Respiratory impedance is correlated with morphological changes in the lungs on three-dimensional CT in patients with COPD

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The forced oscillation technique provides information concerning respiratory impedance, which comprises resistance and reactance of the respiratory system. However, its relationship with morphological changes of the lungs in chronic obstructive pulmonary disease (COPD) remains unclear. Respiratory impedance and spirometric data were evaluated in 98 patients with COPD and 49 reference subjects. Wall thickness (WT) and airway intraluminal area (Ai) of third- to sixth-generation bronchi, and percentage low-attenuation area with less than —950 HU (%LAA) of lungs were measured using three-dimensional computed tomography. COPD patients had higher respiratory impedance, decreased Ai, and increased %LAA compared with reference subjects. Indices of respiratory resistance and reactance and forced expiratory volume in 1 second (FEV1) were correlated with Ai, and the association between percent predicted FEV1 and Ai was predominant in distal bronchi. The difference in respiratory resistance between 5 Hz and 20 Hz (R5–R20) and FEV1/forced vital capacity ratio (FEV1/FVC) were correlated with WT. The %LAA was correlated with the FEV1/FVC ratio and respiratory reactance. Airway function measurements with the forced oscillation technique provide complementary information to spirometry in COPD.

Chronic obstructive pulmonary disease (COPD) is characterized by persistent airflow limitation caused by inhalation of cigarette smoke and/or other noxious particles1. Small airway disease and parenchymal destruction are the key pathologic characteristics of COPD, and lead to air trapping and airflow limitation1–3.

Measurement of pulmonary function based on spirometry is required for the diagnosis of COPD, and is the most reproducible and objective measurement of airflow limitation among several methods available1. However, spirometric measurement is not sufficient for comprehensive assessment of COPD. Guidelines set by the Global Initiative for Chronic Obstructive Lung Disease (GOLD) emphasize the importance of multifaceted assessment for COPD in addition to spirometric assessment1.

Computed tomography (CT) of the chest provides information about pathologic changes in COPD. Studies have demonstrated that airway narrowing and thickening of airway walls on cross-sectional CT of the chest are associated with airflow limitation as assessed by spirometry in COPD patients3–6. Full assessment of airways by conventional CT is difficult because of the complicated structure with the branching framework of the lungs. However, three-dimensional (3D)-CT can be used to provide more detailed information of the airways and their correlations with physiologic function7–11.

The forced oscillation technique (FOT) can provide dynamic information regarding respiratory impedance (which comprises the resistance and reactance of the respiratory system)12–14. In patients with COPD, respiratory impedance measured by the FOT has demonstrated good correlation with spirometric data15–19 as well as with
symptoms\textsuperscript{20,21}. Moreover, respiratory impedance can be used to detect subtle changes in airway function in a manner that is even more sensitive than that seen for conventional spirometry\textsuperscript{21–25}.

However, how and whether structural changes in the lung parenchyma and airways in COPD correlate with respiratory impedance is not known. Respiratory resistance at 5 Hz (R5) and 20 Hz (R20) are supposed to represent total airway resistance and large-airway resistance, respectively\textsuperscript{12,14}. Respiratory reactance is considered to reflect the dynamic elastance and inertia of the respiratory system\textsuperscript{12,14}. However, these interpretations are based not on direct measurement of the lung structure but instead are derived from a physical model or circumstantial evidence from studies that have compared respiratory impedance with other conventional physiologic tests (including spirometry)\textsuperscript{12,14,26}. The association between respiratory impedance and imaging changes in the lungs could aid understanding of COPD pathology.

The aim of the present study was to evaluate potential correlations between respiratory impedance (assessed by the FOT) and airway structures (assessed by 3D-CT) in patients with COPD.

### Results

#### Patient characteristics

Ninety-eight patients with COPD and 49 reference subjects were enrolled (Table 1). Patients with COPD had a median age of 73 years and 92.9% were male. All COPD patients had a history of smoking (median pack-year of 47). The median (range) value for percentage predicted forced expiratory volume in one second (%predicted FEV\textsubscript{1}) for COPD patients was 64.1 (22.9–114.9)% (Table 2). The percentage of GOLD stages I, II, III and IV were 20.4, 53.1, 19.4, and 7.1, respectively. Sixty-seven (68.4%) patients received a long-acting muscarinic antagonist, 76 (77.6%) received a long-acting beta-2 agonist, and 26 subjects (26.5%) received an inhaled corticosteroid. Twenty-eight (28.6%) patients received single inhaler therapy, 63 matched population with age, sex, and smoking status. COPD, chronic obstructive pulmonary disease; BMI, body mass index; FVC, forced vital capacity; FEV\textsubscript{1}, forced expiratory volume in 1 second; MMF, maximum mid-expiratory flow rate; GOLD, Global Initiative for Asthma. LABA, long-acting beta 2 agonist; LAMA, long-acting muscarinic antagonist; ICS, inhaled corticosteroid; N.E., not estimated. *Wilcoxon signed-rank test unless otherwise indicated. ¶Fisher’s exact test.

#### Treatment

| Treatment | All population | Matched population\textsuperscript{1} |
|-----------|----------------|----------------------------------|
| LAMA      | 19 (19.4)      | 13 (18.6)                        |
| LABA      | 9 (9.2)        | 7 (10.0)                         |
| LAMA/LABA | 42 (42.9)      | 27 (38.6)                        |
| ICS/LABA  | 20 (20.4)      | 0 (0)                            |
| ICS/LAMA  | 1 (1.0)        | 1 (1.4)                          |
| ICS/LAMA/LABA | 5 (5.1) | 3 (4.3)                        |

#### Correlation between 3D-CT of lungs and physiologic tests

Among 98 patients with COPD, Spearman rank correlation test was performed (Tables 3, 4, and 5). The airway inner luminal area (Ai; mm\textsuperscript{2}) demonstrated significant correlations with %predicted forced vital capacity (FVC), %predicted FEV\textsubscript{1}, FEV\textsubscript{1}/FVC ratio, and %predicted maximum mid-expiratory flow rate (MMF, Table 3). Among spirometric measurements, %predicted FEV\textsubscript{1} had relatively stronger correlation with the Ai. Correlation coefficients between %predicted FEV\textsubscript{1} and Ai tended to increase gradually towards the distal bronchi. Statistical analysis using Meng-Rosenthal-Rubin method showed that correlation coefficients were significantly different between sixth-generation and third-generation bronchi (p = 0.043), and between sixth-generation and fourth-generation bronchi (p = 0.025) (Supplementary Table S1). As for respiratory impedance, the Ai demonstrated significant inverse correlations with R5, R20, the difference between R5 and R20 (R5–R20), resonant frequency (Fres), and low-frequency reactance area (ALX), and significant correlation with respiratory reactance at 5 Hz (X5).

### Table 1. Patient characteristics. Data are expressed as number (%) or median (range). \textsuperscript{1}Propensity score-matched population with age, sex, and smoking status. COPD, chronic obstructive pulmonary disease; BMI, body mass index; FVC, forced vital capacity; FEV\textsubscript{1}, forced expiratory volume in 1 second; MMF, maximum mid-expiratory flow rate; GOLD, Global Initiative for Asthma. LABA, long-acting beta 2 agonist; LAMA, long-acting muscarinic antagonist; ICS, inhaled corticosteroid; N.E., not estimated. *Wilcoxon signed-rank test unless otherwise indicated. ¶Fisher’s exact test.
### Table 2. Spirometric data. Data are expressed as median (range). 1Propensity score-matched population with age, sex, and smoking status. Abbreviations are as described in Table 1. *Wilcoxon signed-rank test.

| Spirometry | All population | Matched population^1 | COPD (n = 70) | Reference (n = 35) | p-value^* |
|------------|----------------|-----------------------|---------------|-------------------|-----------|
| FVC (L)    | 3.00 (1.14–5.43) | 3.24 (1.80–4.54) | 0.189 | 3.14 (1.14–5.43) | 3.36 (2.19–4.54) | 0.050 |
| FVC, % predicted | 93.1 (54.8–134.9) | 99.8 (80.3–174.7) | 0.009 | 92.6 (57.5–129.7) | 99.2 (80.3–121.4) | 0.034 |
| FEV₁ (L)   | 1.66 (0.45–3.39) | 2.40 (1.68–3.50) | <0.001 | 1.82 (0.54–3.39) | 2.65 (1.69–3.50) | <0.001 |
| FEV₁, % predicted | 64.1 (22.9–114.9) | 97.0 (80.0–163.8) | <0.001 | 65.4 (23.3–112.7) | 95.4 (80.0–110.6) | <0.001 |
| FEV₁/FVC (%) | 55.9 (26.6–69.9) | 77.7 (70.0–95.8) | <0.001 | 60.0 (26.6–69.9) | 76.7 (70.0–95.8) | <0.001 |
| MMF (L/S)  | 0.66 (0.17–7.10) | 2.08 (1.16–8.59) | <0.001 | 0.78 (0.18–7.10) | 2.2 (1.16–4.11) | <0.001 |
| MMF, %predicted | 23.1 (6.3–83.1) | 76.0 (44.4–135.9) | <0.001 | 28.3 (6.3–83.1) | 72.0 (44.4–135.9) | <0.001 |

### Table 3. Correlations between airway luminal areas and physiological function tests in patients with COPD. Data are expressed as Spearman rank correlation coefficient and p-value. R5, respiratory resistance at 5 Hz; R 20, respiratory resistance at 20 Hz; X5, respiratory reactance at 5 Hz; Fres, resonant frequency; ALX, low-frequency reactance area. The other abbreviations are as described in Table 1.

| Spirometry | 3rd-generation bronchi | 4th-generation bronchi | 5th-generation bronchi | 6th-generation bronchi |
|------------|-------------------------|------------------------|------------------------|------------------------|
| r          | r-p-value               | r                      | r                      | r                      |
| FVC, %predicted | 0.349                   | <0.001                 | 0.291                  | 0.005                  | 0.385                 | <0.001                  | 0.418                 | <0.001                  |
| FEV₁, %predicted | 0.375                   | <0.001                 | 0.392                  | <0.001                 | 0.474                 | <0.001                 | 0.548                 | <0.001                  |
| FEV₁/FVC ratio | 0.235                   | 0.024                  | 0.322                  | 0.002                  | 0.339                 | <0.001                 | 0.415                 | <0.001                  |
| MMF, %predicted | 0.321                   | 0.002                  | 0.368                  | <0.001                 | 0.423                 | <0.001                 | 0.497                 | <0.001                  |

### Table 4. Correlations between airway wall thickness and physiological function tests in patients with COPD. Data are expressed as Spearman rank correlation coefficient and p-value. The other abbreviations are as described in Tables 1 and 3.

| Spirometry | 3rd-generation bronchi | 4th-generation bronchi | 5th-generation bronchi | 6th-generation bronchi |
|------------|-------------------------|------------------------|------------------------|------------------------|
| r          | r-p-value               | r                      | r                      | r                      |
| FVC, %predicted | −0.019                  | 0.853                  | 0.020                  | 0.847                  | 0.053                  | 0.617                  | 0.055                  | 0.598                  |
| FEV₁, %predicted | −0.172                  | 0.098                  | −0.189                 | 0.070                  | −0.200                 | 0.054                  | −0.187                 | 0.073                  |
| FEV₁/FVC ratio | −0.269                  | 0.009                  | −0.320                 | 0.002                  | −0.326                 | 0.001                  | −0.323                 | 0.002                  |
| MMF, %predicted | −0.182                  | 0.081                  | −0.221                 | 0.033                  | −0.225                 | 0.030                  | −0.229                 | 0.027                  |

Wall thickness (WT; mm) demonstrated inverse correlations with FEV₁/FVC ratio and %predicted MMF (Table 4). As for respiratory impedance, WT demonstrated positive correlations with R5, R5-R20, Fres, and ALX. There were no correlations between Ai and WT within same generation (Supplementary Table S2). To exclude interdependence between Ai and WT, we performed partial correlation analyses, which demonstrated that Ai and WT were independently correlated with spirometric data and respiratory impedance (Supplementary Tables S3 and 4).
Percentage low-attenuation area (%LAA) demonstrated significant inverse correlation with the FEV1/FVC ratio, %predicted FEV1, and %predicted MMF (Table 5). As for respiratory impedance, the %LAA demonstrated significant correlation with indices of respiratory reactance (X5, Fres, and ALX), whereas correlation with indices of respiratory resistance was not observed.

In contrast, Ai and WT showed no significant correlations with physiologic parameters in 49 reference subjects (Supplementary Table S5).

Propensity score-matched analysis of physiologic tests and 3D-CT of lungs between patients with COPD and reference subjects.

Next, we compared physiologic tests and 3D-CT of lungs between patients with COPD and reference subjects. Because our reference subjects demonstrated a significantly younger age, higher proportion of females, and lower proportion of smokers, compared with patients with COPD (Table 1), we used propensity score matching. After propensity matching with age, sex, and smoking status as co-variables, the two groups had comparable characteristics, except that patients with COPD had significantly lower %predicted FEV1, FEV1/FVC ratio, and %predicted MMF (Table 2). Patients with COPD had significantly higher R5, R20, R5–R20, Fres, and ALX, and significantly lower X5, compared with reference subjects (Table 6). Regarding 3D-CT, patients with COPD had significantly decreased Ai throughout the third- to sixth-generation bronchi, and significantly higher %LAA (Table 7). There was no significant difference in WT between the two groups.

Discussion

In the present study, 3D-CT analyses of lungs revealed that respiratory impedance was correlated with airway narrowing, thickening of airway walls, and emphysema in patients with COPD. Airway narrowing was correlated with respiratory resistance and reactance. Among spirometric parameters, %predicted FEV1 showed a significant correlation with airway narrowing, which indicated a tendency of a gradual increase towards distal bronchi. The FEV1/FVC ratio, R5–R20, and respiratory reactance demonstrated correlations with the WT. Increased %LAA was correlated with the FEV1/FVC ratio and indices of respiratory reactance, but not with indices of respiratory resistance. The respiratory impedance as well as spirometric data may reflect different components of pathogenic changes in COPD, and could be used as complementary approaches for COPD assessment.

Respiratory resistance is considered to reflect the forward pressure of conducting airways 12,27, and so is interpreted as an index of airway caliber 14. The correlation between respiratory resistance and airway narrowing observed in the present study validates this theoretical model. Moreover, serial assessment of airways

| Spirometry | r   | p-value |
|------------|-----|---------|
| FVC, %predicted | −0.153 | 0.133 |
| FEV1, %predicted | −0.464 | <0.001 |
| FEV1/FVC ratio | −0.580 | <0.001 |
| MMF, %predicted | −0.453 | <0.001 |

| Respiratory impedance, whole-breath | r   | p-value |
|------------------------------------|-----|---------|
| R5 | 0.082 | 0.421 |
| R20 | 0.026 | 0.802 |
| R5–R20 | 0.182 | 0.073 |
| X5 | −0.276 | 0.006 |
| Fres | 0.304 | 0.002 |
| ALX | 0.255 | 0.011 |

Table 5. Correlations between percentage of low attenuation area and physiological function tests in patients with COPD. Data are expressed as Spearman rank correlation coefficient and p-value. %LAA, percentage of low attenuation area < −950 HU; The other abbreviations are as described in Tables 1 and 3.

| Respiratory impedance at whole-breath | COPD (n = 98) | Reference (n = 49) | p-value* | COPD (n = 70) | Reference (n = 35) | p-value* |
|--------------------------------------|---------------|--------------------|----------|---------------|--------------------|----------|
| R5 (cmH2O/L/s) | 3.17 (0.52–6.38) | 2.46 (0.97–4.96) | <0.001 | 3.11 (0.52–6.38) | 1.86 (0.97–4.96) | <0.001 |
| R20 (cmH2O/L/s) | 2.51 (0.53–4.55) | 2.11 (1.02–4.60) | 0.004 | 2.45 (0.53–4.55) | 1.83 (1.02–4.60) | 0.002 |
| R5–R20 (cmH2O/L/s) | 0.66 (−0.63–2.14) | 0.37 (−0.52–1.86) | <0.001 | 0.53 (−0.63–1.88) | 0.23 (−0.52–1.86) | 0.001 |
| X5 (cmH2O/L/s) | −0.72 (−5.35–0.37) | −0.33 (−2.18–0.24) | <0.001 | −0.59 (−5.35–0.37) | −0.31 (−2.18–0.24) | 0.005 |
| Fres (Hz) | 11.27 (4.00–30.17) | 7.38 (4.16–16.04) | <0.001 | 10.71 (4.00–30.17) | 7.31 (4.16–16.04) | <0.001 |
| ALX (cmH2O/L/s × Hz) | 3.76 (0–51.26) | 1.14 (0.02–14.59) | <0.001 | 2.66 (0–51.26) | 1.06 (0.02–14.59) | 0.002 |

Table 6. Respiratory impedance at whole-breath. Data are expressed as median (range). *Propensity score-matched population with age, sex, and smoking status. The other abbreviations are as described in Table 3. *Wilcoxon signed-rank test.
reactance is considered to be a marker of flow limitation. The correlation between respiratory reactance and caliber collapses in the expiratory phase because of low elastic recoil pressure due to parenchymal destruction at full-inspiration could not account fully for dynamic changes in the lungs during tidal breathing. Airway reduction in apparent respiratory compliance and a decrease in respiratory reactance. Therefore, respiratory signals cannot pass through flow-limiting segments (“choke points”) and reach alveoli, thereby eliciting marked.

High-frequency waves reflect back from large airways and low-frequency waves travel deep into the lungs. In the present study, R5 and R20 demonstrated almost similar correlations with airway narrowing in third- to sixth-generation bronchi assessed by 3D-CT.

Airway narrowing was also correlated with respiratory reactance. Usually, respiratory reactance reflects the elastic and inertial properties of the respiratory system. However, if flow limitation is present, oscillatory signals cannot pass through flow-limiting segments (“choke points”) and reach alveoli, thereby eliciting marked reduction in apparent respiratory compliance and a decrease in respiratory reactance. Therefore, respiratory reactance is considered to be a marker of flow limitation. The correlation between respiratory reactance and airway narrowing observed in the present study may reflect flow limitation caused by obstruction of peripheral airways. It has been proposed that respiratory reactance reflects small airway function, so detailed information about more distal airways may be necessary for further understanding of respiratory impedance.

In the present study, thickening of airway walls was correlated with respiratory impedance. The correlations between WT and impedance were independent of Ai, and vice versa, according to the partial correlation analysis. Peribronchial fibrosis and consequent remodeling of airway tissue associated with COPD progression reduces airway compliance, which may result in a decrease in respiratory reactance. The correlation between thickening of airways walls and respiratory reactance observed in the present study was compatible with this hypothesis. Airway fibrosis may also affect tissue resistance (a component of respiratory reactance) and contribute to the correlation between thickening of airway walls and respiratory reactance. The increased %LAA was correlated with indices of respiratory reactance. Emphysema is one of the key pathologic characteristics of COPD, which reflects parenchymal destruction.

The increased %LAA was correlated with indices of respiratory reactance. Emphysema is one of the key pathologic characteristics of COPD, which reflects parenchymal destruction. Emphysema displays a patchy distribution and causes heterogeneous airway collapse, which may be associated with respiratory reactance denoting non-uniform ventilation.

The present study had several limitations. First, respiratory impedance cannot be explained entirely by airway structure alone. Respiratory resistance includes tissue resistance and chest-wall resistance (though airway resistance is the major part). Secretion within the airway luminal area (a key component of airway obstruction in COPD) could not be evaluated in our study. These potentially influential factors (other than structural features of airways) should be considered for interpretation of respiratory impedance. Second, the limited resolution of CT could not aid evaluation of peripheral airways more distal than sixth-generation bronchi. Major sites of airway obstruction in COPD are small airways < 2 mm in diameter. We evaluated third- to sixth-generation bronchi, but more distal airways (e.g. terminal bronchioles) are also important in COPD. Third, static CT images obtained at full-inspiration could not account fully for dynamic changes in the lungs during tidal breathing. Airway caliber collapses in the expiratory phase because of the low elastic recoil pressure due to parenchymal destruction (emphysema). Time-series analyses of airways could provide further insights into respiratory impedance.

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### Table 7. Three-dimensional CT imaging of the lungs.

|            | All population | Matched population† |
|------------|----------------|---------------------|
| Ai (mm²)   | COPD (n = 98)  | Reference (n = 49)  | p-value* | COPD (n = 70)  | Reference (n = 35) | p-value* |
| 3rd-generation bronchi | 35.9 (18.2–76.8) | 41.2 (18.2–76.8) | 0.007 | 35.7 (18.2–76.8) | 41.7 (23.2–76.8) | 0.002 |
| 4th-generation bronchi | 9.8 (3.7–20.4)  | 11.1 (5.5–25.1)  | 0.007 | 9.8 (4.6–18.2)  | 11.5 (6.1–25.1) | 0.005 |
| 5th-generation bronchi | 5.8 (2.4–11.7) | 7.1 (3.0–12.0)   | <0.001 | 5.8 (2.6–11.6) | 7.1 (3.8–12.0) | <0.001 |
| 6th-generation bronchi | 3.3 (0.9–7.9)  | 4.3 (1.7–7.9)   | <0.001 | 3.2 (0.9–6.9)  | 4.2 (2.3–7.9) | <0.001 |
| WT (mm)    | COPD (n = 98)  | Reference (n = 49)  | p-value* | COPD (n = 70)  | Reference (n = 35) | p-value* |
| 3rd-generation bronchi | 1.6 (1.2–2.0)  | 1.5 (1.3–1.9)   | 0.005 | 1.6 (1.2–1.9)  | 1.5 (1.3–1.8) | 0.268 |
| 4th-generation bronchi | 1.4 (1.1–1.8)  | 1.3 (1.0–1.5)   | 0.010 | 1.4 (1.1–1.6)  | 1.3 (1.2–1.6) | 0.126 |
| 5th-generation bronchi | 1.2 (0.9–1.5)  | 1.2 (1.0–1.4)   | 0.085 | 1.2 (0.9–1.5)  | 1.2 (1.1–1.4) | 0.407 |
| 6th-generation bronchi | 1.1 (0.9–1.3)  | 1.1 (0.9–1.3)   | 0.194 | 1.1 (0.9–1.3)  | 1.1 (0.9–1.3) | 0.679 |
| %LAA       | COPD (n = 98)  | Reference (n = 49)  | p-value* | COPD (n = 70)  | Reference (n = 35) | p-value* |
| 4th-generation bronchi | 41.4 (17.1–64.2) | 29.6 (11.2–51.6) | <0.001 | 40.9 (20.7–64.2) | 31.5 (14.6–51.6) | <0.001 |

*Wilcoxon signed-rank test.

The increased %LAA was correlated with indices of respiratory reactance. Emphysema is one of the key pathologic characteristics of COPD, which reflects parenchymal destruction. Emphysema displays a patchy distribution and causes heterogeneous airway collapse, which may be associated with respiratory reactance denoting non-uniform ventilation.

Provided new insights regarding interpretation of respiratory reactance according to the proximal–distal axis. High-frequency waves reflect back from large airways and low-frequency waves travel deep into the lungs. Therefore, it has been hypothesized that R5 denotes total airway resistance and R20 denotes central airways. In the present study, R5 and R20 demonstrated almost similar correlations with airway narrowing in third- to sixth-generation bronchi assessed by 3D-CT.

Airway narrowing was also correlated with respiratory reactance. Usually, respiratory reactance reflects the elastic and inertial properties of the respiratory system. However, if flow limitation is present, oscillatory signals cannot pass through flow-limiting segments (“choke points”) and reach alveoli, thereby eliciting marked reduction in apparent respiratory compliance and a decrease in respiratory reactance. Therefore, respiratory reactance is considered to be a marker of flow limitation. The correlation between respiratory reactance and airway narrowing observed in the present study may reflect flow limitation caused by obstruction of peripheral airways. It has been proposed that respiratory reactance reflects small airway function, so detailed information about more distal airways may be necessary for further understanding of respiratory impedance.

In the present study, thickening of airway walls was correlated with respiratory impedance. The correlations between WT and impedance were independent of Ai, and vice versa, according to the partial correlation analysis. Peribronchial fibrosis and consequent remodeling of airway tissue associated with COPD progression reduces airway compliance, which may result in a decrease in respiratory reactance. The correlation between thickening of airways walls and respiratory reactance observed in the present study was compatible with this hypothesis. Airway fibrosis may also affect tissue resistance (a component of respiratory reactance) and contribute to the correlation between thickening of airway walls and respiratory reactance. R5–R20 is considered to reflect non-uniform distribution of ventilation and airway obstruction. In the present study, thickening of airways walls could represent small airway function, so detailed information about more distal airways may be necessary for further understanding of respiratory impedance.

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The present study had several limitations. First, respiratory impedance cannot be explained entirely by airway structure alone. Respiratory reactance includes tissue reactance and chest-wall reactance (though airway resistance is the major part). Secretion within the airway luminal area (a key component of airway obstruction in COPD) could not be evaluated in our study. These potentially influential factors (other than structural features of airways) should be considered for interpretation of respiratory impedance. Second, the limited resolution of CT could not aid evaluation of peripheral airways more distal than sixth-generation bronchi. Major sites of airway obstruction in COPD are small airways < 2 mm in diameter. We evaluated third- to sixth-generation bronchi, but more distal airways (e.g. terminal bronchioles) are also important in COPD. Third, static CT images obtained at full-inspiration could not account fully for dynamic changes in the lungs during tidal breathing. Airway caliber collapses in the expiratory phase because of the low elastic recoil pressure due to parenchymal destruction (emphysema). Time-series analyses of airways could provide further insights into respiratory impedance. Finally, our reference subjects demonstrated a significantly younger age and higher proportion of females. Although the proportion of smoker in the reference subjects was lower than that in COPD patients, relatively few were non-smokers and the majority was smoker, all which may have affected the results. Comparison of patients with COPD and non-smoking reference subjects (only 14 subjects) showed that COPD patients had thicker walls, narrower airways, and higher respiratory impedance, although some of the differences were not significant. In the present study, thickening of airway walls was correlated with respiratory reactance. The correlations between thickening of airway walls and respiratory reactance observed in the present study was compatible with this hypothesis.
study, we used propensity-score matching analysis to minimize the potential influence of heterogeneity among the reference group. After propensity matching with age, sex, and smoking status as co-variables, the two groups had comparable characteristics.

Respiratory impedance was correlated with airway narrowing, thickening of airway walls, and emphysema in patients with COPD. Complementary use of FOT and spirometry may be useful for evaluation of pathogenic changes in COPD.

Methods

Study design. This was a prospective observational study conducted in accordance with the ethical standards described in the Declaration of Helsinki. The study protocol was approved by the Institutional Review Board of Hamamatsu University School of Medicine (Hamamatsu, Japan). Each patient provided written informed consent to be included in the study. The study was registered with the University Hospital Medical Information Network Clinical Trial Registry (identification code: 000013541).

Patient eligibility. Patients who satisfied the definition of COPD set by GOLD were enrolled. Exclusion criteria were: the requirement of treatment change, respiratory-tract infection, and COPD exacerbation 4 weeks before study commencement; long-term oxygen therapy; bronchial asthma; diffuse lung diseases; neuromuscular diseases; congenital anomalies of the bronchial tree; history of thoracic surgery. Subjects without COPD were enrolled as a reference population and were required to: (i) have normal pulmonary function as assessed by spirometry; (ii) not meet the exclusion criteria described above. Propensity score-matched analysis was performed to minimize biases between patients with COPD and reference subjects in one-to-two matching using age, sex, and smoking status as co-variables.

Multi-detector-row computed tomography (MDCT). MDCT was done in the supine position at full-inspiration breath-hold using a 64-slice MDCT scanner (Aquilion-64; Toshiba Medical Systems, Tokyo, Japan). Scanning parameters were: collimation, 64 × 0.5 mm; tube voltage, 120 kV; tube current, 200 mA; rotation time, 0.5 s; pitch, 0.83. These parameters were varied by the scanner system to obtain the optimum radiation dose and image quality. Images were reconstructed using a standard reconstruction algorithm (FC50) for the lung using a slice thickness of 0.5 mm and reconstruction interval of 0.5 mm.

Three-dimensional (3D) CT analysis of lungs. Using a reconstructed 3D-CT method, we obtained images of airways with image-analyzing software (SYNAPSE VINCENT; Fuji Film, Tokyo, Japan). The bronchial

Figure 1. Three-dimensional CT analysis of the airway. (A) The image of the airway tree reconstructed by three-dimensional computed tomography. (B) The longitudinal section of the reformatted airway in the multiplanar reconstruction image. (C) Measurement of the airway in the short axis image. Red and green lines indicate the outer and inner edge of the airway, respectively.
pathway was reconstructed automatically into multiplanar reconstruction images with a window width of 1600 HU and a window level of −600 HU (Fig. 1). Measurements of airways were done in four ways according to methods reported previously11. First, six segmental bronchi (B1, B2, B3, B8, B9, and B10) in the right lung were selected. Middle-lobe bronchi as well as B6 and B7 were excluded from analyses because they are readily associated with atelectasis, airway collapse and mucus hypersecretion. The left lung was also excluded from analyses to avoid artifacts due to transmitted cardiac motion. Second, in each selected bronchus, four levels of the airways (third- (segmental), fourth- (sub-segmental), fifth-, and sixth-generation bronchi) were identified by tracing airway trees peripherally on the same trunk of each segmental bronchus. Third, in the midpoint of each level of airway, WT and Ai perpendicular to the long axis of the airway were computed automatically (Fig. 1). Airway center line was constructed by using minimum spanning tree, and inner and outer airway contours were extracted by using graph cuts method. WT was calculated as the mean distance of the outer to inner edge of the airway assuming that it is a “true” circle. The inner and outer airway contours were extracted automatically by the image-analysis software. If an error occurred during automatic extraction of airway contours, then it was corrected by manual means. Fourth, six images were evaluated at each level of the bronchus, and the results for bronchi from each generation were expressed separately as the means of six airways. The %LAA, defined by the percentage of area below −950 HU in total lung area, was calculated using SYNAPSE VINCENT.

Measurement of spirometry and respiratory impedance. Spirometry and the FOT were performed at the same day of the chest CT scans. The FOT was performed before spirometry to avoid influences of forced breathing. Short-acting β2-agonists were not withheld for at least 12 hours before these tests. Autospirometer System 7 (Minato Medical Science Co., Ltd., Osaka, Japan) was used to measure spirometry according to the standards of the Japanese Respiratory society25. Respiratory impedance was measured using a commercial FOT device (Most-Graph 01; Chest MI, Tokyo, Japan) according to standard recommendations as reported previously17,27. Briefly, impulse oscillatory signals generated by a loudspeaker at intervals of 0.25 s were applied to the respiratory system through the mouthpiece during tidal breathing. Mouth pressure and flow signals were measured and calculated to obtain the resistance and reactance properties against oscillatory frequencies from 4–36 Hz. We evaluated R5, R20, R5–R20, X5, Fres (resonant frequency; where the reactance crosses zero and the airflow limitation is maximal). Data are the median (range) unless indicated otherwise. Statistical tests were two-sided, and p < 0.05 was considered significant. Values were analyzed using JMP v9.0.0 (SAS Institute Japan, Tokyo, Japan), except for partial correlation analyses and Meng-Rosenthal-Rubin method using R version 3.2.4 (The R Foundation for Statistical Computing, Vienna, Austria, 2016).

Statistical analyses. The Wilcoxon signed rank test was used for continuous variables and Fisher’s exact test for categorical groups. Correlation between continuous variables was undertaken using the Spearman rank correlation coefficient. Partial correlation analyses were performed to exclude the potential confounding between Ai and WT. Difference between correlation coefficients was evaluated by Meng-Rosenthal-Rubin method. Propensity score-matched analysis was performed using EZR (Saitama Medical Center, Jichi Medical University, Saitama, Japan), which is a graphical user interface for R (The R Foundation for Statistical Computing, Vienna, Austria).

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