Neuronal Excitability

Sex Differences in the Alcohol-Mediated Modulation of BLA Network States

Alyssa DiLeo,1 Pantelis Antonoudiou,2 Spencer Ha,2 and Jamie L. Maguire2

https://doi.org/10.1523/ENEURO.0010-22.2022

1Program in Neuroscience, Graduate School of Biomedical Sciences, Tufts University, Boston, Massachusetts 02111 and 2Department of Neuroscience, Tufts School of Medicine, Tufts University, Boston, Massachusetts 02111

Abstract

Alcohol use, reported by 85% of adults in the United States, is highly comorbid with mood disorders, like generalized anxiety disorder and major depression. The basolateral amygdala (BLA) is an area of the brain that is heavily implicated in both mood disorders and alcohol use disorder. Importantly, the modulation of BLA network/oscillatory states via parvalbumin (PV)-positive GABAergic interneurons has been shown to control the behavioral expression of fear and anxiety. Further, PV interneurons express a high density of δ subunit-containing GABA<sub>A</sub> receptors (GABA<sub>A</sub>Rs), which are sensitive to low concentrations of alcohol. Therefore, we hypothesized that the effects of alcohol may modulate BLA network states that have been associated with fear and anxiety behaviors via δ-GABA<sub>A</sub>Rs on PV interneurons in the BLA. Given the impact of ovarian hormones on the expression of δ-GABA<sub>A</sub>Rs, we also examined the ability of alcohol to modulate local field potentials in the BLA from male and female C57BL/6J and Gabrd<sup>−/−</sup> mice after acute and repeated exposure to alcohol. Here, we demonstrate that acute and repeated alcohol can differentially modulate oscillatory states in male and female C57BL/6J mice, a process that involves δ-GABA<sub>A</sub>Rs. This is the first study to demonstrate that alcohol is capable of altering network states implicated in both anxiety and alcohol use disorders.

Key words: alcohol; basolateral amygdala; GABA; network states; oscillations; sex differences

Significance Statement

Alcohol use disorder and mood disorders are highly comorbid. The basolateral amygdala (BLA) is implicated in both disorders, but the mechanisms contributing to their shared pathophysiology remain uncertain. Here we demonstrate that acute and repeated alcohol exposure can alter network oscillations in the BLA that control the behavioral expression of fear and anxiety. These data suggest that alcohol may directly influence network states associated with mood. Further, we demonstrate sex differences in the ability of alcohol to modulate BLA network states, an effect involving δ-GABA<sub>A</sub> receptors, which may contribute to sex differences in alcohol intake and comorbid mood disorders. These data potentially point to a novel mechanism mediating the effects of alcohol on affective states.

Introduction

Alcohol is the most widely used drug in the United States, with ~85% of adults reporting alcohol use in their lifetime. Despite this high rate of use, only ~5% will go on to develop an alcohol use disorder while most adults continue to drink without reaching this diagnostic criterion (Substance Abuse and Mental Health Services Administration, 2020). The transition from first drink to alcohol dependence is encouraged by both the positive and negative reinforcing effects of alcohol, each with corresponding neurobiological frameworks (Gilpin and Koob, 2008). Comorbid mood disorders, such as major...
depression and anxiety disorders, contribute to the reinforcing effects of alcohol by pushing individuals to drink to relieve tension in high-stress or high-anxiety situations (Kushner et al., 2011). The basolateral amygdala (BLA) has been identified as a brain region contributing to both alcohol use disorder and anxiety disorders (Silberman et al., 2009; Tye et al., 2011; Agoglia and Herman, 2018).

Accumulating evidence demonstrates a critical role for oscillatory states in the BLA in modulating fear and anxiety-like behaviors (Likhitik et al., 2014; Stujenske et al., 2014; Davis et al., 2017; Antonoudiou et al., 2021). However, the impact of alcohol on these network states has not been explored. Network oscillations within and between brain areas represent a mechanism for the transition between brain and behavioral states. Specifically, particular oscillation frequencies within and between the BLA and medial prefrontal cortex (mPFC) are associated with either a fear (3–6 Hz) or safety (6–12 Hz) state (Davis et al., 2017). This circuit, along with other regions like the hippocampus, has also been shown to contribute to high- and low-anxiety states in mice (Likhitik et al., 2014).

It is well established that the anxiolytic properties of alcohol can motivate consumption and contribute to the high comorbidity of alcohol use disorders and mood disorders (Thomas et al., 2003; Smith and Randall, 2012; Mason et al., 2019). However, it is unclear how alcohol impacts network states underlying modulation of anxiety states. Here we examine the ability of acute, low-dose alcohol to modulate BLA network activity in alcohol-naive mice, using local field potentials (LFPs) to measure network oscillations in the BLA in male and female C57BL/6J mice during acute and repeated exposures to alcohol.

The generation of oscillations is thought to involve the ability of GABAAergic interneurons, particularly parvalbumin (PV)-expressing interneurons, to synchronize populations of principal neurons (Bartos et al., 2007; Sohal et al., 2009; Fuchs et al., 2017). Somatic-targeting, fast-spiking PV interneurons exert powerful control over a large network of excitatory principal cells, and, as such, are capable of generating and synchronizing oscillations to orchestrate network communication (McDonald, 1992; Bocchio and Capogna, 2014). There is a critical role for PV interneurons in oscillation generation within the BLA both ex vivo and in vivo, where PV interneurons can shift oscillatory frequencies and drive behavioral states (Davis et al., 2017; Ozawa et al., 2020; Antonoudiou et al., 2021).

PV interneurons in the BLA express a high density of extrasynaptic δ subunit-containing GABA<sub>A</sub> receptors (GABA<sub>A</sub>Rs), which are uniquely sensitive to alcohol and play a role in regulating both alcohol consumption and anxiety-like behaviors, including anxiety associated with alcohol withdrawal (Glykys et al., 2007; Melón et al., 2019; Antonoudiou et al., 2021). Tonic inhibition mediated by δ-GABA<sub>A</sub>Rs has been shown to control hippocampal oscillations (Mann and Mody, 2010; Pavlov et al., 2014) and loss of the δ subunit in PV interneurons alters gamma oscillations in the CA3 region of the hippocampus (Ferando and Mody, 2013, 2014). Given the evidence that PV interneurons, which modulate oscillations in the BLA, have a high density of δ subunit expression, we further hypothesized that alcohol acts via δ-GABA<sub>A</sub>Rs in the BLA to modulate oscillations associated with the network communication of fear and anxiety. To test this, we examined the ability of alcohol to alter oscillatory states in the BLA of male and female Gabrd<sup><s>−/−</s></sup> mice. Our findings suggest that the ability of alcohol to modulate network states involves δ subunit-containing GABA<sub>A</sub>Rs. We conclude that alcohol can modulate BLA oscillatory states in a sex-specific manner, a process which, in part, involves δ subunit-containing GABA<sub>A</sub>Rs.

Materials and Methods

Animals

Adult male and female C57BL/6J mice, 8–12 weeks of age, were purchased from The Jackson Laboratory (stock #000664) and group housed in temperature- and humidity-controlled housing rooms on a 12 h light/dark cycle (lights on at 7:00 A.M.) with ad libitum food and water. Animals were handled according to protocols and procedures approved by the Tufts University Institutional Animal Care and Use Committee. Female mice are maintained in an acyclic state without exposure to males. Global Gabrd<sup><s>−/−</s></sup> knock-out mice were bred in house (Mihalek et al., 1999, 2001). Mice were single housed and habituated to new cages for 24 h before the start of experiments.

Stereotaxic surgery

All mice undergoing surgery were anesthetized with ketamine/xylazine (90 and 5–10 mg/kg, i.p., respectively) and treated with sustained release buprenorphine (0.5–1.0 mg/kg, s.c.). A lengthwise incision was made to expose the skull and a unilateral craniotomy was performed to lower a depth electrode [paraformaldehyde (PFA)-coated stainless steel wire, A-M Systems] into the BLA (anteroposterior (AP), −1.50 mm; mediolateral (ML), −3.30 mm; dorsoventral (DV), −5 mm), affixed to a head mount (catalog #8201, Pinnacle) with stainless steel screws as ground, reference, and frontal cortex EEG (AP, +0.75 mm; ML, ±0.3 mm; DV, −2.1 mm) electrodes. EMG wires were positioned in the neck muscles.

LFP recordings

LFP recordings were performed in male and female C57BL/6J and Gabrd<sup><s>−/−</s></sup> mice after a week of recovery from implant surgery. LFP recordings were acquired

The authors are supported by National Institutes of Health Grants F31-AA-028410, R01-AA-026256, R01-NS-105628, R01-NS-102937, R01-NS-105628, and R01-MH-122379.

Acknowledgment: We thank Dr. Klaus Miczek, Dr. Laverne Melón, and Dr. Leon Reijmers for feedback and guidance on this project. The visual abstract created with BioRender.com.

Correspondence should be addressed to Jamie L. Maguire at jamie.maguire@tufts.edu.

https://doi.org/10.1523/ENEURO.0010-22.2022

Copyright © 2022 DiLeo et al.

This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license, which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.
using Lab Chart software (AD Instruments) collected at 4 kHz and amplified 100×. Spectral analysis was performed in MATLAB (Antonoudiou et al., 2021) using MatWAND (https://github.com/pantelisantonoudiou/MatWAND), which uses the fast Fourier transform, similar to previous reports (Kruse and Eckhorn, 1996; Frigo and Johnson, 1998; Pape et al., 1998; Freeman et al., 2000). Briefly, recordings were divided into 5 s overlapping segments and the power spectral density for a range of frequencies was obtained (Oppenheim et al., 1999). LFP power was quantified as a power area.

Acute and repeated alcohol exposure
Mice were habituated to new cages with ad libitum food and water for 24 h before starting the experimental paradigm. All injections were performed 2–3 h into the light cycle (on at 7:00 A.M.) at the same time each day across all cohorts. A dose response of four doses (0.5, 1.0, 1.5, and 2.0 g/kg, i.p.) was completed in a group of male C57BL/6J mice to determine the appropriate dose to use in our experiments. The acute exposure consisted of a 60 min baseline period followed by a saline injection (0.9% NaCl, i.p.) and a subsequent 1 g/kg ethanol injection [20% (v/v), i.p.]. The repeated exposure consisted of a 60 min baseline period followed by an intraperitoneal injection of either saline or 1 g/kg ethanol [20% (v/v)] for 5 consecutive days. For the females, the acute ethanol exposure was calculated from the first day of the repeated exposure paradigm.

Blood ethanol concentration measurements
Blood from the submandibular vein was collected from a separate cohort of male and female C57BL/6J mice 15 min after exposure to alcohol (1 g/kg, i.p.) on days 1, 2, and 5 of repeated alcohol exposure. Blood was spun down at 1.8 × g for 15 min at 4°C, and serum was stored at −80°C until blood ethanol concentration (BEC) measurements were performed using the BioAssay Systems EnzyChrom Ethanol Assay Kit (ECET-100) according to the manufacturing protocol. Measurements are reported in milligrams per deciliter.

Immunohistochemistry
Immunohistochemistry was performed as previously reported (Melón et al., 2019) in a separate cohort of C57BL/6J mice 30 min following repeated exposure to vehicle or alcohol for 5 d. Mice were anesthetized with isoflurane, transcranially perfused with 0.9% saline and 4% PFA, and subsequently cryoprotected in 10% and 30% sucrose. The brains were then flash frozen using isopentane and stored at −80°C until cryosectioning. Free-floating 40 μm coronal slices were costained for PV and δ using universal antigen retrieval buffer (catalog #CTS015, R&D Systems) and primary antibodies against δ-GABAAR (1:100; catalog #868A-GDN, Phosphosolutions) and PV (1:1000; catalog #P3088, Sigma-Aldrich) for 72 h at 4°C. The slices were then incubated with a biotinylated goat anti-rabbit (1:1000; catalog #BA1000, Vector Laboratories) and Alexa-Fluor 488-conjugated goat anti-mouse (1:200; catalog #A28181, Thermo Fisher Scientific) for 2 h at room temperature and streptavidin-conjugated Alexa-Fluor 488 (1:200; catalog #S3254, Thermo Fisher Scientific) for 2 h at room temperature. Slices were mounted and coverslipped with antifade hard set mounting medium with DAPI (VECTASHIELD, catalog #H1500, Vector Laboratories). Fluorescent labeling in the BLA was imaged on a Nikon A1R confocal microscope, and z-stacks were acquired using a 20× objective. Camera settings were kept consistent across samples and cohorts. The images were analyzed using ImageJ software by outlining PV-positive interneurons using the ROI manager and measuring the integrated density of PV and δ expression on the outlined PV-positive interneurons. Each cell was considered its own data point within each animal.

Statistical analysis
Data were analyzed using Prism 8 software (GraphPad Software) and MatWAND in MATLAB (MathWorks). To ensure a consistent time period for analysis across cohorts, we analyzed the first 40 min of baseline and the first 35 min of each injection period. Repeated-measures two-way ANOVAs were performed to detect the significance of frequency, treatment, sex, or genotype. A Greenhouse–Geisser correction was applied where necessary. A mixed-effects model was used if values were missing across days. A post hoc Sidák’s multiple-comparison test was performed to identify significant differences of specific frequency ranges. ANOVA results are reported in Extended Data Tables 1-1, 2-1, and 3-1. Multiple comparisons are reported in Extended Data Tables 1-2, 2-2, and 3-2. p values < 0.05 were considered significant. All n values for each treatment group are shown in the figure legends.

Results
Alcohol modulates BLA network states
To characterize the effect of acute alcohol on BLA oscillations in wild-type mice, we recorded LFPs in the BLA of C57BL/6J mice in response to either a vehicle (0.9% saline, i.p.) or alcohol (1 g/kg, i.p.) injection (Fig. 1A,B). We found that vehicle injections in male C57BL/6J mice significantly decreased high theta power (6–12 Hz; p = 0.0117; 95% CI = 0.04658, 0.4029), while increasing the low gamma power (40–70 Hz; p = 0.0027; 95% CI = −0.08079) compared with baseline (Extended Data Fig. 1-1A). However, we did not find any difference between the two vehicle injections, indicating there was no sensitization or adaptation to the second injection. We have previously observed the impact of vehicle injections on oscillatory states in the BLA (Antonoudiou et al., 2021), which likely reflects the network response to the stress of the injection. Therefore, all results are compared with the first vehicle injection within the treatment paradigm. We performed a dose response examining changes in network activity in response to four different doses of alcohol (0.5, 1.0, 1.5, 2.0 g/kg, i.p.; Extended Data Fig. 1-2).
These experiments determined that the 1.0 g/kg dose was capable of significantly altering relevant BLA network state frequencies without producing lethargy or sedation in the mice (6–12 Hz: \( p = 0.030; \) 95% CI = 0.024, 0.450; Extended Data Fig. 1-2A–C). Therefore, we chose this concentration for our experiments throughout this study. In response to alcohol treatment in male C57BL/6J mice, the power in the beta frequency range (15–30 Hz) is decreased compared with vehicle (\( p = 0.034; \) 95% CI = 0.01033, 0.2925; Fig. 1C,E,G, Extended Data Fig. 1–1C). This indicates that alcohol can modulate specific oscillatory frequencies within the BLA that are implicated both in addiction and mood disorders (Jurado-Barba et al., 2020).

Alcohol modulates BLA network states in a sex-dependent manner

Because of the well documented sex differences in alcohol related behaviors (Melón et al., 2013; Barkley-Levenson and Crabbe, 2015; Becker and Koob, 2016; Sneddon et al., 2019), we treated female C57BL/6J to the same acute alcohol paradigm as described in males (Fig. 1B). Similar to the males, we did not find any significant differences between the two vehicle injections in the vehicle/vehicle control experiments in females (Extended Data Fig. 1-1B). We did find that vehicle significantly decreased high theta power compared with baseline (\( p = 0.0273; \) 95% CI = 0.2699, 0.4296; Fig. 1H), similar to what we observed in the males. Additionally, there was no significant difference between the male and female C57BL/6J BLA LFP response to the vehicle injection (figure not shown).

In response to acute alcohol exposure, we found that alcohol significantly decreased the gamma-band power in female C57BL/6J mice compared with vehicle (\( p = 0.0014; \) 95% CI = 0.2955, 0.9441; Fig. 1D,F,H, Extended Data Fig. 1–1D), a unique signature from the males. Interestingly, this reduction in gamma power represents a blunting of the increase in power exhibited by the vehicle injection (Fig. 1f). Although alcohol decreased BLA power in different frequency bands in males and females, there were no direct significant differences between groups (figure not shown). Collectively, these data suggest that acute ethanol modulates the BLA network differently in male and female mice.

Alcohol modulation of BLA network states involves \( \delta \) subunit-containing GABA\(_{4}\)Rs

Previous literature has supported the role of \( \delta \)-GABA\(_{4}\)Rs in mediating the effects of alcohol on tonic inhibition, drinking, and withdrawal behaviors (Wallner et al., 2003; Santhakumar et al., 2007; Darnieder et al., 2019; Melón et al., 2019). Therefore, to test whether alcohol is mediating its effects on BLA network states through \( \delta \)-GABA\(_{4}\)Rs, we repeated the same procedure in male and female Gabrd\(^{−/−}\) mice. We found that vehicle injections significantly increased BLA power at low gamma frequencies only in the vehicle/alcohol condition in male Gabrd\(^{−/−}\) mice compared with baseline (\( p = 0.0121; \) 95% CI = \(-1.062, -0.1299; \) Fig. 2A,B). In both Gabrd\(^{−/−}\) males and females, we did not find any significant difference between vehicle injections (Extended Data Fig. 2-1).

Unlike C57BL/6J males, acute alcohol significantly decreased the low gamma band of Gabrd\(^{−/−}\) males (\( p = 0.020; \) 95% CI = 0.07, 0.78; Fig. 2A,B) and Gabrd\(^{−/−}\) females (\( p = 0.0012; \) 95% CI = 0.1731, 0.5352; Fig. 2C,D) compared with vehicle. This effect was similar to, but not as robust an effect as, that in C57BL/6J females. However, direct comparisons between male C57BL/6J and male Gabrd\(^{−/−}\) mice or between C57BL/6J females and Gabrd\(^{−/−}\) females did not detect significant differences in the ability of alcohol to modulate oscillatory states (figure not shown). Collectively, these data suggest that the loss of the GABA\(_{4}\)R \( \delta \) subunit impacts the network effect of alcohol more profoundly in males and induces a similar network effect as that observed in C57BL/6J females.

Ability of repeated alcohol exposure to modulate BLA network states is dependent on \( \delta \) subunit-containing GABA\(_{4}\)Rs

Since we established that acute alcohol could modulate specific oscillatory frequencies in the BLA, we were interested in how BLA LFPs changed over time in response to repeated doses of alcohol. Male C57BL/6J and Gabrd\(^{−/−}\) mice received vehicle (0.9% saline) or low dose (1 g/kg, i.p.) alcohol for 5 consecutive days (Fig. 3A). We did not find significant effects of repeated vehicle injections across days in either the male C57BL/6J or male Gabrd\(^{−/−}\) mice (Extended Data Fig. 3-1A,C).

Interestingly, in response to repeated alcohol treatment, we found a change in the baseline low gamma power from the first to last day (BASE2-BASE1) in male C57BL/6J mice (\( p = 0.0252; \) 95% CI = 0.03785, 0.6384; Extended Data Fig. 3-2A), which may be an anticipatory change associated with repeated alcohol administration. In response to alcohol treatment, we observed a significant increase in low gamma power from the first to the last day of exposure (EtOH-BASE2: \( p = 0.0305; \) 95% CI = \(-0.7087, -0.0327; \) Fig. 3C) along with an increase in BEC (first, 161.4 mg/dl; last, 189.8 mg/dl; \( p < 0.0001; \) 95% CI = \(-160.4, -95.81; \) Fig. 3A). In contrast, we did not observe significant effects of repeated alcohol on baseline or treatment in male Gabrd\(^{−/−}\) mice (Fig. 3D, Extended Data Fig. 3–2C).

Direct comparison between male C57BL/6J and Gabrd\(^{−/−}\) mice on the first day of alcohol exposure does not reveal any significant changes within baseline (Extended Data Fig. 4-1A), but did find that male C57BL/6J mice had significantly decreased high theta (\( p = 0.0138; \) 95% CI = \(-1.061, -1.088\)) and beta (\( p = 0.0022; \) 95% CI = \(-0.7193, -0.1659\)) BLA power compared with male Gabrd\(^{−/−}\) mice in response to alcohol exposure (Fig. 4A). By the last day, there were significant decreases within the baseline period specifically in the low gamma (\( p = 0.0343; \) 95% CI = \(-0.9892, -0.03576\)) and high gamma (\( p = 0.0062; \) 95% CI = \(-0.5908, -0.09553\)) BLA power in male C57BL/6J mice compared with male Gabrd\(^{−/−}\) mice (Extended Data Fig. 4-1B), again likely attributed to the role of the GABA\(_{4}\)R \( \delta \)
Figure 1. Acute alcohol exposure alters BLA network activity differently in male and female C57BL/6J mice. A, Representative targeting of BLA LFP recordings. B, Acute alcohol exposure paradigm consisted of LFP recordings during baseline (60 min), vehicle injection (0.9% saline, i.p.; 60 min), and a treatment injection (0.9% saline or 1 g/kg alcohol, i.p.; 60 min). This dose was determined through a dose response (Extended Data Fig. 1-2). C, D, Representative male spectrogram of normalized beta power (15–30 Hz; C) and representative female spectrogram of normalized gamma power (40–70 Hz; D) from acute alcohol exposure. E, F, Average...
subunit in the anticipatory effects of repeated alcohol exposure. In response to repeated alcohol administration, we observed a significant increase in the high gamma frequency range in male C57BL/6J mice compared with male Gabrd−/− mice (p = 0.0203; 95% CI = 0.02211, 0.2899; Fig. 4B). Overall, these results suggest a blunted impact of acute and repeated alcohol exposure on BLA oscillatory states in mice lacking the GABAARδ subunit. Further, these data indicate a role for δ-GABAARs in adapting to alcohol exposure over time, as well as anticipating alcohol treatment, as shown by the changes in baseline in male C57BL/6J mice, but not Gabrd−/− mice.

Sex differences in BLA network states in response to repeated alcohol exposure

Repeated alcohol exposure in female C57BL/6J and Gabrd−/− mice involved acute alcohol or vehicle exposure on day 1 and the repeated alcohol exposure on days 2–5 (Fig. 3B). We will be using their second day of exposure in our repeated alcohol comparisons, which were not significantly different in female C57BL/6J mice (Extended Data Fig. 3-3A). Neither day 1 nor day 2 were significantly different from day 5 in female Gabrd−/− mice (Extended Data Fig. 3-3B,C). Therefore, we continued to use day 2 as the first day of repeated exposure in our analysis.

We did not observe significant effects of vehicle exposure across days in female C57BL/6J (Extended Data Fig. 3-1B) or female Gabrd−/− mice (Extended Data Fig. 3-1D). Interestingly, unlike the males, we did not observe any significant effect of repeated alcohol administration in C57BL/6J female and Gabrd−/− females across days (Fig. 3D,F, Extended Data Fig. 3-2B,D) despite an increase in BEC in female C57BL/6J mice from the first to last day of exposure (first, 151.1 mg/dl; last, 257.3 mg/dl; p = 0.0072; 95% CI = −166.0, −46.32; Fig. 3B).

Direct comparison between C57BL/6J males and females did not reveal significant differences at any frequency range within the baseline period (Extended Data Fig. 4-1C) or in the effect of alcohol from baseline (Fig. 4C) on the first day of exposure. However, by the last day, we found an increase in high theta power (p = 0.0435; 95% CI = 0.01858, 1.216) and a decrease in high gamma power within the baseline of males, but not in the females (p = 0.0122; 95% CI = −0.5198, −0.06395; Extended Data Fig. 4-1D). In response to repeated alcohol exposure, males exhibited a significantly reduced power in the high

Figure 2. Acute alcohol exposure produces effects in Gabrd−/− mice similar to those in female C57BL/6J mice. A, C, Representative male (A) and female (C) spectrogram of normalized gamma power from acute injections of vehicle and alcohol (1 g/kg, i.p.; Extended Data Fig. 1-2, dose response). Dots represent the mean, and the shaded region represents the SEM. G, H, Normalized power area for vehicle/alcohol acute exposure in males (n = 11; G) and females (n = 8; H). #p < 0.05 vs baseline; *p < 0.05, **p < 0.01 vs vehicle. Acute vehicle exposure does not alter BLA network activity in C57BL/6J mice (Extended Data Fig. 1-1). Summaries of ANOVA and multiple-comparison tests can be found in Extended Data Tables 1-1 and 1-2.

July/August 2022, 9(4) ENEURO.0010-22.2022 eNeuro.org
Figure 3. Repeated alcohol exposure exaggerates BLA network modulation in male C57BL/6J mice. 

A, Experimental paradigm (left) of the repeated alcohol exposure procedure in male C57BL/6J and Gabrd<sup>−/−</sup> mice of BLA LFP recordings during baseline (60 min) and vehicle or alcohol injections (0.9% saline or 1 g/kg alcohol, i.p.; 60 min) over 5 d. BEC measurements (right) of male C57BL/6J mice (n = 6) taken 15 min after alcohol exposure (1 g/kg, i.p.) on days 1, 2, and 5 of exposure. ***p < 0.0001.

B, Repeated alcohol exposure paradigm (left) for female C57BL/6J and Gabrd<sup>−/−</sup> mice, which includes the acute alcohol exposure and the repeated alcohol exposure as day 2 (first day) to 5 (last day). Justification for using day 2 instead of day 1 is in Extended Data Figure 3-3. BEC measurements (right) of female C57BL/6J mice (n = 5) taken 15 min after alcohol exposure (1 g/kg, i.p.) on days 1, 2, and 5 of exposure.

C–F, Change in the effect of alcohol on the first and last days of exposure in male (first day, n = 8; last day, n = 6; C) and female (first day, n = 10; last day, n = 8) C57BL/6J mice and male (first day, n = 8; last day, n = 8; E) and female (first day, n = 8; last day, n = 8) Gabrd<sup>−/−</sup> mice. *p < 0.05 versus first exposure. Repeated vehicle exposure does not change BLA network activity (Extended Data Fig. 3-1). Baseline network activity is modulated by repeated alcohol exposure in male C57BL/6J mice (Extended Data Fig. 3-2). Summaries of ANOVA and multiple-comparison tests can be found in Extended Data Tables 3-1 and 3-2.
theta frequency range in C57BL/6J males with no effect in females (\(p = 0.0433; 95\% \text{ CI} = 1.085, -0.01596; \text{Fig. 4D}\)). There was no significant difference between male and female BECs on day 1 or day 5, suggesting the same level of alcohol intoxication modulates BLA LFPs differentially in the two sexes. Collectively, these data suggest that male C57BL/6J mice are becoming more sensitive to the sedative effects of alcohol as demonstrated by the significant effects of repeated alcohol administration on gamma frequency ranges, an effect that involves the GABAAR \(\delta\) subunit (Pian et al., 2008).

Repeated alcohol exposure alters \(\delta\)-expression on PV interneurons in the BLA

Alcohol exposure can change the expression of GABAAR subunits (Liang et al., 2004; Olsen and Spigelman, 2012; Lindemeyer et al., 2014; Follesa et al., 2015), and sex differences in GABAAR \(\delta\) subunit expression has been reported (Maguire et al., 2005). Changes in the expression of the GABAAR \(\delta\) subunit, whether through genetic deletions or hormone fluctuations during pregnancy, can alter specific oscillation frequencies in the hippocampus (Ferando and Mody, 2013, 2015). Therefore, we hypothesized that altered expression of the GABAAR \(\delta\) subunit on PV interneurons in the BLA may contribute to our observed sex differences in BLA network states. Thus, we examined whether there were any potential sex differences in GABAAR \(\delta\) subunit expression in the BLA or in \(\delta\) expression associated with alcohol exposure. We observed a higher \(\delta\) expression on PV interneurons in naive female C57BL/6J mice (mean = 1,002,542; SEM = 45,011) compared with naive male C57BL/6J mice (mean = 538,252; SEM = 12,440; \(t_{424} = 10.39, p < 0.0001; \text{Fig. 5B}\)) with no change to PV immunoreactivity (female: mean = 1,464,808; SEM = 51,507; male: mean = 1,448,517; SEM = 97,320; Fig. 5A). Interestingly, vehicle treatment alone reduced PV immunoreactivity in females compared with males (\(p < 0.0001; 95\% \text{ CI} = 267746, 621366; \text{Fig. 5D}\)) and also reduced \(\delta\) expression on PV neurons in females compared with males (\(p < 0.0001; 95\% \text{ CI} = 35,568, 126,727; \text{Fig. 5E}\)). These data demonstrate baseline sex differences in the expression and lability of GABAAR \(\delta\) expression on PV interneurons in the BLA.

Repeated alcohol treatment reduced PV immunoreactivity in males compared with vehicle (\(p < 0.0001; 95\% \text{ CI} = 147,380, 518,402; \text{Fig. 5D}\)). In contrast, repeated alcohol exposure in females did not alter PV immunoreactivity but did significantly reduce GABAAR \(\delta\) expression on PV interneurons compared with vehicle (\(p = 0.0018; 95\% \text{ CI} = 35,568, 126,727; \text{Fig. 5E}\)), an effect that was not observed in males.

Comparing males and females exposed to repeated alcohol, PV immunoreactivity (\(p = 0.0195; 95\% \text{ CI} = 15,583, 98,517; \text{Fig. 5E}\)) and \(\delta\) expression on PV interneurons are reduced in females compared with males (\(p < 0.0001; 95\% \text{ CI} = 54,848, 142,689; \text{Fig. 5E}\)). These data imply that changes in GABAAR \(\delta\) expression on PV interneurons may mediate sex differences and the response to repeated alcohol exposure.
Figure 5. Repeated alcohol exposure reduces δ-GABAA receptor expression on PV interneurons in female C57BL/6J mice. 

A, B. Integrated density of PV immunoreactivity (A) and δ expression (B) on PV interneurons in naïve male (cell, n = 224; animal, n = 4) and female (cell, n = 202; animal, n = 4) C57BL/6J mice. C. Representative images from male and female C57BL/6J mice who received repeated administration of vehicle or alcohol. D, E. Integrated density of PV immunoreactivity (D) and δ expression (E) on PV interneurons in the BLA of male (vehicle: cell, n = 112; animal, n = 3; alcohol: cell, n = 97; animal, n = 3) and female (vehicle: cell, n = 117; animal, n = 3; alcohol: cell, n = 169; animal, n = 3) C57BL/6J mice who received repeated administrations of vehicle or alcohol. *p < 0.05, **p < 0.01, ****p < 0.0001.
Discussion

Network states have been shown to correlate with behavioral states, and accumulating evidence demonstrates that signature oscillatory states in the BLA are associated with fear and anxiety states (Likhit et al., 2014; Stuijenske et al., 2014; Davis et al., 2017; Antonoudiou et al., 2021). In fact, optogenetically driving specific oscillatory states influences the behavioral expression of fear (Ozawa et al., 2020) and learned helplessness (Antonoudiou et al., 2021). However, limited studies have examined the physiological, pathological, or pharmacological mechanisms mediating transitions between network and behavioral states. Recent work has demonstrated that chronic stress can perturb oscillations in the BLA and a clinically effective antidepressant treatment can restore the “healthy” network state (Antonoudiou et al., 2021; Walton et al., 2022). Here, we examine the impact of alcohol on BLA network states. Given the anxiolytic effects of alcohol, we posited that alcohol may be capable of shifting the network state toward the anxiolytic state. We demonstrate that acute alcohol exposure is capable of altering BLA network states and that there are sex differences in the effect of alcohol on BLA network states, affecting different frequencies in males and females. These data are the first to demonstrate that alcohol is capable of modulating network states associated with affective states.

It has been demonstrated that PV interneurons are critical in orchestrating oscillatory states in the BLA (Antonoudiou et al., 2021). PV interneurons in the BLA express a high density of δ-GABA<sub>A</sub>Rs, which have been suggested to be a target for low-dose alcohol (Sundstrom-Poromaa et al., 2002; Wallner et al., 2003; Hanchar et al., 2006; Santhakumar et al., 2007). However, the actions of alcohol directly on these receptors remains somewhat controversial (Borghese et al., 2006; Korpi et al., 2007). It is important to note that the majority of these studies focus solely on principal neurons; GABA<sub>E</sub>ergic interneurons, on the other hand, have a unique receptor subunit composition in which the δ subunit has been shown to partner with the α1 subunit and have been demonstrated to generate tonic GABA<sub>E</sub>ergic currents, which are highly sensitive to low concentrations of ethanol (Glykys et al., 2007). Thus, we proposed that the high expression of δ subunit-containing GABA<sub>A</sub>Rs on PV interneurons in the BLA may confer unique sensitivity to the effects of alcohol and, given the role of these interneurons in coordinating oscillations, may mediate the effects of alcohol on BLA network states. Here we demonstrate that δ-GABA<sub>A</sub>Rs influence the ability of alcohol to alter specific oscillatory states in the BLA, blunting the ability to shift network states. Specifically, we observed a reduction of beta power from acute ethanol in male wild-type mice that was blunted in mice lacking δ-GABA<sub>A</sub>Rs (Figs. 1, 2). Others have found reductions of beta power in the nucleus accumbens shell during alcohol relapse related to reduced synchrony of the local network, suggesting that this reduction we find in response to alcohol may also be from reduced synchrony (Hadar et al., 2017). This power detected in the beta frequency may arise from the neighboring high-theta oscillator, given the lack of a clear beta peak (Extended Data Fig. 1-1C). Regardless, these data suggest that δ subunit-containing GABA<sub>A</sub>Rs are important players in mediating the effects of alcohol on oscillatory states related to mood/anxiety, although it is also possible that other GABA<sub>A</sub>R subtypes are involved. Previous studies demonstrated that the δ subunit has a specific role in lower frequencies compared with higher frequencies (Antonoudiou et al., 2021), which may be true for the effects of alcohol as well. Further studies are required to investigate the impact of other GABA<sub>A</sub>R subtypes in mediating the ability of alcohol to modulate BLA network states given that previous studies have implicated other GABA<sub>A</sub>R subtypes, such as the γ2 subunit, in anxiety-like behavior (Chandra et al., 2005) and alcohol withdrawal severity (Buck and Hood, 1998). It is also possible that the indirect effects of alcohol on receptor expression, neurotransmitter availability, and other neuromodulators could account for the changes in BLA oscillations observed here (Morrow et al., 2001; Fleming et al., 2009; Olsen and Liang, 2017). For example, the effects of alcohol have been suggested to be mediated through the action of neuroactive steroids (Morrow et al., 2001; Finn et al., 2010; Finn and Jimenez, 2018), and, given recent evidence that allopregnanolone can alter BLA network states (Antonoudiou et al., 2021; Walton et al., 2022), this may be an indirect mechanism whereby alcohol could modulate BLA network states. Arguing against this indirect mechanism is the evidence that alcohol exerts unique effects on BLA network states compared with allopregnanolone (Antonoudiou et al., 2021).

Since expression of δ subunit-containing GABA<sub>A</sub>Rs have been shown to be sensitive to ovarian steroid hormone modulation and are implicated in sex differences in alcohol intake (Darnieder et al., 2019), we hypothesized that there may be sex differences in the ability of alcohol to modulate BLA network states through actions on these receptors. In fact, we do observe sex differences in the modulation of BLA network states by alcohol, although both male and female C57BL/6J mice reach similar BAC levels after alcohol exposure (Fig. 3A,B). Interestingly, the loss of the GABA<sub>A</sub>R δ subunit in males shifts the alcohol modulation of the BLA network state toward the signature that we observe for female C57BL/6J mice (Figs. 1, 3), and we believe that the observed sex differences in the expression of δ subunit-containing GABA<sub>A</sub>Rs in the BLA (Fig. 5) may underlie these differences. Further, there are well documented sex differences in responses to alcohol, alcohol-related anxiety-like behavior, and estrous cycle-dependent δ expression (Maguire et al., 2005; Rhodes et al., 2005; Barkley-Levenson and Crabbe, 2015), consistent with our observations of sex differences in the alcohol-induced modulation of network states. Additionally, sex differences have been reported in neural oscillations in major depressive disorder with oscillatory signatures of susceptibility (Thériault et al., 2021). Future studies are required to evaluate the relationship between the capacity of alcohol to modulate network states and voluntary alcohol consumption, the anxiolytic effects of alcohol, and the anxiogenic effects of alcohol withdrawal.

To investigate whether the effect of alcohol on network states may be altered after repeated exposure, we treated mice with low-dose alcohol for up to 5 d. Interestingly, we
found that BLA network states changed before alcohol exposure. Given the evidence that the amygdala is involved in valence encoding and assignment, it is possible that network state changes before the alcohol exposure is reflective of the anticipation or expectation of the event, which has been demonstrated in a pavlovian conditioning paradigm (Pignatelli and Beyeler, 2019; Tallot et al., 2020). We found robust effects of repeated alcohol exposure on the BLA LFP response of male C57BL/6J mice, which was significantly different from the male Gabrd−/− and female C57BL/6J mice. In fact, we found no differences in the extent of the effect of repeated alcohol on network states in female C57BL/6J or Gabrd−/− mice despite similar BEC levels between male and female C57BL/6J mice. It is possible that the change in BLA power in male C57BL/6J mice across repeated alcohol exposure is because of the increase in BEC rather than an adaptation to the injections. Indeed, our dose response data show that higher doses of alcohol do have larger effects on BLA network states. Alcohol administration prominently affected gamma-band oscillations in the BLA, a network activity that has been associated with local network synchrony, affective learning, and memory consolidation, (Bocchio et al., 2017; Antonoudiou et al., 2021; Headley et al., 2021).

Given the critical role of PV interneurons in the generation of BLA gamma oscillations (Antonoudiou et al., 2021; Headley et al., 2021), alcohol may directly modulate PV interneuron signaling. It is possible that reduction in tonic inhibition of PV interneurons in female and Gabrd−/− mice makes PV interneurons more susceptible to the effects of acute ethanol leading to disruption in the generation of gamma network oscillations in BLA.

Since the downregulation of the δ subunit has been thought to confer tolerance to alcohol (Olsen and Liang, 2017), the reduction of δ subunit in female, but not male, C57BL/6J mice could explain the lack of effects on LFPs after repeated alcohol exposure. Further, GABAAR agonists and positive allosteric modulators, like neurosteroids, which exert effects through the δ subunit, can block tolerance to the sedative effects of alcohol (Debatin and Barbosa, 2006; Barbosa and Morato, 2007). This could explain why alcohol does not change the BLA network in female C57BL/6J and Gabrd−/− mice, who have reductions in δ-GABAAR expression, after repeated administration like it does in male C57BL/6J mice. However, this study did not directly measure sensitization or tolerance to the effects of alcohol, and future studies could link network and behavioral changes. Last, we found that δ expression on PV interneurons is increased in naive female C57BL/6J mice compared with males. Because we did not see any baseline differences in BLA network states between male and female C57BL/6J mice, this difference in expression may not impact BLA oscillations, but the expression of δ in females does influence the response to alcohol exposure.

The literature and recent findings demonstrate a strong role for PV interneurons in oscillation generation (Antonoudiou et al., 2021), giving support to the likely fact that the effects of alcohol on PV interneurons are influencing the oscillations. However, because of the heterogeneity of the interneuron population in the BLA, it is possible that other interneuron types, like somatostatin, cholecystokinin, or PKC-δ-expressing cells may be involved in effecting oscillations (Klausberger et al., 2005). Furthermore, another major influence on BLA oscillations are other brain areas with strong network connections to the BLA, such as the mPFC, which is heavily implicated in addiction (Goldstein and Volkow, 2011; Davis et al., 2017; Ozawa et al., 2020).

Forced alcohol injections or alcohol-induced aversion can cause stress to mice, which may contribute to the observed effects (Eckardt et al., 1974). However, our experimental plan was designed to dissociate any stressful or unpleasant effects of the infusion from the effects of alcohol. Although we observed significant effects of vehicle injections on BLA LFP responses compared with baseline (Extended Data Fig. 1-1), we did not find any sensitization or tolerance to the injections in the acute alcohol experiment or across days in the repeated alcohol experiment (Extended Data Figs. 1-1, 2-1, 3-1), similar to what has been reported previously (Antonoudiou et al., 2021). Thus, we are confident that the observed effects of alcohol on BLA oscillatory states is because of the effects of alcohol rather than an aversive experience related to the route of administration. Further, our data suggest that the effects of alcohol may mitigate the stress-induced effects on the BLA network state.

To our knowledge, this is the first demonstration that alcohol can modulate oscillations in the BLA, which have been implicated in governing behavioral states. Numerous studies have investigated the relationship of BLA network states to behavioral states; however, few studies have investigated mechanisms mediating transitions between BLA network states. The current study demonstrates that alcohol can induce a transition between network states associated with fear and anxiety, which may mediate the impact of alcohol on anxiety states. Future work is required to investigate how changes in the BLA relate to other connected areas implicated in alcohol use and anxiety, such as the central amygdala, mPFC, nucleus accumbens, BNST, and ventral striatum (Janak and Tye, 2015). Recordings of oscillations are stable over long periods of time and thus can be examined throughout the addiction cycle from intoxication to withdrawal to preoccupation in specific brain areas to understand how alcohol changes communication between these areas. Thus, this novel approach may demonstrate utility in understanding the trajectory from first exposure to alcohol dependence and the contribution of both the positive and negative reinforcing effects of alcohol.

References

Agoglia A, Herman M (2018) The center of the emotional universe: alcohol, stress, and CRF1 amygdala circuitry. Alcohol 72:61–73.

Antonoudiou P, Colmers PLW, Walton NL, Weiss GL, Smith AC, Nguyen DP, Lewis M, Quirk MC, Barros L, Melón LC, Maguire JL (2021) Allopregnanolone mediates affective switching through modulation of oscillatory states in the basolateral amygdala. Biol Psychiatry 91:293–293.

Barbosa ADE, Morato GS (2007) Influence of epipregnanolone on the modulation of rapid tolerance to ethanol by neurosteroids. Braz J Psychiatry 29:337–345.
Barkley-Levenson AM, Crabbe JC (2015) Genotypic and sex differences in anxiety-like behavior and alcohol-induced anxiety in high drinking in the dark selected mice. Alcohol 49:29–36.

Bartos M, Vida I, Jonas P (2007) Synaptic mechanisms of synchronized gamma oscillations in inhibitory interneuron networks. Nat Rev Neurosci 8:46–56.

Becker JB, Koob GF (2016) Sex differences in animal models: focus on addiction. Pharmacol Rev 68:242–263.

Bocchio M, Capogna M (2014) Oscillatory substrates of fear and safety. Neuon 83:753–755.

Bocchio M, Nabavi S, Capogna M (2017) Synchronous plasticity, engrams, and network oscillations in amygdala circuits for storage and retrieval of emotional memories. Neuon 94:731–743.

Borghese CM, Störstrovu ŞI, Ebert B, Herd MB, Beletti D, Lambert JJ, Marshall G, Wafford KA, Harris RA (2006) The δ subunit of γ-aminobutyric acid type A receptors does not confer sensitivity to low concentrations of ethanol. J Pharmacol Exp Ther 316:1360–1368.

Buck KJ, Hood HM (1998) Genetic association of a GABAA receptor γ2 subunit variant with severity of acute physiological dependence. Mamm Genome 9:975–978.

Chandra D, Korpi ER, Miralles CP, de Blas AL, Homanics GE (2005) GABAA receptor γ2 subunit knockdown mice have enhanced anxiety-like behavior but unaltered hypnotic response to benzodiazepines. BMC Neurosci 6:30.

Debatin T, Barbosa ADE (2006) Effect of isopregnanolone on rapid tolerance to the anxiolytic effect of ethanol. J Pharmacol Exp Ther 316:1360–1368.

Eckardt MJ, Skurdal AJ, Brown JS (1974) Conditioned taste aversion

Fernando I, Mody I (2013) Altered gamma oscillations during pregnancy through loss of δ subunit-containing GABAA receptors on parvalbumin interneurons. Front Neural Circuits 7:144.

Fernando I, Mody I (2014) Altered gamma oscillations following partial and complete ablation of δ subunit-containing GABAA receptors from parvalbumin interneurons. Neuropharmacology 88:91–98.

Finn DA, Jimenez VA (2018) Dynamic adaptation in neurosteroid neurotransmission in response to alcohol. In: Handbook of experimental pharmacology (Grant KA, Lovinger DM, eds), pp 1–24. Berlin, Heidelberg: Springer.

Finn DA, Beckley EH, Kaufman KR, Ford MM (2010) Manipulation of GABAergic steroids: sex differences in the effects on alcohol drinking- and withdrawal-related behaviors. Harm Behav 57:12–22.

Fleming RL, Manis PB, Morrow AL (2009) The effects of acute and chronic ethanol exposure on presynaptic and postsynaptic γ-amino butyric acid (GABA) neurotransmission in cultured cortical and hippocampal neurons. Alcohol 43:603–618.

Follesa P, Floris G, Asuni GP, Ibbá A, Tocco MG, Zicca L, Mercante B, Deriu F, Gorini G (2015) Chronic intermittent ethanol regulates hippocampal GABA(A) receptor delta subunit gene expression. Front Cell Neurosci 9:445.

Freeman WJ, Rogers LJ, Holmes MD, Silbergeld DL (2000) Spatial and complete ablation of parvalbumin interneurons. Front Neural Circuits 7:144.

Fuchs T, Jefferson SJ, Hooper A, Yee P-H, Maguire J, Luscher B (2017) Disinhibition of somatostatin-positive GABAergic interneurons results in an anxiety-like and antidepressant-like brain state. Mol Psychiatry 22:920–930.

Glipin NW, Koob GF (2008) Neurobiology of alcohol dependence focus on motivational mechanisms. Alcohol Res Health 31:185–195.

Glykys J, Peng Z, Chandra D, Homanics GE, Houser CR, Mody I (2007) A new naturally occurring GABA_A receptor subunit partnership with high sensitivity to ethanol. Nat Neurosci 10:40–48.

Goldstein RZ, Volkow ND (2011) Dysfunction of the prefrontal cortex in addiction: neuroimaging findings and clinical implications. Nat Rev Neurosci 12:652–669.

Hadar R, Voget M, Vengeliene V, Haumesser JK, van Riesen C, Avchalumov Y, Spanagel R, Winter C (2017) Altered neural oscillations and elevated dopamine levels in the reward pathway during alcohol relapse. Behav Brain Res 316:131–135.

Hanchar HJ, Chutsrisopon P, Meera P, Supavilai P, Sieghart W, Wallner M, Olsen RW (2006) Ethanol potently and competitively inhibits binding of the alcohol antagonist Ro15-4513 to 4/6 GABAA receptors. Proc Natl Acad Sci U.S.A 103:8546–8551.

Headley DB, Kyriazi P, Feng N, Ssr SS, Paré D (2021) Gamma Oscillations in the Basolateral Amygdala: Localization, Microcircuitry, and Behavioral Correlates. J Neurosci 41:6087–6101.

Janak PH, Tye KM (2015) From circuits to behaviour in the amygdala. Nature 517:284–292.

Jurado-Barba R, Sion A, Martínez-Maldonado A, Domínguez-Centeno I, Prieto-Montalvo J, Navarrete F, Salud García-Gutierrez M, Mancinas J, Rubió G (2020) Neuropsychophysiological measures of alcohol dependence: can we use EEG in the clinical assessment? Front Psychiatry 11:676.

Klausberger T, Marton LF, O’Neill J, Huck JHJ, Dalezios Y, Fuentealba P, Suen WY, Papp E, Kaneko T, Watanabe M, Csicsvari J, Somogyi P (2005) Complementary roles of cholecystokinin- and parvalbumin-expressing GABAergic neurons in hippocampal network oscillations. J Neurosci 25:9782–9793.

Korpi ER, Debus F, Linden AM, Málécot C, Leppä E, Vekovischeva V, Kruse W, Eckhorn R (1996) Inhibition of sustained gamma oscillations and network activity in amygdala circuits for storage and retrieval of emotional memories. Front Cell Neurosci 9:445.

Kruse W, Eckhorn R (1996) Inhibition of sustained gamma oscillations and network activity in amygdala circuits for storage and retrieval of emotional memories. Front Cell Neurosci 9:445.

Kushner MG, Maurer E, Menary K, Thuras P (2011) Vulnerability to between health behaviors, posttraumatic stress disorder, and co-morbidity general anxiety and depression. Cogn Behav Ther 48:184–193.

Lindemeyer AK, Liang J, Cagetti E, Olsen RW, Spigelman I (2014) Ethanol-induced plasticity of GABAergic receptors in the basolateral amygdala. Neurochem Res 39:1162–1170.

Liang J, Cagetti E, Olsen RW, Spigelman I (2004) Altered pharmacology of synaptic and extrasynaptic GABA receptors on CA1 hippocampal neurons is consistent with subunit changes in a model of alcohol withdrawal and dependence. J Pharmacol Exp Ther 310:1234–1245.

Likhitk E, Stjenske JM, Topiwala MA, Harris AZ, Gordon JA (2014) Prefrontal entrainment of amygdala activity signals safety in learned fear and innate anxiety. Nat Neurosci 17:106–113.

Lindemeyer AK, Liang J, Mard VR, Meyer EM, Suryanarayanan A, Olsen RW, Spigelman I (2014) Ethanol-induced plasticity of GABAergic receptors in the basolateral amygdala. Neurochem Res 39:1162–1170.

Mann EO, Mody I (2010) Control of hippocampal gamma oscillation frequency by tonic inhibition and excitation of interneurons. Nat Neurosci 13:205–212.

Mason JE, LeBouthillier DM, Asmundson GJG (2019) Relationships between health behaviors, posttraumatic stress disorder, and co-morbidity general anxiety and depression. Cogn Behav Ther 48:184–199.

McDonald AJ (1992) Projection neurons of the basolateral amygdala: a correlative Golgi and retrograde tract tracing study. Brain Res Bull 28:179–185.
Melón LC, Wray KN, Moore EM, Boehm SL (2013) Sex and age differences in heavy binge drinking and its effects on alcohol responsivity following abstinence. Pharmacol Biochem Behav 104:177–187.

Melón LC, Nasman JT, John AS, Mbonu K, Maguire JL (2019) Internueral δ-GABAA receptors regulate binge drinking and are necessary for the behavioral effects of early withdrawal. Neuropsychopharmacology 44:425–434.

Mihalek RM, Banerjee PK, Korpi ER, Quinlan JJ, Firestone LL, Mi Z-P, Lagenaur C, Trotter V, Sieghart W, Anagnostaras SG, Sage JR, Farselow MS, Guidotti A, Spigelman I, Li Z, Delorey TM, Olsen RW, Homansic GE (1999) Attenuated sensitivity to neuroactive steroids in aminobutyrate type A receptor delta subunit knockout mice. Proc Natl Acad Sci U.S.A 96:12905–12910.

Mihalek RM, Bowers BJ, Wehner JM, Kralic JE, VanDoren MJ, Morrow AL, Homansic GE (2001) GABAa receptor delta subunit knockout mice have multiple defects in behavioral responses to ethanol. Alcoholism Clin Exp Res 25:1708–1718.

Morrow AL, VanDoren MJ, Penland SN, Matthews DB (2001) The role of GABAAergic neuroactive steroids in ethanol action, tolerance and dependence. Brain Res Reviews 37:98–109.

Olsen RW, Spigelman I (2012) GABAA receptor plasticity in alcohol withdrawal. In: Jasper’s basic mechanisms of the epilepsies, Ed 4 (Noebels JL, ed, pp 1–14). Bethesda, MD: National Center for Biotechnology Information.

Olsen RW, Liang J (2017) Role of GABAA receptors in alcohol use disorders suggested by chronic intermittent ethanol (CIE) rodent model. Mol Brain 10:45.

Oppenheim AV, Schafer RW, Buck JR (1999) Discrete-time signal processing, Ed 2. Upper Saddle River, NJ: Prentice Hall.

Ozawa M, Davis P, Ni J, Maguire J, Papouin T, Reijmers L (2020) Experience-dependent resonance in amygdalo-cortical circuits supports fear memory retrieval following extinction. Nat Commun 11:4358.

Pape HC, Paré D, Driesang RB, Paré D, Driesang RB (1998) Two types of intrinsic oscillations in neurons of the lateral and basolateral nuclei of the amygdala. J Neurophysiol 79:205–216.

Pavlov I, Savtchenko LP, Song I, Koo J, Pimashkin A, Rusanov DA, Semyanov A (2014) Tonic GABAA conductance bidirectionally controls interneuron firing pattern and synchronization in the CA3 hippocampal network. Proc Natl Acad Sci U.S.A 111:504–509.

Pian JP, Criado JR, Walker BM, Ehlers CL (2008) Differential effects of acute alcohol on EEG and sedative responses in adolescent and adult Wistar rats. Brain Res 1194:28–36.

Pignatelli M, Beyeler A (2019) Valence coding in amygdala circuits. Curr Opin Behav Sci 26:97–106.

Rhodes JS, Best K, Belknap JK, Finn DA, Crabbe JC (2005) Evaluation of a simple model of ethanol drinking to intoxication in C57BL/6J mice. Physiol Behav 84:53–63.

Santhakumar V, Wallner M, Otis TS (2007) Ethanol acts directly on extrasynaptic subtypes of GABAA receptors to increase tonic inhibition. Alcohol 41:211–221.

Silberman Y, Bajo M, Chappell AM, Christian DT, Cruz M, Diaz MR, Kash T, Lack AK, Messing RO, Siggins GR, Winder D, Roberto M, Mccool BA, Weiner JL (2009) Neurobiological mechanisms contributing to alcohol-stress-anxiety interactions. Alcohol 43:509–519.

Smith JP, Randall CL (2012) Anxiety and alcohol use disorders: comorbidity and treatment considerations. Alcohol Res 23:414–431.

Sneddon EA, White RD, Radke AK (2019) Sex differences in binge-like and aversion-resistant alcohol drinking in C57BL/6J mice. Alcohol Clin Exp Res 43:249–249.

Sohal VS, Zhang F, Yizhar O, Deisseroth K (2009) Parvalbumin neurons and gamma rhythms enhance cortical circuit performance. Nature 459:696–702.

Stujenske JM, Likhtik E, Topiwala MA, Gordon JA (2014) Fear and safety engage competing patterns of theta-gamma coupling in the basolateral amygdala. Neuron 83:919–933.

Substance Abuse and Mental Health Services Administration (2020) Key substance use and mental health indicators in the United States: results from the 2019 national survey on drug use and health. Rockville, MD: Substance Abuse and Mental Health Services Administration.

Sundstrom-Poroma I, Smith DH, Gong QH, Sabado TN, Li X, Light A, Wiedmann M, Williams K, Smith SS (2002) Hormonally regulated aβδ GABAa receptors are a target for alcohol. Nat Neurosci 5:721–722.

Tallot L, Graupner M, Diaz-Mataix L, Doyère V (2020) Beyond freezing: temporal expectancy of an aversive event engages the amygdalo-prefronto-dorsostriatal network. Cereb Cortex 30:5257–5269.

Thériault RK, Manuduca JD, Perreault ML (2021) Sex differences in innate and adaptive neural oscillatory patterns link resilience and susceptibility to chronic stress in rats. J Psychiatry Neurosci 46: E258–E270.

Thomas SE, Randall CL, Carrigan MH (2003) Drinking to cope in socially anxious individuals: a controlled study. Alcohol Clin Exp Res 27:1837–1943.

Tye KM, Prakash R, Kim S-Y, Fenno LE, Grosenick L, Zarabi H, Thompson KR, Gradinaru V, Ramakrishnan C, Deisseroth K (2011) Amygdala circuitry mediating reversible and bidirectional control of anxiety. Nature 471:358–362.

Wallner M, Hancher HJ, Olsen RW (2003) Ethanol enhances aβδ and aβδ γ-aminobutyric acid type A receptors at low concentrations known to affect humans. Proc Natl Acad Sci U.S.A 100:15218–15223.

Walton NL, Antonoudiou P, Barros L, DiLeo A, Gabby J, Howard S, Paracha R, Sánchez EJ, Weiss GL, Kong D, Maguire JL (2022) Impaired endogenous neurosteroid signaling contributes to behavioral deficits associated with chronic stress. BioRxiv. doi:10.1101/2021.12.30.474579.