Calcification, Storm Damage and Population Resilience of Tabular Corals under Climate Change

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Abstract

Two facets of climate change—increased tropical storm intensity and ocean acidification—are expected to detrimentally affect reef-building organisms by increasing their mortality rates and decreasing their calcification rates. Our current understanding of these effects is largely based on individual organisms’ short-term responses to experimental manipulations. However, predicting the ecologically-relevant effects of climate change requires understanding the long-term demographic implications of these organism-level responses. In this study, we investigate how storm intensity and calcification rate interact to affect population dynamics of the table coral Acropora hyacinthus, a dominant and geographically widespread ecosystem engineer on wave-exposed Indo-Pacific reefs. We develop a mechanistic framework based on the responses of individual-level demographic rates to changes in the physical and chemical environment, using a size-structured population model that enables us to rigorously incorporate uncertainty. We find that table coral populations are vulnerable to future collapse, placing in jeopardy many other reef organisms that are dependent upon them for shelter and food. Resistance to collapse is largely insensitive to predicted changes in storm intensity, but is highly dependent on the extent to which calcification influences both the mechanical properties of reef substrate and the colony-level trade-off between growth rate and skeletal strength. This study provides the first rigorous quantitative accounting of the demographic implications of the effects of ocean acidification and changes in storm intensity, and provides a template for further studies of climate-induced shifts in ecosystems, including coral reefs.

On coral reefs, mortality rates of coral colonies are elevated by severe summer storms and cyclones, which dislodge colonies from the substrate, particularly in wave-exposed, highly productive habitats like reef crests [12], [15]. Such mechanical disturbances limit the dominance of fast-growing, mechanically unstable coral growth forms, and facilitates the maintenance of high local diversity in coral assemblages [16]. Predicted increases in the intensity of storms and cyclones [3] are likely to increase mortality in biomechanically vulnerable species. Moreover, the effects of storms may be exacerbated by declining rates of calcification caused by ocean acidification and thermal stress [17]. Decreased calcification by corals, if manifested as slower colony growth rates [18], may also have implications for lifetime reproductive output, which is strongly related to colony size and longevity [19].

Coral species with tabular growth forms are particularly important ecosystem engineers on wave-exposed Indo-Pacific reefs (Figure 1A). They grow and calcify rapidly compared to other growth forms, allowing them to dominate reef crest communities [20], [21]. Consequently, they are major contributors to calcification and reef accretion in these habitats. Table corals are key contributors to reef structural complexity. They harbour distinctive understory communities [21] and provide shelter from predation and high flow for many mobile species, especially small fishes [22] and juvenile parrotfishes, which as
adults play a crucial functional role controlling macroalgae on reefs [23]. Some tabular species are essential prey of corallivorous butterflyfishes, which decline markedly in abundance without them [24] (Figure 1B). However, the same traits that make table corals important ecosystem engineers also make them particularly susceptible to climate change. They are vulnerable to dislodgement by storm-generated waves (Figure 1C) [12], [16] and susceptible to ocean acidification, thermal stress, coral bleaching and disease [25], [26].

In this study, we examine the effects of changes in storm intensity and calcification rates on population growth of the ecologically dominant table coral *Acropora hyacinthus*, a pandemic reef-building species on wave-exposed Indo-Pacific reefs [27]. We compare estimates of coral population growth under atmospheric CO₂ scenarios for the Pre-Industrial Revolution (PIR), present day, and projections under two future climate scenarios. We incorporate into this model: (1) shifts in aragonite saturation state ($\Omega_{arag}$) driven by increasing pCO₂ and sea surface temperature (SST) [28] based on a range of empirically-calibrated relationships between $\Omega_{arag}$ and SST and coral calcification rate [3], [29], [30], and (2) a field-validated mechanistic model relating the severity of tropical storm events to the dislodgment mortality of corals of different sizes [12]. Reductions in calcification may be manifested in a combination of two ways: by reduced colony growth rates, and by reduced skeletal density [17], [4]. A decline in colony growth reduces survival and reproductive rate, since colonies will be in smaller, less fecund, and remain in more vulnerable size classes for longer than under normal growth rates [31]. Conversely, reduced skeletal density increases the vulnerability of larger colonies to dislodgement during storms. We therefore consider both types of responses to calcification, to capture the range within which the true response will lie. Because ocean acidification is expected to decrease the mechanical integrity of reef substrate due to declines in inorganic cementation and increases in bioerosion [32], and this, in turn, increases the risk of colony dislodgement during storms [33], we also consider the potential effects of decreases in the strength of reef substrate. Therefore, we examine 12 scenarios in all, corresponding to combinations of (i) weak, intermediate, or strong relationships between calcification and $\Omega_{arag}$ and SST, (ii) whether growth rate and/or skeletal density decline as calcification decreases, and (iii) whether reef substrate strength is weak or strong, reflecting both contemporary spatial variation in reef lithification [34], and the potential for a future weakening of substrate strength due to ocean acidification.

![Figure 1. Tabular corals that provide habitat structure, shelter and food for associated reef organisms.](https://example.com/figure1.png)

(A) Wave-exposed coral communities are often dominated by tabular corals (photo: Andrew Baird). (B) The obligate corallivore, *Chaetodon trifascialis*, feeds almost exclusively on the pandemic study species, *Acropora hyacinthus* (photo: Morgan Pratchett). (C) Tabular growth forms are particularly vulnerable to mechanical dislodgement during summer storms.

doi:10.1371/journal.pone.0046637.g001
Materials and Methods

Reef Mechanical Environment

The hydrodynamic dislodgement of coral colonies can be expressed as the dimensionless inequality [12]:

\[
\frac{\sigma_y}{U^2 \rho_w} \leq \frac{16}{d_w DMT^3 \pi} \int_{y=0}^{h} y w(y) dy
\]  

(1)

The left-hand side of the equation describes the mechanical environment, where \( \sigma_y \) is the limiting material tensile strength (either reef substrate or coral skeleton; N m\(^{-2}\)), \( U \) is the expected yearly maximum water velocity (ms\(^{-1}\)), and \( \rho_w \) is seawater density (~1025 kg m\(^{-3}\)). The right-hand side—the Colony Shape Factor (CSF)—is a measure of mechanical vulnerability described by the projection of colony shape above the substrate \( (\alpha(y)) \) is the projected width, \( y \) is distance above the substrate and \( h \) is the height of the colony and the basal attachment perpendicular widths \( (d_1 \text{ and } d_2) \) (a consistent length unit is required for all CSF parameters; e.g., meters). Dislodgement of an individual within the population is expected if a DMT generated by a wind event becomes equal to or less than its CSF [12]. The material density and tensile strength \( (\sigma_y) \) of both reef substrate and Acropora hyacinthus were measured at the southeast reef at Lizard Island in an earlier study [33], which found the substrate to be approximately an order of magnitude weaker than the coral and therefore limiting whole-colony mechanical integrity under present-day environmental conditions. The expected yearly maximum water velocity \( (U) \) was calculated at the same reef crest site based on the reconstruction of a 37-year history of wind conditions at the study site [35]. We used this field-validated wind-fetch and wave attenuation model to calculate how changes in storm intensity (equivalent to maximum sustained wind speed) translate into maximum wave orbit water velocity at the substrate. We used the fitted curvilinear relationship between regional winds \( U_{\text{wind}} \) and colony-level water velocities at the reef crest \( (U = -0.0036 U_{\text{wind}}^2 + 0.2139 U_{\text{wind}} - 0.0016, r^2 = 0.9899) \) to calculate how proportional changes in storm intensity translate approximately into maximum water velocity (e.g., a 10% increase in expected storm intensity is assumed to translate in to a 10% increase in yearly maximum regional wind speed). Shifts in storm intensity in the model are based on the expected 2 to 11% increase of global intensities by 2100 [3].

Changes to material strength and yearly colony growth were based on proportional changes to coral calcification \( G \) in response to sea surface temperature \( (\text{SST}) \) and aragonite saturation state \( (\Omega_{\text{arag}}) \). SST and \( \Omega_{\text{arag}} \) estimates were used for a range of stabilized atmospheric CO\(_2\) levels, ranging from historical Pre-Industrial Revolution \( (\text{PIR}) \) conditions \( (280 \text{ ppm}) \), through present-day \( (380 \text{ ppm}) \), to two future climate change scenarios \( (\text{doubling of PIR: } 560 \text{ ppm}; \text{and doubling of present-day: } 750 \text{ ppm}) \) [20]. Proportional changes in calcification (relative to present-day) were estimated in three ways to capture the likely range of calcification responses of reef corals to future mean SST and \( \Omega_{\text{arag}} \) (Figure 2):

a) The “strong” response is based on an empirically-derived equation for whole-reef aragonite precipitation \( (\text{Eq. 6 in [5]}) \), which considers both changing SST and \( \Omega_{\text{arag}} \):

\[
G_{\text{strong}} = \frac{T G_{\text{gross}}(\text{SST}, \Omega_{\text{arag}})}{T G_{\text{gross}}(\text{SST} = 31.8, \Omega_{\text{arag}} = 0)}
\]

where \( T G_{\text{gross}} \) is coral gross calcification and \( \text{SST} = 31.8 \) and \( \Omega_{\text{arag}} = 0 \) are present-day temperature and saturation state values.

b) The “intermediate” response is based on an experimentally derived relationship for reef corals in a microcosm (equation from Fig. 2 caption in [36]), which was later supplemented with other calcification studies \( (\text{Eq. 1 and the first order response in Fig. 9 in [30]) and considers changes in } \Omega_{\text{arag}} \text{ only. The two responses are similar and we refer to [30] from this point on.}

\[
G_{\text{intermediate}} = \frac{25.60 \Omega_{\text{arag}} - 31.8}{25.60 \Omega_{\text{arag}} - 31.8}
\]

c) The “weak” response is based on the linear model fitted to experimental data for a branching congener of Acropora hyacinthus, Acropora intermedia, for two SST and three \( \Omega_{\text{arag}} \) experimental treatments [29], where coefficient estimates were calculated from the paper’s online supporting ANOVA table:

\[
G_{\text{weak}} = \frac{19.82 + 2.04 \Omega_{\text{arag}} - 0.51 \text{SST} - 0.038 \Omega_{\text{arag}} \text{SST}}{19.82 + 2.04 \Omega_{\text{arag}} - 0.51 \text{SST} - 0.038 \Omega_{\text{arag}} - 0.038 \text{SST} 0.038 \Omega_{\text{arag}} - 0.038 \text{SST}}
\]

Other studies of calcification responses for the genus Acropora fall within this range and consider changes in \( \Omega_{\text{arag}} \) only (e.g., [37], [38]).

The proportional change in calcification was applied to coral skeleton density or growth as per the scenarios outlined earlier (e.g., a 10% reduction in calcification relative to present-day translates into either a 10% reduction in skeletal density or a 10% reduction in added planar area per year). Changes in skeletal strength were calculated based on an empirically derived relationship between aragonite density \( \rho_a \) and tensile strength as a function of stabilized atmospheric CO\(_2\) scenarios.

Figure 2. Predicted proportional changes in calcification rate as a function of stabilized atmospheric CO\(_2\) scenarios. Changes are shown relative to present-day (380 ppm) for three published calcification responses: weak [30], intermediate [29] and strong [5].
doi:10.1371/journal.pone.0046637.g002
Little is known about how the material properties of reef substrate are related to ambient levels of SST and $a_{v,ar}$. Therefore, in scenarios with reef substrate weakening, we assumed that substrate density changes similarly to skeleton density (as calcification changes), and we used the reported relationship between density and strength [4] to calculate substrate strength. Our projected estimates based on these assumptions are likely to be conservative, because substrate in the future will be comprised of coral skeleton that is presumably weaker than it is presently, and the processes cementing this skeleton are expected to diminish as aragonite saturation state declines [32].

\[
\sigma_t = 0.49e^{2.24a_v}
\]

Figure 3. The reef coral mechanical environment. Mean expected yearly mechanical threshold ($DMT$) as a function of storm intensity and atmospheric CO$_2$ scenario for reef calcification scenarios: strong (A, D), intermediate (B, E), and weak (C, F) applied to both reef substrate and coral (A–C) and coral only (D–F). The black points represent present-day estimates of mean yearly $DMT$. Shaded lines and areas represent parameters used in the IPM, including the pre-Industrial Revolution and two 2100 scenarios (doubling of Pre-Industrial Revolution [560 ppm] and doubling of present-day [750 ppm]). For reference, coral photographs illustrate $DMT$ levels that would theoretically dislodge tabular colonies based on their shape.

doi:10.1371/journal.pone.0046637.g003

Integral Projection Model

An integral projection model (IPM) [39] was used to translate changes in the mechanical environment into three population-level measures: cover, lifetime reproductive output, and intrinsic population growth rate. Cover is the sum of individual colony areas in a population. Lifetime reproductive output is the reproductive output of a colony or cohort integrated over the average lifespan. The intrinsic population growth rate is the per-capita propensity for population regeneration following a reduction in population size to very low levels (e.g., due to a tropical storm, bleaching event or crown of thorns outbreak). The IPM framework is well-suited to corals, because colony size is treated continuously, avoiding coarse and arbitrary size classifications, and thereby facilitating a more precise characterization of colony growth, fecundity and mechanical vulnerability, all of which are highly dependent on colony size. The parameterization of the projection model is based on a well-studied population of *A. hyacinthus* living on the physically exposed reef crest on the southeast reef at Lizard Island, Great Barrier Reef, Australia. We modeled the number of individuals in the population of size $x$ at time $t + 1$ given the number of individuals of size $x$ at time $t$ [39]:

\[
\frac{dN(t+1|x)}{dt} = R(N(t|x)) - m(N(t|x)) - d(N(t|x))
\]
which is made up of probability functions for yearly growth \(g\), survivorship \(s\), and recruitment \(r\). The growth function was estimated using permanent quadrat data at the study site [Hughes, unpublished data]. The planar area of colonies of standalone (i.e., uncrowded) \(A. \text{hyacinthus}\) was calculated from digitized photo-quadrats from one year to the next to estimate growth rate based on the change in planar area over time \((n = 45)\). Growth in standalone colonies best reflects unconstrained population growth, such as recovery following a storm disturbance while space is not limiting and competitive pressure is small. Colonies missing at \(t+1\) were excluded for calculating yearly growth.

Survivorship was modeled as:

\[
s(x) = (1 - b)\gamma(x),
\]

where \(1 - b\) is the background yearly survivorship (excluding hydrodynamic disturbance mortality), and \(\gamma(x)\) is the probability of dislodgment for a colony size \(x\) due to hydrodynamic disturbance. We develop \(\gamma(x)\) using previously-calculated values of CSF and size (planar area) for \(A. \text{hyacinthus}\) from the outermost, 20 m wide, section of the exposed reef crest at the study site \((n = 220)\) [12]. For a given year, the expected distribution of Dislodgement Mechanical Thresholds (DMTs) was calculated according to Eq. 1. Therefore, the hydrodynamic disturbance survivorship function becomes:

\[
\gamma(x) = \int [1 - dmt(csf)]f(csf|x)\, dcdf,
\]

where \(f(csf | x)\) is the probability density function of CSF for a colony of size \(x\), and \(dmt(csf)\) is the Gumbel cumulative density function of minimum yearly DMT from 1967–2003 reported by [35].

Figure 4. Parameterizing the population model using empirical demographic data. (A) \(A. \text{hyacinthus}\) colony planar area \((m^2)\) at year \(t+1\) plotted against area at year \(t\) at the exposed reef crest study location. The unity line (intercept 0 and slope 1) illustrates that the majority of points fall in the region of increasing size. (B) Colony shape factor (CSF; dimensionless) as a function of colony planar area \((m^2)\) of \(A. \text{hyacinthus}\) colonies. Dashed lines in both panels represent 95% prediction intervals. (C) Log-likelihood profile for integral projection model recruitment parameter. The dashed line shows the log-likelihood 95% confidence bounds. (D) Colony size density distribution of \(A. \text{hyacinthus}\) at the study site (bars) and the best-fit model stable size distribution as a result of optimizing the recruitment parameter. Shaded area illustrates 95% log-likelihood confidence intervals. doi:10.1371/journal.pone.0046637.g004
Yearly background mortality $b$ was estimated from the quadrat data as the proportion of colonies at time $t$ that were dead at $t+1$. Two-thirds of the colonies that were dead at $t+1$ died during periods when Lizard Island experienced large cyclonic wave conditions (1990–1991 [Cyclone Ivor] and 1998–1999 [Cyclone Rona]). Therefore, we calculated the number of colonies that died in each year, out of the total number observed in the data (excluding the two cyclone years), and used binomial likelihood to estimated background mortality probability.

Because $A. hyacinthus$ is a broadcast spawner, we first modeled the population as an open system, which assumed that recruitment is largely independent of the local population and that the majority of recruits are from habitats insensitive to the modeled climate variables. Minimum colony size was set to $10^{-2.5}$ m$^2$ (~7 cm diameter), which is the size of first recruitment in the IPM, corresponding to the approximate size of first reproduction for $A. hyacinthus$ [19], [40]. For a given recruitment rate, we projected the population from year to year for each storm intensity and calcification rate scenario until the population stabilized (typically 5–25 years). When expressed relative to present-day cover, the differences in cover among scenarios were insensitive to initial population structure or recruitment rate. To calculate relative lifetime reproductive output, we seeded the population and projected it without recruitment until it went extinct, and then summed the total area of colonies from each year of the projection. Relative lifetime reproductive output was insensitive to the number of recruits in the projected cohort.

Because changes to storm intensity and calcification are large-scale phenomena, and $A. hyacinthus$ tends to occupy the wave-exposed habitats most vulnerable to such changes, increased mortality or reduced growth of adult colonies is likely to have substantial feedback effects on reproductive output at the metapopulation level, and thus recruitment is also likely to decrease over time. To assess the demographic implications of such feedback effects on recruitment, we used a closed population model, in which the supply of recruits declined in proportion to the reproductive output of the local population. The closed model assumes that we capture the global pool of adults and that variables, such as substrate strength, flow regime and mortality, are similar for all populations. We then assume that the number of colonies entering the population at $t+1$ is proportional to the total planar area of colonies in the population at $t$ [19]. Specifically,

Figure 5. Projected coral cover under alternative future CO2 stabilization scenarios. pCO2 is assumed to affect demographic processes through different mechanisms in each panel: (A) coral and substrate weakening, (B) coral growth decline and substrate weakening, (C) coral weakening only, and (D) coral growth decline only. Curves represent the three published calcification responses to $\Omega_{\text{arag}}$ and SST: low, intermediate and high. Shaded areas capture the 2 to 11% range of predicted increases in future storm intensity [3].

doi:10.1371/journal.pone.0046637.g005
The recruitment was modeled as:

\[
r(x, y) = \begin{cases} 
qx & \text{if } x > 10^{-2.5} \text{ and } y \leq 10^{-2.5} \\
0 & \text{if } x \leq 10^{-2.5} \text{ for all } y
\end{cases}
\]  

(5)

where \( q \) is the number of colonies recruiting back to the population as \( 10^{-2.5} \) m\(^2\) colonies for a given area of adult colony. We estimate the recruitment parameter by varying \( q \) until the predicted colony size distribution (the eigenvector associated with the IPM’s leading eigenvalue [39]) best fit the empirical size probability density distribution for the study site [12]. Maximum log-likelihood was used to find the best-fit parameter \( q \) and the log-likelihood confidence intervals were used as estimates of uncertainty associated with the recruitment parameter. We then used the long-run density-independent population growth rate of this population, the dominant eigenvalue \( \lambda \), to quantify the population-dynamic consequences of increases in storm intensity and declines in calcification rate, because it expresses the different demographic effects of such environmental changes in the common currency of the capacity of a coral colony to contribute to population growth [41].

**Results**

**Reef Mechanical Environment**

The mean expected yearly mechanical threshold (DMT) showed a range of potential trajectories depending on environmental change and coral response scenario (Figure 3). For scenarios in which reef substrate strength declines as a function of calcification potential—and regardless of if coral individuals invest in colony growth or skeletal density (strength)—the mean dislodgement threshold is expected to decrease by up to four-fold (a given colony is four times more likely to be hydrodynamically dislodged) by the end of the century, depending on the calcification response to changing atmospheric CO\(_2\) levels (Figure 3A–C).
decreases in DMT are driven primarily by reduction in material strength and secondarily by increases in maximum yearly water velocities (Eq. 1), and were especially pronounced for the “high” reef calcification scenario (Figure 3A). For scenarios in which reef substrate strength is unaffected by future pCO2 levels, and coral colonies maintain fast colony growth despite declines in skeletal density, decreases in DMT are driven primarily by storm intensity and change relatively little by century’s end. An exception is for the high calcification response scenario, where a point is reached at approximately 600 ppm where coral skeleton became weaker than the reef substrate, and DMT declines precipitously (Figure 3D). For scenarios in which substrate strength is unaffected by future pCO2 levels and coral colonies sacrifice colony growth in order to maintain skeletal strength, decreases in DMT are driven only by storm intensity (identical to that shown in Figure 3E, F).

**Integral Projection Model**

Colony growth for *A. hyacinthus* colonies is well described by a power-law relationship between colony sizes in successive years ($X_{t+1} = 0.032 \times 10^{0.86}$, Figure 4A). The slope of the relationship is less than one (0.86), indicating that added area (as a proportion of size) declines as colonies grow. The relationship between colony mechanical vulnerability and size is also captured well by a power-law relationship (Figure 4B). We estimated the non-cyclone induced background mortality probability as 0.066 per annum (95% confidence intervals: 0.017–0.164), and assume that this level is constant from year to year. This result is consistent with estimates from other studies at Lizard Island [42].

When modeled as an open system, population projections indicate declines in *A. hyacinthus* cover of more than twofold by the end of the century in 8 of the 12 scenarios we examined (Figure 5). Projected lifetime reproductive output showed almost identical patterns relative to present-day. Variation in the magnitude of projected change among our 12 scenarios indicates that the population-level response to increasing atmospheric CO2 depends primarily on the sensitivity of substrate strength to reef calcification rate. Decline is rapid if the strength of the reef substrate diminishes at a rate similar to that of in situ measurements of community calcification rate (red curves, Figure 5A, B). Decline is intermediate if substrate strength diminishes in proportion to measurements of the calcification rate of individual coral colonies (Figure 5A, B; yellow and green curves), while decline is small if substrate strength is insensitive to $\Omega_{arag}$ and thus changes are driven primarily by storm intensity (Figure 5C, D). Regardless of the reef substrate response to acidification, population cover declines less rapidly if colonies also reduce their skeletal density to maintain growth rate (Figure 5A), except when coral skeletal strength falls below that of the reef substrate, at which point population growth rate declines precipitously (Figure 5C, red curve). Similar thresholds occur for the other intermediate and weak calcification response curves in Figure 5C, but these thresholds lie at atmospheric CO2 levels beyond those explored here.

When modeled as a closed system, the IPM eigenvector that best fit the empirical size structure data had a recruitment parameter $q$ at time $t$ of 7.4 recruits per m$^2$ of colony planar area at time $t$ (Figure 4C). The log-likelihood profile 95% confidence interval ranged from 4.7 to 11.7 recruits per year per colony area. Given this best-fit recruitment rate, the model fitted the empirical size distribution data reasonably well (Figure 4D) and the two were statistically indistinguishable (two-sided Kolmogorov-Smirnov test: $D = 0.2$, $p = 0.2719$). Parameterizing the recruitment function with the mean and confidence intervals for $q$, the integrated projection model suggests that populations are currently able to replenish themselves (Figure 6), with a mean annual population growth rate, $\lambda$, of approximately 1.036 (implying a population doubling time of 12 years; 95% confidence bounds ranging from 1.007 to 1.110). By the end of this century, $\lambda$ is predicted to decline below unity, indicating loss of the capacity for self-replenishment, in 7 of the 12 doubling Pre-Industrial Revolution and 9 of the 12 doubling present-day scenarios we examined. In contrast with predictions for coral cover, $\lambda$ is predicted to decline marginally faster as a function of atmospheric CO2 level when corals maintain growth rate and sacrifice skeletal density (Figure 6A, B).

**Discussion**

Our study focuses specifically on two environmental changes likely to be associated with anthropogenic effects on climate (increased storm intensity and decreasing aragonite saturation state due to the interaction between ocean acidification and increasing temperature), and it explores their demographic effects on adult coral growth and mortality from storms. The model translates these environmental changes into individual-level growth and mortality probability, thereby providing a more mechanistic basis for population-level responses than more traditional phenomenological approaches that consider only the aggregate dynamics of coral cover. Our results indicate that the environmental change scenarios we examined will impact levels of cover and population resilience of *A. hyacinthus*. The magnitude of this impact depends primarily on how SST and $\Omega_{arag}$ influences reef substrate mechanical integrity, which is currently poorly understood [32], and secondarily on the degree to which colony calcification diminishes, for which a range of scenarios exist [43]. Change in colony calcification introduces an important demographic trade-off between maintaining growth rate (greater reproductive output) and skeletal density (lower storm-induced mortality), which requires further research to understand the potential for adaptation within the physiological and energetic constraints imposed by this trade-off [44]. Finally, increases in storm intensity expected by 2100 appear to play a relatively minor role in long-term population persistence, compared to calcification responses and individual-level demographic trade-offs.

Our projections are likely to be somewhat conservative because we omit other human impacts that reduce coral reproduction and growth and/or increase mortality [45] (e.g., pollution, overfishing and other effects of rising CO2 and SST). For example, one of the most biologically significant effects of increasing temperatures on corals is increases in the frequency and severity of coral bleaching, which can cause mass mortality, increase susceptibility to disease, and reduce subsequent growth and reproduction [46]. The frequency and intensity of bleaching events has been increasing over the past several decades [46], and, while there is growing evidence of acclimation and adaptation to warmer temperatures [43], it is unlikely that such physiological and evolutionary changes will occur rapidly enough to avoid adverse consequences entirely [6], [29], [43]. Furthermore, our approach does not include any effect of ocean acidification on pre- and post-settlement stages [13], [14]. Finally, the combined impacts of ocean acidification, pollution, overfishing and other environmental changes are likely to change species composition and interactions [23], [26], including reductions in herbivore abundances that mediate shifts from corals to algae or other weedy species [45], [47–49].

By developing explicit mechanistic connections between size-dependent demographic rates and environmental feedbacks, our study makes important advances on earlier projections of the effects of climate change on coral persistence, which model the
show that tabular corals are prone to large and rapid declines in coral cover, and to population collapse, due principally to increased vulnerability to storm-induced dislodgment as a consequence of ocean acidification and decreased lifetime reproductive output. Because the top-heavy growth forms of tabular and arborescent corals makes such species particularly susceptible to dislodgment, these effects are likely to be manifested as a shift towards lower coral cover overall, and towards coral assemblages more dominated by structurally simpler, more mechanically stable species that are less productive and offer less shelter and food for other coral reef organisms.

Acknowledgments

We especially thank D. Allen, M. Baskett, D. Falster and E. Madin for helpful discussions. L. Cao provided SST and \( \Omega_{\text{arag}} \) data.

Author Contributions

Conceived and designed the experiments: JSM SRC. Performed the experiments: JSM. Analyzed the data: JSM. Contributed reagents/materials/analysis tools: JSM TPH. Wrote the paper: JSM SRC TPH.

References

1. Kleypas JA, Buddemeier RW, Archer D, Gattuso JP, Langdon C, et al. (1999) Geochemical consequences of increased atmospheric carbon dioxide on coral reefs. Science 284: 118–120.
2. IPCC (2007) Summary for policymakers. Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change (IPCC). Solomon S, et al., editors. Cambridge: Cambridge University Press.
3. Knutson TR, McBride JL, Chan J, Emanuel K, Holland G, et al. (2010) Tropical cyclones and climate change. Nat Geosci 3: 157–163.
4. Madin JS, O'Donnell MD, Connolly SR (2008) Climate-mediated mechanical changes to post-disturbance coral assemblages. Biol Lett 4: 490–493.
5. Silverman J, Lazar B, Cao L, Caldeira K, Erz J (2009) Coral reefs may start dissolving when atmospheric CO2 doubles. Geophys Res Lett 36: L05606.
6. Baskett ML, Gaines SD, Nisbet RM (2009) Symbiont diversity may help coral reefs survive moderate climate change. Ecol Appl 19: 3–17.
7. Anthony KRN, Maynard JA, Diaz-Pulido G, Mumby PJ, Marshall PA, et al. (2011) Ocean acidification and warming will lower coral reef resilience. Global Change Biol 17: 1798–1808.
8. Helmut B, Kingsolver JG, Carrington E (2005) Biophysics, physiological ecology, and climate change: does mechanism matter? Annu Rev Physiol 67: 177–201.
9. Keane R, Porter W (2009) Mechanistic niche modelling: combining physiological and spatial data to predict species’ ranges. Ecol Lett 12: 334–350.
10. Denny MW, Gaylord B (2010) Marine Ecomechanics. Ann Rev Mar Sci 2: 89–114.
11. Hughes TP (1984) Population dynamics based on size rather than age: a general model with a reef coral example. Am Nat 123: 778–795.
12. Madin JS, Connolly SR (2006) Ecological consequences of major hydrodynamic disturbances on coral reefs. Nature 444: 477–200.
13. Kroeker KJ, Kordas RL, Crum RN, Singh GG (2010) Meta-analysis reveals negative yet variable effects of ocean acidification on marine organisms. Ecol Lett 13: 1419–1434.
14. Albright R, Langdon C (2011) Ocean acidification impacts multiple early life history processes of the Caribbean coral P. astreoides. Global Change Biol 17: 2487–2487.
15. Massel S, Done T (1995) Effects of cyclone waves on massive coral assemblages on the Great Barrier Reef: metroecology, hydrodynamics and demography. Coral Reefs 12: 153–166.
16. Connell JH (1978) Diversity in tropical rain forests and coral reefs. Science 199: 1302–1310.
17. Cooper TF, De’ath G, Fabricius KE, Lough JM (2008) Declining coral calcification in massive Pocillopora in two nearshore regions of the northern Great Barrier Reef. Global Change Biol 14: 529–538.
18. McCulloch M, Faler J, Trotter J, Montagna P (2012) Coral resilience to ocean acidification and global warming through pH up-regulation. Nature Climate Change 2: 1–5.
19. Hall VR, Hughes TP (1996) Reproductive strategies of modular organisms: comparative studies of reef-building corals. Ecology 77: 950–963.
20. Stimson J (1985) The effect of shading by the table coral A. hyacinthus on understory corals. Ecol 66: 40–53.
21. Baird AH, Hughes TP (2000) Competitive dominance by tabular corals: An experimental analysis of recruitment and survival of understory assemblages. J Exp Mar Bio Ecol 251: 117–132.
22. Pratchett MS, Marmane MJ, Berumen ML, Eagle JE, Pratchett DJ (2008) Habitat associations of juvenile versus adult butterflyfishes. Coral Reefs 27: 541–551.
23. Bellwood DR, Hughes TP, Folke C, Nyström M (2004) Confronting the coral reef crisis. Nature 429: 827–833.
24. Berumen ML, Pratchett MS (2008) Trade-offs associated with dietary specialization in coralivorous butterflyfishes (Chaetodontidae: Chaetodon). Behav Ecol Sociobiol 62: 989–994.
25. Marshall PA, Baird AH (2000) Bleaching of corals on the Great Barrier Reef: differential susceptibilities among taxa. Coral Reefs 19: 155–163.
26. Hughes TP, Baird AH, Dinsdale EA, Moltschaniwskyj NA, Pratchett MS, et al. (2012) Assembly rules of reef corals are flexible along a steep climatic gradient. Coral Reefs 22: 1–6.
27. Veron JEN, Wallace CC (1984) Scleractinia of Eastern Australia, Part 5: Acroporidae. Sydney: Australian Institute of Marine Science Monograph Series, Angus and Robertson.
28. Cao L, Caldeira K (2008) Atmospheric CO2 stabilization and ocean acidification. Geophys Res Lett 35: L09609.
29. Anthony KRN, Kline DI, Diaz-Pulido G, Dove S, Hoegh-Guldberg O (2008) Ocean acidification causes bleaching and productivity loss in coral reef builders. Proc Natl Acad Sci U S A 105: 17442–17446.
30. Langdon C, Atkinson MJ (2005) Effect of elevated pCO2 on photosynthesis and calcification of corals and interactions with seasonal change in temperature/irradiance and nutrient enrichment. J Geophys Res 110: C09007.
31. Hughes TP, Jackson JBC (1985) Population dynamics and life histories of foliaceous corals. Ecol Monographs 55: 141–166.
32. Manzello DP, Kleypas J, Budd D, Eakin C, Glynn P, et al. (2008) Poorly cemented coral reefs of the eastern tropical Pacific: Possible insights into reef development in a high-CO2 world. Proc Natl Acad Sci U S A 105: 10450–10455.
33. Madin JS (2005) Mechanical limitations of reef corals during hydrodynamic disturbances. Coral Reefs 24: 630–635.
34. Marshall JF (1985) Cross-shelf and facies related variations in submarine cementation in the Central Great Barrier Reef. Proc Fifth Int Coral Reef Cong Tahiti 3: 221–226.
35. Madin JS, Black KP, Connolly SR (2006) Scaling water motion on coral reefs: from regional to organismal scales. Coral Reefs 25: 635–622.
36. Langdon C (2002) Review of experimental evidence for effects of CO2 on calcification of reef builders. Proc 9th Int Coral Reef Symp 2: 1091–1098.
37. Schneider K, Erz J (2006) The effect of carbonate chemistry on calcification and photosynthesis in the hermatypic coral Acropora hyacinthus. Limnol Oceanogr 51: 1294–1293.
38. Anthony KRN, Kleypas JA, Gattuso JP (2011) Coral reefs modify their seawater carbon chemistry: implications for impacts of ocean acidification. Global Change Biol 17: 3655–3666.
39. EASTERLING MR, Elsner JP, DSS (2000) Size-specific sensitivity: applying a new structured population model. Ecology 81: 694–708.
40. Wallace CC (1985) Reproduction, recruitment and fragmentation in nine species of the coral genus A. Mar Biol 88: 217–233.
41. Caswell H (2001) Matrix population models: Construction, analysis and interpretation, 2nd Edition. Massachusetts: Sinauer Associates.
42. Wakefield M, Done TJ, Johnson CR (2008) Decadal trends in a coral community and evidence of changed disturbance regime. Coral Reefs 27: 1–13.
43. Pandolfi JM, Connolly SR, Marshall DJ, Cohen AL (2011) Projecting coral reef futures under global warming and ocean acidification. Science 333: 418–422.
44. Madin JS, Hoogenboom MO, Connolly SR (2012) Integrating physiological and biomechanical drivers of population growth over environmental gradients on coral reefs. J Exp Biol 215: 968–976.
45. Hughes TP, Baird AH, Bellwood DR, Card M, Connolly SR, et al. (2003) Climate Change, Human Impacts, and the Resilience of Coral Reefs. Science 301: 929–933.
46. Baker AC, Glynn PW, Riegl B (2008) Climate change and coral reef bleaching: An ecological assessment of long-term impacts, recovery trends and future outlook. Estuar Coast Shelf Sci 80: 435–471.
47. Diaz-Pulido G, Gouveia M, Tilbrook B, Dove S, Anthony K (2011) High CO2 enhances the competitive strength of seaweeds over corals. Ecol Lett 14: 156–162.
48. Hughes TP (1994) Catastrophes, phase-shifts, and large-scale degradation of a Caribbean coral reef. Science 265: 1547–1551.
49. Mumby PJ, Dahlgren CP, Harborne AR, Kappel CV, Micheli F, et al. (2006) Fishing, trophic cascades, and the process of grazing on coral reefs. Science 311: 98–101.
50. Annan JD, Hargreaves JC (2006) Using multiple observationally-based constraints to estimate climate sensitivity. Geophys Res Lett 33: L06704.