RESEARCH ARTICLE

Inhibition of *Aspergillus fumigatus* and Its Biofilm by *Pseudomonas aeruginosa* Is Dependent on the Source, Phenotype and Growth Conditions of the Bacterium

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Abstract

*Aspergillus fumigatus* (*Af*) and *Pseudomonas aeruginosa* (*Pa*) are leading fungal and bacterial pathogens, respectively, in many clinical situations. Relevant to this, their interface and co-existence has been studied. In some experiments *in vitro*, *Pa* products have been defined that are inhibitory to *Af*. In some clinical situations, both can be biofilm producers, and biofilm could alter their physiology and affect their interaction. That may be most relevant to airways in cystic fibrosis (CF), where both are often prominent residents.

We have studied clinical *Pa* isolates from several sources for their effects on *Af*, including testing involving their biofilms. We show that the described inhibition of *Af* is related to the source and phenotype of the *Pa* isolate. *Pa* cells inhibited the growth and formation of *Af* biofilm from conidia, with CF isolates more inhibitory than non-CF isolates, and non-mucoid CF > non-mucoid. Preformed *Af* biofilm was more resistant to *Pa*, and inhibition that occurred could be reproduced with filtrates. Inhibition of *Af* biofilm appears also dependent on bacterial growth conditions; filtrates from *Pa* grown as biofilm were more inhibitory than from *Pa* grown planktonically. The differences in *Pa* shown from these different sources are consistent with the extensive evolutionary *Pa* changes that have been described in association with chronic residence in CF airways, and may reflect adaptive changes to life in a polymicrobial environment.
Introduction

Cystic fibrosis (CF) is the result of mutations in the CF transmembrane conductance regulator affecting epithelial chloride and bicarbonate transport. One result is development of thick respiratory secretions, which results in airway obstruction and recurrent episodes of lung inflammation and infection, leading to acute and chronic deterioration of lung function and a shortened lifespan [1]. The affected persons have defective mucociliary clearance and production of thick sticky mucus in which various pathogens can become entrapped. This is a suitable environment for microbial growth and colonization, and these organisms or their soluble metabolites contribute to airway inflammation and subsequent damage.

The most common bacterium and fungus infecting these airways are *Pseudomonas aeruginosa* (*Pa*) [2] and *Aspergillus fumigatus* (*Af*) [3–7], respectively, particularly in the chronically infected older patients. *Pa* evolves in CF airways, producing variants, such as those resulting in mucoid colony types, which are adapted to chronic residence there [2,8,9]. *Af* is ubiquitous in ambient air and the environment, and thus can be inhaled and subsequently establish residency. Both organisms are proficient adapters to environmental stress and relatively resistant to current antimicrobials. They are suspected as important agents in promoting mucus plug formation in the airways, and both are known to form biofilms in vitro and in vivo [2,9–20]. Microbes in biofilms have altered metabolism compared to the same organisms growing planktonically, and biofilms provide microbes with protection from host defenses as well as tolerance to some antimicrobial drugs [21]. The attribution of a role for these microbes in mucus plugging and biofilms stems from the known extracellular production of glycan polymers by *Af* [22] and alginate by *Pa* [2,9,17]. In addition to infection, *Af* can cause allergic bronchopulmonary aspergillosis in up to 15% of CF patients, a complication that causes repeated acute exacerbations, institution of immunosuppressive therapy, and accelerated decline in lung function [23]. As is the case with *Pa*, *Af* also produces secondary metabolites, in the environment as well as *in vivo*, which are known tissue toxins and also have immunosuppressive actions on the host response [24]. Infection with *Pa* or *Af* has been associated with a more rapid decline in CF pulmonary function [17,25–33], with the co-infected patients having the worst prognosis [32,34]. Both pathogens are also important because either can be an opportunist, causing invasive disease [35–39] or other complications [40,41], in lung transplantation, a therapeutic modality offered in debilitating CF.

It is therefore important to study the interactions between these two pathogens. *Pa*-secreted molecules have been well studied for their antifungal activities, a property that has been demonstrated in vitro [42–47]. These inhibitory molecules include homoserine lactones, pyocyanin and other phenazine derivates, pyrrolnitrin and fluorescent green pigments. However, these studies were performed with one or few *Pa* isolates, none representatives of variants that establish chronic residency in CF airways. Moreover, *Pa*-associated factors involved in this interkingdom inhibition continue to be elucidated [48]. The aim of this study was to evaluate the effect of different clinical *Pa* phenotypic variants obtained from CF and non-CF patients on *Af* biofilm formation and preformed *Af* biofilm.

Materials and Methods

Isolates

Any CF isolates from patient respiratory cultures were obtained after written informed consent, for biobanking of the patients’ specimens and subsequent use, approved by the Stanford Institutional Review Board. Other isolates were obtained following clinically indicated cultures. Twenty-six clinical isolates of *Pa* recovered from non-CF patients (n = 16 isolates), or CF
patient sputum (n = 10), from Stanford University Hospital and clinics were evaluated. Among the CF isolates, five were mucoid colony phenotype variants [2,8,9] and five were non-mucoid colony phenotype variants. A list of all isolates studied, and their classification, is given in Tables 1 and 2. We were able to include a mucoid and a non-mucoid Pa isolate obtained from the same CF patient the same day, 2 non-mucoid Pa isolates from another CF patient 6 mos. apart, and 2 Pa isolates with different colonial morphologies from each of 2 non-CF patients obtained the same day, plus another Pa isolate from one of these patients one month later. Af isolate 10AF, a virulent non-CF patient isolate [49,50], was used as the reference Af isolate throughout this study. Nine sputum Af isolates, also identified by molecular methods to be Af sensu stricto [51], were obtained from non-CF patients in a previous study [51] and additionally studied.

Af conidia were obtained as follows: Af was taken from stock suspensions stored at -80°C and then grown for 4 days on Sabouraud Dextrose Agar (Becton Dickinson and Co., Sparks, MD) at 37°C. Conidia were harvested by gently washing with 0.05% Tween-80 (J.T. Baker Chemical Co., Phillipsburg, NJ) in 0.9% saline (Baxter Healthcare Corp., Deerfield, IL). Pa stocks were maintained at -80°C in Microbank microbial storage vials (Pro-Lab Diagnostics, Richmond Hill, Ontario, Canada). Each frozen Pa stock culture was initially inoculated onto Trypticase Soy + 5% sheep blood agar plates (TSA; BBL, Becton Dickinson; subsequent studies indicated the absence of blood in the agar plates did not affect the activity of the Pa harvested) and incubated overnight at 37°C. Pa colonies were then picked, 1–2 loopfuls of the bacteria diluted in RPMI-1640 medium, and the suspension adjusted in the spectrophotometer at A610 to an absorbance of 0.35–0.40 with fresh RPMI-1640 medium. This corresponded to 10^9/ml, with variation over a 3-fold range. A 1:30 dilution was then made in studies of direct Pa action on Af biofilm, or production of Pa culture supernatants, to be described. We found, in the studies to be described, that neither direct Pa action, nor Pa supernatant action, on Af biofilm was particularly sensitive to the size of the initial Pa inoculum in those studies, as variations of at least 10-fold in live Pa cells, and at least 33-fold in the inoculum used to make planktonic Pa supernatants, resulted in no significant differences in activity.

**Inhibition of planktonic Af growth**

Inhibition of growth was assessed by using a conidial inoculum, following guidelines for susceptibility testing of filamentous organisms [52].

**In vitro model of biofilm development**

Biofilms were formed by using a modified in vitro model described previously [53]. To form Af biofilm, sterile polystyrene disks (Biosurface Technologies, Bozeman, MT) were placed in 12-well tissue culture plates (Corning Inc., Corning, NY). Each well contained 3 ml of fresh RPMI-1640 medium (Lonza, Walkersville, MD) with 10^5 Af conidia/ml. Disks were incubated at 37°C for 16 h with shaking at 70 rpm, to allow the fungal cells to attach. Following the attachment phase, disks were gently rinsed in sterile saline (Baxter Healthcare Corp.), transferred to new plates containing fresh RPMI-1640 medium, and incubated for an additional 24 h at 37°C with shaking at 100 rpm (Fig 1I). Biofilm formation here, and in all experimental conditions to be described, was verified by inspection or with a light microscope [16].

**Direct interaction of live Pa on Af biofilm formation**

Suspensions of fungal conidia (3 ml) and bacterial cells (0.1 ml), prepared as described above, were combined in 12-well tissue culture plates in fresh RPMI-1640 medium and incubated at...
Table 1. Clinical isolates.

| Sample identification ID # | CIMR # | P. aeruginosa (Pa) phenotype | Clinical Laboratory ID # Valley Medical Center (VMC) or Stanford University (SU) | Specimen | Date of the positive Pa culture |
|---------------------------|--------|-----------------------------|---------------------------------------------------------------------------------|----------|-------------------------------|
| 1                         | 14–75  | Pa Non CF                   | VMC                                                                              | Respiratory | 1/28/2013                     |
| 2                         | 14–76  | Pa Non CF                   | VMC                                                                              | Respiratory | 2/19/2013                     |
| 3                         | 14–77  | Pa Non CF                   | VMC                                                                              | Respiratory | 2/19/2013                     |
| 4                         | 14–78  | Pa Non CF                   | SU 60370871                                                                     | Respiratory | 2/17/2013                     |
| 9                         | 14–83  | Pa Non CF                   | VMC                                                                              | Respiratory | 3/17/2013                     |
| 12                        | 14–86  | Pa Non CF                   | SU 42266353                                                                     | Respiratory | 4/22/2013                     |
| 13                        | 14–87  | Pa Non CF (Strain #1)       | SU 21548242                                                                     | Respiratory | 8/10/2013                     |
| 14                        | 14–88  | Pa Non CF (Strain #2)       | SU 21548292                                                                     | Respiratory | 8/10/2013                     |
| 16                        | 14–90  | Pa Non CF (Strain #1)       | SU 27917939                                                                     | Respiratory | 9/24/2013                     |
| 17                        | 14–91  | Pa Non CF (Strain #2)       | SU 27917939                                                                     | Respiratory | 9/24/2013                     |
| 19                        | 14–93  | Pa Non CF                   | SU 21548292                                                                     | Respiratory | 9/26/2013                     |
| 20                        | 14–94  | Pa Non CF                   | SU 10790053                                                                     | Non respiratory | 8/14/2013                     |
| 24                        | 14–98  | Pa Non CF                   | SU 41082579                                                                     | Non respiratory | 10/02/2013                    |
| 25                        | 14–99  | Pa Non CF (strain #2)       | SU 22765499                                                                     | Non respiratory | 9/25/2013                     |
| 26                        | 14–100 | Pa Non CF                   | SU 26308684                                                                     | Respiratory | 9/26/2013                     |
| 27                        | 14–101 | Pa Non CF                   | SU 28674323                                                                     | Non respiratory | 9/23/2013                     |
| 5                         | 14–79  | Pa CF Mucoid                | SU 20060455                                                                     | Respiratory | 3/1/2013                      |
| 11                        | 14–85  | Pa CF Mucoid                | SU 60821238                                                                     | Respiratory | 2/21/2013                     |
| 18                        | 14–92  | Pa CF Mucoid                | SU 09710807                                                                     | Respiratory | 7/19/2013                     |
| 21                        | 14–95  | Pa CF Mucoid                | SU 7841943                                                                      | Respiratory | 9/17/2013                     |
| 22                        | 14–96  | Pa CF Mucoid                | SU 41053570                                                                     | Respiratory | 8/10/2013                     |
| 7                         | 14–81  | Pa CF Non Muc               | SU 09710807                                                                     | Respiratory | 1/19/2013                     |
| 8                         | 14–82  | Pa CF Non Muc               | SU 16242976                                                                     | Respiratory | 3/13/2013                     |
| 10                        | 14–84  | Pa CF Non Muc               | SU 7841943                                                                      | Respiratory | 2/01/2013                     |
| 15                        | 14–89  | Pa CF Non Muc               | SU 41053570                                                                     | Respiratory | 8/10/2013                     |
| 23                        | 14–97  | Pa CF Non Muc               | SU 16242976                                                                     | Respiratory | 9/6/2013                      |

Table 2. Aspergillus fumigatus clinical isolates.

| Sample identification ID # | CIMR # | A. fumigatus phenotype |
|---------------------------|--------|-----------------------|
| 1                         | 13–55  | CF                    |
| 2                         | 13–56  | CF                    |
| 3                         | 13–57  | CF                    |
| 4                         | 13–58  | CF                    |
| 5                         | 13–59  | CF                    |
| 6                         | 13–60  | CF                    |
| 7                         | 13–61  | CF                    |
| 8                         | 13–62  | CF                    |
| 9                         | 13–63  | CF                    |
| 10                        | 12–26  | Non CF                |
| 11                        | 12–30  | Non CF                |
| 12                        | 12–48  | Non CF                |

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37°C for 16 h with shaking at 70 rpm. Thus, for the 3 x 10^5 Af/well always involved in biofilm formation, the ratio of Pa to Af was 3.3 x 10^7:3 x 10^5/well. Disks were then rinsed gently with sterile saline, transferred to new plates containing fresh RPMI-1640 medium, and incubated for an additional 24 h at 37°C with shaking at 70 rpm (Fig 1II).
Direct interaction of live Pa on preformed (i.e., established) Af biofilm

Fungal biofilms were formed as described above. After 16 h the disks containing the fungal preformed biofilms were rinsed gently in sterile saline, transferred to new plates containing Pa suspension (3 ml containing 3.3 x 10^7 cells) prepared as described above, and incubated for an additional 24 h at 37°C with shaking at 70 rpm (Fig 1II and 1IV).

Pa planktonic supernatant assay

A Pa suspension was prepared as described above in fresh RPMI-1640 medium. To obtain the planktonic culture filtrates, the bacterial suspension in fresh RPMI described above was incubated in 50 ml conical tubes (Falcon, Brookings, So. Dakota) for 24 h at 37°C with shaking at 70 rpm. The spent medium was removed, filtered to remove any suspended cells or debris. The planktonic supernatant was gently removed, filter sterilized (0.22 μm) (Fisherbrand, Pittsburgh, PA) and used. Filtered supernatant (1.5 ml) was added to selected wells of a 12-well tissue culture plate previously inoculated with 1.5 ml of the standardized Af suspension in fresh RPMI-1640. Fungal biofilms attached and formed as described above. Disks containing biofilms were washed 3 times with sterile saline and transferred to a new plate containing fresh RPMI-1640, and incubated for an additional 24 h at 37°C with shaking at 70 rpm (Fig 1III). Although "supernatants" and "filtrates" are used interchangeably in this paper, when any supernatants were tested they were always first filtered as described above; unfiltered supernatants were never used.

For the preformed biofilm assay, 1.5 ml of the filtered bacterial supernatant was added to selected wells of a 12-well tissue culture plate with 1.5 ml of fresh RPMI-1640. Af biofilms were prepared as described above. Disks containing these fungal preformed biofilms were washed 3 times with sterile saline, transferred to the plate containing the fresh media + the bacterial filtrates, and incubated for an additional 24 h at 37°C with shaking at 70 rpm. Af wells without bacterial supernatant were also included to serve as controls (Fig 1V).

To assay growth during Pa planktonic culture, the Pa planktonic suspension was prepared as described above, adjusted to 10^6/ml, incubated as described above, and at the end of the 24 h, serial dilutions of the supernatant were inoculated onto TSA plates, incubated for 24 h at 37°C, and CFU enumerated.

Pa biofilm filtrate assay

To obtain the Pa biofilm filtrates, a 25 ml suspension of Pa prepared as described above, adhered to 50 ml canted neck tissue culture flasks (BD Biosciences, San Diego) for 2 h at 37°C on a 100 rpm shaker incubator (attachment phase). The liquid was then removed and the flasks were rinsed gently 3 times with sterile saline. Twenty ml of fresh RPMI-1640 was added to the flask and adhered cells formed Pa biofilms for 22 h (total of 24 h), observed by the presence of a layer of bacterial growth on the inner surface of the flask. The spent medium was then removed and processed as described above. Af conidia forming biofilms or preformed Af biofilms were challenged with the Pa biofilm culture filtrate, as described above. Bacterial supernatant-free wells were also included as controls (Fig 1III and 1V). Either Pa planktonic or biofilm supernatants were used when prepared, although studies indicated no change in activity if stored refrigerated for at least one week.

Studies with serum

For sets of experiments with Af and Pa cultures, to be described, fetal bovine serum (Gibco, Grand Island, NY) was added to the RPMI-1640 medium at each step to produce a 10% concentration.
XTT assay

Either when conidia formed biofilm, or when preformed Af biofilm was studied, at the end of the challenge period with either live Pa cells or filtered Pa cultured supernatants (or control media), the Af biofilm was studied with the XTT assay. The tetrazolium salt, XTT (2,3-bis[2-methoxy-4-nitro-5-sulfophenyl]-2H-tetrazolium-5-carboxanilide inner salt) (Sigma, St. Louis, MO) was used to measure the metabolic activity of Af. XTT is reduced by a mitochondrial dehydrogenase to a water-soluble formazan product. Af biofilm discs were rinsed 3 times in sterile saline and transferred to fresh wells containing 3 ml sterile phosphate-buffered saline (PBS) (pH 7.3–7.5, Lonza). Menadione (Sigma) 0.85 gm was added to 5 ml acetone, and mixed in a 1:11 ratio of menadione to XTT (1 mg/ml). The XTT-menadione solution (180 μl) was added to each well and the plates were incubated in the dark for 2 h at 37°C. Following incubation, the contents of the wells were collected and centrifuged for 10 min at 13,300 x g. The pellet was then discarded and the absorbance at 490 nm of the supernatant was determined with a spectrophotometer (Genesys 20, Thermo Scientific, Waltham, MA). Although XTT is a measure of metabolic activity of cells, previous studies of Af have indicated XTT results are linear with mass, and equated XTT result with dry weight [54–56].

For the assay of live Pa cells co-incubated with Af, we determined that the XTT readings only reflected the Af by studying Pa similarly cultured without Af, showing there was no increase of XTT reduction compared to background without any microbes present.

Confocal laser scanning microscopy (CLSM)

Af biofilms were formed on disks as described above. The biofilms were challenged with Pa supernatants obtained from a representative of each of the 3 Pa groups. After incubation at 37°C, the disks were washed three times in sterile PBS and stained using a fluorescent stain (FUN-1; Invitrogen Molecular Probes, Eugene, OR), prepared according to the manufacturer’s instructions. FUN-1 (1 μl) from a 10 mM stock was mixed in 1 ml of PBS. Staining was performed as previously described [11,13]. Three drops of the mixture were added on the top of the biofilm, which was then mounted on a glass slide and covered with a glass coverslip (22 × 22 mm). The disks were incubated for 45 min at 37°C in the dark. The FUN1 visualized the morphology of Af biofilm, a bright green cytoplasmic stain produced after passive diffusion. An additional analysis was made to visualize the effect of bacterial supernatant on the fungal biofilm metabolic activity. A red fluorescence, visualized from intravacuolar staining, was noted, which requires both plasma membrane integrity and metabolic capability (data not shown). Dead cells would have fluoresced bright yellow-green, with no red structures, and were not apparent in our images.

Sections on the xy plane were taken at 1 μm intervals along the z-axis to determine the depth of the biofilms. Microscopic visualization and image acquisition of biofilms were conducted at the Stanford Biofilm Research Center using an upright Leica TCDSP2 scanning confocal laser microscope (Leica Lasertechnik GmbH, Heidelberg, Germany) equipped with an argon/krypton laser and detectors, and filter sets for monitoring of green (excitation 480nm, emission 517nm) and red (excitation 633nm, emission 676). Images were obtained using a 63 x 1.4 Plan-APChromat DIC (Leica, Heidelberg, Germany) objectives. Multichannel simulated fluorescence projection (SFP, a shadow projection) images and vertical cross sections through the biofilm were generated using the IMARIS software package (Bitplane AG, Zürich, Switzerland). Images were processed for display by using Photoshop software (Adobe, Mt. View, CA). Representative images were taken.
Dose-response assay of filtrates

A dose response study was performed using one isolate from each of the 3 Pa groups. Pa planktonic and biofilm culture filtrates were obtained as described above. The culture filtrates or sterile distilled water were diluted in fresh RPMI supplemented with 10% FBS, so that the percent of fresh medium in each test situation was decreased as the spent medium or sterile distilled water increased. Af biofilms were formed in this mixed media, as described above, or planktonic Af growth was studied [52].

Effect of temperature on filtrates

The Pa planktonic and biofilm culture filtrates were prepared as described above. In brief, the final cell-free supernatants were (a) heat treated at 56°C for 30 min and immediately used or (b) frozen at -80°C, stored for 6 days, and thawed. These filtrates were then tested compared to fresh filtrates against either Af conidia forming biofilm, or preformed biofilm, as described above.

Effect of DNase I and Proteinase K on Pa supernatant activity

To determine whether Pa or phage DNA or a Pa protein(s) were involved in the inhibition of Af, we studied the effect of pretreatment of Pa culture filtrates with DNase I (Life Technologies, Grand Island, NY) or Proteinase K (Sigma) and measured the filtrate inhibitory activity on Af biofilm formation (Fig 1I). Pa biofilm culture filtrates were prepared as described using a CF non-mucoid Pa isolate, Af biofilm metabolic activity was measured using the XTT assay.

Pa culture filtrates were treated with DNase I or Proteinase K for 2 hours at 37°C. Ethylenediaminetetraacetic acid disodium salt dehydrate (EDTA) (Sigma) was used to inactivate the DNase I activity. Experiments with DNase I treatment involved growing the Af for 16 hours in the presence of (a) Pa filtrate, (b) DNase I (final concentration 2 U/ml) + Pa filtrate, (c) DNase I + EDTA (final concentration 20mM) + Pa filtrate, (d) EDTA + Pa filtrate, or (e) EDTA alone or (f) DNase + EDTA (in fresh RPMI medium). These were followed by 24 h growth in fresh RPMI-1640 alone.

For Proteinase K pretreatment, conidia were incubated with (a) Pa filtrate, (b) Proteinase K (final concentration 50 μg/ml) + Pa filtrate, or (c) medium treated with Proteinase K.

Materials for other characterization studies

A 30,000 MW cutoff filter was obtained from Millipore Ltd., Tullagreen, Ireland.

Elastolytic production of Pa isolates was assessed as previously described [57].

FeCl₃ was obtained from Sigma.

Statistical analyses

Where n is not stated, each experiment was performed at least two times, with triplicate wells each time. Data from experiments using XTT as the assay parameter were collected in blocks. Each block was comprised of n = 6 values derived from of two sets of triplicate values collected on different days. For the data presented in Figs 2 and 3 statistical analyses were performed as follows. Each condition had five blocks of data, which were comprised of controls, nonCF Pa, mucoid Pa and nonmucoid Pa (Pa was either live cells or spent culture filtrates as indicated). Each block of four conditions was analyzed separately using one-way analysis of variance (ANOVA) followed by a Tukey’s post-hoc test to adjust for the multiple comparisons being performed. Prior to pooling, the data in all blocks were verified to be normally distributed and to have homogenous variances. An initial ANOVA comparison of the five blocks for each
Fig 2. Pa inhibition of Af during biofilm formation. The data presented show the activity of each live Pa isolate or planktonic- or biofilm-generated Pa culture filtrate on Af during formation of biofilm by Af. Individual isolate results are shown in comparison with their respective Af controls (n = 6 for each). There were five non-CF, five mucoid CF and five non-mucoid CF isolates tested; isolate designations are shown on the x-axis. The bars on the far right of each panel represent the pooled data for each type of isolate (n = 30). Bars represent the mean ± SD of the XTT reduction read at 490 nm. (A) Af conidia were exposed to live Pa cells for 16 h. (B) Af conidia were exposed to Pa planktonic supernatant for 16 h. (C) Af conidia were exposed to Pa biofilm supernatant for 16 h. The resulting readings were determined. “XTT metabolic activity” refers to the spectrophotometric absorbance of the formazan reduction product of XTT at 490 nm. Assays were performed in blocks with two runs of triplicates done on different days, for each Pa-Af combination. Panel
condition (e.g., controls) showed the blocks for that condition (e.g., controls) not be significantly different. These results allowed us to pool the data for each condition with a result of n = 30 each. The pooled data were analyzed using one-way ANOVA with a Tukey’s post-test. For comparisons across live cell versus planktonic or biofilm supernatants, control values were first compared and determined to be no different, which was followed by ANOVA and Tukey’s comparisons of the experimental sets. All statistical analyses were done using GraphPad Prism (GraphPad Software, Inc. La Jolla, CA). Statistical significance was considered P<0.05.

Results
A summary of all the studies to now be described is in S1 Table. Our primary aim in these studies was that of a population survey to determine whether Pa from CF or non-CF patients showed differences in their interactions with Af. Thus we designed studies to assess Pa from non-CF patients compared with mucoid and nonmucoid Pa from CF patients. Two experimental designs were used. The first was that of the effects of Pa on the development of biofilm by Af conidia and the second was that of the effects of Pa on preformed Af biofilm. In all of these studies we assessed the effects of live Pa cells and the effects of spent culture filtrates from Pa planktonic or biofilm growth and initially relied on the parameter of metabolic activity of Af, based on XTT reduction. For studies in Figs 2 and 3, five non-CF isolates were selected randomly from the 16 available, and compared to the CF mucoid and non-mucoid isolates.

Inhibition by Pa cells or Pa culture filtrates during Af biofilm formation
The experimental design is shown in Figs 1 and 2, exposing the Af conidia to live Pa cells or filtrates for 16 h followed by an additional 24 h of biofilm growth before metabolic assay of XTT reduction.

Fig 2A summarizes the activity of the Pa cells during Af biofilm formation, as assessed by metabolic activity. Comparisons of effect of individual isolates of Pa on Af with their respective Af control showed that each isolate significantly inhibited Af during the formation of Af biofilm (P<0.001 all comparisons). Pooling of the 5 blocks of data showed that all three groups of Pa isolates significantly inhibited fungal biofilm formation (each group, P<0.001). Of interest, these data indicate that the inhibitory effect of CF mucoid Pa or CF non-mucoid isolates was higher compared to the effect of the non-CF isolates (P<0.001, both comparisons), and CF non-mucoid cells were more inhibitory than CF mucoid cells (P<0.001).

Fig 2B shows the in vitro activity of culture filtrates from Pa grown under planktonic conditions on the fungal biofilm formation, as assessed by metabolic activity. Comparisons showed that the planktonic culture filtrates of each individual isolate of Pa significantly inhibited Af during biofilm formation in comparison with controls (P<0.001). Similarly, the pooled data showed that the planktonic culture filtrates of each of the three groups of Pa isolates significantly inhibited Af (each, P<0.001). The inhibitory effect on Af biofilm formation by the culture filtrates from CF mucoid or CF non-mucoid Pa isolates was higher compared to the effect of non-CF Pa isolates (P<0.001)FAF.
Fig 3. *Pa* inhibition of *Af* biofilm. The data presented show the activity of each live *Pa* isolate or planktonic or biofilm generated culture filtrate on preformed biofilm of *Af*. Individual isolate results are shown in comparison with their respective *Af* controls (n = 6 for each). There were five non-CF, five mucoid CF and five non-mucoid CF isolates tested; isolate designations are shown on the x-axis. The bars on the far right of each panel represent the pooled data for each isolate (n = 30). Bars represent the mean ± SD of the XTT reduction at read at 490 nm. Panel (A) *Af* preformed biofilms exposed to live *Pa* cells for 24 h. (B) *Af* preformed biofilms were exposed to *Pa* planktonic spent supernatant for 24 h. (C) *Af* preformed biofilms were exposed to *Pa* biofilm spent supernatant for 24 h.
of the non-CF filtrates (P<0.001, both comparisons), and culture filtrates from CF non-mucoid Pa cells were more inhibitory than those from CF mucoid cells (P<0.001).

The results of incubation of the conidia with culture filtrates obtained from Pa isolates grown under biofilm conditions are shown in Fig 2C. The culture filtrates of all isolates significantly inhibited Af attempting to form biofilm as compared to their respective controls (P<0.001, all comparisons). Examination of the pooled dated showed that biofilm culture filtrates of Pa isolates from CF or non-CF patients significantly inhibited the metabolic activity of Af in biofilm development compared to the controls (all three groups, P<0.001). In addition, culture filtrates from CF mucoid or non-mucoid Pa isolates were significantly more inhibitory than those from non-CF isolates (P<0.001, both comparisons); CF non-mucoid Pa isolates biofilm culture filtrates were more inhibitory than those from mucoid isolates (P<0.001).

Whereas biofilm formation appears inhibited in the preceding studies, and in CLSM studies and studies of preformed Af biofilm to be described, we cannot completely rule out that differences in Af/biofilm formation are a result of growth inhibition of the fungus. Studies (described below) of Pa inhibition of, by contrast, purely planktonic growth of Af, where there were not significant differences between the three Pa groupings, suggest the inhibitory differences of Pa groups relates more to the effect on Af/biofilm formation.

**Effect of live Pa cells or culture filtrates on preformed Af biofilm**

The experimental design for these studies is shown in Figs 1 and 3, that of allowing Af to form biofilm for 16 h and then exposing this biofilm to live Pa or culture filtrates for an additional 24 h before XTT assessment of Af metabolic activity. Assessment of the inhibitory effects of the live Pa cells showed that only a single non-CF isolate, Pa19, significantly inhibited the preformed Af/biofilm (P<0.01), whereas the other four non-CF isolates had no significant effect (P>0.05). Compared to preformed Af/biofilm controls, all isolates of non-mucoid or mucoid live Pa from CF patients significantly inhibited the metabolic activity of preformed Af/biofilm (P<0.001). Pooling of the five blocks of data showed that non-CF isolates were not inhibitory, whereas mucoid Pa isolates and non-mucoid CF isolates significantly inhibited the metabolic activity of the preformed Af/biofilm in comparisons to controls or non-CF isolates of Pa (P<0.001, both comparisons). Non-mucoid CF isolates were more inhibitory than mucoid CF isolates (P<0.001) (Fig 3A).

Fig 3B shows the activity of planktonic Pa culture filtrates on preformed Af biofilm. Comparisons of individual culture filtrate activity from individual isolates showed that a single non-CF isolate, Pa19, filtrate was inhibitory (P<0.05), whereas all those from individual filtrates from non-mucoid or mucoid Pa from CF patients, grown under planktonic conditions, significantly inhibited the oxidative metabolism of preformed Af/biofilm (P<0.001, all comparisons). The results of analysis of the pooled data showed non-CF planktonic culture filtrates had no inhibitory activity, whereas planktonic filtrates from mucoid or non-mucoid Pa isolates were
more inhibitory than controls or filtrates from non-CF isolates \( (P<0.001, \text{both comparisons}) \). Non-mucoid CF isolate filtrates were more inhibitory than mucoid CF filtrates \( (P<0.001) \).

Fig 3C shows the activity that filtrates from cultures of biofilm-grown Pa isolates had on preformed Af biofilm. No inhibitory activity was exhibited by the non-CF biofilm filtrates \( (P>0.05) \), whereas all biofilm filtrates from mucoid or non-mucoid CF Pa from CF patients significantly inhibited preformed Af biofilm, compared to preformed Af biofilm controls \( (P<0.001) \). Similarly, the comparisons of the pooled data showed that non-CF isolate biofilm filtrates were not inhibitory and both mucoid and non-mucoid isolate biofilm filtrates were more inhibitory than controls or filtrates of non-CF isolates \( (P<0.001, \text{both comparisons}) \). Again, non-mucoid CF isolate biofilm filtrates were more inhibitory than those of mucoid CF isolates \( (P<0.001) \).

**Effect of Pa growth on inhibition by Pa supernatants**

We felt that the effects of the Pa culture filtrates might plausibly be due to differences in growth of the Pa isolates, that isolates with better growth could result in more inhibitory compounds secreted to the culture filtrates. For these and subsequent studies, we chose one representative isolate from each Pa group (non-CF (Pa19), non-mucoid CF (Pa10) and mucoid CF (Pa11)) (Figs 2 and 3). Each was grown planktonically and the 24 h growth quantitated as described (Methods). A \( 10^6 \) inoculum of each grew to \( 3.2 \times 10^8 \) in the case of the two CF isolates, and \( 3.3 \times 10^8 \) for the non-CF isolate. Thus differences in the inhibitory power of the supernatants appear not explained by differences in growth among these isolates.

**Minimal effect of serum on inhibition**

Because components of serum can also be present where there is local inflammation, such as in the airways, to more closely mimic in vivo conditions it is of interest to know whether the interactions described above might be different in the presence of serum. For these studies, serum was added to the medium in all steps (Fig 1), as indicated in Methods, and all 16 non-CF Pa isolates were used, as well as the mucoid and non-mucoid CF isolates.

Overall, the results were virtually the same as those from the studies done in the absence of serum. However, there were exceptions. The Pa live mucoid cells, or the filtrates of such cultures grown planktonically or as Pa biofilm, were, with serum present, not statistically significantly inhibitory, as assessed metabolically, to preformed Af biofilm. Specifically, absent that one exception, the significance and conclusions with (a) mucoid Pa and Af biofilm formation, or (b) non-CF Pa or non-mucoid CF Pa, as live cells or culture supernatants (the supernatants from cultures grown planktonically or as Pa biofilm), were the same as the results in the absence of serum, when tested against either (c) Af biofilm formation or (d) preformed Af biofilm. Again, non-mucoid Pa isolates were most inhibitory in all conditions, and preformed Af biofilm the most resistant. None of the 16 non-CF Pa isolates behaved differently than the rest of that group (data not shown).

In additional experiments (using only the 3 representative Pa isolates, as described in the preceding section, from each Pa group), we omitted serum from Pa while generating supernatants, but added serum during Af biofilm formation or preformed Af biofilm development. We found the same results, as described, as when serum was present or absent in all phases.

Overall, we conclude that serum is not an important enhancing or decremental factor in Pa generation of inhibition in supernatants, nor a factor in the effect on Af.
Comparison of Pa filtrates grown planktonically or as biofilm

All of the studies done with or without serum appeared to indicate that there were differences in inhibitory activity between Pa planktonic and biofilm supernatants. Because there were few experiments, at this point, where these two types of Pa filtrates were compared directly to each other, we approached the comparison, for the studies in Figs 2 and 3, in the statistical fashion described in the Methods section. For the non-CF and the mucoid CF Pa’s, planktonic and biofilm supernatants inhibited Af biofilm formation. However, there was no difference between the planktonic and biofilm culture filtrates from the non-CF Pa, whereas Pa biofilm supernatants from mucoid and non-mucoid CF Pa were more inhibitory than planktonic supernatants (P<0.001, both comparisons). The supernatants of non-mucoid CF Pa were inhibitory to both Af biofilm formation and preformed Af biofilm, and Pa biofilm supernatants were more inhibitory than planktonic supernatants in both cases (P<0.001 for both comparisons). For comparison, a second analysis examined only the studies done in the presence of serum, and with all 16 non-CF isolates. For this second analysis we first compared the controls from each experiment where Pa isolates had been studied (in which only one type of Pa supernatant was used per experiment) by one-way ANOVA to assure the Af control results were not different in those experiments. These analyses showed that there were no significant differences in the Af controls. With this assurance we then compared the inhibition results of the three types of Pa for the two types of Pa supernatants. This allowed 20 experiments with the non-CF Pa’s and 10 experiments each with mucoid and non-mucoid CF Pa to be studied. The conclusions were the same as the prior statistical analysis, that of the studies done in the absence of serum.

These statistical differences in planktonic and biofilm Pa filtrates can also be suggested by visual comparison of Figs 2 and 3, although these differences were more marked when serum was present. In some subsequent experiments, under special conditions to be described subsequently (e.g., comparisons with E. coli), there were direct comparisons of the two kinds of Pa supernatants, and supernatants from planktonically grown Pa were never more inhibitory than supernatants from biofilm-grown Pa, regardless of the source of the Pa.

Confocal microscopy analysis

All the preceding studies assessed metabolic effects on biofilm. We now assessed effects on the physical biofilm itself.

Effects on biofilm thickness

The effect of the Pa spent culture filtrates on Af biofilm thickness and morphology were assessed using CLSM. The thickness results for the Af biofilm after conidia exposure for 16h to the Pa spent supernatants from the representatives of all three bacterial phenotypes are shown (Fig 4). The spent supernatant obtained from all three Pa phenotypes, after Pa growth planktonically or as biofilm, resulted in a significant reduction of the fungal biofilm thickness compared to the untreated control (Fig 4A and 4B). Treatment with the spent supernatants from the non-CF and the CF mucoid isolates had no significant effect on the thickness measurements of preformed Af biofilm, whereas the supernatant of the non-mucoid Pa, grown either planktonically or as biofilm, was inhibitory (Fig 4C and 4D).

Effects on Af biofilm formation

Untreated biofilms showed an architecture formed by a dense filamentous multicellular structure with acute-angle dichotomous branching (Figs 5A and 6A). The Pa supernatants’ effect on Af biofilm was also observed in the morphology studies using CLSM. Fig 5B, 5C and 5D show
the effect of *Pa* culture filtrates on *Af* biofilm formation. Non-CF *Pa* spent supernatant, from *Pa* grown as biofilm, resulted in reduction in filamentation and presence of some "glued" hyphae, without a clear separation of the filamentous elements. A similar morphology pattern was observed when the conidia were treated with planktonic culture filtrates from the same strain (data not shown). Treatment with culture filtrates from the CF mucoid strain culture, grown under biofilm condition, resulted in decreased number of hyphae and presence of some "bulging" structures (possibly conidial remnants) distributed throughout the disperse filaments, and "glued" hyphae (Fig 5C). The CF non-mucoid *Pa* biofilm filtrate resulted in severe effects on hyphal structure, with loss of filamentation, and hyphal tips or branch points appeared to be thin. Several bulges and deposition of amorphous material were also observed. Treatment with planktonic spent supernatant from the CF non-mucoid isolate was associated with a
disruption of the hyphae structure and presence of surface collapse and glued filaments (data not shown).

**Effects on preformed biofilm**

As was suggested by the biofilm thickness data, treatment of *preformed* *Af* biofilm with the spent supernatants from the non-CF and the CF mucoid isolates were not associated with important morphology changes (*Fig 6B* and *6C*). There was a prominent morphology change observed comparing the untreated *Af* control and treatment with the CF non-mucoid *Pa* strain grown under biofilm conditions (*Fig 6D*). We observed severe effects on hyphal structure, with loss of filamentation, and hyphal tips and branch points appeared to be thin after the treatment. Moreover, we observed amorphous structures distributed throughout the intertwined defective filamentous networks.
MW characterization of the inhibitors in filtrates

To better understand the nature of the inhibitor(s) present in the Pa filtrates, we performed a series of studies to assess their physical and chemical nature.

To determine whether the inhibitors in filtrates were related to small molecular species, a sample of inhibitory filtrate from the non-mucoid Pa biofilm culture was passed through an ultrafilter that now excluded all materials >30,000 MW, and tested against Af conidia forming biofilm, in comparison to the same filtrate not further manipulated (as in Fig 1III). These two samples inhibited Af biofilm formation (P<0.001), and were not different from each other. We conclude that the inhibitory component(s) in Pa filtrates appear to be <30,000 MW species.
Specificity

As a brief inquiry into the specificity of the culture filtrates of Pa inhibiting Af, we grew E. coli ATCC 43888 under identical conditions as we had the Pa isolates, and collected these filtrates in the same manner. Supernatant from the non-mucoid Pa was included in these experiments as a positive control. The filtrates of the E. coli, either grown planktonically or as biofilm, did not significantly inhibit Af conidia forming biofilm (conditions under which, in contrast, all Pa filtrates tested could inhibit; see Fig 2), and the positive Pa control filtrate did significantly inhibit. The filtrates of the E. coli, either grown planktonically or as biofilm, also did not inhibit preformed Af biofilm (data not shown).

To ascertain whether the reference Af isolate, 10AF, was representative, three and nine other non-CF Af isolates were tested, by challenging formation of Af biofilm from their conidia, or preformed Af biofilm, respectively (Fig III and IV, respectively), by live Pa cells of the non-mucoid CF Pa. Isolate 10AF was assayed concurrently. The XTT activity of all untreated Af biofilms formed were not significantly different, and all were markedly inhibited ($P<0.001$) by the addition of the Pa cells (data not shown).

We conclude that not all bacteria produce compounds that are inhibitory to Af biofilm and that the inhibition by Pa is not unique to our reference strain of Af.

The effect of Pa culture filtrates on Af is a result of inhibitors in filtrates, not reduction of nutrients

We addressed the possibility that the inhibition of Af biofilm by the Pa filtrates would be due to a reduced nutritional value of the spent culture filtrates diluting out the value of newly added medium during the incubation phase in the Af biofilm assays. The effect of fresh RPMI diluted with Pa culture filtrates (from biofilm-grown Pa) or sterile distilled water (a milieu that offers no nutrient support) on Af biofilm formation was studied. Fungal biofilm metabolic activity was inhibited over a range of ratios of fresh RPMI diluted with culture filtrates obtained from the representative CF and non CF Pa isolates, with inhibition starting at as little as 20% filtrate (80% fresh RPMI) ($P<0.05$) (Fig 7A). In contrast, Af biofilms were not significantly affected after challenge with fresh RPMI diluted in distilled water until the mixture contained only a 20% fresh RPMI concentration (80% distilled water) (Fig 7A); this concentration of RPMI is much lower than that of the 50% fresh RPMI dilution with filtrates as used in the supernatant experiments previously described (Figs 2 and 3). These findings indicate that the presence of inhibitory products in Pa culture filtrates were responsible for the inhibition of the metabolic activity of the fungal biofilms, rather than the inhibition being due to dilution of the nutrients in media by addition of Pa culture filtrate, or removal of the nutrients from the growth media during the Pa growth; i.e., that the inhibition by culture filtrate of Pa is a result of substances ($<30,000$ MW) Pa secretes into the culture filtrate, rather than nutrients removed by Pa.

Although this paper is focused on Af biofilm, it was of interest, for comparison, to see the effect of Pa supernatants on Af growing planktonically, as would occur in tubes, as in classical susceptibility testing [52]. The representative isolates from each of the three Pa groups was studied. When Pa supernatant was diluted in medium 1:1, exactly as in the challenges of biofilm, applied to Af conidia inocula in tubes, and the tubes examined when hyphal growth in matched controls was 4+, visible Af growth in the presence of supernatants did not occur (i.e., 0 growth in the supernatants of the CF isolates, or trace growth in that of the non-CF Pa). To again assure the inhibition seen was a result of toxic factors from the Pa, rather than depletion of media by Pa growth, the dilution series experiment with distilled water was repeated as above (Fig 7B). The result was even more dramatic than seen with Af biofilm targets: any
concentration of supernatant ≥10% (i.e., ≤90% fresh medium) reduced the planktonic growth metabolic activity by XTT virtually completely, whereas it was only when the concentration of distilled water reached 40% (only 60% fresh medium) that any effect at all was seen on that XTT result (Fig 7B). The effect was modestly larger with the non-mucoid CF Pa isolate, but was not statistically significantly greater than the inhibition seen with the supernatants of the other two Pa isolates.

Effect of temperature on Pa supernatants

Two experiments were performed on heat treatment of the Pa filtrates (from planktonic or biofilm cultures), using the 3 representative isolates detailed, and (a) in the absence of serum from all steps or (b) just during the generation of supernatants. Comparison was made to untreated filtrates.

The heat treatment of the supernatants of all 3 Pa isolates, under both test conditions “(a)” and “(b)”, removed all inhibitory activity against Af forming biofilm or preformed Af biofilm, i.e., heated supernatant results same as controls (data not shown). There was one small exception: the non-mucoid CF strain heated supernatant retained significant inhibitory activity against Af forming biofilm, but not against preformed Af biofilm, only when the filtrate was from the Pa biofilm (not planktonic Pa) culture, and only in condition “(b)”.

In another experiment, freeze-thawed supernatant of the non-mucoid CF isolate, either grown planktonically or as biofilm, retained inhibitory activity against Af forming biofilm (both P<0.001 compared to untreated Af control), but this was significantly diminished compared to untreated supernatants (both P<0.001). When either frozen-thawed supernatant was tested against preformed Af biofilm, all activity was lost.
We conclude that the heat treatment used removed essentially all Af-inhibitory activity from supernatants of all types of Pa isolates. Freeze-thawing significantly diminishes inhibitory activity.

**Effect of DNase I and proteinase K on Pa biofilm culture filtrate activity**

To determine whether the inhibitory substances were protein or DNA, we treated a nonmucoid Pa culture filtrate with DNAase or proteinase prior to assay for inhibitory activity. Compared to untreated Pa filtrate, filtrate treated with DNase I showed no significant differences in inhibitory capacity compared to Af control, nor did the DNase I alone have an inhibitory effect (data not shown).

EDTA (a chelator of metal anions, such as Fe; and a component of the DNase assay) alone reduced the metabolic activity of Af biofilm compared to Af biofilm untreated controls ($P<0.001$). The EDTA inhibitory effect was less than that of untreated Pa filtrate. EDTA + DNase I did not affect inhibition of Af biofilm by Pa filtrate. Finally, DNase I had no effect on EDTA inhibition of Af biofilm, as there were no significant differences between EDTA and DNase I + EDTA.

Proteinase K alone did not affect Af biofilm formation. The addition of Proteinase K to Pa filtrates did not affect filtrate inhibition of Af biofilm formation.

**Other characterization of Pa isolates**

Because of the differences we observed among the Pa types tested, we also tested for some phenotypes previously associated with Pa virulence. Elastase activity, a virulence factor in Pa [57], was assessed in the Pa isolates that were used in the Af biofilm studies. The 5 non-mucoid CF isolates more frequently showed a positive test compared to the 5 mucoid CF and 5 non-CF isolates (3 vs. 2 and 1, respectively, at 24 hrs.; 5 vs. 3 and 3 at 48 hrs. with additional incubation at 4°C).

Certain Pa colonial types have been associated with lasR gene (a “master” gene controlling Pa virulence genes) mutations, more severe lung disease and persistence despite eradication attempts, including a wrinkly colony surface, irregular colony edge, metallic sheen, and green colony color [58]. As with elastase, these characteristics were also not significantly associated with our three groups differing in Af inhibition; 2 or 3 (of 5) of the non-mucoid isolates had each of these characteristics, compared to 1 to 3 of the other groups of 5 in each instance.

**Role of iron in inhibition by Pa filtrates**

Iron is a co-factor required for growth and both Af and Pa produce siderophores to acquire iron for their needs. Three experiments were performed to confirm a possible role for iron sequestration by Pa filtrates in the inhibition of Af. A filtrate of the inhibitory non-mucoid Pa, grown under biofilm conditions, was tested (schema as in Fig 1V) alone or with added FeCl$_3$, at 3-fold increasing concentrations from 11 μM through 297 μM, then 10-fold increasing concentrations from 300 μM through 30 mM, against Af forming biofilm, and compared to the usual control (RPMI without filtrate) (Fig 8). In combinations with filtrate, from 11 to 300 μM FeCl$_3$, inhibition by the filtrate was retained, and without significant difference from the inhibition by filtrate in the absence of FeCl$_3$ ($P<0.001$ for inhibition by all, compared to no-filtrate control). At 3000 μM FeCl$_3$, there was no significant inhibition compared to control Af, and the inhibition was significantly less ($P<0.001$) than that with the filtrate without FeCl$_3$. At 30 mM FeCl$_3$, the XTT result showed not only lack of inhibition, but was significantly *enhanced* compared to both the filtrate-free control and filtrate without added FeCl$_3$. 


In separate studies on FeCl₃ alone, reported preliminarily elsewhere [59], we have found, as in Fig 8, FeCl₃ alone at ≥2500 μM to enhance Af biofilm XTT activity. The enhancement of growth in the present study, with 3000 μM FeCl₃ alone, compared to partial inhibition by the same concentration combined with filtrate, confirms the dose-responsive reversal of inhibition by filtrate shown in Fig 8.

We conclude, from the iron reversal of Pa supernatant inhibition, that iron sequestration, such as in the form of Pa siderophores or other Pa iron-binding molecules [60], is at least part of the mechanism of the inhibition of Af biofilm by Pa filtrates.

**Discussion**

Whereas planktonic Af growth appears affected by any Pa isolate, and the inhibition is more intense than on Af biofilm growth, we show that the described inhibition of Af biofilm by Pa [42–45] is related to the source and phenotype of the Pa isolate. Titration studies may have elucidated differences, among Pa types, in inhibition by supernatants on planktonic Af growth, but we chose not to pursue this. The differential effects of Pa types on Af biofilm is consistent with the extensive phenotypic and genomic changes in the mutable Pa organism that, associated with chronic residence in CF airways, have been described [2,8,9,61]. Specifically, we show live Pa cells inhibit the growth and formation of Af biofilm, with CF Pa isolates more inhibitory, and non-mucoid CF Pa isolates most inhibitory. Inhibition did not require the presence...
of Pa cells, as Pa culture filtrates were also inhibitory. The inhibition by filtrates showed the same hierarchy, with CF Pa isolates more inhibitory, and non-mucoid Pa isolates most inhibitory. Preformed Af biofilm was more resistant to Pa; non-mucoid CF Pa isolates were most inhibitory, and, again, that inhibition could be reproduced with culture filtrates. Reproducibility of the three Pa groupings was assured by the repeated experiments of various conditions, utilizing representative isolates of the groupings, as well as the number of isolates studied. Moreover, selection of Pa19, the most inhibitory of the non-CF isolates, as the representative of that group, would have minimized the differences in these studies between the CF and non-CF isolates.

Inhibition of Af biofilm appears dependent on the bacterial growth conditions, since filtrates from Pa grown as biofilm were more inhibitory than from Pa grown planktonically. However, the relationship of the number of Pa cells at the end of biofilm growth vs. the number after planktonic growth is unknown. Differences in biological effect on mammalian cells for molecules produced by planktonic Pa vs. biofilm Pa have previously been described, although planktonic Pa in that study produced more such active molecules [19]. We have assumed the production of substances with different Af-inhibitory power by the three Pa groupings is unrelated to the growth of the Pa, based on our growth studies by representative isolates of each group. However, each of the groups studied may be heterogenous, and more extensive studies, with more members of each group, might then show a relationship between growth and supernatant inhibition.

We chose to measure Af inhibition by the XTT assay of metabolism (which does not exclude the possibility of death of some Af cells), because of the controversies in interpreting CFU reduction in a multinucleate filamentous organism, commonly with incomplete septa between cells [62,63]. Such problems are magnified when attempting to quantitate a biofilm network [64]. However, others have indicated Af is, in fact, killed by Pa [64]. The XTT results would also not exclude a switch by the fungus to non-oxidative, fermentative, metabolism. However, our photomicrographs, and measurements of biofilm thickness, support the concept of actual growth inhibition occurring. The effects we describe on preformed Af biofilm show that Pa effects on Af are not solely on inhibition of the initial adhesion stage of Af biofilm development.

Previous work has indicated many candidate Pa molecules that can explain the various inhibitions we have described [42–47]. We confirm that small Pa-derived molecules, probably not nucleic acids or proteins per our studies, can explain at least most of the inhibition in supernatants. Whereas most previous work has focused on the toxic effects of Pa metabolites on fungi, our work suggests that denial of Fe to Af by Pa products is an important part of the inhibition. This is in contrast to a study of Pa inhibition of Candida albicans, which concluded that inhibition of that fungus was not a result of Fe limitation [46], although these apparent fungal differences could be affected by different experimental conditions. The bacterial-fungal interplay is more complex than just molecules produced by Pa, as recent elegant work has shown that Af can transform Pa metabolites, and thus radically alter the effect on the interaction, including the degree of inhibition [65]. Finally, Fe limitation by Pa could also in part be a result of modulation of Af siderophore production by Pa metabolites [66].

Pa quorum-sensing systems control expression of virulence factors, such as those previously implicated in Af inhibition [42–47]. A study that indicated no difference in quorum-sensing between CF and non-CF Pa isolates [67] might suggest that quorum sensing-induced molecules are at least not solely responsible for the increased inhibitory activity we have described for CF non-mucoid isolates. Some phenotypic factors previously associated with Pa virulence [57,58] that we studied did not appear associated with Af inhibition.
Our findings contrast with others, who reported Af biofilm was not inhibited by Pa [45], a difference likely attributable to their lack of study of CF Pa isolates. Our findings also contrast with the report of heat-resistance of Af-inhibitory factors produced in Pa cultures [45], since we show lability of essentially all the inhibition to heat, as well as significant lability to a freeze-thaw cycle. Such study differences could be a result of different Pa and Af culture and/or heating conditions. These differences support the value of detailing the physical and chemical characterizations of the supernatants described here; these characterizations could lead to further identification of the molecules responsible for the inhibition we have described. Another pathway to this end would be to utilize the many Pa virulence factor mutants [68] that have been described (studies in progress).

A limitation of our studies is that we have only studied one time period, although one adequate for bacterial growth, for production of Pa factors, whereas different regulatory systems may have different kinetics [68]. It may be of interest to study earlier and later time periods of Pa culture, and assess whether inhibitory factors are produced to a greater or lesser extent. We also cannot state at this time whether the enhanced inhibitory activity by biofilm Pa compared to planktonic Pa culture filtrates, or that resulting from non-mucoid CF isolates and their filtrates, is owing to increased production of inhibitory factors, or possibly the production of different factors. Nor is it known whether the inhibition by live cells relies solely on the same inhibitory factors (factors which can be released by Pa cells in close proximity to Af without dilution in culture medium) as in supernatants or whether there are unique factors resulting from or triggered by microbial contact. In this regard, the apparent greater inhibitory activity of live cells compared to supernatants (e.g., Fig 2) could be related to the observation that Pa production of virulence factors can be increased by co-culture with some other microbes [69]. It would also be of interest to study prolonged mixed Pa-fungal biofilm development [64], since the mixed condition is only beginning in our studies of Af forming biofilm challenged with Pa cells. Another issue is that CF patients in proximity to each other can cross-infect Pa isolates [70]; moreover, one Pa clone can therefore come to dominate in patients in one clinic-hospital setting [71]. Thus, without detailed sequencing data, one cannot be sure isolates from multiple CF patients in the same setting are not all very closely related; our observations should be confirmed with isolates from other centers. Finally, a population study, similar to that which we have done for Pa, would be of interest with respect to CF Af isolates, and whether any differ from non-CF, or reference Af, isolates (studies in progress).

Individual patient samples in our cohort shows not only that Pa isolates of varying colonial phenotype can co-exist in the same patient, but isolates with varying Af-inhibitory properties can co-exist; such polymorphism could be a transition state in the evolution of Pa colonization. Overall, from our studies of different Pa phenotypes, we hypothesize the transition of Pa from non-mucoid to mucoid [72–75], a process that usually occurs with time in CF disease, with the latter moving to a deeper zone of the lung [75], ablates Pa’s inhibitory effect on Af biofilm, and thus, helps explain why establishment of Af in the airways usually occurs later in CF disease. Evolution of Pa may act in concert with other factors favoring Af colonization [76], including repeated antibacterial courses.

**Supporting Information**

S1 Table. Compendium of studies reported.

(DOCX)
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Author Contributions
Conceived and designed the experiments: JAGF JAJH KVC AMS DAS. Performed the experiments: JAGF JCP JAJH HN KC. Analyzed the data: JAGF JCP JAJH KVC KC EC DAS. Contributed reagents/materials/analysis tools: RBM AMS NB DAS. Wrote the paper: JAGF JCP RBM JAJH KVC AMS EC DAS. Statistical analyses: JAGF KVC EC DAS.

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