Activities of the peptidyl transferase center of ribosomes lacking protein L27

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ABSTRACT
The ribosome is the molecular machine responsible for protein synthesis in all living organisms. Its catalytic core, the peptidyl transferase center (PTC), is built of rRNA, although several proteins reach close to the inner rRNA shell. In the Escherichia coli ribosome, the flexible N-terminal tail of the ribosomal protein L27 contacts the A- and P-site tRNA. Based on computer simulations of the PTC and on previous biochemical evidence, the N-terminal α-amino group of L27 was suggested to take part in the peptidyl-transfer reaction. However, the contribution of this group to catalysis has not been tested experimentally. Here we investigate the role of L27 in peptide-bond formation using fast kinetics approaches. We show that the rate of peptide-bond formation at physiological pH, both with aminoacyl-tRNA or with the substrate analog puromycin, is independent of the presence of L27; furthermore, translation of natural mRNAs is only marginally affected in the absence of L27. The pH dependence of the puromycin reaction is unaltered in the absence of L27, indicating that the N-terminal α-amino is not the ionizing group taking part in catalysis. Likewise, L27 is not required for the peptidyl-tRNA hydrolysis during termination. Thus, apart from the known effect on subunit association, which most likely explains the phenotype of the deletion strains, L27 does not appear to be a key player in the core mechanism of peptide-bond formation on the ribosome.

Keywords: ribosome; translation; peptide-bond formation; tRNA; ribozyme

INTRODUCTION
In all living organisms, the ribosome catalyzes the sequential polymerization of amino acids into functional proteins. In each round of translation elongation, the ribosome selects an aminoacyl-tRNA (aa-tRNA) corresponding to the mRNA codon presented in the A site. After accommodation of the 3′ end of the A-site tRNA in the peptidyl transferase center (PTC) of the ribosome, the amino group of the aa-tRNA nucleophilically attacks the ester of the peptidyl-tRNA in the P site leading to the formation of a peptide bond and the transfer of the peptidyl chain onto the A-site tRNA. The PTC resides in the large subunit of the ribosome (50S in bacteria) and is constituted mainly by rRNA, which is thought to be responsible for the catalytic activity. However, crystal structures of the Thermus thermophilus ribosome in complex with tRNAs (Selmer et al. 2006; Voorhees et al. 2009) showed that protein L27, which is conserved in prokaryotes, extends far enough into the PTC to potentially contribute to catalysis. L27 is a small protein located at the base of the central protuberance of the 50S subunit. Its long, conserved N-terminal tail protrudes into the PTC, where it was reported to interact with both A-site and P-site tRNAs (Wower et al. 2000; Maguire et al. 2005; Voorhees et al. 2009; Jenner et al. 2010). Peptide-bond formation on the ribosome is for most aa-tRNAs rate-limited by their accommodation into the A site (Pape et al. 1998; Wohlgemuth et al. 2010; Johansson et al. 2011). Therefore, pre-steady state kinetic studies that aimed at determining the reaction mechanism often utilized aa-tRNA analogs, such as puromycin (Pmn) or its derivatives (e.g., C-Pmn [Katunin et al. 2002; Okuda et al. 2005; Brunelle et al. 2006; Wohlgemuth et al. 2006; Beringer and Rodnina 2007]). When Pmn is used as substrate, the peptidyl-transfer (PT) reaction shows a pronounced pH dependence with two ionizing groups, indicating that besides the amino group of the substrate (6.9 for Pmn [Katunin et al. 2002]), another ionizing group, with a pKa of ~7.5, contributes to catalysis (Katunin et al. 2002; Beringer and Rodnina 2007). Based on computer simulations of the pre- and post-PT state of the ribosome, this group has been proposed to be the N-terminal amine of L27 (Trobro and Åqvist 2008; Xiao and Wang 2012). The absence of L27 is thus predicted to alter the pH dependence of PT; however, the contribution of this
ionizable group to the PT reaction has not been experimentally tested so far.

Early biochemical data indicated a functional role for L27 in the PT activity. *Escherichia coli* strains in which L27 was deleted grew six times slower than the wild type (wt), and ribosomes lacking L27 were found to carry sub-stoichiometric amounts of L16, L21, and L20. The PT activity of ΔL27 ribosomes was slightly reduced, both in the presence of Pmn or native A-site substrate (Phe-tRNA^A^{Phe}) (Wower et al. 1998). Truncation of the N-terminal tail of L27 revealed that the absence of as few as the first three residues reduces the PT activity to the level of ΔL27 ribosomes (Maguire et al. 2005). smFRET data indicated that the main function of L27 might be to stabilize the tRNAs on the ribosome (Wang and Xiao 2012; Xiao and Wang 2012), which for some amino acids may contribute to the acceleration of the PT reaction. Furthermore, the α-amino group of L27 was recently suggested to take part in proton transfer during the PT reaction, assisting the 5′-phosphate oxygen of the A-site A76 in the deprotonation of the nucleophile in the transition state (Polikanov et al. 2014). Although L27 is not present in Archaea and Eukaryotes, another ribosomal protein, the archaean L10e and its eukaryotic homolog RPL10, extends a loop in the same position as the L27 tail (Armache et al. 2010); mutations in this conserved loop are lethal (Hofer et al. 2007). Validation of the role of L27 in the PT reaction for all or perhaps only certain incoming amino acids would, however, require detailed biochemical and kinetic studies. Here we use rapid kinetics methods to precisely quantify the contribution of L27 on the activity of the PTC.

**RESULTS AND DISCUSSION**

We purified ribosomes from the *E. coli* strain lacking the gene rpmA encoding L27 (IW312 strain; hereafter referred to as ΔL27) and from the same strain expressing L27 from a plasmid (IW312 + pPOTA1: rpmA; hereafter referred to as wt). Ribosome profiles obtained by sucrose-gradient centrifugation showed the absence of a pronounced 70S peak in the ΔL27 strain, indicating that most ribosomes are dissociated into the subunits at 5 mM Mg^{2+} (Fig. 1A). In accordance with earlier results of sucrose-gradient analysis (Wower et al. 1998), impaired subunit association might explain the slow growth phenotype of this strain. We thus collected fractions containing a mixture of 50S and 70S and confirmed the absence of L27 by quantitative mass spectrometry (Fig. 1B) and Western blotting (Fig. 1C). The active ribosome concentration was determined by preparing 70S initiation complexes in the presence of mRNA, initiation factors, and increasing concentrations of f^[(3)H]Met-tRNA^Met (Fig. 1D). More than 90% of the wt ribosomes formed stable complexes with fMet-tRNA^Met^Met, whereas ΔL27 ribosomes initiated to only ~40%, indicating that the preparation contains significant portion of free 50S subunits and/or incomplete and inactive 70S ribosomes.

Previous reports indicated that in the presence of low concentrations of Pmn, formation of fMet-Pmn is impaired on ribosomes lacking L27 (Maguire et al. 2005). Because Pmn concentrations used in those experiments (1 μM) were far below the K_M value for the ribosome–Pmn complex (~1–30 mM, depending on the tRNA in the P site) (Youngman et al. 2004; Beringer et al. 2005; Wohlgemuth et al. 2008), the observed decrease in the kinetics of the Pmn reaction might be due to a Pmn binding defect, rather than the reduced PT rate. To distinguish between the K_M and k_cat effects, we first reproduced the previous studies using fMet-tRNA^Met and Pmn as P- and A-site substrates. Indeed, the extent of fMet-Pmn formation with ΔL27 ribosomes appeared lower compared to the wt (Fig. 2A); however, when the amount of active ribosomes was taken into account, it turned out that all ribosomes active in initiation were also active in the PT reaction (Fig. 2A). To verify this result, we repeated the experiment with initiation complexes (ICs) purified through a sucrose cushion; as determined by nitrocellulose filtration, ∼40% of the IC formed with ΔL27 ribosomes dissociated upon purification (not shown), indicating that binding of fMet-tRNA^Met^Met was somewhat weaker on the mutant ribosomes. The active concentration of the remaining stable complexes was calculated from the amount of f^[(3)H]Met-tRNA^Met^Met.
Role of L27 in the PTC

FIGURE 2. Peptide-bond formation with Pmn as A-site substrate. Unpurified (A) or sucrose gradient-purified (B) ICs prepared with wt (open circles) or ΔL27 (closed circles) ribosomes (0.25 µM) were mixed with Pmn (1 µM), and the extent of dipeptide reaction was monitored over time. Solid lines represent the results of exponential fitting (see Materials and Methods). (C) Time courses of fMet-Pmn (left panel), fMetPhe-Pmn (middle panel), and fMetPro-Pmn (right panel) formation upon mixing of wt (open symbols) and ΔL27 (closed symbols) IC (50 nM) with high concentrations of Pmn (2.5–10 mM) at pH 6.5 (squares), 7.5 (triangles), and 8.5 (circles). Time courses were normalized for the extent of the reaction to facilitate visual inspection. Solid lines represent the results of exponential fitting of the time points. Rates of the fMet-Pmn reaction for wt and ΔL27 ribosomes, respectively, were 0.11 ± 0.1 and 0.13 ± 0.1 sec⁻¹ at pH 6.5; 1.6 ± 0.1 and 1.4 ± 0.1 sec⁻¹ at pH 7.5; and 7.8 ± 1.7 and 7.1 ± 1.9 sec⁻¹ at pH 8.5. Rates of the fMetPhe-Pmn for wt and ΔL27 ribosomes, respectively, were 1.0 ± 0.2 and 0.8 ± 0.1 sec⁻¹ at pH 6.5; 55 ± 6 and 30 ± 3 sec⁻¹ at pH 7.5; and 55 ± 10 and 49 ± 10 sec⁻¹ at pH 8.5. Rates of the fMetPro-Pmn reaction at pH 7.5 were 0.06 ± 0.01 and 0.02 ± 0.01 sec⁻¹ for wt and ΔL27 ribosome, respectively.

bound. These remaining ribosome complexes showed identical rates of peptide-bond formation (Fig. 2B), which shows that ΔL27 ribosomes do not have a PT defect.

The ribosome group presumably involved in the PT reaction with Pmn has not been identified. If the N-terminal α-amine of L27 takes part in catalysis, one would expect the pH dependence of the Pmn reaction to change in the absence of L27. To test this hypothesis, we compared the kinetics of fMet-Pmn formation at different pH values using ΔL27 and wt ribosomes. This time, we determined the maximal rate of peptide-bond formation at high concentrations of Pmn (Fig. 2C). As expected, the rate of fMet-Pmn formation depends on pH. However, at a given pH, the rates of the reaction were identical on the wt and ΔL27 ribosomes. This finding indicates that not only the PT activity at a defined pH, but also the overall rate/pH profile is not altered in the absence of L27, which further strengthens the notion that the protein itself is completely dispensable for this reaction. A similar result was obtained when we followed the Pmn reaction on post-translocation complexes carrying an fMetPhe-tRNA_Phe in the P site at different pH values (Fig. 2C). Also in this case the rate of fMetPhe-Pmn tripeptide formation was independent of the presence of L27. To check whether L27 might be more important for less reactive substrates such as proline (Wohlgemuth et al. 2008), which causes ribosomal stalling when more than two prolines have to be incorporated into the peptide (Doerfel et al. 2013; Ude et al. 2013), we checked the Pmn reaction with post-translocation complexes carrying a fMetPro-tRNA_Pro in the P site. In this case, the rate of peptide-bond formation was slightly decreased in the absence of L27 (0.02 sec⁻¹, compared with the rate of 0.06 sec⁻¹ obtained with wt ribosomes) (Fig. 2C).

Although L27 seems to be dispensable for PT with A-site tRNA analogs, it might still play a role in the presence of full-length aa-tRNAs. We thus determined the rate of fMet-Phe formation in the absence of L27. ICs were mixed with saturating concentrations of EF-Tu–GTP–Phe-tRNA_Phe ternary complex (TC-Phe) (Fig. 3A). The extent of dipeptide formation was lower on ΔL27 ribosomes (40%) compared with wt ribosomes (60%), as previously observed (Wower et al. 1998). However, the rate was very similar to the wt control (90 sec⁻¹ and 110 sec⁻¹ for ΔL27 and wt, respectively).

To investigate the origin of this small difference, we also measured the rate of dipeptide formation on ribosomes carrying an N-terminal truncated version of L27 (L27 Δ1-6). The dipeptide rate obtained with these ribosomes, 90 sec⁻¹, indicates that the small difference is due solely to the presence of the first six residues of L27 (Fig. 3A).

Apart from the proposed direct participation in the PT mechanism, the L27 tail might be involved in cognate

FIGURE 3. Reactions with natural substrates. (A) PT with cognate aa-tRNA was measured upon mixing wt (open circles), L27Δ1-6 (closed triangles), or ΔL27 (closed circles) ICs (50 nM) containing Met-tRNA in the P site and a UUC codon in the A site with TC-Phe (10 µM). Solid lines represent the results of exponential fitting of the time points, which yielded the rates of 110 ± 13, 90 ± 9, and 90 ± 8 sec⁻¹ for wt, L27Δ1-6, and ΔL27 ribosomes, respectively. (B) PT with near cognate aa-tRNA was measured as in A, using ICs with a CUC codon in the A site. Exponential fitting (solid lines) yielded the rates of 0.017 ± 0.002 and 0.024 ± 0.002 sec⁻¹ for wt and ΔL27 ribosomes, respectively. (C) Peptide release was measured upon mixing wt (open circles) or ΔL27 (closed circles) ICs (75 nM) programmed on a UAA codon with RF2 (1.5 µM). The rates, determined by exponential fitting (solid lines), were 6.6 ± 0.6 and 7.6 ± 0.8 sec⁻¹ for wt and ΔL27 ribosomes, respectively.
aa-tRNA selection. In fact, the crystal structure of the *T. thermophilus* ribosome in complex with a near-cognate ternary complex shows the N-terminal tail of L27 in a different conformation as compared to the cognate tRNA (Jenner et al. 2010). We reasoned that if L27 is involved in tRNA selection, PT to a near-cognate aa-tRNA would be affected by the absence of L27. To test this hypothesis, we reacted ribosomes carrying fMet-tRNA^{Met} in the P site and a CUC codon in the A site (coding for leucine) with an excess of the near cognate Phe-tRNA^{Phe-EF-Tu-GTP} (Fig. 3B). In this case, the extent and the rate of the PT reaction were essentially identical (rates of 0.014 and 0.019 sec^{-1} for wt and ΔL27 ribosomes, respectively). In summary, our data argue against any significant involvement of L27 in the mechanism of peptide-bond formation.

In addition to the PT reaction, the PTC catalyzes the hydrolysis of the peptidyl tRNA during translation termination. This prompted us to measure the impact of L27 in this reaction. We prepared pre-termination complexes with fMet-tRNA^{Met} in the P site and a UAA stop codon in the A site and followed the kinetics of peptide release in the presence of saturating concentrations of RF2. Also in this case, no difference was observed between the wt and ΔL27 ribosomes (Fig. 3C). The rate of peptide release (~7 sec^{-1}) was in agreement with that previously determined using wt (MRE600) ribosomes (Kuhlenkoetter et al. 2011; Indrisiunaite et al. 2015).

To investigate the importance of L27 for the steps of the elongation cycle that are not reflected in our model assays (e.g., translocation, tRNA competition) or for context-dependent effects on peptide-bond formation, we translated a full-length natural mRNA with ΔL27 ribosomes, using the mRNA coding for the *E. coli* CspA as a model (Rudorf et al. 2014). The time courses of translation (Fig. 4A) show that the mutant ribosomes translate the mRNA as efficiently as the wt ribosomes and without significant pausing, confirming that deletion of L27 does not have adverse effects on incorporation of amino acids other than Phe that was tested above. The average elongation rate per codon, as determined by fitting of the intensity change of the full-length product, was 1.3 aa sec^{-1} for the wt and 0.8 aa sec^{-1} for the ΔL27 ribosomes (Fig. 4B).

Taken together, our data strongly argue against an important contribution of L27 to the core mechanism of peptide-bond formation or peptidyl-tRNA hydrolysis at the PTC. Previously reported phenotypes of the mutant ribosomes can, in our opinion, be explained by the fact that the absence of L27 impairs the 50S subunit assembly and subunit association, thereby yielding a nonhomogeneous population of active and inactive ribosomes inside the cell. Indeed, the major phenotype we observed is an impaired 70S formation, which could explain the growth phenotype and the low yield of translation initiation. One small additional effect may be related to the stabilization of the substrates on the ribosome by the N-terminal tail of L27, as suggested by the subtle kinetic effects of L27 on fMetPhe formation and overall translation.

There are currently two main suggestions for the mechanism of ribosomal peptide-bond formation, the so-called proton shuttle (Trobro and Åqvist 2005) and the proton wire (Polikanov et al. 2014) models. Both models are consistent with the experimental data available so far, including the pH/rate profiles for different substrates, mutational analysis, and KIE and KSI measurements (Katunin et al. 2002; Youngman et al. 2004; Beringer and Rodnina 2007; Lang et al. 2008; Wohlgemuth et al. 2008; Hiller et al. 2011; Kuhlenkoetter et al. 2011; Zaher et al. 2011). Both models suggest that the proton from the attacking amine is transferred to the leaving group through the 2′-OH of the P-site tRNA A76; however, the impact of L27 and the exact path of proton transfer is different (we note that at a time when the proton shuttle model was suggested, the positions of the water molecules in the catalytic site were not known with precision). Although L27 does not play a role in the proton shuttle model, the wire model (Polikanov et al. 2014) suggests a contribution of the N-terminal amine of L27 in restricting the access of water molecules by closing the reaction pocket, thereby delaying the loss of a proton from a catalytic water molecule to the bulk solvent. The wire model would predict that the absence of the N terminus of L27 should alter both the rates of intermediate formation and the conversion of the intermediate into products, which is clearly not consistent with the present experiments. However, we cannot exclude that in the absence of L27, some other group (protein, rRNA, or water) takes over the functions of L27 (e.g., closes the reaction pocket). Alternatively, it is also possible that although the wire model is correct as to the path for proton transfer, L27 does not play the proposed
role even though it is located in the vicinity. Finally, both pathways—the shuttle and the wire—may turn out to be isoenergetic; in the absence of L27, the ribosome may favor the shuttle mechanism that does not involve L27.

MATERIALS AND METHODS

Strains, chemicals, and buffers

*E. coli* strains JW312 (ΔL27) and JW312 expressing the full-length L27 (wt) or its N-terminally truncated version (L27 Δ1-6) from the plasmid pPOT1AE were a kind gift from Robert Zimmermann (University of Massachusetts Amherst) (Maguire et al. 2005). Chemicals were from Roche Molecular Biochemicals, Sigma-Aldrich, or Merck. Radioactive compounds were from Hartmann Analytic. MNV mRNA (5′-GGCAAGGAGGUAAUAAUCAAGAUU-3′), in which the coding sequence is in italics and the A-site codon (underlined) is NNN (UUC coding for Phe, CUC coding for Leu, CCG coding for Pro, and UAA as the stop codon), was purchased from IBA. Buffer A: 50 mM Tris–HCl, 30 mM KCl, and 7 mM MgCl₂. Buffer B (HiFi): 50 mM Tris–HCl, pH 7.5, 70 mM NH₄Cl, 30 mM KCl, and 7 mM MgCl₂. Buffer B (HiFi): 50 mM Tris–HCl, pH 7.5, 70 mM NH₄Cl, 30 mM KCl, 3.5 mM MgCl₂, 0.5 mM spermidine, 8 mM putrescine, and 2 mM DTT. The pH of buffers and Pmn stocks was adjusted to 6.5, 7.5, or 8.5 at 37°C.

Ribosomes, initiation, and ternary complexes

For ribosome preparation, cells were grown in LB medium supplied with 50 µg/mL kanamycin, 1 mM IPTG, and, where appropriate, 50 µg/mL ampicillin, as described in Maguire et al. (2005). Ribosomes, initiation factors, EF-Tu, EF-G, tRNAs, and RF2 were prepared as previously described (Rodnina et al. 1995; Gromadski and Rodnina 2004; Kuhlenkoetter et al. 2011). Ternary complex EF-Tu–GTP–Phe-tRNAₔPhe was prepared by incubating EF-Tu (40 µM), GTP (1 mM), phosphoenol pyruvate (3 mM), pyruvate kinase (0.05 mg/mL), and [14C]Phe-tRNAₔPhe or [14C]Pro-tRNAₔPro (20 µM) in buffer A. Initiation complexes (ICs) were prepared by incubating ribosomes (1 µM) with mRNA (3 µM), initiation factors (1.5 µM each), f[3H]Met-tRNA₀Met (2 µM), and GTP (1 mM) in buffer A for 30 min at 37°C. Post-translocation complexes carrying fMetPhe-tRNAₔPhe or fMetPro-tRNAₔPro in the P site were formed by reacting ICs with a twofold excess of the suitable TC and catalytic amounts of EF-G for 5 min at 37°C. Ribosome complexes were purified by centrifugation through sucrose cushions (1.1 M sucrose in 1-6) from the shuttle and the wire—may turn out to be isoenergetic; in the absence of L27, the ribosome may favor the shuttle mechanism that does not involve L27.

Western blot

Twenty-five picomoles of MRE600, wt or ΔL27 70S ribosomes were digested for 1 h at 37°C with 0.1 µg/mL Benzonase RNase (Sigma-Aldrich) before loading on an 18% acrylamide SDS-PAGE gel. The presence of L27 was determined with the use of polyclonal antibodies against L27.

Mass spectrometry

One hundred picomoles of wt and ΔL27 ribosomes were proteolyzed with trypsin and analyzed by LC-ESI MS/MS as described (Davydov et al. 2013). Thermo RAW files were processed with MaxQuant (1.5.2.8) against a Uniprot *E. coli* (K12) database. 50S subunit proteins were quantified by intensity-based label-free quantification using MaxQuant Label Free Quantification (LFQ) values (Cox et al. 2014).

Kinetics experiments

All experiments were carried out at 37°C, if not stated otherwise. The formation of f[3H]Met-Pmn and of f[3H]Met[14C]Phe/Pro-Pmn was measured upon rapid mixing of equal volumes (14 µL) of ICs (50 nM; final concentrations are reported throughout the paper) and Pmn (2.5–10 mM) in a quench-flow apparatus (RQF-3, KinTek Corporation). Samples were quenched with 25% formic acid and f[3H]Met-Pmn was released upon addition of 500 µL of 1.5 M sodium acetate (pH 4.5) saturated with MgSO₄ and extracted in 750 µL ethyl acetate. Radioactivity in 500 µL of the organic phase was counted. Alternatively, samples were quenched with 0.5 M KOH, and f[3H]Met[14C]Phe/Pro-Pmn peptides were separated by reverse-phase HPLC (see below). For reactions with aa-tRNAs, ICs (50 nM) were mixed in the quench-flow with a large excess of the ternary complex EF-Tu–GTP–Phe-tRNAₔPhe (10 µM). Samples were quenched with 0.5 M KOH, hydrolyzed for 30 min at 37°C, and neutralized with one-tenth volume of glacial acetic acid. Dipeptides were analyzed by HPLC according to Katunin et al. (2002). Radioactivity in the eluate was counted after addition of Lumasafe Plus scintillation cocktail (PerkinElmer). Time courses were fitted to an exponential function, \( F = F_\infty + A \times \exp(-k_{app} \times t) \), with a time constant, \( k_{app} \) the amplitude of the signal change, \( A \), the final signal, \( F_\infty \), and the fluorescence signal at time \( t \), \( F \). For fitting of the time courses shown in Figure 2A,B, an additional exponential term was included. Calculations were performed using Prism (GraphPad software). The rates are reported ± SEM.

In vitro translation

Single-round translation of the full-length CspA mRNA (70 aa) was performed according to Doerfel et al. (2013) and Rudorf et al. (2014). wt and ΔL27 ICs were prepared as described above using Bodipy-FL-Met-tRNA₀Met. Translation was initiated by mixing IC (20 nM) with a translation mix containing EF-Tu (39 µM), total aa-tRNA (19.3 µM), EF-G (2 µM), GTP (1 mM), phosphoenol pyruvate (1.3 mM), and pyruvate kinase (0.1 mg/mL) at 37°C in buffer B. The translation product was separated on a 16.5% Tris–tricine–PAGE (Schägger and von Jagow 1987) and visualized by the fluorescent reporter at the N terminus of the peptides. The intensity of the full-length product was quantified with ImageJ. Average translation rates per codon were determined by fitting with a delay phase followed an exponential function (GraphPad software).

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REFERENCES

Armache J-P, Jarasch A, Anger AM, Villa E, Becker T, Blushan S, Jossinet F, Habecck M, Dindar G, Franckenberg S, et al. 2010. Localization of eukaryote-specific ribosomal proteins in a 5.5-Å cryo-EM map of the 80S eukaryotic ribosome. Proc Natl Acad Sci 107: 19754–19759.

Beringer M, Rodnina MV, 2007. Importance of tRNA interactions with the ribosome: studies with substrate analogs. Biol Chem 388: 687–691.

Beringer M, Bruehl C, Xiong L, Pfister P, Bieling P, Katunin VI, Mankin AS, Böttger EC, Rodnina MV. 2005. Essential mechanisms in the catalysis of peptide bond formation on the ribosome. J Biol Chem 280: 36065–36072.

Brunelle JL, Youngman EM, Sharma D, Green R. 2006. The interaction between C75 of tRNA and the a loop of the ribosome stimulates peptidyl transferase activity. RNA 12: 33–39.

Cox J, Hein MY, Luber CA, Paron I, Nagaraj N, Mann M. 2014. Accurate proteome-wide label-free quantification by delayed normalization and maximal peptide ratio extraction, termed MAXLFQ. Mol Cell Proteomics 13: 2513–2526.

Davydov II, Wohlgemuth I, Artamonova II, Urlaub H, Tonevitsky AG, Rodnina MV. 2013. Evolution of the protein stoichiometry in the L12 stalk of bacterial and eukaryotic ribosomes. Nat Commun 4: 1387.

Doerfler LK, Wohlgemuth I, Kothe C, Peske F, Urlaub H, Rodnina MV. 2013. EF-P is essential for rapid synthesis of proteins containing consecutive proline residues. Science 339: 85–88.

Gromadski KB, Rodnina MV. 2004. Kinetic determinants of high-fidelity tRNA discrimination on the ribosome. Mol Cell 13: 191–200.

Hiller DA, Singh V, Zhong M, Strobel SA. 2011. A two-step chemical mechanism for ribosome-catalysed peptide bond formation. Nature 476: 236–239.

Hofer A, Bussiere C, Johnson AW. 2007. Mutational analysis of the ribosomal protein rpl10 from yeast. J Biol Chem 282: 32630–32639.

Indrisiunaite G, Pavlov MY, Heurgué-Hamard V, Ehrenberg M. 2015. The pH dependence of class I RF-dependent termination of mRNA translation. J Mol Biol 427: 1848–1860.

Jenner L, Demeshkina N, Yusupova G, Yusupov M. 2010. Structural rearrangements of the ribosome at the tRNA proofreading step. Nat Struct Mol Biol 17: 1072–1078.

Johansson M, Ieong K-W, Trobro S, Strazewski P, Àqvist J, Pavlov MY, Ehrenberg M. 2011. pH-sensitivity of the ribosomal peptidyl transfer reaction dependent on the identity of the A-site aminoacyl-tRNA. Proc Natl Acad Sci 108: 79–84.

Katunin VI, Muth GW, Strobel SA, Wintermeyer W, Rodnina MV. 2002. Important contribution to catalysis of peptide bond formation by a single ionizing group within the ribosome. Mol Cell 10: 339–346.

Kuhlenkoepter S, Wintermeyer W, Rodnina MV. 2011. Different substrate-dependent transition states in the active site of the ribosome. Nature 476: 351–354.

Lang K, Erlacher M, Wilson DN, Micura R, Polacek N. 2008. The role of 23S ribosomal RNA residue A2451 in peptide bond synthesis revealed by atomic mutagenesis. Chem Biol 15: 485–492.

Maguire BA, Beniaminov AD, Ramu H, Mankin AS, Zimmermann RA. 2005. A protein component at the heart of an RNA machine: The importance of protein L27 for the function of the bacterial ribosome. Mol Cell 20: 427–435.

Okuda K, Sella AC, Strobel SA. 2005. Uncovering the enzymatic pKa of the ribosomal peptidyl transferase reaction utilizing a fluorinated puromycin derivative. Biochemistry 44: 6675–6684.

Pape T, Wintermeyer W, Rodnina MV. 1998. Complete kinetic mechanism of elongation factor Tu-dependent binding of aminoacyl-tRNA to the A site of the E. coli ribosome. EMBO J 17: 7490–7497.

Polikanov YS, Steitz TA, Innis CA. 2014. A proton wire to couple aminoacyl-tRNA accommodation and peptide-bond formation on the ribosome. Nat Struct Mol Biol 21: 787–793.

Rodnina M, Fricke R, Kuhn L, Wintermeyer W. 1995. Codon-dependent conformational change of elongation factor Tu preceding GTP hydrolysis on the ribosome. EMBO J 14: 2613–2619.

Rudorf S, Thommen M, Rodnina MV, Lipowsky R. 2014. Deducing the kinetics of protein synthesis in vivo from the transition rates measured in vitro. PLoS Comput Biol 10: e1003909.

Schägger H, von Jagow G. 1987. Tricine-sodium dodecyl sulfate-polyacrylamide gel electrophoresis for the separation of proteins in the range from 1 to 100 kDa. Anal Biochem 166: 368–379.

Selmer M, Dunham CM, Murphy FV IV, Weixdauber A, Petry S, Kelley AC, Weir JR, Ramakrishnan V. 2006. Structure of the 70S ribosome complexed with mRNA and tRNA. Science 313: 1935–1942.

Trobro S, Àqvist J. 2008. Role of ribosomal protein L27 in peptidyl transfer. Biochemistry 47: 8988–9006.

Ude S, Lassak I, Starosta AL, Krazenberger T, Wilson DN, Jung K. 2013. Translation elongation factor EF-P alleviates ribosome stalling at polyproline stretches. Science 339: 82–85.

Voorhees RM, Weixdauber A, Loakes D, Kelley AC, Ramakrishnan V. 2009. Insights into substrate stabilization from snapshots of the peptidyl transferase center of the intact 70S ribosome. Nat Struct Mol Biol 16: 528–533.

Wang Y, Xiao M. 2012. Role of the ribosomal protein L27 revealed by single-molecule FRET study. Protein Sci 21: 1696–1704.

Wohlgemuth I, Beringer M, Rodnina MV. 2006. Rapid peptide bond formation on isolated 50S ribosomal subunits. EMBO Rep 7: 699–703.

Wohlgemuth I, Brenner S, Beringer M, Rodnina MV. 2008. Modulation of the rate of peptidyl transfer on the ribosome by the nature of substrates. J Biol Chem 283: 32229–32335.

Wohlgemuth I, Pohl C, Rodnina MV. 2010. Optimization of speed and accuracy of decoding in translation. EMBO J 29: 3701–3709.

Wower JK, Wower J, Zimmermann RA. 1998. Ribosomal protein L27 participates in both 50S subunit assembly and the peptidyl transferase reaction. J Biol Chem 273: 19647–19652.

Wower J, Kirillov SV, Wower IK, Guven S, Hixson SS, Zimmermann RA. 2000. Transit of tRNA through the ribosome complexed with mRNA and tRNA. J Mol Biol 297: 737887–73894.

Xiao M, Wang Y. 2012. L27-tRNA interaction revealed by mutagenesis and pH titration. Biophys Chem 167: 8–15.

Youngman EM, Brunelle JL, Kochaniak AB, Green R. 2004. The active site of the ribosome is composed of two layers of conserved nucleotides with distinct roles in peptide bond formation and peptide release. Cell 117: 589–599.

Zaher HS, Shaw JJ, Strobel SA, Green R. 2011. The 2′OH group of the peptidyl-tRNA stabilizes an active conformation of the ribosomal PTC. EMBO J 6: 2445–2453.