The dual role of iNOS in cancer

Frederica Vanini, Khosrow Kashfi, Niharika Nath

1 Department of Physiology, Pharmacology and Neuroscience, Sophie Davis School of Biomedical Education, City University of New York Medical School, New York, NY 10031, United States
2 Department of Life Sciences, New York Institute of Technology, NY 10023, United States

ABSTRACT

Nitric oxide (NO) is one of the 10 smallest molecules found in nature. It is a simple gaseous free radical whose predominant functions is that of a messenger through cGMP. In mammals, NO is synthesized by the enzyme nitric oxide synthase (NOS) of which there are three isoforms. Neuronal (nNOS, NOS1) and endothelial (eNOS, NOS3) are constitutive calcium-dependent forms of the enzyme that regulate neural and vascular function respectively. The third isoform (iNOS, NOS2), is calcium-independent and is inducible. In many tumors, iNOS expression is high, however, the role of iNOS during tumor development is very complex and quite perplexing, with both promoting and inhibiting actions having been described. This review will aim to summarize the dual actions of iNOS-derived NO showing that the microenvironment of the tumor is a contributing factor to these observations and ultimately to cellular outcomes.

© 2015 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).
1. Introduction

Nitric oxide (NO) is a free radical gasotransmitter that regulates various biological functions in the body. After it was identified in the 1980s as a vasoactive small molecule, the cardiovascular activities of NO were more notable mainly related to its vascular relaxation function, and to its anti-thrombotic and anti-inflammatory effects [1,2]. Besides blood flow regulation, NO involvement is recognized in other physiological functions such as neurotransmission, immune-response facilitation and in anti-pathogenic response [3–5].

The production of NO in cells under normal physiological conditions occurs by the conversion of l-arginine to l-citrulline by the enzyme nitric oxide synthase (NOS). There are three isoforms of NOS: neuronal NO synthase (nNOS, also known as NOS1), inducible NO synthase (iNOS or NOS2) and endothelial NO synthase (eNOS or NOS3) [3,6] (Fig. 1). The category called constitutive NOS (cNOS) includes both nNOS and eNOS and when activated only produce nanomolar concentrations of NO for seconds or minutes. However, iNOS the inducible isozyme generates higher amounts of NO, in the micromolar range and for longer intervals such as for hours or days [7]. Both cNOS members depend on increases in calcium ion concentrations for activity; hence, produce low amounts of NO for short durations, whereas iNOS is calcium-independent. In general, the expression levels of iNOS in tissues is also a measure of NO generated in that tissue or its surrounding environment [6].

NO was mainly viewed as an oncogenic molecule for several years. However, the biological functions of the NOS enzymes and the activities of NO have been disseminated over the years with a finer eye. This was possible partly due to pharmacological success of NO releasing compounds against heart disease and also to beneficial effects of these compounds when combined with chemotherapy against certain cancers [3,4]. Among the effects of NO in cancer, it is now evident that NO plays important roles in various stages of carcinogenesis such as DNA damage, oncogene activation, inhibition of DNA repair enzymes and tumor suppressor genes, and the modulation of apoptosis and metastasis [8–11]. Anti-tumor effects of NO produced by the immune-defense system were demonstrated to function against tumors of different human origins in animal models [12], while implications of pro-tumor effects of NO were made by association with expression of enzymes that produce NO in tumor cells in progressing tumors and metastasized tissue [13]. Over the years a dual role of NO in cancer has been acknowledged [9,14,15] and studied with more momentum to dissect the mechanisms leading to these two activities with respect to tumorigenesis [9,14].

1.1. Basic mechanism of action of NO

In order to appreciate the dual role of NO, its signaling pathway deserves some elaboration. Two major pathways are at play regarding signaling mechanisms of NO. One is cGMP-dependent (NO–sGC–cGMP pathway) and the other is cGMP-independent, which is also referred to as the NO oxidative pathway. For the NO–sGC–cGMP signaling pathway in blood vessels, l-arginine is converted by NOS to produce NO which diffuses enzymes to the lumen and in the walls. In leukocytes, NO derived from NOS may diffuse across the plasma membrane and cytoplasm. NO reacts with the active site of soluble guanylate cyclase (sGC) and produces cyclic GMP (cGMP). cGMP activates cGMP-dependent Protein Kinase G (PKG), which phosphorylates multiple substrates [16]. The other two major downstream elements that may also be activated are cGMP dependent gated ion channels, and cGMP dependent phosphodiesterase [16]. In platelets, activation of the cGMP dependent kinase phosphorylates a variety of substrates, and is involved in platelet adhesion and aggregation [17]. In general, an increase in cGMP leads to smooth muscle relaxation (Fig. 1), vasorelaxation and decrease of platelet aggregation [17,18].

The cGMP-independent pathway occurs most commonly through modification of proteins by S-nitrosylation of the cysteine residues [19–21] (Fig. 1). Such post translational modification also affects transcriptional activity by alteration of DNA binding of the protein, notably the transcription factor NF-κB which loses ability to bind DNA due to such modifications [22,23]. Hypoxia-inducible factor-1 (HIF-1) estrogen receptor and NF-κB are also redox sensitive transcription factors that are regulated by S-nitrosylation [24]. Signaling events regulated by S-nitrosylation may lead to either progression or inhibition of cancer. S-nitrosylation of NF-κB and matrix metalloproteinase 9 (MMP9) promotes cell death whereas S-nitrosylation of caspase-3, caspase-9, and c-Jun N-terminal kinase prevents activity and inhibits apoptosis [23]. Death receptors were also found to be regulated by for example, nitrosylation of death receptor DR4 and Fas promotes apoptosis via the death-inducing signal complex [25,26].

NO reacts with oxygen radicals to form peroxynitrite which is a strong oxidative and nitrosative agent, resulting in direct nitrosylation or nitrosation or nitration of signaling proteins. Nitrosative signaling in cancer cells is known to contribute to more proliferation and metastasis, and resistance to therapy and therefore, poor outcome of treatment [27]. On the other hand, apoptotic and necrotic cell death can occur by peroxynitrite via lipid peroxidation, cysteine oxidation, and also by protein nitrosylation [28,29].

2. More than a tale of two concentrations

The presence of NOS isoforms and their ability to produce different levels of NO was the basis for the concept that different

---

**Fig. 1.** Biosynthesis of NO and its mechanism of action. NO is produced by three nitric oxide synthase (NOS) isoforms neuronal, endothelial, and inducible (nNOS, eNOS, and iNOS) that catalyze the oxidation of l-arginine to l-citruline. NO activates the enzyme sGC to increase cGMP production that has downstream signaling effects. NO also has biological action by modification of protein through S-nitrosylation.
cellular activities of NO may be concentration dependent [30,31]. The physiological function of NO is dependent primarily on its concentration. At low concentrations, NO acts as a signaling molecule regulating smooth muscle relaxation and blood flow, neurotransmission, platelet activity, iron homeostasis, cell survival and proliferation whereas at high concentrations it is believed to modulate immune-mediated anti-tumor activities [32,33] (Fig. 2). Considering a cancer cell, the cellular outcome is based on additional factors for response such as the cell or tumor type, duration of exposure, NO flux, and immune and vascular cells. Therefore, the update in NO and cancer biology is that the overall effect of NO is an interplay of its activities emanating from its amount from the tumor and from its microenvironment.

A low concentration of NO of less than 100 nM prevents certain cell types from apoptosis, and thereby favor tumorigenesis and tumor and from its microenvironment.

Fig. 2. NO modulates various cellular activities by altering multiple pathways. Abbrev: PI3K-AKT: Phosphatidylinositol-3-Kinase and Protein Kinase B; MAPK: Mitogen-activated protein kinase; ERK: Extracellular signal-regulated kinase; JNK: c-Jun N-terminal protein kinase; VEGF: Vascular endothelial growth factor; HIF: Hypoxia inducible factor; COX2: Cytochrome Oxidase subunit 2; ILα: Interleukin 8; PGE2: Prostaglandin E2; cyt c: Cytochrome c; Bcl-2: B-cell lymphoma-extra large XIAP: X-linked inhibitor of apoptosis protein; MMP: Matrix metalloproteinase; cGMP: Cyclic guanosine monophosphate; NO: nitric oxide.

threshold amounts. Some exogenous donors have provided accurate measurements of threshold NO exposure and distinct effects on cells. Using spermineNONOate low NO doses of 10–300 nM were obtained in MCF-7 breast cancer cells [46] which induced proliferative events such as ERK phosphorylation and HIF-1α (hypoxia induced factor 1α) accumulation resulting in tumor proliferation and differentiation [46]. Further p53 phosphorylation occurs at high doses of NO (above 300 nM), generally known to be associated with apoptosis induction. Therefore, in addition to high and low doses, different threshold levels of NO are required for activation or stabilization of key proteins involved in carcinogenesis.

Reactive oxygen species (ROS) also influence threshold NO levels within the tumor microenvironment. ROS may modulate signaling by NO as was observed in both tumor and endothelial cells [39]. Another complication is the development of NO resistant cells through low yet continuous exposure to NO. Such tumor cells show cellular adaptation and enhanced survival by altered threshold levels [47]. NO may also form reactive nitrogen species (RNS) through interaction with the superoxide radical and may cause DNA damage and genotoxicity [48]. In an inflammatory process, NO and RNS may therefore stimulate carcinogenesis.

Based on in vitro NO release by donors and their in vivo representation of NO/RNS levels at the sites of tumors, the concentration ranges of NO by NO-donors have also been proposed to be divided into the following two ranges: Moderate concentration range (100–350 μmol/L) and high concentration range (500–1000 μmol/L) [49]. The normal NO levels are generally below the 50 nmol/L level. The cellular responses of the moderate concentration range includes increases in cell proliferation and genomic instability, and decrease in apoptosis and DNA repair [50,51]. The outcomes for the high concentration range include decreases in cell proliferation and increase in apoptosis, DNA damage and stimulation of DNA-damage signaling pathways leading to ATM/ATR-dependent p53 phosphorylation [52] (Fig. 2).
3. Background iNOS and NO

Since presence of three different NOS enzymes in cells are a reflection of levels of NO produced, reports have demonstrated that the relationship of NO to the dynamics of tumor growth is linked to certain determining features. These are; the expression patterns of NOS isoforms in various types of cells, the amount of NO produced by each isoform in the cell type including the duration, and the chemical interactions of NOS-derived NO in the cell [53]. Among the three NOS enzymes in mammalian systems, iNOS stands apart as it generates more NO than the constitutive forms of NOS (eNOS and nNOS) [7,54], is expressed after cytokine exposure, and more specifically modulates important tumor related processes such as malignant transformation, angiogenesis, and metastasis [7,54]. iNOS-derived NO and its tumorigenic or tumoricidal activities are dual roles of considerable interest that are debated in cancer biology [34,55]. Anti-tumor effects of iNOS have been examined by many researchers particularly those involved in the volume of tumor promoting reports of NO. The current understanding is that during the phases of tumor growth, the epithelial cells of the growing tumor have tumorigenic properties via iNOS activity while the environmental stroma community, for example, tumor-associated macrophages provide tumoricidal activities also via iNOS, and a competition sets in and dynamics alter with time [56]. When these dynamics are elucidated to a finer detail, therapies may be tailored for the appropriate activity of NO.

3.1. iNOS gene, polymorphisms and basic regulation

In order to understand the role of iNOS in proliferative or anti-tumor kinetics, a brief background of iNOS gene and iNOS regulation is presented first. The human iNOS gene is located on chromosome 17q11.2-q12. There are several sequences with high homology to the iNOS genes in the human genome [57]. Different polymorphisms have been described in the sequence of the human iNOS promoter [58,59]. Single nucleotide polymorphism of the iNOS gene (NOS2A Ser608Leu) caused a two-fold risk increase for B- and T-cell non-Hodgkin lymphoma and also for the diffuse large B-cell and follicular lymphomas [60,61]. A penta nucleotide repeat polymorphism –CCTTT (n)- that occurs in the human iNOS promoter at position – 2.5 kb with a median number of 12 CCTTTT-repeats, has been correlated with severity of human diseases including gastric cancer [62], glaucoma [63] and urothelial carcinoma [64,65].

One advance in the understanding of gene expression regulation involves epigenetics. In particular, DNA methylation at the C-5 positions of cytosine in the CpG dinucleotide occurs as a common epigenetic mechanism leading to gene silencing. In human alveolar macrophages and monocytes CpG motifs near the transcription start site of the iNOS promoter have been shown to be methylated, which means it is an epigenetically silenced iNOS gene [66]. Histone modifications were also found to be low at the iNOS promoter region suggesting low activity of the gene [66].

Regarding non-genetic regulatory mechanisms, iNOS is primarily regulated at the expression level by inflammatory cytokines (TNF-α, IL-1β, IFN-γ), lipopolysaccharide endotoxin, hypoxia and oxidative stress [67,68] and more recently, by heat shock protein Hsp70 which may function like a cytokine [69]. (Fig. 3). Transcriptional and post-transcriptional mechanisms resulting in the induction of iNOS expression are found to vary in different cell types or species. The most important intracellular signal transduction pathways are the NF-κB and JAK-STAT pathways [70–73]. Inhibition of iNOS expression by numerous agents, such as glucocorticoids, TGF-β1 and antioxidants have been shown to result from inhibition of NF-κB and STAT-1α activation [74–77] (Fig. 3). In addition, MAPK pathway contributes to iNOS gene expression involving the activation of transcription factors such as AP1, ATF2, cAMP-responsive elements, NF-κB, and the transcription factors from the Ets family [34,72,78]. Post-transcriptional regulation of iNOS gene expression predominantly occurs via mechanisms that influence iNOS mRNA stability [59].

4. Tumor iNOS and NO

The roles of NO relating to apoptosis, cell cycle, tumor progression, angiogenesis and metastasis, are currently viewed at the host end and tumor end since NO was found to be actively associated with tumors as well as the tumor environment, for example the vasculature cells and other stromal cells. In earlier studies in cultured cells high levels of NO that were generated by iNOS transfection in tumor cell lines produced a cytostatic or growth inhibitory effects [79–81], whereas in animal models, iNOS overexpression produced pro-tumor or anti-tumor effects on tumor growth, depending on the tumor microenvironment and the tumor type [82–84]. The tumor microenvironment includes stromal cells such as cells of the immune system and vascular tissue, and NO was established to be a component for their activities [84]. Overall, the effect of NO depends on the expression level of iNOS, duration and timing of NO delivery, the microenvironment, the genetic background and the cell type.

4.1. Overexpression of iNOS and anti-tumor activity

Overexpression of iNOS and concomitant changes in cancer cell kinetics were demonstrated to be anti-cancer in potential according to several in vitro and in vivo studies. Prostate cancer cells DU145 and PC3, and colon cancer HT-29 cell produced enhanced amounts of NO upon overexpression of iNOS by gene transfer, which increased these cells' sensitivity to cisplatin-induced cell death [85] or to radiation therapy [86]. In a mouse model of thyroid cancer, iNOS overexpression inhibited tumorigenesis [87]. In animal models of fibrosarcoma, tumor progression slowed down upon expression of iNOS via gene transfer demonstrating potential of tumor growth inhibition by iNOS-derived NO. Introduction of iNOS in conjunction with cytokines IFN-γ in prostate cancer cells also inhibited growth of the cells [88]. iNOS expressing pancreatic cancer cells did not produce tumors or metastases in xenograft mouse models due to NO upregulation and apoptosis [89]. Overexpression by means of other delivery forms of iNOS also produced anti-tumor results. In xenograft models of human colon and ovarian cancers, delivery of iNOS expressing cells by microencapsulation increased apoptosis of
In contrast, reducing iNOS expression by NOS inhibitors in MDA-MB-231 and T47D estrogen receptor (ER)-negative breast cancer cells led to increase in motility and loss of adhesion implying a role of NO in inhibiting the progression of breast cancers [27].

Induction of iNOS derived NO is believed to occur in photodynamic therapy, which is a recent area for treatment of cutaneous tumors. This procedure induces iNOS as a cytoprotective response. It is envisaged that combination of NO-inhibitors and photodynamic therapy may improve its clinical use [91].

4.2. Bystander anti-tumor effect

Bystander effect refers to a non-targeted response and generally holds significance with respect to radiation therapy. However, in gene therapy, it refers to the ability of the transfected cells to transfer death signals to neighboring tumor cells [92]. iNOS gene delivery by designer biomimetic vector and iNOS nanoparticles were found to overexpress iNOS and produce cytotoxicity and cell death in ZR-75-1 breast cancer cells, coupled with killing of bystander cells [93]. Similarly, radiation-induced bystander effects and cell killing are mediated through an iNOS-derived NO [94].

4.3. The mechanisms of anti-tumor effects: cytotoxicity and apoptosis

Typical mechanisms in NO-mediated apoptosis include caspase activation, chromatin condensation, and DNA fragmentation [95,96]. At the molecular level, high concentrations of NO derived from iNOS in macrophages induce p53 phosphorylation resulting in endothelial cell growth arrest, and higher concentrations and prolonged exposure time induce cell death [37]. Prolonged production of NO has been associated with the release of cytochrome c from the mitochondria, activation of caspase, modulation of anti-apoptotic Bcl-2 proteins, and increase in p53 expression [97] (Fig. 2). Alternatively, NO may induce the expression of DNA-dependent protein kinase in DNA repair that confers protection against oxidative and nitrosative stress [98]. A recent study has reported that one of the underlying mechanisms by which NO-mediated NF-κB inhibition suppresses tumor cell resistance and metastasis is through inhibition of the downstream targets Snail and the transcription factor Yin Yang 1 (YY1) and induction of Raf-1 kinase inhibitor protein (RKIP) [99] (Fig. 2). Finally, evidence indicating that endothelial-cell derived NO mediates the elimination of disseminated tumor cells is increasing. For example, it has been shown that endothelial-cell-produced NO had a cytotoxic effect on disseminating tumor cells [100]. A bystander signaling mechanism which depends on ROS or iNOS derived NO has also been examined to produce cell killing effects [49], however it may depend on the cell situation and environment of the tumor cell.

4.4. iNOS expression and pro-tumor activity

iNOS derived NO have been associated with many tumors and in their progression into metastasis. iNOS positivity has been reported consistently in human cancers at a variety of sites, including the lung [101] prostate [102], breast [103], bladder [104], oral cavity, esophagus [105] and colon [106]. In each single case, the effects of NO have to be interpreted with regard to the microenvironment of the tumor. Increased iNOS activity has been positively correlated with the degree of malignancy in gynecological tumors [80], gastric cancer, squamous cell carcinoma, hepatocellular carcinoma, melanoma and leukemia [107], while high iNOS expression is associated with favorable prognoses in ovarian [108] and lung cancers [109]. Osteosarcoma of the jaw in patients was associated with over expression of iNOS as deduced by immunohistochemical analysis [110]. Similarly, oral carcinomas and oral hard tissue sarcoma in patients were also associated with iNOS expression. Interestingly, soft tissue oral sarcomas from this study were negative for iNOS expression [111] and iNOS expression in carcinomas was higher than in sarcomas. In women with estrogen receptor (ER)-negative breast tumors, iNOS expression is an independent marker for prognosis. iNOS positivity related with angiogenesis of tumors, accumulations of p53 mutations and EGFR activation [27]. In other clinical studies of gastric cancer, iNOS correlated with tumor progression and has potential to be of prognostic value [112]. Low levels of expression of iNOS produced tumors in the pancreas and led to liver metastasis and ascites in mice models while higher levels of iNOS expression did not lead to metastasis. When iNOS expression was inhibited, it promoted distant liver metastasis in these mice [113]. Nitric oxide is proposed to remodel the cytoskeleton and lead to metastasis of lung cancer cells with evidence of increased expression of iNOS in these cells.

4.5. Bystander pro-tumor effect and cell situation

NO has context-dependent effects which may be compiled as ‘cell situation’, and when viewed in totality, may affect neoplastic transformation. iNOS derived NO and RNS in cells lead to genomic instability and tumorigenesis in the microenvoronment through a proposed “field effect” where bystander cells are affected. This effect was used to explain BRCA1 downregulation in bystander MCF-10A breast epithelial cells cocultured with iNOS-stimulated macrophages [50]. Further, radiation induced bystander effects include a role of iNOS which appear to be either pro- or anti-tumor depending on the cell situation. Induction of iNOS derived NO is believed to occur in photodynamic therapy, which is a recent area for treatment of cutaneous tumors. This procedure induces iNOS. In animal models of photodynamic therapy, resistance to cell killing has been reported due to increased NO derived from iNOS from breast and prostate tumors [114]. In human prostate cancer cells, there was enhanced proliferation and migration of the photostressed surviving cells, and increased invasion was due to NO-mediated activation of matrix metalloproteinase-9 (MMP-9), an ECM component [114–116]. Therefore pro-survival influences affecting surviving cells after photo-killing imply that iNOS induction is a cell situation consideration, and may be a secondary response to an altered status.

4.6. Cell situation and iNOS/NO effect

Protective or cytotoxic effects depend on the cells situation. In tumor cell lines with wild type p53, DNA damage by NO and/or modification through nitrosative stresses, as in some transformed cells, induces the accumulation of p53 resulting in apoptosis. It was found that p53 binds to the promoter region of the iNOS gene to inhibit protein iNOS expression and further generation of NO [117]. On the other hand, NO also stabilizes p53 leading to apoptosis [118], for example; induction of iNOS expression in tumor cells with wild type p53 resulted in reduced tumor growth [119]. Further, when examined in a p53 negative host, a study showed that iNOS-derived NO suppresses lymphomagenesis by promoting apoptosis and decreasing tumor cell proliferation [120].

Apoptosis induction mediated by NO via iNOS induction also occurs in colorectal [121] and pancreatic cancer cells [122] by IL-2-activated killer lymphocytes upon stimulation with inflammatory cytokines IFN-γ and TNF-α. In general the process was found to require high NO output with a high concentration in the tumor microenvironment.
4.7. iNOS and cell adhesion

NO promotes migration by reduced cell adhesion, and therefore promotes invasion. This occurs through inhibition of integrin expression. In particular NO was found to specifically inhibit α2β1 integrin-mediated platelet adhesion to immobilized collagen [123]. Other molecular events leading to induction of iNOS expression and generation of NO include activation of integrin α9β1 and src tyrosine kinase activity which enhanced cell migration [124]. In addition, NO was found to modulate the matrix metalloproteinase-8 (MMP-8) or procollagenase expression and its activity, and therefore affected tumor cell invasion (Fig. 2). NO has been implicated in the activation pathways of MMP-1, -2, -3, -8, -9 [125–127]. NO inhibits the progression of breast cancers since down-regulation of NOS with NOS inhibitors in MDA-MB-231 and T47D cells was found to lead to increase in motility and loss of adhesion [128].

4.8. Pro-tumor effects of stromal iNOS/NO

Besides the tumor cells, the stromal cells of the host produce NO also. These stromal host cells in the tumor micro-environment constitute immune cells such as macrophages, fibroblasts and vascular cells, and include the basement membrane and extra-cellular matrix as well. There is a constant and dynamic interplay between the tumor cells and the stroma immune cells, which changes according to the change in conditions. In early events of tumorigenesis, macrophages generate high concentrations of NO/ RNS to initiate tumor cell apoptosis and destroy emerging transformed cells. When or as the tumor escapes the immune system and grows, macrophages are reprogrammed by the tumor micro-environment to support the tumor. Low amounts of NO/RNS are pro-angiogenic and support tumor growth and metastasis by inducing growth factors VEGF and matrix metalloproteinases. Studies in gastric cancer (GC) patients found iNOS expression was associated with distant metastasis, vascular and lymph node development. Approximately 54% of the GC tumors displayed iNOS expression associated with tumor progression and thereby has the potential to indicate poor prognosis in GC patients [129]. In another study, iNOS and COX-2 were induced in gastric cancer cells that were co-cultured with gastric stromal fibroblasts and involved interaction of the IL-6-STAT3 pathway from the fibroblasts to promote tumor growth [130], p53 status of tumors also plays an important role. Inactivated p53 in tumors also plays an important role. Inactivated p53 in tumors also plays an important role. Inactivated p53 in tumors also plays an important role. Inactivated p53 in tumors also plays an important role. Inactivated p53 in tumors also plays an important role.

Breast mammography of high density fibroblasts are proposed to produce a stress response and make fibroblasts behave like macrophages based on increased iNOS derived NO production and JNK1 stress kinase signals, leading to an inflammatory process and promoting breast cancer [132]. iNOS expression in tumor parenchyma and stroma in gastric cancer patients showed positive correlation with gastric carcinogenesis, tumor lymphangiogenesis and lymphatic metastases [133]. Treatment with a Toll-like receptor agonist imiquimod in iNOS-knockout mice that were implanted with tumor cells improved tumor suppression compared to control wild-type mice, and led to a better survival rate. Combination of imiquimod and iNOS inhibitor produced similar results [134].

4.9. Anti-tumor effects of stromal iNOS/NO

Anti-tumor activity emanating from stromal iNOS was also demonstrated over the years. An increase in iNOS and iNOS-mediated NO release in stromal macrophages or fibroblasts in the tumor environment was associated with anti-tumor activity in pancreatic cancer [135] and in LPS stimulated macrophages [136]. The interplay and dynamics of tumor-iNOS expression and host-iNOS were also considered, where in wild type and iNOS −/− mice responded differently to iNOS-negative tumor cells; the murine fibrosarcomas in the iNOS −/− mice increased at faster rate than in the wild type host [137]. iNOS were also induced by Lipid A and interleukin 10 which inhibited tumorigenesis in colon and breast cancers in rodent models [138,139]. Delivery of liposomes containing lipopeptides suppressed murine sarcoma hepatic metastasis by induction of iNOS and NO generation [140]. In human ovarian cancer models, an interesting observation was that only NO-producing macrophages exhibited anti-tumor activity. IFN-β-transfected tumor cells which were injected into the peritoneal cavity of nude mice produced no tumors compared to control cell injections, and were found to stimulate high NO levels in the murine macrophages [141]. Therefore, host iNOS or stromal iNOS may provide anti-tumor activity.

4.10. Stromal iNOS and cell situation considerations

iNOS from stromal cells may not be indicative of any one effect. A factor that complicates host versus tumor iNOS performance is NO sensitivity of the tumor cells. Different tumor types may have differential NO sensitivity. It was demonstrated that host iNOS deletion increased metastatic tumor growth of M5076 ovarian sarcomas in the lung, whereas in other examples of iNOS−/− mice, a reduced metastasis of B16-BL6 melanomas occurred [142]. A tissue specific sensitivity is hypothesized where in M5076 cells are sensitive to macrophage-induced NO, whereas B16-BL6 cells are insensitive, implying that host iNOS favors tumor progression of B16-BL6 cells. Another contributing factor affecting tumor outcome is tissue type. The pathways regulating iNOS expression are different depending on cell type [143]. In murine breast carcinoma, tumor cell-iNOS decreases the metastatic development whereas stromal cell-iNOS increased metastasis [144]. In contrast, in human cervical intraepithelial neoplasias increased iNOS in the cervical stroma was associated with poor response to IFN-alpha 2B immunotherapy [145]. Platelets are required for the vascular endothelium adhesion to tumor cells. Platelet aggregation prevents tumor cell removal by immunological attack [146]. NO from tumor cells prevent platelet aggregation. The antitumor actions include loss of malignancy due to inhibition of platelet aggregation through a cGMP-dependent mechanism. A metastatic human colorectal carcinoma cell line has lower iNOS activity but higher platelet aggregation compared with a non-metastatic tumor line that is derived from the same patient [147].

Regarding immune suppression activity, NO inhibits the production of IL-12 from macrophages and dendritic cells [148]. It was believed that NO modulates differentiation of T helper cells and therefore controls the T cell response [149–151] and later it was shown that iNOS from activated T cells selectively modulate T cell differentiation [152].

5. NO molecular pathways and signaling

A collective view of the NO related pathways from the studies reviewed here needs to be highlighted at this point. Increased production of NO may promote tumor progression and metastasis
by direct induction of tumor cells proliferation, migration and invasion and indirectly through the expression of angiogenic factors in tumor cells. First, exposure of cells to various oxidants induces MAPK, such as ERK, JNK/SAPK and p38 kinases [153,154]. A special mention of NFκB is necessary since NF-kB pathway which is a pro-survival mechanism has been found to be a central player and indeed, it is activated in over 50% of all cancers [155,156]. Promotion of metastasis by NF-kB has been reported through increased epithelial to mesenchymal transition [157,158]. In breast cancer cells, other pathways are activated, for example phosphatidylinositol 3-kinase (PI3K/Akt), c-Myc and HIF [159,160]. Others have reported NO activation in β-catenin transcriptional regulation in osteocytes [161] and colorectal cancer cells [162], and in models of colitis [163]. Regarding low level exposure to NO, tumor cells show cellular adaptation and through enhanced survival, particularly by producing IL-10, TGF-β and PGE2 [47].

5.1. Pharmacological inducers of iNOS and cell fate

Statins are cholesterol lowering agents that inhibit HMG-CoA reductase. These agents have been shown to have tumoricidal and apoptotic activity that is mediated by iNOS induction [164]. Epidemiological studies have suggested that statins have anti-cancer activity against many cancers including breast carcinoma [165]. Among the various studies, post-menopausal breast cancer patients using statins had significant benefit when individual classes of statin therapy were used, although not for all classes combined [166]. In MCF-7 breast cancer cells, two representative agents simvastatin and fluvastatin induced iNOS mRNA and protein expression, reduced proliferation and induced apoptosis in these cells. In ER-negative and metastatic triple negative cancer cells, fluvastatin produced anti-proliferative and anti-invasive effects also by increasing iNOS-derived NO [167]. In contrast pitavastatin reduced iNOS mRNA expression and suppressed intestinal polypl development in APC Min mice models of colon carcinogenesis [168]. Considering the fact that statins are used for cholesterol management for more than two decades, it is important to note that there is no data to suggest that patients with statin regimen are at a greater risk of developing any type of cancer. At the same time, iNOS overexpression or its inhibition are not sole indicators of tumor fate.

Cancer chemopreventive agents that have anti-tumor activity have been examined for iNOS induction and inhibition. Interestingly, in accordance with a dual role of iNOS, compounds that induce iNOS are cancer preventive and compounds that inhibit iNOS are also cancer preventive. Induction of iNOS-derived NO production by fenretinide, a retinamide derivative of vitamin A, is necessary for growth inhibition and promotion of apoptosis of metastatic breast cancer cells. The amount of NO related directly to the intensity of these responses. In contrast, reduced iNOS expression by liposomal fenretinide was obtained in animal models of hepatocarcinogenesis [169,170]. The well known chemotherapeutic agent 5-fluouracil (5-FU) induced iNOS derived NO and induced apoptosis in liver cancer cells. Agents that induced NO formation such as L-arginine improved the sensitivity of the cells to 5-FU [171]. Other pharmacological agents that are NO-donors and are chemopreventive agents, such as the NO-donating Non-steroidal anti-inflammatory drugs (NO-NSAIDs), for example NO-donating aspirin, have demonstrated reduction in cytokine-induced iNOS expression associated with growth inhibition of HT-29 colorectal cancer cells [172,173]. The same finding was corroborated in azoxymethane-induced colonic tumors in rats where NO donating-aspirin or ß-luprofen inhibited iNOS expression and activity, and reduced tumor incidence and multiplicity [174]. It is clear that iNOS and NO affects many types of cells and at various stages. To explain many of these phenomenon from cell and animal models and in human cancers, more studies on the interplay between tumor cells, immune cells, tumor-associated macrophages, endothelial and epithelial cells mediated by iNOS in cancer progression and metastasis are needed.

From a speculative point of view it begins to appear that iNOS is likely an adaptive response to cellular stresses and a mimic of certain cell situations. At the cost of appearing simplistic, it may be envisaged that iNOS has similar role as the heat shock protein Hsp70 which is stress inducible with similar dual roles in cancer, such as promoting tumor cells to survive and regulating antitumor immunity. iNOS may cross talk with Hsp70 [69], and HSP70 and iNOS expressions reflect a collective response affecting cell fate [175]. Perhaps iNOS may be given a similar status as Hsp70.

6. Conclusions and perspectives

During past two decades or so much has been written about the dual nature of NO, which strongly suggests a concentration-dependent relationship between NO expression and biological response. This phenomenon is well accepted in pharmacology. What has been the cornerstone of much debate is the role of iNOS in cancer. Does its expression mean that iNOS-derived NO is carcinogenic or anti-carcinogenic? The current thinking based on observations that iNOS expression is high in a number of tumors and that this correlates with poor survival, has led to the conclusion that induction of this NO isoform may somehow be related to tumor genesis or that its expression may be used as marker for not so favorable outcomes. However, we believe that such a conclusion is still not warranted as in totality, it appears that the dual role of iNOS is strongly influenced by the cell situation and is environment dependent, with either induction or inhibition of iNOS having anti-cancer potential based on tumor and cell types. We believe that any approaches to iNOS based therapy against cancer may need to be cell situation based and needs a lot more directed studies.

Grant support

This work was supported in part by NIH grant R24 DA018055. The funding agency had no role in the writing of the manuscript; and in the decision to submit the manuscript for publication.

Conflict of interest

The authors have nothing to disclose.

References

[1] L.J. Ignarro, G.M. Buga, K.S. Wood, R.E. Byrns, G. Chaudhuri, Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. Proc. Natl. Acad. Sci. USA 84 (1987) 9265–9269.
[2] L.J. Ignarro, C. Napoli, J. Loscalzo, Nitric oxide donors and cardiovascular agents modulating the bioactivity of nitric oxide: an overview, Circ. Res. 90 (2002) 21–28.
[3] D.A. Geless, T.R. Billiar, Molecular biology of nitric oxide synthases, Cancer Metastasis Rev. 17 (1988) 7–23.
[4] A.C. Quinn, A.J. Petros, P. Vallance, Nitric oxide: an endogenous gas, Br. J. Anaesth. 74 (1995) 443–451.
[5] C. Bogdan, Nitric oxide and the immune response, Nat. Immunol. 2 (2001) 907–916.
[6] W.K. Alderton, C.E. Cooper, R.G. Knowles, Nitric oxide synthases: structure, function and inhibition, Biochem. J. 357 (2001) 593–615.
[7] T. Michel, O. Peron, Nitric oxide synthases: which, where, how, and why? J. Clin. Investig. 100 (1997) 2146–2152.
[8] P.K. Lala, A. Oruccevic, Role of nitric oxide in tumor progression: lessons from experimental tumors, Cancer Metastasis Rev. 17 (1998) 91–106.
shock protein 70 for inducible nitric oxide synthase induction, Cell. Signal. 25 (2013) 1310–1317.

[70] A.K. Mankan, M.W. Lawless, S.G. Gray, D. Kelleher, R. McManus, NF-κB p50 regulation: the nuclear response, J. Cell. Mol. Med. 13 (2009) 631–643.

[71] S. Akass, A. Hoffmann-Crook via the NF-κB signalling system, Cyto- kine Growth Factor Rev. 18 (2007) 189–197.

[72] H. Kleinert, T. Wallerath, G. Fritz, I. Irgir-Biedert, F. Rodriguez-Pascual, D. A. Geller, Cytokin induction of NO synthase II in human DLD-1 cells: roles of the IKK-STAT3/1 and NF-κB-signalling pathways, Br. J. Pharc- mol. 125 (1998) 193–201.

[73] R.W. Ganster, B.S. Taylor, I. Shao, D.A. Geller, Complex regulation of human inducible nitric oxide synthase gene transcription by Stat and NF-κB p50, Proc. Natl. Acad. Sci. USA 98 (2001) 8638–8643.

[74] H. Kleinert, C. Euchenhofer, I. Ihrig-Biedert, U. Forstermann, Glucocorticoids inhibit the induction of nitric oxide synthase II by down-regulating cytokine- induced activity of transcription factor nuclear factor-κB, Mol. Phar- macol. 49 (1996) 347–356.

[75] G. Pascual, C.K. Glass, Nuclear receptors versus inflammation: mechanisms of transduction, Trends Endocrinol. Metab. 17 (2006) 321–327.

[76] N. Mukaida, M. Morita, Y. Ishikawa, N. Rice, S. Okamoto, T. Kasahara, et al., Novel mechanism of glucocorticoid-mediated gene repression, Nuclear factor-κB is target for glucocorticoid-mediated interleukin 8 gene re- pression, J. Biol. Chem. 269 (1994) 12289–12295.

[77] L. Tedeschi, M. Menegazzi, D. Margotto, H. Suzuki, U. Forstermann, H. Kleinert, Anti-inflammatory actions of St. John’s wort: inhibition of human inducible nitric oxide synthase expression by down-regulating signal transducer and activator of transcription-1alpha (STAT-1alpha) activation, J. Pharmacol. Exp. Ther. 303 (2003) 254–261.

[78] VM. Jansen-Heinig, I. Macara, B.T. Mostmann, Cooperation between oxidants and tumor necrosis factor in the activation of nuclear factor (NF-)κB p50: requirement of Ras/mitogen-activated protein kinase activity in the activation of NF-κB by oxidants, Am. J. Respir. Cell Mol. Biol. 20 (1999) 942–952.

[79] R. Lelchuk, M.W. Radomski, J.F. Martin, S. Moncada, Constitutive and inducible nitric oxide synthases in human megakaryoblastic cells, J. Pharmacol. Exp. Ther. 262 (1992) 1220–1224.

[80] L.L. Thomsen, F.G. Lawton, R.G. Knowles, J.E. Beesley, V. Riveros-Moreno, S. Moncada, Nitric oxide synthase activity in human gynecological cancer, Cancer Res. 54 (1994) 1352–1354.

[81] L.L. Thomsen, D.W. Miles, J. Harperfield, L.G. Bobrow, R.G. Knowles, S. Moncada, Nitric oxide synthase activity in human breast cancer, Br. J. Cancer 72 (1995) 41–44.

[82] K. Xie, S. Huang, Z. Dong, S.H. Jiang, M. Gutman, Q.W. Xie, et al., Transfection with the inducible nitric oxide synthase gene suppresses tumorigenesis and abrogates metastasis by K-1735 murine melanoma cells, J. Exp. Med. 181 (1995) 1333–1343.

[83] P.K. Lala, L. Chakraborty, Role of nitric oxide in carcinogenesis and tumour progression, Lancet Oncol. 2 (2001) 149–156.

[84] R. Zhang, A. Ma, S.J. Urbanis, D.M. McCafferty, Induction of inducible nitric oxide synthase: a protective mechanism in colitis-induced adenocarcinoma, Cancer Research 28 (2007) 1122–1130.

[85] R. Zhang, H.O. Martin, C.A. Coulier, J. Worthington, C. Murphy, T. Robson, et al., Nitric oxide synthase gene therapy enhances the toxicity of cisplatin in cancer cells, J. Gene Med. 11 (2009) 160–168.

[86] P. Chung, T. Cook, K. Liu, V. Vodovotz, R. Zamora, S. Finkelstein, et al., Overexpression of the hepatic acute phase protein apolipoprotein A-I in the induced cytokine:pstatinase gene in mice enhances inflammation-induced apoptosis in colorectal cancer cells via a caspase-dependent mechanism. Nitric Oxide: Biol. Chem. 8 (2003) 119–126.

[87] M.N. Soler, P. Bobe, K. Benihoud, G. Lemaire, B.A. Roos, S. Lausson, Gene therapy of rat medullary thyroid cancer by naked nitric oxide synthase II and hTERT transgene, J. Gene Med. 2 (2000) 344–352.

[88] W.L. Chen, H.J. Feng, J.S. Li, H.G. Li, Expression and pathological relevance of inducible nitric oxide synthase expression in osteosarcoma of the jaws, Int. J. Oral Maxillofac. Surg. 36 (2007) 544–549.

[89] D. Augustine, B. Sekar, S. Murali, B. Ramesh, R.N. Madhavan, S.P. Patil, et al., Expression of inducible nitric oxide synthase in carcinomas and sarcomas affecting the oral cavity, South Asian J. Cancer 4 (2015) 78–82.

[90] H.G. Li, H.M. Xu, Inducible nitric oxide synthase, nitrotyrosine and apoptosis in gastric adenocarcinoma: an in vitro and in vivo correlation with a poor survival, World J. Gastroenterol.: WJG 11 (2005) 2539–2544.

[91] B. Wang, D. Wei, V.E. Crum, E.L. Richardson, H.H. Xiong, Y. Luo, et al., A novel model system for studying the double-edged roles of nitric oxide production in cancer progression, Oncogene 22 (2003) 1771–1782.

[92] A.W. Girotti, Tumor-generated nitric oxide as an antagonist of photodynamic therapy, Photochem. Photobiol. Sci. 14 (2015) 1425–1432.

[93] J.M. Fahey, A.W. Girotti, Accelerated migration and invasion of prostate cancer cells after a photodynamic therapy-like challenge: Role of nitric oxide, Nitric Oxide: Biol. Chem. 49 (2015) 47–55.

[94] R. Bhownick, A.W. Girotti, Signaling events in apoptotic photokilling of 5-aminoavuline acid-treated tumor cells: inhibitory effects of nitric oxide, Free Radic. Biol. Med. 47 (2009) 731–740.

[95] K. Forrester, S. Ambs, S.E. Lupold, R.B. Kapust, E.A. Spiallere, W.C. Weinberg, et al., Nitric oxide-induced p53 accumulation and regulation of inducible nitric oxide synthase expression by wild-type p53, Proc. Natl. Acad. Sci. USA 93 (1996) 2442–2447.

[96] B. Brune, A. von Koenthen, K.B. Sandau, Nitric oxide and its role in apoptosis, Eur. J. Pharmacol. 351 (1998) 261–272.

[97] B.M. Choi, H.O. Pae, S.I. Jiang, Y.M. Kim, H.T. Chung, Nitric oxide as a pro- apoptotic as well as anti-apoptotic modulator, J. Biochem. Mol. Biol. 35 (2002) 116–126.

[98] W. Xu, L. Liu, G.C. Smith, I.G. Charles, Nitric oxide upregulates expression of DNA-PKcs to protect cells from DNA-damaging anti-tumour agents, Nat. Cell Biol. 2 (2000) 339–345.

[99] R. Schneider, S. Bartel, Dual role of NO donors in the reversal of tumor cell resistance and EMT: downregulation of the NF-κB/p50/Snail/YY1/RKIP cir- cuitry. Nitric Oxide: Biol. Chem. 24 (2011) 1–7.

[100] H. Qiu, F.W. Orr, D. Jensen, H.H. Wang, A.R. McIntosh, B.B. Hasinoff, et al., Arrest of B16 melanoma cells in the mouse pulmonary microcirculation in- duces endothelial nitric oxide synthase-dependent nitric oxide release that is cytotoxic to the tumor cells, Am. J. Pathol. 162 (2003) 403–412.

[101] L. Zhang, J. Liu, X. Wang, Z. Li, X. Zhang, P. Caio, et al., Upregulation of cytokinotin and tumor necrosis factor and their role in tumour progression, J. Surg. Oncol. 6853.

[102] F. Vanini et al. / Redox Biology 6 (2015) 334–343
lymphocytes in human oral squamous cell carcinoma, Head Neck 33 (2011) 1301–1308.

S. M. Niedbala, J. C. Alves-Filho, S. Y. Fukuda, S. M. Vieira, A. M. Faneite, F. Sonego, et al., Regulation of type II T helper cell function by nitric oxide during infection, Proc. Natl. Acad. Sci. USA 101 (2004) 19220–19225.

Y. Janjun, R. Zhang, G. Lu, Y. Shen, L. Peng, C. Zhu, et al., T-cell-derived inducible nitric oxide synthase switches off Th17 cell differentiation, J. Exp. Med. 210 (2013) 1447–1462.

N. Ouyang, S. K. Mahtaney, D. Chowdhury, J. Chaudhuri, A. Manna, J. Vinayagan, et al., ICBE3 induces iNOS expression by ROS-dependent JNK and ERK activation for apoptosis of leukaemic cells, Apoptosis 17 (2012) 612–626.

A. Seeger, C. Rosenberg, W. D. Schmitt, C. Denkert, S. Hauptmann, Nitric oxide of human colorectal adenocarcinoma cell lines promotes tumour cell invasion, Br. J. Cancer 86 (2002) 1310–1315.

S. M. F. Eddy, D. H. Sherr, G. E. Sonenshein, NF-kappaB and epithelial to mesenchymal transition of cancer, J. Cell Biochem. 104 (2008) 734–744.

R. L. Pruett, B. J. Boersma, T. M. Howe, J. E. Goodman, D. D. Thomas, L. Ying, et al., Induction and IGF-1 activate the Akt pathway in breast cancer, Int. J. Cancer 120 (2007) 799–805.

H. W. Lo, S. C. Hsu, M. Ali-Sayed, M. Gunduz, W. X. Xia, Y. Wei, et al., Nuclear expression of EGR and STAT3 in the activation of the iNOS/NF-κB pathway, Cancer Cell 7 (2005) 575–585.

A. Santos, A. Bakker, G. Gotz, B. Heidoblau, J. M. de Blije-Hogervorst, J. Klein-Nulend, Early activation of the beta-catennin pathway in osteoclasts is mediated by nitric oxide, phosphatidyl inositol-3 kinase/Akt, and focal adhesion kinase, Biochem. Biophys. Res. Commun. 391 (2010) 364–369.

L. Yu, G. L. Borchert, J.-Y. Yang, Polyoma enhancer activator 1, an et s transcription factor, mediates the induction of cyclooxygenase-2 by nitric oxide in colorectal cancer cells, J. Biol. Chem. 279 (2004) 18694–18700.

H. Wang, R. Zhang, S. Wen, D. M. McCafferty, P. L. Beck, W. K. Maclauran, Nitric oxide increases Wnt-induced secreted protein 1 (WISP-1/CCN1) expression and function in colitis, J. Mol. Med. 87 (2009) 435–445.

S. Kotasrani, C. L. Williams, B. Kalyanaraman, Statin-induced breast cancer cell death: role of inducible nitric oxide and arginine-dependent pathways, Cancer Res. 67 (2007) 7386–7394.

T. P. Ahern, T. L. Lash, P. Damkier, M. P. Christiansen, D. P. Cronin-Fenton, Statins and breast cancer prognosis: evidence and opportunities, Lancet Oncol. 15 (2014) e461–e468.

I. A. Cauley, A. M. McNier, R. J. Rodabough, A. LaCroix, D. C. Bauer, K. L. Margolis, et al., Statin use and breast cancer: prospective results from the Women’s Health Initiative, J. Natl. Cancer Inst. 98 (2006) 700–707.

A. K. Kanugula, P. N. Gollavilli, S. B. Vasamsetti, S. K. Aravind, R. Chopra, R. Ummanni, et al., Statin-induced inhibition of breast cancer proliferation and invasion involves the iron regulatory protein and the iron regulatory protein of the cancer, J. Cell Sci. 119 (2006) 545–555.

W. A. Siegert, C. Rosenberg, W. D. Schmitt, C. Denkert, S. Hauptmann, Nitric oxide mediates epithelial-stromal interactions and promotes breast cancer progression, J. Exp. Med. 203 (2006) 2421–2433.

G. van der Valk, I. van Oosterhout, J. W. ten Berge, E. van der Pols, L. van der Graaf, J. van Schagen, et al., Inhaled nitric oxide reduces bronchoconstriction in patients with asthma, Eur. Respir. J. 21 (2003) 715–723.

J. B. Hejja, M. A. Yang, K. L. Bao, Z. J. Zhang, F. Li, H. Wang, et al., Activation of inducible nitric oxide synthase in human pancreatic carcinoma cell lines by hypoxia and reoxygenation, Cancer Res. 57 (1997) 4606–4612.

R. Seppel, A. J. H. van den Brink, S. C. Reuten, M. G. J. J. van Tol, M. A. J. van Rhoon, et al., Induction of cyclooxygenase-2 by nitric oxide in human esophageal cancer cells, Carcinogenesis 26 (2005) 417–427.

M. A. Simeone, E. Miekecklou, L. D. Broening, E. A. Grimm, A. M. Tari, A novel mechanism by which N-(4-hydroxy/ethyl)retinamide inhibits breast cancer cell growth: the production of nitric oxide, Mol. Cancer Ther. 1 (2002) 1009–1017.

J. Jiang, J. Liu, J. Zhu, C. Yang, A. Zhang, Mechanism of apoptotic effects induced by 5-fluorouracil on human liver carcinoma Bel7402 cell line, Chin. Med. J. 115 (2002) 968–971.

A. Spiegel, T. Hrudynac, C. Jen, G. Gao, N. Ouyang, X. Liu, et al., NO-donating aspirin inhibits both the expression and catalytic activity of inducible nitric oxide synthase in HT-29 human colon cancer cells, Biochem. Pharmacol. 70 (2005) 993–1000.

W. F. Williams, N. Nath, J. Chen, T. R. Hrudynac, J. Gao, L. Kopelovich, et al., Growth inhibition of human colon cancer cells by nitric oxide (NO)-donating aspirin is associated with cyclooxygenase-2 induction and beta-catennin/T-cell factor signaling, nuclear factor-kappaB, and NO synthesis 2 inhibition: implication for tumour suppression, Cancer Prev. Res. 4 (2011) 7613–7618.

C. V. Rao, B. S. Reddy, V. E. Steele, C. X. Wang, X. Liu, et al., Inhibition of nuclear factor-kappaB and Mat1A genes in the early stages of rat liver carcinogenesis, Cancer Res. 26 (2005) 417–427.

M. G. Ostrom, J. E. Stewart, J. A. Milner, R. H. Freudenreich, J. C. O’Leary, M. D. Gimpel, et al., The expression of iNOS and nitric oxide synthase in colon cancer and colonic inflammation, Acta Histochem. 114 (2012) 827–835.

M. A. Huber, N. Azizos, B. Baumann, S. Grunert, A. Sommer, H. Pehamberger, et al., NF-kappaB is essential for epithelial-mesenchymal transition and metastasis in a model of breast cancer progression, J. Clin. Investig. 114 (2004) 569–581.

M. A. Huber, N. Azizos, B. Baumann, S. Grunert, A. Sommer, H. Pehamberger, et al., NF-kappaB is essential for epithelial-mesenchymal transition and metastasis in a model of breast cancer progression, J. Clin. Investig. 114 (2004) 569–581.