Here we describe a “light projector” system that can address, i.e. “read and write” electrochemical reactions on a non-structured macroscopic semiconducting electrode with spatial and temporal resolution. In our approach the illumination of an amorphous silicon electrode/electrolyte interface is spatially defined by means of a ferroelectric micromirror system that gives total freedom on both the two-dimensional light profile (illumination shapes) as well as on the transient times of the projected images. The device has no moving parts and allows for spatial and temporal control of the illumination stimulus driving local changes to the rate of an electrochemical reaction. The performance of the system is assessed by generating microscale patterns of Cu2O on the electrode (“electrochemical writing”) followed by their 2D current mapping (“electrochemical reading”) using methanol electro-oxidation and carbon dioxide electro-reduction. The latter illustrate the electrochemical imaging aspects of the device using two technologically relevant examples.

One could envision a simplification of these rather sophisticated electrochemical imaging techniques by shifting the tenet of one-electrode/one-wire toward one-wire/many-electrodes, doing so by turning an insulator into a transient conductor only at one specific site that is defined by the experimentalist. Some of us10–12 and others13–17 have recently started making use of semiconductor electrodes to drive redox processes in a spatial-selective fashion. In this context, Choudhury et al. recently reported a strategy to spatially confine an electrochemical reaction using the intrinsic semiconducting property of silicon to generate and separate charge carriers when illuminated.10 These charge carriers oxidize or reduce nearby species present in the electrolyte or tethered to the surface. Directing the flux of carriers on the surface is a matter of controlling the illuminated areas, as schematized in Figure 1a. As a proof-of-principle experiment, Choudhury et al. used a light pointer mounted on a travelling x-y stage to guide the illumination in 2D. However, to fully exploit the potential of this “light-addressable” electrochemistry concept it is necessary to develop a method that allows to control the time and shape of the illumination with greater freedom, therefore using an illumination source akin of a microscale projector.

The field of spatial localization of electrochemical reactions at interfaces has made tremendous improvements over the last three decades, and recent conceptual and technical advances in the fields of catalysis, energy and sensing have been linked to our ability to quantify adventitious or deliberate two-dimensional inhomogeneity of an electrified interface.1–5 Addressing an electrochemical reaction with spatial resolution was originally done by the construction of individually-addressable electrochemical arrays, where each electrode is individually connected to an external power source or measuring instrument.6 These arrays use considerable space for the pad connections and the geometry is pre-defined and restricted by the array design, giving limited flexibility to the experimentalist. Moreover, the construction of individually-addressable electrochemical arrays is an expensive and laborious task. One strategy to overcome this problem has been implemented by using a single ultramicroelectrode tip that is mounted on a travelling stage. This is the so-called scanning electrochemical microscopy (SECM), where an ultramicroelectrode is scanned over the sample and without any geometrical restriction. Since the SECM invention,7 other scanning electrochemical probe techniques have been developed and used extensively in research, with notable examples being scanning electrochemical cell microscopy, SECCM,8 and scanning photoelectrochemical microscopy, SPECM.9 These scanning techniques are very powerful instruments, both for surface characterization as well as modification.8 For instance, electrochemical scanning techniques can now claim spatial resolution down to few nanometers,2 and imaging speed as high as few seconds per frame.9 However, scanning electrochemical techniques present the intrinsic characteristics, and to a degree a limitation, of a scanning probe method. They are “sequential” methods, i.e. only one discrete area can be addressed at a certain time, with the moving tip possibly generating convective forces in the electrolyte.9 Furthermore, the investigating ultramicroelectrode “probe” is approaching the electrode from the electrolyte side, hence for instance we can only investigate charge transfer kinetics at an electrified surface indirectly from the electrolyte “point of view”. This is because scanning electrochemical techniques have a critical requisite of a one-electrode/one-wire, i.e. it is only possible to turn on the entire conductive interface and then raster the probe to a specific site.

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Projection System to Spatially Address Electrochemical Reactions

Electrochemical Microscopy Based on Spatial Light Modulators: A Projection System to Spatially Address Electrochemical Reactions at Semiconductors

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Figure 1. Working principle of the device. (a) Illumination of a photoelectrode generates charge carriers that are used to drive electrochemical reactions, in this case the discharge of metal ions with formation of metallic patterns only on illuminated discrete areas of the electrode surface. (b) Diagram of the set-up. This device allows to control illumination on the photoelectrode hence confining the electrochemical reaction in space and time. This concept can be used in semiconductors or photoconductors, for example, to deposit particle micropatterns by simply projecting on the substrate a specific user-defined illumination pattern. (c) SEM image of a Cu$_2$O micropattern deposited using this concept by discharging cupric ions from the electrolyte solution. An array of bright squares 46 $\mu$m in width and separated by gaps of 92 $\mu$m was projected on the surface (d) Picture of the actual set-up here described. The electrodes are connected to a general potentiostat which has been omitted for clarity (see Figure S2 in the supplementary material for a picture including the potentiostat).

Experimental

Overall instrumental design.—The instrument consists of four main parts schematically depicted in Figure 1b: 1) a “microdisplay” that allows spatial and temporal control of the light, which is composed of a light source, a ferroelectric mirror and optics, 2) a single-compartment, three-electrode cell where the photoelectrode is placed and connected to 3) a single-channel potentiostat, and 4) a computer to control both the microdisplay and the potentiostat. Figure 1d shows a picture of the whole device. Supporting Table S1 is a detailed description of all the components.

Microdisplay.—Light from an LED is projected onto a photoconductive amorphous silicon working electrode after being reflected by a ferroelectric micromirror device that acts as spatial light modulator.
Figure 2. Main components of the microdisplay set-up. (a) Scheme of the main parts comprising the microdisplay. The light is represented as a solid red line with an arrow indicating the direction of travel and perpendicular arrows indicating its polarization. (b) Top-view picture of the microdisplay indicating its main components. Parts are numbered according to Table S1 in the supplementary material.

orientation. This gives to the light reflected by each FLCoS element only two statuses, which can be thought as ON/OFF. For the ON status the polarization is changed by 90° allowing the beam to go back toward the polarizer beam splitter and to cross it undisturbed. On the other hand, the OFF status of an FLCoS element does not change the polarization and the light is reflected by the polarizer beam splitter and removed from the system. All the ON statuses after crossing the polarizer beam splitter are collected by the focus lens (6) and finally projected onto the substrate situated in a three electrodes electrochemical cell ((9) and (10) in Figure 2 and Figure 3). The focus lens (6) and the cell holder (9) sit on x-y-z traveling stages (8) to allow focusing and/or magnification/reduction of the projected image. To prevent damages to the liquid crystals, the net bias across it must equal zero over time, that is to say each ferroelectric liquid crystal micromirror must be switched ON and OFF intermittently. Because of this, the projected light is intermittent in nature with a frequency that can be modulated in a range between ca. 500 Hz to ca. 10 kHz. This requirement needs an accurate synchronization between the LED light source and the FLCoS micromirror device which is achieved by means of two separate boards (7); one board for the LED driving and the other is...
Figure 3. a) Scheme and b) picture of the electrochemical cell. The electrochemical cell was CNC-machined from a PTFE block. Two “o-rings” (not shown in the scheme) are placed between the FTO and electrolyte holder and between the working electrode and electrolyte holder to prevent leaking of the electrolyte.

the microdisplay interface for timing, ordering and synchronization of the images uploaded on its memory.

Electrochemical cell and amorphous silicon substrate preparation and derivatization.—Figure 3a shows a blown-up scheme of the electrochemical compartment. The electrochemical cell was machined with a 5-axis CNC from a polytetrafluoroethylene (PTFE) block. The geometry of the electrochemical cell was studied to allow the spatially-modulated light beam coming from FLCoS to be illuminated on the working electrode surface with minor scattering from a liquid/air interface. The light is therefore projected on the working amorphous silicon electrode from the electrolyte side after passing through a conductive layer of FTO deposited on a glass slide. The latter serves also as the counter electrode. The light path through the electrolyte was minimized to a length of ca. 2.5 mm to avoid as much as possible light intensity absorbance from the electrolyte. An Ag/AgCl/KCl (sat.) reference electrode is placed in a small reservoir/port sitting just on the top of the working and counter electrodes. This port is also used to inject electrolyte in the cell and remove air bubbles. The electrochemical cell is then fitted on a holder (9) which is mounted tightly on a travelling stage ((8), see Figures 1d and 2) to keep it at an accurate position and/or allowing for precise changes to its position.

As explained in the introduction section, spatially and temporally addressing electrochemical reactions requires a light stimulus as well as a photoconductive substrate, i.e. a substrate that allows electron transfer only when illuminated. For that we used a photoconductive thin layer (1 μm) of intrinsic amorphous silicon (a-Si hereafter) deposited on a highly-conductive substrate (a Si(100) wafer with a resistivity of 0.001–0.004 Ω cm). The a-Si layer faces the electrolyte, from where it is illuminated, and the Si(100) wafer is back-contacted against a copper plate after applying gallium-indium eutectic to ensure an ohmic contact before connecting to the potentiostat circuit. The a-Si thin layer acts as a dark insulator and a photoconductor; only at the illuminated area the conductivity increases to address localized electrochemical reactions.

Amorphous Silicon (a-Si) preparation method.—P-type Si(100) wafers with a resistivity of 0.001–0.004 Ω cm (boron-doped) were rinsed with DCM (Ajax Finechem), dried under a stream of argon, and immersed in piranha solution (3:1 (v/v) mixture of concentrated sulfuric acid (H2SO4, Baker) and 30% hydrogen peroxide) for 10 min. Samples were then rinsed with Milli-Q water for 10 min before being transferred to hydrofluoric acid (HF, EMSURE grade, Merck) aqueous solution (1:10 HF/water T = 30 °C) for 60 s. Finally, the Si-H wafers were rinsed with Milli-Q water for 1 min. Undoped amorphous silicon was then deposited using an Oxford Instruments Plasmalab 100 plasma-enhanced chemical vapor deposition system. SiH4 (Coregas, 99.999%) was used as the precursor gas and Ar (Coregas, 99.99999%)
Substrate micropatterning with Cu$_2$O particles (a) Image that is projected on the a-Si photoelectrode and used to guide the deposition of the Cu$_2$O pattern shown in panel (b). The projected pattern is comprised of an array of 16 × 16 squares, each square being 46 μm wide and separated from the neighboring ones by gaps of 92 μm. (b) SEM image of the deposited pattern of Cu$_2$O particles that is generated by projecting on the photoelectrode the image shown in panel (a). A single step potential of −0.5 V was applied and stopped when the transferred charge reached 500 μC (ca. 90 s). The electrolyte solution was 0.5 M K$_2$SO$_4$ and 50 mM CuSO$_4$. (c) SEM images of lined-shaped Cu$_2$O patterns. In this case two line shaped images were projected sequentially allowing for changes to the deposition potential of each "line" (the deposition potential is shown as label in figure). These changes guide the shape of the deposited particle. The two sequentially projected images are "lines" of 46 μm in width that are separated by a 230 μm gap. The electrolyte solution was 0.5 M K$_2$SO$_4$ and 50 mM CuSO$_4$ with a trace amount of KCl. All potentials are given against the Ag|AgCl|KCl (sat.) reference electrode.

was employed as the carrier gas. Plasma was generated using a 13.56 MHz 300 W RF generator with auto matching unit. Deposition of amorphous silicon was performed at 5 × 10$^{-2}$ mTorr at 300 °C until a thickness of 1 μm was reached. Silicon functionalization. To ensure protection of the silicon electrode against anodic decomposition$^{18,19}$ we grafted an organic layer of 1,8-nonadiyne by means of UV-assisted hydrosilylation reactions to ensure the silicon stability against anodic decomposition.$^{20}$ The a-Si wafers were cut into 1 × 1 cm pieces, cleaned with dichloromethane and water, immersed in piranha solution (100 °C, 3:1 (v/v) mixture of concentrated sulfuric acid and 30% hydrogen peroxide), rinsed thoroughly with Milli-Q water, etched for 10 min in aqueous ammonium fluoride solution (degassed by bubbling with argon gas for 20 min in the presence of a small amount of ammonium sulfite), rinsed sequentially with Milli-Q water and dichloromethane, dried under a nitrogen stream and placed on a glass sheet before dropping a small sample of 1,8-nonadiyne (ca. 50 μL) on the wafer. To limit evaporation of the diyne, the liquid sample was top-contacted with a quartz slide. The samples were kept under positive nitrogen pressure and under UV light for 2 h (Vilber, VL-215.M, λ = 312 nm, nominal power output of 30 W and positioned approx. 200 mm away from the silicon sample). The chemically-passivated a-Si samples were then rinsed several times with dichloromethane, rested for 12 h in a sealed vial under dichloromethane at +4 °C and blown dry under nitrogen before analysis. Chemicals: Unless noted otherwise all chemicals were of analytical grade. 1,8-Nonadiyne (98%), H$_2$SO$_4$ (PURANAL, 98%), H$_2$O$_2$ (MOS PURANAL, 30%) were obtained from Sigma Aldrich. Argon gas and CO$_2$ gas were obtained from BOC and of Ultra High Purity.

Potentiostat.—A general one-channel electrochemical workstation (CHI650E, CH Instruments) which connects to a three-electrode electrochemical cell (a-Si working, Ag|AgCl|KCl (sat.) reference and FTO on glass counter electrodes) was used for potential control and data acquisition.

Projection system control.—The projected images are constructed as a 1-bit per pixel bitmap files with MATLAB. Each pixel of the generated bitmap file contains the ON/OFF information that controls each individual FLCoS element to generate the final projected image. Therefore, the bitmap file resolution has to match exactly with the micromirror size, i.e. 2048 × 1536 pixels. Once generated, the image or sequence of images to be projected are
uploaded to the memory on the printed circuit board of the micromirror device (7) using the commercial software MetroCon V3.2 (Forth Dimension Displays) and programmed to be linked to a specific light modulation frequency, time and sequence. The number of images to be uploaded is intrinsically limited by the internal memory, 4 Gbit corresponding with 1024 1-bit images at the given resolution. In this way a movie, i.e. a sequence of images linked to the time each one is projected, is generated. The size of each pixel of the projected image at the magnification here used is 4.6 μm, which was measured with a travelling microscope by projecting a square of a known number of pixels and then dividing the length measurement of the square by the number of pixels (known from the uploaded image), hence giving the size of a single projected pixel. Note that a square of 4.6 μm is therefore in the current set-up the minimum area the electrochemical reaction can be resolved.

Results and Discussion

Experimental verification of the device.—In order to experimentally verify the performance of the instrument described above with regards to spatially (2D) resolving electrochemical reactions we selected particle deposition as a proof-of-concept example. The electrochemical reduction of copper (II) ions was used to deposit Cu2O particles arranged in microscale patterns on the a-Si working electrode surface. This reaction, according to published work, takes place via a two-step mechanism with first the reduction of the cupric to cuprous ions, followed by precipitation of Cu2O through the following reaction: Cu2+ + H2O → Cu2O + 2H+. X-ray photoelectron spectroscopy experiments shown in Figure S2 of the supplementary material confirm that the particle composition is Cu2O. The spatial-temporal control on the light achieved by the set-up described here permits to confine electrochemical reactions to discrete regions of the photoelectrode. By generating light patterns on the electrode surface, such as the one shown in Figure 4a, the electrochemical reduction of copper ions will take place only where the light impinges the surface and the deposition can therefore be spatially addressed. This generates Cu2O patterns that accurately follow the projected image. Figure 4b shows a deposited pattern of Cu2O made by projecting the image of Figure 4a. The electrochemical resolution is somewhat restricted by the dimensions of the projected light when a single element of the FLCoS is turned “ON”, which in our system is a 4.6 μm. When compared with scanning electrochemical techniques that nowadays have resolution of few nanometers this resolution seems to be quite low; however, there is the theoretical possibility of improving resolution R toward the diffraction limit, which is given by R = λ/2π, where λ is the light wavelength and NA the numerical aperture. This makes it in principle possible to eventually reach a sub-micrometer resolution. As the active element is an array containing more than 3 million of dynamic mirrors, this micropatterning method does not need photomasks and the deposited pattern is built within few seconds, in contrast to sequential deposition methods such as the SECM that need to “scan” to build the pattern or photolithography which relies in the use of photomasks. However sequential deposition, i.e. depositing one specific area after another, is also possible. This is important as depositing patterns sequentially makes possible to control the deposition conditions, e.g. controlling particle morphology, size or density over the substrate by varying the deposition conditions from one region to another. For example, we found that changes in particle “cubicity” are driven by the applied bias; the particle shape changes progressively from cubes to octahedrons as the potential is made more cathodic. Hence, the possibility to grow particles in different areas while changing the deposition conditions makes this technique of interest in research aimed at a systematic screening of factors such as deposition potential on the evolution of crystals’ shape. For example, in Figure 4c firstly the left “line” was deposited by projecting a rectangular image, i.e. the “line”, while applying a potential of −0.5 V vs Ag/AgCl/KCl (sat.) to the entire substrate and stopping this when a charge of 500 μC had passed. After this, the right “line” region was illuminated while applying a

![Figure 5](image)

Figure 5. Electrochemical imaging for methanol oxidation. (a) Electrochemical image taken with the device here described for the electrooxidation of methanol at +0.5 V (vs Ag/AgCl/KCl (sat.)) by sequentially illuminating square sections of 46 μm one next to the other at 17 Hz. Electrolyte is 0.1 M NaOH and 0.25 M methanol. Each pixel on the image is 46 μm in width. (b) SEM image of the Cu2O “line” showing a match in the width with the electrochemical image. Images in panels (a) and (b) are shown at the same scale. (c) Current-time data of the electrochemical image shown in panel (a) before being converted to the bitmap image file. The time 0 s corresponds with the bottom-left pixel of the electrochemical image in panel (a), and the time 59 s with the top-right pixel. The current intensity data was averaged every 59 ms (59 points at 1 ms sampling interval), corresponding with a single pixel in the electrochemical image. The series of “spikes” in the current transient data correspond to the time when the projected light passes over the Cu2O “line”, which gives a higher current intensity signal compared to the bare photoconductor. The alternate high and low intensity current “spikes” relates to different activities of cubic vs. octahedral particles. (d) Averaged electrochemical 1D current-distance traces sampled by rastering the light stimulus perpendicular to the lines of Cu2O particles (x-direction distance) at different y-distance offsets. The red contour represents the standard deviation.
potential of 0.0 V and stopping after reaching the same charge. The only difference on the deposition conditions of the two “lines” is the deposition potential: at more cathodic potentials the electro-deposited particles grow as cuboctahedrons while at less cathodic potentials the particles’ grow as cubes. This morphology control is interesting as the catalytic and electronic properties of nanoparticles is in general, and for Cu2O in particular, linked to shape.22,23

Cu2O is known to be a good catalyst for a variety of reactions such as for example methanol oxidation and carbon dioxide reduction.24 This aspect is used here to illustrate the “electrochemical reading” potential of the device, that is to say, using a structured and rastering light beam to localize electrical conductivity with microscale resolution to interrogate the electrochemical activity of the surface. Figure 5a shows an electrochemical image for the methanol oxidation taken on a photoelectrode modified with Cu2O particles patterned by the method described above: two “lines” were deposited at two different potentials to obtain cuboctahedral particles (left line) and cubic particles (right line). To construct the electrochemical image from Figure 5a individual light squares of 46 μm in width (equivalent to 10 pixels of the uploaded image) were projected for 59 ms in rapid sequence, one immediately after the other (corresponding to a frequency of 17 Hz). A constant potential of +0.5 V vs Ag/AgCl/KCl (sat.) is applied during the whole sequence which takes a total time of 59 s to record (Figure 5a). The electrochemical image in Figure 5a was generated using the current-time data shown in Figure 5c; the time data was transformed into (x-y) coordinates by simply knowing the amount of time and area each square was projected and associated with the electrochemical current at that specific time-area. Each of the projected squares therefore represent a “pixel” in the electrochemical image. We note that each of the pixels shown in the electrochemical image is formed from a total of 59 averaged data points, as the sample interval used was of 1 ms and each square was projected for 59 ms. The image resolution is only restricted by the micromirror memory which can store up to 1024 images, corresponding for example to an electrochemical image of 32 × 32 pixels. The scan frequency is in principle limited only by the charging time of the double layer.

Remarkably, the electrochemical image that generates from the sample is in very a good agreement with the known physical width of the “lines” as determined by SEM (Figure 5b). Even more interesting, the pattern made up of cubic Cu2O particles shows the highest current intensity toward methanol oxidation, a fact that is apparent to the eye of the observer when looking at the electrochemical image in Figure 5a. Figure 5d shows 2D x-direction current profiles that are generated by “rastering” at different y-direction offsets the light stimuli first across the “line” formed of cuboctahedral particles and then across “line” formed of cubic particles (left to right in Figures 4 and 5) and it reinforces that a higher current intensity is obtained for the cubic-shaped particles. Capacitance measurements (see Figure S3 in the supporting material) suggest this difference in current magnitude is a result of a face-dependent effect rather than due to a difference in the active surface area. This is probably related with the finding of previous reports that assigned a higher catalytic activity to the (100) face of cubic Cu2O particles.24,25 We remark that having the two patterns under scrutiny (i.e. cubes vs. polyhedron) deposited over the same electrode, exposed to exactly the same electrolyte, pH, same illumination intensity and same cell geometry, removes a great deal of experimental ambiguity when attempting a comparison between catalytic properties of the differently shaped particle deposits. This aspect highlights one of the key advantages of electrochemical microscopy principle delivered by our “micro-projector” design. Finally, although the data above demonstrate the electrochemical reading capabilities of this device for electro-oxidation reactions we show that electro-reductions are also possible. Figure 6a shows an electrochemical image for the reduction of CO2 on a photoelectrode modified with Cu2O particles deposited locally as shown in the SEM image of Figure 6b. As for the data in Figure 5, the dimensions of
the electrochemical read-out are in good agreement with the known width of the Cu2O pattern (Figures 6c and 6d).

Conclusions

Here we have described the construction of a device that performs spatially-resolved electrochemical reactions by projecting user-defined light patterns on photoelectrodes. The light generates charge carriers on the photoelectrode allowing the reaction to happen only with an appreciable rate in correspondence of the illuminated area. This device has no moving parts, uses a general one-channel potentiostat and an unstructured surface, giving total freedom on the space and time of the illumination and therefore also on the space and time of the electrochemical reaction. We have experimentally tested this device for micropatterning of particle deposits on a photoconductor amorphous silicon surface. Microscale patterns can be built very rapidly, without the need of photomasks, by projecting an arbitrary light pattern by means of a dynamic array of FLCoS micromirrors. In particular, we showed this micropatterning capability by electrodepositing patterns of Cu2O particles. We also showed that by changing the electrodeposition potential the particle’s degree of “cubicity” can be controlled over space. The ability to use light to localize electrochemical reactions with micron-scale resolution can in turn be used to interrogate the electrochemical activity of the surface. We have shown this by spatially resolving the surface electroactivity of Cu2O particles toward methanol oxidation and CO2 reduction on an amorphous silicon semiconducting electrode. As the device does not present moving parts this “reading” is done rapidly, where the only practical restriction comes from the charging of the double layer. Patterns of different materials such metals, metal oxides, polymers or molecules could be possibly made on many types of semiconducting parts this “reading” is done rapidly, where the only practical restriction comes from the charging of the double layer.

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References

1. A. J. Bard, R. F. Fan, D. T. Pierce, P. R. Unwin, D. O. Wipf, and F. Zhou, Science, 254, 68 (1991).
2. R. F. Fan, F. Ciucci, A. N. Morozovska, S. V. Kalinin, and S. Jesse, Nat. Chem., 3, 707 (2011).
3. S. Bergner, P. Vatsayan, and F. -M. Matsyk, Anal. Chem., Acta, 775, 1 (2013).
4. D. Polcari, P. Dauphin-Ducharme, and J. Mauzeroll, Chem. Rev., 116, 13234 (2016).
5. F. Conzuelo, K. Sliozberg, R. Gutowski, S. Grützke, M. Nebel, and W. Schuhmann, Anal. Chem., 89, 1222 (2017).
6. E. Cortón and S. R. Mikkelsen, Electrochemical Arrays for Bioassay Applications, in: F. -M. Matsyk, (Ed.) Trends Bioelectroanal., Springer International Publishing, 17, 2017, pp. 103.
7. A. J. Bard, F. R. F. Fan, J. Kwak, and O. Lev, Anal. Chem., 61, 132 (1989).
8. D. Momotenko, J. C. Byers, K. McKelvey, M. Kang, and P. R. Unwin, ACS Nano, 9, 8942 (2015).
9. C. Combellas, M. Fermigier, A. Fuchs, and F. Kanouti, Anal. Chem., 77, 7966 (2005).
10. M. H. Choudhury, S. Ciampi, Y. Yang, R. Tavallaiye, Y. Zhu, L. Zarei, V. R. Goncalves, and J. J. Gooding, Chem. Sci., 6, 6769 (2015).
11. Y. Yang, S. Ciampi, Y. Zhu, and J. J. Gooding, J. Phys. Chem. C, 120, 13032 (2016).
12. Y. B. Vogel, N. Darwish, M. B. Kash, J. J. Gooding, and S. Ciampi, Electrochim. Acta, 247, 200 (2017).
13. B. Y. Chow, C. J. Emig, and J. M. Jacobson, Proc. Natl. Acad. Sci. U. S. A., 106, 15219 (2009).
14. T. L. Rose, D. H. Longendorfer, and R. D. Rash, Appl. Phys. Lett., 42, 193 (1983).
15. S. -H. Huang, L. -S. Wei, H. -T. Chu, and Y. -L. Jiang, Sensors, 13, 10711 (2013).
16. Y. -R. Kim, J. -K. Lee, J. -K. Lee, W. Jang, J. -Y. Lee, J. H. Bae, and T. Y. Chang, Energy Environ. Sci., 8, 3654 (2015).
17. N. Liu, F. Wang, L. Liu, H. Yu, S. Xie, J. Wang, Y. Wang, G. -B. Lee, and W. J. Li, Sci. Rep., 6, 32106 (2016).
18. S. Ciampi, M. James, G. Le Saux, K. Gaus, and J. Justin Gooding, J. Am. Chem. Soc., 134, 844 (2012).
19. L. Zhang, Y. B. Vogel, B. B. Noble, V. R. Goncalves, N. Darwish, A. L. Brun, J. J. Gooding, G. G. Wallace, M. L. Coote, and S. Ciampi, J. Am. Chem. Soc., 138, 9611 (2016).
20. Y. B. Vogel, L. Zhang, N. Darwish, V. R. Goncalves, A. L. Brun, J. J. Gooding, A. Molina, G. G. Wallace, M. L. Coote, J. Gonzalez, and S. Ciampi, Nat. Commun., 8, 2066 (2017).
21. K. -S. Choi, Dalton T., 5432 (2008).
22. C. -S. Tan, S. -C. Hsu, W. -H. Ke, L. -J. Chen, and M. H. Huang, Nano Letters, 15, 2155 (2015).
23. H. Mistry, A. S. Varela, S. Kuhl, P. Strasser, and B. R. Cuenya, Nat. Rev. Mater., 1, 16009 (2016).
24. M. B. Gawande, A. Goswami, F. -X. Felpin, T. Asefa, X. Huang, R. Silva, X. Zou, R. Zboril, and R. S. Varma, Chem. Rev., 116, 3722 (2016).
25. R. Venkatasubramian, J. He, M. W. Johnson, I. Stern, D. H. Kim, and N. S. Pesika, Langmuir, 29, 13135 (2013).