SigE Facilitates the Adaptation of Bordetella bronchiseptica to Stress Conditions and Lethal Infection in Immunocompromised Mice

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sigE facilitates the adaptation of *Bordetella bronchiseptica* to stress conditions and lethal infection in immunocompromised mice

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**Abstract**

**Background:** The cell envelope of a bacterial pathogen can be damaged by harsh conditions in the environment outside a host and by immune factors during infection. Cell envelope stress responses preserve the integrity of this essential compartment and are often required for virulence. *Bordetella* species are important respiratory pathogens that possess a large number of putative transcription factors. However, no cell envelope stress responses have been described in these species. Among the putative *Bordetella* transcription factors are a number of genes belonging to the extracytoplasmic function (ECF) group of alternative sigma factors, some of which are known to mediate cell envelope stress responses in other bacteria. Here we investigate the role of one such gene, sigE, in stress survival and pathogenesis of *Bordetella bronchiseptica*.

**Results:** We demonstrate that sigE encodes a functional sigma factor that mediates a cell envelope stress response. Mutants of *B. bronchiseptica* strain RB50 lacking sigE are more sensitive to high temperature, ethanol, and perturbation of the envelope by SDS-EDTA and certain β-lactam antibiotics. Using a series of immunocompromised mice deficient in different components of the innate and adaptive immune responses, we show that SigE plays an important role in evading the innate immune response during lethal infections of mice lacking B cells and T cells. SigE is not required, however, for colonization of the respiratory tract of immunocompetent mice. The sigE mutant is more efficiently phagocytosed and killed by peripheral blood polymorphonuclear leukocytes (PMNs) than RB50, and exhibits decreased cytotoxicity toward macrophages. These altered interactions with phagocytes could contribute to the defects observed during lethal infection.

**Conclusions:** Much of the work on transcriptional regulation during infection in *B. bronchiseptica* has focused on the BvgAS two-component system. This study reveals that the SigE regulon also mediates a discrete subset of functions associated with virulence. SigE is the first cell envelope stress-sensing system to be described in the bordetellae. In addition to its role during lethal infection of mice deficient in adaptive immunity, our results indicate that SigE is likely to be important for survival in the face of stresses encountered in the environment between hosts.

**Keywords:** *B. bronchiseptica*, Extracytoplasmic function sigma factor, Cell envelope stress, Pathogenesis
Background
The cell envelope of bacterial pathogens is critical for survival both in a host during infection and in the environment outside of the host. As the interface between the bacterium and the outside milieu, the cell envelope acts as a barrier protecting the cell against extracellular hazards. Cell envelope structures are also intimately involved in the formation of contacts with host tissues during infection. To safeguard this important compartment, gram-negative bacteria possess an array of stress responses that sense conditions in the cell envelope and alter gene expression to ensure its integrity [1,2]. In many bacterial pathogens, cell envelope stress responses play a multifaceted role. They provide protection against damage caused by components of the immune system, such as complement and antimicrobial peptides that target the cell envelope [3-5]. They regulate the expression of chaperones required for proper assembly of cell envelope-associated structures, including outer membrane porins, pili, and fimbriae [3,6,7]. In addition, cell envelope stress responses can sense the environment around the bacterium and regulate the expression of virulence factors in response to specific cues, ensuring that these factors are expressed at the proper time and location in the host [2,8]. Despite their importance, no cell envelope stress responses have yet been identified or implicated in pathogenesis in Bordetella species.

Bordetella bronchiseptica is a respiratory pathogen that is closely related to Bordetella pertussis and Bordetella parapertussis, the causative agents of whooping cough in humans [9,10]. B. bronchiseptica causes a range of diseases in various mammals that can be chronic, difficult to completely eradicate, and of variable virulence [11-13]. It is the etiological agent of atrophic rhinitis in swine, kennel cough in dogs, and snuffles in rabbits [12,13]. Documented human infections, generally traced to an animal source, have been observed in immunocompromised individuals, and can be serious, systemic infections [11,14].

The B. bronchiseptica, B. pertussis and B. parapertussis genomes encode a large number of putative transcription factors relative to their overall genome size [15], suggesting that these pathogens have the capacity to extensively regulate gene expression in response to environmental and physiological changes. Despite this finding, only a few Bordetella transcription factors have been studied in any detail [16-20]. Among the predicted transcription factors is an ortholog of the cell envelope stress response sigma factor, σE, of E. coli. In bacteria, sigma factors are the subunits of bacterial RNA polymerases required for specific promoter recognition and transcription initiation [21]. Alternative sigma factors, like σF, are activated in response to specific stresses and rapidly reprogram gene expression by replacing the housekeeping sigma factor and directing RNA polymerase to the genes in their regulons [21,22].

σE belongs to the RpoE-like group of extracytoplasmic function (ECF) sigma factors that have been increasingly implicated as key factors contributing to both bacterial stress responses and virulence [23,24]. These sigma factors are widely distributed across bacterial phyla. Where studied, they direct a diverse set of stress responses primarily targeted to the cell envelope [2,8,24,25]. In E. coli and Salmonella enterica serovar Typhimurium, σE controls many genes whose products are required for the proper expression of outer membrane porins and LPS [26,27]. During infection, σE of S. Typhimurium is required for survival and proliferation in epithelial and macrophage cell lines, and in the presence of antimicrobial peptides [6,28,29]. In Pseudomonas aeruginosa, the σF homologue, AlgU, controls the expression of the exopolysaccharide alginate and conversion to mucoidy. AlgU is constitutively activated in many clinical isolates from cystic fibrosis patients [30,31]. In addition, σE is required for the viability of some bacterial species, but not others. The gene encoding σE is essential in E. coli and Yersinia enterocolitica, but is dispensable in the closely related species S. Typhimurium [6,32,33]. These observations suggest that the functions of σE orthologs have been adapted to combat the challenges each organism faces in its particular environmental niche. By exploring the role of σE in diverse bacterial species, we can learn which aspects of this widespread regulatory pathway are universally conserved and which have diverged over the course of evolution.

Here we show that the B. bronchiseptica σE ortholog, encoded by the gene sigE (BB3752), is an active sigma factor that mediates a cell envelope stress response. This is the first demonstration of an envelope stress-sensing system in Bordetella species. Using a murine infection model, we demonstrate that SigE plays an important role during lethal infection in mice lacking adaptive immunity, but not in respiratory tract colonization. This finding has important implications for human disease, given the observation that B. bronchiseptica can cause serious systemic infections in immunocompromised humans [11,14]. This study suggests that SigE is a critical factor in this process, in addition to the BvgAS master virulence regulatory system.

Results
sigE encodes an active sigma factor
The sigE gene of B. bronchiseptica shares a number of conserved residues with other members of the RpoE-like sigma factors, including those in the DNA-binding regions (Figure 1A) [24]. To determine if sigE encodes an active sigma factor, we asked whether it could direct transcription from the σE-dependent rpoHP3 promoter
in E. coli. This promoter shares a high degree of similarity with a consensus promoter proposed for the RpoE-like sigma factors that was determined from both experimental data and predicted promoter sequences (Figure 1C) [24,27]. The sigE gene from B. bronchiseptica strain RB50 was cloned into the pTrc99a expression plasmid and transformed into a derivative of E. coli MG1655 that carries an rpoH::lacZ reporter gene fusion integrated on
the chromosome [34]. When sigE expression was induced, LacZ activity increased, indicating that SigE can initiate transcription from this promoter (Figure 1B). Furthermore, we found that the gene encoding $\sigma^E$, rpoE, which is essential for viability in E. coli, could be deleted when sigE was overexpressed (data not shown, see Materials and Methods).

To provide additional evidence that SigE is a functional sigma factor, N-terminally His-tagged SigE was purified and tested for its ability to initiate transcription in vitro from the E. coli rpoHP3 promoter. Holoenzyme formed with SigE and E. coli core RNA polymerase (ESigE) was able to direct transcription and produced a transcript of equivalent length to that generated by E. coli Eo$^E$ (Figure 1C). The region immediately upstream of the B. bronchiseptica rpoH homologue, encoded by the fam gene, contains a sequence that is similar to the proposed $\sigma^E$-dependent consensus promoter, suggesting that B. bronchiseptica rpoH is regulated by SigE. Indeed, SigE was able to direct transcription from the putative fam promoter region in vitro (Figure 1C). Taken together, these results demonstrate that SigE is a functional sigma factor and can initiate transcription from promoter sequences similar to those utilized by other members of the RpoE-like sigma factor family.

**sigE contributes to the B. bronchiseptica stress response**

To investigate the role of SigE in B. bronchiseptica, an in-frame deletion of the sigE gene was constructed in RB50 (RB50$\Delta$sigE) that removed 176 out of 200 codons of the gene, leaving 22 and 2 codons at the 5' and 3' ends of the gene, respectively. The deletion was confirmed by PCR and Southern blotting methods (data not shown). $\sigma^E$ orthologs are essential in some bacteria, including E. coli and Y. enterocolitica [33,35], yet are not required for viability in many other species, such as S. Typhimurium, P. aeruginosa, and Burkholderia pseudomallei [6,36,37]. Deletions of B. bronchiseptica sigE were readily obtained, suggesting that it falls in the latter class, and is not essential for viability. Furthermore, RB50$\Delta$sigE grew at a rate similar to that of RB50 under standard growth conditions (37°C in Stainer-Scholle broth) (Figure 2A).

To investigate whether SigE mediates a cell envelope stress response in B. bronchiseptica, we used disk diffusion assays to compare the sensitivity of RB50 and RB50$\Delta$sigE to several chemicals that compromise cell envelope integrity and a series of antibiotics that block different steps in peptidoglycan synthesis. The sigE mutant was more sensitive than the wild-type strain to the detergent SDS in combination with EDTA (Figure 2B). The sigE mutant was also more sensitive than wild-type RB50 to the antibiotics mecillinam and ampicillin (Figure 2B), whereas sensitivity to meropenem, aztreonam, and imipenem was not affected (data not shown). Unlike $\sigma^E$ orthologs in other bacteria, SigE was not required for resistance to the cationic antimicrobial peptide polymyxin B, which targets bacterial membranes, or to osmotic stress (Figure 2B and data not shown) [6,36,38,39]. RB50$\Delta$sigE and RB50 were also equally sensitive to antibiotics that inhibit cytoplasmic processes such as translation (chloramphenicol, erythromycin, kanamycin, tetracycline), transcription (rifampicin), and cytoplasmic enzymes such as DNA gyrase (nalidixic acid), and dihydrofolate reductase (trimethoprim) (data not shown). This lack of sensitivity to multiple antibiotics suggests that the sigE mutation does not lead to an overall increase in the permeability of the outer membrane, which would allow more of the antibiotic to enter the cell. These results show that SigE is important for survival in response to specific types of damage to the cell envelope, such as disruption of cellular membranes caused by SDS/EDTA and interference with synthesis of the peptidoglycan layer caused by ampicillin and mecillinam.

We next asked if sigE is important for survival following a shift to high temperature, which perturbs both the cell envelope and cytoplasm. RB50 and RB50$\Delta$sigE were grown at 37°C to an OD$_{600}$ of 0.4, then shifted to 50°C, a lethal temperature for B. bronchiseptica. Cell viability, assessed by CFU/ml, was measured after the shift to 50°C. Survival of the RB50$\Delta$sigE strain was lower than that of RB50 (Figure 2C). In attempting to complement this phenotype, we found that plasmid-encoded sigE did not restore survival during heat shock (data not shown), although it did complement other phenotypes, as described below. Similar variability in complementation of a $\sigma^E$ mutant by a plasmid-encoded rpoE gene has been seen in other bacteria [29,36,40,41]. Work from Burkholderia cenocepacia showed that expressing $\sigma^E$ from a plasmid actually increased sensitivity to heat stress [36]. In S. Typhimurium, an rpoE mutant was sensitive to paraquat and did not survive in stationary phase under anaerobic conditions. Expression of rpoE from a plasmid partially complemented the former phenotype, but not the latter [29]. Because the anti-sigma factor that regulates $\sigma^E$ activity was not included in any of these instances, it is likely that proper regulation of SigE activity is required for optimal response to particular stresses, not merely excess SigE activity, complicating complementation experiments.

Another aspect of the classical heat shock response is thermotolerance. When bacteria are exposed to an elevated but nonlethal temperature, heat shock responses are induced, resulting in increased production of chaperones and proteases that refold or degrade unfolded proteins [42]. Consequently, the cells are preloaded with protective factors and exhibit increased survival following a subsequent shift to a lethal temperature [42]. To investigate the
role of SigE in this phenomenon, RB50 and RB50ΔsigE were grown to an OD₆₀₀ of 0.1 at 37°C, shifted to 40°C for 90 min, then shifted to 50°C. RB50 cultures incubated at 40°C before 50°C survived better at all time points than those directly shifted from 37°C to 50°C. For example, 54% of the RB50 cells pre-adapted at 40°C survived two hours after the shift to 50°C (Figure 2C) compared to 0.1% survival for those shifted directly from 37°C to 50°C (Figure 2C). RB50ΔsigE pre-adapted at 40°C also survived better at 50°C than when directly shifted from 37°C to 50°C. However, only 38% of the RB50ΔsigE cells survived after one hour (compared to 76% of the wild-type RB50), and 5% survived after two hours at 50°C (Figure 2C). These results demonstrate that B. bronchiseptica exhibits a classical thermotolerance response and that SigE contributes to this response.

Both ethanol and heat shock lead to protein unfolding and membrane perturbation and often elicit similar stress responses [43]. To test the role of sigE in response to ethanol stress, RB50 and RB50ΔsigE were subcultured from mid-exponential-phase cultures into fresh Stainer-Scholte broth with or without 3% ethanol. Both strains grew similarly in medium without ethanol, as noted above. RB50 grew significantly slower in medium containing 3% ethanol than in medium without ethanol (compare the growth curve for RB50 in Figure 2D with that in Figure 2A), but eventually reached a cell density only slightly below that of cultures grown without ethanol. Both strains grew similarly in medium without ethanol, as noted above. RB50 grew significantly slower in medium containing 3% ethanol than in medium without ethanol (compare the growth curve for RB50 in Figure 2D with that in Figure 2A), but eventually reached a cell density only slightly below that of cultures grown without ethanol. In contrast, the cell density of RB50ΔsigE grown in the presence of 3% ethanol never surpassed an OD₆₀₀ of around 0.1, even after 24 hours. Expression of plasmid-encoded sigE in RB50ΔsigE complemented this
phenotype, restoring growth in medium with 3% ethanol to nearly that of RB50 (Figure 2D), indicating that sigE is required for survival during ethanol stress.

σE homologues have also been found to play a role during oxidative stress in S. Typhimurium and Burkholderia pseudomallei [29,41]. However, in disk diffusion assays, SigE was not required for survival in the presence of hydrogen peroxide or paraquat, two inducers of oxidative stress (data not shown). Either SigE is not involved in combating oxidative stress in B. bronchiseptica, or other oxidative-stress responsive pathways compensate for SigE when it is absent.

Growth in the murine respiratory tract is not affected by the lack of sigE

*B. bronchiseptica* RB50 colonizes the respiratory tract of immunocompetent mice, causing an asymptomatic infection that is eventually cleared by the immune system. To determine whether *B. bronchiseptica* SigE contributes to colonization and persistence in the respiratory tract, groups of C57BL/6 mice were inoculated with RB50 or RB50ΔsigE. Colonization was measured in the nasal cavity, trachea, and lung on days 0, 3, 7, 14, 28 and 63 post-inoculation. Both wild-type and sigE-deficient RB50 colonized the nasal cavity at comparable levels, peaking on day 3 post-inoculation, and stabilizing at about 10^{5-5} CFU by 2 weeks post-inoculation (Figure 3). Both strains also showed similar colonization kinetics in the lower respiratory tract of C57BL/6 mice, peaking in numbers on days 3 and 7 post-inoculation in the trachea and lungs, respectively, and declining thereafter, with complete clearance in both organs by day 63 post-inoculation (Figure 3). These data indicate that *B. bronchiseptica* SigE is not required for colonization or persistence in the murine respiratory tract.

SigE contributes to lethal *B. bronchiseptica* infection in mice lacking B cells and T cells, but not in mice lacking TLR4 or TNF-α

*B. bronchiseptica* has been observed to cause a range of disease including bronchitis, lethal pneumonia, and even systemic infection [11,12]. Mice with defined immune deficiencies are particularly susceptible to different forms of disease [44-46], facilitating assessment of the roles of specific bacterial factors/functions in interactions with different aspects of the host immune response.

Mice lacking key components of innate immunity, either TLR4 or TNF-α, were challenged with RB50 or RB50ΔsigE and signs of severe disease were monitored. Consistent with published studies, TLR4^{−/−} and TNF-α^{−/−} mice inoculated with 10^{5} CFU of RB50 quickly developed signs of lethal bordetellosis such as ruffled fur, hunched posture, decreased activity, and difficulty breathing, and succumbed 2 to 5 days post-inoculation [46,47]. Mice challenged with RB50ΔsigE also showed similar signs of disease and time to death (data not shown). In a separate experiment, TLR4^{−/−} mice and TNF-α^{−/−} mice infected with RB50 or RB50ΔsigE that were still alive by day 3 post-inoculation were dissected for bacterial enumeration in the respiratory as well as systemic organs. Both strains also grew to higher numbers in the system organs in TLR4^{−/−}, but not TLR4^{+/+} mice (data not shown). Both strains also grew to higher numbers in the

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**Figure 3** Colonization of the respiratory tract of C57BL/6 mice by RB50 and RB50ΔsigE. Groups of three 4–6 week-old C57BL/6 mice were inoculated with 5 × 10^{5} CFU of RB50 (filled squares) and RB50ΔsigE (open triangles). Groups of three mice were sacrificed at each time point. The bacterial load in the indicated organ is expressed as log_{10} CFU ± SE. The dashed line indicates the limit of detection. The experiment was performed twice with similar results and a representative dataset is shown.
lungs of TNF-α−/− mice than in the lungs of C57BL/6 mice and were recovered from systemic organs only in TNF-α−/− mice (data not shown). These data indicate that SigE is not required for B. bronchiseptica to cause lethal infection and colonize systemic organs in mice lacking TLR4 or TNF-α.

B and T cell-deficient Rag1−/− mice succumb to B. bronchiseptica infection, and death is associated with systemic spread of the infection [48]. To assess the role of SigE during infection in hosts deficient in adaptive immunity, groups of Rag1−/− mice were inoculated with 5 × 10^5 CFU of RB50 or RB50ΔsigE. Rag1−/− mice inoculated with RB50 showed symptoms of lethal bordetellosis on day 13 post-inoculation and succumbed between days 14–35 post-inoculation (Figure 4A). However, Rag1−/− mice inoculated with RB50ΔsigE survived without any overt signs of disease and were euthanized on day 122 post-inoculation. The nasal cavity, trachea, lungs, spleen, liver, and kidneys of these mice were excised to enumerate bacterial loads. Although 10^5-7 CFU of RB50ΔsigE were recovered from the respiratory tract, this strain failed to colonize the spleen or kidney, and only 300 CFU were recovered from the liver (Figure 4B, dark gray bars). In a separate experiment, RB50 and RB50ΔsigE-inoculated Rag1−/− mice were sacrificed on day 28 post-inoculation, when some of the RB50-challenged mice were still alive. The bacterial loads of RB50 and RB50ΔsigE in the respiratory tract on day 28 post-inoculation were similar, about 10^5-7 CFU. At this time, 10^1-6 CFU of RB50 were recovered from liver, spleen, and kidney (Figure 4B, white bars). RB50ΔsigE, however, failed to colonize the spleen, kidney or liver (Figure 4B, light gray bars). These results demonstrate that SigE is required for lethal infection by B. bronchiseptica in Rag1−/− mice.

The failure of RB50ΔsigE to colonize distal organs of Rag1−/− mice suggests that this mutant may be defective in getting into or survival in the bloodstream and/or systemic organs. The bloodstream includes many important bactericidal factors of the host immune system, including complement and phagocytes. We first examined whether B. bronchiseptica lacking sigE is more susceptible to complement-mediated killing. 500 CFU of RB50, RB50ΔsigE, or RB50Δwbm, a strain lacking O-antigen, which is known to be susceptible to complement [48], were incubated at 37°C for one hour in PBS with 20% complement-active or complement-inactive serum from naïve mice. The survival of RB50ΔsigE and RB50 was not affected by the presence of either serum (data not shown). In contrast, the RB50Δwbm strain was almost completely killed by complement-active, but not complement-inactive serum (0.7% survival in the presence of complement-active serum compared to 100% survival in the presence of complement-inactive serum). The observation that RB50ΔsigE survived in the presence of serum without B. bronchiseptica-specific antibodies indicates that the defect in causing systemic infection in mice lacking B and T cells is not due to failure to survive the antimicrobial components in serum, including complement.

**SigE contributes to cytotoxicity to macrophages**

We further tested whether RB50ΔsigE interacts differently than RB50 with another major bactericidal component in the bloodstream, phagocytes. B. bronchiseptica is cytotoxic to macrophages, and this toxicity has been attributed to the activities of the type three secretion system (TTSS) [49]. To test the role of SigE in macrophage cytotoxicity, RAW264.7 murine macrophages were incubated for 4 hours at an MOI of 10 with RB50,
RB50 lacking sigE, or RB50 lacking a functional TTSS (WD3). In this experiment, both the RB50 and RB50ΔsigE strains contained the empty cloning vector pEV to allow direct comparisons with the complemented strain, RB50ΔsigE pSigE. Cytotoxicity was determined by measuring LDH release from the treated macrophages. WD3 caused little cytotoxicity, similar to treatment with medium alone. RB50ΔsigE pEV caused approximately 50% less cytotoxicity than wild-type RB50 pEV (Figure 5). This defect in cytotoxicity was complemented by supplying the sigE gene on the plasmid pSigE (Figure 5), indicating that loss of sigE negatively impacts the ability of RB50 to kill macrophages.

RB50ΔsigE is more efficiently phagocytosed and killed by PMNs

To test if RB50ΔsigE is more susceptible to another bactericidal mechanism, phagocytosis by peripheral blood polymorphonuclear leukocytes (PMNs), RB50 and RB50ΔsigE were incubated with freshly isolated human PMNs and attachment to, phagocytosis by, and killing by these cells were measured. PMNs bound RB50ΔsigE more efficiently than RB50 (Figure 6A), and significantly more RB50ΔsigE than RB50 were phagocytosed by PMNs (Figure 6B). However, the number of viable intracellular RB50ΔsigE was ~50% of the numbers of viable RB50 (Figure 6C, left panel). When differences in attachment and phagocytosis were taken into consideration, significantly more internalized RB50ΔsigE were killed compared to RB50 (Figure 6C, right panel). Together, these data indicate that SigE contributes to B. bronchiseptica resistance to phagocytosis and killing by PMNs.

Discussion

The BvgAS system of the bordetellae plays a central role in regulating gene expression during pathogenesis [50-52]. However, other regulators may be required during the infectious disease cycle, as Bordetella genomes have a large number of putative sensory systems.
In this study, we focused on cell envelope sensing systems and investigated the alternative sigma factor, SigE. We found that SigE of *B. bronchiseptica* does indeed mediate a protective cell envelope stress response and that strains lacking SigE do not establish lethal infections in mice lacking adaptive immunity. These data suggest that the role of SigE is to combat stresses to the envelope imposed by the immune system within a host and by harsh conditions in the environment outside a host. This work is the first demonstration of a cell envelope sensing system in the bordetellae. The $\sigma^E$ system has been explored in the most depth in enteric pathogens belonging to the Gammaproteobacteria [23,25,53]. The bordetellae, members of the Betaproteobacteria, encounter distinctly different environments in the respiratory tract and therefore provide an excellent model to study how the SigE system has been adapted throughout evolution to serve the needs of diverse bacterial pathogens.

The entire *sigE* locus (BB3752-BB3750) is identical at the amino acid sequence level among the classical bordetellae, suggesting a conserved role in the human pathogens *B. pertussis* and *B. parapertussis*. However, the lifestyles and, therefore, conditions encountered differ amongst these three species. *B. bronchiseptica* can live outside the host and primarily infects mammals, although it can infect immunocompromised humans [11,14]. In contrast, *B. pertussis* and *B. parapertussis* primarily infect humans and are directly transmitted between hosts [54,55]. As we learn more about the role of SigE in the bordetellae, it will be of interest to determine whether stresses that induce the SigE system and the SigE regulon members are as highly conserved as the *sigE* locus itself among the bordetellae.

Our results define roles for SigE in *B. bronchiseptica* that are only partially overlapping with those for $\sigma^E$ in other pathogens. SigE was important for survival of *B. bronchiseptica* in the face of both global stresses to the cell envelope caused by heat shock, exposure to ethanol and detergent, and specific stresses caused by several beta-lactam antibiotics (Figure 2). Heat shock, ethanol, and detergent are classical stressors used in the laboratory to mimic conditions that lead to unfolded proteins and disrupted lipids during infection and in the environment. In contrast to the *B. cenocepacia* and S. Typhimurium proteins, *B. bronchiseptica* SigE was not required for survival during osmotic stress [6,36]. SigE was also not required for response to oxidative stress or the antimicrobial peptide polymyxin B, unlike the S. Typhimurium $\sigma^E$ ortholog [6,29]. The variations among bacteria in their use of $\sigma^E$ systems likely reflect both differences in stresses encountered in environmental reservoirs and in particular host tissues during infection, as well as differences in the arrays of additional cellular stress responses possessed by each species. These other responses can act along with or in place of $\sigma^E$. The presence of other stress responses may be particularly pertinent to *B. bronchiseptica*. Its genome is predicted to encode six related ECF sigma factors of unknown function in addition to SigE [24] that may have complimentary and redundant functions with SigE. Future studies defining conditions that activate other ECF sigma factors and their roles in *B. bronchiseptica* pathogenesis will provide a more comprehensive understanding of how *B. bronchiseptica* copes with extracytoplasmic stress.

Stress response systems, like the $\sigma^E$ system, rapidly induce the expression of specialized sets of genes. These systems are often tightly regulated and expressed only when needed, because inappropriate expression of their regulons can interfere with other important cellular functions [8,56,57]. We found that SigE was not required for colonization and persistence of RB50 within the respiratory tract of an immunocompetent host (Figure 3), the primary niche of *B. bronchiseptica*. This result suggests that the pathogen does not encounter stresses in the respiratory tract that require a response by the SigE system. However, *B. bronchiseptica* encounters different challenges during infection in Rag1$^{-/-}$ mice lacking B and T cells. In these mice, the infection spreads to the bloodstream, which is under greater immune surveillance and has a different arsenal of antimicrobial factors to attack invaders than the respiratory tract. The defect of RB50Δ*sigE* in lethal infection of Rag1$^{-/-}$ mice, therefore, reveals a specific function for SigE in response to an unknown stress, possibly related to the innate immune response, that the bacteria encounter during infections that proceed beyond colonization of the respiratory tract.

The inability of RB50Δ*sigE* to cause lethal infections in Rag1$^{-/-}$ mice (Figure 4) could be due to failure to enter or survive in the bloodstream and/or systemic organs of these mice. Since the mutation does not affect survival during incubation with serum in vitro, it is unlikely that the *sigE*-deficient strain is more susceptible to complement or other antimicrobial components in serum. The defect in infection of Rag1$^{-/-}$ mice may then be related to altered interactions of the mutant strain with phagocytic cells in the bloodstream. RB50Δ*sigE* is more susceptible to peripheral blood PMNs than RB50 (Figure 6), and is also less cytotoxic to macrophages than RB50 (Figure 5). Either or both of these defects could explain the failure to recover RB50Δ*sigE* from systemic organs of mice lacking adaptive immune responses and the decreased virulence in these mice.

Why does the RB50Δ*sigE* mutant spread systemically and cause lethal infection in TLR4$^{+/-}$ and TNF-α $^{-/-}$ mice, but not Rag1$^{-/-}$ mice? The lower cytotoxicity of the *sigE* mutant and its increased sensitivity to phagocytic killing
does not affect its virulence in mice lacking innate immune functions. This could be because bacterial numbers within the respiratory tract of TLR4<sup>def</sup> or TNF-α<sup>−/−</sup> mice are nearly an order of magnitude higher than in the lungs of Rag1<sup>−/−</sup> mice. As such, the large number of bacteria in TLR4<sup>def</sup> or TNF-α<sup>−/−</sup> mice may overwhelm limiting host antimicrobial defense mechanisms that can contain the lower bacterial numbers in the lungs of Rag1<sup>−/−</sup> mice. Alternatively, although the cytotoxicity of the sigE mutant is reduced, it may still be sufficient to establish lethal infections in the absence of TLR4 or TNF-α. Thus TLR4- and TNF-α-dependent functions, such as efficient phagocytosis and killing, appear to be sufficient to prevent lethal infection by RB50ΔsigE in Rag1<sup>−/−</sup> mice. Although the exact role remains to be elucidated, our results clearly indicate that SigE is required for lethal infection of mice lacking B and T cells.

Although the <i>B. bronchiseptica</i> strain RB50 causes asymptomatic infections in immunocompetent mice, other strains of <i>B. bronchiseptica</i> can cause a wide range of disease severity in other hosts [11-13]. In particular subsets of immunocompromised humans, such as those infected with HIV, severe systemic <i>B. bronchiseptica</i> infections have been observed [14]. These facts, along with the high degree of sequence conservation for the sigE locus in <i>B. pertussis</i> and <i>B. parapertussis</i>, highlights

### Table 1 Strains and plasmids

| Strain name | Genotype | Source, Reference |
|-------------|----------|-------------------|
| **E. coli** |          |                   |
| SEA001      | MG1655 ΔxpoHP3::lacZ ΔlacX74 | [60] |
| SEA5036     | BL21(DE3) ΔslyD:kan pLysS pPER76 | [61] |
| XQZ001      | BL21(DE3) ΔslyD:kan pLysS pXQZ001 | This work |
| SEA4114     | CAG43113 ΔrpoE:kan ΔnadB::Tn10 | [62] |
| SEA008      | SEA001 pTrc99a | [62] |
| SEA5005     | SEA001 pSEB006 | This work |
| XQZ003      | DH5α pXQZ0003 | This work |
| SS1827      | DH5α pSS1827 | [63] |
| **B. bronchiseptica** | | |
| RB50        | RB50     | [58] |
| SEA5516     | RB50ΔsigE | This work |
| MER001      | RB50 pCW505 | This work |
| MER002      | RB50ΔsigE pCW505 | This work |
| SEA5518     | RB50 pEV | This work |
| SEA5520     | RB50ΔsigE pEV | This work |
| SEA5526     | RB50 pSigE | This work |
| SEA5530     | RB50ΔsigE pSigE | This work |
| RB50Δwbm    | RB50Δwbm pSigE pET15b | [64] |
| WD3         | RB50Δwbm pET15b | | |
| **Plasmid name** | **Description** | **Source, Reference** |
| pTrc99a     | Vector, pBR322 ori, Ap<sup>R</sup> | Pharmacia |
| pSEB006     | sigE in pTrc99a | This work |
| pSEB015     | isolated rpoHP3 promoter in pRLG770, Ap<sup>R</sup> | [61] |
| pPER76      | rpoE in T7 expression vector pET15b, Kan<sup>R</sup> | [65] |
| pXQZ001     | sigE in T7 expression vector pET15b, Kan<sup>R</sup> | This work |
| pXQZ002     | ΔsigE in TOPO-TA vector | This work |
| pSS1827     | helper plasmid competent for mating, Ap<sup>R</sup> | [63] |
| pSS3962     | Bordetella-specific allelic exchange vector, Kan<sup>R</sup> | Stibitz, unpublished work |
| pXQZ003     | ΔsigE in pSS3962 | This work |
| pEV         | Vector pJS72, OSpec<sup>C</sup> cassette replaced with Cm<sup>R</sup> | This work |
| pSigE       | sigE in pEV | This work |
| pCW505      | cytoplasmic expression of GFP | [66] |
the importance of understanding the stressors that activate SigE and how the SigE system responds to them during infection.

Conclusions
In this work, we have demonstrated that the *B. bronchiseptica* extracytoplasmic function sigma factor, SigE, is important for surviving global stresses that affect the whole cell, such as heat shock and ethanol stress, specific stresses that target the cell envelope, such as betalactam antibiotics and SDS-EDTA, and in interactions with the host innate immune system, particularly phagocytes. During infection, SigE is not required for colonization of the respiratory tract of immunocompetent mice. However, it is needed for a specific set of functions associated with virulence, particularly those involved in surviving the innate immune response when the infection progresses in immunocompromised mice. Although SigE systems are widely conserved, the details as to which aspects are shared and which have diverged are complex. As evidence accumulates from studies in different bacteria, it is becoming apparent that these sensory modules are important for stress survival, particularly with respect to the cell envelope. However, the nature of the stresses that SigE systems combat varies. During infection, comparisons are even more difficult, since differences are seen not only amongst SigE systems from one pathogen to another, but also within different niches in the host or during the progression of disease for a single pathogen.

Methods
Strains and media
A complete list of strains used in this study can be found in Table 1. *B. bronchiseptica* strains are derivatives of the previously described *B. bronchiseptica* strain RB50 [58]. *B. bronchiseptica* was maintained on Bordet-Gengou (BG) agar (Difco) containing 10% defibrinated sheep blood (Hema Resources) and 20 μg/ml streptomycin. In liquid culture, *B. bronchiseptica* was grown in Stainer-Scholte broth [59] with aeration. Chloramphenicol was used at 20 μg/ml and IPTG at 1 mM where noted. The RB50ΔsigE mutant was constructed as described below. *E. coli* strains used to measure SigE activity are derivatives of MG1655 that carry the σ^70^-dependent rpoH::lacZ reporter (strain SEA001 [34]). *E. coli* strain BL21(DE3) pLysS was used to express constructs for protein purification. *E. coli* were grown in LB broth in a gyratory water bath with aeration. Ampicillin was used at 100 μg/ml, tetracycline at 20 μg/ml, and kanamycin at 15 μg/ml as needed for experiments with *E. coli*.

Plasmid constructions
All plasmids used in this study are listed in Table 1 and oligonucleotide sequences are given in Table 2. Plasmid pSEB006 was constructed to express sigE in *E. coli*. The sigE gene was amplified from RB50 genomic DNA with the primers SigEF and SigER and cloned into the expression vector pTrc99a under the control of the IPTG-inducible trc promoter. To facilitate purification of SigE, the plasmid pXQ001 was constructed by amplifying the sigE gene from RB50 genomic DNA using the primers HisSigEF and HisSigER. The resulting PCR product was cloned into the T7 expression vector pET-15b (Novagen), which adds a 6X-His tag to the N-terminus of recombinant proteins. To express sigE in *B. bronchiseptica*, sigE was amplified from RB50 genomic DNA using primers 72SigEF and 72SigER and ligated into the XbaI and Xhol sites downstream of the plac promoter in pEV to create pSigE. The expression vector pEV was constructed from the broad host range vector pJST2 by replacing the spectinomycin resistance gene with the cat gene encoding chloramphenicol resistance amplified from pKD3 [67] using primers 72ChlorF and 72ChlorR. The exchange of drug markers was necessary because RB50 is naturally resistant to spectinomycin. pEV and pSigE were moved into RB50 and RB50ΔsigE through tri-parental mating on BG agar with MgCl2. Transconjugants were selected on BG containing 60 μg/ml streptomycin and 20 μg/ml chloramphenicol. Plasmid pCW505 (kindly supplied by Dr. Alison Weiss, Cincinnati, Ohio), which induces cytoplasmic expression of GFP without affecting growth or antigen expression, was used to visualize RB50 and RB50ΔsigE in the phagocytosis assays described below [68].

Construction of RB50ΔsigE strain
The sigE gene was deleted from RB50 using a Bordetella-specific allelic exchange procedure to produce strain SEA5516. Primers used in the construction are listed in Table 2. A PCR product containing 637 bp proximal to the 5' end of sigE was amplified from RB50 genomic DNA using primers SigEKO_LeftF and SigEKO_LeftR. A non-overlapping PCR product containing 534 bp proximal to the 3' end of sigE was amplified with primers SigEKO_RightF and SigEKO_RightR. The two fragments were digested with BamHI and ligated. The resulting construct was amplified with primers SigEKO_LeftF and SigEKO_RightR, cloned into the TopoTA vector (Invitrogen), and verified by sequencing to give plasmid pXQ002. In this deletion construct, the 528 bp central region of the sigE gene is deleted leaving 66 bp at the 5' end and 6 bp at the 3' end of the sigE gene. The deletion construct from pXQ002 was then cloned into the EcoRI site of the allelic exchange vector pSS3962 (Stibitz S., unpublished data) to generate pXQ003 and transformed into *E. coli* strain DH5α. Tri-
parental mating with wild-type *B. bronchiseptica* strain RB50, *E. coli* strain DH5α harboring the pXQ003 vector (strain XQ003), and DH5α harboring the helper plasmid pSS1827 (strain SS1827) [69,70] and selection of mutants were performed as previously described [69]. The deletion strain was verified by PCR using primers SigEKO_LeftF and SigEKO_RightR and by Southern blot analysis.

β-galactosidase assays
Overnight cultures were diluted into fresh medium and grown to an OD600 of 0.1-0.2 at 30°C. Where indicated, IPTG was added to a final concentration of 1 mM. Samples were collected 2.5 hours later and β-galactosidase activity from the σE-dependent reporter was assayed as previously described [60,71].

**Complementation of E. coli ΔrpoE by B. bronchiseptica sigE**
The ability of *B. bronchiseptica* sigE to suppress the lethality caused by deletion of *rpoE* in *E. coli* was determined using a cotransduction assay as described [62]. The ΔrpoE::kan ΔnadB::Tn10 allele from strain SEA4114 was moved via P1 transduction into strain SEA5036, which carries sigE on the plasmid pSEB006. Tet-resistant (tetR) transductants were selected and then screened for kanamycin resistance (kanR). Although the nadB and rpoE alleles are tightly linked (>99%), cotransduction resulting in tetR kanR colonies will only occur if rpoE is no longer essential for viability. In transductions with *E. coli* expressing sigE (strain SEA5005) as the recipient strain, 31 out of 32 tetR transductants were also kanR. In contrast, none of the 39 tetR transductants were kanR when *E. coli* carrying the empty cloning vector (strain SEA008) was the recipient strain.

**Protein purification**
N-terminally His-tagged *B. bronchiseptica* SigE and *E. coli* σE were purified from strain XQZ001 and SEA5036, respectively, as previously described for *E. coli* σE [61]. Briefly, cells were grown at 25°C to an OD600 of 0.5, at which point IPTG was added to induce protein production. Following 1.5-3 hours of induction, cells were harvested by centrifugation and resuspended in lysis buffer containing 20 mM Tris–HCl pH 8.0, 500 mM NaCl, 2.5 mM β-mercaptoethanol, 2.5 mM DTT, 1 mM PMSF. Resuspended cells were then lysed by sonication, and the lysate cleared by centrifugation. The supernatant containing soluble His-SigE was loaded onto a Ni-NTA column (Qiagen). Bound proteins were eluted with a stepwise gradient of 20, 60, 100, and 200 mM imidazole in column buffer (20 mM Tris–HCl pH 8.0, 500 mM NaCl, 2.5 mM β-mercaptoethanol). Fractions containing SigE were pooled and dialyzed into 20 mM Tris–HCl pH 8.0, 50 mM NaCl, and 2.5 mM β-mercaptoethanol.

**In vitro transcription**
100 nM *E. coli* core RNA polymerase (Epicentre) was incubated with 400 nM His-SigE or His-σE in transcription buffer (40 mM Tris–HCl pH 8.0, 10 mM MgCl2, 50 mM NaCl, 1 mM DTT, 0.1 μg/ml BSA) for 10 min at 30°C to form holoenzyme. Multi-round transcription reactions were initiated by addition of holoenzyme at a final concentration of 40 nM sigma factor and 10 nM core RNA polymerase, to prewarmed (30°C) transcription mix containing 5.0 nM supercoiled plasmid template pSEB015 [61] or 5.0 nM linear Pfam template, 5% glycerol, 200 mM ATP, 200 mM CTP, 200 mM GTP, 10 mM UTP, and 2.5 mM [α-32P]UTP in transcription buffer. After 10 min at 30°C, reactions were stopped by...
the addition of stop solution (80% formamide, 20 mM EDTA, 0.1% xylene cyanol, and 0.1% bromophenol blue). Samples were electrophoresed on 6% polyacrylamide gels containing 7.5 M urea, and transcripts were visualized by phosphorimaging. The linear Pfam template was generated by amplification of the promoter region of the gene encoding $\sigma^{32}$ in RB50, $fam$, using the primers PFamF and PFamR (Table 2). The sequence logo in Figure 1C was generated using WebLogo version 2.8.2 (http://WebLogo.berkeley.edu, [72]).

**Disk diffusion assays**

*B. bronchiseptica* cultures in mid-log phase were diluted to $6 \times 10^8$ CFU/ml and spread on Stainer-Scholte agar plates to generate a lawn of bacteria. Disks containing 300 IU polymyxin B, 10 $\mu$g ampicillin, 100 $\mu$g mecillinam, 750 $\mu$g sodium dodecyl sulfate (SDS) and 2.9 $\mu$g EDTA, 30 $\mu$g azoneam, 10 $\mu$g imipenem, 10 $\mu$g meropenem, 30 $\mu$g chloramphenicol, 15 $\mu$g erythromycin, 30 $\mu$g kanamycin, 30 $\mu$g nalidixic acid, 150 $\mu$g rifampicin, 23.75 $\mu$g sulfamethoxazole and 1.25 $\mu$g trimethoprim, 30 $\mu$g tetracycline, 3.0 $\mu$g deoxycholate, 3% hydrogen peroxide, or 2% paraquat were applied to the plates and the zones of inhibition were measured after overnight incubation at 37°C.

**Temperature and ethanol stress**

For temperature stress experiments, mid-log phase cultures of RB50 and RB50$\Delta$sigE were diluted to an OD$_{600}$ of 0.01 in fresh Stainer-Scholte broth and incubated at 37°C in a gyratory water bath with shaking. At an OD$_{600}$ of 0.1, cultures were either shifted to 40°C for adaptation or kept at 37°C. After 90 minutes, all cultures were shifted to 50°C, and survival was measured by plating and CFU counts. For ethanol stress experiments, mid-log-phase cultures of the pertinent strains were subcultured into fresh Stainer-Scholte broth with or without 3% ethanol and incubated at 37°C in a gyratory water bath with aeration. Bacterial growth was measured by OD$_{600}$.

**Complement killing assay**

Complement killing assays were performed as previously described [73]. Approximately 500 CU of RB50, RB50$\Delta$sigE, and RB50$\Delta$wbm from mid-log phase cultures were incubated with 45 $\mu$l of diluted serum from C57BL/6 mice or PBS (final volume for incubation was 50 $\mu$l) for 1 hour at 37°C. Bacterial numbers before and after incubation were determined by plating and CFU counts. Each strain was assayed in triplicate.

**Cytotoxicity assay**

Cytotoxicity assays were performed as previously described [44]. Briefly, bacteria were added to RAW 264.7 murine macrophage cells at a multiplicity of infection (MOI) of 10 and incubated for four hours. Percent lactate dehydrogenase (LDH) release, a measure of cytotoxicity, was determined by using Cytotox96 Kit (Promega) according to the manufacturer’s protocol.

**Phagocytosis and killing by polymorphonuclear leukocytes**

Attachment and phagocytosis of the *B. bronchiseptica* strains by peripheral blood polymorphonuclear leukocytes (PMNs) were evaluated as previously described with a few modifications [74]. Briefly, GFP-expressing bacteria were incubated with PMNs at an MOI of 50 for 20 min at 37°C to allow binding. After extensive washing to remove non-attached bacteria, an aliquot was maintained on ice to be used as a bacterial attachment control. The remaining PMNs were further incubated for 30 min at 37°C to allow internalization, at which point phagocytosis was stopped by placing PMNs on ice. Bacteria bound to the cell surface in both aliquots were detected by incubation with RB50 immune serum for 30 min at 4°C, followed by incubation with R-phycocerythrin (RPE)–labeled goat F(ab')$_2$ fragments of anti-mouse IgG at 4°C for 30 min. All incubations were done in the presence of 25% heat-inactivated human serum to prevent nonspecific binding of antibodies. After washing, ten thousand cells per sample were analyzed by flow cytometry. Attachment control samples were also analyzed by fluorescence microscopy using a DMLB microscope coupled to a DC 100 camera (Leica Microscopy Systems Ltd.). Green fluorescence intensity associated with PMNs maintained at 37°C for 20 min has previously been shown to represent bacterial attachment [74]. Phagocytosis was calculated from the decrease in mean red fluorescence intensity of GFP-positive PMNs after the 30 min incubation allowing for internalization, as previously described [75]. Percent phagocytosis was calculated as follows: 100 x (1-RPE2/RPE1), where RPE1 is the mean RPE-fluorescence of the GFP-positive cells after 20 min at 37°C (attachment control) and RPE2 is the mean RPE-fluorescence of the GFP-positive cells after 50 min (internalized bacteria) at 37°C.

Killing of bacteria by PMNs was assessed as follows: after phagocytosis of the bacteria, 400 $\mu$g/ml polymyxin B and 350 $\mu$g/ml chloramphenicol were added to the PMNs for 1 hour to kill the remaining extracellular bacteria and assess intracellular survival. Serial dilutions of samples were plated to determine the number of viable intracellular bacteria per PMN. The relative percent survival of internalized bacteria was calculated from the relative phagocytosis index and taking into account the initial attachment level of each strain, as follows: percent bacterial killing = [1-N/(A x P)] x 100, where A = number of bacteria associated with PMN after 20 min at 37°C (determined by fluorescent microscopy), P = phagocytosis index of bacteria associated with PMN after 4 hours.
index (1-RPE2/RPE1), N = number of viable bacteria per cell after incubation with antibiotics. Control experiments to assess the efficacy of antibiotic bactericidal activity were performed in parallel. Briefly, samples of 5 × 10⁶ bacteria were incubated with antibiotics for 30 min at 37°C and plated. This resulted in a >99% decrease in CFU.

Animal experiments

C57BL/6J, B6.129 S-Tnfα−/− (TNF-α−/−), B6 129S7-Rag1−/− (Rag1−/−), C3H/HeJ (TLR4-def) mice were obtained from Jackson laboratories (Bar Harbor). All mice were bred in our Bordetella-free, specific pathogen-free breeding rooms at The Pennsylvania State University. For inoculation, mice were sedated with 5% isoflurane (Abbott laboratory) in oxygen and 50 μl of PBS containing 10⁵ or 5 × 10⁵ CFU of the indicated bacteria were pipetted onto the external nares [76,77]. This method reliably distributes the bacteria throughout the respiratory tract [76]. Survival curves were generated by inoculating TLR4-def, TNF-α−/− and Rag1−/− mice with either RB50 or RB50AsigE. Mice suffering from lethal bordetellosis as determined by severe hunched posture, ruffled fur, extremely labored breathing and apathy were euthanized to prevent unnecessary suffering [47]. For quantifying bacterial load, mice were euthanized via CO₂ inhalation, and lung, trachea, nasal cavity, spleen, liver and/or kidneys were excised. Tissues were homogenized in PBS, aliquots were serially diluted, plated, incubated at 37°C for 2 to 3 days, and CFU were determined. All protocols were reviewed by the university IACUC and all animals were handled in accordance with institutional guidelines (IACUC approval number: 31297).

Statistical analysis

The mean ±/− standard error (SE) of the geometric mean was determined when appropriate and expressed as error bars. Two-tailed, unpaired Student’s T-tests were used to determine statistical significance between groups. All experiments were performed at least twice with similar results.

Authors’ contributions

SB and SA conceived and designed the molecular and stress experiments, which were performed by SB, XZ and EH conceived and designed the infection studies, which were performed by XZ. SH performed the cytotoxicity experiments and MR performed the phagocytosis experiments. SB, XZ, EH, and SA wrote the manuscript. All authors have read, contributed to editing, and approved the final manuscript.

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