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A novel marine nitrite-oxidizing *Nitrospira* species from Dutch coastal North Sea water

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INTRODUCTION

The ocean is the largest reservoir of fixed nitrogen on Earth containing about five times more fixed nitrogen than terrestrial systems (Gruber, 2008) which renders marine systems of major importance to global nitrogen cycling. Nitrogen, as the bioavailable forms of ammonium and nitrate, is one of the key nutrients in marine waters and may limit primary production especially in coastal systems (Downing, 1997; Wollast, 1998; Zeh and Kiendler, 2011). Most of the fixed organic nitrogen in the ocean is converted to nitrate by remineralization consisting of ammonification and nitrification (Gruber, 2008). In the two-step process of nitrification, ammonia is oxidized first to nitrite by aerobic ammonia-oxidizing microorganisms and then to nitrate by aerobic nitrite-oxidizing microorganisms. The microbial mediators of nitrification have intrigued scientists ever since the hallmark publication by Winogradsky (1890) in which not only the ability of nitrifying organisms to withdraw energy from mineral substances was reported but it was also concluded that these microorganisms assimilate carbon from carbon dioxide. At present, after 120 years of research efforts, many nitrifying microorganisms are available in culture, and therefore amenable to physiological characterization, but marine species are underrepresented. For instance, when evaluating the phylogeny of β-proteobacterial ammonia-oxidizing bacteria (AOB) (Aakra et al. 2001) examined no less than 38 isolates of which only five had a marine origin. In the review by Koops and Pomerening-Röser (2001) on the distribution and ecophysiology of nitrifying bacteria the phylogenetic relationship of 19 cultured AOB species is shown and for only five of those species a preference for a marine habitat is indicated. In marine ecosystems ammonia-oxidizing archa (AOA) species have been shown to outnumber their bacterial counterparts based on direct cell counts and gene (16S rRNA and amoA) copy numbers (Francis et al., 2003; Wuchter et al., 2006; Mincer et al., 2007). Currently, however, the only two cultured AOA species with a marine or estuarine origin are *Nitrospumilus*

Marine microorganisms are important for the global nitrogen cycle, but marine nitrifiers, especially aerobic nitrite oxidizers, remain largely unexplored. To increase the number of cultured representatives of marine nitrite-oxidizing bacteria (NOB), a bioreactor cultivation approach was adopted to first enrich nitrites and ultimately nitrite oxidizers from Dutch coastal North Sea water. With solely ammonia as the substrate an active nitrifying community consisting of novel marine *Nitrosomonas* aerobic ammonia oxidizers (ammonia-oxidizing bacteria) and *Nitrospira* and *Nitrosopumilus* NOB was obtained which converted a maximum of 2 mmol of ammonia per liter per day. Switching the feed of the culture to nitrite as a sole substrate resulted in a *Nitrosopumilus* NOB dominated community (approximately 80% of the total microbial community based on fluorescence in situ hybridization and metagenomic data) converting a maximum of 3 mmol of nitrite per liter per day. Phylogenetic analyses based on the 16S rRNA gene indicated that the *Nitrosopumilus* enriched from the North Sea is a novel Nitrospira species with *Nitrospira marina* as the next taxonomically described relative (94% 16S rRNA sequence identity). Transmission electron microscopy analysis revealed a cell plan typical for *Nitrospira* species. The cytoplasm contained electron light particles that might represent glycogen storage. A large periplasmic space was present which was filled with electron dense particles. *Nitrospira*-targeted polymerase chain reaction analyses demonstrated the presence of the enriched *Nitrospira* species in a time series of North Sea genomic DNA samples. The availability of this new *Nitrospira* species enrichment culture facilitates further in-depth studies such as determination of physiological constraints, and comparison to other NOB species.

Keywords: marine nitrification, enrichment, *Nitrospumilus*, *Nitrospira*, fluorescence in situ hybridization, transmission electron microscopy, 16S rRNA

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Nitrospina

Forty-five responders were more than 65 years old.

45

The elucidation of the ecophysiology of marine nitrite oxidizers is in part complicated by the difficulties in combining molecular data (e.g., presence and abundance of particular genes or species) with cultivation-derived parameters (e.g., proof of physiological capabilities, affinities, growth rates, salt tolerance).

- Nitrite-oxidizing Nitrospina species were visualized with transmission electron microscopy (TEM) and, using a newly designed primer pair targeting Nitrospina species, its presence was detected in a time series (Wuchter et al., 2006; Pitcher et al., 2011) of high molecular weight DNA isolated from the same coastal sampling site.

MATERIALS AND METHODS

ICOLUIM DESCRIPTION AND REACTOR SET-UP

Water representative of Dutch coastal North Sea water (Pitcher et al., 2011) was collected in February 2007 at high tide at the jetty of the Royal Netherlands Institute for Sea Research situated on the island Texel (53°09′25″N, 4°47′27″E). An aliquot of 48 L North Sea water was filtered using a HF80S polysulfone capillary artificial Kidney/Hemofilter (Fresenius Medical Care Nederland BV, Nieuwkoop, the Netherlands). This resulted in 2 L of 24-fold concentrated biomass suspension and a cleared solution (filtrate) devoid of particles. The collected biomass suspension was incubated in a sterile glass and stainless steel reactor (adaptive, 2 L working volume). Heat-sterilized (20 min, 120°C, 15 kPa) filtrate supplemented with ammonium or nitrite (from 1 M sterile stocks of NH4Cl and NaN3) was used as a medium. Oxygen and pH were monitored online using Applikon (Applikon Biotechnology BV, Schiedam, the Netherlands) sensors. Nitrite concentrations were determined offline in liquid samples withdrawn daily from the reactor using Mercckquant test strips (Merck BV, Schiphol-Rijk, the Netherlands) and ammonium as well as nitrite concentration weekly using colorimetric methods (see below). The reactor was kept at a pH 7.8 with solutions of sterile 1 M NaHCO3 and 0.6 M HCl operated at room temperature 22 ± 2°C, stirred at 150 rpm, and supplied with an air flow of 80 ml/min.

ENRICHMENT WITH AMMONIA AS THE SUBSTRATE

The culture was amended with 200 μM NH4Cl and incubated for 19 days as a batch and then for another 2 days after addition of 400 μM NH4Cl. To avoid nitrite toxicity, the reactor system was thereafter switched to a continuous mode of operation using medium containing 750 μM NH4+ at a dilution rate of 0.25 day−1·2. When nitrite disappeared from the culture, indicating activity of nitrite oxidizers, the ammonium concentration was subsequently increased to 1.5 mM after 3 months, and further to 2, 3, and finally 10 mM after 4, 4.5, and 5.5 months, respectively. The reactor was switched to nitrite as the sole substrate after 7 months.

ENRICHMENT WITH NITRITE AS THE SUBSTRATE

To stimulate growth of the nitrite oxidizers, a batch mode of operation was adopted by adding 750 μM NaN3 provided as the substrate. Whenever nitrite was depleted, it was restored to 750 μM. In this manner, a total of 43 mmol of nitrite were supplied in the first 4 months of operation with nitrite as the sole substrate. Wall growth was suspended and the biomass diluted fourfold by replacement of reactor content with medium respectively, 1 week and 1 month after the switch to nitrite. After 1 month, a fed-batch mode of operation was adopted by adding medium containing 30 mM NaN3 at a flow rate starting at 40 ml per day. The pump rate
respectively, by increasing the flow rate from 40 to 100 ml per day. To retain biomass, the reactor content was allowed to settle once for a week after which clarified liquid was removed to maintain a maximum reactor volume of 2 L. Removal of wall growth and fourfold dilution of the biomass were performed as described above after 2 and 4 months. The influent nitrite concentration was raised from 10 to 20, 40, 60, 80 mM and finally 100 mM after 6, 9, 10, 10.5, and 11 months, respectively, by increasing the flow rate from 40 to 100 ml per day in 10 ml steps keeping NO₂⁻ levels below 2 mg/L. In order to prevent suboptimal nitrite oxidizer growth rates due to carbon limitation, the gas flow of 80 ml/min of air was supplemented to 10 ml/min Argon/CO₂ (95%/5%) from 7.5 months onward. It has furthermore been reported that iron as well as phosphate may become limiting compounds for growth (van de Vossenberg et al., 2008) when a medium containing only natural sea salts is used to enrich marine microorganisms. Therefore, the influent was supplemented with 0.261 ml/L 1 M KH₂PO₄ and 0.45 ml/L of a 5 g/L FeSO₄·7H₂O solution from month eight onward. The reactor was operated for 12 months with nitrite as the sole substrate. During the last month the biomass was no longer allowed to settle prior to medium replenishment resulting in an actual dilution rate of 0.05 day⁻¹.

CHEMICAL ANALYSES

To estimate nitrite concentrations liquid samples were measured directly using Merckquant® teststrips (range for nitrate 10–500 mg/L, nitrite 2–80 mg/L, Merck BV, Schiphol-Rijk, the Netherlands). At least once a week, 0.5 ml aliquots were centrifuged (5 min 10,000 × g) and the resulting supernatants used for more elaborate colorimetric analyses to monitor residual ammonium and nitrite concentrations. To measure nitrite, a colorimetric method adapted from Griess-Romijn-van Eck (1966) was used. A mixture of 50 μl with 0.5 ml of reagent A (10 g of sulfanilic acid in 1 L 1 M HCl) and 0.5 ml reagent B (1 g 1- N-naphthylethenediamine dihydrochloride in 1 L distilled water) was incubated for 10 min at room temperature, and measurements performed at 540 nm. Ammonium concentrations were determined using ortho-phthalaldehyde (OPA) reagent (Both, 1971; Taylor et al., 1974). The OPA reagent consisted of 8.54 g of OPA dissolved in 10 ml of absolute ethanol, with 50 μl of β-mercaptoethanol, and filled to 100 ml with sodium phosphate buffer (0.3 M pH 7.3). To measure ammonium concentrations between 0.25 and 5 mM 50 ml sample was mixed with 800 μl OPA reagent, incubated (20 min, room temperature, in the dark), and the extinction measured (420 nm). To measure in the range of 5–300 μM, 100 μl of sample was mixed with 2 ml OPA reagent containing only 0.054 g/100 ml OPA, incubated (20 min, room temperature, in the dark) and measured with a fluorescence spectrophotometer (excitation 411 nm, emission 482 nm, slit size 5 nm, 600 V).

FLUORESCENCE IN SITU HYBRIDIZATION

Biomass was harvested from 20 ml reactor material by centrifugation (10 min 10,000 × g) and fixed for FISH analyses by addition of 4% w/v paraformaldehyde, incubating on ice (2 h), centrifuging (15 min 10,000 × g) and washing the resulting pellet with phosphate buffered saline (PBS, pH 7.2) and finally adding PBS and 100% EtOH (1:1) to reach a volume of 10% of the original sample. Fixed material was stored at −20°C until analysis. FISH analyses on fixed biomass from the start, after 1 and 6 months of the nitrite-fed period were performed as described by Amann et al. (1990), using 10 μl fixed material per hybridization. Vectashield (Vector Laboratories, Inc., Burlingame, CA, USA) mounting medium with DAPI (4,6-diamidino-2-phenylindole) was used to enhance the fluorescent signal and stain all DNA. Specifications and details of probes used in this study are presented in Table 1. Probes were purchased as Cy3-3, Cy5-5, and Cy5-(6-carboxyfluorescein)-N-hydroxysuccinimide-ester (FLUOS) labeled derivatives from Thermohyibaid (Ulm, Germany). To visualize Nitrosomonas AOB and Nitrospira NOB simultaneously, probes NEU 655 (FLUOS) and NTSPA 712 (Cy5) were used together with their respective competitors (competitor probes consisted of unlabeled oligonucleotides) in single hybridizations at a formamide concentration of 35%. To detect Nitrospina sp. NOB, hybridizations were performed at 29% formamide concentration with probe NTSPN693. To stain all bacteria, a mixture of probes EUB338, EUB338 II, and EUB338 III was used for all hybridizations. Microscopic inspections were performed at a 1000-fold magnification. For image acquisition a Zeiss Axioplan 2 epifluorescence microscope (Zeiss, Jena, Germany) was used with the standard software package (version 3.1). Abundance estimates of cells hybridizing with a particular probe were based on visual inspection of three randomly taken FISH microscopy pictures per hybridization.

EXTRACTION HIGH MOLECULAR WEIGHT DNA

Biomass was harvested from 20 ml reactor content by centrifugation (20 min, 2400 × g) after 3.5 months with ammonia, and after 6 months with nitrite as the sole substrate, respectively. Biomass was also harvested from 50 ml reactor content after 12 months with nitrite as the substrate. High molecular weight DNA was extracted using a cetyltrimethyl-ammoniumbromide (CTAB) and sodium dodecyl sulfate (SDS)-lysozyme-based method adapted from Zhou et al. (1996). Biomass was suspended and incubated for 30 min at 37°C in a mixture of 675 μl CTAB extraction buffer (1g/100 ml CTAB, 100 mM Tris, 100 mM EDTA, 100 mM sodium phosphate, 1.5 M NaCl, pH 8), 50 μl lysozyme (10 mg/ml, 66200 U/mg) and 30 μl Rnase A (10 mg/ml, ≥3000 U/mg). After addition of 50 μl of proteinase K (10 mg/ml, 20 U/mg) and incubation for 30 min at 37°C, the mixture was supplemented with 150 μl 10% SDS and incubated at 65°C for 2 h. DNA was recovered by phenol/chloroform extraction and isopropanol precipitation after which it was suspended in 40 μl ultrapure water (MilliQ, Millipore SA, Molsheim, France) and stored at 4°C until use.

PCR REACTIONS, CLONING, SEQUENCING, AND SEQUENCE ANALYSES

Polymerase chain reaction reactions (30 cycles, followed by a final extension for 10 min at 72°C) were performed in a T gradient PCR apparatus (Whatman Biometra, Göttingen, Germany) using
Table 1 | Oligonucleotide specifications.

| Name     | Used for     | Sequence (5′→3′)          | Position* | Target                               | Reference               |
|----------|--------------|---------------------------|-----------|--------------------------------------|-------------------------|
| EU8338 FISH | OCTGCCCCCCCTGAGGAGT  | 338                      | Most Bacteria | Amann et al. (1990)                 |
| EU8338 II FISH | GCAGCCACCCCTGAGGAGT  | 338                      | Most Planctomycetales | Daims et al. (1999)             |
| EU8338 III FISH | GCCGCCACCCCTGAGGAGT | 338                      | Most Verrucomicrobiales | Daims et al. (1999)             |
| NEU653 FISH | CCCCCTGCTGCACTCTAG  | 653                      | Most halophilic and halotolerant | Wagner et al. (1995)       |
| Competitor NEU653 FISH | TTCACCCCCCTGCTGGAG  | 653                      | Nitrosomonas spp |                     |
| NTSFA712 FISH | CCCCCTGCTGCACTCTAG  | 712                      | Most members of the phylum | Daims et al. (2001)             |
| NTSFN683 FISH | TCCCCAATATACGCACAGTT | 683                      | Nitrospina gracilis | Juretschko (2000)                 |
| 616F PCR | AGAGTTTGATYMTGGCTCAG | 8                       | Bacteria | Juretschko et al. (1998)             |
| 630R PCR | CAKAAAGGAGGTGATCC | 1529                     | Bacteria | Juretschko et al. (1998)             |
| NTSFA1158R PCR | CCCGTCTCCCACCGGTGATCC | 1158                    | Most Nitrospira | Masneri et al. (2006)             |
| NSE87F PCR | AGTGGCGAACGGGTGAGGAATA | 87                      | Most Nitrospira | This study                      |
| NSE1124R PCR | TCTCTGCGAGGTCGCCGGCAGTGA | 1124                  | Most Nitrospira | This study                      |
| 810F Sequencing | GTGCCAGCAGCCGGGGGT | 479                      | Most bacteria | Ustunova et al. (2001)             |
| M13F Sequencing | GTAAAACGACGGCCAG | Region flanking cloning site | pGEMT easy vector | –                             |
| M13R Sequencing | CAGGAAACAGCTATGA | Region flanking cloning site | pGEMT easy vector | –                             |

*E. coli numbering.

GoTaq® Green Master Mix (Promega Benelux BV, Leiden, the Netherlands). A PCR cycle consisted of, 1 min at 93°C, 1 min at annealing temperature (T_a) and 1.5 min at 72°C. For each 25 μl volume PCR reaction, 1 μl of 10-fold diluted high molecular weight DNA was used as the template. Resultant products were cloned using the pGEM T-easy vector cloning kit (Promega Benelux BV, Leiden, the Netherlands). Plasmid DNA was extracted using the GeneJet Plasmid Miniprep Kit (Fermentas GMBH, St. Leon-Rot, Germany). Clones were checked by restriction analysis of plasmid DNA (EcoR1, Fermentas GMBH, St. Leon-Rot, Germany). Sequencing (Sanger method) was performed at the division DNA diagnostics of the Human Genetics department of the University Medical Centre Nijmegen St Radboud. The ContigExpress program of the Vector NTI Suite 7.0 software package (InforMax) was used to assemble full-length clone sequences. Cloned 16S rRNA gene sequences were compared with their closest relatives in the GenBank database by BLASTN searches1. Phylogenetic and molecular evolutionary analyses were conducted using MEGA version 4 (Tamura et al., 2007). The Ribosomal Database Project (RDP) Classifier tool2 (RDP Naïve Bayesian rRNA classifier version 2.5, May 2012, RDP 16S rRNA training set 9) was used to evaluate the taxonomic position of sequences (Wang et al., 2007). Pairwise analyses to determine sequence identities were performed using the internet tool from the Georgetown University Medical Center3.

1 http://www.ncbi.nlm.nih.gov/BLAST
2 http://rdp.cme.msu.edu/classifier/classifier.jsp
3 http://pir.georgetown.edu/pirwww/search/pairwise.shtml

16S rRNA gene sequences are available from GenBank under the following accession numbers: KC706457-706479. For sequences sharing at least 99% sequence identity (see “Results”) a representative sequence has been submitted. For the North Sea nitrifier enrichment cultures clone Cb9 (KC706457) represents the Nitrosomonas sp., Cb12 (KC706458) the Nitrospina sp. and Cb 18 (KC706459) “Candidatus Nitrospira salza.” For the North Sea time
series sequences clone P3_4 represents the cluster of 15 *Nitrospira* sequences.

**METAGENOME SEQUENCING AND RECONSTRUCTION OF THE 16S rRNA GENE SEQUENCE OF THE DOMINANT NOB**

DNA extraction performed on 50 ml reactor biomass at the end of the incubation with nitrite as the sole substrate yielded 15 μg DNA based on spectrophotometric estimation using NanoDrop technology (Thermoscientific, USA). Eight microgram was subsequently used for pyrosequencing using the Roche 454 GS FLX Titanium sequencer (Roche, Switzerland) at the Department of Human Genetics Nijmegen at the Center for Molecular Life Sciences, Institute for Genetic and Metabolic Disease of the Radboud University Nijmegen Medical Center. To estimate *Nitrospira* abundance, all generated reads were mapped using CILC Bio Genomics Workbench (version 5.5.1) to a custom 16S rRNA gene sequence database which consisted of all unaligned sequences in release 18.29 (2,328,464 sequences) of the RDP 16S rRNA database (Cole et al., 2009), from which all sequences containing “uncultured” or “unidentified” in the description were removed. The resultant database (available upon request) contained 339,774 16S rRNA gene sequences. Through mapping (cutoff 90% identity over 90% of the read length) of the sequence reads on this database *Nitrospira* sp. 16S rRNA gene sequence reads were identified. The 16S rRNA gene sequence of the dominant *Nitrospira NOB* was reconstructed through a de novo assembly of those reads using the CILC genomics workbench.

**TRANSMISSION ELECTRON MICROSCOPY**

To investigate the cell morphology of the enriched NOB using TEM, biomass harvested from 100 ml reactor content (by centrifugation for 20 min., 2400 × g) after 8 months of operation with nitrite as the sole substrate was taken. Cryofixation was performed by high pressure freezing and was followed by freeze-substitution in acetone containing 2% osmium tetroxide, 0.2% uranyl acetate, and 1% water, embedding in Epon resin and sectioning using an ultramicrotome for TEM analysis. Sample preparation was performed as described previously by van Niftrik et al. (2008).

**DETECTION OF THE ENRICHED *NITROSPIRA NOB* IN COASTAL NORTH SEA WATER**

To verify that the enriched *Nitrospira* originated from the North Sea and was not a contaminant from our laboratory, high molecular weight DNA samples from a North Sea time series (Wuchter et al., 2006; Pitcher et al., 2011) were screened for the presence of *Nitrospira* by PCR analyses. A new primer pair perfectly matching the full-length 16S RNA gene sequence of the enriched *Nitrospira*, was designed (primers NSE87F and NSE1124R). These primers were tested in PCR reactions (Tc: 80°C) using the DNA extracted from the enrichment after 6 months of operation with nitrite as the substrate as a template. In addition, test reactions were performed using high molecular weight DNA extracted from *Nitrospira defluvii* and *Nitrospira marina* cells and plasmid DNAs containing partial (1073 nt) 16S rRNA gene sequences from *Nitrospira* sublineages 1 (*Nitrospira defluvii*-like, 2 plasmids), II (*N. marina*-like, 2 plasmids), and IV (*N. marina*-like, 2 plasmids). To screen the North Sea time series, six pools were prepared from partial aliquots (3 μl of each sample) of the high molecular weight DNA samples from the time series (see Table 2 in the Results). Prior to amplification, 3 μl of each pool was purified by excision of DNA-containing bands from low-melting point agarose gel (Electron wide range, low melting agarose, VWR BiDH Prolabo) after electrophoresis to remove substances possibly interfering with PCR amplification. PCR products were cloned, and for 25 clones (3–5 clones picked per pool) plasmid DNA was extracted and sequencing performed with primers M13F and M13R.

**RESULTS**

**NORTH SEA-NOB AND NOB ENRICHMENT WITH AMMONIA AS THE SUBSTRATE**

After a lag phase of 10 days, microbial ammonia oxidizers became active in the enrichment with ammonia as the substrate. Within 9 days, 500 μM NO₂⁻ was produced from 500 μM NH₄⁺. A second aliquot of 400 μM ammonium induced further nitrite accumulation at a higher rate (400 μM within 2 days) which indicates growth of ammonia oxidizers. After adopting a continuous mode of operation (D = 0.25 per day) to avoid nitrite toxicity, all supplied ammonium (750 μM) was converted to nitrite in a 1:1 ratio up to 3 months of operation. Hereafter, the nitrite concentration dropped to zero within a 14 day period indicating a rapid increase in nitrite oxidizer activity. During the subsequent stepwise increase of the influent NH₄Cl concentration to 3 mM (after 4.5 months of operation), NH₄Cl as well as NO₂⁻ reactor concentrations remained zero indicating complete consumption of both nitrogen species and therefore an active co-culture of ammonia and nitrite oxidizers.

| Sample pool | Time series samples |
|-------------|---------------------|
| P1          | Oct-Nov-Dec 2003:   |
|             | 23-Oct, 27-Oct, 03-Nov, 13-Nov, 17-Nov, 25-Nov, 01-Dec, 12-Dec, 29-Dec |
| P2          | Jan-Feb-Mar 2004:   |
|             | 02-Jan, 14-Jan, 30-Jan, 09-Feb, 15-Feb, 16-Feb, 23-Feb, 01-Mar, 08-Mar, 15-Mar, 22-Mar, 29-Mar |
| P3          | Apr-May 2004:       |
|             | 13-Apr, 21-Apr, 18-May |
| P4          | Aug-Sep 2004:       |
|             | 02-Aug, 09-Aug, 19-Aug, 23-Aug, 16-Sep, 27-Sep |
| P5          | Oct-Nov-Dec 2004:   |
|             | 19-Oct, 08-Nov, 15-Nov, 29-Nov, 06-Dec, 16-Dec, 23-Dec |
| P6          | Jan 2005:           |
|             | 14-Jan, 25-Jan, 31-Jan |

*Pools consisted of a mixture of 3 μl of each time series sample.*
converting 0.75 mmol of nitrogen per liter per day. The raise to 10 mM NH₄Cl (after 5.5 months of operation) resulted in an ammonium and nitrite accumulation to final concentrations (at 6 months of operation) of 1.8 mM and 100 μM, respectively. During the last half month of operation therefore approximately 2 mmol of nitrogen were consumed per liter per day.

The PCR performed with general bacterial primers on DNA extracted from biomass after 5.5 months of operation with ammonium as the substrate yielded correct-sized inserts (1500 nt). The 20 clones picked for plasmid isolation yielded 20 partial (695–845 nt) 16S rRNA gene sequences of which six contained recognizable nitrifier 16S rRNA sequences based on BlastN searches of the National Center for Biotechnology Information (NCBI) database and taxonomic assignment using the Classifier tool of the RDP. Taxonomic assignment of the remaining sequences resulted in five sequences assigned to uncultured bacteria, two to unclassified Planctomycetes, three to the genus Physiophora within the Planctomycetes, two to unclassified α-proteobacteria, one to the genus Phaeobacter within the α-proteobacteria, and one to unclassified Anaerolineae within the Chloroflexi. Nearly full-length 16S rRNA gene sequences generated from the clones containing a recognizable nitrifier sequence resulted in three Nitrosomonas (AOB) sequences (clones Cb9, 10, and 15; >99% shared sequence identity), two Nitrospina (NOB) sequences (clones Cb12 and 16; 99.5% shared sequence identity), and one Nitrospira (NOB) sequence (clone Cb18). The phylogenetic position of the putative Nitrosononas-like AOB is shown in **Figure 1** which illustrates that the sequence from the enrichment culture is related to Nitrosononas marina but does not cluster closely to any cultivated Nitrosononas species. The closest match in the NCBI database (96% sequence identity) with a cultivated species was the 16S rRNA gene sequence of Nitrosononas sp. NM31 (Purkhold et al., 2000). This implies that the enriched AOB may represent a previously uncultured Nitrosononas species. The closest match in the NCBI database (99% sequence identity) was to an unpublished marine clone sequence (JF514271, clone LXE3). For another sequence (FJ628323, clone NitA0631, Schmidova et al., 2009) sharing 99% sequence identity to the sequence of the enriched North Sea AOB it was known that this sequence was retrieved from brackish water from the anoxic fjord Nitinat Lake, which is an environment with an ammonium concentration between 20 and 200 μM. The Nitrosononas (NOB) sequences shared only 92% sequence identity to the 16S rRNA gene sequence of the cultivated species Nitrospina gracilis strain 3/211 (FR865038). The **FIGURE 1** | 16S rRNA gene sequence based phylogenetic tree showing the position of the enriched North Sea AOB (bold) within the betaproteobacterial AOB. This unrooted neighbor-joining bootstrap tree was inferred using the neighbor-joining algorithm. Total of 1365 nucleotides were considered in the alignment. Bootstrap values are shown at the internal nodes. The scale bar is in the unit of the number of base substitutions per site.
next taxonomically described match for the Nitrospira NOB clone sequence was 94% sequence identity with the 16S rRNA sequence of Nitrospira marina strain Nb-295 (X82559, Ehrich et al., 1995).

The FISH analyses revealed that the biomass at the end of the ammonium-fed enrichment (Figure 2A) consisted mainly (approximately 80% of the total population) of bacteria hybridizing with probe NEU 653. This indicates dominance of halotolerant/halophilic Nitrosomonas-like AOB. In addition, around 10% of the bacterial population hybridized with probe NTSPA712 indicating the presence of Nitrospira-like NOB. No hybridization with probe NTSPN693 was observed which suggests that Nitrospira NOB were a minority within the nitrifier community.

**NORTH SEA NOB ENRICHMENT WITH NITRITE AS THE SUBSTRATE**

Within the first month of the nitrite-fed period (manual supply of Nitrospira marina sequence was 94% sequence identity with the 16S rRNA sequence of the antibacterial and halophilic (approximately 80% of the total population) of bacteria hybridizing with probe NTSPA712 indicating the presence of Nitrospira-like NOB. No hybridization with probe NTSPN693 was observed which suggests that Nitrospira NOB were a minority within the nitrifier community.

**METAGENOME SEQUENCING AND RECONSTRUCTION OF THE 16S rRNA GENE SEQUENCE OF THE DOMINANT NOB**

The 343 sequencing run on DNA extracted from biomass at the end of the incubation with nitrite as the sole substrate (after 12 months) generated, after quality trimming, 1,216,565 single reads with an average length of 405 nt. The mapping of all reads to the custom 16S rRNA gene sequence database resulted in 198 mapped reads, of which 147 mapped to Nitrospira sp. 16S rRNA gene sequences. This implies an abundance of Nitrospira sp. 16S rRNA genes within the total population of 74% which is in agreement with the 80% abundance estimated from the FISH analysis after 6 months of operation with nitrite as the substrate.

The 16S rRNA gene sequence of the dominant Nitrospira NOB reconstructed from the 147 Nitrospira sp. reads exhibited 99.9% sequence identity to the earlier obtained (clone Ch18 and AC6) sequences resulting from PCR analysis, suggesting that the same species persisted as the dominant NOB within the reactor.

**TEM ANALYSIS OF THE ENRICHED NORTH SEA Nitrospira sp.**

The biomass was mainly situated in small aggregates in the culture. This was reflected in the electron microscopy pictures generated with the TEM analysis of the biomass, harvested from the enrichment after 8 months. These showed dense clumps of cells seemingly embedded in extracellular material (Figure 4A). Some typical morphological features of a representative cell are pointed out in Figure 4B. Most striking is the large periplasmic space containing many electron dense particles. In addition, large electron light particles are visible in the cytoplasm.

**Figure 2** | Fluorescence microscopy pictures of the abundance of Nitrospira marina strain Nb-295 (X82559, Ehrich et al., 1995). In green: cells hybridizing with probe NEU 653 (targeting most halophilic and halotolerant Nitrospira marina spp.); In red: cells hybridizing with probe NTSPA712 (targeting most members of the phylum Nitrospirae). (A) Biomass after 1 month; (B) after 3 months; (C) after 6 months of operation with nitrite as the only substrate.
DETECTION OF THE ENRICHED Nitrospira NOB IN COASTAL NORTH SEA WATER

Nitrospira-targeted primer pair 616F/intSP11388R did not yield significant amplicons (data not shown) for DNA samples from the North Sea time series, and therefore primers NSE87F and NSE1124R were developed. This primer pair, designed to specifically target the enriched "Candidatus Nitrospira salvia", yielded correct-sized (1073 nt) amplicons with all tested templates (high molecular weight DNA from the North Sea enrichment, Nitrospira defluvii and N. moscoviensis as well as plasmid DNA from respectively sublineage I (Nitrospira defluvii-like), II (N. moscoviensis-like), and IV (N. marina-like). This demonstrates that this primer pair functions well for all tested Nitrospira species and does not specifically target the enriched North Sea species.

Screening of the six separate pools (see Table 2) from the North Sea time series of high molecular weight DNA samples with this primer pair resulted in 25 16S rRNA gene clone sequences of which 22 contained a Nitrospira sequence. Three (clones P3_4, P3_5 and P4_29) clones exhibited a sequence identity of...
The results from 16S rRNA gene-fiers proved successful. Within 6 months a marine assemblage was retrieved from marine sediment (Gillan and Perret, 2007) and shared only 91–92% sequence identity with the 16S rRNA gene sequence of “Candidatus Nitrospira salina”.

Surprisingly, North Sea time series clone P3_7 contained an insert most resembling the 16S rDNA gene sequence of Nitrospira defluviit (99.5% sequence identity to sequence NC_014355, Lücker et al., 2010) and clone P1_15 an insert resembling the 16S rRNA gene sequence of Nitrospira mucoviscida (97% sequence identity to NR_029287, Ehrich et al., 1995), which are Nitrospira species associated with freshwater environments.

**DISCUSSION**

**BIOREACTOR CULTIVATION OF MARINE NORTH SEA NITRIFIERS**

The bio礁actor approach adopted to enrich North Sea nitrifiers proved successful. Within 6 months a marine assemblage of AOB and NOB was obtained by means of cultivation with ammonia as the substrate. The results from 16S rDNA gene-targeted PCR followed by cloning and sequencing indicated the presence of putative Nitrosomonas-like AOB (5/20 clones), Nitrospira-like NOB (2/20 clones), representing novel species. The FISH analysis demonstrated the abundance of the Nitrosomonas-like AOB (98% of the total bacterial population) and Nitrospira-like NOB (96%) of the total bacterial population, but failed to detect Nitrospira cells. Based on these results Nitrosomonas AOB and Nitrospira NOB are assumed responsible for the observed conversion of 2 mmol of ammonium per liter per day. The 16S rDNA gene reads mapping approach and subsequent 16S rDNA gene sequence analysis corroborated (74% of all 16S rDNA gene reads were affiliated with Nitrosomonas (AOB), and Nitrospira species (NOB) in situ water and its contribution to nitrate formation between 20 and 200 μM/month was reported. Maximum ammonium concentrations in the coastal North Sea water from which the enrichment is derived have, however, been reported to range from 10 to 13 μM during the winter months (Pitcher et al., 2011) which implies aerobic ammonia oxidation will likely be catalyzed by microorganisms with even higher affinities for ammonia. Moreover, AOB have been shown to be outnumbered by NOB (based on 16S rDNA and amoA gene copy numbers) in time series of Dutch coastal North Sea water (Vacht et al., 2006; Pitcher et al., 2011). The enriched North Sea Nitrospira species AOB therefore might exhibit a low abundance in coastal North Sea water because the system was started and refreshed with aerated North Sea water. The high (within species range) 16S rDNA gene sequence identity (>98.7%) of “Candidatus Nitrospira salina” to the 16S rDNA gene sequence retrieved from enrichment M1 corroborates this.

The 16S rDNA gene reads mapping approach and subsequent 16S rDNA gene reconstruction based on metagenomic data corroborated (74% of all 16S rDNA gene reads were affiliated with Nitrosospira) the FISH abundance data and showed that the enriched North Sea Nitrospira sp. persisted in the culture converting 3 mmol of nitrite per liter per day after 12 months of operation suggesting a quite stable microbial community. The TEM analysis data fits well with the known cell morphology of Nitrospira species. Watson et al. (1986) reported the presence of a large periplasmic space for Nitrospira marina and the presence of glycogen and polyphosphate deposits in cultures and Spieck et al. (1998) identified electron dense particles from the periplasmic space of Nitrospira mucoviscida as the nitrite-oxidizing enzyme system. In accordance with the aforementioned, the most striking attribute of the enriched North Sea Nitrospira sp. was a large periplasmic space containing putative proteins of the nitrite-oxidizing enzyme system as suggested by the presence of many electron dense particles. In addition, the cytoplasm contained putative storage material, such as glyco- gen, visible as large electron light particles. Most likely, storage of carbon was triggered by a phosphate limitation during the cultivation. This because TEM analysis was performed on biomass after 8 months of operation and to prevent carbon limitation the reactor had been supplemented with additional CO2 from month 7.5 onward. Additional phosphate to prevent phosphate limitation was provided only after 8 months which may have led to an imbalanced situation in which carbon was plentiful but phosphate was limiting.

**RELEVANCE OF THE ENRICHED NITRIFIERS IN DUTCH COASTAL NORTH SEA WATER**

It has been reported by Uecker et al. (2011) that affinity for ammonia varies among members of different lineages within the AOB genus Nitrosomonas but tends to be relatively similar within a specific lineage. The clustering of the enriched North Sea Nitrosomonas AOB 16S rDNA gene sequence with Nitrosomonas species commonly associated with low substrate environments (Figure 1) therefore suggests this species is likewise adapted to relatively low substrate concentrations. This thought is strengthened by the origin of clone sequence FJ628323 (clone Nt40631) which shares 99% sequence identity to the sequence of the enriched North Sea Nitrosomonas NOB. This clone sequence was retrieved from brackish water from an anaerobic fomed Nitrat Lake (Schmidtova et al., 2009), for which an ammonium concentration between 20 and 200 μM was reported. Maximum ammonium concentrations in the coastal North Sea water from which the enrichment is derived have, however, been reported to range from 10 to 13 μM during the winter months (Pitcher et al., 2011) which implies aerobic ammonia oxidation will likely be catalyzed by microorganisms with even higher affinities for ammonia. Moreover, AOB have been shown to be outnumbered by NOB (based on 16S rDNA and amoA gene copy numbers) in time series of Dutch coastal North Sea water (Vacht et al., 2006; Pitcher et al., 2011). The enriched North Sea Nitrospira species AOB therefore might exhibit a low abundance in coastal North Sea water and its contribution to in situ nitrification may be minor. The retrieval of clone sequences (3/20) from the North Sea time series nearly identical to the 16S rDNA gene sequence of “Candidatus Nitrospira salina” proves that this species does occur in the North Sea. The higher abundance of clone sequences (17/22) forming a separate distinct cluster suggests that another Nitrospira species may actually be more abundant and potentially contribute more to in situ nitrification. Moreover, our data suggested that a minor portion of the nitrifier community may have consisted of Nitrospira NOB after 5.5 months of enrichment with ammonium.
Nitrospira species have often been detected in marine environments (e.g., Mincer et al., 2007; Reman et al., 2010; Santoro et al., 2010; Füssel et al., 2011). Based on our present study, we cannot exclude that *Nitrospira* may be present in greater abundance or contributing to a greater extent to *in situ* nitrite oxidation. Cultivation in a bio reactor set-up offers a higher degree of control over environmental parameters (pH, T, substrate concentration, product concentration) than more traditional batch cultivation. Selection for a particular species due to the cultivation conditions, however, cannot be excluded completely. Our enrichment was performed with a maximum nitrite concentration of 750 μM (during the first month of operation). Off et al. (2010) reported nitrite tolerances for different species of *Nitrospira* ranging from a low 3.5 mM for enrichment culture Aa01 derived from a marine sponge to intermediate (6 mM) for *Nitrosopora marina* to high (15–25 mM) for freshwater *Nitrospira* species. Based on this, the present bio reactor cultivation would be expected to yield a *Nitrospira* NOB associated with low levels of nitrite. Indeed the closest relatives (based on 16S rRNA gene sequence analysis) of the enriched North Sea *Nitrospira* originated from recirculation aquaculture systems which were described as relatively low nitrite environments (10–40 μM nitrite reported by Keuper et al., 2011; nitrite below detection reported by Brown et al., 2013). Substrate concentration may have contributed to the dominance of “Candidatus *Nitrodorubilla salus*” in the final enrichment instead of *Nitrospira marina*-like or *Nitrospira*-like NOB.

Strikingly, the only pools (P3 and P4) from which “Candidatus *Nitrodorubilla salus*” sequences were derived were also the only pools consisting of samples from outside the winter months (spring and summer; Table 2). *Nitrospira defluvii* (enriched from wastewater treatment sludge) Speck et al., 2006; Maixner et al., 2008; Lücker et al., 2010) and *Nitrospira moscoviensis* (isolated from a partially corroded area of an iron pipe of a heating system, Ehrich et al., 1995) are commonly associated with freshwater environments. The detection of clone sequences most related (99.5% to *Nitrospira defluvii*, 97% identity to *Nitrospira moscoviensis*, respectively) to 16S rRNA gene sequences from these *Nitrospira* species may be caused by terrestrial input (e.g., riverine influx) at the sampling site.

**OUTLOOK**

Further research, e.g., selective inhibition experiments of AOA versus AOB activity (Yan et al., 2012), may clarify the role of the enriched *Nitrosomonas* AOB species in Dutch coastal North Sea water nitrification. Bio reactor enrichments adopting more stringent substrate levels may result in marine microbial assemblages with a totally different species composition, which would be useful to compare and contrast to the one presently described. Collection of *in situ* abundance data (e.g., by quantitative PCR analyses) for different species of NOB (e.g., “Candidatus *Nitrodorubilla salus*” versus other *Nitrospira* sp. and *Nitrospira* sp.) may help identify which NOB are of relevance to *in situ* nitrification. Seasonality in the abundance of this species may be corroborated by future reactor or laboratory enrichment experiments performed at different temperatures. Screening of a high resolution time series may aid in elucidating temporal changes in NOB community composition. The availability of the new *Nitrospira* species enrichments facilitates further in-depth studies such as determination of physiological constraints and comparison to other NOB species. Such characterizations will increase our understanding of microbial nitrogen cycling.

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in the environment. Appl. Environ. Microbiol. 50, 194–201.
Watson, S. W., Bock, E., Veloso, J. W., Waterbury, J. B., and Selkon, U. (1986). Nitrospira marina gen. nov., sp. nov. a chemolithotrophic nitrite-oxidizing bacterium. Arch. Microbiol. 144, 1–7.
Winogradsky, S. (1890). Sur les organismes de la nitrification. Comptes rendus de l’Academie des Sciences 110, 1013–1016.
Wollast, R. (1998). “Evaluation and comparison of the global carbon cycle in the coastal zone and in the open ocean,” in The Global Coastal Ocean, eds K. H. Brink and A. R. Robinson (Hoboken: John Wiley & Sons), 213–252.
Wuchter, C., Abbas, B., Cooken, M.-J.-I., Harfert, L., van Bleiswijk, J., Timmers, P., et al. (2006). Archaeal nitrification in the ocean. Proc. Natl. Acad. Sci. U.S.A. 103, 12317–12322.
Yam, J., Haaijer, S. C. M., Op den Camp, H. J. M., van Niftrik, L., Stahl, D. A., Kinneke, M., et al. (2012). Mimicking the oxygen minimum zones stimulating interaction of aerobic and anaerobic ammonia oxidizers in a laboratory-scale model system. Environ. Microbiol. 14, 3346–3358.
Zhu, J. P., and Kudela, R. M. (2011). Nitrogen cycle of the open ocean: from genes to ecosystems. Annu. Rev. Mar. Sci. 5, 197–223.
Zhou, J. Z., Bruns, M. A., and Tiedje, J. M. (1996). DNA recovery from soils of diverse composition. Appl. Environ. Microbiol. 62, 316–322.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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