Complete mitochondrial genome of *Oberea diversipes* (Coleoptera, Cerambycidae)

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ABSTRACT

The complete mitochondrial DNA genome of the *Oberea diversipes* was reported. The genome was a circular molecule of 15,499 base pairs (bp), with 13 protein-coding genes, 22 transfer RNA genes, 2 ribosomal RNA genes, and an estimated 830 bp A+T-rich control region. The total base composition of the mitogenome was A–T bias, with 40.9% for A, 37.0% for T, 13.4% for C, and 8.7% for G. The molecular data presented here would be useful for further study of *O. diversipes*.

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KEYWORDS

Cerambycidae; genus *Oberea*; mitochondrial genome; phylogeny

Considering the diversity of the family Cerambycidae encompassing over 33,000 species (Ślipiński and Escalona 2013), about 40 cerambycids’ complete mitochondrial genome from submitted remains rather limited.

*Oberea diversipes* was restored to specific ranks (Li et al. 2016). It is widely distributed in central, south and southwest China, Laos and Vietnam, and could be harmful to Mullbery et al.

The specimen was collected from Longcanggou (N29°69′, E102°84′), Yingjing county, Ya’an city, Sichuan province. The specimen was deposited in College of Plant Protection, Southwest University with an accession number SWU-COL-160618.

The complete mitogenome of *O. diversipes* (GenBank accession number MN709785) was a circular molecule of 15,499 bp. The total base composition of the mitogenome was A–T bias, with 40.9% for A, 37.0% for T, 13.4% for C, and 8.7% for G. It contained 37 genes including 13 PCGs, 22 tRNAs, and 2 rRNA genes, and a large non-coding region, 830 bp A+T-rich control region. The gene order and orientation of *O. diversipes* were identical to those observed in other cerambycids’ mitogenomes (Song et al. 2015; Fang et al.

Figure 1. ML tree based on 13 concatenated PCGs data sets.
Twenty-three genes were encoded on the majority strand (J strand) and the remains 13 on the minority strand (N strand). All the PCGs began with an ATN start codon, five PCGs (ND2, Cox1, Cox2, Atp8, ND1) with an ATT; five genes (Atp6, Cox3, ND4, ND4l, Cob) with an ATG; three genes (ND3, ND5, ND6) with an ATA. Ten PCGs terminated with a TAN (TAA or TAG) stop codon, whereas 3 genes (Cox2, ND5, ND4) terminated with incomplete stop codons (T→). The size of 22 tRNA genes ranged from 63 bp to 72 bp and all the tRNA could be folded in to the typical clover-leaf secondary structure, except for tRNASer (AGN) lack of the dihydrouridine (DHU) stem. The 1286 bp I rRNA and 810 bp srRNA were detected in the genome. The 830 bp A+T-rich region (control region) was located between srRNA and tRNAlle.

The phylogenetic analysis was done based on the 13PCGs of 11 available cerambycids’ mitogenomes and four mitogenomes from Chrysomelidae as outgroups. The 13 individual PCGs were aligned CLUSTAL W (Thompson et al. 1994) within MEGA6 (Tamura et al. 2013), and the conserved regions were selected by using the GBlocks 0.91b (Castresana 2000). The aligned data were concatenated with Sequence Matrix v.1.7.8 (Vaidya et al. 2011). Substitution model selection of each PCG was conducted using PartitionFinder v.1.1.1 (Lanfear et al. 2012). Maximum likelihood (ML) using PhyML 3.0 (Guindon et al. 2010). A phylogenetic tree was constructed based on the ML method. The ML tree (Figure 1) showed that O. diversipes was clustered to Cerambycoides and Lamiinae were monophyly with high value support, which agrees with the traditional classification. However, Sipinilus spinicornis (belonging to subfamily Philinae) and Obrium sp. (belonging to subfamily Cerambycinae) were sister groups, which was different from the current classification (Leschen and Beutel 2014, Chapters 2.1–2.4). Therefore, we need more information and analysis in the further study.

Disclosure statement
The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

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References
Castresana J. 2000. Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. Mol Biol Evol. 17(4): 540–552.
Fang J, Qian L, Xu M, Yang XJ, Wang BD, An YL. 2016. The complete nucleotide sequence of the mitochondrial genome of the Asian longhorn beetle, Anoplophora glabripennis (Coleoptera: Cerambycidae). Mitochondrial DNA A DNA Mapp Seq Anal. 27(5):3299.
Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. 2010. New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Syst Biol. 59(3):307–321.
Guo K, Chen J, Xu CQ, Qiao HL, Xu R, Zhao XJ. 2016. The complete mitochondrial genome of the longhorn beetle Xylotrechus grayii (Coleoptera: Cerambycidae). Mitochondrial DNA A DNA Mapp Seq Anal. 27(3):2133–2134.
Lanfear R, Calcott B, Ho SYW, Guindon S. 2012. PartitionFinder: combined selection of partitioning schemes and substitution models for phylogenetic analyses. Mol Biol Evol. 29(6):1695–1701.
Leschen RAB, Beutel RG, editors. 2014. Handbook of zoology. Band 4: Arthropoda: Insecta, Teilband/Part 40: Coleoptera, beetles, Vol. 3: morphology and systematics (Phytophaga). Berlin: Walter de Gruyter.
Li Z, Tian LC, Cuccodoro G, Chen L, Lu C. 2016. Taxonomic note of Obersa fuscipennis (Chevrolat, 1852) based on morphological and DNA barcode data (Coleoptera, Cerambycidae, Lamiinae). Zootaxa. 4136 (2): 360–372.
Ślipiński A, Escalona H. 2013. Australian Longhorn beetles (Coleoptera: Cerambycidae). Volume 1: introduction and subfamily Lamiinae. Canberra: ABR/; Melbourne: CSIRO Publishing: p. 484.
Song N, Zhang H, Yin XM, Lin AL, Zhai Q. 2015. The complete mitochondrial genome sequence from the longicorn beetle Obrium sp. (Coleoptera: Cerambycidae). Mitochondrial DNA A DNA Mapp Seq Anal. 59(3):326–327.
Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. 2013. MEGA6: molecular evolutionary genetics analysis version 6.0. Mol Biol Evol. 30(12):2725–2729.
Thompson JD, Higgins DG, Gibson TJ. 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. Nucleic Acids Res. 22(22):4673–4680.
Vaidya G, Lohman DJ, Meier R. 2011. Sequence Matrix: concatenation software for the fast assembly of multi-genre datasets with character set and codon information. Cladistics. 27(2):171–180.