Lycopene and male infertility

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Excessive amounts of reactive oxygen species (ROS) cause a state of oxidative stress, which result in sperm membrane lipid peroxidation, DNA damage and apoptosis, leading to decreased sperm viability and motility. Elevated levels of ROS are a major cause of idiopathic male factor infertility, which is an increasingly common problem today. Lycopene, the most potent singlet oxygen quencher of all carotenoids, is a possible treatment option for male infertility because of its antioxidant properties. By reacting with and neutralizing free radicals, lycopene could reduce the incidence of oxidative stress and thus, lessen the damage that would otherwise be inflicted on spermatozoa. It is postulated that lycopene may have other beneficial effects via nonoxidative mechanisms in the testis, such as gap junction communication, modulation of gene expression, regulation of the cell cycle and immunoenhancement. Various lycopene supplementation studies conducted on both humans and animals have shown promising results in alleviating male infertility—lipid peroxidation and DNA damage were decreased, while sperm count and viability, and general immunity were increased. Improvement of these parameters indicates a reduction in oxidative stress, and thus the spermatozoa is less vulnerable to oxidative damage, which increases the chances of a normal sperm fertilizing the egg. Human trials have reported improvement in sperm parameters and pregnancy rates with supplementation of 4–8 mg of lycopene daily for 3–12 months. However, further detailed and extensive research is still required to determine the dosage and the usefulness of lycopene as a treatment for male infertility.

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INTRODUCTION

There has been increasing evidence in recent years that oxidative stress plays a vital role in the pathogenesis of idiopathic male factor infertility. Elucidating the value of antioxidant supplementation as a treatment option for infertility has therefore become a goal for many researchers. Extensive research has been conducted to show that antioxidants like vitamins E and C and carnitines help in reducing oxidative stress by quenching free radicals.1 However, less is known about the effectiveness of carotenoids, especially that of lycopene—a potent antioxidant and singlet oxygen quencher.2 In this review, we will (i) explain how oxidative stress can cause infertility, (ii) synthesize pertinent information on lycopene, (iii) discuss the possibility of lycopene supplementation as a treatment option for idiopathic male factor infertility and (iv) give a detailed analysis of various human and animal studies involving lycopene that have been conducted both in vivo and in vitro.

OXIDATIVE STRESS AND ITS EFFECTS ON MALE REPRODUCTION

A free radical refers to a molecule that has at least one unpaired electron,3,4 which is responsible for the molecule’s short-lived high energy state that causes instability and extreme reactivity.3 These free radicals will take part in propagative chain reactions and generate even more free radicals until two such radicals react and the unpaired electrons are neutralized.4 In the process, membrane lipids, amino acids and carbohydrates in nucleic acids will be attacked by the free radicals and undergo oxidation.4 Examples of highly reactive oxygen radicals include superoxide anions, hydroxyl radicals and hypochlorite radicals; and these are collectively known as reactive oxygen species (ROS).5 ROS are byproducts of oxygen metabolism and physiological amounts play important roles in sperm function, such as in capacitation, acrosome reaction, hyperactivation and sperm-oocyte fusion.5 Under normal conditions, there are natural antioxidants—both enzymatic and nonenzymatic—present in the seminal plasma to ensure that ROS concentrations remain low. Enzymatic antioxidants include glutathione reductase, superoxide dismutase and catalase; while nonenzymatic oxidants include vitamins C, E and B, carotenoids and carnitines. When ROS levels are greatly increased or antioxidant levels substantially decreased such that the delicate balance between ROS and antioxidants is disturbed, oxidative stress occurs.6,7

Several studies have shown a link between oxidative stress and idiopathic male factor infertility.6,7 This is largely due to the fact that infertile patients have been found to produce a greater amount of abnormal spermatozoa, which generate more ROS and less antioxidants, therefore leading to oxidative stress.6,8 Oxidative stress affects spermatozoa in three main ways—membrane lipid peroxidation, DNA damage and induction of apoptosis.9,10 However, the extent of damage caused depends on the nature of the ROS involved and the environment of the sperm.11 Cell membranes of spermatozoa are rich in polyunsaturated fatty acids, especially docosahexaenoic acid, which makes them more susceptible to oxidative damage by free radicals.11,12 Polyunsaturated
fatty acids consist of numerous unconjugated double bonds containing many electrons. These electrons are donated to ROS upon reaction and lead to the generation of lipid peroxides. As a result, the fluidity of the spermatozoal cell membrane is disrupted, thus negatively affecting sperm motility and viability. Sperm motility will be affected because of the decrease in axonemal protein phosphorylation, while viability will decrease due to modification of important membrane proteins and abnormal acrosome reaction that compromises the ability of the sperm to fuse with the oocyte. Since this is a self-propagating cycle, the process also results in amplification, further exacerbating all the associated problems.

Another effect of ROS on spermatozoa is that of DNA damage. This occurs via direct attack on the bases (especially guanine) or the phosphodiester backbones, hence destabilizing the DNA molecule and causing anomalies including, but not limited to, point mutations, polymorphisms, deletions, translocations and even double-stranded breaks. DNA fragmentation will result in abnormal fertilization, reduced implantation and poor embryonic development such that the offspring is likely to have a shorter lifespan and an increased risk of developing cancer. In cases of more severe damage, spermatozoa may undergo apoptosis, resulting in low sperm counts characteristic of idiopathic male factor infertility. However, sperm DNA is less prone to ROS-induced damage than the plasma membrane due to its highly condensed structure, which offers less surface area for attack by ROS.

**LYCOPENE**

**Properties**

Lycopene is one of the many compounds that make up the carotenoid family. Carotenoids are naturally found in fruits and vegetables, and give plants their bright yellow, orange and red colors. They are essential for photosynthesis and provide protection from excessive light. Hence, they are only synthesized by plants and microorganisms, but not humans. Consumption of fruits and vegetables is the only way humans take in carotenoids. Carotenoids are vital components of human diet not only because they are sources of vitamin A, but also because they have antioxidant properties.

Lycopene (C_{36}H_{56}) is a red-pigmented unsaturated linear carotenoid with a molecular weight of 536.85 Da, containing 11 conjugated and two non-conjugated double bonds. It is lipophilic and hence more soluble in organic solvents. The presence of double bonds allows for both cis- and trans-isomeric forms, and conversion between the forms occurs when it is exposed to light, heat or chemical reactions. Although most red-colored fruits and vegetables are lycopene sources, not all red-colored foods contain lycopene. Some common dietary sources of lycopene include tomatoes and processed tomato products, pink grapefruits, watermelons, apricots, guavas, papayas and rosehips, with processed tomato products containing the highest amount of lycopene.

**Metabolism**

Information concerning the in vivo metabolism of lycopene is lacking. Lycopene can be cleaved enzymatically or oxidatively to yield aplycopenals, which could be responsible for some biological activity as well.

**Mechanism of action**

Several mechanisms of action have been proposed as an explanation of how lycopene works to reduce the risk of oxidative stress-mediated/chronic diseases such as cancer, hypertension, cardiovascular disease (CVD), neurodegenerative disease and osteoporosis. These mechanisms of action can be categorized into oxidative and nonoxidative mechanisms, of which the former is more pertinent and more likely to be the mechanism by which lycopene works to alleviate male infertility. An overview of lycopene’s mechanism of action is shown in Figure 1.

**Oxidative mechanisms**

Due to its 11 conjugated double bonds, lycopene contains many electrons which can be donated to free radicals, resulting in their neutralization. In this way, lycopene acts as an antioxidant to trap free radicals and halt the propagative chain reactions, reducing the ROS burden and alleviating oxidative stress, thus preventing oxidative damage to lipids, proteins and DNA.

Lycopene is regarded as one of the most potent singlet oxygen quenchers in the carotenoid family because it is twice as effective as β-carotene and up to 10 times more effective than α-tocopherol. However, it has been reported that a mixture of carotenoids gives a more marked effect than any individual compound, and this synergism was
According to the World Health Organization, infertility is defined as ‘the inability of a sexually active couple (at least three times per month), not using contraception, to achieve pregnancy within one year’.

The inability of a sexually active couple (at least three times per month) to conceive within 12 months of sexual activity is defined as infertility. Male infertility is one of the leading causes of infertility. Male infertility is defined as the inability of a man to achieve pregnancy with a sexually active woman (at least three times per month) who is not using contraception, to achieve pregnancy within one year.

MALE INFERTILITY AND LYCOPENE AS A POSSIBLE TREATMENT STRATEGY

According to the World Health Organization, infertility is defined as ‘the inability of a sexually active couple (at least three times per month), not using contraception, to achieve pregnancy within one year’. About 10%–15% of couples worldwide are affected by this problem, and approximately half of the cases are due to the male factor. The most common cause of male factor infertility is varicocele (approximately 35%), while 25% of the patients are idiopathically infertile. Other causes include urogenital infections, congenital and genetic anomalies, immunologic factors and endocrine disorders.

Although the precise mechanism of action by which lycopene exerts its effect is hitherto unknown, several studies have shown some evidence that lycopene can help to alleviate male infertility. A study that was performed to investigate the effects of lycopene on sperm parameters showed that the tests contained relatively high lycopene concentrations compared to other parts of the body. This suggests that lycopene is likely to play a major physiological role as an antioxidant in the process of spermatogenesis.

A separate study conducted to qualify and quantify the antioxidants present in human seminal plasma further revealed that the concentration of lycopene was significantly lower in infertile men. With less antioxidants in the seminal plasma, there will be more free radicals available to cause oxidative damage, therefore resulting in abnormal spermatozoa that cause infertility. Goyal et al. proved that lycopene concentration in seminal plasma increases with oral supplementation of lycopene. It can therefore be postulated that the intake of lycopene will offer protection from ROS in seminal plasma and decrease oxidative stress, one of the main causes of idiopathic male factor infertility. Hence, there is a strong indication that lycopene, a natural antioxidant, may contribute to the treatment of male infertility.

A few mechanisms have been proposed, but the main one whereby lycopene is thought to aid in the treatment of infertility is via the antioxidation pathway. Antioxidants are usually reducing agents that donate an electron to free radicals in order to quench ROS.

In this way, lycopene reduces the amount of ROS and decreases lipid peroxidation, thus retaining the integrity of the spermatozoal cell membrane. Moreover, since lycopene is lipophilic and frequently found in cell membranes, it is likely to be present in sufficient amounts to protect spermatozoa from damage by oxidative stress.

Other mechanisms that have been suggested include that of indirectly increasing the amount of antioxidant enzymes in the body by activation of the antioxidant system and decreasing the transcription of proinflammatory factors. Figure 2 shows a summary of the proposed mechanism of action of lycopene in restoring male fertility.

Several studies have been performed on humans and animals, both in vivo and in vitro, in an attempt to elucidate the true value of lycopene supplementation as a possible treatment strategy for idiopathic male factor infertility. In this section, we will evaluate these 12 studies, half of which were conducted on human subjects and the other half on animals, by analyzing the outcomes that are related to oxidative stress (Table 1) and sperm parameters (Table 2).

Oxidative stress-related parameters

In order to measure the efficacy of lycopene, studies have measured biomarkers of oxidative stress such as the amount of lipid peroxidation, the DNA fragmentation index which is representative of the extent of DNA damage, and the amount of 8-hydroxy-deoxyguanosine in the urine.

Since malondialdehyde is produced as an end product of lipid peroxidation, by using the thiobarbituric acid reactive substances test, malondialdehyde concentrations can be an indicator of the damage caused to lipid membranes. Two studies have reported a decrease in lipid peroxidation after lycopene was given to the subjects. Sarkar et al. conducted a study where 45 patients and 30 healthy controls were given

Figure 1: General mechanisms of action of lycopene. The proposed mechanisms of action of lycopene (oxidative and nonoxidative) that decreases the risk of oxidative stress-mediated diseases. Lycopene most likely acts via the oxidative mechanism of action to prevent oxidative stress and its detrimental effects on male infertility. ROS: reactive oxygen species.

Most clearly seen when lycopene or lutein was included in the mixture. Besides quenching singlet molecular oxygen, lycopene is also known to act on other free radicals like hydrogen peroxide, nitrogen dioxide and hydroxyl radicals. Furthermore, since lycopene is lipophilic, it tends to accumulate in cell membranes and lipoproteins, thus exerting a more noticeable effect within such components of the cell. In addition to directly neutralizing ROS by acting as a singlet oxygen quencher, hence causing the overall amount of ROS to decrease, lycopene also indirectly decreases oxidative stress by activating other mechanisms that increase antioxidant potential.

Nonoxidative mechanisms

Other nonoxidative mechanisms by which lycopene could exert its effects include the following: aiding in gap junction communication, modulating gene expression, regulating the cell cycle and enhancing the immune system.

It is projected that tumor cells lack gap junction communication and therefore, continue to proliferate without inhibition. By improving communication between cells, lycopene could possibly prevent tumor formation and hence cancer, especially in the prostate, breast and lung. Lycopene also prevents unwanted cell proliferation by disrupting insulin-like growth factor-1 signaling and preventing cell cycle progression.

Lycopene has been shown to have hypcholesterolemic properties because it inhibits hydroxyl-methyl-glutaryl coenzyme A reductase, an important rate-limiting enzyme responsible for cholesterol production. A decrease in cholesterol also contributes to the alleviation of CVD because less atherosclerotic plaque will be present.

The aforementioned nonoxidative mechanisms of action could possibly apply to male infertility too, but as far as the authors are aware, no studies have been conducted in this area. Thus, more research is still required to ascertain the exact mechanism of action by which lycopene exerts its effects in reducing the risk of chronic diseases and that of male infertility.
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Table 1: Summary of studies measuring the effect of lycopene supplementation on oxidative stress-related parameters

| Authors (year)       | Subjects                     | Lycopene source                      | Lycopene dosage (duration) | Oxidative stress-related parameters |
|----------------------|------------------------------|--------------------------------------|----------------------------|-------------------------------------|
|                      |                              |                                      |                            | Lipid peroxidation | DNA damage | Urinary 8-OHdG | Antioxidants | Immunity |
| Goyal et al. (2007)  | 6 healthy human males        | Heated cream of tomato soup           | ≈22.8 mg kg⁻¹ daily (2 weeks) | ↓                     | ↓          | ↑             | ↓           | ↓        |
| Devaraj et al. (2008)| 82 healthy human males       | Synthetic crystalline lycopene (all-trans) | Placebo, 6, 5, 15 or 30 mg daily (8 weeks) | ↓                     | ↓          | ↑             | ↓           | ↓        |
| Mangiagalli et al. (2010) | 25 broiler breeder males     | Supplementation in drinking water     | 0.5 g L⁻¹ or none (multiple samples over 17 weeks) | ↓                     | ↓          | ↑             | ↓           | ↓        |
| Zini et al. (2010)   | 12 healthy human males       | Purchased                             | 0, 2 or 5 mmol L⁻¹ (30 min) | ↓                     | ↓          | ↑             | ↓           | ↓        |
| Aly et al. (2012)    | 24 healthy Wistar rat males  | Purchased                             | 4 mg kg⁻¹ (7 days)          | ↓                     | ↓          | ↑             | ↓           | ↓        |
| Rosato et al. (2012) | 28 healthy hybrid large white turkey males | Preheated Sigma L9879 (purchased) | 0, 0.05 or 0.1 mg mL⁻¹ (48 h for chilled, 2 weeks for cryopreserved) | ↓                     | ↓          | ↑             | ↓           | ↓        |
| Sarkar et al. (2012) | 45 human males with oxidative stress, 30 healthy controls | Lycopene capsule or tomato products (soup, paste, ketchup) | Placebo or 15 mg daily (10 weeks) | ↓                     | ↓          | ↑             | ↓           | ↓        |
| Choi and Seo (2013)  | 40 healthy Mongolian gerbil males | 5% lycopene extracted from tomatoes (purchased) | 4 different experimental diets, one of which is a 0.5 kg⁻¹ lycopene diet (6 weeks) | ↓                     | ↓          | ↑             | ↓           | ↓        |

8-OHdG: 8-hydroxy-2-deoxyguanosine; ↑: increase; ↓: decrease

Table 2: Summary of studies measuring the effects of lycopene supplementation on sperm-related parameters

| Authors (year)       | Subjects                     | Lycopene source                      | Lycopene dosage (duration) | Sperm-related parameters |
|----------------------|------------------------------|--------------------------------------|----------------------------|--------------------------|
|                      |                              |                                      |                            | Concentration (%) | Count (%) | Motility (%) | Morphology (%) | Viability |
| Gupta and Kumar (2002) | 30 infertile human males    | Unnamed                              | 2000 µg twice daily (3 months) | ↑ (66)                  | ↑ (53)     | ↑ (46)       |               |          |
| Mohanty et al. (2001)  | 50 infertile human males    | Unnamed                              | 8 mg daily (until outcome achieved) | ↑ (60)                  | ↑ (70)     | ↑ (54)       | ↑ (38)       |          |
| Zini et al. (2010)    | 12 healthy human males      | Purchased                            | 0, 2 or 5 mmol L⁻¹ (30 min) | ↑                   | ↑ (70)     | ↑ (54)       | ↑ (38)       |          |
| Hekimoglu et al. (2009) | 42 Wistar albino rat males | Purchased                            | 4 mg kg⁻¹ daily (30 days) | ↑                   | ↑           | ↑           | ↑           |          |
| Mangiagalli et al. (2012) | 25 hybrid Martini rabbit males | Supplementation in drinking water | 0, 0.1 or 0.5 g L⁻¹ (8 weeks) | ↑                   | ↑           | ↑           | ↑ quality of sperm after storage |          |
| Mangiagalli et al. (2010) | 25 broiler breeder males   | Supplementation in drinking water    | 0.5 g L⁻¹ or none (multiple samples over 17 weeks) | ↑                   | ↑           | ↑           | ↑           |          |
| Aly et al. (2012)     | 24 healthy Wistar rat males | Purchased                            | 4 mg kg⁻¹ (7 days)          | ↑                   | ↑           | ↑           | ↑           |          |

↑: increase; ↓: decrease; ≈: no significant difference
reduced the damage sustained by DNA when turkey semen samples were refrigerated or cryopreserved. Taken together, these studies imply that lycopene is able to aid in the reduction of DNA damage in spermatozoa, therefore increasing the chances of successful fertilization of the oocyte and better embryo development.\textsuperscript{1,3,3}

Urinary 8-hydroxy-deoxyguanosine, another biomarker of oxidative stress, was only tested for in the study conducted by Devaraj and his coworkers, who evaluated the effects of various doses of lycopene (0, 6.5, 15 or 30 mg daily) on oxidative stress. Urinary 8-hydroxy-deoxyguanosine was quantified with a competitive enzyme-linked immunosorbent assay and showed a 23% decrease after 8 weeks of a daily dose of 30 mg of lycopene;\textsuperscript{22} hence, indicating that DNA damage was significantly reduced with lycopene supplementation.

Other oxidative stress-related parameters that were discussed in the studies we evaluated are the level of antioxidants and overall immunity. Studies conducted by Choi and Seo\textsuperscript{37} and Aly et al.\textsuperscript{38} revealed an increase in antioxidants, such as catalase and glutathione peroxidase, after lycopene supplementation. By measuring bactericidal activity in serum, a study on broiler breeder males also proved that lycopene supplementation improved overall immunity.\textsuperscript{39} Raised immunity and a higher level of antioxidants will help to reduce oxidative stress and improve both the quality and quantity of spermatozoa, therefore improving fertility outcomes.

On the other hand, a study performed by Goyal et al.\textsuperscript{29} indicated that there was no noteworthy increase in the capacity of seminal plasma to scavenge free radicals. In this experiment, six healthy volunteers were instructed to have 400 g of heated cream of tomato soup daily for 2 weeks. Blood and semen samples were analyzed before and after the experiment. The 2,2′azino-bis (3-ethylbenzthiazoline-6-sulfonic acid) assay was used to assess the total radical-trapping antioxidant potential of the semen samples, but no significant change was observed. This could be due to the fact that lycopene is hydrophobic and therefore trapped in the lipid membranes in seminal plasma. Since the experimental conditions were aqueous, lycopene might have been unable to act as it would in vivo. Moreover, the small size of the study could have affected the reliability of the results as well.

As explained earlier, oxidative stress has a negative effect on the male reproductive system by inducing lipid peroxidation and DNA damage, which may eventually lead to apoptosis. Hence, an improvement in the biomarkers of oxidative stress would show a decrease in oxidative stress-related problems, and is likely to aid in the treatment of infertility.

### Sperm parameter-related parameters

In addition to biomarkers of oxidative stress, other studies also observed sperm parameters to make a more direct and specific evaluation of the effectiveness of lycopene on treating male factor infertility. Sperm parameters generally include sperm count and concentration, motility, viability and morphology.

Sperm count was found to increase in four different studies, with Mohanty’s group reporting a significant 70% increase with the administration of 8 mg of lycopene daily.\textsuperscript{9,38,39,42} It can therefore be seen that sperm count will increase with lycopene supplementation.

Of the four studies that measured sperm concentration, only two of them, Gupta and Kumar\textsuperscript{29} and Mohanty et al.\textsuperscript{9} showed an improvement of 66% and 60%, respectively.\textsuperscript{9,29} However, Gupta and Kumar also noted that a baseline sperm concentration of less than 5 × 10^6 mL\textsuperscript{−1} did not show substantial improvement in concentration.\textsuperscript{29} The other two studies did not find any difference in sperm concentration before and after lycopene supplementation. Hence, there is no conclusive evidence for an improvement in sperm concentration of the ejaculate.

Sperm motility was analyzed by five studies, with a majority of them showing that lycopene had a beneficial effect on this parameter. Gupta and Kumar\textsuperscript{29} and Mohanty et al.\textsuperscript{9} conducted studies that showed a marked improvement in patients’ sperm motility of 53% and 54%, respectively, with the former administering 2 mg of lycopene twice a day for 3 months and the latter giving 8 mg of lycopene once daily. Two other studies that were conducted on rats also produced similar positive results.\textsuperscript{36,41} However, Zini et al.\textsuperscript{13} reported that pretreatment with 0, 2 or 5 μmol L\textsuperscript{−1} lycopene for 30 min at 25°C did not preserve sperm motility after oxidative stress was induced. Although different from the other studies, these results are supported in theory, as mentioned earlier, DNA is less susceptible to damage by oxidative stress than membrane lipids due to its highly condensed and compacted structure. Hence, it would be comparatively more difficult to prevent membrane lipid peroxidation with antioxidants. As such, more extensive studies have to be performed in order to determine if lycopene supplementation will improve sperm motility.

Sperm viability was only assessed by a single study, which indicated that lycopene supplementation improved this parameter. In this study, broiler breeders were separated into two groups, and only one group was given lycopene supplementation of 0.5 g l\textsuperscript{−1} in their drinking water. At 42 weeks of age, semen samples were analyzed and it was shown that the group which received lycopene was almost 6% more viable than the control group that did not receive lycopene.\textsuperscript{38} This therefore indicates that lycopene does help in improving sperm viability.

The last sperm parameter analyzed is sperm morphology. Although all three studies that measured this outcome reported improved morphology after lycopene supplementation, the improvement was less than expected and not as significant as that of sperm concentration and motility.\textsuperscript{9,29} Hekimoglu et al.\textsuperscript{38} also showed that lycopene was successful in normalizing the amount of abnormal sperm produced in rat testes.

In general, the human trials analyzed in this review showed that 4–8 mg of daily lycopene supplementation for 3–12 months is sufficient to treat male infertility.\textsuperscript{9,29} This translates to the intake of approximately 150 g of raw tomatoes or 80 g of watermelon daily.\textsuperscript{29} However, more research and clinical trials have to be conducted on humans to determine the most accurate therapeutic dosage.

### CONCLUSION AND FUTURE DIRECTIONS

As demonstrated by the analyses of the various studies above, the only parameters that are conclusively improved with lycopene supplementation are: a decrease in lipid peroxidation and DNA damage, an increase in antioxidants and therefore general immunity, and improved sperm count and viability. These improvements are vital in tackling the problem of oxidative stress, which affects sperm viability, motility and DNA, and therefore causes infertility. The conflicting results of different studies could be due to the lack of standardized protocols and outcome measurements, and further compounded by the relatively small study sizes which could have introduced some bias into the outcomes. Although the results are generally promising, it is evident that more detailed and extensive research has to be done on the efficacy of lycopene, a potent singlet oxygen quencher, in the treatment of idiopathic male factor infertility. Therefore, in order to prove this, a large-scale placebo-controlled clinical trial must be carried out for statistically significant results. Patients should be randomly assigned to receive different daily dosages of lycopene over a specific time period, and the outcomes measured should not only include sperm parameters, but pregnancy rates as well.

### AUTHOR CONTRIBUTIONS

DD conceived of the study, participated in its design, carried out the literature search, participated in the analysis and interpretation of the
REFERENCES

1. Agarwal A, Nallella KP, Allamani SS, Said TM. Role of antioxidants in treatment of male infertility: an overview of the literature. Reprod Biomed Online 2004; 8: 616–27.
2. Rao AR, Rao LG. Carotenoids and human health. Pharmacol Res 2007; 55: 207–16.
3. Sharma RK, Agarwal A. Role of reactive oxygen species in male infertility. Urology 1996; 48: 835–50.
4. Tremelot K. Oxidative stress and male infertility—a clinical perspective. Hum Reprod Update 2008; 14: 243–58.
5. Agarwal A, Hamada A, Esteves SC. Insight into oxidative stress in varicocele–associated male infertility: Part 1. Nat Rev Urol 2012; 9: 678–90.
6. Gomez E, Buckingham DW, Brindle J, Lanzarate F, Irvine DS, et al. Development of an image analysis system to monitor the retention of residual cytoplasm by human spermatozoa: correlation with biochemical markers of the cytoplasmic space, oxidative stress, and sperm function. J Androl 1996; 17: 276–87.
7. Said TM, Agarwal A, Sharma RK, Mascha E, Sikka SC, et al. Human sperm superoxide anion generation and correlation with semen quality in patients with male infertility. Fertil Steril 2004; 82: 871–7.
8. Atken RJ, Irvine DS, Wu FC. Prospective analysis of sperm–ocyt fusion and reactive oxygen species generation as criteria for the diagnosis of infertility. Am J Obstet Gynecol 1991; 164: 542–51.
9. Mohanty N, Kumar S, Jha A, Arora R. Management of idiopathic oligoasthenospermia with lycopene. Indian J Urol 2001; 17: 58–67.
10. Agarwal A, Saleh RA, Bedaiwy MA. Role of reactive oxygen species in the pathophysiology of human reproduction. Fertil Steril 2003; 79: 829–43.
11. Gharagozlo P, Atken RJ. The role of sperm oxidative stress in male infertility and the significance of oral antioxidant therapy. Hum Reprod 2011; 26: 1628–40.
12. Hamada AJ, Montgomery B, Agarwal A. Male infertility: a critical review of pharmacologic management. Expert Opin Pharmacother 2012; 13: 2511–31.
13. Zini A, San Gabriel M, Libman J. Lycopene supplementation in vitro can protect human sperm deoxyribonucleic acid from oxidative damage. Fertil Steril 2010; 94: 1033–6.
14. Stahl W, Sies H. Lycopene: a biologically important carotenoid for humans? Arch Biochem Biophys 1996; 336: 1–9.
15. Rao AV, Ray MR, Rao LG. Lycopene. Adv Food Nutr Res 2006; 51: 99–164.
16. Atasoy N. Biochemistry of lycopene. J Anim Vet Adv 2012; 11: 2605–10.
17. Chauhan K, Sharma S, Agarwal N, Chauhan B. Lycopene of tomato fame: its role in health and disease. JPSR 2011; 10: 99–115.
18. Kaur A, Dhari J, Sharma OP, Gupta GD, Khamb V. Lycopene. JPLT 2011; 3: 1605–22.
19. Rao AV, Agarwal S. Role of lycopene as antioxidant carotenoid in the prevention of chronic diseases: A review. Nutr Res 1999; 19: 305–23.
20. Bohm V, Frohlich K, Bitsch R. Rosehip—a “new” source of lycopene? Mol Aspects Med 2003; 24: 385–9.
21. Cardinaut N, Tsyssander V, Groltier P, Winklhofer-Roob BM, Ribaltova J, et al. Comparison of the postprandial chylomicron carotenoid responses in young and older subjects. Eur J Nutr 2003; 42: 315–23.
22. Wang XD. Lycopene metabolism and its biological significance. Am J Clin Nutr 2012; 96: 1214–22S.
23. Kaplan LA, Lau JM, Stein EA. Carotenoid composition, concentrations, and relationships in various human organs. Clin Physiol Biochem 1990; 8: 1–10.
24. Schmitz HH, Poor CL, Wellman RB, Erdman JW Jr. Concentrations of selected carotenoids and vitamin A in human liver, kidney and lung tissue. J Nutr 1991; 121: 1613–21.
25. Erdman JW Jr. How do nutritional and hormonal status modify the bioavailability, uptake, and distribution of different forms of lycopene? J Nutr 2005; 135: 2046–7S.
26. Palozza P, Catalano A, Simone R, Cittadini A. Lycopene as a guardian of redox signalling. Acta Biochim Pol 2012; 59: 21–5.
27. Debrer D, Lu QY. Overview of mechanisms of action of lycopene. Exp Biol Med (Maywood) 2002; 227: 920–3.
28. Krishnamoorthy G, Selvamalai K, Elumalai P, Venkataraman P, Arunakaran J. Protective role of lycopene on polychlorinated biphenyls (aroclor 1254)-induced adult rat sertoli cell dysfunction by increased oxidative stress and endocrine disruption. Biomed Prev Nutr 2011; 1: 116–25.
29. Gupta NP, Kumar R. Lycopene therapy in idiopathic male infertility—a preliminary report. Int Urol Nephrol 2002; 34: 369–72.
30. Goyal A, Chopra M, Lalveed BA, Birch B, Cooper AJ. The effects of dietary lycopene supplementation on human seminal plasma. BJU Int 2007; 98: 1456–60.
31. Palan P, Naz R. Changes in various antioxidant levels in human seminal plasma related to immunoinfertility. Arch Androl 1996; 36: 139–43.
32. Agarwal A, Sekhon LH. The role of antioxidant therapy in the treatment of male infertility. Hum Fertil (Camb) 2010; 13: 217–25.
33. Rosato MP, Centoducati G, Santacroce MP, Iaffaldano N. Effects of lycopene on in vitro quality and lipid peroxidation in refrigerated and cryopreserved turkey spermatozoa. Br Poult Sci 2012; 53: 545–52.
34. Erdman JW Jr, Ford NA, Lindshield BL. Are the health attributes of lycopene related to its antioxidant function? Arch Biochem Biophys 2009; 483: 229–35.
35. Choi SK, Seo JS. Lycopene supplementation suppresses oxidative stress induced by a high fat diet in gerbils. Nutr Res Pract 2013; 7: 26–33.
36. Oborna I, Malickova K, Fingerova H, Brezinova J, Horka P, et al. A randomized controlled trial of lycopene treatment on soluble receptor for advanced glycation end products in seminal and blood plasma of normospermic men. Am J Reprod Immunol 2011; 66: 179–84.
37. Devaraj S, Mathur S, Bassu A, Aung HH, Vasu VT, et al. A dose-response study on the effects of purified lycopene supplementation on biomarkers of oxidative stress. J Am Coll Nutr 2008; 27: 267–73.
38. Mangiagalli MG, Martino PA, Smajlovic T, Guidobono Cavalcini L, Marelli SP. Effect of lycopene on semen quality, fertility and native immunity of broiler breeder. Br Poult Sci 2010; 51: 152–7.
39. Aly HA, El-Beshbishy HA, Banjar ZM. Mitochondrial dysfunction induced impairment of spermatogenesis in LPS-treated rats: Modulatory role of lycopene. Eur J Pharmacol 2012; 677: 31–8.
40. Sarkar PD, Gupta T, Sahu A. Comparative analysis of lycopene in oxidative stress. J Assoc Physicians India 2012; 60: 17–9.
41. Hekimoglu A, Kurec Z, Aral F, Baba F, Sahna E, et al. Lycopene, an antioxidant carotenoid, attenuates testicular injury caused by ischemia/reperfusion in rats. Tohoku J Exp Med 2009; 218: 141–7.
42. Mangiagalli MG, Cesari V, Ceralini S, Luzi F, Toschi L. Effect of lycopene supplementation on semen quality and reproductive performance in rabbit. World Rabbit Sci 2012; 20: 141–8.