Benefits of soil biochar amendments to tomato growth under saline water irrigation

Dongli She1,2, Xiaoqin Sun3, Agbna H. D. Gamareldawla1, Elshaikh A. Nazar2, Wei Hu1,3, Khaembah Edith3 & Shuang’en Yu1

Biochar amendments have been used in agriculture to improve soil fertility and enhance crop productivity. A greenhouse experiment was conducted to test the hypothesis that biochar amendment could also enhance the productivity of salt-affected soils. The trial was conducted over two consecutive growing seasons to investigate the effect of biochar amendment (four application rates as: B1 = 0%, B2 = 2%, B3 = 4%, and B4 = 8% by mass of soil) on yield and quality of tomatoes grown in a silt loam soil using non-saline water (I0 = 0.7 dS m−1) and saline water (I1 = 1 dS m−1, I2 = 3 dS m−1) irrigation. Furthermore, the study investigated the mechanism by which biochar addresses the salt stress on plant. The results showed that soil productivity as indicated by the vegetative growth and tomato yield components was adversely and significantly affected by saline water irrigation (P < 0.05). Tomato yield decreased from 689 ± 35.6 to 533 ± 79.0 g per plant as salinity of irrigation water increased from I0 to I2. Then, biochar amendment increased vegetative growth, yield, and quality parameters under saline irrigation water regimes, and ameliorated the salt stresses on crop growth. The highest (8.73 ± 0.15 and 4.10 ± 0.82 g kg−1) and the lowest (8.33 ± 0.08 and 2.42 ± 0.76 g kg−1) values of soil pH and soil organic matter were measured at B4I0 and B1I2 treatments, respectively. Also, the highest rate of biochar amendment combining with non-saline water irrigation (B4I0) produced tomato with the highest plant photosynthetic (17.08 ± 0.19 μmol m−2 s−1) and transpiration rate (8.16 ± 0.18 mmol H2O m−2 s−1). Mechanically, biochar amendment reduced transient sodium ions by adsorption and released mineral nutrients such as potassium, calcium, and magnesium into the soil solution. Therefore, biochar amendments have the potential in ameliorating salt stress and enhancing tomato production.

Water shortage represents a serious risk to global food security1-3. The total global water withdrawal for agricultural, domestic and industrial use is expected to increase by 23% from 1995 to 20254. Irrigated agricultural production in a number of the foremost populated areas of the world, such as China and Pakistan are expected to face extreme water crisis in the near future.

Irrigation is hailed as an intervention to enable agricultural production in areas where rainfall is inadequate, but scarcity of fresh water means some farmers are forced to irrigate plants using poor quality water5. This is particularly so where saline water is irrigated and then exacerbating the problem on agricultural fields that suffer the effects of salinization. Currently, about 33% of the global irrigated lands are affected by salinization6. Salinity affects plant growth by influencing plant physiological processes including photosynthesis and transpiration7. Salt stress decreased soil water potential (osmotic stress), causing an ion toxicity and consequently plant death. Decreased osmotic stress reduces water uptake by plants, which usually through closing leaf stomata and decreasing transpiration, and this negatively affects plant growth by decreasing plant photosynthesis8. Therefore, a direct correlation of plant growth with soluble salt concentration and duration of stress has been identified9.

The impact of salt stress and approaches to diminish the negative effects has been a subject of a number of studies. Techniques involved in these studies range from management options to identifying plant characteristics...
that help mitigate the effects of salinity. For example, Jouyban\textsuperscript{10} investigated the use of different techniques i.e. scraping, flushing, and leaching to remove excess salt from root zone of plants. Others have targeted on the utilization of various irrigation techniques in decreasing salinity\textsuperscript{11}. Improvement of salt tolerance/resistance is another technique used to address the constraint of soil salinity\textsuperscript{12}. Researchers have employed methods to improve plant salt tolerance, including inoculating seeds with halotolerant plant-growth-promoting rhizobacteria, using plant growth regulators and developing salt resistant cultivars\textsuperscript{13–15}. However, these approaches can be unprofitable, limited by the high costs and labor-consuming requirements of that are involved in order to counter the salinization problems. More recently, applications of organic conditioners have become a more sustainable and popular approach for enhancing crop productivity in salt-affected soils\textsuperscript{5}.

Recent work has elaborated the potentiality of biochar to enhance soil fertility and improve crop productivity\textsuperscript{16}. Many studies conducted in different parts of the world have reported improved soil water holding capacity, nutrient availability to plants, and plant productivity as a result of using biochar amendment\textsuperscript{1,5,17–19}. Schmidt\textsuperscript{20} reported that a fourfold increase in pumpkin production after soil amended with biochar when mixed with and without cow urine. Kamman\textsuperscript{21} gave a molecular interpretation about the positive effect of biochar on crop production, where he found that development of acid and basic functional groups and organo-mineral complexes on the biochar-matrix surfaces which responsible for nutrients retention. Joseph\textsuperscript{22} also reported that organic coating on biochar explained its ability on nutrients retention. However, a few researches had been done to evaluate/investigate the potential of biochar amendment (BA) in reducing the soil soluble salt under saline water (SW) irrigation as well as enhancing fruit quality. For example, Usman\textsuperscript{5} used conocarpus biochar under SW irrigation to investigate the changes of soil nutrient availability and tomato growth. Additionally, Lashari\textsuperscript{23} concluded that BA increased the plant growth, biomass, and yield and also increased photosynthesis, nutrient uptake, although under salt stress. The feedstock of biochar is of the main factor determining the effects of BA on plant growth.

### Results

#### Effects of BA and SW irrigation on selected soil properties

The analysis of variance results indicated that both BA and SWIrrigation significantly \((P < 0.05)\) influenced soil EC, SOM and pH as presented in Table 1. The pattern across treatments is greater EC with greater biochar composition, irrespective of irrigation treatment. For example, in 2014, the EC values for the B\textsubscript{4} treatment increased in the order of 0.20 \pm 0.01, 0.85 \pm 0.07, 1.64 \pm 0.15 dS m\textsuperscript{-1} for the I\textsubscript{0}, I\textsubscript{1}, and I\textsubscript{2} irrigation treatments, respectively, while the corresponding values for the B\textsubscript{1} treatment were 0.82 \pm 0.07, 1.40 \pm 0.21, and 2.25 \pm 0.07 dS m\textsuperscript{-1}.

Similarly, soil pH and SOM increased significantly \((P < 0.05)\) by increasing BA rate. For instance, in 2014 and 2015, SOM for I\textsubscript{0}B\textsubscript{4} was 2.7 \pm 0.08 and 2.71 \pm 0.07, and these increased to, respectively 3.67 \pm 0.69 and 4.10 \pm 0.82 kg m\textsuperscript{-2} in I\textsubscript{0}B\textsubscript{4} when the soil amended with 8% biochar.

Bulk density (B\textsubscript{4}) increased significantly in SW irrigation treatments compared to I\textsubscript{0}, and decreased with increasing BA rate (Fig. 1). Treatment I\textsubscript{1} resulted in 13% and 14% lower B\textsubscript{4} under the B\textsubscript{4} biochar compared to the respective non-biochar control, for 2014 and 2015 seasons, respectively. The soil field capacity (FC) was also

| Irrigation regime | Biochar treatment | EC (dS m\textsuperscript{-1}) | pH | SOM (g kg\textsuperscript{-1}) |
|------------------|------------------|-----------------|-----|-----------------|
|                  |                  | 2014 | 2015 | 2014 | 2015 |
| I\textsubscript{0} | B\textsubscript{0} | 0.20\textsuperscript{D} | 0.57\textsuperscript{D} | 8.44\textsuperscript{D} | 8.46\textsuperscript{D} | 2.70\textsuperscript{D} | 2.71\textsuperscript{D} |
|                  | B\textsubscript{1} | 0.45\textsuperscript{E} | 0.75\textsuperscript{E} | 8.65\textsuperscript{E} | 8.69\textsuperscript{E} | 3.30\textsuperscript{E} | 4.00\textsuperscript{E} |
|                  | B\textsubscript{2} | 0.72\textsuperscript{F} | 0.72\textsuperscript{F} | 8.68\textsuperscript{F} | 8.71\textsuperscript{F} | 3.51\textsuperscript{F} | 3.81\textsuperscript{F} |
|                  | B\textsubscript{3} | 0.82\textsuperscript{G} | 0.94\textsuperscript{G} | 8.70\textsuperscript{G} | 8.73\textsuperscript{G} | 3.67\textsuperscript{G} | 4.10\textsuperscript{G} |
| I\textsubscript{1} | B\textsubscript{0} | 0.85\textsuperscript{A} | 1.05\textsuperscript{A} | 8.41\textsuperscript{A} | 8.42\textsuperscript{A} | 2.50\textsuperscript{A} | 2.63\textsuperscript{A} |
|                  | B\textsubscript{1} | 1.18\textsuperscript{B} | 1.61\textsuperscript{B} | 8.54\textsuperscript{B} | 8.55\textsuperscript{B} | 2.80\textsuperscript{B} | 3.10\textsuperscript{B} |
|                  | B\textsubscript{2} | 1.28\textsuperscript{C} | 1.90\textsuperscript{C} | 8.56\textsuperscript{C} | 8.56\textsuperscript{C} | 3.14\textsuperscript{C} | 3.34\textsuperscript{C} |
|                  | B\textsubscript{3} | 1.40\textsuperscript{D} | 2.10\textsuperscript{D} | 8.58\textsuperscript{D} | 8.58\textsuperscript{D} | 3.44\textsuperscript{D} | 3.71\textsuperscript{D} |
| I\textsubscript{2} | B\textsubscript{0} | 1.64\textsuperscript{E} | 1.90\textsuperscript{E} | 8.31\textsuperscript{E} | 8.33\textsuperscript{E} | 2.30\textsuperscript{E} | 2.43\textsuperscript{E} |
|                  | B\textsubscript{1} | 1.82\textsuperscript{F} | 1.99\textsuperscript{F} | 8.40\textsuperscript{F} | 8.42\textsuperscript{F} | 2.40\textsuperscript{F} | 2.70\textsuperscript{F} |
|                  | B\textsubscript{2} | 2.15\textsuperscript{G} | 2.40\textsuperscript{G} | 8.43\textsuperscript{G} | 8.44\textsuperscript{G} | 2.45\textsuperscript{G} | 2.73\textsuperscript{G} |
|                  | B\textsubscript{3} | 2.25\textsuperscript{H} | 2.72\textsuperscript{H} | 8.45\textsuperscript{H} | 8.46\textsuperscript{H} | 2.76\textsuperscript{H} | 2.90\textsuperscript{H} |
| Biochar          | ***              | ***              | ns   | ns   | ***      |
| Salinity         | ***              | ***              | *    | *    | ***      |
| Interaction      | ns               | ns               | ns   | ns   | ns       |
affected significantly by both SW irrigation and BA treatments. It was decreased significantly in SW treatment compared to I0 treatment. However, FC was greater in all BA treatments compared to non-biochar. Figure 1 also shows the soil permanent wilting point (PWP) as influenced by SW irrigation and BA treatments. Soil PWP increased significantly by SW irrigation. However, BA decreased PWP in both I1 and I2 soil compared to their non-biochar control.

Available water content (AWC) for plant decreased significantly under I1 and I2 compared to I0, and increased with the increased rate of BA, for all the SW treatments (Fig. 1). Maximum AWC was observed in the combination of B4I0.

The effects of BA and SW irrigation on plant growth. The measured plant growth parameters were significantly ($P < 0.05$) affected by both BA and SW irrigation as shown in Table 2. These growth parameters generally increased significantly with increases in the BA rate and decreased significantly with increases

| Irrigation regime | Biochar treatment | FAGB (g) | DAGB (g) | FBGB (g) | DBGB (g) | LRWC (%) |
|------------------|------------------|---------|---------|---------|---------|----------|
|                  |          | 2014    | 2015    | 2014    | 2015    | 2014    | 2015    | 2014    | 2015    |
| I0               | B1       | 240.2aD | 302.9aD | 127.5aD | 190.5aD | 25.40aC | 6.32aB  | 92.72aD | 94.00aC |
|                  | B2       | 264.3aC | 327.3aC | 146.6aC | 153.8aC | 26.45aC | ND      | 6.33aB  | 93.80aC |
|                  | B3       | 289.7aB | 352.7aB | 156.7aB | 219.7aB | 27.88aB | 7.22aA  | 94.12aB | 95.07aB |
|                  | B4       | 307.3aA | 383.6aA | 165.7aA | 228.7aA | 29.75aA | 7.58aA  | 95.48aA | 96.10aA |
| I1               | B1       | 181.6aD | 259.9aD | 125.2aD | 188.2aD | 22.09aD | 6.18aC  | 91.52aD | 92.14aD |
|                  | B2       | 248.0aC | 311.0aC | 125.5aC | 199.5aC | 25.14aC | ND      | 6.88aB  | 92.04aB |
|                  | B3       | 285.5aB | 348.5aB | 133.8aB | 192.9aB | 25.11aC | 6.71aB  | 92.59aB | 94.54aB |
|                  | B4       | 300.6aA | 372.3aA | 146.1aA | 209.1aA | 27.63aA | 6.92aA  | 94.29aA | 95.24aA |
| I2               | B1       | 172.0aD | 251.5aD | 57.49aD | 143.5aD | 20.99aD | 5.76aB  | 90.61aD | 90.90aC |
|                  | B2       | 288.6aC | 61.5aC  | 147.5aC | 22.80aC | ND      | 6.14aB  | 91.57aC | 92.47aC |
|                  | B3       | 285.5aB | 228.6aB | 70.69aB | 163.7aB | 24.67aB | ND      | 6.34aA  | 92.02aA |
|                  | B4       | 300.6aA | 286.9aA | 67.78aA | 206.8aA | 24.20aA | 6.92aA  | 92.89aA | 93.51aA |

Biochar *** *** *** *** ***
Salinity ** ** ** ** **
Interaction ns ns ns ns ns

Table 2. Mean values of various plant growth parameters for different saline water irrigation regimes and biochar amendments during the 2014 and 2015 tomato growing seasons. Note: fresh above-ground biomass (FAGB), dry above-ground biomass (DAGB), fresh below-ground biomass (FBGB), dry below-ground biomass (DBGB), and leaf relative water content (LRWC). In each column, different uppercase and lowercase letters indicate a significant difference among the biochar application rates and irrigation salinity levels, respectively, at $P < 0.05$ (Least significant difference, LSD); Analysis of variance (ANOVA): ns, not significant; *, **, and ***, denote significance at $P \leq 0.05$, $P \leq 0.01$, and $P \leq 0.001$, respectively. ND, no data. B1, B2, B3, and B4 represent mixtures of soil with 0%, 2%, 4% and 8% of biochar by mass. I0, I1 and I2 represent irrigation water salinity of tap water, 1 and 3 dS m$^{-1}$. 

Figure 1. Bulk density, Bd (a); Field capacity, FC (b); Available water content, AWC (c) and Permanent wilting point, PWP (d) for different irrigation water salinity and biochar treatments for season 2014 and 2015. B1, B2, B3 and B4 represent mixtures of soil with 0%, 2%, 4% and 8% of biochar by mass. I0, I1 and I2 represent irrigation water salinity of tap water, 1 and 3 dS m$^{-1}$. 

Scientific Reports | (2018) 8:14743 | DOI:10.1038/s41598-018-33040-7
salinity of SW ($P < 0.05$). This was reflected by greater biomass observed for I0B4 treatments in both growing seasons. However, among all the measured growth parameters, there were no significant interaction effects of BA and SW.

The $Pn$ and $Tr$ rates were influenced by BA rates and SW irrigation (Fig. 2). Plants irrigated by saline water had significantly lower leaf photosynthetic compared with the control. Within each irrigation treatment, addition of biochar resulted in significant increases in photosynthetic rate. The highest ($17.08 \pm 0.19 \mu$mol m$^{-2}$ s$^{-1}$) and lowest ($13.0 \pm 0.25 \mu$mol m$^{-2}$ s$^{-1}$) $Pn$ rates were observed for treatments I0B4 and I2B1, respectively. Transpiration rate followed a similar trend with I0B4 recording the highest value ($8.16 \pm 0.18$ mmol H$_2$O m$^{-2}$ s$^{-1}$) while the lowest one ($4.85 \pm 0.27$ mmol H$_2$O m$^{-2}$ s$^{-1}$) was recorded under I2B1. The interaction effects of BA and SW on the $Pn$ and $Tr$ rates were statistically not significant.

Effects of BA and SW irrigation on tomato yield. Both BA rate and SW irrigation significantly ($P < 0.05$) influenced tomato yield and the number of fruits per plant (NFP). In both growing seasons, these two yield parameters significantly increased with decreases in irrigation water salinity and increased with increases in BA rate (Fig. 3). For example, in the 2014 control irrigation treatment (I0), the B1 treatment resulted in an increase of 6 more fruits plant$^{-1}$ and 272 g greater mass than the B1 treatment. Increased salinity had a negative effect on yield. For the B1 treatment in 2014 for example, the yield decreased from $689 \pm 35.6$ to $533 \pm 79.0$ g plant$^{-1}$ as salinity increased from I0 to I3, which was due to the decreased NFP from 21 to 15. There was no interaction significant ($P > 0.05$) effect of BA and SW on yield.

Fruit quality as affected by BA and SW irrigation. Generally, the concentrations of total soluble solid (TSS), soluble sugar content (SS), titratable acidity (TA), vitamin C content (VC), and fruit color index (CI) increased with either BA rate increasing or SW irrigation (Table 3). Overall, there were no significant ($P > 0.05$) interaction effects of BA and SW irrigation on fruit quality parameters. Significant differences in the TSS content were observed among both BA and SW irrigation treatments. The results elaborated that the I0B4 combination gave the greatest values of TSS and VC while I2B1 had the lowest values for these quality measures. Statistically there were no significant effects on CI value, but numerically increases in SW decreased the CI value.
Figure 3. Yield (a,b) and number of fruits per plant (c,d) in 2014 and 2015, respectively, for different irrigation water salinity and biochar treatments. For each year, different uppercase and lowercase letters indicate a significant difference among different biochar application rates (for a given irrigation regime) and salinity levels, respectively, at $P < 0.05$. B$_1$, B$_2$, B$_3$ and B$_4$ represent mixtures of soil with 0%, 2%, 4% and 8% of biochar by mass. I$_0$, I$_1$ and I$_2$ represent irrigation water salinity of tap water, 1 and 3 dS m$^{-1}$.

| Irrigation regime | Biochar treatment | TSS (Brix %) | VC (mg 100 g$^{-1}$) | TA (g 100 g$^{-1}$) | SS (g 100 g$^{-1}$) | SS/TA | CI |
|-------------------|-------------------|--------------|----------------------|---------------------|-------------------|-------|----|
|                   |                   | 2014         | 2015                 | 2014                | 2015              | 2014  | 2015 | 2014  | 2015 |
| I$_0$             | B$_1$             | 4.0$^{cC}$   | 5.0$^{cC}$           | 4.5$^{cC}$          | 4.7$^{cD}$        | 0.35$^{cC}$ | 0.33$^{cC}$ | 1.14$^{cC}$ | 1.12$^{cD}$ | 3.3$^{aA}$ | 3.5$^{aA}$ | 1.95$^{aA}$ | 1.96$^{aA}$ |
|                   | B$_2$             | 5.0$^{cA}$   | 5.3$^{cA}$           | 4.7$^{cA}$          | 5.0$^{cA}$        | 0.36$^{cC}$ | 0.37$^{cC}$ | 1.16$^{cB}$ | 1.18$^{cB}$ | 3.2$^{aA}$ | 3.2$^{aA}$ | 1.91$^{aA}$ | 1.97$^{aA}$ |
|                   | B$_3$             | 5.5$^{cA}$   | 5.5$^{cA}$           | 4.9$^{cA}$          | 5.2$^{cA}$        | 0.39$^{cA}$ | 0.40$^{cA}$ | 1.20$^{cA}$ | 1.20$^{cA}$ | 3.0$^{aA}$ | 3.0$^{aA}$ | 1.93$^{aA}$ | 1.97$^{aA}$ |
|                   | B$_4$             | 5.5$^{cA}$   | 5.7$^{cA}$           | 5.3$^{cA}$          | 5.5$^{cA}$        | 0.40$^{cA}$ | 0.41$^{cA}$ | 1.20$^{cA}$ | 1.24$^{cA}$ | 3.0$^{aA}$ | 3.0$^{aA}$ | 1.95$^{aA}$ | 1.99$^{aA}$ |
| I$_1$             | B$_1$             | 5.5$^{cA}$   | 6.0$^{cA}$           | 4.9$^{cC}$          | 5.3$^{cD}$        | 0.37$^{cA}$ | 0.39$^{cA}$ | 1.19$^{cB}$ | 1.28$^{cB}$ | 3.2$^{aA}$ | 3.3$^{aA}$ | 1.86$^{aA}$ | 1.94$^{aA}$ |
|                   | B$_2$             | 5.8$^{cA}$   | 6.0$^{cA}$           | 5.1$^{cC}$          | 5.5$^{cC}$        | 0.41$^{cB}$ | 0.45$^{cB}$ | 1.24$^{cB}$ | 1.29$^{cB}$ | 3.0$^{aA}$ | 2.9$^{aA}$ | 1.88$^{aA}$ | 1.95$^{aA}$ |
|                   | B$_3$             | 6.2$^{cA}$   | 6.2$^{cA}$           | 5.2$^{cA}$          | 5.7$^{cA}$        | 0.44$^{cB}$ | 0.47$^{cB}$ | 1.31$^{cA}$ | 1.31$^{cA}$ | 3.0$^{aA}$ | 2.9$^{aA}$ | 1.89$^{aA}$ | 1.97$^{aA}$ |
|                   | B$_4$             | 6.3$^{cA}$   | 6.0$^{cA}$           | 5.6$^{cA}$          | 5.9$^{cA}$        | 0.46$^{cB}$ | 0.48$^{cB}$ | 1.31$^{cB}$ | 1.34$^{cB}$ | 2.9$^{cA}$ | 2.9$^{cA}$ | 1.90$^{aA}$ | 1.98$^{aA}$ |
| I$_2$             | B$_1$             | 6.7$^{cB}$   | 6.7$^{cB}$           | 5.7$^{cB}$          | 6.1$^{cB}$        | 0.41$^{cB}$ | 0.42$^{cB}$ | 1.33$^{cA}$ | 1.38$^{cA}$ | 3.2$^{aA}$ | 3.2$^{aA}$ | 1.85$^{aA}$ | 1.90$^{aA}$ |
|                   | B$_2$             | 6.7$^{cB}$   | 6.8$^{cB}$           | 6.1$^{cB}$          | 6.2$^{cB}$        | 0.45$^{cB}$ | 0.46$^{cB}$ | 1.38$^{cA}$ | 1.42$^{cA}$ | 3.1$^{aA}$ | 3.1$^{aA}$ | 1.85$^{aA}$ | 1.90$^{aA}$ |
|                   | B$_3$             | 7.0$^{cB}$   | 7.0$^{cB}$           | 6.4$^{cB}$          | 6.7$^{cB}$        | 0.50$^{cA}$ | 0.51$^{cB}$ | 1.39$^{cA}$ | 1.42$^{cA}$ | 2.8$^{cA}$ | 2.8$^{cA}$ | 1.86$^{aA}$ | 1.91$^{aA}$ |
|                   | B$_4$             | 7.3$^{cB}$   | 7.0$^{cB}$           | 6.8$^{cB}$          | 6.9$^{cB}$        | 0.50$^{cA}$ | 0.52$^{cA}$ | 1.43$^{cA}$ | 1.46$^{cA}$ | 3.1$^{aA}$ | 3.1$^{aA}$ | 1.88$^{aA}$ | 1.93$^{aA}$ |
| Biochar           |                   | *            | ns                   | ***                 | ***               | ***    | ***   | ***    | ***   | *      | ***    | ns     | ns    |
| Salinity          |                   | ***          | ***                  | ***                 | ***               | ***    | ***   | ***    | ***   | ***    | ***    | ns     | ns    |
| Interaction       |                   | ns           | ns                   | ns                  | ns                | ns     | ns    | ns     | ns    | ns     | ns     | ns     | ns    |

Table 3. Mean values of tomato quality parameters including total soluble solids (TSS), soluble sugar (SS), titratable acids (TA), vitamin C (VC), color index (CI), and sugar/acid content ratio (SS/TA) for different saline water irrigation treatments and biochar amendments during the 2014 and 2015 growing seasons. Note: In each column, different uppercase and lowercase letters indicate a significant difference among the biochar application rates and irrigation salinity levels, respectively, at $P < 0.05$ (Least significant difference, LSD); Analysis of variance (ANOVA): ns, not significant; *, **, and *** denote significance at $P \leq 0.05$, $P \leq 0.01$, and $P \leq 0.001$, respectively. B$_1$, B$_2$, B$_3$ and B$_4$ represent mixtures of soil with 0%, 2%, 4% and 8% of biochar by mass. I$_0$, I$_1$, and I$_2$ represent irrigation water salinity of tap water, 1 and 3 dS m$^{-1}$. 
Biochar application rates and irrigation salinity levels, respectively, at P ≤ 0.05 (Least significant difference, LSD); Analysis of variance (ANOVA); ns, not significant; *, **, and *** denote significance at P ≤ 0.05, P ≤ 0.01, and P ≤ 0.001, respectively. B1, B2, and B3 represent mixtures of soil with 0%, 2%, and 4% of biochar by mass. I0, I1, and I2 represent irrigation water salinity of tap water, 1 and 3 dS m⁻¹, respectively.

Table 4. Mean values of soil cation contents (g kg⁻¹) for different biochar application rates (B) and irrigation water salinity levels (I). Note: In each column, different uppercase and lowercase letters indicate a significant difference among the biochar application rates and irrigation salinity levels, respectively, at P ≤ 0.05 (Least significant difference, LSD); Analysis of variance (ANOVA); ns, not significant; *, **, and *** denote significance at P ≤ 0.05, P ≤ 0.01, and P ≤ 0.001, respectively. B1, B2, and B3 represent mixtures of soil with 0%, 2%, and 4% of biochar by mass. I0, I1, and I2 represent irrigation water salinity of tap water, 1 and 3 dS m⁻¹.

| Irrigation regime | Biochar treatment | Ca    | K     | Mg    | Na    |
|-------------------|-------------------|-------|-------|-------|-------|
| I0                | B1                | 0.07a | 0.01bc| 0.01bc| 0.05ab|
|                   | B2                | 0.35ab| 0.09b | 0.04a | 0.04ab|
|                   | B3                | 0.49a | 0.23ab| 0.05a | 0.02a |
might cause the plant to regulate metabolic processes that decreased the production of sucrose and organic acids and/or reallocated them to different plant parts, particularly to the fruits, and thus increased the concentration gradient of sucrose from leaves to fruits45. Similarly, values of the fruit quality parameters increased significantly as BA rate increased. Of particle note is that BA increased the TA content in all the irrigation treatments. These enhancements in fruit quality may be attributed to the BA which affected the root distribution46. Although rooting depth was not measured in this study, the BA increased root biomass. Usman4 obtained similar results, who found that BA increased both TSS and VC slightly under both SW and non-SW irrigation treatments. Moreover, Akhtar47 and Agebna16 reported that BA improved the quality of tomato under deficit irrigation.

Mechanisms by which BA ameliorates salt stress on plants. This study provided evidences that BA can be utilized in salt-affected soils and/or when irrigation water is of low quality. It is probable that BA ameliorated negative impacts of salinity in the tomato plants by three main mechanisms: 1) reducing transient sodium ions by adsorption; 2) releasing mineral nutrients such as potassium, calcium, and magnesium into the soil solution (Table 4); and 3) decreasing osmotic stress by improving the soil AWC35. Novák47 stated that BA has strong absorptive characteristics binding refer to its high porosity, surface area and cation exchange capacity. By adsorbing toxic ions, especially sodium, and/or by releasing more beneficial ions18, BA can therefore reduce the negative effects of salt stress on plants, either by decreasing the exposure of plants to stress agents or by mitigating the stress responses of plants. Soil water content can be increased because that BA increased soil water holding capacity, especially increasing the proportion of larger pores where water is held at lower potentials allowing plants to uptake water more readily (Fig. 1)47. Increasing water availability might also interpret the alleviation of salt stress observed46. Some studies on photosynthetic responses to biochar additions have illustrated the increased water use efficiency at the leaf or whole plant scale46–38.

Conclusions
Saline water irrigation resulted remarkable decrement in tomato growth parameters and physiological processes such as Pn and Tr rates. Soil BA reduced the adverse impacts of salt stress, particularly at higher biochar application rates. The positive effects of BA on tomato growth were due to its capability to adsorb sodium ions and thus reducing its hazards and releasing mineral nutrients such as potassium, calcium, and magnesium into soil solution. Therefore, BA could be used on salt affected soils to improve tomato yield and quality, which would apply to saline-sodic soils elsewhere in the world. In particular, BA has the potential to be widely used combination with SW irrigation in agricultural production to struggle with fresh water crisis.

Materials and Methods
Study site. The experiment was carried out in a greenhouse from March 2014 to August 2015 at the Water-Saving Park of Hohai University (31°57′N, 118°50′E), China. The climate in the area is humid subtropical monsoon with annual mean pan evaporation and precipitation of 900 and 1073 mm, respectively. The mean daily temperature inside the greenhouse was 27.8°C. More details about this area can be found in Agebna44. The physio-chemical soil properties are displayed in Table 1.

Experimental design. To evaluate the combination effects of soil BA and SW irrigation on tomato growth, the experiment was conducted as a 3 × 4 factorial laid out in a completely randomized design with three replicates. Irrigation (I) water with three different salt concentrations were evaluated; I0 (control; tap water had electrical conductivity (EC) of 0.7 dS m−1), I1 (EC = 1 dS m−1) and I2 (EC = 3 dS m−1). Sodium chloride (NaCl) was mixed with tap water to prepare the irrigation water with a given salinity. Soil BA treatments comprising four different proportions of biochar in biochar-mixtures i.e. 0%, 2%, 4%, and 8% and designated as B0, B2, B4, and B8, respectively, were evaluated. The wheat straw biochar was used in this study, which was pyrolyzed at 350–550°C. Typically, 30% dry matter of the wheat straw would be converted into biochar48. The initial biochar properties are presented in Table 5.

Agronomic practices. Tomato (Lycopersicon esculentum Mill, Yazhoufenwang), which is a pink tomato, infinite growth variety, was chosen as the experimental crop. Seeds were sown on March 6 (2014, season 1) and March 10 (2015, season 2) in a nursery, respectively. The Seedlings were transplanted when plants had reached four-leaf stage (April 14 in 2014 and April 19 in 2015, respectively). During the growing period, the weigh method was utilized to verify water losses and then irrigating replace the lost amount of water. The total amounts of water irrigated in each treatment were 147.2 mm and 142.3 mm for 2014 and 2015 growing seasons, respectively. The agricultural applications during the tomato growth periods were the same as Agebna44.

Measurements of soil physio-chemical properties. At the end of each growing seasons (August 12 in 2014 and August 5 in 2015, respectively), soil physio-chemical properties were measured. For each treatment, a disturbed soil sample (~1 kg) was collected from the upper 20 cm layer, and air-dried. The air-dried samples were packed down to pass through a 1-mm mesh before being extracted using a 5:1 soil:leaching liquor to analyze soil pH, EC and cation contents (K+, Na+, Ca2+, Mg2+)49–54. Cation exchange capacity was determined using the ammonium acetate method52. Soil organic matter (SOM) content was measured using oil bath K2Cr2O7 titration method49–52. Soil total nitrogen (TN) was determined using the method described by Bremner53, and the total phosphorus (TP) was determined following Olsen and Sommers54. An undisturbed soil core was removed from the surface (0–20 cm) layer at each treatment for soil bulk density (Bd) measurement55. The core sampler had an internal diameter of 5 cm and height of 5 cm56. Field capacity (FC) and soil permanent wilting point (PWP) were determined as described by Michael47. Soil available water content (AWC) was obtained by subtracting values of PWP from FC58.
| Soil Properties | Soil | Biochar |
|-----------------|------|---------|
| pH              | 7.7  | 9.9     |
| Electrical Conductivity (dS m⁻¹) | 1.42 | 1.0 |
| Ca (g kg⁻¹)     | 0.16 | 0.016   |
| Mg (g kg⁻¹)     | 0.07 | 1.3     |
| Cl (g kg⁻¹)     | 0.11 | 1.44    |
| HCO₃ (g kg⁻¹)   | 0.20 | 0.85    |
| total N (g kg⁻¹) | 1.8  | 59      |
| total P (g kg⁻¹) | 0.66 | 144.3   |
| total K (g kg⁻¹) | 0.4  | 115     |
| CEC (cmol kg⁻¹) | 14.94| 217     |
| Bulk density (g cm⁻３) | 1.35 | 0.4    |
| Field capacity (%) | 25.8 | —      |
| Silt (%)        | 30.1 | —      |
| Sand (%)        | 50.2 | —      |
| Clay (%)        | 19.7 | —      |

Table 5. Basic physical and chemical properties of biochar and the soil in the study area.

Measurements of tomato growth and yield. Tomato growth parameters, including fresh above-ground (FAGB) and below-ground (FBGB) ground biomass, dry above- (DAGB) and below- (DBGB), were measured at the end of each growing seasons, following the details at Agebna. Plant leaf relative water content (LRWC) was measured at one day closing to the middle of the growing season (June 13 in 2014 and July 18 in 2015), utilizing the method describe by Smart. Two mature leaves in plant upper canopy of each treatment were selected to measure plant photosynthesis (Pn) and transpiration (Tr) rates. Four measurements were carried out in the growing season of 2014 (May 23, June 11, July 6 and July 17, respectively) using a portable photosynthesis system (Li-Cor, Lincoln, NE, USA) during the period of 09:00–12:00 h. Mean Pn and Tr rates were calculated from the four measurements.

Tomato fruits were harvested from July 18 to August 12 in 2014 and July 14 to August 5 in 2015. The number of fruits/plant (NFP) was accounted and weighed immediately to determine fruit mass (Yield). Fruits were homogenized using a blender, and chemical composition was quantified to determine fruit quality parameters, including parameters of total soluble solid (TSS, Brix %), titratable acidity (TA, g 100 g⁻¹), vitamin C content (VC, mg 100 g⁻¹ fresh mass, as ascorbic acid), soluble sugar content (SS, g 100 g⁻¹) and fruit color index (CI).

Statistical analysis. All the treatments were subjected to a two way analysis of variance (ANOVA) using the MSTATC statistical package. The Least Significant Difference (LSD; P ≤ 0.05) was used to determine differences between means.

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**Acknowledgements**
We acknowledge and are grateful for the financial support provided by the National Key Research and Development Program of China (2017YFC0403300-04), by the National Natural Science Foundation of China through grant No. 41471180, by the CAS “Light of West China” Program, by the Fundamental Research Funds for the Central Universities (2018B12214), and by the Qing Lan Project of Jiangsu Province.

**Author Contributions**
Dongli She and Shuangèn Yu wrote the main manuscript text. Xiaqin Sun and Agbna H. D. Gamaedalwa done the experiment. Elshaikh A. Nazar prepared figures. Wei Hu and Khaembah Edith revised the English.

**Additional Information**

**Competing Interests:** The authors declare no competing interests.

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