**IL-1α Expression in Pancreatic Ductal Adenocarcinoma Affects the Tumor Cell Migration and Is Regulated by the p38MAPK Signaling Pathway**

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**Abstract**

The interplay between the tumor cells and the surrounding stroma creates inflammation, which promotes tumor growth and spread. The inflammation is a hallmark for pancreatic adenocarcinoma (PDAC) and is to high extent driven by IL-1α. IL-1α is expressed and secreted by the tumor cells and exerting its effect on the stroma, i.e. cancer associated fibroblasts (CAFs), which in turn produce a massive amount of inflammatory and immune regulatory factors. IL-1α induces activation of transcription factors such as nuclear factor-κB (NF-κB), but also activator protein 1 (AP-1) via the small G-protein Ras. Dysregulation of Ras pathways are common in cancer as this oncogene is the most frequently mutated in many cancers. In contrast, the signaling events leading up to the expression of IL-1α by tumor cells are not well elucidated. Our aim was to examine the signaling cascade involved in the induction of IL-1α expression in PDAC. We found p38MAPK, activated by the K-Ras signaling pathway, to be involved in the expression of IL-1α by PDAC as blocking this pathway decreased both the gene and protein expression of IL-1α. Blockage of the p38MAPK signaling in PDAC also dampened the ability of the tumor cell to induce inflammation in CAFs. In addition, the IL-1α autocrine signaling regulated the migratory capacity of PDAC cells. Taken together, the blockage of signaling pathways leading to IL-1α expression and/or neutralization of IL-1α in the PDAC microenvironment should be taken into consideration as possible treatment or complement to existing treatment of this cancer.

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**Introduction**

A highly inflammatory environment is a hallmark for the gastrointestinal malignancy pancreatic adenocarcinoma (PDAC) including a rapid progression and a 5 year survival rate of less than 5% [1,2]. A massive fibrotic stroma encloses and infiltrates the malignant cells [3] and the cellular composition of PDAC microenvironment supports the recruitment of infiltrating immune cells such as T cells, macrophages and dendritic cells (DCs) [4,5]. The CAFs play an important role in tumor progression and this is supported by the fact that many tumors fail to develop unless the stroma is modified [6] and these cellular modifications are induced in a paracrine manner by adjacent tumor cells [7,8]. Proinflammatory factors such as IL-1, TNF-α, and COX-2 induce the expression of inflammatory genes in CAFs and immune cells present in the tumor [4,9].

Inflammation is strongly connected to most types of cancer and involve activation of oncogenes and/or inactivation of tumor suppressor genes that influence the proinflammatory transcriptional programs by the malignant cells [10]. In the case for PDAC, several factors have been shown to be involved in tumor and stroma interactions including CXCL8, TGF-β and metalloproteases [11,12,13], all observed in our PDAC-CAF cross talk system [9]. The inflammation in PDAC is to high extent driven by IL-1α, expressed and secreted by the tumor cells and affecting the stroma cells, i.e. CAFs, which produce massive amount of inflammatory and immune regulatory factors both in vitro and in vivo [5,9]. The signaling event induced by IL-1α is well known and starts with IL-1 binding to and signaling through the IL-1 receptor followed by a subsequent activation of the p38 mitogen activated protein kinase (MAPK) [14]. This occur via the small G protein Ras that becomes associated with IRAK, TRAF6, and TAK-1, which facilitate the p38MAPK activation by IL-1α [15]. In contrast, until very recently the signaling events leading up to the expression of IL-1α by the tumor cells had not been elucidated. Ling et al showed for the first time involvement of the K-Ras mutation in codon 12D in induction of IL-1α expression via the transcription factor AP-1 [16]. Moreover, IL-1α activated NF-κB and its target genes IL-1α and p62 to initiate IL-1α/p62 feed forward loops,
which induced and sustained the NF-κB activity [16]. Dysregulation of Ras pathways is common in cancer as this oncogene is the most frequently mutated in human cancers and contribute to cancer cell survival [10]. Activating K-Ras mutations are present in nearly all PDACs (up to 90%) and occur very early and are the most frequent mutations in pancreatic cancer, followed by mutation or silencing of p53, p1, and DPC4/smad4 [17,18]. For pancreatic cancer, K-Ras mutations are a negative prognostic factor after surgery and adjuvant chemoradiation [19]. The mitogen activated protein kinases (extracellular signal-regulated kinase (ERK), Jun N-terminal kinase (JNK), and p38MAPK) are the best characterized signal pathways in transduction of Ras activity and their oncogenic functions are mostly based on their ability to activate AP-1 [20,21]. Ras/Raf/MAPK pathway is involved in many cellular processes such as cell cycle regulation, wound healing, cell migration, cell growth, division, and differentiation [10].

So far, there is no selective specific inhibitor of K-Ras available for routine clinical use and downstream targets such as MAPKs are then interesting targets for inhibition of K-Ras signaling, ERK, JNK, and p38MAPK are three major MAPKs and have key roles in inflammation, tissue homeostasis, proliferation, differentiation, migration and survival of cells. ERK is activated by mitogens, whereas JNK and p38MAPKs are activated by cellular stress for instance via the K-Ras signaling pathway. The p38MAPK signaling has been shown to affect proliferation, differentiation, and migration and is associated with cancers both in human and mouse [20].

The aim of this study was to examine the signaling cascade involved in the induction of IL-1α expression in PDAC. We hypothesized that tumor cells creates an inflammatory microenvironment by inducing their expression of IL-1α through downstream targets of mutated K-Ras and deciphering this could be of relevance to determine targets for treatment.

We found that the p38MAPK, activated by the K-Ras signaling pathway, to be involved in the expression of IL-1α by PDAC cells, as blocking this pathway decreased both the gene and protein expression of IL-1α. Blockage of the p38MAPK signaling in PDAC also dampened the ability of the tumor cell to induce inflammation in CAFs and CAFs ability to enhance tumor cell migration. Noteworthy, the IL-1α autocrine signaling regulated the migratory capacity in PDAC cells. Taken together, the blockage of signaling pathways leading to IL-1α expression and/or neutralization of IL-1α in the PDAC microenvironment should be taken into consideration as possible treatment or complement to existing treatment of this cancer.

Results

p38MAPK/ERK is Involved in Tumor Cell Associated IL-1α Expression

The IL-1α positive primary PDAC cell line PC013 [9] cultured in 1% FCS were exposed to 0–150 µM p38MAPK (SB203580) inhibitor, 0–50 µM ERK (U0126) inhibitor, and 0–75 µM JNK (SP600125) inhibitor for 24 h. We assessed if the inhibitors asserted any negative effect on cell viability and found no effects on the cell viability. The expression levels of IL-1α mRNA decreased after exposure to the p38MAPK inhibitor, while only a minor decrease was seen after ERK inhibitor, while the JNK inhibitor slightly induce IL-1α expression (Figure 1A–C). 100 µM of p38MAPK inhibitor SB203580 was found to be optimal to use for subsequent experiments. The mRNA expression levels of IL-1β and IL-1RA decreased after exposure to the p38MAPK inhibitor (Figure 1D–E) and had similar expression curves as IL-1α. We have previously shown that the IL-1α positive PDAC cell lines secrete IL-1α protein and this is in accordance with findings from other tumor cell lines [22]. In addition, our primary PDAC cell lines do not express the IL-1β protein only the mRNA [9] and therefore did we only examining the IL-1α protein expression and found that it was significantly decreased after 24 h (p<0.005), 48 h (p<0.005), and 72 h (p<0.001) treatment with p38MAPK inhibitor compared to vehicle treated cells (Figure 1F). We had to use high concentration of SB305380 to block IL-1α and as this inhibitor is known to also work on other MAPKs at high concentration did we confirm the role of p38MAPK in the regulation of tumor cell associated IL-1α by using SB220025, which is considered to be a very specific inhibitor of p38 MAPK than SB203580. This inhibitor significantly reduced the mRNA expression of IL-1α (P = 0.003) and IL-1β (P = 0.003), but did not affect the expression levels of IL-1RA (Figure 1G–I). In addition, we found that p38 MAPK and ERK was phosphorylated in our primary PDAC cell lines PC013, PC065 and that the inhibition of p38 MAPK with SB220025 did not reduce the phosphorylation of this MAPK, whereas the phosphorylation of ERK was reduced by the ERK inhibitor (data not shown). The phosphorylation of p38 MAPK is in accordance with previous findings for pancreatic cancer cell lines [23,24], which indicate that this pathway is constitutively activated in PDAC tumor cells.

The p38MAPK Inhibition had Different Effects on PDACs and CAFs Inflammatory Profiles

The p38MAPK inhibitors had in our study significantly inhibitory effects on PDAC associated IL-1α expression and the p38MAPK pathway have previously been found to be involved in the regulation of chronic inflammation [20]. To examine the regulatory role of p38MAPK on other inflammatory factors we investigated the direct effects of SB203580 on PC013 cells and CAFs. The results for the primary PDAC cell line PC013 showed significant reduced levels of IL-1α (P<0.001), IL-1β (P<0.001), IL-1RA (P = 0.001), and CXCL1 (P = 0.04), whereas CCL20, and TNF-α levels were decreased but not significant (Figure 2A). Levels of VEGFA, CXCL3 and COX-2 increased but only VEGFA to a significant level (P = 0.01) (Figure 2A). Blockage of p38MAPK pathway in CAFs for 24 h resulted in increased mRNA expression levels of CXCL2 (P = 0.02), CXCL3 (P = 0.01) and CXCL13 (P = 0.05) in CAFs (Figure 2B). Furthermore, the CAF gene expression levels of CXCL5 (P = 0.024), CXCL6 (P<0.001), CCL20 (P = 0.03), VEGFA (P = 0.02), COX-2 (P<0.001), IL-6 (P<0.001), and IL-24 (P<0.001) decreased after blockage of p38MAPK (Figure 2B). This clearly shows that the p38MAPK signaling regulates the expression of inflammatory factors differently in PDAC cells compared to CAFs.

Tumor Cell p38MAPK Signaling and IL-1 Regulation Affects the Tumor-CAF Crosstalk

IL-1α has previously been established as an important factor involved in the communication between tumor cells and CAFs in PDAC [9]. Our findings of an inhibiting role of p38MAPK inhibitors on PDAC tumor cell IL-1α expression could have the potential to affect the crosstalk between tumor cells and CAFs. We examined the inflammatory profile of CAFs conditioned with supernatants derived from PC013 first pretreated with p38MAPK inhibitor for 72 h than recultured 48 h without inhibitor. The conditioned CAFs showed significantly decreased gene levels of
CXCL1 (P = 0.01), CXCL2 (P = 0.001), CXCL3 (P = 0.012), CXCL5 (P = 0.023), CXCL8 (P = 0.004), CCL20 (P = 0.001), COX-2 (P = 0.004), IL-6 (P < 0.001), and IL-24 (P = 0.002) (Figure 3A) compared to supernatants from vehicle treated PC013 cells. To relate these findings in CAFs to tumor cell associated IL-1α, as a downstream result of p38MAPK signaling, we analyzed the IL-1α levels in the PC013 supernatants 2 days after the exposure to p38MAPK inhibitor and found significantly reduced levels of IL-1α (P = 0.038) (Figure 3B). Next was the concentration of IL-1α in the p38MAPK pretreated PC013 derived supernatants returned to original levels by addition of exogenous rhIL-1α and used to treat CAFs for 48 h. The CAF gene profile showed increased levels of inflammatory factors (Figure 3C). These results indicate an important role for p38MAPK in regulating PDAC cell-CAF crosstalk through the induction/upregulation of IL-1α expression in PDAC.
** = P

CXCL8, CCL20, VEGFA, IL6, COX-2, TNF, IL-24, IL-1 that acted on PDAC and CAFs by the Autocrine and Paracrine Effects Exerted by IL-1

The migratory properties of the tumor cells is enhanced by the autocrine and paracrine effects exerted by IL-1α on PDAC and CAFs

We further investigated if the decreased level of inflammatory factors in CAFs, treated with supernatants derived from p38MAPK signaling inhibited PC013 cells, affects the ability of the tumor cells to migrate. PC013 cells exposed to supernatants derived from CAFs cocultured with PC013 cells pretreated with p38MAPK (IL-1α low) had significantly decreased migration compared to cells exposed to supernatants derived from CAFs cocultured with untreated PC013 cells (P = 0.0038) (Figure 4A). The decreased levels of IL-1α in PC013 cells exposed to p38MAPK inhibitor should reduce IL-1α autocrine feedback and modulate the functions such as tumor cell mobility. We investigated this by neutralized IL-1α with rhIL-1RA and found a significantly reduced migration of PC013 cells (P = 0.002) (Figure 4B). To in depth elucidate the effect of IL-1α on tumor cell migration we used PC077, a primary PDAC cell line that is IL-1α negative [9]. In addition, rhIL-1RA was added to all groups in this experiment to eliminate any direct effects of IL-1α on the tumor cell migration. Supernatants from untreated CAFs (P = 0.025) significantly enhanced the migration of IL-1RA treated PC077 cells, while supernatants from exogenous IL-1α activated CAFs even further increased migration of PC077 cells compared to both IL-1RA/IL-1α (P = 0.009) treated, IL-1α treated, and untreated CAF supernatants (P = 0.030) (Figure 4C). Moreover, no difference was found between rhIL-1RA and IL-1α/rhIL-1α treated PC077 cells. Taken together, this indicates that IL-1α not only induce autocrine direct effects on cancer cell migration, but also enhance migration through paracrine signaling and activation of CAFs that obtain the ability to stimulate tumor cell migration by altering the tumor cell phenotype, including increased expression of metalloproteases and factors involved in epithelial mesenchymal transition (work in progress).

rhIL-1α in Combination with p38MAPK Inhibitor Effectively Reduced the Levels of Inflammatory Factors Induced in CAFs by PDAC Cells

IL-1α has previously been shown to decrease the levels of inflammatory factors in both single and CAFs cocultured with PDAC cells [9]. Here we examined the effects treatment with IL-1α and p38MAPK inhibitor alone or as a combination therapy had on PC013 and CAF cocultures. Blockage of p38MAPK signaling significantly decreased the levels of CXCL1, CXCL5, CXCL6, IL-6, and IL-24 (P<0.001), CXCL2, and CXCL20 (P = 0.002), CXCL3 (P = 0.003), CXCL5 (P = 0.004), and VEGFA (P = 0.009), (Figure 5) in CAFs cocultured with PDAC cells compared to untreated controls (2–29 fold decrease) (Table 1). Neutralization of IL-1α by rhIL-1RA as a single agent drastically reduced the levels of all the inflammatory factors (2–4076 fold) compared to untreated cocultured CAFs (CXCL1, CXCL5, CXCL6, CXCL20, IL-6, IL-24 and COX-2 (P<0.001), CXCL2, CXCL3, and CXCL6 (P = 0.002), and VEGFA (P = 0.02) (Figure 5 and Table 1). The combination of p38MAPK inhibitor and rhIL-1α decreased the inflammation 4.5–7895 folds compared to untreated controls (CXCL1, CXCL2, CXCL5, CXCL6, CXCL20, COX-2, IL-6, and IL-24 (P<0.001), CXCL3, CXCL5, and VEGFA (P = 0.02) (Figure 5 and Table 1). Moreover, the combination therapy decreased the levels of the inflammatory factors compared to rhIL-1α treated cocultured CAFs (CXCL1, CXCL2, CXCL6 (P<0.001), CXCL3 (P = 0.06), CXCL5 (P = 0.003), CXCL8 (P = 0.04), CCL20 (P = 0.007), VEGFA (P = 0.02), IL-6 (P = 0.009) and IL-24 (P = 0.003)) (Figure 5).

**Discussion**

We have demonstrated that the p38MAPK signaling pathway is involved in the expression of IL-1α by PDAC cells and that IL-1α initiates a change in the PDAC phenotype and properties, making the tumor cells more prone to migrate. The p38MAPK signaling in tumor cells were shown to be involved in the upregulation of inflammation in CAFs via the induction of IL-1α and to our knowledge has this not been shown previously. Our previous finding that IL-1α overexpression correlated with poor survival in PDAC patients was confirmed in the study by Ling et al [16]. These findings suggest that inhibition of IL-1α expression and activity in PDAC will drastically decrease the levels of inflammatory factors in CAFs, and reducing and/or neutralizing the effects of IL-1α could have the potential to reduce tumor spread and improve the clinical outcome for the patients [5,9].

The signaling pathways/events giving rise to constitutive expression of IL-1α have been elusive. A recent study by Ling et al [16] using a mouse model where mutation in K-Ras G12D was used to induce PDAC showed that this activating K-Ras
mutation induced activation of NF-kB, which was required for PDAC development, and expression of IL-1\(\alpha\) in the tumor cells and that this create a intrinsic inflammatory response that promote a pro-tumorigenic microenvironment through the expression of inflammatory mediators, e.g. cytokines such as IL-1\(\alpha\) [16]. The proposed mechanism for NF-kB activation by K-Ras is through AP-1 induced IL-1\(\alpha\) overexpression [16]. The expression of IL-1\(\alpha\) by tumor cells is detected in 90% of the PDAC patients [9] and could correlate to mutations in the oncogene K-Ras as they also are present in up to 90% of the PDAC cases [25,26,27]. These findings clearly demonstrate the role of inflammation in the development of PDAC. As all attempts of developing a drug that directly blocks the mutated K-Ras oncogene has been unsuccessful [28], did we target downstream factors activated by K-Ras. The signaling cascades through K-Ras lead to many different events inside the cell and several pathways, i.e. RAS/RAF/MAPK RAS/P13K/AKT, are well characterized [29]. In response to cellular stress and cytokine stimulation mediated through K-Ras do p38MAPK kinases (MKK3 and MKK6) and JNK kinases (MKK4 and MKK7) phosphorylate p38MAPK and JNK, respectively [15].

PDAC have a very elevated expression of multiple inflammatory genes and the main cell in the tumor stroma, i.e. CAFs, are the major producers of these factors [5,9]. According to our own data and the study by Ling et al [16], IL-1\(\alpha\) is the pro-inflamatory factor responsible for inducing the expression of the inflammation. We found that the IL-1\(\alpha\) gene and protein expression by primary PDAC cell lines was down regulated only when the signaling through p38MAPK was blocked. This clearly show that signaling trough activated p38MAPK highly influence the IL-1\(\alpha\) expression and also eliminates ERK and JNK as contributors to the regulation of IL-1\(\alpha\) in PDAC cells. In addition, the inhibition of p38MAPK in PDAC lead to a modulated gene profile for several
inflammatory genes, some factors decreased whereas others increased.

Interfering with the p38MAPK signaling pathway also directly affected the inflammatory profile of primary CAFs, but the profile differed from the one induced in PDAC, explained by other pathways affected in CAFs compared to PDAC cells. The CAFs play an important role in tumor progression and this is supported by that many tumors fail to develop unless the stroma is modified and these changes are induced in a paracrine manner by adjacent tumor cells [7,8]. The paracrine action of IL-1α, expressed by the tumor cells, seems to be important in the case for developing PDAC [16]. Both soluble and cellular factors are involved in tumor and stroma interactions, including CXCL8, TGFβ, and metalloproteases [9,11,12,13]. The crosstalk between PDAC cells and CAFs induce a high level of inflammation and it was significantly diminished when the PDAC had been pretreated with p38MAPK inhibitors. The down regulation of the inflammatory profile in CAFs was due to the reduced levels of IL-1α expressed by the PDAC cells as replenishing this proinflammatory factor restored the inflammatory responses.

The expression of IL-1α by PDAC affects the tumor cell function, making them more prone to migrate as neutralizing of IL-1α with IL-1RA decreased the migratory capacity. In addition to the autocrine effects of IL-1α on tumor cell migration, the protein also acts in a paracrine manner by promoting a CAF phenotype supporting the tumor cell migration. Induction of IL-1α expression in PDAC cell lines enhance their ability to form metastasis and invade tissue in orthotopic mouse models and in vitro experimental systems [2,30]. Similar effects with increased ability to metastasize has also been described for tumor cells expressing IL-1β [31,32].

The PDAC creates a microenvironment that makes this tumor hard to treat and reducing the inflammation could be one way to improve this. Down regulation of IL-1α expression by p38MAPK in combination with IL-1RA more or less abolished the inflammation in PDAC and CAFs cocultures. Consequently, the inhibition of signaling pathways leading to IL-1α expression and/or neutralization of IL-1α in the PDAC microenvironment should be taken into consideration as a possible treatment or as a complement to existing treatments of this cancer.

Materials and Methods

Ethical Statement

The primary PDAC cell lines PC013 and PC077, and primary CAFs were propagated from PDAC tumor tissue biopsies as described elsewhere [9]. The study protocol and patient consent documents were approved by the Regional Ethics Committee in Linköping, Sweden (Dnr. M38-06). The consent was written and obtained from all participants involved in the study.

Table 1. CAF inflammatory profile after single and combination therapy with rhIL-1RA and p38MAPK inhibitor SB220025.

|       | CXCL1 | CXCL2 | CXCL3 | CXCL5 | CXCL6 | CXCL8 | CCL20 | VEGFA | COX-2 | IL-6 | IL-24 |
|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|------|-------|
| SB220025 | 4     | 9     | 14    | 8     | 4     | 5     | 2     | 2     | 5     | 11   | 29    |
| rhIL-1RA | 1016  | 159   | 564   | 22    | 412   | 4876  | 144   | 2     | 29    | 26   | 79    |
| SB220025/rhIL-1RA | 3214  | 1131  | 1430  | 684   | 1236  | 7892  | 695   | 5     | 28    | 325  | 612   |

The data is presented as fold decrease compared to untreated control CAF.

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The primary PDAC cell lines PC013 and PC077, and primary CAFs were cultured in RPMI 1640 (Fisher Scientific, Pittsburgh, PA), supplemented with 1% FCS (Invitrogen), 2 mM HEPES (Invitrogen), 30 μg/ml Gentamycin (Invitrogen) (R10). For all assays, at 6×10^5 PC013 and PC077 cells were seeded per well and cultured for 24 h before adding the different inhibitors.
Culturing PDAC Cells and CAFs with p38MAPK, ERK and JNK Inhibitors

PC013 cells were cultured in R10 containing vehicle (DMSO), 0.1–150 μM p38MAPK inhibitor SB203580 (Cayman Chemicals, US), p38MAPK inhibitor SB220025, ERK inhibitor U0126, or JNK inhibitor SP2600125 (SigmaAldrich, Sweden) for 24–72 h. In other sets of experiments, PC013 cells were cultured with 100 μM p38MAPK inhibitor SB203580 or vehicle (DMSO) for 72 h, washed 3 times and recultured in 2 ml R10 for 48 h. The 48 h supernatants were transferred and used to culture CAFs for 72 h. The PC013-SB203580-CAF (C20) and PC013-vehicle-CAF (C30) conditioned medium were normalized to number of cells/group (1×10⁷ cells/ml medium) to adjust for differences in the total amount of cells. To confirm the role of IL-1α in tumor cell-CAF cross-talk, the levels of IL-1α was measured in the supernatants from SB203580 and vehicle treated PC013 cells and equalized with rhIL-1α (R&D Systems, UK). The IL-1α normalized medium was added to CAFs for 72 h. The cells were lysed with RLT buffer (Qiagen) and RNA prepared as described elsewhere [9].

Treatment of PDAC and CAFs with p38MAPK Inhibitor and IL-1RA

The effects of p38 inhibitor (SB220025) and IL-1RA on tumor cell/CAF cross-talk were investigated and PC013 cells and CAFs were cocultured in inserts (0.4 μm) (BD, USA) for 5 days in R10 medium supplemented with rhIL-1RA (10 μg/ml) (Kineret 100 mg, Biovitrum AB) and/or 1 μM SB220025.

RNA Extraction and Quantification

Total RNA purification and cDNA was prepared as previously described [9]. Quantitative PCR was performed with Fast SYBR Green Master Mix (Version 9/2007; Applied Biosystems, Foster City, CA) on 7900 Fast Real-Time PCR system with 7900 System SDS 2.3 Software (Applied Biosystems). In the negative controls cDNA were replaced by distilled water. Specific primers for CXCL8, CCL20, IL-1α, IL-1β, IL-6, IL-24, IL-1RA, TGFB1, VEGF-A (CyberGene AB), and COX-2 (Invitrogen) were used. Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (CyberGene AB) and actin were utilized as housekeeping genes. The primers were designed using Primer Express (Applied Biosystems) if not otherwise indicated. Real-time PCRs for the detection of CXCL chemokines and TNFα were performed using TaqMan® Gene Expression Assays (Applied Biosystems) according to the manufacturer’s protocol. All reactions were performed in triplicates including none-template controls and endogenous control probes. FAM conjugated, gene specific assays were

\[
\text{Hs00236937}\_\text{m1 (CXCL1), Hs00236966}\_\text{m1 (CXCL2), Hs00171061}\_\text{m1 (CXCL3), Hs00171085}\_\text{m1 (CXCL5), Hs00237017}\_\text{m1 (CXCL6), and Hs99999043}\_\text{m1 (TNFα).}
\]

The results were analyzed using the ΔΔCt method [33] and presented as either normalized data or as relative gene expression.

ELISA

Supernatants were collected and cells harvested and counted before lysis with lysis buffer pH 7.5. The lysates and supernatants were analyzed for the concentration of IL-1α by ELISA (Nordic Biosite, Sweden) according to the manufacturers’ protocols. This ELISA measures precursor, secreted, and membrane-associated forms of IL-1α. The levels of IL-1α are presented as pg/1×10⁷ cells.

Migration Assay

Transwell migration assay were conducted as described elsewhere [34]. Briefly, PC077 and PC013 were starved in 1% FBS overnight and seeded on 6-well plates Transwell filters (Costar) (8 µm pore size) precoated with 10 μg/ml fibronectin (Sigma, St. Louis, MO). PC077 cells were incubated for 40 h in 1% FBS medium containing 100 ng/ml IL-1α and either 500 pg/ml IL-1α, CAF supernatant (7 days), or IL-1α (500 pg/ml) activated CAF supernatant (7 days). PC013 cells were incubated for 40 h in 1% FBS and/or 100 ng/ml IL-1α, C20, and C30 conditioned medium. The upper compartment was removed and the medium discarded. The adherent cells were washed in PBS, fixed in 4% formaldehyde and visualized using crystal violet (SigmaAldrich, Sweden). The sample identification was blinded and all attached cells were manually counted using 10× magnifications in an inverted microscope (Leica). The data was obtained from 3 experiments (mean ± standard error) and presented as the mean cells/cm².

Statistical Analysis

The statistical analysis was performed with GraphPad Prism 5 (GraphPad Software). P<0.05 was considered statistically significant and error bars throughout indicate standard error of the mean (SEM). The data were analyzed using the paired t-test and unpaired t-test was used for normalized data.

Author Contributions

Conceived and designed the experiments: VT ML. Performed the experiments: VT ML. Performed the analysis: VT ML LB. Wrote the paper: ML VT DM PS AS CB.

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