Evaluation of a Bovine Rapid Visual PAG ELISA Test and Transabdominal Ultrasonography for Early Pregnancy Diagnosis in Awassi Sheep

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ABSTRACT
The aim of this study was to evaluate two different methods (a bovine rapid visual PAG ELISA test and transabdominal ultrasonography) for early diagnosis of pregnancy in Awassi sheep. The ewes used in this study (n=50) were hand-mated in June of 2019 and lambed in October of 2019. Pregnancy diagnosis was performed 34–38 days after mating by rapid visual PAG ELISA test and transabdominal ultrasonography in standing position. The diagnostic test characteristics of both methods were calculated using lambing results as a gold standard. According to the lambing results, it was determined that, of the 50 ewes, 42 (84%) were pregnant and 8 (16%) were not. The sensitivity, specificity, positive predictive value, negative predictive value and accuracy of rapid visual PAG ELISA and transabdominal ultrasonography were 97.6%, 62.5%, 93.2%, 83.3%, 92% and 95.2%, 100%, 100%, 80%, 96%, respectively. In McNemar’s analysis, no difference was found between the pregnancy diagnosis methods and lambing results. In conclusion, both rapid visual PAG ELISA and transabdominal ultrasonography in standing position proved to be reliable and practical methods for pregnancy diagnosis as of day 34 post-mating in Awassi ewes.

Keywords
Pregnancy diagnosis
Pregnancy–associated glycoproteins
Transabdominal ultrasound
Awassi sheep

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INTRODUCTION

The sheep industry stands out as a promising economic sector. Sheep breeding has an important place in Turkey's economy in terms of meat, milk, wool and leather production. The milk yield of Awassi sheep, which is one of the highest milk-producing sheep breeds native to Turkey, can reach up to 250–300 kg in breeder flocks (Taşan, 2016). The fattening performance of male lambs can vary from 35 kg to 42 kg depending on the rearing period and system (Kul and Akan, 2002; Tekel et al., 2007; Şireli and Tekel, 2013). Recent studies have shown that while consumer demand for different types of meat is changing, sheep meat consumption is increasing worldwide (Kandemir et al., 2019). Especially the increased demand for lamb meat requires increased reproductive performance. The early diagnosis of pregnancy in sheep enables strategic interventions in herd management, including the nutrition, culling or separation of sheep. Non-pregnant ewes can be either re-bred in the breeding season, fed only for wool and leather production or culled to reduce feeding costs. Non-pregnant lambs can be marketed at a higher price before transitioning into adulthood. Furthermore, the involuntary culling of pregnant ewes can be prevented. The separation of the flock into pregnant and non-pregnant groups might reduce productive and reproductive losses caused by abortions, stillbirths and birth of weak lambs (Karen et al., 2001; Ulusoy and Kaymaz, 2009; Chaves et al., 2017).

Several methods can be used for pregnancy diagnosis in sheep, and transabdominal ultrasonography is one of the most common. Transabdominal ultrasonography can be used with a sensitivity of 100% as from the 39th day of pregnancy (Jones et al., 2016), and with an accuracy of 100% as from the 40th day of pregnancy (Aziz and Lazim, 2012). Despite its high efficiency in the diagnosis of pregnancy in sheep, ultrasonography also has some practical limitations related to the facilities required for examination, positioning of ewes for examination, availability and portability of equipment, type and frequency of transducer, and cost of equipment (Roberts et al., 2019).

Pregnancy-associated glycoproteins (PAGs) are produced in the fetomaternal interface of the ruminant placenta and released into the intercotyledony space during pregnancy (Sousa et al., 2006). The presence of PAGs in the maternal circulation allows for an early diagnosis of pregnancy in cows (Akköse et al., 2019; Akköse and Çebi Şen, 2019), sheep (Uçar et al., 2018; Alkan et al., 2020) and goats (Gonzalez et al., 1999; Tandiya et al., 2013). Enzyme-linked immunosorbent assay (ELISA) tests, which can be used in the field, have been developed to detect PAGs in the maternal circulation (serum, plasma or whole blood samples) and milk of cattle (BioPRYN, Idexx Bovine Pregnancy Test, Idexx Visual Pregnancy Test, Idexx Milk Pregnancy Test, Idexx Rapid Visual Pregnancy Test). While Xie et al. (1991) detected immunological and structural similarities between ovine and bovine PAG molecules, it has also been reported that bovine and ovine PAGs can exhibit spatially and temporally distinct expression patterns during pregnancy at species or breed level (Green et al., 2000; Lobago et al., 2009; Mercadante et al., 2013). PAG ELISA pregnancy tests commercially validated for cattle have been used in various Brazilian and European sheep breeds (Rovani et al., 2016; Chaves et al., 2017; Steckeler et al., 2018). It is stated that PAG profiles in the peripheral blood can vary between sheep breeds; therefore, determining the specific PAG profile of breeds native to Turkey is important for improved reproductive management (Kaplan et al., 2019). The aim of this study was to evaluate the diagnostic reliability of a bovine rapid visual PAG ELISA test and transabdominal ultrasonography in standing position for early pregnancy diagnosis in Awassi sheep using lambing results as a gold standard.

MATERIAL and METHOD

Material

Animal Management

This study was approved by the Local Ethics Committee for Animal Experiments of Harran University (Protocol no: 2019/006/01–05).

The trial was conducted from June to December 2019, in the Sanliurfa province located in the Southeaster Anatolia Region of Turkey, using Awassi sheep. Fifty Awassi ewes, aged 2–5 years and weighing 40–60 kg, were used in this study. All animals were maintained under the same environmental and nutritional conditions. All ewes had access to pasture (wheat stubble) during the summer and were fed additional concentrate.

Estrus detection was performed twice a day using teaser rams, and ewes in heat were hand-mated. Ewes, to be mated by certain rams, were listed beforehand. The mating and lambing records of all ewes were maintained in the herd management system of the enterprise.

Method

Blood Collection and Rapid Visual PAG ELISA

Blood samples were collected by jugular venipuncture into 10-mL dry vacutainer tubes (Hematube). Transabdominal ultrasonography was performed after blood collection. Sera were separated 1–2 hours after blood collection at room temperature.

Rapid visual PAG ELISA (Idexx rapid visual pregnancy test) was performed in the field, according to the manufacturer’s instructions. Briefly, 100 µL serum and positive and negative controls of assay were added into anti-PAG coated wells. Next, the detector
solution was added to each well and incubated for 7 minutes at room temperature. Following incubation, the wells were washed 3 times with distilled water. Subsequently, the conjugate solution was added to all wells and incubated for 7 minutes at room temperature. After incubation, the wells were washed 3 times with distilled water. Later, the substrate solution was added to all wells and allowed to incubate for 7 minutes at room temperature. At the end of the incubation period, a stop solution was added to all wells, and the test wells were compared to the positive and negative controls for colour change. Rapid visual PAG ELISA results are interpreted based on the colour reaction in the wells. A positive well is blue, while a negative well is transparent. If the colour of the test well is bluer than that of the negative control, the result is positive (pregnant). However, if the colour of the test well is the same with that of the negative control or more transparent, the result is negative (non-pregnant).

**Transabdominal Ultrasonography**

For pregnancy diagnosis, right inguinal transabdominal ultrasonography was performed in all ewes in standing position between days 34–38 post-mating using an ultrasound device (Hasvet 838). Pregnancy was diagnosed based on the observation of a fluid-filled uterus and placenta or a fetus. All ultrasonographic examinations were performed by the same veterinarian.

**Statistical Analysis**

Lambing results were used as a gold standard to test the accuracy of both methods in pregnancy diagnosis. For the prediction of the diagnostic characteristics of rapid visual PAG ELISA and transabdominal ultrasonography, data were classified as follows: correct negative (a), false positive (b), false negative (c) and correct positive (d) results. Subsequently, sensitivity \( [d/(d+c) \times 100] \), specificity \( [a/(a+b) \times 100] \), positive predictive value (PPV) \( [d/(d+b) \times 100] \), negative predictive value (NPV) \( [a/(a+c) \times 100] \), and accuracy \( [(a+d)/(a+b+d+c) \times 100] \) were calculated for each pregnancy diagnosis method. Kappa analysis was performed to determine the agreement of rapid visual PAG ELISA and transabdominal ultrasonography with the lambing results. McNemar’s test and the comparison of the receiver operating characteristic (ROC) curves were used to determine whether there was any significant difference between the results of the two pregnancy diagnosis methods and lambing results (Yağanoğlu and Topal, 2018; Roberts et al., 2019). The Kappa coefficients were classified as indicating a poor \( (< 0.20) \), fair \( (0.21–0.40) \), moderate \( (0.41–0.60) \), good \( (0.61–0.80) \) and very good \( (0.81–1.00) \) agreement (Altman, 1991). Statistical calculations were performed using Statistical Package for Social Sciences (SPSS) version 24.0 at significance level of \( p<0.05 \).

**RESULTS and DISCUSSION**

Out of the 50 ewes used in the present study, 42 lambed and 8 did not lamb. To evaluate the pregnancy outcomes of each pregnancy diagnosis method in ewes, a 2x2 contingency table was constructed (Table 1) and test characteristics were calculated for transabdominal ultrasonography and rapid visual PAG ELISA (Table 2). While 2 ewes were diagnosed false negative with transabdominal ultrasonography, 3 ewes were diagnosed false positive and 1 ewe was diagnosed false negative with rapid visual PAG ELISA for pregnancy.

| Lambing records (Kuzulama sonuçları) | Rapid Visual PAG ELISA (Hızlı Görsel PAG ELISA) | Transabdominal Ultrasonography (Transabdominal Ultrasonografi) |
|-------------------------------------|-----------------------------------------------|-------------------------------------------------|
|                                    | Non-pregnant (Gebe değil) | Pregnant (Gebe) | Total (Toplam) | Non-pregnant (Gebe değil) | Pregnant (Gebe) | Total (Toplam) |
| Did Not Lambed (Kuzulamadı)        | 5 (cn) | 3 (fp) | 8 | 8 (cn) | 0 (fp) | 8 |
| Lambed (Kuzuladı)                  | 1 (fn) | 41 (cp) | 42 | 2 (fn) | 40 (cp) | 42 |
| Total (Toplam)                     | 6 | 44 | 50 | 10 | 40 | 50 |

**Table 1. A 2x2 contingency table for rapid visual PAG ELISA and transabdominal ultrasonography.**

| Lambing records (Kuzulama sonuçları) | Rapid Visual PAG ELISA (Hızlı Görsel PAG ELISA) | Transabdominal Ultrasonography (Transabdominal Ultrasonografi) |
|-------------------------------------|-----------------------------------------------|-------------------------------------------------|
|                                    | Non-pregnant (Gebe değil) | Pregnant (Gebe) | Total (Toplam) | Non-pregnant (Gebe değil) | Pregnant (Gebe) | Total (Toplam) |
| Did Not Lambed (Kuzulamadı)        | 5 (cn) | 3 (fp) | 8 | 8 (cn) | 0 (fp) | 8 |
| Lambed (Kuzuladı)                  | 1 (fn) | 41 (cp) | 42 | 2 (fn) | 40 (cp) | 42 |
| Total (Toplam)                     | 6 | 44 | 50 | 10 | 40 | 50 |

In this study, pregnancy diagnosis was performed 34–38 days after mating. Upon determining that the uterus hung from the intrapelvic area to the abdominal cavity on the 32nd day of pregnancy in Akkaraman sheep, Küplülü et al. (2002) suggested the use of the transabdominal technique for the diagnosis of pregnancy by ultrasonad examination, as of day 32. The application of transabdominal ultrasonography in the sitting position has been reported to produce better results than the standing position for early pregnancy diagnosis (Gürler and Kaymaz, 2011). However, maintaining animals in a sitting position is time-consuming and laborious. In the present study, sheep were restrained in the
standing position by an employee, and the hairless area of the right inguinal region was scanned with a transducer for the diagnosis of pregnancy by ultrasound examination.

The sensitivity and specificity of transabdominal ultrasonography in standing position were determined to be 95.2% and 100%, respectively. Similarly, Jones et al. (2016) reported that, in sheep the sensitivity and specificity of this method were 83% and 100%, respectively, on day 30 post-insemination, and were both 100% on day 39 post-insemination. The positive and negative predictive values of transabdominal ultrasonography in standing position in the present study were 100% and 80%, respectively. The uterus could still be located in the pelvic cavity before the 40th day of pregnancy (Crilly et al., 2017). In the present study, the false negative results (two ewes were diagnosed false negative) were attributed to the uterus having not descended into the abdominal cavity at the time of pregnancy diagnosis. Aziz and Lazim (2012) reported that the accuracy of transabdominal ultrasonography increases from 80% between days 28–32 of pregnancy to 100% between days 40–43 days in Awassi sheep. In agreement with this report, the accuracy of transabdominal ultrasonography was determined to be 96% in the present study.

| Table 2. Diagnostic test characteristics of rapid visual PAG ELISA and transabdominal ultrasonography |
|-----------------------------------------------|-----------------------------------------------|
| Rapide goşel PAG ELISA (Hızlı Görsel PAG ELISA) | Transabdominal Ultrasonography (Transabdominal Ultrasonografi) |
| Sensitivity (%) (Duyarlılık (%)) | 97.6 | 95.2 |
| Specificity (%) (Özgürlük (%)) | 62.5 | 100 |
| Positive predictive value (%) (Pozitif prediktif değer (%)) | 93.2 | 100 |
| Negative predictive value (%) (Negatif prediktif değer (%)) | 83.3 | 80 |
| Accuracy (%) (Doğruluk (%)) | 92 | 96 |

The sensitivity determined for rapid visual PAG ELISA (97.6%) in the present study was in agreement with sensitivities previously reported in Santa Ines ewes 26–30 days after mating (Chaves et al., 2020), in Dorper, Shropshire, Hampshire and crosbred (Dorset x Polypay) sheep 30–90 days after mating (Roberts et al., 2019) and in German Blackhead and Merino sheep in mid-pregnancy (range: 71–120 days) (Steckeler et al., 2018). Furthermore, sensitivities similar to the sensitivity determined for the rapid visual PAG ELISA in the present study were reported for the "Idexx visual pregnancy test" in Santa Ines ewes 30 days after laparoscopic insemination (Chaves et al., 2017) and for the "Idexx bovine pregnancy test" in Corriedela and Ile de France x Texel sheep throughout pregnancy (Rovani et al., 2016). On the other hand, Kaplan et al. (2019) reported a low sensitivity for the "Idexx bovine pregnancy test" before the 56th day of pregnancy in Konya Merino sheep. In addition, Chaves et al. (2017) determined a lower sensitivity for the "Idexx visual pregnancy test" on day 26 post-mating compared to day 28 post-mating (73.17 versus 92.68, respectively).

In the present study, the specificity of rapid visual PAG ELISA was low (62.5%). This was attributed to early embryonic deaths having occurred before the day of pregnancy diagnosis. As PAGs are pregnancy-specific molecules, sheep have a long anestrus period, and controlled mating is practiced at farm level, PAGs detected in the maternal circulation are associated with pregnancy and cannot be of residual nature. Furthermore, in ruminants, PAGs can persist in maternal circulation several days after embryonic death, which may result in false positive results (Hussein et al., 2017). Based on this information, it can be said that pregnancy occurred but was not maintained in ewes diagnosed false positive (3 of the 8 non-pregnant ewes).

Rapid visual PAG ELISA and transabdominal ultrasonography were found to be in strong agreement with the lambing results (Table 3). Roberts et al. (2019) reported a strong agreement for "Idexx rapid visual pregnancy test" and transabdominal ultrasonography with lambing results (0.900 and 0.974, respectively) 30–90 days after breeding in sheep. Similar Kappa coefficients were computed for the two early pregnancy diagnosis methods in the present study.

In the Kappa analysis, values between 0.61 and 0.80 indicated a good agreement and values between 0.81 and 1.0 indicated a very good agreement (Altman, 1991). In the present study, while there was a very good agreement between transabdominal ultrasonography and the lambing results, the agreement between rapid visual PAG ELISA and the lambing results was slightly lower. This lower agreement was attributed to pregnancy losses having occurred before pregnancy diagnosis. In research in cattle, it was determined that PAGs had a long half-life and could be detected in the maternal circulation for a few days after the death of the embryo (Pohler et al., 2016). Therefore, embryonic deaths cause false
positive results. In research in sheep, lower serum PAG levels were measured in Romanov crossbred ewes with pregnancy loss (Hussein et al., 2017). However, further research is needed on the serum PAG profile of ewes in the event of embryonic death.

Table 3. Kappa analysis results of rapid visual PAG ELISA and transabdominal ultrasonography versus lambing for pregnancy diagnosis in Awassi sheep 34-38 days after mating

|                         | Rapid Visual PAG ELISA (Hızlı Görsel PAG ELISA) | Transabdominal Ultrasonography (Transabdominal Ultrasonografi) |
|-------------------------|--------------------------------------------------|---------------------------------------------------------------|
| Degree of Agreement     | 0.669                                             | 0.865                                                         |
| p-value                 | <0.001                                           | <0.001                                                        |

In McNemar’s analysis, there was no significant difference between either rapid visual PAG ELISA or transabdominal ultrasonography and the lambing results. According to the comparison of the ROC curves (Table 4, Figure 1), the area under the curve (AUC) was larger for transabdominal ultrasonography compared to rapid visual PAG ELISA. However, the difference between the two pregnancy diagnosis methods for AUC (0.176) was statistically insignificant.

The two pregnancy diagnosis methods having determined to show a strong agreement and insignificant differences indicates that both methods are reliable for early pregnancy diagnosis in sheep. The choice of method depends on several factors, including among others the cost of diagnosis, the access of shepherds to veterinary services, and herd management. Rapid visual PAG ELISA costs nearly 4$ per analysis. The cost of pregnancy diagnosis by ultrasonography varies between 8$ and 15$ per sheep, according to the base prices determined by the Chamber of Veterinary Surgeons. Although its cost is higher, when performed by a veterinarian, ultrasonography enables the detection of possible reproductive pathologies. Thereby, such cases can be managed at an early stage, and no additional fee is paid for this diagnosis. However, the transport costs of highland shepherds remote from veterinary services add up to the cost of pregnancy diagnosis. In view of such situations, further research is needed to analyse the cost-effectiveness of pregnancy diagnosis methods in small ruminants.

Figure 1. Receiver operating characteristic (ROC) curves of each pregnancy diagnosis method

|                         | AUC     | SE      | 95% CI          |
|-------------------------|---------|---------|-----------------|
| Rapid visual PAG ELISA  | 0.801   | 0.0923  | 0.663 to 0.900  |
| (Hızlı Görsel PAG ELISA)|         |         |                 |
| Transabdominal Ultrasonography | 0.976   | 0.0166  | 0.887 to 0.999  |
| (Transabdominal Ultrasonografi)|       |         |                 |

AUC: Area under curve (eğri altında kalan alan); SE: Standard error (Standart hata); 95% CI: 95% Confidence interval (%95 güven sınırları)

CONCLUSION
In conclusion, both rapid visual PAG ELISA and transabdominal ultrasonography are practical and reliable for pregnancy diagnosis as of day 34 post-mating in Awassi ewes. Rapid visual PAG ELISA can be easily applied in field conditions. Further studies are needed to exhaustive investigate the effects of embryonic deaths on serum PAG profile in sheep.
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REFERENCES
Akköse M, Cebi-Sen C, Kirmi A, Korkmaz O, Yazlık MO, Gocen M, Kuru M, Akçay E 2019. Pregnancy-associated glycoproteins (PAGs) and progesterone concentrations in Holstein heifers following two methods of estrus synchronization. Veterinaria México OA, 6(2): 1-11. https://doi.org/10.22201/fmvz.24486760.e.2019.2.646
Akköse M, Cebi-Sen C 2019. Sütçu sığırlarda erken gebelik tanısı. Hayvansal Üretim, 60(2): 171-179. https://doi.org/10.29185/hayuretim.458620
Alkan H, Kivrak MB, Satilmis F, Tekindal MA, Dinc DA 2020. Detection of twin pregnancies in ewes by pregnancy-associated glycoprotein assay and transabdominal ultrasonography. 72, 106399. https://doi.org/10.1016/j.domaniend.2019.106399.
Altman DG 1991. Practical statistics for medical research. Chapman and Hall, London, p., 404.
Aziz DM, Lazim EH 2012. Transabdominal ultrasonography in standing position for pregnancy diagnosis in Awassi ewes. Small Ruminant Research, 107(2-3): 131–135. http://dx.doi.org/10.1016/j.smallruminres.2012.05.007
Chaves CMS, Costa RLD, Duarte KMR, Machado DC, Paz CCP, Beltrame RT 2017. Visual ELISA for detection of pregnancy-associated glycoproteins (PAGs) in ewe serum. Theriogenology, 97: 78–82. https://doi.org/10.1016/j.theriogenology.2017.04.026
Chaves CMS, da Costa RLD, Duarte KMR, Beltrame RT, Quirino CR 2020. Evaluation of a cattle rapid test for early pregnancy diagnosis in sheep. Tropical Animal Health and Production, 52(3): 1345–1349. https://doi.org/10.1007/s11250-019-02130-7
Crilly JP, Politis AP, Hamer K 2017. Use of ultrasonographic examination in sheep veterinary practice. Small Ruminant Research, 152: 166-173. http://dx.doi.org/10.1016/j.smallruminres.2016.12.021
Güler H, Kaymaz M (2011). Akkaranın ikir koyunlarında transrektal ve transabdominal muayene tekniği kullanılarak embryonik ve fetal yaşam belirlenmesi. Ankara Üniversitesi Veteriner Fakültesi Dergisi, 58:99 – 104. https://doi.org/10.1501/Vetfak_0000002457
Hussein MS, Wael EB, Deghedy A, El-Desouky AM, Ramoun AA 2017. Serum concentration of pregnancy-associated glycoproteins (PAGs) as a predictor for embryonic/fetal losses and fetal numbers in cross-bred ewes. Life Science Journal, 14(5):106-111. https://doi.org/10.7537/marslsj140517.15
Jones AK, Gately RE, McFadden KK, Zinn SA, Govoni KE, Reed SA 2016. Transabdominal ultrasound for detection of pregnancy, fetal and placental landmarks, and fetal age before day 45 of gestation in the sheep. Theriogenology, 85(5): 939–945. 10.1016/j.theriogenology.2015.11.002
Kandemir Ç, Adanacioğlu H, Taşkın T, Koşun N 2019. Türkiye’de koyun ve koyun eti fiyatlarının bölgeore göre çok boyutlu ölçüleme analizi ile karşılaştırılması. Journal of Tekirdag Agricultural Faculty, 16(2): 315-327. https://doi.org/10.33462/jotaf.529109
Kaplan Y, Özyurtlu N, Köse M, Atlı MO, Küküçaksan I, Kirbaş M 2019. Gebe Konya Merinosu koyunlarında erken gebelikte gebelik iliskili glikoproteinlerin plazma profilinin belirlenmesi. Atatürk Üniversitesi Veteriner Biliimleri Dergisi, 14(3):307-314. . https://doi.org/10.17094/ataunivbd.588666
Karen A, Kovacs P, Beckers JF, Szenci O 2001. Pregnancy diagnosis in sheep: Review of the most practical methods. Acta Veterinaria. Brno, 70(2): 115–126. https://doi.org/10.2754/avb20017002115
Kul S, Akcak A 2002. Ivesi ve Ost-Friz x Ivesi Melez (F1) kuzularda besi performansı, kesim ve karkas özellikleri. Journal of Research in Veterinary Medicine, 21(1): 1-7.
Küplülü Ş, Çetin Y, Macun HC, Taşdemir U 2002. Akkaranın ikir koyunlarında transrektal ve transabdominal ultrasonografi yöntemle erken gebelik tani sınırlarının belirlenmesi. Lalahan İveteğine Araştırmalar Enstitüsü Dergisi, 42:25-33.
Lobago F, Bekama M, Gustafsson H, Beckers JF, Yohannes G, Aster Y, Kindahl H 2009. Serum profiles of pregnancy-associated glycoprotein, oestrone sulphate and progesterone during gestation and some factors influencing the profiles in Ethiopian Borana and crossbred cattle. Reproduction in Domestic Animals, 44(4): 685-692. 10.1111/j.1439-0531.2007.01049
Mercadante PM, Waters KM, Mercadante VR, Lamb GC, Elzo MA, Johnson SE, Rae DO, Yelic JV, Ealy AD 2013. Subspecies differences in early fetal development and plasma pregnancy-associated glycoprotein concentrations in cattle. Journal of
Animal Science, 91(8):3693-3701. 10.2527/jas.2012-6130

Pohler KG, Pereira MH, Lopes FR, Lawrence JC, Keisler DH, Smith MF, Vasconcelos JL, Green JA 2016. Circulating concentrations of bovine pregnancy-associated glycoproteins and late embryonic mortality in lactating dairy herds. Journal of Dairy Science, 99(2):1584-1594. http://dx.doi.org/10.3168/jds.2015-10192

Roberts J, May K, Ajani O, Kaneene J 2019. A comparison of pregnancy diagnosis methods in commercial sheep using lambing as a gold standard. Clinical Theriogenology. 11(2): 107-113.

Rovani MT, Cezar AS, Rigo ML, Gasperin BZ, Nobrega Júnior JE, Torres FD, Gonçalves, PBD, Ferreira R 2016. Evaluation of a bovine pregnancy-associated glycoprotein enzyme-linked immunosorbent assay kit for serological diagnosis of pregnancy in sheep. Ciencia Rural. 46(2): 362-367. http://dx.doi.org/10.1590/0103-8478cr20150270

Sousa NM, Ayad A, Beckers JF, Gajewski Z 2006. Pregnancy-associated glycoproteins (PAG) as pregnancy markers in the ruminants. Journal of Physiology and Pharmacology, 57(Supp 8):153-171.

SPSS 2016. IBM SPSS Statistics 24.0 for Windows. Armonk, NY.

Steckeler P, Weber F, Zerbe H, Rieger A, Voigt K 2018. Evaluation of a bovine visual pregnancy test for the detection of pregnancy-associated glycoproteins in sheep. Reproduction in Domestic Animals, 54(2): 280-288. https://doi.org/10.1111/rda.13356

Şireli HD, Tekel N 2013. İvesi erkek kuzuların besi performansı ve karkas özelliklerine süt emme dönemiinde farklı büyüme sistemlerinin etkisi. Journal of Agricultural Sciences, 19(1): 63-70. https://doi.org/10.1501/Tarimbil_0000001228

Tandiya U, Nagar V, Yadav VP, Ali I, Gupta M, Dangi SS, Hyder I, Yadav B, Bhakat M, Choukat VS, Khan FA, Maurya VP, Sarkar M 2013. Temporal changes in pregnancy-associated glycoproteins across different stages of gestation in the Barbari goat. Animal Reproduction Science, 142:141–148. https://doi.org/10.1016/j.anireprosci.2013.09.016

Taşan A 2016. Halk Elinde Yetiştirilen Ivesi Koyunlarında Laktabiyon Süt Verim Özelliklerinin Belirlenmesi ve Genetik Parametrelerin Tahmini. Harran Üniversitesi Fen Bilimleri Enstitüsü Zootekni Anabilim Dalı, Yüksek Lisans Tezi, 46 sy.

Tekel N, Şireli HD, Vural ME 2007. Besi süresinin ivesi erkek kuzuların besi performansı ve karkas özelliklerine etkisi. Journal of Agricultural Sciences, 13(4): 372-378. https://doi.org/10.1501/Tarimbil_0000000389

Uçar U, Köse M, Atlı MO 2018. Konya Merinosu koyunlarında gebelik ilişkili glikoproteinlerin gebelikteki plazma profili ve erken gebelik tamsında kullanılabilirliği. Dicle Üniversitesi Veteriner Fakültesi Dergisi, 11(2): 77-82.

Ulusoy H, Kaymaz M 2009. Koyunlarda gebelik tamsı. Veteriner Hekimleri Derneği Dergisi, 80(1):31-36.

Xie S, Low RC, Nagel RJ, Kramer KK, Anthony RV, Zoli AP, Beckers JF, Roberts RM 1991. Identification of the major pregnancy-specific antigens of cattle and sheep as inactive members of the aspartic proteinase family. Proceedings of the National Academy of Sciences of the United States of America, 88(22):10247-10251. https://doi.org/10.1073/pnas.88.22.10247

Yağanoğlu AM, Topal M 2018. Morkaraman-İvesi ve Tuj irkı koyunlarda gebelik testlerinin Roc analizi ile karşılaştırılması. Alınteri Journal of Agriculture Sciences, 33(1):29-36. https://doi.org/10.28955/alinterizbd.347633