Latest Trend of Milk Derived Exosomes: Cargos, Functions, and Applications

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Exosomes are nanosized phospholipid bilayer vesicles released to the extracellular environment. Exosomes from various tissues or cells are being studied and there has been a growing interest in milk exosomes research due to their emerging role as messengers between cells and the fact that it can be produced in large quantities with rich source of milk. Milk derived exosomes (MDEs) contain lipids, microRNAs, proteins, mRNAs as well as DNA. Studies of exosome cargo have been conducted widely in many research areas, especially exosomal miRNAs. In this paper, we reviewed the current knowledge in isolation and identification, cargos, functions mainly in intestinal tract and immunity system of MDEs. Its application as drug carriers and diseases biomarker are also discussed. Furthermore, we also consider critical challenges of MDEs application and provide possible directions for future research.

Keywords: milk, exosome, cargo, function, application

INTRODUCTION

Extracellular vesicles (EVs) are lipid bound vesicles secreted into the extracellular space by cells. Based on the biogenesis, release pathways, size, content, and function, EVs are differentiated into three subtypes, including microvesicles, exosomes, and apoptotic bodies (1). Exosomes are nanosized (40–100 nm diameter) phospholipid bilayer vesicles released to the extracellular environment through multivesicular bodies after budding with the plasma membrane (2). Compared with other extracellular vesicles (EVs), cargos from exosomes are sorted in a regulated, non-random way and play essential roles in cell-to-cell communication (3). MDEs are regarded as one of the most important signalsomes mediating cellular communication between mother and her offspring.

Milk exosomes have been successfully separated from bovine colostrum and milk (4), porcine milk (5), rat milk (6), goat milk (7), wallaby milk (8), and human breast milk (9) (Table 1). The membrane structure of the exosome is crucial to the cargos inside. Benefiting from the phospholipid bilayer protection, separated exosomes are stable in terms of size and biological activities when stored frozen (−80°C) (38). The membrane allows miRNAs within exosomes to avoid degradation in the gastrointestinal tract and to be further absorbed in the intestine (38, 39).

The aim of this review is to comprehensively summarize and discuss the current research status of milk exosomes including isolation and identification, cargos as well as application in the scientific area. Future research and further application of MDEs is also discussed (Figure 1).
ISOLATION AND IDENTIFICATION

Ultracentrifugation is the most popular method to separate exosomes from various fluids (40). However, it requires great amount of time to complete. For example, the centrifugation with subsequent sucrose density gradient ultracentrifugation required more than 24 h (4). Ultrafiltration procedures were also used to successfully purifying exosomes and do not require special equipment (41). Combined use of membrane filters (0.1–0.2 s) with differential centrifugation could better eliminate the large vesicles and obtain relatively pure exosomes. In addition, HPLC-based method could obtain highly pure exosomes but these processes need dedicated equipment and are not easy to scale up (42). Exosomes can also be obtained by using precipitation methods, such as using polyethylene glycol, then the precipitate can be isolated using low-speed centrifugation or filtration. Commercially precipitation solutions such as ExoQuick has been used by many researchers as it increases exosome recovery and is relatively rapid (40). To obtain more specific isolation of exosomes, immunoaffinity capture methods based on affinity with antibodies to exosomal proteins or specific saccharide residues on the exosome surface have also been used by many researchers.

Exosomes are identified by size, morphology, and membrane proteins. Physical analysis are done using nanoparticle tracking analysis (NTA), dynamic light scattering (DLS), electron microscopy, and tunable resistive pulse sensing (TRPS) in order to determine particle size or concentration. Chemical or biochemical analysis are done via staining, immunoblotting, or proteomic analysis to give information on the exosomal contents (43). Regardless of source, exosomes have a buoyant density range of 1.13–1.21 g/ml. Using Transmission electron microscopes (TEM), Chen et al. (5) observed a greater density at the center of the milk exosomes by ultracentrifugation. Under TEM, MDEs normally exhibit a round morphology and uniform, unimodal distribution in size.

The exosomal proteins are heavily dependent on the tissue or cell type from which is it derived. Due to their endosomal origin, exosomes normally contain membrane transport and fusion proteins, tetraspanins, heat shock proteins, proteins involved in multivesicular body biogenesis, as well as lipid-related proteins and phospholipases (44). Due to different sources, wide variations regarding proteins exist across exosomes. Both colostrum and mature MDEs contain surface marker proteins including tetraspanins such as CD9, CD63 and CD81, milk fat globule-epidermal growth factor-factor 8 (MFG-E8), raft-associated proteins including flotillin-1 as well we internal markers such as tumor susceptibility gene 101 (Tsg101), ALG-2-interacting protein X (Alix), and heat shock protein 70 (HSP70; (45, 46). The endosomal sorting complex and other associated proteins such as Alix and Tsg101 are involved in exosome biogenesis thus are observed (47). Exosomes do not have surface markers such as integrin-β1, p-selectin, CD40, and endoplasmic reticulum (ER) marker calnexin which are considered as markers for other multivesicular bodies (48). The antibody-based techniques including western blot or ELISA are popular for rapid confirmation of exosomal proteins.

CARGOS OF MILK DERIVED EXOSOMES

Similar to exosomes isolated from other fluids or tissues, cargos of MDEs include lipids, microRNAs, proteins, mRNAs, and DNA (3, 49) (Figure 2). The exosomal cargos are sorted in a regulated, non-random process and they play essential roles in cell-to-cell communication (3).

MicroRNAs

MicroRNAs (miRNAs) are small regulatory RNA molecules consisting of 19–24 nucleotides in length and play important roles in a wide range of physiologic and pathologic processes (50, 51). MDEs were found to contain a considerable amount of mammary gland and immune-related microRNAs. Hata et al. (4) found that RNA below 200 nt was more concentrated in the MDEs than the supernatant. Among the 24 human breast milk exosome samples, miRNAs had a range of 9.7–228.2 ng/ml (52). Porcine MDEs were reported to contain 176 known and 315 novel mature miRNAs (5). Similarly, this was also reported in bovine Colostrum and milk by Hata et al. (4). Lin et al. (53) reported increased miRNAs in piglet serum by orally ingestion of bovine and porcine milk exosomes. Colostrum had higher expression of immune-related miRNAs than mature milk. Compared to piglets given in mature milk, 9 of 13 immune-related miRNAs had higher expression in the serum of piglets given Colostrum (54). Izumi et al. (39) reported that immune (miR-15b, miR-27b, miR-106b, miR-155, and miR-223) and development related miRNAs (miR-27b, miR-34a, and miR-130a) are significantly higher in Colostrum than in mature milk.

Several abundant exosomal miRNAs from human and porcine milk were shared between species. The conservation of the miRNAs shared among species is both in sequence homology and their incorporation in milk extracellular vesicles which indicate that they are evolutionarily selected to benefit the newborn (55). MiRNAs profiles are reported to differ among different species (39, 51). Expression of miRNAs in milk exosomes is affected by many other factors, such as host health condition and lactation stage. Up-regulation of miR-142-5p, miR-223, miR-183, and miR-99a-5p and down-regulation of miR-2285-3p and miR-101 were detected when the bovine mammary gland was challenged with Staphylococcus aureus infection (56). Chen et al. (51) identified 245 miRNAs in raw milk and individual miRNA can be significantly altered at different periods of lactation. Interestingly, the expression of seven miRNAs are relatively constant throughout the lactation process. Izumi et al. (39) reported that mothers’ background (race and lifestyles) can also influence milk miRNAs profile.

Proteins

Various researches have been focusing on the exosomal miRNAs. Proteins cargos also play important roles in physiology and pathology. Wang et al. (57) reported that exosomal proteins promote pre-metastatic niche formation and modulate the site-specific metastasis of tumor cells by inducing lymphangiogenesis, angiogenesis, and permeability. Exosome also delivers cargos including proteins and RNAs to the brain and it was shown that feeding an exosome- and RNA-depleted diet impaired...
### TABLE 1 | Summary of the isolation and validation techniques of milk derived exosomes in literature published since 2017.

| Sample Source                  | Isolation Strategy                          | Validation                                      | Comments                                                                 | Reference |
|--------------------------------|---------------------------------------------|-----------------------------------------------|--------------------------------------------------------------------------|-----------|
| Human breast milk              | Centrifugation with ExoQuick               | NTA, TEM, Western blot (Hsp70, CD9)           | Exosomes prevent necrotizing enterocolitis by reducing inflammation and injury as well as restoring tight junction proteins. | He et al. (12) |
| Human breast milk              | Ultracentrifugation                         | NTA, TEM, Western blot (CD81, CD63)           | Exosomal miR-148a was negatively associated with infant weight, fat mass, and fat free mass, while miR-30b was positively associated with infant weight, percent body fat, and fat mass at 1 month. | Shah et al. (13) |
| Human breast milk              | Centrifugation with exosome isolation kit   | TEM, NTA, Western blot (CD81, CD63)           | Exosomes attenuated purine nucleotide catabolism and improved energy status in oxidatively stressed IEC-6 cells | Wang et al. (15) |
| Cow milk                       | Ultracentrifugation and size exclusion chromatography | TEM, Western blot (Tsg101, Flot-1, Alix, CD63) | Milk exosomes can be taken up by intestinal epithelial cells and mediated functional intracellular delivery of siRNA. | Warren et al. (13) |
| Cow milk                       | Ultracentrifugation and size exclusion chromatography | Western blot (Hsp 90, CD63, Tsg101, NTA)     | Milk exosomes can be used as nanocarriers of functional miRNAs in RNA-based therapy. | Pozo-Acebo et al. (14) |
| Cow milk                       | Ultracentrifugation                         | TEM, DLS, Western blot (CD63, CD9)           | Exosomes successfully delivered epicatechin gallate into SHYSYS cells and exhibited enhanced neuroprotective effects. | Luo et al. (16) |
| Cow milk and colostrum         | Ultracentrifugation                         | Western blot (CD9, CD63milk only, Hsp70)     | Exosomes and miRNA can cross the placenta and promote embryo survival in mice | Sadri et al. (22) |
| Cow milk and yak milk          | Ultracentrifugation with rennet precipitation | TEM, DLS, Western blot (CD63, Hsp70, Tsg101) | Exosomes prevent necrotizing enterocolitis by reducing inflammation and injury as well as restoring tight junction proteins. | He et al. (12) |
| Porcine colostrum and mature milk | Ultracentrifugation and SEC                | NTA, TEM, Western blot (Tsg101)              | The pathways related to homeostasis are upregulated in colostrum exosomes whereas pathways related to endothelia cell development and lipid metabolism are upregulated in milk exosomes. | Ferreira et al. (19) |
| Caprine colostrum and mature milk | Ultracentrifugation                         | DLS, TEM, Western blot (CD9, CD63)           | miR-30a-5p, miR-22-3p, and miR-26a are highly conserved in colostrum and mature milk in cows, caprines and humans. | Yun et al. (20) |
| Colostrum powder               | Ultracentrifugation                         | DLS, NTA, AFM                                 | Exosomes can be taken up by human intestinal epithelia cells, not cytotoxic | Ross et al. (17) |
| Porcine colostrum and mature milk | Ultracentrifugation and SEC                | NTA, TEM, Western blot (Tsg101)              | The pathways related to homeostasis are upregulated in colostrum exosomes whereas pathways related to endothelia cell development and lipid metabolism are upregulated in milk exosomes. | Ferreira et al. (19) |
| Cow milk                       | Ultracentrifugation                         | DLS, SEM, Western blot (CD63, CD9)           | Exosomal miR-148a was negatively associated with infant weight, fat mass, and fat free mass, while miR-30b was positively associated with infant weight, percent body fat, and fat mass at 1 month. | Shah et al. (13) |
| Human breast milk              | Centrifugation with ExoQuick               | NTA, TEM, Western blot (Hsp70, CD9)           | Exosomes successfully delivered epicatechin gallate into SHYSYS cells and exhibited enhanced neuroprotective effects. | Luo et al. (16) |
| Human milk                     | Ultracentrifugation                         | TEM, NTA, Western blot (CD81, CD63)           | Exosomes and miRNA can cross the placenta and promote embryo survival in mice | Sadri et al. (22) |
| Cow milk                       | Centrifugation with exosome isolation kit   | TEM, NTA, Western blot (CD81, CD63)           | Exosomal miR-148a was negatively associated with infant weight, fat mass, and fat free mass, while miR-30b was positively associated with infant weight, percent body fat, and fat mass at 1 month. | Shah et al. (13) |
| Cow milk                       | Ultracentrifugation                         | TEM, NTA, Western blot (CD9, CD63)           | Exosomes successfully delivered epicatechin gallate into SHYSYS cells and exhibited enhanced neuroprotective effects. | Luo et al. (16) |
| Skim milk                      | Ultracentrifugation                         | TEM, NTA, Western blot (CD9, Tsg101, CD63)   | Exosomes can protect macrophages from chemotherapeutic drug-induced cytotoxicity. | Matic et al. (23) |
| Cow milk                       | Differential centrifugation                | TEM, NTA, Western blot (CD9, Tsg101, CD63)   | Exosomal miR-148a was negatively associated with infant weight, fat mass, and fat free mass, while miR-30b was positively associated with infant weight, percent body fat, and fat mass at 1 month. | Shah et al. (13) |
TABLE 1 | Continued

| Sample Source | Isolation Strategy | Validation | Comments | Reference |
|---------------|-------------------|------------|----------|-----------|
| Cow milk      | Ultracentrifugation | Western blot (CD9, CD63, Tsg101) | IL-2 and IL-12 enhanced IFN-γ production with cow milk exosomes but not alone. | Komine-Aizawa et al. (24) |
| Cow milk      | Differential centrifugation | TEM, NTA, Western blot (Tsg101, Alix, CD9, CD91) | Milk exosome based drug delivery system constructed showed controlled drug release, biocompatibility and effective in treating OSCC | Zhang et al. (25) |
| Cow milk      | Ultracentrifugation | TEM, NTA, Western blot (Tsg101, Alix, CD9, CD91) | Exosomes can alter murine gut microbiota and SCFA in feces, as well regulate local intestinal immunity. | Tong et al. (26) |
| Canine colostrum | Ultracentrifugation | Western blot (Alix, Tsg101, Hsp70) | Exosomes modified the proliferation and secretory profiles in canine mesenchymal stem cells. | Villatoro et al. (27) |
| Human Breast milk | Centrifugation and filtration with ExoQuick reagent | Electrophoresis, DLS | Human milk-derived exosomes induced proliferation- and epithelial mesenchymal transformation-related changes. MDEs inhibited proliferation and DNMT1 expression in cells with knockdown of mRNA-148a | Reif et al. (28) |
| Cow milk      | Ultracentrifugation | DLS, TEM, Western blot (CD63, CD81, ANXA5, FLOT1, ICAM, Tsg101) | Exosomes can be absorbed as intact particles from the gastrointestinal tract via the Fc receptor. It can be modified with ligands to promote retention in target tissues. | Betker et al. (29) |
| Cow milk      | Ultracentrifugation | TEM, DLS, Western blot (CD9, CD63) | Milk exosomes contain IncRNA which participate in immunity, development, and reproduction. LncRNA had different expression patterns during different stages of lactation. | Zeng et al. (30) |
| Cow milk      | Ultracentrifugation | NTA, TEM, Flow cytometry (CD63, Hsp70, CD9, CD81) | Milk exosomes prevented experimental NEC-induced intestinal injury by increasing goblet cell production and ER function. | Li et al. (31) |
| Yak milk and cow milk | Ultracentrifugation/ ultracentrifugation with rennet precipitation | TEM, DLS, Western blot (CD63, Hsp70, Tsg101) | Yak milk exosomes has a more efficient effect on IEC-6 cell growth under hypoxic conditions than cow milk exosomes. | Gao et al. (32) |
| Breast milk   | Ultracentrifugation | NTA, Western blot (CD81, clathrin) | Human breast milk-derived exosomes allow IECs to be protected from oxidative stress. | Martin et al. (33) |
| Skim milk     | Ultracentrifugation | NTA, TEM, Western blot (CD9, Tsg101, CD63) | Milk exosomes are bioavailable and miRNAs have unique tissue distribution patterns | Manca et al. (34) |
| Cow milk      | Ultracentrifugation | DLS, NTA, SEM, AFM, Western blot (CD63, CD81, Tsg101, Alix) | Milk exosomes can deliver chemotherapeutic drug paclitaxel and effectively inhibit tumor growth. | Agrawal et al. (35) |
| Cow colostrum | Density gradient centrifugation | Western blot (Alix, Tsg101) | Colostrum derived exosomes are enriched with proteins regulating the immune response and growth. | Samuel et al. (36) |
| Buffalo milk  | Centrifugation Exiqon isolation kit | SEM, DLS | As delivery vehicle, exosome encapsulation enhances the stability, solubility and bioavailability of curcumin. | Vashisht et al. (37) |

OSCC, oral squamous cell carcinoma; TEM, transmission electron microscope; NTA, nanoparticle tracking analysis; NEC, necrotizing enterocolitis; ER, endoplasmic reticulum; DLS, dynamic light scattering; SEM, sanning electron microscopy; AFM, atomic force microscopy; AFM, atomic force microscopy.

spatial learning and memory in mice (58). The β-amyloid and α-synuclein are related to the propagation and diagnosis of Alzheimer’s Disease and Parkinson’s Disease and exosomes appear to participate in the spread of these two proteins (59). Rahman et al. (60) reported that proteins within milk extracellular vesicles provide information of host physiology and immunology. They observed 118 differentially expressed proteins between uninfected cattle and bovine leukemia virus-infected cattle. These proteins are involved in diverse biological activities such as metabolic processes, cellular processes, catalytic activities, and so on. Samuel et al. (36) using quantitative proteomics analysis showed that the proteomic cargo of exosomes change based on the lactation state of cows. Compared to that mature MDEs which were enriched with proteins related to transport and apoptosis, colostrum derived exosomes are more enriched with proteins implicated in the immune response, inflammatory response, antimicrobial peptides, cell growth and complement activation.

mRNA, rRNA, and Long Non-coding RNAs (lncRNA)

Microarray analysis showed that most of the milk mRNAs were present in exosomes. Hata et al. (4) observed a considerable amount of mRNAs in milk exosomes. Admyre et al. (52) reported that ribosomal RNA (18S and 28S) is very low in human breast MDEs. Presence of exosomal mRNAs such as CD63, CD36, ErI, FAS, MFG-E8, MHC-II were confirmed in both milk and colostrum by RT-PCR (45). Zeng et al. (30) identified 3,475 novel lncRNAs and 6 annotated lncRNAs in bovine milk exosomes. They also observed that expression of lncRNAs vary
across the stages of lactation. The exosomal RNAs can exert functional effects because of their stability as they are packaged in membranes (61).

**STABILITY**

Cargos encapsulated in exosomes membranes are protected against enzymatic and non-enzymatic degradation. MiRNAs and mRNAs were detected with a considerable amount in the milk although RNase concentration was extremely high (4). The miRNAs and mRNAs have been found in commercial dairy products, such as infant formula, which have undergone stringent industrial processes (51). These RNAs are also stable under harsh treatments including low pH, RNase, and freezing condition (4, 39). The miRNAs in breast MDEs are still high after RNase, freeze-thawing and acidic (pH = 1) treatments (62), thus allowing dietary intake of miRNAs by infants. However, synthetic miRNAs were rapidly degraded under the degradative conditions whereas the endogenous milk miRNAs were resistant to treatment (39). Commercial dairy milk is highly enriched with bta-miR-223 and bta-miR-125b. Pieters et al. (63) observed relatively small differences in expression levels of immune-related miRNAs between raw milk and commercial milk suggesting that processing did not affect most of the miRNAs in the milk. The stability of MDEs makes it highly resistant against degradative conditions in the intestinal tract, thus enabling its uptake by epithelial cells. Bovine milk exosomal lncRNAs, similar to miRNAs, were stable during *in vitro* digestion, no matter with what kind of digestive juices, such as saliva, gastric juice, pancreatic juice, and bile juice (30). Benmoussa et al. (64) simulated gastrointestinal tract conditions and tested how *Bos Taurus* bta-miR-223 and bta-miR-125b withstood digestion. The authors found out that although the miRNAs were decreased most in the stomach, a large number of miRNAs in the upper small intestine compartments suggested their bioaccessibility.

Although it was experimentally proved that the exosomal cargos were effectively protected by the exosomal membrane, the exosomal miRNAs in artificial formula are deficit compared to the raw milk (65). The pasteurization process can decrease exosomes concentrations by ∼50%, due to membranes disruption and cargos degradation (66) by disrupting. The levels of miRNAs are significantly lower in the infant formula compared to raw milk, especially the extensively
hydrolyzed formula (39). A loss of milk miRNAs and exosomes caused by ultra-heat treatment was observed previously (67). Ultrasonication can also affect exosome morphology and cause substantial loss in RNA cargos. In milk, miR-29b and miR-200c are among the most abundant miRNAs. Howard et al. (66) reported that pasteurization and homogenization can cause miR-200c and miR-29b loss whereas heating in the microwave caused loss of miR-29b but not miR-200c. Furthermore, they can be degraded by adding detergent or bacterial fermentation (66). Surface protein removal from exosomes can decrease the exosome uptake by intestinal and vascular endothelial cells (68). Bacterial fermentation can degrade exosome membrane by attacking on exosome proteins, resulting in exosomal miRNA degradation by RNases in the environment (69).

Other than maintaining the stability of the inside components, the lipid membrane of the exosomes also has bioactivity similar the lipids found in other cellular membranes including cholesterol, phospholipids, and sphingolipids (70). To data, limited information on bovine milk exosomal lipids has been reported.

**FUNCTIONS**

**Intestinal Tract**

Milk exosomes play an important role in the development of the digestive tract. Many studies including in mouse models or in vitro cell cultures showed that bovine MDEs can enter the cytoplasm by endocytosis and then release their miRNA cargos across the basolateral membrane (59). MDEs promoted goblet cell expression indicated by increasing mucin production and trefoil factor 3 (TFF3) and mucin 2 (MUC2) (31). Chen et al. (71) observed that villus height and crypt depth of the duodenum and jejunum of mice were increased with daily administration of porcine MDEs. The possible reasons were attributed to increased expression of several transcription and proliferation factors (CDX2, IGF-1R) and reduced expression of protein p53 which regulate cell death (71). Similarly, rat milk exosomes were reported to stimulate intestinal epithelial cells' (IEC-18) viability, enhance proliferation by increasing expression of proliferating cell nuclear antigen (PCNA), and stimulate stem cell activity by increasing leucine-rich repeat-containing G-protein coupled receptor 5 (Lgr5) gene expression (6). The authors explained that MDEs travel within the intestinal epithelial cells and then can be taken up by stem cells thus enhance cell proliferation and stem cell activity. Milk exosomes can be taken up by human macrophages and vascular endothelial cells as well and it may cross the intestinal mucosa and biologically activate the response signal (32, 61). It was observed that IEC-6 cells uptake more yak MDEs than cow MDEs which may be due to the fact that the yak milk exosomes had significantly higher expression of membrane proteins than cow milk exosomes. Yak MDEs increased the survival rate of IEC-6 cells in hypoxic conditions by up to 29% whereas cow milk derived milk only had 22% (32). The possible reasons were attributed to increased expression of oxygen-sensitive propyl hydroxylase-1 (PHD-1) and decreased expression of hypoxia-inducible factor-α (HIF-α), its downstream target vascular endothelial growth factor (VEGF), and p53 (32).
Milk exosomes and their cargo miRNAs are absorbed in the upper intestine and accumulate mainly in the liver in mice (68). Many miRNAs are involved in gut health by exerting certain physiological functions (53). Inflammatory intestinal tissues had lower expression of miRNAs which are related to intestinal goblet cell differentiation and intestinal epithelium intact. Interestingly these miRNAs are highly present in MDEs (72). As components of milk exosomes, miR-148 and miR-155 can suppress intestinal T cells and may have a preventive effect on colorectal cancer carcinogenesis (73). MiR-200b can promote intestinal epithelial cell proliferation by inhibiting the epithelial-mesenchymal transition via TGF-β (74). MiR-146b can alleviate intestinal inflammation through the NF-κB pathway in a mouse colitis model and improve epithelia barrier function (75). Infant formula is miRNA-deficient and this may have negative effects on long-term immunological and metabolic programming of the infants (69). Short chain fatty acids (SCFA) in the gut are the major energy sources of intestinal epithelial cells and promote intestinal health by increasing epithelial absorption cells. Oral administration of MDEs increased SCFA production and number of epithelial absorption cells (26). The abundance of Lachnospiraceae and Ruminococcaceae were also increased as they are associated with the maintenance of gut health. Ross et al. (17) evaluated the effects of bovine colostrum and milk derived exosomes from high, average and low responder cows on human colorectal adenocarcinoma epithelial (Caco-2) cells. They found that co-incubation with exosomes maintain Caco-2 cells metabolic activity. MetaBio activity after incubation with exosomes from high responder cows was significantly greater than that from the low responder cows. It indicates that milk from cows with different immune response genetics might have different effects on gut health. Furthermore, the author pointed out that both colostrum and milk exosomes enhanced cell viability but did not stimulate oncogenic proliferation of Caco-2 cells in vitro.

Exosomes can inhibit the activation of toll-like receptor 4 (TLR4) which was involved in intestinal inflammation and progression of necrotizing enterocolitis (NEC) (76). In normal colonic epithelial cells, MDEs changed the cells from classic cuboid shape to a mesenchymal-like shape and contributed to cell proliferation but this was not observed in the tumor cells (28). The underlying mechanisms were attributed to the fact that MDEs upregulated the expression of collagen type I and downregulated twist1 gene expression and phospatase and tensin homolog (PTEN) protein in normal colonic epithelial cells but not in tumor cells (28). Exosomes gavage also prevented the necrotizing enterocolitis by preventing the ileal morphological injury and reduction in MUC2+ goblet cells and glucose-regulated protein 94 (GRP94+) cells per villus. MiR-200a-3p is a negative regulator of the pro-inflammatory chemokine ligand 9. It was observed that feeding diet depleted of milk exosomes in Mdr1a−/− mice can elicit depletion of miR-200a-3p, elevated cecal inflammation as well as chemokine ligand 9 expression (77). Mice fed regular diet had less intestinal lesions and lower score for gland hyperplasia and stromal collapse compared with the mice on the exosome-depleted diet (77). Martin et al. (33) pointed out that human breast MDEs can attenuate epithelial cell death from oxidative stress induced by H2O2 but not cycloheximide. However, the underlying mechanism is still not clear.

**Immune Function**

Widely spread among eukaryotes, miRNAs represent key components of a conserved system of RNA-based gene regulation. MiRNAs play important roles in the process of cellular proliferation and differentiation, tissue development and differentiation, and immune response (78). Chen et al. (5) identified 176 miRNAs out of 491 miRNAs target genes in transcription, immunity, and metabolism resources in porcine MDEs. The top 14 miRNAs participate in regulation of the IgA immune network and target about 20 immune-related genes such as CD40, CD80, MADCAM1, SLA, and among others. The immune-related miRNAs are similar in cow milk and breast milk (51). Compared to serum, all of the immune-related miRNAs such as miR-181a, miR-155, and miR-223 were abundantly expressed in milk, especially in colostrum, indicating that they may play a critical role in the biogenesis and development of immune system in infants (51). The higher immune-boosting effects of colostrum compared with mature milk may be correlated with higher levels of immune-related miRNAs and gene transcripts (52).

The milk derived EVs can influence the milk recipients’ immune system with the immune-regulatory miRNAs present (63). Exosomes affect intercellular communication through exosomal surface antigens with target cell receptors or via transferring exosomal RNAs and proteins to target cells (79). Under inflammatory condition, TGF-beta is required for inducing the pathogenic T-helper17 cells. TGF-beta on milk derived EVs can modulate T cell differentiation and play a role in the immune system (63). MDEs were reported to effectively alleviate the inflammatory response (increase in anti-inflammatory cytokine GM-CSF) caused by lipopolysaccharide (LPS). The lung and liver NF-κB levels were reduced by 30–40% suggesting the cargos inside exosomes such as immune factors, miRNAs and proteins could be exerting these protective effects (45). The expression levels of immune-related miRNAs in the first 6 months of breast milk are high (62). Breast MDEs were reported to increase the number of Foxp3+, CD4+ CD25+ regulatory T cells in infants (52) and induce B-cell differentiation (62). Milk exosomes can effectively prevent allergy of infants and they are critical for the maturation of the immune system during early infancy (52, 73). Bovine milk miRNAs can affect gene expression in peripheral blood mononuclear cells in human volunteers (68). Oral gavage of MDEs increased gene expression of Muc2, GATA4, RegIIγ, and MyD88 (gene linking gut microbiota and intestinal immunity) which are all related to intestinal immunity (26). Hata et al. (4) reported that RNAs from bovine milk-derived microvesicles might involve in the development of calf’s gastrointestinal and immune systems after being transferred to living cells. The authors observed that acid treatment of milk did not drain the miRNAs, so it is possible that the miRNAs can reach innate and acquired immune cells in gut-associated lymphoid tissues of suckling calves. Izumi et al. (61) observed that bovine milk exosomes were incorporated into differentiated human monocytic leukemia THP-1 cells by using...
flow cytometry and fluorescent microarray techniques and the results indicated that the exosomes might affect human cells through the RNA contents. In addition, cow milk exosomes modulate immunity-related disease possibly by the methylation of cells through miRNA transfer (80, 81). Naqvi et al. (82) demonstrated that higher content of miRNA-30b can inhibit phagocytosis in myeloid inflammatory cells.

Others Functions
Other than effects on intestinal and immune functions widely studied, milk exosomes also have beneficial effects on other areas. In human cells and in mice, bacteria invasion can induce the ADAM10 bearing exosome secretion and the exosomes can serve as decoys to bind bacterially produced toxins thus to protect the host cells. Keller et al. (83) demonstrated that ATG16L1 and other ATG proteins can provide protection against α-toxin through exosomes by releasing ADAM10. Arntz et al. (84) reported that oral delivery of bovine milk derived extracellular vesicles can ameliorate experimental arthritis in IL-1 receptor antagonist−/− and DBA1/J mouse models. Skeletal muscle growth and development can also be regulated by milk exosomes. Milk exosomal miRNAs such as miR21 and miR29a can enhance muscle protein synthesis by amplifying mTOR signaling pathway (85). Spatial learning and memory depends on purinergic receptor signaling (87), Manca et al. (34) observed that exosome and their cargos can also accumulate in the brain which may explain why that dietary depletion of milk exosomes can impair spatial learning and memory due to the aberrant metabolism of purines. Furthermore, in bones, bovine milk exosomes were observed to increase osteocyte number and woven bone formation, promoting osteoblast differentiation in mice (88). Reif et al. (28) reported dual effects of miR-148a on different tumor cells: inhibiting cell proliferation in hepatocellular carcinoma and esophageal cancer whereas promoting cell growths in glioblastoma. Further studies are needed to explore the mechanisms of MDEs on different cells.

APPLICATIONS OF MILK DERIVED EXOSOMES

Drug Carriers
Many factors such as high cost, difficulty in production in sufficient quantities, and toxicity/intolerance issues have prevented the clinical introduction of many natural and synthetic materials. As ideal nanoparticles, it should have attributes including long circulation time, evasion of the host immune system, precise targeting for specific cells, minimal off site toxicity, and ability to carry versatile therapeutics (89). The application of MDEs is very intriguing and they are being explored as nanodevices for the development of new chemotherapeutic/chemopreventive carriers. Compared to other synthetic carriers, nature-derived nanoparticles such as exosomes from milk have more advantages: (1) well-tolerated in the body as it exists in various biological fluids and it exhibited cross-species tolerance with no adverse immune and inflammatory response; (2) longer circulating half-life; (3) can be internalized by other cells (90). Somiya et al. (91) demonstrated that administration of MDEs did not result in systemic toxicity and serial administration did not cause any anaphylaxis effect. Curcumin encapsulated in milk exosomes was found to resist human digestion and possess enhanced endothelial permeability due to its elevated stability, solubility as well as bioavailability (37). Another important trait for MDEs is that it can be produced in bulk with the rich source of milk. In addition, long storage had no significant changes in the physical and biological properties of milk-derived exosomes. Bovine milk exosomes are bioavailable and distribute widely among murine tissues, accumulating mainly in the liver, and spleen. Betker et al. (29) suggested that milk exosomes are absorbed from the gut as intact particles via the neonatal Fc receptor and the intact particles can be modified with ligands to retain in target tissues. For those unstable or poorly bioavailable drugs, MDEs are considered scalable vehicles to deliver them effectively. To use milk exosomes as a vehicle for drug, identifying the target tissues is important. Three hours after intravenous injection, exosome concentrations in liver and spleen peaked and then slightly decreased. In contrast, by oral gavage, the exosome signal in the liver peaked 24 h but no signal after 48 h (34).

Administration of bovine MDEs as systemic drug delivery were successful on tumor bearing mice (35). The authors found that paclitaxel loaded with exosomes inhibited tumor growth by 60% whereas paclitaxel itself only had 31%. The possible reason could be attributed to the sustained release of the drugs and the remarkably lower systemic and immunologic toxicities with the exosomes. Zhang et al. (25) developed a new milk exosome based pH/light sensitive drug delivery system and the system can control the drug release and are proved to be effective in treating oral squamous cell carcinomas (OSCC). With a bi-lipid membrane and an aqueous core, milk exosomes can be potentially used as carriers for both hydrophilic and lipophilic drugs (92). Munagala et al. (45) demonstrated that milk-derived exosomes can deliver both hydrophilic and lipophilic small molecules such as chemo drugs. In cell culture studies against lung tumor and breast cancer, drug-loaded exosomes showed significantly higher efficacy than free drug. The possible reason could be that drug loaded in exosomes had increased stability and higher cellular uptake. Furthermore, milk exosomes had no adverse immune and inflammatory response and the author suggests it can used as a biocompatible and cost effective tool to enhance oral bioavailability and improve efficacy and safety of drugs (45). Folic acid (FA) and other vitamin receptors have been extensively explored to achieve tumor targeting because normally they are overexpressed in many cancers. Milk exosomes can functionalize with tumor targeting ligand to further improve specificity and eliminate off-target side effects of drugs on healthy cells (45).

Markers of Disease
Mastitis in dairy industry, either clinical or subclinical, has caused big economic loss due to the restraint sale of the milk from mastitis cows. Mastitis is usually caused by microbial infection, such as *staphylococci*, *streptococci*, and *coli*form bacteria (93).
Sun et al. (56) reported higher levels of bta-miR-142a and bta-miR-223 in the milk from *S. aureus* challenged cows compared to the control cows, suggesting that these two miRNAs can be used as biomarkers of bacterial infection. Cai et al. (94) analyzed milk exosomes from three healthy and three mastitis cows and identified 18 miRNAs differently expressed between the two groups. Among the 14 differentially expressed miRNAs, nine were upregulated in the cows with mastitis (miR-142-5p, miR-142-3p, miR-103, miR-147, miR-23a, miR-223, miR-146a, miR-146b, and miR-221). The authors indicated that the differently expressed miRNAs, especially miR-223 and miR-142-5p can be used as potential marker candidates for mastitis. Chen et al. (51) suggested using seven milk-associated miRNAs (miR-26a, miR-26b, miR-200c, miR-21, miR-30d, miR-99a, and miR-2148a) as potential biomarkers for the quality control of raw milk and other milk-related products. A potential biomarker for quality control in bovine milk and human milk, miR-148a, was found to be highly expressed in Yorkshire sows (51) but a moderately in Landrace pigs (5). In human, milk derived miRNAs were suggested to be used as markers to identify the novel mechanisms involved in genetic variation for breast function such as SLC20A2 (95). MiR-21, known as an oncomiR, is related to many kinds of cancer such as malignant melanoma, prostate cancer, and hepatocellular carcinoma (96, 97). The miRNAs, especially the oncomiRs deserve more investigation in diseases studies.

**FUTURE RESEARCH**

Milk exosomes could also be explored to load and deliver potentially other macromolecules such as siRNA, miRNAs, plasmid DNA, cDNA, and proteins (antioxidant enzymes, etc). Additionally, the protective effects of milk exosomes per se are very intriguing and suggest utility of these nanovesicles against many inflammation-based diseases. For example, exosomes from bovine milk and colostrum could be exploited as additives in formula milk and thus potentially serve as immune booster in infants and could also be used for immune-compromised cancer patients undergoing chemotherapy. Further, tissue targeting or site-specific delivery of drug loaded exosomes can be explored by adding a wide variety of tumor-targeting ligands such as antibodies (e.g., VEGF, EGFR), peptides (e.g., transferrin, integrins, Her2), or receptor-targets (e.g., FA, biotin and hyaluronic acid) to the milk exosomes.

The gut microbiota can be affected by dietary interventions and composition of microbiota change as diet changes (98). By oral gavage it was observed that 75% of exosomes not absorbed in the upper intestine enter the large intestine and cecum (34). It is noteworthy that exosomes amount entering the digestive tract can largely affect absorption as well as their interaction with gut microbiota. Understanding the biological effects of milk exosomes and their cargos on gut microbes is also intriguing and studies on effects of exosome on gut microbiota and their interactions are scarce. Oral administration of MDEs could modulate the host gut microbiota (26). Exosomes and their cargos participate in the crosstalk between gut microbes and host by altering microbial communities. A growth advantage of the bacteria cultured with milk exosome was observed *in vitro* (99). The gut microbes might act as transmitters or amplifiers of dietary exosome signals because changes in the microbial communities can cause changes in the production of microbial metabolites. Exosomes supplementation also affected hepatic concentrations of purine metabolites as well as muscle grip strength and how these phenotypes depending on the gut microbial, indirectly by exosomes, is interesting.

Genetic selection of high performance and pregnancy-dependent E2 production cause enrichment of miR-148a and miR-21. Reif et al. (28) reported upregulation of miR-148a can inhibit tumor cell proliferation. To the contrary of most reported findings, Melnik and Schmitz (73) pointed out that continuous intake of milk exosomes may pose a risk for chronic diseases including obesity, type 2 diabetes mellitus, osteoporosis, Parkinson’s disease, and common cancers, mainly due to the miRNAs inside the exosomes: such as miR-148a which suppress inhibitor of adipogenesis, miR-29b which belongs the diabetogenic miR family, miR-155 which can promote the initiation and progression of Parkinson’s disease in humans, and miR-21 which promotes tumor progression. The miRNA-21, which was observed in milk exosomes, can enhance mTORC1-driven metabolic processes by attenuating the inhibitory effects of various tumor suppressor proteins on mTORC1-signaling (85, 86). The authors mentioned that as human breast milk is the ideal food for infant, persistent high cow milk signaling during adolescence and adulthood may promote diseases of civilization. In that paper, most of the finding are based on outcomes from consuming milk not on exosomes only, thus the negative effects of MDEs need to be further investigated.

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REFERENCES

1. Zaborowski MP, Balal J, Breakefield XO, Lai CPK. Extracellular vesicles: composition, biological relevance, and methods of study. Bioessence. (2015) 65:783–97. doi: 10.1016/j.bioessence.2015.01.008

2. van Dommelen SM, Vader F, Lakhal S, Kooijmans SA, van Solinge WW, Wood MJ, et al. Microvesicles and exosomes: opportunities for cell-derived membrane vesicles in drug delivery. J Control Release. (2012) 161:635–44. doi: 10.1016/j.jconrel.2011.11.021

3. Zempler J, Lozano A, Sadr M, Sukreet S, Manca S. Biological activities of extracellular vesicles and their cargos from bovine and human milk in humans and implications for infants. J Nutr. (2017) 147:3–10. doi: 10.3945/jn.116.238949

4. Hata T, Murakami K, Nakatani H, Yamamoto Y, Matsuda T, Aoki N. Isolation of bovine milk-derived microvesicles carrying mRNAs and microRNAs. Biochem Biophys Res Commun. (2010) 396:528–33. doi: 10.1016/j.bbrc.2010.04.135

5. Chen T, Xi QY, Ye RS, Cheng X, Qi QE, Wang SB, et al. Exploration of microRNAs in porcine milk exosomes. BMC Genom. (2014) 15:100. doi: 10.1186/1471-2164-15-100

6. Hock A, Miyake H, Li B, Lee C, Ermini L, Koike Y, et al. Breast milk-derived exosomes maintain intestinal epithelial barrier integrity. J Pediatr Surg. (2017) 52:753–9. doi: 10.1016/j.jpedsurg.2017.01.032

7. Chen Z, Luo J, Sun S, Cao D, Shi H, Lour JJ. miR-148a and miR-17-5p synergistically regulate milk TAG synthesis via PPARGC1A and PPARA in goat mammary epithelial cells. RNA Biol. (2017) 14:326–38. doi: 10.8002/1547-6986.2016.1276149

8. Modepalli V, Kumar A, Hinds LA, Sharp JA, Nicholas KR, Lefevre C. Differential temporal expression of milk miRNA during the lactation cycle of the marsupial tammar wallaby (Macropus eugenii). BMC Genomics. (2014) 15:1012. doi: 10.1186/1471-2164-15-1012

9. Mirza AH, Kaur S, Niels IB, Stirling I, Yaran R, Roursagard M, et al. Breast milk-derived extracellular vesicles enriched in exosomes from mothers with type 1 diabetes contain aberrant levels of microRNAs. Front Immunol. (2019) 10:2543. doi: 10.3389/fimmu.2019.02543

10. Shah KB, Chernausek SD, Garman LD, Pezant NP, Plows JF, Kharoudi M, et al. Comparative proteome profiling in exosomes derived from porcine colostrum and caprine colostrum and mature milk. J Anim Sci. (2015) 63:593–602. doi: 10.1109/jast.2021.139.2021.e39

11. Kandimalla R, Aqil F, Alahakeem SS, Jeyabalan J, Tyagi N, Agrawal A, et al. Targeted oral delivery of paclitaxel using colostrum-derived exosomes. Cancers. (2021) 13:3700. doi: 10.3390/cancers13133700

12. Shah KB, Chernausk SD, Garman LD, Pezant NP, Plows JF, Kharoudi M, et al. Comparative proteome profiling in exosomes derived from porcine colostrum and caprine colostrum and mature milk. J Proteomics. (2021) 249:104338. doi: 10.1016/j.jprot.2021.104338

13. Yun B, Kim Y, Park DJ, Oh S. Comparative analysis of dietary exosome-derived microRNAs from human, bovine and caprine colostrum and mature milk. J Anim Sci. (2015) 63:593–602. doi: 10.1109/jast.2021.139.2021.e39

14. Del Pozo-Acebo L, Hazas MLL, Tomé-Carneiro J, Gil-Cabrerizo P, Yanes P, M泉州 SJ, et al. Bovine milk-derived exosomes from colostrum are enriched with microRNAs from mothers with different biological effect on normal fetal colon epithelial cells compared to colon tumor cells in a miRNA-dependent manner. J Transl Med. (2019) 17:325. doi: 10.1186/s12967-019-2072-3

15. Wang L, Wang X, Shi Z, Shen L, Zhang J, Zhang J. Bovine milk-derived exosomes attenuate the alteration of purine metabolism and energy status in IEC-6 cells induced by hydrogen peroxide. Biochem Biophys Res Commun. (2021) 52:755–9. doi: 10.1016/j.bbrc.2021.05.065

16. Luo S, Sun X, Huang M, Ma Q, Du L, Cui Y. Enhanced neuroprotective effects of milk exosomes with enhanced mucus penetrability for oral delivery of siRNA. Biosci. Biotechnol Biochem. (2021) 85:97–105. doi: 10.1080/09168451.2020.1840568

17. Ross M, Atalla H, Karrow N, Mallard BA. The bioactivity of colostrum and milk exosomes of high, average, and low immune responder cows on human intestinal epithelial cells. J Dairy Sci. (2021) 104:2499–510. doi: 10.3168/jds.2020-17489

18. Gao HN, Hu H, Wen PC, Lian S, Xie XL, Song HL, et al. Yak milk-derived exosomes alleviates lipopolysaccharide-induced intestinal inflammation by inhibiting PI3K/AKT/C3 pathway activation. J Dairy Sci. (2021) 104:8411–24.
39. Izumi H, Kosaka N, Shimizu T, Sekine K, Ochiya T, Takase M. Bovine milk contains microRNA and messenger RNA that are stable under degradative conditions. *J Dairy Sci.* (2012) 95:4831–41. doi: 10.3168/jds.2012-5348

40. Yamada T, Inoshima Y, Matsuda T, Ishiguro N. Comparison of methods for isolating exosomes from bovine milk. *J Vet Med Sci.* (2012) 74:1523–5. doi: 10.1292/jvms.2011-9251

41. Cheruvanky A, Zhou H, Pisitkun T, Kopp JB, Knepper MA, Yuen PST, et al. A subset of extracellular vesicles expresses immunoregulatory TGF-beta. *Proteomics* (2011) 11:2017–24. doi: 10.1002/pmic.2010000836

42. van Herwijnen MJC, Driedonks TAP, Snoek BL, Kroon AMT, Kleijnaja, et al. Exosomes contain miRNA and miRNA and mRNA and are taken up by human macrophages. *J Dairy Sci.* (2015) 98:2920–33. doi: 10.3168/jds.2014-9076

43. Lin D, Chen T, Xie M, Li M, Zeng B, Sun R, et al. Oral administration of exosomes secreted by hepatocytes. *J Proteome Res.* (2010) 10:7303–13. doi: 10.1021/pr100293a

44. Admyre C, Johansson SM, Qazi KR, Filen JJ, Lahesmaa R, Norman M, et al. Oral administration of exosomes for genetic information transfer and gene therapy. *Hum Mol Genet.* (2012) 21:R125–34. doi: 10.1093/hmg/ddr317

45. Baietti MF, Zhang Z, Mortier E, Melchior A, Degeest G, Geeraerts A, et al. Syndecan-syntenin-ALIX regulates the biogenesis of exosomes. *Nat Cell Biol.* (2012) 14:677–85. doi: 10.1038/ncb2502

46. Mathivanan S, Ji H, Simpson RJ. Exosomes: extracellular organelles important in intercellular communication. *J Proteomics.* (2010) 73:1907–20. doi: 10.1016/j.jprot.2010.06.006

47. Tili E, Michaille JJ, Calin GA. Expression and function of micro-RNAs in their role in protein trafficking and biomarker potential in Alzheimer's disease. *Int J Mol Sci.* (2016) 17:173. doi: 10.3390/ijms17020173

48. Rahman MM, Takashima S, Kamatari YO, Badr Y, Kitamura Y, Shimizu K, et al. Proteomic profiling of milk small extracellular vesicles from bovine leukemia virus-infected cattle. *Sci Rep.* (2011) 1:2291. doi: 10.1038/srep00229

49. Leiferman A, Shu J, Upadhyaya B, Cui J, Zempleni J. Storage of extracellular vesicles in human milk, and miRNA profiles in human milk exosomes and infant formulas. *Pediatr Gastroenterol Nutr.* (2019) 69:235–8. doi: 10.1097/MPG.0000000000002363

50. Howard KM, Jati Kusuma R, Baier SR, Friemel T, Markham L, Vanamala J, et al. Loss of miRNAs during processing and storing of cow’s (Bos taurus) milk. *J Agric Food Chem.* (2015) 63:588–92. doi: 10.1021/jf505526w

51. Kirchner B, Pfaffl MW, Dummer J, von Mutius E, Ege MJ. microRNA in native and processed cow's milk and its implication for the farm milk effect on asthma. *J Allergy Clin Immunol.* (2016) 137:1893–8. doi: 10.1016/j.jaci.2015.10.028

52. Baire SR, Nguyen C, Xie F, Wood JR, Zempleni J. microRNAs are absorbed in biologically meaningful amounts from nutritionally relevant doses of cow milk and affect gene expression in peripheral blood mononuclear cells, HEK-293 kidney cell cultures, mouse livers. *J Nutr.* (2014) 144:1495–500. doi: 10.3945/jn.114.196436

53. Yu S, Zhao Z, Sun L, Li P. Fermentation results in quantitative changes in milk-derived exosomes and different effects on cell growth and survival. *J Agric Food Chem.* (2017) 65:12208–8. doi: 10.1021/acs.jafc.6b05002

54. Ortega-Anaya J, Jimenez-Flores R. Symposium review: the relevance of bovine milk phospholipids in human nutrition: Evidence of the effect on infant gut and brain development. *J Dairy Sci.* (2019) 102:2738–48. doi: 10.3168/jds.2018-15342

55. Chen T, Xie MY, Sun J, Ye RS, Cheng X, Sun RP, et al. Porcine milk-derived exosomes promote proliferation of intestinal epithelial cells. *J Proteome Res.* (2016) 15:363862. doi: 10.1021/acs.jprot.6b03862

56. Ng PC, Chan KY, Leung KT, Tam YH, Lam HS, et al. Comparative MiRNA expression profiles and molecular networks in human small bowel tissues of necrotizing enterocolitis and spontaneous intestinal perforation. *PLoS ONE.* (2015) 10:e0135737. doi: 10.1371/journal.pone.0135737

57. Melinck BC, Schmitz G. Exosomes of pasteurized milk: potential pathogens of Western diseases. *J Transl Med.* (2019) 17:3. doi: 10.1186/s12967-018-1760-8

58. Chen Y, Xiao Y, Ge W, Zhou K, Wen J, Yan W, et al. miR-200b inhibits TGF-beta-induced epithelial-mesenchymal transition and promotes growth of intestinal epithelial cells. *Cell Death Dis.* (2019) 10:e34541. doi: 10.1038/sddx.2013.22

59. Kata T, Fujiya M, Ueno N, Morichi K, Konishi H, Tanabe H, et al. microRNA-146b improves intestinal injury in mouse colitis by activating nuclear factor-kappaB and improving epithelial barrier function. *J Gene Med.* (2013) 15:249–60. doi: 10.1002/jgm.2717

60. Good M, Sodhi CP, Egän CE. Breast milk protects against the development of necrotizing enterocolitis through inhibition of toll-like receptor 4 in the intestinal epithelium via activation of the epidermal growth factor receptor. *Microb Pathog.* (2015) 81:166–79. doi: 10.1016/j.micpath.2015.05.002

61. Wu D, Kittana H, Chu J, Kachman SD, Cui J, Ramer-Tait AE, et al. Dietary Depletion of Milk Exosomes and Their MicroRNA Carriers Elicits a Depletion of miR-200a-3p and Elevated Inflammatory Inflammation and Parkinson's Disease. *J Mol Sci.* (2016) 17:173. doi: 10.3390/jms17020173
Chemokine (C-X-C Motif) Ligand 9 Expression in Mdr1a<sup>−/−</sup> Mice. *Curr Dev Nutr.* (2019) 3:n2212. doi: 10.1093/cdn/nzz122

87. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell.* (2009) 136:215–33. doi: 10.1016/j.cell.2009.01.002

88. Camussi G, Deregibus MC, Bruno S, Cantaluppi V, Biancone L. Exosomes/microvesicles as a mechanism of cell-to-cell communication. *Kidney Int.* (2010) 78:838–48. doi: 10.1038/ki.2010.278

89. Benmoussa A, Gotti C, Bourassa S, Gilbert C, Provost P. Identification of protein markers for extracellular vesicle (EV) subsets in cow's milk. *J Proteomics.* (2019) 192:78–88. doi: 10.1016/j.jprot.2018.08.010

90. Melnik BC, Schmitz G. MicroRNAs: Milk's epigenetic regulators. *Best Pract Res Clin Endocrinol Metab.* (2017) 31:427–42. doi: 10.1016/j.beem.2017.10.003

91. Naqui AR, Fordham JB, Nares S. miR-24, miR-30b, and miR-142-3p regulate phagocytosis in myeloid inflammatory cells. *J Immunol.* (2015) 194:1916–27. doi: 10.4049/jimmunol.1401893

92. Keller MD, Ching KL, Liang FX, Dhabaria A, Tam K, Ueberheide BM, et al. Decoy exosomes provide protection against bacterial toxins. *Nature.* (2020) 579:260–4. doi: 10.1038/s41586-020-2066-6

93. Arntz OJ, Pieters BC, Oliveira MC, Broeren MG, Bennink MB, de Vries M, et al. Oral administration of bovine milk derived extracellular vesicles attenuates arthritis in two mouse models. *Mol Nutr Food Res.* (2015) 59:1701–12. doi: 10.1002/mnfr.201500222

94. Melnik BC, John SM, Schmitz G. Milk is not just food but most likely a genetic transfection system activating mTORC1 signaling for postnatal growth. *Nutr J.* (2013) 12:103. doi: 10.1186/1475-2891-12-103

95. Lu Y, Zheng Z, Yuan Y, Pathak JL, Yang X, Wang L, et al. The emerging role of exosomes in oral squamous cell carcinoma. *Front Cell Dev Biol.* (2021) 9:628103. doi: 10.3389/fcell.2021.628103

96. Duster R, Prickaerts J, Blokland A. Purinergic signaling and hippocampal long-term potentiation. *Curr Neuropharmacol.* (2014) 12:37–43. doi: 10.2174/1570159X113119990045

97. Oliveira MC, Arntz OJ, Blaney Davidson EN, van Lent PL, Koenders MI, van der Kraan PM, et al. Milk extracellular vesicles accelerate osteoblastogenesis but impair bone matrix formation. *J Nutr Biochem.* (2016) 30:74–84. doi: 10.1016/j.jnutbio.2015.11.017

98. Lai RC, Yeo RW, Tan KH, Lim SK. Exosomes for drug delivery - a novel application for the mesenchymal stem cell. *Biotechnol Adv.* (2013) 31:543–51. doi: 10.1016/j.biotechadv.2012.08.008

99. Sun D, Zhuang X, Xiang X, Liu Y, Zhang S, Liu C, et al. A novel nanoparticle drug delivery system: the anti-inflammatory activity of curcumin is enhanced when encapsulated in exosomes. *Mol Ther.* (2010) 18:1606–14. doi: 10.1038/mt.2010.105

100. Somiya M, Yoshioka Y, Ochiya T. Biocompatibility of highly purified bovine milk-derived extracellular vesicles. *Extracell Vesicles.* (2018) 7:1440132. doi: 10.1080/20013078.2018.1440132

101. Vlassov AV, Magdaleno S, Setterquist R, Conrad R. Exosomes: current knowledge of their composition, biological functions, and diagnostic and therapeutic potentials. *Biochim Biophys Acta.* (2012) 1820:940–8. doi: 10.1016/j.bbagrm.2012.03.017

Conflict of Interest: XZ was employed by company Foshan Nanhai Poultry Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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