Bay and camphor leaf powders as inhibitors for hyperlipidemia and hepatotoxicity associated with industrial trans-fats consumption in male rats

Doha A. Omar, Samah A. El-Hashash, Amira L. Abd Allah

Nutrition and Food Science Dept., Faculty of Home Economics
Al-Azhar University, Egypt

Abstract

Trans-fats are found in large numbers of food products as well as naturally in some animal sources in small amounts. Their consumption was associated with a lot of health problems. The main objective of this study was to investigate the hypolipidemic and hepatoprotective effects of bay and camphor leaf powders (BLP and CLP, respectively) in trans-fat enriched diet-fed rats. Thirty-six adult male albino rats were used and divided into six equal groups, including the control group while groups 2 to 6 were fed for six weeks on trans-fats enriched diet (TFED, 12% hydrogenated vegetable fat) only (group 2), TFED containing 0.5 and 1% of BLP (groups 3 and 4, respectively), or TFED containing 0.5 and 1% of CLP (groups 5 and 6, respectively). At the end, body weight gain was calculated. Lipid profile, liver functions, pancreatic lipase and insulin levels were determined in sera, while cholesterol and phospholipids levels and oxidative stress markers were determined in liver tissue. Moreover, the histopathological examination of liver was performed and phenolic compounds in both powders were determined. Feeding TFED resulted in overweight, hyperlipidemia and liver dysfunction along with oxidative stress, which was confirmed histopathologically. Due to their content of phenolic compounds rather than other antioxidant agents, BLP and CLP exerted anti-obesity and hypolipidemic protective properties and alleviated the risky effects of TFED on hepatocytes. Thus, bay and camphor leaf powders can be considered efficient inhibitors for hyperlipidemia and hepatotoxicity associated with trans-fats consumption.

Keywords: Hyperlipidemia, hepatotoxicity, hydrogenated fats, bay leaves, camphor leaves, rats

Introduction

Trans-fats are composed of trans-fatty acids (TFAs) which are the stereoisomers of the naturally occurring cis-fatty acids (CFAs). They are produced industrially during hydrogenation of unsaturated oils via a process called “hardening” (Emken, 1984). This process reduces most of the double bonds in unsaturated fatty acid moieties of the oils but isomerizes some of the cis double bonds to a trans configuration, resulting in improved stability, a longer shelf life and the acquisition of desirable tactile, functional and sensory properties. Trans-fats are found in foods originating from ruminant animals, such as cows and sheep, and are found in foods containing partially hydrogenated vegetable oils (PHVO). Industrial trans-fats are widely used to produce a variety of foods, including margarines, cookies, pastries, salad dressings and cooking oils (Korver and Katan, 2006).
Approximately 540,000 deaths each year can be attributed to intake of industrially produced trans-fatty acids (Wang et al., 2016). High trans-fat intake increases the risk of death from any cause by 34%, coronary heart disease deaths by 28% (De Souza et al., 2015). Trans-fat increases low density lipoprotein cholesterol (LDL-c) levels while lowering high density lipoprotein cholesterol (HDL-c) levels (De Souza et al., 2015; Mozaffarian et al., 2006).

Animal trans-fat levels can comprise up to 6% of a product’s fat content in ruminant foods, and industrial trans-fat levels can comprise up to 60% of a product’s fat content in foods containing partially hydrogenated vegetable oil (PHVO). The discovery of adverse effects on the blood cholesterol profile and the increased risk of coronary heart disease of industrial trans-fat (Mozaffarian et al., 2006; De Souza et al., 2015) have led to public health recommendations to lower total trans-fat intake to below 1% of total energy intake, primarily by the removal of industrial trans-fat (FAO/WHO, 2010).

On the other hand, high intake of dietary trans-fats has been reported to enhance the steatogenic and pro-fibrotic properties of the western diet in mice (Hansen et al., 2017). Trans-fats likely sensitize mice to the effects of high fat diet by increasing insulin resistance, hepatic lipogenesis and oxidative stress (Machado et al., 2010).

Plants are natural source of treatment and are used from ages for food and medicine (Shah et al., 2021). Herbal plants have also been found to nourish the body and provide vitamins, minerals, and many trace elements that are easy to absorb (Melkegna and Jonah, 2021). Laurus nobilis L. (L. nobilis, bay) leaves are used mainly as a seasoning in cooking. They are rich in active compounds such as phenols, flavonols and flavones, and have antioxidant and antimicrobial effects (Palazzo et al., 2020).

L. nobilis has been positively evaluated by various researchers and explored wide range of pharmacological activities. Antioxidant and wound-healing effects of the ethanolic extract of L. nobilis have been well reported (Vardapetyan et al., 2013). Bay leaves have many other biologic activities such as antibacterial, antiviral, immunostimulant, anticholinergic, antifungal, insect repellent, anticonvulsant, anti-mutagenic, analgesic and anti-inflammatory activities (Batool et al., 2020). L. nobilis was also used for preventing and treating type II diabetes because it reduces the level of serum glucose (Khan et al., 2009).

On the other hand, Cinnamomum camphora L. (C. camphora) is an ever green tree native in China and distributed to some Asian countries. C. camphora is famous for its ornamental, economic, and medicinal value (Chen and Dai, 2015). The leaves are considered one of the most important parts of this plant. They are rich in bioactive compounds (Zhang et al., 2020) which have strong fumigant and anthelmintic activities (Chen et al., 2014). Also, the oil of the seed kernel has antioxidant, anti-inflammatory (Fu et al., 2016), anti-aging and anti-bacterial effects (Chen et al., 2018; He et al., 2018). However, little information is available regarding the preventive influences of Laurus nobilis L. and Cinnamomum camphora L. leaf powders on hyperlipidemia and hepatotoxicity associated with industrial trans-fats consumption in vivo; hence this study was carried out.
Materials and Methods

Materials:

Plant leaves:
Dry leaves of bay (*Laurus nobilis* L.) were purchased from Arab company for Pharmaceutical and Medicinal plants, MEPACO, Egypt. Fresh champhor (*Cinnamomum camphora*) leaves were sampled from several parts of Nawag village, Tanta City, Gharbiya Governorate, Egypt.

Animals:
A total of 36 adult male albino rats (*Sprague-Dawley* strain) weighing 150±10 g were obtained from the animal colony, Helwan farm, Vaccine and Immunity Organization, Ministry of Health, Cairo Governorate, Egypt.

Chemicals and other ingredients:
Casein (≥ 80% protein), cellulose, vitamins, minerals, corn starch, DL-methionine, choline chloride, formalin, diethyl ether and other required chemicals were obtained from Elgomhouria Company for Trading Drugs, Chemicals and Medical Appliances, Cairo, Egypt. Soybean oil and the vegetable ghee were purchased from the local market, Tanta City, Gharbiya Governorate, Egypt.

Methods:

Drying of fresh camphor leaves:
Fresh camphor leaves were washed thoroughly, allowed to drain and then subjected to solar energy for drying.

Preparation of leaf powder:
Dry leaves of the studied plants were milled to fine powders, sieved and stored at room temperature in closed glass bottles in dark place until used.

Phenolic profile identification of plant samples:
Phenolic compounds of leaf powders were identified and determined by high-performance liquid chromatography (HPLC) according to Goupy et al. (1999).

Experimental diets:
Basal diet was prepared from fine ingredients per 100g. It had the following composition: Casein (≥ 80% protein) 14%, soybean oil 4%, cellulose 5%, mineral mixture 3.5%, vitamin mixture 1%, choline chloride 0.25%, DL-methionine 0.3% and corn starch up to 100g (Reeves et al., 1993).

Trans-fats enriched diet (TFED) had the same composition of basal diet mentioned formerly with substituting soybean oil with a source of industrial trans-fats(vegetable ghee) and increasing the percentage from 4% to 12% according to Angelis-Pereira et al. (2017). For treated groups, TFED was supplemented with the powder of either bay or camphor leaves (BLP or CLP, respectively) at two percentages (0.5 and 1%)

Experimental design:
Animals were kept in clean wire cages under hygienic conditions in a room maintained at a relative humidity 40-60%, 20–25°C and 12/12 h day light/darkness cycle. Feed was introduced (ad libitum) to the rats for adaptation for one week in special food containers to avoid scattering. Similarly,
fresh water was provided ad-libitum and checked daily. After that, rats were randomly assigned to 6 equal groups. The 1st group was fed on basal diet only as a negative control group (-veg), while the 2nd group was fed on trans-fats enriched diet (TFED) only as a positive control group (+veg). The 3rd and 4th groups were fed on TFED containing BLP (0.5 and 1%, respectively), while the 5th and 6th groups were fed on TFED containing CLP (0.5 and 1%, respectively).

The experiment lasted for six weeks. Meanwhile, rats were weighed weekly. At the end, animals were weighed, fasted overnight, and then sacrificed under very light ether anesthesia. Blood samples were collected from hepatic portal vein of each rat into dry clean centrifuge tubes. Serum was carefully separated by centrifugation of blood samples at 3500 round per minute for 15 minutes at room temperature, transferred into dry clean Eppendorf tubes, then kept frozen at -20°C for biochemical determinations. Livers were removed from rats by careful dissection, washed in saline solution (0.9%) and dried using filter paper. A specimen of each liver was kept in formalin solution (10%) for later histopathological examination, while other specimen was kept at (-80 °C) for later homogenization and analysis.

**Calculation of body weight gain:**

Body weight gain (BWG) in grams was calculated by subtracting the initial weight of each rat from its final weight.

**Determination of lipid profile:**

Triglycerides (TG), total cholesterol (T.C) and high density lipoprotein cholesterol (HDL-c) were determined in serum according to the method described by Trinder and Ann (1969), Richmond (1973) and Lopes- Virella et al. (1977), respectively. On the other hand, very low and low density lipoprotein cholesterol (VLDL-c and LDL-c, respectively) were calculated using the equations of Friedwald et al. (1972). Atherogenic coefficient (AC) was calculated according to Brehm et al. (2004) and Nimmanapalli et al. (2016) using the following equation: AC = (T.C - HDL-c)/HDL-c.

**Determination of pancreatic lipase:**

Pancreatic lipase (PL) was determined in serum according to the method described by Lykidis et al. (1994).

**Determination of liver cholesterol and phospholipids:**

Cholesterol and phospholipids were determined in liver tissue homogenate (Liv. Cho and Liv. PhLs) according to the methods of Richmond (1973) and Ray et al. (1969), respectively.

**Determination of total antioxidant capacity and malondialdehyde level in liver tissue homogenate:**

In liver tissue homogenate, total antioxidant capacity (TAC) was determined according to Koracevic et al. (2001). Also, lipid peroxidation, expressed as malondialdehyde (MDA), was determined in liver tissue homogenate according to the method described by Ohkawa et al. (1979).
**Determination of liver functions:**

The activities of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) were determined in serum according to the method described by Reitman and Frankel (1957), while total protein (T. P) and albumin (ALB) were estimated according to Sonnenwirth and Jaret (1980).

**Histopathological examination:**

After sacrificing rats, livers were removed and immersed in 10% buffered neutral formalin solution. The fixed specimens were then trimmed, washed and dehydrated in ascending grades of alcohol. After that, they were cleared in xylol, embedded in paraffin, cut in sections of 4-6 microns thickness and stained with haematoxylin and eosin (Drury and Wallington, 1980).

**Statistical analysis:**

Statistical analysis was carried out using one way analysis of variance (ANOVA) test followed by Duncan test through the program of statistical packages for the social science (SPSS). Results were expressed as mean ± standard deviation (mean ± SD). The differences among means at p < 0.05 were considered significant (Snedecor and Cochran, 1989).

**Results and Discussion**

**Phenolic profile in both bay and camphor leaves:**

Bay and camphor leaf powders were analyzed by HPLC for their phenolic compounds. The obtained results were shown in table 1. Except for chlorogenic acid, bay leaf powder recorded lower content of all analyzed phenolic compounds than camphor leaf powder. Chlorogenic acid was not found in camphor leaf powder, while it was found in bay leaf powder (133.49 ppm). Phenolic compounds that were found in high concentrations in bay leaf powder (> 500 ppm) were catechin and ellagic acid (1536.12 and 586.95 ppm, respectively). This is in accordance with El-Gawishet al. (2021) who showed that Laurus nobilis recorded higher content of catechin and ellagic acid. On the other hand, the presence of some phenolic compounds such as pyrogallol as well as vanillic and ferulic acids is in agreement with Muñiz-Márquez et al. (2013) and Muchuweti et al. (2007). In camphor leaf powder, phenolic compounds that were found in high concentrations (> 1000 ppm) were ellagic acid, catechin, catechol and P-OH-benzoicacid (3474.51, 2918.73, 2307.79 and 1340.86 ppm, respectively). Thus, it can be concluded that the two compounds found in both powders in great amounts are ellagic acid and catechins, but in different arrangement. Regarding animal studies, ellagic acid was proved to have a significant hypolipidemic effect in mice liver (Xu et al., 2021). Catechins in green tea were found to induce a specific hypolipidemic effect in rats (Kim et al., 2011).
Table (1)

Analysis of phenolic compounds in both bay and camphor leaf powder samples

| Phenolic compounds (ppm)          | Bay leaf powder | Camphor leaf powder |
|-----------------------------------|----------------|---------------------|
| Catechin                          | 1536.12        | 2918.73             |
| Ellagic acid                      | 586.95         | 3474.51             |
| Ferulic acid                      | 403.05         | 513.65              |
| Caffeine                          | 271.67         | 321.14              |
| Catechol                          | 271.09         | 2307.79             |
| Chlorogenic acid                  | 133.49         | --                  |
| Pyrogallol                        | 105.17         | 715.81              |
| Coumarin                          | 84.82          | 438.25              |
| Vanillic acid                     | 65.84          | 227.38              |
| Caffeic acid                      | 55.78          | 519.23              |
| P-OH-benzoic acid                 | 30.63          | 1340.86             |
| 4-Aminobenzoic acid               | 15.77          | 69.33               |
| Gallic acid                       | 3.14           | 77.69               |

Body weight gain & serum lipid profile:

Data presented in table 2 showed the effect of feeding rats TFED containing bay and camphor leaf powders on body weight gain, serum lipid profile as well as atherogenic coefficient (AC). The mean values of BWG, TG, T.C, LDL-c, VLDL-c and AC were significantly increased in TFED-fed group compared with the control group. Feeding TFED supplemented with bay or camphor leaf powder (0.5, 1%) caused significant decrease compared with feeding on TFED only. In contrast, the mean value of HDL-c was significantly lower in TFED-fed group compared with the control. TFED containing bay or camphor leaf powders (0.5, 1%) led to significant increase compared with TFED alone. In general, the best results were obtained in the groups fed on TFED+1% of BLP, followed by TFED+ 1% CLP, with no significant difference between them.

The present results were in agreement with Kavanagh et al. (2007) who revealed that monkeys fed on a trans-fat diet gained 7.2% of their body weight, as compared to 1.8% for monkeys on a monounsaturated fat diet. Trans-fats enriched diet has an indirect dyslipidemic action. It increases the fat mass in the body (Kavanagh et al., 2007), which in turn is associated with insulin resistance (Arita et al., 1999; Ishibashi et al., 2018). It is thought that in the insulin-resistant state, dyslipidemia occurs (DeFronzo and Ferrannini, 1991).

The hypolipidemic effect of bay leaf powder, in the present study, was in agreement with Al-Samarrai et al. (2017). Gasparyan et al. (2015) also mentioned that extract of bay leaves decreased serum T.C and TG in carbon tetrachloride–intoxicated male Wistar rats. Similarly, Casamassima et al. (2017) showed that bay leaves improved lipid profile in hyperlipidemic rabbits. In fact, nearly no studies were found on the hypolipidemic action of camphor leaves. However, other parts of the plant were analyzed and studied. Fu et al. (2016) found that levels of blood total cholesterol, triglycerides and free fatty acid in the Cinnamomum camphora seed kernel oil group were decreased significantly compared to lard and soy groups.
This hypolipidemic action of the both powders can be attributed to its phenolic content. As mentioned above, the two compounds found in both powders in great amounts were ellagic acid and catechins. Both compounds were reported to have a significant hypolipidemic action in animal studies (Xu et al., 2021; Kim et al., 2011). Of phenolics, flavonoids and their derivatives play an important role in enhancing lipid profile. For example, quercetin was found to decrease serum LDL-c and TG levels in rats (Rafieian-Kopaei et al., 2013). Alchalabi et al., (2020) supported the previous suggestion, as they attributed the hypolipidemic action of Laurus nobilis alcoholic extract in diabetic rats to flavonoids found in the leaves, since they have the ability to manage lipid profile.

| Parameters | BWG (%) | TG (mg/dl) | T.C (mg/dl) | HDL-c (mg/dl) | VLDL-c (mg/dl) | LDL-c (mg/dl) | AC          |
|------------|---------|------------|-------------|---------------|----------------|---------------|-------------|
| Control(-ve g) | 6.78± 0.57 | 63.80± 7.33 | 72.50± 4.70 | 44.05± 3.40 | 12.76± 1.47 | 15.56± 1.75 | 13.12± 1.46 |
| TFED(+ve g)  | 17.1± 1.25 | 176.98± 4.09 | 129.90± 2.03 | 35.40± 3.33 | 66.13± 3.19 | 37.73± 1.99 |
| TFED + BLP (0.5%) | 15.03± 0.68 | 108.30± 8.91 | 87.93± 4.59 | 21.66± 1.79 | 31.84± 1.79 | 22.61± 1.79 |
| TFED + BLP (1%)  | 11.12± 1.30 | 96.95± 2.47 | 76.13± 6.53 | 19.39± 0.49 | 20.44± 2.64 | 20.04± 1.98 |
| TFED + CLP (0.5%) | 14.78± 0.93 | 121.73± 7.66 | 90.48± 6.53 | 35.05± 2.87 | 24.35± 4.99 | 32.06± 2.64 |
| TFED + CLP (1%)  | 11.82± 1.31 | 108.20± 4.78 | 79.85± 3.83 | 21.64± 3.05 | 22.66± 0.96 | 22.29± 2.05 |

Values that have different letters in each column differ significantly, while the difference among those with similar letters is not significant (p<0.05).

Pancreatic lipase in serum:

The effect of bay and camphor leaf powders on pancreatic lipase activity in serum of TFED-fed rats was presented in table 3. The activity of pancreatic lipase (PL) was significantly increased in serum of TFED-fed group compared with the control group. Marked improvement was noticed as a result of supplementing TFED with bay or camphor leaf powder (0.5, 1%). In general, the best results were obtained in the groups fed on TFED supplemented with the higher percentages of bay or camphor leaf powder.

TFED, used through the present study, is a type of high fat diet (HFD) as it consisted of 12% fats versus 4% only in basal diet. So, the present results were in agreement with El-Hashash (2014). Previously, Gidez (1973) demonstrated that levels of pancreatic lipase were increased when the fat content of the diet was raised from about 5% to 15–22%.

Inhibitory effect of some plant extracts on pancreatic lipase was presented by Gholamhoseinian and co-worker, (2010), where they have shown the percent (%) inhibition of pancreatic lipase. According to the data given in a research paper percent inhibition of Laurus nobilis leaves against pancreatic lipase was 20.5% (Gholamhoseinian et al., 2010). Dietary fibers exert inhibitory actions on pancreatic lipase (Dukehart et al., 1989). Thus, the high content of crude fibers in bay leaf powder (26.3± 0.33/100 g) according to Tawfek and Ali (2022) can account the reduced effect of BLP.
Doha A. Omar et al

containing TFED compared with TFED alone on serum pancreatic lipase.

On the other hand, the decreasing effect of both powders on pancreatic lipase activity is a cause for their hypolipidemic effect.

Table (3)

| Groups                  | Parameters | PL (U/L) |
|-------------------------|------------|----------|
| Control(-veg)           |            | 10.07 ± 1.34a |
| TFED(+veg)              |            | 16.70 ± 1.40a |
| TFED + BLP (0.5%)       |            | 12.63 ± 0.91b |
| TFED + BLP (1%)         |            | 10.37 ± 0.40c |
| TFED + CLP (0.5%)       |            | 13.40 ± 1.41b |
| TFED + CLP (1%)         |            | 10.53 ± 0.85c |

-Values that have different letters in each column differ significantly, while the difference among those with similar letters is not significant (p<0.05).

Lipid profile and oxidation indices in liver tissue homogenate:

The mean values of cholesterol and malondialdehyde in liver tissue homogenate (Liv. Cho and MDA, respectively) were significantly increased in TFED-fed group compared with the control group. In contrast, the mean values of phospholipids and total antioxidant capacity in liver tissue homogenate (Liv. PhLs and TAC, respectively) were significantly decreased. Feeding TFED supplemented with bay or camphor leaf powder (0.5, 1%) improved all these indices significantly as compared to feeding on TFED only, except for TAC level as TFED+0.5% CLP could not affect it significantly. In general, the best results were obtained in the groups fed on TFED+1% of BLP, followed by TFED+1% CLP (Table 4).

In line with the present results, Bravo et al. (2011) demonstrated that the high fat diet used to induce nonalcoholic fatty liver disease in rats caused an increase in liver TG (× 2.6) and cholesterol (+30%). Regarding the oxidative effect of HFD, Also, Machado et al. (2010) reported that trans-fats may sensitize mice to the effects of high fat diet by increasing insulin resistance, hepatic lipogenesis and oxidative stress. Moreover, Dhibi et al. (2011) revealed that the rates of hepatic lipid peroxidation were markedly higher in margarine-fed groups than in the control group. It was indicated that trans-fatty acids impair fat cell membrane fluidity. When they are incorporated into cell membranes, the membrane fluidity is reduced and the cells do not function as well. The resulting effect is then to promote further production of reactive oxygen species causing lipid peroxidation.

On the other hand, the antioxidant effect of bay leaves were in agreement with Fang et al. (2005) who reported that sesquiterpene lactones identified in bay leaf were found to have different pharmacological properties including enhancement of liver glutathione S-transferase (GST) activity. Elmastas et al. (2006) explained that the antioxidant activity of ethanol extract of L. nobilis leaves may be due to phenolic compounds present in the extract. Thus, it can be concluded that managing of lipid profile in liver tissue and the antioxidant effect of both studied leaf powders are due to their content of phenolic compounds such as ellagic acid (Xu et al., 2021).
Effect of bay and camphor leaf powders on lipid profile and oxidative indices in liver tissue homogenate of TFED-fed rats

| Groups                  | Liv. Cho (mg/dl) | Liv. PhLs (mg/dl) | TAC (mm/L) | MDA (nmol/g tissue) |
|-------------------------|------------------|-------------------|------------|---------------------|
| Control(-veg)           | 77.67± 6.43      | 305.67± 19.56     | 0.46± 0.06 | 39.33± 5.51         |
| TFED(+veg)              | 149.33± 3.06     | 115.33± 8.33      | 0.18± 0.08 | 99.00 ± 6.56        |
| TFED + BLP (0.5%)       | 111.00± 14.18    | 190.67± 9.45      | 0.30± 0.05 | 75.00± 8.19         |
| TFED + BLP (1%)         | 86.67± 2.89      | 249.00± 16.70     | 0.37± 0.05 | 57.00± 9.00         |
| TFED + CLP (0.5%)       | 123.33± 2.08     | 157.67± 8.14      | 0.20± 0.04 | 81.67 ± 9.45        |
| TFED + CLP (1%)         | 98.00± 3.61      | 238.00± 9.54      | 0.33± 0.05 | 65.33 ± 14.57      |

-Values that have different letters in each column differ significantly, while the difference among those with similar letters is not significant (p<0.05).

Liver functions:

The activities of transaminases (ALT and AST) were increased significantly in serum of TFED group compared with the control group. In contrast, the mean values of total protein and albumin (T.P and ALB) were significantly decreased. Feeding TFED supplemented with bay or camphor leaf powder (0.5, 1%) improved all these indices significantly as compared to feeding on TFED only. The protective effects of bay and camphor leaf powders, as noticed here, increased by increasing their concentrations (Table 5).

These results were in agreement with Dhibi et al. (2011) who found that the activities of transaminases, alkaline phosphatase and lactate dehydrogenase increased significantly in margarine-consumed group compared to the control group. Lin et al. (2010) concluded that higher saturated fat intake is significantly associated with the presence of high albuminuria. As a result, serum albumin decreased significantly. Regarding the effective role of Laurus nobilis, the present results were in line with Alchalabi et al. (2020) who mentioned that statistical reduction of the ALT, AST and ALP activities in diabetic group given Laurus nobilis alcoholic extract compared to normal control group and this was attributed to the antioxidant compounds in the extract. Also, Casamassima et al. (2017) reported a decrease in the activities of ALT and AST in New Zealand white growing rabbits as a result of feeding on dried leaves of Laurus nobilis-contaminated meals. In rats, Mohammed et al. (2021) mentioned that albumin and total protein levels in L. nobilis extract group were insignificantly increased compared to untreated diabetic group. In general, the hepatoprotective effects of either bay or camphor leaf powder can be attributed to its antioxidant and hypolipidemic effects.
Table(5)

Effect of bay and camphor leaf powderson liver function indices in serum of TFED-fed rats

| Groups                  | Parameters | ALT (U/L) | AST (U/L) | T.P (g/dl) | ALB (g/dl) |
|-------------------------|------------|-----------|-----------|------------|------------|
| Control(-veg)           | 11.35±2.85 | 86.75±8.66 | 7.30±0.23 | 4.27±0.15  |
| TFED(+veg)              | 39.58±4.55 | 197.58±17.26 | 5.18±0.66 | 2.95±0.41  |
| TFED + BLP (0.5%)       | 11.75±1.74 | 85.73±7.72  | 6.73±0.20 | 4.01±0.10  |
| TFED + BLP (1%)         | 12.00±1.84 | 82.40±6.90  | 6.92±0.53 | 4.08±0.21  |
| TFED + CLP (0.5%)       | 16.43±2.54 | 90.28±7.41  | 6.11±0.24 | 3.72±0.22  |
| TFED + CLP (1%)         | 11.43±2.77 | 89.65±8.13  | 6.88±0.32 | 4.06±0.19  |

-Values that have different letters in each column differ significantly, while the difference among those with similar letters is not significant (p<0.05).

Histological examination results:

Histological examination results were illustrated in the following figures:

Figs. A&B: Microscopic pictures of H&E stained liver sections show normal hepatocytes arranged in radiating plates around a central vein (CV) with normal sinusoids (s) in the control group.

Figs. C&D: Liver sections from TFED-fed group show prominent centrilobular macrovesicular steatosis (black arrow) in hepatocytes around congested central vein (red arrow) with occluded sinusoids and the other hepatocytes show hydropic degeneration (yellow arrow). Low magnification X: 100 bar 100, high magnification X: 400 bar 50.
Figs. E&F:
Microscopic pictures of H&E stained liver sections of rats fed on TFED containing 0.5% BLP show milder centrilobular macrovesicular steatosis (black arrow) in hepatocytes around mildly congested central vein (red arrow) compared to TFED-fed group. Figs. G&H: Liver sections from rats fed on TFED containing 1% BLP show retained normal histological picture of hepatocytes, central vein and sinusoids (s). Low magnification X: 100 bar 100, high magnification X: 400 bar 50.

Figs. I&J:
Microscopic pictures of H&E stained liver sections of rats fed on TFED containing 0.5% CLP show mild hydropic degeneration (yellow arrow) in hepatocytes with occluded sinusoids and mild congestion (red arrow). Figs. K&L: Liver sections from rats fed on TFED containing 1% CLP show greatly improved histological picture with very few large cytoplasmic vacuoles (black arrow) and normal sinusoids (s). Low magnification X: 100 bar 100, high magnification X: 400 bar 50.
The abnormal changes noticed in hepatocytes of TFED group were in accordance with many animal studies. Popescu et al. (2013) found that HFD consumption for one month by male mice was associated with some abnormalities in hepatocytes including hepatocyte swelling, sometimes associated with karyomegaly, granular or finely vacuolated cytoplasm, numerous activated Kupffer cells, indistinct sinusoids (no visible lumen), because of the volume increase of the hepatocytes, unaltered portal fields. These lesions were consistent with a medium hepatopathy (granular and vacuolar degeneration of the hepatocytes). Similarly, El-Hashash (2014) reported that liver sections from rats fed on high saturated fat diet showed vacuolation of cytoplasm, kupffer cells activation, fatty degeneration of hepatocytes, congestion of portal vein, proliferation of bile ductules and fibroplasia in portal triad that was associated with a portal infiltration with inflammatory cells (H and E ×400). Insulin resistance and oxidative stress are the most important mechanisms through which high fat diet can negatively affect hepatocytes (Mehta et al., 2002). So, the protective effects noticed in the groups fed on TFED containing either BLP or CLP can be attributed to their content of polyphenolic compounds, rather than other antioxidants, which were proved to decrease insulin resistance and alleviate oxidative stress (Fang et al. 2005; Elmastaş et al. 2006).

**Conclusion**
Bay and camphor leaf powders can be considered efficient semi protective agents against hyperlipidemia and hepatotoxicity associated with TFED consumption

**References**

Al-Samarrai, O.R.; Naji, N.A. and Hameed, R.R. (2017):
Effect of Bay leaf (*Laurus nobilis* L.) and its isolated (flavonoids and glycosides) on the lipids profile in the local Iraqi female rabbits. Tikrit Journal of Pure Science, 22 (6):72-75.

Alchalabi, S.; Majeed, D. M.; Jasim, A. and Al-Azzawi, K.S.A. (2020):
Benefit effect of ethanolic extract of Bay leaves (*Laura nobilis*) on blood sugar level in adult diabetic rats induced by alloxan monohydrate. Annals of Tropical Medicine and Public Health, 23(16). DOI:10.36295/ASRO.2020.231608

Angelis-Pereira, M.C.; Barcelos, M.F.P.; Pereira, J.A.R.; Pereira, R.C. and Souza, R.V. (2017):
Effect of different commercial fat sources on brain, liver and blood lipid profiles of rats in growth phase. Acta Cir. Bras., 32(12):1013-1025. DOI: 10.1590/s0102-865020170120000003

Arita, Y.; Kihara, S.; Ouchi, N.; Takahashi, M.; Maeda, K.; Miyagawa, J.; Hotta, K.; Shimomura, I.; Nakamura, T.; Miyaoka, K.; Kuriyama, H.; Nishida, M.; Yamashita, S.; Okubo, K.; Matsubara, K.; Muraguchi, M.; Ohmoto, Y.; Funahashi, T. and Matsuzawa, Y. (1999):
Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. Biochem. Biophys. Res. Commun., 257(1):79-83.

Batool, S.; Khera, R.A.; Hanif, M.A. and Ayub, M.A. (2020):
Bay leaf. Medicinal Plants of South Asia, 2020:63–74. DOI: 10.1016/B978-0-08-102659-5.00005-7
Bravo, E.; Palleschi, S.; Aspichueta, P.; Buqué, X.; Rossi, B.; Cano, A.; Napolitano, M.; Ochoa, B. and Botham, K.M. (2011):
High fat diet-induced nonalcoholic fatty liver disease in rats is associated with hyperhomocysteinemia caused by down regulation of the transsulphuration pathway. Lipids in Health and Disease, 10:60.

Brehm, A.; Pfeiler, G.; Pacini, G.; Vierhapper, H. and Roden, M. (2004):
Relationship between serum lipoprotein ratios and insulin resistance in obesity. Clin. Chem., 50:2316-2322.

Casamassima, D.; Palazzo, M.; Vizzarri, F.; Coppola, R.; Costagliola, C.; Corino, C. and Di Costanzo, A. (2017):
Dietary effect of dried bay leaves (Laurus nobilis) meal on some biochemical parameters and on plasma oxidative status in New Zealand white growing rabbit. Journal of Animal Physiology and Animal Nutrition, 101(5):175-184.

Chen, C.; Zheng, Y.; Zhong, Y.; Wu, Y.; Li, Z. and Xu, L. A. (2018):
Transcriptome analysis and identification of genes related to terpenoid biosynthesis in Cinnamomum camphora. BMC Genomics, 19:550. DOI: 10.1186/s12864-018-4941-1

Chen, H.P.; Yang, K.; You, C.X.; Lei, N.; Sun, R.Q.; Geng, Z.F.; Ma, P.; Cai, Q.; Du, S.S. and Deng, Z.W. (2014):
Chemical constituents and insecticidal activities of the essential oil of Cinnamomum camphora leaves against Lasiodermaserricorne. Journal of Chemistry, Volume 2014, Article ID 963729, 5 pages. DOI: 10.1155/2014/963729

Chen, Y. and Dai, G. (2015):
Acaricidal activity of compounds from Cinnamomum camphora (L.) Presl against the carmine spider mite, Tetranychuscinnabarinus. Pest Manage. Sci., 71:1561–1571. DOI: 10.1002/ps.3961

DeFronzo, R.A. and Ferrannini, E. (1991):
Insulin resistance: A multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. Diabetes Care, 14(3):173-194.

De Souza, R.J.; Mente, A.; Maroleanu, A.; Cozma, A.I.; Ha, V.; Kishibe, T.; Uleryk,E.; Budylowski, P.; Schünemann, H.; Beyene, J.; Anand, S.S. (2015):
Intake of saturated and trans unsaturated fatty acids and risk of all cause mortality, cardiovascular disease, and type 2 diabetes: Systematic review and meta-analysis of observational studies. BMJ, 351:h3978. DOI: 10.1136/bmj.h3978

Dhibi, M.; Brahmi, F.; Mnari,A.;Houas,Z.; Chargui,I.; Bchir,L.; Gazzah,N.; Alsaif,M.A.; and Hammami, M. (2011):
The intake of high fat diet with different trans fatty acid levels differentially induces oxidative stress and nonalcoholic fatty liver disease (NAFLD) in rats. Nutr. Metab. (Lond), 8: 65. DOI: 10.1186/1743-7075-8-65
Doha A. Omar et al

Drury, R.A.B. and Wallington, E.A. (1980): 
Carlton’s Histological Techniques, 5th edition. Oxford University Press. London, New York, Toronto, p 344-345.

Dukehart, M.R.; Dutta, S.K. and Vaeth, J. (1989): 
Dietary fiber supplementation: Effect on exocrine pancreatic secretion in man. Am. J. Clin. Nutr., 50(5): 1023-1028.

El-Gawish, A.M.; EL-Gezawy, E.S. and Zeima, N.M. (2021): 
The potential protective effect of alcoholic extracts of some herbs on hepatotoxicity induced by paracetamol in experimental rats.

Journal of Research in the Fields of Specific Education, 7(37): 1349-1378. DOI:10.21608/jedu.2021.77041.1354

El-Hashash, S.A. (2014): 
Effect of Ficus sycomorus L. leaves on high fat diet-fed rats: Possible mechanisms behind the prevention of obesity and its related disorders. IOSR Journal of Environmental Science, Toxicology and Food Technology, 8:7-16.

Elmastaş, M.; Gülçin, I.; İşildak, Ö.; Küfrevioğlu, Ö.; İbaoğlu, K. and Aboul-Enein, H. (2006): 
Radical scavenging activity and antioxidant capacity of bay leaf extracts. Journal of the Iranian Chemical Society, 3:258–266.

Emken, E.A. (1984): 
Nutrition and biochemistry of trans and positional fatty acid isomers in hydrogenated oils. Annu. Rev. Nutr., 4:339–376.

Fang, F.; Sang, S.; Chen, K.; Gosslau, A.; Ho, C. and Robert, T. (2005): 
Rosen isolation and identification of cytotoxic compounds from Bay leaf (Laurus nobilis). Food Chem., 93:497–501.

FAO/WHO, 2010 
(Food and Agriculture Organization/ World Health Organization): Fats and Fatty Acids in Human Nutrition.WHO Press; Geneva, Switzerland. Report of an Expert Consultation.

Friedwald, W.T; Levee, R.I. and Fredrickson, D.S. (1972): 
Estimation of the concentration of low-density lipoprotein separated by three different methods. Clin. Chem., 18:499-502.

Fu, J.; Zeng, C.; Zeng, Z.; Wang, B. and Gong, D. (2016): 
Cinnamomum camphora seed kernel oil ameliorates oxidative stress and inflammation in diet-induced obese rats. J. Food Sci., 81:H1295–H1300. DOI: 10.1111/1750-3841.13271

68
Gasparyan, G.; Tiratsuyan, S.; Kazaryan, S. and Vardapetyan, H. (2015):  
Effect of *Laurus nobilis* extract on the functioning of liver against CCl4 induced toxicity. Journal of Experimental Biology and Agricultural Sciences, 3(2):174-183.

Gholamhoseinian, A.; Shahouzehi, B. and Sharifi-Far, F. (2010):  
Inhibitory effect of some plant extracts on pancreatic lipase. IJP-International Journal of Pharmacology, 6(1), 18-24.

Gidez, L.I. (1973):  
Effect of dietary fat on pancreatic lipase levels in the rat. J. Lipid Res., 14(2):169-177.

Goupyp, P.; Hugues, M.; Biovin, P. and Amiot, M.J. (1999):  
Antioxidant composition and activity of barley (Hordeum Vulgare) and malt extract and of isolated phenolic compounds. J. Sci. Food Agri., 79:1625-1634.

Hansen, H.H.; Feigh, M.; Veidal, S.S.; Rigbolt, K.T.; Vrang, N. and Fosgerau, K. (2017):  
Mouse models of nonalcoholic steatohepatitis in preclinical drug development. Drug Discovery Today, 22(11):1707-1718. DOI: 10.1016/j.drudis.2017.06.007

He, H.; Qin, J.; Cheng, X.; Xu, K.; Teng, L. and Zhang, D. (2018):  
Effects of exogenous 6-BA and NAA on growth and contents of medicinal ingredient of Phellodendron chinense seedlings. Saudi J. Biol. Sci., 25:1189–1195. DOI: 10.1016/j.sjbs.2017.11.037

Ishibashi, K.; Takeda, Y. and Atsumi, G. (2018):  
Effect of trans fatty acid on insulin responsiveness and fatty acid composition of lipid species of 3T3-L1 adipocytes. In: Szablewski, L. (ed.), Adipose Tissue [Internet]. London: IntechOpen [cited 2022 Jun 24]. 278 p. Available from: https://www.intechopen.com/books/6581.DOI: 10.5772/intechopen.71377

Kavanagh, K.; Jones, K.L.; Sawyer, J.; Kelley, K.; Carr, J.J.; Wagner, J.D. and Rudel, L.L. (2007):  
Trans-fat diet induces abdominal obesity and changes in insulin sensitivity in monkeys. Obes. Res., 15:1675–1684. DOI: 10.1038/oby.2007.20

Khan, A.; Zaman, G. and Anderson, R. A. (2009):  
Bay leaves improve glucose and lipid profile of people with type 2 diabetes. Journal of Clinical Biochemistry and Nutrition, 44(1):52–56. DOI: 10.3164/jcbn.08-188

Kim, A.; Chiu, A.; Barone, M.k.; Avino, D.; Wang, F.; Coleman, C.I. and Phung, O.J. (2011):  
Green tea catechins decrease total and low-density lipoprotein cholesterol: a systematic review and meta-analysis. J. Am. Diet Assoc., 111(11):1720-1729.DOI: 10.1016/j.jada.2011.08.009

Koracevic, D.; Koracevic, G.; Djordjevic, V.; Andrejevic, S. and Cosic, V. (2001):  
Method for the measurement of antioxidant activity in human fluids. J. Clin. Pathol., 54: 356–361.
Doha A. Omar et al

Korver, O. and Katan, M.B. (2006):
The elimination of trans fats from spreads: How science helped to turn an industry around. Nutr. Rev., 64:275–279.

Lin, J.; Judd, S.; Le, A.; Ard, J.; Newsome, B.B.; Howard, G.; Warnock, D.G. and McClellan, W. (2010):
Associations of dietary fat with albuminuria and kidney dysfunction. Am. J. Clin. Nutr., 92(4): 897–904. DOI: 10.3945/ajcn.2010.29479

Lopes-Virella, M. F.; Stone, S.; Ellis, S. and Collwell, J. A. (1977):
Cholesterol determination in high-density lipoprotein separated by three different methods. Clin. Chem., 23(5): 882.

Lykidis, A.; Mougios, V. and Arzoglou, P. (1994):
Pancreatic lipase assays with triglycerides as substrate: Contribution of each sequential reaction to product formation. Clinical Chemistry, 40(11 Pt 1):2053-2056. DOI: 10.1093/clinchem/40.11.2053

Machado, R.M.; Stefano, J.T.; Oliveira, C.P.M.S.; Mello, E.S.; Ferreira, F.D.; Nunes, V.S.; de Lima, V.M.R.; Quintão, E.C.R.; Catanozi, S.; Nakandakare, E.R. and Lottenberg, A.M.P. (2010):
Intake of trans fatty acids causes nonalcoholic steatohepatitis and reduces adipose tissue fat content. The Journal of Nutrition, 140(6): 1127–1132. DOI: 10.3945/jn.109.117937

Mehta, K.; Van Thiel, D.H.; Shah, N. and Mobharan, S. (2002):
Nonalcoholic fatty liver disease: Pathogenesis and the role of antioxidants. Nutr. Rev., 60(9): 289–293.

Melkegna, T.H. and Jonah, S.A. (2021):
Elemental analysis of medicinal plants used for the treatment of some gastrointestinal diseases in Ethiopia using INAA technique. Biol. Trace Elem. Res.,1207-1212.DOI: 10.1007/s12011-020-02236-2

Mohammed, R.R.; Omer, A.K.; Yener, Z.; Uyar, A. and Ahmed, A.K. (2021):
Biomedical effects of Laurus nobilis L. leaf extract on vital organs in streptozotocin-induced diabetic rats: Experimental research. Ann. Med. Surg. (Lond), 61: 188–197. DOI: 10.1016/j.amsu.2020.11.051

Mozaffarian, D.; Katan, M.B.; Ascherio, A.; Stampfer, M.J. and Willett, W.C. (2006):
Trans fatty acids and cardiovascular disease. N. Engl. J. Med., 354:1601–1613. DOI: 10.1056/NEJMra054035.

Muchuweti, M.; Kativu, E.; Mupure, C.H.; Chidewe, C.; Ndhlala, A.R. and Benhura, M.A.N. (2007):
Phenolic composition and antioxidant properties of some spices. American Journal of Food Technology, 2: 414-420. DOI:10.3923/ajft.2007.414.420

70
Muñiz-Márquez, D.B.; Rodríguez, R.; Balagurusamy, N.; Carrillo, M.L.; Belmares, R.; Contreras, J.C.; Nevárez, G.V. and Aguilar, C.N. (2013):
Phenolic content and antioxidant capacity of extracts of *Laurus nobilis* L., *Coriandrum sativum* L. and *Amaranthus hybridus* L., CyTA - Journal of Food, DOI:10.1080/19476337.2013.847500

Nimmanapalli, H.D.; Kasi, A.D.; Devapatla, P.K. and Nuttakki, V. (2016):
Lipid ratios, atherogenic coefficient and atherogenic index of plasma as parameters in assessing cardiovascular risk in type 2 diabetes mellitus. International Journal of Research in Medical Sciences, Int. J. Res. Med. Sci., 4(7):2863-2869.DOI: 10.18203/2320-6012.ijrms20161966

Ohkawa, H.; Ohishi, N. and Yagi, K. (1979):
Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Analytical Biochemistry, 95:351–358.

Palazzo, M.; Vizzarri, F.; Arvay, J.D.; Alessandro, A.G.; Casamassima, D.; Ratti, S.; Corino, C. and Rossi, R.(2020):
Dietary effect of dried bay leaves (*Laurus nobilis*) meal on selected productive performances and on quality meat traits in growing rabbits. Livestock Science, 242:104301.

Popescu, L.A.; Virgolici, B.; Lixandru, D.; Miricescu, D.; Condu⁰, E.; Timnea, O.; Ranetti, A. E.; Militaru, M.; Mohora, M. and Zăgrean, L. (2013):
Effect of diet and omega-3 fatty acids in NAFLD. Rom. J. Morphol. Embryol., 54(3 Suppl):785–790.

Rafieian-Kopaei, M.; Baradaran, A. and Rafieian, M. (2013):
Oxidative stress and the paradoxical effects of antioxidants. J. Res. Med. Sci., 18 (7): 628.

Ray, T.K.; Xlcipski, V.P.; Barclay, M.; Essner, E. and Archibald, F.M. (1969):
Lipid composition of rat liver plasma membranes. J. Biol. Chem.,244: 5528-5536.

Reeves, P.G.; Nielsen, F.H. and Fahey, G.C. (1993):
AIN-93 purified diets for laboratory rodents: Final report of the American Institute of Nutrition Ad Hoc Writing Committee on the Reformulation of the AIN-76A Rodent Diet. J. Nutr., 123:1939-1951.

Reitman, S. and Frankel, S. (1957):
A colorimetric method for the determination of serum glutamic pyruvic transaminase. J. Clin. Pathol., 28: 56-63.

Richmond, N. (1973):
Enzymatic colorimetric test for cholesterol determination. Clin. Chem., 19:1350-1356.

Shah, S.; Nisar, Z.; Nisar, J.; Akram, M.; Ghotekar, S. and Oza, R. (2021):
Nanobiomedicine: A new approach of medicinal plants and their therapeutic modalities. J. Mater. Environ. Sci., 12(1): 1-14.
Doha A. Omar et al

Snedecor, G.W. and Cochran, W.G. (1989): Statistical Methods, 8th Edition. Iowa State University Press, Ames.

Sonnenwirth, A. and Jaret, L. (1980): Grad Wholes Clinical Laboratory Methods and Diagnosis, 18th edition. Mosby, London, p 258-259.

Tawfek, M.A. and Ali, A.R.M. (2022): Effectiveness of cardamom (Elettaria cardamomum) or bay leaf (Laurus nobilis L.) powder in improving the quality of Labneh. ActaSci.Pol. Technol. Aliment., 21 (1): 39-52. DOI: 10.17306/J.AFS.2022.0984

Trinder, P. and Ann, S. (1969): Enzymatic colorimetric test with lipid clearing factor to determine triglycerides. Clin. Biochem., 6:24-27.

Vardapetyan, H.; Tiratsuyan, S.; Hovhannisyan, A.; Rukhkyan, M. and Hovhannisyan, D. (2013): Phytochemical composition and biological activity of Laurus nobilis L. leaves collected from two regions of South Caucasus. Journal of Experimental Biology and Agricultural Sciences, 1:45-51.

Wang, Q.; Afshin, A.; Yakoob, M.Y.; Singh, G.M.; Rehm, C.D.; Khatibzadeh, S, Micha, R.; Shi, P. and Mozaffarian, D. (2016): Impact of nonoptimal intakes of saturated, polyunsaturated, and trans fat on global burdens of coronary heart disease. Journal of the American Heart Association, 5(1):e002891.

Xu, Q.; Li, S.; Tang, W.; Yan, J.; Wei, X.; Zhou, M. and Diao, H. (2021): The effect of ellagic acid on hepatic lipid metabolism and antioxidant activity in mice. Front. Physiol., 12:751501. DOI: 10.3389/fphys.2021.751501

Zhang, Z.; Wu, M.; Lai, Y.; Li, X.; Zhang, D. and Chen, Y. (2020): Efficient extraction of bioenergy from Cinnamomum camphora leaves. Frontiers in Energy Research, 8:90.
مسحوقى أوراق الغار والكافور كموانع لارتفاع دهون الدم والتسمم الكبدي المرتبطين باستهلاك الدهون المتحولة الصناعية في ذكور الجرذان

ضحى عبد الفتاح عمر، سماح أحمد الحشاش، أميرة لطفي عبد الله
قسم التغذية وعلوم الأطعمة – كلية الاقتصاد المنزلي – جامعة الأزهر – مصر

الملخص العربي

توجد الدهون المتحولة في عدد كبير من المنتجات الغذائية كما تواجد طبيعيا بكميات بسيطة في بعض المصادر الحيوانية، وقد ارتبط استهلاكها بكثير من المشاكل الصحية. الهدف الرئيسي من هذه الدراسة هو بحث التأثيرات الخفيفة للدهون والوفيرة للنومز مسحوقى أوراق الغار والكافور في الجرذان التي تعمل على خفض غلي الدهون المنظورة. تم استخدام عدد 36 من ذكور جرذان الألبينو حيث تم تقسيمها إلى ست مجموعات متساوية العدد تشمل المجموعة الضابطة بينما تم تغذية المجموعات من 2 إلى 6 لمدة ستة أسابيع على غذاء غلي الدهون المتحولة (والذي تتألف من 12% من الزيوت النباتية المهدرة) فقط بالنسبة للمجموعة الثانية أو محتوئا على مسحوق أوراق الغار بنسبة 0.5، 1% بالنسبة للمجموعتين الثالثة والرابعة على الترتيب أو محتويًّا على مسحوق أوراق الكافور بنسبة 0.5، 1% بالنسبة للمجموعتين الثالثة والرابعة على الترتيب. تم حساب الزيادة المكتسبة في وزن الجسم، كما تم تقييم الدورة ووظائف الكبد وليزونgéت البنكرياس والإنسولين في السيرم، ومستويات الكوليسترول والفسفور في الدم، ودلال الإجهاد الكبدي في نسيج الكبد. علاوة على ذلك تم إجراء الفحص الهستوباثولوجي للكبد، وتقييم المركبات الفينولية في كلا المسحوقين. أظهرت النتائج زيادة في وزن الجسم وارتفاع في دهون الدم وانخفاض في وظائف الكبد ووظائف الكبد الانتقائي وهي ما أدى الفحص الهستوباثولوجي للكبد، وتدفقه المركبات الفينولية في كلا المسحوقين. أظهرت النتائج زيادة في وزن الجسم وانخفاض في وظائف الكبد. ولاليف الإجهاد الكبدي في نسيج الكبد. أظهرت النتائج زيادة في وزن الجسم وانخفاض في وظائف الكبد. ولاليف الإجهاد الكبدي في نسيج الكبد. أظهرت النتائج زيادة في وزن الجسم وانخفاض في وظائف الكبد. ولاليف الإجهاد الكبدي في نسيج الكبد. أظهرت النتائج زيادة في وزن الجسم وانخفاض في وظائف الكبد.

الكلمات المفتاحية: ارتفاع دهون الدم، التسمم الكبدي، الدهون المتحولة، أوراق الغار، أوراق الكافور، الجرذان.