Is insulin-like growth factor-1 involved in Parkinson’s disease development?

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Abstract
Parkinson’s disease (PD) is a neurodegenerative disorder that results in the death of dopaminergic neurons within the substantia nigra pars compacta and the reduction in dopaminergic control over striatal output neurons, leading to a movement disorder most commonly characterized by akinesia or bradykinesia, rigidity and tremor. Also, PD is less frequently depicted by sensory symptoms (pain and tingling), hyposmia, sleep alterations, depression and anxiety, and abnormal executive and working memory related functions. On the other hand, insulin-like growth factor 1 (IGF-1) is an endocrine, paracrine and autocrine hormone with several functions including tissue growth and development, insulin-like activity, proliferation, pro-survival, anti-aging, antioxidant and neuroprotection, among others. Herein this review tries to summarize all experimental and clinical data to understand the pathophysiology and development of PD, as well as its clear association with IGF-1, supported by several lines of evidence: (1) IGF-1 decreases with age, while aging is the major risk for PD establishment and development; (2) numerous basic and translational data have appointed direct protective and homeostasis IGF-1 roles in all brain cells; (3) estrogens seem to confer women strong protection to PD via IGF-1; and (4) clinical correlations in PD cohorts have confirmed elevated IGF-1 levels at the onset of the disease, suggesting an ongoing compensatory or “fight-to-injury” mechanism.

Keywords: Parkinson’s disease, IGF-1, Aging, Dopamine, Estrogens, Central nervous system

Background: aspects and clinical features of Parkinson’s disease
Parkinson’s disease (PD) was first medically described as a neurological syndrome by James Parkinson in 1817. Over 50 years later, Jean Martin Charcot was more exhaustive in his descriptions and distinguished bradykinesia as a separate cardinal feature of this illness [1]. PD is the second most common neurodegenerative disorder, being age one of the main risk factors for its development [2, 3]. Despite years of study, much is still unknown about its etiology and pathogenesis, although genetic and environmental factors are involved in the establishment of the disease. However, the identification of such factors, their interaction and the associated molecular pathways that lead to neurodegeneration are weakly comprehended.

There are multiple motor and non-motor symptoms that can develop in the course of PD (Fig. 1). This disorder is characterized by the appearance of progressive motor deficit, having four cardinal symptoms grouped in the acronym “TRAP”: tremor at rest, rigidity, bradykinesia and postural instability. To these, postural changes in flexion (camptocormia) and freezing are added as the most characteristic signs of the disease, impeding daily patient’s daily life activities [4, 5].

Tremor is the most common symptom of PD characterized by being at rest with unilateral and distal initiation at a frequency of 4–6 Hz, with a movement in prone-supine position like “counting coins”. This sign may occur in arms, legs, lips and jaw [5–8]. The rigidity observed in PD is described as an increase in muscle tone manifested by a resistance to passive movement. It is usually accompanied by a phenomenon called cogwheel
Bradykinesia is depicted by difficulty in movement planning, initiation and execution [10]. Other manifestations associated with bradykinesia are the loss of spontaneous movements, hypomimia, hypophonia and deficit in arm-accompanying movement when walking [10, 11]. Postural instability appears late in the course of PD and is described as the loss of postural reflexes. These last alterations are frequent in PD patients, mainly camptocormia, which is characterized by neck, trunk, elbows and knees flexion. Nonetheless, other postural changes may occur, e.g. Pisa syndrome [10].

The freezing phenomenon is described as an inability to achieve movements, typically occurred during walking, producing severe disability. It usually lasts less than 10 s, but it can be prolonged. Freezing is accompanied by a festive march and it can be aggravated when turning around during the march or in closed spaces [9].

**Physiopathology of Parkinson’s disease**

The function of basal ganglia is determined by the balance between direct and indirect pathways activated by gamma-aminobutyric acid (GABA) neurotransmitter. The direct pathway starts with the activation of D1 dopamine receptors by dopamine in the striatum, which inhibit (via GABA) the globus pallidus pars interna. This last brain structure, in the absence of dopamine would inhibit the thalamus. Consequently, the direct pathway hinders the inhibition of the thalamus, thus stimulating the cortex (Fig. 2) [12]. On the other hand, the indirect pathway starts with the activation of dopamine D2 receptors in the striatum, inhibiting the globus pallidus pars externa, which in turn would inhibit (via GABA) the subthalamic nucleus. Due to the lack of inhibition by globus pallidus, the subthalamic nucleus activates the globus pallidus pars interna, which hinders (via GABA) the thalamus, hence inhibiting the cortex (Fig. 2) [12]. As follows, the direct pathway results in an excitation of the cortex, as opposed to the indirect pathway, where an inhibition of the cortex occurs.

The balance between these two pathways, regulated by dopaminergic afferent neurons in the substantia nigra pars compacta ascending to the basal ganglia, is progressively affected in PD. Thus, gradual loss of dopaminergic projections hinder the direct pathway, stimulating the globus pallidus pars interna that inhibit (via GABA) the thalamus leading to cortical dysfunction (Fig. 2) [12].

Motor activation alterations in PD patients reflect basal ganglia projection dysfunctions towards motor cortex as a result of the nigrostriatal degeneration, typified by progressive cell death in the central nervous system (CNS). The abnormal neuronal activity in areas receiving excessive inhibition of thalamo-cortical projections in PD has been evident in neuroimaging studies where they are hypoactive, such as the primary motor area (M1), supplementary motor area (AMA) and the lateral prefrontal cortex (DLPFC) [12].

Nonetheless, PD pathophysiology is not fully understood. It can be assumed that PD establishment depends upon various mechanisms, such as dopamine depletion, direct and indirect pathways alterations of the basal ganglia, dysfunction of the basal ganglia to the cortex and abnormal cortical plasticity [12]. PD motor
manifestations begin focally, typically in one limb segment, when dopamine concentrations fall below 60–70% in the contralateral striatum [12].

The initial pathogenesis of PD consists in a cascade of events leading to cell death, which includes oxidative stress, excitotoxicity via glutamate pathways, impaired mitochondrial functions, protein misfolding and aggregation due to ubiquitin-proteasomal system dysfunction, altered lysosome and chaperone-mediated autophagy and the development of cytoplasmic inclusion bodies (called Lewy bodies) (Fig. 3, left panel) [13]. Inflammation and humoral immune reactions may contribute to cell death processes through apoptosis [12].

Lewy bodies contain neuro-filament proteins and ubiquitinated α-synuclein, being their presence an important factor that allows to classify pathological changes in PD progression: stage 1, involves changes in dorsal motor nucleus and olfactory bulb associated with the loss of olfactory function, commonly seen in pre-clinical PD; stage 2, includes Lewy body formation in pons and medulla; stages 3 and 4, produce clinical motor symptoms; and stages 5 and 6, involve damage in neocortical areas, that result in cognitive problems and dementia [14].

Oxidative stress is an alteration observed in PD and other neurodegenerative diseases, e.g. amyotrophic lateral sclerosis and Alzheimer’s disease. The brain is especially vulnerable to this oxidative environment, due to several factors: (1) the large fraction of oxygen consumed by this organ [15]; (2) its high metal ion levels that can react with specific reactive oxygen species (ROS) to form hydroxyl radicals responsible for the oxidative damage [16, 17]; and (3) the elevated prevalence

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**Fig. 2** Basal ganglia dysfunction and physiopathology of Parkinson’s disease. The progressive loss of ascending dopaminergic projections is the key factor in the establishment of Parkinson’s disease. Continuous lines represent the normal function of basal ganglia; dotted lines represent the normal function of brain segments without the previous inhibition/activation pathway; and red arrows represent the alterations in basal ganglia observed in PD.
Cellular and molecular mechanisms in Parkinson’s disease

1. Mitochondrial function and mitophagy
   • Parkin
   • PINK1
   • DJ-1
   • CHCHD2
   • POLG1
   • SREBF1

2. Lysosome autophagy pathway
   • LRRK2
   • VP35
   • DNAJC13 (REM-8)
   • ATP13A2
   • GBA
   • SCARB (LIMP-2)

3. Ubiquitin-proteasome system
   • Parkin
   • FBXO7

4. Neurites structure
   • LRRK2
   • MAPT (tau)

5. Protein aggregation
   • SNCA (α-synuclein)
   • MAPT (tau)

6. Protein and membrane trafficking
   • VPS35
   • DNAJC13 (REM-8)
   • LRRK2
   • GAK
   • RAB7L1
   • RAB39B

7. Synaptic function and dopamine neurotransmission
   • SNCA (α-synuclein)
   • LRRK2
   • SYNJ1 (synaptojanin1)
   • GCH1
   • STX1B (syntaxis-1B)

8. Prion-like transmission:
   • SNCA (α-synuclein)

9. Oxidative damage

Cellular and molecular mechanisms in IGF-1 deficiency

1. Mitochondrial dysfunction sensitive to IGF-1 therapy: a) decreased MMP; b) increased mitochondrial ROS production; c) reduced ATP synthesis

2. Reduced gene expression related to mitochondrial protection and cytoprotection.
   • Bcl2
   • Gadd45a
   • Gadd45b
   • Serpinb1b
   • Serpinb1

3. Over expression of pro-death and pro-oxidative genes in brain of mice with IGF-1 partial deficiency.
   • Ptgs2
   • Nos1
   • Saa1
   • Crlf1
   • Tgfb2

4. Mitochondrial damage and pro-apoptotic genes over expression.
   • Bid
   • Apaf1
   • Grb2
   • Parp1
   • Ulk1

5. α-synuclein modulation:
   • Hyperexpression of α-synuclein in mice with partial IGF-1 deficiency.
   • Direct interaction of α-synuclein and AKT regulates IGF-1 signaling.

6. Neurotrophic factors in IGF-1 deficiency:
   • Overexpression: Bdnf, Ntf3
   • Reduced expression: Manf, Cntf
   • Reduced expression of signaling pathway gene (GDNF Family): Ret

7. Oxidative damage: Increased oxidative damage that reverted to normal by IGF-1 therapy.

Fig. 3  Parallelism between the cellular and molecular mechanisms involved in the pathogenesis of Parkinson’s disease and IGF-1 deficiency (modified from Kalia et al. [8])
of polyunsaturated fatty acids in brain membranes, which are particularly vulnerable to oxidative damage.

Recently, several studies in animal models have shown that various neurotrophic factors, when administered exogenously, reduce the vulnerability of dopaminergic neurons. One of these factors is the glial cell line-derived neurotrophic factor (GDNF), which seems to be effective in the treatment of PD [18].

As aforementioned, age is the major known risk factor for PD. It is important to notice that endogenous levels of some neurotrophic factors decrease with age. This idea suggests that neurotrophic factors loss due to aging may contribute to the etiology of PD [19].

**Prevalence of Parkinson’s disease**

The prevalence of PD increases with age. To date, several meta-analyses have been conducted to obtain worldwide age and sex-related estimates for PD. In 2016, approximately 6.1 million people had PD, most belonging to high and middle socio-demographic index (SDI) countries, such as North America and Canada (Table 1) [3]. This rate is 2.4 times higher than in 1990 (2.5 million people with PD). However, such increase in PD patients worldwide between 1990 and 2016 is not explained entirely by the augmentation in elder people [3].

Age-standardized prevalence of PD shows that this disorder is more frequent between 65 and 79 years, especially in men, being this ratio 1.40 times higher in men than women in 2016 (Table 2) [3, 20–27]. Consequently, additionally to elder age, male sex is recognized as a prominent risk factor in the development of PD. Also, PD incidence in men continues to rise after 80 years old; being the age of PD onset about 2 years later in women than men [28].

**Parkinson’s disease, estrogens and sex differences.**

**Estrogen actions via IGF-1 activity in brain**

Estrogens are steroid hormones that promote female sex characteristics and reproductive capability in several organs, including brain. These hormones can cross the blood brain barrier and also the brain can endogenously produce estrogen from cholesterol [29].

Estrogens have been first identified for their role in female reproductive cycle. Nevertheless, their protective functions against chronic and degenerative diseases are becoming more relevant. These new roles may be responsible for the long-recognized advantages that women have over men in retaining their general health and achieving greater longevity [30].

Most of the circulating estrogens in premenopausal women are produced mainly by the ovaries, and in small quantities by other peripheral tissues, such as liver, adrenal gland, mammary glands, adipose tissue and brain [31]. In males, testes produce only 20% of circulating estrogens, being mostly locally produced by adipose tissue, brain, skin and bone. Herein, aromatases convert testosterone to estrogen [32].

Estrogen receptors have beneficial effects on memory and cognition, being widely distributed in male and female brain [33].

**Table 1 Worldwide prevalence of Parkinson’s disease**

| Country       | General population | References |
|---------------|--------------------|------------|
| High SDI countries | 2.1 million (34.4%) | [3]        |
| High-middle or middle SDI countries | 3.1 million (50.8%) |          |
| Low SDI countries    | 0.9 million (14.8%) |            |
| Worldwide           | 6.1 million Female: 2.9 million (47.5%) Male: 3.2 million (52.5%) |            |

SDI socio-demographic index

**Table 2 Worldwide age-related prevalence of Parkinson’s disease**

| Age-related prevalence of Parkinson’s disease | Male | References |
|----------------------------------------------|------|------------|
| 4–49 years                                    | 3.26 per 10,000 person-years | 3.57 per 10,000 person-years | Hirsch et al. [21] |
| > 80 years                                    | 103.48 per 10,000 person-years | 258.47 per 10,000 person-years |

**Age of Parkinson’s disease patients in countries with high prevalence of the disease**

| Country                  | Age general population | References |
|--------------------------|------------------------|------------|
| United States of America (USA) | 70.5                   | Van Den Eeden [22] |
| Japan                    | 68.7 ± 10.3            | Yamawaki et al. [26] |
| Europe                   | 61.6 ± 9.73            | Pagano et al. [24] |
| China                    | 58.7 ± 10.55           | Song et al. [25] |
| Mexico                   | 56.9 ± 11.8            | Rodriguez-Violante et al. [23] |
female brains [33]. To date, several effects of estrogens in neurodegenerative diseases, such as PD, have been described [34]. Some of them are implicated in cognition, synaptic plasticity, memory, neurogenesis and neuroprotection [35]. Numerous studies have suggested that the molecular mechanism by which estradiol exerts its neuroprotective effects involves phosphatidyl inositol-3-kinase (PI3K) signaling pathway activation. This cascade is activated by insulin and insulin-like growth factor 1 (IGF-1) in brain [36]. As follows, there is a certain association between IGF-1 and estrogens and their protective effects exerted in nigrostriatal dopamine neurons after 6-hydroxdopamine lesions; suggesting them as an useful therapy to palliate PD symptoms [37].

In this manner, women with PD typically have a more benign phenotype with slower progression of the disease than men, and the incidence and prevalence of PD is higher in postmenopausal than in premenopausal women of similar age [38]. This could be due to the higher estrogen activity, which increases dopamine levels in the striatum, thus promoting neuron survival and neuroprotective actions [38, 39]. In addition, estrogens might prevent Lewy body deposition through specific α-synuclein anti-aggregation and fibril destabilization properties [40]. In the last decade, the modulation of α-synuclein aggregation is emerging as an innovative therapeutic strategy for PD treatment. In this fashion, small organic molecules, e.g. polyphenols such as curcumin, have been widely tested for their ability to inhibit α-synuclein aggregation [41, 42].

Until now, estrogen deficiency has been associated with PD. In industrialized countries approximately 0.3% of the entire population and near 1% of people older than 60 years have PD related to estrogen deficiency due to loss of function mutations [43]. For example, in European countries this prevalence ranges from 65.6/100,000 to 12,500/100,000 people, with an incidence from 5 to 346/100,000 [44–46].

Additionally, a range of behavioral and lifestyle preferences associated with gender differences, e.g. diet, exercise, smoking and caffeine, are emerging as potential PD risk modifiers [47]. The nigrostriatal dopaminergic pathway may thus underlie the biological sex differences vulnerability, and could also be responsible for the dimorphic sexual actions of estradiol, which shelter females against striatal dopamine loss in experimental PD. However, in males such pathway fails to protect or may even worsen striatal lesions. Studies in animal models with estrogen deprivation exhibit dopaminergic neuron loss, and altered dopaminergic metabolism and transporter uptake, which can be partially reverted by exogenous estrogen administration [48]. In consequence, there is a link between longer estrogen exposure during lifetime and both decreased PD risk and milder features in women [48]. All these results open a door to potential hormone-based therapies as novel approaches to develop treatments which can delay and possibly halt the progression of the disease [27].

**Physiological roles of insulin-like growth factor 1**

IGF-1 is a 70-amino acid polypeptide hormone with several functions [49]. IGF-1 described roles include tissue growth and development, insulin-like activity, proliferation, pro-survival, anti-aging, antioxidant and neuroprotective, among others. Also, IGF-1 is a major relevant hormone in embryological and postnatal states [50]. Although it is mainly produced by the liver for endocrine activities, virtually every tissue is able to secrete IGF-1 for autocrine and/or paracrine purposes [51].

Until now, the best-known conditions of IGF-1 deficiency in humans are growth hormone insensitivity (GHI), advanced liver cirrhosis and aging, including cardiovascular and neurological diseases associated to aging [52]. More recently, intrauterine growth restriction [53] and metabolic syndrome [54] seem to be other forms of IGF-1 deficiency. Besides these conditions, several diseases have been recently proposed to be either consequence or cause of both systemic or local IGF-1 deficiency [52].

**Expression of IGF-1 in central nervous system**

IGF-1 is produced by all major central nervous system (CNS) cell types, with an area-specific pattern production, being higher in the brain stem, cerebellum, cerebral cortex and the striatum [55, 56]. This hormone peaks during development, declining with age. IGF1 receptor (IGF1R) is expressed in both neural stem cells and all neural cells throughout life [57, 58], being highly produced among the choroid plexus, hippocampus and parahippocampal areas [59]; but also to a less extent in the amygdala, cerebellum and cortex [60–62].

Several studies have shown that IGF-1 expression is increased in frontal cortex in PD compared to controls, while IGF1R is reduced in white matter and amygdala [63, 64]. As aforementioned, IGF-1 levels and signaling cascade are reduced during aging, due to an increased hyper-phosphorylation of insulin receptor substrates (IRSs), one of the first proteins activated in insulin and IGF-1 pathways, thus impairing the activation of downstream molecules such as PI3K [65–67].

IGF-1 mRNA expression levels in brain are relatively low compared to IGF1R, suggesting an important role for peripheral IGF-1 [68]. It was first reported in rats more than 20 years ago that systemic IGF-1 could cross the blood–brain-barrier by transcytosis [59] and via the choroid plexus [68]. Nonetheless, there is no significant
correlation between serum and cerebrospinal fluid IGF-1 levels and therefore, it seems that systemic IGF-1 is not a major source of IGF-1 for the CNS [69]. For this reason, the endocrine role of serum IGF-1 is still a matter of debate.

**IGF-1 actions in the central nervous system**

Little is known about the mechanisms that regulate IGF-1 expression in brain, but there is evidence that growth factors, nutrition, exercise and external insults (e.g. hypoxic/ischemic, demyelination, stereotactic, electrolyte and cryogenic injuries) influence IGF-1 expression in vivo [59, 70–74].

IGF-1 is a potent neuroendocrine regulator of the CNS development, maturation and plasticity [75], even being responsible for up to 40% of brain’s weight [76–80]. This hormone also has a role in brain differentiation, either from mesenchymal stem cells into neuronal lineages [81] or from neural stem cells towards neuronal or glial phenotypes [81]. IGF-1 has been found to exert profound effects on neurogenesis in the developing [78] and adult [82] brain, as well as in axonal development, synaptogenesis [68, 83] and migration [68, 84, 85]. Similarly, IGF-1 has shown to be indispensable for exercise-induced neurogenesis [86].

Several studies have found increased IGF-1 levels in the CNS following an injury, acting as an endogenous protective mechanism [87, 88]. This is in accordance with the increased IGF-1 levels observed at the beginning of PD pathophysiology (Table 3), suggesting that those whose IGF-1 concentrations do not rise enough will present a worst outcome. Similarly, murine models of peripheral IGF-1 concentrations do not rise enough will present a pathophysiology (Table 3), suggesting that those whose increased IGF-1 levels observed at the beginning of PD have to be determined, not only in neurons but in the entire web, particularly in astrocytes, in order to avoid overlooking results mediated through them. In this sense, astrocytes express IGF1R in the developing brain [107], and to a less extent in the adult [58], hence activating IGF-1 actions, such as proliferation [108], growth factor production [109, 110], cytoskeletal protein levels [108], intercellular communication and messengers [111–113], glutamate transport [114], glucose metabolism [115–117], and mitochondrial and membrane enzyme activity [118, 119].

The role of IGF-1 in adult astrocytes seems to be limited to modulate tissue damage response, where reactive astrocytes express high levels of IGF1R [120, 121] and IGF-1 [122]. Likewise, hepatocytes in damaged liver express IGF1R, a circumstance not observed under normal conditions [123–125]. In this context, pro-inflammatory signals activate astrocytes, turning them into activated pro-inflammatory cells that will secrete several factors (e.g. inducible nitric oxide synthase 2 -iNOS2-, cyclooxygenase 2 -Cox2-, tumor necrosis factor α -TNF-α, etc.) contributing to neuron damage. IGF-1 anti-inflammatory capabilities transform these cells into anti-inflammatory ones, ceasing to produce pro-inflammatory signals and thus, enhance the production of neuroprotective molecules, such as humoral superoxide dismutase (SOD), neurotrophic factors, etc. Both anti- and pro-inflammatory cell types converge on the phosphatase calcineurin, that turns gene expression to anti-inflammatory ones, ceasing to produce pro-inflammatory signals and thus, enhance the production of neuroprotective molecules, such as humoral superoxide dismutase (SOD), neurotrophic factors, etc. Both anti- and pro-inflammatory cell types converge on the phosphatase calcineurin, that turns gene expression to one of this phenotypes [126]. Moreover, it has also been reported a neuroprotective role for IGF-1 associated to mitochondrial protection and antioxidant defenses in aging animals [127].

Data from our group suggest that the mere partial IGF-1 deficiency in mice (heterozygous mice, Igf1+/−,...
| Patients | n   | Age (SD) | Sex | Treatment | UPDRS-III stage (SD) | Design                      | Results                                                                 | Peripheral blood IGF-1 (ng/mL) PD baseline/controls (SD) | References |
|----------|-----|----------|-----|-----------|----------------------|-----------------------------|------------------------------------------------------------------------|----------------------------------------------------------|------------|
| Early (< 3.5 years) vs. moderate (>= 4 years) PD | 37  | 64 (7)   | 15F/22M | Levodopa | 22 (9) (early)/38 (15) (moderate) | Longitudinal prospective cohort (3.5 years) | PD patients in moderate, but not early stages, showed significantly increased baseline IGF-1 levels. PD-1 level was higher in patients with PD and inversely correlated with the UPDRS-III score (r = -0.77) among PD patients. IGFBP-3 unchanged. IGF-1 level was not correlated with the UPDRS-III score and IGFBP-3 level was not significantly different between PD patients and controls. | 130 (26)/106 (24) p = 0.017 | Bernhard et al. [171] |
| Newly diagnosed idiopathic PD (Germany) | 15  | 69 (8.3) | 6F/9M | Drug-naive | 14.30 (3.3) | Cross-sectional cohort | IGF-1 level was higher in patients with PD and inversely correlated with the UPDRS-III score (r = -0.77) among PD patients. IGFBP-3 unchanged. IGF-1 level was not related to motor function in the healthy group. | 158.4 (40.4)/129.2 (29.1) p = 0.004 | Godau et al. [172] |
| Idiopathic PD (Germany) | 18  | 67 (9)   | 8F/22M | Levodopa-treated vs. untreated | 24.1 (8.3) (treated)/16.2 (4.1) (untreated) | Longitudinal (6 months) | IGF-1 was significantly higher in treated PD patients than in controls at all time points (all p < 0.001). IGF-1 levels were correlated with shorter disease duration (r = 0.56, p < 0.01). In the patient group, higher IGF-1 levels were correlated with shorter disease duration (r = 0.56, p < 0.01). In the healthy control group, higher IGF-1 levels were correlated with slightly impaired motor performance (r = 0.46, p < 0.05). In the untreated patient group, IGF-1 levels were significantly higher than healthy controls (p < 0.001). Treatment did not alter GH levels. | 149.06 (30.9)/98.96 (23.2) | Godau et al. [173] |
| PD | 38  | 68 (10)  | F and M | – | – | Cross-sectional cohort | Serum and CSF IGF-1 and IGFBP levels were higher in PD patients than controls (p < 0.001). CSF: 5.97 pg/mL (0.93)/4.40 pg/mL (0.58). Serum: 120.19 (40.86)/207.97 (19.51). | 130.3 (14.6)/144.5 (15.9) | Mashayekhi et al. [174] |
| PD (75), multiple system atrophy (MSA, 25), and progressive supranuclear palsy (PSP, 16) (Japan) | 116 | 68.1 (1.1) | 44F/35M | Drug-naive vs. levodopa-treated | 26.9 (1.8) | Cross-sectional cohort | Serum IGF-1 levels tended to be higher in early PD patients than controls. There was a negative correlation between serum IGF-1 levels and age in PD patients and controls. There was no significant correlation between disease duration and serum IGF-1 levels in PD patients. In controls, there was no significant correlation between serum IGF-1 levels and UPDRS part III. In PD and PSP patients, there was a negative correlation between serum IGF-1 levels and UPDRS part III. In early and drug-naive PD patients there was no significant correlation between serum IGF-1 levels and UPDRS part III. IGF-1 serum levels in PD patients with HY stage 2 were significantly higher than those in PD patients with HY stages 3-5. | 130.3 (14.6)/144.5 (15.9) | Numao et al. [175] |
### Table 3 (continued)

| Patients | n    | Age (SD)       | Sex     | Treatment | UPDRS-III stage (SD) | Design                  | Results                                                                 | Peripher al blood IGF-1 (ng/ml) PD baseline/controls (SD) | References |
|----------|------|----------------|---------|-----------|----------------------|-------------------------|----------------------------------------------------------------------|--------------------------------------------------------|------------|
| > 3 years PD with weight loss (11) vs. PD without weight loss (16) | 27   | 63.5 (8.8)/60.5 (8.6) | 6F, SM/8F, BM | Levodopa | 43.45 (17.26)/37.75 (22.17) | Cross-sectional cohort     | BMI was lower in all PD patients Serum leptin levels were lower in all PD patients Serum GH and IGF-1 levels were higher in all PD patients, mostly in PD with weight loss and without weight loss, respectively Serum active ghrelin levels were positively correlated with serum IGF-1 levels in the control group (p < 0.05; r = 0.67) but not among PD patients | With weight loss 191.73 (33.94), without weight loss 152.19 (49.62)/144.17 (24.24) (p < 0.05 between PD patients and PD patients with weight loss vs. controls) | Frizer et al. [176] |
| PD       | 25   | 67.9 (9.4)     | 5F/20M  | Treated, drug not specified | –         | Cross-sectional cohort     | IGF-1 and IGFBP-3 serum levels in PD patients showed no correlation with the duration and severity of the disease | 132 (42)/113 (51) | Tunsel et al. [177] |
| Early PD (< 2 years) (Italy) | 65   | 59.7 (8.3)     | 26F/39M | Drug-naive | 14.5 (6.7) | 2-year follow-up prospective cohort     | At baseline, serum IGF-1 levels were significantly increased as compared to healthy controls A positive correlation between IGF-1 levels and a specific executive function (phonological fluency) assessing cognitive flexibility was found After a 2-year follow-up, IGF-1 levels were positively related to verbal episodic memory, visuospatial abilities and attention/executive functions Low IGF-1 levels at baseline were independently associated to poor performance on specific cognitive tasks assessing verbal episodic memory and executive functions after 2 years | 91.6 (34.0)/79.1 (23) (p = 0.019) | Pellecchia et al. [178] |
| Early PD (< 2 years) (Italy) | 37   | 59.4 (8)       | 15F/22M | Drug-naive | 14.6 (7.1) | 12-month follow-up prospective cohort     | At baseline, serum IGF-1 levels were moderately increased Patients at the highest IGF-1 quartile presented higher mean dopaminergic scores (worse outcome) | 94.5 (37.5)/79.1 (23) (p < 0.011) | Picillo et al. [179] |
| Early PD (< 2 years) | 405  | 61.20 (9.7)    | 141F/264M | Drug-naive | 20.25 (8.93) | 5-year follow-up prospective cohort     | IGF-1 levels were similar in PD and controls Lower serum IGF-1 levels were associated to poor performances in cognitive tasks assessing executive function, attention and verbal memory | 136.6 (56.1)/134.45 (56.13) | Picillo et al. [180] |
| Meta-analysis covering de novo, drug-naive idiopathic PD patients | 166  | –             | –       | Drug-naive | –         | –                                     | Significantly higher serum IGF-1 levels among de novo, drug-naive idiopathic PD patients at baseline | –                                                   | Li et al. [181] |

HY Hoehn and Yahr score, MDS-UPDRS-III Movement Disease Society-modified UPDRS-III scale, SD standard deviation, UPDRS-III unified Parkinson’s disease rating scale

* Age, sex and body mass index (BMI, kg/m²), as well as the presence or absence of other medical factors known to affect IGF-1 levels, termed medical confounders: diabetes mellitus (reported in medical history or inferred by anti-diabetic medication intake), beta-adrenergic medication, depression (and/or antidepressant medication), neuroleptic medication, thyroid dysfunction, inflammatory diseases and cancer. All of them were taken into account when the study was carried out.
where one of the alleles of the Igf1 gene is disrupted) increased brain oxidative damage, inflammation, edema, apoptosis and impaired learning and memory capabilities; being all these alterations restored by IGF-1 replacement therapy [129]. Consistently, these IGF-1 deficient mice have an alteration in the blood–brain barrier due to a distorted genetic expression of extracellular matrix and intercellular adhesion proteins (unpublished data). Furthermore, IGF-1 deficiency showed in this experimental model, an altered gene expression pattern encoding neurotrophic factors and their receptors [130].

While most of the results found in the literature have been discovered using in vitro studies, other were discovered using animal models of CNS injury or IGF-1 deficiency. Even though these experimental models have been really advantageous in unravelling the neurobiology of IGF-1, clinical data is very scarce and so is our understanding of its therapeutic potential. Nevertheless, tremendous possibilities have been appointed towards its usage. Herein we will focus on PD; however, the degenerative neurological conditions with the larger impact on population have been somehow linked with IGF-1 alterations [131–134].

Neurodegenerative diseases related to aging as IGF-1 deficiency conditions

An increasing number of disorders have been reported highlighting the role of IGF-1 in many different organs and system functions. The knowledge concerning “IGF-1 deficiency conditions” or “GHI”, in which replacement therapy could be considered as an effective therapeutic strategy, is only recent.

The best-known condition of IGF-1 deficiency is Laron syndrome (also known as Laron dwarfism or GHI) [135, 136], predominantly found in children, characterized by alterations in growth hormone receptors (GHR) in the liver, and hence, reducing IGF-1 synthesis. In these patients, IGF-1 replacement therapy improves growth and development [137, 138].

Another IGF-1 deficiency condition, in this case in adults, is liver cirrhosis [139], where decreased IGF-1 serum levels were found in these patients. However, during the last decade, our research team has disclosed the causal link between IGF-1 deficiency and the severe malnutrition syndrome associated to liver cirrhosis progression. In fact, the exogenous administration of low doses of IGF-1 in experimental cirrhosis improved nutritional status and nitrogen balance [140], intestinal absorption [141–144], osteopenia [145, 146], testicular atrophy [147, 148], restored somatostatinergic tone controlling growth hormone (GH) secretion [149] and induced hepatoprotective actions in the liver associated with a reduction in oxidative damage and enhancing mitochondrial protection [150–153]. In addition, IGF-1 replacement therapy increased albumin circulating levels in cirrhotic patients [154].

Thirdly, aging may be considered as a recognized IGF-1 deficiency condition [155–157]. Although several studies have suggested that reduced GH/IGF-1 activity promotes longevity [158–160], a significant amount of evidence indicates that IGF-1 might play a role in several pathological conditions [102, 161–165] commonly seen during aging, associated with oxidative tissue damage. Precisely, several reports suggest that age-related changes in cellular and tissue function are linked to a decrease in anabolic hormones, GH and IGF-1 levels [155–157, 166]. Accordingly, previous studies from our group reported that administration of low doses of IGF-1 in aging rats improved glucose and lipid metabolism, reducing insulin resistance, increased testosterone levels and serum total antioxidant capability of the cell, and reduced oxidative damage in brain and liver, all of them associated with a normalization of antioxidant enzyme activities and mitochondrial function [167, 168].

In addition to these well-known IGF-1 deficiency conditions and its correlation with aging and cognition, several neurodegenerative disorders show low IGF-1 serum levels, such as PD, Alzheimer’s disease and cerebrovascular disease [132, 134]. Conversely, several neurodegenerative diseases, with the exception of PD, exhibit reduced GH-stimulation test response (with clonidine, arginine and growth hormone stimulating hormone, GHRH) [169]. This result could partly explain why in PD, high serum levels of IGF-1 are found (Table 3), in opposition with its related diseases. Moreover, a study disclosed that 5-HT 1-receptor-mediated growth hormone secretion neurotransmission was absent in PD patients, suggesting an alteration in serotonin signaling [170].

As described before, the GH/IGF-1 axis is involved in brain development, growth and function; so its progressive decrease during aging could be embroiled in the establishment of several human cerebrovascular diseases, that lead to neuronal degeneration and dysfunction, such as PD, multiple system atrophy, Lewy body disease, Alzheimer’s disease, vascular dementia, amyotrophic lateral sclerosis, stroke, etc. [131–133].

Theoretical relationship between Parkinson’s disease and IGF-1 deficiency

Noticably, there is a clear relationship between IGF-1 and PD (Fig. 3). The preclinical data presented in this review strongly suggests a neuroprotective role for IGF-1. As Table 3 depicts, IGF-1 levels in almost all PD cohorts are higher in both sera and cerebrospinal fluid of these patients, particularly at the symptom’s onset (UPDRS-III) and then equals out as the disease progresses.
These IGF-1 concentrations inversely correlate with the UPDRS-III score (i.e. with motor function), verbal episodic memory, visuospatial abilities and attention/executive functions; been not found such correlation on healthy subjects [171–181]. As it has been shown, lower IGF-1 levels at the beginning of PD are associated with a worse disease progression and prognosis. However, IGF-1 correlation with the disease onset, duration, severity, etc., still remains a hot topic.

Few genetic alterations have been linked to PD, being the most common the so-called idiopathic PD. Among them are mutations affecting α-synuclein [182–184], which leads to familial PD. However, the exact role of α-synuclein in the pathophysiology of PD, besides its aggregation to form Lewy bodies, remains unclear [185]. Recently, α-synuclein has been found to be essential for IGF-1 and other neurotrophic factors signaling activation in brain via protein kinase B (PKB or Akt). Actually, it has been hypothesized that α-synuclein long-known role in brain could be to transport, localize and solubilize Akt for neurotrophic factor receptors in brain cells [186]. In vitro studies have disclosed that decreased Akt phosphorylation due to lack of functional α-synuclein lead to poor IGF-1 signaling in neuronal and non-neuronal cells. This molecule, Akt, is a key survival signaling mediator of IGF1R in all cells, suggesting that IGF-1 protective effects in brain could be halted at the PD etiology itself.

Also, it has been proved that α-synuclein aggregation neurotoxicity is mediated by dopamine, and that IGF-1 confers protection against such aggregation and promotes signaling rescue through Akt, even in presence of dopamine [187]. Of interest, a recent study found an accurate micro-RNA profile alteration in post-mortem microdissected dopaminergic neurons. In this study, miR-126 particularly blocked IGF-1 signaling pathway, increasing cell vulnerability to 6-hydroxydopamine neurotoxicity [188]. In consequence, all this data demonstrates that cell response capability is decreased may be due to α-synuclein, miR-126, or dopamine cytotoxicity, dysregulating IGF-1 signaling pathway that lead to cell death and neurodegenerative features observed during the course of PD.

Several epidemiological studies have demonstrated a significantly increased risk for developing PD in individuals with central obesity [189, 190], an important result because IGF-1 and insulin signaling pathways are known to be dysregulated under metabolic derangements [54]. This suggests that insulin/IGF-1 signaling deficit could decrease neuroprotection and render the brain vulnerable to oxidative damage [191]. Furthermore, epidemiological studies have shown an association between the exposure to environmental toxins and an increasing incidence of PD in younger people [192–194]. Modern era exposure to toxins causes an elevated chronic grade of inflammation, alike obesity, that in turn antagonizes insulin/IGF-1 signaling, generating resistance and rendering the body without its physiological, trophic and protective effects. This assertion has been recently intimately linked to cardiovascular diseases [195].

As aforementioned, results from PD cohorts illustrated in Table 3 shows that lower IGF-1 levels at the beginning of the disease are associated with a worse progression and prognosis, being IGF-1 concentrations higher in PD cohorts at the symptom’s onset. These data could act as a compensatory mechanism due to the lack of IGF-1 signaling feedback and, also, it could be an endogenous protective factor released in response to cell death and oxidative damage promoted by the disease itself. Such hypothesis is further sustained by a progressive PD model study where IGF-1 was found to protect the nigrostriatal pathway, and that this protection was preceded by activations of key pro-survival signaling cascades, such as Akt [181, 196].

Conclusions
There are strong lines of evidence to correlate IGF-1 with PD establishment and development. Herein, the present review tries to summarize how IGF-1 and estrogen levels, together with their neuroprotective and anti α-synuclein aggregation properties, act in a harmonized manner to maintain brain homeostasis, particularly during aging process. From our experience, the mere partial IGF-1 deficiency in a murine model increases brain oxidative damage, inflammation, edema and apoptosis, also impairing learning and memory capabilities and the blood–brain barrier integrity. All these alterations are restored by exogenous IGF-1 replacement therapy.

As it is known, the pathogenesis of PD, the second most common neurodegenerative disorder, consists in a cascade of events including oxidative stress, excitotoxicity via glutamate pathways, impaired mitochondrial functions, protein misfolding and aggregation due to ubiquitin-proteasomal system dysfunction, altered lysosome and chaperone-mediated autophagy and the development of cytoplasmic Lewy bodies, eventually leading to cell death. Due to its neuroprotective functions, in the last decade several studies have been conducted in order to understand and resolve the linkage between IGF-1 and PD. In this way, numerous basic and translational reports have appointed brain protection and injury recovery functions of IGF-1 in CNS injuries and/or PD patients. These results highlight the protective and homeostatic key roles of IGF-1 in brain, along with its anti-apoptotic, pro-survival and protective features on cellular metabolism.
The aging process, where IGF-1 serum levels decrease with age, is one of the risk factors for PD development. However, several clinical correlations in PD patients have reported an augmentation in IGF-1 levels at the onset of the disease, which suggests, given the preclinical results, an ongoing compensatory or "fight-to-injury" mechanism, since as the disease progresses such elevation dissipates. Additionally, lower IGF-1 levels at the beginning of PD are associated with a worse disease progression and prognosis.

Together with age, male sex is another crucial risk factor for PD establishment. Estrogens seem to confer women a strong protection to PD due to their beneficial effects on memory and cognition. This estrogen related neuroprotection is achieved via IGF-1 signaling pathway, as estrogens activate PI3K to stimulate neuron survival and protection. Nevertheless, more experimental studies need to be performed to better understand the molecular mechanisms from IGF-1 cascade involved in PD.

In this fashion, it could be reasonable to think in a combined therapy with estrogens and IGF-1, since both molecules can cross the blood–brain barrier activating the IGF-1 signaling pathway to exert its favorable actions in brain and also to prevent α-synuclein aggregation (Lewy body deposition), thus avoiding PD development. This hypothesis is in accordance with recent studies where polyphenols, such as curcumin, are used to inhibit α-synuclein aggregation and prevent oxidative damage related alterations. Also, GDNF, a neurotrophic factor, could be used as a therapy for PD, because it can reduce the vulnerability of dopaminergic neurons. However, in order to solve this question, more experimential and clinical studies need to be done.

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Authors’ contributions
GAA, GFR, LE and IME participated in compiling data and elaborating the manuscript. ICC designed the review content, contributed to its writing and revised the conceptual contents until the final approval. All authors read and approved the final manuscript.

References
1. Goetz CG. Charcot on Parkinson’s disease. Mov Disord. 1986;1(1):27–32.
2. Erkkinen MG, Kim M-O, Geschwind MD. Clinical neurology and epidemiology of the major neurodegenerative diseases. Cold Spring Harb Perspect Biol. 2018;10(4):a033118.
3. GBD 2016 Parkinson’s Disease Collaborators. Global, regional, and national burden of Parkinson’s disease, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. Lancet Neurol. 2018;17(11):939–53.
4. Jankovic J. Parkinson’s disease: clinical features and diagnosis. J Neurol Neurosurg Psychiatry. 2008;79(4):368–76.
5. Rodriguez-Oroz MC, Jahanshahi M, Krack P, Litvan I, Macias R, Bezard E, et al. Initial clinical manifestations of Parkinson’s disease: features and pathophysiological mechanisms. Lancet Neurol. 2009;8(12):1128–39.
6. Olanow CW, Obeso JA. The significance of defining preclinical or prodromal Parkinson’s disease. Mov Disord. 2012;27(5):666–9. https://doi.org/10.1002/mds.25019.
7. Antony PMA, Diederich NJ, Krüger R, Balling E. The hallmarks of Parkinson’s disease. FEBS J. 2013;280(11):939–53. https://doi.org/10.1111/febs.12335.
8. Kalia LV, Lang AE. Parkinson’s disease. Lancet. 2013;383(9926):896–912.
9. Berg D, Lang AE, Postuma RB, Maertler W, Deuschl G, Gasser T, et al. Changing the research criteria for the diagnosis of Parkinson’s disease: obstacles and opportunities. Lancet Neurol. 2013;12(5):514–24.
10. Dimberg G, Jahanshahi M. Executive dysfunction in Parkinson’s disease: a review. J Neuropsychol. 2013;7(2):193–224. https://doi.org/10.1111/jnp.12028.
11. Juri C, Rodriguez-Oroz MC, Burguera JA, Guidi J, Obeso JA. Pain and dyskinesia in Parkinson’s disease. Mov Disord. 2010;25(1):130–2.
12. Sabatini U, Boulanouar K, Fabre N, Martin F, Carel C, Colonneso C, et al. Cortical motor reorganization in akinetic patients with Parkinson’s disease: a functional MRI study. Brain. 2000;123(Pt 2):394–403.
13. Olanow CW. The pathogenesis of cell death in Parkinson’s disease—2007. Mov Disord. 2007;22(SUPPL 1):S33–42.
14. Braak H, Ghebremedhin E, Rüb U, Bratzke H, Del Tredici K. Stages in the development of Parkinson’s disease-related pathology. Cell Tissue Res. 2004;318(1):121–34.
15. Mink JW, Blumenshine RJ, Adams DB. Ratio of central nervous system to body metabolism in vertebrates: its constancy and functional basis. Ann J Physiol. 1981;241(3):R203–12.
16. Connor JR, Benkovic SA. Chemical, biochemical, and molecular considerations. Ann Neurol. 1992;32(SUPPL):S51–61.

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17. Mills E, Dong XP, Wang F, Xu H. Mechanisms of brain iron transport: insight into neurodegeneration and CNS disorders. Front Med Chem. 2010;2(1):51–64.
18. Patel NK, Pavese N, Javed S, Hotton GR, Brooks DJ, Gill SS. Benefits of putaminal GDNF infusion in Parkinson disease are maintained after GDNF cessation. Neurology. 2013;81(11):1176–8.
19. Parain K, Murer MG, Yan Q, Faucheux B, Agid Y, Hirsch E, et al. Reduced expression of brain-derived neurotrophic factor protein in Parkinson's disease substantia nigra. Neuroreport. 1999;10(3):557–61.
20. Pringsheim T, Jette N, Frohlich A, Steeves TD. The prevalence of Parkinson's disease: a systematic review and meta-analysis. Mov Disord. 2014;29(13):1583–90.
21. Hirsch L, Jette N, Frohlich A, Steeves T, Pringsheim T. The incidence of Parkinson's disease: a systematic review and meta-analysis. Neuroepidemiology. 2016;46(4):292–300.
22. Van Den Eeden SK. Incidence of Parkinson's disease: variation by age, gender, and race/ethnicity. Am J Epidemiol. 2003;157(11):1015–22.
23. Rodriguez-Violante M, Villar-Velarde A, Valencia-Ramos C, Cervantes-Arriaga A. Características epidemiológicas de pacientes con enfermedad de Parkinson Arch Neuropsiquiatr (Mex) INNN. Arch Neuropsiquiatr. 2011;64:64–4.
24. Pagano P, Ferrara N, Brooks DJ, Pavese N. Age at onset and Parkinson disease phenotype. Neurology. 2016;86(5):1400–7.
25. Song Y, Gu Z, Lan P. Gender differences on motor and non-motor symptoms of de novo patients with early Parkinson disease. Neurol Sci. 2014;35(12):1991–6.
26. Yamawaki M, Kusumi M, Kowa H, Nakashima K. Changes in prevalence and incidence of Parkinson's disease in Japan during a quarter of a century. Neuroepidemiology. 2009;32(4):263–9.
27. Gillies GE, Pienaar IS, Vohra S, Qamhawi Z. Sex differences in Parkinson's disease. J Neurol Neurosurg Psychiatry. 2010;81(13):1176–8.
28. Smith KM, Dahodwala N. Sex differences in Parkinson’s disease and other movement disorders. Exp Neurol. 2014;259:44–56.
29. Le Roith D. Seminars in medicine of the Beth Israel Deaconess Medical Center. Insulin-like growth factors. N Engl J Med. 1997;336(9):633–40.
30. Mcmahon T, Van Zijl PCM, Gilad AA. NIH Public Access.
31. González C, Díaz F, Alonso A. Neuroprotective effects of estrogens: a systematic review. Front Neuroendocrinol. 2014;35(3):370–84.
32. Pollock LA, Ianov L, Foster TC. Estrogen receptors, the hippocampus, and ageing. Front Neuroendocrinol. 2016;2016(43):44–59.
33. Reinhardt RR, Bonyd CA, Lee WH. Patterns of insulin-like growth factor I and II receptor gene expression in the brain. Functional implications. Ann N Y Acad Sci. 1993;692:315–28.
34. Bach MA, Shen-Orr Z, Lowe W, Roberts CT, LeRoith D. Insulin-like growth factor I mRNA levels are developmentally regulated in specific regions of the rat brain. Brain Res Mol Brain Res. 1991;10(1):43–8.
35. Tong M, Dong M, de la Monte SM. Brain insulin-like growth factor and amyloid-β peptide: potential role of manganese neurotoxicity. J Alzheimers Dis. 2009;16(3):585–99.
36. Le Roith D. Seminars in medicine of the Beth Israel Deaconess Medical Center. Insulin-like growth factor-I (IGF-1) deficiency. J Transl Med. 2012;513(1):20–4.
37. Sonntag WE, Ramsey M, Carter CS. Growth hormone and insulin-like growth factor-1 (IGF-1) and their influence on cognitive aging. Ageing Res Rev. 2005;4(2):195–212.
38. Adem A, Jossan SS, D'Argy R, Gillberg PG, Nordberg A, Winblad B, et al. Insulin-like growth factor 1 (IGF-1) receptors in the human brain: quantitative autoradiographic localization. Brain Res. 1989;503(2):299–303.
39. Lewitt MS, Boyd GW. The role of insulin-like growth factors and insulin-like growth factor-binding proteins in the nervous system. Biochem Insights. 2019;17(12):11786241984217. https://doi.org/10.1177/117862419842176.
40. Tong M, Dong M, de la Monte SM. Brain insulin-like growth factor and neurotrophic resistance in Parkinson's disease and dementia with Lewy bodies: potential role of manganese neurotoxicity. J Alzheimers Dis. 2009;16(3):585–99.
41. Yang L, Wang H, Liu L, Xie A. The role of insulin/IGF-1/PI3K/Akt/GSK3β signaling in Parkinson's disease dementia. Front Neurol. 2018;9:20. https://doi.org/10.3389/fnnins.2018.00073/full.
66. Morris JK, Zhang H, Guptha AA, Bomhoff GL, Stanford JA, Geiger PC. Measures of striatal insulin resistance in a 6-hydroxydopamine model of Parkinson's disease. Brain Res. 2008;1240:185–95.

67. Talbot K, Wang H-Y, Kazi H, Han L-Y, Bakshi KP, Stucky A, et al. Demonstrated brain insulin resistance in Alzheimer's disease patients is associated with IGFI resistance. IRS-1 dysregulation, and cognitive decline. J Clin Investig. 2012;122(4):1316–38.

68. Fernandez AM, Torres-Alemán I. The many faces of insulin-like peptide signalling in the brain. Nat Rev Neurosci. 2012;13(4):225–39.

69. Backström M, Hall K, Safari V. Somatomedin levels in cerebrospinal fluid from adults with primary disorders. Acta Endocrinol. 1984;107(2):171–8.

70. Carro E, Trejo JL, Gomez-Isla T, LeRoith D, Torres-Alemán I. Serum insulin-like growth factor I regulates brain amyloid-beta levels. Nat Med. 2002;8:1390–7.

71. Kalmar B, Greensmith L. Induction of heat shock proteins for protection against oxidative stress. Adv Drug Deliv Rev. 2009;61(4):310–8.

72. Garcia-Fernandez M, Sierra E, Puche JE, Guerra L, Castilla-Cortazar I. Liver mitochondrial dysfunction is reverted by insulin-like growth factor II (IGF-II) in aging rats. J Transl Med. 2011;9:123.

73. Castilla-Cortazar I, Garcia-Fernandez M, Delgado G, Puche JE, Sierra I, Barhoum R, et al. Hepatoprotection and neuroprotection induced by low doses of IGF-II in aging rats. J Transl Med. 2011;9:103.

74. Duffy KR, Partridge WM, Rosenfield RG. Human brain-blood barrier insulin-like growth factor I receptor. Metabolism. 1986;37(2):136–40.

75. Dyer AH, Vahdatpour C, Sanfelice A, Tropea D. The role of Insulin-Like Growth Factor 1 (IGF-1) in brain development, maturation and neuroplasticity. Neuroscience. 2016;325:89–99.

76. Joseph D’Ecole A, Ye P. Minireview: expanding the mind: insulin-like growth factor I and brain development. Endocrinol. 2008;149(12):5958–62.

77. Ye P, Li L, Richards RG, DiAugustine RP, D’Ecole AJ. Myelination is altered in insulin-like growth factor-I null mutant mice. J Neurosci. 2002;22(14):6041–51.

78. Popken GJ, Hodge RD, Ye P, Zhang J, O’Kusky JR, et al. In vivo effects of insulin-like growth factor-I (IGF-I) on prenatal and early postnatal development of the central nervous system. Eur J Neurosci. 2004;19(8):2056–68.

79. Ye P, D’Ecole AJ. Insulin-like growth factor actions during development of neural stem cells and progenitors in the central nervous system. J Neurosci Res. 2006;83(1):1–6.

80. O’Kusky JR, Ye P. Neurodevelopmental effects of insulin-like growth factor signaling. Front Neuroendocrinol. 2012;33(3):250–51.

81. Hsu T-J, Khan AA, Pali S, Mustafa Z, Abdullah JM, Jaafar H, et al. Selective induction of neocortical GABAergic neurons by the PDK1-activating insulin-like growth factor 1 (IGF1) and its active peptide (1–3)IGF1 enhance cellular mechanisms. Neurosci Lett. 2012;520(1):51–6.

82. Trejo JL, Llorens-Martín M, Torres-Alemán I, Trejo JL. Mechanisms mediating brain plasticity: IGF-I and adult hippocampal neurogenesis. Neuroscientist. 2009;15(2):134–48.

83. Tropea D, Kreiman G, Lyckman A, Mukherjee S, Yu H, Horng S, et al. Gene expression changes and molecular pathways mediating activity-dependent plasticity in visual cortex. Nat Neurosci. 2006;9(5):660–8.

84. Torres-Alemán I. Insulin-like growth factors as mediators of functional plasticity in the adult brain. Horm Metab Res. 1999;31(2–3):114–9.

85. Jacobo SMP, Kazaiauxas A. Insulin-like growth factor 1 (IGF-1) stabilizes nascent blood vessels. J Biol Chem. 2015;290(10):6349–60.

86. Bonfanti L. PSA-NCAM in mammalian structural plasticity and neurogenesis. Prog Neurobiol. 2006;80(3):129–64.

87. Monzo HJ, Park TH, Dieriks BV, Jansson D, Faull RLM, Dragunow M, et al. Insulin and IGF-1 modulate turnover of polysynaptized neural cell adhesion molecule (PSA-NCAM) in a process involving specific extracellular matrix components. J Neurochem. 2013;126(6):758–70.

88. McCreath RH, McCrea K, Zunic S, Dantzer R, Boussard SR, Johnson RW, et al. Insulin-like growth factor-I enhances the biological activity of brain-derived neurotrophic factor on cerebrocortical neurons. J Neuroimmunol. 2006;179(1–2):186–90.

89. Mednezed P, Azcoitia J, Garcia-Segura LM. Interdependence of oestrogen and insulin-like growth factor-I in the brain: potential for analysing neuroprotective mechanisms. J Endocrinol. 2005;185(1):11–7.

90. Cardona-Gómez GP, Trejo JL, Fernandez AM, Garcia-Segura LM. Estrogen receptors and insulin-like growth factor-I receptors mediate estrogen-dependent synaptic plasticity. Neuropathol Exp Neurobiol. 2000;11(8)1735–8.

91. Trejo JL, Carro E, Lopez-Lopez C, Torres-Alemán I. Role of serum insulin-like growth factor I in mammalian brain aging. Growth Horm IGF Res. 2004;14(Suppl A):S39–43.

92. Svensson J, Diez M, Engel J, Wass C, Twenst A, Jansson J-O, et al. Endocrine, liver-derived IGF-I is of importance for spatial learning and memory in old mice. J Endocrinol. 2006;189(3):617–27.

93. Lupien SB, Bluhm EJ, Ishii DN. Systemic insulin-like growth factor-I administration prevents cognitive impairment in diabetic rats, and brain IGF regulates learning/memory in normal adult rats. J Neurosci Res. 2003;74(4):512–23.

94. Hoshaw BA, Malberg JE, Lucki I. Central administration of IGF-I and BDNF leads to long-lasting antidepressant-like effects. Brain Res. 2005;1033(1–2):204–8.

95. Castro-Alamancos MA, Torres-Alemán I. Learning of the conditioned eye-blink response is impaired by an antisense insulin-like growth factor-I oligonucleotide. Proc Natl Acad Sci USA. 1994;91(21):10203–7.

96. Bluthé R-M, Kelley KW, Dantzer R. Effects of insulin-like growth factor-I on cytokine-induced sickness behavior in mice. Brain Behav Immun. 2006;20(1):57–63.

97. Ang LC, Braumick B, Jurlink BH. Neurite promoting activity of insulin, insulin-like growth factor I and nerve growth factor on spinal motoneurones. Brain Res. 2003;974(1):83–8.

98. Pomerance M, Gavaret JM, Breton M, Pierre M. Effects of growth factors on spinal motoneurons and plasminogen activators by insulin-like growth factor-I and epidermal growth factor. Brain Res. 1994;63(6):206–13.
112. Bradshaw SL, Han VK. Hormonal regulation of astroglial insulin-like growth factor (IGF)-binding protein gene expression by IGFs and insulin. Endocrinology. 1993;133(4):1767–77.

113. Aberg ND, Blomstrand F, Aberg MA, Bjerklund U,Carlsson B,Carlsson-Skvrnut C, et al. Insulin-like growth factor-I increases astrocyte intercellular gap junctional communication and connexin43 expression in vitro. J Neurosci Res. 2003;74(1):12–22.

114. Suzuki K, Ikegaya Y, Matsuura S, Kanai Y, Endou H, Matsuki N. Transient upregulation of the glial glutamate transporter GLAST in response to fibroblast growth factor, insulin-like growth factor and epidemial growth factor in cultured astrocytes. J Cell Sci. 2001;114(Pt 20):3717–25.

115. Sonnewald U, Wang AY, Petersen SB,Westergaard N, Schousboe A, Erikson R, et al. 13 C NMR study of IGF-I and insulin-effects on mitochondrial function in cultured brain cells. NeuroReport. 1995;6(6):880–7.

116. Dingen R, Hamprecht B. Glucose, insulin, and insulin-like growth factor I regulate the glycogen content of astroglia-rich primary cultures. J Neurochem. 1992;58(2):511–7.

117. Masten BA, Werner H, Roberts CT, LeRoith D, Raizada MK. Developmental regulation of insulin-like growth factor-I-stimulated glucose transporter in rat brain astrocytes. Endocrinology. 1991;128(5):2548–57.

118. Spina Purrello V, Cormaci G, Denaro L, Reale S, Costa A, Lalicata C, et al. Effect of growth factors on nuclear and mitochondrial ADP-ribosylation processes during astroglial cell development and aging in culture. Mech Ageing Dev. 2002;123(5):111–20.

119. Matsuda T, Murata Y, Kawamura N, Hayashi M, Tamada K, Takuma K, et al. Selective induction of alpha 1 isoform of (Na+ + K+) -ATPase by insulin/insulin-like growth factor-I in cultured rat astrocytes. Arch Biochem Biophys. 1993;307(1):175–82.

120. Ridet JL, Berry M, Hill DJ, Logan A. Spatial and temporal changes of the glial cytoplasmic astroglial function in cultured brain cells. NeuroReport. 1995;6(7):523–6.

121. Walter HJ, Berry M, Hill DJ, Logan A. Spatial and temporal changes of the glial cytoplasmic astroglial function in cultured brain cells. NeuroReport. 1995;6(7):523–6.

122. Antoniades HN, Galanopoulos T, Neville-Golden J, Maxwell M. Expression of insulin-like growth factor-I and the insulin receptor genes in human primary astrocytomas and meningomas; in vivo studies using situs hybridization and immunocytochemistry. Int J Cancer. 1995;59(2):215–22.

123. Lara-Diaz VJ, Castilla-Cortazar I, Martin-Estal I, Garcia-Magariño M, Aguirre GA, Puche JE, et al. IGF-I modulates gene expression of proteins involved in inflammation, cytoskeleton, and liver architecture. J Physiol Biochem. 2017;73:245–58.

124. Morales-Garza LA, Puche JE, de la Garza RG, Muñoz-Morón Ú, Castilla-Cortázar I. Experimental approach to IGF-1 therapy in acute liver damage: acute injury by carbon tetrachloride in controls and mice with partial IGF-1 deficiency. J Hepatol. 1998;28(1):122–31.

125. Conchillo M, Castilla-Cortázar I, Puche JE, Delgado G, Diaz-Sánchez M, et al. Effects of IGF-1 treatment on osteopetrosis in rats with advanced liver cirrhosis: improvement by Insulin-Like Growth Factor I. BMC Gastroenterol. 2004;4:12.

126. García-Fernández M, Castilla-Cortazar I, Díaz-Sánchez M, Navarro I, Pérez R, García-Fernández M, Díaz-Sánchez M, Puche JE, Delgado G, Conchillo M, et al. Mitochondrial protection by low doses of insulin-like growth factor I. Gastroenterology. 1997;113(4):1180–7.

127. Cemborain A, Castilla-Cortázar I, García M, Quirós J, del Olmo J, Puche JE, García-Magariño M, et al. Antioxidant effects of insulin-like growth factor-I (IGF-I) in rats with advanced liver cirrhosis. BMC Gastroenterol. 2005;5:20.

128. Castilla-Cortázar I, Aliaga-Montilla MA, Salvador J, García M, Picardi A, et al. Neurotrophic factors and their receptors are altered in the cellular and molecular aspects of the insulin-like growth factor-I receptor. In: Endocrine system: a comprehensive guide to endocrine function, and hypogonadism in rats with experimentally induced cirrhosis. J Hepatol. 1998;28(1):122–31.

129. Cemborain A, Castilla-Cortazar I, García M, Quirós J, del Olmo J, Puche JE, García-Magariño M, et al. Antioxidant effects of insulin-like growth factor-I (IGF-I) in rats with advanced liver cirrhosis. J Hepatol. 1998;28(1):122–31.

130. Castilla-Cortázar I, Aliaga-Montilla MA, Salvador J, García M, Picardi A, et al. Neurotrophic factors and their receptors are altered in the cellular and molecular aspects of the insulin-like growth factor-I receptor. In: Endocrine system: a comprehensive guide to endocrine function, and hypogonadism in rats with experimentally induced cirrhosis. J Hepatol. 1998;28(1):122–31.

131. Cemborain A, Castilla-Cortazar I, García M, Quirós J, del Olmo J, Puche JE, García-Magariño M, et al. Antioxidant effects of insulin-like growth factor-I (IGF-I) in rats with advanced liver cirrhosis. J Hepatol. 1998;28(1):122–31.

132. Gubbi S, Quipildor GF, Barzilai N, Hoffman DM, Milman S. 40 years of IGF-1. J Mol Endocrinol. 2016;5(1):T171–85.
insulin-like growth factor-I in cirrhotic rats. Biochim Biophys Acta. 2001;1536(2–3):185–95.

153. Castilla-Cortazar I, Garcia M, Muguerza B, Quiroga J, Perez R, Sanford T, et al. Hepatoprotective effects of insulin-like growth factor-I in rats with carbon tetrachloride-induced cirrhosis. Gastroenterology. 1997;113(5):1682–91.

154. Conchillo M, de Knecht RJ, Payeras M, Quiroga J, Sangro B, Herrero J, et al. Insulin-like growth factor-I (IGF-I) replacement therapy increases albumin concentration in liver cirrhosis: results of a pilot randomized controlled clinical trial. J Hepatol. 2005;43(4):630–6.

155. Ceda GP, Dall’Aglio E, Maggio M, Lauretani F, Bandinelli S, Falzoi C, et al. Clinical implications of the reduced activity of the GH-IGF-I axis in older men. J Endocrinol Investig. 2005;28(11 Suppl Proceedings):96–100.

156. Sun LY, Al-Regaiey K, Masternak MM, Wang J, Bartke A. Local expression of GH and IGF-I in the hippocampus of GH-deficient long-lived mice. Neurobiol Aging. 2005;26(6):929–37.

157. Sonntag WE, Bennett C, Ingram R, Donahue A, Ingraham J, Chen H, et al. Human conditions of insulin-like growth factor-1 axis: relation to caloric restriction. FASEB J. 2005;19(7):2920–32.

158. Shimokawa I, Higami Y, Tsuchiya T, Otani H, Komatsu T, Chiba T, et al. Holzenberger M. The GH/IGF-I axis and longevity. Eur J Endocrinol. 2001;145(7):2920–32.

159. Sonntag WE, Lynch CD, Cooney PT, Hutchins PM. Decreases in life span extension by reduction of the growth hormone-insulin-like growth factor-1 axis: relation to caloric restriction. FASEB J. 2003;17(9):1108–9.

160. Holzenberger M, The GH/IGF-I axis and longevity. Eur J Endocrinol. 2004;151(Suppl):S23–7.

161. Sonntag WE, Bennett C, Ingram R, Donahue A, Ingraham J, Chen H, et al. Hormone growth and IGF-I modulate local cerebral glucose utilization and ATP levels in a model of adult-onset growth hormone deficiency. Am J Physiol Endocrinol Metab. 2006;291(3):E604–10.

162. Maggio M, Lauretani F, Ceda GP, Bandinelli S, Basaria S, Paolisso G, et al. Association of hormonal dysregulation with metabolic syndrome in older women: data from the InCHIANTI study. Am J Physiol Endocrinol Metab. 2007;292(1):E353–8.

163. Groban L, Pales NA, Bennett CDL, Carter CS, Chappell MCC, Kitzman DW, et al. Growth hormone replacement attenuates diastolic dysfunction and cardiac angiotensin II expression in senescent rats. J Gerontol A Biol Sci Med Sci. 2006;61(1):28–35.

164. Li Q, Wu S, Li S-Y, Lopez FL, Du M, Kajstura J, et al. Cardiac-specific overexpression of insulin-like growth factor 1 attenuates age-associated cardiac diastolic contractile dysfunction and protein damage. Am J Physiol Heart Circ Physiol. 2007;292(3):H398–403.

165. Puche JE, Castilla-Cortazar I, Human conditions of insulin-like growth factor-I (IGF-I) deficiency. J Transl Med. 2012;10(1):224.

166. Tollef-Egnell P, Flores-Morales A, Odobele J, Lundeberg J, Norstedt G. Differential cloning of growth hormone-regulated hepatic transcripts in the aged rat. Endocrinology. 2000;141(3):910–21.

167. Puche JE, Garcia-Fernandez M, Munetani J, Rioja J, Gonzalez-Baron S, Cortazar IC. Low doses of insulin-like growth factor-I induce mitochondrial protection in aging rats. Endocrinology. 2008;149:2620–7.

168. Garcia-Fernandez M, Delgado G, Puche JE, Gonzalez-Baron S, Cortazar IC. Low doses of insulin-like growth factor-I improve insulin resistance, lipid metabolism, and oxidative damage in aging rats. Endocrinology. 2008;149:2432–43.

169. Pellecchia MT, Pivonello R, Salvatore E, Faggiano A, Barone P, et al. Differential cloning of growth hormone-phosphoinositide 3-kinase signaling. Neurobiol Aging. 2004;25:1023–32.

170. Volpi R, Caffarra P, Scaglioni A, Boni S, Saginario A, Chiodera P, et al. Direct interactions of alpha-synuclein with insulin-like growth factor. J Proteomics. 2009;73(1):10–29.

171. Kao S-Y, Rescue of alpha-synuclein cytotoxicity by insulin-like growth factors. Biochem Biophys Res Commun. 2009;385(3):434–8.

172. Kim W, Lee Y, McKenna ND, Yi M, Simunovic F, Wang Y, et al. 3-methyladenine contributes to Parkinson's disease by dysregulating the insulin-like growth factor/phosphoinositide 3-kinase signaling. Neurobiol Aging. 2014;35(7):1712–21.

173. Liu D-H, He Y-C, Quinn TJ, Liu J. Serum insulin-like growth factor-I in patients with de novo, drug naive Parkinson's disease: a meta-analysis. PLoS ONE. 2015;10(12):e0144753.

174. Singleton AB, Farrer M, Johnsson J, Singleton A, Hague S, Kachergus J, et al. alpha-Synuclein locus triplication causes Parkinson's disease. Science. 2003;302(5646):841.

175. Kumari U, Tan EK. LRRK2 in Parkinson's disease: genetic and clinical studies from patients. FEBS J. 2009;276(22):6453–65.

176. Licker V, Kovári É, Hochstätter DF, Burkhard PR. Proteomics in human Parkinson's disease research. J Proteomics. 2009;73(1):10–29.

177. Spillantini MG, Schmidt ML, Lee VM, Trojanowski JQ, Jakes R, Goedert M. Alpha-synuclein in Lewy bodies. Nature. 1997;388(6645):839–40.

178. Chung J-Y, Lee S-J, Lee S-H, Jung YS, Ha N-C, Seol W, et al. LRRK2 in Parkinson's disease: genetic and clinical studies from patients. FEBS J. 2009;276(22):6453–65.

179. Chung J-Y, Lee S-J, Lee S-H, Jung YS, Ha N-C, Seol W, et al. Direct interaction of alpha-synuclein with AKT regulates IGF-1 signaling: implication of Parkinson disease. Neurosignals. 2011;19(2):86–96.

180. Caffarra P, Scaglioni A, Boni S, Saginario A, Chiodera P, et al. Direct interactions of alpha-synuclein with insulin-like growth factors. Biochem Biophys Res Commun. 2009;385(3):434–8.

181. Kim W, Lee Y, McKenna ND, Yi M, Simunovic F, Wang Y, et al. 3-methyladenine contributes to Parkinson's disease by dysregulating the insulin-like growth factor/phosphoinositide 3-kinase signaling. Neurobiol Aging. 2014;35(7):1712–21.

182. Duillo AG, Montano J-P. Obesity in Parkinson's disease patients on electrotherapy: collateral damage, adiposity rebound or secular trends? Br J Nutr. 2005;93(4):417–9.

183. Chen H, Zhang SM, Schwarzschild MA, Hernán MA, Willett WC, Ascherio A. Obesity and the risk of Parkinson's disease. Am J Epidemiol. 2004;159(6):547–55.

184. Zaganas I, Kapetanaki S, Mastorodemos V, Kanavouras K, Colosio C, Di Monte DA. The environment and Parkinson's disease: is the nigrostriatal system preferentially targeted by neurotoxins? Lancet Neurol. 2003;2(9):531–8.
195. Aguirre GA, González-Guerra JL, Espinosa L, Castilla-Cortazar I. Insulin-like growth factor 1 in the cardiovascular system. In: Rev Physiol Biochem Pharmacol. 2018.
196. Zuo L, Motherwell MS. The impact of reactive oxygen species and genetic mitochondrial mutations in Parkinson’s disease. Gene. 2013;532(1):18–23.

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