Phylogeography and resistome of pneumococcal meningitis in West Africa before and after vaccine introduction

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Abstract
Despite contributing to the large disease burden in West Africa, little is known about the genomic epidemiology of Streptococcus pneumoniae which cause meningitis among children under 5 years old in the region. We analysed whole-genome sequencing data from 185 S. pneumoniae isolates recovered from suspected paediatric meningitis cases as part of the World Health Organization (WHO) invasive bacterial diseases surveillance from 2010 to 2016. The phylogeny was reconstructed, accessory genome similarity was computed and antimicrobial-resistance patterns were inferred from the genome data and compared to phenotypic resistance from disc diffusion. We studied the changes in the distribution of serotypes pre- and post-pneumococcal conjugate vaccine (PCV) introduction in the Central and Western sub-regions separately. The overall distribution of non-vaccine, PCV7 (4, 6B, 9V, 14, 19F and 23F) and additional PCV13 serotypes (1, 3, 5, 6A, 19A and 7F) did not change significantly before and after PCV introduction in the Central region (Fisher’s test P value 0.27) despite an increase in the proportion of non-vaccine serotypes to 40% (n=6) in the post-PCV introduction period compared to 21.9% (n=14). In the Western sub-region, PCV13 serotypes were more dominant among isolates from The Gambia following the introduction of PCV7, 81% (n=17), compared to the pre-PCV period in neighbouring Senegal, 51% (n=27). The phylogeny illustrated the diversity of strains associated with paediatric meningitis in West Africa and highlighted the existence of phylogeographical clustering, with isolates from the same sub-region clustering and sharing similar accessory genome content. Antibiotic-resistance genotypes known to confer resistance to penicillin, chloramphenicol, co-trimoxazole and tetracycline were detected across all sub-regions. However, there was no discernible trend linking the presence of resistance genotypes with the vaccine introduction period or whether the strain was a vaccine or non-vaccine serotype. Resistance genotypes appeared to be conserved within selected sub-clades of the phylogenetic tree, suggesting clonal inheritance. Our data underscore the need for continued surveillance on the emergence of non-vaccine serotypes as well as chloramphenicol and penicillin resistance, as these antibiotics are likely still being used for empirical treatment in low-resource settings. This article contains data hosted by Microreact.

DATA SUMMARY
The sequencing reads for the genomes analysed have been deposited in the European Nucleotide Archive and the accession numbers for each isolate are listed in Table S1 (available with the online version of this article). A phylogenetic tree and associated metadata are available on Microreact: https://microreact.org/project/HJcM_9lxf.

INTRODUCTION
Streptococcus pneumoniae (the pneumococcus) is one of the top three causes of acute bacterial meningitis, the most severe form of meningitis [1]. Despite the availability of effective vaccines, acute bacterial meningitis due to the pneumococcus continues to be a major cause of morbidity and mortality in sub-Saharan Africa, especially among children under 5 years old [2]. The 10- or 13-valent pneumococcal conjugate vaccine (PCV10 and PCV13 respectively), which target the most prevalent serotypes in serious disease, have been introduced in 39/47 countries in

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sub-Saharan Africa. PCVs have been effective at reducing invasive disease caused by vaccine serotypes in high-income countries and low-resource settings [3–7].

Whole-genome sequencing has emerged as a useful technique for typing bacteria because it provides high-resolution inter-isolate comparisons, which can provide insights into pneumococcal transmission and evolution [8, 9]. For example, in 2016, 3 years post-PCV13 introduction, we identified a novel clade of sequence type (ST) 303 serotype 1 pneumococcus, which caused a meningitis outbreak in Ghana [10]. Access to genome sequencing and a well-defined global phylogeny of serotype 1 strains [11] allowed us to place the outbreak isolates in a global context. Unfortunately, in most West and Central African settings there remains a paucity of genomic data for pneumococcal serotypes associated with invasive disease.

An added advantage of genome sequencing is that it allows the prediction of resistance to commonly prescribed antibiotics [12]. For example, macrolide, chloramphenicol and tetracycline resistance can be predicted by the presence of the horizontally acquired \emph{ermB} and \emph{ormef}, \emph{cat} and \emph{tetM} genes, respectively [12–15]. Similarly, rifampicin, penicillin and co-trimoxazole resistance can be inferred based on known mutations in \emph{rpoB}, penicillin-binding protein genes \emph{pbp1A}, \emph{pbp2B} and \emph{pbp2X}, and \emph{folA/folP} genes, respectively [10, 12–16]. Through genomics, we can identify the emergence and spread of multidrug-resistant clones that could pose a serious public-health threat, especially in high disease burden settings.

We present genomic analysis of 185 pneumococcal isolates from children with confirmed meningitis in West Africa between 2010 and 2016. The collection of these isolates was facilitated through the World Health Organization (WHO) Invasive Bacterial Vaccine-Preventable Disease (IB-VPD) sentinel surveillance network. Our analysis probes the effect of PCV introduction on the genomic epidemiology of pneumococci causing paediatric meningitis. We also studied the potential effects of PCV introduction on the distribution of antibiotic-resistance genes. These data provide the first baseline data on the genomic epidemiology of pneumococcal paediatric meningitis in West Africa pre- and post-PCV introduction.

**Impact Statement**

\textit{Streptococcus pneumoniae} causes severe diseases including meningitis and pneumonia which are leading causes of morbidity and mortality in sub-Saharan Africa, particularly among children. Resource constraints make it difficult to carry out paediatric infectious disease surveillance in this setting. We compiled a genomic dataset of \textit{S. pneumoniae} strains associated with paediatric bacterial meningitis in West and Central Africa, collected through sentinel surveillance. This unique dataset has allowed us to study the genetic determinants of antimicrobial resistance in the pre- and post-PCV introduction periods. We also show novel insights into the phylogenetic landscape of \textit{S. pneumoniae} associated with paediatric meningitis and highlight patterns consistent with the localization of some sublineages by geographical sub-region.

**METHODS**

**Study design**

The WHO IB-VPD surveillance is supported by the WHO Collaborating Centre (WHO CC) for New Vaccines Surveillance hosted by the Medical Research Council Unit The Gambia at the London School of Hygiene and Tropical Medicine (MRCG). Within this framework, vaccine preventable meningitis surveillance has been on-going in 11 countries across 17 sentinel sites since 2010. As part of the surveillance, bacterial isolates recovered from suspected paediatric meningitis cases were sent to the...
WHO CC hosted at the MRCG for confirmation, serotyping, antibiotic-susceptibility testing and whole-genome sequencing in collaboration with the Global Pneumococcal Sequencing project (https://www.pneumogen.net/gps/).

Study population
The study participants were aged between 6 days and 59 months old, the mean age was 20.8 months and the median age was 14 months (age was recorded for 129 patients). This dataset also included two older children aged 8 and 16 years old. Patients admitted with suspected meningitis at paediatric teaching and referral hospitals were enrolled into the surveillance. Isolates for whole-genome sequencing were received from sentinel sites in The Gambia, Senegal, Ivory Coast, Ghana, Togo, Benin, Niger, Cameroon and the Democratic Republic of Congo (Table 1).

Bacteriology
Lumbar puncture was performed to collect cerebrospinal fluid (CSF) specimens. For some patients, specimens were also collected from other clinical sites, including blood, lung/pleural aspirate and pus. Our dataset includes isolates from CSF (151, 81.6%), blood (21, 11.4%), lung/pleural aspirate (9, 4.9%), pus (1, 0.5%) and 3 (1.6%) from an unknown source. The pneumococcal isolates were cultured using standard techniques and sent to the WHO Regional Reference Laboratory (RRL) for further analysis.

Clinical specimens were streaked onto Columbia agar with 5% defibrinated sheep blood and incubated overnight in 5% CO₂ at 37°C. Suspected colonies of the pneumococcus were confirmed by sensitivity to optochin (Oxoid) and stored in 16% (v/v) glycerol broth at −70°C for shipment to the WHO RRL. Confirmation of species and characterization of serotypes were done by PCR and latex agglutination, as described elsewhere [16]. Antimicrobial susceptibility to cefotaxime, chloramphenicol, meropenem, vancomycin, co-trimoxazole, rifampicin, tetracycline, oxacillin (for penicillin) and erythromycin was performed by disc diffusion, and interpreted according to the Clinical and Laboratory Standards Institute (CLSI) guidelines. Confirmatory Etest was done for cefotaxime and ceftriaxone by colleagues at the US Centers for Disease Control and Prevention (CDC). All the disc diffusion assays were performed at the WHO RRL. Minimum inhibitory concentrations (MICs) could not be assessed due to the inhibitory cost of the kits. The WHO RRL participates in external quality assurance (EQA) programmes organized by the United Kingdom National External Quality Assessment Service (UK NEQAS), the National Institute for Communicable Diseases (NICD), South Africa, and the United States Centers for Disease Control and Prevention (US CDC).

Whole-genome sequencing
Genomic DNA was extracted and purified from fresh overnight cultures of the stored pneumococcal isolates using a modified Qiagen extraction protocol as previously described [10]. Paired-end sequencing on the Illumina HiSeq platform was performed at the Wellcome Sanger Institute, Cambridge, UK [17].

Genomic analysis
The in silico serotype and the multilocus sequence type (MLST) were determined from the genomes using SeroBA [18] and a local alignment of de novo contigs against MLST alleles in pubMLST [19], respectively. Sequencing reads were mapped to the pneumococcal strain ATCC 700669 reference genome (accession no. FM211187) using bwa (version 0.7.17) with default parameters [20], the BAM files were sorted and duplicates were marked using Picard. The mpileup command in SAMtools (version 1.2.1) was used to call bases at all sites with at least five reads mapped, and output the calls in the variant call format (VCF) [21]. A consensus sequence was generated and SNPs were called where at least 75% of reads

Table 1. Counts of study isolates from each country by vaccine era

| Sub-region  | Country         | PCV introduction* | Pre-PCV [n (%)] | Post-PCV [n (%)] | Unknown [n (%)] | Total |
|-------------|-----------------|-------------------|-----------------|------------------|-----------------|-------|
| Eastern     | Cameroon        | 2011              | 2 (6.9)         | 27 (93.1)        | 0 (0)           | 29    |
| DR Congo    | 2011            | 0 (0)             | 0 (0)           | 1 (100)          | 1               |
| Central     | Ghana           | 2012              | 17 (65.4)       | 9 (34.6)         | 0 (0)           | 26    |
| Benin       | 2011            | 3 (60)            | 2 (40)          | 0 (0)            | 5               |
| Ivory Coast | 2014            | 2 (100)           | 0 (0)           | 0 (0)            | 2               |
| Niger       | 2014            | 23 (92)           |                 | 2 (8)            | 25              |
| Togo        | 2014            | 19 (82.6)         | 4 (17.4)        | 0 (0)            | 23              |
| Western     | Senegal         | 2013              | 53 (96.4)       | 2 (3.6)          | 0 (0)           | 55    |
| The Gambia  | 2009            | 0 (0)             | 19 (100)        | 0 (0)            | 19              |
| Total       |                 |                   | 119 (64.3)      | 63 (34.1)        | 3 (1.6)         | 185   |

*Post-PCV period commences the year after the introduction of a PCV vaccine of any valency.
mapped to the alternative allele. The consensus sequences from all genomes were amalgamated into a multiple sequence alignment FASTA file and variable sites were extracted using the SNP-sites program (version 2.5.1) [22]. The maximum-likelihood phylogeny was reconstructed from 99474 variable sites in the core genome using RAXML (version 8.2.8) with a general time reversible model and 100 bootstrap replicates [23]. The phylogenetic tree and associated metadata are publicly available on Microreact: https://microreact.org/project/HJcM_9bf.

De novo assemblies were generated from sequencing using The Sanger Institute assembly pipeline that is described elsewhere [24]. Assemblies were annotated using Prokka (version 1.14.5) [25] and the pan genome was inferred using Roary (version 3.12) [26]. Genomes were clustered based on the presence and absence of accessory genes using PANINI [27] and visualized in a scatter plot using Microreact. PANINI uses a Student’s t-distributed stochastic neighbour embedding machine learning algorithm to compute similarities in the accessory genome. Antimicrobial resistance was predicted based on the presence of known genotypes at loci that are associated with antibiotic resistance (rpoB, pbp1A, pbp2B, pbp2B and folA/folP genes) using an antibiotic-resistance calling pipeline designed for S. pneumoniae [12, 28–31]. The ermA/mefA, cat and tetM genes were linked to erythromycin, chloramphenicol and tetracycline resistance, respectively. Genotypic variants of pbp and folA/folP genes known to confer resistance were attributed to penicillin and co-trimoxazole resistance, respectively.

Statistical analysis
Data were compiled in Excel and uploaded onto RStudio (version 1.2.5033) for statistical analysis. The analyses were carried out separately on genomes from the Western and Central sub-regions. Paired t-test, chi-square (χ²) test and Fisher’s exact test were used to analyse the differences in the distribution of vaccine and non-vaccine serotypes in the pre- and post-PCV introduction periods. There were only two isolates from the pre-PCV period in the Eastern sub-region, so no comparisons could be made.

RESULTS
We analysed 185 genomes of pneumococci isolated from paediatric meningitis patients across West Africa and parts of Central Africa. The isolates were clustered based on country into three sub-regions: isolates from Gambia and Senegal were clustered as the Western sub-region; Ghana, Togo, Ivory Coast, Benin and Niger were clustered as the Central West sub-region; and Cameroon and the Democratic Republic of Congo represented the Eastern sub-region. We defined the pre-PCV period as the time period preceding the first introduction of the PCV (any valency) in a given country and post-PCV as the time post-introduction of PCV. A third of the isolates were recovered from cases that occurred post-PCV introduction and two thirds were pre-PCV (Table 1).

Serotype distribution across sub-regions
We classified our strains into serotypes and MLSTs based on the whole genome, and visualized their distribution in the three sub-regions as a means of studying sub-regional diversity. There were 32 serotypes identified among the cases, which included 76 unique Sequence Types (STs). Most isolates belonged to serotypes 1 (47, 25.4%), 14 (18, 9.7%), 5 (16, 8.6%) and 23F (14, 7.6%). Other commonly identified serotypes included 19F (10, 5.4%), 6A (10, 5.4%), 12F (9, 4.9%), 6B (9, 4.9%) and 15B/15C (6, 3.2%). Serotype 1 was dominant in the West and Central sub-regions, but it was not detected in the Eastern sub-region (Fig. 1a, b). Most serotypes were uncommon (20, 62.5%) and were only detected in one or two patients (Fig. 1a).

In the Central sub-region, isolates were sequenced from 64 and 15 patients in the pre- and post-PCV introduction periods, respectively. The overall distribution of non-vaccine serotypes, PCV7 serotypes (4, 6B, 9V, 14, 18C, 19F and 23F) and additional PCV13 serotypes (1, 3, 5, 6A, 19A and 7F) did not change significantly before and after PCV introduction in the Central region: the proportion of non-vaccine serotypes in the post-PCV period was 40% (n=6) compared to the pre-PCV period, 21.9% (n=4) (Fisher test P value 0.27) (Fig. 1c). The proportion of isolates with the non-vaccine serotypes 24, 12F, 35B and 23B increased by 13.3, 8.7, 6.3 and 5.1%, respectively, in the post-PCV period (Fig. 1d, Table S2). In the Western sub-region, all pre-PCV isolates were from Senegal (n=53), while 90% (n=19) of the post-PCV introduction isolates were from The Gambia after the introduction of PCV7, which precluded a comparison based on the PCV era.

Phylogenetic analysis and sub-regional clustering
The phylogenetic tree and a scatter plot depicting similarity of accessory genomes content were annotated with sub-region and serotype in order to visualize potential clustering of strains from the same sub-region. Phylogeographical clustering was observed within serotypes, with isolates from the same sub-region clustering and sharing similar accessory genome content (Fig. 2a). In West Africa, sub-regional clustering was observed within the serotype 1 clade: ST3081 was found only in the Western sub-region, while ST303 was the predominant genotype in the Central sub-region. Serotype 6A isolates formed two divergent clades with markedly different accessory genome content. The ST3324 subclade was prevalent in the Western sub-region and a ST5547 subclade was prevalent in the Central sub-region.

Serotype 5 was not found in the Central sub-region, but in the Western sub-region it was the second most common serotype. Three unique STs were present in the Western sub-region, but in the Eastern sub-region only ST289 was reported. Serotypes 5, 12F, 14 and 19F had a highly conserved serotype specific accessory genome and did not vary with ST or sub-region of origin (Fig. 2b).
Resistance genotypes in the context of sub-region, PCV introduction period and phylogeny

Our data confirm the presence of antibiotic-resistance genotypes known to confer resistance to penicillin ($n=38$, 20.5%), chloramphenicol ($n=18$, 9.7%), co-trimoxazole ($n=101$, 54.6%), erythromycin ($n=3$, 1.6%) and tetracycline ($n=111$, 60%). There did not appear to be a significant correlation between the presence of antibiotic resistance and sub-region (Fig. 3). Likewise, there was no significant trend linking the presence of resistance genotypes to the vaccine introduction period or whether the strain was a vaccine or non-vaccine serotype (Fig. 3). The genotypic predictors of resistance among the isolates were examined in the context of the phylogeny. Resistance genotypes appeared to be conserved within selected sub-clades of the phylogenetic tree (Fig. 4). Within serotype 1, isolates harbouring both tetM and folA/ folP resistance genotypes were common in the ST303/ST217 sub-clade ($n=8$, 42%) but absent in the ST3081 ($n=0$), and only four isolates in the ST618 sub-clade (20%) bore the tetM resistance gene (Fig. 4). Prediction of antimicrobial susceptibility from the genome has been found to have good correlation with phenotypic data [12]. However, we noted 23 (12.4%) chloramphenicol-resistant cases but the cat gene was only detected in 13 (56.3%) cases. Similarly, among 33 penicillin-resistant cases, 24 (72.7%) had a pbp genotype known to confer resistance. Unfortunately, we were unable to perform phenotypic antimicrobial susceptibility retesting to confirm the results.

DISCUSSION

Our dataset offers important insights into the genomic epidemiology of $S. pneumoniae$ associated with meningitis in West and Central Africa, a high-burden region with a paucity of genomic and epidemiological data. Phylogeographical clusters of isolates causing meningitis in the same geographical sub-region shared similar accessory genome content. The pneumococcus has an open pan genome that can readily acquire accessory genes from microbes within its ecological niche, making it highly adaptable to its environment [32, 33]. The accessory genome content plays a major role in determining the fitness of pneumococcal lineages and re-shaping the perturbed bacterial population structure following major clinical interventions like vaccination [34, 35]. Further work needs to be done to determine whether children in some parts of West and Central Africa are at a greater risk of developing meningitis.
Fig. 2. Phylogeny of *S. pneumoniae* genotypes causing paediatric meningitis in West and Central Africa, and a scatter plot showing accessory genome similarity. (a) A phylogenetic tree annotated with branches coloured by serotype, with metadata rings to show sub-region of origin, vaccine era and vaccine type with ST displayed as text on the outer ring. (b) A PANINI accessory genome scatter plot where each point, representing one isolate, is coloured by serotype, and distances between points are proportional to accessory genome similarity. The PANINI plot is by major serotypes (in bold) and the STs that demonstrate geographical clustering.
meningitis due to the presence of virulent pneumococcal lineages.

Although our dataset was not powered to study the impact of PCV on the epidemiology of pneumococcal bacterial meningitis among children, we gleaned some observations on the distribution of serotypes before and after the introduction of PCV. Serotype 1 remained a leading cause of paediatric pneumococcal meningitis. In the Central sub-region, the prevalence of serotype 1 decreased by 17.9% following the introduction of PCV13. Serotype 1 has been causing invasive disease for a long time and has evolved into distinct lineages that have adapted to specific geographical localities [11, 36]. In sub-Saharan Africa, serotype 1 encompasses highly virulent clones, some of which are capable of causing lethal outbreaks of meningitis [10, 37–39].

The high burden of serotype 1 in this sub-region may contribute to the lag in decreasing the burden of serotype 1 disease in the post-PCV13 period. In Ghana, where a three-dose regimen of PCV13 was introduced without a booster, serotype 1 remained a common cause of invasive pneumococcal disease up to 3 years after the introduction of PCV13, and in that period caused a meningitis outbreak among older children and adults in Ghana [7, 10, 40]. Experts have previously suggested that a booster may be effective in expediting the decline of vaccine serotypes

Fig. 3. Trends in the presence and absence of antibiotic-resistance genotypes in the context of sub-region and PCV introduction period. (a) A column plot showing the proportion of genomes bearing antibiotic-resistance genes among serotypes, which were grouped according to whether they were PCV7 serotypes, additional PCV13 serotypes or non-vaccine serotypes. (b) A column plot showing the proportion of isolates bearing antibiotic-resistance among isolates from the pre- and post-PCV introduction periods in each sub-region. Note that only two isolates from the Eastern sub-region in the pre-PCV introduction period were available.
in sub-Saharan Africa [41]. In South Africa, a three-dose regimen with a booster dose was effective in rapidly decreasing the burden of PCV13 serotypes, including serotype 1 [42]. A nuanced analysis of the role of boosters is needed based on robust epidemiological data or a clinical trial.

The replacement of vaccine serotypes causing invasive diseases by non-vaccine serotypes has been reported in several countries following the introduction of PCVs [7, 43–46]. The Global Pneumococcal Sequencing consortium has identified serotype 12F among the top five leading serotypes of the post-PCV13 period in at least two different countries [47]. In the Central sub-region, the prevalence of non-PCV serotypes, including serotype 12F, increased following the introduction of PCV13 serotypes. This underscores the need for continued surveillance in West Africa, to carefully monitor the role of non-vaccine serotypes in the post-PCV13 introduction period.

Antibiotic resistance can contribute to treatment failures and negatively impact clinical outcomes [48]. There was no clinically defined resistance to cefotaxime and ceftriaxone, which are the primary WHO-recommended antibiotics [49]. Isolates bearing the penicillin-resistance pbp genotypes were recovered from all sub-regions; this is an important finding because 49 when neither cefotaxime nor ceftriaxone are available, penicillin may be used to treat meningitis.

Championing improved infection management, diagnosis and antibiotic stewardship in sub-Saharan Africa may curb the emergence and spread of paediatric pneumococcal infections that are resistant to the recommended drugs [50–52]. This is challenging to implement in a low-resource setting with infrastructural constraints; and warrants special attention [53]. Vaccines have been proposed as a potential mechanism for reducing the burden of antibiotic resistance by lowering the risk of infection [54]. This was not reflected in this dataset, as a decrease in the prevalence of antibiotic-resistance genes was not associated with the post-PCV introduction era, and there were no discernible trends regarding the presence of resistance genotypes in vaccine and non-vaccine serotypes. While this may be reassuring, continued genomic surveillance is required to

Fig. 4. Antibiotic resistance and resistance genotype patterns in the context of the whole genome phylogeny. A phylogenetic tree with branches coloured by serotype and metadata blocks corresponding to sub region, vaccine introduction period, phenotypic antibiotic resistance patterns and presence of antibiotic resistance genes for penicillin (PBP, OX), chloramphenicol (cat, C), erythromycin (mef/ermB, E), co-trimoxazole (folP/folA, SXT), tetracycline (tetM, TE) and cefotaxime (CTX).
monitor these trends and to understand the underlying evolutionary mechanisms. We may be missing the contribution of novel resistance mechanisms that have not been discovered yet and resistance mechanisms that are difficult to predict with whole-genome sequencing, such as efflux pump upregulation and cell wall permeability changes [55, 56]. Additionally, our dataset may be affected by the use of antibiotic prior to hospital admission.

**Conclusion**

Our study underscores the need for continued surveillance for monitoring the emergence of non-vaccine serotypes as well as residual serotype 1. Genomic surveillance of pneumococcal disease and carriage in the African meningitis belt may provide an evidence-base to inform future strategies to enhance the control of the pneumococcus. Likewise, monitoring of antimicrobial resistance using genomic approaches may enhance the detection of drug resistant strains and contribute to ensuring that patients receive optimal treatment in low-resource settings.

**Study limitations**

A limitation of this study is that PCVs were introduced in the countries which participated in the surveillance at different time points and the countries did not use the same PCV formulations i.e. PCV10 or PCV13. This made it difficult to get a representative subset of pre- and post-PCV isolates across all three sub-regions. The number of isolates recovered from some countries was very low, e.g. there were single isolates from Ivory Coast and Democratic Republic of Congo. Grouping the isolates into sub-regions made the analysis possible, but this introduced potential bias as serotype distributions likely differ between and within countries. Furthermore, a limitation of sentinel surveillance is that we likely missed a significant proportion of pneumococcal meningitis cases that did not attend the sentinel sites. This is further compounded by the low culture-recovery rates for the pneumococcus across the sub-region. Coordinating surveillance in a low-resource setting is extremely challenging. This dataset highlights a need to develop laboratory capacity in this sub-Saharan Africa. The inability to perform retrospective confirmatory MIC testing may have potentially contributed to the discrepancies between MICs and genomic predictions.

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**Author contributions**

M. A., B. A. K.-A., J. M., M. B., M. S., M. H. K., A. A. G., F. H., B. H., A. G. B., T. S. D., E., T. N. O., I. F., F. O., T. D., L. A. R., S. K. O., B. A. D. A., J. A., B. S., B. F., S. C., B. A. N., A. B. and R. N. conducted the surveillance. K. P. K., R. F. B., L. M. and S. D. B. led the Global Pneumococcal Sequencing Project. P.-E. T., C. O., S. S., E. F.-N., S. J. and C. E. carried out the bacteriology work and DNA purification. M. S., A. K. W., B. K. and M. A. conceptualized the genomic and statistical analysis. R. A. G. and S. W. L. performed the genome assembly, annotation, serotyping, MLST analysis, genotyping and resistance prediction. M. S. and B. A. K.-A. drafted the first draft of the manuscript. All authors contributed to the drafting and review of the manuscript.

**Conflicts of interest**

The authors declare that there are no conflicts of interest.

**Ethical statement**

The standard operating procedures and protocols for the genomic sequencing were approved by The joint Gambia Government/MRC The Gambia Ethics Committee. Approval was granted by Ministries of Health of the participating countries to characterise isolates collected as part of ongoing routine surveillance for vaccinepreventable diseases.

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