Density- and trait-mediated effects of a parasite and a predator in a tri-trophic food web

Aabir Banerji, Alison Duncan, Joanne Griffin, Stuart Humphries, Owen Petchey, Oliver Kaltz

To cite this version:

Aabir Banerji, Alison Duncan, Joanne Griffin, Stuart Humphries, Owen Petchey, et al.. Density- and trait-mediated effects of a parasite and a predator in a tri-trophic food web. Journal of Animal Ecology, Wiley, 2015, 84 (3), pp.723-733. 10.1111/1365-2656.12317. hal-02353410

HAL Id: hal-02353410

https://hal.umontpellier.fr/hal-02353410

Submitted on 7 Nov 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.
Density- and trait-mediated effects of a parasite and a predator in a tri-trophic food web

Aabir Banerji, Alison B. Duncan, Joanne S. Griffin, Stuart Humphries, Owen L. Petchey and Oliver Kaltz

1Institut für Evolutionsbiologie und Umweltwissenschaften, Universität Zürich, Winterthurerstrasse 190, Zürich CH-8057, Switzerland; 2Institut des Sciences de l’Evolution, UMR 5554, Université Montpellier II, Place Eugène Bataillon, 34095 Montpellier Cedex 05, France; and 3School of Life Sciences, University of Lincoln, Brayford Pool, Lincoln, Lincolnshire LN6 7TS, UK

Summary

1. Despite growing interest in ecological consequences of parasitism in food webs, relatively little is known about effects of parasites on long-term population dynamics of non-host species or about whether such effects are density or trait mediated.

2. We studied a tri-trophic food chain comprised of (i) a bacterial basal resource (Serratia fonticola), (ii) an intermediate consumer (Paramecium caudatum), (iii) a top predator (Didinium nasutum) and (iv) a parasite of the intermediate consumer (Holospora undulata). A fully factorial experimental manipulation of predator and parasite presence/absence was combined with analyses of population dynamics, modelling and analyses of host (Paramecium) morphology and behaviour.

3. Predation and parasitism each reduced the abundance of the intermediate consumer (Paramecium), and parasitism indirectly reduced the abundance of the basal resource (Serratia). However, in combination, predation and parasitism had non-additive effects on the abundance of the intermediate consumer, as well as on that of the basal resource. In both cases, the negative effect of parasitism seemed to be effaced by predation.

4. Infection of the intermediate consumer reduced predator abundance. Modelling and additional experimentation revealed that this was most likely due to parasite reduction of intermediate host abundance (a density-mediated effect), as opposed to changes in predator functional or numerical response.

5. Parasitism altered morphological and behavioural traits, by reducing host cell length and increasing the swimming speed of cells with moderate parasite loads. Additional tests showed no significant difference in Didinium feeding rate on infected and uninfected hosts, suggesting that the combination of these modifications does not affect host vulnerability to predation. However, estimated rates of encounter with Serratia based on these modifications were higher for infected Paramecium than for uninfected Paramecium.

6. A mixture of density-mediated and trait-mediated indirect effects of parasitism on non-host species creates rich and complex possibilities for effects of parasites in food webs that should be included in assessments of possible impacts of parasite eradication or introduction.

Key-words: density-mediated indirect interaction, Didinium, Holospora, Paramecium, trait-mediated indirect interaction

Introduction

Parasitism is ubiquitous. Parasites infect hosts across all trophic positions and can drastically alter host behaviour, morphology and life-history patterns (Hatcher, Dick & Dunn 2006; Lafferty et al. 2008; Sukhdeo 2010). In so doing, they affect food web properties such as stability, species interaction strengths and energy flow (Lafferty, Dobson & Kuris 2006; Lafferty & Kuris 2009; Hatcher & Dunn 2011). Some of these food web level effects of
parasitism are likely caused by effects of parasites on non-host species. Such indirect effects of parasites can occur via density and/or trait mediation (Hudson, Dobson & Lafferty 2006; Hatcher, Dick & Dunn 2012, 2014). Density-mediated effects are likely pervasive and large, since parasitism commonly has negative effects at the population level that are at least as large as the effects of predation (Lefèvre et al. 2009; Watson 2013). Trait-mediated effects of parasitism can also be important. For example, parasitism can change the behaviour of hosts, affecting their likelihood of being consumed (Lagrué et al. 2007; Yanoviak et al. 2008), their feeding rates (Crompton 1984; Dick et al. 2010), and their activity as ecosystem engineers (Mouritsen & Poulin 2005). Understanding the roles of parasites in food webs therefore requires knowledge about the density- and trait-mediated indirect effects that parasite species can have via direct effects on host species. This knowledge would also aid assessments of possible impacts of parasite introductions or removals on non-host species (Torchin et al. 2002; Koop et al. 2011; Barry et al. 2014).

While trait-mediated effects of parasites can manifest quickly (i.e. faster than a generation), density-mediated effects (caused by reduced host reproductive rate, for example), will manifest over longer time-scales (i.e. time-scales equivalent to multiple host generations). Consequently, studying trait- and density-mediated effects empirically often entails long-term investigation to obtain long-term data. Such data are scarce for naturally occurring food webs (Williams 2009) and also present difficulty when assigning observed patterns to processes. One approach to circumventing these obstacles is to employ manipulative experiments with rapidly reproducing organisms such as protozoa and bacteria. In their review of microcosm studies, Jessup et al. (2004) highlight important contributions that such experiments have recently made to our understanding of ecological/evolutionary processes and note that microcosm studies have also played a historical role in the shaping of ecology.

To allow for diverse effects of parasitism on non-host species, our experimental community was a tri-trophic food web, with the host (Paramecium caudatum Ehrenberg) occupying the intermediate trophic level. Hence, the host can be both a consumer and a resource. The base of the food chain was the bacterium Serratia fonticola, and the top of the food chain was the predator Didinium nasutum Stein. The parasite was the Paramecium-specific Holospora undulata (Fig. 1). Didinium nasutum and P. caudatum predator–prey dynamics in the absence of parasites are well studied (Li et al. 2013) and have been simulated using recent mathematical models (Harrison 1995; Kozlova, Singh & Easton 2002). Both species are cosmopolitan freshwater ciliates that reproduce via binary fission. Didinium feeds exclusively on other ciliates and mainly on Paramecium, while Paramecium is primarily bacterivorous (Berger 1979).

Harrison (1995) and others obtained quantitative agreement between observed and predicted dynamics by assuming that Didinium exhibits a sigmoidal functional response and a delayed numerical response on Paramecium (Kozlova, Singh & Easton 2002). We found, however, that a more recent model developed by Li et al. (2013) was the most conducive for the purposes of the present study. Though less parsimonious than its predecessors, the Li et al. model is structured in a way that ensures that all parameter estimates are biologically realistic (i.e. within ranges that have been empirically derived). Moreover, the culture methods used in the experiment to which the model had originally been tailored were very similar to those employed in the present study. This model therefore provided an ideal baseline of comparison for quantifying the effects of parasitism of Paramecium by the bacterium H. undulata.

Holospora undulata is a sessile, single-host, bacterial parasite (Gromov & Ossipov 1981) with two developmental stages: a long (c. 20 μm), tindle-shaped infectious form and a smaller (5 μm), round reproductive form. While foraging, Paramecium ingests the infectious form, which – if it survives – ends up in the micronucleus and differentiates into the reproductive form. The reproductive form multiplies, fills the nucleus and begins to differentiate again. Reproductive forms are vertically transmitted to the daughter cells of Paramecium during mitosis. Infectious forms are released for horizontal transmission during mitosis or when Paramecium dies (Fokin 2004). Holospora has been observed in multiple locations throughout the range of its host, indicating that it, too, is cosmopolitan (Fujishima 2009).

Holospora affects Paramecium morphology, reproduction and behaviour – causing, for example, earlier onset of clonal decline, reduced division rates, shrunken buccal (‘mouth’) cavity and shortened cell length (Table S1, Supporting information). Finding that Holospora reduces Paramecium’s dispersal capacity, Fellous et al. (2011) speculated that Holospora may also reduce Paramecium’s per capita mobility. Via these direct effects on Paramecium, Holospora can theoretically have indirect effects on the long-term abundances of Didinium and Serratia. For example, by reducing Paramecium abundance, Holospora may indirectly cause Didinium to grow slower and consume fewer Paramecium cells. This, in turn, may allow both Paramecium and Didinium to persist, with Didinium failing to achieve sufficient abundance to eliminate all of its prey (Salt 1979). Since Paramecium consumes Serratia, Holospora’s reduction of Paramecium abundance may also indirectly cause an increase in Serratia abundance. These would be density-mediated indirect effects (Abrams 2007). Trait-mediated indirect effects of Holospora could yield similar results.

The behaviour and morphology of Paramecium affect both its vulnerability to predators (Salt 1979; Hewett 1988) and its foraging ability (Hall et al. 1976; Fenchel 1980). Paramecium responds to certain physical and chemical predator cues with sudden changes in direction and bursts of speed (Knoll, Haacke-Bell & Plattmer 1991; Hamel et al. 2011). Paramecium size determines its rate of...
capture and handling by *Didinium* (Hewett 1980) and very likely its encounter rate with bacterial prey (Fenchel 1980; Shimeta & Jumars 1991; Verity 1991). To summarize, *Paramecium* has a suite of traits that influence and are influenced by both parasitism and predation, and this creates the potential for its predator, parasite and prey to each have trait-mediated indirect effects on one another.

The objectives of the present study were to (i) quantify the effects of predation and parasitism with respect to the population dynamics of the intermediate consumer (*Paramecium*) and its resource (*Serratia*), (ii) determine whether effects of predation and parasitism function additively or synergistically, (iii) assess whether these effects are predominantly density or trait mediated and (iv) evaluate potential density- and trait-mediated indirect interactions between the predator (*Didinium*) and parasite (*Holospora*). Controlled predation experiments, mathematical modeling, and semi-automated image analysis of *Paramecium* behaviour and morphology were used to achieve these objectives.

**Materials and methods**

**CULTURING OF MICROBIAL SPECIES**

Species were cultured in growth medium consisting of 0.55 g Carolina Biological Supply protozoan pellets, 0.5 mL concentrated Chalkley’s medium and 1 L reverse osmosis-purified water. Methyl cellulose (0.2041 g L⁻¹) was added to the medium to increase its viscosity to c. 5 cP at 20 °C, a technique shown to prolong coexistence between *Didinium* and *Paramecium* by reducing the swimming speed of both species equally without poisoning either species or providing refuge space for *Paramecium* (Luckinbill 1973; Veilleux 1979). Although studies have also specified the importance of using low-nutrient growth medium to prolong *Didinium-Paramecium* coexistence (Harrison 1995), nutrient concentrations below that which was used in the present study were not enough to sustain our infected *Paramecium* cultures (data not shown). Once autoclaved and allowed to cool to c. 20 °C, the medium was inoculated with *S. fonticola*.

Isogenic stock cultures were prepared from clonal lines of parasite-free and *Holospora*-infected *Paramecium* by adding one uninfected or one infected individual to the growth medium. We used a *Paramecium* strain originally collected near Venice, Italy and infected in the laboratory (see Duncan et al. 2010). *Didinium* was obtained from Sciento (strain P220), and an isogenic stock culture was prepared by rearing a *Didinium* cell and its subsequent offspring in petri dishes containing growth medium inoculated with c. 150 cells mL⁻¹ of uninfected *Paramecium* (of a separate strain of uncertain origin).

**EXPERIMENTAL MANIPULATION OF PARASITISM AND PREDATION**

To quantify the separate and combined effects of predation (by *Didinium*) and parasitism (by *Holospora*) on *Paramecium* and *Serrata*...
ratia abundance and the indirect effect of parasitism (of Paramecium by Holospora) on Didinium abundance, a two-way fully factorial design was used, with one factor being the presence/absence of Didinium and the other being the presence/absence of Holospora. As a control for changes in Serratia abundance independent of the effects of other species in the system, Serratia was also grown alone. For brevity, we use the following treatment codes: the letter S if Serratia was present (which was the case in all microcosms), P if Paramecium was present, D if Didinium was present, H if Holospora was present, and a hyphen (−) in place of D or H if either or both species were absent. The letter S by itself denotes the Serratia-only control.

The four treatment combinations (SP-, SPD-, SP-H, SPDH) and S were thrice replicated for a total of 15 experimental units. Each replicate was in a 50-mL Falcon tube (Becton, Dickinson and Company – BD Biosciences © 2013, 1 Becton Dr., Franklin Lakes, NJ, USA) containing 30 mL of growth medium that had been inoculated with Serratia 2 days prior to Day 0 of the experiment. On Day 0 of the experiment, 1 mL was extracted from each replicate tube to estimate the starting abundances of Serratia. Manipulation of the presence/absence of Holospora was performed by replacing this 1 mL with either 1 mL from the uninfected Paramecium stock culture or 1 mL from the Holospora-infected Paramecium stock culture, as appropriate to the treatment. S replicates were inoculated with 1 mL from the same Serratia-containing growth medium that had been used to establish the Paramecium stock cultures, so that they remained the same volume and experienced the same handling conditions as the treatment replicates.

Manipulation of the presence/absence of Didinium was performed by adding 30 Didinium cells via micropipette to each replicate of the SPD- and SPDH treatments after Paramecium had exited its exponential growth phase (Day 11). Subsequently, 1 mL from each tube was sampled once a day, for 3 days a week (every second day, followed by a 2-day break), and replaced with fresh bacterized medium at the end of each week. Upon sampling and upon replacement, microcosms were mixed thoroughly. These procedures created semi-continuous culture conditions that permitted long-term observation of dynamics (McGrady-Steed, Harris & Morin 1997; Banerji & Morin 2009). All tubes were stored in the same compartment of the same incubator, which kept a constant temperature of 20 °C and a 12-h light/dark cycle.

The abundances of healthy and infected Paramecium and of Didinium were estimated via direct counts using a light microscope. If individuals of either species were not detected in the 1-mL sample of a replicate, a systematic search was performed of the entire volume of the replicate. Serratia abundance was estimated via serial dilution and plating. Once a week, 5–20 cells from the infected populations of Paramecium were isolated, stained with lacto-aceto-orcein (1%; Görtz & Dieckmann 1980) and examined under the microscope at 400× magnification to verify that these populations remained infected throughout the experiment.

Variation in species abundances was analysed using repeated-measures generalized linear mixed models, with time as a continuous linear covariate and replicate identity as a random factor. Note, however, that treating time as a factor gives the same ecological interpretation. We used a logistic regression approach, with population size of Paramecium, Didinium or Serratia as response variables, and an underlying Poisson error and log link function. We fitted fully factorial models, containing all possible interactions between experimental treatments and the day covariate. Analyses were restricted to the time interval between Day 13 (2 days after the introduction of Didinium) to Day 43 (last date before global Didinium extinction). Complementary analyses that allowed for auto-regressive error structure and therefore accounted for temporal autocorrelation in the data did not significantly improve model fits (not shown).

MODELLING OF PREDATOR–PREY DYNAMICS

To evaluate which of the processes underlying the predator–prey interaction of Didinium and Paramecium are less likely to be altered by the presence of Holospora, we fit to the data the deterministic model developed by Li et al. (2013). This approach also enabled us to explicitly account for observed variation in abundances of Paramecium at the time that Didinium was introduced to the Didinium-containing microcosms. The Li et al. model was specifically designed for interpreting the predator–prey dynamics of Didinium and Paramecium in growth medium thickened with methyl cellulose. We assumed that four parameters of the model could be affected by parasitism: the growth rate of Paramecium, the carrying capacity of Paramecium, the maximum attack rate of Didinium on Paramecium and the maximum birth rate of Didinium feeding on Paramecium. Details of the model, including the methods that were used to incorporate the potential effects of Holospora, are given in Appendix S1 (Supporting information). The model was fit to the population dynamics via maximum-likelihood estimation, fit was assessed using AIC, and estimates of effects of parasitism on parameter estimates were tested with z-tests.

EFFECTS OF PARASITISM ON PARAMECIUM BEHAVIOUR AND MORPHOLOGY

Movement pattern, swimming speed, cell shape and cell size were extracted from video recordings of Paramecium cells with differing levels of infection – ‘overtly infected’, ‘covertly infected’ or ‘uninfected’. Cells categorized as overtly infected were drawn from the infected stock culture and were conspicuous due to the presence of massively inflated micronuclei (loaded with high numbers of infectious forms) that could already be identified as opaque spots in the cytoplasm at low magnification under the microscope. Covertly infected cells did not exhibit obvious outward symptoms of infection, and their less inflated micronuclei carried fewer, if any, infectious forms (Fig. S1, Supporting information; see also Kaltz & Koella 2003).

Recordings were acquired, analysed and processed in accordance with the workflow proposed by Pennekamp & Schtickzelle (2013), using a stereomicroscope (Leica M205 C; Leica Mikrosysteme Vertrieb GmbH, Ernst-Leitz-Strasse 17-37, 35578 Wetzlar, Germany) and mounted digital CMOS camera (Hamamatsu C11440) in combination with the software programs ImageJ (Abramoff et al. 2004) and x (R Development Core Team 2008). Paramecium cells were transferred via micropipette into 1 mL of growth medium spread across a glass Sedgewick–Rafter counting cell. Videos were recorded for 5 s with a 40-ms field delay and 10-ms exposure (giving 25 frames per second) at low (7.8×) magnification. To minimize blur and achieve highest optical resolution and contrast, the image was set to grey scale, and high-intensity external illumination was placed around the stage plate (Schott VisiLED MC 1500; SCHOTT AG, Hattenbergstrasse 10, 55122 Mainz, Germany). So that the software could visually sep-
brate Paramecium from artefacts, the video recordings were converted to 8-bit format, and a size threshold of 10–255 pixel lengths was specified for Paramecium. Videos in which Paramecium ceased swimming, swam vertically or exited the field of view were discarded. ImageJ’s Particle Analyzer and Particle Tracker returned \( x,y \) coordinates (pixel locations within the field of view) for cells within each frame of each video – along with estimates of length and width in pixels, aspect ratio (dimensionless ratio of length to width), and cross-sectional area in square pixels.

Mobility was quantified in terms of the frequency of turns made by Paramecium and the average swimming speed of Paramecium in each video. Turns were defined as movements that caused Paramecium to deviate at least 45 degrees from its original trajectory. If, throughout the video, Paramecium deviated >45 degrees from its original trajectory, its movement pattern was defined as being linear (having no turns).

Swimming speed was defined as spatial displacement over time. This was calculated by taking the square root of the sum of the squared change in position along the \( X \)-axis and the squared change in position along the \( Y \)-axis (Pythagorean Theorem). The resulting measure was converted from pixels per frame to millimetres per second and then averaged for each Paramecium cell. To ensure that this measure of swimming speed was not confounded by how often Paramecium made turns and how much it slowed down or sped up at the beginning or end of a turn, only linear parts of trajectories were used to make the calculation. Also omitted were videos in which Paramecium was undetected or misidentified by ImageJ’s Particle Tracker (e.g. due to quick turns and loss of frames resulting from deletion of background artefacts), leaving a total of 55 videos upon which to base the calculation. Estimates of cross-sectional length, width, area and aspect ratio were calculated for these same remaining videos by taking the mean of the output of ImageJ’s Particle Analyzer across frames for each cell. Effects of infection level on swimming speed, aspect ratio, total per capita turns and cross-sectional area were assessed using one-way analysis of variance (ANOVA).

To assess whether differences in swimming speed and cell length among infected and uninfected Paramecium were enough to create differences in Paramecium’s predicted rate of encounter of Serratia, we employed two separate methods of estimating encounter rates: one developed by Fenchel (described in Shimeta & Jumars 1991), the other by Verity (1991). Details regarding the terms used in these equations are given in Appendix S2 (Supporting information).

Results

Protist and Bacterial Abundances

Paramecium–Didinium Dynamics

Despite the addition of methyl cellulose to the growth medium, the population dynamics of Didinium and Paramecium did not exhibit sustained oscillations in any of the replicates. In one replicate of the SPDH treatment, Didinium went extinct immediately after its introduction to the system. This replicate was therefore excluded from analyses.

After \( c \) week, Paramecium abundance reached carrying capacity. Abundances in the predator- and parasite-free control populations (- -) remained high for \( c \) 4 weeks and then declined. Infection with Holospora (SP-H populations) reduced abundance by about one order of magnitude (Fig. 2). Statistical analysis of the temporal dynamics revealed a significant day*predator*parasite interaction (\( F_{1,139} = 9.71, P = 0.0022 \)), indicating different trajectories of infected and uninfected populations after the introduction of Didinium on day 11. Initially, predation by Didinium led to similar rates of decline in Paramecium abundance in both infected and uninfected populations. After Day 22, the decline stabilized in two uninfected (SPD-) populations, while the third went extinct. In contrast, at the same time, abundance began to increase in the two infected (SPDH) populations to levels observed in corresponding uninfected populations with the predator, and even exceeding abundance levels of infected (SP-H) populations in the predator-free treatment (Fig. 2).

Didinium abundance increased initially, reaching peak levels 10–12 days after introduction. However, all populations went extinct over the following 20 days. Didinium generally reached higher abundances feeding on uninfected (SPD-) Paramecium populations than on infected (SPDH) Paramecium populations (\( F_{1,3} = 28.46, P = 0.0129 \)). This difference was particularly pronounced during the initial period after introduction (Days 11–22).

Serratia Dynamics

The abundance of the bacterial resource Serratia was generally lower in the presence of the intermediate consumer Paramecium than in the Serratia-only (S) controls (\( F_{1,13} = 792, P < 0.0001 \)). This reduction was almost 100-fold, when Paramecium populations were at their peak density (Fig. 2). We further detected a significant day*predator*parasite interaction (\( F_{1,139} = 8.39, P = 0.0044 \)): in the absence of Didinium, Serratia abundance was mostly lower when Paramecium was infected (SP-H) than when Paramecium was uninfected (SP-). However, this difference disappeared \( c \) 10 days after the addition of the Didinium predator, such that equivalent Serratia abundances were observed with infected (SPDH) and uninfected (SPD-) Paramecium populations (Fig. 2).

In other words, the combined action of parasite infection (Holospora) and predation (Didinium) on the abundance of the intermediate consumer (Paramecium), as well as that of its resource (Serratia), was non-additive. Specifically, predation by Didinium reduced the negative effect of Holospora infection not only on Paramecium abundance, but also on Serratia abundance – such that the presence of the predator removed the population size advantage observed in uninfected Paramecium populations.

Modelling

Maximum-likelihood estimation produced the parameter estimates listed in Table 1. The best fit of the model to the SPD replicates yielded a negative log likelihood of 104.434. Without Holospora’s potential effects on Didini-
um’s functional response, the best fit of the model to the SPDH replicates yielded a negative log likelihood of 73.196 (Fig. 3). Inclusion of effects of parasitism produced a best fit with a negative log likelihood of 71.599. Only the effects of parasitism on $r$ and $K$ were statistically distinguishable from 0 at the 0.05 level. Based on AIC, the model with effects on Didinium’s functional and numerical responses was less parsimonious than the model with only density-related effects ($\text{dAICc} = 4.9$, d.f. = 5, weight = 0.078).

**Paramecium behaviour and morphology**

Overtly infected, covertly infected and uninfected Paramecium cells did not significantly differ in terms of their total per capita turning frequency ($F_{2,77} = 2.634$, $P = 0.078$) or cross-sectional area ($F_{2,63} = 1.091$, $P = 0.342$). Swimming speed ($F_{2,63} = 4.459$, $P = 0.016$) and aspect ratio ($F_{2,63} = 95.95$, $P < 0.01$), however, did differ among the infectious groups. Swimming speed was higher in covertly infected Paramecium cells than overtly or uninfected cells. Aspect ratio was lowest in overtly infected Paramecium, moderate among covertly infected Paramecium and highest among uninfected Paramecium – meaning that – as is consistent with the literature (Fokin 1985), Paramecium cells became shorter and fatter with increasing parasite load (Fig. 4).

Given the differences in swimming speed and cell length among infected and uninfected Paramecium, both Fenchel’s method and Verity’s method of estimation predict higher encounter rates with Serratia for infected Paramecium than for uninfected Paramecium (Fig. S2, Supporting information).

**Discussion**

We investigated the mechanisms and consequences of parasite-mediated effects in experimental food webs, containing two protozoans (predator and prey), a bacterial parasite and a bacterial prey species. The observed population dynamics of the different protagonists revealed a mix of direct and indirect effects along the food web,
including a modulation of parasitic effects in the presence of a predator. Below, we explore some potential explanations for and implications of the observed results, involving density-mediated and trait-mediated mechanisms.

**Paramecium–Didinium dynamics**

As expected, both parasitic infection by *Holospora* and predation by *Didinium* had a strong negative impact on
Paramecium abundance. Initially, the introduction of the predator led to a massive decrease in abundance of both infected and uninfected Paramecium, and a concomitant increase in Didinium. Didinium grew to lower peak density and tended to die out earlier when preying on infected Paramecium. Since Didinium does not feed on Holospora independently of Paramecium, and Holospora cannot infect Didinium, one can infer that Holospora’s effect on Didinium abundance was mediated via Paramecium. Most likely, this was due to the abundance of infected Paramecium being generally lower than that of uninfected Paramecium. However, the methods employed in our study do not entirely rule out the possibility of infection reducing the nutritional quality of Paramecium. Butzel & Bolten (1968) demonstrated that there is a link between prey nutritive status and Didinium population dynamics in a study that involved D. nasutum and a species of Paramecium in the aurelia complex. They found that Didinium exhibits decreased fission rates, abnormal cell formation and inability to encyst when fed Paramecium aurelia that have been progressively starved or malnourished. Future studies should explore the likelihood of Holospora-infected Paramecium being less nutritious to Didinium than uninfected Paramecium and of this being the basis of Holospora’s effects on Didinium population dynamics.

There were signs of synergism between the effects of parasitism and predation during the decline phase of the Didinium. In particular, the infected Paramecium populations recovered and achieved the same if not higher abundance than their uninfected counterparts. This suggests some kind of predator-related buffering or overcompensation for parasitic effects on the part of Paramecium. A similar effect has been observed in another study involving Paramecium and Holospora, wherein a certain type of stochastic environmental fluctuation allowed infected populations to maintain the same density as uninfected populations (Duncan et al. 2013). This phenomenon may be generalizable and therefore warrants further investigation.

Image analysis revealed significant effects of infection on Paramecium aspect ratio and swimming speed. Cell length decreased with increasing parasite load, meaning that heavily infected Paramecium cells become smaller and fatter. Theoretically, this could increase vulnerability to predation by Didinium, given that Didinium feeds more readily on smaller cells (Hewett 1980). On the other hand, we found that infection also tended to increase swimming speed, at least in covertly infected individuals with more moderate parasite loads. This may counter the disadvantage of being smaller, as faster prey have better chances of escaping encounters with predators like Didinium (Knoll, Haacke-Bell & Plattner 1991; Hamel et al. 2011). This enhanced activity is intriguing in that it contrasts with previous findings of reduced short-distance dispersal of infected Paramecium (Fellous et al. 2011). Moreover, the modified aspect ratio it is associated with should make the energetic cost of locomotion higher for infected Paramecium (Roberts 1981), which (in combination with infected Paramecium having higher rates of encounter with Serratia) could explain Paramecium’s enhanced negative effect on Serratia abundance in the presence of Holospora (discussed further below).

In a follow-up experiment, we evaluated the potential net effect of these parasite-induced modifications on predation risk. To this end, the feeding rate of individual Didinium cells facing 10 infected or uninfected Paramecium cells was measured over the course of several hours (further details provided in Fig. S3 caption, Supporting information). This experiment revealed no significant effects of infection status on Didinium feeding rate (Fig. 4).
S3, Supporting information), suggesting that the above trait modifications either played no role in terms of predation risk or cancelled each other out. Future studies might re-evaluate this conclusion based on results of a rigorous functional response experiment that systematically varies the level of Paramecium abundance to which Didinium is exposed.

The modelling results were in line with the above observations. We found that Holospora’s effects on Didinium abundance and on Didinium and Paramecium’s predator–prey dynamics were more likely due to density-mediated indirect effects than to a combination of density- and trait-mediated indirect effects. Altogether, reducing Paramecium’s growth rate and carrying capacity was sufficient to produce a close fit of the model to the dynamics of Didinium and Holospora-infected Paramecium, implying that, under these circumstances, Holospora’s density-mediated indirect effects were the most important.

**serratia dynamics**

While Serratia abundance was indeed highest in the complete absence of Paramecium, it was lower in the presence of infected Paramecium than in the presence of uninfected Paramecium. This suggests that infection with Holospora may increase Paramecium’s per capita feeding rate, despite Holospora’s generally negative effect on Paramecium fitness. If so, this may represent a compensatory response to Holospora’s depletion of energy/nutrients, but may also reflect an adaptive parasite strategy. Holospora may, for example, actively induce feeding in Paramecium to obtain more resources for its own reproduction (Lefèvre et al. 2008).

Similar to what it did in the case of Paramecium abundance, the presence of the predator also reduced the negative effect of infection on Serratia abundance. Indeed, a week after introduction of the Didinium, Serratia abundances in infected Paramecium populations had caught up with those in uninfected populations, which was not the case in predator-free populations. The reduction in abundance due to Didinium may have had greater weight in the case of the infected Paramecium populations due to Holospora’s enhancement of Paramecium’s reduction of Serratia.

**Implications for parasite dynamics**

One important finding regarding parasite (Holospora) dynamics is the buffering effect of predation. Predation by Didinium ultimately had a net beneficial effect on infected Paramecium populations, allowing them to recover from critically low abundances reached towards the end of the experiment. The present experiment did not address the consequences for parasite (horizontal) transmission and epidemic spread. However, it is clear that maintaining relatively higher population density may further increase the force of infection, as the number of infected hosts is directly related to the frequency of new infections. Moreover, a second follow-up experiment provided evidence for a direct impact of predation on Holospora transmission. Twenty-four hours after the introduction of 10 Didinium to high-density infected populations, we observed a significant increase in the concentration of free infectious forms of Holospora in the medium which remained in effect 48 h later ($F = 10.32$, $P < 0.01$; Fig. S4, Supporting information; further details provided in Fig. S4 caption, Supporting information). A possible explanation is that these parasite transmission stages are released while Didinium devours and digests infected Paramecium. This may directly enhance the chance of transmission in a population under predator attack. Similar results have been reported in systems comprising the predator Chaoborus and its prey Daphnia (Cáceres, Knight & Hall 2009; Duffy et al. 2011).

**Perspectives**

Parasite effects in multitrophic communities are poorly understood and sometimes greatly underappreciated (Morand & Gonzalez 1997; Lafferty et al. 2008; Poulin 2010). Our study illustrates how the addition of a single additional antagonist (here Didinium) can have complex, and partly unexpected, demographic feedbacks on host-parasite interactions and vice versa. We also found that such effects can be mirrored at lower levels of a food chain. The broader significance of these results is that they point to possible complications for pest management in agriculture and conservation, when additional players in the natural community are certain to come into play. For example, the use of certain parasites as biological control agents may be inadvisable, if another species (here Didinium) interacting with the target obstructs the expected effects due to the target’s behavioural and physiological responses.

Here, we described the impact of a single episode of proliferation and prey reduction by a predator which subsequently went extinct. Building on this simple framework, future work may address more complex scenarios, such as the spread and maintenance of an epidemic or the occurrence of co-evolution, when all interacting species are maintained over longer time-scales.

**Acknowledgements**

Support for this work came from the University of Zurich Institute of Evolutionary Biology and Environmental Studies. We thank David Montagnes, Dennis Hansen, Yves Choffat, Dan Flynn, Thomas Mussie, Marco Plebani, Gian Marco Palamara, Mahbub Mohiabdi and the members of the Princeton Disease Modeling Group for helpful comments and technical assistance.

**Data accessibility**

Data available from the Dryad Digital Repository: http://dx.doi.org/10.5061/dryad.3kb72 (Banerji et al. 2014).
Supporting Information

Additional Supporting Information may be found in the online version of this article.