Seleção de fungos entomopatogênicos nativos contra Cosmopolites sordidus (Germar) em laboratório

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ABSTRACT
The objective of this study was to select native isolates of entomopathogenic fungi against adult Cosmopolites sordidus in laboratory conditions. Four isolates of Beauveria bassiana, two isolates of Metarhizium anisopliae, and two isolates of Paecilomyces lilacinus were tested against C. sordidus. The entomopathogenic fungi that resulted in mortality rates over 50% were M. anisopliae (MA-CsCha and MA-Carabid) and B. bassiana (BB-CsCha and BB-CsLp). The most virulent isolates were MA-CsCha, BB-CsCha, and MA-Carabid with median lethal times (LT50) of 4.82, 5.4, and 5.79 days, respectively. In conclusion, the MA-CsCha, BB-CsCha, MA-Carabid, and BB-CsLp isolates are viable candidates to be tested in banana fields.

KEYWORDS: banana root borer, Beauveria bassiana, Metarhizium anisopliae, biocontrol.

RESUMO
Este estudo teve como objetivo selecionar isolados de fungos entomopatogênicos contra adultos de Cosmopolites sordidus em condições de laboratório. Quatro isolados de Beauveria bassiana, dois isolados de Metarhizium anisopliae e dois isolados de Paecilomyces lilacinus foram testados contra C. sordidus. Os fungos entomopatogênicos que resultaram em mais de 50% da mortalidade de adultos de C. sordidus foram M. anisopliae (MA-CsCha e MA-Carabid) e B. bassiana (BB-CsCha e BB-CsLp). Os isolados mais virulentos foram MA-CsCha, BB-CsCha e MA-Carabid com um tempo letal médio (TL50) de 4.82, 5.4 e 5.79 dias, respectivamente. Em conclusão, os isolados MA-CsCha, BB-CsCha, MA-Carabid e BB-CsLp são candidatos para serem testados em plantações de banana.

PALAVRAS-CHAVE: Moleque-da-bananeira, Beauveria bassiana, Metarhizium anisopliae, biocontrole.

The banana crop is important because it contributes to food security with an annual production of about 127 million tons worldwide (NYINE & PILLAY 2007, FAOSTAT 2020). However, the commercial crop faces problems caused by pests, and the Banana Root Borer (BRB), Cosmopolites sordidus (Germar), which infests the cortical tissues, can generate losses of up to 100% of the harvest (AKELLO et al. 2008, OLIVEIRA et al. 2017).

In this context, entomopathogenic fungi are an integrated pest management tool and an important alternative to insecticides against the BRB (OLSON 2015). There is a wide distribution of entomopathogenic fungi, which means they have a high genetic variability and distinct levels of virulence and local adaptability (NIASSY et al. 2013). Therefore, suitable isolates should be selected to control BRB populations according to the abovementioned aspects, since these studies are scarce under Peruvian conditions.

The entomopathogens Beauveria bassiana (Bals.) Vuill. (now Cordyceps bassiana) and Metarhizium anisopliae (Metsch.) Sorokin have been shown to be the most promising biocontrol agents against BRB due to their cosmopolitan distribution and high lethality caused by their production of mycotoxins and growth which blocks the insect’s digestive tract (TINZAARA et al. 2015, SALUSTINO et al. 2019). These species are
reported to cause infections for more than 300 species of pest insects (MOREIRA et al. 2017), which implies that this high genetic variability should be studied in order to select efficient entomopathogenic fungi adapted to the conditions where it is necessary to control BRB. Furthermore, the discovery of new native isolates with high virulence and infectivity can add value to biological control methods by reducing production costs and the use of conventional insecticides in banana fields (MEMBANG et al. 2020).

The objective of this study was to select native entomopathogenic fungi as potential biocontrol agents of *C. sordidus* populations in a humid tropic climate. The hypothesis was that at least one entomopathogenic fungus in the study would reach a 50% mortality rate within seven days after inoculation. Therefore, the results of this study offer new opportunities to discover potential entomopathogenic fungi that can be used in field using local and exotic isolates.

The study was conducted in the Laboratory of Entomopathogens at the Universidad Nacional Agraria de la Selva (UNAS), Tingo María, Peru (9°18'48” S, 75°59'45” W, 668.6 m above sea level), which has a humid tropical climate (Af) based on the Köppen-Geiger classification (PEEL et al. 2007). The eight entomopathogenic fungi tested in this study were obtained from the Servicio Nacional de Sanidad Agraria (SENASA) and UNAS collections. In the laboratory, four isolates of *Beauveria bassiana*, two isolates of *Metarhizium anisopliae*, and two isolates of *Paecilomyces lilacinus* (Tom) Samson were tested against *C. sordidus*. Information about the collections is presented in Table 1.

The entomopathogenic fungi were cultured in Petri dishes (8.5 cm diameter) containing Potato Dextrose Agar (PDA) at 24±5 °C for ten days. The Petri dishes and PDA were previously autoclaved at 121 °C, 15 lbs for 15 minutes.

After ten days, entomopathogenic fungi were sown in an artificial substrate. The artificial substrate had been prepared as follows: 200 g rice and 40 mL water placed in a polypropylene bag (8 x 12 cm); the bags were then folded, stapled closed, and autoclaved at 121 °C, 15 lbs for 15 minutes. Entomopathogenic fungi were sown in an artificial substrate in a laminar flow cabinet.

The bags with entomopathogenic fungi were stored on metal shelves in a room at 24 ± 5 °C and incubated for ten days. The bags were shaken daily to homogenize the internal content.

| Isolate codes | Scientific name | Origin |
|---------------|----------------|--------|
| BB-Hh | *Beauveria bassiana* | *Hypothememus hampei* collected in Sania Province, Peru (SENASA, commercial code: CCB-LE265) |
| BB-CsCha | *Beauveria bassiana* | *Cosmopolites sordidus* collected in Chanchamayo Province, Peru (SENASA, commercial code: CCB-LE2118) |
| BB-CsLp | *Beauveria bassiana* | *Cosmopolites sordidus* collected in Leoncio Prado Province, Peru (UNAS) |
| BB-CsVr | *Beauveria bassiana* | *Cosmopolites sordidus* collected in Villa Rica District, Peru (UNAS) |
| PL-MiIt | *Paecilomyces lilacinus* | *Meloidogyne incognita* collected in Italy (SENASA, commercial code: CCB-LE701) |
| PL-CsChi | *Paecilomyces lilacinus* | *Cosmopolites sordidus* collected in Chiclayo Province (UNAS) |
| MA-Carabid | *Metarhizium anisopliae* | Unidentified Carabid specimen in Peru (SENASA, commercial code: CCB-LE302) |
| MA-CsCha | *Metarhizium anisopliae* | *Cosmopolites sordidus* collected in Chanchamayo Province, Peru (SENASA, commercial code: CCB-LE319) |
| Control | - | Sterile water |

For the conidial suspension, a ten-day-old fungal colony was diluted in sterile distilled water and mixed with Tween 80® (0.05%). Then, the mixture was filtered through sterilized gauze to separate the conidia from the mycelium and other impurities. The quantity of conidia contained in the mixture were registered using a Neubauer chamber and then adjusted to a concentration of 10^8 conidia mL^-1. This concentration was used for the findings obtained by MEMBANG et al. (2020).

Adult *Cosmopolites sordidus* were collected in a banana field at Los Milagros Village, Tingo Maria, Peru. Traps formed by 10 cm banana stalks were used to attract *C. sordidus* in the field. One week later, adult *C. sordidus* were collected from the traps and placed in plastic boxes to be transported. In the laboratory, the adult *C. sordidus* were sprayed with and immersed in sodium hypochlorite (0.5%) for disinfection and then rinsed with sterile water three times. Only the more active ones of these insects were...
A completely randomized design with nine treatments was adopted to determine the mortality rates and lethal times of entomopathogenic fungi isolates (Table 1) against *C. sordidus* in the laboratory.

For each isolate, an experiment was conducted with three replicates. One replicate consisted of a group of 15 healthy adult *C. sordidus* that were individually sprayed with the solution containing the entomopathogens until they were dripping. A total of 45 adult *C. sordidus* were used per isolate. The sprayed insects were then conditioned in Petri dishes (10 cm diameter) containing banana stalks, which had been disinfected with 0.5% sodium hypochlorite solution for five min, in order to feed.

The number of active *C. sordidus* were evaluated for 21 days after infection. The mortality rate was calculated by the percentage of dead insects from total insects. Dead insects were individually maintained in Petri dishes lined with moistened filter paper to allow fungal sporulation.

The mortality rate percentage (x) under laboratory conditions was transformed by arcsine \( \sqrt{\frac{x}{100}} \). To obtain the median lethal times (LT50) of the isolates against *C. sordidus*, a Probit analysis was conducted. Data were analyzed using the R software for Windows (R CORE TEAM 2016). Cumulative mortality rates were corrected using Abbott's formula. Normality and equality of variance were tested by the Kolmogorov-Smirnov and the Levene’s tests, respectively (p ≤ 0.05). The mortality rate percentage was analyzed as a response variable in one-way (isolate) ANOVA. Whenever a significant F-test (p ≤ 0.05) was detected, the mean values for mortality were compared with Tukey’s test (HSD) at (p ≤ 0.05). The values of LT50 and LT90 of the isolates were estimated at 95% confidence levels (CL) using the Probit analysis with the “ecotoxicology” Package (GAMA 2015).

In total, 878 adult *C. sordidus* were collected for the study. The experiment revealed the ability of entomopathogenic fungi isolates to control *C. sordidus* adults under laboratory conditions. The MA-CsCha isolate resulted in a 57.78% mortality rate for adult *C. sordidus* five days after inoculation in the laboratory (Table 2). However, the MA-CsCha, BB-CsCha, MA-Carabid, and BB-CsLp isolates resulted in mortality rates of over 50% for adult *C. sordidus* seven days after inoculation.

According to the Probit regression, the MA-CsCha isolate resulted in a 50% mortality rate for *C. sordidus* 4.82 days after inoculation (Figure 1A). However, the BB-CsCha isolate resulted in a 90% mortality rate for *C. sordidus* in 8.04 days (Figure 1C). The differences between isolates are explained by the slope indicating that the rate of mortality increases 7.44 times per day for adult *C. sordidus* inoculated with BB-CsCha.

### Table 2. Mortality rates (%) for adult *Cosmopolites sordidus* infected with entomopathogenic fungi in the laboratory.

| Treatments      | Days after infection |
|-----------------|---------------------|
|                 | 5       | 6       | 7       |
| MA-CsCha        | 57.78a  | 71.11a  | 82.22a  |
| BB-CsCha        | 24.44b  | 77.78a  | 82.22ab |
| MA-Carabid      | 8.89b   | 73.33a  | 84.44a  |
| BB-Hh           | 13.33b  | 28.89b  | 33.33cd |
| BB-CsLp         | 13.33b  | 24.44bc | 53.33bc |
| PL-Milt         | 4.44b   | 4.44c   | 4.44e   |
| PL-CsChi        | 4.44b   | 6.67bc  | 11.11de |
| BB-CsVr         | 2.22b   | 6.67bc  | 11.11de |

Mean values followed by different letters in the same column are significantly different based on Tukey’s test (p ≤ 0.05).

*Metarhizium anisopliae* and *B. bassiana* infect a wide range of hosts in numerous insect orders, including Orthoptera, Lepidoptera, Dermaptera, Diptera, and Coleoptera (MAINA et al. 2018). However, in order to select isolates and use them for biological control, it is fundamental to understand the mortality rate and lethal time (kill at least 50% individuals) of an entomopathogenic fungus on a specific target pest (BAYISSA et al. 2017, MWEKE et al. 2018, MAHOT et al. 2019). This study reports on two isolates of *M. anisopliae* (MA-CsCha, MA-Carabid) and two isolates of *B. bassiana* (BB-CsCha, BB-CsLp), which can cause mortality rates of over 50% for adult *C. sordidus* under laboratory conditions. In Brazil, FANCELLI et al. (2013) reported similar results, wherein native isolates of *B. bassiana* promoted mortality rates (14 - 96%) for adult *C. sordidus* in laboratory experiments. The most virulent isolates achieved a LT50 of 4.82, 5.40, and 5.79 days (Figure 1), which were faster than the most virulent isolates reported by FANCELLI et al. (2013),
GONZÁLEZ et al. (2018), and LOPES et al. (2011).

On the other hand, some studies have reported that exotic isolates can possibly be more effective than native isolates (LOPES et al. 2013). Although it is not a rule, this study used exotic isolates from Sania (BB-Hb), Villa Rica (BB-CsVr), Chiclayo Province (PL-CsChi), and Italy (PL-MiIt), which did not present relevant results.

The findings also indicate variations between and within entomopathogenic fungi isolates, as reported by CHENG et al. (2016). Therefore, an individual belonging to the same fungal species is not a guarantee for similar virulence, because isolates can use different types and quantities of enzymes and metabolites to infect the insect host (ABDELAZIZ et al. 2018).

Future studies should use combinations of different species of biological control agents so as to increase the effects against pests, reduce insecticide use, and minimize environmental pollution risks and pest resistance (REDDY et al. 2014, CARVALHO 2017).

The results of this study indicate suitable entomopathogenic candidates to be tested in banana crops against *C. sordidus*. In conclusion, the native isolates of *Metarhizium anisopliae* (MA-CsCha and MA-Carabid) and *Beauveria bassiana* (BB-CsCha and BB-CsLp) were selected as potential biocontrol agents against *C. sordidus*.

Figure 1. Mortality rates for adult *Cosmopolites sordidus* caused by infection from isolates MA-CsCh (A), BB-CsCh (B), and MA-Carabid (C) over time, according to the Probit regression.

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