Effect of oxaliplatin plus 5-fluorouracil or capecitabine on circulating and imaging biomarkers in patients with metastatic colorectal cancer: a prospective biomarker study

Reem Mahmood (reem.mahmood@nhs.net)  
Christie NHS Foundation Trust  
https://orcid.org/0000-0003-3683-7972

Danielle Shaw  
Clatterbridge Cancer Centre NHS Foundation Trust

Tine Descamps  
Cancer Research UK

Cong Zhou  
Cancer Research UK

Robert D. Morgan  
Christie NHS Foundation Trust

Saifee Mullamitha  
Christie NHS Foundation Trust

Mark Saunders  
Christie NHS Foundation Trust

Nerissa Mescalado  
Christie NHS Foundation Trust

Alison Backen  
Christie NHS Foundation Trust

Karen Morris  
Cancer Research UK

Ross A. Little  
The University of Manchester

Susan Cheung  
The University of Manchester

Yvonne Watson  
The University of Manchester

James P. B. O'Connor  
The University of Manchester

Alan Jackson  
The University of Manchester

Geoff J. M. Parker  
The University of Manchester

Caroline Dive  
Cancer Research UK

Gordon C. Jayson  
Christie NHS Foundation Trust

Research article

Keywords: colorectal cancer, angiogenesis, biomarkers, bevacizumab

DOI: https://doi.org/10.21203/rs.3.rs-44821/v2

License: © This work is licensed under a Creative Commons Attribution 4.0 International License.  Read Full License
Abstract

Background: Patients with metastatic colorectal cancer are treated with cytotoxic chemotherapy supplemented by molecularly targeted therapies. There is a critical need to define biomarkers that can optimise the use of these therapies to maximise efficacy and avoid unnecessary toxicity. However, it is important to first define the changes in potential biomarkers following cytotoxic chemotherapy alone. This study reports the impact of standard cytotoxic chemotherapy across a range of circulating and imaging biomarkers.

Methods: A single-centre, prospective, biomarker-driven study. Eligible patients included those diagnosed with colorectal cancer with liver metastases that were planned to receive first line oxaliplatin plus 5-fluorouracil or capecitabine. Patients underwent paired blood sampling and magnetic resonance imaging (MRI), and biomarkers were associated with progression-free survival (PFS) and overall survival (OS).

Results: 20 patients were recruited to the study. Data showed that chemotherapy significantly reduced the number of circulating tumour cells as well as the circulating concentrations of Ang1, Ang2, VEGF-A, VEGF-C and VEGF-D from pre-treatment to cycle 2 day 2. The changes in circulating concentrations were not associated with PFS or OS. On average, the MRI perfusion/permeability parameter, $K_{\text{trans}}$, increased in response to cytotoxic chemotherapy from pre-treatment to cycle 2 day 2 and this increase was associated with worse OS (HR 1.099, 95% CI 1.01-1.20, p=0.025).

Conclusions: In patients diagnosed with colorectal cancer with liver metastases, treatment with standard chemotherapy changes cell- and protein-based biomarkers, although these changes are not associated with survival outcomes. In contrast, the imaging biomarker, $K_{\text{trans}}$, offers promise to direct molecularly targeted therapies such as anti-angiogenic agents.

Background

Colorectal cancer is the fourth most common cancer in the United Kingdom, with around 42,000 new cases diagnosed each year (1). For patients presenting with metastatic disease, overall survival remains poor, with only around 10% alive five years after their diagnosis (2). The management of patients with metastatic colorectal cancer has evolved over the past decade with the additional use of molecularly targeted therapies in combination with cytotoxic chemotherapy.

For patients diagnosed with colorectal cancer with liver metastases, first line standard cytotoxic chemotherapy includes: FOLFOX (5-fluorouracil/folinic acid plus oxaliplatin), FOLFIRI (5-fluorouracil/folinic acid plus irinotecan) or CAPOX (capecitabine plus oxaliplatin) (3,4). Targeted therapies against vascular endothelial growth factors (VEGF) e.g. bevacizumab, and epidermal growth factor receptors (EGFR) e.g. cetuximab or panitumumab, are also recommended for first line management of patients with metastatic colorectal cancer in combination with cytotoxic chemotherapy (3,4). Indeed, bevacizumab, cetuximab and panitumumab have been shown in randomised phase III trials to prolong progression-free survival (PFS) (5–13) and overall survival (OS) (5–7,12). Circulating biomarkers, such as CK18, have been shown to be predictive of prognosis and progression in colorectal cancer (14,15). In addition, use of genetics can guide the use of EGFR treatment, with patients that are KRAS/BRAF mutant not benefitting from these drugs (16–18). However, there are no validated circulating or imaging biomarkers to guide the use of VEGF and EGFR therapies, which are expensive and associated with toxicity.

Early phase trials have assessed the effect of traditional cytotoxic chemotherapy in combination with bevacizumab using magnetic resonance imaging (MRI) (19–22) and circulating biomarkers (23–25). However, in order to better understand the data reported for combination therapy, the effects of cytotoxic chemotherapy alone need to be assessed as a control. In this study, MRI and blood-based biomarkers were investigated in patients undergoing standard cytotoxic chemotherapy. Data from this study may improve the understanding of the utility of these biomarkers for future trials incorporating molecularly targeted therapies.

Methods

This was a prospective, single-centre, biomarker-driven study recruiting patients that were treated at the Christie NHS Foundation Trust for colorectal cancer with liver metastases. Ethical approval was obtained from the local ethics committee (see supplementary information). All patients gave written informed consent to participate in the study.

Study Participants

Eligible participants included those with histologically-proven colorectal cancer; liver metastases measuring at least 30mm in the longest axis; 18 years of age or older; a World Health Organization (WHO) performance status of 0 to 2; were planned to commence primary therapy with oxaliplatin plus 5-fluorouracil (5-FU) or capecitabine; white cell blood count ≥4x10⁹/l; platelet count ≥100x10⁹/l; serum total bilirubin concentration ≤1.5×upper limit of normal (ULN); serum alkaline phosphatase concentration ≤5×ULN and; a calculated glomerular filtration rate ≥50ml per minute.

Patients were excluded if MRI was contra-indicated due to standard criteria relating to metal implants or allergy to MRI contrast; use of adjuvant chemotherapy within 12 months prior to study enrolment; a personal medical history including any non-colorectal malignancy within 5 years of study enrolment; concurrent use of other investigational medicinal product or; pregnant or breast-feeding women.

Study Drugs

Patients were treated with either oxaliplatin plus 5-FU (oxaliplatin 85mg/m² of body surface area [BSA] plus folinic acid 350mg and 5-FU 400mg/m² on day 1 followed by 5-FU 2,400mg/m² intravenous infusion [46 hours] every two-week cycle) or oxaliplatin plus capecitabine (oxaliplatin 130mg/m² on day 1 and capecitabine 1,000mg/m² on day 1 to 14 every 3-week cycle) for a maximum of 6 cycles.
Clinical Endpoints

Clinical endpoints included progression-free survival (PFS) and overall survival (OS). Progressive disease was defined as the time interval from the date of study registration to the date of either clinical or radiological progression or death. On imaging, progressive disease was measured using the response evaluation criteria in solid tumours (RECIST) version 1.1 (26). OS was defined as the time interval from the date of study registration to the date of death. All patients were followed up until they reached the PFS efficacy endpoint; no censoring was present in the dataset.

Computed tomography (CT) was performed every 8 weeks as part of standard tumour assessment. As part of standard treatment, plasma carcinoembryonic antigen (CEA) and lactate dehydrogenase (LDH) concentrations were measured at the start of each cycle of chemotherapy. Both can be used to predict prognosis and response to treatment in metastatic colorectal cancer (27,28).

Biomarker Schedule

A detailed description of the methodology used to for the imaging and circulating biomarkers is provided in the supplementary information.

Study time points for dynamic contrast-enhanced MRI (DCE-MRI) and diffusion weighted MRI (DW-MRI) included pre-treatment, cycle 1 day 2, cycle 1 day 8, cycle 2 day 2 of chemotherapy and following 12 weeks of chemotherapy. Pre-treatment, MRI scans were carried out twice, at least 24 hours apart, to determine the repeatability of the imaging biomarkers. Regions of interest (ROIs) within the liver were defined manually by a trained operator, in order to determine whole tumour volume (WTV) from T1- and T2-weighted images as well as the DCE-MRI images. Parameters derived from DCE-MRI included the transfer coefficient (ktrans), volume of extravascular extracellular space (ve) and vascular plasma volume (vp). For DWI-MRI, the apparent diffusion coefficient (ADC) was derived.

Blood samples for circulating tumour cells (CTCs) and a panel of plasma-derived circulating protein biomarkers were collected at the same time points as MRI including Ang2, VEGF-A, VEGF-C, VEGF-D, VEGFR1, VEGFR2, IL6, IL8, Tie2, KGF, PIGF, FGFb, HGF, PDGFbb, SDF1b, E-selectin, M65 and VCAM-1.

Statistical Analysis

The target recruitment for the study was 20 patients. All biomarkers were assessed for normality and transformed when necessary. To identify whether biomarker concentrations changed significantly from pre-treatment to cycle 2 day 2, paired Student’s t-tests were performed. Cycle 2 day 2 of chemotherapy was selected for significance testing in order to determine the early effects of cytotoxic chemotherapy. A correlation network analysis was performed to examine the relationship between multiple biomarkers without the requirement to conduct multiple sequential analyses (23,25). This was done based on Pearson correlations and build from the qgraph package in R.

Cox proportional hazard regression was used for survival analysis, respecting the proportionality and linearity assumptions. Kaplan Meier curves were constructed using dichotomized data (longitudinal increase versus decrease in biomarker concentration), and the median PFS and OS intervals in each group were calculated. Statistical significance was determined using p-values, with a cut off of 0.025 being considered statistically significant in order to reduce the impact of multiple testing. More stringent adjustment for multiple comparisons was not considered due to the limited sample size. Analysis was carried out using R 3.5.0.

Results

Patient Characteristics

Between October 2011 and November 2013, 20 patients were recruited to the study. Patient demographics are shown in Table 1. The mean age of participants was 69 years and the majority were male (85%). During the study, the best radiological response to chemotherapy included: 12 patients (60%) had RECIST complete or partial response (CR/PR), 2 patients (10%) had RECIST stable disease (SD) and 6 patients (30%) had disease progression. Across the entire cohort, the median PFS and OS were 8.7 and 17.3 months, respectively. Twelve patients completed all scanning protocols at chemotherapy cycle 6 and attrition occurred due to falling performance status throughout the trial. The imaging protocols were well tolerated and provided repeatable results.

Table 1: Pre-treatment patient demographics
### Patient Demographic

| Value          |
|----------------|
| Total patients | 20             |
| Sex: number (percentage) | Male Female |
| Age: (years)   |
| WHO performance status: number (percentage) |
| Pre-treatment CEA (µg/L): | Mean Range |
| Pre-treatment LDH (IU/L): | Mean Range |
| Chemotherapy regimen: number (percentage) |

### Pre-treatment Biomarkers

Pre-treatment characteristics including age, WHO performance status and pre-treatment concentrations of CEA and LDH were not associated with PFS or OS. Pre-treatment, CTCs were detectable in all 20 patients. The mean number of CTCs was 4 per 7.5ml of blood.

Evaluation of the association between survival outcomes and pre-treatment concentrations of circulating biomarkers showed that lower concentrations of Ang2 (HR 0.41, 95%CI 0.19-0.86, \(p=0.019\)) and VEGF-A (HR 0.41, 95%CI 0.19-0.87, \(p=0.021\)) were associated with a significantly reduced PFS. No other pre-treatment biomarkers were found to be associated with PFS or OS.

### Biomarkers on Treatment

Data showed that the plasma concentration of most circulating angiogenesis-related biomarkers reduced from pre-treatment to cycle 2 day 2, with significant reductions in Ang1, Ang2, VEGF-A, VEGF-C, and VEGF-D (Table 2). VCAM-1 was the only circulating biomarker to significantly increase (\(p=0.0194\)). The increase of VCAM-1 and decrease of all other circulating biomarkers from pre-treatment to cycle 2 day 2 was not associated with PFS or OS.

The mean number of CTCs significantly reduced from pre-treatment to cycle 2 day 2 (\(p=0.0021\)). A higher number of CTCs at cycle 2 day 2 was associated with significantly worse OS (HR 2.82, 95%CI 1.3-6.1, \(p=0.008\)).

MRI data showed that WTV, enhancing tumour volume (ETV) and T1 decreased significantly from pre-treatment to cycle 2 day 2 (\(p=0.002, p=0.0003\) and \(p=0.0008\), respectively). The ADC significantly increased from pre-treatment to cycle 2 day 2 (\(p=0.017\)) (Table 2). However, none of these parameters were associated with PFS or OS. On average, there was an increase in \(K_{\text{trans}}\) from pre-treatment to cycle 2 day 2. In those patients whose \(K_{\text{trans}}\) increased at cycle 2 day 2, there was a significantly worse OS outcome when compared to those patients whose \(K_{\text{trans}}\) did not increase at cycle 2 day 2 (HR 1.099, 95%CI 1.01-1.20, \(p=0.025\)) (Figure 1).

All patients had an increased CEA concentration from pre-treatment to cycle 2, but this was not associated with PFS nor OS (PFS: \(p=0.521\), OS: \(p=0.638\)). There was no significant difference between the mean increase in CEA concentration between patients with an increased \(K_{\text{trans}}\) and those with a decreased \(K_{\text{trans}}\) (6.79 [95% CI 6.17 - 7.36] versus 6.51 [95% CI 6.21 - 6.81], respectively, \(p=0.43\)).

### Table 2: Significant changes in circulating and imaging biomarkers from pre-treatment to cycle 2 day 2

| Biomarker type | Biomarker name | Mean difference from pre-treatment to C2D2 [95% CI] | \(p\)-value |
|----------------|----------------|-----------------------------------------------|--------------|
| Circulating    | VEGF-C         | -0.932 [-1.333, -0.531]                      | 0.0002       |
|                | FGFb           | -0.866 [-1.262, -0.469]                      | 0.0003       |
|                | VEGF-A         | -0.788 [-1.16, -0.415]                       | 0.0004       |
|                | M65            | -0.57 [-0.845, -0.296]                       | 0.0004       |
|                | Ang1           | -0.722 [-1.084, -0.36]                       | 0.0006       |
|                | VEGF-D         | -0.316 [-0.489, -0.143]                      | 0.0014       |
|                | CTCs           | -1.313 [-2.074, -0.552]                      | 0.0021       |
|                | PDGFbb         | -0.44 [-0.74 - -0.14]                        | 0.0070       |
|                | IL8            | -0.47 [-0.82 - -0.12]                        | 0.0124       |
|                | VCAM-1         | 0.32 [0.07 - 0.58]                           | 0.0149       |
|                | E-selectin     | -0.3 [-0.54 - -0.06]                         | 0.0183       |
| Imaging        | WTV (mm³)      | -0.523 [-0.751, -0.295]                      | 0.0002       |
|                | ETV (mm³)      | -0.56 [-0.813, -0.307]                      | 0.0003       |
|                | T1 (ms)        | -0.144 [-0.217, -0.071]                      | 0.0008       |
|                | ADC (x10⁻³mm²/s) | 0.91 [0.60 - 0.92] | 0.0170 |
|                | \(v_e\)        | 0.04 [0.01 - 0.07]                           | 0.0254       |
|                | \(K_{\text{trans}}\) | 3.38 [0.05 - 6.70] | 0.0469       |

The correlation network analysis showed that, across all patients, the change in the circulating concentrations of angiogenesis-related proteins in response to chemotherapy was similar. The interaction between these proteins is undisturbed by cytotoxic chemotherapy, shown by the close clustering of angiogenic proteins.
biomarkers both at pre-treatment and at cycle 2 day 2 (Figure 2).

See supplementary information for the full data set.

**Discussion**

In this study we have investigated the impact of cytotoxic chemotherapy on circulating proteins and CTCs as surrogate markers of tumour vasculature and cellularity, in parallel with MRI, to document changes in tumour perfusion and tumour volume. This study was limited by the small numbers of participants and so should be regarded as exploratory only.

A key finding of this study was that an increase in $K^{\text{trans}}$ at cycle 2 day 2 was associated with a worse OS. In contrast, trials of anti-angiogenic or vascular disrupting agents show near universal early decrease in $K^{\text{trans}}$ (19). This is considered to reflect reduction in either perfusion or permeability of tumour blood vessels, or a combination of the two processes, rather than a systemic effect. The process by which $K^{\text{trans}}$ changes with a cytotoxic chemotherapy regimen is more complex and indirect. It is possible that is that in these patients, the tumour adapts to chemotherapy by increasing in its vasculature. Another possibility is that these patients have tumours which partly respond to chemotherapy, as shown by the reduction in WTV, but the remaining tumour mass consists of well-perfused chemotherapy-resistant tissue. This process could be explained by a “healing response” to chemotherapy, causing an increase in tumour vascular function and reflected by an increased $K^{\text{trans}}$ value. As the network analysis does not describe changes that are associated with individual biomarkers in isolation, but rather global reductions in angiogenesis biomarkers, the implication is that there is not a dynamic biological response to chemotherapy. Thus, chemotherapy is killing sensitive tumour cell populations leaving behind more resistant clones; characterised here with high $K^{\text{trans}}$. However, since massive cell death of both tumour and non-tumour cells resulting from chemotherapy can lead to the release of damage associated molecular pattern (DAMPS) proteins, it is possible that any resultant systemic inflammatory response could lead to increase in vasculature permeability, detected an increased $K^{\text{trans}}$.

Interestingly, the standard of care marker of treatment response, CEA concentration, increased in all patients from pre-treatment to cycle 2. As mentioned, there was no significant difference between the mean increase in CEA concentration between patients with an increased $K^{\text{trans}}$ and those with a decreased $K^{\text{trans}}$. This suggests that $K^{\text{trans}}$ could be more useful than CEA to predict poorer responses to cytotoxic chemotherapy. This finding may also direct earlier introduction of anti-angiogenic agents, such as bevacizumab.

This study also shows an association between lower pre-treatment circulating concentrations of Ang2 and VEGF-A and worse PFS. There is inconsistency in the value of pre-treatment biomarkers of angiogenesis reported in the literature (29–31). Results differ between studies (Table 3) but are also difficult to compare due to different angiogenesis-related proteins investigated and methods of sampling/analysis used. Hence, our study focussed on the significance of dynamic response to treatment rather than pre-treatment concentrations.

**Table 3: Correlation between pre-treatment circulating angiogenesis-related biomarkers and survival outcomes in studies which recruited patients with metastatic colorectal cancer (22-24)**

| Study Title | No. of patients | Treatment received | Pre-treatment biomarker | Correlation with survival outcomes |
|-------------|-----------------|--------------------|-------------------------|-----------------------------------|
| Prognostic/predictive value of 207 serum factors in colorectal cancer treated with cediranib and/or chemotherapy* (22) | 582 | FOLFOX or CAPOX chemotherapy + cediranib/placebo | VEGF-D | Low pre-treatment concentration correlated with improved PFS and OS regardless of treatment received |
| | | | VEGFR-1 | |
| | | | Tie-2 | |
| | | | Ang2 | No correlation |
| Changes in circulating VEGF levels in relation to clinical response during chemotherapy for metastatic cancer (23) | 90 | Camptothecin | VEGF-165 | Patients with high pre-treatment concentration were more likely to have progressive disease during treatment |
| Phase II Trial of Infusional Fluorouracil, Irinotecan, and Bevacizumab for Metastatic Colorectal Cancer: Efficacy and Circulating Angiogenic Biomarkers Associated With Therapeutic Resistance (24) | 43 | FOLFIRI + bevacizumab | VEGF-2 | No correlation |

**Conclusion**

In conclusion, the response to cytotoxic chemotherapy treatment in patients with colorectal cancer with liver metastases showed a maintained robust relationship between angiogenic biomarkers. In some patients, poor outcome was associated with the early detection of well-perfused tissue in smaller tumours suggesting that chemotherapy was unable to kill the remaining component of a tumour, presumably because of increased clearance of cytotoxic agents. These findings identify a group of patients whose tumour does not respond well to traditional cytotoxic chemotherapy alone and who might benefit from early addition of molecularly targeted therapies.

**Abbreviations**

5-FU - fluorouracil
ADC - apparent diffusion coefficient
Ang – angiopoetin
BSA - body surface area
CAPOX - capecitabine plus oxaliplatin
CEA - carcinoembryonic antigen
CI – confidence interval
CK18 – cytokeratin-18
CT - computed tomography
CTC - circulating tumour cells
CR - complete response
DAMPs - damage associated molecular pattern
DCE-MRI - dynamic contrast-enhanced magnetic resonance imaging
DWI-MRI - diffusion weighted magnetic resonance imaging
EGFR - epidermal growth factor receptors
ETV – enhancing tumour volume
FGFb – basic fibroblastic growth factor
FOLFIRI - fluorouracil/folinic acid plus irinotecan
FOLFOX - fluorouracil/folinic acid plus oxaliplatin
HGF – hepatocyte growth factor
HR – hazard ratio
iAUC – incremental area under the curve
IL8 – interleukin 8
KGF – keratinocyte growth factor
\( K^{\text{trans}} \) - endothelial contrast agent transfer coefficient
LDH - lactate dehydrogenase
MRI - magnetic resonance imaging
NHS – National Health Service
OS - overall survival
PDGFbb – platelet derived growth factor beta
PFS - progression-free survival
PIGF – placental growth factor
PR - partial response
RECIST - response evaluation criteria in solid tumours
ROI - regions of interest
SD - stable disease
SDF1b – stromal cell-derived factor 1
ULN - upper limit of normal

VCAM1 – vascular cell adhesion molecule 1

\(\nu_e\) - fractional extravascular extracellular volume

VEGF – vascular endothelial growth factor

VEGFR – vascular endothelial growth factor receptor

\(\nu_p\) - fractional blood plasma volume

WHO - World Health Organization

WTV - whole tumour volume

Declarations

Ethics approval and consent to participate

This trial was performed with local ethics committee approval from the NRES Committee North West - Liverpool East, under the NHS Health Research Authority (11/NW/0118)(32). Local research and development department approval was obtained and laboratory work was carried out in accordance with the principles of Good Clinical Laboratory Practice (World Health Organization 2009).

Consent for publication

Not applicable

Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request

Competing interests

GJMP is a director and shareholder in Bioxydyn Limited, a company with an interest in advanced MRI biomarkers.

No other competing interests

Funding

Funding received from Cancer Research UK, Manchester NIHR Biomedical Research Centre and AstraZeneca Pharmaceuticals.

Authors' contributions

R.D.Mahmood and D.S. contributed equally as 1st authors to the interpretation of results and write up of the manuscript. G.C.J. oversaw the entire project from conduct of the study, interpretation of data and manuscript write up. T.D. and C.Z. performed statistical analysis of the data and reviewed the finished manuscript. D.S., N.M., M.S. and S.M. conducted the trial and oversaw recruitment, patient management, imaging studies and sample collection. A.B. conducted the ELISA analyses throughout the project. C.D. is director of the circulating biomarkers laboratory and oversaw the conduct and analysis of all ELISAs. A.J., J.P.B.O’C., and G.J.M.P. oversaw the advanced imaging, from acquisition, standardization, data acquisition, analysis, and Q.C. standards. R.A.L., Y.W. and S.C. were responsible for application of DCE-MRI protocols and definition of regions of interest. R.D.Morgan, J.P.B.O, G.J.M.P and A.B all had significant input in substantive revisions of the manuscript and all authors have approved the final manuscript.

Acknowledgements

The authors would like to thank the patients involved in this study and their families. We gratefully acknowledge the support funding received from Cancer Research UK, AstraZeneca Pharmaceuticals and Manchester NIHR Biomedical Research Centre and the support of the host institutions, CRUK Manchester Institute, and the Wolfson Molecular Imaging Centre, Manchester. We would like to thank The Systemic Therapy Research Group at The Christie NHS Foundation Trust for their assistance with preparation of the manuscript. The sponsor for this study was The Christie Hospital NHS Foundation Trust.

References

1. UK CR. Bowel Cancer Statistics [Internet]. 2016. Available from: https://www.cancerresearchuk.org/health-professional/cancer-statistics/statistics-by-cancer-type/bowel-cancer

2. Office for National Statistics. Cancer survival in England - adults diagnosed [Internet]. Available from: https://www.ons.gov.uk/peoplepopulationandcommunity/healthandsocialcare/conditionsanddiseases/datasets/cancersurvivalratescancersurvivalinenglish

3. Van Cutsem E, Cervantes A, Adam R, Sobero A, Van Krieken JH, Aderka D, et al. ESMO consensus guidelines for the management of patients with metastatic colorectal cancer. Ann Oncol [Internet]. 2016 Aug;27(8):1386–422. Available from:
https://linkinghub.elsevier.com/retrieve/pii/S0923753419347544

4. National Comprehensive Cancer Network. NCCN Clinical Practice Guidelines in Oncology: Colon Cancer. 2019;

5. Hurwitz H, Fehrenbacher L, Novotny W, Carwight T, Hainsworth J, Heim W, et al. Bevacizumab plus Irinotecan, Fluorouracil, and Leucovorin for Metastatic Colorectal Cancer. N Engl J Med [Internet]. 2004 Jun 3;350(23):2335–42. Available from: http://www.nejm.org/doi/abs/10.1056/NEJMoa032691

6. Cunningham D, Lang I, Marcuello E, Lorusso V, Ocvirk J, Shin DB, et al. Bevacizumab plus capecitabine versus capecitabine alone in elderly patients with previously untreated metastatic colorectal cancer (AVEX): an open-label, randomised phase 3 trial. Lancet Oncol [Internet]. 2013 Oct;14(11):1077–85. Available from: https://linkinghub.elsevier.com/retrieve/pii/S1470204513701542

7. Kabbavarf F, Irl C, Zurlo A, Hurwitz H. Bevacizumab Improves the Overall and Progression-Free Survival of Patients with Metastatic Colorectal Cancer Treated with 5-Fluorouracil-Based Regimens Irrespective of Baseline Risk. Oncology [Internet]. 2008;75(3–4):215–23. Available from: https://www.karger.com/Article/FullText/163850

8. saltz LB, Clarke S, Diaz-Rubio E, Scheithauer W, Figer A, Wong R, et al. Bevacizumab in Combination With Oxaliplatin-Based Chemotherapy As First-Line Therapy in Metastatic Colorectal Cancer: A Randomized Phase III Study. J Clin Oncol [Internet]. 2008 Apr 20;26(12):2013–9. Available from: http://ascopubs.org/doi/10.1200/JCO.2007.14.9930

9. Cremolini C, Loupakis F, Antoniotti C, Lupi C, Sensi E, Lonardi S, et al. FOLFOXIRI plus bevacizumab versus FOLFIRI plus bevacizumab as first-line treatment of patients with metastatic colorectal cancer: updated overall survival and molecular subgroup analyses of the open-label, phase 3 TRIBE study. Lancet Oncol [Internet]. 2015 Oct;16(13):1306–15. Available from: https://linkinghub.elsevier.com/retrieve/pii/S1470204515001229

10. Van Cutsem E, Nowacki M, Lang I, Cascarini S, Shchepotin I, Maurel J, et al. Randomized phase III study of irinotecan and 5-FU/FA with or without cetuximab in the first-line treatment of patients with metastatic colorectal cancer (mCRC): The CRYSTAL trial. J Clin Oncol [Internet]. 2007 Jun 20;25(18_suppl):4000–4000. Available from: http://ascopubs.org/doi/10.1200/JCO.2007.25.18_suppl.4000

11. Van Cutsem E, Kühne C-H, Hritie E, Zaluski J, Chang Chien C-R, Makhson A, et al. Cetuximab and Chemotherapy as Initial Treatment for Metastatic Colorectal Cancer. N Engl J Med [Internet]. 2009 Apr 2;360(14):1408–17. Available from: http://www.nejm.org/doi/10.1056/NEJMoa0805019

12. Douillard J-Y, Siena S, Cassidy J, Tabemero J, Burkes R, Barugel M, et al. Randomized, Phase III Trial of Panitumumab With Infusional Fluorouracil, Leucovorin, and Oxaliplatin (FOLFOX4) Versus FOLFOX4 Alone As First-Line Treatment in Patients With Previously Untreated Metastatic Colorectal Cancer: The PRIME Study. J Clin Oncol [Internet]. 2010 Nov 1;28(31):4697–705. Available from: http://ascopubs.org/doi/10.1200/JCO.2009.27.4860

13. Tebbutt NC, Wilson K, Gabei VJ, Cummins MM, Zannino D, van Hazel GA, et al. Caepticabine, Bevacizumab, and Mitomycin in First-Line Treatment of Metastatic Colorectal Cancer: Results of the Australasian Gastrointestinal Trials Group Randomized Phase III MAX Study. J Clin Oncol [Internet]. 2010 Jul 1;28(19):3191–8. Available from: http://ascopubs.org/doi/10.1200/JCO.2010.10.7723

14. Greystoke A, Dean E, Saunders MP Cummings J, Hughes A, Ranson M DC and RA. Multi-level evidence that circulating CK18 is a biomarker of tumour burden in colorectal cancer. Br J Cancer. 2012;107(9):1518–24.

15. Scott LC, Evans TRJ, Cassidy J, Harden S, Paul J, Ullah R, et al. Cytokeratin 18 in plasma of patients with gastrointestinal adenocarcinoma as a biomarker of tumour response. Br J Cancer [Internet]. 2009 Aug 14;101(3):410–7. Available from: http://www.nature.com/articles/6605175

16. Douillard J-Y, Oliner KS, Siena S, Tabemero J, Burkes R, Barugel M, et al. Panitumumab–FOLFOX4 Treatment and RAS Mutations in Colorectal Cancer. N Engl J Med [Internet]. 2013 Sep 12;369(11):1023–34. Available from: http://www.nejm.org/doi/10.1056/NEJMoa1305275

17. Ciardiello F, Lenz H-J, Kohne C-H, Heinemann V, Tejpar S, Melezinek I, et al. Treatment outcome according to tumor RAS mutation status in CRSTAL study patients with metastatic colorectal cancer (mCRC) randomized to FOLFIRI with/without cetuximab. J Clin Oncol [Internet]. 2014 May 20;32(15_suppl):3506–3506. Available from: http://ascopubs.org/doi/10.1200/JCO.2014.32.15_suppl.3506

18. Bokemeyer C, Kohne C-H, Ciardiello F, Lenz H-J, Heinemann V, Klinkhardt U, et al. Treatment outcome according to tumor RAS mutation status in OPUS study patients with metastatic colorectal cancer (mCRC) randomized to FOLFIRI with/without cetuximab. J Clin Oncol [Internet]. 2014 May 20;32(15_suppl):3505–3505. Available from: http://ascopubs.org/doi/10.1200/JCO.2014.32.15_suppl.3505

19. O’Connor JPB, Jackson A, Parker GJM, Roberts C, Jayson GC. Dynamic contrast-enhanced MRI in clinical trials of antivascular therapies. Nat Rev Clin Oncol [Internet]. 2012 Mar 14;9(3):167–77. Available from: http://www.nature.com/articles/nrclonc.2012.2

20. Anzidei M, Napoli A, Zaccagna F, Cartoccio G, Saba L, Menichini G, et al. Liver Metastases From Colorectal Cancer Treated With Conventional and Antiangiogenetic Chemotherapy. J Comput Assist Tomogr [Internet]. 2011 Mar;35(2):167–77. Available from: https://insights.ovid.com/crossref?an=00004728-201111000-00006

21. O’Connor JPB, Carano RAD, Clamp AR, Ross J, Ho CCK, Jackson A, et al. Quantifying Antivascular Effects of Monoclonal Antibodies to Vascular Endothelial Growth Factor: Insights from Imaging. Clin Cancer Res [Internet]. 2009 Nov 1;15(3):1674–82. Available from: http://clincancerres.aacrjournals.org/cgi/doi/10.1158/1078-0432.CCR-09-0731

22. Koh D-M, Scurr E, Collins D, Kanber B, Norman A, Leach MO, et al. Predicting Response of Colorectal Hepatic Metastasis: Value of Pretreatment Apparent Diffusion Coefficients. Am J Roentgenol [Internet]. 2007 Apr;188(4):1001–8. Available from: http://www.ajronline.org/doi/10.2214/AJR.06.0601

23. Jayson GC, Zhou C, Backen A, Horsley L, Marti-Marti K, Shaw D, et al. Plasma Tie2 is a tumor vascular response biomarker for VEGF inhibitors in metastatic colorectal cancer. Nat Commun [Internet]. 2018 Dec 7;9(1):4672. Available from: http://www.nature.com/articles/s41467-018-07174-1

24. Zhou C, Clamp A, Backen A, Berzuini C, Renahan A, Banks RE, et al. Systematic analysis of circulating soluble angiogenesis-associated proteins in ICON7 identifies Tie2 as a biomarker of vascular progression on bevacizumab. Br J Cancer [Internet]. 2016 Jul 28;115(2):228–35. Available from: http://www.nature.com/articles/bjc2016194

25. Backen A, Renehan AG, Clamp AR, Berzuini C, Zhou C, Oza A, et al. The Combination of Circulating Ang1 and Tie2 Levels Predicts Progression-Free Survival Advantage in Bevacizumab-Treated Patients with Ovarian Cancer. Clin Cancer Res [Internet]. 2014 Sep 1;20(17):4549–58. Available from:
26. Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, et al. New response evaluation criteria in solid tumours: Revised RECIST guideline (version 1.1). Eur J Cancer [Internet]. 2009 Jan;45(2):228–47. Available from: https://linkinghub.elsevier.com/retrieve/pii/S0959804908008733

27. Li G, Wang Z, Xu J, Wu H, Cai S, He Y. The prognostic value of lactate dehydrogenase levels in colorectal cancer: a meta-analysis. BMC Cancer [Internet]. 2016 Dec 25;16(1):249. Available from: http://bmccancer.biomedcentral.com/articles/10.1186/s12885-016-2276-3

28. Duffy M., van Dalen A, Haglund C, Hansson L, Klapdor R, Lamerz R, et al. Clinical utility of biochemical markers in colorectal cancer. Eur J Cancer [Internet]. 2003 Apr;39(6):718–27. Available from: https://linkinghub.elsevier.com/retrieve/pii/S0959804902008110

29. Spencer SKM, Pommer AJC, Morgan SR, Barry ST, Robertson JD, Hoff PM, et al. Prognostic/predictive value of 207 serum factors in colorectal cancer treated with cediranib and/or chemotherapy. Br J Cancer [Internet]. 2013 Nov 22;109(11):2765–73. Available from: http://www.nature.com/articles/bjc2013649

30. Lissoni P, Rovelli F, Malugani F, Brivio F, Fumagalli L, Gardani GS. Changes in circulating VEGF levels in relation to clinical response during chemotherapy for metastatic cancer. Int J Biol Markers [Internet]. 2003;18(2):152–5. Available from: http://www.biological-markers.com/article(changes-in-circulating-vegf-levels-in-relation-to-clinical-response-during-chemotherapy-for-metastatic-cancer-art005450

31. Kopetz S, Hoff PM, Morris JS, Wolff RA, Eng C, Glover KY, et al. Phase II Trial of Infusional Fluorouracil, Irinotecan, and Bevacizumab for Metastatic Colorectal Cancer: Efficacy and Circulating Angiogenic Biomarkers Associated With Therapeutic Resistance. J Clin Oncol [Internet]. 2010 Jan 20;28(3):453–9. Available from: http://ascopubs.org/doi/10.1200/JCO.2009.24.8252

32. UK Legislation. The Medicines for Human Use (Clinical Trials) Regulations 2004 [Internet]. 2004 [cited 2020 May 12]. Available from: http://www.legislation.gov.uk/uksi/2004/1031/pdfs/uksi_20041031_en.pdf