Type 2 Diabetes Mellitus (T2DM): Biological Overview from Pathways to Organelles and its Translation toward a Torpid Wound Healing Process

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Abstract

T2DM is a heterogeneous group of metabolic diseases characterized by hyperglycemia resulting from defects in insulin secretion, insulin action, or both. Hyperglycemia may simply represent the tip of a broad series of molecular events from mitochondrial damages, to epigenetic and metabolic pathways deregulations. At the same time, hyperglycemia appears as the most proximal trigger for the onset and perpetual progression of multi-organ complications even under normoglycemic conditions. Thus, the initial hyperglycemic hit translates into a permanently harmful cellular imprinting as has been demonstrated in diabetic donors’ cells after several passages and cultured in ideal conditions. The wound healing failure along with the inability of the innate immunity to control peripheral infections is the hybrid that determines that 85% of all non-traumatic lower extremity amputations are practiced in diabetic subjects. Diabetic wounds exhibit a complex networking of inflammatory cytokines, local proteases, cytotoxic reactive oxygen and nitrogen species and a polymicrobial biofilm that impose a stagnant phenotype. All these ingredients negatively impact on fibroblasts, endothelial cells and keratinocytes while paradoxically perpetuate the immuno-inflammatory infiltrate. Although the molecular fundamentals toward chronification have not been elucidated, it seems that different gene simultaneously converge to impose the wound cells a pro-senescent, pro-catabolic and pro-apoptotic phenotype given the lack of a “physiological tuning” of tyrosine kinase-dependent receptors due to their limited activation by insulin and local growth factors. Although recombinant growth factors and smart devices have been introduced during the last years the figures of amputations are still discouraging. Faults have been committed while selecting the appropriate growth factor and because of the “chronic” instinct to treat the chronic wounds topically, where bioavailability of the active principle is compromised by wound and bacterial biofilm proteases. The periodic intralesional infiltration of epidermal growth factor has proved to overcome this hurdle. Granulation tissue growth stimulation and wound healing capacity has been restored in diabetic patients by this procedure in several clinical trials and common clinical practice studies.

Keywords: Diabetes; Ulcer; Amputation; Granulation tissue; Re-epithelialization

Introduction

The current understanding on the molecular mechanisms impairing wound healing in diabetic subjects has progressively expanded over the last 20 years. Although we are still far from detailing the pathways toward wound chronification, diabetes- molecular factors that enforce fibroblasts, pericytes, keratinocytes and endothelial cells to precocious senescence, arrest and apoptosis have been identified to some extent [1-3].

Diabetes has rapidly increased in global prevalence, morbidity and mortality [4]. Diabetic foot ulcers (DFUs) are estimated to occur in 15 % of all patients with diabetes [5] and precede 84 % of all diabetes-related lower-leg amputations [6]. These figures suggest the existence of a gap between the bench-derived molecular progresses and wound management in daily practice. Diabetic foot ulcer healing represents a considerable investigational and clinical challenge and the treatment requires in addition to education and resources, an integral multidisciplinary approach to succeed in reducing amputation rates [7].

The general outcome of diabetic foot ulcers management is poor and there is continuing uncertainty concerning optimal approaches to confront it [8]. As a response, judicious strategies as the molecular-guided approaches in a style of “personalized wound management” have started to hatch [9].

This review would hopefully offer a comprehensive notion about the entangled biochemical pathways of T2DM and its relationship with torpid healing as one of its complications. Special focus is addressed on the current lines of evidences about the toxic resonance of acute and long term exposure to high glucose on granulation tissue building blocks: fibroblasts and endothelial cells. We have included a characterization of the diabetic granulation tissue organizational disorders. The rationale and fundamentals for a unique pharmacological intervention to deal with high-grade ulcers are also presented. This EGF-based intervention is distinguished by the fact that its delivery procedure is based on the ulcer bed infiltration thus ensuring bioavailability to wound-responsive cells. The literature search was based on English language

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**General Overview of Type 2-Diabetes Mellitus (T2DM)**

T2DM may be conceptually defined as a heterogeneous group of metabolic diseases characterized by hyperglycemia resulting from defects in insulin secretion, insulin action, or both. It is important to note however that hyperglycemia may simply represent a distal and terminal aspect of a broad series of events, involving peripheral insulin resistance, rupture of glycogen homeostasis, and relative insulin secretion response failure. It is generally accepted that T2DM is a polygenic disorder composed of subtypes whereby genetic susceptibility is strongly associated with environmental factors. Thus, this metabolic epidemic explosion represents the phenotypic expression to a collision between genes and environment[10].

Evolutionarily glucose is a major energy source and is an important substrate for complex carbohydrates, protein and lipid synthesis in mammalian cells. It provides energy in the form of ATP through glycolysis and the citric acid cycle. It is also used in the formation of glyceraldehyde 3-phosphate under hypoxia and anaerobic pathways, thus providing the principal source of cellular energy and substrate storage. Glucose hydrophilic nature limits its trespassing through the cellular lipid bilayer, thus specific transporters are required for cells to incorporate it. Glucose carrier proteins (GLUTs) mediate insulin-stimulated glucose uptake. Nevertheless, glucose clearance is also mediated via insulin-independent mechanisms derived from the ability of plasma glucose to influence its own clearance by a mass action effect[11].

The regulation of glucose storage and endogenous production is central to blood glucose control. Gluconeogenesis and glucogenolysis are tightly regulated by hormones, enzymes, signaling pathways and cellular organelles. At the system level, organs like the liver, skeletal muscle, kidneys and adipose tissue play essential roles in glucose homeostasis. GLUTS mediate insulin-stimulated glucose uptake in these organs and tissues by a mechanism involving translocation between cellular compartments. GLUT isozyme expression is regulated by insulin and other factors such as hypoxia, oxidative phosphorylation and osmotic stresses[12].

T2DM is characterized by a reduction of total body glucose clearance as compared with age and weight-matched controls[13]. This phenomenon includes insulin-independent[12] and insulin-dependent glucose clearance[14] as has been demonstrated in humans at risk of diabetes. Insulin action is not solely restricted to GLUTS regulation, but also to glucose homeostasis through hepatic glucose production suppression and stimulation of peripheral combustion[15]. At the end, the decline in peripheral glucose utilization and/or excessive hepatic production constitutes a major ingredient in the so-called insulin resistance whose major consequence is chronic hyperglycemia and/or hyperinsulinemia[16].

Different studies have shown that insulin resistance occurs at the hepatic and skeletal muscle levels. Insulin action signal transduction is impaired in skeletal muscle from T2DM subjects underscoring the contribution of molecular defects underlying the insulin resistant phenotype[12]. The fact that this process has been observed in cultured myoblasts after several passages suggests a genetic origin in connection with a heightened state of oxidative stress[17]. Insulin resistance is also a determinant factor in pancreatic β-cells failure. Factors limiting β-cell ability to respond to an increasing metabolic demand remain unknown, but likely involve genetic factors as well as glucotoxicity itself[18].

Hyperglycemia, along with insulin resistance and abnormal insulin and glucagon secretion, may alter glucose fluxes through specific pathways and cause glucose carbon to be shunted away from normal sites of storage. Furthermore, routes of glucose disposal are abnormal in diabetic subjects since glucose oxidation appears significantly reduced. Among other factors, an impaired mitochondrial activity might represent a basic defect in T2DM[19]. Thus, abnormal glucose storage and utilization in skeletal muscle and other tissues characterize T2DM, rendering an aberrant and uncontrolled pattern of proteins glycosylation[20]; which is a major contributing factor for the multi-system complications.

**Enzymes and pathways underlying T2DM**

As stated in the brilliant review by Bouche and co-workers (20) T2DM is a heterogeneous process so that multiple defects are necessary and permissive to the development of hyperglycemia[20]. Molecular approaches using gene expression profiling have demonstrated modifications in those encoding key enzymes involved in glycolysis, oxidative metabolism, and mitochondrial function associated with progressive insulin resistance and diabetes[20]. Some relevant examples of disease-associated gene expression modifications are shown in table 1.

The first step in glucose metabolism is its phosphorylation, rendering glucose 6-phosphate (G-6-P) by hexokinases. Hexokinase II levels have been found to be low in skeletal muscle of diabetic subjects as compared with matched controls. A loss-of-function mutation of glucokinase has also been identified in some families with maturity onset diabetes of the young (MODY)[45]. These patients demonstrate decreased glucose phosphorylation and decreased insulin secretion[46]. On the other hand, increased endogenous glucose production is a consistent feature of T2DM which may be associated to an increased glucose-6-phosphatase activity[47].

The pyruvate dehydrogenase complex determines the transformation of pyruvate to acetyl-CoA (coenzyme A). This enzyme complex is inactivated by ATP when cellular energy stores are high and by pyruvate dehydrogenase kinase (PDK). Pyruvate dehydrogenase activity is less responsive to insulin stimulation both in patients with diabetes and in their offspring[48]. Furthermore, increased PDK4 activity has been observed in insulin resistance and T2DM subjects[27]. Other studies have suggested that inhibition of the glycolytic enzyme GAPDH (glyceraldehyde-3-phosphate dehydrogenase) could functionally divert upstream glycolytic metabolites into three major pathways mediating hyperglycemic damage: activation of protein kinase C (PKC) isoforms, hexosamine pathway flux, and advanced glycation end-product formation[49]. This concomitant impairment in the glucose oxidative pathway could render explanation as to why pyruvate-lactate interconversion rate is so greatly enhanced in T2DM patients. The hyperlactinemia may amplify insulin resistance by increased liver gluconeogenesis and decreased muscle glucose uptake[50].

Abnormal activation of glycogen synthase (GS) by insulin action has also been ascribed to T2DM individuals. Whether this is due to hyperglycemia or to a genetic defect has not been elucidated. In support to the first notion is the fact that GS activation was not has also been ascribed to T2DM individuals. Whether this is due to hyperglycemia or to a genetic defect has not been elucidated. In support to the first notion is the fact that GS activation was not
| Gene name and REFSEQ mRNAs | Protein localization | Biological function | Relevance in the pathway of insulin resistance (IR) | Reference |
|----------------------------|----------------------|---------------------|---------------------------------|----------|
| Hexokinase 2 (HK2 or HKII) NM_000189.4 | Enzyme localized in various compartments (cytoplasm, membrane, mitochondrial outer membrane) | Hexokinases phosphorylate glucose to produce glucose 6-phosphate, thus committing glucose to the glycolytic pathway. The expression of this gene is insulin responsive. | Non-insulin dependent diabetes mellitus (NIDDM) patients are characterized by a reduced activity and a reduced gene expression of HK2 in muscle, which may be secondary to the metabolic perturbations. | [21,22] |
| Glucokinase (hexokinase 4) (GCK) NM_001161587.1 | Enzyme localized in various compartments (cytosol, nucleoplasm, cytoplasm, nucleus). | Hexokinases phosphorylate glucose to produce glucose 6-phosphate, the first step in most glucose metabolism pathways. This enzyme regulates carbohydrate metabolism by acting as a glucose sensor. It is not inhibited by its product glucose-6-phosphate but remains active while glucose is abundant. | Mutations in this gene have been associated with: NIDDM, Maturity onset diabetes of the young, type 2 (MODY2), persistent hyperinsulinemic hypoglycemia of infancy (PHHI). | [23,24] |
| Pyruvate dehydrogenase lipoylamine kinase isozyme 4 (PDK4) NM_002612.3 | Enzyme localized in various compartments (cytoplasm, mitochondrial inner membrane, mitochondrial matrix). | It plays an important role in maintaining normal blood glucose levels under starvation, and is involved in the insulin signaling cascade. | PDK4 plays a crucial role in glucose utilization and lipid metabolism by regulating the pyruvate dehydrogenase complex (PDC) and is an emerging therapeutic target for type 2 diabetes. | [25,26] |
| Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) | Enzyme localized in various compartments (cytoplasm, mitochondrial matrix). | This gene encodes a member of the pyruvate dehydrogenase isozyme family. The encoded protein phosphorylates pyruvate dehydrogenase, down-regulating the activity of the mitochondrial pyruvate dehydrogenase complex. | Plays an important role in maintaining normal blood glucose levels under starvation, and is involved in the insulin signaling cascade. Overexpression of this gene may play a role in cell proliferation by regulating carbohydrate and fatty acid metabolism. | [27] |
| Protein kinase C (PKC) isoforms NM_002737.2 | Localized in various compartments (condensed chromosome, condensin complex, condensin core heterodimer, cytoplasm, etc). | GAPDH is a key enzyme in glycolysis that catalyzes the first step of the pathway by converting the D-glyceraldehyde 3-phosphate (G3P) into 3-phospho-D-glyceroyl phosphate. | GAPDH activates the major pathways implicated in diabetic complications, including advanced glycation end products (AGEs), protein kinase C, and hexosamine pathway. | [28] |
| Glycogen synthase (UDP-glucose:glycogen glucosyltransferase) (GYS1) | Localized in key tissues as skeletal muscle and adipose tissue; as in various compartments as mitochondrial and cytoplasm. | This enzyme converts excess glucose residues one by one into a polymeric chain for storage as glycogen. It is a key enzyme in glycogenesis. | Activation of diacylglycerol (DAG)-protein kinase C (PKC) pathway enhances polyol pathway, increases oxidative stress, and the accumulation of advanced glycation end products; all proposed as potential cellular mechanisms by which hyperglycemia induces many vascular abnormalities in renal, retinal, and cardiovascular tissues in diabetic animals and patients. | [29,30] |
| Glycogen synthase kinase (GSK-3) | Protein is localized predominantly in the cytoplasm but is also found in the nucleus. Its subcellular localization is changed in response to stimuli. | Serine/threonine kinase with important roles in the regulation of glycogen synthesis, protein synthesis, gene transcription, and cell differentiation in various cell types. Constitutively active protein kinase that acts as a negative regulator in the hormonal control of glucose homeostasis. | Patients with type 2 diabetes normally exhibit low glycogen storage levels because of impairments in insulin-stimulated glycogen synthesis and suppression of glycolysis. Insulin stimulates glycogen synthase by inhibiting glycogen synthase kinases or/and activating protein phosphatase 1 (PP1) among other mechanisms. | [31] |
| Glycogen phosphorylase (PYGL) | Protein is localized in various compartments (cytoplasm, soluble fraction). | Phosphorylation is an important allosteric enzyme in carbohydrate metabolism. PYGL breaks up glycogen into glucose subunits. This enzyme participates in the glycolysis pathways, Insulin signaling pathway, and sucrose metabolism. | Glycogen phosphorylase activity is critical for normal skeletal muscle function. Mutations in liver PYGL inhibit the conversion of glycogen to glucose and results in moderate hypoglycemia. The inhibition of PYGL has been proposed as one method for treating type 2 diabetes. Inhibiting the release of glucose from the liver glycogen supply appears to be a valid approach. | [32-34] |
T2DM patients [51]. Furthermore, studies of muscle biopsy samples after in vivo insulin stimulation showed decreased activation of the insulin receptor along with the PI 3-kinase activity in skeletal muscle of the patients as compared with control non-diabetic subjects [52]. Concomitantly GSK-3 (glycogen synthase kinase) protein levels and activity are elevated in skeletal muscle [53].

Of relevance for T2DM pathophysiology is the hexosamine pathway since the final step in hexosamine biosynthesis is the formation of UDP-N-acetylglucosamine and other nucleotide hexoamines. These are major substrates for protein glycosylation participating in glucotoxicity and glucose-induced insulin resistance. Glycosylation is a major player in diabetes systemic complications and biochemical disturbances, including gene transcription, insulin signaling, signaling pathways, and systemic endothelial dysfunction. In vivo experiments have shown that increasing the flux into the hexosamine pathway by various means induces defects in insulin secretion and action, including diminished insulin-stimulated glucose uptake [20]. Glutamine - fructose-6-P aminotransferase (GFAT) levels appear elevated in skeletal muscle of T2DM patients and chronic hyperglycemia is also associated with increased enzyme activity [54].

Finally, transcription factors that coordinately regulate genes encoding mitochondrial function alter the expression of those regulating oxidative phosphorylation. PGC-1a is a PPAR (peroxisome proliferator-activated receptor) coactivator that promotes mitochondrial biogenesis and is a powerful regulator of oxidative energy metabolism under conditions of both health and disease [55]. PGC-1a levels are altered both in diabetes patients and in their offspring, who are at high risk for developing the disease [56]. Figure 1 summarizes current concepts on T2DM pathophysiological cascade.

**Cellular organelles involved in T2DM pathogenesis**

The connection between hyperglycemia/insulin resistance and oxidative stress has been acknowledged for years. Moreover, an intrinsic impairment in oxidative capacity, increased oxidative stress and reactive oxygen species (ROS) levels are an important trigger and perpetuating factors for insulin resistance [57].

Insulin resistance is associated with reduced insulin-stimulated mitochondrial activity as the result of blunted mitochondrial plasticity. The seminal observation that insulin becomes intimately bound to mitochondria and microsomes, incited to postulate that the hormone affects oxidative phosphorylation. Ulterior studies showed a clear defect in oxidative phosphorylation in mitochondria from diabetic individuals, which appeared ameliorated following insulin intervention in both in vivo and in vitro systems [58]. Further investigations demonstrated that mitochondrial DNA depleted cells are not insulin responsive [59], suggesting that the insulin signal must interact with this organelle as a response checkpoint. In general terms T2DM individuals present acquired or inherited reductions of mitochondrial activity as the result of blunted mitochondrial plasticity and glycogen metabolism [56]. Further investigations demonstrated that mitochondrial DNA depleted cells are not insulin responsive [59], suggesting that the insulin signal must interact with this organelle as a response checkpoint. In general terms T2DM individuals present acquired or inherited reductions of mitochondrial activity as the result of blunted mitochondrial plasticity and glycogen metabolism [56].

Hyperglycemia increases oxidative stress through several pathways. A major mechanism appears to be the overproduction of the superoxide anion by the mitochondrial electron transport chain. Mitochondria per se are a rich source of oxidative end products. In parallel, mitochondrial superoxide anion is severely implicated in insulin resistance. Reversion of O2− production has proved to restore insulin sensitivity [62]. Insulin-resistant animal models have higher levels of superoxide production [63].

Diminished mitochondrial content in circulating cells is associated with decreased insulin sensitivity and could be a marker of altered content in insulin-responsive tissues. More specifically, mitochondrial area and function are decreased in muscle of obese and diabetic persons [64], and mitochondrial activity correlates closely to measures of insulin sensitivity. Likewise, mitochondrial dysfunction and decreased

### Table 1: List of genes involved in glycolysis, oxidative metabolism, and mitochondrial function associated with progressive insulin resistance and diabetes.

| Gene Symbol | Description | Function |
|-------------|-------------|----------|
| Glucagon-like peptide 1 receptor (GLP1R) | Protein is localized in various compartments (integral to membrane). | Promotes the protein encoded by this gene as a member of the glucagon receptor family of G protein-coupled receptors. | GLP1R is known to be expressed in pancreatic beta cells. Activated GLP1R stimulates the adenylyl cyclase pathway which results in increased insulin synthesis and release of insulin. Consequently GLP1R has been suggested as a potential target for the treatment of diabetes. | [39,40] |
| Aldolase B, fructose-bisphosphate (ALDOB) | Protein is localized in various compartments (cytosol, lysosome, microsome and others). | Fructose-1,6-bisphosphate aldolase is a tetrameric glycolytic enzyme that catalyzes the reversible conversion of fructose-1,6-bisphosphate to glyceraldehyde 3-phosphate and dihydroxyacetone phosphate. | ALDOB is involved in cellular response to extracellular stimulus; cellular responses to insulin, glycolysis. It has been associated with type2-diabetes. | [41,42] |
| Phosphofructokinase (PFK); 4 alternative transcripts in the liver: NM_002095.5, NM_001166686.1, NM_001166687.1, NM_001166688.1 | Protein is localized in various compartments (cytosol, membrane, cytosol, soluble fraction). | PFK is the most important enzyme in glycolysis. Three phosphofructokinase isozymes exist in humans: muscle, liver and platelet. These isozymes function as subunits of the mammalian tetramer phosphofructokinase, which catalyzes the phosphorylation of fructose-6-phosphate to fructose-1,6-bisphosphate. | A deficiency of phosphofructokinase can be inherited due to the genetic disorder glycosylation type VII Tarui's disease. Research has shown that this disease can lead to insulin resistance and reduced insulin secretion by beta cells. | [43,44] |

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The endoplasmic reticulum (ER) is a membranous synthesis, metabolism, and transport organelle with vital roles in multiple processes that include the synthesis and processing of proteins and lipids, regulation of glucose and calcium homeostasis. ER acts as a quality control organelle to ensure that the newly made proteins are folded into their correct configuration. A failure in this control leads to accumulation of unfolded proteins that are toxic to cells resulting in a state known as ER stress. The relationship between ER stress and diabetes derived from 1980’s investigations showing that ER function inducing compounds can normalize glucose–insulin homeostasis and eliminate insulin resistance and T2DM. Today it is acknowledged that ER stress plays a central role in peripheral insulin resistance and T2DM at the molecular, cellular, and organism levels [68]. The well-known diabetic glucotoxicity is a metabolic stress that induces ER stress and the unfolded protein response (UPR), which, up to a limit, can afford erroneous protein synthesis reduction and an increase in chaperones manufacture, required for proper protein folding. The disrupted ER function also affects the fate of glucose and promotes hyperglycemia through insulin resistance, stimulation of hepatic glucose production, and suppression of glucose disposal [69] by affecting insulin receptor signaling via c-Jun N-terminal kinase (JNK) hyperactivation and subsequent serine phosphorylation of insulin receptor substrate-1 (IRS-1) [70]. The ER stress is also a major source of reactive oxygen species, which establishes a link with mitochondrial dysfunction, proinflammatory cytokine expression and insulin resistance via activation and nuclear translocation of nuclear factor-κB (NFκB) [71].

**Diabetic Complications- The Wound Healing Failure**

Diabetic chronic and uncontrolled hyperglycemia can result in significant morbidity and mortality from macro- and microvascular complications to lower limb amputation [4]. The significance of these complications is emphasized by the fact that someone in the world dies from diabetes-related complications every seven seconds [72] and diabetes is now the leading cause of kidney failure and lower limb amputations globally [73]. Although a thorough incursion into the molecular basis of diabetic multiorgan complications is not the main goal of this review, a brief mention to Michael Brownlee’s unifying hypothesis is worthy. Hyperglycemia, conceptually defined as the proximal trigger of diabetic complications, has been authenticated through large-scale prospective studies such as the Diabetes Control and Complications Trial (DCCT/EDIC) and the UK Prospective Diabetes Study (UKPDS) [74,75]. According to the unifying hypothesis hyperglycemia unleashes the mitochondrial ROS overproduction causing tissue damage by five major mechanisms: increased flux of glucose and other sugars through the polyol pathway, increased intracellular formation of advanced glycation end-products (AGEs), increased expression of the receptor for advanced glycation-end-products (RAGE) and the cytokines mediating the state of low grade inflammation. Mounting evidences leave no room to doubt on the deleterious role of the advanced glycation-end-products (AGE) accumulation. AGEs induce inflammation, apoptosis and cellular aging. On the other hand, hyperglycemia per se as in combination with pro-inflammatory cytokines reduces the cellular pool of anti-oxidant reserves and creates a pro-oxidant environment by the mitochondrial over-production of reactive oxygen species (ROS). These free radicals hamper the mitochondrial oxidative phosphorylation and provoke DNA damages. Under these oxidant and toxic environment, the cells activate signaling pathways and transcription factors that when their “fine tuning” is out of control lead to further amplification of cellular damages. In parallel, other gene/proteins involved in cells defense, anabolism, and survival become downregulated. It is worth noting to mention that hyperglycemia itself is toxic enough to reduce the expression of some of these genes or its derived proteins as e-NOS. As a consequence, cells become predisposed to onset a precocious senescent program, become refractory to programmed turnover and prone to apoptosis. These “cellular illness” translates into tissue and organs irreversible complications.

As stated above, depletion of mtDNA causes insulin resistance with a drastic reduction in basal and insulin-stimulated glucose uptake and GLUT4 translocation in myocytes. The depletion of mtDNA causes a selective and reversible reduction in insulin receptor substrate (IRS-1) mRNA and protein levels and in insulin-stimulated IRS-1 and Akt2/PKB phosphorylation [49]. Alterations in nuclear-encoded genes that regulate mitochondrial biogenesis, such as PGC-1α, AMP kinase, and CAM IV are considered as genetic basis for T2DM inheritance [66]. Although defects in insulin-stimulated muscle glucose transport and phosphorylation appear as very early events in the pathogenesis of T2DM, controversial data exist in the current literature regarding skeletal muscle mitochondrial biology in the pathogenesis of insulin resistance. Conclusively, if mitochondrial dysfunction is not the major culprit in insulin resistance and T2DM pathophysiology; the organelle dysfunction is at least an active player [67].

As shown in the Figure 1, mitochondrial dysfunction is at least an active player [67].

**Figure 1: Molecular ingredients in T2DM and its complications pathophysiology.** Studies based on cultured cells, gene transcriptional expression and epigenetic regulation have progressed to show that a number of gene/proteins appear to be “imprinted” as to predispose or to lead to insulin resistance and T2DM. Epidemiological studies have also shown the negative impact of environmental factors as life and diet styles. High glucose burden has proved to be toxic to cultured cells as to multiorgan systems under both acute and chronic exposure conditions. A line of evidences has shown that in humans and experimental animals, hyperglycemia is the proximal trigger for the activation of acute phase hepatic reactants and pro-inflammatory cytokines elevation in circulation. In a perpetuate vicious circle, the pro-inflammatory cytokines perturbs insulin receptor signal transduction and thus amplifies hyperglycemia. A cross-activation has been described between the receptor for advanced glycation-end products (RAGE) and the cytokines mediating the state of low grade inflammation in diabetes. Mounting evidences leave no room to doubt on the deleterious role of the advanced glycation-end products (AGE) accumulation. AGEs induce inflammation, apoptosis and cellular aging. On the other hand, hyperglycemia per se as in combination with pro-inflammatory cytokines reduces the cellular pool of anti-oxidant reserves and creates a pro-oxidant environment by the mitochondrial over-production of reactive oxygen species (ROS). These free radicals hamper the mitochondrial oxidative phosphorylation and provoke DNA damages. Under these oxidant and toxic environment, the cells activate signaling pathways and transcription factors that when their “fine tuning” is out of control lead to further amplification of cellular damages. In parallel, other gene/proteins involved in cells defense, anabolism, and survival become downregulated. It is worth noting to mention that hyperglycemia itself is toxic enough to reduce the expression of some of these genes or its derived proteins as e-NOS. As a consequence, cells become predisposed to onset a precocious senescent program, become refractory to programmed turnover and prone to apoptosis. These “cellular illness” translates into tissue and organs irreversible complications.

**Flux through the tricarboxylic acid cycle have been demonstrated to play an important role in the insulin resistance of aging [65].**
The refractoriness to heal and the propensity of diabetic wounds for chronication even in metabolically compensated diabetic patients can be explained by virtue of the metabolic memory. Recent conclusive evidence has emerged from zebrafish diabetic models showing that impaired fin regeneration is a heritable process driven by metabolic memory-mediated epigenetic changes and not from hyperglycemia acute consequences as ROS and AGEs accumulation [78].

**The diabetic wound fibroblast**

The fibroblast is central to the wound healing process by secreting, contracting and remodeling the extracellular matrix (ECM). They also secrete growth factors as important messengers for mesenchymal-to-mesenchymal and epithelial-mesenchymal communication, especially to establish the emerging basement membrane and subsequent re-epithelialization. Therefore, any impediment to fibroblast function is deterrent for normal healing and may result in chronic, non-healing wounds. Under the high glucose burden imposed by diabetes, cutaneous and extra cutaneous fibroblasts appear perturbed. For many years, in vitro models recreating “clinical hyperglycemia” have proved to disrupt normal fibroblasts physiology and derange the secretion of extracellular matrix ingredients. These experiments have suggested that high glucose concentration is the detonator of a downstream cascade of molecular disturbances for the cutaneous fibroblast [79]. Rowe and co-workers pioneered the in vitro models that demonstrated reduced; synthetic, proliferative and secreting capabilities in diabetic cutaneous fibroblasts [80]. In other parallel studies, high glucose concentrations proved to inhibit fibroblast proliferation, while the cells turned refractory to proliferate to growth factors such as insulin-like growth factor type-1 (IGF-I) and epidermal growth factor (EGF) [81]. Following these attractive targets, Goldstein’s findings allowed to establish the hypothesis that diabetic fibroblast replicative life span proportionally decline with subject predisposition under normal glucose concentrations, concluding that persistent, heritable abnormality is present in mesenchymal tissues of overt diabetic and genetically predisposed subjects. Years later, Goldstein also announced that cells obtained from insulin-dependent or insulin-independent diabetic people not only exhibit abnormal replicative capacity in vitro, but that the aging process appeared more precoceously than in non-diabetic counterparts [82]. Other studies showed that the addition of conditioned media from non-insulin-dependent diabetes mellitus wound fibroblasts induced a dose-dependent inhibition in normal fibroblast proliferation apparently related to elevated L-lactate levels [83]. This replicative refractoriness of diabetic fibroblasts has been reproduced by different groups in subsequent years, thus confirming the need for additional external supplements to ensure cell cycle progression [84].

In addition to the onset of a quiescent and senescent phenotype of diabetic wound fibroblasts, their ability for horizontal and vertical migration is also dramatically impaired when compared to normal donor cells in different migration assays [85]. Most of these attributes are reproduced under acute exposures to high glucose concentrations so that migration speed is reduced by ~40 % associated to a decrease in cell directionality and to non-productive protrusive events, such as loss of cell polarization, consistent with the increased activity of Rac1 and the projection of multiple lamellipodia. This experiment concluded that the generation of reactive oxygen species (ROS) may lie behind these abnormalities as they were partially or completely rescued by treatment with N-Acetyl-Cysteine (NAC) [86].

As a consequence of the cutaneous accumulation of advanced glycation-end products (AGEs), the skin collagen increases its biological age. Fibroblasts therefore are not excluded from the onset of a pro-senescent phenotype. Fibroblasts cultured and exposed to an AGE precursor reduce the ability to efficiently migrate because of reduced adherence to the matrix. This observation appeared associated to a higher level of misfolded proteins suggesting an ER stress along with extracellular regulated kinase 1/2 (ERK1/2) and Akt pathways downregulation. AGEs precursors also induce oxidative stress and apoptosis via a cascade of apoptotic proteins as FOXO1, BIM and caspase-3 activation in cutaneous fibroblasts [87]. Attention has been paid during the last years to the role of FOXO1 activation in diabetic connective tissue cells as a major apoptotic effector of AGEs and tumor necrosis factor-alpha (TNF-α). FOXO1 limits wound healing by inhibiting fibroblasts proliferation and promoting cell death [88, 89]. Interestingly, insulin inactivates FOXO1 via Akt leading to its nuclear export and degradation. Defective insulin action in the skin has been proposed as an important mechanism contributing to wound healing defects in diabetes. Perhaps the assorted constellation of the hormone’s pharmacological benefits (increased expression of endothelial nitric oxide synthase, vascular endothelial growth factor, and stromal-derived factor-1α/SDF-1α) observed in experimental and clinical wounds when insulin is topically administered may be attributable to FOXO1 neutralization. Curiously, the acceleration of wound healing occurs in parallel to a local recovery in the expression of proteins involved in insulin signaling pathways [90].

Despite the broad molecular information supporting the diabetic wound fibroblast phenotype of migratory and proliferative arrest, intriguing evidences demand further research. For instance, what is the mechanism underlying the proliferative arrest and the pro-senescent phenotype of the cells even under ideal culture conditions and successive culture passages? It is likely that the emerging findings on the impact of metabolic memory and its epigenetic imprinting make those cells to remind where they used to indwell.

**Diabetic wound endothelial cells**

Angiogenesis is a comprehensive term which indicates the physiological process involving the growth of new blood vessels or neovascularization. This is a vital process for embryological growth, tissue development, and wound healing. Different growth factor families as vascular endothelial growth factors (VEGF), fibroblast growth factor (FGF), angiopoietins, platelet-derived growth factor (PDGF), transforming growth factor-β (TGF-β), in collaboration with other proteins such as integrins, cadherins and ephrins regulate angiogenesis by promoting endothelial cells recruitment, proliferation, migration, co-opting and collar stabilization.

There is an enormous and ever-growing body of evidence indicating the close correlation between hyperglycemia and the abnormalities in endothelial morphology and function [91]. The UK Prospective Diabetes Study (UKPDS) and Diabetes Control of Complications Trial (DCCT) found microvascular disease and hyperglycemia to be intrinsically related. Thus, anomalous angiogenesis is a hallmark of both type forms of diabetes which is clearly and early observable during the process of granulation tissue growth, condition that has been successfully reproduced in animal models [92]. Furthermore, insulin has a dramatic impact on the endothelial homeostasis by its ability to stimulate NO release via a cascade that involves activation of the phosphatidylinositol 3-kinase (PI3K)-Akt signaling and endothelial nitric oxide synthase (e-NOS) phosphorylation. The later being of paramount importance in angiogenesis and wound healing as described below [93]. As depicted for fibroblasts, high glucose and the glycated by-products exert a toxic effect on endothelial cells and the
vascular wall in general. In parallel, the endothelial cells per se seem to be a very sensitive target to high glucose. Endothelial dysfunction is intricately related to insulin resistance through the stimulatory effects of insulin on glucose disposal and NO production in the endothelium. High glucose ambient has been shown to disturb endothelial cells cycle, disrupt tube formation, increase DNA damage, delay endothelial cells replication, and induce apoptosis [94]. In vitro models simulating “normoergicenia” and “hyperglycemia” have demonstrated that under a high glucose ambient, proliferation of dermal microvascular endothelial cells appear impaired [95] while apoptosis is enhanced [96] which may be related to the concomitant up-regulation of pro-inflammatory cytokines as TNF-α, death receptors TNF-R1 and Fas ligand in a variety of cultured endothelial cells [97].

Hyperglycemia and the accumulation of AGEs disturb the role of angiogenic growth factors as VEGF, its receptor, its signaling pathway, thus affecting endothelial proliferation, migration, and endothelial progenitor cells (EPCs) release and recruitment. Insulin resistance interrupts the NO-mediated angiogenic positive regulation over angiogenic growth factors such as VEGF, FGF, and TGF-β [98]. Compelling evidences indicate that at least a portion of the hyperglycemia and AGEs-mediated endothelial damages and dysfunctions are associated with an impaired mitochondrial activity resulting in mutations of mitochondrial DNA by a disproportionate reactive oxygen radical production leading to an inflammatory reaction and apoptosis [99]. In a similar manner many of the metabolic conditions associated with diabetes stem from a failure in NO synthesis or its precarious degradation. In this respect, the integrity of the Akt/e-NOS coupling pathway for a normal endothelial function appears compulsory [100].

Endothelial progenitor cells (EPCs) are active players for the maintenance and repair of endothelial cells. They participate in angiogenesis as they proliferate, migrate and differentiate, and are a source for proangiogenic factors and cytokines. Multiple evidences indicate that the number of circulating EPCs is decreased under both clinical forms of diabetes, which is likely to be involved in the pathogenesis of vascular complications [101]. Under experimental diabetic conditions the EPCs number appears significantly decreased in the bone marrow as in the peripheral blood which was reverted by treating the mice with insulin [102]. In general the bone marrow-derived EPCs in the diabetic patients are considered as dysfunctional, producing fewer endothelial cells and with reduced replicative and migratory potential [103]. As to fully divert the physiological role of EPCs in tissue repair and angiogenesis, the duet hyperglycemia-ROS stimulate the EPCs to produce pro-angiogenic cytokines and to shift NO production by elevating iNOS and decreasing eNOS [104]. As described for other cells, AGEs treatment disrupts EPCs physiology thus leading to a downregulation of e-NOS and Bcl-2 expression, as well as an elevation in cyclo-oxygenase-2, Bax, NF-xB, and caspase-3 in a MAPK (ERK/P38/JNK)-dependant manner [105].

The diabetes-mediated vascular damage is perhaps the most outspoken and ancestrally identified emblem of this disease. It is varied and broad as it is the concept of systemic endothelial dysfunction. Diabetes distorts the angiogenic program to ironically culminate with a misdistribution of soluble angiogenic factors: shortage where and when required (lower extremities skin) but overproduced where and when not needed (retina). It is also challenging to understand how and why microvascular morphological changes that recreate chronic, life-time processes are readily identified in a 7-days old granulation tissue fragment, even in compensated patients. This incites to investigate which are the diabetes operational local and/or systemic forces that disrupt vascular morphogenesis.

### The diabetic granulation tissue

Tissue regenerative capacity has been neglected along the species evolution. Thus, scarring process has emerged as an urgent alternative to favor the structural and functional restoration of a wounded zone. Within these events, the process of granulation tissue formation is pivotal as it constitutes a sort of living-temporary aggregate of cells and proteins, acting as a welding material until tissue continuity is restored. However, the reluctance to trigger and sustain the out-growth of a productive granulation tissue with an appropriate extracellular matrix is typical in diabetic patients, particularly if ischemia concurs. As mentioned, these wounds are characterized by a proliferative arrest, pro-inflammatory, pro-oxidant and pro-degradative phenotype in which a spill-over of proteases degrades extracellular matrix ingredients, growth factors and their receptors [106].

This stubbornness to heal in diabetes is conditioned by systemic and local factors that in complicity counteract intrinsic reparative mechanisms. In a broad systemic context, inflammation and the anabolic deficit can be conceptually mentioned. Diabetic patients with foot ulceration confront alterations of the immune status with an active upregulation of circulating levels of acute-phase proteins, cytokines,
and chemokines that impose a chronic systemic inflammatory profile, and amplify local wound inflammatory networks. The systemically elevated levels of pro-inflammatory response markers and the wound expression of cytokines and chemokines are among the culprits of the abnormal repair mechanism [106]. Another factor to be considered is that diabetes per se is a metabolic disease in which fuel metabolism is perturbed given the rupture of one of the most important anabolic axis of the organism: insulin/insulin-like growth factor type-I. The role of insulin in wound healing is well known by its anabolic effect on wound protein balance favoring synthesis and preventing degradation [107]. Both insulin and IGF-1 appear to act in part by the induction of ATF4 (CREB2), essential for the activation of mammalian target of the rapamycin complex 1 (m-TORC1), which in turn is required for protein synthesis via FOXO-dependent genes repression. At the end, protein synthesis and cellular anabolism prevail [108]. It seems that a finely tuned, constitutive activity level for the insulin and other growth factor tyrosine-kinase receptors is indispensable for cell physiology including the events encompassed within the healing process (Figure 2).

Fibroblasts are the main source of collagen, and the number of fibroblasts can be taken as a measure of repair by their collagen synthesis ability. It is much likely that the growth factors such as epidermal growth factor (EGF), transforming growth factor beta (TGF-β1), insulin-like growth factor (IGF-I), and platelet-derived growth factor (PDGF) that stimulate fibroblasts proliferation, transdifferentiation and the synthesis of matrix components, appear in deficit in diabetic foot ulcers resulting in a scarce extracellular matrix formation. Numerous growth factors (TGF-β1, IGF-I, PDGF) regulate the balanced expression of matrix metalloproteases and their tissue inhibitors (MMPs /TIMPs), while most of them exhibit an altered expression in diabetic foot ulcers. Moreover, the imbalance in the diabetic foot ulcers milieu between TGF-β1 and TGF-β3 in which the former appears down-regulated, may explain fibroblasts quiescence in terms of proliferation and secretion [109]. This phenomenon represents the deficit of one of the most potent pro-fibrogenic and fibroblast-mitogenic growth factors, which at the same time down regulates macrophages activation.

One of the main challenges for the diabetic wound healing is the structuring of a normal matrix in quantity and quality. In general, a poor extracellular matrix formation distinguishes diabetic foot ulcers, which can result from: (a) diminished synthesis, (b) increased degradation by proteolytic enzymes, (c) toxicity due to glycated by-products accumulation, and (d) toxicity by biofilm bacterial contaminants diffusion [110].

The diabetic granulation process does not generally exhibit the orderly cascade of events that characterize normal wound healing. This has been confirmed through the histopathological analysis of granulation tissue biopsies by Loots and co-workers who described the lesions as “frozen” in a chronic low-grade inflammatory state associated to a scarce provisional extracellular matrix [111]. Our group’s serial biopsies from both neuropathic and ischemic ulcers-derived granulation tissue have identified histological differences for both types of wounds in the absence of clinical infection. Polymorphonuclear cells (PMN) infiltration is intense and prolonged particularly in neuropathic wounds, co-existing with a scarce extracellular matrix accumulation in which collagen deposit is impoverished (Figure 3). Under more mature stages, neuropathic lesions may also show an abnormal sprout of new small vessels and capillaries that may derive not from a normal angiogenic response but due to arteriovenous shunts. Our observations remind those of Black and co-workers who demonstrated that neuropathic patients exhibit a decrease in fibroblast proliferation and a scarce amount of collagen accumulation within the wound bed [112]. On the contrary, a broadly spread infiltration of round cells predominate in those patients suffering from wound bed ischemia, associated to a fibro-hyaline matrix of “hardened” aspect and abnormal angiogenesis in which vascular wall cellular mosaicism, precocious media thickening, endothelial nuclei hypertrophy and many other defects can be identified (Figure 4). It is likely that the combination of arterial hyperperfusion and glucose toxic derivatives imprint a particular pattern of damage to the morphogenesis of vessels within the wound [113]. These observations incite to speculate that the biochemical microenvironment in ischemic and neuropathic diabetic wounds is different and that the inflammatory “badge” is in correspondence with the wound’s most prevalent pathogenic component. In contrast to acute wounds in non-diabetic subjects, the inflammatory reaction in diabetic appears prolonged, which sharply delays granulation tissue formation and maturation. Data derived from murine diabetic models indicate that the exaggerated inflammatory reaction is related to the prolonged expression of macrophage inflammatory protein-2 (MIP-2) and macrophage chemoattractant protein-1 (MCP-1) [114]. Furthermore, the down-regulation of the anti-inflammatory cytokine IL-10 in diabetic ulcers environment represents the collapse of an important inflammatory repressor [115]. Other evidences indicate...
that PMNs are critical cells toward the acquisition and perpetuation of inflammation and a degradative phenotype. The granulocytes secrete TNF-α and IL-1β, which act as a triggering signal for MMP expression via the common NF-κB signaling pathway. Importantly, the perpetuated homing of PMN within the wound bed is associated to high local levels of elastase secretion, ROS and reactive nitrogen species. High circulating and PMN-associated elastase levels are attributable to poor glycemia control and are currently considered as a risk marker for the development of diabetic angiopathy [116].

Wound contraction failure is a clinical hallmark of diabetic granulation tissue. Fibroblast-to-myoﬁbroblast transdifferentiation represents a key event during wound healing and tissue repair. The contractile force generated by myofibroblasts as a highly specialized cell, speeds the healing process of dermal wounds in healthy humans, accounting for 80-90% of scar tissue reduction [117]. In diabetes, all the biological factors that instrument the onset of a contractile phenotype/activity become reduced, which seems to be mostly attributed to the deleterious activities of TNF-α within the wound, thus suppressing α-SMA expression [118].

As granulation tissue growth is the corollary of numerous cellular concerted events and converging soluble forces as growth factors, many more unanswered questions still remain as a challenge: 1) What are the driving forces underlying the microscopic structural differences between neuropathic and ischemic ulcer beds? 2) What explains the “inheritance” of vascular changes such as the Monckeberg’s media thickening in nascent arteries within an incipient granulation tissue? 3) Why granulation tissue is histomorphologically abnormal even in metabolically compensated patients?

The Rationale for a Novel Delivery Procedure. EGF Intralesional Infiltration is Justified and Improves Diabetic Wound Healing Response

Topical application of recombinant human growth factors (GFs) dates back to almost 30 years. Their arrival sparked hopes as tissue healing magic bullets. To our understanding the setback stemmed from a rigorous clinical trial in which EGF was topically administered to acute, controlled and experimentally induced wounds in healthy volunteers [119] somewhat turned off the early medical enthusiasm. Accordingly, this clinical failure warned about the need for additional research in GFs pharmacology as in wounds biochemical environment. The study also suggested that even those acute, clean and controlled wounds in healthy subjects may not represent a ‘comfortable’ substrate for GFs physical and chemical integrity. Previous clinical evidences had already rendered disappointing results possibly because of local bioavailability limitations [120]. Unquestionably, the need to precondition the wound bed to preserve GFs local bioavailability for subsequent receptor stimulation and downstream signaling activation emerged as novel paradigmatic concepts [121]. This concept introduced by Mast and Schultz more than 20 years ago revolutionized wound healing pharmacology [122] and hitherto, a myriad of novel approaches to treat chronic wounds have been devised. Despite these historical messages, the topical application of growth factors onto the wound surface still remains as the most popular delivery method for growth factors and peptides [8].

Chronic wounds, particularly diabetic ones appear as a rich source of serine proteases, especially elastase which prompted the notion that optimal healing of chronic wounds may require to attenuate excessive proteolysis. Furthermore, diabetic wounds exhibit an exaggerated susceptibility to host an abnormal bacterial burden which produces proteases that degrade growth factors, their receptors, amplify the inflammatory response and deter growth factor storage within the extracellular matrix [123]. The existence of a peculiar biofilm that rapidly regenerate following sharp debridement, abundant in glycation-derived toxins, and characterized by a difficult to eradicate pathogenic polymicrobial community [124,125], is another significant characteristic of diabetic foot ulcers (DFU). The understanding of the pathogenic significance of the biofilm in chronic wounds render current explanation on why topically administered growth factors may have failed in healing some chronic wounds. Today, decades of efforts toward growth factor pharmaceutical development may be considered a failure. The above mentioned data provides support for the premise that impaired GFs availability may act as a rate limiting factor in diabetic wound healing [126]. According to Smill’s analysis of 4 randomized studies, combining surgical debridement with the daily topical application of PDGF-BB a modest 15% improvement in the rate of fully healed diabetic ulcers was revealed as compared to placebo [127]. Other growth factors which at a moment were clinically investigated in the USA [128] vanished along the way because of lack of efficacy even when topically applied to acute, controlled, partial-thickness wounds in healthy volunteers. All these pieces of knowledge encouraged the search for more effective treatments and delivery strategies to ensure growth factor optimal activity. So far, these have included: (a) growth factor coding gene delivery to the wound cells through an adenoviral vector [129], (b) use of agents that inhibits metalloproteinases and TNF-α converting enzyme (TACE) [130], (c) wound bed preparation as to diminish wound microenvironment hostility [121].

The above described pieces of knowledge contributed to shape the idea that injecting EGF deep into the wound base and contours would allow for a larger pharmacodynamic response in terms of granulation tissue growth and wound closure. Another factor that nourished the idea of intralesional infiltration was the seminal ﬁnding of Cross and Roberts who showed very limited diffusion of radio labeled growth factors administered onto the wound surface, thus remaining into the upper levels of the granulating tissue [131]. In other words, these authors demonstrated that growth factors downward diffusion is actually hindered by a number of physical and chemical factors; thus becoming limitedly available for the responsive cells. It is noteworthy to mention that according to classic studies, the interaction between EGF and its receptor requires a prolonged time window interaction in order to commit wound cells for proliferation [132]. These ideas along with the animal experiences described below provided the rationale of the intralesional delivery via local infiltration.

Experiments from our group suggested a possible reduction of EGF bioavailability by proteases derived from non-infected, acute, controlled experimental wounds. It was somewhat surprising as other studies had already established proteolysis on GFs and their receptors in chronicity circumstances [133]. Further evaluations of cumulative profiles of radiolabeled EGF in a rat full-thickness wound model showed EGF degraded subspecies and a short residence time within the wound contours [134]. We had the experience of injecting EGF locally into denervated rat hind limbs upon sciatic nerve full-thickness cut. In addition to significantly assist in neurological restoration, the treatment enhanced limb peripheral soft tissue survival by delaying or preventing the onset of plantar ulcers and toe necrosis. The experiments offered an important lesson: locally injected EGF could stimulate the survival and repair of cutaneous and adjacent soft tissues in a context of circulatory neurogenic deterioration, somewhat similar to the diabetic lower limb...
clinical condition [135]. Trophic ulcers appeared to be prevented. We subsequently showed, by a series of experiments that single or repeated systemic or local EGF injections exerted 'clear-cut' cytoprotective and proliferative responses, supporting the intrinsic ability of EGF at supraphysiologic concentrations to unleash biological events required for tissue repair [136]. These experiments in animals rendered the first pharmacological rationales for its investigational use in the treatment of diabetic foot ulcers, including ulcers under ischemic condition: (a) EGF contributed to attenuate lipid peroxidation and free radicals cytotoxicity, (b) EGF reduced local mediators of inflammation, (c) prevented cellular demise under episodes of ischemia/reperfusion, and (d) stimulated epithelial cells proliferation. Globally speaking, injected EGF could control three major ingredients of diabetic ulcers: inflammation, ischemia and oxidant cytotoxicity. The biological rationale for the intra and peri-lesional infiltration has been recently reviewed. The major fundamental is based on the finding of three major cellular stratum among the horizontal axis from the surface to the bottom of the granulation tissue in diabetic ulcers. The deeper layer expresses far more EGF receptors and far more cyclin D1 than the superficial counterpart. By the contrary, fibroblasts on the surface express far more AGE, prohibitin and elastase than those located at the depth. Thus, the infiltration not only by-passes chemical and physical barriers for degradation but also turns available EGF to those cells with larger EGF receptors expression and proliferative capability [137]. The first clinical evidences on EGF infiltrative treatment involved diabetic foot ulcers and amputation residual bases. All the lesions were chronic, complex and recalcitrant to heal, staged as grades III or IV of the Wagner’s scale. The efficacy showed in this type of wounds [138] paved the way for a solid clinical development, which successfully culminated with a multicenter, double-blinded, placebo-controlled phase III clinical trial in which an unprecedented efficacy and safety profile were reported for high-grade ulcers [139-144]. Ongoing studies from our group (Jorge Berlanga-Acosta, unpublished data) have shown that locally infiltrated EGF is able to trigger acute and long term gene responses. Early gene response includes the up-regulation of proliferative, pro-survival and pro-anabolic genes. At the long term response (21 days after initiation) EGF induces the expression of a constellation of other growth factors that contribute to the healing process. In other words, the locally infiltrated EGF restores the deficit of growth factors brought about by diabetes and resumes the activity of tyrosine kinase receptors which appear crucial for the healing process, especially in the diabetic context where they are depressed [145].

A freshly published review by Lopez-Saura and co-workers in which the post-marketing clinical experience in more than 2000 subjects receiving intraleisional EGF for advanced lesions was analyzed, confirmed the results of the clinical trials with 75 % probability of complete granulation response, 61 % healing, and a 16 % absolute benefit includes ischemic patients and again a broad safety profile were reported for high-grade ulcers [139-144]. Ongoing studies from our group (Jorge Berlanga-Acosta, unpublished data) have shown that locally infiltrated EGF is able to trigger acute and long term gene responses. Early gene response includes the up-regulation of proliferative, pro-survival and pro-anabolic genes. At the long term response (21 days after initiation) EGF induces the expression of a constellation of other growth factors that contribute to the healing process. In other words, the locally infiltrated EGF restores the deficit of growth factors brought about by diabetes and resumes the activity of tyrosine kinase receptors which appear crucial for the healing process, especially in the diabetic context where they are depressed [145].

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Concluding Remarks

Despite the presence of novel therapeutic agents developed against T2DM; an ordinary disease of the past has turned into a modern day pandemic. Since hyperglycemia is just the clinical tip of the iceberg and given the disease pathogenic complexity, T2DM poses as an investigational and clinical challenge. In addition to the broad constellation of enzymes, signaling switchers, transcription factors and mitochondria-controlling genes involved in the pathogenic cascade of T2DM; organelles like mitochondria and the endoplasmic reticulum are also committed. Notwithstanding the boundaries between cause and consequence and the individual commitment of each ingredient within the disease’s puzzle remain blurry and elusive.

It is known that high glucose burden is toxic for fibroblasts, endothelial cells and keratinocytes. All these cells become arrested in migration and proliferation, gain a pro-senescent phenotype and may develop apoptosis. However, the fact that cultured fibroblasts and myocytes exhibit behavioral traits that have proved useful in predicting the onset of diabetes; and that some of these traits are shown by cultured cells from clinically healthy descendants of diabetic progenitors, indicate the responsibility of innate epigenetic and genetic heritable elements whose tunings is largely associated with external factors. In support to this notion is the recent enrichment of the Brownlee’s unifying hypothesis with epigenetic changes as major rulers in the metabolic memory. The propensity of diabetic wounds for chronification in compensated patients is influenced by the matrix and cell metabolic memory. This relatively new concept remains treatment-orphan, appears irreversible and very much responsible for the multi-system complications progression. Thus, it is likely that T2DM and its complications will continue their striking course for years ahead.

Although the molecular fundamentals of diabetic wound chronification are somewhat understood, the marriage between wound healing failure and diabetic innate immunity deficit tribute 80 % of all non-traumatic amputations worldwide. This simple notion indicates that first and second line/high technology interventions such as recombinant molecules, bioactive dressings, subatmospheric pressure, and others, still exhibit an insufficient therapeutic impact. Oversized neuropathic ulcers are hard and long to heal. The ischemic ones, even undergoing satisfactory reperfusion procedures may take months to a year for re-epithelialization at the most optimistic scenario. Pharmacological doses of growth factors are intrinsically capable of resetting the diabetic wound bed cell biological competence by resuming the activation of different tyrosine kinase axis. The bioavailability limitations imposed by the wound environment in which the growth factors and their receptors are degraded, and those imposed by the limited physical diffusion of the proteins downward within the granulation tissue have limited growth factor incorporation to the clinical armamentarium. Thus far, the EGF infiltrative intervention stands as the unique pharmacological intervention jerking outsize and ischemic poor-prognosis wounds towards a rapid and sustained healing response.

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