The Intestinal Chemokine Thymus-expressed Chemokine (CCL25) Attracts IgA Antibody-secreting Cells

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Abstract

Immunoglobulin A (IgA) provides protection against pathogens at mucosal surfaces. Chemotactic responses have been hypothesized to target IgA plasma cells involved in mucosal immune responses. We show here that thymus-expressed chemokine (TECK, CCL25) is a potent and selective chemoattractant for IgA antibody-secreting cells (ASC), efficiently recruiting IgA-producing cells from spleen, Peyer’s patches, and mesenteric lymph node. Cells secreting IgA antibody in response to rotavirus, an intestinal pathogen, also respond well. In contrast, IgG- and IgM-ASC respond poorly. Epithelial cells in the small intestines, a principal site of IgA-ASC localization and IgA production in the body, highly and selectively express TECK. The migration of IgA-ASC to the intestinal epithelial cell chemokine TECK may help target IgA-producing cells to the gut wall, thus helping define and segregate the intestinal immune response.

Key words: homing • migration • chemokine • B cell • isotype

Introduction

Humoral immunity in mucosal tissues, especially the gastrointestinal mucosa, is characterized by local production and secretion of IgA, and the intestinal lamina propria (LP)* harbors many IgA-producing plasma cells initially recruited from circulating antibody-secreting cells (ASC; for a review, see references 1 and 2). In contrast, IgG-ASC predominate in systemic (nonmucosal) chronic inflammatory sites, for example in arthritis or soft tissue inflammation, and IgM-ASC, which produce the initial wave of low affinity antibodies during the primary immune response, are prominent within lymphoid organs (3). Early studies demonstrating mucosal lymphoid-derived ASC migration to factors in mouse colostrum led to the hypothesis that ASC might be targeted to mucosal surfaces by epithelium-derived chemoattractants (4). Since chemoattractant cytokines (chemokines) play major roles in the in vivo homing of other leukocyte subsets to lymphoid and extralymphoid tissues (5, 6), we set out to assess the chemokine responsiveness of cells secreting antibodies of different isotypes.

Materials and Methods

Chemotaxis. Lymphocytes from mouse spleen, Peyer’s patch, mesenteric lymph node, and bone marrow (BM) from male and female C57Bl6/J mice were isolated and migration assays were performed as described previously (7). Briefly, 2 × 10^5 lymphocytes were placed in 5-μ Transwell inserts (Costar) in wells with medium alone (basal) or medium containing chemokines at concentrations shown previously to be optimal for chemotaxis of various responding BM and peripheral leukocyte populations (7) (100 nM mKC, 100 nM monokine induced by IFN-γ (MIG), 50 nM stromal cell–derived factor (SDF-1α), 500 nM mBLC, 100 nM mMIP-1α, 1 nM mEotaxin, 100 nM hTARC, 100 nM mRANTES, 100 nM hMIP-3α, 100 nM hELC, 100
IgG-conjugated Armenian hamster anti–mouse IgM (BD PharMingen), IgG1–FITC (Clone R-35–95; BD PharMingen), or rat IgG2a–FITC as a negative control (Clone R-35–95; BD PharMingen). In one experiment, anti–mouse CD11c was added to this depleting antibody cocktail to exclude possible dendritic cell contamination. Biotinylated antibodies were visualized with streptavidin-Cy-Chrome (BD PharMingen) and FITC-staining was amplified using the Alexa Fluor 488 Signal Amplification kit for Fluorescein-conjugated probes (Molecular Probes). The sort gate for ASC was set for B220(hi), Thy-1.2–, IgD–, CD11b/Mac-1– ± CD11c– large lymphocytes and the FITC amplified background was determined using rat IgG2a–FITC stained cells. Sort gates for IgA+ and IgG+ ASC were identical. Sorted cells (generally 3,500–5,000 IgA+ or IgG+ ASC phenotype cells) were lysed in guanidinium buffer and the total RNA isolated by phenol/chloroform extraction. The RNA was reverse transcribed using the Advantage RT-for-PCR kit (CLON-TECH). PCR and Southern Blot analysis were performed as described previously (11). The intron-spanning PCR primers for mouse CC chemokine receptor (CCR)9 were 5'-CCTC-TCTTTACAGACCAGAC-3' and 5'-GTCATGGTCTTT-ACTCTTGTCG-3'. As a loading control, PCR products for mouse GAPDH were created using intron-spanning PCR primers that were 5'-CCATGTTTGTGATGGGTGTG-3' and 5'-CCTCTTTGATGTCATCATAC-3'.

Isolation of and Chemotaxis by LP Lymphocytes. LP lymphocytes (LPLs) were isolated from normal murine small intestine as described in reference 12. Briefly, 4–5 small intestines were cleared of Peyers' patches, cut open longitudinally, cut into short 5-mm segments, and washed at room temperature with vigorous shaking four times in divalent cation-free HBSS supplemented with 5 mM EDTA, 25 mM Hepes, and 2.5% antibiotic-antimycotic (Sigma-Aldrich) to remove epithelial cells and intramural lymphocytes until no more shedding occurred. Intestines were then washed twice in RPMI 1640 with 10% FCS/15 mM Hepes/2.5% antibiotic-antimycotic. LPLs were isolated by shaking the intestinal pieces in the RPMI 1640 supplemented with 20% BCS/25 mM Hepes/antibiotic-antimycotic and 300 U/ml collagenase Type VIII (Sigma-Aldrich) for three 40-min sessions. At the end of each 40-min incubation, released cells were immediately washed in RPMI 1640/10% BCS containing penicillin-streptomycin to remove and neutralize the collagenase. LPLs were allowed to recover in RPMI 1640/10% FCS containing penicillin-streptomycin in a CO2 incubator for 2 h before any analysis. Migration of LPLs was performed in RPMI 1640 supplemented with 0.5% BSA instead of serum to decrease background migration of lymphocytes. Responding cells were stained as outlined in the figure legend.

ASC Enrichment for Chemotaxis Assays. Mesenteric lymph node cells were harvested 10 d after conventional oral inoculation of mice with RV (8, 9). The cell preparation was enriched for ASC (B220(hi)/IgA–, IgD–, nonT cells) by precoating goat anti–rat IgG-conjugated Dynalbeads (Dynal) with rat anti–mouse CD4, CD8, and IgD antibodies (all FITC conjugated) individually. The individually coated beads were combined, washed, and incubated with the cell preparation for 45 min at 4°C. CD4+, CD8+, and IgD+ cells were removed with a magnet and the unbound ASC-enriched population was transferred and placed in culture for 1 h at 37°C to recover before assay.

Results and Discussion

We initially analyzed ASC from spleen, a central filtering and collecting organ for circulating immune cells that comprises plasma cells of each isotype. Spleen cells were isolated, placed in an upper Transwell chamber, and allowed to migrate to individual chemokines placed in the bottom well. ASC in the starting population and in the bottom
well after migration to specific chemokines or control media were enumerated by conventional ELISPOT assays, and the percent of ASC migrated was determined. The migration of naive follicular B cells, a population that circulates through secondary lymphoid tissues but not the gut wall, is presented for comparison.

Fig. 1 A shows that IgM-ASC failed to respond well to any chemokine tested, although they migrated above background to SDF-1α, (CXCL12), a widely expressed chemokine ligand for CXC chemokine receptor (CXCR)4 that is active on many lymphocyte subsets. IgG–ASC (Fig. 1 A) displayed a detectable but weak SDF-1α response, as well, but also migrated significantly to MIG (CXCL9), a ligand for CXCR3. CXCR3 and its ligands have been implicated in inflammatory T cell migration (13); thus, migration to MIG may help IgG–ASC enter diverse tissue sites of inflammation.

The most dramatic chemotactic responses, however, were displayed by IgA–ASC (Fig. 1 A, right panel). These IgA-producing cells respond well to SDF-1α (a chemo-
kine whose receptor is expressed by most leukocytes), but also to TECK (CCL25). In control experiments, incubation with TECK had no effect on the numbers of IgA–ASC detected or on the amount of IgA produced per ASC during overnight cultures (data not shown), indicating that the ASC arrived in the bottom chemoattractant wells by migration. A representative ELISPOT well illustrating IgA–ASC migration to TECK is presented as Fig. 1 B. Consistent with their efficient response to TECK, sorted IgA⁺ (but not IgG⁺) ASC express abundant messenger RNA for the TECK receptor, CCR9 (Fig. 1 C; note the presence of two bands, indicating expression of both CCR9 splice variants).

Unlike most circulating naive and memory lymphocytes including follicular (IgD⁺) B cells (5, 7) (illustrated in Fig. 1), neither IgA– nor IgG–ASC migrated to the lymphoid tissue-expressed CCR7 ligands ELC/CCL19 (Fig. 1) or SLC/CCL21 (data not shown), chemokines implicated in homing to lymphoid organs. These results are consistent with recent studies showing that ASC phenotype (B220⁻/CD138⁺) splenocytes from alum/nitrophenyl chicken γ-globulin–immunized mice migrate to SDF-1α but not ELC or SLC (14). Similarly, Wehrli et. al. has suggested that early ASC phenotype cells can migrate to ELC and SDF-1α, but that they largely lose these responses in association with exit from their lymphoid sites of generation (15). Thus most ASC may be programmed to migrate to extralymphoid sites, rather than recirculate through secondary lymphoid tissues.

IgA–ASC are thought to be induced in the intestine-associated Peyer’s patch and mesenteric lymph nodes. They then travel via the lymphatic system through the thoracic duct into the bloodstream. Many localize to or pass through the spleen on their way to populating the intestinal LP (16, 17). Some IgA-ASC also migrate to and reside in the BM, where they contribute to systemic IgA production (18). As shown in Fig. 2, TECK responsiveness is a common feature of IgA–ASC harvested from Peyer’s patches and mesenteric lymph nodes, as well as spleen. On average, BM IgA–ASC responded relatively less well to TECK (although the difference observed is not statistically significant) (Fig. 2). In fact, migration of BM IgA–ASC to TECK and SDF was variable and in some experiments was significantly less than migration by IgA–ASC from MLN or spleen. These differences may reflect variability in the proportion of terminal differentiated plasma cells in the BM (see below), where mature plasma cells are expected to be present at a higher frequency than in lymphoid organs.

The development of naive B cells into differentiated IgA–ASC is associated with the gain of surface IgA expression and a progressive decrease in B220 levels; few if any ASC express B220 at the high levels seen on naive or germinal center B cells, and many terminally differentiated tissue plasma cells are B220neg (19). The gut LP is a major site of IgA–ASC recruitment and the relative high percentage of IgA–ASC in this organ allowed us to address the B220 levels of the TECK-responsive IgA–ASC. (The cytological and histochemical terms “plasmablasts” and “plasma cells” have been used inconsistently in reference to ASC expressing varying levels of B220 [references 14, 15, 19, and 20], and will not be used here). Fig. 3 A illustrates that both IgA⁺/B220intermediate (B220mid) and IgA⁺/B220high large lymphocytes are found in the LP, and we have confirmed that both of these populations comprise IgA–ASC (unpublished data). Fig. 3 B demonstrates that the IgA⁺/B220mid LP IgA–ASC population responds much better to TECK than IgA⁺/B220high ASC. The LP IgA⁺/B220high ASC population had also lost its responsiveness to SDF-1α (similar to a B220high/CD138hi lymph node population described by Wehrli et. al., reference 15). Splenic TECK-responsive IgA–ASC were also restricted to the IgA⁺/B220mid but not IgA⁺/B220high ASC populations (data not shown). IgA⁺/B220high ASC may represent cells that have terminally homed to the site of their final residence with no further need for migratory receptors.

We next asked whether cells producing antibody in response to a well-defined intestinally restricted pathogen could also migrate to TECK. RV, a double-stranded RNA virus, exclusively infects the small intestine villous epithelium in humans and mice, causing diarrhea (21). RV is responsible for up to a million childhood deaths per year. Protection against RV in immunodeficient mice can be conferred by transfer of immune B cells, and immunity correlates with anti-RV IgA but not anti-RV IgG levels in
the mouse model (22, 23). Mesenteric lymph node cells were harvested from mice 10 d after RV infection, and cells migrating to TECK or SDF-1α were tested for the isotype of anti-RV antibody production by modified (antigen-specific) ELISPOT assay (8, 9). RV-specific IgA-ASC represent ~10–12% of total IgA ASC in mesenteric nodes at this time (9). Fig. 4 shows that these RV-specific IgA-ASC migrate strongly to TECK. In contrast, in an experiment in which mesenteric node IgG-ASC were assessed in parallel, RV-specific IgG-ASC did not respond to TECK above background (data not shown). The frequency of RV-specific IgA-ASC migration to TECK was comparable to that of the total IgA-ASC population in the same experiments. We conclude that IgA-ASC induced by small intestinal infection with RV display efficient chemotaxis to TECK.

Migration to the widely expressed chemokine SDF-1α is a property shared with most mature leukocytes, and many other cell types as well (24). Such a general response may contribute to motility within tissues or retention within a tissue (14), but is unlikely to control differential tissue and subset-specific cell homing. In contrast, recent studies of TECK tissue expression patterns reveal that it is critically positioned to contribute to IgA–ASC localization in the gut. TECK mRNA is highly expressed in the thymus, where it is hypothesized to participate in T cell development (25, 26). In the periphery however, TECK mRNA is largely restricted to the gastrointestinal tract, especially the small intestinal mucosa (27, 28). In situ hybridization and immunohistochemistry reveal TECK expression by epithelial cells in the jejunum, duodenum and ileum, especially but not exclusively in the crypt regions near the base of intestinal villi (28, 29). This region is enriched in MadCAM-1+ venules supporting extravasation of homing lymphocytes, as well. Additional chemoattractants, possibly including SDF-1α (30), may support further migration and dispersal of ASC throughout the intestinal LP. In contrast, TECK is poorly expressed by nonintestinal epithelial tissues (28). Thus, the studies presented here lead to the hypothesis that selective expression of TECK by intestinal epithelial cells may serve to efficiently recruit IgA–ASC to the gastrointestinal LP, and it will be of interest to assess the role of TECK in circulating ASC interactions with LP venules, and in diapedesis into the LP, in future in situ studies. Interestingly, TECK is expressed at lower levels in the colon than in the small intestines. It could contribute to ASC migration there as well, but other chemoattractants may play parallel roles in the homing of IgA–ASC to the colon and other mucosal tissues.

Our results provide the first demonstration that ASC respond to chemokines and demonstrate further the specialization of chemokine responses can be associated, at the population level, with ASC specialization in terms of isotype expression. The selective recruitment of IgA–ASC by the intestinal chemokine TECK not only provides a mechanism for the recruitment and/or retention of mucosal ASC in the intestinal wall as originally hypothesized by Lamm (4), but may also provide a paradigm in which tissue-selective chemokine expression by specialized epithelia helps determine the character of local humoral and cellular immune responses.
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