Use of a Silver-Impregnated Vascular Graft: Single-Center Experience

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Abstract: Introduction: Vascular graft infection is a life threatening situation with significant morbidity and mortality. Bacterial graft infection can lead to false aneurysms, bleeding and sepsis. There are a lot of risky situations where grafts can become infected. It is therefore highly desirable to have a vascular graft that is resistant to infection. In this retrospective clinical study, a silver-impregnated vascular graft was evaluated in various indications. Methods: Our study included a total of 71 patients who received a silver-impregnated vascular graft during the period from 2013 to 2018. Patients had an aortoiliac localization of vascular graft in 61 cases (86%), and a peripheral localization on the lower limbs in 10 cases (14%). Indications for the use of these special vascular grafts were trophic lesions or gangrene in the lower limbs in 24 cases (34%), suspicious mycotic abdominal aortic aneurysm (mAAA) in 4 cases (5.5%), salmonella aortitis or aneurysms in 4 cases (5.5%), infection of the previous vascular graft in 11 cases (15.5%), other infections in 12 cases (17%), AAA rupture in 10 cases (14%) and other reasons (pre-transplant condition, multiple trauma, graft-enteric fistula) in 6 cases (8.5%). Thirty-day mortality, morbidity, the need for reintervention and amputation, primary and secondary graft patency, and finally the presence of a proven vascular graft infection were evaluated. Results: The 30-day mortality was 19.7%, and morbidity was 42.2%. The primary patency of the vascular graft was 91.5%. Reoperation was necessary in 10 cases (14%) and amputation was necessary in 10 cases (14%). The median length of hospital stay was 13 days and the mean follow-up period was 48 ± 9 months. During the follow-up period, six patients (8.5%) died from reasons unrelated to surgery or without any relation to the vascular graft. Secondary patency after one year was 88%. Infection of the silver graft was observed in three patients (4.2%). Conclusions: Based on our results, the silver graft is a very suitable alternative for solving infectious, or potentially infectious, situations in vascular surgery. In particular, in urgent or acute cases, a silver graft is often the only option.

Keywords: antibiotics; graft patency; silver-impregnated vascular graft; vascular graft infection

1. Introduction

Despite ever-improving vascular grafts, better antibiotic prophylaxis and better and earlier specific diagnostic methods, prosthetic vascular graft infection remains one of the worst surgery procedure-related complications. A way to prevent and possibly address these serious situations is still being discussed. The use of venous autografts is obviously limited, and the use of allografts carries its own specific problems and complications [1–6].

Creating an artificial vascular graft that is resistant to infection is one of the goals of current research in the field of vascular surgery. Infection in an artificial vascular graft has significant mortality and morbidity [7]. Bacterial graft infection can lead to false aneurysms, bleeding, chronic infected wounds and sepsis. These are situations that directly threaten the patient’s life, and infection of the vascular reconstruction in the lower limbs is often associated with the risk of limb loss. Despite obvious antibiotic prophylaxis, we observe this complication in 1.5–6% of patients [3,7,8]. The most common infectious species are...
Staphylococcus aureus, methicillin-resistant Staphylococcus aureus, Psedomonas aeruginosa, Salmonella enteritidis and others. Infections caused by fungi are less common [2,9]. According to most authors, the permanent and definitive treatment of an artificial vascular graft infection is impossible [4,10,11]. Therefore, graft explantation and replacement is almost always necessary. However, this is associated with the problem of replacing an infected graft. In the aortoiliac region, venous grafts cannot be used, with a few exceptions [4,5]. Operative techniques using the superficial femoral vein are still uncommon [12]. We are forced to continue using an artificial vascular graft or arterial allograft, which is often not available in urgent or acute cases [13]. It is therefore highly desirable to have a vascular graft that is as resistant to infection as possible. Implantation of these specific grafts has more indications than just the replacement of infected vascular prostheses. The most common are vascular reconstructions in lower limbs with trophic lesions or gangrene, acute aortitis, mycotic aneurysms (Figure 1), aortoduodenal fistula, and others [7,14]. Antibiotic-impregnated vascular grafts were introduced many years ago; this option is still commonly used and we have a lot of literature data with relatively satisfactory results [15–19]. The problem is the relatively short period in which antibiotics are released from the graft. Grafts impregnated with silver salts have been introduced relatively recently. The silver salts, firmly bound in polyester vascular grafts, act directly on the phospholipid layer of the cytoplasmic membrane of bacteria and also directly attack bacterial DNA and prevent its replication. Therefore, this is a different mechanism compared to the release of antibiotics from antibiotic-bound grafts [20]. The microbicidal function of silver should therefore significantly increase resistance to infection [15,21], but recently published studies are also controversial [22,23]. The aim of this work is to present the results of an observational retrospective clinical study conducted in a single center using silver-impregnated vascular grafts in various indications. As for us, the novelty of the study is the use of silver-impregnated grafts in various risky indications with very good results. We did not avoid the use of silver grafts, even in severe cases, such as a confirmed mycotic aneurysm, infection of a regular vascular graft with methicillin-resistant Staphylococcus aureus or major intra-abdominal trauma requiring vascular reconstruction. The authors also discuss the question of necessary long-lasting antibiotic treatments in these specific situations.

Figure 1. Types of reconstructions included in the study.

2. Methods

This is a monocentric retrospective clinical study that retrospectively analyzed the data of patients in whom a silver-impregnated vascular graft was implanted for various indications.

Our study included a total of 71 patients, 53 men (75%) and 18 women (25%), who received a silver-impregnated vascular graft during the period from 2013 to 2018 at the Vascular Surgery Dpt., University Hospital, in Pilsen.

The mean age of the patients was 65.7 years. Patients had an aortoiliac localization of the vascular graft (aortobifemoral bypass, aortoarotic, aortofemoral, iliacofemoral bypass) in 61 cases (86%), and a peripheral localization in the lower limbs (femoropopliteal, femorofemoral or femorocrural bypass) in 10 cases (14%) (Figure 1). We have summarized crucial
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Data from the cohort in Table 1. Indications for the use of these special vascular grafts were trophic lesions or gangrene in the lower limbs in 24 cases (34%), suspicious mycotic abdominal aortic aneurysm (mAAA) in 4 cases (5.5%) (Figure 2), salmonella aortitis or aneurysms in 4 cases (5.5%), infection of the previous vascular graft in 11 cases (15.5%), other infections in 12 cases (17%), AAA rupture in 10 cases (14%) and other reasons (pre-transplant condition, multiple trauma, graft-enteric fistula) in 6 cases (8.5%) (Figure 3). All surgical procedures were performed under general anesthesia; antibiotic therapy was used in all cases. In 35 cases (49.5%), only prophylactic wide spectrum antibiotics were used, which included two doses perioperatively. In the remaining 36 cases (50.5%), antibiotic therapy continued for longer, at least for one week, due to severe infection in various localizations. Cefazolin was used as a prophylactic antibiotic, while in other cases the antibiotics were used specifically based on previous culture results (cefuroxime, vancomycin, ciprofloxacin and combinations of them).

Table 1. Cohort of patients in the study.

| Age (y) | Male | Female | Total |
|---------|------|--------|-------|
| Age 40–60 | 11   | 6    | 17   |
| Age 60–70 | 25   | 10   | 35   |
| Age 70–80 | 14   | 2    | 16   |
| Age above 80 | 3 | 0   | 3    |
| Comorbidity | | | |
| DM | 16 | 3 | 19 |
| CHHF | 18 | 5 | 23 |
| CHRF | 13 | 1 | 14 |
| AH | 39 | 11 | 50 |
| ISS | 5 | 0 | 5 |
| ATB | | | |
| Cefazolin (prophylaxis) | 25 | 10 | 35 |
| Cefuroxime | 5 | 2 | 7 |
| Vancomycin | 10 | 3 | 13 |
| Ciprofloxacin | 2 | 1 | 3 |
| Combination | 11 | 2 | 13 |
| Type of Reconstruction | | | |
| Aortoiliac region | 49 | 12 | 61 |
| Peripheral region | 4 | 6 | 10 |

AH—arterial hypertension, DM—diabetes mellitus, CHRF—chronic renal failure, ISS—immunosuppression situation, CHHF—chronic heart failure.

The reasons for using silver grafts are described in Figure 3.

All data were obtained from the University Hospital clinical database. Thirty-day mortality, morbidity, the need for reintervention and amputation, primary and secondary graft patency, and finally the presence of a proven vascular graft infection were evaluated.
Figure 2. Silver-impregnated graft implanted after mycotic AAA resection.

Figure 3. Indications for using silver-impregnated vascular grafts.

3. Results

30-day mortality in our cohort was 19.7% (14 patients), the most common causes of death being hemorrhagic shock, sepsis and multiorgan failure. Our mortality rate was fully comparable with literature data on these major vascular emergencies [4,5,24]. Morbidity was 42.2% and most commonly included sepsis, multiorgan failure, respiratory failure, and
wound infection. This data corresponded to similar cohorts of patients (rupture of AAA, mycotic AAA, vascular graft infection) where silver grafts had not been used [25,26]. In the same time period, 186 regular non-silver grafts were implanted in these indications. A summary of the morbidity/mortality results in the silver graft group is presented in Table 2.

Table 2. Morbidity/mortality data in our cohort.

|                         | Male | Female | Total (%) |
|-------------------------|------|--------|-----------|
| 30-Day Mortality        | 11   | 3      | 14 (19.7%)|
| Hemorrhagic shock       | 3    | 0      | 3         |
| Multiorgan failure      | 6    | 2      | 8         |
| Sepsis                  | 1    | 1      | 2         |
| Morbidity               | 23   | 7      | 30 (42.2%)|
| Respiratory failure     | 11   | 6      | 17        |
| Multiorgan failure      | 10   | 3      | 13        |
| Wound infection         | 15   | 3      | 18        |
| Sepsis                  | 6    | 0      | 10        |

The primary patency rate of vascular grafts at 1 year was 91.5%. Early graft closure (within 30 days) occurred in six cases. Predictors of primary patency loss were the presence of critical limb ischemia, diabetes mellitus and the necessity of below-the-knee bypasses. There was no difference in patency between the silver graft and regular graft cohorts. Reoperation was necessary in 10 cases (14%); the reason was mainly graft occlusion (8.5%), bleeding (10%), wound infection (8.5%) or a combination of the above. Amputation was necessary in 10 cases (14%). The median length of hospital stay was 13 days and the mean follow-up period was 48 ± 9 months. During the follow-up period, six patients (8.5%) died from reasons unrelated to surgery or without any relation to the vascular graft. Secondary patency after 1 year was 88%. All of this data corresponded with data for common cohorts, where a normal graft was used (our own unpublished data).

Proven infection of the silver graft was only observed in three patients (4.2%) during the follow-up period (Table 3). The infections were confirmed by positron emission tomography with computed tomography (PET/CT)—high 18F-fluoro-2-deoxy-D-glucose (FDG) uptake was observed in vascular grafts and also by positive cultures. **Staphylococcus aureus** was cultured in two cases (haemoculture) and **Salmonella enteritidis** in one case as a monoinfection not only from blood cultures but also from the silver graft. Thus, there was no doubt that these agents were the cause of infection. In one case, the vascular graft was explanted and replaced with an aortic allograft. Figure 4 shows the PET/CT of the infected silver graft before explantation, where the cultivated bacteria were **Salmonella enteritidis**. The probable source of infection was diarrhea 2 months before clinical signs of sepsis occurred. The other two cases of peripheral silver graft infection were treated conservatively with very good results (vancomycin 0.5 mg/day intravenously for 10 days, then ciprofloxacin 800 mg/day orally for a minimum of 6 weeks). Regular CT and PET/CT examinations were performed to follow the treatment success. All three patients with silver graft infections recovered very well without any severe complications. Patients with an aortic allograft had no clinical or PET/CT signs of re-infection, and both patients with left infected peripheral vascular reconstructions had no clinical signs of infection, although the PET/CT images indicated persistent high uptake values of 18F-fluoro-2-deoxy-glucose (18F-FDG). They are, at present, without antibiotic treatment.
4. Discussion

Infection of any artificial materials implanted into the human body is always a very serious problem, whether it is an artificial joint, heart valve, hernia mesh or artificial vascular graft. Especially in vascular surgery, infection of an artificial vascular graft is a nightmare for every vascular surgeon. These situations are associated with significant morbidity and mortality [27]. The use of resistant vascular grafts may be beneficial as it may reduce the need for reoperation caused by vascular graft infection and can improve the outcome of surgeries in potentially infectious areas (trophic lesions of the lower limbs, gangrene, aortitis, aortoduodenal fistula, ongoing bacteremia, etc.).

The use of cryopreserved or fresh allografts is one of the alternatives for managing these catastrophic situations. According to a number of authors, the results are very good and comparable to the use of silver grafts based on the results of a number of studies [28]. The problem is an obvious lack of these grafts in acute and urgent cases. Antibiotic-impregnated grafts have not met expectations in the past, and according to recent data, are inferior to silver-impregnated grafts [1,24]. In in vitro studies, different types of grafts were each contaminated separately with various micro-organisms. For all micro-organisms tested (authors have tested Staphylococcus epidermidis, methicillin-resistant Staphylococcus aureus (MRSA), Escherichia coli, and Candida albicans), the silver graft demonstrated a more sustainable and efficient 7-day antimicrobial activity than the rifampicin-soaked graft [15,29]. This is probably related to the rapid release of antibiotics with early peak values in the blood and a rapid decline of blood levels. There are many in vitro studies showing the bactericidal effect of silver [29–32].
Our own clinical experiences with antibiotic grafts are also very poor. We only implanted 12 rifampicin-soaked vascular grafts in this time period, but the results were unsatisfactory; six patients showed signs of reinfection within the 1-year follow up.

On the other hand, a number of data are available that demonstrate the very good results of silver grafts, the good long-term patency of these vascular grafts and especially a low percentage of infection [1,7,22]. These data are often based on large multicentric trials or meta-analyses with similar inclusion criteria, so their results are very comparable and their conclusions are credible.

Our retrospective study focused on the long-term results of silver graft use in various indications. The primary aim was to obtain information about the long-term patency of these vascular grafts, other complications associated with the grafts and especially about their infection. Our values of primary and secondary patency (91% and 88%, respectively) are fully in line with the use of common vascular grafts in non-risk situations. We did not observe any allergic reactions to the silver grafts in our cohort. Infection in the graft occurred only in three patients (4.2%), despite very risky infectious situations. In one of them, we had to explant the graft and use a fresh aortic allograft, while in the remaining two patients conservative therapy was used (long-term antibiotic treatment).

Our very good experience with silver grafts makes us consider the possible expansion of indications for the use of these highly-resistant vascular grafts; for example, in immunosuppressed patients or in combination procedures, where the risk of infection is multiplied (AAA resection plus another procedure e.g., cholecystectomy, colectomy, liver resection, etc.). The novelty of the study is the use of silver-impregnated grafts in various risky indications with very good results.

The immediate availability of silver grafts in their various variants is of particular importance, especially in the field of urgent and acute vascular surgery. It is known that the floridity of the inflammatory process in the vascular graft area can fluctuate, and thus in some cases, the situation can be managed with long-term antibiotic therapy. However, there are situations where a sudden onset of an attenuated infection does not give time to plan an elective solution and obtain an allograft for urgent surgery, especially in the case of sudden bleeding, e.g., from a rupture of a pseudoaneurysm in one of the anastomoses. A similar situation can occur in the case of the development of prosthetic-enteric fistula with associated, mostly massive, bleeding into the affected part of the gastrointestinal tract. Equally urgent are cases of AAA rupture, where we perioperatively find the cause of the aortitis, most often of salmonella origins. Similarly, a previously unknown infection in a stent-graft implanted in AAA can only manifest by its rupture, especially in non-compliant patients who have not been monitored on a regular basis. Infection in stent-grafts is often mentioned as a severe EVAR complication [33]. To avoid it, strict aseptic conditions during stent-graft delivery are necessary.

However, silver grafts are not only used in aortoiliac surgery, but also in peripheral blood vessels. It is well-known that many patients do not have a venous graft available for vascular reconstruction, whether that is due to gracility, varicosity or saphenous vein trunk stripping. Here again, the use of silver grafts is available as a suitable alternative, especially in patients with trophic lesions in the lower limbs, whether of ischemic, diabetic, venous or combined etiology.

The use of silver grafts is contraindicated in known allergies to silver and palladium, which is very rare in the population. We have not observed this situation. The disadvantage is the slightly higher price of silver grafts compared to a conventional vascular grafts, but when compared with the costs associated with obtaining an allograft or the treatment of infectious complications of common vascular grafts, the price of silver grafts is still relatively lower [28]. More recently, triclosan (5-chloro-2-(2,4-dichlorophenoxy)phenol) has been combined together with silver in a new generation of grafts with promising results [34].
5. Conclusions

Based on our results, silver grafts are a very suitable alternative for solving infectious or potentially infectious situations in vascular surgery. High primary and secondary patency, low graft-infection rate and optimal costs are the crucial factors that allow us to draw such conclusions. Especially in urgent or acute cases, silver grafts are often the only option. This is, in our opinion, the biggest advantage of using silver-impregnated grafts. Even if it was just a bridge to buy time for a possible final solution, it is worth having this method in our portfolio. Furthermore, the physical condition of the patient must be considered in the decision of which strategy will be selected. Our study shows very reasonable and acceptable results in situations where infection would certainly occur using a common vascular graft. A large randomized trial in this field would certainly be desirable; the problem is if this study is feasible with the heterogeneity of the group of patients in whom silver graft use is indicated. Therefore, a number of biases can logically be expected. Based on our own experience, and also on the basis of a number of literary data, the use of silver-impregnated grafts is beneficial. Future results can be even better thanks to a new generation of vascular grafts where silver salts are combined with other bactericidal substances.

Author Contributions: J.M. and V.T., supervision, manuscript writing; V.O., K.H., B.C. and J.B., data validation. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Charles University Research Fund Progres Q39 and by project No. NU20-02-00368 awarded by Czech Health Research Council and by the Ministry of Health of the Czech Republic.

Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki, and approved by the Institutional Review Board of Faculty of Medicine in Pilsen 1.3.2021.

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: All data are available in University Hospital medical records.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Batt, M.; Feugier, P.; Camou, F.; Coffy, A.; Senneville, E.; Caillon, J.; Calvet, B.; Chidiac, C.; Laurent, F.; Revest, M.; et al. A Meta-Analysis of Outcomes after In Situ Reconstructions for Aortic Graft Infection. *Angiology* 2018, 69, 370–379. [CrossRef] [PubMed]
2. Gharamti, A.; Kanafani, Z.A. Vascular Graft Infections An update. *Infect. Dis. Clin. N. Am.* 2018, 32, 789–809. [CrossRef] [PubMed]
3. Seeger, J.M. Management of patients with prosthetic vascular graft infection. *Am. Surg.* 2000, 66, 166–177. [PubMed]
4. Blaha, L.; Bulejčik, J.; Riha, D. Vascular graft infection in the aortoiliac territory—Our view in the light of European Society for Vascular Surgery Guidelines—The retrospective observation study. *Perspect. Surg.* 2021, 100, 330–338. [CrossRef]
5. Sebesta, P.; Šádler, P.; Sedivý, P.; Zdráhal, P.; El Samman, K.; Jindrák, V.; Syrúček, M. Radikální operace infekce cévní protézy v aortofemorální pozici s pomocí cerstvého tepenného allograftu: Nase střednědobé zkoušení [Radical operation of infected aortofemoral prosthesis using fresh arterial allograft: Our mid-term experience]. *Rozhl. V Chir.* 2011, 90, 4–13. (In Czech)
6. Shiraev, T.P.; de Boer, M.; Joseph, S.; Loa, J.; Qasabian, R. Aortic graft explants—A single institution analysis of incidence and outcomes. *Vascular* 2022. [CrossRef]
7. Rico, J.-B. InterGard silver bifurcated graft: Features and results of a multicenter clinical study. *J. Vasc. Surg.* 2006, 44, 339–346. [CrossRef]
8. El-Gamel, A. Proximal Aortic Graft Infection: Time for Treatment in an Aortic Centre? *Heart Lung Circ.* 2022, 31, 4–6. [CrossRef]
9. Puges, M.; Bérand, X.; Caradu, C.; Accoceberry, I.; Gabriel, F.; Cazanave, C. Fungal Vascular Graft and Endograft Infections Are Frequently Associated with Aorto-Enteric Fistulas. *Eur. J. Vasc. Endovasc. Surg.* 2021, 62, 819–820. [CrossRef]
10. Rufa, M.; Ursulescu, A.; Stan, A.; Göbel, N.; Albert, M.; Franke, U.F.W. Cryopreserved aortic homograft, lifeline treatment for infected vascular prosthesis with mediastinal abscess in a re-redo case. *J. Surg. Case Rep.* 2022, 2022, rjab44. [CrossRef]
11. Kouijzer, I.J.; Van der Jagt, M.F.; Bleeker-Rovers, C.P.; Dirven, M.; de Mast, Q.; Poyck, P.P. Outcome in Patients after Autologous Femoral Vein Reconstruction for Primary Aortic Infection and Aortic Graft Infection: A Case Series. *Ann. Vasc. Surg.* 2021, 18. [CrossRef]
12. Neufang, A.; Savvidis, S. Operative technique and morbidity of superficial femoral vein harvest. Gefäßchirurgie 2016, 21, 45–54. [CrossRef]

13. Spacek, M.; Měřička, P.; Janoušek, L.; Štádle, P.; Adamec, M.; Vlachovský, R.; Guňka, I.; Navrátil, P.; Thieme, F.; Špunda, R.; et al. Current vascular allograft procurement, cryopreservation and transplantation techniques in the Czech Republic. Adv. Clin. Exp. Med. 2019, 28, 529–534. [CrossRef]

14. Larena-Avellaned, A.; Russmann, S.; Fein, M.; Debus, E.S. Use of the silver-acetate–coated graft in arterial occlusive disease: A retrospective, comparative study. J. Vasc. Surg. 2009, 50, 790–798. [CrossRef]

15. Berard, X.; Puges, A.; Cazanave, C.; Stecken, L.; Bordenave, L.; Pereyre, S.; M’Zali, F. In vitro Evidence of Improved Antimicrobial Efficacy of Silver and Triclosan Containing Vascular Grafts Compared with Rifampicin Soaked Grafts. Eur. J. Vasc. Endovasc. Surg. 2019, 57, 424–432. [CrossRef]

16. Berger, P.; Van Herwaarden, J.A.; Harkissen, S.; De Vries, J.P.; Ekkelkamp, M.; Moll, F.L. Surgical treatment of infected aortic grafts. J. Cardiovasc. Surg. 2012, 53, 719–734.

17. McGuinness, B.; Ali, K.P.; Phillips, S.; Stacey, M. A Scoping Review on the Use of Antibiotic-Impregnated Beads and Applications to Vascular Surgery. Vasc. Endovasc. Surg. 2019, 54, 147–161. [CrossRef]

18. Clemens, M.S.; Stull, M.C.; Hata, K.W.; Heafner, T.A.; Watson, J.D.B.; Arthurs, Z.M.; Propper, B.W. Antimicrobial-bonded graft patency in the setting of a polymicrobial infection in swine (Sus scrofa). J. Vasc. Surg. 2017, 66, 1210–1216. [CrossRef]

19. Bisdas, T.; Beckmann, E.; Marsch, G.; Burgwitz, K.; Wilhelmi, M.; Kuehn, C.; Haverich, A.; Teebken, O. Prevention of Vascular Graft Infections with Antibiotic Impregnation Prior to Implantation: In Vitro Comparison between Daptomycin, Rifampin and Nebacitin. Eur. J. Vasc. Endovasc. Surg. 2012, 43, 448–456. [CrossRef]

20. Mufty, H.; Eynde, J.V.D.; Meuris, B.; Metsemakers, W.-J.; Van Wijngaerden, E.; Vandendriessche, T.; Steenackers, H.P.; Fourneau, I. Pre-clinical in vivo Models of Vascular Graft Coating in the Prevention of Vascular Graft Infection: A Systematic Review. Eur. J. Vasc. Endovasc. Surg. 2021, 62, 99–118. [CrossRef]

21. Honig, S.; Seeger, P.; Rohde, H.; Kölbel, T.; Debus, E.S.; Diener, H. Efficacy of antiseptic impregnation of aortic endografts with rifampicin compared to silver against in vitro contamination with four bacteria that frequently cause vascular graft infections. JVS Vasc. 2020, 1, 181–189. [CrossRef]

22. Szeberin, Z.; Münch, Z.; Fehevrári, M.; Biró, G.; Entz, L.; Ácsády, G. Ezüst-acetáttal bevont Dacron grafttal végzett rekonstrukció értékelése [Mid-term results of silver-coated Dacron graft implantation into aortic and lower extremity revascularization]. Magy. Sebészeti 2010, 63, 369–373. [CrossRef]

23. Mufty, H.; Eynde, J.V.D.; Steenackers, H.P.; Metsemakers, W.-J.; Van Wijngaerden, E.; Vandendriessche, T.; Steenackers, H.P.; Fourneau, I. A systematic review of preclinical data regarding commercial silver-coated vascular grafts. J. Vasc. Surg. 2021, 74, 1386–1393.e1. [CrossRef]

24. Hassen-Khodja, R.; Sadaghianloo, N.; Jean-Baptiste, É. Matériaux de reconstruction aortique résistants à la contamination bactérienne [Aortic reconstruction with graft materials resistant to bacterial infections]. Bull. l’academie Natl. Med. 2013, 197, 979–999. discussion 991. (In French)

25. Kessler, V.; Klopjf, J.; Eilenberg, W.; Neumayer, C.; Brostjan, C. AAA Revisited: A Comprehensive Review of Risk Factors, Management, and Hallmarks of Pathogenesis. Biomedicines 2022, 10, 9. [CrossRef]

26. Golemovic, M.; Skific, M.; Haluzan, D.; Pavic, P.; Cepulic, B.G. Ten-year experience with cryopreserved vascular allografts in the Croatian Cardiovascular Tissue Bank. Cell Tissue Bank. 2022, 21–18. [CrossRef]

27. Wilson, W.R.; Bower, T.C.; Creager, M.A.; Amin-Hanjani, S.; O’Gara, P.T.; Lockhart, P.B.; Darouiche, R.O.; Ramlawi, B.; Derdeyn, C.; et al. Management, and Hallmarks of Pathogenesis. Biomedicines 2022, 10, 9. [CrossRef]

28. Bisdas, T.; Wilhelmi, M.; Haverich, A.; Teebken, O.E. Cryopreserved arterial homografts vs silver-coated Dacron grafts for abdominal aortic infections with intraoperative evidence of microorganisms. J. Vasc. Surg. 2011, 53, 1274–1281.e4. [CrossRef]

29. Hardman, S.; Cope, A.; Swann, A.; Bell, P.; Naylor, A.; Hayes, P. An In Vitro Model to Compare the Antimicrobial Activity of Silver-Coated Versus Rifampicin-Soaked Vascular Grafts. Ann. Vasc. Surg. 2004, 18, 308–313. [CrossRef]

30. Tilmacu, C.-M.; Mathieu, M.; Lavigne, J.-P.; Toupet, K.; Guerero, G.; Ponche, A.; Amalric, J.; Noël, D.; Mutin, P.H. In vitro and in vivo characterization of antibacterial activity and biocompatibility: A study on silver-containing phosphonate monolayers on titanium. Acta Biomater. 2015, 15, 266–277. [CrossRef]

31. Mohseni, M.; Shamlooo, A.; Aghababaei, Z.; Vossoughi, M.; Moravvej, H. Antimicrobial Wound Dressing Containing Silver Sulfadiazine with High Biocompatibility: In Vitro Study. Artif. Organs 2016, 40, 765–773. [CrossRef] [PubMed]

32. Rozhin, A.; Batasheva, S.; Kryuchkova, M.; Cherednichenko, Y.; Rozhina, E.; Fakhrullin, R. Biogenic Silver Nanoparticles: Synthesis and Application as Antibacterial and Antifungal Agents. Micromachines 2021, 12, 1480. [CrossRef] [PubMed]

33. Al-Zoubi, N.A.; Al-Shawwa, Z. Complications of endo-vascular aortic repair for abdominal aortic aneurysm: A retrospective single-centre experience. Ann. Med. Surg. 2021, 64, 102219. [CrossRef] [PubMed]