An engineered non-oxidative glycolytic bypass based on Calvin-cycle enzymes enables anaerobic co-fermentation of glucose and sorbitol by *Saccharomyces cerevisiae*

Aafke C. A. van Aalst, Robert Mans and Jack T. Pronk*

**Abstract**

**Background:** *Saccharomyces cerevisiae* is intensively used for industrial ethanol production. Its native fermentation pathway enables a maximum product yield of 2 mol of ethanol per mole of glucose. Based on conservation laws, supply of additional electrons could support even higher ethanol yields. However, this option is disallowed by the configuration of the native yeast metabolic network. To explore metabolic engineering strategies for eliminating this constraint, we studied alcoholic fermentation of sorbitol. Sorbitol cannot be fermented anaerobically by *S. cerevisiae* because its oxidation to pyruvate via glycolysis yields one more NADH than conversion of glucose. To enable re-oxidation of this additional NADH by alcoholic fermentation, sorbitol metabolism was studied in *S. cerevisiae* strains that functionally express heterologous genes for ribulose-1,5-bisphosphate carboxylase (RuBisCO) and phosphoribulokinase (PRK). Together with the yeast non-oxidative pentose-phosphate pathway, these Calvin-cycle enzymes enable a bypass of the oxidative reaction in yeast glycolysis.

**Results:** Consistent with earlier reports, overproduction of the native sorbitol transporter Hxt15 and the NAD$^+$-dependent sorbitol dehydrogenase Sor2 enabled aerobic, but not anaerobic growth of *S. cerevisiae* on sorbitol. In anaerobic, slow-growing chemostat cultures on glucose–sorbitol mixtures, functional expression of PRK-RuBisCO pathway genes enabled a 12-fold higher rate of sorbitol co-consumption than observed in a sorbitol-consuming reference strain. Consistent with the high $K_m$ for CO$_2$ of the bacterial RuBisCO that was introduced in the engineered yeast strains, sorbitol consumption and increased ethanol formation depended on enrichment of the inlet gas with CO$_2$. Prolonged chemostat cultivation on glucose–sorbitol mixtures led to loss of sorbitol co-fermentation. Whole-genome resequencing after prolonged cultivation suggested a trade-off between glucose-utilization and efficient fermentation of sorbitol via the PRK-RuBisCO pathway.

**Conclusions:** Combination of the native sorbitol assimilation pathway of *S. cerevisiae* and an engineered PRK-RuBisCO pathway enabled RuBisCO-dependent, anaerobic co-fermentation of sorbitol and glucose. This study demonstrates the potential for increasing the flexibility of redox cofactor metabolism in anaerobic *S. cerevisiae* cultures and, thereby, to extend substrate range and improve product yields in anaerobic yeast-based processes by enabling entry of additional electrons.

**Keywords:** Ethanol, NADH, Redox engineering, Sorbitol, Yeast, CO$_2$, Fermentation, Electrons
Background

With an estimated global output of 103 billion litres in 2021 [1], fuel ethanol produced from plant carbohydrates with the yeast *Saccharomycyes cerevisiae* remains the largest process in microbial biotechnology based on product volume. Yeast-based ethanol production is predominantly performed in the USA and Brazil, using corn starch and cane sugar, respectively, as feedstocks [1, 2]. Since the carbohydrate feedstock can contribute up to 70% to the overall process costs of industrial ethanol production, optimization of the yield of ethanol on carbohydrates is of paramount importance for process economics [3, 4].

Hydrolysis of corn starch yields glucose as fermentable sugar, while sucrose, the predominant sugar in sugar cane, is hydrolysed to glucose and fructose by yeast invertase [5, 6]. In *S. cerevisiae*, conversion of these hexoses to ethanol and carbon dioxide occurs via the Embden–Meyerhof glycolysis and the fermentation enzymes pyruvate decarboxylase and alcohol dehydrogenase. By producing two moles of ethanol per mole of hexose, this pathway conserves the entire degree of reduction of the substrate in ethanol and, thereby, reaches the theoretical maximum yield of ethanol on hexose sugars [7]. In practice, this theoretical maximum is approached at near-zero growth rates in anaerobic retentostat cultures, in which the impact of yeast biomass formation on carbon and redox metabolism is negligible [8].

To achieve ethanol yields above 2 mol per mole hexose, additional electrons would have to be fed into alcoholic fermentation, for example in the form of NADH. However, the configuration of the metabolic network of wild-type *S. cerevisiae* precludes this option. This constraint is illustrated by experiments in which formate was co-fed to anaerobic, glucose-limited cultures of *S. cerevisiae* strains overproducing the native NAD⁺-dependent formate dehydrogenase Fdh1. In these cultures, the additional electrons provided by formate were channelled into the glycolytic intermediate dihydroxyacetone phosphate to glycerol-3-phosphate by NAD⁺-dependent glycerol-3-phosphate dehydrogenase (Gpd1 or Gpd2). This redox reaction is followed by dephosphorylation of glycerol-3-phosphate by glycerol-3-phosphate phosphatase (Gpp1 or Gpp2) [10, 11]. In anaerobic cultures of wild-type *S. cerevisiae* strains, glycerol formation is essential for re-oxidation of ‘surplus’ NADH generated in biosynthetic reactions and has an economically significant negative impact on ethanol yields in industrial processes [12].

The rigidity of the *S. cerevisiae* metabolic network that prevents use of formate-derived NADH for alcoholic fermentation and necessitates glycerol production anaerobic for redox balancing, also prevents anaerobic fermentation of polyols such as mannitol and sorbitol. Mannitol is a main component of brown seaweed, which is investigated as a potential feedstock for ethanol production [13]. Sorbitol occurs in flowering plants [14] and is industrially produced by catalytic hydrogenation of glucose [15]. Although *S. cerevisiae* genomes harbour structural genes for polyol transporters and dehydrogenases, aerobic growth on mannitol and sorbitol typically requires prolonged adaptation [16, 17]. Instantaneous aerobic growth is observed upon combined overexpression of either of the native hexose-transporter genes *HXT13, HXT15* or *HXT17* and a native gene encoding mannitol dehydrogenase (*MAN1* or *MAN2*) or sorbitol dehydrogenase (*SOR1* or *SOR2*) [18, 19]. Since these polyol dehydrogenases are NAD⁺-dependent, conversion of mannitol or sorbitol to pyruvate yields one more NADH than glucose upon conversion to pyruvate via the glycolytic pathway. Use of polyols as (co-)substrates therefore provides an interesting model to explore metabolic engineering strategies for feeding additional electrons into yeast-based ethanol production. Such additional electrons could alternatively be provided by, for example, co-feeding of electrochemically produced formate [20, 21] or cathode-associated electrobiotechnology [22, 23].

Our group explored expression of heterologous genes encoding the Calvin-cycle enzymes ribulose-5-phosphate kinase (PRK) and ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO) in yeast to re-route re-oxidation of ‘surplus’ NADH from glycerol formation to ethanol formation [24]. This metabolic engineering strategy encompasses a bypass of the NADH-yielding glyceraldehyde-3-phosphate-dehydrogenase reaction in glycolysis, involving the native non-oxidative pentose-phosphate pathway, PRK and RuBisCO. This bypass allows for redox-neutral synthesis of 3-phosphoglycerate from glucose and CO₂. Subsequent conversion of 3-phosphoglycerate to ethanol via the regular yeast pathway for alcoholic fermentation then enables re-oxidation of NADH. Implementation of this strategy in engineered strains led to strongly reduced glycerol yields and correspondingly increased ethanol yields on sugar [24]. Strains were further optimized by combined overexpression of non-oxidative pentose-phosphate pathway enzymes [25] to increase supply of ribulose-5-phosphate and deleting the structural gene encoding for the Gpd2 isoenzyme of glycerol-3-phosphate dehydrogenase. This approach yielded *S. cerevisiae* strains with an over 10% higher ethanol yield on glucose in anaerobic batch cultures, while showing the same rates of growth and ethanol production as a non-engineered parental strain [26].
The goal of the present study was to explore whether introduction of a functional PRK-RuBisCO bypass can accommodate the NADH generated upon the entry of sorbitol into glycolysis and, thereby, enable anaerobic (co−)fermentation of this polyol. To this end, Cas9-mediated genome editing was used to construct S. cerevisiae strains containing overexpression cassettes for HXT15 and SOR2 with or without a simultaneously introduced PRK-RuBisCO bypass. Anaerobic growth and product formation of the resulting engineered strains were quantitatively analysed in anaerobic mixed-substrate batch and chemostat cultures on glucose and sorbitol.

Results

Theoretical analysis of glucose or sorbitol fermentation by wild-type and engineered S. cerevisiae

Introduction of a PRK-RuBisCO-based ‘bypass’ of the oxidative reaction in glycolysis, as previously applied for improving ethanol yields of anaerobic glucose-grown cultures [24, 26], could theoretically enable redox-neutral fermentation of sorbitol (Fig. 1). Simultaneous operation of the native yeast glycolytic pathway and this bypass should then be redox-cofactor balanced according to the following ‘redox half reactions’:

\[
3.5 \text{sorbitol} \rightarrow 7 \text{ethanol} + 7 \text{CO}_2 + 3.5 \text{NADH} + 7 \text{ATP} \text{via native yeast glycolysis}
\]

\[
2.5 \text{sorbitol} + 3.5 \text{NADH} \rightarrow 6 \text{ethanol} + 3 \text{CO}_2 \text{via PRK-RuBisCO bypass}
\]

The combined reaction would then provide a redox-balanced, net ATP-generating pathway for anaerobic fermentation of sorbitol (Fig. 1) or, by analogy, mannitol:

![Fig. 1](image-url)
6 sorbitol → 13 ethanol + 10 CO2 + 7 ATP combined

Functional expression of this pathway in a host organism could, in the absence of growth, support a theoretical maximum yield of \(13.6 = 2.17\) mol of ethanol per mole sorbitol, which is 8.5% higher than the theoretical yield of ethanol on glucose. Compared to alcoholic fermentation of glucose, this pathway for sorbitol fermentation would yield 42% less ATP per mole of substrate. Provided that sufficient rates of alcoholic fermentation can be achieved to maintain industrially relevant productivities, a low ATP yield on sorbitol could be interesting as it should divert carbon substrate from biomass formation to ethanol production [27, 28].

To further assess the predicted impact of the proposed metabolic engineering strategy, it was implemented in a stoichiometric model of the core metabolic network of \(S.\) \(cerevisiae\) [27, 29]. The model was then used to calculate biomass and ethanol yields at different specific growth rates. Calculations were based on the assumption that NADH from biomass formation [24, 26], as well as NADH from the reaction catalysed by sorbitol dehydrogenase (Fig. 1), was exclusively re-oxidized by ethanol formation via the PRK-RuBisCO route. Consistent with the calculations presented above, sorbitol fermentation via the engineered pathway was predicted to result in a theoretical maximum yield of 2.17 mol ethanol (mol sorbitol)\(^{-1}\) (Fig. 2, Additional file 5: Table S1). Up to a specific growth rate of 0.1 h\(^{-1}\), the predicted molar yield of ethanol on sorbitol remained above the theoretical maximum yield on glucose (Fig. 2, Additional file 5: Table S1). As anticipated based on the lower ATP yield from sorbitol fermentation, predicted biomass yields on this substrate (g biomass (mol sorbitol)\(^{-1}\)) were 42% lower at all specific growth rates than corresponding biomass yields in glucose-grown cultures. At the same specific growth rate, the required biomass-specific rate of sorbitol fermentation was therefore predicted to be 71% higher than in glucose-grown anaerobic cultures.

**Characterization of \(S.\) \(cerevisiae\) strains overexpressing HXT15 and SOR2**

Consistent with results from an earlier study [19], aerobic batch cultures of \(S.\) \(cerevisiae\) IME611, which carried overexpression cassettes for \(HXT15\) and \(SOR2\), grew on synthetic medium (SM) with sorbitol as sole carbon source at a specific growth rate of 0.23 h\(^{-1}\). Under the same conditions, cultures of the congenic reference strain IME324 showed virtually no growth (Additional file 5: Table S2). Strain IMX2506, in which overexpression of \(HXT15\) and \(SOR2\) was combined with deletion of \(GPD2\) and introduction of a PRK-RuBisCO pathway optimized for reduced glycerol production in anaerobic conditions.
glucose-grown batch cultures [26], showed a similar growth rate on sorbitol (0.25 h⁻¹). Specific growth rates of these strains on sorbitol were approximately 33% lower than on glucose (0.36 h⁻¹ and 0.35 h⁻¹, respectively; Table S2). However, despite the fast aerobic growth of strain IMX2506 on sorbitol, no growth was observed after up to 50 days of anaerobic incubation in SM with sorbitol as sole carbon source.

**Co-utilization of glucose and sorbitol in anaerobic batch cultures**

The inability of *S. cerevisiae* IMX2506 to grow anaerobically on sorbitol as sole carbon source suggested that the in vivo capacity of Hxt15, Sor2 and/or the engineered PRK-RuBisCO bypass was too low to sustain the rate of ATP production required for cellular maintenance. Such a scenario might still allow for anaerobic co-consumption.

**Fig. 3** Growth, glucose consumption, sorbitol consumption, ethanol formation and glycerol formation in anaerobic bioreactor batch cultures of *S. cerevisiae* strains IME324 (reference strain) (A), IME611 (overexpression cassettes for HXT15 and SOR2) (B), IMX1489 (optimized PRK-RuBisCO bypass and gpd2Δ mutation, [26]) (C) and IMX2506 (optimized PRK-RuBisCO bypass and gpd2Δ mutation, overexpression cassettes for HXT15 and SOR2) (D). Cultures were grown anaerobically at pH 5 and at 30 °C on synthetic medium containing 20 g L⁻¹ glucose and 30 g L⁻¹ sorbitol as carbon sources.
of sorbitol and glucose. We therefore investigated growth in anaerobic bioreactor batch cultures on a mixture of 20 g L\(^{-1}\) glucose and 30 g L\(^{-1}\) sorbitol (Fig. 3, Table 1). No co-consumption of sorbitol was observed in mixed-substrate cultures of strains IME324 (reference) and IMX1489, which expressed an optimized PRK-RuBisCO bypass and carried a gpd2Δ mutation (Fig. 3, Table 1). In contrast, upon reaching stationary phase, strain IME611, which carried overexpression cassettes for HXT15 and SOR2 had consumed 1.5 g L\(^{-1}\) (4.3 mmol g\(_x\)\(^{-1}\); subscript \(x\) denotes biomass) sorbitol. Co-consumption coincided with a higher glycerol production (16.3 mmol g\(_x\)\(^{-1}\)) than observed in strains IME324 and IMX1489 (11.8 mmol g\(_x\)\(^{-1}\) and 3.1 mmol g\(_x\)\(^{-1}\), respectively, Table 1). This observation indicated that, in strain IME611, the surplus NADH generated during sorbitol co-fermentation (Fig. 1) was predominantly re-oxidized by glycerol formation. In contrast, when strain IMX2506, which combined the genetic modifications carried by strains IME611 and IMX1489, reached stationary phase, consumption of 2.3 g L\(^{-1}\) sorbitol (5.4 mmol g\(_x\)\(^{-1}\)) was accompanied by production of only 3.5 mmol g\(_x\)\(^{-1}\) glycerol. In addition, co-consumption of sorbitol by strain IMX2506 coincided with a higher apparent ethanol yield on glucose than observed for the three other strains (Table 1). These results indicated that overexpression of HXT15 and SOR2 in a strain with an active PRK-RuBisCO pathway enabled a modest co-fermentation of glucose and sorbitol in anaerobic batch cultures.

### Table 1: Yields of biomass and ethanol on glucose, sorbitol consumption, stoichiometric relationships between glycerol production and biomass formation and specific growth rates in anaerobic bioreactor batch cultures of S. cerevisiae strains IME324 (reference strain), IME611 (overexpression cassettes for HXT15 and SOR2), IMX1489 (optimized PRK-RuBisCO bypass and gpd2Δ mutation, [26]) and IMX2506 (optimized PRK-RuBisCO bypass and gpd2Δ mutation, overexpression cassettes for HXT15 and SOR2)

| Strain                  | IME324 | IME611 | IMX1489 | IMX2506 |
|-------------------------|--------|--------|---------|---------|
| HXT15 and SOR2 cassettes| no     | yes    | no      | yes     |
| PRK-RuBisCO bypass and gpd2Δ | no     | no     | yes     | yes     |
| Specific growth rate (h\(^{-1}\)) | 0.31 ± 0.01 | 0.31 ± 0.00 | 0.30 ± 0.00 | 0.27 ± 0.00 |
| Biomass yield on glucose (g mol\(^{-1}\)) | 16.2 ± 0.3 | 16.0 ± 0.2 | 16.3 ± 0.9 | 17.4 ± 0.5 |
| Ethanol yield on glucose (mol mol\(^{-1}\)) | 1.47 ± 0.02 | 1.51 ± 0.01 | 1.64 ± 0.04 | 1.80 ± 0.05 |
| Ethanol yield on substrates (mol mol\(^{-1}\)) | 1.47 ± 0.02 | 1.42 ± 0.01 | 1.64 ± 0.04 | 1.65 ± 0.05 |
| Glycerol produced (mmol (g\(_x\)\(^{-1}\))) | 11.8 ± 0.2 | 16.3 ± 0.2 | 3.1 ± 0.1 | 3.5 ± 0.0 |
| Sorbitol consumed (mmol (g\(_x\)\(^{-1}\))) | < 0.3 | 4.3 ± 0.1 | < 0.3 | 5.4 ± 0.2 |
| Degree of reduction recovery (%) | 100-102 | 100-101 | 100-104 | 95-99 |

Cultures were grown on 20 g L\(^{-1}\) glucose and 30 g L\(^{-1}\) sorbitol. 'Substrates' refers to the combination of glucose and sorbitol. Specific growth rates and stoichiometries were calculated from at least 7 sampling points in the exponential growth phase. Values represent averages ± mean deviations of measurements on independent duplicate cultures for each strain. Degree of reduction balances yielded electron recoveries between 95 and 104%.

* These yield values on glucose also include ethanol and biomass formed by the additional consumption of sorbitol.

### Co-fermentation of sorbitol by anaerobic mixed-substrate chemostat cultures

In anaerobic mixed-substrate batch cultures of strain IMX2506, sorbitol consumption predominantly occurred when the supplied glucose was already nearly consumed (Fig. 3, panel D). Based on this observation, we hypothesized that glucose and sorbitol competed for Hxt transport proteins. Sorbitol co-consumption at low glucose concentrations was investigated in anaerobic chemostat cultures. In chemostat cultures, culture broth is removed at a fixed flow rate (\(F_{\text{out}}\)) while the culture volume (\(V_L\)) is kept constant by continuous supply of fresh medium, thus controlling the specific growth rate [30, 31]. The anaerobic chemostat cultures were grown on a mixture of 10 g L\(^{-1}\) glucose and 10 g L\(^{-1}\) sorbitol at a dilution rate (\(D_{\text{out}}/V_L\)) which, in steady-state cultures, equals specific growth rate of 0.025 h\(^{-1}\). In these chemostat cultures, residual glucose concentrations were below 0.036 g L\(^{-1}\). In chemostat cultures of strain IMX2506, anaerobic sorbitol conversion rates were 12-fold higher than in cultures of strain IME611, which carried overexpression cassettes for HXT15 and SOR2 but did not harbour a PRK-RuBisCO bypass (Table 2). In cultures of the congenic reference strain IME324, which carried no PRK-RuBisCO bypass or HXT15 and SOR15 overexpression cassettes (Table 2), glucose was virtually completely consumed but sorbitol concentrations in the chemostat cultures equalled those
This observation reflected the reference strain’s inability to anaerobically consume sorbitol. However, the PRK-RuBisCO-expressing reference strain IMX1489, which did not contain overexpression cassettes for $HXT15$ and $SOR15$, did co-consume sorbitol (Table 2, Fig. 4B).

### Table 2

Yields of biomass and ethanol on glucose, sorbitol consumption, biomass-specific sorbitol uptake rates and stoichiometric relationships between glycerol production and biomass formation in anaerobic bioreactor chemostat cultures of $S.\, cerevisiae$ strains IME324 (reference strain), IME611 (overexpression cassettes for $HXT15$ and $SOR2$), IMX1489 (optimized PRK-RuBisCO bypass and gpd$\Delta$ mutation, [26]) and IMX2506 (optimized PRK-RuBisCO bypass and gpd$\Delta$ mutation, overexpression cassettes for $HXT15$ and $SOR2$). ‘Substrates’ refers to the combination of glucose and sorbitol. Cultures were grown at a dilution rate of 0.025 h$^{-1}$ on 10 g L$^{-1}$ of glucose and 10 g L$^{-1}$ of sorbitol (pH 5).

| Strain | IME324 | IME611 | IMX1489 | IMX2506 |
|--------|--------|--------|---------|---------|
| HXT15 and SOR2 cassettes | no | yes | no | yes |
| PRK-RuBisCO bypass and gpd$\Delta$ | no | no | yes | yes |
| Biomass yield on substrates (g$_{\text{X}}$ mol$_{-1}^{-1}$) | 14.2 ± 0.1 | 14.3 ± 0.7 | 12.0 ± 0.9 | 13.3 ± 0.4 |
| Yield of ethanol on glucose (mol mol$^{-1}$)$^a$ | 1.62 ± 0.03 | 1.66 ± 0.07 | 2.96 ± 0.00 | 2.86 ± 0.02 |
| Ethanol yield on substrates (mol mol$^{-1}$) | 1.63 ± 0.04 | 1.61 ± 0.06 | 1.76 ± 0.01 | 1.83 ± 0.04 |
| Glycerol/biomass (mmol g$_{\text{X}}$)$^{-1}$ | 8.1 ± 0.2 | 10.8 ± 0.5 | 0.6 ± 0.2 | 1.0 ± 0.0 |
| Sorbitol consumed (mmol g$_{\text{X}}$)$^{-1}$ | < 0.5 | 2.2 ± 1.1 | 33.9 ± 3.0 | 27.0 ± 0.1 |
| Biomass-specific sorbitol uptake rate (mmol g$_{\text{X}}$ h$^{-1}$)$^{-1}$ | < 0.02 | 0.06 ± 0.03 | 0.89 ± 0.08 | 0.67 ± 0.00 |
| Degree of reduction recovery (%) | 99–100 | 98–102 | 98–99 | 100–101 |

Values represent averages ± mean deviations of measurements on independent steady-state triplicate cultures of strain IME611 and duplicate cultures of strain IMX2506.

$^a$ These apparent yield values on glucose also include ethanol and biomass formed by the additional consumption of sorbitol.

Fig. 4: Sorbitol consumption by anaerobic chemostat cultures of $S.\, cerevisiae$ strains IMX2506 (optimized PRK-RuBisCO bypass and gpd$\Delta$ mutation, overexpression cassettes for $HXT15$ and $SOR2$) (A) and IMX1489 (optimized PRK-RuBisCO bypass and gpd$\Delta$ mutation [26]) (B). Chemostat cultures were grown at a dilution rate of 0.025 h$^{-1}$ on 10 g L$^{-1}$ of glucose and 10 g L$^{-1}$ of sorbitol. A Average residual sorbitol concentration ± standard deviation in four chemostat cultures of strain IMX2506. After 400 h, the CO$_2$ content of the inlet gas was reduced to zero in two of the four cultures. The dotted line represents expected wash-in kinetics of sorbitol in the absence of sorbitol consumption: $c(t) = c_0 - (c_0 - c_{400})e^{-D\cdot t}$, with $c$ = residual sorbitol concentration, $c_{400}$ = sorbitol concentration at 400 h, $c_0$ = sorbitol concentration in medium feed and $D$ = dilution rate. B Average sorbitol concentration ± standard deviation in two chemostats of strain IMX1489. CO$_2$ supplementation was stopped at 700 h.
10% CO₂ was switched to pure N₂ during fermentation runs. This switch led to an instantaneous, progressive increase of the residual sorbitol concentration, with a profile that closely corresponded to wash-in kinetics in the complete absence of sorbitol consumption (Fig. 4). These results, in combination with the absence of sorbitol consumption in strain IME611 lacking the PRK-RuBisCO pathway (Table 2), confirmed involvement of in vivo RuBisCO activity in anaerobic sorbitol co-fermentation.

**Prolonged continuous cultivation on a glucose–sorbitol mixture does not select for improved sorbitol fermentation**

During anaerobic chemostat cultivation of strain IMX2506 on sorbitol and glucose, sorbitol co-consumption increased during the first 10 volume changes. After this point, it stabilized for approximately 3 volume changes (Fig. 4A), leaving ~ 4 g L⁻¹ of sorbitol unused. Based on the assumption that improved co-consumption of sorbitol by spontaneous mutants would confer a selective advantage, two new anaerobic continuous cultures were grown on a glucose and sorbitol mixture for over 80 generations. During the first approximately 40 generations, sorbitol consumption by the mixed-substrate chemostat cultures improved. However, contrary to expectation, sorbitol co-consumption deteriorated rather than improved further in both evolution experiments. In evolution line 1, sorbitol co-consumption completely ceased, while in evolution line 2 its rate declined by approximately 3.5-fold (Fig. 5A). In both prolonged continuous cultures, glycerol production increased, indicating a decline of the in vivo activity of the PRK-RuBisCO pathway (Fig. 5B).

To explore underlying mechanisms for the reduced sorbitol co-consumption observed during prolonged mixed-substrate cultivation, whole-genome sequencing was performed on culture samples. After 86 generations, sequence data from both chemostat cultures showed an identical non-synonymous point mutation in the open-reading frame of the spinach prk sequence, which caused an alanine-to-aspartate change at position 193 in the Prk protein (A193D). In evolution line 1, in which sorbitol co-consumption was virtually completely abolished, 73% of the prk sequence reads carried the mutation. A lower percentage (45%) of reads carrying this mutation was identified in evolution line 2, in which some sorbitol co-consumption was still observed after 86 generations. Sequence alignment showed that the mutation in prk involves a highly conserved amino acid residue in type II PRK proteins from 49 different species (Archaea, Eukaryotes and Cyanobacteria; position 372 in the multiple sequence alignment as identified in Additional file 5: Fig. S6 of the publication by Gurrieri et al. [33]). In these dimeric type II PRK enzymes (including the spinach PRK expressed in this study), an alanine residue is invariably found at this position, while the bacterial allosterically regulated and octameric type I PRK enzymes, either have an alanine or a threonine in this position. For both PRK types, this residue is located on the edge of a β-sheet [33, 34].

![Fig. 5 Sorbitol consumption (A) and glycerol production (B) in duplicate anaerobic chemostat cultures of strain IMX2506 (optimized PRK-RuBisCO bypass and gpd2Δ mutation, overexpression cassettes for HXT15 and SOR2). For the first 20 generations, the cultures were grown on a mixture of 10 g L⁻¹ sorbitol and 10 g L⁻¹ glucose at a dilution rate of 0.05 h⁻¹. After 20 h, the dilution rate was decreased to 0.025 h⁻¹ and after 50 generations the substrate mixture was changed to 20 g L⁻¹ sorbitol and 10 g L⁻¹ glucose.](image-url)
hypothesize that exchange of a neutral residue (alanine/threonine) for a negatively charged residue (aspartate) in a highly conserved position disrupted protein folding and thereby reduced or even abolished PRK activity.

No sequence changes were found in other coding regions in the analysed culture samples where sorbitol consumption was greatly reduced after prolonged cultivation. However, sequence data from both evolution experiments displayed segmental aneuploidy in multiple chromosomes (Additional file 5: Fig. S1, Table S3). Some amplifications differed between the two experiments or, based on read coverages, only occurred in part of the population. Since the duplicated regions carried multiple genes, no definitive interpretation is possible without extensive reverse engineering studies [35, 36]. However, we note that ca. 100 kb duplication of a fragment of chromosome IV (~1,075,000–1,175,000) that was found in both evolution experiments carried the HXT6 and HXT7 genes. These genes, which encode the major high-affinity glucose transporters of S. cerevisiae, are highly expressed in glucose-limited chemostat cultures [37]. A large duplication of a centromeric region of chromosome XV (~0–495,000) carried the GPD2 locus into which the overexpression cassettes for six genes encoding non-oxidative pentose-phosphate-pathway enzymes had been integrated. In addition, this region harboured HXT11 which, like HXT15, encodes a functional sorbitol transporter [19].

**Discussion**

Introduction of functional Calvin-cycle enzymes in S. cerevisiae was previously shown to enable coupled re-oxidation of ‘surplus’ NADH, generated during biomass formation in anaerobic cultures, to ethanol formation, thereby reducing glycerol production and enhancing ethanol yield on sugars [24, 26, 38, 39]. The present study demonstrates that expressions of a non-oxidative, PRK-RuBisCO bypass of glycolysis enabled a 12-fold higher rate of sorbitol conversion in anaerobic chemostat cultures grown on glucose–sorbitol mixtures than observed in cultures of a reference S. cerevisiae strain.

As reported for aerobic growth of S. cerevisiae on sorbitol [19], overexpression of HXT15 and SOR2 was required to achieve a modest co-utilization of sorbitol by anaerobic batch cultures of S. cerevisiae grown on glucose–sorbitol mixtures (Fig. 3). In contrast, constitutive overexpression of these genes was not required for a much more extensive sorbitol co-consumption by slow-growing, glucose-limited anaerobic chemostat cultures of PRK-RuBisCO-expressing strains (Fig. 4B). Acquisition of the ability to aerobically consume sorbitol in batch cultures was previously attributed to mutations in the Tup1-Cyc8 complex, that relieve transcriptional repression of many genes, including HXT15 and SOR2 [40]. However, whole-genome sequencing did not identify such mutations in steady-state chemostat cultures of the PRK-RuBisCO-expressing reference strain IMX1489, which co-consumed sorbitol but did not contain overexpression cassettes for HXT15 and SOR15. Gene expression patterns in S. cerevisiae are strain dependent [41] and glucose concentrations strongly affect transcriptional regulation of HXT transporter genes [37]. Furthermore, expression of HXT13 and HXT15 (two polyol transporters) has been linked to growth on non-fermentable carbon sources [42]. We hypothesize that the conditions in our chemostat setup, which combined low residual glucose concentrations with the presence of sorbitol, stimulated expression of native HXT13 and HXT15 genes and thereby enabled sorbitol utilization in strain IMX1489 (Fig. 4B).

Although expression of the PRK-RuBisCO bypass enabled anaerobic co-consumption of sorbitol and glucose, anaerobic growth of S. cerevisiae on sorbitol as sole carbon source was not yet achieved. In order for sorbitol fermentation to sustain anaerobic growth, the biomass-specific rate of ATP formation should exceed the cellular ATP requirement of anaerobic cultures for cellular maintenance processes (approximately 1 mmol ATP (g biomass)−1 h−1; [8]). At an ATP stoichiometry of 1.17 mol ATP (mol sorbitol)−1 calculated for the PRK-RuBisCO-dependent pathway, the threshold biomass-specific rate of sorbitol fermentation for anaerobic growth would then equal 0.86 mmol (g biomass)−1 h−1. The biomass-specific sorbitol uptake rate of strain IMX2506 in anaerobic mixed-substrate chemostat cultures of 0.67 mmol sorbitol g−1 biomass h−1 (Table 4) remained below this threshold.

In aerobic batch cultures of HXT15- and SOR2-overexpressing S. cerevisiae strains (IME611 and IMX2506) (Additional file 5: Fig. S2) a specific growth rate of 0.22 h−1 was observed. At an estimated biomass yield of 0.07 g−1 (mmol sorbitol)−1 in these respiro-fermentative cultures, this growth rate would correspond to a biomass-specific sorbitol-consumption rate of 3.0 mmol (g biomass)−1 h−1. In anaerobic cultures, such an in vivo activity of sorbitol transport and oxidation to fructose would be over threefold higher than required to meet maintenance-energy requirements.

Alternatively, in vivo capacity of the PRK-RuBisCO pathway might be too low to support anaerobic growth on sorbitol alone. Since form-II RuBisCO enzymes such as T. denitrificans cbbM exhibit low kcat values [32] and require chaperones for functional expression, high-level expression is required to support the in vivo fluxes required for anaerobic growth on sorbitol as sole carbon source. Implementation of RuBisCO variants with a
higher $k_{cat}$ [43], potentially in combination with increased expression of PRK, may enable anaerobic growth on sorbitol.

Instead of targeted engineering of the PRK-RuBisCO pathway, we tried to use adaptive laboratory evolution to identify key genes involved in its in vivo capacity. While, in two parallel evolution experiments, the degree of sorbitol co-consumption initially increased, it subsequently deteriorated. This deterioration was accompanied by an increase of the glycerol concentration in the cultures that indicated loss of functionality of the PRK-RuBisCO pathway. No mutations or reduction of copy number of the $cbbM$ expression cassettes were observed upon prolonged cultivation. This observation suggested that protein burden caused by RuBisCO overexpression was unlikely to be a key factor in the unexpected loss of pathway functionality. Instead, a mutation in the prk gene, duplication of overexpression cassettes for non-oxidative pentose phosphate pathway enzymes and duplication of a genomic region carrying the $HXT6$ and $HXT7$ genes that both encode the high-affinity glucose transporters, were found. These mutations may indicate a trade-off between kinetics of sorbitol fermentation via the PRK-RuBisCO bypass and kinetics of glucose uptake at the very low residual glucose concentrations in the chemostat cultures. Further research is required to investigate whether this trade-off is impacted by the reported toxic effects of PRK overexpression [44].

**Conclusion**

This study provides a proof of principle for engineering a redox-cofactor-neutral bypass of glycolysis to enable entry of additional electrons in the main yeast alcoholic fermentation pathway. This strategy enabled a higher theoretical maximum yield of ethanol than possible with sugars as sole source of electrons. In addition to further analysis of the rate-controlling steps in the PRK-RuBisCO pathway, alternative bypasses of the oxidative step in glycolysis with a better ATP stoichiometry may be explored. A particularly interesting option is offered by the combined expression of a heterologous phosphoketolase, phosphotransacetylase and acetylating acetaldehyde dehydrogenase [45, 46]. In contrast to the PRK-RuBisCO strategy, this redox cofactor-neutral bypass of glyceraldehyde-3-phosphate dehydrogenase has a net positive ATP yield [27]. In addition to improving polyol fermentation, further research on extending flexibility of redox cofactor balancing in yeast may ultimately enable the co-consumption of auxiliary electron donors such as formic acid and/or hydrogen [9, 47, 48] to boost ethanol yields on sugars beyond current limits.

**Methods**

**Strains and maintenance**

*Saccharomyces cerevisiae* strains used in this study (Table 3) belong to the CEN.PK lineage [49, 50] and were propagated in YPD medium (10 g L$^{-1}$ Bacto yeast extract, 20 g L$^{-1}$ Bacto peptone, 20 g L$^{-1}$ glucose). *Escherichia coli* XL-1 Blue-derived strains were propagated in LB medium (10 g L$^{-1}$ Bacto tryptone, 5 g L$^{-1}$ Bacto yeast extract, 20 g L$^{-1}$ glucose) supplemented with 100 µg mL$^{-1}$ ampicillin. After addition of sterile glycerol (30% v/v) to late exponential-phase *S. cerevisiae* cultures or overnight *E. coli* cultures, 1-mL aliquots were frozen and stored at $-80$ °C.

**Construction of plasmids and expression cassettes**

Cas9-based genome editing [51] was employed for markerless integration of expression cassettes in the intergenic region X-2 [52]. Oligonucleotide primers used in this

---

**Table 3**  *S. cerevisiae* strains used in this study

| Strain name | Relevant genotype | Parental strain | Reference |
|-------------|------------------|-----------------|-----------|
| CEN.PK113-7D | MATa URA3 GPD2 | - | [49] |
| IMX581 | MATa ura3-32 GPD2 can1::cas9-natNT2 | CEN.PK113-5D | [51] |
| IME324 | MATa ura3-32 GPD2 can1::cas9-natNT2 pA26-TEF (URA3) | IMX581 | [26] |
| IMX1489 | MATa ura3-32 GPD2 can1::cas9-natNT2 pA26-TEF (CPS1) | IMX581 | [26] |
| IMX2411 | MATa ura3-32 GPD2 can1::cas9-natNT2 X-2::pTEF1-HXT13-tCYC1, pACT1-SOR2-tCPS1 | IMX581 | This study |
| IMX2495 | MATa ura3-32 GPD2 can1::cas9-natNT2 X-2::pTEF1-HXT13-tCYC1, pACT1-SOR2-tCPS1 | IMX1489 | This study |
| IMX2506 | MATa ura3-32 GPD2 can1::cas9-natNT2 X-2::pTEF1-HXT13-tCYC1, pACT1-SOR2-tCPS1 pA26-TEF (URA3) | IMX2495 | This study |
| IME611 | MATa ura3-32 GPD2 can1::cas9-natNT2 X-2::pTEF1-HXT13-tCYC1, pACT1-SOR2-tCPS1 pA26-TEF (URA3) | IMX2411 | This study |
study are listed in Table S1. Plasmid fragments and yeast integration cassettes were PCR amplified with Phusion High Fidelity DNA polymerase (Thermo Fisher Scientific, Waltham MA) and Dreamtaq polymerase (Thermo Fisher). Yeast genomic DNA was isolated as described by [53]. PCR-amplified DNA fragments were purified from agarose gels with a Zymoclean Gel DNA kit (Zymo Research, Irvine CA) or purified directly from the PCR mix with a GeneJET kit (Thermo Fisher). Purified plasmid backbone and insert fragments were assembled with a Gibson assembly cloning kit (Thermo Fisher). Purified plasmid backbone and insert fragments were assembled with a Gibson assembly cloning kit (New England Biolabs, Ipswich MA), downscaled to 5-µL reaction volume. After plasmid assembly, 1 µL of the resulting mixture was used for heat-shock transformation of E. coli XL-1 Blue [54]. Plasmids were isolated from E. coli using Sigma GenElute Plasmid kit (Sigma-Aldrich).

Plasmids used and constructed in this study are listed in Table 4. A pTEF1-HXT15-tCYC1 expression cassette was obtained by assembling three DNA fragments. A TEF1 promoter fragment and a CYC1 terminator fragment were amplified from p426-TEF [55] with primer pairs 16711/16712 and 17032/16712, respectively. The coding region of HXT15 ORF was amplified with primer pair 16705/16706 from genomic DNA of strain CEN.PK113-7D. Gibson assembly of KpnI-linearized pUDE885 with the resulting DNA fragment yielded pUDE941. Amplification of the pACT1-SOR2-tCPS1 expression cassette from pUDE941 with primer pair 16715/16716 added terminal sequences homologous to the X-2 integration site on Chromosome X of S. cerevisiae [52] and to synthetic homologous recombination sequence A (SHR-A). These terminal sequences allowed for simultaneous in vivo assembly and integration into the X-2 locus with the pTEF1-HXT15-tCYC1 cassette.

**Table 4 Plasmids used and constructed in this study**

| Name       | Characteristics | Origin |
|------------|-----------------|--------|
| pUDR538    | Hyg, gRNA.X-2-2 µm-gRNA.X-2 | [56]   |
| p426-TEF   | 2 µm, URA3, pTEF1-tCYC1 (empty vector) | [55]   |
| pUD968     | 2 µm, URA3     | This study |
| pUDE885    | 2 µm, URA3, pACT1-tCPS1 | This study |
| pUDE941    | 2 µm, URA3, pACT1-SOR2-tCPS1 | This study |

Yeast genome editing

The lithium acetate method [57] was used for yeast transformation. S. cerevisiae IMX2411 was constructed by transforming strain IMX581 with pUDR538, along with pACT1-SOR2-tCPS1, pTEF1, HXT15 and tCYC1 DNA fragments. Transformants were selected on YPD plates (10 g L⁻¹ Bacto yeast extract, 20 g L⁻¹ Bacto peptone, 20 g L⁻¹ glucose and 20 g L⁻¹ agar) supplemented with 200 mg L⁻¹ hygromycin B. After verification of correct assembly by diagnostic PCR, single colony isolates were restreaked thrice and stored. The empty-vector reference strain IME611 was constructed by transforming strain IMX2411 with p426-TEF (URA3). Uracil prototrophic transformants were selected on synthetic medium with vitamins [SM, [58]] supplemented with 20 g L⁻¹ agar and 20 g L⁻¹ sorbitol. A pTEF1-HXT15-tCYC1 expression cassette was amplified from genomic DNA of strain IMX2411 with primer pair 16711/16712. Strain IMX2495 was obtained by transforming strain IMX1489 with pUDR538 together with pACT1-SOR2-tCPS1 and pTEF1-HXT15-tCYC1 fragments. Transformants were selected on YPD-hygroycin plates. Strain IMX2506 was obtained by transforming strain IMX2495 with p426-TEF (URA3). Transformants were selected on SM plates supplemented with 20 g L⁻¹ sorbitol.

Shake-flask cultivation

Aerobic shake-flask cultures were grown in 500-mL round-bottom shake flasks containing 100 mL medium, placed in an Innova incubator shaker (Eppendorf Niederland B.V., Nijmegen, The Netherlands) at 30 °C and 200 rpm. Anaerobic cultures were grown in 50-mL round-bottom shake flasks containing 30 mL medium, incubated at 30 °C in a Bactron anaerobic chamber (Sheldon Manufacturing Inc., Cornelius, OR) with an atmosphere of 5% (v/v) H₂, 6% (v/v) CO₂ and 89% (v/v) N₂. Flasks were shaken on a IKA KS 260 basic
shaker (Dijkstra Verenigde BV, Lelystad, The Netherlands) at 200 rpm [59].

Bioreactor cultivation
Anaerobic chemostat cultures and batch cultures were grown at 30 °C in 2-L bioreactors (Applikon, Delft, The Netherlands), with 1-L working volume. Anaerobic chemostat cultures were grown at a dilution rate of 0.025 L h⁻¹ and the effluent pump was controlled by an electrical level sensor to maintain a 1.0-L working volume. The pH was kept constant at 5.0 by automatic addition of 2 M KOH. Chemostat cultures were grown on SM with 10 g L⁻¹ glucose and 10 g L⁻¹ sorbitol and bioreactor batch cultures were grown on SM with 20 g L⁻¹ glucose and 30 g L⁻¹ sorbitol. Media was supplemented with the anaerobic growth factors Tween 80 (420 mg L⁻¹) and ergosterol (10 mg L⁻¹) [59], and 0.2 g L⁻¹ antifoam C (Sigma-Aldrich). Unless otherwise indicated, cultures were sparged at 0.5 L min⁻¹ with an N₂/CO₂ (90/10%) gas mixture. The outlet gas stream was cooled to 4 °C in a condenser to minimize evaporation. Oxygen diffusion was minimized by use of Norprene tubing (Saint-Gobain, Amsterdam, The Netherlands) and Viton O-rings (ERIKS, Haarlem, The Netherlands). Steady state was assumed when, after at least 5 volume changes of operation under constant conditions, biomass dry weight, ethanol, glycerol and acetate concentrations varied by less than 5% over at least two additional volume changes. Inocula for chemostat and bioreactor batch cultures were prepared in 500-mL shake flasks containing 100-mL SM with 20 g L⁻¹ glucose. A first ‘wake-up’ culture was inoculated with frozen stock culture, grown aerobically for 3–5), these were used as inocula for bioreactor cultures at the position of yeast biomass [62].

Analytical methods
Biomass dry weight measurements, analysis of metabolite concentrations and correction for ethanol evaporation were performed as described previously [24]. Metabolite concentrations in steady-state chemostat cultures were analysed after rapid quenching of the culture samples using cold steel beads [60]. Carbon recoveries could not be accurately calculated due to the high concentration of CO₂ in the inlet gas of bioreactor cultures of PRK-RuBisCO expressing strains [24, 26]. Instead, balances of degree of reduction of substrates and products [61] were calculated. These calculations were based on concentrations of relevant components in medium feed and culture samples and on a published value for the elemental composition of yeast biomass [62].

High-throughput analysis of specific growth rates
A Growth-Profiler system (EnzyScreen, Heemstede, The Netherlands) was used for parallel analysis of specific growth rates of multiple S. cerevisiae strains. Cultures were grown under air on SM supplemented with either 20 g L⁻¹ glucose or 20 g L⁻¹ sorbitol, in a culture volume of 250 μL, at 30 °C and at an agitation rate of 250 rpm. The measurement interval was set at 30 min and specific growth rates were calculated from raw green values [63].

Whole-genome sequencing
100 mL aerobic shake-flask cultures S. cerevisiae strains IMX1489 and IMX2506 on YPD were centrifuged for 10 min at 5,000 × g in late exponential phase (OD₆₆₀ of 10–15). For genomic DNA isolation from chemostat cultures of strain IMX1489, 50 mL samples were harvested. Genomic DNA extracted with a Qiagen Blood & Cell Culture NDA kit and 100/G Genomics-tips (Qiagen, Hilden, Germany) was quantified with a Qubit Fluorometer 2.0 (Thermo Fisher). The genome of strain IMX1489 was sequenced in-house as described previously [64] on an Illumina Miseq sequencer (Illumina, San Diego, CA, USA) with a minimum of 50-fold read coverage. Custom paired-end sequencing of genomic DNA of duplicate chemostat cultures of IMX1489 was performed by Macrogen (Amsterdam, The Netherlands) on a 350-bp PCR-free insert library using Illumina SBS technology. Genomic DNA of strain IMX2506 was sequenced by Genomescan (Leiden, The Netherlands) with Illumina SBS technology yielding 151 bp reads with at least 50-fold read coverage. Sequence reads were mapped against the genome of S. cerevisiae CEN.PK113-7D [65] to which a virtual contig containing the sequences of pDAN-prk-tPGK1, pTDH3-cbbm-tCYC1, pTEF1-groEL-tACT1 and pTP11-groES-tPGI1 had been added, and processed as described previously [64].

Stoichiometric analysis
Quantitative estimates of the ethanol yield, biomass yield and biomass-specific rate of substrate consumption were obtained using a stoichiometric model of the core metabolic network of S. cerevisiae [29]. This model was adjusted as described previously, by including a PRK-RuBisCO-based bypass of the oxidative reaction in glycolysis (Fig. 1, [27]). For sorbitol-grown cultures, three additional reactions were implemented, corresponding to a sorbitol facilitator (Eq. 1), a sorbitol dehydrogenase (Eq. 2) and a fructose kinase (Eq. 3).
sorbitol (external) → sorbitol (cytosolic)  \[1\]
sorbitol + NAD+ → fructose + NADH  \[2\]
fructose + ATP → fructose – 6P + ADP  \[3\]

Supplementary Information
The online version contains supplementary material available at https://doi.org/10.1186/s13068-022-02200-3.

Acknowledgements
We thank Mickel Jansen for many stimulating discussions and Walter van Gulik for his assistance in modelling the impact on product yields and substrate uptake rates. We thank Maaike Remeijer for her help in the construction of strain IMX2411 and Nicolò Baldi for the construction of pUD968 and pUD888.

Author contributions
AA: validation, methodology, formal analysis, investigation, writing—original draft, writing—review and editing. JP: conceptualization, supervision, writing—review and editing. RM: conceptualization, supervision, writing—original draft, writing—review and editing. All authors read and approved the final manuscript.

Funding
DSM Bio-based Products & Services B.V. (Delft, The Netherlands).

Availability of data and materials
DNA sequencing data of the *Saccharomyces cerevisiae* strains IMX1489, IMX2506 and evolved population of evolution line 1 (deposited as IMS1232) and evolution line 2 (deposited as IMS1233) were deposited at NCBI under BioProject accession number PRJNA184599. All measurement data used to prepare Fig. 3, Tables 1 and 2, Fig. 5, Figure S2 and Tables S2 and S3 of the manuscript are available in Additional file 1, Additional file 2, Additional file 3 and Additional file 4, respectively.

Declarations

Ethics approval and consent to participate
Not applicable.

Consent for publication
Not applicable.

Competing interests
The PhD project of AA is funded by DSM Bio-based Products & Services B.V. (Delft, The Netherlands). Royal DSM owns intellectual property rights of technology discussed in this paper. AA, JP and RM are co-inventors on patent applications related to the present work.

Received: 11 May 2022 Accepted: 17 September 2022
Published online: 17 October 2022

References
1. Renewable Fuels Association. Annual ethanol production. https://ethanolrfa.org/markets-and-statistics/annual-ethanol-production. Accessed 7 Jan 2022.
2. Lopes ML, de Lima Paulillo SC, Godoy A, Cherubin RA, Lorenzi MS, Giometti FHC, et al. Ethanol production in Brazil: a bridge between science and industry. Braz J Microbiol. 2016;47:64–76.
3. Pfromm PH, Amanor-Boadu V, Nelson R, Vladians P, Madl R. Bio-butanol vs. bio-ethanol: a technical and economic assessment for corn and switchgrass fermented by yeast or Clostridium acetobutylicum. Biomass Bioenergy. 2010;34:515–24.
4. Maorella B, Blanch H, Wilke C. Economic evaluation of alternative ethanol fermentation processes. Biotechnol Bioeng. 1984;26:1003–25.
5. Marques WL, Raghavendiran V, Stambuk BJ, Gombert AK. Sucrose and *Saccharomyces cerevisiae*: a relationship most sweet. FEMS Yeast Res. 2006;6:1193–203.
6. Gascón S, Lampen JQ. Purification of the internal inverterase of yeast. J Biol Chem. 1968;243:1567–72.
7. Dugar D, Stephanopoulos G. Relative potential of biosynthetic pathways for biofuels and bio-based products. Nat Biotechnol. 2011;29:1074–8.
8. Boender LG, de Hulster EA, van Maris AJ, Daran-Lapujade PA, Pronk JT. Quantitative physiology of *Saccharomyces cerevisiae* at near-zero specific growth rates. Appl Environ Microbiol. 2009;75:5607–14.
9. Geertman J-MA, Van Dijken JP, Pronk JT. Engineering NADH metabolism in *Saccharomyces cerevisiae*: formate as an electron donor for glycerol production by anaerobic, glucose-limited chemostat cultures. FEMS Yeast Res. 2006;6:1193–203.
10. Verduyn C, Postma E, Scheffers WA, Van Dijken JP. Energetics of *Saccharomyces cerevisiae* in anaerobic glucose-limited chemostat cultures. Microbiology. 1990;136:405–12.
11. Van Dijken JP, Scheffers WA. Redox balances in the metabolism of sugars by yeasts. FEMS Microbial Rev. 1986;1:199–224.
12. Nissen TL, Hamann CW, Kielland-Brandt MC, Nielsen J, Villadsen J. Anaerobic and aerobic batch cultures of *Saccharomyces cerevisiae* mutants impaired in glycerol synthesis. Yeast. 2000;16:463–74.
13. Sharma S, Horn SJ. Enzymatic saccharification of brown seaweed for production of fermentable sugars. Biores Technol. 2016;213:155–61.
57. Gietz RD, Woods RA. Genetic transformation of yeast. Biotechniques. 2001;30:816–31.
58. Verduyn C, Postma E, Scheffers WA, Van Dijken JP. Effect of benzoic acid on metabolic fluxes in yeasts: a continuous-culture study on the regulation of respiration and alcoholic fermentation. Yeast. 1992;8:501–17.
59. Mooiman C, Bouwknegt J, Dekker WJ, Wiersma SJ, Ortiz-Merino RA, De Hulster E, et al. Critical parameters and procedures for anaerobic cultivation of yeasts in bioreactors and anaerobic chambers. FEMS yeast Res. 2021;21:faob035.
60. Mashego M, Van Gulik W, Vinke J, Heijnen J. Critical evaluation of sampling techniques for residual glucose determination in carbon-limited chemostat culture of Saccharomyces cerevisiae. Biotechnol Bioeng. 2003;83:395–9.
61. Roels J. Simple model for the energetics of growth on substrates with different degrees of reduction. Biotechnol Bioeng. 1980;22:33–53.
62. Lange H, Heijnen J. Statistical reconciliation of the elemental and molecular biomass composition of Saccharomyces cerevisiae. Biotechnol Bioeng. 2001;75:334–44.
63. Baldi N, de Valk SC, Sousa-Silva M, Casal M, Soares-Silva I, Mans R. Evolutionary engineering reveals amino acid substitutions in Ato2 and Ato3 that allow improved growth of Saccharomyces cerevisiae on lactic acid. FEMS Yeast Res. 2021;21:faob033. https://doi.org/10.1093/femsyr/foab033.
64. Perli T, Vos AM, Bouwknegt J, Dekker WJ, Wiersma SJ, Mooiman C, et al. Identification of oxygen-independent pathways for pyridine nucleotide and Coenzyme a synthesis in anaerobic fungi by expression of candidate genes in yeast. MBio. 2021;12:e00967-e1021.
65. Salazar AN, Gorter de Vries AR, van den Broek M, Wijsman M, de la Torre CP, Brickwedde A, et al. Nanopore sequencing enables near-complete de novo assembly of Saccharomyces cerevisiae reference strain CEN. PK113–7D. FEMS Yeast Res. 2017;17:fox074. https://doi.org/10.1093/femsyr/fox074.

Publisher's Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.