Polymorphism of the DNA repair genes RAD51 and XRCC2 in smoking- and drinking-related laryngeal cancer in a Polish population

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Abstract

Introduction: Cigarette smoke and alcohol can generate reactive oxygen species, which may induce DNA double-strand breaks (DSBs), the most serious DNA lesion. In humans, DSBs are repaired mainly by non-homologous end joining and homologous recombination repair (HRR). Several polymorphisms in the DNA repair gene have been extensively studied in the association with various human cancers. In the present work we investigated the association between polymorphisms of two HRR genes, XRCC2 and RAD51, and tobacco- and alcohol-related larynx cancer in a Polish population.

Material and methods: Two polymorphisms of the XRCC2 gene, −41657C>T (rs718282) and 31479G>A (rs3218536), as well as one polymorphism of the RAD51 gene, −135G>C (rs1801320), were investigated by PCR-RFLP in 253 patients with larynx cancer and 253 age- and sex-matched non-cancer controls.

Results: Analysis of the gene-smoking and -drinking interactions revealed a weak association between larynx cancer and the −41657C>T polymorphisms of the XRCC2 gene among the moderate alcohol drinkers. The C allele of the −135G>C polymorphism of RAD51 increased cancer risk in the smoker group. Increased risk was also found for heavy drinkers. Additionally, there were no significant differences between distributions of genotypes in subgroups assigned to different TNM stages and grades.

Conclusions: The results indicated that the −135G>C polymorphism of the RAD51 gene may be associated with smoking- and drinking-related larynx cancer in Poland.

Key words: RAD51, XRCC2, laryngeal cancer, polymorphism.

Introduction

Larynx cancer is strongly associated with smoking and high alcohol consumption, as well as occupational and environmental exposure to carcinogens [1-3]. Cigarette smoke can generate reactive oxygen species, which are capable of inducing DNA double-strand breaks (DSBs) [4],
the most serious DNA damage, which, if not repaired or misrepaired, may result in genomic instability, cancer transformation or cell death [5-7].

Double-strand breaks are repaired by two major mechanisms: homologous recombination repair (HRR) and nonhomologous end joining (NHEJ) [8, 9].

RAD51 is a protein playing a central role in HRR through direct interaction with XRCC2, XRCC3, BRCA1, BRCA2 and other proteins to form a complex essential for the repair of DSBs and cross-links in DNA [10].

The RAD51 gene has been mapped to chromosome 15q14-15 and is highly polymorphic. The involvement of RAD51 in DNA repair determines its potential role in maintaining genetic stability, which is disturbed in cancer. Therefore the problem of genetic variability of the RAD51 gene in cancer is worth studying. A substitution G/C at position 135 in the 5' untranslated region (UTR) of the RAD51 gene (the 135 G/C 5' UTR polymorphism) has been reported as a possible factor to affect RAD51 mRNA stability [11, 12]. –135G>C polymorphism of RAD51 is located in the regulatory element of the RAD51 promoter and is suggested to be associated with messenger RNA expression. A study of women matched for BRCA1 mutation revealed that the C allele of this polymorphism is associated with a 2-fold lowering of breast and ovarian cancer risk as compared with the wild-type G allele [13]. The results of several previous studies suggested that RAD51 –135G>C polymorphism might be used as a predictive factor of precancerous lesions for head and neck cancer in a Polish population [14].

The XRCC2 gene, located at q36.1, is an essential part of the homologous recombination repair pathway and a functional candidate for involvement in cancer progression [15]. Common variants within XRCC2, including single nucleotide polymorphisms (SNPs) in exon 3 (Arg188His, R188H, rs3218536 or 31479G>A), have been identified as potential cancer susceptibility loci in recent studies, although association results are controversial. The Arg188His polymorphism has been proposed to be a genetic modifier for smoking-related pancreatic cancer and was associated with an increased risk of pharyngeal cancer and oral cancer risk [16-18]. The 188His allele of this polymorphism may be associated with a significantly increased risk of breast cancer, but not with bladder cancer, colorectal adenoma, or skin cancer [19-22].

In the present study the association between the –41657C>T and 31479G>A polymorphisms of the XRCC2 gene and the –135G>C polymorphism of the RAD51 gene and smoking- and proteinase K digestion. The final samples were kept in Tris-EDTA buffer, pH 8, at –20°C until use.

**Material and methods**

**Patients**

Blood samples were obtained from 253 patients: 194 men and 59 women with larynx cancer (squamous cell carcinoma) treated in the Department of Head and Neck Cancer, Medical University of Lodz, Lodz, Poland and the Department of Otolaryngology, Medical University of Poznan, Poland in 2005-2009 and 253 cancer-free age- and sex-matched controls. The patients ranged in age from 48 to 86 years (median: 63 years, mean ± SD: 63.9 ±8.4). There were 138 cases (55%) of grade 1, 104 cases (41%) of grade 2, and 11 cases (4%) of grade 3 in total. According to TNM staging there were 60 cases (24%) of stage I, 33 cases (13%) of stage II, 113 cases (45%) of stage III, 41 cases (16%) of stage IVA, and 6 cases (2%) of stage IVB. The study was approved by the Bioethics Committee of the Medical University of Lodz and each patient gave written consent.

**DNA isolation**

Peripheral blood lymphocytes (PBLs) were isolated by centrifugation in a density gradient of Histopaque-1077 (15 min, 280 g). Genomic DNA was isolated from PBLs by phenol/chloroform extraction and proteinase K digestion. The final samples were compared by Hardy-Weinberg equilibrium using the standard x^2^ test or Fisher’s exact test (for N ≤ 10). Genotype frequencies in cases and controls were compared by x^2^ tests. The genotype-specific risks
were estimated as odds ratios (ORs). In all cases wild type genotype served as a reference group.

Results

**XRCC2 and RAD51 gene polymorphisms**

Table I displays the distribution of genotypes and frequency of alleles of the −41657C>T, 31479G>A and −135G>C polymorphisms in patients with larynx cancer and controls. All distributions of genotypes and alleles were in Hardy-Weinberg equilibrium. We did not find any significant difference in genotype or allele frequencies in patients with cancer and controls (p > 0.05).

Table II shows the haplotype distribution of XRCC2. The haplotype analysis according to wild

| Genotype | Patients (N = 253), n (%) | Controls (N = 253), n (%) | OR (95% CI) | Value of p^b |
|----------|--------------------------|---------------------------|-------------|--------------|
| 31479G>A XRCC2 |                          |                            |             |              |
| G/G      | 230 (90.9)               | 240 (94.9)                | 1.00 Ref.   |              |
| G/A      | 22 (8.7)                 | 13 (5.1)                  | 1.76 (0.86-3.59) | 0.157        |
| A/A      | 1 (0.4)                  | 0                         | NE          |              |
| G/A + A/A | 23 (9.1)               | 13 (5.1)                  | 1.84 (0.91-3.73) | 0.119        |
| G        | 482 (95)                 | 493 (97)                  | 1.00 Ref.   |              |
| A        | 24 (5)                   | 13 (3)                    | 1.88 (0.95-3.75) | 0.093        |

| −41657C>T XRCC2 |                          |                            |             |              |
| C/C      | 88 (34.8)                | 75 (29.6)                  | 1.00 Ref.   |              |
| C/T      | 148 (58.5)               | 168 (66.4)                 | 0.75 (0.51-1.09) | 0.165        |
| T/T      | 17 (6.7)                 | 10 (4.0)                   | 1.44 (0.63-3.35) | 0.507        |
| C/T + T/T | 165 (65.2)              | 178 (70.4)                 | 0.79 (0.54-1.14) | 0.254        |
| C        | 324 (64)                 | 318 (63)                   | 1.00 Ref.   |              |
| T        | 182 (36)                 | 188 (37)                   | 0.95 (0.74-1.23) | 0.740        |

| −135G>C RAD51 |                          |                            |             |              |
| G/G      | 174 (68.8)               | 190 (75.1)                 | 1.00 Ref.   |              |
| G/C      | 69 (27.3)                | 58 (22.9)                  | 1.29 (0.87-1.94) | 0.145        |
| C/C      | 10 (3.9)                 | 5 (2.0)                    | 2.18 (0.73-6.51) | 0.242        |
| G/C + C/C | 79 (31.2)              | 63 (24.9)                  | 1.37 (0.92-2.02) | 0.138        |
| G        | 417 (82)                 | 438 (86)                   | 1.00 Ref.   |              |
| C        | 89 (18)                  | 68 (14)                    | 1.37 (0.97-1.94) | 0.082        |

NE – not estimated, ^aCrude odds ratio (OR), 95% CI – confidence interval at 95%, ^bχ2

| Haplotypes | Patients (N = 253), n (%) | Controls (N = 253), n (%) | OR (95% CI) | Value of p^b |
|------------|--------------------------|---------------------------|-------------|--------------|
| XRCC2-31479–41657 |                          |                            |             |              |
| G/G-C/C    | 79 (31.2)                | 74 (29.2)                  | 1.00 Ref.   |              |
| G/A-C/C    | 9 (3.6)                  | 3 (1.2)                    | 2.81 (0.73-10.78) | 0.207        |
| A/A-C/C    | 0                        | 0                         | NE          |              |
| G/G-C/T    | 136 (53.8)               | 158 (62.4)                 | 0.81 (0.54-1.19) | 0.327        |
| G/A-C/T    | 13 (5.1)                 | 8 (3.2)                    | 1.52 (0.59-3.88) | 0.516        |
| A/A-C/T    | 1 (0.4)                  | 0                         | NE          |              |
| G/G-T/T    | 15 (5.9)                 | 8 (3.2)                    | 1.75 (0.70-4.38) | 0.319        |
| G/A-T/T    | 0                       | 2 (0.8)                    | NE          |              |
| A/A-T/T    | 0                       | 0                         | NE          |              |

NE – not estimated, ^aCrude odds ratio (OR), 95% CI – confidence interval at 95%, ^bχ2
type of G31479G-C41657C showed a lack of association with laryngeal cancer.

In order to evaluate whether the polymorphisms were associated with the progression of larynx cancer, individuals were categorized into groups according to the cancer staging system of the TNM Classification of Malignant Tumours.

We did not find any association of the XRCC2 and RAD51 polymorphisms in the patient group with cancer progression assessed by TNM staging (p > 0.05) (data not shown).

There were no significant differences between distributions of genotypes in subgroups assigned to a particular histological grade (p > 0.05).

Gene-smoking and gene-alcohol drinking interaction

All patients and controls were divided into four groups: non-smokers (never), subjects smoking 10 cigarettes per day for 10 years (ever), those smoking 20 cigarettes per day for 20 years (moderate), and those smoking 20 cigarettes per day for 30 years (heavy). Among patients, 110 were ever smokers, 41 were moderate smokers, and 79 were heavy smokers. There were 76 ever, 31 moderate and 23 heavy smokers in the control group. The group of never smokers comprised 123 controls and 32 patients.

We investigated the relationship between smoking and the risk for larynx cancer, independent of genotype. We compared cancer patients and controls according to their smoking habits. Smoking significantly increased the risk of larynx cancer. Individuals who declared that they were moderate smokers had a slightly smaller odds ratio (OR = 8.57; 95% CI: 5.19-13.98) than individuals who declared that they were former smokers (ever group) (OR = 9.89; 95% CI: 6.84-14.28). Alcohol consumption also significantly increased the risk of larynx cancer. However, the effect of alcohol consumption was not so strong as smoking; for example, heavy smokers had OR almost five times greater (OR = 29.10; 95% CI: 17.37-49.45) than heavy drinkers (OR = 6.26; 95% CI: 3.58-11.03).

Table III shows the genotype distribution in cases and controls. Odds ratio analysis showed a statistically significant positive association of larynx cancer risk and smoking status for carriers of the C allele of the −135G>C polymorphism of the RAD51 gene. The risk of larynx cancer increased significantly in heavy smokers.

We also investigated the association between the polymorphisms and alcohol consumption (Table IV). Drinking habits were categorized in terms of non-drinking, moderate and heavy drinkers. We defined "moderate" drinking as 1-2 drinks/day, and "heavy" drinking as 3 or more drinks/day. In the control group there were 112 non-drinkers, 121 moderate and 20 heavy drinkers. Among patients, 60 were non-drinkers, 150 moderate and 43 heavy drinkers.

We found a statistically significant increase in risk of laryngeal cancer in the group of heavy alcohol drinkers carrying the C allele of the −135G>C polymorphism of the RAD51 gene. An observed increased risk of larynx cancer associated with this polymorphism for moderate alcohol drinkers was not statistically significant. We found a positive association between the risk of larynx cancer and the C/T genotype of the −41657C>T polymorphism of the XRCC2 gene in moderate drinkers.

The findings indicated that a statistically significant increased risk of head and neck cancer in non-drinkers was associated with heterozygous G31479A and the combined G31479A-A31479A genotype. The variant A allele of XRCC2 increased cancer risk in this group (Table IV).

Discussion

An attempt was undertaken in the present study to determine whether single nucleotide polymorphisms (SNPs) in the DNA repair pathway (XRCC2 −41657C>T, XRCC2 31479G>A, and RAD51 −135G>C) were associated with the risk of smoking- and drinking-related larynx cancer.

Tobacco smoking and alcohol drinking generate oxidative DNA damage which may contribute to larynx cancer. Tobacco smoke contains high quantities of chemical carcinogens, such as polycyclic aromatic hydrocarbons, aromatic amines, N-nitroso compounds and reactive oxygen species (ROS). These chemicals can form bulky adducts after activation by specific enzymes [25], and can induce a variety of oxidative damage [25-28].

A causal association has been established between alcohol consumption and cancers of the oral cavity, larynx, liver, colon, rectum, esophagus, pancreas and lung as well as breast cancer in women. The effect of alcohol can be modulated by polymorphisms in genes encoding enzymes for ethanol metabolism (alcohol dehydrogenases, aldehyde dehydrogenases, and cytochrome P450 2E1), folate metabolism, and DNA repair [29, 30]. Alcohol consumption is increasing in many countries and is an important cause of cancer worldwide [31].

Several studies have reported that the genes involved in DNA repair and in the maintenance of genome integrity play a crucial role in protecting against mutations that lead to cancer. Single nucleotide polymorphisms have been identified in several DNA repair genes, such as XRCC2 and RAD51, but the influence of specific genetic variants on repair phenotype and cancer risk has not yet been clarified. The polymorphisms chosen for this study have been shown to have functional sig-
Table III. Distribution of genotypes and odds ratios (OR) of the 31479G>A and 41657C>T polymorphisms of the XRCC2 gene and the –123G>C polymorphism of the RAD51 gene in patients with larynx cancer and the control group according to smoking status.

| Genotype | Non-smokers | Ever smokers | Moderate smokers | Heavy smokers | p | OR* (95% CI) |
|----------|-------------|--------------|-----------------|--------------|---|--------------|
|          | Patients (N = 32) | Controls (N = 123) | Patients (N = 10) | Controls (N = 76) | Patients (N = 41) | Controls (N = 31) | Patients (N = 79) | Controls (N = 23) | p |
| G/G      | 29 (91) | 119 (97) | 100 Ref | 89 (88) | 70 (92) | 100 Ref | 37 (90) | 30 (97) | 100 Ref | 75 (95) | 21 (91) | 100 Ref |
| G/A      | 3 (9) | 11 (11) | 1.44 (0.51-4.09) | 6 (8) | 4 (10) | 3.24 (0.34-30.57) | 2 (9) | 0.56 (0.09-3.27) | 0.13 (< 0.001) |
| A/A      | 0 (0) | 0 (0) | – | 0 (0) | 0 (0) | – | 0 (0) | 0 (0) | – | – | – |
| G/A + A/A | 3 (9) | 12 (12) | 1.57 (0.56-4.40) | 6 (8) | 4 (10) | 3.24 (0.34-30.57) | 2 (9) | 0.56 (0.09-3.27) | 0.13 (< 0.001) |
| G        | 61 (95.3) | 242 (98.4) | 100 Ref | 189 (93.6) | 146 (96.1) | 100 Ref | 78 (95) | 61 (98.4) | 100 Ref | 154 (97.5) | 44 (95.6) | 100 Ref |
| A        | 3 (4.7) | 13 (6.4) | 1.67 (0.62-4.51) | 6 (3.9) | 4 (5) | 3.12 (0.34-28.71) | 2 (4.4) | 0.57 (0.10-3.23) | 0.11 (< 0.001) |
| –41657C>T XRCC2 |
| C/C      | 9 (28) | 32 (26) | 100 Ref | 35 (35) | 19 (25) | 100 Ref | 13 (32) | 12 (39) | 100 Ref | 31 (39) | 12 (52) | 100 Ref |
| C/T      | 20 (63) | 86 (70) | 1.00 Ref | 60 (59) | 52 (68) | 100 Ref | 23 (56) | 19 (61) | 100 Ref | 45 (57) | 11 (48) | 1.58 (0.62-4.04) | 0.31 (0.001) |
| T/T      | 3 (9) | 5 (4) | 1.23 (0.43-10.68) | 6 (6) | 5 (7) | 0.65 (0.17-2.41) | 5 (12) | 0 (0) | – | 3 (4) | 0 (0) | – |
| C/T + T/T | 23 (72) | 91 (74) | 0.89 (0.37-2.14) | 66 (65) | 57 (75) | 0.62 (0.32-1.22) | 28 (68) | 19 (61) | 1.36 (0.51-3.61) | 0.31 (0.001) |
| C        | 38 (59.4) | 150 (60.9) | 100 Ref | 130 (64) | 90 (59.2) | 100 Ref | 49 (59.8) | 43 (69.4) | 100 Ref | 107 (67.7) | 35 (76.1) | 100 Ref |
Table III. Cont.

| Genotype       | Non-smokers | Ever smokers | Moderate smokers | Heavy smokers |
|----------------|-------------|--------------|------------------|---------------|
|                | Patients (N = 32) | Controls (N = 123) | Crude OR (95% CI) (p) | Adjusted OR* (p) | Patients (N = 101) | Controls (N = 76) | Crude OR (95% CI) (p) | Adjusted OR* (p) | Patients (N = 41) | Controls (N = 31) | Crude OR (95% CI) (p) | Adjusted OR* (p) | Patients (N = 79) | Controls (N = 23) | Crude OR (95% CI) (p) | Adjusted OR* (p) |
| T              | 26 (40.6) | 96 (39.1) | 1.06 (0.61-1.87) (0.32) | 1.06 (0.61-1.87) (0.32) | 72 (36) | 62 (40.8) | 0.81 (0.52-1.24) (0.380) | 1.94 (0.198) | 33 (40.2) | 19 (30.6) | 1.52 (0.76-3.06) (0.312) | 2.88 (0.071) | 51 (32.3) | 11 (23.9) | 1.51 (0.71-3.22) (0.365) | 1.59 (0.658) |
| -135G>C RADS1  |             |              |                  |                   |              |              |                  |                   |              |              |                  |                   |              |              |                  |                   |              |              |
| G/G            | 20 (63) | 91 (74) | 100 Ref | 70 (69) | 56 (74) | 100 Ref | 27 (66) | 24 (77) | 100 Ref | 57 (72) | 21 (91.3) | 100 Ref |                  |                   |              |              |                  |                   |              |              |
| G/C            | 10 (31) | 29 (24) | 1.57 (0.66-3.73) (0.427) | 0.16 (< 0.001) | 30 (30) | 18 (24) | 1.33 (0.67-2.63) (0.511) | 2.12 (0.024) | 12 (29) | 7 (23) | 1.52 (0.52-4.49) (0.624) | 171 (0.288) | 17 (22) | 2 (8.7) | 3.13 (0.66-14.72) (0.110) | 6.60 (< 0.001) |
| C/C            | 2 (6) | 3 (2) | 3.03 (0.47-19.36) (0.239) | 0.18 (< 0.001) | 1 (1) | 2 (3) | 0.40 (0.03-4.52) (0.424) | 0.58 (0.663) | 2 (5) | 0 (0) | – | 5 (6) | 0 (0) | – |
| G/C + C/C      | 12 (37) | 31 (26) | 1.76 (0.72-4.01) (0.256) | 0.13 (< 0.001) | 31 (31) | 20 (27) | 1.24 (0.64-2.41) (0.639) | 196 (0.040) | 34 (34) | 7 (23) | 1.77 (0.62-5.13) (0.420) | 2.01 (0.015) | 22 (28) | 2 (8.7) | 4.05 (0.87-18.74) (0.103) | 8.20 (< 0.001) |
| G              | 50 (78.1) | 211 (85.8) | 100 Ref | 170 (84) | 130 (85.5) | 100 Ref | 66 (80.5) | 55 (88.7) | 100 Ref | 131 (83) | 44 (95.7) | 100 Ref |                  |                   |              |              |                  |                   |              |              |
| C              | 14 (21.9) | 35 (14.2) | 168 (0.84-3.37) (0.193) | 0.10 (< 0.001) | 32 (16) | 22 (14.6) | 1.11 (0.62-2.00) (0.841) | 1.86 (0.036) | 16 (19.5) | 7 (11.3) | 1.90 (0.73-4.96) (0.269) | 2.12 (0.019) | 27 (17) | 2 (4.3) | 4.53 (1.03-19.84) (0.029) | 8.25 (< 0.001) |

*Odds ratio adjusted by age, sex and drinking
### Table N. Distribution of genotypes and odds ratios (OR) of the 31479G>A and the 41657C>T polymorphisms of the XRCC2 gene and the −135G>C polymorphism of the RAD51 gene in patients with larynx cancer and the control group according to drinking status

| Genotype        | Non-drinkers | Moderate drinkers | Heavy drinkers |
|-----------------|--------------|------------------|---------------|
|                 | Patients (N = 60) | Controls (N = 112) | Patients (N = 150) | Controls (N = 121) | Patients (N = 43) | Controls (N = 20) |
|                 | n (%) | Crude OR (95% CI) | Adjusted OR* (p) | n (%) | Crude OR (95% CI) | Adjusted OR* (p) | n (%) | Crude OR (95% CI) | Adjusted OR* (p) |
| 31479G>A XRCC2  |              |                  |                  |              |                  |                  |              |                  |                  |
| G/G             | 52 (87) | 1.00 Ref. |                  | 139 (93) | 1.00 Ref. |                  | 39 (91) | 1.00 Ref. |                  |
| G/A             | 8 (13)  | 4.15 (1.19-14.43) | 0.37 (0.041) | 11 (7)  | 1.11 (0.44-2.81) | 179 (0.267) | 3 (7)  | 1.46 (0.14-15.00) | 3.65 (0.279) |
| A/A             | 0 (0)   |                  |                  | 0 (0)   |                  |                  | 1 (2)  |                  |                  |
| G/A + A/A       | 8 (13)  | 4.15 (1.19-14.43) | 0.37 (0.041) | 11 (7)  | 1.11 (0.44-2.81) | 0.55 (0.200) | 4 (9)  | 1.94 (0.20-18.65) | 3.81 (0.061) |
| T/T             | 3 (5)   | 1.23 (0.27-5.67) | 1.95 (0.590) | 11 (7)  | 1.43 (0.42-4.93) | 1.03 (0.31) | 3 (7)  | 1.33 (0.12-14.87) |                  |
| T/T + C/T       | 40 (67) | 1.15 (0.59-2.24) | 0.74 (0.119) | 100 (66) | 0.54 (0.32-0.95) | 1.13 (0.538) | 25 (58) | 0.92 (0.31-2.73) | 2.14 (0.061) |
| C               | 77 (64.1) | 1.00 Ref. |                  | 189 (63) | 1.00 Ref. |                  | 78 (62) | 1.00 Ref. |                  |

**−41657C>T XRCC2**

| Genotype        | Non-drinkers | Moderate drinkers | Heavy drinkers |
|-----------------|--------------|------------------|---------------|
|                 | Patients (N = 60) | Controls (N = 112) | Patients (N = 150) | Controls (N = 121) | Patients (N = 43) | Controls (N = 20) |
|                 | n (%) | Crude OR (95% CI) | Adjusted OR* (p) | n (%) | Crude OR (95% CI) | Adjusted OR* (p) | n (%) | Crude OR (95% CI) | Adjusted OR* (p) |
| C/C             | 20 (33)  | 1.00 Ref. |                  | 50 (33) | 1.00 Ref. |                  | 18 (42) | 1.00 Ref. |                  |
| C/T             | 37 (62)  | 1.14 (0.59-2.24) | 0.92 (0.29-3.03) | 89 (59) | 0.51 (0.29-0.89) | 1.19 (0.410) | 22 (51) | 0.88 (0.29-2.67) | 2.16 (0.071) |
| T/T             | 3 (5)    | 1.23 (0.27-5.67) | 1.95 (0.590) | 11 (7)  | 1.43 (0.42-4.93) | 1.03 (0.31) | 3 (7)  | 1.33 (0.12-14.87) |                  |
| T/T + C/T       | 40 (67)  | 1.15 (0.59-2.24) | 0.74 (0.119) | 100 (66) | 0.54 (0.32-0.95) | 1.13 (0.538) | 25 (58) | 0.92 (0.31-2.73) | 2.14 (0.061) |
| C               | 77 (64.1) | 1.00 Ref. |                  | 189 (63) | 1.00 Ref. |                  | 78 (62) | 1.00 Ref. |                  |
Table IV. Cont.

| Genotype     | Non-drinkers | Moderate drinkers | Heavy drinkers |
|--------------|--------------|-------------------|----------------|
|              | Patients    | Controls          | Crude OR (95% CI) | Adjusted OR* (p) | Patients    | Controls          | Crude OR (95% CI) | Adjusted OR* (p) | Patients    | Controls          | Crude OR (95% CI) | Adjusted OR* (p) |
|              | (N = 60) n (%) | (N = 112) n (%) | (p)              |                   | (N = 150) n (%) | (N = 121) n (%) | (p)              |                   | (N = 43) n (%) | (N = 20) n (%) | (p)              |                   |
| T            | 43 (35.9)    | 76 (33.9)         | 1.79 (1.09-2.95) | 0.029            | 0.78 (0.121)    | 111 (37)        | 99 (40.9)        | 0.84 (0.59-1.20) | 0.399        | 123 (0.540)   | 53 (38)          | 13 (35)          | 1.41 (0.67-2.98) | 0.471          | 2.44 (0.068)    |
| –135G>C RAD51 | G/G 44 (73)  | 81 (72)           | 1.00 Ref         |                   | 100 (67)       | 92 (76)        | 1.00 Ref         |                   | 30 (70)       | 19 (95)        | 100 Ref          |                   |
|              | G/C 16 (27)  | 29 (26)           | 1.01 (0.49-2.07) | 0.887            | 0.32 (0.001)   | 42 (28)         | 26 (21)         | 1.48 (0.84-2.61) | 0.216        | 2.37 (0.004)  | 11 (26)          | 1 (5)            | 6.96 (0.83-58.41) | 0.041          | 3.29 (0.089)    |
|              | C/C 0 (0)    | 2 (2)             | –                |                   | 8 (5)          | 3 (2)          | 2.45 (0.63-9.52) | 0.305        | 3.48 (0.098)  | 2 (5)            | 0 (0)            | –                |
|              | G/C + C/C 16 (27) | 31 (28)      | 0.95 (0.47-1.92) | 0.888            | 0.38 (0.001)   | 50 (33)         | 29 (23)         | 1.58 (0.92-2.72) | 0.120        | 2.36 (0.002)  | 13 (31)          | 1 (5)            | 8.20 (0.99-68.15) | 0.022          | 3.43 (0.073)    |
|              | G 104 (86.7) | 191 (85.3)       | 1.00 Ref         |                   | 242 (80.7)     | 210 (86.8)     | 1.00 Ref         |                   | 71 (82.5)     | 39 (97.5)     | 100 Ref          |                   |
|              | C 16 (13.3)  | 33 (14.7)        | 0.89 (0.47-1.69) | 0.841            | 0.32 (0.001)   | 58 (19.3)       | 32 (13.2)       | 1.57 (0.98-2.52) | 0.074        | 2.35 (0.002)  | 15 (17.5)        | 1 (2.5)          | 8.23 (1.05-64.75) | 0.039          | 3.42 (0.071)    |

*Odds ratio adjusted by age, sex and smoking.
Polymorphism of the DNA repair genes RAD51 and XRCC2 in smoking- and drinking-related laryngeal cancer in a Polish population

nificance and may be responsible for a low DNA repair capacity phenotype characteristic of cancer patients including larynx carcinoma.

Cancer of the larynx is a head and neck cancer. These include tumors of the nasal cavities, paranasal sinuses, oral cavity, nasopharynx, oropharynx, hypopharynx, and larynx.

The contribution of polymorphisms of various DNA repair genes in development of head and neck carcinoma is controversial.

In the present work we investigated the frequencies of alleles of the XRCC2 –41657C>T, 31479G>A, and RAD51 –135G>C polymorphisms in blood from patients with larynx cancer and blood samples from healthy individuals. We investigated the relationship between genotype and the risk for larynx cancer, independent of smoking and drinking. In our study, none of these SNPs was found to individually contribute to larynx cancer risk. We demonstrated that there were no differences in the frequencies of the alleles between patients and controls. We also analyzed the distribution of genotypes and frequency of alleles in groups of patients suffering from larynx cancer according to different cancer staging by the TNM classification. We did not find any association of the XRCC2 –41657C>T, 31479G>A or RAD51 –135G>C polymorphisms in patients with cancer progression assessed by tumor size (T) and node status (N). Our results suggest that these SNPs may not be associated with the occurrence of larynx cancer. At least two concepts can be considered: these polymorphisms are not involved in larynx cancer at all, or they are dependent markers of the disease (gene-gene and gene-environment interactions may modify laryngeal cancer risk). In the second case it is important to find out which factors act in connection with XRCC2 –41657C>T, 31479G>A, and RAD51 –135G>C polymorphisms in larynx cancer. Therefore, as a high risk factor for larynx cancer occurrence we analyzed patients with positive smoking and drinking status within the larynx cancer group.

Our previous results confirm the significance of smoking and alcohol consumption in larynx cancer development. Drinking as well as smoking significantly increased the risk of laryngeal carcinoma in a Polish population [2].

In the reported study, we found an association between RAD51 –135G>C and XRCC2 –41657C>T polymorphism and the incidence of drinking-related larynx cancer, but found no significant association with XRCC2 31479G>A polymorphism. Instead, a significant association was identified between RAD51 –135G>C polymorphism and smoking-related larynx cancer.

Our results are in agreement with recent data obtained for smoking- and drinking-related laryngeal cancer in a Polish population [14] and opposed to those obtained by Werbrouck et al. [32], which showed a protective effect of the –135G>C polymorphism on larynx cancer occurrence. On the other hand, a protective effect was obtained in our study for all polymorphisms in the group of never smokers and never drinkers.

Some reports have documented that the XRCC2 –41657C>T variation was related to increased esophageal squamous cell carcinoma (ESCC) and gastric cardia adenocarcinoma (GCA) risk [24]. This study provides evidence that links –41657C>T polymorphism of the XRCC2 gene with the risk of larynx cancer in moderate drinkers. An increased risk of larynx cancer observed in carriers of heterozygous variants of the –41657C>T polymorphism in a group of moderate drinkers should be interpreted carefully because we did not obtain any association for the T/T homozygotes or for individuals carrying at least one T allele of this polymorphism. This may be due to a small number of variant alleles of this polymorphism obtained for a Polish population.

In our earlier studies we investigated SNPs in base excision repair genes in laryngeal carcinoma. We found that the Cys326 allele of the hOGG1 gene may increase the risk of larynx cancer associated with smoking or alcohol consumption [2]. The Ser/Cys and Cys/Cys genotypes increased the risk of larynx cancer among heavy smokers, but did not change the risk in former smokers or moderate smokers. These genotypes also increased the risk of larynx cancer in moderate and heavy drinkers.

Moreover, we analyzed XRCC2 and RAD51 polymorphisms in breast, endometrial and colorectal cancer [33-37]. Our results suggested that the –135G>C polymorphism of the RAD51 gene may be linked to endometrial and colorectal cancer and can be considered as an additional marker of this disease. However, we did not find any association of the XRCC2 polymorphisms with endometrial, breast or colon cancer progression. Śliwiński et al. examined the role of SNPs in genes that participate in the HR pathway (XRCC3 and RAD51), which repairs double strand DNA breaks and risk of head and neck cancer. The presence of the XRCC3 or RAD51 polymorphic variants allele was associated with the increased risk of cancer progression in smoking and alcohol drinking patients [14].

Flores-Obando et al. found no statistically significant association between smoking, alcohol, human papillomavirus and race/ethnicity and DNA repair gene polymorphisms (XRCC1- base excision repair (BER) and XPD- nucleotide excision repair (NER)) and head and neck cancer [38]. Also, the analysis of a Hungarian population suggested that XRCC2 polymorphism is not correlated with head and neck cancer [39].
In contrast, in the Polish population the combined Arg194Trp-Arg399Arg genotype of base excision repair gene XRCC1 may be associated with head and neck cancer and may have an impact on identification of a high-risk cancer population [40]. It is known that polymorphisms in XRCC2 may modify individual susceptibility to smoking-related cancers. Unfortunately, it is difficult to find in the literature reports directly linking SNPs in the DNA repair gene XRCC2 with head and neck cancer occurrence. Only in single studies does the research suggest that potential combination of XRCC1, XRCC2, XRCC3 and XRCC4 SNPs may have an impact on identification of an oral cancer high-risk population [35].

Benhamou et al. [16] found a correlation between the XRCC2-Arg188His polymorphism and cancers of the upper aerodigestive tract. The XRCC2 188His allele was associated with a significantly increased risk of pharyngeal cancer. The XRCC2 Arg188His polymorphism has been proposed to be a genetic mod- ifier for smoking-related pancreatic cancer [17] and was associated with an increased risk of oral cancer risk [36]. In contrast, Arg188His and three SNPs in the 5' promoter region have been associated with reduced risk of bladder cancer [36].

RAD51 and XRCC2 gene polymorphisms have been studied as a risk factor for various cancers including larynx cancer. In conclusion, the present study provides further evidence for the significance of XRCC2 and RAD51 polymorphism in larynx cancer.

The obtained data suggest that the –135G>C polymorphism in RAD51, involved in the DNA repair pathway, is associated with larynx cancer risk in smoking and drinking patients. Finally, it is postu- lated that –135G>C polymorphism of the RAD51 gene may be used as a predictive factor for smoking- and drinking-related larynx cancer in Poland. Further studies, conducted on a larger group, are suggested to clarify this point.

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