Signatures Based On Tumor Microenvironment Genes Can Reliably Predict The Prognosis of Laryngeal Cancer Patients

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Abstract

**Purpose:** To develop a tumor microenvironment (TME) related genes based prognostic model for laryngeal cancer patients risk prediction.

**Methods:** A innovative prognostic model was generated based on TME related genes (760 genes). Based on the model, laryngeal cancer patients were categoried into high and low-risk group. The immune status including immune cell infiltration ratio as well as checkpoints have been explored.

**Results:** It can be shown here 15 genes demonstrate significant differences, which can better predict laryngeal cancer patient prognosis. The accuracy of the model prediction is evaluated and approved by the AUC value. From the immune cell infiltration ratio analysis, there is a significant difference in the infiltration degree of several types of immune cells and 6 immune checkpoints between high and low-risk laryngeal cancer patients. At the same time, the close related genes as well as TME pathways have been also investigated.

**Conclusion:** This study has explored a potential prognostic biomarker and developed a novel TME-associated prognostic model for laryngeal cancer, which provides a valuable reference for future clinical research.

Introduction

The incidence of laryngeal cancer accounts for about 1 to 5% of human systemic tumors[1]. In the field of otolaryngology, it is second only to nasopharyngeal cancer and nasal cavity as well as sinus cancer[2]. Laryngeal cancer occurs more commonly in men than in women (5.8 cases per 100,000 vs 1.2 per 100,000, respectively)[3]. The incidence of laryngeal cancer has a male to female ratio of 5:1. With the decrease in tobacco use, the incidence of laryngeal cancer has been decreasing 2.4% each year for the last 10 years[4]. However, the 5-year survival rate of 60.9% has changed little over the past several years. The major risk factor for laryngeal cancer is tobacco[5]. In addition to tobacco, there are still some other factors that can induce the development of laryngeal cancer Laryngopharyngeal reflux, human papillomavirus infection, environmental or occupational exposures, and alcohol are some other major contributors to laryngeal cancer development[6–8]. Treatment for laryngeal cancer has evolved from radical resections and intensive radiation or chemoradiation. Despite significant advances in early diagnosis and multidisciplinary cancer treatment, the long-term prognosis is still far from satification[9]. Therefore, it is urgent to identify the sensitive and specific molecular markers in patients with laryngeal cancer.

Recent studies have shown that the tumor microenvironment (TME), composed of a variety of immune cells and stromal cells, is critical in the occurrence and development of cancer[10]. The multiple immune cells, according to the innate and adaptive systems, as well as other cytokines and signaling molecules, contribute to the tumor microenvironment. Although some immune cells possess antitumoral properties,
others function to suppress antitumor immunity and are manipulated by the tumor to evade the immune system\textsuperscript{[11]}. Increasing evidence indicates that innate and adaptive immune mediators in the tumor microenvironment play an important role in tumor progression and development of laryngeal carcinomas\textsuperscript{[12]}. However, the relationship between TME-related genes and the prognosis of laryngeal cancer patients has rarely been reported.

Based on the fact of poor accuracy of prognosis, the laryngeal cancer has brought great difficulty to clinical treatment. Since TME has attracted more and more attention in the cancer research currently, a more comprehensive approach to build a TME-associated prognostic model for laryngeal cancer is required. To address these issues, in this study, we analyzed the expression of 760 tumor microenvironment-related genes in laryngeal cancer patients and their prognostic value in laryngeal cancer patients. Moreover, the 22 specific immune cell infiltration as well as 6 immune checkpoints in laryngeal cancer have been also studied comprehensively. All of these promising outcomes enriched the precise prognosis of the disease, which provide tremendous help for future laryngeal cancer study.

**Material And Methods**

**Data Source**

We downloaded the gene information profile and corresponding clinical information of 513 head and neck squamous cell carcinoma (HNSC) patients from the database: The Cancer Genome Atlas (TCGA, https://tcga-data.nci.nih.gov/tcga/). Of which, 180 of these 513 patients were laryngeal cancer patients. Excluding samples with incomplete survival information, 169 laryngeal cancer patients were used for the following analysis.

In addition, we also downloaded a data set from the Gene Expression Omnibus (Geo https://www.ncbi.nlm.nih.gov/GEO/) database, GSE27020, which contains 109 laryngeal cancer patients with complete survival information. Samples in the GEO Dataset were examined using the Affymetrix Human Genome U133A Array platform.

**LASSO Cox regression analysis**

Based on the expression values of 760 TME related genes in laryngeal carcinoma samples, a single factor Cox regression analysis for prognosis of laryngeal carcinoma was developed, with a threshold of $P < 0.01$. Then the glmnet package of R language was generated for LASSO Cox regression analysis in order to further select the genes associated with the prognosis of laryngeal carcinoma. The selected genes were established to calculate the risk score for each sample by the following formula:

$$\text{Risk Score} = \sum_{i=1}^{n} \text{Coef}_i \times X_i$$
where $\text{Coe}$ is the risk coefficient of each factor calculated by the LASSO-Cox model; $X_i$ is the expression value of each factor, in this study refers to the expression value of the gene. Then the patients were divided into high-risk group and low-risk group according to the median of the risk score.

**Survival analysis**

The survival analysis was performed using the R language survival package and survminer (https://CRAN.R-project.org/package=survminer). The package estimates the overall survival rate of different TCGA groups based on the Kaplan-Meier method, and uses log-rank test to investigate the significance of survival difference between different groups.

**Calculation of immune cell infiltration ratio**

The software CIBERSORT was utilized to calculate the relative proportion of 22 immune cells in HCC patients\cite{13}. The CICERSORT software could use the deconvolution algorithm with the preset 547 barcodes to characterize the composition of immune infiltrating cells according to the gene expression matrix. The sum of the proportions of all estimated immune cell types in each sample is equal to 1.

**Establishment of Nomogram prognostic prediction model**

Nomogram is widely used to predict the prognosis of cancer. In order to predict the survival probability of patients at 1 year, 3 years as well as 5 years, we preformed the R language RMS package to establish the prediction model based on all independent prognostic factors determined by multi-factor Cox regression analysis nomogram, draw the calibration curve of nomogram, and analyze the relationship between the predicted probability of Nomogram and the actual incidence rate.

**Differential gene analysis**

The differentially expressed genes analysis were based on the limma function package\cite{14} of the R language (version3.5.2), with the difference factor greater than 1.5 times and FDR $\leq 0.05$ as a criteria.

**Functional enrichment analysis**

For the obtained differentially expressed genes, we use the "clusterProfiler" \cite{15} function package in the R language to perform Gene Ontology (GO, including Biological Process, Molecular Function and Cellular Component) terms and Kyoto Encyclopedia of Genes and Enrichment analysis of Genomes (KEGG) pathways. The significantly enriched GO term and KEGG pathway were further investigated using the P-value $< 0.05$ corrected by the "BH" method as the standard.

**Statistical analysis**

The multi-factor Cox regression model was built to analyze whether risk score could predict the survival of patients with laryngeal carcinoma independently of all other factors. The Wilcoxon signed rank sum test method was used to compare the differences of immune cell infiltration in different groups, with $P < 0.05$ as the significant threshold. The statistical analysis was established by R software, with version number v3.5.2.

**Results**
Construction and verification of the prognosis model

The single-factor cox regression analysis was developed by the TCGA data set samples with 760 TME-related gene expression values as continuous variables. The P-value < 0.01 was used as a threshold. 50 genes were finally screened out. Protective genes with HR value less than 1 are favorable for prognosis, while risk genes with HR value greater than 1 are unfavorable for prognosis. 2 of the 50 genes are protective genes, and the remaining 48 genes are risk genes. The forest diagram of the top 20 genes with the smallest P-value among the 50 genes is shown in Fig. 1A.

After that, the 50 selected genes were subjected to LASSO Cox regression analysis. According to the lambda values of the LASSO Cox analysis, we have determined that the optimal number of genes is 15 (Fig. 1B, whereas the lambda value is the smallest). The 15 genes are BCAT1, RANBP1, CORO2B, RASIP1, EZH2, TCEAL4, FST, CD63, GEMIN7, SPOCK1, S100A11, CXCL3, RUVBL2, ETV5 and EID1. Then the gene expression was weighted with the regression coefficients of the LASSO Cox regression analysis to establish a risk score model for predicting patient survival, according to Risk Score = (0.1459482*GABARAPL2)+(0.1549568*SAR1A)+(0.1554117*ST13)+(0.1892597*GAPDH)+(0.1121593*FADD)+(0.0857282*LAMP1). We calculated the risk score of each patient, and divided the samples of the TCGA data set and GEO data set into high-risk groups and low-risk groups according to the median of the risk scores. Based on the survival analysis, we found that in the TCGA data set and GEO validation set, high-risk laryngeal cancer samples have poor overall survival compared with lower-risk group (Fig. 1C).

In addition, it can be demonstrated from the time-dependent ROC that the AUC of the 1-year, 3-year, and 5-year survival period of the TCGA data set are 0.7478, 0.8688 and 0.7829 respectively (Fig. 1D), indicating the risk model can effectively predict the prognosis of patients with laryngeal cancer. At the same time, we found that the 15 genes were differentially expressed between the high and low-risk groups (Fig. 1E).

In general, the risk score constructed by 15 TME-related genes: BCAT1, RANBP1, CORO2B, RASIP1, EZH2, TCEAL4, FST, CD63, GEMIN7, SPOCK1, S100A11, CXCL3, RUVBL2, ETV5, and EID1 can better predict the prognosis of patients with laryngeal cancer.

Risk score is an independent prognostic marker of HCC

We included age, sex, TNM stage, HPV index and risk score in multivariate Cox regression analysis to determine whether the risk score was an independent prognostic indicator. The results are shown in Fig. 2A. It was found that risk score was significantly associated with overall survival, and the samples with high risk score had a higher risk of death and were unfavorable for prognosis (HR = 5.50, 95% CI: 3.54–8.6, P < 0.001).

In order to further explore the prognostic value of risk score in laryngeal cancer patients with different clinicopathological factors (including age and TNM stage), we regrouped patients, performing a Kaplan-Meier survival analysis. It could be demonstrated that the overall survival rate of the high-risk group is significantly lower than that of the low-risk group of the samples in different ages and stages (Fig. 2B-
2C). These results confirm that the risk score can be used as an independent indicator to predict the prognosis for HCC patients.

**Nomogram model can better predict the prognosis of HCC patients**

We use four independent prognostic factors: age, gender, radiotherapy status as well as risk score to construct the nomogram model (Fig. 3A). For each patient, draw three lines upwards to determine the points obtained from each factor in the Nomogram. The "Total Points" axis is determined by the sum of these points, of which draw a line to generate the probability of HCC patients surviving 1, 3, and 5 years. The one-year and two-year corrected curves in the calibration chart are relatively close to the ideal curves (the 45-degree line that passes through the origin of the coordinate axis with a slope of 1), indicating that the predicted results of the model in one, three, and five years are in good agreement with the actual results (Fig. 3B-3D).

**Differential gene analysis and functional enrichment results of patients with laryngeal cancer patients**

In order to further investigate the differences of gene expression between high-risk and low-risk groups, we conducted a differential gene analysis and a functional enrichment analysis between the two sets of samples. Compared to the low-risk group, a total of 673 genes display a specific expression manner, including 583 up-regulated genes and 90 down-regulated genes (Fig. 4A).

We performed GO and KEGG enrichment analysis for these 673 genes. It was found that these 673 genes were significantly enriched in GO terms such as extracellular matrix organization and KEGG Pathway such as ECM-receptor interaction. The top 20 most significantly enriched genes and pathways are shown in Fig. 4B and 4C.

**Immune status of laryngeal cancer patients in high and low-risk groups**

We used the CIBERSORT method combined with LM22 feature matrix to estimate the difference of immune infiltration between 22 immune cells in high-risk and low-risk groups of patients with laryngeal cancer. Figure 5A summarized the results of immune cell infiltration in 169 laryngeal cancer patients. There are significant differences in the infiltration ratios of 6 types of immune cells, such as B.cells.naive, between high and low-risk groups (Fig. 5B).

Since the expression of immune checkpoints has become a biomarker of immunotherapy for laryngeal cancer patients, we analyzed the correlation between patient risk score and key immune checkpoints (CTLA4, PDL1, LAG3, TIGIT, IDO1, TDO2). It could be seen that the risk score is closely associated with all the 6 checkpoints, which demonstrate the significant differences between the high and low-risk groups of laryngeal cancer patients (Fig. 5C).

**Discussion**

Laryngeal cancers represent one-third of all head and neck cancers and maybe a significant source of morbidity and mortality. They are most often diagnosed in patients with significant smoking history\(^{[16]}\). Currently, researchers have been trying to find breakthroughs in the prevention, early screening, diagnosis
and treatment of laryngeal cancer, and have made many progress in these areas, but the prediction of its prognosis is still inferior\cite{17}. In this study, we established a innovative prognostic analysis based on TME-related gene expression. Among 760 TME-related genes, 50 of them display specific expression. Interestingly, majority of the differentially expressed genes are risk genes. The tumor microenvironment is established by a wide array of cells from both adaptive and innate immune systems. Based on their phenotypical and functional characteristics, these cells can be subclassified as antitumoral or protumoral\cite{18}. Based on our results, the protumoral may play a key function in the laryngeal cancer development as well as progress. These cells promote tumor cell immune evasion by dampening the immune response and generating immune tolerance\cite{19}. As previously suggested, they include regulatory T cells (Tregs), myeloid-derived suppressor cells (MDSC) and tumor-associated macrophages (TAM)\cite{20}. This is consistent with our finding shown in Fig. 5A. These types of cells may be involved in tumor immune escape mechanisms. At the same time, there are evidences showing the increased amount of protumoral immune cells in the peripheral blood of laryngeal cancer patients compared with healthy donors\cite{18}. Eventhough, the percentage of protumoral immune cells contribute the tumor size of laryngeal cancer\cite{21}.

Here, we deeply investigate the immune cell infiltration in different groups of laryngeal cancer patients, where differences in the proportion of immune cell infiltration may be an intrinsic feature of individual differences receiving immunotherapy. In addition, the correlation between different types of immune cells is weak, which indicates that there is a large heterogeneity in the infiltration of different immune cells in tumor patients. Here, in this study, all the 6 immune checkpoints demonstrate significant differences in the high and low-risk groups. Immune checkpoints function through the regulation of the extent of T cells activation and to prevent excessive activation and autoimmunity\cite{22}. These inhibitory pathways are critical in the tumor microenvironment and have been employed by tumor cells as a mechanism of immune evasion\cite{23}. The relation between expression immune checkpoints in the peripheral blood and aggressive tumor growth in patients with laryngeal cancer has also been described by several studies\cite{24,25}, indicating that the poor prognosis of laryngeal cancer patients with high risk may be due to the immunosuppressive microenvironment.

Here, using TME-related gene expression as an independent variable, we generated a risk score model aiming for laryngeal cancer prognostic prediction. The accuracy of the model prediction is evaluated and approved by the AUC value, which demonstrates not only the rationality of our method on the one hand but also the importance of the prediction model for future laryngeal cancer prognosis. In addition to the prognosis model establishment for laryngeal cancer, another feature of this study is to explore multiple key associated genes. BCAT stands for branched-chain amino-acid transaminase enzymes. It has been found that up-regulation of BCAT1 is associated with poor prognosis in numerous types of tumors, such as gastric cancer\cite{26}. The RanBP1 protein, which binds Ran and regulates its interaction with effectors, is overexpressed in many cancer types\cite{27}. Several observations indicate that RanBP1 contributes to regulate the function of the mitotic apparatus\cite{28}. Enhancer of zeste homolog 2 (EZH2), function as a master regulator of chromatin, provides several molecular-based evidences in cancer therapy. Genetic,
transcriptional, and posttranscriptional dysregulation of EZH2 is frequently observed in many cancer types\cite{29,30}. The elevated expression of SPOCK1 has been shown to correlate with EMT-related markers in human gastric cancer tissue, clinical metastasis and a poor prognosis in patients with gastric cancer\cite{31}. In addition, knockdown of SPOCK1 expression significantly inhibits the invasion and metastasis of gastric cancer cells in vitro and in vivo. CXCL3 belongs to the CXC-type chemokine family and is known to play a multifaceted role in various human malignancies\cite{32}. RUVBL2 represents RuvB-like 2 protein, deregulation of which in hepatocellular carcinoma is influenced at the genomic, epigenetic and transcriptional levels. It has been suggested as a promising prognostic marker as well as a therapeutic target for hepatocellular carcinoma\cite{33}. Beside these, the connection between the other genes and laryngeal cancer are not clear yet. All of these deserve further investigation.

In conclusion, in the light of the fact that there remains no gold standard prognosis and no reliable disease-specific prediction for laryngeal cancer, we establish a innovative prognostic model based on TME-related gene expression. According to the model, several related genes and immune cells as well as 6 key immune checkpoints demonstrate significant difference in high and low-risk group. Overall we shed light on questions and challenges posed by the laryngeal cancer, and we establish a innovative prediction target which can provide great help for future understanding of the disease.

Declarations

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Conflict of interest statement

The authors declare no competing interests.

References

1. de Miguel-Luken MJ, Chaves-Conde M, Carnero A. 2016. A genetic view of laryngeal cancer heterogeneity. 15:1202-12
2. Forastiere A, Koch W, Trotti A, Sidransky D. 2001. Head and neck cancer. The New England journal of medicine 345:1890-900
3. Britt CJ, Gourin CG. 2017. Contemporary management of advanced laryngeal cancer. Laryngoscope investigative otolaryngology 2:307-9
4. Bradley PJ. 2016. Laryngeal cancer in nondrinker nonsmoker young patients: a distinct pathological entity? Current opinion in otolaryngology & head and neck surgery 24:140-7
5. Nocini R, Sanchis-Gomar F, Lippi G. 2019. Physical activity and laryngeal cancer. Annals of translational medicine 7:791
6. Obid R, Redlich M, Tomeh C. 2019. The Treatment of Laryngeal Cancer. Oral and maxillofacial surgery clinics of North America 31:1-11
7. Salvador-Coloma C, Cohen E. 2016. Multidisciplinary Care of Laryngeal Cancer. Journal of oncology practice 12:717-24
8. Kinshuck AJ, Shenoy A, Jones TM. 2017. Voice outcomes for early laryngeal cancer. Current opinion in otolaryngology & head and neck surgery 25:211-6
9. Haigentz M, Jr., Silver CE, Hartl DM, Takes RP, Rodrigo JP, et al. 2010. Chemotherapy regimens and treatment protocols for laryngeal cancer. Expert opinion on pharmacotherapy 11:1305-16
10. Arneth B. 2019. Tumor Microenvironment. Medicina 56
11. Frankel T, Lanfranca MP, Zou W. 2017. The Role of Tumor Microenvironment in Cancer Immunotherapy. Advances in experimental medicine and biology 1036:51-64
12. Kerkar SP, Restifo NP. 2012. Cellular constituents of immune escape within the tumor microenvironment. Cancer research 72:3125-30
13. Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, et al. 2015. Robust enumeration of cell subsets from tissue expression profiles. Nature methods 12:453-7
14. Friedman J, Hastie T, Tibshirani R. 2010. Regularization Paths for Generalized Linear Models via Coordinate Descent. Journal of statistical software 33:1-22
15. Yu G, Wang LG, Han Y, He QY. 2012. clusterProfiler: an R package for comparing biological themes among gene clusters. Omics : a journal of integrative biology 16:284-7
16. Steuer CE, El-Deiry M, Parks JR, Higgins KA, Saba NF. 2017. An update on larynx cancer. CA: a cancer journal for clinicians 67:31-50
17. Garcia-Leon FJ, Garcia-Estepa R, Romero-Tabares A, Gomez-Millan Borrachina J. 2017. Treatment of advanced laryngeal cancer and quality of life. Systematic review. Acta otorrinolaringologica espanola 68:212-9
18. Sun W, Li WJ, Fu QL, Wu CY, Lin JZ, et al. 2015. Functionally distinct subsets of CD4(+) regulatory T cells in patients with laryngeal squamous cell carcinoma are indicative of immune deregulation and disease progression. Oncology reports 33:354-62
19. Dewyer NA, Wolf GT, Light E, Worden F, Urba S, et al. 2014. Circulating CD4-positive lymphocyte levels as predictor of response to induction chemotherapy in patients with advanced laryngeal cancer. Head & neck 36:9-14
20. Gabrilovich DI, Nagaraj S. 2009. Myeloid-derived suppressor cells as regulators of the immune system. Nature reviews. Immunology 9:162-74
21. Starska K, Forma E, Lewy-Trenda I, Wos J, Papiez P, et al. 2013. Expression of CTLA-4 and Foxp3 in peripheral blood T cells of patients with squamous cell laryngeal carcinoma. Contemporary oncology 17:370-7
22. Erfani N, Khademi B, Haghshenas MR, Mojtaba Z, Khademi B, Ghaderi A. 2013. Intracellular CTLA4 and regulatory T cells in patients with laryngeal squamous cell carcinoma. Immunological
23. Locy H, de Mey S, de Mey W, De Ridder M, Thielemans K, Maenhout SK. 2018. Immunomodulation of the Tumor Microenvironment: Turn Foe Into Friend. Frontiers in immunology 9:2909

24. Hirata E, Sahai E. 2017. Tumor Microenvironment and Differential Responses to Therapy. Cold Spring Harbor perspectives in medicine 7

25. Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, et al. 2006. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. Science 313:1960-4

26. Xu Y, Yu W, Yang T, Zhang M, Liang C, et al. 2018. Overexpression of BCAT1 is a prognostic marker in gastric cancer. Human pathology 75:41-6

27. Ikari N, Serizawa A, Mitani S, Yamamoto M, Furukawa T. 2019. Near-Comprehensive Resequencing of Cancer-Associated Genes in Surgically Resected Metastatic Liver Tumors of Gastric Cancer. The American journal of pathology 189:784-96

28. Rensen WM, Roscioli E, Tedeschi A, Mangiacasale R, Ciciarello M, et al. 2009. RanBP1 downregulation sensitizes cancer cells to taxol in a caspase-3-dependent manner. Oncogene 28:1748-58

29. Mu X, Chen M, Xiao B, Yang B, Singh S, Zhang B. 2019. EZH2 Confers Sensitivity of Breast Cancer Cells to Taxol by Attenuating p21 Expression Epigenetically. DNA and cell biology 38:651-9

30. Yamagishi M, Uchimaru K. 2017. Targeting EZH2 in cancer therapy. Current opinion in oncology 29:375-81

31. Chen D, Zhou H, Liu G, Zhao Y, Cao G, Liu Q. 2018. SPOCK1 promotes the invasion and metastasis of gastric cancer through Slug-induced epithelial-mesenchymal transition. Journal of cellular and molecular medicine 22:797-807

32. Qi YL, Li Y, Man XX, Sui HY, Zhao XL, et al. 2020. CXCL3 overexpression promotes the tumorigenic potential of uterine cervical cancer cells via the MAPK/ERK pathway. Journal of cellular physiology 235:4756-65

33. Yan T, Liu F, Gao J, Lu H, Cai J, et al. 2019. Multilevel regulation of RUVBL2 expression predicts poor prognosis in hepatocellular carcinoma. Cancer cell international 19:249

**Figures**
Figure 1

Construction of a prognostic model for laryngeal cancer. (A) Forest plot of the top 20 most significant TME genes related to the prognosis of laryngeal cancer. HR is the Hazard ratio, and 95% CI is the 95% confidence interval. (B) The graph of determining the tuning parameter lambda in the LASSO regression model. The horizontal axis is log (lambda), and the vertical axis is the partial likelihood deviation value respectively. The lambda value corresponding to the smallest value is the best. (C) Kaplan Meier survival
curve in the TCGA data set. The horizontal axis indicates time, while the vertical axis indicates survival rate. Different colors represent different groups. The P-value is based on log-rank test. (D) The time-dependent ROC curve. The horizontal axis is the false positive rate, while the vertical axis is the true positive rate respectively. The accuracy of the prediction is evaluated by the AUC (area under the ROC curve) value. (H) The mRNA expression heat map of the 15 genes selected in the TCGA data set and GEO validation data set.

Figure 2

Risk score is an independent prognostic marker for laryngeal cancer. (A) Multivariate Cox regression analysis forest plot. Compared with reference samples, samples with Hazard ratio greater than 1 have a higher risk of death, and samples with Hazard ratio less than 1 have a lower risk of death risk of death. (B-C) Kaplan Meier survival curve of HCC patients ≤60 years old and >60 years old.
Figure 5

Immune infiltration of laryngeal cancer patients in the high and low-risk groups. (A) The relative proportion of immune infiltrating cells in all patients. (B) The violin diagram of immune cells with significant difference in high and low-risk group. The horizontal axis represents high and low-risk group, the vertical axis represents relative infiltration ratio of immune cells respectively. The P-value is calculated by wilcoxon method. (C) The Chord diagram of the correlation between the risk score and the expression of
6 prominent immune checkpoints. The thicker the line between them, the stronger the correlation between them.