Review Article

Liver Cirrhosis: Evaluation, Nutritional Status, and Prognosis

Hiroki Nishikawa\(^1\)\(^2\) and Yukio Osaki\(^1\)

\(^1\)Department of Gastroenterology and Hepatology, Osaka Red Cross Hospital, Osaka, Japan
\(^2\)Division of Hepatobiliary and Pancreatic Disease, Department of Internal Medicine, Hyogo College of Medicine, Hyogo, Japan

Correspondence should be addressed to Hiroki Nishikawa; nishikawa_6392@yahoo.co.jp

Received 19 May 2015; Revised 8 July 2015; Accepted 13 July 2015

Academic Editor: Ekihiro Seki

The liver is the major organ for the metabolism of three major nutrients: protein, fat, and carbohydrate. Chronic hepatitis C virus infection is the major cause of chronic liver disease. Liver cirrhosis (LC) results from different mechanisms of liver injury that lead to necroinflammation and fibrosis. LC has been seen to be not a single disease entity but one that can be graded into distinct clinical stages related to clinical outcome. Several noninvasive methods have been developed for assessing liver fibrosis and these methods have been used for predicting prognosis in patients with LC. On the other hand, subjects with LC often have protein-energy malnutrition (PEM) and poor physical activity. These conditions often result in sarcopenia, which is the loss of skeletal muscle volume and increased muscle weakness. Recent studies have demonstrated that PEM and sarcopenia are predictive factors for poorer survival in patients with LC. Based on these backgrounds, several methods for evaluating nutritional status in patients with chronic liver disease have been developed and they have been preferably used in the clinical field practice. In this review, we will summarize the current knowledge in the field of LC from the viewpoints of diagnostic method, nutritional status, and clinical outcomes.

1. Introduction

The liver is the major organ for the metabolism of three major nutrients: protein, fat, and carbohydrate [1, 2]. Chronic hepatitis C virus (HCV) infection affects about 170 million people worldwide and is the most common cause of chronic liver disease. Of these HCV-infected individuals, 20–30% eventually develop liver cirrhosis (LC) or hepatocellular carcinoma (HCC). In our country, about 30,000 persons per year die from HCC, with 70–80% of these deaths ascribed to HCV [3, 4].

LC results from different mechanisms of liver injury that lead to necroinflammation and fibrosis. Histologically, LC is characterized by diffuse nodular regeneration surrounded by dense fibrotic septa with subsequent collapse of liver structures and thus causes pronounced distortion of vascular architecture in the liver [5].

Increasingly, LC has been seen to be not a single disease entity but one that can be graded into distinct clinical stages related to prognosis [5]. In addition, the economic and social burden of LC is immense considering decreased quality of life, the disability of labor, poorer physical activity, and need for frequent hospitalizations in patients with LC.

In terms of diagnostic methods for LC, several noninvasive methods have been developed and these methods have been used for predicting prognosis in patients with LC; these include serum markers such as aspartate aminotransferase to platelet ratio index (APRI), FIB-4 index, aspartate aminotransferase (AST) to alanine aminotransferase (ALT) ratio, or modalities such as acoustic radiation force impulse (ARFI), transient elastography (TE), and magnetic resonance elastography [6–13].

On the other hand, subjects with LC often have protein-energy malnutrition (PEM) and poor physical activity. These conditions often result in sarcopenia, which is the loss of skeletal muscle volume and increased muscle weakness. Recent studies have demonstrated that PEM and sarcopenia are predictive factors for poorer survival in patients with LC [14]. Based on these backgrounds, several methods for evaluating nutritional status in patients with chronic liver disease such as indirect calorimetry, dual-energy X-ray absorptiometry (DEXA), bioimpedance analysis (BIA),
and anthropometry have been developed and they have been preferably used in the clinical settings [15].

In this review, we will summarize the current knowledge in the field of liver cirrhosis from the view of diagnostic method, nutritional status, and clinical outcomes.

2. Conventional Classification and Prognostic Assessment of Liver Cirrhosis

Currently, the most commonly used classification of liver function for patients with LC is Child-Pugh classification. This was originally designed to predict mortality during surgery in patients with LC [16]. This has been demonstrated to be useful in determining patient prognosis and thus several staging system for HCC including Japan Integrated Staging (JIS), Barcelona Clinic Liver Cancer (BCLC), and Cancer of Liver Italian program (CLIP) use this system as prognostic determinant [17–20]. The Model for End-Stage Liver Disease (MELD) score was originally developed as a prognostic model of early mortality in LC patients undergoing a transjugular intrahepatic portosystemic shunt (TIPS) [21]. This score includes variables of serum concentrations of bilirubin and creatinine and international normalized ratio for prothrombin time (INR) and, in most liver transplantation centers, MELD score has replaced the Child-Pugh score for priority of organ allocation due to superiority of prognostic ability of MELD score [21, 22]. On the other hand, serum sodium has been demonstrated to be an independent risk factor for mortality in LC patients with or without HCC [23, 24]. Kim et al. reported that the addition of the serum sodium to generate the MELD-Na score was more accurate than MELD for predicting short-term mortality on the waiting list of liver transplantation in LC patients [23]. Moreover, in their study, they estimated that the use of MELD-Na might have prevented 7% of deaths that occurred within 90 days of listing for liver transplantation [23]. In our previous study (n = 1170), we demonstrated that lower serum sodium concentration is a useful predictor in HCC patients complicating with LC [24]. Hepatic venous pressure gradient as determined by subtraction of the free-hepatic venous pressure from the wedged hepatic venous pressure has also been demonstrated to be an independent predictor in patients with LC [25]. However, unfortunately, these models did not include nutritional status of the LC patients.

On the other hand, D'Amico et al. have classified compensated LC into clinical stages 1 and 2 and decompensated LC in clinical stages 3 and 4 based on a systematic review of 118 reports [26]. They defined patients in clinical stage 1 as neither varices nor ascites and reported that the 1-year mortality was only 1%, and if patients develop varices (clinical stage 2), the 1-year mortality increases up to 3.4%. Furthermore, with decompensated LC and onset of ascites (clinical stage 3), the 1-year mortality increases up to 20%, and, following a variceal bleeding (clinical stage 4), the 1-year mortality was higher than 50% [26]. Currently, a proposal has been made to include two more additional clinical stages to this classification system: clinical stage 5, LC patients with bacterial infections (such as spontaneous bacterial peritonitis or bacteremia) as 1-year mortality increases from 49% to 66%, and clinical stage 6, patients with renal failure as mortality at 1-year could be around 70% [27, 28]. These classification systems are promising for predicting prognosis in patients with LC.

3. Noninvasive Methods for Predicting LC or LC Related Complications

Noninvasive markers of LC can be radiologic or serum based. Although liver biopsy remains the reference standard for evaluating the extent of liver fibrosis in patients with chronic liver diseases, several noninvasive methods such as TE and ARFI have been developed as alternatives to liver biopsies. Recent reports have focused on assessing the performance of noninvasive methods through long-term follow-up studies with clinical outcomes associated with LC [29–31].

Vergniol et al. reported that noninvasive tests for liver fibrosis (measurement of liver stiffness (FibroScan), FibroTest, APRI, and FIB-4 index) can predict 5-year survival of patients with chronic hepatitis C (n = 1457) [29]. Singh et al. demonstrated in their meta-analysis (n = 7058) that the degree of liver stiffness using elastography is associated with risk of decompensated cirrhosis, HCC, and death in patients with chronic liver diseases and it might be used in risk stratification [30]. A recent Japanese study demonstrated that measurements of spleen stiffness using ARFI can be used to identify patients with cirrhosis with esophageal varices (EVs) or high-risk EVs [31].

On the other hand, we previously reported that the GSA index as defined by the uptake ratio of the liver to the liver plus heart at 15 min to the uptake ratio of the heart at 15 min to that at 3 min ratio calculated from 99mTc-labeled diethylene triamine pentaacetate-galactosyl human serum albumin (99mTc-GSA) scintigraphy yielded the highest area under the receiver operating curve (AUROC) for predicting histologically proven cirrhosis with a level of 0.786 at an optimal cut-off value of 1.37 (sensitivity: 65.9%; specificity: 79.0%) in HCV-related HCC patient treated with surgical resection (SR) (n = 213) and it can be a useful predictor for HCC recurrence after surgery [32]. Furthermore, in non-B and non-C HCC patients treated with SR (n = 118), we have shown that the FIB-4 index yielded the highest AUROC for histologically proven cirrhosis with a level of 0.887 at an optimal cut-off value of 2.97 (sensitivity: 92.3%; specificity: 69.6%), and FIB-4 index >2.97 (P = 0.044) was a significant independent factor linked to HCC recurrence [33].

Recently, the Wisteria floribunda agglutinin-positive human Mac-2-binding protein (WFA+-M2BP) was demonstrated to be a liver fibrosis glycobiomarker with a unique fibrosis-related glycoalteration [34]. Yamasaki et al. reported that WFA+-M2BP can be applied as a useful surrogate marker for not only liver fibrosis but also the risk of HCC development [35].

4. Nutritional Status and Nutritional Assessment in Liver Cirrhosis

Cirrhosis, which develops over a long period of time, is frequently complicated with PEM [1, 2]. In our data, the proportion of PEM in LC patients was around 30% (unpublished
Sarcopenia is characterized by the depletion of skeletal muscle mass [47, 48]. In general, skeletal mass is maintained by a balance between synthesis and breakdown of protein [49]. LC patients have insufficient glycogen stores because of deterioration of liver function and energy generation pattern in these patients after an overnight fast is reported to be equivalent to that observed in healthy controls after 2 or 3 days of starvation [49]. These catabolic states increase the consumption of amino acids as an energy source and accelerate the breakdown in skeletal muscle to release amino acids, eventually leading to sarcopenia [49]. Recently, some studies have indicated that hyperammonemia can cause sarcopenia [50].

On the other hand, sarcopenia has become a key clinical entity for understanding the impact of aging on health outcomes. In 1989, Rosenberg first introduced the term “sarcopenia” to refer to age-related loss of skeletal muscle mass and volume [51]. Similar to bone, when persons reach around 50 years of age, they lose about 1-2% of their muscle mass per year [52]. Sarcopenia is a common disorder in aged populations contributing to functional decline, disability, and frailty [51, 52]. Several studies reported the increased risk of chronic metabolic disorders and mortality in persons with low muscle mass [53, 54]. Aging-related sarcopenia is defined as primary sarcopenia, whereas LC is a cause of secondary sarcopenia. Hiraoka et al. reported that in their analysed 988 subjects with chronic liver disease and 372 normal control subjects, presarcopenia as defined by less than two standard deviations below the mean psoas muscle area index (psoas muscle area at the mid-L3 level in CT (cm²)/height(m)² value in the controls) was observed in 15.3% of patients with chronic hepatitis, 24.4% of those with Child-Pugh A, 37.7% of those with Child-Pugh B, and 37.1% of those with Child-Pugh C and the frequency of presarcopenia was higher in chronic hepatitis regardless of age as compared with normal controls [55].

5.1. Assessment Methods for Sarcopenia. Several methods for sarcopenia assessment in patients with LC have been proposed.

5.1.1. Handgrip Strength. Hirsch et al. demonstrated in their controlled trial that handgrip strength was a useful marker for the assessment of nutritional status in LC patients [56]. Currently, The European Working Group on Sarcopenia in Older People (EWGSOP) recommends measurement of handgrip strength as a practical measure of muscle strength [57]. However, it should be kept in mind that this method has not been well established as considerable variation in the measurement methods has the potential to lead to measurement errors.

5.1.2. Imaging Studies: CT. According to the recent investigations, the psoas muscle area or thickness can be measured on the axial CT scan at the various levels of lumbar spine such as L3 vertebral level, L4 vertebral level, and at the level of umbilicus for assessing sarcopenia [58–61]. These studies showed the good correlations of clinical outcomes and the psoas muscle mass [58–61]. The psoas muscle can be easily identified and easily measured on a CT scan, as it is surrounded by retroperitoneal fat tissue and vertebra, and is not susceptible to the compression of ascites or splenomegaly.
Thus, measurement of psoas muscle area at L3 vertebral level has been preferably used for assessing sarcopenia. However, no consensus value for CT-based sarcopenia has been well established in Asian populations.

5.1.3. Bioimpedance Analysis. BIA is a noninvasive technique that measures electrical resistance and reactance [39, 62–64]. In recent studies, electrical BIA has been proposed for body composition analysis in patients with chronic liver disease [39, 62–64]. This method is based on the principle that body fat and no fat mass have specific components, such as water, proteins, and minerals [39, 62–64]. Electrical bioimpedance consists in the delivery of a low-intensity electric current which flows through the body by the ions movements [39, 62–64]. Fernandes et al. reported that the assessment using BIA presented a statistically significant correlation with Child-Pugh classification [39]. On the other hand, BIA did not demonstrate the ability to distinguish between minimal and advanced degrees of hepatic fibrosis in patients with chronic HCV infection [63].

5.1.4. Dual-Energy X-Ray Absorptiometry. DEXA through a low-dose X-ray can be used to measure fat, total body bone mineral, and fat-free soft tissue mass [65, 66]. In healthy persons, excellent agreement is found between data obtained using DEXA and data obtained from the more established reference methods [66]. However, this technique is not accurate for evaluating body composition in LC patients with fluid retention.

5.1.5. Other Methods. Skin-fold thickness measurement using a caliper is the method that quantify fat mass in the upper arm (midarm muscle area) [67]. However, there have been conflicting reports for the accuracy for predicting malnutrition in LC patients because of its interobserver variability, and this method did not correlate with Child-Pugh classification [66, 67].

6. Sarcopenic Obesity

The current global obesity epidemic has created a new condition: the combination of obesity and sarcopenia, described as sarcopenic obesity [68]. As LC patients occasionally have sarcopenia (around 40%) and obesity (around 30%), it can be deduced that a considerable number of LC patients may have sarcopenic obesity [69]. In addition, obesity is often accompanied by nonalcoholic fatty liver disease (NAFLD), and the prevalence of this liver disease is increasing in industrialized countries. NAFLD can progress to nonalcoholic steatohepatitis and LC [70]. The increase in obesity prevalence rates in elderly patients is also of concern, given the associated disease risks such as coronary heart disease and more limited treatment options available in this age group. Sarcopenic obesity has been also found to be related to poorer survival in patients with solid tumors of the respiratory and gastrointestinal tracts [71]. Sarcopenic obesity may become a major condition in LC patients in the future.

Sarcopenic obesity is assuming a significant role as a risk factor due to the double metabolic burden derived from excess adiposity (obesity) and low muscle mass (sarcopenia). Obesity also induces systemic inflammation and insulin resistance and both prompt hypercatabolism and impairs the anabolic effect of muscles, leading to protein breakdown stimulation and muscle synthesis suppression [53]. Skeletal muscle plays a significant role in insulin sensitivity as a primary tissue associated with whole body insulin-mediated glucose uptake [53]. Several studies reported that low skeletal muscle mass is linked to obesity, metabolic syndrome, and dysglycemia, and the reverse was demonstrated in large populations with higher muscle mass associated with better insulin resistance and a lower risk of developing diabetes [53, 54, 72]. Moreover, a recent study demonstrated that sarcopenic obesity is more closely linked to insulin resistance than obesity or sarcopenia alone [54]. Taken together, this new condition may lead to accelerating sarcopenia progression.

On the other hand, sarcopenic obesity is a newly recognized clinical entity following living donor liver transplantation [73]. Choudhary et al. reported that 82 patients are undergoing liver transplantation and 72 patients (88%) developed sarcopenic obesity and metabolic syndrome despite resuming routine exercise after liver transplantation [73]. In LC patients, who receive liver transplantation, appropriate nutrition and exercise after transplantation may be required.

BMI is a simple anthropometric index calculated from individual height and weight and is widely used. However, BMI is limited anthropometrically in that it does not evaluate individual components of body weight such as muscle volume or regional fat distribution. Body mass can be grossly divided into two compartments. These are fat mass and fat-free mass. In a multicompartment body composition model, fat-free mass may be partitioned into skeletal and integument and skeletal muscle and visceral organs and total body water. Total body water is further partitioned into intracellular and extracellular water [74]. Regional fat distribution plays an essential role especially in patients with metabolic syndrome [75]. Taking these into consideration, BMI may not be suitable for evaluating sarcopenic obesity.

7. Nutritional Support in LC Patients with Sarcopenia

The aims of nutritional therapy in cirrhotic patients are the support of liver regeneration, the prevention or correction of specific nutritional deficiencies, and the prevention and/or treatment of the LC related complications [76]. The recommendations in nutritional intervention target the optimal supply of adequate substrates related to requirements linked to protein, energy, lipids, carbohydrates, vitamins, and minerals. Early identification and treatment of malnutrition in LC patients have the potential to lead to better clinical outcome and prevent LC related complications [77].

7.1. Vitamin. Vitamin deficiencies such as vitamin A, B, D, and E in LC patients are in general associated with disorders of liver function and diminished reserves and with increasing severity of the disease. They are related to inadequate dietary
intake and/or malabsorption. Fat soluble vitamin deficiencies are common manifestations in LC patients [78]. Thus, vitamin supplementation may be essential for advanced LC patients [78].

7.2. Minerals. Zinc is an essential trace element required for normal cell growth, development, and differentiation and zinc deficiency is common in LC patients [79]. Zinc supplementation is demonstrated to reverse clinical signs of zinc deficiency in LC patients [79, 80]. Furthermore, zinc supplementation produced metabolic effects and trends toward improvements in liver functional reserve, hepatic encephalopathy, and general nutritional status [80, 81].

7.3. BCAA. In LC patients, the plasma level of branched-chain amino acid (BCAA) is positively correlated with the serum albumin level. Such a correlation is seen only in patients with chronic liver diseases such as cirrhosis [1, 2, 76]. The albumin-BCAA correlation and the inability of cirrhotic patients to maintain an adequate plasma level of BCAA with diet alone serve as the theoretical rationale for the use of BCAA granules for the treatment of cirrhosis. In cirrhotic patients, BCAA uptake in skeletal muscle is increased for ammonia detoxification and energy production and, in turn, the plasma level of BCAA and albumin production decrease [1, 2, 76]. BCAA granules (LIVACT, Ajinomoto Pharma, Tokyo, Japan) contain L-valine, L-leucine, and L-isoleucine at a ratio of 1:2:2:1. L-leucine induces albumin synthesis in hepatic cells via transcription factors such as mammalian target of rapamycin (mTOR) [1, 76, 82–88]. BCAA granules were originally developed for the treatment of hypoalbuminemia associated with decompensated cirrhosis. However, later studies found a variety of other pharmacological actions of this drug. BCAA granules therapy not only improves hypoalbuminemia but also inhibits cirrhosis-related complications such as esophageal varices and ascites, reduces insulin resistance and oxidative stress, improves fatty acid metabolism, stimulates the immune system, and inhibits angiogenesis [1, 76, 82–88]. The 2010 guidelines for comprehensive treatment of hepatitis virus-related cirrhosis in Japanese patients recommend the use of BCAA granules to preserve liver function and inhibit hepatic carcinogenesis [89]. Furthermore, Hanai et al. recently reported that BCAA supplementation improved the survival of sarcopenic LC patients in their subgroup analysis \( (P < 0.01) \) [36]. Conversely, the American Society for Parenteral and Enteral Nutrition (ASPEN) and the European Society for Clinical Nutrition and Metabolism recommend that BCAA supplementation be carried out only in cirrhotic patients with chronic hepatic encephalopathy that is refractory to pharmacotherapy [90, 91]. There may be differences of indications for BCAA therapy in LC patients between Japan and Western countries.

7.4. Carnitine. Carnitine deficiency has been demonstrated to be linked to LC [92]. Administration of L-carnitine, which is a derivative with high bioactivity in carnitine derivatives, has been suggested as a safe alternative treatment for LC patients [93, 94]. In field clinical practice, Malaguarnera et al. reported that LC patients treated with L-carnitine showed greater reductions in serum ammonia levels and improvements of neuropsychological functioning in comparison with placebo [93]. On the other hand, Nakanishi et al. demonstrated that L-carnitine reduces muscle cramps in LC patients [94]. However, whether L-carnitine improves sarcopenia in LC patients remains unclear. Further examination will be needed to confirm these results.

8. Sarcopenia, Patient Performance Status, and HCC Prognosis

Severe muscle wasting or sarcopenia is one of the most common and frequently hidden complications in HCC patients with LC, which negatively has an effect on survival and quality of life. These complications may potentially lead to deterioration of performance status (PS). The PS scale measures how the daily living ability is affected by the underlying disease. The PS scale recommended by the Eastern Cooperative Oncology Group (ECOG) is widely used by clinicians to assess the functional status in patients with various cancers [95]. It also serves as an indicator of cancer therapy and predictor of patient survival. The PS scale is a major survival determinant in patients with HCC and is specifically included in the BCLC staging system as an essential parameter for treatment guidance for HCC [96]. In our previous study of PS on survival in HCC patients \( (n = 1003) \), a worse PS was significantly associated with age, gender, Child-Pugh classification, HCC stage, JIS score, initial treatment option for HCC, maximum tumor size, alanine aminotransferase value, hypoalbuminemia, hyperbilirubinemia, renal insufficiency, hyponatremia, and prothrombin and poorer PS was an independent predictor linked to OS with a hazard ratio of 1.773 \( (P < 0.001) \). Thus, we concluded that PS was closely associated with status of HCC patients with LC and could be an important predictor for these populations [20]. In our recent another study, we proposed PS combined JIS system in HCC patients with LC and demonstrated that it can be a useful prognostic system for HCC patients complicating with LC as compared with other classification systems such as original JIS system, BCLC, and CLIP \( (n = 1170) \) [97].

8.1. HCC and Impact of Sarcopenia. Fujiwara et al. retrospectively investigated the effect of body composition components on survival in HCC patients \( (n = 1257) \) and demonstrated that sarcopenia, intramuscular fat (IMF) deposition, and high VSR (called visceral adiposity) were significantly associated with mortality, independent of HCC stage or Child-Pugh classification and their multivariate analysis revealed that sarcopenia (hazard ratio (HR), 1.52; 95% confidence interval (CI), 1.18–1.96; \( P = 0.001 \)), IMF deposition (HR, 1.34; 95% CI, 1.05–1.71; \( P = 0.020 \)), and visceral adiposity (HR, 1.35; 95% CI, 1.09–1.66; \( P = 0.005 \)) but not BMI were significant predictive factors linked to survival [98].
8.2. Prognosis in Sarcopenic HCC Patients according to Treatment Modality for HCC

8.2.1. Surgical Resection. The quality of skeletal muscle has attracted much attention as a novel indicator of sarcopenic HCC patients. Recently, Hamaguchi et al. demonstrated in their large study (447 HCC patients) that preoperative quality of skeletal muscle as evaluated by intramuscular adipose tissue content using preoperative CT imaging was well linked to postoperative mortality and HCC recurrence [99].

In addition, a recent study demonstrated that sarcopenia, as assessed by total psoas major volume, was an independent factor predictive of postoperative complications for primary hepatic cancers (HR: 3.06) [100]. Another recent study revealed that, in 109 HCC patients undergoing hepatectomy, sarcopenic HCC patients (n = 59) had significantly shorter median overall survival than nonsarcopenic HCC patients (52.3 months versus 70.3 months; \( P = 0.015 \)) and, in their multivariate analysis, sarcopenia was revealed to be an independent predictor of poorer overall survival (HR = 3.19; \( P = 0.013 \)) and disease-free survival (HR = 2.60; \( P = 0.001 \)) [101]. Otsuji et al. reported that preoperative sarcopenia increased the morbidity rate including the rate of developing liver failure in patients treated with major hepatectomy with extrahepatic bile duct resection (n = 256) [102].

8.2.2. Transcatheter Arterial Therapies. Dodson et al. demonstrated that sarcopenia was an independent predictor of mortality following transcatheter intra-arterial therapy with sarcopenic patients having a twofold increased risk of mortality in patients with liver malignancies (n = 216) [103].

8.2.3. Molecular Targeted Therapy. Recently, sarcopenia, regardless of the presence of weight loss, has been identified as an independent adverse predictor for systemic chemotherapy toxicity. Mir et al. reported that in advanced HCC patients with Child-Pugh A (n = 40), sarcopenia predicts the occurrence of dose limiting toxicities within the first month of sorafenib therapy [104].

9. Conclusion

Several noninvasive methods for evaluating the degree of liver fibrosis and nutritional status have been developed and these methods have been used for predicting prognosis in patients with LC. LC patients often have PEM and poor physical activity. These conditions often result in sarcopenia, affecting negatively the survival. Sarcopenic obesity, which is recently recognized as novel clinical entity, may lead to accelerating sarcopenia progression. Thus, adequate nutritional support and exercise management may be essential for such patients. In HCC patients complicating LC, sarcopenia is also a significant problem due to its prognostic impact (Figure 1).

Conflict of Interests

The authors have no conflict of interests to declare.

References

[1] H. Moriwaki, Y. Miwa, M. Tajika, M. Kato, H. Fukushima, and M. Shiraki, " Branched-chain amino acids as a protein- and energy-source in liver cirrhosis," Biochemical and Biophysical Research Communications, vol. 313, no. 2, pp. 405–409, 2004.
[2] M. R. Charlton, “Branched-chain amino acid enriched supplements as therapy for liver disease,” Journal of Nutrition, vol. 136, no. 1, supplement, pp. 2955–2985, 2006.
[3] Y. Imai, S. Tamura, H. Tanaka et al., “Reduced risk of hepatocellular carcinoma after interferon therapy in aged patients with chronic hepatitis C is limited to sustained virological responders,” Journal of Viral Hepatitis, vol. 17, no. 3, pp. 185–191, 2010.
[4] Y. Arase, K. Ikeda, F. Suzuki et al., “Long-term outcome after interferon therapy in elderly patients with chronic hepatitis C,” Interferon, vol. 50, no. 1, pp. 16–23, 2006.
[5] D. Schuppan and N. H. Afdhal, “Liver cirrhosis,” The Lancet, vol. 371, no. 9615, pp. 838–851, 2008.
[6] L. Castera, “Invasive and non-invasive methods for the assessment of fibrosis and disease progression in chronic liver disease,” Best Practice and Research: Clinical Gastroenterology, vol. 25, no. 2, pp. 291–303, 2011.
[7] L. Chrostek and A. Panasiuk, “Liver fibrosis markers in alcoholic liver disease,” World Journal of Gastroenterology, vol. 20, no. 25, pp. 8018–8023, 2014.
[8] Y. Sumida, A. Nakajima, and Y. Itoh, “Limitations of liver biopsy and non-invasive diagnostic tests for the diagnosis of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis,” World Journal of Gastroenterology, vol. 20, no. 2, pp. 475–485, 2014.
[9] J. O. Smith and R. K. Sterling, “Systematic review: non-invasive methods of fibrosis analysis in chronic hepatitis C,” Alimentary Pharmacology and Therapeutics, vol. 30, no. 6, pp. 557–576, 2009.
[10] M. D’Onofrio, S. Crosara, R. de Robertis et al., “Acoustic radiation force impulse of the liver,” World Journal of Gastroenterology, vol. 19, no. 30, pp. 4841–4849, 2013.
[11] Y. K. Mariappan, K. J. Glaser, and R. L. Ehman, “Magnetic resonance elastography: a review,” Clinical Anatomy, vol. 23, no. 5, pp. 497–511, 2010.
[12] M.-L. Yu, S.-M. Lin, C.-M. Lee et al., “A simple noninvasive index for predicting long-term outcome of chronic hepatitis C after interferon-based therapy,” Hepatology, vol. 44, no. 5, pp. 1086–1097, 2006.
H. Nishikawa, R. Kita, T. Kimura et al., “Clinical implication of...

J. Vergniol, J. Foucher, E. Terrebonne et al., “Noninvasive tests for fibrosis and liver stiffness predict 5-year outcomes of patients with chronic hepatitis C,” Gastroenterology, vol. 140, no. 7, pp. 1970–1979, 2011.

S. Singh, L. L. Fujii, M. H. Murad et al., “Liver stiffness is associated with risk of decompensation, liver cancer, and death in patients with chronic liver diseases: a systematic review and meta-analysis,” Clinical Gastroenterology and Hepatology, vol. II, no. 12, pp. 1573–1584, 2013.

Y. Takuma, K. Nouso, Y. Morimoto et al., “Measurement of spleen stiffness by acoustic radiation force impulse imaging identifies cirrhotic patients with esophageal varices,” Gastroenterology, vol. 144, no. 1, pp. 92–101, 2013.

H. Nishikawa, Y. Osaki, H. Komekado et al., “Clinical implication of the preoperative GSA index in 99mTc-GSA scintigraphy in hepatitis C virus-related hepatocellular carcinoma,” Oncology Reports, vol. 33, no. 3, pp. 1071–1078, 2015.

H. Nishikawa, Y. Osaki, H. Komekado et al., “Clinical significance of the FIB-4 index for non-B non-C hepatocellular carcinoma treated with surgical resection,” Oncology Reports, vol. 33, no. 1, pp. 88–94, 2015.

T. Toshima, K. Shirabe, T. Ikegami et al., “A novel serum marker, glycosylated Wisteria floribunda agglutinin-positive Mac-2 binding protein (WFA'-MBBP), for assessing liver fibrosis,” Journal of Gastroenterology, vol. 50, no. 1, pp. 76–84, 2015.

K. Yamasaki, M. Tateyama, S. Abiru et al., “Elevated serum levels of Wisteria floribunda agglutinin-positive human Mac-2 binding protein predict the development of hepatocellular carcinoma in hepatitis C patients,” Hepatology, vol. 60, no. 5, pp. 1563–1570, 2014.

T. Hanai, M. Shiraki, K. Nishimura et al., “Sarcopenia impairs prognosis of patients with liver cirrhosis,” Nutrition, vol. 31, no. 1, pp. 193–199, 2015.

T. M. Johnson, E. B. Overgard, A. E. Cohen, and J. K. Dibaise, “Nutrition assessment and management in advanced liver disease,” Nutrition in Clinical Practice, vol. 28, no. 1, pp. 15–29, 2013.

J. Hasse, S. Strong, M. A. Gorman, and G. Liepa, “Subjective global assessment: alternative nutrition-assessment technique for liver-transplant candidates,” Nutrition, vol. 9, no. 4, pp. 339–343, 1993.

S. A. Fernandes, L. Bassani, F. F. Nunes, M. E. D. Aydos, A. V. Alves, and C. A. Marroni, “Nutritional assessment in patients with cirrhosis,” Arquivos de Gastroenterologia, vol. 49, no. 1, pp. 19–27, 2012.

J. Ignacio de Ulibarri, A. González-Madroño, N. G. P. de Villar et al., “CONUT: a tool for controlling nutritional status. First validation in a hospital population,” Nutrition Hospitalaria, vol. 20, no. 1, pp. 38–45, 2005.

A. González-Madroño, A. Mancha, F. J. Rodriguez, J. Culebras, and J. I. de Ulibarri, “Confirming the validity of the CONUT system for early detection and monitoring of clinical undernutrition; comparison with two logistic regression models developed using SGA as the gold standard,” Nutrición Hospitalaria, vol. 27, no. 2, pp. 564–571, 2012.

W.-T. Chang, C.-G. Ker, H.-C. Hung et al., “Albumin and prealbumin may predict retinol status in patients with liver cirrhosis,” Hepato-Gastroenterology, vol. 55, no. 86–87, pp. 1681–1685, 2008.

C. Roongpisuthipong, A. Sobhonsilsuk, K. Nantiruj, and S. Songchitsomboon, “Nutritional assessment in various stages of liver cirrhosis,” Nutrition, vol. 17, no. 9, pp. 761–765, 2001.

F. Gunsar, M. L. Raimondi, S. Jones et al., “Nutritional status and prognosis in cirrhotic patients,” Alimentary Pharmacology and Therapeutics, vol. 24, no. 4, pp. 563–572, 2006.
A. Abad-Lacruz, E. Cabr´e, F. Gonz´alez-Huix et al., “Routine tests of renal function, alcoholism, and nutrition improve the prognostic accuracy of Child-Pugh score in nonbleeding advanced cirrhotics,” American Journal of Gastroenterology, vol. 88, no. 3, pp. 382–387, 1993.

N. Panagaria, K. Varma, S. Nijhawan, A. Mathur, and R. R. Rai, “Comparison of nutritional status between patients with alcoholic and non-alcoholic liver cirrhosis,” Tropical Gastroenterology, vol. 27, no. 2, pp. 75–79, 2006.

P. Tandon, M. Ney, I. Irwin et al., “Severe muscle depletion in patients on the liver transplant wait list: its prevalence and independent prognostic value,” Liver Transplantation, vol. 18, no. 10, pp. 1209–1216, 2012.

A. J. Montano-Loza, J. Meza-Junco, C. M. M. Prado et al., “Muscle wasting is associated with mortality in patients with cirrhosis,” Clinical Gastroenterology and Hepatology, vol. 10, no. 2, pp. 166–173, 2012.

S. Dasarathy, “Consilience in sarcopenia of cirrhosis,” Journal of Cachexia, Sarcopenia and Muscle, vol. 3, no. 4, pp. 225–237, 2012.

M. J. Englesbe, S. P. Patel, K. He et al., “Sarcopenia and mortality after liver transplantation,” Journal of the American College of Surgeons, vol. 211, no. 2, pp. 271–278, 2010.

I. H. Rosenberg, “Sarcopenia: origins and clinical relevance,” Journal of Nutrition, vol. 127, supplement 5, pp. 9905–9915, 1997.

C. Wang and L. Bai, “Sarcopenia in the elderly: basic and clinical issues,” Geriatrics and Gerontology International, vol. 12, no. 3, pp. 388–396, 2012.

S. Hirsch, D. Bunout, P. de la Maza et al., “Bioelectrical impedance analysis for the evaluation of hepatic fibrosis in patients with chronic hepatitis C infection,” Digestive Diseases and Sciences, vol. 53, no. 7, pp. 1957–1960, 2008.

F. A. F. Figueiredo, R. M. Perez, M. M. Freitas, and M. Kondo, “Comparison of three methods of nutritional assessment in liver cirrhosis: subjective global assessment, traditional nutritional parameters, and body composition analysis,” Journal of Gastroenterology, vol. 41, no. 5, pp. 476–482, 2006.

P. Fiore, M. Merli, A. Andreoli et al., “A comparison of skinfold anthropometry and dual-energy X-ray absorptiometry for the evaluation of body fat in cirrhotic patients,” Clinical Nutrition, vol. 18, no. 6, pp. 349–351, 1999.

M. Zamboni, G. Mazzali, F. Fantin, A. Rossi, and V. Di Francesco, “Sarcopenic obesity: a new category of obesity in the elderly,” Nutrition, Metabolism and Cardiovascular Diseases, vol. 18, no. 5, pp. 388–395, 2008.

M. Shiraki, S. Nishiguchi, M. Saito et al., “Nutritional status and quality of life in current patients with liver cirrhosis as assessed in 2007–2011,” Hepatology Research, vol. 43, no. 2, pp. 106–112, 2013.

H. Nishikawa and Y. Osaki, “Non-B, non-C hepatocellular carcinoma (review),” International Journal of Oncology, vol. 43, no. 5, pp. 1333–1342, 2013.

C. M. M. Prado, J. C. K. Wells, S. R. Smith, B. C. M. Stephan, and M. Sierra, “Sarcopenic obesity: a Critical appraisal of the current evidence,” Clinical Nutrition, vol. 31, no. 5, pp. 583–601, 2012.

S. Misra and A. S. Kalamangla, “Relative muscle mass is inversely associated with insulin resistance and prediabetes: findings from the third National Health and Nutrition Examination Survey,” The Journal of Clinical Endocrinology & Metabolism, vol. 96, pp. 2898–2903, 2011.

G. M. Chertow, E. G. Lowrie, D. W. Wilmore et al., “Nutritional assessment with bioelectrical impedance analysis in maintenance hemodialysis patients,” Journal of the American Society of Nephrology, vol. 6, no. 1, pp. 75–81, 1995.

S. M. Grundy, H. B. Brewer Jr., J. I. Cleeman, S. C. Smith Jr., and C. Lentfant, “Definition of metabolic syndrome: report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition,” Arteriosclerosis, Thrombosis, and Vascular Biology, vol. 24, pp. e13–e18, 2004.

H. Nishikawa and Y. Osaki, “Clinical significance of therapy using branched-chain amino acid granules in patients with liver cirrhosis: subjective global assessment, traditional nutritional parameters, and body composition analysis,” Journal of Gastroenterology, vol. 41, no. 5, pp. 476–482, 2006.

K. Norman, M. Pirlich, J. Sorensen et al., “Bioimpedance vector analysis as a measure of muscle function,” Clinical Nutrition, vol. 28, no. 1, pp. 78–82, 2009.

F. Antaki, M. M. French, D. K. Moonka, and S. C. Gordon, “Bioelectrical impedance analysis for the evaluation of hepatic fibrosis in patients with chronic hepatitis C infection,” Digestive Diseases and Sciences, vol. 53, no. 7, pp. 1957–1960, 2008.

F. A. F. Figueiredo, R. M. Perez, M. M. Freitas, and M. Kondo, “Comparison of three methods of nutritional assessment in liver cirrhosis: subjective global assessment, traditional nutritional parameters, and body composition analysis,” Journal of Gastroenterology, vol. 41, no. 5, pp. 476–482, 2006.
cirrhosis and hepatocellular carcinoma,” *Hepatology Research*, vol. 44, no. 2, pp. 149–158, 2014.

[77] C. Bémeur and R. F. Butterworth, “Nutrition in the management of cirrhosis and its neurological complications,” *Journal of Clinical and Experimental Hepatology*, vol. 4, no. 2, pp. 141–150, 2014.

[78] H. Andersen, M. Borre, J. Jakobsen, P. H. Andersen, and H. Vilstrup, “Decreased muscle strength in patients with alcoholic liver cirrhosis in relation to nutritional status, alcohol abstinence, liver function, and neuropathy,” *Hepatology*, vol. 27, no. 5, pp. 1200–1206, 1998.

[79] M. Hayashi, K. Ikezawa, A. Ono et al., “Evaluation of the effects of combination therapy with branched-chain amino acid and zinc supplements on nitrogen metabolism in liver cirrhosis,” *Hepatology Research*, vol. 37, no. 8, pp. 615–619, 2007.

[80] K. Katayama, M. Saito, T. Kawaguchi et al., “Effect of zinc on liver cirrhosis with hyperammonemia: a preliminary randomized, placebo-controlled double-blind trial,” *Nutrition*, vol. 30, no. 11-12, pp. 1409–1414, 2014.

[81] M. H. Somi, P. Rezaeifar, A. O. Rahimi, and B. Moshere, “Effects of low dose zinc supplementation on biochemical markers in non-alcoholic cirrhosis: a randomized clinical trial,” *Archives of Iranian Medicine*, vol. 15, no. 8, pp. 472–476, 2012.

[82] T. Kawaguchi, N. Izumi, M. R. Charlton, and M. Sata, “ Branched-chain amino acids as pharmacological nutrients in chronic liver disease,” *Hepatology*, vol. 54, no. 3, pp. 1063–1070, 2011.

[83] Y. Muto, S. Sato, A. Watanabe et al., “Effects of oral branched-chain amino acid granules on event-free survival in patients with liver cirrhosis,” *Clinical Gastroenterology and Hepatology*, vol. 3, no. 7, pp. 705–713, 2005.

[84] H. Moriwaki, M. Shiraki, H. Fukushima et al., “Long-term outcome of branched-chain amino acid treatment in patients with liver cirrhosis,” *Hepatology Research*, vol. 38, no. 1, pp. S102–S106, 2008.

[85] S. Hayashi, H. Chung, M. Kudo et al., “Oral branched-chain amino acid granules reduce the incidence of Hepatocellular carcinoma and improve event-free survival in patients with liver cirrhosis,” *Digestive Diseases*, vol. 29, no. 3, pp. 326–332, 2011.

[86] H. Yoshiji, N. Noguchi et al., “Attenuation of insulin-resistance-based hepatocarcinogenesis and angiogenesis by combined treatment with branched-chain amino acids and angiotensin-converting enzyme inhibitor in obese diabetic rats,” *Journal of Gastroenterology*, vol. 45, no. 4, pp. 443–450, 2010.

[87] H. Yoshiji, N. Noguchi, M. Kitade et al., “Branched-chain amino acids suppress insulin-resistance-based hepatocarcinogenesis in obese diabetic rats,” *Journal of Gastroenterology*, vol. 44, no. 5, pp. 483–491, 2009.

[88] T. Ohno, Y. Tanaka, F. Sugawaka et al., “Suppressive effect of oral administration of branched-chain amino acid granules on oxidative stress and inflammation in HCV-positive patients with liver cirrhosis,” *Hepatology Research*, vol. 38, no. 7, pp. 683–688, 2008.

[89] H. Kumada, T. Okanoue, M. Onji et al., “Guidelines for the treatment of chronic hepatitis and cirrhosis due to hepatitis C virus infection for the fiscal year 2008 in Japan,” *Hepatology Research*, vol. 40, no. 1, pp. 8–13, 2010.

[90] ASPEN Board of Directors and the Clinical Guidelines Task Force, “Guidelines for the use of parenteral and enteral nutrition in adult and pediatric patients,” *Journal of Parenteral and Enteral Nutrition*, vol. 26, no. 1, supplement, pp. 1SA–138SA, 2002.

[91] M. Plauth, E. Cabré, O. Riggio et al., “ESPEN guidelines on enteral nutrition; liver disease,” *Clinical Nutrition*, vol. 25, no. 2, pp. 285–294, 2006.

[92] S. Krähnenbühl and J. Reichen, “Carnitine metabolism in patients with chronic liver disease,” *Hepatology*, vol. 25, no. 1, pp. 148–153, 1997.

[93] M. Malagnanera, M. P. Gargante, E. Cristaldi et al., “Acetyl-L-carnitine treatment in minimal hepatic encephalopathy,” *Digestive Diseases and Sciences*, vol. 53, no. 11, pp. 3018–3025, 2008.

[94] H. Nakanishi, M. Kurosaki, T. Tsuchiya et al., “L-carnitine reduces muscle cramps in patients with cirrhosis,” *Clinical Gastroenterology and Hepatology*, vol. 13, no. 8, pp. 1540–1543, 2015.

[95] M. M. Oken, R. H. Creech, and D. C. Tormey, “Toxicology and response criteria of the Eastern Cooperative Oncology Group,” *American Journal of Clinical Oncology: Cancer Clinical Trials*, vol. 5, no. 6, pp. 649–655, 1982.

[96] J. Bruix, M. Sherman, and Practice Guidelines Committee for AASLD, “Management of hepatocellular carcinoma,” *Hepatology*, vol. 42, no. 5, pp. 1208–1236, 2005.

[97] H. Nishikawa, R. Kita, T. Kimura et al., “Proposal of performance status combined Japan integrated Staging system in hepatocellular carcinoma complicated with cirrhosis,” *International Journal of Oncology*, vol. 46, no. 6, pp. 2371–2379, 2015.

[98] N. Fujitaka, H. Nakagawa, Y. Kudo et al., “Sarcopenia, intramuscular fat deposition, and visceral adiposity independently predict the outcomes of hepatocellular carcinoma,” *Hepatology*, vol. 63, no. 1, pp. 131–140, 2015.

[99] Y. Hamaguchi, T. Kaido, S. Okumura et al., “Preoperative intramuscular adipose tissue content is a novel prognostic predictor after hepatectomy for hepatocellular carcinoma,” *Journal of Hepato-Biliary-Pancreatic Sciences*, vol. 22, no. 6, pp. 475–485, 2015.

[100] V. Valero III, N. Amini, G. Spolverato et al., “Sarcopenia adversely impacts postoperative complications following resection or transplantation in patients with primary liver tumors,” *Journal of Gastrointestinal Surgery*, vol. 19, no. 2, pp. 272–281, 2015.

[101] T. Voron, L. Tselikas, D. Pietrasz et al., “Sarcopenia impacts on short- and long-term results of hepatectomy for hepatocellular carcinoma,” *Annals of Surgery*. In press.

[102] H. Otsuji, Y. Yokoyama, T. Ebata et al., “Preoperative sarcopenia negatively impacts postoperative outcomes following major hepatectomy with extrahepatic bile duct resection,” *World Journal of Surgery*, vol. 39, no. 6, pp. 1494–1500, 2015.

[103] R. M. Dodson, A. Firoozmand, O. Hyder et al., “Impact of sarcopenia on outcomes following intra-arterial therapy of hepatocellular carcinoma,” *Annals of Surgery*, vol. 217, no. 12, pp. 2123–2132, 2013.

[104] O. Mir, R. Coriat, B. Blanchet et al., “Sarcopenia predicts early dose-limiting toxicities and pharmacokinetics of sorafenib in patients with hepatocellular carcinoma,” *PLoS ONE*, vol. 7, no. 5, Article ID e37563, 2012.