Tumefactive demyelination in a patient with relapsing-remitting MS on ocrelizumab

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Ocrelizumab is an intravenously infused humanized monoclonal antibody that induces antibody-dependent cellular cytolysis and complement-mediated lysis of CD20-expressing B cells. Ocrelizumab reduced gadolinium-enhancing activity 94% and 95% compared with subcutaneous interferon beta-1a in relapsing-remitting MS (RRMS).1 We report a case of tumefactive demyelination (TD) in an ocrelizumab-treated patient with RRMS.

Case report

The patient initially presented in April 2009, at age 27 years, with an episode of numbness in the right arm and torso. In the following months, he developed intermittent lower extremity weakness, requiring bilateral assistance to ambulate. In June 2010, he was diagnosed with RRMS after a brain MRI showed 20 nonenhancing lesions periventricularly and in the white matter on fluid-attenuated inversion recovery (FLAIR) sequence. Cervical and thoracic MRI revealed diffuse cord T2 hyperintensity, and a lumbar puncture (LP) showed 17 oligoclonal bands. The patient began interferon beta-1a subcutaneously but transitioned to cyclophosphamide 3 months later after requiring wheelchair assistance and developing bladder and bowel incontinence. While on cyclophosphamide, brain MRIs in 2011 and 2012 revealed 8 new enhancing lesions and a small enhancing left corona radiata lesion, respectively. He transitioned to dimethyl fumarate in October 2013. A brain MRI in September 2015 demonstrated new nonenhancing FLAIR hyperintensities. He transitioned to ocrelizumab in October 2017 receiving two 300 mg doses 2 weeks apart and then received a 600 mg maintenance dose in May 2018 (preinfusion serum CD19 = 0.6% lymphocytes).

One day before next scheduled infusion in November 2018, the patient reported exacerbated weakness, incoordination, and difficulty concentrating. Ocrelizumab was infused (preinfusion serum CD19 = 0.7% lymphocytes). In the following weeks, the patient developed intermittent weakness in his left upper extremity. He presented to the emergency department. While there, he experienced generalized tonic-clonic seizures and was sedated and intubated. After cessation of seizure activity and extubation, he was transferred to our institution. His neurologic examination was as previously observed except for increased weakness in the left upper extremity and ataxia with finger-nose-finger testing. An MRI demonstrated a 4.5 × 6.7 × 6.7 cm heterogeneous right parietal lobe hyperintensity, which exhibited peripheral enhancement on T1 sequences after gadolinium administration. Magnetic resonance (MR) perfusion imaging found decreased cerebral blood volume (CBV) in the area of interest (figure, B–D). An LP found the following: 16 white cells, 0 red cells, glucose = 63 mg/dL, and protein = 81 mg/dL. High-sensitivity JC virus (JCV) PCR assay was negative. No malignant cells were identified. A stereotactic-guided brain biopsy of the lesion was performed. Pathology was consistent with active demyelinating lesion (figure, E–H). Immunohistochemistry for polyoma virus was negative, excluding progressive multifocal leukoencephalopathy (PML). The patient received 3 days of 1 g IV methylprednisolone and then began oral methotrexate 20 mg weekly.

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Discussion

TD has not been reported with ocrelizumab or rituximab, a less humanized anti-CD20 monoclonal antibody. It has occurred with other MS drugs including fingolimod, natalizumab, and alemtuzumab. Our patient has particularly severe MS, as evinced by his rapidly worsening clinical course, high MRI disease burden, and continued accrual of new MRI lesions. It is possible that ocrelizumab was unable to control this patient’s fulminant disease. Alternately, it may be that a limited reconstitution of B cells (preinfusion serum CD19 = 0.7%) was sufficient for this patient to mount a potent autoimmune response. We believe that human antichimeric (HACA) antibodies or a direct effect from ocrelizumab can be excluded. One would expect little suppression of CD19 count if HACA antibodies were present. A more proximate relationship between infusion and TD development would be expected if ocrelizumab was causative, and mechanistically, this does not seem likely.

Questions emerge regarding treatment. Because inadequate serum suppression of B cells might have been a contributor, one wonders whether altering dosing to target continuous full CD20 B-cell suppression (preinfusion serum CD19 = 0) is appropriate for patients with particularly inflammatory disease. Methotrexate was combined with ocrelizumab based on a report by Tak et al. that methotrexate offered additional clinical benefit in rheumatoid arthritis without additional side effects. Further work would be beneficial to elucidate whether this is a viable strategy in ocrelizumab patients experiencing breakthrough disease.

Several factors contributed to the decision to perform a brain biopsy. MRI appearance was consistent with PML, tumeffactive MS, or glioma. In these conditions, MR perfusion CBV is typically increased rather than decreased, as seen in our patient. PML was entertained, given the patient’s previous immunosuppressive medication exposure and a false-negative rate.
with JCV CSF PCR testing. In addition, it seemed implausible that such florid demyelination could occur with ocrelizumab. It is our hope that this report will alert readers to this possibility.

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**Appendix (continued)**

| Name                          | Location                                                                 | Role                      | Contribution                                                                 |
|-------------------------------|--------------------------------------------------------------------------|---------------------------|-------------------------------------------------------------------------------|
| Vanessa F. Moreira Ferreira, MD | Brigham and Women’s Hospital, Partners MS Center, Harvard Medical School; Boston, MA, USA | Author                    | Designed and conceptualized the study; analyzed the data; and drafted the manuscript for intellectual content |
| David Meredith, MD            | Brigham and Women’s Hospital, Partners MS Center, Harvard Medical School; Boston, MA, USA | Author                    | Major role in the acquisition of histopathologic data                          |
| James Stankiewicz, MD         | Brigham and Women’s Hospital, Partners MS Center, Harvard Medical School; Boston, MA, USA | Author                    | Designed and conceptualized the study; analyzed the data; and interpreted the data and revised the manuscript for intellectual content |
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