Insights into TREM2 biology by network analysis of human brain gene expression data

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Rare variants in TREM2 cause susceptibility to late-onset Alzheimer’s disease. Here we use microarray-based expression data generated from 101 neuropathologically normal individuals and covering 10 brain regions, including the hippocampus, to understand TREM2 biology in human brain. Using network analysis, we detect a highly preserved TREM2-containing module in human brain, show that it relates to microglia, and demonstrate that TREM2 is a hub gene in 5 brain regions, including the hippocampus, suggesting that it can drive module function. Using enrichment analysis we show significant over-representation of genes implicated in the adaptive and innate immune system. Inspection of genes with the highest connectivity to TREM2 suggests that it plays a key role in mediating changes in the microglial cytoskeleton necessary not only for phagocytosis, but also migration. Most importantly, we show that the TREM2-containing module is significantly enriched for genes genetically implicated in Alzheimer’s disease, multiple sclerosis, and motor neuron disease, implying that these diseases share common pathways centered on microglia and that among the genes identified are possible new disease-relevant genes.

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1. Introduction

Alzheimer’s disease (AD) is the most important cause of presenile dementia worldwide. In the last 3 years, there has been growing evidence for the importance of genetic risk factors in AD pathogenesis with the discovery of 10 new loci from genome-wide association studies (GWAS) (Hollingworth et al., 2011; Naj et al., 2011), and, most recently, the identification of heterozygous rare variants in TREM2 as a cause of increased susceptibility to late-onset AD (Guerreiro et al., 2012; Jonsson et al., 2012). Given that the TREM2 variants reported are associated with an odds ratio of 2.90 to 5.05, similar to ApoE4, and given that this is an entirely new AD locus, these findings are of potentially high importance to the understanding of AD pathogenesis (Guerreiro et al., 2012; Jonsson et al., 2012). However, TREM2 is yet to be extensively studied in this context, and its functions in human brain are not fully understood at present.

TREM2 encodes a trans-membrane glycoprotein, consisting of an extracellular immunoglobulin-like domain, a transmembrane domain, and a cytoplasmic tail. It is known to be expressed on the cell membrane of a subset of myeloid cells, including dendritic cells, osteoclasts, tissue macrophages, and, most importantly for AD, microglia, the resident macrophages of the central nervous system (Colonna, 2003). Since TREM2 itself has no intracellular signaling motif, it is thought to be completely dependent on the adaptor protein TYROBP (also known as DAP12) for its downstream effects. Consistent with this view is the fact that homozygous loss-of-function mutations in either TREM2 or TYROBP result in Nasu–Hakola disease (Online Mendelian Inheritance in Man...
number 605086), a rare disease characterized by painful bone cysts, psychotic symptoms, and progressive presenile dementia (Paloneva et al., 2002). Upon TYROBP stimulation, an ITAM-src kinase-signaling pathway is initiated leading to cellular activation via a range of downstream cascades (Linnartz et al., 2010) and among other possible functions, phagocytosis. Although the endogenous ligand of TREM2 remains unknown, TREM2 has been shown to recognize lipopolysaccharides in the cell wall of bacteria and, on activation, triggers the phagocytic uptake of bacteria and the release of reactive oxygen species (Daws et al., 2003; N’Diaye et al., 2009). Other studies have implicated TREM2 in the phagocytosis of apoptotic material both in vitro and in experimental autoimmune encephalomyelitis, an animal model for multiple sclerosis. The build-up of apoptotic material and the inability of microglia with reduced TREM2 expression to clear it may lead to a frustrated pro-inflammatory phenotype, which would explain why microglia with reduced TREM2 expression, when challenged with apoptotic material also had increased expression of pro-inflammatory cytokines (Stuart et al., 2003).

Despite this information, there is clearly still a great deal that we do not yet know about TREM2, and in particular, there is only limited information about its functions in the human brain. Understanding the predominant biological functions of TREM2 in the brain is clearly of immense importance in the identification of potential therapeutic targets for AD and new candidate genes. In this study, we use a systems-biology approach (Lee et al., 2004; Stuart et al., 2003), based on whole-transcriptome gene expression analysis, to identify co-expressed genes with TREM2 and to begin to address these questions in human brain tissue. We expression-profiled 788 brain samples from 101 neurologically and neuropathologically normal individuals (10 distinct brain regions each) as part of the UK Human Brain Expression Consortium (UKBEC), and used weighted gene co-expression network analysis (WGCNA) to group genes into modules in an unsupervised manner (Langfelder and Horvath, 2008; Oldham et al., 2006; Oldham et al., 2008; Zhang and Horvath, 2005). A large and growing body of evidence now exists to suggest that this approach is extremely useful in identifying modules of biologically related genes that are not just co-expressed but co-regulated (Konopka et al., 2011; Oldham et al., 2008; Rosen et al., 2011; Winden et al., 2009). In this article, we focus our analysis on the TREM2-containing modules and compare through preservation statistics the overlap between TREM2-containing modules in different brain regions and different data sets to better understand the function of this gene and its relationship to other known AD-related genes.

2. Methods

2.1. Human brain samples and analysis of Affymetrix Human Exon 1.0 ST array

Brain samples were collected by the Medical Research Council (MRC) Sudden Death Brain and Tissue Bank (Millar et al., 2007), Edinburgh, UK. All individuals were confirmed to be neuro-pathologically normal by histology performed on sections prepared from paraffin-embedded brain tissue blocks, and the diagnosis was determined by a consultant neuropathologist. All samples had fully informed consent for retrieval and were authorized for ethically approved scientific investigation (Research Ethics Committee number 10/H0716/3).

Total RNA was isolated from human post-mortem brain tissues using the miRNeasy 96 well kit (Qiagen, Manchester, UK). The quality of total RNA was evaluated by the 2100 Bioanalyzer (Agilent Technologies, Wokingham, UK) and RNA 6000 Nano Kit (Agilent Technologies, Wokingham, UK) before processing with the Ambion WT Expression Kit and Affymetrix GeneChip Whole Transcript Sense Target Labeling Assay, and hybridization to the Affymetrix Exon 1.0 ST Arrays (Affymetrix UK Ltd, High Wycombe, UK) following the manufacturers’ protocols. Hybridized arrays were scanned on an Affymetrix GeneChip Scanner 3000 7G (Affymetrix UK Ltd, High Wycombe, UK) and visually inspected for hybridization artefacts. Further details regarding tissue collection, RNA isolation, quality control, and processing have been previously reported (Trabzuni et al., 2011).

All arrays were pre-processed using Robust Multi-array Average (quantile normalization, probe set summary by median polish) with GC background correction (GC-RMA) algorithm (Wu et al., 2004) in Partek Genomics Suite v6.6 (Partek Inc, St. Louis, MO). After re-mapping the Affymetrix probe sets onto human genome build 19 (GRCh37) as documented in the NetAffx annotation file (HuEx-1.0-st-v2 Probeset Annotations, Release 31), we restricted analysis to probe sets that had gene annotation and contained at least 3 probes that hybridized uniquely to a genome position. The resulting expression data were corrected for individual effects (within which are nested post-mortem interval, brain pH, sex, age at death, and cause of death) and experimental batch effects (date of hybridization). Transcript-level expression was calculated for 26,993 genes using Winsorized means (Winsorizing the data below 10% and above 90%). We further restricted to 15,409 transcripts that passed the Detection Above Background (DABG) criteria ($p < 0.001 in at least 50% of samples in at least 1 brain region), had a coefficient of variation >5%, and expression values of at least 5 in samples in at least 1 brain region.

2.2. Analysis of publicly available GEO data sets

Human microarray data sets were downloaded from the Gene Expression Omnibus (GEO; (Edgar et al., 2002)) and ArrayExpress (Parkinson et al., 2011) repositories. The accession numbers of the data sets used were GSE30272 (human brain tissue, Illumina Human 49K Oligo array [HHEBO-7 set], 23), E-MTAB-945 (monocytes, Illumina HumanHT-12 v4.0 expression beadchip (Fairfax et al., 2012), E-TABM-733 (macrophages, Affymetrix GeneChip Human Genome U133 Plus 2.0 (Smith et al., 2009), and GSE43151 (dendritic cells, Illumina HumanHT-12 v4.0 expression beadchip, 26). In the case of all 4 data sets, we restricted our analysis to probes with gene annotation and evidence of detection above background. With regard to the 2 illumina HumanHT-12 v4.0 data sets, gene annotation was based on the re-mapping of probes as performed by the ReMOAT pipeline (Barbosa-Moraes et al., 2010) and probes were required to pass a detection p value of <0.01 in at least 50% of samples in a given data set. In the case of the macrophage data set generated using the Affymetrix GeneChip Human Genome U133 Plus 2.0, gene annotation was based on the current NetAffx annotation file (HG-U133_Plus_2.233.annot.csv), and probes were required to pass a detection p-value of <0.01 in at least 50% of samples with the data set. Finally, in the case of the brain data set (GSE30272), gene annotation was based on the annotation files provided within GEO, and further detection-based probe filtering was performed as described by Colantuoni et al. (Colantuoni et al., 2011). In all cases, the resulting expression data were corrected for experimental batch effects and individual effects (including sex and age when available).

2.3. Identification and removal of outlier samples for network analyses

Gene co-expression analyses are particularly sensitive to the presence of outlier samples, and therefore rigorous quality control procedures were used to ensure the highest possible level of quality
for each dataset. Outliers were identified by visual inspection after clustering the samples using hierarchical clustering with Euclidean distance as the distance measure. Transcripts clusters showed no relation with other covariates (e.g., age, gender, cause of death, post-mortem interval, pH). We also confirmed that the majority of the identified outliers had low interarray correlation, which is defined as the Pearson correlation coefficient of the expression levels for a given pair of transcripts using all available data available (i.e., <3 standard deviations [SDs] of the average interarray correlation). After outlier removal, we repeated the process to check for additional outliers.

2.4. Network construction

We followed a step-by-step network construction and module detection. The network was constructed from the pairwise Pearson correlations between all genes, raising these correlations to a power of 12 while maintaining the information on the correlation signs between genes, thus constructing a “signed” network (Mason et al., 2009). Under this design, only positive correlations among genes are used to construct the network. The selected power used to weight the correlations between genes was chosen to recapitulate scale-free topology (Zhang and Horvath, 2005).

Topological overlap measure (TOM), a pairwise measure of node similarity (i.e., how close the neighbors of a gene are to the neighbors of the other gene) was then calculated. Unlike correlation or adjacency measures, which consider each pair of genes in isolation, TOM considers each pair of genes in relation to all other genes in the network, and it reflects the relative interconnectedness of 2 nodes within their local neighbourhood. The TOM measure allows us to create more robust co-expression relationships (Yip and Horvath, 2007), serving as a filter to exclude spurious or isolated connections during network construction.

Then, by defining a dissimilarity matrix based on TOM (1-TOM), and creating a hierarchical clustering tree based on this dissimilarity matrix, a dynamic tree-cutting algorithm was used to “cut” each dendrogram and define an initial set of modules for each network as branches of the cluster trees (the minimum module size was arbitrarily set to 30 genes). After an initial set of modules was created, correlations among module eigengenes (MEs), defined as the first principal component of the expressions of the genes within the module, are used to merge close modules (we choose a height cut of 0.1 of the ME dendrogram, corresponding to correlation of 0.9, to merge modules). MEs summarize the characteristic expression pattern of a module, and modules with very similar expression profiles will show highly correlated eigengenes. This procedure allows the identification of modules of densely interconnected, coexpressed genes (Oldham et al., 2006; Ravasz and Barabasi, 2003; Zhang and Horvath, 2005), and has been shown to be placing functionally related genes into groups (Eisen et al., 1998).

2.5. Preservation analysis

To validate module construction for a reference network, it is desirable to show that modules are reproducible (or preserved) in an independent network. To this end, a simple cross-tabulation analysis of module memberships, and the related Fisher’s exact test p value can be calculated. In general, to determine whether a module found in a reference data set can also be found in a test data set, module preservation statistics implemented in the WGCNA R software can be used.

For each pair of brain tissues, the overlap between all modules was calculated along with the probability of observing such overlap by chance. The significance of module overlap was assessed using a Fisher’s exact test (also known as hypergeometric test) for each of the pairwise overlap. To account for multiple comparisons, a Bonferroni correction was applied on the basis of the number of modules in the comparison networks.

To validate modules consistency across tissues, and to evaluate specifically how well the TREM2-containing modules were preserved across tissues, we calculated a composite Z summary statistic that aggregates different module preservation statistics (Langfelder et al., 2011). The composite Z summary statistic can be used to determine whether module nodes remain highly connected in the test network, and to determine whether the connectivity pattern between nodes in the reference network is similar to that in the test network. We used a permutation-based procedure in WGCNA to calculate a Z summary for each module (specifically, randomly permuting the module labels in the test network and calculate corresponding preservation statistics). The higher the preservation Z summary is for a given module in a specific brain tissue, the stronger the evidence that the module is preserved in another brain region. Under the null hypothesis of no module preservation, the preservation Z summary follows an approximately standard normal distribution, and comprehensive simulation studies led to the following thresholds: a module shows no evidence of preservation if its Z summary is less than 2; a Z summary greater than 5 (or 10) indicates moderate (strong) module preservation (Langfelder et al., 2011).

2.6. Network connectivity and module memberships

For each network the whole network connectivity (also known as “degree”) for all genes can be calculated, and it describes the relative importance of each gene in the network (Zhang and Horvath, 2005). To identify the most highly connected, or most central genes within each module, referred to as “hubs”, measure of module memberships (MMs) can be used. MMs are derived by first summarizing the expression levels of all genes in each module by the corresponding ME (that is, the first principal component obtained by singular value decomposition). MMs for each gene with respect to each module are defined as the Pearson correlation between the expression level of a given gene and the module ME. This quantity describes the extent to which a gene “belongs” to a module. Values of MM range from −1 (perfect anti-correlation) to 1 (perfect correlation). We used MMs to characterize the importance of TREM2 within each module for each brain tissue.

2.7. Gene set enrichment analysis with DAVID

To evaluate the biological and functional relevance of coexpressed genes within the TREM2-containing modules, we used DAVID v6.7 (http://david.abcc.ncifcrf.gov/), the database for annotation, visualization, and integrated discovery (Huang da et al., 2009b). We examined the overrepresentation (i.e., enrichment) of the 3 Gene Ontology (GO) categories (biological processes, cellular components, and molecular function) for each list of coexpressed genes with TREM2 for each tissue by comparing numbers of significant genes annotated with this biological category with chance.

3. Results

3.1. Regional distribution of TREM2 mRNA expression in human brain

The human data used here were provided by the UK Human Brain Expression Consortium (Trabzuni et al., 2011) and consisted of 101 control post-mortem brains. All samples originated from individuals with no significant neurological history or neuropathological abnormality, and all were collected by the MRC Edinburgh Brain Bank (Millar et al., 2007) ensuring a consistent dissection
protocol and sample handling procedure. A summary of the available demographic details of these samples, including a thorough analysis of their effects on array quality, is provided by Trabzuni et al. (Trabzuni et al., 2011). In brief, samples originated from individuals with a male: female ratio of 77:24 (3.2), mean age at death of 51 years (range, 16–83 years), and modal cause of death of ischemic heart disease (59%; full details provided in Supplementary Tables 1 and 2). Up to 10 brain regions were sampled from each individual, resulting in 788 Affymetrix Human Exon 1.0 ST arrays after rigorous removal of outliers. The regions sampled were BA 8 and 9 (frontal cortex [FCTX]), BA17 (occipital cortex [OCTX]), BA 21 and 22 (temporal cortex [TCTX]), intralobular white matter (WHMT), hippocampus (HIPP), thalamus (at the level of the lateral geniculate nucleus [THAL], putamen (at the level of the anterior commissure [PUTM]), substantia nigra (SNIG), the inferior olivary nucleus (sub-dissected from the medulla [MEDU]) and the cerebellar cortex (CRBL).

Fig. 1A shows the regional distribution of mRNA expression for TREM2 at the gene level for human brains. This demonstrated significant regional differences in TREM2 gene expression with a 4.96-fold difference (paired t test p value = 6.63 × 10^{−32}) between white matter (the highest-expressing region) and cerebellum (lowest-expressing region). Given the age-dependent and sex-biased incidence of Alzheimer’s disease (AD), we investigated TREM2 expression in each brain region for age- or sex-related changes in expression, while accounting for the potential confounding effects of post-mortem interval, cause of death, brain pH, and experimental batch effects. We found suggestive age-related increases in TREM2 expression in 3 brain regions: MEDU (p = 0.043), THAL (p = 0.037), and SNIG (p = 0.048, Fig. 1B). Although these findings could not be replicated using the only other publicly available data set covering these brain regions (Kang et al., 2011) (Supplementary Fig. 1), it should be noted that this particular data set contains samples originating from only 18 adults, whereas our
study is based on samples originating from 101 individuals. There was no evidence of sex-biased expression of TREM2 in any brain region.

3.2. Weighted Gene Co-expression Network Analysis identifies a TREM2-containing module, which is highly preserved in all brain regions

To gain insights into the functional organization of the brain transcriptome, we used the WGCNA package (Langfelder and Horvath, 2008; Zhang and Horvath, 2005) and focused on TREM2. This analytic approach uses the degree of gene neighbourhood sharing, as defined on the basis of co-expression relationships, to identify groups or modules of genes that are highly co-expressed and (by implication) functionally related (Geschwind and Konopka, 2009; Miller et al., 2008; Oldham et al., 2008; Winden et al., 2009). We used WGCNA to analyze gene expression data on 15,409 transcripts equating to 13,706 genes (passing filtering as described in Methods) to construct signed weighted gene co-expression networks for each of the 10 brain regions (see Methods). On this basis, we identified between 13 and 34 modules per region, depending on the brain region (Table 1). Whole network connectivity for each tissue and TREM2-specific connectivity is summarized in the Supplementary Table 3. Before proceeding to investigate TREM2-containing modules, we checked the consistency of all the identified modules across the 10 brain regions and particularly within the 3 available neocortical regions (FCTX, TCTX, and OCTX). As would be expected, we observed particularly high module conservation among the 3 cortical regions, as shown in (Fig. 2A and B). These findings are consistent with the study by Oldham et al. (Oldham et al., 2008) that showed similar co-expression preservation in 3 different regions in the brain.

Having established the robustness of the whole network, we checked the preservation statistics for the TREM2-containing modules across all 10 brain regions. Table 2 shows the number of shared genes (and percentages) among the TREM2-containing modules from each region. We identified 77 genes present in all 10 regional TREM2-containing modules. We used Fisher’s exact test to test for significant overlap in module membership and found that all 45 pairwise comparisons were highly significant \( p < 1.1 \times 10^{-3} = 0.05/45 \) after Bonferroni correction, indicating a high overlap in module membership among all TREM2-containing modules. We also used a permutation test procedure implemented in the WGCNA R package to evaluate preservation across multiple tissues. This procedure produces a summary preservation Z statistic (plotted in Fig. 3) that takes into account connectivity patterns between nodes, but not just overlaps in module membership on cross- tabulation (see Methods) (Langfelder et al., 2011). We found that the TREM2-containing module is highly conserved in all brain regions.

3.3. Independent human brain dataset replicates gene membership found within the TREM2-containing module

For validation purposes, we used an independent and publicly available dataset (Colantuoni et al., 2011), collected from human dorsolateral prefrontal cortex in an extensive series of samples collected from fetal development through aging (GSE30272). We restricted our analysis to adult brain samples (individuals aged 16 years and more, \( n = 175 \)) to maximize comparability with the UKBEC dataset. Using the same parameters for network construction as were applied to the UKBEC data set, we constructed a signed WGCNA for 15,250 transcripts (representing 10,740 gene symbols). On this basis, we identified 46 modules (size ranging between 37 and 1,969 transcripts; whole network connectivity between 4.6 and 146.0). The TREM2-containing module in this dataset contained 255 unique genes symbols (representing 315 transcripts). We observe a highly significant overlap between genes in this module and the genes within the corresponding TREM2-containing module in the frontal cortex of the UKBEC dataset. More specifically, we observe 95 overlapping gene symbols (2.2 genes expected by chance, Fisher’s exact test \( p < 7 \times 10^{-12} \)).

3.4. TREM2-containing module is partially preserved in blood-derived macrophages

Given the importance of TREM2 in the pathogenesis of AD, there is an urgent need to better understand its functions. Although ideally this would be achieved by studying adult human microglia, practically this is challenging. This led us to assess the validity of using other more accessible and potentially relevant human cell types, namely monocytes, macrophages, and dendritic cells. To do this, we used publicly available human whole-genome gene expression data generated on the Illumina HT12 v4 beadchip array and the Affymetrix U133 Plus 2.0 array with large numbers of healthy individuals: (monocytes, E-MTAB-945, \( N = 188 \) (Fairfax et al., 2012); macrophages, E-TABM-733, \( N = 16 \) (Smith et al., 2009); dendritic cells, GSE34151, \( N = 69 \) (Barreiro et al., 2012)). In each case, we started by checking the evidence for TREM2 expression in these cell types. In keeping with the existing literature (on the basis of microarray data), we were unable to convincingly detect TREM2 expression in monocytes (percentage of samples passing a detection \( p \text{ value} > 0.01 = 4.1\% \)), but could detect expression in nonstimulated macrophages and dendritic cells.

We next used WGCNA to assess gene expression data on 11,658 genes in macrophages and 8300 genes in dendritic cells (passing filtering as described in Methods) to construct signed weighted gene co-expression networks (see Methods). Focusing on the TREM2-containing module in macrophages and dendritic cells, we found that TREM2 had high module membership (macrophages and dendritic cells, 61\(^{\text{th}}\) quantile) but was not a “hub” gene in either cell type. We next assessed preservation of the TREM2-containing module across the 2 cell types and in post-mortem human brain tissue. We found that whereas there was no significant preservation of the TREM2-containing module between dendritic cells and macrophages (Fisher’s exact test \( p \text{ value} = 0.38 \)) or dendritic cells and brain (Fisher’s exact test \( p \text{ value} = 0.77 \)), there were 13 genes in common between the TREM2-containing
modules in macrophages and hippocampus (5.62 genes expected by chance, Fisher’s exact test $p = 4.52 \times 10^{-3}$). Although this does suggest significant preservation of the TREM2-containing module across these 2 tissue/cell types, it is much weaker than that observed between the 2 brain data sets analyzed (Fisher’s exact test $p$ value $< 7 \times 10^{-142}$).

### 3.5. TREM2-containing modules in human brain correspond to a microglial signature

Consistent with the expression of TREM2 in microglia, the known microglia markers CCR1, ITGAM, AIF1, FCER1G, and CD68 were present within the TREM2-containing module in all brain regions.
including the independent replication data set. In the case of each of these genes, we measured their module membership (MM), as defined by how well the expression pattern of each gene within the module correlates with the first principal component of gene expression for that module, termed the module eigengene (see Methods). In all cases, the mean MM across all brain regions was above the median (CX3CR1 56th; ITGAM 93rd; AIF1 82nd, FCER1G 78th, and CD68 97th quantile). These findings and, in particular, the high MM of microglia markers, suggest that the TREM2-containing modules in human brain strongly relate to the gene expression signature of microglia. It should also be noted that searching for genes related to the major neurotransmitter signaling systems (namely glutamatergic, GABAergic, dopaminergic, cholinergic, serotonergic, histaminergic, glycineergic, purinergic, adenosinergic, noradrenergic, and adrenergic) demonstrated only the presence of the purinergic and adenosinergic receptors (ADORA3, P2RX4, P2RY12, and P2RY13) within the TREM2-containing modules, suggesting that this module is very unlikely to relate to any neuronal population. Similarly, no oligodendrocyte markers (OLIGO1, OLIGO2, MAG, MOG, and MOBP) were detected and of the astrocyte markers (GFAP, S100B, and ALDOC); only GFAP (a marker of activated astrocytes) was present in the TREM2-containing module in white matter (quantile = 38th).

We also investigated gene membership within the TREM2-containing modules to see if we could infer the predominant microglial activation state relevant to these modules. Despite the distinct origin of microglia (Ginhoux et al., 2010), these cells (like peripheral macrophages) have been classified as M1- or M2-like (with sub-classification of M2a, M2b, and M2c). Whereas the M1 phenotype is adopted in response to activation with lipopolysaccharide or interferon-γ and denotes a pro-inflammatory state, the

Table 2
Number of genes in the TREM2-containing module for each tissue and overlap with TREM2-containing modules in other tissues

| Tissue | Genes in module (n) | CRBL | FCTX | HIPP | MEDU | OCTX | PUTM | SNIG | TCTX | THAL | WHMT |
|--------|---------------------|------|------|------|------|------|------|------|------|------|------|
| CRBL   | 226                 | —    | 132  | 145  | 145  | 208  | 147  | 113  | 196  | 151  | 218  |
| FCTX   | 151                 | 132  | —    | 117  | 144  | 149  | 127  | 144  | 100  | 143  | 122  |
| HIPP   | 175                 | 145  | 117  | —    | 112  | 142  | 127  | 144  | 100  | 143  | 122  |
| MEDU   | 560                 | 208  | 144  | 142  | —    | 146  | 176  | 176  | 122  | 176  | 126  |
| OCTX   | 186                 | 147  | 127  | 136  | 176  | —    | 106  | 119  | 139  | 135  | 176  |
| PUTM   | 133                 | 113  | 100  | 101  | 112  | 106  | —    | 119  | 106  | 104  | 125  |
| SNIG   | 377                 | 196  | 143  | 158  | 283  | 162  | 119  | —    | 173  | 165  | 294  |
| TCTX   | 257                 | 151  | 134  | 196  | 139  | 106  | 173  | —    | 134  | 194  | 154  |
| THAL   | 187                 | 151  | 125  | 176  | 104  | 104  | 134  | —    | 177  | —    | —    |
| WHMT   | 728                 | 218  | 149  | 169  | 426  | 176  | 294  | —    | 194  | 177  | —    |

Percentages are quoted as proportion of the number of genes in the TREM2-containing module in the tissue indicated in the rows. Overlaps were tested using Fisher's exact test at a Bonferroni-adjusted value of \( p < 1.1 \times 10^{-3} = 0.05/45 \). All overlaps were found to be highly significant.

Key: CRBL, cerebellum; FCTX, frontal cortex; HIPP, hippocampus; MEDU, medulla (specifically inferior olivary nucleus); OCTX, occipital cortex (specifically primary visual cortex); PUTM, putamen; SNIG, substantia nigra; TCTX, temporal cortex; THAL, thalamus; WHMT, intralobular white matter.

Fig. 3. Preservation statistic (Z summary) of each regional TREM2-containing module across the equivalent modules in all other human brain regions.
M2 phenotype is adopted in response to interleukin-4 (Gordon, 2003). We investigated features of 62 genetic markers (as described elsewhere (David and Kroner, 2011); Supplementary Table 4) of the M1 and M2 activation states. Of these, 17 genes were members of at least 1 of the regional TREM2-containing modules. These included the following: 5 genes associated with the M2a activation state (IGF1, PDGFB, PDGFC, TGFBI, and MSRI); 7 genes contributing to the major histocompatibility complex (MHC) class II protein complex which has been associated with both the M1 and M2b activation state (CD74, HLA-DMA, HLA-DMB, HLA-DPA1, HLA-DPB1, HLA-DQA2, and HLA-DRA); and 4 genes associated with the M1 activation state (FCGR2A, FCGR2B, FCGR3B, and CXCL10); and finally CD86, which (like MHC class II) has been associated with both the M1 and M2b states. Because we detected markers for both the M1 and M2 activation states, we used MM as a basis for hierarchical clustering of genes to better assess microglia state in different brain regions. These data are presented in Fig. 4 and show that the clustering of genes to better assess microglia state in different brain regions. These data are presented in Fig. 4 and show that the clustering of genes to better assess microglia state in different brain regions.

3.6. TREM2 is a “hub” gene within the TREM2-containing module in the hippocampus and white matter among other brain regions

Within each TREM2-containing module, WGCNA allows us to identify the most highly connected genes, or central genes, referred to as “hubs,” as defined by a high MM (as described above). The MM for TREM2 was above the median in all brain regions, including the replication data set (88th quantile within this module). In fact, TREM2 was in the top 90th quantile in the hippocampus, white matter, medulla, putamen, and substantia nigra (Table 3), indicating that TREM2 is a clearly a hub gene in these 5 tissues.

Based on this definition of “hub,” namely, MM in the top 90th quantile in at least 5 brain regions, we identified a further 15 genes of interest: APBB1IP, C3AR1, CYBB, DOCK2, DOCK8, NCKAP1L, ADORA3, CD68, FYB, PTPRC, TBXAS1, ADAM28, ITGAM, METTL9, and CD84. This would suggest that although 980 genes are identified as members of the TREM2-containing module across all 10 brain regions, TREM2 is actually part of a select group of genes driving module function.

3.7. TREM2-containing modules in the human brain are enriched for genes implicated not only in the innate but also the adaptive immune system

Gene Ontology (GO) enrichment analysis was performed on the TREM2-containing modules in each brain region using the Database for Annotation, Visualization and Integrated Discovery (DAVID) (Huang da et al., 2009a, 2009b). The results of this analysis are summarized in Table 4 and provided in full in Supplementary Table 5. This demonstrated that the TREM2-containing modules from all 10 brain regions were highly enriched for genes related to immune response (minimum Bonferroni-corrected p value = 1.65 × 10^{-45}). This finding is clearly consistent with the known expression of TREM2 on microglia (Colonna, 2003). In keeping with the existing literature, GO enrichment analysis provided strong evidence for a role for TREM2 in the innate immune response (minimum Bonferroni-corrected p value = 3.44 × 10^{-12}) and, more specifically, lysosome activity in all brain regions. This was demonstrated by the significant enrichment of genes classified by GO to lytic vacuoles and lysosomes in 7 of the 10 regional TREM2-containing modules including the hippocampus, frontal cortex, and white matter (minimum Bonferroni-corrected p value = 2.96 × 10^{-4}). Interestingly, we found that despite the weak (but significant) preservation of the TREM2-containing module in macrophages, there was a significant overrepresentation of genes related to lytic vacuoles and lysosomes in these data too (minimum Bonferroni-corrected p value in human brain = 2.96 × 10^{-4}; macrophages = 2.10 × 10^{-2}), suggesting that modulation of lysosome activity is a well-preserved TREM2-related function though the underlying gene network may be different (Supplementary Fig. 2).

We also found evidence in support of links to the adaptive immune system. There was significant enrichment of genes involved in the regulation of lymphocyte activation in all 10 regional TREM2-containing modules (minimum Bonferroni-corrected p value = 6.42 × 10^{-7}) with evidence for interactions with both B and T cells. The GO term “B cell mediated immunity” was significant in all 10 regional TREM2-containing modules (minimum Bonferroni-corrected p value = 9.16 × 10^{-5}), whereas “T cell activation” was featured in 9 of 10 modules (minimum Bonferroni-corrected p value = 8.75 × 10^{-5}). Other major microglial functions were also

![Fig. 4. Hierarchical clustering of microglial markers across all 10 brain regions based on module membership. Genetic markers of microglial state that were not detected in a given regional TREM2-containing module are depicted in gray (and have a module membership of 0).](image-url)
highlighted in this analysis with “antigen processing and presentation via MHC class II” appearing as a highly significant term in all 10 regional TREM2-containing modules (minimum Bonferroni-corrected $p$ values $= 1.15 \times 10^{-3}$) and “the regulation of cytokine production” significant in 8 of 10 modules (minimum Bonferroni-corrected $p$ values $= 7.68 \times 10^{-7}$). Investigating the TREM2-containing modules for the enrichment of GO molecular functions, highlighted the involvement of G-protein-coupled nucleotide receptor signalling in 8 of 10 brain regions (including the hippocampus, Bonferroni corrected $p$ value $= 3.46 \times 10^{-4}$). Interestingly, genes related to purinergic signaling featured more prominently in the TREM2-containing modules than those involved in IgG binding, cytokine signaling, or MHC class II receptor activity.

3.8. Insights into TREM2-related signaling pathways

To obtain insights into TREM2-related signaling, we chose to focus on a “core set” of 156 TREM2-related genes (defined on the

### Table 3

Summary of the module membership (MM) within the TREM2-containing modules for each brain tissue for TREM2 and all transcripts assigned to TREM2-containing modules

| Data set | Genes in module (n) | TREM2 | Summary statistics of the MM of the transcripts within the TREM2-containing module |
|----------|--------------------|-------|-----------------------------------------------------------------|
|          | MM                 | 1−p<sup>th</sup> quantile | Min | First quartile | Median | Mean | Third quartile | Max |
| UKBEC    | CRBL 226           | 0.82  | 0.17 | 0.29 | 0.55 | 0.68 | 0.67 | 0.78 | 0.95 |
|          | FCTX 151           | 0.78  | 0.26 | 0.32 | 0.64 | 0.72 | 0.71 | 0.78 | 0.94 |
|          | HIPP 175           | 0.89  | 0.09 | 0.20 | 0.68 | 0.76 | 0.74 | 0.85 | 0.93 |
|          | MEDU 560           | 0.92  | 0.03 | 0.17 | 0.51 | 0.68 | 0.65 | 0.8  | 0.95 |
|          | OCTX 186           | 0.76  | 0.25 | 0.25 | 0.55 | 0.66 | 0.65 | 0.76 | 0.92 |
|          | PUTM 133           | 0.88  | 0.05 | 0.4  | 0.64 | 0.73 | 0.71 | 0.79 | 0.92 |
|          | SNIG 377           | 0.88  | 0.07 | 0.31 | 0.55 | 0.67 | 0.67 | 0.8  | 0.94 |
|          | TCTX 257           | 0.79  | 0.15 | 0.22 | 0.57 | 0.65 | 0.66 | 0.75 | 0.94 |
|          | THAL 187           | 0.86  | 0.12 | 0.41 | 0.63 | 0.74 | 0.71 | 0.81 | 0.94 |
|          | WHMT 728           | 0.9   | 0.03 | 0.03 | 0.43 | 0.6  | 0.59 | 0.75 | 0.95 |
|          | Colantuoni et al. 315 | 0.8  | 0.12 | 0.20 | 0.51 | 0.62 | 0.62 | 0.74 | 0.92 |

1−p<sup>th</sup> quantile = rankMM(TREM2)/module size. The p<sup>th</sup> quantiles > 90<sup>th</sup> quantile are highlighted in boldface type. For these tissues, TREM2 shows a major role within the module (hub gene).

Key: CRBL, cerebellum; FCTX, frontal cortex; HIPP, hippocampus; MEDU, medulla (specifically inferior olivary nucleus); OCTX, occipital cortex (specifically primary visual cortex); PUTM, putamen; SNIG, substantia nigra; TCTX, temporal cortex; THAL, thalamus; UKBEC, UK Human Brain Expression Consortium; WHMT, intralobular white matter.

### Table 4

Summary of the results of GO enrichment analysis performed on the TREM2-containing modules in each brain region

| Term                                | No. of Regions | Bonferroni corrected p-Value |
|-------------------------------------|----------------|-----------------------------|
| CRBL                                | 2,080-34       | 3,165-23                   |
| FCTX                                | 3,165-23       | 3,165-23                   |
| HIPPO                               | 3,165-23       | 3,165-23                   |
| OCTX                                | 3,165-23       | 3,165-23                   |
| PUTM                                | 3,165-23       | 3,165-23                   |
| SNIG                                | 3,165-23       | 3,165-23                   |
| TCTX                                | 3,165-23       | 3,165-23                   |
| THAL                                | 3,165-23       | 3,165-23                   |
| WHMT                                | 3,165-23       | 3,165-23                   |

Key: CRBL, cerebellum; FCTX, frontal cortex; HIPPO, hippocampus; MEDU, medulla (specifically inferior olivary nucleus); OCTX, occipital cortex (specifically primary visual cortex); PUTM, putamen; SNIG, substantia nigra; TCTX, temporal cortex; THAL, thalamus; WHMT, intralobular white matter.
basis of module membership above the 80th quantile in at least 1 brain region). We used this set to look for enrichment of canonical gene sets defined by the Kyoto Encyclopedia of Genes and Genomes (KEGG) and relating to known pathways (Kanehisa and Goto, 2000; Kanehisa et al., 2012). We identified 11 pathways with a Bonferroni-corrected p value of < 0.05. After accounting for significant overlaps in the gene memberships of these pathways (as calculated within the Molecular Signatures Database v3.1, http://www.broadinstitute.org/gsea/msigdb/index.jsp), we identified 2 independent pathways of interest, the “Fc gamma R-mediated phagocytosis” (Bonferroni-corrected p value = 4.72 \times 10^{-4}) and “Systemic lupus erythematosus” pathways (Bonferroni-corrected p value = 3.46 \times 10^{-4}). These are depicted in Fig. 5A and B with the proteins/complexes containing “core” TREM2-related genes highlighted with a red box (Kanehisa et al., 2012; Kanehisa and Goto, 2000).

Fig. 5. Enrichment of “core” TREM2-related genes within Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways. (A) “Fc gamma R-mediated phagocytosis” pathway with proteins/complexes containing “core” TREM2-related genes highlighted with a red box (Kanehisa et al., 2012; Kanehisa and Goto, 2000). (B) “Systemic lupus erythematosus” pathway with proteins/complexes containing “core” TREM2-related genes highlighted with a red box (Kanehisa et al., 2012; Kanehisa and Goto, 2000).
Combining these pathways together would explain how TREM2 located on microglia might act to respond to apoptotic material and immune complexes in their micro-environment through the recognition of the C1Q complex and signal via the ITAM-src-signaling pathway to initiate phagocytosis of the material, but also potentially other processes (including antigen presentation). Because 2 ITAM-containing adaptor proteins are members of the “core” gene set, namely FCER1G and TYROBP, in theory at least either could associate with TREM2 protein. Similarly, on the basis of TREM2-module membership, downstream signaling is most likely to occur through the src kinase, LYN (a member of the “core” gene set). However, it is worth noting that HCK is a member of the TREM2-containing module in medulla and white matter suggesting potential regional heterogeneity in the use of src kinases.

Because the use of canonical pathways is limited by our existing knowledge base, we also chose to visualize gene interconnection within the TREM2-containing module using VisANT. Connections
among genes was defined on the basis of TOM and genes were “connected” if the TOM was greater than 0.10. Given the involvement of TREM2 in late-onset AD, we focused on the TREM2-containing module in the hippocampus, a region that undergoes severe neurodegeneration, and contrasted this with a relatively unaffected region, the cerebellum (Fig. 6A and B). Based on a TOM of >0.10, we identified 45 genes connected to TREM2 in hippocampus and similarly 45 genes in cerebellum. As would be expected, both gene lists included members already known to be part of the TREM2 intracellular signaling pathway (including TYROBP and SYK) and many genes that were previously identified within the canonical KEGG pathways. Interestingly, both lists also included 2 purinergic receptors, P2RY12 and P2RY13. The P2RY12 is already known to be expressed on microglia and has been shown to play a critical role in the chemotactic response to ATP released by apoptotic and dying cells (Haynes et al., 2006; Pocock and Kettenmann, 2007). In the case of the hippocampus, this list included 9 genes (NCKAP1L, APBB1IP, CSF1R, DOCK2, DOCK8, ITGAX, C1QB, LAPTM5, and METTL9) with “direct” connection to TREM2 (as marked in pink and using the cut off of TOM >0.10). Only 2 such genes were identified in cerebellum, NCKAP1L and ITGAX. Interestingly, 5 of the genes with “direct” connection in hippocampus have been implicated in cytoskeletal rearrangements and cell adhesion. NCKAP1L (also known as HEM1) encodes a member of the HEM family of tissue-specific trans-membrane proteins and forms part of the Scar/WAVE complex, which plays an important role in regulating cell shape and has been shown to be essential for actin polymerization and myosin regulation during neutrophil chemotaxis (Weiner et al., 2006). Similarly DOCK2 is known to be involved in remodeling of the actin cytoskeleton required for lymphocyte migration (Sanui et al., 2003), and the biological functions of DOCK8 include regulation of cell migration and morphology (Harada et al., 2012; Mizesko et al., 2013). APBB1IP (also known as RIAM) encodes the Rap1-GTP-interacting adaptor molecule (RIAM) and has been implicated in mediating changes in the actin cytoskeleton after integrin-mediated adhesion acting through the Vav2-RhA-ROCK-myosin light chain pathway (Hernandez-Varas et al., 2011; Lefaucheur et al., 2004). Finally, ITGAX encodes the integrin alpha X chain protein, which forms part of the alpha X beta 2 complex that has been implicated in the adherence of neutrophils and monocytes to stimulated endothelium cells, and in the phagocytosis of complement-coated particles (Hynes, 1992; Schoenwaelder and Burridge, 1999; van Lookeren Campagne et al., 2007). Although, the remaining 4 genes (CSF1R, LAPTM5, C1QB, and METTL9) had no obvious common features, the identification of CSF1R is notable, as the TREM2-TYROBP protein complex has recently been implicated in CSF1R signaling providing a biological basis for the high connection between CSF1R and TREM2 (Otero et al., 2009). C1QB has already been highlighted within the canonical pathway for “Systemic lupus erythematosus” and LAPTM5 (lysosomal protein transmembrane 5) is a transmembrane receptor that is associated with lysosomes. Using the information provided by both the investigation of canonical pathways and data-driven network construction, it is possible to build a potential TREM2-signaling pathway for human microglia (Fig. 6C) characterized by a central role for TREM2 in the response to C1Q-containing complexes and ATP/ADP in the micro-environment and the subsequent cytoskeletal changes that are required not only for phagocytosis, but also surveillance and chemotaxis.

3.9. Relative enrichment of genes already linked to Alzheimer’s disease among genes within the TREM2-containing modules

The identification of variants in TREM2 as an important cause of late-onset Alzheimer’s disease raises the possibility that genes that are highly co-expressed with TREM2 might also be candidate genes for AD. One way of assessing the validity of this approach is to look at the enrichment of genes already genetically linked to AD among genes within the TREM2-containing modules in human brain. Using HUGEnavigator (http://www.hugenavigator.net/HuGENavigator/home.do) (Yu et al., 2008), which uses text mining algorithms to search PubMed and to create an up-to-date knowledge base for exploring genetic associations, we can identify 1499 genes that have been linked to AD. From this list, 1004 genes were studied within our WGCNA analysis of human brain. We looked for enrichment of these AD-related genes within a “core set” of 156 TREM2-related genes (defined on the basis of module membership above the 80th quantile in at least 1 brain region). We found a highly significant enrichment of AD-related genes among the “core set” of TREM2-connected genes ($\chi^2$ test with Yates correction $p$ value $= 3.67 \times 10^{-10}$). Among the genes accounting for this enrichment were MS4A6A, which is strongly implicated in AD through recent GWAS (Hollingworth et al., 2011; Naj et al., 2011) and APBB1IP (Morgan et al., 2007) (full details provided in Supplementary Table 6). We used a similar approach to look for enrichment of genes implicated in other neurological disorders, namely Parkinson’s disease (PD), multiple sclerosis (MS), and motor neuron disease (MND), on the basis that TREM2-related microglial activity might be a general contributing factor in the pathogenesis of multiple disorders (Supplementary Table 6). Although there was no evidence for enrichment of PD-related genes ($\chi^2$ test with Yates correction $p$ value $= 0.92$), there was evidence for enrichment of MS ($\chi^2$ test with Yates correction $p$ value $= 7.50 \times 10^{-15}$) MND-related genes ($\chi^2$ with Yates correction, $p = 4.85 \times 10^{-5}$). These findings suggest that among the list of “core genes,” there might be as-yet-unidentified AD-related genes, and also (albeit indirectly) validate previously identified AD candidate genes.

4. Discussion

The discovery of variants in TREM2 as a predisposing factor for late-onset AD has opened up a potential new avenue for the understanding of this disease (Guerreiro et al., 2012; Jonsson et al., 2012). To realize that potential an improved appreciation for the “normal” function of TREM2 in the human brain is required, particularly because increased risk is thought to be mediated through reduced TREM2 function. To this end, we used a large microarray data set generated from neuropathologically control post-mortem tissue to analyze TREM2 expression, and we used WGCNA analysis to identify highly co-expressed genes in 10 brain regions, including the hippocampus. This approach, albeit not without its drawbacks, allowed us to obtain insights into TREM2 function, not in an isolated cell type but in the context of whole human brain tissue. We demonstrate that a highly preserved TREM2-containing module of gene expression exists in all brain regions and relates to a microglial expression signature. Importantly, TREM2 is a hub gene in this module in 5 brain regions, including the hippocampus, suggesting that it is capable of driving functional processes related to TREM2, but also the adaptive immune system widens the functional processes related to TREM2. Furthermore, close inspection of the genes with the highest connectivity to TREM2, provided insights into TREM2-related signaling, suggesting that this gene has a central role in mediating changes in the cytoskeletal structure in response to damage-related signals (such as C1Q and ATP/ADP) within the microglial micro-environment. Finally, we demonstrate significant enrichment of candidate genes for AD, MS, and, to a lesser extent, MND within the core genes of the TREM2-containing
modules, implying, first, that these neurological diseases may share common pathways centred on microglial function and, second, that among the list of genes identified by this study are as-yet-unknown disease-relevant genes.

The basis of this study was microarray data generated from 788 human brain samples originating from 101 neurologically and neuropathologically control individuals collected from a single brain bank, the MRC Sudden Death Brain Bank in Edinburgh, and covering 10 brain regions. These data set allowed us to confidently quantify TREM2 expression across the human brain, demonstrate significant regional differences, and assess age-related changes in expression. Given that previous studies, not to mention this one, have found that TREM2 is almost exclusively expressed on microglia in human brain (Colonna, 2003), it seems likely that the regional differences in TREM2 expression relate to regional differences in either overall microglial density or a subpopulation of microglia. Certainly the higher expression of TREM2 in white matter versus gray matter samples, and the high TREM2 expression in hippocampus, mirror known regional differences in microglial density in human brain (Mittelbronn et al., 2001). This interpretation of the TREM2 expression data would imply that the suggestive age-related increase in TREM2 expression found in 3 brain regions may also reflect changes in microglial density, again perhaps of a specific sub-population. Although we were unable to find evidence in support of this expression pattern within existing publicly accessible data sets, there is evidence in support of age-related increases in microglia in the normal human brain (Conde and Streit, 2006) and a growing literature to suggest age-related changes in microglial function (Miller and Streit, 2007).

We next used WGCNA to group genes into modules that have strongly co-varying expression patterns, to identify biologically relevant features of the data. This approach allowed us to detect a TREM2-containing gene module, which was not only well preserved within all 10 brain regions studied but which could also be replicated in an independent human brain expression data set generated using an unrelated expression platform. In keeping with previous published analyses, this gene module, despite the use of heterogeneous samples (e.g., cerebellar cortex), related to a specific cell type, namely microglia, as demonstrated by the presence of multiple microglial markers within the TREM2-containing modules in all human brain samples. We were unable to clearly establish the predominant activation state of the microglia, with the detection of both markers of the M1 activation state (FCGR2A, FCGR2B, FCGR3B, and CXCL10) and M2a activation state in a subset of regions (IGF1, PDGFB, PDGFC, TGFB1, and MSR1). This complexity is likely to arise at least in part from the use of post-mortem brain tissue and the related variability in the mode of death of the individuals sampled. Although all samples originated from individuals undergoing sudden death, some modes of death may be associated with specific forms of microglial activation either due to common preceding events (e.g., angina preceding a myocardial infarction) or more directly. However, it should also be recognized that the understanding of microglial activation states is incomplete (Hanisch and Kettenmann, 2007; Kettenmann et al., 2013), making the identification and use of state-specific markers difficult.

Because studying human microglial function is challenging, we also used publicly available monocyte, dendritic cell, and macrophage microarray data sets to explore TREM2 expression and assess the validity of these cell types in modeling TREM2 function in human brain (Barreiro et al., 2012; Fairfax et al., 2012; Smith et al., 2009). In agreement with the existing literature, we detected TREM2 in dendritic cells and macrophages, but not in monocytes (Colonna, 2003). Although we were able to identify a TREM2-containing module in both dendritic cells and macrophages, there was no significant overlap in gene membership among the 2 cell types. There was evidence for weak but significant overlap in the TREM2-containing modules in macrophages and brain (as represented by the TREM2-containing module in the hippocampus). On deeper inspection, the major common element between these modules was the significant enrichment of lysosome-related genes. These findings suggest that using macrophages and dendritic cells to study TREM2 function in human brain may not be helpful. Furthermore, these findings highlight the unique properties of microglia, as compared to related myeloid cell types, in keeping with their distinct developmental origin (Chan et al., 2007).

As a result of these findings, we focused our efforts on studying the features of the TREM2-containing modules in human brain. We found that TREM2 was a hub gene in 5 brain regions, namely, the hippocampus, white matter, substantia nigra, and putamen, suggesting that TREM2 is capable of driving microglial functions in these regions. Although these findings do not fully explain the regional distribution of pathology in either Nasu-Hakola disease or AD, the “hub” role of TREM2 in white matter and hippocampus is certainly consistent with the severe leukoencephalopathy in the former and the selective loss of hippocampal neurons that characterize the latter. Using GO enrichment analysis to investigate module membership, we confirmed the well-recognized role of TREM2 in the innate immune response and related that to lysosome activity. However, this analysis also revealed the significant enrichment of genes related to lymphocyte activity (T and B cell), antigen presentation and cytokine release, which may be equally important in human brain.

To obtain further insights into TREM2-related signaling, we focused on a “core” group of 156 TREM2-related genes and used this gene list to look for enrichment of canonical gene sets defined by KEGG and relating to known pathways. Using this approach we identified 2 independent pathways of interest, the “Fc gamma R-mediated phagocytosis” and “Systemic lupus erythematosus” pathways. Because canonical pathways are limited by our existing knowledge base, we also visualized gene connectivity within the TREM2-containing module using VisANT in hippocampus and cerebellum. This approach not only identified genes highlighted within the KEGG pathways, but also identified additional genes of interest based on their high connectivity to TREM2 particularly in the hippocampus. The predominant common theme among these genes is their involvement in cytoskeletal regulation. For example, NCKAP1L (also known as HEM1), a member of the Scar/WAVE complex, plays an important role in regulating cell shape, and is essential in neutrophil chemotaxis (Weiner et al., 2006). Similarly DOCK2 is known to be involved in remodeling of the actin cytoskeleton required for lymphocyte migration (Sanui et al., 2003) and the biological functions of DOCK8, include regulation of cell migration (Harada et al., 2012; Jabara et al., 2012; Lambe et al., 2011). Interestingly, disruptions in this gene have also been linked to mental retardation and developmental disabilities (Griggs et al., 2008). ABB1IP encodes the Rap1–GTP-interacting adaptor molecule (RIAM) and is involved in mediating changes in the actin cytoskeleton after integrin-mediated adhesion, and is already a candidate gene for late-onset AD based on linkage analyses at chromosome 10 (Hernandez-Varas et al., 2011; Lafuente et al., 2004; Morgan et al., 2007).

This approach also highlighted a range of damage-related signals, namely C1Q complexes, ATP/ADP, and CSF-1, through the C1Q complex, plays an important role in regulating cell shape, and is essential in neutrophil chemotaxis (Weiner et al., 2006). ABB1IP encodes the Rap1–GTP-interacting adaptor molecule (RIAM) and is involved in mediating changes in the actin cytoskeleton after integrin-mediated adhesion, and is already a candidate gene for late-onset AD based on linkage analyses at chromosome 10 (Hernandez-Varas et al., 2011; Lafuente et al., 2004; Morgan et al., 2007).
phagocytic cells, including microglia, and this gene is of particular interest because the TREM2-TYROB protein complex has recently been implicated in CSFIR signaling (Otero et al., 2009). Furthermore, mutations in this gene are a cause of hereditary diffuse leukoencephalopathy with spheroids (HDSLs), an autosomal-dominant central nervous system white matter disease with some features in common with Nasu-Hakola disease (Rademakers et al., 2011). Thus, combining these 2 major themes (i.e., damage-related signals and cytoskeletal regulation) would suggest that TREM2 plays a key role in mediating changes in the microglial cytoskeleton required to initiate not only phagocytosis of apoptotic material but also other processes such as microglial migration to injury or plaque deposition sites (Nimmerjahn et al., 2005).

Finally, but most importantly, we demonstrate a significant enrichment of genes already implicated in AD, MS, and, to a lesser extent, MND (as documented by the HUGEnavigator database (Yu et al., 2008)) among the core genes of the TREM2-containing gene modules of human brain. This finding is important for 3 reasons. First, it places a number of known AD risk genes, namely MS4A6A and TREM2, in the same functional gene module, allowing us to begin to identify important pathophysiological pathways as opposed to single genes. Second, the enrichment of known MS, and to a lesser extent, MND-related genes (but not PD-related genes) suggests that the traditional classification of primary and secondary neurodegenerative diseases may be artificial and that the underlying pathophysiological processes converge (in at least a subset of neurological disorders) and center on microglial function. Third, this analysis implies that among the list of identified genes, there may as yet unidentified risk genes for AD and potentially other neurological disorders. Thus, this study provides important insights into TREM2-related signaling, which could be used as a platform for further experiments into the pathophysiological basis of late-onset Alzheimer’s disease.

**Disclosure statement**

The authors have no conflicting financial, personal, or professional interests.

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**Definitions**

**Adjacency matrix**

The adjacency matrix defines the co-expression network. It is constructed here from the pairwise Pearson correlations between all genes, raising these correlations to a power beta while maintaining the information on the correlation signs between genes, thus constructing a signed weighted co-expression network.

**Connectivity**

For each gene, the connectivity (also known as degree) is defined as the sum of its adjacencies with the other network genes. In co-expression networks, the connectivity measures how correlated a gene is with all other network genes.

**Topological overlap matrix (TOM)**

Topological overlap measure (TOM) is a pairwise measure of node similarity, that measures how close the neighbors of a gene are to the neighbors of the other gene. Unlike correlation or adjacency measures, which consider each pair of genes in isolation, TOM considers each pair of genes in relation to all other genes in the network, and it reflects the relative interconnectedness of 2 nodes within their local neighbourhood.

**Module eigengene (ME)**

The module eigengene is defined as the first principal component of the expression profiles of all genes in a given module.

**Module membership (MM)**

For each gene, the module membership (also known as eigengene-based connectivity) is calculated as the correlation of the gene expression profile with the module eigengene (ME) of a given module. If MM of a gene for a given module is close to 0, it means that the gene is not part of that module. On the other hand, if MM is close to 1 or 1, it is highly connected to the module genes. The sign of module membership encodes whether the gene has a positive or a negative relationship with the module eigengene. Highly connected intramodular hub genes tend to have high module membership values to the respective module.

**Appendix A. Supplementary data**

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.neurobiolaging.2013.05.001.

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