Cytisine, a Partial Agonist of α4β2 Nicotinic Acetylcholine Receptors, Reduced Unpredictable Chronic Mild Stress-Induced Depression-Like Behaviors

Jing Han1†, Dong-sheng Wang2†, Shui-bing Liu1* and Ming-gao Zhao1*  
1Department of Pharmacology, School of Pharmacy, The Fourth Military Medical University, Xi’an 710032,  
2Department of Orthopedics, Jinling Hospital, Clinical School of Nanjing, Second Military Medical University, Nanjing 210002, China

Abstract
Cytisine (CYT), a partial agonist of α4β2-nicotinic receptors, has been used for antidepressant efficacy in several tests. Nicotinic receptors have been shown to be closely associated with depression. However, little is known about the effects of CYT on the depression. In the present study, a mouse model of depression, the unpredictable chronic mild stress (UCMS), was used to evaluate the activities of CYT. UCMS caused significant depression-like behaviors, as shown by the decrease of total distances in open field test, and the prolonged duration of immobility in tail suspension test and forced swimming test. Treatment with CYT for two weeks notably relieved the depression-like behaviors in the UCMS mice. Next, proteins related to depressive disorder in the brain region of hippocampus and amygdala were analyzed to elucidate the underlying mechanisms of CYT. CYT significantly reversed the decreases of 5-HT1A, BDNF, and mTOR levels in the hippocampus and amygdala. These results imply that CYT may act as a potential anti-depressant in the animals under chronic stress.

Key Words: Cytisine, Depression, Nicotinic receptor, Chronic stress

INTRODUCTION
Neuron nicotinic acetylcholine receptors (nAChRs), belonging to the ligand-gated ion channel superfamily of neurotransmitter receptors. Varying combinations of nAChR subunits (α1-α10, β1-β4, γ, δ, and ε, α2-α7 and β2-β4 are expressed in the brain) assemble into pentameric ion channels, allowing for diverse pharmacological properties (Lukas et al., 1999), have been investigated for developing drugs that can potentially treat various central nervous system disorders. The cholinergic hypothesis of depression proposes that hyperactivity of the cholinergic system over that of the adrenergic system leads to depression. Several lines of evidence from rodent and human studies support this hypothesis (Janowsky et al., 1972). Additionally, a number of key antidepressants such as selective serotonin reuptake inhibitor antidepressants (SSRIs, e.g. fluoxetine, sertraline, paroxetine, and citalopram), the norepinephrine reuptake inhibitor reboxetine, the norepinephrine, imipramine, and nortriptyline) have been shown to possess partial antagonistic activities at nAChRs (Hennings et al., 1997; Fryer and Lukas, 1999; López-Valdés and García-Colunga, 2001; Arias et al., 2010). The α4β2 heteropentameric and α7 homopentameric subtypes are the two major nAChR subtypes expressed in the brain (Picciotto et al., 1998). The α4β2-nAChRs are widely distributed in the brain regions implicated in depression, including the thalamus, hippocampus, striatum, hypothalamus, amygdala, ventral tegmental area (VTA), locus coeruleus, and dorsal raphe nucleus (Philip et al., 2010), and thought to regulate the release of monoamine neurotransmitters through action in these areas (Hogg et al., 2003; Gotti et al., 2006).

Cytisine (CYT), a natural plant alkaloid, has been used as an inexpensive smoking-cessation aid for 50 years in Eastern Europe (Picciotto et al., 1995; Tutka and Zatoński, 2006). Cytisine, like varenicline, is a partial agonist selectively binding to the α4β2-nAChRs that appear to mediate nicotine dependence (Cahill et al., 2013). CYT behaves like a weak...
Fig. 1. Effects of CYT on depressive-like behaviors. CYT treatment blocked UCMS-induced depressive-like behaviors. (A) Timeline of the UCMS exposure, drug treatment, and behavioral tests. (B) Treatment of CYT (1 mg/kg) affected the total distance traveled in the OF. The total distance means the distance in whole area of the open field. (C) UCMS significantly increased the immobility time in the TST; CYT treatment decreased the immobility time in UCMS-exposed mice. (D) UCMS significantly increased the immobility time in the FST; CYT treatment decreased the immobility time in UCMS-exposed mice. n=6/group. *p<0.05, **p<0.01 compared with the control mice. #p<0.05 compared to the UCMS mice treated with saline.

partial agonist, mimicking effects of nicotine to a limited degree (Radchenko et al., 2015). Preclinical data support the antidepressant efficacy of CYT when used in conjunction with other primary antidepressants (e.g. SSRIs) (Philip et al., 2012). Cytisine is reported to reduce alcohol consumption and nicotine-induced alcohol drinking (Bell et al., 2009; Sajja and Rahman, 2013). Emerging evidence suggests that Cytisine significantly reduces depression-like behaviors in preclinical models that mimic major depressive disorder and co-morbid alcohol or nicotine use disorder (Rahman, 2015). However, little is known about the effects of CYT on the chronic stress-induced depression-like behaviors.

Nicotine increases 5-hydroxytryptamine (5-HT) release in the cortex, striatum, hippocampus, dorsal raphe nucleus (DRN), hypothalamus, and spinal cord (Ma et al., 2005). In addition, nicotine has antidepressant, anxiolytic, anorexic, antinociceptive (Cheeta et al., 2001; Schmidt et al., 2001; Seth et al., 2002), and antihypnotic properties (Salín-Pascual et al., 1996). While the diverse and complex effects of nicotine are not yet fully understood, considerable evidence suggests that 5-hydroxytryptamine (5-HT) may play a role in affective disorders and drug addiction. The effects involve stimulation of 5-HT1A receptors. The 5-HT1A receptors play a role in mediating the anti-depression effects of nicotine (Seth et al., 2002), however, little has been reported on the alteration of antidepressant action under chronic treatment in hippocampus and amygdala. Brain-derived neurotrophic factor (BDNF) functions in both the peripheral and central nervous systems (CNS) and has a beneficial impact on supporting the survival of existing neurons and promoting the growth of newly differentiated neurons and synapses (Bergami and Berninger, 2012). Several lines of evidence indicate that chronic stress and low level of BDNF are the key components of depression pathology (Yan et al., 2015).

The mammalian target of rapamycin (mTOR) signaling pathway is disturbed with major depressive disorder, and the activation of mTOR signaling is required for rapid antidepressant action in the hippocampus and other brain regions (Zhong et al., 2014). Unpredictable chronic mild stress (UCMS) decreases the phosphorylation levels of mTOR and its downstream signaling components, i.e., Akt-1 (Wang et al., 2015). This is accompanied by suppressed synaptic plasticity.

In the present study, we sought to understand whether CYT, a partial agonist of α4β2-nAChRs, attenuated depression-like behaviors in the UCMS mice and the underlying mechanisms. Here, we showed that CYT acted as an effective anti-depressive drug by regulating the serotonin receptor, BDNF, and mTOR signaling.

**MATERIALS AND METHODS**

**Drugs**

All chemicals were purchased from Sigma (St. Louis, MO, USA) unless otherwise stated. CYT was purchased from the Shanghai Pureone Biotechnology Co., Ltd (Shanghai, China). Anti-5-HT1A and anti-BDNF antibodies were purchased from Chemicon (Temecula, CA, USA). Anti-mTOR, p-mTOR, anti-AKT, p-AKT, anti-S6K, p-S6K, anti-CREB, and p-CREB antibodies were purchased from Abcam (Cambridge, UK). Anti-β-actin antibody was purchased from Sigma. All of the chemicals and reagents used were commercially available and of standard biochemical quality.

**Animals**

Male C57BL/6J mice (6-8 weeks, Laboratory Animal Center of the Fourth Military Medical University) were habituated for 1 week. Mice were housed under standard conditions (temperature 24 ± 2°C, humidity 50-60%, 12:12-h light/dark cycle, lights on at 08:00 h). Food and water were available ad libitum. Mice were marked on their tails with a permanent marker for identification and were randomly assigned to one of the different treatment groups (control, UCMS-saline, and UCMS-CYT; n=6/group). Each animal received the same treatment throughout the experiments. The Fourth Military Medical University Animal Care and Use Committee approved the animal protocols.
UCMS paradigm and drug treatments

UCMS mice were exposed to various stressors for 5 weeks, and time-matched control mice did not receive any stressors. The stressors include restraint (2 h), inversion of day/night light cycle (light off and light on), cold (in a cold room at 4°C for 1 h), 45° tilted cage (overnight), cage rotation (20 min), wet bedding (250 ml of water added into cage, overnight), no bedding (overnight), food and water deprivation (overnight), forced swimming (cold water 4°C for 6 min), and overcrowding (overnight) (Willner et al., 1987; Koo and Duman, 2008). On average, two stressors were administered per day. Non-stressed controls were handled only for cage changes and behavioral tests. The stressors and time course of UCMS have been detailed in Fig. 1. Mice were exposed to UCMS for a total of 5 weeks. At the beginning of the third week, UCMS-exposed mice were given CYT (1 mg/kg, i.p.) for two weeks. Cytisine (1-3 mg/kg) partially substituted for nicotine and at the highest dose tended to antagonize nicotine’s discriminative stimulus effects (Radchenko et al., 2015). In present study, 1 mg/kg of CYT (i.p.) was selected for the behavioral experiments.

Open field test (OF)

Mice were placed individually in one corner of the open field (50 cm length×45 cm width×30 cm deep box) (Shanghai Jiliang Software Technology, JLBhv-LAR-1, Shanghai, China) and allowed to freely explore the arena during a 15-min test session.

Tail suspension test (TST)

Mice were suspended by the tail with a paper clip attached with adhesive tape about 5 mm from the end of the tail (Shanghai Bio-will Co., Ltd., BW-DTS203, Shanghai, China). Time spent immobile was recorded over the 6 min test. The time spent immobile during the last 4 min was scored by an observer blind to treatments. After completion of the test, mice were returned to the home cage a holding cage until all cage-mates were tested.

Forced swim test (FST)

Mice were placed individually into glass cylinders (13 cm diameter, 25 cm tall) filled to a depth of 18 cm with water (25 ± 1.0°C) (Shanghai Jiliang Software Technology, JLBhv-FSR-1, Shanghai, China). The mice were placed in the cylinders for 6 min. The time spent immobile during the last 4 min was scored by an observer blind to treatments. Immobility was defined as the cessation of all movements (e.g., climbing, swimming) except those necessary for the mouse to keep its head above water (i.e., floating).

Western blot analysis

Tissue samples from the bilateral amygdala and hippocampus were dissected from the brain slices (300 μm) under the anatomical microscope. Total homogenates of amygdala and hippocampus sample were got from 6 mice respectively, and each sample of the mouse was examined by Western blot. Equal amounts of protein (30 μg) from the hippocampus and amygdala were separated and electro-transferred onto PVDF membranes (Invitrogen), which were probed with antibodies for 5-HT1A (dilution ratio 1:1,000), BDNF (dilution ratio 1:1,000), AKT (dilution ratio 1:1,000), p-AKT (dilution ratio 1:1,000), and BDNF (dilution ratio 1:1,000).
tio 1:1,000), p-S6K (dilution ratio 1:1,000), S6K (dilution ratio 1:1,000), mTOR (dilution ratio 1:1,000), p-mTOR (dilution ratio 1:1,000), CREB (dilution ratio 1:1,000), p-CREB (dilution ratio 1:1,000), and with β-actin (dilution ratio 1:10,000) as a loading control. The membranes were incubated with horseradish peroxidase-conjugated secondary antibodies (anti-rabbit/anti-mouse IgG for the primary antibodies), and bands were visualized using an ECL system (Lightning Blot System, Perkin Elmer, Waltham, MA, USA). For data quantification, band intensity of each blot was calculated as ratio relative to the β-actin. The intensity ratio of control group was set as 100%, and the intensity ratios of other treatment groups were expressed as percentage to the control group.

Data analysis and statistics
Results are expressed as mean ± SD. Data were evaluated using one-way analysis of variance (ANOVA) for post hoc comparisons (SPSS 13.0). Data that passed the homogeneity test were analyzed by the one-way ANOVA least significant difference (LSD) test. Data that did not pass the homogeneity test were analyzed by the one-way ANOVA Dunnett’s T3 test. p<0.05 was considered statistically significant.

RESULTS

Anti-depression like behaviors of CYT in UCMS model
Mice were exposed to UCMS for a total of 5 weeks. At the beginning of the third week, UCMS-exposed mice were given saline or CYT (i.p., 1 mg/kg) every day for 2 weeks (Fig. 1A). OF was used to determine the locomotor activity. Reduced activity in the total distance of an open field has been correlated with depression-like behaviors in rodents (El Yacoubi et al., 2003). Treatment of CYT increased the total distance traveled in UCMS mice during the 15-min test session in the OF (F(2, 15)=5.861, p=0.013, LSD test; Fig. 1B). In the tail suspension test (TST) and forced swim test (FST), the mean immobility period of the UCMS mice was significantly longer than that of the control group (TST: F(2, 15)=6.735, p=0.008, LSD test; Fig. 1C) (FST: F(2, 15)=6.456, p=0.009, LSD test; Fig. 1D). Treatment of CYT (1 mg/kg) decreased immobility time of UCMS mice in the TST (Fig. 1C) and FST (Fig. 1D).

CYT increases the levels of 5-HT1A in UCMS mice
Serotonin receptors play pivotal roles in depression disorders (Seth et al., 2002). UCMS decreased the levels of 5-HT1A receptor in homogenates of hippocampus and amygdala, whereas treatment of CYT reversed the alteration of 5-HT1A
Han et al. Anti-Depression Effects of Cytisine

**Fig. 5.** CYT increased mTOR signaling activities in the amygdala. (A) Representative Western blot of proteins in amygdala. (B) Treatment of CYT (1 mg/kg) up-regulated the ratio of p-AKT/ Akt in UCMS mice. (C) Treatment of CYT (1 mg/kg) up-regulated the ratio of p-mTOR/ mTOR in UCMS mice. (D) Treatment of CYT (1 mg/kg) up-regulated the ratio of p-S6K/S6K in UCMS mice. (E) Treatment of CYT (1 mg/kg) up-regulated the ratio of p-CREB/CREB in UCMS mice. n=6/group. *p<0.05 compared with the control mice. *p<0.05 compared to the UCMS mice treated with saline.

receptor (Hippocampus: F(2, 15)=10.29, p=0.002, LSD test; Fig. 2A) (Amygdala: F(2, 15)=12.95, p=0.001, LSD test; Fig. 2B).

**CYT up-regulates levels of BDNF in UCMS mice**

Levels of BDNF is decreased in depression disorders (Yan et al., 2015). UCMS decreased the expression levels of BDNF in hippocampus and amygdala of UCMS mice, whereas treatment of CYT increased the BDNF levels in the hippocampus and amygdala (Hippocampus: F(2, 15)=25.798, p<0.001, LSD test; Fig. 3A) (Amygdala: F(2, 15)=17.735, p<0.001, LSD test; Fig. 3B).

**Treatment of CYT increases the activities of mTOR signaling in UCMS mice**

mTOR signaling pathway is down-regulated in the major depressive disorder (Zhong et al., 2014). Proteins related to the mTOR signaling in the hippocampus and amygdala were detected by the western blot. We found that the total levels of AKT, mTOR, S6K, and CREB were not changed in the hippocampus of UCMS mice, however, the ratio of p-AKT/AKT (F(2, 15)=25.757, p<0.001, LSD test; Fig. 4A, 4B), p-mTOR/mTOR (F(2, 15)=21.342, p<0.001, LSD test; Fig. 4C), p-S6K/S6K (F(2, 15)=14.961, p<0.001, LSD test; Fig. 4D), or p-CREB/CREB (F(2, 15)=6.682, p=0.008, LSD test; Fig. 4E) was significantly decreased. Treatment of CYT could reverse above alterations of p-AKT/AKT, p-mTOR/mTOR, p-S6K/S6K, or p-CREB/CREB. Similar results were found in the region of amygdala. Treatment of CYT could ameliorate the decrease of ratio of p-AKT/ AKT (F(2, 15)=12.01, p=0.001, LSD test; Fig. 5A, 5B), p-mTOR/ mTOR (F(2, 15)=9.452, p=0.002, LSD test; Fig. 5C), p-S6K/S6K (F(2, 15)=9.293, p=0.002, LSD test; Fig. 5D), or p-CREB/CREB (F(2, 15)=11.491, p=0.001, LSD test; Fig. 5E) in the amygdala.

**DISCUSSION**

Nicotinic agonists, through activation of neuronal nAChR, improve cognitive performance in both animals and humans. nAChRs, including α4β2-nAChRs, influence synaptic plasticity through the facilitation of presynaptic and postsynaptic mechanisms (Glassman et al., 1990; Salin-Pascual et al., 1995; Semba et al., 1998). In present study, mice were exposed to UCMS for a total of 5 weeks exhibited significant depressive behaviors, as shown by the decreased locomotor activity and the longer mean immobility period in the tail suspension test and forced swim test. Cytisine (1 mg/kg) could alleviate the depressive-like behaviors in the chronic stress mice. Present study indicates that CYT, a partial agonist of α4β2-nicotinic re-
ceptors, may act as a potential anti-depressant in the animals under chronic stress.

Nicotinic receptors are pentameric, ligand-gated ion channels comprised of heteromeric or homomeric subunits encoded by nine α (α2-10) and three β (β2-4) genes in the human brain. nAChR blockers potentiate the effects of selective serotonin reuptake inhibitors (SSRIs) in some treatment-resistant patients (Mineur et al., 2015). Interactions between the serotonergic and cholinergic systems are quite related to mood disorders. The effect of nicotine is attributed to an activation of several β2- and β4-containing receptors, as well as homomeric α7 nicotine receptors, which are broadly distributed in the CNS (Damaj et al., 2003; Millar, 2003; Woolorton et al., 2003), Presynaptic nAChRs regulate neurotransmitter release, while postsynaptic nAChRs activate intracellular signaling and gene transcription. Nicotine exposure is known to have multiple effects on the 5-HT system, and thus the expression of nAChRs by 5-HT neurons may play an important role in the 5-HT abnormalities. CYT, as a partial agonist of αβ2-nAChRs, up-regulates the levels of 5-HT1A receptors in hippocampus and amygdala and produces antidepressant-like effects in the present UCMS model.

Stress and depression are associated with neuronal atrophy and decrease of synaptic connections and leads to decreased expression and release of BDNF in the prefrontal cortex, limbic brain regions, and hippocampus (Nasca et al., 2013). Present study demonstrates that BDNF levels are decreased in the hippocampus and amygdala. Treatment of CYT up-regulates BDNF levels, and improves stress-induced cognitive and behavioral alterations.

The mTOR signaling pathway is implicated in the pathophysiology of depression and in the antidepressant-like effects of different compounds (Abelaira et al., 2014; Zhong et al., 2014). Our results found that UCMS induced a significant decrease of p-AKT, p-S6K, p-mTOR and p-CREB levels in the hippocampus and amygdala. CYT administration significantly ameliorated the decrease of ratio of p-AKT/AKT, p-mTOR/mTOR, p-S6K/S6K, or p-CREB/CREB. PI3K/AKT pathway and mTOR signaling play an important role in the production of BDNF, implicating the activities of CYT in the antidepressant response (Duman and Voleti, 2012).

In summary, the present study shows that CYT produces antidepressant-like effects through modulating the 5-HT1A, BDNF, and mTOR signaling in the hippocampus and amygdala of UCMS model. This study is helpful to elucidate the mechanisms underlying the antidepressant effects of CYT and for the clinical treatment of depression by traditional herbs.

ACKNOWLEDGMENTS

This work was supported by grants from the Certificate of China Postdoctoral Science Foundation (2014M552632) and the National Science Foundation of China (31271144, 31271126, 81325022).

REFERENCES

Abelaira, H. M., Réus, G. Z., Neotti, M. V. and Quevedo, J. (2014) The role of mTOR in depression and antidepressant responses. Life Sci. 101, 10-14.

Arias, H. R., Rosenberg, A., Targowska-Duda, K. M., Feuerbach, D., Joziwka, K., Moaddel, R. and Wainer, I. W. (2010) Tricyclic antidepressants and mecamylamine bind to different sites in the human alpha4beta2 nicotinic receptor ion channel. Int. J. Biochem. Cell Biol. 42, 1007-1018.

Bell, R. L., Eiler, B. J., 2nd, Cook, J. B. and Rahman, S. (2009) Nicotinic receptor ligands reduce ethanol intake by high alcohol-drinking HAD-2 rats. Alcohol 43, 581-592.

Bergami, M. and Berninger, B. (2012) A fight for survival: the challenges faced by a newborn neuron integrating in the adult hippocampus. Dev. Neurobiol. 72, 1016-1031.

Cahill, K., Stevens, S., Perera, R. and Lancaster, T. (2013) Pharmacological interventions for smoking cessation: an overview and network meta-analysis. Cochrane Database Syst. Rev. 5, CD009329.

Cheeta, S., Irvine, E. E., Kenny, P. J. and File, S. E. (2001) The dorsal raphe nucleus is a crucial structure mediating nicotine's anxiolytic effects and the development of tolerance and withdrawal responses. Psychopharmacology (Berl.) 155, 78-85.

Damaj, M. I., Kao, W. and Martin, B. R. (2003) Characterization of spontaneous and precipitated nicotine withdrawal in the mouse. J. Pharmacol. Exp. Ther. 307, 526-534.

Duman, R. S. and Voleti, B. (2012) Signaling pathways underlying the pathophysiology and treatment of depression: novel mechanisms for rapid-acting agents. Trends Neurosci. 35, 47-56.

El Yacoubi, M., Bouali, S., Popa, D., Naudon, L., Leroux-Nicollet, I., Hamon, M., Costentin, J., Adrien, J. and Vaugeois, J. M. (2003) Behavioral, neurochemical, and electrophysiological characterization of a genetic mouse model of depression. Proc. Natl. Acad. Sci. U.S.A. 100, 6227-6232.

Fryer, J. D. and Lukas, R. J. (1999) Antidepressants noncompetitively inhibit nicotinic acetylcholine receptor function. J. Neurochem. 72, 1117-1124.

Glassman, A. H., Helzer, J. E., Covey, L. S., Cottler, L. B., Steiner, F., Tipp, J. E. and Johnson, J. (1990) Smoking, smoking cessation, and major depression. JAMA 264, 1546-1549.

Gotti, C., Riganti, L., Vailati, S. and Clementi, F. (2006) Brain neuronal nicotinic receptors as new targets for drug discovery. Curr. Pharm. Des. 12, 407-428.

Hennings, E. C., Kiss, J. P. and Vizi, E. S. (1997) Nicotinic acetylcholine receptor antagonist effect of fluoxetine in rat hippocampal slices. Brain Res. 759, 292-294.

Hogg, R. C., Raggenbass, M. and Bertrand, D. (2003) Nicotinic acetylcholine receptors: from structure to brain function. Rev. Physiol. Biochem. Pharmacol. 147, 1-46.

Janowsky, D. S., el-Yousef, M. K., Davis, J. M. and Sekerke, H. J. (1972) A cholinergic-adrenergic hypothesis of mania and depression. Lancet 2, 632-635.

Koo, J. W. and Duman, R. S. (2008) IL-1beta is an essential mediator of the antineurogenic and anhedonic effects of stress. Proc. Natl. Acad. Sci. U.S.A. 105, 751-756.

López-Valdés, H. E. and García-Colunga, J. (2001) Antagonism of nicotinic acetylcholine receptors by inhibitors of monoamine uptake. Mol. Psychiatry 6, 511-519.

Lucas, R. J., Changeux, J. P., Le Novère, N., Albuquerque, E. X., Balfour, D. J., Berg, D. K., Bertrand, D., Chiappinelli, V. A., Clarke, P. B., Collins, A. C., Dani, J. A., Grady, S. R., Kellar, K. J., Lindstrom, J. M., Marks, M. J., Quik, M., Taylor, P. W. and Wonnacott, S. (1999) International Union of Pharmacology. XX. Current status of the nomenclature for nicotinic acetylcholine receptors and their subunits. Pharmacol. Rev. 51, 397-401.

Ma, Z., Streckner, R. E., McKenna, J. T., Thakkar, M. M., McCarley, R. W. and Tao, R. (2005) Effects on serotonin of (-)-nicotine and dimethylphenylpipеразин in the dorsal raphe and nucleus accumbens of freely behaving rats. Neuroscience 135, 949-958.

Millar, N. S. (2003) Assembly and subunit diversity of nicotinic acetylcholine receptors. Biomembr. Soc. Trans. 31, 869-874.

Mineur, Y. S., Einstein, E. B., Bentham, M. P., Wigsten, M. B., Blake- man, S., Newbold, S. A. and Picciotto, M. R. (2015) Expression of the 5-HT1A serotonin receptor in the hippocampus is required for social stress resilience and the antidepressant-like effects induced by the nicotinic partial agonist cytisine. Neuropsychopharmacology 40, 936-946.
Nasca, C., Xenos, D., Barone, Y., Caruso, A., Scaccianoce, S., Mattisiano, F., Battaglia, G., Mathé, A. A., Pittaluga, A., Lionetto, L., Simmaco, M. and Nicoletti, F. (2013) L-acetylcarnitine causes rapid antidepressant effects through the epigenetic induction of mGlur2 receptors. Proc. Natl. Acad. Sci. U.S.A. 110, 4804-4809.

Philip, N. S., Carpenter, L. L., Tyrka, A. R. and Price, L. H. (2010) Nicotinic acetylcholine receptors and depression: a review of the preclinical and clinical literature. Psychopharmacology (Berl.) 212, 1-12.

Philip, N. S., Carpenter, L. L., Tyrka, A. R. and Price, L. H. (2012) The nicotinic acetylcholine receptor as a target for antidepressant drug development. ScientificWorldJournal 2012, 104105.

Picciotto, M. R., Zoli, M., Léna, C., Bessis, A., Lallemand, Y., Le Novère, N., Vincent, P., Pich, E. M., Brület, P. and Changeux, J. P. (1995) Abnormal avoidance learning in mice lacking functional high-affinity nicotine receptor in the brain. Nature 374, 65-67.

Picciotto, M. R., Zoli, M., Rimondini, R., Léna, C., Marubio, L. M., Pich, E. M., Fuxe, K. and Changeux, J. P. (1998) Acetylcholine receptors containing the beta2 subunit are involved in the reinforcing properties of nicotine. Nature 391, 173-177.

Radchenko, E. V., Dravolina, O. A. and Bespalov, A. Y. (2015) Agonist and antagonist effects of cytisine in vivo. Neuropharmacology 95, 206-214.

Rahman, S. (2015) Targeting brain nicotinic acetylcholine receptors to treat major depression and co-morbid alcohol or nicotine addiction. CNS Neurol. Disord. Drug Targets 14, 647-653.

Sajja, R. K. and Rahman, S. (2013) Cytisine modulates chronic voluntary ethanol consumption and ethanol-induced striatal up-regulation of DeltaFosB in mice. Alcohol 47, 299-307.

Salin-Pascual, R. J., de la Fuente, J. R., Galicia-Polo, L. and Drucker-Colin, R. (1995) Effects of transdermal nicotine on mood and sleep in nonsmoking major depressed patients. Psychopharmacology (Berl.) 121, 476-479.

Salin-Pascual, R. J., Rosas, M., Jimenez-Gench, A., Rivera-Meza, B. L. and Delgado-Parra, V. (1996) Antidepressant effect of transdermal nicotine patches in nonsmoking patients with major depression. J. Clin. Psychiatry 57, 387-389.

Schmidt, B. L., Tambelli, C. H., Gear, R. W. and Levine, J. D. (2001) Nicotine withdrawal hyperalgesia and opioid-mediated analgesia depend on nicotine receptors in nucleus accumbens. Neuroscience 106, 120-136.

Seba, J., Mataki, C., Yamada, S., Nankai, M. and Toru, M. (1998) Antidepressant-like effects of chronic nicotine on learned helplessness paradigm in rats. Biol. Psychiatry 43, 389-391.

Seth, P., Cheeta, S., Tucci, S. and File, S. E. (2002) Nicotinic–serotonergic interactions in brain and behavior. Pharmacol. Biochem. Behav. 71, 796-805.

Tutka, P. and Zatorowski, W. (2006) Cytisine for the treatment of nicotine addiction: from a molecule to therapeutic efficacy. Pharmacol. Rep. 58, 777-798.

Wang, W., Lu, Y., Xue, Z., Li, C., Wang, C., Zhao, X., Zhang, J., Wei, X., Chen, X., Cui, W., Wang, Q. and Zhou, W. (2015) Rapid-acting antidepressant-like effects of acetyl-l-carnitine mediated by PI3K/AKT/BDNF/VGF signaling pathway in mice. Neuroscience 285, 281-291.

Willner, P., Towell, A., Sampson, D., Sophokleous, S. and Muscat, R. (1987) Reduction of sucrose preference by chronic unpredictable mild stress, and its restoration by a tricyclic antidepressant. Psychopharmacology (Berl.) 93, 358-364.

Wooltorton, J. R., Pidoplichko, V. I., Broide, R. S. and Dani, J. A. (2003) Differential desensitization and distribution of nicotinic acetylcholine receptor subtypes in midbrain dopamine areas. J. Neurosci. 23, 3176-3185.

Yan, W. J., Tan, Y. C., Xu, J. C., Tang, X. P., Zhang, C., Zhang, P. B. and Ren, Z. Q. (2015) Protective effects of silibinin and its possible mechanism of action in mice exposed to chronic unpredictable mild stress. Biomed. Ther. (Seoul) 23, 245-250.

Zhong, P., Wang, W., Pan, B., Liu, X., Zhang, Z., Long, J. Z., Zhang, H. T., Cravatt, B. F. and Liu, Q. S. (2014) Monoacylglycerol lipase inhibition blocks chronic stress-induced depressive-like behaviors via activation of mTOR signaling. Neuropsychopharmacology 39, 1763-1776.