Dual arginine recognition of LRRK2 phosphorylated Rab GTPases

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ABSTRACT Parkinson’s-disease-associated LRRK2 is a multidomain Ser/Thr kinase that phosphorylates a subset of Rab GTPases to control their effector functions. Rab GTPases are the prime regulators of membrane trafficking in eukaryotic cells. Rabs exert their biological effects by recruitment of effector proteins to subcellular compartments via their Rab-binding domain (RBD). Effectors are modular and typically contain additional domains that regulate various aspects of vesicle formation, trafficking, fusion, and organelle dynamics. The RBD of effectors is typically an α-helical coiled coil that recognizes the GTP conformation of the switch 1 and switch 2 motifs of Rabs. LRRK2 phosphorylates Rab8α at T72 (pT72) of its switch 2 α-helix. This post-translational modification enables recruitment of RILPL2, an effector that regulates ciliogenesis in model cell lines. A newly identified RBD motif of RILPL2, termed the X-cap, has been shown to recognize the phosphate via direct interactions between an arginine residue (R132) and pT72 of Rab8α. Here, we show that a second distal arginine (R130) is also essential for phospho-Rab binding by RILPL2. Through structural, biophysical, and cellular studies, we find that R130 stabilizes the primary R132:pT72 salt bridge through favorable enthalpic contributions to the binding affinity. These findings may have implications for the mechanism by which LRRK2 activation leads to assembly of phospho-Rab complexes and subsequent control of their membrane trafficking functions in cells.

INTRODUCTION

Rab GTPases belong to the Ras superfamily of GTP/GDP-binding molecular switches that regulate membrane trafficking in eukaryotic cells (1). GTP/GDP exchange factors (GEFs) convert Rabs into their GTP form and are involved in directing them onto distinct subcellular compartments via their prenylated C-terminal cysteine residues. The switch 1 and 2 regions of Rabs undergo local conformational changes during the nucleotide exchange that enable recruitment of GTP-specific cytosolic effector proteins (2). Rabs are turned off through the action of GTPase activating proteins (GAPs) that aid the hydrolysis of the γ-phosphate of GTP. In recent years, several Rab GTPases have been shown to be targets of protein kinases (3). Phosphorylation of Rabs has emerged as a mechanism for tuning Rab functions through control of the strength of their interactions with GEFs, GAPs, and effector proteins. Many Rabs link directly or indirectly to motor proteins (myosin, dynein, and kinesin) and their associated adaptors (4). Rab connections to the cytoskeleton control processes such as vesicle trafficking, organelle structure, and dynamics (2,5). However, the molecular basis for phosphodependent control of membrane trafficking by Rab GTPases is poorly understood.

We have recently described a novel X-cap motif that enables the RILP family of effectors to bind to phosphorylated Rab8α. Here, we show that recognition of pT72 involves a dual arginine motif involving R130 and R132 from the RH2 domain of RILPL2. Thermodynamic and structural studies suggest that the phosphospecific interactions are highly sensitive to the upstream region, which includes the N-terminal RH1 domain that binds to myosin Va. The dual arginine motif may facilitate the dynamic nature of membrane trafficking processes involving phosphorylation of Rabs.
Rab GTPases (6). The N-terminal RH1 (RILP homology) domain of the cytosolic adaptor Rab-interacting lysosomal protein-like 2 (RILPL2) binds to the globular tail domain of myosin Va (7). The C-terminal RH2 domain of RILPL2 encodes a Rab-binding domain (RBD) that adopts a parallel α-helical coiled-coil structure. The N-terminus of the RBD forms an X-cap motif that resembles a hook that winds around its partner. This motif is stabilized by a short antiparallel β-strand-like interaction and has an overall X-shaped conformation that caps the α-helical coiled coil. In the structure of pRab8a with a minimal RBD (residues 129–165), the X-cap enables projection of an Arg residue (R132RL2) from each monomer toward a phosphorylated threonine residue of Rab8a (pT72R8). In cells, the kinase LRRK2 phosphorylates Rab8a and Rab10 at conserved threonine residues in the switch 2 region, and the post-translational modification tunes the strength of Rab interactions with their binding partners (8). Rab8a/10 phosphorylated at pT72/pT73 recruits RILPL2 to membranes and affect the growth of primary cilia in model cell lines (9). In previous work, we also showed that myosin Va binding to RILPL2 enhances its affinity for pRab8a through an unknown mechanism (6). LRRK2 is the leading cause of inherited Parkinson’s disease, a disorder of the central nervous system that manifests as a progressive degeneration of motor mobility, balance, and tremors. Phosphodependent recruitment of RILPL2 by pRab8a/10 is a part of the underlying molecular pathways that connect LRRK2 to neuronal degeneration (10). Therefore, it is critical to understand the biophysical processes that underpin Rab regulatory pathways associated with Parkinson’s disease.

Here, we have determined the structure of pRab8a in a complex with an extended RBD of RILPL2 (117–165; abbreviated as RL2117) at 1.9-Å resolution. The structure includes residues preceding the X-cap that were not mapped in our previous work. In addition, we have determined the structure of the phosphomimetic T72E mutant of Rab8a (Rab8aTE) in complex with the minimal RBD of RILPL2 (129–165; RL2129) at 1.7-Å resolution. These two structures are analyzed in comparison to the previously published structure of pRab8a in complex with the minimal RBD of RILPL2 (129–165; RL2129). We find that the overall structures are identical in all three complexes with no significant differences in intermolecular contacts. However, there are dramatic conformational changes localized to two arginine residues (R130RL2, R132RL2) in the X-cap of RILPL2. The complex with the extended RBD (RL2117) has two well-ordered stacked arginines in which R130 appears to stabilize the conformation of R132, which directly forms a salt bridge to the pT72R8. The other two complexes display various levels of disorder, suggesting an influence of the upstream residues of RILPL2 on pRab8a recognition. Calorimetry reveals that RL2117 has a significantly higher affinity for pRab8a relative to RL2129. We conclude that a robust interaction between the RILP family of effectors with cognate phospho-Rabs is dependent on a dual arginine recognition motif. Mutation of distal R130 to lysine/glutamine is sufficient to abolish interactions with pRab8a in cells. The apparent sensitivity of the dual arginine motif to the upstream region of RILPL2 suggests a mechanism by which myosin:RILPL2 complex formation may regulate its recruitment to phospho-Rab8a membranes.

**MATERIALS AND METHODS**

*Escherichia coli* expression constructs

The *E. coli* expression optimized cDNA for Rab8a was obtained from GenScript and covers the residues 1–181 of human Rab8a (Q67L; Q67L + T72E), as previously described (6). The Q67L mutation maintains the Rab8a in the active GTP-bound conformation. The cDNA was cloned into the PET28a vector using *Nde*I/HindIII cloning sites and provides a hexahistidine tag that is cleavable by thrombin. The cDNA for human RILPL2117–165 was amplified by PCR from a human full-length RILPL2 construct in PET28a (6) using the following primers: 5'-C AG GGA GCC TCG TGG GGC ACG AAC AAG ATG GTG G-3' (forward); 5'-GC AAA GCA CGG GCC TCG TTA ACC GCT TTT GTA GCA TGG CAG-3' (reverse). Similarly, the cDNA for human RILPL2129–165 was made using the following primers: 5'-C AG GGA GCC TCG TGG GGC ACG AAC CTT CGT GTT ACC C-3' (forward); 5'-GC AAA GCA CGG GCC TCG TTA ACC GCT TTT GTA GCA TGG CAG-3' (reverse). The resulting DNA was cloned using ligation-independent methods into the pLIC-MBP vector (11). The construct was sequenced and transformed into *E. coli* BL21(DE3) for expression as His6-MBP-(TEV)-RILPL2117–165.

**Protein production**

Expression of Rab8a was carried out in LB (Miller) broth supplemented with antibiotic (34 µg/mL kanamycin or 100 µg/mL ampicillin) at 37°C. Media, chemicals, and antibiotics were obtained from FOMEDIUM. At an OD600 of 0.6, the culture was cooled to 18°C for ~1 h and induced with 0.5 mM IPTG, after which cells were grown overnight at 18°C. Expression of His6-MBP-RILPL2117–165 was carried out in LB (Miller) medium supplemented with 100 µg/mL ampicillin (FOMEDIUM) at 37°C. At an OD600 of 0.6–0.8, the culture was induced with 0.5 mM IPTG. Cells were grown for an additional 4 h at 37°C. After induction, cells were harvested by centrifugation, and the pellets were resuspended in His-tag extraction buffer (20 mM Tris-HCl, 300 mM NaCl, 10 mM imidazole, and 10 mM β-mercaptoethanol (pH 8.0); ≥5 mM MgCl2 for Rab constructs). Cells were lysed by sonication, and the cell lysate was centrifuged at 26,000 × g for 45 min at 4°C to remove cellular debris. The supernatants were loaded onto an Ni2+-agarose resin (Qiagen). The resin was washed with a 10-fold excess of extraction buffer followed by washing with a five-fold excess using the same buffer supplemented with 40 mM imidazole before elution of the bound proteins using extraction buffer supplemented with 200 mM imidazole. The eluted proteins were dialyzed against 20 mM Tris-HCl, 100 mM NaCl, 5 mM MgCl2, and 1 mM DTT (pH 7.5). Overnight incubation at 4°C with thrombin (GE Healthcare) or TEV protease was used to remove the N-terminal hexahistidine or hexahistidine-MBP tags from each protein. After cleavage, the proteins were run through a second Ni2+-agarose column. The flow-through fractions were collected, while the uncleaved proteins remained bound to the resin. The peptide corresponding to residues 129–165 of RILPL2 was synthesized with an N-terminal hexahistidine tag (GenScript). The peptide was solubilized in aqueous buffer (20 mM Tris-HCl, 100 mM NaCl, 5 mM MgCl2, 1 mM DTT (pH 7.5)) before crystallization trials.

The second nickel procedure was repeated to minimize the amount of uncleaved His6-MBP-RILPL2 precursor. Cut RL2117 was dialyzed into aqueous buffer (20 mM Tris-HCl, 100 mM NaCl, 5 mM MgCl2, 1 mM DTT (pH 7.5)) before crystallization trials.

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Cocrystallization

Cocrystals of Rab8TE:RL2129 complex were obtained in a 1:2 molar ratio of protein/peptide at a total of 12 mg/mL. Crystals were grown in 100 mM HEPES buffer (pH 7.5), 10% PEG 4000, and 10% 2-propanol. Plate-like crystals were harvested in precipitant supplemented with 25% glycerol and stored frozen in liquid nitrogen. X-ray data were collected under a cryogenic nitrogen stream at 100 K (beamline 24-ID-C; Advanced Photon Source).

For in vitro pulldowns, hexahistidine-tagged full-length RILPL2 (1–211) or hexahistidine-MBP-tagged truncated RILPL2 constructs were used. Rab8 and RILPL2 constructs (10 μM each) were mixed together in 1.5-mL centrifuge tubes with 25 μL Ni²⁺-agarose resin in a final volume of 1 mL of binding buffer (20 mM Tris (pH 8.0), 300 mM NaCl, 20 mM imidazole, 5 mM MgCl₂, 10 mM β-mercaptoethanol). The reaction mixture was mildly shaken for 15 min followed by gentle centrifugation (1000 rpm). The resin was washed three times with 1 mL of the binding buffer. After release of proteins from resin with 50 μL elution buffer (20 mM Tris-Cl (pH 8.0),

| TABLE 1 Crystallographic data and refinement statistics |
|--------------------------------------------------------|
| **Rab8TE** | **Rab8TE:RILPL2 (129–165)** | **pRab8a:RILPL2 (117–165)** |
| Beamsline | NSLS FI MX | NECAT APS, 24-ID-C | NECAT APS, 24-ID-E |
| Wavelength (Å) | 0.9789 | 0.9789 | 0.97918 |
| Space group | P 2₁ | P 2₁, 2₁, 2₁ | C 2 2 2 1 |
| Asymmetric unit | 2 × Rab8a | 2 × Rab8a, 2 × RILPL2 | 1 × Rab8a, 1 × RILPL2 |
| Cell (a, b, c, Å) | 36.09, 118.38, 39.6 | 60.698, 71.733, 116.39 | 62.773, 68.43, 128.1 |
| Resolution (Å) | 29.29–1.72 (1.76–1.72) | 61.07–1.684 (1.744–1.684) | 46.26–1.9 (1.968–1.9) |
| Number of reflections: total (unique) | 144,140 (32,822) | 325,450 (10,484) | 183,379 (15,819) |
| Completeness (%) | 96 (85.2) | 96.86 (79.50) | 96.02 (90.3) |
| <I/σ(I)> | 6.9 (2.6) | 20.31 (2.44) | 16.79 (0.35) |
| Multiplicity | 4.4 (3.9) | 5.7 (2.3) | 8.6 (7.9) |
| Rmerge | 0.135 (0.441) | 0.0543 (0.403) | 0.05713 (4.301) |
| CC1/2 | 0.986 (0.806) | 0.998 (0.584) | 0.999 (0.54) |

**Refinement**

| Rwork | 0.1865 (0.2184) | 0.1815 (0.3305) | 0.2187 (0.5266) |
| Rfree | 0.2302 (0.2690) | 0.2028 (0.3577) | 0.2669 (0.5685) |
| Rfree size (Å) | 1632 (163) | 2867 (217) | 1040 (111) |
| RMSD bond lengths (Å) | 0.006 | 0.011 | 0.014 |
| RMSD bond angles (°) | 0.86 | 1.32 | 1.33 |
| Average overall B-factor | 19.83 | 24.93 | 57.94 |
| Mean B-factors (Å²) protein/GTP/waters | 18.92/21/25 | 23.44/14.15/36.21 | 58.28/48.17/53.94 |
| Ramachandran analysis favored/allowed (%) | 97.96/2.04 | 95.63/2.72 | 97.17/2.83 |
| PDB accession code | 6whe | 6sq2 | 7lw8 |

Values in parentheses correspond to the statistics in the highest resolution bin. $R_{merge} = \sum hkl |I_{hkl} - \langle I_{hkl} \rangle| / \sum hkl I_{hkl}$, $R_{work} = \sum hkl |F_{o,hkl} - F_{c,hkl}| / \sum hkl F_{o,hkl}$, $R_{free}$.
300 mM NaCl, 200 mM imidazole), samples were subjected to SDS-PAGE and visualization with 0.5% Coomassie Brilliant Blue.

Calorimetry experiments were performed in triplicate. The N-terminal polyhistidine-tagged RILPL2 peptide (His6-RILPL2129–165), or recombinantly expressed RILPL2117–165, was dialyzed together with pRab8a/ Rab8TE (20 mM Tris-HCl, 150 mM NaCl, 5 mM MgCl2, and 1 mM DTT (pH 7.5)). Protein concentrations were calculated based on their absorbance at 280 nm using an ND-1000 NanoDrop spectrophotometer (Thermo Scientific).

Protein concentrations were calculated based on their absorbance at 280 nm using an ND-1000 NanoDrop spectrophotometer (Thermo Scientific). Samples were centrifuged at 13,200 rpm for 10 min before the determination of protein concentration and ITC analyses. Injection of Rab8a/pRab8a into RILPL2 was performed, rather than the other way around. RILPL2 injections into buffer revealed a large heat of dilution, which complicated estimates of binding enthalpy and Kd. The concentrations of proteins for injection were between 400 and 600 μM Rab8a and 40–60 μM RILPL2. Data were processed using Origin 7.0 with the ITC plug-in.

**Plasmids for cellular assays**

The plasmids used for coimmunoprecipitation experiments were acquired from MRC PPU Reagents and Services (https://mrcppureagents.dundee.ac.uk/reagents-proteins/overview/): GFP-empty pcDNA5 FRT/TO (DU13156); Flag-LRRK2 Y1699C pcCMV (DU13165); HA-Rab8a WT pcCMV (DU35414); RILPL2-GFP WT pcDNA5 FRT/TO (DU27481); RILPL2-GFP P128A pcDNA5 FRT/TO (DU68411); RILPL2-GFP D127A pcDNA5 FRT/TO (DU68427); RILPL2-GFP D127N pcDNA5 FRT/TO (DU68428); RILPL2-GFP T126A pcDNA5 FRT/TO (DU68412); RILPL2-GFP D124A pcDNA5 FRT/TO (DU68429); RILPL2-GFP D124A pcDNA5 FRT/TO (DU68413).

**Antibody reagents**

Antibodies used in this study were diluted in 5% w/v bovine serum albumin in TBS supplemented with 0.1% Tween-20 (TBS-T) and 0.03% w/v sodium azide. The rabbit monoclonal antibody for total LRRK2 (N-terminus) was purchased at the University of Dundee (16). Anti-GFP (PABG1; Chromotek, used at 1:1000), anti-HA (3F10; Merck, used at 1:1000), anti-pT72-Rab8a (MjF-R20; Abcam, used at 0.5 μg/mL), anti-LRRK2 C-terminal (N241A/S; Neuronab, used at 1:1000), and anti-α-tubulin (3873S; CST, used at 1:5000). Secondary antibodies used were LI-COR IRDye for 800CW goat anti-rabbit (925–32211), goat anti-mouse (926–32210), and 680LT goat anti-rabbit (925–68029) and goat anti-mouse (926–68020), all used at 1:10,000 dilution in TBS-T.

**Culture and transfection of cells**

HEK293 cells were cultured in Dulbecco’s modified Eagle medium (Gluta-max; Gibco) supplemented with 10% fetal bovine serum (Sigma), 100 U/mL penicillin, and 100 μg/mL streptomycin. Transient transfections were performed 40–48 h before cell lysis using polyethylenimine PEI (Polysciences) at around 60–70% confluence. Transfections for coimmunoprecipitation experiments were done in 10-cm round cell culture dishes using 3 μg of Flag-LRRK2 Y1699C, 1 μg of HA-Rab8a, and 1 μg of GFP control or RILPL2-GFP cDNA construct per dish diluted in 1 mL of OPTIMEM media and supplemented with 20 μg of PEI incubated for 20 min before being added to the cell media. At 90 min before lysis, cells were treated with 500 nM of MLI-2 inhibitor or 0.1% DMSO control.

**Comunoprecipitation of Rab8a and RILPL2**

Cells were washed with phosphate-buffered saline and lysed in lysis buffer—50 mM Tris–HCl (pH 7.5), 1 mM EDTA, 10 mM sodium β-glycerophosphate, 50 mM sodium fluoride, 5 mM potassium phosphate × 10H2O, 0.27 M sucrose—and supplemented fresh before lysis with 1% v/v Triton-x100, one tablet of cOmplete Mini (EDTA-free) protease inhibitor (1183671001; Merck) per 10 mL of buffer, 0.1 μg/mL of microcystin, and 1 μM of sodium orthovanadate. Lysates were clarified by centrifugation at 17,000 × g for 10 min. For GFP immunoprecipitation, lysates were incubated with nanobody α-GFP binder Sepharose from MRC PPU Reagents and Services for 1 h at 4°C (15 μL of packed resin/0.5 mg of lysate). Bound complexes were recovered by washing the beads three times with wash buffer (50 mM Tris–HCl (pH 7.5), 150 mM NaCl) before eluting with 2× SDS-PAGE sample buffer supplemented with 1% v/v 2-mercaptoethanol. The samples were denatured at 70°C for 10 min, and the resin was separated from the sample by centrifugation through a 0.22-μm Spinex column (CLS8116; Sigma). Samples were subjected to immunoblotting.

**Gel electrophoresis and immunoblot analysis**

Samples were run on gels consisting of a 4% w/v acrylamide stacking gel (4% w/v acrylamide, 0.125 M Tris–HCl (pH 6.8), 0.2% v/v tetramethylizedenediamine, and 0.08% w/v ammonium persulfate (APS)) and 10% w/v acrylamide separating gel (10% w/v acrylamide, 0.375 M Bis-Tris (pH 6.8), 1% v/v tetramethylizedenediamine, and 0.05% w/v APS) in MOPS buffer (50 mM MOPS, 50 mM Tris, 1 mM EDTA, 0.1% w/v SDS) at 90–120 V. Proteins were electrophoretically transferred onto nitrocellulose membranes (Amersham Protran 0.45 μm nitrocellulose; GE Healthcare) at 90 V for 90 min on ice in transfer buffer (48 mM Tris/HCl, 39 mM glycine, 20% v/v methanol). Transferred membranes were blocked with 5% w/v nonfat dry milk dissolved in TBS-T (20 mM Tris/HCl (pH 7.5), 150 mM NaCl, and 0.1% v/v Tween 20) at room temperature for 1 h. Membranes were then incubated with primary antibodies overnight at 4°C. After washing membranes in TBS-T × 15 min, membranes were incubated with secondary antibodies at room temperature for 1 h. After washing membranes in TBS-T × 15 min, membranes were scanned using LI-COR Odyssey CLx. Protein band intensities were analyzed using LI-COR Image Studio Lite software.

**RESULTS AND DISCUSSION**

**Structural comparisons of Rab8a complexes with RILPL2**

The structure of GTP-bound pRab8(Q67L) in complex with RL2117 forms crystals with one molecule of pRab8a and one molecule of RILPL2 as the repeating unit. The biological heterotetramer is generated by a twofold crystallographic axis down the length of the central parallel α-helical dimer of RL2117. In the biological assembly, the effector bridges two molecules of Rab8 via hydrophobic and polar interactions. The X-cap is formed at the top of the α-helices, and it is critical for positioning the effector to enable direct contacts between R132RL2 and pT72R8. The structure resembles the reported complex of pRab8a in complex with RL2129 (Fig. 1 B), in which the asymmetric unit was the biologically relevant heterotetramer (6). In this previous structure with the minimal RBD, the salt bridge between pT72R8 and R132RL2 was well ordered, but the distal R130RL2 displayed considerable flexibility. In contrast, the structure of pRab8a in complex with RL2117 reveals a well-ordered R130RL2 that stacks against R132RL2 (Fig. 1 C). This stable arrangement of the dual arginines may enable formation of a symmetric twofold crystallographic axis. In the ensuing discussions, stacking will refer to planar π–π interactions of the guanidine side chains of arginines. Crystallographic details are shown in Table 1.
Although the RBD of the complex begins at 117, residues 117–125 are disordered. The model contains residues 126–161 of RILPL2 and resembles closely the minimal RBD (129–161) of the previous structure (pRab8a:RILP129 complex). There are no additional interactions with pRab8a, and upon superposition, equivalent backbone residues have a root mean-square deviation of 1.44 Å. The only significant difference between the complexes is the conformation of R130RL2, which is discussed in more detail below. As a further comparison, the structure of a phosphomimetic mutant Rab8a-T72E (Rab8TE) in complex with RILP129 has also been determined. The complex Rab8TE:RILP129 crystallizes in the same space group as pRab8a:RILP129 (Table 1), and again, there are no significant differences at the interface.

The three complexes afford an excellent opportunity to investigate the structural and thermodynamic details of phospho-Rab binding by effectors. Electron density maps reveal that one side of the E72R8:R132RL2 interaction in the Rab8TE:RL2129 complex is well ordered and mimics the pT72R8:R132RL2 salt bridge. However, the opposite side shows some flexibility as evidenced by the poor density and geometry (Fig. 2A). In contrast, the key pT72R8:R132RL2 interaction in the pRab8a:RILP129 complex is well ordered, suggesting a stronger salt bridge with phosphothreonine (Fig. 2B). There is also partial stacking of the R132/R130 side chains in the two complexes with the minimal (RILP129) RBD. R130RL2 in the complex pRab8:RILP129 reveals better stacking relative to Rab8TE, but there remains considerable flexibility as evidenced by electron density maps (Fig. 2B).

Strikingly, the newly determined structure of pRab8a:RL2117 reveals an ordered R130 side chain that forms strong stacking interactions with the guanidino group of R132RL2. As a measure of the close contacts, the
distance between R130(Nε) and R132(NH2) is 3.4 Å. In contrast to the other complexes, pRab8a:RILPL2117 forms a symmetric twofold axis down the length of the coiled coil. Therefore the two salt bridges between pT72R8 and R132 are identical by symmetry in the biological heterotetramer. Rab8TE:RL2129 and pRab8a:RL2129 assemble as heterotetramers in the lattice. As suggested previously, it is possible that conformational flexibility in R130/R132 may prevent a symmetric twofold axis from forming in the other two complexes. Previously, we reported that R130RL2 was essential for complex formation in cells. In the context of full-length RILPL2, even a mutation to lysine (R130K) significantly reduced the interaction with pRab8a (6). However, the contribution of this distal arginine to complex formation was unclear because the side chain was relatively disordered. Given the short RBD (129–165) in previous work, we could not exclude the possibility that both R130 and R132 might directly interact with pT72R8 in longer variants of RILPL2. Here, the structure of pRab8a with an extended RBD–RL2117 suggests that R130RL2 contributes to p-p stacking interactions with R132RL2 and stabilizes its salt bridge with pT72R8. A lysine residue (R130K) cannot substitute for this interaction, thus providing a rationale for the inability of a positive charge to maintain binding to pRab8a.

**Thermodynamics of complex formation**

In vitro pulldowns were performed to qualitatively assess the affinities of the three complexes (Fig. 3A). These experiments suggested that full-length RILPL2 (1–211) has less affinity to pRab8a relative to the truncated variant RL2117. Although the in vitro reduction in affinity appears to be modest, the finding is surprising because effectors are modular and RBD interactions with Rabbs are generally independent of other domains (5). Isothermal titration calorimetry (ITC) was then exploited to provide more insight into the binding affinities (Fig. 3B; Table 2). The data reveal that the affinity between pRab8a and RL2117 is significantly stronger than other complexes, with a more robust enthalpy contributing to complex formation (Fig. 3B). As discussed previously, comparisons of the three complexes—Rab8TE:RL2129, pRab8a:RL2129, and pRab8a:RL2117—reveal no significant differences apart from the side chain of R130. Therefore, the increased enthalpic contribution to the binding affinity can be attributed to a stronger salt bridge between pT72R8 and R132RL2 (Fig. 3C). Although the distal R130RL2 is relatively far from the nearest phosphate oxygen (5.6 Å), it may also contribute to the enthalpy through long-range electrostatic interactions. The bar graph of free-energy signatures (ΔH, ΔTS) also indicates that the entropic term contributes favorably toward formation of all of the complexes. However, the enthalpic gains are countered by a reduced entropic contribution, which can partially be attributed to ordered guanidino side chains. The complexes Rab8TE:RL2129 and pRab8a:RL2129 have a similar Kd, but favorable enthalpy for the pT72:R132 interaction is offset by a reduction in the entropic gains (Fig. 3D; Table 2). Intriguingly, relative to the truncated RL2117 variant, full-length RILPL2 has a reduced affinity to pRab8a with less favorable enthalpy. The N-terminal RH1 domain appears to antagonize the interactions between pRab8a and the RH2 domain by an unknown mechanism that requires further investigation.

In addition to complexes, we also determined the structure of uncomplexed Rab8TE. This was performed to support the hypothesis that thermodynamic parameters can be attributed to complex formation rather than intrinsic structural changes.
from the negative charge at switch 2. The structure of Rab8\textsubscript{TE} reveals no significant conformational differences in the overall fold relative to the complex Rab8\textsubscript{TE}:RL2\textsubscript{129} (Fig. S1).

\begin{table}[h]
\centering
\begin{tabular}{|c|c|c|c|c|}
\hline
 & K\textsubscript{D} (\textmu M) & \Delta H (kcal/mol) & \text{-}T\Delta S (cal/k\textdegree mol) & N \tabularnewline \hline
Rab8\textsubscript{TE}(Q67L) & & & & \\
RL2\textsubscript{129} & 3.4 \pm 0.5 & -2.7 \pm 0.3 & -4.6 \pm 0.5 & 1.03 \pm 0.05 \\
Rab8a(Q67L) & & & & \\
RL2\textsubscript{129} & 3.3 \pm 0.4 & -5.1 \pm 0.4 & -2.2 \pm 0.4 & 0.92 \pm 0.2 \\
RL2\textsubscript{117} & 0.54 \pm 0.1 & -7.4 \pm 0.2 & -1 \pm 0.3 & 1.1 \pm 0.15 \\
RL2-FL & 1.4 \pm 0.3 & -1.4 \pm 0.3 & -6.4 \pm 1.2 & 1.04 \pm 0.1 \\
\hline
\end{tabular}
\caption{Thermodynamics of RILPL2 binding to pRab8a/Rab8\textsubscript{TE}}
\end{table}

Similarly, the recent structure of phosphorylated Rab8a at S111 is identical to the unphosphorylated variant (17). Although the structure of uncomplexed pRab8a-pT72 has not yet been determined, it is likely that phosphorylated Rabs do not undergo significant conformational changes upon complex formation with RILPL2.

\textbf{Mutagenesis and cellular assays}

Mutational and cellular analyses of RILPL2 residues preceding R130\textsuperscript{RL2} were performed to assess the contribution of this segment to pRab8a binding (Fig. 4). GFP-tagged full-length RILPL2 variants were overexpressed in HEK293 cells together with HA-tagged Rab8a and Flag-tagged LRRK2.
Y1699C to ensure maximal phosphorylation of Rab8a. LRRK2 specific inhibitor Mli-2 was used at 500 nM concentration for 90 min as a control. RILPL2 was immunoprecipitated using anti-GFP resin, and the samples were subjected to immunoblotting. These studies reveal that single mutants D124ARL2 and D127ARL2 are partially defective in binding to pRab8a (Fig. 4, A and B). The side-chain carboxylate of D127RL2 is within 3 Å of the backbone NH groups of N129RL2 and R130RL2. Although the geometry is not ideal, these interactions may nevertheless contribute to stabilization of the backbone conformation. It would also explain why D127NRL2 does not have the same defect as D127ARL2 (Fig. 4A). The effects of mutations are mapped as hotspots on a sequence alignment of the predicted X-cap of effector proteins (Fig. 4, B and C). There are modest sequence similarities in the upstream region preceding R130 within the RILP family. Red circles denote hotspots in which any mutation abolishes binding in cells (explored in our previous publication (6)), whereas yellow circles denote an intermediate phenotype. The molecular basis for the contribution of D124RL2 to phospho-Rab recognition is unknown because the segment 117–125 is flexible in electron density maps.

**CONCLUSION**

Phosphorylation of Rab GTPases has emerged as a regulatory mechanism for membrane trafficking in a variety of contexts (3). The switch 2 α-helix of Rab GTPases is a hotspot for phosphorylation, and its modification is likely to influence subsequent interactions with GAPs, GEFs, and effector proteins. Here, we show that the conformation of arginine residues from RILPL2 involved in pRab8a recognition is sensitive to the region preceding the RBD. In particular, distal R130RL2 is flexible, without a stabilizing influence from upstream residues. However, the full-length effector comprising both RH1/RH2 domains dramatically reduces the enthalpy of complex formation, suggesting that the distal R130 is disordered. The structural and thermodynamic studies also suggest that the enthalpic gain from the salt bridge between R132/pT72 is offset by a reduction in favorable entropy arising from the ordering of the guanidino side chains of R130/R132. A video is shown with the three complexes as a trajectory (Rab8TE:RILP129/pRab8:RILP117) using the Morph application in Chimera (Video S1: 18). Despite the lack of any additional interactions, the upstream segment of RILPL2 appears to stabilize the conformations of the two arginines for optimal interactions with pRab8a. Cation-π and π-π interactions that involve aromatic residues are widely known for their stabilizing influence in a variety of contexts. However, arginine clusters are increasingly being recognized for their contribution to protein complex formation (19–21). Conformational dynamics of RILPL2 have also previously been observed in its interactions with Rab36 and myosin Va (22).
The structures of numerous signaling complexes involving phosphorylated peptides have been determined. In general, the peptides adopt an extended conformation, and the phosphorylated side chains are deeply embedded, with interactions on both sides of the phosphate moiety (Fig. 5, A and B). In contrast, the RILPL2 interaction with the phosphorylated switch 2 helix is more peripheral (Fig. 5 C). This more subtle interaction motif in the pRab8a:RILPL2 complex may be influenced by the upstream hinge region that connects the RH1 domain to the RH2 domain. We have shown previously that myosin Va binding to the RH1 domain enhances the affinity of the RH2 domain to pRab8a (6). An attractive mechanistic model is that myosin binding facilitates a conformation for the hinge region that enables recognition of pRab8a by a dual arginine motif. Myosin binding to the RH1 domain may enable ordering of the distal arginine (R130), which would enhance the stability of the salt bridge between R132 and pT72 of Rab8a. One prediction from this model is that myosin:RILPL2 complexes are more likely to be recruited to phospo-Rab8a membranes downstream of LRRK2 activation.

**SUPPORTING MATERIAL**

Supporting material can be found online at https://doi.org/10.1016/j.bpj.2021.03.030.

**AUTHOR CONTRIBUTIONS**

D.W. and E.P. analyzed data, performed the research, and wrote the article. A.R.K. designed the project, analyzed data, performed research, and wrote the article.

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