miR-34a−/− Mice are Susceptible to Diet-Induced Obesity

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Objective: MicroRNA (miR)−34a regulates inflammatory pathways, and increased transcripts have been observed in serum and subcutaneous adipose of subjects who have obesity and type 2 diabetes. Therefore, the role of miR-34a in adipose tissue inflammation and lipid metabolism in murine diet-induced obesity was investigated.

Methods: Wild-type (WT) and miR-34a−/− mice were fed chow or high-fat diet (HFD) for 24 weeks. WT and miR-34a−/− bone marrow-derived macrophages were cultured in vitro with macrophage colony-stimulating factor (M-CSF). Brown and white preadipocytes were cultured from the stromal vascular fraction (SVF) of intrascapular brown and epididymal white adipose tissue (eWAT), with rosiglitazone.

Results: HFD-fed miR-34a−/− mice were significantly heavier with a greater increase in eWAT weight than WT. miR-34a−/− eWAT had a smaller adipocyte area, which significantly increased with HFD. miR-34a−/− eWAT showed basal increases in Cd36, Hmgcr, Lxrα, Pgc1α, and Fasn. miR-34a−/− intrascapular brown adipose tissue had basal reductions in c/ebpα and c/ebpβ, with in vitro miR-34a−/− white adipocytes showing increased lipid content. An F4/80high macrophage population was present in HFD miR-34a−/− eWAT, with increased IL-10 transcripts and serum IL-5 protein. Finally, miR-34a−/− bone marrow-derived macrophages showed an ablated CXCL1 response to tumor necrosis factor-α.

Conclusions: These findings suggest a multifactorial role of miR-34a in controlling susceptibility to obesity, by regulating inflammatory and metabolic pathways.

Introduction

The growing incidence of obesity worldwide has presented new therapeutic challenges, making it critical to understand the underlying mechanisms of obesity-associated pathology. In 2014, the World Health Organization estimated that 12.9% of adults have obesity (BMI ≥30 kg/m², age ≥18 years) worldwide, with the Americas (26.8%) and Europe (23%) showing the highest percentage. Obesity is associated with increased low-grade, chronic inflammation that predisposes individuals to a number of comorbidities, e.g., type 2 diabetes (T2D), dyslipidemia, cardiovascular diseases (CVDs), and cancers (1,2).

The main proponent of weight gain during obesity is white adipose tissue (WAT). However, far from being just a lipid storage vessel, WAT can modulate the inflammatory response through secretion of inflammatory cytokines, including CCL2, CXCL8, tumor necrosis factor (TNF)-α, interleukin (IL)−1β, and IL-6 (1-3). In opposition, brown adipose tissue (BAT) expends fatty acids (FA) to produce heat through mitochondrial action and uncoupled protein 1 (UCP1), maintaining body temperature. Peroxisome proliferator-activated receptor (PPAR)γ coactivator (PGC)−1α is important for BAT adaptive thermogenesis, increasing mitochondrial biogenesis and UCP1 expression and promoting brown adipocyte differentiation (4).

MicroRNAs (miRNAs/miRs) are small (19–25 nt), noncoding RNAs, which primarily bind the 3’ untranslated region of target mRNA, mediated by Argonaute proteins within the cytoplasmic RNA-inducing silencing complex (RISC). Before RISC loading, miRNAs form a duplex consisting of a mature (5p) and star (3p) strand, but usually only the 5p is loaded. miRNA is silenced through inhibition of translation or degradation (5).

The miR-34 family consists of 34a, 34b, and 34c, with miR-34a encoded on chromosome 1p36.22 and miR-34b and c on chromosome 11q23.1 (6). Interestingly, miR-34a appears to be dysregulated during obesity, with increased serum miR-34a transcripts found in...
patients who have excess weight (BMI ≥ 25 kg/m²) with T2D, compared with patients who have excess weight without diabetes (7). In addition, studies have shown a correlation with increasing BMI and miR-34a transcripts in human subcutaneous WAT (scWAT) and increased miR-34a transcripts over subcutaneous adipocyte differentiation in vitro (8,9). Further to its pro-diabetes role, miR-34a transcripts have been positively associated with destruction of primary β-islet cells and cell lines, upon inflammatory stimulation, with decreased insulin secretion, through inhibition of VAMP2 (10,11).

Inflammatory roles of miR-34a have been proposed, with lipopolysaccharide-induced inflammation reducing miR-34a expression in macrophage cell lines, and miR-34a mimics reducing IL-6 and TNF-α expression (12). Furthermore, miR-34a has been shown to target the soluble IL-6 receptor (13).

Adipose tissue macrophages (ATMs) represent a major immune infiltrate into the obese WAT (predominantly visceral), increasing from around 12% to 41% (14). Increasing inflammatory cytokines, particularly CCL2, recruits monocytes into the WAT that differentiate into ATMs (15). A high percentage are proinflammatory F4/80⁺ CD11b⁺ CD11c⁻ M1 ATMs in high-fat diet (HFD)-fed rodents, and depletion of these cells has shown a reversal in inflammation and insulin resistance (16). Other immune cells, such as neutrophils, have also been shown to contribute to insulin resistance and obesity-induced inflammation (17).

Therefore, the aim of this study was to examine the metabolic and inflammatory role of miR-34a in a murine, diet-induced obesity model, using miR-34a⁻/⁻ mice; specifically, examining miR-34a’s role in adipocytes and macrophages during obesity. Our findings show that miR-34a⁻/⁻ mice are susceptible to weight gain, likely through basal metabolic gene changes in adipose caused by dysregulation of PGC-1α.

**Methods**

**Murine studies**

All in vivo studies used 7-week-old, male, wild-type (WT) or miR-34a⁻/⁻ (B6.Cg-Mir34atm1Lhe/J) C57BL/6 mice (Jackson Labs, Sacramento, CA) bred in-house. Mice were fed normal chow (chow) or HFD (0.15% cholesterol and 21% lard: Special Diet Services, Essex, UK) ad libitum for 24 weeks. Weekly weights, monthly fasting blood glucose, and glucose tolerance test (GTT) measurements were recorded. Fasting (16–18 h) blood glucose measurements were recorded using an Accu-Chek® Mobile glucose monitor (Roche, Burgess Hill, UK). GTT measurements were taken every 30 min following I.P. administration of 1 g/kg glucose, in 0.9% saline solution. All studies were carried out in a registered research facility, under UK Home Office guidelines.

**Human studies**

Omental adipose tissue biopsies were collected from six patients with obesity (five women and one man), T2D (HbA1c = 7.5 ± 0.97%), and/or sleep apnea, undergoing bariatric surgery. Patients were on metformin (BMI: 42.13 ± 3.24; age: 50.17 ± 8.08 years). All patients gave written informed consent before inclusion in the study, with approval from the West of Scotland Research Ethics Service (REC Ref 12/WS/0158).

Additional methods can be found in Supporting Information.

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**Results**

**Expression of miR-34a in obese adipose tissue and in vitro bone marrow-derived macrophages**

*In situ* hybridization showed miR-34a distribution within visceral WAT was ubiquitous throughout the epididymal (e)WAT of WT mice on chow and HFD and omental adipose from bariatric surgery patients with obesity (Figure 1A). Similarly, no change was observed in miR-34a or miR-34a* transcripts between WT chow and HFD groups (Figure 1B). However, we observed increases in both miR-34a and 34a* transcripts within the liver (P = 0.0088, P = 0.0027) and intrascapular (i)BAT (P = 0.0433, P = 0.0399) during HFD feeding (Figure 1C). Furthermore, we observed an increase in miR-34a (P = 0.0086) and miR-34a* (P = 0.0461) transcripts in WT bone marrow-derived macrophages (BMDMs) when stimulated with TNF-α (Figure 1D). We examined the other miR-34 family members but found no expression changes (Supporting Information Figure S1).

**Endogenous miR-34a regulated body weight and adipocyte size**

To investigate the role of endogenous miR-34a in regulating obesity, we measured the weight and metabolic parameters of WT and miR-34a⁻/⁻ mice over 24 weeks on chow or HFD (Figure 2A). Before the study began, it was noted that miR-34a⁻/⁻ mice were heavier at week 0 (chow: 6.71%; P = 0.1311 and HFD: 8.07%; P = 0.0377) (Supporting Information Figure S2A). This was maintained in the chow group at week 2 (6.11%; P = 0.0312) and 3 (6.86%; P = 0.0798) but increased further in HFD-fed miR-34a⁻/⁻ mice at week 2 (12%; P < 0.0001) and 3 (11.60%; P = 0.0009). HFD-fed miR-34a⁻/⁻ mice were significantly heavier (9.22 ± 0.56%; P = 0.0075) than WT counterparts over 24 weeks, by AUC analysis (Figure 2B, C). Body weights of chow-fed miR-34a⁻/⁻ mice were similar to WT controls. Interestingly, when we examined the total body fat percentage (AUC_final), both HFD-fed WT and miR-34a⁻/⁻ mice showed similar fat content (41.7 ± 1.04% vs. 43.5 ± 0.68%) (Figure 2D). These weight differences were not explained by food intake during metabolic cage studies (Supporting Information Figure S2B) or fasting serum leptin levels (Supporting Information Figure S2C).

However, ex vivo investigations revealed a significant increase in eWAT weight in HFD-fed miR-34a⁻/⁻ mice (0.61 ± 0.10 g vs. 1.80 ± 0.08 g; P < 0.0001) not observed in WT controls (Figure 2e). Liver weight remained unchanged (Supporting Information Figure S2D). Histological examination of the eWAT revealed smaller, more densely packed adipocytes within the miR-34a⁻/⁻ chow tissues, compared with WT (Figure 2F, G). When stressed with a HFD, the eWAT from miR-34a⁻/⁻ mice showed a significant increase in cell area (P = 0.0030) and decrease in cell number per field (P = 0.0013) not seen in WT, bringing them to WT levels.

To examine alterations in glucose homeostasis, we measured monthly fasting blood glucose and performed a GTT at week 24. However, we did not observe any changes between HFD-fed WT and miR-34a⁻/⁻ mice, both showing hyperglycemia from week 12 (P < 0.001) (Supporting Information Figure S2E) and similar glucose handling (Supporting Information Figure S2F). This was further compounded by no change in fasting serum insulin or resistin between HFD groups at week 24 (Supporting Information Figure S2F). Additionally, there was no change in fasting serum cholesterol.
and triglyceride measurements between WT and miR-34a<sup>−/−</sup> groups on either chow or HFD (Supporting Information Table S1).

miR-34a<sup>−/−</sup> mice showed basal metabolic gene changes in eWAT and iBAT on chow diet

To investigate the changes in adipose weight and adipocyte size, we examined metabolic gene expression in eWAT <em>ex vivo</em>. The following FA and cholesterol metabolism genes’ expression was increased in chow-fed miR-34a<sup>−/−</sup> mice compared with WT chow counterparts: <em>Cd36</em> (<em>P</em> = 0.0036), 3-hydroxy-3-methyl-glutaryl-CoA reductase (<em>Hmgcr</em>; <em>P</em> = 0.0012), liver X receptor (<em>Lxr</em><sub>a</sub>; <em>P</em> < 0.0012), <em>Pgc-1α</em> (<em>P</em> = 0.0138), and fatty acid synthase (<em>Fasn</em>; <em>P</em> = 0.0027). In addition, decreased <em>Ppar-γ</em> (<em>P</em> = 0.0471), acetyl-CoA carboxylase (<em>Acc-x</em>; <em>P</em> = 0.0018), and <em>Fasn</em> (<em>P</em> = 0.0274) transcripts were observed in miR-34a<sup>−/−</sup> mice when fed HFD (Figure 3A). We observed no
miR-34a<sup>−/−</sup> mice are susceptible to diet-induced obesity. (A) Diagrammatic representation of the 24-week in vivo study structure: high-fat diet (HFD), fasting blood glucose (FBG), and glucose tolerance test (GTT). Data in panels B–F are from WT and miR-34a<sup>−/−</sup> (KO) mice on chow vs. HFD over 24 weeks of diet. (B) Representative image of HFD-fed WT and KO mice at week 24 of study. (C) Body weight measurements over 24 weeks; n = 10 for KO chow and WT HFD and n = 9 for WT chow and KO HFD groups. (D) MRS analysis of mouse percentage body fat at week 24, calculated from integration ratios for lipid (AUCH2O) to water (AUCH2O) peaks; n = 6 for WT chow and HFD, n = 4 for KO HFD, and n = 3 for chow HFD groups. (E) Weight of excised epididymal (e)WAT at week 24 of study; n = 10 for KO chow and WT HFD and n = 9 for WT chow and KO HFD groups. (F) Representative H&E staining of eWAT excised in panel E. (G) Quantification of adipocyte area and number in panel F, with averages taken from ≥5 fields per section of n = 10 for KO chow and WT HFD and n = 8 for WT chow and KO HFD groups. Area measurements were converted from pixels to square centimeters using the image scale. All graphs represent mean values with SEM. **P < 0.01, ****P < 0.0001, one-way ANOVA, with Bonferroni’s multiple comparisons post-test.
changes in other eWAT metabolic genes examined (Supporting Information Figure S3A).

Furthermore, we investigated the expression of metabolic genes in iBAT. Interestingly, we observed decreased expression of Cebpa (P < 0.01) and Cebpβ (P < 0.01) from both miR-34a<sup>−/−</sup> groups and increased expression of Pgc1α (P = 0.0072) in miR-34a<sup>−/−</sup> mice when fed HFD (Figure 3B). However, Ucp1 expression remained unchanged. Additionally, we saw few changes in hepatic metabolic genes (Supporting Information Figure S3B–D).

eWAT macrophages from HFD-fed miR-34a<sup>−/−</sup> mice expressed an F4/80<sup>high</sup> phenotype, with an increase in type 2 cytokines

Given that miR-34a transcripts were increased in WT BMDMs stimulated with TNF-α, they may contribute to the in vitro metabolic profile. Therefore, using FACS we gated on the larger, more granular F4/80<sup>+</sup> cells from WT and 34a<sup>−/−</sup> eWAT to identify macrophages, after 24 weeks on chow or HFD (Figure 4A). These cells were CD45<sup>+</sup>, CD11b<sup>+</sup>, MHCII<sup>+</sup>, and CD86<sup>+</sup>. Further analysis revealed a population of F4/80<sup>high</sup> cells within the miR-34a<sup>−/−</sup> HFD group not observed in WT (Figure 4B). We observed a significant increase in F4/80 surface expression on miR-34a<sup>−/−</sup> cells, when fed HFD (P = 0.0497), and a lower F4/80<sup>+</sup> macrophage content within the eWAT of miR-34a<sup>−/−</sup> chow mice, compared with WT chow (P = 0.0078) (Figure 4C). There were no changes in the other macrophage M1/M2 surface markers examined (Supporting Information Figure S4A), or M1/M2 genes within the eWAT, except a basal increase in miR-34a<sup>−/−</sup> chow eWAT Nos2 (P = 0.0500) and decreased HFD-fed miR-34a<sup>−/−</sup> eWAT Retnla (P = 0.0006) transcripts, over WT diet controls (Supporting Information Figure S4B). To check whether an increase in cytosolic lipid could contribute to the F4/80<sup>high</sup> macrophage phenotype in miR-34a<sup>−/−</sup> HFD eWAT, we back-gated on this population for side-scatter, a measurement of internal complexity, but observed no difference in side-scatter MFI between any of the groups (Supporting Information Figure S4C).

Bassaganya-Riera et al. identified a subset of F4/80<sup>high</sup> ATMs that produced high levels of anti-inflammatory IL-10 (18). Therefore, we examined eWAT cytokine transcripts and observed increased IL-10 in miR-34a<sup>−/−</sup> HFD eWAT, compared with WT HFD and miR-34a<sup>−/−</sup> chow eWAT (P < 0.05) (Figure 4D). Additionally, serum cytokine levels of IL-5 (P = 0.0103) and chemokine CXCL9 (P = 0.0303) were increased in HFD-fed miR-34a<sup>−/−</sup> mice, over WT controls (Figure 4E). However, serum IL-10 was not detected in any group. miR-34a<sup>−/−</sup> and WT in vitro BMDMs were stimulated with TNF-α to assess macrophage-mediated modulation of metabolic and inflammatory pathways. Interestingly, TNF-α stimulation of miR-34a<sup>−/−</sup> macrophages did not upregulate transcripts of the inflammatory, neutrophil chemotactic cytokine Cxcl1 (P = 0.0168), or secreted protein (P = 0.0470), as in WT controls (Figure 4F). As a corollary, we observed a reduction in splenic neutrophil percentage (CD45<sup>+</sup> CD11b<sup>+</sup> F4/80<sup>+</sup> CD11c<sup>−</sup> Ly6c<sup>−</sup>Ly6g<sup>+</sup>) in miR-34a<sup>−/−</sup> mice from both chow (P = 0.0003) and HFD (P = 0.0356) groups, compared with WT controls at 24 week of diet (Figure 4G). However, the same was not observed in the eWAT (Supporting Information Figure S4D).

Examination of Cd36, Hmgcr, Lxra, Ppara, Fasn, and Accα transcripts in BMDMs revealed no change (Supporting Information Figure S4E, F). However, higher basal transcript expression of Pgc1α (P = 0.0024) was observed in miR-34a<sup>−/−</sup> BMDMs, which decreased to WT levels upon TNF-α stimulation (P = 0.0139) (Figure 4H). Alternatively, other serum and BMDM supernatant cytokines from miR-34a<sup>−/−</sup> and WT mice did not change (Supporting Information Figure S5).

In vitro miR-34a<sup>−/−</sup> adipocytes showed changes in lipid content

To examine the function of miR-34a in adipose tissue, we differentiated primary white and brown adipocytes in vitro. We observed significant upregulation of miR-34a transcripts (P = 0.0330) by day 8 of differentiation in WT white adipocyte precursors (Figure 5A), with upregulation of miR-34b<sup>+</sup> and 34c<sup>+</sup> at day 4 (P < 0.05), which decreased by day 8 (Supporting Information Figure S6A).

Therefore, we examined differentiation genes in WT and miR-34a<sup>−/−</sup> white adipocytes. Examination of Cebpα, Ppara, and Fabp4 over the 8-day differentiation revealed only a small increase in Cebpα
expression ($P = 0.0396$) at day 2 in miR-34a$^{-/-}$ adipocytes (Figure 5B–D). Interestingly, we observed increased lipid content in day 8 miR-34a$^{-/-}$ white adipocytes ($P = 0.0280$) (Figure 5E) by FACS, with no change in the percentage of lipid-positive cells (Figure 5F).

Surprisingly, miR-34a$^{-/-}$ white adipocytes had increased supernatant leptin at day 0 ($P = 0.0458$), with lower levels at day 6 ($P = 0.0009$) of differentiation, compared with WT (Figure 5G). Furthermore, supernatant insulin was decreased at day 2 ($P = 0.0041$) of

**Figure 4** High-fat diet (HFD)-fed miR-34a$^{-/-}$ mice show an F4/80$^{high}$ phenotype, with increased type 2 cytokines and a reduction in splenic neutrophils. (A) Representative dot-plots showing the gating strategy used to identify macrophages in the SVF from murine epididymal (e)WAT at week 24 in WT and miR-34a$^{-/-}$ (KO) mice on chow vs. HFD. (B) Representative dot-plots showing the F4/80$^{high}$ macrophage phenotype in KO eWAT after HFD feeding, from the same study as panel A. (C) Quantification of panel B, showing median fluorescence intensity (MFI) values representing surface expression and percentage F4/80$^{+}$ cells in the macrophage FSC-SSC gate; $n = 9$ for all groups, except $n = 10$ for KO chow. (D) RT-qPCR data of interleukin (IL)-10 gene expression in eWAT from same study as panel A, normalized to 18s rRNA; $n = 6$ for all groups, except $n = 5$ for KO HFD and WT chow. (E) Significant changes from cytokine Luminex data of serum from mice fasted for 16 to 18 h from the same study as panel A; $n = 6$ for KO chow and WT HFD groups and $n = 5$ for WT chow and KO HFD groups. (F) RT-qPCR gene expression and supernatant, cytokine Luminex protein data for CXCL1, from WT and KO in vitro bone marrow-derived macrophages (BMDM) ± 45.45 ng/mL tumor necrosis factor (TNF)-α for 24 h; $n = 3$. RT-qPCR data normalized to 18s rRNA. (G) FACS quantification of the percentage of neutrophils (CD45$^{+}$ CD11b$^{+}$ F4/80$^{-}$ CD11c$^{-}$ Ly6c-Ly6g$^{-}$) in the FSC-SSC population from spleens of mice in the same study as panel A. Gating strategy is shown in Supporting Information Figure S4D; $n = 8$ for all groups, except $n = 4$ for WT chow and $n = 6$ for WT HFD, KO HFD, and WT Tnfα groups. (H) RT-qPCR quantification of peroxisome proliferator-activated receptor-γ coactivator (PGC)-1α and c/ebpα transcripts in the same samples as panel F; $n = 3$. All graphs represent mean values with SEM, except for RT-qPCR, represented as relative quantification (RQ) with RQmin – RQmax values. *$P < 0.05$, **$P < 0.01$, ***$P < 0.001$, one-way ANOVA, with Bonferroni’s multiple comparisons post-test.
miR-34a increases over in vitro white adipocyte differentiation, and miR-34a−/− white adipocytes show increased lipid content at day 8. (A) RT-qPCR quantification of miR-34a transcripts over in vitro differentiation of SVF white preadipocytes (day 0–8) from WT murine epididymal (e)WAT, with mature adipocyte fraction (WAT) as a positive control, normalized to RNU6B; three mice were pooled together for each replicate of *n* = 3. (B–D) RT-qPCR gene expression over in vitro differentiation of SVF white preadipocytes (day 0–8) from miR-34a−/− (KO) and WT murine eWAT, with mature adipocyte fraction (WAT) as a positive control, normalized to 18s rRNA; three mice were pooled together for each replicate of *n* = 3. (E,F) Quantification of lipid in day 8 in vitro adipocytes by FACS and BODIPY, represented as geometric mean fluorescence intensity (MFI) values representing content, and percentage positive cells; *n* = 3. (G,H) Adipokine Luminex measurements of leptin and insulin from supernatants of cultures in panels B–D; *n* = 3. All graphs represent mean values with SEM, except for RT-qPCR, represented as 1/ΔCt. Statistics were calculated using a two-way ANOVA, with Bonferroni’s multiple comparisons post-test, or an unpaired Student’s *t*-test for FACS lipid measurements. A one-way ANOVA and Dunnett’s multiple comparisons post-test was used to calculate statistics for panel A, compared with day 0. *P* < 0.05, **P < 0.01, ***P < 0.001.
miR-34a−/− white adipocyte differentiation, compared with WT (Figure 5H). We observed no change in resistin (Supporting Information Figure S6B). In day 8 brown adipocytes, we did not observe any changes in leptin, insulin, or resistin (Supporting Information Figure S6C). Additionally, obesity proteome array analysis of day 8 supernatants from white and brown adipocyte cultures did not reveal any other cytokine or adipokine differences (Supporting Information Figure S7).

Figure 6.

Obesity miR-34a−/− Mice are Susceptible to Obesity Lavery et al.
To further explore the metabolic changes in miR-34a−/− iBAT, we examined miR-34a expression over *in vitro* differentiation of WT brown adipocyte precursors, but found no change in miR-34a (Figure 6A). However, we observed increases in miR-34b and 34c− at day 4 ($P = 0.0305$) and both miR-34b and 34c− at day 6 ($P < 0.05$) and day 8 ($P < 0.01$) of differentiation (Figure 6B).

WT and miR-34a−/− day 8 brown adipocytes were stimulated with noradrenaline to examine the effect of β-adrenergic stimulation on Ucp1 and Pgc1α transcripts. However, we observed no difference in Ucp1 and Pgc1α transcript expression (Figure 6C, D). Additionally, we quantified cellular mitochondrial content by FACS but found no difference (Figure 6E, F). However, FACS analysis identified an increase in the percentage of noradrenaline-stimulated, miR-34a−/− lipid-positive brown adipocytes ($P = 0.0445$) (Figure 6G), with no difference in cellular lipid content (Figure 6H).

We examined the expression of key brown adipocyte differentiation genes, but found no difference in Pgc1α, Ucp1, Cebpα, Cebpβ, or Ppar-γ transcripts between WT and miR-34a−/− brown adipocytes (Supporting Information Figure S8A). Finally, we observed no change in white adipocyte Cd36, Hmgcr, Lxrα, Pgc1α, Fasn, and Acc-α metabolic genes (Supporting Information Figure S8B).

**Discussion**

The regulation of chronic, low-grade inflammation of adipose tissue during obesity is poorly understood and this study aimed to address the role of miR-34a in this process. Our findings showed that miR-34a−/− mice were heavier than WT controls and gained more weight when challenged with a HFD, suggesting they were more susceptible to weight gain. miR-34a−/− eWAT showed smaller, more numerous adipocytes, with a lower tissue mass on chow that increased to WT levels on HFD. It is surprising that there was no difference in these characteristics between HFD-fed miR-34a−/− and WT mice, despite a clear difference in body weight. With no change in liver weight, this could suggest increases in other WAT depots (e.g., scWAT), BAT, or muscle tissue. One gene that was increased in liver weight, this could suggest increases in other WAT depots caused by chronic overexpression of peroxisome proliferator-activated receptor-γ coactivator (PGC)-1α during HFD feeding.

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Respiratory uncoupling by UCPs has been linked to increased FA oxidation (24). It may seem counterproductive that in miR-34a−/− eWAT we saw basal increases in genes involved in lipid uptake: Cδδ, de novo lipogenesis: Fasn, and cholesterol biosynthesis: Hmgr, which caused increases in adipocyte total and free cholesterol, associated with omental adipocyte hypertrophy in pri-mates (25-27). However, MPGC-1z TGs have increased expression of Cδδ and Fasn, with Fasn expression driven by Pgc1z’s increase in Lxrα expression, and increased intracellular lipid (28). The authors suggest that this response “refuels” the cell’s lipid stores for continued FA oxidation. Therefore, there may be an imbalance in lipolysis/FA oxidation vs. lipid uptake/lipogenesis, pushing metabolism to the latter in adipocytes of miR-34a−/− mice, due to chronic Pgc1z upregulation, which predisposes these mice to weight gain when stressed with a HFD. Additionally, this would explain why miR-34a−/− mice are heavier before HFD feeding. In fact, when MPGC-1z TGs were fed HFD they developed insulin resistance and were not protected from diet-induced obesity, showing increased lipid uptake and lipogenesis genes in skeletal muscle, including Cδδ, and no change in resting serum insulin, triglycerides, cholesterol, glucose, and FFAs (29). The increased lipid content observed in mature *in vitro* miR-34a−/− white adipocytes supports this...
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Paradoxically, Bassaganya-Riera et al. suggested that F4/80 high macrophages, inhibiting NF-κB, which increased further upon LPS-stimulation (18). These reports supported data showing ablated upregulation of CXCL1 in TNF-α-vitro stimulated micR-34a in response in micR-34a. Systemically, we observed characteristics of a type 2 immune response in HFD-fed micR-34a−/− mice, with increased intracellular IL-6, TNF-α, higher levels of MHCII, CX3CR1, CD11c, CCR2, and PPAR-γ in db/db mice, which others have shown represses TNF-α. Thus, a type 2 immune response in HFD-fed micR-34a−/− mice may be initiated as a compensatory effect during overnutrition, to regain metabolic homeostasis. A study limitation is that we did not examine micR-34a expression in BMDMs from HFD-fed mice or TNF-α-stimulated adipocytes.

In summary, this study has demonstrated that micR-34a regulates the development of diet-induced obesity in mice, with micR-34a−/− mice showing a susceptibility to weight gain, likely through dysregulation of PGC1α (Figure 6). It is clear that the role of micR-34a in metabolism is complex and likely to vary depending on context of treatment and model, with potentially contrasting roles in different cell types. This study has highlighted that chronic micR-34a inhibition, as a treatment for obesity, may have undesirable metabolic and immunoregulatory consequences. Further research is required to fully elucidate the metabolic and inflammatory pathways regulated by micR-34a.

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