Animal Movement and Establishment of Vaccinia Virus Cantagalo Strain in Amazon Biome, Brazil

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To understand the emergence of vaccinia virus Cantagalo strain in the Amazon biome of Brazil, during 2008–2010 we conducted a molecular and epidemiologic survey of poxvirus outbreaks. Data indicate that animal movement was the major cause of virus dissemination within Rondônia State, leading to the establishment and spread of this pathogen.

Outbreaks of vaccinia virus (VACV) infection in dairy cows and dairy workers have been frequently reported in Brazil during the past decade, mainly within the southeastern region (1–6), except for central Goiás (2) and Tocantins State in the northeastern boundary of the Brazilian Amazon biome (Figure 1) (7). VACV Cantagalo strain (CTGV) was first detected in 1999 and associated with several outbreaks (1,5,7). Related strains have been reported in subsequent outbreaks (2–4,6).

Mato Grosso State is partially inserted into the southeastern Amazon biome (Figure 1) and has the largest cattle population of Brazil, ≈26 million (8). It is the main connection with the Amazon states, especially with Rondônia. Rondônia has nearly 11.2 million cattle and is the largest milk producer in the region, yielding >720 million liters/year (8). Therefore, intense livestock trade occurs through the so-called Amazon Gate of Mato Grosso and Rondônia. Both states had previously been considered poxvirus disease–free.

The Study

From June 2008 through June 2010, we investigated outbreaks of poxvirus-related disease on 56 dairy farms of Mato Grosso and Rondônia (Figure 1). A probable case was defined as illness observed in any cattle from these farms that showed clinical signs consistent with a poxvirus-

Figure 1. Location of vaccinia virus Cantagalo strain (CTGV)–related outbreaks. A) Brazilian states where CTGV–related outbreaks have been reported. RJ, Rio de Janeiro; SP, São Paulo; ES, Espírito Santo; MG, Minas Gerais; GO, Goiás; TO, Tocantins; MT, Mato Grosso; RO, Rondônia. B) An enlarged map of Rondônia showing the location of the outbreaks along highway BR-364. The 2 largest dairy regions and the municipalities referred in this article are also shown: 1, Ouro Preto D’Oeste; 2, Teixeirópolis; 3, Urupá; 4, Jaru; 5, Cacaulândia; 6, Ariquemes; 7, Cacoal; 8, Nova Brasilândia D’Oeste; 9, Espigão D’Oeste. Green shading, Amazon biome; light yellow shading, main dairy region; light orange shading, second-largest dairy region. Thin red lines indicate the Brazilian highway network and thick red lines the main highways of the Amazon region. Maps were constructed by using ArcGIS version 9.3.1 (www.esri.com) based on the location coordinates. ARAV, vaccinia virus Araçatuba strain; PSTV, vaccinia virus Passatempo strain.

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related infection: vesicopustular lesions on the udder and/or teats of cows or on muzzle and/or tongue of suckling calves, fever, lymphadenopathy, and remission after ≈3 weeks. A confirmed case was any probable case that was laboratory confirmed by PCR or that had an epidemiologic link to a farm with PCR-positive results. We considered as epidemiologic links the migration of dairy workers, animal movements, and location of neighboring farms. Each inspection generated an epidemiologic investigation form by the animal health agencies of these states. Human infection was noted with a description of lesions on hands, arms, and face.

A total of 52 samples of scabs were collected from cattle on 23 farms. DNA was isolated as described (5) and subjected to PCR by using the EACP primers to detect the full-length hemagglutinin (HA) gene, a marker and subjected to PCR by using the EACP primers to detect the full-length hemagglutinin (HA) gene, a marker for orthopoxvirus infection (1,5,7). To specifically detect CTGV, we used a reverse primer that annealed to the flanking regions of an 18-nt deletion in the HA gene as well as the 15-nt deletion within K2L (1,3,5). DNA from reference CTGV (I) was used as a positive control, and VACV-WR and cowpox virus strain Brighton Red were used as negative controls (5). This strategy successfully identified CTGV-like isolates, as confirmed by DNA sequencing (5,7). Thirty-five samples, representative of the 23 farms (Table 1), were positive for both orthopoxvirus (full-length HA) and CTGV infections. This result strongly suggests that CTGV was the etiologic agent of disease in these farms. For further confirmation, we sequenced the HA, K2L, and C7L genes from 6 samples, as described (7). All sequences showed the 18-nt deletion in the HA gene as well as the 15-nt deletion within K2L found in all CTGV-like isolates (7,9). The relatedness of the virus isolates with CTGV-like isolates was confirmed by analysis of the nucleotide identity matrix (Table 2) and phylogenetic inference (data not shown).

An epidemiologic investigation was conducted by using data available from the 56 epidemiologic investigation forms, analyzing cattle movement forms issued during this period, and interviewing farmers and animal health agency
veterinarians. In addition to the 23 farms with positive samples, 33 other farms had epidemiologic links to CTGV-positive farms. Therefore, our data suggest that CTGV infection was the cause of all outbreaks.

Analysis showed disease prevalence among lactating cows and suckling calves to be 11.83% (1,231/10,407) (Table 1). These rates were lower than those for farms of southeastern states, which reached 50% to 80% (10). Factors that may account for this difference were more efficient surveillance systems in Mato Grosso and Rondônia and the notably larger herd size on Mato Grosso and Rondônia farms. In this study, 55.36% of the affected farms had >100 cattle (mean=253), in contrast to 90% of southeastern farms, which had ≤100 cattle (3,10). The economic losses were substantial because milk production was the major activity at 76.79% of the farms in this study; the income of 93.02% of those was solely based on dairy activities. In addition, 67.85% of the farms reported at least 1 infected dairy worker.

BR-364 is a 4,141.5-km highway with a major veterinary inspection station, the Amazon Gate, at the Mato Grosso–Rondônia border. Affected farms were mostly located along this main road connecting Mato Grosso with the Amazon region (Figure 1, panel A). Despite that, we could not establish an epidemiologic link between the outbreaks in Mato Grosso and Rondônia. In contrast, the spread and establishment of CTGV within Rondônia were evidently related to animal movement along BR-364. Outbreaks in Rondônia (Figure 1, panel B) were first reported in August 2009 after the First Rondônia Dairy Cattle Auction in the municipality of Ji-Paraná (Figures 1, 2), which is part of Rondônia’s major dairy region (11). According to auction records, animals were not traded from other states for this event. Therefore, the circumstances of CTGV entry into Rondônia are still uncertain, but informal animal trade should be considered as well as the pre-event translocation of temporary dairy workers from Mato Grosso, as reported by some farmers.

Cattle trade during the auction launched virus spread throughout the state, which was amplified by subsequent trading and human translocation (Figure 2). Animal movement was associated with the disease spread on 48.57% of farms, whereas migrating dairy workers and neighboring farms were associated with 22.86% and 28.57% of the cases, respectively. Conversely, in Mato Grosso, 13.04% of the farms were related to the virus spread by animal movement, whereas 60.87% were associated with migrating workers. On southeastern farms, dairy workers have also been associated with virus spread, but no quantitative data have been reported (5,10). The distinct pattern in Rondônia could be related to extraordinary intrastate cattle movement. Analysis of 459,884 cattle movement forms issued in 2009 revealed an intrastate translocation of 6,875,031 cattle, which corresponded to 29.34 cattle/km² in contrast to 17.44 cattle/km² in Mato Grosso. The occurrence of new CTGV outbreaks in 2010 links between farms are indicated by arrow colors (red for animal trade, blue for migration of workers, green for neighboring farms) but do not infer geographic distances. Farms were located in these municipalities: JP, Ji-Paraná; JA, Jaru; CCO, Cacoal; OPO, Ouro Preto D'Oeste; ARI, Ariquemes; CAU, Cacaúlandia; TX, Teixeirópolis; NBO, Nova Brasilândia D’Oeste; URU, Urupá. Green star indicates a farm where animals showed clinical signs December 13, 2009, but the owner did not notify the state agency until January 11, 2010.
supports a nonsporadic pattern of infection in Rondônia, highlighting the establishment of this zoonotic infection within the state.

Conclusions

This study investigated the appearance, establishment, and spread of CTGV infection into the Amazon biome. Our findings suggest that animal movement was the main cause of virus long-distance dissemination in Rondônia, whereas the migration of dairy workers was involved in the focal spread. Animal movement plays an essential role in cross-border spread of several disorders worldwide, including bovine tuberculosis, foot-and-mouth disease, and bovine diarrhea viruses (12–15). We suggest animal movement is also a notable risk factor for CTGV dissemination.

According to the inspection station database at the Amazon Gate, 287,906 cattle were moved into Rondônia through BR-364 in 2008–2009, of which 12,501 stayed in Rondônia. Attention is needed to address the westward spread of this zoonosis into the Amazon biome, toward the third largest dairy region of Rondônia surrounding Porto Velho (Figure 1, panel B). Therefore, new regulations and intensified surveillance should be implemented to restrain CTGV spread, particularly in dairy regions with elevated rates of cattle trade for heifer replacement.

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References

1. Damaso CR, Esposito JJ, Condit RC, Moussatche N. An emergent poxvirus from humans and cattle in Rio de Janeiro State: Cantagalo virus may derive from Brazilian smallpox vaccine. Virology. 2000;277:439–49. DOI: 10.1006/viro.2000.0603

2. Nagasse-Sugahara TK, Kisielius JJ, Ueda-Itô M, Curti SP, Figueiredo CA, Cruz AS, et al. Human vaccinia-like virus outbreaks in São Paulo and Goiás States, Brazil: virus detection, isolation and identification. Rev Inst Med Trop Sao Paulo. 2004;46:315–22. DOI: 10.1590/S0036-46652004000600004

3. de Souza Trindade G, da Fonseca FG, Marques JT, Nogueira ML, Mendes LC, Borges AS, et al. Arcatuba virus: a vaccinialike virus associated with infection in humans and cattle. Emerg Infect Dis. 2003;9:155–60.

4. Leite JA, Drumond BP, Trindade GS, Lobato ZI, da Fonseca FG, dos SJ, et al. Passatempo virus, a vaccinia virus strain, Brazil. Emerg Infect Dis. 2005;11:1935–8.

5. Damaso CR, Reis SA, Jesus DM, Lima PS, Moussatche N. A PCR-based assay for detection of emerging vaccinia-like viruses isolated in Brazil. Diagn Microbiol Infect Dis. 2007;57:39–46. DOI: 10.1016/j.diagmicrobio.2006.07.012

6. Megid J, Appolinario CM, Langoni H, Pituco EM, Okuda LH. Vaccinia virus in humans and cattle in southwest region of São Paulo State, Brazil. Am J Trop Med Hyg. 2008;79:647–51.

7. Medaglia ML, Pessoa LC, Sales ER, Freitas TR, Damaso CR. Spread of Cantagalo virus to northern Brazil. Emerg Infect Dis. 2009;15:1142–3. DOI: 10.3201/eid1507.081702

8. Instituto Brasileiro de Geografia e Estatística. Profile of Brazilian municipalities 2010 [cited 2010 Sep 16]. http://www.ibge.gov.br/english/estatistica/economia/perfilmunic/2008/default.shtm

9. Moussatche N, Damaso CR, McFadden G. When good vaccines go wild: feral orthopoxviruses in developing countries and beyond. J Infect Dev Ctries. 2008;2:156–73. DOI: 10.3855/jidc.258

10. Donatele DM, Travassos CEFP, Leite JA, Kroon EG. Epidemiologia da poxvirose bovina no Estado do Espírito Santo, Brasil. Braz J Vet Res Anim Sci. 2007;44:8.

11. Brazilian Ministry of Development and Industry. Ministério do Desenvolvimento, Indústria e Comércio Exterior. 2010 [cited 2010 Sep 17]. http://www.desenvolvimento.gov.br/sitio

12. Fèvre EM, Bronsvoort BM, Hamilton KA, Cleaveland S. Animal movements and the spread of infectious diseases. Trends Microbiol. 2006;14:125–31. DOI: 10.1016/j.tim.2006.01.004

13. Randolph SE, Rogers DJ. The arrival, establishment and spread of exotic diseases: patterns and predictions. Nat Rev Microbiol. 2010;8:361–71. DOI: 10.1038/nrmicro2336

14. Sutmoller P, Casas Olascoaga R. The risks posed by the importation of animals vaccinated against foot and mouth disease and products derived from vaccinated animals: a review. Rev Sci Tech. 2003;22:823–35.

15. Moennig V, Houe H, Lindberg A. BVD control in Europe: current status and perspectives. Anim Health Res Rev. 2005;6:63–74. DOI: 10.1017/S1079247405000035

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