Wnt Genes in Wing Pattern Development of Coliadinae Butterflies

Jennifer Fenner1,2, Caleb Benson2, Luis Rodriguez-Caro2,3, Anna Ren4, Riccardo Papa5,6, Arnaud Martin4, Federico Hoffmann7, Ryan Range1 and Brian A. Counterman2*

1 Department of Biological Sciences, College of Sciences and Mathematics, Auburn University, Auburn, AL, United States, 2 Department of Biological Sciences, Mississippi State University, Starkville, MS, United States, 3 Division of Biological Sciences, College of Humanities and Sciences, University of Montana, Missoula, MT, United States, 4 Department of Biological Sciences, Columbian College of Arts and Sciences, The George Washington University, Washington, DC, United States, 5 Department of Biology, College of Natural Sciences, University of Puerto Rico—Rio Piedras, San Juan, Puerto Rico, 6 Molecular Sciences and Research Center, University of Puerto Rico, San Juan, Puerto Rico, 7 Department of Biochemistry, Molecular Biology, Entomology, and Plant Pathology, College of Agriculture and Life Sciences, Mississippi State University, Starkville, MS, United States

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*Correspondence: Brian A. Counterman bcounterman@biology.msstate.edu

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INTRODUCTION

Wnt genes are responsible for a variety of vital developmental processes and are highly conserved across animals. From axis formation in embryogenesis to insect wing development, Wnt genes are a major part of the developmental toolkit. There are 13 subfamilies of Wnt ligand genes that code for a set of secreted glycoproteins rich in cysteine. The transduction of these Wnt ligands is controlled by the canonical Wnt/β-catenin pathway and the non-canonical Wnt/JNK planar cell polarity (PCP) and Wnt/Ca2+ pathways (Rao and Kühl, 2010). The generalized function of Wnt...
Wnts in these pathways has been studied in detail over the past century (Nusse, 2012); however, the explicit function and contribution of each Wnt gene throughout development remains poorly understood, and most of what is known comes from model organisms such as mice and Drosophila. Attempts to extrapolate Wnt gene function from one model organism to another species have proven difficult, not only due to differences in the Wnt gene repertoires among species, but also the potential functions of different Wnt genes interacting in multiple Wnt pathways (Murat et al., 2010; Range, 2018). To better understand the role of Wnt ligands in the evolution of developmental processes, a systematic analysis of Wnt gene content and activity is needed in clades where Wnt ligands may have played an important role in morphological diversification.

Studies of the characterization and function of Wnt signaling among arthropods are limited outside of Drosophila. Among arthropod species that have been examined, the numbers of Wnt gene content vary from six in Acrithosiphon pisum to nine in Tribolium castaneum (Ding et al., 2019). Among holometabolous insects, there are eight Wnt genes that are most commonly found: Wnt1, Wnt5, Wnt6, Wnt7, Wnt9, Wnt10, Wnt11, and WntA. Drosophila lacks only WntA; the functions of its seven Wnt genes include axis patterning in embryogenesis, control of the sexual dimorphic development of reproductive organs, development of neuroblast identity, and wing formation (Murat et al., 2010). Lepidoptera typically harbor a repertoire of eight Wnt genes, Wnt1, Wnt5, Wnt6, Wnt7, Wnt9, Wnt10, Wnt11, and WntA that are expressed in various tissues and stages (Ding et al., 2019; Holzem et al., 2019). In all arthropods (Murat et al., 2010) and in other animal phyla (Somorjai et al., 2018) Wnt9–Wnt1–Wnt6–Wnt10 are arranged in a tandem array, forming a syntenic block of paralogs. Not only are these genes linked on a single chromosome, but some (Wnt1–Wnt6–Wnt10) also share similarities in expression patterns on the wing discs of wing-spotted fruit flies, Drosophila guttata (Koshikawa et al., 2015) and the Euphydryas chalcides butterfly (Martin and Reed, 2014). Such consistent expression patterns between diptera and lepidoptera suggest a possible ancestral role of these Wnts in wing development. In addition, Wnt7–Wnt5–WntA form a syntenic arrangement of genes that is conserved across insects (Murat et al., 2010), and seem to originate from an ancestral Wnt5-7 cluster that pre existed at the root of Bilateria (Somorjai et al., 2018). Finally, Wnt11 is not located within either lepidopteran Wnt cluster, but is on another chromosome separate from the two main clusters (Ding et al., 2019).

Recently, Wnt genes have been implicated in the development and evolution of color patterns on butterfly wings. Early studies in Heliconius butterflies used genetic mapping, gene expression assays, and pharmacological manipulation to identify WntA as the gene responsible for major differences in melanin sexy wing patterns across the genus (Martin et al., 2012). Recently, the functional role of WntA in wing pattern development was shown in twelve Heliconius species using CRISPR gene-editing (Concha et al., 2019). Further, the role of WntA in wing pattern development has also been explored across other nymphaid butterflies, which all showed unique changes in wing pattern, including the white vein contour markings of Monarch wings (Gallant et al., 2014; Martin and Reed, 2014; Mazo-Vargas et al., 2017). Beyond this work in nymphaid butterflies, little is known about the importance of the Wnt landscape in various aspects of insect development (Murat et al., 2010).

Within Lepidoptera, the most well-characterized Wnt gene beside WntA, is Wnt1, for its role in color pattern development. Wnt1, designated as wingless when it was first discovered in Drosophila, is responsible for the development of imaginal discs, and is therefore vital for the formation of both wings and eyes (Baker, 1988). Wnt1 is perhaps most extensively studied of the Wnt genes due to its role in segment polarity (Baker, 1987). In the silkworm, Bombyx mori, Wnt1 has likely retained this ancestral role, as Wnt1 knockouts with CRISPR/Cas9 reveal its necessity during embryogenesis (Edayoshi et al., 2015; Zhang et al., 2015). In nymphaid butterflies, Wnt1 retains its expected embryonic expression (Holzem et al., 2019) and is also expressed in the peripheral tissue of the developing wings (Macdonald et al., 2010), as in other holometabolous insects (Tomoyasu et al., 2009). Within those developing wing tissues, butterfly Wnt1 also pre-patterns the “Discalis” ground plan elements on the wing across several lepidopteran lineages (Martin and Reed, 2010). In at least one nymphaid butterfly, Wnt6 and Wnt10 wing disc expression is concurrent with Wnt1 in both the peripheral tissue and discal pattern elements of the larval wing disk (Martin and Reed, 2014); an overlap that is consistent with the previously described functional redundancy between these three paralogs in Drosophila and Tribolium (Bolognesi et al., 2008; Murat et al., 2010). Quantitative PCR profiles of the silkworm Wnt repertoire also suggest that Wnt1, Wnt6, Wnt7, Wnt10, and WntA are all expressed in larval wing discs (Ding et al., 2019). Similarly, an RNA-seq study of pupal wing development in passionvine butterflies revealed expression of Wnt1, Wnt5, Wnt6, Wnt7, and WntA (Hanly et al., 2019). To our knowledge, Wnt9 and Wnt11 have not been shown to be expressed during lepidopteran wing development, and Wnt9 (syn. DWnt4) has been implicated in differences in male and female reproductive development in Drosophila (Cohen et al., 2002; Murat et al., 2010).

Here, we characterize the repertoire of Wnt genes in a pierid butterfly, Zerene cesonia, and explore their expression and potential role during wing pattern development. Z. cesonia is a Coliadinae butterfly, which are characterized by their strikingly bright yellow wings that contrast sharply with black margins often found on the wing edge. They are also noted for the bright ultraviolet (UV) reflectance of male wings, produced by specialized nano-structures on yellow pterygial scales in the medial region of the forewing (Fenner et al., 2019). Among Colidinae species, UV brightness is the top indicator of male mating success (Papke et al., 2007) and we are interested in whether Z. cesonia may play similar roles in pattern development in Z. cesonia as to what has been observed in other butterflies.

Pierid butterflies diverged from better studied Nymphalidae, which includes Monarchs and Heliconius butterflies, ~51 million years ago, and from silkmoths > 95 million years ago, providing a comparative framework in Lepidoptera to explore Wnt evolution and function in wing development (Kawahara et al., 2019). Using an RNA-seq approach, we detect the expression of four out of eight Wnt genes during the wing development of Z. cesonia.
Interestingly, we find different Wnt genes are expressed in Z. cesonia relative to nymphalid wings, including no expression of WntA, despite its involvement in wing pattern development of several nymphalid species. Using pharmaceutical assays similar to studies of nymphalids, we observed a similar expansion of melanic scales across the forewing in two species of pierid butterflies, Z. cesonia and its close relative Colias eurytheme. Importantly, we also observed a loss of the UV reflective pattern in Z. cesonia. Our findings suggest differences in the way the Wnt repertoire is used for color patterning during the wing development of pierids and nymphalids.

MATERIALS AND METHODS

Gene Expression Assays
RNA was extracted using TRIZOL from developing wing tissue of male Z. cesonia from fifth instar larvae and every day of pupation till emergence (i.e., days 1–6). A minimum of three males were used for each stage (with a maximum of five replicates on day 4 of pupation). Total RNA was extracted and mRNA was isolated using oligo(dT) magnetic beads from NEBNext Poly(A) mRNA Magnetic Isolation Module (NEB, Ipswich, MA, United States). cDNA was prepared using a modified version of the Smart-seq2 protocol (Picelli et al., 2014) and libraries were prepared using a Nextera XT DNA Library Preparation Kit (Illumina, San Diego, CA, United States). Libraries were sequenced as 150 base pair paired-end reads, at an average of 18.9 million reads per library (minimum of 13.2 million reads and maximum of 28.19 million reads). RNA-seq reads were preprocessed using AfterQC (Chen et al., 2017) and Trimomatic (Bolger et al., 2014). Reads were then mapped to gene models from the reference genome using Bowtie2 and gene expression was estimated using RSEM (Li and Dewey, 2011) with default parameters. Wnt expression profiles were identified for Wnt1/wingless, Wnt6, Wnt7, and Wnt11, but no RNA-seq reads mapped to WntA, Wnt10, Wnt5, or Wnt9. Expression profiles are reported as mean FPKM (fragments per kilobase of transcript per million mapped reads) values across individuals for each developmental stage.

Gene Content Analyses
We used a combination of bioinformatic approaches to identify the repertoire of Wnt genes in representative Lepidoptera, using the Drosophila melanogaster wnt repertoire as a reference. We first surveyed the Ensembl Metazoa database (Howe et al., 2020) for the full set of wnt paralogs in B. mori, Danaus plexippus, Heliconius melpomene, and Melitaea cinxia. We then used BLAST (Altschul et al., 1990) to search for the Wnt paralogs in Bicyclus anynana, Pieris rapae, and Vanessa tameamea, which are available at NCBI. Wnt genes for Pieris napi and Phoebis sennae were acquired using BLAST tools and available genome assemblies at lepbase.org (Challi et al., 2016). Wnt genes were identified in Z. cesonia using the annotated genome assembly v.1.0 (NCBI Accession JAAHXH000000000) (Rodriguez-Caro et al., 2020) and confirmed through sequence similarity searches and alignments with BLAST and MUMmer3.23 (Altschul et al., 1990; Kurtz et al., 2004). A Zerene eurydice, the sister species to Z. cesonia, individual (collected in San Bernardino, CA, United States) was sequenced (150 bp paired end Illumina reads, 6 Gb of data yielding 20X coverage) and reads were mapped to the Z. cesonia genome assembly v1.0. Z. eurydice Wnt gene sequences were extracted using the Z. eurydice genotypes. For C. eurytheme, a 10X Genomics linked-read library was constructed for a single individual (collected in Osborne, MS, United States) according to protocol (10X Genomics, San Francisco, CA, United States) and sequenced by Novogene yielding 10.6 Gb of sequence data, which was assembled with Supernova (Weissenfeld et al., 2017). Assembly scripts and information are available through a github repository1. To identify the Wnt genes sequences in C. eurytheme, BLAST and MUMmer were used to search and align Z. cesonia Wnt gene sequences with the 10X assembly.

Phylogenetic Analyses
We aligned the Wnt paralogs for silkmoth (B. mori), four species of nymphalids (B. anynana, H. melpomene, Junonia coenia, V. tameamea), and five species of pierid butterflies (C. eurytheme, Phoebis sennae, P. napi, P. rapae, Z. cesonia, and Z. eurydice) based on their conceptual amino acid translation using MUSCLE (Edgar, 2004) as implemented in MEGA (Kumar et al., 2018). We then reconstructed phylogenetic relationships among lepidopteran Wnt genes to resolve orthology. We inferred phylogenies in a maximum likelihood framework using IQ-TREE ver 1.6.6 (Nguyen et al., 2015) in the implementation available from the IQ-Tree web server (Trifinopoulos et al., 2016) last accessed on December 2019, selecting the best-fitting model of substitution identified with the ModelFinder subroutine from IQ-Tree (Kalyaanamoorthy et al., 2017). Support for the nodes was evaluated with the Shimodaira–Hasegawa approximate likelihood-ratio test (SH-aLRT), the aBayes test (Anisimova et al., 2011) and 1000 pseudoreplicates of the ultrafast bootstrap procedure (Hoang et al., 2018). We then removed the D. melanogaster sequences and used the resulting maximum likelihood tree to estimate differences in selective pressure among the lepidopteran wnt paralogs as evidenced by differences in the ratio of the rate of non-synonymous substitution per non-synonymous site (dN) to the rate of synonymous substitution per synonymous site (dS) using the codon-based maximum likelihood approach proposed by Goldman and Yang (1994) as implemented in PAML ver 4.9j (Yang, 2007). We explored variation in the dN/dS ratio among the different lepidopteran wnt paralogs using the branch models implemented in the program codeml implemented in PAML.

Pharmaceutical Assays
Zerene cesonia females were collected from a local Mississippi prairie (33°30’36.98”N, 88°44’14.57”W). Females were allowed to oviposit eggs on Dalea purpurea. Upon hatching, larvae were raised on an artificial diet (Shelby et al., 2020) under 16:8 photoperiod and 28°C conditions. A laboratory colony was then maintained over several generations under these conditions. At 12 h after pupation, Z. cesonia individuals (N = 25) were injected with either 0.5 µg of heparin sodium salt (Sigma–Aldrich) diluted...
in water, or with water as a control, and *C. eurytheme* (*N* = 5) were injected with 0.25 µg of heparin. Injections were conducted with a 10 µl Hamilton syringe with a 26-gauge needle into the right side in the interstice that separates the baso-posterior sections of the developing wings. The outer pupal casing was sterilized with ethanol prior to injections and the pupa were hung in a pupal chamber until emergence. Individuals were preserved for photography and microscopy following eclosion.

**Photography and Microscopy**

Individuals were photographed using an UV-IR converted Nikon D7000 camera with an AF-S Micro Nikkor 105 mm Lens for visual color images. UV color images used the same lens and camera but with the addition of a BAADER U-Filter, 60 nm IR, and VIS filter. Scanning electron microscopy was performed on a JEOL JSM-6500F FE-SEM at 5 kV. Wing sections of *Z. cesonia* and specimens of *Heliconius sara* from Martin et al. (2012) were mounted on aluminum stubs with silver paste and coated with 15 nm of platinum.

**RESULTS**

**Wnt Gene Repertoire and Evolution in Butterflies**

The *Wnt* gene content is conserved across all Lepidopteran genomes studied. Our phylogenetic analyses grouped the Lepidopteran *Wnt* paralogs into eight strongly supported clades that correspond to previously described *Wnt* genes in nymphaid butterflies (*Wnt1/wingless, Wnt5, Wnt6, Wnt7, Wnt9, Wnt10, Wnt11, and WntA*). In the *Z. cesonia* assembly v1.0, the eight *Wnt* genes are distributed in clusters on three chromosomes: *Wnt 9-1-6-10* on chromosome 1, *Wnt A-5-7* on chromosome 25, and *Wnt11* on chromosome 11 (Figure 1 and Supplementary Table S1). This was not the case for *Wnt11* and *WntA*, which have been secondarily lost in the genome of *D. melanogaster*. The *D. melanogaster Wnt 7* paralog was placed sister to the clade that included the lepidoptera *Wnt11* and *Wnt7* paralogs, likely as a result of deep divergence among these *Wnt* paralogs.

Our analyses of patterns of molecular evolution failed to find significant differences in estimates of synonymous and non-synonymous substitution rates (ω) among the different lepidoptera paralogs. The model that assigns a single estimate of ω to all codons in the alignment along all branches in the tree could not be rejected in any of our tests (Supplementary Table S2), with very low value for this parameter (ω < 0.01). These findings suggest *Wnt* genes are evolving very slowly in Lepidoptera and are likely under stringent purifying selection due to their highly important and conserved functions during organismal development.

**Wnt Gene Expression During Wing Development**

Expression of *Wnt* genes during wing development of *Z. cesonia* was similar and divergent from patterns described in other lepidopteran species. Fifth instar larval wing disks showed strong expression of *Wnt1* and *Wnt6*, consistent with their known expression in lepidoptera wing peripheral tissues (Macdonald et al., 2010; Martin and Reed, 2010, 2014) as well as in the *Drosophila* wing disk dorso-ventral boundary (Janson et al., 2001). In *Z. cesonia*, both genes showed decreased, but detectable levels during the pupal stages, which may be related to color patterning in the first 2 days after puparium formation (APF). *Wnt10* is co-expressed with *Wnt1* and *Wnt6* in *D. guttifera* and in the nymphaid *E. chalcedon* (Martin and Reed, 2014; Koshikawa et al., 2015), but was not recovered in our pierid dataset. *Wnt7* was expressed in both the larval stage and the first pupal day, as previously reported in two *Heliconius* species (Hanly, 2017). Following this initial peak, expression sharply decreased peaked again later during pupal development starting at the day 2 stage before rising again shortly at 5–6 days APF. *Wnt11* has been associated with *Wnt*/INK PCP pathway in mammals (Uysal-Onganer and Kypta, 2012), and its expression was detected during mid-pupal stages when scale cytoskeletal structures are forming. However, *Wnt11* transcript abundance was at least twofold lower than any other *Wnt* gene suggesting expression may be very spatially and/or time restricted during wing development (Figure 2). Most noteworthy, is the lack of *WntA* expression, which is involved in wing pattern development among all the nymphalid butterflies assessed so far (Mazo-Vargas et al., 2017). For those *Wnt* genes not detected from the wing RNA-seq data, we also surveyed RNA-seq data from adult head and thorax previously generated to help annotate the *Z. cesonia* genome, and we were able to confirm expression of *Wnt5*, *Wnt10*, and *WntA*, but not *Wnt9*, suggesting the expression of *Wnt9* is restricted to specific tissues and stages as hinted from other studies (Bolognesi et al., 2008; Murat et al., 2010; Chen et al., 2016).

**Pharmaceutical Assays Implicate Morphogens in Wing Pattern Development**

Early studies using the pharmaceutical Heparin, which can bind to *Wnt* ligands and facilitate their secretion and transport, induced a possible *Wnt* gain-of-function effect in butterfly wing patterns. These Heparin injection individuals expressed phenotypes that exhibited an extension of the basal, discal, and external symmetry systems (Serfas and Carroll, 2005). Pupal injection of Heparin in coliadinae butterflies produced melanic expansions that were reminiscent of the effects of Heparin in other butterflies (Serfas and Carroll, 2005; Martin et al., 2012; Sourakov, 2018). Heparin caused the apical melanic wing region to expand broadly across dorsal forewings in *Z. cesonia* females. In males, dorsal forewings showed only moderate levels of melanization but underwent instead a complete loss of the UV patterns. The dorsal hindwing melanic border was expanded in females, but unaffected in males. Both males and females exhibited ectopic expression of melanin patterning on the ventral fore and hindwings, with particularly strong melanic coloration on the apical region of the forewing. A similar trend of forewing melanin expansion was observed in *C. eurytheme* Heparin injected female individuals (Figure 3). Only a single
FIGURE 1 | Wnt gene content and synteny in butterfly genomes. Phylogenetic analyses reconstruct clades for each of eight Wnt genes present in lepidoptera genomes. Inset shows chromosomal linkage and order of Wnt genes in Z. ceconia genome assembly (Supplementary Table S1), which is known to vary within clusters across Lepidoptera (Holzem et al., 2019).
male *C. eurytheme* injected with Heparin survived, but did not eclose properly, therefore we were unable to assay Heparin’s impact on UV reflectance in *Colias*. These experiments show that Heparin will perturb both pigment and structural coloration in Coliadinae butterflies in a manner similar to nymphalids, despite the lack of *WntA* expression. They also reveal sexually dimorphic effects of heparin that were unexpected. It is plausible that those differences highlight intrinsic divergence in the female and male patterning architecture, a hypothesis that will require further investigation but establish *Z. cesonia* as an interesting model system for developmental studies of sex-specific color patterning.

Inspection of scales on *Z. cesonia* wings revealed broad organizational and scale shape changes (*Figure 3*). In the expanded melanic region of Heparin injected individuals, the melanic scales had their characteristic shapes and structures, in place of yellow scales that would typically be present. Heparin injected males that lost UV patterning did develop cover scales that appear to have the tightly organized and stacked lamellae ridges responsible for reflecting UV. However, these scales had highly reduced widths, exposing much of the ground scales that lack the UV-reflecting structures. The result was a weakly reflective UV patch on the forewings of Heparin injected male *Z. cesonia* (*Figure 3*). Scale size was highly reduced among Heparin treated individuals across multiple wing regions and both sexes (*Figure 3B*), suggesting that the Heparin had a systemic impact on scale development across the wings. This microscopic inspection of the impacts of Heparin demonstrates clear changes in scale structure that had not been previously documented in nymphalids.

Microscopic examination of Heparin treated *Heliconius* butterflies (Family: Nymphalidae) did not reveal the systemic scale reduction observed across the wings of *Z. cesonia*, suggesting that Heparin may affect scale development differently in the two families of butterflies. In *H. sara*, Heparin induced changes in both pigment and scale structure across the wing, with a complete loss of yellow scales. The black and yellow scales of *H. sara* are distinguishable by the shape and organization of their lamellar ridges, as seen at 5000X in *Figure 4*. SEM imaging revealed that Heparin induced melanic scales (*Figure 4*) had the expected ridge architecture of wildtype melanic scales. Similar changes in both pigment and scale structure were also observed in the CRISPR-edits of *WntA* among *Heliconius*.
butterflies (Concha et al., 2019), although the mutant color patterns typically altered in more narrow wing regions, since heparin is thought to also affect other Wnt ligands (Martin and Reed, 2014; Mazo-Vargas et al., 2017). In Z. cesonia, we did not see a concordant change in pigment and structure across the wing. Instead, melanic patterns expanded as expected and scale structures were altered across all color of scales (Figure 3). The melanic scales of Heparin treated female wings had an ultrastructure that was distinct from both black and yellow scales of control wings, with triple pronged leading edges that are not commonly found across Z. cesonia wings (Figure 3, blue box).

It is worth noting the large difference in Heparin dosage among nymphaid and pierid butterfly species; for example, 5 µg of Heparin was required to observe wing pattern changes in Heliconius, while only 0.5 µg was used in Z. cesonia, and higher dosage always resulted in pupal mortality (n = 25). This suggests that pierids may have a more sensitive response to Heparin, possibly due to Heparin binding to different ligands in Z. cesonia than in H. sara. Collectively, these Heparin results show that although Heparin appeared to produced similar wing pattern phenotypes across butterfly families, the effects of Heparin on scale development and dosage responses differ dramatically. Collectively, these findings suggest that targets of Heparin impact...
FIGURE 4 | Heparin impacts both scale pigmentation and structure in *H. sara*. Wings from Martin et al. (2012). Heparin injections result in yellow-to-blue/black scale color shifts, and SEM imaging reveals concordant changes in ultrastructure (5000X).
both scale pigmentation and structure across nymphalid and pierid butterfly wings, but also indicate that pierids may use a different set of Wnts or other signaling molecules to pre-pattern wing coloration.

**DISCUSSION**

### Wnts in Butterfly Wing Pattern: Conservation and Divergence

*Wnt* gene content, clustering, and amino acid sequence appear to be highly conserved among pierids, as well as other Lepidoptera. However, the deployment of these genes appears to be highly divergent among different butterfly lineages. Interestingly several *Wnt* genes displayed divergent expressions between developing wings of nymphalid and pierid butterflies. The expression profiles for *Wnt* genes in *Z. cesonia* clearly showed that they do not deploy the same *Wnt* genes as nymphalids in wing pattern development.

Most surprising was the lack of *WntA* expression during wing development in *Z. cesonia*, despite its importance in wing pattern development of nymphalid butterflies (Mazo-Vargas et al., 2017; Concha et al., 2019). In nymphalids, *WntA* expression in larval wing discs prefigures adult wing elements and diminishes during the first days of AF (Martin et al., 2012; Martin and Reed, 2014). *Z. cesonia* completely lacked *WntA* expression, suggesting that this pierid lacks the *WntA*-positive stripe system of nymphalids called the Central Symmetry System (CSS). Of note, classic work in comparative morphology has established that this pattern is divergent and unusually reduced in Pieridae (Schwanwitsch, 1956) and it will be interesting to further investigate if *WntA* expression has been completely lost in additional pierids to test for generalization. It is possible that *WntA* may be expressed in levels too low to detect in the *Z. cesonia* transcriptome, with an average of 18.9 million reads per library, but if so such low levels of expression, it is unlikely that *WntA* would be serving the same vital function in *Z. cesonia* as it is in nymphalid butterflies where it is more highly expressed.

Differences in *Wnt* gene deployment during wing development among lepidoptera lineages likely result from changes in regulatory regions of the *Wnt* genes. Across Lepidoptera, there is evidence of strong purifying selection on *Wnt* gene amino acid sequences, demonstrating that both *Wnt* gene content and sequences are highly conserved (Martin et al., 2012; Mazo-Vargas et al., 2017; Holzem et al., 2019). In *Heliconius* butterflies, differences in the spatial expression of *WntA* across developing wings are associated with genetic changes in non-coding regions near *WntA*. Van Belleghem et al. (2017) speculate that these reflect putative cis-regulatory elements (CREs) that are responsible for initiating *WntA* expression in narrow wing regions that correspond to pattern elements in the adult wings. Similarly, Van Belleghem et al. identified CREs near the gene *optix* that were associated with red color patterning in *Heliconius*, which (Lewis et al., 2019) recently used CRISPR to edit the CREs and demonstrate their function in red pattern development. We suggest that evolution of CREs near the *Wnt* gene clusters is likely responsible for differences in *Wnt* gene deployment and pattern divergence of lepidoptera wings.

### Wnt Pathway Involved in Structural Coloration

Heparin treatments consistently lead to an expansion of melanism across butterfly wings. However, if Heparin is impacting melanism via the Wnt pathway, it is likely affecting different Wnt ligands in different butterfly species. In nymphalid butterflies, *WntA* has been shown to be spatially expressed in wing regions that melanism expands and is a likely candidate target of Heparin treatments. However, in *Z. cesonia* wing discs, *WntA* is not expressed, and at the time of Heparin injections only *Wnt1*, *Wnt6*, and possibly *Wnt7* ligands appear to be present in wing tissue. Also different from nymphalid butterflies, the expansion of melanism was greatest in the anterior portion of the wing in *Z. cesonia* and *C. eurytheme*, where in nymphalids, melanin expansion largely occurred in elements of the CSS of the wing (Martin et al., 2012). Lastly, we note that the melanin response to Heparin was sex dependent, with females exhibiting greater expansions than males. This sexual dimorphism has not been observed in nymphalids; however, in species such as *Heliconius*, sexual dimorphism in wing color pattern is rare. Among pierids, wing color patterns are often sexually dimorphic, most notably in UV reflective patterns which are often present in males, but not females, as is found in *Z. cesonia* (Silberglied and Taylor, 1973; Stella et al., 2018; Fenner et al., 2019). Interestingly, in *Drosophila* *Wnt7* expression is sexually dimorphic and regulated by *doublesex* during embryonic gonad development of male specific pigment cells (Deshpande et al., 2016). Collectively, the gene expression and Heparin treatment results suggest that *Wnt7* could be a candidate ligand involved in sexually dimorphic wing development in a pierid butterfly.

Previous studies using Heparin assays have largely focused on pattern level changes across the wing, here we show how Heparin impacts the development of the wing scales. The Heparin assays suggest that the Wnt pathway may play a role in the development of structural as well as pigment coloration. Although much is known about the genes responsible for pigment colors such as melanin, little is known about the pathways responsible for generating structural colors (Zhang et al., 2017). Here we show that males lose the ability to produce their UV patterns, but not due to cell fate changes, but do to overall scale shape changes. In nymphalid butterflies, *WntA* has recently been implemented as a modulator of both scale structure and color (Concha et al., 2019). Since *WntA* is not expressed in *Z. cesonia*, but both scale and pattern phenotypes were altered, this suggests that the Wnt pathway in general may play an important role in the development of both scale coloration and overall structure.

### Multiple Pathways Involved in Wing Pattern Development

To date, there are at least three different Wnt signaling pathways: the canonical Wnt/beta pathway, the non-canonical Wnt/JNK (Wnt/PCP), and the Wnt/Ca2+ pathway (Nusse, 2012). In the last decade, it has become evident that two or more of these Wnt signaling pathways often signal within the same cell or
territories to influence cell fate (Range et al., 2013; Range, 2018; Martinez-Bartolomé and Range, 2019). Here we characterized the expression of four Wnt genes in Z. cesonia throughout pupal wing development. As stated above, our Heparin injections showed perturbations in both scale color, which is likely due to transcriptional regulation, and scale structure, which is due to subcellular changes in the actin-myosin cytoskeleton. In many cases, the non-canonical Wnt signaling pathways are involved in morphogenetic processes; therefore, it is possible that multiple Wnt pathways are activated during wing development. We realize that individual Wnts can activate canonical and/or non-canonical pathways depending on the cellular context; it is interesting to note that Wnt11 has been shown to activate the Wnt/JNK pathway in Drosophila development (Uysal-Onganer and Kypta, 2012). Although the expression of Wnt11 is low during wing development in Z. cesonia, its peak expression occurs during days 2 and 3 when scale formation is taking place. On the other hand, Wnt7 expression during wing development, like WntA, has only been seen in butterfly wings and thus the Wnt pathways they may initiate remain unresolved. Together, the handful of Wnt genes expressed suggest that multiple Wnt pathways could be concordantly regulating various aspects of wing development. Further characterization needs to be conducted in order to address which Wnt pathways are deployed during wing development in Z. cesonia.

In addition, other signaling pathways are likely involved and may contribute to the heparin-induced wing changes. Heparin binding is not limited to the Wnt family and impact the diffusion of other signaling genes, such as fibroblast growth factor (FGF), Hedgehog (Hh), and transforming growth factor-β (TGFβ) families. In Z. cesonia Hh, TGFβ-activated kinase 1 (Tak1), branchless (FGF family), and decapentaplegic (dpp) (FGF family) are all expressed during wing development and could have been impacted by heparin treatments. In Junonia butterflies, Hh has been implicated in eyespot pattern development (Tong et al., 2012); however, the morphogens Tak1, branchless, and dpp have yet to be investigated in another butterfly species. Therefore, although Wnt genes have played a major role in Nymphalid wing pattern development and diversification (Concha et al., 2019), the results of this paper suggest that Pierids express Wnts differently and further characterization is needed for other families of morphogens that may be involved in wing patterning.

DATA AVAILABILITY STATEMENT

The names of the repository/repositories and accession number(s) can be found at: NCBI [BIOPROJECT PRJNA587792], GitHub [https://github.com/bcount/Colias10X].

AUTHOR CONTRIBUTIONS

JF and BC designed the study, conducted the pharmaceutical assays, and wrote the manuscript draft. JF collected microscopy and wing image data and prepared RNA-seq libraries. CB and LR-C conducted RNA-seq analyses. RP provided sequencing of the RNA-seq libraries. AR conducted pharmaceutical assays for C. eurytheme. FH conducted phylogenetic and molecular evolution analyses. AM and RR contributed to experimental design and data interpretation. All authors contributed to preparation and editing of the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fevo.2020.00197/full#supplementary-material

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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