Gorilla Class I Major Histocompatibility Complex
Alleles: Comparison to Human and Chimpanzee
Class I
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Summary
14 gorilla class I major histocompatibility complex (MHC) alleles have been isolated, sequenced, and compared to their counterparts in humans and chimpanzees. Gorilla homologues of HLA-A, -B, and -C were readily identified, and four Gogo-A, four Gogo-B, and five Gogo-C alleles were defined. In addition, an unusual Gogo class I gene with features in common with HLA-A and its related pseudogene, HLA-H, is described. None of the gorilla alleles is identical or even closely related to known class I alleles and each encodes a unique antigen recognition site. However, the majority of polymorphic substitutions and sequence motifs of gorilla class I alleles are shared with the human or chimpanzee systems. In particular, elements shared with HLA-A2 and HLA-B27 are found in Gogo-A and -B alleles. Diversity at the Gogo-B locus is less than at the Gogo-A locus, a trend the opposite of that seen for HLA-A and -B. The Gogo-C locus also appears to have limited polymorphism compared to Gogo-A. Two basic Gogo-C motifs were found and they segregate with distinctive sets of HLA-C alleles. HLA-A alleles are divided into five families derived from two ancient lineages. All chimpanzee A alleles derived from one of these lineages and all gorilla alleles derive from the other. Unlike chimpanzee Patr-A alleles, the Gogo-A alleles do not clearly partition with one of the HLA-A families but have similarities with two. Overall, gorilla class I diversity appears from this sampling to show more distinctions from class I HLA than found for chimpanzee class I.

The number of class I genes in the MHC varies greatly between species, as does the extent of their polymorphism (1, 2). In humans, some 17 class I genes, pseudogenes, and gene fragments have been mapped to the HLA region (3), and of these, HLA-A, -B, and -C have the widespread tissue distribution and well-developed polymorphism that characterize genes involved in presenting antigens to CD8+ T cells (4). Sequence variability is focused on amino acid substitutions that change the peptide binding specificity of the antigen recognition site (5), and suggests the polymorphism results from selection for molecules that present different environmental antigens.

Comparison of alleles of the HLA-A, -B, and -C loci suggests various mutagenetic events have contributed to the accumulated polymorphism. In addition to point substitutions and intragenic recombinations (6) are recombinatorial mechanisms that move short segments of homologous sequence between alleles of the same locus and, to a lesser extent, alleles of different loci (5). Although certain alleles can easily be related in this way, others differ by highly distinctive structural motifs for which evolutionary intermediates no longer exist. Further understanding of the evolution of class I HLA alleles may be obtained by comparison to their homologues in related species. For humans the closest relatives are the chimpanzees and the gorillas, species estimated to have all shared a common ancestor 7–10 million years ago (7–9).

Previous analysis of 10 chimpanzee class I alleles revealed a remarkable degree of similarity with HLA-A, -B, and -C alleles (10, 11). No species specific features were found and particular similarities between individual alleles in the two species were identified. Despite the similarities no example of an allele that is identical in the two species has been discovered and almost all the substitutions are at positions of the molecule predicted to alter peptide presentation. In addition, the impression obtained from what is still a limited sampling of alleles is that diversity and polymorphism in chimpanzees is but a fraction of that found in humans (12). To provide an independent assessment of these issues we now examine the class I alleles of the gorilla.

Materials and Methods
EBV-transformed cell lines were established from Calabar, Inaki, Banga, and Oko, lowland gorillas (Gorilla gorilla) housed at Yerkes Regional Primate Center (Atlanta, GA). Details concerning the
individuals (age, sex, and tissue typing) and the derivation of the cell lines have been described (12). Although the ancestry of these individuals is not precisely known, there is no evidence to suggest they are related. Analysis of the class I alleles provides the following genotypes: Banga, Gogo-A*0401, -B*0101, -B*0103, -C*0101, and -C*0201; Oko, Gogo-Oko, -B*0201, and -C*0202; Calabar, Gogo-A*0101, -B*0102, -B*0103, -C*0102, and -C*0203; Inaki, Gogo-A*0201, -A*0501, -B*0101, and -B*0201. Cell lines have been deposited with the American Type Culture Collection (Rockville, MD).

The conditions for RNA extraction, first-strand cDNA synthesis, and PCR were as previously described (13). For this study, a universal 5' amplification primer, which hybridizes to all HLA class I alleles, was used. The 3' amplification primers were designed to hybridize to locus-specific regions within the 3' untranslated sequence. The reagents and procedures used by Ennis et al. (13) to target specific human class I loci were used in this study and amplification performed using the universal 5' primer in conjunction with the "locus specific" primers consisted largely of product from the target locus. Sequences of the amplification primers are: HLA-A2loc, 5'-CCGCAAGCTTCTGGGGAGGAAACACAGGTCAGCATGGGAGGGAGCACAGGTCAC-CG'IGGGAAG-3*; HLA-B44loc, 5'-AACGAAACGCAGGCAGAGGAG-CACAGGTCAGTGTGGGGAC-3'.

The amplification protocol consisted of 30 cycles of denaturation (60 s at 94°C), annealing (60 s at 65°C), and extension (90 s at 72°C). Extension times were increased by 2 s with each cycle. After the final cycle, the amplification tubes were incubated at 72°C for 10 min. The amplification products were extracted and back-extracted with phenol/chloroform, ethanol precipitated, and resuspended in H2O before sequential digestions with Sall and HindIII. The digested products were ligated to similarly cut M13mpl8 and mp19 before transformation of Escherichia coli JM109. Plaques were inoculated onto fresh cultures of JM109 in 2x yeast tryptone (YT) for overnight incubation, and single-stranded DNA was prepared from the phage lysates. Preliminary DNA sequencing was performed to group clones with identical sequence. At least three clones per locus were sequenced from the phage lysates. Preliminary DNA sequencing was performed to identify sequence differences (16-57 nucleotides) within that defined by pairwise comparisons among HLA-A alleles (2-62 nucleotides). This is true for some of the gorilla A-like sequences but not all. In particular, the Gogo-Oko sequence greatly extends the range of differences in pairwise comparisons and inspection shows this sequence has unusual features.

Closely linked to HLA-A are at least two structurally related pseudogenes with sequence motifs similar to HLA-A at the locus-specific positions and in the 3' untranslated region. Best characterized of these pseudogenes is HLA-H, which is postulated to have been an antigen-presenting locus that was inactivated through the acquisition of one or two single base pair mutations (19). Of interest here is that the Gogo-Oko sequence has similarities with HLA-H. Most striking is a 210-bp segment of exon 2 (positions 74-286) that is nearly identical in Gogo-Oko and HLA-H (Fig. 1). At the 3' end of the coding region, there are additional similarities including a termination codon distinct from those found in HLA-A and other class I HLA genes.

The patterns of substitution at locus-specific positions indicate that Gogo-Oko is equally related to both HLA-A and HLA-H and a similar conclusion is reached from overall comparisons of the sequence. For example, the range of differences between Gogo-Oko and HLA-A alleles is 62-94 nucleotides compared to 76-81 nucleotides for HLA-H alleles. Comparison of individual exons shows that Gogo-Oko is more similar to HLA-A (and other Gogo-A) alleles in exon 3 and to HLA-H alleles in exon 2 (Table 1). It is highly unlikely that the unusual pattern of sequence motifs in Gogo-Oko is the result of a PCR or cloning artifact, as an almost identical allele has been isolated by Watkins et al. (20) from another individual gorilla. These data suggest three possibilities for the nature of Gogo-Oko: first, that it is an allele of the Gogo-A locus that has been the recipient of a gene conversion(s) from an HLA-H-like gene; second, that it is an allele of an HLA-H-like gene that has been the recipient of a gene conversion(s) from a Gogo-A-like gene; and third, that the patterns of nucleotide substitution at a set of 46 "locus-specific" positions (18). Testing the 14 gorilla class I sequences by these two complementary criteria clearly shows that five segregate with HLA-A sequences, four with HLA-B sequences, and five with HLA-C sequences (Figs. 2 and 3). At the 46-locus-specific positions, two of the gorilla alleles, Gogo-A*0201 and Gogo-A*0401, have patterns of substitution identical to the corresponding HLA locus, and none have <37 identities (Fig. 3). Furthermore, the similarities seen in the coding region extend into the 3' untranslated sequence, a region in HLA-A, -B, and -C sequences that is divergent between loci but relatively homogenous within the alleles of a locus (Fig. 4). We find therefore that the gorilla, like the chimpanzee, has clearly recognizable HLA-A, -B, and -C-like class I MHC loci. These results demonstrate that the origin of this basic arrangement of expressed class I genes predates the divergence of humans, chimpanzees, and gorillas.

Results

The Gorilla MHC Has Class I Genes Similar to HLA-A, -B, and -C. 14 different class I heavy chain cDNA were cloned from four gorilla EBV-transformed B cell lines, and their nucleotide sequences were determined (Fig. 1). That locus-specific oligonucleotides based on the sequences of either HLA-A, -B, or -C alleles efficiently primed polymerase chain amplification from gorilla cDNA indicated the presence of gorilla genes related to these human class I HLA genes, and sequence comparison showed this to be the case. Alleles of the HLA-A, -B, and -C loci can be distinguished unequivocally on the basis of overall levels of sequence similarity and
### Comparison of Gorilla Class I Alleles to Human and Chimpanzee Class I

| Gorilla | Human | Chimpanzee |
|---------|-------|------------|
| Gogo-A*0101 | TCGGCGGGTAC | CCCACCCAGAC | GACCACCTTCG | CATGAGGACG | TGGCGGCCG | GAGGAGGACC | TCCCGCCTTG | GACCCTGCAG | GCAGGCTTG |
| Gogo-A*0201 | TGGGCGGGTAC | CCCACCCAGAC | GACCACCTTCG | CATGAGGACG | TGGCGGCCG | GAGGAGGACC | TCCCGCCTTG | GACCCTGCAG | GCAGGCTTG |
| Gogo-A*0401 | TGGGCGGGTAC | CCCACCCAGAC | GACCACCTTCG | CATGAGGACG | TGGCGGCCG | GAGGAGGACC | TCCCGCCTTG | GACCCTGCAG | GCAGGCTTG |
| Gogo-A*0501 | TGGGCGGGTAC | CCCACCCAGAC | GACCACCTTCG | CATGAGGACG | TGGCGGCCG | GAGGAGGACC | TCCCGCCTTG | GACCCTGCAG | GCAGGCTTG |

**Table Notes:**
- **Gogo** represents Gorilla Class I alleles.
- **HLA-H** represents human Class I alleles.
- **Patr** represents chimpanzee Class I alleles.

**Sequence Comparison:**

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| Gorilla | Human | Chimpanzee |
|---------|-------|------------|
| Gogo-A*0101 | TCGGCGGGTAC | CCCACCCAGAC | GACCACCTTCG | CATGAGGACG | TGGCGGCCG | GAGGAGGACC | TCCCGCCTTG | GACCCTGCAG | GCAGGCTTG |
| Gogo-A*0201 | TGGGCGGGTAC | CCCACCCAGAC | GACCACCTTCG | CATGAGGACG | TGGCGGCCG | GAGGAGGACC | TCCCGCCTTG | GACCCTGCAG | GCAGGCTTG |
| Gogo-A*0401 | TGGGCGGGTAC | CCCACCCAGAC | GACCACCTTCG | CATGAGGACG | TGGCGGCCG | GAGGAGGACC | TCCCGCCTTG | GACCCTGCAG | GCAGGCTTG |
| Gogo-A*0501 | TGGGCGGGTAC | CCCACCCAGAC | GACCACCTTCG | CATGAGGACG | TGGCGGCCG | GAGGAGGACC | TCCCGCCTTG | GACCCTGCAG | GCAGGCTTG |

**Note:** The sequences are aligned to highlight similarities and differences across the class I alleles.

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**Figure Description:**

1. **Sequence Alignment:** Comparison of Gorilla Class I alleles (Gogo) with Human (HLA-H) and Chimpanzee (Patr) Class I alleles.
2. **Alignment Criteria:**
   - **G** for Gorilla
   - **H** for Human
   - **P** for Chimpanzee
3. **Alignment Approach:** Use of dashes (-) to indicate gaps.

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Figure 1. The nucleotide sequences of the coding region of 14 gorilla class I alleles. Sequences are compared to Gogo-A*0101. Dashes signify identity and dots denote gaps introduced for optimal alignment. In bold are substitutions from the consensus in the regions of identity between Gogo-Oko and HLA-H, the polymorphism in exon 5 that is shared by Gogo-B and Patr-B alleles, and the unusual termination codon shared by Gogo-Oko and Patr-B*01

1. The nucleotide sequences of the coding region of 14 gorilla class I alleles. Sequences are compared to Gogo-A*0101. Dashes signify identity and dots denote gaps introduced for optimal alignment. In bold are substitutions from the consensus in the regions of identity between Gogo-Oko and HLA-H, the polymorphism in exon 5 that is shared by Gogo-B and Patr-B alleles, and the unusual termination codon shared by Gogo-Oko and Patr-B*01
it represents an allele of a locus that is distinct from Gogo-A and not the direct homologue of either HLA-A or HLA-H. Further insight into the unusual divergence of Gogo-Oko is gained by an examination of the relationship of A-like sequences of the gorilla to individual families of HLA-A alleles.

On the basis of nucleotide sequence, all HLA-A alleles can be grouped into five families (A2, A3, A9, A10, and A19), which are shaded solid; those from pairs of alleles from the same locus (A v A, B v B, C v C) are stippled. Differences of 25-62 nucleotides between families (Fig. 5a).

Figure 2. Pairwise comparison of gorilla (Gogo) class I alleles with HLA-A, -B, and -C alleles. (A) The frequency histogram obtained from all pairwise comparisons between 74 HLA-A, -B, and -C alleles. Differences from pairs involving alleles of different loci (A v B, A v C, B v C) are shaded solid; those from pairs of alleles from the same locus (A v A, B v B, C v C) are stippled. (B) The range of differences when the groups of Gogo-A, -B, or -C alleles are compared in a pairwise fashion to HLA-A, -B, or -C alleles.

3, 14 in exon 4, and 11 in exons 5-8) were identified. Although only two alternative nucleotides are found at any of these positions, their combination provides an unequivocal fingerprint for each family (Fig. 6). The motifs at the family-specific positions further substantiate the similarities between the A3 and A9 families on the one hand and the A2, A9, and A10 families on the other. These observations suggest that five modern HLA-A alleles derive from two ancestral families: one that gave rise to the HLA-A3 and HLA-A9 families, the other to the HLA-A2, A10, and A19 families. Analysis of the pattern of substitution at the family-specific positions in the Gogo-A sequences substantiates this hypothesis.

Figure 3. Patterns of substitution for ape class 1 alleles at 46 locus-specific positions defined from comparisons of HLA-A, -B, and -C alleles. At each locus-specific position, all alleles of an individual HLA locus are identical, but differences exist between the loci. The number of these positions has been reduced from the 62 compared in a previous analysis (18), due to the inclusion of a recently discovered group of divergent HLA-C alleles. For each locus, the ape sequences are compared to the human consensus; dark lines indicate identity. Sequences from gorilla (Gogo), chimpanzee (Patr), and orangutan (Popy) are included. The position numbers correspond to those of Fig. 1.
Table 1. Range of Nucleotide Differences with Gogo-Oko

|                        | HLA-A   | Gogo-A  | HLA-H  | Popy-A | Patr-A |
|------------------------|---------|---------|--------|--------|--------|
| Entire coding region   | 62-94*  | 69-85   | 76-81  | 78     | 68-75  |
| Exon 1                 | 5-8     | 3-6     | 9      | 6      | 6-7    |
| Exon 2                 | 24-40   | 26-40   | 12-20  | 29     | 30-34  |
| Exon 3                 | 12-25   | 15-23   | 29-34  | 27     | 17-19  |
| Exon 4                 | 7-12    | 8-11    | 10-11  | 6      | 9-11   |
| Exon 5                 | 4-9     | 7-8     | 7      | 6      | 4-5    |
| Exon 6                 | 1-3     | 1       | 1      | 1      | 0-2    |
| Exons 7 and 8          | 2-3     | 2       | 3-5    | 3      | 0-2    |

* The range of nucleotide differences when Gogo-Oko is compared to individual alleles of the designated class I loci. A single Popy-A allele was compared.

-A*0201, -A*0401, and -A*0501 at both locus-specific and family-specific positions argues strongly that these cDNA sequences represent alleles of a gorilla Gogo-A locus that is the homologue of HLA-A.

In contrast, Gogo-Oko has a motif at the family-specific positions that is quite distinct from the Gogo-A sequences and that does not clearly segregate with any of the HLA-A families. In fact, its motif is closest to the consensus, suggesting this gene diverged before separation of any of the A families. This provides support for the possibility that Gogo-Oko is the product of a locus different from Gogo-A and

Figure 5. The relationship of ape A alleles to the different HLA-A families. Families of HLA-A alleles can be distinguished on the basis of overall sequence comparison. (a) In the upper panel are shown the frequency distributions of differences when all pairwise comparisons between HLA-A alleles are made. The middle panel shows the distribution for pairs of alleles derived from different families; the bottom panel shows the distribution for pairs of alleles from the same family. (b) A tree plotted from parsimony analysis (17) showing the segregation of HLA-A alleles into five families and two extended ancestral families. A bovine class I MHC sequence, B13-6, is the outlier for the analysis (36).
| Gorilla molecule | Similar molecule | Region of similarity | Gorilla molecule | Similar molecule | Region of similarity |
|------------------|------------------|----------------------|------------------|------------------|----------------------|
| A’0101 | A10s, A2s, A’2301, Patr-A’04 | Leader peptide | Gogo-A’0201 | Patr-A’04 | α2 (98–162) |
| Popy-A’01 | α1 (1–48) | | A’3301 | α2 (98–140) |
| Gogo-A’0201 | α1 (1–55) | | A’0201, A’2901 | α2-α3 | |
| A’2301, A’2401 | α1 (1–61) | | A’3201, A’2901 | α2-α3 | |
| Patr-A’04 | α1 (1–61) | | (153–188) | |
| A’2501, A’2601 | α1 (10–75) | | A’3201, A’2901 | α3 (203–274) | |
| A’3301 | α2 (10–72) | | Gogo-A’0401 | α3 (203–274) | |
| Gogo-A’0501 | α1-α2 | | & B’0103 | Patr-B’03 | Leader peptide |
| (84–170) | | B’0103 | α1 (1–64, 71–90) | |
| Gogo-A’0201 | α2 (98–162) | | B’2702 | α1-α2 | 71–94) | |
| A’3301, A’3101 | α2 (98–140) | | Bw4+ | αα (79–83) | |
| B’1401, B’1402 | α2 (153–181) | | Patr-B’01 | B’2601 | α2 (117–155) | |
| Gogo-A’0401 | α3 (2 distinctions) | | | A’2601 | α1-α2 | |
| Gogo-A’0201 | α3 (2 distinctions) | | Patr-B’02, -B’03, -B’04 | α1 (34–78) | |
| A2s, A10s, A19s | α3 (few distinctions) | | B’4001 | B’2601 | α1 (1–31) | |
| A’0201, A’3201 | Leader peptide | | many HLA-Bs | α1 (50–75) | |
| Patr-A’04 | α1 (1–55) | | A’2601 | α1-α2 | |
| A’2301, A’2401 | α1 (1–55) | | A’0101 | α2 (76–101) | |
| A’0201, A’0203 | α1 (10–73) | | B’0103 | α1-α2 | (76–96) | |
| Gogo-A’0101, A’0501 | α2 (98–162) | | Patr-B’03 | α2 (117–155) | |
| A’3301, A’3101 | α2-α3 | | B’5701 | α2-α3 | (157–259) | |
| A’0201 | (164–264) | | Patr-B’01, -B’03 | tm and cyt. | (275–339) | |
| A’0401 | A10s, A2s, A’2301 | Leader peptide | C’0101 & C’0102 | Most HLA-Cs | Leader peptide |
| Patr-A’04 | α1 (1–51) | | Gogo-C’0201, Cw6p | α1-α2 | |
| A’2601, A’2501 | α1 (1–55) | | B’5701 | α2-α3 | |
| B’5701, B’5801 | α1 (57–69) | | Cw’0701 | cyt. | |
| Gogo-A’0101 | α1-α2 | | | | |
| A’0201 | (71–140) | | | | |
| A’0501 | α3 (3 distinctions) | | | | |
| A10s, A2s, A’2301 | Leader peptide | | | | |
| Patr-A’04, A’2301 | α1 (1–51) | | | | |
| Patr-B’02, -04, -05 | α1 (64–72) | | | | |
| Bw4+ | α1 (79–83) | | | | |
| A’2501 | α1-α2 | | | | |
| (76–104) | | | | | |
| Gogo-A’0101 | α1-α2 | | | | |
| (84–170) | | | | | |
The gorilla molecules were assessed for regions of similarities with human, chimpanzee, and gorilla class I proteins. Numbers in parenthesis indicate the positions of the class I molecule where similarity is evident.

Table 2. (continued)

| Gorilla molecule | Similar molecule | Region of similarity |
|------------------|------------------|----------------------|
| C*0201, C*0202 & C*0203 | Most HLA-Cs | Leader peptide |
| Cw*0601, Cw'0101 | α1 (10-76) | |
| Cw'06p | α1-α2 | (25-96) |
| Cw*0701, Cw*0702, HLA4 | α1-α2 | (81-113) |
| Cw*0701 | α2-α3 | (163-266) |
| Cw*0701 | tm and cyt. | (295-342) |

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tm., transmembrane; cyt., cytoplasmic.

that diverged from Gogo-A before diversification of the families.

that none of the Gogo-A sequences segregates with a single family of HLA-A alleles is a quite different situation from that observed for chimpanzee Patr-A sequences. In that species all the Patr-A alleles clearly show a close relationship with the HLA-A3 family. This can be seen both from overall comparisons and from the pattern of substitution at the family-specific positions. Interestingly, the presence of an A9-specific position at position 807 in Patr-A4 may belie the shared ancestry of the A3 and A9 families. Perhaps of greatest importance is the conclusion emerging from these analyses that the A locus molecules in gorillas and chimpanzees derived from two distinct ancestral lineages of A alleles, both of which are represented in humans.

Proteins Motifs Shared by Gogo-A, Patr-A, and HLA A Alleles. As expected from the family relationships, none of the Gogo-A sequences is particularly similar to individual HLA-A alleles: the most similar pair being Gogo-A*0101 and HLA-A*6801, which differ at 27 nucleotide positions. Instead, they are formed of a pieture of structural motifs, many of which can be recognized in class I HLA and Patr sequences (Table 2).

In pairwise comparisons, the Gogo-A*0101, -A*0201, and -A*0401 proteins differs by 13–14 amino acid substitutions. Although distributed throughout the heavy chain, they are predominantly found in α1 and α2, and in particular at peptide binding positions of those domains (Fig. 7). The number of differences and their distributions is comparable to that seen, for example, between HLA-A*0201 and -A*6801.

The α1 domain of Gogo-A*0201 is almost the same as that of HLA-A*0201, and some similarities extend through the rest of these molecules. In particular, the motif at residues 62–65 of the α1 helix, which is similar in HLA-A*0201, HLA-B*5701, and -B*5801 and gives rise to a shared alloantigenic site, is present (22). This motif is also present in Gogo-A*0401, but interestingly in that case, the flanking amino acids are characteristic of those found in HLA-B*5701 and -B*5801.

The Gogo-A*0101 protein has an α1 domain and the first half of α2, which are clearly related to HLA-A*3301. Gogo-A*0501 differs from Gogo-A*0101, -A*0201, and -A*0401 by 27–29 amino acids. Substitutions in the α1 helix are the feature that make this allele distinct (17 of the 21 residues from positions 63–83 differ from those in Gogo-A*0101, -A*0201, and -A*0401). The sequence in this region can be divided into three segments: positions 62–72, which are characteristic of motifs found in HLA-B but not HLA-A alleles; positions 73–76, which are unique; and positions 77–83, which comprise a motif common to HLA-A, -B, and -H alleles. The latter motif is responsible for the Bw4 public epitope found in certain HLA-A and -B molecules (23). In these B-like regions, Gogo-A*0501 most closely resembles HLA-B*4901.

| Position | Gogo-B*0101 | Gogo-B*0102 | Gogo-B*0103 | Gogo-B*0201 |
|----------|-------------|-------------|-------------|-------------|
| 95       | I           | F           | W           | I           |
| 97       | R           | R           | Y           | R           |
| 99       | F           | F           | M           | Y           |
| 103      | V           | V           | M           | M           |

Differences between the three subtypes of Gogo-B*01. The residues at these positions for Gogo-B*0201, a molecule that is divergent elsewhere, are also included as its motif is related to that found for Gogo-B*0103. * Peptide-interacting residues.
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B*0102, which differ by a single nucleotide that substitutes phenylalanine for isoleucine at position 95.

The one substitution that is not at a peptide binding position, methionine at position 103 in Gogo-B*0103, is associated with substitutions at two neighboring peptide-binding residues (97 and 99), and its presence may be the result of a "hitchhiking" effect in which the product of a recombination event was selected because of the substitutions at positions 97 and 99. Alternatively, this substitution may have functional effects not yet appreciated.

Their close relationship strongly indicates that Gogo-B*0101, -B*0102, and -B*0103 derive from a common ancestor. The most related class I gene is HLA-B*4901, which differs at 40 or 41 nucleotide positions. Comparison with other class I alleles reveals elements of sequence that are shared with human and chimpanzee molecules (Table 2). For most of the leader peptide and residues 1-64 of the al domain, Gogo-B*0101, -B*0102, and -B*0103 are identical to the chim-
panzee B locus molecule, Patr-B*03. The COOH-terminal part of the al domain, residues 70-90, is identical to HLA-B*2702, a feature that may explain the high frequency with which HLA-B27 alloantibers react with gorillas. Residues 65-70 of the al helix have a unique motif not seen in other class I molecules.

Characteristically, it is difficult to identify evolutionary relationships in the al domain of HLA-B due to the extensive reassortment of substitutions in different molecules. The same is true for Gogo-B. In the COOH-terminal, two-thirds of the al domain, Gogo-B*0101, -B*0102, and -B*0103 are similar to HLA-B*1401, the differences being due to one substitution that is unique to Gogo-B*0101, -B*0102, and -B*0103 (arginine at 176) and another (histidine 171) shared by HLA-B*1401 and HLA-B5 molecules (Table 2).

Gogo-B*0201: a Divergent Allele. The Gogo-B*0201 allele differs from the Gogo-B*01 group by 58-60 nucleotide substitutions that give rise to 36-38 amino acid differences. These include 15-16 differences in peptide binding or TCR-interacting residues of the antigen recognition site. The Gogo-B*0201 heavy chain bears elements in common with Patr-B*03 and the HLA-B40 group of alleles in the NH2-terminal 76 residues of the al domain. The COOH-terminal part of the al has a motif not found in the HLA-C alleles. This region produces the alternative Bw4 or Bw6 epitopes, one of which is found on all HLA-C molecules. The Gogo-B*0201 molecule has neither motif in this region, instead having a sequence that is characteristic of HLA-A molecules.

HLA-B alleles are characterized by their homogeneity of sequence in exons 4-8, a feature that is not so strong for the Gogo-B alleles (18). Gogo-B*0201 differs by five amino acids from Gogo-B*01 in al and has a distinctive motif with an additional amino acid in the transmembrane region. This transmembrane region polymorphism is also seen in two other Patr-B alleles, Patr-B*01 and -B*03 (10, 11), and its origins may thus predate the divergence of the ape species (Fig. 1).

Preservation of Two Divergent Groups of C Alleles in Gorillas and Humans. The pattern of highly related alleles seen at the Gogo-B locus is reiterated with the five Gogo-C alleles. They divide into two groups: one consisting of Gogo-C*0101 and -C*0102; the other of Gogo-C*0201, -C*0202, and
-C*0203. In the first group, Gogo-C*0101 and -C*0102 are only distinguished by a single nucleotide substitution at position 933 in exon 5. This change does not affect the amino acid sequence and thus both alleles encode the same protein. It is improbable that this difference results from natural selection and is more likely the result of a neutral mutation that entered the population by genetic drift. Indeed, an analogous, single, silent substitution between two HLA-Cw2 alleles has been described (25). Among alleles of the second group, Gogo-C*0201 differs from Gogo-C*0203 by a single nucleotide in exon 1 that substitutes proline for leucine at position -13 in the leader peptide. This is unlikely to have functional effects upon the mature protein and is probably another example of a neutral mutation.

The Gogo-C*0202 allele differs from Gogo-C*0203 by a cluster of five nucleotide substitutions at positions 496 and 572 in exon 3. These lead to five amino acid substitutions at residues 142, 147, 152, 156, and 167 of the helix of the α2 domain. With the exception of 142, all are peptide binding residues. This difference between Gogo-C*0202 and Gogo-C*0203 is possibly the result of a single recombination event and it is therefore likely that these differences are the result of natural selection. A potential donor gene for either sequence motif in the region exchanged has not been identified. That two tryptophan residues in Gogo-C*0101 are replaced by two residues (leucine and serine) with much smaller side chains is likely to significantly modify the architecture of the peptide binding groove.

The two groups of Gogo-C sequences are exceptionally divergent. For example, the prototypical sequences, Gogo-C*0101 and Gogo-C*0201, exhibit 62 nucleotide substitutions and 35 amino acid differences. There is, however, a paucity of substitutions in the α1 helix, a feature shared by HLA-C alleles and that contrasts with the considerable variation of HLA-A and -B. The Gogo-C*0101 and C*0201 sequences differ by a single amino acid difference at position 77 of the α1 helix.

Differences between the two groups of Gogo-C alleles are found throughout the coding region with a bias towards the 3' exons encoding the α3, transmembrane, and cytoplasmic domains (Fig. 8). Such a pattern of differences is atypical of that seen between alleles of a class I HLA locus and more characteristic of that seen between class I loci. Human HLA-C alleles also divide into two groups: one comprising Cw*0701, Cw*0702, and HLA4, and the other Cw*0101-0601 and Cw*1201. Comparison of the Gogo-C and HLA-C alleles reveals that the Gogo-C*02 group is more closely related to HLA-Cw*0701, -Cw*0702, and HLA4 than they are to Gogo-C*01. Correspondingly, the Gogo-C*01 alleles are more closely related to HLA-Cw*0101-0601 and Cw*1201 than to the Gogo-C*02 alleles. These two families of HLA-C sequences, which can readily be distinguished on the basis of substitutions at 14 positions in exons 4-8, must therefore predate the divergence of humans and gorillas (Fig. 8 b). Their existence in chimpanzees is uncertain as just a single Patr-C allele has been characterized from that species. It does, however, group with the Gogo-C*01 group of sequences.

The diversity in the 3' exons of the two groups of Gogo-C and HLA-C sequences is unusual and suggests they may not be true alleles but the products of distinct but closely related genes. A precedent for such a phenomenon is provided by the class I genes of the H-2D region of the mouse (26). Some evidence to support this hypothesis comes from the studies of Duceman et al. (27), who distinguished the HLA-C locus from that encoding Y328 by restriction fragment length polymorphisms. Against this hypothesis is the absence of evidence for any HLA haplotypes expressing more than one HLA-C gene, and the possibility that there is a single locus with two highly distinctive families of alleles is still a reasonable possibility. In that case, they provide a further example of trans-species evolution of class I polymorphism (28).

**Diversification of the Antigen Recognition Site.** The heavy chain proteins predicted to be encoded by the 14 Gogo class I alleles have features common to HLA-A, -B, and -C alleles,

![Figure 8](image-url)
including the two disulphide bonds and the single glycosylation site at position 86. There is every indication from comparison with the HLA-A, -B, and -C protein sequences that all the Gogo class I heavy chains will assemble with ~/2-microglobulin (~/2-m) and peptide to form molecules that present antigens to T cells. Previously, we found gorilla ~/2-m to be identical in protein sequence to both human and chimpanzee ~/2-m (12).

Our previous comparison of chimpanzee and human class I genes showed that a majority of the substitutions found at positions of variation in the chimpanzee alleles were also found in human alleles, and that extended motifs of sequences were preserved in the two species (12, 18, 29). This pattern is consistent with a scenario in which the lifetime of alleles and polymorphic motifs can be longer than those of species and in which new species can inherit multiple alleles from ancestral species. A survey of the variation in the Gogo-A, -B, and -C alleles can be found in HLA-A, -B, and -C alleles (78%), Patr-A, -B, and -C alleles (51%) or either (82%). There are, however, substitutions that are unique to the gorilla lineage (Fig. 9), and as these frequently lead to amino acid replacements in the antigen recognition site, they are likely to be the result of natural selection for novel antigen-presenting properties. The number of these unique substitutions found in the gorilla class I heavy chains significantly exceeds that seen for the chimpanzee heavy chains (Fig. 9).

Although individual polymorphisms, and to some extent motifs, are shared by the class I alleles of humans, gorillas, and chimpanzees, there is no example of identical alleles, or of alleles encoding identical heavy chains, in the different species. Even the most related alleles (Patr-A and the HLA-A3 family) have amino acid substitutions in the antigen recognition site, and distinctive motifs in the residues in the antigen recognition site are characteristic of class I heavy chains from different species (Fig. 10). In addition, comparison of residues at the functional residues of the antigen recognition site clearly illustrates the relative conservation in sequence at TCR-interacting residues compared to peptide binding-residues (this is seen both within and between the species). These results suggest that no class I heavy chain present in the common ancestor of humans, chimpanzees, and gorillas has survived the last 5-10 million yr with its amino acid sequence and its antigen recognition sites unchanged.

**Discussion**

*Gorilla gorilla, Pan troglodytes,* and *Homo sapiens* are three closely related primate species that are estimated to have diverged from a common ancestor 7-10 million yr ago. Molecular analysis of homologous protein and nucleic acid sequences reveals so few differences between the three species that the issue as to which two of the species are most closely

| Gogo-A*0101 | ----- | ----- | E-F | ----- | ----- | I | S | I |
| Gogo-A*0201 | ----- | ----- | E-R | K | E-F | ----- | I | S | I |
| Gogo-A*0401 | ----- | ----- | E-R | ----- | ----- | L | ----- | I |
| Gogo-A*0501 | V | ME | EL | ----- | S | ----- | RQ | K | ----- | K |
| Gogo-Oko | TR | T | ----- | ----- | S | ----- | R- |----- |
| Gogo-B*0101 | ----- | ----- | ----- | ----- | ----- | R | ----- | E |
| Gogo-B*0102 | ----- | ----- | ----- | ----- | ----- | R | ----- | E |
| Gogo-B*0103 | ----- | ----- | ----- | ----- | ----- | R | ----- | E |
| Gogo-B*0201 | Q | W | T | ----- | ----- | E | ----- | R- |----- |
| Gogo-C*0101 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Gogo-C*0102 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Gogo-C*0201 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Gogo-C*0202 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Gogo-C*0203 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-A*01 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-A*02 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-A*03 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-A*04 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-B*01 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-B*02 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-B*03 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-B*04 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-B*05 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |
| Patr-C*01 | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- | ----- |

![Figure 9](image-url)  
Figure 9. Gorilla and chimpanzee amino acid polymorphisms that have not been identified in human class I molecules. Polymorphisms in bold lettering are found in only one molecule. Positions are numbered corresponding to Fig. 7 and those that are predicted to interact with peptide, TCR, or both are marked by downward triangle, upward triangle, or diamond, respectively.
The number of class I genes and their patterns of variation between class I alleles can be far greater than expected that final resolution of the trichotomy will come with comparison of extensive segments of homologous DNA. Factors that may have contributed to differences in the related has been highly controversial and remains unresolved. At present, the most common view is that chimpanzees and humans share a more recent common ancestor than either species does with the gorilla. It is to be expected that final resolution of the trichotomy will come with comparison of extensive segments of homologous DNA that are subject to minimal or no selection.

DNA encoding classical class I heavy chains is not of this type. These sequences are selected for antigen-presenting function and much of their variation and polymorphism can be attributed to that selection. Within species there is variation between class I alleles that can be far greater than seen between species for many other genes. Furthermore, it is clear that the number of class I genes and their patterns of expression change on the evolutionary timescale. Alleles at a particular class I locus may number into the hundreds and their distribution within a population can be quite even. Factors that may have contributed to differences in the class I alleles and loci of gorillas, chimpanzees, and humans are the pools of alleles and haplotypes inherited by new species from ancestral species, the nature and course of selection along different lineages, and neutral divergence.

In humans, HLA-A, -B, and -C are the class I loci implicated in antigen presentation and T cell immunity. Homologues of each of the three loci are expressed in the gorilla and chimpanzee, and the similarities between the homologous loci in the three species are such that no species-specific patterns of substitution can be discerned. For the chimpanzee class I sequences so far analyzed, all have conformed with the patterns established from analysis of class I HLA genes. This is not the case for the gorilla class I alleles, which in various ways are seen to break out of, or extend, the HLA patterns. For example, the number of unique substitutions found with the gorilla alleles significantly exceeds those found with the chimpanzee, and there is some unusual mixing of polymorphic motifs between the loci.
Most striking is the Gogo-Oko gene. Although more related to HLA-A than to either HLA-B or -C, it is not clear if this gene is an allele of Gogo-A, the gorilla homologue of the HLA-A locus. It has extended sequence identities with HLA-H, an HLA-A-related pseudogene, and does not segregate with any of the established families of A alleles. Watkins et al. (20) have isolated a similar clone to Gogo-Oko and suggest it was formed by a segmental exchange that introduced a segment of an HLA-H-like gene into an A allele. Our analysis does not favor this interpretation over plausible alternatives, in which Gogo-Oko is an allele of a locus distinct from Gogo-A. One possibility is for this locus to be the homologue of HLA-H, in which case the divergence of their sequences might be attributed to the profound difference that in the human lineage this gene (HLA-H) became nonfunctional, whereas in the gorilla lineage it may have retained function. A second possibility is that Gogo-Oko represents a third A-related gene that has not been described, or does not exist, in humans.

A second feature that distinguishes the gorilla class I loci is in differences between the A and B loci. By various parameters (the numbers of alleles, their relative frequency, and the diversity of the antigen recognition site), HLA-B appears a more functionally diversified locus than HLA-A (5). Furthermore, the greater diversity of HLA-B alleles in exons 2 and 3 encoding the antigen recognition site is associated with homogeneity in exons 4–8. In contrast, HLA-A alleles that are less diverse in exons 2 and 3 exhibit greater diversity than HLA-B in exons 4–8 (18). From our, albeit small, sampling of gorilla alleles, we see the opposite trends. Excluding Gogo-Oko, we obtained four very distinctive Gogo-A alleles from four genes isolated, whereas four Gogo-B genes yielded just two distinctive allelic motifs. However, in this small sampling of Gogo-B alleles, there is greater diversity in exon 4 than found in HLA-B. The Gogo-C locus has a lack of diversity similar to Gogo-B, and if the gene organization is like that in humans, then these two loci will be closely linked.

In comparisons of HLA-A, -B, and -C alleles, it has proved easier to track the lineages of HLA-A alleles than either the HLA-B or -C alleles. This is primarily due to the presence of polymorphisms in exons 3–8, particularly exon 4, which are absent in HLA-B. None of these polymorphisms are at functionally important positions of the protein (the antigen recognition site and CD8 binding site); a proportion are silent substitutions and they are thus likely to be selectively neutral.

The HLA-A alleles divide into five families themselves derived from two ancient lineages. One lineage led to the HLA-A2, A10, and A19 families, the other to the HLA-A3 and A9 families. The chimpanzee A alleles (Patr-A) conform more closely to the families defined in the human than do the Gogo-A alleles. Patr-A alleles clearly segregate with the HLA-A3 family, whereas the Gogo-A alleles are a mix of the A10 and A19 families and appear to have diverged from the human lineage before separation of these two families. This difference does not provide evidence that gorillas are more distantly diverged from humans than chimpanzees because the times at which the families diverged along the 2 lineages are likely to be different.

It is, however, intriguing that the Gogo-A alleles derive from one ancient lineage whereas the Patr-A alleles derive from the other. Thus, although both types of alleles were present in the common ancestor of the humans, gorillas, and chimpanzees, only one type has survived in each of the ape lineages, whereas both have survived in humans. Thus, in comparing A alleles one can see that Gogo-As and Patr-As correspond to two distinct parts of the distribution obtained with the HLA-A alleles (Fig. 11). In considering just this single, highly polymorphic, antigen-presenting locus, humans might be said to be more like both chimpanzees and gorillas than the ape species are to each other.
References

1. Klein, J., and F. Figueroa. 1986. Evolution of the major histocompatibility complex. CRC Crit. Rev. Immunol. 6:295.
2. Watkins, D.I., F.S. Hodi, and N.L. Letvin. 1988. A primate species with limited major histocompatibility complex class I polymorphism. Proc. Natl. Acad. Sci. USA. 85:7714.
3. Orr, H.T., and R. DeMars. 1988. Mapping of class I DNA sequences within the human major histocompatibility complex. Immunogenetics. 18:489.
4. Lawlor, D.A., J. Zemmour, P.D. Ennis, and P. Parham. 1990. Evidence of class-I MHC genes and proteins: from natural selection to thymic selection. Annu. Rev. Immunol. 8:23.
5. Parham, P., C.E. Lomen, D.A. Lawlor, J.P. Ways, N. Holmes, H.L. Coppin, R.D. Salter, A.M. Wan, and P.D. Ennis. 1988. Nature of polymorphism in HLA-A, -B, and -C molecules. Proc. Natl. Acad. Sci. USA. 85:4005.
6. Holmes, N., and P. Parham. 1985. Exon shuffling in vivo can generate novel HLA class I molecules. EMBO (Eur. Mol. Biol. Organ.) J. 4:2849.
7. Sibley, C.G., and J.E. Ahlquist. 1984. The phylogeny of the hominoid primates, as indicated by DNA-DNA hybridization. J. Mol. Evol. 20:22.
8. Hasegawa, M., H. Kishino, and T. Yano. 1987. Man's place in hominoidea as inferred from molecular clocks of DNA. J. Mol. Evol. 26:132.
9. Ruvolo, M., T.R. Disotell, M.W. Allard, W.M. Brown, and R.L. Honeycutt. 1991. Resolution of the African hominoid trichotomy by use of a mitochondrial gene sequence. Proc. Natl. Acad. Sci. USA. 88:1570.
10. Lawlor, D.A., F.E. Ward, P.D. Ennis, A.P. Jackson, and P. Parham. 1988. HLA-A and B polymorphisms predate the divergence of humans and chimpanzees. Nature (Lond.). 335:268.
11. Mayer, W.E., M. Jonker, D. Klein, P. Ivanyi, G. van Seventer, and J. Klein. 1988. Nucleotide sequences of chimpanzee MHC class I alleles: evidence for trans-species mode of evolution. EMBO (Eur. Mol. Biol. Organ.) J. 7:2765.
12. Lawlor, D.A., E. Warren, F.E. Ward, and P. Parham. 1990. Comparison of class I MHC alleles in humans and apes. Immunol. Rev. 113:147.
13. Ennis, P.D., J. Zemmour, R.D. Salter, and P. Parham. 1990. Rapid cloning of HLA-B cDNA by using the polymerase chain reaction: frequency and nature of errors produced in amplification. Proc. Natl. Acad. Sci. USA. 87:2833.
14. Staden, R. 1980. A new computer method for the storage and manipulation of DNA gel reading data. Nucleic Acids Res. 8:3673.
15. Devereux, J., P. Haebler, and O. Smithies. 1984. A comprehensive set of sequence analysis programs for the VAX. Nucleic Acids Res. 12:387.
16. Saitou, N., and M. Nei. 1986. The number of nucleotides required to determine the branching order of three species, with simony, version 3.0. A computer program distributed by the Illinois Natural History Survey, Champaign, IL.
17. Parham, P., D.A. Lawlor, C.E. Lomen, and P.D. Ennis. 1989. Diversity and diversification of HLA-A,B,C alleles. J. Immunol. 142:3937.
18. Zemmour, J., B.H. Koller, P.D. Ennis, D.E. Geraghty, D.A. Lawlor, H.T. Orr, and P. Parham. 1990. HLA-AR, an inactivated antigen-presenting locus related to HLA-A. Implications for the evolution of the MHC. J. Immunol. 144:3619.
19. Watkins, D.I., Z.W. Chen, T.L. Garber, A.L. Hughes, and N.L. Letvin. 1991. Segmental exchange between MHC class I genes in a higher primate: recombination in the gorilla between the ancestor of a human non-functional gene and an A locus gene. Immunogenetics. 34:185.
20. Kato, K., J.A. Trapani, J. Allopenna, B. DePonte, and S.Y. Yang. 1987. Molecular analysis of the serologically defined HLA-Aw19 antigens: a genetically distinct family of HLA-A antigens comprising A29, A31, A32, and A33, but probably not A30. J. Immunol. 143:3371.
21. Ways, J.P., J.B. Rothbard, and P. Parham. 1986. Amino acid residues 56 to 69 of HLA-A2 specify an antigenic determinant shared by HLA-A2 and HLA-B17. J. Immunol. 137:217.
22. Parham, P., D.A. Lawlor, C.E. Lomen, and P.D. Ennis. 1989. Multiple mechanisms producing diversity of HLA-C alleles. J. Immunol. 142:3937.
23. Lutz, C.T., D.A. Jensen, J. Schiffenbaner, D.K. Didier, B.D. Schwartz, and C.S. Davis. 1990. Multiple mechanisms produce diversity of HLA-C alleles. Hum. Immunol. 28:27.
24. Sease, L.R., R.M. Horton, J.K. Pullen, and Z. Cai. 1991. Structure and origin of diversity in the major histocompatibility complex. CRC Crit. Rev. Immunol. In press.
25. Duceman, B.W., D. Ness, R. Rendi, M.J. Chorney, R. Srivastava, D.S. Greenspan, J. Pan, S.M. Weissman, and F.C. Grunet. 1986. HLA-JY 328: mapping studies and expression of a polymorphic HLA class I gene. Immunogenetics. 23:90.
26. Parham, P., D.A. Lawlor, C.E. Lomen, and P.D. Ennis. 1989. Diversity and diversification of HLA-A,B,C alleles. J. Immunol. 142:3937.
27. Zemmour, J., B.H. Koller, P.D. Ennis, D.E. Geraghty, D.A. Lawlor, H.T. Orr, and P. Parham. 1990. HLA-AR, an inactivated antigen-presenting locus related to HLA-A. Implications for the evolution of the MHC. J. Immunol. 144:3619.
28. Watkins, D.I., Z.W. Chen, T.L. Garber, A.L. Hughes, and N.L. Letvin. 1991. Segmental exchange between MHC class I genes in a higher primate: recombination in the gorilla between the ancestor of a human non-functional gene and an A locus gene. Immunogenetics. 34:185.
29. Kato, K., J.A. Trapani, J. Allopenna, B. DePonte, and S.Y. Yang. 1987. Molecular analysis of the serologically defined HLA-Aw19 antigens: a genetically distinct family of HLA-A antigens comprising A29, A31, A32, and A33, but probably not A30. J. Immunol. 143:3371.
30. Ways, J.P., J.B. Rothbard, and P. Parham. 1986. Amino acid residues 56 to 69 of HLA-A2 specify an antigenic determinant shared by HLA-A2 and HLA-B17. J. Immunol. 137:217.
31. Parham, P., D.A. Lawlor, C.E. Lomen, and P.D. Ennis. 1989. Multiple mechanisms producing diversity of HLA-C alleles. J. Immunol. 142:3937.
32. Lutz, C.T., D.A. Jensen, J. Schiffenbaner, D.K. Didier, B.D. Schwartz, and C.S. Davis. 1990. Multiple mechanisms produce diversity of HLA-C alleles. Hum. Immunol. 28:27.
33. Sease, L.R., R.M. Horton, J.K. Pullen, and Z. Cai. 1991. Structure and origin of diversity in the major histocompatibility complex. CRC Crit. Rev. Immunol. In press.
34. Duceman, B.W., D. Ness, R. Rendi, M.J. Chorney, R. Srivastava, D.S. Greenspan, J. Pan, S.M. Weissman, and F.C. Grunet. 1986. HLA-JY 328: mapping studies and expression of a polymorphic HLA class I gene. Immunogenetics. 23:90.
35. Klein, J. 1987. Origin of major histocompatibility complex polymorphism: the trans-species hypothesis. Immunogenetics. 23:90.
36. Parham, P., D.A. Lawlor, C.E. Lomen, and P.D. Ennis. 1989. Diversity and diversification of HLA-A,B,C alleles. J. Immunol. 142:3937.
special reference to the human-chimpanzee-gorilla divergence. J. Mol. Evol. 24:189.
33. Hughes, A.L., and M. Nei. 1988. Pattern of nucleotide substitution at major histocompatibility complex class I loci reveals overdominant selection. Nature (Lond.). 335:167.
34. Hedrick, P.W., and G. Thomson. 1983. Evidence for balancing selection at HLA. Genetics. 104:449.
35. Townsend, A., and H. Bodmer. 1989. Antigen recognition by class I-restricted T lymphocytes. Annu. Rev. Immunol. 7:601.
36. Ennis, P.D., A.P. Jackson, and P. Parham. 1988. Molecular cloning of bovine class I MHC cDNA. J. Immunol. 141:642.
37. Bjorkman, P.J., and P. Parham. 1990. Structure, function, and diversity of class I major histocompatibility complex molecules. Annu. Rev. Biochem. 59:253.