SASRINTM, a Versatile Tool in Peptide Synthesis and Solid-Phase Organic Chemistry

Monika Mergler*, Jacques Gosteli, Peter Grogg, Rolf Nyfeler, and Rudolf Tanner
SANDMEYER Prize Winners 1998

Abstract. The synthesis of the 4-(3-methoxy-4-(hydroxymethyl)phenoxymethyl) derivative of polystyrene-co-divinylbenzene (SASRINTM), its derivatives and its application in solid-phase peptide synthesis will be briefly reviewed in this paper. Solid-phase synthesis of fully protected peptide fragments may be the most important, but by far not the only application, as cleavage with nucleophiles or solid-phase synthesis on SASRIN derivatives will yield a range of useful C-terminally modified peptides, such as peptide hydrazides or p-nitroanilides.

1. Introduction

Synthetic bioactive peptides have found wide application in biology, biochemistry, medicine and related areas. Such compounds are made available by two principally different synthetic methods [1]: either by 'classical' convergent synthesis in solution, which is rather time-consuming, or by stepwise solid-phase peptide synthesis (SPPS) employing an insoluble carrier, a more recent development by MERRIFIELD [2]. Even though peptides can be obtained far more rapidly by SPPS, the 'classical' solution synthesis still has its merits such as concomitant isolation, characterization and purification of intermediates, whereas in stepwise SPPS these operations are restricted to the final product. 'Convergent SPPS' [3] combines the advantages of both methods: fully protected peptide fragments obtained by SPPS on suitable resin derivatives followed by characterization and purification are coupled either to fragments in solution [4], or to resin-bound fragments [5]. Thus, crude products of better quality can be obtained, as usually side products stemming from incomplete fragment coupling are removed more easily than deletion sequences (i.e., fragments lacking merely a single amino acid) accumulated during stepwise SPPS. Fully protected peptide fragments can be obtained by SPPS only if the side-chain ('permanent') protecting groups are left intact during cleavage from the resin. Based on the work of SHEPPARD et al. [6], we at BACHEM developed and successfully marketed the Super Acid-Sensitive Resin (SASRINTM, 2-methoxy-4-alkoxybenzyl-alcohol resin 2) [7][8]. The award of the SANDMEYER prize occasioned the writing of the present short and inevitably incomplete review of SASRIN's manifold applications, including part of our recent work.

Initially, SASRIN was conceived for the synthesis of fully tert-butyl-type-protected fragments using temporary, base-labile Nα-[9H-fluoren-9-yl]methoxy-carbonyl (Fmoc) protection. Cleavage from SASRIN merely requires short repetitive treatments with 0.5 to I% trifluoroacetic acid (TFA) in methylene chloride (DCM) [8][9]. SASRIN esters turned out to be sufficiently (but not excessively) acid-labile, so that the more acid-sensitive moieties Lys(Boc), Tyr(Bu) and His(Trt) are left intact during cleavage, whereas Fmoc amino acids and hydroxybenzotriazole (HOBT) do not cause premature cleavage under normal coupling conditions. Naturally, SASRIN can be used in stepwise SPPS employing Fmoc/Bu-protection as well. Even an additional advantage is gained: samples can be cleaved rapidly at any stage of the synthesis for monitoring purposes. By characterizing these samples an unambiguous batch-documentation system is obtained, which showed its value in solving synthetic problems and optimizing large-scale syntheses. The completed fully protected peptide obtained from SASRIN can be purified before cleaving the permanent protecting groups or, if...
desired, be further modified in solution, e.g., by iodine treatment to cleave Cys(Acm) and to form an S-S-bridge. Total deblocking of a SASRIN-bound peptide can be performed in two stages: i) cleavage from the resin with 0.5–1% TFA in DCM, and ii) removal of side-chain protecting groups in solution, thus facilitating the fine-tuning of cleavage conditions. A two-stage deprotection is recommended when cleaving acid-sensitive peptides such as O-sulfated peptides.

SASRIN and SASRIN derivatives have found many other applications including use in solid-phase organic synthesis since its development in 1986; some of them will be dealt with below.

2. Synthesis of SASRINTM and Derivatives Thereof

As can be taken from Scheme 1, the SASRIN linker is obtained by Viilsmeier formalization of 3-methoxyphenol [6] followed by NaBH₄ reduction [7]. The Na-phenolate of the linker 1 is reacted with chloromethyl polystyrene. The conversion is nearly complete as only traces of chloromethyl groups (which enhances nucleophilic displacement) are present on the polystyrene. The reaction is followed by the addition of NaOMe in DMA, r.t. [8].

Resin 2 (i.e., SASRIN) is loaded with a Fmoc/tBu-protected amino acid. The conditions of esterification had to be optimized to get a high load whilst keeping concomitant racemization low [9] (cf. Scheme 1). But even then, several amino acid derivatives, especially derivatives of Cys and His, could only be esterified at the risk of considerable losses of optical purity, as these compounds racemize too rapidly upon activation. Hence, an alternative coupling method had to be found. Protected amino acids can be linked to Merrifield resin (i.e., chloromethylated crosslinked polystyrene) via nucleophilic substitution, excellent results with virtually no concomitant racemization are obtained by employing their cesium salts [10]. Fortunately, SASRIN could be readily converted into the corresponding chloride 3 [12] by treatment with triphenylphosphine dichloride (generated in situ from triphenylphosphine and tetrachloromethane [13]). The chloride 3 reacts with the dry cesium salt of the desired Fmoc-amino acid and sodium iodide (which enhances nucleophilic displacement) to yield the SASRIN derivative. High conversions and, as expected, minimal racemization could be obtained [14]. Up to now, a broad range of Fmoc-amino acid derivatives of SASRIN could be made commercially available including side-chain linked Asp and Glu derivatives. The chloride 3 will also alkylate other nucleophiles, such as amines, leading to rather acid-labile derivatives [15].

Scheme 1. Synthesis of SASRIN and Loading with Protected Amino Acids

3. Synthesis of Peptides Using SASRINTM

Except for the cleavage procedure, all synthetic protocols developed for Wang resin [16][17] can be applied to SASRIN, likewise the equipment for manual and automated SPPS. 'Maximum protection' is recommended, the following commonly used side-chain-protecting groups will be left intact during acidolytic cleavage from SASRIN (if properly performed): Trt, Mtt, (Asn, Gln); OrBu (Asp, Glu); Pmc (Arg); Trt, Acm (Cys); Trt (His); Boc (Lys, Trp); rBu (Ser, Thr, Tyr) and Boc or Fmoc for the N-terminus. When synthesizing peptides with a C-terminal Pro, the Nα-deblocking at the dipeptide stage will lead to rapid diketopiperazine (piperazine-2,5-dione) formation and concomitant cleavage, i.e., loss of load, which can be circumvented by coupling a Fmoc dipeptide at the risk of racemization.

SASRIN lends itself especially to the synthesis of acid-sensitive peptide derivatives such as O-sulfated or O-glycosylated peptides. Under certain precautions, the hydroxy amino acids Tyr, Thr and Ser can be coupled without side-chain protection. Then, the following couplings must proceed unambiguously generating only amide bonds, otherwise selectively cleavable O-protecting groups such as trialkylsilyl or allyl have to be chosen. Allyl
allyloxy carbonyl side-chain-protecting groups could be cleaved as described in [18] leaving the peptide-resin bond intact. For O-sulfation of Tyr, the otherwise fully protected resin-bound peptide is treated with a large excess of sulfur trioxide/pyridine complex in dry DMF/pyridine 2:1 followed by thorough washing which includes treatment with diluted aqueous sodium carbonate in DMF to obtain the less acid-labile sodium salt. The sulfation should be monitored by analyzing the cleavage products obtained from resin samples; sulfation may be repeated with fresh reagent in case of unsatisfactory conversion. If two or more amino acids have to be modified, better results may be obtained by coupling the corresponding amino acid derivative (e.g., Fmoc-Tyr(SO$_3$Na)OH) if available. This approach will equally benefit from the acid-lability of SASRIN.

Oxidation of resin-bound peptides containing cysteines leading to intramolecular disulfide bridge formation will only succeed if the bridge is also formed smoothly in solution. Moreover, proper swelling and reduced loads are essential prerequisites. When oxidizing with iodine (e.g., 8 equiv. of iodine in DCM/MeOH/water 60:25:4 [19]), a base such as DIPEA has to be added to neutralize hydrogen iodide formed during the oxidation. Scheme 2 summarizes various synthetic strategies for cystine-containing peptides employing SASRIN.

The analysis of samples taken, washed and cleaved turned out to be the only reliable method of monitoring the synthesis of $\Psi$-[CH$_2$NH] pseudopeptides, i.e., peptides containing reduced peptide bonds, on SASRIN. The $\Psi$-[CH$_2$NH] bond is created via reductive alkylation [20], i.e., the amino terminus is reacted with a Fmoc-aminoaldehyde to form the Schiff base which is reduced with NaBH$_3$CN and a small amount of acetic acid. Then, chain elongation cannot be monitored unambiguously by the color tests applied in standard SPPS [16][17] anymore. Albeit less reactive than the $\alpha$-amino group, the $\Psi$-[CH$_2$NH] group can be acylated, thus precautions have to be taken.

4. Cleavage of Fully Protected Fragments

It cannot be overemphasized that all polar impurities have to be washed out carefully with methylene chloride before starting the cleavage procedure, which only then will proceed rapidly and smoothly. Treatments with 0.5–1% TFA/DCM should be kept short (2–5 min), with rapid filtration and immediate neutralization of the solution, e.g., with pyridine, to follow [3b][8]. Upon first treatment with diluted TFA the donor groups of the peptide such as the amide bonds are protonated, consuming part of the TFA and solubilizing the protected fragment. Thus, additional TFA is needed for rapid cleavage whilst neutralization may suffice to precipitate the fragment. In the course of cleavage, the resin turns deeply violet. If Trp, Met, Cys are present, the color will change less dramatically, if at all, as the 'violet species' resulting from the cleavage will alkylate the indole and thioether moieties, respectively, and thus bind the peptide irreversibly. In this case cleavage yields can be improved by employing Trp(Boc) or adding a suitable cation scavenger, e.g., ethane-1,2-dithiol [21]. Good yields have also been obtained by cleaving the peptide resin with mixtures of hexafluoroisopropanol (HFIP) and DCM [22]. Treatment with HFIP/DCM 1:4 (v/v) for 3 to 5 h or HFIP/DCM 3:7 (v/v) for less than 1 h leaves Tyr(tBu) and Lys(Boc) intact, whereas His(Trt) is partially cleaved. Side-chain alkylation, as described above, may occur as well. The cleavage rate slightly depends on the C-terminal amino acid; the highest rate has been observed with C-terminal Gly [23], the lowest with C-terminal His(Trt). This cleavage procedure is especially suitable for large-scale SPPS and for fragments to be coupled, as neither...
acids nor bases have to be added (and, in the course of work-up, thoroughly removed).

5. Cleavage with Nucleophiles

(Fully) protected short peptides with a modified C-terminus can be obtained from SASRIN by cleavage with nucleophiles. Scheme 3 summarizes the options of this cleavage method. Cleavage rates are significantly influenced by steric hindrance (except for reductive cleavage), i.e., by the type of α-substituent of the C-terminal amino acid as well as by the bulkiness of the attacking nucleophile. The ease of cleavage decreases approximately in the order:

| Gly > Asp(OrBu) ≥ Ser(OrBu) ≥ Phe | > Leu > Pro > Val > Thr(OrBu) ≥ Ile |
| Met | Glu(OrBu) | Tyr(OrBu) |
| Arg(Pmc) | Lys(Boc) | Trp |
| Cys(Acm) | Ala |

Good swelling facilitates the attack of the nucleophile. But if the cleavage rate is low, base-catalyzed C-terminal epimerization may become a problem, especially if the C-terminal amino acid is prone to racemization. So, peptides with a C-terminal Phe may epimerize considerably. Asp(OrBu)-containing peptides are susceptible to side reactions, if treated with bases. Especially sequences such as -Asp(OrBu)-Gly- will yield mixtures even under most carefully controlled conditions, whereas Glu(OrBu) and Asn(Mtt) tend to be less sensitive. Nα-Fmoc is removed under most of the conditions applied, but the N-terminus need not be protected. An unambiguous correlation between the length of the fragment and cleavage yield has not been noted, but in some cases a decreasing rate with increasing length (up to ten amino acids) has been observed. The procedure can be repeated; repetitive shorter treatments with the nucleophile may be more effective than a prolonged cleavage.

The rate of hydrazinolysis with hydrazine hydrate [24] moderately depends on the nature of the solvent: N,N'-dimethylpropyldiene urea > N-methylpyrrolidone ≥ N,N-dimethylacetamide > N,N-dimethylformamide > N,N,N'-tetramethylurea. But except for DMA, these solvents slowly react with hydrazine hydrate. A higher rate, e.g., at cleaving peptides with β-branched C-termini, can be achieved by using anhydrous hydrazine. A fully protected peptide azide is generated from the corresponding hydrazide, and this highly reactive species is coupled in situ to a suitable peptide fragment (or to its own deprotected N-terminus, i.e., cyclization, cf. Scheme 3). Thus, as protected peptide hydrazides and protected ‘free acids’ can be obtained rapidly from the same batch of peptide resin, the most suitable activation/fragment-coupling method can be chosen without additional synthetic work.

Ammonolysis/ammonolysis yielding the fully protected (N-alkyl)-amides [25] can be performed with neat primary N-alkylamine (under pressure in case of ammonia, methylamine, and ethylamine [26]). Fortunately, cleavage with secondary amines proceeds extremely slowly, so that standard Fmoc cleavage by repetitive short treatments with piperidine/DMF 1:4 will cause only negligible losses of load when synthesizing medium-sized peptides (up to 30 amino acids).

Base-catalyzed transesterification yields fully protected peptide esters [27]. Rate and yield can be increased considerably by adding a solvent for proper swelling such as DMA, as the peptide resin does not swell sufficiently in alcohols. Unfortunately, the concomitant racemization of the C-terminal amino acid will also be accelerated (Phe >>= Ala >= Val >= Leu >= Pro). Lithium bromide/1,8-diazabicyclo[5.4.0]-undec-7-ene [28] and potassium cyanide

Scheme 3. Cleavage with Nucleophiles

| Azide coupling | Free N-terminus: head to tail cyclization via azide |
| -------------- | ----------------------------------------------- |
| Fully protected peptide-NHNH₂ | Fully protected peptide-NH₂ |
| Fully protected peptide alcohol | Fully protected peptide-O-SASRIN |
| Fully protected peptide aldehyde | Fully protected peptide-OR' |
| coupling to carrier | further modification |
| e.g. peptide NH₂-NHNH₂ |

i) hydrazine hydrate/DMA 1:9 or 1:4. ii) NH₃, NH₄Cl (or LiCl), under pressure. iii) RNNH₂ (neat if possible). iv) NH₃R', NH₄R' (neat if possible). v) R'OH, DMA, KCN (or LiBr/diaza-bicycloundecene 5:2 (molar ratio). vi) NaBH₄, LiCl, THF (or THF/EtOH). vii) pyridine · SO₃, Et₃N, DMSO.
Scheme 4. Synthesis of Fully Protected Peptide Hydrazides via SASRIN carbazate

\[
\begin{align*}
\text{SASRIN-OH} & \xrightarrow{\text{i}} \text{SASRIN-OCO-N} \xrightarrow{\text{ii}} \text{SASRIN-OCONHNH}_2 \\
& \xrightarrow{\text{iii}} \text{SASRIN-OCONHNHCOCR} \xrightarrow{\text{iv, v}} \text{Prot NH...CONHRCOONHNH}_2
\end{align*}
\]

Prot = protecting group different from Fmoc

i) carbonyldimidazole (10 equiv.), THF. ii) hydrazine hydrate/DMF 1:4. iii) FmocNHCHRCOOH, coupling reagent. iv) standard Fmoc/tBu SPPS, last coupling Prot-NHCHR'COOH. v) 1% TFA/DCM or HFIP/DCM 1:4, -CO₂.

Scheme 5. Synthesis of Peptide p-Nitroanilides 7

\[
\begin{align*}
\text{SASRIN-OH} & \xrightarrow{\text{i}} \text{SASRIN-OCO-N} \xrightarrow{\text{ii}} \text{SASRIN-OCONHNH}_2 \\
& \xrightarrow{\text{iii}} \text{SASRIN-OCONHNHCOCR} \xrightarrow{\text{iv, v}} \text{Prot NH...CONHRCOONHNH}_2 \\
& \xrightarrow{\text{vi}} \text{Boc NH...CONHRCOONHNH}_2 \xrightarrow{\text{vii}} \text{Boc NH...CONHRCOONHNH}_2 \\
& \xrightarrow{\text{viii}} \text{Boc NH...CONHRCOONHNH}_2 \xrightarrow{\text{viii}} \text{Boc NH...CONHRCOONHNH}_2
\end{align*}
\]

i) p-nitrophenyl isocyanate, THF/DMF 1:1. ii) 2M Sn Cl₂ : 2 H₂O, DMF, 8 h. iii) FmocNHCHRCOOH, O-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphato, DiPEA or collidine [40], DMF. iv) standard Fmoc/tBu SPPS, last coupling Boc NHCHR'COOH. v) 1% TFA/DCM, or HFIP/DCM 1:4, -CO₂. vi) deblocking, e.g., 95% eq. TFA. vii) NaB0₃ · 4 H₂O (10 equiv.), AcOH, 8–16 h.

[29] proved to be the most effective catalysts of transesterification, thus, cleavage with the former catalysts proceeds more rapidly, whereas removal of the latter is more convenient.

The ease of reductive cleavage does not depend on the nature of the C-terminal amino acid, Ile reacts as smoothly as Gly [30]. The choice of the lithium salt (chloride, bromide) and the composition of the solvent, i.e., the amount of ethanol added to the THF, determine the reactivity of the system, which has to be optimized to suppress side reactions. Special care has to be taken to avoid reductive cleavage of peptide bonds if the peptide contains Pro or N-alkylamino acids, as the tertiary amide bond is quite susceptible towards reduction. Asp(OrBu) may be slowly reduced to yield homoserine, whereas Glu(OrBu) is more resistant to reduction. Cleavage yields may decrease with increasing peptide length. Fully protected peptide alcohols can be oxidized to yield the corresponding aldehydes, e.g., with sulfur trioxide/pyridine complex, dimethylsulfoxide and triethylamine (Parikh-Doering method), whereas the attempt to obtain these compounds directly by reductive cleavage with disobutyl aluminum hydride at low temperature failed.

6. SASRIN™ Derivatives

The scope of applications of SASRIN can even be further extended by employing reactive derivatives such as 4 (see Scheme 4) [31]. The carbazate 5 (available from 4 and hydrazine hydrate) has been used for the synthesis of fully protected peptide hydrazides, as the cleavage conditions do not differ from those described in Sect. 4. SASRIN carbazate lends itself especially to the synthesis of protected hydrazides containing Asp(OrBu), C-terminal Pro (no formation of diketopiperazine), or a β-branched C-terminus.

Peptide p-nitroanilides which have found wide application as chromogenic enzyme substrates can be rapidly obtained by SPPS on (p-aminophenyl)aminocarbonyl oxymethyl polystyrene followed by oxidation of the peptide p-aminoanilide cleaved from the resin [32]. But, as 4 reacted only sluggishly with weak bases such as 4-phenylenediamine, a different synthetic pathway had to be found to obtain the p-aminoanilide derivative of SASRIN 6 [33]. The analogous derivative of hydroxymethylated polystyrene has already been synthesized by a method not applicable to SASRIN [32]. The synthesis of 6 avoiding bifunctional reagents and its use is outlined in Scheme 5. Unexpectedly, the nitro derivative could be smoothly reduced with stannous chloride without premature cleavage from the resin.

Burdick’s resin [32] and the SASRIN derivative have been employed for the synthesis of peptide p-aminoanilides to be oxidized in solution to yield the peptide p-nitroanilides 7, compounds having gained importance as enzyme substrates. Again, the mild cleavage procedure (1% TFA/DCM works as well as HFIP/DCM) turns out to be an additional advantage. Trp(Boc)-containing, protected peptide p-nitroanilides could be obtained by perborate oxidation, whereas the unprotected indole moiety would have been attacked.
N-SASRfN-yl ethylamine can be obtained smoothly by treating 3 with ethylamine/DMF 1:2, Fmoc/Bu SPPS, followed by cleavage with 95% TFA. Yields of the deprotected peptide N-ethylamide [34] for the N-ethylamide derivative of SASRfN is considerably less acid-sensitive than the SASRfN esters (cleavage of N-unsubstituted amides from the corresponding SASRfN derivative proved even more difficult, only low yields of peptide amides were obtained, e.g., with TFA/thioanisol 95:5 [35]).

The synthesis of acid-labile SASRfN ethers has not been tackled yet, whereas the less acid-sensitive SASRfN thiocethers, e.g., the derivative of N-Fmoc-cysteamine, can be rapidly obtained by reacting SASRfN chloride 3 with thiols and a tertiary amine.

7. Conclusions and Outlook

In this review we have demonstrated the versatility of SASRfN and several derivatives thereof. Besides its standard applications in solid-phase peptide synthesis, such as synthesis of fully protected peptide fragments, a broad range of peptide derivatives can be obtained either by variation of the cleavage method or by employing SASRfN derivatives such as ρ-(SASRfN-oxy carbonylamino)aniline 6. SASRfN found further applications in combinatorial chemistry. So SASRfN was used as a carrier in solid-phase organic synthesis, e.g., of ρ-substituted benzoic acids via Suzuki coupling on solid phase [36], of quinazoline-2,4-diones [37], and of β-lactams [38]; solid phase reactions were monitored by recording the FTIR and Raman spectra of a single bead [39]. The development of further SASRfN derivatives will be stimulated certainly not only by the needs of peptide synthesis but also by the demands of combinatorial chemistry.

Received: November 17, 1998

[1] E.g., J. Jones, 'The Chemical Synthesis of Peptides', Clarendon Press, Oxford 1991.
[2] R.B. Merrifield, J. Am. Chem. Soc. 1963, 85, 2149.
[3] a) P. Lloyd-Williams, F. Albericio, E. Giralt, Tetrahedron 1993, 49, 11065; b) F. Albericio, P. Lloyd-Williams, E. Giralt, Methods Enzymol. 1997, 289, 313.
[4] E.g., A.M. Felix, Z. Zhao, T. Lambros, M. Ahmad, W. Liu, A. Daniewski, J. Michalowski, E.P. Heimer, J. Peptide Res. 1998, 52, 153.
[5] E.g., D. Gatos, S. Patrjinaakou, O. Hatzi, K. Barlos, Lett. Pept. Sci. 1997, 4, 177.
[6] R.C. Sheppard, B.J. Williams, Int. J. Peptide Protein Res. 1982, 20, 451.
[7] M. Mergler, J. Gosteli, P. Grogg, R. Tanner, Tetrahedron Lett. 1988, 29, 4005; M. Mergler, J. Gosteli, P. Grogg, EP 0292729 (to Bachem AG), 1992; M. Mergler, J. Gosteli, P. Grogg, US Pat. 4,831,084 (to Bachem AG), 1989; M. Mergler, J. Gosteli, P. Grogg, US Pat. 4,914,151 (to Bachem AG), 1990.
[8] For a similar anchor and an improved cleavage protocol see: A. Flörsheimer, B. Riniker, in 'Peptides 1990', Proc. 21st European Peptide Symposium, Eds. E. Giralt, D. Andreu, Escom, Leiden 1992, 131; P. Riniker, A. Flörsheimer, H. Fretz, P. Sieber, B. Kamber, Tetrahedron 1993, 49, 9307; see also [3b] for an excellent cleavage protocol.
[9] M. Mergler, R. Nyfeler, R. Tanner, J. Gosteli, P. Grogg, Tetrahedron Lett. 1988, 29, 4009.
[10] Cf. G. Lu, S. Mojsov, J.P. Tum, R.B. Merrifield, J. Org. Chem. 1981, 46, 3433. For an alternative synthesis see A.R. Katritzky, D. Toader, K. Watson, J.S. Kiely, Tetrahedron Lett. 1997, 38, 7849.
[11] D.F. Gisun, Helv. Chim. Acta 1973, 56, 1476.
[12] M. Mergler, R. Nyfeler, J. Gosteli, Tetrahedron Lett. 1989, 30, 6741. Recently, another mild method to convert SASRfN into SASRfN chloride has been described by: D.A. Nugel, D.A. Wacker, G.A. Nemeth, Tetrahedron Lett. 1997, 38, 5789. When treating SASRfN with triphenylphosphate and tetrabromomethane (or triphenylphosphine dibromide), the desired SASRfN bromide is obtained, but side reactions such as benzyl ether cleavage will reduce the quality of the resin; cf. A.M. Fivush, T.M. Willson, Tetrahedron Lett. 1997, 38, 7151.
[13] R. Appel, Angew. Chem., Int. Ed. 1975, 14, 801.
[14] M. Mergler, R. Nyfeler, J. Gosteli, R. Tanner, Tetrahedron Lett. 1989, 30, 6745.
[15] Cf. K. Ngu, D.V. Patel, Tetrahedron Lett. 1997, 38, 973.
[16] a) i.e., ρ-Alkoxybenzyl alcohol resin, see: S.S. Wang, J. Am. Chem. Soc. 1973, 95, 1328; b) G.B. Fields, R.L. Noble, Int. J. Peptide Protein Res. 1990, 35, 161.
[17] M. Mergler in 'Peptide Synthesis Protocols', Eds. M.W. Pennington, B.M. Dunn, Humana Press, Totowa NJ, 1994, p. 287–301.
[18] A. Loettf, H.X. Zhang, Int. J. Peptide Protein Res. 1993, 42, 346.
[19] B. Kamber, A. Hartmann, K. Eislter, B. Riniker, H. Rink, P. Sieber, W. Rittel, Helv. Chim. Acta 1980, 63, 899.
[20] D.H. Coy, S.J. Hocart, Y. Sasaki, Tetrahedron 1998, 44, 835.
[21] B. Kamber, B. Riniker, Proc. 12th Amer. Pept. Symp. 1991, Eds. J.A. Smith, J.E. Rivier, Escom, Leiden 1992, 551.
[22] B. Kamber, R. Nyfeler, Proc. 2nd Int. Symp. Solid Phase Synthesis 1991, Ed. R. Epton, Intercept, Andover 1992, 429.
[23] M. Mergler, R. Nyfeler, Proc. 4th Int. Symp. Solid Phase Synthesis 1995, Ed. R. Epton, Mayflower Scientific, Birmingham 1996, 481.
[24] M. Mergler, R. Nyfeler, Proc. 3rd Int. Symp. Solid Phase Synthesis 1993, Ed. R. Epton, Mayflower Worldwide, Birmingham 1994, 599.
[25] D.J. Burdick, M.E. Struble, J.P. Burnier, Tetrahedron Lett. 1993, 34, 2589.
[26] M. Mergler, F. Dick, J. Gosteli, R. Nyfeler, Poster presented at the 8th Int. Conf. Polymer-Based Technology, Maile Hachamisha 1998.
[27] Cf. D. Sarantakis, J.J. Bicksler, Tetrahedron Lett. 1997, 38, 7325.
[28] M. Mergler, unpublished work. The N-substituted derivatives (amines and amides) are currently under investigation.
[29] J.W. Guiles, S.G. Johnson, W.V. Murray, J. Org. Chem. 1996, 61, 3169.
[30] M.F. Gordeev, H.C. Hui, E.M. Gordon, D.V. Patel, Tetrahedron Lett. 1997, 38, 1729.
[31] B. Ruhlhand, A. Bhandari, E.M. Gordon, M.A. Gallop, J. Am. Chem. Soc. 1996, 118, 253, B. Ruhlhand, A. Bombnin, M.A. Gallop, J. Org. Chem. 1997, 62, 7820.
[32] S.S. Rahman, D.J. Busby, D.C. Lee, J. Org. Chem. 1998, 63, 6196.
[33] L.A. Carpino, A. El-Faham, C.A. Minor, F. Albericio, J. Chem. Soc., Chem. Commun. 1994, 201; L.A. Carpino, A. El-Faham, J. Org. Chem. 1994, 59, 695.