Complement 3 and the Prognostic Nutritional Index Distinguish Kawasaki Disease from Other Fever Illness with a Nomogram

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Abstract: Objective: This study aimed to establish a model to distinguish Kawasaki disease (KD) from other fever illness using the prognostic nutritional index (PNI) and immunological factors. Method: We enrolled a total of 692 patients (including 198 with KD and 494 children with febrile diseases). Of those, 415 patients were selected to be the training group and 277 patients to be the validation group. Laboratory data, including the neutrophil-to-lymphocyte ratio (NLR), the platelet-to-lymphocyte ratio (PLR), the prognostic nutritional index (PNI), and immunological factors, were retrospectively collected for an analysis after admission. We used univariate and multivariate logistic regressions and nomograms for the analysis. Result: Patients with KD showed significantly higher C3 and a lower PNI. After a multivariate logistic regression, the total leukocyte count, PNI, C3, and NLR showed a significance (p < 0.05) and then performed well with the nomogram model. The areas under the ROC in the training group and the validation group were 0.858 and 0.825, respectively. The calibration curves of the two groups for the probability of KD showed a near agreement to the actual probability. Conclusions: Compared with children with febrile diseases, patients with KD showed increased C3 and a decreased nutritional index of the PNI. The nomogram established with these factors could effectively identify KD from febrile illness in children.

Keywords: Kawasaki disease; nomogram model; white blood cell; prognostic nutritional index; platelet-to-lymphocyte ratio; neutrophil-to-lymphocyte ratio; complement 3

1. Introduction

Kawasaki disease (KD), also known as mucocutaneous lymph node syndrome (MCLS), is a form of acute febrile and systemic vasculitis that commonly occurs in children under 5 years old [1]. The primary pathological changes of KD are systemic non-specific vasculitis involving small and medium arteries and the most serious complication is a coronary artery lesion (CAL) including artery aneurysms, coronary artery stenosis, thrombosis, myocardial infarction, and sudden death [2]. With an increasing number of patients in recent years, KD has become the main cause of acquired heart disease in children [3].

Both the etiology and pathogenesis of KD remain unknown but may be the result of combined effects from genetic heredity, infection (bacteria, mycoplasma, virus, COVID-19, fungus, etc.), and the immune response [4]. It is believed that certain pathogenic
microbial infections and an immune response imbalance may lead to KD [5,6]. Therefore, speculation has indicated that inflammatory markers may have the potential to identify KD and the disease prognosis [7]. Recent studies have shown that the peripheral blood neutrophil-to-lymphocyte ratio (NLR) and the platelet-to-lymphocyte ratio (PLR) were associated with the severity of KD and the involvement of CALs [8].

The serum level of albumin is a negative acute-phase protein marker that decreases during inflammation or malnutrition. Inflammation reduces the albumin concentration by lowering its rate of synthesis, causing leakage due to the increased permeability of the blood vessels, and is associated with higher catabolic rates [9]. The serum levels of albumin play an important role in diagnosing incomplete KD according to the American Heart Association (AHA) supplementary criteria and predicting IVIG-resistant KD [10,11]. Onodera’s prognostic nutritional index (Onodera’s PNI) is composed of a serum albumin value (ALB) and a peripheral blood lymphocyte count (TLC) with the formula PNI = ALB (g/L) + 5 × TLC (10⁹/L). This index was first proposed by Buzby and later established by Onodera [12–14]. The PNI is a scoring index used to assess the nutritional status of patients and predict the risk of surgery and the prognosis of a variety of malignancies [15]. The PNI has been found to be a potentially important predictor of disease activities and complications of autoimmune diseases [16]. Tai et al. reported that the PNI could be a candidate as an adjunctive predictor of a CAL as well as IVIG resistance. Together with a low PNI, factors such as intravenous immunoglobulin (IVIG) resistance, male gender, and platelet count contribute to high odds for predicting a CAL within six months of illness [17].

A KD diagnosis primarily depends on the clinical presentation and the exclusion of other clinically similar cases with known causes. The prompt administration of IVIG treatment can reduce the incidence of coronary artery aneurysms from 20–25% to 3–5% [10]. The aim of this study is to explore the clinical value of the PNI combined with immune factors in the identification of KD.

2. Materials and Methods

2.1. Study Participants

KD children hospitalized in Shenzhen Baoan Women’s and Children’s Hospital from August 2016 to July 2019 were enrolled in this study. Febrile children who were hospitalized on the same day were also enrolled as a control group. Patients with autoimmune diseases, sepsis, or incomplete data were excluded for a total of 51 KD patients and 105 fever controls. The clinical indicators were collected and compared between the two groups. The clinical diagnosis of KD was based on the revised diagnostic criteria of KD by the American Heart Association (AHA) in 2017. IVIG responsiveness was defined as the abatement of fever within 48 h after completing IVIG treatment and no return of fever (> 38 °C) for at least 7 days after with a marked improvement or normalization of the comorbid signs of inflammation. Incomplete KD was defined as those who had less than four symptoms and were finally diagnosed as KD. The clinical data of gender, age, weight, and clinical manifestations of the enrolled children were collected for the analysis. We also recorded the laboratory results including white blood cell (WBC), neutrophil, lymphocyte, platelet, albumin (ALB), immunoglobulin A (IgA), IgG, IgM, C₃, C₄, the neutrophil-to-lymphocyte ratio (NLR), the platelet-to-lymphocyte ratio (PLR), and the PNI for the analysis. The training group was selected 60% randomly from the entire sample studied.

2.2. Statistical Analysis

We used The statistical analyses and graphics were performed with IBM SPSS 13.0 (SPSS Inc, Armonk, NY) and R 3.5.1 (The R Foundation for Statistical Computing, Vienna, Austria) with the rms statistical packages. for the statistical analysis including a single factor analysis and a multifactor analysis, and expressed normal distribution measurement data as mean and standard deviation. An independent sample t-test was used to compare the two groups. The median and interquartile range (IQR) were used to describe the measurement
3. Results

3.1. Clinical Features

We included a total of 692 children in this study including 422 male children (60.98%) and 270 female children (39.02%). The KD group consisted of 118 boys (59.6%) and 98 girls (40.4%) with a median age of 21 (13–17) months. The febrile control group had 304 boys (61.54%) and 190 girls (38.46%) with a median age of 21 (11–41) months. Among them, 415 (60%) children were randomly selected as the training group and 295 (40%) as the verification group. The comparison of variables between the groups is shown in Table 1. No significant difference was observed between the training group and the verification group ($p > 0.05$).

| Variable          | Total (N = 692) | Training Group (N = 415) | Verification Group (N = 277) | p-Value |
|-------------------|-----------------|--------------------------|-----------------------------|---------|
| Age (month)       | 21 (11–40)      | 22 (12–40)               | 19 (11–40)                  | 0.366   |
| Male gender, N (%)| 422 (60.98)     | 248 (59.76)              | 174 (62.82)                 | 0.419   |
| Body weight (Kg)  | 12.48 ± 4.73    | 12.66 ± 4.88             | 12.21 ± 4.5                 | 0.215   |
| WBC (*10^9/L)     | 10.4 (7.22–14.85)| 10.2 (7.4–14.96)         | 11 (7.05–14.74)             | 0.955   |
| IgA (g/L)         | 0.59 (0.4–1.01) | 0.61 (0.41–1.03)         | 0.57 (0.37–0.96)            | 0.291   |
| IgG (g/L)         | 7.27 ± 2.31     | 7.39 ± 2.33              | 7.1 ± 2.28                  | 0.107   |
| IgM (g/L)         | 1.04 (0.79–1.32)| 1.04 (0.79–1.32)         | 1.03 (0.77–1.32)            | 0.766   |
| C3 (g/L)          | 1.29 ± 0.27     | 1.28 ± 0.25              | 1.29 ± 0.28                 | 0.532   |
| C4 (g/L)          | 0.4 ± 0.13      | 0.4 ± 0.13               | 0.41 ± 0.14                 | 0.401   |
| NLR               | 1.3 (0.68–2.44) | 1.31 (0.68–2.54)         | 1.27 (0.68–2.31)            | 0.371   |
| PLR               | 83.85 (58.01–124.29) | 83.07 (56.37–125.6)      | 84.98 (60.59–123)           | 0.576   |
| PNI               | 60.92 ± 16.02   | 60.84 ± 15.28            | 61.05 ± 17.09               | 0.866   |

WBC: white blood cell; IgA: immunoglobulin A; IgG: immunoglobulin G; IgM: immunoglobulin M; C3: complement 3; C4: complement 4; NLR: neutrophil-to-lymphocyte ratio; PLR: platelet-to-lymphocyte ratio; PNI: prognostic nutritional index.

3.2. Univariate Analysis

A single variable analysis of all variables in the training group revealed that gender, age, body weight, IgA, IgG, IgM, and C4 were not statistically significant between the KD group and the control group ($p > 0.05$). WBC, the PNI, NLR, PLR, and C3 demonstrated a significant difference between the KD patients and the febrile controls ($p < 0.05$) (Table 2).
### Table 2. Comparison between the Kawasaki disease patients and the febrile controls (the training group).

|                          | Febrile Controls (N = 295) | KD (N = 120) | p-Value |
|--------------------------|-----------------------------|--------------|---------|
| Age (month)              | 22 (11–41)                  | 22 (15–37)   | 0.537   |
| Male gender, N (%)       | 178 (60.34)                 | 70 (58.33)   | 0.706   |
| Body weight (Kg)         | 12.87 ± 5.39                | 12.16 ± 3.27 | 0.104   |
| WBC (*10^9/L)            | 9.1 (6.8–12.7)              | 14.36 (10.52–17.94) | <0.001 * |
| IgA (g/L)                | 0.59 (0.4–1.03)             | 0.64 (0.44–1.05) | 0.473   |
| IgG (g/L)                | 7.43 ± 2.28                 | 7.27 ± 2.46  | 0.530   |
| IgM (g/L)                | 1.04 (0.79–1.31)            | 1.05 (0.79–1.32) | 0.895   |
| C3 (g/L)                 | 1.21 ± 0.23                 | 1.44 ± 0.25  | <0.001 * |
| C4 (g/L)                 | 0.4 ± 0.12                  | 0.39 ± 0.13  | 0.343   |
| NLR                      | 1 (0.57–1.89)               | 2.7 (1.55–4.69) | <0.001 * |
| PLR                      | 76.86 (52.18–115.23)        | 99.49 (70.25–162.26) | <0.001 * |
| PNI                      | 62.41 ± 14.8                | 56.97 ± 15.8 | <0.001 * |

KD: Kawasaki disease; WBC: white blood cell; IgA: immunoglobulin A; IgG: immunoglobulin G; IgM: immunoglobulin M; C3: complement 3; C4: complement 4; NLR: neutrophil-to-lymphocyte ratio; PLR: platelet-to-lymphocyte ratio; PNI: prognostic nutritional index; * p < 0.05.

### 3.3. Multivariate Logistic Regression Analysis

Significant indicators of the univariate analysis including WBC, C3, NLR, PLR, and the PNI were included in the multivariate analysis to screen out the independent risk factors of KD and a nomogram was established based on the results of the multivariate analysis. The statistics showed that WBC, C3, NLR, and the PNI were independent risk factors of KD (Table 3).

### Table 3. Multivariate logistic regression analysis.

|                          | p-Value | OR       | 95% CI for OR |
|--------------------------|---------|----------|---------------|
| WBC (*10^9/L)            | < 0.0001| 1.201    | 1.121–1.288   |
| C3 (g/L)                 | < 0.0001| 22.631   | 6.867–74.585  |
| NLR                      | 0.025   | 1.218    | 1.025–1.446   |
| PNI                      | 0.002   | 0.958    | 0.932–0.984   |
| Constant                 | < 0.0001| 0.006    |               |

WBC: white blood cell; C3: complement 3; NLR: neutrophil-to-lymphocyte ratio; PNI: prognostic nutritional index; OR: odds ratio; CI: confidence interval.

### 3.4. Scoring System for Predicting KD

The logistic regression results of the training group were used to make the nomogram (Figure 1). In the model, the maximum scores corresponding with each predictor were WBC (100 points), C3 (51 points), NLR (60 points), and the PNI (75 points). The occurrence probability of KD corresponding with the scores is shown in Table 4.
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Figure 1. The nomogram for the probability of Kawasaki disease. WBC: white blood cell; C3: complement 3; NLR: neutrophil-to-lymphocyte ratio; PNI: prognostic nutritional index. KD: Kawasaki disease.

Table 4. The incidence risk of Kawasaki disease corresponding with the total score.

| Total Points | Risk of KD |
|--------------|------------|
| 61           | 0.01       |
| 69           | 0.02       |
| 79           | 0.05       |
| 87           | 0.1        |
| 96           | 0.2        |
| 102          | 0.3        |
| 107          | 0.4        |
| 111          | 0.5        |
| 115          | 0.6        |
| 120          | 0.7        |
| 126          | 0.8        |
| 135          | 0.9        |
| 143          | 0.95       |
| 161          | 0.99       |
| 186          | 0.999      |
| 194          | 0.9995     |

3.5. Performance of the Nomogram

The ROC curve was applied to verify the nomogram results in the training group and the validation group and showed a good differentiation with an area under the curve of the ROC of 0.815 (95% confidence interval: 0.815–0.901) in the training group and 0.825 (95% confidence interval: 0.769–0.881) in the validation group (Figure 2). The nomogram was calibrated using the calibration curve [18]. A calibration curve of the nomogram for the training group and the validation group is presented in Figure 3, which shows that the prediction of KD by the nomogram agreed well with the actual probabilities in both groups. The calibration curves for the KD outcome in the two groups demonstrated almost no apparent departure from the fit with a good correspondence between the predicted outcome and the actual outcome.
Figure 2. Receiver operating characteristic (ROC) curve of the nomogram for the training group and the validation group.

Figure 3. The calibration curves for the nomogram of the training group and the validation group. The black dashed line is the reference line for where an ideal nomogram would lie. The red dotted line is the performance of the nomogram and the blue solid line corrects for any bias in the nomogram. (a) The training group; (b) the validation group.

3.6. ROC Curve and the Cutoff Value of C3 and the PNI

The area under the curve of the ROC of C3 was 0.761 (95% confidence interval: 0.72–0.801), 0.63 (95% confidence interval: 0.584–0.677) in the PNI, and 0.788 (95% confidence interval: 0.749–0.826) in the two variables combined (Figure 4). The cutoff value of C3 and the PNI was 1.38g/L and 52.03, respectively (Table 5).
4. Discussion

The accurate diagnosis of KD remains a challenge for clinicians because its clinical manifestations are often similar to or overlap with other febrile infectious diseases in children and no specific laboratory test is currently available to confirm the diagnosis. A recent study reported the existence of the overdiagnosis of KD [19]. The challenge for clinicians is to prevent the occurrence of coronary artery aneurysms (CAAs) based on the accurate diagnosis and precise treatment of KD. Therefore, establishing a prediction model to identify KD from other febrile infectious diseases is crucial. In this report, we reviewed the clinical data of 216 patients with KD and 394 patients with other febrile infectious diseases and established a new prediction model with a high accuracy.

Peripheral blood total WBC is one of the predictors in this model and it increases in the acute phase of KD. WBCs can be used as a non-specific inflammatory indicator in combination with clinical manifestations to predict KD [20]. WBCs may also be able to predict the severity of systemic inflammation and IVIG non-reactivity in KD patients [21]. Other studies have shown that a WBC count greater than $16\times10^9$/L is positively correlated with heart damage [22]. Therefore, although the specificity of WBCs is not high for KD, it is widely used in clinical practice and has a practical significance for the clinical diagnosis of KD [23].

In this model, NLR (the ratio of the neutrophil count to the lymphocyte count of the peripheral blood) is an important predictor for identifying KD. The immune response to inflammation includes neutrophils moving to the site of inflammation, releasing inflammatory cytokines, and activating T cells, which play a key role in the development of vascular inflammation. Lymphocytes are produced by lymphoid organs and play an important role in the body’s immune response; they can also be used as a marker of immune regulation. Therefore, NLR is a reflection between the inflammatory response and the immunity balance. A few studies have shown that the higher the NLR value, the heavier
the inflammatory response [24,25]. Recent studies have indicated that a high level of NLR is an independent influencing factor of IVIG resistance in KD [26].

Onodera’s PNI is an index reflecting the nutritional status. The PNI has been reported to be a strong indicator for predicting the prognosis of patients with malignant tumors and has been widely used in predicting the prognosis, postoperative complications, and quality of life of a variety of tumors [27,28]. The PNI was found to be a novel surrogate independent predictor for IVIG-resistant KD according to a recent study [29]. In KD patients, the ALB levels were significantly lower than those of the febrile control group and even lower in KD with CAL formation [30].

Recent studies have reported that a reduced lymphocyte count can serve as an independent predictor for IVIG resistance in KD [31]. Onodera’s PNI score is calculated based on these two indicators of lymphocytes and albumin and can reflect the nutritional status and immune function. In this study, Onodera’s PNI was an important predictor for distinguishing KD from other febrile diseases.

The plasma level of C3 in the KD group was significantly higher than in the febrile controls and was also one of the important indicators for distinguishing KD from the controls. C3 is involved in the three complement pathways (classical, lectin, and alternative) and plays an important role in the innate immune response. Yan et al. found that, compared with a fever control group, the level of C3 was significantly higher in a KD group and it was higher in an IVIG-sensitive group compared with an IVIG-non-responsive group [7]. Dysregulation or overactivation of the complement system is the pathogenesis of vascular inflammation and aortic aneurysm formation [32,33]. However, few studies have addressed the complement pathway of KD [34,35]. Katayama et al. reported that a Ficolin 1 inhibitory antibody injection improved vasculitis of a KD mouse model, further suggesting that the lectin pathway may be involved in the pathogenesis of KD [36].

This study is a single-center retrospective study with a relatively small number of cases and a randomized controlled study with a larger sample of multiple centers is needed to further verify the value of the prediction model.

5. Conclusions

Our study demonstrated that a nomogram has a good prediction ability with WBC, NLR, C3, PNI, and other predictors. This report is the first to use C3 and the PNI as predictive factors to distinguish KD from febrile disease. This paper clarified the importance of C3 in KD and provided direction for further research on the pathogenesis of KD.

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Institutional Review Board Statement: This study was conducted according to the guidelines of the Declaration of Helsinki and approved by the ethics committee of Shenzhen Baoan Women’s and Children’s Hospital (protocol code IRB No. LLSC2020-07-32-01-KS in 2020.07.10).

Data Availability Statement: The dataset containing the results from this article is available from the corresponding author upon request.

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References
1. Burns, J.C. The riddle of Kawasaki disease. *N. Engl. J. Med.* 2007, 356, 659–661. [CrossRef]
2. Tsuda, E.; Hamaoka, K.; Suzuki, H.; Sakazaki, H.; Murakami, Y.; Nakagawa, M.; Takasugi, H.; Yoshibayashi, M. A survey of the 3-decade outcome for patients with giant aneurysms caused by Kawasaki disease. *Am. Heart J.* 2014, 167, 249–258. [CrossRef]
3. Gorczyca, D.; Postepski, J.; Olesinska, E.; Lubieniecka, M.; Lachor-Motyka, I.; Opoka-Winiarska, I.; Gruenpeter, A. The clinical profile of Kawasaki disease of children from three Polish centers: A retrospective study. *Rheumatol. Int.* 2014, 34, 875–880. [CrossRef]
4. Kuo, H.C. Preventing coronary artery lesions in Kawasaki disease. *Biomed. J.* 2017, 40, 141–146. [CrossRef]
5. Shulman, S.T.; Rowley, A.H. Kawasaki disease: Insights into pathogenesis and approaches to treatment. *Nat. Rev. Rheumatol.* 2015, 11, 475–482. [CrossRef] [PubMed]
6. Chen, K.Y.H.; Messina, N.; Germano, S.; Bonnici, R.; Freyne, B.; Cheung, M.; Goldsmith, G.; Kollmann, T.R.; Levin, M.; Burdner, D.; et al. Innate immune responses following Kawasaki disease caused and toxic shock syndrome. *PLoS ONE* 2018, 13, e0191830. [CrossRef]
7. Ding, Y.; Li, G.; Xiong, L.J.; Yin, W.; Liu, J.; Liu, F.; Wang, R.G.; Xia, K.; Zhang, S.L.; Zhao, L. Profiles of responses of immunological factors to different subtypes of Kawasaki disease. *BMC Musculoskelet. Disord.* 2015, 16, 315. [CrossRef] [PubMed]
8. Kawamura, Y.; Takeshita, S.; Kanai, T.; Yoshida, Y.; Nonoyama, S. The Combined Usefulness of the Neutrophil-to-Lymphocyte Ratio and Platelet-to-Lymphocyte Ratios in Predicting Intravenous Immunoglobulin Resistance with Kawasaki Disease. *J. Pediatr.* 2016, 178, 281–284.e1. [CrossRef] [PubMed]
9. Don, B.R.; Kayser, G. Serum albumin: Relationship to inflammation and nutrition. *Semin. Dial.* 2004, 17, 432–437. [CrossRef]
10. McCrindle, B.W.; Rowley, A.H.; Newburger, J.W.; Burns, J.C.; Bolger, A.F.; Gewitz, M.; Baker, A.L.; Jackson, M.A.; Takahashi, M.; Shah, P.B.; et al. Diagnosis, Treatment, and Long-Term Management of Kawasaki Disease: A Scientific Statement for Health Professionals From the American Heart Association. *Circulation* 2017, 135, e927–e999. [CrossRef]
11. Song, R.; Yao, W.; Li, X. Efficacy of Four Scoring Systems in Predicting Intravenous Immunoglobulin Resistance in Children with Kawasaki Disease in a Children’s Hospital in Beijing, North China. *J. Pediatr.* 2017, 184, 120–124. [CrossRef] [PubMed]
12. Wei, G.B.; Lu, Y.Y.; Liao, R.W.; Chen, Q.S.; Zhang, K.Q. Prognostic nutritional index predicts prognosis in patients with metastatic nasopharyngeal carcinoma. *Onco Targets Ther.* 2016, 9, 5955–5961. [CrossRef]
13. Onodera, T.; Goseki, N.; Kosagi, K. [Prognostic nutritional index in gastrointestinal surgery of malnourished cancer patients]. *Nihon Geka Gakkai Zasshi* 1984, 85, 1001–1005.
14. Buzby, G.P.; Mullen, J.L.; Matthews, D.C.; Hobbs, C.L.; Rosato, E.F. Prognostic nutritional index in gastrointestinal surgery. *Am. J. Surg.* 1980, 139, 160–167. [CrossRef]
15. Sakurai, K.; Tamura, T.; Toyokawa, T.; Amano, R.; Kubo, N.; Tanaka, H.; Muguruma, K.; Yashiro, M.; Maeda, K.; Ohira, M.; et al. Low Preoperative Prognostic Nutritional Index Predicts Poor Survival Post-gastrectomy in Elderly Patients with Gastric Cancer. *Ann. Surg. Oncol.* 2016, 23, 3669–3676. [CrossRef] [PubMed]
16. Correa-Rodriguez, M.; Pocovi-Gerardino, G.; Callejas-Rubio, J.L.; Fernandez, R.R.; Martin-Amada, M.G.; Ortego-Centeno, N.; Rueda-Medina, B. The Prognostic Nutritional Index and Nutritional Risk Index Are Associated with Disease Activity in Patients with Systemic Lupus Erythematosus. *Nutrients* 2019, 11, 638. [CrossRef] [PubMed]
17. Tai, I.H.; Wu, P.L.; Guo, M.M.; Lee, J.; Chu, C.H.; Hsieh, K.S.; Kuo, H.C. Prognostic nutritional index as a predictor of coronary artery aneurysm in Kawasaki Disease. *BMC Pediatr.* 2020, 20, 203. [CrossRef]
18. Alba, A.C.; Agorittas, T.; Walsh, M.; Hanna, S.; Iorio, A.; Devereaux, P.J.; McGinn, T.; Guyatt, G. Discrimination and Calibration of Clinical Prediction Models: Users’ Guides to the Medical Literature. *JAMA* 2017, 318, 1377–1384. [CrossRef]
19. Coon, E.R.; Wilkes, J.; Bratton, S.L.; Srivastava, R. Paediatric overdiagnosis modelled by coronary abnormality trends in Kawasaki disease of children from three Polish centers: A retrospective study. *Acta Paediatr.* 2018, 103, 937–941. [CrossRef]
20. Parthasarathy, P.; Agarwal, A.; Chawla, K.; Tofighi, T.; Mondal, T.K. Upcoming biomarkers for the diagnosis of Kawasaki disease: A review. *Clin. Biochem.* 2015, 48, 1188–1194. [CrossRef]
21. See, Y.M.; Kang, H.M.; Lee, S.C.; Yu, I.W.; Kil, H.R.; Rim, J.W.; Han, J.W.; Lee, K.Y. Clinical implications in laboratory parameter values in acute Kawasaki disease for early diagnosis and proper treatment. *Korean J. Pediatr.* 2018, 61, 160–166. [CrossRef] [PubMed]
22. Koyanagi, H.; Yanagawa, H.; Nakamura, Y.; Yashiro, M. Leukocyte counts in patients with Kawasaki disease: From the results of nationwide surveys of Kawasaki disease in Japan. *Acta Paediatr.* 1997, 86, 1328–1332. [CrossRef]
23. Kourtidou, S.; Skee, A.E.; Bruce, M.E.; Wren, H.; Mangione-Smith, R.M.; Portman, M.A. Kawasaki Disease Substantially Impacts Health-Related Quality of Life. *J. Pediatr.* 2018, 193, 155–163.e5. [CrossRef] [PubMed]
24. Guthrie, G.J.; Charles, K.A.; Roxburgh, C.S.; Horgan, P.G.; McMillan, D.C.; Clarke, S.J. The systemic inflammation-based neutrophil-lymphocyte ratio: Experience in patients with cancer. *Crit. Rev. Oncol. Hematol.* 2013, 88, 218–230. [CrossRef] [PubMed]
25. Cakmak, H.A.; Dingez Cakmak, B.; Abide Yayla, C.; Inci Coskun, E.; Erturk, M.; Keles, I. Assessment of relationships between novel inflammatory markers and presence and severity of preeclampsia: Epididymal fat thickness, pentraxin-3, and neutrophil-to-lymphocyte ratio. *Hypertens. Pregnancy* 2017, 36, 233–239. [CrossRef]
26. Wu, S.; Long, Y.; Chen, S.; Huang, Y.; Liao, Y.; Sun, Y.; Zhang, Q.; Zhang, C.; Yan, H.; Qi, J.; et al. A New Scoring System for Prediction of Intravenous Immunoglobulin Resistance of Kawasaki Disease in Infants Under 1-Year Old. *Front. Pediatr.* 2019, 7, 514. [CrossRef]
27. Kanda, M.; Mizuno, A.; Tanaka, C.; Kobayashi, D.; Fujiwara, M.; Iwata, N.; Hayashi, M.; Yamada, S.; Nakayama, G.; Fujii, T.; et al. Nutritional predictors for postoperative short-term and long-term outcomes of patients with gastric cancer. *Medicine* 2016, 95, e3781. [CrossRef]
28. Okada, I.; Shirahata, A.; Soda, H.; Saitou, M.; Kigawa, G.; Nemoto, H.; Sanada, Y.; Hibi, K. Significance of Onodera’s prognostic nutritional index for treating unresectable or recurrent colorectal cancer with chemotherapy. *Gan Kagaku Ryoho* 2012, 39, 231–235.

29. Li, G.; Xu, X.; Chen, P.; Zeng, R.; Liu, B. Prognostic value of pretreatment prognostic nutritional index in intravenous immunoglobulin-resistant Kawasaki disease. *Heart Vessels* 2021, 36, 1366–1373. [CrossRef]

30. Dominguez, S.R.; Friedman, K.; Seewald, R.; Anderson, M.S.; Willis, L.; Glode, M.P. Kawasaki disease in a pediatric intensive care unit: A case-control study. *Pediatrics* 2008, 122, e786–e790. [CrossRef]

31. Piram, M.; Darce Bello, M.; Tellier, S.; Di Filippo, S.; Boralevi, F.; Madhi, F.; Meinzer, U.; Cimaz, R.; Piedvache, C.; Kone-Paut, I. Defining the risk of first intravenous immunoglobulin unresponsiveness in non-Asian patients with Kawasaki disease. *Sci. Rep.* 2020, 10, 3125. [CrossRef] [PubMed]

32. Chimenti, M.S.; Ballanti, E.; Triggianese, P.; Perricone, R. Vasculitides and the Complement System: A Comprehensive Review. *Clin. Rev. Allergy Immunol.* 2015, 49, 333–346. [CrossRef] [PubMed]

33. Zhou, H.F.; Yan, H.; Bertram, P.; Hu, Y.; Springer, L.E.; Thompson, R.W.; Curci, J.A.; Hourcade, D.E.; Pham, C.T. Fibrinogen-specific antibody induces abdominal aortic aneurysm in mice through complement lectin pathway activation. *Proc. Natl. Acad. Sci. USA* 2013, 110, E4335–E4344. [CrossRef]

34. Biezeveld, M.H.; Geissler, J.; Weverling, G.J.; Kuipers, I.M.; Lam, J.; Ottenkamp, J.; Kuijpers, T.W. Polymorphisms in the mannose-binding lectin gene as determinants of age-defined risk of coronary artery lesions in Kawasaki disease. *Arthritis Rheumatol.* 2006, 54, 369–376. [CrossRef] [PubMed]

35. Okuzaki, D.; Ota, K.; Takatsuki, S.I.; Akiyoshi, Y.; Naoi, K.; Yabuta, N.; Saji, T.; Nojima, H. FCN1 (M-ficolin), which directly associates with immunoglobulin G1, is a molecular target of intravenous immunoglobulin therapy for Kawasaki disease. *Sci. Rep.* 2017, 7, 11334. [CrossRef] [PubMed]

36. Katayama, M.; Ota, K.; Nagi-Miura, N.; Ohno, N.; Yabuta, N.; Nojima, H.; Kumanogoh, A.; Hirano, T. Ficolin-1 is a promising therapeutic target for autoimmune diseases. *Int. Immunol.* 2019, 31, 23–32. [CrossRef] [PubMed]