miRNA-223 upregulated by MYOD inhibits myoblast proliferation by repressing IGF2 and facilitates myoblast differentiation by inhibiting ZEB1

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Skeletal muscle differentiation can be regulated by various transcription factors and non-coding RNAs. In our previous work, miR-223 is differentially expressed in the skeletal muscle of chicken with different growth rates, but its role, expression and action mechanism in muscle development still remains unknown. Here, we found that MYOD transcription factor can upregulate miR-223 expression by binding to an E-box region of the gga-miR-223 gene promoter during avian myoblast differentiation. IGF2 and ZEB1 are two target genes of miR-223. The target inhibition of miR-223 on IGF2 and ZEB1 are dynamic from proliferation to differentiation of myoblast. miR-223 inhibits IGF2 expression only in the proliferating myoblast, whereas it inhibits ZEB1 mainly in the differentiating myoblast. The inhibition of IGF2 by miR-223 resulted in the repression of myoblast proliferation. During myoblast differentiation, miR-223 would be upregulated owing to the promoting effect of MYOD, and the upregulation of miR-223 would inhibit ZEB1 to promote myoblast differentiation. These results not only demonstrated that the well-known muscle determination factor MYOD can promote myoblast differentiation by upregulate miR-223 transcription, but also identified that miR-223 can influence myoblast proliferation and differentiation by a dynamic manner regulates the expression of its target genes.

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MicroRNAs (miRNAs) are small non-coding RNA of 20–25 nucleotides that mainly transcript from introns or intergenic regions, having critical roles in the regulation of gene expression at posttranscriptional levels.¹ It has been reported that miRNAs widely participate in chicken embryo growth, maturation, muscle cell proliferation and differentiation, cell migration, dosage compensation effect of Z chromosome and a series of life activities.²⁻⁷ In our preliminary study, we have used high-throughput RNA sequencing to study breast muscle transcriptome in Recessive White Rock (fast-growing chicken) and Xinghua chicken (slow-growing chicken), and pointed out that miR-223 can be used as a candidate gene associating with broiler growth.⁸

The miR-223 is highly conserved among vertebrates. As the first miRNA identified in human hemopiestic system,⁹ miR-223 was confirmed to participate in the regulation of hematopoietic cell proliferation and differentiation,¹⁰ and become a research hotspot. There are some competitive binding sites for NFI-A and C/EBP alpha in the upstream of miR-223. C/EBP alpha enhances miR-223 transcription activity, whereas NFI-A inhibits the transcription activity of miR-223.¹⁰ The upregulation of miR-223 expression will promote granulocyte and megakaryocyte generation, and the decline of miR-223 expression will cause hematopoietic stem cells to differentiate into erythrocyte.¹¹⁻¹² In addition, miR-223 was confirmed to regulate cell proliferation, differentiation, migration and signal transduction in mammals.¹³ However, the roles of miR-223 in muscle development still remain unclear.

Insulin-like growth factor-2 (IGF2) is a member of insulin-like growth factor family that can play a role in all insulin-activated target cells.¹⁴ It is well-known that IGF2 can be acted as the intermediate messenger of growth hormone, which can transfer the growth hormone to target organs, and regulate the development of organisms.¹⁵ IGF2 is conserved during evolution, the gene structure and exons location in chicken are similar to that in mammal, although the non-coding sequences were very different.¹⁶ In mammals, IGF2 is a kind of multifunctional cell regulation factor that has an important role in the muscle cell proliferation and differentiation.¹⁷ In chicken, IGF2 is significantly associated with broilers growth traits and carcass traits, such as birth weight, breast muscle weight, abdominal fat weight and glandular stomach weight.¹⁸⁻²⁰

Zinc finger E-box binding homeobox 1 (ZEB1) is a crucial nuclear transcription factor, it can directly bind to the E2-box [5-CATTC(G)] sequence in different gene promoter region, repressing the activity of gene transcription.²¹ In mammal, ZEB1 has three inhibiting zones.²² Zone-I mainly regulates T-lymphocyte differentiation. No inhibitory activity was found in zone-II. Zone-III is associated with muscle differentiation. In chicken, ZEB1 is also called DeltaEF1 (delta-crystallin/E2-box factor 1), which is highly homologous with mice and can also recognize the E2-box sequence (CATTG).²³ In the process

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miR-223 expression is related to skeletal muscle development. (a) Relative expression of miR-223 in Xinghua chicken breast muscle from embryo day 10 (E10) to E19. (b) Fluorescence in situ hybridization of miR-223 in Xinghua chicken breast muscle from E10 to E19. (c) H-E staining of breast muscle fiber cross section from E10 to E19 in Xinghua chicken

Figure 1  miR-223 expression is related to skeletal muscle development. (a) Relative expression of miR-223 in Xinghua chicken breast muscle from embryo day 10 (E10) to E19. (b) Fluorescence in situ hybridization of miR-223 in Xinghua chicken breast muscle from E10 to E19. (c) H-E staining of breast muscle fiber cross section from E10 to E19 in Xinghua chicken
of chicken embryo development, ZEB1 was expressed in somite, notochord, urogenital system, nervous system and other parts of the chicken embryo, suggesting that it may involve in cell development process. In addition, ZEB1 can inhibit the activity of viral gene transcription by targeting the E2-box sequence exists in the promoter region of chicken infectious anemia virus, result in the decrease of the virus transcription activity. MYOD and MYOG are critical transcription factors in skeletal muscle differentiation and can regulate the transcription of most myogenesis-related genes. During myoblast differentiation, both of these two factors are upregulated their expression. MYOD and MYOG regulates their target genes transcription by binding to E-box promoter elements containing the site CANNTG. Recently, many myogenic miRNAs were found to be regulated by MYOD and MYOG.
In our previous miRNA sequencing data, we found miR-223 expression is related to skeletal muscle development. Results found another regulatory pathway of MYOD to shape myoblast proliferation than in proliferating myoblast. These results identified a role for miR-223 in avian skeletal muscles of chickens with different growth rate. To further understand the relationship between miR-223 and skeletal muscle development, we detected its expression in breast muscle during chicken embryonic development. miR-223 was gradually upregulated its expression from embryonic day 10 (E10) to E13. However, its expression was continually downregulated after E13 (Figure 1a). In addition, in situ hybridization results also showed that the miR-223 expression was upregulated from E10 to E13 and downregulated from E14 to E19 (Figure 1b). Haematoxylin–eosin staining of the breast muscle showed that the muscle fibers are blury, irregular and may in the stage of cell differentiation and fusion between E10 and E13 (Figure 1c). After E13, the cross sections of muscle fibers were relatively regular and becoming more clearly defined (Figure 1c). Together with the expression data of miR-223, these results suggesting that miR-223 upregulated its expression during the active stage of muscle cell differentiation and fusion, whereas it downregulated its expression during muscle fiber maturation.

miR-223 regulates myoblast proliferation and differentiation. In our previous miRNA sequencing data, we found miR-223 overexpression inhibited chicken primary myoblast proliferation. (a) CCK-8 assay indicates miR-223 overexpression inhibited chicken primary myoblast (CPM) proliferation. (b) CCK-8 assay indicates miR-223 inhibition promoted CPM proliferation. (c) CPM was overexpressed with miR-223 and the negative control (NC) mimic, and the cell cycle phase was then analyzed. (d) Relative mRNA expression of the cell cycle-related genes after transfection of miR-223 and NC mimic. (e) CPM was transfected with miR-223 mimic and the NC inhibitor, and the cell cycle phase was then analyzed. (f) Relative mRNA expression of the cell cycle-related genes after transfection of miR-223 mimic and NC mimic. (g) miR-223 expression was upregulated from E10 to E13. However, its expression was continually downregulated after E13 (Figure 1a). In addition, by analyzing cell cycle of the transfected myoblasts, we found that miR-223 overexpression can decrease cell population in the S phase and increase cell population in the G1/0 phase (Figure 2c). miR-223 overexpression can also inhibit the expression of cell cycle-promoting genes and enhance cell cycle-inhibiting genes (Figure 2d). On the contrary, inhibition of miR-223 significantly increased cell population in the S phase and decreased cell population in the G1/0 phase (Figure 2e). miR-223 inhibition also enhanced the expression of cell cycle-promoting genes and repressed cell cycle-inhibiting genes (Figure 2f). EdU staining showed that the proliferation rate of miR-223-transfected cells was significantly reduced compared with that of the control cells (Figure 2g and h), whereas miR-223 loss-of-function promoted cell proliferation rate (Figure 2g and h). Furthermore, these roles of miR-223 were also existed in the qm-7 cells (Figure 2i-p), indicating that miR-223 can inhibit avian myoblast proliferation.

The inhibitory effect of miR-223 on myoblast proliferation was achieved by its target gene IGF2. IGF2 is an important growth factor that can functioning as growth promoting hormone during cell development. Here, we found that the 3'-untranslated regions (3'-UTR) of IGF2 mRNA has a potential binding site of miR-223 (Figure 3a). To validate whether IGF2 is a target gene of miR-223, we constructed two dual-luciferase reporters with the wide-type and mutant 3'-UTR of IGF2, respectively. Results show that miR-223 significantly repressed the luciferase activity of the wide-type reporter, whereas it has no effect on the luciferase activity of the mutant reporter (Figure 3b). In addition, miR-223 significantly inhibited the mRNA and protein expression of IGF2 gene in CPM (Figure 3c), and the inhibition of miR-223 released IGF2 mRNA and protein expression (Figure 3d). The above results indicated that IGF2 is a direct target gene of miR-223.

Although IGF2 is a well-known growth promoting-gene, its roles in chicken myoblast proliferation have not been examined. In this study, we transfected pSDS-IGF2 and three IGF2 siRNAs in poultry myoblast to overexpressing or inhibiting IGF2 expression, respectively. pSDS-IGF2 transfection significantly increased IGF2 mRNA and protein expression, whereas the si-170, which is specifically designed for
chicken IGF2 mRNA, significantly repressed IGF2 mRNA and protein expression (Figure 3e-h). IGF2 overexpression promotes myoblast proliferation through CCK-8 assay (Figure 3i and j) and cell cycle analysis (Figure 3k and l), whereas IGF2 inhibition repressed myoblast proliferation (Figure 3m-p). Furthermore, the rescue assay shown that miR-223 overexpression can inhibit the myoblast proliferation, and the transfection of IGF2 overexpression vector could counteract
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Figure 3 The inhibitory effect of miR-223 on myoblast proliferation was achieved by its target gene IGF2. (a) The potential binding site of miR-223 in the IGF2 mRNA 3' UTR. (b) Dual-luciferase reporter assay indicated that miR-223 can bind to the predicted binding site of the IGF2 mRNA 3' UTR. (c) miR-223 overexpression significantly inhibited IGF2 mRNA and protein expression in CPM. (d) miR-223 inhibition significantly promoted IGF2 mRNA and protein expression in CPM. (e) IGF2 overexpression significantly promoted IGF2 mRNA and protein expression in CPM. (f) IGF2 inhibition significantly inhibited IGF2 mRNA and protein expression in CPM. (g) IGF2 overexpression significantly promoted IGF2 mRNA and protein expression level in qm-7. (h) IGF2 inhibition significantly inhibited IGF2 mRNA and protein expression level in qm-7. (i) CCK-8 assay indicates that IGF2 overexpression significantly promoted myoblast proliferation in CPM. (j) CCK-8 assay indicates that IGF2 overexpression significantly promoted myoblast proliferation in CPM. (k) CPM was overexpressed with IGF2 and the control vector, and the cell cycle phase was then analyzed. (l) qm-7 was overexpressed with IGF2 and the control vector, and the cell cycle phase was then analyzed. (m) CPM was co-transfected with miR-223 and pSDS-IGF2, miRNA mimic NC and control vector, miR-223 and control vector, respectively, and the cell cycle phase was then analyzed. Results are shown as the mean ± S.E.M. of three independent experiments. Independent sample t-test was used to analysis the statistical differences between groups. *P < 0.05; **P < 0.01

the inhibitory effect of miR-223 on myoblast proliferation (Figure 3q). Together, these results argue that the inhibitory effect of miR-223 on myoblast proliferation was achieved by its target gene IGF2.

miR-223 promotes myoblast differentiation. miR-223 was upregulated its expression during the active stage of chicken skeletal muscle differentiation and fusion. Next, we examined the expression and function of miR-223 during myoblast differentiation and fusion. miR-223 expression was significantly upregulated from the proliferation to the differentiation of both CPM and qm-7 cells (Figure 4a and b), suggesting its involvement in these processes. Therefore, we transfected miR-223 mimic and inhibitor into the myoblasts. After transfection, the cells were induced to differentiation and fusion. Overexpression of miR-223 significantly increased the expression of muscle differentiation marker genes (Figure 4c and d), whereas the inhibition of miR-223 decreased the expression of these marker genes (Figure 4e and f). In addition, miR-223 promotes the formation of myotubes (Figure 4g), and the size of the myotube area was significantly increased after transfection of miR-223 (Figure 4h). Therefore, these results demonstrated that miR-223 can promote myoblast differentiation.

ZEB1 is another miR-223 target gene, functioning as an inhibitor of myoblast differentiation. By using the TargetScan online software to predict the target genes of miR-223, we found that the 3'-UTR of ZEB1 mRNA has a potential binding site of miR-223 (Figure 5a). Notably, this binding site is conserved among vertebrates (Figure 5b). The dual-luciferase report assay suggested that miR-223 directly binds to the predicted target site of ZEB1-3'-UTR (Figure 5c). miR-223 overexpression inhibited the mRNA and protein expression level of ZEB1 (Figure 5d and e), and the inhibition of miR-223 promoted the mRNA and protein expression level of ZEB1 (Figure 5f and g). Therefore, the above results indicated that ZEB1 is another target gene of miR-223.

During myoblast proliferation and differentiation, ZEB1 protein was gradually downregulated its expression (Figure 5h). To test the function of ZEB1 in myoblast differentiation, we synthesized siRNAs specifically target for chicken ZEB1 mRNA. Transfection of these siRNAs significantly inhibited the mRNA and protein expression level of ZEB1 in CPM and qm-7 (Figure 5i and j). In addition, ZEB1 inhibition significantly increased the expression of muscle differentiation marker genes in both of these two cells (Figure 5k and l), suggesting ZEB1 is a repressor of myoblast differentiation. Rescue assay shown that ZEB1 overexpression is able to counteract the promotion effect of miR-223 on myoblast differentiation (Figure 5m). Therefore, ZEB1 is another miR-223 target gene that can function as an inhibitor of myoblast differentiation.

The inhibition of miR-223 on IGF2 and ZEB1 is different between myoblast proliferation and differentiation. The above results shown that both IGF2 and ZEB1 are target genes of miR-223. However, IGF2 is able to promote myoblast differentiation in mammal myoblast.32 Its expression was increased during CPM and qm-7 differentiation (Figure 6a and b). This expression trend is similar to miR-223, which can directly inhibit IGF2 expression. In addition, specifically inhibition of IGF2 by siRNA significantly downregulated muscle differentiation marker genes (Figure 6c and d), indicating its promoting effect in myoblast differentiation. These results suggested that miR-223 cannot restrict IGF2 expression and function during myoblast differentiation.

On the other hand, ZEB1, another miR-223 target gene that can inhibit myoblast differentiation, is unable to regulate cell cycle in proliferating myoblast (Figure 6e and f). Therefore, we were interest in how the miR-223 regulates the ZEB1 and IGF2 expression in the proliferating and differentiating myoblast, respectively. For ZEB1, miR-223 inhibits its expression in both proliferation and differentiation stages of myoblast (Figure 6g), and the inhibitory effect of miR-223 on ZEB1 has no significant difference between these two stages. For IGF2, miR-223 inhibits its expression only in the proliferation stage of myoblast (Figure 6g), and the miR-223 expression shown no significant difference between the proliferation and differentiation stages (Figure 6h). To further validate these results, we transfected dual-luciferase reporter inserted with the 3'-UTR of ZEB1 or IGF2 into the proliferating and differentiating myoblasts, respectively. The reporter activity inserted with ZEB1-3'-UTR was significant lower in differentiating myoblasts compared with that in proliferating myoblasts (Figure 6i). However, the reporter activity inserted with IGF2-3'-UTR was significant higher in differentiating myoblasts compared with that in proliferating myoblasts (Figure 6i). Together, these results suggested that the inhibition of miR-223 on IGF2 and ZEB1 is different between myoblast proliferation and differentiation.

MYOD regulates miR-223 transcription by binding to the E-box 1 region. To further understand the structure of the
gga-miR-223 gene, we isolated its full-length pri-miR-223 by using 5' and 3' rapid amplification of cDNA ends (RACE). The obtained gga-miR-223 gene was 2228 bp in length (Figure 7a). Next, we analyzed the upstream region of the gga-miR-223 gene to find the core promotor region. Four fragments, including 668-bp, 1020-bp, 1508-bp and 1932-bp upstream regions of the gga-miR-223 transcription start site (TSS) were amplified and cloned into the pGL3-basic vector. Forty-eight hours after transfected these reporter vectors into the myoblasts, the relative luciferase activity was measured to analyze the promoter activity of these four reporters. Results shown that the four reporters have similar relative luciferase activity (Figure 7b), suggesting that the shortest one or no reporter have the core promoter region. In addition, we found that there are two E-boxes located in the 1932-bp upstream regions of the gga-miR-223 TSS (Figure 7a). It is well-known that the MYOG and MYOD transcription factors can bind to the E-box region and promote the transcription activity of myogenic genes. Considering that MYOD, MYOG and miR-223 are all

Figure 4 miR-223 promotes myoblast differentiation. (a) Relative miR-223 expression during CPM differentiation. (b) Relative miR-223 expression during qm-7 differentiation. (c) Relative expression of muscle differentiation marker genes after CPM transfected with miR-223 and NC. (d) Relative expression of muscle differentiation marker genes after qm-7 transfected with miR-223 and NC. (e) Relative expression of muscle differentiation marker genes after CPM transfected with miR-223 inhibitor and NC inhibitor. (f) Relative expression of muscle differentiation marker genes after qm-7 transfected with miR-223 inhibitor and NC inhibitor. (g) MyHC staining of CPM at 72 h after transfection of miR-223 or NC mimic. (h) Myotube area (%) at 72 h after transfection of miR-223 or NC mimic. Results are shown as the mean ± S.E.M. of three independent experiments. Independent sample t-test was used to analysis the statistical differences between groups. *P < 0.05; **P < 0.01
upregulated their expression during myoblast differentiation, we next examined whether MYOD and MYOG can bind to these two E-box regions and promote gga-miR-223 transcription. By mutated these two regions, respectively, we found that the relative luciferase activity was significantly reduced when we mutated E-box 1, whereas there is no change of the luciferase activity when we mutated E-box 2 (Figure 7b). This result suggesting that the E-box 1 is
important for the transcription activity of the promoter region. Next, chromatin immunoprecipitation assay (ChIP) results indicated that the MYOD can bind to the E-box 1 region (Figure 7c). To avoid the effect of endogenous MYOD, we used DF-1 cell to test the regulation of MYOD on the gga-miR-223 gene promoter and miR-223 transcription. MYOD overexpression promoted the relative luciferase activity of the pGL3-1932 reporter, but this overexpression had no effect on the luciferase activity of the mutated pGL3-1932 reporter (Figure 7d). In addition, MYOD overexpression promoted miR-223 expression (Figure 7e). For primary myoblast, MYOD loss-of-function not only inhibited the expression of miR-223 and muscle differentiation marker genes, but also promoted the expression of miR-223 target genes.
Figure 7  MYOD regulates miR-223 transcription by binding to the E-box 1 region. (a) The location of the two E-boxes in the 1932-kb upstream region of the gga-miR-223 gene TSS. (b) Identification of the core region in the gga-miR-223 gene promoter by luciferase reporter assays. (c) ChIP-qPCR analysis using anti-MYOG, anti-MYOD or chicken IgG antibodies, and the values showed that MYOD could bind to the R1 region of the chicken gga-miR-223 gene in myoblasts. A region from the GAPDH gene was amplified as a NC to verify the specificity of the enrichment. (d) MYOD overexpression promoted the relative luciferase activity of the pGL3-1932 reporter in DF-1 cell. (e) MYOD overexpression upregulated miR-223 expression in DF-1 cell. (f) Relative expressions of miR-223 and its downstream genes after transfection of si-MYOD in chicken primary myoblast. (g) CCK-8 assay indicated that MYOG loss-of-function significantly reduced myoblast proliferation in chicken primary myoblast. Results are shown as the mean ± S.E.M. of three independent experiments. Independent sample t-test was used to analysis the statistical differences between groups. *P<0.05; **P<0.01
genes IGF2 and ZEB1 (Figure 7f). Furthermore, myoblast proliferation was also being inhibited when the cell was transfected with si-MYOD (Figure 7g). Together, these results demonstrated that the MYOD regulates gga-miR-223 transcription by binding to the E-box 1 region.

**Discussion**

In this study, we identified miR-223 as another miRNA that has a role in avian skeletal muscle development. miR-223 regulates myoblast proliferation and differentiation by balancing the expression of its target genes IGF2 and ZEB1. In addition, MYOD promotes miR-223 transcription during myoblast differentiation by binding to the gga-miR-223 gene promoter region (Figure 8). These findings not only revealed a new miRNA-mediated pathway functions on skeletal muscle differentiation, but also found an interest regulatory mode of miRNA in its target genes.

MYOD belongs to the myogenic basic-helix-loop-helix transcription factors, and is an important regulator during skeletal muscle differentiation. MYOD has the ability to initiate a lot of muscle-related genes transcription by recruitment of transcription factors and histone acetyltransferases. Many muscle development related genes have been found to be regulated by MYOD during myoblast differentiation. In this study, we found that MYOD can bind to the E-box, which is located in the promoter region of miR-223 gene, and promote miR-223 expression in chicken myoblast. In addition, the expression of miR-223 and MYOD are upregulated during chicken myoblast differentiation, demonstrating a positive association between these two regulators. Besides, some of the muscle-specific miRNAs, such as miR-1, miR-133 and miR-206, can also be regulated by MYOD during myoblast differentiation. Therefore, these findings suggested that MYOD is able to regulate myoblast differentiation through activate various myogenic miRNAs expression.

ZEB1 is a transcriptional repressor. During muscle differentiation, ZEB1 is able to repressed muscle gene transcription, and the inhibition of ZEB1 can promote myoblast differentiation and fusion. Although the repression effect of ZEB1 in muscle differentiation has been well documented, however, its expression pattern during muscle differentiation still remains unknown. In this study, we have shown that ZEB1 protein was gradually downregulated during chicken myoblast differentiation, and the decrease of ZEB1 is, at least in part, owing to the inhibition of miR-223. In addition, ZEB1 can also repress myoblast differentiation in avian, but it cannot regulate myoblast proliferation, even though it has been shown to have positive role in many cancer cell proliferation. MYOD can occupy G/C-centered E-boxes in the promoters of many muscle-related genes and initiate the transcription of these genes during myoblast differentiation. Interestingly, ZEB1 can also bind to the G/C-centered E-boxes in myoblast, but its binding results in the repression of muscle-related gene expression. Therefore, our results suggesting that MYOD inhibits ZEB1 expression by promoting miR-223 expression during myoblast differentiation.

IGF2 has long been established to have important roles in muscle development, and it is also a critical growth factor for chicken. However, the function of IGF2 in avian myoblast has not been verified before. Here, we reported that IGF2 can both promote avian myoblast proliferation and differentiation, and we also found that miR-223 has the ability to inhibit IGF2 expression. Notably, miR-223 inhibits IGF2 expression only in the proliferating myoblast. Its inhibition effect on IGF2 expression was significantly reduced from proliferating myoblast to differentiating myoblast. In addition, the inhibitory effect of miR-223 on ZEB1 in the proliferating myoblast was more significant than that in the differentiating myoblast. It is generally understood that a miRNA can bind to multiple target genes, and the inhibiting effect of miRNA on these target genes would be different because of their target sites with different functional properties. However, it is rare to see that a miRNA has different binding efficiency on one target gene during cell development. Previous study has shown that synaptic stimulation can relieve the repression of miRNA to its target genes. A miRNA-mediated inhibition of target gene translation can also be relieved when the cells subjected to different stress conditions, and this relief is depend on the binding of HuR, which is an AU-rich element RNA-binding protein (RBP), to the 3′ UTR of the target gene. Dead end 1 (Dnd1) is another RBP evolutionary conserved among vertebrates, and also has the ability to counteracts the inhibition roles of several miRNAs by binding to the 3′ UTR of target genes. However, the precise mechanism of how miR-223 dynamic regulates IGF2 and ZEB1 during myoblast proliferation and differentiation still need to be explored. Collectively, these and our findings suggest that the binding of miRNA to its target genes is dynamic in different cellular environment, and this binding can be rapidly responded to the specific cellular needs.

In summary, this work has shown that the avian myoblast proliferation and differentiation are regulated by MYOD-miR-223-IGF2/ZEB1 pathway. In the proliferating avian myoblast, miR-223, which is upregulated by MYOD, inhibits IGF2 expression to represses myoblast proliferation. In the differentiating avian myoblast, the upregulation of MYOD promotes miR-223 expression, and miR-223 inhibits ZEB1 rather than IGF2 to facilitate myoblast differentiation and fusion.
RNA extraction, cDNA synthesis and quantitative real-time PCR. The total RNA was extracted from tissue samples using the RNeasy Mini Kit (Qiagen, Hilden, Germany). cDNA synthesis was performed using the High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA, USA) according to the manufacturer’s instructions. Quantitative real-time PCR was performed using the ABI 7500 real-time PCR system (Life Technologies, Carlsbad, CA, USA). The expression levels of miR-223 were normalized to U6 snRNA levels.

Transfections. Transfections were performed with Lipofectamine 3000 reagent (Invitrogen) according to the manufacturer’s instructions. Nucleic acids were diluted in OPTI-MEM Medium (Gibco). All experiments were carried out at least three times independently.

Cell cycle analysis. After 36 h transfection, primary myoblast or QM-7 cells cultured in 12-well plates were fixed in 70% ethanol overnight at −20 °C. The Cell Cycle Analysis Kit (Thermo Fisher Scientific, Waltham, MA, USA) was used to perform DNA content analysis. The data were acquired by a BD Accuri C6 flow cytometer (BD Biosciences, San Jose, CA, USA). All the data were acquired by averaging the results from four independent repeats.

CCK-8 assay. Primary myoblast or QM-7 cells were cultured in 96-well plates. A total of 10 μl of CCK-8 solution was added to each well and incubated for 1 h. The absorbance was measured at 450 nm by a Model 680 Microplate Reader (Bio-Rad). All the data were acquired by averaging the results from five-independent experiments.

miRNA in situ hybridization assay. A 15-μm thick cryosection from breast muscle tissues of Xinhua chicken was fixed in 4% PFA for 15 min at room temperature, and then treated for 10 min with Proteinase K. miRNA in situ hybridization was performed using the FISH kit (Exonbio Lab, Guangzhou, China). DNA was counterstained with DAPI (1 μg/ml). Images of miRNA signals in slides were captured by a Leica DMi8 fluorescent microscope.
EdU assay. EdU assay was performed as previously described with the following modifications. Twenty-four hours after transfection, primary myoblasts were exposed to 10 μM 5-ethynyl-2'-deoxyuridine (EdU; RiboBio) for 24 h at 37 °C, and the QM-7 cells were exposed to 50 μM EdU for 2 h at 37 °C. In addition, for primary myoblasts, 1× Apollo reaction cocktail (RiboBoo) was added to the cells and incubated for 30 min. Whereas, in QM-7 cells, 1× Apollo reaction cocktail was added to the cells and incubated for 20 min. The EdU-stained cells were visualized under a Leica DMi8 fluorescent microscope. The proliferation rate was calculated by the number of EdU-stained cells normalized to the number of Hoechst 33342-stained cells.

Statistical analysis. Each experiment was repeated three times, and all results are represented as the mean ± S.E.M. Independent sample t-test was used to perform the statistical significant difference between groups.

Ethics standards. All experimental protocols were approved by the South China Agricultural University Institutional Animal Care and Use Committee (approval ID: SCAU1014). And the methods were carried out in accordance with the regulations and guidelines established by this committee.

Conflict of Interest
The authors declare no conflict of interest.

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1. Bartel DP, MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 2004; 116: 281–297.
2. Rengaraj D, Lee BR, Lee SI, Seo HW, Han JY. Expression patterns and miRNA regulation of DNA methyltransferases in chicken primordial germ cells. PLoS ONE 2011; 6: e18254.
3. Gischwend AR, Weingartner LA, Moore RC, Ming R. The sex-specific region of sex chromosomes in animals and plants. BMC Genomics 2012; 13: 362.
4. Huang HY, Liu RR, Zhao GP, Li QH, Zheng MQ, Zhang JJ et al. Integrated analysis of miRNA and mRNA expression profiles in adipose tissues in chickens. Sci Rep 2015; 5: 16132.
5. Han B, Lian L, Li X, Zhao C, Qu L, Liu C et al. Chicken gga-miR-103-3p targets CCNE1 and TFDP2 and inhibits MDCC-MSB1 cell migration. G3 (Bethesda) 2016; 6: 1277–1285.
6. Li Y, Wang X, Ju J, Shao F, Zhang Y, Lu X et al. MiR-122 targets the vanin 1 gene to regulate xenopus embryogenesis. Cell Death and Disease 2012; 13: 1491–1493.
7. Seok R, Mura K, Funahashi J, Kamachi Y, Muta K, Abe Y, Muta K et al. Identification of three single nucleotide polymorphisms in the chicken insulin-like growth factor 1 and 2 genes and their associations with growth and feeding traits. Poult Sci 2003; 82: 1485–1493.
8. Tang S, Sun D, Ou J, Zhang Y, Xu G, Zhang Y. Evaluation of the Igfs (IGF1 and IGFD) genes as candidates for growth, body measurement, carcass, and reproduction traits in Beijing You and Silkie chickens. Anim Biotechnol 2010; 21: 104–113.
9. Gholami M, Erbe M, Garce R, Preisinger R, Weigend A, Weigend S et al. Population genomic analyses based on 1 million SNPs in commercial egg layers. PLoS ONE 2014; 9: e89882.
10. van Grunsven LA, Telaesen V, Michiels C, Opdecamp K, Huybrechts B, Bellesford EJ. deltaEf1 and SIPI are differentially expressed and have overlapping activities during Xenopus embryogenesis. Dev Dyn 2006; 235: 1491–1500.
11. Postigo AA, Dean DC. Differential expression and function of members of the zfh-1 family of zinc finger homeodomain repressors. Proc Natl Acad Sci USA 2000; 97: 6391–6396.
12. Li Y, Wang X, Yu J, Shao F, Zhang Y, Lu X et al. Identification of three single nucleotide polymorphisms in the chicken insulin-like growth factor 1 and 2 genes and their associations with growth and feeding traits. Poult Sci 2003; 82: 1485–1493.
13. Seok R, Mura K, Funahashi J, Kamachi Y, Muta K, Abe Y, Muta K et al. Identification of three single nucleotide polymorphisms in the chicken insulin-like growth factor 1 and 2 genes and their associations with growth and feeding traits. Poult Sci 2003; 82: 1485–1493.
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45. Brennecke J, Stark A, Russell RB, Cohen SM. Principles of microRNA-target recognition. PLoS Biol 2005; 3: e85.
46. Ashraf SI, McLoon AL, Sciarici SM, Kunes S. Synaptic protein synthesis associated with memory is regulated by the RISC pathway in Drosophila. Cell 2006; 124: 191–205.
47. Schratt GM, Tsubing F, Ngh EA, Karre CG, Sabatini ME, Kiebler M et al. A brain-specific microRNA regulates dendritic spine development. Nature 2006; 439: 283–289.
48. Bhattacharyya SN, Habermacher R, Martine U, Closs EI, Filipowicz W. Relief of microRNA-mediated translational repression in human cells subjected to stress. Cell 2006; 125: 1111–1124.
49. Kedde M, Strasser M, Boldajipour B, Oude VJ, Slanchev K, le Sage C et al. RNA-binding protein Drd1 inhibits microRNA access to target mRNA. Cell 2007; 131: 1273–1286.
50. Luo W, Li G, Yi Z, Nie Q, Zhang X. E2F1-miR-20a-5p/20b-5p auto-regulatory feedback loop involved in myoblast proliferation and differentiation. Sci Rep 2016; 6: 27904.

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