Towards robust in vivo quantification of oscillating biomagnetic fields using Rotary Excitation based MRI

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Spin-lock based functional magnetic resonance imaging (fMRI) has the potential for direct spatially-resolved detection of neuronal activity and thus may represent an important step for basic research in neuroscience. In this work, the corresponding fundamental effect of Rotary EXcitation (REX) is investigated both in simulations as well as in phantom and in vivo experiments. An empirical law for predicting optimal spin-lock pulse durations for maximum magnetic field sensitivity was found. Experimental conditions were established that allow robust detection of ultra-weak magnetic field oscillations with simultaneous compensation of static field inhomogeneities. Furthermore, this work presents a novel concept for the emulation of brain activity utilizing the built-in MRI gradient system, which allows REX sequences to be validated in vivo under controlled and reproducible conditions. Via transmission of Rotary EXcitation (tREX), we successfully detected magnetic field oscillations in the lower nano-Tesla range in brain tissue. Moreover, tREX paves the way for the quantification of biomagnetic fields.

According to the 2019 Global Burden of Diseases, Injuries, and Risk Factors Study, mental disorders were among the ten most common causes of global burden of diseases, as measured by disability-adjusted life-years. According to analysis of data of previous years, taking into account related suicides, mental disorders could even have a significantly more severe impact. An increase in the prevalence of mental disorders as a result of the influence of the COVID-19 pandemic is considered likely. One of the great goals of current research is to understand the functional principles of the brain, so that those disorders can be studied to understand the underlying mechanisms and to improve their treatment.

In recent years, brain imaging has made great progress. For example, the anatomical structures of the brain are well known from in vivo measurements such as diffusion-weighted magnetic resonance imaging (MRI). Diagnostics currently well-established for functional investigations include techniques that directly measure neuronal activity, such as electroencephalography (EEG) and magnetoencephalography (MEG). Indirect measurement via hemodynamic response is provided by blood oxygenation level dependent (BOLD) functional MRI (fMRI). However, for the precise investigation of functional connectivity and in-depth analysis of brain activity, noninvasive methods with improved spatial and temporal resolution compared to existing methods are required. Consequently, developing a novel method that both meets the requirements of spatiotemporal resolution and direct detection of neuronal activity is of great importance for the progress of brain research. A young branch of MRI research is therefore addressing direct mapping of neuronal activity based on spin-locked MRI techniques.

Direct imaging of neuronal currents occurring during certain activity patterns of the brain was initially attempted by detecting modulations of the relaxation time constant $T_1^*$ or a phase imposition. Since the main magnetic field in clinical scanners is in the range of 1.5...7 T, the system is resonant to oscillating fields in the RF range (radiofrequency, ≈ 1 MHz) and is therefore insensitive to neural oscillations in the Hz range. In a system with a field strength of about ≈ 1 μT, resonant energy absorption of fields with frequencies at the magnitude of Hz

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is possible. In such low-fields, however, the magnetization $M_0$ in thermal equilibrium is very small and thus poor signal-to-noise-ratio (SNR) is to be expected. In 2008, Witzel et al. proposed a completely new approach (SIRS, Stimulus-Induced Rotary Saturation) for detecting tiny oscillating magnetic fields in the Hz range. Instead of using a low-field device, Witzel exploited the rotary saturation effect first described by Redfield in 1955. Within the so-called spin-lock (SL) condition, in the presence of a continuous wave RF pulse rotating at the Larmor frequency, sensitivity to magnetic field fluctuations in the low frequency range can be created while maintaining high SNR at clinical field strengths. Here, the resonance condition can be shifted by adjusting the SL field strength. In subsequent studies, a modification of the SIRS sequence was proposed, which shows a significantly improved signal scaling behavior and provides sensitivity to the oscillation phase. Most importantly, this technique is not based on magnetization saturation, but on excitation of magnetization during spin-lock interaction. This concept, which we refer to hereafter as Rotary Excitation (REX), holds great potential for the direct detection of neuronal activity, since ultra-weak magnetic oscillations act like pseudo RF pulses in the rotating frame and thus can be detected via sinusoidal signal variations in a positive contrast evaluation (REX weighted contrast). A brief summary of the available studies on rotary saturation and rotary excitation targeting fMRI is provided in the Supplementary Material (S.1).

One of the major challenges in employing the REX technique to detect neuronal activity is its susceptibility to static field inhomogeneities. Improvements of basic SL techniques known from $T_1\rho$ quantification concerning the compensation of static field inhomogeneities, have not yet been investigated in the context of SIRS or REX experiments. However, compensation of static field inhomogeneities is essential for SL based detection of ultra-weak field oscillations, since static system imperfections are of the same order of magnitude as the dynamic components to be measured. Thus, low susceptibility to inhomogeneities combined with high sensitivity to dynamic magnetic field components is required. Most SIRS studies and REX studies were performed without elaborated compensation techniques. For SIRS, a ramped SL technique compensating for $B_0$ and $B_1^\perp$ imperfections has been introduced, while for REX measurements the widely used composite SL (C-SL) technique has already been applied. However, the influence of such compensation techniques on the measured REX signals has not been thoroughly investigated so far and furthermore approaches like rotary-echo SL (RE-SL), balanced SL (B-SL) and adiabatic excitation, have not been considered. In particular, there is evidence of a strong dependence of REX signals on the SL parameters used (e.g., choice of spin-lock pulse duration $t_{SL}$) and adiabatic excitation, which has not yet been considered for compensated REX techniques.

Another major challenge is the experimental validation of new SIRS or REX based methods. Since the interaction of a SL pulse with a weak magnetic field oscillation has to be measured, phantom experiments have been performed so far in which the existing MRI setup had to be equipped with additional hardware. Examples include field-generating loop-coils or dipole electrodes, which need precise calibration and RF shielding within the MRI room. Furthermore, accurate synchronization with the MRI sequence is required. These circumstances slow down technical development and pose a major challenge to reproducibility. In addition, new sequences cannot be tested on humans under in vivo conditions, as this would usually require certification of the developed hardware. Thus, in vivo validations under controlled conditions were not performed and the highly challenging measurements of neuronal biomagnetism have been attempted directly within the fMRI setup.

In this study, the basic REX technique is advanced in three steps. First, we investigate how SL compensation techniques can be used to robustly measure and quantify ultra-weak magnetic field oscillations to achieve direct detection of biomagnetic fields. Thereby, the detected REX signals could potentially be increased while reducing the influence of static field inhomogeneities. The different preparation characteristics of most common compensation techniques were examined by Bloch simulations. Particular attention was paid to the critical parameter of SL duration. Second, we propose a novel method for transmitting the rotary excitation effect onto the object under investigation (tREX, transmitted-Rotary-Excitation). Using ultra-weak gradient waveforms generated by the built-in MRI gradient system, the proposed method allows to mimic brain activity by transmitting artificial magnetic fields onto the living brain tissue. This not only enables reproducible, rapid validation measurements in phantoms but also a comparison of the compensation techniques under in vivo conditions in humans. Third, we demonstrate how this new technique can pave the way for quantification of biomagnetism, as tREX enables signal calibration in vivo.

Methods

In the present study, a thorough investigation of the rotary excitation effect for established spin-lock compensation techniques was performed using Bloch simulations, phantom validations and in vivo experiments in human brain tissue. To allow reproduction of our experiments, all measurements were performed using the new tREX concept. Furthermore, the MRI sequences used were implemented in the hardware-independent open-source Pulsed sequence framework based on Matlab (R2018b, The MathWorks Inc., Natick, Massachusetts, USA). The measurements were carried out on a clinical 3.0 T scanner (MAGNETOM Skyra, Siemens Healthcare, Erlangen, Germany) using a 20-channel head coil and the standard gradient system (max. gradient strength = 45 mT/m, max. slew rate = 200 T/m/s).

**REX preparation modules.** Spin-lock preparation modules are commonly classified by their compensation approaches for $B_0$ and $B_1^\perp$ field inhomogeneities. In this study, the following preparatory sequences were considered:

- standard continuous wave spin-lock (S-SL), no compensation
- rotary-echo spin-lock (RE-SL), $B_1^\perp$ compensation
• composite spin-lock (C-SL), single refocused \( B_0 \) and \( B_1^+ \) compensation\(^{19} \)
• balanced spin-lock (B-SL), double refocused \( B_0 \) and \( B_1^+ \) compensation\(^{21} \)

The sequence diagrams of the corresponding preparation modules are shown in the Supplementary Material (Supplementary Fig. 1). All four modules start with a 90° excitation pulse (‘y’ axis) rotating the magnetization to the x’ axis (notation in rotating frame coordinates). It is known from Definition that adiabatic excitation prior to spin-locking is highly beneficial as it allows preventive compensation of \( B_1^+ \) imperfections\(^{22} \). An identical \( B_1^+ \) optimized adiabatic-half-passage pulse (AHP, hyperbolic secant shape, duration 3 ms) was therefore used for all modules. Since the REX method aims at measuring ultra-weak magnetic field oscillations (e.g. \( \Delta B_0 \text{stim} = 0 \ldots 100 \text{nT} \)), the compensation of static \( B_0 \) inhomogeneities (e.g. \( \Delta B_0 \text{static} = 300 \text{nT} \) for 0.1 ppm at 3 T) during SL interaction is thus of great importance (\( \Delta B_0 \text{static} > \Delta B_0 \text{stim} \)). Among the modules studied, C-SL and B-SL provide compensation with respect to \( B_0 \) inhomogeneities due to their 180° refocusing pulses. Contrary to further assumptions or simplifications, the sinusoidal signal or by calculating the standard deviation of the signal for \( N \) different \( \phi \):

\[
S_{\text{REX}}(r, \phi) = a(r) \cdot \sin[\phi \cdot m(r) + \varphi_0(r)] \cdot \sqrt{2} + b(r)
\]

Since, off-resonance effects cannot be completely avoided during REX preparation or imaging, the measured signal might preserve an offset \( b(r) \) and can manifest a phase shift \( \varphi_0(r) \) with respect to \( \phi \). The sinusoidal signal is 2π-periodic only in the exact resonance case and therefore a modulation factor \( m(r) \) was introduced. The amplitude \( a(r) \) is primarily influenced by the strength of the oscillating field and is used as the measure of magnetic stimulus detection. This can be determined either by signal fitting with the sinusoidal function (Eq. 1), or by calculating the standard deviation of the signal for \( N \) different \( \phi \):

\[
A_{\text{REX}}(r) = |a(r)| = \sqrt{\frac{1}{N-1} \sum_{n=1}^{N} [S_{\text{REX}}(r, \phi_n) - S_{\text{REX}}(r)]^2}
\]

A complete REX experiment thus consists of the acquisition of typically \( N = 5 \ldots 20 \) REX weighted images with different phases \( \phi \) and the evaluation of the amplitude \( A_{\text{REX}} \) as an indicator for neuronal activity according to Eq. (2) in pixel-wise evaluations. In order to scale the REX amplitude, the magnitude was normalized with the reference signal \( S_0 \) of the imaging sequence without preparation. Simulation results were normalized accordingly with \( M_0 = 1 \).

Concept of tREX experiments. An essential part of the present study is the realization and validation of REX experiments without using external hardware such as loop-coils or dipole-electrodes for field generation\(^{15} \). Instead, we propose a new approach to study the interaction of ultra-weak magnetic field oscillations with spin-locking fields under in vivo conditions using exclusively the standardized hardware on a clinical MRI system. For this purpose, the built-in gradient system was utilized. The gradients, which usually cause spin dephasing for the aim of spatial encoding, were controlled to generate small magnetic oscillations in selected off-center slices. Thus, the effect of rotary excitation was transmitted (tREX) in the field of view (FOV) within an off-center slice to the magnets isocenter (Fig. 1b). Magnetic field oscillations with amplitudes in the range \( 1 \ldots 100 \text{nT} \) at a frequency of \( f_{\text{stim}} = 50 \text{Hz} \) were aspired in \( 10 \text{mm} \) offcenter slices. The gradient waveform \( G_z(t) \) was calculated as follows:

\[
G_z(t) = \frac{\Delta B_0 \text{stim}}{\Delta z} \cdot \sin[2\pi f_{\text{stim}} \cdot t + \phi]
\]

The tREX concept can also be carried out with the x- or y-gradient or with a superposition of all three gradients in an oblique slice. Here, tREX benefits from the very precise control of gradient coils and enables the full implementation of REX experiments within the Pulseq framework.

Bloch simulation of REX modules. The four REX modules were investigated with regard to their different preparation characteristics with respect to \( B_0 \) and \( B_1^+ \) inhomogeneities using numerical Bloch simulations. The work of Ueda et al. on the dynamics of magnetization under stimulus-induced rotary saturation serves as an important reference\(^{25} \). The basic concepts can be adopted for REX. In contrast to Ueda\(^{25} \), the Bloch equations, which are described for the REX case by a system of nonlinear differential equations, were solved numerically. A Runge–Kutta method (4th order, RK4) implemented in Matlab was used for this purpose. Details of the simulation can be found in the Supplementary Material (S.3).

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Figure 1. Sequence concept for validation of ultra-weak magnetic field detection using tREX for phantom or in vivo experiments (RF = radio-frequency event, $G_x$, $G_y$, $G_z$ = gradient field event). The upper part of the figure shows the pulse sequence using the B-SL preparation module. In the in vivo experiments, the $k$-space acquisition was performed with ramped flip angles and four interleaved spirals were used for a fast readout of the REX weighted contrast. In the phantom experiments, the spiral readouts were replaced by TSE readouts. Subfigures (a–c) explain the individual steps of magnetization behavior. After fat saturation, an adiabatic excitation pulse flips the magnetization to the transverse plane as shown in (a). This is followed by one of the four spin-lock preparation modules (B-SL is illustrated exemplarily). During SL preparation, the interaction with the ultra-weak magnetic field oscillation takes place. In the tREX approach the oscillating field is generated by sinusoidal waveforms of the built-in gradient system. To obtain a non-zero amplitude, the measurement must be performed in an offcenter slice, as shown in (b). The excitation process of the magnetization perpendicular to the spin-lock axis is dependent on the relative phase between spin-lock pulse and tREX waveform. At the beginning of the spin-lock pulse ($t_{SL} = 0$) the magnetization is aligned along the $x'$-axis. The perpendicular component increases due to rotary excitation effect and decays simultaneously with the relaxation time constants $T_{1\rho}$ and $T_{2\rho}$. After the preparation module, remaining transverse magnetization is crushed. The resulting longitudinal magnetization in (c) depends on the relative phase according to the trajectories shown in (b).
The first simulation investigates the influence of the spin-lock parameters $t_{\text{SL}}$ and $f_{\text{SL}}$ on the amplitude $A_{\text{REX}}$. When examining the behavior regarding $f_{\text{SL}}$, a resonance peak at $f_{\text{SL}} = f_{\text{stim}}$ is expected according to Eq. (5). In the second simulation, the $t_{\text{SL}}$ characteristics for variable $f_{\text{stim}}$ was investigated. In further simulations presented in the Supplementary Material, it was tested whether the relaxation times $T_{10}$ and $T_{20}$ or the field strength of the stimulus $\Delta B_0^{\text{stim}}$ have an influence on the $t_{\text{SL}}$ characteristics.

In a final simulation, the performance of the REX amplitude in the presence of $B_0$ field imperfections was investigated. For this purpose, a static magnetic field inhomogeneity component $\Delta B_0^{\text{static}} = 2\pi \Delta B_0^{\text{static}} = 2\pi \delta B_0^{\text{static}}$ was assumed (Eq. 5–6, Supplementary Material S.3).

**Phantom experiments.** For the phantom experiments, a gel phantom was constructed on agar–agar basis. The Agarose (Agar Kobe I, A2113, A. Hartenstein GmbH) was dissolved in demineralized water at a concentration $c = 0.75%$ and doped with 0.25 mmol/L Gd-DTPA (Gadopentetat-Dimeglumin, Magnograf). The gel was prepared in a cylindrical sample tube ($O = 9$ cm, $h = 16$ cm, $\approx 1$ L).

In initial preparatory experiments, it was investigated whether the ultra-weak gradient waveforms required for the tREX method can be generated with the standard built-in gradient system. The target was the transmission of magnetic field oscillations with $\Delta B_0^{\text{stim}} = 1$ … 100 nT, e.g. in a 10 mm offcenter slice. For this purpose, trajectory measurements were performed according to Eq. (3) in the Agarose phantom. Sinusoidal gradient waveforms (Eq. (3)) with a frequency $f_{\text{stim}} = 50$ Hz and amplitudes in the range 0.02 … 20 nT/mm (15 logarithmically spaced steps) were generated. The relative phase of the oscillating gradients was varied in the range 0 … 2$n\pi$ (10 linearly spaced steps). For the measurement of the gradient trajectories realized by the setup, the phase evolution was recorded in a thin offcenter slice (2 mm thickness) at a distance of 25 mm from the isocenter for $t_{\text{ADC}} = 30$ ms (50 kHz bandwidth). To correct for static off-resonance effects, the observed phase evolution was rectified by a reference scan (tREX gradients switched off). The gradient trajectory was subsequently determined from the discrete derivative of the phase evolution. The results of the trajectory measurements were compared with the set gradient stimuli.

For the verification of the rotary excitation effect transmitted by gradient oscillations, a series of experiments was performed in an axial tREX slice (thickness 5 mm, distance from the isocenter 10 mm) in the Agarose phantom with $\Delta B_0^{\text{stim}} = 50$ nT. REX weighted images with 30 relative phases $\phi = 0$ … $3\pi$ were acquired. For the pixel-wise determination of REX amplitude maps, both the standard deviation of the measured signal and a sinusoidal fit were evaluated in the range 0 … 2$n\pi$ for the first 20 phases. Control measurements were performed with the tREX gradients switched off (stimulus off). Additionally, the characteristic of the measured REX amplitude for different $t_{\text{SL}}$ was investigated and compared with the results of the Bloch simulation. Finally, the $t_{\text{SL}}$ characteristic was investigated in a ROI (region-of-interest) based evaluation. For imaging of the REX signals, a turbo spin echo (TSE) readout with centric encoding was used in all phantom experiments ($T_e = 12$ ms). Due to the phase encoding, REX experiments with identical relative phases had to be repeated several times. The FOV was $150 \times 150$ mm² and the matrix $96 \times 96$. The turbo factor of the TSE readout was 4, so $96/4 = 24$ identical REX preparations were performed for the acquisition of a single REX weighted image. A delay $T_{\text{ADC}} = 1$ s was used between the individual preparations with TSE readouts for longitudinal magnetization recovery.

In the present study, the $B_0$ field homogeneity was adjusted in the tREX slice by an extended 2nd order shimming routine prior to the actual field detection experiments. $B_0^+ + B_0^-$ mapping was performed in the same tREX slice with identical FOV settings for control. Thus, the spatially dependent off-resonance $\Delta f_0^{\text{static}} (\vec{r})$ was determined in the measured FOV. In order to study REX amplitude behavior under the influence of well-defined $B_0$ field inhomogeneities, an additional constant gradient was applied in $x$ direction during the SL interaction. The gradient amplitude was varied in different experiments in the range 0 … 32 Hz/cm (5 steps), corresponding to a maximum gradient strength of $\approx 75$ nT/mm.

**In vivo measurements.** All in vivo experiments were performed in brain tissue of a healthy volunteer. The participant was screened for contraindications to MRI and written informed consent was obtained. The study complied with the Helsinki Declaration on Human Experimentation and was approved by the local institutional ethics board (Ethics Committee of the Institute for Psychology of the Faculty for Human Sciences, University of Würzburg). For the tREX experiment, a slice (thickness 5 mm, distance from the isocenter 10 mm) was chosen that partly intersects the frontal sinus, which exhibits increased field inhomogeneities$^{27}$, in the frontal lobe. $B_0^+$ and $B_0^-$ mapping was carried out for control. Thus, the REX effect was observed in vivo under controlled conditions and the interference of natural field inhomogeneities with the detection of magnetic field oscillations in brain tissue could be investigated in detail.

For fast in vivo imaging of the REX effect, spiral acquisitions were carried out. In order to reduce the influence of $T_2^*$ relaxation and off-resonance effects, short readout durations ($t_{\text{ADC}} = 11$ ms, $T_X = 16$ ms) were used and the k-space was sampled in four segments using interleaved spirals and ramped flip angles ($\alpha_k = 30^\circ, 35^\circ, 45^\circ, 90^\circ$) for slice excitation (Fig. 1). This succession of flip angles produces an approximately uniform transverse magnetization in each interleave, while maximizing the measurement signal. A similar approach has already been used for $T_1$ quantification$^{28}$. Hereby, an approximate pure REX weighted contrast can be generated in the final images. For longitudinal magnetization recovery, a delay $T_{\text{ADC}} = 1$ s was inserted between each individual REX scan.

The tREX method was investigated in vivo in a comparative study considering best and worst case choice of SL pulse durations. The results of the Bloch simulation regarding $t_{\text{SL}}$ characteristics (Eq. 6) were taken into account for the choice of $t_{\text{SL}}$. In the best case measurements, the following $t_{\text{SL}}$ values were used: 77.5 ms (S-SL), 70 ms (RE-SL), 80 ms (C-SL), 80 ms (B-SL). For the worst case measurements, the following values were used: 82.5 ms (S-SL), 80 ms (RE-SL), 70 ms (C-SL), 60 ms (B-SL). For each measurement, pixel-wise REX amplitude
maps were calculated using the standard deviation (Eq. 2). In addition, for the best case measurements, amplitude maps were computed from the complex signal fit (Eq. 1).

Finally, it was examined whether the tREX method can be utilized as a calibration technique for the quantification of magnetic field oscillations. Measurements based on the B-SL preparation module were repeated for different stimulus field strengths. The measured amplitudes \( A_{\text{AREX}} \) were tested for correlation with \( \Delta B_{\text{stim}}^0 \) in a ROI-based evaluation and compared with the control amplitude (stimulus off). Linear regression was performed to test whether tREX is suitable for spatially dependent calibration of REX amplitudes and thus can be used for in vivo quantification of neuronal activity.

Ethics approval and consent to participate. All experimental procedures were in accordance with institutional guidelines and were approved by local authorities.

Results

Bloch Simulation: prediction of \( A_{\text{AREX}} \) maxima. Figure 2 shows the results of the \( t_{\text{SL}}/f_{\text{SL}} \) characteristics as heat maps for the different preparation modules. For all modules, as expected, the highest REX amplitudes were observed at \( f_{\text{SL}} = f_{\text{stim}} = 50 \) Hz in the resonance condition. However, the behavior of \( A_{\text{AREX}} \) when varying \( t_{\text{SL}} \) is more complex. In the resonance case, alterations of \( A_{\text{AREX}} \) can be observed. These are particularly distinct for the modules equipped with compensation techniques, e.g. with B-SL local maxima occur at \( t_{\text{SL}} \approx 40, 80, 120, 160 \) ms, whereas minima (\( A_{\text{AREX}} = 0 \)) occur at \( t_{\text{SL}} = 60, 100, 140, 180 \) ms. Thus, the periodic intervals \( \Delta t_{\text{SL}} \) of \( A_{\text{AREX}} \) maxima are: 9.8 ms for S-SL, 19.9 ms for RE-SL, 20.0 ms for C-SL, 40 ms for B-SL. The effect of minima/maxima formation was visually illustrated in the Supplementary Material (Supplementary Fig. 2).

In Fig. 3, the oscillation phenomenon can be examined for variable \( f_{\text{stim}} \). Here it can be seen that the periods are shortest for S-SL and doubled for RE-SL and C-SL. The period of B-SL is quadrupled compared to S-SL. The length of the periods depends on \( f_{\text{stim}} \) and a module dependent factor \( \eta \). The period length can be described by \( \Delta t_{\text{SL}} = \frac{\eta}{2f_{\text{stim}}} \). For S-SL and RE-SL, the positions of the maxima are shifted. The following approximation was empirically derived for the prediction of the \( k \)-th \( A_{\text{AREX}} \) maximum:

\[
t_{\text{SL}}(k) = \frac{\eta}{2f_{\text{stim}}} \cdot (k - \kappa)
\]

with \( k \in \mathbb{N} \),

\( \eta = 1 \) for S-SL, \( \eta = 2 \) for RE-SL, \( \eta = 2 \) for C-SL, \( \eta = 4 \) for B-SL, \( \kappa = 1/4 \) for S-SL, \( \kappa = 1/2 \) for RE-SL, \( \kappa = 0 \) for C-SL, \( \kappa = 0 \) for B-SL.

This rule applies for the modules RE-SL, C-SL and B-SL with a mean deviation of \( \approx 0.05\% \) in the simulated range. For S-SL, where minima and maxima are not distinct, the prediction is less accurate (\( \approx 4.6\% \)). The results concerning the influence of the \( T_{1\rho} \) and \( T_{2\rho} \) relaxation times on the position of the maxima are shown in the Supplementary Material (Supplementary Fig. 3). Here, only minor deviations were found with regard to the positions...
of the maxima. Essentially, short relaxation times hamper the measurement at high $t_{\text{SL}}$ times. The magnitude of the stimulus $\Delta B_0^{\text{stim}}$ determines the magnitude of the observed amplitude $A_{\text{REX}}$ with a clear linear correlation ($R^2 = 0.998$, Supplementary Fig. 4). No significant influence on $t_{\text{SL}}$ characteristic was found. Thus, Eq. (4) describes the occurrence of $A_{\text{REX}}$ maxima in a general way. Therefore, for $f_{\text{stim}} = 50$ Hz, the following $t_{\text{SL}}$ times were used in further simulations and measurements: 77.5 ms for S-SL, 70 ms for RE-SL, 80 ms for C-SL, 80 ms for B-SL.

**Phantom measurements.** The validation of ultra-weak gradient waveforms is reported in the Supplementary Material (Supplementary Fig. 5). The measurements confirm that the built-in gradient system enables magnetic field oscillations to be generated precisely with amplitudes 1 ... 20 nT/mm ($R^2 > 0.99$). In this range, both amplitude and phase could be set precisely. The mean deviation of the adjusted amplitude was $\pm 1.16\%$. A waveform with 0.1 nT/mm represents the limit. Here, a slight oscillation with high noise level could still be observed in the trajectory measurement. However, $R^2 = 0.67$ was found this case. Below this limit, no oscillation of the tREX gradient could be detected. Thus, tREX experiments up to 1 nT can be performed within a 10 mm offcenter slice.

Figure 4 presents the proof-of-concept of the tREX method. The expected sinusoidal signal for different phases $\phi$ could be clearly observed ($R^2 = 0.999$). However, a strong formation of banding artifacts is evident in the REX-weighted images for S-SL, which was significantly reduced for B-SL. All measurements were performed in the expected $A_{\text{REX}}$ maximum (according to Bloch simulations). Yet, differences in the calculated $A_{\text{REX}}$ maps are evident. B-SL ($A_{\text{REX}} = 0.1994 \pm 0.0089$) shows a more homogeneous and on average higher REX amplitude than S-SL ($A_{\text{REX}} = 0.173 \pm 0.033$). The REX amplitudes of RE-SL and C-SL were $A_{\text{REX}} = 0.165 \pm 0.037$ and $A_{\text{REX}} = 0.193 \pm 0.017$. The control measurement (stimulus off using B-SL) exhibited no significant oscillation ($A_{\text{off}} = 0.0060 \pm 0.0038$). From the pre-calibration scans, the following $B_0$ and $B_1^\phi$ field deviations were observed within the tREX slice: $\delta B_0 = -2.3 \pm 3.5$ Hz and $B_1^\phi = 96.4 \pm 6.8$ Hz. Here, $B_1^\phi$ represents the ratio of the measured to the nominal $B_1^\phi$ field strength. In the Supplementary Material, the corresponding maps $a(\vec{r}), b(\vec{r}), \phi(\vec{r})$ and $m(\vec{r})$ fitted with Eq. (1) were compared and the respective $\Delta B_0$ and $B_1^\phi$ field maps are attached (Supplementary Figs. 6–7).

Figure 5 depicts comparisons between the measured $t_{\text{SL}}$ characteristics and the Bloch simulation results for the resonance condition. The data show high agreement, confirming the existence of $t_{\text{SL}}$ minima and maxima. The following $\Delta t_{\text{SL}}$ periods were obtained in good agreement with the simulation results: 18.5 ± 0.8 ms for RE-SL, 18.8 ± 1.2 ms for C-SL, 38.6 ± 0.6 ms for B-SL. The shift of RE-SL vs C-SL was 9.3 ± 0.4 ms. No values were determined for S-SL due to the weak oscillation strength.

The results of the REX amplitude performance in presence of $B_0$ inhomogeneities are shown in Fig. 6 and were compared with the corresponding results of the Bloch simulation. Both measurements and simulations show a decrease in $A_{\text{REX}}$ with increasing $\delta B_0^{\text{static}}$, confirming that static field inhomogeneities hamper the measurement of dynamic field oscillations. However, the four preparation modules show distinctly different sensitivities to $B_0$ inhomogeneities. A halving of $A_{\text{REX}}$ was observed in the simulation at the following $\Delta f_0$ values: ±24.0 Hz for S-SL, ±31.7 Hz for RE-SL, 26.6 Hz for C-SL, 36.7 Hz for B-SL ($\pm 0.19$ ppm, $\pm 0.25$ ppm, $\pm 0.21$ ppm, $\pm 0.29$ ppm).
m, respectively at 3 T). Thus, B-SL exhibits the highest and S-SL the lowest sensitivity (improvement ≈ 53%). In the phantom experiments we observed (25 ± 2) Hz, (32 ± 2) Hz, (27 ± 2) Hz and (37 ± 2) Hz, respectively. These results agree well in simulations and experiments. To minimize the influence of deviation in the experimental validation, a ROI with small deviation (100.6 ± 3.2%) was evaluated.

In vivo measurements. Figure 7 presents the proof-of-concept of the tREX method in vivo in brain tissue. The expected sinusoidal signal of rotary excitation was clearly observed in the REX weighted images (R² > 0.99). No significant REX variation was detected in the control measurements (gradient stimulus off). In the calculated A⁰ maps, structures of brain tissue can be roughly distinguished. In two different ROIs, we observed A⁰ = 0.1192 ± 0.0037 and A⁰ = 0.0912 ± 0.0037. In the control experiments the reference amplitude was A⁰ = 0.00868 ± 0.00068 and A⁰ = 0.0076 ± 0.0025, respectively. The regions of skull bone and fat produced significantly reduced amplitudes.
The impact of tSL choice for B₀ and B⁺₁ compensated modules is demonstrated by the AREX maps in Fig. 8. While only minor differences were observed for S-SL, the remaining modules show clear differences between the best and worst case experiments. However, significant differences between the modules are also evident in the best case measurements. Local reductions in AREX were observed near the frontal sinus (for S-SL, RE-SL and C-SL) and a significant reduction is noticeable for RE-SL near the cranial bone. This is also evident in the box plot in Fig. 8, where AREX was examined throughout the whole brain tissue. B-SL shows the best overall performance and a clear delineation between best/worst case measurements. The amplitude parameter |a| fitted according to Eq. (1) yields highly comparable detection maps with approximately identical characteristics. On average over all maps, AREX was 3.3% above |a|. The R² values in brain tissue (best case) of the complex signal fit yielded values: 0.982 ± 0.012 for S-SL, 0.985 ± 0.014 for RE-SL, 0.9907 ± 0.0071 for C-SL, 0.9929 ± 0.0033 for B-SL. The remaining parameters of the fitted maps and the corresponding ΔB₀ and B⁺₁ field maps are attached in the Supplementary Material (Supplementary Figs. 9–10). Supplementary Fig. 8 presents the REX weighted images of the best/worst case measurements for all modules.

In the tREX calibration study, a clear linear correlation between the stimulus field strength ΔB⁺stim and the measured REX amplitude was observed under in vivo conditions. In Fig. 9, three exemplary ROIs were analyzed by means of linear regression and the agreement with a simple linear model yielded R² > 0.99 in each case. Pixel-wise linear regression revealed R² = 0.9977 ± 0.0094 in a whole brain tissue ROI. In addition, the whole brain tissue was evaluated specifically for small stimuli (ΔB⁺stim ≤ 10 nT). A slight rotary excitation effect could still be detected at 1 nT. Here, the average signal increase of A_{REX} compared to the reference measurement (stimulus

\[ \text{Figure 5. Experimental verification of Bloch simulation tSL characteristics in the phantom experiment. In (a), the results of the Bloch simulation are plotted. (b) The } A_{REX} \text{ amplitudes experimentally determined by a tSL variation measured in the Agarose phantom. Here, } t_{SL} \text{ was varied from 0…160 ms for each preparation module. To achieve feasible measurement times, six relative phases } \phi \text{ were varied per amplitude calculation. The respective } \Delta t_{SL} \text{ periods are noted in blue. Due to the low amplitude of the oscillation, no period duration was determined for the phantom results of the S-SL module. The qualitative behavior of } A_{REX} \text{ maxima could be observed in the phantom measurements in good agreement with the simulation results. The period lengths of the measurements are slightly smaller than in the simulation, which could be due to a deviation of the actual spin-lock amplitude (} B⁺_{inhomogeneity}). \text{ In the simulation and experiment, the periods are shortest for S-SL and doubled for RE-SL and C-SL. The period of B-SL is quadrupled.} \]
off) was 7.3%. However, significant delineation to the reference signal in the brain tissue ROI was only possible at \( \Delta B_{B_{\text{stim}}} \leq 2.5 \text{ nT} \).

Discussion

In the present work, a basic concept for the direct detection and quantification of neuronal activity by MRI has been advanced in three interdependent steps. The major concerns of the REX technique raised by Coletti\(^2\), namely the susceptibility to \( B_0 \) and \( B_+ \) inhomogeneities, were addressed by the implementation of established compensating spin-locking techniques\(^19\text{--}21\). Bloch simulations were used to progress the principle understanding of REX and its parameter dependency. The simulation results indicating significantly higher robustness of \( B_0 \) and \( B_+ \) compensated modules were confirmed in phantom experiments. For experimental validation purpose, the novel tREX approach was introduced. The transmission of rotary excitation using the built-in gradient system enables fast sequence development and validation with a high degree of precision and reproducibility without the need of external hardware. Based on this principle, initial validations of the REX technique were successfully performed in vivo and an approach for the quantification of biomagnetism can be proposed.

Experimental validation of the tREX waveforms could confirm that ultra-weak magnetic field oscillations can be generated with the standard built-in gradient system. While the timing of the phases is fully adequate for the purpose of the REX experiments, the amplitudes of the oscillations are limited, due to the precision of the hardware components. Ultimately, digital quantization of the electronic gradient control is the lower boundary. Minimum gradients of 0.1 nT/mm could only be just generated, whereby the detected waveforms were extensively
noisy ($R^2 = 0.67$). Thus, an oscillation of 1 nT within the isocenter near slice $\Delta z = 10\text{ mm}$ is currently the limit of feasibility. Above this limit (1 ... 20 nT/mm), reliable amplitude control and linearity could be demonstrated and the oscillations show the desired sinusoidal profile ($R^2 > 0.99$). These technical conditions must be considered in tREX experiments especially if measurements are to be performed in more distant offcenter slices. In addition, tREX represents only a simple mimic of the in general spatially complex neuronal fields, because only z-components are generated and phase dispersion within a voxel is not produced.

The simulation and measurement results concerning REX based field detection show that an appropriate choice of SL pulse duration depending on the stimulus frequency is mandatory for the compensated RE-SL, C-SL, and B-SL modules. For this, a prediction rule (Eq. 4) was empirically developed. The accuracy of the prediction is reliable with a deviation of $-0.05\%$. Only for S-SL the prediction is less accurate (deviation of $-4.6\%$). Here, relaxation effects seem to systematically shift the position of the $\text{AREX}_{\text{max}}$ maxima. For the compensated modules, no significant influence of relaxation on the positions of local maxima could be found. Here, spin dynamic effects are crucial, since by reversing the SL direction or by applying refocusing pulses, the magnetization trajectory is abruptly bifurcated. Thus, constellations exist in which the final magnetization is projected either completely on the $z$-axis (maximum condition) or completely on the $y'$-axis (minimum condition). This phenomenon was illustrated in the Supplementary Material (Supplementary Fig. 2). The positions of the maxima conditions could be largely confirmed in the phantom experiments. The average deviation from the predicted positions was 5.5\%. This can be explained mainly by a variation of the actual $B_0^{\text{inh}}$ field, whereby the SL amplitude is not exactly 50 Hz.

The present work addressed the question whether dynamic magnetic field oscillations can be measured with a simultaneous compensation of static field inhomogeneities. Bloch simulations verified that B-SL, for example, exhibits a significant increase in $B_0^{\text{inh}}$ robustness. A halving of the measured REX amplitude occurs from $\Delta B_0^{\text{inh}} = \pm 0.29$ ppm at 3 T. Thus, the REX method has functionality for typical $B_0$ distributions in brain tissue after standard shimming. This was also evident in vivo, as B-SL still provided high REX sensitivity even in regions of increased $B_0$ imperfections. The impact of $B_0^{\text{inh}}$ inhomogeneities was preventively compensated by using adiabatic excitation pulses as proposed in22. However, this only improves the initial conditions prior to the actual REX effect. A deviation in $B_0^{\text{inh}}$ still leads to a violation of the REX resonance condition ($f_{\text{SL}} \neq f_{\text{stim}}$). This cannot be fully compensated by any of the SL modules considered. Yet, in the simulations and measurements, B-SL indicated broader peaks in the $\text{AREX}$ characteristics, which effectively compensates for higher $B_0^{\text{inh}}$ deviations. This effect was also visible in the phantom experiment (Fig. 4). Despite an approximately optimal shim throughout the entire phantom ($\Delta f_0 = -2.3 \pm 3.5$ Hz), a decrease in $\text{AREX}$ was observed at the edge of the phantom. In this region, deviations of the $B_0^{\text{inh}}$ field ($\approx 90\%$, Supplementary Fig. 7) were present. In contrast to S-SL, B-SL generated
For separate 2D scans, slice-selective higher order shimming is recommended. Another alternative is to adjust the carrier frequency of the spin-lock pulse to the local Larmor frequency of a specific ROI in the cortex. The findings of simulation and phantom measurements were used to perform initial proof-of-concept experiments for oscillating magnetic field detection in vivo. Here, a spiral readout was used consistently, generating one REX-weighted image for each preparation. In contrast to the method proposed in, not a true single-shot readout was used, but an interleaved acquisition with ramped flip angles. This step makes the readout less susceptible to off-resonance effects and still provides a quasi-single-shot in the context of the preparation. Thus, the sequence presented in this work can be applied for the detection of neuronal activity, where the relative phase of the stimulus is unknown and full k-space sampling must be carried out for each preparation. Another advantage of this technique is the reduced \( T_2^* \) effect due to the shorter individual readouts. This could also be beneficial for the robust detection of neuronal fields at high field strength, as the influence of the BOLD effect is effectively reduced by shorter spiral acquisition times. A disadvantage of our readout is the additional impact of \( T_1 \) relaxation between the readouts and potential artifacts due to incorrect flip angles. Here, the use of Shinnar–Le-Roux optimized RF pulses and parallel imaging approaches could be explored for further improvement and acceleration.

Although in vivo REX detection with the simple S-SL module was successful in principle, it was demonstrated that the use of \( B_0 \) and \( B_1^+ \) compensated modules provides significant improvement. In the detailed evaluation of the measured REX signals by the fit according to Eq. (1), significant increase in performance can be observed for C-SL and B-SL (Supplementary Fig. 8). In brain tissue these techniques yield homogeneous \( A_{\text{AREX}} \) maps, low offsets of REX oscillation, and the signals are approximately 2π-periodic. Another finding was that RE-SL provided slightly better detection than C-SL in the region near the frontal sinus. This can be explained by the characteristics of the \( B_0 \) susceptibility, where RE-SL clearly differs from the other modules and shows improved sensitivity at
some high offresonance values. However, both in phantom and in vivo, RE-SL shows poor REX sensitivity at the edge of the object which is possibly linked to the $B^+_{1}$ distribution. Hence, in conclusion, we recommend to use at least the single refocused C-SL module or the double refocused B-SL module for in vivo REX experiments.

At present, a major challenge for REX detection lies in relatively high noise of the reference scans. In the reconstruction and evaluation of REX weighted images, the influence of respiration has not been corrected yet, which leads to increased $A_{\text{off}}^{\text{REX}}$ and hampers separability at low $\Delta B_{0}^{\text{stim}}$. Therefore, as has been indicated by Truong et al., the true limit of sensitivity of the REX technique might be lower than 1 nT and can be further improved in vivo. This important point needs to be investigated in future studies, for example, using t-test statistics and repetitive REX experiments. Further investigations should be carried out whether other physiological processes such as blood flow impact REX detection and whether, e.g., triggered measurements or measurements during breath hold are beneficial. For motion correction, image registration techniques could be adapted from established BOLD fMRI routines.

An important finding of our work is that tREX experiments establish a linear correlation ($R^2 > 0.99$) between the artificial stimulus field strength and the measured REX amplitude in vivo. Thus, tREX offers high potential as a calibration technique for the amplitude quantification of biomagnetic fields. A reasonable approach for a REX fMRI protocol would be to measure two $A_{\text{REX}}$ maps using tREX at different $\Delta B_{0}^{\text{stim}}$ and to calculate a tissue

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**Figure 9.** In vivo correlation measurements of the $A_{\text{REX}}$ amplitude and the stimulus field strength $\Delta B_{0}^{\text{stim}}$. The $A_{\text{REX}}$ values averaged within the marked ROIs for $\Delta B_{0}^{\text{stim}}$ between 1 and 100 nT (14 steps) from the maps in (a) are plotted against field strength in subfigure (b). Linear regression yielded $R^2$ values > 0.99 for all three exemplary ROIs. The slope of the tREX calibration varies in different ROIs. The corresponding pixel-wise $R^2$ map of the linear fit is also shown in (b). The averaged $R^2$ in the whole brain tissue ROI (dashed line) was 0.9977 ± 0.0094. Due to the high agreement with the linear fit, tREX experiments enable in vivo calibration of REX amplitudes. This fact can be exploited in a fMRI experiment for quantifying neuronal magnetic field oscillations. The $A_{\text{REX}}$ amplitude evaluated in the whole brain tissue, was examined more closely in the lower field strength range 1…10 nT in (c). The signal obtained for 1 nT cannot be significantly distinguished from the reference measurement (stimulus off) in the whole brain tissue. However, mean and median are both above the reference measurement amplitude (~7.3%). Above 2.5 nT, the results provide a significant increase in $A_{\text{REX}}$. A reason for the low separability at 1 nT could be the influence of respiration, as this increases the reference signal due to motion issues.
specific and spatially dependent REX calibration function. This can be utilized for the quantification of magnetic field oscillations in real measurements of neuronal activity by linking the measured REX amplitudes to the linear calibration function. It is essential to note that the REX calibration depends on the selected offcenter slice and ultra-low stimulus field strengths (<10 nT) cannot be generated throughout the whole volume of the brain due to the limited precision of the gradient system. For this reason, we propose for a robust determination of the calibration function to perform tREX measurements at stimulus field strengths well above this limit (e.g., 20 nT and 40 nT) and to exploit the linear correlation of the REX effect for the quantification of neuronal fields. However, a fixed limitation of the tREX method is that no calibrations can be performed in an isocenter slice. Thus, future fMRI studies should begin to trial tREX calibrations and subsequent REX detections in basic single-slice experiments near the isocenter.

In future studies, to achieve the next step for direct detection and quantification of neuronal fields, concepts for fMRI experiments based on a reliable method for modulating neural oscillations needs to be developed. One option for this is the state-dependent enhancement of alpha waves as suggested in4 or by triggering steady state visually evoked potentials (SSVEPs) with flickering stimuli34,35. Another promising application is the localization of the seizure onset zone in focal epilepsy36. While the expected stimulus frequencies in alpha activity and SSVEPs are below the 50 Hz oscillation used in this work, frequencies > 100 Hz can be expected in focal epilepsy. The stimulus frequency dependent prediction rule of \(A_{\text{REX}}\) maxima conditions found in this work can serve for optimization of the respective sequence parameters. As the relative phase with respect to the spin-lock is generally unknown, the REX amplitude must be determined by the standard deviation of random phase signals as discussed in12. A potential but technically complex alternative is the simultaneous recording of an EEG for retrospective evaluation and a subsequent detection by the REX fit presented in this work37. In this case, the R² value of the REX fit could be used for an improved validation and higher statistical significance of the measured amplitudes. For these ambitious goals, the present work provides basic MRI concepts, specific optimization approaches and a pulse sequence that is already fully suitable for in vivo studies.

Data availability
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Code availability
The original Pulseq MRI sequences for in vivo REX detection used in this work are provided in an open-access GitHub repository (https://github.com/maximiliangram/Pulseq_tREX).

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Author contributions

P. Albertova and M. Gram contributed equally to this work. Guarantors of integrity of entire study: P. Nordbeck, PM. Jakob; study concepts and study design: all authors; data acquisition and ethic approval: M. Gram, P. Albertova, M. Blaimer, M. Gamer, MJ Herrmann; experimental studies: M. Gram, P. Albertova, V. Schirmer; data analysis: M. Gram, P. Albertova; manuscript drafting and manuscript editing: all authors.

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Competing interests

The authors declare no competing interests.

Additional information

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