Liver X Receptor Modulates Diabetic Retinopathy Outcome in a Mouse Model of Streptozotocin-Induced Diabetes

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Endothelial progenitor cells (EPCs), critical for mediating vascular repair, are dysfunctional in a hyperglycemic and/or hypercholesterolemic environment. Their dysfunction contributes to the progression of diabetic macro- and microvascular complications. Activation of “cholesterol-sensing” nuclear receptors, the liver X receptors (LXRα/LXRβ), protects against atherosclerosis by transcriptional regulation of genes important in promoting cholesterol efflux and inhibiting inflammation. We hypothesized that LXR activation with a synthetic ligand would correct diabetes-induced EPC dysfunction and improve diabetic retinopathy. Studies were performed in streptozotocin (STZ)-injected DBA/2J mice fed a high-fat Western diet (DBA/STZ/WD) and treated with the LXR agonist GW3965 and in LXRα+/−, LXRβ+/−, and LXRαβ+/− mice. Retinas were evaluated for number of acellular capillaries and glial fibrillary acidic protein (GFAP) immunoreactivity. Bone marrow EPCs were analyzed for migratory function and gene expression. Compared with vehicle-treated DBA/STZ/WD mice, GW3965 treated mice showed fewer acellular capillaries and reduced GFAP expression. These mice also exhibited enhanced EPC migration and restoration of inflammatory and oxidative stress genes toward nondiabetic levels. LXRα−/−, LXRβ−/−, and LXRαβ−/− mice developed acellular capillaries and EPC dysfunction similar to the DBA/STZ/WD mice. These studies support a key role for LXR in retinal and bone marrow progenitor dysfunction associated with type 1 diabetes. LXR agonists may represent promising pharmacologic targets for correcting retinopathy and EPC dysfunction.

The liver X receptors (LXRs) are widely known for their important roles in modulating whole-body cholesterol homeostasis (1), LXRα (NR1H3) and LXRβ (NR1H2) belong to the nuclear receptor superfamily of ligand-activated transcription factors. LXRβ (NR1H2) is ubiquitously expressed, whereas LXRα (NR1H3) expression is highest in the liver, kidney, intestine, and adrenal gland. LXRs initiate a feedback response to elevated cholesterol levels by increasing the transcription of genes involved in cholesterol catabolism and efflux when activated by their endogenous ligands (oxysterols) (2). Direct LXR target genes include the ATP-binding cassette (ABC) transporters, ABCA1 and ABCG1, which govern cellular efflux of cholesterol from macrophages, as well as apolipoprotein E (ApoE) (1). LXR is also a key modulator of systemic inflammation (3). Because LXRs promote reverse cholesterol transport and suppress inflammatory responses, activation of LXR has been considered as a prime drug target for the treatment of the macrovascular disease atherosclerosis (4). Currently available synthetic LXR ligands (i.e., GW3965) have been shown to be effective at reducing plaque formation in animal models of atherosclerosis (5).

Inflammation and oxidative stress are central to the pathogenesis of diabetic retinopathy, a common microvascular complication of diabetes. Studies have shown inflammatory proteins are increased in the retinas or vitreous humor of diabetic animals and patients (6–8). Moreover, blocking inflammation and oxidative stress in diabetic animals prevents the development of lesions characteristic of retinopathy (9). Inflammatory molecules that have been shown to contribute to structural or functional alterations that are characteristic of retinopathy include nuclear factor (NF)-κB, inducible nitric oxide synthase (iNOS), cyclooxygenase (COX), intracellular adhesion molecule (ICAM), 5-lipoxygenase, interleukin (IL)-1β, and tumor necrosis factor (TNF)-α (10,11), and may represent good therapeutic targets. Likewise, the renin-angiotensin system (RAS) contains an anti-inflammatory and vascular protective axis, activated through Mas1, the putative receptor for angiotensin (Ang)-1–7, the product of ACE2 (12). Activation of this protective axis has been shown to prevent the development of diabetic retinopathy (13). To date, the effect of LXR activation on the ACE2/Ang-(1–7)/Mas1 pathway is unknown.

Although endothelial dysfunction is central to the pathogenesis of diabetic retinopathy, equally important is the lack of endothelial regeneration and repair. Bone marrow (BM)–derived endothelial progenitor cells (EPCs) are key cells responsible for maintenance and repair of the vasculature. EPCs have been shown to promote revascularization of ischemic tissues (14) and have been used for cell-based therapies to induce vascular repair, specifically for treatment of myocardial infarction (15) and peripheral arterial disease (16). EPCs function by direct incorporation into injured areas and by providing a source of growth factors and cytokines to enhance the growth potential of the resident vasculature. Cluster of differentiation (CD)34+ cells are a commonly used EPC population in humans, whereas...
lineage-negative (Lin⁻) stem-cell antigen-1 (Sca-1⁺) and c-Kit⁺ double-positive cells are accepted markers for the early hematopoietic stem cell progenitors, which are the source of EPCs that give rise to endothelium and participate in vascular repair in mice (17).

EPC dysfunction has been implicated in the development of the vasodegenerative phase of diabetic retinopathy (18), and numbers of circulating EPCs are low in individuals with nonproliferative diabetic retinopathy (19). In a diabetic state, BM-derived progenitors are dysfunctional, producing fewer endothelial cells with reduced proliferative, migratory, integrative, and vasculogenic potential (20). Differentiation of BM-derived progenitor cells can be shifted away from endothelial precursors toward a proinflammatory phenotype that express a deleterious repertoire of cytokines (21).

Enhanced oxidative stress in diabetes contributes to progenitor cell dysfunction, leading to cellular and replicative senescence (22). The main components of the superoxide-generating NADPH oxidase enzyme include the membrane-associated cytochrome b558, the catalytic gp91phox (Nox2), the regulatory p22phox subunit, and cytosolic components (23). The main isoform of Nox present in BM-derived progenitors is Nox2. Increased activity of NADPH oxidase results in reduced migratory function in diabetic progenitor cells (24). Activation of the protective axis of the RAS by Ang-(1–7) via stimulation of the Mas 1 receptor results in reduced NADPH oxidase activity and improved vascular and EPC function (25).

Although there are established links between vascular disease and decreased EPCs, to date, there has been no direct association between LXR expression in EPCs and the presence of micro- or macrovascular complications. Therefore, we asked whether LXR agonists could prevent the progression of diabetic retinopathy and correct diabetes-induced EPC dysfunction in a novel accelerated model of diabetic complications, the DBA/2J mouse treated with streptozotocin (STZ) and fed a high-fat Western diet (DBA/STZ/WD).

**LXR AGONISTS MODULATE RETINOPATHY**

**RESEARCH DESIGN AND METHODS**

**Animals.** All animal procedures were in compliance with the National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals, and with the Association for Research in Vision and Ophthalmology Statement for the Use of Animals in Ophthalmic and Vision Research, and were approved by the institutional animal care and use committee at the University of Colorado, the University of Toronto, and the University of Florida (UF). DBA/2J male mice were purchased from Jackson Laboratories (Bar Harbor, ME). At 8 weeks old, mice were treated for 5 consecutive days with STZ at a concentration of 40 mg/kg of body weight in 2.94% (w/v) sodium citrate (pH 4.5). Mice were fed a diet containing vehicle or the LXR agonist GW3965 (at a daily dose of 10 mg/kg) for 12 additional weeks. All DBA/2J mice were transferred to a Western (high-fat/high-cholesterol) diet (Harlan, TD 88137) for the last 4 weeks of the experiment. The Western diet was used to accelerate the progression of diabetic retinopathy and correct diabetes-induced EPC dysfunction by adding hyperlipidemia to hyperglycemia, as has been previously described (20). Male wild-type (WT) and LXRα⁻/⁻, LXRβ⁻/⁻, and LXRαβ⁻/⁻ mice on a mixed-strain background (C57BL/6J:129SvEv) were maintained ad libitum on a 2016-chow diet (Harlan) in a temperature and light-controlled environment. Experiments were performed when the animals were 5–8 months old.

**Tyrosin digest and quantitation of acellular capillaries in the retina.** We examined the retinas of DBA/WD control mice and DBA/STZ/WD, untreated or treated with GW3965 (n = 6 for each group), and WT, LXRα⁻/⁻, and LXRαβ⁻/⁻ mice (n = 3 for each group) using trypsin digests as previously described (27,28).

**Assessment of inflammatory cell and activated retinal glia.** Selected eyes from control and 20-week STZ-diabetic DBA/WD animals were processed as previously described (28), and the sections were reacted with antibody to glial fibrillary acidic protein (GFAP, BD Biosciences, San Jose, CA) for detection of Muller glia, CD45 (BD Biosciences), or to CD11b (Wako Chemicals) for detection of inflammatory monocytes and ionized calcium binding adaptor molecule 1 (Iba1) (Wako Chemicals) as a marker of microglia, followed by appropriate species-specific fluorescent-conjugated secondary antibodies. Sections were examined and image were captured using epifluorescence. Numbers of positive cells were counted by observers masked to the treatment from at least three random sections per eye. Counts from multiple sections were averaged, and that value was considered as a single sample for that particular eye. Three eyes from each group were assessed in this manner.

**BM progenitor isolation and migration experiments.** The femur and tibia of each mouse were flushed, and progenitors were enriched from BM mononuclear cells with mouse hematopoietic progenitor cell enrichment kits according to the manufacturer's instructions (STEMCELL Technologies Inc., Vancouver, BC, Canada). Progenitors were tested for their migratory ability toward 100 nM/LCWM cell-derived growth factor (SDF)-1α (R&D Systems, Minneapolis, MN) with the QCM Chemotaxis cell migration assay (Chemicon International, Inc., Temecula, CA) according to the manufacturer's instructions.

**Mouse model of oxygen-induced retinopathy (OIR).** C57BL6/J timed-pregnant mice were obtained from Jackson Laboratories and were housed in the UF Animal Care facilities. In the neonatal mouse model of OIR, 7-day-old mice were placed with their nursing dams in a 75% oxygen atmosphere for 5 days. On postnatal day (PND) 12 and continuing through PND 17, mouse pups received twice-daily gavage (10 μL/gavage) of vehicle (0.9% sodium chloride) or GW3965 (0.05 mg/mouse/day). On the fifth day after return to normoxia (PND 17), the animals were euthanatized. The eyes were processed and sections were stained with hematoxylin and eosin (H&E) and analyzed as previously described (29). The efficacy of treatment was calculated as the average percentage of nuclei per section in the eye of the GW3965-treated pups versus the eye of vehicle-treated pups. Each data point represents a minimum of one eye each from six pups.

**Retinal histology in LXR⁻/⁻ mice.** Eyes from WT, LXRα⁻/⁻, LXRβ⁻/⁻, and LXRαβ⁻/⁻ mice (n = 3/group) were enucleated, fixed, and sectioned for H&E staining as previously described (29). Three individuals who were masked to the identity of the H&E sections performed a pathologic examination of the slides.

**Retinal flat mounts and collagen IV staining.** Neural retinas (n = 3/group), consisting of WT, LXRα⁻/⁻, LXRβ⁻/⁻, and LXRαβ⁻/⁻, were processed as previously described and stained with rabbit anticallogen IV (Abcam, San Francisco, CA), followed by goat anti-rabbit IgG conjugate to DyLight 549 (Abcam) to label vasculature (18). Laser scanning confocal microscope (Leica TCS SP2, Leica Microsystems, Buffalo Grove, IL) was used, and six random fields were captured for each retina. Total numbers of junction points of each field and branch length was measured from the compressed images using ImageJ software.

**Real-time quantitative PCR.** Total RNA was extracted from mouse BM or from human CD34⁺ EPCs and reverse transcribed as described (30). Real-time PCR was carried out using TaqMan Gene Expression Assays (Applied Biosystems, Carlsbad, CA; hβ-actin, hNkx2, hIL-1β, hNos2, hMus1) or SYBR green (mABCA1, mABCG1, mCyclophilin, mIL-1β, mLXRa, mLRXβ, mNF-kB, mNos2), both with universal PCR Master Mix (Applied Biosystems). Primer sequences are available upon request. Relative mRNA levels were calculated using the comparative Ct method normalized to cyclophilin for mouse studies and β-actin for human studies.

**TABLE 1**

| Metabolic parameters of DBA/2J mice fed a WD |
|-------------------------------------------|
|               | DBA/WD + veh | DBA/WD + GW3965 | DBA/STZ/WD + veh | DBA/STZ/WD + GW3965 |
| Body weight (g) | 35.3 ± 0.3   | 38.7 ± 0.7*     | 21.6 ± 0.7#     | 20 ± 1#             |
| Plasma glucose (mg/dL) | 231 ± 70   | 269 ± 48        | 727 ± 22#       | 820 ± 16#           |

*P < 0.05 compared with vehicle-treated mice. #P < 0.05 compared with DBA/WD of the same treatment group.*
**Cholesterol content.** The cholesterol content of an isolated neural retina from an individual mouse was determined by liquid chromatography tandem mass spectrometry (LC/MS/MS) after lipid extraction using a modified Folch procedure. Each sample was spiked with 0.35 μg cholesterol-d7 to normalize for extraction efficiency. Dried lipid extracts were resuspended in 100 μL high-performance liquid chromatography-grade methanol before injection of 1 μL to the LC/MS/MS. LC/MS/MS-run parameters used were as previously described (31). The peak area for cholesterol was normalized to that of cholesterol-d7 and then normalized to the total amount of RNA extracted to account for any differences in tissue input.

**Flow cytometry and in vitro treatment of total BM cells with GW3965.** Total BM-derived cells from WT and LXRα/β-/- mice (8 months old) were stained with CD117 (c-Kit, PerCP-Cy5.5-conjugated 2B8; BD Biosciences), 0.25 μg/10⁶ cells and Ly6A/E (Sca-1, PE-Cy7-conjugated D7; BD Biosciences),

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![Image of retinal vasculature with acellular capillaries](image-url)

**FIG. 1.** GW3965 treatment reduces acellular capillaries in the retina of STZ-diabetic mice. **A-D:** Representative images of retinal vasculature from vehicle- (A and C) and GW3965- (B and D) treated DBA/WD (A and B) and DBA/STZ/WD mice (C and D). GW3965 was incorporated into the diet for the duration of the study. The retinas were prepared by trypsin digestion and stained with hematoxylin and periodic acid Schiff. Reduced numbers of acellular capillaries (arrowheads) were observed in retinal vasculature isolated from GW3965-treated DBA/STZ/WD (D) compared with vehicle-treated (C). **E:** Quantitative measurements of acellular capillaries. Data have been plotted as mean ± SEM (n = 6 per group). *P < 0.05 compared with vehicle treated. #P < 0.05 compared with DBA/WD of same treatment group. (A high-quality digital representation of this figure is available in the online issue.)
1 µg/10⁶ cells, followed by LIVE/DEAD Fixable Dead Cell Stain (Invitrogen), 1 µL/10⁶ cells. Samples were fixed with IC Fixation Buffer (eBioscience) and stored at 4°C until analysis. Sample acquisition was performed in a FACS Aria cytometer (BD Biosciences), and analysis was performed using FlowJo (Tree Star). For in vitro treatment experiments, total BM-derived cells from WT and LXRαβ⁻/⁻ mice (5 months old) were cultured for 48 h in culture medium (DMEM + 10% FBS + 1 x Fungizone ≤ 1 µmol/L GW3965) and then collected for RNA extraction and gene expression analysis.

**CD34⁺ EPC isolation and treatment.** CD34⁺ cells were isolated from the peripheral blood of healthy human donors after UF institutional review board approval. The collected blood was layered on top of Ficoll (GE Healthcare, Uppsala), and mononuclear cells were enriched for CD34⁺ using a human progenitor cell enrichment kit (STEMCELL Technologies, Inc.). Cells were suspended in media (Stem Span, STEMCELL Technologies, Inc.) with Stem Span CC100 cytokine cocktail (STEMCELL Technologies, Inc.), treated with GW3965 (0, 0.1, or 1 µmol/L) overnight and used for subsequent experiments.

**Statistical analysis.** A Student t test was used for comparisons between two groups. One-way ANOVA, followed by the Tukey post hoc test, was used for multiple comparisons. All values are expressed as mean ± SEM. A value of P < 0.05 was considered to be statistically significant. Statistical tests were performed using statistics software (SPSS Inc., Chicago, IL, or GraphPad Software, La Jolla, CA).

**RESULTS**

**Chronic treatment of STZ diabetic mice with LXR agonist (GW3965) prevents the development of diabetic retinopathy and reduces inflammatory cells in the retina.** Mice of the DBA/2J strain spontaneously develop ocular abnormalities, including glaucomatous loss of retinal ganglion cells, Müller glia activation, and at 8 and

![FIG. 2. Activated glia and inflammatory monocytes in vehicle-treated and GW3965-treated STZ-diabetic DBA/WD mouse retinas.](image)

A–D: Representative images of GFAP expression in vehicle- (A and C) and GW3965- (B and D) treated DBA/WD (A and B) and DBA/STZ/WD (C and D) retinas. In DBA/STZ/WD mice (12 weeks of hyperglycemia, 4 weeks of WD), GFAP expression was elevated in Müller glial cells in retinas of the vehicle-treated animals (C), whereas the expression was reduced in retinas from GW3965-treated animals (D). E: Quantification of CD11b⁺, CD45⁺, and Iba1⁺ cells shows a significant increase in all three inflammatory cell types in the retinas of 20-week-old DBA/STZ/WD mice. GW3965 returns the numbers of these inflammatory cells to control levels when DBA/STZ/WD mice are fed the LXR agonist. Data have been plotted as mean ± SEM (n = 3 per group). *P < 0.05 compared with DBA/STZ/WD mice. #P < 0.05 compared with DBA/WD of same treatment group. (A high-quality digital representation of this figure is available in the online issue.)
11 months, signs of neoangiogenesis (32). We reasoned that in this model, the superimposition of STZ-induced diabetes and feeding of the high-fat WD would accelerate the vascular phenotype typically associated with diabetes. The metabolic characteristics of these mice are described in Table 1. Although GW3965 has been reported to improve hyperglycemia in a high-fat diet model of diabetes (5), we did not observe a change in glycemic control with our 12-week study (Table 1). Compared with control (Fig. 1A), the STZ-treated mice (Fig. 1C) showed a dramatic increase in acellular capillaries (Fig. 1E). GW3965 treatment resulted in a 50% reduction in the number of acellular capillaries ($P < 0.05$) in the treated group (Fig. 1D) compared with vehicle-treated diabetic mice (Fig. 1C). Nondiabetic mice treated with GW3965 (Fig. 1B) showed no change in the number of acellular capillaries compared with the vehicle-treated nondiabetic mice (Fig. 1A).

To determine whether this change in the number of acellular capillaries was associated with reduced inflammation, GFAP expression in Müller glial cells was assessed in retinas from vehicle- and GW3965-treated DBA/STZ/WD mice (Fig. 2). Nondiabetic DBA/WD mice (Fig. 2A) showed low levels of GFAP expression, as did the GW3965-treated nondiabetic mice (Fig. 2B). In contrast, there was a marked increase in GFAP expression in the DBA/STZ/WD mice (Fig. 2D). GW3965 treatment also reduced diabetes-induced ocular inflammation to baseline levels, as assessed by a significant decrease in CD11b+ cells (Fig. 2E, left panel), CD45+ cells (Fig. 2E, center panel), and Iba1+ microglia (Fig. 2E, right panel) in the DBA/STZ/WD mice.

**LXR agonist GW3965 enhances EPC function in STZ-diabetic mice.** One mechanism to assess the overall function of EPCs is to measure their ability to migrate toward chemoattractants. Treatment with GW3965 resulted in a significant increase ($P < 0.05$) in migration of EPCs toward SDF-1α (100 nmol/L) isolated from WT C57BL/6J mice (Fig. 3A, left black bar). Similarly, treatment with GW3965 resulted in a significant increase ($P < 0.05$) in migration of EPCs toward SDF-1α (100 nmol/L) isolated from WT C57BL/6J mice (Fig. 3A, left black bar). Similarly, treatment with GW3965
resulted in an increase ($P < 0.05$) in migration of the EPCs isolated from DBA/WD mice (Fig. 3A, center black bar) and DBA/STZ/WD mice (Fig. 3A, right black bar) compared with cells isolated from vehicle-treated mice (Fig. 3A, white bars). These data demonstrate that the improved EPC function observed after LXR activation occurs independent of the mouse model used.

To determine whether the functional differences could be accounted for by changes in expression levels of Nox2 (Fig. 3B, left panel), Mas1 (Fig. 3B, center panel) and IL-1β (Fig. 3B, right panel), we analyzed BM-derived cells from each of the DBA/2J treatment groups described above (Fig. 3). Quantitative PCR analysis demonstrated a significant increase in the expression of IL-1β in DBA/STZ/WD mice compared with DBA/WD mice. This increase in STZ-injected mice was significantly attenuated in the diabetic mice treated with GW3965 (Fig. 3B). Likewise, treatment with GW3965 in DBA/STZ/WD mice resulted in a 10-fold reduction in Nox2, the main isoform of Nox expressed in BM cells that is responsible for superoxide generation, while increasing the mRNA expression of the protective Mas1 receptor (Fig. 3B), supporting a direct association between LXR activation and the protective axis of the RAS.

**Supplementation with GW3965 results in a reduction of preretinal neovascularization in the OIR model.**

Rodent models of STZ diabetes do not develop retinal angiogenesis, as is characteristic of advanced human disease. Therefore, we used a well-established model of retinal angiogenesis in mice, the OIR model. In this model, nursing dams and their pups were placed at 75% oxygen on PND 7. The high oxygen level interrupts normal retinal blood vessel development. On PND 12, the mice were reared, because LXR activation in the retina may lead to pathology. This was confirmed, because LXRα−/−, LXRβ−/−, and LXRαβ−/− mice exhibited increased numbers of acellular capillaries compared with WT mice (P < 0.05; Fig. 5B). There were no significant differences in the levels of the inflammatory markers IL-1β and NF-κB in the retinas of LXRαβ−/− mice compared with WT mice (data not shown). However, the expression of Nox2 was increased in the LXRαβ−/− mice (P = 0.05; Fig. 5B), suggesting that increased reactive oxygen species may be present in the retinas of these mice.

Our gain of function studies using the LXR agonist clearly demonstrated protection from the development of retinopathy; thus, we reasoned that the loss of LXR activation in the retina may lead to pathology. This was confirmed, because LXRα−/−, LXRβ−/−, and LXRαβ−/− mice showed repression of both IL-1β and NF-κB, and Nox2 mRNA (Fig. 6C). Furthermore, in vitro LXR agonist treatment of BM-derived cells from WT and LXRαβ−/− mice showed repression of both IL-1β and Nox2 in the WT mice, while having no effect in the LXRαβ−/− mice, as expected (Fig. 6D). No significant change was observed with IL-1β in any of the groups (data not shown).

**LXR agonists modulate retinopathy**

LXRαβ−/− mice have a greater number of acellular capillaries and reduced numbers of reparative EPCs with decreased migratory capacity. Consistent with the role of LXRs in modulating cholesterol content, we found the cholesterol content of the neural retina was increased in the LXRαβ−/− mice compared with WT (P = 0.05; Fig. 5A). Previous studies have shown that LXRαβ−/− mice have elevated intracellular cholesterol levels in numerous tissues, including the liver, adrenal gland, and macrophages (33–35). However, cholesterol and triglyceride levels in the serum are lower in the LXRαβ−/− mice compared with WT mice (36), and the LXRαβ−/− mice are normoglycemic and tend to be more insulin-sensitive compared with WT mice (30). In LXRαβ−/− mice, cholesterol can accumulate in tissues because there is an impaired capacity to efflux the cholesterol back into circulation. We measured the gene expression of two cholesterol transporters important in cholesterol efflux (ABCA1 and ABCG1) using quantitative PCR. ABCA1 and ABCG1 were both strongly repressed in the neural retinas from LXRαβ−/− mice (P < 0.05; Fig. 5B). Our gain of function studies using the LXR agonist clearly demonstrated protection from the development of retinopathy; thus, we reasoned that the loss of LXR activation in the retina may lead to pathology. This was confirmed, because LXRα−/−, LXRβ−/−, and LXRαβ−/− mice exhibited increased numbers of acellular capillaries compared with WT mice (Fig. 5C, D). To exclude the possibility that the LXR knockout mice had intrinsic abnormalities in their retinas, we performed H&E staining of retinal cross-sections of WT, LXRα−/−, LXRβ−/− and LXRαβ−/− mice. We did not observe any difference in retinal thickness or morphology between the experimental groups (data not shown). We also performed retinal flat mounts to examine the integrity and pattern of the retinal vasculature. No differences were detected in the vasculature between groups of mice, as assessed by numbers of junctions and branch lengths (data not shown).

We next assessed whether the LXRαβ−/− mice would have a defect in their EPCs that could be contributing to the development of retinopathy. LXRαβ−/− mice had reduced numbers of Sca-1+/c-Kit+ progenitor cells within their BM compared with WT mice (Fig. 6A). Furthermore, Lin− Sca-1+ progenitor cells isolated from LXRαβ−/− and LXRαβ−/− mice exhibited significantly reduced migration to SDF-1α compared with Lin− Sca-1+ cells isolated from WT mice (Fig. 6B). Gene expression analyses from total BM of WT versus LXRαβ−/− mice demonstrated that LXRαβ−/− cells had significantly reduced LXRα and LXRβ mRNA, as expected, while having increased levels of NF-κB and IL-1β, and Nox2 mRNA (Fig. 6C). Furthermore, in vitro LXR agonist treatment of BM-derived cells from WT and LXRαβ−/− mice showed repression of both NF-κB and Nox2 in the WT mice, while having no effect in the LXRαβ−/− mice, as expected (Fig. 6D). No significant change was observed with IL-1β in any of the groups (data not shown).

LXR agonist reduces a key marker of oxidative stress in human CD34+ EPCs. Human CD34+ EPCs isolated from healthy donors and treated ex vivo with the LXR agonist GW3965 (0, 0.1, or 1 μmol/L) showed a marked

**FIG. 4.** GW3965 treatment reduces preretinal neovascularization in the OIR model. Mouse pups treated with GW3965 have a significant reduction in preretinal neovascularization compared with vehicle treatment. Data have been plotted as mean ± SEM (n = 6 per group). *P < 0.05 compared with vehicle treatment.
FIG. 5. Loss of LXR leads to retinal vascular injury and formation of acellular capillaries. 

A: Cholesterol content for neural retinal cells is increased in LXRα/β−/− mice compared with WT mice. Relative cholesterol levels measured by LC/MS/MS are calculated after normalizing for the extraction efficiency of an internal standard (cholesterol-d7) and for the RNA content of the extracted tissue (n = 4–6 per group).

B: Gene expression analysis of neural retinas derived from WT and LXRα/β−/− mice and normalized to the WT group for each gene (n = 3–6 per group).

C: Representative images of retinal vasculature from WT, LXRα−/−, LXRβ−/−, and LXRα/β−/− mice. Acellular capillaries are indicated with arrowheads. Increased numbers of acellular capillaries were observed in retinal vasculature isolated from LXR knockout mice compared with WT, with the greatest degree of acellularity shown by the LXRα/β−/− mice.

D: Quantitative measurements of acellular capillaries (n = 3 per group). Data have been plotted as mean ± SEM. *P < 0.05 compared with WT. (A high-quality digital representation of this figure is available in the online issue.)
reduction of Nox2 mRNA expression (Fig. 7), similar to that seen in DBA/STZ/WD mouse BM-derived cells treated with GW3965 (Fig. 3B).

**DISCUSSION**

Although there are established links between vascular disease and decreased EPC number and function, this is the first study to examine the association between LXR expression in EPCs and the presence of retinopathy. DBA/STZ/WD mice treated with the LXR agonist GW3965 showed fewer acellular capillaries and reduced GFAP expression in the retina. BM-derived progenitor cells from DBA/STZ/WD mice treated with GW3965 showed improved migratory function compared with cells isolated from vehicle-treated DBA/STZ/WD mice. Although we observed a protective effect of LXR activation, the absence of LXRα/β<sup>-/-</sup> mice showed increased acellular capillaries in the retina along with fewer Sca-1<sup>+</sup>/c-Kit<sup>+</sup> cells in their BM; furthermore, Lin<sup>-</sup>/Sca-1<sup>+</sup> cells from LXRα/β<sup>-/-</sup> mice exhibited decreased migratory capacity compared with those from WT mice. Nox2, the catalytic subunit of NADPH oxidase, was increased in BM-derived cells of DBA/STZ/WD origin, and treatment with the LXR agonist, GW3965, resulted in a dramatic reduction in Nox2 mRNA expression (Fig. 7B). Importantly, activation of LXR using GW3965 in human CD34<sup>+</sup> cells similarly resulted in downregulation of Nox2 mRNA (Fig. 7). Thus, LXR activation would serve to reduce oxidative damage, and these results suggest that LXR agonists may represent promising pharmacologic targets for correcting diabetes-induced retinopathy and EPC dysfunction.

There is increasing evidence for a link between cholesterol levels and EPC function (37,38). Hypercholesterolemic patients, free of diabetes, have been found to have a reduced number of circulating EPCs with impaired function (39,40). A dose-dependent decrease in EPC survival and function was observed with exposure to oxidized LDL, and numerous in vitro and in vivo studies demonstrated that hydroxymethylglutaryl-CoA reductase inhibitors (statins) potently enhance EPC numbers and function (41–43). Hypercholesterolemia has also been linked to monocyteosis in atherosclerosis (44). Animal studies have shown that increased monocyteosis in response to granulocyte-macrophage
FIG. 7. Human EPCs respond to GW3956 similar to mouse EPCs. Human peripheral blood CD34+ cells obtained from healthy donors and treated ex vivo with GW3965 show a significant decrease in Nox2 mRNA expression compared with cells exposed to vehicle alone. Data have been plotted as mean ± SEM (n = 3 donors per group). aP < 0.05 compared with vehicle treatment.
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