Association between p53 Expression and Amount of Tumor-Infiltrating Lymphocytes in Triple-Negative Breast Cancer

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Triple-negative breast cancer (TNBC) is a molecular subtype of breast cancer that lacks expression of estrogen and progesterone receptors and does not show overexpression of human epidermal growth factor 2. This profile has no targeted therapy or standard biomarkers, and its aggressive phenotype, lower overall survival, and shorter relapse-free intervals than other breast cancer subtypes make its clinical management a challenge.1,2 Different biomarkers for TNBC as prognostic or predictive indicators and possible targets for novel therapy have been studied.1,2,3 Insight into the connection between the immune system and breast cancer may improve treatments and outcomes.

We previously demonstrated that TNBC tumors characteristically possess more tumor-infiltrating lymphocytes (TILs) than other breast cancer subtypes and TILs are positively correlated with endoplasmic reticulum stress-associated molecules.4-8 The endoplasmic reticulum is responsible for protein folding in cells.9,10 In tumor environments, the combination of high cancer cell proliferation rates, nutrient deficiency, and hypoxia lead to accumulation of unfolded or misfolded proteins in the endoplasmic reticulum that induce endoplasmic reticulum stress. Tumor cells activate the unfolded protein response and various downstream endoplasmic reticulum stress signaling pathways to adapt to this environment.9,10 Recent studies show that endoplasmic reticulum stress-associated molecules play important roles in tumorigenicity in TNBC.9

The TP53 gene is a tumor suppressor gene that regulates the cell cycle, cell proliferation, DNA repair, cellular senescence, and death by apoptosis.11 Cells with somatic TP53 mutations can avoid apoptosis and progress to malignant tumor cells. Tumors with TP53 mutations are highly invasive, poorly differentiated, and have a high histologic grade, showing poor response to chemotherapy.12 In solid cancers, mutations affecting the protein-
encoding reading frame, often referred to as null mutations, result in p53 protein absence.11 On the contrary, TP53 missense mutations may lead to the production of a mutant p53 protein, which has a prolonged half-life relative to the normal isoform that leads to its accumulation in tumor cells and makes it readily detectable by immunohistochemistry.11,13-15 Mutant p53 proteins not only lose the tumor suppressor function of wild-type p53 but also acquire new functions not present in the wild-type protein, termed gain-of-function properties, that promote tumorigenesis. So far, mutant p53 gain-of-function properties have been shown to stimulate tumor cell proliferation, migration, invasion, survival, chemoresistance, cancer metabolism, and tissue architecture disruption.16

Both types of mutations (null and missense) have been observed in the same cancer type.11 TP53 mutations are seen in 18%–25% of primary breast cancers2,17 and in approximately 80% of TN-BCs, which is markedly more frequent than in other breast cancer subtypes.2,18,19 Mutations in TP53 are predominantly missense mutations, producing mutant p53 proteins. Furthermore, as the mutant protein in malignant cells is less susceptible to degradation than wild-type p53, its accumulation establishes the TP53 mutation as an attractive therapeutic target for TNBC.20,21

Considering that TNBC is highly correlated with TIL levels, endoplasmic reticulum stress-associated molecules, and TP53 mutation rate, we examined the correlations among TIL levels, endoplasmic reticulum stress-associated molecules, and expression of p53 in TNBC.

MATERIALS AND METHODS

Patients and tissue specimens

A total of 678 TNBC patients who underwent surgery for primary breast cancer between 2004 and 2010 at Asan Medical Center in Seoul, Korea were retrospectively selected. In this group, 470 patients did not present with lymph node metastasis, and they received four cycles of adjuvant anthracycline (60 mg/m² adriamycin) and cyclophosphamide (600 mg/m²). The remaining 208 patients presented with lymph node metastases and were treated with four cycles of Adriamycin, followed by either four cycles of paclitaxel (175 mg/m²) or four cycles of docetaxel (75 mg/m²). The remaining 208 patients presented with lymph node metastases and were treated with four cycles of adriamycin, followed by either four cycles of paclitaxel (175 mg/m²) or four cycles of docetaxel (75 mg/m²). In total, 548 patients (80.8%) received radiotherapy. The median follow-up period was 78.3 months. Clinicopathologic information and survival data were obtained from medical records and surgical pathology reports. Exemption from informed consent after de-identification of information was approved by the Institutional Review Board of Asan Medical Center (2013-0866).

Histological evaluation

Two pathologists (H.J.L. and G.G.) reviewed whole sections of the hematoxylin and eosin-stained slides for histologic grade, pT category, pN category, and necrosis in the invasive area. Additionally, the levels of stromal TILs were evaluated, using full sections in 10% increments (defined as the mean percentage of plasma cells and lymphocytes in stroma of invasive carcinoma; if < 10% area, then 0, 1, or 5% level criteria were used).22

Tissue microarray construction and immunohistochemical evaluation

Available formalin-fixed paraffin-embedded blocks of 678 cases were arrayed with a tissue-arraying instrument, as previously described.8 Tissue microarray sections were stained with an automatic immunohistochemical staining device (Benchmark XT, Ventana Medical Systems, Tucson, AZ, USA). Antibodies to target phospho-eukaryotic initiation factor 2a (p-eIF2a), protein kinase RNA-like endoplasmic reticulum kinase (PERK), X-box binding protein-1 (XBP1), and CD8 were used, and their expressions in the tumor cells were determined, as previously described.8,23 An additional antibody to target p53 (1:1,500, Dako, Glostrup, Denmark) was also used. The level of p53 expression was ranked on a 4-point intensity scale (0, none; 1, mild; 2, moderate; and 3, intense). The percentage of nuclear expression of the p53 was also measured. An “immunoreactive score” was generated as the product of the intensity and the percentage of positive cells.

Statistical analysis

All statistical analyses were performed using SPSS statistical software ver. 18.0 (SPSS, Chicago, IL, USA). The chi-squared test, linear-by-linear association test, Spearman’s correlation, Mann-Whitney U test, and log-rank test were used to evaluate the data. All tests were two-sided and statistical significance was set at p < .05.

RESULTS

Clinicopathologic characteristics of the study population

All 678 patients were female, and their median age was 47 years at diagnosis (range, 23 to 76 years). Histologic grades 1 and 2 occurred in 160 cases (23.6%), and grade 3 in 518 cases (76.4%). There were 299 pT1 tumors (44.1%), 353 pT2 tumors (52.1%), 25 pT3 tumors (3.7%), and one pT4 tumor (0.1%). Most (n = 470) of the patients did not have pathologic lymph node metastasis (pN0, 69.3%) while 120 tumors (17.7%) were
pN1, 46 tumors (6.8%) were pN2, and 42 tumors (6.2%) were pN3. The tumors were categorized into four groups based on TIL amounts: < 10% TILs (164 patients, 24.2%), ≥ 10%; < 30% TILs (154 patients, 22.7%); ≥ 30%; and < 60% TILs (148 patients, 21.8%), ≥ 60% TILs (212 patients, 31.3%). Necrosis in the invasive area was identified in 488 of the patients (72.0%).

Classifying p53 expression as none, low, or high

Immunohistochemical analysis of p53 expression was scored as follows (Fig. 1): 0, no expression (299 cases, 44.1%); > 0 to 240, low expression (136 cases, 20.1%); and > 240 to 300, high expression (243 cases, 35.8%).

Characteristics of tumors according to p53 expression

Compared to the low p53 expression group, the high p53 expression group was significantly associated with higher histologic grade (p < .001), higher TIL levels (p = .009), increased presence of necrosis in the invasive area (p = .010), and higher expression of two endoplasmic reticulum stress-associated molecules (p-eIF2a, p = .013 and XBP1, p = .007) (Table 1, Fig. 2). Compared to the no p53 expression group, the high p53 expression

| Parameter                        | p53 expression level | Low (%) | No (%) | High (%) | p-value   |
|----------------------------------|----------------------|---------|--------|----------|-----------|
|                                  |                      | Low vs no | Low vs high | No vs high |           |
| Histologic grade                 |                      | .011     | < .001 | .088     |           |
| 1 and 2                          | 48 (35.3)            | 70 (23.4) | 42 (17.3) |           |
| 3                                | 88 (64.7)            | 229 (76.6) | 201 (82.7) |           |
| pT category                      |                      | .103     | .430   | .408     |           |
| 1                                | 70 (51.4)            | 121 (40.5) | 108 (44.5) |           |
| 2                                | 61 (44.9)            | 168 (56.2) | 124 (51.0) |           |
| 3                                | 5 (3.7)              | 9 (3.0)   | 11 (4.5)    |           |
| 4                                | 0                    | 1 (0.3)   | 0          |           |
| pN category                      |                      | .510     | .636   | .161     |           |
| 0                                | 95 (69.9)            | 199 (66.6) | 176 (72.4) |           |
| 1–3                              | 41 (30.1)            | 100 (33.4) | 67 (27.6)    |           |
| TIL                              |                      | .524     | .009   | .032     |           |
| < 10%                            | 39 (28.7)            | 72 (24.1)   | 53 (21.8)    |           |
| ≥ 10% and < 30%                  | 33 (24.3)            | 77 (25.8)   | 44 (18.1)    |           |
| ≥ 30% and < 60%                  | 33 (24.3)            | 65 (21.7)   | 50 (20.6)    |           |
| ≥ 60%                            | 31 (22.7)            | 85 (28.4)   | 96 (39.5)    |           |
| Necrosis in the invasive area    |                      | .579     | .010   | .015     |           |
| Negative                         | 46 (33.8)            | 92 (30.8)   | 52 (21.4)    |           |
| Positive                         | 90 (66.2)            | 207 (69.2)  | 191 (78.6)   |           |
| p-eIF2a                          |                      | .174     | .013   | .191     |           |
| Low                              | 79 (59.3)            | 151 (51.4)  | 108 (45.4)   |           |
| High                             | 55 (41.0)            | 143 (48.6)  | 130 (54.6)   |           |
| PERK                             |                      | .300     | .740   | .487     |           |
| Low                              | 77 (57.0)            | 153 (51.5)  | 132 (54.8)   |           |
| High                             | 58 (43.0)            | 144 (48.5)  | 109 (45.2)   |           |
| XBP1                             |                      | .137     | .007   | .088     |           |
| Low                              | 86 (64.7)            | 167 (56.6)  | 119 (50.0)   |           |
| High                             | 47 (35.3)            | 128 (43.4)  | 119 (50.0)   |           |

TIL, tumor-infiltrating lymphocyte.
group was significantly associated with higher TIL levels ($p = .032$). Compared to the low p53 expression group, the no p53 expression group was significantly associated with higher histologic grade ($p = .011$). The TIL levels of the no p53 expression group did not show significant differences from the TIL levels of the low p53 expression group ($p = .524$).

There were differences in the amounts of TILs (Fig. 3A) and the average number of CD8$^+$ T cells (Fig. 3B) among the three groups of TNBC patients classified according to p53 expression level. The high p53 expression group had significantly higher amounts of TILs ($p = .009$ and $p = .015$) and more CD8$^+$ T cells ($p = .001$ and $p = .015$) than the low and no expression groups, respectively. However, there was no significant difference between the no and low p53 expression groups in the amounts of stromal TILs ($p = .492$) and the number of CD8$^+$ T cells ($p = .133$).
Association between high histologic grade and endoplasmic reticulum stress-associated molecules

Since tumors with high histologic grades have an increased proliferation rate, which induces endoplasmic reticulum stress, we also analyzed the relationship between histologic grade and expression of endoplasmic reticulum stress-associated molecules. High histologic grade was significantly associated with two out of three endoplasmic reticulum stress-associated molecules (p-eIF2a, p = .036 and XBP1, p < .001, but not PERK, p = .928) (Table 2).

Prognostic significance of p53 expression in TNBC

Univariate analysis was undertaken to elucidate the prognostic significance of p53 expression. There was no significant difference in disease-free (p = .406) or overall survival rates (p = .444) among the three groups of no, low, and high p53 expression (Fig. 4).

Prognostic significance of TILs among groups based on p53 expression in TNBC

Since the level of TILs in TNBC patients has been recognized as a significant prognostic factor,24 the effect of TILs in each p53 expression group was examined. In the high p53 expression group, the higher the TIL level, the better the disease-free (p = .006) and overall survival rates (p = .005) (Fig. 5). Disease-free (p = .001) and overall survival curves (p = .001) were more significantly separated when the TILs were divided by 10%. In the no p53 expression group, the higher the TIL level, the higher the disease-free (p = .044) and overall survival rates (p = .046). Conversely, in the low p53 expression group, disease-free (p = .092) and overall survival rates (p = .108) were not significantly different, according to the TIL levels.

DISCUSSION

Our previous studies demonstrated that TILs are highly associated with interferon- and endoplasmic reticulum stress-associated molecules.5,8,25 Based on these results, we hypothesized that high p53 expression, which is linked with a high histologic grade (frequent mitosis, large cell and nuclear size, and poor differentiation), might induce endoplasmic reticulum stress and subsequent interferon signaling pathway activation, as well as the influx of TILs. Moreover, high p53 expression, which is related to necrosis, might induce the release of damage-associated molecules, an immune response, and the influx of TILs. The present research analyzed the clinical and pathological significance of p53 expression in TNBC. In comparison to low p53 expression, high p53 expression was shown to be associated with high histologic grade and endoplasmic reticulum stress-associated molecules.
p53 and Tumor-Infiltrating Lymphocytes

Darb-Esfahani et al. revealed a significant association between p53 protein expression characterized by immunohistochemical staining intensity ("wild-type pattern", tumor cell nuclei stained as variable and weak intensity; "overexpression", ≥ 60% of tumor cell nuclei stained as uniformly strong or moderate intensity; "loss," tumor cell nuclei stained completely negative) and the TP53 mutation, with 80% of the p53 overexpression group possessing the missense mutation. Also, in TNBC, missense TP53 mutations were significantly linked with higher levels of stromal TILs (p = .028) and CD8A gene expression (p = .020), and tended to be associated with a better survival (p = .093) compared to all other types of mutations.

TNBC has a higher TIL level than other breast cancer subtypes. However, the level of TIL varies within TNBC. TP53 mutations occur more frequently in TNBC (80%) than other breast cancer subtypes. Of the TP53 mutations, missense mutations produce a new mutant protein that can be presented on the cell surface via major histocompatibility complex (neoantigen), triggering the immune system and leading to a TIL response. This hypothesis is supported by our finding that the TIL levels of p53 overexpression group (probably due to missense mutation) were significantly higher than the TIL levels in the p53 low expression group (wild-type TP53) (p = .009). In contrast, null mutations of the TP53 gene simply do not produce normal levels of p53 protein, and the p53 function is lost. Since it does not make a new mutant protein to act as a neoantigen, it does not trigger the immune response, resulting in unchanged TIL levels. This hypothesis is supported by our finding that the no p53 expression group (probably due to null mutation) did not show a significant difference in TIL levels compared to those of the low p53 expression group (p = .524), while TIL levels in the no p53 expression group showed a significant difference compared to TIL levels in the high p53 expression group (p = .032).

Although TNBC has a high mutation burden and other gene mutations can enhance immunogenicity, our findings suggest an important role of TP53 mutation in the immunogenicity of TNBC.

In TNBC patients, a high TIL level is associated with good prognosis. In our study, p53 overexpression was related to TIL levels, but not with prognosis. Although there have been many attempts to correlate TP53 mutation status and clinical outcomes, such as overall survival rates in TNBC patients, conflicting findings have often emerged, with either poor survival in TNBC patients with TP53 mutation or no significance. This is

Fig. 5. Kaplan-Meier survival analyses of triple-negative breast cancer patients according to tumor-infiltrating lymphocytes levels in low (A, D), no (B, E), and high (C, F) p53 expression groups.
probably due to the beneficial effects of mutant p53 gain-of-function for tumorigenesis that offsets the favorable prognostic impacts of high TIL levels.

Mutant p53 proteins have been regarded as attractive targets for cancer therapy. Most of the strategies developed to target mutant p53 proteins involve restoration to wild-type p53 activity and depletion of the mutant p53 protein. Small molecules, such as 2,2-bis(hydroxymethyl)-quinuclidin-3-one or zinc metallo-chaperone-1, are representative restoration methods to wild-type p53 activity. Among exemplary methods for depletion of the mutant p53 protein, geldanamycin, 17AAG, and ganetespib are inhibitors of heat shock protein 90, which, in turn, inhibits the degradation of mutant p53 protein mediated by carboxyl terminus of heat shock cognate protein 70–interacting protein and murine double minute 2 (MDM2).16,33

In the current study, overall and disease-free survival rates were much better when the level of TILs was ≥ 10% compared with < 10% in the high p53 expression group, suggesting that the TP53 missense mutation is associated with TIL influx and some TILs can recognize mutant p53. Therefore, in the high p53 expression groups with TIL levels < 10%, the identification of T cells with T cell receptors (TCRs) reactive to mutant p53 and development of engineered T cell adoptive immunotherapy targeting mutant p53 may lead to successful immunotherapy. For instance, Lheureux et al.33 found immunogenic TP53 “hotspot” mutations as well as T cells with mutant p53-reactive TCRs in seven ovarian cancer patients. Therefore, in TNBCs with TP53 mutations, genetic transfer of the TP53 mutation-specific TCRs into autologous lymphocytes could be used to generate cells for use in cancer adoptive cell transfer immunotherapy.

The present study demonstrated close associations between the expression of p53 and the molecules associated with endoplasmic reticulum stress and TIL influx. Further studies targeting mutant p53 could facilitate the development of efficient immuno-therapeutic agents.

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**Conflicts of Interest**

The authors declare that they have no potential conflicts of interest.

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