IncRNA OXCT1-AS1 Promotes Metastasis in Non-Small-Cell Lung Cancer by Stabilizing LEF1, In Vitro and In Vivo

Binru Li, Libo Zhu, Linlin Li, and Rui Ma

Department of Thoracic Medicine, Cancer Hospital of China Medical University, Liaoning Cancer Hospital and Institute, Shenyang, Liaoning 110042, China

Correspondence should be addressed to Rui Ma; marui@cancerhosp-ln-cmu.com

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1. Introduction

Non-small-cell lung cancer (NSCLC) is the main type of lung cancer and one of the most frequently occurring cancers worldwide [1]. According to the Global Cancer Statistics 2018, lung cancer cases account for about 11.6% of the total number of cancer diagnoses; furthermore, among the total number of cancer-related deaths, 18.4% are attributable to lung cancer [2]. Although progress has been made in NSCLC treatment, metastatic NSCLC is still associated with a high rate of mortality [3, 4], indicating the critical need for the identification of new therapeutic targets. Metastasis is a complex multistep process, and the invasion and migration capability of cancer cells determines the metastasis of NSCLC [5, 6]. Furthermore, a gain of mesenchymal characteristics as well as loss of epithelial features frequently accompany the attainment of metastatic ability in NSCLC; this epithelial-mesenchymal transition involves numerous molecules such as long noncoding RNA (lncRNA), transcription factor, and circular RNA [7, 8]. However, the precise mechanisms underlying NSCLC metastasis have not yet been identified.

lncRNA is a class of RNA without protein-coding functions. Emerging researches have shown that many lncRNAs are dysregulated in NSCLC, and that their dysregulation contributes to metastasis in cancer [9, 10]. Recently, Liao et al. [11] found that the lncRNA CCHE1 increased the proliferation, metastasis, and invasion of NSCLC cells, and was predictive of poor survival in NSCLC patients. Chen et al. [12] identified a novel lncRNA OXCT1-AS1 through microarray experiments, and showed that its expression was elevated in lymph node metastasis. Furthermore, these authors found that lncRNA OXCT1-AS1 suppressed miR-455-5p to promote bladder cancer proliferation and invasion. However, the exact role of lncRNA OXCT1-AS1 in the metastasis of NSCLC, and the mechanisms involved, remain poorly understood.

Growing evidence indicates that lncRNAs participate in the regulation of gene expression at the transcription or post-transcription levels, as well as that of pathological and...
cellular physiological processes [13, 14]. In addition, lncRNAs regulate protein stability [15], as competing endogenous RNAs [16], and also regulate mRNAs’ stability [17]. Jiang et al. [18] recently reported that IncRNA SNHG15 interacts with and stabilizes Slug by blocking its ubiquitination to promote colon cancer progression. Furthermore, Xue et al. [15] found that IncRNA LINRIS blocked ubiquitination of IGFBP2 in colorectal cancer, maintaining its stability. Therefore, we speculated that IncRNA OXCT1-AS1 may play a potential role in migration, invasion, and metastasis of NSCLC by influencing the stability of target protein.

Lymphoid enhancer-binding factor 1 (LEF1), which is characterized by a high expression in tumors, is a key transcription factor of the WNT pathway and participates in the regulation of cell metastasis and proliferation [19]. Nguyen et al. [20] found that the WNT/TCF pathway through HOXB9 and LEF1 mediates metastasis in lung adenocarcinoma. Furthermore, results from Zhao et al.’s study [21] revealed that in esophageal squamous cell carcinoma, elevated level of LEF1 was obviously associated with lymph node metastasis, histologic grade, TNM stage, and poor prognosis. Therefore, we speculate that IncRNA OXCT1-AS1 may promote NSCLC metastasis by stabilizing LEF1.

To the best of our knowledge, the present study is the first to profile the expression of IncRNA OXCT1-AS1 in NSCLC cells. Additionally, we explored the role of IncRNA OXCT1-AS1 in regulating migration, invasion, and metastasis of NSCLC and investigated the interactions between IncRNA OXCT1-AS1 and LEF1, with the aim of identifying novel therapeutic targets for metastatic NSCLC.

2. Methods

2.1. Cell Culture. Human normal epithelial cell line BEAS-2B and human NSCLC cell lines (A549, H1299, H23, and HCC827) were obtained from the American Type Culture Collection (Manassas, VA, USA). NSCLC cell lines were cultured in RPMI 1640 medium (Thermo Fisher Scientific, Inc., Waltham, MA, USA) adding 10% fetal bovine serum (FBS; Thermo Fisher Scientific). BEAS-2B cells were cultured in BEGM medium (Thermo Fisher Scientific) adding 10% FBS. All cells were incubated in a humidified incubator at 37°C with 5% CO₂ and saturated humidity. When cell confluence had reached 70%-80%, the cells were detached with trypsin (0.25%) to obtain a single-cell suspension.

2.2. Lentiviral Vector System, Plasmids, and Cell Transfection. shRNAs targeting IncRNA OXCT1-AS1 (defined as sh-IncRNA) or LEF1 mRNA (defined as sh-LEF1) were ligated into the vector. The empty vector, which was used as the negative control, was named sh-NC. The lentiviruses were synthesized by Wuhan GeneCreate Biological Engineering Co., Ltd. (Wuhan, Hubei, China). Cells were transfected with these lentiviruses. To obtain stably transfected cell lines, H1299 and A549 cells were treated with 2–3 μg/mL puromycin (14 days). Knockdown efficiency was evaluated by qRT-PCR and western blot analyses. The expression vectors for Flag-tagged MS2 coat protein (MCP) and MS2-tagged IncRNA OXCT1-AS1, NARF, and LEF1 were provided by GeneCreate Biological Engineering, and Flag-tagged expression vectors for full-length IncRNA OXCT1-AS1, full-length LEF1 mRNA, and IncRNA OXCT1-AS1 fragments were provided by Sangon Biotech Co., Ltd. (Shanghai, China). The plasmids were transfected into A549 cells with Lipofectamine 3000 (Thermo Fisher Scientific), as recommended by the manufacturer. Transfection efficiency was assessed by qRT-PCR and western blot analyses. The shRNAs’ sequences are as follows: IncRNA OXCT1-AS1—forward, 5'-CAC CGC TTA CAT AGA GTA AGT TTG CCG AAG CAA ACT TAC TCT ATG TAA GC-3'; reverse, 5'-AAA AGC TTA CAT AGA GTA AGT TTG CTT CGT CGT AGA TTG AAG GC-3'; reverse, 5'-AAA AGC TTA CAT ATG TAA GC-3'; LEF1—forward, 5'-CAC CGC CTT AAA TCT ACG CAG AAG ACG AAT TTT CTG CTT CGT AGA TTG AAG GC-3'; reverse, 5'-AAA AGC TTA CAT ATG TAA GC-3'; NARF—forward, 5'-CAC CGC CTT AAG ATT CGT CTT CTG CTT CGT AGA TTG AAG GC-3'; reverse, 5'-AAA AGC TTA CAT ATG TAA GC-3'.

2.3. RNA Extraction and qRT-PCR Analysis. Total RNA was extracted from cells of both cell lines using a Total RNA extraction kit (Beyotime Institute of Biotechnology, Shanghai, China). The primer sequences for IncRNA OXCT1-AS1, GAPDH, and LEF1 were designed and synthesized by Sangon Biotech Co., Ltd. Total RNA was reverse transcribed into cDNA using a Reverse Transcriptase Kit (Beyotime Institute of Biotechnology), as recommended by the manufacturer. Then, qRT-PCR was performed using SYBR Premix Ex Taq II (Takara Biotechnology Co., Ltd., Dalian, Liaoning, China). The primer sequences were as follows: IncRNA OXCT1-AS1—forward, 5'-CTG AAC GTC AAG AGG TGG TGG CAA T-3'; reverse, 5'-GGT TGG TAC ACC TGG TGC TCA T-3'; GAPDH—forward, 5'-GGT TGG TAC ACC TGG TGC TCA T-3'; reverse, 5'-GGT TGG TAC ACC TGG TGC TCA T-3'. GAPDH was used as the endogenous control. Gene expression data were analyzed with the 2^-ΔΔCt method [22].

2.4. Isolation of Nuclear and Cytoplasmic Fractions. In accordance with the manufacturer’s protocol of the Nuclear and Cytoplasmic Extraction Reagent Kit (Beyotime Institute of Biotechnology), nuclear-cytoplasmic fractionation was conducted. Briefly, after washing in PBS, cells were suspended in cytoplasmic extraction reagent I (0.2 mL), followed by the addition of cytoplasmic extraction reagent II (11 μL). The suspension was then incubated on ice for 60 s, followed by centrifugation (16,000 g, 5 min). The cytoplasmic extract was the supernatant fraction, and the crude nuclei formed the pellet fraction.
2.5. Transwell Assays and Cell Proliferation Assays. For migration, 10,000 cells were placed into the upper chamber (8 µm pore size) (Corning Incorporated, Corning, New York, USA). For invasion, 1×10^5 cells were placed into the upper chamber with Matrigel (Corning Incorporated). RPMI 1640 medium (10% FBS) was supplemented to the lower chamber. After incubation (24 h), the cells were removed. Then, cells through the membrane were fixed with methanol, followed by crystal violet staining (0.1%). Images were acquired, and then cell numbers were counted under an inverted microscope (Olympus Corporation, Tokyo, Japan). For assessment of cell proliferation, 3000 cells were plated in 96-well plate. After incubation for different times (24, 48, and 72 h), cell viability was determined by the addition of CCK-8 (10 µL), using a CCK-8 kit (Dojindo, Mashikimachi, Kumamoto, Japan). The absorbance (450 nm) was measured using a microplate reader (Beijing Potenov Technology Co., Ltd., Beijing, China).

2.6. Western Blot Analysis. Using a RIPA kit (Beyotime), total protein was extracted. After protein lysis and centrifugation, the protein concentration was detected by a bicinchoninic acid kit (Beyotime Institute of Biotechnology). Subsequently, cell lysates were separated by SDS-PAGE and transferred onto PVDF membranes. After blocking with 5% skim milk (60 min), it was probed with primary antibodies overnight (4°C). After washing with PBST (4 times), secondary antibodies were added for 60 min (25°C). After washing again with PBST (4 times), enhanced chemiluminescence substrates (Beyotime Institute of Biotechnology) were applied to visualize the protein bands in a dark room. Quantification of protein bands was performed by ImageJ software (National Institutes of Health, Bethesda, MD, USA). The relative protein expression was normalized to β-actin. The following antibodies were used: Mouse Anti-E-cadherin antibody (ab231303, Abcam, Cambridge Biomedical Campus, Cambridge, UK; 1/1000), Mouse Anti-N-cadherin antibody (ab98952, Abcam, 1/1000), Rabbit Anti-vimentin antibody (ab137321, Abcam, 1/2000), Rabbit Anti-Snail antibody (ab216347, Abcam, 1/1000), Rabbit Anti-LEF1 antibody (ab137872, Abcam, 1/2000), Mouse Anti-SIRT1 antibody (ab110304, Abcam), Mouse Anti-Flag tag antibody (ab125243, Abcam), Rabbit Anti-NARFL antibody (ab241215, Abcam, 1/1500), Mouse Anti-HA tag antibody (ab18181, Abcam, 1/1500), and Rabbit Anti-β-actin antibody (ab179467, Abcam, 1/500), Goat Anti-Mouse IgG H&L (HRP) (ab205719, Abcam, 1/50,000), and Goat Anti-Rabbit IgG H&L (HRP) (ab205718, Abcam, 1/50,000).

2.7. Database Analysis. The targets of lncRNA OXCT1-AS1 were acquired through the RNA Interactome Database (http://www.rna-society.org/rnainter/home.html/).
and PRIdictor websites (http://bclab.inha.ac.kr/pridictor/pridictor.html). Through the Protein Lysine Modifications Database (PLMD, http://plmd.biocuckoo.org/), lysine modification sites were acquired.

2.8. RNA Pull-Down Assays. Biotinylated lncRNA OXCT1-AS1, antisense lncRNA OXCT1-AS1, and lncRNA OXCT1-AS1 fragments were transcribed in A549 and 293T cells. The RNA products treated with RNase-free DNase I (Thermo Fisher Scientific) were purified using a Total RNA Purification Kit (Sangon Biotech Co., Ltd.). Then, 4 μg of biotin-labelled RNAs were denatured (65°C, 300 s) in PA buffer and slowly cooled. Next, the folded RNA was supplemented with 2 U/mL RNasin (Solarbio) and streptavidin Dynabeads (Thermo Fisher Scientific) and then incubated for 60 min at 4°C. After washing, beads were boiled in 40 μL 1x SDS loading buffer for 600 s. Then, lncRNA OXCT1-AS1 interacting proteins were subjected to western blot analysis.
2.9. RNA Immunoprecipitation Assay. A549 cells were lysed in 500 μL of RNA immunoprecipitation buffer (Beyotime Institute of Biotechnology) containing protease inhibitors and an RNAase inhibitor. After centrifugation (8,000 g, 600 s). The supernatants were incubated with anti-Flag, anti-LEF1, anti-mouse IgG, or anti-rabbit IgG antibody for 120 min at 4°C (gentle rotation). Then, protein A/G beads (40 mL) were added and incubated for 60 min (4°C). After washing in RIPA buffer (3 times) and PBS (once), RNA was extracted and qRT-PCR was performed to detect the level of gene expression.

2.10. Coimmunoprecipitation (CO-IP) Assay. A549 cells were lysed in 500 μL of RNA immunoprecipitation buffer (Beyotime Institute of Biotechnology) containing protease inhibitors. After preclearing with 30 μL protein G/A-plus agarose beads (Thermo Fisher Scientific) for 60 min (4°C), centrifugation was performed (3,000 g, 5 min, 4°C). Supernatants were incubated with antibody (2 μg) for 240 min (4°C). Then, the immunoprecipitates were incubated with protein G/A-plus agarose beads (30 μL) overnight. After centrifugation (3,000 g, 5 min, 4°C), the precipitates were washed (5 × 10 min) with bead wash solution, followed by resuspension in loading buffer (60 μL) and incubation for 300–600 s (100°C). Next, western blot was performed.

2.11. In Vivo Tumorigenesis in Nude Mice. Animal studies were implemented following the principles and procedures of the National Institutes of Health Guide for the Care and Use of Animals. Four-to-five-week-old BALB/c-nude mice (50% male and 50% female) were obtained from the Laboratory Animal Center of Jilin University (SCXK (Ji) 2016-0001) (Changchun, Jilin, China). The research procedures were reviewed and approved by the Medical Ethics Committee of China Medical University. Mice were allowed free access to food as well as water and maintained at 20 ± 2°C. For in vivo assay of cancer cell metastasis (ten mice per group), negative control or lncRNA OXCT1-AS1- (or LEF1-) knockdown A549 cell suspensions (1 × 10⁶ cells, 100 μL) were injected into the tail veins of nude mice. BALB/c-nude mice were euthanized as soon as the following symptoms were detected: (a) inability to obtain water or food; (b) general lack of moving activities; (c) severe cachexia (weight loss approaching 25%); (d) pale appearance, body coat looking scruffy and unhealthy; (e) infection at the injection site; and (f) breathing problem. After 3 weeks, BALB/c-nude mice were anesthetized with 1% isoflurane and sacrificed by decapitation. Their lungs were harvested for hematoxylin and eosin (H&E) staining. The lung nodules were quantified microscopically by three observers. The body weights of nude mice were measured regularly.

2.12. Statistical Analysis. Each experiment was repeated in triplicate independent experiments. The data are shown as the mean ± SEM. GraphPad software 8 (GraphPad Software, Inc., La Jolla, CA, USA) and SPSS (IBM Corp., Armonk, NY, USA) were used for statistical analyses. Student’s t-test was performed to evaluate the difference between two groups. The significance of the difference between multiple groups was analyzed by one-way analysis of variance followed by
Figure 4: Continued.
Figure 4: lncRNA OXCT1-AS1 interacts with LEF1 in NSCLC cells. Subcellular fractionation assay and qRT-PCR were carried out to evaluate the distribution of lncRNA OXCT1-AS1 in the nucleus and cytoplasm of the H1299 (a) and A549 cells (b). The downstream targets were predicted using the RNAInter website. The binding sites of lncRNA OXCT1-AS1 in LEF1 (d) and the binding sites of LEF1 in lncRNA OXCT1-AS1 (e) were predicted using PRIdictor. (f, g) LEF1 protein expression in lncRNA OXCT1-AS1 stably depleted and negative control H1299 and A549 cells was measured by western blot. (h) Schematic presentation of the MS2-RNA pull-down strategy used to assess the interaction between LEF1 and lncRNA OXCT1-AS1. (i) Whole-cell lysates of A549 cells transfected with antisense-lncRNA-MS2 or IncRNA-MS2 were incubated with amylase beads and MBP-MS2 purified protein, followed by western blot analysis. GAPDH served as a negative control. (j) RNA pull-down assay using 293T cells cotransfected with Flag-LEF1 and IncRNA OXCT1-AS1-MS2 or antisense IncRNA OXCT1-AS1-MS2, followed by western blot analysis. (k) RNA immunoprecipitation assays were performed in A549 cell extracts using a LEF1 antibody, followed by qRT-PCR. GAPDH mRNA and FGF4 mRNA were used as the negative and positive control, respectively. (l) Deletion mapping of the LEF1-binding domain in IncRNA OCT1-AS1. Diagrams of full-length IncRNA OXCT1-AS1 and the deletion fragments. (m) Immunoblot analysis of LEF1 in A549 cells pulled down using different IncRNA OXCT1-AS1 fragments. GAPDH was used as the negative control. The bottom image shows each IncRNA OXCT1-AS1 fragment.

3. Results

3.1. lncRNA OXCT1-AS1 Is Upregulated in NSCLC Cell Lines. To further confirm the effects of lncRNA OXCT1-AS1 on metastasis in vivo, lncRNA OXCT1-AS1-downregulated and negative control A549 cells were injected into the tail vein of nude mice. As shown in Figures 3(a)–3(c), lncRNA OXCT1-AS1 depletion dramatically suppressed the development of pulmonary metastasis. HE staining indicated that the number of metastatic nodules in the lung was significantly lower in the sh-NC group compared with that in the sh-lncRNA group (Figures 3(b) and 3(c)). Interestingly, the body weights of mice were measured and were shown to be stable (Figure 3(d)). These results demonstrate that IncRNA OXCT1-AS1 might act as a key factor to promote metastasis in NSCLC.

3.4. lncRNA OXCT1-AS1 Interacts with LEF1 and Upregulates Its Expression. To identify the mechanism underly丙 lncRNA OXCT1-AS1-induced metastasis in NSCLC, the distribution of lncRNA OXCT1-AS1 was first investigated in H1299 and A549 cells. The results shown in Figures 4(a) and 4(b) indicated that lncRNA OXCT1-AS1 was enriched in the cytoplasm. Then, downstream targets of lncRNA OXCT1-AS1 were predicted using bioinformatics databases. As shown in Figure 4(c), among the 429 downstream targets, transcription factor LEF1, which was identified using the RNAInter (http://www.rna-society.org/main/ en/home.html/) and PRIdictor websites (http://bclab.http://inha.ac.kr/pridictor/pridictor.html/), was capable of binding lncRNA OXCT1-AS1. The binding sites

Tukey’s post hoc test. Values of $P < 0.05$ were considered to indicate statistical significance. *$P < 0.05$ and **$P < 0.01$ compared with BEAS-2B or sh-NC cells. ***$P < 0.05$ and ****$P < 0.01$ compared with MG132 treatment.
| Position | Peptides | Type         |
|----------|----------|--------------|
| 283      | DSDLMHVQHQRKR | Acetylation  |
| 283      | DSDLMHVQHQRKR | Ubiquitination |
| 301      | PKRPHKSLNATFL | Ubiquitination |
| 311      | NAIFIBMOVRANMV | Ubiquitination |
| 382      | KKKRRRREQHESAG | Acetylation  |
| 382      | KKKRRRREQHESAG | Ubiquitination |
| 54       | ECDLADERSYNES | Ubiquitination |
| 86       | SQEYHDDRHEPDID | Acetylation  |
| 86       | SQEYHDDRHEPDID | Ubiquitination |
| 95       | REHPDEGRPDGGLY | Acetylation  |
| 95       | REHPDEGRPDGGLY | Ubiquitination |

(a) Relative expression of LEF1

(b) sh-NC sh-lncRNA

(c) MG132 - MG132 +

(d) HA-UB Flag-LEF1 IncRNA MG132

(e) CHX treated (h)

(f) IP Flag Input

(g) Ubiquitin LEF1 β-Actin

(h) Relative expression of protein

(i) Relative expression of protein

(j) Relative expression of protein

Figure 5: Continued.
on LEF1 and lncRNA OXCT1-AS1 are shown in Figures 4(d) and 4(e). Next, we explored the relationship between lncRNA OXCT1-AS1 and LEF1, and found that lncRNA OXCT1-AS1 knockdown attenuated LEF1 protein expression in H1299 and A549 cells (Figures 4(f) and 4(g)). To confirm the interaction between lncRNA OXCT1-AS1 and LEF1 protein, RNA pull-down assays were performed in A549 cells. The results manifested that lncRNA OXCT1-AS1 pulled down LEF1 but not GAPDH (Figures 4(h) and 4(i)). Additionally, RNA pull-down assays in 293T cells transfected with Flag-LEF1, lncRNA OXCT1-AS1, or its antisense lncRNA revealed an ability of lncRNA OXCT1-AS1 to pull down Flag-LEF1 (Figure 4(j)). To further validate this interaction between lncRNA OXCT1-AS1 and LEF1, RNA immunoprecipitation assays were performed. As Figure 4(k) displays, LEF1 was associated with lncRNA OXCT1-AS1 but not control GAPDH mRNA. Deletion-mapping analysis indicated that the lncRNA OXCT1-AS1 fragments 1201 nt and 1731 nt were involved in the interaction with LEF1 (Figures 4(l) and 4(m)). We conclusively demonstrate that lncRNA OXCT1-AS1 interacts with LEF1 in NSCLC cells.

3.5. lncRNA OXCT1-AS1 Stabilizes LEF1 by Blocking NARF-Mediated Ubiquitination. To elucidate the potential mechanisms underlying the role of lncRNA OXCT1-AS1 in regulating LEF1 stability, we investigated the ubiquitination sites of LEF1 using the Protein Lysine Modifications Database (http://plmd.biocuckoo.org/index.php). The results of Figure 5(a) show that LEF1 had a ubiquitination site at the 54 position, which was close to the binding site of lncRNA OXCT1-AS1 on LEF1 at the 55 position. A549 cells were treated with MG132 to suppress LEF1 degradation. As shown in Figures 5(b) and 5(c), LEF1 levels were significantly reduced following lncRNA OXCT1-AS1 knockdown in the presence of MG132. This suggests that an enhancement of LEF1 stability by lncRNA OXCT1-AS1 is related to proteasomal degradation. Subsequently, we explored the role of lncRNA OXCT1-AS1 on LEF1 expression in the presence of CHX, as shown in Figures 5(d) and 5(e). We found that lncRNA OXCT1-AS1 knockdown shortened the half-life of LEF1. Additionally, lncRNA OXCT1-AS1 overexpression in 293T cells decreased the levels of LEF1 ubiquitination (Figure 5(f)), which was also confirmed in A549 cells (Figure 5(g)). To further investigate the mechanism by which lncRNA OXCT1-AS1 regulates LEF1 ubiquitination, E3 ubiquitin-ligase NARF was knocked down and overexpressed in A549 cells. As shown in Figures 5(h)–5(j), NARF knockdown resulted in a significant increase in LEF1 levels.

**Figure 5:** lncRNA OXCT1-AS1 stabilizes LEF1 by inhibiting NARF-mediated LEF1 ubiquitination. (a) The ubiquitination sites of LEF1 were predicted using Protein Lysine Modifications Database. (b, c) A549 cells were transfected with shRNA against lncRNA OCT1-AS1, followed by MG132 treatment. Then, LEF1 expression was detected by western blot analysis. (d, e) A549 cells were transfected with shRNA against lncRNA OCT1-AS1, followed by CHX treatment (100 μg/mL) for different times (0, 1, and 2 h). Then, LEF1 expression was detected by western blot analysis. (f) After transfection with HA-UB, Flag-LEF1, or lncRNA OCT1-AS1 in combination or separately, and one day posttranscription, 293T cells were treated with MG132 (6 h), followed by CO-IP using the anti-LEF1 antibody. Then, western blot analysis was performed. (g) lncRNA OCT1-AS1 stably depleted A549 cells were treated with MG132 for 6 h, followed by CO-IP using the anti-LEF1 antibody; then, western blot was performed. (h–j) Lysates from A549 cells with NARF knockdown or NARF overexpression were used to detect the levels of NARF and LEF1 by western blot analysis. (k, l) 293T cells were transfected with Flag-NARF, HA-LEF1, and sh-lncRNA, followed by treatment with MG132 (6 h); next, cell lysates were immunoprecipitated using the antibody against Flag or HA. The precipitates and inputs were analyzed by western blot analysis. (m, n) NARF expression in H1299 and A549 cells with lncRNA OXCT1-AS1 knockdown was detected by western blot.
Figure 6: Continued.
(P < 0.01); however, NARF overexpression resulted in a remarkable decrease (P < 0.01). Additionally, lncRNA OXCT1-AS1 overexpression weakened the interaction between LEF1 and NARF in A549 cells, including NARF coimmunoprecipitating less LEF1 and LEF1 coimmunoprecipitating less NARF (Figures 5(k) and 5(l)). Interestingly, downregulation of lncRNA OXCT1-AS1 did not affect the levels of NARF (P > 0.05), as shown in Figures 5(m) and 5(n). Therefore, these results confirm that lncRNA OXCT1-AS1 abrogates NARF-mediated LEF1 ubiquitination by blocking the interaction between LEF1 and NARF, preventing LEF1 proteasomal degradation.

### 3.6. LEF1 Is Involved in Migration and Invasion of NSCLC Cells

To evaluate whether LEF1 is functionally involved in NSCLC cell metastasis, A549 and H1299 cells with LEF1 knockdown were established. The transfection efficiency of sh-LEF1 was determined by western blot assay (Figures 6(a) and 6(b)). CCK-8 assays showed that the cell viability of H1299 cells with sh-LEF1 transfection was significantly decreased compared with that in the sh-NC group cells (P < 0.01, Figure 6(c)). Similar results were obtained in A549 cells (P < 0.01, Figure 6(d)). In functional experiments, LEF1 knockdown demonstrated a stronger ability to decrease the migration of H1299 and A549 cells compared with the control cells (Figures 6(e) and 6(f)). Importantly, sh-LEF1 markedly inhibited the invasion of H1299 and A549 cells (Figures 6(g) and 6(h)). Further, we investigated the correlation between LEF1 and proteins associated with epithelial-mesenchymal transition. Western blot assay revealed that LEF1 knockdown significantly downregulated the expression of N-cadherin, vimentin, and Snail, while significantly upregulating E-cadherin expression, in H1299 and A549 cells (Figures 6(i)–6(l)).

### 3.7. LEF1 Is Involved in Metastasis of NSCLC

To investigate the possibility of acquired tumor metastasis in vivo, LEF1-downregulated and negative control A549 cells were injected...
LEF1 knockdown significantly inhibited lung metastasis (Figures 7(a)–7(c)). H&E staining indicated that the number of metastatic nodules in the lung was obviously lower in the sh-LEF1 group than in the sh-NC group (Figures 7(b) and 7(c)). Interestingly, the body weights of mice were shown to be stable (Figure 7(d)). These results collectively suggest that LEF1 plays a role in NSCLC cell metastasis.

Figure 7: LEF1 knockdown inhibits NSCLC metastasis. (a) The images of metastatic nodules in the lungs of nude mice after tail vein injection with LEF1-downregulated and negative control A549 cells. (b) H&E staining in nude mice after tail vein injection with LEF1-downregulated and negative control A549 cells. (c) The average number of metastatic nodules in the lungs in the tail vein injection model. (d) The body weights of nude mice were measured.

Figure 8: A schematic illustration of the role and molecular mechanism underlying lncRNA OXCT1-AS1 in promotion of NSCLC metastasis. lncRNA OXCT1-AS1 stabilized LEF1 by blocking NARF-mediated ubiquitination and subsequently promoted migration and invasion of NSCLC.
4. Discussion

The 5-year survival rate of NSCLC is lower than 15%, primarily because of the occurrence of distant metastatic disease [23, 24]. Protein modification is closely associated with metastasis in NSCLC [25]. Among the various complex regulatory networks of cancer metastasis, IncRNAs also play a key role in regulating tumor fate [26]. Despite the rapidly increasing number of IncRNAs with growing knowledge about their underlying mechanisms, studies of IncRNAs in NSCLC metastasis remain inadequate. Here, we characterize, for the first time, the functions of IncRNA OXCT1-AS1 in NSCLC metastasis and the mechanisms involved. Our findings identify potential novel therapeutic targets for cancer metastasis.

In this study, we demonstrated that knockdown of IncRNA OXCT1-AS1 inhibited the invasion and migration in NSCLC cells. Additionally, the function and pathological significance of IncRNA OTUD6B-AS1 in NSCLC metastasis were also confirmed in vivo. Subsequently, we investigated the potential molecular mechanism by which IncRNA OXCT1-AS1 maintains LEF1 stability, and found that this involves the blockade of NARF-mediated LEF1 ubiquitination and proteasomal degradation in vitro. Moreover, we found that LEF1 stimulated cell migration and invasion in vitro and in vivo (Figure 8).

An increasing number of studies have demonstrated that IncRNA is involved in tumorigenesis and development of NSCLC. For example, results from Jiang et al.’s study [27] revealed that the IncRNA HOTAIR contributes to tumorigenesis and metastasis in NSCLC by upregulating miR-613 expression. Therefore, the identification of IncRNA signatures may be of clinical value in the diagnosis, treatment, and prognostic prediction of NSCLC. Recently, a study by Chen et al. [12] highlighted the important roles of IncRNA OXCT1-AS1 in the complex molecular processes that contribute to bladder cancer cell aggressiveness. IncRNA OXCT1-AS1 imprinted 1.7 kb is highly expressed in bladder cancer, metastatic lymph node tissues, and multiple bladder cancer cell lines [12]. A key finding of our study was that the expression of IncRNA OXCT1-AS1 was significantly enhanced in NSCLC cells compared with that in normal lung epithelial cells, especially in the H1299 and A549 cell lines, suggesting the clinical significance of this IncRNA in NSCLC. This observation is additionally consistent with Chen et al.’s report [12].

Extensive research has demonstrated the crucial roles of IncRNAs in NSCLC metastasis [28, 29]. IncRNA PCAT6, a well-studied IncRNA, contributes to tumorigenesis and metastasis in NSCLC by binding to EZH2 [30]. Furthermore, results from He et al.’s study [31] showed that IncRNA AFAP1-AS1, an oncogene, promotes cell migration by increasing AFAP1 expression in NSCLC. In this study, loss-of-function experiments revealed that depletion of IncRNA OXCT1-AS1 strongly inhibited NSCLC cell migration and invasion in vitro. Furthermore, in vitro function assays showed that IncRNA OXCT1-AS1 knockdown significantly reduced the number of metastatic nodules in the lung. These data confirmed that IncRNA OXCT1-AS1 induces NSCLC metastasis. Silencing IncRNA OXCT1-AS1 may represent a promising strategy to inhibit NSCLC metastasis.

IncRNAs have been shown to exert their functions through a variety of mechanisms, by directly interacting with proteins, mRNAs, and/or microRNAs [12, 32]. In the present study, we performed cell nuclear and cytoplasmic RNA isolation and found that IncRNA OXCT1-AS1 was enriched in the cytoplasm, which suggests that IncRNA OXCT1-AS1 plays a role in metastasis in a posttranscriptional-dependent manner. However, the mechanisms by which IncRNA OXCT1-AS1 induces metastasis remain largely unknown, with numerous studies proposing that protein regulation is involved [33, 34]. Using bioinformatics databases, 429 downstream targets were predicted. Considering that transcription factor LEF1 is reported to be involved in lung cancer metastasis [20], and there is a steric hindrance effect that the binding of IncRNA OXCT1-AS1 to the 55 position of LEF1 affects the ubiquitination of LEF1 at the 54 position, we selected LEF1 as a target. Interestingly, we found that IncRNA OXCT1-AS1 knockdown resulted in the attenuation of LEF1 protein expression. LEF1, a downstream factor of the Wnt/β-catenin pathway, regulates gene transcription independently [35]. Extensive research has shown that LEF1 plays an essential role in the epithelial-mesenchymal transition by activating the transcription of N-cadherin, vimentin, and Snail [34, 35]. Bleckmann et al. [36] reported that LEF1 overexpression correlates with poor prognosis in cerebral metastasis of lung adenocarcinomas. Therefore, we hypothesized that IncRNA OXCT1-AS1 maintains LEF1 stability to contribute to NSCLC metastasis. To verify this hypothesis, validation experiments were conducted, and showed that IncRNA OXCT1-AS1 directly interferes with LEF1 through its fragments 800 nt and 1201 nt, supporting our hypothesis. These findings also suggest that LEF1 stability is maintained by IncRNA OXCT1-AS1 at the posttranslational level.

Accumulating evidence shows that the stability of LEF1 can be controlled by its ubiquitination [37] and sumoylation [38]. Interestingly, recent research indicates that IncRNAs are capable of regulating protein ubiquitination [34, 39]. For example, Tang et al. [40] found that the IncRNA GLCC1 stabilizes c-Myc by blocking ubiquitination via direct interaction with the HSP90 chaperone protein. In this study, we found that IncRNA OXCT1-AS1 blocked LEF1 ubiquitination, thereby preventing LEF1 proteasomal degradation. Previous studies have indicated that nemo-like kinase-associated ring protein NARF regulates the ubiquitylation and degradation of LEF1 [41]. Therefore, we explored the role of NARF in IncRNA OXCT1-AS1-mediated LEF1 stability, and found that IncRNA OXCT1-AS1 knockdown facilitates the interaction between NARF and LEF1. These findings show that IncRNA OXCT1-AS1 blocks NARF-mediated LEF1 ubiquitination by inhibiting the interaction between NARF and LEF1, thereby preventing LEF1 proteasomal degradation. Our findings are largely consistent with previous studies, in which constitutively activated STAT5a was shown to recruit the E3 ubiquitin-ligase NARF to LEF1 and lead to LEF1 ubiquitination and degradation [42]. These findings highlight the important role of IncRNA OXCT1-AS1 in NSCLC metastasis. Although IncRNA OXCT1-AS1...
metastasis. Although these diagnosis, prognostic evaluation, and therapy of NSCLC a foundation for the application of lncRNA OXCT1-AS1 in metastasis of NSCLC. These functional roles of LEF1 were further confirmed by loss-of-function assays in vivo. In line with previous findings in esophageal squamous cell carcinoma [21], these results confirm that lncRNA OXCT1-AS1 promotes NSCLC metastasis by stabilizing LEF1.

In conclusion, the present study reveals, for the first time, that upregulation of the lncRNA OXCT1-AS1 induces NSCLC metastasis in vitro and in vivo. Mechanistically, our findings demonstrate that lncRNA OXCT1-AS1 blocks NARF-mediated LEF1 ubiquitination by inhibiting the interaction between LEF1 and NARF, thereby preventing LEF1 proteasomal degradation. Furthermore, LEF1 was confirmed to be involved in NSCLC metastasis. These findings establish a foundation for the application of lncRNA OXCT1-AS1 in diagnosis, prognostic evaluation, and therapy of NSCLC metastasis. Although these findings may lack clinical evidence, our study still have implications for treatment strategies of NSCLC metastasis.

Data Availability
All data generated or analyzed during this study are included in this published article. The datasets generated and/or analyzed during the current study are available from the corresponding author on reasonable request.

Conflicts of Interest
The authors declare that they have no competing interests.

Authors’ Contributions
Binru Li conceived and together with Rui Ma designed the study. Binru Li and Libo Zhu were involved in data collection. Libo Zhu and Linlin Li performed the statistical analysis and preparation of figures. Binru Li drafted the paper. Rui Ma contributed substantially to its revision. Binru Li and Libo Zhu contributed equally to this work. All authors have reviewed and approved of the article prior to submission and have read and approved the final article.

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