ORIGINAL ARTICLE

The shaping and cleaning abilities of self-adjusting files in the preparation of canals with isthmuses after glidepath enlargement with ISO or ProTaper Universal NiTi files

Zhaohui Liu, Jia Liu, Lisha Gu, Wei Liu, Xi Wei*, Junqi Ling*

Department of Operative Dentistry and Endodontics, Guanghua School of Stomatology and Institute of Stomatological Research, Sun Yat-sen University, Guangzhou, Guangdong, China

Received 2 August 2015; Final revision received 31 August 2015
Available online 4 January 2016

KEYWORDS

- glidepath enlargement
- ISO files
- isthmus
- micro-computed tomography
- NiTi rotary files
- ProTaper Universal files
- self-adjusting files

Abstract

Background/purpose: This study compared the shaping and cleaning ability of self-adjusting files (SAF) after glidepath enlargement with ISO NiTi files (Group I) or ProTaper Universal (PTU; Group II) for preparing maxillary premolar canals with isthmuses.

Materials and methods: Twenty-eight teeth containing isthmuses were scanned preoperatively after glidepath enlargement and preparation with SAF (n = 14). Changes in canal and isthmus volume, prepared surface, debris volume, and transportation were determined. Data were compared by t test between groups and paired t test within each group.

Results: No difference was observed regarding changes in canal volume or prepared surface between the two groups (P > 0.05). Paired t tests showed that the coronal and middle prepared areas of the canal after using SAF in Group I were statistically larger than those using ISO files, whereas the use of an adjuvant preparation with SAF after PTU resulted in a significant increase in the apical prepared area in Group II. ISO and SAF systems deviated less from the center than did PTU and SAF systems at most sections. After the use of SAF, isthmus volumes were significantly increased within each group (P < 0.05). Although less debris resulted from the use of SAF instruments as compared to glidepath instruments, there was no significant difference between both groups (P > 0.05).
Introduction

The goal of endodontic therapy is the removal of all infected or necrotic tissue, microorganisms, and microbial byproducts from the root canal system. However, it is difficult to achieve thorough debridement of the entire root canal system due to the intricate nature of the morphological anatomy, particularly of teeth containing isthmuses, anastomoses, and/or other canal irregularities.1-3 Isthmuses are defined as narrow ribbon-, fin-, or web-shaped communications between two root canals. In roots with two canals, isthmuses are highly prevalent, such as the mesial roots of maxillary and mandibular molars, the distal root of mandibular molars, the maxillary and mandibular first and second premolars, and mandibular incisors.4-6 Furthermore, the buccolingual orientation of isthmuses does not permit radiographic detection before the endodontic procedures, further aggravating the difficulties inherent in root canal debridement.

As the most widely used files for root canal preparation, rotary files have a spiral blade arranged in a helical formation. When rotated, they machine a central portion of the root canal into a round cross-section, but the buccal/lingual areas of flat root canals and areas facing the isthmus in tear-shaped canals cannot be adequately prepared.7,8 Furthermore, hard-tissue debris in the root canal system mainly accumulates in the isthmus area, as discovered using microcomputed tomography (MCT) or histologic evaluation.1,8 Although untouched areas and debris can theoretically be treated by the following chemical debridement, these isthmuses and other canal irregularities make irrigant penetration difficult, resulting in ineffective dissolution of hard-/soft-tissue remnants and microorganism destruction. Consequently, isthmuses often harbor tissue, microbes, and debris after instrumentation, potentially providing space and nutrition for bacterial growth and/or future bacterial recontamination of the root canal.1,8

The self-adjusting file (SAF; ReDent-Nova, Ra’anana, Israel) was designed to address the shortcomings of traditional rotary files by adjusting itself three-dimensionally to the shape of the root canal. This instrument is operated with reciprocating vibrating handpieces, and its hollow design allows for continuous delivery of irrigant throughout the procedure via a special rinsing unit.9 To date, root canal preparation with the SAF has been quantitatively and qualitatively described in several studies.10-13 MCT-based studies showed that the percent of root canal area affected by the SAF method is higher than that affected by popular rotary instrumentation.10,12,14 Additionally, SAF is significantly more effective than rotary NiTi instrumentation in eliminating debris and viable Enterococcus faecalis cells in long oval root canals.15-19 Consequently, it is supposed that SAF has superior potential to treat more area of the isthmus and eliminate debris produced by rotary canal preparation.

According to the description by Metzger et al.,9,20 a 1.5-mm SAF may easily be compressed to the extent where it can be inserted into any canal previously prepared or negotiated with a #20 K-file, and a 2.0-mm file can easily be compressed into a canal prepared with a #30 K-file. In fact, the average diameter of a biological apex is 0.08 mm to 0.3 mm,21 therefore, the use of a SAF alone is unable to adequately prepare the root canal. Currently, the glidepath used for SAF preparation is ISO K files.9,12 However, insufficient apical preparation and inadequate apical irrigation with SAF after ISO K files instrumentation has been reported.22 This observation suggests that root canal debridement is improved with progressively larger instrumentation through removing more contaminated dentin and providing better access for efficient irrigation and disinfection of the canal system. The purpose of this study was to compare the shaping and cleaning abilities of the SAF after glidepath enlargement with ISO NiTi files or Pro-Taper Universal (PTU) for preparing maxillary premolar canals with isthmuses.

Materials and methods

Selection of teeth

Mature single-root maxillary premolars were collected from a native Chinese population. All teeth were extracted for orthodontic reasons, and written consent was obtained from the patients. Teeth were excluded if there were extensive caries, fractures, or prior endodontic treatment. Attached soft tissue and calculus were removed using an ultrasonic scaler, and the teeth were stored in 0.1% thymol solution at 4°C until further use.

Radiographs were taken of the teeth from the mesiodistal directions and scanned using a desktop X-ray microfocus CT scanner (ZKKS-MCT-Sharp, Guangzhou, China) at an isotropic resolution of 20 μm. The system consisted of a sealed X-ray tube operated at 75 kV/133 μA. Teeth containing type IV isthmuses described by Hsu and Kim23 were selected and accessed using high-speed diamond burs. The instruments were gently introduced by hand to working length in ascending order beginning with #8 K files (Dentsply Maillefer, Ballaigues, Switzerland), and the first binding file was termed as the initial apical file. Canal curvatures were assessed according to Schneider’s technique.24 A total of 28 teeth with curvatures of < 20° and initial apical files of < #15 were selected. Canal lengths and patency were determined using size #10 K files, and the working length (WL) was established at 1 mm.
shorter than the length of the root. The glidepath of all of the teeth was confirmed to a size #15 K file (Dentsply Maillefer). Specimens were then randomly allocated to two groups (n = 14 each) based on the preparation protocols used. The sample size was based on a pilot study and related research. All root canal treatments were carried out by a single doctor who had been specifically trained to operate the SAF instrument and had more than 10 years of clinical experience with NiTi rotary instruments.

**Group I: Glidepath enlargement with ISO NiTi files and SAF preparation**

Canals in Group I were first prepared to the WL using ISO instruments 25/02 and 30/02 (Hero642; Micromega, Besançon, France). The average WL in this group was 20.3 mm (range, 19–22.5 mm). NaOCl (1.3%) and EDTA (17%) were delivered in a syringe with a 30-gauge needle as deep as possible to 1 mm from working length as the irrigant (2 mL NaOCl and 6 mL EDTA per canal each time). After being prepared to size #30, canals were irrigated with 5 mL 1.3% NaOCl. The teeth were then subjected to a postoperative MCT scan. Thereafter, the canals were prepared using 2.0-mm diameter self-adjusting files (SAF-2.0; ReDent-Nova). The SAF was operated using an in-and-out vibrating handpiece at 5000 vibrations/min and 0.4-mm amplitude. A special irrigation device (VATEA; ReDent-Nova) was connected to the irrigation hub on the file, enabling irrigant to be delivered at a flow rate of 5 mL/min. During the first 2 minutes, 1.3% NaOCl was used, which was followed with 1 minute of EDTA irrigation and another 1 minute of NaOCl irrigation per canal. The total volume of NaOCl and EDTA used for this group was 24 mL and 17 mL per canal, respectively. The teeth were scanned using the MCT.

**Group II: Glidepath enlargement with PTU and SAF preparation**

Canals in Group II were prepared with six PTU rotary instruments (Dentsply Maillefer) in the sequence recommended by the manufacturer (Sx, S1, S2, F1–F3). The average WL in this group was 20.5 mm (range, 18–22.5 mm). Two milliliters of 1.3% NaOCl and 17% EDTA were used to irrigate the canal between each instrument according the method described for Group I. After F3 instrumentation, the canal was irrigated with 7 mL of 1.3% NaOCl and scanned with MCT. Then canals were prepared using SAF-2.0 under irrigation with 1 minute of EDTA and 1 minute of 1.3% NaOCl per canal and delivered as previously described. The volume of NaOCl and EDTA used for this group was 24 mL and 17 mL per canal, respectively. The teeth were then scanned for a third time with the MCT.

**Evaluation**

MCT data were reconstructed from the biological apex to the level of the cementoenamel junction. Precise repositioning of pre-preparation and various post-preparation images was ensured by combining a custom-made SAF preparation after glidepath enlargement.
mounting device with a software-controlled iterative superimposition algorithm (the ImageFusion module in the software MedINRIA, Paris, France). The resulting color-coded root canal models (in which red indicates preoperative canal surfaces, green indicates postoperative canal surfaces, and yellow indicates hard-tissue debris) enabled quantitative comparison of the matched root canals before and after shaping. Subsequently, changes in canal volume, prepared canal surface, isthmus volume, debris volume, and transportation were measured after the use of each instrument system, and the apical diameter was designated as the maximum mesiodistal diameter of the canal at the section 1 mm from the apex.

Matched images of the surface areas of the canals, before and after preparation, were examined to evaluate three-dimensionally the amount of instrumented area. Prepared root canal surface was expressed as a percentage of the number of affected surface voxels to the total number of surface voxels. Isthmuses were segmented according to the principle that isthmuses with mesiodistal widths equal to one quarter of the diameter of the main canals were identified as the starting and ending points in the oro-palatal direction,1 and the volumes of the isthmuses were calculated. The debris after instrumentation was identified and calculated as follows: black-colored voxels were identified as soft tissue, liquid, or air (black color) in the preoperative scan; in the second scan, voxels that had changed from black to white were assumed to be dentin debris. The amount of transportation at the 1-, 2-, 3-, 4-, 6-, and 10-mm points from the apical foramen was measured from the cross-sectional images before and after the canal preparation according to the following method modified by Gergi et al.25 The formula:

\[
(\frac{a_1}{C_0} - \frac{a_2}{C_0}) - (\frac{b_1}{C_0} - \frac{b_2}{C_0})
\]

was used to calculate the amount of transportation: \(a_1\) was the shortest distance from the mesial edge of the uninstrumented canal to the mesial edge of the root, and \(a_2\) was the shortest distance from the mesial edge of the instrumented canal to the mesial edge of the root; \(b_1\) was the shortest distance from the distal edge of the instrumented canal to the distal edge of the root, and \(b_2\) was the shortest distance from the distal edge of the instrumented canal to the distal edge of the root.

Because normality assumptions were verified using the Shapiro-Wilk test, the results were statistically analyzed using a \(t\) test between groups I and II, and a paired \(t\) test within groups. The null hypothesis was set at 5%.

Results

The average preoperative volumes (mm\(^3\)) of canals in Groups I and II were 15.303 ± 2.900 and 14.589 ± 3.644, respectively, and the volumes of the corresponding isthmuses were 0.931 ± 0.221 and 0.948 ± 0.360, respectively. The apical diameter (mm) of the first, second, and third scans was 0.132 ± 0.048, 0.327 ± 0.098, and 0.404 ± 0.074, respectively, for Group I, and 0.138 ± 0.059, 0.323 ± 0.090, and 0.408 ± 0.087, respectively, for Group II. There was no significant difference between groups regarding canal and
isthmus volume and apical diameter before treatment ($P > 0.05$).

No significant difference between Groups I and II was observed regarding the increases in volume and prepared rate ($P > 0.05$). Paired $t$ tests showed that coronal and middle prepared areas of the canal obtained after using SAF in Group I were statistically higher than those obtained using ISO files, but no difference was observed for the apical prepared area within this group. By contrast, additional preparation with SAF following PTU resulted in a significant increase in the apical prepared area, while no difference was observed for the coronal and middle parts. Within-group isthmus volumes were significantly increased after the use of SAF instruments for both groups ($P < 0.05$; Table 1). Compared with using the glidepath system, relatively less debris was present after using SAF instruments, although no significant difference was noted ($P > 0.05$; Table 1). Neither technique removed debris completely from the canals or the isthmuses.

Canal transportation was $0.046 \pm 0.059$ and $0.122 \pm 0.110$ in Groups I and II, respectively. At 1 mm, 2 mm, 3 mm, 4 mm, 6 mm, and 10 mm from the apical foramen, the mean degrees of buccal or palatal canal transportation observed after using various instruments are shown in Figure 1. There was no difference in the extent of transportation between buccal and palatal canals ($P > 0.05$). ISO and SAF systems deviated less from the center as compared to PTU and SAF systems at most of the sections, especially at sections in the middle part of the canal ($P < 0.05$; Figure 1).

Color-coded superimposed models of pre- and post-operative MCT data demonstrated an obvious increase in

**Figure 2** Three-dimensional reconstruction and superimposition of MCT scans of the root canal system of the maxillary premolar under investigation. (A,B) The initial canal configuration showing the complex isthmus area. The top root canals were prepared using (A1) ProTaper Universal as glidepath enlargement and then using (A2) SAF, and the bottom root canals were prepared using (B1) ISO files as glidepath enlargement and then using (B2) SAF. (A1,2, B1,2) Superimposed MCT reconstructions after instrumentation. Prepared areas, untouched areas, and hard-tissue debris are indicated in green, red, and yellow, respectively. ISO = ISO NiTi files; MCT = microcomputed tomography; SAF = self-adjusting files.
prepared canal surface and volume, and a decrease in debris when canals were prepared using SAF in Group I. In Group II, the morphology slightly changed and the amount of debris notably decreased (Figures 2 and 3).

Discussion

The SAF is a hollow file designed as a compressible, thin-walled, pointed cylinder composed of a 120-mm-thick nickel-titanium lattice. This tool adapts itself to the shape of a canal both longitudinally and along the cross-section, and has many other advantages, including less instrument separation and less preparation error. According to the instructions, glidepath must be confirmed to size #20 or #30 before using SAF. In previous studies, K files were usually used for glidepath enlargement, and changes in the root canal before and after SAF preparation were seldom investigated. Compared with K files, rotary files had been found to be more efficient, with less transportation observed. Therefore, ISO rotary files were chosen, and the shaping and cleaning efficiency of ISO and SAF were evaluated and compared with that of PTU and SAF in this study. MCT scanning has been increasingly used in recent years for studies of root canal preparation with different chemomechanical techniques. Exact superimposition ensured by software offers a noninvasive reproducible technique for three-dimensional quantitative and qualitative assessment of increases in volume and debris in prepared areas of root canal systems.

The ISO and SAF system preparation removed a relatively uniform dentin layer from the canal circumference, and also deviated less at most sections relative to the PTU or PTU and SAF system. After instrumentation with larger taper PTU files, adjuvant preparation with SAF exhibited little impact on the morphology of the middle and coronal parts. This result suggested that assisted preparation with SAF after ISO files can personalize the shaping and cleaning of the middle and coronal parts of the canal according to the anatomy of each root canal, thereby reducing the occurrence of complications, such as transportation, caused by larger tapered files.

It was reported that SAF preparation can result in insufficient apical preparation and inadequate apical irrigation. Consistent with these results, we also observed less prepared areas in the apical part of the canal compared with the coronal and middle parts. However, adjuvant preparation with SAF after PTU significantly increased the preparation rate of the apical part. The increased preparation rate might be due to the expanded coronal part produced by larger tapered files, which provided better access for apical preparation with SAF. Therefore, the PTU and SAF hybrid techniques could be recommended to prepare wider root canals.

Regarding the preparation of isthmuses, paired t tests showed that volumes of isthmuses after SAF preparation increased significantly in both groups. Although debris was found to accumulate in isthmuses after rotary file preparation in several studies, only a few evaluated the volume changes of isthmus after mechanical preparation. Markvart et al. found that the unprepared areas of isthmuses were 17.6%, and a minor reduction in the isthmus volume was noticed, which might result from the accumulation of debris and crossing the isthmus of root canal instruments. The canals selected for the present study mainly contained type IV isthmuses. The SAF adjusted itself three-dimensionally to the shape of the root canal and removed the dentin from the tear-shaped areas of the isthmuses. However, the SAF reduced the accumulation of hard-tissue debris in the isthmus due to the application of continuous irrigation through the hollow file, which resulted in the volume increase observed in the isthmuses.
It was recently reported that the use of SAF in the mesial roots of mandibular molars resulted in less hard-tissue debris accumulation as compared to when rotary files were used. This could be attributed either to avoiding rotary motion in the canals or the continuous irrigation applied through the hollow file throughout the procedure, or both. However, in our study, no significant difference was observed in debris volume, either within or between groups. The possible reason for this might be that the isthmuses of maxillary premolars are wider and more mutable than those of mandibular molars, which results in amounts of debris too low to compare.

In conclusion, the SAF following glidepath enlargement with ISO NiTi files improved the preparation of the coronal and middle part of the canals, and resulted in less transport. Adjuvant preparation with SAF after the use of large-tapered instruments increased the apical prepared area, and can be recommended to prepare wider root canals. Both techniques improved the debridement efficacy with relation to isthmuses, but a small amount of debris was also observed in the isthmuses after both techniques.

Conflicts of interest

The authors have no conflicts of interest relevant to this article.

Acknowledgments

This study was supported by Key Clinical Program of the Ministry of Health, China [No. (2010) 439].

References

1. Adcock JM, Sidow SJ, Looney SW, et al. Histologic evaluation of canal and isthmus debridement efficacies of two different irrigant delivery techniques in a closed system. J Endod 2011;37:544–8.
2. Paqué F, Balmer M, Attin T, Peters OA. Preparation of oval-shaped root canals in mandibular molars using nickel-titanium rotary instruments: a microcomputed tomography study. J Endod 2010;36:703–7.
3. Susin L, Liu Y, Yoon JC, et al. Canal and isthmus debridement efficacies of two irrigant agitation techniques in a closed system. Int Endod J 2010;43:1077–90.
4. Gu L, Wei X, Ling J, Huang X. A microcomputed tomographic study of canal isthmuses in the mesial root of mandibular first molars in a Chinese population. J Endod 2009;35:353–6.
5. Verma P, Love RM. A Micro CT study of the mesiobuccal root canal morphology of the maxillary first molar tooth. Int Endod J 2011;44:210–7.
6. Mannocci F, Peru M, Sherriff M, Cook R, Pitt Ford TR. The isthmuses of the mesial root of mandibular molars: a microcomputed tomographic study. Int Endod J 2005;38:558–63.
7. Peters OA, Peters CJ, Schönenberger K, Barbakow F. ProTaper rotary root canal preparation: effects of root canal anatomy on final shape analyzed by micro CT. Int Endod J 2003;36:86–92.
8. Endal U, Shen Y, Knut A, Gao Y, Haapasalo M. A high-resolution computed tomographic study of changes in root canal isthmus area by instrumentation and root filling. J Endod 2011;37:223–7.
9. Metzger Z, Teperovich E, Zary R, Cohen R, Hof R. The self-adjusting file (SAF). Part 1: respecting the root canal anatomy – A new concept of endodontic files and its implementation. J Endod 2010;36:679–90.
10. Peters OA, Boessler C, Paqué F. Root canal preparation with a novel nickel-titanium instrument evaluated with micro-computed tomography: canal surface preparation over time. J Endod 2010;36:1068–72.
11. Versiani MA, Pécora JD, de Sousa-Neto MD. Flat-oval root canal preparation with self-adjusting file instrument: a microcomputed tomography study. J Endod 2011;37:1002–7.
12. Peters OA, Paqué F. Root canal preparation of maxillary molars with the self-adjusting file: a microcomputed tomography study. J Endod 2011;37:53–7.
13. Akman M, Akbulut MB, Aydinkelba HA, Belli S. Comparison of different irrigation activation regimes and conventional irrigation techniques for the removal of modified triple antibiotic paste from root canals. J Endod 2015;41:720–4.
14. Solomonov M, Paqué F, Fan B, Eliat Y, Berman LH. The challenge of C-shaped canal systems: a comparative study of the self-adjusting file and ProTaper. J Endod 2012;38:209–14.
15. Siqueira Jr JF, Alves FR, Almeida BM, de Oliveira JC, Rôças IN. Ability of chemomechanical preparation with either rotary instruments or self-adjusting file to disinfect oval-shaped root canals. J Endod 2010;36:1860–5.
16. Alves FR, Almeida MA, Neves MA, Rôças IN, Siqueira Jr JF. Time-dependent antibacterial effects of the self-adjusting file used with two sodium hypochlorite concentrations. J Endod 2011;37:1451–5.
17. De-Deus G, Souza EM, Barino B, et al. The self-adjusting file optimizes debridement quality in oval-shaped root canals. J Endod 2011;37:701–5.
18. Adigüzelo O, Yigit-Ozer S, Kaya S, Uysal I, Ganidagli-Ayaz S, Akkus Z. Effectiveness of ethylenediaminetetraacetic acid (EDTA) and MTAD on debris and smear layer removal using a self-adjusting file. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2011;112:803–8.
19. Kaya S, Yigit-Ozer S, Adigüzel O. Evaluation of radicular dentin erosion and smear layer removal capacity of self-adjusting file using different concentrations of sodium hypochlorite as an initial irrigant. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2011;112:524–30.
20. Metzger Z, Teperovich E, Cohen R, Zary R, Paqué F, Hülsmann M. The self-adjusting file (SAF). Part 3: removal of debris and smear layer – A scanning electron microscope study. J Endod 2010;36:697–702.
21. Marending M, Schicht OO, Paqué F. Initial apical fit of K-files versus LightSpeed LSX instruments assessed by microcomputed tomography. Int Endod J 2012;45:169–76.
22. Paranjpe A, de Gregorio C, Gonzalez AM, et al. Efficacy of the self-adjusting file system on cleaning and shaping oval canals: a microbiological and microscopic evaluation. J Endod 2012;38:226–31.
23. Hsu YY, Kim S. The resected root surface. The issue of canal isthmuses. Dent Clin North Am 1997;41:529–40.
24. Schneider SW. A comparison of canal preparations in straight and curved root canals. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1971;32:271–5.
25. Gergi R, Rjeily JA, Sader J, Naaman A. Comparison of canal preparations in straight and curved root canals. Int Endod J 2012;45:1451–8.
26. Hof R, Perevalov V, Eltanani M, Zary R, Metzger Z. The self-adjusting file (SAF). Part 2: mechanical analysis. J Endod 2010;36:891–6.
27. Paqué F, Al-Jadaa A, Kfir A. Hard-tissue debris accumulation created by conventional rotary versus self-adjusting file instrumentation in mesial root canal systems of mandibular molars. Int Endod J 2012;45:413–8.
28. Markvart M, Darvann TA, Larsen P, Dalstra M, Krellborg S, Björndal L. Micro-CT analyses of apical enlargement and molar root canal complexity. Int Endod J 2012;45:273–81.