Abstract: The purpose of this study was to examine the effect of desensitizing dentifrices containing stannous fluoride (SnF$_2$) on dentinal tubule occlusion. Two experimental dentifrices with the same ingredients but different SnF$_2$ concentrations (Group II, 0.4% w/w; Group III, 0.454% w/w) were used; distilled water was used in the control group (Group I). Third molars were collected from Japanese and American dental patients. The crowns were removed and sectioned to obtain dentin discs, which were further cut into quarters. Thirty-six specimens each from Japanese and American patients were divided into three sets (n = 12 each) and assigned to each of the three treatment groups. The specimens were brushed for 10 s twice per day for 4 days. After treatment, the discs were observed by scanning electron microscopy, and the extent of dentinal tubule occlusion in the images was expressed on a five-point categorical scale. Group II and III specimens from Japanese and American patients showed greater dentinal tube occlusion than those from Group I, but the differences were not statistically significant. The present results suggest that both SnF$_2$ concentrations mitigate dentin hypersensitivity, regardless of patient ethnicity.

Keywords: dentin hypersensitivity; stannous fluoride; dentifrice; dentinal tubule occlusion; four-day occlusion method.

Introduction
Dentin hypersensitivity—short, sharp pain induced by external stimulation—is a common oral health problem (1,2). The hydrodynamic theory of the mechanism of dentin hypersensitivity is widely accepted (3,4). External stimuli such as thermal, tactile, chemical, or osmotic pressure provoke movement of intratubular dentinal fluids in exposed dentin because of the absence of cementum after gingival recession (5,6). The symptoms of dentin hypersensitivity can be mitigated by modifying the surface of exposed dentin and occluding dentinal tubules (7). Several approaches, such as application of chemical agents or substances similar to hydroxyapatite, filling or sealing of cavities with resin-based materials or glass-ionomer cements, and use of gingival grafts, have been recommended to reduce hypersensitivity in dental clinics (8-10). Chemical agents such as potassium nitrate and potassium chloride are thought to suppress pain by penetrating dentinal tubules and elevating potassium ion levels (8). In addition, organic and inorganic chemicals and their compounds have been used to block dentinal fluid flow, thereby reducing hydraulic conductance across dentinal tubules (8-10). However, no single method or desensitizing agent is ideal for the treatment of hypersensitivity. Therefore, a treatment strategy based on hypersensitivity stage should be developed and implemented.

Use of dentifrices that include a chemical compound...
to occlude dentinal tubules or desensitize the peripheral sensory nerves in the dental pulp is the treatment of choice because of its low cost and the easy availability of these substances in daily tooth brushing (8). Potassium oxalate, aluminum lactate, strontium chloride, and complex fluorides are commonly included in dentifrices, to mitigate pain (9). However, these dentifrices occasionally fail to reduce pain or may lose their effect on dentin hypersensitivity over time. Therefore, a more reliable and longer-lasting dentin-desensitizing dentifrice is needed.

Dentifrices containing stannous fluoride (SnF$_2$) have anti-caries and anti-gingivitis effects and have been used for over 50 years (11). Moreover, previous studies reported that these dentifrices may reduce enamel erosion (12-14), breath odor (15), and dentin hypersensitivity (16-22). Nevertheless, concerns have been expressed regarding the formulation of these dentifrices, particularly with respect to the stability of stannous ions versus bioavailability, the tendency to enhance extrinsic dental stains, and the unpleasant taste (23). To prevent degradation of SnF$_2$ into insoluble and inactive stannic hydroxides and oxides and mitigate dental staining, an anhydrous SnF$_2$ formulation that includes the chelating agent sodium tripolyphosphate was developed (21). However, the effect of anhydrous SnF$_2$-containing dentifrices on dentin hypersensitivity is not clear.

The present study used a 4-day occlusion method to examine the effect of a newly formulated dentin-desensitizing dentifrice including SnF$_2$. The effect was evaluated by using a categorical occlusion scale for dentinal tubules and scanning electron microscopy (SEM) of samples collected from patients who were asked to brush their teeth with this dentifrice twice daily for up to 4 days. In addition, the teeth of Japanese and American patients were compared to investigate the effect of patient ethnicity on the ability of the dentifrice to occlude dentinal tubules. The null hypotheses tested were that the effectiveness of experimental desensitizing dentifrices is not influenced by SnF$_2$ concentration and that the status of occluded dentinal tubules would not differ between Japanese and American patients.

### Materials and Methods

#### Study materials

Two experimental dentifrices with the same ingredients but different percentages of SnF$_2$ (Group II, 0.4% w/w SnF$_2$; Group III, 0.454% w/w SnF$_2$) were used (Table 1). Distilled water was used for the control group (Group I).

#### Collection of human teeth

To investigate the influence of patient ethnicity on occlusion of dentinal tubules by the dentifrices, third molars were collected from Japanese and American dental patients. Caries-free third molars extracted for orthodontic or other medical/dental reasons were obtained from Japanese patients who visited the dental hospital at the Nihon University School of Dentistry. Informed consent for the study was obtained from all patients. In addition, unerupted third molars from American patients (with ethical committee approval from GlaxoSmithKline) were sourced through an appropriate contracted supplier of GlaxoSmithKline Consumer Health. The study protocol was reviewed and approved by the Ethics Committee for Human Studies of the Nihon University School of Dentistry (2015-03). Partially and fully erupted molars were not included in this study. Unerupted third molars of regular size and shape were selected, and teeth with any signs of cracking in the enamel or across the base were excluded. Specimens for which the extraction procedure caused slight scuffing of the enamel were included. After extraction, soft tissue attached to the root was immediately removed with hand instruments.

| Group | Experimental dentifrice (Lot No.) | Main components | Manufacturer |
|-------|---------------------------------|-----------------|--------------|
| I     | —                               | Distilled water | —            |
| II    | (1470B302)                      | 0.4% w/w stannous fluoride, glycerin, PEG-8, hydrated silica, pentasodium triphosphate, sodium lauryl sulphate, flavor, titanium dioxide, polyacrylic acid, cocamidopropyl betaine, sodium saccharin | GlaxoSmithKline Consumer Healthcare, Middlesex, UK |
| III   | (1470B283)                      | 0.454% w/w stannous fluoride, glycerin, PEG-8, hydrated silica, pentasodium triphosphate, sodium lauryl sulphate, flavor, titanium dioxide, polyacrylic acid, cocamidopropyl betaine, sodium saccharin | GlaxoSmithKline Consumer Healthcare, Middlesex, UK |

PEG-8: polyethylene glycol 400.

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and the tooth was then immersed in distilled water for 6 h. The extracted teeth were stored frozen (−20°C) until further experiments.

**Specimen preparation**

The crowns of the teeth were sectioned perpendicular to the long axis of the roots, just above the enamel-dentin junction, with a diamond disc saw (Isomet 1000; Buehler Ltd, Lake Bluff, IL, USA) and copious water cooling, to remove most of the crown. The specimens were rinsed with distilled water to remove any debris and stored in 0.1% w/w thymol solution in a sealed container. Dentin disc specimens free from cracks, staining, and other imperfections were prepared as follows: the removed crowns were sectioned parallel to the cut surface with a diamond saw to obtain a dentin disc (thickness, 1.0-1.5 mm). Only one disc could be cut from each crown. Each dentin disc specimen was then cut into quarters with a diamond bur. One quarter from each dentin disc was placed in a 25-mm mold, with the crown side of the specimen facing down, and covered with acrylic resin to a depth of 10 mm (Resin tray II; Shofu Inc., Kyoto, Japan). After the resin had hardened at room temperature and atmospheric pressure, the specimen was removed from the mold, and the dentin face was polished sequentially with 800-, 1200-, and 2,500-grit silicon carbide papers (Struers Inc., Cleveland, OH, USA), and finally with a 0.25-μm diamond polishing suspension (DP-Paste; Struers, Ballerup, Denmark), to a mirror finish. The specimens were cleaned in an ultrasonic bath (Quantrex 310; L&R Mfg., Co., Kearny, NJ, USA) for 120 s with 300 ppm of an ultrasonic cleaner (RC-100, Mettler-Toledo International Inc., Tokyo, Japan). The specimens were rinsed with distilled water for 10 s, thus ensuring that all visible paste was removed. The brush head was cleaned in distilled water with an ultrasonic bath, and new paste was applied for each group. For dentin disc treatment, an electric toothbrush (Braun Oral-B 5000 with wireless smartguide, Oral-B FlossAction head; Procter & Gamble, Cincinnati, OH, USA) was used in daily clean mode (Fig. 1). Each specimen was brushed after wetting the brush head with distilled water and applying 1.1 ± 0.1 g of test dentifrice to the brush head. Brushing was performed for each group. For dentin disc treatment, the specimens were left for 30 s before being gently rinsed in distilled water for 10 s. All specimens from one country of origin were immersed in a beaker containing 50 mL of artificial saliva (Table 2) for 60 min with no stirring. After 60 min, the first brushing was performed for each group. For dentin disc treatment, an electric toothbrush (Braun Oral-B 5000 with wireless smartguide, Oral-B FlossAction head; Procter & Gamble, Cincinnati, OH, USA) was used in daily clean mode (Fig. 1). Each specimen was brushed after wetting the brush head with distilled water and applying 1.1 ± 0.1 g of test dentifrice to the brush head. Brushing was performed uniformly for 10 s across a resin plug containing three dentin specimens assigned to a single treatment group. An equivalent brushing force of approximately 100 g was used, as measured with an electronic scale (AE163, Mettler-Toledo International Inc., Tokyo, Japan). The specimens were left for 30 s before being gently rinsed in distilled water for 10 s, thus ensuring that all visible paste was removed. The brush head was cleaned in distilled water with an ultrasonic bath, and new paste was applied for each separate treatment. The specimens were then returned to 50 mL of fresh artificial saliva and stored for at least 4 h.

The second brushing was carried out as described above. At the end of days 1-3, the specimens were placed in a sealed pot on tissue paper wetted with distilled water for storage overnight at 5 ± 2°C, for treatment the next day.

| Table 2 Components of artificial saliva |
|----------------------------------------|------------------|
| Potassium chloride                     | 30 mM            |
| Calcium chloride di-hydrate            | 3 mM             |
| Potassium di-hydrogen orthophosphate   | 10 mM            |
| Sodium chloride                        | 13 mM            |
| Type II Porcine stomach mucin (M2378 Sigma) | 0.22% w/w     |
| Sodium azide as preservative          | 0.02% w/w        |
day. At the end of day 4, the specimens were removed, washed with distilled water, and allowed to dry overnight at room temperature and atmospheric pressure in preparation for SEM.

**Scanning electron microscopy (SEM)**

All dentin discs were observed after treatment by field-emission SEM (ERA-8800FE; Elionix Inc., Tokyo, Japan). The specimens were coated with a thin film of gold (Au) in a Quick Coater vacuum evaporator (Type SC-701; Sanyu Electron Co., Tokyo, Japan). To ascertain the uniformity of the dentin profile, five predesignated areas of the dentin were imaged in each dentin section. At least three representative SEM micrographs were acquired at ×1,500 magnification (three images per disc quarter). Observations were performed at an operating voltage of 10 kV.

**Evaluation of dentinal tubule occlusion**

Each SEM image was assessed for the extent of dentinal tubule occlusion, as expressed on a five-point categorical scale (Fig. 2A-2E), by a single examiner from the Department of Operative Dentistry, Nihon University School of Dentistry. Before the study began, the examiner was calibrated against a set of standard micrographs encompassing the entire grading scale. The examiner was blinded to the type of treatment and did not observe the images being taken.

**Energy-dispersive X-ray spectroscopy (EDX)**

The elements present in representative dentin surfaces from each group were analyzed with SEM/EDX (GENESIS 2000; EDAX Ametek Co., Tokyo, Japan) at 20 kV and ×1,500 magnification. Specimens brushed with distilled water (controls) were also analyzed. Measurements were taken perpendicular to the prepared dentin specimens. The elemental content (wt%) of the Au-coated surface was measured at three regions, as close as possible to the center, in five specimens from each group, and the mean value was determined for each group. The analysis was performed with ZAF correction (atomic number, absorption, and fluorescence), based on standard-less correction (24). 

**Statistical analysis**

Treatment differences were analyzed by using the Kruskal-Wallis test followed by the Steel-Dwass test at a significance level of 0.05, and differences in relation to national origin were analyzed by using the Mann-Whitney U test with Bonferroni correction at a significance level of 0.05. In SEM/EDX analysis, data from each group and each detected element were analyzed with the Kruskal-Wallis test followed by the Steel-Dwass test at a significance level of 0.05. All statistical analysis was done with the Sigma Plot, ver. 11.0, software program (SPSS Inc., Chicago, IL, USA).
Results

Evaluation of dentinal tubule occlusion
Figures 3 and 4 show the categorization of specimens, by group. All Group I specimens (regardless of national origin) were scored as 5 (unoccluded dentinal tubules). In Group II, all specimens from Japanese patients were scored as 3 or 4, and all specimens from American patients were scored as 2, 3, or 4. In Group III, specimens from Japanese and American patients were scored as 1 through 4. In Groups II and III, the numbers of specimens with a score of 3 far exceeded the numbers of specimens with other scores, regardless of patient ethnicity.

Table 3 shows the results of the Kruskal-Wallis test followed by the Steel-Dwass test, for comparison of treatment, and the results of the Mann-Whitney U test with Bonferroni correction, for comparison of specimen origin. Although values from Groups II and III did not significantly differ ($P > 0.05$) for Japanese specimens, dentinal tube occlusion was significantly greater in these groups than in Group I ($P < 0.05$). The trend was similar for American specimens, i.e., dentinal tube occlusion was significantly greater in Groups II and III than in Group I ($P < 0.05$). In addition, the results did not significantly differ in relation to specimen origin ($P > 0.05$).

SEM observation
Regardless of patient ethnicity, all dentinal tubules in the initial dentin specimens were completely opened, and smear plugs were absent after mirror polishing and ultrasonic cleaning for 120 s, indicating that the dentinal tubules were free from debris (Figs. 5A, 6A). The morphological appearance of specimens in treatment Group I (without dentifrice) was similar to that of the initial specimens from both ethnic groups (Figs. 5B, 6B). No deposit or debris in the intertubular dentin or dentinal tubules was noted. In contrast, the morphological characteristics of American and Japanese specimens in treatment Groups II and III differed from those in Group I and the initial dentin specimens (Figs. 5CD, 6CD). Some deposits derived from the dentifrice were observed in the intertubular dentin and dentinal tubules. In particular, the dentinal tubules in Group III appeared to be more occluded than those in Group II (Figs. 5D, 6D). Under higher magnification, aggregated small particles in the dentinal tubules and granular deposits on the dentin surface were observed in specimens from Group III.

Energy-dispersive X-ray microanalysis
The elements present in representative dentin surfaces from each group are shown in Table 4. Si and Sn were detected in Groups II and III but not in Group I. No significant differences in elemental profile were noted between Groups II and III, regardless of specimen origin. The control specimens from Japanese and American patients had the same elements and similar concentrations of those elements.

Discussion
To overcome the drawbacks of earlier dentifrices containing SnF$_2$, advanced technologies have been devel-
oped to stabilize the compound and reduce tooth staining (21,25). These technologies have led to a re-evaluation of the use of SnF2 to reduce caries, gingivitis, and enamel erosion and mitigate dentin hypersensitivity (12-22). Several studies around the world have investigated advanced SnF2-containing dentifrices (12-22). A recent 8-week clinical trial of dentifrices containing anhydrous 0.454\% w/w SnF2 showed a significant (>20\%) reduction in sensitivity as compared with a control dentifrice containing 1,000 ppm sodium fluoride (19). Another clinical trial reported a statistically significant reduction in dentin hypersensitivity with a 0.454\% w/w stabilized SnF2 dentifrice as compared with a negative control dentifrice. The desensitizing effect increased in magnitude with continued use (20).

The present results show that a SnF2 dentifrice (Groups II and III) induced significantly greater dentinal tube occlusion than did distilled water (Group I). The SnF2 desensitizing mechanism is attributable to formation of insoluble stannous compounds due to rapid oxidization (Sn2+ to Sn4+) and hydrolysis of SnF2 in the presence of water or saliva (19). These compounds contribute to the formation of a layer over the dentin and reduce its permeability by creating precipitates within the dentinal tubules (26). Using an ultrasonic device, Endo et al. investigated the effect of a calcium phosphate desensitizer on dentin

| Japanese   | Group I | Group II | Group III |
|------------|---------|----------|-----------|
| Carbon     | 21.6 (2.6)\% | 25.7 (0.8)\% | 22.9 (2.9)\% |
| Nitrogen   | 13.2 (1.0)\% | 14.3 (0.8)\% | 12.3 (0.9)\% |
| Oxygen     | 33.1 (1.3)\% | 33.1 (3.8)\% | 30.3 (3.9)\% |
| Sodium     | 0.7 (0.2)\% | 0.6 (0.1)\% | 0.6 (0.1)\% |
| Magnesium  | 0.4 (0.1)\% | 0.4 (0.1)\% | 0.3 (0.1)\% |
| Silicon    | 0.7 (0.1)\% | NA        | 0.8 (0.1)\% |
| Phosphorus | 11.6 (0.6)\% | 10.4 (0.7)\% | 12.0 (2.1)\% |
| Tin        | 1.8 (0.3)\% | NA        | 1.7 (0.4)\% |
| Calcium    | 17.6 (1.1)\% | 15.7 (3.3)\% | 17.8 (1.0)\% |

Table 4. Effect of dentifrices on elemental composition (wt\%) on the dentin surface

- Group I: Initial group
- Group II: Treatment Group I (×1,500 and ×20,000)
- Group III: Treatment Group II (×1,500 and ×20,000)

Values in parentheses are standard deviations.

Fig. 5. Representative SEM images of Japanese dentin specimens showing morphological differences in relation to treatment. 5A. Initial specimen of dentin (×1,500). 5B. Treatment Group I (×1,500 and ×20,000). 5C. Treatment Group II (×1,500). 5D. Treatment Group III (×1,500 and ×20,000).

Fig. 6. Representative SEM images of American dentin specimens showing morphological differences in relation to treatment. 6A. Initial specimen of dentin (×1,500). 6B. Treatment Group I (×1,500 and ×20,000). 6C. Treatment Group II (×1,500). 6D. Treatment Group III (×1,500 and ×20,000).
hypotheses and found that sonic velocity was significantly higher for the desensitizer-applied specimens than for negative and positive control specimens (27). They suggested that this higher sonic velocity might account for the small precipitates inside the dentinal tubules and the presence of a layer of granular precipitate on the dentin surface, and that occlusion of dentinal tubules might reduce dentinal fluid movement (27). The present SEM images from Groups II and III are consistent with these earlier findings and revealed deposits in intertubular dentin and dentinal tubules. The aggregates of small particles were presumed to be SnF$_2$ compounds on the basis of detection of tin in EDX analysis.

The most common SnF$_2$ concentration in dentifrices is 0.454% w/w; however, 0.4% w/w SnF$_2$ has long been widely used as a topically applied gel. Although the Japanese Ministry of Health, Labor and Welfare began permitting the use of dentifrices containing up to 1,500 ppm fluoride in 2017, almost all commercially available dentifrices in Japan contain less than 1,000 ppm fluoride. However, it remains to be determined if dentifrices containing 0.454% w/w and 0.4% w/w SnF$_2$ differ in relation to dentinal tubule occlusion. The present results showed no significant difference in dentinal tubule occlusion between these SnF$_2$ concentrations. Hence, the first null hypothesis—that the effectiveness of an experimental desensitizing dentifrice would not be influenced by the concentration of SnF$_2$—was not rejected.

Dentifrices containing 0.454% w/w or 0.4% w/w SnF$_2$ may be used to mitigate dentin hypersensitivity. Dentinal tubule occlusion did not significantly differ in relation to specimen origin under any of the treatment conditions. Hence, the second null hypothesis—that the effectiveness of an experimental desensitizing dentifrice would not be influenced by the concentration of SnF$_2$—was not rejected. Dentifrices containing 0.454% w/w or 0.4% w/w SnF$_2$ may be used to mitigate dentin hypersensitivity.

Dentinal tubule occlusion did not significantly differ in relation to specimen origin under any of the treatment conditions. Hence, the second null hypothesis—that the effectiveness of an experimental desensitizing dentifrice would not be influenced by the concentration of SnF$_2$—was not rejected. Dentifrices containing 0.454% w/w or 0.4% w/w SnF$_2$ may be used to mitigate dentin hypersensitivity.

Tooth erosion related to wear can result in diminished esthetic appearance, functional occlusion, dentin hypersensitivity, and tooth fracture. Progression of root caries is caused by dentin exposure with gingival recession and poor oral hygiene and is common among elderly adults, because of their decreased saliva secretion. In addition to mitigating dentin hypersensitivity, dentifrices containing SnF$_2$ will likely slow progression of erosion and root caries because they induce remineralization and have anti-plaque and anti-gingivitis properties.

The present results suggest that dentinal tubule occlusion was significantly greater for dentifrices containing SnF$_2$ than in controls. In addition, dentinal tubule occlusion was greater for the dentifrice with a higher SnF$_2$ concentration than for that with a lower concentration, although the difference was not significant. No significant difference in occlusion between Japanese and American specimens was noted after any of the present treatment methods. In conclusion, these results suggest that 0.454% w/w and 0.4% w/w SnF$_2$ are effective in mitigating dentin hypersensitivity, regardless of patient ethnicity.

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**Conflict of interest**

This study was funded by GlaxoSmithKline. The sponsor had no control over the interpretation, writing, or publication of this work.

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