Arginase: Mechanisms and Clinical Application in Hematologic Malignancy

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Compared to normal tissues and cells, the metabolic patterns of tumor illnesses are more complex, and there are hallmarks of metabolic reprogramming in energy metabolism, lipid metabolism, and amino acid metabolism. When tumor cells are in a state of fast growth, they are susceptible to food shortage, resulting in growth suppression. Using this metabolic sensitivity of tumor cells to construct amino acid consumption therapy does not harm the function of normal cells, which is the focus of metabolic therapy research at the moment. As a non-essential amino acid, arginine is involved in numerous crucial biological processes, including the signaling system, cell proliferation, and material metabolism. Rapidly dividing tumor cells are more likely to be deficient in arginine; hence, utilizing arginase to consume arginine can suppress tumor growth. Due to the absence of arginine succinate synthase, arginine succinate lyase, and ornithine carbamoyl transferase in some blood tumors, arginases may be employed to treat blood tumors. By investigating the mechanism of arginase treatment and the mechanism of drug resistance in greater depth, arginase treatment becomes more successful in hematological cancers and a new anti-cancer agent in clinical practice.

Keywords: arginase, hematologic malignancy, mechanisms, clinical application, therapy

INTRODUCTION

As a heterogeneous disease, the metabolic patterns of tumors differ significantly from normal tissues (1). Metabolic reprogramming of tumor cells refers to the metabolic changes caused by gene mutations and structural changes in tumor cells, such as enhanced glycolysis exemplified by the ‘Warburg effect,’ increased glucose uptake and consumption, increased amino acid uptake and catabolism such as glutamine, and increased lipid and protein synthesis. These metabolic alterations will contribute to the malignant proliferation of tumor cells and the adaptation to an unfavorable living environment (2–6). The distinction between tumor cell and normal cell metabolic pathways encourages the development of antitumor therapies based on metabolic pathways (7). Metabolic reprogramming allows tumor cells to thrive in the microenvironment of nutritional deficiency and hypoxia, but there are also metabolic vulnerabilities. Compared to normal cells, tumor cells are dependent on exogenous nutrients, especially amino acids (8, 9). The metabolic reliance of tumor
cells on exogenous amino acids has become the theoretical foundation for the creation of amino acid deprivation therapy (9).

Arginine is a multifunctional amino acid, involved in many important biological functions in the body, including cell proliferation, signal transduction, muscle growth, nerve transmission, etc., involved in protein synthesis, nucleotide synthesis, urea cycle and other important metabolic pathways, and as a prerequisite for the synthesis of many molecular substances such as citrulline, nitric oxide, glutamic acid, proline biosynthesis (10, 11). Arginine is also classified as a semi-essential amino acid in human body. Under normal physiological conditions, there are endogenous and exogenous sources of arginine, that is, the production of arginine from the starting through ornithine cycle and the uptake of extracellular arginine by cells through membrane protein transporters (12). The key enzymes for arginine synthesis and metabolism mainly include arginine succinate synthase (ASS), arginine succinate lyase (ASL) and ornithine carbamoyl transferase (OCT) (Figure 1). The low expression of these key enzymes makes it impossible for many malignant tumors to synthesize arginine endogenously and become arginine deficient tumor cells, including many hematological malignancies (13). Arginine auxotrophic tumors can be treated by arginine-depleting enzymes without interfering with normal cell physiology, thus becoming a potential antitumor strategy (14).

As an enzyme that depletes arginine, arginase has become a crucial component of arginine depletion treatment. It has been shown to have an anti-tumor impact on hematological malignancies, mostly by triggering autophagy, apoptosis, oxidative stress, and even interfering with the progression of the cell cycle to induce tumor cell death. As a result, this introduction focuses on the current state of arginase application in hematological cancers, associated research mechanisms, and its potential as a treatment for hematological tumors.

ARGinine NUTRITIONAL DEFICIENCIES AND DEPRIVATION THERAPY

The fundamental aspect of tumor cells is metabolic reprogramming, which is to adapt to their rapid multiplication by modifying their metabolic processes, as well as the survival milieu of hypoxia and nutritional shortage (15). During the process of metabolic reprogramming, it is also susceptible to dietary deficiencies, particularly for the growth of essential
nutrients like amino acids (16). Currently, amino acid deprivation therapy for amino acid nutritional deficit has become a promising anti-tumor therapy and has yielded positive outcomes in numerous tumor treatments. The concept of amino acid deprivation was developed in the therapy of tumors, and L-asparaginase was launched more than 50 years ago for the treatment of juvenile leukemia. L-asparaginase is currently one of the most important medications for the combination chemotherapy of juvenile acute lymphoblastic leukemia, which enhances the remission rate of this disease (17, 18). As a result of the efficacy of asparaginase in the therapy of leukemia, there are numerous metabolic enzymes that investigate amino acid nutritional deficits, including enzymes that target arginine metabolism, glutamine, and cysteine.

As a semi-essential amino acid, arginine participates in numerous cellular biological processes, transforms into other amino acids (such as glutamine, proline, etc.), and creates proteins, urea, and agmatine (19). On a biological level, the primary sources of arginine in plasma are dietary digestion and absorption and other protein modification, whereas intestinal-kidney axis arginine synthesis contributes for just 5-15 percent. In the de novo synthesis of arginine, ornithine and carbamoyl phosphate are catalyzed by OCT to produce citrulline, which is subsequently catalyzed by ASS1 and ASL to produce L-arginine. The rate-limiting enzymes of endogenous arginine synthesis are ASS1 and ASL, which are typically expressed in normal cells and unaffected by arginine consumption (19, 20).

Due to the low or non-expression of essential enzymes in arginine endogenous synthesis, especially ASS1, ASL, and OCT, in numerous cancers, tumor cells cannot ordinarily generate endogenous arginine and are highly dependent on exogenous arginine; these tumors are referred to as arginine-auxotrophic. Studies reveal that more than 70 percent of cancers, including melanoma, hepatocellular carcinoma, renal cell carcinoma, glioblastoma, and prostate cancer, as well as a variety of hematological tumors, have arginine shortage (21–23). Cai Y et al. investigated the expression level of the ASS1 gene in various malignancies using the UALCAN database and determined that the expression level of ASS1 in acute myeloid leukemia (AML) was the lowest of all cancers, thereby making AML a typical arginine auxotrophic tumor (24). In addition to Hodgkin’s lymphoma and non-Hodgkin’s lymphoma, low expression of ASS1 was observed in additional solid hematological malignancies (22).

Current studies on the genetic mechanism of low expression of ASS1 gene have confirmed that methylation of CPG regions within the ASS1 gene promoter may be a common epigenetic mechanism of ASS1 gene silencing expression (23), or may include other mechanisms such as inhibition of ASS1 promoter by hypoxia inducible factor HIF-1α. Through the study of glioblastoma cases, Syed N et al. found that the CpG island of ASS1 with methylation was up-regulated in the sensitivity to arginine deprivation therapy, further verifying that the expression of ASS1 was affected by the promoter of the gene promoter (25). In the study of melanoma cells by Tsai WB et al., it was found that the regulation of ASS1 expression was caused by the interaction between the negative regulator HIF-1α and the positive transcription regulator c-Myc and Sp4. HIF-1α combined with the E-box site at the promoter of ASS1 gene could induce the silence of ASS1 expression and cause arginine consumption sensitivity (26). These related studies further induced epigenetic mechanisms by exploring the expression of ASS1 and ASL genes, thus providing ideas for the development of new treatments.

Arginine depletion therapy for arginine-auxotrophic tumors by arginine depletion enzyme (ADE) paves the way for a new anti-tumor treatment. In scientific research of arginine metabolism, the primary degrading enzymes are arginine decarboxylase (ADC), arginine deiminase (ADI), and arginase. ADI is mostly generated from prokaryotes, and ornithine cannot be synthesized via the ADI route as a source of energy for their growth. Mycoplasma-derived ADI is the first antitumor medication to be documented. Even though it has some therapeutic effects, it is easily detected and inactivated by the immune system because of its immunogenicity. So ADI must be bioengineered into PEG-ADI in clinical settings to minimize its immunogenicity (27). ADC, an additional arginine-depriving enzyme, is mostly expressed in mammals. Due to its cytotoxicity to healthy normal cells, it is not employed as a cancer treatment at this time. In contrast to these two enzymes, the arginases currently under investigation are derived from humans and lack immunogenicity and high toxicity. Currently, it has been demonstrated that they have potential therapeutic benefits on a variety of solid and hematological malignancies (11, 28).

ARGINASE AND ITS APPLICATION IN HEMATOLOGICAL MALIGNANCY

Arginase is a manganese-dependent metal ion hydrolase that catalyzes arginine metabolism to generate non-toxic products ornithine and urea. Human tissues express two distinct kinds of arginase, arginase I and arginase II, which are encoded by two distinct genes on separate human chromosomes (29). Human arginase I is mostly found in the cytoplasm of liver cells and is primarily responsible for the consumption of excess ammonia in human tissues via the urea cycle, whereas human arginase II is a mitochondrial enzyme with high expression e.g. in kidney tissue. Arginase II is primarily involved in the creation of nitric oxide and polyamines, as opposed to the arginine urea cycle (30, 31).

In the initial investigation, the arginase was purified from animal liver, however, due to the high dose need of purified arginase treatment, further research and application are limited (32). Human arginase was PEGylated to produce recombinant PEGylated arginase (rhArg-peg), and the catalytic activity of arginase was enhanced by substituting cobalt ions for manganese ions. Its half-life and activity time were lengthened, and its immunogenicity was diminished (33, 34). RhArg has therapeutic potential for malignancies lacking OCT or ASS expression. According to studies, arginine deiminase is
ineffective in tumors expressing ASS, mostly because arginine can be synthesized from citrulline via ASS and ASL (33, 35).

Although arginase therapy in hematological malignancies has not yet reached the approved clinical stage, many studies have confirmed the potential therapeutic activity of arginase in hematological malignancies. Francis Mussai et al. confirmed that pegylated recombinant human arginase (BCT-100) led to rapid consumption of arginine inside and outside cells, thereby inducing cell cycle arrest and cell death. They further revealed that it is because of defective expression of ASS and OCT in AML cells (36). In addition to the therapeutic effect in AML cells, BCT-100 treatment can block the expression of cyclin D3 in cells, thereby inducing apoptosis of T- acute lymphoblastic leukemia (T-ALL) cells in vitro and in vivo, reflecting the potential therapeutic effect of BCT-100 on T-cell malignancies by consuming arginine (28). In addition, Carmela De Santo et al. showed that BCT-100 combined with dexamethasone had obvious cytotoxicity on acute lymphoblastic leukemia (ALL) cells. Although ALL cells up-regulated the expression of arginine transporter CAT-1 and endogenous ASS or OCT, it still could not prevent the cytotoxicity of BCT-100 (37). The combination of BCT-100 and cytarabine has been effective in the treatment of AML and ALL in vivo and in vitro, but drug resistance easily develops, so that further research is needed. In hematological solid tumors, Zeng X et al. demonstrated that recombinant human arginase can inhibit the growth of non-Hodgkin’s lymphoma (NHL) cells by arginine deprivation, which may play an anti-tumor role by cell cycle arrest and caspase-dependent apoptosis (38).

**EFFECT OF ARGINASE TREATMENT ON HEMATOLOGICAL MALIGNANCY**

Using recombinant arginase to treat nutritionally deficient hematological cancers is expected to trigger cell death, which may be mediated by autophagy, cell cycle arrest, or oxidative stress. After treatment with recombinant human arginase, studies have demonstrated that apoptosis results in cell death in ALL, but AML exhibits caspase-independent and non-apoptotic cell death (39). In the research of the influence of BCT-100 on the cell cycle and proliferation, Francis Mussai et al. discovered that the G0/G1 phase of AML cells treated with BCT-100 remained unchanged. Compared to untreated cells, the expression of cyclin A was substantially elevated, whereas the expression of cyclins B and E was lowered, resulting in cell cycle arrest, necrosis, and death (36).

The increased generation of ROS is the cause of tumor cell death produced by BCT-100. Studies demonstrate that BCT-100 therapy further induces ROS generation in bladder cancer cells, blocking phosphorylation of AKT and mTOR and triggering autophagy and death of tumor cells (39). S. Xu et al. also discovered that rhArg slows tumor cell proliferation via autophagy, cell cycle arrest, and oxidative stress in the research on hepatocellular carcinoma and breast cancer (40).

**STUDY ON THE RESISTANCE MECHANISM OF ARGINASE**

In arginase-treated cells, the expression of ASS1 is restored and up-regulated, allowing arginine-deprived tumor cells to recover endogenous arginine synthesis. c-MY, C is a positive regulator of ASS1 expression, and its activation is the primary cause of ASS1 reexpression (41). Arginine deprivation enhances ROS generation in the tumor microenvironment, which in turn activates Gas6/Axl to initiate the PI3K/Akt/GSK3B pathway, resulting in c-Myc oncogene expression upregulation (42). By replacing the negative regulator HIF-1a of the ASS1 gene promoter and competitively occupying the E box of the promoter, up-regulated c-Myc restores and up-regulates the expression of the ASS1 gene, thereby restoring the endogenous synthesis of arginine (43).

Dietary restriction can trigger autophagy in cells. Through autophagy, cells can recruit damaged proteins and defective organelles into autophagosomes for digestion and breakdown misfolded proteins or organelles by binding to lysosomes, thereby supplying the energy and materials necessary for cell survival. Also present in arginase treatment is autophagy. Short-term arginine deprivation therapy decreases ATP and NO levels, induces endoplasmic reticulum stress by activating the mTOR pathway, promotes cell autophagy, and nourishes cells with critical nutrients. In the absence of arginine for an extended period of time, cancer cells can activate the Bax-mediated apoptotic pathway or re-express the ASS gene to produce resistance to arginases (44, 45). Zeng X et al. confirmed that recombinant human arginase activated autophagy in lymphoma cells, human promyelocytic leukemia cells, and human T-ALL cells, which served a protective function (38). In practice, the combination of autophagy inhibitors (such as 3-methyladenine and chloroquine) can augment the anti-tumor activity of arginase and induce apoptosis in tumor cells (46).

In addition to the aforementioned potential resistance mechanisms, arginase therapy is associated with the activation of several metabolic pathways. During the development of arginase resistance, proteomics reveals that the cellular metabolism shifts from glutamine dependence to glucose dependence (47, 48). Several investigations have demonstrated that once the arginase consumes arginine, cell growth becomes dependent on aspartic acid. Aspartic acid is the precursor of pyrimidine nucleotides and can be used as the substrate for arginine synthesis by ASS. Therefore, arginase therapy should be combined with different metabolic inhibition treatments to achieve better therapeutic effects, which requires additional research (48).

**CONCLUSION**

As a semi-essential amino acid in the human body, arginine participates in a variety of vital biological processes. By arginine-degrading enzymes, arginine depletion therapy can...
limit the growth of nutritionally deficient tumors. Hematological cancers, including AML, ALL and various lymphomas, have been shown to respond favorably to this treatment. Through recombinant engineering, arginase has been transformed into pegylated recombinant human arginase, which has a longer half-life, and less adverse effects. Currently, the application of arginase to hematological cancers, including AML, ALL and various lymphomas, have been shown to respond favorably to this treatment. Furthermore, pegylated arginase, which has a longer half-life, and less adverse effects, has entered the clinical research phase, and additional investigation is required to confirm the therapeutic benefit. Drug resistance mechanisms of arginase therapy is still in its exploratory phase, and more study is required to clarify it. To develop more effective treatment options for hematological cancers based on arginase, more in-depth discussion and study is required.

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REFERENCES

1. Judge A, Dodd MS. Metabolism. Essays Biochem (2020) 64:607–47. doi: 10.1042/EBC20190041
2. Hanahan D, Weinberg RA. Hallmarks of Cancer: The Next Generation. Cell (2011) 144:646–74. doi: 10.1016/j.cell.2011.02.013
3. Callao V, Montoya E. Toxohormone-Like Factor From Microorganisms With Impaired Respiration. Science (1961) 134:2041–2. doi: 10.1126/science.134.3495.2041
4. Lunt SY, Vander Heiden MG. Aerobic Glycolysis: Meeting the Metabolic Requirements of Cell Proliferation. Annu Rev Cell Dev Biol (2011) 27:441–64. doi: 10.1146/annurev-cellbio-092910-154237
5. Xia L, Oyang L, Lin J, Tan S, Han Y, Wu N, et al. The Cancer Metabolic Reprogramming and Immune Response. Mol. Cancer (2021) 20:28. doi: 10.1186/s12943-021-01136-8
6. Sun L, Song L, Wan Q, Wu G, Li X, Wang Y, et al. Cmyc-Mediated Activation of Serine Biosynthesis Pathway is Critical for Cancer Progression Under Nutrient Deprivation Conditions. Cell Res (2015) 25:429–44. doi: 10.1038/cr.2015.33
7. Zhang Y, Yang JM. Altered Energy Metabolism in Cancer: A Unique Opportunity for Therapeutic Intervention. Cancer Biol Ther (2013) 14:81–9. doi: 10.4161/cbt.22958
8. Icard P, Lincert H. A Global View of the Biochemical Pathways Involved in the Regulation of the Metabolism of Cancer Cells. Biochim Biophys Acta (2012) 1826:423–33. doi: 10.1016/j.bcbi.2012.07.001
9. Scott S, Lamb J, Smith S, Wheatley DN. Single Amino Acid (Arginine) Deprivation: Rapid and Selective Death of Cultured Transformed and Malignant Cells. Br J Cancer (2000) 83:800–10. doi: 10.1054/bjoc.2000.1355
10. Oberkersch RE, Santoro MM. Role of Amino Acid Metabolism in Angiogenesis. Vascular Pharmacol (2019) 112:17–23. doi: 10.1016/j.vph.2018.11.001
11. Feun LG, Kuo MT, Savaraj N. Arginine Deprivation in Cancer Therapy. Curr Opin Clin Nutr Metab Care (2015) 18:78–82. doi: 10.1097/MCO.0000000000000122
12. Jahani M, Noroznezhad F, Mansouri K. Arginine: Challenges and Opportunities of This Two-Faced Molecule in Cancer Therapy. BioMed Pharmacother (2018) 102:594–601. doi: 10.1016/j.biopha.2018.02.109
13. Caldwell RB, Toque HA, Narayan SP, Caldwell RW. Arginase: An Old Enzyme With New Tricks. Trends Pharmacol Sci (2015) 36:395–405. doi: 10.1016/tpis.2015.03.006
14. Lukey MJ, Katt WP, Cerione RA. Targeting Amino Acid Metabolism for Cancer Therapy. Drug Discov Today (2017) 22:796–804. doi: 10.1016/j.drudis.2016.12.003
15. Borougah LK, DeBerardinis RJ. Metabolic Pathways Promoting Cancer Cell Survival and Growth. Nat Cell Biol (2015) 17:351–9. doi: 10.1038/ncb3124
16. Garcia-Bermudez J, Williams RT, Guarecuco R, Birsoy K. Targeting Extracellular Nutrient Dependencies of Cancer Cells. Mol Metab (2020) 33:67–82. doi: 10.1016/j.molmet.2019.11.011
17. Jaffe N, Traggis D, Das L, Frauenberger G, Hann HW, Kim BS, et al. Favorable Remission Induction Rate With Twice Weekly Doses of L-Asparaginase. Cancer Res (1973) 33:1–4.
18. Richards NG, Kilberg MS. Asparagine Synthetase Chemotherapy. Annu Rev Biochem (2006) 75:629–54. doi: 10.1146/annurev.biochem.75.103004.142520
19. Fultang L, Varden A, De Santo C, Mussai F. Molecular Basis and Current Strategies of Therapeutic Arginine Depletion for Cancer. Int J Cancer (2016) 139:501–9. doi: 10.1002/ijc.30501
20. Husson A, Brasse-Lagnel C, Fairand A, Renouf S, Lavoinne A. Argininosuccinate Synthetase From the Urea Cycle to the Citrulline-NO Cycle. Eur J Biochem (2003) 270:1887–99. doi: 10.1046/j.1432-1033.2003.03559.x
21. Asciero PA, Scala S, Castello G, Daponte A, Simeone E, Ottaiano A, et al. Pegylated Arginine Deiminase Treatment of Patients With Metastatic Melanoma: Results From Phase I and II Studies. J Clin Oncol (2005) 23:7660–8. doi: 10.1200/JCO.2005.02.0933
22. Delage B, Luong P, Maharaj L, O’Riain C, Syed N, Crook T, et al. Promoter Methylation of Argininosuccinate Synthetase-1 Sensitises Lymphomas to Arginine Deiminase Treatment, Autophagy and Caspase-Dependent Apoptosis. Cell Death Dis (2012) 3:e342. doi: 10.1038/cddis.2012.83
23. Qiu F, Huang J, Sui M. Targeting Arginine Metabolism Pathway to Treat Arginine-Dependent Cancers. Cancer Lett (2015) 364:1–7. doi: 10.1016/j.canlet.2015.04.020
24. Cai Y, Chow JPH, Leung YO, Lu X, Yuen CH, Lee WL, et al. NEI-01-Induced Arginine Deprivation Has Potent Activity Against Acute Myeloid Leukemia Cells Both In Vitro and In Vivo. Mol Cancer Ther (2021) 20:2218–27. doi: 10.1158/1535-7163.MCT-21-0120
25. Syed N, Langer J, Janczar K, Singh P, Lo Nguro C, Lattanzio L, et al. Epigenetic Status of Argininosuccinate Synthetase and Argininosuccinate Lyase Modulates Autophagy and Cell Death in Glioblastoma. Cell Death Dis (2013) 4:e458. doi: 10.1038/cddis.2012.197
26. Tassov WB, Aiba I, Lee SY, Feun L, Savaraj N, Kuo MT. Resistance to Arginine Deiminase Treatment in Melanoma Cells is Associated With Induced Argininosuccinate Synthetase Expression Involving C-Myc/HIF-1alpha/Spl4. Mol Cancer Ther (2009) 8:2323–33. doi: 10.1158/1535-7163.MCT-09-0794
27. Veronesi FM, Mero A. The Impact of PEGylation on Biological Therapies. BioDrugs (2006) 22:315–29. doi: 10.2165/00063030-200822050-00004
28. Hernandez CP, Morrow K, Lopez-Barcons LA, Zabaleta J, Sierra R, Velasco C, et al. Pegylated Arginase I: A Potential Therapeutic Approach in T-ALL. Blood (2011) 115:5214–21. doi: 10.1182/blood-2009-12-258822
29. Vockley JG, Jenkinson CP, Shukla H, Kern RM, Grody WW, Cederbaum SD. Cloning and Characterization of the Human Type II Arginase Gene. Genomics (1996) 38:118–23. doi: 10.1006/geno.1996.0606
30. Krebs HA, Hems R, Lund P. Accumulation of Amino Acids by the Perfused Rat Liver in the Presence of Ethanol. Biochem J (1973) 134:697–705. doi: 10.1042/bj1340697
31. Morris SM Jr, Bhamidipati D, Kepka-Lenhart D. Human Type II Arginase: Sequence Analysis and Tissue-Specific Expression. Gene (1997) 193:157–61. doi: 10.1016/S0378-1119(97)00099-1
32. Bach SJ, Hawkins RA, Swaine D. A Short Method for the Purification of Arginase From Ox Liver. Biochem J (1963) 89:263–5. doi: 10.1042/bj0890263
33. Cheng PN, Lam TL, Lam WM, Tsai SM, Cheng AW, Lo WH, et al. Pegylated Recombinant Human Arginase (Rharg-Peg5,000mw) Inhibits the In Vitro and In Vivo Proliferation of Human Hepatocellular Carcinoma Through Arginine Depletion. Cancer Res (2007) 67:309–17. doi: 10.1158/0008-5472.CAN-06-1945
34. Stone EM, Glazer ES, Chantranupong L, Cherukuri P, Breece RM, Tierney DL, et al. Replacing Mn(2+) With Co(2+) in Human Arginase I Enhances Cytotoxicity Toward L-Arginine Auxotrophic Cancer Cell Lines. ACS Chem Biol (2010) 5:333–42. doi: 10.1021/cb900267
35. Yoon JK, Frankel AE, Feun LG, Ekemcioglu S, Kim KB. Arginine Deprivation Therapy for Malignant Melanoma. Clin Pharmacol (2013) 5:11–9. doi: 10.2147/CPPAA.S37350
36. Mussafi E, Egan S, Higginbotham-Jones J, Perry T, Beggs A, Odintsova E, et al. Arginine Dependence of Acute Myeloid Leukemia Blast Proliferation: A Novel Therapeutic Target. Blood (2015) 125:2386–96. doi: 10.1182/blood-2014-09-600643
37. De Santo C, Booth S, Vardon A, Cousins A, Tubb V, Perry T, et al. The Arginine Metabolome in Acute Lymphoblastic Leukemia can be Targeted by the Pegylated-Recombinant Arginase I BCT-100. Int J Cancer (2018) 142:1490–502. doi: 10.1002/ijc.31170
38. Zeng X, Li Y, Fan J, Zhao H, Xian Z, Sun Y, et al. Recombinant Human Arginase Induced Caspase-Dependent Apoptosis and Autophagy in non-Hodgkin’s Lymphoma Cells. Cell Death Dis (2013) 4:e840. doi: 10.1038/cddis.2013.359
39. Zhao Z, Zhang P, Li W, Wang D, Ke C, Liu Y, et al. Pegylated Recombinant Human Arginase I Induces Autophagy and Apoptosis via the ROS-Activated AKT/mTOR Pathway in Bladder Cancer Cells. Oxid Med Cell Longev (2021) 2021:5510663. doi: 10.1155/2021/5510663
40. Xu S, Lam SK, Cheng PN, Ho JC. Recombinant Human Arginase Induces Apoptosis Through Oxidative Stress and Cell Cycle Arrest in Small Cell Lung Cancer. Cancer Sci (2018) 109:3471–82. doi: 10.1111/cas.13782
41. Long Y, Tsai WB, Wangsachiitr M, Tsukamoto T, Savaraj N, Feun LG, et al. Arginine Deiminase Resistance in Melanoma Cells is Associated With Metabolic Reprogramming, Glucose Dependence, and Glutamine Addiction. Mol Cancer Ther (2013) 12:2581–90. doi: 10.1158/1535-7163.MCT-13-0302
42. Kuo MT, Long Y, Tsai WB, Li YY, Chen HHW, Feun LG, et al. Collaboration Between RSK-EphA2 and Gas6-AXL RTK Signaling in Arginine Starvation Response That Confers Resistance to EGFR Inhibitors. Transl Oncol (2020) 13:355–64. doi: 10.1016/j.tranon.2019.12.003
43. Roeker Omewin S, Navasumrit P, Waraprasi S, Parthob V, Sricharunrat T, Bhudesawasi V, et al. Decreased ArgininosuccinateSynthetase Expression in Thai Patients With Cholangiocarcinoma and the Effects of ADI-PEG20 Treatment in CCA Cell Lines. Oncol Lett (2018) 16:1529–38. doi: 10.3892/ol.2018.8807
44. Chen CL, Hsu SC, Ann DK, Yen Y, Kung HJ. Arginine Signaling and Cancer Metabolism. Cancers (Basel) (2021) 13:3541. doi: 10.3390/cancers13143541
45. Changou CA, Chen YR, Xing L, Yen Y, Chuang FY, Cheng RH, et al. Arginine Starvation-Associated Atypical Cellular Death Involves Mitochondrial Dysfunction, Nuclear DNA Leakage, and Chromatin Autophagy. Proc Natl Acad Sci USA (2014) 111:14147–52. doi: 10.1073/pnas.1404171111
46. Li Y, Zeng X, Wang S, Fan J, Wang Z, Song P, et al. Blocking Autophagy Enhanced Leukemia Cell Death Induced by Recombinant Human Arginase. Tumour Biol (2016) 37:6627–35. doi: 10.1007/s13277-015-4253-x
47. Brashears CB, Barlin M, Ehrhardt WR, Rathore R, Schultz Z, Tzeng SC, et al. Systems Level Profiling of Arginine Starvation Reveals MYC and ERK Adaptive Metabolic Reprogramming. Cell Death Dis (2020) 11:662. doi: 10.1038/s41419-020-02899-8
48. Rabinovich S, Adler L, Yizhak K, Sarver A, Silberman A, Agorn S, et al. Diversion of aspartate in ASS1-deficient tumors fosters de novo pyrimidine synthesis. Nature (2015) 527:379–83. doi: 10.1038/nature15529

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