ABSTRACT
Current management of neuromyelitis optica (NMO) is noncurative and only partially effective. Immunosuppressive or immunomodulatory agents are the mainstays of maintenance treatment. Safer, better-tolerated, and proven effective treatments are needed. The perceived rarity of NMO has impeded clinical trials for this disease. However, a diagnostic biomarker and recognition of a wider spectrum of NMO presentations has expanded the patient population from which study candidates might be recruited. Emerging insights into the pathogenesis of NMO have provided rationale for exploring new therapeutic targets. Academic, pharmaceutical, and regulatory communities are increasingly interested in meeting the unmet needs of patients with NMO. Clinical trials powered to yield unambiguous outcomes and designed to facilitate rapid evaluation of an expanding pipeline of experimental agents are needed. NMO-related disability occurs incrementally as a result of attacks; thus, limiting attack frequency and severity are critical treatment goals. Yet, the severity of NMO and perception that currently available agents are effective pose challenges to study design. We propose strategies for NMO clinical trials to evaluate agents targeting recovery from acute attacks and prevention of relapses, the 2 primary goals of NMO treatment. Aligning the interests of all stakeholders is an essential step to this end. Neurology® 2015;84:1805–1815

GLOSSARY
AQP4 = aquaporin-4; IgG = immunoglobulin G; MS = multiple sclerosis; NMO = neuromyelitis optica; PLEX = plasma exchange.

Challenges and opportunities in designing clinical trials for neuromyelitis optica

Neuromyelitis optica (NMO) is a rare, chronic, autoimmune disease involving the CNS, specifically optic nerves and spinal cord. The specific biomarker aquaporin-4 (AQP4)–immunoglobulin G (IgG) is detected in most patients, but assays for AQP4-IgG vary in sensitivity and specificity. In vivo and in vitro studies and favorable response to plasma exchange (PLEX) suggest that AQP4-IgG is pathogenic. Some patients with NMO may develop symptoms implicating involvement of other CNS regions such as the area postrema and hypothalamus. Others have...
limited variants of NMO (e.g., recurrent optic neuritis or recurrent myelitis). Such patients are collectively referred to as having NMO spectrum disorders. In seropositive patients, a confident diagnosis can be made after a single clinical event. While AQP4-IgG detection facilitates diagnosis and predicts relapse, an important proportion of cases are seronegative.10,11

Acute NMO attacks are generally more severe than those of multiple sclerosis (MS) and may be fatal.12,13 Mortality estimates have improved from 30% at 5 years,12 to 9% at 6 years.13 Unlike MS, disability in NMO occurs as cumulative sequelae of attacks rather than during a secondary progressive phase.14 Thus, minimizing the frequency and severity of attacks is a primary therapeutic goal. IV corticosteroids and, in refractory cases, PLEX are currently used to treat acute attacks. Immunosuppressive agents (e.g., azathioprine, mycophenolate mofetil, rituximab, and corticosteroids) are prescribed to reduce attack frequency, based on small, uncontrolled studies.15–17 These agents are collectively referred to as “empiric” treatments herein, to avoid any suggestion that a standard of NMO therapy has been established.

Several obstacles have historically limited clinical trials in NMO. However, these barriers are now mitigated by important advances that increase the feasibility of informative trials (table 1). An increasing number of therapeutic candidates, including those repurposed for evaluation in NMO, and emerging candidates specific to AQP4 (e.g., aquaporumab),18 create an immediate need for definitive NMO clinical trials. Promising repurposed agents include the following:

1. Rituximab (anti-CD20 monoclonal antibody targeting B cells)19,20
2. Eculizumab (anti-C5 monoclonal antibody targeting complement)21
3. Tocilizumab (anti-IL-6R monoclonal antibody targeting T- and B-cell activation, Th17 differentiation, and plasmablast survival)22–24

Clinical trial templates developed for MS are not directly applicable to NMO (table 2). Herein, we review issues pertinent to clinical trial development in NMO based on discussions among clinicians, researchers, industry partners, and patients at research-focused meetings of the Guthy-Jackson Charitable Foundation. Key concepts were further refined by all coauthors, including one serving in a regulatory capacity.

OVERARCHING CONCEPTS FOR NMO CLINICAL TRIAL DESIGN

No drug or treatment has been proven to be safe and effective in NMO in randomized, controlled studies, and none has received regulatory approval.

Opinions vary widely among investigators regarding ethics of placebo-controlled studies for maintenance treatment of NMO. NMO-associated transverse myelitis and optic neuritis attacks can result in devastating neurologic consequences.12,13 Moreover, retrospective analyses suggest that current empiric therapies may be moderately effective.20,25,26 However, ascertainment bias and biases inherent in treatment discontinuation and reporting of attacks in retrospective studies preclude conclusions regarding efficacy of any currently used agent. Furthermore, rigorous studies frequently contradict theoretical benefit27 or reveal unexpected toxicity,28 especially when agents are tested in combination.29 Some investigators and regulatory agencies regard the evidence supporting effectiveness of current empiric treatments sufficiently inadequate that placebo-controlled studies in NMO are justified for several reasons, including:

1. Noninferiority studies require an established standard of treatment that currently does not exist.
2. NMO clinical trials must address safety as well as efficacy both for acute and chronic exposure, considering that long-term therapy is likely necessary.
3. Sensitive and meaningful NMO-customized outcomes measures are needed to inform treatment.

Attacks, rather than neurodegeneration, are the principal contributors to NMO-related disability.14 Thus, frequency and severity of attacks are the most suitable primary endpoints in NMO clinical trials. Validated criteria, including imaging evidence, that define specific signs or symptoms of an acute attack are necessary for optimal trial design. Whether nonattack-related effects of the disease (e.g., changes in cognitive function, behavior, or other neurologic functions) occur in NMO is uncertain. Likewise, laboratory-based biomarkers reflecting or predicting NMO attack frequency, severity, or therapeutic efficacy must be validated before consideration as surrogate outcomes.30–34 Poor cross-sectional correlation of AQP4 autoantibody titers with NMO disease activity and severity exemplifies this issue. Longitudinal assessment of titers within individuals may be more promising than between individuals, although this has not been adequately explored.35,36 Similar
considerations apply to serum cytokine levels, immune cell profiles,\(^{34}\) or surrogates of tissue-specific injury (e.g., CSF glial fibrillary astrocytic protein\(^{37}\)). CSF parameters are difficult to measure serially, and MRI signals are highly variable among individuals. Brain MRI abnormalities in NMO occur in selective regions, are difficult to quantify, and have inconsistent association with clinical disability. Changes in retinal nerve fiber layer thickness over time, as measured by optical coherence tomography, are more striking in NMO than in MS.\(^{38}\) However, some patients with NMO spectrum disorders do not have optic nerve events, and changes in optical coherence tomography are typically delayed after acute events.

### Table 1 Obstacles and advances influencing development of clinical trials for NMO

| Obstacles                                                                 | Advances                                                                 |
|--------------------------------------------------------------------------|--------------------------------------------------------------------------|
| Rarity                                                                   | Improved diagnosis                                                       |
| Prevalence estimated to be 1–5/100,000\(^{48,49}\)                         | Increased sensitivity, attributable to novel biomarker, AQP4-IgG         |
| 1%–5% of demyelinating diseases in western countries; >50% in some Asian countries\(^{50}\) | Appreciation of NMO spectrum disorder with resulting expansion of pool of eligible patients |
| Diagnostic uncertainty                                                   | Increased specificity of diagnosis                                        |
| Mimics: MS, neurosarcoïdosis, paraneoplastic disorders; mitochondrial encephalomyelopathies | More sensitive and specific assays for AQP4-IgG                          |
| Confusing nomenclature: Devic disease; optosarcipal MS; syndromic nomenclature (e.g., recurrent transverse myelitis) | Improved ability to distinguish between NMO and MS                      |
| Heterogeneity                                                            | Greater availability of natural history data\(^{13}\) and data reflecting impact of maintenance therapy |
| AQP4-IgG seropositive vs seronegative\(^{50}\)                           | Identification of relevant therapeutic targets\(^{51,52}\)               |
| Monophasic vs relapsing\(^{1,2}\)                                        | AQP4 antibodies. Protective antibodies engineered by protein modification to abrogate complement- and cell-dependent cytotoxicity\(^{28}\) |
| Variability in sensitivity and specificity in AQP4-IgG assays\(^{8}\)  | Specific B-cell population markers and B-cell/plasmablast survival factors (IL-6)\(^{22,23,34}\) |
| Challenges of placebo-controlled design                                  | Downstream effectors of AQP4-IgG interaction with AQP4, including complement,\(^{21}\) neutrophils, and eosinophils\(^{53,54}\) |
| Risk of severe and irreversible disability from attacks                  | Th17 pathway\(^{55}\)                                                   |
| Generally accepted “standard treatments” for attacks and preventative treatments,\(^{3,5}\) despite lack of rigorous proof in NMO-dedicated placebo-controlled studies |                                                                 |
| Variable and unpredictable disease course                                 |                                                                           |
| Heterogeneous risk of events following enrollment; not easily predicted at current time |                                                                 |
| Changing prognosis with changes in diagnostic criteria, early recognition, and treatment |                                                                 |
| “Benign” cases of NMO\(^{57}\)                                           |                                                                           |
| Lack of prospectively validated outcome measures designed for use in NMO |                                                                           |
| Different evolution and more limited involvement of CNS than MS; Expanded Disability Status Scale and related functional systems not immediately translatable |                                                                 |
| Lack of imaging or laboratory biomarkers reflecting disease activity and response to therapy |                                                                 |
| Pathogenesis incompletely understood; optimum targets unclear             |                                                                           |
| Complexities of delivery of novel antigen-directed therapeutics\(^{18}\) |                                                                           |
| Achieving adequate concentrations and sufficient half-life to provide protection at site of target antigen |                                                                 |

Abbreviations: AQP4 = aquaporin-4; IgG = immunoglobulin G; IL-6 = interleukin 6; MS = multiple sclerosis; NMO = neuromyelitis optica.

### Table 2 Selected features differentiating NMO from MS

| Progressive disease uncommon; disability accumulates as a result of attacks\(^{14}\) | Attacks are more severe in NMO |
|--------------------------------------------------------------------------|-------------------------------|
| Absence of informative MRI changes that serve as primary outcome of proof-of-concept phase II clinical trials in MS\(^{56}\) | Predilection of specific regions of CNS in NMO (optic nerve, spinal cord, area postrema, and circumventricular organs) |

Abbreviations: MS = multiple sclerosis; NMO = neuromyelitis optica.
AQP4-IgG, AQP4, complement, and granulocytes. Typically, acute events are treated with high-dose pulse corticosteroids, and with PLEX in refractory cases. The following considerations apply to clinical trials of treatment of acute NMO episodes.

**Inclusion/exclusion criteria.** An issue common to all potential NMO clinical trials is whether participation should be confined to AQP4-IgG–seropositive individuals (table 3). Restricting enrollment to seropositive patients would enhance cohort homogeneity and diagnostic certainty, excluding NMO mimics, such as sarcoidosis or paraneoplastic disorders. The majority of potential study participants are seropositive. However, current serologic assays have imperfect sensitivity and may not predict treatment response. To address potential for differential efficacy, patients may need to be randomized based on serostatus.

**Clinical trial designs.** The potential for severe and irreversible morbidity during acute NMO attack is particularly relevant in evaluating the acceptability and ethical correctness of trial design (Table 3 and figure 1).

**Single agent vs placebo.** Randomized placebo-controlled trials established that corticosteroids shorten recovery of acute optic neuritis and MS relapses (figure 1A). Although no comparable studies exist in NMO, widespread corticosteroid use and acceptable short-term adverse event profile make it difficult to prohibit their use. Following treatment with corticosteroids, subjects are randomized to an experimental drug vs placebo, either after or concurrent to their treatment with corticosteroid therapy. PLEX was superior to sham pheresis in a controlled study of acute, severe attacks of CNS demyelinating disease that failed corticosteroid treatment; that study included 2 patients with NMO. Subsequent uncontrolled prospective and retrospective clinical studies in patients with NMO also reported favorable outcomes. In the only sham-controlled study, patients failing the initial randomized treatment crossed over to the comparator, allowing both intra- and interindividual comparisons of PLEX efficacy with sham. The opportunity for favorable outcomes may decline as the interval from treatment initiation lengthens and neurologic injury becomes irreversible; therefore, it is necessary to consider the sequence of treatment in the analysis. Crossover of subjects who do not improve is also a risk mitigation and recruitment facilitation strategy by guaranteeing access to the active

### Table 3 Pros and cons of design issues in NMO clinical trials

| Study measure | Pro | Con |
|---------------|-----|-----|
| **Study design** | | |
| Placebo control | Scientifically rigorous; required sample size smaller assuming current empiric therapies are effective; fewer total attacks required assuming current therapies effective | Potential for greater risk of attacks in placebo group if current empiric therapies are efficacious and withheld; addresses superiority to placebo, not to current empiric regimens; risk of attack for a given patient in control arm potentially greater than in treatment arm |
| Placebo control as add-on to standard therapy | Rigorous, while allowing for potential benefits of empiric therapy | Incomplete consensus on details of empiric therapy makes uniformity difficult; empiric therapy may have failed in a patient who is then continued on the same regimen; difficult to disentangle effects of experimental and empiric therapy; may reduce event number and necessitate a larger sample size or a longer study than if compared with placebo; potential additive toxicity of experimental and empiric treatment |
| Comparison to empiric therapy | May mitigate risk to individual patient and enhance acceptability or enrollment; addresses whether new therapy offers incremental benefit to current practice | Empiric therapy not established to be efficacious; superiority required; if the standard therapy is effective, a larger sample size or longer study may be necessary if compared with placebo; specified standard therapy may be ineffective or even harmful; may be unacceptable for regulatory approval in some jurisdictions |
| **Inclusion criteria** | | |
| AQP4-IgG+ | Enhanced homogeneity; majority of patients are AQP4-IgG+ | Assay incompletely sensitive; some may seroconvert in follow-up; excludes 30% of patients classified as NMO spectrum disorder and thereby prolongs enrollment; effects of treatment on AQP4-IgG seronegative subjects not addressed; may require licensing of AQP4-IgG as codiagnostic and may restrict access after approval to patients who are AQP4-IgG seronegative |
| **Endpoint** | | |
| Time to first event | Enables early exit in treatment failure | Latency to onset of action uncertain and treatment failure declaration may be premature; loss of potential information from each patient; attack definition may not be sufficiently rigorous to distinguish true from pseudo-attacks |
| Annualized relapse rate | More information for each enrolled patient because all observed attacks used in analysis | Early withdrawals pose major problem for study power and interpretability of results |
| Composite of relapse severity and frequency | May add additional important information that could be an important aspect in evaluation of drug efficacy | No validated method exists to evaluate attack severity |

Abbreviations: AQP4 = aquaporin-4; IgG = immunoglobulin G; NMO = neuromyelitis optica.
treatment in either the first or second treatment phase.

Add-on to corticosteroid treatment. A randomized, placebo-controlled study of an add-on therapy may be appropriate for agents with theoretical additive or synergistic benefit (e.g., addition of a granulocyte inhibitor or complement inhibitor to pulse corticosteroid treatment) (figure 1B). If combination therapy proves efficacious, follow-up studies would be necessary to evaluate superiority of the experimental drug as a single agent.

Outcome measures. Evaluation of attack-specific outcomes requires that neurologic deficits be assessed and recovery quantified with pertinent impairment scales. An integrated estimate of recovery (e.g., mild, moderate, marked) could be pooled regardless of the site of the attack. The proportion of patients who achieve these levels of recovery could be compared across treatments. Thus, patients having a variety of neurologic deficits (e.g., visual, motor, or sensory) could be included in a single study. Recovery of the patient-specific “targeted neurologic deficit” is a focused and relevant outcome and is typically assessed within 2 to 4 weeks after an event. Favorable treatment-related outcomes generally occur rapidly but delayed benefit is also of interest.

Attack prevention. Synopsis. Attack prevention studies are typically offered to patients in remission who may more favorably consider participation in a placebo-controlled trial. Current preventive regimens are nonspecific, and include low-dose oral corticosteroids and lymphocyte-directed immunosuppressants. Newer strategies may target upstream mechanisms involved in AQP4-IgG generation, such as specific plasmablast expansion, CNS trafficking, and autoreactive T-cell activation. The following considerations are relevant to clinical trials evaluating maintenance treatments.

Inclusion/exclusion criteria. Restricting enrollment to seropositive individuals was discussed in the context of acute disease (table 3). Comorbidities, such as infections, immunodeficiencies, and agent-specific risk factors, typically exclude participation. A minimum level of recent disease activity is required to exclude subjects with quiescent disease. However, targeting patients with high levels of recent disease activity for enrollment to enhance study power may limit the ability to generalize results. Furthermore, the attack frequency may regress to a lower mean event rate on study.

Clinical trial designs. Three potential clinical trial strategies are considered for phase IIb/III maintenance studies.

Experimental agent vs placebo. Placebo-controlled studies are the standard for pivotal trials (figure 2A). We propose several mitigating strategies to reduce the risk of a disabling attack during placebo exposure, including the following:

1. Liberal “escape” criteria in the event of treatment failure (e.g., a single attack)
2. Ratio of subjects randomized to experimental treatment vs placebo >1:1, reducing the odds that a given patient will be assigned to the placebo group
3. Early recognition and uniform aggressive treatment of attacks
4. Limited duration of placebo-controlled period after which patients may receive active study drug or empiric treatment.

(A) An experimental treatment (Expt Tx) is compared with placebo. At enrollment while in relapse, a patient would either discontinue prior maintenance treatment (placebo-only design) or not (placebo-controlled add-on). The study design could allow patients whose initial treatment failed to cross over to the other treatment regimen. (B) An experimental treatment is added to a standard regimen for management of an acute attack such as pulse high-dose IV corticosteroids or plasma exchange (PLEX). The treatment may be used along with the other empiric treatment from the outset or after a predefined period of initial treatment in patients who have met failure criteria.
Studies should be designed to observe the minimum number of events necessary for achieving interpretable and robust outcomes. Placebo-controlled studies require fewer observed attacks to achieve adequate statistical power than those comparing experimental and an efficacious empiric therapy; however, the risk of an attack for an individual patient may be greater in a placebo-controlled than in an active treatment comparator study. Placebo-controlled studies do not address superiority to current empiric treatments that may be perceived as moderately effective and relatively inexpensive. To address this concern, one or more empiric therapy arms could be added to a placebo-controlled study or subsequently evaluated against an experimental agent that is proven to be superior to placebo.

Add-on experimental agent vs placebo. Subjects are randomized to experimental agent or placebo, while also receiving a protocol-defined empiric regimen (figure 2B). Investigators may terminate patient participation after a first on-study relapse. However, extended participation in the randomized phase beyond the first event is more acceptable to some stakeholders in an add-on than in a pure placebo-controlled trial. Extended participation increases power by allowing each patient to contribute more than a single event. Potential limitations include possible additive toxicity of multiple agents and unwarranted attribution of benefit to combined therapy when one agent might be solely responsible. Effective empiric therapy may also blur outcome specificity, thereby requiring recruitment of additional subjects or observation of more events than in a pure placebo trial. However, the per-subject risk of an event may be lower. As stated above, if a combination regimen proves efficacious, ensuing studies would be required to assess the experimental agent alone (figure 2B) vs the combination.

Experimental agent vs existing therapy. Direct comparison of an investigational agent to an existing empiric...
therapy circumvents concerns about pure placebo exposure (figure 2C). Multiple options for empiric therapy during the trial may be necessary, as subjects may have failed one or more of comparator treatments. A priori consensus of empiric therapy mitigates, but does not eliminate, unwanted variability in the comparator. Active comparator studies require large sample sizes to detect modest differences in efficacy. Also, unrecognized deleterious effects of an unproven comparator treatment may convolute evaluation of efficacy of the experimental treatment. A variation of this approach is comparison of 2 empiric therapies to evaluate superiority (figure 2D).

Outcome measures. Three study endpoints are potential measures of attack frequency or severity (table 3):

**Time to first on-study event.** Ending the randomized phase at the first on-study attack mitigates the risk of placebo exposure, allowing subjects access to alternative therapies when treatment fails. Time to first attack is a continuous measure with inherent statistical advantages. Subjects experiencing an attack may be eligible for an open-label investigational drug or other empiric off-protocol treatment, but could no longer contribute to efficacy evaluation. However, withdrawal of patients after a first event sacrifices information about efficacy and safety as compared to assessment of the annualized relapse rate over the entire observational period. Early relapses could result in underestimation of efficacy for agents with delayed effectiveness. To minimize the consequences of delayed effectiveness, a protocol might define a period of initial treatment during which attacks do not qualify as endpoints, either for primary or secondary analysis. Alternatively, all study participants could receive corticosteroids for a defined period based on the pharmacodynamics of the experimental agent to minimize the risk of early relapse before the experimental agent becomes effective. While any combination of treatments could confound the interpretation of trial results, this approach equally distributes any beneficial signal attributable to corticosteroid treatment among subjects in all treatment limbs.

**Annualized attack rate.** This metric, a common primary outcome in MS clinical trials, annualizes net frequency of all attacks. However, variation in the time over which relapse rate is calculated is problematic when dropout rates are high, and when they differ between treatment groups.

**Attack frequency/severity composite.** Worsening in NMO reflects the cumulative result of attack-related disability. Although attacks are generally more severe than in MS, milder attacks may also occur. Heightened awareness of new or worsening symptoms in prospective clinical trials exaggerates the risk of recording events that are not true attacks. For example, in the recent open-label study of eculizumab, 2 on-study events (back pain alone in one subject and a one-line change of visual acuity in a second) were of equivocal significance. Although experience with stratification of attack severity is limited, a simple system for rating optic nerve and spinal cord attacks has been applied to retrospective studies. This strategy is similar to well-accepted clinical scales used in MS and focuses on visual, pyramidal, sensory, and sphincter functions that are most consistently affected in NMO. Theoretically, each NMO attack could be rated based on attack severity at nadir or after a fixed period of recovery (e.g., 1 month), and a composite developed to reflect attack frequency and severity (i.e., weighted composite relapse severity/frequency score = \( \sum \text{attack, } \times \text{severity/time, } i = 1 \rightarrow n \)). Such an integrated attack frequency and severity metric would minimize the impact of mild equivocal events. Alternatively, the distribution of attacks using a categorical prespecified scale (mild, moderate, severe) is compared using nonparametric statistical tests to mitigate concerns surrounding variability of unvalidated severity scoring scales.

A masked adjudication panel empowered to evaluate all relevant data in real time, including clinical, laboratory, and imaging results, to independently confirm attacks and grade their severity might reduce study site-related variation in the application of an attack frequency endpoint. Furthermore, an independent adjudication panel would resolve potential conflicts of site investigators who may be inclined to interpret an equivocal event as an attack to facilitate treatment of the patient. The decisions of the adjudication panel would not influence treatment decisions for the trial subject, but could define the primary endpoint.

**OTHER ISSUES** Phase of study. The phase of clinical trial for an experimental NMO treatment relates to antecedent experience. Safety profiles of repurposed agents are better established than those of new therapies developed specifically for NMO, possibly eliminating the need for early-phase trials. Early-phase clinical trials would be required for de novo drug development; however, given the recent emphasis on drug repurposing, phase II and III clinical trial designs may be most relevant to planned NMO clinical trials:

**Phase IIa exploratory studies.** Phase IIa or target engagement studies are conducted for promising agents for which adequate safety data are available (figure 3). Patients enrolled in these studies typically have active NMO that has failed one or more empiric treatments. Dramatic reductions in attack frequency compared with pretreatment in single-arm, open-label studies (e.g., as revealed in patients treated with rituximab \( n = 8 \)), eculizumab \( n = 14 \), or...
tocolizumab \(n = 7\) may portend favorable results in controlled trials. However, “regression to the mean” may be an alternative explanation, and decline in event rates in placebo groups may exceed the difference between active and control groups. Nonetheless, phase IIa exploratory studies may be useful for screening novel NMO therapies in cohorts of 10 to 15 patients with recently active NMO whose other treatments have failed. If safety criteria are met and most subjects’ treatment has not failed, the agent may be deemed “promising” and considered for phase IIb and III studies. Sensitive biomarkers, if validated, could accelerate drug development as has detection of new MRI lesions for MS.

Phase IIb and III studies. Agents surviving the futility criteria of phase IIa may be examined in subsequent controlled studies to rigorously evaluate efficacy (figure 2, A–D). First-line treatments should be interrogated in relatively unselected populations representative of their intended use. However, second-line treatments, including potent agents with less favorable safety profiles, may be appropriate for severe or refractory NMO cohorts. Because no treatment is currently proven to be efficacious in NMO, superiority rather than noninferiority to the comparator is required.

Washout interval. Washout periods enhance the likelihood that efficacy is attributed to the experimental agent rather than to residual effects of prior treatment. Patients with NMO receiving immunosuppressive drugs may exhibit long-lasting carryover effects. For example, rituximab-induced B-cell depletion may persist for more than 1 year. However, patients are potentially susceptible to relapse during washout periods, negatively impacting safety and enrollment. To address this concern, randomization may need to be stratified based on prior treatment.

Masking. Definitive studies should be double-masked by ensuring that the placebo has similar characteristics (frequency and route of administration, color, etc.) and by providing appropriate concomitant medications to prevent infusion reactions. Where adverse effects result in unavoidable unmasking, an “evaluating” physician may be designated to assess the primary outcome while another physician evaluates safety, adverse effects, and decides whether termination based on safety is necessary, a frequently used strategy in MS trials.

Clinical trial pipeline. Innovative strategies for coordinating NMO clinical trials to evaluate multiple agents concurrently are desirable for phase I or IIa studies (figure 3). In addition, if standardized inclusion criteria and study procedures are adopted, control subjects might be shared among multiple phase IIb and III controlled clinical trials. Theoretically, this strategy might provide a larger comparator, minimize exposure to placebo, and enhance efficient use of scarce patient resources. Moreover, subjects involved in trials of “failed” therapeutic candidates could be redirected into new studies through a streamlined crossover mechanism to facilitate enrollment and accrual. There are many hurdles to be overcome, however. Diagnostic criteria continue to evolve and have led to expansion of NMO disease spectrum. Differences in inclusion and exclusion criteria based on prior treatment and other subject antecedents vary among studies and could affect the feasibility of a common control group. Investigators, industry sponsors, and regulators would need to be willing to adopt standardized inclusion and exclusion criteria and common study procedures across trials to facilitate this initiative. However, it may be cost-effective to all parties to facilitate multiple phase IIa and possibly phase III studies proceeding simultaneously. Thus, an NMO clinical trial pipeline envisions a cooperative approach for reducing risk to patients, optimizing enrollment, offering patients greatest access to the most promising drug candidates, and promoting long-term resources (e.g., generation of clinical information and biological sample resources for investigational studies), ultimately advancing NMO care.
DISCUSSION  NMO presents unique opportunities and challenges for clinical trial design. Among the most difficult aspects of such trials is implementation of placebo-controlled studies, given the potential severity of NMO attacks. Preventing attacks and limiting their sequelae are highest priorities. Trial designs must be acceptable to all stakeholders and protect patient interests, while being informative and robust. Careful definition of attack frequency and severity, and discovery of surrogate efficacy biomarkers should facilitate these goals. Innovative approaches, such as shared placebo groups, may expand and speed the investigational pipeline.

AUTHOR CONTRIBUTIONS
Dr. B. Weinschenker participated in the analysis, interpretation, writing, and critical review of the manuscript for important intellectual content. Mr. G. Barron participated in the critical review of the manuscript for important intellectual content. Ms. J. Behne participated in the critical review of the manuscript for important intellectual content. Dr. J. Bennett participated in the critical review of the manuscript for important intellectual content. Dr. P. Chin participated in the critical review of the manuscript for important intellectual content. Dr. B. Clee participated in the critical review of the manuscript for important intellectual content. Dr. B. Greenberg participated in the critical review of the manuscript for important intellectual content. Dr. S. Higashi participated in the critical review of the manuscript for important intellectual content. Dr. D. Knappertz participated in the critical review of the manuscript for important intellectual content. Dr. M. Levy participated in the critical review of the manuscript for important intellectual content. Ms. A. Melia participated in the critical review of the manuscript for important intellectual content. Dr. J. Palacu participated in the critical review of the manuscript for important intellectual content. Dr. T. Smith participated in the analysis, interpretation, writing, and critical review of the manuscript for important intellectual content. Dr. M. P. Sohrani participated in the critical review of the manuscript for important intellectual content. Dr. S. VanMeter participated in the critical review of the manuscript for important intellectual content. Dr. P. Vilholakad participated in the critical review of the manuscript for important intellectual content. Dr. M. Wasilewski participated in the critical review of the manuscript for important intellectual content. Dr. D. Wingerdurch participated in the critical review of the manuscript for important intellectual content. Dr. M. Yeaman participated in the analysis, interpretation, writing, and critical review of the manuscript for important intellectual content.

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DISCLOSURE
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REFERENCES

1. Lennon VA, Wingerchuk DM, Kryzer TJ, et al. A serum autoantibody marker of neuromyelitis optica: distinction from multiple sclerosis. Lancet 2004;364:2106–2112.

2. Jarius S, Wildemann B. Aquaporin-4 antibodies (NMO-IgG) as a serological marker of neuromyelitis optica: a critical review of the literature. Brain Pathol 2013;23:661–683.

3. Bradl M, Misu T, Takahashi T, et al. Neuromyelitis optica: pathogenicity of patient immunoglobulin in vivo. Ann Neurol 2009;66:630–643.

4. Saadoun S, Waters P, Bell BA, Vincent A, Verkman AS, Richard S. Prione: tolerability, efficacy, and predictors of benefit in neuromyelitis optica. Mult Scler 2013;19:1237–1240.

5. Zhang H, Bennett JL, Verkman AS. Ex vivo spinal cord slice model of neuromyelitis optica reveals novel immunopathogenetic mechanisms. Ann Neurol 2011;70:943–954.

6. Watanebe S, Nakashima I, Misu T, et al. Therapeutic efficacy of plasma exchange in NMO-IgG-positive patients with neuromyelitis optica. Mult Scler 2013;19:1281–1286.

7. Misu T, Fujihara K, Nakashima I, Sato S, Inoyama Y. Intrathecal activation of B cells and RNA from multiple sclerosis. J Neurol Sci 2010;284:29–36.

8. Waters P, Bell BA, Vincenzi A, Verkman AS. AQP4-IgG predicts relapse following longitudinally extensive transverse myelitis. Ann Neurol 2006;59:569–576.

9. Jiao Y, Fryer JP, Lennon VA, et al. Updated estimate of AQP4-IgG serostatus and disability outcome in neuromyelitis optica. Mult Scler 2013;20:1197–1204.

10. Waters PJ, McKeon A, Leite MI, et al. Serologic diagnosis of NMO: a multicenter comparison of aquaporin-4 IgG assays. Neurology 2012;78:665–671.

11. Wingerchuk DM, Hogan camp WF, O’Brien PC, Weinschenker BG. The clinical course of neuromyelitis optica (Devic’s syndrome). Neurology 1999;53:1107–1114.

12. Kiley J, Leite MI, Nakashima I, et al. Prognostic factors and disease course in aquaporin-4 antibody-positive patients with neuromyelitis optica spectrum disorder from the United Kingdom and Japan. Brain 2012;135:1834–1849.

13. Wingerchuk DM, Pittock SJ, Lucchinetti CF, Lennon VA, Weinschenker BG. A secondary progressive clinical course is uncommon in neuromyelitis optica. Neurology 2007;68:603–605.

14. Kimbrough DJ, Fujihara K, Jacob A, et al. Treatment of neuromyelitis optica: review and recommendations. Mult Scler Relat Disord 2012;1:180–187.

15. Trebst C, Berthele A, Jarius S, et al. Diagnosis and treatment of neuromyelitis optica: consensus recommendations of the Neuromyelitis Optica Study Group [in German]. Nervenarzt 2011;82:768–777.

16. Sellner J, Boggild M, Clenet M, et al. EFNS guidelines on diagnosis and management of neuromyelitis optica. Eur J Neurol 2010;17:1019–1032.

17. Tradrantip L, Zhang H, Saadoun S, et al. Anti-aquaporin-4 monoclonal antibody blocker therapy for neuromyelitis optica. Ann Neurol 2012;71:314–322.

18. Cree BAC, Lamb S, Morgan K, Chen A, Waubant E, Genain C. An open label study of the effects of rituximab in neuromyelitis optica. Neurology 2008;65:1443–1448.

19. Jacob A, Weinschenker BG, Violich I, et al. Treatment of neuromyelitis optica with rituximab: retrospective analysis of 25 patients. Arch Neurol 2008;65:1443–1448.

20. Pittock SJ, Lennon VA, McKeon A, et al. Eculizumab in AQP4-IgG-positive relapsing neuromyelitis optica spectrum disorders: an open-label pilot study. Lancet Neurol 2013;12:554–562.

21. Kieseier B, Stuve O, Dehmel T, et al. Disease amelioration with tocilizumab in a treatment-resistant patient with neuromyelitis optica: implication for cellular immune responses. JAMA Neurol 2013;70:390–393.

22. Ayzenberg I, Kleiter I, Schroder A, et al. Interleukin 6 receptor blockade in patients with neuromyelitis optica nonresponsive to anti-CD20 therapy. JAMA Neurol 2013;70:394–397.

23. Araki M, Matsuoka T, Miyamoto K, et al. Efficacy of the anti-IL-6 receptor antibody tocilizumab in neuromyelitis optica: a pilot study. Neurology 2014;82:1302–1306.

24. Costanzi C, Mariello M, Lucchinetti CF, et al. Azathioprine: tolerability, efficacy, and predictors of benefit in neuromyelitis optica. Neurology 2011;77:659–666.

25. Jacob A, Mariello M, Weinschenker BG, et al. Treatment of neuromyelitis optica with mycophenolate mofetil: retrospective analysis of 24 patients. Arch Neurol 2009;66:1228–1313.

26. Echt DS, Liebson PR, Mitchell LB, et al. Mortality and morbidity in patients receiving eneainide, flecainide, or placebo. N Engl J Med 1991;324:781–788.

27. Hofmeyr GJ, Gulmezoglu AM, Novikova N, Lawrie TA. Postpartum misoprostol for preventing maternal mortality and morbidity. Cochrane Database Syst Rev 2013;7:CD008982.

28. Engebretsen O, Edvardsen H, Løkkevik E, et al. Gefitinib in combination with weekly docetaxel in patients with metastatic breast cancer caused unexpected toxicity: results from a randomized phase II clinical trial. ISRN Oncol 2012;2012:716789.

29. Hinson SR, McKeon A, Fryer JP, Apiwatthanakul M, Lennon VA, Pittock SJ. Prediction of neuromyelitis optica attack severity by quantitation of complement-mediated injury to aquaporin-4-expressing cells. Arch Neurol 2009;66:1164–1167.

30. Ishizu T, Oseogawa M, Mei FJ, et al. Intrathecal activation of the IL-17/IL-8 axis in opticospinal multiple sclerosis. Brain 2005;128:988–1002.

31. Uzawa A, Mori M, Ara K, et al. Cytokine and chemokine profiles in neuromyelitis optica: significance of interleukin-6. Mult Scler 2010;16:1443–1452.

32. Uzawa A, Mori M, Sato Y, Manuda S, Kawabara S. CSF interleukin-6 level predicts recovery from neuromyelitis optica relapse. J Neurol Neurosurg Psychiatry 2012;83:339–340.

33. Chihara N, Aranami T, Sato W, et al. Interleukin 6 signaling promotes anti-aquaporin 4 autoantibody production from plasma cells in neuromyelitis optica. Proc Natl Acad Sci USA 2011;108:3701–3706.
35. Jarius S, Aboul-Enein F, Waters P, et al. Antibody to aquaporin-4 in the long-term course of neuromyelitis optica. Brain 2008;131:3072–3080.

36. Takahashi T, Fujihara K, Nakashima I, et al. Anti-aquaporin-4 antibody is involved in the pathogenesis of NMO: a study on antibody titre. Brain 2007;130:1235–1243.

37. Takano R, Misu T, Takahashi T, Sato S, Fujihara K, Inuyama Y. Astrocytic damage is far more severe than demyelination in NMO: a clinical CSF biomarker study. Neurology 2010;75:208–216.

38. Green AJ, Cree BA. Distinctive retinal nerve fibre layer and vascular changes in neuromyelitis optica following optic neuritis. J Neurol Neurosurg Psychiatry 2009;80:1002–1005.

39. Magana SM, Keegan BM, Weinshenker BG, et al. Beneficial plasma exchange response in central nervous system inflammatory demyelination. Arch Neurol 2011;68:870–878.

40. Temple RJ; Center for Drug Evaluation and Research, US Food and Drug Administration. Enrichment strategies for clinical trials. 2013. Available at: http://www.fda.gov/downloads/Drugs/UCM345394.pdf. Accessed August 1, 2014.

41. Beck RW, Cleary PA. Optic neuritis treatment trial: one-year follow-up results [see comments]. Arch Ophthalmol 1993;111:773–775.

42. Kinkel R. Methylprednisolone. In: Rudick R, Goodkin D, editors. Multiple Sclerosis Therapeutics. London: Martin Dunitz; 1999:349–370.

43. Weinshenker BG, O’Brien P, Petterson T, et al. A randomized trial of plasma exchange in acute CNS inflammatory demyelinating disease. Ann Neurol 1999;46:878–886.

44. Varrin-Doyer M, Spencer CM, Schuyle-Topphoff U, et al. Aquaporin 4-specific T cells in neuromyelitis optica exhibit a Th17 bias and recognize clostridium ABC transporter. Ann Neurol 2012;72:53–64.

45. Collongues N, Cabre P, Marignier R, et al. A benign form of neuromyelitis optica: does it exist? Arch Neurol 2011;68:918–924.

46. Kurtzke JF. Rating neurological impairment in multiple sclerosis: an Expanded Disability Status Scale (EDSS). Neurology 1983;33:1444–1452.

47. Spector R, Park GD. Regression to the mean: a potential source of error in clinical pharmacological studies. Drug Intell Clin Pharm 1985;19:916–919.

48. Bizzoco E, Lolli F, Repice AM, et al. Prevalence of neuromyelitis optica spectrum disorder and phenotype distribution. J Neurol 2009;256:1891–1898.

49. Asgari N, Lillevang ST, Skejoe HPB, Falah M, Stenager E, Kyrk KO. A population-based study of neuromyelitis optica in Caucasians. Neurology 2011;76:1589–1595.

50. Sirirho S, Nakashima I, Takahashi T, Fujihara K, Prayoonwiwat N. AQP4 antibody-positive Thai cases: clinical features and diagnostic problems. Neurology 2011;77:827–834.

51. Van Herle K, Behne JM, Van Herle A, Blaschke TF, Smith TJ, Yeaman MR. Integrative continuum: accelerating therapeutic advances in rare autoimmune diseases. Annu Rev Pharmacol Toxicol 2012;52:523–547.

52. Verkman AS, Ratelade J, Rossi A, Zhang H, Tradtrantip L. Aquaporin-4: orthogonal array assembly, CNS functions, and role in neuromyelitis optica. Acta Pharmacol Sin 2011;32:702–710.

53. Saadoun S, Waters P, MacDonald C, et al. Neutrophil protease inhibition reduces neuromyelitis optica-immunoglobulin G-induced damage in mouse brain. Ann Neurol 2012;71:323–333.

54. Zhang H, Verkman AS. Eosinophil pathogenicity mechanisms and therapeutics in neuromyelitis optica. J Clin Invest 2013;123:2306–2316.

55. Axtell RC, Raman C, Steinman L. Interferon-beta exacerbates Th17-mediated inflammatory disease. Trends Immunol 2011;32:272–277.

56. Matthews L, Marasco R, Jenkinson M, et al. Distinction of seropositive NMO spectrum disorder and MS brain lesion distribution. Neurology 2013;80:1330–1337.
Challenges and opportunities in designing clinical trials for neuromyelitis optica
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