Development of a multi-locus sequence typing scheme for Laribacter hongkongensis, a novel bacterium associated with freshwater fish-borne gastroenteritis and traveler's diarrhea

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Abstract

Background: Laribacter hongkongensis is a newly discovered, facultative anaerobic, Gram-negative, motile, sea gull-shaped rod associated with freshwater fish-borne gastroenteritis and traveler’s diarrhea. A highly reproducible and discriminative typing system is essential for better understanding of the epidemiology of L. hongkongensis. In this study, a multilocus sequence typing (MLST) system was developed for L. hongkongensis. The system was used to characterize 146 L. hongkongensis isolates, including 39 from humans and 107 from fish.

Results: Fragments (362 to 504 bp) of seven housekeeping genes were amplified and sequenced. Among the 3068 bp of the seven loci, 332 polymorphic sites were observed. The median number of alleles at each locus was 34 [range 22 (ilvC) to 45 (thiC)]. All seven genes showed very low $d_{x}$/d$_{y}$ ratios of < 0.04, indicating that no strong positive selective pressure is present. A total of 97 different sequence types (STs) were assigned to the 146 isolates, with 80 STs identified only once. The overall discriminatory power was 0.9861. eBURST grouped the isolates into 12 lineages, with six groups containing only isolates from fish and three groups only isolates from humans. Standardized index of association ($I_{A}$) measurement showed significant linkage disequilibrium in isolates from both humans and fish. The $I_{A}$ for the isolates from humans and fish were 0.270 and 0.636, indicating the isolates from fish were more clonal than the isolates from humans. Only one interconnected network (acnB) was detected in the split graphs. The P-value (P = 0) of sum of the squares of condensed fragments in Sawyer’s test showed evidence of intragenic recombination in the rho, acnB and thiC loci, but the P-value (P = 1) of maximum condensed fragment in these gene loci did not show evidence of intragenic recombination. Congruence analysis showed that all the pairwise comparisons of the 7 MLST loci were incongruent, indicating that recombination played a substantial role in the evolution of L. hongkongensis. A website for L. hongkongensis MLST was set up and can be accessed at http://mlstdb.hku.hk:14206/MLST_index.html.

Conclusion: A highly reproducible and discriminative MLST system was developed for L. hongkongensis.

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Background

Laribacter hongkongensis is a newly discovered, facultative anaerobic, Gram-negative, motile, sea gull-shaped rod that belongs to the Neisseriaceae family of β-proteobacteria. It was first recovered from the blood and empyema thoracis of a patient with alcoholic liver cirrhosis in Hong Kong in 2001 [1]. Since the patient's underlying disease and the presence of ascites suggested that the gastrointestinal tract may be a possible source of infection, L. hongkongensis was intensively sought in human fecal specimens. During a period of two months, the bacterium was recovered from the stool of three patients with community-acquired gastroenteritis and traveler's diarrhea [4]. L. hongkongensis is likely to be globally distributed, as travel histories from patients suggested that it is present in at least four continents, including Asia, Europe, Africa and Central America [3,4]. Recently, L. hongkongensis has also been reported from another coastal province in mainland China [5]. In a recent review, L. hongkongensis, together with enterotoxigenic Bacteroides fragilis and Klebsiella oxytoca, were included as newly appreciated agents associated with acute diarrhea [6]. Although the causative role of L. hongkongensis in gastroenteritis is yet to be established [7], these data provide strong evidence that the bacterium is a potential diarrheal pathogen that warrants further investigations.

L. hongkongensis has been found in the intestines of healthy freshwater fish but not other studied animals that are commonly used for cooking in Hong Kong [4,8,9]. The bacterium was recovered from the guts of 24% of 360 freshwater fish studied, with the highest recovery rates from grass carp (60%) and bighead carp (53%) and during spring and summer [6,7]. Moreover, L. hongkongensis has also been recovered from drinking water reservoirs in Hong Kong [10]. The presence of a heterogeneous population of L. hongkongensis by pulsed-field gel electrophoresis (PFGE) among isolates from freshwater fish [9] and the association of L. hongkongensis gastroenteritis with fish consumption [4] suggested that freshwater fish is likely the major reservoir of the bacterium and the source of human infections.

A highly reproducible and discriminative typing system is essential for better understanding of the epidemiology of L. hongkongensis. Previously, we have used PFGE for typing L. hongkongensis [4,7,8]. However, due to experimental variations, PFGE patterns are difficult to compare among different laboratories. As multi-locus sequence typing (MLST) is well known to be highly reproducible and discriminative for bacteria, we developed such a typing system for L. hongkongensis using the sequence information of the L. hongkongensis complete genome sequence project. In this article, we report the development of an MLST scheme for L. hongkongensis using 146 isolates from humans and fish.

Methods

L. hongkongensis isolates

A total of 146 L. hongkongensis isolates, including 39 isolates recovered from humans and 107 isolates from fish, were used in this study (Additional file 1). L. hongkongensis isolate HKU1 was recovered from the blood culture and empyema pus of a patient with bacteremic empyema thoracis [1]. The other 38 isolates from humans (isolates HLHK2 to HLHK39) were recovered from the stool of patients with community-acquired gastroenteritis [2,4]. Isolates HLHK 2–4 were recovered from patients in Switzerland while the rest were from patients in Hong Kong. The 107 isolates from fish were recovered from the guts of freshwater fish sampled from retail food markets in Hong Kong [4,9]. These included 50 isolates (FLHK1–8, FLHK25–26, FLHK36–43, FLHK50–59, FLHK61–71, FLHK77–84 and FLHK94–96) recovered from grass carp (Ctenopharyngodon idella), 42 isolates (FLHK9–14, FLHK27–33, FLHK44–49, FLHK72–76, FLHK85–93, FLHK97–100 and FLHK103–107) from bighead carp (Aristichthys nobilis), 12 isolates (FLHK15–21, FLHK34–35, FLHK60 and FLHK101–102) from mud carp (Cirrhina molitorella) and three isolates (FLHK22–24) from large-mouth bass (Micropterus salmoides). The identification of all L. hongkongensis isolates were confirmed phenotypically by standard conventional biochemical methods and genotypically by 16S rRNA gene sequencing [1,4].

DNA extraction

Bacterial DNA extraction was modified from our previous published protocol [1]. Briefly, 800 μl of NaOH (0.05 M) was added to 200 μl of bacterial cells suspended in distilled water and the mixture was incubated at 60 °C for 45 min, followed by addition of 240 μl of Tris-HCl (pH 7.0), achieving a final pH of 8.0. The resultant mixture was diluted 100× and 0.5 μl of the diluted extract was used for PCR.

PCR amplification and sequencing

Extracted DNA from the 146 isolates of L. hongkongensis was used as the template for amplification of seven housekeeping genes [transcription termination factor Rho (rhu); aconitate hydratase (acnB); cell division protein (ftsH); anthranilate synthase component I (trpE); ketol-acid reductoisomerase (ilvC); thiamin biosynthesis protein ThiC (thiC); enolase (eno)], using primers listed in Table
1. The seven housekeeping genes were chosen because either the gene itself or other genes in the same metabolic pathway has been used in MLST schemes of other bacteria. The sequences of the seven genes were obtained from our on-going L. hongkongensis complete genome sequence project (unpublished data).

The PCR mixture (100 μl) contained denatured L. hongkongensis DNA, PCR buffer (10 mM Tris-HCl pH 8.3 and 50 mM KCl), 2 mM MgCl₂, 200 μM of each deoxynucleoside triphosphates and 2.5 U Ampli Taq Gold DNA polymerase (Applied Biosystems, Foster City, CA, USA). For rho, trpE, ilvC, thiC and eno, the sample was amplified in 40 cycles of 94°C for 1 min, 55°C for 1.5 min and 72°C for 2 min, and with a final extension at 72°C for 10 min in an automated thermal cycler (Applied Biosystems, Foster City, CA, USA). For acnB and ftsH, the sample was amplified using a reannealing temperature of 60°C. Twenty microliters of each amplified product was electrophoresed in 2% (w/v) agarose gel, with a molecular size marker (GeneRuler™ 50 bp DNA ladder, MBI Fermentas, Canada). Electrophoresis in Tris-borate-EDTA buffer was performed at 120 volts for 40 min. The gel was stained with ethidium bromide (0.5 μg/ml) for 15 min, rinsed and photographed under ultraviolet light illumination.

The PCR product was gel-purified using the QIAquick PCR purification kit (QIagen, Hilden, Germany). Both strands of the PCR product were sequenced using BigDye Terminator Cycle Sequencing kit version 3.1 with an ABI Prism 3700 DNA Analyzer according to manufacturers’ instructions (Applied Biosystems, Foster City, CA, USA) and the PCR primers. BioEdit version 7.0.5.2 was used for reading the sequences and aligning the forward and backward reads [11].

### Allele and sequence type assignment

The nucleotide sequences of the seven gene loci used for MLST in all the L. hongkongensis isolates were aligned and compared with those of isolate HLHK1 using Clustal W multiple alignment [12] implemented in BioEdit version 7.0.5.2 [11]. An arbitrary number was assigned to each distinct allele at a locus. The numbered alleles at each locus were combined in order to establish the sequence type (ST) for each isolate. Each ST was numbered in the order of identification (ST-1, ST-2, etc.). The data have been deposited in our Laribacter hongkongensis complete genome sequence and MLST database [http://mlstdb.hku.hk:14206/MLST_index.html](http://mlstdb.hku.hk:14206/MLST_index.html).

### Sequence analysis

The proportions of nucleotide alterations that led to a change in the amino acid sequence (non-synonymous substitution, \(d_a\)) and the proportions of nucleotide alterations that did not lead to a change in the amino acid sequence (synonymous substitution, \(d_s\)) were calculated with START2 [http://pubmlst.org/software/analysis/](http://pubmlst.org/software/analysis/)[13]. Phylogenetic analysis was performed using ClonalFrame algorithm with the software package ClonalFrame version 1.1, using 50,000 burn-in cycles and 100,000 further iterations [14]. Over 500 trees were generated from which a 75% majority-rule consensus tree was derived with MEGA version 4.0 [15]. STs were grouped into lineages with eBURST [16]. The members of an eBURST lineage were defined as groups of two or more independent isolates where each had identical alleles at six or more loci with at least one member of the group. The linkage disequilibrium between alleles at the seven gene loci was measured using the standardized index of association (\(F_{ST}\)) with LIAN 3.5 [http://pubmlst.org/analysis/](http://pubmlst.org/analysis/)[17,18]. Split decomposition analysis was performed using the SplitsTree program (version 4.10) [19]. Sawyer’s test analysis for intragenic recombination was performed with START2 [http://pubmlst.org/software/analysis/](http://pubmlst.org/software/analysis/)[13]. Gene tree congruence analysis was performed using the Shimodaira-Hasegawa (SH) test [20] as implemented in PAUP 4.0b10 using the REL method and 10000 bootstrap replicates [21]. Ninety-seven STs were selected and used in the SH test. Maximum-likelihood trees for each MLST gene of the 97 STs were inferred under a general time-reversible model, with an estimated gamma distribution, using PHYML v3.0 [22].

### Table 1: Primers for amplification and sequencing of the seven housekeeping genes in L. hongkongensis

| Gene locus | Forward | Reverse | Amplicon size (bp) |
|------------|---------|---------|-------------------|
| rho        | LPW868 5'-ATCGTCTCNNTGATTGACGAG-3' | LPW869 5'-GATGTGATGGCCNGGAA-3' | 468 |
| acnB       | LPW870 5'-GCCAGGCCTCTGATTGACGAG-3' | LPW871 5'-GGCNGCCCGGACNTACTG-3' | 561 |
| ftsH       | LPW1712 5'-CGTGCAAACGCCACGGG-3' | LPW1713 5'-ACCGNCGATCTGAC-3' | 480 |
| trpE       | LPW1714 5'-CGTGCAAACGCCACGGGACCA-3' | LPW1715 5'-GGCNGCCCGGACNTACTG-3' | 626 |
| ilvC       | LPW1716 5'-ATCGTCTCNNTGATTGACGAG-3' | LPW1717 5'-GGCNGCCCGGACNTACTG-3' | 473 |
| thiC       | LPW3137 5'-CGCTGCGGGGCGGGAAC-3' | LPW2555 5'-CCAGATCAGCGAGTGG-3' | 557 |
| eno        | LPW866 5'-ACCGNCGATCTGAC-3' | LPW867 5'-GGCNGCCCGGACNTACTG-3' | 517 |
The median number of alleles at each locus was 34 [range 22 to 45] (thiC). Allelic profiles were assigned to the 146 isolates of L. hongkongensis (Additional file 1). The alleles defined for the MLST system were based on sequence lengths of between 362 bp (ilvC) and 504 bp (acnB). The median number of alleles at each locus was 34 [range 22 to 45 (thiC)].

The $d_n/d_s$ ratio for the seven gene loci are shown in Table 2. All seven genes showed very low $d_n/d_s$ ratios of $< 0.04$ (median 0.0154, range 0.0000 – 0.0355), indicating that no strong positive selective pressure is present.

**Results**

**Variation at the seven MLST loci**

Single bands of the expected sizes were observed for each gene locus amplified using the specific primers. Among the 3068 bp of the seven loci, a total of 332 polymorphic sites were observed in the 146 isolates of L. hongkongensis. Two hundred and sixty-five and 246 polymorphic sites were observed in the 39 isolates from humans and 107 isolates from fish respectively. No insertion, deletion or premature termination was observed in any of the polymorphic sites. Allelic profiles were assigned to the 146 isolates of L. hongkongensis (Additional file 1). The alleles defined for the MLST system were based on sequence lengths of between 362 bp (ilvC) and 504 bp (acnB). The median number of alleles at each locus was 34 [range 22 (ilvC) to 45 (thiC)].

The $d_n/d_s$ ratio for the seven gene loci are shown in Table 2. All seven genes showed very low $d_n/d_s$ ratios of $< 0.04$ (median 0.0154, range 0.0000 – 0.0355), indicating that no strong positive selective pressure is present.

**Relatedness of L. hongkongensis isolates**

A total of 97 different STs were assigned to the 146 L. hongkongensis isolates, with 80 of the 97 STs identified only once (Additional file 1). Thirty-eight different STs were observed in the 39 isolates from humans and 62 different STs observed in the 107 isolates from fish, with 29 STs in 50 grass carp, 30 STs in 42 bighead carp, 11 STs in the 12 mud carp and two STs in three large mouth bass. Three STs (ST-7, ST-23 and ST-26) were found in both isolates from humans and fish. The most common ST (ST-41) was identified nine times, followed by ST-42 (eight isolates) and ST-45 (seven isolates). The overall discriminatory power for the 146 isolates was 0.9861, that for the isolates from 39 humans was 0.9987 and for the isolates from fish was 0.9755. ClonalFrame was used to construct a dendrogram using the concatenated nucleotide sequences of the seven gene loci of the 146 isolates (Fig. 1).

Table 2: Characteristics of loci and Sawyer’s test analysis for intragenic recombination in L. hongkongensis isolates

| Locus | Size of sequenced fragment (bp) | No. of alleles identified | No. (%) of polymorphic nucleotide sites | % G + C | $d_n/d_s$ | SCF (P-value) | MCF (P-value) |
|-------|---------------------------------|---------------------------|----------------------------------------|---------|----------|---------------|---------------|
| rho   | 399                             | 31                        | 40 (10.0%)                             | 58.7%   | 0.0000   | 160937 (0)   | 39 (1)        |
| acnB  | 504                             | 39                        | 45 (8.9%)                              | 66.6%   | 0.0043   | 281863 (0)   | 43 (1)        |
| ftsH  | 428                             | 43                        | 46 (10.7%)                             | 63.4%   | 0.0126   | 392301 (0.53) | 43 (1)        |
| trpE  | 448                             | 34                        | 44 (9.8%)                              | 59.4%   | 0.0265   | 174730 (0.46) | 37 (1)        |
| ilvC  | 362                             | 22                        | 16 (4.4%)                              | 58.3%   | 0.0154   | 11688 (0.55) | 14 (1)        |
| thiC  | 473                             | 45                        | 101 (21.4%)                            | 63.3%   | 0.0355   | 954286 (0)   | 92 (1)        |
| eno   | 454                             | 34                        | 40 (8.8%)                              | 60.5%   | 0.0266   | 118330 (0.18) | 33 (1)        |

$^a$SSCF, sum of the squares of condensed fragments

$^b$P-value indicating statistically significant (P < 0.05) evidence for recombination are marked with asterisks

$^c$MCF, maximum condensed fragment

eBURST grouped the isolates into 12 lineages, with 14 STs in group 1, 12 STs in group 2, seven STs in group 3, three STs in groups 4–6 and two STs in groups 7–12, whereas 43 STs did not belong to any of the 12 groups (Fig. 2 and Additional files 1 and 2). These 43 singleton STs were isolated from 25 patients and 19 fish (one ST was found in both). All these 12 groups were also observed as clusters in the dendrogram (Fig. 1). Groups 2, 3, 7, 8, 11 and 12 contained only isolates from fish, group 1 contained 34 isolates from fish and two isolates from humans, group 4 contained three isolates from fish and one isolate from human, group 9 contained one isolate from fish and two isolates from humans, and groups 5, 6 and 10 contained only isolates from human. $P_A$ measurement showed significant linkage disequilibrium in both isolates from humans and fish. The $P_A$ for the isolates from humans and fish were 0.270 (0.243 if the three isolates from Switzerland were removed and 0.251 if the allelic profiles of the 38 unique STs of the isolates from humans were used) and 0.636 (0.469 if the allelic profiles of the 59 unique STs of the isolates from fish were used), indicating that the isolates from fish were more clonal than the isolates from humans. Only one interconnected network (acnB) was detected in the split graphs (Fig. 3). The P-value (P = 0) of sum of the squares of condensed fragments in Sawyer’s test showed evidence of intragenic recombination in the rho, acnB and thiC loci, but the P-value (P = 1) of maximum condensed fragment in these gene loci did not show evidence of intragenic recombination (Table 2). Congruence analysis showed that all the pairwise comparisons of the 7 MLST loci were incongruent, indicating that recombination played a substantial role in the evolution of L. hongkongensis. (Table 3).

No relationships were observed among the L. hongkongensis isolates with respect to their years of isolation; the locations of the hospitals, age and sex of the patients and the presence of plasmids in the isolates from patients [23]; nor to the species of the fish and the locations of the markets where the fish were purchased.
Figure 1
Phylogenetic tree showing the relationships of the 97 STs of *L. hongkongensis* in this study. The genetic relatedness among the 97 STs was assessed by ClonalFrame algorithm based on the pair-wise differences in the allelic profiles of the seven housekeeping genes. Numbers immediately to the right of the dendrogram show the eBURST clonal clusters to which the STs belong.
Discussion

A highly discriminative MLST scheme was developed for *L. hongkongensis*. Seven housekeeping genes with very low \( d_{s}/d_{r} \) ratios of the range of 0.0000 – 0.0355, similar to the housekeeping genes in other MLST schemes, were employed to produce a highly discriminative MLST scheme, with discriminatory power of 0.9861, comparable to the MLST schemes of other pathogenic bacteria, for molecular typing of *L. hongkongensis*. When the same *L. hongkongensis* isolate was subcultured 50 times, no difference was observed between the sequences of the seven gene loci in the original isolate and the one after 50 subcultures (data not shown). Therefore, these seven loci are discriminative enough for typing, but not evolving too rapidly to an extent that will mask genetic relatedness, as in the case of *Helicobacter pylori*, another urease positive, S-shaped and motile alimentary tract microbe [24,25].

The *L. hongkongensis* isolates recovered from fish were clustered. In our previous study on ecoepidemiology of *L. hongkongensis* using PFGE of *Sph*I digested chromosomal DNA, it was found that the isolates from fish and humans were clustered separately [9]. Analysis of the sequences of the seven gene loci using both dendrogram and eBURST groups revealed a similar phenomenon to the previous ecoepidemiology study, although the clustering pattern of the isolates in the present study was different from that in the previous one (data not shown). eBURST group analysis showed that six of the 12 groups consisted exclusively of isolates from fish, whereas three of the 12 groups consisted exclusively of isolates from humans (Fig. 2). All
these 12 eBURST groups were also found in clusters in the dendrogram (Fig. 1), although $F_{A}$ measurement showed that the isolates from fish were probably more clonal than the isolates from humans. All these results of clustering of isolates from fish and humans into different groups observed in both the previous PFGE and the present MLST studies suggested that some clones of L. hongkongensis could be more virulent than others.

Although the isolates from fish appeared more clonal than the isolates from humans, a heterogeneous population of L. hongkongensis existed in the same ecosystem. STs recovered from the same species of fish or the same fish market did not cluster together. Over 80% of freshwater fish consumed in Hong Kong are imported from fish farms in mainland China, whereas the remaining 20% are locally reared in fish farms in rural areas of Hong Kong.

Table 3: Shimodaira-Hasegawa test for congruency among tree topologies for the seven loci and their concatenated sequence

| Locus | Concatenation | rho | acnB | ftsH | trpE | ilvC | thiC | eno |
|-------|---------------|-----|------|------|------|------|------|-----|
| rho   | 0.0001*       | 0.0000* | 0.0000* | 0.0000* | 0.0000* | 0.0000* | 0.0000* |
| acnB  | 0.0000*       | 0.0000* | 0.0000* | 0.0000* | 0.0000* | 0.0000* | 0.0000* |
| ftsH  | 0.0003*       | 0.0002* | 0.0002* | 0.0003* | 0.0002* | 0.0002* | 0.0003* |
| trpE  | 0.0001*       | 0.0000* | 0.0001* | 0.0000* | 0.0000* | 0.0000* | 0.0000* |
| ilvC  | 0.0075*       | 0.0090* | 0.0064* | 0.0048* | 0.0056* | 0.0059* | 0.0072* |
| thiC  | 0.0000*       | 0.0000* | 0.0000* | 0.0000* | 0.0000* | 0.0000* | 0.0000* |
| eno   | 0.0008*       | 0.0003* | 0.0008* | 0.0003* | 0.0008* | 0.0008* | 0.0008* |

*P values (*, P < 0.05) represent differences in likelihood score between the maximum likelihood topology of each locus.
Since the same species of freshwater fish in a particular market is usually obtained from the same fish farm and multiple STs were present in *L. hongkongensis* isolates recovered from the same species purchased from the same market, it implied that multiple clones of *L. hongkongensis* probably existed in the same aquaculture farm in mainland China or Hong Kong.

**Conclusion**

Seven housekeeping genes with very low $d_{s}/d_{i}$ ratios were employed to produce a highly discriminative MLST scheme for molecular typing of *L. hongkongensis*.

**Abbreviations**

bp: base pairs; $d_{s}$: non-synonymous substitution; $d_{i}$: synonymous substitution; DNA: deoxyribonucleic acid; $I_{S_A}$: standardized index of association; M: molar; min: minutes; ml: milliliter; MLST: multilocus sequence typing; PCR: polymerase chain reaction; PFGE: pulsed-field gel electrophoresis; ST: sequence type; μl: microliter.

**Authors’ contributions**

PCYW conceived the study and drafted the manuscript. PCYW, JLLT, SKPL and KYY participated in the design of the study. PCYW and JLLT supervised the study. PCYW, JLLT and AKLT analyzed the data. HT constructed the database and website. KMC, EKYL, JKHC, SSLM, DMWT and LMWC carried out the PCR and sequencing experiments. SKPL and KYY corrected the manuscript. All authors read and approved the final manuscript.

**Additional material**

Additional file 1  
**Characteristics of *L. hongkongensis* isolates used in the present study.** The tabulated data describe the background epidemiological and MLST characteristics of the 146 *L. hongkongensis* isolates in this study. Click here for file [http://www.biomedcentral.com/content/supplementary/1471-2180-9-21-S1.doc](http://www.biomedcentral.com/content/supplementary/1471-2180-9-21-S1.doc)

Additional file 2  
**eBURST groups of *L. hongkongensis* isolates.** The tabulated data provide the detailed compositions of each eBURST group of *L. hongkongensis* isolates. Click here for file [http://www.biomedcentral.com/content/supplementary/1471-2180-9-21-S2.doc](http://www.biomedcentral.com/content/supplementary/1471-2180-9-21-S2.doc)

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