The Correction Methods of the Intestinal Microflora in Chronic Colonic Stasis by Siphon Enema and Probiotics as a Means of the Hirschsprung-associated Enterocolitis Prevention

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Abstract

AIM: The purpose of the current article is to study the state of the intestinal microflora in the context of chronic colonic stasis and assesses the methods of its correction by means of siphon enema and probiotics’ implication (bifidumbacterin and lactobacterin) to prevent the development of HAEC.

METHODS: This study was conducted in the Astana City Children’s Hospital No. 2 and involved 60 children who applied for medical help with symptoms of chronic constipation. They were admitted to the hospital in a planned or emergency manner with suspected Hirschsprung’s disease in the period from 2015 to 2017, after approval of the Ethics Committee. An X-ray contrast study of the large intestine (irrigography) was performed in all the children for a diagnostic purpose. The following parameters of the state of the large intestine were studied: The function of the ileocecal valve (Bauhin’s valve), the diameter and shape of the large intestine, the presence of pathological formations, and symptoms of intestinal obstruction. Statistical analysis was carried out using Student’s test for dependent and independent samples by means of the BioStat Software.

RESULTS: After treatment, the bacteriological examination of stool samples demonstrated an increase in the number of beneficial microorganisms and a decrease in the number of opportunistic microorganisms. The growth of the ileocecal valve (Bauhin’s valve), the diameter and shape of the large intestine, the presence of pathological formations, and symptoms of intestinal obstruction. Statistical analysis was carried out using Student’s test for dependent and independent samples by means of the BioStat Software.

CONCLUSION: The developed method of correcting the quantitative composition of the intestinal microflora is very effective and can be implemented in clinical practice.

Introduction

Hirschsprung’s disease is a congenital malformation of the colon with a lack or absence of ganglion cells in the intestinal segment, manifested mainly in symptoms of chronic colonic stasis. The reported incidence of the disease is 1:5000 among newborns [1], [2], [3]. Despite improved surgical results, complications of the disease occur with a frequency of 22.7% to 38.5% [4].

The most severe complication of Hirschsprung’s disease is Hirschsprung-associated enterocolitis (HAEC). HAEC occurs against the background of pronounced local immune perturbation of the intestinal wall and microbiocenosis disturbance, resulting in the development of pathogenic microflora and leading to the disorder of microcirculation with subsequent perforation of the intestinal wall and development of peritonitis [5], [6], [7], [8]. Complications can develop both before and after surgical treatment. The incidence of enterocolitis before surgery varies from 6% to 50% of cases, while after surgery – from 2% to 35% [9].

The lack of methods to correct these disorders leads to complications in the anastomosis zone in 10–15% of cases [10]. The mortality rate remains high – 2.6–12.0%, especially in young children – 31–35% [11], [12], [13].

The available data show that the prevention of HAEC remains a relevant issue. This work aimed at studying the state of the intestinal microflora in the context of chronic colonic stasis and assessing the methods of its correction by siphon enema and probiotics (bifidumbacterin and lactobacterin) to prevent the development of HAEC. The proposed method was not used before, so there are no scientific studies on the peculiarities of implication and the obtained results.

Materials and Methods

The prospective study was conducted in the Astana City Children’s Hospital No. 2, involving 60...
children who applied for medical help with symptoms of chronic constipation. They were admitted to the hospital in a planned or emergency manner with suspected Hirschsprung’s disease during the period from 2015 to 2017, after approval of the Ethics Committee (Extract from the No. 5 min of the Ethics Committee meeting).

The age of the children ranged from 2 months to 14 years. There were 36 boys (60%) and 24 girls (40%). The distribution of patients by age and gender is presented in Table 1.

Table 1: Distribution of children by age and gender

| Age/gender | Boys | Girls |
|------------|------|-------|
| 1–6 months | 2    | 1     |
| 6–12 months| 3    | 1     |
| 1–3 years old| 8    | 7     |
| 3–6 years old| 11   | 9     |
| 6–11 years| 10   | 6     |
| Older than 11 years| 2    | 0     |
| Total      | 36   | 24    |

For diagnostic purposes, all the children underwent an X-ray contrast study of the large intestine (irrigography). According to the results of irrigography, they were divided into the following nosological forms (Figure 1).

![Figure 1: Results of irrigography](image)

The study was conducted in two stages:

1st stage – analysis of a series of cases

To study the effect of bowel cleansing by means of a siphon enema on the quantitative composition of the intestinal microflora in chronic colonic stasis, a series of cases have been analyzed. The bacteriological examination of stool samples showed that the number of normal intestinal microflora microorganisms (bifidobacteria, lactic acid bacteria, and *E. coli* with normal enzyme activity) in patients with symptoms of chronic colonic stasis was below the minimum normal values. Before the administration of siphon enemas, such opportunistic microorganisms as hemolyzing *Escherichia coli*, *Staphylococcus aureus*, lactose-negative *E. coli*, enterococci, *Proteus mirabilis*, and Candida yeast-like fungi were detected during the bacteriological examination of stool samples. To prepare the large intestine for irrigography and surgical intervention, siphon enemas were used. The volume of fluid for enema was determined by the age of the patients. The patients had indications for siphon enemas implication like complaints about the absence or delay of stool for several days over a long period of time with a lack of effect from a cleansing enema in the anamnesis.

Cleansing enemas were given to patients under the age of 2 months. The average duration of preparation was 4.2 days.

2nd stage – nonrandomized controlled research

At this stage, based on the results of a series of cases, the quantitative composition of the intestinal microflora was corrected. Forty patients with symptoms of chronic constipation were equally divided into two groups (with implementing of the randomization of patients using a random number table).

The main group involved 20 children. In this group, to prepare for irrigography and surgical intervention, the intestinal microflora was corrected by the proposed method. Probiotics were administered orally and by an enema with direct irrigation of the colon cavity at an age dosage given in the instructions: For children up to a year – 1 pack 2–3 times a day, from a year and older – 1 pack 3–4 times a day.

The control group involved 20 children. They received probiotics only enterally.

Bifidumbacterin (containing live bifidobacteria at no <50 million CFU/g) and lactobacterin (containing at least 10 million live acidophilic lactobacilli) preparations were used as a probiotic medicine in the form of powder:

- Bifidumbacterin forte regulates the balance of the intestinal microflora. The effect comes from the high concentration of bifidobacteria occluded on absorbent carbon particles, being antagonists of a wide range of pathogenic (including *Shigella* spp., *Salmonella* spp., and *S. aureus*), and opportunistic microorganisms (including *Proteus* spp. and *Klebsiella* spp.). It is produced by JSC “Firma Vitafarma,” Russia.
- Lactobacterin (powder) includes live lactobacilli with antagonistic activity against a wide range of pathogenic and opportunistic bacteria (including staphylococci, *Proteus*, and enteropathogenic *E. coli*), normalizes the digestive activity of the gastrointestinal tract, improves metabolic processes, and promotes the restoration of natural immunity. It is produced by Microgen NPO, Russia.

The average duration of treatment was 11.3 days for the main group and 11.2 days for the control group.

The statistical processing of the results was carried out using Student’s t-test for dependent and independent samples by means of the BioStat Software.
Results

After the administration of siphon enemas at the first stage of the survey (is described in details above), the bacteriological examination of stool samples showed a marked decrease in the number of beneficial microorganisms of the intestinal microflora with a statistically significant difference (p < 0.001) (Table 2).

As can be seen from Table 2, in the period after the administration of siphon enemas, there was a decrease in the number and CFU/g concentrations of opportunistic microorganisms in the intestine (hemolyzing E. coli, S. aureus, lactose-negative E. coli, enterococci, and Candida yeast-like fungi). At the same time, there was a decrease in the number of beneficial microorganisms (bifidobacteria, lactic acid bacteria, and E. coli with normal enzyme activity).

At the second stage in the period before the correction of the quantitative composition of the intestinal microflora, the results of the bacteriological examination of stool samples in both groups corresponded to the results of the first stage before the administration of siphon enemas: A decrease in the number of microorganisms of the obligate intestinal microflora and the detection of certain species of opportunistic microorganisms.

The normalization of the number of beneficial microorganisms and the reduction in the number of opportunistic microorganisms (Table 3) was observed in patients of the main group after treatment by the developed method.

After treatment, the concentration of all kinds of beneficial microorganisms in the intestine, namely bifidobacteria, lactic acid bacteria, and E. coli with normal enzyme activity, increased to normal values with a statistically significant difference: p < 0.001, p < 0.01, and p < 0.05, respectively. A statistically significant decrease in the concentration of opportunistic enterobacteria in the intestine such as hemolytic active E. coli, lactose-negative E. coli, and P. mirabilis was revealed.

In the bacteriological examination of stool samples obtained after treatment in the control group of patients, despite a slight improvement in the ratio of beneficial microorganisms in the intestine, the number of opportunistic microorganisms (S. aureus, Klebsiella pneumoniae, Citrobacter diversus, and P. mirabilis) increased (Table 4).

Despite treatment, there was a decrease of bifidobacteria with obvious statistical significance (p < 0.001). Other types of beneficial microorganisms (lactic acid bacteria, E. coli with normal enzyme activity) increased. However, the statistical processing revealed that these parameters had no significant difference (p > 0.05) and did not reach the normal level (p < 0.001).

When analyzing the number of opportunistic enterobacteria in the control stool, one can note an increase of pathogens above the norm and concentration in the intestine with the appearance of associations between them with a statistically significant difference (p < 0.001).

A comparative analysis of the results of the bacteriological examination of the intestinal microflora in patients of the main and control groups revealed that the

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Table 2: Results of the study of the intestinal microflora before and after administration of siphon enemas

| Microorganisms                      | Norm in children (Industry standard 91500.11.0004-2003) | Number of microorganisms before administration of siphon enemas (n=20) | Number of microorganisms after administration of siphon enemas (n=20) | p       |
|-------------------------------------|----------------------------------------------------------|---------------------------------------------------------------------|---------------------------------------------------------------------|---------|
|                                     | M ± m | CFU/g |                   | M ± m | CFU/g |                   |                   |         |
| Bifidobacteria                     | 10^-10 | 2,37×10^8 ± 9,04×10^5 CFU/g | 2,23±10^8 ± 9,11×10^5 CFU/g | 0,0001 |
| Lactic acid bacteria                | 10^-10 | 5,95×10^8 ± 1,05×10^5 CFU/g | 5,45±10^8 ± 1,08×10^5 CFU/g | 0,0001 |
| E. coli with normal enzyme activity | 10^-10 | 3,25×10^8 ± 9,24×10^5 CFU/g | 2,8±10^8 ± 8,4×10^5 CFU/g | 0,01    |
| Opportunistic enterobacteria       | Norm in children (Industry standard 91500.11.0004-2003) | Number of patients with bacteria above the norm | Number of patients with bacteria above the norm |         |
|                                    | CFU/g | abs (%) | CFU/g | abs (%) |         |         |
| Hemolytic active E. coli           | 0 | 10^4 | 1 (2) | 10^4 | 1 (2) |         |         | p<0,000 |
| S. aureus                          | 0 | 10^2 | 1 (2) | 10^2 | 1 (2) |         |         | p<0,000 |
| Candida yeast-like fungi            | <10^-10 | 10^3 | 2 (10) | 10^3 | 1 (2) |         |         | p<0.05  |
| Lactose-negative E. coli           | <10^-10 | 10^4 | 1 (2) | 10^4 | 1 (2) |         |         | p<0.000 |
| Enterooccoci and P. mirabilis      | 0 | 10^4 | 1 (2) | 10^4 | 1 (2) |         |         | p<0.001 |

1. # – degree of reliability in relation to the norm
2. ## – degree of reliability at p<0.01; ### – degree of reliability at p<0.001; """" – degree of reliability at p<0.001 between the value groups in CFU/g, abs – absolute value

Table 3: Results of the study of the intestinal microflora before and after treatment of patients in the main group

| Microorganisms                      | Norm in children (Industry standard 91500.11.0004-2003) | Number of microorganisms before treatment (n=20) | Number of microorganisms after treatment (n=20) | p       |
|-------------------------------------|----------------------------------------------------------|---------------------------------------------------------------------|---------------------------------------------------------------------|---------|
|                                     | M ± m | CFU/g |                   | M ± m | CFU/g |                   |                   |         |
| Bifidobacteria                     | 10^-10 | 1,02×10^10 ± 6,9×10^9 CFU/g | 1,18±10^10 ± 4,8×10^9 CFU/g | 0,001 |
| Lactic acid bacteria                | 10^-10 | 7,7×10^9 ± 8,8×10^8 CFU/g | 2,62×10^8 ± 8×10^8 CFU/g | 0,01    |
| E. coli with normal enzyme activity | 10^-10 | 2,53×10^9 ± 9,67×10^8 CFU/g | 3,20×10^9 ± 9×10^8 CFU/g | 0,05    |
| Opportunistic enterobacteria       | <10^-3 | Number of patients with bacteria above the norm | Number of patients with bacteria above the norm |         |
|                                    | CFU/g | abs (%) | CFU/g | abs (%) |         |         |
| Hemolytic active E. coli           | 0 | 10^-2 | 2 (10) | 10^-2 | 1 (5) |         |         | p<0,05  |
| S. aureus                          | 0 | 10^-2 | 1 (5) | 10^-2 | 1 (5) |         |         | p<0,000 |
| Candida yeast-like fungi and enterococci | <10^-3 | 10^-2 | 1 (5) | 10^-2 | 1 (5) |         |         | p<0,05  |
| Lactose-negative E. coli           | <10^-3 | 10^-2 | 1 (5) | 10^-2 | 1 (5) |         |         | p<0,000 |
| P. mirabilis                       | <10^-3 | 10^-2 | 1 (5) | 10^-2 | 1 (5) |         |         | p<0,000 |

1. # – degree of reliability in relation to the norm
2. ## – degree of reliability at p<0.01; ### – degree of reliability at p<0.001; """" – degree of reliability at p<0.001 between the value groups in CFU/g, abs – absolute value
number of microorganisms in stool samples significantly differed in each group with a statistically significant difference. The data obtained in this comparative analysis for the main and control groups are presented in Table 5.

Table 5 shows that before treatment, there was no statistically significant difference of beneficial and pathogenic intestinal microorganisms between groups (p > 0.05). While in the period after treatment, in patients of the main group, in comparison with the control group, the number of beneficial microorganisms was higher, and there was a decrease in the number of opportunistic enterobacteria (p < 0.001).

When studying the number of microorganisms in the intestinal microflora and comparing them with the degree of dysbiosis (the degrees of dysbiosis were determined according to the criteria of the sectoral standard of the Ministry of Health of Russian Federation “Protocol of treatment. Intestinal dysbiosis” [14]), the following results were revealed (Figure 2):

- Before treatment the 1st-degree intestinal dysbiosis was detected in 14 patients of the main group (70%) and in 15 patients of the control group (75%); 2nd-degree dysbiosis was detected in the remaining six patients of the main group (30%) and in the remaining five patients of the control group (25%);
- After treatment in the main group, the number of patients with 1st-degree intestinal dysbiosis decreased to three cases (15%), and the number of patients with 2nd-degree dysbiosis decreased twofold – three patients (15%), while the state of the intestinal microflora in the remaining 14 patients (70% of cases) corresponded to the norm;
- In the same period after treatment in the control group, the number of patients with 1st-degree intestinal dysbiosis decreased to four patients (20%), due to an increase in patients with 2nd-degree dysbiosis (the remaining 16 patients – 80% of cases). No patients had the normal parameters of the intestinal flora.

Figure 2 shows that in the period before treatment, there was no statistically significant difference

![Figure 2: Distribution of patients of the main and control group by the degree of intestinal dysbiosis before and after treatment](image)

Note: # – degree of reliability at p=0.000 in patients of the main and control group with 2nd-degree intestinal dysbiosis).

Table 4: Results of the study of the intestinal microflora before and after treatment of patients in the control group

| Microorganisms          | Norm in children (Industry standard 91500.11.0004-2003) | Number of microorganisms before treatment (n=20) | Number of microorganisms after treatment (n=20) | p     |
|-------------------------|-------------------------------------------------------|-----------------------------------------------|---------------------------------------------|-------|
|                         | M ± m                                                  | M ± m                                        |                                            |       |
| **Bifidobacteria**      | 10⁶⁻¹⁰⁶ CFU/g                                          | 1.207×10⁶ ± 6.9×10⁵ CFU/g                   | 2.577×10⁶ ± 8.19×10⁵ CFU/g                 | p<0.001|
| Lactic acid bacteria    | 10⁶⁻¹⁰⁶ CFU/g                                          | 6.3×10⁵ ± 1.05×10⁵ CFU/g                     | 8.8×10⁵ ± 1.08×10⁵ CFU/g                   | p=0.05 |
| E. coli with normal enzyme activity | 10⁻¹⁰⁶ CFU/g                                      | 2.08×10⁶ ± 7.84×10⁵ CFU/g                   | 5.01×10⁶ ± 8.86×10⁵ CFU/g                 | p=0.05 |
| Opportunistic enterobacteria | <10⁶                                              | Number of patients with bacteria above the norm |                                        |       |
|                         | M ± m                                                  | CFU/g                                        | abs (%)                                     |       |
| Hemolytic active E. coli | <10⁶                                               | -                                            | -                                          |       |
| S. aureus               | <10⁶                                               | 10⁶                                          | 10⁶                                        |       |
| Candida yeast-like fungi | <10⁻¹⁰⁶                                        | 10⁶                                          | 10⁶                                        |       |
| Lactose-negative E. coli | <10⁶                                              | -                                            | -                                          |       |
| Enterococci             | <10⁻¹⁰⁶                                            | 10⁶                                          | 10⁶                                        |       |
| K. pneumoniae           | <10⁶                                               | -                                            | -                                          |       |
| C. diversus             | <10⁶                                               | -                                            | -                                          |       |
| P. mirabilis            | <10⁶                                               | -                                            | -                                          |       |
| S. aureus and K. pneumoniae | 0                                                   | -                                            | -                                          |       |
| C. diversus and P. mirabilis | 0                                                   | -                                            | -                                          |       |

1. # – degree of reliability in relation to the norm:
   - # – degree of reliability at p<0.05;
   - ## – degree of reliability at p<0.01;
   - #### – degree of reliability at p<0.001;

2. # – degree of reliability at p=0.000 in patients of the main and control group with 2nd-degree intestinal dysbiosis.

Table 5: Comparative analysis of the results of the bacteriological examination of the intestinal microflora in patients of the main and control groups

| Microorganisms | Number of microorganisms before treatment (n=20) | p | Number of microorganisms after treatment (n=20) | p |
|----------------|-----------------------------------------------|---|-----------------------------------------------|---|
|                | M ± m                                        | M ± m                                |                                          | M ± m                                | |
| **Bifidobacteria** | 1.205×10⁶ ± 6.9×10⁵ CFU/g                   | p=0.05                              | 1.18×10⁶ ± 4.86×10⁵ CFU/g              | p=0.001                             |
| Lactic acid bacteria | 6.3×10⁵ ± 1.05×10⁵ CFU/g                     | p=0.05                              | 2.62×10⁶ ± 8×10⁵ CFU/g                 | p=0.001                             |
| E. coli with normal enzyme activity | 2.08×10⁶ ± 7.84×10⁵ CFU/g                   | p=0.05                              | 3.20×10⁶ ± 9×10⁵ CFU/g                 | p<0.01                             |
| Opportunistic enterobacteria | 5.01×10⁶ ± 8.86×10⁵ CFU/g                   | p=0.05                              | 1.502×10⁵ ± 3.4×10⁵ CFU/g              | p<0.001                             |

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in the degree of intestinal dysbiosis in patients of the main and control groups (p > 0.05). After treatment in the main group, in comparison with the control group, there was an increase in the number of patients with the normal intestinal microflora (14 patients – 70%) as well as a decrease in the number of patients with intestinal dysbiosis with an obvious statistically significant difference (p < 0.001).

Discussion

There are studies in the already existed scientific literature which dwell on the usage of probiotics only enterally [15], [16], [17], [18]. Shen et al. investigated the number of lactobacilli, probiotic bacteria, and bifidobacteria in 30 children with Hirschsprung disease (10 without enterocolitis, 20 with enterocolitis) and 15 healthy children in the control group. They found a statistically significant decrease in the level of probiotic bacteria in all 30 children and, moreover, revealed that the number of bifidobacteria is lower in patients with enterocolitis compared with patients without enterocolitis. Based on this, the authors concluded that a deficiency of bifidobacteria and lactobacilli in patients with enterocolitis may lead to a decrease in the barrier function of the intestinal epithelium and may be a predisposing factor in the development of HAEC; hence, treatment with prebiotics or probiotics of such patients may have a positive result [19].

However, there are studies that reject such a conclusion. For example, El-Sawaf et al. involved 32 patients with Hirschsprung’s disease to his study, they took the probiotics orally and after the calculations of the results, the authors concluded that taking probiotics does not reduce the risk of developing HAEC [20].

Against this background, the surveys on the implication of siphon enema and drug enema in the prevention of HAEC are not found at all. Hence, the use of probiotics by drug enema to reduce the risk of developing HAEC is a completely new way of treatment. The patent for the invention of the “Method for the prevention of HAEC in children” (No. 32679, dated 01.29.2018 by the Ministry of Justice of the Republic of Kazakhstan) was obtained for the developed method.

Limitation

The effectiveness of the developed method is reduced within Hirschsprung’s disease patients with an intestinal stoma since the presence of an intestinal stoma makes it difficult to perform a medicinal enema with probiotics of full value. Non-positive results might be obtained.

Conclusions

The usage of the developed method of biocenosis correction allows managing the negative effect of siphon enemas and normalizing the ratio of microorganisms inhabiting the large intestine. The method of preventing the inflammatory process in the intestine – HAEC – in accordance with the developed method of correcting the quantitative composition of the intestinal microflora by syphon enemas and probiotics is effective and can be introduced into clinical practice.

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