ABSTRACT

Introduction: Absence epilepsy is a brief non-convulsive seizure associated with sudden abruptness in consciousness. Because of the unpredictable occurrence of absence seizures and the ethical issues of human investigation on the pathogenesis and drug assessment, researchers tend to study animal models. This paper aims to review the advantages and disadvantages of several animal models of nonconvulsive induced seizure.

Methods: The articles that were published since 1990 were assessed. The publications that used genetic animals were analyzed, too. Besides, we reviewed possible application methods of each model, clinical types of seizures induced, purposes mechanism of epileptogenesis, their validity, and relevance to the absence epileptic patients.

Results: The number of studies that used genetic models of absence epilepsy from years of 2000 was noticeably more than pharmacological models. Genetic animal models have a close correlation of electroencephalogram features and epileptic behaviors to the human condition.

Conclusion: The validity of genetic models of absence epilepsy would motivate the researchers to focus on genetic modes in their studies. As there are some differences in the pathophysiology of absence epilepsy between animal models and humans, the development of new animal models is necessary to understand better the epileptogenic process and, or discover novel therapies for this disorder.

Keywords: Epilepsy, Absence, Animal models, Seizures, Genetic models
1. Introduction

Absence epilepsy, as a childhood disease, is characterized by generalized epileptic activities in both hemispheres of the brain and accompanied by unconsciousness. Absence epilepsy is a non-convulsive seizure and can be classified into typical and atypical forms (Berg et al., 2010; Manning, Richards, & Bowery, 2003; Snead, 1995). Typical absence seizures are defined behaviorally by a paroxysmal loss of consciousness without aura or postictal mood and accompanied with bilateral synchronous spike and wave discharges (SWDs) about 3 Hz in the electroencephalogram (EEG). Usually, the duration of the seizure persists for 3-10 seconds (Figure 1). Misinterpretation with day-dreaming happens in such episodes because they are associated with a fixed and gaze, especially in children (Manning et al., 2003).

Atypical absence seizures are described with complex orofacial automatisms, abnormal neurodevelopmental outcomes, and cognitive impairment in patients. Such atypical episodes are more frequent and lasted more than typical seizure duration. SWDs frequency is less than 3 Hz. Seizure onset and offset are gradual, and there is a minor correlation between the EEG and behavioral changes. For instance, during seizure episodes children might be able to walk or talk (Manning et al., 2003).

The incidence of absence epilepsy in children up to the age of sixteen years is about 2 to 8 out of every 100000 children and with a prevalence of 10% of children with any form of epilepsy (Crunelli & Leresche, 2002). Since the precise etiology of absence epilepsy is unknown and multiple mechanisms are involved in the pathophysiology (Karimzadeh et al., 2017), hopefully, most of our knowledge about mechanisms and treatment of human absence epilepsy arise from the use of appropriate chemical and genetic animal models. This study aimed to review the pathophysiology of absence epilepsy as well as different kinds of experimental models. Besides, the validity and similarity of different absence epilepsy models have been illustrated.

1.1. Search strategy

We searched for the relevant publications in PubMed, Wiley, and Google scholar. The keywords of “absence epilepsy”, “absence seizure”, “animal model”, “pharmacological animal models”, “chemical models”, and “genetic models” were used in the title and abstract of articles.

The articles were searched from 1990 to 2018. To explore most researchers’ focus on the genetic models, the number of publications that used genetic animals were analyzed, too.

1.2. Statistical analysis

The number of publications was analyzed by 1-way Analysis of Variance (ANOVA) followed by Tukey post hoc test in SPSS V. 20. The significance level was established by $P \leq 0.05$.

1.3. Pathogenesis of absence epilepsy

Studies reveal that absence epilepsy arises from an aberration of the interplay between some areas of the cortex, especially the somatosensory cortex and the thalamus. Investigations in human and animal models identified hypersynchronization in the thalamocortical circuit that generates SWDs in absence seizure (Avoli, Gloor, Kostopoulos,
In typical absence seizures, the main components of the thalamocortical circuit contribute to the creating SWDs involving the reticular and ventrobasal (medial and lateral) nuclei of the thalamus and somatosensory perioral region of the somatosensory cortex (Meeren, van Luijtelaar, da Silva, & Coenen, 2005). Besides, the channelopathy and imbalance of excitatory and inhibitory receptors in the other thalamocortical neurons such as intrathalamic and dorsal thalamic nuclei are involved in SWDs generation (Karimzadeh et al., 2017). The atypical absence seizure involves both the thalamocortical circuit and the limbic system (Cortez, McKerlie, & Snead, 2001).

Absence epilepsy may be related to several malfunctions in the nervous system. Hyperexcitability of the cortex and hyperfunction of the GABAergic tonus in the thalamus trigger hyperactivity of low-threshold T-type calcium currents activated by hyperpolarization in the thalamus. Naturally, there are reciprocal excitatory glutamatergic and inhibitory GABAergic projections between the cortex and thalamus. These neurons can produce both tonics and burst firings in different conditions which are specialized for the flow of information and sleep spindles at the network, but hyperactivation of the GABAergic neurons in the reticular nucleus of the thalamus leads to a burst of synchronized firing and causing inhibitory postsynaptic potentiation in thalamocortical neurons (Figure 1). Gamma-aminobutyric acid (GABA) induces hyperpolarization, especially through GABAB receptors in the specific relay nuclei and the reticular nuclei of the thalamus and then hyperactivation of the low-voltage-activated T-type calcium currents that create a rebound burst of fast spikes leading to the next cycle of the oscillation (Budde, Pape, Kumar, & Huguenard, 2006; Meeren, Pijn, Van Luijtelaar, Coenen, & da Silva, 2002; Polack, Guillemain, Daransart, Depaulis, & Charpier, 2007).

### 1.4. Animal models of absence epilepsy

The validity of animal models in the experimental studies is identified by three characters of reproducibility, predictability, and quantifiability. There are several animal models for absence epilepsy such as electrical stimulation, chemical injection, and genetic models of Drosophila, mice, rats, baboons, and cats. They should be comparable behaviorally, electroencephalographic, ontogenetic, etiologic, and pharmacologically with human absence epileptic disease. These models must indicate SWDs in the EEG as the hallmark of absence epilepsy in humans, but there are some differences in some clinical criteria in the animal models that are presented in Table 1. Therefore, researchers should select the models based on the aim of the study considering the limitation of each model (Fisher, 1989; Manning et al., 2003; Pitkänen, Schwartzkroin, & Moshé, 2005).

### 1.5. Pharmacological animal models of absence epilepsy

Several pharmacological compounds induce acute or chronic absence seizures (Table 2).

Penicillin, low-dose pentylenetetrazole (L-PTZ), 4,5,6,7 tetrahydroisoxazolo (4,5-c) pyridine-3-ol (THIP), and gamma-hydroxybutyrate (GHB) are the most common compounds for the induction of typical absence epilepsy. Also, AY-9944 (trans-1, 4-bis[2-chloro-benzylamino-ethyl] cyclohexane dihydrochloride) and methylazoxymethanol acetate (MAM)-AY-9944 are administered for the induction of atypical absence seizures (Tang, & Loke, 2011).

### Table 1. Characteristics of experimental animal models for absence seizures

| Most Important Feature                  | Pharmacologic Animal Models | Genetic Animal Models                                      |
|----------------------------------------|----------------------------|-----------------------------------------------------------|
| EEG and behavior similar to the human condition | LD-PTZ, GHB, THIP, AY-9944, MAM-A | GAERS, WAG/Rij, Lethargic, slow-wave-epilepsy mouse, tottering mouse |
| Reproducibility and predictability     | LD-PTZ, GHB, THIP, AY-9944, MAM-A | GAERS, WAG/Rij, lethargic                                  |
| Quantifiable                          | LD-PTZ, GHB, THIP, AY-9944, MAM-A | GAERS, WAG/Rij, lethargic                                  |
| Appropriate pharmacology              | LD-PTZ, GHB, PLC, AY9944     | GAERS, WAG/Rij, Lethargic, slow-wave epilepsy, tottering   |
| Unique developmental profile          | LD-PTZ, GHB, AY9944          | GAERS, WAG/Rij, lethargic                                  |
| Exacerbated by GABAergic drugs        | LD-PTZ, GHB, AY9944, MAM-A   | GAERS, WAG/Rij, lethargic                                  |
| Involvement of thalamocortical mechanisms | LD-PTZ, GHB, PLC, AY9944, MAM-A | GAERS, WAG/Rij, lethargic                                  |
| Blocked by GABAB receptor antagonists | LD-PTZ, GHB, AY9944, MAM-A   | GAERS, WAG/Rij                                           |

Jafarian, M., et al. (2020). Experimental Models of Absence Epilepsy. Basic and Clinical Neuroscience, 11(6), 715-726.
1.6. Chemical models of typical absence epilepsy

1.6.1. Penicillin

Local administration of some antibiotics such as cephalosporin and penicillin onto the cortex or systemically (250000–6000000 U/kg) at high doses can induce a seizure in cats and rodents. Penicillin acts as a GABA agonist and has limited effects in rodents rather than cats. Because of the unstable penetration of penicillin through the blood-brain barrier, it produces multifocal spikes with only occasional bursts of bilaterally synchronous SWDs associated with the attention deficit. In other words, seizures initiated through parenteral penicillin have minor similarities to clinical absence seizures in rats.

1.6.2. Low-dose pentylenetetrazole

PTZ, a tetrazole derivative, is the most commonly used epileptic induced agent for systemic convulsive epilepsy. PTZ is a GABAA receptor antagonist and interacts with GABAA receptors at the picrotoxin binding site. The extensive use of PTZ in different types of epilepsy has made this model an identifier of anticonvulsants and proconvulsants. The low dose of PTZ (20–30 mg/kg) injected intraperitoneally (IP) or subcutaneously produces freezing and is used to induce absence-like seizures from 3 weeks of age. L-PTZ produces twitching of vibrissae at the frequency of 6-7 Hz SWD on the EEG in rodents. Generally, seizure initiates 2-10 min after injection and depends on the dose and species. SWDs duration is about 2-3 s in rodents (Deyn et al., 1992; MacDonald & Barker, 1977; Tang & Loke, 2011).

1.6.3. GHB and g-butyrolactone (GBL) models

GHB is a GABA metabolic substance that is produced naturally in the mammalian brain. Endogenous GHB exists in low concentration in different areas of the nervous system. Auto-radiographic studies have shown the presence of GHB receptors in the cortex, thalamus, striatum, hypothalamus, substantia nigra, and hippocampus. The affinity of GHB for GABAB receptors has been shown, especially in the thalamo-cortical network. Following the IP administration of GHB, electrographic and behavioral events occur similar to generalized absence seizures in cats, rats, and monkeys (Deyn et al., 1992; Pitkänen et al., 2005). The intravenous administration of GHB (200 mg/kg) induces the absence epileptic attacks in the same frequency of human SWDs (2.5 Hz) and clinical symptoms such as head drops, behavioral immobility, pupillary dilation, staring, rhythmic eye movements, and stereotypical automatisms. This model is considered a valid model because it is predictable, reproducible, and produces electrographic and behavioral symptoms similar to the human absence epilepsy condition. Besides, as the kinetics of the GHB is detected well then it is a good model for the study of some drugs with unknown pharmacokinetic features (Cortez, & Snead, 2006).

Furthermore, GBL as a pro-drug of GHB increases the reproducibility and predictability of the GHB model of absence seizures. GBL is used more because of the consistency and rapidity of onset of its effects (Bearden, Snead, Healy, & Pegram, 1980) and has been shown to produce the same EEG and behavioral effect as that of GHB. A single dose application of GBL generates absence seizures through the activation of the GHB-related neuromodulation or interaction with GABAergic inhibitory and excitatory neurotransmission in the cortico-thalamo-cortical circuit (Bearden et al., 1980; Pitkänen et al., 2005).

Administration of GHB (240 mM) changes the rhythmicity of EEG in the GHB models. The peak components of GBL rise 1 minute after GBL injection and fell rapidly to undetectable levels within 5 minutes. GBL-induced SWDs can be quantitated as cumulative duration (in seconds) per 20-min cycle or as a percentage of control SWDs duration. This pharmacological rat model is an appropriate experimental model for the study of the mechanisms of bilaterally synchronous SWDs production and screen for the anti-seizure activity of antiepileptic drugs in generalized absence seizures (Depaulis, Snead, Marescaux, & Vergnes, 1989; Karamahmutoğlu et al., 2013; Tang & Loke, 2011).

1.7. 4,5,6,7 Tetrahydroisoxazolo (4,5-c) pyridine-3-ol (THIP)

THIP, as a GABA agonist, induces bilaterally synchronous SWDs in rats. THIP is a selective GABA agonist for the extrasynaptic delta-subunits of GABAA receptors. Delta-subunits of GABAA receptors were abundantly found in the thalamus and neocortex. The optimum dose of THIP to induce bilaterally synchronous SWDs is 5-10 mg/kg and lasted 7 to 9 s. The dose of THIP to elicit SWDs is 7.5 mg/kg (IP). The model is quantified similarly to the GHB model (Depaulis et al., 1989; Fariello & Golden, 1987; Fisher, 1989; Pitkänen et al., 2005).

1.8. Chemical Models of atypical absence epilepsy

1.8.1. AY-9944

AY-9944 is a common compound for induction of atypical absence seizure. Subcutaneous administration of AY-9944 (7.5 mg/kg) inhibits the reduction of 7-dehydrocholesterol to cholesterol and leads to an abnormal cognitive
outcome. It causes seizure onset in the prepubescent and maximum peak in the adult periods. Anti-absence drugs decrease these seizures and phenytoin and GABA agonist drugs exacerbate them. The AY-treated rat can be used to investigate the pathogenesis and treatment of this malignant disorder (Chan et al., 2004; Cortez, Cunnane, & Snead, 2002; Cortez et al., 2001).

In some atypical absence epilepsy studies, antimitotic agent MAM with AY-9944 is used to produce seizures. In the MAM-AY model (double-hit model), rats display bilaterally synchronous SWDs with a frequency of 4 to 6 Hz. The MM-AY-induced atypical absence seizures; however, ethosuximide, as well as sodium valproate, suppress the induced seizures. The histopathologic assessment showed the MAM-treated rat brains have hippocampal heterotopias, atrophy, and abnormalities of cortical lamination. The MAM-AY-treated rat is a reproducible model for studying refractory atypical absence seizures in children with brain maldevelopment (Pitkanen et al., 2005; Serbanescu, Cortez, McKerlie, & Snead, 2004).

### 1.8.2. Genetic models of absence epilepsy

During recent decades, it has been revealed that genetic factors play a critical role in idiopathic generalized epilepsies, including absence epilepsy. Some evidence emphasizes the role of genes in the pathogenesis of Childhood Absence Epilepsy (CAE). It has been reported that monozygotic twins compared with dizygotic twins suffered more from CAE (Berkovic, Howell, Hay, & Hopper, 1994; Marini et al., 2003). Although there are several types of chemical animal models for CAE, genetic models are more valid for research. Spontaneous absence seizures and reproduced symptoms in the genetic models could be considered as the most important advantages compared to chemical models and they are reliable to clarify the pathophysiology of the human condition.

Some genetic models of rats and mutant mice are currently used in most experiments. Six mutant mice are suitable models for absence epilepsy, including lethargic, tottering, mocha, stargazer, slow-wave-epilepsy, and ducky.
1.8.3. Genetic mutant mouse models

Spontaneous mutations in mice associated with other epilepsy models and generated by genetic engineering approaches are critical models to comprehend the pathogenesis of human epilepsies. They were considered as a reproducible biological model for experimental strategies to prevent or reverse the onset of seizures. Spontaneous SWDs have also been observed in mutant mouses, which is usually accompanied by other neurological disorders (ataxia). A Mendelian basis underlies epileptic phenotype in most of the mice models. Since epileptic mutant mice is a favored model in neurobiology and neuropharmacology for the study of abnormal brain synchronization, it has a pioneering role in the neurogenic analysis of seizure disorders. These models were a key role to figure out the significance of single genes in the hereditary transmission of epilepsy in the mammalian nervous system. Yet there are several absence epileptic mutants mouse (tottering, lethargic, stargazer, mocha, slow-wave-epilepsy, and ducky), but their number are restricted by the rate of mutations and feasible dysfunctions (Crunelli & Leresche, 2002; Pitkänen et al., 2005).

1.8.4. Tottering mouse

The phenotype of tottering mice includes motor seizures and ataxia (Tokuda et al., 2007). There were no abnormalities in their EEG recordings associated with motor seizures (Tokuda et al., 2007). The mutation in these mice is the tottering locus on chromosome 8 (Campbell & Hess, 1996). This chromosomal mutation creates an abnormal α1A subunit of voltage-gated Ca$^{2+}$ channel. This subunit forms the core of P/Q Ca$^{2+}$ channels and this malformation decreases the Ca$^{2+}$ current (Bourinet et al., 1999). It has been believed that a decrease in the Ca$^{2+}$ current in Purkinje cells has been involved in the pathophysiology of ataxia (Rhyu, Abbott, Walker, & Sotelo, 1999). Tottering mice are the best model for ataxia.

The manifestation of tottering seizures (behavioral arrest, assuming a fixed staring posture) is the same as CAE in humans. The frequency of SWDs in these mice
is higher (6-7 Hz) than humans and absence seizures are suppressed by anti-absence drugs (Heller, Dichter, & Sidman, 1983; Kaplan, Seyfried, & Glaser, 1979).

1.8.5. Lethargic mouse

The lethargic mouse has a spontaneous mutation on chromosome 2 in the gene encoding the β4 subunit of voltage-gated calcium channels (Frankel, 1999). This subunit is an auxiliary part of voltage-gated calcium channels that regulate Ca\(^{2+}\) influx (Burgess, Jones, Meisler, & Noebels, 1997). The phenotype of a lethargic mouse appears after 15 days of age by ataxia and lethargic behavior accompanied by focal motor seizures. Generalized cortical SWDs are the second seizure type which is recorded in their EEG (Pinault et al., 1998). The characteristic of their absence seizures is the same as human CAE and tottering absence seizures (Pinault et al., 1998).

There are no structural and pathological changes in the brain and spinal cord but some immunological discrepancies such as splenic and thymic degeneration and lymphocytopenia are seen (Dung, 1977). Interestingly, after two months of age, their immune function and loss of body weight would be recovered but their fertility is reduced (Devanagondi, Egami, LeDoux, Hess, & Jinnah, 2007).

1.8.6. Stargazer mouse

This mutant was named stargazer because of mutation in stargazing protein (γ2 subunit of T-type or Cav3.1 calcium channel). A single mutation occurs in the gene encoding stargazing on mouse chromosome 15 (Lettas et al., 1997; Noebels, Qiao, Bronson, Spencer, & Davisson, 1990). This protein has a modulatory role in voltage-gated calcium channels (Burgess & Noebels, 1999). Also, γ2 subunit is necessary for AMPA (α-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid) receptors for synaptic targeting in the ionotropic glutamate receptors (Chen et al., 2000).

An increase in calcium currents in thalamic relay neurons may have a crucial role in the absence seizure initiation in stargazer mice (Snead, 1995). The phenotype of stargazer is similar to the other mutant mice that are ataxia, paroxysmal dyskinesia, absence seizure, and head tossing (Sharp et al., 2001).

1.8.7. Mocha mouse

The characteristic of absence seizures in mocha is the same as a stargazer mouse. The frequency of SWDs is 6-7 Hz and phenotype is accompanied by ataxia, tonic-clonic seizures, pigment dilution, and increased bleeding time (Sarkisian, 2001). The gene encoding the AP-3δ protein on chromosome 10 has been mutated in these mice (Noebels, 2003). The AP-3 complex is a heterodimer that regulates lysosome trafficking and other related organelles (Di Pietro et al., 2006; Kantheti et al., 1998).

1.8.8. Slow-wave epilepsy mouse

SWDs in these mutants appear with a frequency of 3 to 4.5 Hz during absence seizures (Cox et al., 1997). It has been considered as a valid model for absence epilepsy because of seizure phenotype and anti-absence-drug responses (Blumenfeld, 2005). The main mutation underlying SWDs generation in the spike-wave epilepsy mice has been occurred in the gene encoding the ubiquitous sodium hydrogen exchanger on chromosome 4 (Papale et al., 2000).
al., 2009). This exchanger regulates the hemostasis, cell volume, and mitogenic responses to growth factors.

These mutants have some extra phenotype as the same as the other mice models for absence epilepsy. Ataxia, tonic-clonic seizures, and cerebellum degeneration are some phenotypes that appear in these mutants in addition to absence seizures (Qiao & Noebels, 1993).

1.8.9. Ducky mouse

The frequency of SWDs in ducky mice like most of the mice model of absence epilepsy is 6 Hz (Barclay et al., 2001). Ataxia, limb dyskinesia, and developmental dysgenesis of some brain areas like the cerebellum, medulla, and spinal cord are some additional phenotypes that exist in these mice. The gene encoding r2 and a 6 subunit of high-voltage activated Ca2+ channels on chromosome 9 have been mutated (Porter et al., 1993). These channels are extremely permeable to Ca2+ in excitatory and non-excitatory cells and have a critical role in the absence seizures genesis (Zhang, Mori, Burgess, & Noebels, 2002).

1.8.10. Genetic rat models of absence epilepsy

In the mutant mice models, generally, a single gene underlies the epileptic phenotype (SWDs); however, in humans, a polygenetic mode of inheritance occurs. So it seems that polygenic rat models are more valid in comparison to other models (Sarkisova & van Luijtelaar, 2011). Genetic absence epileptic rats from Strasbourg (GAERS) and WAG/Rij rats represented spontaneous absence seizure attacks for the first time about 25 years ago. It is believed that a single gene might be responsible for causing absence seizure attacks (Rudolf et al., 2004) but different genes are involved in the characteristics of absence seizures and SWDs (Rudolf et al., 2004). Different studies with various approaches revealed that these two rats have a similar feature to human absence epilepsy. Statistical data showed fabulous information about absence epilepsy that was published with the use of these two rats (more than 500 articles; Figure 2).

1.8.11. Genetic Absence Epilepsy Rat From Strasbourg (GAERS) rats

In France (Strasbourg), a fully inbred strain of rats was derived from an outbred Wiistar colony. It is notable that, 100% of animals show the EEG and behavioral characteristics of absence seizures in three-month-old rats. However, the first detectable SWDs appear around 30 days of age. In the first seizures, the SWDs appear rarely and in short duration (1-3 s). During a lifetime, the severity of seizures (duration and frequency) increases gradually. The most SWDs occur at around 4 to 6 months old. There are not any differences in the characteristics of SWDs between males and females, that represent autosomal genetic transmission (Coenen & Van Luijtelaar, 2003; Crunelli & Leresche, 2002).

The frequency of SWDs in these rats is 7-10 Hz (Seidenbecher & Pape, 2001). The behavioral manifestation is the same as a typical absence seizure such as loss of cautiousness and some chewing motion during the seizure. No pathological and structural abnormalities have been reported in these rats (Avanzini, Vergnes, Spreafico, & Marescaux, 1993). Anti-absence drugs can suppress seizure attacks. The most important difference between absence seizures in GEARS and human is the higher frequency of SWDs (Avanzini et al., 1993).

It is believed that polygenetic mutation is involved in the absence seizures genesis in this rat model (Rudolf et al., 2004). A polygenic mutation on the chromosomes of 4, 7, and 8 seems that regulate the SWDs initiation in the GAERS (Rudolf et al., 2004). Single nucleotide mutation in the gene encoding T-type Ca2+ channel has occurred in GAERS. Future studies are necessary to determine the exact chromosomal mutations (Powell et al., 2009).

1.8.12. WAG/Rij rats

The WAG/Rij strain is a subline of the WAG strain that was derived from Wistar supply by A. L. Bacharach at the Glaxo Laboratories in London (UK) in 1924. In other words, WAG/Rij is an inbred strain in which brother-sister mating has been carried for more than 130 generations. The behavioral study showed that WAG/Rij rats have a short latency to move out from the home cage into familiar and new environments (Coenen & Van Luijtelaar, 2003; Sarkisova & van Luijtelaar, 2011). Many similarities in the cognitive deficits, as well as behavioral disturbances, have been shown in the WAG/Rij rats and CAE patients (Karson, Utkan, Balç, Arıcıoğlu, & Ateş, 2012).

SWDs are emerged at around 60 to 80 days age in WAG/Rij (Seidenbecher & Pape, 2001). At the age of 3 months, half of the WAG/Rij display fully developed SWDs, and at 6 months of age all of the animals show SWDs and it occurs alternatively until the death of the animals (Pitkänen et al., 2005). SWDs in the WAG/Rij rats are very age-dependent. In 6-month-old rats, the number of SWDs is about 16–18 discharges per hour.

Sex differences are very little (Coenen & Van Luijtelaar, 1987). Generally, burst activities last 1–30 s with a spike-wave frequency of 7–11 Hz in the cortical EEG of
adult WAG/Rij rats (Sarkisova & van Luijtelaar, 2011). The behavioral manifestation of seizures (facial myoclonic jerks, eye twitching, and head tilting) is the same as GAERS rats (Sitnikova & Van Luijtelaar, 2007). It is noticeable that besides the high frequency of SWDs, the age of SWDs initiation is different from humans. The pharmacological responses to anti-absence drugs in these rats are the same as humans (Karimzadeh et al., 2013). A polygenetic mutation (chromosome 5 and 9) control the SWDs in these rats (Gauguier et al., 2004).

It seems that channelopathy of slow and fast Ca\(^{2+}\) channels has a critical role in the SWDs generation of WAG/Rij rats as well as the other genetic animal models (Gorji, Mittag, Shahabi, Seidenbecher, & Pape, 2011). Besides, some cognitive deficits, as well as pathological changes (neuronal injury and cell death) in several brain areas (hippocampus and neocortex) have been reported in these rats (Jafarian et al., 2015).

1.8.13. The quantitative comparison of genetic animal models of absence epilepsy

The articles which used genetic models to study absence epilepsy were counted from 2000 to 2018. The articles included 293 WAG/Rij, 74 GEARS, 21 stargazers, 19 tottering mice, 7 ducky mice, 3 lethargic, and 3 slow-wave epilepsy. The number of articles that used WAG/Rij rats was significantly more than the others (P<0.001).

2. Conclusion

Our findings of many diseases have been found from valid animal models. Although there are several models for the screening, quantification, and evaluation of absence epilepsy, the important issue is still the reproducibility of the full clinical syndrome and pathogenesis as well as different etiology of the absence seizures. In this regard, it should be noted that how much the existing models are useful for discovering new drugs for 30% of medically refractory epilepsy. It seems that current animal models are designed based on the underlying mechanism of seizure, not epileptogenesis. Besides, epilepsy is classified as a chronic disorder and most animal models are not appropriate for long-term chronic induction of diseases (Havasimehr, Saffarzadeh, Divanbeigi, & Karimzadeh, 2018). As regards, childhood absence epilepsy has multifactorial genetic etiology, so it seems that genetic animal models are more suitable than chemical models, because of the close correlation of their EEG features and clinical appearances to the human condition. Among genetic models of mice and rats, the WAG/Rij and GAERS strains of Wiślar rats have asserted to be valid and predictive models of human absence epilepsy. Although there are some differences in seizure characteristics such as frequency, ontogeny, and the time of seizure developing, the pharmacologic re-activities and the pathogenesis explained in both strains are very similar to human absence epilepsy. Differences between the two strains suggest the involvement of several genetic mechanisms in the SWDs pathogenesis. Multidisciplinary studies of these two strains lead to finding trustable information about the role of the cortex and the thalamus, and other subcortical circuits. As statistics show, the most published papers in the absence epilepsy was designed based on the WAG/Rij rats. It should be mentioned that pharmacological models such as the GHB model are used to assess the mechanisms of receptors in the pathogenesis and new anti-seizure compounds.

As all of these models have some shortcomings and limitations, so researchers should be open-minded and enthusiastic to more valid animal models for a better understanding of the epileptogenic process and or discovery of novel therapies for this heterogeneous disorder.

Ethical Considerations

Compliance with ethical guidelines

This study was approved by the Ethics Committee of the Iran University of Medical Sciences.

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Authors’ contributions

Writing – original draft: Maryam Jafarian; Data collection: Mohammad Esmaeil Alipour; Data analysis: Fariba Karimzadeh.

Conflict of interest

The authors declared no conflicts of interest.

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