High pretransplant HBV level predicts HBV reactivation after kidney transplantation in HBV infected recipients

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INTRODUCTION

HBV infection is more prevalent among patients in dialysis and in renal transplant recipients than in the general population. In Korea, the positivity rate for HBsAg among people aged 10 years or older was 3.7% of the total population being infected with HBV [1].

Pre-existing HBV infection is considered a relative contraindication to kidney transplantation. The use of immunosuppressive drugs enhances viral replication and may reactivate HBV, and this may accelerate liver injury and progression to liver failure [2]. HBV reactivation is a major risk factor for morbidity and mortality in HBsAg-positive renal transplant recipients. Moreover, HBV-associated glomerulonephritis may recur or develop de novo in the graft, reducing function and possibly resulting in graft failure [3].

Past studies established that HBsAg-positive kidney recipients are at increased risk for mortality and graft failure compared with HBsAg-negative recipients [4,5]. In the last decade, however, introduction of effective oral antiviral therapies for HBV
and judicious use of immunosuppressive drugs in recipients with viral hepatitis have altered management of HBV-infected recipients [6].

In this study, we aimed to identify the outcomes of HBsAg-positive recipients who received preemptive antiviral agents after successful kidney transplantation and to analyze the risk factors for HBV reactivation.

**METHODS**

**Patients**

Lamivudine has been used at Samsung Medical Center to treat HBsAg-positive renal transplant recipients since 1999. Data for 944 patients who underwent kidney transplantation between September 1999 and September 2010 were retrieved from the electronic medical recording system and reviewed. Forty-two patients (4.4%) were categorized as HBV-infected when seropositivity for hepatitis B surface antigen was recorded on the transplant registration form. Children (<18 years old), retransplantation patients, foreigner, recipients of multiorgan transplants, and those with pretransplantation HCV infection were excluded. Data were collected for gender, age, cause of end-stage renal disease, type and duration of dialysis, HBV DNA titer before transplantation, type of transplantation, human leukocyte antigen (HLA) mismatches, induction immunosuppression and types of calcineurin inhibitor and antiviral agents used. HBV DNA was quantified using the COBAS TaqMan HBV test (detection limit 12 IU/mL Roche Molecular Systems, Branchburg, NJ, USA). HBV DNA titers in sera were measured using a real-time PCR assay. Recipients who had taken antiviral agents before the transplantation continued with these medicines after the operation without change. Those for whom antiviral agents were not prescribed before the transplantation began lamivudine (100 mg/day) preemptively after the surgery.

All rejection episodes were confirmed by percutaneous renal biopsy. Delayed graft function (DGF) was defined by the need for hemodialysis in the first week posttransplant. Bacterial infections were diagnosed by culture results. Viral infections were diagnosed by culture results, rapid antigen detection tests, changes in serology, PCR, and cerebral spinal fluid abnormalities. Graft loss was defined as a return to dialysis, a graft nephrectomy, or retransplantation.

HBV reactivation was defined as a serum ALT concentration two times greater than the upper limit of normal, accompanied by a new detection of HBV DNA without any other cause of hepatic dysfunction. Detection of the YMDD mutation was defined as lamivudine resistance, and adefovir 10 mg/day was then added to lamivudine or patients were switched to entecavir 0.5 mg/day after consultation with a hepatologist. Survival and clinical data recorded at the time of patient death, kidney transplantation and last clinic visit up to July 2011 were assessed. This study complies with the standards of Declaration of Helsinki and current ethical guidelines.

**Immunosuppression**

All recipients received quadruple therapy: induction agents (ATGAM or Thymoglobulin), steroids (methylprednisolone), mycophenolate mofetil, and cyclosporine or tacrolimus. ATGAM was given at 15 mg/kg per dose for 5–7 doses, and Thymoglobulin was given at 1.5 mg/kg per dose for 5–7 doses. All doses of ATGAM and thymoglobulin were given postoperatively. Steroids were started at 500 mg per day intravenously, then tapered to 16 mg per day at 1 month posttransplantation, then to 8 mg per day at 3 months. Mycophenolate mofetil was started 1.5 g per day orally on the day of the transplantation. Cyclosporin or tacrolimus was started on posttransplant day 3.

**Anti-infectious prophylaxis**

Trimethoprim-sulfamethoxazole was administered for primary prophylaxis of *Pneumocystis jiroveci* pneumonia for at least 1 year after kidney transplantation. Other prophylactic antibiotics and antifungal drugs were not routinely administered because of the concern about emergence of resistant pathogens. All recipients received intravenous ganciclovir for 7 days. The dose of intravenous ganciclovir depended on the status of renal function in patients. They were monitored monthly for cytomegalovirus (CMV) infection by CMV antigenemia assay. If the CMV antigenemia assay showed more than 50/400,000 leukocytes, recipients were preemptively treated with intravenous ganciclovir until the CMV antigenemia assay became negative.

**Statistical analysis**

All statistical analyses were performed with IBM SPSS ver. 19.0 (IBM Co., Armonk, NY, USA). Continuous variables were reported as median and range or mean ± standard deviation. Cutoff value in HBV DNA was calculated by receiver operating characteristics (ROC) curve. Continuous variables were compared using Mann-Whitney U tests and categorical variables were compared using Fisher exact test. HBV reactivation-free survival was analyzed using the Kaplan-Meier method and backward in Cox regression analysis was used to determine factors related to the development of HBV reactivation. A P-value less than 0.05 was considered statistically significant.

**RESULTS**

**Clinical characteristics for HBsAg-positive recipients**

The clinical characteristics of kidney transplant recipients were summarized Table 1. Forty-two patients, including 28 men and 14 women, with a median age of 43 years (23–60 years),
fulfilled the inclusion criteria. Median follow-up duration for these 42 recipients was 52.5 months (8–136 months). The underlying kidney disease was chronic glomerulonephritis in 9 patients (21.4%), diabetes in 7 patients (16.7%), hypertension in 5 patients (11.9%), IgA nephropathy in 4 patients (9.5%), unknown in 15 patients (35.7%), and others in 2 patients (4.8%). Most patients (n = 29, 69%) received hemodialysis over a median time span of 11 months (1–188 months). The mean HBV-DNA level in these patients prior to transplantation was $1.58 \times 10^7$ IU/mL. Twenty-one patients (50%) received kidney allografts from a living donor and the other patients, from deceased donors. However, the proportion of living donor kidney transplantation (LDKT) in HBsAg-negative recipients was higher than in HBsAg-positive (P = 0.001).

### Graft function and graft survival

The incidence of DGF in HBsAg-positive recipients was higher than in HBsAg-negative recipients, but there was no statistically significant difference between the two groups (4.8% in HBsAg-positive vs. 1.9% in HBsAg-negative, P = 0.205). The serum creatinine in HBsAg-positive recipients was higher than in HBsAg-negative at 2 weeks (P = 0.010) because the proportion

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**Table 1. Clinical characteristics of kidney transplant recipients with and without HBsAg**

| Characteristic                              | HBsAg positivity | P-value |
|--------------------------------------------|------------------|---------|
|                                            | No (n = 902)     | Yes (n = 42) |
| Gender                                     |                  |         |
| Male                                       | 504 (55.9)       | 28 (66.7) |
| Female                                     | 398 (44.1)       | 14 (33.3) |
| Age (yr)                                   | 41.9 ± 10.7      | 43.3 ± 9.2 |
| Causes of end-stage renal disease          |                  |         |
| Unknown<sup>a</sup>                        | 336 (37.3)       | 15 (35.7) |
| Diabetes                                   | 120 (13.3)       | 7 (16.7) |
| Glomerulonephritis                         | 121 (13.4)       | 9 (21.4) |
| IgA nephropathy                            | 99 (11.0)        | 4 (9.5) |
| Hypertension                               | 146 (16.3)       | 5 (11.9) |
| Others<sup>b</sup>                         | 80 (8.9)         | 2 (4.8) |
| Type of dialysis                           |                  |         |
| Hemodialysis                               | 623 (69.1)       | 29 (69.0) |
| Continuous ambulatory peritoneal dialysis   | 156 (17.3)       | 8 (19.0) |
| None                                       | 122 (13.5)       | 5 (11.9) |
| Duration of dialysis (mo)                  | 27.1 ± 35.2      | 27.2 ± 37.8 |
| Panel reactive antibody                    |                  |         |
| Class I (≥30%)                             | 29 (3.2)         | 2 (4.8) |
| Class II (≥30%)                            | 5 (0.6)          | 0 (0) |
| HLA mismatch ≥4                            | 661 (73.4)       | 36 (85.7) |
| CMV status                                 |                  |         |
| CMV D–/R–                                  | 1 (0.1)          | 0 (0) |
| CMV D+/R+                                  | 854 (94.7)       | 38 (90.5) |
| CMV D+/R–                                  | 21 (2.3)         | 0 (0) |
| CMV D–/R+                                  | 26 (2.9)         | 4 (9.5) |
| Type of transplantation                    |                  |         |
| LDKT                                       | 678 (75.2)       | 21 (50.0) |
| DDKT                                       | 224 (24.8)       | 21 (50.0) |
| Calcineurin inhibitor                      |                  |         |
| Cyclosporin                                | 456 (51.9)       | 24 (57.1) |
| Tacrolimus                                 | 422 (48.1)       | 18 (42.9) |
| Follow-up duration (mo)                    | 64.4 ± 38.6      | 53.7 ± 34.6 |

Values are presented as number (%) or mean ± standard deviation. HLA, human leukocyte antigen; CMV, cytomegalovirus; D, donor; R, recipient; LDKT, living donor kidney transplantation; DDKT, deceased donor kidney transplantation.

<sup>a</sup>Unknown indicates those with unidentified causes of end-stage renal disease in pretransplant. <sup>b</sup>Others included polycystic kidney disease, systemic lupus erythematosus, purpura, and etc.
of deceased donor kidney transplantation was higher than that in LDKT in HBsAg-positive recipients. We found no significant difference in the median serum creatinine levels at 1-, 3-, 6-, 9-, or 12-month posttransplant between the two groups ($P = \text{not significant}$) (Fig. 1). The 1 year acute rejection rates in HBsAg-positive recipients and HBsAg-negative recipients was 38.8% and 28.0%, respectively, but there was no statistically significant difference between the two groups ($P = 0.197$). Infection complications in 1-year posttransplant are shown in Table 2. However, the difference in the incidence of infections between the two groups did not achieve statistical significance.

We compared the death-censored graft survival rates between HBsAg-positive recipients and HBsAg-negative recipients. The 1-, 3-, and 5-year death-censored graft survival rates of HBsAg-negative recipients were 98.2%, 96.7%, and 94.3%, respectively, and those graft survival rates of HBsAg-positive recipients were 100%, 100%, and 94.7%, respectively. There was no statistical difference between the two groups ($P = 0.398$) (Fig. 2). Only one of the 42 HBsAg-positive patients experienced graft rejection and resumed dialysis. Graft loss was due to chronic rejection.

### Outcomes of posttransplant antiviral therapy

Most patients ($n = 35$, 85.3%) received lamivudine after kidney transplantation and the other patients received adefovir ($n = 1$) or entecavir ($n = 6$); these antiviral agents were started in patients with detectable HBV DNA or as preemptive therapy in HBsAg-positive dialysis patients (Fig. 3). During antiviral treatment, most of the patients ($n = 35$, 83.3%) were considered to have stable liver status and 7 patients had potential deterioration due to HBV reactivation and YMDD mutation. The 1-, 3-, and 5-year HBV reactivation-free survival rates were 95.0%, 82.6%, and 78.4%, respectively. Alternative or add-on therapies such as adefovir ($n = 3$) or entecavir ($n = 4$) were administered when HBV DNA titers rose and lamivudine resistance were detected. Five patients who received lamivudine showed

![Fig. 1. Serum creatinine levels after transplantation. The serum creatinine in HBsAg-positive recipients was higher than in HBsAg-negative at 2 weeks ($P = 0.010$) because the proportion of deceased donor kidney transplantation was higher than that in living donor kidney transplantation in HBsAg-positive recipients. We found no significant difference in the median serum creatinine levels at 1-, 3-, 6-, 9-, or 12-month posttransplant between the two groups ($P = \text{not significant}$).](image1)

![Fig. 2. Death-censored graft survival between HBsAg-positive recipients and HBsAg-negative recipients.](image2)

![Fig. 3. Schematization of antiviral prophylaxis.](image3)

### Table 2. Infection complications during the 1-year posttransplant period

| Variable               | HBsAg positivity | P-value |
|------------------------|------------------|---------|
|                        | No ($n = 902$)   | Yes ($n = 42$) |
| Herpes zoster          | 62 (6.9)         | 5 (11.9) | 0.213 |
| Bacterial infection    | 40 (4.4)         | 0 (0)    | 0.252 |
| Tuberculosis           | 9 (1.0)          | 1 (2.4)  | 0.367 |
| Fungal infection       | 11 (1.2)         | 0 (0)    | 0.472 |
| Parvovirus infection   | 145 (16.1)       | 7 (16.7) | 0.832 |
| BK virus infection     | 188 (20.8)       | 14 (33.3) | 0.080 |
| Urinary tract infection| 30 (3.3)         | 2 (4.8)  | 0.649 |
| CMV infection          | 323 (35.8)       | 16 (38.1) | 0.745 |

Values are presented as number (%). CMV, cytomegalovirus.
lamivudine resistance, but liver functions were stable. Adefovir was used as a combination therapy with lamivudine in 3 patients and entecavir was replaced of lamivudine in 2 patients. The liver functions of patients who have HBV reactivation or YMDD mutation were well controlled by lamivudine and adefovir combination or entecavir switching.

All patients were under treatment with antiviral agents at the time of last assessment. Among these patients, only one was taking adefovir and lamivudine combination therapy and the others were on monotherapy. In the monotherapy group, 23 patients were treated with lamivudine, 6 with adefovir, and 12 with entecavir. At the end of follow-up, HBV reactivation was well controlled by switch or combination therapy in all patients with the disease. All patients were alive at the end of follow-up and none developed end-stage liver disease or hepatocellular carcinoma.

### Comparison of kidney recipients with and without HBV reactivation

Baseline demographic and clinical characteristics of the two

| Variable                                | HBV reactivation | P-value |
|------------------------------------------|------------------|---------|
|                                          | No (n = 35)      | Yes (n = 7) |       |
| Gender                                   |                  |          |
| Male                                     | 25 (71.4)        | 3 (42.9)  | 0.197  |
| Female                                   | 10 (28.6)        | 4 (57.1)  |         |
| Age (yr)                                 | 43.9 ± 8.4       | 40.6 ± 13.0| 0.417  |
| Causes of end-stage renal disease        |                  |          |
| Unknown                                  | 12 (34.3)        | 3 (42.9)  | 0.639  |
| Diabetes                                 | 7 (20.0)         | 0 (0)     |         |
| Glomerulonephritis                       | 8 (22.9)         | 1 (14.3)  |         |
| IgA nephropathy                          | 3 (8.6)          | 1 (14.3)  |         |
| Hypertension                             | 3 (8.6)          | 2 (28.6)  |         |
| Others                                   | 2 (5.8)          | 0 (0)     |         |
| Type of dialysis                         |                  |          |
| Hemodialysis                             | 23 (65.7)        | 6 (85.7)  | 0.372  |
| Continuous ambulatory peritoneal dialysis| 8 (22.9)         | 0 (0)     |         |
| None                                     | 4 (11.4)         | 1 (14.3)  |         |
| Duration of dialysis, months             | 27.9 ± 39.1      | 22.8 ± 32.1| 0.559  |
| HBV DNA before transplantation ≥ 5 × 10\(^{4}\) IU/mL | 5 (14.3) | 5 (71.4)  | 0.005  |
| HBeAg-positive before transplantation    | 8 (22.9)         | 4 (57.1)  | 0.088  |
| Type of transplantation                  | 0.679            |           |
| LDKT                                     | 18 (51.4)        | 3 (42.9)  |         |
| DDKT                                     | 17 (48.6)        | 4 (57.1)  |         |
| HLA mismatch > 3                         | 29 (82.9)        | 7 (100)   | 0.567  |
| Induction immunosuppression              |                  |          |
| None                                     | 15 (42.9)        | 3 (42.9)  | 1.000  |
| Basiliximab                              | 14 (40.0)        | 2 (28.6)  | 0.570  |
| Antithymocyte globulin                   | 6 (17.1)         | 2 (28.6)  | 0.482  |
| Calcineurin inhibitor                    | 1.000            |           |
| Cyclosporin                              | 20 (57.1)        | 4 (57.1)  |         |
| Tacrolimus                               | 15 (42.9)        | 3 (42.9)  |         |
| Antiviral agents                         | 0.432            |           |
| Adefovir                                 | 1 (2.9)          | 0 (0)     |         |
| Entecavir                                | 6 (17.1)         | 0 (0)     |         |
| Lamivudine                               | 28 (80.0)        | 7 (100)   |         |
| Follow-up duration (mo)                  | 52.3 ± 35.0      | 60.9 ± 34.0| 0.578  |

Values are presented as number (%) or mean ± standard deviation.

LDKT, living donor kidney transplantation; DDKT, deceased donor kidney transplantation; HLA, human leukocyte antigen.

\(^{a}\)Unknown indicates those with unidentified causes of end-stage renal disease in pretransplant. \(^{b}\)Others included polycystic kidney disease, systemic lupus erythematosus, purpura, and etc.
groups are shown in Table 3. ROC curve showed that the cutoff value of HBV DNA was $5 \times 10^4$ IU/mL ($R^2 = 77.1\%$ and $P = 0.025$, respectively) (Fig. 4). The HBV DNA levels before transplantation in patients who experienced HBV reactivation were higher than in those who did not have HBV reactivation (median, 12 IU/mL; range, 11–4.25 $\times 10^6$ IU/mL vs. median, $8.32 \times 10^7$ IU/mL; range, 12–2.92 $\times 10^8$ IU/mL; $P = 0.023$) (Fig. 5). All patients with HBV reactivation had received lamivudine after transplantation, but the relative effectiveness of lamivudine (as compared to other drugs) on DNA levels or reactivation risk did not show statistical significance because only small numbers of patients received adefovir or entecavir. Multivariate analysis revealed that increased pretransplant HBV DNA levels ($>5 \times 10^4$ IU/mL) were closely associated with HBV reactivation in HBsAg-positive recipients after kidney transplantation (hazard ratio, 1.559; 95% confidence Interval, 1.126–2.158; $P = 0.007$).

**DISCUSSION**

One important finding in our study was that treatment with antiviral drugs such as lamivudine, adefovir, and entecavir may suppress to nearby undetectable levels the viral activity in HBsAg-positive kidney transplant recipients. Among HBsAg-positive patients who did not receive preemptive treatments, the 10-year graft survival was significantly lower than in HBsAg-negative recipients [7]. However, HBsAg-positive recipients treated with lamivudine showed significantly higher 10-year graft and patient survival than HBsAg-positive recipients who did not receive antiviral treatments [8].

As compared to patients in previous series, the HBsAg-positive kidney transplant recipients in this study showed better patient and allograft survival [9,10]. In addition, there was no statistical difference in graft survival between HBsAg-positive recipients and HBsAg-negative recipients. At the end of follow-up, all patients were alive and only one patient experienced graft failure. We attribute these favorable outcomes to suppression of viral activities through antiviral treatments.

The use of immunosuppressive drugs enhances viral replication, which may accelerate liver injury and progression to liver failure in HBsAg-positive kidney transplant recipients [2]. The HBV genome contains a glucocorticoid-sensitive receptor. Glucocorticoid drugs increase activity at this receptor binding site and promote HBV transcription [11] followed by chronic HBV reactivation [12]. Our study revealed no deaths related to liver failure or portal hypertension; and this underscores the importance of sustained viral suppression in maintaining hepatic function and preventing acute-on-chronic hepatic failure even with concurrent immunosuppressive treatment.

Anti-HBV treatments, including pegylated interferon and antiviral agents such as lamivudine, adefovir, entecavir, or tenofovir, have changed the prognosis for HBV infection. Interferon cannot be used in renal transplant recipients because of its low effectiveness, direct nephrotoxicity and the associated risk of acute allograft rejection [13]. The first-generation antiviral agents lamivudine and adefovir effectively suppress HBV. Preemptive or prophylactic treatment of HBsAg-positive renal transplant recipients with lamivudine before hepatic dysfunction provides favorable short-term outcomes, including reduction in viral load, improvement in liver chemistry and prevention of irreversible histologic deterioration [2]. Prolonged use of lamivudine, however, may result in drug resistance. The most common mutation associated with lamivudine resistance involves the replacement of methionine by valine or isoleucine in the tyrosine-methionine-aspartate-aspartate (YMDD) motif of the HBV DNA polymerase (the “YMDD mutation”) [14]. Genotypic resistance can be detected in 14%–32% of immunocompetent patients with chronic hepatitis B after one year of lamivudine treatment [15].

![Fig. 4](image1.png)

**Fig. 4.** Receiver operating characteristics curve showed that the cutoff value of HBV DNA was $5 \times 10^4$ IU/mL ($R^2 = 77.1\%$ and $P = 0.025$, respectively).

![Fig. 5](image2.png)

**Fig. 5.** Difference of HBV DNA levels before transplantation between patients with HBV reactivation and without HBV reactivation.
Increased HBV DNA level (≥5 × 10^5 IU/mL) to suppress HBV DNA to undetectable levels. This may greatly reduce risk for HBV reactivation after transplantation.

The limitations of our study include a retrospective design, a very small number of HBsAg-positive recipients to provide data. But given the high liver-related morbidity and mortality associated with rapid viral replication in conjunction with immunosuppressive therapy, a randomized controlled study that included HBsAg-positive renal transplant recipients without antiviral treatment would be unethical. Thus we compared HBsAg-positive recipients with HBsAg-negative in our data.

In conclusion, the development and application of potent antiviral agents has significantly increased graft and patient survival among HBV-infected recipients, although liver failure still occurs more frequently in this group compared with uninfected patients. This study showed that effective viral suppression in HBsAg-positive renal transplant recipients using antiviral agents improves both patient and allograft survival.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

REFERENCES

1. Korean Association for the Study of the Liver. KASL Clinical Practice Guidelines: management of chronic hepatitis B. Clin Mol Hepatol 2012;18:109-62.
2. Kalia H, Fabrizi F, Martin P. Hepatitis B virus and renal transplantation. Transplant Rev (Orlando) 2011;25:102-9.
3. Ivanyi B. A primer on recurrent and de novo glomerulonephritis in renal allografts. Nat Clin Pract Nephrol 2008;4:446-57.
4. Aroldi A, Lampertico P, Montagnino G, Passerini P, Villa M, Campise MR, et al. Natural history of hepatitis B and C in renal allograft recipients. Transplantation 2005;79:1132-6.
5. Fabrizi F, Martin P, Dixit V, Kanwal F, Dulai G. HBsAg seropositive status and survival after renal transplantation: meta-analysis of observational studies. Am J Transplant 2005;5:2913-21.
6. Perrillo RP. Hepatitis B and renal transplantation: securing the sword of damocles. Hepatology 2002;36:1041-5.
7. Mathurin P, Mouquet C, Poynard T, Sylla C, Benalia H, Fretz C, et al. Impact of hepatitis B and C virus on kidney transplantation outcome. Hepatology 1999;29:257-63.
8. Ahn HJ, Kim MS, Kim YS, Kim SI, Huh KH, Ju MK, et al. Clinical outcome of renal transplantation in patients with positive pre-transplant hepatitis B surface antigen. J Med Virol 2007;79:1655-63.
9. Degos F, Lugassy C, Degott C, Debure A, Carnot F, Theis V, et al. Hepatitis B virus and hepatitis B-related viral infection in renal transplant recipients: a prospective study of 90 patients. Gastroenterology 1988;94:151-6.
10. Parfrey PS, Forbes RD, Hutchinson TA, Beaudoin JG, Dauphinie WD, Hollomby DJ, et al. The clinical and pathological course of hepatitis B liver disease in renal transplant recipients. Transplantation 1984;37:461-6.
11. Tur-Kaspa R, Shaul Y, Moore DD, Burk RD, Okret S, Poellinger L, et al. The glucocorticoid receptor recognizes a specific nucleotide sequence in hepatitis B virus DNA causing increased activity of the HBV enhancer. Virology 1988;167:650-3.
12. Cheng AL, Hsiung CA, Su IJ, Chen PJ, Chang MC, Tsao CJ, et al. Steroid-free chemotherapy decreases risk of hepatitis B virus (HBV) reactivation in HBV-carriers with lymphoma. Hepatology 2003;37:1320-8.
13. Theret E, Pol S, Legendre C, Gagnadoux MF, Cavalcanti R, Kreis H. Low-dose recombinant leukocyte interferon-alpha treatment of hepatitis C viral infection in renal transplant recipients: a pilot study. Transplantation 1994;58:625-8.
14. Allen MI, Deslauriers M, Andrews CW,
Tipples GA, Walters KA, Tyrrell DL, et al. Identification and characterization of mutations in hepatitis B virus resistant to lamivudine. Lamivudine Clinical Investigation Group. Hepatology 1998;27:1670-7.

15. Lai CL, Chien RN, Leung NW, Chang TT, Guan R, Tai DI, et al. A one-year trial of lamivudine for chronic hepatitis B. Asia Hepatitis Lamivudine Study Group. N Engl J Med 1998;339:61-8.

16. Lok AS, Lai CL, Leung N, Yao GB, Cui ZY, Schiff ER, et al. Long-term safety of lamivudine treatment in patients with chronic hepatitis B. Gastroenterology 2003;125:1714-22.

17. European Association for the Study of the Liver. EASL clinical practice guidelines. Management of chronic hepatitis B. Gastroenterol Clin Biol 2009;33:539-54.

18. Marzano A, Gaia S, Ghisetti V, Carenzi S, Premoli A, Debernardi-Venon W, et al. Viral load at the time of liver transplantation and risk of hepatitis B virus recurrence. Liver Transpl 2005;11:402-9.

19. Beckebaum S, Sotiropoulos GC, Klein CG, Broelsch CE, Saner F, Paul A, et al. Predictive factors of outcome in patients transplanted for hepatitis B. Transplantation 2009;87:872-81.