Physicochemical Properties of Low-Fat Yoghurt with Whey Protein Isolates as Fat Alternative

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Abstract: In this research, the effect of whey protein isolates (WPI) as a fat alternative on the physicochemical properties of low-fat (1-2 %) yoghurt samples during 14 days of storage was determined. The samples were analyzed for their chemical composition, syneresis, tyrosine, firmness, color, and free fatty acids. Yoghurt having 2 % of WPI showed significantly higher amount of tyrosine but lower syneresis; as total solids, protein, and fat were higher than the low-fat yoghurts (1% WPI and low-fat control, p < 0.05). However, WPI addition decreased the white and green tones but increased the yellow; thus, the addition of WPI didn’t affect the opacity and brightness of low-fat yoghurt. The addition of WPI also gave rise to the amounts butyric, capric, and oleic acids during storage (p < 0.05). Herein we propose 2 % WPI as a fat alternative to improve the physicochemical properties of low-fat for a storage duration of 14 days.

Keywords: low-fat yoghurt; whey protein isolates; physicochemical properties; free fatty acids; fat replacer

INTRODUCTION

The market for functional and convenience food is widening day-by-day as the public is getting aware of healthy diets (Lasik et al., 2016). Yoghurt has gained considerable economic importance as it is healthy and nutritious (Guggisberg et al., 2009). It is a type of miscellaneous, lactic acid–fermented milk product which suits all palates and meal occasions. In Turkey, it is frequently consumed either as a sole meal or as supplement. Turkish yoghurt production has been increased up to 4.5 % from 2015 to 2016 i.e. 1173 million tons that makes a 25.2 % market share of the dairy products. The average yoghurt consumption/person in Turkey is about 30 kg/year that is gradually increasing (Anonymous, 2016; Isleten and Karagül-Yuceer, 2006).

Fat plays an important role in the structural integrity and mouth feeling of yoghurt as it interacts with casein micelles. Less amount of fat in yoghurt leads to increased syneresis, poor texture, weak body, undesirable taste, and lower total solid content. Using whey proteins as a fat alternative in yoghurts could be a good practice due to their good nutritional and functional properties. Recently, the use of heat-treated whey protein concentrate (HPWC) as a fat alternative in non-fat goat milk yoghurt has suggested it as a possible fat alternative to improve the consistency of non-fat yoghurt. However, while the main constituent i.e. β-lactoglobulin is the same in WPI and WPC, WPI has a higher-purity whey protein then WPC. Also, WPI is high in calcium, minerals, and branched-chain amino acids such as valine, leucine, and isoleucine (Torres et al., 2018; Zhang et al., 2015).

These additional properties may increase the textural quality of yoghurt i.e. firmness, creaminess, viscosity, along with its health benefits and functional properties (Torres et al., 2018; Zhang et al., 2015). However, very limited work has been done on WPI usage as a fat alternative in yoghurt production. According to Torres et al. (2018), the addition of whey protein microparticles (MPWP) improved texture and rheological properties. But using MPWPs with a non-microparticulated source as WPI resulted in low-viscosity yoghurt. Zhang et al. (2015) suggested that WPC could be used as a fat replacer for goat milk yoghurt production to improve the texture and water holding capacity of yoghurt. More work is needed to explore WPI usage in reduced and whole-fat yoghurt production during storage.

Since free fatty acids (FFA) variation gives rise to changes in the organoleptic and nutritional properties of dairy products, they are important to be evaluated in yoghurt. Lipolysis occurs during storage where the FFAs are liberated

Sorumlu Yazar: demet.karaman@adu.edu.tr
Geliş Tarihi: 15 Temmuz 2019
Kabul Tarihi: 17 Aralık 2019
and short chain fatty acids (SCFA) provides the sensory quality to the dairy products. Acetic, butyric, and oleic acids show antibiotic and anticancer properties that are important nutritional aspects of fatty acids (ReguŁa 2007). FFAs in yoghurt are influenced by the type of starter culture, quality of raw milk, and technological treatments such as incubation, cooling, and storage (Güler and Gürsoy-Balci 2011). Furthermore, milk protein usage in yoghurt as a fat replacer could be an effective FFA variation during storage. To the best of our knowledge, there is no report on this subject. Although, a number of authors have studied the free fatty acids in yoghurt (Güler 2007; Güler and Gürsoy-Balci 2011; Sumarmono et al., 2015) or the structural and sensory properties of yoghurt with WPI (Guggisberg, 2009; Guggisberg et al., 2007; Ibrahim et al., 2017; Matumoto-Pintro et al., 2011; Onsekiizoglu Bagci and Gunasekaran 2016; Patocka et al., 2006; Shi et al.,2017; Walsh-O’grady et al., 2001; Wang et al., 2015; Zhang et al., 2015), scientific literature characterizing FFAs in WPI-added yoghurt is rare.

This study was aimed to: i) determine the variations that may develop in low-fat yoghurt’s physicochemical properties by WPI addition during storage, ii) understand whether or not the FFA profiles of yoghurt were related to WPI addition, and iii) to investigate a novel approach to developing yoghurt using WPI as a fat replacer.

MATERIALS AND METHODS

Materials

Raw cow’s milk for analysis was bulk collected from Omur Sut Mam. Ldt. Sti (Aydın, Turkey). Whey protein isolate (WPI) was obtained from Danisco Food International (Turker Teknik Company, İstanbul, Turkey), with an approximate composition of 96 % protein and 2 % total fat, as stated by the manufacturer. A freeze-dried direct vat set thermophilic yoghurt culture (Yoflex: Express 1.0), composed of a mixture of Lactobacillus delbrueckii subsp. bulgaricus and Streptococcus thermophilus, was kindly provided by Maysa Starter Culture Company (İstanbul, Turkey).

Methods

Yoghurt production

Fatty raw milk was preheated to 55 °C and standardized at 3 % (fatty) and 0.5 % (reduced fat) fat levels. After being heated to 95 °C for 10 min, the milk was cooled to 45 °C. At this stage, WPI was added at different levels (1 and 2 % w/w) into the reduced-fat milk samples. Thus, four yoghurt samples were prepared as follows: 3 % fat (Control, fatty yoghurt, FY), 0.5 % fat (low-fat yoghurt, LY), 0.5 % fat + 1 % WPI (low-fat yoghurt with 1% WPI, LY1), and 0.5 % fat + 2 % WPI (low-fat yoghurt with 2 % WPI, LY2). After mixing in a blender, commercial yoghurt culture was added at a concentration of 3 % (after pre-activation). The inoculated milk samples were poured into 250 g plastic cups with lids and incubated at 43–45 °C. Incubation was ended when the experimental yoghurt samples reached pH 4.6–4.7. The fermentation times of all 4 types of set yoghurt were approximately 4 h. Following incubation, yoghurt samples were cooled and stored at 3-5 °C for 14 days (Figure 1). Sample production and all the analyses were performed in triplicate.

Figure 1. Flowchart illustrating the production of yogurts.

FY: Fatty yogurt (% 3 fat+% 0 WPI); LY: Low fat yogurt (≤ % 0.5 fat+% 0 WPI); LY1: 1% WPI added yoghurt (≤ % 0.5 fat+1 WPI); LY2: 2% WPI added yoghurt (≤ % 0.5 fat +2% WPI).

Some physicochemical properties of milk samples

The basic chemical composition of raw milk (total solid and fat) was determined by gravimetric and Gerber methods (Anonymous 1994). Protein contents were analyzed according to the Kjeldahl method (AOAC 2010). Milk pH was measured using a pH meter (Adwa, Romania) with a combined glass electrode.

Yogurths analyses

The total solid and fat contents of the yoghurt samples were determined by the standard Turkish methods (Anonymous 1989). The titratable acidity (TA, %) and ash content were detected according to AOAC standards (AOAC 2000, 2010). Total nitrogen was determined by the Kjeldahl method and protein content was calculated using a
conversion factor of 6.38. Tyrosine value and syneresis were measured according to Hull (1947) and Guggisberg et al. (2011) respectively. The firmness (F35 mm) of yoghurt samples was measured using a Universal Testing Machine equipped with a 500 N force sensor (Zwick/Roel Z.05 TH, Zwick, Germany) and a cylinder (h=12.5 cm, Ø=6 cm) (Guggisberg et al., 2011). A Hunter Lab Color Flex EZ spectrophotometer (S/N CFEZ 1209 Model, Hunter Associates Laboratory, Inc., Reston, VA, USA) was used to measure the yoghurt’s whiteness (L), greenness (a) and yellowness (b). All L, a, and b values were taken per single sample in triplicate at different sites, and the average was calculated. Free fatty acids were analyzed using an Agilent GC (model GC 6890N) equipped with a capillary column (300 x 250µm x 0.25µm, Agilent 19091F-433 HP-FFAP, CA, USA). The extraction was obtained according to reported method (Yıldız-Akgül 2018). The gas chromatography (GC) injection volume was 2 mL while the temperature of the GC oven was increased from 120 to 230 °C at a rate of 10 °C/min. The split was set at 1:10. Fatty acid standards supplied as samples (Sigma-Aldrich, Germany) were prepared at 50, 100, 150, 200, and 250 ppm and injected for free fatty acid identification.

**Statistical analysis**

The data were analyzed using the SPSS (Version 18.0, SPSS Inc., USA) commercial statistical package. A critical level of significance at p=0.05 was used throughout the study. Analysis of variance was performed on each attribute and treatment by 14 days of storage interactions. Any significant treatment, time, or interaction effect was calculated. Pearson’s correlation coefficient test was used for multiple comparisons.

**RESULTS AND DISCUSSION**

**Physicochemical characteristics of milk**

The physicochemical composition of raw cow’s milk was found as follows: fat (3.87 ± 0.64 %), total solids (12.42 ± 0.77 %), protein (3.28 ± 0.03 %), ash (0.68 ± 0.02 %), pH (6.62 ± 0.09), and density (1.030 ± 0.01 g/mL). The gross composition of the raw milk was in accordance with the Turkish Codex’s Standard for Raw Milk (Anonymous 1994). Physicochemical characteristics of yoghurt during storage

| Samples* | Total solids (%) | Fat (%) | Protein (%) | Ash (%) |
|----------|------------------|---------|-------------|---------|
| FY       | 11.09±0.17       | 3.00±0.00 | 2.86±0.15  | 0.69±0.00 |
| LY       | 9.42±0.07        | 0.60±0.00 | 3.33±0.33  | 0.70±0.00 |
| LY1      | 10.36±0.07       | 0.60±0.00 | 4.25±0.19  | 0.71±0.00 |
| LY2      | 11.74±0.13       | 0.70±0.00 | 5.39±0.30  | 0.72±0.00 |

*FY: Fatty yogurt (% 3 fat%+% 0 WPI); LY: Low fat yogurt (≤ % 0.5 fat%+% 0 WPI); LY1: 1% WPI added yogurt (≤ % 0.5 fat%+% 1 WPI); LY2: 2% WPI added yogurt (≤ % 0.5 fat%+% 2 WPI).

Superscript lowercase letters means significant difference (p<0.05).
storage as a result of the accumulation of lactic acid produced by the bacteria similar to the related fermented foods (Damin et al., 2009). Tyrosine detection tracks released α-amino groups due to proteolysis of the milk proteins, that indicates the proteolytic activity of the starter culture. Exceeding from 0.5 mg/mL can cause certain flavor defects, such as bitterness, depending on the storage time, culture type, and protein structure in the yoghurt (Kesenkaş et al., 2011). In this study, the tyrosine levels of the yoghurt samples during the 14-day storage period were observed in the range of 0.32-0.47 mL/5g. The tyrosine value of the yoghurt samples did not reach a certain threshold value; thus the flavor of the yoghurt maintains at the end of the storage.

Similarly, storage time and WPI addition significantly affected the tyrosine values in this study (p < 0.05). Tyrosine levels in the WPI-containing yoghurts (LY1 and LY2) were significantly higher (p < 0.05) as compared to the controls (FY and LY) at the beginning and end of the storage period (Table 2). This can be related to the higher total solids content of the samples LY1 and LY2 vs. the controls (Table 1). The amount of tyrosine in all yoghurt samples was higher at the end of the storage than on day 1 with significant fluctuations over time (p < 0.05). Similar results were obtained by Şenel et al. (2011).

Texture is one of the main factors affecting yoghurt quality. Poor texture and syneresis reduce its appeal to consumers (Lasik et al., 2016). Figure 2 shows the changes in syneresis over 14 days of storage. The syneresis was influenced by WPI addition and fat content (p < 0.05). No effect of storage was detected (p > 0.05). The 2 % WPI added yoghurt (sample LY2) had the lowest level of syneresis at the beginning and at the end of the storage. High amount of total solids (11.74 %, Table 1) in LY2 partially responsible for the low syneresis observed during the whole storage period (4.08 and 4.75, Figure 2). On the other hand, sample LY (low-fat) consistently displayed higher syneresis (between 6.92 and 6.50) compared with the other yoghurts (p < 0.05), which could be related to the low total solids (9.42 %), as well as the least fat content (0.6 %) (Table 1). Such behavior can be explained by their higher solids and fat concentration; thus the possibility of building a more cohesive polymer network to reduce the syneresis. This observation is in agreement with the results of (Kaminarides et al., 2007) comparing the yoghurts containing 6.6 and 0.9 % fat content. Since pH decreases during storage (Table 2), unexpectedly, there were abrupt ups and downs in syneresis; although these changes (throughout the storage) were statistically insignificant (p >

Table 2. Physicochemical compositions of yoghurt samples during storage for 14 days (n=3)

| Parameter* | Samples | Day 1 | Day 7 | Day 14 | Mean /Average |
|------------|---------|-------|-------|--------|---------------|
| pH         | FY      | 4.25±0.02cA | 4.20±0.03bA | 4.18±0.03aA | 4.21±0.04A |
|            | LY      | 4.26±0.01cB | 4.23±0.02bB | 4.21±0.02bB | 4.23±0.03B |
|            | LY1     | 4.34±0.04cC | 4.29±0.02bC | 4.28±0.02aC | 4.30±0.04C |
|            | LY2     | 4.42±0.03cD | 4.28±0.02bD | 4.26±0.02aD | 4.32±0.08D |
|           | FY      | 0.76±0.02aA | 0.79±0.02bA | 0.85±0.00cA | 0.80±0.04A |
| Titratable | LY      | 0.79±0.03aB | 0.79±0.02bB | 0.85±0.01cB | 0.81±0.03B |
| acidity, LA | LY1     | 0.85±0.01aC | 0.85±0.01bC | 0.93±0.04cC | 0.87±0.05C |
|            | LY2     | 0.92±0.03aD | 0.96±0.02bD | 1.03±0.01cD | 0.97±0.05D |
|           | FY      | 0.32±0.01aA | 0.36±0.01bA | 0.37±0.01cA | 0.35±0.02A |
| Tyrosine,  | LY      | 0.35±0.01aB | 0.37±0.01bB | 0.37±0.01cB | 0.36±0.01B |
| mg/5 g     | LY1     | 0.44±0.01aC | 0.45±0.01bC | 0.45±0.01cC | 0.45±0.01C |
|            | LY2     | 0.44±0.02aC | 0.44±0.02bC | 0.47±0.02cC | 0.45±0.03C |

* FY: Fatty yoghurt (% 3 fat+% 0 WPI); LY: Low fat yoghurt (% ≤ 0.5 fat+% 0 WPI); LY1: 1% WPI added yoghurt (% ≤ 0.5 fat+1 % WPI); LY2: 2% WPI added yoghurt (% ≤ 0.5 fat+2 % WPI).

a,b,c The same column with different superscripts among yoghurt samples significantly differ (P<0.05)
A,B,C,D The same row with different superscripts among yoghurt samples significantly differ (p<0.05)
Increases (Onsekizoglu Bagci and Gunasekaran, 2016) and decreases (Isleten and Karagul-Yuceer, 2006; Matumoto-Pintro et al., 2011) in the syneresis of WPI-added yoghurts during storage have also been reported in previous studies. The results of textural examinations of firmness are presented in Figure 3. The firmness of the control fatty yoghurt (FY) was significantly (p < 0.05) higher than the others at the beginning of the storage. This could be related to the firmness of yoghurts increasing as the fat content increased. The lower firmness of sample LY2 (2% WPI added), LY1 and LY can be an outcome of the lower syneresis (Figure 2) and higher total solids (Table 1) as observed previously (Kaminarides et al., 2007; Wang et al., 2015). Similarly, the firmness changed during storage (p < 0.05). The firmness values of sample FY (fatty) control yoghurts decreased but increased for other samples at the end of the storage. Yoghurt color is another powerful quality descriptor and consumer acceptance quality attribute. The color values depend on the type of milk used in the yoghurt-making, the chemical composition of the yoghurt, and the yoghurt-making technique since the gel opacity is related to fat content, casein ratio, and the aggregation level of fat and casein (Güler and Park 2011).

On the first day of storage, the color of the yoghurt samples was significantly influenced (p < 0.05) by WPI addition (Figure 4). The L (whiteness) values of WPI-containing yoghurt (LY1-LY2) were significantly lower (p < 0.05) than the low-fat and fatty control yoghurts (LY, FY). However, during storage, the L value decreased for the low-fat and WPI-containing yoghurts (LY, LY1, LY2) but increased for the sample (fatty) control yoghurt (FY). The highest fat level of FY yoghurt may have increased the whiteness of the yoghurt.

All the yoghurt samples showed a negative value, indicating greenness. WPI addition contributed more to the green color compared to the low-fat control yoghurt (without WPI). However, alteration was observed after 7 days of storage. At day 1 of storage, WPI addition implied more yellowing (higher b value) than the control yoghurt. Further increases in the b values were statistically insignificant for any of the yoghurt samples at the end of the storage (p > 0.05).
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0.05). So, WPI addition was characterized by significantly lower whiteness, lower greenness, and higher yellowness than the fatty control yoghurts (FY) possibly due to the presence of more fat globules and lower protein levels (Table 1) than other samples at day 1. At the end of the storage, WPI-containing yoghurts (LY1, LY2) had higher greenness (a) but less yellowness (b) and whiteness (L) than sample LY (low fat); that means opacity and higher brightness wasn’t observed with WPI addition (Figure 4). However, there were no statistically significant differences between the first and last day of storage on the yellowness (b) of all samples (p > 0.05). Yıldız-Akgül 2018 studied the color change of WPI-added Torba yoghurt during 14 days of storage. The author observed a decrease in whiteness (L) and greenness (a) but an increase in yellowness (b) during storage. Fermented beverages prepared from milk with an increased proportion of whey proteins to caseins were also observed to have a significantly higher value of yellowness (b) and greater lightness (Lasik et al., 2016).

Free fatty acid profiles of yoghurts during storage

The samples having free fatty acids from C4 to C18:1 are given in Table 3. The most abundant FFAs in the samples were palmitic (C16) and myristic (C14) acids with concentrations of 97.55-44.26 and 31.76-18.07 ppm, respectively. The short- (C4-C10, SCFA), medium- (C12-C14, MCFAs), and long-chain (C16, C18, LCFA) FFAs represented 3.22-11.54, 0.94-31.54, and 4.12-97.55 ppm, respectively. Quantifying the levels of short-chain FFAs is important since their concentration can cause flavor changes and defects. However, despite the quantitative importance of medium and long-chain FFAs, they are not the main contributors of flavor to the dairy products (Güler and Park, 2011).

In the present study, the short-chain fatty acids i.e. butyric, caproic, caprylic, and capric acids were most abundant in the FY (fatty) control yoghurt at day 1. The addition of WPI affected (p < 0.05) the SCFA levels. Yoghurts containing 1 and 2 % WPI- (LY1, LY2) had lower levels than fatty and low-fat control yoghurts (without WPI). Similar trends were observed at day 7 and 14. Caproic (C6) and caprylic (C8) levels in all yoghurt samples decreased during the storage (p < 0.05). However, the butyric (C4) and capric acid (C10) levels of only LY2 (2 % WPI added) were significantly increased from day 1 up to 14. This could be related to not only the formation of volatile fatty acids, which are responsible for the formation of free fatty acids from lipolysis, but also to amino acid degradation (Beshkova et al., 1998; Güler and Park, 2011). Similar results were also obtained after 21-day storage of yoghurt by Güler and Gürsoy-Balcı (2011).

When it comes to the medium-chain free fatty acids, lauric (C12) and myristic (C14) acids presented similar behavior in all samples at the beginning of storage. MCFAs levels in all samples were significantly affected by the WPI addition and storage time (p < 0.05). Regardless of WPI addition, the levels of lauric and any myristic acids (C12-C14) were significantly lower in the WPI-containing yoghurts (LY1, LY2) than in non-WPI yoghurts (FY, LY). Lauric acid increased steadily in all the yoghurts during storage. However, no regular trend of increasing and decreasing was observed in myristic acid (C14) levels during the storage of all samples. Similar results were obtained in a study related to strained yoghurt during storage (Şenel et al., 2011).

Regarding long-chain fatty acids (LCFAs), palmitic (C16), stearic (C18), and oleic acid (C18:1) levels were significantly affected by the WPI addition during the first and last day of storage (p < 0.05). Being fatty, the FY (without WPI) control yoghurt had more LCFAs than other yoghurts. Similar to MCFAs, the LY2 (2 % WPI added) was lower than all samples in terms of LCFAs. The levels of LCFAs in all yoghurts were significantly (p < 0.05) affected by the storage period, with markedly lowered stearic acid (C18) levels while the palmitic (C16) and oleic acid (C18:1) levels were increased. In the literature, there are conflicting data concerning the lipolytic activity of yoghurt starters. Guler and Gürsoy-Balcı (2011) demonstrated the decrease in LCFAs in yoghurt, while Rao and Reddy (1984) found an increase in stearic and oleic acids. Oleic acids showed the same profile as palmitic acid in yoghurts and it was the second most abundant LCFA in the samples. The release of free fatty acids from triglycerides continues to occur during the process of lipolysis (Sumarmono et al., 2015).

Generally, it can be surmised that WPI addition affects the direction and intensity of changes in FFA levels during storage. The 2 and 1 % WPI-added yoghurts (LY2 and LY1) had the least short-, medium-, and long-chain fatty acids. However, irregular changes were observed during the storage. Contrary to expectations, there were increases in SCFAs (butyric and capric acid), MCFAs (myristic acid), and LCFAs (palmitic and oleic acid) during storage in the 2 % WPI-added sample (LY2). This could be related to the higher fat content in LY2 compared to LY and LY1 (Table 1). So, using WPI as a fat alternative in low-fat yoghurt affected FFA variation in storage.

CONCLUSION

The result obtained contributes to the physicochemical properties and free fatty acid composition of WPI-added low-fat yoghurts. Significant differences were found in the WPI-added yoghurts at the beginning and at the end of storage period (p < 0.05). A 2 % WPI addition increased total solids, protein, and fat levels of low-fat yoghurt. Similarly, the titratable acidity and tyrosine levels were higher with lower syneresis and texture enhancements, comparatively. However, WPI addition decreased the whiteness (L) and greenness (a) with an increase in the yellowness (b). Using a WPI generally resulted in a decrease of short-, medium-, and long-chain fatty acids. However, during the storage time, the levels of butyric, capric, myristic, palmitic, and oleic acid increased in the 2 % WPI-added yoghurts. These results indicate that WPI could be used as a fat alternative in low-fat yoghurt. However, more work is needed to gain detailed information about the microstructures, sensory characteristics, and microbial characteristics of low-fat yoghurt with 2 % WPI.
Table 3. Fatty acid profiles of yoghurt samples during 14 days of storage, (n=3) (ppm).

| Parameter* | Samples | Day 1 | Day 7 | Day 14 | Mean/Average |
|------------|---------|-------|-------|--------|--------------|
|            |         | Mean  | Mean  | Mean   |              |
|            |         | Values| Values| Values  |              |
|            |         |       |       |        |              |
| Butyric, C4| FY      | 5.17±0.82aB | 5.24±0.60aB | 7.17±0.51bB | 5.86±1.14B   |
|            | LY      | 3.67±0.20aA | 3.22±0.17aA | 3.32±0.10bA | 3.40±0.25A   |
|            | LY1     | 3.39±0.28aA | 3.22±0.20aA | 3.38±0.16bA | 3.33±0.21A   |
|            | LY2     | 3.35±0.33aA | 3.71±0.21aA | 3.70±0.13bA | 3.58±0.27A   |
|            | FY      | 11.30±0.26cA | 10.53±0.28bA | 8.57±0.20aA | 10.14±1.24A  |
|            | LY      | 10.83±0.61cA | 9.92±0.30bA | 9.04±0.93aA | 9.93±0.97A   |
|            | LY1     | 10.40±0.17cA | 9.85±0.32bA | 9.76±0.34aA | 10.00±0.39A  |
|            | LY2     | 10.01±0.35cA | 9.57±0.27bA | 9.77±0.12aA | 9.78±0.30A   |
|            | FY      | 9.89±0.27cC | 9.16±0.05bC | 8.31±0.57aC | 9.12±0.75C   |
|            | LY      | 9.62±0.24bC | 8.12±0.14bB | 7.79±0.18aB | 8.51±0.86B   |
|            | LY1     | 9.04±0.20cA | 7.55±0.45bA | 7.35±0.11aA | 7.98±0.84A   |
|            | LY2     | 8.15±0.12cA | 7.53±0.24bA | 7.66±0.14aA | 7.78±0.32A   |
|            | FY      | 11.54±0.34bC | 10.69±0.28aC | 8.69±0.38aC | 10.31±1.30C  |
| Capric, C10| LY      | 10.46±0.13bA | 8.66±0.25aA | 8.64±0.17aA | 9.25±0.92A   |
|            | LY1     | 9.93±0.29bB | 9.24±0.40aB | 10.16±0.14aB | 9.78±0.49B   |
|            | LY2     | 9.26±0.27bA | 8.55±0.52aA | 9.53±0.23aA | 9.11±0.54A   |
| Lauric, C12| FY      | 4.31±0.51aD | 4.40±0.46abD | 4.67±0.20bD | 4.46±0.39D   |
|            | LY      | 1.52±0.17aC | 1.60±0.12abC | 1.69±0.17bC | 1.60±0.15C   |
|            | LY1     | 1.30±0.08bA | 1.27±0.19abB | 1.47±0.06bB | 1.35±0.14B   |
|            | LY2     | 0.94±0.03aA | 0.96±0.03abA | 1.14±0.11bA | 1.01±0.11A   |
|            | FY      | 31.54±1.47aD | 30.97±1.32aD | 31.76±0.71aD | 31.42±1.11D  |
| Myristik, C14| LY     | 23.72±0.40aC | 28.30±1.42aC | 27.85±0.39aC | 26.62±0.32C   |
|            | LY1     | 22.58±0.34ab | 18.69±0.30aB | 18.83±0.06aB | 20.03±1.93B  |
|            | LY2     | 18.61±0.19aA | 17.58±0.29aA | 18.07±0.15aA | 18.09±0.48A  |
|            | FY      | 91.74±5.99aD | 96.38±3.55bD | 97.55±2.03bD | 95.22±4.50D  |
| Palmitic, C16| LY     | 68.30±1.48aC | 71.70±0.44bc | 71.91±0.41bc | 67.64±1.93C  |
|            | LY1     | 54.03±2.80aB | 56.92±3.10bb | 59.08±1.38bb | 56.68±3.11B  |
|            | LY2     | 44.26±0.86aA | 50.07±1.71ba | 51.92±0.62ba | 48.75±3.60A  |
|            | FY      | 28.36±0.21cC | 27.06±0.09bc | 25.53±0.42aC | 26.98±1.25C  |
| Stearic, C18| LY      | 5.34±0.44aB | 5.31±0.26bB | 5.15±0.14aB | 5.27±0.28B   |
|            | LY1     | 4.25±0.19aC | 4.26±0.06aB | 4.19±0.07aA | 4.24±0.11A   |
|            | LY2     | 4.12±0.09aC | 4.18±0.07bA | 4.12±0.03aA | 4.14±0.06A   |
|            | FY      | 29.15±0.98aD | 30.89±0.29bD | 31.89±0.05cD | 30.64±1.30D  |
| C18:1      | LY      | 13.10±0.13aC | 13.83±0.10bc | 14.20±0.05cc | 13.71±0.49C  |
|            | LY2     | 12.82±0.28aB | 13.04±0.28bB | 13.37±0.31cB | 13.08±0.35B  |
|            | FY      | 11.78±0.21aA | 12.44±0.09aB | 12.76±0.08ac | 12.33±0.45A  |

* FY: Fatty yoghurt (% 3 fat+% 0 WPI); LY: Low fat yoghurt (≤ 0.5 fat+% 0 WPI); LY1: 1% WPI added yoghurt (≤ 0.5 fat+% 1 WPI); LY2: 2% WPI added yoghurt (≤ 0.5 fat + % 2 WPI).

** The same column with different superscripts among yoghurt samples significantly differ (P<0.05).

*** The same row with different superscripts among yoghurt samples significantly differ (P<0.05)

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