Quantitative Genetic Analyses of Male Color Pattern and Female Mate Choice in a Pair of Cichlid Fishes of Lake Malawi, East Africa

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Abstract

The traits involved in sexual selection, such as male secondary sexual characteristics and female mate choice, often co-evolve which can promote population differentiation. However, the genetic architecture of these phenotypes can influence their evolvability and thereby affect the divergence of species. The extraordinary diversity of East African cichlid fishes is often attributed to strong sexual selection and thus this system provides an excellent model to test predictions regarding the genetic architecture of sexually selected traits that contribute to reproductive isolation. In particular, theory predicts that rapid speciation is facilitated when male sexual traits and female mating preferences are controlled by a limited number of linked genes. However, few studies have examined the genetic basis of male secondary sexual traits and female mating preferences in cichlids and none have investigated the genetic architecture of both jointly. In this study, we artificially hybridized a pair of behaviorally isolated cichlid fishes from Lake Malawi and quantified both melanistic color pattern and female mate choice. We investigated the genetic architecture of both phenotypes using quantitative genetic analyses. Our results suggest that 1) many non-additively acting genetic factors influence melanistic color patterns, 2) female mate choice may be controlled by a minimum of 1–2 non-additive genetic factors, and 3) $F_2$ female mate choice is not influenced by male courting effort. Furthermore, a joint analysis of color pattern and female mate choice indicates that the genes underlying these two traits are unlikely to be physically linked. These results suggest that reproductive isolation may evolve rapidly owing to the few genetic
factors underlying female mate choice. Hence, female mate choice likely played an important role in the unparalleled speciation of East African cichlid fish.

Introduction

The genetic architecture of traits experiencing sexual selection influences phenotypic evolution and speciation [1]. The pattern and rate of response of a trait to selection depends on several factors including the strength of selection, the presence of genetic variation, the mode of inheritance, genetic correlations with other traits, the numbers of genes and alleles underlying a phenotype, the distribution of allelic effects, and patterns of pleiotropy, dominance, and epistasis [1–4]. Despite their importance, empirical studies examining these factors within the context of sexual selection and speciation are sparse, particularly in vertebrates. Here we investigate the genetic architecture of melanistic coloration and female mate choice in the context of the diversification of East African cichlids.

The radiation of East Africa’s cichlid fishes has produced the largest extant vertebrate diversification identified to date. More than 2000 cichlid species have evolved in East Africa within the past 10 million years [5]. This unparalleled vertebrate radiation is primarily concentrated in the three East African Great Lakes: Lake Tanganyika, Lake Victoria, and Lake Malawi. Within Lake Malawi, over 600 cichlid species [6] have evolved from a single common ancestor since the formation of the lake within the past one to four million years [5, 7]. A hallmark of this diversification is the extraordinary diversity of male color patterns. The diversity in color patterns of East African cichlids has long been attributed to sexual selection via female mate choice [8–10] suggesting that strong sexual selection significantly contributed to their speciation [11–15]. Although some have suggested that color patterns may have evolved in response to ecological pressures [16–19] or male-male competition [20–22], many empirical studies have documented the role that male color patterns, i.e. colored ornamentation [23], body hue [24, 25], and melanistic patterns [26–28], play in sexual selection via female mate choice.

Speciation via sexual selection can be greatly facilitated by the physical linkage of the genes contributing to female mate choice and the preferred male phenotype [29, 30]. In many models of the diversification of East African cichlids, incipient species chose their mates based on male color patterns [10, 31, 32]. This targeted mate selection impedes gene flow and contributes to reproductive isolation [33–36]. Speciation can be further expedited when the genes underlying male secondary sexual traits and female mate choice are physically linked. Tight physical linkage can reduce the likelihood of recombination events, which can lead to the accumulation of genetic incompatibilities, and facilitate speciation [10, 29, 31, 37–41].
Despite their importance, the genetic mechanisms contributing to male coloration and female mate choice involved in cichlid diversification remain poorly understood. The genetics of male color pattern in cichlids are relatively well studied [11–13, 42] and several independent studies suggest that male coloration [43–45] is controlled by a small number of genes. However, much less is known about the genetics of female mate choice in this system. One study suggests that female mate choice is influenced by a small number of genes [29]. Additional studies in this or other systems are rare since accurate measurements of female mate choice are difficult to obtain and are time consuming [46, 47]. As a result, even fewer studies have performed joint genetic analyses of male sexual traits and female mate choice [48].

To examine the quantitative genetic basis of male color pattern and female mate choice, we employed a well-established method to quantify color pattern [49] and designed a novel assay to test female mate choice in cichlids. We performed quantitative genetic analyses on both phenotypes to understand the mode of gene action and number of loci involved in these evolutionary significant traits. Further, we tested for physical linkage between color pattern and female mate choice. Specifically, we hypothesized that if the genes underlying color pattern and female mate choice were physically linked, the phenotypes would co-segregate in the hybrid F2 generation. If such linkage is observed, it may provide insights into the mechanisms facilitating the rapid divergence of cichlid species [30].

Materials and Methods

Focal species and their hybrids

The Baylor University Institutional Animal Care and Use Committee approved this research. The protocol number is 08–09. Lake Malawi National Park, the Malawi Fisheries Department granted our sampling permit. *Maylandia zebra* and *Maylandia benetos* are a pair of sympatric rock-dwelling cichlid species from Lake Malawi. While *M. zebra* is a cosmopolitan species that occurs at many locations across the lake, *M. benetos* only inhabits Mazinzi Reef in the southeastern arm of the lake [18]. At Mazinzi Reef, these two species coexist in sympatry but do not hybridize: in over 400 hours of observation, no interspecific courtship or intermediate hybrid individuals were observed (Danley, pers. obs.). The two study species differ primarily in their melanistic markings [28]; *M. zebra* has a bright blue ground color with 5–7 dark body bars, a black cheek, and dark banding on the pelvic fin, whereas *M. benetos* has a bright blue ground color and faint body barring and pelvic fin markings. Although territorial males of both species maintain a bright blue hue, the melanisitic markings of *M. zebra* become more conspicuous when the males are territorial and fade when the male is stressed (Fig. 1). In contrast the melanistic markings of *M. benetos* become faint when territorial and their melanistic markings only become apparent when stressed. Females and subdominant males of both species are drab brown (*M. zebra*) or drab olive (*M. benetos*). Female *M. zebra* display similar melanisitic markings as
observed in male *M. zebra*; however, the markings are much less conspicuous than those of the territorial *M. zebra* males [28]. A previous study of this species pair found that visual cues alone are sufficient for conspecific mate recognition and that melanistic patterning may play a significant role in the reproductive isolation of these species [28]. The mating behaviors of the two species are nearly identical [50] and involve elements such as quiver, lateral display and circling. Both species mate year round, are maternal mouthbrooders, and have a reproductive cycle that varies from 28 to 30 days at 28°C under laboratory conditions (Ding, pers. obs.).

Since the two species do not hybridize naturally, hybrid F₁ offspring were obtained by artificial fertilization. Mature eggs from *M. benetos* and sperm from *M. zebra* were obtained by gently compressing the abdomen of the fish. The gametes were mixed for five minutes in a petri dish (136 mm diameter × 14 mm deep) filled with tank water and the fertilized eggs were transferred into a glass incubator where they were incubated at 28°C until the larvae could swim independently. The reciprocal F₁ cross, *M. zebra* (♀) × *M. benetos* (♂), was created with the same method. F₂ hybrids were obtained by intercrossing the F₁
individuals derived from the cross between *M. zebra* (♀) × *M. benetos* (♂). The 11 *M. benetos* (♀) × *M. zebra* (♂) F₁ broods produced only F₁ females thus no F₂ hybrids could be obtained from this direction. Backcrosses were generated by crossing *M. zebra* (♀) × *M. benetos* (♂) F₁ hybrids to the two parental species. Prior to the F₂ reaching sexual maturity, male and female F₂ hybrids were isolated to avoid any sexual experience. Both parental stocks were reared under laboratory conditions for at least 10 generations. Water temperature was kept between 26° and 28°C. Light was kept at a 12-hour day/night cycle using timer controlled fluorescent lights. Fish were fed a mixture of food flakes twice daily.

**Quantification of color pattern**

Fish were allowed to grow for a minimum of one year. Scales and pelvic fins were collected to quantify the differences in color pattern between the two species. Both males and females were sampled to compare the color pattern within each species and hybrid brood. Scales from the first complete body bar (one scale from immediately below the lateral line) and the distal third of the pelvic fin were collected (Fig. 1). Scale and pelvic fin samples were then emerged in K⁺ rich aggregating fluid to contract melanophores [49]. Melanophore quantity was assessed by counting the number of melanophores which occur in a 0.25 mm² area from both tissues at 30x magnification through a Nikon stereo microscope (SMZ1500).

**Phenotyping of female mate choice**

Mate choice experiments were performed in 110 cm × 28 cm × 30 cm tanks split into three compartments: two male compartments that flanked a central female compartment (Fig. 2). Plastic grating composed of 15 mm squares (‘egg crate’) was used to divide the compartments and produce a false bottom to the tank. The false bottom prevented females from collecting their eggs during mating. Each divider separating the three compartments contained an opening sufficiently large to allow free movement of the test females between compartments, but small enough to prevent the males from escaping their respective areas. A male *M. zebra* was randomly placed in one flanking compartment and a male *M. benetos* was placed in the opposing flanking compartment. Both males were allowed to acclimate for at least 24 hours. In all experimental trials, males were size-matched for standard length (<2 mm difference). The side allocated to the male of each species was randomized to control for side bias. Ten size-matched pairs of males were used in this experiment.

Making accurate measurements of female mate choice can be challenging and time consuming [46, 47]. Several methods have been developed to quantify cichlid fish mate choice including (listed in order of increasing measurement confidence [51]: time in association with a male [27], behavioral assays [29] and egg counts [52]). Because egg counts are a direct rather than a proxy measure of female mate choice, this was the primary metric of female mate choice used in our experiment.
The amount of time a female spent in association with each male in their respective compartments was recorded to evaluate the reliability of association time as an indicator of female mate choice. In addition, male courting effort was quantified to estimate its effects on female mate choice.

In each mate choice trial, a single reproductive female, as determined by its swollen abdomen and slightly protruded genital papilla, was introduced to the central compartment. The behaviors of both males and the female were video recorded for two hours after the introduction of the female to the trial arena. After reviewing the video and verifying that female had visited both male compartments, a 20 minute video clip was used to quantify male courting effort. Within the 20 minute video, three elements of male courting behavior were scored: quiver, lateral display, and circling. While quivering, a male swiftly swims toward a female, exaggeratedly beats its tail, and flexes its body along its long axis. During a lateral display, the male erects its fins and orients parallel or perpendicular to the female in the water column above its territory. If the female does not break off the courtship event, the pair will begin circling which occurs when the male and female swim in tight circles over the spawning area [53]. Thirty randomly selected F2 female mate choice behavior videos were scored to evaluate the effect of male courtship effort on F2 female mate choice. The number of times each male performed a quiver, lateral display, or circling behavior was used to quantify male courting effort (S1 Video. Female mate choice assay tank).

Female mate choice was quantified based on patterns of egg laying. The number of eggs laid beneath each of the three compartments (M. zebra compartment, M. benetos compartment, and/or the central compartment that lacked a stimulus male) was recorded; trials were discarded if eggs were laid in the center partition. Females typically laid eggs within 24 hours of their introduction to the test tank. Two hundred thirty one females were evaluated for their mate choice. Among these, 10 females from each parental species were tested once, whereas 29 F1 and
182 F₂ females were each tested three times against three different pairs of males in three different arenas. The mate choice of the backcrosses was not examined.

**Statistical analyses**

To compare male and female color pattern differences in scale and pelvic fin melanophore counts of different generations, a two-factor analysis of variance (ANOVA) was performed.

We used quantitative genetic methods to investigate the genetic architecture of color pattern and female mate choice. In order to investigate the mode of gene action for melanistic color pattern, we compared the additive and the additive-dominance model using the joint-scaling test [54]. Tests for epistasis on melanistic color pattern were conducted following Lynch and Walsh [55]. Using this method, the test statistic, $\Delta$, is calculated as the difference in means between the F₂ and the weighted means of the parentals and F₁. The joint scaling test and tests for epistasis were not used to investigate the mode of gene action for female mate choice as female mate choice in the parentals was not normally distributed (specifically, the parental variances equaled 0). The number of genetic factors underlying both coloration phenotypes and female mate choice were estimated using the Castle-Wright estimator following Lynch and Walsh [55]. Briefly, using the means and variances of the parental, F₁ and F₂ phenotypic distributions, the number of segregating genetic factors that are likely to be responsible for the quantitative differences of the traits can be inferred. These factors were then adjusted using Zeng’s correction [56]. Assuming an additive genetic effect on female mate choice, we expect the mean of female mate choice in the reciprocal F₁ hybrids and F₂ to be 0. We tested this hypothesis with a one sample t-test.

Female mate choice was quantified using a mate choice index expressed as the number of eggs laid in the *M. zebra* compartment minus the number of eggs laid in the *M. benetos* compartment, divided by the total number of eggs laid in the trial. This index ranges from +1 (females that exclusively choose *M. zebra* males) to −1 (females that exclusively choose *M. benetos* males). The average of the mate choice index was used as an overall estimate of F₁ and F₂ female mate choice.

Following a similar method used by Haesler and Seehausen [29], male courting effort was quantified by regressing the number of male displays (lateral display, quiver, circling) in a 20 minute interval against the amount of time that the female spent in the male’s compartment. The residuals of the male courting effort are independent of the time that the female was present in each male compartment.

To test whether female mate choice was influenced by male courting effort, we separately examined the effect of each male courting effort (lateral display, quiver, circling) on female mating preference: males that successfully mated would be expected to have exerted more effort than those males that were rejected. A Mann-Whitney-Wilcoxon test was used to test for this difference. This test was also performed to evaluate the time spent in association with successful and rejected males. The videos of 24 F₂ females (10 exclusively mated with *M. zebra* and 14...
exclusively mated with *M. benetos*) were selected to evaluate the effect of male courting efforts and the utility of female time in association as a predictor of female mate choice. The six females that mated with both males were excluded from this analysis.

To evaluate female maternal effects, we tested for possible differences in mate choice between the F1 reciprocal crosses with a Welch two sample t-test. To test whether the mate choice in the F2 female population deviates from random mating, a $\chi^2$ goodness of fit test was performed.

Finally, we investigated the phenotypic correlation between color pattern and female mate choice by comparing the color patterns of the F2 females that exclusively mated with *M. zebra* to those that exclusively mated with *M. benetos*. Only those females that mated exclusively with one species were used as these provided the most reliable estimate of mate choice. If the genes contributing to female mate choice were physically linked with the genes influencing color pattern, we would expect the co-segregation of color pattern and mate choice phenotypes in the F2: the F2 females which exclusively mated with *M. benetos* (*M. zebra*) males would resemble *M. benetos* (*M. zebra*) in their color pattern. An ANOVA was used to test for this phenotypic difference between F2 females which exclusively mated with either *M. benetos* or *M. zebra*. All melanophore count data were square root transformed to improve normality. The data analyses were performed in R [57].

**Results**

**Color pattern general statistics**

556 fish were phenotyped in both scale and fin tissues to quantify color pattern phenotypes. No statistically significant difference was found between males and females in both scale ($F=0.264, d.f.=1, p=0.607$) and fin melanophore counts ($F=0.683, d.f.=1, p=0.409$) across all lines. As a result, male and female melanophore data were pooled for subsequent analysis.

Of the 556 individuals phenotyped for color pattern, 86 (61 males, 25 females) were *M. zebra* and 47 (34 males, 13 females) were *M. benetos*. Ninety four F1 hybrids (48 males, 46 females) derived from five independent broods (2 *M. zebra* ($\varphi$) $\times$ *M. benetos* ($\sigma$) broods with 17 individuals and 3 *M. benetos* ($\varphi$) $\times$ *M. zebra* ($\sigma$) broods with 77 individuals) were phenotyped for color pattern. Two hundred seventy F2 individuals (62 males, 208 females) from three independent crosses were phenotyped for color pattern. A total of 60 backcross individuals were included in the color pattern analysis: 39 individuals (20 males, 19 females) were the result of an F1 (*M. zebra* ($\varphi$) $\times$ *M. benetos* ($\sigma$)) backcross to *M. zebra*, while 21 individuals (14 males, 7 females) were derived from F1 (*M. zebra* ($\varphi$) $\times$ *M. benetos* ($\sigma$)) backcross with *M. benetos*.

Scale melanophore counts varied across the broods. *Maylandia zebra* had the highest mean melanophore count (mean (SD)=79.45 (19.92)). The *M. zebra* backcross had the next largest mean (SD) scale melanophore count at 68.56...
(14.91). Both directions of the F₁ were intermediate in their scale melanophore counts. The melanophore count for F₁ (M. benetos (♀) × M. zebra (♂)) was 43.75 (15.68). The melanophore count for F₁ (M. zebra (♀) × M. benetos (♂)) was 38.28 (13.44). No statistically significant differences were observed in the scale melanophore counts in the F₁ based on the direction of the cross (t = -1.61, d.f. = 45.14, p = 0.12). The mean (SD) count of both directions of the F₁ was 39.95 (14.29). The F₂ melanophore scale counts were intermediate as well and had a mean (SD) of 41.06 (13.77) melanophores. The M. benetos backcross, (M. zebra (♀) × M. benetos (♂)) with M. benetos, had a mean (SD) scale melanophore count of 23.16 (5.19). With a mean (SD) scale melanophore count of 17.48 (9.72), M. benetos had the fewest number of scale melanophores (Fig. 3).

Fin melanophore counts were distributed in a similar pattern as the scale melanophores. Maylandia zebra had the highest mean (SD) melanophore count (110.5 (29.51)). The backcross to M. zebra (F₁ (M. zebra (♀) × M. benetos (♂)) backcross to M. zebra) had a mean (SD) of 77.24 (26.92) melanophores on the fins. The melanophore count for M. benetos (♀) × M. zebra (♂) F₁ cross was 88.70 (26.55) while the melanophore count for the M. zebra (♀) × M. benetos (♂) F₁ cross was 85.56 (28.97). No significant differences were observed in the fin melanophore counts in the F₁ reciprocal crosses (t = -0.50, d.f. = 53.16, p = 0.618). The overall F₁ hybrid mean (SD) was 86.49 (28.16) melanophores on the fin. F₂ individuals had a mean (SD) fin melanophore count of 70.00 (23.63). With 45.16 (19.76) mean (SD) number of melanophores on the fin, the backcross (M. zebra (♀) × M. benetos (♂) backcross to M. benetos) began to resemble M. benetos which had a mean (SD) fin melanophore count of 19.73 (9.95) (Fig. 4).

The ANOVA results indicated that both scale (F = 149.913, d.f. = 5, p = 2e⁻¹⁶) and fin (F = 118.772, d.f. = 5, p = 2e⁻¹⁶) melanophore densities were significantly different among parental, F₁, F₂ and backcross generations. Tukey’s post-hoc test revealed that melanophore counts in both scale (p < 1e⁻¹⁰) and fin (p < 1e⁻¹⁰) were much higher in M. zebra than that in M. benetos. Hybrid individuals had intermediate melanophore counts on both tissue types (Table 1, 2). For scale melanophore counts, Tukey’s multiple comparisons suggested that M. zebra was slightly, though significantly, different from the backcross with M. zebra (p = 0.045), while no significant difference existed between M. benetos and the backcross with M. benetos (p = 0.083). Further, no statistically significant difference was found between the F₁ and F₂ generations in scale melanophore count (p = 0.977). For the melanophore count in fin tissues, M. zebra fin melanophore counts were significantly greater than the backcross with M. zebra (p < 1e⁻¹⁰). Likewise, M. benetos had significantly fewer fin melanophores than the backcross to M. benetos (p < 1e⁻¹⁰). Significant differences between F₁ and F₂ generations in melanophore count in fin was observed as well (p < 1e⁻⁵).

Quantitative genetic analysis of color pattern

Neither the additive model (Table 1; χ²₁ = 22.19, p = 0.00018) nor the additive-dominance model (Table 1; χ²₂ = 22.10, p = 6.22e⁻⁵) explained the pattern of
Furthermore, the additive-dominance model did not significantly improve the fit of the model to the observed data (Table 1; $\Lambda_1=0.09$, $p=0.76$). The inheritance of melanophore in the pelvic fin tissues showed a similar pattern: both the additive model (Table 2; $\chi^2_1=101.33$, $p<10^{-10}$) and additive-dominance model were rejected (Table 2; $\chi^2_2=18.58$, $p=0.0003$). However, the additive-dominance model did significantly improve the fit of the model to the observed data (Table 2; $\Lambda_1=82.74$, $p<10^{-10}$). The
rejection of the additive and additive-dominance models suggests the action of epistasis, which was supported for both the scale ($\frac{|\Delta|}{\sqrt{Var(\Delta)}} = 0.019 < 1.96$) and fin ($\frac{|\Delta|}{\sqrt{Var(\Delta)}} = 0.16 < 1.96$) phenotypes. Genetic factor analysis suggested that a large number of genetic factors control melanophore patterning of the scales and pelvic fins. The estimated minimum number of genetic factors ranged from $-5$ (SD=11.02) to $-30$ (SD=55.09) for scale and 29 (SD=229.27) to 141 (SD=1496.37) for the pelvic fin depending on the size of allelic effects (Table 3).

**Test for association between male courting effort and female mate choice**

No statistically significant effect of male courtship effort on female preference was observed for any of the three male behaviors: lateral display ($p=0.17$, Fig. 5a),

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### Table 1. The square root transformed means (standard errors) of melanophore counts (within a 0.25 mm$^2$ area for scales (Fig. 1).

| Scale                  | Model                     | Expected Additive | Expected Additive-Dominance |
|------------------------|---------------------------|-------------------|----------------------------|
| **Observed**           |                           |                   |                            |
| *M. zebra*             | 8.84 (0.12)               | 9.01 (0.10)       | 9.03 (0.11)                |
| *M. benetos*           | 3.97 (0.21)               | 3.63 (0.11)       | 3.67 (0.16)                |
| F$_1$                  | 6.22 (0.12)               | 6.32 (0.05)       | 6.30 (0.10)                |
| F$_2$                  | 6.31 (0.07)               | 6.32 (0.05)       | 6.32 (0.05)                |
| Backcross to *M. zebra*| 8.23 (0.15)               | 7.67 (0.06)       | 7.66 (0.06)                |
| Backcross to *M. benetos*| 4.78 (0.13)            | 4.98 (0.07)       | 4.98 (0.07)                |

$\chi^2_{2}=22.10, p<0.0001$ $\chi^2_{1}=22.19, p<0.0001$ $\Lambda_1=0.09, p=0.76$

Given are empirical counts, expected values assuming an additive genetic model, and expected values assuming an additive-dominance model. $\chi^2$ tests were used to compare the genetic models to the observed data. A likelihood-ratio test was used to test if the additive-dominance model better explained the data than the additive model ($\Lambda$).

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### Table 2. The square root transformed means (standard errors) of melanophore counts (within a 0.25 mm$^2$ area for pelvic fins (Fig. 1).

| Fin                     | Model                     | Expected Additive | Expected Additive-Dominance |
|-------------------------|---------------------------|-------------------|----------------------------|
| **Observed**            |                           |                   |                            |
| *M. zebra*              | 10.42 (0.16)              | 11.00 (0.13)      | 10.25 (0.15)               |
| *M. benetos*            | 4.30 (0.17)               | 5.13 (0.13)       | 4.35 (0.16)                |
| F$_1$                   | 9.17 (0.16)               | 8.07 (0.07)       | 9.06 (0.12)                |
| F$_2$                   | 8.25 (0.09)               | 8.07 (0.07)       | 8.18 (0.06)                |
| Backcross to *M. zebra* | 8.66 (0.25)               | 9.53 (0.08)       | 9.65 (0.08)                |
| Backcross to *M. benetos*| 6.55 (0.35)            | 6.59 (0.09)       | 6.70 (0.09)                |

$\chi^2_{2}=18.58, p<0.001$ $\chi^2_{1}=101.33, p<0.0001$ $\Lambda_1=82.74, p=0.0001$

Given are empirical counts, expected values assuming an additive genetic model, and expected values assuming an additive-dominance model. $\chi^2$ tests were used to compare the genetic models to the observed data. A likelihood-ratio test was used to test if the additive-dominance model better explained the data than the additive model ($\Lambda$).

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quiver ($p=0.20$, Fig. 5b) and circle ($p=0.89$, Fig. 5c). However, females did spend significantly more time associating with males they eventually mated with compared to the males that were rejected: in 21 of 24 trials the female spent more time with the preferred male ($p=1.82e^{-6}$, Fig. 5d).

**Quantitative genetic analysis of female mate choice**

The mean (SD) female mate choice index for *M. zebra* and *M. benetos* was 1(0) and −1(0), respectively and all hybrid crosses had intermediate means with varying amounts of dominance in the direction of *M. zebra* (Fig. 6). The mean (SD) mate choice for the 17 F₁ females derived from the cross between *M. benetos* (♀) × *M. zebra* (♂) equaled 0.42 (0.64) and was significantly greater than 0 (i.e. there was significant dominance in the direction of *M. zebra*) ($t=2.6887$, $d.f.=16$, $p=0.008$). The mean (SD) of mate choice for the 12 F₁ hybrid derived from the cross between *M. zebra* (♀) × *M. benetos* (♂) was 0.89 (0.24) and was significantly greater than 0 ($t=12.93$, $d.f.=11$, $p=2.687e^{-8}$). The Welch two sample t-test showed a significant difference between the two reciprocal crosses ($t=2.74$, $d.f.=21.61$, $p=0.023$). As only one direction of the cross (*M. zebra* (♀) × *M. benetos* (♂)) could be used to make F₂ hybrids, genetic factor analyses of female mate choice, which requires F₁ data, utilized only the F₁ data from this cross. The mean of the mate choice index for F₂ hybrids was 0.097 (0.68) and was significantly greater than 0 ($t=1.9387$, $d.f.=181$, $p=0.027$) (Fig. 6). The number of F₂ hybrids that mated exclusively with *M. zebra* or *M. benetos* was 39 and 31, respectively. One hundred twelve F₂ females mated with both *M. zebra* and *M. benetos* (Table 4). This pattern of female mate preference in the F₂ significantly deviated from random mating ($\chi^2=65.68$, $d.f.=2$, $p<0.0001$). The minimum number of genetic factors affecting female mate choice was 1.16 (0.02). After Zeng’s correction [56] as described in Lynch and Walsh [55] was applied, the estimated number of genetic factors ranged from 1.2 to 1.9, depending on the distribution of allelic effects (Table 5).

**Test for phenotypic correlation between female mate choice and male color pattern**

When the melanophore counts of those F₂ that mated exclusively with *M. benetos* were compared to the melanophore counts of those F₂ that mated exclusively with *M. zebra* no significant differences were observed in either the scale ($F=1.4727$, $d.f.=1$, $p=0.2296$) or fin data ($F=0$, $d.f.=1$, $p=0.9999$) (Fig. 7a, b).

**Discussion**

**Genetic factors influencing melanistic patterning**

The genetic architecture of sexually selected traits remains poorly understood. To gain a better understanding of the number of genetic factors involved, the modes of gene action, and the genetic correlation between male signaling and female
mating preference phenotypes, we conducted a biometric analysis of an East African cichlid pair. Our results suggest that many loci are involved in melanistic patterns of scales and pelvic fins. This is indicated by the large number of genetic factors that was estimated for fin melanophore counts and the negative number of genetic factors for scale melanophores. The estimated negative numbers of factors may be a result of many genes of small effect contributing to a phenotype or violations of the assumptions of the Castle-Wright method. The Castle-Wright method assumes that alleles are alternatively fixed in two parental lines and have

| $C_a$ | Scale $n_a$ | Fin $n_a$ |
|-------|-------------|-----------|
| 0     | -5.33       | 29.02     |
| 0.25  | -6.92       | 36.02     |
| 1     | -11.67      | 57.03     |
| 4     | -30.69      | 141.08    |

Table 3. The estimated effective number of factors (standard deviation) influencing scale and fin melanophore counts for a range of allelic effects ($C_a=0$ assumes the equivalence of allelic effects; $C_a=1$ assumes a normal distribution of allelic effects; $C_a=4$ assumes the allelic effects have a negative exponential distribution; $C_a=4$ assumes that allelic effects have a leptokurtic distribution).

Fig. 5. Comparisons of the residuals derived from the regression of male lateral displays (a), quivers (b), and circles (c) against the amount of time that female stayed in each side of the tank between the successfully mated males and rejected males; further comparison of the time in association (d) that female spent with the successful mated males and rejected males is shown.

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equal allelic effects [55]. However, our results clearly suggest that genes with partially dominant effects underlie the melanistic patterning and female mate choice. In addition, a relatively small sample size for the backcross lines can affect the estimates as well [55]. Therefore, the estimated number of genetic factors should only be considered as a lower bound of the genetic factors underlying these traits.

Our finding differs from previous investigations of the genetic architecture of coloration in East African cichlids. Using a similar biometric approach, Barson et al. [58] found that blue body coloration of cichlids was influenced by 4–7 genes,

![Figure 6. Mean (x axis) and variance (y axis) of the female mate choice index in the parentals, and F1 and F2 hybrids.](image)

### Table 4. Results of the female mate choice assay.

|                | M. zebra | M. benetos | M. zebra & M. benetos | Total (Mean of the mate choice index) |
|----------------|----------|------------|-----------------------|--------------------------------------|
| M. zebra       | 10       | 0          | 0                     | 10 (1)                               |
| M. benetos     | 0        | 10         | 0                     | 10 (−1)                              |
| F1 (M. benetos ♀ × M. zebra ♂) | 7       | 0          | 10                    | 17 (0.42)                            |
| F1 (M. zebra ♀ × M. benetos ♂) | 9       | 0          | 3                     | 12 (0.89)                            |
| F2             | 39       | 31         | 112                   | 182 (0.097)                          |

Females of M. zebra, M. benetos, F1 and F2 were tested in a two way mate choice tank (Fig. 2). The numbers of individuals tested for female mate choice and the mean of the mate choice in each generation are given.

![Table 4](image)
Magalhaes and Seehausen [45] found that 2–4 genes appear to determine red coloration in a hybrid cross of Lake Victoria cichlids, and O’Quin et al. [43] found that 1–4 genes control the pigmentation differences between a pair of Lake Malawi cichlids one of which, *M. zebra*, is also a focal species of this study. Additional efforts by O’Quin et al. (2013) identified one QTL associated with pelvic fin melanophore pigmentation but failed to identify any significant QTL for melanophore counts in scales and a number of additional tissue types including the caudal fin and cheek. The authors suggested that multiple genes with small effects were underlying the number of melanophores in these areas of the body which is consistent with our findings. A recent mapping study identified 41 QTL and 13 epistatic interactions were underlying the melanistic patterning in cichlids [59]. All of these studies consistently demonstrated that the additive genetic model is inadequate for explaining the observed phenotypic variance. In our experiment, both the additive and the additive-dominance models of inheritance were rejected when examining the distribution of melanistic phenotypes in the hybrid crosses, while the presence of epistatic interactions was supported.

Table 5. The estimated effective number of factors influencing cichlid female mate choice for a range of allelic effects (*C_a* = 0 assumes the equivalence of allelic effects; *C_a* = 0.25 assumes a normal distribution of allelic effects; *C_a* = 1 assumes the allelic effects have a negative exponential distribution; *C_a* = 4 assumes that allelic effects have a leptokurtic distribution).

| *C_a* | n  | Standard deviation |
|-------|----|--------------------|
| 0     | 1.2| 0.02               |
| 0.25  | 1.2| 0.04               |
| 1     | 1.3| 0.09               |
| 4     | 1.9| 0.57               |

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![Fig. 7. Comparison of F2 scale (a) and fin (b) phenotypes between individuals that exclusively mated with *M. zebra* or *M. benetos*.](image)

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Male courting effort versus female mate choice

Parental investment is largely asymmetrical in haplochromine cichlids, including the vast majority of Lake Malawi cichlids: females are promiscuous mouth brooders which produce relatively large, energetically expensive eggs that are fertilized by males that contribute little other than sperm [60]. This asymmetrical investment in parental care is thought to have led to the evolution of strong female mate choice [61] and vigorous male courtship [62] in this system [9, 63, 64].

In our mate choice trials, females were allowed to interact with males. As a result, females that stayed longer in one side of the arena tended to elicit more male courting efforts. Many studies suggest that vigorous male courting effort can significantly affect female mate choice [65–67]. In our study, by examining the effect of male courtship effort via statistically removing the effect of time spent with a female, we found that none of the three male courting behaviors significantly influenced female mate choice. Consistent with Haesler and Seehausen [29], these results suggest that female mate choice probably occurs prior to the onset of intense courtship [50]. Alternatively, male courtship displays may have an indirect effect on reproductive success or may be maintained through pleiotropic effects. Nonetheless, we did find a significant difference between the times that female spent with successful males compared to the rejected males. Time spent in association may be a good indicator of female preference particularly in interspecific comparisons. However this measure lacks the necessary accuracy when examining individual preferences within F2 populations.

Segregation of female mate choice among F2 female hybrids

Quantifying the genetic basis of female mate choice can improve our understanding of reproductive isolation and speciation. In our study, females of the parental species, *M. zebra* and *M. benetos*, mated exclusively with conspecific males which is consistent with the previous study of these species [28].

Despite significant differences in the mate choice of the reciprocal hybrid F1 females, F1 hybrids derived from both directions exhibited a preference for *M. zebra* males and an additive genetic effect in the F1 was rejected. Likewise, the F2 females exhibited a dominant preference for the *M. zebra* phenotype; however, the degree of dominance in the F2 was much smaller than in either of the F1 lines. Why the degree of dominance of mate choice diminished in the F2 is unclear. It may be the result of a ‘parent of origin’ epigenetic phenomenon [68–71] or masking effects by maternal imprinting on an intermediate F1 phenotype in the F2 [72] (early imprinting would not have occurred in the F1 as these were the only lines raised entirely outside of a female’s buccal cavity and would have escaped any developmental imprinting). Nonetheless, female mate choice significantly deviates from random mating in the F2 population indicating that female mate choice has a strong heritable component. In addition, it is clear that the alleles influencing female mate choice in the two parental species have segregated in the
F₂ population to recapitulate the parental phenotypes. The strong genetic component of female mate choice is further corroborated by our behavioral experiments in which no statistical correlation between male courting effort and female mate choice was observed.

Future studies should further explore the dominance of *M. zebra* mate preferences. Such additional studies could provide valuable insight into the diversification of Lake Malawi’s cichlids. Directional selection is believed to drive the evolution of dominance [73–75] and the dominance of *M. zebra* mate preferences may reflect ongoing directional selection operating on mate preferences. In addition, dominance could affect the tempo of genetic isolation of diverging lineages as a partially dominant allele will increase in frequency faster than a recessive allele [76]. Furthermore, the pattern of dominance biased preference for *M. zebra* would lead to biased introgression into the *M. zebra* gene pool. Lastly, the observed dominance, if confirmed, may be, one of the few documented empirical examples of “Haldane's sieve” [77] in female mate choice. “Haldane’s sieve” predicts that if species-level sexual isolation results primarily from directional selection, then interspecific hybrid females should discriminate against males of either or both parental types [73]. It is also worth noting that only female hybrids were produced from the 11 *M. benetos* (♀) × *M. zebra* (♂) F₁ broods in our attempt to make the reciprocal cross. This might suggest that postzygotic isolation has started to evolve between the studied species. If further experiments corroborate our observation, this might be among the first experimental evidence for Haldane’s rule in cichlids [78].

**Genetic factors influencing female mate choice**

Female mate choice is thought to accelerate the rate of species divergence [79]. Quantitative genetic models of mate choice [38, 39, 80] and empirical studies [37, 40, 81] have suggested that the number of genes contributing to a phenotype influences the pace of phenotypic evolution. As such, quantifying the genetic elements influencing female mating preference in East African cichlids may provide a genetic mechanism which may have contributed to the extraordinary speed of this species radiation. In our study, we found that female mate choice is controlled by few genetic factors. Our result is consistent with an empirical study of a pair of Lake Victoria cichlids [29] and suggests that the rapid diversification of East African cichlids may have been facilitated by few genetic factors underlying female mate choice.

**Correlation between female mate choice and color pattern**

In a previous study, visual cues, and melanistic markings in particular, were sufficient for female mate choice in this species pair [28]. Furthermore, quantitative genetic models and empirical studies suggest that a physical linkage of female mate choice and male secondary sexual traits likely accelerates speciation in cichlids [3, 30, 37, 82]. However in this study, female mate choice and
melanistic color pattern were not associated with each other in F_{2} females and there was no evidence of linkage between the male signal and female mate preferences. However, as noted by Van der Sluijs et al. [48], if many genes underlie both female mate choice and male secondary traits, it may be difficult to detect weakly linked genes. Nonetheless, our results suggest that factors other than the physical linkage of melanistic patterns and mate choice have contributed to the divergence of this species pair. Future experiments are needed to examine the role of physical linkage of genes contributing to preferred male phenotypes and female mate preferences in the broader East African cichlid radiation.

Conclusions

The genetic mechanisms underlying the extraordinary East African cichlids radiation still remain elusive. In this study, using a pair of closely related, yet reproductively isolated Malawian cichlids as the model, we investigated the genetic architecture of two evolutionary significant traits, male color pattern and female mate choice, with quantitative genetic analyses. Our results suggest that melanistic color patterns are influenced by many non-additively acting genetic factors, while female mate choice may be controlled by a few non-additive genetic factors. Female mate choice is a heritable trait and male courting effort had little influence on it. Furthermore, our joint analysis of color pattern and female mate choice suggests that the genes underlying these two traits are unlikely to be physically linked. Cichlids reproductive isolation may evolve rapidly owing to the few genetic factors underlying the female mate choice. The genetic nature of male color pattern is even more complex, multiple loci with dominant and epistatic interaction were involved in its formation. Further studies are needed to identify the loci underlying female mate choice and male color pattern to understand the role these evolutionary significant traits played in cichlid speciation.

Supporting Information

S1 Video. Female mate choice assay tank.
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Author Contributions
Conceived and designed the experiments: BD AEH PDD. Performed the experiments: BD DWD. Analyzed the data: BD MC. Contributed reagents/materials/analysis tools: BD PDD. Wrote the paper: BD MH PDD.

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