Investigation on the immunomodulatory activities of *Sarcodon imbricatus* extracts in a cyclophosphamide (CTX)-induced immunosuppressed mouse model

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**Aims:** *Sarcodon imbricatus*, an edible fungus, is widely used in Asian medicine because of its significant pharmacological activities. In the present study, we investigated the immunomodulatory effects of polysaccharide-enriched *S. imbricatus* extracts (SP) in cyclophosphamide (CTX)-induced immunosuppressed mice.

**Results:** Astragalus polysaccharide (AP) was used as a positive control. Compared with CTX-induced immunosuppressed mice, thirty-day SP treatment strongly enhanced the organ indexes of spleen and thymus and suppressed hind paw swelling. Both AP and SP increased the serum levels of immunoglobulin (IgA, IgG, and IgM), and suppressed the overproduction of interleukin-2 (IL-2). Moreover, SP reduced methane dicarboxylic aldehyde levels, and increased the total antioxidant capacity, superoxide dismutase, and glutathione peroxidase in both serum and liver tissues of CTX-induced immunosuppressed mice.

**Conclusion:** *S. imbricatus* extracts significantly improved immune function in CTX-induced immunosuppressed mice via modulation of oxidative systems.

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1. Introduction

Immunity is a physiological function of human bodies that identifies “self” and “non-self” components, undermines and excludes antigenic material, and maintains human health (Viladomiu et al., 2016; Halim et al., 2017). Immunological homeostasis, immune surveillance, and immune defense are three immune system functions. Immunosuppression is observed in various pathophysiological and clinical conditions (Wang et al., 2015; Chouhan et al., 2015). Cyclophosphamide (CTX) is clinically used for tumor therapy and exhibits strong immunosuppressive effects in patients, by causing damage to the immune system and hematopoietic function (Safi et al., 2015a,b; Díaz-Montero et al., 2012). In addition, CTX causes senescence by disturbing the balance of the oxidation system.

Commonly used regulatory medicines may exhibit adverse effects, including general malaise and/or neurotoxicity (Wang et al., 2015; Luo et al., 2016; Pionka, 2013; Markman, 1996). Numerous studies have demonstrated that active ingredients derived from herbs, such as polysaccharides, alkaloids, and sapo-nins, can modulate immune function in humans and animals (Kuang et al., 2011). By regulating the oxidation system and relieving inflammation, *Cordyceps militaris* fruit bodies improved membranous glomerulonephritis in rats (Song et al., 2016; Vine et al., 2015a, 2015b; Muhammad et al., 2017). Previously, we found that *Tricholoma matsutake* enhanced immunity in CTX-induced immunosuppressed mice by promoting the secretion of immunomodulatory molecules. *Sarcodon imbricatus*, a well-known edible fungus, has been widely used in Asian medicine because of its pharmacological activities (Khalig et al., 2016; Vizzini et al., 2012). However, most studies regarding *S. imbricatus* have focused on the separation and analysis of its components (Sułkowska-Ziaja et al., 2014; Ishaq and Jafri, 2017); its immunomodulatory function has not previously been systematically studied.

In the present study, the immunomodulatory activities of *S. imbricatus* were determined in CTX-induced immunosuppressed...
mice. The mechanisms of S. imbricatus-mediated immunomodulatory effects were determined by analyzing the levels of immunoglobulin, inflammatory factors, and oxidation related kinases.

2. Methodology

2.1. Animal experiments

S. imbricatus fruit bodies were extracted twice with 20-fold double distilled (D.D.) water at 95 °C for 4 h. The supernatant was collected and freeze dried prior to use. Polysaccharide-enriched S. imbricatus extracts were denoted as SP.

The experimental protocol was approved by the Institution Animal Ethics Committee of Jilin University. Six-week-old KunMing (KM) mice (18–22 g) were purchased from the Experimental Research Center of Medical Animal (Guangdong, China, SCXK (YUE)2013-0002), and housed under a 12-h light/dark cycle at 23 ± 1 °C with water and food available ad libitum.

Sixty mice were injected with CTX (40 mg/kg) subcutaneously for seven days. An additional 12 mice were injected with normal saline and used as a control group (CTRL). CTX-treated immunosuppressed mice were randomly separated into five groups (n = 12), and orally treated with either 80 mg/kg of Astragalus polysaccharide (AP + CTX group; positive control mice) or 50, 100, or 200 mg/kg of SP (SP + CTX groups) for thirty days. Control mice were orally treated with 0.1 mL/kg of normal saline for thirty days. The body weight of each mouse was measured on days 1, 10, and 30.

After delayed-type hypersensitivity testing, blood was sampled from each group. The mice were then sacrificed via injection with 200 mg/kg of pentobarbital. The liver, spleen, and thymus of each mouse were removed and weighed. Organ indexes were calculated as the organ weight divided by the total body weight.

### Table 1
Effects of SP and AP on hind paw swelling and organ indexes of spleen and thymus.

| Treatment | Spleen index (g/kg) | Thymus index (g/kg) | Hand paw swelling (mm) |
|-----------|---------------------|--------------------|------------------------|
| CTRL      | 2.0 ± 0.2           | 1.8 ± 0.1          | 0.9 ± 0.3              |
| Model     | 1.6 ± 0.2**         | 1.1 ± 0.4**        | 0.3 ± 0.2***           |
| SP (mg/kg)| 50                  | 1.8 ± 0.4**        | 1.6 ± 0.4              |
|           | 100                 | 1.9 ± 04**         | 1.7 ± 0.3              |
|           | 200                 | 2.1 ± 0.5**        | 1.8 ± 0.4             |
| AP (mg/kg)| 80                  | 1.9 ± 0.3          | 1.8 ± 0.7              |

Data are expressed as mean ± S.D. (n = 12). CTX: cyclophosphamide. SP: S. imbricatus extracts. AP: Astragalus polysaccharides.

##** P < 0.05 versus control group.
##*** P < 0.001 versus control group.
* P < 0.05 versus CTX mice.
** P < 0.01 versus CTX mice.

**Fig. 1.** SP-regulated serum levels of (A) IgA, (B) IgG, (C) IgM, and (D) IL-2 in CTX-induced immunosuppressed mice. Data are expressed as mean ± S.D. (n = 12). # P < 0.05 and ### P < 0.001 versus control group. * P < 0.05 and *** P < 0.001 versus CTX mice. CTX: cyclophosphamide. SP: S. imbricatus extracts. AP: Astragalus polysaccharides.
2.2. Delayed-type hypersensitivity (DTH) tests

On day 26, the mice were sensitized via intraperitoneal injection with 0.2 mL of 2% (v/v) sheep red blood cells (SRBC) for four days. The sensitized mice were challenged via injection with 20 μL of 20% (v/v) SRBC into the measured hind paw. The thickness of the hind paw was recorded at 24 h post-injection.

2.3. Biochemical index measurements

The levels of oxidation factors (methane dicarboxylic aldehyde (MDA), total antioxidant capacity (T-AOC), superoxide dismutase (SOD), and glutathione peroxidase (GSH-Px)), immunoglobulin (IgA, IgG, and IgM), and interleukin-2 (IL-2) in serum and/or liver tissues were determined using enzyme-linked immunosorbent assay (ELISA) kits (Calbiotech, USA) following the manufacturer’s protocols.

2.4. Statistical analyses

Data were expressed as mean ± S.D. A one-way analysis of variance (ANOVA) followed by Dunn’s test using SPSS 17.0 software (IBM Corporation, Armonk, USA) were used to determine statistical significance. *P < 0.05 was recognized as significant.

3. Results and discussion

3.1. Effects of SP on organ indexes and hind paw swelling

Low thymus and spleen indexes were exhibited by CTX-induced immunosuppressed mice (*P < 0.05; Table 1). In contrast, SP at 100 and 200 mg/kg increased spleen indexes, and AP at 80 mg/kg and SP at all of the administered doses improved thymus indexes (*P < 0.05; Table 1). The thymus transports activated T cells into blood circulation and promotes mast cell development (Li et al., 2015; Rahman et al., 2017). The spleen synthesizes immune effector molecules and promotes phagocytosis of granulocytes (Hassanpour et al., 2013; Atta et al., 2017). Hence, the enhanced organ indexes exhibited by SP-treated mice are beneficial to immunoregulatory functions.

SP and AP significantly suppressed hind paw swelling compared to CTX-induced immunosuppressed mice (*P < 0.05; Table 1). Hind paw swelling indicates the strength of delayed allergy (DTH), which is mediated by T cells involved in cellular immunity (Huang et al., 2015; Dai et al., 2014; Sarfraz et al., 2017). Data suggest that the immunomodulatory activity of SP may be related to cellular immunity.

3.2. Upregulation of immunoglobulin and IL-2 by SP

Low immunoglobulin levels (including IgA, IgG, and IgM) were found in CTX-induced immunosuppressed mice. Immunoglobulin levels were significantly increased after thirty-day SP treatment (*P < 0.05; Fig. 1A–C). Foreign substances can be identified and neutralized by immunoglobulin. During humoral immune response, opsonophagocytosis is initiated by increased IgA and IgG levels upon antigen exposure. Thus, humoral immunity may be involved in SP-mediated immunomodulatory effects in CTX-induced immunosuppressed mice.

Lower levels of IL-2 were determined in CTX mice than in non-treated mice (*P < 0.05; Fig. 1D). Thirty-day SP treatment resulted in >25% enhancement of IL-2 levels (*P < 0.05; Fig. 1D). Cytokines are necessary for the differentiation of memory T cells. Perturbations

Fig. 2. SP-regulated levels of (A) MDA, (B) T-AOC, (C) SOD, and (D) GSH-Px in serum and liver tissues of CTX-induced immunosuppressed mice. Data are expressed as mean ± S.D. (n = 12). *P < 0.05 and **P < 0.01 versus control group. †P < 0.05 and ††P < 0.01 versus CTX mice. CTX: cyclophosphamide. SP: S. imbricatus extracts. AP: Astragalus polysaccharides.
in cytokines levels may cause various pathologies, including immunodeficiency, autoimmunity, and atopic disease (Sun et al., 2011). IL-2, a pleiotropic cytokine, drives T-cell growth, augments NK cytolytic activity, induces the differentiation of regulatory T cells, and mediates activation-induced cell death (Zhao and Ashraf, 2016; Liao et al., 2011). Therefore, the immune regulatory activities of SP may be partially attributed to the regulation of immunoglobulin and IL-2 levels.

3.3. SP regulation of oxidative factors

MDA levels directly indicate the degree of lipid oxidation in the body and indirectly indicate the degree of cell damage. SOD, GSH-Px, and AOC levels represent the ability of an organism to remove oxygen free radicals. Compared to control mice, CTX enhanced MDA levels, and significantly reduced the activities of SOD, GSH-Px, and AOC in serum and liver tissues (P < 0.05; Fig. 2). These levels were normalized by SP treatment (P < 0.05; Fig. 2).

Reactive oxygen species (ROS) are highly reactive molecules containing oxygen, and can modify DNA and proteins (Zafar et al., 2016). Under normal conditions, ROS are balanced by the antioxidation defense system, including SOD and GSH-Px. When an imbalance between oxidants and antioxidants occurs, oxidative stress is reached (Portal-Nunez et al., 2016). SOD and GSH-Px protect against oxidative cell damage by clearing excessive MDA and ROS (Ma et al., 2016; Sies, 1997). Previously, we demonstrated that Cordyceps militaris protects rats against membranous glomerulonephritis by relieving oxidative damage (Liu et al., 2015; Sarfraz et al., 2016; Song et al., 2016). Overall, SP-mediated immunomodulatory activity may be attributed to its modulation of the antioxidation system.

4. Conclusion

S. imbricatus extracts enhanced the immunity of CTX-induced immunosuppressed mice by increasing immunoglobulin and immune factors in serum, enhancing organ indexes of thymus and spleen, and neutralizing oxidative stress. Further investigation is necessary to determine the applicability of SP to immune-related diseases.

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