Brief Report

Comprehensive Clinical and Genetic Analysis of CHEK2 in Croatian Men with Prostate Cancer

Kira Kirchner 1,†, Marija Gamulin 2,†, Tomislav Kulis 3, Bianca Sievers 1, Zeljko Kastelan 3 and Davor Lessel 1,*

1 Institute of Human Genetics, University Medical Center Hamburg-Eppendorf, 20246 Hamburg, Germany
2 Department of Oncology, University Hospital Center Zagreb, University of Zagreb School of Medicine, 10000 Zagreb, Croatia
3 Department of Urology, University Hospital Centre Zagreb, University of Zagreb School of Medicine, 10000 Zagreb, Croatia
* Correspondence: d.lessel@uke.de
† These authors contributed equally to this work.

Abstract: Germline pathogenic and likely pathogenic (P/LP) variants in CHEK2 have been associated with increased prostate cancer (PrCa) risk. Our objective was to analyze their occurrence in Croatian PrCa men and to evaluate the clinical characteristics of P/LP variant carriers. Therefore, we analyzed CHEK2 in 150 PrCa patients unselected for age of onset, family history of PrCa or clinical outcome, and the frequency of identified variants was compared to findings in 442 cancer-free men, of Croatian ancestry. We identified four PrCa cases harboring a P/LP variant in CHEK2 (4/150, 2.67%), which reached a statistical significance (p = 0.004) as compared to the control group. Patients with P/LP variants in CHEK2 developed PrCa almost 9 years earlier than individuals with CHEK2 wild-type alleles (8.9 years; p = 0.0198) and had an increased risk for lymph node involvement (p = 0.0047). No association was found between CHEK2 status and further clinical characteristics, including the Gleason score, occurrence of aggressive PrCa, the tumor or metastasis stage. However, carriers of the most common P/LP CHEK2 variant, the c.1100delC, p.Thr367Metfs15*, had a significantly higher Gleason score (p = 0.034), risk for lymph node involvement (p = 0.0001), and risk for developing aggressive PrCa (p = 0.027). Thus, in a Croatian population, CHEK2 P/LP variant carriers were associated with increased risk for early onset prostate cancer, and carriers of the c.1100delC, p.Thr367Metfs15* had increased risk for aggressive PrCa.

Keywords: CHEK2; prostate cancer; aggressive prostate cancer

1. Introduction

CHEK2 (Checkpoint kinase 2), encoding the CHK2 protein, is a serine/threonine protein kinase and one of the key components of the DNA damage response (DDR) pathway. CHK2 has pleiotropic functions regulating cell cycle progression after endogenous or exogenous DNA damage or the blockade of DNA replication, thereby preventing the entry of damaged cells into mitosis [1]. Given its biological role, it is not unexpected that genetic studies have implicated CHEK2 as a multiorgan tumor susceptibility gene. Indeed, CHEK2 has been associated with an increased risk for testicular germ-cell tumors, breast cancer, and colorectal cancers [2–4].

Moreover, CHEK2 has been implicated in conferring an increased risk of prostate cancer (PrCa) [5], especially early onset [6] and metastatic PrCa [7]. In light of the therapeutic actionability of pathogenic and likely pathogenic (P/LP) CHEK2 variants in metastatic castration-resistant prostate cancer (mCRPC) [8], the identification of germline variants that not only predict the disease risk, but also have implications for the clinical outcome, has increasing relevance, especially since recent large-scale analyses have demonstrated that P/LP variants in cancer susceptibility genes are identified in more than 50% of the cases who do not meet formal germline testing criteria [9].
Prompted by our recent findings where we established CHEK2 as the first moderate-penetrance testicular germ-cell tumor (TGCT) susceptibility gene and identified the low-penetrance CHEK2 p.Ile157Thr variant as a likely founder missense variant in Croatian men [2], this study aimed to evaluate the PrCa risk conferred by the P/LPs in CHEK2 along with its clinical implications in a Croatian population.

2. Materials and Methods

2.1. Study Participants

A total of 150 Croatian patients with a primary prostate cancer (Supplementary Table S1), unselected for age of onset, family history of PrCa or clinical outcome, were ascertained within a single year by the Departments of Oncology and Urology at the Clinical Hospital Centre Zagreb, Croatia. Clinical data on all cases were uniformly obtained from an oncology specialist using a structured clinical table. Aggressive cases (n = 25) were defined as previously described [10]. Briefly, the following criteria defined an aggressive PrCa: (i) cause of death (regardless of T-stage or Gleason score), (ii) stage 4 (regardless of T-stage or Gleason score), or (iii) T-stage 3 and Gleason score ≥ 8. The control group consisted of 442 cancer-free Croatian men in whom we had previously analyzed the occurrence of P/LP variants in CHEK2 [2]. Written informed consent was obtained from each individual after a detailed explanation of the purpose of the study. The study was approved by the ethics committee of the Clinical Hospital Centre Zagreb, Zagreb, Croatia, 8.1-19/125-3.

2.2. Genetic Analyses

The extraction of genomic DNA from whole-blood samples was performed by a standard salting-out method, as described previously [11]. A PCR amplification and Sanger sequencing of all 14 coding exons of CHEK2 was performed as previously described [2]. In addition to the previously reported protocols for the detection of exon 9_10 deletion [2], the occurrence of this deletion was verified by PCR amplification as described previously [12]. The clinical interpretation of identified CHEK2 variants followed the American College of Medical Genetics and Genomics (ACMG) criteria, as previously described [2].

2.3. Statistical Analyses

For the statistical analysis, an unpaired t-test or a Fischer’s exact test were performed, using GraphPad Prism 8.

3. Results

A detailed clinicopathological characterization of the 150 Croatian prostate cancer (PrCa) patients included in the study is shown in Table 1 and in more detail in Supplementary Table S1. In summary, the mean age at onset of PrCa was 68.17 years (ranging from 48 to 84 years). Eighteen cases developed additionally another tumor (12%). Regarding the PrCa, the mean size of the primary tumor (T-stage) was 2.10, 18 cases developed a primary metastasis (12%), and in 4 cases the PrCa was spread to lymph nodes (2.7%), whereas the mean Gleason score was 6.99 (ranging from 4 to 10). Twenty-five cases developed aggressive PrCa (16.7%), out of which eleven deceased due to PrCa-related complications (7.3%). A family history of tumor occurrence was available for only 119 PrCa patients (79.3%). Notably, none had a family history of PrCa and six had a family history of any malignancy (5%). Out of the latter, three had a mother who had developed a breast cancer (2.5%).

Utilizing a direct Sanger sequencing approach followed by a PCR amplification for the detection of exon 9_10 deletion, we identified pathogenic or likely pathogenic (P/LP) CHEK2 germline variants in four PrCa cases (4/150, 2.67%). In more detail, two cases harbored the c.1100delC, p.Thr367Metfs15*, and two cases harbored the c.1169A>C, p.Tyr390Ser (Figure 1). Comparing these data to the control group [2], the difference reached statistical significance (4/150 vs. 0/442, p = 0.004; Table 2). We additionally identified two cases harboring the low-penetrance variant c.470T>C, p.Ile157Thr. However, contrary to
our findings in TGCT’s [2], the p.Ile157Thr among Croatian men with PrCa was statistically insignificant compared with ethnically matched noncancer controls (2/150 vs. 6/442, p = 0.99; Table 2). Moreover, we identified a single PrCa case harboring c.1270T>C, p.Y424H, a variant classified as a variant of unknown significance (Figure 1).

Table 1. Clinicopathological characteristics of the 150 patients included in this study.

| Status   | Total | Cases n = 150 |
|----------|-------|---------------|
| Mean age at diagnosis (mean +/− SEM; range) | 68.17 (+/−0.62; 48–84) |
| Cases with other tumors (%) | 18 (12) |
| T-stage (%) |       |               |
| T1       | 26 (17.33) |
| T2       | 83 (55.33) |
| T2/T3    | 2 (1.33)   |
| T3       | 32 (21.33) |
| T4       | 4 (2.66)   |
| Unknown  | 3 (2)      |
| Mean (+/−SEM; range) | 2.10 (+/−0.06) |
| Presence of metastasis (M-stage; %) |       |
| Yes: 132 (88) | |
| No: 18 (12) |
| Spread to lymph nodes (N-stage; %) |       |
| Yes: 142 (94.67) | |
| No: 4 (2.67) |
| Unknown: 4 (2.67) |
| Gleason score (%) |       |
| 4        | 1 (0.67)   |
| 6        | 51 (34)    |
| 7        | 58 (38.67) |
| 8        | 23 (15.33) |
| 9        | 10 (6.67)  |
| 10       | 3 (2)      |
| Unknown  | 4 (2.67)   |
| Mean (+/−SEM; range) | 6.99 (+/−0.08; 4–10) |
| Aggressive PrCa cases (%) | 25 (16.7) |
| Death due to PrCa (%) | 11 (7.3) |
| Family history of prostate cancer (available for cases) | 0 (119) |
| Family history of any tumor (available for cases; %) | 6 (119; 5) |

Figure 1. Location of identified CHEK2 variants. Schematic protein structure of CHK2 showing conserved domains. SQ/TQ: SQ/TQ cluster domain; FHA: forkhead-associated domain; kinase domain; NLS: nuclear localization signal. Pathogenic and likely pathogenic variants are shown in black above the protein structure; the other variants are shown in blue.
Table 2. Enrichment analysis: frequency of CHEK2 variants identified in Croatian prostate cancer patients compared to cancer-free Croatian men. P/LPs: pathogenic and likely pathogenic variants (c.1100delC, p.T367Mfs*15; c.1169A>C, p.Y390S). Significant p-values are depicted in bold.

| CHEK2 Variants | Genotype Count | OR (95% CI) | p-Value |
|----------------|----------------|------------|---------|
|                | Controls | Cases | Per Allele |
| P/LPs          | 0/0/442  | 0/4/146 | infinity (2.97 to infinity) | 0.004 |
| c.470T>C, p.I157T | 0/6/436 | 0/2/148 | 0.98 (0.2–4.08) | 0.99 |

We next performed a case–case analysis comparing the clinical characteristics of PrCa cases in our study, stratified by CHEK2 variant status. The mean age of onset for P/LPs carriers was 59.5 ± 6.75 compared to 68.4 ± 0.60 in noncarriers, a finding that reached statistical significance (8.9 years difference; p = 0.0198). Moreover, P/LP carriers had a statistically increased risk for lymph node involvement (0.25 ± 0.25 vs. 0.02 ± 0.01, p = 0.0047). A further comparison regarding the Gleason score, the occurrence of aggressive PrCa, at the tumor and metastasis stage, all revealed statistically insignificant p-values (Table 3). We further compared the clinical characteristics of the two carriers of the most common P/LP CHEK2 variant, c.1100delC, p.Thr367Mfs15* [13], to our PrCa cohort. The carriers of this variant had a 10-year earlier age of onset (58.0 ± 10 vs. 68.3 ± 0.61), a finding which reached a borderline significance (p = 0.056). Moreover, there was a statistically significant enrichment regarding the development of aggressive PrCa by the carriers of this variant (2/25, 8% vs. 0/125, 0%, p = 0.027), and the carriers had a higher Gleason score (8.5 ± 0.5 vs. 6.97 ± 0.08, p = 0.034) and increased risk for lymph node involvement (0.5 ± 0.5 vs. 0.02 ± 0.01, p < 0.0001).

Table 3. Clinicopathological characteristics of carriers of P/LP CHEK2 variants and noncarriers. Significant p-values are depicted in bold.

|                  | CHEK2 WT | CHEK2 P/LP Carriers | Chek2 non c.1100delC Carriers | CHEK2 c.1100delC Carriers | p-Value |
|------------------|----------|---------------------|-------------------------------|---------------------------|---------|
| Age of diagnosis | 68.4 ± 0.60 | 59.5 ± 6.75          | 68.3 ± 0.61                  | 58.0 ± 10                 | 0.0556  |
| Gleason score    | 6.97 ± 0.09 (142/144 patients) | 7.5 ± 0.65            | 6.965 ± 0.08                | 8.5 ± 0.5                | 0.0336  |
| T-stage          | 2.10 ± 0.06 (143/144 patients) | 2.25 ± 0.48           | 2.09 ± 0.06                 | 3                        | 0.0704  |
| M-stage          | 0.12 ± 0.03 | 0                   | 0.12 ± 0.03                | 0                        | 0.6020  |
| N-stage          | 0.02 ± 0.01 | 0.25 ± 0.25         | 0.02 ± 0.01                | 0.5 ± 0.5                | <0.0001 |
| Aggressive PrCa  | 23/146    | 2/4                 | 23/148                       | 2/2                      | 0.0268  |

4. Discussion

Prostate cancer (PrCa) is one of the most common male cancers worldwide, the second tumor-associated cause of death in men, and one of the most heritable human cancers [14,15]. In line with the strong heritability of PrCa, genome-wide association studies (GWAS) have reported more than 260 common low-penetrance genetic variants associated with PrCa risk [16]. Moreover, pathogenic or likely pathogenic germline variants (P/LPs) in several susceptibility genes have been shown to confer a moderate-to-high increase in risk for PrCa [17]. In the past, the analysis of the germline susceptibility genes was primarily utilized to identify individuals at high risk for developing cancer, who would benefit from enhanced cancer surveillance and early screening strategies. However, recent studies established the additional utility of germline genetic testing in PrCa both for the prediction of the clinical outcome [7,10] and for germline-directed treatment [8].
P/LPs variants in the tumor suppressor gene CHEK2 have been associated with increased PrCa risk [5,18,19] and belong to the group of germline variants with therapeutic actionability [8]. Previous studies, analyzing the implications of CHEK2 P/LP variants for clinical care have utilized cohorts enriched for cases with metastatic PrCa [7], lethal PrCa [13,20], aggressive PrCa [10], early onset [6], or primarily analyzing ethnic-specific founder effects [21].

Here, we performed the first comprehensive CHEK2 analysis in Croatian men with PrCa that were unselected for the age of onset, family history of PrCa, or its clinical outcome. According to recent epidemiological data, we included around 10% of all patients in Croatia who developed PrCa within one year [22,23]. We found CHEK2 P/LP in 2.67% of unselected PrCa cases that were significantly enriched compared to the control group. Thus, in line with previous studies [5–7,12], our findings further support CHEK2 being a moderate PrCa susceptibility gene. Moreover, despite the rather small sample size, our findings may be informative for the clinical management of PrCa cases, at least in Croatia. Namely, the cases harboring a CHEK2 P/LP presented almost 9 years earlier than noncarriers and had an increased risk for lymph node involvement. Furthermore, we identified two recurrent P/LPs, c.1100delC, p.Thr367Metfs15*, and c.1169A>C, p.Tyr390Ser, that should be included in the local PrCa germline screening, as both of these variants enable a genotype-directed treatment. Interestingly, out of these two P/LPs only the c.1100delC, p.Thr367Metfs15* was associated with aggressive PrCa, a finding previously observed in PrCa cases of European ancestry [13]. Moreover, we found that carriers of this variant had a significantly higher Gleason score, and similarly to overall P/LP carriers, a statistically increased risk for lymph node involvement.

One of the limitations of this study is the absence of familial PrCa cases along with cases having a family history of any malignancy. It is, however, worth noting that the mother of one of the cases harboring a P/LP variant developed breast cancer. Although her DNA was not available for testing, a recent study suggested that a family history of breast cancer and high Gleason score could be predictors to identify PrCa cases harboring P/LP germline variants in one of the DNA repair genes [24].

In addition, the low-penetrance variant c.470T>C, p.Ile157Thr, was not overrepresented in Croatian PrCa cases. Thus, our data might point to p.Ile157Thr being a specific risk factor for the testicular germ-cell tumors (TGCT) in the Croatian population.

5. Conclusions

Taken together, our study suggested CHEK2 P/LPs as a moderate PrCa-susceptibility gene associated with early age of onset in the Croatian population. In addition, we provided further evidence for the association of the c.1100delC, p.Thr367Metfs15* with aggressive PrCa.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/genes13111955/s1, Table S1: The 150 prostate cancer patients included in this study.

Author Contributions: K.K. and B.S. carried out the genetic analysis. M.G., T.K. and Z.K. collected patient material and clinical data. K.K. performed the statistical analysis. D.L. designed the study with input from the co-authors. K.K., M.G. and D.L. wrote the first draft of the manuscript. All authors revised the manuscript for intellectual content, approved the version to be published, and agreed to be accountable for all aspects of the work in ensuring that questions related to the integrity of any part of the work were appropriately investigated and resolved. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded through the German Cancer Aid (Deutsche Krebshilfe) to D.L.

Institutional Review Board Statement: The study was performed in accordance with the Declaration of Helsinki protocols and performed in accordance with protocols approved by the Ethics Committee of the Clinical Hospital Centre Zagreb, Zagreb, Croatia, 8.1-19/125-3.
Informed Consent Statement: All biological samples were obtained following written informed consent from the parents of the affected individuals.

Data Availability Statement: Not applicable.

Acknowledgments: We thank all individuals for their participation in the study.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Zannini, L.; Delia, D.; Buscemi, G. CHK2 kinase in the DNA damage response and beyond. *J. Mol. Cell Biol*. 2014, 6, 442–457. [CrossRef] [PubMed]
2. AlDubayan, S.H.; Pyle, L.C.; Gamulin, M.; Kulis, T.; Moore, N.D.; Taylor-Weiner, A.; Hamid, A.A.; Reardon, B.; Wubbenhorst, B.; Godse, R.; et al. Association of Inherited Pathogenic Variants in Checkpoint Kinase 2 (CHEK2) With Susceptibility to Testicular Germ Cell Tumors. *JAMA Oncol*. 2019, 5, 514–522. [CrossRef] [PubMed]
3. Cybulski, C.; Wokolorczyk, D.; Jakubowska, A.; Huzarski, T.; Byrski, T.; Gronwald, J.; Masoij, B.; Dębiak, T.; Górski, B.; Blecharz, P.; et al. Risk of breast cancer in women with a CHEK2 mutation with and without a family history of breast cancer. *J. Clin. Oncol*. 2011, 29, 3747–3752. [CrossRef] [PubMed]
4. Xiang, H.P.; Geng, X.P.; Ge, W.W.; Li, H. Meta-analysis of CHEK2 1100delC variant and colorectal cancer susceptibility. *Eur. J. Cancer* 2011, 47, 2546–2551. [CrossRef] [PubMed]
5. Dong, X.; Wang, L.; Taniguchi, K.; Wang, X.; Cunningham, J.M.; McDonnell, S.K.; Qian, C.; Marks, A.F.; Slager, S.L.; Peterson, B.J.; et al. Mutations in CHEK2 Associated with Prostate Cancer Risk. *Am. J. Hum. Genet*. 2003, 72, 270–280. [CrossRef]
6. Cybulski, C.; Wokolorczyk, D.; Kluzniak, W.; Jakubowska, A.; Gorski, B.; Gronwald, J.; Huzarski, T.; Kashyap, A.; Byrski, T.; Dębniak, T.; et al. An inherited NBK mutation is associated with poor prognosis prostate cancer. *Br. J. Cancer* 2013, 108, 461–468. [CrossRef]
7. Pritchard, C.C.; Mateo, J.; Walsh, M.F.; De Sarkar, N.; Abida, W.; Beltran, H.; Garofalo, A.; Gulati, R.; Carreira, S.; Eeles, R.; et al. Inherited DNA-Repair Gene Mutations in Men with Metastatic Prostate Cancer. *N. Engl. J. Med*. 2016, 375, 443–453. [CrossRef]
8. De Bono, J.; Mateo, J.; Fizazi, K.; Saad, F.; Shore, N.; Sandhu, S.; Chi, K.N.; Sartor, O.; Agarwal, N.; Olmos, D.; et al. Olaparib for Metastatic Castration-Resistant Prostate Cancer. *N. Engl. J. Med*. 2020, 382, 2091–2102. [CrossRef]
9. Mandelker, D.; Zhang, L.; Kemel, Y.; Stadler, Z.K.; Joseph, V.; Zehir, A.; Pradhan, N.; Arnold, A.; Walsh, M.F.; Liying, Z.; et al. Mutation Detection in Patients With Advanced Cancer by Universal Sequencing of Cancer-Related Genes in Tumor and Normal DNA vs Guideline-Based Germline Testing. *JAMA* 2017, 318, 825–835. [CrossRef]
10. Nguyen-Dumont, T.; MacInnis, R.J.; Steen, J.A.; Theys, D.; Tsimiklis, H.; Hammet, F.; Mahmoodi, M.; Pope, B.J.; Park, D.J.; Mahmood, K.; et al. Rare germline genetic variants and risk of aggressive prostate cancer. *Int. J. Cancer* 2020, 147, 2142–2149. [CrossRef] [PubMed]
11. Lessel, D.; Gamulin, M.; Kulis, T.; Toliat, M.R.; Gracic, M.; Friedrich, K.; Žunec, R.; Balija, M.; Nürnberg, P.; Kastelan, Z.; et al. Replication of genetic susceptibility loci for testicular germ cell cancer in the Croatian population. *Carcinogenesis* 2012, 33, 1548–1552. [CrossRef]
12. Cybulski, C.; Wokolorczyk, D.; Huzarski, T.; Byrski, T.; Gronwald, J.; Gorski, B.; Dębniak, T.; Maseso, B.; Jakubowska, A.; Gliniewicz, B.; et al. A large germline deletion in the Chek2 kinase gene is associated with an increased risk of prostate cancer. *J. Med. Genet.* 2006, 43, 863–866. [CrossRef] [PubMed]
13. Wu, Y.; Yu, H.; Zheng, S.L.; Na, R.; Mamawala, M.; Landis, T.; Wiley, K.; Petkewicz, J.; Shah, S.; Shi, Z.; et al. A comprehensive evaluation of CHEK2 germline mutations in men with prostate cancer. *Prostate* 2018, 78, 607–615. [CrossRef] [PubMed]
14. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2019. *CA Cancer J. Clin.* 2019, 69, 7–34. [CrossRef] [PubMed]
15. Mucci, L.A.; Hjelmborg, J.B.; Harris, J.R.; Czene, K.; Havelick, D.J.; Scheike, T.; Graff, R.E.; Holst, K.; Möller, S.; Unger, R.H.; et al. Familial Risk and Heritability of Cancer Among Twins in Nordic Countries. *JAMA J. Am. Med. Assoc.* 2016, 315, 68–76. [CrossRef] [PubMed]
16. Contí, D.V.; Darst, B.F.; Moss, L.C.; Saunders, E.J.; Sheng, X.; Chou, A.; Schumacher, F.R.; Al Olama, A.A.; Benlloch, S.; Dadavan, T.; et al. Trans-ancestry genome-wide association meta-analysis of prostate cancer identifies new susceptibility loci and informs genetic risk prediction. *Nat. Genet.* 2021, 53, 65–75. [CrossRef]
17. Heifand, B.T.; Xu, J. Germline Testing for Prostate Cancer Prognosis. *Urol. Clin. N. Am.* 2021, 48, 401–409. [CrossRef]
18. Cybulski, C.; Huzarski, T.; Gorski, B.; Maseoj, B.; Mierzejewski, M.; Dębniak, T.; Gliniewicz, B.; Matyjasik, J.; Złowocka, E.; Kurzawski, G.; et al. A Novel Founder CHEK2 Mutation is Associated with Increased Prostate Cancer Risk. *Cancer Res.* 2004, 64, 2677–2679. [CrossRef]
19. Seppälä, E.H.; Ilonen, T.; Mononen, N.; Autio, V.; Rökk, A.; Matikainen, M.P.; Tammela, T.L.J.; Schleutker, J. CHEK2 variants associate with hereditary prostate cancer. *Br. J. Cancer* 2003, 89, 1966–1970. [CrossRef]
20. Rantapero, T.; Wahlfors, T.; Kähler, A.; Hultman, C.; Lindberg, J.; Tammela, T.L.J.; Nykter, M.; Schleutker, J.; Wiklund, F. Inherited DNA Repair Gene Mutations in Men with Lethal Prostate Cancer. *Genes* 2020, 11, 314. [CrossRef]
21. Brandão, A.; Paulo, P.; Maia, S.; Pinheiro, M.; Peixoto, A.; Cardoso, M.; Silva, M.P.; Santos, C.; Eeles, R.A.; Kote-Jarai, Z.; et al. The CHEK2 Variant C.349A>G Is Associated with Prostate Cancer Risk and Carriers Share a Common Ancestor. Cancers 2020, 12, 3254. [CrossRef]

22. Kuliš, T.; Krhen, I.; Kaštelan, Ž.; Znaor, A. Trends in prostate cancer incidence and mortality in Croatia, 1988 to 2008. Croat. Med. J. 2012, 53, 109–114. [CrossRef]

23. Reljic, A.; Cukelj, P.; Tomaskovic, I.; Ruzic, B.; Sekerija, M. Epidemiology of Prostate Cancer in Croatia—Situation and Perspectives. Acta Clin. Croat 2018, 57 (Suppl. S1), 27–34. [CrossRef]

24. Giri, V.N.; Hegarty, S.E.; Hyatt, C.; O’Leary, E.; Garcia, J.; Knudsen, K.E.; Kelly, W.K.; Gomella, L.G. Germline genetic testing for inherited prostate cancer in practice: Implications for genetic testing, precision therapy, and cascade testing. Prostate 2019, 79, 333–339. [CrossRef] [PubMed]