Identification of sources and bioaccumulation pathways of MeHg in subantarctic penguins: a stable isotopic investigation

Marina Renedo1,2, David Amouroux2, Zoyne Pedrero2, Paco Bustamante1,4 & Yves Cherel3

Seabirds are widely used as bioindicators of mercury (Hg) contamination in marine ecosystems and the investigation of their foraging strategies is of key importance to better understand methylmercury (MeHg) exposure pathways and environmental sources within the different ecosystems. Here we report stable isotopic composition for both Hg mass-dependent (e.g. $\delta^{202}$Hg) and mass-independent (e.g. $\Delta^{199}$Hg) fractionation (proxies of Hg sources and transformations), carbon ($\delta^{13}$C, proxy of foraging habitat) and nitrogen ($\delta^{15}$N, proxy of trophic position) in blood of four species of sympatric penguins breeding at the subantarctic Crozet Islands (Southern Indian Ocean). Penguins have species-specific foraging strategies, from coastal to oceanic waters and from benthic to pelagic dives, and feed on different prey. A progressive increase to heavier Hg isotopic composition ($\delta^{202}$Hg and $\Delta^{199}$Hg, respectively) was observed from benthic (1.45 ± 0.12 and 1.41 ± 0.06‰) to epipelagic (1.93 ± 0.18 and 1.77 ± 0.13‰) penguins, indicating a benthic-pelagic gradient of MeHg sources close to Crozet Islands. The relative variations of MeHg concentration, $\delta^{202}$Hg and $\Delta^{199}$Hg with pelagic penguins feeding in Polar Front circumpolar waters (1.66 ± 0.11 and 1.54 ± 0.06‰) support that different MeHg sources occur at large scales in Southern Ocean deep waters.

As a result of its severe toxicity, mercury (Hg) is considered as a worldwide pollutant of major concern for humans and wildlife1. It is present in all compartments of the Earth, transported over long distances and accumulated in the environment2. Anthropogenic pressure has perturbed the global Hg cycle and tripled Hg concentrations in oceanic surface waters since pre-industrial periods3. The elemental form of Hg (Hg0) from the atmosphere can be oxidized into Hg2+ and deposited in the surface of the ocean, where one part can be rapidly reduced back to Hg0. Mercury is present principally as dissolved Hg0 or inorganic Hg2+ in oceanic waters, but inorganic forms can be methylated by microbiological4,5 or abiotic processes6, leading to the incorporation of methylmercury (MeHg) in organisms and its consequent biomagnification in marine food webs. Vertical profiles of MeHg oceanic distribution7–9, including in the Southern Ocean10, showed that surface waters consistently present lower MeHg concentrations due to more rapid photodegradation, and that increasing MeHg concentrations are found with depth, peaking at low-oxygen and/or microbially active intermediate waters8,11,12. Consequently, an increasing gradient of total Hg (THg) concentrations was found in fish caught from the surface (epipelagic zone) to deeper waters (mesopelagic zone)13–15.

Seabirds are meso- to top predators within marine ecosystems and are therefore exposed to elevated concentrations of MeHg via dietary uptake. They are recognized as effective bioindicators of marine Hg contamination at different spatial scales according to their life cycle16,17. Most seabirds disperse or migrate from the breeding grounds during the inter-nesting period, during which they use different food and feeding ecology strategies. Flying seabirds can cover long distances at that time, thus resulting in the integration of Hg originated from both...
the breeding and inter-breeding foraging zones in their tissues. Therefore, the use of flying seabirds as bioindicators requires a good knowledge of their feeding ecology over the entire annual cycle to better interpret their Hg levels and exposure pathways. Compared to flying birds, the flightless diving penguins exploit relatively spatially restricted foraging zones all year long and are thus representative of Hg contamination in more limited areas, which make them interesting models for biomonitoring studies.

Investigating several penguin species provide access to different marine environmental compartments since they have species-specific foraging ecologies. They feed on a large diversity of prey in different oceanographic ecosystems, both horizontally (from the neritic to the oceanic domains) and vertically, as they forage at different depths of the water column (from the epipelagic to the mesopelagic zones). In this study, we investigated the four sympatric penguin species that breed at the subantarctic Crozet Islands (Southern Indian Ocean): king _Aptenodytes patagonicus_, gentoo _Pygoscelis papua_, macaroni _Eudyptes chrysocome_ and eastern rockhopper _E. chrysoceome filholi_ penguins. The king penguin (KP) is a large oceanic species that feed on mesopelagic fish (mackerels) at deep depths (100–300 m) in distant southern foraging grounds located in the vicinity of the Polar Front. In contrast, the medium-sized gentoo penguin (GP) is a coastal neritic species that dive both pelagically and benthically to feed opportunistically on a large diversity of prey, including swarming crustaceans and benthic fish. The smaller and closely-related macaroni (MP) and rockhopper (RP) penguins forage in offshore waters where they primarily target swarming crustaceans (euphausiids and hyperiids) in the top 70 m of the water column. Here, we measured Hg isotopes in penguin blood samples with the main objective of exploring potentially different MeHg trophic sources due to the bird contrasted foraging ecology. Blood Hg is known to reflect recent Hg exposure (over the last weeks preceding sampling) and penguins are more restricted to areas near their colonies at the time of sample collection (near the end of the breeding period). Hence, Hg isotopic composition of penguin blood samples was considered as indicative of Hg values in waters surrounding the Crozet Islands.

Mercury has seven stable isotopes that undergo mass dependent fractionation (MDF, δ²⁰⁷Hg) as a result of many physical, chemical or biological processes, namely volatilization, reduction, methylation or demethylation, photochemical reactions, and trophic. Because of different combinations of all these processes in the environment, Hg MDF occurs with different degrees of magnitude, thus providing information about Hg processes and specific reservoirs of ecosystems. However, due to the complexity of the Hg biogeochemical cycle, Hg sources cannot be easily differentiated in a given environment, especially when using top predators as bioindicators because they integrate spatio-temporally the trophic web. Moreover, photochemical reactions induce significant Hg mass independent fractionation (MIF, here δ¹⁹⁹Hg), wherein principally odd isotopes are enriched or depleted in reaction products relative to the even isotopes. Recent observations also reported MIF of Hg even-mass isotopes, mainly in samples derived from the atmosphere. Contrary to Hg MDF, no substantial Hg MIF has been observed during trophic processes, meaning that MIF of odd isotopes can be used as a conservative tracer of MeHg sources in predators. This allows investigating photochemical processes before MeHg uptake in the food web. Therefore, the combination of MDF with MIF is used as an effective double tracer of both Hg sources and processes in the environment.

An increasing number of studies have successfully applied Hg isotope analysis for elucidating sources and pathways of MeHg in the environment. In aquatic ecosystems, Hg MIF is highly sensitive to photochemical reactions and varies as a function of the extent of light penetration at different locations or depths. For example, higher photodemethylation rates in surface waters leads to higher Hg MIF in epipelagic than in mesopelagic fish. Fish MIF signature reveals that MeHg without MIF is produced at the pycnocline, thus diluting the MIF of MeHg exported from the surface mixed layer. Higher magnitudes of MIF have also been observed in oceanic versus coastal organisms. This gradient is mainly attributed to enhanced MeHg photodemethylation in oceanic waters due to higher light penetration, while higher water turbidity and benthic MeHg inputs lead to lower extent of photodemethylated Hg in coastal waters. Consequently, we hypothesized that Hg MIF values in penguins should present an increasing gradient from benthic to oceanic, and from mesopelagic to epipelagic foragers. Stable isotopes of carbon (δ¹³C) and nitrogen (δ¹⁵N) are used as relevant proxies of foraging habitat and trophic position of consumers, respectively. The usefulness of the stable isotope method has already been deeply investigated in the Southern Ocean, with seabird δ¹³C values indicating their latitudinal foraging grounds and depicting both offshore versus inshore consumers and benthic versus pelagic ones. Since consumers are enriched in the δ¹⁵N over their food, δ¹⁵N values indicate the trophic position of consumers within a given trophic web, thus allowing making interpretations about trophic relationships in ecological studies. Thus, we used δ¹³C and δ¹⁵N values to help explaining the potential variations in Hg stable isotopes in penguins and to trace the origin, trophic transfer and bioaccumulation processes of Hg in marine food webs, as it was previously demonstrated in fish.

The main objective of this work was to investigate the effectiveness of Hg stable isotope ratios in seabird tissues to discern and quantify MeHg sources and exposure pathways in the different marine compartments in which they forage. We hypothesized that contrasted ecological strategies among penguins would determine the uptake of distinct environmental MeHg sources and would lead to interspecies differences in blood Hg isotopic composition. The exploration of these Hg isotopic ratios in combination with blood δ¹³C and δ¹⁵N values were expected to provide new insights into the complex factors controlling Hg biogeochemical processes and help identifying the sources of MeHg ultimately accumulated in marine top predators.

**Results**

**Blood Hg concentrations and Hg isotopic composition.** Blood THg concentrations differed amongst penguin species (Kruskal Wallis, H = 30.09, p < 0.0001), with king (KP) and gentoo (GP) penguins presenting higher concentrations than macaroni (MP) and rockhopper (RP) penguins (Table 1). Accordingly, blood MeHg was overall different (H = 28.87, p < 0.0001), with KP and GP showing higher concentrations (both 1.9 μg g⁻¹)
Table 1. Food and feeding ecology, including blood δ¹³C (as a proxy of foraging habitat) and δ¹⁵N (as a proxy of trophic position) values, together with blood THg, MeHg and Hg isotopic composition (all isotopes and individual data in Supplementary Table S1) of subantarctic penguins from Possession Island, Crozet Archipelago (n, number of individuals). Values are means ± SD. Values not sharing the same superscript letter are statistically different.

| Species                  | Diet                        | Diving behaviour | n  | δ¹³C (%) | δ¹⁵N (%) | THg (µg g⁻¹) | MeHg (µg g⁻¹) | MeHg (%) | δ¹⁰¹Hg | Δ¹⁹⁹Hg | Δ¹⁹⁹Hg/Δ²⁰¹Hg | ratio |
|--------------------------|-----------------------------|------------------|----|---------|----------|--------------|-------------|----------|--------|--------|----------------|-------|
| King penguin             | fish                        | mesopelagic      | 11 | −21.8 ± 0.4⁴ | 10.1 ± 0.2⁴ | 2.01 ± 0.29⁴ | 1.88 ± 0.28⁴ | 93 ± 1   | 1.49 ± 0.11⁴ | 1.60 ± 0.04⁴ | 1.16 ± 0.04⁴ |
| Gentoo penguin           | crustaceans, fish           | epipelagic, benthic | 11 | −18.6 ± 0.3⁴ | 8.2 ± 0.6⁴ | 2.04 ± 1.00⁴ | 1.89 ± 0.93⁴ | 93 ± 1   | 1.45 ± 0.12⁴ | 1.41 ± 0.06⁴ | 1.18 ± 0.04⁴ |
| Macaroni penguin         | crustaceans (fish)          | epipelagic       | 10 | −20.0 ± 0.7⁴ | 8.6 ± 0.4⁴ | 1.06 ± 0.16⁴ | 1.01 ± 0.15⁴ | 96 ± 2   | 1.66 ± 0.11⁴ | 1.54 ± 0.06⁴ | 1.14 ± 0.02⁴ |
| Eastern rockhopper penguin | crustaceans (fish)          | epipelagic       | 10 | −20.8 ± 0.3⁰ | 8.6 ± 0.4⁰ | 0.97 ± 0.20⁰ | 0.93 ± 0.19⁰ | 95 ± 1   | 1.93 ± 0.18⁰ | 1.77 ± 0.13⁰ | 1.15 ± 0.04⁰ |

Figure 1. Blood δ¹⁵N and δ¹³C values of subantarctic penguins from Possession Island, Crozet Archipelago. Abbreviations: GP, gentoo penguin; KP, king penguin; MP, macaroni penguin; RP, rockhopper penguin.

than MP and RP (1.0 and 0.9 µg g⁻¹, respectively). All individual penguins presented a large predominance of MeHg in their blood (94 ± 2%, range: 91–98%, n = 42).

Blood samples showed large ranges of individual δ²⁰²Hg (MDF) and Δ¹⁹⁹Hg (MIF) values (1.28–2.12‰ and 1.31–1.95‰, respectively, n = 42). Both blood δ²⁰¹Hg and Δ¹⁹⁹Hg values differed among penguins (H = 26.94 and 32.27, respectively, both p < 0.0001), except KP and GP that showed identical δ²⁰¹Hg values. Blood δ²⁰²Hg and Δ¹⁹⁹Hg values increased in the order GP < MP < RP and GP ≈ MP < KP < RP, respectively (Table 1). RP notably showed high inter-individual variability in their blood Δ¹⁹⁹Hg values (from 1.55 to 1.93‰).

Measured blood δ²⁰²Hg values followed the predicted theoretical MDF line (Supplementary Fig. S1A). In contrast, measured blood δ²⁰¹Hg values diverged from the δ¹⁹⁹Hg theoretical MDF line (Supplementary Fig. S1B), indicating that all blood samples showed MIF of the δ¹⁹⁹Hg (and δ²⁰¹Hg) odd isotopes. Overall penguin blood samples displayed a Δ¹⁹⁹Hg/Δ²⁰¹Hg slope of 1.16 ± 0.05 (R² = 0.98, p < 0.0001) (Supplementary Fig. S2). Because of the low inter-species range and high intra-species homogeneity of Hg isotopic values, the measured Δ¹⁹⁹Hg/Δ²⁰¹Hg slopes for each penguin species were not accurate. Hence, we calculated the mean Δ¹⁹⁹Hg/Δ²⁰¹Hg ratios for each penguin species (Table 1). Blood Δ²⁰⁰Hg values were not significantly different from zero and no statistical differences of Δ²⁰⁰Hg values were found between populations (Supplementary Fig. S1A). Therefore, no MIF of even Hg isotopes was detected in penguin blood, as already mentioned in a previous publication on avian blood and feathers⁰⁷.

Blood δ¹³C and δ¹⁵N values. Penguins were segregated by their δ¹³C and δ¹⁵N values (Fig. 1). The four penguin species presented distinct δ¹³C values (Kruskal Wallis, H = 35.30, p < 0.0001), with a progressive δ¹³C enrichment from KP to GP (Table 1). Blood δ¹⁵N values were also different (H = 25.24, p < 0.0001). They allow splitting species into two groups, with MP, RP and GP differing from KP by their 1.5–1.8‰ lower δ¹⁵N values (Fig. 1).

Relationship between blood Hg isotopic composition and δ¹³C and δ¹⁵N values. Excluding KP values, highly negative correlations were found for δ¹³C values with δ²⁰²Hg values (R² = −0.73, p < 0.0001) and with Δ¹⁹⁹Hg values (R² = −0.71, p < 0.0001) (Fig. 2). In contrast, no significant correlation was observed between blood δ²⁰²Hg and δ¹⁵N values (R² = −0.10, p = 0.52) and between blood Δ¹⁹⁹Hg and δ¹⁵N values (R² = 0.03, p = 0.30) (Supplementary Fig. S2).
**Discussion**

MeHg is the predominant Hg species in blood of penguins, irrespective of THg concentrations or species. Consequently, measured Hg isotopic composition of this tissue corresponds almost exclusively to MeHg. The predominance of MeHg in blood allows (i) the direct comparison of blood THg as a proxy of MeHg among the four penguin species, and (ii) the exploration of MDF and MIF values measured on THg to trace MeHg pathways in each marine compartment used by penguins. In the following sections, variations of Hg isotopic composition among penguins are discussed and interpreted as a function of their species-specific ecological characteristics in order to estimate sources and processes involving MeHg in the Crozet marine environment. We first interpreted blood $\delta^{13}$C values to define the foraging habitats of penguins and then combined $\delta^{13}$C with $\delta^{202}$Hg and $\Delta^{199}$Hg values to depict potentially different Hg sources and/or processes between the inshore and offshore environments. The relationship between blood $\delta^{13}$N values and Hg isotopes was also explored to test the potential influence of trophic processes on penguin $\delta^{202}$Hg and $\Delta^{199}$Hg values.

**Variations in foraging habitats ($\delta^{13}$C) between penguin populations.** In waters surrounding subantarctic islands, $\delta^{13}$C values decrease from neritic to oceanic waters (inshore-offshore $\delta^{13}$C gradient), and from warm to cold waters (latitudinal $\delta^{13}$C gradient), thus allowing using $\delta^{13}$C values to assess the main foraging habitats of seabirds. A large range of $\delta^{13}$C values was observed across penguin species with decreasing values from gentoo (GP) to king (KP) penguin. KP exhibited much lower $\delta^{13}$C values than the other two oceanic penguins (macaroni MP and rockhopper RP), which is in agreement with their well-known southern foraging grounds from Crozet Islands down to the Polar Front. RP showed significantly lower $\delta^{13}$C values than MP, indicating that they forage at more southern latitudes than MP during the breeding period. Finally, the more positive $\delta^{13}$C values of GP compared to the other penguins are in agreement with the inshore feeding habits of the species. Species-specific $\delta^{13}$C values clearly demonstrate that each of the four penguins breeding at the Crozet Islands document a different compartment of the marine environment, thus allowing studying the processes affecting Hg behavior and its fate in various ecosystems from the Southern Ocean.

**Relation between Hg isotopes and penguin trophic ecology ($\delta^{15}$N).** Differences in dietary composition amongst pelagic penguins are well illustrated by $\delta^{15}$N values being higher in the fish-consumer species.
∆advection and iron fertilization. The lack of correlation between blood δ13C values (Fig. 1) indicate that KP foraged in an oceanic ecosystem that is not closely connected to the Crozet archipelago, where the carbon pump is driven by the local advection and iron fertilization. The lack of correlation between blood ∆199Hg and δ15N values is consistent with the absence of MIF during trophic transfer35,48–50. While MDF could be produced during trophic transfer35,37,48, no correlation was observed between ∆202Hg and δ15N values. Based on the existing knowledge of Hg isotopic fractionation dynamics, we concluded that the observed MIF (∆199Hg) variations between penguin species are linked to distinct MeHg sources having undergone different degrees of photochemical reactions. Since no effect of trophic level (δ15N) was found for ∆202Hg and ∆199Hg values, the most plausible explanation of Hg isotopic variations is the consequence of species-specific foraging habitats within the marine ecosystems.

Hg isotopic values and penguin species-specific foraging habitats. MIF characteristics and penguin foraging depths. Photochemical reactions of MeHg and inorganic Hg in the water column are the main drivers of Hg odd-MIF values and each process is characterized by a different ∆199Hg/∆201Hg ratio. Since this ∆199Hg/∆201Hg ratio is assumed to be preserved in the food chain after MeHg assimilation by primary producers and during biomagnification up the food web, it is used to identify mechanisms involving MIF variations41. Theoretical slopes have been experimentally designed in aquatic systems34, corresponding to 1.36 ± 0.02 for MeHg photodemethylation and 1.00 ± 0.02 for inorganic Hg photoreduction in freshwater with natural DOC. Although studies in freshwater fish also reported ∆199Hg/∆201Hg slopes close to 1.348,49,51, slightly lower ∆199Hg/∆201Hg slopes have been observed in marine organisms41,43,52–55. This effect may be the result of different ligands associated with Hg in aqueous solutions, such as dissolved organic matter34,56,57, or dissolved cations or halogens in seawater40. Indeed, variable ∆199Hg/∆201Hg slopes have been recently documented for MeHg photodemethylation under different types and concentrations of DOC37, indicating that low concentrations of DOC relative to MeHg could lead to lower ∆199Hg/∆201Hg slopes. The overall ∆199Hg/∆201Hg slope for penguin blood samples (1.16 ± 0.05) (Supplementary Fig. S2) is consistent with those previously reported in marine fish muscle41,42,55 and seabird eggs41. Due to the predominance of MeHg in penguin blood, and assuming a potential diminution in MIF slope due to low DOC concentrations in subtropical waters, the obtained ratio indicates an accumulation of residual MeHg that has principally undergone photochemical demethylation.

Hg MIF in marine fish has been used to estimate the relative proportion of MeHg formed in the open ocean that is photochemically degraded prior its entry into the food web41. Thus, it has been shown to be an effective proxy of fish foraging depths41. Assuming MeHg photodemethylation as the major photochemical process, we estimated the percentage of presumed photodemethylated MeHg before entering the food web based on experimental studies34,57 (detailed in SI). The estimated extent of MeHg photodemethylation varied slightly between penguin species (~13 to ~16%), a difference that is surprisingly low when taking into account the range of habitats used by the penguins (from coastal benthic/pelagic feeders to mesopelagic feeders). Considering the existence of higher amounts of DOC in benthic waters relative to the pelagic domains, more substantial differences were expected between compartments, even for similar MeHg concentrations. The photodemethylation extent estimated in an Antarctic coastal ecosystem (~13–18%)34 based on the same experimental models57 was similar to our findings. Slightly higher range of MeHg photodemethylation was observed in the Arctic Ocean from ice-covered to non-ice-covered marine areas (~8–16%)52. Another study in fish from the Gulf of Mexico estimated that MeHg in coastal fish was ~10–20% degraded in contrast to oceanic fish whose percentage of photodemethylated MeHg was ~40–65%42. Even higher in surface waters, photodegradation appears overall limited in the Southern Indian Ocean compared to subtropical waters (e.g. Gulf of Mexico), as a consequence of lower sunlight extent and slighter angle of incidence.

Rapid light attenuation with depth leads to the inhibition of MeHg photodemethylation and thus to lower Hg odd isotope MIF values41. Although such Hg MIF is much more sensitive to photochemical reactions, photodemethylation also induces MDF with the remaining MeHg enriched in heavier isotopes41. In the North Pacific Ocean, Blum et al.41 documented a Δ199Hg offset of ~5% between fish feeding at the surface mixed layer and at 600 m depth. Here, we aimed to assess the correlation between Hg MIF and penguin foraging depths in the Southern Indian Ocean. A gradual increase of Δ199Hg values was observed from the coastal and mixed benthic/pelagic forager (GP) to the two pelagic penguins that feed in Crozet waters, with RP having higher values than MP. Although both MP and RP forage in near surface waters, the significantly different blood Δ199Hg values between species (~0.20‰) suggest differences in their foraging depth intervals during the breeding period. On the other hand, KP are representative of Polar Front waters during the sampling period and they were out of this Hg MIF-foraging depth trend. Despite their mesopelagic behaviour, KP presented higher Δ199Hg values than epipelagic MP (mean difference 0.06‰), which is likely associated to a specific relation between Hg MIF and the characteristics of ocean waters either close to Crozet (MP) or to the Polar Front (KP).

Tracing distinct MeHg sources over inshore-offshore and benthic-pelagic gradients. For a complete understanding of the exposure pathways to MeHg relative to species-specific foraging habitats in penguins, both in horizontal (inshore-offshore) and in vertical (benthic-pelagic) dimensions, we combined Hg isotopic discrimination using both MDF (∆202Hg) and MIF (∆199Hg) values. A gradual increase of both ∆202Hg and ∆199Hg values was observed in the three penguin species foraging in Crozet waters, in the order GP < MP < RP (Fig. 3). The overall range of ∆202Hg values (1.28 to 2.12‰) of penguins is slightly higher than those observed in pelagic and benthic fish from the Gulf of Mexico42, estuarine waters of the Labrador Sea46 and Hawaii coastal and marine areas55. However,
penguin $\Delta^{199}\text{Hg}$ values (1.31 to 1.95‰) fall within the range of the values observed in fish of these three mentioned regions.

Experimentally determined $\Delta^{199}\text{Hg}/\delta^{202}\text{Hg}$ ratios during aquatic MeHg photodemethylation exhibited a slope of $2.43 \pm 0.10$‰ whereas microbial demethylation and reduction only affect $\delta^{202}\text{Hg}$ values ($\Delta^{199}\text{Hg}/\delta^{202}\text{Hg}$ slope $\sim 0.49$‰) in subantarctic and Polar Front waters when compared to other marine ecosystems.

Benthic-pelagic gradient of MeHg sources in Crozet Islands: Previous studies have already documented Hg isotopic differences between coastal and oceanic marine organisms$^{35,42,43}$, indicating the existence of contrasted environmental MeHg sources with distinct Hg isotopic baselines and different extent of aquatic photochemistry. Significantly lower $\delta^{202}\text{Hg}$ and $\Delta^{199}\text{Hg}$ values exhibited by benthic/pelagic GP relative to pelagic foragers is indicative of their higher accumulation of Hg with a sediment origin. Sediment Hg isotopic composition is characterized by (i) a different Hg isotopic baseline if compared to oceanic waters (ii) close to zero MIF and negative MDF extent and (iii) further no or low photochemical reactions of benthic MeHg. Therefore, lighter Hg isotopic values are typically found in benthic coastal biota relative to oceanic organisms as a result of a higher continental influence$^{42,43}$. No sediment Hg isotopic data are available from the Crozet Islands, but it is likely that $\delta^{202}\text{Hg}$ values are negative and $\Delta^{199}\text{Hg}$ values are close to zero, as commonly observed in sediments from other sites such as in the Arctic Ocean ($\delta^{202}\text{Hg}: -1.37 \pm 0.38$‰; $\Delta^{199}\text{Hg}: -0.02 \pm 0.07$‰$^{39}$) and the Antarctic coasts ($\delta^{202}\text{Hg}: -0.39 \pm 0.49$‰; $\Delta^{199}\text{Hg}: 0.71 \pm 0.43$‰$^{39}$). Indeed, Zheng et al. observed similar MIF values between historical sediment profiles and penguin and seal fresh faeces, suggesting that faeces were the dominant sources of Hg to the sediments at different time periods. Due to the huge penguin (and other seabirds) populations, a significant fraction of Hg accumulated in coastal sediments from the Crozet Islands could be of ornithogenic origin, thus showing similar isotopic values as other Antarctic sediments$^{39}$. In coastal ecosystems, a higher turbidity reduces light penetration, thus limiting Hg photochemistry. This phenomenon, together with the influence of benthic Hg inputs, may also contribute to the lower $\Delta^{199}\text{Hg}/\delta^{202}\text{Hg}$ ratio in subantarctic and Polar Front waters when compared to other marine ecosystems.

The significant correlations between blood $\delta^{13}\text{C}$ and Hg isotopic values (Fig. 2) suggest a progressive transition from terrestrial to marine values along both a horizontal (inshore-offshore) and a vertical (benthic-pelagic) gradient. This is in agreement with in situ Hg methylation in sediment and reduced exposure to sunlight (due to turbidity and/or depth) as the main mechanisms lowering Hg isotopic values in benthic ecosystems. Meanwhile, a gradual increase in $\Delta^{199}\text{Hg}$ (and $\delta^{202}\text{Hg}$) was observed from inshore to offshore waters as a consequence of higher magnitude of photochemical processes in more opened areas. Nevertheless, the low variations in Hg isotopes values between benthic and pelagic penguins compared to previous inshore-offshore values measured in other marine ecosystems$^{42,43}$ seems to indicate a higher degree of mixing between benthic and pelagic MeHg sources. The remote location of the Crozet Islands, which are surrounded by deep oceanic waters (4000–5000 m), could
explain a lower impact of the sediment-derived MeHg inputs compared to continental coastal zones. Moreover, these islands have a plateau of around 150 km wide that interacts with different water masses derived from the Antarctic Circumpolar Current, thus potentially favouring the recirculation and mixing of MeHg from different sources. Indeed, the relatively low range of $\delta^{13}$C values and the similar $\delta^{202}\text{Hg}/\Delta^{199}\text{Hg}$ ratio of the three penguins seem to be coherent with common environmental MeHg production sources, with the resulting MeHg accumulating either in the benthic or pelagic food webs in Crozet waters. MeHg in offshore marine ecosystems likely derives primarily from inorganic Hg deposited from the atmosphere. However, the high degree of oceanic recirculation within the water column in the vicinity of Crozet Islands could favour the redistribution of MeHg between the surface to deeper zones of the water column and its combination with the MeHg originating from the benthic zones.

Different pelagic MeHg sources between Crozet and distant (Polar Front) waters: The significant trend between MeHg concentrations and both $\delta^{202}\text{Hg}$ and $\Delta^{199}\text{Hg}$ values (Fig. 4) clearly illustrates a common mixing source for the four subantarctic penguins with a dominant sediment MeHg source in benthic/pelagic penguins with higher Hg levels (GP) and a more diluted pelagic MeHg source with lower concentrations at the sea surface (RP). Significantly higher MeHg concentrations in KP, as discussed above, could also be associated to their higher trophic level compared to MP and RP. However, the lower Hg isotopic values of KP suggest that they feed on a distinct food web that is MeHg-enriched compared to the pelagic ecosystem close by the Crozet Islands. Both seasonal release of nutrients and summer algal bloom lead to higher primary productivity in Polar Front waters, which favours bacterial activity compared to northern waters. Therefore, greater methylation yields at the Polar Front can also contribute to the higher MeHg concentrations accumulated in KP compared to MP and RP. Moreover, algal blooms may lead to higher accumulation and availability of organic matter close to the photic zone of the water column, and its higher photodemethylation under exposure to sunlight. This hypothetical effect should provide higher MIF signatures in MeHg accumulated in mesopelagic food webs of the Polar Front.

In conclusion, RP, MP and GP seem to be connected to the same dominant benthic Hg source, which appears to be associated to the Crozet shelf sediments diluted and mixed in nearby waters with a pelagic source. On the other hand, Hg stable isotope values indicate that KP is relying on a different and unique Hg isotopic baseline due to specific inorganic Hg source and methylation/demethylation pathways in such offshore open waters.

Measuring Hg isotopic composition in sympatric penguins of the Crozet Islands allows demonstrating that penguins are exposed to different levels and sources of MeHg when they forage in three distinct food webs (i.e.

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**Figure 4.** Relationships between Hg isotopes ($\delta^{202}\text{Hg}$ and $\Delta^{199}\text{Hg}$) and MeHg concentration values. (A) Blood $\delta^{202}\text{Hg}$ and inverse of MeHg concentration, and (B) blood $\Delta^{199}\text{Hg}$ and inverse of MeHg concentration of subantarctic penguins from Possession Island, Crozet Archipelago. Abbreviations: GP, gentoo penguin; KP, king penguin; MP, macaroni penguin; RP, rockhopper penguin.
of tetramethylammonium hydroxide (25% TMAH in H₂O, Sigma Aldrich) 65. Hg species analyses were carried out by using an advanced Hg analyzer (AMA-254, Altec) for the intercomparison with total Hg concentrations obtained by Hg speciation analyses, i.e. the sum of inorganic and organic Hg. Hg isotopic composition was determined using cold-vapor generator (CVG)-MC-ICPMS (Nu Instruments), as detailed previously35. Hg isotopic values were reported as delta notation, calculated relative to the bracketing standard SRM-997 thallium standard solution was used for the instrumental mass-bias correction using the exponential law (details of calculation in SI). Secondary standard NIST RM-8160 (previously UM-Almadén standard) was used for validation of the analytical session (Supplementary Table S2). Details of Hg isotopic composition analyses are included in the SI and are further detailed elsewhere27.

For the validation of the analytical results, four certified reference material were analysed: human hair IAEA-086 and NIES-13, tuna fish ERM-CE-464 and dogfish liver DOLT-4. An internal reference sample was prepared with pooled samples collected from different individuals of king penguins from the Crozet Islands (RBC-KP, red blood cells). It was analysed at each analytical session. Analytical uncertainty for delta values was calculated using SD typical errors for reference materials (Supplementary Table S2), as recommended by reference publications for standard reporting of Hg isotopic ratio uncertainties56,67.

Carbon and nitrogen stable isotopes analyses. Blood samples were freeze-dried and powdered, and subsamples were weighed with a microbalance and packed in tin containers. Carbon (δ¹³C) and nitrogen (δ¹⁵N) stable isotope ratios were determined in red blood cells with a continuous flow mass spectrometer (Thermo Scientific Delta V Advantage) coupled to an elemental analyser (Thermo Scientific Flash EA 1112) in the laboratory LIENSs (La Rochelle, France) (aliquots mass: ~0.3 mg). Results are in delta notation relative to Vienna PeeDee Belemnite and atmospheric N₂ for δ¹³C and δ¹⁵N, respectively. Replicate measurements of internal laboratory standards (acetanilide) indicated measurement errors <0.15‰ for both δ¹³C and δ¹⁵N values.

Statistical analyses. Statistical tests were performed using R 3.3.2 (RStudio)98. Before analyses, data were checked for normality of distribution and homogeneity of variances using Shapiro–Wilk and Breusch-Pagan tests, respectively. Since data groups did not meet specificities of normality and homoscedasticity, non-parametrical tests (Kruskal–Wallis with Conover-Iman test) were performed. Statistically significant results were set at α = 0.05. Values are means ± SD. We examined the correlations between MeHg concentrations, δ¹³C, δ¹⁵N and both Hg MDF (δ²⁰²Hg) and MIF (δ¹⁹⁹Hg) using linear regressions and Pearson correlation rank tests. Hg MIF δ¹⁹⁹Hg and Δ²⁰⁵Hg values were regressed to determine if MIF ratio was consistent with photochemical degradation of MeHg and in good agreement with previous studies on marine organisms (e.g.42,43,59).

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**Author Contributions**

D.A., P.B. and Y.C. participated in the design of the study. M.R. performed mercury speciation and isotopic analyses and drafted the manuscript. D.A., Z.P. and M.R. carried out the interpretation of mercury isotopic results. P.B., Y.C. and M.R. investigated ecological and trophic aspects. All authors contributed to manuscript revision.

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