Biogenic amines degradation by *Lactobacillus plantarum*: toward a potential application in wine

Vittorio Capozzi, Pasquale Russo, Victor Ladero, María Fernández, Daniela Fiocco, Miguel A. Alvarez, Francesco Grieco and Giuseppe Spano*  

*Correspondence: Giuseppe Spano, Department of Food Science, University of Foggia, Via Napoli 25, Foggia 71122, Italy. e-mail: g.spano@unifg.it

Biogenic amines (BA) in wine represent a toxicological risk for the health of the consumer, with several trade implications. In this study 26 strains of *Lactobacillus plantarum* were analyzed for their ability to degrade BA commonly found during wine fermentation. Two strains of *L. plantarum* were selected in reason of their ability to degrade putrescine and tyramine. The degradation was assessed *in vitro*, both in presence of the BA and in presence of the specific chemical precursor and of producer bacteria. The two *L. plantarum* biotypes were found capable to work synergically. In addition, the survival in wine-like medium and the aptitude to degrade malic acid after alcoholic fermentation of the selected *L. plantarum* strains was analyzed. Our results suggest the potential application of wine *L. plantarum* strains to design malolactic starter cultures able to degrade BA in wine.

Keywords: lactic acid bacteria, amine degradation, biogenic amines, malolactic fermentation, wine, *Lactobacillus plantarum*, putrescine, tyramine

INTRODUCTION

Biogenic amines (BA) are low molecular weight organic bases with aliphatic, aromatic, and heterocyclic structures commonly found in fish, fish products, meat, dairy products, vegetables, wine, cider, and beer (Suzzi and Gardini, 2003; Spano et al., 2010). In general, foods likely to contain high levels of BA are fermented foods or foodstuff exposed to microbial contamination during food process or storage (Ali, 1996). Histamine, tyramine, cadaverine, 2-phenylethylamine, spermine, spermidine, putrescine, tryptamine, and agmatine are considered to be the most important BA occurring in foods. The main bacteria responsible for BA production in fermented food matrices are the lactic acid bacteria (LAB; Lonvaud-Funel, 2001). In fact, LAB can catabolize (principally decarboxylate) amino acids into amine-containing compounds. The physiological role of BA synthesis mainly seems to be related to defense mechanisms used by bacteria to withstand acidic environments (Spano et al., 2010).

Indigenous BA compounds are also naturally produced in different human tissues because of their biological role in processes such as synaptic transmission, blood pressure control, allergic response, and cellular growth control (Russo et al., 2010). The gastro-intestinal tract, as a function of dietary intake of food containing BA, therefore represents an exogenous source of these molecules for humans. This exogenous source of BA can provoke high levels in the human organism, and in reason of their importance in physiological processes, with negative consequences to human health (Ladero et al., 2010; Russo et al., 2010). Human sensitivity fluctuates according to the correct functioning of the detoxification systems since BA are generally metabolized in the human gut through the action of amine oxidases (Spano et al., 2010). Under physiological conditions this system has a protective role against the absorption of dietary BA. For this reason, in individuals with a pathological deficiency of amine oxidases activities, the ingestion of food products containing excess of BA may lead to high levels of BA in the organism (Kuefner et al., 2004). Additionally, the ethanol and some drugs have been found to inhibit the detoxification systems (Zimaktin and Anichtchik, 1999; Horton et al., 2005). In the wine industry, the occurrence of BA has been receiving increasingly attention in reason of the potential threats of toxicity to humans, connected with alcohol content (Smit et al., 2008). There are trade implications due to the recommended or suggested existing limits for histamine in wine in some European countries such as Germany (2 mg/l), Holland (3 mg/l), Finland (5 mg/l), Belgium (5–6 mg/l), France (8 mg/l), Switzerland, and Austria (10 mg/l; Smit et al., 2008). In addition, last year, the Panel on Biological Hazards of the European Food Safety Authority release a Scientific Opinion on risk based control of BA formation in fermented foods 9 [EFSA Panel on Biological Hazards (BIOHAZ), 2011], testifying the attention of European institutions to the subject.

In reason of the toxicity and of the existing legislative limits, several technological strategies have been proposed in order to control BA in foods (Naila et al., 2010). The main measure to prevent BA formation is the microbial control through cold treatments. However, it is difficult to operate with low temperature in the case of fermented foods, where a microbial growth is crucial to obtain the final products. Secondary control measures to prevent BA formation in foods or to reduce their levels once formed need to be considered as alternatives. Methods to limit microbial growth may include hydrostatic pressures, irradiation, controlled atmosphere...
Packaging, or the use of food additives (Naila et al., 2010). Furthermore, BA could be controlled, especially in fermented foods, modulating microbial resources. Indeed, fermented foods producers commonly utilize a specific starter culture, that is “a microbial preparation of large numbers of cells of at least one microbial species to be added to a raw material to produce a fermented food by accelerating and steering its fermentation process” (Leroy and De Vuyst, 2004). In general, the choice of starter cultures is crucial to guarantee the quality of the final products and microbial starter unable to produce BA might dominates the autochthonous microbiota, reducing the occurrence of BA producers (Dapkevicius et al., 2008; Latorre-Moratalla et al., 2007; Naila et al., 2010; Spano et al., 2010). Additionally, starter may control BA in food via their amine oxidase activity because oxidizes BA into aldehyde, hydrogen peroxide, and ammonia (Suzzi and Gardini, 2003; Naila et al., 2010; Spano et al., 2010). In red wine, the most abundant amines are usually putrescine, histamine, tyramine, and cadaverine (Beneduce et al., 2010), the last being considered the most important reason for wine intolerance (Konakovskiy et al., 2011). Inoculation with starter cultures that are unable to produce BA is a feasible option for the control of these chemical substances in wine (Martín-Álvarez et al., 2005; Spano et al., 2010). However, it does not represent the definitive solution when the dominance of starter cultures on the indigenous BA-producer microflora is not sufficient. Recently, García-Ruiz et al. (2011) isolated some LAB from wine and other oenological source and tested their ability to degrade BA. Whereas a negative influence of the wine matrix was observed, their findings indicated a potential application of wine LAB in order to reduce BA in wine (García-Ruiz et al., 2011).

In this paper we report the selection of autochthonous strains of Lactobacillus plantarum able to degrade BA and their suitability to be used as malolactic starter in wine production.

MATERIALS AND METHODS

BACTERIAL STRAINS AND GROWTH CONDITIONS

The LAB strains included in this study were routinely cultured on MRS (de Man-Rogosa–Sharp, Oxoid, Milan, Italy) medium. The pH of the medium was adjusted to pH 5.8 with KOH before sterilization for 15 min at 121°C. Samples from 10 natural vinifications of “Nero di Troia” wines were aseptically collected at the early stages of malolactic fermentation from winery located in Apulia region (Italy). For microbiological analyses, 10 ml of each sample were homogenized with 90 ml of a saline–peptone water, after which serial dilutions were prepared. For the isolation of LAB, MRS agar containing 100 mg/l of cycloheximide was used, and the plates were incubated under anaerobic conditions (BBL, GasPack-System) at 30°C for 72 h. Isolates were identified as bacilli and putative LAB by positive Gram staining and negative catalase assay. All strains were stored at −80°C in MRS supplemented with glycerol (20% v/v).

STRAINS IDENTIFICATION

Genomic DNA of putative LAB was isolated using the Microbial DNA extraction kit (Cabru, Milan, Italy) according to manufacturer’s procedure.

Preliminary identification of L. plantarum strains was performed by amplification of the recA gene. Primers planF and pREV were used in the PCR tests (Torriani et al., 2001). For PCR amplification about 50 ng of genomic DNA were added to a 50 μl PCR mixture and amplified with Taq DNA Polymerase (Qiagen, Milan, Italy). The temperature profile was as follows: 94°C for 5 min; 30 cycles of 94°C for 0.5 min, 46°C for 0.5 min, 72°C for 0.5 min; 72°C for 5 min. Furthermore, the isolated strains were identify by partially sequencing the 16S rRNA-encoding gene. Most of the gene encoding the 16S ribosomal RNA was amplified by PCR using primers pA and pH according to Edwards et al. (1989). The amplicons were purified using QIAquick PCR Purification Kit (Qiagen, Milan, Italy) and sequenced. The resulting sequences were compared with sequences available at NCBI database (GenBank) using the standard nucleotide–nucleotide homology search basic local alignment search tool (BLAST; http://www.ncbi.nlm.nih.gov/BLAST).

In order to verify if the bacteria analyzed in this study harbor genes encoding enzymes involved in production of BA such as tyramine, histamine, and putrescine, a multiplex-PCR-based assay was carried out to detect the presence of tyrosine decarboxylase (tdc), histidine decarboxylase (hdc), and ornithine decarboxylase (ode) genes (Coton et al., 2010). About 50 ng of genomic DNA was added to a 50 μl PCR mixture and amplified with Taq DNA Polymerase (Qiagen, Milan, Italy). The reaction mix was cycled through the following temperature profile: 94°C for 5 min; 15 cycles of 94°C for 1 min, 55°C for 1.3 min, 72°C for 1 min; 12 cycles of 94°C for 1 min, 58°C for 1.3 min, 72°C for 1 min; 72°C for 5 min.

PCR fragments were analyzed by agarose gel electrophoresis carried out by loading 10 μl of sample on 1.2% agarose gels.

ANALYSIS OF BA BY HPLC AND PRELIMINARY SCREENING

In order to test the individual degrading capacity of isolated strains, BA concentration was quantified in MRS broth samples supplemented with putrescine (1 mM), tyramine (1 mM), cadaverine (1 mM), or histamine (2.5 mM) after 24 h with or without strain inoculation (2 × 10⁷ CFU ml⁻¹; total volume 20 ml). One milliliter of each sample was centrifuged and the supernatant was filtered through 0.45 μm PTFE filters (WVR, USA). Five microliters were derivatized using dansyl chloride, separated, and analyzed using the conditions described by Ladero et al. (2011).

VALUATION OF DEGRADING BACTERIA IN VITRO

MRS broth samples (1 ml) supplemented with putrescine (1 mmol l⁻¹), tyramine (1 mmol l⁻¹), cadaverine (1 mmol l⁻¹), and histamine (2.5 mmol l⁻¹) were collected after 24 h with and without degrader strain inoculation (starting from an OD₆₀₀ of about 0.05; total volume 1 l). In a separate trial the two strains were co-inoculated (starting from an OD₆₀₀ of about 0.05 for each strain; total volume 1 l). MRS broth samples (1 ml) supplemented with tyrosine (15 mmol l⁻¹) or agmatine (4.38 mmol l⁻¹; Arena et al., 2011) were collected after 24 h from inoculation with degrader strain (starting from an OD₆₀₀ of about 0.05) and producer strain (starting from an OD₆₀₀ of about 0.05; total volume 1 l). BA content was determined as previously indicated and degration activity was expressed as a percentage of the control without degrader strain. The producer strains were Lactobacillus brevis IOEB 9809 (Lucas et al., 2007; Arena et al., 2010) and Enterococcus.
**Table 1** Degradation (in percentage) of putrescine, tyramine, and histamine by *Lactobacillus plantarum* strains isolated from wine after 24 h of culture in MRS broth (20 ml) supplemented with putrescine (1 mM), tyramine (1 mM), histamine (2.5 mM), and cadaverine (1 mM).

| Strain            | Putrescine | Tyramine | Histamine | Cadaverine |
|-------------------|------------|----------|-----------|------------|
| *L. plantarum* NDT 03 | 7.15       | n.d.     | 8.63      | n.d.       |
| *L. plantarum* NDT 09 | 3.47       | 22.12    | 2.21      | 1.83       |
| *L. plantarum* NDT 10 | 11.05      | n.d.     | 3.66      | n.d.       |
| *L. plantarum* NDT 16 | 31.09      | 2.75     | 7.49      | 3.41       |
| *L. plantarum* NDT 21 | 8.72       | 13.33    | 7.19      | 6.68       |

Degradation activity is expressed as a percentage of control without strain. The data presented are mean values from three separate experiments; n.d., no detected effect.
Survival in Wine-Like Medium and Malolactic Fermentation of *L. Plantarum* NDT 09 and *L. Plantarum* NDT 16

In order to analyze the survival and fermentation suitability of *L. Plantarum* strains able to degrade BA, the strains were inoculated in a wine-like medium. The synthetic must was subjected to fermentation by a commercial oenological yeast strain to achieve 12.1% v/v of ethanol content. In three independent experiments, MLF was induced with *L. Plantarum* strains NDT 09 and NDT 16 previously adapted at pH 3.5. The results obtained by the quantification of bacterial biomass and l-malic acid degradation are reported in Figure 3. The sampling times were 0, 1 h, 1, 2, 3, 7, 14, and 21 days after strain inoculation. Neither degradation of l-malic acid nor spontaneous growth was observed within the control wine (uninoculated wine). The *L. Plantarum* NDT 09 and *L. Plantarum* NDT 16 strains were able to survive in the wine-like medium (Figure 3). An initial decrease in the survival with respect of expected population (2 × 10⁶ CFU ml⁻¹) was noticed after the inoculation in wine, probably due to the harsh conditions imposed to bacterial cells. Bacterial population remained stable after 1 h. Subsequently, LAB biomass concentration diminished progressively up to the seventh day post inoculation (Figure 3). The strain *L. Plantarum* NDT 16 better survive in wine-like medium than *L. Plantarum* NDT 09 (Figure 3). At the 14th day after inoculation, no viable cell was detected. Both strains were found able to degrade l-malic acid. The efficient consumption of this dicarboxylic acid was conducted up to the 14th day after inoculation (Figure 3). *L. plantarum* NDT 16 was found to better degrade l-malic acid than *L. plantarum* NDT 09. After the 14th day, consistently to the findings in the survival, also l-malic acid degradation stopped. The growth level and the l-malic acid consumption rates showed to be strain-dependent. Overall, the results reported demonstrated that the analyzed strains could be tested as malolactic strains for wine with low l-malic acid contents.

**DISCUSSION**

The biotechnological approach reported in this paper, aimed to conciliate malolactic fermentation and BA degradation within the same biotechnological resources: the malolactic starter cultures. The main species involved in malolactic fermentation in wine is *Oenococcus oeni*. However, as reported by García-Ruiz et al. (2011), in the wine environment, the highest potential for BA degradation among LAB seem to be related to *Lactobacillus* and *Pediococcus* species, while *O. oeni* demonstrated low and rare degradation characteristics (García-Ruiz et al., 2011). In the last years, *Lactobacillus* spp. received increasing attention as an important resource for the design of a new generation of malolactic fermentation starter cultures (du Toit et al., 2010), insomuch as we found in commerce malolactic starter formulate using *L. plantarum* V22 strain released in 2010 by Lallemant (Cho et al., 2011; Miller et al., 2011). Other than malic acid decarboxylation, the main advantages of *L. plantarum* employed were: resistance to the stressing wine conditions, production of wine aroma compounds, production of plantaricins, all important factors to compete with spoilage LAB (du Toit et al., 2010).

In this work, we selected two *L. plantarum* strains (named NDT 09 and NDT 16) from a pool of *L. plantarum* strains isolated from red wine undergoing malolactic fermentation able to degrade BA such as putrescine and tyramine. The degradation rates observed was higher than that previously reported for *L. plantarum* strains (Leuschner et al., 1998; García-Ruiz et al., 2011). Furthermore, the strains were evaluated in wine-like medium for...
FIGURE 3 | Evolution of the bacterial population during MLF in wine-like medium separately carried out by the two selected Lactobacillus plantarum strains. Samples were taken at the time of inoculation (time zero) and after 1 h, 1, 2, 3, 7, 14, and 21 days. L. plantarum NDT 09 (open squares, continuous line) and L. plantarum NDT 16 (filled rhombi, continuous line) counts on de Man Rogosa Sharpe agar were reported in figure. Also the evolution of l-malic acid consumption during MLF was monitored (L. plantarum NDT 09, open squares, dotted line; L. plantarum NDT 16, filled rhombi, dotted line). Data are the mean ± SD, for n = 3, Figure 1.

their suitability to degrade malic acid. Although MLF was incompletely performed, the two biotypes showed a respectable aptitude to degrade malic acid, indicating a possible application in reason of wine/must malic acid content. As BA production ability was found to be both species- and strain-dependent (Bover-Cid and Holzapfel, 1999; Russo et al., 2010; Ladero et al., 2012), our work confirmed the scientific evidence indicating a species- and strain dependence also in BA degradation by LAB. Our findings provided the basis for an innovative approach that might be of particular interest for hot climate regions where grapes are harvested at high maturity usually resulting in wine with pH higher than pH 3.5, high ethanol concentration, low malic acid content, and with considerable spoilage-associated risks. Further studies are ongoing in order to select new L. plantarum strains able to degrade histamine, and to analyze the influence of wine conditions on BA degradation and malic acid consumption by L. plantarum NDT 09 and L. plantarum NDT 16. Indeed, as indicated by García-Ruiz et al. (2011), wine chemicals such as ethanol (12%) and polyphenols (75 mg/l), and additives like SO2 (30 mg/l) may reduce the degrading ability of LAB strains.

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