A ZYG-12–dynein interaction at the nuclear envelope defines cytoskeletal architecture in the C. elegans gonad

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Abbreviations used in this paper: ECM, embryonic culture medium; KASH, Klarsicht/Anc-1/Syne-1 homology; MTOC, microtubule-organizing center; SUN, Sad1/unc-84; TEM, transmission EM.

Changes in cellular microtubule organization often accompany developmental progression. In the Caenorhabditis elegans embryo, the centrosome, which is attached to the nucleus via ZYG-12, organizes the microtubule network. In this study, we investigate ZYG-12 function and microtubule organization before embryo formation in the gonad. Surprisingly, ZYG-12 is dispensable for centrosome attachment in the germline. However, ZYG-12–mediated recruitment of dynein to the nuclear envelope is required to maintain microtubule organization, membrane architecture, and nuclear positioning within the syncytial gonad. We examined γ-tubulin localization and microtubule regrowth after depolymerization to identify sites of nucleation in germ cells. γ-Tubulin localizes to the plasma membrane in addition to the centrosome, and regrowth initiates at both sites. Because we do not observe organized microtubules around zyg-12(ct350) mutant nuclei with attached centrosomes, we propose that gonad architecture, including membrane and nuclear positioning, is determined by microtubule nucleation at the plasma membrane combined with tension on the microtubules by dynein anchored at the nucleus by ZYG-12.

Introduction

Tissue architecture and organ architecture are key to biological function. Tissue architecture, in turn, is largely determined by constituent cell architecture, including global cell shape as well as subcellular positioning of the nucleus and other organelles. The cytoskeleton is responsible for generating and maintaining morphology and organelle organization in cells with a variety of shapes and functions (for review see Li and Gundersen, 2008). We do not yet understand how diverse cells use a common set of cytoskeletal elements and similar basic tools to manipulate these elements to generate distinct architectures. Cells manipulate these cytoskeletal elements using regulatory proteins, including nucleators, depolymerases, severing proteins, and other binding proteins. Additionally, motor proteins transport cargo along cytoskeletal tracks, and anchored motor proteins pull on the cytoskeleton itself (Nguyen-Ngoc et al., 2007) to achieve the desired organization.

Although all cytoskeletal components make significant contributions to the establishment and maintenance of architectural organization, dramatic changes in microtubule organization upon differentiation and the extensive diversity in microtubule organization among cell types speaks to their key role in cell architecture. The concept of a microtubule-organizing center (MTOC), a site of γ-tubulin localization that nucleates microtubule formation, is used to explain the origins of various microtubule arrangements. In dividing cells, the nucleus-attached centrosome nucleates a characteristic array with the plus ends of the microtubules extending radially toward the cell periphery. However, differentiated cells have various alternative organizations in which microtubules are nucleated at noncentrosomal sites via mechanisms that are less well understood. For example, during differentiation of myoblasts, the microtubule cytoskeleton is remodeled from a centrosome-nucleated radial structure to a series of parallel arrays along the long axis of the myotube that are likely nucleated at nuclei with plus ends extending to the poles (Tassin et al., 1985; Bugnard et al., 2005). Polarized epithelia provide a second dramatic example of an alternative microtubule cytoskeleton; epithelial cells contain linear arrays of microtubules oriented with minus ends at the apex and plus ends extending toward the cell periphery. The concept of a MTOC is used to explain the origins of these microtubule arrangements. In dividing cells, the nucleus-attached centrosome nucleates a characteristic array with the plus ends of the microtubules extending radially toward the cell periphery. However, differentiated cells have various alternative organizations in which microtubules are nucleated at noncentrosomal sites via mechanisms that are less well understood. For example, during differentiation of myoblasts, the microtubule cytoskeleton is remodeled from a centrosome-nucleated radial structure to a series of parallel arrays along the long axis of the myotube that are likely nucleated at nuclei with plus ends extending to the poles (Tassin et al., 1985; Bugnard et al., 2005). Polarized epithelia provide a second dramatic example of an alternative microtubule cytoskeleton; epithelial cells contain linear arrays of microtubules oriented with minus ends at the apex and plus ends extending toward the cell periphery.

Changes in cellular microtubule organization often accompany developmental progression. In the Caenorhabditis elegans embryo, the centrosome, which is attached to the nucleus via ZYG-12, organizes the microtubule network. In this study, we investigate ZYG-12 function and microtubule organization before embryo formation in the gonad. Surprisingly, ZYG-12 is dispensable for centrosome attachment in the germline. However, ZYG-12–mediated recruitment of dynein to the nuclear envelope is required to maintain microtubule organization, membrane architecture, and nuclear positioning within the syncytial gonad. We examined γ-tubulin localization and microtubule regrowth after depolymerization to identify sites of nucleation in germ cells. γ-Tubulin localizes to the plasma membrane in addition to the centrosome, and regrowth initiates at both sites. Because we do not observe organized microtubules around zyg-12(ct350) mutant nuclei with attached centrosomes, we propose that gonad architecture, including membrane and nuclear positioning, is determined by microtubule nucleation at the plasma membrane combined with tension on the microtubules by dynein anchored at the nucleus by ZYG-12.

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extending to the basal side (Bre et al., 1990; Meads and Schroer, 1995). In epidermal development, epidermal cells progress from a stage with a centrosomal array to a stage with a parallel array as microtubule-anchoring proteins are relocalized from the centrosome to the cell junctions near the apical membrane (Lechler and Fuchs, 2007; Meng et al., 2008). Thus, dramatic alterations in microtubule organization often accompany differentiation. To comprehend the events that underlie cell differentiation and development of tissue architecture, it is important to understand how diverse microtubule arrays are organized.

We use the *Caenorhabditis elegans* hermaphrodite gonad as a model to probe the molecular mechanisms that establish and maintain tissue architecture. The gonad consists of an orderly arrangement of nuclei within a large syncytium that is surrounded by somatic gonad sheath cells (Fig. 1A). The global architecture of the gonad as well as the cell cycle state of the nuclei changes as nuclei progress from the distal gonad tips to the proximal region. A distal tip cell caps each gonad arm and acts to maintain a mass of mitotic nuclei. As new cell divisions push nuclei proximally and away from the distal tip cells, nuclei initiate a meiotic program in the transition zone. Each meiotic nucleus is maintained at the periphery of the gonad in a partially enclosed membrane compartment with a cytoplasmic bridge that connects to a central cytoplasmic pool, the rachis (Hirsh et al., 1976; Hall et al., 1999). Nuclei progress through the pachytene region into the loop of the gonad where changes in cell shape, including acquisition of extra cytoplasm, and full cellularization lead to production of large oocytes. A transverse view of the distal gonad arm reveals ~12 germline nuclei abutting the gonad sheath and surrounding the rachis (Fig. 1B). When viewed from the lateral side, germline nuclei are evenly spaced at the periphery throughout the distal gonad arm (Fig. 1C). This regularly arrayed pattern of germline nuclei within the gonad seems important for proper oogenesis and reproduction. Nuclear position in *C. elegans*, including that of germline nuclei, is subject to active control by the cytoskeleton and by extracellular matrix proteins (Hedgecock and Thomson, 1982; Vogel and Hedgecock, 2001; Thompson et al., 2002; Lu et al., 2008). However, the mechanisms that maintain the evenly distributed peripheral pattern of germline nuclei are mysterious.

Nuclear positioning in other *C. elegans* tissues and in other systems often depends on a conserved pair of protein families called Sad1/unc-84 (SUN) and Klarsicht/Anc-1/Syne-1 homology (KASH) domain proteins (for review see Starr, 2007). Current models posit that conserved SUN domain proteins bind chromatin or the nuclear lamina in the nucleus and span the inner nuclear membrane to bind to a conserved KASH domain protein partner in the inter membrane space. The KASH domain proteins span the outer nuclear membrane and interact with cytoskeletal elements in the cytoplasm. Thus, a SUN–KASH pair can facilitate interaction of the nucleus with a cytoskeletal system, including the microtubules (Starr and Han, 2002; Fischer et al., 2004; Yu et al., 2006; Zhang et al., 2007).

In the *C. elegans* embryo, centrosome–nucleus attachment is maintained via the SUN–KASH pair, SUN-1 and ZYG-12 (Malone et al., 2003). Two of three alternative ZYG-12 isoforms have a transmembrane-containing KASH domain and are localized to the nuclear envelope by SUN-1, but the third ZYG-12 isoform lacks the KASH domain and localizes to the centrosome (Malone et al., 2003). Interaction of these nuclear and centrosomal isoforms is proposed to mediate centrosome–nucleus attachment, and nuclear migration clearly depends on the connection between the nucleus and the centrosome MTOC. Dynein also plays a role in embryonic pronuclear migration; depletion of *C. elegans* dynein subunits DLI-1 or DHC-1 causes a penetrant pronuclear congression defect (Gonczy et al., 1999; Yoder and Han, 2001). These data are consistent with a model in which dynein provides the force for movement of the centrosome, whereas ZYG-12 provides centrosome–nucleus attachment, permitting nuclear movement. ZYG-12 also interacts directly with DLI-1, the light intermediate chain of dynein, and recruits DLI-1 to the nuclear envelope in the early embryo and the gonad (Malone et al., 2003), suggesting that ZYG-12 functions as a molecular anchor on the nuclear envelope for cytoplasmic dynein. However, the role of dynein at the nuclear envelope in embryos is not clear; a zyg-12 allele (or577) that abrogates ZYG-12 homodimerization but retains the dynein interaction has migration and centrosome detachment defects as severe as those of zyg-12(ct350), which disrupts the dynein interaction (Malone et al., 2003), suggesting that the key function for ZYG-12 in the embryo is centrosome–nucleus attachment.

Both SUN-1 and ZYG-12 are expressed in the germline, but neither the role of these proteins nor the role of the microtubules in germline architecture, nuclear positioning, or centrosome attachment has been investigated in the gonad. Therefore, we started our investigation of germline cytoarchitecture with ZYG-12. In this study, we reveal that, in the gonad, ZYG-12 is dispensable for centrosome attachment. However, ZYG-12 recruitment of dynein to the germ cell nuclear envelope is critical for nuclear positioning. Because both ZYG-12 function and nuclear positioning in the gonad are independent of the centrosome, we examined alternative sites of microtubule nucleation. Based on the localization of γ-tubulin and microtubule regrowth initiation at the cytoplasmic membrane after depolymerization, we propose that gonad architecture, including germline nuclear positioning, is determined by microtubule nucleation at the plasma membrane in combination with tension on these microtubules provided by dynein that is anchored to the nuclear membrane by ZYG-12.

**Results**

**Mutation of zyg-12 results in nuclear position and oogenesis defects in the *C. elegans* gonad**

In the embryo, nuclear positioning requires ZYG-12 at the nuclear membrane. ZYG-12 is also localized to the nuclear envelope of germline nuclei in the syncytial *C. elegans* gonad (Malone et al., 2003). To determine whether ZYG-12 has essential function in the gonad, we examined zyg-12 mutant gonads for defects. To avoid the early embryonic lethality associated with disruption of ZYG-12 function, we used a temperature-sensitive allele of zyg-12. We shifted postembryonic, first larval stage (L1) zyg-12(ct350 ts) animals to the restrictive temperature (25°C) and examined the gonad of the adults. Intriguingly, we observed...
disruption of the regular arrangement of nuclei within the syncytial gonad of all mutant animals. Although germline nuclei are arranged in a regular array at the periphery of the gonad in wild type (Fig. 1, A–C), germline nuclei appear randomly distributed within the gonad of the zyg-12(ct350) mutants (Fig. 1 D). Furthermore, large oocytes with multiple nuclei form in the proximal gonad arm of the mutants (Fig. 1 F) in contrast to the single nucleus in each wild-type oocyte (Fig. 1 E).

We used immunofluorescence and confocal microscopy to further characterize nuclear mispositioning. We extracted gonads from zyg-12(ct350) or wild-type animals at the restrictive temperature and performed immunofluorescence with antibodies against ZYG-12 to visualize nuclear envelopes and against α-tubulin to visualize cytoplasm. In wild type, germline nuclei are evenly arranged directly under the surface of gonad (Fig. 2 A), and no nuclei occupy the central rachis (Fig. 2 B). In contrast, in the zyg-12(ct350) mutant, the nuclei are disarrayed at the surface (Fig. 2 D), and large numbers of nuclei have collapsed into the central rachis (Fig. 2 E). We also observed abnormal clusters of small nuclei that do not appear to have undergone oogenesis in the proximal gonad (Fig. 2 F).

Because zyg-12 is required for centrosome–nucleus attachment in the embryo, it is required for embryonic mitotic divisions (Malone et al., 2003). To rule out a mitotic defect in the rapidly proliferating germline as the underlying cause of the disruption in nuclear arrangement, we examined gonads at shorter intervals after the shift to the restrictive temperature. We transferred zyg-12(ct350) egg-laying hermaphrodites that had been maintained at 15°C to the restrictive temperature (25°C) for 1, 4, or 8 h and examined the postmitotic region of their gonads. Germline nuclei retained their position after 1 h at 25°C (Fig. 2 G). However, within 4 h, germline nuclei were displaced in the distal (Fig. 2 H) and the proximal (Fig. 2 K) gonad. 8 h after the temperature shift, the gonad was heavily disrupted (Fig. 2, I and L). The majority of the distal nuclei are unlikely to have undergone mitosis during the time course of the experiment. Moreover, the nuclei in the pachytene zone of the proximal gonad are postmitotic before the temperature shift and yet become mispositioned. Wild-type germline nuclei maintained their well-aligned position when cultured at the restrictive temperature starting in L1 (Fig. 1 C) or after 6 h at the restrictive temperature (Fig. 2, A–C). The rapid nuclear...
repositioning throughout the gonad after 4 h at the restrictive temperature suggests that the phenotype is independent of any mitotic defect.

Germ cell membranes are also disrupted in zyg-12(ct350) mutant gonads

To further characterize the disrupted gonad phenotype caused by zyg-12(ct350), we examined cell membranes. We used a germ cell membrane–localized syntaxin-GFP fusion, GFP::SYN-4 (Jantsch-Pfunger and Glotzer, 1999), to visualize the membranes that partially surround each germ cell nucleus in the syncytial gonad. In parallel with the disruption of nuclear position, the membranes are rearranged in the zyg-12(ct350) mutants. In wild type, the membranes that partially surround each germ cell nucleus are well organized in a honeycomb-like structure (Fig. 3 A). However, within 6 h at the restrictive temperature, zyg-12(ct350) membranes are disorganized (Fig. 3, B and C) and, in the proximal gonad, sometimes incorporate >1 nucleus (Fig. 3, E and F). We conclude that zyg-12 is involved in a mechanism that helps to maintain the well-aligned, evenly spaced peripheral position of the germline nuclei and the regularly arrayed surrounding membranes within the gonad syncytium.

ZYG-12-dynein and ZYG-12-SUN-1 interactions are important for normal gonad architecture

To understand how disruption of ZYG-12 could alter both nuclear position and membrane architecture, we tested alleles of zyg-12 that affect distinct protein–protein interactions (Fig. 4 J). ZYG-12 interacts with itself and with dynein light intermediate chain DLI-1 in yeast two-hybrid assays (Fig. 4 K; Malone et al., 2003). The zyg-12(ct350) allele mutation (Q44P) lies in the N-terminal region (Fig. 4 J) and renders both the ZYG-12–ZYG-12 and ZYG-12–DLI-1 interactions sensitive to high temperature in a yeast two-hybrid assay (Malone et al., 2003). In contrast, the mutation (Q367P) present in another zyg-12 ts allele, or577, lies in the coiled-coil domain of ZYG-12 (Fig. 4 K) and abrogates the ZYG-12–ZYG-12 interaction without affecting the ability of ZYG-12 to bind DLI-1 in a yeast two-hybrid assay (Malone et al., 2003). Although zyg-12(or577ts) embryos exhibit detached centrosomes and promiscuous migration defects similar to those of zyg-12(ct350ts) embryos (see Fig. 6, B and C), we surprisingly found that zyg-12(or577ts) mutants do not display the gonad defect of zyg-12(ct350). The gonads of zyg-12(or577) hermaphrodites shifted to 25°C for 6 h are indistinguishable from
wild-type animals (Fig. 4, A–C), zyg-12(or577) hermaphrodites shifted to the restrictive temperature during L1 also have evenly spaced germline nuclei and oocytes with single nuclei as adults (Fig. S1, A and B). Using an antibody to the heavy chain of cytoplasmic dynein, DHC-1, we confirmed that DHC-1 was recruited to the nuclear membrane of all germline nuclei in wild-type (Fig. 4 D) but not zyg-12(ct350) mutant gonads at 25°C (Fig. 4 E; Malone et al., 2003). zyg-12(or577) animals had normal DHC-1 localization at the nuclear membrane (Fig. 4 F).

The lack of a gonad phenotype in zyg-12(or577), which binds to and recruits dynein to the nuclear envelope, compared with the severe nuclear positioning and membrane morphology defects of zyg-12(ct350), which fails to bind dynein or recruit it to the nuclear envelope, suggests that a ZYG-12 interaction with dynein is important for maintenance of gonad architecture. To further investigate the role of dynein, we used RNAi to deplete DHC-1. dhc-1(RNAi) causes a larval arrest phenotype (unpublished data). However, adult animals that escape larval arrest have a gonad phenotype similar to that of zyg-12(ct350) animals. In the distal gonad of dhc-1(RNAi) hermaphrodites, germline nuclei were displaced from the periphery of the gonad (Fig. 4 G) as previously noted (Wolke et al., 2007). Phenotypes in the proximal gonad in dhc-1(RNAi) were somewhat variable. However, there were many oocytes with multiple nuclei (Fig. 4 H), and the arrangement of the oocytes during their formation is abnormal (Fig. 4 I).

In the zygote and the gonad, the localization of ZYG-12 to the outer nuclear membrane is dependent on the inner nuclear envelope protein SUN-1 (Malone et al., 2003; Penkner et al., 2007). As expected, depletion of sun-1 via RNAi also causes disorganization of the nuclei in the distal gonad and clumping of multiple nuclei in the proximal gonad (Fig. S2). We conclude that in addition to ZYG-12, both known binding partners of ZYG-12, SUN-1 and DLI-1, are required for maintenance of gonad architecture.
Therefore, we determined whether zyg-12(ct350) at the non-permissive temperature resulted in loss of the centrosome from its perinuclear position in the gonad. We examined attachment of the centrosome to the nucleus at the restrictive temperature in wild-type and zyg-12(or577) gonads, in which germline nuclei are always properly positioned, and in zyg-12(ct350) gonads, in which the germline nuclei are mispositioned. We confirmed that both maintenance of microtubule organization within the gonad is centrosome independent. The best-characterized MTOC is the centrosome, and ZYG-12 is required for the juxtaposition of the centrosome and the nucleus in the embryo. Thus, one simple hypothesis to explain disruption of microtubules in zyg-12(ct350) gonads is that mispositioning of the centrosomal MTOC underlies the microtubule disorganization. Therefore, we determined whether zyg-12(ct350) at the non-permissive temperature resulted in loss of the centrosome from its perinuclear position in the gonad. We examined attachment of the centrosome to the nucleus at the restrictive temperature in wild-type and zyg-12(or577) gonads, in which germline nuclei are always properly positioned, and in zyg-12(ct350) gonads, in which the germline nuclei are mispositioned. We confirmed that both...
attachment. Misplaced mutant nuclei maintain their centrosome attachments (Fig. 6 E), and misplaced nuclei with attached centrosomes are sometimes completely outside any microtubule network (Fig. 5 C). Thus, the centrosome is not sufficient to organize the global microtubule architecture in the postmitotic gonad, and the mechanism for maintaining nuclear positioning and microtubule organization within the gonad is likely centrosome independent.

\( \gamma \)-Tubulin localization suggests plasma membrane nucleation of microtubules in the distal gonad

As the centrosome is not the key to the loss of microtubule organization in \( zyg-12(\text{ct350}) \) mutants, we hypothesized that another major MTOC is present in the gonad. To identify potential
were marked with GFP. Filamentous microtubules are found in the space between the nuclear envelope and plasma membrane, with some overlap at each membrane, before treatment with the drug (Fig. 7 G). Microtubules were effectively depolymerized after 1 h of nocodazole treatment (Fig. 7 H). Consistent with the idea that the microtubule network is critical for maintaining germline nuclear position and germ cell membrane organization, both are disrupted upon microtubule depolymerization (Fig. 7 H). Within 30 min of nocodazole removal, staining for microtubules reveals numerous puncta that are associated with the plasma membrane (Fig. 7 I; and Fig. S5, C–E). No microtubules are observed along the circumference of the nuclear membrane at these early time points (Fig. 7 I; and Fig. S5, C and E) with the exception of those at the centrosome (Fig. S5, G and H). Microtubules do localize along the circumference of the nuclear membrane by 1 h after nocodazole removal (Fig. S5 F). This temporal pattern of microtubule regrowth suggests that the plasma membrane is a primary microtubule nucleation site. Microtubule nucleation at the centrosome is also observed in disorganized gonads of the zyg-12(ct350) mutants (Fig. S5 I), again suggesting that the centrosome has the capacity to nucleate some microtubules but is insufficient to organize the global germline microtubule array. We propose that the majority of microtubules in the gonad are nucleated at the plasma membrane and that sites of microtubule nucleation, we visualized γ-tubulin, the core component of the canonical microtubule nucleation pathway. We first visualized γ-tubulin using live animals expressing γ-tubulin::GFP (Dammermann et al., 2004). Although the γ-tubulin::GFP signal is weak, it demonstrates that the bulk of the γ-tubulin in the distal region of the gonad syncytium localizes to the germ cell membranes and to the centrosome (Fig. S5 A). In the proximal gonad, we see γ-tubulin on the nuclear and plasma membranes of oocytes with some signal emanating from structures in the cytoplasm as well (Fig. S5 B).

As a more sensitive method to confirm the expression pattern, we used immunofluorescence with a polyclonal GFP antibody in combination with a nuclear pore complex antibody to mark the nuclear membrane. As observed in live animals, γ-tubulin::GFP localizes to both the germ cell membranes and to the nucleus-associated centrosome in the distal gonad (Fig. 7, A–C). In the zyg-12(ct350) mutant at the restrictive temperature, γ-tubulin::GFP still localizes to the cell membranes and to the centrosomes (Fig. 7, D–F). Based on the localization of γ-tubulin and the phenotype of zyg-12(ct350), we hypothesized that the plasma membrane may be a major site of microtubule nucleation in the distal gonad.

To investigate this hypothesis, we used nocodazole to depolymerize microtubules and then removed the drug to observe regrowth in gonads in which the plasma and nuclear membranes were marked with GFP. Filamentous microtubules are found in the space between the nuclear envelope and plasma membrane, with some overlap at each membrane, before treatment with the drug (Fig. 7 G). Microtubules were effectively depolymerized after 1 h of nocodazole treatment (Fig. 7 H). Consistent with the idea that the microtubule network is critical for maintaining germline nuclear position and germ cell membrane organization, both are disrupted upon microtubule depolymerization (Fig. 7 H). Within 30 min of nocodazole removal, staining for microtubules reveals numerous puncta that are associated with the plasma membrane (Fig. 7 I; and Fig. S5, C–E). No microtubules are observed along the circumference of the nuclear membrane at these early time points (Fig. 7 I; and Fig. S5, C and E) with the exception of those at the centrosome (Fig. S5, G and H). Microtubules do localize along the circumference of the nuclear membrane by 1 h after nocodazole removal (Fig. S5 F). This temporal pattern of microtubule regrowth suggests that the plasma membrane is a primary microtubule nucleation site. Microtubule nucleation at the centrosome is also observed in disorganized gonads of the zyg-12(ct350) mutants (Fig. S5 I), again suggesting that the centrosome has the capacity to nucleate some microtubules but is insufficient to organize the global germline microtubule array. We propose that the majority of microtubules in the gonad are nucleated at the plasma membrane and that
Second, organelles such as mitochondria cross the cytoplasmic bridge from germ cell to rachis, illustrating the need for holding the larger nucleus in place and restricting its proximity to the rachis bridge (Fig. 8). Third, microtubules are maintained in close proximity to the outer membrane of the germ cell (Fig. 8 A). Fourth, microtubule organization in the germ cells is nonuniform. There are relatively few microtubules, and they are not found in parallel arrays. They crisscross (Fig. 8 C) and approach both the plasma membrane and the nucleus, specifically contacting the nucleus at areas of electron density (Fig. 8 B) in proximity to the nuclear pores (not depicted). Microtubules also contact (unpublished data) the rachis along its structural support feature, “the rachis coat” (Vogel and Hedgecock, 2001; Hall and Altun, 2008). The coat is an electron-dense structure (Fig. 8, A, D, and E) that covers the proximal gonad.

Ultrastructural analysis of gonad subcellular architecture

We also used ultrastructural analysis to examine gonad architecture and microtubule organization to gain a deeper understanding of how microtubules relate to other organelles and major features of the gonad. Electron micrographs of the proximal gonad reveal several interesting features. First, there is a striking uniformity in cytoplasmic contents in the partially cellularized germ cells and the rachis, consisting primarily of an abundance of free ribosomes, some mitochondria, and rather little ER or yolk (Fig. 8). Second, organelles such as mitochondria cross the cytoplasmic bridge from germ cell to rachis, illustrating the need for holding the larger nucleus in place and restricting its proximity to the rachis bridge (Fig. 8). Third, nuclei are maintained in close proximity to the outer membrane of the germ cell (Fig. 8 A). Fourth, microtubule organization in the germ cells is nonuniform. There are relatively few microtubules, and they are not found in parallel arrays. They crisscross (Fig. 8 C) and approach both the plasma membrane and the nucleus, specifically contacting the nucleus at areas of electron density (Fig. 8 B) in proximity to the nuclear pores (not depicted). Microtubules also contact (unpublished data) the rachis along its structural support feature, “the rachis coat” (Vogel and Hedgecock, 2001; Hall and Altun, 2008). The coat is an electron-dense structure (Fig. 8, A, D, and E) that covers the proximal gonad.

Figure 7. Plasma membrane as the nucleation site of microtubules in the distal gonad. (A–E) γ-Tubulin localizes to germ cell membrane in both wild-type (WT) and zyg-12(ct350) gonads. (A) In the distal gonad of wild-type animals, γ-tubulin localizes to germ cell membranes and centrosomes (white arrows). (B) The nuclear pore complex marks the nuclei. (C–F) In zyg-12(ct350), neither the membrane nor the centrosomal localization of γ-tubulin::GFP is altered. (F) However, nuclear position has been disrupted. (G–I) Microtubules regrow from the plasma membrane (marked by GFP::SYN-4) after the removal of depolymerizing drug from cultured gonads. (G) Filamentous microtubules occupy the cytoplasm and partially colocalize with the plasma membrane in a cultured gonad not subjected to nocodazole. (H) 1 h of nocodazole treatment effectively depolymerizes microtubules in the gonad. (I) 30 min after the removal of nocodazole, microtubules regrow at the plasma membrane.
outer surface of the rachis, with slight thickening near each germ cell opening. The coat is comprised principally of hemicentin, which is required to prevent early germ cell membranes from encapsulating multiple nuclei (Vogel and Hedgecock, 2001). Mitochondria (and other potential cargoes) are often observed to lie very near to microtubules by transmission EM (TEM; Fig. 8 E). Finally, bundles of microtubules cross the cytoplasmic bridge between the germ cell and the rachis (Fig. 8 D).

Discussion

Distinct molecular interactions by ZYG-12 are required for gonad and embryonic functions

Recruitment of ZYG-12 to the nuclear envelope is critical to embryonic development (Malone et al., 2003). In this study, we show that ZYG-12 at the nuclear envelope is crucial for maintenance of the global architecture of the gonad as well. Interestingly, the key binding partners for ZYG-12 in the embryo appear to be distinct from those in the gonad.

In the embryo, a homodimeric interaction between ZYG-12 proteins localized separately to the nuclear envelope and the centrosome is presumed to mediate the necessary attachment between the nucleus and the centrosome (Malone et al., 2003). In the gonad, ZYG-12 has no role in nucleus–centrosome attachment. ZYG-12 is not localized to the centrosome in the gonad (Penkner et al., 2007; unpublished data), and nucleus–centrosome attachment is maintained in the gonad in both zyg-12(ts) alleles examined. Furthermore, ZYG-12–ZYG-12 interactions are dispensable in the gonad, as indicated by the normal gonad phenotype of the ZYG-12 homodimerization-defective mutant zyg-12 (or577). This is further illustrated by comparing the gross phenotype of zyg-12(ct350), which is sterile, indicating a germline defect, and zyg-12(or577), which is embryonic lethal but not sterile, suggesting normal gonad and germline function. Thus, we have revealed a ZYG-12–independent mechanism for centrosome attachment in the postmitotic gonad. The protein emerin appears to function in nucleus–centrosome attachment in human fibroblasts (Salpingidou et al., 2007). It will be interesting to determine whether C. elegans emerin shares this function in the gonad or whether other as yet unidentified proteins attach germ-line nuclei and perhaps nuclei in other tissues to centrosomes.

In the germline and embryos, ZYG-12 appears to function as a molecular anchor for recruiting dynein to the outer nuclear membrane. Although the function of the dynein at the nuclear envelope in embryos may play a minor role in centrosome attachment (Malone et al., 2003), we have shown that recruitment of dynein to the nuclear membrane by ZYG-12 is critical for maintenance of gonad architecture; loss of ZYG-12–DLI-1 interactions likely underlies the zyg-12(ct350) mutant phenotype. Although we cannot rule out the possibility that an interaction between ZYG-12 and another unknown protein is simultaneously disrupted by the ct350 allele, our data are consistent with a central role for dynein. Disruption of dynein expression mimics the zyg-12(ct350) phenotype, and the lack of dynein recruitment to the nuclear envelope is correlated with the zyg-12(ct350) phenotype. ZYG-12 has also been implicated in meiotic chromosome pairing (Penkner et al., 2007), and it remains to be determined which molecular interactions of ZYG-12 are key to this function.

Model for architecture of the distal gonad

We propose that cellular architecture in the distal gonad is dictated by microtubules nucleated at the plasma membrane by γ-tubulin and captured at the nuclear envelope by dynein, whose motor activity, directed toward microtubule minus ends, creates tension between the nucleus and plasma membrane (Fig. 9). This tension could explain the major structural features of the distal gonad, including the peripheral position of the nuclei in the
gonad, with a central rachis devoid of nuclei, and the architecture of the plasma membrane that appears to be pulled around each nucleus (Hall et al., 1999). The predicted resulting haphazard arrangement of microtubules in the cytoplasm is supported by the ultrastructural analysis.

If the only considerations in the distal gonad were positioning nuclei near the edge and bringing plasma membrane extensions between nuclei, other arrangements of microtubules might work equally well. For example, microtubules could be nucleated at or near the nucleus and be pulled on by interactions with the cell cortex. This general arrangement is used frequently (Siegrist and Doe, 2007). However, the distal gonad is not simply a storage shed for future oocyte nuclei; it is hypothesized to function as a nurse compartment that provides material to the oocyte (Wolke et al., 2007). Oocytes, which are transcriptionally quiescent, are packed with material for early embryogenesis via actin-based cytoplasmic streaming from the rachis (Wolke et al., 2007). The transcriptionally active germline nuclei distal to the developing oocytes are thought to synthesize the rachis cytoplasm (Starck, 1977; Gibert et al., 1984; Wolke et al., 2007). The high free ribosome density observed in electron micrographs is consistent with a role for these cells in synthesizing material for loading into the oocyte during cytoplasmic streaming. Thus, most trafficking from the peripheral nuclei should be directed into the central rachis of the gonad, and this transport is likely to be microtubule based. Microtubule nucleation at the plasma membrane and tension along the side of microtubules by dynein at the nuclear envelope allows the tips of microtubules to extend past the nuclei into the rachis (Fig. 8 D and Fig. 9), whereas other microtubule arrangements that could account for peripheral localization of nuclei as mentioned previously, would not. Interestingly, microtubules appear to make closest contact with the nucleus at sites containing electron-dense projections that lie in proximity to the nuclear pore (Fig. 8 D and not depicted). This may provide a link for trafficking from the nucleus to the rachis. Microtubule extension from the peripheral germ cells into the rachis has been described previously (Wolke et al., 2007), but the organizing factors were not known. Interactions between microtubules and the rachis coat are also likely to contribute to the integrity of the membrane in the area of the bridge to the rachis. The rachis coat appears to reinforce the tubular shape of the rachis and probably the cytoplasmic bridges between the germ cells and the rachis, as the coat covers the rachis surface with slight thickening at each opening but never extends over the surface of the germ cells (Hall et al., 1999). Interactions between this coat and local microtubules could maintain each rachis opening between a germ cell and the syncytium as the maturing germ cell migrates proximally along the rachis.

Several other biologically important features of the gonad are likely to be facilitated by anchoring nuclei at its outside edge. Unanchored nuclei could float through cytoplasmic bridges to the rachis, as observed for mitochondria (Fig. 8). This might block the bridge or disrupt cytoplasmic streaming in the rachis. It could also result in multiple nuclei streaming into the oocyte as seen in zyg-12(ct350) (Fig. 1 F and Fig. 2 F). Other mutants are known to cause similar defects in the distal gonad, such as nuclei in the central rachis (Thompson et al., 2002), chains of membrane-covered germ-line cells with extended cytoplasmic bridges connecting them to a malformed rachis (Lu et al., 2008), or germ line cells containing multiple nuclei per membranous compartment but connected normally to the rachis (Vogel and Hedgecock, 2001). The gonads of these mutants appear superficially similar to that of zyg-12(ct350). However, molecular analyses suggest that these phenotypes are caused by different mechanisms such as a mitotic defect or malformed membranes.

Aspects of the phenotype observed after loss of dynein from the nuclear envelope in zyg-12(ct350) are also consistent with a plasma membrane–nucleus tension model for architecture of the distal gonad. After loss of dynein from the nuclear envelope in the zyg-12(ct350) mutant, the distal gonad is rearranged such that the microtubule network encapsulates groups of nuclei while excluding others. The excluded nuclei with no surrounding microtubules clearly illustrate the lack of microtubule-organizing activity by the nuclei and their attached centrosomes. The nuclei...
that are encapsulated by the microtubule network no longer pull membrane around themselves and are not pulled to the periphery because tension between nuclei and the membrane is lost. Although the proposed model for microtubule arrangement seems to make sense for the biology of the *C. elegans* gonad, it appears to conflict with what is known about microtubule arrays in other cell types. Typically, microtubule minus ends are thought to be near the center of the cell and plus ends at the periphery (Steinhauer and Kalderon, 2006), although in this case, the principles that underlie the arrangement of microtubules have not been elucidated (Steinhauer and Kalderon, 2006). Further afield, minus ends are also known to be far from the cell center in mammalian (Baas et al., 1988) and *Drosophila* (Stone et al., 2008) neurons. In all of these examples, the positioning of the nucleation sites is not well understood. This system will provide an opportunity to identify, using a simple genetic organism, how these sites are regulated and to test whether this regulation is the key to setting up different types of architecture in different regions of a single organ.

### Materials and methods

#### Maintenance and culture of *C. elegans*

Wild-type N2 animals were maintained at 20 or 15°C. The temperature-maintenance systems of *C. elegans* have been described previously (Malone et al., 2003). Full-length *dhc-1* was PCR amplified from cDNA *yk1024* using primers 5′-GGCTTATGCCCAACACTGCAAC-3′ and 5′-GTCCGCGATGTCATGCACTGTCGCGG-3′ cloned into Ndel and Pal sites of vector pGAKT7 and shifted to the pGADT7 vector. Plasmids were introduced into the yeast AH1109 strain, and selection was performed on synthetic defined/-Leu/-Trp plates. Cotransformants were inoculated into liquid YPD medium and grown to OD600 0.8. 1 ml culture was centrifuged and washed once with 1 ml of water and resuspended in 1 ml of water. 10 µl of a 1:100 dilution in water was inoculated on a synthetic defined/-Leu/-Trp/-His/-Ade plate and incubated at 25°C.

#### Microtubule depolymerization and regrowth

We cultured gonads from strain syn-4(ok372); xla88 (gfp::syn-4) for drug treatments as described previously [Wolke et al., 2007]. We placed animals in embryonic culture medium (ECM; 84% Leibovitz L-15 medium [Invitrogen], 9.3% FBS [Hyclone], 4.7% sucrose, 0.01% levamisole, and 2 mM EGTA) on a coverslip and cut at the pharynx. We transferred extruded gonads to a 1.5-ml centrifuge tube containing ECM and 10 µg/ml nocodazole. After 1 h, gonads were washed twice with 1 ml ECM then resuspended in ECM and incubated for 0, 20, 30, or 60 min. All gonads were fixed and stained with anti-tubulin antibody as described in Immunofluorescence and microscopy.

#### Ultrastructural analysis

Intact young adult nematodes were fixed and embedded for electron microscopy by conventional methods (Hall, 1995). A two-step fixation was used: animals were first fixed in buffered aldehydes followed by post-fixation with osmium tetroxide and en bloc staining with uranyl acetate. Serial thin sections were examined with an electron microscope (CM10; Philips). The distal gonad was observed in lengthwise sections to better display the relationship between the developing germline, the rachis, and the overlying somatic gonad. Additional TEM images of the gonad in animal NS53 can be viewed at http://www.wormimage.org.

#### Online supplemental material

Fig. S1 shows that the *zyg-12(ok350)* has a normal gonad at 25°C. Fig. S2 demonstrates that *sun-1* RNAi disrupts germline nuclear position. Fig. S3 presents confocal images of microtubule architecture in the germline of wild type and *zyg-12(ct350)* grown at 25°C. Fig. S4 shows 3D projection images of germline microtubules. Fig. S5 shows live and GFP antibody–immunostained stages of the germline of animals expressing *tubulin::GFP* and shows imaging regrowth of microtubules at time points after nocodazole removal. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200902101/DC1.

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