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Characterization of a Feline Influenza A(H7N2) Virus

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During December 2016–February 2017, influenza A viruses of the H7N2 subtype infected ≈500 cats in animal shelters in New York, NY, USA, indicating virus transmission among cats. A veterinarian who treated the animals also became infected with feline influenza A(H7N2) virus and experienced respiratory symptoms. To understand the pathogenicity and transmissibility of these feline H7N2 viruses in mammals, we characterized them in vitro and in vivo. Feline H7N2 subtype viruses replicated in the respiratory organs of mice, ferrets, and cats without causing severe lesions. Direct contact transmission of feline H7N2 subtype viruses was detected in ferrets and cats; in cats, exposed animals were also infected via respiratory droplet transmission. These results suggest that the feline H7N2 subtype viruses could spread among cats and also infect humans. Outbreaks of the feline H7N2 viruses could, therefore, pose a risk to public health.

Influenza A viruses are endemic in humans and enzootic in other mammalian species including swine and horses; occasional infections of other mammalian species including whales, seals, sea lions, felidae in zoos, and other species have been reported (7). Reports of influenza A virus infections in dogs and cats were rare until 2004, when equine influenza viruses of the H3N8 subtype caused outbreaks in greyhounds in Florida (2). Since then, influenza viruses of the H3N8 and H3N2 subtypes have caused several outbreaks in dogs in the United States and South Korea (3–5). Until recently, only 1 major influenza A virus outbreak had been reported in cats (6). This changed in December 2016 with the outbreak of low pathogenic avian influenza A viruses of the H7N2 subtype in animal shelters in New York. Approximately 500 cats were infected in December 2016–February 2017; most of which experienced a mild illness with coughing, sneezing, and runny nose from which they recovered fully. Severe pneumonia developed in 1 elderly animal with underlying health issues, which was euthanized. A veterinarian who had treated an infected animal also became infected with the feline influenza A(H7N2) virus and experienced a mild, transient illness, suggesting the potential for these viruses to infect humans. While this manuscript was being prepared, Belser et al. reported that the H7N2 subtype virus isolated from the human case caused a mild disease in mice and ferrets, but was not transmitted among ferrets (7). We assessed feline H7N2 subtype viruses isolated from infected cats during the outbreak for their replicative ability, pathogenicity, and transmissibility in mammals; in contrast to the findings recently published by Belser et al. (7), we detected productive infection of co-housed ferrets, although with low efficiency. We also conducted extensive pathology and transmission studies in cats, and detected feline virus transmission via respiratory droplets to exposed cats. Our study provides additional data on the risk that the feline H7N2 subtype viruses pose to public health.

Methods

Cells and Viruses

The origins and growth conditions of all cell lines used in this study are described in the online Technical Appendix (https://wwwnc.cdc.gov/EID/article/24/1/17-1240-Techapp1.pdf). The feline H7N2 subtype viruses used in this study were isolated from swabs collected from cats with influenza-like symptoms during the outbreak in an animal shelter in New York in December 2016. We obtained A/chicken/New York/22409–4/1999 (H7N2, A/chicken/...
NY/99) virus from the Agricultural Research Service, US Department of Agriculture (8). We deposited the viral gene sequences obtained in this study to GenBank. We amplified the feline virus in Madin-Darby canine kidney (MDCK) cells and the A/chicken/NS/99 virus in 10-day-old embryonated chicken eggs.

**Growth Kinetics of Viruses in Cell Culture**

We infected cells with viruses at a 0.005 multiplicity of infection, incubated them for 1 hour at 37°C, washed twice, and cultured with 1× minimal essential medium containing 0.3% bovine serum albumin and trypsin treated with L-1-tosylamide-2-phenylethyl chloromethyl ketone at 33°C and 37°C (37°C and 39°C for chicken embryo fibroblast cells) for various periods. We determined virus titers at the indicated time points by use of plaque assays in MDCK cells. The statistical analyses are described in the online Technical Appendix.

**Infection of Animals**

To determine the pathogenicity of the viruses in infected animals, we anesthetized three 6-week-old female BALB/c mice (Jackson Laboratory, Bar Harbor, ME, USA) for each virus with isoflurane and inoculated intranasally with 10-fold serially diluted virus in a 50-µL volume. The mice were monitored daily for 14 days and checked for changes in body weight and morbidity and mortality. We euthanized animals if they lost more than 25% of their initial bodyweight.

To determine the pathogenicity of the viruses in infected ferrets and cats, we inoculated 6-month-old female ferrets (Triple F Farms, Sayre, PA, USA; 3 per group; serologically negative by hemagglutination inhibition assay for currently circulating human influenza viruses), and unvaccinated 4- to 5-month-old female specific-pathogen-free cats (Liberty Research, Waverly, NY, USA; 3 per group; serologically negative by hemagglutination inhibition assay for circulating human influenza viruses) with 10^6 PFU (mice) in 0.5 ml of phosphate-buffered saline. We monitored the animals daily for changes in bodyweight, body temperature, and clinical signs for 14 days.

For virus replication in organs and pathology analyses, we worked with groups of mice (12 per group), ferrets (6 per group), and cats (6 per group). We inoculated the animals intranasally with 10^5 PFU (mice) in 0.05 ml of phosphate-buffered saline or 10^6 PFU (ferrets and cats) of viruses in 0.5 ml of phosphate-buffered saline. On days 3 and 6 postinfection, we euthanized 6 mice, 3 ferrets, and 3 cats in each group for pathological analysis and virus titration in organs (by use of plaque assays in MDCK cells).

**Virus Transmission Studies in Ferrets and Cats**

For direct contact transmission experiments, we housed 3 ferrets per group in regular ferret cages and 3 cats per group in large dog transporter cages (online Technical Appendix Figure 1), and infected them intranasally with 10^6 PFU (500 µL) of viruses. One day later, we housed 1 virus-naive animal with each infected animal. We collected nasal washes from the infected ferrets and nasal swabs from the infected cats on day 1 after infection, and from the exposed animals on day 1 after exposure and then every other day (for up to 11 days). We determined virus titers in the nasal washes and swabs by performing plaque assays in MDCK cells. We monitored all animals daily for disease symptoms and changes in bodyweight and temperature for 14 days.

We performed airborne transmission experiments by using ferret isolators (Showa Science, Tokyo, Japan) (9–11) or regular cat cages. In these settings, there was no directional airflow from the infected to the exposed animals. We inoculated 3 animals per group intranasally with 10^6 PFU (500 µL) of viruses. One day after infection, we placed 3 immunologically naïve animals in a cage adjacent to an infected animal. This setting prevented direct and indirect contact between animals but allowed spread of influenza virus by respiratory droplet. We spaced the ferret cages 5 cm apart and the cat cages 35 cm apart. We monitored the animals and assessed virus titers as described above.

**Results**

**Genetic and Phylogenetic Analysis of Feline Influenza(H7N2) Viruses Isolated in Animal Shelters in New York, December 2016**

We obtained swabs (collected on the same day) from 5 cats that experienced influenza-like symptoms during the outbreak at an animal shelter in New York, NY, in December 2016. After inoculation of these samples into MDCK cells, we isolated 5 pleomorphic influenza A viruses of the H7N2 subtype (Table 1; online Technical Appendix Figure 2). The HA consensus sequences of the 5 isolates (established by Sanger sequence analysis) displayed >99.9% similarity at the nucleotide level (Table 1). Phylogenetic analyses demonstrated that the 8 viral RNA segments of the 5 feline H7N2 viruses are most closely related to poultry influenza A(H7N2) viruses detected in the New York area in the late 1990s through early 2000s (Figure 1; online Technical Appendix Figures 3–9), suggesting that the 2016 feline H7N2 virus isolates descended from viruses that circulated more than a decade ago in the northeastern United States.

The HA protein of the 2016 feline H7N2 subtype virus encodes a single arginine residue at the hemagglutinin cleavage site (PEKPKPR↓G; the arrow indicates the cleavage site that creates the HA1 and HA2 subunits), indicative of low pathogenicity in chickens. Antigenically, A/feline/New York/WVDL-14/2016 (A/feline/NS/16) differs from other, closely related H7 viruses (online Technical Appendix Table 1); for example, its HA deviates by 27 aa from the closely related A/chicken/NS/22409–4/1999 HA. The
neuraminidase (NA) and ion channel (M2) proteins of the H7N2 viruses do not encode amino acids that confer resistance to neuraminidase or ion channel inhibitors. Inspection of the remaining feline H7N2 viral proteins revealed an absence of the most prominent amino acid changes known to facilitate adaptation to mammals, such as PB2–627K (16). These data thus suggest the 2016 feline H7N2 subtype viruses are avian-derived influenza viruses of low pathogenicity in avian and mammalian species.

Replication of Feline and Avian H7N2 Subtype Viruses in Cultured Cells

To characterize the replicative ability of the 2016 feline H7N2 viruses in cultured cells, we compared A/feline/NY/16 (which encodes the consensus amino acid sequence of the 5 isolates) with a closely related 1999 avian influenza virus, A/chicken/NY/22409–4/1999 (H7N2, A/chicken/NY/99) (Figure 1; online Technical Appendix Figures 3–9), which was isolated from a chicken in a live-bird market in New York state in 1999 (8). There are a total of 97 aa differences between A/feline/NY/16 and A/chicken/NY/99 viruses (12 aa differences in polymerase basic 2 (PB2), 7 in polymerase basic 1 (PB1), 12 in polymerase acidic (PA), 27 in hemagglutinin (HA), 8 in nucleoprotein (NP), 11 in neuraminidase (NA), 7 in matrix protein 1 (M1), 4 in matrix protein 2 (M2), and 9 in nonstructural protein 1 (NS1). Canine, human, feline, and chicken cells were infected at a multiplicity of infection of 0.005 at temperatures mimicking those of the upper and lower respiratory tracts of the respective species (i.e., 37°C and 39°C for chicken cells; 33°C and 37°C for the remaining cells) (Figure 2). In canine MDCK, feline Clone81, and human Calu-3 cells, A/feline/NY/16 replicated at least as efficiently as A/chicken/NY/99 virus, while both viruses replicated to low titers in human A549 cells. Of note, A/feline/NY/16 virus replicated less efficiently than A/chicken/NY/99 virus in feline lung Fc2Lu cells. In chicken embryo fibroblast cells, A/feline/NY/16 virus replicated more slowly than A/chicken/NY/99 virus at early time points and reached its highest titers at later time points. When we compared virus growth at the 2 temperatures tested (i.e., 37°C and 39°C for chicken cells; 33°C and 37°C for the remaining cells), we observed similar trends (for example, in MDCK cells, A/feline/NY/16 replicated more efficiently than A/chicken/NY/99 at both temperatures tested).

Replication and Pathogenicity of Feline and Avian H7N2 Subtype Viruses in Mice

To assess the replication of A/feline/NY/16 and A/chicken/NY/99 viruses in mice, 3 mice per group were inoculated intranasally with 10-fold dilutions of viruses, and their bodyweight and morbidity and mortality were monitored daily for 14 days. Mice infected with A/feline/NY/16 virus did not experience weight loss or signs of disease, whereas infection with 10⁶ PFU of A/chicken/NY/99 virus caused severe weight loss and required euthanasia (online Technical Appendix Figure 10).

A/feline/NY/16 replicated efficiently in the nasal turbinates and less efficiently in the lungs of infected animals (online Technical Appendix Figure 11); no virus was isolated from the other organs tested (i.e., brains, kidneys, livers, and spleens; data not shown). A/chicken/NY/99 replicated more efficiently in the lungs than in the nasal turbinates, consistent with immunohistochemistry analyses that detected A/feline/NY/16 virus antigens mainly in the upper respiratory organs of infected mice, whereas A/chicken/NY/99 virus antigens were detected more frequently in the lower respiratory organs (online Technical Appendix Figure 12).

Replication and Pathogenicity of Feline and Avian H7N2 Subtype Viruses in Ferrets

Ferrets intranasally infected with 10⁶ PFU of A/feline/NY/16 or A/chicken/NY/99 virus did not lose bodyweight (online Technical Appendix Figure 13) but 2 of the ferrets infected with A/chicken/NY/99 virus had high fevers on day 1 postinfection. Both viruses replicated efficiently in the nasal turbinates and were also isolated from the trachea and lungs of some animals (Table 2), consistent with similar antigen distributions for both viruses (online Technical Appendix Figure 14). No viruses were isolated from any of the other organs tested.

Replication and Pathogenicity of Feline and Avian H7N2 Subtype Viruses in Cats

The infection of ≈500 cats with H7N2 subtype viruses in animal shelters in New York in December 2016 suggested

| Virus | PB2 | PB1-F2 | PA | NA | NS2 |
|-------|-----|-------|----|----|-----|
| A/feline/New York/WVDL-3/2016 | S    | C     | Q  | Y  | C   |
| A/feline/New York/WVDL-9/2016 | N    | Y     | R  | H  | C   |
| A/feline/New York/WVDL-14/2016 | S    | C     | Q  | Y  | C   |
| A/feline/New York/WVDL-16/2016 | S    | C     | Q  | Y  | F   |
| A/feline/New York/WVDL-20/2016 | S    | C     | Q  | Y  | C   |

*Consensus sequences among the 5 H7N2 subtype viruses are shown in bold. Amino acids: C, cysteine; D, aspartic acid; E, glutamic acid; F, phenylalanine; H, histidine; L, leucine; N, asparagine; Q, glutamine; R, arginine; S, serine; Y, tyrosine. Viral proteins: NA, neuraminidase; NS, nonstructural protein; PA, polymerase acidic; PB, polymerase basic.
Figure 1. Phylogenetic tree of influenza A viral hemagglutinin segments from New York, NY, USA, compared with reference viruses. Phylogenetic analysis was performed for selected influenza A viruses representing major lineages. The evolutionary history was inferred using the neighbor-joining method (12). The optimal tree with the branch length sum of 1.22521320 is shown. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (500 replicates) is shown next to the branches (13). The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were computed using the Tamura 3-parameter method (14) and are in the units of the number of base substitutions per site. The analysis involved 44 nt sequences. Codon positions included were 1st + 2nd + 3rd + noncoding. All positions containing gaps and missing data were eliminated. The final dataset contained a total of 1,612 positions. Evolutionary analyses were conducted in MEGA7 (15).
Figure 2. Growth properties of A/feline/NY/16 and A/chicken/NY/99 influenza A(H7N2) viruses in mammalian and avian cells at different temperatures, New York, NY, USA. Cells were infected with viruses at a multiplicity of infection of 0.005 and incubated at 33°C and 37°C (or at 37°C and 39°C for avian CEF cells). Supernatants were harvested at the indicated time points. Virus titers were determined by use of plaque assays in Madin-Darby canine kidney (MDCK) cells. The species from which the cell lines are derived are shown. The values presented are the averages of 3 independent experiments ±SD. Statistical significance was determined as described in the online Technical Appendix (https://wwwnc.cdc.gov/EID/article/24/1/17-1240-Techapp1.pdf). *p<0.05; **p<0.01; ***p<0.001; ****p<0.0001. A549, human lung carcinoma epithelial cells; Clone81, cat kidney fibroblast cells; Fc2Lu, cat lung cells; CEF, chicken embryo fibroblast cells.
efficient replication of these viruses in felines. However, it was unclear whether these viruses were restricted to the respiratory organs or caused systemic infection. Cats intranasally infected with 10⁶ PFU of A/feline/NY/16 or A/chicken/NY/99 did not lose bodyweight (Figure 3); however, fever was detected in 1 animal infected with A/feline/NY/16, and 1 infected with A/chicken/NY/99 virus; and a different animal infected with A/feline/NY/16 sneezed intensely on day 3 postinfection, but recovered fully.

A/feline/NY/16 virus replicated efficiently in the nasal turbinates, trachea, and lungs of infected cats (with the exception of 1 cat with a virus-negative lung sample on day 3 postinfection; Table 2). We isolated A/chicken/NY/99 virus mostly from nasal turbinates, with limited replication in the trachea and lung. These findings are consistent with the detection of A/feline/NY/16 antigen in both the upper and lower respiratory organs of infected cats, whereas A/chicken/NY/99 antigen was detected mainly in the nasal turbinates (Figure 4). A/feline/NY/16 and A/chicken/NY/99 viruses were also isolated from the jejunum or colon of some of the infected animals (Table 2), although viral antigen was not detected in the intestines of cats infected with A/chicken/NY/99 or A/feline/NY/16 virus. These results demonstrate that the feline H7N2 virus replicates efficiently in the upper and lower respiratory tract of cats, reflecting adaptation of the virus to its new host.

All cats infected with the A/feline/NY/16 virus exhibited histologic lesions in their nasal turbinates, tracheas, and lungs. Nasal turbinate pathology was moderate to severe in 5 of 6 cats with multifocal to diffuse distribution of lesions (Figure 5, panel A). The tracheas of these cats exhibited mild to moderate histopathology (Figure 5, panel B), whereas the lungs exhibited multifocal to coalescing histopathology centered mostly on the bronchioles, with 3 of 6 cats possessing moderately severe lesions (Figure 5, panel C). Similar histopathological changes were found in cats infected with A/chicken/NY/99 virus. Appreciable histopathology was also noted in the small intestine (duodenum) of cats infected with A/feline/NY/16 and A/chicken/NY/99 viruses (Figure 5, panel D; cat ID nos. 1, 2, 4, 8, 10, and 12 in Table 2), although virus was detected in the intestines of only 3 cats (cat ID nos. 4, 6, and 9 in Table 2). The correlation between virus replication and histologic lesions in cat intestines is currently unknown.

Transmission of Feline and Avian H7N2 Subtype Viruses in Ferrets and Cats

The fulminant spread of the feline H7N2 subtype viruses among cats, and the confirmed H7N2 virus infection of a veterinarian who treated the animals, indicate that these originally avian influenza viruses have the ability to transmit among mammals. To test the transmissibility of feline and

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### Table 2. Virus titers in organs of ferrets and cats infected with A/feline/NY/16 or A/chicken/NY/99 influenza A(H7N2) viruses, New York, NY, USA

| Species and virus   | Days postinfection | Animal ID no. | Nasal turbinates | Trachea | Lung | Small intestine | Colon | Other organs†      |
|---------------------|--------------------|---------------|------------------|---------|------|-----------------|-------|-------------------|
| Ferret              |                    |               |                  |         |      |                 |       |                   |
| A/feline/NY/16      | 3                  | 1             | 4.4              | 3.3     | 4.7  | –               | –     | –                 |
|                     |                    | 2             | 5.2              | –       | 2.4  | –               | –     | –                 |
|                     |                    | 3             | 5.4              | –       | –    | –               | –     | –                 |
|                     | 6                  | 4             | 3.1              | –       | –    | –               | –     | –                 |
|                     |                    | 5             | 5.8              | –       | –    | –               | –     | –                 |
|                     | 6                  | 6             | 6.0              | –       | –    | –               | –     | –                 |
| A/chicken/NY/99     | 3                  | 7             | 5.9              | 3.3     | –    | –               | –     | –                 |
|                     |                    | 8             | 6.0              | –       | –    | –               | –     | –                 |
|                     | 6                  | 9             | 6.6              | 3.4     | –    | –               | –     | –                 |
|                     | 10                 | 10            | 4.2              | –       | –    | –               | –     | –                 |
|                     | 11                 | 11            | 4.5              | 5.7     | –    | –               | –     | –                 |
|                     | 12                 | 12            | 4.4              | –       | –    | –               | –     | –                 |
| Cat                |                    |               |                  |         |      |                 |       |                   |
| A/feline/NY/16      | 3                  | 1             | 3.9              | 4.8     | –    | –               | –     | –                 |
|                     |                    | 2             | 4.1              | 6.6     | 5.8  | –               | –     | –                 |
|                     |                    | 3             | 6.9              | 7.0     | 5.8  | –               | –     | –                 |
|                     | 6                  | 4             | 6.3              | 6.2     | 6.1  | –               | 2.3   | –                 |
|                     |                    | 5             | 7.7              | 7.8     | 4.7  | –               | –     | –                 |
|                     | 6                  | 6             | 5.9              | 6.2     | 6.7  | 3.8             | –     | –                 |
| A/chicken/NY/99     | 3                  | 7             | 6.4              | 5.8     | 3.9  | –               | –     | –                 |
|                     |                    | 8             | 2.0              | –       | –    | –               | –     | –                 |
|                     | 9                  | 9             | 6.1              | –       | –    | 4.9             | –     | –                 |
|                     | 6                  | 10            | 6.4              | –       | 5.0  | –               | –     | –                 |
|                     |                    | 11            | 4.6              | –       | –    | –               | –     | –                 |
|                     | 12                 | 12            | 6.7              | 4.0     | –    | –               | –     | –                 |

†, no virus detected.
††, Brain, spleen, kidneys, liver, and pancreas.
avian H7N2 subtype viruses in ferrets, 3 animals per group (each placed in a separate cage) were infected intranasally with $10^6$ PFU (500 μL) of A/feline/NY/16 or A/chicken/
NY/99 virus. One day later, we housed 1 naive ferret with each of the infected ferrets (direct contact transmission experiment), or placed naive ferrets in wireframe cages (within

Figure 3. Body weight and temperature changes in cats infected with A/feline/NY/16 and A/chicken/NY/99 influenza A(H7N2) viruses, New York, NY, USA. Three cats per group were infected intranasally with $10^6$ PFU of viruses and monitored for bodyweight and temperature changes.

Figure 4. Immunohistochemistry findings in cats infected with influenza A(H7N2) virus, New York, NY, USA. Shown are representative sections of nasal turbinates and lungs of cats infected with the indicated viruses on days 3 and 6 postinfection. Three cats per group were infected intranasally with $10^6$ PFU of virus, and tissues were collected on days 3 and 6 postinfection. Type A influenza virus nucleoprotein (NP) was detected by a mouse monoclonal antibody to this protein. For nasal turbinate sections: –, no NP-positive cells; +/-, NP-positive cells detected in 1–2 focal regions; +, NP-positive cells detected in >2 focal regions; 2+, NP-positive cells in large regions. For bronchus and alveolar sections: –, no NP-positive cells; +/-, ≥5 NP-positive cells; +, >6 NP-positive cells. NP-positive cells were detected in focal, but not in diffuse bronchial and alveolar sections. For all analyses, the entire sections were evaluated. Scale bars indicate 50 μm (nasal turbinates) or 100 μm (lung).
In lungs, moderately severe histopathologic changes are present in the lower airways. The lamina propria of bronchi (B) and bronchioles (Br) and the surrounding interstitium are infiltrated by numerous histiocytes, lymphocytes, and plasma cells (*), which also extend into and expand neighboring alveolar septa. The infiltrates extend into and expand nearby alveolar septa. The lumina of bronchioles are filled with numerous foamy macrophages, viable and degenerating neutrophils, proteinaceous fluid, and sloughed respiratory epithelial cells.

Hyperplasia of bronchiolar-associated lymphoid tissue (open arrow) and perivascular edema (solid arrow) are present. Scale bar indicates 500 µm. B) In nasal cavities, copious amounts of exudate are present comprising numerous degenerating and necrotic neutrophils, cellular debris, proteinaceous fluid, and strands of mucin. The respiratory epithelium covering the nasal turbinates (T) is extensively eroded. The underlying lamina propria appears diffusely bluish-purple due to infiltration by moderate-to-large numbers of histiocytes, neutrophils, lymphocytes, and plasma cells (*). Scale bar indicates 500 µm. C) In the trachea, a locally extensive focus of inflammation is present in the underlying lamina propria. Moderate numbers of histiocytes, neutrophils, and plasma cells are also infiltrating the respiratory epithelium. Scale bar indicates 500 µm. D) In the tracheal wall, moderate numbers of histiocytes, lymphocytes, and plasma cells (*), which also extend into and expand nearby alveolar septa. The infiltrates extend into and expand nearby alveolar septa. The lumina of bronchioles are filled with numerous foamy macrophages, viable and degenerating neutrophils, proteinaceous fluid, and sloughed respiratory epithelial cells. Scale bar indicates 500 µm.

**Figure 5.** Pathology findings in cats infected with A/feline/NY/16 influenza A(H7N2) virus on day 6 postinfection, New York, NY, USA. A) The submucosa of the duodenum are present between and around Brunner’s glands (BG). Scale bar indicates 100 µm.

| Virus and transmission mode | Pair | Action | 1     | 3     | 5     | 7     | 9     | 11    | Serocconversion, HI titer† |
|----------------------------|------|--------|-------|-------|-------|-------|-------|-------|--------------------------|
| A/feline/NY/16             | 1    | Infection | 4.2   | 5.6   | 5.0   | –     | –     | –     | 320                       |
|                           | 2    | Infection | 4.6   | 4.3   | 5.4   | –     | –     | –     | 320                       |
|                           | 3    | Infection | 5.3   | 5.0   | 4.8   | 2.8   | –     | –     | 640                       |
| Direct contact             | 1    | Infection | 3.6   | 4.1   | 5.0   | 2.0   | –     | –     | 640                       |
|                           | 2    | Infection | 5.5   | 5.1   | 4.3   | 1.3   | –     | –     | 320                       |
|                           | 3    | Infection | 5.0   | 5.2   | 5.2   | 2.9   | –     | –     | 640                       |
| A/chicken/NY/99            | 1    | Infection | 5.8   | 4.0   | 4.3   | –     | –     | –     | 160                       |
|                           | 2    | Infection | 5.6   | 4.2   | 3.5   | –     | –     | –     | <10                       |
|                           | 3    | Infection | 5.1   | 3.7   | 3.5   | –     | –     | –     | 320                       |
| Respiratory droplets       | 1    | Exposure  | –     | –     | –     | –     | –     | –     | 160                       |
|                           | 2    | Exposure  | –     | –     | –     | –     | –     | –     | <10                       |
|                           | 3    | Exposure  | –     | –     | –     | –     | –     | –     | 160                       |
| Direct contact             | 1    | Exposure  | –     | –     | –     | –     | –     | –     | 160                       |
|                           | 2    | Exposure  | –     | –     | –     | 3.8   | 4.3   | 3.4   | 160                       |
|                           | 3    | Exposure  | –     | –     | –     | 2.1   | –     | –     | 10                        |
|                           | 1    | Infection | 4.3   | 4.3   | 3.0   | –     | –     | –     | 160                       |
|                           | 2    | Infection | 4.2   | 3.8   | 3.8   | –     | –     | –     | 160                       |
|                           | 3    | Infection | 4.9   | 3.9   | 4.3   | –     | –     | –     | 320                       |

*HI, hemagglutination inhibition; –, no virus detected.
†Serum specimens were collected on day 18 after infection, exposure, or contact, and examined using an HI assay. The HI titer is the inverse of the highest dilution of serum that completely inhibited hemagglutination.
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We conducted the transmission study in cats in the same way as the study in ferrets; we spaced cages 35 cm apart to prevent direct contact between the inoculated and exposed animals (online Technical Appendix Figure 1, panel A). All infected cats secreted viruses for 5–7 days after infection and seroconverted, except for 1 cat infected with A/chicken/NY/99 virus, which seroconverted but did not shed virus (Table 4). We did not isolate A/chicken/NY/99 virus from contact or exposed animals, although these animals seroconverted (Table 4). Direct contact transmission of A/feline/NY/16 virus was detected in all 3 pairs of cats, with both seroconversion and virus isolation in 2 pairs (Table 4). Respiratory droplet transmission of A/feline/NY/16 occurred in 2 pairs of animals, with high virus titers detected in the nasal secretions of the exposed animals on days 9 and 11 postexposure, respectively; both of the exposed animals also seroconverted (Table 4). In the third transmission pair, the exposed animal did not shed virus or seroconvert. Taken together, we demonstrated that A/feline/NY/16 virus has the ability to transmit among cats via contact and respiratory droplets; the relative contribution of these modes of transmission to the H7N2 subtype virus outbreaks in cat shelters in New York is unknown.

Receptor-Binding Specificity of Feline and Avian H7N2 Subtype Viruses

Avian influenza viruses isolated from their natural reservoir (i.e., wild aquatic birds) are often restricted in their ability to infect mammalian cells because of their preferential binding to α2,3-linked sialic acids, whereas most human influenza viruses preferentially bind to α2,6-linked sialic acids (17–19). We performed glycan array analysis with A/feline/NY/16, A/chicken/NY/99, and Kawasaki/173-PR8, a control virus possessing the HA and NA genes of the seasonal human A/Kawasaki/173/2001 (H1N1) virus and the remaining genes from A/PR/8/34 (H1N1) virus. As expected, Kawasaki/173-PR8 virus bound to α2,6-linked sialosides (Figure 6; online Technical Appendix Table 2). A/chicken/NY/99 virus bound to both α2,6- and α2,3-linked sialosides, consistent with the dual avian/human receptor-binding specificity of influenza viruses isolated from land-based poultry (1). Of note, A/feline/NY/16 virus bound strongly to α2,3-linked sialosides (i.e., avian-type receptors) with negligible binding to human-type receptors.

Next, we examined the prevalence of α2,3- and α2,6-linked sialosides in the feline airway and intestines of an immunologically naive cat by using lectins that detect α2,3-linked (i.e., MAA I and MAA II) and α2,6-linked sialosides (i.e., SNAI). MAA I and MAA II bound to epithelial cells throughout the feline airway, whereas SNAI binding was detected only in the trachea and bronchus (Figure 7), consistent with the findings of other research groups (20–22). We did not detect sialosides in the cat intestine. The predominance of avian-type receptors in

| Table 4. Influenza A(H7N2) virus titers in nasal swab samples from cat transmission studies, New York, NY, USA* |
|---|---|---|---|---|---|---|---|---|---|---|---|
| Virus and transmission mode | Pair | Action | Virus titers in nasal swab samples by days after infection, log10 PFU/mL | Seroconversion, HI titer† |
|---|---|---|---|---|---|---|---|---|---|---|---|
| A/feline/NY/16 | Respiratory droplets | 1 | Infection | 5.6 | 4.7 | 4.3 | 3.0 | – | – | – | 320 |
| | | 2 | Exposure | 4.5 | 2.7 | 5.0 | 4.6 | 5.2 | – | – | 160 |
| | | 3 | Infection | 4.8 | 3.2 | 5.3 | 3.6 | – | – | – | 320 |
| | Direct contact | 1 | Infection | 5.9 | 4.6 | 3.4 | – | – | – | 320 |
| | | 2 | Contact | 6.0 | 5.0 | 4.6 | – | – | – | 640 |
| | | 3 | Infection | 4.9 | 4.9 | 4.4 | 4.2 | – | – | – | 640 |
| A/chicken/NY/99 | Respiratory droplets | 1 | Infection | 4.0 | 3.7 | 4.7 | – | – | – | 320 |
| | | 2 | Infection | 2.6 | 1.6 | 2.3 | – | – | – | 80 |
| | Direct contact | 1 | Infection | 4.5 | 2.4 | 4.5 | – | – | – | 80 |
| | | 2 | Infection | 3.4 | 4.8 | 4.1 | 3.6 | – | – | – | 160 |
| | | 3 | Infection | 3.4 | 3.5 | 3.2 | 3.3 | – | – | – | 160 |

*HI, hemagglutination inhibition; –, no virus detected.
†Serum samples were collected on day 18 after infection, exposure, or contact, and examined by use of an HI assay. The HI titer is the inverse of the highest dilution of serum that completely inhibited hemagglutination.

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the upper respiratory tract of felines may have led to the selection of feline H7N2 virus HA proteins with preferential binding to α2,3-linked sialosides.

**Sensitivity to Neuraminidase Inhibitors**

To test whether infections with the feline H7N2 viruses could be treated with neuraminidase (NA) inhibitors, we assessed the sensitivity of A/feline/NY/16 and A/chicken/NY/99 to several NA inhibitors (i.e., oseltamivir, zanamivir, and laninamivir) by determining the 50% inhibitory concentration (IC$_{50}$) of the NA enzymatic activity. We used A/Anhui/1/2013 (H7N9) virus as an NA inhibitor–sensitive control and its NA inhibitor–resistant variant, A/Anhui/1/2013-NA-R294K, as an NA inhibitor–resistant control (online Technical Appendix Table 4) or by the small number of animals used in these studies. We also performed transmission studies in cats and detected feline H7N2 subtype virus transmission via direct contact and respiratory droplets. However, the group size used is a potential limitation of our study.

Cats are not a major reservoir of influenza A viruses, but can be infected naturally or experimentally with influenza viruses of different subtypes (23). Serologic surveys suggest high and low rates of seroconversion to seasonal
human and highly pathogenic avian influenza viruses, respectively. Natural infections most likely result from close contact with infected humans or animals, and most of these infections appear to be self-limiting.

Few cases of human infections with influenza viruses of the H7 subtype were reported until 2013, and they typically caused mild illness; however, infection of a veterinarian with a highly pathogenic avian H7N7 virus had fatal consequences (24,25). Since 2013, influenza viruses of the H7N9 subtype have caused more than 1,300 laboratory-confirmed infections in humans, with a case-fatality rate of ≈30%. Although the current H7N9 and feline H7N2 subtype viruses do not exclusively bind to human-type receptors and do not transmit efficiently among humans, the spread and biologic properties of these viruses should be monitored carefully.

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Characterization of a Feline Influenza A(H7N2) Virus

Technical Appendix

Supplementary Methods

Cells and Viruses

Madin-Darby canine kidney (MDCK) cells (obtained from ATCC) were maintained in Eagle’s minimal essential medium (MEM) containing 5% newborn calf serum and antimicrobial drugs. Human lung carcinoma epithelial A549 cells were propagated in a 1:1 mixture of Dulbecco’s modified Eagle’s medium (DMEM) and Ham’s F12 medium containing 10% fetal calf serum (FCS) with antimicrobial drugs. Human airway epithelial cells (Calu-3, obtained from Raymond Pickles, University of North Carolina, Chapel Hill, NC, USA) were cultured in DMEM/F12 medium supplemented with 10% FCS and antimicrobial drugs. Chicken embryo fibroblast (CEF) cells were prepared from 10-day-old chicken embryos and cultured in DMEM with 10% FCS and antimicrobial drugs. Cat kidney fibroblast Clone81 (ECACC 90031403) and cat lung Fc2Lu (ECACC 90112712) cells were purchased from the European Collection of Authenticated Cell Cultures (ECACC). Clone81 cells were cultured in DMEM with 10% FCS and antimicrobial drugs. Fc2Lu cells were maintained in MEM with 1% non-essential amino acids (NEAA), 10% FCS, and antimicrobial drugs. All cells were maintained at 37 °C with 5% CO₂ unless otherwise stated.

The viral genomic sequences of the 5 feline H7N2 viruses have been deposited in GenBank under the following accession numbers: A/feline/New York/WVDL-3/2016: MF978390–MF978397; A/feline/New York/WVDL-9/2016: MF978398–MF978405; A/feline/New York/WVDL-14/2016: MF978406–MF978413; A/feline/New York/WVDL-16/2016: MF978414–MF978421; A/feline/New York/WVDL-20/2016: MF978422–MF978429. The sequences of the HA, NA, M, and NS segments of A/chicken/NY/22409–4/1999 virus were available in GenBank (accession nos. AY240896, AY254122, AY241605, and AY241644, respectively) (7). We (re)sequenced all 8 viral segments and deposited the sequences of the PB2,
PB1, PA, and NP segments in GenBank under accession nos. MF988320–MF988323. The sequences of the HA and NA segments differed from AY240896 and AY254122 at the nucleotide, but not at the amino acid level, and were deposited in GenBank under accession nos. MF988323 and MF98825, respectively. The sequences of the M and NS segments were identical to AY241605 and AY241644, respectively, and therefore were not submitted to GenBank.

**Negative Staining**

MDCK cells were infected with A/feline/NY/16 and cultured in 1× MEM containing 0.3% bovine serum albumin and trypsin treated with L-1-tosylamide-2-phenylethyl chloromethyl ketone at 37°C. Forty-eight hours later, the supernatants were harvested and cell debris was removed by centrifugation at 1,750 x g. The virion-containing supernatants were adsorbed to Formvar-coated copper mesh grids, negatively stained with 2% phosphotungstic acid solution, and air dried. Digital images of virions were taken with a Tecnai F20 electron microscope (FEI, Tokyo, Japan) at 200 kV.

**Animal Experiments**

All experiments with mice, ferrets, and cats were performed in accordance with the guidelines set by the Institutional Animal Care and Use Committee at the University of Wisconsin–Madison, which also approved the protocols used (protocol numbers V00806 and V01190). The facilities where this research was conducted are fully accredited by the Association for the Assessment and Accreditation of Laboratory Animal Care International. The animal experiments described in this study were not designed to generate datasets for statistical analysis; hence, the sample size was small and randomization and blinding were not performed.

**Immunohistochemistry**

Tissues excised from animal organs preserved in 10% phosphate-buffered formalin were processed for paraffin embedding and cut into 5- and 3-μm-thick sections for hematoxylin and eosin staining and immunohistological staining, respectively. One section from each tissue sample was stained using a standard hematoxylin and eosin procedure; another sample was processed for immunohistological staining with a mouse monoclonal or rabbit polyclonal antibody for type A influenza nucleoprotein antigen (prepared in our laboratory) that reacts comparably with all of the viruses used in this study. Specific antigen–antibody reactions were visualized with 3,3′- diaminobenzidine tetrahydrochloride staining by using the DAKO LSAB2
system (Agilent, Santa Clara, CA, USA). Our negative controls (not shown) included sections from mock-infected animals. As a positive control (also not shown), we used formalin-fixed, paraffin-embedded lung sections from humans infected with seasonal influenza viruses.

Detection of α2,3- and α2,6-linked Sialosides in Cat Organs

To detect α2,3- and α2,6-linked sialosides (2–4), the tissues of a naive cat were fixed in 4% paraformaldehyde–phosphate-buffered saline (PBS) and embedded in paraffin. The paraffin blocks were cut into 3-μm-thick sections and mounted on silane-coated glass slides. The sections were pretreated with 0.05% trypsin (Difco Laboratories, Detroit, MI, USA) at 37°C for 15 min and then with 0.3% hydrogen peroxide at room temperature for 30 min. They were then incubated at 4°C overnight with biotin-conjugated Sambucus nigra lectin I (SNA I; EY Laboratories, San Mateo, CA, USA) to detect α2,6-linked sialosides, or with biotinylated-Maackia amurensis lectin I and II (MAAI and II; Vector Laboratories, Burlingame, CA, USA) to detect α2,3-linked sialosides. After being washed, the sections were incubated with horseradish peroxidase-conjugated streptavidin and visualized by staining with 3,3-diaminobenzidine (DAB).

Neuraminidase Inhibition Assay

Diluted viruses were mixed with different concentrations of oseltamivir carboxylate (the active form of oseltamivir), zanamivir, or laninamivir (all obtained from Daiichi Sankyo Co., Ltd, Tokyo, Japan) (5,6). Samples were incubated for 30 min at 37°C, followed by the addition of methylumbelliferyl-N-acetylneuraminic acid (Sigma, St Louis, MO) as a fluorescent substrate (7,8). After incubation for 1 h at 37°C, the reaction was stopped with the addition of sodium hydroxide in 80% ethanol. The fluorescence of the solution was measured at an excitation wavelength of 360 nm and an emission wavelength of 465 nm, and the 50% inhibitory concentration (IC50) was calculated.

Glycan Array Analysis

Glycan array analysis was performed on a glass slide microarray containing 6 replicates of 130 diverse sialic acid-containing glycans, including terminal sequences and intact N-linked and O-linked glycans found on mammalian and avian glycoproteins and glycolipids (9). Viruses were amplified in MDCK cells. Supernatants collected from infected cells were centrifuged at 1,462 × g for 30 min to remove cell debris. Viruses were inactivated by mixing the supernatants
with 0.1% β-propiolactone (final concentration). Virus supernatant was laid over a cushion of 30% sucrose in PBS, ultracentrifuged at 76,755 × g for 2 h at 4°C, and then resuspended in PBS for storage at −80°C. Virus samples (equivalent of 128 hemagglutination units) were incubated on the array surface for 1 h at room temperature, and labeled with mouse monoclonal anti-H7/H1 IgG and goat anti-mouse IgG-Alex Fluor 488 antibodies for sequential 1-hour incubations. Slide scanning to detect virus bound to glycans was conducted using an Innoscan1100AL (Innopsys, Carbone, France) fluorescent microarray scanner. Fluorescent signal intensity was measured using Mapix (Innopsys, Carbone, France) and mean intensity minus mean background of 4 replicate spots was calculated. A complete list of the glycans on the array is presented in Technical Appendix, Table 2.

Hemagglutination Inhibition (HI) Assay

To detect hemagglutination inhibition (HI) activity (https://www.cdc.gov/flu/professionals/laboratory/antigenic.htm), serum samples were treated with receptor-destroying enzyme (Denka Seiken Co., Ltd., Tokyo, Japan) at 37°C for 16–20 hours, followed by receptor-destroying enzyme inactivation at 56°C for 30–60 min. The treated sera were serially diluted 2-fold with PBS in 96-well U-bottom microtiter plates (Thermo Scientific, Rochester, NY, USA) and mixed with the amount of virus equivalent to eight hemagglutination units, followed by incubation at room temperature (25°C) for 30 min. After 50 μL of 0.5% turkey erythrocytes was added to the mixtures, they were gently mixed and incubated at room temperature for a further 45 min. HI titers are expressed as the inverse of the highest antibody dilution that inhibited hemagglutination.

Statistical Analyses

We compared the values obtained for each strain, using a 2-way ANOVA, and creating a matrix of contrasts to compare each time-point separately. We then adjusted the p values by using Holm’s method to account for family-wise errors; we considered the differences significant if we obtained p values <0.05.

Phylogenetic Analysis

Phylogenetic analyses were carried out for selected avian and human influenza A viruses representing major lineages. The evolutionary history was inferred using the neighbor-joining method (10). The optimal trees were selected and the percentages of replicate trees in which the
associated taxa clustered together in the bootstrap test (500 replicates) (11) were identified. The
trees were drawn to scale, with branch lengths in the same units as those of the evolutionary
distances used to infer the phylogenetic tree. The evolutionary distances were computed using
the Tamura 3-parameter method (12) and are in the units of the number of base substitutions per
site. Codon positions included were 1st + 2nd + 3rd + Noncoding. All positions containing gaps
and missing data were eliminated. Evolutionary analyses were conducted in MEGA7 (13).

**Biosafety and Biosecurity**

All recombinant DNA protocols were approved by the University of Wisconsin–
Madison’s Institutional Biosafety Committee after risk assessments were conducted by the
Office of Biologic Safety. In addition, the University of Wisconsin–Madison Biosecurity Task
Force regularly reviews the research program and ongoing activities of the laboratory. The task
force has a diverse skill set and provides support in the areas of biosafety, facilities, compliance,
security, and health. Members of the Biosecurity Task Force are in frequent contact with the
principal investigator and laboratory personnel to provide oversight and assure biosecurity. The
H7N2 viruses used in this study are low pathogenicity avian viruses according to the definition
by the US Department of Agriculture and experiments with these viruses can be conducted in
Biosafety Level 2+ (BSL2+) containment. For animal experiments with the feline H7N2 viruses,
staff wore personal protective equipment including disposable coveralls, double gloves,
dedicated shoes with disposable shoe covers, and powered air-purifying respirators that HEPA
filter the air for extra protection. Ferret studies were conducted in BSL3 containment. All
personnel working in BSL3 containment complete rigorous biosafety, BSL3, and Select Agent
(for the US laboratory) training before participating in research studies. The principal
investigator participates in training sessions and emphasizes compliance to maintain safe
operations and a responsible research environment. The laboratory occupational health plans are
in compliance with the policies of the University of Wisconsin–Madison.

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**Technical Appendix Table 1.** Antigenic characterization of H7 viruses by use of monoclonal antibodies

| Virus (Subtype)                                | Hemagglutination inhibition (HI) titers* |
|-----------------------------------------------|-----------------------------------------|
|                                              | HA from A/Seal/Mass/1/80 (H7N7)          |
|                                              | HA from A/Netherlands/219/03 (H7N7)      |
|                                              | HA from A/Anhui/1/2013 (H7N9)            |
|                                              |                                         |
| A/feline/New York/WVDL-14/2016 (Feline/NY/16) | 46/6                                    |
| A/chicken/NY/22409–4/1999 (Chicken/NY/99)   | 55/3                                    |
| A/duck/Hong Kong/301/1978 H7N2                | 58/2                                    |
|                                              | 8/4                                     |
|                                              | 10C6                                    |
|                                              | 2–20–20                                 |
|                                              | 19–17–20                                |
| A/turkey/England/1963 H7N3                   | 200                                     |
| A/turkey/Oregon/1971 H7N3                    | 400                                     |
| A/turkey/Tennessee/1/1978 H7N3               | 800                                     |
| A/chicken/Japan/1925 H7N7                   | 200                                     |
| A/equine/Prague/1/1956 H7N7                 | 400                                     |
| A/equine/New Market/1/1977 H7N7             | 800                                     |
| A/seal/Massachusetts/1/1980 3200             | 6400                                    |
| A/turkey/England/1963 H7N3                   | 800                                     |
|                                             | 800                                     |
| A/turkey/Oregon/1971 H7N3                    | 200                                     |
| A/turkey/Tennessee/1/1978 H7N3               | 400                                     |
| A/chicken/Japan/1925 H7N7                   | 800                                     |
| A/equine/Prague/1/1956 H7N7                 | 200                                     |
| A/equine/New Market/1/1977 H7N7             | 400                                     |
| A/seal/Massachusetts/1/1980 3200             | 6400                                    |
| A/turkey/England/1963 H7N3                   | 800                                     |
| A/turkey/Oregon/1971 H7N3                    | 200                                     |
| A/turkey/Tennessee/1/1978 H7N3               | 400                                     |
| A/chicken/Japan/1925 H7N7                   | 800                                     |
| A/equine/Prague/1/1956 H7N7                 | 200                                     |
| A/equine/New Market/1/1977 H7N7             | 400                                     |
| A/seal/Massachusetts/1/1980 3200             | 6400                                    |
| A/turkey/England/1963 H7N3                   | 800                                     |
| A/turkey/Oregon/1971 H7N3                    | 200                                     |
| A/turkey/Tennessee/1/1978 H7N3               | 400                                     |
| A/chicken/Japan/1925 H7N7                   | 800                                     |
| A/equine/Prague/1/1956 H7N7                 | 200                                     |
| A/equine/New Market/1/1977 H7N7             | 400                                     |
| A/seal/Massachusetts/1/1980 3200             | 6400                                    |
| A/turkey/England/1963 H7N3                   | 800                                     |
| A/turkey/Oregon/1971 H7N3                    | 200                                     |
| A/turkey/Tennessee/1/1978 H7N3               | 400                                     |
| A/chicken/Japan/1925 H7N7                   | 800                                     |
| A/equine/Prague/1/1956 H7N7                 | 200                                     |
| A/equine/New Market/1/1977 H7N7             | 400                                     |
| A/seal/Massachusetts/1/1980 3200             | 6400                                    |
| A/turkey/England/1963 H7N3                   | 800                                     |
| A/turkey/Oregon/1971 H7N3                    | 200                                     |
| A/turkey/Tennessee/1/1978 H7N3               | 400                                     |
| A/chicken/Japan/1925 H7N7                   | 800                                     |
| A/equine/Prague/1/1956 H7N7                 | 200                                     |
| A/equine/New Market/1/1977 H7N7             | 400                                     |
| A/seal/Massachusetts/1/1980 3200             | 6400                                    |
| A/turkey/England/1963 H7N3                   | 800                                     |
| A/turkey/Oregon/1971 H7N3                    | 200                                     |
| A/turkey/Tennessee/1/1978 H7N3               | 400                                     |
| A/chicken/Japan/1925 H7N7                   | 800                                     |

*Hemagglutination inhibition (HI) assays were carried out as follows: 2-fold serial dilutions of antibodies were mixed with the amount of virus equivalent to 8 hemagglutination units of virus in 96-well U-bottom microtiter plates, followed by incubation at room temperature for 60 min. After an equal volume of 0.5% chicken red blood cells was added, the mixtures were gently mixed and then incubated for a further 60 min at 4°C. HI titers were determined as the inverse of the highest antibody dilution that inhibited the hemagglutination.

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**Technical Appendix Table 2. List of glycans used for arrays**

| No | M#   | S#   | Common Name                                                                 | Linkage | NeuAC/Neu Gc(A/B), both -C | Structure |
|----|------|------|------------------------------------------------------------------------------|---------|----------------------------|-----------|
| 1  | M040 | M040 | Galβ(1–4)-GlcNAcβ-ethyl-NH2                                                  | –       | –                          |           |
| 2  | M221 | WJ-5–149–1 | Galβ(1–4)-GlcNAcβ(1-3)-Galβ(1-3)-GlnNac-Thr-NH2 | –       | –                         |           |
| 3  | M222 | 152Sp14 | Galβ(1–4)-GlcNAcβ(1-6)[Galβ(1-3)]-GlnNac-Thr-NH2 | –       | –                         |           |
| 4  | M223 | 144Sp14 | Galβ(1–4)-GlcNAcβ(1-3)-GlnNac-Thr-NH2                                      | –       | –                         |           |
| 5  | M224 | 21Sp14 | Galβ(1–4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-6)]-GlnNac-Thr-NH2              | –       | –                         |           |
| 6  | M225 | 119Sp14 | Galβ(1–4)-GlcNAcβ(1-6)-GlnNac-Thr-NH2                                       | –       | –                         |           |
| 7  | M009 | M009 | Galβ(1–4)-GlcNAcβ(1-2)-Manβ(1-3)[Galβ(1-4)-GlcNAcβ(1-2)-Manβ(1-6)]-Manβ(1-4)-GlcNAcβ(1-5)-GlcNAcβ-Asn-NH2 | –       | –                         |           |
| 8  | M226 | 375Sp22 | Galβ(1–4)-GlcNAcβ(1-2)-Manβ(1-3)[Galβ(1-4)-GlcNAcβ(1-2)-Manβ(1-6)]-Manβ(1-4)-GlcNAcβ(1-4)[Fuc(1-6)-GlcNAcβ-Asn-Ser-Thr-NH2 | –       | –                         |           |
| No | M#     | S#     | Common Name                                                                 | Linkage | NeuAC/Neu Gc(A/B), both -C | Structure |
|----|--------|--------|------------------------------------------------------------------------------|---------|---------------------------|-----------|
| 9  | M227   | 487Sp1 | NeuAc(1-3)Galβ(1-4)GlcNAcβ(1-2)Manα(1-3)(Galβ(1-4)GlcNAcβ(1-2)) | –       | –                         | ![Structure](image) |
| 10 | M228   | 517Sp  | NeuAc(1-3)Galβ(1-4)GlcNAcβ(1-2)Manα(1-3)(Galβ(1-4)GlcNAcβ(1-2)) | –       | –                         | ![Structure](image) |
| 11 | M001   | M001   | NeuAc(2-3)Galβ(1-4)-6-O-sulfoGlcNAcβ-propyl-NH2                          | 3       | A                         | ![Structure](image) |
| 12 | M037   | M037   | NeuAc(2-3)-Galβ(1-4)-6-O-sulfo-GlcNAcβ-propyl-NH2                        | 3       | A                         | ![Structure](image) |
| 13 | M039   | M039   | NeuAc(2-3)-6-O-sulfo-Galβ(1-4)-GlcNAcβ-ethyl-NH2                         | 3       | A                         | ![Structure](image) |
| 14 | M036   | M036   | NeuAc(2-3)-6-O-sulfo-Galβ(1-4)-(Fucα(1-3))-GlcNAcβ-propyl-NH2           | 3       | A                         | ![Structure](image) |
| 15 | M038   | M038   | NeuAc(2-3)-Galβ(1-3)-6-O-sulfo-GlcNAcβ-propyl-NH2                      | 3       | A                         | ![Structure](image) |
| No | M#  | S#  | Common Name                  | Linkage | NeuAC/NeuGc(A/B), both - C | Structure |
|----|-----|-----|------------------------------|---------|-----------------------------|-----------|
| 16 | M011| M011| NeuAca(2→3)-βGal(1→4)-αGlcβ-ethyl-NH₂ | 3       | A                           |           |
| 17 | M012| M012| NeuAca(2→3)-βGal(1→4)-αGlcNAcβ-ethyl-NH₂ | 3       | A                           |           |
| 18 | M014| M014| NeuAca(2→3)-βGal(1→4)-αGlcNAcβ(1→3)-βGal(1→4)-αGlcNAcβ-ethyl-NH₂ | 3       | A                           |           |
| 19 | M035| M035| NeuAca(2→3)-βGal(1→4)-αGlcNAcβ(1→3)-βGal(1→4)-αGlcNAcβ(1→3)-βGal(1→4)-αGlcNAcβ-ethyl-NH₂ | 3       | A                           |           |
| 20 | M013| M013| NeuAca(2→3)-βGalNAcβ(1→4)-αGlcNAcβ-ethyl-NH₂ | 3       | A                           |           |
| 21 | M010| M010| NeuAca(2→3)-βGal(1→3)-αGlcNAcβ-ethyl-NH₂ | 3       | A                           |           |
| 22 | M032| M032| NeuAca(2→3)-βGal(1→3)-αGlcNAcβ(1→3)-βGal(1→4)-αGlcNAcβ-ethyl-NH₂ | 3       | A                           |           |
| 23 | M033| M033| NeuAca(2→3)-βGal(1→3)-αGlcNAcβ(1→3)-βGal(1→4)-αGlcNAcβ-ethyl-NH₂ | 3       | A                           |           |
| 24 | M028| M028| NeuAca(2→3)-βGal(1→3)-βGalNAcβ(1→3)-βGal(1→4)-αGlcβ-ethyl-NH₂ | 3       | A                           |           |
| 25 | M045| M045| NeuAca(2→3)-βGal(1→3)-βGalNAcβ-Thr-NH₂ | 3       | A                           |           |
|   | M#  | WJ  | Sequence                  | 3' NeuAc LN Core 1/2 | 3' NeuAc DILN Core 1 | 3' NeuAc TriLN Core 1 | 3' NeuAc TetraLN Core 1/2 | 3' NeuAc PentaLN Core 1 | Notes |
|---|-----|-----|---------------------------|-----------------------|-----------------------|------------------------|---------------------------|-------------------------|-------|
| 26 | M120| WJ-6–121–1 | 3' NeuAc LN Core 1 (1163) | A                     |                       |                        |                           |                         |       |
| 27 | M128| WJ-6–153–1 | 3' NeuAc DILN Core 1 (1528) | A                     |                       |                        |                           |                         |       |
| 28 | M153| WJ-8–145–1 | 3' NeuAc TriLN Core 1 (1894) | A                     |                       |                        |                           |                         |       |
| 29 | M142| WJ-8–101–1 | 3' NeuAc TetraLN Core 1 (2259) | A                     |                       |                        |                           |                         |       |
| 30 | M143| WJ-8–103–1 | 3' NeuAc PentaLN Core 1 (2624) | A                     |                       |                        |                           |                         |       |
| 31 | M050| M050 | NeuAc\(2–3\)–Gal\[(1–4)–GlcNAc\[(1–6)\]–Gal\[(1–3)\]–GalNAc–Thr–NH\_2 | A                     |                       |                        |                           |                         |       |
| 32 | M053| M053 | NeuAc\(2–3\)–Gal\[(1–4)–GlcNAc\[(1–3)\]–Gal\[(1–4)–GlcNAc\[(1–6)\]–Gal\[(1–3)\]–GalNAc–Thr–NH\_2 | A                     |                       |                        |                           |                         |       |
| 33 | M202| WJ-9–41–1 | 3' NeuAc TriLN Core 2 (1894) | A                     |                       |                        |                           |                         |       |
| 34 | M152| WJ-8–141–1 | 3' NeuAc TetraLN Core 2 (2259) | A                     |                       |                        |                           |                         |       |
| 35 | M149| WJ-8–131–1 | 3' NeuAc PentaLN Core 2 (2624) | A                     |                       |                        |                           |                         |       |
| 36 | M195| WJ-9–13–1 | 3' NeuAc TetraLN TriLN Core 2 (3645) | A                     |                       |                        |                           |                         |       |
|   |   |   |   |   |   |
|---|---|---|---|---|---|
| 37 | M156 | WJ-8–155–1 | 3' NeuAc TetraLN Core 2 (4376) | 3 | A |
| 38 | M055 | M055 | NeuAc(2–3)-Gal[1(1–4)-GlcNAc[1(1–3)]GalNAc-Thr-NH2 | 3 | A |
| 39 | M057 | M057 | NeuAc(2–3)-Gal[1(1–4)-GlcNAc[1(1–3)]Gal[1(1–4)-GlcNAc[1(1–3)]GalNAc-Thr-NH2 | 3 | A |
| 40 | M186 | WJ-8–99–1 | 3' NeuAc TriLN Core 3 (1732) | 3 | A |
| 41 | M178 | WJ-8–83–1 | 3' NeuAc TetraLN Core 3 (2097) | 3 | A |
| 42 | M177 | WJ-8–77–1 | 3' NeuAc PentaLN Core 3 (2462) | 3 | A |
| 43 | M059 | M059 | NeuAc(2–3)-Gal[1(1–4)-GlcNAc[1(1–3)]NeuAc(2–3)-Gal[1(1–4)-GlcNAc[1(1–3)] | 3 | A |
| 44 | M061 | M061 | NeuAc(2–3)-Gal[1(1–4)-GlcNAc[1(1–3)]NeuAc(2–3)-Gal[1(1–4)-GlcNAc[1(1–3)] | 3 | A |
| 45 | M185 | WJ-8–97–1 | 3' NeuAc TriLN Core4 (3118) | 3 | A |
| 46 | M180 | WJ-8–87–1 | 3' NeuAc TetraLN Core4 (3848) | 3 | A |
| 47 | M179 | WJ-8–85–1 | 3' NeuAc PentaLN Core4 (4579) | 3 | A |
|   |   |   |   |   |
|---|---|---|---|---|
| 48 | M182 | WJ-8–91–1 | 3' NeuAc TetraLN Core6 (2097) | 3 | A |
| 49 | M184 | WJ-8–95–1 | 3' NeuAc PentaLN Core6 (2462) | 3 | A |
| 50 | M102 | WJ-10–71–1 | 3' NeuAc LecLN I-Antigen (2104) | 3 | A |
| 51 | M098 | WJ-10–61–1 | 3' NeuAc TriLN I-Antigen (2855) | 3 | A |
| 52 | M026 | M026 | NeuAc(2–3)-Gal[(1–4)-GlcNAc](1-2)-Manu(1–3)-(NeuAc(2–3)-Gal)[1-] | 3 | A |
| 53 | M041 | M041 | NeuAc(2–3)-Gal[(1–4)-GlcNAc](1-3)-Gal[(1–4)-GlcNAc](1–2)-Manu(1–3)- | 3 | A |
| 54 | M043 | M043 | NeuAc(2–3)-Gal[(1–4)-GlcNAc](1-3)-Gal[(1–4)-GlcNAc](1–2)-Manu(1–3)-Gal[1-] | 3 | A |
| 55 | M107 | WJ-5–21–1 | 3' NeuAc DiLN Bi-(3594) | 3 | A |
| 56 | M110 | WJ-5–35–1 | 3' NeuAc TriLN Bi-(4324) | 3 | A |
| 57 | M112 | WJ-5–39–1 | 3' NeuAc TetraLN Bi-(4828) | 3 | A |
| 58 | M114 | WJ-5–45–1 | 3' NeuAc PentaLN Bi-(5556) | 3 | A |
| Page | M132 | WJ-6–41–1 | 3' NeuAc TriLN Bi-CF(4470) | 3 | A |
|------|------|------------|-----------------------------|---|---|
| 60   | M122 | WJ-6–13–1 | 3' NeuAc TetraLN Bi-CF(5200) | 3 | A |
| 61   | M118 | WJ-6–117–1| 3' NeuAc DiLN Tri-(4615)     | 3 | A |
| 62   | M119 | WJ-6–119–1| 3' NeuAc TriLN Tri-(5716)    | 3 | A |
| 63   | M141 | WJ-7–47–1 | 3' NeuAc TetraLN Tri-(6808)  | 3 | A |
| 64   | M125 | WJ-6–149–1| 3' NeuAc DiLN Tri-CF(4761)   | 3 | A |
| 65   | M127 | WJ-6–151–1| 3' NeuAc TriLN Tri-CF(5858)  | 3 | A |
| 66   | M068 | 701_WJ-10–91–1 | Gn/3'SLN/3'SLN-TriN | 3 | A |
| 67   | M031 | M031 | NeuAca(2–3)-[GalNAcβ(1–4)]-Galβ(1–4)-GlcNAcβ-ethyl-NH₂ | 3 | A |
| 68   | M016 | M016 | NeuAca(2–3)-[GalNAcβ(1–4)]-Galβ(1–4)-Glcβ-ethyl-NH₂ | 3 | A |
| 69   | M017 | M017 | Galβ(1–3)-GalNAcβ(1–4)-[NeuAca(2–3)]-Galβ(1–4)-Glcβ-ethyl-NH₂ | 3 | A |
|    | M002 | M002 | NeuAc(2-3)-Galβ(1-4)-[Fuca(1-3)]-GlcNAc-propyl-NH₂ | 3 | A |
|----|------|------|---------------------------------------------------|---|---|
| 70 | M029 | M029 | NeuAc(2-3)-Galβ(1-3)-[Fuca(1-4)]-GlcNAc(1-3)-Galβ(1-4)-[Fuca(1-3)] | 3 | A |
| 71 | M022 | M022 | NeuAc(2-3)-Galβ(1-4)-[Fuca(1-3)]-GlcNAc(1-3)-Galβ(1-4)-[Fuca(1-3)] | 3 | A |
| 72 | M015 | M015 | NeuAc(2-3)-Galβ(1-4)-[Fuca(1-3)]-GlcNAc(1-3)-Galβ(1-4)-[Fuca(1-3)]-GlcNAc(1-3)-Galβ(1-4)-[Fuca(1-3)] | 3 | A |
| 73 | M206 | WJ-9-7-1 | 3' SLeX TriLN Core 1(2332) | 3 | A |
| 74 | M147 | WJ-8-127-1 | 3' SLeX TriLN Core 3(2170) | 3 | A |
| 75 | M146 | WJ-8-125-1 | 3' SLeX TriLN Core 4(3994) | 3 | A |
| 76 | M219 | WJ-119-1 | NeuAc(2-3)Galβ(1-4)[Fuca(1-3)]-GlcNAc(1-2)Manα(1-3)[NeuAc(2-3)Galβ(1-4)[Fuca(1-3)]-GlcNAc(1-2)Manα(1-6)]-Manβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)GlcNAcβ(1-4)[Fuca(1-3)]-GlcNAc-propyl-NH₂ | 3 | A |
| 77 | M215 | WJ-12-79-1 | NeuAc(2-6)-Galβ(1-4)-(6S)GlcNab-ethyl-NH₂ | 6 | A |
| 78 | M003 | M003 | NeuAc(2-6)-Galβ(1-4)-6-O-sulfo-GlcNAc-ethyl-NH₂ | 6 | A |
| M018 | M018 | NeuAcα(2–6)-Galβ(1–4)-Glcβ-ethyl-NH₂ | 6 | A | ![Image](https://via.placeholder.com/150) |
| M019 | M019 | NeuAcα(2–6)-Galβ(1–4)-GlcNAcβ-ethyl-NH₂ | 6 | A | ![Image](https://via.placeholder.com/150) |
| M021 | M021 | NeuAcα(2–6)-Galβ(1–4)-GlcNAcβ(1–3)-Galβ(1–4)-GlcNAcβ(1–3)-Galβ(1–4)-GlcNAcβ | 6 | A | ![Image](https://via.placeholder.com/150) |
| M020 | M020 | NeuAcα(2–6)-GalNAcβ(1–4)-GlcNAcβ-ethyl-NH₂ | 6 | A | ![Image](https://via.placeholder.com/150) |
| M121 | M121 | WJ-6–123–1 | 6 | A | ![Image](https://via.placeholder.com/150) |
| M129 | M129 | WJ-6–155–1 | 6 | A | ![Image](https://via.placeholder.com/150) |
| M135 | M135 | WJ-7–149–1 | 6 | A | ![Image](https://via.placeholder.com/150) |
| M148 | M148 | WJ-8–13–1/WJ-7–107–1 | 6 | A | ![Image](https://via.placeholder.com/150) |
| M051 | M051 | NeuAcα(2–6)-Galβ(1–4)-GlcNAcβ(1–6)-Galβ(1–3)-GlcNAc-Tyr-Thr-NH₂ | 6 | A | ![Image](https://via.placeholder.com/150) |
| #  | Code  | Type             | Structure                                              | Anno. | Pm. |
|-----|-------|------------------|--------------------------------------------------------|-------|-----|
| 91  | M054  | M054             | NeuAc(2–6)-Galβ(1–4)-GlcNAcβ(1–3)-Galβ(1–4)-GlcNAcβ(1–6)-Galβ(1–4)-GlcNAcβ(1–3)-GalNAc-Thr-NH₂ |       | A   |
| 92  | M201  | WJ-9–39-1        | 6° NeuAc TrLN Core 2 (1894)                            |       | A   |
| 93  | M159  | WJ-8–23-1        | 6° NeuAc TetraLN Core 2 (2259)                         |       | A   |
| 94  | M157  | WJ-8–17-1        | 6° NeuAc PentaLN Core 2 (2624)                         |       | A   |
| 95  | M163  | WJ-8–33-1        | 6° NeuAc TetraLN TrLN Core 2 (3645)                    |       | A   |
| 96  | M161  | WJ-8–29-1        | 6° NeuAc PentaLN TetraLN Core 2 (4376)                 |       | A   |
| 97  | M056  | M056             | NeuAc(2–6)-Galβ(1–4)-GlcNAcβ(1–3)-GalNAc-Thr-NH₂      |       | A   |
| 98  | M058  | M058             | NeuAc(2–6)-Galβ(1–4)-GlcNAcβ(1–3)-Galβ(1–4)-GlcNAcβ(1–3)-GalNAc-Thr-NH₂ |       | A   |
| 99  | M172  | WJ-8–65-1        | 6° NeuAc TrLN Core 3 (1732)                            |       | A   |
| 100 | M166  | WJ-8–49-1        | 6° NeuAc TetraLN Core 3 (2097)                         |       | A   |
| 101 | M164  | WJ-8–35-1        | 6° NeuAc PentaLN Core 3 (2462)                         |       | A   |
| Page | M060 | M060 | 102 | NeuAc[2–6]-Gal[1–4]-GlcNAc[1–3]–[NeuAc[2–6]-Gal[1–4]-GlcNAc[1–6]] | 6 | A |
|------|------|------|-----|-------------------------------------------------|---|---|
|      | M062 | M062 | 103 | NeuAc[2–6]-Gal[1–4]-GlcNAc[1–3]–Gal[1–4]-GlcNAc[1–3]–[NeuAc[2–6]-Gal[1–4]-GlcNAc[1–6]] | 6 | A |
|      | M174 | WJ–8–73–1 | 104 | NeuAc TrLN Core4 (3118) | 6 | A |
|      | M170 | WJ–8–61–1 | 105 | NeuAc TetraLN Core4 (3848) | 6 | A |
|      | M168 | WJ–8–57–1 | 106 | NeuAc PentaLN Core4 (4579) | 6 | A |
|      | M181 | WJ–8–89–1 | 107 | NeuAc TetraLN Core6 (2097) | 6 | A |
|      | M183 | WJ–8–93–1 | 108 | NeuAc PentaLN Core6 (2462) | 6 | A |
|      | M097 | WJ–10–59–1 | 109 | NeuAc TrLN I-Antigen (2856) | 6 | A |
|      | M104 | WJ–10–77–1 | 110 | NeuAc DiLN I-Antigen (2104) | 6 | A |
|      | M006 | M006 | 111 | Gal[1–4]–GlcNAc[1–2]–Man[1–3]–[NeuAc[2–6]-Gal[1–4]-GlcNAc[1–2]] | 6 | A |
|      | M007 | M007 | 112 | NeuAc[2–6]-Gal[1–4]-GlcNAc[1–2]-Man[1–3]-Gal[1–4]-GlcNAc[1–2] | 6 | A |
|   |   | GlcNAc(1→2)-Man(1→3) \[NeuAc(2→6)-Gal(1→4)\]-GlcNAc(1→2)-Man(1→6) | 6 |
|---|---|---|---|
| 113 | M008 | M008 | A |

|   |   | NeuAc(2→6)-Gal(1→4)-GlcNAc(1→2)-Man(1→3) \[NeuAc(2→6)-Gal(1→4)\]-GlcNAc(1→2)-Man(1→6) | 6 |
|---|---|---|---|
| 114 | M004 | M004 | A |

|   |   | NeuAc(2→6)-Gal(1→4)-GlcNAc(1→2)-Man(1→3) \[NeuAc(2→6)-Gal(1→4)\]-GlcNAc(1→2)-Man(1→6) | 6 |
|---|---|---|---|
| 115 | M042 | M042 | A |

|   |   | NeuAc(2→6)-Gal(1→4)-GlcNAc(1→3)-Gal(1→4)-GlcNAc(1→2)-Man(1→6) \[NeuAc(2→6)-Gal(1→4)\]-GlcNAc(1→2)-Man(1→6) | 6 |
|---|---|---|---|
| 116 | M109 | WJ-5-33-1 | A |

|   |   | NeuAc(2→6)-Gal(1→4)-GlcNAc(1→3)-Gal(1→4)-GlcNAc(1→2)-Man(1→6) \[NeuAc(2→6)-Gal(1→4)\]-GlcNAc(1→2)-Man(1→6) | 6 |
|---|---|---|---|
| 117 | M044 | M044 | A |

|   |   | NeuAc(2→6)-Gal(1→4)-GlcNAc(1→3)-Gal(1→4)-GlcNAc(1→2)-Man(1→6) \[NeuAc(2→6)-Gal(1→4)\]-GlcNAc(1→2)-Man(1→6) | 6 |
|---|---|---|---|
| 118 | M089 | JP-3–8-1 | A |

|   |   | NeuAc (3594) | 6 |
|---|---|---|---|
| 119 | M081 | JP-3–12-1 | A |

|   |   | NeuAc TetraLN Bi-(4828) | 6 |
|---|---|---|---|
| 120 | M083 | JP-3–16-1 | A |

|   |   | NeuAc PentaLN Bi-(5556) | 6 |
|---|---|---|---|
| 121 | M085 | JP-3–20-2 | A |

|   |   | NeuAc DiLN Bi-CF(3740) | 6 |
|---|---|---|---|
| 122 | M087 | JP-3–24-1 | A |

|   |   | NeuAc TriLN Bi-CF(4470) | 6 |
|---|---|---|---|
Technical Appendix Table 3. Virus sensitivity to NA inhibitors

| NA inhibitors          | Chicken/NY/99 | Feline/NY/16 | A/Anhui/1/2013<sup>§</sup> (H7N9) | A/Anhui/1/2013<sup>¶</sup>-NA-R294K<sup>¶</sup> (H7N9) |
|------------------------|---------------|--------------|-----------------------------------|-----------------------------------------------------|
| Oseltamivir carboxylate<sup>†</sup> | 1.6           | 1.0          | 3.6                               | 64,000                                               |
| Zanamivir              | 5.6           | 8.2          | 8.1                               | 340                                                  |
| Laninamivir<sup>‡</sup> | 15            | 17.5         | 3.4                               | 210                                                  |

<sup>*IC<sub>50</sub> value: mean nmol/L of duplicate reactions.</sup><br />
<sup>†Oseltamivir carboxylate is the active form of oseltamivir.</sup><br />
<sup>‡Laninamivir is the active form of laninamivir octanoate.</sup><br />
<sup>§A/Anhui/1/2013 (H7N9): NA inhibitor-sensitive virus.</sup><br />
<sup>¶A/Anhui/1/2013-NA-R294K (H7N9): NA inhibitor-resistant virus (14).</sup>
**Technical Appendix Table 4.** Amino acid differences among A/feline/NY/16 virus and human H7N2 isolate (A/New York/108/2016)  

| Virus                  | PA | HA | NA |
|------------------------|----|----|----|
|                        | 57 | 9  | 127| 156| 40 | 362|
| A/feline/NY/16         | Q  | T  | S  | T  | Y  | R  |
| A/New York/108/2016*  | R  | I  | N  | A  | H  | K  |

*The sequences were obtained from GISAID (accession nos. EPI944622–EPI944629). PA, polymerase; HA, hemagglutinin; NA, neuraminidase.*
Technical Appendix Figure 1. Cage settings for virus transmission studies in cats. All cat transmission experiments were conducted at the Charmany Instructional Facility, School of Veterinary Medicine, University of Wisconsin–Madison, under controlled conditions of temperature and humidity. (A and B) Cages and racks used for respiratory droplet transmission studies. Cats were housed individually in regular cat cages. The two racks holding infected and naïve cats were spaced 35 cm apart to prevent direct and indirect contact between animals while allowing respiratory droplet transmission of influenza viruses. (C and D) Cages used for direct contact transmission studies. Large dog transporter cages with a perch/resting platform were used. One infected and one naïve cat were housed together in one cage.
Technical Appendix Figure 2. Images of feline H7N2 virions observed by negative-staining electron microscopy. Virions negatively stained with 2% phosphotungstic acid solution were observed under an electron microscope. (A and B) Higher magnification of virus particles. (C) Lower magnification of virus particles. Scale bar = 100 nm.
Technical Appendix Figure 3. Phylogenetic tree of influenza A viral PB2 segments. The optimal tree with the sum of branch length = 1.59092622 is shown. The analysis involved 48 nt sequences. The final dataset contained a total of 2,260 positions.
Technical Appendix Figure 4. Phylogenetic tree of influenza A viral PB1 segments. The optimal tree with the sum of branch length = 1.3928728 is shown. The analysis involved 47 nt sequences. The final dataset contained a total of 2,263 positions.
Technical Appendix Figure 5. Phylogenetic tree of influenza A viral PA segments. The optimal tree with the sum of branch length = 1.51709379 is shown. The analysis involved 48 nt sequences. The final dataset contained a total of 2,090 positions.
Technical Appendix Figure 6. Phylogenetic tree of influenza A viral NP segments. The optimal tree with the sum of branch length = 1.44906153 is shown. The analysis involved 50 nt sequences. The final dataset contained a total of 1,444 positions.
Technical Appendix Figure 7. Phylogenetic tree of influenza A viral NA segments. The optimal tree with the sum of branch length = 0.72173357 is shown. The analysis involved 31 nt sequences. The final dataset contained a total of 1,343 positions.
Technical Appendix Figure 8. Phylogenetic tree of influenza A viral M segments. The optimal tree with the sum of branch length = 0.72235656 is shown. The analysis involved 41 nt sequences. The final dataset contained a total of 971 positions.
Technical Appendix Figure 9. Phylogenetic tree of influenza A viral NS segments. The optimal tree with the sum of branch length = 1.97636652 is shown. The analysis involved 79 nt sequences. The final dataset contained a total of 811 positions.
Technical Appendix Figure 10. Pathogenicity of A/feline/NY/16 and A/chicken/NY/99 viruses in mice. Bodyweight changes in mice infected with A/feline/NY/16 and A/chicken/NY/99 viruses. Three mice per group were infected intranasally with A/feline/NY/16 and A/chicken/NY/99 virus in amounts of 10–10^6 PFU. Bodyweight and morbidity and mortality were monitored daily for 14 days.
Technical Appendix Figure 11. Virus titers in the organs of infected mice. Six mice per group were infected intranasally with $10^5$ PFU of A/feline/NY/16 and A/chicken/NY/99 viruses. Three mice in each group were euthanized on days 3 and 6 postinfection, and organs including brains, lungs, nasal turbinates, kidneys, livers, and spleens were collected. Viruses were isolated only from the lungs and nasal turbinates of infected animals; therefore, the other organs tested are not shown in the figure.
Technical Appendix Figure 12. Immunohistochemical findings in mice infected with A/feline/NY/16 or A/chicken/NY/99 virus. Shown are representative sections of nasal turbinates and lungs of mice infected with the indicated viruses on days 3 and 6 postinfection. Three mice per group were infected intranasally with $10^6$ PFU of virus, and tissues were collected on days 3 and 6 post-infection. Influenza virus antigens were detected by a mouse monoclonal antibody for NP. For nasal turbinate sections: -, 0 NP-positive cells; +/−, NP-positive cells detected in 1 focal region; +, NP-positive cells detected in $>$3 focal regions. For bronchus and alveolar sections: -, 0 NP-positive cells; +: $>$6 NP-positive cells. NP-positive cells were detected in focal, but not in diffuse bronchial and alveolar sections. For all analyses, the entire sections were evaluated. Left: H&E staining. Right: immunohistochemical staining for NP. Scale bars, 50 μm (nasal turbinates), 100 μm (lung).
Technical Appendix Figure 13. Bodyweight and temperature changes in ferrets infected with $10^6$ PFU of A/feline/NY/16 or A/chicken/NY/99 virus. Bodyweight and temperature were monitored daily for 14 days. A and B) Bodyweight and temperature changes for 3 ferrets per group infected with A/feline/NY/16 virus. C and D) Bodyweight and temperature changes for 3 ferrets per group infected with A/chicken/NY/99 virus.
Technical Appendix Figure 14. Immunohistochemical findings in infected ferrets. Shown are representative sections of nasal turbinates and lungs of ferrets infected with the indicated viruses on days 3 and 6 postinfection. Three ferrets per group were infected intranasally with $10^6$ PFU of virus, and tissues were collected on days 3 and 6 postinfection. Influenza virus nucleoprotein was detected by a rabbit polyclonal antibody to this protein. For nasal turbinate sections: -, 0 NP-positive cells; +/−: NP-positive cells detected in 1 focal region; +, NP-positive cells detected in >3 focal regions. For bronchus and alveolar sections: -, 0 NP-positive cells. For all analyses, the entire sections were evaluated. Left: H&E staining. Right: immunohistochemical staining for NP. Scale bars, 50 µm (nasal turbinates), 100 µm (lung).