REVIEW

Preprocessing of 2-Dimensional Gel Electrophoresis Images Applied to Proteomic Analysis: A Review

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Abstract Various methods and specialized software programs are available for processing two-dimensional gel electrophoresis (2-DGE) images. However, due to the anomalies present in these images, a reliable, automated, and highly reproducible system for 2-DGE image analysis has still not been achieved. The most common anomalies found in 2-DGE images include vertical and horizontal streaking, fuzzy spots, and background noise, which greatly complicate computational analysis. In this paper, we review the preprocessing techniques applied to 2-DGE images for noise reduction, intensity normalization, and background correction. We also present a quantitative comparison of non-linear filtering techniques applied to synthetic gel images, through analyzing the performance of the filters under specific conditions. Synthetic proteins were modeled into a two-dimensional Gaussian distribution with adjustable parameters for changing the size, intensity, and degradation. Three types of noise were added to the images: Gaussian, Rayleigh, and exponential, with signal-to-noise ratios (SNRs) ranging 8–20 decibels (dB). We compared the performance

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Introduction

Proteomics is the analysis of the complete set of proteins (i.e., the proteome) produced in a cell, tissue, or organism, at a given time. A number of different aspects of protein analysis are covered in proteomics, including the analysis of protein expression. The presence or absence of proteins, and the direct measurement of relative protein abundances can help understand cellular processes, and may be useful for identifying drug targets and diagnostic/prognostic markers [1]. As such, it is one of the most active fields of biological research, given its wide range of applications.

One of the most commonly used techniques in comparative proteomic studies is two-dimensional gel electrophoresis (2-DGE) [2], a technique capable of resolving thousands of proteins in a single run. Using 2-DGE, proteins are firstly separated according to their isoelectric points (pI) in one dimension, and then, according to their molecular weights in a second dimension [3]. Afterward, the 2-DGE gels are stained for protein visualization and analyzed with computer-assisted image evaluation systems for a comprehensive qualitative and quantitative examination of the proteomes [4].

2-DGE was initially employed for protein separation and analysis approximately in 1975 [5]. Even though it was introduced more than four decades ago, nowadays 2-DGE is still widely used for whole proteome analysis [6,7], comparative analysis of proteome changes [6,8], biomarker discovery, cancer research [9,10], as well as for the identification of protein isoforms and post-translational modifications [11], among other purposes. Its popularity could be attributed to several factors. It is possible to resolve and visualize thousands of proteins in a single gel as mentioned above. In addition, 2DGE is very affordable compared to other techniques, which does not require complex laboratory equipment but offers reliable and reproducible results. 2GDE is optimal for cases when multiple analyses are required on replicate samples. More importantly, it is compatible with mass spectrometry and other downstream analyses. Therefore, 2GDE is often used as one of the first steps in describing the protein composition of a particular sample type, at a certain time point, and under a particular set of conditions.

Many computational applications are available for processing and analyzing 2-DGE images, such as MELANIE, PDQuest, Z3, Progenesis Workstation, ProteomeWeaver, ProteinMine, Delta2D, and DeCyder [12,13]. Given that one cell can express around ten thousand proteins, 2-DGE needs an effective computational tool that can process large volumes of information [14]. It is often necessary to apply appropriate preprocessing techniques to 2-DGE images, as the ultimate performance of these analysis tools strongly depends on the quality of the images to be processed [4,15].

For 2-DGE image analysis, techniques are required to detect protein spots, to segment and to quantify the protein expression level based on the number of pixels [15,16]. An additional step of image alignment is performed in order to match the corresponding protein spots from different images [17]. However, due to technical difficulties inherent to 2-DGE, anomalies are often found in gel images, such as noise around protein spots, vertical and horizontal streaking, saturation of certain protein spots, presence of very faint protein spots, as well as non-linear intensity of protein spots [14,18]. Therefore, 2-DGE image analysis is frequently perturbed by the presence of different types and levels of noise. For instance, the impulsive noise can spectrally interfere in all frequencies when the sample is digitalized. The background of 2-DGE images can also vary among samples, depending on the technical specifications of the imaging system used to capture and digitalize the images [15]. These variations and anomalies often complicate the analysis of 2-DGE images and affect the reproducibility of the results obtained [19,20]. A preprocessing phase could be effective in reducing or eliminating these anomalies, thus reducing errors in the subsequent spot detection. Omitting this preprocessing stage may profoundly affect the results of the subsequent analysis, as noise could be falsely identified as a protein, whereas real proteins of interest could be missed [21]. Therefore, it is important to review the advances in 2-DGE image preprocessing, in order to identify areas of improvement that may be of great interest to researchers in the field.

There are three common objectives for the 2-DGE image preprocessing, i.e., pixel intensity regularization (image normalization), background correction, and noise reduction (filtration) [15]. In the subsequent sections, we describe in detail the preprocessing strategies to achieve these objectives, their limitations, and recent advances. We also compare the most representative noise reduction techniques using synthetic gel images for a quantitative evaluation. Finally, we conclude with recommendations to be considered for successful automatic 2-DGE image processing.

Preprocessing of 2-DGE images

2-DGE images inevitably exhibit anomalies caused by sample preparation techniques and the imaging system used to acquire the digital images [22]. The most common anomalies in 2-DGE images are oversaturated, faint, or fuzzy spots, vertical and horizontal streaking, overlapping spots, and noise [13,23].
Figure 1 shows a 2-DGE image with the most common anomalies. The image was obtained in our laboratory from prostate tissue, the challenges of image preprocessing, and thus, the requirement for robust algorithms in order to achieve proper preprocessing are apparent.

Image normalization

Intensity normalization of 2-DGE images aims to normalize the intensity of protein spots to facilitate processing. The spot intensities may vary due to differences in the exposure of the gel during the staining process and data saturation, among other reasons [15]. As a step of image processing [24], intensity normalization is also known as a preprocessing technique in various literatures [25]. In fact, some researchers recommend eliminating saturated spots before processing in order to achieve the optimal results for the identification of proteins in the gel [14,26].

To avoid saturation caused by high protein concentrations, it is necessary to make sure that most of the data are represented by levels slightly below the maximum possible intensity. Highly abundant proteins often mask fainter spots that represent proteins with low abundance. Usually, these low-abundance proteins are also relevant, but pose difficulties in accurate quantification, if the spots are too faint [27].

The normalization process helps to reduce noise, which distorts the detection of protein spots in 2-DGE images according to criteria of statistical significance, thus allowing the outline of the spots of interest to be delineated [28]. However, in some circumstances, the intensity of pixels does not present a clear tendency. This type of standardization is based on information from various replicate gels, generating a statistical average of the intensity of each pixel. With these statistical averages, the intensities of the entire image are normalized, using normalization algorithms such as Cyclic Loess, Contrast, Cyclic Linear, and Quantile [28]. Image regularization with multiple replicate gels has yielded satisfactory results, since changes in patterns of protein expression among samples are small compared to the overall pattern of protein spots. The major limitation of this approach lies in obtaining and aligning a representative number of gel images, which may result in the unwanted removal of faint protein spots [15]. With image regularization, it is possible to minimize distortions present in the images; however, low-abundance proteins are often eliminated in the process. Therefore, it is necessary to apply discriminatory algorithms to these regularization techniques, so that potentially faint but important protein spots would not be removed during image regularization.

Background correction

The aim of 2-DGE image processing is to detect spots, and define the location, border, and intensity of each spot. However, in some conditions, defining the limits of the spot border is difficult due to the varied background, noise, and saturation of protein spots [14,29,30]. Background correction is thus necessary, as the background signal of 2-DGE images is not uniform and varies locally according to protein intensities. The greater the intensity of a protein spot, the greater the noise signal around it, which introduces errors in detection. In addition, anomalies such as vertical streaking must be removed from the image, as protein spots contained within these streaks are not identifiable or quantifiable [15,31].

Three background correction techniques are available to reduce the influences of these phenomena. These include adjustment by polynomials, local and global minima, and histograms.

Background correction with polynomial adjustment has been widely accepted in the scientific community [32]. This technique seeks smoothing the image background, removing noise from overlapping spots, and minimizing the effects of intensity degradation. The biggest limitation of background correction is the selection of a background sample for the adjustment. A poor selection of the background sample can generate significant image distortion, or even lead to the improper softening of background texture [33]. Local and global minima are the simplest technique for background correction, making it possible to effectively mitigate noise in the flatter areas (without significant changes in pixels) and smooth discontinuities. However, a global threshold is not sufficient to detect background variations [15].

For background adjustment using histograms, the distribution of intensities in an image is used to identify the background. The maximum peak in the intensity distribution is the intensity of background pixels. The challenge of this technique is to establish the intensity threshold of the background that is usually set by an optimization process [34]. The selection of parameters is a significant drawback for users with little knowledge of optimization techniques, considering that the model implemented to describe the background may be insufficient or cause a significant loss of faint but important protein spots [35]. Although studies on the implementation of background correction techniques for 2-DGE images have lost momentum, a variety of techniques [36] in this field are yet to be explored. In addition, new techniques have also been developed that are efficient for the analysis of images with features similar to those of 2-DGE images, such as combination of Gaussian mixture models and machine learning approach [37], which may be promising options for 2-DGE image processing.
Noise reduction

2-DGE image filtering is the most commonly used approach within the scientific community, with the most diversified techniques [15]. These filtering techniques can be classified into two broad categories: linear and nonlinear filters.

The first techniques reported for 2-DGE image preprocessing were based on linear filters, which are the most widely used techniques in commercial software programs. These kinds of filters compute the output pixel as a linear relation of the surrounding pixels [38]. In general, linear filters tend to blur images, affecting the edges, lines, and fine details of the protein spots [39]. While linear filtering techniques, such as the Gaussian filters, have an acceptable performance under general conditions, they are also likely to reduce the density of protein spots by smoothing the contour [40].

Orthogonal regression methods have been used as a simple and efficient option that can adapt to a linear model [41]. However, the edges of protein spots of interest in 2-DGE images are often distorted during filtering [25,42]. The linear filters based on orthogonal regression techniques, which are most commonly used for noise reduction in 2-DGE images, are based on principal component analysis (PCA) and Wiener filters. PCA is a robust orthogonal regression technique, which shows a great advantage in terms of spot edge maintenance. However, small details, such as low-abundance protein spots, can be removed in the process [43,44].

With the Wiener filter, it is possible to obtain satisfactory results with a low level of distortion of the image, but the noise reduction is compromised [38]. When taking into account the overall advantages of noise removal, a certain level of signal degradation is generally acceptable. However, the noise reduction results in the blurred image with poor edge definition [15]. In general, these methods allow the user to select the number of times the smoothing filter is applied, and the size of the convolution window to adjust the level of smoothing [45].

Nonlinear filters have fewer limitations, but generally increase the degree of complexity and the need to define parameters more thoroughly. Some nonlinear filtering techniques used in 2-DGE images include contourlet transform [46], total variation [47], and wavelet transform [19]. The contourlet transform performs a decomposition of the data in frequency and space by means of two decomposition methods [46]. These methods allow a multiple scale of space and frequency, as well as a high degree of directionality. Satisfactory results have been achieved using these methods, in addressing the low signal-to-noise ratio [48], and loss of relevant information, thereby surpassing the wavelet transform [48].

The total variation technique, proposed by Chan [49] and Xin [50], is very advantageous in terms of the extraction of geometric features and in terms of the preservation of edges. Using this technique, different degrees of smoothing on different points can be achieved, which exemplifies its anisotropy. Although able to capture all the directional information, this technique leads to staircase effects in the noise removal process [51]. Therefore, this technique does not perform as well as the wavelet transform technique [41,50]. Wavelet transform is the most commonly used technique for noise reduction, with a high percentage of filtering and acceptable level of loss of relevant information [19,43,52–57]. This method has become increasingly popular in the preprocessing of 2-DGE images since 2004, as reported in the study by Kaczmarek and others [19].

Despite the simplicity of noise suppression methods based on spatial filtering, the image tends to be severely distorted after processing. As these types of images have local features with high variation, it is almost impossible to discriminate between information of interest and noise, when the processing considered only the spatial or spectral domain [41]. For this reason, it is necessary to use a filter with a joint time–frequency domain [57] that exceeds spatial filtering, in both signal/noise ratio and distortion. Using the wavelet transform method, the image can be decomposed into space-frequency components and evaluated point by point. It is also possible to combine several images in order to obtain an average value and thus eliminate noise more effectively [56]. For a correct operation, it is necessary to determine the scale of the wavelet transform. However, it is difficult to properly define the contours, resulting in a significant distortion of the information of interest [50].

Different derivations of the wavelet transform [58,59] are available to deal with its limitations. For instance, Barlaud and colleagues [60] presented a very useful technique called Wavelet Transform Quincunx, to obtain a multiresolution scale, thereby improving the performance around the edges and enhancing filtering effect [41]. A hybrid technique, WTTV, combines the power of wavelets with the advantages of Total Variation (TV) [50], to reduce the loss of relevant information and increases the level of filtering. The wavelet transform can be applied first, followed by a total variation routine, thus minimizing distortions caused by the wavelet transform. In this case, it is necessary to carefully define the parameters for both components, since false information is generally introduced due to limitations of the directional information. As a result, the geometric captures can be affected, creating a distortion of the borders around the spots of interest [46].

Due to the limitations of these techniques and the need for parameter adjustments, advanced techniques such as genetic algorithms [52] have been applied. However, it is noteworthy that this entails an increase in computational complexity. Although methods are available to minimize the computational cost of genetic algorithms, these add a higher level of complexity to the whole process [61].

Validation measures for processing 2-DGE images

The lack of performance indicators with which the various techniques can be fairly compared greatly hinders the selection of preprocessing techniques. In fact, performance comparison of many commonly used techniques has not yet been reported in the literature [15]. The most commonly used indicator is the signal-to-noise ratio (SNR) obtained from the mean square error (MSE):

\[
\text{MSE}_n = \frac{\sum_{i=1}^{n} (x_i - \hat{x}_i)^2}{\sum_{i=1}^{n} (x_i)^2}
\]

(1)

\[
\text{SNR} = 10 \times \log_{10} \frac{1}{\text{MSE}_n}
\]

(2)

However, with the SNR indicator, important factors regarding the retention of relevant information are not considered. Daszykowski thus proposed a measure called false discovery rate (FDR), defined as a ratio of the number of
stable variables from the permuted data to the number of stable variables from the original dataset [25]. While this measure is focused on the object detection process, it does not list the spots of interest that were removed during the preprocessing routine. FDR can be estimated by:

$$FDR = \frac{N_1}{N_2 \times 100} \quad (3)$$

where $N_1$ is the number of falsely identified significant features in the permuted data for a given threshold value and $N_2$ is the number of significant features in the experimental data set for the same threshold.

Another measure is spot efficiency [41] as defined below, which establishes a relationship between the false spots that were generated or prevailed, and spots that were lost with the technique.

$$\text{Spot efficiency} = \frac{\text{Number of spots detected} - \text{Number of false spots}}{\text{Number of spots detected} + \text{Number of lost spots}} \quad (4)$$

Quantitative comparison of noise reduction methods

Comparison of synthetic images

The performance of several noise reduction techniques was compared using synthetic images. Experiments were developed under controlled conditions and the quantitative indicators SNR and spot efficiency were used to determine the performance of the filtering techniques with a perturbation of impulsive-type noise. To generate a cloud of spots, we used a model of uniform distribution, which simulates proteins in 2-D gels, and allowed overlapping between spots. The synthetic spot model is a Gaussian distribution [62], with adjustable parameters for size, intensity, and degradation (see Figure 2A). We used a 512 x 512 synthetic image with 250 protein spots. For each protein, the standard deviation was randomly set between 0.3 and 0.8, and its maximum intensity was constrained between 0.6 and 1. Different types of noise were added to the synthetic images to examine how the change of distribution affects the behavior of the techniques being evaluated. The noise distributions applied to the images are: Gaussian noise, Rayleigh type noise, and exponential type noise [63]. Table 1 presents the parameters used for each noise type to simulate images with SNR between 8 and 20 dB. An example of a synthetic image with noise is presented in Figure 2B.

To obtain images with a blurring effect, a linear approximation of the camera movements in pixels ($N$) was performed, using image processing techniques with a filter of displacement vector [64]. We used synthetic images with linear displacement from 5 to 30 pixels and a representative synthetic image with blurring effect is shown in Figure 2C. In addition, we also generated images with distortions between 5% and 30% in the background to represent a non-uniform illumination system, as exemplified by the image in Figure 2D.

Figure 2 Synthetic protein spots modeled as a 2-D Gaussian distribution
A. Example of synthetic image. B. Synthetic image with noise obtained by adding random values with Gaussian distribution. C. Synthetic image with blurring using a filter of displacement of 5 pixels. D. Synthetic image with distortion that change background intensity until 5%.
The SNR and spot efficiency for the images with noise and filtered with nonlinear techniques wavelet, contourlet, TV, and WVTV are presented in Table 2 and Table 3, respectively. The technique with the best performance for each noise distribution, variance, and indicator is highlighted. As expected, the performance of each of these techniques decreases with the increasing noise level. In terms of SNR, wavelet is the best technique for images with Gaussian noise (Table 2). For images with Rayleigh and exponential noise, wavelet presents the best performance for low-level noise. Conversely, TV was better for high level of Rayleigh and exponential noise (8–12 dB).

In terms of spot efficiency, wavelet and WVTV techniques have a lower sensitivity to noise than the contourlet and TV techniques, as reflected by the best performance of wavelet and WVTV techniques. Wavelet was the best for images with noise levels between 12 and 20 dB, and WVTV was best for images with high-level noise. Figure 3 presents an example of filtering results obtained for the image with Gaussian noise equal to 10 dB.

### Table 1 Parameters for Gaussian, Rayleigh and exponential noise of synthetic images

| Distribution | Parameter | Noise intensity (dB) |
|--------------|-----------|----------------------|
|              |           | 20       | 18       | 16       | 14       | 12       | 10       | 8        |
| Gaussian     | $a$       | 0.0766   | 0.0966   | 0.1214   | 0.1530   | 0.1929   | 0.2430   | 0.3091   |
| Rayleigh     | $b$       | 0.0277   | 0.0398   | 0.0539   | 0.0725   | 0.0980   | 0.1335   | 0.1858   |
| Exponential  | $a$       | 14.0361  | 10.0361  | 8.3389   | 6.8749   | 5.6862   | 4.6894   | 3.8549   |

Note: $a$, standard deviation for Gaussian distribution with median $\mu = 0$; $b$, scale for Rayleigh distribution with shifting parameter $a = 0$; $a$, scale parameter for exponential distribution.

### Table 2 Performance of noise reduction techniques evaluated using SNR

| Noise type | Noise reduction technique | Noise intensity (dB) |
|------------|--------------------------|----------------------|
|            |                          | 20       | 18       | 16       | 14       | 12       | 10       | 8        |
| Gaussian   | Wavelet                  | 27.61    | 26.70    | 25.55    | 24.24    | 22.82    | 24.15    | 19.29    |
|            | Contourlet               | 25.80    | 24.08    | 22.36    | 20.79    | 19.41    | 18.26    | 17.33    |
|            | TV                       | 26.29    | 24.48    | 22.64    | 20.79    | 18.92    | 16.99    | 15.02    |
|            | WVTV                     | 20.01    | 19.97    | 19.90    | 18.76    | 18.75    | 18.63    | 18.46    |
| Rayleigh   | Wavelet                  | 27.54    | 24.76    | 21.04    | 17.77    | 14.95    | 12.35    | 9.86     |
|            | Contourlet               | 25.75    | 20.03    | 19.90    | 16.97    | 14.42    | 11.96    | 9.25     |
|            | TV                       | 26.11    | 23.87    | 20.73    | 17.69    | 15.01    | 12.50    | 10.50    |
|            | WVTV                     | 20.30    | 19.59    | 18.15    | 16.20    | 14.12    | 11.73    | 8.78     |
| Exponential| Wavelet                  | 25.15    | 6.69     | 24.63    | 21.38    | 18.30    | 15.41    | 12.72    |
|            | Contourlet               | 23.63    | 22.95    | 20.84    | 18.35    | 16.05    | 13.76    | 11.58    |
|            | TV                       | 23.53    | 24.78    | 24.20    | 22.28    | 19.86    | 17.20    | 14.60    |
|            | WVTV                     | 18.28    | 18.71    | 18.47    | 17.60    | 16.22    | 14.37    | 12.29    |

Note: SNR, signal-to-noise ratio; TV, total variation; WVTV, wavelet-total variation. The values in bold indicate the best SNR for each noise level.

### Table 3 Performance of noise reduction techniques evaluated using spot efficiency

| Noise type | Noise reduction technique | Noise intensity (dB) |
|------------|--------------------------|----------------------|
|            |                          | 20       | 18       | 16       | 14       | 12       | 10       | 8        |
| Gaussian   | Wavelet                  | 90.71    | 90.00    | 90.36    | 87.86    | 88.57    | 86.07    | 71.79    |
|            | Contourlet               | 89.29    | 88.57    | 87.50    | 87.51    | 78.93    | 65.00    | 44.64    |
|            | TV                       | 87.14    | 89.64    | 86.79    | 85.00    | 72.21    | 37.89    | 25.12    |
|            | WVTV                     | 87.14    | 86.43    | 86.43    | 85.71    | 86.79    | 87.14    | 85.16    |
| Rayleigh   | Wavelet                  | 90.36    | 89.64    | 89.64    | 91.43    | 89.64    | 87.86    | 88.57    |
|            | Contourlet               | 89.64    | 88.57    | 88.57    | 86.07    | 89.29    | 84.64    | 72.36    |
|            | TV                       | 87.86    | 87.86    | 85.36    | 88.57    | 85.43    | 87.59    | 51.25    |
|            | WVTV                     | 86.43    | 86.79    | 87.86    | 85.00    | 86.79    | 86.07    | 83.57    |
| Exponential| Wavelet                  | 90.00    | 90.00    | 88.93    | 88.21    | 88.21    | 88.21    | 77.14    |
|            | Contourlet               | 90.36    | 87.14    | 87.86    | 84.29    | 72.50    | 61.84    | 48.25    |
|            | TV                       | 90.71    | 89.29    | 87.50    | 86.07    | 82.86    | 75.00    | 62.37    |
|            | WVTV                     | 85.71    | 85.36    | 85.36    | 85.71    | 84.29    | 85.57    | 83.93    |

Note: TV, total variation; WVTV, wavelet-total variation. The values in bold indicate the best spot efficiency for each noise level.
Table 4 presents SNR and spot efficiency for images with the blurring effect and non-uniform background distribution filtered with nonlinear techniques wavelet, contourlet, TV, and WVTV. Contourlet and wavelet transform have the best performance with blurred images and images with a non-uniform background distribution. Both filtering techniques achieved comparable results for the different levels of blurring and distortion.

Comparison with a real 2-DGE image

A real 2-DGE image taken from the LECB 2-D PAGE Gel Images Datasets was used to compare the performance of the aforementioned filtering techniques [65]. Sample 19, an annotated 2-DGE image of human leukemia (Figure 4), in which many proteins have been previously identified was
selected for computing the rate of detection of these known proteins before and after filtering using the different techniques. Table 5 presents the rate of detected proteins from the original image, and of the image filtered by wavelet, contourlet, TV, and WVTV. It can be seen that wavelet, contourlet, and WVTV improved the rate of detection in comparison with the non-filtered image. A detection rate of 71.2% was achieved using TV, whereas a maximum detection rate of 87.9% was achieved using wavelet.

### Table 5: Performance of noise reduction techniques evaluated using the rate of detected proteins in a real 2-DGE image

| Image               | Detection rate |
|---------------------|----------------|
| Original            | 75.8%          |
| Filtered by wavelet | 87.9%          |
| Filtered by contourlet | 81.8%        |
| Filtered by TV      | 71.2%          |
| Filtered by WVTV    | 78.8%          |

*Note: The value in bold indicates the best detection rate achieved. Data source: LECB 2-D PAGE Gel Images Dataset.*

### Table 6: Summary of preprocessing techniques for 2-DGE images

| Objective             | Approach               | Disadvantage                                |
|-----------------------|------------------------|---------------------------------------------|
| Image normalization   | Multiple-gel           | Requiring alignment of samples              |
| Background correction | Adjustment of polynomial | Requiring selection of background sample   |
|                       | Local and global minima | Depending on threshold selection            |
|                       | Adjustment using histogram | Requiring setting of optimization parameters |
| Filtering             | Wavelet                | Requiring scale selection                   |
|                       | Contourlet             | Requiring scale selection                   |
|                       | TV                     | Resulting in the staircase effect           |
|                       | WVTV                   | Depending on parameter of both wavelet and TV|

Remarks

The first step in most commercial 2-DGE image analysis software programs is spot detection. Often, false positive spots are detected due to noise and other artifacts in the image, and conversely, some real data may be missed. Therefore, image preprocessing is a fundamental step for proteomic analysis using 2-DGE, as the performance of the subsequent analysis is directly affected by the quality of the image. The development of image acquisition systems and optimal sample preparation protocols greatly improves image quality, reducing the intensity and frequency of unwanted anomalies. However, some anomalies are unavoidable and affect the accuracy and reproducibility of the proteomic characterization of the sample. Each of the preprocessing approaches meets particular needs. Nonetheless, the order these preprocessing stages should be implemented in, and whether all of them are actually necessary or some could be excluded without affecting the final performance, is still not clear. In conclusion, a methodology that adequately integrates these preprocessing approaches is needed.

Even though several specialized image processing software programs are available for 2-DGE, these still require considerable human intervention, which affects reproducibility of the results as well. The wide range of anomalies in 2-DGE images, such as fuzzy spots, vertical and horizontal streaking, and noise, make it difficult to process the image. Therefore, it is important to implement advanced preprocessing techniques that could help mitigate the effect of these anomalies. In this review, we grouped three approaches of preprocessing: filtering, image regularization, and background correction, which are suited to different purposes such as noise reduction, regularization of the intensity of the pixels, and background correction. If these preprocessing procedures are not implemented, negative effects are observed during the image processing stage, such as image distortion, loss of relevant information, and changes in the contour of the spots of interest, yielding non-representative data. Table 6 summarizes the most common techniques, and the disadvantages of the three main groups of approaches for 2-DGE image pre-processing.

Noise reduction is successful when filtering techniques are applied. Although linear filters are easy and simple for reducing noise in 2-DGE images, these filters tend to distort the edges of the spots of interest. Techniques such as Gaussian filters, orthogonal regression, and PCA often remove small details such as low-abundance proteins in the process. Thus, nonlinear noise reduction techniques are presented as more
appropriate solutions. Nonlinear filters are more robust, but also more complex to use; they require the definition of parameters that depend on the image. It is clear that there is still a great scope for improving the performance of preprocessing techniques for 2-DGE images, as well as for proposing new ones, since the decision to implement these techniques in 2-DGE image processing is based on their performance with other types of images. Therefore, 2-DGE images are an attractive alternative for testing new algorithms for noise reduction, background correction, and image adjustment.

In this article, we validated the quantitative performance of the most representative preprocessing techniques for noise reduction of 2-DGE images. By comparing the performance of wavelet, contourlet, TV, and WVT V techniques using SNR and spot efficiency indicators, we conclude that spot efficiency is more appropriate for evaluating the noise reduction techniques applied to 2-DGE images. This index provides information about the effect of the noise reduction technique on the process of protein detection. Quantitative performance comparison using synthetic images indicates that wavelet filtering is the best technique under the test conditions, which achieves good results for the images with different types and levels of noise evaluated.

Taking into account the results obtained for the images with a blurring effect, and the fact that this anomaly is recurrent [66], we highlight the importance of high-quality image acquisition equipment, as this blurring effect is not mitigated effectively by the preprocessing techniques evaluated in this review. Blurring dramatically affects the performance of detection algorithms, and hence, specialized deblurring algorithms could be included in the preprocessing stage of 2-DGE images. However, cautions should be taken when including these techniques since they can lead to a significant distortion of the image [67].

Competing interests

The authors have declared no competing interests.

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References

[1] Shruthi BS, Vinodhkumar Selvamani P. Proteomics: a new perspective for cancer. Adv Biomed Res 2016;5:67.
[2] Magdeldin S, Enany S, Yoshida Y, Xu B, Zhang Y, Zureena Z, et al. Basics and recent advances of two dimensional- polyacrylamide gel electrophoresis. Clin Proteomics 2014;11:16–26.
[3] Friedman DB, Hoving S, Westermeier R. Isoelectric focusing and two-dimensional gel electrophoresis. Methods Enzymol 2009;463:515-40.
[4] Rabilloud T, Lelong C. Two-dimensional gel electrophoresis in proteomics: a tutorial. J Proteomics 2011;74:1829-41.
[5] Klowe J. Protein mapping by combined isoelectric focusing and electrophoresis of mouse tissues. A novel approach to testing for induced point mutations in mammals. Humangenetik 1975;26:231–43.
[6] Mitra A, Boroujeni MB. Application of gel-based proteomic technique in female reproductive investigations. J Hum Reprod Sci 2015;8:18-24.
[7] Curreem SOT, Watt RM, Lau SKP, Woo PCY. Two-dimensional gel electrophoresis in bacterial proteomics. Protein Cell 2012;3:346-63.
[8] Murphy S, Dowling P, Ohslandeck K. Comparative skeletal muscle proteomics using two-dimensional gel electrophoresis. Proteomes 2016;4:27.
[9] Huang CH, Chiou SH. Clinical proteomics identifies potential biomarkers in Helicobacter pylori for gastrointestinal diseases. World J Gastroenterol 2014;20:1529-36.
[10] Alvarez-Chaver P, Otero-Estévez O, de la Cadena MP, Rodríguez-Berrocal FJ, Martinez-Zorrano VS. Proteomics for discovery of candidate colorectal cancer biomarkers. World J Gastroenterol 2014;20:3804-24.
[11] Rogowska-Wrzesinska A, Le Bihan MC, Thayyen-Andersen M, Roepstorff P. 2D gels still have a niche in proteomics. J Proteomics 2013;4:13.
[12] Bandow JE, Baker JD, Berth M, Painter C, Sepulveda OJ, Clark JA, et al. Improved image analysis workflow for 2-D gels enables large-scale 2-D gel-based proteomics studies-COPD biomarker discovery study. Proteomics 2008;8:3030-41.
[13] Sengar RS, Upadhyay AK, Singh M, Gadre VM. Analysis of 2-D gel images for detection of protein spots using a novel non-separable wavelet based method. Biomed Signal Process Control 2016;25:62-75.
[14] Shamekhi S, Miran Baygi MH, Azarian B, Gooya A. A novel multi-scale Hessian based spot enhancement filter for two dimensional gel electrophoresis images. Comput Biol Med 2015;66:154–69.
[15] Rye M, Fargestad EM. Preprocessing of electrophoretic images in 2-DE analysis. Chemom Intell Lab Syst 2012;117:70-9.
[16] Nhek S, Tessema B, Indahl UG, Martens H, Mosleth EF. 2D electrophoresis image segmentation within a pixel-based framework. Chemom Intell Lab Syst 2015;141:33-46.
[17] Kostopoulou E, Katsigiannis S, Maroulis D. 2-D gel spot detection and segmentation based on modified image-aware grow-cut and regional intensity information. Comput Methods Programs Biomed 2015;122:26-39.
[18] Smith R. Two-dimensional electrophoresis: an overview. Methods Mol Biol 2009;519:1–16.
[19] Kaczmarek K, Walczak B, De Jong S, Vandeginste BGM. Preprocessing of two-dimensional gel electrophoresis images. Proteomics 2004;4:2377-89.
[20] Rabilloud T, Chevallet M, Luche S, Lelong C. Two-dimensional gel electrophoresis in proteomics: past, present and future. J Proteomics 2010;73:2064-77.
[21] Tsakanikas P, Manolakos ES. Improving 2-DE gel image denoising using contourlets. Proteomics 2009;9:3877-88.
[22] Nhek S, Mosleth E, Høy M, Griessl M, Tessema B, Indahl U, et al. Nonlinear visualisation and pixel-based alignment of 2D electrophoresis images. Chemom Intell Lab Syst 2012;118:97-108.
[23] Savelonas MA, Mylona EA, Maroulis D. Unsupervised 2D gel electrophoresis image segmentation based on active contours. Pattern Recognit 2012;45:720–31.
[24] Chang J, Van Remmen H, Ward WF, Regnier FE, Richardson A, Cornell J. Processing of data generated by 2-dimensional gel electrophoresis for statistical analysis: missing data, normalization, and statistics. J Proteome Res 2004;3:1210-8.
[25] Dasykowski M, Wrobel MS, Bierczynska-Krzysik A, Silberring J, Lubec G, Walczak B. Automatic preprocessing of electrophoretic images. Chemom Intell Lab Syst 2009;97:132-40.
[26] Berth M, Moser FM, Kolbe M, Bernhardt J. The state of the art in the analysis of two-dimensional gel electrophoresis images. Appl Microbiol Biotechnol 2007;76:1223-43.
[27] Hortin GL, Sviridov D. The dynamic range problem in the analysis of the plasma proteome. J Proteomics 2010;73:629-36.
[28] Keeping AJ, Collins RA. Data variance and statistical significance in 2D-gel electrophoresis and DIGE experiments: comparison of the effects of normalization methods. J Proteome Res 2011;10:1353–60.

[29] Rasliwan S, Faheem T, Sarhan A, Youssef BA. A fuzzy-watershed based algorithm for protein spot detection in 2-DGE images. JCSNS Int J Comput Sci Netw Secur 2010;10:254–60.

[30] Banerjee T, Kambhampati C. An image analysis suite for spot detection and spot matching in two-dimensional electrophoresis gels. Electrophoresis 2008;29:706–15.

[31] Faergestad EM, Rye M, Waleczk B, Gidskehaug L, Wold JP, Grove H, et al. Pixel-based analysis of multiple images for the identification of changes: a novel approach applied to untravely proteome patterns of 2-D electrophoresis gel images. Proteomics 2007;7:3450–61.

[32] Eilers PHC. Parametric time warping. Anal Chem 2004;76:404–11.

[33] Kaczmarek K, Waleczk B, de Jong S, Vandeginste GM. Baseline reduction in two dimensional gel electrophoresis images. Acta Chromatogr 2005;15:82–96.

[34] Peep P, Corzo LG. Local pixel value collection algorithm for spot segmentation in two-dimensional gel electrophoresis research. Comp Funct Genomics 2007;2007:89596.

[35] Brauner JM, Groemer TW, Stroebel A, Grosse-Holz S, Oberstein T, Wiltfang J, et al. Spot quantification in two dimensional gel electrophoresis image analysis: comparison of different approaches and presentation of a novel compound fitting algorithm. BMC Bioinformatics 2014;15:181.

[36] Picardi M. Background subtraction techniques: a review. IEEE Inter Conf Syst Man Cybern 2005;3099–104.

[37] Li Y, Li X. A background correction method based on lazy snapping. Proc 7th Int Conf Image Graphics 2012;1–5.

[38] Dowsey AW, English JA, Lisacek F, Morris JS, Yang GZ, Dunn MJ. Image analysis tools and emerging algorithms for expression proteomics. Proteomics 2010;10:4226–57.

[39] Camistraci CV, Abbas A, Gao X. Median Modified Wiener Filter for nonlinear adaptive spatial denoising of protein NMR multidimensional spectra. Sci Rep 2015;5:8017.

[40] Worz S, Winz ML, Rohr K. Geometric alignment of 2D gel electrophoresis images using physics-based elastic registration. Proc 5th IEEE Int Symp Biomed Imaging: Nano Macro 2008;1135–8.

[41] Sengar RS, Kumar A. Segmentation of two dimensional electrophoresis gel image using the Wavelet Transform and the Watershed Transform. Proc 2012 National Conf Commun 2012:1–5.

[42] Van Hulle S, Vandewalle J. The total least squares problem: computational aspects and analysis. 1st ed. Philadelphia: SIAM; 1991.

[43] Daszykowski M, Stanimirova I, Bodzon-Kulakowska A, Silvering J, Lubec G, Waleczk B. Start-to-end processing of two-dimensional gel electrophoretic images. J Chromatogr 2007;1158:306–17.

[44] Locantore N, Marron JS, Simpson DG, Tripoli N, Zhang JT, Cohen KL, et al. Robust principal component analysis for functional data. Test 1999;8:1–73.

[45] Appel RD, Vargas JR, Palagi PM, Walther D, Hochstrasser DF. Melanie II-a third-generation software package for analysis of two-dimensional electrophoresis images: II. Algorithms. Electrophoresis 1997;18:2735–48.

[46] Do MN, Vetterli M. The contourlet transform: An efficient directional multiresolution image representation. IEEE Trans Image Process 2005;14:2091–106.

[47] Oliveri G, Anselmi N, Massa A. Compressive sensing imaging of non-sparsely distributed scatterers by a total-variation approach within the Born approximation. IEEE Trans Antennas Propag 2014;62:5157–70.

[48] Takanikas, Manolakos. Effective denoising of 2D gel proteomics images using contours. Proc 2007 IEEE Int Conf Image Process 2007:269–72.

[49] Chan TF, Osher S, Shen J. The digital TV filter and nonlinear denoising. IEEE Trans Image Process 2001;10:231–41.

[50] Xin H, Zhao F. Effective denoising methods for two-dimensional gel electrophoresis images. Proc 4th Int Conf Biomed Eng Informatics 2011:1571–4.

[51] Catté F, Lions PL, Morel JM, Coll T. Image selective smoothing and edge detection by nonlinear diffusion. SIAM J Numer Anal 1992;29:182–93.

[52] Faheem MT, Rashwan S, Sarhan A, Youssef BA. De-noising 2D gel images using wavelet transform based on genetic algorithm. Proc 10th WSEAS Int Conf Appl Comput Appl Comput Sci 2011:187–94.

[53] Bartášek P, Piinasil J, Směkal Z. Wavelet-based de-noising techniques in MRI. Comput Methods Programs Biomed 2011;104:480–8.

[54] Munsell BC, Barsanti RJ. Electrophoresis 2D gel residual encoding using adaptive wavelets for image segmentation. Proc IEEE Southeast Conf 2005:163–8.

[55] Pasti L, Waleczk B, Massart DL, Reschiglian P. Optimization of signal denoising in discrete wavelet transform. Chemom Intell Lab Syst 1999;48:21–34.

[56] Cheung KW, Po LM. Low complexity preprocessing for 2D discrete multiwavelet transform. Proc 1997 IEEE Int Symp Circuits Syst 1997:0–3.

[57] Mallat S. A Theory for multiresolution signal decomposition: the wavelet representation. Pattern Anal Mach Intel IEEE Trans 1989;11:674–93.

[58] Yora P, Chudasama N. Different image fusion techniques and parameters: a review. Int J Comput Sci Inf Technol 2015;6:889–92.

[59] Chang SG, Yu B, Vetterli M. Adaptive wavelet thresholding for image denoising and compression. IEEE Trans Image Process Publ IEEE Signal Process Soc 2000;9:1532–46.

[60] Barlaud M, Sole P, Gaidot T, Antonini M, Mathieu P. Pyramidal lattice vector quantization for multiscale image coding. IEEE Trans Image Process 1994;3:367–81.

[61] Klein R, Faust O. Genetic algorithms. In: Supply chain management and advanced planning. Berlin: Springer Berlin Heidelberg; 2015. p. 537–44.

[62] Marengo E, Demartini M, Robotti E, Bobba M. A new algorithm for the simulation of sodium dodecil sulfate two-dimensional polyacrylamide gel electrophoresis data sets. J Proteome Res 2010;9:1864–72.

[63] Gonzalez RC, Woods RE. Digital image processing. 2nd ed. New Jersey: Prentice Hall; 2002.

[64] Yitzhaky Y, Kopeika NS. Identification of blur parameters from motion blurred images. Graph Model Image Process 1997;59:310–20.

[65] Lester EP, Lemkin PF, Lipkin LE. Protein indexing in leukemias and lymphomas. Ann N Y Acad Sci 1984;428:158–72.

[66] Jonak K, Ječsaiak K, Puszynski K. Application of the Born approximation. IEEE Trans Antennas Propag 2010:9:1864–72.

[67] Khan A, Yin H. Quality measures for blind image deblurring. Proc 2012 IEEE Int Conf Imaging Syst Tech 2012:456–9.