The Effect of Ceftriaxone in Valproic Acid-Induced Mouse Model of Autism

Gamze Gur1, Ruhan Deniz Topuz1,*, Gulnur Kizilay2

1Department of Medical Pharmacology, Faculty of Medicine, Trakya University, 22030-Edirne, Turkey. 2Department of Histology Embryology, Faculty of Medicine, Trakya University, 22030-Edirne, Turkey.

Abstract
Purpose: Autism is a multifactorial neurodevelopment disease and it has not been disclosed as a hypoglumatergic or hyperglutamathergic disease. Ceftriaxone is an antibiotic that increases glutamate transporter-1 (GLT-1) expression in the brain in chronic use. In our study we aimed to investigate the effects of different doses of ceftriaxone in postnatal period in male mice exposed to valproic acid (VPA) at 12.5th day of pregnancy.

Methods: A total of 96 BALB/c male mice were divided into 12 groups (n=8 animals per group). Ceftriaxone (50, 100, 200 mg/kg/d) or saline was given to the male offsprings born from pregnant mice administered VPA and/or saline, between days 47 and 55. Dihydrokainic acid (10 mg/kg), a GLT-1 inhibitor, was administered intraperitoneally to evaluate whether GLT-1 mediates the effect of ceftriaxone. Three chamber sociability and social interaction test and the rota rod test were performed in all groups on days 54 and 55. GLT-1 levels in the hippocampus were measured by immunohistochemistry (IHC) and western blotting (WB).

Results: In our study, autism-like behaviors were observed in male offsprings that were exposed to VPA in the intrauterine period. Chronic ceftriaxone administration has no curative effect on behavioral impairment seen in autism.

Conclusion: Our results show that ceftriaxone did not exert significant therapeutic effect on VPA-induced mouse model of autism.

Introduction
Autism spectrum disorder (ASD) is a common and serious neurodevelopmental disorder characterized by two main symptoms; lack of social communication skills and repetitive behavior. Patients may exhibit various symptoms such as stereotyped or repetitive movements or speech, reduced sharing of interests, emotions or affect, abnormalities in eye contact, insistence on sameness and inflexible adherence to routines. Since ASD is a multisymptomatic disease, it is difficult to diagnose.

The etiology of autism has not been fully elucidated. Prenatal exposure to viruses, toxic substances and various pathogens may cause ASD. Likewise, gestational diabetes, maternal infections (rubella, cytomegalovirus, etc), maternal inflammation and drugs such as valproic acid (VPA), thalidomide, misoprostol, heavy metals and ethanol may be associated with ASD.3,4

Central nervous system develops in between embryonic days 7.5 and 18.5 in mice.5 Autism-like behaviors are reported in mice exposed to different doses of VPA during this period. Offsprings whose mothers received a single dose of 600 mg/kg VPA injection at day 12.5 of gestation are particularly considered to be autistic and used in experimental studies.6 It is not known exactly how VPA causes autistic behaviors. Determination of folic acid metabolism, inhibition of histone deacetylation and increased oxidative stress may play roles in this process.6-8

The pathophysiology of autism is unclear and unbalance of GABA and glutamate ratios may cause ASD.9 In autistic patients, it has been shown that plasma GABA levels are higher and glutamate/GABA ratio is lower.10 Although there are studies reporting the contrary,11 Glutamate is the most important excitatory neurotransmitter in the brain, and glutamate transporters regulate extracellular glutamate homeostasis. Glutamate transporter-1 (GLT-1) is responsible for 90% of glutamate uptake and organizes synaptic transmission of glutamate.12 Beta-lactam antibiotics stimulate GLT-1 expression by increasing the transcription of the GLT-1 gene.13,14 It has been shown that when beta lactam antibiotic, ceftriaxone, administered to animals; expression of GLT-1 in brain and its biochemical and functional activity were increased. Also it exerts neuroprotective effects by stimulating GLT-1 expression.13 In addition we showed that chronic ceftriaxone injection increased GLT-1 expression in hippocampus in mice.15

Based on this information we aimed to investigate the effect of ceftriaxone in male offspring exposed to VPA in the embryonic period and determine whether ceftriaxone-
induced increase in GLT-1 levels in the brain has any effect on behavioral impairment in ASD.

Material and Methods

**Animals**

Male and female BALB/c mice, weighing 20–30 g at the beginning of the experiments, were used in this project. Animals were housed in a quiet room, and water and food were provided *ad libitum*.

**Drugs and experimental procedures**

Mice were mated for 48 hours, and the presence of a vaginal plug was considered as gestational day 0. Pregnant mice received a single intraperitoneal injection of 600 mg/kg VPA (Sigma-Aldrich, China) or saline on gestational day 12.5. Only male offspring born from mothers exposed to saline or VPA were used. The male offspring were separated from the dams at 21st day of life and the rota rod test, three chamber sociability and social interaction test (3 CHM) were performed on postnatal day 54-55.

Ceftriaxone (50–200 mg/kg, i.p.) was administered on postnatal days 47–55 for 8 days. Ceftriaxone (Rocephin, Roche) was diluted from commercial preparations and dissolved in saline. Dihydrokainic acid 10 mg/kg (Sigma Aldrich, USA), a GLT-1 blocker, was co-administered with ceftriaxone 200 mg/kg group. All drugs and saline were administered i.p. in a volume of 0.1 mL/10 g body weight.

After having completed all behavioral tests, mice were euthanized and brain tissues were dissected for western blotting (WB) and immunohistochemistry (IHC) analysis.

**Behavioral Tests**

**Three chamber sociability and social interaction test**

Three chamber sociability and social interaction test (3 CHM) was performed with minimal modification as previously reported. The test apparatus consists of a plexiglas box with a total size of 60 × 40 × 20 cm divided into three chambers. There are openings between the compartments, that allow exploring of all the arena and two wire cages. The test was performed on two days; on first day, mice was free in three chamber boxes for 30 minutes for habituation. On the second day the task consisted of three sessions. The first session was the habituation period for 5 minutes. Subject animal was left in the center area and allowed to roam freely. After habituation, the second session (sociability test) lasted for 10 minutes and a novel animal was introduced in the wire cage of the left compartment. In this part, the time spent in compartments and the sniffing duration of the familiar/unfamiliar animals were calculated in the last part for evaluation of social novelty. All records were analyzed using EthoVision XT 11.5 (Noldus, The Netherlands), Mouse Behaviour Module.

**Rota rod performance test**

A rota rod apparatus (Commat, Ankara, Turkey) was used to evaluate locomotor coordination. The mouse was left on a rotating bar and the time to fall from the bar was recorded. The cut-off time was set to 180 seconds.

**Western blotting**

Western blot analyses were performed as described previously. Briefly, samples were loaded into NuPAGE Novex 4–12% Bis-Tris gel (Life Technologies, Invitrogen, Carlsbad, CA, USA), electrophoretically separated and electroblotted onto a nitrocellulose membrane (The iBlot™ Gel Transfer Stack, Nitrocellulose, Invitrogen). The blocking solution was used for blocking membrane for 30 minutes to reduce nonspecific binding (Western Breeze Chemiluminescent Western Blot Immunodetection Kit Anti-rabbit; Invitrogen). Subsequently, the membranes were incubated overnight at 4°C with rabbit polyclonal EAAT2/GLT1 (1:1000 dilution in blocking solution, Novus Biologicals, Littleton, CO). The membrane was washed two times for 10 minutes with wash buffer and then incubated with secondary antibody for 30 minutes (Western Breeze Chemiluminescent Western Blot Immunodetection Kit Anti-rabbit; Invitrogen). Following washing three times for 10 minutes with wash buffer, the protein was visualized by ChemiDoc™ MP Imaging System (Universal Hood 3; Bio-Rad, Hercules, CA, USA) with enhanced chemiluminescence substrate (Invitrogen).

Then, the same membranes were washed according to the protocol using Restore Western Blot Stripping Buffer (Thermo Fisher Scientific, Fremont, CA). The same membranes were then incubated with rabbit polyclonal β-actin Antibody (Novus Biologicals) at a dilution of 1/1000 in a shaker overnight at + 4°C. The procedures for GLT-1 were repeated in the same order and imaging was performed. Immunoblot bands for GLT-1 and β-actin were quantified using an ImageJ 1.48v program (Wayne Rasband, National Institutes of Health, Bethesda, MD, USA). The ratio of the density of GLT-1 to β-actin was calculated for each sample.

**Immunohistochemistry**

For immunohistochemical examinations, 5 µm thick sections were taken from paraffin-embedded brain tissue samples and incubated overnight in a 56°C oven. Subsequently, deparaffinization with toluene (Merck Millipore, Darmstadt, Germany) and rehydration with graded series of ethanol (70-100%) were boiled in citrate buffer (pH 6, Invitrogen) for antigen recovery. The sections were then exposed to H2O2 (Abcam, Cambridge, USA) to remove endogenous peroxidase activity. In order to prevent nonspecific binding, the
blocking antibody (Invitrogen) was used for 10 minutes. The incubated sections were incubated overnight at +4°C in EAAT2/GLT1 antibody (Novus Biologicals) prepared with antibody dilution solution (Invitrogen) at room temperature. Biotinylated secondary antibody (Invitrogen) against the species from which the primary antibody is produced is administered for 10 minutes. and finally treated with HRP-streptavidin (Invitrogen) for 10 minutes. Staining reactions were visualized using DAB (Invitrogen), then counterstained with hematoxylin. Sections were examined using a ×200 objective on a BX-51 Olympus microscope (Olympus, Tokyo, Japan) and photographed. EAAT2/GLT1 immunoreactivity were semi-quantitatively evaluated using a histological score (HSCORE) value, average score was used for statistical analysis, described previously.17

Statistical analysis

Three chamber sociability and social interaction test, the rota rod test, immunohistochemical examination and WB results were analyzed using t test in two groups comparisons, while one-way variance analysis (ANOVA) and post hoc Bonferroni test was used in multiple comparisons. The analyzes were performed on GraphPad Prism 6.0 for Mac OS X software and P < 0.05 was considered significant.

Results

In this study, the effects of ceftriaxone (50, 100, 200 mg/kg) were evaluated on the VPA-induced mouse model of autism. In addition, changes in GLT-1 levels in the hippocampus were evaluated by IHC and WB analysis. Rota rod test, three chamber sociability and social interaction test were performed in all groups on days 54 and 55. The data obtained from the study will be presented in four sections:

Effect of prenatal VPA administration

In the VPA exposed offsprings the time spent near the full cage was significantly lower than the saline group (P = 0.0229; unpaired t test), but no difference was observed in the sniffing times of empty or full cages (P = 0.2959; unpaired t test) in the sociability phase of 3 CHM test. In VPA exposed group, sniffing time of the full cage was also lower compared to sniffing time of empty cage (P = 0.0324; paired t test) (Figure 1A and 1B). In the social interaction phase the time spent near the unfamiliar animal (P = 0.0375; unpaired t test) and the sniffing time of the unfamiliar animal (P = 0.0335; unpaired t test) were lower in the VPA exposed offsprings compared to saline groups; sniffing duration of the unfamiliar animal was also lower compared to familiar animal in VPA exposed group (P = 0.0243; paired t test) (Figure 1C and 1D). There were no differences between the groups in the rota rod test (data not shown).

Effects of ceftriaxone on intrauterine saline exposed offsprings

There were no differences in the time spent near full/empty cage among groups in the first part of the 3 CHM. In this stage of the experiment, the time spent near the full cage were higher than the empty cage in the 50 mg/kg and 100 mg/kg cef groups. However, ceftriaxone decreased sniffing times of the empty cage at all doses used (P = 0.0003; ANOVA). In the second part of the test ceftriaxone had no effect in any parameter, however the time spent with the unfamiliar mice was greater than the time spent with the familiar mice in the 50 mg/kg cef

Figure 1. Effects of intrauterine VPA injection in three chamber sociability and social interaction test. (A) Time spent near the empty/full cage; (B) Total sniffing duration of the empty/full cage; (C) Time spent near the familiar/unfamiliar animal; (D) Total sniffing duration of the familiar/unfamiliar animal. (*P < 0.05; ** P < 0.01 vs saline exposed offsprings, unpaired t test, # P < 0.05 vs same group, paired t test. Vertical lines indicate SEM)
group \((P=0.0108\) paired \(t\) test). The time spent on rod was not different among groups (data not shown).

**Effects of postnatal ceftriaxone treatment on intrauterine VPA exposed offsprings**

In the sociability phase of 3 CHM test, there were no difference in the time spent near the full/empty cages, but sniffing times of the empty cage was increased in 50 mg/kg ceftriaxone group \((P=0.0506\); ANOVA). There was also a difference in terms of the time spent in the near full/empty cage in the control group \((P=0.0487\); paired \(t\) test) (Figure 2A and 2B). In the social novelty phase, there were no significant differences in the time spent near the full/empty cage or sniffing time of the familiar/unfamiliar animals at any doses of ceftriaxone compared to saline group (Figure 2C and 2D). Ceftriaxone had no effect in the time spent on rod in the rotarod test (data not shown).

**Effects of postnatal dihydrokainic acid injection on intrauterine VPA exposed offsprings**

There was an increase in the dihydrokainic acid + 200 mg/kg cef group in terms of the time spent near the full cage (Figure 3A and 3B, \(P=0.0304\), ANOVA) In the second part of 3 CHM, no difference was observed in any parameters (Figure 3C and 3D).

**Western blotting and immunohistochemistry results**

GLT-1 levels were higher in the VPA exposed offsprings than the saline exposed groups in WB \((P<0.0001\); unpaired \(t\) test), but there were no differences between these groups.
in IHC (Table 1, Figure 4A and Figure 5).

In WB, GLT-1 level was higher in 200 mg/kg chronic ceftriaxone group compared to control in saline exposed offspring ($P < 0.0001$; ANOVA) (Figure 5D); however, there were no differences in IHC findings (Table 1 and Figure 5).

GLT-1 level did not change in VPA exposed offsprings in WB (Figure 5B), but were higher than control groups in 200 mg/kg ceftriaxone group in IHC ($P = 0.0001$; ANOVA) (Table 1 and Figure 5).

WB (Figure 4C) and IHC data (Table 1 and Figure 5) show that dihydrokainic acid had no effect on GLT-1 level.

### Discussion

In this study, we aimed to investigate the effect of ceftriaxone, a beta lactam antibiotic, in VPA-induced mouse model of autism. Autism-like abnormal behavior was induced with a single injection (600 mg/kg) of VPA on the 12.5th day of gestation. Increased doses of ceftriaxone treatment had no effect on autism-like abnormal behavior. GLT-1 levels were increased in 200 mg/kg ceftriaxone groups in intrauterine saline exposed offsprings, but this increase did not lead to any behavioral changes.

Autism is a common neurodevelopmental disease; however, the pathophysiological process leading to autism is not clear. Abnormalities in many neurotransmitters such as GABA and glutamate have been observed in ASD, and these results have been directly associated with the pathogenesis of ASD. Plasma GABA concentration was higher and glutamate/GABA ratio was lower in autistic individuals compared to normal.10 In contrast, other studies have shown a decrease in GABA and GABA/glutamate ratios in frontal lobe imaging in autistic individuals.11 Zhang et al18 declared that bumetanide treatment caused a decrease in the symptoms of individuals with autism by decreasing the GABA/glutamate ratio. Today, there are two theories that explain the relationship between autism and glutamate. The first theory is introduced by Carlsson in 1998 and suggests that autism is a hypoglutamatergic disease. Carlsson attributed this theory to the presence of autism-like symptoms in healthy people who were treated with NMDA antagonists and to the detection of brain damage in glutamate-rich neurons in autism.19 According to the second theory, autism is a hyperglutamatergic disease.20 Fatemi et al20 explained his theory with three findings; serum glutamate levels of individuals with autism are higher than normal individuals; the glutamic acid decarboxylase enzyme, which is responsible for the destruction of glutamate, is reduced in the brain tissues of autistics and glial fibrillary acidic protein, a marker of gliosis in the brain tissues and astroglial-microglial

### Table 1. Immunoreactivity of GLT-1 in all groups (mean ± SD)

| Groups               | GLT-1 immunoreactivity |
|----------------------|------------------------|
| Saline (A)           | 160.0 ± 15.58          |
| Vpa (B)              | 155.0 ± 15.21          |
| Vpa+Saline (C)       | 175.6 ± 7.763          |
| Vpa+50 mg/kg cef (D) | 156.9 ± 18.50          |
| Vpa+100 mg/kg cef (E)| 175.6 ± 30.76          |
| Vpa+200 mg/kg cef (F)| 216.9 ± 26.04          |
| Vpa+dhc acid (G)     | 187.9 ± 11.85          |
| Vpa+dhc acid+200 mg/kg cef (H)| 192.5 ± 16.04 |
| Saline+saline (I)    | 186.9 ± 22.19          |
| Saline+50 mg/kg cef (J)| 158.1 ± 20.69         |
| Saline+100 mg/kg cef (K)| 191.9 ± 27.25         |
| Saline+200 mg/kg cef (L)| 191.3 ± 18.08         |

Figure 4. Levels of GLT-1 in western blot. (**P < 0.0001 vs control unpaired t test, vertical lines indicate SEM)
activation, levels are higher in autistic individuals. Astroglia take glutamate from both the extracellular domain and synthesize glutamate from glutamine.\textsuperscript{20,22} Glutamate is a very important neurotransmitter for regulation of behavior; it is released from synaptic terminals and removed by glutamate transporters, especially GLT-1, in the hippocampus.\textsuperscript{23} GLT-1 plays a key role in regulation of glutamate concentration and disorders of its amount and function can cause abnormal behaviors.\textsuperscript{24} Recent studies have shown that chronic use of ceftriaxone, a beta-lactam antibiotic, increases the expression of GLT-1 depending on the dose.\textsuperscript{25} The inducing effect of ceftriaxone on GLT-1 expression has also been shown by our group.\textsuperscript{15} Moreover many beta-lactam antibiotics, such as ampicillin, cefazolin, and cefoperazone exert increase in GLT-1 expression and enhance its function.\textsuperscript{26}

In the present study, we predicted that if autism is a hyperglutamatergic disease; increased GLT-1 levels, induced by ceftriaxone, can ameliorate the behavioral symptoms of autism. However, we showed that GLT-1 levels was higher in VPA exposed offsprings without ceftriaxone treatment. Studies indicate that VPA increases the mRNA of EATTs in astrocytes and oligodendrocytes. VPA makes this effect via inhibition of histone deacetylase enzyme and increases the expression of not only GLT-1 but also other glutamate transporters.\textsuperscript{27} VPA increases GLT-1 expression in the hippocampus and cortex but decreases the expression in the cerebellum.\textsuperscript{28} There were increased GLT-1 levels both in VPA exposed offspring and saline treated groups in WB. We showed for the first time that VPA applied in the intrauterine period may lead to increase in GLT-1 levels in the postnatal period. Postnatal ceftriaxone treatment on day 47-55 did not increase GLT-1 levels in VPA-induced autistic mice; however, GLT-1 levels were increased in saline exposed 200 mg/kg ceftriaxone treated group in WB. This increase may also be due to elevation in GLT-1 levels by VPA exposure in the prenatal period.

In this study, chronic ceftriaxone treatment did not exert any effect in VPA exposed offspring. There is no research on the effect of ceftriaxone in autism model of animals; however, a case report reported that beta lactam, cefixime, treatment reduced aggressive behaviors in an autistic child.\textsuperscript{29} Although the improvement in aggressive behavior was explained by the drug interaction of beta cefixime, which increases the antiepileptic drug level.\textsuperscript{30} This case report was one of the important reasons for starting this research project.

In our study, dihydrokainic acid, a GLT-1 inhibitor, was administered to evaluate whether ceftriaxone effect was related with GLT-1. In the first part of 3 CHM test, sniffing time of the empty/full cage was decreased when dihydrokainic acid was administered alone and in combination with ceftriaxone (200 mg/kg); this effect can be explained as decreased sociability. However, there was a decrease in sniffing time of the unfamiliar animal in the combination group in the second part of 3 CHM, but there was no increase in sniffing time of the familiar animals; this effect can also be speculated as decrease in sociability. It was reported that dihydrokainic acid injection into the central nucleus of amygdala elicited increased anxiety and depressive behavior in normal rats,\textsuperscript{21} but there is no
research yet on effects of dihydrokainic acid in autism.

In our study, GLT-1 expression was evaluated using WB and IHC analysis. In WB, there was a significant increase in GLT-1 levels in VPA exposed offspring. Recent research indicates that VPA increase GLT-1 levels due to its epigenetic effects. However, no study examined the effect of a single dose of VPA administered in the intrauterine period on GLT-1 levels in the postnatal period. Our research showed that the enhancing effect of VPA on GLT-1 levels in the intrauterine period continued in the postnatal period; however, we did not observe this increase in IHC analysis. It is remarkable that there is no correlation in the results of WB and IHC. The difference between these two analyses may be due to the difference between sampling methods. Punch biopsy was taken randomly from the hippocampus for WB analysis, whereas IHC was performed separately in different regions of the hippocampus. In addition, four out of eight animals in each group were allocated to the right and the other four to the left hemispheres for WB and IHC. The difference in outcome may be due to differences in right and left hemispheres in autism. Concerning our future research our expectation is that removing hippocampus totally and performing analyses in the same hemisphere will ensure the reliability of our results.

Two theories can be developed by evaluating the results of our study. First, if VPA injection during the intrauterine period causes autism-like behavior because of the increase in GLT-1 levels, it can be a hypoglutamatergic disease. This is because increased GLT-1 will lead to decrease in glutamate concentration in the synaptic cleft. However, the question that should be kept in mind is whether the effect a single VPA exposure during the intrauterine period will continue for so long. The second theory is that autism is a hyperglutamatergic disease and GLT-1 levels increase as a compensation mechanism. If autism is a hyperglutamatergic disease, drugs that increase GLT-1 levels such as ceftriaxone may provide a hope for treatment of autism. However, to support these theories, studies on different autism models are needed.

Conclusion
Our results show that ceftriaxone did not exert significant therapeutic effect on VPA-induced mouse model of autism. But our study has some limitations so different studies are needed.

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Ethical Issues
The experimental protocol of this study was approved by the local ethics committee. (Protocol no: TUHADYEK-2016/43).

Conflict of Interest
The authors have no conflicts of interest to declare.

References
1. Bhandari R, Palival JK, Kuhad A. Neuropsychopathology of autism spectrum disorder: complex interplay of genetic, epigenetic, and environmental factors. *Adv Neurobiol* 2020;24:97-141. doi: 10.1007/978-3-030-30402-7_4
2. Chaste P, Leboyer M. Autism risk factors: genes, environment, and gene-environment interactions. *Dialogues Clin Neurosci* 2012;14(3):281-92. doi: 10.31887/DCNS.2012.14.3/pchaste
3. Dufour-Rainfray D, Vourc’h P, Tourrette S, Guilloteau D, Chalon S, Andres CR. Fetal exposure to teratogens: evidence of genes involved in autism. *Neurosci Biobehav Rev* 2011;35(5):1254-65. doi: 10.1016/j.neubiorev.2010.12.013
4. Ornoy A, Weinstein-Fudim L, Ergaz Z. Genetic syndromes, maternal diseases and antenatal factors associated with autism spectrum disorders (ASD). *Front Neurosci* 2016;10:316. doi: 10.3389/fnins.2016.00316
5. Chen VS, Morrison JP, Southwell MF, Foley JF, Bolon B, Elmore SA. Histology atlas of the developing prenatal and postnatal mouse central nervous system, with emphasis on prenatal days E7.5 to E18.5. *Toxicol Pathol* 2017;45(6):705-44. doi: 10.1177/0192623317728134
6. Kataoka S, Takuma K, Hara Y, Maeda Y, Ago Y, Matsuda T. Autism-like behaviours with transient histone hyperacetylation in mice treated prenatally with valproic acid. *Int J Neuropsychopharmacol* 2013;16(10):91-103. doi: 10.1017/s1461145711001714
7. de Theije CG, Koelink PJ, Korte-Bouws GA, Lopes da Silva S, Korte SM, Olivier B, et al. Intestinal inflammation in a murine model of autism spectrum disorders. *Brain Behav Immun* 2014;37:240-7. doi: 10.1016/j.bbi.2013.12.004
8. Nicolini C, Fahnestock M. The valproic acid-induced rodent model of autism. *Exp Neurol* 2018;299(Pt A):217-27. doi: 10.1016/j.expneurol.2017.04.017
9. Horder J, Petrinovic MM, Mendez MA, Buns A, Takumi T, Spooren W, et al. Glutamate and GABA in autism spectrum disorder—a translational magnetic resonance spectroscopy study in man and rodent models. *Transl Psychiatry* 2018;8(1):106. doi: 10.1038/s41398-018-0155-1
10. El-Ansary A, Al-Ayadhli L. GABAergic/glutamatergic imbalance relative to excessive neuroinflammation in autism spectrum disorders. *J Neuroinflammation* 2014;11:189. doi: 10.1186/s12974-014-0189-0
11. Harada M, Taki MM, Nose A, Kubo H, Mori K, Nishitani H, et al. Non-invasive evaluation of the GABAergic/glutamatergic system in autistic patients observed by MEGA-editing proton MR spectroscopy using a clinical 3 tesla instrument. *J Autism Dev Disord* 2011;41(4):447-54. doi: 10.1007/s10803-010-1065-0
12. Danbolt NC. Glutamate uptake. *Prog Neurobiol* 2001;65(1):1-105. doi: 10.1016/S0301-0082(00)00067-8
13. Rothstein JD, Patel S, Regan MR, Haenggeli C, Huang YH, Bergles DE, et al. Beta-lactam antibiotics offer neuroprotection by increasing glutamate transporter expression. *Nature* 2005;433(7021):73-7. doi: 10.1038/nature03180
14. Su ZZ, Leszczy nieceka M, Kang DC, Sarkar D, Chao W, Volsky DJ, et al. Insights into glutamate transport regulation in human astrocytes: cloning of the promoter for excitatory amino acid transporter 2 (EAAT2). *Proc Natl Acad Sci U S A* 2003;100(4):1955-60. doi: 10.1073/pnas.0313655100
15. Karaman I, Kizilay-Ozfidan G, Karadag CH, Ulugol A. Lack of effect of ceftriaxone, a GLT-1 transporter activator, on spatial memory in mice. *Pharmacol Biochem Behav* 2013;108:61-5. doi: 10.1016/j.pbb.2013.04.013
16. Kaidanovich-Beilin O, Lipina T, Vukobradovic I, Roder J, Woodgett JR. Assessment of social interaction behaviors. *J Vis Exp* 2011(48):2473. doi: 10.3791/2473
17. Kizilay G, Cakmak H, Yen CF, Atabekoglu C, Arici A, Kayisli...
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UA. Expression and regulation of c-Jun N-terminal kinase (JNK) in endometrial cells in vivo and in vitro. *Histocherm Cell Biol* 2008;130(4):761-71. doi: 10.1007/s00418-008-0421-z

18. Zhang L, Huang CC, Dai Y, Luo Q, Ji Y, Wang K, et al. Symptom improvement in children with autism spectrum disorder following bumetanide administration is associated with decreased GABA/glutamate ratios. *Transl Psychiatry* 2020;10(1):9. doi: 10.1038/s41398-020-0692-2

19. Carlsson ML. Hypothesis: is infantile autism a hypoglutamatergic disorder? Relevance of glutamate - serotonin interactions for pharmacotherapy. *J Neural Transm (Vienna)* 1998;105(4-5):525-35. doi: 10.1007/s007020050076

20. Fatemi SH, Halt AR, Stary JM, Kanodia R, Schulz SC, Realmuto GR. Glutamic acid decarboxylase 65 and 67 kDa proteins are reduced in autistic parietal and cerebellar cortices. *Biol Psychiatry* 2002;52(8):805-10. doi: 10.1016/s0006-3223(02)02143-0

21. Shinohe A, Hashimoto K, Nakamura K, Tsuji M, Iwata Y, Tsuchiya KJ, et al. Increased serum levels of glutamate in adult patients with autism. *Prog Neuropsychopharmacol Biol Psychiatry* 2006;30(8):1472-7. doi: 10.1016/j.pnpbp.2006.06.013

22. Laurence JA, Fatemi SH. Glial fibrillary acidic protein is elevated in superior frontal, parietal and cerebellar cortices of autistic subjects. *Cerebellum* 2005;4(3):206-10. doi: 10.1080/14734220500208846

23. Lehre KP, Danbolt NC. The number of glutamate transporter subtype molecules at glutamatergic synapses: chemical and stereological quantification in young adult rat brain. *J Neurosci* 1998;18(21):8751-7. doi: 10.1523/jneurosci.18-21-08751.1998

24. Mei YY, Wu DC, Zhou N. Astrocytic regulation of glutamate transmission in schizophrenia. *Front Psychiatry* 2018;9:544. doi: 10.3389/fpsyt.2018.00544

25. Yan H, Li CM, Li YL, Gong ZH. [Effect of spinal glutamate transporter 1 on chronic constriction injury of sciatic nerve and morphine tolerance of rats]. *Yao Xue Xue Bao* 2009;44(6):581-5.

26. Rao PS, Goodwani S, Bell RL, Wei Y, Boddu SH, Sari Y. Effects of ampicillin, cefazolin and cefoperazone treatments on GLT-1 expressions in the mesocorticolimbic system and ethanol intake in alcohol-prefering rats. *Neuroscience* 2015;295:164-74. doi: 10.1016/j.neuroscience.2015.03.038

27. Martínez-Lozada Z, Guillem AM, Robinson MB. Transcriptional regulation of glutamate transporters: from extracellular signals to transcription factors. *Adv Pharmacol* 2016;76:103-45. doi: 10.1016/bs.pharma.2016.01.004

28. Perisic T, Holsboer F, Rein T, Zschocke J. The CpG island shore of the GLT-1 gene acts as a methylation-sensitive enhancer. *Glia* 2012;60(9):1345-55. doi: 10.1002/glia.22353

29. Ghanizadeh A, Berk M. Beta-lactam antibiotics as a possible novel therapy for managing epilepsy and autism, a case report and review of literature. *Iran J Child Neurol* 2015;9(1):99-102.

30. Zhu L, Wang H. Commentary on “beta-lactam antibiotics as a possible novel therapy for managing epilepsy and autism.” *Iran J Child Neurol* 2018;12(3):139-40.

31. John CS, Sypek EI, Carlezon WA, Cohen BM, Öngür D, Bechtholt AJ. Blockade of the GLT-1 transporter in the central nucleus of the amygdala induces both anxiety and depressive-like symptoms. *Neuropsychopharmacology* 2015;40(7):1700-8. doi: 10.1038/npp.2015.16