5** Logistic regression analysis of most significantly associated SNPs of each associated gene, adjusted with age, gender and BMI in combined population. (DOC)

**Table S6** One Way ANOVA of blood glucose (fasting and post prandial) with TCF7L2 gene SNPs. (DOC)

**Table S7** Genotype Interaction analysis of significantly associated SNPs. (DOC)

**Table S8** Distribution of epidemiological parameters between diabetes patients and Controls populations for studied three populations of India. (DOC)

### Acknowledgments

Authors acknowledge the participant in the study for their support and Massarray Facility at SMVDU, Katra for assistance in data generation.

### Author Contributions

Conceived and designed the experiments: RNKB SS SA. Performed the experiments: RC NK AB AM. Analyzed the data: SA SS SM. Contributed reagents/materials/analysis tools: RNKB ASB MKD SG GBNC PS. Wrote the paper: SS RA RNKB.

### References

1. Shaw JE, Sicree RA, Zimmet PZ (2010) Global estimates of the prevalence of diabetes for 2010 and 2030. Diabetes Res Clin Pract 87: 4–14.

2. Zimmer P, Alberti KG, Shaw J (2001) Global and societal implications of the diabetes epidemic. Nature 414: 782–787.

3. Wild S, Roglic G, Green A, Sicree R, King H (2004) Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. Diabetes Care 27: 1047–1053.

4. Cornels MC, Hu FB (2012) Gene-environment interactions in the development of type 2 diabetes: recent progress and continuing challenges. Annu Rev Nutr 32: 245–259.

5. Poulsen P, Kyvik KO, Vaag A, Beck-Nielsen H (1999) Heritability of type II (non-insulin-dependent) diabetes mellitus and abnormal glucose tolerance—a population-based twin study. Diabetologia 42: 139–145.

6. Florez JC, Hirschhorn J, Al Shabuler D (2003) The inherited basis of diabetes mellitus: implications for the genetic analysis of complex traits. Annu Rev Genomics Hum Genet 4: 257–291.

7. Permut MA, Wasson J, Cox N (2005) Genetic epidemiology of diabetes. J Clin Invest 115: 1431–1439.

8. Horikawa Y, Oda N, Cox NJ, Li X, Orho-Melander M, et al. (2000) Genetic variation in the gene encoding calpain-10 is associated with type 2 diabetes mellitus. Nat Genet 26: 163–175.

9. Meyre D, Boutaia-Naji N, Toumani A, Samson C, Lecoeur C, et al. (2005) Variants of ENPP1 are associated with childhood and adult obesity and increase the risk of glucose intolerance and type 2 diabetes. Nat Genet 37: 863–867.

10. Love-Gregory LD, Wasson J, Ma J, Jin CH, Glaser B, et al. (2004) A common polymorphism in the upstream promoter region of the hepatocyte nuclear factor-4 alpha gene on chromosome 20q is associated with type 2 diabetes and appears to contribute to the evidence for linkage in an ashkenazi jewish population. Diabetes 53: 1134–1140.

11. Al Shabuler D, Hirschhorn JN, Klammernark M, Lindgren CM, Voelhl MC, et al. (2000) The common PPARGamma Pro12Ala polymorphism is associated with decreased risk of type 2 diabetes. Nat Genet 26: 76–80.
36. Mulholland DJ, Dedhar S, Coetzee GA, Nelson CC (2005) Interaction of
positively regulates endostatin expression by interacting with the
cell surface. J Biol Chem 280: 1457–1464.
37. Schulze MB, Al-Hasani H, Boening F, Eibinger F, Doring F, et al. (2007) Variation
in the transcription factor 7-like 2 (TCF7L2) gene reconfirms associations with type 2
diabetes in the prospective, population-based EPIC-Potsdam cohort. Diabetologia 50:
2405–2407.
38. Seta KA, Rother RA (1997) Overexpression of insulin degrading enzyme: cellular
localization and effects on insulin signaling. Biochem Biophys Res Commun
141: 717–717.
39. Fakhrai-Rad H, Nikeshlah A, Kamei A, Fernstrom M, Zierath JR, et al. (2000)
Insulin-degrading enzyme identified as a candidate diabetes susceptibility gene in
GK rats. Hum Mol Genet 9: 2149–2158.
40. Kurochkin IV (2001) Insulin-degrading enzyme: embarking on amyloid
deposition. Trends Biochem Sci 26: 421–425.
41. Gu HF, Efendic S, Nordman S, Osenton CG, Brismar K, et al. (2004) Common variants in the TCF7L2 gene are strongly associated with type 2 diabetes susceptibility loci identified through large-scale association studies of variants in genes encoding the pancreatic beta-cell.
42. Gu HF, Efendic S, Lonnerdal B, Nordman S, Ostenson CG, Brismar K, et al. (2004)
A variant in CDKAL1 influences insulin response and risk of type 2 diabetes. Nat Genet 39: 770–775.
43. Zeggini E, Weedon MN, Langen DJ, Frayling TM, Elliott KS, et al. (2007)
Replication of genome-wide association signals in UK samples reveals risk loci for type 2 diabetes. Science 316: 1336–1341.
44. Zeggini E, Weedon MN, Langen DJ, Frayling TM, Elliott KS, et al. (2007)
A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316: 889–894.
45. Zeggini E, Scott LJ, Saxena R, Voight BF, Marchini JL, et al. (2007) Meta-
analysis of genome-wide association data and large-scale replication identifies additional susceptibility loci for type 2 diabetes. Nat Genet 40: 608–614.
46. Kasuwa K, Misayu K, Horikoshi T, Hara K, Osawa H, et al. (2008) Common variants in KCNQ1 are associated with susceptibility to type 2 diabetes mellitus. Nat Genet 40: 1092–1097.
47. Shetty P (2012) Public health: India’s diabetes time bomb. Nature 485: S14–16.
48. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, et al. (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316: 889–894.
49. Steinthorsdottir V, Thorleifsson G, Reynisdottir I, Benediktsson R, Jonsdottir T,
et al. (2008) SNPs in KCNQ1 are associated with susceptibility to type 2 diabetes mellitus. Nat Genet 40: 1092–1097.
50. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, et al. (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316: 889–894.
51. Tabara Y, Osawa H, Kawamoto R, Onuma H, Shimizu I, et al. (2009) Replication study of candidate genes associated with type 2 diabetes based on genome-wide screening. Diabetes 58: 493–498.
52. Chauhan G, Spurgeon CJ, Tabassum R, Bhaskar S, Kulkarni SR, et al. (2010)
Impact of common variants of PPARG, KCNJ11, TCF7L2, SLC30A8, HHEX, CDKN2A, IGF2BP2, and CDKAL1 on the risk of type 2 diabetes in 5,164 North Indian population. Diabetes 58: 2137–2147.
53. Takeuchi F, Serizawa M, Yamamoto K, Fujisawa T, Nakashima E, et al. (2009)
A systematic meta-analysis of genetic association studies for diabetic retinopathy. Diabetes 58: 2137–2147.
54. Gamboa-Meleendez MA, Huerta-Chagoya A, Moreno-Macias H, Vazquez-
Camarinha P, Ordonez-Sanchez ML, et al. (2012) Contribution of Common Genetic Variation to the Risk of Type 2 Diabetes in the Mexican Mestizo Population. Diabetes.
55. Gupta V, Vinay DG, Rajig S, Kranthikumar MV, Janipalli CS, et al. (2012) Association analysis of 31 common polymorphisms with type 2 diabetes and its diabetic retinopathy traits in 700 Indian sib units. Diabetologia 55: 349–357.
56. Rong R, Hanson RL, Ortiz D, Wiedrich C, Koles K, et al. (2009) Confirmation of multiple risk Loci and genetic impacts by a genome-wide association study of type 2 diabetes in the Japanese population. Hum Genet 122: 1092–1097.
57. Shetty P (2012) Public health: India’s diabetes time bomb. Nature 485: S14–16.
58. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, et al. (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316: 889–894.
59. Steinthorsdottir V, Thorleifsson G, Reynisdottir I, Benediktsson R, Jonsdottir T,
et al. (2008) SNPs in KCNQ1 are associated with susceptibility to type 2 diabetes mellitus. Nat Genet 40: 1092–1097.
60. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, et al. (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316: 889–894.
61. Tabara Y, Osawa H, Kawamoto R, Onuma H, Shimizu I, et al. (2009) Replication study of candidate genes associated with type 2 diabetes based on genome-wide screening. Diabetes 58: 493–498.
62. Chauhan G, Spurgeon CJ, Tabassum R, Bhaskar S, Kulkarni SR, et al. (2010)
Impact of common variants of PPARG, KCNJ11, TCF7L2, SLC30A8, HHEX, CDKN2A, IGF2BP2, and CDKAL1 on the risk of type 2 diabetes in 5,164 North Indian population. Diabetes 58: 2137–2147.
63. Takeuchi F, Serizawa M, Yamamoto K, Fujisawa T, Nakashima E, et al. (2009)
A systematic meta-analysis of genetic association studies for diabetic retinopathy. Diabetes 58: 2137–2147.
64. Gamboa-Meleendez MA, Huerta-Chagoya A, Moreno-Macias H, Vazquez-
Camarinha P, Ordonez-Sanchez ML, et al. (2012) Contribution of Common Genetic Variation to the Risk of Type 2 Diabetes in the Mexican Mestizo Population. Diabetes.
65. Gupta V, Vinay DG, Rajig S, Kranthikumar MV, Janipalli CS, et al. (2012) Association analysis of 31 common polymorphisms with type 2 diabetes and its diabetic retinopathy traits in 700 Indian sib units. Diabetologia 55: 349–357.
66. Rong R, Hanson RL, Ortiz D, Wiedrich C, Koles K, et al. (2009) Confirmation of multiple risk Loci and genetic impacts by a genome-wide association study of type 2 diabetes in the Japanese population. Hum Genet 122: 1092–1097.
67. Shetty P (2012) Public health: India’s diabetes time bomb. Nature 485: S14–16.
68. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, et al. (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316: 889–894.
69. Steinthorsdottir V, Thorleifsson G, Reynisdottir I, Benediktsson R, Jonsdottir T,
et al. (2008) SNPs in KCNQ1 are associated with susceptibility to type 2 diabetes mellitus. Nat Genet 40: 1092–1097.
70. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, et al. (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316: 889–894.
71. Tabara Y, Osawa H, Kawamoto R, Onuma H, Shimizu I, et al. (2009) Replication study of candidate genes associated with type 2 diabetes based on genome-wide screening. Diabetes 58: 493–498.
72. Chauhan G, Spurgeon CJ, Tabassum R, Bhaskar S, Kulkarni SR, et al. (2010)
Impact of common variants of PPARG, KCNJ11, TCF7L2, SLC30A8, HHEX, CDKN2A, IGF2BP2, and CDKAL1 on the risk of type 2 diabetes in 5,164 North Indian population. Diabetes 58: 2137–2147.
73. Takeuchi F, Serizawa M, Yamamoto K, Fujisawa T, Nakashima E, et al. (2009)
A systematic meta-analysis of genetic association studies for diabetic retinopathy. Diabetes 58: 2137–2147.
74. Gamboa-Meleendez MA, Huerta-Chagoya A, Moreno-Macias H, Vazquez-
Camarinha P, Ordonez-Sanchez ML, et al. (2012) Contribution of Common Genetic Variation to the Risk of Type 2 Diabetes in the Mexican Mestizo Population. Diabetes.