Changing pattern of infectious agents in postneurosurgical meningitis

Abstract

Background: The rates of postneurosurgical infections, particularly meningitis and the most common pathogens have been reported variable during the recent years. The aim of this research was to determine the prevalence of postneurosurgical meningitis and its current spectrum of infectious agent.

Methods: In a descriptive study, the postneurosurgical patients’ cerebrospinal fluid was cultured on blood and MacConkey agar plates and evaluated at Microbiology Laboratory. 55 significant isolates as species level were recognized by bacteriological techniques.

Results: Acinetobacter baumannii (A. baumannii) was the most common organism [29(52.7%)], followed by Klebsiella pneumoniae [8(14.5%)], Methicillin-resistant Staphylococcus aureus [6(10.9%)], Staphylococcus saprophyticus [5(9.1%)], Pseudomonas aeruginosa [3(5.5%)], Methicillin-sensitive Staphylococcus aureus [3(5.5%)], and Enterococcus faecalis [1(1.8%)]. The majority of A. baumannii isolates were carbapenem-resistant.

Conclusion: Our research revealed that the rate of postneurosurgical meningitis due to carbapenem-resistant A. baumannii has been increased. This finding emphasized the importance of preventive strategies against A. baumannii. The changing pattern of infectious agents in postneurosurgical meningitis over time suggests the necessity of other studies.

Keywords: Acinetobacter Baumannii, Post Neurosurgical Meningitis, Pattern, Infectious Agents

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Neurosurgical procedures, for example, craniotomy, cerebrospinal fluid (CSF) shunting, and external ventricular drain (EVD) insertion may lead to central nervous system (CNS) infections. Meningitis is the most common infectious complication caused by the different kinds of pathogens (1, 2). This complication is rare with accompanying high morbidity and mortality without immediate treatment (2). The rates of postneurosurgical meningitis have been reported from 0.3% to 10% (1-7). Because of subtle and unusual manifestations, nonspecific CSF parameters, and aseptic meningitis due to meningeal inflammation after neurosurgical procedures, the definite diagnosis is challenging (8, 9). Furthermore, some studies have shown the high rates of negative CSF smears or cultures in patients with bacterial meningitis (10, 11). Previous studies found some risk factors for postneurosurgical CNS infections, such as implantation of foreign body, CSF leakage, previous neurosurgical infection, absence of antibiotic prophylaxis, duration of surgery over 4 hours, interventions involving nasal sinuses, emergency surgeries, and prior to radiation therapy (2, 3, 12).
The most common pathogen in many reports was *Staphylococci*, particularly *Staphylococcus aureus* (1-3, 7, 9, 13-15), whereas some recent reports have shown the higher rates of gram-negative organisms (10, 11, 16). The changing pattern of pathogens necessitates more research. Also, the locally published data are lacking. Thus, we conducted this study to determine the prevalence of postneurosurgical meningitis and its current spectrum of infectious agents in a private hospital in Tehran, Iran.

The aim of this research was to determine the prevalence of postneurosurgical meningitis and its current spectrum of infectious agents.

**Methods**

We conducted a descriptive study from March 2010 through March 2012 in a private hospital in Tehran, Iran. We selected the patients that underwent neurosurgical procedures, followed by fever, headache, loss of consciousness, or meningismus. Then, the patients’ CSF was obtained and inoculated into blood and MacConkey agar plates at the Microbiology Laboratory. We incubated these plates at 37°C for 24 hours, and recognized the significant isolates as species level by common bacteriological techniques. The negative cultures were excluded. Totally, 55 pathogens were isolated at the Microbiology Laboratory from the patients’ specimens. The control organisms used were *Staphylococcus aureus* ATCC 29213 and 25923; *Methicillin-resistant Staphylococcus aureus* (non-MRSA); ATCC 43300 (MRSA); *Enterococcus faecalis* ATCC 51299; *Staphylococcus epidermidis* ATCC 12228; *Staphylococcus saprophyticus* ATCC BAA-750; *Enterococcus faecalis* ATCC 29212; *Streptococcus pneumoniae* ATCC 49619; *Escherichia coli* ATCC 35218; *Klebsiella pneumoniae* ATCC 700603; *Pseudomonas aeruginosa* ATCC 27853; *Acinetobacter baumannii* ATCC BAA-747; and *Enterobacter aerogenes* ATCC 13048 to control the quality of media and evaluate color stability. Three control American Type Culture Collection (ATCC) organisms were used for each test, and we examined each test for each new group varied (17). Susceptibility pattern was determined by disk diffusion method based on Kirby-Bauer method on Mueller-Hinton agar plate for gram-negative pathogens (18).

The following disks were used: Amoxicillin clavulanate (30 μg/disk) was placed in the center of plate while ceftriaxone (30 μg/disk), cefixime (5 μg/disk), cefotaxime (30 μg/disk), carbenicillin (100μg/disk), cephalexin (30μg/disk), trimethoprim-sulfamethoxazole (25μg/disk), amikacin (30 μg/disk), gentamicin (10 μg/disk), ciprofloxacin (5 μg/disk), imipenem (10 μg/disk), tetracycline (30 μg/disk), nalidixic acid (30 μg/disk), piperacillin-tazobactam (110 μg/disk), and colistin (10 μg/disk) were situated around the central disc. Five disks were been placed on each plate. The Clinical and Laboratory Standards Institute (CLSI) recommends a zone size of ≥ 5mm difference between the cephalosporin disk with and without clavulanic acid which is considered as significant and Extended Spectrum Beta Lactamase (ESBL) producing bacteria (19). For *Staphylococcus aureus* isolates, methicillin and vancomycin susceptibility were determined by using the CLSI microdilution methods (BHI). Portion (10μg) of 0.5 McFarland suspensions were pipetted onto brain heart infusion agar plates (17).

The following data obtained in patients with positive CSF cultures: Age, gender, ward, type of neurosurgery, the use of device in CNS, the days of meningeal signs and symptoms appearance after the neurosurgery, type of organism, antimicrobial susceptibility, and outcome.

**Results**

From March 2010 through March 2012, 1593 patients underwent neurosurgical procedures in our hospital. 90% of patients had a kind of device (68% EVD, 20% Shunt, and 1% Prosthesis). 176(11%) patients were suspected to have postneurosurgical meningitis (fever, headache, loss of consciousness, or meningismus). 55 patients (3.5%) had positive CSF culture. 31 patients were males (56%) and 24 were females (44%). The mean age of patients was 32.4±20.9. The youngest patient was 1 year of age, the oldest 78. Most of the patients belonged to the young age group between 20-29 years (20%) (table 1).

28 patients (51%) had an emergency neurosurgery. 44 patients (80%) were admitted to Intensive Care Unit (ICU), and the others were hospitalized in neurosurgery ward. The mean days of the average time from onset of meningeal signs and symptoms after surgery was 1.3±0.5 (Minimum 1 day and maximum, 3 days). All patients received antibiotics before the signs and symptoms appear. As shown in table 2, craniotomy in terms of the neurosurgical procedures, accounted for the greatest number [47(85.5%)]. 16.5% of Prosthesis, 13.5% of Spinal fusions, 13% of CSF Shunts, 11% of Laminectomies, 9.5% of Craniotomies, 7.5% of...
EVDs, and 3% of Discectomies led to meningitis. In meningitis group, a kind of device was used for 48 (87.3%) patients. Most of them were EVD [31 (56.4%)], followed by CSF shunt [16 (29.1%)], and prosthesis [1 (1.8%)].

Table 1. The Frequency (Percentage) of Patients’ Age Group in Post Neurosurgical Meningitis

| Age Group | Frequency | %  |
|-----------|-----------|----|
| <10 years | 10        | 18.2 |
| 10-19 years | 7        | 12.7 |
| 20-29 years | 11       | 20 |
| 30-39 years | 10       | 18.2 |
| 40-49 years | 5        | 9.1 |
| 50-59 years | 4        | 7.3 |
| ≥60 years | 8         | 14.5 |
| Total     | 55        | 100 |

Table 2. The Frequency (Percentage) of Neurosurgical Procedures in Patients with Post Neurosurgical Meningitis

| Procedure      | Frequency | Percentage |
|----------------|-----------|------------|
| Craniotomy     | 47        | 85.5       |
| Spinal Fusion  | 3         | 5.5        |
| Laminectomy    | 3         | 5.5        |
| Discectomy     | 2         | 3.5        |
| Total          | 55        | 100        |

Acinetobacter baumannii was the most common organism [29 (52.7%)], followed by Klebsiella pneumoniae [8 (14.5%)], MRSA [16 (10.9%)], Staphylococcus saprophyticus [5 (9.1%)], Pseudomonas aeruginosa [3 (5.5%)], Methicillin-sensitive Staphylococcus aureus (MSSA) [3 (5.5%)], and Enterococcus faecalis [1 (1.8%)].

67% of gram-negative bacilli isolates were ESBL. 76% of A. baumannii isolates were carbapenem-resistant (only resistant to colistin) and the others (24%) were multi-drug resistant (only sensitive to carbapenems and colistin). 62% of A. baumannii infections led to death. 2 patients with A. baumannii infection had recurrence.

Unfortunately, the mortality rate was 45%. 57% of infections due to gram-negative bacteria were accompanied with death, whereas, only two patients (13%) with gram-positive infection died. 28 patients (51%) were treated, and 2 (3.5%) of them had a recurrence.

Discussion

In our study, the incidence of postneurosurgical meningitis was 3.5%, which has been similar to the recent reports (11, 16). All patients received prophylactic antibiotics after the surgery. Several articles have noted the importance of this matter (2, 11). The rate of emergency surgeries in McClelland and Hall (15%) and Logigan et al.’s studies (16%) was remarkably lower than our research (51%) (2, 15). McClelland and Hall showed 1.1% of craniotomies, 1.6% of CSF shunts and none of laminectomy, discectomy, and spinal fusion led to meningitis (2). In Logigan et al.’s study, 55% and 32% patients with postneurosurgical meningitis underwent craniotomy and CSF shunt insertion, respectively (15). We found the intermediate rates: 9.5% of craniotomies, 13% of CSF shunts, 13.5% of spinal fusions, 11% of laminectomies, and 3% of discectomies.

In this study, gram-negative bacilli, particularly A. baumannii was the most common organism. The most common pathogen in Van de Beek et al.’s (1), McClelland and Hall (2), Korinek et al. (3, 13), Ermann et al. (7), Zarrouk et al. (9), Apisarnthanarak et al. (14), Logigan et al.’s (15) reports was Staphylococci, particularly Staphylococcus aureus, whereas some recent reports have shown the higher rates of gram-negative organisms (10, 11, 16, 20).

The present study showed that 76% of A. baumannii isolates were carbapenem-resistant. These rates were remarkably higher than Gaynes and Edwards (21), Rahbar and Hajian (22), Metan et al. (23), Logigan et al. (15), Baek-kim et al. (24), Cescio et al. (25), Vahdani et al. (26), Aminzadeh and Yaghubi (27), Khan et al. (28), and Moon et al.’s (29) findings. Mortality rate varies in different studies from 5% to 39% (8, 15, 16, 28-30). Unfortunately, the mortality rate of our patients was higher (45%). The majority of meningitis caused by A. baumannii led to death.

Our research and other studies (21-29) revealed that the rate of postneurosurgical meningitis due to A. baumannii and its resistance to broad-spectrum antibiotics, particularly to carbapenems has been increased. Carbapenem-resistant A. Baumannii is a matter of great concern in terms of treatment (23-25, 27). The major predisposing factors are previous carbapenems administrations, ICU admittance, emergency neurosurgeries, total parenteral nutrition, using device, and long duration of hospitalization (31, 32). Baek-Kim showed that the postneurosurgical infections due to A. baumannii are more resistant (24). Inappropriate use of antibiotics as a prophylaxis or empirical treatment, contamination of
operation room, ICU, or laboratory, and changing pattern of infectious agents in postneurosurgical meningitis are the possible reasons that necessitate further research (10).

In conclusion, in our study we found a high rate of post neurosurgical meningitis due to *Acinetobacter baumannii* resistant to carbapenems. This finding emphasizes the importance of preventive strategies against *gram-negative bacilli*, especially *A. baumannii*. The changing pattern of infectious agents in postneurosurgical meningitis over time suggests the necessity of other studies to give the most up-to-date insight to physicians.

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**References**

1. Van de Beek D, Drake JM, Tunkel AR. Nosocomial Bacterial Meningitis. N Engl J Med 2010; 362: 146-54.
2. McClelland S 3rd, Hall WA. Postoperative Central Nervous System Infection: Incidence and Associated Factors in 2111 Neurosurgical Procedures. Clin Infect Dis 2007; 45: 55-9.
3. Korinek AM, Baugnon T, Golmard JL, et al. Risk Factors for Adult Nosocomial Meningitis after Craniotomy: Role of Antibiotic Prophylaxis. Neurosurgery 2006; 59: 126-33.
4. NNIS System. National Nosocomial Infection Surveillance (NNIS) System Report, Data Summary from January 1992 through June 2003, issued August 2003. Am J Infect Control 2003; 31: 481-98.
5. Chang SM, Parney IF, McDermott M, et al. Glioma Outcomes Investigators. Perioperative Complications and Neurological Outcomes of First and Second Craniotomies among Patients Enrolled in the Glioma Outcome Project. J Neurosurg 2003; 98: 1175-81.
6. Vernet E, Adell C, Trilla A, et al. Usefulness of Risk Indexes for the Prediction of Surgical Site Infection in Patients Undergoing Neurosurgical Procedures. Med Clin (Barc) 2004; 122: 92-5. [In Spanish]
7. Erman T, Demirhindi H, Gocer AI, et al. Risk Factors for Surgical Site Infections in Neurosurgery Patients with Antibiotic Prophylaxis. Surg Neurol 2005; 63: 107-13.
8. Tängden T, Enblad P, Ullberg M, Sjölin J. Neurosurgical Gram-Negative Bacillary Ventriculitis and Meningitis: A Retrospective Study Evaluating the Efficacy of Intraventricular Gentamicin Therapy in 31 Consecutive Cases. Clin Infect Dis 2011; 52: 1310-6.
9. Zarrouk V, Vassor I, Bert F, et al. Evaluation of the Management of Postoperative Aseptic Meningitis. Clin Infect Dis 2007; 44: 1555-9.
10. Talebi Taher M, Molla Hosseini R, Fotokian AA. The Frequency of Bacterial Meningitis after Brain Surgery, Firoozgar Hospital (1999-2004). Razi J Med Sci 2006; 13: 151-8. [In Persian]
11. Malekpour-Afsar R, Karamoozian S, Shafei H. Post Traumatic Meningitis in Neurosurgery Department. Am J Infect Dis 2009; 5: 21-5.
12. Korinek AM, Golmard JL, Elcheick A, et al. Risk Factors for Neurosurgical Site Infections after Craniotomy: A Critical Reappraisal of Antibiotic Prophylaxis on 4,578 Patients. Br J Neurosurg 2005; 19: 155-62.
13. Korinek AM. Risk Factors for Neurosurgical Site Infections after Craniotomy: A Prospective Multicenter Study of 2944 Patients. The French Study Group of Neurosurgical Infections, the SEHP, and the C_CLIN Paris-Nord Service Epidemiologie Hygiene et Prevention. Neurosurgery 1997; 41: 1073-81.
14. Apisarnthanarak A, Jones M, Waterman BM, et al. Risk Factors for Spinal Surgical-Site Infections in a Community Hospital: A Case-Control Study. Infect Control Hosp Epidemiol 2003; 24: 31-6.
15. Logigan C, Mihalache D, Turcu T. Clinical Study of 57 Cases of Nosocomial Meningitis. J Prevent Med 2008; 16: 59-68.
16. Srinivas D, Veena Kumari HB, Somanna S, Bhagavatula I, Anandappa CB. The Incidence of Postoperative Meningitis in Neurosurgery: An Institutional Experience. Neurol India 2011; 59: 195-8.
17. Cockerill FR, Wilkler MA, Alder J, et al. Clinical and Laboratory Standards Institute (CLSI). M07-A9 Methods
for dilution antimicrobial susceptibility tests for bacteria that grow aerobically, approved standard – ninth edition, vol 32, No2. 2012. Available at: URL: http://shopping.netsuite.com/c.1253739/site/sample-pdf/m07A9-sample.pdf. Accessed May 10, 2014.

18. Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a single disc method. Am J Clin Pathol 1966; 45: 493-6.

19. Cockerill FR, Wikler MA, Older J, et al. Clinical and Laboratory Standard Institute (CLSI). M100-S17. Performance standards for antimicrobial susceptibility testing: seventeenth informational supplement; vol32, No 3. 2012. Available at: URL: http://antimicrobans.com.ar/ATB/wp-content/upload/2012/11/M100S22E.pdf. Accessed May 15, 2014.

20. Brown EM. The Management of Neurosurgical Patients with Postoperative Bacterial or Aseptic Meningitis or External Ventricular Drain-Associated Ventriculitis Infection in Neurosurgery working party of the British society for Antimicrobial Chemotherapy. Br J Neurosurg 2000; 14: 7-12.

21. Gaynes R, Edwards JR. National Nosocomial infections surveillance system. Overview of Nosocomial Infections Caused by Gram-Negative Bacilli. Clin Infect Dis 2005; 41: 848-54.

22. Rahbar M, Hajian M. Detection and Quantitation of the Etiologic Agents of Ventilator-Associated Pneumonia in Endotracheal Tube Aspirates from Patients in Iran. Infect Control Hosp Epidemiol 2006; 27: 884-5.

23. Metan G, Alp E, Aygen B, Sumerkan B. Carbapenem-Resistant Acinetobacter Baumannii: An Emerging Threat for Patients with Post-Neurosurgical Meningitis. Int J Antimicrob Agents 2007; 29: 112-3.

24. Kim BN, Peleg AY, Lodise TP, et al. Management of Meningitis Due to Antibiotic-Resistant Acinetobacter Species. Lancet Infect Dis 2009; 9: 245-55.

25. Cascio A, Conti A, Sinardi L, et al. Post-Neurosurgical Multidrug Resistant Acinetobacter Baumannii Meningitis Successfully Treated with Intrathecal Colistin. A New Case and a Systemic Review of the Literature. Int J Infect Dis 2010; 14: 572-9.

26. Vahdani P, Yaghoubi T, Aminzadeh Z. Hospital Acquired Antibiotic-Resistant Acinetobacter Baumannii Infections in a 400-Bed Hospital in Tehran, Iran. Int J Prev Med 2011; 2: 127-30.

27. Aminzadeh Z, Yaghoubi T. Drug-Resistant Post-Neurosurgical Nosocomial Acinetobacter Baumannii Meningitis in Two Iranian Hospitals. Afr J Biotechnol 2012; 11: 4083-4.

28. Khan FY, Abukhattab M, Baager K. Nosocomial Postneurosurgical Acinetobacter Baumannii Meningitis: A Retrospective Study of Six Cases Admitted to Hamad General Hospital, Qatar. J Hosp Infect 2012; 80: 176-9.

29. Moon C, Kwak YG, Kim BN, Kim ES, Lee CS. Implications of Postneurosurgical Meningitis Caused by Carbapenem-Resistant Acinetobacter Baumannii. J Infect Chemother 2013 19: 916-2.

30. O’Neil E, Humphreys H, Phillips J, Smyth EG. Third-Generation Cephalosporin Resistance among gram-Negative Bacilli Causing Meningitis in Neurosurgical Patients: Significant Challenges in Ensuring Effective Antibiotic Therapy. J Antimicrob Chemother 2006; 57: 356-9.

31. Baran G, Erbay A, Bodur H, et al. Risk Factors for Nosocomial Imipenem-Resistant Acinetobacter Baumannii Infections. Int J Infect Dis 2008; 12: 16-21.

Cisneros JM, Rodriguez-Bano J, Fernandez-Cuenca F, et al. Risk Factors for the Acquisition of Imipenem-Resistant Acinetobacter Baumannii in Spain: A Nationwide Study. Clin Microbiol Infect 2005; 11: 874-9.