Forensic Application of Epidermal Ubiquitin Expression to Determination of Wound Vitality in Human Compressed Neck Skin

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UBIQUITIN is a member of the heat shock protein family and is rapidly induced by various types of stimuli, including ischemic and mechanical stress. However, its significance in determining wound vitality of neck compression skin in forensic pathology remains unclear. We immunohistochemically examined the expression of ubiquitin in the neck skin samples to understand its forensic applicability in determining wound vitality. Skin samples were obtained from 53 cases of neck compression (hanging, 42 cases; strangulation, 11 cases) during forensic autopsies. Intact skin from the same individual was used as the control. Ubiquitin expression was detected in 73.9% of keratinocytes in intact skin samples, but only in 21.2% of keratinocytes in the compression regions, with statistical differences between the control and compression groups. This depletion in the case of neck compression may be caused by the impaired conversion of conjugated to free ubiquitin and failure of de novo ubiquitin synthesis. From a forensic pathological perspective, immunohistochemical examination of ubiquitin expression in the skin of the neck can be regarded as a valuable marker for diagnosing traces of antemortem compression.

Keywords: ubiquitin, compression, neck skin, immunohistochemistry, forensic pathology

INTRODUCTION

In forensic practices, differentiating antemortem injury from postmortem damage is one of the important issues, which is a classical but still modern topic (1–3). Thus, there are numerous forensic studies exploring available markers for the determination of wound vitality using biochemical, histochemical, immunohistochemical and molecular biological techniques (4–6). Moreover, there are several recent studies focusing on omics sciences and micro RNA (7, 8).

Fatal asphyxia due to neck compression is often encountered during forensic autopsies. Neck ligature marks can result from various lesions, including manual strangulation, ligature strangulation, strangulation, direct striking, armlocks, and cord entanglement (9, 10). Ligature mark is the most important finding on the neck for forensic pathologists. The mark is in the
form of a furrow or groove in the tissue, pale in color and may later change from tan to dark brown. The dryness and desiccation of the abraded skin makes the mark hard and parchment-like (11). In contrast, if a soft material, such as a towel, is used, or if a beard or a part of the cloth is between the ligature and the skin, the ligature mark may be faint or inconspicuous. Microscopy of the thyroid and salivary glands usually reveals focal interstitial hemorrhages, and the lymph gland shows congestion, supporting the antemortem nature of neck compression (12). However, only few studies have focused on the relationship between the ligature compression and neck tissue, and very few markers have been investigated as an evidence for neck compression (13–15).

Ubiquitin is an 8.5 kDa protein containing 76 amino acids and is commonly found in eukaryotic cells (16). Ubiquitin has seven lysine and methionine sites at its N-terminus, which tend to get self-ubiquitinated and extend to form different types of polyubiquitin chains (17). Ubiquitination is closely associated with many cellular processes such as cell cycle regulation, immune response, inflammatory response, and apoptosis. Major chronic degenerative diseases in humans, including Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis, are primarily associated with the nervous system, and ubiquitin is incorporated into many inclusion bodies that characterize these neurodegenerative diseases (18). Additionally, ubiquitin is involved in the response to acute cell injury. Transcription of ubiquitin mRNA in mammalian cells is induced by heat shock and other stresses (19, 20). In contrast, short-term ischemia causes depletion of free ubiquitin in gerbil hippocampal neurons, which are the most vulnerable to ischemic injury (21). Moreover, ubiquitin gene expression after ischemia/reperfusion has been studied in the rat brains, where it was observed to initially decrease after reperfusion; however, the expression increased after the blood flow was restored (22), suggesting that ubiquitin expression may serve as an indicator of the ischemic stress.

In the field of forensic pathology, there are several studies have investigated whether ubiquitin expression can be used as an indicator for forensic diagnosis. An immunohistochemical study of ubiquitin in the human locus coeruleus has shown that the number of neurons with ubiquitin expression is significantly higher in the case of long-term death struggles (23). In human kidney tissues, ubiquitin-immunopositive tubular epithelial cells are higher than the other groups involving subjects who died due to fire, blunt injury, sharp injury, and fatal hypothermia, suggesting that ubiquitin positivity is a characteristic of death due to injury and hypothermia (24, 25). In addition, the ubiquitin immune responsiveness of pigmented substantia nigra neurons in the midbrain has been suggested to be triggered by severe, deadly stress from asphyxia, drowning, and fire (26, 27). We also have previously reported that ubiquitin would be one of the markers for wound age estimation (28). However, there have been no immunohistochemical or forensic diagnostic studies pertaining to ubiquitin using compressed neck skin. In this study, we investigated the immunohistochemical expression of ubiquitin in neck skin specimens from autopsy cases and discussed whether it can be a useful marker for the forensic diagnosis of compression.

**MATERIALS AND METHODS**

**Antibodies**

The following polyclonal antibodies (pAbs) were used for immunohistochemical analysis in the present study: rabbit anti-ubiquitin pAbs (UBA52 Ab, AF0289, Affinity Biosciences, Jiangsu, China).

**Human Ligature Marks**

A total of 53 ligature marks (hanging, 42 cases; strangulation, 11 cases) with a postmortem interval of <96 h were obtained from forensic autopsies at our institute. In each case, the cause of death was carefully determined based on autopsy, histopathological findings and toxicological data. Intact skin from the same individual was used as the control. The detailed profiles of all cases (sex, age, and postmortem intervals) are shown in Table 1.

**Immunohistochemical Analysis**

Skin specimens were fixed in 4% formaldehyde solution buffered with PBS for 4–7 days, embedded in paraffin, and sectioned at a thickness of 4 μm. Briefly, deparaffinized sections were incubated with PBS containing 1% normal goat serum and 1% bovine serum albumin (BSA) to reduce non-specific reactions. Thereafter, sections were further incubated with anti-ubiquitin pAbs (dilution 1:100) for 12–17 h at 4°C. After incubation with biotinylated secondary antibodies, immune complexes were visualized using Catalyzed Signal Amplification System (Dako, Kyoto, Japan) according to the manufacturer’s instructions. As a negative control, sections were incubated with normal rabbit serum instead of the primary antibodies; and no positive signal was detected in this case, thus indicating the specificity of the antibodies.

**Morphometrical Analysis**

To evaluate ubiquitin expression in the skin, the ratio of ubiquitin-positive keratinocytes to the total number of corresponding keratinocytes was calculated in five randomly selected high-power fields (×400). The average values were evaluated as an indicator for ubiquitin expression. Morphometric evaluation was blindly performed by two investigators without the prior knowledge of the samples.

**Statistical Analysis**

The mean and standard error of the means (SEM) were calculated. Statistical analysis was performed using analysis of variance or Mann-Whitney U-test. Statistical significance was set at $P < 0.05$. Correlation analysis was performed using the non-parametric Spearman's correlation coefficient. Statistical significance was set at $P < 0.05$.

| Number | Male/Female | Age (y) Range | Mean | Postmortem interval (h) Range | Mean |
|--------|-------------|---------------|------|------------------------------|------|
| 53     | 32/21       | 16–90         | 59.2 | 10–84                        | 35.8 |
Ethical Approval

This study was approved by the Research Ethics Committee of Wakayama Medical University (No. 3313). All the procedures were performed in accordance with the Declaration of Helsinki Principles. Moreover, this study was conducted using autopsy records from the past, and we could not obtain informed consent from the bereaved family for the use of these records. Therefore, in accordance with the "Ethical Guidelines for Medical Research Involving Human Subjects (enacted by the Ministry of Health, Labor, and Welfare in Japan), section 12–1 (2) (a) (c)." Since this was a de-identified retrospective study of archived autopsy-derived tissue, the review board of the Research Ethics Committee of Wakayama Medical University waived the need for written informed consent from the relatives of the individuals studied.

RESULTS

Immunohistochemical Analysis of Ubiquitin in Autopsy Samples

We examined the distribution of ubiquitin in the skin samples. Consistent with previous observations (28), ubiquitin-positive signals were observed predominantly in predominantly keratinocytes of uninjured skin samples (Figure 1A). However, in most ligature marks, ubiquitin was not detected in the keratinocytes (Figure 1B).

DISCUSSION

It is an essential work to determine wound vitality and wound age in forensic autopsy cases. To achieve the purpose, advanced biological techniques are applied to forensic pathology (4–8). In forensic medicine, the compression mark on the neck is one of the most important criteria for determining whether the

The Expression of Ubiquitin Was Lower in Neck Compression Cases

As shown in Figure 2, the ratio of ubiquitin expression in keratinocytes was significantly suppressed in compressed skin samples, compared to that in the control samples. There were no significant differences among sex, age, and postmortem intervals in terms of ubiquitin expressions (Figure 3).
neck has been affected by the contractile force. Asphyxiation in the forensic context is associated with mechanical asphyxia through various mechanisms such as strangulation, hanging, smothering, choking and aspiration. Direct causes of death include systemic hypoxia due to airway obstruction, as well as possible neurological effects of cerebral ischemia and neck pressure. In general, it is not difficult to observe such marks upon gross examination. However, in certain situations, such as when the force is weak or the vital reaction is uncertain, it may be necessary to confirm the presence of the compression mark using a staining method during the histological examination of the neck skin. Immunohistochemical analyses may provide reliable information for the estimation of vitality using compression marks (2, 3, 15).

There are lot of studies on the determination of wound vitality of ligature marks (15). Turillazzi et al. have investigated the immunohistochemical expression of various cytokines in skin specimens from autopsy cases of death by hanging (29). Previous studies have reported that high expression of IL-15 in the skin of the neck may be a reliable marker of ligature mark vitality. IL-15 is known to activate neutrophils, which are major players in inflammation of damaged tissues. It has been reported that neutrophils were first observed in human skin wounds aged about 20–30 min (30). On the other hand, Kondo et al. have demonstrated that neutrophils were observed primarily at wound sites aged approximately 4–12 h, and were a source of cytokines and chemokines such as IL-1α, IL-8, CCL2, and CCL3 (31, 32). In an experimental study of cytokine expression during skin wound healing in mice, infiltration of numerous neutrophils producing IL-1α, IL-1β, IL-6, and TNF-α was observed at the wound sites 3 and 6 h after injury (33). Therefore, it has been suggested that inflammatory cells and cytokines may act as markers for the determination of wound age and vitality (34–36). Actually, several lines of accumulating evidence implied that IL-1β, one of the representative inflammatory cytokines, would be a candidate molecule for the discrimination between antemortem-postmortem hangings in both animal experiments and human samples. Grellner could confirm that IL-1β immunostaining of epidermal cells was a useful tool to discriminate antemortem and post-mortem hanging (37, 38).

Additionally, Maiese et al. demonstrated that intracytoplasmic depletion of FLIP playing as an inhibitor of apoptosis was evident in the epidermal layers of antemortem neck compression (39). Pérez et al. (40) reported a higher frequency of cells positive to cathepsin D and P-selectin was found in subcutaneous injured skin. Tryptase and CD15 were also useful for the determination of wound vitality in ligature marks (29). Focardi et al. (41, 42) focused on Langerhans cells determined by both antigens of MHC-II and CD1A and revealed that Langerhans cells and their-derived iNOS was significantly higher in ligature marks with vitality. Alternatively, De Matteis et al. (43) examined neck muscle tissues not neck skin samples, and evaluated the use of Troponin I—as fast skeletal muscle (TNNI2) to perform differential diagnoses about vitality in suicide by hanging and simulated hanging.

Prangenberg et al. (4) showed heat shock protein was widely applied to forensic pathology such as fire-related death, hypothermia, cardiac death, drowning, excited delirium, trauma and SIDS. Ubiquitin is known as a heat-shock protein induced by several stress conditions (18). Therefore, ubiquitin may contribute to the degradation of denatured proteins produced under various stress conditions. Several lines of accumulating evidence demonstrated the availability of ubiquitin in the postmortem forensic diagnosis of pathophysiology such as asphyxia, drowning, fire, and hypothermia (24–27). Moreover, only our group examined the expression of ubiquitin in human mechanical skin wounds such as stab wounds, cut wounds, surgical wounds and lacerations (28). Subsequently, we have observed an increase of ubiquitin expression in antemortem skin wounds, and demonstrated that a significant ubiquitin positivity rate of above 30% in human skin wounds may indicate the wound age to be of 7–14 days (28). These observations prompt us to examine the expression of ubiquitin in compressed skin samples with a hypothesis that ubiquitin expression might be enhanced after antemortem compression. Unexpectedly, we have found the suppression of ubiquitin expression in compressed skin samples, compared with intact skin samples. Actually, in the present study, we found that 73.9% of the keratinocytes in the control specimens were ubiquitin-positive, whereas only 21.2% of the keratinocytes in the compressed specimens were ubiquitin-positive with a statistical difference. These observations were similar to those of Maiese’s study that intracytoplasmic depletion of FLIP was found in the epidermal layers of antemortem ligature marks (39).

Ubiquitin gene expression after ischemia/reperfusion has been studied in the rat brains, wherein it was observed to initially decrease after reperfusion, but increased before the blood flow was restored to normal levels (22). These results suggest that ubiquitin may act as a useful marker of ischemic stress. In addition, Morimoto et al. have demonstrated that short-term ischemia causes depletion of free ubiquitin in gerbil hippocampal neurons, which are the most vulnerable to ischemic injury (21). Thus, the discrepancy would result from the difference of wound type between previous (28) and present studies. Although our previous study examined open skin wounds such as cutting and stabbing (28), we employed compressed skin samples indicating closed skin wounds in the present study. Compression to the neck skin can cause more severe local ischemia, eventually resulting in the decrease of ubiquitin expression at the compressed skin area including ligature marks. In other words, a substantial reduction in ubiquitin expression in the skin of the neck may be characteristic of the compression.

From a forensic safety standpoint, it is not sufficient to make a diagnosis using a single marker. For example, immunohistochemical detection of aquaporin-3 (AQP3) in the skin of the neck can be considered a valuable marker for diagnosing traces of antemortem compression (14). Thus, it is emphasized that several different markers reported previously should be examined in forensic practices in order to prevent over-diagnosis or missing of wound vitality. In addition to examining the skin tissue of the neck, markers of neck compression of lung tissue were examined in another study. There were partial differences in the level of immunohistochemical staining for AQP5 among the causes of death such as choking, choking, and
sudden cardiac death (44). Moreover, increased thyroglobulin, total T3, and free T3 levels in postmortem blood samples may suggest neck compression (45–47). Recently, Neri et al. showed an increase in the expression of miRNAs recognized as regulators of the inflammatory response in skin lesions such as miR125a-5p and miR125b-5p, implying that regulation of miRNAs as new tool for cutaneous vitality lesions demonstration in ligature marks (48).

CONCLUSION

We have showed that the detection of ubiquitin in the neck skin is possible with the accuracy required for forensic purposes. This fact is especially true for soft-marks, which are particularly difficult to assess based on gross examination and conventional histological analysis using HE staining.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Research Ethics Committee of Wakayama Medical University (No. 3313). Written informed consent from the participants’ legal guardian/next of kin was not required to participate in this study in accordance with the national legislation and the institutional requirements.

REFERENCES

1. Cecchi R. Estimating wound age: looking into the future. Int J Legal Med. (2010) 124:523–36. doi: 10.1007/s00414-010-0505-x
2. Kondo T, Ishida Y. Molecular pathology of wound healing. Forensic Sci Int. (2010) 203:93–8. doi: 10.1016/j.forsciint.2010.07.004
3. Ishida Y, Nosaka M, Kondo T. Bone marrow-derived cells and wound age estimation. Front Med (Lausanne). (2022) 9:228572. doi: 10.3389/fmed.2022.228572
4. Prangenberg J, Doberentz E, Mawick A, Madae B. Mini review: the forensic value of heat shock proteins. Front Med (Lausanne). (2022) 8:900100. doi: 10.3389/fmed.2021.900100
5. Bertozzi G, Ferrara M, La Russa R, Pollice G, Gurgoglione G, Frisoni P, et al. Wound vitality in decomposed bodies: new frontiers through immunohistochemistry. Front Med (Lausanne). (2021) 8:802841. doi: 10.3389/fmed.2021.802841
6. Prangenberg J, Doberentz E, Madae B. Mini review: forensic value of aquaporines. Front Med (Lausanne). (2021) 8:793140. doi: 10.3389/fmed.2021.793140
7. De Simone S, Giacani E, Bosco MA, Vittorio S, Ferrara M, Bertozzi G, et al. The role of miRNAs as new molecular biomarkers for dating the age of wound production: a systematic review. Front Med (Lausanne). (2022) 8:803067. doi: 10.3389/fmed.2021.803067
8. Ros AC, Bacci S, Luna A, Legaz I. Forensic impact of the omics science involved in the wound: a systematic review. Front Med (Lausanne). (2022) 8:786798. doi: 10.3389/fmed.2021.786798
9. Knight B, Saukko PJ. Fatal pressure on the neck. 3rd ed. In: Knight’s Forensic Pathology. London: Arnold (2004). p. 368–94.
10. Di Maio DJ, Di Maio Vincent JM. Asphyxia. In: Forensic Pathology. New York, NY: Elsevier (1989). p. 257–77.
11. Bardale R. Violent asphyxia. In: Principles of Forensic Medicine and Toxicology. London: Jaypee brothers medical publishers (2011). p. 284–313.
12. Dixit PG, Mohite PM, Ambade VN. Study of histopathological changes in thyroid, salivary gland and lymph nodes in hanging. J Forensic Med Toxicol. (2001) 18:1–4.
13. Advenier AS, de la Grandmaison GL. Traumatic rupture of deep neck structures in hanging: two case reports. Am J Forensic Med Pathol. (2014) 35:189–92. doi: 10.1097/PAF.0000000000000114
14. Ishida Y, Kuninaka Y, Nosaka M, Shimada E, Hata S, Yamamoto H, et al. Forensic application of epidermal AQP3 expression to determination of wound vitality in human compressed neck skin. Int J Legal Med. (2018) 132:1375–80. doi: 10.1007/s00414-018-1780-1
15. Maiese A, Del Duca F, Santoro P, Pellegrini L, De Matteis A, La Russa R, et al. An overview on actual knowledge about immunohistochemical and molecular features of vitality, focusing on the growing evidence and analysis to distinguish between suicidal and simulated hanging. Front Med (Lausanne). (2022) 8:793539. doi: 10.3389/fmed.2021.793539
16. Shen W, Zhang Z, Ma J, Lu D, Lyu L. The ubiquitin proteasome system and skin fibrosis. Mol Diagn Ther. (2021) 25:29–40. doi: 10.1007/s40291-020-00509-2
17. Roque W, Summer R, Romero F. Fine-tuning the ubiquitin-proteasome system to treat pulmonary fibrosis. Connect Tissue Res. (2019) 60:50–61. doi: 10.1080/03008207.2018.1529174
18. John Mayer R, Landon M, Lowe J. Ubiquitin and the molecular pathology of human disease. In: Peter JM, Harris JR, Finley D editors. Ubiquitin and the Molecular Biology of the Cell. New York, NY: Plenum Press (1996). p. 429–62.
19. Bond U, Schlesinger MJ. Ubiquitin is a heat shock protein in chicken embryo fibroblasts. Mol Cell Biol. (1985) 5:949–56. doi: 10.1128/mcb.5.5.949-956.1985
20. Bond U, Schlesinger MJ. The chicken ubiquitin gene contains a heat shock promoter and expresses an unstable mRNA in heat-shocked cells. Mol Cell Biol. (1986) 6:4602–10. doi: 10.1128/mcb.6.12.4602-4610.1986
21. Morimoto T, Ide T, Ibara Y, Tamura A, Kirino T. Transient ischemia depletes free ubiquitin in the gerbil hippocampal CA1 neurons. Am J Pathol. (1996) 148:249–57.

AUTHOR CONTRIBUTIONS

SZ, YI, and TK formulated the hypothesis and designed the project. SZ, MN, YK, and AK performed the main experiments. AI, SH, and HiY provided technical assistance and discussion. YH, JM, and HaY helped with some experimental procedures. YI and TK oversaw the experiments and provided the main funding for the project. YI, FF, and TK participated in writing the manuscript. All authors contributed to the article and approved the submitted version.

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22. Noga M, Hayashi T. Ubiquitin gene expression following transient forebrain ischemia. Brain Res Mol Brain Res. (1996) 36:261–7. doi: 10.1016/0169-328x(95)00260-y

23. Piette MH, Pieters SE, De Letter EA. Evaluation of the agonal stress: can immunohistochemical detection of ubiquitin in the locus coeruleus be useful? Int J Legal Med. (2011) 125:333–40. doi: 10.1007/s00414-010-0434-8

24. Ishikawa T, Zhu BL, Li DR, Zhao D, Michiue T, Maeda H. Immunohistochemical investigation of ubiquitin and myoglobin in the kidney in medicolegal autopsy cases. Forensic Sci Int. (2007) 171:136–41. doi: 10.1016/j.forsciint.2006.10.016

25. Shimizu K, Ohtani S, Shiono H, Fukusima T, Sasaki M. Expression of ubiquitin protein in each organ at death from hypothermia. Forensic Sci Int. (1997) 86:61–8. doi: 10.1016/s0379-0738(97)02117-8

26. Quan L, Zhu BL, Ishida T, Oritani S, Taniguchi M, Fujita MQ, et al. Intraneuronal ubiquitin immunoreactivity of the pigmented neurons of the substantia nigra in fatal acute mechanical asphyxiation and drowning. Int J Legal Med. (2001) 115:6–11. doi: 10.1007/s0041400000200

27. Quan L, Zhu BL, Oritani S, Ishida K, Fujita MQ, Maeda H. Intraneuronal ubiquitin immunoreactivity in the pigmented neurons of the substantia nigra in fire fatalities. Int J Legal Med. (2001) 114:310–5. doi: 10.1007/s004140000192

28. Kondo T, Tanaka J, Ishida Y, Mori R, Takayasu T, Ohshima T. Ubiquitin expression in skin wounds and its application to forensic wound age determination. Int J Legal Med. (2002) 116:267–72. doi: 10.1007/s00414-002-0322-y

29. Turillazzi E, Vacchiano G, Luna-Maldonado A, Neri M, Pomara C, Rabozzi R, et al. Tryptase, CD15 and IL-15 as reliable markers for the determination of soft and hard ligature marks vitality. Histol Histopathol. (2010) 25:1539–46. doi: 10.14670/HH-25.1539

30. Betz P. Histological and enzyme histochemical parameters for the age estimation of human skin wounds. Int J Legal Med. (1994) 107:60–8. doi: 10.1007/BF01225491

31. Kondo T, Ohshima T, Mori R, Guan DW, Ohshima K, Eisenmenger W. Immunohistochemical detection of chemokines in human skin wounds and its application to wound age determination. Int J Legal Med. (2002) 116:87–91. doi: 10.1007/s0041401000260

32. Kondo T, Ohshima T, Eisenmenger W. Immunohistochemical and morphometrical study on the temporal expression of interleukin-1α (IL-1α) in human skin wounds for forensic wound age determination. Int J Legal Med. (1999) 112:249–52. doi: 10.1007/s004140050244

33. Kondo T, Ohshima T. The dynamics of inflammatory cytokines in the healing process of mouse skin wound: a preliminary study for possible wound age determination. Int J Legal Med. (1996) 108:231–6. doi: 10.1007/BF01369816

34. Kondo T. Timing of skin wounds. Leg Med (Tokyo). (2007) 9:109–14. doi: 10.1016/j.legalmed.2006.11.009

35. Peyron PA, Colomb S, Becas D, Adriansen A, Gauchotte G, Tiers L, et al. Cytokines as new biomarkers of skin wound vitality. Int J Legal Med. (2021) 135:2357–45. doi: 10.1007/s00414-021-02659-z

36. Casse JM, Martirelle L, Vignaud JM, Gauchotte G. Skin wounds vitality markers in forensic pathology: an updated review. Med Sci Law. (2016) 56:128–37. doi: 10.1177/0258245415590175

37. Balandiz H, Pehlivan S, Çiçek AF, Tuğcu H. Evaluation of vitality in the experimental hanging model of rats by using immunohistochemical IL-1β antibody staining. Am J Forensic Med Pathol. (2015) 36:317–22. doi: 10.1016/j.ajfmp.20000000000199

38. Grrellner W. Time-dependent immunohistochemical detection of proinflammatory cytokines (IL-1β, IL-6, TNF-α) in human skin wounds. Forensic Sci Int. (2002) 130:90–6. doi: 10.1016/s0379-0738(02)00342-0

39. Maiese A, De Matteis A, Bolino G, Turilliazzi E, Frati P, Fineschi V. Hypo-expression of flice-inhibitory protein and activation of the caspase-8 apoptotic pathways in the death-inducing signaling complex due to ischemia induced by the compression of the asphyxiogenic tool on the skin in hanging cases. Diagnostics (Basel). (2020) 10:938. doi: 10.3390/diagnostics10110938

40. Legaz Pérez I, Falcón M, Gimenez M, Diaz FM, Pérez-Cárceles MD, Osuna E, et al. Diagnosis of vitality in skin wounds in the ligature marks resulting from suicide hanging. Am J Forensic Med Pathol. (2017) 38:211–8. doi: 10.1097/PAF.000000000003322

41. Focardi M, Puliti E, Grifoni R, Palandrini M, Bugelli V, Pinchi V. Immunohistochemical localization of Langerhans cells as a tool for vitality in hanging mark wounds: a pilot study. Aust J Forensic Sci. (2020) 52:393–405. doi: 10.1080/00450618.2019.1567811

42. Focardi M, Bugelli V, Venturini M, Bianchi I, Defraia B, Pinchi V, et al. Increased expression of iNOS by Langerhans cells in hanging marks. Aust J Forensic Sci. (2022) 54:2; 177–86. doi: 10.1080/00450618.2020.1817556

43. De Matteis A, dell’Aquila M, Maiese A, Frati P, La Russa R, Bolino G, et al. The Troponin-I fast skeletal muscle is reliable marker for the determination of vitality in the suicide hanging. Forensic Sci Int. (2019) 301:284–8. doi: 10.1016/j.forsciint.2019.05.055

44. Wang Q, Ishikawa T, Michiue T, Zhu BL, Guan DW, Maeda H. Intrapulmonary aquaporin-5 expression as a possible biomarker for discriminating smothering and choking from sudden cardiac death: a pilot study. Forensic Sci Int. (2012) 220:154–7. doi: 10.1016/j.forsciint.2012.02.013

45. Palmieri C, Tettamanzi C, Scarpelli MP, Rousseau G, Egger C, Bongiovanni M. Postmortem biochemical investigation results in situations of fatal mechanical compression of the neck region. Leg Med (Tokyo). (2018) 30:59–63. doi: 10.1016/j.legalmed.2017.11.002

46. Takaso M, Tsuibo H, Komeda N, Morimoto M, Ikegaya H. Value of the serum thyroglobulin level for diagnosing neck compression in postmortem cases. J Forensic Leg Med. (2018) 58:126–9. doi: 10.1016/j.jflm.2018.04.016

47. Tamaki K, Katsumata Y. Enzyme-linked immunosorbent assay for plasma thyroglobulin following compression of the neck. Forensic Sci Int. (1990) 44:193–201. doi: 10.1016/0379-0738(90)90250-3

48. Neri M, Fabbri M, D’Errico S, Di Paolo M, Frati P, Gaudio RM, et al. Regulation of miRNAs as new tool for cutaneous vitality lesions demonstration in ligature marks in deaths by hanging. Sci Rep. (2019) 9:20011. doi: 10.1038/s41598-019-56682-7

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