The return of Pfeiffer’s bacillus: Rising incidence of ampicillin resistance in *Haemophilus influenzae*

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**Abstract**

*Haemophilus influenzae*, originally named Pfeiffer’s bacillus after its discoverer Richard Pfeiffer in 1892, was a major risk for global health at the beginning of the 20th century, causing childhood pneumonia and invasive disease as well as otitis media and other upper respiratory tract infections. The implementation of the Hib vaccine, targeting the major capsule type of *H. influenzae*, almost eradicated the disease in countries that adapted the vaccination scheme. However, a rising number of infections are caused by non-typeable *H. influenzae* (NTHi), which has no capsule and against which the vaccine therefore provides no protection, as well as other serotypes equally not recognised by the vaccine. The first line of treatment is ampicillin, but there is a steady rise in ampicillin resistance. This is both through acquired as well as intrinsic mechanisms, and is cause for serious concern and the need for more surveillance. There are also increasing reports of new modifications of the intrinsic ampicillin-resistance mechanism leading to resistance against cephalosporins and carbapenems, the last line of well-tolerated drugs, and ampicillin-resistant *H. influenzae* was included in the recently released priority list of antibiotic-resistant bacteria by the WHO. This review provides an overview of ampicillin resistance prevalence and mechanisms in the context of our current knowledge about population dynamics of *H. influenzae*.

**DATA SUMMARY**

The data for Fig. 1 for different serotypes per country is derived from the ECDC (https://ecdc.europa.eu/en/invasive-haemophilus-influenzae-disease/atlas), the data for the Hib vaccine implementation from the WHO (http://www.who.int/immunization/monitoring_surveillance/data/en/). Fig. 2 shows a summary of several studies, all data is available from the previously published studies and reports, and is clearly cited in the text and listed in the bibliography.

**INTRODUCTION**

*H. influenzae* is a diverse species and has traditionally been classified by its polysaccharide capsule, which can either be entirely absent (non-typeable *H. influenza*, NTHi [1]), or one of six serotypes (a–f; [1–3]). In the 1980s, one of the most successful vaccines to date, the Hib vaccine, was widely implemented against the major disease-causing form of *H. influenzae*, serotype b; this serovar almost disappeared in countries which introduced Hib into their vaccination scheme [4] (Fig. 1a–c). The introduction of the vaccine in 73 countries supported by the Global Alliance for Vaccines and Immunizations (GAVI) was estimated to save 1.4–1.7 million lives 2011–2020 [5], and was introduced into all GAVI-supported countries by 2014 [6] (Fig. 1d, e). The success of the Hib vaccine is accompanied by rising numbers of NTHi as well as, to a much lesser extent, other serotypes [7–9], which fill the niche left by the strongly reduced prevalence of *H. influenzae* [10–13] (Fig. 1b). In post-Hib settings NTHi is now the main pathogenic lineage, causing upper respiratory tract and ear infections and chronic obstructive pulmonary disease (COPD) infections [14, 15] which is the third leading cause of death world-wide [16], and is one of the three leading causes of acute otitis media (AOM; together with *Streptococcus pneumoniae* and *Moraxella catarrhalis* [17]). NTHi is also the main type causing invasive disease, including sepsis and meningitis [11, 12, 17–19]. Even though invasive disease, especially meningitis, was mostly driven by Hib in the pre- and sometimes also the post-vaccination period [20, 21], several reports show NTHi meningitis cases comprise up to 69% of the *H. influenzae*-derived meningitis [19, 22–25]. For detailed discussion of virulence and interaction with the host the readers are pointed to several recent expert reviews [10–13, 17, 26], these aspects will not be covered in detail here.

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**Abbreviations:** BLNAR, beta-lactamase negative ampicillin resistant; BLPAR, beta-lactamase positive ampicillin resistant; COPD, chronic obstructive pulmonary disease; Hib, Haemophilus influenzae serotype B vaccine; NTHi, non-typeable Haemophilus influenzae; PBP, penicillin binding protein; PG, peptidoglycan.
The main treatment against *H. influenzae* is ampicillin, a member of the beta-lactams. These interfere with the biosynthesis of the Gram-negative cell wall through binding of the enzymes which cross-link the peptidoglycan subunits, the penicillin-binding proteins (PBP2, PBP3) [27, 28]. Key resistance mechanisms are either acquired enzymes that hydrolyse beta-lactams (beta-lactamases), or target modifications through changes in the PBP sequences. The majority of high-risk multi-drug-resistant Gram-negative pathogens to date acquired beta-lactamases, which mainly spread through mobile elements [29, 30]. There has been a steady increase in *H. influenzae* isolates resistant to ampicillin, including a high number of intrinsic resistance cases, and tracking of the epidemiology is challenging due to the genomic diversity of *H. influenzae* and its high prevalence of recombination. The importance of an increasing level of resistance amongst *H. influenzae* isolates has been recognised by the WHO, and it was included in the recently released list of high-priority antimicrobial-resistant pathogens [31]. This review will profile this currently understudied pathogen with an emphasis on the increasing prevalence of ampicillin resistance and the rising number of resistance mechanisms against other antimicrobial classes in the context of its complex population.

**Plasmid-derived ampicillin resistance**

Although the first report of resistance occurred in 1972 [32], *H. influenzae* is still largely treatable by most major groups of antimicrobials. The first beta-lactam-resistance mechanisms observed relied on the acquisition of the beta-lactamases blaTEM-1/2 and blaROB-1 [33, 34], and blaTEM-1 is the beta-lactamase most prevalent gene. It moved via the associated Tn2/3 transposon onto a cryptic plasmid already present in *H. influenzae*, which can also integrate into the chromosome and carries several resistances [18, 35]. More recently distribution of mobile resistances was also described on small conjugative plasmids, and transfer to *Escherichia coli*, as shown in vitro, indicates the possibility of inter-species resistance spread [36]; others of these small plasmids lack the conjugation machinery. However, other common *H. influenzae* plasmids were highly unstable when transformed into *E. coli* [37]; possibly of higher relevance as plasmid reservoirs for *H. influenzae* are closely related species like *Pasteurella multocida*, *Haemophilus haemolyticus* and *Haemophilus parasuis* [35, 37, 38]. Studies have shown the presence of conjugative plasmids with blaROB-1 shared amongst these species; however, these plasmids have a significant impact on the fitness of *H. influenzae*, which might explain the low prevalence of blaROB-1 [37]. However, due to the highly permeable cell envelope of *H. influenzae* [33, 39], there is an increased influx of beta-lactams [39]. The otherwise widespread blaTEM-1 enzyme thus needs to be modified to confer resistance in *H. influenzae*, either through modification of the promoter to achieve higher expression or through modification of the enzyme itself [40–43].

**Intrinsic resistance against ampicillin and other beta-lactams**

*H. influenzae* encodes four PBPs, however several studies have shown that there is no correlation between resistances and mutations in PBPs 1, 2 and 4 [44, 45]. PBP3 however, encoded by the gene ftsI, clearly confers resistance against ampicillin, and includes several mutations around functional areas of the protein. Ubukata et al. [46] initiated a typing scheme which can be used to classify PBP3 according to its mutations, and currently comprises four classes [47, 48]. The main resistance-conferring mutations are in one of the two key functional sites, disrupting the Lys–Thr–Gly (KTG) or the Ser–Ser–Asn (SSN) motif, which lead to a decrease in binding of ampicillin to PBP3 [45–47]. The thus intrinsically resistant strains against ampicillin are referred to as BLNAR, or beta-lactamase-negative ampicillin-resistant (Fig. 2); BLNAR strains that acquired a beta-lactamase in addition are referred to as beta-lactamase-positive amoxicillin-clavulanate resistant (BLPACR), however this group is usually only found at low prevalence. Whilst initially BLNAR was mainly observed in Japan, where Hib was only introduced in 2009 to the vaccination scheme [49], BLNAR has spread across the globe in the past 20 years (Fig. 2). NTHi, the, to date, most prevalent *H. influenzae* type causing disease, is often associated with considerable proportions of resistance in recent studies [50], with similar rates of acquired and intrinsic (BLNAR) resistance mechanisms [51–54]. BLNAR strains are grouped, according to their resistances, into low-level (MIC 1 µg ml$^{-1}$; sensitive=MIC 0.25 µg ml$^{-1}$) and high-
level (MIC $2\mu$g ml$^{-1}$) resistance. High-level-resistant strains also show resistance against cephalosporins [46, 47] and have increased in prevalence in parts of Asia [48, 55–59]. In addition, upregulation of AcrAB, combined with PBP3 changes, can lead to further resistance to cephalosporins as well as carbapenems, the last-line drugs that are widely applicable against Gram-negative infections [47, 60–64].

**Rising incidence of resistances against other drugs through intrinsic mechanisms**

Given the increasing trend in resistances, the treatment options for *H. influenzae* are becoming increasingly limited. Azithromycin, a macrolide, is widely prescribed in respiratory disease, although there is need for caution against over-use to prevent selection for resistance [65]. Similar to ampicillin resistance, *H. influenzae* modifies the macrolide targets (50S ribosomal RNA and ribosome-binding proteins), leading to high levels of resistance [66, 67]. This mechanism is in addition to the ubiquitously present AcrAB multidrug-efflux pump which provides a baseline reduced susceptibility, and the authors of recent studies have reported changes in the AcrAB sequence to further contribute to resistance [68]. Another major alternative are quinolones, however, recently there has been an accumulation of reports of fluoroquinolone resistance [69–72] again through mutations in the targets, DNA gyrase (encoded by *gyrA* and *gyrB*) and topoisomerase IV (encoded by *parC* and *parE*); and thus again conferring resistance through a plasmid-independent mechanism, making it easy for resistances to arise independently through point mutations and/or spread through recombination.
**H. influenzae population structure**

Not only recognised as a pathogen very early on, *H. influenzae* was also the bacterium that opened the area of whole-genome sequencing, representing the first complete genome from a free-living organism sequenced (in 1995) [73]. One of the main challenges of NTHi is its great diversity; whilst the capsulated lineages are mainly clonal expansions [74], with each serotype presenting a monophyletic lineage, initial multi-locus sequence typing (MLST) studies already recognised the increased recombination, and as a consequence higher diversity, of NTHi [75]; and in population studies, there are often almost as many STs in a dataset as NTHi sequences [76–78]. NTHi can also persist in a host over several years, which not only leads to changes in virulence genes as well as potential vaccine targets [79, 80], but also increases its exposure to antimicrobials to treat unrelated infections of the host, and is another risk factor in the acquisition or spread of resistances through point mutations and recombination, respectively. Recombination in *H. influenzae* is elevated for the whole species through its natural competence, a bacterial mechanism increasing the uptake of DNA from the same species. This is achieved by a DNA uptake machinery located at the cell envelope that shows a strong bias toward certain short DNA sequence motifs, and the genomes of the respective bacteria are highly enriched in the preferred sequences, mostly equally distributed throughout the genome and also reflected in the encoded peptides [81–83]. This leads to an increased ability to recombine between different strains [78, 84], and thus also facilitates easy exchange of chromosomal resistance mechanisms, for example *ftsI* [85]. In addition, recombination between related species has been observed, and leads to spread of resistance through exchange of the chromosomally encoded target gene *ftsI* [86].

**Interactions with other species**

*H. influenzae* shares its niche with *Streptococcus pneumoniae* and *Moraxella catarrhalis* in the nasopharynx, three of the four primary pathogens (with *Pseudomonas aeruginosa*) causing COPD [87], and the three leading pathogens of AOM [17]. Similar to the success of the Hib vaccine, the main disease-causing serovars of *S. pneumoniae* were successfully controlled with several vaccines, but also here we
see an increase in serovars not targeted by the available vaccines and in and drug resistances, including penicillin as well as macrolides [88–91]. Mixed infections further provide the potential to share resistance mechanisms across species without actual gene transfer: *Moraxella catarrhalis* can secrete outer membrane vesicles (OMVs) containing beta-lactamase which hydrolyse beta-lactams, and thus protect de facto sensitive *S. pneumoniae* and *H. influenzae* strains from the antimicrobial [92], as well as complement resistance factors, protecting from the immune system [87]. A similar effect was observed with beta-lactamase-containing OMVs secreted by *H. influenzae* protecting group A Streptococci, another common component of the nasopharynx microbiota [93]. More studies for a better understanding are also required to understand the impact of the widely used pneumococcal vaccines. Results from a recent study have indicated that introduction of the *S. pneumoniae* vaccine resulted in increased carriage of NTHi, although not due to the expansion of a particular lineage/sequence type [94]. This study is, however, in contrast to other reports, where a reduction of NTHi carriage following vaccination with pneumococcal vaccine was observed [95–98]. Other studies reported no change in NTHi following pneumococcal vaccination; given the different study designs and the differences in pneumococcal vaccines, some of which include the *H. influenzae* protein D as carrier, more data will be required to better understand the impact of different pneumococcal vaccines on *H. influenzae* [17, 99, 100].

**Conclusion**

*H. influenzae* is a major pathogen, considering respiratory as well as invasive disease. The success of the Hib vaccine has been followed by a global increase of NTHi, which receives little attention from public media and is not included in major resistance surveillance programs. This is likely to be since NTHi can still readily be treated in most settings, but there is an alarming rise in intrinsic resistances against beta-lactams, as well as other classes. While a lot of focus is placed on acquired resistance mechanisms, the epidemiology of intrinsic resistance is much less understood; this is additionally challenging for NTHi, which has a highly diverse population structure and can undergo recombination, including exchange of resistance determinants, within the species as well as with related species. Intrinsic resistance, especially against beta-lactams, is also a significant problem for future drug developments. A major effort to control the current crisis in antimicrobial resistance is the development of beta-lactamase and carbapenemase inhibitors [101–103] used in combination with beta-lactams and carbapenems, respectively, to revert cells with these acquired enzymes to be de facto susceptible. There are currently 10 inhibitors in clinical development, but changes in PBP3 and AcrAB escape these treatment options [104]. Given the limited amount of genomic surveillance data, and that *H. influenzae* resistance is not included in current antimicrobial-resistance monitoring programs [105], the extent of the increase in resistant isolates, as well as the population dynamics leading to the spread of resistance, are not clear. There is an urgent need for better surveillance of resistant *H. influenzae*, understanding of the population dynamics, including recombination events, and the influence of pneumococcal vaccines.

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**Conflicts of interest**

The authors declare that there are no conflicts of interest.

**References**

1. Pittman M. Variation and type specificity in the bacterial species *Haemophilus influenzae*. J Exp Med 1931;53:471–492.
2. Kroll JS, Moxon ER. Capsulation and gene copy number at the cap locus of *Haemophilus influenzae* type b. J Bacteriol 1988;170: 859–864.
3. Stuy HJ. On the nature of nontypable *Haemophilus influenzae*. Antonie van Leeuwenhoek 1978;44:367–376.
4. World Health Organization. 2018. Immunization, vaccines and biologicals - data, statistics and graphics. www.who.int/immunization/monitoring_surveillance/data/en/.
5. Lee LA, Franzel L, Atwell J, Datta SD, friberg IK et al. The estimated mortality impact of vaccinations forecast to be administered during 2011–2020 in 73 countries supported by the GAVI Alliance. Vaccine 2013;31:B61–B72.
6. GAVI. 2018. *Haemophilus influenzae* type b vaccine support. www.gavi.org/support/rv/s/hib/.
7. Ulanova M, Tsang RSW. *Haemophilus influenzae* serotype a as a cause of serious invasive infections. Lancet Infect Dis 2014;14: 70–82.
8. Nitta DM, Jackson MA, Burry VF, Olson LC. Invasive *Haemophilus influenzae* type f disease. Pediatr Infect Dis J 1995;14:157–160.
9. Desai S, Jamieson FB, Patel SN, seo CY, Dang V et al. The epidemiology of invasive *Haemophilus influenzae* non-serotype B disease in Ontario, Canada from 2004 to 2013. PLoS One 2015; 10:e0142179.
10. Gilsdorf JR. What the pediatrician should know about non-typeable *Haemophilus influenzae*. J Infect 2015;71:S10–S14.
11. Duell BL, Su YC, Riesbeck K. Host-pathogen interactions of non-typeable *Haemophilus influenzae* from commensal to pathogen. FEMS Lett 2016;590:3840–3853.
12. Van Eldere J, slack MPE, Ladhani S, Cripps AW. Non-typeable *Haemophilus influenzae*, an under-recognised pathogen. Lancet Infect Dis 2014;14:1281–1292.
13. Jalaifand F, Riesbeck K. Update on non-typeable *Haemophilus influenzae*-mediated disease and vaccine development. Expert Rev Vaccines 2018;17:503–512.
14. Finney Lj, Ritchie A, Pollard E, Johnston SL,mallia P. Lower airway colonization and inflammatory response in COPD: a focus on *Haemophilus influenzae*. Int J Chron Obstruct Pulmon Dis 2014;9:1119–1132.
15. Dagan R, Leibovitz E. Bacterial eradication in the treatment of otitis media. Lancet Infect Dis 2002;2:593–604.
16. World Health Organization. 2018. Global health estimates 2016: deaths by cause, age, sex, by country and by region, 2000–2016. www.who.int/news-room/fact-sheets/detail/the-top-10-causes-of-death.
17. Cerqueti M, Giufrè M. Why we need a vaccine for non-typeable *Haemophilus influenzae*. Hum Vaccin Immunother 2016;12:2357–2361.
18. Tristram S, Jacobs MR, Appelbaum PC. Antimicrobial resistance in *Haemophilus influenzae*. Clin Microbiol Rev 2007;20:368–389.
patients’ background factors. J Infect Chemother 2015;21:247–256.

55. Honda H, Sato T, Shinagawa M, Fukushima Y, Nakajima C et al. Multiclonal expansion and high prevalence of β-lactamase-neg-
ative high-level ampicillin-resistant Haemophilus influenzae in Japan, and susceptibility to quinolones. Antimicrob Agents Chemother 2018;AAC.00851-18.

56. Park C, Kim KH, Shin NY, Byun JH, Kwon EY et al. Genetic diver-
sity of the ftsl gene in β-lactamase-nonproducing ampicillin-resistant and β-lactamase-produces amoxicillin-clavulanic acid-resistant nasopharyngeal Haemophilus influenzae strains isolated from children in South Korea. Microb Drug Resist 2013; 19:224–230.

57. Sanbongi Y, Suzuki T, Osaki Y, Senju N, Ida T et al. Molecular evolution of β-lactam-resistant Haemophilus influenzae: 9-year surveillance of penicillin-binding protein 3 mutations in isolates from Japan. Antimicrob Agents Chemother 2006;50:2487–2492.

58. Hashida K, Shimori T, Hohchi N, Muratani T, Mori T et al. Nasopharyngeal Haemophilus influenzae carriage in Japanese children attending day-care centers. J Clin Microbiol 2008;46: 876–881.

59. Hagiwara E, Baba T, Shinohara T, Nishihira R, Komatsu S et al. Antimicrobial resistance genotype trend and its association with host clinical characteristics in respiratory isolates of Haemophilus influenzae. Chemotherapy 2012;58:352–357.

60. Seyama S, Wajima T, Nakamimi H, Noguchi N. Clarithromycin resistance mechanisms of epidemic β-lactamase-nonproducing ampicillin-resistant Haemophilus influenzae strains in Japan. Antimicrob Agents Chemother 2014;60:3207–3210.

61. Kitaoka K, Kimura K, Kitanaka H, Banno H, Jin W et al. Carbape-
nem-nonsusceptible Haemophilus influenzae with penicillin-binding protein 3 containing an amino acid insertion. Antimicrob Agents Chemother 2018;62:e00671-18.

62. Skaare D, Anthonisen IL, Kahlmeter G, Matuschek E, Nata-
Fuursted K, Hartmeyer GN, Stegger M, Andersen PS, Justesen et al. Emergence of clonally related multidrug resistant Haemophi-
lus influenzae with penicillin-binding protein 3-mediated resistance to extended-spectrum cephalosporins, Norway, 2006 to 2013. Euro Surveill 2014;19:20986.

63. Fuursted K, Hartmeyer GN, Stegger M, Andersen PS, Justesen US. Molecular characterisation of the clonal emergence of high-
level ciprofloxacin-monoresistant Haemophilus influenzae in the region of Southern Denmark. J Glob Antimicrob Resist 2016;5: 67–70.

64. Cherkaoui A, Diene SM, Renzoni A, Emonet S, Renzi G et al. Imi-
penem heteroresistance in non-typeable Haemophilus influenzae is linked to a combination of altered PBP3, slow drug influx and direct efflux regulation. Clin Microbiol Infect 2017;23:118.e9–118.e19.

65. Serisier DJ. Risks of population antimicrobial resistance associ-
ated with chronic macrolide use for inflammatory airway dis-
bases. Lancet Respir Med 2013;1:262–274.

66. Peric M, Bozdogan B, Jacobs MR, Appelbaum PC. Effects of an efflux mechanism and ribosomal mutations on macrolide sus-
cceptibility of Haemophilus influenzae clinical isolates. Antimicrob Agents Chemother 2003;47:1017–1022.

67. Bogdanovich T, Bozdogan B, Appelbaum PC. Effect of efflux on telithromycin and macrolide susceptibility in Haemophilus influ-
enezae. Antimicrob Agents Chemother 2006;50:893–898.

68. Seyama S, Wajima T, Nakamimi H, Noguchi N. Amino acid substitution in the major multidrug efflux transporter protein AcrB contributes to low susceptibility to azithromycin in Haemophi-
lus influenzae. Antimicrob Agents Chemother 2017;61:e01337-17.

69. Seyama S, Wajima T, Yanagisawa Y, Nakamimi H, Ushio M et al. Rise in Haemophilus influenzae with reduced quinolone susceptibility and development of a simple screening method. Pediatr Infect Dis J 2017;36:263–266.

70. Kondo H, Ito S, Hatazaki K, Horige K, Nakane K et al. Gyrase and/or ParC alterations of Haemophilus influenzae strains isolated from the urethra of men with acute urethritis. J Infect Chemother 2018;24:232–235.

71. Kuo SC, Chen PC, Shiu YR, Wang HY, Lai JF et al. Levofloxacin-resistant Haemophilus influenzae, Taiwan, 2004–2010. Emerg Infect Dis 2014;20:1386–1390.

72. Puig C, Tirado-Vélez JM, Calatayud L, Tubau F, Garmendia J et al. Molecular characterization of fluoroquinolone resistance in nontypeable Haemophilus influenzae clinical isolates. Antimicrob Agents Chemother 2015;59:661–666.

73. Fleischmann RD, Adams MD, White O, Clayton RA, Kirkness EF et al. Whole-genome random sequencing and assembly of Hae-
mophilus influenzae Rd. Science 1995;269:496–512.

74. Musser JM, Kroll JS, Granoff DM, Moxon ER, Brodeur BR et al. Global genetic structure and molecular epidemiology of encap-
sulated Haemophilus influenzae. Rev Infect Dis 1990;12:75–111.

75. Meats E, Feil EJ, Stringer S, Cody AJ, Goldstein R et al. Charac-
terization of encapsulated and noncapsulated Haemophilus influ-
enezae and determination of phylogenetic relationships by multilocus sequence typing. J Clin Microbiol 2003;41:1623–1636.

76. Kaur R, Chang A, Xu Q, Casey JR, Pichichero ME. Phylogenetic relatedness and diversity of non-typable Haemophilus influenzae in the nasopharynx and middle ear fluid of children with acute otitis media. J Med Microbiol 2011;60:1841–1848.

77. de Chiara M, Hood D, Muzzi A, Pickard DJ, Perkins T et al. Genome sequencing of disease and carriage isolates of non-
typeable Haemophilus influenzae identifies discrete population structure. Proc Natl Acad Sci USA 2014;111:5439–5444.

78. Power PM, Bentley SD, Parkhill J, Moxon ER, Hood DW. Investi-
gations into genome diversity of Haemophilus influenzae using whole genome sequencing of clinical isolates and laboratory transformants. BMC Microbiol 2012;12:273.

79. Pettigrew MM, Ahearn CP, Gent JF, Kong Y, Gallo MC et al. Haemophi-
lus influenzae genome evolution during persistence in the human airways in chronic obstructive pulmonary disease. Proc Natl Acad Sci USA 2018;115:E3256–E3265.

80. Gallo MC, Kirkham C, Eng S, Bebawee RS, Kong Y et al. Changes in IgA protease expression are conferred by changes in genomes during persistent infection by nontypeable Haemo-
philus influenzae in chronic obstructive pulmonary disease. Infect Immun 2018;86:e00313-18.

81. Findlay WA, Redfield RJ. Coevolution of DNA uptake sequences and bacterial proteomes. Genome Biol Evol 2009;1:45–55.

82. Smith HO, Gwinn ML, Salzberg SL. DNA uptake signal sequen-
ces in naturally transformable bacteria. Res Microbiol 1999;150: 603–616.

83. Chen I, Dubnau D. DNA uptake during bacterial transformation. Nat Rev Microbiol 2004;2:241–249.

84. Mell JC, Shumilina S, Hall IM, Redfield RJ. Transformation of natural genetic variation into Haemophilus influenzae genomes. PLoS Pathog 2011;7:e1002151.

85. Skaare D, Anthonisen IL, Caugant DA, Jenkins A, Steinbak M et al. Multilocus sequence typing and ftsl sequencing: a powerful tool for surveillance of penicillin-binding protein 3-mediated beta-lactam resistance in nontypeable Haemophilus influenzae. BMC Microbiol 2014;14:131.

86. Witherden EA, Bajanca-Lavado MP, Tristram SG, Nunes A. Role of inter-species recombination of the ftsl gene in the dissemi-
nation of altered penicillin-binding-protein-3-mediated resistance in Haemophilus influenzae and Haemophilus haemolyticus. J Antimicrob Chemother 2016;69:1501–1509.

87. Perez AC, Murphy TF. Potential impact of a Moraxella catarrhalis vaccine in COPD. Vaccine 2017.

88. Lai C-C, Lee K, Xiao Y, Ahmad N, Veeraraghavan B et al. High burden of antimicrobial drug resistance in Asia. J Glob Antimicrob Resist 2014;2:141–147.
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