Abstract. This study aimed to investigate the therapeutic effect of berberine on renal ischemia-reperfusion injury in rats and its effect on Bax and Bcl-2. Sixty adult SD rats were randomly divided into four groups: control group A, renal ischemia-reperfusion group B, berberine group C and berberine + exendin-(9-39) treatment group D. In group A, right kidney was resected and left renal pedicle was separated, but left renal artery was not blocked. Renal ischemia-reperfusion model was established in other groups. Rats in group C were not subjected to any treatment after model construction. Rats in group C and D were subjected to intraperitoneal injection of berberine 7 days before the experiment. Besides that, intraperitoneal injection of exendin-(9-39) was performed at day 1 and 4 after model construction. Automatic biochemical analyzer was used to measure serum creatinine (SCr) and blood urea nitrogen (BUN). Malondialdehyde (MDA) in renal cortex was measured by enzyme-linked immunosorbent assay and contents of Bax and Bcl-2 in renal tissue were measured by western blot analysis. Apoptosis of rat renal cells was detected by TUNEL assay. The results showed that levels of SCr, BUN, MDA and Bax were significantly higher in group B than in other groups (P<0.05). Levels of Bcl-2 in group B were significantly higher than those in group A but significantly lower than those in group C and D. Compared with group A, apoptosis of renal cells was more severe in group B. Compared with group B, apoptosis of renal cells was significantly improved in group C and D, but was still more severe than that in group A. In conclusion, berberine can effectively improve renal function in rats with renal ischemia-reperfusion injury by inhibiting Bax expression and promoting Bcl-2 expression.

Introduction

Ischemia-reperfusion injury (IRI) is caused by blood flow perfusion after organ ischemia, which can bring damages to tissues and organs (1). IRI is a common physiological process in clinical practices. Studies have shown that patients with renal transplantation or renal surgery are likely to be affected by IRI. After renal transplantation, IRI may lead to rejection to cause renal failure, which in turn shortens the survival of patients (2-4). Renal IRI is one of the causes of acute kidney injury. Although great efforts were made to treat this disease, treatment outcomes are still unsatisfactory, and the incidence and mortality are relatively high (5). Therefore, how to reduce the morbidity and mortality is a major task for researchers.

Berberine, also known as jamaicin, is a common type of alkaloids (isoquinolines). The active ingredients are extracted from the roots of the rhizome. Studies have shown that this drug has promising therapeutic effects in the treatment of digestive diseases and diarrhea, but its absorption through oral administration is not satisfactory, and the duration of plasma concentration is short (6,7). Exendin-(9-39), as an antagonist of GLP-1R, can effectively attenuate Nesfatin-1 and delay the emptying of the stomach and reduce the intake of berberine and effectively maintain the retention of berberine (8). In recent years, it has been found that berberine has anti-inflammatory functions and more and more studies confirmed that berberine played a certain regulatory role in inhibiting the body's immune response and anti-apoptosis response (9). In this study, rat model of renal IRI was established to explore the therapeutic effects of berberine in the treatment of renal IRI with the expectation of providing references for clinical treatment.

Materials and methods

Laboratory animals and materials. A total of 40 healthy male SD rats weighing 250-300 g were purchased from Wuhan MultiSciences (Lianke) Biotech Co., Ltd. (Hangzhou, China). Rabbit polyclonal Bcl-2 antibody (dilution 1/500; cat. no. ab59348) and rabbit monoclonal Bax antibody (dilution 1/500; cat. no. ab32503) were purchased Abcam (Cambridge, MA, USA). Malondialdehyde (MDA) kit was from Beyotime Biotechnology (Jiangsu, China). Exendin-(9-39) kit was purchased from Bachem (Bubendorf, Switzerland). Terminal deoxynucleotidyltransferase-mediated dUTP nick...
end labeling (TUNEL) cell apoptosis kit was purchased from Trevigen (New York, NY, USA).

**Animal grouping.** Forty adult SD rats were randomly divided into four groups: sham-operation group A, renal ischemia-reperfusion group B, berberine treatment group C and berberine + exendin-(9-39) treatment group D, 10 rats in each group. Rats were housed in a temperature controlled room (21 ± 2°C) on a 12:12-h light/dark cycle (lights on at 06:00 a.m.). All rats had free access to water and food. Rats in group A and B were intraperitoneally injected with 2 ml saline from 7 days before surgery. Rats in group C and D were subjected to intraperitoneal injection of berberine (0.4 mg/kg/day) 7 days before the experiment (berberine powder was dissolved in 2 ml of saline). Besides that, intraperitoneal injection of exendin-(9-39) was performed in group D at day 1 and 4 at a dose of 45 µg/kg/3 days after model construction. In group A, right kidney was resected and left renal pedicle was separated, but left renal artery was not blocked. Renal ischemia-reperfusion model was established in other groups.

Establishment of animal model. Rats were fasted for 12 h before animal model establishment, but were allowed to access water freely. Intra-peritoneal injection of 10% chlorhexinate hydrate (350 mg/kg) was performed for anesthesia. Rats were fixed and disinfected. An incision was made on the middle of the abdomen, and lateral peritoneum was cut to expose right kidney. Left ureter was protected, and left renal pedicle was separated. A non-destructive vascular clamp was used for left renal artery occlusion until kidneys turned from red to violet-black. Then vascular clamp was removed and the successful establishment of rat model of renal IRI is indicated by the changes of renal color from violet-black to red. The study was approved by the Ethics Committee of People's Hospital of Lishui Animal Center (Zhejiang, China).

**Sample collection and testing.** Blood samples were collected from the orbital veins of rats before left kidney resection. Before centrifugation at 2,500 x g for 5 min at 4°C, level of MDA in plasma was determined according to the instructions of MDA kit, and levels of serum creatinine (SCr) and blood urea nitrogen (BUN) were detected using an automatic biochemical analyzer after left kidney resection was performed, and 500 mg renal tissue was washed three times with NaCl, and stored at -4°C. Western blot analysis was used to detect the expression of Bax and Bcl-2 in renal tissues of rats. Tissue apoptosis was detected by TUNEL assay.

**Western blot analysis.** Total protein was extracted, quantified, and mixed with buffer. After electrophoresis, transmembrane was performed on ice under 100 V for 2 h. Membranes were blocked with 5% skim milk for 1 h, and incubated with primary rabbit polyclonal Bcl-2 antibody (dilution 1:500; cat. no. ab59348) and rabbit monoclonal Bax antibody (dilution 1:500; cat. no. ab32503) overnight. After that, membranes were further incubated with secondary goat anti-rabbit (HRP) IgG antibody (dilution 1:2,000; cat. no. ab6721) at room temperature for 1 h. After washing with TBST, BandScan 5.0 software (Glyko, Inc., Novato, CA, USA) was used to measure gray value of protein bands and calculate the gray ratio.

Table I. Comparison of renal cell apoptosis among 4 groups.

| Groups | n  | Percentage (%) |
|--------|----|----------------|
| A      | 9  | 3.40±1.20      |
| B      | 8  | 57.41±13.65    |
| C      | 9  | 17.2±4.2       |
| D      | 10 | 15.1±2.7       |

**TUNEL.** TUNEL assay kit was used to detect the apoptotic cells in kidney of each group, and the operation was carried out in strict accordance with the instructions of the kit. Cells were counted under optical microscopy. Five visual fields were randomly selected under optical microscope.  Apoptosis rate (AI) = total apoptotic cells/total cell number x100%. Determination of apoptotic cells: nuclei of apoptotic renal cells showed yellow or yellow-brown color, and nuclei of normal renal cells showed blue after staining.

**Statistical analysis.** In this study, SPSS22.0 software package was used to analyze all the data. Measurements were expressed as mean ± standard deviation (mean ± SD), and comparison were performed by ANOVA analysis followed by Least Significant Difference post hoc test. P<0.05 was considered to indicate a statistically significant difference.

**Results**

**Model construction.** In this study, rat model of renal IRI was established. During model construction, 1 rat died in group A, 2 in group B, 1 in group C and none in group D.

**Comparison the renal functions among groups.** Comparisons of levels of BUN, Scr and MDA among groups showed that levels of BUN, Scr and MDA in group B were significantly increased compared with those in other groups (P<0.05), while levels of BUN, Scr and MDA were significantly lower in group A than in other groups (P<0.05). No significant differences in levels of BUN, Scr and MDA were found between groups C and D (P>0.05) (Figs. 1-3).

**Comparison of levels of Bax and Bcl-2 among groups.** Group C and D pretreated with berberine showed significantly reduced expression level of Bax compared with that in group B, which was not treated with berberine (P<0.05). In contrast, expression level of Bcl-2 was significantly increased in group C and D compared with group B (P<0.05). No significant difference was found between C and D (P>0.05) (Fig. 4).

**Comparison of renal cell apoptosis.** Apoptosis of rat renal cells was detected by TUNEL method (the positive nucleus was brown and the negative was blue). Apoptosis was found in a small portion of cells in group A (3.40±1.20%). Compared with group A, cell apoptosis was significantly increased in group B (57.41±13.65%, P<0.05). Compared with group B, apoptosis of renal cells in group C and D was significantly improved (17.2±4.2 and 15.1±2.7%, P<0.05). Compared with group A, apoptosis of renal cells in group C and D was
significantly increased (17.2±4.2 and 15.1±2.7%, P<0.05). No significant difference was found between group C and D (P>0.05) (Table I).

**Discussion**

Kidney is a high perfusion organ that is highly sensitive to ischemia, hypoxia and reperfusion. IRI is caused by blood flow perfusion after long-term organ ischemia, which can delay recovery of organ functions or even bring damages to tissues and organs (10). Reperfusion may also cause tissue damage, cell apoptosis and necrosis, which has great damage to the tissues and organs of patients (11). Occurrence of renal IRI may cause unsatisfactory treatment outcomes of shock, renal transplantation and cardiopulmonary bypass, and is also a main cause of the failure of renal transplantation (12). At present, the pathogenesis of IRI is still unclear, and mainstream theories include calcium overload, leukocyte action and ATP deficiency (13).

Renal IRI is a complex pathological process with cell necrosis and apoptosis involved (14). Cell necrosis refers to non-programmed cell death mediated by the functions of cell lysates, protein denaturation, and activation of inflammatory factor receptor. Most of the cell necrosis is caused by reversible organ damage (15). Apoptosis is a process of programmed cell death that occurs through the regulation of genes, activation of extracellular molecules and external factors (16). Bcl-2, as one of the cytokines with anti-apoptotic function, can inhibit free radicals to induce intracellular calcium overload, causing blocked mitochondrial membrane permeability and caspase activation, thereby inhibiting apoptosis (17). Bax is a class of genes that can promote apoptosis through the regulation of cytokine, and studies have shown that the ratio of Bax/Bcl-2 can directly reflect cell apoptosis (18).

In this study, rat IRI model was established. Expression levels of Scr and BUN in rats of group A that were treated with saline were significantly lower than those in group B (P=0.027). Levels of BUN in group A were significantly lower than those in group C and D (P=0.042 and P=0.045). BUN expression levels in C and D groups were significantly lower than those in B group (P=0.038 and P=0.041). There was no significant difference between C and D groups (P>0.05). *P<0.05, compared with group B.

Expression levels of MDA in group A were significantly lower than those in group B (P=0.032 and P=0.035). Scr expression levels in group A were significantly lower than those in group C and D (P=0.024 and P=0.020). There was no significant difference between C and D groups (P>0.05). *P<0.05, compared with group B.

Expression levels of Scr in group A were significantly lower than those in group B (P=0.001). Scr expression levels in group A were significantly lower than those in group C and D (P=0.032 and P=0.035). Scr expression levels in C and D groups were significantly different from those in B group (P=0.024 and P=0.020). There was no significant difference between C and D groups (P>0.05). *P<0.05, compared with group B.

Expression levels of BUN in group A were significantly lower than those in group B (P=0.027). Levels of BUN in group A were significantly lower than those in group C and D (P=0.042 and P=0.045). Expression levels of MDA in group A were significantly lower than those in group C and D (P=0.036 and P=0.040). There were significant differences between group B and group C and D (P=0.044 and P=0.039). There was no significant difference between group C and D (P>0.05). *P<0.05, compared with group B.

Expression levels of Scr in group A were significantly lower than those in group B (P=0.001). Scr expression levels in group A were significantly lower than those in group C and D (P=0.032 and P=0.035). Scr expression levels in C and D groups were significantly different from those in B group (P=0.024 and P=0.020). There was no significant difference between C and D groups (P>0.05). *P<0.05, compared with group B.
will increase under renal damage (19). Our study showed that berberine pretreatment can effectively inhibit the occurrence of IRI (19). MDA, as the final products of lipid in oxidation reactions under high levels of oxidative free radicals, can reflect the degree of oxidation reaction (20). Therefore, plasma level of MDA was detected. MDA level was significantly increased in group B compared with other groups, indicating that free radical-induced peroxide is one of the causes of tissue damage. Levels of MDA in group B were significantly higher than those in groups C and D, which demonstrate that pretreatment with berberine can effectively reduce the expression of MDA. Studies have shown that (21) Bcl-2 family members play pivotal roles in cell apoptosis, and regulation of Bax and Bcl-2 protein expression can effectively reduce the apoptosis. In this study, western blot analysis results showed that berberine pretreatment significantly reduced the expression levels of Bax protein and increased the expression of Bcl-2 protein in group C and D compared with group B. Studies have shown that (22) Bax expression is upregulated under cell apoptosis, Bax overexpression can promote the cell apoptosis, while Bax expression inhibition can inhibit cell apoptosis. Bcl-2 can bind to Bax to form heterodimers, thereby decreasing the expression of Bax and inhibiting cell apoptosis. TUNEL assay showed that berberine pretreatment significantly improved cell apoptosis in group C and D compared with group B. However, cell apoptosis in group C and D was still more severe than that in group A. Therefore, berberine pretreatment can effectively reduce the severity of IRI, but cannot inhibit its occurrence.

However, this study is still limited by some shortcomings. Sample size is small, and the effects caused by different surgical methods and different ways of anesthesia were not excluded. In addition, only animal experiments were performed, but clinical studies were not included. Those shortcomings will be improved in our future studies.

In conclusion, berberine can effectively improve renal function in rats with renal IRI possibly by inhibiting Bax expression and promoting Bcl-2 expression.

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Availability of data and materials
All data generated or analyzed during this study are included in this published article.

Authors' contributions
HZ and JuL designed the study. JiL and LL performed the animal experiments. HZ, JuL and LL analyzed the animal experiment data. All authors read and approved the final manuscript.

Ethics approval and consent to participate
The study was approved by the Ethics Committee of People's Hospital of Lishui Animal Center (Zhejiang, China).

Patient consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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