Influence of body mass index and weight lifting on bicep brachii muscle and distal bicep tendon stiffness evaluated using ultrasound elastography

Mahdi Al-Qahtani1, Omar Altuwaijri1, Meteb Altaf2, Majed Al-Enezi1, Mahmoud Abulmeaty3 and Ravish Javed1*

Abstract

Background: This study aimed to investigate the relationship between stiffness of the bicep brachii muscle (BBM) and distal bicep tendon (DBT) and effects of weight lifting (pre- to post-workout changes) among groups with different body mass indexes (BMI).

Methods: Participants were divided into four groups according to BMI: A, underweight (< 18.5 kg/m²); B, normal (18.5–24.9 kg/m²); C, overweight (25.0–29.9 kg/m²); and D, obese (> 30.0 kg/m²). All participants were males who were untrained and had sedentary lifestyle without involvement in sports activities for the past 12 months. Ultrasonographic measurements to determine muscle and tendon stiffness was performed on the dominant side (i.e., right side) of the upper extremities in all participants.

Results: Twenty-one healthy and untrained males volunteered to participate in this study; 14 were nonsmokers and 7 were smokers. The mean age and BMI were 22.5 ± 1.5 years and 23.8 ± 6.3 kg/m², respectively. Groups A, B, C, and D had four, ten, four, and three participants, respectively. The BBM thickness did not increase with increase in BMI and was not significantly different (P > .05) between groups. The BBM stiffness was significantly different (all P < .05) from pre- to post-workout values in all groups, whereas DBT stiffness did not follow the same trend.

Conclusions: Our study revealed that the BBM thickness is independent of BMI. After weight lifting, BBM stiffness in groups A and B increased for BBM compared to those in groups C and D. A similar trend was also recorded for DBT. Weight lifting in concentric and eccentric motions affects the stiffness of the BBM and DBT, thus weight lifting plays a role in adjusting the stiffness of the BBM and DBT.

Trial registration The study was approved by ethics committee of the College of Applied Medical Sciences (CAMS 080-3839; March 14, 2018).

Keywords: Ultrasound, Elastography, Strain ratio, Bicep brachii muscle, Distal bicep tendon, Body mass index

Background

In the upper limbs of the human body, the biceps brachii muscle (BBM) and distal bicep tendon (DBT) play a significant role in simple elbow joint movements, such as lifting and picking up objects and maintaining typing, writing, throwing, and resting positions. The BBM is the...
longest muscle in the arm, crosses two joints, and is connected to two tendons (long and short) at the proximal side and one tendon at the distal side. Elbow flexion and forearm supination are the most authoritative functions of the BBM and DBT [1], whereas supination of the forearm at the level of the elbow joint is performed solely by the DBT [2]. The BBM can be easily activated with optimum control and full recruitment of the motor unit when subjected to contraction. Pathological conditions, such as shoulder pain, usually involve both the long and short tendons of the BBM [3]. However, the long head of the biceps, due to its anatomical particularities and close functional relationship with the rotator cuff, is more frequently injured than the short head. While injuries of the BBM are rare, overuse and lifting excessive weight can cause muscle fibers to break, leading to severe pain. Injuries of the DBT are also uncommon; only 3% of all bicep tendon injuries (proximal and distal) are caused by partial or complete DBT rupture [4, 5]. Diagnosis of a partial tear of the DBT is complicated, and numerous approaches can be employed, such as ultrasonography (US) [6] and magnetic resonance imaging (MRI) [7]. Complete tears of the DBT are most commonly diagnosed by simply retracting the muscle belly and tendon [8]. Surgical treatment is usually recommended for DBT rupture as the results of conservative treatments are considered unreliable [9].

The BBM is used during upper-limb motion, in which the connected tendons directly and indirectly bear weight and force. Thus, any jerking and sporadic movement can cause damage to both the BBM and DBT. The intactness and strength of the elbow joint, thus the BBM and DBT, are critical to both athletes and non-athletes for optimal elbow function. Individuals who are actively involved in physical or sports activities and those with sedentary lifestyle are at a higher risk of elbow joint-related injuries, due to excessive use in physically active individuals and underuse in individuals with sedentary lifestyle. The stiffness of the BBM and DBT is correlated with body mass index (BMI) and physical activity. The combination of low BMI and sedentary lifestyle is associated with reduced stiffness of the BBM and DBT compared with high BMI and sedentary lifestyle. In contrast, individuals who are physically active with a lower BMI tend to exhibit similar or higher stiffness of the BBM and DBT than those with higher BMI who do not perform any physical activity. The stiffness of the BBM and DBT alone cannot depict the strength or weakness of the muscle independent of BMI, as this is also linked with physical activity; a healthy and lean body is achieved by regular exercise [10]. Muscle strength can be described as the force required in a single effort by a muscle to be used against some form of resistance [11]. Strength training or weight lifting can lead to significant variations in skeletal muscle [12, 13].

Presently, US elastography (USE) is the most appropriate and reasonable approach to measure the stiffness of the BBM and DBT. However, imaging modalities, such as magnetic resonance elastography (MRE) and USE, can be used to identify damage to the BBM and DBT caused by exercise [14, 15] or other physical activities. Both MRE and USE tend to deliver good results. Compared with US, MRE is time-consuming and expensive; however, the traditional US cannot diagnose tissue stiffness. Therefore, USE was developed as an inexpensive, quick, real-time, mobile, and safe alternative to MRI in evaluating tissue stiffness. The method works by identifying the strain induced by applying compression, either manually or automatically, at a particular area in the surrounding tissues.

Previous studies investigating muscle and tendon stiffness provide limited data. To date, there is no study corroborating the relationship of the stiffness of the BBM and DBT and the effects and benefits of weight lifting in individuals with sedentary lifestyle and different BMIs involving external reference material. The present study investigated the association between weight lifting and the stiffness of the BBM and DBT in healthy individuals with sedentary lifestyle. This study aimed to correlate the benefit of weight lifting (15 repetitions each with 2-kg and 5-kg dumbbells) in different groups and its effect on elastographic strain ratios.

**Methods**

**Study model**

This prospective study was conducted at the Biomedical Technology Department, College of Applied Medical Sciences, King Saud University. The study and its protocols were approved by the ethical review board (number CAMS 080-3839) of the College of Applied Medical Sciences, King Saud University. The guidelines of the Declaration of Helsinki were strictly followed in all experimental settings. Written informed consent was obtained from all participants of the study.

**Participants**

The current study recruited male students from the university. Volunteers with any history of corticosteroid treatment, hormone therapy, inflammatory/metabolic diseases, elbow surgery, upper-limb surgery/pain, and weight lifting or, specifically, patients with previous upper-limb injury were excluded from the study. The inclusion criteria were a sedentary lifestyle, lack of involvement in any type of sport or physical activity in the previous year, and Saudi descent. All participants were healthy and active, apart from participating...
in weight lifting or sports. All participants had right-side dominance.

A mandatory examination session regarding the inclusion criteria was conducted on all volunteers before obtaining actual readings on each volunteer. Thus, 21 participants were finally enrolled and divided into four groups according to BMI: A, underweight (<18.5 kg/cm²); B, normal weight (18.5–24.9 kg/cm²); C, overweight (25.0–29.9 kg/cm²); and D, obese (>30.0 kg/cm²). The mean age of the volunteers was 22.5 ± 1.5 years to analyze the muscle and tendon in close (or twenties) age range.

Familiarization to experimental protocols or orientation session for participants
A familiarization session was conducted in front of all potential participants to brief them about the experimental settings and protocols of US and USE. Additionally, one measurement and reading was performed by one of the authors following experimental protocols. Readings of the familiarization session were not included in the analysis. It was clarified that any concerns and questions from the participants related to experimental protocols would be addressed.

Body mass index and bioelectric impedance analysis
We calculated BMI by measuring the height (in cm) using an immobile stadiometer. Then, this value was entered into body composition analysis equipment (InBody 720, Body Composition Analyser; InBody Corporation Limited, Cerritos, CA, USA), which works on the principle of bioelectrical impedance [16–18]. Besides BMI, skeletal muscle mass (SMM) was also measured using the body composition analyzer.

Sonographic examinations
All US examinations were conducted using a high-frequency probe, linear transducer L14-5/38, Ultrasound SonixTouch Q+ (Analogic Corporation, 8 Centennial Drive, Peabody, MA, USA). Before examination (or measurements) in the current study, all volunteers were subjected to a thorough inspection of the upper limb, starting from the glenohumeral (shoulder) joint until approximately the midpoint of the radius bone for each participant. Moreover, all volunteers were asked to perform all fundamental and possible movements with their arm and forearm to identify if any abnormality in the proximal and distal tendons was present. Participants were seated in an adjustable-height chair next to a table. The dominant arm was extended 180° and placed on the table with a pillow underneath for comfort. Thickness was measured at the midpoint of the biceps brachii (Fig. 1), with the US probe notch facing the glenohumeral joint over the skin without exerting any pressure. Earlier, volunteers were asked to perform maximum elbow flexion, and the midpoint was marked at the center of the bicep brachii belly. All parameters of the US machine were kept constant, even the pillow used for the study was the same in all volunteers, as reproducibility is operator dependent and readings can change with slight variations. Thus, readings were obtained one after another without any break. To ascertain that a fixed pressure is applied on the skin and the same amount of strain is produced each time all measurements were obtained at four-bar indication, which can be seen on the bottom left of Fig. 4, it is the strain level indicator provided by the manufacturer of the US machine.

Strain elastography measurements
Strain elastography (SE) measurements for the BBM and DBT were obtained using the same instrument (selecting Elasto Mode) as previously used for US measurements. All SE readings of the BBM were recorded by placing the transducer over the external reference material (with dimensions 79 × 24.5 × 2 mm) on the belly of the BBM. The elasticity of the external reference material was 4.01 ± 1.13 MPa. Marks were made around the borders of the reference material with the use of a tape (Fig. 2). The reference material is commercially available. The use of external reference material is considered a more valid and reliable approach, especially when the number of participants is more than one, to overcome the limitation of subcutaneous fat tissue, which changes in every participant. The external reference material had a constant elasticity, in contrast to the BBM. The US transducer was placed in the longitudinal plane over the bicep brachii belly, with the notch of the US probe facing the shoulder of the examinee. All SE readings were measured at the
center of the probe placement. A similar protocol was followed for the measurement of the DBT by SE (Fig. 3). To minimize the chance of error and avoid under- or overestimation of stiffness for all volunteers, the same external reference material was used and strain values were measured at four-bar indication, which can be seen on the bottom left of Fig. 4, which is the strain level indicator provided by the manufacturer of the US machine. However, there can still be a chance of little or no variation when measuring the strain ratios due to the amount of subcutaneous tissue that can differ for each individual.

**Bicep brachii muscle measurement arrangement**

We conducted SE readings in three phases of the BBM. In the initial phase, three readings were obtained at the BBM for each participant before lifting any weight (Fig. 4). In the second phase, after rest, participants were asked to lift a 2-kg dumbbell 15 times in a relaxed seated position. While seated, each participant was instructed to start from an elbow extension angle of approximately 0° to an elbow flexion of approximately 90°. Fifteen repetitions were performed in this manner, and three SE readings of the BBM were obtained in the same position. In the final phase, after a brief rest, a 5-kg dumbbell was lifted 15 times as previously described, and three SE readings were obtained for the BBM. Dumbbell weights of 2 and 5 kg were used in all groups as this weight range is usually encountered in daily life, regardless of whether an individual is involved in weight lifting or has a sedentary lifestyle, and thus can easily be lifted by all participants. Fixed dumbbell weights were used in all groups to ensure the same amount of weight-lifting impact on the BBM. The mean of the three readings was calculated for each phase to increase intraobserver reliability and decrease deviation and risks of error.

**Distal bicep tendon measurement arrangement**

To measure the DBT by SE, each participant underwent three phases (as for the BBM). The DBT readings were measured by asking the participant to move their forearm in maximum supination for better visualization. This protocol was followed in all volunteers for DBT measurements. The SE readings for all three phases were calculated for each participant just after taking readings of the BBM, with similar protocol and settings (Fig. 5).

**Inclusion of external reference material**

An external reference was used to maintain consistency and set a common reference for all SE values (for both the BBM and DBT). A silicon rubber was purchased from a commercial source. The dimensions and elasticity of the reference material were $80 \times 24 \times 2$ and $4.01 \pm 1.13$ MPa, respectively. Two regions of interest were selected in calculating the value of SE in the case of BBM (BBM/adjacent external reference material; Fig. 4). Similarly, Fig. 5 shows the two regions of interest selected for calculating...
the SE value for the DBT (DBT/adjacent external reference material).

Statistical analysis

We used SPSS Statistics version 25 software for Windows (IBM Corporation, Armonk, NY, USA) in the statistical analysis. One-way analysis of variance was performed using Tukey’s honestly significant difference test to evaluate the level of significance among the three phases for each group. Differences between the groups were found using a t-test, while post hoc Tukey’s test was performed to determine the level of significance between them. All values were presented as mean ± standard deviation with a P value of 0.05 indicating significant difference.

Results

We enrolled 21 participants aged 22.5 ± 1.5 years (range 21–28 years). The mean weight, height, and BMI of all 21 participants were 70.1 ± 18 kg, 171.8 ± 6.0 cm, and 23.8 ± 6.3 kg/cm², respectively. The age ranges for groups A (n = 4), B (n = 10), C (n = 4), and D (n = 3) were 22–23, 21–28, 21–24, and 22–22 years, respectively; seven participants were smokers and 14 were nonsmokers. The mean BMIs for each group was 17.4 ± 0.7, 21.6 ± 2.1, 27.2 ± 0.8, and 35.4 ± 6.3 kg/cm² for groups A, B, C, and D, respectively. The SE values for the BBM and DBT before and after weight lifting showed that strain values significantly differed according to BMI. A statistically significant difference was observed for most factors between the four groups.

The mean BMI significantly differed between all groups, except for groups A and B (Fig. 6).

The mean SMM measurements significantly differed between groups (Fig. 7).

Figure 8 shows the mean BBM thickness for each group before weight lifting. Although the thickness increased from groups A to B and B to C, the mean thickness of group D was found to be decreased compared with that of group C. None of the differences were statistically significant.
The mean strain ratio of the BBM for group A showed an increasing trend after lifting of 2- and 5-kg dumbbells (Fig. 9). Additionally, a significant difference was noted between each phase.

The mean strain ratio of the BBM for group B also showed an increasing trend after lifting the 2- and 5-kg dumbbells (Fig. 10). Furthermore, a significant difference was observed before and after lifting the 2-kg weight and before and after lifting the 5-kg weight. No significant difference was observed between lifting the 2-kg and 5-kg weight.

In group C, the strain ratio of the BBM decreased when subjected to weight lifting. The strain ratios in each phase were significantly different before and after lifting 2 kg, before and after lifting 5 kg, and between lifting 2 kg and 5 kg weight (Fig. 11).

The mean strain ratio for the BBM in group D dramatically decreased when subjected to weight lifting (Fig. 12). The strain ratios were significantly different before and after lifting 2 kg, and after lifting 5 kg. No significant difference was observed between lifting 2 kg and 5 kg weight.

Figure 13 shows the mean strain ratio of the DBT for group A. The values showed variation when subjected to lifting 2 and 5 kg dumbbells. A significant difference was observed from before lifting to after lifting 5 kg and after lifting 2 kg to lifting 5 kg. However, no significant difference was observed before lifting and after lifting 2 kg dumbbells.

In group B, the mean strain ratios for the DBT increased when subjected to weight lifting, although these differences were not significant (Fig. 14). A significant difference was noted between the strain ratios before lifting and after lifting 5 kg.

As Fig. 15 shows, the mean strain ratios for the DBT in group C showed no significant difference in any phase, although the value decreased across the three phases.

The mean strain ratios of the DBT for group D showed no significant difference when subjected to weight lifting.
but exhibited irregular changes when subjected to lifting 2 or 5 kg dumbbells compared with that before lifting. A significant difference was noted among groups A, B, C, and D for mean values of strain ratios in BBM before weight lifting but not after lifting 2 and 5 kg. In the case of DBT, the variation in mean values of strain ratios was only statistically significant in groups B and D after lifting 2 kg (Table 1).

Discussion

The aim of this study was to scrutinize the association of the effects of weight lifting on the BBM and DBT concerning BMI. The current study results revealed that SE measurements of BBM showed significant variation when exposed to weight lifting, while SE measurements of DBT showed significant variation in only a few scenarios. Variations in the mean SE values seemed to be correlated with the mean BMI of various groups, all of them having a sedentary lifestyle. To date, there are no data on the effect of weight lifting (in concentric and eccentric motions, using two specific weights) on the SE (evaluated using an external reference) of the BBM and DBT.

In groups A and B, there was a significant increase in BBM mean strain ratio (or stiffness) before and after lifting 2- and 5-kg dumbbells. Newham et al. [19] reported that, in the first 3 days of exercises, eccentric movements can cause instant ultrastructural damage to the sarcomeres of the muscle fibers, affecting myofibrillar structures. The intramuscular water content level increases following repetitive muscle contractions [20, 21]. Thus, the increase in BBM stiffness is due to a combination of factors, including change in the osmotic grade of the vascular and extravascular spaces, production of metabolites in the affected muscle, and induced edema coupled to increased blood flow (intramuscular) due to increased permeability and capillary pressure [22–24]. Weight lifting causes microfiber injury in the BBM that

![Graph](image-url)

**Fig. 14** Mean strain ratios of the distal bicep tendon for group B. *P<0.05

![Graph](image-url)

**Fig. 15** Mean strain ratios of the distal bicep tendon for group C

![Graph](image-url)

**Fig. 16** Mean strain ratios of the distal bicep tendon for group D

| Table 1 | Mean strain ratios of bicep brachii muscle and distal bicep tendon in all groups |
|---------|-----------------------------------------------------------------------------|
| Groups  | A (Underweight) | B (Normal) | C (Overweight) | D (Obese) |
|---------|-----------------|------------|---------------|-----------|
| Before lifting weight | 0.024<sup>‡</sup> | 0.054<sup>‡,#</sup> | 0.166<sup>‡,#</sup> | 0.293<sup>‡,#</sup> |
| After lifting 2 kg | 0.060 | 0.158 | 0.101 | 0.244 |
| After lifting 5 kg | 0.110 | 0.163 | 0.073 | 0.092 |
| Before lifting weight | 0.175 | 0.207 | 0.293 | 0.169 |
| After lifting 2 kg | 0.161 | 0.285<sup>‡</sup> | 0.153 | 0.115<sup>‡</sup> |
| After lifting 5 kg | 0.284 | 0.440 | 0.149 | 0.143 |

Key: Symbols *, †, ‡, §, ¶, Δ indicate the presence of significant difference (P<0.05) between groups
led to an increase in stiffness in groups A and B. This results in increased muscular pressure due to fluid accumulation. Microscopic muscle damage is a consequence of increased stress in the muscle fiber, which produces microtears as the muscle is stretched by intense concentric and eccentric muscle movements [25]. Few studies have confirmed that concentric and eccentric muscle contractions cause increased muscle stiffness [26, 27]. It has been observed that weight lifting is strenuous to unaccustomed exercises or movements that affect the stiffness and mechanical properties of muscles and the joint activity and the movements of an individual for 24–72 h. In a study by Agten et al. [28], sarcolemmal breakup and opening of mechanosensitive cell membrane channels triggered by eccentric exercise cause accumulation of intracellular Ca$^{2+}$ and Na$^{+}$ ions. This causes increased cellular damage, inflammation, and edema and transient reduction in functionality; however, these ion influxes are also responsible for stimulus-induced muscle hypertrophy [29]. To remove necrotic cells, inflammatory cells (primarily neutrophils) penetrate skeletal muscles in concentric and eccentric exercises of 45 min to 2 h in duration [30, 31]. In the present study, the increased stiffness observed in groups A and B might be due to the increase in blood flow and extracellular muscle edema following weight lifting. The increase in stiffness of BBM after weight lifting was approximately equal in group A from before lifting to after lifting 2 kg and from after lifting 2 kg to after lifting 5 kg. The significant difference in BBM stiffness across all three phases could be explained by the absence of sufficient SMM to counter the concentric and eccentric exercise movement. Sufficient skeletal muscle comprises 40% of human body weight [32] and was found to be not present in group A volunteers by the body composition analyzer. In group B, a significant difference was observed in BBM stiffness from before lifting to after lifting 2 kg, the difference from after lifting 2 kg to after lifting 5 kg was not significant in group B, and group B has higher SMM than group A, as it was measured using the body composition analyzer. The results of a previous study by Tomlinson et al. relating to increased adiposity and muscle stiffness among overweight individuals compared with normal or underweight individuals corroborate our findings [33]. Groups C and D have higher BMI; thus, the presence of higher fat infiltration [34] and higher SMM might be the cause of decrease in stiffness in these two groups. Moreover, in the earlier study, weight lifting causes decreased SMM [35]. The hypothesis is that the greater the BMI, the lower the BBM stiffness. This may be attributed to obesity-related changes in skeletal muscles. In group C, the presence of higher SMM and a decrease in stiffness values with activity were recorded. In group D, no significant difference was observed in BBM stiffness before lifting and after lifting 2 kg because of the presence of higher SMM, which was also higher than group C.

The current study reveals that DBT stiffness varies depending on the BMI. A previous study by Bohm et al. [36] reported that tendons are responsive to loading procedures, which corroborates our findings by showing different stiffness values for DBT readings. Another study by Kubo et al. [37] reported stiffness of the tendon to be proportional to the time spent exercising; thus, after lifting 2 and 5 kg weights, there was a difference in DBT stiffness among all groups. A study by Siu et al. [38] revealed that stiffness of the tendon is increased by exercise, which validates our findings that groups A and B exhibited significant differences in DBT stiffness following exercise. However, this result was not observed in groups C and D, indicating that tendon composition is different than in groups A and B due to BMI; a previous tendon study validates our results [39]. In the present study, stiffness decreased with the increased weight used for concentric and eccentric weight lifting. Groups C and D represent overweight and obese participants, respectively; thus, their tendon has more stiffness and less likely to be affected by 2- and 5-kg dumbbell weight lifting. A possible reason for the decreased stiffness in overweight and obese individuals is the changes in the structure of the tendon that impairs the movement of interstitial fluid in response to weight lifting [40]. The lack of significant differences in BBM and DBT stiffness in groups C and D is caused by the difference in the SMM. Despite the increase in the SMM, obesity is associated with an increased amount of type II muscle fiber, reduction in satellite cell activation, and insulin sensitivity, which impairs muscular regeneration. Moreover, obesity-associated low-grade chronic inflammation could impair skeletal muscle protein synthesis [41].

Our study involved Saudi men in their twenties with sedentary lifestyle. As muscles and tendons [42] develop and grow differently in females than males, future studies on muscles and tendon stiffness in females would be informative. To further strengthen the relationship between BBM and DBT stiffness with BMI, new studies involving children, elderly participants, athletes, and unhealthy individuals are required, as they were excluded in the present study. Furthermore, the limited reproducibility is a weakness of USE as it is operator dependent. Moreover, an increased or equal sample size for each group is highly recommended for future studies, as the present study has a limited number of volunteers in one group. Another study can be conducted to relate the stiffness with the subcutaneous soft tissue to overcome the limitations of the BMI measurements. However, the same amount of subcutaneous soft tissue present in two
different individuals can lead to similar or different stiffness readings.

Conclusions
Our study provides insight into the stiffness of muscles and tendons following weight lifting and is the first to analyze this parameter of the BBM and DBT after weight lifting concerning BMI using SE. Generally, weight lifting in concentric and eccentric motions affects the stiffness of the BBM and DBT, and the direction of this effect depends on the BMI, with stiffness increasing in individuals with lower BMI and decreasing in individuals with higher BMI. In this study, we attempted to establish an association between the BBM and muscle stiffness and track the changes in the SMM by bioelectric impedance analyzer during weight lifting. Weight lifting plays a role in adjusting (increasing or decreasing) the BBM and DBT stiffness.

Abbreviations
BBM: Bicep brachii muscle; DBT: Distal bicep tendon; BMI: Body mass index; US: Ultrasound; MRI: Magnetic resonance imaging; MRE: Magnetic resonance elastography; SMM: Skeletal muscle mass; USE: Ultrasound elastography; SE: Strain elastography.

Acknowledgements
The authors extend their appreciation to the Deanship of Scientific Research at King Saud University for funding this work through the research group (No. RG-1440-117).

Authors’ contributions
MQ and RJ conceived the research. RJ, OA, and MAe analyzed the clinical and imaging data. RJ designed the study. RJ and MQ drafted the manuscript. RJ, Mal, and Mab conducted the statistical analysis. RJ reviewed the final manuscript. All authors read and approved the final manuscript.

Funding
The authors extend their appreciation to the Deanship of Scientific Research at King Saud University for funding this work through the research group (no. RG-1440-117). The funder had no involvement or influence whatsoever in the study design at any stage, collection of the data or its analysis and interpretation, writing and preparation of the manuscript, or its submission for publication.

Availability of data and materials
The datasets generated and/or analyzed in the current study are not publicly available due to patient privacy protection but are available from the corresponding author on reasonable request.

Ethics approval and consent to participate
Ethical standards were strictly followed in all experimental procedures as approved by the institutional ethics board of the College of Applied Medical Sciences Research Ethics Committee—King Saud University bearing ethics approval number CAMS 0800-3839, and the study complied with the principles of the Declaration of Helsinki. All participants provided written informed consent before the experimental procedures.

Consent for publication
Not applicable.

Competing interests
The authors have no competing interest to declare.

References
1. Simons DG, Travell JG, Simons LS. Myofascial pain and dysfunction: the trigger point manual. Upper half of body, vol. 1. London: Lippincott Williams & Wilkins; 1999.
2. Pierce JL, Nacey NC. Elbow ultrasound. Curr Rad Rep. 2016;4(10):51.
3. Frost A, Zafar MS, Maffulli N. Tenotomy versus tenodesis in the management of pathologic lesions of the tendon of the long head of the biceps brachii. J Am Soc Surg Sports Med. 2009;37(4):828–33.
4. Mazzocca AD, Spang JT, Anciero RA. Distal biceps rupture. Orthop Clin North Am. 2008;39(2):237–49.
5. Shields E, Olsen JR, Williams RB, Rouse L, Maloney M, Volarini I. Distal biceps brachi tendon repair: a single-incision technique using a cortical button with interference screw versus a double-incision technique using suture fixation through bone tunnels. J Am Soc Surg Sports Med. 2015;43(5):1072–6.
6. Sconfinenza LM, Albano D, Allen G, Bazzocchi A, Bignotti B, Chianva V, et al. Clinical indications for musculoskeletal ultrasound updated in 2017 by European Society of Musculoskeletal Radiology (ESSR) consensus. Eur Radiol. 2018;28(12):5338–51.
7. Huec J, Moinard M, Liqou J, Zopoli B, Chauveaux D, Liberellier A. Distal rupture of the tendon of biceps brachii. J Bone Joint Surg. 1996;78:767–70.
8. Belli P, Costantini M, Mirk P, Leone A, Pastore G, Marano P. Sonographic diagnosis of distal biceps tendon rupture: a prospective study of 25 cases. J Ultrasound Med. 2001;20(6):587–95.
9. Al-Qahtani M, Aljaser E, Alajmi A, Altwijri O, Mirza H. Effect of training on stiffness of distal biceps tendon: a pilot study. Kuwait Med J. 2018;50(2):220–5.
10. Buys R, Avila A, Cornelissen VA. Exercise training improves physical fitness in patients with pulmonary arterial hypertension: a systematic review and meta-analysis of controlled trials. BMC Pulm Med. 2015;15(1):40.
11. Enoka RM. Muscle strength and its development. New perspectives. Sports Med. 1988;6(3):146–68.
12. Jones DA, Rutherford OM, Parker DF. Physiological changes in skeletal muscle as a result of strength training. Q J Exp Physiol Transl Integr. 1989;74(3):233–56.
13. Evans JW. Periodized resistance training for enhancing skeletal muscle hypertrophy and strength: a mini-review. Front Physiol. 2019;10:13.
14. Nitsu M, Michizaki A, Endo A, Takei H, Yanagisawa O. Muscle hardness measurement by using ultrasound elastography: a feasibility study. Acta Radiol. 2011;52(1):99–105.
15. Lacourpaile L, Nordez A, Hug F, Cournier A, Dibie C, Guillet H. Time-course effect of exercise-induced muscle damage on localized muscle mechanical properties assessed using elastography. Acta Physiol. 2014;211(1):135–46.
16. Macaulay TR, Uhler CD, Schoeder ET. Evaluation of body composition measurements obtained using whole-body plethysmography: 752 board # 13 May 30–2. Med Sci Sports Exer. 2018;50(5S):161.
17. Andreacci JL, Nocera V, Heinrich C, Fradkin AJ, Dixon CB. Validation of the InBody 770 for the assessment of percent body fat in young adults: 765 board # 26 May 30–2. Med Sci Sports Exer. 2018;50(5S):165.
18. Ranasinghe C, Gamage P, Katulanda P, Andraweera N, Thilakarathne S, Tharanga P. Relationship between body mass index (BMI) and body fat percentage, estimated by bioelectrical impedance, in a group of Sri Lankan adults: a cross sectional study. BMC Public Health. 2013;13(1):797.
19. Newham DJ, McPhail G, Mills KB, Edwards BH. Ultrastructural changes after concentric and eccentric contractions of human muscle. J Neurol Sci. 1983;61(1):109–22.
20. Yanagisawa O, Kudo H, Takahashi N, Yoshioka H. Magnetic resonance imaging evaluation of cooling on blood flow and oedema in skeletal muscles after exercise. Eur J Appl Physiol. 2004;91(5–6):737–40.
21. Yamauchi J, Hargens AR. Effects of dynamic and static handgrip exercises on hand and wrist volume. Eur J Appl Physiol. 2008;103(1):41–5.
22. Ploutz-Snyder LL, Convertino VA, Dudley GA. Resistance exercise-induced fluid shifts: change in active muscle size and plasma volume. Am J Physiol Regul Integr Comp Physiol. 1995;269(3 Pt 2):R536–43.
23. Fotedar LK, Slopis JM, Narayana PA, Fenstermacher MJ, Pivarnik J, Butler BJ. Proton magnetic resonance of exercise-induced water changes in gastrocnemius muscle. J Appl Physiol. 1990;69(5):1695–701.
24. Sjøgaard G, Saltin B. Extra- and intracellular water spaces in muscles of man at rest and with dynamic exercise. Am J Physiol Regul Integr Comp Physiol. 1982;243(3):R271–80.
25. Fridén J, Sjöström M, Ekblom B. Myofibrillar damage following intense eccentric exercise in man. Int J Sports Med. 1983;4(3):170–6.
26. Howell JN, Chleboun GS, Conatser RR. Muscle stiffness, strength loss, swelling and soreness following exercise-induced injury in humans. J Physiol. 1993;464(1):183–96.
27. Chleboun GS, Howell JN, Conatser RR, Giesey JJ. Relationship between muscle swelling and stiffness after eccentric exercise. Med Sci Sports Exerc. 1993;25(4):S29–35.
28. Achten CA, Buck FM, Dyer L, Fluck M, Pfirrmann CW, Roesskopf AB. Delayed-onset muscle soreness: temporal assessment with quantitative MRI and shear-wave ultrasound elastography. AJR Am J Roentgenol. 2017;208(2):402–12.
29. McKune AJ, Semple SJ, Peters-Futre E. Acute exercise-induced muscle injury. Biol Sport. 2012;29(1):3–10.
30. Mäntyranta DL, Reid WD, Lyster DM, McKenzie DC. Different effects of strenuous eccentric exercise on the accumulation of neutrophils in muscle in women and men. Eur J Appl Physiol. 2000;81(2):47–53.
31. Fielding RA, Manfredi TJ, Ding W, Fiatarone MA, Evans WJ, Cannon JG. Acute phase response in exercise. III. Neutrophil and IL-1 beta accumulation in skeletal muscle. Am J Physiol Regul Integr Comp Physiol. 1993;265(1 Pt 2):R166–72.
32. Jespersen J, Kjaer M, Schjerling P. The possible role of myostatin in skeletal muscle atrophy and cachexia. Scand J Med Sci Sports. 2006;16(2):74–82.
33. Tomlinson DJ, Erskine RM, Morse CI, Winwood K, Onambélé-Pearson G. The impact of obesity on skeletal muscle strength and structure through adolescence to old age. Biogerontology. 2016;17(3):467–83.
34. Kennedy PF, Barnhill E, Gray C, Brown C, Van Beek EJR, Roberts N, et al. Magnetic resonance elastography (MRE) shows significant reduction of thigh muscle stiffness in healthy older adults. GeroScience. 2020;42(1):311–21.
35. Chomentowski P, Dubé JJ, Amati F, Stefanovic-Racic M, Zhu S, Toledo FG, et al. Moderate exercise attenuates the loss of skeletal muscle mass that occurs with intentional caloric restriction-induced weight loss in older, overweight to obese adults. J Gerontol S Biomed Sci Med Sci. 2009;64(5):575–80.
36. Bohm S, Mersmann F, Arampatzis A. Human tendon adaptation in response to mechanical loading: a systematic review and meta-analysis of exercise intervention studies on healthy adults. Sports Med Open. 2015;1(1):7.
37. Kubo K, Ikebukuro T, Maki A, Yata H, Tsuonoda N. Time course of changes in the human Achilles tendon properties and metabolism during training and detraining in vivo. Eur J Appl Physiol. 2012;112(7):2679–91.
38. Siu WL, Chan CH, Lam CH, Lee CM, Ying M. Sonographic evaluation of the effect of long-term exercise on Achilles tendon stiffness using shear wave elastography. J Sci Med Sport. 2016;19(11):883–7.
39. Taş S, Yılmaz S, Onur MR, Soylu AR, Altuntaş O, Korkusuz F. Patellar tendon mechanical properties change with gender, body mass index and quadriceps femoris muscle strength. Acta Orthop Traumatol Turc. 2017;51(1):54–9.
40. Wearing SC, Hooper SL, Grigg NL, Nolan G, Smeathers JE. Overweight and obesity alters the cumulative transverse strain in the Achilles tendon immediately following exercise. J Bodyw Mov Ther. 2013;17(3):316–21.
41. Kim J, So WY. High body mass index is associated with the extent of muscle damage after eccentric exercise. Int J Environ Res Public Health. 2018;15(7):1378.
42. Miller BF, Hansen M, Olesen JL, Schwarz P, Babraj JA, Smith K, et al. Tendon collagen synthesis at rest and after exercise in women. J Appl Physiol. 2007;102(2):402–12.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.