Extraction of volatile organic compounds from leaves of Ambrosia artemisiifolia L. and Artemisia annua L. by headspace-solid phase micro extraction and simultaneous distillation extraction and analysis by gas chromatography/mass spectrometry

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Abstract This study was designed to analyze the volatile organic compounds in the leaves of Ambrosia artemisiifolia L. and Artemisia annua L. from Korea. For extraction of volatile compounds, headspace-solid phase micro extraction (HS-SPME) and simultaneous distillation extraction (SDE) were applied and analyzed by gas chromatography/mass spectrometry (GC/MS). From the results, SDE extraction was found to give the highest concentration of volatile compounds with an average concentration of 1,237.79 mg/kg for A. annua L. leaves compared to 1,122.73 mg/kg by HS-SPME technique. A total of 116 volatile organic compounds were identified, including 76 similar volatile organic compounds detected by both the methods of extraction in leaves of subject species at varying concentrations. Among these 33 volatile organic compounds were reported for the first time from the subject plant species. Thus the present research findings extend the characterization of volatile organic compounds from leaves of A. annua L. and A. artemisiifolia L. species and reported some distinguishing compounds which may be used for their discrimination.

Keywords Ambrosia artemisiifolia L. leaves · Artemisia annua L. leaves · Volatile organic compound · Headspace-solid phase micro extraction (HS-SPME) · Simultaneous distillation extraction (SDE) · Gas chromatography/mass spectrometry (GC/MS)

Introduction
Ambrosia artemisiifolia L. (common ragweed) leaves are highly allergenic pollens, causing great agricultural losses (Molinaro et al., 2016). These have negative impact on the diversity of plant species, richness and the composition of their vegetation is a topic of on-going discussion (Sölter
et al., 2012). This species is indigenous to North America (Basset and Crompton, 1975) but also found abroad in South Asia, such as South Korea (Kil et al., 2004) and China (Sang et al., 2011; Xie et al., 2010). The allopathic action of *A. artemisiifolia* L. leaves has already been reported in literature, while knowledge of their volatile organic compounds was still lacking.

In comparison, *Artemisia annua* L. (sweet wormwood) is a medicinal herb; native to South East Asia, including South Korea, China, and India (Wu et al., 2017). This plant has been used for many centuries in the treatment of malaria, fever, flavouring of sprit, perfumes, folk medicine and industrial purposes (Bilia et al., 2008; Ma et al., 2007; Nekoei et al., 2012). Recently, *A. annua* L. leaves were found to be effective against human leukemia, tumour, small-cell lung carcinomas, and breast cancer carcinomas (Efferth and Willmar, 2007). Due to the similar appearance, consumers and manufacturers are confused to separate *A. annua* L. leaves in the market from *A. artemisiifolia* L. leaves.

Headspace-solid phase micro extraction (HS-SPME) is a quick, solvent less, and simple method for analysis of volatile organic compounds (Holt, 2001; Kataoka et al., 2000; Zhang et al., 1994). This technique is a non-invasive and non-destructive that avoids solvent impurity contamination (Heath and Reineccius, 1986). It combines sampling, extraction, and concentration into a simple continuous process (Adam et al., 2005). In contrary, Likens and Nickerson developed simultaneous distillation–extraction (SDE) is a traditional and widely used method for extraction of volatile organic compounds (Islam et al., 2020). Gu et al. (2009) indicated that SDE has outstanding high performance and reproducibility compared to other conventional methods such as hydro distillation and steam distillation.

Some wild toxic plants are morphologically very similar to important medicinal plants. When such wild toxic plants are accidentally taken, instead of medicinal plants for different purposes, they cause many problems. According to Kim and Jang (2019), in South Korea, 42 patients faced abdominal pain after intake of *A. artemisiifolia* L. leaves instead of *A. annua* L. leaves. So, it was very important to understand the volatile organic compounds characteristics of leaves of both the subject species selected for current study and to find out the distinguishing marker compounds. From literature search it was come to know that HS-SPME and SDE techniques were not used to isolate volatile organic compounds from *A. artemisiifolia* L. leaves and also there was no comparative study found on the volatile organic compounds of *A. artemisiifolia* L. leaves and *A. annua* L. leaves. In the current analysis, therefore both HS-SPME and SDE techniques coupled to GC/MS analysis were applied to establish the comparative results of volatile organic compounds profile of *A. artemisiifolia* L. leaves and *A. annua* L. leaves. It was also attempted to report distinguishing marker volatile organic compounds which may be used for their identification.

## Materials and methods

### Sample collection

Figures and taxonomy of the sampled plants are shown in Fig. 1. In February 2018, the authentic *A. artemisiifolia* L. leaves and *A. annua* L. leaves were collected in triplicate from Korea native plants growers farm association corporation, Seongnam-si, Republic of Korea. These two leave varieties were thoroughly washed with clean water, followed by rinsing with distilled water. Then, using a blender (MR 350CA, Braun, Spain) the leaves were crushed, transferred to polythene bags, properly labelled and used for volatile compounds extraction and GCMS analysis.

### Chemicals and reagents

All chemicals and reagents used in this study were purchased from Sigma-Aldrich Co. (St. Louis, MO, USA) and Thermo Fisher Scientific (Waltham, MA, USA). The water purification system (Millepore Corporation, Beds., USA) was used to prepare ultra-pure water. For the re-distillation of the organic solvents (n-pentane and diethyl ether), a wire spiral packed double distilling apparatus (Normschliff Geratebau, Wertheim, Germany) was used. Anhydrous sodium sulphate was burned overnight at 650 °C in furnace (F 6000, Barnsted Thermolyne Co., IA., USA) and then used for dehydration of organic solvent.

### Extraction of volatile compounds using HS-SPME technique

Around 2.0 g of each homogenized sample of *A. artemisiifolia* L. leaves and *A. annua* L. leaves were taken into 15.0 mL headspace clear glass vials (Supelco, PA., USA) and then injected 50.0 μL (100 ppm in n-pentane) n-butyl benzene as an internal standard. After that, the headspace glass vial was hold on for 30 min in equilibrium temperature at 70 °C and then SPME fibre (50/30 μm DVB/CAR/PDMS, Supelco) was exposed for 5 min to absorb volatile organic compounds. Finally, the temperature of the injector was maintained at 250 °C, and the fibre was kept in GC/MS objector for 5 min to analyse the volatile organic compounds.
Extraction of volatile compounds using SDE technique

Every 30.0 g homogenized sample was mixed with 1,000 mL of distilled water and the pH was adjusted at 7.0 by dilute NaOH/HCl solution. For the quantitative analysis of volatile organic compounds, 100 ppm, 10 mL n-butyl benzene was added as an internal standard. Volatile organic compounds were extracted from both leaves by modified simultaneous distillation extraction (SDE, Likens & Nickerson types) apparatus with 100 mL redistilled n-pentane:diethyl ether (1:1, v/v) mixture. The experiment was maintained under normal atmospheric pressure for 3 h (Schultz et al., 1977). The SDE extract was dehydrated overnight by adding 10 g of anhydrous Na₂SO₄. Finally, the vigreux column was used to concentrate the extract volume to 1.5 mL. After that, it was again concentrated to 0.5 mL under N₂ gas mild flush. Finally, the concentrated extract was injected into the GC/MS system for determination of volatile organic compounds (Khan et al., 2015).

Analysis of volatile organic compounds by GC/MS

The quantitative analysis of volatile organic compounds was carried out by Shimadzu GC/MS, QP-2010 (Shimadzu, Japan) with the EI (electron impact) mode. The ionization voltage was 70 eV, and temperatures of the injector and of ion source were maintained at 250 °C and 220 °C, respectively. The volatile organic compounds were isolated by GC column DB-5 (60 m × 0.25 mm i.d., film thickness 0.25 μm, Phenomenex, USA) and the mass spectra scanned were from 41 to 500 m/z. The GC/MS oven temperature was set as follows; 40 °C (5 min isothermal) raised to 220 °C at 2 °C/min and then to 280 °C at 10 °C/min (10 min isothermal). Helium gas at flow rate of 1.0 mL/min was used as the carrier gas. The sample injector volume was 1.0 μL and the split ratio was 1:20 (Islam et al., 2020).

Identification and quantification of volatile organic compounds

The A. artemisiifolia L. leaves and A. annua L. leaves volatile organic compounds were identified by the spectral databases, including NIST 12, FFNSC 2012, and WILEY 7. In addition, our own mass spectral data books were applied (Davies, 1990). Moreover, evaluations of retention indices to reference data were studied (Adams, 2007). The retention time of solutes of standard n-alkanes (C₈–C₂₀) mixture was used as an external reference (Jeon et al., 2017). The quantitative assessment of volatile organic compounds was carried out with the help of peak area percentage of the internal standard by using the formula:

\[
\text{Volatile organic compounds amount (mg/kg)} = \frac{P_c \times 1000}{P_i \times A_s}
\]

where, \(P_i\) = Peak area (internal standard).
\(A_s\) = Amount of sample (g).
\(P_c\) = Peak area (component in sample).

Statistical Analysis

The volatile organic compounds were calculated in triplicate, and the data was evaluated using the Software Version 20 (IBM, New York, USA) of Statistical Package for Social Sciences (SPSS). The final results were reported as mean ± standard deviation (mg/kg).
Results and discussions

The GC/MS chromatograms of volatile compounds of various extracts from A. artemisiifolia L. leaves and A. annua L. leaves via HS-SPME and SDE techniques were as displayed in Fig. 2a–d. The identified volatile organic compounds detail such as retention index (RI), compound names, molecular formulae (MF), molecular weights (MW), peak areas %, and their concentrations were as shown in Table 1. From the results it was found that both the leaves contain a wide range of volatile organic compounds. In published literature, it has been reported that both the leaves contain around 30–35 compounds (Nekoei et al., 2012; Wang et al., 2006).

In the current analysis, leaves of both plants were extracted for volatile compounds by HS-SPME and SDE. The extracts were analyzed by GC/MS, which identified more compounds compared to already reported studies (Haghighian et al., 2008). A total of 116 volatile organic compounds were identified from the subject leaves and this study thus reported 33 volatile organic compounds for the first time in comparison to published literature (Table 1, marked bold). The total amount of volatile organic compounds for A. artemisiifolia L. leaves and A. annua L. leaves ranged from 651.53 mg/kg to 1,122.73 mg/kg and 566.1 mg/kg to 1,237.79 mg/kg through HS-SPME and SDE techniques, respectively.

Volatile compounds of A. artemisiifolia L. leaves by HS-SPME technique

A total of 46 volatile organic compounds were identified in A. artemisiifolia L. leaves amounting 651.53 mg/kg (Table 1). These included 28 hydrocarbons (76.49%, 499.20 mg/kg), 7 alcohols (11.62%, 75.81 mg/kg), 4 ketones (3.92%, 25.59 mg/kg), 3 ethers (4.09%, 26.70 mg/kg), 3 aldehydes (0.61%, 3.97 mg/kg), and 1 ester (3.1%, 20.26 mg/kg) (Table 2). The terpene group compounds (99.12%, 646.91 mg/kg) were the maximum in A. artemisiifolia L. leaves; which included 21 sesquiterpene hydrocarbons (67.58%, 441.00 mg/kg), 6 oxygenated sesquiterpenes (12.73%, 83.12 mg/kg), 8 oxygenated monoterpenes (9.9%, 64.59 mg/kg), and 7 monoterpene hydrocarbons (8.91%, 58.20 mg/kg) (Table 3). The highest concentration of volatile organic compounds were of β-caryophyllene (12.08%, 78.64 mg/kg), followed by γ- muurolene (6.49%, 42.39 mg/kg), spathulenol (6.13%, 40.04 mg/kg), and δ-cadinene (4.5%, 29.40 mg/kg). A. artemisiifolia L. leaves gave 6 compounds including phenyl acetaldehyde, (E)-β-ocimene, artemisia alcohol, germacrene B, palustrol, and γ-humuleneepoxide II by HS-SPME which were not detected by SDE extraction (Table 1).

Volatile compounds of A. annua L. leaves by HS-SPME technique

From Table 1, the amount of volatile organic compounds from A. annua L. leaves was found as 566.10 mg/kg by HS-SPME method. These included a total of 41 volatile organic compounds, with 17 hydrocarbons (27.67%, 153.84 mg/kg), 7 ketones (21.96%, 122.07 mg/kg), 7 alcohols (8.48%, 37.92 mg/kg), 5 aldehydes (1.19%, 6.57 mg/kg), 3 ethers (39.96%, 229.35 mg/kg), and 2 esters (2.97%, 16.35 mg/kg). As per Table 3, terpene group compounds were having the highest concentration (97.26%, 540.18 mg/kg), which consisted of 5 oxygenated sesquiterpenes (47.93%, 275.01 mg/kg), 13 sesquiterpene hydrocarbons (26.25%, 146.04 mg/kg), 13 oxygenated monoterpenes (21.66%, 111.33 mg/kg), and 4 monoterpane hydrocarbons (1.42%, 7.80 mg/kg). The major volatile organic compounds were caryophyllene oxide (31.90%, 186.48 mg/kg) followed by camphor (11.80%, 65.76 mg/kg), β-selinene (8.29%, 46.65 mg/kg), β-caryophyllene (5.76%, 31.68 mg/kg), alloaromadendrene epoxide (5.33%, 29.31 mg/kg), phytone (5.02%, 27.99 mg/kg), and aromandendrene (5.02%, 27.99 mg/kg). The HS-SPME method found 6 compounds such as cuminaldehyde, benzyl pentanoate, α-acoradiene, alloaromadendrene epoxide, 4-cadinene-7-ol, and phytone, which were not detected by SDE analysis (Table 2). In literature other studies have also reported artemisia ketone, yomogi alcohol, camphor, and β-selinene in A. annua L. leaves (Nekoei et al., 2012; Reale et al., 2011). Thus current research findings are line with the published research studies.

Volatile compounds of A. artemisiifolia L. leaves by SDE technique

The amount of volatile organic compounds in A. artemisiifolia L. leaves by SDE extraction was 1,122.73 mg/kg. A total of 49 volatile organic compounds were reported (Table 1), belonging to different chemical groups such as 26 hydrocarbons (51.76%, 577.58 mg/kg), 12 alcohols (32.29%, 360.52 mg/kg), 5 aldehydes (3.42%, 38.16 mg/kg), 4 ketones (1.79%, 19.96 mg/kg), 1 ether (9.42%, 105.12 mg/kg), and 1 ester (1.92%, 21.39 mg/kg). The terpene group (94.67%, 1,056.58 mg/kg) was the major class of volatile organic compounds in A. artemisiifolia L. leaves, with 22 sesquiterpene hydrocarbons (45.01%, 502.32 mg/kg), 5 oxygenated sesquiterpenes (31.84%, 355.39 mg/kg), 6 oxygenated monoterpenes (11.07%, 123.61 mg/kg), and 37 monoterpene hydrocarbons (6.75%, 75.26 mg/kg) (Table 3). The major volatile
organic compounds detected were spathulenol (12.45%, 138.92 mg/kg), caryophyllene oxide (9.42%, 105.12 mg/kg), α-cumene (5.75%, 64.22 mg/kg), valencene (5.26%, 58.96 mg/kg), borneol (5.09%, 56.82 mg/kg), and β-caryophyllene (4.61%, 51.45 mg/kg). A. artemisiifolia L. leaves were found to have 12 volatile organic compounds such as sulcatone, benzene acetaldehyde, linalool, β-bourbonene, β-cubebene, neryl acetone, β-sesqui sabine, junenol, and α-guaiol which were reported by SDE and not by HS-SPME technique (Table 1). Wang et al., (2006) have reported by steam distillation method that the germacerene D, limonene, and α-pinene are present in A. artesimisifolia L. leaves.

Volatile compounds of A. annua L. leaves by SDE extraction

As shown in Table 1, the amount of volatile organic compounds in A. annua L. leaves was 1,237.79 mg/kg by SDE method. These were 71 compounds belonging to specific chemical functional groups including 26 alcohols (26.47%, 323.25 mg/kg), 22 hydrocarbons (31.35%, 390.32 mg/kg), 11 aldehydes (1.87%, 23.41 mg/kg), 7 ketones (16.01%, 199.05 mg/kg), 3 ethers (21.49%, 267.14 mg/kg), 1 ester (2.16%, 26.88 mg/kg). The highest proportion was of terpene group compounds (92.02%, 1,138.84 mg/kg) which included 23 oxygenated monoterpenes, 11 sesquiterpene hydrocarbons, 2 oxygenated sesquiterpenes, and 11 monoterpenic hydrocarbons having 40.17%, 493.56 mg/kg; 27.80%, 346.09 mg/kg; 20.50%, 254.96 mg/kg and 3.55%, 44.23 mg/kg, respectively.
| No | R.I. | Compound name          | M.F. | M.W. | HS-SPME | SDE               |
|----|------|------------------------|------|------|---------|-------------------|
|    |      |                        |      |      |         |                  |
|    |      | *Ambrosia artemisiifolia L* |      |      |         |                  |
|    |      |                        |      |      |         |                  |
|    |      | *Artemisia annua L.* |      |      |         |                  |
|    |      |                        |      |      |         |                  |
| 1  | 706  | 3-Methylbutanalb       | C₅H₁₀O | 87   | –       | –               |
| 2  | 719  | 1-Pentene-3-ol        | C₅H₁₀O | 86   | –       | –               |
| 3  | 727  | Pentanalb             | C₅H₁₀ | 86   | –       | –               |
| 4  | 729  | 2-Ethylfuran         | C₆H₁₂  | 96   | –       | –               |
| 5  | 754  | 2-Methyl-1-butanol   | C₅H₁₀O | 88   | –       | –               |
| 6  | 767  | (E)-2-Pentenalb      | C₅H₁₀O | 84   | –       | –               |
| 7  | 775  | 1-Pentanolb           | C₅H₁₀O | 88   | –       | –               |
| 8  | 794  | 2-Hexanone           | C₅H₁₂O | 100  | –       | –               |
| 9  | 802  | Hexanalb              | C₅H₁₀O | 100  | 0.15 ± 0.00 | 0.95 ± 0.04   |
| 10 | 836  | Furfuralb             | C₅H₁₀O | 96   | –       | –               |
| 11 | 855  | (E)-2-Hexenalb       | C₅H₁₀O | 98   | –       | –               |
| 12 | 857  | (Z)-3-Hexen-1-ol     | C₅H₁₀O | 100  | –       | –               |
| 13 | 871  | 1-Hexanolb           | C₅H₁₂O | 102  | –       | –               |
| 14 | 880  | Heptanalb             | C₆H₁₂O | 114  | –       | –               |
| 15 | 894  | 2-Heptanone          | C₆H₁₂O | 114  | –       | –               |
| 16 | 928  | *-Thujene             | C₆H₁₄O | 136  | –       | –               |
| 17 | 935  | *-Pinene             | C₆H₁₄O | 136  | 1.87 ± 0.00 | 12.19 ± 0.02   |
| 18 | 950  | Camphene             | C₆H₁₄O | 136  | 0.15 ± 0.00 | 1.00 ± 0.00   |
| 19 | 956  | 2,4-Thujadiene       | C₆H₁₄O | 134  | –       | –               |
| 20 | 960  | (E)-2-heptenalb      | C₆H₁₄ | 134  | –       | –               |
| 21 | 965  | Benzaldehyde         | C₆H₁₄O | 196  | 0.29 ± 0.02 | 1.88 ± 0.02   |
| 22 | 975  | Sabine               | C₆H₁₄O | 136  | 0.10 ± 0.00 | 0.67 ± 0.00   |
| 23 | 979  | *-Pinene             | C₆H₁₄O | 136  | 0.12 ± 0.00 | 0.79 ± 0.00   |
| 24 | 990  | Salicytone           | C₇H₈O | 126  | –       | –               |
| 25 | 992  | *-Myrcene            | C₇H₈O | 136  | 1.80 ± 0.02 | 11.73 ± 0.08   |
| 26 | 1001 | Yomogi alcohol       | C₈H₁₀O | 154  | –       | –               |
| 27 | 1015 | (E,E)-2,4-Heptadienalb | C₈H₁₀O | 110  | –       | –               |
| 28 | 1019 | *-Terpinene          | C₈H₁₄O | 136  | –       | –               |
| 29 | 1027 | p-Cymene             | C₈H₁₄O | 134  | –       | –               |
| 30 | 1031 | Limonene             | C₈H₁₄ | 136  | 3.89 ± 0.02 | 25.40 ± 0.04   |
| 31 | 1035 | 1,8-Cineole          | C₈H₁₄O | 154  | 0.47 ± 0.00 | 3.04 ± 0.02   |
| 32 | 1040 | Benzyl alcohol       | C₈H₁₀O | 108  | –       | –               |
| 33 | 1049 | Benzene acetaldehyde | C₉H₁₂O | 120  | –       | –               |
| 34 | 1050 | Phenyl acetaldehyde | C₉H₁₂O | 120  | 0.17 ± 0.00 | 1.14 ± 0.00   |
| 35 | 1052 | (E,)-β-Ocimene      | C₉H₁₄O | 136  | 0.98 ± 0.02 | 6.42 ± 0.04   |
| LS | 1059 | n-Butylbenzene      | C₉H₁₄O | 134  | –       | –               |
| No. | Rf | Compound name | M.W. | HS-SPME | M.W. | SDE | Area (%) | Area (%) | Area (%) |
|-----|----|----------------|------|---------|------|-----|----------|----------|----------|
| 36  | 0.06 | 2-Terpineol | 136 | 0.06 | 0.06 | - | - | - | - |
| 37  | 0.07 | 2-Acetophenone | 152 | 0.07 | 0.07 | - | - | - | - |
| 38  | 0.08 | 2-Hexanone | 154 | 0.08 | 0.08 | - | - | - | - |
| 39  | 0.11 | 2-Methylpropanoic acid | 150 | 0.11 | 0.11 | - | - | - | - |
| 40  | 0.13 | 2-Butanone | 150 | 0.13 | 0.13 | - | - | - | - |
| 41  | 0.15 | 2-Methylbutanone | 150 | 0.15 | 0.15 | - | - | - | - |
| 42  | 0.17 | 2-Heptanone | 150 | 0.17 | 0.17 | - | - | - | - |
| 43  | 0.20 | 2-Octanone | 154 | 0.20 | 0.20 | - | - | - | - |
| 44  | 0.22 | 2-Nonanone | 158 | 0.22 | 0.22 | - | - | - | - |
| 45  | 0.24 | 2-Decanone | 162 | 0.24 | 0.24 | - | - | - | - |
| 46  | 0.27 | 2-Tridecanone | 166 | 0.27 | 0.27 | - | - | - | - |
| 47  | 0.30 | 2-Tetradecanone | 170 | 0.30 | 0.30 | - | - | - | - |

Table 1 continued
| No | R.I. | Compound name | M.F. | M.W. | HS-SPME | SDE |
|----|------|---------------|------|------|----------|-----|
|    |      |               | C15H24 | 204 |          |     |
| 72 | 1391 | (Z)-β-Elemene | 2.14 ± 0.00 | 13.95 ± 0.10 | – | – |
| 73 | 1392 | Modiphenene   | 0.23 ± 0.00 | 1.26 ± 0.00 | – | – |
| 74 | 1395 | Benzyl pentanoate | 0.26 ± 0.01 | 1.44 ± 0.02 | – | – |
| 75 | 1396 | β-Bourbonene   | 0.42 ± 0.00 | 4.67 ± 0.02 | – | – |
| 76 | 1399 | β-Cubebene    | 2.60 ± 0.02 | 28.96 ± 0.16 | – | – |
| 77 | 1409 | (Z)-Jasmone    | 0.60 ± 0.02 | 7.41 ± 0.30 | – | – |
| 78 | 1421 | α-Gurjumene   | 0.96 ± 0.00 | 10.73 ± 0.08 | – | – |
| 79 | 1424 | β-Caryophyllene | 4.61 ± 0.04 | 51.45 ± 0.22 | 7.80 ± 0.04 | 97.04 ± 0.33 |
| 80 | 1441 | β-Copaene     | 1.19 ± 0.02 | 13.27 ± 0.06 | 0.59 ± 0.02 | 7.40 ± 0.20 |
| 81 | 1443 | (Z)-α-Bergamotene | 3.21 ± 0.04 | 35.82 ± 0.10 | – | – |
| 82 | 1444 | Aromandrened  | – | – | – | – |
| 83 | 1455 | Germacrene D  | 0.59 ± 0.00 | 6.62 ± 0.02 | – | – |
| 84 | 1456 | Dihydropseudoionone | 0.14 ± 0.00 | 0.75 ± 0.00 | – | – |
| 85 | 1461 | (E)-β-Farnesene | 1.27 ± 0.04 | 15.85 ± 0.06 | – | – |
| 86 | 1466 | α-Caryophyllene | 2.92 ± 0.04 | 32.58 ± 0.12 | 1.31 ± 0.02 | 16.31 ± 0.08 |
| 87 | 1468 | (Z)-4,5-Muroladiene | 2.92 ± 0.04 | 32.58 ± 0.12 | 1.31 ± 0.02 | 16.31 ± 0.08 |
| 88 | 1471 | α-Acoradiene  | – | – | – | – |
| 89 | 1483 | α-Curcumene   | 5.75 ± 0.04 | 64.22 ± 0.20 | – | – |
| 90 | 1488 | γ-Murolene    | 1.01 ± 0.02 | 11.28 ± 0.08 | 3.79 ± 0.01 | 47.18 ± 0.44 |
| 91 | 1490 | Valencene     | 5.26 ± 0.04 | 58.69 ± 0.18 | – | – |
| 92 | 1494 | Germacrene D  | 2.14 ± 0.02 | 26.66 ± 0.24 | – | – |
| 93 | 1496 | (E)-β-Ionone  | – | – | – | – |
| 94 | 1500 | β-Seineine    | 1.62 ± 0.02 | 18.10 ± 0.08 | 4.20 ± 0.03 | 52.23 ± 0.13 |
| 95 | 1505 | β-Seineine    | 0.90 ± 0.00 | 10.05 ± 0.02 | – | – |
| 96 | 1506 | α-Murolene    | 1.37 ± 0.02 | 15.25 ± 0.04 | – | – |
| 97 | 1509 | β-Isobolene   | 1.70 ± 0.02 | 18.96 ± 0.06 | – | – |
| 98 | 1514 | γ-Cadinene    | 1.18 ± 0.01 | 14.71 ± 0.04 | – | – |
| 99 | 1517 | β-Sesquisabinene | 1.83 ± 0.02 | 20.46 ± 0.06 | – | – |
| 100 | 1518 | α-Cadinene    | 2.75 ± 0.05 | 30.74 ± 0.10 | 2.47 ± 0.01 | 30.77 ± 0.10 |
| 101 | 1523 | (E)-Cadin-1,4-diene | 1.41 ± 0.02 | 15.68 ± 0.05 | – | – |
| 102 | 1529 | α-Calacorene  | 1.82 ± 0.02 | 20.27 ± 0.03 | – | – |
| 103 | 1536 | (E)-Nerolidol | 3.04 ± 0.00 | 33.91 ± 0.08 | – | – |
| 104 | 1537 | (Z)-Nerolidol | – | – | – | – |
| 105 | 1553 | Caryophyllene oxide | 2.11 ± 0.02 | 20.80 ± 0.04 | – | – |
| 106 | 1564 | Germacrene B  | – | – | – | – |
| No | R.I | Compound name | M.F | M.W | HS-SPME | SDE |
|----|-----|---------------|-----|-----|---------|-----|
|    |     |               |     |     | Ambrosia artemisiifolia L | Artemisia annua L | Ambrosia artemisiifolia L | Artemisia annua L |
|    |     |               | Area (%) | (mg/kg) | Area (%) | (mg/kg) | Area (%) | (mg/kg) |
| 108| 1570| Junenol<sup>a</sup> | C<sub>15</sub>H<sub>26</sub>O | 222 | – | – | 2.88 ± 0.04 | 32.19 ± 0.08 |
| 109| 1571| α-Guaiol<sup>a</sup> | C<sub>15</sub>H<sub>26</sub>O | 222 | – | – | 4.05 ± 0.06 | 45.25 ± 0.06 |
| 110| 1576| Palustrol<sup>b</sup> | C<sub>15</sub>H<sub>26</sub>O | 222 | 0.27 ± 0.00 | 1.73 ± 0.01 | – | – |
| 111| 1578| Spathulenol<sup>c</sup> | C<sub>15</sub>H<sub>26</sub>O | 220 | 6.13 ± 0.10 | 40.04 ± 0.22 | – | – |
| 112| 1610| Salvial-4(14)-en-1-one<sup>d</sup> | C<sub>15</sub>H<sub>25</sub>O | 220 | 1.81 ± 0.08 | 11.84 ± 0.20 | 2.58 ± 0.02 | 14.22 ± 0.06 |
| 113| 1616| α-Humuleneoxide II<sup>e</sup> | C<sub>15</sub>H<sub>25</sub>O | 220 | 0.31 ± 0.00 | 2.05 ± 0.02 | – | – |
| 114| 1626| Alloaromadendrene epoxide<sup>f</sup> | C<sub>15</sub>H<sub>25</sub>O | 220 | – | – | 5.33 ± 0.08 | 29.31 ± 0.04 |
| 115| 1649| 4-Cadinene-7-ol<sup>g</sup> | C<sub>15</sub>H<sub>25</sub>O | 222 | – | – | 3.10 ± 0.02 | 17.01 ± 0.06 |
| 116| 1847| Phytone<sup>h</sup> | C<sub>18</sub>H<sub>36</sub>O | 268 | – | – | 5.02 ± 0.02 | 27.99 ± 0.11 |
|    |     | **Total**     |     |     | 100.00 | 651.53 | 100.00 | 566.10 |

Bold mark compounds name = Firstly reported volatile organic compounds in this study, – = Not detected

<sup>a</sup> Alcohol
<sup>b</sup> Aldehyde
<sup>c</sup> Ester
<sup>d</sup> Hydrocarbon
<sup>e</sup> Ketone
<sup>f</sup> Ether
<sup>g</sup> Miscellaneous

Data were reported by mean ± standard deviation (n = 3); 0.00 = The value is less than 0.01, *RI Retention index, **M.F Molecular formula, ***M.W Molecular weight, I.S. Internal standard;
Table 3. The major volatile organic compounds were caryophyllene oxide (18.83%, 234.16 mg/kg) followed by camphor (10.66%, 132.61 mg/kg), \( \beta \)-caryophyllene (7.80%, 97.04 mg/kg), (E)-pinocarveol (5.27%, 65.55 mg/kg), and \( \beta \)-selinene (4.20%, 52.23 mg/kg). In the current study, 14 volatile organic compounds were found only by SDE compared to HS-SPME method. Both extractions of A. artemisiifolia L. leaves identified compounds such as \( \gamma \)-terpinene, phenethyl alcohol, \( \alpha \)-campholenal, camphene hydrate, (Z)-chrysanthenol, p-cymen-8-ol, \( \alpha \)-terpineol, (Z)-carveol, indole, 2-methoxy-4-vinylphenol, eugenol, (Z)-jasmone, and (Z)-nerolidol (Table 1). In literature, similar compounds such as erythritol, camphor, germacrene D, artesimia ketone, \( \alpha \)-caryophillene, \( \alpha \)-cuvebene have already been detected by steam distillation and hydrodistillation in A. artemisiifolia L. leaves (Bilia et al., 2008; Haghighian et al., 2008; Juteau et al., 2002; Tzenkova et al., 2010; Vidic et al., 2018).

**Comparison of extraction techniques and identification marker compounds**

Headspace-solid phase micro extraction (HS-SPME) is used to extract volatile organic compounds with short time and to extract more volatile monoterpenes. Simultaneous distilled extraction (SDE) is capable of extracting higher amount of volatile organic compounds and requires longer extraction time at high temperature. Both techniques have been used for volatile compounds in the past around the world (Majcher and Jelenı́, 2009). Some researchers have compared HS-SPME with other conventional methods and have reported that the HS-SPME method as more significant than other traditional methods for the determination of volatile organic compounds in herbs (Majcher and Jelenı́ 2009; Yang et al., 2011). The SDE technique may subsequently increase the low volatile organic compounds with a high molecular weight, such as straight-chain acids, and sesquiterpenes.
In the current study, 116 volatile organic compounds were reported, with 11 compounds (hexanal, α-pinene, benzaldehyde, limonene, borneol, α-copaene, β-caryophyllene, α-caryophyllene, β-selinene, δ-cadinene, and caryophyllene oxide) identified in the leaves of both species applying the 2 extraction methods. From Table 3, the sesquiterpene hydrocarbons (67.58%, 441 mg/kg) were relatively in higher content in A. artemisiifolia L. leaves whereas oxygenated sesquiterpenes (47.93%, 275.01 mg/kg) were detected in higher concentrations in A. annua L. leaves by HS-SPME technique. Both HS-SPME and SDE methods extracted more terpene hydrocarbons from A. artemisiifolia L. leaves and oxygenated terpene compounds from A. annua L. leaves.

Volatile organic compounds are liable for flavour, aroma and bioactivity. In addition, these have importance for chemical fingerprinting. The chemical elements have various forms of plant essential oils, which are regularly used to identify the variety or to improve the chemotypes. In the current study, both leaves were found to consist of mainly hydrocarbons, ethers, and alcoholic type of volatile organic compounds.

Each country and possibly each region may have a distinct type of A. annua L. leaves and A. artemisiifolia L. leaves essential oil, called chemotypes. Different chemotypes such as erythritol, champhore, germacrene D, artemisia ketone, α-caryophyllene, and α-cuvebene have been reported for most of the A. annua L. leaves essential oil in literature (Bilia et al., 2008; Haghighian et al., 2008; Juteau et al., 2002; Tzenkova et al., 2010; Vidic et al., 2018; Wang et al., 2006). In this study, spathulenol, caryophyllene oxide, and β-caryophyllene chemotypes were observed in A. artemisiifolia L. leaves whereas caryophyllene oxide, champhor, and β-selinene chemotypes were detected in A. annua L. leaves. This variation may be due to the geographical and ecological conditions, time of harvesting, and age of the plant (Bagamboula et al., 2004).

In conclusion, this study determined the volatile organic compounds from A. artemisiifolia L. leaves and A. annua L. leaves using HS-SPME and SDE extraction methods coupled with GC/MS analysis. The results demonstrated remarkable differences in identified volatile compounds concentration in both leaves by 2 extraction methods. The highest concentration of volatile organic compounds was detected in A. annua L. leaves (1,237.79 mg/kg) by SDE technique whereas the lowest value was observed in the same leaves (566.1 mg/kg) by HS-SPME extraction. A total of 116 volatile components were identified, extracted by SDE compared to 71 volatile organic compounds by HS-SPME technique, while 33 volatile organic compounds were detected for the first time compared to published literature on A. artemisiifolia L. leaves and A. annua L. leaves. The common major volatile organic compounds (caryophyllene oxide and champhor) were detected in leaves of both samples. The identified marker compounds in A. artemisiifolia L were γ-muurolene and spathulenol while β-selinene and caryophyllene were reported in A. annua L. leaves. Thus, the present findings extend the characterization of volatile organic compounds of A. annua L. leaves and A. artemisiifolia L. leaves and reported some distinguishing compounds which may be used for their characterization.

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Compliance with ethical standards

Conflict of interest Authors have no conflict of interest.

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