Trends in ExPEC serogroups in the UK and their significance

H. Ciesielczuk1,2 · C. Jenkins3 · M. Chattaway3 · M. Doumith2 · R. Hope4 · N. Woodford1,2 · D. W. Wareham1

Received: 26 April 2016 / Accepted: 6 June 2016 © The Author(s) 2016. This article is published with open access at Springerlink.com

Abstract Extra-intestinal pathogenic Escherichia coli are a significant cause of urinary tract infection and bacteraemia within the UK. We sought to identify the serogroups of 658 E. coli isolates collected in the UK between January 2011 and March 2012, to better understand the ExPEC population and understand the relevance of serogroups in this pathotype. Isolates were typed and serogroup identified using established phenotypic and molecular methods. Sixty-two serogroups were identified; 54 among urinary isolates and 35 among bloodstream isolates. However, serogroups O25, O6, and O2 dominated both infection types. These serogroups were linked to the major ExPEC STs as follows: ST131-O25, ST73-O6, ST127-O6, and ST95-O2. The serogroup data from this study have increased our understanding of the ExPEC population in the UK, but also highlighted key ST–serogroup relationships within the major ExPEC clones. These data can be used to guide vaccine design and in the development of laboratory diagnostic tests targeting the ExPEC population.

Introduction

Multi-locus sequence typing (ST) and phylogenetic grouping (phylogroup) have become established methods for typing extra-intestinal pathogenic Escherichia coli (ExPEC) strains. In the UK the ST131/B2, ST127/B2, ST95/B2, ST73/B2, and ST69/D lineages are the most frequently recovered from bloodstream infections (BSI) and urinary tract infections (UTI) [1–3]. However, prior to the widespread use of these molecular methods, serotyping was extensively used to characterise pathogenic E. coli, [4, 5] as well as to inform the development of E. coli vaccines [6].

In the UK, the incidence of E. coli BSI is increasing year on year (https://www.gov.uk), while E. coli remains the most common cause of UTI [7]. Therefore, a successful ExPEC vaccine has the potential to reduce the incidence of BSI and UTI attributable to dominant strains in the UK and around the world [8–10].

Over time a number of associations between serotypes, STs, and E. coli pathotypes have been observed, such as ExPEC O25b:H4 (ST131), enterohaemorrhagic E. coli (EHEC) O157:H7 (ST11), EHEC O104:H4 (ST678), and avian pathogenic E. coli (APEC) O1 (ST95) [11–14]. Serotyping remains an important method in identifying clinically-significant ExPEC strains and for the study of local ExPEC populations in laboratories lacking molecular capability [4]. However, few UK studies of ExPEC have evaluated the importance of serotype in relation to ST, phylogroup, and infection type [1–3, 15].
Therefore, we resolved to understand the relevance of serogroup within the UK ExPEC population and identify important links with infection type and sequence type.

Materials and methods

Bacterial strains

The 658 UK ExPEC isolates studied were part of a larger collection described previously [16]. They comprised 379 urinary isolates collected predominantly from patients in East London (Barts Health NHS Trust) and 279 bloodstream isolates collected as part of the BSAC Bacteraemia Resistance Surveillance Programme (2011). This included isolates from 38 centres located around the UK and Republic of Ireland [17]. The purpose of these two collections was to identify any common trends among ExPEC isolates recovered from urine versus isolates recovered from BSI.

The urinary isolates were recovered from episodes of complicated \((n = 170, 45 \%)\) and uncomplicated UTI \((n = 161, 42 \%)\), as well as cases of asymptomatic bacteruria \((n = 48, 13 \%)\). For bloodstream isolates, the presumed focus of BSI was genitourinary \((n = 118, 42 \%)\), gastrointestinal \((n = 38, 14 \%)\), chest \((n = 14, 5 \%)\), catheter-related line \((n = 4, 1 \%)\) or skin and soft-tissue infection \((n = 2, <1 \%)\), with the primary source unknown in 37 \(% (n = 103)\).

Phenotypic serogrouping, phylogrouping and sequence typing PCR

Conventional serogrouping was performed using the method of Gross and Rowe [18]. Phylogroup was determined by PCR [19]. The major ExPEC STs (ST131, ST95, ST73 and ST69) were also identified by PCR [20], along with an additional primer pair to detect ST127. The forward \((5'-CGCATAACAGGATTGTCTGGG-3')\) and reverse \((5'-GCTATTCTACGGGCATTGTG-3')\) primers for ST127 generated an amplicon of 404 bp.

| Table 1 Patient characteristics for all the study isolates |
|-----------------|-----------------|-----------------|
|                 | Urinary isolates (%) | Bloodstream isolates (%) |
| **Sex**         | Female           | Male            |
|                 | 312 (82)         | 149 (53)        |
|                 | 67 (18)          | 130 (47)        |
| **Age (years)** | Range            | 1–99            | 0–97            |
|                 | Median           | 39              | 73              |
| Healthcare setting | Community-associated | 337 (89) | 190 (68) |
|                 | Hospital-associated | 42 (11)  | 89 (32)       |
| **Total**       | 379              | 279             |

The number of isolates is listed for each category, with the percentage in parenthesis.
The proportion of isolates comprising the most prevalent serogroups (O2, O6, and O25) differed by infection type. Twenty-nine percent (n = 110/379) of urinary isolates comprised these serogroups, with O6 the most commonly identified, followed by O25. This pattern was reversed in the 43% of bloodstream isolates comprising these three serogroups (n = 120/279), with O25 the most commonly identified, followed by O6.

Antibiotic susceptibility data was performed as part of a previous study [16] and was reviewed for these three serogroups. Serogroup O25 was the most resistant of the three serogroups, followed by O2. Notably, BSI isolates within serogroups O6 and O25 were more resistant than the urinary isolates, but for O2 isolates this was reversed and the urinary isolates were more resistant than those from BSI.

Relationship of serogroup to phylogroup

Phylogroups A, B1, B2, and D comprised 31, 10, 31, and 22 different serogroups respectively (Fig. 1). It is noteworthy that 19 serogroups were only detected in phylogroups A and B1, while 27 serogroups were only detected in phylogroups B2 and D (Fig. 1). Within all four phylogroups, the urinary isolates displayed a greater variety of serogroups than BSI isolates. However, in phylogroups B2 and D an even greater number of serogroups were identified in both infection types. The most common serogroups in each of the phylogroups were O8 (A and B1), O25 (B2), and O44 (D).

Relationship between phylogroup and ST

The phylogroup and ST of the isolates studied are detailed in Table 2. ST131, ST95, and ST73 were only detected in phylogroups B2 and D, while ST69 was detected among all phylogroups except A, and ST127 was detected in phylogroups A and B2. With the exception of ST131, all other STs were identified more frequently in urinary isolates than BSI isolates.

One isolate was not assigned a phylogroup or a major ExPEC ST, but was identified as having serogroup O18ab.

---

**Fig. 1** Serogroups identified in each of the four *E. coli* phylogroups. ‘Multiple’ lists the serogroups identified in three or four of the phylogroups. Only O86 was identified in all four phylogroups. Serogroups only identified in urinary isolates are coloured blue, serogroups only identified in bloodstream isolates are coloured red and serogroups identified in both are in bold. The most common serogroup in each of the phylogroups was O8 (A and B1), O25 (B2) and O44 (D).
**Relationship of serogroup to ST**

The major ExPEC STs demonstrated reduced serogroup diversity, with just 29 of the 62 serogroups identified, as detailed in Table 3. ST131 and ST127 displayed least serogroup diversity (n = 5 each), while ST69 displayed the greatest diversity of serogroups (n = 15). With regard to infection type, both ST131 and ST73 had greater serogroup diversity within their BSI isolates than urinary isolates, whereas this pattern was reversed for ST95 and ST69, while ST127 isolates had equal distribution of serogroups within both infection types. Five serogroups were only identified in the major ExPEC STs: O17, O19a, O110, O125ab, and O136. Within the three most prevalent serogroups; ST95 comprised the majority of O2 isolates (n = 25/44, 57 %), and ST73 comprised more than half the O6 isolates (n = 42/75, 56 %), followed by ST127 (n = 21/75, 28 %). ST131 comprised the vast majority of O25 isolates (n = 89/104, 86 %).

**Discussion**

This is the first large-scale UK study reporting on the serogroup diversity of E. coli causing UTI and BSI and the relationship between serogroup, phylogroup, and major ExPEC STs. Urinary isolates demonstrated greater serogroup diversity than bloodstream isolates, representing the ability of many E. coli strains to invade the bladder, but with fewer able to invade the bloodstream successfully [8]. Similarly, the less pathogenic phylogroups A and B1 comprised mostly urinary isolates, with 15 of the 19 phylogroup-specific serogroups (79 %) detected, while a smaller proportion of phylogroup-specific serogroups (n = 16/27, 59 %) were detected within the virulent phylogroups B2 and D bloodstream isolates.

In keeping with this pattern, the major ExPEC STs comprised mostly phylogroup B2 and D isolates, and displayed a limited repertoire of serogroups. This was notable for ST131/B2 (O25) and ST73/B2 (O6) which were prevalent among bloodstream isolates, while ST95/B2 (O2) and ST69/D (O77) were most frequently identified in urinary isolates. The link between phylogroup and major ExPEC ST is well established [21–23], but data from this study highlights that the relationship between serogroup and ST is just as important, as evidenced by the dominant serogroups found: ST131-O25 (B2), ST73-O6 (B2), and ST95-O2 (B2). This important link between serogroup and ST may have developed either through clonal expansion (ST131-O25) [13] or due to virulence-associated traits (ST127-O6 and ST73-O6) [24], but either way, identification of serogroup in E. coli isolates provides an alternative means to identify major ExPEC clones and understand the local ExPEC population.

While the serogroup repertoire for the major UK ExPEC STs may be limited, it appears to be changing when compared to previously published reports [1, 25]. This could represent the large sample size examined here or the fact that the UK ExPEC STs have evolved. Away from the major known STs, analysis of this UK collection identified 21 serogroups not previously documented among ExPEC isolates, again highlighting a limitation of studies that have focused on small populations or referred strain collections. These newly identified ExPEC serogroups included O12, O31, O32, O36, O37, O39, O40, O46, O56, O62, O81, O83, O87, O93, O110, O118, O135, O150, O158, O162, and O182 [4, 26, 27]. These novel ExPEC serogroups highlight the importance of fully understanding the local ExPEC population, clonal or otherwise, for future vaccine development, for genotypic

---

**Table 2** ExPEC phylogroup and sequence type for all study isolates

| Phylogroup | Urinary isolates (%) | Bloodstream isolates (%) |
|------------|----------------------|--------------------------|
| A          | 54 (14)              | 22 (8)                   |
| B1         | 24 (6)               | 7 (3)                    |
| B2         | 215 (57)             | 202 (72)                 |
| D          | 86 (23)              | 47 (17)                  |
| NK         | 0                    | 1 (<1)                   |
| Major ST   |                      |                          |
| ST131      | 41 (11)              | 74 (27)                  |
| ST127      | 16 (4)               | 12 (4)                   |
| ST95       | 56 (15)              | 34 (12)                  |
| ST73       | 40 (11)              | 39 (14)                  |
| ST69       | 50 (13)              | 14 (5)                   |
| Other      | 176 (46)             | 106 (38)                 |

**Table 3** Serogroups identified in the major ExPEC sequence types by infection type

| ST131 | ST127 | ST95 | ST73 | ST69 |
|-------|-------|------|------|------|
| Urinary |       |      |      |      |
| O16 O25 | O1 O6 O31 O110 O150 O153 | O4 O158 | O17 O45 O73 O102 O117 O150 O153 |
| Bloodstream | O19a O136 O153 | O11 O27 | O8 O18ac O27 | O27 O125ab |
| Both | **O16 O25** | **O6** | **O2** O4 O16 O18ac | **O2** O6 O22 O25 | **O11 O15 O25 O44 O77 O86** |

Serogroups in bold text comprised >50 % of all isolates in that ST and the most common serogroups are underlined.
laboratory tests targeting this particular microbial population, and to further understand the clinical importance and virulence of emerging strains of *Escherichia coli* and the diseases they cause.

To conclude, this large study detailed the serogroup diversity of ExPEC isolates causing UTI and BSI in the UK, with particular reference to major STs. Data collected here can be used to expand our knowledge of ExPEC, as well as to inform vaccine development.

**Acknowledgments** We thank the British Society for Antimicrobial Chemotherapy for kindly providing *E. coli* bloodstream isolates from the BSAC Bacteraemia Resistance Surveillance Programme (2011), and all the staff at PHE’s Gastrointestinal Bacteria Reference Unit for their guidance and patience during the serogrouping process.

The multilocus sequence type PCR is currently being evaluated for commercialisation (Patent No.1315527.0) by Public Health England.

**Compliance with ethical standards**

**Funding** This work was performed as part of a PhD study funded by PHE.

**Conflict of interest** The authors declare that they have no conflict of interest.

**Ethical approval** This article does not contain any studies with human participants or animals performed by any of the authors.

**Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

**References**

1. Lau S, Reddy S, Chessbrough J, Bolton F, Willshaw G, Cheasty T et al (2008) Major uropathogenic *Escherichia coli* strain isolated in the North West of England identified by multilocus sequence typing. J Clin Microbiol 46(3):1076–1080

2. Gibreel T, Doddson A, Chessbrough J, Fox A, Bolton F, Upton M (2012) Population structure, virulence potential and antibiotic susceptibility of uropathogenic *Escherichia coli* from Northwest England. J Antimicrob Chemother 67(2):346–356

3. Horner C, Fawley W, Morris K, Parnell P, Denton M, Wilcox M (2014) *Escherichia coli* bacteraemia: 2 years of prospective regional surveillance (2010–12). J Antimicrob Chemother 69(1):91–100

4. Emanghorashlu F, Farshad S, Kalani M (2011) Relationship between O serotype and virulent genes in *Escherichia coli* causing urinary tract infections. Iran J Kidney Dis 5(4):234–237

5. Johnson J, Russo T (2002) Uropathogenic *Escherichia coli* as agents of diverse non-urinary tract extraintestinal infections. J Infect Dis 186(6):859–864

6. Poolman JT, Wacker M (2016) Extraintestinal pathogenic *Escherichia coli*, a common human pathogen: challenges for vaccine development and progress in the field. J Infect Dis 213(1):6–13

7. Chakupurakal R, Ahmed M, Sobithadevi D, Chinnappan S, Reynolds T (2010) Urinary tract pathogens and resistance pattern. J Clin Pathol 63(7):652–654

8. Banerjee R, Johnston B, Lohse C, Chattopadhyay S, Tchesnokova V, Sokurenko EV et al (2013) The clonal distribution and diversity of extraintestinal *Escherichia coli* varies according to patient characteristics. Antimicrob Agents Chemother 57(12):5912–5917

9. Abdallah KS, Cao Y, Wei DJ (2011) Epidemiologic investigation of extra-intestinal pathogenic *E. coli* (ExPEC) based on PCR phylogenetic group and fimH single nucleotide polymorphisms (SNPs) in China. Int J Mol Epidemiol Genet 2(4):339–353

10. Russo T, Johnson J (2000) Proposal for a new inclusive designation for extraintestinal pathogenic isolates of *Escherichia coli*: ExPEC. J Infect Dis 181(5):1753–1754

11. Underwood AP, Dallman T, Thomson NR, Williams M, Harker K, Perry N et al (2013) Public health value of next-generation DNA sequencing of enterohemorrhagic *Escherichia coli* isolates from an outbreak. J Clin Microbiol 51(1):232–237

12. Monecke S, Mariani-Kurkdjian P, Ewig FX, Bielec C, Slickers P et al (2011) Presence of enterohemorrhagic *Escherichia coli* strain isolated in France prior to 2011. Appl Environ Microbiol 77(24):8784–8786

13. Nicolas-Chanoine M, Blanco J, Leflon-Guibout V, Demarty R, Alonso M, Canica M et al (2008) Intercontinental emergence of *Escherichia coli* clone O25:H4-ST131 producing CTX-M-15. J Antimicrob Chemother 61(2):273–281

14. Johnson JR, Johnston B, Clabots CR, Kuskowski MA, Roberts E, DeRoy C (2008) Virulence genotypes and phylogenetic background of *Escherichia coli* serogroup O6 isolates from humans, dogs, and cats. J Clin Microbiol 46(2):417–422

15. Day M, Doumith M, Abernethy J, Hope R, Reynolds R, Wain J et al (2016) The population structure of *Escherichia coli* causing bacteraemia in the UK and Ireland between 2001 and 2010. J Antimicrob Chemother [Accepted for publication]

16. Ciesielczuk H, Hornsey M, Choi V, Woodford N, Wareham D (2016) Extraintestinal pathogenic isolates of *Escherichia coli* phylogenetic groups. J Clin Microbiol 54(3):755–765

17. Reynolds R, Hope R, Williams L (2008) Survey, laboratory and ethical approval. This article does not contain any studies with human participants or animals performed by any of the authors.

18. Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

19. Gross R, Rowe B (1985) Serotyping of *Escherichia coli*. In: Sussman M (ed) The virulence of *Escherichia coli*. Cambridge University Press, Cambridge, pp 345–360

20. Doumith M, Day M, Hope R, Wain J, Woodford N (2012) Improved multiplex PCR strategy for rapid assignment of the four major *Escherichia coli* phylogenetic groups. J Clin Microbiol 50(9):3108–3110

21. Doumith M, Day M, Ciesielczuk H, Hope R, Underwood A, Reynolds R et al (2014) Rapid identification of major *Escherichia coli* sequence types causing urinary tract and bloodstream infections. J Clin Microbiol 53(1):160–166

22. Johnson JR, Owens KL, Clabots CR, Weissman SJ, Cannon SB (2006) Phylogenetic relationships among clonal groups of extraintestinal pathogenic *Escherichia coli* as assessed by multi-locus sequence analysis. Microbes Infect 8(7):1702–1713

23. Novais A, Vuotto C, Pires J, Montenegro C, Donelli G, Coque TM et al (2013) Diversity and biofilm-production ability among isolates of *Escherichia coli* phylogroup D belonging to ST69, ST393 and ST405 clonal groups. BMC Microbiol 13:144

24. Cagnacci S, Gualco L, Debbia E, Schito G, Marchese A (2008) European emergence of ciprofloxacin-resistant *Escherichia coli* clonal groups O25:H4-ST131 and O15:K52:H1 causing community-acquired uncomplicated cystitis. J Clin Microbiol 46(8):2605–2612
24. Alghoribi MF, Gibreel TM, Dodgson AR, Beatson SA, Upton M (2014) Galleria mellonella infection model demonstrates high lethality of ST69 and ST127 uropathogenic E. coli. PLoS One 9(7): e101547

25. Manges A, Tabor H, Tellis P, Vincent C, Tellier P (2008) Endemic and epidemic lineages of Escherichia coli that cause urinary tract infections. Emerg Infect Dis 14(10):1575–1583

26. Melzer M, Petersen I, Cheasty T (2008) The difference in serotypes between extended-beta-lactamase (ESBL) and non-ESBL-producing E. coli blood culture isolates at a UK district general hospital. J Hosp Infect 68(4):367–369

27. Ananias M, Yano T (2008) Serogroups and virulence genotypes of Escherichia coli isolated from patients with sepsis. Braz J Med Biol Res 41(10):877–883