Age of Both Parents Influences Reproduction and Egg Dumping Behavior in *Drosophila melanogaster*

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Abstract

Trans-generational maternal effects have been shown to influence a broad range of offspring phenotypes. However, very little is known about paternal trans-generational effects. Here, we tested the trans-generational effects of maternal and paternal age, and their interaction, on daughter and son reproductive fitness in *Drosophila melanogaster*. We found significant effects of parent ages on offspring reproductive fitness during a 10 day postfertilization period. In daughters, older (45 days old) mothers conferred lower reproductive fitness compared with younger mothers (3 days old). In sons, father’s age significantly affected reproductive fitness. The effects of 2 old parents were additive in both sexes and reproductive fitness was lowest when the focal individual had 2 old parents. Interestingly, daughter fertility was sensitive to father’s age but son fertility was insensitive to mother’s age, suggesting a sexual asymmetry in trans-generational effects. We found the egg-laying dynamics in daughters dramatically shaped this relationship. Daughters with 2 old parents demonstrated an extreme egg dumping behavior on day 1 and laid >2.35× the number of eggs than the other 3 age class treatments. Our study reveals significant trans-generational maternal and paternal age effects on fertility and an association with a novel egg laying behavioral phenotype in Drosophila.

Key words: age, carry-over effects, fertility, parental, paternal, trans-generational

Investigations of aging effects on fitness, particularly fertility, have largely focused on the influence of an organism’s age on reproductive senescence within a generation, for example, offspring viability (Kern et al. 2001). For example, studies in *Drosophila melanogaster* have shown increasing female age to be associated with a range of reproductive phenotypes and behaviors including the numbers of eggs laid and egg to adult viability (reviewed in Miller et al. 2014). Very little attention has been aimed at understanding trans-generational effects of parental age on offspring fitness traits (but see Koch et al. 2018), in spite of a growing body of evidence that trans-generational effects are prevalent throughout model animal study systems (Mousseau and Dingle 1991; Sartorius and Nieschlag 2010; Rando 2012) and can impact human health (D’Onofrio et al. 2014; McGrath et al. 2014).

One of the first examples of trans-generational effects of maternal age on offspring quality was observed in mice (Wang and vom Saal 2000) in which maternal age at first pregnancy was associated with body weight, testes size, and epididymis size in offspring. The conclusions from that study were that hormone concentrations in utero could have imprinting effects on the offspring and these effects
could change with changing female physiology, and therefore age. More recently, investigations have revealed that epigenetic information of parental diet can be inherited through the male germline (mouse: Carone et al. 2010) and that paternal diet per se can influence offspring characteristics (D. melanogaster: Valtonen et al. 2012). Hereafter, we refer to the broad term “epigenetics” as chemical modifications to DNA and/or histones that are stably maintained, and while not changing the DNA sequence, can heritably alter gene expression (reviewed in Feil and Fraga 2012).

Although maternal effects (Mousseau and Dingle 1991) and in utero environment can influence offspring phenotypes, paternal effects are more difficult to detect, since males contribute little more than sperm and seminal fluid in the ejaculate when paternal parental care is absent (Clutton-Brock 1991; Carone et al. 2010). In spite of this limited provisioning to offspring, paternally derived seminal proteins exert a large impact on female reproductive physiology and egg-laying behavior (Avila et al. 2011) and these effects can be detected in the next generation (Priest et al. 2008). For example, paternal tetracycline exposure has been shown to affect offspring sperm viability in pseudoscorpions Cordylochernes scorpioides (Zeh et al. 2012). Also in Drosophila, paternal age has been shown to influence olfactory memory (Burns and Mery 2010); consistent with various psychiatric disorder associations with advanced paternal age in humans (D’Onofrio et al. 2014; McGrath et al. 2014).

An early study in Drosophila serrata revealed maternal age effects could impact offspring fitness, and were cumulative in effect across 2 generations of old mothers (Hercus and Hoffmann 2000). A more recent study found that offspring viability in D. melanogaster is influenced by a complex 3-way interaction between parental relatedness, parental age, and gametic age at successive developmental stages (Tan et al. 2013). A maternal age × paternal age interaction has also been noted in D. melanogaster daughters and the highest reproductive fitness was found in the old mother: young father combination (Nystand and Dowling 2014). Focusing more specifically on paternal age, a previous comparison between young (2 days old), intermediate (13–14 days old), and old (32–33 days old) paternal ages in D. melanogaster revealed that offspring of old fathers suffered significantly lower larval viability (egg-to-adult survival) than those from young fathers (Price and Hansen 1998). In the same study, there were no significant effects of paternal age class on daughter fecundity; measured as the number of eggs laid, and male fertility was not directly assessed. Instead, son mating ability (presence or absence of copulation) was measured revealing no significant effect of paternal age treatment.

Beyond fertility, other studies have found no consistent evidence across various genotypes that paternal age influences longevity (D. melanogaster: Priest et al. 2002), although there is some evidence that paternal age can affect longevity in humans (Gavrilo et al. 1997). Reproductive potential declines with age in insects (Parsons 1964; David et al. 1975) and also in humans (Tarín et al. 2000). The aim of this investigation was to test whether there was any influence of paternal age on offspring fertility in an F1 generation, to understand whether paternal age can alter offspring phenotypic traits and carry-over between generations. Here, using the fruit fly D. melanogaster, we tested whether there was an association between the age of a focal fly’s parents and the fitness of the focal fly, measured as the number of eggs laid, the total number of offspring produced and the egg-to-adult viability. We tested fitness in both sexes; with 4 experimental parent age treatments (Supplementary Figure S1). Briefly, the parents of the focal individuals were either 3 days old or 45 days old. The experimental parental classes (4 in total) were: 1) young mother; young father; 2) old mother; young father; 3) young mother; old father; and 4) old mother; old father. This study design allowed us to test whether there is any trans-generational influence of parental age on offspring fertility and disentangle whether there is an effect of sex of the parent that was old or young. As a result, we measured the trans-generational effects of parental age on fitness and not the direct fitness of aged individuals because all the daughter and son (F1) individuals in the fertility assay were the same age (3 days old at the start of the 10 day egg lay).

Materials and Methods

Fly Husbandry

We used one genotype throughout the experiment: an inbred Oregon R strain produced by balancer chromosome replacement whose construction was completed by backcrossing for 3 generations to homogenize the genetic background (Montooth et al. 2010). Flies were maintained throughout the experiment at 25 °C on a 12 h: 12 h light: dark regime, with humidity at 70% ± 5%. Before the experimental procedures, flies were density controlled (25 females and 25 males; 4-day egg lay) for 2 generations on standard lab fly food. The recipe for the food in these holding vials was identical to that described below (see Fertility Assay section), except 2% autolyzed yeast was used instead of 6%, and the volume of food was 10 mL.

Experimental Design

The aim of the experiment was to manipulate the age of parents of the focal daughters and sons that were used in the fertility assays (see design scheme in Supplementary Figure S1). To achieve this, we isolated virgin males and females from the density-controlled stocks and housed them in same sex vials for 45 days. During this same 45-day period, the flies from the stocks were maintained (density-controlled) to then become the parents of young (3-day-old) flies, which were used in the experimental crosses. For the 45-day aging treatment virgin flies were transferred every fourth day onto a fresh food source (2% yeast w/v; as above) with additional sprinkled yeast on the surface. Previous life span studies in the same genotype suggest ~85% of the population is alive at day 45 on a similar food type (Villa-Cuesta et al. 2014).

At the stage of generating the focal experimental flies (below), young and old flies were allocated evenly across all aged and young treatment crosses. This was to ensure there was no bias of vial of origin at the parental generation stage and any inherent (vial) variation in quality was spread across all treatments. The “young” or “old” flies were then mated to produce daughters and sons for the experimental generation in a reciprocal design.

Estimating Genetic Variation in Our Sample Population

We conducted a supplementary analysis using RNA-seq reads from a previous study (Mossman et al. 2016, 2017) to estimate the numbers of SNPs within this genetic stock and estimated low levels of genetic variation throughout the transcriptome. These analyses were based on ~87.1 Mb of exome sequence. For brevity here, we have described this experiment in detail in the Supplementary Materials (section iv) and provide a brief description of the results in the main article.

Generation of Experimental Flies

Parents of the focal (experimental) generation were either young (3 days old) or old (45 days old) when they were mated. When the aged flies were 45 days old and the young flies were 3 days old, they were mated in their mating treatment.
These were:

(i) 3-day-old mother × 3-day-old father
(ii) 45-day-old mother × 3-day-old father
(iii) 3-day-old mother × 45-day-old father
(iv) 45-day-old mother × 45-day-old father

A schematic of the crosses and experimental procedures are shown in Supplementary Figure S1. The offspring eclosing from these crosses were used in the fertility assay. Adult flies eclosed and were immediately collected as virgins for the fertility assay. Males and females were separated as virgins and aged for 3 days before the fertility assay was conducted (Supplementary Figure S1).

Where possible, an individual virgin daughter or son was randomly selected from the vial of eclosing adults. In the treatments in which fewer than 23 of the vials produced offspring, we sampled multiple virgin males and virgin females from the same vial. Vial ID was fit as a random term in models to account for this shared vial provenance of some flies.

To test the effects of parental age treatment on male fertility, we used the same virgin tester female type across alternative male types. For example, 3-day-old virgin sons from the treatments (i–iv (above)) were all mated to a 3-day-old virgin female who emerged from the treatment (i). This young mother: young father treatment was considered a reference group, since 2 young flies (3 days old) mating is the most likely of our 4 scenarios to occur in natural populations, based on reproductive fitness relationships with age (Ashburner 1989) and very short sexual maturation times in D. melanogaster (Miller et al. 2014). For daughter fertility measures, we used 3-day-old virgin daughters that were the products of treatments i–iv (above) and mated these to 3-day-old virgin males from treatment (i) (Supplementary Figure S1).

Fertility Assay

The fertility assay was conducted on 6% yeast w/v food with no additional sprinkled yeast on the food surface. The full food recipe is as follows: 11% sugar, 6% autolyzed yeast, 5.2% cornmeal, agar 0.79% w/v in water, and 0.1% tegosept-methyl 4-hydroxybenzoate, from Sigma (St. Louis, MO). Food (approximately 5 mL) was added to each vial (glass narrow shell).

The aim of the fertility assay was to obtain an estimate of offspring productivity of each treatment for both daughter and son fertility over a 10-day period. We mated the treatment isofemale or isomale to the tester male or female individual, respectively, for 24 h. Each vial contained 1 male and 1 female. After 24 h exposure, the male was removed from the female and the female was transferred into a fresh vial. The female was transferred onto a fresh 6% food vial at the same time of day, daily, for 10 days (Supplementary Figure S1). Daughters and sons that died during the egg lay were excluded from the analysis. A summary of the numbers of flies that were not included in the analysis are included in Supplementary Table S4 (Supplementary Materials).

Sample sizes (number of single-fly egg-laying vials) for each treatment were: 1) young mother: young father (daughters n = 18, sons n = 28); young mother: old father (daughters n = 23, sons n = 24); old mother: young father (daughters n = 25, sons n = 28); and old mother: old father (daughters n = 23, sons n = 19) (Supplementary Table S4). The number of eggs laid by each isofemale was counted daily to allow estimates of egg-to-adult viability, in addition to total offspring counts.

Offspring Counts

After 10 days of transfers onto fresh food, each isofemale was discarded from the vial and the offspring were allowed to eclose. After 14 days, all the offspring had eclosed and were counted daily. We counted the number of males and females, to provide resolution to investigate any sex ratio distortion effects over time and between treatments. None were detected (data not shown).

Statistics—MCMCglmm

We fitted zero-inflated poisson error structure to our count data models (egg number and total offspring number) because there were zero values in our data sets, which we wanted to model since they may be of biological nature. Zero values may also indicate that flies had not successfully mated to either male or female tester flies, or could represent random infertility among the age treatments. We fit a binomial error structure (“multinomial2”) in the model of the proportion of flies that survived from egg to adulthood (egg-to-adult survival) and used only female and male samples that were successful in producing at least one offspring. We analyzed egg count data, offspring count data and egg-to-adult viability using the (MCMCglmm) package (v 2.19) (Hadfield 2010). MCMCglmm applies Markov Chain Monte Carlo generalized linear mixed models (MCMCglmm) in a Bayesian framework. Using generalized linear models (glm), we identified there was over-dispersion in our count data (ratio of residual deviance/residual df) >1. To account for this, MCMCglmm applies an observation level random effect to deal with data over-dispersion. In some of the experimental treatments, there were not enough independent vials to source a single focal isomale or isofemale to have a target of ~25 vials per treatment (in the case of young mother: old father and old mother: old father). This was only evident in the focal age-treatment flies and not the standard 3-day-old male or female that was mated to the treatment fly. Each young mother: young father son or daughter fly was sourced from an independent vial.

To account for shared vial of origin statistically, we fitted the “vial of origin” of the treatment flies as a random effect. In all models the factors: 1) mother’s age, 2) father’s age and 3) their interaction were fitted as fixed effects. The old age class was 45 days old and the young age class was 3 days old. Vial of origin of the experimental fly was the random effect. Results of zero-inflated poisson models for the egg and offspring count data are reported in the main text. We also conducted an analysis on the same count data using zero-truncated models and these are reported in the Supplementary Materials.

Prior Specification

MCMCglmm uses a Markov Chain Monte Carlo algorithm with an inverse Wishart before fit generalized linear mixed models. We applied the default MCMCglmm prior shape for the zero-inflated poisson models, and an inverse gamma prior (prior = list(R = list(nu = 0.002, V = 1)), G = list(G1 = list(nu = 0.002, V = 1))) for the egg-to-adult viability assay.

Markov Chain Parameters

Prior estimation of the model fixed and random effects were based on Markov chains of 600 000 iterations, with a 150 000 iteration burn-in and a thinning interval of 50. This resulted in a posterior estimated based on 9000 samples from the chain. We report posterior means, posterior modes, the highest posterior density (95% credible interval [CI]), number of effective samples and the associated pMCMC value for each fixed effect. Fixed effects were
considered statistically significant if the 95% CIs excluded zero, with the associated $P$-value ($p_{MCMC}$) being <0.05.

Model Convergence Checking

Models were checked for convergence by 2 methods: 1) visual inspection of the model traces, and 2) using the Gelman–Rubin statistic (Gelman and Rubin 1992) implemented in the [coda] R package (Plummer et al. 2006). Visual inspection of traces of the sampled posterior revealed there were no cases of reducible chains (e.g., chains did not get stuck in regions of the parameter space). The Gelman–Rubin statistic was calculated using the [gelman.diag] command in R. We compared the between- and within-chain posterior distribution variance with 3 independent MCMC chains of the same model (600 000 iterations; burn-in: 150 000; thinning interval: 50). Chain convergence is evident when the potential scale reduction factor (PSR) is <1.1 (Gelman and Rubin 1992). The PSR was exactly 1.0 in all analyses and we were therefore confident the chains had converged. In Table 1 and Supplementary Table S1, we report the results of the first of the 3 chains in each analysis.

Data Availability

In accordance with the *Journal of Heredity* data archiving policy (Baker 2013), we have deposited the primary data underlying the analyses as follows: Phenotype data are uploaded as Supplementary Materials.

Results

Numbers of Eggs

In focal daughters, there was a significant interaction between maternal and paternal ages ($P < 0.01$: Table 1(i)) on egg numbers, suggesting maternal age had statistically different effects when mated to different paternal age classes (and vice versa). Focal daughters with younger mothers produced greater numbers of eggs overall (Figure 1A). These results are largely influenced by a distinct egg laying phenotype in daughters with 2 old parents (see below and Figure 2A).

In focal sons, age of parent either alone or in combination did not influence the number of eggs laid by tester females (Table 1(iv); Figure 1B).

Table 1. Trans-generational age effects on offspring (F$_1$) fertility traits

| Focal sex | Trait | Term against the intercept | Posterior mode | Posterior mean | Lower 95% CI | Upper 95% CI | Effective samples | pMCMC |
|-----------|-------|---------------------------|----------------|---------------|-------------|-------------|-------------------|-------|
| Daughters | (i) Total egg number | Intercept | 4.843 | 4.862 | 4.639 | 5.091 | 9475 | <0.0001 |
|           |       | Mother age (young) | 0.433 | 0.421 | 0.108 | 0.734 | 9000 | 0.01 |
|           |       | Father age (young) | 0.441 | 0.403 | 0.103 | 0.706 | 9649 | 0.01 |
|           |       | Mother age (young) × Father age (young) | −0.569 | −0.592 | −1.041 | −0.154 | 9000 | <0.01 |
| (ii) Total offspring number | Intercept | 4.158 | 4.172 | 3.943 | 4.395 | 7906 | <0.0001 |
|           |       | Mother age (young) | 0.601 | 0.614 | 0.310 | 0.915 | 8125 | <0.001 |
|           |       | Father age (young) | 0.554 | 0.604 | 0.322 | 0.904 | 9000 | <0.0001 |
|           |       | Mother age (young) × Father age (young) | −0.541 | −0.507 | −0.933 | −0.100 | 9651 | 0.02 |
| (iii) Egg-to-adult survival | Intercept | −0.343 | −0.348 | −0.667 | −0.009 | 8708 | 0.04 |
|           |       | Mother age (young) | 0.848 | 0.833 | 0.398 | 1.272 | 8508 | <0.001 |
|           |       | Father age (young) | 0.966 | 0.950 | 0.535 | 1.396 | 9779 | <0.0001 |
|           |       | Mother age (young) × Father age (young) | −0.153 | −0.294 | −0.895 | 0.310 | 9000 | 0.34 |
| Sons      | (iv) Total egg number | Intercept | 4.880 | 4.868 | 4.648 | 5.095 | 9000 | <0.0001 |
|           |       | Mother age (young) | 0.146 | 0.179 | −0.105 | 0.479 | 9305 | 0.23 |
|           |       | Father age (young) | 0.294 | 0.246 | −0.046 | 0.517 | 9000 | 0.09 |
|           |       | Mother age (young) × Father age (young) | −0.001 | −0.038 | −0.426 | 0.333 | 9000 | 0.85 |
| (v) Total offspring number | Intercept | 4.450 | 4.434 | 4.160 | 4.707 | 9000 | <0.0001 |
|           |       | Mother age (young) | 0.095 | 0.132 | −0.210 | 0.498 | 9000 | 0.46 |
|           |       | Father age (young) | 0.368 | 0.375 | 0.054 | 0.731 | 8709 | 0.03 |
|           |       | Mother age (young) × Father age (young) | −0.084 | −0.064 | −0.517 | 0.377 | 9000 | 0.79 |
| (vi) Egg-to-adult survival | Intercept | 0.508 | 0.487 | 0.065 | 0.918 | 9000 | 0.03 |
|           |       | Mother age (young) | 0.148 | 0.161 | −0.374 | 0.709 | 9000 | 0.55 |
|           |       | Father age (young) | 0.415 | 0.422 | −0.071 | 0.982 | 9000 | 0.11 |
|           |       | Mother age (young) × Father age (young) | −0.411 | −0.278 | −0.963 | 0.440 | 9000 | 0.43 |

Three traits: (i, iv) total egg number; (ii, v) total offspring number; and (iii, vi) egg-to-adult survival of focal individuals were tested using generalized linear mixed models (glmm) implemented in the [MCMCglmm] R package (Hadfield 2010). Results of the Bayesian analyses are shown for each sex. Error distributions were modeled as “zero-inflated poisson,” “zero-inflated poisson,” and binomial (“multinomial2”), respectively. The random effect was modeled as the vial of origin. Markov chains were run for 600 000 iterations with burn-in: 150 000, and thinning interval: 50. Any reduction in sampling is also shown and significance was judged as 95% CIs excluding zero and $p_{MCMC}$ values <0.05 (bold).
There was a significant interaction between mother’s and father’s age on total offspring in focal daughters (Table 1(ii); Figure 1C). The combination of 2 young parents was associated with the greatest offspring numbers, and 2 old parents conferred the lowest offspring numbers, across all 4 age treatments. The effects of father’s age are conditional on the age of the mother (and vice versa).

In focal sons, only father’s age was associated with the number of offspring produced (Table 1(v); Figure 1D). Males with younger fathers produced statistically greater numbers of offspring than males with older fathers. Interestingly, mother’s age and its interaction with father’s age were not associated with total offspring numbers, consistent with no differences in egg numbers.

**Egg-to-Adult Survival**

In focal daughters, there was a significant large effect of both mother and father age on egg-to-adult survival. Younger mothers and younger fathers conferred increased egg-to-adult viability (Table 1(iii) and Figure 1E). There was no interaction of maternal and paternal age ($P = 0.34$), suggesting aged parents were influencing egg-to-adult survival in the same way, regardless of the age of the other parent.
In contrast, in focal sons there were no significant effects of maternal ($P = 0.55$) or paternal age ($P = 0.11$), or their interaction ($P = 0.43$) on egg-to-adult survival (Table 1(vi); Figure 1F).

**Phenotypic Variation over Time**

All 3 phenotypes showed variation over the 10-day experiment. In daughters, we discovered a striking effect of having 2 old parents in egg laying, offspring production, and egg to adult viability. Figure 2 describes these relationships. Using the same experimental protocol for egg laying and offspring production, we routinely observe *Drosophila* to exhibit an initial increase in egg and offspring production up to days 2–3, then a gradual decrease over time (Supplementary Figure S9 and Camus and Dowling 2018). Daughters with at least 1 young parent showed a typical trend in
egg and offspring production over time. By contrast, daughters with 2 old parents demonstrated a unique egg dumping behavior on day 1 and laid, on average, $>2.35 \times$ the number of eggs than the other 3 age class treatments (Supplementary Table S5). This high number of eggs translated to relatively high numbers of offspring on day 1, then a rapid drop until day 6, when egg and offspring numbers returned to similar levels to the daughters of the other 3 age categories. Egg-to-adult survival was routinely lower in offspring of daughters with 2 old parents across all time points (Figure 2).

In sons, we observed a more subtle pattern of phenotypic changes over time. Two old parents conferred the lowest (although nonsignificant) number of eggs on day 1 in sons; the opposite effect to that observed in daughters. Over time, the number of eggs in the old mother: old father cross increased to similar values to the other age treatments (Figure 2). In general, the egg-to-adult survival decayed at a constant rate in males and there was no appreciable difference between the 4 treatments, either over time, or as a cumulative 10 day total.

Estimates of Genetic Variation in Our Population

Using a trio analysis (Supplementary Materials), we estimate that there are extremely low levels of genetic variation in the flies used in this study. We estimate that between 51 (conservative estimate) and 266 (relaxed thresholds) are found in the RNA-seq libraries of the same genotypes used in the present study, corresponding with approximately 0.51 and 2.66 mutations per generation in the transcriptome. Given the number of nucleotides in the transcriptome that were shared across all 3 libraries, we estimate the mutation rate of our fly line to range from $2.93 \times 10^{-9}$ and $1.53 \times 10^{-8}$ mutations per site per generation.

Discussion

We found that fertility of daughters and sons is affected by maternal and paternal age in different ways. Reproductive fitness measures in daughters were influenced to a large degree by mother and father age, which was associated with a unique egg dumping phenomenon. Parental age effects in males were restricted to father’s age, and mother’s age only influenced daughter reproductive fitness.

Another study on trans-generational effects in Drosophila (Nystrand and Dowling 2014) found that daughter reproductive output (number of eclosed offspring over 4 days) was sensitive to a strong interaction between maternal age and paternal age. This finding was in a different direction to that observed in the current study. There are several possible reasons why the results differ between studies. The young and old age classes in Nystrand and Dowling’s study (Nystrand and Dowling 2014) were 4 and 14 days, respectively, while in the current study these ages were 3 and 45 days. The age classes in the current investigation are more protracted and likely influence different physiological and/or genetic differences in the offspring (particularly in offspring of older parents).

Measures of reproductive fitness across 4 days may not be directly comparable (estimated as $R^2 = 0.82$ in Nystrand and Dowling 2014) with measures over longer durations (10 days in the present study). In the present study, we found a similar strong positive correlation between offspring production in days 1–4, and the total offspring over 10 days ($r = 0.91$, $df = 184$, $t = 15.31$, $P < 0.0001$). The study design of Nystrand and Dowling (Nystrand and Dowling 2014) also incorporated an immune challenge factor into their experiment, which was included in significant higher order interaction effects, thus influencing the results of mother and father age as first-order effects. Nevertheless, significant age effects have been observed in both studies, suggesting aged Drosophila parents influence offspring fitness, even between young and old flies with only 10 days age difference (Nystrand and Dowling 2014). Interestingly, there are qualitative differences between these studies. Nystrand and Dowling (2014) found that daughters of younger fathers and young mothers had the lowest reproductive fitness overall, an opposite effect to the results we found in the present study; where daughters of young mothers and young fathers produced the highest overall offspring numbers.

Different genetic backgrounds were used between studies and this may explain the trans-generational effect differences. We found low levels of genetic variation in the Oregon R genetic stock used in this experiment based on exome-wide SNP data from a previous study (Mossman et al. 2016, 2017) (Supplementary Materials), however, we cannot rule out the opportunity for cohort selection on age-associated alleles in the old parent treatments. Theoretically, isoegenic lines do not exist after one generation of stock creation due to de novo (spontaneous) mutation. The rate of fitness associated mutations ($U$) is estimated in D. melanogaster to be in the order of 1.2 per generation per diploid genome (Haag-Liautard et al. 2007). The Oregon R stock used in this study was approximately 100 generations old and this estimate suggests 120 fitness-associated mutations may be segregating in our Oregon R stock population. Other estimates of the spontaneous mutation rate in Drosophila are $2.7 \times 10^{-9}$ to $3.5 \times 10^{-8}$ per site per generation (Keightley et al. 2009), suggesting at least $-105$ mutations would be present (mutation rate per site per generation × genome size $(1.5 \times 10^8$ bases) $\times 2$ ploidy $\times 100$ generations). We estimated the numbers of SNPs that were segregating in the transcriptome of our Oregon R population to be between 51 and 266 (Supplementary Materials). These analyses suggest our initially created stock was isoegenic or near isoegenic. The mutations that have accumulated since the culture was created are approximately equal to those predicted from spontaneous mutation.

During the 45 day aging of flies, it is a reasonable assumption that any deleterious age-associated alleles in the population would be purged, essentially purifying for neutral or beneficial alleles at those variable sites. However, expression of any allelic variation in crosses between aged flies would depend on the dominance characteristics of those mutations. Daughters and sons of older parents would be expected to harbor less genetic variation since their parents have been subject to age-selection in the previous generation. Parental mating between old and young flies would therefore likely increase the expression of heterosis, when compared with matings between 2 old parents. In contrast, 2 young parents would be expected to produce more genetically variable offspring. We found good evidence that overall reproductive fitness over the 10 days deteriorated with increasing numbers of old parents from 0 old parents (highest fitness: average offspring = 113.8 [females], 106.8 [males]) > 1 old parent (average offspring= 112.3 [females], 99.5 [males]) >2 old parents (lowest fitness: average = 57.7 [females], 77.9 [males]). This overall result is consistent with a cohort selection effect, however, we cannot rule out other nonmutually exclusive mechanisms of parent age-associated offspring fitness. We did not investigate lifespan and we therefore cannot make any assumptions about correlations between reproductive effects and lifespan in these flies (sensu Lansing effects: Lansing 1947).

The differences in fertility of the parental flies were expected, since old flies were in the declining phase of fertility and these produced fewer offspring for the experimental assay. However, the fitness effects we observed in their offspring (the focal experimental
phenotype (Santos et al. 1994; Hoffmann and Loeschcke 2006). We found the opposite effect whereby those offspring from the weakest cross with the lowest larval densities (old mother × old father treatment) also possessed the lowest fertility. This effect suggests larval densities are unlikely to have influenced our main results because we observe the opposite effect to that which would be expected under a conventional density dependent phenotype (Santos et al. 1994; Hoffmann and Loeschcke 2006). Our results may be slightly desensitized estimates of trans-generational age effects because we could not fully control possible larval density effects. We also acknowledge that treatment groups had small sample sizes.

The trans-generational effects of parental age on offspring fitness we found were both maternal and paternal in origin in daughters, while son fitness was sensitive to father’s age alone. We show that paternal age can modify daughter and son fitness. While maternal age effects have been suggested to modify the fitness of the adult insects via oviposition behavior [e.g., egg mass provisioning and its influence on the imaginal disc (Labeyrie 1988) and egg size and offspring development time (Vijendravarma et al. 2010)], paternal effects on daughter fitness are more difficult to pinpoint even though they may be large and have consequences for fitness. If paternal age effects are common there may be genetic benefits of females mating to younger males if the sperm haploid genome is higher quality in younger males.

Old fathers—poor fertility

There are a number of nonmutually exclusive reasons why flies with old fathers may have poorer reproductive performance. The integrity of the sperm nucleus of an old father may be compromised, either through oxidative stress (Wallace and Melov 1998), pre- and post-meiotic sperm senescence (reviewed in Pizzari et al. 2008), or possibly epigenetic modification (Rando 2012). All 3 possibilities could impair the quality of the embryo (and adult offspring) since half of the diploid genome of the son or daughter focal fly is aged and/or has been exposed to increased germ cell mitotic replication in the previous generation. Furthermore, increased organism age corresponds to increased chance of mutations to accumulate in the germline (Crow 2000), which could also modify the effects of genetic imprinting in the offspring, especially if the organism was to receive a “double dose” of aged haploid genomes.

Old, Old Parents

Fifty-five days old is likely to be beyond the physiological extreme of fertility experienced in natural settings, although surprisingly little is known about life history traits in wild fruit fly populations (Lachaise and Silvain 2004; Mansourian et al. 2018). That parental age influences offspring fertility, and this process may be mediated by gamete nucleus quality (Pizzari et al. 2008), it is possible that reproductive senescence may serve as an evolutionary filter to maintain high gamete quality in populations. One consequence of aged gametes in populations may be an influence on mating behavior in females (polyandry), where females mate with >1 male, to ensure high quality gametes from younger males are available for fertilization (Radwan 2003). Old fruit flies, with potentially compromised gamete integrity and increased mutation load (Crow 2000) are not as fit, and their offspring also have compromised fertility to the extent that a double dose of possibly poor quality haploid DNA from parents is associated with significantly reduced fertility.

Trans-generational Reproductive Plasticity?

Selection for age at reproduction has been shown to affect mating frequency in females (Sgrò et al. 2000), which can have a trans-generational effect on daughter lifetime reproductive success (LRS) (Priest et al. 2008). In the present study, we assumed the flies used in this experiment had been likely selected for early life reproduction, which is known to affect mating frequency; old females have lower mating frequency than young females (Miller et al. 2014). We do not make any assumptions about the quality of the offspring as a function of mating frequency, since this was not manipulated in any way in the experiment.

All daughters, regardless of their parent ages were mated to a standard 3-day-old male whose parents were both young. The effects we detected in daughters of 2 old parents are therefore likely to be the result of a male–female interaction involving components of the male ejaculate. Male Drosophila can plastically alter female reproductive investment and ~22% of this variation is of male genetic origin (Pischedda et al. 2011). Seminal fluid peptides transferred to females during copulation (Gromko et al. 1984) [e.g., sex peptide (SP)] are known to exert large influence over female reproductive behavior in Drosophila (Wolfner 1997; Chapman 2001; Ram and Wolfner 2009; Avila et al. 2011), including suppressing mating receptivity and increasing egg-laying for several days after mating (Rubinstein and Wolfner 2013). Seminal proteins effect other physiological changes in females such as relaxation of oviduct musculature, along with increasing octopamine signaling which increases ovulation rate (Rubinstein and Wolfner 2013). One such seminal protein, ovulin (Acp26Aa), is transferred from males, interacts with the female reproductive tract and increases octopamine signaling thus increasing egg release from the ovary (Rubinstein and Wolfner 2013). Ovulation rate is maximized during the 24 h after mating as a result of ovulin transfer, and octopamine stimulation (Herndon
and Wolfner 1995), although these eggs show lower hatchability (Chapman et al. 2001). The amount of ovulin transferred to mated females is significantly lower than the amount transferred to virgin females (Sirot et al. 2011), however the influence of large-scale male age differences on ovulin transfer is unknown.

Egg laying by daughters of 2 old parents phenocopies a maximal ovulation rate, along with low egg-to-adult survival, and is observed to decline rapidly after 24 h, consistent with the effects of ovulin. We therefore hypothesize that transgenerational age effects are influencing the central nervous system of female *Drosophila* via overstimulation of octopaminergic and/or other neurotransmitter networks (Rodríguez-Valentin et al. 2006) or increasing sensitivity to seminal proteins during the first day postmating. As a result, daughters of 2 old parents dump large quantities of eggs, as observed in the present study. This hypothesis requires testing, but opens a new avenue of research on a potential role of parent age on the neurophysiology of daughter reproductive behavior.

**Concluding Remarks**

Whether trans-generational age effects are prevalent in human societies is a growing issue, since increasing numbers of humans are delaying the onset of parenthood (Sartorius and Nieschlag 2010). Further investigations in mammalian models are required to investigate whether there is a need for concern of human fertility being impacted by advancing parental ages, and what possibly genetic, epigenetic or environmental mechanisms may underpin such effects. When the consequences of aging are viewed as trans-generational phenomena, many potentially illuminating areas of research emerge.

**Supplementary Material**

Supplementary material is available at *Journal of Heredity* online.

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**Authors’ Contributions**

J.A.M. designed the study and analyzed the data. R.M.S.B., E.B., N.M., and J.A.M. conducted the egg and offspring counts. J.A.M. and D.M.R. wrote the article. All authors gave final approval for publication.

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**Data Accessibility**

The datasets supporting this article have been uploaded as part of the Supplementary Materials.

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