Subchronic oral toxicity of silver nanoparticles

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Abstract

Background: The antibacterial effect of silver nanoparticles has resulted in their extensive application in health, electronic, consumer, medicinal, pesticide, and home products; however, silver nanoparticles remain a controversial area of research with respect to their toxicity in biological and ecological systems.

Results: This study tested the oral toxicity of silver nanoparticles (56 nm) over a period of 13 weeks (90 days) in F344 rats following Organization for Economic Cooperation and Development (OECD) test guideline 408 and Good Laboratory Practices (GLP). Five-week-old rats, weighing about 99 g for the males and 92 g for the females, were divided into four groups (10 rats in each group): vehicle control, low-dose (30 mg/kg), middle-dose (125 mg/kg), and high-dose (500 mg/kg). After 90 days of exposure, clinical chemistry, hematology, histopathology, and silver distribution were studied. There was a significant decrease (P < 0.05) in the body weight of male rats after 4 weeks of exposure, although there were no significant changes in food or water consumption during the study period. Significant dose-dependent changes were found in alkaline phosphatase and cholesterol for the male and female rats, indicating that exposure to more than 125 mg/kg of silver nanoparticles may result in slight liver damage. Histopathologic examination revealed a higher incidence of bile-duct hyperplasia, with or without necrosis, fibrosis, and/or pigmentation, in treated animals. There was also a dose-dependent accumulation of silver in all tissues examined. A gender-related difference in the accumulation of silver was noted in the kidneys, with a twofold increase in female kidneys compared to male kidneys.

Conclusions: The target organ for the silver nanoparticles was found to be the liver in both the male and female rats. A NOAEL (no observable adverse effect level) of 30 mg/kg and LOAEL (lowest observable adverse effect level) of 125 mg/kg are suggested from the present study.

Background

The antibacterial activity exhibited by silver in a range of studies [1-5] has resulted in the widespread use of silver nanoparticles in bedding, washing machines, water purification, toothpaste, shampoo and rinse, nipples and nursing bottles, fabrics, deodorants, filters, kitchen utensils, toys, and humidifiers [6,7], where the main body or inner surface of the product is mixed or coated with germ-resistant nano-silver to prevent the growth of fungi and bacteria. Despite such widespread use of silver-nanoparticle-containing products, subchronic and chronic toxicity data on silver nanoparticles remain rare. The lack of exposure data on silver nanoparticles in the workplace and silver nanoparticles released from consumer products or released into the environment makes it difficult to assess the risks of using these materials. Limited data have been reported for silver nanoparticles by inhalation and oral routes of exposure. The target organs for silver nanoparticles by subchronic inhalation are the lungs and liver in male and female rats [8]. This study suggested a NOAEL (no observable adverse effect level) of 100 μg/m³. A LOAEL (lowest observable adverse effect level) and NOAEL have been reported for a 28-day oral toxicity study using Sprague-Dawley rats to be 300 mg/kg and 30 mg/kg, respectively [9]. In this study, F344 rats were exposed to silver nanoparticles following the Organization for Economic Cooperation and Development (OECD) test guideline 408 (OECD, 1998), using a 13-week repeated-oral-dose toxicity protocol. The study was conducted under OECD Good Laboratory Practices (GLP). Clinical chemistry, histopathology, and distribution of
silver nanoparticles were investigated in blood, lungs, kidneys, brain, liver, and other organs.

**Materials and methods**

**Silver Nanoparticles**

Silver nanoparticles (CAS No. 7440-22-4) were purchased from NAMATECH, Ltd. (Daejeon, Korea), and were at least 99.98% pure. Count median diameter and geometric standard deviation of silver nanoparticles in 0.5% aqueous carboxymethylcellulose (CMC, Sigma USA) analyzed by transmission electron microscopy were 56 nm and 1.46, respectively (Figure 1).

**Transmission Electron Microscopy**

The filters on which the silver nanoparticles in the 0.5% CMC were filtered were coated with carbon, mounted on an electron microscope grid (200 mesh, Veco, Eerbeek, Holland), and visualized under a transmission electron microscope (TEM, Hitachi 7100). The diameters of 400 randomly selected particles were measured at 50,000 x magnification, and the silver particles were analyzed using an energy-dispersive x-ray analyzer (EDX-200, Horiba, Japan) at an accelerating voltage of 75 kV.

**Animals and Conditions**

Four-week-old male and female, specific-pathogen free (SPF) Fisher 344 rats were purchased from Japan SLC Inc. (Japan) and acclimated for 7 days before starting the experiments. During the acclimation and experimental periods, the rats were housed in polycarbonate cages (maximum of 3 rats per cage) in a room with controlled temperature and humidity.
temperature (22.2 ± 1.7°C) and humidity (48.4 ± 6.0%), and a 12-h light/dark cycle. The rats were fed a rodent diet (Harlan Teklad, USA) and filtered water ad libitum. The rats were divided into 4 groups (10 rats in each group): vehicle control (0.5% carboxymethylcellulose, CMC), low-dose group (30 mg/kg/day), middle-dose group (125 mg/kg/day), and high-dose group (500 mg/kg/day). When the rats reached five weeks of age, they were exposed to silver nanoparticles following OECD test guideline 408 [10] by gavage for 13 weeks of repeated oral administration (dosing volumes were 10 ml/kg). Dose levels were selected based on previous observations in a 28-day oral toxicity study by Kim et al. [9]. The study was conducted under OECD Good Laboratory Practices.

Clinical Chemistry and Hematology
At the conclusion of the 13-wk experiment, the rats were 18 wks old. Before necropsy, food was withheld for 24 h and the rats were anesthetized with CO2 gas. Blood was then drawn from the abdominal aorta, collected in heparinized vacutainers, and analyzed for ALB (albumin), ALP (alkaline phosphatase), Ca (calcium), CHO (cholesterol), CRE (creatinine), gamma-GT (gamma-glutamyl transpeptidase), GLU (glucose), GOT (glutamic oxaloacetic transaminase), GPT (gamma-glutamyl transpeptidase), IP (inorganic phosphorus), LDH (lactate dehydrogenase), MG (magnesium), TP (total protein), UA (uric acid), BUN (blood urea nitrogen), TBIL (total bilirubin), CK (creatine phosphokinase), Na (sodium), K (potassium), Cl (chloride), TG (triglyceride), and A/G (ratio of albumin to globulin) using a biochemical blood analyzer (Hitachi 7180, Hitachi, Japan). The blood was also analyzed for the WBC (white blood cell count), RBC (red blood cell count), Hb (hemoglobin concentration), HTC (hematocrits), MCV (mean corpuscular volume), MCH (mean corpuscular hemoglobin), MCHC (mean corpuscular hemoglobin concentration), RDW (red cell distribution width), PLT (platelet count), MPV (mean platelet volume), NE# (number of neutrophils), NE% (percent of neutrophils), LY# (number of lymphocytes), LY% percent of lymphocytes), MO# (number of monocytes), MO% (percent of monocytes), EO# (number of eosinophils), EO% (percent of eosinophils), BA# (number of basophils), and BA% (percent of basophils) using a blood cell counter (Hemavet 0950, CDC Tech., USA).

Organ Weights and Histopathology
After collecting blood samples, the rats were killed by cervical dislocation. Adrenal glands, bladder, testes, ovaries, uterus, epididymis, seminal vesicle, heart, thymus, thyroid gland, trachea, esophagus, tongue, prostate, lungs, nasal cavity, kidneys, spleen, liver, pancreas, and brain were removed carefully, weighed, and fixed in a 10% formalin solution containing neutral phosphate-buffered saline. Thereafter, the organs were embedded in paraffin, stained with hematoxylin and eosin, and examined under light microscopy.

Determination of Tissue Silver
Tissues were digested with conc. nitric acid by using a microwave digestion system (MARS 230/60, CEM). The concentration of silver in digested fluid was analyzed with a flameless method using an atomic absorption spectrophotometer equipped with a Zeeman graphite furnace (Perkin Elmer 5100ZL, Zeeman Furnace Module, USA) based on the NIOSH 7300 method [11]. The concentration of silver in the tissue was expressed as μg/g wet weight.

Statistical Analysis
Statistical analysis was performed with SPSS (Version 12). Statistical evaluation was performed by analysis of two-tailed Student’s t-test or analysis of variance (ANOVA) following multiple comparison tests with Duncan’s method. The level of statistical significance was set at p < 0.05.

Results
Animal Observation, Food Consumption, and Effect on Body and Organ Weights
There were no significant differences in food consumption and water intake between treated male and female rats and the control group (data not shown). There were no significant dose-related changes in the body weight of female rats; however, there were significant (P < 0.05) dose-related decreases in the body weight of high-dose male rats at 4, 5, and 7 weeks of exposure at the conclusion of the study 13 weeks, and middle-dose male rats at 10 weeks of exposure (Figure 2). No significant organ-weight changes were observed in either the male or female rats after 90 days except for an increase (P < 0.05) in the weight of the left testis for the high-dose male rats, and for decreases (P < 0.05) in the weight of right kidney for the low-and middle-dose female rats (Tables 1 and 2).

Effects on Clinical Chemistry and Hematology
There appeared to be increase ALP for male rats in the middle and high-dose groups statistically not significant (Table 3). However, there was a significant increase (P < 0.01) in alkaline phosphatase (ALP) for female rats in the high-dose groups (Table 4). A significant increase (P < 0.01) in cholesterol was also found in the middle and high-dose male rats and the high-dose female rats (Table 3 and 4). A significant increase (P < 0.05) in total bilirubin was noted in the middle-dose male rats.
Figure 2 Body weight changes during 90-day oral administration of silver nanoparticles. A. Male. a Significant difference vs. control, \( p < 0.05 \) (10 weeks). b Significant difference vs. control, \( p < 0.05 \) (4, 5, 7–13 weeks). B. Female.

**Table 1 Relative organ weights for male rats after 90-day oral administration of silver nanoparticles (mean ± S.D., \( n \geq 9 \))**

| Dose (mg/Kg) | 0 | 30 | 125 | 500 |
|--------------|---|----|-----|-----|
| Testis (Left) | 0.47 ± 0.02 | 0.48 ± 0.03 | 0.49 ± 0.02 | 0.51 ± 0.01* |
| Testis (Right) | 0.47 ± 0.02 | 0.48 ± 0.02 | 0.48 ± 0.02 | 0.49 ± 0.02 |
| Spleen | 0.21 ± 0.03 | 0.21 ± 0.01 | 0.22 ± 0.01 | 0.20 ± 0.02 |
| Liver | 2.96 ± 0.10 | 2.91 ± 0.16 | 2.92 ± 0.09 | 2.87 ± 0.11 |
| Pituitary gland | 0.002 ± 0.001 | 0.003 ± 0.006 | 0.005 ± 0.004 | 0.003 ± 0.003 |
| Adrenal gland (Left) | 0.011 ± 0.002 | 0.010 ± 0.001 | 0.008 ± 0.001 | 0.009 ± 0.003 |
| Adrenal gland (Right) | 0.008 ± 0.001 | 0.007 ± 0.001 | 0.008 ± 0.001 | 0.009 ± 0.003 |
| Prostate | 0.13 ± 0.06 | 0.16 ± 0.05 | 0.15 ± 0.04 | 0.16 ± 0.05 |
| Lungs | 0.42 ± 0.02 | 0.41 ± 0.04 | 0.42 ± 0.03 | 0.43 ± 0.03 |
| Brain | 0.62 ± 0.02 | 0.61 ± 0.03 | 0.62 ± 0.02 | 0.63 ± 0.02 |
| Heart | 0.27 ± 0.02 | 0.26 ± 0.02 | 0.27 ± 0.01 | 0.26 ± 0.02 |
| Thymus | 0.10 ± 0.02 | 0.10 ± 0.02 | 0.11 ± 0.02 | 0.10 ± 0.02 |
| Kidney (Left) | 0.28 ± 0.01 | 0.28 ± 0.02 | 0.29 ± 0.01 | 0.28 ± 0.01 |
| Kidney (Right) | 0.29 ± 0.02 | 0.28 ± 0.02 | 0.29 ± 0.01 | 0.29 ± 0.01 |

*Significant difference vs. control, \( p < 0.05 \)

**Table 2 Relative organ weights for female rats after 90-day oral administration of silver nanoparticles (mean ± S.D., \( n \geq 9 \))**

| Dose (mg/Kg) | 0 | 30 | 125 | 500 |
|--------------|---|----|-----|-----|
| Ovary (Left) | 0.027 ± 0.004 | 0.026 ± 0.005 | 0.023 ± 0.003 | 0.024 ± 0.007 |
| Ovary (Right) | 0.072 ± 0.134 | 0.025 ± 0.006 | 0.026 ± 0.003 | 0.028 ± 0.005 |
| Spleen | 0.22 ± 0.02 | 0.21 ± 0.01 | 0.21 ± 0.01 | 0.21 ± 0.02 |
| Liver | 2.67 ± 0.18 | 2.61 ± 0.07 | 2.53 ± 0.12 | 2.62 ± 0.17 |
| Pituitary gland | 0.007 ± 0.002 | 0.006 ± 0.001 | 0.006 ± 0.001 | 0.007 ± 0.002 |
| Adrenal gland (Left) | 0.015 ± 0.003 | 0.014 ± 0.002 | 0.014 ± 0.001 | 0.015 ± 0.003 |
| Adrenal gland (Right) | 0.014 ± 0.002 | 0.014 ± 0.001 | 0.014 ± 0.001 | 0.015 ± 0.003 |
| Uterus | 0.233 ± 0.044 | 0.257 ± 0.090 | 0.216 ± 0.040 | 0.195 ± 0.040 |
| Lungs | 0.54 ± 0.06 | 0.55 ± 0.04 | 0.50 ± 0.05 | 0.54 ± 0.05 |
| Brain | 0.99 ± 0.05 | 0.98 ± 0.05 | 0.99 ± 0.06 | 1.00 ± 0.05 |
| Heart | 0.32 ± 0.02 | 0.32 ± 0.02 | 0.31 ± 0.02 | 0.32 ± 0.03 |
| Thymus | 0.14 ± 0.02 | 0.13 ± 0.02 | 0.13 ± 0.02 | 0.15 ± 0.02 |
| Kidney (Left) | 0.31 ± 0.02 | 0.30 ± 0.02 | 0.30 ± 0.01 | 0.30 ± 0.02 |
| Kidney (Right) | 0.31 ± 0.02 | 0.29 ± 0.01 | 0.30 ± 0.02 | 0.01* |

*Significant difference vs. control, \( p < 0.05 \)
(Table 3). A significant decrease in magnesium, protein, and inorganic phosphorus was found in the middle and high-dose female rats (Table 4). No significant changes in the hematological parameters were noted except for a significant increase (p < 0.05) in monocytes in the high-dose group (Figure 3). A significant decrease in magnesium, protein, and total cholesterol was noted (Table 4). A significant decrease in magnesium, protein, and total cholesterol was noted (Table 4). A significant decrease in magnesium, protein, and total cholesterol was noted (Table 4). A significant decrease in magnesium, protein, and total cholesterol was noted (Table 4). A significant decrease in magnesium, protein, and total cholesterol was noted (Table 4).
Table 5 Hematological values for male rats after 90-day oral administration of silver nanoparticles (mean ± SD)

| Dose (mg/Kg) | 0 (n = 9) | 30 (n = 9) | 125 (n = 10) | 500 (n = 10) |
|--------------|-----------|------------|-------------|--------------|
| WBC          | 8.33 ± 1.25 | 8.22 ± 1.44 | 8.17 ± 1.28 | 8.21 ± 1.07  |
| RBC          | 9.08 ± 0.32 | 9.13 ± 0.66 | 8.88 ± 0.24 | 9.07 ± 0.35  |
| Hb           | 16.67 ± 0.40 | 16.76 ± 1.10 | 16.47 ± 0.53 | 16.93 ± 0.68 |
| HCT          | 36.96 ± 1.41 | 37.80 ± 2.90 | 36.97 ± 1.46 | 37.84 ± 2.12 |
| MCV          | 41.17 ± 0.58 | 41.38 ± 0.72 | 41.62 ± 0.77 | 41.72 ± 0.92 |
| MCH           | 18.58 ± 0.34 | 18.94 ± 0.40 | 18.55 ± 0.58 | 18.67 ± 0.37 |
| MCHC          | 45.13 ± 1.08 | 44.38 ± 1.31 | 44.59 ± 1.69 | 44.79 ± 1.25 |
| RDW           | 17.98 ± 0.78 | 17.83 ± 0.68 | 17.83 ± 1.04 | 18.10 ± 0.86 |
| MPV           | 738.22 ± 53.33 | 731.56 ± 105.79 | 725.40 ± 45.43 | 718.50 ± 72.05 |
| NEU          | 26.13 ± 3.30 | 26.72 ± 3.14 | 26.76 ± 4.14 | 27.12 ± 2.62 |
| LYO           | 69.77 ± 3.39 | 68.21 ± 3.45 | 68.93 ± 5.30 | 69.34 ± 3.13 |
| MONO          | 57.30 ± 7.80 | 54.57 ± 1.12 | 53.98 ± 1.19 | 53.35 ± 0.80 |
| EOS           | 0.25 ± 0.04 | 0.39 ± 0.50 | 0.25 ± 0.21 | 0.15 ± 0.08 |
| BASO          | 0.08 ± 0.14 | 0.12 ± 0.22 | 0.09 ± 0.08 | 0.04 ± 0.03 |

Note: WBC(K/μL), White blood cells; RBC(M/μL), Red blood cells; Hb(g/dL), Hemoglobin; HCT(%), Hematocrits; MCV(fl), Mean corpuscular volume; MCH (pg), Mean corpuscular hemoglobin; MCHC(g/dL), Mean corpuscular hemoglobin concentration; RDW(%), Red cell distribution width; PLT(μL), Platelets; MPV(μm), Mean platelet volume; NEU(%), Neutrophils; LYO(%), Lymphocytes; MONO(%), Monocytes; EOS(%), Eosinophils; BASO(%), Basophils.

Table 6 Hematological values for female rats after 90-day oral administration of silver nanoparticles (mean ± SD)

| Dose (mg/Kg) | 0 (n = 8) | 30 (n = 10) | 125 (n = 10) | 500 (n = 10) |
|--------------|-----------|------------|-------------|--------------|
| WBC          | 10.00 ± 4.69 | 8.40 ± 4.72 | 6.10 ± 2.13 | 5.80 ± 2.74  |
| RBC          | 12.22 ± 4.84 | 6.70 ± 3.74* | 12.10 ± 5.82 | 11.90 ± 3.90 |

*Significant difference vs. control, p < 0.05

10, and 2/10 of the control, low, middle, and high-dose female rats, respectively (Table 8 and 4D). The higher incidence of bile-duct hyperplasia, with or without necrosis, fibrosis, and/or pigmentation, in the treated animals also suggests a treatment-related effect. Although there appeared to be a slight increase of minimal tubular basophilia in the kidneys of high-dose male rats, these changes were not statistically significant and thus not considered to be test article exposure related. Tubular basophilia were also more prevalent in the male rats compared to the female rats (Table 8). Minimum or mild renal unilateral or bilateral mineralization was observed in 5/10, 8/10, 7/10, and 9/10 of the control, low, middle, and high-dose female rats, respectively, indicating a treatment-related effect.

Histopathologic examination of lung tissue did not show any treatment-related effects.

In the intestines, pigmentation of villi was observed in 0/10, 0/10, 8/10, and 8/10 of the control, low, middle, and high-dose male rats, respectively (Table 8). This dose-dependent increase in the pigmentation of the villi indicated an apparent treatment-related effect (Figure 5). In contrast, there was a slight treatment-related increase in pigmentation of intestinal villi in female rats (Table 9).

Silver Distribution in Tissues

There was a statistically significant (P < 0.01) dose-dependent increase in the silver concentration of all the tissue samples from the groups exposed to silver nanoparticles in this study (Table 10). In addition, a two-fold higher accumulation of silver in the kidneys of female rats when compared with the male rats occurred across all the dose groups indicating a marked gender-dependent distribution.

Discussion

Silver nanoparticles are widely used as bactericidal agents in consumer products, but their potential effects in humans remain poorly understood. Silver at doses below levels that cause argyria or argyriosis are generally considered to be relatively non-toxic [12]. Wijnhoven et al. have hypothesized that the toxic effects of silver
are proportional to free silver ions, but it is unclear how this relates to silver nanoparticles. Absorption of silver after oral administration has been shown to be subject to a first-pass effect through the liver, resulting in excretion into the bile [13]. Uncleared silver has been shown to be deposited in the renal glomerular basement membrane [14-17], mesangium [18], Kupffer cells, and sinusoid endothelium cells in the liver [15].

Unlike the 28-day study of Kim et al. [9], the present study shows intestinal pigmentation and effects from exposure to silver nanoparticles. Dose-dependent increases in the silver concentrations in the intestinal villi, as observed in this and other reports [19], and in the blood, indicate that the orally absorbed silver from nanoparticles is able to enter the blood circulation and be distributed to other organs. Since similar effects have not been previously reported for soluble silver, there is a possibility that these effects are due to particles as opposed to ionized silver.

Increases in renal silver reported in the current study are consistent with literature showing that ionized silver is deposited in the renal glomerular basement membrane [14-17] and mesangium [18]. The gender-related distribution of silver nanoparticles in the kidneys was also consistent with the results from the 28-day [9], 90-day oral, and 90-day inhalation studies [8]. The average nanoparticle sizes used in the 28-day and 90-day oral toxicity studies was 60 nm and 56 nm, respectively. Common treatment-related endpoints and distribution were found in all these studies indicating that distribution and toxicity do not appear to be dependent on particle size in the tested range or route of administration.

In previous reports, the target organs for silver nanoparticles were shown to be the liver in a 28-day oral toxicity study [9] and the liver and lungs in a 90-day inhalation study [8]. The results of the current study are consistent with previous reports with respect to the liver as the target organ and the tissue distribution of silver originating from nanoparticles. Liver toxicity as evaluated by histopathology included bile-duct hyperplasia and increased foci, which was consistent with the pathologic observations in the 28-day [9] oral toxicity and 90-day inhalation [8] toxicity studies. Increases in alkaline phosphatase and cholesterol were also consistent with the liver toxicity reported by Kim et al. and Sung et al.

In contrast to the 90-day inhalation study, no coagulation effects on peripheral blood were observed. Since there is similar hepatic pathology in the two studies, it is not clear whether increased coagulation times were due to hepatic injury in the inhalation study or some other mechanism.

Inhaled silver nanoparticles have been shown to cause lung inflammation [8]. These effects may be due to particle size since gold nanoparticles (which are not

Figure 3 Coagulation times for male and female rats: A, male prothrombin time (PT); B, male active partial thromboplastine time (APPT); C, female PT; D, female APPT. (Error bars indicate standard deviation.)
likely to be ionized) show similar effects [20]. Our very limited data makes it tempting to hypothesize that effects from silver nanoparticles are not only due to ionization of silver from the surface of silver nanoparticles, but may originate (at least in part) from direct effects of nanoparticles. The relationship between particle effect and effect from ionized silver could be tested by quantitatively examining target tissues for the presence of nanoparticles with a concurrent measurement of tissue ionized silver as opposed to measuring total tissue silver alone. If silver nanoparticles have particle-related effects which are tissue specific and significant compared to effects from ionized silver, it will be important to incorporate such differences into future occupational and environmental risk assessments.

Table 8 Histopathological findings for male rats after 90-day oral administration of silver nanoparticles

| GROUP | Control | Low | Middle | High |
|-------|---------|-----|--------|------|
| Liver | No microscopic findings | 5/10 | 3/10 | 0/10 |
| Abnormality | 5/10 | 7/10 | 10/10 | 100 |
| Hyperplasia | bile duct | minimum | 4/10 | 7/10 | 8/10 | 6/10 |
| Vacuolation | hepatocellular | minimum | 1/10 | 0/10 | 0/10 | 0/10 |
| Necrosis | focal | minimum | 0/10 | 4/10 | 3/10 | 4/10 |
| Lobular | moderate | 0/10 | 0/10 | 1/10 | 10/10 |
| Multifocal | moderate | 0/10 | 0/10 | 1/10 | 0/10 |
| Hemorrhage | 0/10 | 0/10 | 1/10 | 10/10 | 0/10 |
| Pigmentation | 0/10 | 0/10 | 1/10 | 10/10 | 2/10 |
| Kidneys | No microscopic findings | 10/10 | 10/10 | 100 |
| Abnormality | 0/10 | 0/10 | 1/10 | 100 |
| Basophilia | tubular | minimum | 0/10 | 0/10 | 1/10 | 2/10 |
| Inflammation | focal | minimum | 0/10 | 0/10 | 1/10 | 10/10 |
| Tubular | minimum | 0/10 | 0/10 | 0/10 | 1/10 |
| Lungs | No microscopic findings | 9/10 | 9/10 | 100 |
| Abnormality | 1/10 | 10/10 | 100 |
| Histocytois | focal | minimum | 1/10 | 0/10 | 0/10 | 1/10 |
| Mineralization | minimum | 0/10 | 0/10 | 0/10 | 2/10 |
| Anthracosis | 0/10 | 0/10 | 0/10 | 1/10 |
| Intestines | No microscopic findings | 10/10 | 10/10 | 100 |
| Abnormality | 0/10 | 0/10 | 8/10 | 8/10 |
| Pigment | villi | yellow | 0/10 | 0/10 | 1/10 | 8/10 |
| Heart | No microscopic findings | 10/10 | 10/10 | 100 |
| Abnormality | 0/10 | 0/10 | 2/10 | 2/10 |
| Inflammation | left ventricle | minimum | 0/10 | 0/10 | 2/10 | 1/10 |
| Right ventricle | minimum | 0/10 | 0/10 | 0/10 | 1/10 |
| Ears | No microscopic findings | 10/10 | 10/10 | 100 |
| Abnormality | 0/10 | 0/10 | 0/10 | 1/10 |
| Inflammation | Harderian gland | minimum | 0/10 | 0/10 | 0/10 | 1/10 |
| Pancreas | No microscopic findings | 10/10 | 10/10 | 100 |
| Abnormality | 0/10 | 0/10 | 0/10 | 2/10 |
| Inflammation | pancreas | minimum | 0/10 | 0/10 | 0/10 | 2/10 |
Figure 4 Histopathological findings in liver: (A) control liver, (B) multiple foci of inflammatory cell infiltrates, including eosinophils, especially around central vein and portal areas (C) bile duct hyperplasia, and (D) several foci of inflammatory cell infiltration, especially around central vein and sinusoid on hepatic lobules. Bar indicates 100 μm. A. Control (×100). B. High dose (×100). C. High dose (×100). D. High dose (×200).

Figure 5 Histopathological findings in intestines of male rats dosed with 500 mg/kg silver nanoparticles for 90 days. A. Control male (×40). B. Control male (×100). C. High-dose male (×40). D. High-dose male (×100).
### Table 9 Histopathological findings for female rats after 90-day oral administration of silver nanoparticles

| GROUP   | Control | Low   | Middle | High  |
|---------|---------|-------|--------|-------|
|         | N  | %     | N  | %     | N  | %     | N  | %     |
| Liver   |     |       |     |       |     |       |     |       |
| No microscopic findings | 5/10 | 50 | 2/10 | 20 | 2/10 | 20 | 3/10 | 30 |
| Abnormality |     |       |     |       |     |       |     |       |
| Hyperplasia |     |       |     |       |     |       |     |       |
| bile duct | 3/10 | 30 | 7/10 | 70 | 8/10 | 80 | 7/10 | 70 |
| Necrosis |     |       |     |       |     |       |     |       |
| focal | 0/10 | 0 | 2/10 | 20 | 2/10 | 20 | 1/10 | 10 |
| central vein | 0/10 | 0 | 0/10 | 0 | 0/10 | 0 | 1/10 | 10 |
| Fibrosis |     |       |     |       |     |       |     |       |
| minimum | 2/10 | 20 | 1/10 | 10 | 2/10 | 20 | 1/10 | 10 |
| Pigmentation |     |       |     |       |     |       |     |       |
| 0/10 | 0 | 0/10 | 0 | 4/10 | 40 | 1/10 | 10 |
| Kidneys |     |       |     |       |     |       |     |       |
| No microscopic findings | 5/10 | 50 | 2/10 | 20 | 3/10 | 30 | 1/10 | 10 |
| Abnormality |     |       |     |       |     |       |     |       |
| Mineralization |     |       |     |       |     |       |     |       |
| unilateral | 4/10 | 40 | 7/10 | 70 | 5/10 | 50 | 5/10 | 50 |
| bilateral | 1/10 | 10 | 1/10 | 10 | 1/10 | 10 | 4/10 | 40 |
| Lungs   |     |       |     |       |     |       |     |       |
| No microscopic findings | 10/10 | 100 | 10/10 | 100 | 10/10 | 100 | 5/10 | 50 |
| Abnormality |     |       |     |       |     |       |     |       |
| Histocytosis |     |       |     |       |     |       |     |       |
| 0/10 | 0 | 0/10 | 0 | 1/10 | 10 | 0/10 | 0 |
| Intestines |     |       |     |       |     |       |     |       |
| No microscopic findings | 10/10 | 100 | 10/10 | 100 | 10/10 | 100 | 5/10 | 50 |
| Abnormality |     |       |     |       |     |       |     |       |
| Pigment villi |     |       |     |       |     |       |     |       |
| yellow | 0/10 | 0 | 0/10 | 0 | 0/10 | 0 | 5/10 | 50 |
| faint yellow | 0/10 | 0 | 0/10 | 0 | 0/10 | 0 | 5/10 | 50 |
| Eyes    |     |       |     |       |     |       |     |       |
| No microscopic findings | 8/10 | 80 | 9/10 | 90 | 9/10 | 90 | 9/10 | 90 |
| Abnormality |     |       |     |       |     |       |     |       |
| Inflammation Harderian gland |    |       |     |       |     |       |     |       |
| minimum | 2/10 | 20 | 1/10 | 10 | 1/10 | 10 | 1/10 | 10 |
| Pancreas |     |       |     |       |     |       |     |       |
| No microscopic findings | 10/10 | 100 | 10/10 | 100 | 10/10 | 100 | 9/10 | 90 |
| Abnormality |     |       |     |       |     |       |     |       |
| Inflammation pancreas |     |       |     |       |     |       |     |       |
| minimum | 0/10 | 0 | 0/10 | 0 | 0/10 | 0 | 1/10 | 10 |

**Significant difference vs. control, p < 0.01. (ANOVA)**

* Significant difference vs. distinction of sex, p < 0.05. (T-test)

### Table 10 Tissue silver content after 90-day oral administration of silver nanoparticles

| Dose/Sex | Testes (μg/g wet weight) | Liver (μg/g wet weight) | Kidneys (μg/g wet weight) | Brain (μg/g wet weight) | Lungs (μg/g wet weight) | Blood (μg/g wet weight) |
|----------|--------------------------|-------------------------|---------------------------|-------------------------|-------------------------|-------------------------|
| 0 mg/Kg  | Male 0.04 ± 0.02          | Male 0.02 ± 0.01        | Female 0.03 ± 0.01        | Male 0.02 ± 0.01        | Female 0.05 ± 0.02      | Male 0.001 ± 0.000      |
|          | Female 0.01 ± 0.01        |                        |                          | Female 0.01 ± 0.01      |                        | Female 0.002 ± 0.002    |
| 30 mg/Kg | Male 6.56 ± 0.33**        | Male 4.20 ± 1.57**      | Male 1.49 ± 0.37**       | Male 0.47 ± 0.18**      | Male 1.94 ± 0.64**      | Male 0.111 ± 0.016**    |
|          | Female 8.56 ± 3.22**      | Female 7.98 ± 0.91**    | Female 0.38 ± 0.05**     | Female 4.97 ± 0.90**    | Female 0.087 ± 0.017**  |                        |
| 125 mg/Kg| Male 11.84 ± 1.62**       | Male 10.19 ± 2.09**     | Female 8.82 ± 2.13**     | Male 0.69 ± 0.06**      | Female 10.97 ± 3.81**   | Male 0.191 ± 0.037**    |
|          | Female 29.13 ± 9.74**     | Female 37.09 ± 17.44**  | Female 0.77 ± 0.11**     | Female 17.64 ± 9.06**   | Female 0.122 ± 0.010**  |                        |
| 500 mg/Kg| Male 23.75 ± 9.13**       | Female 68.65 ± 33.59**  | Female 99.19 ± 32.82**   | Male 3.54 ± 1.44**      | Female 56.04 ± 51.14    | Male 0.419 ± 0.083**    |
|          | Female 98.75 ± 31.65**    | 226.88 ± 55.64**        | 37.0 ± 1.17**            | 45.83 ± 11.43**         | 0.303 ± 0.099**         |                        |

**Significant difference vs. control, p < 0.01. (ANOVA)**

* Significant difference vs. distinction of sex, p < 0.05. (T-test)
Conclusions
A LOAEL (lowest observable adverse effect level) of 125 mg/kg and a NOAEL (no observable adverse effect level) of 30 mg/kg are suggested from the current 90-day oral toxicity study. The LOAEL from this study is significantly lower than the LOAEL of 300 mg/kg derived from the 28-day oral toxicity study (Kim et al., 2008). However, the NOAEL derived from this study is consistent with the NOAEL of 30 mg/kg derived from the 28-day oral study (Kim et al., 2008).

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Authors’ contributions
UIY headed the study and performed pathologic analyses together with MYS, YHC, HMC, IJK and drafted the manuscript with YSK and BK. YSK headed all animal treatments with MYS, KSS, and HRR. JDP and JHL contributed to the study. All authors reviewed and interpreted data and read and approved the final manuscript.

Competing interests
The authors declare that they have no competing interests.

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