| Title          | Myostatin Increases Smad2 Phosphorylation and Atrogin-1 Expression in Chick Embryonic Myotubes |
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Skeletal muscle mass is an important trait in poultry meat production. In mammals, myostatin, a negative regulator of skeletal muscle growth, activates Smad transcription factors and induces the expression of atrogin-1 by regulating the Akt/FOXO pathway. Although the amino acid sequence of chicken myostatin is known to be completely identical to its mammalian counterpart, previous studies in chicken skeletal muscles have implied that the physiological roles of chicken myostatin are different from those of mammals. Furthermore, it remains to be elucidated whether myostatin affects cellular signaling factors and atrogin-1 expression. In this study, using chick embryonic myotubes, we found that myostatin significantly increased the phosphorylation rate of Smad2 and mRNA levels of atrogin-1. No significant change was observed in the phosphorylation of Akt and FOXO1. These in vitro results suggest that the molecular mechanisms underlying myostatin-induced expression of atrogin-1 might be different between chickens and mammals.

**Key words:** atrogin-1, chick myotube, myostatin, Smad

**Introduction**

Skeletal muscle mass is an important trait in poultry meat production. Since it is controlled by a balance between protein synthesis and degradation, a number of studies have been carried out to clarify the mechanisms underlying protein metabolism in poultry skeletal muscle.

Numerous lines of evidence indicate that Akt-mediated signaling pathways play critical roles in regulating protein synthesis and proteolysis in muscle cells (Glass, 2005; Sandri, 2008, 2013; Schiaffino et al., 2013; Sanchez et al., 2014). For example, activation of the Akt/mTOR pathway stimulates protein synthesis. The phosphorylated Akt directly phosphorylates and inactivates FOXO, which is a major transcription factor of atrophy-related genes, atrogenes, such as atrogin-1 and MuRF-1 (Foletta et al., 2011; Gumucio and Mendias, 2013). Atrogin-1 and MuRF-1, two muscle-specific E3 ubiquitin ligases, induce the degradation of myofibril proteins. Therefore, the activation of Akt-mediated pathways and expression of atrogenes have been generally examined to understand protein metabolism in skeletal muscle.

Myostatin, a member of the transforming growth factor-β (TGF-β) superfamily, has been demonstrated to inhibit muscle mass development in mammals (McPherron et al., 1997; Huang et al., 2011; Rodriguez et al., 2014). For example, individual muscles of myostatin null mice weigh 2–3 times more than those of wild-type mice, and the increase in skeletal muscle mass appears to result from a combination of hyperplasia and hypertrophy (McPherron et al., 1997). Naturally occurring mutation in the myostatin gene leads to a hypermuscular phenotype in humans, mice, dogs, sheep, and cattle (Rodriguez et al., 2014). In addition, myostatin plays a fundamental role in regulating adult muscle growth and size (Rodriguez et al., 2014).

Accumulating evidence from previous studies have revealed the manner in which myostatin affects skeletal muscle growth. For example, myostatin downregulates the Akt/mTOR pathway and inhibits protein synthesis in C2C12 myotubes and mammalian skeletal muscles (Taylor et al., 2001; Welle et al., 2006; Amirouche et al., 2009; Morissette et al., 2009; Lipina et al., 2010). A recent in vitro study showed that high concentrations (2–3 μg/mL) of myostatin significantly inhibited the mTOR pathway and protein synthesis, whereas low concentrations (0.01–1 μg/mL) of myostatin significantly induced the phosphorylation of AMP-activated protein kinase (AMPK) and inhibited protein synthesis without affecting the mTOR pathway (Deng et al., 2017). In addition to the inhibition of protein synthesis, myostatin promotes the transcription of atrogin-1 by regu-
lating Smads and the Akt/FOXO pathway (Gumucio and Mendias, 2013; Lokireddy et al., 2011; McFarlane et al., 2006). Smads are transcription factors that are phosphorylated after myostatin binds to its receptors (Huang et al., 2011; Rodriguez et al., 2014). A previous in vivo study reported that Smad3 was sufficient to induce an increase in atrogin-1 promoter activity in murine skeletal muscle that was co-transfected with Smad3 and atrogin-1 promoter (Goodman et al., 2013). However, several in vitro and in vivo studies have shown that myostatin inhibits protein synthesis and the Akt/mTOR pathway without affecting protein degradation and expression of atrogin-1 and MuRF-1 (Taylor et al., 2001; Welle et al., 2006; Amirouche et al., 2009). Collectively, results suggest that myostatin inhibits skeletal muscle growth mainly by downregulating Akt phosphorylation in mammals.

The amino acid sequence of mature myostatin is identical among humans, mice, rats, pigs, dogs, chickens, and turkeys (Huang et al., 2011). However, previous studies have suggested that the roles of myostatin in skeletal muscles differ between mammals and chickens. For example, knockdown of the myostatin gene by RNA interference increased body, but not carcass, weight in chickens (Bhattacharya et al., 2017), and myostatin expression was not altered in lines of poultry exhibiting myofiber hyper- and hypoplasia (Mott and Ivarie, 2002). In contrast to the results from previous studies on mammalian skeletal muscles which showed that myostatin expression increased due to food deprivation or restriction (Allen et al., 2010; Zhao et al., 2016), myostatin mRNA levels decreased due to fasting in chicken skeletal muscles (Guernec et al., 2004; Saneyasu et al., 2015). Furthermore, it remains to be elucidated as to how myostatin affects the cellular signaling factors and expression of atrogenes in chicken skeletal muscle. To clarify the mechanism underlying species-specific roles of myostatin in chicken skeletal muscle, we investigated the effects of myostatin on the mRNA and protein levels of protein metabolism-related factors in chick embryonic myotubes.

Materials and Methods

Cell Culture

Breast muscles dissected from 14-day-old chick embryos were minced using surgical scissors and digested with HBSS (+) (Nacalai Tesque, Inc., Kyoto, Japan) containing 0.2% collagenase (Worthington Biochemical Corp., Lakewood, NJ, USA) for 20 min at 37°C. The cells were collected by centrifugation and resuspended in DMEM (Nacalai Tesque, Inc., Kyoto, Japan) supplemented with 15% serum (Fetal Clone III, GE Healthcare Bio-Sciences AB, Uppsala, Sweden), 1 × non-essential amino acid solution (Nacalai Tesque, Inc., Kyoto, Japan), and 1 × gentamicin/amphotericin solution (Life Technologies, Carlsbad, CA, USA). The cell suspension was passed through a cell strainer to remove tissue debris and then transferred to an uncoated flask to allow attachment of fibroblasts. After 1 h, the unattached cells were transferred to another uncoated flask and this procedure was repeated 2–3 times. The unattached cells were counted and plated onto collagen I-coated 12-well plates at 1 × 10⁵ cells/well. The cells were incubated in the medium described above at 37°C, with 5% CO₂ in a humidified chamber until the formation of myotubes. Myotubes were treated with or without 20 nM recombinant human/rat/mouse myostatin (PeproTech Inc., Rocky Hill, NJ, USA) for 2 h, in the absence or presence of serum.

Real-time PCR Analysis

The cells were washed with phosphate-buffered saline (PBS). Then, total RNA was extracted from the cells using Sepazol-RNA I (Nacalai Tesque, Inc., Kyoto, Japan). First-strand cDNA was synthesized from total RNA using ReverTra Ace® qPCR RT Master Mix with gDNA Remover (Toyobo Co. Ltd, Osaka, Japan). The levels of mRNA were quantified for each primer (Table 1) using TB Green Premix Ex Taq II (Tli RNaseH Plus; Takara Bio Inc., Otsu, Japan) according to the supplier’s recommendations, in an Applied Biosystems 7300 Real-Time PCR System (Applied Biosystems, Foster City, CA, USA). The expression levels of target genes were normalized to that of ribosomal protein S17 (RPS17).

Western Blot Analysis

Western blot analysis was performed as previously described by Saneyasu et al. (2017). Briefly, cells were rinsed with ice-cold PBS. Then, cells were scraped into 100 μL of lysis buffer. The cell lysates were ultrasonicated and centrifuged at 17,900 × g for 15 min at 4°C. The supernatants were stored at −80°C. Protein concentrations were determined using the Pierce BCA Protein Assay Kit (Pierce Biotechnology, Rockford, IL, USA). The lysates (4 μg of protein) were subjected to sodium dodecyl sulfate-polyacryl-

| Gene name   | Forward primer | Reverse primer | Accession number |
|-------------|----------------|----------------|-----------------|
| Atrogin-1   | 5′-cac ctt ggg aga agc ctt cca-3′ | 5′-cgg gga gtc gat gat a-3′ | NM_001030956 |
| FOXO1       | 5′-tct ggt cag gga aat gg-3′ | 5′-gcg gga gtc gat gat g-3′ | NM_204328 |
| MuRF-1      | 5′-tgtt aga aga tgg agc aag gct at-3′ | 5′-gct agg tgc tga aga ctg act-3′ | XM_424369 |
| Myostatin   | 5′-acc cac caa gat gtc ccc ta-3′ | 5′-acc cgc aac gat cta caa cc-3′ | NM_001001461 |
| RPS17       | 5′-ggc ggt gat gat cca gaa gt-3′ | 5′-gct ctt gtt ggt gtg aag t-3′ | NM_204217 |

FOXO, forkhead box class O; MuRF-1, muscle ring-finger protein 1; RPS17, ribosomal protein S17. The primers of atrogin-1, FOXO1, MuRF-1, and RPS17 have been used in previous studies (Saneyasu et al., 2015, 2016, 2017).
amidine gel electrophoresis (SDS-PAGE) and western blotting using HorizBlot (ATTO Co., Tokyo, Japan), according to the manufacturer’s instructions. Bands were detected by Chemi-Lumi one Super (Nacalai Tesque, Inc., Kyoto, Japan), visualized with the LumiCube (Liponics Inc., Tokyo, Japan), and quantified using the CS Analyzer software (ATTO Co., Tokyo, Japan). Anti-Akt (#9272), anti-phospho-Akt (pAkt) (Ser473) (#9271), anti-pAMPKα (Thr172) (#2531), anti-AMPKα (#2532), anti-FOXO1 (#9454), anti-pFOXO1 (Ser256) (#9461), anti-pSmad2 (Ser465/467)/ Smad3 (ser423/425) (#8828), anti-Smad2/3 (#8685), anti-β-actin (#4967), and horseradish peroxidase (HRP)-conjugated anti-rabbit IgG (#7074) were purchased from Cell Signaling Technology (Danvers, MA, USA). The species cross-reactivity is described in the manufacturer’s data sheet. Additionally, we confirmed with the manufacturer that each antigen sequence for #2532, #8828, and #8685 has 88%, 100%, and 100% homology with that of chicken, respectively. When detecting pSmad2/3, FOXO1, and pFOXO1, antibodies were diluted with Can Get Signal (Toyobo Co. Ltd, Osaka, Japan).

Statistical Analysis

Data were analyzed using the Dunnett’s test or t-test. All statistical analyses were performed using Excel 2013 (Microsoft, USA), with Statcel 3 add-in software (OMS, Tokyo, Japan).

Results

Myostatin significantly induced phosphorylation of Smad2 and expression of FOXO1 and atrogin-1 in chick embryonic myotubes cultured in the presence of serum, whereas no significant change was observed in the phosphorylation of Smad3, Akt, AMPK, and FOXO1, and in mRNA levels of MuRF-1 (Fig. 1). Although FOXO1 mRNA levels significantly increased, FOXO1 protein levels did not significantly change (Fig. 1). These results suggest that myostatin upregulates the expression of atrogin-1 and FOXO1 via Smad activation in an Akt/FOXO1 and AMPK pathway-independent manner.

The treatment with serum-free medium significantly increased mRNA levels of myostatin, atrogin-1, and MuRF-1 in chick myotubes (Fig. 2A). AMPK phosphorylation was significantly induced by serum-free treatment, whereas Akt phosphorylation was significantly inhibited (Fig. 2B). Unexpectedly, no significant change was observed in Smad2 and FOXO1 phosphorylation (Fig. 2B). In addition, FOXO1 mRNA levels were significantly decreased by serum-free treatment (Fig 2A), although no significant change was observed in its protein levels (Fig. 2B). All these findings suggest that serum-free treatment upregulates transcription of myostatin, atrogin-1, and MuRF-1, possibly via AMPK activation.

Similar to the results observed in the presence of serum, myostatin significantly increased Smad2 phosphorylation and FOXO1 expression in chick myotubes cultured under serum-free conditions, and no significant change was observed in the phosphorylation of Smad3, Akt, and AMPK, and in mRNA levels of MuRF-1 (Fig. 2). Moreover, no significant change was observed in FOXO1 protein levels and its phosphorylation (Fig. 2A). However, no significant change was observed in atrogin-1 mRNA levels (Fig. 2A). These results suggest that Smad activation did not enhance serum-free-induced upregulation of atrogin-1 expression.

Myostatin mRNA levels were dramatically decreased by myostatin treatment under serum-free conditions (Fig. 2B). Similarly, although no significant difference was observed, mRNA levels of myostatin decreased to 58% in myostatin-treated cells under serum-supplemented conditions (Supplementary Figure 1). Therefore, the present results suggest that myostatin expression is negatively regulated via myostatin-induced Smad activation in chicken skeletal muscle.

Discussion

A previous in vitro study using C2C12 myotubes showed that myostatin increases mRNA levels of FOXO1 and atrogin-1, but not MuRF-1 (McFarlane et al., 2006). Similarly, overexpression of activin receptor-like kinase 5 (which is activated by the binding of myostatin to an activin type II receptor and then phosphorylates Smads) or Smad3 increases the activity of atrogin-1 but not MuRF-1 promoter, in murine skeletal muscle (Goodman et al., 2013; Sartori et al., 2009). Smad3 can bind directly to DNA at the 5’-AAGAC-3’ sequence known as Smad-binding element, whereas Smad2 cannot bind directly to DNA (Matsuzaki 2011). However, Sartori et al. (2009) showed that RNAi-mediated knockdown of Smad3 alone or of both Smad2 and Smad3 induces a 10% and 22% increase in the cross-sectional area of murine muscle fiber, respectively. Smad2, therefore, also appears to be involved in the inhibition of muscle growth. In this study, using chick embryonic myotubes, similar results were observed, except Smad3 phosphorylation. These findings therefore suggest that the myostatin/Smad pathway plays a role in regulating the expression of proteolysis-related factors in chicken skeletal muscles, but the contributing isoform may be different from that in mammalian ones.

Previous studies using mammalian cells have investigated the long-term (12–48 h) effects of myostatin (McFarlane et al., 2006; Deng et al., 2017; Lokireddy et al., 2011). McFarlane et al. (2006) showed that myostatin-induced promotion of atrogin-1 expression is dependent on FOXO1. Additionally, Lokireddy et al. (2011) found that active Smad signaling is required for myostatin to increase the protein levels of FOXO1 and atrogin-1. In contrast, the present study investigated the relatively short-term (2h) effects of myostatin. Therefore, the difference in incubation time might be one of the reasons why no significant change was observed in the levels of Akt and FOXO1. Importantly, the present study showed that myostatin induced Smad2 phosphorylation and atrogin-1 expression without affecting the Akt/FOXO1 pathway. Smads are known to cooperate with other DNA binding proteins to elicit specific transcriptional response (Shi and Massagué, 2003). For example, Smad2/3 can bind members of the FOXO family of transcription factors to regulate gene expression (Gumucio and Mendias, 2013). Therefore, the present results might provide new
Fig. 1. (A) Effects of myostatin on the phosphorylation of Smad, Akt, FOXO1, and AMPK, (B) Effects of myostatin on the mRNA levels of FOXO1, atrogin-1, and MuRF-1, in the chick myotubes. The myotubes were treated with 15% serum-containing medium with or without (Con) 20 nM myostatin (Mstn), for 2 h. Data are expressed as mean ± standard error of the mean of five (A) and six (B) wells in each group. Student’s t-test was used to analyze the differences. ** Significant with respect to control group (**P < 0.01).
Fig. 2.  (A) Effects of serum-free conditions and myostatin treatment on the mRNA levels of myostatin, FOXO1, atrogin-1, and MuRF-1.  (B) Effects of serum-free conditions and myostatin treatment on the phosphorylation of Smad, Akt, FOXO1, and AMPK, in the chick myotubes. The myotubes were treated with either 15% serum-containing medium (Serum), serum-free medium (SF) with or without 20 nM myostatin (Mstn), for 2 h. Data are expressed as mean ± standard error of the mean of six wells in each group. The Dunnett’s test was used to analyze the differences between SF and Serum or Mstn groups. *, ** Significant with respect to SF group (* \( P < 0.05 \); ** \( P < 0.01 \)).
hypotheses, suggesting that Smads are able to promote atrogin-1 transcription in cooperation with FOXO1 independently of Akt-mediated regulation, and that Smad cooperates with other unknown factors to regulate atrogin-1 expression.

Serum-free treatment increases mRNA and protein levels of atrogin-1 and MuRF-1 and decreases phosphorylation of Akt and FOXO in C2C12 and L6 myotubes (Kim et al., 2015; Sandri et al., 2004), indicating that the increase in atrogin-1 and MuRF-1 expression is controlled by the Akt/FOXO pathway. However, a previous study in C2C12 myotubes showed that an AMPK activator increased atrogin-1 and MuRF-1 mRNA levels without affecting Akt phosphorylation (Egawa et al., 2014). In the present study, phosphorylation of AMPK, but not of FOXO1, was significantly induced by serum-free treatment with the increase in atrogin-1 and MuRF-1 expression. Therefore, it is possible that AMPK regulates atrogin-1 and MuRF-1 expression without affecting the Akt/FOXO pathway in both mammals and birds.

An AMPK activator reportedly increased the levels of myostatin protein in L6 myotubes (Lee et al., 2015). Levels of myostatin mRNA significantly increased in skeletal muscles of Smad3-null mice and in C2C12 myotubes treated with a Smad3 inhibitor (McFarlane et al., 2014). In the present study, AMPK appeared to increase myostatin expression in chick myotubes, whereas Smad appeared to inhibit myostatin expression even when AMPK was activated. Therefore, these findings suggest that AMPK and Smad control myostatin expression in chicken myotubes cultured in vitro, in the same manner as seen in mammals. In contrast to the results from in vitro studies, previous in vivo studies have suggested a different mechanism for myostatin expression in the skeletal muscle, between mammals and chickens. Fasting significantly promoted myostatin expression in mice (Allen et al., 2010) and inhibited its expression in chickens (Guerne et al., 2004; Saneyasu et al., 2015). Although the exact reason for this difference remains unknown, based on the present results, fasting-induced decrease in myostatin expression in chicken skeletal muscle might be caused by the activation of the Smad signaling pathway. Interestingly, a previous study found that myostatin is present extracellularly in murine skeletal muscle as uncleaved pro-myostatin and proposed that extracellular cleavage of pro-myostatin to mature myostatin represents a novel control point for myostatin synthesis. Biochemical and Biophysical Research Communications, 494: 278–284, 2017.

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