Novel Single Nucleotide Polymorphisms in Interleukin 6 Affect Tacrolimus Metabolism in Liver Transplant Patients

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Abstract

Background: Tacrolimus is the first-line immunosuppressant after organ transplantation. It is mainly metabolized by cytochrome P450, family 3, subfamily A (CYP3A) enzymes, but there are large individual differences in metabolism. Interleukin 6 (IL6) has been shown to cause a pan-suppression of mRNA levels of ten major CYP enzymes in human hepatocyte cultures. IL6 has been shown to provide hepatoprotection in various models of liver injury. Rs1800796 is a locus in the IL6 gene promoter region which regulates cytokine production. We speculated that IL6 rs1800796 polymorphisms may lead to individual differences in tacrolimus metabolism by affecting CYP3A enzymes levels and liver function after liver transplantation.

Methodology/Principal Findings: Ninety-six liver transplant patients receiving tacrolimus were enrolled in the study. Two single nucleotide polymorphisms (SNP), CYP3A5 rs776746 and IL6 rs1800796, were genotyped in both donors and recipients. The effects of SNPs on tacrolimus concentration/dose (C/D ratio) at four weeks after transplantation were studied, as well as the effects of donor IL6 rs1800796 polymorphisms on liver function. Both donor and recipient CYP3A5 rs776746 allele A showed association with lower C/D ratios, while donor IL6 rs1800796 allele G showed an association with higher C/D ratios. Donor CYP3A5 rs776746 allele A, IL6 rs1800796 allele C, and recipient CYP3A5 rs776746 allele A were associated with fast tacrolimus metabolism. With increasing numbers of these alleles, patients were found to have increasingly lower tacrolimus C/D ratios at time points after transplantation. Donor IL6 rs1800796 allele G carriers showed an association with higher glutamic-pyruvic transaminase (GPT) levels.

Conclusions: Combined analysis of donor CYP3A5 rs776746, IL6 rs1800796, and recipient CYP3A5 rs776746 polymorphisms may distinguish tacrolimus metabolism better than CYP3A5 rs776746 alone. IL6 may lead to individual differences in tacrolimus metabolism mainly by affecting liver function.

Introduction

Tacrolimus is the first-line immunosuppressant after organ transplantation, reducing rejection and improving graft and recipient survival. However, it is also characterized by a narrow therapeutic window, and large individual differences in metabolism [1,2]. Indeed, after administration of “standard” doses, some patients show no therapeutic effects or serious side effects. Individualization of medical treatment could result in great improvement in therapeutic efficacy, and reduction of side effects, as well as reduction of the cost of treatment [3,4]. At present, it is difficult to institute individualized medicine in the early postoperative period. Genetic factors such as polymorphisms can be closely related to drug metabolism. There is growing interest in the field of pharmacogenomics which focuses on the relationship between host genetics and drug metabolism [5]. Large clinical studies have shown that genetic factors can guide individualized medication of warfarin and clopidogrel [6,7]. Pharmacogenomics research on tacrolimus could contribute to individualized medication in the early postoperative period of liver transplantation.

CYP3A enzymes, which are mainly expressed in liver and intestine, are the major metabolic enzymes of tacrolimus [8,9]. The rs776746 polymorphisms in intron 3 of CYP3A5 have been correlated with altered gene expression due to a splicing defect. These CYP3A5 rs776746 GG genotype carriers are associated with slow tacrolimus metabolism [10,11]. However, the effect by CYP3A5 rs776746 still does not completely explain individual differences in tacrolimus metabolism [12,13].
IL6 has been demonstrated to cause a pan-suppression of mRNA of ten major CYP enzymes in human hepatocyte cultures [14]. IL6 could promote hepatic survival by stimulating liver regeneration and providing hepatoprotection in various models of liver injury [15], so it may be relevant to liver function after liver transplantation. The rs1800796 locus, which is in the IL6 gene promoter region, could regulate cytokine production and has been proved to be a functional SNP [16,17].

The aim of the study was to investigate the relationship between tacrolimus metabolism and IL6 rs1800796 in a large liver transplant cohort to evaluate the possibility of individualizing tacrolimus treatment in the early postoperative period of liver transplantation.

**Materials and Methods**

**Patients**

A total of 96 patients (16 female and 80 male) who underwent liver transplantation at Shanghai Jiao Tong University Affiliated First People’s Hospital between July 2007 and February 2011 were enrolled in this study. One patient was excluded from further analysis because the genotyping failed. The patients were all Han Chinese. The average age of the patients was 47.8 ± 8.3 years, and the average weight was 63.0 ± 10.4 kg. All of the patients received tacrolimus-based immunosuppressive regimens. Tacrolimus (Prograf, Astellas Pharma, Japan) was administered orally twice daily at an initial dose of 0.06 mg/kg/d. The dose of tacrolimus was adjusted according to the target blood concentration of 7 to 10 ng/ml during the first month after transplantation.

**Ethics Statement**

This research was approved by the Ethics Committee of Shanghai Jiao Tong University, and informed written consent was obtained according to the Declaration of Helsinki and its amendments.

**Data Collection**

Blood samples were collected half an hour before tacrolimus was administered, and trough concentrations (ng/ml) were then detected by PRO-TracTM II Tacrolimus ELISA kit (Diasorin, USA) with microparticle enzyme immunoassay (ELx800NB analyzer, BioTek, USA). C/D ratio was calculated with trough concentration and weight standardized 24-hour tacrolimus dose (mg/kg/d). We calculated C/D ratios at four weeks after transplantation, and we used the median of C/D ratios at weeks 1 to 4 to measure weekly changes in tacrolimus metabolism. We also used the median of glutamic-pyruvic transaminase levels (U/L) at weeks 1 to 4 after transplantation to measure weekly liver function.

**Extracting genomic DNA and genotyping**

Using an AllPrep DNA/RNA Mini Kit (Qiagen, Germany) according to the manufacturer’s instructions, genomic DNA was isolated from both donor and recipient liver tissue, where were previously stored at −80°C. Genotyping of SNPs was conducted by the Sequenom MassARRAY SNP genotyping platform (Sequenom, USA) [18]. The protocols included DNA and primer preparation, PCR amplification, SAP treatment, primer extension, resin cleanup, spotting primer extension products on SpectroCHIP, and detection primer extension products by mass spectrometer.

**Statistical Analysis**

Hardy-Weinberg equilibrium and allele frequency were analyzed using PLINK v1.07 (http://pngu.mgh.harvard.edu/purcell/plink/). Quantitative data between two groups were compared using Mann-Whitney U tests, and among several groups by Kruskal-Wallis. Non-parametric tests were performed in SPSS v17.0 (SPSS, USA). Two-sided tests were used in all analysis, and P < 0.05 was considered statistically significant.

**Results**

**Gene polymorphisms**

For CYP3A5, rs776746 allele A (28.4%) was found to be the minor allele, while allele G (71.6%) was the major allele. For IL6, rs1800796 allele G (30.3%) was found to be the minor allele, while allele C (69.7%) was the major allele. Both SNP frequencies were in accordance with Hardy-Weinberg equilibrium (P > 0.05). There were no differences in allele frequencies of the two SNPs between donors and recipients. Genotype frequencies of the two SNPs are shown in Table 1.

**Associations between CYP3A5 rs776746, IL6 rs1800796 polymorphisms and tacrolimus C/D ratios**

The effects of donor CYP3A5 rs776746 and IL6 rs1800796 polymorphisms on tacrolimus C/D ratios at four weeks after transplantation are shown in Table 2. Tacrolimus C/D ratios of donor CYP3A5 rs776746 allele A carriers at weeks 1, 3, and 4 were 199.0, 88.7, and 85.6, respectively, while C/D ratios of non-carriers were 295.1, 121.2, and 148.8, respectively. These differences between donor CYP3A5 rs776746 allele A carriers and non-carriers were significant (P = 0.006, 0.028, 0.001, respectively). Tacrolimus C/D ratios of donor IL6 rs1800796 allele G carriers at weeks 2 and 3 were 132.3 and 127.4, respectively, while C/D ratios of non-carriers were 105.2 and 97.4, respectively, and the differences were significant (P = 0.032, 0.021, respectively). Thus, donor CYP3A5 rs776746 allele A and IL6 rs1800796 allele G carriers were associated with fast tacrolimus metabolism.

The effects of recipient CYP3A5 rs776746 and IL6 rs1800796 polymorphisms on tacrolimus C/D ratios at four weeks after transplantation are shown in Table 3. Tacrolimus C/D ratios of recipient CYP3A5 rs776746 allele A carriers at weeks 1, 2, 3, and 4 were 179.0, 100.6, 97.0, and 90.2, respectively, while C/D ratios of non-carriers were 290.5, 133.2, 123.7, and 136.5, respectively, and the differences were significant (P = 0.003, 0.018, 0.030, 0.017, respectively). There was no significant difference in tacrolimus C/D ratio between recipient IL6 rs1800796 allele G carriers and non-carriers.

**Table 1. Genotype frequencies of CYP3A5 rs776746 and IL6 rs1800796.**

| SNPs         | Genotypes | Frequency of | Frequency of |
|--------------|-----------|--------------|--------------|
|              | donors    | recipients   |              |
| rs776746     | GG        | 41(0.432)    | 40(0.448)    |
|              | GA+AA     | 54(0.568)    | 50(0.526)    |
| rs1800796    | CC        | 49(0.516)    | 45(0.474)    |
|              | CG+GG     | 40(0.432)    | 46(0.484)    |

*Ref* is for referenced genotype, which is constituted of major allele and major allele. *Mut* is for mutated genotypes, which are constituted of major allele and minor allele or minor allele and minor allele.

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**Table 2. Tacrolimus C/D ratios of donor CYP3A5 rs776746 and IL6 rs1800796 polymorphisms.**

| SNPs         | Carrier | Non-carrier | P-value |
|--------------|---------|-------------|---------|
| rs776746     | 199.0   | 295.1       | 0.006   |
|              | 88.7    | 121.2       | 0.028   |
|              | 85.6    | 148.8       | 0.001   |
| rs1800796    | 132.3   | 105.2       | 0.032   |
|              | 127.4   | 97.4        | 0.021   |

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**Table 3. Tacrolimus C/D ratios of recipient CYP3A5 rs776746 and IL6 rs1800796 polymorphisms.**

| SNPs         | Carrier | Non-carrier | P-value |
|--------------|---------|-------------|---------|
| rs776746     | 179.0   | 290.5       | 0.003   |
|              | 100.6   | 133.2       | 0.018   |
|              | 97.0    | 123.7       | 0.030   |
|              | 90.2    | 136.5       | 0.017   |
carriers and non-carriers. Thus, recipient CYP3A5 rs776746 allele A is also associated with fast tacrolimus metabolism.

**Associations between combined polymorphisms and tacrolimus C/D ratios**

Donor CYP3A5 rs776746 allele A, IL6 rs1800796 allele C, and recipient CYP3A5 rs776746 allele A were shown to be associated with fast tacrolimus metabolism as stated above. Therefore, these three alleles were further investigated in a combination analysis. The associations between the number of alleles associated with fast metabolism and tacrolimus C/D ratios are shown in Table 4. With increasing numbers of alleles associated with fast metabolism, patients were found to have increasingly lower tacrolimus C/D ratios at all time points through the four weeks (P=0.001, 0.001, <0.001, <0.001, respectively).

**Associations between donor IL6 rs1800796 polymorphisms and GPT**

The effects of donor IL6 rs1800796 polymorphisms on GPT at four weeks after transplantation were studied. GPT of donor IL6 rs1800796 allele G carriers at weeks 1, 2, 3, and 4 were 167.0, 45.5, 30.0, and 34.5, respectively, while GPT of non-carriers were 142.5, 29.0, 20.0, and 20.0, respectively. The differences between donor IL6 rs1800796 allele G carriers and non-carriers at weeks 2, 3, and 4 were significant (P=0.004, 0.002, 0.006, respectively), but not at week 1 (P=0.141).

**Discussion**

The CYP3A enzymes including three functional enzymes as CYP3A4, CYP3A5, and CYP3A7 are responsible for the oxidative metabolism of over 50% of the drugs in widespread use [19]. CYP3A4 is responsible for most CYP3A-mediated drug metabolism [20]. There have been previous studies on the associations between CYP3A4 gene polymorphisms and tacrolimus metabolism [21–23], but there are no definitive conclusions comparing CYP3A4 gene polymorphisms and tacrolimus metabolism [24]. Our study is the first to demonstrate that both donor and recipient CYP3A5 gene polymorphisms in liver transplant patients [10,11]. Our study has verified that both donor and recipient CYP3A5 rs776746 were associated with tacrolimus metabolism in a transplant population.

It has been shown previously that CYP3A5 rs776746 could predict tacrolimus metabolism to a certain extent, but could not completely explain the individual differences [12,13]. Presently, about two hundred cytokines produced by many cell types have been found, and they usually act by autocrine and paracrine mechanisms [25]. In recent years, more attention has been paid to relationships between cytokines and tacrolimus metabolism. There have been reports that donor and recipient IL10 gene polymorphisms were associated with tacrolimus metabolism and IL10 was thought to reduce tacrolimus C/D ratios through up regulation of P-glycoprotein [26,27]. Our study is the first to demonstrate that donor IL6 rs1800796 polymorphisms are associated with tacrolimus metabolism.

Previous studies have shown that cytokines affect pharmacokinetic and pharmacodynamic behaviors of drugs [25,26]. IL6 has been shown to suppress mRNA of CYP3A5 in human hepatocyte cultures [14]. Cytokines also play a central role in immunologic events that occur after transplantation [29]. The liver often undergoes ischemia/reperfusion and immune injury during the transplantation and postoperative time. In this regard, IL6 activates the STAT3 signaling pathway through the gp130-IL6R complex on hepatocytes, and this process promotes liver

**Table 2.** The effects of donor CYP3A5 rs776746 and IL6 rs1800796 polymorphisms on tacrolimus C/D ratios.

| SNPs Genotypes | Week 1 | Week 2 | Week 3 | Week 4 |
|----------------|--------|--------|--------|--------|
|                | N      | C/D ratio | P       | C/D ratio | P       | C/D ratio | P       | C/D ratio | P       |
| rs776746 GG    | 46     | 295.1±413.2 | 0.006   | 130.8±130.9 | 0.086   | 121.2±81.1 | 0.028   | 148.8±126.9 | 0.001   |
| rs776746 GA+AA | 49     | 199.0±209.8 |         | 102.8±101.1 |         | 88.7±102.5 |         | 85.6±75.5    |         |
| rs1800796 CC   | 41     | 221.2±275.0 | 0.314   | 105.2±91.1  | 0.032   | 97.4±76.1  | 0.021   | 95.1±83.7    | 0.113   |
| rs1800796 GA+GG| 54     | 240.9±318.7 |         | 132.3±119.0 |         | 127.4±125.3|         | 133.2±119.3 |         |

**Table 3.** The effects of recipient CYP3A5 rs776746 and IL6 rs1800796 polymorphisms on tacrolimus C/D ratios.

| SNPs Genotypes | Week 1 | Week 2 | Week 3 | Week 4 |
|----------------|--------|--------|--------|--------|
|                | N      | C/D ratio | P       | C/D ratio | P       | C/D ratio | P       | C/D ratio | P       |
| rs776746 GG    | 50     | 290.5±283.4 | 0.003   | 133.2±107.8 | 0.018   | 123.7±113.0 | 0.030   | 136.5±114.0 | 0.017   |
| rs776746 GA+AA | 45     | 179.0±237.5 |         | 100.6±105.7 |         | 97.0±82.9  |         | 90.2±83.0    |         |
| rs1800796 CC   | 49     | 282.6±338.3 | 0.217   | 133.3±142.5 | 0.344   | 110.6±110.8 | 0.604   | 133.1±144.4 | 0.135   |
| rs1800796 GA+GG| 46     | 208.7±260.8 |         | 115.0±64.7  |         | 109.6±90.4  |         | 95.7±94.0    |         |

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regeneration, the acute-phase response, and hepatoprotection against Fas and toxic damage in liver [15]. Cytokine production may differ among different people due to gene polymorphisms [30,31]. The rs1800796 locus in IL6 promoter, which can regulate its production, was shown to be a functional SNP [16,17]. The C to G variation at IL6 rs1800796 has been shown to decrease transcriptional activity of the IL6 promoter, and carriers of CC genotype are high-expressors of IL6 [16,17,32]. Thus, we speculate that carriers of CC genotype have higher levels of IL6, leading to increased suppression of CYP3A5 expression and hepatoprotection. The current study showed that carriers of donor IL6 rs1800796 CC genotype were associated with faster recovery of liver function after transplantation, which is consistent with the fact that IL6 is a key molecule for liver regeneration and repair [33]. Suppression of CYP3A5 expression and hepatoprotection have opposite effects on tacrolimus metabolism. In the current study, carriers of donor IL6 rs1800796 CC genotype had a lower tacrolimus C/D ratios compared with non-carriers, indicating that the hepatoprotection effect of IL6 was greater than the suppression effect on CYP3A5 in liver transplant patients in the early postoperative period. The ability of donor IL6 rs1800796 to predict tacrolimus metabolism was less than CYP3A5 rs776746. The reason for this observation may be the opposing effects on tacrolimus metabolism of IL6 which could suppress CYP3A5 expression or provide hepatoprotection.

The current study demonstrated that donor IL6 gene polymorphisms were associated with tacrolimus metabolism, not the recipient. Donor IL10 gene polymorphisms have been reported to be associated with tacrolimus metabolism [8]. Expression of IFNG and IL10 has been found to be up regulated in hepatocytes from allograft tissue after orthotopic liver transplantation [34]. There is evidence that hepatocytes can express IL6 in the HBV-infected liver microenvironment [35]. The liver has the unique capacity to regulate its growth and mass after liver injury [36]. The current data are consistent with the concept that hepatocyte secretion of IL6 may facilitate liver repair and protection after transplantation. Li et al [27] have found that serum IL18 levels were associated with tacrolimus C/D ratios and that recipient IL1B gene polymorphisms were not associated with tacrolimus C/D ratios and serum IL18 levels. We suspect that donor IL1B gene polymorphisms may be associated with tacrolimus metabolism as has been shown for IL6 in the current study.

At present, genotyping of CYP3A5 rs776746 polymorphisms has been used in clinical applications, however, it could not predict tacrolimus metabolism accurately. Our study showed that genotyping of donor IL6 rs1800796 polymorphisms could assist CYP3A5 rs776746 to predict tacrolimus metabolism more effectively. Various populations in other regions of the world may differ in certain features of genetic architecture. Whether IL6 rs1800796 is applicable to other populations with regards to tacrolimus metabolism requires further investigation. The gene-drug observations in the current study suggest that genetic factors may also affect liver transplant patient outcomes including graft and recipient survival rates. Proof of such associations requires further study.

**Conclusions**

Donor gene polymorphisms of IL6 play a more important role than those of recipient. Combined polymorphisms of donor CYP3A5 rs776746, IL6 rs1800796, and recipient CYP3A5 rs776746 have a greater effect on tacrolimus metabolism than CYP3A5 rs776746 alone. IL6 levels may lead to individual differences in tacrolimus metabolism mainly by affecting liver function.

**Author Contributions**

Conceived and designed the experiments: DC JF ZW ZP. Performed the experiments: FG SQ. Analyzed the data: DC JF. Contributed reagents/materials/analysis tools: SQ. Wrote the paper: DC.

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