Catecholamines Facilitate Fuel Expenditure and Protect Against Obesity via a Novel Network of the Gut-Brain Axis in Transcription Factor Skn-1-deficient Mice

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A B S T R A C T

Taste signals and nutrient stimuli sensed by the gastrointestinal tract are transmitted to the brain to regulate feeding behavior and energy homeostasis. This system is referred to as the gut-brain axis. Here we show that both brush cells and type II taste cells are eliminated in the gastrointestinal tract of transcription factor Skn-1 knockout (KO) mice. Despite unaltered food intake, Skn-1 KO mice have reduced body weight with lower body fat due to increased energy expenditure. In this model, 24-h urinary excretion of catecholamines was significantly elevated, accompanied by increased fatty acid β-oxidation and fuel dissipation in skeletal muscle and impaired insulin secretion driven by glucose. These results suggest the existence of brain-mediated energy homeostatic pathways originating from brush cells and type II taste cells in the gastrointestinal tract and ending in peripheral tissues, including the adrenal glands. The discovery of food-derived factors that regulate these cells may open new avenues for the treatment of obesity and diabetes.

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1. Introduction

Taste signals and nutrient stimuli sensed by the gastrointestinal (GI) tract are transmitted to the central nervous system, including the nucleus of the solitary tract and hypothalamus, via afferent neurons and hormonal mediators, thereby controlling feeding behavior and energy homeostasis. This system is referred to as the gut-brain axis (Cummings and Overduin, 2007; Furness, 2012). This information is subsequently conveyed from the brain to peripheral tissues via efferent sympathetic and parasympathetic nerves including preganglionic sympathetic splanchnic input to the adrenal glands. For example, gut incretins such as glucose-dependent insulinotropic peptide (GIP) and glucagon-like peptide-1 (GLP-1), which are released from the intestinal enteroendocrine cells in response to nutrient ingestion, potentiate glucose-stimulated insulin secretion (GSIS) from pancreatic β-cells (Wu et al., 2015). GLP-1 also activates the GLP-1 receptors expressed on vagal afferent nerve terminals in the portal vein and in the brain across...
the blood-brain barrier, which is involved in the regulation of appetite (Katsurada et al., 2014; Kinzig et al., 2002; Vahl et al., 2007). Gavage of agonists for bitter taste receptor increases food intake via the secretion of ghrelin, a hormone that potentiates hunger sensation (Janssen et al., 2011).

The Skn-1 (also known as Pou2f3) gene, which encodes the POU homeodomain transcription factor, was originally identified as a regulator of the differentiation of epidermal keratinocytes (Andersen et al., 1993, 1997). We previously reported that Skn-1a is expressed in sweet, umami (savory), and bitter-sensing taste cells, which are referred to as type II taste cells (Matsumoto et al., 2011). Type II taste cells are completely eliminated in Skn-1 knockout (KO) mice, resulting in loss of electrophysiological and behavioral responses to sweet, umami, and bitter tastes. Thus, Skn-1a is critical for generating type II taste cells. Skn-1a is also required for the generation of Trpm5-expressing solitary chemosensory cells (SCCs) in the nasal respiratory epithelium and microvillus cells in the main olfactory epithelium (Ohmoto et al., 2013; Yamaguchi et al., 2014). Skn-1a is expressed in several tissues including the stomach, but not in the brain (Yukawa et al., 1993).

There are four major cell types in the small intestine: enterocytes and Goblet, Paneth, and enteroendocrine cells (van der Flier and the adult mouse intestinal epithelium (Gerbe et al., 2012). Brush cells to as tuft cells or caveolated cells) constitute a minor fraction (0.4%) of are supposed to be chemosensory cells (Young, 2011) and express transient receptor potential melastatin 5 (Trpm5), Doublecortin-like kinase 1 (Dclk1), and choline acetyltransferase (Bezencon et al., 2008; Gerbe et al., 2009, 2011; Saqui-Salces et al., 2011; Schütz et al., 2015). The mechanisms of differentiation and the function of brush cells remain elusive.

In the present study, we found that Skn-1a regulates differentiation of Trpm5-expressing brush cells in the GI tract. Skn-1 KO mice exhibited reduced body weight with lower body fat than wild-type (WT) littermates. Despite unaltered food intake, Skn-1 KO mice exhibited increased energy expenditure, caused by augmented catecholamine secretion. Our work raises the concept that taste cells receiving sweet, bitter, and umami tastes as well as brush cells receiving food-borne chemicals are involved in regulating body weight and body fat. Collectively, the present study provides new insights into the regulation of energy homeostasis originating from brush cells and taste cells in the GI tract and signaling to peripheral tissues including the adrenal glands, via the brain.

2. Material and Methods

2.1. Experimental Animals

All animal experiments were approved by the Animal Care and Use Committee at The University of Tokyo. Skn-1/Pou2f3-deficient mice with a mixed 129 × C57BL/6J background were generated as previously described (Matsumoto et al., 2011) and then backcrossed to C57BL/6J for >10 generations. Male Skn-1 KO mice and their WT littermates generated by crossing the heterozygous mice were used in the present study. Mice were fed a normal chow diet (Lab MR Breeder, Nusan Co., Yokohama, Japan) after weaning or High-Fat Diet 32 (CREA Japan Inc., Tokyo, Japan) from 4 weeks of age for ~12 weeks to induce dietary obesity. Mice were maintained in a room with a constant temperature of 22 ± 1°C under a 12-h light-dark cycle (lights on at 8 a.m.). Tissue samples were collected from 12- to 16-week-old mice that were fasted for 18 h.

2.2. X-ray Computed Tomography (CT) Scan Analysis

Mice older than 16 weeks were anesthetized using an isoflurane nebulizer (Muromachi Kidai Co., Ltd., Tokyo, Japan); a Latheta LCT-200 CT instrument (Aloka-Hitachi LCT-200, Tokyo, Japan) was then used to scan abdominal fat and muscle mass and extract thighbone density.

2.3. Blood Chemical Parameters

Blood was collected from approximately 20-week-old mice by cardiac puncture at 10 a.m. under ad libitum feeding condition and after 18-h fasting. Serum chemical parameters and hormones (PTH, T3, and T4) were analyzed by Nagahama Life Science Laboratory (Shiga, Japan). Plasma insulin and leptin were measured using the Lumines 200™ System (Lumines, Austin, TX, USA) by GeneticLab Co., Ltd. (Sapporo, Japan). Plasma adiponectin and FGF21 were measured using mouse adiponectin ELISA (BioVendor, Brno, Czech Republic) and mouse and rat FGF-21 ELISA (BioVendor), respectively.

2.4. Indirect Calorimetry

Indirect calorimetry was performed on 20- to 28-week-old mice using an indirect calorimeter (ARCO-2000, Arco Systems Inc., Chiba, Japan) and spontaneous motor activity was measured using a pyroelectric infrared ray sensor (NS-AS01, Neuroscience Inc., Tokyo, Japan) for 3–4 days after 2 days of habituation essentially as described by Tschop et al. (2011) or with an indirect calorimeter (MK-5000RQ, Muromachi Kidai Co. Ltd.) as previously described (Watanabe et al., 2014). The respiratory exchange ratio (RER) was calculated as the molar ratio of VCO2/VO2. Energy expenditure (kcal per hour) was calculated as heat (kcal/h) = (3.815 + 1.232 × RER) × VO2.

2.5. Mitochondrial DNA Copy Number

To measure the mitochondrial DNA copy number, the ratio of mitochondrial DNA to genomic DNA was calculated by a previously described method (Watanabe et al., 2014). In brief, total DNA was extracted from the gastrocnemius of 14- to 19-week-old mice that were fasted for 16 h using a QiAamp DNA mini kit (QIAGEN Ltd., Tokyo, Japan) and used for qPCR (Applied Biosystems Japan Ltd., Tokyo, Japan). We used the TaqMan Gene Expression Assay (Applied Biosystems Foster City, USA; ACTB, Mm00607939_s1; CYTB, Mm044225274). The PCR cycling conditions were 50 °C for 2 min, 95 °C for 10 min, and 40 cycles of 95 °C for 15 s and 60 °C for 1 min. The mitochondrial DNA copy number is presented relative to nuclear DNA following amplification of the mitochondrial gene region (cytochrome b vs. the nuclear endogenous control region, β-actin).

2.6. DNA Microarray

DNA microarray analysis using total RNA extracted from the gastrocnemius of 12- to 16-week-old mice that were fasted for 18 h was performed as previously described (Nakai et al., 2008) with minor modifications. In brief, Total RNA samples (100 ng each) were prepared and processed for microarray analysis using a 3' IVT Express kit and GeneChip® Mouse Genome 430 2.0 Array (Affymetrix, Santa Clara, CA, USA) according to the manufacturer’s standard protocols. The raw microarray data (CEL files) were quantified by the Factor Analysis for Robust Microarray Summarization (FARMS) (Hochreiter et al., 2006) using the statistical language R (Team and Computing, 2005) and Bioconductor (Gentleman et al., 2004). To identify differentially expressed genes, the rank products (RP) method (Bretling et al., 2004) was applied to the FARMS-quantified data. All microarray data are MIAME compliant and have been deposited in a MIAME-compliant database, the National Center for Biotechnology Information (NCBI) Gene Expression Omnibus (http://www.ncbi.nlm.nih.gov/geo/, GEO Series accession number GSE76936), as described in more detail on the MGED Society web site (http://www.mged.org/Workgroups/MIAME/miame.html).
2.7. Fecal TG and Total Energy Content

Fecal lipids were extracted from dried feces of 15-week-old mice fed a HFD according to the Folch method (Folch et al., 1957), and the TG concentration was measured using the Wako triglyceride E-Test (Wako Pure Chemical Industries, Ltd., Osaka, Japan). The total energy content of feces collected for 8 weeks was measured by bomb calorimetry.

2.8. Measurement of Urine Catecholamines

Twenty-four-hour urine (for catecholamine measurements) was collected into collection vials from 11- to 12-week-old mice individually housed in metabolic cages. To avoid spontaneous oxidation of catecholamine, 60 μL of hydrochloric acid (6 mol/L) was added to the collection vials (Moreira-Rodrigues et al., 2012). The collected urine was purified using MonoSpin PBA (GL Sciences, Tokyo, Japan) and urine catecholamines were measured by HPLC as previously described (Tsunoda et al., 2011). In brief, Chromatography was performed using an Inertsil ODS-4 column (5 μm, 250 × 20 mm i.d., GL Science) with acetate-citrate buffer at a flow rate of 0.5 mL/min and detected by electrochemical detection. The concentration of urine creatinine was measured using a LabAssay Creatinine kit (Wako Pure Chemical Industries, Ltd.).

2.9. qRT-PCR in Adrenal Glands

Total RNA was extracted from adrenal glands using RNeasy mini-columns (Qiagen, Venlo, Netherlands), followed by genomic DNA

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**Fig. 1.** Brush cells are eliminated in the GI tract of Skn-1 KO mice. (a) Tissue distribution of Skn-1a. RT-PCR analysis revealed that Skn-1a was abundantly expressed in the circumvallate papillae, stomach, duodenum, jejunum, and ileum but absent or only faintly expressed in all other tissues examined, including the pancreas, islets of Langerhans, brain, liver, skeletal muscle, white adipose tissue (WAT), and brown adipose tissue (BAT). (b) Immunostaining revealed strong Skn-1a signals in the corpus beneath the limiting ridge in the stomach (upper panel) and in a subset of cells in the small intestine (lower panel). Inset in the upper right corner of each panel shows higher magnification image of the area enclosed by the dashed box. (c) In situ hybridization revealed that Trpm5 mRNA was expressed in the jejunum of WT mice (upper panel) but not in Skn-1 KO mice (lower panel). (d) Immunostaining revealed Dclk1 signals in WT mice (upper panel) but not Skn-1 KO mice (lower panel). The arrowheads indicate signals. (e) Double immunostaining demonstrated that nearly all Trpm5-positive cells were also positive for Skn-1a. The arrows indicate cells expressing Trpm5 and Skn-1a. Scale bars: 100 μm and 25 μm (inset). See also Fig. S1.
digestion using RNase-free DNase (Qiagen). First-strand cDNA was generated by reverse transcription from total RNA (Superscript III Reverse Transcription Kit; Life Technologies, Gaithersburg, MD). Levels of mRNA transcripts were determined by qPCR (ABI Prism 7000 Sequence Detection System, Applied Biosystems, Foster City, CA). cDNA levels were normalized against β-actin. PCR amplification was performed using TaqMan technology (TaqMan Gene Expression Assays, Applied Biosystems). The TaqMan probe IDs were as follows: tyrosine hydroxylase (Th; Mm00447557_m1), dopa decarboxylase (Ddc; Mm00516688_m1), dopamine β-hydroxylase (Dbh; Mm00460472_m1), phenylethanolamine N-methyltransferase (Pnmt; Mm00476993_m1), and β-actin (Mm00607939_s1). The delta-delta method was used for relative quantification (Livak and Schmittgen, 2001).

Fig. 2. Metabolic phenotypes of Skn-1 KO mice fed a normal chow diet. (a) Changes in body weight under normal chow diet conditions. When fed a normal chow diet, Skn-1 KO mice had lower body weight than their WT littermates for 13 weeks after weaning (n = 75). Also see Fig. S2A. (b) Skn-1 KO mice had a lower body fat percentage than their WT littermates, whereas there was no significant difference in muscle mass percentage between the two genotypes (n = 10–19). (c and d) Serum levels of total ketone bodies were higher in Skn-1 KO mice than their WT littermates under ad libitum feeding (n = 18–28) (c) and 18-h fasting conditions (n = 13) (d). (e) Skn-1 KO and WT littermates had similar food intake at 8 weeks of age (n = 5–8). (f) Skn-1 KO mice exhibited increased energy expenditure compared with their WT littermates (n = 9). (g) qRT-PCR analysis revealed that the mitochondrial DNA content in skeletal muscle was significantly higher in Skn-1 KO mice than in their WT littermates. The data are presented as the mean ± SEM; **p<0.01 and *p<0.05. See also Fig. S2.
2.10. OGTT, IPGTT, ITT, and Measurement of Insulin, GIP, and GLP-1

Gavage administration of glucose (3.0 mg/g of body weight for measurement of blood glucose, plasma insulin, and plasma total GIP; 5.0 mg/g of body weight for measurement of plasma active GLP-1; 2.0 mg/g of body weight for measurement of blood glucose and plasma insulin in mice fed a HFD) was performed on 16-week-old WT and Skn-1 KO mice at 10 a.m. after 18-h fasting (n = 20–30 animals per group) using a syringe attached to a feeding needle (KN-349-M1, Natsume Seisakusho Co., Ltd., Tokyo, Japan) inserted through the mouth into the stomach. For IPGTT or ITT, mice were fasted for 18 or 4 h, then injected intraperitoneally with glucose (1.5 mg/g of body weight) or insulin (0.75 μIU/g of body weight), respectively. Tail blood samples were collected from the tail vein before gavage (time 0) and 15, 30, 45, 60, 90, and 120 min after gavage. For measurement of GIP and GLP-1, tail blood samples were collected into chilled tubes containing DPP-IV inhibitor (Merck Millipore, Billerica, MA) and/or EDTA and aprotinin (for GLP-1; Wako Pure Chemical Industries, Ltd.) before gavage (time 0) and 15, 30, and 60 min after gavage (for GIP) or 10, 20, and 40 min after gavage (for GLP-1). Blood glucose, plasma insulin, plasma total GIP, and plasma active GLP-1 were measured by Glucose Pilot (Iwai Chemicals Co. Ltd., Tokyo, Japan), Ultra Sensitive Mouse Insulin ELISA Kit (Morinaga Institute of Biological Science Inc., Kanagawa, Japan), Rat/Mouse GIP (Total) ELISA Kit (EZRMICP-55K, Merck Millipore), and GLP-1 (Active) ELISA Kit (AKMGP-011, Shibayagi Co. Ltd., Gunma, Japan), respectively.

2.11. RT-PCR

First-strand cDNAs from 11 different tissues except the circumval- luate papillae (Genostaff, Tokyo, Japan) were used for RT-PCR. For the circumvallate papillae, total RNA was extracted and reverse-transcribed to cDNA with an oligo dT primer. Approximately 500 bp of coding region encompassing multiple exons of Skn-1a was amplified from each cDNA for 30 cycles at an annealing temperature of 60 °C. The volume of PCR products for Skn-1a loaded on the agarose gel was approximately normalized using the PCR product for GAPDH.

2.12. Islet Isolation and In Vitro Insulin Release From Mouse Islets

Isolated pancreatic islets were prepared by liberase (Roche Diagnos- tics, Basel, Switzerland) digestion, manually selected, and incubated in RPMI 1640 tissue culture medium (Life Technologies) supplemented with 10% (vol/vol) fetal bovine serum at 37 °C overnight. Insulin secretion in vitro was measured in static incubations. Prior to experiments, islets were pre-incubated for 60 min at 37 °C in Krebs-Ringer (KR) buffer composed of 119 mM NaCl, 4.7 mM KCl, 2.5 mM CaCl2, 1.2 mM MgCl2, 1.2 mM KH2PO4, 25 mM NaHCO3, and 10 mM HEPES (pH 7.4) with 5% bovine serum albumin. The medium was gassed with 95% O2 and 5% CO2 to maintain constant pH and oxygenation. The groups of 10 islets were incubated for 60 min at 37 °C in Krebs-Ringer buffer solution containing 2.5 mM glucose (low glucose). The islets were transferred to new low glucose Krebs-Ringer buffer and incubated for 60 min. Subsequently, the islets were incubated in Krebs-Ringer buffer containing 25 mM glucose (high glucose) for 30 min. The supernatants from the second low-glucose solution and from the high-glucose solution were collected for the insulin ELISA assay and stored at −80 °C. Insulin concentrations were measured using an Ultra Sensitive Mouse Insulin ELISA Kit (Morinaga Institute of Biological Science).

2.13. Immunohistochemistry

Rabbits were immunized with peptides corresponding to residues 1140–1153 (C+PASARDREYLESGL) of Trpm5 (Operon Biotechnologies, Tokyo, Japan). Preincubation of the antibody with peptide antigen (10 ng/mL) abolished the brush cell staining, confirming the specificity of the antibody (data not shown). The other primary antibodies used were as follows: rabbit anti-Skn-1a antibody (5sc-330, Santa Cruz Biotechnology, Santa Cruz, CA), rabbit anti-Dclkl antibody (#AP7219b, Abgent, San Diego, CA), rabbit anti-ChgA antibody (#20085, ImmunoStar, Hudson, WI), and goat anti-GLP-1 antibody (5sc-7782, Santa Cruz Biotechnology). Immunohistochemistry was performed as previously described (Ishimaru et al., 2006). In brief, fresh-frozen sections (10-μm thick) were fixed in 4% paraformaldehyde (PFA), permeabilized with ice-cold methanol, blocked with phosphate-buffered saline (PBS) containing 5% skim milk, and incubated with rabbit anti-Skn-1a antibody (1/1000 dilution), rabbit anti-Trpm5 antibody (1/200 dilution), rabbit anti-Dclkl antibody (1/50 dilution), or rabbit anti-ChgA antibody (1/200 dilution), followed by incubation with Alexa Fluor 488- or 594-conjugated secondary antibodies (Santa Cruz Biotechnology). Nuclear staining was performed using DAPI (4′,6-diamidino-2-phenylindole) (Roth Biocell, Saint-Aubin, France). The sections were mounted with ProLong Gold Antifade reagent with DAPI (Invitrogen). Images were captured using a Zeiss LSM 780 confocal microscope.

Table 1

| Gene symbol | Gene name | Probe ID |
|-------------|-----------|----------|
| ABCD2       | ATP-binding cassette, sub-family D (ABD), member 2 | 1419748_at |
| Ociad2      | OCA-I domain containing 2 | 1435917_at |
| Ptcrt7      | PTC7 protein phosphatase homolog (S. cerevisiae) | 1455958_8_at |
| 4930402E16Rik | RIKEN cDNA4930402E16 Rik gene | 1458669_x_at |
| 8430406C28Rik | RIKEN cDNA8430406C28 gene | 1433837_at |
| Tbc1d15     | TBC1 domain family, member 15 | 1416062_at |
| Xaf1        | XIAP associated factor 1 | 1443608_at |
| Col4a3d3p   | Collagen, type IV, alpha 3 (Goodpasture antigen) binding protein | 1420384_at |
| Dsp         | Desmoplakin | 1435493_at, 1435494_at |
| DltA        | Dihydrolipoamide S-acetyltransferase (E2 component of pyruvate dehydrogenase complex) | 1426264_at, 1426265_x_at |
| Fdx1        | Ferredoxin 1 | 1449108_at |
| Gpam        | Glyceraldehyde-3-phosphate dehydrogenase, mitochondrial | 1419499_at |
| Hk2         | Hexokinase 2 | 1422612_at |
| Coq10b      | Coenzyme Q10 homolog B (S. cerevisiae) | 1406510_x_at |
| Inmmt       | Inner membrane protein, mitochondrial | 1429533_at |
| Ide         | Insulin degrading enzyme | 1423121_at, 1435140_at |
| Krt5        | Keratin 5 | 1424096_at |
| Klf1b       | Kinesin family member 1B | 1423995_at, 1451642_at, 1453200_at, 1425770_at |
| Elovl6      | ELOVL family member 6, elongation of long chain fatty acids (yeast) | 1417403_at |
| Cide1       | Cold shock domain containing E1, RNA binding | 1423997_at |
| Prkca       | Protein kinase C, alpha | 1437393_at, 1446598_at, 1450945_at |
| Ppm1K       | Protein phosphatase 1K (PP2C domain containing) | 1441988_at |
| Ppp3ca      | Protein phosphatase 3, catalytic subunit, alpha isoform | 1426401_at, 1438478_x_at |
| Reep1       | Receptor accessory protein 1 | 1433509_s_at |
| NDS3        | NADH dehydrogenase subunit 5 | 1426088_at |
| Opa1        | Optic atrophy 1 | 1434880_at |
| Slc25a12    | Solute carrier family 25 (mitochrondrial carrier, Aralar), member 12 | 1436440_at |
| Slc25a25    | Solute carrier family 25 (mitochondrial carrier, phosphate carrier), member 25 | 1424735_at, 1447836_x_at |
| Tmnc1       | Transmembrane and tetratricopeptide repeat containing 1 | 1435261_at |
| Tap1        | Transporter 1, ATP-binding cassette, sub-family B (MDR/TAP) | 1416016_at |
| Ucp3        | Uncoupling protein 3 (mitochondrial, proton carrier) | 1420658_at |
Fluor 488- (#A21206) or Alexa Fluor 555-conjugated anti-rabbit IgG (#A31572, Life Technologies). For GLP-1, fresh-frozen sections (10-μm thick) were fixed with 4% PFA, blocked with PBS containing 5% normal donkey serum and 0.2% Triton X-100, and incubated with goat anti-GLP-1 antibody (1/150 dilution) followed by incubation with Alexa Fluor 555-conjugated anti-goat IgG (#A21432, Life Technologies).

For double staining, fresh-frozen sections (10-μm thick) were fixed with 4% PFA and permeabilized with ice-cold methanol followed by antigen retrieval. The sections were then blocked with Tris-buffered saline with 0.05% Tween 20 (TBST) containing 5% skim milk and incubated with rabbit anti-Trpm5 antibody (1/200 dilution), followed by incubation with horseradish peroxidase (HRP)-conjugated anti-rabbit IgG (#31460, Life Technologies) and subsequent reaction with tyramide signal amplification (TSA)-Alexa Fluor 488 (Life Technologies) as a substrate for peroxidase. After antigen retrieval, the sections were blocked with TBST containing 5% skim milk and incubated with rabbit anti-Skn-1a antibody (1/300 dilution) or rabbit anti-Dclk1 antibody (1/50 dilution), followed by incubation with Cy3-conjugated anti-rabbit IgG (#711-165-152, Jackson ImmunoResearch, West Grove, PA).

Fig. 3. Metabolic phenotypes of Skn-1 KO mice fed a HFD. (a) Changes in body weight under HFD conditions. When fed a HFD for 12 weeks after 4 weeks of age, Skn-1 KO mice exhibited dramatically reduced body weight compared with their WT littermates (n = 54). (b) Skn-1 KO mice had a lower body fat percentage than their WT littermates, whereas there were no significant differences in muscle mass percentage between the two genotypes (n = 18–26). (c) Skn-1 KO mice had lower tissue weights of epididymal and perirenal WAT (n = 6–12). (d) Skn-1 KO and WT littermates had similar food intake at 8 weeks of age (n = 8–10). (e) Skn-1 KO mice exhibited increased energy expenditure compared with their WT littermates (n = 4). The data are presented as the mean ± SEM; **p < 0.01 and *p < 0.05. See also Fig. S3.
2.14. In Situ Hybridization

In situ hybridization was performed as previously described (Ishimaru et al., 2005). In brief, fresh-frozen sections (10-μm thick) of the small intestine, including the duodenum, jejunum, and ileum, were placed on MAS-coated glass slides (Matsunami Glass, Osaka, Japan) and fixed in 4% PFA in PBS. Prehybridization (at 58 °C for 1 h), hybridization (at 58 °C, 2 O/N), washing (0.2× SSC at 58 °C), and development (NBT-BCIP) were performed using a digoxigenin-labeled probe for Trpm5. Stained images were obtained using a fluorescence microscope (BX51, Olympus, Tokyo, Japan) equipped with a cooled CCD digital camera (DP71, Olympus).

2.15. Data Analysis and Statistical Methods

All data are presented as the mean ± SEM. Statistically significant differences were assessed using Student's t-test.

3. Results

3.1. Brush Cells in the GI Tract Are Abolished in Skn-1 KO Mice

We first investigated whether Skn-1a plays important roles in the regulation of cell differentiation in the stomach and small intestine. To examine the mRNA expression of Skn-1a in the GI tract, we performed RT-PCR analysis of mRNA from several tissues of WT mice. Skn-1a was abundantly expressed in the circumvallate papillae, stomach, duodenum, jejunum, and ileum, but it was absent or only faintly expressed in all other tissues examined, including the pancreas, islets of Langerhans, brain, liver, skeletal muscle, white adipose tissue (WAT), and brown adipose tissue (BAT) (Fig. 1a). RT-PCR of GAPDH as a control revealed comparable levels in all the tissues other than islets of Langerhans and WAT, in which slightly less intense bands were detected (Fig. 1a). To examine the tissue distribution of Skn-1a in the stomach and intestine at the cellular level, we next performed immunohistochemistry using an anti-Skn-1a antibody. In the stomach, Skn-1a signals were abundantly detected in the corpus beneath the limiting ridge (Fig. 1b). In the duodenum, jejunum, and ileum, Skn-1a signals were observed in approximately 100 cells per section (Fig. 1b and data not shown). These results suggest that Skn-1a functions as a transcription factor that regulates cell differentiation in the stomach and intestine as well as in the taste buds, nasal respiratory epithelium, and main olfactory epithelium (Matsumoto et al., 2011; Ohmoto et al., 2013; Yamaguchi et al., 2014).

We next examined the expression of Trpm5 in the small intestine. In situ hybridization analysis using a probe for Trpm5 revealed that Trpm5 signals scattered throughout the small intestine including the duodenum, jejunum, and ileum, of WT mice (Fig. 1c and data not shown), consistent with the previous reports (Bezencon et al., 2008). Intriguingly, no signals of Trpm5 were observed in Skn-1 KO mice (Fig. 1c). Immunohistochemistry using an anti-Trpm5 antibody confirmed the presence of the Trpm5 protein in the duodenum of WT mice but the absence of Trpm5 protein in Skn-1 KO mice (Fig. S1a).

Doublecortin-like kinase 1 (Dclk1) antibody marks brush cells (Gerbe et al., 2009, 2011; Saqui-Salces et al., 2011). Chromogranin A (ChgA) is a marker for enteroendocrine cells (Rindi et al., 2004). GLP-1 is an incretin, a peptide hormone, secreted from enteroendocrine L cells [Wu et al., 2015]. Signals of Dclk1 were detected in WT mice but not in Skn-1 KO mice (Fig. 1d). By contrast, signals of ChgA and GLP-1 were observed in WT and Skn-1 KO mice (Fig. S1a). Finally, we performed double staining using antibodies against Trpm5 and Skn-1a or Dclk1. Nearly all Trpm5-positive cells were also positive for Skn-1a or Dclk1 (Figs. 1e and S1b). Taken together, these results demonstrate that brush cells expressing Trpm5 and Dclk1 are abolished in Skn-1 KO mice and that Skn-1a is a crucial transcription factor for generating brush cells in the GI tract.

3.2. Metabolic Phenotypes of Skn-1 KO Mice Fed a Normal Chow Diet

To examine the effect of eliminating type II taste cells and brush cells in the GI tract on energy homeostasis, we characterized the metabolic phenotypes of Skn-1 KO mice. Whereas there was no significant difference in body weight at birth between the Skn-1 KO and WT littermates (Fig. S2a), Skn-1 KO mice exhibited lower body weight compared to their WT littermates when weaned at 3 weeks of age (Figs. 2a and S2a). When fed a normal chow diet, Skn-1 KO mice exhibited significantly lower body weight than their WT littermates throughout the 13 weeks after weaning (Fig. 2a). For example, the body weights of the Skn-1 KO and WT littermates at 16 weeks of age were 27.4 ± 0.5 g and 29.8 ± 0.4 g, respectively. X-ray computed tomography (CT) analysis of body composition revealed that Skn-1 KO mice had a lower body fat percentage than their WT littermates at approximately 20 weeks of age, whereas there was no significant difference in muscle mass percentage between the two genotypes (Fig. 2b). Among blood chemical parameters, the concentration of serum total ketone bodies was significantly higher in Skn-1 KO mice than in their WT littermates under ad libitum feeding and 18-h fasting conditions (Fig. 2c and d). In addition, some parameters related to lipid metabolism, such as cholesterol and non-esterified fatty acid (NEFA) levels, tended to be higher in Skn-1 KO mice compared with their WT littermates (Tables S1 and S2). Collectively, these results demonstrate that lipid metabolism, such as lipid degradation and β-oxidation of fatty acids, is accelerated in Skn-1 KO mice.

Despite the lower body weight and body fat percentage of Skn-1 KO mice, there was no difference in food intake between the Skn-1 KO and WT littermates at 8 weeks of age (Fig. 2e) and in water intake between the two genotypes (data not shown). Analysis of the metabolic rates of the mice revealed that the energy expenditure of Skn-1 KO mice was higher than that of their WT littermates (Fig. 2f). By contrast, there were no significant differences in the respiratory exchange ratio (RER)

![Fig. 4. Enhanced catecholamine secretion in Skn-1 KO mice fed a HFD. (a) Serum triiodothyronine (T3), tetraiodothyronine (T4), and thyroid stimulating hormone (TSH) were comparable between Skn-1 KO and WT littermates (n = 7–8). (b) Urinary epinephrine and dopamine were higher in Skn-1 KO mice than their WT littermates (n = 13–14). The data are presented as the mean ± SEM; *p < 0.05. See also Fig. S4.](image)
or spontaneous motor activity between the two genotypes (Figs. S2b and S2c). These results demonstrate that increased energy expenditure causes reduced body weight with lower body fat percentage in Skn-1 KO mice.

Quantitative real-time PCR (qPCR) analysis revealed that the mitochondrial DNA content in the skeletal muscle was significantly higher in Skn-1 KO mice than in their WT littermates (Fig. 2g). Moreover, DNA microarray analysis revealed enhanced expression of genes involved in mitochondrial function, such as uncoupling proteins 3 (Ucp3), in Skn-1 KO mice (Table 1). These results suggest that energy expenditure is increased in the skeletal muscle of Skn-1 KO mice.

3.3. Metabolic Phenotypes of Skn-1 KO Mice Fed a HFD

We next examined the metabolic phenotypes of Skn-1 KO mice fed a HFD. When fed a HFD for >12 weeks after 4 weeks of age, the body weight of Skn-1 KO mice was dramatically lower than that of their WT littermates (Fig. 3a). For example, the body weights of Skn-1 KO and
WT littersmates at 16 weeks of age were 30.4 ± 0.9 g and 39.5 ± 0.9 g, respectively. CT analysis of body composition and measurement of tissue mass revealed that Skn-1 KO mice had a lower body fat percentage, including epididymal and perirenal white adipose tissues, than their WT littersmates at approximately 20 weeks of age, whereas there were no significant differences in the weight ratios of other tissues, such as muscle, between the two genotypes (Fig. 3b and c and Table S3).

Although there was no significant difference in food intake between the Skn-1 KO and WT littersmates at 8 weeks of age (Fig. 3d), energy expenditure was higher in Skn-1 KO mice than in their WT littersmates, but spontaneous motor activity was similar between the two genotypes (Fig. 3e and data not shown). In addition, we observed no differences in food intake or fecal triacylglycerol (TG) between the Skn-1 KO and WT littersmates at 15 weeks of age (Figs. S3a and S3b). Fecal energy content was also comparable between the Skn-1 KO and WT littersmates (Fig. S3c). These results suggest that increased energy expenditure is the primary cause of reduced adiposity in Skn-1 KO mice fed a HFD.

Finally, we measured plasma adiponectin, FGF21, insulin, and leptin in Skn-1 KO and WT littersmates fed a HFD (Table S4). Plasma leptin tended to be lower in Skn-1 KO mice compared with their WT littersmates (p = 0.08).

3.4. Catecholamine Secretion is Augmented in Skn-1 KO Mice Fed a HFD

We examined whether the levels of thyroid hormones, including triiodothyronine (T3), tetraiodothyronine (T4), and thyroid stimulating hormone (TSH), and catecholamines, including norepinephrine, epinephrine, and dopamine, were altered in Skn-1 KO mice fed a HFD. Serum T3, T4, and TSH were comparable between Skn-1 KO and WT littersmates (Fig. 4a). We next measured 24-h urinary excretion of catecholamines. Urinary epinephrine and dopamine were significantly higher and norepinephrine tended to be higher (p = 0.06) in Skn-1 KO mice compared to their WT littersmates (Fig. 4b). qPCR analysis revealed that the gene expression of catecholamine biosynthetic enzymes, including tyrosine hydroxylase (Th), dopa decarboxylase (Ddc), dopamine-β-hydroxylase (Dbh), and phenylethanolamine N-methyltransferase (Pnmt), in adrenal glands was comparable between Skn-1 KO and their WT littersmates (Fig. S4). These results suggest that catecholamines are over-secreted from adrenal glands in Skn-1 KO mice.

3.5. Insulin Secretion is Reduced in Skn-1 KO Mice

To examine carbohydrate metabolism in Skn-1 KO mice, we performed an oral glucose tolerance test (OGTT). Blood glucose concentrations in Skn-1 KO and WT littersmates were comparable for 120 min after oral glucose administration (Fig. 5a). By contrast, the increase in plasma insulin 60 min after glucose administration was smaller in Skn-1 KO mice than in their WT littersmates (Fig. 5b). However, plasma GIP and GLP-1, which potentiate insulin secretion from pancreatic β-cells (Wu et al., 2015), were comparable between Skn-1 KO and WT littersmates after oral glucose administration (Figs. S5a and S5b).

We next examined the insulin secretion ability of pancreatic β-cells in Skn-1 KO mice. First, RT-PCR analysis revealed that Skn-1a was not expressed in the pancreas or islets of Langerhans of WT mice (Fig. 1a). Second, insulin secretion from isolated islets in response to 25 mM glucose was comparable between Skn-1 KO and WT littersmates (Fig. S5c). Third, in the intraperitoneal glucose tolerance test (IPGTT), blood glucose concentrations were comparable between Skn-1 KO and WT littersmates for 120 min after administration (Fig. S5d). Plasma insulin concentrations were also comparable between Skn-1 KO and WT littersmates before and at 15 min after intraperitoneal administration (Fig. S5e). Collectively, these results demonstrate that insulin secretion from pancreatic β-cells is normal in Skn-1 KO mice. Finally, the insulin tolerance test (ITT) revealed that Skn-1 KO and WT littersmates had comparable insulin sensitivity (Fig. S5f).

When fed a HFD, blood glucose was significantly lower in Skn-1 KO mice than their WT littersmates for 120 min after oral glucose administration (Fig. 5c), demonstrating that impaired glucose tolerance in WT littersmates fed a HFD is improved in Skn-1 KO mice. In addition, plasma insulin levels were lower in Skn-1 KO mice than in their WT littersmates before glucose administration (p < 0.05) and tended to be lower at 15 min after administration (p = 0.10. Fig. 5d). ITT revealed that the Skn-1 KO mice had improved insulin resistance compared with their WT littersmates (Fig. 5e). Collectively, these results demonstrate that

![Fig. 6. Schematic representation of the presumed metabolic pathways originating from brush cells and type II taste cells in the GI tract. Food components and digested nutrients are detected by the tongue and GI tract. In Skn-1 KO mice, the lack of type II taste cells and brush cells results in increased catecholamine secretion. Lipolysis in WAT, serum levels of total ketone bodies, and thermogenesis in muscle are increased, whereas insulin secretion from the pancreas is reduced. Consequently, Skn-1 KO mice have reduced body weight with lower body fat percentage due to higher energy expenditure.](image-url)
the insulin resistance and impaired glucose tolerance observed in WT mice fed a HFD are ameliorated in Skn-1 KO mice.

4. Discussion

In the present study, we demonstrated that brush cells and type II taste cells in the GI tract (Matsumoto et al., 2011) are unexpectedly abolished in Skn-1 KO mice. This finding indicates that Skn-1 is critical for the development of Trpm5-expressing chemosensory cells in mice. The Skn-1 KO mice had reduced body weight with a lower body fat percentage, primarily due to increased energy expenditure. Furthermore, catecholamine secretion was increased in Skn-1 KO, accompanied by decreased insulin secretion. We propose brain-mediated mechanisms of energy homeostasis originating from brush cells and taste cells in the GI tract to peripheral tissues including the adrenal glands (Fig. 6). In detail, taste signals and nutrient stimuli, which are sensed by taste cells and brush cells in the GI tract are transmitted to the brain via afferent neurons and/or humoral mediators and subsequently conveyed to the adrenal glands that secrete catecholamines via efferent sympathetic nerves. In Skn-1 KO mice, loss of taste and/or nutrient sensing may intensify the activity of the efferent sympathetic nerves, resulting in exaggerated secretion of catecholamines. Based on the network of the gut-brain axis originating from brush cells and taste cells in the GI tract, the present study suggests an approach to the treatment of obesity.

Skn-1a is expressed in Trpm5-expressing chemosensory cells. Loss-of-function of Skn-1a results in the elimination of these cell types (Matsumoto et al., 2011; Ohmoto et al., 2013; Yamaguchi et al., 2014). In the present study, we demonstrated that Skn-1a is expressed in Trpm5-expressing brush cells in the GI tract including stomach and intestine. It should be noted that these cells are abolished in the GI tract of Skn-1 KO mice. Genes encoding sweet taste receptors (T1R2 and T1R3) and downstream signaling molecules such as gustducin are expressed in enteroendocrine L cells that secrete GLP-1 (Jang et al., 2007). When glucose was orally administered, the plasma GLP-1 concentration was not augmented in gustducin KO mice compared with WT mice, which resulted in a lack of a rapid increase in plasma insulin levels and an impaired glucose tolerance (Jang et al., 2007). We showed that GLP-1 signals were similar between WT and Skn-1 KO mice in immunohistochemical analyses, consistent with normal incretin release after oral glucose administration in Skn-1 KO mice. These results demonstrate that Skn-1a is also required for the generation of Trpm5-expressing brush cells, but not for that of enteroendocrine L cells, in the GI tract.

Trpm5 is expressed not only in the chemosensory cells but also in the mouse pancreatic β-cell line MIN6, pancreatic islets of Langerhans, and pancreatic β-cells (Colson et al., 2010; Nakagawa et al., 2009; Oya et al., 2011; Prawitt et al., 2003; Yamamoto and Ishimaru, 2013). Trpm5 plays an important role in the regulation of GSIS from pancreatic β-cells via two major mechanisms: triggering and amplifying pathways (Brixel et al., 2010; Colson et al., 2010; Liman, 2010). Trpm5 KO mice, like Skn-1 KO mice, gain significantly less body weight and fat mass and are more sensitive to insulin than WT mice when fed either a HFD or a high-carbohydrate diet (Larsson et al., 2015). T1R3 is the common subunit of sweet (T1R2 + T1R3) and umami (T1R1 + T1R3) taste receptors (Yarmolinsky et al., 2009). When fed a HFD, T1R3 KO mice had reduced fat mass, in spite of unaltered body weight (Simon et al., 2014). T1R3 is expressed in the gut, pancreas, and adipose tissue as well as in the tongue (Yamamoto and Ishimaru, 2013). In contrast, Skn-1a was not expressed in the pancreas, islets of Langerhans, or brain, suggesting that the tissue distribution of Skn-1a is more restricted than that of Trpm5 and T1R3. In this context, Skn-1 KO mice are unique models for investigating unexplored mechanisms underlying the gut-brain axis and whole-body energy metabolism.

Skn-1 KO mice exhibited lower body weight than their WT littermates after weaning when fed either a normal chow diet or a HFD, whereas body weight was indistinguishable between Skn-1 KO and WT littermates at birth. Notably, the Skn-1 KO mice had much lower body weight than their WT littermates when fed a HFD; the body weight gain of Skn-1 KO mice fed a HFD for 12 weeks was nearly equivalent to that of WT mice fed a chow diet during the same period, and the Skn-1 KO mice did not become obese. In Skn-1 KO mice fed a HFD, blood glucose levels and insulin secretion as well as insulin tolerance were improved, and plasma leptin, an indicator of body fat mass, tended to be lower than in their WT littermates. These results demonstrate that Skn-1 KO mice are resistant to HFD-induced metabolic dysregulation.

The reduced body weight of Skn-1 KO mice was not due to decreased food intake or nutrient absorption but to increased energy consumption, which resulted in a lower body fat ratio. Skn-1 KO mice exhibited slightly but significantly higher urinary epinephrine and dopamine, implying an increase in the total amounts of these hormones released in 24 h. Because there was no difference in the expression of the biosynthetic enzymes for catecholamine in the adrenal glands, it is unlikely that this significant increase in catecholamine excretion was caused by catecholamine overproduction. Because peripheral secretion of catecholamine is induced by activation of the sympathetic nervous system (Watanabe et al., 1988) and Skn-1a is not expressed in the brain, it is tempting to speculate that the sympathetic nervous system in Skn-1 KO mice is activated by unidentified signals from the GI tract to the brain and not alteration of brain function.

In particular, epinephrine and norepinephrine are critical to maintain energy production under hypoglycemia and malnutrition through lipolysis, ketogenesis (Bahnson et al., 1984; Steiner et al., 1991), and increased mitochondrial in skeletal muscle (Miura et al., 2007). In adipocytes, catecholamines induce lipolysis, but insulin inhibits lipolysis (Nielsen et al., 2014). Consistent with previous reports, the copy number of mitochondrial DNA and the concentration of serum total ketone bodies were increased in Skn-1 KO mice, in accordance with an increase in energy consumption in this model. These results indicate that the appropriate elevation of catecholamine secretion in Skn-1 KO mice may protect against obesity and metabolic syndrome. Acute psychosocial stress induces changes in catecholamine sensitivity (Strahler et al., 2015). Pheochromocytoma is a tumor arising from adrenomedullary chromaffin cells that automatically overproduce catecholamines (Lenders et al., 2014). Patients with pheochromocytoma with similar levels of circulating catecholamines exhibit variable phenotypes, possibly due in part to differences in hormone sensitivity to the corresponding receptors (Markou et al., 2015). Catecholamine sensitivity in peripheral tissues may be similarly enhanced by stress signals from the brain in Skn-1 KO mice.

In the OGTT, Skn-1 KO mice exhibited a smaller increase in plasma insulin compared with their WT littermates, whereas blood glucose concentrations were indistinguishable between Skn-1 KO and WT littermates. Why did the Skn-1 KO mice exhibit such a reduced increase in plasma insulin despite similar changes in blood glucose? First, plasma concentrations of incretins, such as GIP and GLP-1, were comparable between Skn-1 KO and WT littermates after oral glucose administration, which is consistent with the immunohistochemical analyses revealing that GLP-1 signals were similar between WT and Skn-1 KO mice. Second, insulin secretion from pancreatic β-cells was normal in Skn-1 KO mice. Third, Skn-1 KO and WT littermates showed comparable insulin sensitivity in the ITT. By contrast, urinary epinephrine and dopamine were significantly higher in Skn-1 KO mice than their WT littermates. Catecholamines inhibit insulin secretion from pancreatic β-cells (Barth et al., 2007). Overexpression of pancreatic alpha2A-adrenergic receptor in the diabetic Goto-Kakizaki rat mediates adrenergic suppression of insulin secretion (Rosengren et al., 2009). Higher catecholamine secretion may inhibit insulin secretion in Skn-1 KO mice. Collectively, our results demonstrate that enhanced catecholamine secretion in Skn-1 KO mice induces lipolysis in WAT, accelerates β-oxidation of fatty acids in the liver and skeletal muscle, and, consequently, thermogenesis in skeletal muscle, and inhibits insulin secretion from the pancreas. A range of diverse effects contribute to resistance to HFD-induced obesity and fuel dyshomeostasis (Fig. 6). Further studies are warranted to elucidate the molecular mechanisms underlying the regulation of energy metabolism.
originating from brush cells and type II taste cells in the GI tract, e.g., identifying which chemosensory receptors are expressed in the brush cells of the GI tract.

5. Conclusions
In a network of the gut-brain axis, the loss of brush cells and taste cells in the GI tract alters energy metabolism in Skn-1 KO mice. We thus propose the concept that taste-receiving cells in the oral cavity and/or food-borne chemicals—receiving brush cells in the gut are involved in regulation of the body weight and adiposity. The discovery of food-derived factors that regulate these cells may open new avenues for the treatment of obesity and diabetes.

Conflicts of Interest
The authors declare that they have no conflicts of interest.

Author Contributions
S.U., Y.I., and K.A. designed the experiments. S.U., Y.I., M.N., H.M., and K.A. performed experiments and analyzed the data. C.K. and M.T. guided the experimental methods. N.O., T.A., and H.M. participated in discussions. S.U., Y.I., M.N., H.M., and K.A. wrote the paper.

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Appendix A. Supplementary data
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