Prognostic and clinicopathological significance of MUC expression in head and neck cancer: a systematic review and meta-analysis

Hao Lu1,2,*, Dan Liang3,*, Yun Zhu1,2, Wanlin Xu1,2, Kaishuo Zhou1,2, Limin Liu1,2, Shengwen Liu1,2 and Wenjun Yang1,2

1 Department of Oral Maxillofacial-Head and Neck Oncology, Ninth People’s Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China
2 Shanghai Research Institute of Stomatology and Shanghai Key Laboratory of Stomatology, Shanghai, China
3 State Key Laboratory of Oral Diseases, National Clinical Research Center for Oral Diseases, West China Hospital of Stomatology, Sichuan University, Chengdu, China
* These authors have contributed equally to this work

Correspondence to: Wenjun Yang, email: ywjdoctor@outlook.com
Correspondence to: Shengwen Liu, email: wubenzhimu17@163.com

Keywords: MUC, head and neck cancer, biomarkers, prognosis, meta-analysis

Received: December 23, 2016    Accepted: February 28, 2017    Published: July 27, 2017

ABSTRACT

The prognostic value of mucins expression in patients with head and neck cancer (HNC) remains controversial. To address this, a meta-analysis was performed to systematically evaluate prognostic significance of mucins expression in HNC. Electronic and manual searches were performed and a total of 20 studies including 2046 patients were selected for the final analysis. Increased mucins expression was associated with unfavorable overall survival in HNC patients (HR=1.83, 95% CI: 1.43-2.33, p=0.000). Mucins overexpression was also in correlation with more advanced TNM stage (RR=0.84, 95% CI: 0.73-0.97, p=0.017), higher risk of lymph node metastasis (RR=0.69, 95% CI: 0.57-0.84, p=0.000) and deeper invasion (RR=0.58, 95% CI: 0.44-0.76, p=0.000). These results suggested that elevated mucins expression was significantly associated with worse prognosis and more detrimental clinicopathological outcomes, revealing the promising potential of mucins as biomarkers for HNC management.

INTRODUCTION

Head and neck cancer (HNC) is a group of biologically similar cancers originating from the oral cavity, nasopharynx, oropharynx, hypopharynx and larynx, which mainly behave as squamous cell carcinoma histologically. HNC is the sixth most frequent type of malignant tumor, causing more than 400,000 deaths annually worldwide [1-5]. Its high mortality rate and the disfigurement or functional deficiency that survivors may suffer result in a considerable global public health burden. Despite great advance in multidisciplinary combined diagnosis and treatment, only 30-50% patients with HNC survive over 5 years after initial diagnosis worldwide [6]. Clinically, the classic tumor, node, and metastasis (TNM) staging system is widely used for the initial diagnosis but failed to reflect the inherent biological heterogeneity especially in atypical early symptoms or concealed metastasis patients. Therefore, novel biomarkers involved in cancer development are greatly needed to stratify patients with poor prognosis of HNC in order to make optimal individualized therapy.

Much attention has been focused on the involvement of mucins (MUC) in tumor carcinogenesis and metastasis recently. MUC is a family of high O-glycosylated protein, characterized by a basic structure including a central polymorphic tandem repeat region [7-8]. Only expressing on the apical surfaces of various luminal and glandular normal epithelial cells, MUC play an important role in cell-cell adhesion, immune response and alteration of intracellular signaling [9]. However, the tightly regulated homeostatic expression may be disrupted by various factors such as cancer cells, in particular. The observation of subcellular distribution and biochemical features...
changes during malignant transformation and tumor progression suggest that MUC may be the key point in carcinogenesis and subsequent metastasis of cancers [10-14]. Therefore, aberrant MUC expression may be predictive biomarkers in HNC.

Over the past decade, numerous independent studies have evaluated the clinical and the prognostic value of MUC protein expression in HNC. Yet, the results of these reports remain controversial and no clear consensus has been achieved so far [15-19]. Limited to small sample size some publications may draw inconsistent results due to potential random errors. Therefore, we conducted a systematic review and meta-analysis to address the association between MUC expression and prognostic value and the common clinicopathological parameters of HNC.

RESULTS

Eligible studies

A total of 655 potential relevant studies were retrieved after the primary database searches and two additional studies were obtained from the reference lists. After careful screening of the titles and abstracts, 609 articles were excluded as shown in Figure 1. Then eventual 48 publications underwent elaborately full-text evaluation. Eventually, 20 observational studies consisting of 2064 cases were satisfied for subsequent pooling calculation [15-19, 26-40].

![PRISMA 2009 Flow Diagram](image)

**Figure 1:** Flow diagram of the study selection process.
Table 1: Characteristics of eligible studies included in the meta-analysis.

| First Author | Year | Country | Case Number/positivity rate | Cancer Type | Mean age | Male/Female | AT* | Methode | Ab used | Expression Location | Cutoff | C. Features\# | Prognostic \# | Analyzes Methode | NOS* + Score |
|--------------|------|---------|-----------------------------|-------------|----------|------------|-----|----------|---------|---------------------|--------|--------------|------------|-----------------|-----------|
| 1 Nitta T    | 2000 | Japan   | 77 (59.7%)                  | OSCC        | NA       | NA         | No  | IHC     | DF3, IF3, D3        | Membrane and/ or cytoplasm | 5%     | D,T,N,S,I       | NA         | NA              | 8         |
| 2 Alex L     | 2005 | Spain   | 80 (100%/ 95%)              | MEC         | 53.0     | 18/22      | AR/A  | IHC     | Maek95, IHC        | Membrane and cytoplasm          | 5%     | D             | sur. curve | M               | 8         |
| 3 Haendel LA | 2005 | France  | 63 (71%/ 79%)               | MEC         | 52.2     | 20/43      | NA   | IHC     | Maek95, IHC        | Membrane and cytoplasm          | Median | P             | sur. curve | M               | 8         |
| 4 Wieruszewski VB | 2004 | USA     | 74 (31%)                    | PTC         | NA       | 28/46      | No   | IHC     | V64/5, V64/5        | Membrane and cytoplasm          | Score ≥2 | NA           | HR         | M               | 7         |
| 5 Hamada T   | 2012 | Japan   | 206 (39%)                   | OSCC        | 66.3     | 133/73     | No   | IHC     | DF3                   | Membrane and cytoplasm          | 5%     | G,D,T,N,S,P,I   | HR         | U/M             | 8         |
| 6 Cros J     | 2013 | France  | 114 (26.3%)                 | MSGT        | 52+18    | NA         | AR/A  | IHC     | Polyclonal, Cytoplasm | Cytoplasm | Score ≥2 | NA           | HR         | U              | 8         |
| 7 Masha MA   | 2016 | USA     | 87 (78%)                    | HNSCC       | 59.5     | 61/25      | NA   | IHC     | V64/5, Cytoplasm | Cytoplasm | Score ≥1 | G,D,N,S       | NA         | NA             | 8         |
| 8 Gorani EC  | 2016 | Iran    | 280 (75%)                   | PTC and FC | NA       | NA         | NA   | IHC     | V64/5, Cytoplasm | Cytoplasm | Score ≥3 | G,T,S,A,I     | NA         | NA             | 7         |
| 9 Baek SK    | 2017 | Korea   | 87 (72%)                    | PTC         | 48.6     | 15/72      | NA   | IHC     | V64/5, V64/5        | Membrane and cytoplasm          | Score ≥3 | G,T,N,S,A,I   | NA         | NA             | 8         |
| 10 Croce MV  | 2018 | Argentina | 125 (82%)                  | HNSCC       | 59.9     | 98/27      | NA   | IHC     | CT33, Cytoplasm     | Membrane and cytoplasm          | Median | G,D,T,N,S       | NA         | NA             | 7         |
| 11 Li X      | 2015 | China   | 59 (66.1%)                  | HC          | 58.2     | 48/11      | No   | IHC     | Neovlab             | Membrane and cytoplasm          | 50%    | G,D,N,S        | NA         | NA             | 6         |
| 12 Song L    | 2014 | China   | 62 (92%)                    | MEC         | 56.2     | 48/14      | No   | IHC     | Neovlab             | Membrane and cytoplasm          | 25%    | G,T,N,P        | HR         | M               | 8         |
| 13 Renaud F  | 2014 | France  | 94 (49%)                    | PTC         | 42.3     | 23/71      | NA   | IHC     | M8                     | Membrane and cytoplasm          | 20%    | G,N,A,I        | sur. curve | M               | 8         |
| 14 Liu S     | 2014 | China   | 357 (69%)                   | MEC         | 45.9     | 162/195    | No   | IHC     | Neovlab             | Membrane and cytoplasm          | 25%    | G,D,T,P        | HR         | M               | 8         |
| 15 Paleri V   | 2014 | UK      | 30 (43%)                    | LSCC        | 66.0     | NA         | AR   | IHC     | Neovlab             | Nucleus and/or cytoplasm         | Score ≥1 | NA           | HR         | M               | 6         |
| 16 Hamada T   | 2012 | Japan   | 150 (41%)                   | OSCC        | 64.5     | 97/53      | No   | IHC     | -/ BS7               | Membrane and cytoplasm          | 5%     | G,D,N,S,T,I     | HR         | U/M             | 8         |
| 17 He F      | 2019 | China   | 53 (57%)                    | PTC         | 50±11/ 45±12 | 16/37    | No   | IHC     | DF3                   | Membrane and cytoplasm          | 25%    | G,N,A,S        | NA         | NA             | 7         |
| 18 Weed DT   | 2001 | USA     | 40 (85%)                    | HNSCC       | 63       | NA         | No   | IHC     | -/ 1SH10             | Membrane and cytoplasm          | 10%    | D,T,N,S        | NA         | NA             | 7         |
| 19 Weed DT   | 2004 | USA     | 28 (54% for 16%, 79% for 1SH10) | MEC         | 54.3     | 11/17      | No   | IHC     | -/ 1SH, 1SH10        | Membrane and cytoplasm          | 30%    | D             | sur. curve | M               | 7         |
| 20 Croce MV  | 2015 | UK      | 29 (69%)                    | HNSCC       | 58.1     | 23/6       | No   | IHC     | CS95, Cytoplasm     | Membrane and cytoplasm          | Median | G,N,T,G,A     | NA         | 8               | 8         |

*AT: Adjuvant therapy; \#C. Features: clinicopathological features; AR: adjuvant radiotherapy; AC: adjuvant chemotherapy; NA: not available

**NOS: Newcastle-Ottawa Scale; Sur. curve: survival curve; IHC: immunohistochemistry; ISH: in situ hybridization; HR: hazard ratio; A: age; G: gender; T: TNM stage; D: differentiation; S: tumor size; N: lymph node metastasis; P: perineural invasion; I: mode of invasion

OSCC: oral squamous cell carcinoma; MEC: mucoepidermoid carcinoma; PTC: papillary carcinoma; MSGT: malignant tumors of the salivary glands

FC: follicular carcinoma; HNSCC: head and neck squamous cell carcinoma; HC: hypopharyngeal carcinoma; LSCC: laryngeal squamous cell carcinoma
Demographic characteristics of included studies

Altogether, the sample-size ranged from 28 to 357, with a median of 103.2 participants. The median age of patients in all studies was 55.6 years old (range, 42.3-66.3) and the mean proportion of male patients was about 52.9%. The matching criteria varied among the enrolled studies. Studies concerning MEC and PTC occupied the largest proportion of cancer types among all primary literatures ($n = 5$, respectively), followed by HNSCC ($n = 4$), OSCC ($n = 3$) and remaining types of solid neoplasm.

The MUC1 and MUC4 expression were evaluated by IHC staining in paraffin-embedded tissue blocks using several monoclonal antibodies as DF3, Ma695, VU4H5 for MUC1 and 1G8, 8G7, 15H10 for MUC4 respectively.

As for the clinicopathological factors, seventeen eligible studies were divided into seven subgroups: 5 for age, 11 for gender, 10 for TNM tumor stage, 11 for tumor size, 13 for lymph node metastasis, 12 for tumor differentiation and 6 for mode of invasion. With respect to the quality of the including studies, none of the eligible entries scored less than six stars by the Newcastle-Ottawa Scale system, indicating a relative high methodological quality across all studies. Other detailed information were extracted and summarized in Table 1.

Impact of MUC expression on survival rates of patients in HNC

Seven observational trials including 993 participants offered original data on survival rates in terms of different MUC expression (Table 2). It demonstrated that increased MUC activity was associated with unfavorable survival rate ($HR = 1.83$, 95%CI: 1.43-2.33, $P < 0.001$). There was no significant heterogeneity among these studies ($I^2 = 0.0\%$, $p$ (Q-test) = 0.577), so that a fixed-effect model was used to combine the HR and 95% CI (Figure 2).

Furthermore, subgroup analysis stratified by nationality, cancer type, adjuvant therapy, sample size, methods, cut-off value, MUC subtype, tumor location, antibody for MUC1 and antibody for MUC4 were performed respectively.

With regard to nationality, a worse survival rate was strongly linked to MUC positivity in Asian patients ($n = 4$, $HR = 1.92$, 95% CI: 1.46-2.52, $p = 0.000$, $I^2 = 0.0\%$), while over-expression of MUC in western countries was irrelevant with poor prognosis ($n = 3$, $HR = 1.50$, 95% CI: 0.88-2.57, $p = 0.140$, $I^2 = 6.5\%$).

In the subgroup analysis by cancer type, a worse survival rate was strongly linked to MUC positivity in salivary tumors ($n = 3$, $HR = 2.04$, 95% CI: 1.36-3.07, $p = 0.001$, $I^2 = 10.0\%$) than in non-salivary tumors ($n = 4$, $HR = 1.72$, 95% CI: 1.27-2.33, $p = 0.000$, $I^2 = 0.0\%$).

As to adjuvant therapy (AT) and surgery operation, elevated MUC expression was referred to a worse prognostic role in non-AT ($n = 5$, $HR = 1.96$, 95% CI: 1.51-2.53, $p = 0.000$, $I^2 = 0.0\%$). Nevertheless, higher MUC activity was irrelevant with AT ($n = 2$, $HR = 1.06$, 95% CI: 0.51-2.19, $p = 0.874$, $I^2 = 0.0\%$).

In the subgroup analysis by sample size, higher MUC expression status was significantly associated with poorer survival rate both in sample size > 100 group ($n = 4$, $HR = 1.76$, 95% CI: 1.34-2.31, $p = 0.000$, $I^2 = 0.0\%$)

Figure 2: Funnel plot of hazard ratios (HR) for overall survival associated with high level of MUC expression.
and sample size < 100 group (n = 3, HR = 2.12, 95% CI: 1.25-3.61, p = 0.006, I² = 22.6%).

There were two stratified subgroup in terms of detecting method among papers. Higher MUC expression was related to poorer survival rate according to IHC method (n = 6, HR = 1.88, 95% CI: 1.47-2.41, p = 0.000, I² = 0%), but not to ISH method (n = 1, HR = 0.85, 95% CI: 0.24-3.02, p = 0.802).

Subgroups analysis by different cut-off values indicating that high MUC expression was associated to a worse prognosis no matter in cut-off > 10% (n = 5, HR = 1.95, 95% CI: 1.38-2.77, p = 0.000, I² = 0%) or cut-off < 10% (n = 2, HR = 1.72, 95% CI: 1.22-2.41, p = 0.002, I² = 0%).

Stratified by MUC subtype of the included papers, MUC1 positive status was identified as a worse prognosis marker in HNC (n = 5, HR = 2.09, 95% CI: 1.51-2.89, p = 0.000, I² = 0%), while MUC4 detected by other antibodies were irrelevant with poor prognosis (n = 1, HR = 0.85, 95% CI: 0.24-3.02, p = 0.802).

Correlation of MUC expression with clinicopathological parameters

We also investigated the association of high MUC expression and clinicopathological features. As reported in Table 3, higher MUC expression was significantly associated with more advanced TNM stage (I+II vs. III+IV, n = 10, RR = 0.84, 95% CI: 0.73-0.97, p = 0.017, Figure 3C), lymph node metastasis (negative vs. positive, n = 13, RR = 0.69, 95% CI: 0.57-0.84, p = 0.000, Figure 3E), and mode of invasion (1 to 3 vs. 4c + 4d, n = 6, RR = 0.58, 95% CI: 0.44-0.76, p = 0.000, Figure 3F). However, MUC over-expression was not significantly associated
with gender (male vs. female, \(n = 11\), RR = 0.98, 95% CI: 0.90-1.08, \(p = 0.707\)), age (\(<45\) yr vs. \(\geq 45\) yr, \(n = 5\), RR = 0.91, 95% CI: 0.75-1.09, \(p = 0.304\)), grade of differentiation (well + moderate vs. poor, \(n = 12\), RR = 1.16, 95% CI: 0.98-1.37, \(p = 0.075\)) and tumor size (T1 + T2 vs. T3 + T4, \(n = 11\), RR = 0.90, 95% CI: 0.75-1.07, \(p = 0.236\)).

Subgroup analysis stratified by different MUC subtype was performed to compare the sensitivity of MUC subtype as biomarkers. With regard to UICC stage, elevated MUC4 expression was referred to play a worse prognostic role in head and neck cancers (\(n = 1\), RR = 0.67, 95% CI: 0.46-0.97, \(p = 0.036\)) (Table 3).

### Sensitivity analysis

Pooled estimates of the relation of MUC overexpression to prognostic outcomes and clinicopathological parameters were not substantially altered according to the ‘leave-one-out’ method, demonstrating the reliability of our results (Figure 4A-4H).

### Publication bias

Analysis of survival rate and clinicopathological features demonstrated no obvious asymmetry in the funnel plots for publication bias (\(p > 0.05\)) (Figure 5A-5H). More sensitive Egger’s regression test confirmed these results, indicating that our pooled results had no significant publication bias.

Table 3: Meta-analysis of the association between MUC expression and clinicopathological features of head and neck cancer.

| Clinicopathological variables | Study N. | Samples | Stat. | Pooled data | Test of heterogeneity |
|------------------------------|----------|---------|-------|-------------|-----------------------|
|                              |          |         | RR (95% CI) | \(p\) value | \(\chi^2\) | \(p\) value | \(I^2\) (%) |
| Gender (male/female)         | All studies | 11 | 1502 (726/776) | FEM | 0.98 (0.90-1.08) | 0.707 | 9.52 | 0.392 | 6.2 |
| Subgroup                     | MUC1     | 8  | 1266 (568/698) | FEM | 0.98 (0.89-1.08) | 0.676 | 8.53 | 0.272 | 0.0 |
|                             | MUC4     | 2  | 236 (158/78)   | FEM | 1.00 (0.79-1.28) | 0.993 | 0.86 | 0.443 | 0.0 |
| Age (<45 yr / \(\geq 45\) yr) | All studies | 5  | 514 (239/275)  | FEM | 0.91 (0.75-1.09) | 0.304 | 0.12 | 0.998 | 0.0 |
| Subgroup                     | MUC1     | 5  | 514 (239/275)  | FEM | 0.91 (0.75-1.09) | 0.304 | 0.12 | 0.998 | 0.0 |
|                             | MUC4     | 0  |         |         |           |         |     |       |     |
| UICC stage (I + II / III+IV) | All studies | 10 | 1206(806/490) | REM | 0.84 (0.73-0.97) | 0.017 | 26.76 | 0.002 | 66.4 |
| Subgroup                     | MUC1     | 9  | 1146(702/444)  | REM | 0.87 (0.76-1.00) | 0.052 | 20.20 | 0.010 | 60.4 |
|                             | MUC4     | 1  | 150(104/46)    | REM | 0.56 (0.39-0.80) | 0.002 | 0.00 | —     | —     |
| Differentiation (well + moderate / poor) | All studies | 12 | 1233(902/331) | REM | 1.16 (0.98-1.37) | 0.075 | 21.89 | 0.025 | 49.7 |
| Subgroup                     | MUC1     | 9  | 961(690/271)   | REM | 1.12 (0.90-1.38) | 0.272 | 19.74 | 0.011 | 59.5 |
|                             | MUC4     | 3  | 272(212/60)    | REM | 1.23 (0.96-1.58) | 0.099 | 2.06 | 0.357 | 2.90 |
| Lymph node (with / without metastasis) | All studies | 13 | 1101(424/677) | REM | 0.69 (0.57-0.84) | 0.000 | 58.22 | 0.000 | 79.4 |
| Subgroup                     | MUC1     | 11 | 871(338/533)   | REM | 0.66 (0.53-0.83) | 0.000 | 55.07 | 0.000 | 81.8 |
|                             | MUC4     | 2  | 720(144)       | REM | 0.82 (0.65-1.22) | 0.317 | 2.89 | 0.089 | 65.4 |
| Mode of invasion (1 to 3 / 4c + 4d) | All studies | 6  | 745(498/247)  | REM | 0.58 (0.44-0.76) | 0.000 | 18.25 | 0.003 | 72.6 |
| Subgroup                     | MUC1     | 5  | 595(389/206)   | REM | 0.56 (0.40-0.78) | 0.001 | 17.42 | 0.002 | 77.0 |
|                             | MUC4     | 1  | 150(109/41)    | REM | 0.67 (0.46-0.97) | 0.036 | 0.00 | —     | —     |
| Tumor size (T1 + T2 / T3 + T4) | All studies | 11 | 1046(660/386) | REM | 0.90 (0.75-1.07) | 0.236 | 33.37 | 0.000 | 70.0 |
| Subgroup                     | MUC1     | 9  | 816(601/315)   | REM | 0.92 (0.76-1.12) | 0.416 | 23.21 | 0.003 | 65.5 |
|                             | MUC4     | 2  | 230(159/71)    | REM | 0.79 (0.41-1.52) | 0.477 | 9.44 | 0.002 | 89.4 |

Stat: Statistic models; FEM: fixed-effect model; REM: random-effect model; RR: relative risk; 95% CI: 95% confidence interval
DISCUSSION

As revealed by the laboratorial evidence, aberrant overexpression, mislocation profiles and truncated glycans of tumor-associated MUC (TA-MUC) were commonly observed in variety of epithelial cancers. The structural and functional complexity of TA-MUC emphasizes its pivotal value in the pathogenesis and progression of cancer. However, some conflicting conclusions were reported in clinic. Although the majority of studies demonstrated that high level of MUC indicated worse clinicopathological parameters and poor prognosis, there are still a number of studies which showed the opposite conclusion [16, 17, 19]. Thus, a comprehensive study is urgently demanded.

To our knowledge, the present study is the first and most full-scale meta-analysis systemically exploring the possible prognostic role of MUC up-regulation in...
HNC. Here, 20 eligible studies including 2064 cases were combined to yield statistics, indicating that higher MUC expression was strongly related to poorer overall survival (HR = 1.83, 95% CI: 1.43-2.33, \( p = 0.000 \)) in patients with HNC. Subgroup analysis was consistent with the pooled OS regardless of cancer type, sample size, staining cut-off value and MUC subtypes. Among patients accepted adjuvant therapy (AR and/or AC) along with surgery operation, no significant association was identified between positive MUC expression and poor OS, exhibiting a sharp contrast with the patients without AT (HR = 1.96, 95% CI: 1.51-2.53, \( p = 0.000 \)). The discrepancy may be, partially, explained by the advanced radiotherapy technique and platinum-based therapeutic regimen for advanced HNC, achieving a significant improvement in 5-year OS [41]. As for western countries and ISH subgroups stratified by nationality and detecting methods respectively, no significant association was identified though a tendency was shown. This situation was possibly due to the relatively limited studies enrolled in the subgroups. As shown in Table 2, only three qualified studies for western countries and one for ISH detecting method studies were subjected to subgroup analysis, which may affect the real results. Based on these points,

\[ \text{Figure 4: Sensitivity analysis based on stepwise omitting one study at a time for overall survival (OS). (A) HR (B) Gender (C) Age (D) UICC stage (E) Differentiation (F) Lymph node metastasis (G) Mode of invasion (H) Tumor size.} \]
large-scaled studies are still needed to strength our results in the future. Stratified by the location of tumor, aberrant expression of MUC was significantly associated with poor OS in OSCC subgroup, which exhibited a sharp contrast with the patients that suffering from LSCC. The discrepancy may be, partially, explained by the morphologic diversity between oral cavity and laryngeal regions combined with different carcinogens. The inherent biological heterogeneity induces different disease evolution process though both of them originate from epithelial cells. Varying results were noted with regard to different antibodies used for detecting MUC1 or MUC4 specificity. DF3 for MUC1 may be a more tumor-specific antibody in immunohistochemistry than others.

Figure 5: Begg’s funnel plot for the evaluation of potential publication bias on overall estimate of overall survival (OS). (A) HR (B) Gender (C) Age (D) UICC stage (E) Differentiation (F) Lymph node metastasis (G) Mode of invasion (H) Tumor size.
Figure 6: Trial sequential analysis of studies reporting the association between MUC protein expression and (A) UICC stage, (B) Lymph node metastasis and (C) Mode of invasion. The solid blue line represents the cumulative Z-curve. The dashed red line represents the trial sequential monitoring boundary. TSA indicates that no further trials are required.
(i.e. VU4H5) in HNC especially for paraffin-embedded tissues, as the same to 8G7 for MUC4. Perhaps the best explanation for staining sensitivity differences among these antibodies may be the different binding epitopes. For example, aberrantly underglycosylated TA-MUC1 tissues may somehow mask the carbohydrate epitopes recognized by VU4H5, while still permitting reaction with DF3 which binds to peptide epitopes within the variable number of tandem repeat (VNTR) domain [42]. Although different survival rate were exhibited by these antibodies, the same tendency revealed that the increased expression of MUC protein was associated with unfavorable OS in patients with HNC.

Consistent with the theoretical inference, our results also demonstrated that overexpression of MUC was tightly associated with advanced TNM stage (I+II vs. III+IV, RR = 0.84, 95%CI: 0.73-0.97, p = 0.017 ), high risk of cervical lymph node metastasis (negative vs. positive, RR = 0.69, 95%CI: 0.57-0.84, p = 0.000 ) and the depth of invasion (1 to 3 vs. 4c+4d, RR = 0.58, 95%CI: 0.44-0.76, p = 0.000 ). Given that more advanced TNM stage, more positive cervical lymph node metastasis and deeper invasion are adverse prognostic features, the pooled analysis may explain why positive MUC expression was associated with poor survival rate in patients with HNC. Several studies have uncovered unique roles of the MUC on the oncogenic and pro-metastasis effects, including promoting proliferation, metabolism, angiogenesis, invasion, metastasis, epithelial to mesenchymal transition (EMT) and resistance to apoptosis [9, 14, 43-47]. For example, multiple studies provide direct evidence for the role of MUC1 in EMT process, which enable cancer cells acquire their invasive and metastasis potential. Notably, MUC1 can directly inhibit the expression of E-cadherin, but upregulation the EMT inducers, such as Snail, Slug, Vimentin and Twist [43-44]. Similarly, MUC4 can specifically potentiate phosphorylation of the receptor tyrosine kinase ErbB2, which results in up-regulation of the cyclin-dependent kinase inhibitor p27kip and silence of protein kinaseB/Akt pathways [47]. Taken together, these mechanisms can explain the association between high MUC expression and adverse clinical pathological features in HNC patients. Therefore, MUC can serve as a good candidate for predicting the status and prognosis of HNC, making up the deficiency of the current physical detecting methods.

Apart from the inspiring outcomes, several limitations still existed in this meta-analysis. First, the lack of unified standardized protocol and evaluation system for detecting MUC status influence the accurate estimation of prognosis for HNC. Second, the numbers of studies and patients pooled into analysis were limited due to insufficient data. Third, the missing information from negative studies which were less frequently published could lead to publication bias, although significant heterogeneity was not detected from the current analysis. Finally, despite the usage of random-effect model and subgroup analysis, some heterogeneity still existed, which may weaken our pooled conclusions.

In conclusion, despite the abovementioned limitations, our meta-analysis indicated that MUC expression was significantly associated with poorer OS, more advanced TNM stage, higher risk of cervical lymph node metastasis, and deeper invasion in patients with HNC for the first time. These findings revealed that the status of MUC could not only distinguish normal and precancerous cells but also could differentiate of a less metastatic cancer form its highly aggressive form. Therefore, patients with highly expression of MUC in HNC may need more radical treatment and rigorous monitoring, especially for those without typical symptoms or signs of metastasis. However, prospective clinical trials that are well-designed, using standardized methods, with long-term follow up, are required to verify the MUC hypothesis in HNC directly.

MATERIALS AND METHODS

Search strategy

This meta-analysis was conducted according to the PRISMA statement [20]. A systematic literature search of PubMed, Embase, and Web of Science database was performed with the following strategy: (“mucin-1” OR “muc-1” OR “episialin”) AND (“oral” OR “mouth” OR “tongue” OR “gingival” OR “pharynx” OR “larynx”) AND (“tumor” OR “cancer” OR “carcinoma” OR “neoplasm” OR “malignant”) AND (“prognostic” OR “prognosis” OR “outcome” OR “survival”) up to September 1, 2016. Citation lists of retrieved articles, including review articles, were additionally reviewed to guarantee the sensitivity of the search process.

Inclusion/exclusion criteria

The criteria for inclusion were listed as following: (1) studies detected MUC expression by immunohistochemistry (IHC) or in situ hybridization (ISH) analysis in human cancer tissues; (2) sufficient information was provided to evaluate the relationship between MUC expression and clinicopathological parameters and/or cancer prognosis; (3) the hazard ratios (HRs) with 95% confidence intervals (CIs) of overall survival (OS) were provided or can be extracted from the Kaplan-Meier curves based on the method reported before [21].

Studies were excluded if they were: (1) reviews, case reports, meta-analyses, letters, conference abstracts without original data; (2) articles which could not extract the relevant data; (3) overlapping articles and those with duplicated data.
Data extraction

The valuable data including surname of the first author, mean age, gender, cancer type, anti-body used, clinicopathological features, prognosis and other relevant data were extracted by two investigators (Lu and Liang) independently and illustrated in Table 1. When the prognosis was only plotted as a Kaplan-Meier curve in some articles, Engauge Digital 9.3 software (from https://sourceforge.net/projects/digitizer/) was applied to digitize and extract the data. A joint decision was offered in the case of any disagreement.

Quality assessment

The quality of included studies was assessed by two independent reviewers (Zhu and Xu) on the basis of the Newcastle-Ottawa scale (NOS) system, which is a 9-point scoring system by judging three main categories: including selection, comparability and outcome. Studies with scores≥6 were regarded as high-quality studies [22-23].

Statistical analysis

Pooled HR with their 95% CI was used to assess the association between MUC expression levels and the cancer prognosis (OS). Meanwhile, the impact of MUC expression on clinicopathological parameters was performed by RR with their 95% CIs. The heterogeneity among the included studies was checked by the chi-squared Q test. \( P > 0.10 \) or \( I^2 < 50\% \) indicated that the differences between the results of various studies were due to chance, and then a fixed-effect model was used. Otherwise, a random-effects model was used. Subgroup analyses were performed to explore the source of heterogeneity. The sensitivity analysis and funnel plots were carried out to evaluate the robustness and the possible publication bias respectively. All statistical tests were performed using Stata version 12.0 (Stata Corp, College Station, TX, USA) with two-tails \( p \) values. A \( p \)-value < 0.05 was considered to be statistically significant.

Trial sequential analysis (TSA)

TSA was employed to test whether the current findings were reliable and conclusive, based on diversity-adjusted threshold for statistical significance. We conducted TSA with assumptions including a plausible overall 5% risk of type I error with a power of 80%. When the cumulative z-curve crosses the trial monitoring boundary or enters the futility zone before the required power is reached, future trials are superfluous considering a sufficient level of evidence is reached [24-25]. Otherwise, the current evidence is insufficient for drawing a conclusion. These analyses were performed using Trial Sequential Analysis Software version 0.9 beta (www.ctu.dk/tsa).

ACKNOWLEDGMENTS

We would like to acknowledge the helpful comments on this paper received from our reviewers. Additionally, we would like to thank all our colleagues in the Department of Oral Maxillofacial-Head and Neck Oncology, Ninth People’s Hospital, Shanghai Jiao Tong University School of Medicine.

CONFLICTS OF INTEREST

All authors declared that they had no conflicts of interests.

GRANT SUPPORT

This work was supported by the Research Grants (16ZR1418800) from Science and Technology Commission of Shanghai Municipality.

REFERENCES

1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. International journal of cancer. 2015; 136: E359-E386.
2. Toporcov TN, Znaor A, Zhang ZF, Yu GP, Winn DM, Wei Q, Vilensky M, Vaughan T, Thomson P, Talamini R, Szeszenia-Dabrowska N, Sturgis EM, Smith E, et al. Risk factors for head and neck cancer in young adults: a pooled analysis in the INHANCE consortium. International journal of epidemiology. 2015; 441: 169-85.
3. Hashibe M, Brennan P, Benhamou S, Castellsague X, Chen C, Curado MP, Dal Maso L, Daudt AW, Fabianova E, Fernandez L, Wünsch-Filho V, Franceschi S, Hayes RB. Alcohol drinking in never users of tobacco, cigarette smoking in never drinkers, and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. Journal of the National Cancer Institute. 2007; 99: 777-790.
4. Little MA, Pokhrel P, Murphy KL, Kawamoto CT, Suguitan GS, Herzog TA. The reasons for betel-quid chewing scale: assessment of factor structure, reliability, and validity. BMC oral health. 2014; 14: 1.
5. Choudhury JH, Ghosh SK. Gene-environment interaction and susceptibility in head and neck cancer patients and in their first-degree relatives: a study of Northeast Indian population. Journal of Oral Pathology & Medicine. 2015; 44: 495-501.
6. Jayaprakash V, Rigual NR, Moysich KB, Loree TR, Nasca MA, Menezes RJ, Reid ME. Chemoprevention of head and neck cancer with aspirin: a case-control study. Archives of Otolaryngology-Head & Neck Surgery. 2006; 132: 1231-1236.

7. Hollingsworth MA, Swanson BJ. Mucins in cancer: protection and control of the cell surface. Nature Reviews Cancer. 2004; 4: 45-60.

8. Yolken RH, Peterson JA, Vonderfecht SL, Fouts ET, Midthun K, Newburg DS. Human milk mucin inhibits rotavirus replication and prevents experimental gastroenteritis. Journal of Clinical Investigation. 1992; 90: 1984.

9. Hattrup CL, Gendler SJ. Structure and function of the cell surface (tethered) mucins. Annu Rev Physiol. 2008; 70: 431-457.

10. Singh PK, Hollingsworth MA. Cell surface-associated mucins in signal transduction. Trends in cell biology. 2006; 16: 467-476.

11. Krishn SR, Kaur S, Smith LM, Johansson SL, Jain M, Patel A, Gautam SK, Hollingsworth MA, Mandel U, Clausen H, Lo WC, Fan WT, Manne U, et al. Mucins and associated glycan signatures in colon adenoma-carcinoma sequence: Prospective pathological implication (s) for early diagnosis of colon cancer. Cancer letters. 2016; 374: 304-314.

12. Rakha EA, Boyce RW, Abd El-Rehim D, Kurien T, Green AR, Paish EC, Robertson JF, Ellis IO. Expression of mucins (MUC1, MUC2, MUC3, MUC4, MUC5AC and MUC6) and their prognostic significance in human breast cancer. Modern Pathology. 2005; 18: 1295-1304.

13. Awaya H, Takeshima Y, Yamasaki M, Inai K. Expression of MUC1, MUC2, MUC5AC, and MUC6 in atypical adenomatous hyperplasia, bronchioloalveolar carcinoma, adenocarcinoma with mixed subtypes, and mucinous bronchioloalveolar carcinoma of the lung. American journal of clinical pathology; 2004, 121: 644-653.

14. Kimura T, Finn OJ. MUC1 immunotherapy is here to stay. Expert opinion on biological therapy. 2013; 13: 35-49.

15. Liu S, Ruan M, Li S, Wang L, Yang W. Increased expression of MUC1 predicts poor survival in salivary gland mucopidermoid carcinoma. Journal of Cranio-Maxillofacial Surgery. 2014; 42: 1891-1896.

16. Weed DT, Gomez-Fernandez C, Pacheco J, Ruiz J, Hamilton-Nelson K, Arnold DJ, Civantos FJ, Zhang J, Yasin M, Goodwin WJ, Carraway KL. MUC4 and ERBB2 expression in major and minor salivary gland mucopidermoid carcinoma. Head & neck. 2004; 26: 353-364.

17. Weed DT, Gomez-Fernandez C, Bonfante E, Lee TD, Pacheco J, Carvajal ME, Goodwin WJ, Carraway KL. MUC4 (sialomucin complex) expression in salivary gland tumors and squamous cell carcinoma of the upper aerodigestive tract. Otolaryngology—Head and Neck Surgery. 2001; 124: 127-141.

18. Siyi L, Shengwen L, Min R, Wenjun Y, Lizheng W, Chenping Z. Increased expression of MUC1 has close relation with patient survivor in highgrade salivary gland mucopidermoid carcinoma. Journal of Oral Pathology & Medicine. 2014; 43: 579-584.

19. Alos L, Lujan B, Castillo M, Nadal A, Carreras M, Caballero M, de Bolos C, Cardesa A. Expression of membrane-bound mucins (MUC1 and MUC4) and secreted mucins (MUC2, MUC5AC, MUC5B, MUC6 and MUC7) in mucopidermoid carcinomas of salivary glands. The American journal of surgical pathology. 2005; 29: 806-813.

20. Kleijnen J, Moher D. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. 2009.

21. Tierney JF, Stewart LA, Gherzi D, Burdett S, Sydes MR. Practical methods for incorporating summary time-to-event data into meta-analysis. Trials. 2007; 8: 1.

22. Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. European journal of epidemiology. 2010; 25: 603-605.

23. Cochrane handbook for systematic reviews of interventions. Chichester: Wiley-Blackwell. 2008.

24. Wetterslev J, Thorlund K, Brok J, Glud C. Trial sequential analysis may establish when firm evidence is reached in cumulative meta-analysis. Journal of clinical epidemiology. 2008; 61: 64-75.

25. Thorlund K, Devereaux PJ, Wetterslev J, Guyatt G, Ioannidis JP, Thabane L, Gluud LL, Als-Nielsen B, Glud C. Can trial sequential monitoring boundaries reduce spurious inferences from meta-analyses? International Journal of Epidemiology. 2009; 38: 276-286.

26. Nitta T, Sugihara K, Tsuyama S, Murata F. Immunohistochemical study of MUC1 mucin in premalignant oral lesions and oral squamous cell carcinoma. Cancer. 2000; 88: 245-254.

27. Handra-Luca A, Lamas G, Bertrand JC, Fouret P. MUC1, MUC2, MUC4, and MUC5AC expression in salivary gland mucoepidermoid carcinoma: diagnostic and prognostic implications. The American journal of surgical pathology. 2005; 29: 881-889.

28. Wreesmann VB, Sieczka EM, Socci ND, Hezel M, Belbin TJ, Childs G, Patel SG, Patel KN, Tallini G, Prystowsky M, Shaha AR, Kraus D, Shah JP, et al. Genome-wide profiling of papillary thyroid cancer identifies MUC1 as an independent prognostic marker. Cancer research. 2004; 64: 3780-3789.

29. Hamada T, Nomura M, Kamikawa Y, Yamada N, Batra SK, Yonezawa S, Sugihara K. DF3 epitope expression on MUC1 mucin is associated with tumor aggressiveness, subsequent lymph node metastasis, and poor prognosis in patients with oral squamous cell carcinoma. Cancer. 2012; 118: 5251-5264.
30. Cros J, Sbidian E, Hans S, Roussel H, Scotte F, Tartour E, Bransu D, Laurent-Puig P, Bruneval P, Blons H, Badoual C. Expression and mutational status of treatment-relevant targets and key oncogenes in 123 malignant salivary gland tumours. Annals of oncology. 2013; 24: 2624-2629.

31. Macha MA, Rachagani S, Pai P, Gupta S, Lydiatt WM, Smith RB, Johansson SL, Lele SM, Kakar SS, Farghaly H, Lee JH, Meza J, Ganti AK, et al. MUC4 regulates cellular senescence in head and neck squamous cell carcinoma through p16/Rb pathway. Oncogene. 2015; 34: 1698-1708.

32. Morari EC, Silva JR, Guilhen ACT, Cunha LL, Marcello MA, Soares FA, Vassallo J, Ward LS. Muc-1 expression may help characterize thyroid nodules but does not predict patients’ outcome. Endocrine pathology. 2010; 21: 242-249.

33. Baek SK, Woo JS, Kwon SY, Lee SH, Chae YS, Jung KY. Prognostic significance of the MUC1 and MUC4 expressions in thyroid papillary carcinoma. The Laryngoscope. 2007; 117: 911-916.

34. Croce MV, Rabassa ME, Pereyra A, Segal-Eiras A. Differential expression of MUC1 and carbohydrate associated antigens in primary and secondary head and neck squamous cell carcinoma. Head & neck. 2008; 30: 647-657.

35. Li XZ, Tian J, Zhang LQ, Wang TD. [Expression of MUC1, MUC2, and Met in papillary thyroid carcinoma and their clinical significance]. Journal of Shandong University Health Sciences. [Article in Chinese]. 2005; 43: 346-349.

36. Renaud F, Gnemmi V, Devos P, Aubert S, Crépin M, Coppin L, Ramdane N, Bouchindhomme B, d’Herbomez M, Van Seuningen I, Do Cao C, Pattou F, Carnaille B, et al. MUC1 expression in papillary thyroid carcinoma is associated with BRAF mutation and lymph node metastasis; the latter is the most important risk factor of relapse. Thyroid. 2014; 24: 1375-1384.

37. Paleri V, Pearson JP, Bulmer D, Jeannon JP, Wight RG, Wilson JA. Expression of mucin gene products in laryngeal squamous cancer. Otolaryngology—Head and Neck Surgery. 2004; 131: 84-88.

38. Hamada T, Wakamatsu T, Miyahara M, Nagata S, Nomura M, Kamikawa Y, Yamada N, Batra SK, Yonezawa S, Sugihara K. MUC4: a novel prognostic factor of oral squamous cell carcinoma. International Journal of Cancer. 2012; 130: 1768-1776.

39. He F, Li H, Li WS, Dong XH. [Expression of mucin-1 and -catenin in papillary thyroid carcinoma and the clinical significance thereof]. [Article in Chinese]. Zhonghua Yixue Zazhi. 2009; 89: 393-396.

40. Croce MV, Rabassa ME, Price MR, Segal-Eiras A. MUC1 mucin and carbohydrate associated antigens as tumor markers in head and neck squamous cell carcinoma. Pathology & Oncology Research. 2001; 7: 284-291.

41. National Comprehensive Cancer Network. NCCN Clinical Practice Guidelines in Oncology: Head and Neck Cancers. (Version 1.2007). 2007d.

42. Rossi EA, McNeer R, Price-Schiavi SA, Van den Brande JM, Komatsu M, Thompson JF, Carraway CA, Fregien NL, Carraway KL. Sialomucincomplex, a heterodimeric glycoprotein complex: expressionas a soluble, secretable form in lactating mammary glandand colon. J Biol Chem. 1996; 271: 33476-33485.

43. Roy LD, Sahraei M, Subramani DB, Besmer D, Nath S, Tindel TL, Bajaj E, Shanmugam K, Lee YY, Hwang SI, Gendler SJ, Mukherjee P. MUC1 enhances invasiveness of pancreatic cancer cells by inducing epithelial to mesenchymal transition. Oncogene. 2011; 30: 1449-1459.

44. Mohr AM, Bailey JM, Lewallen ME, Liu X, Radhakrishnan P, Yu F, Tapprich W, Hollingsworth MA. MUC1 regulates expression of multiple microRNAs involved in pancreatic tumor progression, including the miR-200c/141 cluster. PloS one. 2013; 8: e73306.

45. Hayashi T, Takahashi T, Motoya S, Ishida T, Itoh F, Adachi M, Hinoda Y, Imai K. MUC1 mucin core protein binds to the domain 1 of ICAM-1. Digestion. 2001; 63: 87-92.

46. Hollingsworth MA, Swanson BJ. Mucins in cancer: protection and control of the cell surface. Nature Reviews Cancer. 2004; 4: 45-60.

47. Jepson S, Komatsu M, Haq B, Arango ME, Huang D, Carraway CA, Carraway KL. Muc4/sialomucin complex, the intramembrane ErbB2 ligand, induces specific phosphorylation of ErbB2 and enhances expression of p27 (kip), but does not activate mitogen-activated kinase or protein kinaseB/Akt pathways. Oncogene. 2002; 21: 7524-7532.