Progress in Research on the Role of FGF in the Formation and Treatment of Corneal Neovascularization

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Corneal neovascularization (CNV) is a sight-threatening disease usually associated with inflammatory, infectious, degenerative, and traumatic disorders of the ocular surface. Fibroblast growth factor (FGF) family members play an important role in angiogenesis to induce corneal neovascularization, which significantly affects the differentiation, proliferation, metastasis, and chemotaxis of vascular endothelial cells. Both acidic fibroblast growth factor (aFGF) and basic fibroblast growth factor (bFGF) demonstrate positive staining in capillaries and induce corneal stromal cells. The anabolism of endothelial cells is induced by bFGF in corneal neovascularization. FGFs exert their effects via specific binding to cell surface-expressed specific receptors. We believe that both anti-FGF antibodies and anti-FGF receptor antibodies represent new directions for the treatment of CNV. Similar to anti-vascular endothelial growth factor antibodies, subconjunctival injection and eye drops can be considered effective forms of drug delivery.

Keywords: corneal neovascularization, fibroblast growth factor, development, treatment, drug delivery

CORNEAL NEOVASCULARIZATION

The incidence rate of corneal neovascularization (CNV), a sight-threatening condition, is approximately 1.4 million patients per year (Bonini, 2016). It is usually associated with inflammatory, traumatic, or infectious disorders of the ocular surface. Corneal neovascularization can cause vision damage and even blindness, the rate of which is as high as 57.4% (Chang et al., 2001). The cornea needs to be transparent to allow the passage of light into the retina. When CNV occurs, abnormal blood vessels directly block light, indirectly diffract light as canals for inflammatory cells, and damage the structure of the cornea by depositing lipids and proteins into the corneal stroma. Various mechanisms of corneal neovascularization have been studied. A balance exists between angiogenic factors (such as fibroblast growth factor and vascular endothelial growth factor) and anti-angiogenic factors (such as angiotatin, endostatin, and pigment epithelium-derived factor) in the cornea (Senturk et al., 2016). An imbalance leads to CNV, which can be divided into superficial neovascularization and deep stromal neovascularization. In the normal cornea, heparan sulfate prevents the release of potent angiogenic cytokines, such as fibroblast growth factor (FGF) and vascular endothelial growth factor (VEGF). However, under stimulation, cytokines may be released, causing an imbalance (Gurung et al., 2018).
FGF

FGF

Fibroblast growth factor participates in cell proliferation, migration, and tissue repair in adults (Itoh and Ornitz, 2004). FGFs can stimulate many cell types, including endothelial cells and nerve cells. The FGF family comprises 22 members (FGF1-23, except FGF15 because mouse FGF15 is the orthologue of human and chick FGF19) (Beenken and Mohammadi, 2009). Many of these FGFs, particularly FGF-1 (acidic FGF) and FGF-2 (basic FGF), have been shown to influence angiogenesis in several tissues in vivo (Table 1) by acting on endothelial cells. Different factors affect FGFs in different tissues. FGFs are produced by endothelial cells and are stored in the extracellular matrix. They show a high affinity for heparin (Klagsbrun, 1990).

FGFs activate transmembrane tyrosine kinases and their coupled intracellular signalling pathways to stimulate biological activities and transduce extracellular signals through the PI3K, MAPK, and phospholipase Cγ (PLCγ) pathways (Klagsbrun, 1990; Zhou et al., 2009; Francavilla et al., 2013; Huang et al., 2016). A dual-receptor system comprising a family of four receptor tyrosine kinases (FGFRs) and heparan sulfate proteoglycans (HSPGs) mediates the stimulation of cellular metabolism by FGFs. Thus, many of the FGF isoforms bind several FGFs. HSPG receptors may provide additional specificity (Klagsbrun, 1990). ERK1/2, JNK1/2, and p38 alpha/beta are three main MAPK subgroups (Zhang et al., 2009). Phospholipase C can receive the signal from FGFs and then activate PLCγ1 to induce cell proliferation and migration in vascular smooth muscle cells by cleaving the phospholipid phosphatidylinositol 4,5-bisphosphate into diacylglycerol and inositol 1,4,5-triphosphate (Michel, 1998). Studies have suggested that heparan may modulate the activity of FGFs (Ornitz, 2000). Additionally, studies have suggested that membrane-type 1 matrix metalloproteinase (MT1-MMP) plays an important role in modulating FGF-mediated signal pathways (Zhou et al., 2000). It controls FGF signalling by reducing the amount of FGF bound to the cell surface, increasing the proteolysis of FGFs and the downregulation of FGFR-1 and -4 (Tassone et al., 2015).

**TABLE 1 | Fibroblast growth factors (FGFs) associated with angiogenesis.**

| FGFs | Pathway/influencing factor | Related tissue/disease |
|------|---------------------------|-----------------------|
| FGF-1 | S156C-TIMP3 mutation | Choroid (Loren et al., 2009; de Oliveira Dias et al., 2011) |
| | PS3 | Inflamed tissue (Zhou et al., 2009) |
| | Erk and MMP-7 | Colon cancer (Jin et al., 2016) |
| | S100A13 | Endometriosis (Francavilla et al., 2013) |
| FGF-2 | VEGF | Corneal, choroid, and retina (Senturk et al., 2018) |
| | NDY1/KDM2B-mir101-EZH2 | Tumor tissue (Akpek et al., 2004) |
| | Interleukin-1β | Chondrocytes (Baradaran-Rafii et al., 2019) |
| | AKT/MMP-2 | Human umbilical vein (Mimura et al., 2011) |
| FGF-3 | Erk and MMP-7 | Tumor tissue (Nor et al., 2001) |
| FGF-8 | Co-expression of VEGF | Prostate cancer (Stevenson et al., 2012) |
| FGF-9 | VEGF-A | Bone (Kooijvink et al., 1996) |
| FGF-18 | Wnt/β-catenin | Hepatocellular carcinoma (Zheng et al., 2003) |
| FGF-21 | Dynamin-2 and Rab6 | Kidney (Kim et al., 2013) |

**FGFs in the Eye**

mRNAs encoding basic FGF are produced by corneal epithelial cells, stromal fibroblasts, and corneal endothelial cells (Oladiippo et al., 2014). FGF-2 may have different functions in the three primary cell types of the cornea. Studies have shown that in corneal epithelial cells, FGF-2 stimulates the proliferation of corneal epithelial cells and increases the rate of epithelial wound healing through autocrine and paracrine effects (Kanayama et al., 2007). Rajesh et al. showed that FGF-2 promotes corneal stromal wound healing by increasing cellular proliferation (stimulated by TGF-beta via paracrine actions) (Moioli et al., 2006) and cellular motility (significant enhancement of stromal fibroblast motility by 100 ng/ml FGF-2 in animal experiments) (Rao et al., 1992). Cdc42 activation, Rho inactivation, and the phosphatidylinositol 3-kinase pathway in corneal endothelial cells (CECs) are essential for FGF-2-induced wound healing (Lee and Kay, 2006), and endothelial mesenchymal transformation can be mediated by FGFs in CECs.

Additionally, FGFs can mediate the proliferation and regeneration of the lens (FGF-1 and FGF-2) (Zhu et al., 2012) and retinal cells (FGF-2, FGF-5, and FGF-9) through different signalling pathways (Gong et al., 2014). FGF-9, FGF-21, and FGF-23 are also expressed in choroidal endothelial cells and affect choroid plexus epithelial cell behaviour (Loren et al., 2009; de Oliveira Dias et al., 2011).

FGFs also play an important role in early mammalian eye development. The neuroepithelium of the optic vesicle separates into NR and RPE domains in a manner mediated by extrinsic factors that emanate from the surface ectoderm, for which fibroblast growth factors are prime candidates (Bassnett and Sikic, 2017).

**FGFS AND RELATED CORNEAL DISEASES**

Many FGFs affect angiogenesis, but FGF-2 is the most common isoform to be detected in the eye and to cause CNV in different eye diseases. One of these diseases is ocular chemical burn. We focused on alkali burns because of their severity. CNV can be caused by severe corneal alkali burn (Nominato et al., 2018), in which deep corneal stroma or full-thickness corneal injury is involved. In this situation, angiogenic factors play an important role in CNV. For example, experimental data have shown that on the second day after alkaline burn, b-FGF is obviously expressed in the corneal epithelium, substantia propria layer, and endothelium (Xiao et al., 2012). Additionally, studies have shown that the upregulation of related genes (microRNA-296) after alkali burn is positively correlated with the expression of related FGF isoforms (FGF-23). FGF-23 may influence corneal...
inflammatory responses by participating in cytokine-cytokine receptor interaction pathways (Hayashi et al., 1996). Keratitis is another important cause of corneal neovascularisation (Soiberman et al., 2017). In the initial stage of infection with herpes simplex virus type 1 (HSV-1), both virus and immune cells are present in the cornea. Meanwhile, various cytokines and growth factors (FGF-2 and Ang-2), which also permeate the cornea, lead to further inflammation. The origin of FGF-2 may not fibroblasts (keratocytes), epithelium, endothelium, blood endothelial cells, and lymphatic endothelial cells rather than leukocytes (Xu et al., 2019). In the late stage, immune cells and growth factors continue to be effective while the virus is removed from the cornea. Furthermore, FGF-2 mediates the expression of other cytokines (such as VEGF-A, IL-6, and Ang-2), which are crucial for HSV-1-induced corneal neovascularization (Klagsbrun, 1990).

Patients who undergo corneal transplantation can develop corneal neovascularization. Particularly, after high-risk keratoplasty, intense CNV outgrowth is a common phenomenon in the early postoperative period (Kelly et al., 2011). Corneal neovascularization can lead to an increased risk of graft rejection caused by an imbalance between angiogenic factors and anti-angiogenic factors. Additionally, FGF is an important angiogenic factor. Presently, no study has shown how FGFs cause CNV after corneal transplantation and which FGF isoform is involved, but this phenomenon warrants further research.

ROLE OF FGFS IN CNV FORMATION

Angiogenesis is the process by which new blood vessels grow by sprouting from established blood vessels (Carmeliet and Jain, 2011). In the cornea, these existing blood vessels can be part of the vascular plexus around the limbus of the anterior ciliary artery. Corneal angiogenesis occurs because of the release of proangiogenic factors, such as FGF-2, VEGF, and several other chemokines, from hypoxic or inflammatory cells. The binding of angiogenic factors to corresponding receptors on vascular endothelial cells leads to many events, including the following: (a) injury to endothelial cell junctions through the activation of non-receptor Src family kinases and increased expression of integrins; (b) the promotion of endothelial cell proliferation by mitogen-activated protein kinase and phosphoinositide 3’ kinase; (c) the secretion of metalloproteinases (MMPs) by endothelial cells to promote basal membrane disruption and pericyte detachment; and (d) blood vessel destabilization caused by the release of angiopoietin 2 (ANG 2) from endothelial cell granules (Figure 1). Additionally, murine tissue inhibitor of metalloproteinase-4 (TIMP-4) expression in the cornea may play a role in regulating extracellular matrix remodelling associated with corneal wound healing and angiogenesis; however, the mechanism is unclear.

Vascular endothelial cells are stimulated directly by FGFs from angiogenic tissues. FGF-producing cells also release FGFs through autocrine or nuclear actions (Figure 1), and secondary angiogenic factors then act on the vasculature. The vascular endothelium can induce the secretion of secondary regulatory molecules in the same way, and such secretion can also be influenced by other angiogenic factors, such as VEGF. The specific role of FGFs depends on the environment in which they are located (Beenken and Mohammadi, 2009).

PROGRESS IN FGF RESEARCH FOR CNV TREATMENT

Current Effective Treatments for CNV Presently, laser treatment, drug treatment, and surgical treatment are available to treat corneal neovascularization. Drugs used for treatment include glucocorticoids, immunosuppressive agents, and various vascular growth factor inhibitors. While the topical administration of steroidal anti-inflammatory drugs is the first-line treatment for corneal neovascularization, many side effects

![FIGURE 1 | Role of FGF-2 in corneal angiogenesis. (A) Activation for the destruction of endothelial cell junctions via MMP secretion. (B) Disruption of the basal membrane and pericytes via ANG2. (C) Secretion from vascular endothelial cells to induce blood vessel destabilization. (D) Corneal epithelial cell production through autocrine or nuclear actions during events such as trauma and hypoxia. (E) Limbus capillaries: activation of Src family kinases.](https://www.frontiersin.org)
may be beneficial for the recovery of vision. Researchers established a model of anti-FGF-2 antibody-treated mouse corneal neovascularization and then evaluated the corneal sensitivity and visual acuity of these mice to assess their functional vision. Time-course experiments revealed partial recovery of visual acuity in mice treated with an anti-FGF-2 antibody compared with control mice (Zheng et al., 2001). This finding supports anti-VEGF therapy for corneal neovascularization, but further experimental confirmation is needed.

**FGF as a Drug Source and the Administration Route**

Several anti-FGF2 monoclonal antibodies (mAbs) that can neutralize the activities of FGF2 *in vitro* and *in vivo* have been identified (Jin et al., 2016). However, to our knowledge, no anti-FGF2 mAb has entered clinical trials. Because of the similarity between VEGF and FGF antagonists, we can presume the likely effect of FGF antagonists in treating CNV based on the effects of anti-VEGF drugs.

A previous study demonstrated that the subconjunctival injection of anti-VEGF drugs (1.25 mg/0.05 ml bevacizumab) significantly reduces inflammation and fibroblast activity to inhibit corneal neovascularization in the cornea of a rat model of alkali burn. Rats in the test group were treated with a subconjunctival injection (Kim et al., 2013). Another researcher advocated combining subconjunctival (1.25 mg/0.05 ml) and intracorneal injection (1.25 mg/0.05 ml). They confirmed that combining subconjunctival and intracorneal injection did not damage corneal endothelial cells (Lichtinger et al., 2014). No clinical case featuring the administration of anti-FGF drugs through subconjunctival and intracorneal routes for the treatment of CNV exists, but we can refer to these modes of administration. Therefore, the drug concentration and side effects remain unclear, and further research is needed.

Recently, research on new eye drops has progressed. Considering the effects of anti-VEGF drugs, in the future, Avastin eye drops may be shown to exhibit the same anti-VEGF effects with fewer complications than intravitreal use (Kim et al., 2013). Bevacizumab eye drops seem to inhibit corneal neovascularization without inducing obvious corneal epithelial side effects. Bevacizumab eye drops are prepared by adding 0.9% physiological saline to the drug at a concentration of 5 mg/ml. The minimum storage temperature of the eye drops is –20°C (before opening), the maximum storage temperature is 4°C (after opening), and the maximum storage time is 14 days; after opening, they needs to be used within 1 day (Bock et al., 2008). Therefore, we believe that preparations of anti-FGF drugs as eye drops can be applied for the clinical treatment of corneal neovascularization. The effects of the FGF antagonist tecogalan sodium against corneal neovascularization were tested in animal experiments, and tecogalan sodium demonstrated dose-dependent antiangiogenic activity. The inhibitory effect of 100 ng terconazole sodium was weak, and the effect of 250 ng was marked, indicating that corneal neovascularization induced by bFGF can be inhibited by the topical instillation of tecogalan sodium (Murata et al., 1995). However, no data regarding the
new research direction. The authors believe that the route of administration of anti-FGF drugs will become a focus. The discovery of additional FGF antagonists has advantages. The authors also thank Professor Qianying Gao from the State Key Laboratory of Sun Yat-sen University.

AUTHOR CONTRIBUTIONS

MC and HZ participated in drafting the manuscript. LB, MZ, and JC provided technical assistance. MC and HZ revised the manuscript. HZ supervised the project and provided financial support. MC and HZ wrote the main part of the paper. All of the authors read and approved the final manuscript.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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