Quantified Coexpression Analysis of Central Amygdala Subpopulations

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Abstract

Molecular identification and characterization of fear controlling circuitries is a promising path towards developing targeted treatments of fear-related disorders. Three-color in situ hybridization analysis was used to determine whether somatostatin (SOM, Sst), neurotensin (NTS, Nts), corticotropin-releasing factor (CRF, Crf), tachykinin 2 (TAC2, Tac2), protein kinase c-δ (PKC-δ, Prkcd), and dopamine receptor 2 (DRD2, Drd2) mRNA colocalize in male mouse amygdala neurons. Expression and colocalization was examined across capsular (CeC), lateral (CeL), and medial (CeM) compartments of the central amygdala. The greatest expression of Prkcd and Drd2 were found in CeC and CeL. Crf was expressed primarily in CeL, while Sst-, Nts-, and Tac2-expressing neurons were distributed between CeL and CeM. High levels of colocalization were identified between Sst, Nts, Crf, and Tac2 within the CeL, while little colocalization was detected between any mRNAs within the CeM. These findings provide a more detailed understanding of the molecular mechanisms that regulate the development and maintenance of fear and anxiety behaviors.

Key words: Amygdala; Coexpression; Crf; Sst; Prkcd; Tac2; Nts; fear; Somatostatin

Significance Statement

Functional and behavioral analysis of central amygdala microcircuits has yielded significant insights into the role of this nucleus in fear and anxiety related behaviors. However, precise molecular and locational description of examined populations is lacking. This publication provides a quantified regionally precise description of the expression and coexpression of six frequently examined central amygdala population markers. Most revealing, within the most commonly examined region, the posterior central lateral amygdala (CeL), four of these markers are extensively coexpressed, suggesting the potential for experimental redundancy. These data clarify circuit interaction and function and will increase relevance and precision of future cell type-specific reports.

Introduction

The amygdala comprises a wide array of molecularly, electrophysiologically, and functionally distinct cell populations (Cassell et al., 1999; Ciocchi et al., 2010; Yu et al., 2016). Recent evidence suggests that distinct subpopulations play differential roles in fear and extinction learning...
(Herry et al., 2008; Haubensak et al., 2010; Li et al., 2013a; Kim et al., 2017). The characterization of molecularly identifiable neuronal populations is an important early step in developing improved treatments for fear and anxiety-related disorders (McCullough et al., 2016).

Previous work has shown that the central lateral amygdala (CeL) contains a mutually inhibitory circuit that gates fear expression via the inhibition of central medial amygdala (CeM) output neurons (Herry et al., 2008; Ehrlitch et al., 2009; Ciocchi et al., 2010; Letzkus et al., 2015). The CeL is often conflated with the central capsular division of the amygdala (CeC) although these regions have distinct projection patterns and potentially different roles in fear and anxiety; in the present manuscript we discuss these two regions separately (Jolkkonen and Pitkänen, 1998; Bourgeais et al., 2001).

The protein kinase c-δ (PKC-δ, Prkcd)-expressing neuron population has previously been shown to directly inhibit CeM output neurons, reducing activity in response to conditioned stimuli (CS) following fear conditioning and thus playing an important role in fear extinction learning (Herry et al., 2008; Ciocchi et al., 2010; Haubensak et al., 2010; Cai et al., 2014). The somatostatin (SOM, Sst) expressing population appears to be a counterpart of the PKC-δ population; activity of SOM neurons increases in response to CS following fear conditioning and activity in this population is both necessary and sufficient for the production of fear and defensive behaviors (Li et al., 2013b; Penzo et al., 2014; Yu et al., 2016). The tachykinin 2 (TAC2, Tac2) population plays a complementary role to the SOM population; activity of the Tac 2 expressing population is both necessary and sufficient for fear learning (Andero et al., 2014; Andero et al., 2016).

In addition to SOM and TAC2, other neuropeptides have been implicated as playing critical roles in fear circuitry. In particular corticotropin-releasing factor (CRF, Crf)- and neurotensin (NTS, Nts)-expressing neurons are expressed in populations ideally situated and connected to participate in the central amygdala fear controlling circuit (Petrovich and Swanson, 1997). Both NTS and CRF have been shown to play important roles in fear learning and expression (Merali et al., 1998; Thompson et al., 2004; Yamauchi et al., 2007; Shilling and Feifel, 2008; Gafford and Ressler, 2015).

Dopamine plays a critical role in fear and extinction learning. Specifically, the differential distributions of dopamine receptors may have important implications for mediating fear behaviors (de la Mora et al., 2010; Abra-
The dopamine receptor 2 (DRD2, Drd2) has been suggested to label a large population of neurons implicated in the development and maintenance of fear behaviors (Perez de la Mora et al., 2012; Kim et al., 2017).

Considering the large numbers of CeA cell populations that play parallel or complementary roles in fear behaviors, it is important to determine the extent to which these populations overlap. While much work has been completed identifying markers for behaviorally relevant neuronal populations, less has been done to examine the extent to which each of these populations is distinguishable on the basis of gene expression patterns. In the present investigation three-color in situ hybridization was used to determine the extent of overlap in expression of Prkcd, Sst, Nts, Tac2, Crf, and Drd2. Importantly, significant differences are found in distribution and overlap across the anterior-posterior (A-P) axis of the CeA, thus results are provided as both compressed across the CeA and split into anterior (A-P -0.8 to -1.2) and posterior (A-P -1.3 to -1.8) fractions (McDonald, 2003). Results suggest that within the CeC, Prkcd and Drd2 label large nonoverlapping populations. Within the posterior CeL, Sst, Tac2, Nts, and Crf populations largely overlap. Of these Sst labels the largest population that contains the others markers to varying extents. Within the CeL, the Prkcd and Drd2 populations largely do not overlap with each other or the other populations examined. The CeM has modestly sized Sst, Tac2, Nts, and Crf populations, but is largely devoid of Prkcd- and Drd2-labeled cells. Notably, unlike within the CeL, within the CeM, the Sst, Tac2, Nts, and Crf populations largely do not overlap suggesting important differences in the functional populations labeled by these markers in the CeL and CeM.

Materials and Methods

Animals

C57BL/6J mice were obtained from The Jackson Laboratory. All ten male mice were adult (10 weeks) at the time of tissue collection. All mice were group housed and maintained on a 12/12 h light/dark cycle. Mice were housed in a temperature-controlled colony and given unrestricted access to food and water. All procedures conformed to National Institutes of Health guidelines and were approved by McLean Hospital Institutional Animal Care and use Committee. All animals were killed, and tissue was collected together during light cycle at zeitgeber 3:00 P.M.

RNA scope staining

Staining for mRNA of interest was accomplished using RNA Scope Fluorescent Multiplex 2.5 labeling kit. Probes used for staining are: mm-Nts-C1, mmNts-C2, mm-Tac2-C1, mm-Tac2-C2, mm-Sst-C1, mm-Sst-C2, mm-Crh-C1, mm-Prkcd-C1, mm-Prkcd-C3, mmDrd2-C3. Brains were extracted and snap-frozen in methyl-butane on dry ice. Sections were taken at a width of 16 μm. Staining procedure was completed to manufacturers specifications.

Image acquisition

Images were acquired with experimenter blinded to probes used. Sixteen-bit images of each section were acquired on a Leica SP8 confocal microscope using a 10× objective. Within a sample images were acquired with identical settings for laser power, detector gain, and amplifier offset. Images were acquired as a z-stack of 10 steps of 0.5 μm each. Max intensity projections were then created and analyzed.

Data analysis

The expression and coexpression of mRNA of different markers of interest was quantified in three areas CeC, CeL, and CeM. Images (approximate area) of regions were taken bilaterally from a minimum of one section from each of four animals for each marker pair (n > 8 amygdala/marker pair). Individual cells were identified based on DAPI staining of the nucleus. Cells were determined to be expressing marker when more than five fluorescent dots or an area of staining sufficient to contain five dots was clearly associated with a single nucleus. The width of a cell was considered to be twice the diameter of the nucleus. The distribution of cells across CeA nuclei was determined by dividing the number of labeled cells in a nucleus by the total number of labeled cells across all nuclei. The percentage of cells in a nucleus expressing a certain mRNA was determined by dividing the number of positive cells in a nucleus by the total number of DAPI-labeled nuclei in the nucleus. Experimenters blinded to the identity of probes completed all counts.

Statistical analysis

Determination of the percentage of cells within a subcompartment expressing marker of interest was accomplished by dividing the total number of cells expressing the marker by the total number of DAPI-positive nuclei in the area and multiplying by 100. Determination of the percentage of a labeled population found in a certain subcompartment was accomplished by dividing the number of labeled cells found in a compartment by the total number of labeled cells found in all compartments and multiplying by 100. Statistical analysis of the whether a labeled population was significantly different from the coexpressing component of that population was performed using the nonparametric Mann–Whitney test with GraphPad Prism software package.

Results

Patterns of mRNA expression observed with in situ staining for colocalization of three marker experiments were identical to those observed from single labeling of each marker. Staining patterns were consistent with those observed in the literature and with those produced by the Allen Brain Institute. All six probes produced strong staining in the central CeA.

Each marker was examined individually to characterize its distribution across subcompartments of the CeA. Determination of subcompartment location was accomplished through examination of DAPI staining patterns. As the most popular available brain atlases (Paxinos and Allen Institute) differ somewhat on the locations of CeA...
subcompartments across the A-P axis of the amygdala, the reference atlas provided through the Allen Brain Institute was used as a primary guide (Lein et al., 2007).

**Distribution of labeled cells**

The distribution of total cells expressing mRNAs of interest was examined across CeA subcompartments (Fig. 1A,B) Prkcd staining was almost entirely contained within the CeC (49.8 ± 3.6% of labeled cells) and CeL (44.4 ± 3.8%) with only minority populations found within the CeM (5.7 ± 2.8%). Drd2 was strongly expressed within the CeC (43.3 ± 3.3%) and CeL (32.4 ± 3.3%) with a smaller population within the CeM (24.3 ± 2.8%). Crf was primarily expressed within CeL (69.9 ± 3.8) with smaller populations in CeM (20.7 ± 5.1%) and CeC (9.4 ± 4.1%). Sst, Tac2, and Nts populations are primarily found in CeL (61.3 ± 3.7%, 48.8 ± 6.0%, and 33.3 ± 4.6%, respectively) and CeM (42.0 ± 5.9%, 48.9 ± 5.9%, and 59.5 ± 4.0%, respectively) with only small numbers of cells labeled in CeC (6.7 ± 0.8%, 2.3 ± 1.3%, and 7.2 ± 2.4%, respectively).

**Prevalence of labeled cells**

Single labeling by marker mRNAs was examined to determine their prevalence within a subcompartment (Fig. 1C). This was completed by determining the proportion of labeled cells to the total number of DAPI-positive cells within a compartment. Drd2 and Prkcd label large proportions of cells within the CeC (28.1 ± 2.0% of total DAPI-positive nuclei and 25.0 ± 2.5%, respectively) while other
markers Sst, Tac2, Nts, and Crf label minority populations (5.2 ± 0.6%, 1.0 ± 0.5%, 2.4 ± 0.4%, 3.2 ± 0.3%, respectively). Prkcd, Drd2, Sst, Tac2, Nts, and Crf each label significant populations within the CeL (25.1 ± 2.9%, 18.9 ± 2.3%, 25.9 ± 1.8%, 14.2 ± 1.9%, 13.3 ± 1.4%, and 19.3 ± 1.5%, respectively). Sst, Tac2, and Nts label moderate populations within the CeM (11.1 ± 0.6%, 10.2 ± 1.0%, and 11.3 ± 0.9%, respectively), while Prkcd, Drd, and Crf label smaller proportions of cells (1.1 ± 0.2%, 8.7 ± 1.3%, and 4.1 ± 0.5%, respectively).

Colocalization of CeA markers

Colocalization among markers was examined within each CeA subcompartment. Triple-labeled images were analyzed only for colocalization between pairs of markers due to practical limitations on the number of probe combinations. Additionally, although colocalization was examined at a variety of A-P positions (A-P -0.8 to -1.8), coexpression data are presented in Figures 1–7 is collapsed across A-P -0.8 to -1.8. This may lead to an underestimation of the colocalization of some markers at certain positions (discussed below), but nonetheless provides an indication of overall colocalization between markers in the CeA. Tables 1, 2 provide quantification of expression and colocalization at both anterior (A-P -0.8 to -1.2) and posterior (A-P -1.3 to -1.8) positions.

Sst/Tac2/Prkcd

Sst appears to mark the largest population of CeL cells (Figs. 1C, 2C). This population overlaps to a great extent with Tac2 (Fig. 2D) within the CeL, but not the CeC or CeM (Fig. 2F). Quantification of coexpression reveals that the total Sst labeled population is significantly larger than the coexpressing Sst/Tac2 population in all subcompartments; however, within the CeL the Tac2 population is not significantly different from the coexpressing Sst/Tac2.

Figure 3. Coexpression of Crf, Nts, and Prkcd (A-P -1.5). A, Map of area examined. B, DAPI stain (gray) of area examined. C, Crf expression (green) is found strongly in the CeL and moderately in the CeM. D, Nts expression (red) is found strongly in the CeL and CeM. E, Prkcd expression (cyan) is found strongly in the CeC and CeL. F, Overlay of B–E reveals strong overlap in expression of Crf and Nts in CeL but not CeM. Prkcd does not highly coexpress in any area. Scale bar: 50 μm. G, Quantification of single expressing cells and coexpressing Crf and Nts cells in CeC, CeL, and CeM. Bars represent the mean number of (co)expressing cells in each subcompartment. H, Quantification of single expressing cells and coexpressing Crf and Prkcd cells in CeC, CeL, and CeM. Bars represent the mean number of (co)expressing cells in each subcompartment. I, Quantification of single expressing cells and coexpressing Nts and Prkcd cells in CeC, CeL, and CeM. Bars represent the mean number of (co)expressing cells in each subcompartment. Data presented as mean ± SEM where *p < 0.05 difference between single and double-labeled populations (Mann–Whitney U test).
These data suggest that the larger Sst population may entirely contain the Tac2 population at this A-P range. Prkcd exhibits a typical dense CeC and CeL expression (Fig. 2E). Within the CeL total Sst, Tac2 and Prkcd populations are larger than coexpressing populations, suggesting these RNAs mark separate populations (Fig. 2H, I). Within CeC and CeM total populations are significantly larger than coexpressing populations except in cases where total population is very small.

**Crf/Nts/Prkcd**

Crf labels a large population of CeL cells with somewhat sparser labeling in the CeM (Fig. 3C). Nts marks a large population within the CeL and a moderate population within the CeM (Fig. 3D). Quantification of coexpression reveals the total Crf population is significantly larger than the coexpressing Crf/Nts population in the CeL and CeM; however, within the CeL the Nts population is not significantly different from the coexpressing Crf/Nts population (Fig. 3G). This suggests that within the CeL, the Nts population may be contained within the Crf population, while within the CeM, these populations are distinct. Prkcd demonstrates a similar expression pattern to that seen in Figure 2 (Fig. 3E). The Prkcd population is separately expressed from the Crf and Nts populations in all areas where an appreciable number of marked cells are found (Fig. 3H, I).

**Nts/Sst**

Examination of coexpression of Nts and Sst reveals similar patterns. Within the CeL the Nts population appears to be contained within the Sst population (Fig. 4A–F). While within the CeM these mRNAs mark distinct populations (Fig. 4F).

**Crf/Tac 2**

Likewise, when Crf and Tac2 are examined for coexpression, neither of the total labeled populations is significantly larger than the colabeled Crf/Tac2 populations (Fig. 5C–F). However, within the CeM these mRNAs mark separate populations (Fig. 5C–F).

These results suggest a hierarchical organization within the CeL wherein Sst/H11022, Crf/H11011, Tac2/H11022, and Nts/H11044. This is in contrast to the CeM where all total labeled populations are found to be significantly different from their colabeling with any other marker.

Examination of coexpression at a variety of A-P positions reveals that the zone of highest coexpression between Sst, Tac2, Nts, and Crf is constrained to A-P -1.3 to -1.8. Examination of more anterior positions (A-P -0.8 to -1.2) demonstrates that these populations are coexpressed at lower rates and found in different subcompartments in the anterior CeA.

**Prkcd/Nts/Drd2**

At anterior positions (A-P ~0.9) Drd2 labels a large population of CeC cells while Prkcd cells are found in a cluster in the ventral aspect of the CeC (Fig. 6C,E). At this A-P position Nts is found primarily within the CeM (Fig. 6D). These populations largely do not overlap (Fig. 6F–I).
At a similar A-P position (~0.8) Crf densely labels the CeL (Fig. 7C). Very little Tac2 staining is found within the CeL; however, labeled Tac2 cells are found in the CeM and the dorsal aspect of the main intercalated mass (Im) located ventrally to the BLA (Fig. 7D). Prkcd is found in the ventral CeC (Fig. 7E). These populations largely do not overlap (Fig. 7F).

Slightly more posteriorly (A-P ~1.2), the densely labeled CeL seen in more posterior sections begins to appear (Fig. 7G–L). Sst densely labels the CeL and more sparsely the CeM (Fig. 7I). Prkcd begins to form the typical CeC and CeL expression pattern (Fig. 7K); however, Tac2 does not densely label the CeL at this position and markers continue to be coexpressed at low levels (Fig. 7J–L). These results highlight that the zone of dense coexpression in CeL is constrained to more posterior aspects of the CeA.

The overall percentages of coexpression for all mRNA pairs examined at anterior and posterior positions is presented in Table 1. Table 2 contains quantification of total numbers of labeled and colabeled cells as well as labeled cells as a percentage of total DAPI-labeled cells. Descriptive summary of localization and colocalization of examined CeA mRNA markers is found in Figure 7M.

**Discussion**

The central amygdala plays a pivotal role in the control of a wide range of behaviors, including those reflecting fear and anxiety (Maren and Fanselow, 1996; Johansen et al., 2011; Pare and Duvarci, 2012; Fadok et al., 2017; Lezak et al., 2017). As such, the connectivity, cytoarchitecture and expression profiles of cells in the various subdivisions of this nucleus have been widely studied, especially in rats (McDonald, 1982, 1984; Jolkkonen and Pitkänen, 1998; McDonald, 1998; Cassell et al., 1999). To date, a number of molecularly identified populations have been described as playing distinct roles in the control of behavior. Additionally, the distributions of these populations have been mapped using immunohistochemistry or in situ hybridization (Cassell et al., 1986; Cassell et al., 1999; Andero et al., 2014; Andero et al., 2016). However, minimal data are available on the extent to which these population markers overlap, especially in mice, leading to ambiguity in the specificity of identified and manipulated populations. The present data provide a novel and important advance by examining the coexpression of Prkcd, Sst, Nts, Tac2, Crf, and Drd2 in the CeA of mice. Additionally, these data begin to address the critical need for parsimonious description of both molecular and locational identity of neuron populations examined in the CeA.

Our methods for identifying cells expressing an mRNA involved a binary system so that all cells reaching minimum cutoff (five fluorescent puncta within twice the nuclear diameter) were considered to be expressing. Thus, the representation of low expressing transcripts may be under sampled in exchange for increased confidence. This is most relevant for Drd2, which has the greatest apparent range in expression strength. All animals were the same age and brains were obtained under the same conditions at the same time. This approach was used to...
characterize the baseline identity of cells; however, it ignores a wealth of data concerning levels of expression at the time of sacrifice and dynamic (e.g., circadian or following behavior) changes in expression level. One clear example is that both Tac2 and Sst are clearly expressed at different levels in different populations. Cells appear to express Tac2 and Sst at both high (bright) and moderate/low (dimmer) levels within the same subnucleus. Future studies examining static differences and dynamic changes in mRNA expression level may yield important information regarding the functional roles of these mRNAs.

Our data confirm in mice previous immunohistochemical analyses conducted in rats by demonstrating that within the CeL there is a high degree of overlap between Sst, Nts, Tac2, and Crf. Remarkably, this overlap is observed only within a constrained posterior section of the CeL between A-P -1.4 and -1.8. Examination of these populations across the A-P axis suggests that Crf most consistently marks a CeL population while Sst, Nts, and Tac2 most consistently label cells in the anterior CeM before densely marking the CeL at posterior positions. Within the posterior CeL, these populations are highly overlapping. Sst-expressing cells represent the largest population containing the majority of cells expressing Nts and Tac2 in CeC, CeL, and CeM. Bars represent the mean number of (co)expressing cells in each subcompartment. Data presented as mean ± SEM where *p < 0.05 difference between single and double-labeled populations (Mann–Whitney U test).

Figure 6. Coexpression of Prkcd, Nts, and Drd2 (A-P -0.9). Examination of markers at anterior positions within CeA reveals differential distributions across subcompartments and reduced coexpression. A, Map of area examined (A-P -0.9). B, DAPI stain (gray) of area examined. C, Prkcd expression (green) is found strongly in ventral CeC. D, Nts expression (red) is found strongly in the CeM with limited expression in medial ventral CeC. E, Drd2 expression (cyan) is found strongly in the more dorsal elements of the CeC. F, Overlay of B–E reveals limited overlap in expression of any marker examined. Scale bar: 50 μm. G, Quantification of single expressing cells and coexpressing Nts and Prkcd cells in CeC, CeL, and CeM. Bars represent the mean number of (co)expressing cells in each subcompartment. H, Quantification of single expressing cells and coexpressing Drd2 and Prkcd cells in CeC, CeL, and CeM. Bars represent the mean number of (co)expressing cells in each subcompartment. I, Quantification of single expressing cells and coexpressing Nts and Drd2 cells in CeC, CeL, and CeM. Bars represent the mean number of (co)expressing cells in each subcompartment. Data presented as mean ± SEM where *p < 0.05 difference between single and double-labeled populations (Mann–Whitney U test).
ple, at anterior positions the Allen Brain atlas identifies the location of the dense Crf population as the CeL while the Paxinos and Franklin (2013) atlas identifies this region as the interstitial nucleus of the posterior limb of the anterior commissure (IPAC). While these may be semantic differences, the consistency of nucleus identification has important implication for the quantification of coexpression. Our decision to adhere more closely to the Allen Brain Atlas Reference Atlas may have led to an underestimation of the extent of coexpression of examined markers within the CeL.

Drd2 appears to consistently mark a large CeC and CeL population that is contiguous with the Ast. This is in contrast to Prkcd, which at anterior positions marks a very ventral population of CeC cells before moving more dorsally to mark a very constrained population of CeC and CeL cells at posterior positions. Consistent with previously published work, neither the Prkcd nor the Drd2 populations is highly overlapping with any others examined. This finding validates the identification of these populations as potentially markers for functionally distinct subpopulations (Kim et al., 2017).

Literature identifying functionally distinct CeA populations has been inconsistent in identifying precisely the CeA subcompartment where neurons of interest reside. Such a specific delineation is especially critical in the case of Sst, Nts, Tac2, and Crf, where the identification of these populations within the CeL may be redundant to previous work. Conversely, lack of coexpression in the CeM highlighted by the present findings may indicate a more specialized role for these cells. Future studies using intersectional approaches may yield clear and parsimonious

Figure 7. Coexpression of Crf, Tac2, and Prkcd; and Sst, Tac2, and Prkcd (A-P -0.8 and -1.22). A, Map of area examined (A-P −−0.8). B, DAPI stain (gray) of area examined. C, Crf expression (green) is found strongly in CeL. D, Tac2 expression (red) is found in the CeM. E, Prkcd expression (cyan) is found strongly in ventral CeC. F, Overlay of B–E reveals limited overlap in expression of any marker examined. Scale bar: 200 μm. G, Map of area examined (A-P −−1.2). H, DAPI stain (gray) of area examined. I, Sst expression (green) is found strongly in CeL and CeM. J, Tac2 expression (red) is found in the CeL and CeM. K, Prkcd expression (cyan) is found strongly in CeL and CeC with limited expression in the CeM. L, Overlay of H–K reveals limited overlap in expression of any marker examined. Scale bar: 50 μm. M, Summary figure demonstrating localization and colocalization of examined CeA mRNA markers.
descriptions of the distinct functional roles of single expressing and coexpressing populations in the CeL and CeM (Dymecki et al., 2010; Hirsch et al., 2013; Jensen and Dymecki, 2014; Okaty et al., 2015). Additionally, unlike in the CeL where examined populations label a majority of total cells, in the CeM, examined populations make up less than half of total cells indicating many additional populations that remain to be described.

Table 1. Coexpression of examined mRNAs across CeA subcompartment in anterior and posterior CeA

|       | Anterior                  |       | Posterior                |
|-------|---------------------------|-------|--------------------------|
|       | CeC | Sst (%)| Tac2 (%)| Cf (%)| Nts (%)| Prkcd (%)| Drd2 (%)| CeC | Sst (%)| Tac2 (%)| Cf (%)| Nts (%)| Prkcd (%)| Drd2 (%)|
| Parent Population |     |       |         |       |       |           |         |     |       |         |       |       |           |         |
| CeL   | Sst | 0.0   | 1.7     | 1.25% | 1.75% | 2.95%     | 3.2%    |     |       |         |       |       |           |         |
|       | Tac2| 0.0   | 0.0     | 0.0   | 0.0   | 0.0       | 0.0     |     |       |         |       |       |           |         |
|       | Cf  | 0.0   | 1.7     | 1.25% | 1.75% | 2.95%     | 3.2%    |     |       |         |       |       |           |         |
|       | Nts | 4.3   | 4.3     | 4.3   | 4.3   | 4.3       | 4.3     |     |       |         |       |       |           |         |
|       | Prkcd | 0.0 | 0.0     | 0.0   | 0.0   | 0.0       | 0.0     |     |       |         |       |       |           |         |
|       | Drd2 | 0.0  | 0.0     | 0.0   | 0.0   | 0.0       | 0.0     |     |       |         |       |       |           |         |
| CeM   | Sst | 0.0   | 1.7     | 1.25% | 1.75% | 2.95%     | 3.2%    |     |       |         |       |       |           |         |
|       | Tac2| 0.0   | 0.0     | 0.0   | 0.0   | 0.0       | 0.0     |     |       |         |       |       |           |         |
|       | Cf  | 0.0   | 1.7     | 1.25% | 1.75% | 2.95%     | 3.2%    |     |       |         |       |       |           |         |
|       | Nts | 4.3   | 4.3     | 4.3   | 4.3   | 4.3       | 4.3     |     |       |         |       |       |           |         |
|       | Prkcd | 0.0 | 0.0     | 0.0   | 0.0   | 0.0       | 0.0     |     |       |         |       |       |           |         |
|       | Drd2 | 0.0  | 0.0     | 0.0   | 0.0   | 0.0       | 0.0     |     |       |         |       |       |           |         |

Table 2. Expression and coexpression of examined mRNAs across CeA subcompartment and anterior and posterior axis

|       | CeC | Anterior | Posterior | CeC | Anterior | Posterior | CeC | Anterior | Posterior |
|-------|-----|----------|-----------|-----|----------|-----------|-----|----------|-----------|
|       |     | Average # | % Total   |     | Average # | % Total   |     | Average # | % Total   |
|       |     |           |           |     |           |           |     |           |           |
|       | Cf  | 10.8     | 4.99%     |     | 4.5      | 21.75%    |     | 4.5      | 17.89%    |
|       | Nts | 6.7      | 2.89%     |     | 10.8     | 6.92%     |     | 7.8      | 16.80%    |
|       | Prkcd | 38.3 | 15.48%    |     | 6.0      | 3.83%     |     | 70.5     | 31.18%    |
|       | Tac2 | 1.6     | 0.69%     |     | 7.4      | 2.69%     |     | 46.1     | 17.39%    |
|       | Sst  | 6.3      | 2.91%     |     | 11.7     | 7.76%     |     | 80.2     | 29.73%    |
|       | Drd2 | 79.3     | 31.24%    |     | 45.8     | 28.11%    |     | 39.3     | 16.16%    |
|       | Cf / Nts | 0.6 | 0.26%     |     | 4.4      | 2.55%     |     | 20.7     | 8.99%     |
|       | Cf / Prkcd | 0.4 | 0.18%     |     | 1.9      | 1.06%     |     | 5.3      | 2.33%     |
|       | Cf / Tac2 | 0.4 | 0.20%     |     | 0.9      | 0.32%     |     | 23.1     | 9.24%     |
|       | Cf / Sst | 0.0     | 0.00%     |     | 0.0      | 0.00%     |     | 0.0      | 0.00%     |
|       | Cf / Drd2 | 0.3 | 0.13%     |     | 1.0      | 4.74%     |     | 2.4      | 1.38%     |
|       | Nts / Prkcd | 1.9 | 0.76%     |     | 0.6      | 0.74%     |     | 3.9      | 1.68%     |
|       | Nts / Tac2 | 1.3 | 0.26%     |     | 0.6      | 2.97%     |     | 5.0      | 9.42%     |
|       | Nts / Sst | 0.3     | 0.13%     |     | 0.2      | 0.14%     |     | 37.2     | 13.87%    |
|       | Nts / Drd2 | 4.4  | 1.25%     |     | 0.3      | 0.06%     |     | 1.7      | 0.80%     |
|       | Prkcd / Tac2 | 0.0 | 0.00%     |     | 0.0      | 0.00%     |     | 4.5      | 1.97%     |
|       | Prkcd / Sst | 0.0     | 0.00%     |     | 0.0      | 0.00%     |     | 0.0      | 0.00%     |
|       | Prkcd / Drd2 | 5.5  | 1.78%     |     | 0.0      | 0.00%     |     | 4.0      | 1.06%     |
|       | Tac2 / Sst | 0.3     | 0.06%     |     | 0.3      | 0.13%     |     | 40.5     | 13.70%    |
|       | Tac2 / Drd2 | 0.3  | 0.13%     |     | 0.0      | 0.00%     |     | 1.1      | 0.40%     |
|       | Sst / Drd2 | 0.0     | 0.00%     |     | 0.0      | 0.00%     |     | 0.7      | 0.00%     |

Values presented of average number of positive cells found within a subcompartment in either anterior or posterior region. Additionally, these values are presented as a percentage of total DAPI stained nuclei in each area.
These results represent a starting point in a more comprehensive characterization of the many possible markers for CeA subpopulations. The receptors of the protein products of several of the mRNAs examined may be promising markers for specific subpopulations (Cfr1, Cfr2, Tacr2, Sstr1-5). Additionally, further research into the coexpression of various neuropeptides and other identified markers such as putative adenylate cyclase-activating polypeptide (PACAP), vasoactive intestinal peptide, cholecystokinin, neuropeptide Y, dynorphin, enkephalin, and substance P (all of which have also been shown to also play important roles in fear and anxiety behaviors) will, in the future, be necessary to identify the extent to which additional populations colocalize within the CeA. These types of analyses provide a more thorough understanding of the molecular basis of amygdala function and may facilitate the development of innovative approaches, such as those target-specific cell types by exploiting their unique patterns of receptor expression, to treat fear and anxiety-related disorders.

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