Urinary proteomic profiling in severe obesity and obstructive sleep apnoea with CPAP treatment

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ABSTRACT

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Introduction: Obstructive sleep apnoea (OSA) is common in obesity and is associated with cardiovascular and metabolic complications. Continuous positive airway pressure (CPAP) in OSA may lead to physiological changes reflected in the urinary proteome. The aim of this study was to characterise the urinary proteome in severely obese adult subjects with OSA who were receiving CPAP compared with severely obese subjects without OSA.

Methods: Severely obese subjects with and without OSA were recruited. Subjects with OSA were receiving CPAP. Body composition and blood pressure measurements were recorded. Urinary samples were analysed by Capillary Electrophoresis–Mass Spectrometry (CE-MS).

Results: Twenty-seven subjects with OSA-on-CPAP (age 49±7 years, BMI 43±7 kg/m²) and 25 controls without OSA (age 52±9 years, BMI 39±4 kg/m²) were studied. Mean CPAP use for OSA patients was 14.5±1.0 months. Metabolic syndrome was present in 14(52%) of those with OSA compared with 6(24%) of controls (p=0.039). A urinary proteome comprising 15 peptides was identified showing differential expression between the groups (p<0.01). Although correction for multiple testing did not reach significance, sequences were determined for 8 peptides demonstrating origins from collagens, fibrinogen beta chain and T-cadherin that may be associated with underlying cardiovascular disease mechanisms in OSA.

Conclusions: The urinary proteome is compared in OSA with CPAP and without OSA in severe obesity. The effects of CPAP on OSA may lead to changes in the urinary peptides but further research work is needed to investigate the potential role for urinary proteomics in characterising urinary peptide profiles in OSA.

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1. Introduction

In obstructive sleep apnoea (OSA), episodes of recurrent upper airway obstruction cause intermittent hypoxia, with frequent sleep arousals and fragmented sleep, resulting in excessive daytime hypersomnolence [1]. OSA has been increasing in prevalence, and it has been estimated that 13% of men and 6% of women between the ages of 30 and 70 years have moderate to severe forms of OSA [2]. This has been linked with the growing obesity prevalence as it is known that obesity is a risk factor for OSA. The prevalence of OSA in severe obesity has been estimated to be between 40% and 90% [3,4]. OSA is associated with hypertension, cardiovascular disease, type 2 diabetes and metabolic impairment [5].

It is known that OSA is independently associated with arterial hypertension and this may be related to repeated hypoxia and reoxygenation episodes that may cause ischaemia–reperfusion injury, as well as increased sympathetic activity, inflammation and oxidative stress [6]. The kidney has a role in blood pressure regulation and may be sensitive to the effects of intermittent hypoxia [7]. Nocturnal hypoxia may be associated with loss of kidney function [8], and a positive relationship has been observed between impaired glomerular filtration and desaturation frequency, suggesting an association between OSA and chronic kidney impairment [9]. Histological changes in renal glomeruli have been reported in severely obese patients with OSA [7], that may be related to pressor responses and endothelial dysfunction in OSA [10].

Capillary electrophoresis coupled to mass spectrometry (CE–MS) is a sensitive proteomic analytical technique that is robust with high reproducibility of results in an acceptable time frame [11]. In CE–MS, capillary zone electrophoresis is interfaced with high resolution mass spectrometry; electrophoretic separation of peptides is performed by CE according to charge and size, and the peptides are separated by application of high voltage and analysed in the mass spectrometer [11]. The accuracy, precision, selectivity, sensitivity, reproducibility, and stability of the CE–MS measurements have been previously demonstrated [12]. CE–MS has been widely used to study biomarker panels of urinary peptides for conditions such as coronary artery disease [13] and chronic kidney disease [14] that may indicate the presence of the relevant condition [15].

We have previously used CE–MS to characterise the urinary proteome in severe obesity with and without OSA, and to determine if there were significant differences in urinary peptide profiles that may potentially relate to OSA. 24 peptides were found to be differentially distributed between the two groups although these differences did not reach significance after correction for multiple testing. Sequences were determined for 8 peptides demonstrating origins from collagens and fibrinogen alpha [16]. The proteome in OSA prior to treatment may have been influenced by the effects of OSA potentially influencing the peptide panel and it was important to ascertain if there were changes in the urinary peptide panel with continuous positive airway pressure (CPAP) treatment.

CPAP therapy can improve glomerular hyperfiltration in OSA patients [17], perhaps by reducing renal renin-angiotension system (RAS) activity, suggesting a potential effect of CPAP on kidney function [18]. The sensitivity of the kidney to repetitive intermittent hypoxia may influence the urinary proteome in OSA, and it is conceivable that the effects of CPAP treatment in OSA may lead to physiological changes reflected by the urinary proteome. Defining this in patients on CPAP may be useful as it would potentially reflect effects of CPAP on various mechanisms, such as RAS activity, oxidative stress, and endothelial dysfunction [19].

The use of urinary CE–MS analyses to study the urinary proteome in adult OSA and severe obesity with CPAP therapy has not been previously investigated. In this paper, we sought to investigate the hypothesis that the urinary peptide patterns were different between the two groups. To achieve this goal, we characterised the urinary proteome in severely obese subjects with OSA who were receiving CPAP compared to subjects without OSA.

2. Methods

2.1. Research design

This was a case-control study comparing patients who were on effective CPAP treatment with subjects without OSA. The research involved patient recruitment and sampling, and subsequent CE–MS urinary proteome analyses.

2.2. Ethics statement

The study was approved by the research ethics committee (NRES 13/NW/0589) and was performed in accordance with the Declaration of Helsinki 2008. All patients gave written informed consent.

2.3. Participants

Severely obese patients from the weight management and sleep clinics at University Hospital Aintree were recruited from March 2012 to January 2013. Patients were selected according to the inclusion and exclusion criteria as assessed according to the clinical history, physical examination and analysis of the medical notes. At baseline, all subjects had spirometry testing and overnight polysomnography to confirm or refute the diagnosis of OSA. All subjects with OSA at baseline were previously untreated for their OSA and they were offered CPAP. All OSA subjects on CPAP and non-OSA subjects were followed-up from September 2013 to February 2014.

Patients were eligible if they were ≥ 21 years old and had body mass index (BMI) ≥ 35 kg/m². Patients who were overweight or had mild obesity were not included because the majority of the patients were recruited from the hospital weight management service which manages patients with severe obesity. Exclusion criteria were patients with cardiorespiratory disease; current smokers or those with more than 10 pack years smoking history; kidney and liver disease; acute illnesses and pregnancy. Based on the exclusion criteria, we sought to control for potential confounders that may affect the interpretation of the urinary proteome including coronary artery disease, coexisting respiratory illness, renal...
and liver disease, as well as other comorbidities. OSA patients who were not CPAP treatment or not compliant with their CPAP treatment were excluded from the analysis. Likewise, patients who had bariatric surgery were excluded from subsequent analysis as our aim was to study urinary proteome in subjects who were receiving CPAP treatment (CPAP ≥ 4 h per night) and non-OSA subjects.

2.4. Sample size

Although no previous studies evaluating the effect of CPAP on the adult urinary proteome using CE–MS in OSA and obesity have been performed, previous urinary proteomic work in paediatric OSA have investigated sample sizes of n=11–30 per group [20,21]. Therefore, we aimed for a minimum of 25 subjects per group.

2.5. Protocol

All patients attended a study visit day between 0800 and 1000 h and underwent a detailed history and physical examination. Body composition measurements, anthropometry, fasting blood and urine sampling and venous blood gases (to assess for hypercarbia) were performed. Daytime somnolence was assessed using an Epworth Sleepiness Scale Questionnaire (ESS) where a score >10 indicated increased sleepiness. To ensure comparability and uniformity of assessments of patients, body composition, anthropometry and sampling were carried out at the same time at each study visit.

2.5.1. Blood pressure

Blood pressure was measured at the arm in a sitting position after a rest for at least 5 min at 1 min intervals between each measurement, using an oscillometric digital blood pressure monitor (HEM-705CP, Omron, Japan). The mean of three measurements was calculated.

2.5.2. Body measurements

All measurements were performed in triplicate. Weight and height were measured without shoes and with light clothing. Other measurements included neck circumference at the level of the laryngeal prominence; waist circumference midway between the lower rib and iliac crest; and hip circumference was measured horizontally over the widest part of the gluteal region. The tape measure was ensured to be snug and not compressing the skin, parallel to the floor with measurement at the end of a normal expiration. Body composition measurements used bioimpedance scales (TBF-521, TANITA, Tokyo, Japan). This method has been previously validated [22].

2.5.3. Biochemical measurements

Serum samples were collected using standard phlebotomy vials and immediately sent to the pathology laboratory for analysis in accordance with local protocol and standards. Serum biochemistry was measured using standard laboratory assays (Roche, UK). Blood gases were analysed with a Cobas Blood Gas Analyser (Roche, UK).

2.5.4. Metabolic syndrome

Subjects were assessed for metabolic syndrome according to the National Cholesterol Education Program (NCEP) guidelines [23]. Patients had metabolic syndrome if three or more risk factors were present: waist circumference (males >102 cm; females >88 cm), triglycerides ≥ 1.7 mmol/l, high density lipoprotein (HDL) cholesterol (males <1.04 mmol/l; females <1.3 mmol/l), blood pressure ≥ 130/≥ 85 mmHg, and fasting glucose ≥ 6.1 mmol/l.

2.5.5. Baseline sleep studies and spirometry

All subjects previously had overnight multichannel respiratory limited polysomnography (PSG) (Somnoscreen Digital PSG acquisition system, Version 2.0, Somnomedics, Randersacker, Germany) and spirometry (Spiro Air LT system, Medisoft, Sorinnes, Belgium) to confirm or refute OSA. Apnoea was defined as a cessation of airflow for >10 s. Hypopnoea was defined as a 50% reduction in airflow accompanied by a >4% desaturation and a reduction in chest wall movement. OSA was diagnosed if the apnoea-hypopnoea index (AHI) was ≥ 5.

2.5.6. CPAP therapy

Patients with OSA received standard CPAP therapy with S8/S9 Escape machines (ResMed, Abingdon, UK). CPAP compliance was based upon usage in hours per night (h/night) at the prescribed pressure. Compliance data was recorded by the CPAP machines and was downloaded using ResScan software (Version 4.2, ResMed, Abingdon, UK). This was assessed at each patient’s most recent routine CPAP compliance/adherence clinic. Adequate compliance was defined as a usage time of >4 h/night on >70% of nights in the treatment group.

2.6. CE–MS materials and methods

2.6.1. Urine sample preparation

Spontaneously voided urine samples for CE–MS analyses were collected at the study visit using urine Monovettes (Sarstedt, Nümbrecht, Germany). Second void samples were collected at the same time each morning (~09:00 h) after an overnight fast for 8 h, at their study visit prior to the administration of any medication. The samples were stored at −80 °C in urine Monovettes until the sample preparation stage. Sample preparation for proteomic analysis was performed as previously described [24]. In this process, each 0.7 mL aliquot of urine was thawed immediately before use and diluted with 0.7 mL of 2 M urea, 10 mM NH₄OH containing 0.02% SDS. Each sample was ultrafiltered (2000 g, 60 min, 4 °C) to remove higher molecular mass proteins, such as albumin and immunoglobulin G using Centrisart ultracentrifugation filter devices (20 kDa MW) (Sartorius stedim Biotech, Goettingen, Germany) until 1.1 mL of filtrate was obtained. The filtrate from each sample was then applied onto separate PD-10 desalting columns (GE Healthcare, Uppsala, Sweden) equilibrated in 0.01% NH₄OH in HPLC-grade H₂O to decrease matrix effects by removing urea, electrolytes, and salts, and to enrich polypeptides present. Finally, all samples were lyophilised, and stored at 4 °C.
2.6.2. CE-MS analysis
Samples were re-suspended in HPLC-grade H2O shortly before CE–MS analyses, as described [24]. CE–MS analysis was performed using a P/ACE MDQ capillary electrophoresis system (Beckman Coulter, Fullerton, USA) using a 90 cm 360 μm OD, 50 μm ID non-coated silica capillary with a tapered tip (New Objective, Woburn, USA) coupled to a microTOF MS (Bruker Daltonic, Bremen, Germany) [24]. A solution of 20% acetonitrile (Sigma-Aldrich, Taukirchen, Germany) in HPLC-grade water with 0.94% formic acid (Sigma-Aldrich) was used as running buffer. The ESI sprayer (Agilent Technologies, Palo Alto, CA, USA) was grounded, and the ion spray interface potential was set between −4 and −4.5 kV. Data acquisition and MS acquisition methods were automatically controlled by the CE via contact-close-relays. For each sample, spectra were accumulated every 3 s, over a range of m/z 350–3000 over 60 min [24]. The accuracy, precision, selectivity, sensitivity, reproducibility, and stability of the CE–MS measurements have been previously demonstrated [12].

2.6.3. CE–MS data processing
Mass spectra were processed using MosaiquesVisu software (Mosaiques Diagnostics, Germany), including peak picking, deconvolution and deisotoping [25]. The software automatically examined all mass spectra from a CE–MS analysis for signals above the threshold (Signal-Noise-Ratio > 4). Only signals that were present in 3 consecutive spectra were accepted. The isotopic distribution was assessed and charge was assigned based on the isotopic distribution, using a matched filtering algorithm. This operation resulted in a list of signals defined by mass/charge, charge, migration time, and signal intensity (relative abundance defined by ion counts). This list was transformed into a dataset containing only mass, migration time, and signal intensity; signals that represent the same compound but with a different charge state were combined.

To allow for compilation and comparison of samples, normalised signal intensity was used as a measure of relative abundance. CE migration time and ion signal intensities of the samples were calibrated and normalised using internal polypeptide standards by linear regression [26]. ‘Housekeeping’ peptides that consistently appear in urine samples and are unaffected by disease states provided ideal reference mapping points for CE migration times [15]. Data sets were accepted only if they passed a strict quality control criteria: a minimum of 950 chromatographic features (mean number of features minus one standard deviation) must be detected with a minimal MS resolution of 8000 (required resolution to resolve ion signals with z ≤ 6) in a minimal migration time interval (the time window, in which separated signals can be detected) of 10 min. After calibration, the mean deviation of migration time (compared to reference standards) was below 0.35 min [24]. The resultant peak list included molecular mass (Da), normalised CE migration time (min) and normalised signal intensity for each peptide. All results were annotated into a Microsoft SQL database for statistical analysis.

Peptide profiles for OSA and non-OSA patients were compared for significant differences. Subsequently, the identity of the peptides was determined by matching against an validated database that was previously developed using liquid chromatography–MS/MS analysis on a quadrupole time-of-flight mass spectrometer [15]. These human urinary peptide sequences can be accessed at (http://mosaiques-diagnostics.de/diapatpcms/mosaiquescms/front_content.php?idcat=257).

2.7. Statistical analysis
Patient demographics were summarised by OSA group as means (SD) (or median (IQR) if non-normal) for continuous variables and frequencies (%) for categorical variables. Comparisons between groups were performed using a t-test for continuous variables which were normally distributed and a Mann–Whitney test for those that were not. Categorical variables were tested using a Chi-Square or Fishers Exact test depending on the expected frequencies. Assessment for normal distribution of patient data was by Shapiro–Wilk testing. Statistical significance for patient data was assigned at p<0.05. Statistical analyses were carried out using SPSS version 20 (IBM Corp, Armonk, NY, USA).

For proteomics analyses, the non-parametric Wilcoxon test has been shown to be well-suited for proteomics data sets [27], and was used to determine significance in peptide abundance between the groups. Statistical significance was assigned at p ≤ 0.01. Given the large number of possible peptides present, assessment of differences included an adjustment for multiple testing. Correction for multiple testing to account for false positives (false discovery rate) was performed using the Benjamini–Hochberg test.

3. Results
Fifty-three subjects who had been assessed at baseline were recruited. One urinary sample could not be analysed by CE–MS and therefore was excluded from the analysis. Therefore, 52 patients were entered into the study; 27 with OSA who were on CPAP and 25 without OSA (non-OSA) (Fig. 1). All subjects were of white European ethnicity. Patients attended their follow-up after a median of 15 (IQR 13,16) months. For patients with OSA, the mean CPAP use was 14.5 months (SD 1.0) and mean CPAP use per night was 4.6 h (SD 0.6). Mean CPAP pressure was 10 cmH2O (SD 1.0).

Clinical and biochemical parameters of the 52 patients according to those without OSA (non-OSA) and OSA-on-CPAP groups are presented in Table 1. The groups did not significantly differ in age, BMI and gender. More OSA subjects on CPAP had metabolic syndrome, particularly meeting the waist circumference, blood pressure, HDL and triglycerides components of the criteria. Although OSA subjects tended to have higher waist circumference, neck circumference, waist: hip ratio and body fat percentage by bioimpedance, these differences were not statistically significant. Smoking, diabetes, alcohol consumption and medication use were similar between groups.
Changes in vasoactive peptides and proinflammatory cytokines, and activity of enzymes that process procollagen precursors to mature collagen including procollagen protein fragments of collagen alpha-1 (I) and (III) in addition to fibrinogen beta chain and cadherin 13 (T-Cadherin) peptides. Fibrinogen beta chain has a role in fibrinogen synthesis and haemostasis, blood viscosity as a binding surface for proteins and as an extracellular matrix component, and may be associated with cardiovascular risk factors [28]. T-Cadherin is highly expressed in the vasculature including endothelial cells, has been implicated in the modulation of angiogenic activities and is associated with circulating levels of adiponectin that is important for vascular homoeostasis. Additionally, links between T-cadherin and blood pressure, lipids, metabolic syndrome, type 2 diabetes and ischaemic stroke have been found in genome-wide association studies [29]. It should be noted that the mean amplitude of fibrinogen beta chain and cadherin-13 was lower in the CPAP-treated patients and it is possible that this may reflect decreased breakdown or differential expression with treatment.

The extracellular matrix contains a fibrillar collagen network that is regulated by fibroblasts and myofibroblasts. Changes in vasoactive peptides and proinflammatory cytokines, and activity of enzymes that process procollagen precursors to mature collagen including procollagen proteinases and collagen degradation enzymes (matrix
metalloproteinases) may account for the excretion of urinary collagen fragments [30]. Furthermore, it is known that alterations of the myocardial collagen matrix may occur in cardiovascular disease that may result in perturbations in vascular function [30]. In this context, it is known that OSA is associated with cardiovascular disease [31], and the peptides in the peptide panel may be a reflection of underlying cardiovascular changes associated with CPAP.

The mechanisms underlying the observed urinary peptide profile in this study remain unclear, but may be related to the treatment effects of CPAP on intermittent hypoxia and systemic hypertension. Post-translational modification of proteins has been postulated as a mechanism by which protein function is influenced by chronic intermittent hypoxia, and there is evidence that this induces changes in the phosphorylation state of proteins associated with transcriptional activation, signal transduction pathways, and neurotransmitter synthesis [32]. Multiple lines of evidence underscore the importance of CPAP treatment in lowering BP in patients with severe OSA [33]. Additionally, OSA-mediated hypertension may be influenced by the effects of CPAP on reducing renin-angiotensin system activity with changes in renal perfusion [17,18]. It is conceivable that the treatment effects of CPAP on OSA disease progression may lead to the modification of proteins and changes in protease activity, and renal effects that may be reflected in the urinary proteome [15].

This study was limited by the lack of serial measurements of urine samples throughout the follow-up period and by

| Table 1 – Patient demographics were summarised by treated OSA and non-OSA groups as means (SD) (or median (IQR) if non-normal) for continuous variables and frequencies (%) for categorical variables. Significant p-values are highlighted in bold. |
|---------------------------------------------------------------|
| **Patient characteristics**                                   |
| **Non-OSA**                                                   | **OSA on CPAP**                                 | **p-Value** |
| **Age (years)** 52 (9)                                        | 49 (7)                                         | 0.2252      |
| **Gender: male** 13 (52%)                                     | 15 (56%)                                      | 0.7972      |
| **Smoked in past** 6 (24%)                                    | 7 (26%)                                       | 0.8727      |
| **Alcohol intake: <8 units per week**                         |                                               |             |
| **BMI (kg/m²)** 17(68%)                                       | 19 (70%)                                      | 0.8532      |
| **Neck circumference (cm)** 42.1(4.3)                          | 44.2 (4.6)                                    | 0.0916      |
| **Waist hip ratio** 0.94 (0.1)                                 | 0.98 (0.1)                                    | 0.2069      |
| **Body fat (%) TANITA scales** 42.8 (8.8)                      | 44.4 (8.4)                                    | 0.5029      |
| **Diabetes** 5 (20%)                                          | 9 (33%)                                       | 0.2788      |
| **Hypertension** 5 (20%)                                      | 6 (22%)                                       | 0.8446      |
| **BP-lowering medications** 5 (20%)                           | 6 (22%)                                       | 0.8446      |
| **Glucose-lowering therapy** 5 (20%)                          | 9 (33%)                                       | 0.2788      |
| **Lipid-lowering medications** 8 (32%)                        | 9 (33%)                                       | 0.9184      |
| **Metabolic syndrome** 6 (24%)                                 | 14 (52%)                                      | 0.0391      |
| **Cardio-respiratory parameters**                             |                                               |             |
| **Systolic BP (mmHg)** 128 (11)                               | 129 (11)                                      | 0.9684      |
| **Diastolic BP (mmHg)** 82 (8)                                 | 79 (7)                                        | 0.2276      |
| **Mean Arterial Pressure (mmHg)** 100 (91,104)                 | 98 (93,101)                                   | 0.161       |
| **Heart rate (beats per min):** 71 (10)                        | 74 (10)                                       | 0.2775      |
| **CPAP use per night (h)**                                    |                                               |             |
| **CPAP pressures (cmH2O)**                                    |                                               |             |
| **Epworth sleep score:** 4 (3, 9)                             | 3 (2, 11)                                     | 0.5015      |
| **O2 saturations (%):** 97 (96,99)                            | 97 (96,98)                                    | 0.9186      |
| **PCO2 (kPa)** 5.4 (5.1, 5.6)                                 | 5.3 (5.1, 5.5)                                | 0.3882      |
| **AHI at baseline** 2.4 (1.6, 3.7)                            | 24.6 (14.0, 43.6)                             | < 0.001     |
| **Metabolic parameters**                                      |                                               |             |
| **Albumin Creatinine Ratio (mg/mmol)** 0.4 (0.2, 0.5)          | 0.5 (0.3, 0.8)                                | 0.1475      |
| **Creatinine (micromol/l)** 77 (13)                            | 78 (15)                                       | 0.8252      |
| **MDRD-GFR (mL/min/1.73 m²)** 87 (13)                          | 89 (15)                                       | 0.542       |
| **Low density lipoprotein (mmol/l)** 2.7 (1.1)                 | 2.6 (0.9)                                     | 0.7637      |
| **High density lipoprotein (mmol/l)** 1.3 (0.4)                | 1.2 (0.3)                                     | 0.3228      |
| **HbA1c (mmol/mol)** 37 (34, 40)                               | 38 (35, 48)                                   | 0.1466      |
| **Total cholesterol (mmol/l)** 4.7 (1.1)                       | 4.6 (1.0)                                     | 0.7119      |
| **Fasting glucose (mmol/l)** 5.4 (5.1, 5.9)                    | 5.4 (4.7, 6.0)                                | 0.8905      |
| **Thyroid stimulating hormone (mUI/l)** 2.2 (1.3)              | 2.1 (0.9)                                     | 0.9275      |
| **Triglycerides (mmol/l)** 1.2 (0.8, 2.0)                      | 1.6 (1.0,2.6)                                 | 0.1634      |
| **Bicarbonate (mmol/l)** 24.0 (1.8)                            | 24.2 (2.1)                                    | 0.6896      |
| **Highly sensitive C-Reactive Protein (hsCRP) (mg/L)** 3.3 (1.1, 6.8) | 3.0 (1.6, 5.8)                               | 0.8189      |

Abbreviations: AHI: Apnoea-Hypopnoea index, BMI: body mass index, BP: blood pressure, CPAP: continuous positive airway pressure, HbA1c: glycated haemoglobin, MDRD-GFR: Glomerular Filtration Rate using Modification of Diet in Renal Disease Equation, PCO2: partial pressure of carbon dioxide.
the presence of potential confounders such as metabolic syndrome and patient medications. It may be argued that the excretion of urinary peptides may vary during the day, because of physical activity, diet, or medications taken. We sought to control for this by sampling all participants at the same time each morning, at second void, after an overnight fast at their study visit prior to the administration of any medication. We envisaged that it was important for the urinary polypeptide panel to reflect ‘real-life’ clinical patients for it to be relevant and applicable. Therefore, controlling for every potential confounding factor would not have been pragmatic and would have limited recruitment of potential participants. Repeat polysomnography was not conducted to prove if the AHI was sufficiently reduced on CPAP but the patients were managed according to local hospital protocol and were already established on their treatment at follow-up, with improved symptoms and ESS scores as assessed at their compliance/adherence clinic visits.

There were more patients in the CPAP-treated OSA group who had metabolic syndrome than non-OSA patients. This...
| Peptide ID | Peptide mass (Da) | CE time (min) | Unadjusted Wilcoxon p-value | Area under the curve | Benjamini and Hochberg p-value | Sequence | Protein name | Start amino acid | Stop amino acid | SwissProt name Accession number | Frequency OSA on CPAP | Mean amplitude OSA on CPAP (SD) | Frequency non-OSA | Mean amplitude non-OSA (SD) |
|-----------|------------------|--------------|-----------------------------|---------------------|---------------------------|----------|--------------|----------------|----------------|-----------------------------|----------------|--------------------------|----------------|--------------------------|
| 61039     | 1813.715         | 31.69499     | 0.000532                    | 0.77                | 0.4099525                 | GmpGSpGSpGmpGSpGSpGSpG | Collagen alpha-1 (III) chain | 537            | 555            | CD3A1_HUMAN g124056490 | 1.0            | 1557 (856)               | 1.0            | 2616 (1196)               |
| 55756     | 1700.721         | 31.99699     | 0.00096                     | 0.76                | 0.4099525                 | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 816            | 854            | CD1A1_HUMAN g124056487 | 0.8            | 699 (487)                | 0.9            | 1237 (625)                |
| 130747    | 3359.578         | 31.89787     | 0.001473                    | 0.75                | 0.511072                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 800            | 810            | CD1A1_HUMAN g124056487 | 0.7            | 69 (58)                  | 0.3            | 24 (38)                  |
| 167576    | 4759.125         | 29.94851     | 0.002237                    | 0.73                | 0.582167                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 180            | 198            | CD3A1_HUMAN g124056490 | 1.0            | 259 (87)                 | 1.0            | 200 (112)                 |
| 11249     | 980.2951         | 35.57353     | 0.000419                    | 0.73                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 430            | 455            | CD1A1_HUMAN g124056487 | 0.9            | 942 (69)                 | 1.0            | 1311 (538)                |
| 54269     | 1675.708         | 35.20131     | 0.000473                    | 0.72                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 32874          | 1333.416       | FGB protein (Fragment) | 0.7            | 186 (155)                | 0.8            | 321 (184)                 |
| 131294    | 3375.574         | 33.91691     | 0.000508                    | 0.72                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 91249          | 2401.078       | CD3A1_HUMAN g296434458 | 0.7            | 46 (50)                  | 0.3            | 16 (26)                  |
| 90651     | 2385.054         | 33.94877     | 0.000609                    | 0.72                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 57384          | 1738.76        | Cadherin-13             | 0.8            | 711 (477)                | 1.0            | 1115 (196)                |
| 88299     | 2339.085         | 23.66349     | 0.000638                    | 0.72                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 87411          | 2321.165       | FGB protein (Fragment) | 0.4            | 42 (69)                  | 0.7            | 113 (125)                 |
| 91249     | 2401.078         | 23.05962     | 0.000784                    | 0.70                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 19969          | 1132.569       | Cadherin-13             | 0.7            | 49 (48)                  | 0.4            | 20 (29)                  |
| 57384     | 1738.76          | 25.0428      | 0.000928                    | 0.71                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 1221           | 1232           | CD3A1_HUMAN g296434458 | 0.7            | 49 (48)                  | 0.4            | 20 (29)                  |
| 87411     | 2321.165         | 22.05962     | 0.000784                    | 0.70                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 33274          | 1333.416       | FGB protein (Fragment) | 0.4            | 42 (69)                  | 0.7            | 113 (125)                 |
| 90651     | 2385.054         | 33.94877     | 0.000609                    | 0.72                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 87411          | 2321.165       | FGB protein (Fragment) | 0.4            | 42 (69)                  | 0.7            | 113 (125)                 |
| 88299     | 2339.085         | 23.66349     | 0.000638                    | 0.72                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 91249          | 2401.078       | CD3A1_HUMAN g296434458 | 0.7            | 49 (48)                  | 0.4            | 20 (29)                  |
| 91249     | 2401.078         | 23.05962     | 0.000784                    | 0.70                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 19969          | 1132.569       | Cadherin-13             | 0.7            | 49 (48)                  | 0.4            | 20 (29)                  |
| 57384     | 1738.76          | 25.0428      | 0.000928                    | 0.71                | 0.645249                  | GmpGSpGSpGSpGSpGSpG | Collagen alpha-1 (III) chain | 1221           | 1232           | CD3A1_HUMAN g296434458 | 0.7            | 49 (48)                  | 0.4            | 20 (29)                  |

SD: standard deviation. FGB: fibrinogen beta chain. OSA-on-CPAP: subjects with obstructive sleep apnoea receiving CPAP. Non-OSA: subjects without obstructive sleep apnoea.
was attributable to a combination of factors including waist circumference, blood pressure and lipid profiles. It would be difficult to separate metabolic syndrome from OSA in severe obesity as these conditions are closely associated. The incorporation of such factors in the discovery process may potentially lead to peptide profiles that may be more representative of patients with OSA in the severely obese population.

This study was limited by the relatively small sample size. Additionally, we did not directly compare the urine proteome in OSA patients directly before commencing CPAP treatment with findings after CPAP. However, it must be emphasised that this was foremost a discovery study, with the aim to use CE–MS to investigate urinary profiles in severely obese subjects with OSA who were receiving CPAP in relation to subjects without OSA. Our findings provide a novel insight into urinary peptide patterns in a well-characterised cohort of severely obese patients with OSA who received CPAP, and subjects without OSA. As this study was based on samples collected from patients at a specific point in time, a cause-and-effect relationship between the observed urinary proteome and CPAP itself cannot be established. Nevertheless, it would have been ethically questionable to perform a randomised placebo-CPAP controlled study comparable to our duration of CPAP use (>1 year) which is a strength of this study.

5. Conclusion

The findings from this study provide a unique insight of the urinary proteome in OSA with CPAP. The identified peptide panel includes collagen, cadherin and fibrinogen subtypes that may be related to mechanisms underlying cardiovascular disease in OSA. This may be associated with the treatment effects of CPAP on OSA disease progression influencing the expression of urinary peptides. There is a potential role for urinary proteomics in characterising urinary peptide profiles in OSA. Further work is still needed to investigate the use of proteomic methods in defining peptide profiles that reflect OSA and in the presence of CPAP that may potentially support CPAP treatment monitoring and progress or relating to the changes in cardiovascular risk profiles, potentially translating into better monitoring of OSA treatment and aid in the care of patients with sleep disorders.

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Conflict of interest statement

AT is employed by Mosaiques Diagnostics GmbH. HM is co-founder and co-owner of Mosaiques Diagnostics GmbH. All other authors have nothing to declare.

Contribution

IS, RP, SC, ND, KH, JB, JW contributed to the design, analyses and conduct of the study. IS performed the CE–MS experiments. IS, JB, AT, AA, WM, HM contributed to the proteomic analyses. All authors contributed to the writing of the manuscript.

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