Somatic Embryogenesis: Identified Factors that Lead to Embryogenic Repression. A Case of Species of the Same Genus

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Abstract

Somatic embryogenesis is a powerful biotechnological tool for the mass production of economically important cultivars. Due to the cellular totipotency of plants, somatic cells under appropriate conditions are able to develop a complete functional embryo. During the induction of somatic embryogenesis, there are different factors involved in the success or failure of the somatic embryogenesis response. Among these factors, the origin of the explant, the culture medium and the in vitro environmental conditions have been the most studied. However, the secretion of molecules into the media has not been fully addressed. We found that the somatic embryogenesis of *Coffea canephora*, a highly direct embryogenic species, is disrupted by the metabolites secreted from *C. arabica*, a poorly direct embryogenic species. These metabolites also affect DNA methylation. Our results show that the abundance of two major phenolic compounds, caffeine and chlorogenic acid, are responsible for inhibiting somatic embryogenesis in *C. canephora*.

Introduction

The production of a whole plant from a single somatic cell is a clear confirmation that cells contain the genetic information necessary to develop a new and functional plant [1]. This process is known as somatic embryogenesis (SE), and is a powerful biotechnological process that allows the mass production of economically important cultivars. Furthermore, SE is also an attractive system to study the morphology, biochemistry, genetic and molecular mechanisms of embryo development [2]. This system is built on the concept of cellular totipotency, which is the ability of a single cell to divide and produce a complete and functional plant. A certain
subset of cells, under specific induction conditions, can generate embryogenic cells that lead to the initiation of the somatic embryo [1,3].

There are different factors involved in the success or failure of the SE response. Among these factors are the species, the origin of the explant, the culture medium, the type and concentration of the growth regulators used, the nitrogen and carbon sources, and the in vitro environmental conditions [4–6]. Another important factor that affects the SE response is the release of organic molecules by the explants into the culture medium. Some of the secreted molecules that have been demonstrated to induce or modulate the SE response [7–9] have been classified as polysaccharides, amino acids, growth regulators, vitamins [10], proteins and lipophilic molecules [11–13]. Other compounds, mostly secondary metabolites, have been found to inhibit the embryogenic response of the cells. For instance, in Daucus carota, SE is inhibited by a chemical compound of low molecular mass that is released in response to high density cell cultures [14]. This inhibitory compound was identified as 4-hydroxybenzyl alcohol (4-HBA) [15] and its accumulation in embryogenic cell cultures suppresses the rapid division of cells, mainly at early globular stages, inhibiting SE. Subsequently, it was observed that during embryogenic induction in Larix leptolepis, some inhibitory factors present in the medium culture also affected the SE response [16]. These inhibitory molecules were isolated and identified as vanillyl benzyl ether (VBE) [17] and 4-[(phenylmethoxy) methyl] phenol (4-PMP) [18]. VBE causes aberrations, particularly in the suspensor, which arrest the nutrient supply and growth regulator flux, inhibiting the development of somatic embryos in L. leptolepis. Besides VBE, the presence of 4-PMP in the culture medium strongly suppresses SE [18]. The molecular mechanism by which secondary metabolites affect the SE response is not understood. However, it seems that epigenetic modulation could be involved [19–22].

It has been shown that DNA methylation is important for somatic embryo development [23–26]. For instance, in Pinus nigra, the higher levels of DNA methylation correspond to a non-embryogenic line, but low levels promote SE [27]. Similar events have also been reported in other species, such as Rosa x hybrid [28] and Eleuterococcus senticosus [29]. Furthermore, in both D. carota and Cucurbita pepo, the formation of embryogenic cells is related to an increase in DNA methylation levels [23,24]. It was also found that during the generation of pro-embryogenic mass (Pm), the methylation of DNA decreases, but it gradually reverts in accordance with embryo development [23]. More recently, similar events were observed during the SE of C. canephora [26], where the maturation of the embryo was marked by an increase in the levels of DNA methylation. In contrast, disturbances in DNA methylation patterns caused by 5-azacytidine (5-AzaC, demethylating agent) cause the loss of embryogenic potential in cultures of Medicago truncatula and D. carota [30,31] as well as in the early stages of C. canephora SE [26]. On the other hand, it has been shown that there exists a negative relation between phenolic compounds and the DNA methylation levels [32]. Several plant-derived compounds have been proposed as DNA methylation modifiers, including polyphenols, alkaloids, organosulfur compounds and terpenoids, which have been found to have important functions [21,22]. However, the role of secreted molecules into the media has not been fully addressed.

C. arabica and C. canephora are the most important species of the genus Coffea. Some of the distinctive characteristics of these species are that C. canephora is resistant to some diseases, while C. arabica produces coffee of high quality. These species combined produce 100% of the commercial coffee around the world. The biology of the species makes them incompatible and unable to produce hybrids. However, plant tissue culture can be used to resolve this issue.

SE of C. arabica has been explored by several groups around the world [33–36]. The systems used by these groups take a large amount of time to produce somatic embryos. We have developed a SE method that produces viable embryos within a few weeks in C. canephora [37]. However, under the same induction conditions, C. arabica is not able to produce somatic embryos.
This background provides evidence that compounds released by somatic cells could determine whether the embryogenic process succeeds or not, somehow affecting DNA methylation. Therefore, the central aim of the present study is to assess whether the secondary metabolites secreted by *C. arabica* into the culture medium affect the SE response and DNA methylation in *C. canephora*, a highly embryogenic species. We found that the SE of *C. canephora* is disrupted by the metabolites secreted from *C. arabica*, which also affect DNA methylation.

### Results

#### Differential response to somatic embryogenesis in coffee

In the present work, the plantlets from both species were subjected to the same *in vitro* preconditioning treatment with the growth regulators 1-naphthalene acetic acid (NAA) and kinetin (KIN), and then the explants were incubated with 6-benzyladenine (BA) to induce SE (see Material and Methods; Fig 1). After 56 days under these conditions, SE development in both species was compared. In *C. arabica* (Fig 2A), the first morphological change observed in the explants, around 14 days after induction (dai), was a small amount of growth of dedifferentiated tissue around the explant, which was most visible between 21 and 28 dai. At 35 dai, an increase in the growth of dedifferentiated tissue was observed, but from day 42 to day 56 dai, cell proliferation stopped and both the callus-like structures and the explants showed a rapid phenolic oxidation and died. However, in the case of *C. canephora* (Fig 2B), the SE response was very efficient and the first changes in the explants were observed between 7 and 14 dai, with a thickening in the edge of the explant and the beginning of cell proliferation, which quickly increased between 21 and 28 dai, generating the so-called Pm. The high rate of cell division in Pm allowed the start of the first embryogenic stage, the globular (G), at 35 dai. The development of the next embryogenic structures, heart (H) and torpedo (T), were predominant from 42 to 49 dai and finally, at 56 dai, the cotyledonary (C) embryos were completely developed.

In order to evaluate SE development in both species more closely, the SE process was observed every week by scanning electron microscopy (SEM) (Fig 3). In the case of *C. arabica* (Fig 3A), while cell proliferation starts at seven dai, at 14 days the cellular growth is more evident, and it can be observed that the growth initiates in the wounded cells. Between 28 and 35 dai, the accumulation of cell proliferation begins to break the epidermal cells of the explant, and between 42 to 49 dai, it is possible to observe semi-organized structures, which are more evident at 56 dai. These structures could correspond to early embryogenic stages such as globular-like structures. However, we observed several morphological defects; for instance, the presence of non-embryogenic cells as well as the absence of protoderm (Fig 3A, 56 dai), which is crucial to normal embryo development [8].

On the other hand, when we observed, through SEM, the development of SE in *C. canephora* (Fig 3B), we found interesting similarities and important differences in comparison with *C. arabica* (Fig 3A). For instance, SE in *C. canephora* proceeds similarly to *C. arabica* during the first 14 dai. However, cell growth and proliferation in *C. canephora* was more active and abundant, allowing the establishment of the Pm between 21 and 28 dai. At day 35 dai in *C. canephora*, the cells’ organization leads to the formation of the G stage, which is characterized by its spherical form and the presence of the protoderm. Then, by 42 dai, it can be observed that the embryos previously in the G stage have developed into H embryos, the Pm has increased, and the generation of new G embryos has also increased. During the last two weeks, important changes were observed in the SE of *C. canephora*. First, at the H stage, the elongation of the embryo and the development of cotyledonary primordia allow the formation of the T stage, which is the quickest transition into the C embryo stage. Since SE is an asynchronous
process, by 56 dai it is possible to observe all the embryogenic stages, and yet it is possible to distinguish the proliferation of new G embryos (Figs 2B and 3B).

Compounds secreted by *Coffea arabica* explants inhibit SE in other species

Previous reports have indicated that one of the reasons why an explant does not respond to *in vitro* culturing is the secretion of inhibitory compounds into the culture medium [38,39]. In this context, we wondered whether the impaired SE in *C. arabica* is due to extracellular compounds released into the medium. Therefore, we added 21 dai *C. canephora* explants into the
medium of 7 dai C. arabica and co-cultured them together for two weeks (Fig 4B; See Fig 1, step 1). Surprisingly, we found that the co-culture of C. arabica and C. canephora negatively affects the development of the SE process in C. canephora; this negative effect was reflected by a significant reduction in cell proliferation of the Pm and a rapid phenolic oxidation around the explant, which was more evident three weeks after the co-culturing (Fig 4C). This finding strongly suggests that the explants of C. arabica release inhibitory molecules into the culture medium, impairing even a highly embryogenic process.

To determine whether the inhibitors in the conditioned medium (CM) of C. arabica and, therefore, their effects accumulate during the period of culture, we cultured explants of C. canephora of 21 dai in CM of C. arabica of 7, 14 and 21 dai without any additional treatment (Fig 5; See Fig 1 step 2). The embryogenic response of C. canephora was evaluated every seven days (Fig 5A), and the results of the formation of total somatic embryos on the CM were accounted for after 35 days of incubation (Fig 5B). It was found that the CM of C. arabica at any time point was able to inhibit the embryogenic process in C. canephora. The CM from seven dai inhibited the SE of C. canephora more strongly (~93%), while the CM from 14 and 21 days reduced the embryogenic response by 80% in comparison with the control (Fig 5B). Additionally, in order to know whether the inhibitory compounds were heat-resistant, the CM
Identified Factors that Lead to Embryogenic Repression

from 7, 14 and 21 dai was autoclaved (+Δ) and used as in the previous experiment. Under this condition, a small increase in the embryogenic response was observed, such as a gradual increase in the number of somatic embryos, depending on the age of the CM used. For instance, when the CM was heated, the amount of *C. canephora* embryos in the H stages was higher in comparison with the CM that was not heated, although the embryos at T or C stages did not change in either condition (Fig 5B), indicating that the secreted inhibitory compounds from the CM of *C. arabica* are heat-resistant.

Since when we add CM we are also adding inorganic salts, we performed an experiment to rule out the effect of the inorganic components of the Yasuda medium. We extracted the CM with ethyl acetate and used the components in the organic phase to inhibit SE in *D. carota* (S1 Fig). The inhibitory effect was the same as that of the CM without the ethyl acetate partition, suggesting that the inhibition was not due to the presence of additional inorganic salts.

It has been reported that chemical factors implicated in the impairment of SE are substances of low molecular weight [14,16]. Therefore, in order to test this theory, the CM of *C. arabica* from 7, 14, 21 and 28 dai was separated by molecular mass, through a 5 kDa cut-off membrane...
The LmmCM was added separately to the embryogenic culture of 21 dai C. canephora, and the embryogenic response was evaluated at day 35. It was observed that the presence of LmmCM from 7 dai dramatically reduced the embryonic response, up to 95.5%, whereas the LmmCM from 14, 21 and 28 dai caused an inhibition between 81% and 83%, respectively (Fig 6B). In the case of the embryogenic culture of D. carota, the LmmCM was added at the beginning of the embryogenic induction, and the embryogenic response was evaluated after 14 days. The embryogenic response in D. carota was also inhibited (Fig 6A), particularly the LmmCM collected from 7 dai, which strongly stops the development of the embryogenic structures (Fig 6C). These results indicate that the inhibitory molecule(s) present in the CM of C. arabica can...
impair the embryogenic development of highly embryogenic cultures such as *C. canephora* and *D. carota*.

Identification of the chemical nature of the secreted compounds by *Coffea arabica* explants

In order to identify which compounds are present in the LmmCM fraction from seven dai, we analyzed, by GC-MS and UPLC-ESI-ITMS, the CM from *C. arabica* as described in Materials and Methods. It was found by GC-MS analysis that the LmmCM is composed of one prominent compound, which was identified as caffeine, whereas hydroxybenzoic acid and trans-cinnamic acid showed lower signal intensity (S2 Fig). The analysis of the LmmCM fraction by UPLC-ESI-ITMS led to the identification of the eight compounds listed in Table 1. As all of the compounds were tentatively identified as phenolics, we included 34 phenolic compounds as standards (S1 Table). The compounds 1–32 were run in negative ionization mode, whereas compounds 27, 33 and 34 were operated in positive ionization mode. Using the standards as a guide, as well as the retention time and extracted ion chromatograms, we detected and

| Phenolic compound       | LmmCM1 (μmoles флask⁻¹) | LmmCM2 (μmoles флask⁻¹) |
|-------------------------|-------------------------|-------------------------|
| Benzoic acid            | 0.26                    | 0.39                    |
| Caffeine                | 11.042                  | 10.982                  |
| Caffeic acid            | 0.017                   | 0.039                   |
| Catequine               | 0.034                   | 0.087                   |
| Chlorogenic acid        | 1.715                   | 1.986                   |
| Epicatequine            | 0.029                   | 0.050                   |
| Ferulic acid            | 0.0178                  | 0.024                   |
| Salicylic acid          | 0.186                   | 0.275                   |

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quantified eight phenolic compounds (Fig 7) from two independent experiments. Among the identified compounds, the benzoic, ferulic, salicylic and caffeic acids, as well as catequine and epicatequine, were present in low concentrations in the LmmCM, whereas chlorogenic acid and caffeine represent 98% of the phenolic compounds identified (Table 1).

Effect of caffeine, chlorogenic, hydroxybenzoic and trans-cinnamic acids on the formation of somatic embryos on *C. canephora*

We tested whether the two most abundant metabolites, caffeine and chlorogenic acid, as well as hydroxybenzoic acid and trans-cinnamic acids found in very low concentration, exert a negative response on the SE of *C. canephora*. Caffeine, chlorogenic acid, hydroxybenzoic acid and trans-cinnamic acid were added (Fig 1, step 4) in the induction medium of *C. canephora* at different concentrations (1, 10, 100 and 1000 μM; Fig 8). It was found that the addition of 1 μM of caffeine into the induction medium of *C. canephora* reduced the embryogenic potential by 41.3% (Fig 8A) whereas at 10 μM and 100 μM, the inhibition was 75 and 81%, respectively. Interestingly, it was observed that caffeine at the highest concentration (1,000 μM) still allowed the formation of some embryos. In the case of chlorogenic acid, its addition to the medium at concentrations of 1 and 10 μM caused a reduction in the embryogenic response of 52 and 65%, respectively, compared with the control (only with DMSO) (Fig 8B). The data show that the presence of chlorogenic acid in the induction medium strongly inhibited the transition of the H stages to T and C stages, while the same compound added at 100 and 1,000 μM caused a total SE inhibition. In addition, the presence of hydroxybenzoic acid at concentrations of 1, 10 and 100 μM moderately decreased the total number of somatic embryos (3.4, 27.6 and 41.3%, respectively) in comparison with the control (Fig 8C), whereas trans-cinnamic acid at the same concentrations has a more inhibitory effect on SE, mostly in the T and C stages (Fig 8D). However, it was also observed that both compounds added at 1,000 μM totally abolished embryo formation.
LmmCM inhibits SE and decreases DNA methylation in *Coffea canephora*

Recent reports have shown a correlation between DNA methylation, one of the most studied epigenetic mechanisms, and the embryogenic capacity in plants [25,31,40,41]. It has also been documented that plant-derived compounds, such as polyphenolic compounds, have the ability to act as DNA methyltransferase inhibitors [22]. In our analysis several phenolic compounds were detected from LmmCM (Fig 7, Table 1). Therefore, we wondered whether the inhibition of SE in *Coffea canephora* by LmmCM is due to changes in DNA methylation. We analyzed global DNA methylation of both *C. arabica* and *C. canephora* explants every seven days, under embryogenic conditions, for 49 days (Fig 9A). It was observed that during the embryogenic induction process of *C. canephora*, 5-methyl-2'-deoxycytosine (5m dC) levels were higher than those of *C. arabica*, except for day 21, when both species had 22.5% DNA methylation (Fig 9A). Day 21 is the day related to the beginning of cell proliferation on the edge of the explant (Figs 3 and 4). However, unlike *C. canephora*, which maintains DNA methylation levels during the SE process, *C. arabica* presented a rapid decrease in DNA methylation levels, from 22.5% to 12% (Fig 9B).
9A). Once we indeed found changes in DNA methylation levels between *C. arabica* and *C. canephora* (Fig 9A), we assessed whether the LmmCM of *C. arabica* from 7 dai affects the DNA methylation in *C. canephora* at 14, 21 and 56 days after the induction of SE (Fig 9B). We compared the effect of the LmmCM with that of 5-AzaC at 10 μM (Fig 1, step 5). It was observed that the effect of LmmCM on DNA methylation was higher than that of 5-AzaC 10 μM at 14 and 21 dai, and this effect remained after 56 dai.

**Discussion**

SE is a powerful and useful tool to understand the morphogenetic processes that occur during the development of plant embryogenesis [3]. Although the molecular changes by which somatic embryos are generated have been investigated [42–46], the mechanism by which the somatic cell transforms into the somatic embryo is still unknown. SE induction in some species seems to be more responsive to the embryogenic pathway than in others, even those from the same genus [47]. It has been proposed that the failure of SE induction in some plants is due to the secretion of compounds into the medium [15,17,18,39,48]. Therefore, we evaluated the SE response of *C. canephora* (a highly direct embryogenic species) in the presence of *C. arabica* (a deficient direct embryogenic species) explants (Fig 4), and it was observed that in the presence of *C. arabica*, SE in *C. canephora* was disrupted. This finding indicates that some substance in the surrounding medium of *C. arabica* affects SE in *C. canephora*. Some reports have even highlighted that the inhibition of SE development is produced by the accumulation of low molecular mass molecules in cultures with high cell density [14,16]. Our results show that in *C. arabica*, the SE-inhibitory compounds accumulate in the culture medium as early as seven dai (Fig 5), and they have a molecular mass lower than 5 kDa (Fig 6). In cultures of carrot and *L. leptolepis*, low molecular mass compounds such as 4-HBA, VBE and 4-PMP were identified as...
inhibitors of the SE response, suppressing rapid cell divisions and the development of the sus-
pensor, respectively [15,17,18]. Therefore, we tested the 4-HBA to see whether this compound,
which was not detected in any of the samples tested (S3 Fig), affects the SE of *C. canephora* (S4
Fig). We found that 4-HBA moderately reduced the number of embryogenic structures in *C.
canephora* when it was added at the beginning of the induction, in comparison to the response
of the LmmCM isolated from *C. arabica*. Unlike 4-HBA, which seems to be very specific to car-
rot, the LmmCM of *C. arabica* has a more broadly negative effect, as was demonstrated in car-
rot as well as coffee explants (Fig 6).

Phenolic compounds can accumulate in the culture medium, interfering with the SE process
[39,49]. Therefore, from the metabolites identified by GC-MS and UPLC-ESI-ITMS (Fig 7,
Table 1 and S2 Fig), we selected four compounds (caffeine, chlorogenic acid, hydroxybenzoic
acid and trans-cinnamic acid), two very abundant in the LmmCM and two present in low
amounts, and evaluated them in a pure form during the embryogenic response of *C. canephora*
(Fig 8). We found that caffeine, even at the lowest concentration of 10 μM, strongly decreases
the SE process in *C. canephora* (Fig 8A) and this compound is secreted into the culture medi-
um, particularly in *C. arabica* (Fig 7). In contrast to chlorogenic acid, hydroxybenzoic acid and
trans-cinnamic acid, caffeine even at the highest concentration (1,000 μM) allows somatic em-
bryos to develop in *C. canephora* (Fig 8A). Similar results have also been observed during the
development of zygotic embryos in coffee; these embryos have the ability to avoid caffeine
autotoxicity [50]. The growth of the hypocotyl allows the root tip to move away from the caf-
eine-rich endosperm and facilitates the initiation of cell divisions in the root tip. On the other
hand, caffeine accumulates in the cotyledons only after the cell division ends there, suggesting
that the accumulation of caffeine is separated by space and time as a way to avoid autotoxicity
[50]. At the molecular level, it has been found that the application of caffeine during the growth
of the root meristem of *Allium cepa* induces chromosomal aberrations during cell proliferation,
especially during the G2 phase and the mitotic prophase [51]. Furthermore, several studies in-
dicate that this compound also inhibits cell plate formation during cytokinesis in plants [52–
54].

It is worth noting that when chlorogenic acid was applied to *C. canephora* cultures, it was
observed that its presence drastically decreased embryo production, up to 52% with a concen-
tration of 1 μM (Fig 8B). This compound also interrupts embryo development beyond the H
stage, whereas at concentrations ≥ 100 μM the SE process was completely inhibited. At pres-
ent, the physiological role of chlorogenic acid in SE is not clear; however, it is known that high
concentrations of caffeine are accompanied by a considerable accumulation of chlorogenic
acid, as a pathway to form a complex to store the caffeine in the vacuole [55]. It was also ob-
served that during coffee germination the concentration of chlorogenic acids decreases and the
caffeine is released, whereas during leaf expansion an accumulation of both compounds occurs
[56].

It was found that hydroxybenzoic acid at 100 μM concentration moderately inhibited SE
(Fig 8C), indicating that this compound is not the responsible for the inhibition found in the
SE of *C. canephora*. This result is consistent with previous reports, which indicate that the pres-
ence of hydroxybenzoic acid in the embryogenic cell suspension of carrot slightly decreases so-
matic embryo formation [48,57]. Moreover, the addition of trans-cinnamic acid to the cultures
of *C. canephora* at the same concentration (100 μM) caused an important reduction in the for-
mation of embryogenic structures, principally at the T and C stages, which are practically sup-
pressed in comparison to the other treatments used (Fig 8D). Similar observations were also
reported by Liu et al. [58], who found that embryos in the globular stage of *Brassica juncea*
treated with trans-cinnamic acid (an auxin polar transport inhibitor) induce the fusion of coty-
ledons. This suggests that the correct polar transport of auxin from the early globular embryo
stages is crucial for their transition to more mature embryogenic stages [59]. In addition, it has been found that hydroxycinnamic acid is accumulated principally in lines susceptible to browning, affecting SE in date palm [60]. On the other hand, the reduction of the cinnamic acid content stimulates cell division and promotes the development of early embryo structures in oak [61]. According to these results, a high concentration of trans-cinnamic acid, like that used with *C. canephora* (Fig 8D) could be related to the phenolic oxidation of the explants, leading to aberrations at the beginning of Pm development and preventing the generation of somatic embryos. It has been also suggested that hydroxycinnamic acid as well as the chlorogenic acid could be required to reinforce the cell wall, since these compounds, and their derivatives, are necessary to the lignification process [56,62]; probably modifying cell plasticity and interfering with the embryogenic response [60]. The results suggest that differential accumulation of phenolic compounds, in the culture medium of *C. arabica*, could be a major cause of the interruption in the cell division pattern affecting cellular proliferation and the SE process.

On the other hand, the regulatory mechanism by which secondary metabolites interfere with the SE process is still unknown. However, it has been reported that polyphenols, alkaloids, terpenoids and catechins, among others, can induce the loss of DNA methylation [21,22,32]. Our analysis allowed us to determine that LmmCM is composed of caffeine and chlorogenic acid, which both represent 98% of the phenolic compounds in the CM, as well catechin, epicatechin, and caffeic acid (Table 1), compounds that have been shown to have similar effects on DNA methylation [21,63,64]. Therefore, we assessed whether the LmmCM has any effect on DNA methylation and, thus, could be a mechanism by which SE is prevented in coffee (Fig 9).

It was found that there is an important difference in the methylation levels between *C. canephora* and *C. arabica* (Fig 9A). *C. arabica* presented a 5% less DNA methylation rate than *C. canephora* at seven days after induction of SE. This result suggests that the inhibitory molecules present in the LmmCM could stimulate directly or indirectly the loss of DNA methylation (even more than 5-AzaC, Fig 9B), by a mechanism that needs to be further investigated.

It has been observed that caffeic acid and chlorogenic acid, two phenols isolated from coffee plants, inhibit the activity of DNA methyltransferases, particularly through a non-competitive mechanism due to an increase in the levels of S-adenosyl-L-homocysteine (SAH, an inhibitor of DNA methylation) [21]. Furthermore, the accumulation of SAH has also been reported during the biosynthesis of caffeine [65]. Since DNA methylation and the biosynthesis of caffeine require the same reserves of SAM, the reduction of SAM to SAH can generate a feedback inhibition of various SAM-dependent methylation processes [21]. In addition, there are reports indicating that other phenolic compounds can also inhibit DNA methyltransferase activity [21,22]. These findings suggest that the molecules present in the LmmCM, mainly phenolic compounds, could interfere with the activity of DNA methyltransferases in *C. arabica* and, therefore, disrupt the SE process. However it could also be possible that the decrease in DNA methylation is a consequence of the impaired SE process, with the phenolics acting upstream.

In conclusion, our results provide evidence that extracellular molecules of low molecular mass can negatively modulate the SE response and affect DNA methylation. Moreover, we showed that the release of caffeine and chlorogenic acid into the culture medium of *C. arabica* strongly impaired the SE process in *C. canephora*. Further investigations are needed to determine how these compounds affect DNA methyltransferases in this plant, and which genes are epigenetically regulated.

### Materials and Methods

#### Plant materials and growth conditions

The buds used in this study were collected from plants cultivated in our research institute, Centro de Investigación Científica de Yucatán (Scientific Research Center of Yucatan), in...
consequence no specific permissions were required for these locations/activities. This research
does not involve endangered or protected species.

Segments from buds of C. arabica derived from plants in the field were washed and disinfected with 5% Extran and 6% NaClO₃ and rinsed with sterile distilled water. The sterile buds were cultured in vitro in shoot induction medium MS salts [66] supplemented with 2.96 μM thiamine-HCl, 550 μM myo-inositol, 4.06 μM nicotinic acid, 2.43 μM pyridoxine, 26.64 μM glycine, 31.6 μM cysteine, 116.85 μM sucrose, 62.5 μM BA and 0.25% (w/v) gelrite, pH 5.8 and cultured at 25 ± 2°C under a 16/8h (light/darkness) photoperiod (150 μmol m⁻² s⁻¹). Plantlets obtained from shoot induction were isolated and cultured in MS medium supplemented with 11.86 μM thiamine-HCl, 550 μM myo-inositol, 158 μM cysteine, 8.12 μM nicotinic acid, 87.64 mM sucrose and 0.25% (w/v) gelrite, pH 5.8. C. canephora in vitro plants were cultured under the same conditions as C. arabica.

D. carota seeds were sterilized with 70% ethanol and 20% NaClO₃ for 3 min and rinsed three times with distilled sterile water and cultured in vitro in semisolid B₅ medium [67] for 7 days at 25 ± 2°C under a 16/8h (light/darkness) photoperiod (150 μmol m⁻² s⁻¹). Segments 2–4 mm in length were excised from seedlings’ hypocotyls, and then two grams of tissue were transferred onto liquid B₅ medium supplemented with 11.4 μM 2,4-dichlorophenoxyacetic acid and maintained at 100 rpm under the same conditions described above to generate cell suspension. Cell suspensions were subcultured every 14 days.

Somatic embryogenesis induction

Plantlets of C. arabica and C. canephora were subjected under the same conditions of culture for direct embryogenic induction. Briefly, both species were pre-conditioned for 14 days in MS medium supplemented with 0.54 μM NAA and 2.32 μM KIN under the same growth conditions described above. Leaves of plantlets were cut into segments of 0.25 cm², and five explants were inoculated in 50 mL liquid Yasuda modified medium contained in 250 mL flasks as previously described [37] in the presence of 5 μM BA and cultured at 25 ± 2°C in dark conditions at 55 rpm.

For embryogenic induction of D. carota, small clusters of cells were collected from seven-day-old high-density cell suspension through a stainless steel sieve of 50 μm, and then the cells were washed three times with liquid B₅ medium without 2,4-dichlorophenoxyacetic acid and suspended in this same medium. Finally, 50 μL of packed volume cells were inoculated in 50 mL of this same medium, producing low-density cultures.

Electron microscopy

Leaf explants of both coffee species under embryogenic induction were collected every 7 days during a temporal course of 56 days. Both sets of samples were fixed and treated as described previously [26] and analyzed with a scanning electronic microscope (GEOL JSM 6360 LV).

Bioassays with conditioned medium of Coffea arabica

To evaluate the role of possible factors involved in the inhibition of SE response in C. arabica, we used the following strategies: first, the embryogenic explants of C. canephora 21 dai were co-cultured together with explants of C. arabica in the same medium seven dai (Fig 1). Second, the explants from 7, 14 and 21 dai under embryogenic conditions of C. arabica were removed and the medium (conditioned medium; CM) was used to culture embryogenic explants of C. canephora of 21 dai as shown in Fig 2B. Third, the CM from 7, 14 and 21 days of C. arabica was autoclaved and this medium was used to culture the embryogenic explants of 21 dai of C. canephora. Fourth, the CM of each flask (50 mL of medium) of C. arabica from 7, 14, 21 and
28 dai were fractionated through a membrane of cut-off 5 kDa (Amicon Ultra-15), and the obtained <5 kDa fractions of CM of C. arabica (LmmCM) were lyophilized and resuspended with 5 mL of water and sterilized by filtration with a 0.22-μm membrane. This solution was added into the culture medium of C. canephora at 21 dai and D. carota at the moment of embryogenic induction. C. canephora and D. carota cultured at induction conditions without the CM medium were used as controls and the biological response for each species was evaluated at 35 days and at 14 dai, respectively. The number of somatic embryos of D. carota was determined from three aliquots of 0.5 mL of culture for each flask.

Analytical procedure for the metabolite profiling of LmmCM from C. arabica by UPLC-ESI-MS

Secondary metabolites were extracted from 400 mL of CM, 7 dai of SE in C. arabica. In brief, the culture medium was lyophilized until the volume reached 50 mL and then extracted three times with ethyl acetate. The extract was reduced to one mL and stored at -20°C prior to analysis. All chemicals were of analytical reagent grade (Sigma-Aldrich). HPLC-grade acetonitrile, ethanol (Fisher Scientific) and deionized water (18.2 MΩ cm, Labconco, USA) were used throughout. Formic acid, ammonium formate and phenolic compounds used as standards were Sigma reagents. Metabolite profiling was performed as described elsewhere [68], using an UltiMate 3000 liquid chromatography (Dionex, Thermo Scientific) UHPLC system equipment with a chromatographic column Luna C18 (150 x 2 mm, 3 μm) from Phenomenex and coupled with mass spectrometry detection (an ion-trap mass spectrometer AmaZon SL fitted with ESI source; Bruker Daltonics). The LC–MS system was controlled by Hystar V3.2 software, where the data were processed by Data Analysis V4.1 SP2 and QuantAnalysis V2.0 SP2 (Bruker Daltonics). The mixed standard solutions of the 34 phenolics compounds (S1 Table) containing 0.1, 0.5, 1, 5, 10 and 20 mg L⁻¹ of each compound were prepared in ethanol. All solutions were filtered, the injection volume was 10–30 μL and three replicates were always carried out. Gradient elution with two mobile phases (A—ammonium formate 10 mM + 0.2% v/v formic acid, pH 2.9; B—acetonitrile + 0.2% v/v formic acid) was as follows: 0–1 min 15% B; 1–6 min 50% B; 6–8 min 60% B; 8–11 min 60% B; 11–13 min 90% B; 14–17 min 15% B; column thermostat was set at 30°C and a total flow rate 0.2 mL min⁻¹ was applied. The ESI source was operated in negative ionization mode for the compounds 1–32 and in positive ionization mode for compounds 27, 33, and 34, with the following parameters: alternate spray voltage 4,500 V; plate voltage 500 V; nebulizer gas pressure 26 psi (N₂); dry gas 6 L min⁻¹ (N₂); source temperature 200°C and capillary exit voltage 140 V. The mass spectra were obtained by means of an Ultrascan mode in the m/z scan range 70–400, with an ion charge control (ICC) target setting 100,000 and a maximum accumulation time 100 ms. Tuning was performed for the mixed standard solution of all 34 compounds (0.1 mg L⁻¹ each) using smart parameter setting (SPS); target m/z 200, compound stability 100%, trap drive level 100%. Total ion chromatograms were acquired; base peak and extracted ion chromatograms were generated (m/z window for EIC ± 0.3 Da). For quantification, Bruker QuantAnalysis software was used, calculating the areas under the [M-H]⁻ ions ([M+H]⁺ for positive ESI mode) from respective EIC. External calibration was carried out using a series of mixed standard solutions and the linear quadratic regression fit yielded r² > 0.999 (S1 Table); on-column detection limits evaluated based on signal-to-noise ratio 3:1 were in the range 1–22 ng (10–220 μg L⁻¹ as referred to the injected solution). The two samples were dissolved in 1 mL deionized water, the results presented in Table 1 correspond to the concentrations found in these solutions.
4-hydroxybenzyl alcohol and phenolic compounds assays on *C. canephora* somatic embryogenesis

Embryogenic cultures of *C. canephora* were treated with 4-hydroxybenzyl alcohol (Sigma, H20806) and phenolic compounds: hydroxybenzoic acid (Sigma, H6761), trans-cinnamic acid (Sigma, C6004), caffeine (Sigma, C0750) and chlorogenic acid (Sigma, C3878). Chlorogenic acid was dissolved in dimethyl sulfoxide (Sigma, D4540), while the other compounds were dissolved in the same induction medium. Then, the solutions were filter-sterilized through a membrane of 0.22 μm. 4HBA was added to the medium at the beginning of SE induction (seven dai) at concentrations of $10^{-6}$ M and $10^{-4}$ M and at 14 dai at a concentration of $10^{-4}$ M. Hydroxybenzoic acid, trans-cinnamic acid, caffeine and chlorogenic acid were added to the medium at seven dai at concentrations of 1, 10, 100 and 1,000 μM, and the total number of somatic embryos in every developmental stage was counted at 56 dai.

DNA methylation

Genomic DNA extraction was carried out according to the method described by Echevarría-Machado et al. [69]. Briefly, 100 mg of leaf explants of *C. arabica* and *C. canephora* under embryogenic conditions, as shown in Fig 2, were collected every seven days from 0 to 49 days. The DNA digestion and the separation of nucleosides were performed as described in detail elsewhere [70], and the HPLC analysis was performed using the chromatographic column Kromasil C18 (250 x 4.6 mm, 5 μm from Phenomenex), and an Agilent series 1200 HPLC equipped with a pump quaternary and diode array detector. 5mdC quantification was obtained by applying the formula below to the peak areas: % 5mdC = $C_{5mdC}/(C_{5mdC} + C_{dC}) \times 100$, where $C$ is the concentration of 5mdC and 2′-deoxycytosine (dC).

Effects of LmmCM and 5-azacytidine on DNA methylation during the somatic embryogenesis of *C. canephora*

Embryogenic cultures of *C. canephora* were treated in the absence (control) or presence of 10 μM of 5-AzaC added every seven days from day 7 to day 49, whereas the addition of the LmmCM was done only one time, at day 7 after the induction. Then, the explants from the control and each treatment (5-AzaC and LmmCM) were re-collected at 14, 21 and 56 dai for DNA methylation analysis. An assessment of the global DNA methylation levels under the effects of either 5-AzaC or LmmCM was performed as described above.

Statistical analysis

All the data were processed and analyzed using analysis of variance (ANOVA). The significance grade among the mean values was carried out using the Tukey test. Differences were considered to be significant at $P \leq 0.01$. Data were analyzed using Origin V8 software (Data Analysis and Graphing Software).

Supporting Information

S1 Fig. Comparative effect of the LmmCM and the total molecules extracted with ethyl acetate from the conditioned medium (CM) of *Coffea arabica* on the embryogenic cultures of *Daucus carota*. A. Schematic representation of the experimental procedure. CM of *C. arabica* was separated and extracted as described in Materials and Methods. The two different fractions: the low molecular mass of conditioned medium (LmmCM) and the ethyl acetate phase were added separately to the embryogenic cultures of *Daucus carota*. B. Effects of LmmCM
and ethyl acetate phase in the somatic embryogenesis process of D. carota. (TIF)

S2 Fig. Gas chromatography profiles of compounds extracted from the conditioned medium of 7-day-old C. arabica culture. A. Extraction of the LmmCM fraction with ethyl acetate. Peaks marked as 1, 2 and 3 correspond to the fragmentation patterns of trans-cinnamic acid (B), hydroxybenzoic acid (C) and caffeine (D), respectively. (TIF)

S3 Fig. Gas chromatography profile of the phenolic compounds extracted with ethyl acetate at day 7 from conditioned medium of Coffea arabica. Each peak was identified by mass spectroscopy as shown in S1 Fig. A chromatogram of a standard sample of 4-hydroxymethyl alcohol was overlapped on the chromatogram of the ethyl acetate extracted conditioned medium in order to show the retention time of this compound and its absence in the analyzed sample. (TIF)

S4 Fig. Effect of 4-hydroxybenzyl alcohol on the embryogenic cultures of Coffea canephora. A. *4HBA at 10⁻⁶ M and 10⁻⁴ M was added at the beginning (7 days) of the embryogenic induction of C. canephora. **4HBA at 10⁻⁴ M was added at 14 days after embryogenic induction of C. canephora. B. The number of embryos at different developmental stages was counted at 56 days. The control was cultivated in the absence of 4HBA. Error bars represent the SE (n = 3). Different letters in bars represent the statistical significance of mean differences between each embryogenic stage at a given time by the Tukey test (P ≤ 0.01). The experiment was carried out three times. (TIF)

S1 Table. Analytical parameters and detection limits (DL) evaluated for 34 phenolic compounds based on linear regression calibration and concentration determined in LmmCM samples from C. arabica by UPLC-ESI-ITMS. (DOCX)

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Author Contributions

Conceived and designed the experiments: VMLV CDLP. Performed the experiments: GINC AAM KW RMGA. Analyzed the data: VMLV CDLP GINC KW. Contributed reagents/materials/analysis tools: VMLV CDLP KW. Wrote the paper: VMLV CDLP GINC.

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