The aim of this study was to determine the frequencies and specificities of “enzyme-only” detected red blood cell (RBC) alloantibodies in the routine antibody screening and antibody identification in patients hospitalized in Austria. Routine blood samples of 2420 patients were investigated. The antibody screening was performed with a 3-cell panel in the low-ionic strength saline- (LISS-) indirect antiglobulin test (IAT) and with an enzyme-pretreated (papain) 3-cell panelfully automated on the ORTHO AutoVue Innova System. The antibody identification was carried out manually with an 11-cell panel in the LISS-IAT and with an enzyme-pretreated (papain) 11-cell panel. In total 4.05% (n = 98) of all patients (n = 2420) had a positive RBC antibody screening result. Of them 25.51% (25/98) showed “enzyme-only” detected specific or nonspecific RBC alloantibodies. Rhesus and Lewis system antibodies were found the only specificities of “enzyme-only” RBC alloantibodies: all in all 4.8% (4/98) were detected with anti-E, 3.06% (3/98) with anti-Le a, 3.06% (3/98) with anti-D after anti-D prophylaxis and 1.02% (1/98) with anti-e. In total, 14.29% (14/98) showed a nonspecific RBC alloantibody result with the enzyme test. The results of the present study demonstrate that a high number of unwanted positive reactions with the enzyme technique overshadows the detection of “enzyme-only” RBC alloantibodies. (Trial Registration: K-37-13).

1. Introduction

Pretransfusion blood grouping, red blood cell (RBC) antibody screening, and compatibility testing are essential to prevent incompatible blood transfusion and alloimmunization. The Nobelist Karl Landsteiner, discoverer of the first human marker locus, published the results of a complete cross testing of the RBCs and sera of six people (including himself) in his 1901 paper [1–3]. Since then numerous other human blood group antigens have been described and categorized. Alloimmunization can cause a hemolytic transfusion reaction in individuals lacking the corresponding blood group antigen on their erythrocytes [4, 5]. RBC alloimmunization correlates with the number of transfusions [6–8], and the immunogenicity of the blood group antigens is crucial [5, 9]. About 25–28 antigens are known to cause hemolytic transfusion reactions and should be detected with the pretransfusion RBC antibody screening test [10]. The Rhesus (Rh), Kell (K), Duffy (Fy), and Kidd (Jk) antigens are some of these clinically significant blood group antigens [11].

Pretransfusion compatibility testing involves ABO grouping, Rh typing, RBC antibody screening, RBC antibody identification, and also cross matching the RBC unit designed to be transfused [12]. Hemagglutination is still the classical method for antigen testing and antibody screening [13]. The indirect antiglobulin test (IAT) is considered to be a reliable and effective method to detect clinically relevant
RBC alloantibodies [14]. In the last few years, pretransfusion testing practices have shifted from tube to gel technology. The gel test is more sensitive than the conventional tube method [15, 16]. It has been well known for a long time that the enzyme treatment of RBCs modifies the erythrocyte surface [17–19] and that some Rh antibodies occur only in the enzyme (papain) technique [20, 21]. The main argument for the use of the enzyme technique in the routine testing would be to detect clinically significant RBC alloantibodies, but published works on this topic are rare [22]. In Austria, hospital blood banks without donation, production and screening facilities, and the so-called "blood-depositories" are mainly managed by specialists for anesthesiology, laboratory medicine, transfusion medicine, and internal medicine [23]. Among the blood-depositories, differences of opinion exist regarding the use of enzyme-pretreated RBCs.

The aim of this study was to determine the frequencies and specificities of "enzyme-only" detected erythrocyte alloantibodies in the routine RBC antibody screening and identification in patients hospitalized in Austria.

2. Materials and Methods

The ethical approval for this study was provided by the Ethical Committee of Upper Austria, Linz, Austria (Trial Registration no.: K-37-13). The study period was from January 17, 2013 to May 17, 2013.

2.1. Patient Material. Blood samples of 2420 hospitalized patients, who underwent routine blood grouping and RBC antibody screening at the Department of Laboratory Medicine in the Central Hospital Steyr (Austria), were investigated. The patients were mainly adults and in very few instances children. Ethylenediaminetetraacetic acid (EDTA) plasma was used for the analysis. All the patients were tested for the ABO blood group, the Rh antigen D, and RBC alloantibodies.

2.2. Blood Group Determination. The fully automated ABO/Rh typing was performed using the gel technique on the ORTHO AutoVue Innova System (Ortho Clinical Diagnostics, Raritan, New Jersey). According to the Austrian Guidelines for Blood Group Serology and Transfusion Medicine (latest version July 2000), the RBC antigens A, B, and Rh D, as well as the presence of isoagglutinins, were tested. All patients with a positive result in the RBC antibody screening were tested for the Rh antigens C, c, E, and e.

2.3. RBC Antibody Screening. The RBC antibody screening was carried out in a fully automated process with a 3-cell panel (0.8% Surgiscreen RBCs reagents 1, 2, and 3 [Ortho Clinical Diagnostics, Raritan, New Jersey]) in the low-ionic strength saline- (LISS-) IAT. Additionally an enzyme test with 3 enzyme-pretreated (papain) red test cells (ID-DiaCell PI, PI, PI Rh [BIO-RAD, Hercules, California]) was performed on the ORTHO AutoVue Innova System (Ortho Clinical Diagnostics, Raritan, New Jersey).

2.4. RBC Antibody Identification. If the RBC antibody screening showed a positive result in the LISS-IAT and/or with the enzyme-test (papain), the antibody identification was carried out manually with an 11-cell panel (ID-DiaPanel 1-11 [BIO-RAD, Hercules, California]) in the LISS-IAT and with an enzyme-pretreated (papain) 11-cell panel (ID-DiaPanel P 1-11 [BIO-RAD, Hercules, California]). If a specific RBC alloantibody was identified, the corresponding blood group antigen was determined to confirm the RBC antibody identification result. The autocontrol was made in the LISS-IAT. Serological reactions were graded from 0 to 4+.

2.5. Data Analysis. Descriptive statistics were performed to analyze the serological ABO and Rh phenotype frequencies and the frequencies and specificities of the RBC alloantibodies.

3. Results

3.1. ABO and Rh D Phenotype Frequencies. Of all patients (n = 2420), 38.18% (924/2420) had blood group O, 44.42% (1075/2420) blood group A, and 12.44% (310/2420) blood group B and 4.96% (120/2420) showed the AB phenotype. All in all 84.42% (2043/2420) were Rh antigen D positive and 15.58% (377/2420) were Rh antigen D negative.

3.2. RBC Antibody Screening Results. In total 4.05% (n = 98) of all patients (n = 2420) showed a positive RBC antibody screening result with the 3-cell panel in the LISS-IAT and/or with the enzyme-pretreated (papain) 3-cell panel. Of them 82.65% were women and 17.35% were men. The median age of patients with a positive RBC antibody result was 42 (range: 17–91) years. In total 95.95% (n = 2322) had a negative test result in the LISS-IAT as well as with the enzyme test.

3.3. RBC Antibody Identification Results. The frequencies and specificities of RBC alloantibodies detected in the LISS-IAT ± enzyme (papain) are illustrated in Table I: in total, 66.32% (n = 65) of all patients with a positive RBC antibody screening result (n = 98) presented a specific RBC alloantibody identification result. The most common specific RBC alloantibodies were directed against Rhesus (Rh), Lewis (Le), and Kell (K) antigens. Six patients were found with combined specific RBC alloantibodies: three patients had combined anti-D and anti-C, one patient combined anti-D, anti-C, and anti-K, one patient combined anti-E, anti-K, and anti-Fya, and one patient had combined anti-c, anti-K, and anti-Fyb. Eight patients were found with a nonspecific RBC alloantibody identification in the LISS-IAT ± enzyme (papain). The frequencies and specificities of "enzyme-only" detected RBC alloantibodies are shown in Table 2: all in all eleven patients showed a specific RBC alloantibody result with the enzyme test. The identified "enzyme-only" RBC alloantibodies were Rhesus and Lewis system antibodies. Fourteen patients showed a nonspecific RBC alloantibody result with the enzyme test. All specific and nonspecific RBC alloantibody detection results in the LISS-IAT and with the enzyme test are summarized in Table 3.
Table 1: Frequencies and specificities of RBC alloantibodies detected in the LISS-IAT ± enzyme (papain) (n = 73).

| Antibody-specificities | Frequencies | Percent |
|------------------------|-------------|---------|
| Anti-D after anti-D prophylaxis | 32 | 32.66 |
| Anti-E | 3 | 3.06 |
| Anti-Le\(^a\) | 4 | 4.08 |
| Anti-D | 5 | 5.10 |
| Anti-K | 5 | 5.10 |
| Anti-D and anti-C | 3 | 3.06 |
| Anti-Fy\(^a\) | 2 | 2.04 |
| Anti-Kp\(^b\) | 2 | 2.04 |
| Anti-M | 2 | 2.04 |
| Anti-C\(^w\) | 1 | 1.02 |
| Anti-S | 1 | 1.02 |
| Anti-D, anti-C, and anti-K | 1 | 1.02 |
| Anti-E, anti-K, and anti-Fy\(^a\) | 1 | 1.02 |
| Anti-c, anti-K, and anti-Fy\(^a\) | 1 | 1.02 |
| Anti-I | 1 | 1.02 |
| Anti-II | 1 | 1.02 |
| Nonspecific RBC alloantibodies | 8 | 8.17 |
| Total | 73 | 74.49 |

RBC: red blood cell; Le\(^a\): Lewis\(^a\); K: Kell; Fy\(^a\): Duffy\(^a\); LISS: low-ionic strength saline; IAT: indirect antiglobulin test.

Table 2: Frequencies and specificities of “enzyme-only” detected RBC alloantibodies (n = 25).

| Antibody specificities | Frequencies | Percent |
|------------------------|-------------|---------|
| Anti-E | 4 | 4.08 |
| Anti-Le\(^a\) | 3 | 3.06 |
| Anti-D after anti-D prophylaxis | 3 | 3.06 |
| Anti-e | 1 | 1.02 |
| Nonspecific RBC alloantibodies | 14 | 14.29 |
| Total | 25 | 25.51 |

RBC: red blood cell; Le\(^a\): Lewis\(^a\).

4. Discussion

The ABO and Rh D frequency data of our study population are comparable with the described results in the literature. A study about blood group frequencies in Germany with the data of more than 600,000 donors demonstrated similar results: all in all 41.21% had blood group O, 43.26% blood group A, 10.71% blood group B, and 4.82% blood group AB. 82.71% were Rh D positive and 17.29% were Rh D negative [24].

In the present study 3.06% (n = 3) of all patients with a positive RBC antibody screening result (n = 98) showed an “enzyme-only” anti-Le\(^a\) in the antibody identification. In general Le antibodies are not considered clinically significant [11]. This is probably because the most of the antibodies in the Lewis system are not active at 37°C [34].

In this study a high number of unwanted positive enzyme reactions were detected. In total 14.29% (n = 14) of patients with a positive antibody testing result (n = 98) had a nonspecific RBC alloantibody identification result with the enzyme test. These findings confirm the results of other works on this topic. The detection of clinically significant “enzyme-only” erythrocyte antibodies was overshadowed by the high number of nonspecific or clinically insignificant antibodies [29]. Many specialists working on this as a daily routine are convinced that the enzyme test in the RBC antibody screening is not justified because of the high workload in investigating all the initial positive enzyme-method reactions compared with the low number of clinically significant erythrocyte antibodies identified [32].

The limitation of this study is the low number of patients. In a greater study population more “enzyme-only” RBC
alloantibodies may be detected and the results of the present study could be substantiated.

5. Conclusions

The data presented in this study demonstrate that Rhesus and Lewis system antibodies were the only specificities of "enzyme-only" detected RBC alloantibodies in patients hospitalized in Austria accompanied by a high number of unwanted nonspecific enzyme reactions. Compared to the existing literature the majority of "enzyme-only" detected RBC alloantibodies in patients hospitalized in Austria accompanied by a high number of unwanted positive enzyme-reactions. In clinical practice extra time and additional personnel and labor costs for investigating a high number of unwanted positive enzyme-reactions must be calculated. Based on these facts each hospital "blood-depository" should make its own pragmatic decision whether the enzyme test for routine RBC antibody screening is justified or not in their patient population.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

[1] R. Owen, "Karl Landsteiner and the first human marker locus," Genetics, vol. 155, no. 3, pp. 995–998, 2000.
[2] T. Makarovska-Bojadzieva, M. Blagoevska, P. Kolevski, and S. Kostovska, "Optimal blood grouping and antibody screening for safe transfusion," Prilozi, vol. 30, no. 1, pp. 119–128, 2009.
[3] J. P. Aymard, "Karl Landsteiner (1868–1943) and the discovery of blood groups," Transfusion Clinique Et Biologique, vol. 19, no. 4-5, pp. 244–248, 2012.
[4] D. R. Branch, "Solving the dilemma of prevention of red cell alloimmunization," Immunotherapy, vol. 4, no. 9, pp. 903–905, 2012.
[5] S. Zalpuri, J. J. Zwaginga, and J. G. van der Bom, "Risk Factors for Alloimmunisation after red blood Cell Transusions (RF-FACT): a case cohort study," British Medical Journal Open, vol. 2, no. 3, Article ID e001150, 2012.
[6] V. M. Alves, P. R. Martins, S. Soares et al., "Alloimmunization screening after transfusion of red blood cells in a prospective study," Revista Brasileira De Hematologia e Hemoterapia, vol. 34, no. 3, pp. 206–211, 2012.
[7] R. de Oliveira Cruz, M. A. Mota, F. M. Conti et al., "Prevalence of erythrocyte alloimmunization in polytransfused patients," Einstein, vol. 9, no. 2, pp. 173–178, 2011.
[8] K. M. Byrne, G. S. Booth, J. Y. Lee, and K. R. Ravenell, "The importance of antibody detection and identification in the chronically transfused patient," Laboratory Medicine, vol. 41, no. 5, pp. 261–263, 2010.
[9] C. A. Tormey and G. Stack, "Immunogenicity of blood group antigens: a mathematical model corrected for antibody evanescence with exclusion of naturally occurring and pregnancy-related antibodies," Blood, vol. 114, no. 19, pp. 4279–4282, 2009.
[10] S. Pathak, M. Chandrashekhar, and G. R. Whankhede, "Type and screen policy in the blood bank: is AHG cross-match still required? A study at a multispeciality corporate hospital in India," Asian Journal of Transfusion Science, vol. 5, no. 2, pp. 153–156, 2011.
[11] M. H. Yazer, "The blood bank "black box" debunked: pre-transfusion testing explained," Canadian Medical Association Journal, vol. 174, no. 1, pp. 29–32, 2006.
[12] K. A. Downes and I. A. Shulman, "Pretransfusion testing practices in North America, 2005–2010: an analysis of the College of American Pathologists Interlaboratory Comparison Program J-Survey Data, 2005–2010," Archives of Pathology and Laboratory Medicine, vol. 136, no. 3, pp. 294–300, 2012.
[13] M. E. Reid, "Transfusion in the age of molecular diagnostics," Hematology American Society of Hematology Education Program, pp. 171–177, 2009.
[14] J. Cid, N. Nogués, R. Montero, M. Hurtado, A. Briega, and R. Parra, "Comparison of three microtube column agglutination systems for antibody screening: DGGel, DiaMed-ID and Ortho BioVue," Transfusion Medicine, vol. 16, no. 2, pp. 131–136, 2006.
[15] M. A. Hassan, F. H. Al Gahtani, and A. M. A. Gader, "The frequency of alloantibodies in pregnant and multi-transfused patients—a comparative study between the ID-Micro Typeing Gel System and the conventional tube method," Journal of Applied Hematology, vol. 2, no. 3, pp. 234–242, 2011.
[16] E. Delaflor-Weiss and V. Chizhevsky, "Implementation of gel testing for antibody screening and identification in a community hospital, a 3-year experience," Laboratory Medicine, vol. 36, no. 8, pp. 489–492, 2005.
[17] K. Aho and C. L. Christian, "Studies of incomplete antibodies. I. Effect of papain on red cells," Blood, vol. 27, no. 5, pp. 662–669, 1966.
[18] K. Sagisaka, K. Tokiwa, and N. Yoshioka, "Actions of proteolytic enzymes on agglutinability of M and N blood-group red cells," Tohoku Journal of Experimental Medicine, vol. 106, no. 2, pp. 191–197, 1972.
[19] K. Sagisaka and K. Takahashi, "On the agglutinogens of red cells developed with proteolytic enzymes and neuraminidase," Tohoku Journal of Experimental Medicine, vol. 120, no. 2, pp. 169–175, 1976.
[20] B. E. Dodd and D. A. Eeles, "Rh antibodies detectable only by enzyme technique," Immunology, vol. 4, pp. 337–345, 1961.
[21] F. Gundorf, "Rhesus antibodies demonstrable only by enzyme technique," Vox Sanguinis, vol. 24, no. 4, pp. 380–381, 1973.
[22] A. Pereira, R. Mazzara, A. Gelabert, and R. Castillo, "Two-stage papain-indirect antiglobulin test and LISS direct agglutination are not appropriate for pretransfusion screening for unexpected antibodies," Haematologica, vol. 76, no. 6, pp. 475–478, 1991.

Table 3: RBC alloantibody detection in the LISS-IAT and with the enzyme test.

|                     | LISS-IAT + enzyme | LISS-IAT | “Enzyme-only” |
|---------------------|-------------------|----------|---------------|
| Specific RBC alloantibodies | 60                | 5        | 11            |
| Nonspecific RBC alloantibodies | 7                 | 1        | 14            |

RBC: red blood cell; LISS: low-ionic strength-saline; IAT: indirect antiglobulin test.
[23] H. Schennach, C. Gabriel, D. Schönitzer, and B. Blauhut, "Transfusion medicine in Austria," Transfusion Medicine and Hemotherapy, vol. 33, no. 5, pp. 364–373, 2006.

[24] F. F. Wagner, D. Kasulke, M. Kerowgan, and W. A. Flegel, "Frequencies of the blood groups ABO, Rhemus, D category VI, Kell, and of clinical relevant high-frequency antigens in South-Western Germany," Infusionstherapie und Transfusionsmedizin, vol. 22, no. 5, pp. 285–290, 1995.

[25] S. P. Masouredis, "Pretransfusion tests and compatibility: questions of safety and efficacy," Blood, vol. 59, no. 5, pp. 873–875, 1982.

[26] Q. Wang, Q. Yang, Y. Bai, C. Zhang, Y. Diao, and D. Fang, "Frequency of RBC alloantibodies in Chinese surgical patients," Transfusion Medicine and Hemotherapy, vol. 39, no. 4, pp. 283–286, 2012.

[27] Y. Lin, K. Pavenski, E. Saidenberg, and D. R. Branch, "Blood group antigens and normal red blood cell physiology: a Canadian blood services research and development symposium," Transfusion Medicine Reviews, vol. 23, no. 4, pp. 292–309, 2009.

[28] G. Garratty, M. J. Telen, and L. D. Petz, "Red cell antigens as functional molecules and obstacles to transfusion," Hematology American Society of Hematology Education Program, vol. 2002, no. 1, pp. 445–462, 2002.

[29] D. Castellá, J. Cid, M. Panadés, and C. Martín-Vega, "One thousand seventy antibodies detected only by a 2-stage papain test: wanted and unwanted positive reactions," Immunohematology, vol. 17, no. 4, pp. 122–124, 2001.

[30] J. Conne, P. Schneider, and J.-D. Tissot, "Role of enzyme-treated cells in RBC antibody screening using the gel test: a study of anti-RH1, -RH2, and -RH3 antibodies," Journal of Clinical Laboratory Analysis, vol. 21, no. 2, pp. 61–66, 2007.

[31] M. Pujol, J. M. Sancho, and M. A. Zarco, "The gel enzyme technique in pretransfusion antibody screening," Haematologica, vol. 87, no. 10, pp. 1119–1120, 2002.

[32] P. D. Issitt, M. R. Combs, S. J. Bredehoeft et al., "Lack of clinical significance of “enzyme-only” red cell alloantibodies," Transfusion, vol. 33, no. 4, pp. 284–293, 1993.

[33] M. Contreras, M. De Silva, P. Teesdale, and P. L. Mollison, "The effect of naturally occurring Rh antibodies on the survival of serologically incompatible red cells," British Journal of Haematology, vol. 65, no. 4, pp. 475–478, 1987.

[34] G. Daniels, "ABO, Hh, and Lewis systems," in Human Blood Groups, G. Daniels, Ed., pp. 7–98, Blackwell Science, 2002.