C/EBP β Mediates Endoplasmic Reticulum Stress Regulated Inflammatory Response and Extracellular Matrix Degradation in LPS-Stimulated Human Periodontal Ligament Cells

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Abstract: Periodontitis is an oral inflammatory disease that not only affects the integrity of local tooth-supporting tissues but also impacts systemic health. A compositional shift in oral microbiota has been considered as the main cause of periodontitis; however, the potential mechanism has not been fully defined. Herein, we investigated the role of CCAAT/enhancer-binding protein β (C/EBP β), a member of the C/EBP family of transcription factors, in human periodontal ligament cells (hPDLCs) exposed to Porphyromonas gingivalis (P. gingivalis) lipopolysaccharide (LPS). RT-PCR and Western blotting analysis showed that the expression of C/EBP β was significantly increased in hPDLCs stimulated with LPS stimuli. Overexpression of C/EBP β by the recombinant adenoviral vector pAd/C/EBP β markedly increased the expression of the pro-inflammatory cytokines IL-6 and IL-8, and matrix metalloproteinases (MMP)-8 and -9 in hPDLCs in response to LPS. Furthermore, the activation of endoplasmic reticulum (ER) stress was confirmed in LPS-stimulated hPDLCs by measuring the expression of the ER stress marker molecules protein kinase-like ER kinase (PERK), eIF2α, GRP78/Bip, and C/EBP homologous protein (CHOP). The ER stress inhibitor salubrinal repressed, but inducer tunicamycin enhanced, the production of IL-6, IL-8, MMP-8, and MMP-9 in hPDLCs stimulated with LPS stimuli. Overexpression of C/EBP β by the recombinant adenoviral vector pAd/C/EBP β markedly increased the expression of the pro-inflammatory cytokines IL-6 and IL-8, and matrix metalloproteinases (MMP)-8 and -9 in hPDLCs in response to LPS. Furthermore, the activation of endoplasmic reticulum (ER) stress was confirmed in LPS-stimulated hPDLCs by measuring the expression of the ER stress marker molecules protein kinase-like ER kinase (PERK), eIF2α, GRP78/Bip, and C/EBP homologous protein (CHOP). The ER stress inhibitor salubrinal repressed, but inducer tunicamycin enhanced, the production of IL-6, IL-8, MMP-8, and MMP-9 in hPDLCs. Additionally, ER stress inducer tunicamycin significantly increased the expression level of C/EBP β in hPDLCs. Blocking of C/EBP β by siRNA resulted in a significant decrease in the secretion of IL-6 and IL-8 and expression of MMP-8 and MMP-9 induced by tunicamycin treatment in hPDLCs. Taken together, ER stress appears to play a regulatory role in the inflammatory response and extracellular matrix (ECM) degradation in hPDLCs in response to LPS stimuli by activating C/EBP β expression. This enhances our understanding of human periodontitis pathology.

Keywords: periodontitis; C/EBP β; endoplasmic reticulum stress; human periodontal ligament cells; P. gingivalis LPS

1. Introduction

Periodontitis is an oral inflammatory disease that affects the tooth-supporting tissues, including the alveolar bone, gingiva, parodontium, and cementum. A compositional shift in oral microbiota, especially Porphyromonas gingivalis (P. gingivalis), Treponema denticola, and Tannerella forsythia, has been
considered as the main cause of periodontitis [1]. The accumulation of virulent oral bacteria on
adjacent teeth results in the loss of connective tissue and causes bone loss, eventually affecting
the integrity of the supporting structures. Dysfunction of the periodontium not only disrupts the local oral
conditions but also affects systemic health [2,3]. Those patients with periodontitis face an increased
risk of atherosclerosis, aspiration pneumonia, rheumatoid arthritis, and cancer. Hence, knowledge
about the pathogenic mechanisms related to periodontal disease is urgently needed to improve the
therapeutic strategies for human periodontitis.

Endoplasmic reticulum (ER) stress, a disequilibrium between the demand for ER function and the
capacity of the ER, is caused by various factors, such as viral infection, genetic mutations, and other
environmental factors. An elevated load of unfolded proteins in the ER induces the cells’ response to
ER stress by activating the unfolded protein response (UPR) pathway to restore the normal function
of ER [4]. It has been increasingly recognized that ER stress is associated with multiple biological
processes, including inflammation, bone loss, cell apoptosis, and extracellular matrix degradation,
and that it is involved in the progression of various diseases, such as inflammatory diseases and
diabetes mellitus [4,5]. Upregulation of ER stress has also been observed in periodontitis. ER stress
induced by \( P.\ gingivalis \) was proven to be involved in alveolar bone resorption in an experimental
periodontitis mouse model [6]. Additionally, ER stress was reported to be activated by advanced
glycation end products and to mediate inflammation in human periodontal ligament cells [7]. However,
the precise mechanisms by which ER stress mediates inflammatory responses in periodontal disease
remain unknown.

CCAAT/enhancer-binding protein \( \beta \) (C/EBP \( \beta \)) is a member of the family of transcription factors
that contain a basic leucine zipper (bZIP) domain at the C terminus. The C/EBP members play roles in
a wide range of cellular processes, such as cellular apoptosis, proliferation, adipocyte differentiation,
carbohydrate metabolism and inflammation [8,9]. There is emerging evidence that C/EBP \( \beta \) is
associated with ER stress and contributes to tumor cell death and migration [10,11]. Nevertheless, the
interplay and potential roles of C/EBP \( \beta \) and ER stress in human periodontitis remain to be elucidated.

In the present study, we aimed to determine the expression and role of C/EBP \( \beta \) in periodontal
ligament cells exposed to lipopolysaccharide (LPS). Furthermore, we investigated the role of ER stress,
and addressed whether and how ER stress interacts with C/EBP \( \beta \) in periodontal ligament cells. The
data obtained will enhance our understanding of human periodontitis pathology.

2. Results

2.1. C/EBP \( \beta \) Gene Expression Is Increased in Human Periodontal Ligament Cells (hPDLCs) Exposed to
Lipopolysaccharide (LPS)

To elucidate the role of C/EBP \( \beta \) in periodontal disease, we investigated the expression of the
C/EBP \( \beta \) gene in hPDLCs exposed to LPS. The response of hPDLCs to stimulation with \( P.\ gingivalis \)
LPS and \( E.\ coli \) LPS is shown in Figure 1A. Gene expression levels of pro-inflammatory cytokines IL-6
and IL-8 were markedly upregulated after stimulation with all stimuli. The increase in the levels of
IL-6 and IL-8 in hPDLCs in response to stimulation with 1 \( \mu \)g/mL \( P.\ gingivalis \) LPS was higher than
that in response to \( E.\ coli \) LPS (Figure 1A), indicating the suitability of LPS as a stimulant of hPDLCs.
Further, the mRNA and protein expression levels of C/EBP \( \beta \) were significantly increased in hPDLCs
stimulated with LPS in comparison with untreated (NT) cells. The increase in the expression level
of C/EBP \( \beta \) in hPDLCs in response to \( P.\ gingivalis \) LPS was higher than \( E.\ coli \) LPS (Figure 1B,C).
These data suggest that C/EBP \( \beta \) is aberrantly induced by LPS stimuli in hPDLCs. Thus, 1 \( \mu \)g/mL
\( P.\ gingivalis \) LPS was used in subsequent experiments.
with pAd/C/EBPβ Degradation Induced by LPS in hPDLCs

As seen in Figure 2B, C/EBPβ (Figure 2D). Consistent with the results in Figure 2B, we also observed that C/EBPβ overexpression increased the expression of cytokine IL-6 and IL-8 in hPDLCs not with different extent, suggesting that C/EBPβ is involved in the increase of IL-6 and IL-8 levels induced by LPS in hPDLCs.

Furthermore, we investigated the expression level of MMP-8 and MMP-9 in hPDLCs treated with pAd/C/EBPβ and LPS by Western blotting assay. The results shown in Figure 2C indicate that MMP-8 and MMP-9 expression were detected in hPDLCs stimulated by LPS. Following pAd/C/EBPβ transduction, the levels of MMP-8 and MMP-9 in hPDLCs were significantly increased. We then performed gelatizymography assay to determine the effect of C/EBPβ on MMP-9 activity. Compared to the control, the secretion of MMP-9 was significantly increased by C/EBPβ overexpression (Figure 2D). Consistent with the results in Figure 2B, we also observed that C/EBPβ overexpression increased the expression of MMP-8 and MMP-9 around 1.5-fold in unstimulated hPDLCs (data not shown), suggesting the regulatory role of transcription factors C/EBPβ in hPDLCs in response to LPS stimuli.

2.2. C/EBPβ Induces the Production of Pro-Inflammatory Cytokines and Enhances the Activation of Matrix Metalloproteinases (MMPs) in hPDLCs Stimulated by LPS

As an inflammatory response and extracellular matrix (ECM) degradation are the main pathological processes of periodontitis [12,13], we clarified the effect of C/EBPβ on the production of pro-inflammatory cytokines and the activation of MMPs, major enzymes that are responsible for the degradation of ECM, in hPDLCs stimulated by LPS. Firstly, a recombinant adenoviral vector pAd/C/EBPβ was constructed to overexpress C/EBPβ in cultured hPDLCs. The levels of C/EBPβ in cells treated with pAd/C/EBPβ transduction and LPS stimulation was confirmed using a Western blotting assay (Figure 2A). Then the levels of IL-6 and IL-8 were evaluated in hPDLCs by ELISA. As seen in Figure 2B, C/EBPβ overexpression increased the expression of cytokine IL-6 and IL-8 in hPDLCs stimulated by LPS or not with different extent, suggesting that C/EBPβ is involved in the increase of IL-6 and IL-8 levels induced by LPS in hPDLCs.

2.3. Endoplasmic Reticulum (ER) Stress Regulates Inflammatory Response and Extracellular Matrix Degradation Induced by LPS in hPDLCs

Previous studies revealed that ER stress plays a crucial role in the inflammatory response and metabolism of ECM during normal physiological and pathological processes [2,12]. C/EBPβ is associated with ER stress and contributes to cell death, migration, and inflammation [4,14]. Therefore we attempted to determine the association between C/EBPβ and ER stress in the progression

Figure 1. Expression of CCAAT/enhancer-binding protein β (C/EBPβ) gene in human periodontal ligament cells (hPDLCs) stimulated by lipopolysaccharide (LPS). (A) hPDLCs were incubated with 1 μg/mL Porphyromonas gingivalis (P. gingivalis) LPS (Pg LPS) or E. coli LPS (Ec LPS) for 24 h. The levels of IL-6 and IL-8 in hPDLCs were measured using an ELISA kit; then, the mRNA (B) and protein (C) levels of C/EBPβ in hPDLCs were analyzed by RT-PCR and Western blotting, respectively (* p < 0.05, ** p < 0.05 vs. not treated (NT); # p < 0.05 vs. Pg LPS).
of periodontitis. Firstly, activation of the ER stress signaling pathway induced by LPS stimuli was observed in hPDLCs by measuring the activation of ER stress-related marker molecules protein kinase-like ER kinase (PERK) and eIF2α. As seen in Figure 3A, the phosphorylation of both PERK and eIF2α was significantly upregulated. Next, we examined the expression of GRP78/Bip and CHOP, another two marker genes of ER stress, in hPDLCs. Western blotting assay indicated that GRP78/Bip and CHOP expression increased in hPDLCs after stimulation by LPS (Figure 3B), implying the activation of ER stress.

Figure 2. Effect of C/EBP β on production of pro-inflammatory cytokines and activation of matrix metalloproteinases (MMPs) in hPDLCs exposed to LPS. hPDLCs were transfected with a recombinant adenoviral vector pAd/C/EBP β or Ad-GFP control vector for 48 h, followed by stimulation with P. gingivalis LPS for another 24 h. (A) The protein level of C/EBP β in hPDLCs was determined by Western blotting; (B) The levels of IL-6 and IL-8 were analyzed by ELISA assay in hPDLCs transfected with adenoviral vector pAd/C/EBP β or Ad-vector in the presence or absence of LPS; (C) MMP-8 and MMP-9 levels in hPDLCs stimulated with LPS were analyzed by Western blotting assay; (D) gelatin zymography was performed to determine MMP-9 activity in hPDLCs stimulated with LPS. Untransfected hPDLCs were stimulated by P. gingivalis LPS and used as the control in (A,C,D). ($p < 0.05$ vs. NT; # $p < 0.05$ vs. LPS; * $p < 0.05$, ** $p < 0.05$ vs. control).

We then employed the ER stress inhibitor salubrinal (Calbiochem, San Diego, CA, USA) and inducer tunicamycin (Sigma, St. Louis, MO, USA) to elucidate the role of ER stress in LPS-treated hPDLCs. The inhibitory and inducing efficiency of salubrinal and tunicamycin were confirmed by detecting the expression of GRP78/Bip and CHOP (Figure 3C). Further analysis showed that salubrinal treatment successfully repressed, but tunicamycin increased, the secretion of IL-6 and IL-8 in hPDLCs stimulated by LPS or not, with different extents, suggesting that ER stress is involved in the increase of IL-6 and IL-8 levels induced by LPS in hPDLCs (Figure 3D). The expression of MMP-8 and MMP-9 were...
also decreased by salubrinal, but increased by tunicamycin, in hPDLCs stimulated by LPS (Figure 3E). Consistent with the results in Figure 3D, we also observed that the expression of MMP-8 and MMP-9 in unstimulated hPDLCs was only modestly affected by ER stress inhibition/induction (data not shown). These results suggest that LPS-induced ER stress plays a regulatory role in the inflammatory response and ECM degradation in hPDLCs in response to LPS stimuli.

**Figure 3.** The role of ER stress induced by LPS in hPDLCs. hPDLCs were incubated with 1 μg/mL *P. gingivalis* LPS for 24 h. (A) The phosphorylation of protein kinase-like ER kinase (PERK), eIF2α and (B) the expression of GRP78/Bip and C/EBP homologous protein (CHOP) in hPDLCs were measured by Western blotting (*p < 0.05 vs. NT); (C) hPDLCs were pretreated with 5 μg/mL tunicamycin or 10 μM salubrinal for 2 h, followed by stimulation with *P. gingivalis* LPS for 24 h. The levels of GRP78/Bip and CHOP in hPDLCs were measured by Western blotting to confirm the efficiency of salubrinal and tunicamycin; (D) the levels of IL-6 and IL-8 were analyzed by ELISA in hPDLCs treated with salubrinal or tunicamycin in the presence or absence of LPS; and (E) Western blotting was used to measure the expression of MMP-8 and MMP-9 in hPDLCs stimulated with LPS. Untreated hPDLCs were stimulated by *P. gingivalis* LPS and used as the control. ($p < 0.05$ vs. NT; # $p < 0.05$ vs. LPS; * $p < 0.05$ vs. control).

### 2.4. C/EBP β Mediates the Regulatory Role of ER Stress in hPDLCs

To further investigate the interplay between ER stress and C/EBP β in hPDLCs in response to LPS stimuli, the levels of C/EBP β were confirmed using Western blotting in hPDLCs treated with ER stress inducer tunicamycin or inhibitor salubrinal in the presence of LPS stimulation. As shown in Figure 4A, tunicamycin treatment significantly increased the expression level of C/EBP β in hPDLCs. Compared with untreated cells, however, C/EBP β expression was markedly downregulated in hPDLCs after incubation with the ER stress inhibitor salubrinal.
Figure 4. Interplay between C/EBPβ and endoplasmic reticulum (ER) stress in hPDLCs. (A) hPDLCs were pretreated with 5 μg/mL tunicamycin or 10 μM salubrinal for 2 h, followed by stimulation with P. gingivalis LPS for 24 h. The expression of C/EBPβ in hPDLCs was then analyzed by Western blotting; (B) hPDLCs were transfected with C/EBPβ siRNA or non-targeting control siRNA (s-Mock) for 48 h, followed by stimulation with P. gingivalis LPS for another 24 h. The protein level of C/EBPβ was determined by Western blotting to confirm the efficiency of siRNA; (C, D) C/EBPβ siRNA-transfected hPDLCs were pretreated with tunicamycin, followed by stimulation with P. gingivalis LPS. The levels of IL-6 and IL-8 and the expression of MMP-8 and MMP-9 in hPDLCs were then analyzed by ELISA and Western blotting, respectively. Untransfected hPDLCs treated with LPS were used as the control. (*p < 0.05, **p < 0.05 vs. control).

Furthermore, we introduced the C/EBPβ siRNA to verify whether or not C/EBPβ is responsible for the ER stress signaling pathway in hPDLCs. The blocking efficiency of C/EBPβ siRNA was confirmed by Western blotting assay (Figure 4B). The results showed that C/EBPβ siRNA transfection resulted in a significant decrease in the enhanced secretion of IL-6 and IL-8 and expression of MMP-8 and MMP-9 induced by tunicamycin treatment in hPDLCs (Figure 4C, D). Consistent with the results in Figures 2B and 3D, we also observed that the expression of MMP-8 and MMP-9 in unstimulated hPDLCs was only modestly affected by C/EBPβ/ER stress manipulation (data not shown). Taken together, these results provide evidence that LPS-induced ER stress in hPDLCs activates the expression of C/EBPβ, which is involved in the regulation of ER stress in hPDLCs in response to LPS stimuli.

3. Discussion

Periodontitis is a chronic infectious disease that affects the integrity of tooth-supporting periodontium. Within the periodontium, the periodontal ligament is the key connecting tissue
among teeth and other tooth-supporting tissues, including alveolar bone, gingiva, parodontium and cementum alveolar bone [2]. Periodontal ligament cells (PDLs) are fibroblast-like cells that can express several ECM-related molecules. hPDLs also exhibit increased pro-inflammatory mediators and cytokines production in response to bacterial infection [15]. Generally, periodontal health requires a maintainable microbiota homeostasis and immuno-inflammatory state in the periodontium, in which hPDLs play crucial roles [16]. In periodontitis, the host immune homeostasis is dysregulated by oral bacteria [17,18]. Disruption of oral microorganisms has been considered the traditional cause of periodontitis. *P. gingivalis* is recognized as the major cause of periodontitis. Therefore, most studies have primarily used *P. gingivalis* as the model pathogen to stimulate hPDLs in vitro when investigating the periodontal host immune response to microbial stimuli [19]. In the present study, *P. gingivalis* LPS was proven to stimulate hPDLs to produce pro-inflammatory cytokines IL-6 and IL-8, and was used as a stimulus for hPDLs in subsequent experiments.

C/EBPβ, a member of the C/EBP family of transcription factors, regulates the expression of target genes that are involved in cellular proliferation, apoptosis, and migration. Accumulating evidence has demonstrated the role of C/EBPβ in mediating adipocyte differentiation and neoplastic metabolism [20–22]. Increased C/EBPβ expression in mice is capable of inducing hepatic steatosis that may lead to the occurrence of nonalcoholic steatohepatitis [23]. C/EBPβ also induces MMP1/3 expression via the p38 mitogen-activated protein kinase (MAPK) pathway to mediate tumor necrosis factors (TNF)-α-induced cancer cell migration, contributing to the regulation of cancer metastasis [24]. In terms of inflammation, C/EBPβ plays key roles in the control of inflammation and host immunity by regulating the functions of immunologic effector cells (macrophages and granulocytes) and target cells [8,9]. Zimmermann et al. recently reported that C/EBPβ is involved in the regulation of serotonin transporter 5-HTT expression in macrophages in response to bacterial LPS stimuli [21]. Additionally, the C/EBP family of transcription factor also have regulatory roles in periodontal development and pathology. Savage et al. proved that p20C/EBPβ, a negative C/EBP isoform, reduced alveolar bone mass and site-specific dentin dysplasia in transgenic mice [25]. C/EBPα, another member of the C/EBP family, was reported to regulate the lineage commitment of osteoclasts, which play a critical role in periodontal diseases [26]. Our data here show that the expression level of C/EBPβ was significantly upregulated in hPDLs after stimulation by LPS, and C/EBPβ overexpression increased the secretion of pro-inflammatory cytokines and the activation of MMPs, which are major mediators of the pathological progression of periodontitis, suggesting the important role of the transcription factor C/EBPβ in periodontal diseases.

Another approach we took to identify the role of C/EBPβ was to elucidate the interplay between ER stress and C/EBPβ in hPDLs in response to LPS stimuli. ER stress is a self-adaptive and survival pathway of eukaryotic cells in response to nervous physiological states or stimuli. It is well known that ER stress is involved in multiple biological processes and various diseases, including periodontitis [12,27]. In mammalian cells, three ER-localized transmembrane proteins: double-stranded RNA-dependent protein kinase-like ER kinase (PERK), inositol-requiring 1 (IRE-1), and activating transcription factor 6 (ATF6), are primary initiators and transducers of ER stress signaling cascades. In inflammatory diseases, the PERK, IRE-1, and ATF6 pathways have been identified to play crucial roles in ER stress-mediated inflammatory response by regulating NF-κB and c-Jun N-terminal kinase (JNK) activation [4]. Emerging evidence has shown that C/EBPβ is associated with ER stress and contributes to cell death, migration, and inflammation. Meir et al. reported that C/EBPβ regulates ER stress-induced apoptosis in mouse and human cells [14]. In addition, C/EBPβ knockdown moderates inflammation and lipid accumulation in the mouse model of nonalcoholic steatohepatitis [23]. Nevertheless, the interaction and potential roles of C/EBPβ and ER stress in human periodontitis are not well defined. Herein, we determined that ER stress is induced by LPS and plays a regulatory role in the inflammatory response and ECM degradation in hPDLs in response to LPS stimuli. Furthermore, C/EBPβ was found to be activated by ER stress in hPDLs after stimulation with LPS and, thus, might be involved in the regulation of ER stress in hPDLs in response to LPS.
stimuli. These observations are consistent with a previous study demonstrating that the expression of human C/EBP β is activated by ER stress via the unfolded protein response element downstream of the coding gene [28]. However, our study is not without its limitations. How C/EBP β is involved in ER stress should be investigated in a subsequent study.

In summary, we have demonstrated the novel role of C/EBP β in hPDLCs exposed to LPS, and shown that ER stress induced by LPS plays a regulatory role in the inflammatory response and ECM degradation in hPDLCs in response to LPS stimuli by activating C/EBP β expression. This provides new insights into the pathology of human periodontitis.

4. Experimental Section

4.1. Cell Culture

Human periodontal ligament cells (hPDLCs) were obtained from healthy donors undergoing orthodontic treatment as described previously [19]. Briefly, after tooth extraction, the middle portion of root surfaces was scraped to obtain human periodontal ligament (hPDL) tissue. The PDL tissue was cut into small pieces and washed with Dulbecco’s modified Eagle medium (DMEM) (Sigma, St. Louis, MO, USA), followed by incubation in DMEM medium containing collagenase for 2 h at 37 °C. The isolated cells were then washed three times and cultured in DMEM, supplemented with 10% fetal bovine serum (FBS) (Invitrogen, Carlsbad, CA, USA) and 1% penicillin/streptomycin (Sigma) at 37 °C in a 5% CO₂ atmosphere. Cells from passages 3–5 were used for subsequent experiments. Approval was obtained from the Ethics Committee of School of Stomatology of Fourth Military Medical University (24 September 2014, NO. 20140911). All subjects gave informed consent.

4.2. LPS Preparation and Stimulation

Ultrapure P. gingivalis LPS and Escherichia coli LPS were obtained from Invivogen (San Diego, CA, USA). The hPDL-derived cells were grown to approximately 80% confluence in 24-well plates and stimulated with 1 µg/mL P. gingivalis LPS or E. coli LPS for 24 h. Cells were then harvested for RNA and protein extraction.

4.3. Cell Transfection

The recombinant adenoviral vector pAd/C/EBP β was constructed. Briefly, full-length C/EBP β cDNA was amplified by RT-PCR from total RNA and subcloned into the pAdTrack-cytomegalovirus (CMV) vector. Then the recombinant pAd/C/EBP β vector was generated according to the manufacturer’s protocol. Adenoviruses without apparent cytotoxicity were used at 20 multiplicity of infection (moi) for 48 h for hPDLCs transduction. Adenovirus vector (Ad-GFP) encoding green fluorescent protein (GFP) gene was used as the control.

The siRNA targeting C/EBP β mRNA was synthesized by Takara (Takara, Dalian, China). hPDLCs were cultured in 96-well plates to approximately 80% confluence and transfected with C/EBP β siRNA or non-targeting negative control siRNA using Lipofectamine 2000 (Invitrogen) for 48 h. Cells were then stimulated with P. gingivalis LPS for another 24 h and harvested for subsequent experiments. The transfection efficacy was determined by Western blotting assays.

4.4. Quantitative RT-PCR

Trizol reagent (Sigma) was applied to isolate total RNA in hPDLCs. Expression of C/EBP β was detected using the CellAmp Direct RNA Prep kit (Takara) for qPCR and the Protein Analysis kit (Takara). The primers were as follows: C/EBP β: 5'-CAGCGCCGTCCTCTCCC-3' (forward) and 5'-CAATGAAAAACCCCAAGGAAAC-3' (reverse); Glyceraldehyde-3-phosphate dehydrogenase (GAPDH): 5'-ACCACATGCATGCCCATC-3' (forward) and 5'-ACCACATCCATGCATCAC-3' (reverse). The reaction was performed as follows: 10 min 95 °C; 40 cycles of 1 min 95 °C, 2 min 63 °C,
1 min 72 °C; and 10 min 72 °C for final annealing. Cycle threshold (Ct) values of GAPDH were used as the internal control.

4.5. Western Blotting

Cultured cells were lysed with lysis buffer (20 mmol/L 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES), 5 mmol/L KCl, 25 mmol/L MgCl₂, 1% Triton X-100, and 0.5% phenylmethylsulfonyl fluoride (PMSF) to obtain total proteins. Equal amounts of proteins were separated by SDS-PAGE gels (Invitrogen), then electrophoretically transferred to polyvinylidene fluoride (PVDF) membrane. The blots were incubated with primary antibodies (mouse anti-C/EBP β 1:1000 dilution, Abcam, Cambridge, UK; rabbit anti-matrix metallopeptinases (MMP)-9 1:2000 dilution, rabbit anti-PERK, GRP78/Bip 1:3000, Abcam; mouse anti-CHOP 1:1000, (Pierce, Rockford, IL, USA); mouse anti-phospho-PERK, phospho-eIF2α 1:1000 dilution (Cell Signaling Technology, Danvers, MA, USA) and mouse anti-β-actin 1:3000 dilution (ABclonal, College Park, MD, USA)) overnight at 4 °C, and labeled with horseradish peroxidase-conjugated secondary antibodies for 1 h at room temperature. Protein bands were then detected using enhanced chemiluminescence (ECL, Amersham Pharmacia, Piscataway, NJ, USA) and measured by Image Quant software ev2003.03 (GE Healthcare, Pittsburgh, NJ, USA).

4.6. ELISA

The hPDL-derived cells were stimulated with 1 µg P. gingivalis LPS or E. coli LPS for 24 h at 37 °C. The treated cells were measured for the secretion of IL-6 and IL-8 using an ELISA kit (Boster, Wuhan, China) according to the manufacturer’s instructions.

4.7. Gelatin Zymography

MMP-9 activity was measured by gelatin zymography assay as previously described [29]. In brief, proteins (80 mg) were collected from cells and separated by SDS-PAGE containing 0.1% gelatin at 4 °C. Then the gels were washed and incubated in buffer (10 mM CaCl₂, 50 mM Tris–HCl (pH 7.5)) at 37 °C. After staining and destaining, bands were analyzed using Image Quant software.

4.8. Statistical Analysis

The data are presented as mean ± SEM. Statistical differences were analyzed by Student’s t test using GraphPad Prism 6 (La Jolla, CA, USA). All experiments were performed at least three times. A value of p < 0.05 was considered to be statistically significant.

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References

1. Hajishengallis, G. Periodontitis: From microbial immune subversion to systemic inflammation. Nat. Rev. Immunol. 2015, 15, 30–44. [CrossRef] [PubMed]
2. Darveau, R.P.; Tanner, A.; Page, R.C. The microbial challenge in periodontitis. Periodontology 1997 1997, 14, 12–32. [CrossRef]
3. Pink, C.; Kocher, T.; Meisel, P.; Dorr, M.; Markus, M.R.; Jablonowski, L.; Groteventd, A.; Nauck, M.; Holtfreter, B. Longitudinal effects of systemic inflammation markers on periodontitis. J. Clin. Periodontol. 2015, 42, 988–997. [CrossRef] [PubMed]
4. Kezhong, Z.; Kaufman, R.J. From endoplasmic-reticulum stress to the inflammatory response. Nature 2008, 454, 455–462.

5. Uehara, Y.; Hirose, J.; Yamabe, S.; Okamoto, N.; Okada, T.; Oyadomari, S.; Mizuta, H. Endoplasmic reticulum stress-induced apoptosis contributes to articular cartilage degeneration via C/EBP homologous protein. Osteoarthr. Cartil. 2014, 22, 1007–1017. [CrossRef] [PubMed]

6. Yamada, H.; Nakajima, T.; Domon, H.; Honda, T.; Yamazaki, K. Endoplasmic reticulum stress response and bone loss in experimental periodontitis in mice. J. Periodontal Res. 2015, 50, 500–508. [CrossRef] [PubMed]

7. Xu, J.; Xiong, M.; Huang, B.; Chen, H. Advanced glycation end products upregulate the endoplasmic reticulum stress in human periodontal ligament cells. J. Periodontol. 2015, 86, 440–447. [CrossRef] [PubMed]

8. Ko, C.Y.; Chang, W.C.; Wang, J.M. Biological roles of CCAAT/enhancer-binding protein β during inflammation. J. Biomed. Sci. 2015, 22, 1–8. [CrossRef] [PubMed]

9. Poli, V. The role of C/EBP isoforms in the control of inflammatory and native immunity functions. J. Biol. Chem. 1998, 273, 29279–29282. [CrossRef] [PubMed]

10. Chiribau, C.B.; Gaccioli, F.; Huang, C.C.; Yuan, C.L.; Hatzoglou, M. Molecular symbiosis of CHOP and C/EBP β isoform LIP contributes to endoplasmic reticulum stress-induced apoptosis. Mol. Cell. Biol. 2010, 30, 3722–3731. [CrossRef] [PubMed]

11. Arensdorf, A.M.; Rutkowski, D.T. Endoplasmic reticulum stress impairs IL-4/IL-13 signaling through C/EBP expression dependent on p38 MAPK. J. Cell Biochem. 2015, 116, 2766–2777. [CrossRef] [PubMed]
25. Savage, T.; Bennett, T.; Huang, Y.F.; Kelly, P.L.; Durant, N.E.; Adams, D.J.; Mina, M.; Harrison, J.R. Mandibular phenotype of p20C/EBPβ transgenic mice: Reduced alveolar bone mass and site-specific dentin dysplasia. *Bone* 2006, 39, 552–564. [CrossRef] [PubMed]

26. Wei, C.; Guochun, Z.; Liang, H.; Mengrui, W.; Hongliang, C.; Yi-Ping, L. C/EBPα regulates osteoclast lineage commitment. *Proc. Natl. Acad. Sci. USA* 2013, 110, 7294–7299.

27. Domon, H.; Takahashi, N.; Honda, T.; Nakajima, T.; Tbeta, K.; Abiko, Y.; Yamazaki, K. Up-regulation of the endoplasmic reticulum stress-response in periodontal disease. *Clin. Chim. Acta* 2009, 401, 134–140. [CrossRef] [PubMed]

28. Chen, C.; Dudenhausen, E.E.; Pan, Y.X.; Zhong, C.; Kilberg, M.S. Human CCAAT/enhancer-binding protein β gene expression is activated by endoplasmic reticulum stress through an unfolded protein response element downstream of the protein coding sequence. *J. Biol. Chem.* 2004, 279, 27948–27956. [CrossRef] [PubMed]

29. Fiorentini, C.; Bodei, S.; Bedussi, F.; Fragni, M.; Bonini, S.A.; Simeone, C.; Zani, D.; Berruti, A.; Missale, C.; Memo, M. GPNMB/OA protein increases the invasiveness of human metastatic prostate cancer cell lines DU145 and PC3 through MMP-2 and MMP-9 activity. *Exp. Cell Res.* 2014, 323, 100–111. [CrossRef] [PubMed]