Hepatic differentiation of mouse bone marrow-derived mesenchymal stem cells using a novel 3D culture system

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Abstract. The development of novel culture systems that mimic the in vivo microenvironment may be beneficial for inducing the differentiation of stem cells and promoting liver function. In the present study, spheroid cultures and decellularized liver scaffolds (DLSs) were utilized to obtain differentiated hepatocyte-like cells. Mouse bone marrow (BM)-derived mesenchymal stem cells (MSCs) self-aggregated into spheroids under low-attachment conditions and implanted into the DLSs via a negative pressure suction device. The Albp-ZsGreen adenoviral vector, which is driven by the albumin (ALB) promoter, was utilized for real-time monitoring of the differentiation status of hepatocytes from stem cells. The findings of the present study may be useful for cell transplantation purposes.

Introduction

Liver transplantation is the primary treatment for patients with acute liver failure, end-stage liver disease and inherited liver-based metabolic disorders. However, the demand for suitable organs for transplantation far exceeds the available donor organs (1). Tissue engineering and regenerative medicine-based strategies are a promising alternative to organ transplantation (2-4).

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Tissue-specific cells and scaffolding biomaterials are essential for liver tissue engineering (5). As primary hepatocytes exhibit poor proliferative potential in vitro, it may be more feasible to generate hepatocytes via the differentiation of mesenchymal stem cells (MSCs). MSCs are commonly cultured as 2D monolayers using conventional tissue culture techniques, which may result over time in loss of replicative ability, reduced colony-forming efficiency and poor differentiation capacity (6,7). The microenvironment has a crucial influence on stem cell biology. Therefore, the present study investigated spheroid culture, which has been reported to improve cell-cell contact and interactions of cells with the extracellular matrix (ECM) compared with traditional monolayer methods (8). As cells exist in their native morphology, significant differences in phenotype and responses have been observed between monolayer and spheroid cultures (9,10). Our previous study revealed that 3D spheroid cultures of MSCs enhanced cell yield and maintained stemness, in addition to osteogenic and adipogenic differentiation efficiencies (11).

In the last decade, advances in organ and tissue decellularization have made it possible to obtain tissue-specific ECM from whole organs via the perfusion of the organs with various detergents (12-15). Whole organ decellularization represents a potential strategy for the fabrication of scaffolds for the engineering of tissues and organs, as the decellularized scaffolds maintain their microarchitecture and retain numerous bioactive signals that are difficult to replicate artificially (16). Decellularized liver scaffolds may act as anchors for hepatocyte-like cells derived from stem cells, and aid their attachment, proliferation and organization (17-19). In addition, decellularized liver scaffolds may be an alternative option for heterotopic hepatocyte transplantation (20).

In the present study, spheroid culture and decellularized liver scaffolds (DLSs) were utilized to establish a novel 3D culture system to promote maturation of hepatocyte-like cells from mouse bone marrow (BM)-derived MSCs. The Albp-ZsGreen adenoviral vector, which is driven by the albumin (ALB) promoter, was utilized for real-time monitoring of the differentiation status of hepatocytes from stem cells. The findings of the present study may be useful for cell transplantation purposes.
Materials and methods

Animals. The study was approved by the Ethics Committee of Sichuan University (Chengdu, China). Three livers were isolated from 6-month-old male Bama miniature pigs weighing 10-15 kg for perfusion decellularization. Male C57BL/6 mice (n=3; age, 8 weeks; weight, 20-25 g) were used for hepatocyte isolation. All animals were obtained from the Animal Experiment Center of Sichuan University (Chengdu, China). The mice and Bama miniature pigs were maintained on an alternating 12-h light/dark cycle, fed regular chow, and given water ad libitum.

The surgeries were performed under ketamine (6 mg/kg body weight, administered intramuscular; Kelun, Chengdu, China) and xylazine (10 mg/kg intramuscular; Kelun) anesthesia. Under deep anesthesia, a laparotomy was performed and the liver was exposed. After systemic heparinization through the inferior vena cava, the hepatogastric ligament was carefully dissected. The proximal PV was catheterized. The hepatic artery and common bile duct were ligated and transected. All perihepatic ligaments were severed. Simultaneously, the liver was slowly perfused with 2 l deionized water containing 0.1% EDTA (Kelun) through a cannula in the PV, and the SHIVC was transected, allowing outflow of the perfusate. Following blanching, the liver was stored at -80°C overnight. The Bama miniature pigs were sacrificed during the perfusion process due to an excessive amount of blood loss.

Cultivation of mouse BM-MSCs. Commercial mouse BM-MSCs were purchased from Cyagen Biosciences Inc. (Guangzhou, China). C57BL/6 mouse MSC growth medium (cat. no. MUBMX-90011; Cyagen Biosciences Inc.) was utilized to culture cells and was replaced at least every 2 days. Cells at passage 4-6 were used for subsequent experiments.

Formation of BM-MSCs spheroids. For spheroid cultures, the harvested BM-MSCs were suspended in 10 ml serum-free medium at 1x10^6 cells/ml and cultured in glass spheroid dishes (13x8x4 cm), which were coated with Sigmacote® (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany). Spheroid dishes were incubated with continuous rocking at 0.167 Hz for 24 h to induce spheroid formation. BM-MSC spheroids were stained with 4',6-diamidino-2-phenylindole (DAPI; Santa Cruz Biotechnology, Inc., Dallas, TX, USA). The viability of BM-MSC spheroids was assessed using the FluoroQuench™ fluorescent stain (One Lambda; Thermo Fisher Scientific, Inc., Waltham, MA, USA) according to the manufacturer's protocol. Samples were imaged using a Leica DFC495 fluorescence microscope (Leica Microsystems GmbH, Wetzlar, Germany).

Evaluation of decellularized porcine liver. DLSs were obtained as previously described (21). DLS samples were fixed on day 21 and ALB levels were measured using a Leica DM40000B microscope (Leica Microsystems GmbH). Following DAPI staining, the percentage of ZsGreen-positive cells was determined using ImageJ software version 1.48 (National Institutes of Health, Bethesda, MD, USA). Three samples of both 2D and 3D undifferentiated cells were incubated with adenovirus as controls.

Western blot analysis. Differentiated cells in 2D and 3D culture systems were homogenized to generate protein lysates using radioimmunoprecipitation assay lysis buffer (Beyotime Institute of Biotechnology, Haimen, China) with protease inhibitors (Beyotime Institute of Biotechnology). Equal quantities of protein (80 µg) were separated by 10% SDS-PAGE. Proteins were transferred onto polyvinylidene difluoride membranes (EMD Millipore, Billerica, MA, USA). Membranes were blocked in 5-10% non-fat milk in Tris-buffered saline containing 0.1% Tween-20 for 1 h at room temperature and then incubated overnight with anti-ALB (1:1,000; rabbit monoclonal; cat. no. ab207327; Abcam, Cambridge, UK) and anti-GAPDH (1:1,000) secondary antibodies (both from Abcam) for 2 h at room temperature. Protein bands were visualized using an enhanced chemiluminescence kit (Thermo Scientific, Inc., Waltham, MA, USA).

ALB and urea production. Conditioned media from the differentiated BM-MSCs of 2D and 3D culture systems was collected on day 21 and ALB levels were measured using a mouse ALB ELISA kit (cat. no. E90-134; Bethyl Laboratories, Inc., Montgomery, TX, USA) according to the manufacturer's
protocol. A total of 2 mM heavy, diazonium-enriched ammonium chloride (Cambridge Isotope Laboratories, Inc., Tewksbury, MA, USA) was added to the medium to determine the metabolic ability of differentiated cells. The total urea concentration and proportion of diazonium-enriched urea and natural urea in the medium were measured to determine the source of urea synthesis. Supernatants were quantified by capillary gas chromatography and mass spectrometry, as previously reported (25). Freshly isolated mouse primary hepatocytes were included as a control. Hepatocytes were isolated using a two-step perfusion method as previously described (25).

**Immunofluorescence analysis.** Differentiated cells from 2D and 3D groups were fixed in 4% formaldehyde in PBS and permeabilized with 0.1% Triton X‑100 for 15 min at room temperature. Following permeabilization, samples were blocked with 2% bovine serum albumin (Sigma‑Aldrich; Merck KGaA) in PBS (blocking buffer) for 1 h and subsequently treated with primary antibodies diluted in blocking buffer overnight at 4˚C. The antibodies utilized were sheep anti‑ALB (1:1,000; cat. no. ab8940; Abcam), rabbit anti‑α-fetoprotein (1:1,000; AFP; cat. no. AF5134; Affinity Biosciences, Cell Signal Transduction, Cambridge, UK) and rabbit anti‑cytokeratin‑19 (CK19; 1:1,000; cat. no. AF0192; Affinity Biosciences, Cell Signal Transduction). Alexa Fluor 488-conjugated rabbit anti-sheep (1:400; cat. no. ab150181; Abcam) and goat anti-rabbit (1:400; cat. no. ab150077; Abcam) secondary antibodies were incubated with samples at room temperature for 1 h in the dark. Following nuclear staining with DAPI, slides were mounted and observed under a Leica DMI6000 fluorescence microscope (Leica Microsystems GmbH).

**Gene array analysis.** Sample labeling and array hybridization were performed with Whole Mouse Genome Oligo Microarray (cat.no.G4122F; 4x44K; Agilent Technologies, Inc., Santa Clara, CA, USA). Briefly, total RNA was extracted from BM‑MSCs, primary mouse hepatocytes and 3D hepatocyte‑like cells using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer’s protocol, and used for synthesis of cRNAs, which were labeled with Cyanine 3‑UTP. The concentration of cRNA was measured using a NanoDrop ND‑1000 (Thermo Fisher Scientific, Inc., Wilmington, DE, USA). The hybridized arrays were performed and scanned using the Agilent DNA Microarray Scanner (Agilent Technologies, Inc.). Data were normalized and analyzed using the TIGR MultiExperiment Viewer version 4.8.1 (Institute of Genomic Research, Rockville, MD, USA). Gene ontology (GO) analysis was performed using the Database for Annotation, Visualization and Integrated Discovery (DAVID; https://david.ncifcrf.gov) to determine the functions of the predicted target genes and to uncover the miRNA-target gene regulatory network based on the predicted biological processes and molecular functions; P<0.01 was used as the threshold.

**Statistical analysis.** Data are expressed as the mean ± standard error of three independent experiments. Data were analyzed using SPSS software version 17.0 (SPSS, Inc., Chicago, IL, USA). A one-way analysis of variance followed by the Dunnett’s post hoc test was performed to compare groups. P<0.05 was considered to indicate a statistically significant difference.

**Results**

**Characterization of BM‑MSC spheroids.** To generate BM‑MSC spheroids, BM‑MSCs were harvested at passage 4-6 (Fig. 1A). Following optimization of cell number and growth conditions, spheroid formation was observed under a scanning electron microscope within 24 h (Fig. 1B). Based on DAPI staining, the cells within the spheroids were in close proximity (Fig. 1C). The FluoroQuench™ fluorescent staining assay
revealed that the viability of BM-MSCs in spheroids remained >95% in 3D culture (Fig. 1D-F).

**Characterization of DLSs.** Whole-organ decellularization was achieved by portal perfusion using sodium dodecyl sulfate and Triton X-100. Following decellularization, the porcine liver parenchyma became semi-transparent (Fig. 2A). Hematoxylin and eosin staining revealed no visible cell nuclei and cellular material in the decellularized liver scaffolds (Fig. 2B). Decellularized tissue sections were observed under a scanning electron microscope (Fig. 2C).
Identification of hepatocyte-like cells. The hepatocyte-like cells were generated from mouse BM-MSCs as shown in Fig. 3A. The expression of the Albp-ZsGreen adenovirus was not observed in non-hepatic cells (Fig. 3B); however, ALB synthesis was observed in mature hepatocytes, which suggested that hepatic differentiation had occurred in 2D and 3D cells (Fig. 3C). A greater percentage of ZsGreen-positive cells was observed in the 3D group, which suggested that the 3D culture system provided an improved external microenvironment for differentiation (Fig. 3D). Similarly, the expression levels of ALB in 3D cells were greater compared with the 2D cells following induction (Fig. 3E). In addition, immunocytochemistry demonstrated that 3D cells expressed hepatocyte-like cell markers, including ALB, AFP and CK19, to a greater extent than 2D cells (Fig. 4).

Metabolic activity of differentiated cells in 2D or 3D culture. Compared with the 2D group, cumulative ALB secretion by differentiated cells in the 3D group was significantly greater compared with the 2D group (P<0.05; Fig. 5A). Clearance of heavy urea, which contained ammonium chloride, was greater in the 3D group compared with the 2D group (P<0.05; Fig. 5B). The proportion of heavy urea to total urea produced by the 2D group, 3D group and primary hepatocytes fluctuated from 9.6 to 11.8%, 19.8 to 22.1% and 39.8 to 42.1%, respectively (Fig. 5B).

Gene expression in 3D hepatocyte-like cells. The results of the present study demonstrated the advantage of 3D culture in promoting hepatic differentiation of mouse BM-MSCs.
The cDNA microarray showed that differentiated 3D hepatocyte-like cells resembled mouse primary hepatocytes more than mouse BM-MSCs (Fig. 6A). GO enrichment analysis revealed that signaling pathways that associated with liver function were significantly upregulated, including those involved in fat metabolism, amino acid metabolism and drug metabolism (Fig. 6C), whereas signaling pathways associated with the cell cycle were significant downregulated in the 3D group compared with BM-MSCs (Fig. 6B).

**Discussion**

A limitation of traditional methods for the induction of differentiation of adherent monolayer hepatocyte-like cells is low differentiation efficiency (26). In the present study, a novel 3D culture system was established to obtain differentiated hepatocyte cells from spheroid cultures and DLSs.

Using a simplified portal vein perfusion procedure, porcine liver was effectively decellularized and examined under a scanning electron microscope. Subsequently, the 3D model was utilized. In our previous study, spheroids were pipetted onto the center of the DLS to allow spontaneous attachment. However, certain spheroids detached from the DLS in the early stages of culture (8,22). In the present study, a negative pressure suction device was utilized to ensure the efficient attachment of spheroids to the DLS.

A simple, efficient tool for real-time monitoring of hepatocyte differentiation from stem cells is useful. As the adenoviral vector is commonly utilized for gene expression studies due to its efficient transduction, an Albp-ZsGreen adenoviral vector was constructed to detect hepatocyte-like cells. Following the second induction period, green fluorescence was detected at an earlier stage in 3D cells compared with 2D cells and the fluorescence intensity in 3D cells was significantly greater in a time-dependent manner (data not shown).

Ammonia is a product of protein metabolism and requires clearance by the liver. A functional urea cycle is an important characteristic of mature hepatocytes. To assess urea production and the detoxification of ammonia via the urea cycle, heavy ammonium chloride was added to the culture medium. The concentration of urea produced by differentiated cells in the 3D group was greater compared with the 2D group. In addition, the results of the present study demonstrated that ~20% of urea was produced by urea cycle activity in 3D cells, which was significantly greater compared with 2D cells.
The liver is the largest internal organ providing essential metabolic, exocrine and endocrine functions. These functions include production of bile, metabolism of dietary compounds, detoxification, regulation of glucose levels via glycogen storage and control of blood homeostasis by secretion of clotting factors and serum proteins, including ALB. In the present study, GO analysis revealed that the majority of upregulated genes were associated with liver function, whereas cell cycle-associated pathways were significantly downregulated.

In conclusion, the results of the present study suggested that a 3D culture system may promote hepatic differentiation of mouse MSCs, to generate high yields of mature hepatocytes. This miniaturized culture system may possess unique advantages over previous methods and may provide a potential strategy for cell transplantation and drug research.

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