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ABSTRACT
Immunoglobulin G3 (IgG3) is the predominant IgG subclass elicited in response to polysaccharide antigens in mice. This specific subclass has been shown to crosslink its fragment crystallizable (Fc) regions following binding to multivalent polysaccharides. Crosslinking leads to increased affinity through avidity, which theoretically should lead to more effective protection against bacteria and yeast displaying capsular polysaccharides on their surface. To investigate this further we have analyzed the binding characteristics of two IgG monoclonal antibody (mAb) subclass families that bind to the capsular polysaccharide (CPS) of Burkholderia pseudomallei. The first subclass family originated from an IgG3 hybridoma cell line (3C5); the second family was generated from an IgG1 cell line (2A5). When the Fc region of the 3C5 IgG3 is removed by proteolytic cleavage, the resulting F(ab’)2 fragments exhibit decreased affinity compared to the full-length mAb. Similarly, when the parent IgG3 mAb is subclass-switched to IgG1, IgG2b, and IgG2a, all of these subclasses exhibit decreased affinity. This decrease in affinity is not seen when the 2A5 IgG1 mAb is switched to an IgG2b or IgG2a, strongly suggesting the drop in affinity is related to the IgG3 Fc region.

KEYWORDS
antibody; binding affinity; capsule; melioidosis; polysaccharide

Introduction
Burkholderia pseudomallei is a soil-dwelling bacillus that causes melioidosis, a severe disease common to Southeast Asia and northern Australia. Melioidosis cases in northeast Thailand are particularly severe; from 1997–2007 the mortality rate was nearly 43%.1,2 In addition, Limmathurotsakul and colleagues estimated there to be 165,000 melioidosis cases globally per year resulting in 89,000 deaths.3 The high mortality rate is in part due to the difficulty in diagnosing melioidosis and the inherent resistance of B. pseudomallei to commonly prescribed antibiotics.3

B. pseudomallei produces a number of virulence factors that enhance pathogenesis, chief among them being the capsular polysaccharide (CPS).2,4 Polysaccharide capsules are found on many pathogenic bacteria and fungi, and contribute to virulence by inhibiting complement activation and preventing phagocytosis.5-8 They are high molecular weight antigens with repeating epitopes that are displayed on bacterial and fungal cell surfaces.9-12 The B. pseudomallei CPS is comprised of an unbranched homopolymer of 1,3-linked 2-O-acetyl-6-deoxy-β-D-mannoheptopyranose residues.13 CPS has been shown to inhibit phagocytosis by reducing the amount of complement factor C3b that is deposited on the bacterial cell surface.4 Mutation of specific genes comprising the B. pseudomallei CPS operon results in the production of mutant strains attenuated for pathogenesis in animal models.14-16

Anti-capsular antibodies are an important mechanism for host defense, thus capsules are appealing vaccine candidates; however, polysaccharide antigens do not illicit a robust humoral immune response by themselves. Normally, humoral immunity is induced in a T-cell dependent manner,17 however, polysaccharides utilize a T-cell independent pathway and stimulate B-cells by crosslinking multiple cell surface antigen receptors. T-cell independent responses produce a short-lived and weak humoral immune response.18 To circumvent this weak response polysaccharides can be conjugated to immunogenic proteins or toxoids.19-21 For example, the Haemophilus influenzae capsule elicits a much stronger immune response when it is conjugated to tetanus toxoid.19

The IgG subclass produced in response to T-cell independent polysaccharide antigens in mice is restricted to
IgG3.22-24 This begs the question of whether this subclass restriction is helpful when the immune system encounters an encapsulated pathogen. To address this question, we produced subclass switch families of polysaccharide-specific mouse monoclonal antibodies and analyzed their immunochromic interactions. These subclass families possess identical variable regions (Fv), but different heavy chain constant regions. Subclass switching of hybridoma clones occurs infrequently in vitro,25,26 although different clones secreting different subclasses can be isolated and expanded, allowing for efficient study of subclass-switch mAb families.

Two subclass-switch mAb families that bind to B. pseudomallei CPS were isolated and used in this study. The first family was derived from an IgG3 hybridoma cell line (3C5) that was isolated following immunization with heat-killed B. pseudomallei.27 The additional IgG 3C5 subclass cell lines were isolated from the parental IgG3 cell line (IgG3→IgG1→IgG2b→IgG2a). Our group previously determined that mAb 3C5 is a relatively high affinity murine IgG3 that provides passive protection in a murine model of pulmonary melioidosis.27 The second family was derived from an IgG1 cell line (2A5) and includes 3 IgG subclasses (IgG1→IgG2b→IgG2a). IgG1 mAb 2A5 has a similar affinity to mAb 3C5, but was generated with a CPS glycoconjugate.

Our study supports previous findings that murine IgG3 Fc regions have the ability to enhance affinity through Fc-Fc interactions when binding to antigens that contain repeating epitopes, such as polysaccharides.28-30 Our experimental design includes binding and affinity studies of 2 subclass switch families of B. pseudomallei capsule-specific IgG mAbs via ELISA, Western blot and surface plasmon resonance (SPR). More specifically, we show that IgG3 Fc regions contribute to antibody binding to B. pseudomallei CPS, resulting in increased affinity.

Materials and methods

Immunization of mice and production of mAbs

Generation of mAb 3C5 IgG3 has been described.27 Briefly, B. pseudomallei strain 1026b was incubated overnight under BSL-3 containment at 37°C in brain heart infusion (BHI) broth. Bacteria were heat-inactivated at 80°C for 2.5 h and confirmed killed by establishing no-growth in BHI broth and back-plating on BHI agar (each for 3 days). BALB/c mice were immunized via intraperitoneal (i.p.) injections with $2 \times 10^8$ heat-inactivated bacteria every 2 weeks for 8 weeks total. An ELISA was used to assess antibody titers to B. pseudomallei. A final boost was administered 3 d prior to splenectomy. Hybridoma cells were produced as previously described.31 Western blot analysis was performed to identify hybridoma cell lines that were producing mAbs reactive with purified CPS.

Purified CPS was conjugated to cationized bovine serum albumin (cBSA; Pierce) as previously described.24 In brief, B. pseudomallei LPS O-antigen mutant strain RR2683 was grown at 37°C in Luria Bertani-Lennox (LBL) broth and the CPS extracted via hot aqueous-pH.nol. CPS and rough LPS were separated on a Sephadex G-50 column and the purified CPS activated with sodium meta-periodate (NaIO$_4$; Pierce). cBSA was added, followed by sodium cyanoborohydride (NaBH$_3$CN). Aliquots were incubated at room temperature for 4 d. Sodium borohydride (NaBH$_4$) was added and the conjugate was lyophilized for later use. This conjugate was used to immunize BALB/c mice and produce mAb 2A5 IgG1. Mice were immunized via i.p. injections with 5 µg of CPS-BSA every 2 weeks, for 6 weeks total. Antibody titers were measured by ELISA (see below) and hybridoma cell lines were generated as previously described.31

Isolation of subclass-specific hybridoma cell lines

A modified protocol based on the method of Spira et al.25 was used to isolate subclass-switch mAb families.26 Switching from one subclass to another follows the germline order of heavy chain exons (IgG3, IgG1, IgG2b, IgG2a). The procedure was done in a sequential manner to obtain hybridoma clones that secrete each subclass. Briefly, the hybridoma cell line secreting the parent mAb (e.g. IgG3 mAb 3C5) was plated at 1000 cells/well in a 96-well tissue culture plate. The supernatant from these wells was added to an ELISA plate that contained goat anti-mouse IgG1 in the solid phase. A horseradish peroxidase (HRP)-labeled goat anti-mouse secondary antibody was used to identify the wells that contained IgG1 antibody. Selected high-positive hybridoma wells were then diluted to 100 cells/well and the ELISA was repeated. The dilutions were continued to 10 followed by 1 cell/well at which time multiple IgG1 clones were isolated. An IgG1 cell line was grown in culture and the cells were plated as previously described at 1000 cells/well to isolate an IgG2b secreting cell line. This protocol was repeated for isolation of each subclass. Cell lines were grown in Integra CL 1000 culture flasks (Integra Biosciences) with RPMI media containing 4.5 g glucose, 4 mM L-glutamine, 50 µM 2-mercaptoethanol, 20 mM HEPES, 1 mM sodium pyruvate, and 15% low IgG fetal bovine serum. Antibodies were purified by affinity chromatography over a protein-A column.
**F(ab′)2 fragments**

Full-length mAb 3C5 IgG3 was digested with pepsin from porcine gastric mucosa (Sigma) to obtain F(ab′)2 fragments. Briefly, mAbs (5 mg/mL) were incubated with shaking at 37°C for 30 min with pepsin at a final concentration of 0.2 μg/mL in 20 mM NaOAc, pH 4.4. Next, 10% (v/v) of Tris (2 M) was used to stop the reaction. F(ab′)2 fragments were purified over a Superose 12 (GE Healthcare) molecular sieve column. Eluted fractions were assessed by non-denaturing sodium dodecyl sulfate polyacrylamide (12%) gel electrophoresis (SDS-PAGE) with Coomassie blue staining. Fractions that contained F(ab′)2 fragments (showing typical reduction in molecular weight vs. full length antibody) were combined.

**Variable region sequencing of mAbs**

Heavy and light chain variable regions were sequenced as previously described. Briefly, total mRNA was isolated from each hybridoma cell line with an RNeasy Mini Kit (QIAGEN). cDNA was synthesized using a First Strand cDNA Synthesis Kit (Thermo Scientific) and amplified with a Mouse Ig-Primer Set (Novagen). PCR products were TA cloned into the pGem-T vector (Promega), and sequenced. Two independent clones were sequenced for each subclass switch cell line.

**Western blot**

Isolated subclass switch mAbs were confirmed to bind to purified CPS via Western blot as previously described. Briefly, 1 μL of a 10x concentrated B. pseudomallei 1026b lysate, 1.1 x 10⁵ inactivated whole cells of B. mal­lei China 7 (BEI Resources), 8 x 10⁶ inactivated whole cells of B. thailandensis E264 (BEI Resources), or 0.5 μg purified CPS (see above) were incubated with 1 volume of proteinase K at 3.3 mg/mL for 1 hr at 60°C. Next, samples were separated by SDS PAGE (BioRad) at 160 V for 1 hr, followed by transfer to a nitrocellulose membrane (BioRad) via a TransBlot Turbo (BioRad). Membranes were blocked in Tris-buffered saline plus Tween 20 (TBST, 50 mM Tris-HCl, pH 7.6; 150 mM NaCl, 0.1% Tween 20) supplemented with, 5% milk overnight at 4°C (blocking solution). Membranes were then probed with mAbs at 0.1 μg/mL diluted in blocking solution for 1 hr while rocking at room temperature. Membranes were washed 3 times for 15 min with TBST followed by incubation for 30 min at room temperature with HRP-conjugated goat anti-mouse kappa chain antibody (Southern Biotech) diluted 1:10,000 in blocking solution. Membranes were washed 3 additional times and binding was detected with SuperSignal West Femto Chemilumi­nescent Substrate (Pierce). Binding was visualized with a Chemidoc imaging system (BioRad). Western blots were also performed to compare the binding activity of each mAb subclass. A. pseudomallei whole cell lysate (87 μg/gel) was added to a 7.5% SDS PAGE gel that contained one regular sized well for the molecular weight marker and one large well (well sides cut out) for the B. pseudomallei whole cell lysate. Electrophoresis and blotting was performed as above. A miniblotter (with separate lane chambers for probing) was used so different concentrations of each subclass mAb could be used to probe the same blot. The nitrocellulose membranes were probed with either 1:100 or 1:1000 (stock solution of 1 mg/ml) dilutions of each subclass mAb as above. Membranes were washed, probed with a secondary antibody and imaged as above.

**Surface plasmon resonance**

Binding affinity was determined by surface plasmon resonance (SPR) with a BIAcore X100 (GE Healthcare). Purified CPS (see above) was benzoquinone-activated and conjugated to biotin as previously described, and immobilized onto a streptavidin (SA) sensor chip at 30 response units (RU). A second flow cell was unmodified and used for reference subtractions. Affinity was evaluated with mAbs diluted in HBS-EP+ running buffer (10 mM HEPES, 150 mM NaCl, 3 mM EDTA, and 0.05% surfactant P20, pH 7.4) at a range of either 0.33–333 nM or 12–3333 nM. Two-fold serial dilutions of mAb were injected over the immobilized CPS at 30 μL/min for 60 s followed by 120 s of passive dissociation. The sensor chip surface was regenerated between each concentration with 10 mM HCl, pH 1.5. Each experiment was performed at least twice, except for 3C5 IgG3 F(ab′)2 and 2A5 IgG2a. In both cases, the availability of purified reagent limited us to one assessment at several (6 or 8) different antibody concentrations. Dissociation constants (Kd) were calculated using BIAevaluation software (GE Healthcare). The apparent kinetic constants were determined using the Bivalent Analyte model in BIAevaluation software. All evaluations passed the statistical internal quality controls of BIAevaluation software.

**ELISA**

Subclass-switch mAbs were evaluated via a direct antigen binding ELISA. Polystyrene plates (Thermo Scientific) were coated for 90 min at 37°C with 0.005% (w/v) Poly-L-Lysine (Sigma) diluted in PBS. Plates were washed with PBS and incubated overnight with 4 μg/mL of purified CPS. Next, plates were washed with PBS+0.05%
Western blot. The IgG3 subclass of mAb 3C5 and the subclass-switch variants of mAb 3C5 were generated with a modified sequential sib selection protocol where individual hybridoma clones were screened by ELISA to identify subclass-switch families of both mAbs were generated with a modiﬁed sequential sib selection protocol where individual hybridoma clones were screened by ELISA to identify subclass-switch clones. To verify that the heavy and light chain variable regions were identical for all subclasses, total mRNA was isolated from each hybridoma cell line and used for cDNA synthesis and PCR ampliﬁcation followed by sequencing. Results indicate that the variable region sequences between each family are identical, and the sequences between each family (3C5 vs. 2A5) are diﬀerent (data not shown).

Western blot analysis was performed to verify that the parent mAb 2A5 IgG1 bound to CPS (Fig. 1). Our previous study conﬁrmed that IgG3 mAb 3C5 is reactive with CPS from B. pseudomallei and B. mallei.27 The CPS is a high molecular weight antigen comprised of an unbranched polymer of -3)-2-O-acetyl-6-deoxy-β-D-manno-heptopyranose-(1- residues that can be visualized by a characteristic high molecular weight smear via Western blot.27 The IgG3 subclass of mAb 3C5 and the IgG1 subclass of mAb 2A5 both bound to the same proteinase K-resistant high molecular weight antigen found in B. pseudomallei, B. mallei and puriﬁed CPS preparations. Although a small percentage of B. thailandensis strains do produce CPS, E264 does not and both mAbs were not reactive to this strain (Fig. 1).34,35

Results

We previously generated IgG3 mAb 3C5, from BALB/c mice immunized with heat-inactivated whole B. pseudomallei.27 IgG1 mAb 2A5 was generated from BALB/c mice immunized with puriﬁed CPS conjugated to BSA;12 this glycoconjugate induces high titers of CPS-speciﬁc IgG antibodies in mice.21 Subclass-switch families of both mAbs were generated with a modiﬁed sequential sib selection protocol where individual hybridoma clones were screened by ELISA to identify subclass-switch clones.25 To verify that the heavy and light chain variable regions were identical for all subclasses, total mRNA was isolated from each hybridoma cell line and used for cDNA synthesis and PCR ampliﬁcation followed by sequencing. Results indicate that the variable region sequences within each family are identical, and the sequences between each family (3C5 vs. 2A5) are diﬀerent (data not shown).

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Binding kinetics of each mAb were measured by SPR to determine if Fc regions from each IgG subclass contribute to mAb afﬁnity. Puriﬁed CPS was conjugated to biotin33 and immobilized on a streptavidin-coated SPR sensor chip. Total immobilization of CPS was 30 response units (RU). Injection of mAbs occurred over a 60 second pulse and a titratable increase in RU was observed (Fig. 2). A steady state model was applied to each graph to determine the dissociation constant for each mAb (Fig. 3). The calculated binding afﬁnity of each mAb is in Table 1. Notably, the subclass-switch variants of mAb 3C5 have a substantially lower afﬁnity for CPS than the parent IgG3 (8 to 20-fold). To determine if this afﬁnity change was in part due to Fc region variation, F (ab’)2 fragments were produced from the 3C5 IgG3. The F(ab’)2 fragment afﬁnity is comparable to the 3C5 non-IgG3 mAbs (29-fold reduction compared to IgG3 mAb 3C5). However, the 2A5 mAb subclasses show minor afﬁnity variation, meaning the overall afﬁnity is similar to that of IgG3 mAb 3C5.

The SPR data indicated that the non-IgG3 3C5 antibodies had lower afﬁnity, while the 2A5 subclass mAbs had comparable afﬁnity. As further support we performed a Western blot experiment to compare binding of the 3C5 and 2A5 subclass switch mAbs. A miniblotter (with separate lane chambers for probing) was used so diﬀerent concentrations of each subclass mAb could be used to probe the same blot containing
Figure 2. Binding specificity of subclass-switch family mAbs and 3C5 IgG3 F(ab')2 fragments given as a function of response units (RU) generated over time. Data are shown from a representative experiment. A BIAcore X100 instrument was used to determine the affinity of each mAb for CPS. CPS purified from Bp RR2683 was conjugated to biotin and immobilized on a SA sensor chip. Binding was analyzed by injecting 8 samples diluted 2-fold (dilutions; 0.33–333 or 12–3,333 nM) for 60 s, followed by 120 s of passive dissociation.
Figure 3. Binding affinity of subclass-switch family mAbs and 3C5 IgG3 F(ab')2 fragments given as a function of RUs generated by concentration. Data are shown from a representative experiment. The steady-state model from BLAevaluation software was applied to each graph in Fig. 2 to determine the dissociation constant ($K_D$) of each mAb. A smaller $K_D$ corresponds to a higher affinity. Note the higher mAb concentrations needed for calculation of affinity for the 3C5 non-IgG3 mAbs.
a *B. pseudomallei* lysate. As shown in Fig. 4A at equal mAb probing concentrations the most CPS reactivity was seen with mAb 3C5 IgG3 with far less intense reactivity for the non-IgG3 subclasses. Subclass-switching mAb 2A5 generated mAbs that bound to CPS at similar signal levels as the parent IgG1 (Fig. 4B). The exposure time for Western blots in panel A and B were different to prove this point.

A ELISA was performed to compare the relative antigen binding characteristics of the 3C5 and 2A5 subclass families (Fig. 5). Purified CPS was incubated in the solid phase at 4 μg/mL and 2-fold serial dilutions of mAbs were added in the fluid phase starting at 2,000 μg/mL. Each line on the plot represents a different mAb subclass. These plots were used to calculate the half-maximal effective concentration value (EC50), which is the antibody

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**Table 1. Summary of antibodies generated for this study.**

| mAb   | IgG Subclass | VH Family | VL Family | $k_a \times 10^{-3}$ (μM$^{-1}$s$^{-1}$)$^a$ | $k_d \times 10^{-3}$ (s$^{-1}$)$^b$ | $K_d$ (nM)$^c$ | EC50 (μg/mL) |
|-------|--------------|-----------|-----------|------------------------------------------|---------------------------------|----------------|---------------|
| 3C5   | IgG3         | Vh6       | IgK V19/28| 280 ± 2.0                                 | 1.1 ± 0.05                      | 73 ± 6.0       | 0.9 ± 0.03     |
|       | IgG1         | Vh6       | IgK V19/28| 16 ± 0.3                                  | 2.4 ± 0.14                      | 1460 ± 81      | 18 ± 1.7       |
|       | IgG2a        | Vh6       | IgK V19/28| 27 ± 0.3                                  | 2.4 ± 0.09                      | 790 ± 120      | 12 ± 0.8       |
|       | IgG2b        | Vh6       | IgK V19/28| 43 ± 0.7                                  | 4.1 ± 0.11                      | 550 ± 120      | 12 ± 1.0       |
|       | F(ab')$_2$   | —         | —         | 38 ± 2.0                                  | 5.9 ± 0.29                      | 2100 ± 610     | 383 ± 130      |
| 2A5   | IgG1         | Vh6       | IgK V21   | 1800 ± 120                                | 24 ± 1.6                        | 43 ± 6.0       | 0.37 ± 0.02    |
|       | IgG2a        | Vh6       | IgK V21   | 1100 ± 50                                 | 12 ± 0.51                       | 36 ± 4.3       | 0.36 ± 0.03    |
|       | IgG2b        | Vh6       | IgK V21   | 720 ± 30                                  | 9.3 ± 0.35                      | 58 ± 8.1       | 0.76 ± 0.08    |

$^a$Association rate constant

$^b$Dissociation rate constant

$^c$Dissociation constant

$^d$IgG3 F(ab')$_2$

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**Figure 4.** Comparison of binding activity within each mAb subclass by Western blot. *B. pseudomallei* Bp82 total cell lysate (87 μg) was separated by SDS-PAGE and transferred onto nitrocellulose membrane. The membranes were probed with mAb 3C5 family (panel A) and 2A5 family (panel B) using a miniblottor. 3C5 IgG3 shows substantially higher reactivity to CPS compared to the other subclasses. The reactivity between different subclasses of mAb 2A5 and CPS was comparable. 2A5 and 3C5 antibody concentration was prepared from 1mg/ml stock. Exposure time for Western blots in panel A and B were different.
concentration that generates a response halfway between the baseline and maximum (Table 1). The calculated EC\textsubscript{50} value for the parent 3C5 IgG3 mAb is 0.9 µg/mL, whereas the calculated EC\textsubscript{50} values for the 3C5 subclass-switch mAbs are higher, by roughly 10 to 20-fold. In addition, F(ab\textsuperscript{′})\textsubscript{2} fragments were generated from 3C5 IgG3 and the EC\textsubscript{50} values of these fragments were substantially higher than IgG3 mAb 3C5. Finally, the EC\textsubscript{50} values of the 2A5 subclass-switch mAbs were comparable. The calculated EC\textsubscript{50} values of the 2A5 subclasses ranged from 0.36–0.76 µg/mL, which is similar to 3C5 IgG3.

Discussion

Many pathogenic microorganisms produce capsular structures, including *Haemophilus influenzae*, \textsuperscript{36} *Neisseria meningitidis*, \textsuperscript{37} *Bacillus anthracis*, and *Cryptococcus neoformans*.\textsuperscript{38} These capsules are comprised of polysaccharides or, in the case of *B. anthracis*, a polypeptide.\textsuperscript{39} Capsules are antiphagocytic and are generally required for virulence.\textsuperscript{40} Antibodies that target capsules are opsonic and protective in many cases.\textsuperscript{40} As such, capsules are ideal vaccine targets, however, most capsular polysaccharides do not elicit a strong immune response, especially in infants and young children.\textsuperscript{41} Capsular polysaccharides are high molecular weight antigens comprised of identical repeating units, making them T-cell independent type 2 antigens (TI-2).\textsuperscript{42} They can activate B-cells by multivalent cross-linking of B-cell receptors, which produces a very specific antibody response.\textsuperscript{18} Mice generally produce IgG3 in response to capsular polysaccharides;\textsuperscript{22-24} humans generally produce IgG2.\textsuperscript{43} As such, it is important to study the immunochemistry between antibodies and microbial capsules to develop effective vaccines and immunotherapeutics that elicit the appropriate humoral response against encapsulated pathogenic microbes.

Subclass-switch families are antibodies with identical Fv regions but different heavy chain constant regions, thereby making them invaluable for determining how the Fc region contributes to affinity and protection.\textsuperscript{26,44,45} Previous research has shown that subclass-switch antibodies bind to their target with variable specificity and affinity, despite having identical Fv regions. Seminal research by Greenspan and colleagues established the enhanced binding of murine IgG3 to polysaccharides over that of non-IgG3 subclass-switch mAbs containing identical variable regions.\textsuperscript{28-30} Specifically, it was shown that a partial murine IgG subclass family (IgG3→IgG1→IgG2b) interacts with the same Group A streptococcal antigen in very different ways. Murine IgG3 bound cooperatively and thus possessed a higher affinity than IgG1 or IgG2b. In addition, IgG3-derived F(ab\textsuperscript{′})\textsubscript{2} fragments did not show cooperative binding and bound with a much lower affinity, similar to that of the IgG1 and IgG2b.\textsuperscript{28} Our recent studies in *Bacillus anthracis* have shown that an IgG3 mAb possessed greater binding affinity to the capsular polypeptide\textsuperscript{26} when compared to all 3 other switched subclasses. The reduced binding affinity of the non-IgG3 subclass mAbs results from alterations in the CH2 and CH3 domains, which comprise the Fc region in IgG molecules. When altering this same murine IgG3 Fc region by engineering human chimeric antibodies (chAbs), the resulting chAbs have significantly reduced binding affinities for the capsular polypeptide.\textsuperscript{46} Together, these studies strongly support that changes in the Fc region can alter antibody-antigen interactions.

There have been multiple studies showing passive transfer of antibodies can provide protection in animal models of melioidosis,\textsuperscript{27,47-52} however, there have been no studies to date comparing different antibody subclasses. In the current study, we generated 2 unique subclass-switch families that bind to the CPS of *B.
**Disclosure of potential conflicts of interest**

No potential conflicts of interest were disclosed.

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