Enhanced nitrogen removal and energy saving in a microalgal–bacterial consortium treating real municipal wastewater

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ABSTRACT

The optimization of total nitrogen removal from municipal wastewater was investigated in a laboratory-scale photo-sequencing batch reactor (PSBR) operated with a mixed microalgal-bacterial consortium spontaneously acclimatized to real wastewater. No external aeration was provided in the PSBR to reduce energy consumption: oxygen was only supplied by the microalgal photosynthesis. The enhancement of total nitrogen removal was achieved through: (1) feeding of wastewater in the dark phase to provide readily biodegradable COD when oxygen was not produced, promoting denitrification; (2) intermittent use of the mixer to favor simultaneous nitrification-denitrification inside the dense flocs and to achieve 41% energy saving with respect to continuous mixing. Efficient COD removal (86 ± 2%) was observed, obtaining average effluent concentrations of 37 mg/L and 22 mg/L of total COD and soluble COD, respectively. TKN removal was 97 ± 3%, with an average effluent concentration of 0.5 ± 0.7 mg NH₄⁺-N/L. Assimilation of nitrogen by heterotrophic bacteria accounted only for 20% of TKN removal, whilst the major part of TKN was nitrified. In particular, the nitrification rate was 1.9 mgN L⁻¹ h⁻¹ (specific rate 2.4 mgN gTSS⁻¹ h⁻¹), measured with dissolved oxygen near zero, when the oxygen demand was higher than the oxygen produced by photosynthesis. Total nitrogen of 6.3 ± 4.4 mgN/L was measured in the effluent after PSBR optimization.

Key words | microalgae, microalgal–bacterial consortia, nitrification–denitrification, nitrogen removal, photobioreactor, real wastewater

INTRODUCTION

The treatment of real wastewater with microalgal–bacterial consortia has gained increasing attention in the last few years because the synergistic effects of microalgae and heterotrophic bacteria can be exploited for carbon and nitrogen removal through nitrification–denitrification, ensuring a significant energy saving (Su et al. 2012; Zhang et al. 2012; He et al. 2013; Abinandan & Shanthakumar 2015; Wang et al. 2016b; Arcila & Buitrón 2017; Quijano et al. 2017).

However, when ammonium removal occurs with nitrifiers in activated sludge, a huge amount of electrical energy is required to supply air in aerobic bioreactors. This energy consumption, represents up to half of the energy expenditure of a wastewater treatment plant (WWTP) based on activated sludge. In particular, considering the annual energy consumption by the air supply in aerated tanks in the range of 4.7–28 kWh PE⁻¹ y⁻¹ (Foladori et al. 2013), a medium-size WWTP (20,000 population equivalent, PE) may consume 94–560 MWh/y, which corresponds to an annual cost of 11,000–64,000 €/y (average EU electricity price for industrial consumers of 0.114 €/kWh; Eurostat 2017).

These figures emphasize the need to explore new alternatives aimed at reducing energy consumption and associated costs of wastewater treatment in WWTPs. In this context, the development of heterogeneous
microalgal–bacterial consortia offers the possibility of moving towards quasi-zero-energy consumption; in particular, the oxygen produced by photosynthetic microorganisms may support the oxidation of organic matter and ammonium by heterotrophic and nitrifying bacteria (Van den Hende et al. 2014). Suspended-biomass reactors operating as photo-sequencing batch reactors (PSBR) represent a common configuration for the laboratory-scale implementation of microalgal–bacterial consortia (Arcila & Buitrón 2016; Wang et al. 2016b) because of the easy setup with small pumps and timers.

With regard to nitrogen removal in microalga–bacterial consortia, the main mechanisms are biomass assimilation, nitrification–denitrification and, when high pH values are reached, ammonia volatilization (Delgadillo-Mirquez et al. 2016; Wang et al. 2016b). González-Fernández et al. (2011) observed that microalgal assimilation of nitrogen is frequently overestimated. Su et al. (2011), treating municipal wastewater with a mixed population of microalgae and bacteria, observed that denitrification was considered to be a possible removal mechanism, while ammonia volatilization was excluded due to the pH being lower than 8.5.

In spite of the advantage of oxygen production by photosynthetic microorganisms, dissolved oxygen (DO) released in the bulk liquid may become excessive and may inhibit the transformation of nitrates into nitrogen gas through heterotrophic denitrification, which requires strictly anoxic conditions. Thus, it may be difficult to meet the legal requirements for total nitrogen in the effluent if the operational strategies in the microalgal system are not optimized. Wang et al. (2015) achieved 90% of nitrogen removal by alternating light and dark periods and without artificial aeration, but to promote denitrification during the dark period, an organic carbon source was added.

The present experimental study focuses on the optimization of total nitrogen removal in a laboratory-scale PSBR fed with real wastewater and operated with a mixed consortium of microalgae, cyanobacteria and bacteria. The phases in the PSBR were feeding + react + settling + draw, but no idle phase was included in the cycles. The length of the phases and the photoperiod (light/dark) were combined so as to affect the duration of the aerobic and anoxic periods and promote nitrification and denitrification, without the need for an additional external carbon source. This paper contributes to the optimization of a PSBR, in terms of enhancement of nitrogen removal, reduction of the time of treatment, and energy saving where possible, thus improving the global sustainability of the system.

**MATERIALS AND METHODS**

**Influent wastewater**

Influent municipal pre-settled wastewater was collected after the primary settler of the Trento Nord WWTP (100,000 PE) and fed in to a laboratory-scale photobioreactor. No filtration of the influent wastewater was performed before feeding in to the reactor. This was to allow the microorganisms naturally present in the influent wastewater to enter the reactor, thus significantly affecting the composition of the microalgal–bacterial consortium developed in the system. Table 1 shows the concentrations of the main parameters in the influent pre-settled wastewater: chemical oxygen demand (COD), soluble COD (sCOD), total Kjeldahl nitrogen (TKN), ammonium nitrogen (NH4+-N), nitrite nitrogen (NO2--N), nitrate nitrogen (NO3--N), TN, total phosphorus (TP), orthophosphate phosphorus (PO43--P) and total suspended solids (TSS). The average concentrations are in agreement with typical values expected in pre-settled wastewater (Tchobanoglous et al. 2003).

**Photo-sequencing batch reactor**

The PSBR setup consisted of a cylindrical bench-scale reactor made of Pyrex glass with diameter of 0.13 m, height of 0.29 m and a working volume of 2 L (Colaver, Italy) not sealed off from atmosphere (Figure 1(a)). The system was equipped with:

- peristaltic pumps (Kronos Seko, Italy) for pumping the influent and discharging the effluent; 0.7 L of influent wastewater was fed per cycle;
- cool-white lamp, 8 LEDs × 0.5 W, 0.18 m high and 0.065 m wide (Orion, Italy), arranged on one side of the PSBR.

| Parameter | Concentration (mg/L) |
|-----------|----------------------|
| COD       | 257 ± 91             |
| sCOD      | 120 ± 42             |
| TKN       | 55 ± 20              |
| NH4+-N    | 50 ± 14              |
| NO2--N    | 0.1 ± 0.1            |
| NO3--N    | 1.2 ± 0.9            |
| TN        | 54 ± 22              |
| TP        | 4.8 ± 1.5            |
| PO43--P   | 2.8 ± 0.9            |
| TSS       | 184 ± 122            |

Table 1 | Characterization of influent pre-settled wastewater (average ± standard deviation; number of samples = 30)
- magnetic mixer set at 200 rpm (AGE, Velp, Italy) used to maintain the biomass in suspension during the react phase, but avoiding excessive turbulence and reoxygenation from the air;
- timers (TR 612 top2, Theben, Germany) to control on/off of pumps, lamp and mixer.

A light intensity of 30 μmol m$^{-2}$ s$^{-1}$ (expressed as photosynthetically active radiation, PAR) was measured inside the reactor near the top of the liquid surface, at the most exposed point. Light originated from solar light penetration into the laboratory and the lamp, but the reactor was never exposed to direct sunlight. This light intensity is in the same order of magnitude as other studies in the literature (for example 63 μmol m$^{-2}$ s$^{-1}$ in Karya et al. 2015; 90 μmol m$^{-2}$ s$^{-1}$, in Mennaa et al. 2015) but lower than the PAR in outdoor plants which surpasses 200 μmol m$^{-2}$ s$^{-1}$ (light irradiances that saturate photosynthesis). It is worth noting that no external aeration was provided in the PSBR (in order to reduce energy consumption and move towards a zero-energy system); the oxygen available in the reactor was supplied only by microalgal photosynthesis.

The temperature of the mixed liquor during the experiment was 22.8 °C on average.

An acclimatized microalgal–bacterial consortium that had spontaneously developed over more than a year in the PSBR was employed in this study, which was carried out from January to July 2017. Microscopic observations of the biomass showed the presence of *Chlorella*, diatoms, many filamentous cyanobacteria and heterotrophic bacteria embedded in flocs.

**Operational strategy in the PSBR**

The typical PSBR cycle was based on the following phases:
(1) feeding, with duration of 5 min; (2) react, 47.5 hours; (3) settling, 30 min; (4) draw, 5 min. The photoperiod was 16 hours of light + 8 hours of dark and the sequence of the light and dark periods is summarized in Figure 1(b). A constant photoperiod using artificial light allowed us to control the photoperiod and understand the process clearly. The sequence of light and dark causes the alternation of high and low oxygen in the bulk liquid and thus the alternation of aerobic and anoxic conditions, which can be
advantageously exploited for optimizing nitrogen removal from wastewater.

In order to enhance total nitrogen removal and save energy in the PSBR, four different operational regimes were applied during the experiment (Figure 1(b)):

- **LOW**: feeding at the beginning of the light phase (at 8:00) and continuous mixing during the react phase. This basic operational regime was used as the control/starting point, but it did not meet the EU discharge limits in sensitive areas for total nitrogen (15 mg N/L for 10,000 < PE < 100,000; 10 mg N/L for PE > 100,000), requiring progressive optimizations.

- **FED**: feeding at the beginning of the dark phase (at 23:30) and continuous mixing in the react phase; the aim was to create an anoxic phase at the beginning of the cycle (to exploit readily biodegradable COD, RBCOD, and the anoxic conditions of the influent wastewater) in order to promote denitrification and thus to enhance total nitrogen removal.

- **MIX1**: feeding at the beginning of the dark phase (at 23:30) with the mixer off until the beginning of the light phase (at 07:00); the aim was to create an anoxic phase at the beginning of the cycle and to reduce the energy consumption by stopping the mixing for 7.5 hours in the cycle (16% energy saving).

- **MIX2**: feeding at the beginning of the dark phase (at 23:30) with the mixer off during all the dark phases and before the end of the react phase (at 19:00); the aim was to create longer anoxic conditions to enhance denitrification and to reduce the energy consumption by stopping the mixing for 19.5 hours in the cycle (41% energy saving).

Due to the good settleability of the biomass in all the operational regimes, the duration of the settling phase (30 min) was appropriate and remained unchanged.

### Analytical methods

The chemical parameters COD, sCOD, TKN, NH$_4^+$-N, NO$_2^-$-N, NO$_3^-$-N, TP, PO$_4^{3-}$-P and TSS were analyzed in the influent and effluent wastewater according to *Standard Methods* (APHA 2012). Influent TN was estimated as the sum of TKN and NO$_x$-N. The parameter sCOD was measured after filtration of the sample through a 0.45 µm membrane. Fractionation of COD in the influent wastewater aimed at distinguishing soluble and particulate biodegradable COD was performed by respirometry (Vanrolleghem et al. 1999; Ziglio et al. 2001). TSS in the mixed liquor (APHA 2012) was measured to determine the concentration of the microalgal–bacterial consortium in the PSBR. DO and temperature were measured with a Multi3410 meter and an FDO®925 sensor (WTW, Germany). Oxidation–reduction potential (ORP) and pH were measured continuously (every 10 min) with a pH3310 meter coupled with the electrodes Sentix®ORP and Sentix®41, respectively (all from WTW, Germany). PAR was measured with a quantum sensor SQ-520 (Apogee Instruments, USA). Microscopic observations were performed using a Nikon Optiphot EFD-3 Microscope (Nikon, Japan) for the morphological characterization of the microalgal–bacterial consortium.

### Track studies

Track studies were performed in the PSBR to measure the dynamics of the N-forms during a typical cycle. Samples were collected every hour, then filtered and finally analyzed for NH$_4^+$-N, NO$_2^-$-N and NO$_3^-$-N. The volumetric removal of NH$_4^+$-N (or the production of NO$_3^-$-N) expressed as mg N L$^{-1}$ h$^{-1}$ was estimated using the slope of the straight line that interpolates the experimental concentrations over time during the track study. The corresponding specific rate, expressed as mg N g TSS$^{-1}$ h$^{-1}$, was obtained by dividing the volumetric rate by the TSS concentration in the mixed liquor. The removal rate was also expressed per unit of surface, which was useful for appreciating the footprint of the system, taking into account the height of mixed liquor in the reactor.

### Statistical analysis

A Student’s t test, producing a $p$ value, was used to evaluate the significant difference in results. Significance was assumed at $p < 0.05$.

### RESULTS AND DISCUSSION

**PSBR ensures robust and efficient COD removal (>85%) in all the operational regimes**

The profiles of COD and sCOD concentrations in the influent and effluent wastewater during the 6-month experiment are shown in Figure 2. Although large variations of COD concentration in real wastewater were common (257 ± 91 mg/L), the average concentrations in the effluent were 37 ± 7 mg COD/L (min–max range equal to 21–54 mg COD/L), without statistically significant variations among the operational regimes. These results largely met...
This COD fractionation suggests that the effluent soluble COD (22 ± 6 mg/L) was mainly composed of the soluble inert fraction of the influent wastewater (SS) which passed unchanged through the PSBR, while the rest may have been soluble microbial products (SMPs) originating from substrate metabolism and biomass decay (Azami et al. 2012).

The particulate COD in the effluent was associated with some suspended solids floating and leaving the system during discharge. The effluent TSS concentration was around 15 mg TSS/L during the whole experiment, indicating very good sedimentation of the biological flocs developed in the PSBR.

With regard to the removal of nutrients in the whole experiment, average P concentration decreased from 4.8 ± 1.5 mg/L in the influent to 2.5 ± 0.8 mg/L in the effluent (average removal efficiency of 47 ± 19%), while N removal is discussed in depth in the following sections.

**TKN removal >95% occurs in the PSBR at near-zero DO**

The influent TKN (55 ± 20 mg TKN/L) was almost entirely removed in the PSBR, leading to effluent concentrations of 1.5 ± 2.1 mg TKN/L, 0.5 ± 0.7 mgNH₄-N/L and 0.1 ± 0.4 mg NO₂-N/L.

There are three ways in which TKN and ammonium can be transformed by the microalgal–bacterial consortium: ammonia volatilization, assimilation by biomass and nitrification. In particular, volatilization was negligible in this PSBR because the pH in the reactor was 7.6 ± 0.3 on average during the whole experiment, and ammonia volatilization requires relatively high pH values. Nitrogen is also assimilated and incorporated into biomass through cell synthesis during the growth of heterotrophs and, to a lesser extent, autotrophs (Tchobanoglous et al. 2003). The real amount of N removed by net microbial biomass growth was estimated taking into account sludge production, which was 0.03 g TSS/d. Considering the specific content of N in biomass (0.10 g N/g VSS), an amount of 2.4 mg N/L was used daily for the net growth. It accounts for about 4% of the TKN transformed in the system. This highlights that sludge production and nutrient uptake in PSBRs applied to the highly efficient treatment of real municipal wastewater may be significantly lower than that expected in PSBRs specifically designed to maximize algal growth.

The N mass balance reveals that the major role in TKN removal in the PSBR was played by autotrophic nitrification. Karya et al. (2013) also observed that nitrification accounted for more than 80% of ammonium removal during the
treatment of synthetic artificial wastewater in an open photobioreactor inoculated with nitrifying activated sludge and a pure culture of *Scenedesmus* sp.

The very low concentration of NH$_4^+$-N in the effluent wastewater (<1.0 mg/L in 89 samples, with the exception of only three cases) demonstrates that nitrification was complete during the 6-month experiment and not negatively affected by the different operational regimes implemented in the PSBR. As shown in Figure 3, the average TKN removal efficiency was 97 ± 3% and very stable during the whole period, even when the influent TKN concentration reached very high values of around 100 mg TKN/L (Figure 3).

On the basis of these considerations, the nitrification kinetic in the PSBR reactor was investigated in depth. Track studies were carried out in the reactor after the addition of influent wastewater with a known amount of ammonium and the time profiles of NH$_4^+$-N, NO$_2$-N, NO$_3$-N were measured. The results obtained in a typical track study carried out during the light phase of the LOW operational regime (used as control) are shown in Figure 4. The decrease of ammonium occurred rapidly and the initial concentration of 14 mg NH$_4^+$-N/L was completely removed in less than 7 hours, corresponding to a nitrification rate of 1.9 mg N L$^{-1}$ h$^{-1}$. Considering that the biomass in the PSBR was 0.8 g TSS/L, a specific nitrification rate of 2.4 mg NH$_4^+$-N g TSS$^{-1}$ h$^{-1}$ was calculated. Similar results for nitrification kinetics were found during the other operational regimes (FED, MIX1, MIX2), which did not significantly affect ammonium removal.

Considering the hourly nitrification rate, a daily rate of 30 mg NH$_4^+$-N L$^{-1}$ d$^{-1}$ can be calculated for a daily photoperiod of 16/24 hours, which corresponds to a surface removal rate of 5.4 g NH$_4^+$-N m$^{-2}$ d$^{-1}$ (considering the reactor's diameter of 0.13 m, top surface area of 0.0133 m$^2$ and volume of 0.0024 m$^3$). This specific value gives an approximate estimate of the footprint. For a comparison, 10.2 g NH$_4^+$-N m$^{-2}$ d$^{-1}$ was found in a flat PSBR with irradiance an order of magnitude higher than in this experiment (Rada-Ariza *et al.* 2017).

The specific value of 2.4 mg N gTSS$^{-1}$ h$^{-1}$ found for the microalgal–bacterial consortium is similar to the specific nitrification rate reported in the literature for activated sludge systems (Tchobanoglous *et al.* 2003). However, the important difference is that the microalgal–bacterial consortium works with a limited TSS concentration (around 1 g TSS/L) to allow light to penetrate in the biomass, while activated sludge works at 4 g TSS/L. This results in a significant difference in the volumetric nitrification rate of microalgal consortia and activated sludge. However, a further investigation is needed at a real scale to evaluate the volumetric nitrification rate of microalgal–bacteria consortia under more realistic conditions.

From Figure 4 it can be observed that the decrease of ammonium occurred steadily over a long period (>5 hours) with DO concentrations near zero. Since no external aeration was provided to the PSBR, only photosynthetic activity was responsible for oxygen production during the light phase. In the presence of NH$_4^+$, the oxygen demand required for nitrification surpassed the amount produced by photosynthesis, resulting in a near-zero DO concentration. Then, when ammonium was completely depleted, the DO concentration increased gradually, reaching values above 5 mg O$_2$/L.
Concerning the profile of NO₃-N, the initial concentration of 14 mg N/L increased to 19.5 mg N/L at the end of the track study, indicating that only 40% of the ammonium removed was found as nitrates. The explanation of this low increase in nitrates is that simultaneous nitrification–denitrification occurred in this PSBR. The availability of soluble COD, which decreased from 121 mg/L in the influent wastewater to 17 mg/L at the end of the track study, may support denitrification. Analogous observations were found by Tiron et al. (2015) in activated algae granules treating low-strength wastewater in a PSBR. In the present reactor, dense clusters of microalgae, bacteria and abiotic solids were also observed, and this may contribute to initiating conditions for partial denitrification.

Although the near-zero DO phase may suggest the absence of oxygen, it must not be confused with a completely anoxic phase, because photosynthesis occurred and oxygen was continuously produced in the system, but there was not enough oxygen to satisfy the requirements of the biomass. The advantage of nitrification in municipal wastewater by exploiting the oxygen produced in a microalgae–bacterial consortium, without the need of artificial aeration, may offer very powerful opportunities in the evolution of WWTPs. However, further research is needed to prove the feasibility of the process under real conditions in order to evaluate the influence of the main parameters’ variations (e.g. light supply, mass transfer and temperature).

**Total nitrogen <10 mgN/L in the effluent through the enhancement of denitrification in the PSBR**

The LOW operational regime, used as the control, produced a concentration of TN in the effluent wastewater in the range 6.8–40.0 mg N/L, which was over the limits of 10–15 mg N/L (as an annual average) for discharge in sensitive areas imposed by EU Directive 91/271/EEC (Figure 5). The high TN concentration in the effluent, largely composed by nitrates, was the result of: (1) slight assimilation of N by biomass and (2) only partial denitrification.

To enhance TN removal, efforts aimed at increasing the assimilation of N by pure microalgae do not seem to be so effective, because assimilation permits moderate N removal rates, often requires a longer reaction time and thus larger volumes for the treatment of wastewater.

The great advantage of using microalgae–bacterial consortia instead of pure microalgal strains is that bacterial nitrification can be exploited to speed up ammonium removal. Then a higher TN removal can be achieved by ensuring a denitrification step (Rada-Ariza et al. 2017).

In our case, the microalgae–bacterial consortium removed 30 mg N L⁻¹ d⁻¹ for a daily photoperiod of 16/24 hours, which was noticeably higher than those found by Álvarez-Díaz et al. (2017) (N removal of 6.6 mg N L⁻¹ d⁻¹) treating secondary effluents under 14/24 hours of light with Chlorella sorokiniana. A similar N removal (6.63 mg N L⁻¹ d⁻¹) was reported by Cabanelas et al. (2015) in the case of Chlorella vulgaris treating urban wastewater under 14/24 hours of light.

Concerning denitrification, there could still be room for improvement, when suitable conditions were applied in the PSBR. In the LOW operational regime the ORP profile (data not shown) assumed negative values only for 2–4 hours per cycle after the feeding phase, not long enough to create the conditions for full denitrification in the system. Therefore, the enhancement of TN removal was investigated through the optimization of the denitrification process, but without worsening the effluent quality in terms of COD and NH₄-N. This was achieved with the following modifications:

![Figure 5](http://iwaponline.com/wst/article-pdf/78/1/174/475325/wst078010174.pdf)

**Figure 5** | (a) Profiles of total nitrogen in the influent and effluent wastewater compared with the discharge limit (10–15 mgN/L; EU Directive 91/271/EEC); (b) average effluent concentration and standard deviation for each operational regime.
1. The time of feeding (FED operational regime): wastewater was fed during the dark period instead of the light period, in order to provide readily biodegradable COD, required for denitrification, during the dark period when oxygen is consumed but not produced.

2. The intermittent use of the mixer (MIX operational regime): switched on during the light phases, when the biomass requires complete exposure to the light for photosynthesis, and switched off during the night, to avoid turbulence, favor denitrification inside flocs and energy saving; in this phase the biomass settled but the process was not negatively affected.

As shown in Figure 5, the application of the FED and MIX operational regimes resulted in a gradually decreasing profile of TN. The average TN concentration in the effluent was 21 ± 8.8 mg N/L in the LOW operational regime (not optimized), 16.2 ± 3.8 mg N/L in the FED regime, and 10.6 ± 2.9 mg N/L in the MIX1 regime, reaching the lowest average value of 6.3 ± 4.4 mg N/L in the MIX2 regime, which met the EU regulation limits. The enhancement of total N removal due to the progressive optimization of the PSBR was evident only when the MIX regimes were applied. In fact, no statistically significant difference was observed between the LOW and FED regimes, while effluent TN decreased significantly in the MIX1 and MIX2 regimes (p < 0.05).

The decrease of TN occurred along with the extension of the period with negative ORP in the cycle. ORP values in the range −50 to −200 mV, which are favorable for denitrification, were measured for periods of 7–8 hours during the night in the MIX2 regime. In this case, the mixer was switched off in the light phase (at 19:00, for 4 hours before settling and discharge), in order to move toward anoxic conditions before feeding, thus favoring denitrification at the beginning of the subsequent cycle.

**CONCLUSIONS**

The total nitrogen removal in a PSBR fed with real wastewater and operated with a mixed microalgae–bacterial consortium was investigated and optimized to achieve effluent TN concentrations in accordance with EU requirements for discharge in sensitive areas.

COD and TKN removal in the whole experiment were 86 ± 2% and 97 ± 3%, producing an average effluent concentration of 37 mg COD/L and 0.5 ± 0.7 mg NH₄-N/L. In spite of long periods with DO near zero during the PSBR cycle, due to an oxygen demand higher than photosynthesis production, the nitrification rate was 1.9 mg N L⁻¹ h⁻¹, corresponding to a specific rate of 2.4 mg N g TSS⁻¹ h⁻¹. PSBR optimization was aimed at enhancing denitrification through the feeding during the dark and the intermittent use of the mixer; in this way, the total nitrogen in the effluent during the MIX2 regime reached 6.3 ± 4.4 mg N/L.

In conclusion, the PSBR allowed the nitrification of municipal wastewater under laboratory conditions without the need for artificial aeration and with an important energy saving, while simultaneous denitrification was exploited to enhance total nitrogen removal. However, considering the ability of microalgae or associated bacteria to synthesize N₂O, further research is needed to couple the nitrogen balance with a greenhouse gas mass balance.

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