The feasibility of $[^{18}\text{F}]$EF5-PET/CT to image hypoxia in ovarian tumors: a clinical study

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Abstract

Rationale: Evaluation of the feasibility of $[^{18}\text{F}]$EF5-PET/CT scan in identifying hypoxic lesions in ovarian tumors in prospective clinical setting.

Methods: Fifteen patients with a suspected malignant ovarian tumor were scanned with $[^{18}\text{F}]$EF5 and $[^{18}\text{F}]$FDG-PET/CT preoperatively. The distribution of $[^{18}\text{F}]$EF5-uptake, total intraabdominal metabolic tumor volume (TMTV), and hypoxic subvolume (HSV) were assessed.

Results: $[^{18}\text{F}]$EF5-PET/CT suggested hypoxia in 47% (7/15) patients. The median HSV was 87 cm³ (31% of TMTV). The $[^{18}\text{F}]$EF5-uptake was detected in primary tumors and in four patients also in intra-abdominal metastases. The $[^{18}\text{F}]$EF5-uptake in cancer tissue was low compared to physiological excretory pathways, complicating the interpretation of PET/CT images.

Conclusions: $[^{18}\text{F}]$EF5-PET/CT is not feasible in ovarian cancer imaging in clinical setting due to physiological intra-abdominal $[^{18}\text{F}]$EF5-accumulation. However, it may be useful when used complementarily to FDG-PET/CT.

Keywords: EF5-PET/CT, Ovarian cancer, Hypoxia

Introduction

Ovarian cancer (OC) is the most lethal gynecological malignancy, and the majority of patients are diagnosed at an advanced stage [1]. Although OC is initially chemosensitive, most women experience multiple and finally chemoresistant relapses. The survival odds have not markedly improved despite extensive research, and completeness of surgery is still a major prognostic factor [2].

The presence of hypoxic regions in solid tumors is associated with a poor prognosis for many cancer types [3–5]. Hypoxia-mediated chemoresistance is also the greatest clinical challenge in OC [6, 7].

There is a considerable need for non-invasive imaging of tumor hypoxia since it provides additional information, which could be integrated into strategies of treatment [8]. The method can improve therapeutic outcomes by predicting chemoresistance and selecting potentially treatment-resistant tumors for targeted surgery.

$^{18}$F-nitroimidazolpentafluoropropylacetamide ($[^{18}\text{F}]$EF5) is one of the extensively investigated and clinically tested tracers of tissue hypoxia [9, 10]. $^{18}$F$[^{18}\text{F}]$EF5 belongs to the nitroimidazole group and has considerable membrane permeability and capability to accumulate in viable hypoxic, though not in apoptotic or necrotic cells [11–13].

Since there is no systematic data evaluating the eligibility of hypoxia-imaging among patients with ovarian malignancy, we conducted the prospective clinical study to evaluate the feasibility of $[^{18}\text{F}]$EF5-PET/CT scan in identifying hypoxic lesions in ovarian tumors.

Material and methods

Study population

This prospective non-randomized study was conducted at Turku University Hospital, Finland, between November 2017 and June 2019. Patients between 38 and 79 years of age with ovarian tumor, who were not pregnant, nursing, or had a history of previous malignancies were included.
Ethical approval was obtained from the institutional review board (18.10.2016§443), and all subjects signed an informed consent form, ClinicalTrials.gov identifier: NCT04001023.

A whole-body contrast-enhanced $^{18}$F-FDG-PET/CT and $^{18}$F-EF5-PET/CT of the abdomen were performed on separate days preoperatively. PET/CT images were then evaluated by a nuclear medicine specialist and gynecological oncologist to assess the distribution of the cancer and to determine regions of suspected hypoxia in the intraabdominal tumor load for targeted biopsies for future research.

**PET/CT scanning procedure**

The PET/CT studies were performed with a digital PET/CT scanner: Discovery MI (General Electric Medical Systems, Milwaukee, WI, USA). It has combined PET/CT-scanners with a 128-slice CT and a 3D PET imaging capability. The PET imaging field of view (FOV) was 70 cm in diameter and 20 cm in axial length. To obtain attenuation correction for 511 keV photon distribution, the transmission scan was performed using a low-dose (noise index 30, automatic 3D current modulation, 10–120 mAs, and 120 kVp) CT protocol.

The patients received an intravenous injection of 370 MBq of $^{18}$F-EF5. A static emission scan was acquired 180 min from the tracer injection to cover the entire abdomen (3 bed positions, 7.5 min/bed). The patients voided prior to the scan. The sinogram data was corrected for dead-time, decay, and photon attenuation and reconstructed in a 256 \times 256 matrix. Image reconstruction followed the

### Table 1 Patient characteristics and PET information

| Nr. | Age at diagnosis (years) | Weight (kg) | Histology | Stage | MTV (cm$^3$) | Hypoxia in $^{18}$F-EF5 PET/CT | Hypoxic subvolume (cm$^3$, % of MTV) | Injected $^{18}$F-EF5 activity (Mbq) |
|-----|--------------------------|-------------|-----------|-------|-------------|--------------------------------|-------------------------------------|-------------------------------------|
| 1   | 62                       | 106         | high grade serous IIIC | 571   | yes         | 364 (64%)                      | 381                                 |
| 2   | 56                       | 81          | endometrioid IIB       | 250   | yes         | 121 (49%)                      | 367                                 |
| 3   | 65                       | 100         | sarcoma IIIB           | 1908  | yes         | 42 (2%)                        | 374                                 |
| 4   | 38                       | 93          | mucinous IA            | 158   | no          | 367                            | 367                                 |
| 5   | 73                       | 54          | carcinosarcoma IIIC    | 267   | yes         | 17 (6%)                        | 301                                 |
| 6   | 79                       | 68          | high grade serous IVB  | 615   | yes         | 202 (33%)                      | 220                                 |
| 7   | 67                       | 63          | high grade serous IIIC | 955   | no          | 385                            | 385                                 |
| 8   | 56                       | 70          | fibroma                | -     | 224         | no                             | 377                                 |
| 9   | 50                       | 103         | clear cell IIIC        | 116   | no          | 378                            | 378                                 |
| 10  | 69                       | 77          | high grade serous IVA  | 324   | no          | 380                            | 380                                 |
| 11  | 45                       | 91          | serous BOT IA          | 66    | no          | 374                            | 374                                 |
| 12  | 65                       | 69          | high grade serous IIIC | 273   | no          | 377                            | 377                                 |
| 13  | 50                       | 152         | endometrioid IIA       | 173   | no          | 363                            | 363                                 |
| 14  | 68                       | 59          | high grade serous IIIC | 1288  | yes         | 87 (7%)                        | 362                                 |
| 15  | 64                       | 60          | endometrioid IIA       | 346   | yes         | 33 (10%)                       | 177                                 |

*BOT borderline ovarian tumor, MTV metabolic tumor volume, HSV hypoxic subvolume*
Q.Clear method (a Bayesian-penalized likelihood reconstruction algorithm for PET) incorporating random and scatter correction with $\beta$ value of 350. The final in-plane FWHM (full-width half-maximum) of the systems was < 5 mm.

A whole-body [$^{18}$F]FDG-PET/CT with low-dose CT combined with diagnostic contrast-enhanced imaging for anatomical reference was performed following the standard institutional protocol for OC. The details of the protocol used for [$^{18}$F]FDG-PET/CT are the same as abovementioned [$^{18}$F]EF-5 PET/CT protocol except the larger scan area starting 60 min post injection from the base of the scull to the middle thigh (6 bed positions, 2 min/bed) with 4 MBq/kg of [$^{18}$F]FDG.

The scans were performed in a random order depending on the availability of the [$^{18}$F]EF5 and camera. The mean interval between scans was 2 (range 1–7) days.

Fig. 2 MTV (metabolic tumor volume, cm$^3$) and HSV (hypoxic subvolume, cm$^3$) of a particular patient (nr 1–15). Under the x-axis, the same patient’s distribution of the disease is presented.

![Graph showing MTV and HSV values](image)
Image analyses
Total metabolic tumor volume (MTV), maximal standardized $[^{18}F]$FDG, and $[^{18}F]$EF5-uptake values (SUVmax) were assessed with the PET VCAR (Volume Computer-Assisted Reading) program. For $[^{18}F]$FDG-PET/CT analyses the SUVmax values were corrected for body weight and injected dose. TMTV was defined as the sum volume of all lesions.

A hypoxic voxel was defined using a threshold tumor to gluteus maximus muscle ratio (TMR) for $[^{18}F]$EF5-uptake of 1.5, based on earlier experience [14]. The hypoxic subvolume (HSV) was defined as the sum volume of all lesions with TMR over 1.5.

Statistical analyses
Statistical analyses were performed using JMP Pro 13 software from SAS. Continuous variables were compared using a Wilcoxon rank-sum test. A non-parametric Spearman rank correlation test was used to evaluate the association between SUVmax values. Two-tailed $P$ values $< 0.05$ were considered statistically significant.

Results
Fifteen patients were enrolled. EF5-PET/CT suggested hypoxia in 46% (7/15) of the patients. In those patients the mean HSV was 87 [CI95% (9.5–238)] cm$^3$ and proportionally 31 [CI95% (2–47)%] of MTV.

Patients’ clinical and imaging characteristics are presented in Table 1. The distribution of $[^{18}F]$EF5-avid lesions was variable. In 7 of the 15 (46%) patients, $[^{18}F]$EF5PET/CT was suggestive for hypoxia. All of these patients had $[^{18}F]$EF5-accumulation inside the primary tumor and 4 also had $[^{18}F]$EF5-avid metastatic lesions. Seven patients had omental metastases, and 3 had $[^{18}F]$EF5-accumulation in the omental cake.

All of the $[^{18}F]$EF5-avid tumors also had increased uptake of $[^{18}F]$FDG; however, only a weak statistical correlation between $[^{18}F]$EF5 and $[^{18}F]$FDG-uptake in tumors was detected (Spearman’s rho 0.33, $p = 0.057$).
SUVmax values of EF5 and FDG uptake in the malignant primary tumors and metastases of 13 patients are presented in Fig. 1.

Seven patients had FDG-avid peritoneal carcinosis, a finding typical of OC. Only 1 patient had [18F]EF5-accumulation in the peritoneal carcinosis (thickness < 1 cm). The distribution of the disease, MTV, and HSV values are presented in Fig. 2. None of the patients had [18F]EF5-avid metastases when the primary tumor PET-finding was not suggestive of hypoxia. There was no statistical difference between [18F]EF5-uptake in the primary tumor and metastases (p = 0.72).

Our study included two patients with non-malign tumors (patient nr 8 and 11), which presented no [18F]EF5-accumulation.

The physiological uptake of [18F]EF5 in the gall bladder/bile, small intestine, and urinary bladder was notably higher than in the tumors (Fig. 3).

A demonstrative EF5- and FDG-PET/CT images of a patient with advanced ovarian cancer are presented in Fig. 4.

**Discussion**

Hypoxia is a common phenomenon in cancer with 50–60% of solid tumors containing hypoxic regions [15]. While hypoxia has a well-established role in promoting hematogenous metastases of cancer cells [16], the hematogenous spread is rare in OC at the time of diagnosis [17]. It should also be noted that the role of hypoxia in the transcoelomic spread to the peritoneum and omentum (common to OC) has not been widely investigated. On the basis of our study, [18F]EF5-PET/CT suggested hypoxia in half of the patients and the distribution of [18F]EF5-uptake was variable. [18F]EF5-uptake was detected mainly inside the ovarian tumor and less often in metastases. One preclinical study suggested a hypoxic environment to induce omental/peritoneal metastases [18]. Another study [19] which included two OC patients detected EF5-uptake and severe hypoxia in a peritoneal carcinosis biopsied laparoscopically promptly after the injection of EF5. Our cases with widespread peritoneal carcinosis typically had several [18F]FDG-avid areas but only one patient had [18F]EF5-avid peritoneal lesion.

The previous hypoxia imaging studies are conducted mostly on solid and locally advanced tumors [3, 5, 14, 20, 21], while half of our patients had disseminated cancer in the abdominal cavity hampering the comparison of our results to the other studies. However, [18F]EF5-uptake values in our study were comparable to previously reported [18F]EF5 SUVmax values in head and neck tumor studies, where the tumor hypoxia was confirmed with immunohistochemical staining [3, 14, 20]. Our study suggests that hypoxia is not a common phenomenon OC in transcoelomic metastases.

According to our study, the feasibility of [18F]EF5 PET/CT in ovarian cancer imaging in clinical practice appears to be limited. An uptake of [18F]EF5 in tumors was relatively weak and as the tumors were also [18F]FDG-avid, the visual estimation of [18F]-EF5-PET/CT – maximum intensity projection

TMR – tumor:gluteus maximus muscle ratio

**Fig. 4** An image of a patient with advanced ovarian cancer. All three metastases are FDG-avid (2b) and 2 of them have also accumulated EF5 (1b). The SUVmax-values in [18F]EF5-avid (tumor:gluteus maximus muscle ratio ≥ 1.5) are remarkably lower (1b) compared to the physiological uptake in bile (1).
CT was strongly lead by [18F]FDG-PET/CT. The avidity to both [18F]EF5 and [18F]FDG can be explained with Warburg’s effect [22], where cancer cells rely widely on glycolysis and reduce their respiration regardless of tissue oxygenation level. Unlike OC, head and neck and lung cancer are isolated tumors that are not surrounded by physiologically EF5-affine tissues. Our study is prospective, and two tumors eventually appeared to be benign. Nevertheless, we consider it important to present them especially as they showed no EF5 uptake.

Previously, two excretory paths of highly lipophilic [18F]EF5-tracer have been demonstrated [23, 24]. In the latter, it was assumed that due to slow tracer biliary excretion, only small amounts of activity would be seen in the small intestine. However, our study revealed an excessive [18F]EF5-uptake in the bile and small intestine. In contrast to the supradiaphragmatic tumors, this phenomenon imposes limitations on assessing tumors presenting weak [18F]EF5-uptake. Especially when located near to the intestine, metastases may be easily be mistaken to physiological uptake and remain unnoticed.

Conclusion
Non-invasive hypoxia imaging with [18F]EF5-PET/CT is possible, but its clinical use is restrained by the weak tumor uptake of the tracer compared to the non-specific uptake in excretory organs. The potential usefulness of with [18F]EF5-PET/CT in OC could be complementary to FDG-PET/CT with the intent to determine high-risk patients. The role of hypoxia in OC is intensively studied and [18F]EF5-PET/CT forms an attractive tool for patient stratification.

Abbreviations
OC: Ovarian cancer; [18F]EF5: 18F-nitroimidazolpentafluoropropylacetamide; MTV: Total metabolic tumor volume; VCAR: Volume Computer-Assisted Reading; SUVmax: Maximal standardized uptake value; TMR: Tumor to gluteus maximus muscle ratio; HSV: Hypoxic subvolume

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Authors’ contributions
SH, MS, ML and JH conceptualized the paper; ML managed patient’s diagnostic workup; SF was responsible for the tracer production; MS, TN, and ML analyzed and reported the imaging findings; ML drafted the manuscript; and all the authors revised and commented on the paper and approved the final version of the manuscript.

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Availability of data and materials
The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate
Ethical approval was obtained from the review board of Turku University Hospital (18.10.2016/443), and all subjects signed an informed consent form.

Consent for publication
A consent to publish individual persons’ data was obtained from that person.

Competing interests
The authors report no conflicts of interest.

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