Review Article

The Relationship between Circulating ANGPTL8/Betatrophin Concentrations and Adult Obesity: A Meta-Analysis

Jingjing Ye, Yu Qin, Dong Wang, Ling Yang, and Guoyue Yuan

Department of Endocrinology, Affiliated Hospital of Jiangsu University, 438 Jiefang Road, Zhenjiang, Jiangsu 212001, China

Correspondence should be addressed to Guoyue Yuan; yuanguoyue1@126.com

Received 3 May 2019; Revised 2 September 2019; Accepted 17 September 2019; Published 22 October 2019

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In this study, we evaluated the relationship between circulating betatrophin levels and obesity. Obesity is a common public health problem that is increasing globally. Betatrophin, a newly identified protein, is predominantly expressed in white and brown fat tissues and in the liver. Growing evidence suggests that betatrophin plays a pivotal role in metabolism, including the synthesis and degradation of lipids in cells, and adipocyte differentiation. Previous studies have assessed the association between circulating betatrophin levels and obesity; however, this relationship remains unclear. Therefore, our study is aimed at examining the impact of betatrophin on obesity using a meta-analysis of the current evidence. We performed a meta-analysis to quantify the relationship between betatrophin levels and obesity. A literature search was conducted through the EMBASE, Web of Science, and MEDLINE databases. Retrieved studies were screened, without any language restrictions to identify relevant literature published up to December 2018. Observational studies, in which the association between circulating concentrations of betatrophin and obesity was evaluated, were considered suitable for the systematic review. Of the 65 manuscripts retrieved, 9 datasets from 6 studies, involving 681 participants, detected an association between circulating betatrophin and obesity. Circulating betatrophin levels of obese subjects were higher than those of nonobese subjects (random effects weighted mean difference (WMD) = 0.250 μg/mL, 95% CI: 0.048–0.451, \( I^2 = 94.8\% \), \( p = 0.015 \)), yet with significant between-study heterogeneity. This heterogeneity appeared to be modified by glycemic status but not by age, the ELISA kits used, sample source, or body mass index. The high circulating betatrophin concentration may directly increase the risk of obesity in adults. Betatrophin may serve as a therapeutic target for obesity in adults.

1. Introduction

Obesity causes considerable health and socioeconomic problems in many countries. Previous epidemiological studies have reported conflicting results regarding betatrophin levels and obesity, but the biological mechanisms underlying this relationship remain obscure.

Obesity is a common public health problem and is on the rise worldwide. This is apparent from the 2014 World Health Organization (WHO) report, which stated that over 0.6 billion people were obese, and more than 1.9 billion adults were overweight [1]. In recent years, obesity has become a significant burden on global healthcare expenditures. It is frequently associated with chronic disorders, including heart disease, polycystic ovary syndrome, breast cancer, and diabetes. Globally, obesity and overweight contribute to at least 2.8 million deaths each year [2].

Adipose tissue, recognized as an important endocrine organ, has attracted much attention due to its effect on few metabolic processes such as glucose homeostasis, lipid metabolism, inflammation, and blood pressure. Betatrophin, also known as “ANGPTL8,” “RIFL,” “C19ORF80,” “TD26,” and “lipasin,” is a novel adipokine secreted from the adipose tissue and the liver [3]. Recent studies have indicated that betatrophin is linked to glucolipid disorders [4]. In this case, circulating betatrophin levels might be affected by nutritional status that alters the body weight [5]. The study done by Ren et al. on mice demonstrated that the transcript level of angioptotin-like protein 8 (ANGPTL8) was higher for subjects with obesity [6]. However, the findings of multiple
epidemiological studies have not been entirely consistent. Several studies showed that circulating betatrophin levels were not significantly different in adults with obesity [7, 8]. By contrast, some studies demonstrated a positive relationship between circulating betatrophin levels and obesity [9–12], whereas other studies demonstrated an inverse relationship [13–15]. Hence, more reliable association between circulating betatrophin concentration and the risk of obesity is warranted.

Thus far, a meta-analysis has not been performed specifically to assess the association between circulating betatrophin concentration and the risk of obesity. Therefore, the aim of the present study was to systematically review all eligible studies that have assessed the association between betatrophin levels and the risk of obesity, thereby providing a quantitative estimation of the association.

2. Materials and Methods

2.1. Ethical Approval and Patient Consent. This study is a meta-analysis. Hence, approval by an ethics committee and informed patient consent were not required.

2.2. Data Sources. A comprehensive literature search was performed in the EMBASE, Web of Science, and MEDLINE databases up to December 2018, without any language restrictions. The search strategy included the following query terms: “ANGPTL8,” “betatrophin,” “RIFL,” “C19ORF80,” “TD26,” “lipasin,” “obesity,” and “obese.” Furthermore, the reference lists of the identified papers were reviewed for comprehensive search results. The study was conducted in accordance with the MOOSE group standards for reporting meta-analysis of observational studies. Titles, abstracts, and full-text papers were screened by 2 independent reviewers with expertise in conducting systematic reviews. Disagreements were resolved by consulting a 3rd reviewer, who was not involved in the initial procedure.

2.3. Study Selection

2.3.1. Inclusion and Exclusion Criteria. Observational studies were considered suitable for the systematic review, in which the association between circulating concentrations of betatrophin and obesity was evaluated. If conference abstracts included sufficient data to extract effect estimates, they were also included. The exclusion criteria were as follows: (1) review paper, (2) search not restricted to humans, and (3) studies lacking a nonobese control group. For overlapping publications, the most complete data were included.

2.3.2. Data Extraction. Data from each eligible study were independently collected by two researchers, using a predesigned data extraction form. Any disagreements were checked by a third investigator. General characteristics of each study were extracted: year of publication, the first author’s last name, the mean age and body mass index (BMI), country, cases and control or cohort size, glycemic status, and betatrophin measurement method. The quality of each study was evaluated using elements of the Newcastle-Ottawa Scale (NOS) [16]. In addition, when data were missing or when there was doubt regarding information in the publications, we communicated with the authors to obtain further information.

2.3.3. Statistical Analysis. The data on betatrophin levels in each study were adopted to mean difference ± standard deviation (SD). Standard error of the mean was converted to SD. The weighted mean differences (WMDs) of total betatrophin levels were calculated for all the eligible studies in the meta-analysis and were used in fixed-effects or random-effects models. The $\chi^2$ test and $I^2$ test evaluated heterogeneity in the results of the studies ($p < 0.1$ and $I^2 > 50\%$ was considered to indicate a statistically significant heterogeneity) [17]. Visual inspection of publication bias by a funnel plot of the data was not applied, as it may be interpreted incorrectly if studies were heterogeneous [18, 19]. Begg’s funnel plot and Egger’s regression asymmetry test was used to estimate the publication bias [20]. Moreover, sensitivity analysis was undertaken with the exclusion of the studies with borderline eligibility. Subgroup analyses were performed by geographic region, sample source, the studies employing different betatrophin ELISA kits, and the studies with different mean age, BMI, and glycemic status. The statistical analysis and meta-analysis were performed with Stata 11.0 (Stata, College Station, TX, USA). The present study was conducted in accordance with the MOOSE guidelines for meta-analysis [21].

3. Results

3.1. Search Results. A PRISMA flow chart of the search procedure is presented in Figure 1. The search strategy identified 65 potentially relevant papers, with 59 being excluded after deduplication and title/abstract screening. After further analysis of the identified papers, we finally included 9 datasets from 6 studies (2 papers reported results for 5 separate studies) [7, 8, 10–12, 14]. Table 1 presents the main characteristics of the 9 studies. Only circulating full-length betatrophin concentration was measured in these studies.

3.2. Main Analysis. The meta-analysis included 9 datasets from 6 studies [7, 8, 10–12, 14]. Overall, 307 healthy controls and 374 patients with obesity were included in the meta-analysis. Cochran’s Q test was used to assess the heterogeneity among the 9 studies. Since significant heterogeneity was found between the studies ($Q = 154.39$, d.f. = 8, $p < 0.05$), the random-effects model was adopted. All of the studies were included in the random-effects model. We observed that circulating betatrophin levels of obese subjects were higher than that of nonobese subjects (random-effects WMD = 0.25 pg/mL, 95% CI: 0.048–0.451, $I^2 = 94.8\%$, $p = 0.015$). Figure 2 shows the WMD and 95% CIs for each individual study.

Since there was a high level of heterogeneity among the included studies, we subsequently performed subgroup analyses by all characteristics to assess whether they can explore the potential source of heterogeneity between circulating betatrophin levels and obesity. Subgroup analyses indicated that potential variables such as age, different sample sources (serum vs. plasma), different betatrophin ELISA kits (ELISA
kits from Wuhan EIAAB Science vs. other companies), and geographical area demonstrated no significant difference (Table 2). However, when we stratified the results by glycemic status, this indicated that glycemic status may have partially been the source of heterogeneity.

We further conducted a sensitivity analysis to assess the stability of these results. The sensitivity analysis was conducted by sequentially deleting each study to check the influence of an individual study on the results. The conclusions of the sensitivity analysis revealed no significant alteration in the direction of effect, which indicated the stability of our meta-analysis.

Begg’s test and Egger’s regression asymmetry test were conducted to evaluate the publication bias. Begg’s test \( (p = 0.452) \) and Egger’s regression asymmetry test \( (p = 0.297) \) demonstrated that publication bias was not detected.

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**Table 1: Characteristics of studies included in the meta-analysis.**

| Study                        | Country | Sample | Glycemic status | Case group Betatrophin (ng/mL) | Mean BMI (kg/m²) | Control group Betatrophin (ng/mL) | Mean BMI (kg/m²) |
|------------------------------|---------|--------|-----------------|-------------------------------|-----------------|-----------------------------------|-----------------|
| Gómez-Ambrosi et al. (2014 A) [14]^a | Spain   | Serum  | NG              | 75 26.9 ± 15.4               | 39.4 ± 6.7      | 33 45.1 ± 24.4                   | 23.6 ± 1.2      |
| Gómez-Ambrosi et al. (2014 B) [14]^a | Spain   | Serum  | IGT             | 30 18.3 ± 10.7               | 43.7 ± 7.5      | 33 45.1 ± 24.4                   | 23.6 ± 1.2      |
| Gómez-Ambrosi et al. (2014 C) [14]^a | Spain   | Serum  | T2DM            | 15 13.5 ± 8.8                | 39.0 ± 3.6      | 33 45.1 ± 24.4                   | 23.6 ± 1.2      |
| Fenzl et al. (2014)          | Austria | Plasma | NG              | 19 0.973 ± 0.402             | 46.9 ± 1.4      | 20 1.2 ± 0.54                    | 24.6 ± 0.6      |
| Fu et al. (2014)             | USA     | Serum  | NG              | 29 4.42 ± 0.58               | —                | 24 1.96 ± 0.98                   | —               |
| Guo et al. (2015 A) [8]^b     | China   | Serum  | NG              | 30 33.91 ± 46.64             | 32.95 ± 2.84    | 17 21.01 ± 18.25                 | 22.35 ± 1.48    |
| Guo et al. (2015 B) [8]^b     | China   | Serum  | T2DM            | 19 27.1 ± 28.59              | 33.47 ± 2.96    | 17 21.01 ± 18.25                 | 22.35 ± 1.48    |
| Abu-Farha et al. (2016)      | Kuwait  | Plasma | NG              | 62 1.15 ± 0.108              | 34.76 ± 3.22    | 82 0.775 ± 0.046                 | 24.66 ± 2.86    |
| Crujeiras AB et al. (2016)   | Spain   | Plasma | Unknown         | 95 1.24 ± 0.43               | 35.7 ± 4.5      | 48 0.97 ± 0.69                   | 22.9 ± 2.2      |

BMI: body mass index; NG: norm glycemic; IGT: impaired glucose tolerance. Data are presented as means ± SD. ^aFrom the same study. ^bFrom the same study.
Study ID  WMD (95% CI)  % Weight
Javier Gómez-Ambrosi A (2014)  -18.20 (-27.23, -9.17)  0.05
Javier Gómez-Ambrosi B (2014)  -26.80 (-35.96, -17.64)  0.05
Javier Gómez-Ambrosi C (2014)  -31.60 (-41.04, -22.16)  0.05
Anna Fenzl (2014)  -0.23 (-0.52, 0.07)  20.77
Zhiyao Fu (2014)  2.46 (1.44, 3.48)  3.56
Kaifeng Guo A (2014)  12.90 (-5.95, 31.75)  0.01
Kaifeng Guo B (2014)  6.09 (-9.47, 21.65)  0.02
AB Crujeiras (2016)  0.27 (0.24, 0.30)  37.78
Abu-Farha (2016)  0.38 (0.35, 0.40)  37.72
Overall (I-squared = 94.8%, p = 0.000)  0.25 (0.05, 0.45)  100.00

Note: weights are from random effects analysis

Figure 2: Forest plot for the association of circulating betatrophin levels and obesity. The summary estimates were analyzed using a random-effects model. (a) The study by Gómez-Ambrosi et al. [14], where A represents data from individuals with normal glycemic values, B from individuals with impaired glucose tolerance, and C from individuals with diabetes mellitus. (b) The study by Guo et al. [8], where A represents data from individuals with normal glycemic values and B from individuals with diabetes mellitus.

Table 2: Meta-analysis of circulating betatrophin levels and obesity by study characteristic.

| Characteristic                  | No. of studies | Case | Control | WMD (95% CI) | Heterogeneity tests |
|--------------------------------|----------------|------|---------|--------------|---------------------|
| Geographic region              |                |      |         |              | i²                  |
| Europe                         | 4              | 215  | 147     | -18.76 (-37.049, -0.465) | 0.96 |
| Asia                           | 3              | 111  | 116     | 1.16 (-2.579, 4.891)     | 0.096 |
| Other                          | 2              | 48   | 44      | 1.11 (-1.522, 3.744)     | 0.99 |
| Sample source                  |                |      |         |              |                    |
| Serum                          | 6              | 198  | 157     | -9.972 (-25.131, 5.187)  | 0.955 |
| Plasma                         | 3              | 176  | 150     | 0.172 (-0.123, 0.467)    | 0.878 |
| Mean BMI (kg/m²)               |                |      |         |              |                    |
| <35                            | 3              | 111  | 116     | 1.16 (-2.579, 4.891)     | 0.096 |
| ≥35                            | 5              | 234  | 167     | -2.408 (-4.114, -0.703)  | 0.96 |
| Unknown                        | 1              | 29   | 24      | 2.460 (2.015, 2.905)     | 0.00 |
| ELISA kit                      |                |      |         |              |                    |
| Wuhan ELAAB Science Co.        | 3              | 110  | 126     | 0.541 (-0.077, 1.158)    | 0.946 |
| Other                          | 6              | 264  | 181     | -10.478 (-24.627, 3.67)  | 0.948 |
| Mean age                       |                |      |         |              |                    |
| ≥45                            | 6              | 230  | 222     | -7.656 (-11.645, -3.667) | 0.955 |
| <45                            | 2              | 49   | 37      | -0.224 (-0.521, 0.074)   | 0.465 |
| Unknown                        | 1              | 95   | 48      | 0.270 (0.244, 0.296)     | 0.0  |
| Glycemic status                |                |      |         |              |                    |
| NG                             | 5              | 215  | 176     | 0.534 (-0.313, 1.380)    | 0.92 |
| Hyperglycemia                  | 3              | 64   | 83      | -18.547 (-37.148, 0.054) | 0.88 |
| Unknown                        | 1              | 95   | 48      | 0.270 (0.244, 0.296)     | 0.0  |
4. Discussion

Betatrophin, a newly founded adipokine, was believed to play a pivotal part in metabolism. Betatrophin was a major focus in obesity research, for which relevant research results have been published continuously. Several epidemiologic studies have demonstrated that there is an independent and positive association of circulating betatrophin with obesity, whereas other studies reported no association. Inconsistent results were obtained because of the relatively small sample size in the studies, which may be affected by numerous factors. Hence, we conducted a meta-analysis to further evaluate the relationship between circulating betatrophin levels and obesity.

The present study is the first comprehensive meta-analysis that quantitatively explores a possible relationship between circulating betatrophin levels and obesity. Our study showed that circulating betatrophin levels increased in individuals with obesity.

Obesity has increased rapidly over the past few decades, which is associated with a significantly increased risk of premature death. The increasing prevalence of obesity is a challenge to public health care in both developed and developing countries. This obesity epidemic is a major driving force for the increase in the risk of chronic metabolic complications, including type 2 diabetes [22]. Recent studies have demonstrated that a number of adipokines (adiponectin, vaspin, leptin, chemerin, etc.) have been associated with inflammatory responses and metabolic disorders in patients with obesity [23]. However, few data are available regarding betatrophin.

Betatrophin, also called as angiopoietin-like protein 8 (e.g., ANGPTL8), hepatocellular carcinoma-associated protein TD26 and chromosome 19 open reading frame 80 (C19orf80), TD26, refeeding induced fat and liver (RIFL) protein, and lipasin, is a novel protein predominantly secreted in the liver and the adipose tissue. Previous studies in mouse models implicated an important role for betatrophin in a number of metabolic-related pathways, such as lipid metabolism and energy balance in mice [10]. In this meta-analysis of 9 observational studies, individuals with obesity had higher betatrophin levels. There is emerging evidence that supports a positive relationship between circulating betatrophin levels and obesity. Our study showed that betatrophin levels have a close relationship with inflammatory markers, including high-sensitivity C-reactive protein (HsCRP) levels [26], which is central to the development of obesity [27]. Second, betatrophin overexpression was shown to relate to hypertriglyceridemia, while the absence of betatrophin was shown to decrease triglyceride levels without affecting glucose levels [28, 29]. Third, insulin resistance and enhanced insulin contribute to the upregulation of betatrophin levels [30, 31]. Inflammation, lipid metabolism, and insulin resistance are potential modifiable risk factors for obesity [32–34]. Given the connection between obesity and inflammation, lipid metabolism, and insulin resistance, it is obvious that high circulating betatrophin levels are positively correlated with obesity risk.

Since a substantial heterogeneity existed among the analyzed studies, we analyzed the potential contributing factors that might explain why the results varied between the studies. Subgroup and sensitivity analyses were conducted to investigate the potential sources of heterogeneity. BMI is frequently used to measure obesity but does not seem to offer an explanation for this heterogeneity. Betatrophin is predominantly secreted by the liver in humans. Some studies have shown that serum betatrophin levels were significantly higher in patients with nonalcoholic fatty liver disease (NAFLD) compared to those without NAFLD. However, BMI failed to account for the accumulation of fat within the liver. A previous study showed that a higher percentage of visceral fat typically correlates with a higher risk of NAFLD [35]. More studies are needed to elucidate the relationship between circulating betatrophin levels and visceral fat.

Recently, several researchers have raised a concern regarding the potential influence of glycemic status differences on betatrophin levels [36, 37]. Stratified subgroup analyses were conducted to evaluate between-study variations in glycemic status. Notably, the impact of glycemic status seems to partially explain the heterogeneity observed. However, the underlying mechanisms remain unknown and need to be elucidated. Sample source and the ELISA kits used are considered to affect measured betatrophin levels [9, 38]. However, our findings showed that neither were contributing factors to the heterogeneity between studies. In addition, we used a sensitivity analysis in a stepwise manner to reanalyze the statistics of each paper. The results show that the meta-analysis results are stable and provide more reliable interpretation results.

There are several limitations in the present study, which should be addressed in accounting for these results. First, there was a lot of variation in sample sizes of the included studies, which might impact the statistical heterogeneity observed in our meta-analysis. Therefore, high-quality studies consisting of sufficient sample sizes were needed for further evaluation regarding the association between circulating betatrophin levels and adult obesity in the future. Second, the study is based on observational studies, which are particularly vulnerable to information or selection bias. Third, exclusion of grey literature may increase the risk of publication bias. However, no sign of such bias was detected in either Begg’s test or Egger’s regression asymmetry test. Fourth, different forms of betatrophin display different roles,
but in our study, only full-length betatrophin levels were detected in the study population. Fifth, circulating betatrophin concentration might be affected by sex or lipid profiles, which cannot be taken into account because of lack of data. Finally, only fasting, circulating betatrophin was detected. As a food intake-induced hormone, postprandial betatrophin after a standard diet could be more meaningful to be analyzed in future investigations [39].

5. Conclusions

In summary, this meta-analysis demonstrated that circulating betatrophin levels in patients with obesity are higher than those of healthy controls. Thorough understanding of the role of betatrophin in the progression of obesity is helpful to better guide patient treatment. Moreover, further studies are required to explore the possible physiologic mechanisms of betatrophin and its role in obesity.

Disclosure

The funders were not involved in the manuscript writing, editing, approval, or decision to publish.

Conflicts of Interest

The authors declare that there is no conflict of interest regarding the publication of this article.

Acknowledgments

This work was supported by the Foundation for the Author of the National Natural Science Foundation of China (grant numbers 81570721 and 81870548), the Social Development Project of Jiangsu Province (BE2018692), the Natural Science Foundation of Jiangsu Province, China (BK20151331 and BK20191222), the High Caliber Medical Personnel Foundation of Jiangsu Province (LGY2016053), the Six Talent Peaks Project in Jiangsu Province (2015-WSN-006, 2016-SWYY-011), the Scientific Research Projects of Jiangsu Health and Family Planning Commission (Y2018109), and the Science And Technology Support Social Development Project of Zhenjiang City, Jiangsu Province (grant numbers SH2015028 and SH2015042).

References

[1] N. N. Sun, T. Y. Wu, and C. F. Chau, "Natural dietary and herbal products in anti-obesity treatment," *Molecules*, vol. 21, p. 1351, 2016.

[2] M. H. Rouhani, F. Haghighatdoost, P. J. Surkan, and L. Azadbakht, "Associations between dietary energy density and obesity: a systematic review and meta-analysis of observational studies," *Nutrition*, vol. 32, pp. 1037–1047, 2016.

[3] F. J. Kong, L. L. Ma, G. Li, Y. X. Chen, and J. Q. Zhou, "Circulating betatrophin levels and gestational diabetes mellitus: a systematic review and meta-analysis," *PLoS One*, vol. 12, article e0169941, 2017.

[4] D. Liu, H. Qu, H. Wang et al., "Relationship between serum betatrophin levels and the first-phase of glucose-stimulated insulin secretion," *Obesity Research & Clinical Practice*, vol. 12, pp. 9–15, 2018.

[5] M. Ejarray, M. Borlaug, N. Vilarrasa et al., "Angiopoietin-like protein 8/betatrophin as a new determinant of type 2 diabetes remission after bariatric surgery," *Translational Research*, vol. 184, pp. 35–44, 2017.

[6] G. Ren, J. Y. Kim, and C. M. Smas, "Identification of RIFL, a novel adipocyte-enriched insulin target gene with a role in lipid metabolism," *American Journal of Physiology-Endocrinology and Metabolism*, vol. 303, pp. E334–E351, 2012.

[7] A. Fenzl, B. K. Itariu, L. Kosi et al., "Circulating betatrophin correlates with atherogenic lipid profiles but not with glucose and insulin levels in insulin-resistant individuals," *Diabetologia*, vol. 57, pp. 1204–1208, 2014.

[8] K. Guo, J. Lu, H. Yu et al., "Serum betatrophin concentrations are significantly increased in overweight but not in obese or type 2 diabetic individuals," *Obesity*, vol. 23, pp. 793–797, 2015.

[9] S. Wu, H. Gao, Y. Ma, L. Fu, C. Zhang, and X. Luo, "Characterisation of betatrophin concentrations in childhood and adolescent obesity and insulin resistance," *Pediatric Diabetes*, vol. 17, pp. 53–60, 2016.

[10] M. Abu-Farha, D. Sirraman, P. Cherian et al., "Circulating ANGPTL8/betatrophin is increased in obesity and reduced after exercise training," *PLoS One*, vol. 11, article e0147367, 2016.

[11] Z. Fu, F. Berhane, A. Fite, B. Seyoum, A. B. Abou-Samra, and R. Zhang, "Elevated circulating lipasin/betatrophin in human type 2 diabetes and obesity," *Scientific Reports*, vol. 4, p. 5013, 2015.

[12] A. B. Crujeiras, M. A. Zulet, I. Abete et al., "Interplay of atherogenic factors, protein intake and betatrophin levels in obese–metabolic syndrome patients treated with hypocaloric diets," *International Journal of Obesity*, vol. 40, pp. 403–410, 2016.

[13] H. Tuhan, A. Abaci, A. Anik et al., "Circulating betatrophin concentration is negatively correlated with insulin resistance in obese children and adolescents," *Diabetes Research and Clinical Practice*, vol. 114, pp. 37–42, 2016.

[14] J. Gómez-Ambrosi, E. Pascual, V. Catalán et al., "Circulating betatrophin concentrations are decreased in human obesity and type 2 diabetes," *The Journal of Clinical Endocrinology & Metabolism*, vol. 99, pp. E2004–E2009, 2014.

[15] S. Barja-Fernández, C. Folgueira, L. M. Seoane et al., "Circulating Betatrophin Levels Are Increased in Anorexia and Decreased in Morbidly Obese Women," *The Journal of Clinical Endocrinology & Metabolism*, vol. 100, p. E1188–E1196, 2015.

[16] G. A. Wells, B. Shea, D. O’Connell, and Ottawa Health Research Institute, *The Newcastle-Ottawa Scale (NOS) for assessing the quality of non-randomised studies in meta-analyses*, 2018, May 2018, http://www.ohri.ca/programs/clinical_epidemiology/oxford.asp.

[17] B. C. Zhang, W. J. Liu, W. L. Che, and Y. W. Xu, "Serum total adiponectin level and risk of cardiovascular disease in Han Chinese populations: a meta-analysis of 17 case-control studies," *Clinical Endocrinology*, vol. 77, pp. 370–378, 2012.

[18] M. Egger, G. Davey Smith, M. Schneider, and C. Minder, "Bias in meta-analysis detected by a simple, graphical test," *BMJ*, vol. 315, pp. 629–634, 1997.
[19] N. Terrin, C. H. Schmid, J. Lau, and I. Olkin, “Adjusting for publication bias in the presence of heterogeneity,” Statistics in Medicine, vol. 22, pp. 2113–2126, 2003.

[20] C. B. Begg and M. Mazumdar, “Operating characteristics of a rank correlation test for publication bias,” Biometrics, vol. 50, pp. 1088–1101, 1994.

[21] D. F. Stroup, J. A. Berlin, S. C. Morton et al., “Meta-analysis of Observational Studies in Epidemiology: A Proposal for Reporting,” JAMA, vol. 283, pp. 2008–2012, 2000.

[22] N. Dali-Youcef, M. Mecili, R. Ricci, and E. Andres, “Metabolic inflammation: connecting obesity and insulin resistance,” Annals of Medicine, vol. 45, pp. 242–253, 2013.

[23] T. Yamauchi, J. Kamon, H. Waki et al., “The fat-derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity,” Nature Medicine, vol. 7, pp. 941–946, 2001.

[24] M. Abu-Farha, J. Abubaker, and J. Tuomilehto, “ANGPTL8 (betatrophin) role in diabetes and metabolic diseases,” Diabetology and Metabolism Research and Reviews, vol. 33, article c2919, 2017.

[25] M. Abu-Farha, I. Al-Khairi, P. Cherian et al., “Increased ANGPTL3, 4 and ANGPTL8/betatrophin expression levels in obesity and T2D,” Lipids in Health and Disease, vol. 15, p. 181, 2016.

[26] M. Abu-Farha, J. Abubaker, I. Al-Khairi et al., “Circulating angiopeitoin-like protein 8 (betatrophin) association with HsCRP and metabolic syndrome,” Cardiovascular Diabetology, vol. 15, p. 25, 2016.

[27] F. Vohra, F. Alkhudhairey, A. A. Al-Kheraif, Z. Akram, and F. Javed, “Peri-implant parameters and C-reactive protein levels among patients with different obesity levels,” Clinical Implant Dentistry and Related Research, vol. 20, pp. 130–136, 2017.

[28] F. Quagliarini, Y. Wang, J. Kozlitina et al., “Atypical angiopeitin-like protein that regulates ANGPTL3,” Proceedings of the National Academy of Sciences, vol. 109, pp. 19751–19756, 2012.

[29] Y. Wang, F. Quagliarini, V. Gušarova et al., “Mice lacking ANGPTL8 (betatrophin) manifest disrupted triglyceride metabolism without impaired glucose homeostasis,” Proceedings of the National Academy of Sciences, vol. 110, pp. 16109–16114, 2013.

[30] H. Wang, L. Du, T. Wu et al., “Circulating betatrophin is associated with insulin resistance in humans: cross-sectional and interventional studies in vivo and in vitro,” Oncotarget, vol. 8, pp. 96604–96614, 2017.

[31] P. Lu, X. Chen, Z. Zhang et al., “Insulin upregulates betatrophin expression via PI3K/Akt pathway,” Scientific Reports, vol. 7, p. 5594, 2017.

[32] N. M. Bazhan, A. V. Baklanov, J. V. Piskunova, A. J. Kazantzseva, and E. N. Makarova, “Expression of genes involved in carbohydrate-lipid metabolism in muscle and fat tissues in the initial stage of adult-age obesity in fed and fasted mice,” Physiological Reports, vol. 5, article e13445, 2017.

[33] J. M. Perkins, N. G. Joy, D. B. Tate, and S. N. Davis, “Acute effects of hyperinsulinemia and hyperglycemia on vascular inflammatory biomarkers and endothelial function in overweight and obese humans,” American Journal of Physiology-Endocrinology and Metabolism, vol. 309, pp. E168–E176, 2015.

[34] J. N. Fain, “Release of inflammatory mediators by human adipose tissue is enhanced in obesity and primarily by the nonfat cells: a review,” Mediators of Inflammation, vol. 2010, Article ID 513948, 20 pages, 2010.

[35] P. M. Graffy and P. J. Pickhardt, “Quantification of hepatic and visceral fat by CT and MR imaging: relevance to the obesity epidemic, metabolic syndrome and NAFLD,” The British Journal of Radiology, vol. 89, article 20151024, 2016.

[36] S. Ozayzgan, B. Onal, E. M. Kurtulus, H. Uzun, G. Akkan, and D. Konukoglu, “Plasma betatrophin levels of subjects classified with normal, impaired, and diabetic glucose tolerance, and subjects with impaired fasting glucose,” Hormone and Metabolic Research, vol. 49, pp. 434–439, 2017.

[37] T. Gao, K. Jin, P. Chen et al., “Circulating betatrophin correlates with triglycerides and postprandial glucose among different glucose tolerance statuses—a case-control study,” PLoS One, vol. 10, article e0133640, 2015.

[38] Z. Fu, A. B. Abou-Samra, and R. Zhang, “An explanation for recent discrepancies in levels of human circulating betatrophin,” Diabetologia, vol. 57, pp. 2232–2234, 2014.

[39] S. Qiu, X. Cai, H. Yin et al., “Association between circulating irisin and insulin resistance in non-diabetic adults: A meta-analysis,” Metabolism, vol. 65, pp. 825–834, 2016.