HAUS5 Is A Potential Prognostic Biomarker With Functional Significance in Breast Cancer

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Background: Breast cancer (BRCA) has become the most frequently appearing, lethal, and aggressive cancer with increasing morbidity and mortality. Previously, it was discovered that the HAUS5 protein is involved in centrosome integrity, spindle assembly, and the completion of the cytoplasmic division process during mitosis. By encouraging chromosome misdivision and aneuploidy, HAUS5 has the potential to cause cancer. The significance of HAUS5 in BRCA and the relationship between its expression and clinical outcomes or immune infiltration remains unclear.

Methods: Pan-cancer was analyzed by TIMER2 web and the expression differential of HAUS5 was discovered. The prognostic value of HAUS5 for BRCA was evaluated with KM plotter and confirmed with Gene Expression Omnibus (GEO) dataset. Following that, we looked at the relationship between the high and low expression groups of HAUS5 and breast cancer clinical indications. Signaling pathways linked to HAUS5 expression were discovered using Gene Set Enrichment Analysis (GSEA). The relative immune cell infiltrations of each sample were assessed using the CIBERSORT algorithm and ESTIMATE method. We evaluated the Tumor Mutation Burden (TMB) value between the two sets of samples with high and low HAUS5 expression, as well as the differences in gene mutations between the two groups. The proliferation changes of BRCA cells after knockdown of HAUS5 were evaluated by fluorescence cell counting and colony formation assay.

Result: HAUS5 is strongly expressed in most malignancies, and distinct associations exist between HAUS5 and prognosis in BRCA patients. Upregulated HAUS5 was associated with poor clinicopathological characteristics such as tumor T stage, ER, PR, and HER2 status. Mitotic prometaphase, primary immunodeficiency, DNA replication, cell cycle related signaling pathways were all enriched in the presence of elevated HAUS5 expression, according to GSEA analysis. The BRCA microenvironment’s core gene, HAUS5, was shown to be related with invading immune cell subtypes and tumor cell...
INTRODUCTION

Female breast cancer (BRCA) has surpassed lung cancer as the most frequent cancer, with an estimated 2.26 million new cases (11.7%) (1), and is the leading cause of cancer deaths among women globally (2). Because of advancements in early screening and the development of anti-cancer strategies, BRCA therapy has vastly improved, however, the recurrence and metastasis rate of BRCA remains high (3, 4). Due to tumor heterogeneity and complexity, existing biomarkers are limited in predicting BRCA prognosis (5). Therefore, it is urgent to explore new molecular biomarkers as prognostic indicators in the field of clinical research to enhance prognosis and guide individualized treatment strategies (6).

HAUS5 (Augmin like complex subunit 5), also known as Dgt5 or KIAA0841, is one of the eight subunits of the augmin complex (7). During cell division, it mainly participates in spindle assembly, centrosome integrity, and cytoplasmic division (8). Abnormal expression of HAUS5 can induce microtubule fragmentation of centrosome and increment of centrosome volume, leading to chromosome dislocation and functional deficits of bipolar spindle (9), which in turn may induce tumor formation. It suggests that HAUS5 may be involved in the onset and progression of breast cancer. However, there are currently inadequate research on the influence of HAUS5 on breast cancer. As a result, greater research into the relevance and function of HAUS5 in BRCA pathogenesis is required.

In the study, we investigated the link between HAUS5 mRNA expression and BRCA clinical characteristics, as well as the possible role of HAUS5 in BRCA patients’ prognosis and immune regulations. In vitro, the impact of HAUS5 on BRCA cells was investigated further. Our study highlights the potential carcinogenic role of HAUS5 in breast cancer. Targeting HAUS5 may be a promising prognostic and chemotherapy target in the future.

METHOD

Datasets and Data Preprocessing

TCGA data breast cancer expression profile, mutation, and clinical data were all downloaded using the function provided by TCGAbiolinks (10), which contained a total of 1092 samples, including 113 normal samples and 979 breast cancer samples. Cases with sufficient or missing data were deleted from subsequent data processing. BRCA patients were classified into low- and high-HAUS5 expression groups according to the median HAUS5 expression value. GSE21653 was downloaded from the GEO database and validated for survival analyses (11).

Data of protein interaction networks are collected from STRING databases. KEGG channel information from GSEA database (https://www.gsea-KEGG.org/gsea/index.jsp). The oncogene list was collected from the COSMIC (12) and OncKB (13) database, the immune-related gene list was collected from IMMPORT (14) database, and the gene information related to breast cancer was collected from CTD (15) database.

GO Enrichment Analysis

Pan cancer was analyzed by TIMER2 (tumor immune estimation resource, version 2) web (http://timer.cistrome.org/) and observed the expression difference of HAUS5 between tumor and adjacent normal tissues for the different tumors or specific tumor subtypes of the TCGA project. The protein expression of HAUS5 was obtained from the human protein atlas (HPA) (http://www.proteinatlas.org/) database.

Survival Analysis

To explore whether the expression of the HAUS5 gene is related to the prognosis of breast cancer, we divided the cancer samples into two groups with high and low expression according to the median expression value of HAUS5 and then drew the K-M curve. Both overall survival and breast cancer-specific survival were used as endpoints. The prognostic value of HAUS5 for BRCA was evaluated with KM plotter (https://kmplot.com/analysis/).

Demographic and Clinical Variables

The relationship between HAUS5 expression and clinical characteristics, including tumor status, lymph node status, distant metastasis, pathologic stage, histological type, PR status, ER status, HER2, and age was analyzed by using χ2 statistics.

Function and Pathway Analysis by Gene Set Enrichment Analysis (GSEA)

Differentially expressed genes (DEGs) between Low- and high-HAUS5 expression groups were identified by using the DESeq2 R
package (version 1.26.0). In this study, GSEA was performed using the ggplot2 R package (v 3.3.3) to demonstrate the significant functions and pathways between the two groups. The expression level of HAUS5 was used as a phenotype label. An adjusted p-value < 0.05, normalized enrichment score (|NES|) > 1, and false discovery rate (FDR) < 0.25 were considered as significant difference.

**TIMER Database Analysis**
To explore whether the expression of HAUS5 is related to the tumor immune microenvironment, we first used the ESTIMATE algorithm to estimate the content of stromal cells and immune cells in tumor samples. Secondly, CIBERSORT algorithm was used to evaluate the invasion level of 22 immune cells in the sample and to evaluate the difference in the invasion level of each immune cell between the high and low expression groups (http://timercistrome.org/) (16).

**TMB and Mutation Analysis**
The mutation data of HAUS5 was obtained from the cbioPortal (https://www.cbioportal.org/) web platform. In the study, we explored the genomic profiles of HAUS5 with a z-score threshold ± 1.5. TMB was obtained by calculating the number of tumor mutations per Mb in each sample (17), and the TMB threshold is 10 mut/Mb. The relationship between HAUS5 and TMB was analyzed. After comparing the TMB value between the two groups with high and low expression of HAUS5, we further analyzed the differences of gene mutations between the two groups of samples.

**PPI Network and KEGG/GO Analysis**
To further explore the functional role of HAUS5 in cancer, we collected 100 genes directly interacting with HAUS5 from five protein-interaction network databases: STRING (18) Mentha (19), BioGRID (20), HPRD (21), and IntAct (22). At the same time, to show the function of the HAUS5 gene more directly, we collected the list of genes related to breast cancer from the CTD database, the list of oncogenes from the COSMIC and OncKB database, and the list of immune-related genes from IMMPORT database. Finally, all the information was annotated into the interaction network, and Cytoscape (23) was used to visualize the gene interaction network. To further explore the functions of this interaction network module dominated by HAUS5, we conducted go and KEGG enrichment analysis on these genes.

**Cell Culture**
The human breast cancer cell line (MDA MB 231 and HCC1937) was obtained from Shanghai Cell Bank (Shanghai, China) and cells were cultured in DMEM high glucose and supplemented with 10% fetal bovine serum (HyClone), 100 units/ml penicillin (HyClone) and 100 μg/ml streptomycin (HyClone) at 37°C in a humidified chamber with 5% CO2.

**RNA Interference and Cell Counting Assay**
Lentiviral vector (Shanghai GeneChem Co., Ltd., Shanghai, China). Subsequently, 293T cells were co-transfected with lentiviral vector carrying the shHAUS5 or negative control shRNA and packaged plasmids. The lentivirus were then harvested and the virus titer was determined. In addition, lentiviral vectors carried the green fluorescent protein (GFP) gene to label tumor cells. After 24 h of transfection, the medium was changed and cultured at 37°C for another 72 h. The images of infected cells were observed under a phase-contrast and fluorescence microscope. Logarithmic growth phase HCC1937 and MDA-MB 231 cells infected with lentivirus carrying shHAUS5 or shCtrl, were seeded into plates and incubated at 37°C with 5% CO2 for up to 5 days. The cells were then counted. The experiments were performed in triplicate and repeated at least three times independently.

**Colony Formation Assay**
HCC1937 and MDA-MB 231 cells (1,000 cells/well) were seeded into 6-well plates. After adhesion cells were cultured in DMEM at 37°C for 14 days. During this period, the cells were washed with PBS every 3 days, fixed with paraformaldehyde (4%, 15 minutes), and stained with crystal violet (0.1%, 15 minutes) and the numbers of colonies with > 50 cells were counted. All experiments were repeated at least three times.

**Statistical Analysis**
We described the baseline characteristics of the patients and treatment using summary statistics. Differences between qualitative variables and continuous variables were analyzed using χ² statistics, t-test, and analysis of variance (ANOVA), respectively. Survival was estimated using the Kaplan–Meier method and compared between the different groups using the log-rank test. All the tests above were 2-tailed, and a p-value of less than 0.05 was considered statistically significant.

**RESULTS**

**HAUS5 Was Up-Regulated in Pan-Cancer and Has Prognostic Value in BRCA**
To explore the possible role of HAUS5 in carcinogenesis, we first analyzed the expression of the HAUS5 gene in 37 human cancers of TCGA using the TIMER database. As shown in Figure 1A, HAUS5 was significantly overexpressed in 14 cancers compared with normal samples, including BLCA, BRCA, CHOL, COAD, ESCA, HNSC, KIRC, KIRC, LIHC, LUAD, LUSC, PRAD, READ, and STAD. We compared the expression of the HAUS5 gene in cancer and normal samples, and it was illustrated in the boxplot in Figure 1B that HAUS5 is significantly overexpressed in cancers. The prognostic value of HAUS5 for BRCA was evaluated with KM plotter (Figure 1C) and confirmed with GSE21653 (Figure 1D). There was a significant difference in OS and DFS between the high-low expression groups of the HAUS5 gene, indicating that there was a correlation between the expression of this gene and the prognosis of patients.

**Clinical Correlation of HAUS5 in Breast Cancer**
Subsequently, we compared the correlation between the high and low expression group of HAUS5 and the clinical indicators
of breast cancer. As can be seen from Table 1, there were significant differences in tumor T stage, ER, PR and HER2 status between the high and low expression groups. These results also indicate the potential value of HAUS5 for clinical staging.

**Gene Set Enrichment Analysis (GSEA)**

To elucidate the biological functions of HAUS5, we analyzed the DEGs between the low- and high-expression HAUS5 groups based on the median HAUS5 expression value. We also performed a GSEA pathway analysis (Supplementary Table 1). The results showed that high HAUS5 expression was mainly enriched in mitotic prometaphase, primary immunodeficiency, DNA replication, cell cycle related signaling pathways (Figure 2A). At the same time, we completed the GO analysis (Supplementary Table 2). BP terms were associated with development, including “skin development”, “epidermis development”, “keratinocyte differentiation”, “keratinization”, and “cornification”. The top 5 CC terms were cell cytoskeleton related, such as “intermediate filament cytoskeleton”, “intermediate filament”, “keratin filament”, “cornified envelope”, and “Golgi lumen”. The top 5 MF terms were associated with “receptor ligand activity”, “transcription activator activity”, “arachidonic acid monoxygenase activity”, “structural constituent of the epidermis”, and “arachidonic acid epoxygenase activity” (Figure 2B).

**Analysis of HAUS5 Expression, Copy Number and Related Immune Microenvironment**

In many cancers, the mRNA expression level of HAUS5 was significantly increased, and the tumor tissue contains not only tumor cells, but also a variety of different types of cells, such as stromal cells, fibroblasts, and immune cells, which together constitute the tumor microenvironment. As shown in Figure 3A, the stromal cell score in the group with high HAUS5 expression was significantly lower than that in the group with low HAUS5 expression (P<0.001). Many cancer studies had confirmed that stromal cells play an important role in the development, metastasis and drug resistance of tumors (24). However, there was no difference in immune scores between the high and low expression groups, which may be due to the different composition proportions of different immune cells between the two groups, leading to the insignificant difference. Therefore, in order to further explore the relationship between HAUS5 and tumor immune cells, we used different immune invasion algorithms to calculate the invasion levels of different immune cells.

Figure 3B shows that the infiltration levels of CD8+ T Cell, CD4+ T Cell, Macrophage and Neutrophil change with the change of HAUS5 copy number. We had also constructed a stacked bar chart in the Supplementary Figure 1, which showed the different relative proportions somatic Copy Number Alteration (sCNA) status of HAUS5 in all TCGA cancer types. As can be seen from Figure 3C, Macrophage, CD4+ T Cell and CD8+ T Cell showed significant difference in immune infiltration levels among HAUS5 expression groups (P<0.001). However, there was no difference between the B cell, Myeloid Dendritic cell, and Neutrophil, which confirmed our above conjecture.

According to the Figure 3D boxplot, B cells naive, Macrophages M0, Macrophages M2, Mast cells resting, NK cells activated, Plasma cells, and T cells follicular helper, the infiltration
TABLE 1 | Correlation between HAUS5 expression and clinical features.

| Variables                  | n=978 | HAUS5 | p value (Chi-squared test) |
|----------------------------|-------|-------|---------------------------|
| Age (years)                | 978   |       |                           |
| <50                        | 256   | 132   | 26.99%                    |
| ≥50                        | 722   | 357   | 73.01%                    |
| Tumor status (T)           |       |       |                           |
| T1                         | 250   | 106   | 21.68%                    |
| T2                         | 566   | 300   | 61.35%                    |
| T3                         | 126   | 68    | 13.91%                    |
| T4                         | 33    | 13    | 2.66%                     |
| unknown                    | 3     | 2     | 0.41%                     |
| Lymph node status (N)      |       |       |                           |
| N0                         | 469   | 246   | 50.31%                    |
| N1                         | 312   | 156   | 31.90%                    |
| N2                         | 108   | 44    | 9%                        |
| N3                         | 72    | 35    | 7.16%                     |
| unknown                    | 17    | 8     | 1.64%                     |
| Distant metastasis (M)     |       |       |                           |
| M0                         | 804   | 390   | 79.75%                    |
| M1                         | 20    | 11    | 2.25%                     |
| unknown                    | 154   | 88    | 18%                       |
| Cancer stage (C)           |       |       |                           |
| I                          | 161   | 73    | 14.93%                    |
| II                         | 554   | 296   | 60.53%                    |
| III                        | 223   | 103   | 21.06%                    |
| IV                         | 18    | 9     | 1.84%                     |
| unknown                    | 22    | 8     | 1.64%                     |
| ER Status                  |       |       |                           |
| Negative                   | 216   | 135   | 27.61%                    |
| Positive                   | 725   | 338   | 69.12%                    |
| unknown                    | 37    | 16    | 3.27%                     |
| PR Status                  |       |       |                           |
| Negative                   | 312   | 180   | 36.81%                    |
| Positive                   | 626   | 291   | 59.51%                    |
| unknown                    | 40    | 18    | 3.68%                     |
| HER2 Status (R)            |       |       |                           |
| Negative                   | 503   | 254   | 51.94%                    |
| Positive                   | 141   | 48    | 9.82%                     |
| unknown                    | 334   | 187   | 38.24%                    |

*p <= 0.05, **p <= 0.01, ***p <= 0.001, ****p <= 0.0001.

ER, estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor receptor 2; TNM, tumor, node, metastasis.

FIGURE 2 | GSEA analysis results. (A) Enrichment of genes in the representative pathways by GSEA function analysis. (B) GO and KEGG analysis of DEGs in low- and high- HAUS5 expression samples.
levels of the seven kinds of immune cells were significantly different between the high and low expression groups.

The above results have confirmed the correlation between the HAUS5 gene and stromals and immune cells in the immune microenvironment. The expression or copy number variation of HAUS5 gene will affect the tumor microenvironment and may play an important role in the occurrence, development, metastasis, and immune response of cancer.

TMB Analysis
We used cBioPortal (https://www.cbioportal.org/) to analyze the HAUS5 expression and its mutation in BRCA. As was shown in Figures 4A, C, a relatively high mutation rate of HAUS5 was observed in BRCA patients. In the 1084 sequenced BRCA patients, the genetic alteration was found in 32 BRCA patients and the mutation rate was 3%.

Although PD-1/PD-L1 immune checkpoint blocking therapy has been approved for the treatment of a variety of malignancies, only a small number of patients benefit from treatment (25). How to screen out patients who can benefit from the massive number of patients is particularly important. TMB is an emerging biomarker for the prediction of immune efficacy. Due to the High mutation load of a tumor, patients with TMB High produce a lot of antigens in the body and have a more adequate response to immunotherapy, with greater clinical benefits. Therefore, we continue to analyze the relationship between HAUS5 and TMB. As can be seen from the violin diagram in Figure 4B, TMB in the low expression group was significantly higher than that in the high expression group (P<0.001), which further emphasized the correlation between the HAUS5 gene and immunity.

15 genes with the significant difference in mutation frequency was shown in the forest map (Figure 4D, P<0.01). The mutation frequency of TP53, FANCD2, DAB1 and UNC5D genes in the high expression group was significantly higher than that in the low expression group. The mutation frequency of GATA3, ANK3, CDC42BPA and AFF2 genes in the low expression group was significantly higher than that in the high expression group. CDC42BPA binds to the cell cycle division protein CDC42 as a downstream effector (26). CDC42 can play an important role as a gene for targeted therapy of breast cancer (27, 28). GATA3 is a member of the transcription factor GATA family and can be used as a molecular marker of breast cancer (29). Somatic mutation of GATA3 is associated with clinicopathological features and expression of TCGA in breast cancer patients (30). FANCD2 (31), DAB1 (32) and ANK3 (33) are all supported by literature to be associated with breast cancer.

HAUS5 Interaction Network and Functional Analysis
Among the 100 genes directly interacting with HAUS5 (Supplementary Table 3), gray refers to the genes not annotated by the above database, while the genes in other colors are all breast cancer-related genes, including 14 pink genes, 4 blue genes, and purple genes, which are both cancer genes and immune-related genes in Figure 5A. More or less, these key genes have been proved to be closely related to breast cancer by relevant studies. For example, studies have shown that GSK3B gene is a potential drug target of triple negative breast cancer, which can regulate epithelial mesenchymal transformation and tumor stem cell characteristics (34). This result also indicates that HAUS5 may play an important role in the occurrence and development of cancer or the immune response of tumor, and can be used as a potential target.

We performed a GO analysis (Supplementary Table 4), and Figure 5B shows the top 5 enrichment results respectively. Many of these biological processes are closely related to cell division, for example, organelle fission, nuclear division, sister chromatid segregation. The molecular function enrichment involved in chemotaxis, such as tubulin binding, microtubule binding, motor activity, chemokine activity and histone kinase activity. The cellular component enrichment involved in spindle and centromeric region. In addition, pathway information collected in KEGG database was used for functional enrichment, with a total of 5 pathways shown in Figure 5B including cell cycle, cellular senescence, oocyte meiosis, PPAR signaling pathway. These pathways are also closely involved in cell cycle regulation. These results suggest that this functional module may play an important role in cell proliferation, division, and cell cycle regulation.

Knockdown of HAUS5 Inhibits BRCA Cells Progression In Vitro and the Protein Expression of HAUS5 in HPA
Immunohistochemistry-based antibody-specific staining scores in breast tumors were obtained from the Human Protein Atlas, in which tumor-specific staining was divided into four grades: high, medium, low and undetected (Figures 6A–D). To explore the oncogenic role of overexpression HAUS5 in BRCA, HCC1937 and MDA-MB-231 cells were transfected with sh-HAUS5 or sh-NC and performed colony formation assay (Figures 6E–H) and fluorescence cell counting (Figure 7). The findings showed that knockdown of HAUS5 significantly suppressed the proliferation ability of BRCA cell lines.

DISCUSSION
In this study, we first found that HAUS5 mRNA overexpressed in breast cancer. Meanwhile, HAUS5 mRNA expression also plays a huge role in the diagnosis of BRCA. In addition, by mining the clinical value of HAUS5 mRNA expression in different clinical variables, including tumor status, lymph node status, distant metastasis, pathologic stage, histological type, PR status, ER status, HER2. Besides, HAUS5 mRNA expression predicted poor prognosis and served as an independent prognostic factor for overall survival.

To further analyze the function of HAUS5, we conducted immune microenvironment analysis and found that there was a significant difference in the proportion of stromal cells between high and low expression groups. Although there was no difference in immune score, the infiltration level of different
cells was significantly correlated with the copy number of HAUS5 from the point of view of immune cell infiltration level. The expression of HAUS5 also affects the infiltration level of different immune cells. Moreover, there were significant differences in TMB between high and low expression groups. TMB has been proved to be related to the response rate of the PD-1 antibodies in many previous studies (35). The higher the TMB is, the higher the PD-1 antibody.
response rate (36). This indicates that the high expression of this gene may cause breast cancer patients to have no immune response to the PD-1 antibody, which may be a prognostic marker of the PD-1 antibody. In addition, the copy number variation of the gene is also related to the infiltrating immune cells in the tumor tissue (37). There are fewer infiltrating immune cells in the tumor tissue with increased or decreased copy number, indicating that the copy number variation of the gene may affect the immune cell infiltration, which needs to be further studied. On the other hand, the gene may affect the expression of chemokine receptors in tumor tissue. Therefore, the different changes of immune cell infiltration caused by affecting the expression of chemokine receptors may be one of the mechanisms of copy number variation of the gene affecting immune cell infiltration. The lower the transcription level of the gene, the higher the infiltration of CD8+ T cells. It may be that the low expression of the gene is related to the increase of TMB and produces more new tumor antigens. Therefore, it can recruit newer antigen specific CD8+ T cells to infiltrate into tumor tissues and produce killing effects.

Subsequently, we analyzed the difference in mutation frequency between the high and low expression groups, and found that the mutation frequency of TP53 and other genes in the high expression group was significantly higher than that in the low expression group, while the mutation frequency of GATA3 and other genes in the low expression group was higher. Therefore, the expression level of HAUS5 is related to the level of immunity and mutation of tumor cells.
To further analyze the function of HAUS5, we collected 100 genes directly interacting with HAUS5 from the protein interaction network database, and jointly constructed an interaction network module dominated by HAUS5 genes. From the interaction network diagram, we can see that several oncogenes and immune-related genes directly interact with HAUS5, and many genes have been supported to be related to breast cancer, indicating that the HAUS5 gene we studied may have a very important function in breast cancer. Subsequently, we conducted GO and KEGG enrichment to explore the function of this module. From the results of our enrichment analysis, it can be easily seen that this module plays an important role in the regulation of cell cycle.

In *in vitro* study, we also investigated the role of HAUS5 in the proliferation of BR cells. We found that the knockout of HAUS5 suppressed the growth of breast cells by fluorescence cell counting and colony formation assay. However, there is some limitation in this study. For example, it might be interesting to study HAUS5 in breast cancer cells at the single cell level (38, 39). In addition, identifying drugs...
targeting HAUS5 might push the results into clinical practice (40). Finally, we need more in vitro and in vivo experiment to validated the exact role of HAUS5 in BRCA and how does it work.

CONCLUSION

HAUS5 can be used as a potential marker and deserves further analysis to explore its role in the process of tumor occurrence, development, metastasis, and differentiation associated with immune infiltration from the experimental level.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

AUTHOR CONTRIBUTIONS

CY and YH conceived and designed the study. ZH, JY, WQ, and JH performed the experiments. ZH and ZC analyzed the data.
ZH and JH wrote the manuscript. All authors have read and approved this manuscript.

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