CXCL9 as a Prognostic Inflammatory Marker in Early-Stage Lung Adenocarcinoma Patients

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Background: This study was performed to evaluate the value of inflammatory biomarkers in predicting the prognosis of early-stage (stage IA-IIB) lung adenocarcinoma.

Methods: Ten inflammatory biomarkers were tested with a Luminex bead-based assay in early-stage lung adenocarcinoma patients who underwent resection.

Results: A total of 152 early-stage lung adenocarcinoma patients were analyzed in this study. The mean patient age (SD) was 59.9 (9.4) years. In total, 58.6% of patients were females, and never smokers accounted for 84.0%. Lung adenocarcinoma patients with high CXCL9 levels had a 71% reduced risk of recurrence relative to patients with low CXCL9 levels (HR = 0.29, 95% CI: 0.13–0.64, p = 0.0021). After Bonferroni correction, CXCL9 remained significantly related to the risk of early-stage lung adenocarcinoma recurrence. Lung adenocarcinoma patients with high CXCL9 levels had an 80% reduced risk of death relative to patients with low CXCL9 levels (HR = 0.20, 95% CI: 0.05–0.78, p = 0.021), and those in the TCGA validation cohort were at a 29% reduced risk of death (HR = 0.71, 95% CI: 0.45–0.99, p = 0.044).

Conclusion: Our results demonstrate for the first time that the CXCL9 level is a protective factor for both disease-free survival (DFS) and overall survival (OS) in early-stage lung adenocarcinoma patients.

Keywords: CXCL9, inflammation, early stage, lung adenocarcinoma, prognosis

INTRODUCTION

Non-small cell lung cancer (NSCLC) accounts for approximately 85% of all lung cancers and is the leading cause of death from cancer worldwide (1). Adenocarcinoma is the most frequent subtype of NSCLC, and its incidence continues to increase (2–4). Lung cancer patients diagnosed in the early stages have a good prognosis, but approximately 30–55% of patients develop recurrence despite resection (5). Therefore, the identification of biomarkers able to predict which early-stage patients have a high risk for recurrence and death in subsequent years is urgently needed.

There is a growing body of evidence showing that chronic inflammation is a hallmark of lung carcinogenesis (6–8). Some broad investigations have also proven that inflammatory markers are associated with the risk of lung cancer (9–11). One of our previous studies evaluated the relationship between 10 inflammatory markers and early-stage lung adenocarcinoma risk (12), and the results showed that four inflammatory markers were significantly associated with the risk of
early-stage lung adenocarcinoma: CXCL13 (C-X-C motif chemokine ligand 13), CCL22 (C-C motif chemokine ligand 22), CXCL9 (C-X-C motif chemokine ligand 9), and IL-10 (interleukin 10). After Bonferroni correction, only CXCL13 and CCL22 were found to be independently related to the risk of early-stage lung adenocarcinoma.

However, few studies have used a broad panel to evaluate the prognostic value of inflammatory markers in early-stage lung adenocarcinoma. To fill this gap, the present study therefore aimed to investigate the association between these 10 inflammatory markers and the prognosis of early-stage lung adenocarcinoma patients with a median follow-up time of 60.6 months. The 10 inflammatory biomarkers were measured in all study participants, including CRP (C-reactive protein), CCL22, CXCL13, CXCL9, TNFRII (Tumor necrosis factor receptor II), IL-1b (interleukin 1 beta), IL-6 (interleukin 6), IL-10, IFN-r (interferon-gamma), and TGF-a (transforming growth factor alpha), and they were previously reported to be associated with lung cancer (13–18).

MATERIALS AND METHODS

Study Population

Patients who were histologically confirmed as having lung adenocarcinoma by lung resection according to the classification criteria of the World Health Organization (19) at Shanghai Chest Hospital between September 2013 and March 2015 were included in the present study. The pathologic stage was determined according to the International Association for the Study of Lung Cancer (IASLC) tumor-node-metastasis (TNM) classification, 8th edition (20). The inclusion criteria were as follows: (1) patients with a TNM stage IA to IIB; (2) available serum samples before diagnosis; and (3) complete follow-up data. In contrast, patients were excluded if they were missing baseline data. All the enrolled patients signed informed consent documents. This study was approved by the ethics committee of Shanghai Chest Hospital.

Follow-Up Assessment

Follow-up was performed annually after discharge at the outpatient department of our hospital or through a telephone

### Table 1 | Baseline characteristics of lung adenocarcinoma patients.

| Variables          | Non-recurrence cases | Recurrence cases | P-value |
|--------------------|----------------------|------------------|---------|
| Age (years), mean (SD) | 59.6 (9.6)           | 58.2 (10.7)      | 0.61    |
| Gender, N (%)       |                      |                  |         |
| Female             | 70 (64.2)            | 19 (44.2)        | 0.024   |
| Male               | 39 (35.8)            | 24 (55.8)        |         |
| Smoking status, N (%) |                |                  |         |
| Never smokers      | 94 (86.2)            | 34 (79.1)        | 0.28    |
| Ever smokers       | 15 (13.8)            | 9 (20.9)         |         |
| TNM stage, N (%)   |                      |                  |         |
| IA                 | 82 (75.2)            | 10 (23.3)        | <0.001  |
| IB                 | 11 (10.1)            | 6 (14.0)         |         |
| II                 | 16 (14.7)            | 27 (62.8)        |         |
| Adjuvant therapy   |                      |                  |         |
| Yes                | 21 (19.3)            | 13 (30.2)        | 0.14    |
| No                 | 88 (80.7)            | 30 (69.8)        |         |
| Biomarkers(pg/mL), mean (SD) |      |                  |         |
| CRP                | 2199847.0 (371273.9) | 22226112 (323723.1) | 0.97   |
| CXCL13             | 48.7 (25.6)          | 45.0 (27.0)      | 0.43    |
| CCL22              | 90.0 (48.1)          | 93.5 (42.1)      | 0.67    |
| CXCL9              | 31.0 (32.1)          | 26.9 (23.4)      | 0.39    |
| TNFRII             | 175.3 (81.6)         | 169.8 (83.5)     | 0.71    |
| IL-6               | 12.3 (7.6)           | 12.6 (7.0)       | 0.82    |

SD, standard deviation; N (%), number (percentage). P was calculated by the t-test or the Mann-Whitney U test or the Chi² test where appropriate. TNM stage, tumor-node-metastasis stage.

### Table 2 | Risk prediction of inflammation biomarkers for disease-free survival (DFS) of early stage lung adenocarcinoma.

| Biomarkers, pg/mL | Non-recurrence cases N (%) | Recurrence cases N (%) | HR (95% CI) | P* |
|-------------------|----------------------------|------------------------|-------------|----|
| CXCL9             | ≤7.8 7 (6.4)               | 8 (18.6)               | 1           | 0.0021 |
|                   | >7.8 102 (93.6)            | 35 (81.4)              | 0.29 (0.13–0.64) |    |
| CXCL13            | ≤21.4 15 (13.8)            | 7 (16.3)               | 1           | 0.33  |
|                   | >21.4 94 (86.2)            | 36 (83.7)              | 0.67 (0.30–1.52) |    |
| CCL22             | ≤77.7 55 (50.5)            | 16 (37.2)              | 1           | 0.27  |
|                   | >77.7 54 (49.5)            | 27 (62.8)              | 1.40 (0.76–2.56) |    |
| TNFRII            | ≤275.4 99 (90.8)           | 39 (90.7)              | 1           | 0.89  |
|                   | >275.4 10 (10.2)           | 4 (9.3)                | 1.10 (0.37–3.13) |    |
| IL-6              | ≤14.1 66 (60.6)            | 23 (53.5)              | 1           | 0.47  |
|                   | >14.1 43 (39.4)            | 20 (46.5)              | 0.80 (0.43–1.47) |    |
| CRP               | ≤888244.7 66 (60.6)        | 20 (46.5)              | 1           | 0.23  |
|                   | >888244.7 43 (39.4)        | 23 (53.5)              | 1.45 (0.79–2.65) |    |
| IL-10             | Undetectable 90 (82.6)     | 35 (81.4)              | 1           | 0.40  |
|                   | Detectable 19 (17.4)       | 8 (18.6)               | 1.39 (0.64–3.03) |    |
| IFN-r             | Undetectable 67 (61.5)     | 26 (60.5)              | 1           | 0.30  |
|                   | Detectable 42 (38.5)       | 17 (39.5)              | 1.38 (0.15–2.56) |    |
| TGF-a             | Undetectable 92 (84.4)     | 38 (88.4)              | 1           |       |
|                   | Detectable 17 (15.6)       | 5 (11.6)               | 0.87 (0.34–2.22) | 0.77  |

HR, hazard ratio; 95% CI, 95% confidence interval. P was adjusted for age, gender, smoking, TNM stage, and adjuvant therapy under multivariate Cox proportional hazards model.
interview or post mail if the patient failed to show at the scheduled time. The main endpoint was disease-free survival (DFS), defined as the time from lung resection to the date of disease relapse or death from any cause, whichever occurred first. Patients who did not experience any event were censored at the date of the last follow-up in April 2019. The secondary endpoint was overall survival (OS), defined as the time from surgery until the date of death or the last follow-up, whichever occurred first.

**Laboratory Methods**

The circulating levels of 10 inflammatory markers were measured in serum specimens collected at baseline (processed at 2,400–3,000 rpm for 15 min), and then the samples were frozen within 2 h of collection and stored at −80°C. Inflammatory markers were measured using a Luminex bead-based assay (12). Concentrations were calculated using a four- or five-parameter standard curve. All the samples were assayed in duplicate, and the detection results were averaged for analysis.

**Statistical Analysis**

Data are summarized as the mean ± standard deviation (SD) for continuous variables and as counts and percentages for categorical variables. Their differences were compared by the Wilcoxon rank-sum test, Student’s t-test, Pearson’s X2 test or Fisher’s exact test, where appropriate. A survival tree analysis was employed to determine the optimal cutoff point of each biomarker as described previously (21, 22) (STREE software, available at http://c2s2.yale.edu/software/stree/).

For inflammatory biomarkers, measurements below the lowest limit of detection (LLOD) were assigned a value of half the LLOD, as described previously (12, 13). Biomarkers in which the levels were over the LLOD in more than 25% of subjects were dichotomized by the optimal cutoff points determined by the survival tree analysis. Biomarkers in which the levels were over the LLOD in 10–25% of individuals were categorized as detectable and undetectable.

The impact of each biomarker on DFS or OS in lung adenocarcinoma patients was evaluated by a multivariate Cox proportional hazards model and is as expressed as the hazard ratio (HR) and its 95% confidence interval (CI) after controlling for age, sex, smoking status, TNM stage, and neoadjuvant therapy. The Kaplan–Meier (K–M) method and log-rank test were used to compare DFS and OS between patient groups defined by each dichotomized biomarker. Stratified analyses were performed separately according to age, sex, and smoking status. The findings were validated using mRNA expression and OS data from The Cancer Genome Atlas (TCGA) for lung adenocarcinoma (www.cbioportal.org; accessed July 2019).

All statistical analyses were completed with SPSS software version 22.0 for Windows (IBM SPSS, Inc., Chicago, IL, USA) unless indicated. Statistical significance was taken as a two-sided P < 0.05, and multiple testing was controlled by Bonferroni correction.
RESULTS

Patient Characteristics

A total of 152 early-stage lung adenocarcinoma patients were ultimately analyzed in this study, and the patient demographics are shown in Table 1. The mean patient age (SD) was 59.9 (9.4) years. Females accounted for 58.6% of the total population, and 84.0% were never smokers. According to TNM stage, 71.7% of patients were at stage I, and 28.3% of patients were at stage II. The median values of the inflammatory markers were comparable between the recurrence and non-recurrence groups (all \( p > 0.05 \)). The optimal cutoff points for CXCL9, CXCL13, CCL22, TNFRII, IL-6, and CRP were 7.8 pg/ml, 21.4 pg/ml, 77.7 pg/ml, 275.4 pg/ml, 14.1 pg/ml, and 888244.7 pg/ml, respectively. The median follow-up time was 60.6 months.

| Variables          | Non-recurrence cases N | Recurrence cases N | HR (95% CI) | \( P^* \) |
|--------------------|------------------------|--------------------|-------------|----------|
| Age < 60 Low/High  | 6/52                   | 6/28               | 0.34 (0.13–0.91) | 0.031    |
| Age ≥ 60 Low/High  | 1/50                   | 2/42               | 0.08 (0.15–0.46) | 4.3*10^-3 |
| Female Low/High    | 5/65                   | 4/15               | 0.18 (0.06–0.62) | 6.6*10^-3 |
| Male Low/High      | 2/37                   | 4/20               | 0.33 (0.11–1.00) | 0.051    |
| Never smokers Low/High | 5/89                   | 7/27               | 0.10 (0.04–0.26) | 3.0*10^-6 |
| Ever smokers Low/High | 2/13                   | 1/8                | 1.10 (0.09–13.60) | 0.96     |

*HR, hazard ratio; 95% CI, 95% confidence interval.

P was adjusted for age, gender, smoking, TNM stage, and adjuvant therapy under multivariate Cox proportional hazards model.

Association of Inflammatory Markers With DFS

In the multivariate Cox analyses, only CXCL9 was significantly associated with DFS after adjusting for age, sex, smoking status, stage, and adjuvant therapy (Table 2). Lung adenocarcinoma patients with high CXCL9 levels had a 71% reduced risk of recurrence relative to patients with low CXCL9 levels (\( HR = 0.29, 95\% CI: 0.13–0.64, p = 0.0021 \)). K–M analysis showed that lung adenocarcinoma patients with high CXCL9 levels had a significantly longer median DFS time than those with low CXCL9 levels (log-rank test \( p = 0.014 \); Figure 1A). While other markers, such as CXCL13, CCL22, and IL-10, did not show significant differences in DFS of lung adenocarcinoma patients between high and low groups (Figures 1B–D). After Bonferroni correction, CXCL9 remained significant, with a significance level of 0.5% (0.05/10, here 10 refers to 10 biomarkers under study).

The association of CXCL9 with DFS in lung adenocarcinoma patients was stratified by age, sex, and smoking status (Table 3). Elevated CXCL9 levels were found to predict a good prognosis in never smokers, females, and both young and old lung adenocarcinoma patients. The K–M analysis showed that increased CXCL9 levels were still significantly correlated with longer DFS in never smokers and young lung adenocarcinoma patients (Figures 2A,B).

Influence of CXCL9 Expression on OS and TCGA Validation

Lung adenocarcinoma patients with high CXCL9 levels had an 80% reduced risk of death (\( HR = 0.20, 95\% CI: 0.05–0.78, p = 0.021 \)) relative to patients with low CXCL9 levels (Table 4), although K–M analysis did not reach statistical significance (log-rank test \( p = 0.082 \); Supplement Figure 1).

After analysis of the TCGA database, 492 lung adenocarcinoma patients were enrolled in the validation cohort, and their characteristics are summarized in Supplement Table 1. Patients with high CXCL9 levels in the validation cohort had a
Our findings demonstrate for the first time that the CXCL9 level is as a protective factor for both the DFS and OS of early-stage lung adenocarcinoma patients. The application of CXCL9 as a predictive and prognostic biomarker of early-stage lung adenocarcinoma patients will be investigated in the future with large, well-designed, multicenter cohort studies, along with in vitro and in vivo functional experiments.

DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/Supplementary Material.

ETHICS STATEMENT

This study was approved by the Ethics Committee of Shanghai Chest hospital, and all participants provided written informed consent.

AUTHOR CONTRIBUTIONS

BH, TC, and YZ contributed to the conception and design of the research. BS and MH contributed to the experiment. YZ, JQ, YL, and JL contributed to the analysis and interpretation of the data. XZ and HW contributed to the clinical data collections. YZ and JQ wrote the first draft of manuscript. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2020.01049/full#supplementary-material

Supplement Figure 1 | The Kaplan-Meier curves for the OS among early stage lung adenocarcinoma patients according to CXCL9.

Supplement Table 1 | Baseline characteristics of TCGA validation cohort patients.
REFERENCES

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin. (2019) 69:7–34. doi: 10.3322/caac.21551
2. Molina JR, Yang R, Cassivi SD, Schild SE, Adjei AA. Non-small cell lung cancer: epidemiology, risk factors, treatment, and survivorship. Mayo Clin Proc. (2008) 83:584–94. doi: 10.4065/mcp.2008.06116x1(60)0735-0
3. Lorret-Tieulent J, Soerjomataram I, Ferlay J, Rutherford M, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer. (2015) 136:39–76. doi: 10.1002/ijc.29217
4. Lortet-Tieulent J, Soerjomataram I, Ferlay J, Rutherford M, Weiderpass E, et al. Increasing incidence of lung cancer in the world: a global study. Lung Cancer. (2014) 84:13–22. doi: 10.1016/j.lungcan.2014.01.009
5. Nakamura H, Saij H. Worldwide trend of increasing primary adenocarcinoma of the lung. Surg Today. (2014) 44:1004–12. doi: 10.1007/s11295-013-0636-6
6. Carnio S, Novello S, Papotti M, Loiacono M, Scagliotti GV. Prognostic and predictive biomarkers in early stage non-small cell lung cancer: tumor based approaches including gene signatures. Transl Lung Cancer Res. (2013) 2:372–81. doi: 10.3978/j.issn.2218-6751.2013.10.05
7. Shiels MS, Katki HA, Hildesheim A, Pfeiffer RM, Engels EA, Williams M, et al. Circulating inflammation markers and prospective risk for lung cancer: a Gi- and phospholipase C-dependent pathway and not via activation of Mek/p44/p42 MAPK nor Akt/PI-3 kinase. Blood. (2003) 102:1959–65. doi: 10.1182/blood-2002-12-3945
8. Muthuswamy R, Urban J, Lee JJ, Reintart TA, Bartlett D, Kalinski P. Ability of mature dendritic cells to interact with regulatory T cells is impaired during maturation. Cancer Res. (2008) 68:5972–8. doi: 10.1158/0008-5472.CAN-07-6818
9. Ikeda A, Aoki N, Kido M, Iwamoto S, Nishiura H, Maruoka R, et al. Progression of autoimmune hepatitis is mediated by IL-18-producing dendritic cells and hepatic CXCL9 expression in mice. Hepatology. (2014) 60:224–36. doi: 10.1002/hep.27087
10. Holt AP, Haughton EL, Lalar PF, Filer A, Buckley CD, Adams DH. Liver myofibroblasts regulate infiltration and positioning of lymphocytes in human liver. Gastroenterology. (2009) 136:705–14. doi: 10.1053/j.gastro.2008.10.020
11. Vanderappellen J, Van Damme J, Stryuf S. The role of CXC chemokines and their receptors in cancer. Cancer Lett. (2008) 267:226–44.
12. Shahabuddin S, Ji R, Wang P, Brailoiu E, Dun N, Yang Y, et al. CXCR3 chemokine receptor-induced chemotaxis in human airway epithelial cells: role of p38 MAPK and PI3K signaling pathways. Am J Physiol Cell Physiol. (2006) 291:C34–9. doi: 10.1152/ajpcell.00441.2005
13. Murakami T, Kawada K, Iwamoto M, Akagami M, Hara K, Nakashima Y, et al. The role of CXCR3 and CXCR4 in colorectal cancer metastasis. Int J Cancer. (2013) 132:276–87. doi: 10.1002/ijc.27670
14. Frohlich S, Martinez-Romo A, O’Connor JR, Gil-Benroso R, San-Miguel T, Terradillos L, et al. Intracellular coexpression of CXC- and CC-chemokine receptors and their ligands in human melanoma cell lines and dynamic variations after xenotransplantation. BMC Cancer. (2014) 14:118. doi: 10.1186/1471-2407-14-118
15. Ding Q, Lu P, Xia Y, Ding S, Fan Y, Li X, et al. CXCL9: evidence and contradictions for its role in tumor progression. Cancer Med. (2016) 5:3246–59. doi: 10.1002/cam4.934
16. Lieber S, Reintart S, Raifer H, Finkenagel F, Dreyer T, Bronger H, et al. Prognosis of ovarian cancer is associated with effector memory CD8(+) T cell accumulation in ascites, CXCL9 levels and activation-triggered signal transduction in T cells. Oncoimmunology. (2017) 6:e124672. doi: 10.2105/20160204.2018.124672
17. Chang KP, Wu CC, Fang KH, Tsai CY, Chang YL, Liu SC, et al. CXCL9 and total cholesterol are significant predictors of overall survival in patients with oral cavity squamous cell carcinoma. Oncol Rep. (2013) 29:1105–10. doi: 10.3892/or_00004019
18. Addison CL, Arendgen DA, Morris SR, Yue XY, Burdick MD, Mulligan MS, et al. The CXCL9 chemokine, monokine induced by interferon-gamma, inhibits non-small cell lung carcinoma tumor growth and metastasis. Hum Gene Ther. (2000) 11:247–61. doi: 10.1089/10430300050015996
36. Kowalczyk O, Burzykowski T, Niklinska WE, Kozlowski M, Chyczewski L, Niklinski J. CXCL5 as a potential novel prognostic factor in early stage non-small cell lung cancer: results of a study of expression levels of 23 genes. *Tumour Biol.* (2014) 35:4619–28. doi: 10.1007/s13277-014-1605-x

37. Nakanishi T, Imaizumi K, Hasegawa Y, Kawabe T, Hashimoto N, Okamoto M, et al. Expression of macrophage-derived chemokine (MDC)/CCL22 in human lung cancer. *Cancer Immunol Immunother.* (2006) 55:1320–9. doi: 10.1007/s00262-006-0133-y

38. Bodelon C, Polley MY, Kemp TJ, Pesatori AC, McShane LM, Caporaso NE, et al. Circulating levels of immune and inflammatory markers and long versus short survival in early-stage lung cancer. *Ann Oncol.* (2013) 24:2073–9. doi: 10.1093/annonc/mdt175

39. Zhang Y, Hu M, Han B. EP1.03-22 prognostic value of serum inflammation biomarkers in early stage lung adenocarcinoma. *J Thorac Oncol.* (2019) 14:S961–2. doi: 10.1016/j.jtho.2019.08.2103

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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