Insight on CD151 (Cluster of Differentiation 151)

Tetraspanins are four-transmembrane-spanning proteins with short cytoplasmic N- and C-termini and one small and one large extracellular domain, namely (EC1 and EC2), with a unique cysteine motif in EC2 domain. They are highly expressed on cell surface and/or intracellular vesicle. Palmitoylation of intracellular and juxtamembrane cysteine of tetraspanins along with specific integrins contributes to tetraspanin complex formation. This complex formation protects tetraspanins from lysosomal degradation and promotes increased cell–cell interaction.

CD151 (glycoprotein-27 (GP-27)/Red blood cell antigen 2 (MER 2)/platelet-endothelial tetraspan antigen-3 (PETA-3)/Raph blood group antigen (RAPH)/Membrane Glycoprotein SFA-1 (SFA-1)/tetraspanin-24 (TSPAN-24)) is a plasma membrane protein that belongs to tetraspanin superfamily. CD151 forms tetraspanin web with integrins such as α3β1, α6β1, α7β1, and α6β4; membrane receptors; intracellular signaling molecules such as phosphoinositide 4-kinase (PI4K) and protein kinase C (PKC); immunoglobulin super family proteins and other tetraspanins such as CD9, CD81 and CD63. These tetraspanin-enriched microdomains (TEMs) serve as molecular facilitator.

CD151 contains a potential tyrosine-based sorting motif in the C-terminal domain. The YXXΦ (tyrosine linked with hydrophobic amino acid) motif (where Φ is a hydrophobic residue) is required for endocytosis and sorting of proteins from the trans-Golgi network (TGN) to lysosomes for degradation. A mutation in the CD151 YXXΦ motif diminishes CD151 internalization and affects integrin-dependent cell migration. The major functions of CD151 are maintenance of epithelial cell integrity, wound healing, platelet aggregation, regulation of membrane fusion, trafficking, cell motility, angiogenesis and tumor metastasis. It is normally expressed in endothelial cells and platelets and frequently overexpressed in cancer cells where it is functionally associated with cancer progression and metastasis.

Genetics of CD151

CD151 (Raph blood group) gene is represented as CD151, which is located on the short (p) arm of chromosome 11 at position 15.5. More precisely, CD151 gene is located from base pair 832, 951 to base pair 838, 834 on chromosome 11. It is an autosomal gene with sexually dimorphic expression, involved in signal transduction, cell proliferation and death. It is also involved in differential neural development, cognitive function and neurological diseases.

Association of CD151 with Integrins and Other Proteins

Metastasis is facilitated by cell–cell interactions between tumor cells and endothelium in which cell-adhesion molecules, such as integrins and selectins, play an important role. Hemler has demonstrated that CD151 interacts directly with α3β1 and α6β4 integrins through their palmitoylated...
cysteine residues. CD151 acts as master regulators of α6β4, α6β1 and α3β1 integrin-assembly into TEMs.13 The major laminin-332 cellular receptors with α3β1 and α6β4 integrin-dependent adhesion, migration and signaling are impaired or altered by CD151 excision, indicating the vital role of CD151 in association with integrins.14,15 Mapping of the integrin-CD151 association was performed using chimeric α6/α3 integrins and CD151/NAG2 TM4SF proteins, which showed an interaction of amino acids from 570 to 705 at the extracellular sites of α6 integrin and amino acids from 186 to 217 on a large extracellular loop of CD151.16 Devbhandari et al17 reviewed the role of CD151/integrin β1 complex in cancer metastasis. Yauch et al1 reported that specific site on the large extracellular loop of CD151 associates with α3β1 integrin with unusually high stoichiometry, proximity and stability. In the same year, Sterk et al18 reported that CD151 uses the same site for interaction with α6β1, α6β4 and other integrins.

**Role of CD151 in De Novo Carcinogenesis**

CD151, in association with laminin and laminin-binding integrins, involves in the regulation of carcinogenesis. Li et al19 evaluated the role of CD151 in de novo carcinogenesis, multiplicity and progression of skin squamous cell carcinoma (SCC) in comparison with normal cells. CD151 supports survival and proliferation of keratinocytes by activation of transcription factor STAT3, a regulator of cell proliferation and apoptosis. CD151 regulates α6β4 distribution by enhancing protein kinase Cα (PKCα)—α6β4 integrin association and PKC-dependent β4 S1424 phosphorylation. CD151 in complex with PKCα enhances invasive behavior by phosphorylating β4 integrin, thus affects subcellular localization and epithelial disruption. In addition, CD151 knockout mice showed a decrease in tumor latency, tumor incidence, multiplicity and size. Hence, CD151 targeting may be therapeutically beneficial in cancer therapy.

**CD151 and Cancer Metastasis**

Deregulation of various tetraspanins is reported in human malignancy.20 Overexpression of CD151 was also reported to associate with poor prognosis of lung,21 colon,22 esophageal,23 pancreatic24 and endometrial cancers.25 In addition, several evidences have supported the contribution of CD151 in cancer metastasis.26 Loss of CD151 decreased the integrin-mediated cell migration, spreading, and invasion through FAK and Rac1-mediated signaling.13 Earlier reports have shown that down-regulation of CD151 by short-hairpin RNA decreased the tumorigenicity and communication between tumor and endothelial cells. This emphasizes CD151 as a potential prognostic marker.27 CD151 also modulates the activity of cytokine-like transforming growth factor-β (TGF-β). CD151 in association with dimerized TGF-β receptor promotes invasion and metastasis through the activation of Smad2/3, c-Akt, Erk1/2, JNK, JUN and matrix metalloproteinase-9 (MMP-9) signaling pathways (Fig. 1). Sadej et al reported that CD151 is a positive regulator of TGF-β-induced signaling in cancer metastasis.28

Studies on CD151-null mice showed impaired pathological angiogenesis without vascular defect during normal development.29 Adhesion-dependent activation of endothelial cells caused diminished expressions of PKC/c-Akt, e-NO, Rac and Cdc42, which are important signaling pathways of angiogenesis and cytoskeleton reorganization without altering the expression of Raf, ERK, p38, MAP kinase, FAK and Src. CD151 regulates cytoskeletal reorganization, invasion and cell-adhesion functions of endothelial cells during pathological angiogenesis by modulating laminin-binding integrin-mediated activation of PI3K/Akt, JNK and PKC pathways (Fig. 2). Evaluation of prognostic significance of CD151 expression with estrogen receptor (ER), progesterone receptor (PR), human epidermal growth factor receptor-2 (“HER-2”) and E-cadherin indicated a strong concordance of CD151...
CD151—a striking marker for cancer therapy

Biomarkers in Cancer 2015:7

Kohno et al demonstrated that CD151 also enhanced cell motility, invasion and metastasis through focal adhesion kinase (FAK). Haeuw et al reported that monoclonal antibodies (mAbs) targeted to CD151 inhibited cell motility. Similarly, Fei et al reported that CD151-AAA mutant inhibited cell-proliferation, migration and chemotaxis in HepG2 cells. Genetic ablation of CD151 inhibited metastasis in a transgenic mouse model without showing any noticeable effect on expression of markers associated with proliferation, apoptosis or angiogenesis in primary tumors. Shanmukhappa et al have demonstrated for the first time that CD151siRNA and anti-CD151 antibodies significantly reduced the porcine reproductive and respiratory syndrome virus (PRRSV) infection. One recent study showed that simultaneous inhibition of CD9/CD81 and CD151 has a profound inhibitory effect on cancer metastasis through α3β1-PKCα-mediated signaling in MDA-MB 231 breast cancer cells.

Several studies have demonstrated that CD151 is involved in cell-adhesion and formation of hemidesmosomes; thus, inhibition of CD151 expression by hypoxia decreased the cell–cell interaction and cell–matrix adhesion. A study by RR Malla et al have demonstrated that silencing of cathepsin B and uPAR using siRNA decreased the interactions of CD151 with laminin–binding integrin α3β1 and inhibits cell-adhesion and invasion in glioma.

**CD151 Pathogenesis and Clinical Studies**

CD151 is implicated in pathological processes associated with cancer progression, neoangiogenesis and epithelial-mesenchymal transition (EMT). CD151 overexpressing cancer cells acquired high migratory abilities, which are crucial for tumor cell invasion. Overexpression of CD151 was also reported to associate with intercellular adhesiveness and wound healing. CD151 plays a crucial role in the interaction of podocyte-GBM binding adhesion receptor and integrin α3β1. Deletion of CD151 leads to reduced adhesiveness at cell–matrix interface, leading to glomerular nephropathy.

CD151-null mice showed diminished tumor cell residence in the lungs after injection with Lewis lung carcinoma, which may be because of inhibition of angiogenesis. In vitro studies showed that tumor-endothelial adhesion, tumor-transendothelial migration and tumor-induced permeability were defective in CD151-null endothelial cells. Blocking of CD151 using specific mAbs inhibited invasion without affecting primary tumor growth and tumor cell arrest or growth at the secondary site. Knockdown of CD151 inhibited downstream signaling through Akt, ErK1/2 and FAK and markedly sensitizes ErbB2+ cancers.

Kwon et al reported that overexpression of CD151 may be a potential molecular therapeutic target in advanced stages of breast cancer. Recent clinical studies have demonstrated that a positive correlation exists between CD151 expression and progression of cancer cells. An interesting fact has been reported recently that integrin-free CD151 can promote tumor cell migration without binding to integrin. This study suggests that CD151 can control migration, independent of integrin association.

**Expression Levels of CD151**

Comparative analysis of tetraspanin expression in various types of cancers revealed that CD151 is highly expressed in solid tumor compared with non-solid tumor (Table 1). CD151 is usually localized to the basal and lateral junctions of tumor...
cells.\(^5\) Solid tumors exhibit heterogeneity of neoplastic and normal cells at histological, genetic and gene expression levels. Considerable heterogeneity of CD151 expression was reported in various tumor tissues. The intensity of staining was noticeably weaker in well-differentiated cells of oral SCC. The gradient of CD151 expression was particularly prominent in the invasive front of tumors.\(^6\) Overexpression of CD151 can be correlated with large tumor size, depth of invasion and advanced stage of tumor.\(^7\) A number of investigations have suggested that high CD151 gene expression in cancer is associated with poor prognosis.\(^8\) Tokuhara et al have reported that survival rate of patients with CD151-positive tumors was much lower than that of CD151-negative patients.\(^9\) For optimal separation between low and high risk for overall survival, prognostic factors are usually considered as an optimized cutoff point.\(^10\) The cutoff point of CD151 in ER negative breast cancer patients was reported as 14%\(^11\) and 31%,\(^12\) in lung cancer patients as 50%,\(^13\) and in gastric cancer patients as >50%,\(^14\) using immunohistochemical and RT-PCR analysis. These studies show the cutoff values to identify CD151 positivity, which may vary based on cancer type. However, large-scale prospective and retrospective studies on various cancers established CD151 as a prognostic marker for cancer therapy with minimal side effects on normal cells.\(^15\)

**Future Prospects**

This review explores the importance of CD151 in the maintenance of cellular integrity and cell communication. It also imparts the role of CD151 in cancer cell migration, invasion, and metastasis. Thus, CD151 is a crucial target for cancer therapy. Disruption of tetraspanin web by targeting CD151 may affect the signaling pathways involved in cell survival and cancer progression. CD151 can be downregulated using mAbs, shRNA or gene knockout, which may find application in cancer therapy. Understanding the mechanism of palmitoylation of CD151 may give a wide scope for future cancer research.

### Abbreviations

CD151, cluster of differentiation 151; GP-27, glycoprotein-27; PETA-3, platelet-endothelial tetraspan antigen-3; TSPAN-24, tetraspanin-24; TEM, tetraspanin-enriched microdomains; ECM, extracellular membrane; MMP, matrix metalloproteinase.

### Author Contributions

Conceived and designed the experiments: RM. Analyzed the data: SK. Wrote the first draft of the manuscript: GV. Contributed to the writing of the manuscript: AB. Agree with the results and conclusions: SK. Jointly developed the structure and arguments for the paper: RM. Made critical revisions and approved final version: RM and VRD.

All authors reviewed and approved of the manuscript.

### REFERENCES

1. Orłowski E, Chaud R, Yip J, et al. A platelet tetraspanin superfamily member, CD151, is required for regulation of thrombus growth and stability in vivo. *J Thromb Haemost*. 2009;7:2074–2084.
2. Sterck LM, Gueijen CA, Van Den Berg JG, Claessen N, Weening JJ, Sonnenberg A. Association of the tetraspanin CD151 with the laminin-binding integrins α3β1, α6β1, α6β4 and 67β1 in cells in culture and in vivo. *J Cell Sci*. 2002;115:1161–1173.
3. Yauch RL, Berditchevski F, Harler MB, Reichner J, Hemler ME. Highly stoichiometric, stable and specific association of integrin α3β1 with CD151 provides a major link to phosphatidylinositol 4-kinase and may regulate cell migration. *Mutat Biol Cell*. 1998;9:2751–2765.
4. Zhang XA, Bontrager AL, Hemler ME. TM4SF proteins associate with activated PKC and link PKC to specific β1 integrins. *J Biol Chem*. 2001;276:25005–25013.
5. Zoller M. Tetraspanins: push and pull in suppressing and promoting metastasis. *Nat Rev Cancer*. 2009;9:40–55.
6. Wang H-X, Li Q, Sharma C, Knoblich K, Hemler ME. Tetraspanin protein contributions to cancer. *Biochim Biophys Acta*. 2011;1814:547–552.
7. Liu L, He B, Liu WM, Zhou D, Cox JV, Zhang XA. Tetraspanin CD151 promotes cell migration by regulating integrin trafficking. *J Biol Chem*. 2007;282:31631–31642.
8. Baleato RM, Guthrie PL, Gubler MC, Ashman LK, Roselli S. Deletion of CD151 results in a strain-dependent glomerulonephritis due to severe alterations of the glomerular basement membrane. *Am J Pathol* 2008;173:927–937.
9. Yoo SH, Lee K, Cha JY, Moon KC. CD151 expression can predict cancer progression in clear cell renal cell carcinoma. *Histopathology*. 2011;58(2):191–197.
10. Whittock NV, McLean WH. Genomic organization, amplification, fine mapping, and intragenic polymorphisms of the human hemidesmosomal tetraspanin CD151 gene. *Biochim Biophys Acta*. 2001;2812(2):425–430.
11. Armstrong C, Moreira D, Bödinger K, Jimenez G, Taniguchi S, Tsai HW. Identification of sexually dimorphic genes in the neonatal mouse cortex and hippocampus. *Brain Res*. 2014;1562:23–38.
12. Hemler ME. Integrin-associated proteins. *Curr Opin Cell Biol*. 1998;10:578–585.
13. Yang XY, Richardson AL, Torres-Arzayus M, et al. CD151 accelerates breast cancer by regulating alpha 6 integrin functions, signalling, and molecular organization. *Cancer Res*. 2008;68(9):3204–3213.

| TYPE OF CANCER | CD9 | CD63 | CD81 | CD82 | CD151 | REFERENCE |
|---------------|-----|------|------|------|------|-----------|
| Breast cancer | ++  | +    | +++  | +    | +++  | Penas et al, 2000 \(^4\) |
| Lung cancer   | +++ | ++   | ++   | +++  | +++  | Funakoshi et al, 2003 \(^5\) |
| Colon cancer  | +++ | +    | +++  | +    | +++  | Le Naour et al, 2006 \(^6\) |
| Burkitt lymphoma | +  | +++  | +    | +    | +    | Ferrer et al, 1998 \(^7\) |

Notes: Expression levels of tetraspanin proteins (CD9, CD63, CD81, CD82, and CD151) involved in cancer progression in different cancer cells. +, low; ++, medium; ++++, high.

| TYPE OF CANCER | CD9 | CD63 | CD81 | CD82 | CD151 | REFERENCE |
|---------------|-----|------|------|------|------|-----------|
| Breast cancer | ++  | +    | +++  | +    | +++  | Penas et al, 2000 \(^4\) |
| Lung cancer   | +++ | ++   | ++   | +++  | +++  | Funakoshi et al, 2003 \(^5\) |
| Colon cancer  | +++ | +    | +++  | +    | +++  | Le Naour et al, 2006 \(^6\) |
| Burkitt lymphoma | +  | +++  | +    | +    | +    | Ferrer et al, 1998 \(^7\) |
33. Copeland BT, Bowman MJ, Ashman LK. Genetic ablation of the tetraspanin
32. Fei Y, Wang J, Liu W, et al. CD151 promotes cancer cell metastasis via integrins
31. Haeuw JF, Goetsch L, Bailly C, Corvaia N. Tetraspanin CD151 as a target for
30. Kohno M, Hasegawa H, Miyake M, Yamamoto T, Fujita S. CD151 enhances
28. Sadej R, Romanska H, Kavanagh D, et al. Tetraspanin CD151 regulates trans
25. Voss MA, Gordon N, Maloney S, et al. Tetraspanin CD151 is a novel prognostic
24. Sterk LM, Geuijen CA, Oomen LC, Calafat J, Janssen H, Sonnenberg A. The
23. Devbhandari RP, Shi GM, Ke AW, et al. Profiling of the tetraspanin CD151
22. Yauch RL, Kazarov AR, Desai B, Lee RT, Hemler ME. Direct extracellular contact between integrin alpha (3) beta(1) and TM4SF protein CD151. J Biol Chem. 2000;275(13):9230–9238.
21. Derbhandari RP, Sterk LM, Wang W, et al. Profiling of the tetraspan protein CD151 web and conspiracy of CD151/integrin β1 complex in the progression of hepatocellular carcinoma. PLoS One. 2011;6(9):e24901.
20. Sterk LM, Geuijen CA, Oomen LC, Calafat J, Janssen H, Sonnenberg A. The tetraspan molecule CD151, a novel constituent of hemidesmosomes, associates with the integrin alpha beta4 and may regulate the spatial organization of hemidesmosomes. J Cell Biol. 2000;149:969–982.
19. Li Q, Yang XH, Xu F, et al. Tetraspanin CD151 plays a key role in skin squamous cell carcinoma. Oncogene. 2013;32(14):1772–1783.
18. Sterk LM, Geuijen CA, Oomen LC, Calafat J, Janssen H, Sonnenberg A. The tetraspan molecule CD151, a novel constituent of hemidesmosomes, associates with the integrin alpha beta4 and may regulate the spatial organization of hemidesmosomes. J Cell Biol. 2000;149:969–982.
17. Devbhandari RP, Shi GM, Ke AW, et al. Profiling of the tetraspanin CD151
16. Yauch RL, Kazarov AR, Desai B, Lee RT, Hemler ME. Direct extracellular contact between integrin alpha (3) beta(1) and TM4SF protein CD151. J Biol Chem. 2000;275(13):9230–9238.
15. Hasegawa M, Furuya M, Kasuya Y, et al. CD151 dynamics in carcinoma-stroma interaction: integrin expression, adhesion strength and proteolytic activity. Lab Invest. 2007;87(9):882–893.
14. Geary SM, Cowin AJ, Copeland B, Baleato RM, Miyazaki K, Ashman LK. The role of the tetrapsin CD151 in primary keratinocyte and fibroblast functions: implications for wound healing. Exp Cell Res. 2008;314(11–12):2165–2175.
13. Hasegawa M, Furuya M, Kasuya Y, et al. CD151 dynamics in carcinoma-stroma interaction: integrin expression, adhesion strength and proteolytic activity. Lab Invest. 2007;87(9):882–893.
12. Yauch RL, Kazarov AR, Desai B, Lee RT, Hemler ME. Direct extracellular contact between integrin alpha (3) beta(1) and TM4SF protein CD151. J Biol Chem. 2000;275(13):9230–9238.
11. Derbhandari RP, Sterk LM, Wang W, et al. Profiling of the tetraspan protein CD151 web and conspiracy of CD151/integrin β1 complex in the progression of hepatocellular carcinoma. PLoS One. 2011;6(9):e24901.