Incision Wound Healing Test of Ethanolic Extract Gel from Salaon (Parsonsia alboflavescens [Dennst.] Mabb.) Leaves in Male Rats

Nilsya Zebua*, Muharni Saputri, Winda Giovana Sijabat, Inda Aritika Sri Retno Wulandari, Ira Nofriani, Winda Agustiani Zai, Ramadani Arda Arista, Muhammad Suhaime, Aula Firsia

Department of Pharmacy, Faculty of Pharmacy, Tjut Nyak Dhien University, Sumatera Utara, Indonesia

Abstract

BACKGROUND: An incision wound is a wound caused by a sharp object. One of the plants that can be used as a traditional medicine for an incision wound is salaon leaves. The prepared form chosen is the gel because it is easy to use and its distribution is faster on the skin.

AIM: The aim of the study was to explore whether salaon leaf extract gel meets the quality evaluation requirements and to know the effectiveness of ethanol extract gel of salaon (Parsonsia alboflavescens [Dennst] Mabb.) leaves to cure a scar on male rats.

METHODS: Experimental method with salaon leaves as sample. Simplicia of salaon leaves extracted by maceration method, and then arenga gum gel-based formulation, afterward evaluation, and testing of its effectiveness on the healing of incision wound of male rats. Normal data were analyzed by the One Way Analysis of variance (ANOVA) and Post Hoc Tuckey Method. Kruskal-Wallis test used for abnormal data.

RESULTS: The whole formula shows homogenous results, normal pH, stability, dissemination and good viscosity, less irritating, and with good adhesion. The results of a statistical analysis of the wound on the 8th day with ANOVA test indicate a significant difference between the five groups $p = 0.000$ ($p < 0.05$) and Post-Hoc Tukey Honest Significant Difference test results obtained a concentration of 5% not significantly different from the positive control Placenta extract gel. The optimum dose that can heal incision wound in male rats is 5%.

CONCLUSION: The whole formulation qualifies for the quality of the gel formulation. Ethanol extract gel of salaon leaves can heal wound on male rats and the optimal dose is 5%.

Introduction

A wound is an end result of a bodily harm that reasons harm and penetrates into the layers of the skin [1]. The basic principle of top-rated wound healing is to minimize tissue damage and provide good enough tissue perfusion and oxygenation, the right nutrients, and a moist wound healing environment to restore the anatomical continuity and characteristic of the affected part [2].

Research on natural ingredients on substances that may accelerate wound healing is one among the items that are presently developing and is being tried by researchers and ancient practitioners around the world. The use of treatments with natural ingredients is becoming increasingly popular today because it has fewer side effects than conventional preparations [3]. The time course of healing for an acute wound usually ranges from 5 to 10 days, or within 30 days [4].

One of the plants that can be used as traditional medicine is salaon (Parsonsia alboflavescens [Dennst] Mabb.) leaves. Salaon is derived from the Apocynaceae family. Empirically salaon leaves have been used to treat bruises, gastric wounds, as well as open wounds. In conventional medication inside the Asia Pacific region, the species Apocynaceae is used to deal with fever, malaria, pain, diabetes, and gastrointestinal diseases [5].

In general, the preparation of wound drugs in the form of liquid or semi-solid form. Half-solid dosage forms such as ointments, cream, and gel are options for all the better ones including longer drug contact and protecting wounds from extraordinary diagnostic. The dosage form of the gel is easier to use and the spread on the skin is faster. In addition, the gel has cooling, moisturizing properties, and easily penetrates the skin to provide a healing effect. Gel preparations can protect the skin from excessive dehydration. The formulation and selection of the right base on the manufacture of gels will affect the number and speed of the active substance to be absorbed. Ideally, the base and carrier should be easy to wear on the skin, not irritating, and safe to use on [6].

In this study, the base of the gel used is gum derived from Arenga pinnata. Gum has been widely used as a thickener, emulsion stabilizer as well as additive in various food and medicine industries. Gum is also known to have antimicrobial and antioxidant properties [7].
Materials and Methods

**Materials**

Salaon (*P. alboflavescens* [Dennst.] Mabb.) leaves, aquadest, Arenga pinnata, ethanol 96% (v/v), ethanol 70% (v/v), ethanol p.a 99.5% (v/v), glycerol 99.5%, lidocaine 2%, methyl paraben, propylene glycol.

**Collection of materials**

The samples used are salaon leaves from Sidamanik Subdistrict, Simalungun Regency, North Sumatra, Indonesia, collected from December 2020 to January 2021.

**Making of simpliscia powder**

Salaon leaves are cleaned of dirt, then dried in a dryer cabinet with a temperature of ±45°C then mashed with a blender and weighed the weight of dry powder.

**Arenga gum extraction**

Arenga gum was weighed then washed thoroughly with aqueducts and then smoothed with a ratio of 1:10 water. Once smooth, the sugar palm fruit is filtered with mesh 18 and then put in the refrigerator at 4°C for 24 h. Then a cross-union of supernatant and ethanol 96% with a ratio of 1:2, after which it is stored in the refrigerator at 4°C for 24 h. The formed sediment is filtered, then soaked in ethanol p.a 99.5% for 5 min then strain and squeeze until dry. Once dry is crushed until slightly smooth, put it in a container and store in the desiccator until dry [8].

**Basic gel formulation**

The basic formula of the standard gel contains 5 g Carboxymethyl cellulose- Na (CMC-Na), 10 g glycerol 5 g, propylene glycol, and aqua dest till 100 ml [9].

**Modification gel formulation**

The modified gel formula is made by replacing CMC-Na as thickener and gel shaper with arenga gum, the addition of extracts, and addition of methylparaben as preservatives with formula 5 g arenga pinnata, 10 g glycerol, 5 g propylene glycol glycerol, aqua dest till 100 ml, and salaon leaves extract with some concentration (2.5%, 5%, and 7.5%) (Table 1).

The material is weighed according to the weight of each formula. In beaker, glasses are filled with 100 ml of aqua dest, and added arenga gum then homogenized using a magnetic stirrer on the hotplate until a homogeneous period is formed. Furthermore, methylparaben, glycerol, and propylene glycol are added so that a homogeneous gel base is obtained. After the base of the gel was homogeneous then added ethanol extract salaon leaves then homogenized.

**Gel evaluation**

Gel evaluation includes organoleptic, homogeneity test, pH test, cycling test, viscosity test, spread resistance test, and irritation test for volunteers.

**Organoleptic**

Gel inspection and description consist of an examination of form, color, and odor [10].

**Homogeneity**

All developed gels were tested for homogeneity by visual inspection. Its appearance and the presence of aggregates were tested [11], [12].

**Measurement of pH**

The pH of the gel was measured by using a digital pH meter. Digital pH meter is dipped in a diluted gel sample, let stand for a while and the result is adjusted to the standard pH meter [13].

**Stability test**

Each formula is placed in a transparent glass container. The gel is stored at a temperature of 4 ± 2°C for 24 h, then transferred to an oven at 40°C for 24 h (1 cycle). Testing was conducted as many as six cycles or for 12 days and observed the occurrence or absence of changes that occur in each formula. Gel conditions compared during experiments with previous gel conditions.

**Table 1: Modification gel formulation**

| Material                   | Formula and composition | Blanko | F1     | F2     | F3     |
|----------------------------|-------------------------|--------|--------|--------|--------|
| Extract ethanol of salaon leaves | 5                       | 2.5    | 5      | 7.5    |
| Arenga gum (g)             | 5                       | 5      | 5      | 5      |
| Glycerol (g)               | 10                      | 10     | 10     | 10     |
| Propylene glycol (g)       | 5                       | 5      | 5      | 5      |
| Methyl paraben (g)         | 0.1                     | 0.1    | 0.1    | 0.1    |
| Aquadest (ml)              | Till 100                | Till 100 | Till 100 | Till 100 |

**Spreadability**

The gel is weighed as much as 0.5 g and then placed in the middle of a scaled round glass. On top of the gel is placed other round glass or other transparent materials and ballast so that the weight of the glass is round and ballast 150 g, silenced 1 min, then noted the diameter of its spread [14].

Open Access Maced J Med Sci. 2021 Sep 04; 9(A):776-781.
Measurement of viscosity

A total of 100 ml of gel is put in a tube-shaped container and then installed spindle 4. Spindles should be submerged in test preparations. Viscometers are turned on and mounted rotors can rotate at a speed of 60 rpm.

Irritation test

The irritation test is done with a closed patch test in which the cream is applied to the inside of the arm, then covered with gauze, after 24 h, watch for any symptoms. This trial was conducted on 25 female volunteers aged 18–25 years, with the condition: Healthy woman, 18–25 years old, no history of allergy, participate voluntarily, healthy physically and mentally [15].

Table 2: Organoleptic gel test results

| Formulation          | Form       | Color  | Odor        |
|----------------------|------------|--------|-------------|
| Blanko               | Semi-solid| Peach  | Arenga odor |
| Placenta extract gel | Semi-solid| White  | Gel odor    |
| F1                   | Semi-solid| Black  | Leaf odor   |
| F2                   | Semi-solid| Black  | Leaf odor   |
| F3                   | Semi-solid| Black  | Leaf odor   |

Table 3: Gel homogeneity test results

| Formulation          | Homogeneity |
|----------------------|-------------|
| Blanko               | Homogeneous |
| Placenta extract gel | Homogeneous |
| F1                   | Homogeneous |
| F2                   | Homogeneous |
| F3                   | Homogeneous |

Table 4: pH gel test results

| Formulation          | pH value |
|----------------------|----------|
| Blanko               | 6.13     |
| Placenta extract gel | 6.12     |
| F1                   | 6.12     |
| F2                   | 6.12     |
| F3                   | 6.12     |

Adhesion test

The sample of 0.25 g aped between two glass objects and a load of 1 kg for 5 min. The load is large and the load is 80 g on the appliance. The time at which the gel comes off the glass object.

Preparation of experiment animal

The FMIPA USU Animal Research Ethics Committee has approved the protocol study with Ethical clearance number 0151/KEPH-FMIPA/2021. The test animal used is a male rat. The test animals prepared were as many as 25 heads that were randomly divided into 5 groups of animals and then adapted to environmental conditions for 1 week. Criteria of test animals include age 2–3 months, male sex, weight 200 g, a healthy condition characterized by active movement, rat food in the form of pellets and for beverages used bottles as much as 20–45 ml per day and mice placed in one cage.

Table 6: Viscosity results

| Formulation          | Viscosity (cPa.s) |
|----------------------|-------------------|
| Blanko               | 99.9              |
| Placenta extract gel | 72.6              |
| F1                   | 71.8              |
| F2                   | 44.9              |

Table 7: Irritation test results

| Formulation          | Volunteer | Symptoms    |
|----------------------|-----------|-------------|
| Blanko               | 1         | No symptoms |
| Blanko               | 2         | No symptoms |
| Blanko               | 3         | No symptoms |
| Blanko               | 4         | No symptoms |
| Blanko               | 5         | No symptoms |
| Placenta extract gel | 6         | No symptoms |
| Placenta extract gel | 7         | No symptoms |
| Placenta extract gel | 8         | No symptoms |
| Placenta extract gel | 9         | No symptoms |
| Placenta extract gel | 10        | No symptoms |
| F1                   | 11        | No symptoms |
| F1                   | 12        | No symptoms |
| F1                   | 13        | No symptoms |
| F1                   | 14        | No symptoms |
| F1                   | 15        | No symptoms |
| F2                   | 16        | No symptoms |
| F2                   | 17        | No symptoms |
| F2                   | 18        | No symptoms |
| F2                   | 19        | No symptoms |
| F2                   | 20        | No symptoms |
| F3                   | 21        | No symptoms |
| F3                   | 22        | No symptoms |
| F3                   | 23        | No symptoms |
| F3                   | 24        | No symptoms |
| F3                   | 25        | No symptoms |

Table 5: Spreadability results

| Formulation          | Spreadability (cm) |
|----------------------|--------------------|
| Blanko               | 5                  |
| Placenta extract gel | 5.1                |
| F1                   | 5.2                |
| F2                   | 5.3                |
| F3                   | 6.6                |

Incision wound healing test

The hair around the area to be injured is shaved and cleaned using a 70% alcoholic cotton swab. The need is done after the previous rats were annexed using lidocaine. In the area is made an incision in the back area using scalpel along 1 cm to the subcutaneous layer.

Table 8: Adhesion test results

| Formulation          | Adhesion     |
|----------------------|--------------|
| Blanko               | Homogeneous  |
| Placenta extract gel | Homogeneous  |
| F1                   | Homogeneous  |
| F2                   | Homogeneous  |
| F3                   | Homogeneous  |

A total of 25 male rats Wet divided into 5 groups, each group consisting of 5 heads. The rat group was given an ethanol extract gel of saaloo leaves.

a. Group I: Negative control i.e. rats given gel base
b. Group II: Positive control i.e. mice given comparison gel.
c. Group III: Rats were given ethanol extract gel of saaloo leaves concentration of 2.5%
d. Group IV: Mice were given ethanol extract gel of saaloo leaves concentration of 5%
e. Group V: Mice were given ethanol extract gel of saaloo leaves concentration of 7.5%

In male rats that suffered cuts, the gel was thinly applied 1 (one) time a day (the wound was not closed). Observation of wounds is done every day visually by measuring the length of the wound until the wound is considered healed (the wound is dry and the length of the wound is reduced.)
Data analysis

Data collection and analysis were conducted based on the length of the cut and the time it took for the wound in mice to heal using a formulation of saloon leaf extract gel with different concentrations and gel comparison for the cuts on the market. Data analysis using SPSS 26.0 Free trial method one way Analysis of variance (ANOVA) and Post-Hoc Tukey Honest Significant Difference (HSD). Kruskal-Wallis tests are performed for normal non-distributed data.

Results

Incision wound healing test of ethanol extract gel of saloon leaves aims to know the presence of an effect on the decrease in the length of the wound and the percentage of wound healing. This test was conducted on male rat test animals. The cut wounds that have been made are then given therapy using saloon leaf ethanol extract gel according to the treatment group namely negative control group (gel base), positive control group (Placenta extract gel), F1 (concentration group 2.5%), F2 (concentration group 5%), and F3 (concentration group 7.5%) (Tables 2-7).

Measurement of the length of the cut in all treatment groups is done visually from the first day to the eighth day. The average measurement of the length of the cut wounds of each group of male rats is shown in Tables 8-10.

Table 8: Adhesion test results

| Formulation               | Adhesion (s) |
|---------------------------|--------------|
| Blanko                    | 25           |
| Placenta extract gel      | 38           |
| F1                        | 37           |
| F2                        | 34           |
| F3                        | 24           |

Table 9: Average length of wound

| Days | Treatment      | Average length of wound (cm) |
|------|----------------|------------------------------|
|      | G1             | G2                          |
| 1    | 0.96           | 0.94                        |
| 2    | 0.98           | 0.9                         |
| 3    | 0.92           | 0.74                        |
| 4    | 0.84           | 0.62                        |
| 5    | 0.82           | 0.56                        |
| 6    | 0.74           | 0.4                         |
| 7    | 0.64           | 0.24                        |
| 8    | 0.54           | 0.06                        |

The results of the adhesion test showed that the entire formula had an adhesion of more than 4 s. Standard gel adhesiveness is not <4 s. The longer the gel’s adhesion time the better because it allows the active substance to be fully eliminated. From the data obtained all formulas meet the standards of gel adhesiveness [20].

Irritant tests are performed on the skin to determine whether or not there are side effects of gel preparations. The formula is declared safe in case of no allergic reactions or irritations to the skin. Based on the results of the irritant test obtained showing the absence of allergic reactions and irritations caused by the preparations, it can be concluded that all formulas are safe to use.

The results of the adhesion test showed that the entire formula had an adhesion of more than 4 s. Standard gel adhesiveness is not <4 s. The longer the gel’s adhesion time the better because it allows the active substance to be fully eliminated. From the data obtained all formulas meet the standards of gel adhesiveness [20].

Discussion

The results of the homogeneity test of the gel preparation showed that the entire gel formula has good homogeneity that does not contain any grain in a piece of glass. Flattened gel, if there are no granules in a piece of glass, it can be said that the preparation is homogeneous [16]. The results of experiments that have been conducted on gel preparations from saloon leaves ethanol extract there are no granules on a piece of glass, so the preparation is said to be homogeneous.

pH test results show all formulas have a pH that is close to the normal pH range of the skin so that all formulas can meet good and safe tropical preparation criteria if applied to damaged skin such as cuts. pH preparations that meet the pH criteria of the skin are in the range of 5–8 [17]. If the pH value of the composition preparation is too acidic, it will cause skin irritation, and if the pH value of the composition preparation is too alkaline, it will cause skin dryness and irritation [18].

Stability test results showed that all preparations were stable, with no discoloration and shape change after storage for six cycles. The results of the spread test showed that the spreadability of all gel preparation formulas tested was in the skin’s spread range of 5.0–7.0 cm, so it can be concluded that the gel preparations meet the requirements of good gel spreadability [17]. The therapeutic effect of gels depends on their distribution. The distribution of gels contributes to the uniform application of gels on the skin. Therefore, the produced gels should have good distribution and ideal quality for topical application [19].
ethanol extract has less healing power compared to gels containing 5% salaon leaves extract. Then ethanol extract gel of salaon leaves 5% is the most effective to accelerate the healing of wounds (Figure 1).

The data of the measurement of the length of the incision wound obtained is then obtained statistically using SPSS 26.0 Free trial. Statistical analysis of the measurement data of the length of the incision wound includes formality test, One Way ANOVA test, Post-Hoc Tukey HSD test, and Kruskal-Wallis. Test normality data using the Shapiro-Wilk test. If the data is distributed normally, a homogeneity test is conducted with the Levene test. If the variance of normal and homogeneous distributed data is continued by the parametric test method, namely One Way ANOVA tests. When data are not distributed normally, a Kruskal-Wallis non-parametric test is used. Hypotheses are considered meaningful when the value p < 0.05 if in the test ANOVA produced a value of p < 0.05 then continued by conducting a Post Hoc Tukey HSD analysis.

Based on the data that has been obtained, normally distributed data is data on the 3rd, 4th, 5th, 6th, 7th, and 8th days with (p > 0.05). The data meet the requirements of one way ANOVA test while for normal non-distributed data, data on the 1st and 2nd day are conducted Kruskal-Wallis test. Based on the results of the Kruskal-Wallis test found that broadly found a significant difference that is the value of p > 0.05.

The results of the analysis of the measurement data of the length of the cut on the 8th day obtained from the One Way ANOVA test showed a significance value of p < 0.05 which is p = 0.000, which means there is a meaningful difference in each treatment group. To find out there are significant differences in each treatment group followed by a Post-Hoc Tukey HSD test.

Based on the results of the Post Hoc Tukey HSD test it is known that the length of the gel cut with a concentration of 5% does not differ significantly from the positive control of Placenta extract gel. Quantitative analysis of preparations containing ethanol extract of salaon leaves concentration is 5% more effective compared to concentrations of 2.5% and 7% due to a faster decrease in the length of cuts.

Figure 1: Wound length measurement graph

Conclusion

All preparation formulas of gel extract ethanol leaves salaon meet the quality requirements of gel evaluation. The preparation of salaon leaf ethanol extract gel can heal wounds in male rats. Post-Hoc Tukey HSD test results obtained a concentration result of 5% not significantly different from Placenta extract gel as a positive control. The optimum dose of ethanol extract gel of salaon leaves that can cure incision wound in male rats in the dosage form of the gel is 5%.

References

1. Er塞尔 M, Uyanikgil Y, Akarca FK, Ozete E, Altunci YA, Karabey F, et al. Effects of silk Sericin on incision wound healing in a dorsal skin flap wound healing rat model. Med Sci Monit. 2016;22:1064-78. https://doi.org/10.12659/msm.897981 PMid:27032876
2. Gadekar R, Saurabh MK, Thakur GS, Saurabh A. Study of formulation, characterisation and wound healing potential of transdermal patches of curcumin. Asian J Pharm Clin Res. 2012;5(4):225-30.
3. Tabassum N, Ahmad F. Role of natural herbs in the treatment of hypertension. Pharmacogn Rev. 2011;5(9):30-40. https://doi.org/10.4103/0973-7847.79097 PMid:22096316
4. Velnar T, Bailey T, Smrkolj V. The wound healing process: An overview of the cellular and molecular mechanisms. J Int Med Res. 2009;37(5):1528-42. https://doi.org/10.1177/147327530803700531 PMid:19930861
5. Chan EW, Wong SK, Chan HT. Apocynaceae species with antiproliferative and/or antiplasmoidal properties: A review of ten genera. J Integrat Med. 2016;14(4):269-84. https://doi.org/10.1016/s2095-4964(16)60261-3 PMid:27417173
6. Farid N, Awaluddin N, Hamzah S, Yusuf M, Rahmania R. Formulation and antibacterial activity test of hand sanitizer ethanol extract of basil (Ocimum Sanctum. L) against Escherichia coli and Streptococcus aureus. Med Kesehatan Politeknik Kesehatan Makassar. 2020;15(2):228-37. https://doi.org/10.32382/medkes.v15i2.1764
7. Sayuti K, Neswati N, Hijra R, Effendi E. Antioxidant activity of “Kolang Kaling” Jam which is added with “Pucuk Merah” (Syzygium olea) fruit juice. Asian J Appl Res Community Dev Empowerment. 2020;4(1):28-33. https://doi.org/10.29165/ajarcde.v4i1.36
8. Kaban J, Revery J, Tarigan J, Zebua NF. Modified extraction and purity test of Arenga pinnata Gum. Asian J Pharm Clin Res. 2018;11:148-50. https://doi.org/10.22159/ajpcr.2018.v11i1.26593
9. Maswadeh HM, Semreen MH, Naddaf AR. Anti-inflammatory activity of Achillea and Ruscus topical gel on carrageenan-induced paw edema in rats. Acta Pol Pharm. 2006;63(4):277-80. PMid:17203864
10. Leyni L, Ginting EE, Hafiz I. Formulation and evaluation of candidenut (Aleurites moluccana L.) oil in gel preparation. Asian J Pharm Res Dev. 2020;8(5):41-3. https://doi.org/10.22270/ajprv.v8i5.842
11. Gupta R, Gupta DG. Formulation development and evaluation of ant-inflammatory potential of Cordia obliqua topical gel on animal model. Pharmacogn J. 2017;9(6):s93-8. https://doi.org/10.5530/pj.2017.6s.163
12. Saiesh P, Shabaraya AR, Shripathy D, Soman L. Formulation and evaluation of topical gel containing econazole nitrate. Int J Pharm Bio Sci. 2014;3(3):82-91.
13. Febriyenti F, Putri RF, Suharti N. Formulation and evaluation of patchouli oil gel. J Sains Farmasi Klinis. 2019;6(3):191-4. https://doi.org/10.25077/jsfk.6.3.191-194.2019
14. Garg A, Aggarwal D, Garg S, Sigla AK. Spreading of semisolid formulation: An update. Pharm Tecnol. 2002;9(2):84-102.
15. Hasri NM, Zebua NF, Sudewi S. Test of burn wounds healing effects of collagen from snakehead fish (Channa striata) bone in the preparation of cream on male white rats (Rattus norvegicus). Indones J Pharm Clin Res. 2020;3(1):62-75. https://doi.org/10.32734/idprcl.v3i1.3977
16. Naziniwaty N, Karsono K, Zebua NF, Nerdy N. Antioxidant activity and antiaging gel formulation grapefruit peel (Citrus maxima Merr.) ethanolic extract. Der Pharm Lett. 2016;8(20):84-94.
17. Zebua NF, Putra ED, Harahap U, Kaban J. Durian seed utilization as a base material of tropical gel. Asian J Pharm Clin Res. 2018;11(1):174-7. https://doi.org/10.22159/ajpcr.2018.v11s1.26600
18. Ariffin B, Nasution R, Desriani N, Marianne M, Helwati H. Antimicrobial activity of hand lotion of flower Mimusops elengi. Open Access Maced J Med Sci. 2019;7(22):3748-56. https://doi.org/10.3889/oamjms.2019.496 PMid:32127968
19. Dantas MG, Reis SA, Damasceno CM, Rolim LA, Rolim-Neto PJ, Carvalho FO, et al. Development and evaluation of stability of a gel formulation containing the monoterpene borneol. ScientificWorldJournal. 2016;2016:7394685 https://doi.org/10.1155/2016/7394685 PMid:27247965
20. Lumentut N, Edi HJ, Rumondor EM. Formulasi dan uji stabilitas fisik sediaan krim ekstrak buah pisang goroho (Musa Acuminata L.) konsentrasi 12.5% sebagai Tabir Surya. J MIPA. 2020;9(2):42-6. https://doi.org/10.35799/jmuo.9.2.2020.28248