Leptospira interrogans endostatin-like outer membrane proteins bind host fibronectin, laminin and regulators of complement.

Permalink
https://escholarship.org/uc/item/79q308r5

Journal
PloS one, 2(11)

ISSN
1932-6203

Authors
Stevenson, Brian
Choy, Henry A
Pinne, Marija
et al.

Publication Date
2007-11-14

DOI
10.1371/journal.pone.0001188

Peer reviewed
Leptospira interrogans Endostatin-Like Outer Membrane Proteins Bind Host Fibronectin, Laminin and Regulators of Complement

Brian Stevenson1,*, Henry A. Choy2,3, Marija Pinne2,3, Matthew L. Rotondi2,4, M. Clarke Miller4,5, Edward DeMoll4, Peter Kraiczy5, Anne E. Cooley1, Trevor P. Creamer5, Marc A. Suchard7, Catherine A. Brissette1, Ashutosh Verma1,8, David A. Haake3,9

1 Department of Microbiology, Immunology, and Molecular Genetics, University of Kentucky College of Medicine, Lexington, Kentucky, United States of America, 2Division of Infectious Diseases, Veterans Affairs Greater Los Angeles Health Care System, Los Angeles, California, United States of America, 3Department of Medicine, University of California Los Angeles School of Medicine, Los Angeles, California, United States of America, 4Department of Chemistry, University of Kentucky, Lexington, Kentucky, United States of America, 5Institute of Medical Microbiology and Infection Control, University Hospital of Frankfurt, Frankfurt am Main, Germany, 6Department of Molecular and Cellular Biochemistry, University of Kentucky College of Medicine, Lexington, Kentucky, United States of America, 7Department of Biomathematics, University of California Los Angeles School of Medicine, Los Angeles, California, United States of America, 8Department of Veterinary Sciences, University of Kentucky, Lexington, Kentucky, United States of America

The pathogenic spirochete Leptospira interrogans disseminates throughout its hosts via the bloodstream, then invades and colonizes a variety of host tissues. Infectious leptospires are resistant to killing by their hosts’ alternative pathway of complement-mediated killing, and interact with various host extracellular matrix (ECM) components. The LenA outer surface protein (formerly called LfhA and Lsa24) was previously shown to bind the host ECM component laminin and the complement regulators factor H and factor H-related protein-1. We now demonstrate that infectious L. interrogans contain five additional paralogs of lenA, which we designated lenB, lenC, lenD, lenE and lenF. All six genes encode domains predicted to bear structural and functional similarities with mammalian endostatins. Sequence analyses of genes from seven infectious L. interrogans serovars indicated development of sequence diversity through recombination and intragenic duplication. LenB was found to bind human factor H, and all of the newly-described Len proteins bound laminin. In addition, LenB, LenC, LenD, LenE and LenF all exhibited affinities for fibronectin, a distinct host extracellular matrix protein. These characteristics suggest that Len proteins together facilitate invasion and colonization of host tissues, and protect against host immune responses during mammalian infection.

Citation: Stevenson B, Choy HA, Pinne M, Rotondi ML, Miller MC, et al (2007) Leptospira interrogans Endostatin-Like Outer Membrane Proteins Bind Host Fibronectin, Laminin and Regulators of Complement. PLoS ONE 2(11): e1188. doi:10.1371/journal.pone.0001188

INTRODUCTION

Leptospirosis is a zoonotic disease of humans caused by the spirochete Leptospira interrogans and several other members of that genus [1]. The prevalence of leptospirosis in many parts of the world is due to chronic kidney infection of a wide variety of domestic, peridomestic and wild reservoir host mammals, including rodents, pigs, cattle, horses and dogs. Colonization of the renal tubules of carrier animals results in shedding of virulent leptospires in the urine. Leptospires persist in fresh water until infection of a new host occurs via the conjunctiva, breaks in the skin or by invasion of mucous membranes in the respiratory or digestive tract. A hallmark of leptospirosis infection is early and widespread hemoglobinuria dissemination manifested by fever, myalgia, conjunctivitis, meningitis, uveitis and/or jaundice. Between 5 and 10% of patients progress to the more dangerous, icteric phase of leptospirosis, which may lead to death due to acute renal failure, pulmonary hemorrhage, intracerebral hemorrhage, and multiorgan system failure [1]. Infectious Leptospira spp. are endemic in many tropical and temperate areas of the world, presenting health threats to inhabitants of both rural and urban areas, as well as military personnel, aid workers, and tourists.

Presumably as mechanisms that facilitate tissue invasion and colonization, pathogenic leptospires interact with a variety of host extracellular matrix (ECM) components, and some of the bacterial adhesins have been identified [reference [2–6] and this work]. L. interrogans is highly resistant to the alternative pathway of host complement activation [7–12], a feature that is associated with binding of factor H to the bacterial outer membrane, degradation of C3b and C3 convertase, and inhibition of membrane-attack complex formation [11,12]. The capacities to bind host ECM and factor H are associated with virulence, as those traits are held by infectious Leptospira species but are lacking from non-infectious species of Leptospira [2,11,12].

A previous study which screened an L. interrogans expression library for proteins capable of binding host factor H identified an approximately 25 kDa outer membrane protein, designated LfhA (leptospiral factor H-binding protein) [12]. LfhA was also found...
to bind human factor H-related protein 1 (FHR-1), a distinct protein whose carboxy-terminus is very similar to that of factor H [12,13]. LfhA did not bind human factor H-like protein 1 (FHL-1), a splice variant of the same gene as factor H which consists of the first seven short consensus repeat units of factor H [12,13].

A subsequent study by Barbosa and colleagues identified an L. interrogans outer membrane laminin-binding protein, which they designated Lsa24 [leptospiral surface adhesin 24kDa] [2]. Intriguingly, Lsa24 and LfhA are the same protein, indicating that this single protein is able to bind host factor H, FHR-1 and laminin.

As described in the present report, L. interrogans carries 5 additional genes homologous to LfhA/Lsa24. Modeling of the predicted proteins encoded by these six paralogous genes indicated substantial similarities to mammalian endostatins. In order to unify the nomenclatures of these leptospiral genes, we renamed LfhA/Lsa24 as “lenB” (leptospiral endostatin-like protein A) and named the newly-described paralogy lenA, lenC, lenD, lenE and lenF. At the present time, there are no tools available to specifically mutate L. interrogans, so it is impossible to study protein function in situ by deletion and complementation mutagenesis. That being so, in order to better understand these genes and their roles in leptospiral pathogenesis, we characterized relationships of len genes among several distinct L. interrogans strains, performed biophysical characterization of the proteins, and examined interactions between Len and host ECM and complement-regulatory proteins.

RESULTS
Paralogous L. interrogans len genes
In a previous study [12], members of our laboratories demonstrated that the lenA (LfhA) genes of serovar Pomona strain JEN4 and serovar Lai strain 56601 encode a factor H/FHR-1 binding protein. Sequence analyses of additional L. interrogans strains from five other serovars indicated that all contain lenC loci. The lenC gene sequences of serovars Lai, Copenhageni, Grippotyphosa and Hardjo are all highly conserved and are predicted to encode membrane-associated lipoproteins (Fig. 1) [14,15]. The lenC loci of serovars Pomona, Bratislava and Canicola are nearly identical to those of the aforementioned serovars, but include mutations that would preclude their synthesis of intact lipoproteins.

The sequenced genomes of L. interrogans serovar Copenhageni strain Fiocruz L1-130 and serovar Lai strain 56601 both contain an additional locus with size and sequence similar to lenA, which we named lenB (Table 1, Fig. 1). All other L. interrogans strains we examined also contain a lenB locus, bordered by sequences similar to those of serovars Copenhageni and Lai. The lenB loci of serovars Bratislava and Hardjo each encode a protein with a lipoprotein leader polypeptide and a cleavage/lipidation sequence (Fig. 1) [14,15]. However, all other analyzed serovars lack the initial 19 codons of lenB, as well as a substantial portion of upstream DNA, and appear to be defective genes. Thus, only serovars Bratislava and Hardjo are predicted to produce membrane-associated LenB lipoproteins. The 5’ non-coding regions of the Bratislava and Hardjo lenB loci contain extensive homologies to the 5’ non-coding regions of lenC loci (Fig. 1D).

Four additional paralogues of lenA and lenB were identified in the genome sequences of Copenhageni Fiocruz L1-130 and Lai 56601, which we named lenC, lenD, lenE, and lenF (Table 1, Fig. 1). The latter four genes were sequenced in their entirety from five additional L. interrogans isolates. Each of the four genes in all seven strains was intact and predicted to encode a protein of approximately 50kDa. The nearly two-fold increase in size compared to LenA and LenB was accounted for by the presence of two motifs that each resemble LenA and LenB, which we refer to as “Len-motifs” (Fig. 1A). Sequence relatedness between paired Len-motifs of lenC, lenD, lenE, and lenF indicates that the motif duplications arose intragenically. In all four predicted proteins, the Len-motifs are separated by a proline-rich, 5–20 amino acid linker sequence (Fig. 1B and Fig. S1).

Phylogenetic analyses indicated that, in most cases, sequence differences between homologous len loci of the different L. interrogans serovars occurred after the evolutionary diversification of the loci themselves. As examples, all lenC genes are very closely related to each other, as are the identified alleles of lenB, lenD and lenE (Fig. 1C). However, the lenC gene sequences of serovars Bratislava, Copenhageni, Grippotyphosa, Hardjo and Lai are very Bratislava to each other, but are divergent from the lenC genes of serovars Canicola and Pomona, which are also nearly identical to each other (Fig. 1B and C). In the below-described studies of Len functions, we refer to the former group as lenC-1, and the latter as lenC-2. The lenF genes also fell into two distinct clades, designated lenF-1 and lenF-2 (Fig. 1B and C). Computational analyses indicated a stronger likelihood of a recombination event having given rise to the two distinct lenF groups, as opposed to a shared history, with a log10 Bayes factor of approximately 8 (a log10 Bayes factor >2 is considered to be strong evidence [16]). The lenF-1 group of serovars Bratislava, Canicola, Lai and Pomona apparently arose from a recombination event in which both len-motifs of a primordial lenF gene were replaced by motifs of the lenC lineage. The other three examined serovars, Copenhageni, Grippotyphosa and Hardjo, form the distinct lenF-2 group, which does not exhibit evidence of such a recombination event. The lenF-1 recombination event appears to have involved only the len-motifs of that group: the sequences of the lenF-1 and lenF-2 groups are virtually identical in both the 5’ coding and noncoding regions, and the 3’ noncoding region, and those regions differ in sequence composition from analogous regions of the purported lenC donor (Fig. 1B and data not shown). These data indicate that proteins of the LenF-1 group are likely to be mosaics of two parental lineages: lenF-1-derived in the amino-terminal region of the protein, including the signal peptide, and lenC-derived in the two Len-motifs.

Comparisons of len gene sequences also revealed evidence of DNA transfer between leptospires. For example, serovars Pomona and Lai contain very similar lenF-1 genes, but Pomona contains a lenF-2 locus while Lai contains a distinct, lenC-1 variant of that gene (Fig. 1C). Alignments of non-coding regions preceding lenC, lenD, lenE, and lenF loci did not reveal any obvious sequence similarities with each other or with lenA or lenB (data not shown). That result suggests that the lenC, lenD, lenE, and lenF operons might be regulated independently of each other and lenA/lenB, since each operon’s 5’ non-coding DNAs could bind distinct regulatory factors. Supporting that hypothesis, cultured L. interrogans serovar Copenhageni Fiocruz L1-130 produced detectable levels of LenD protein, but not of LenC, LenE, or LenF (Fig. 2 and data not shown), and len gene transcript levels are affected differently when L. interrogans is cultivated under varying conditions [17,18].

Southern blot analyses indicated that lenA genes are carried by all examined species of pathogenic leptospires, but absent from non-infectious Leptospira species [12]. Further evidence of widespread maintenance of len genes among infectious Leptospira species was provided by immunoblot analyses. Polyclonal rabbit antisera raised against recombinant LenD recognized LenD but none of the other L. interrogans Len paralogs (Fig. 2A and data not shown). That antisera identified an approximately 50kDa protein in whole-cell lysates of all examined pathogenic Leptospira species, including strains of L. kirschneri, L. noguchii, L. santarosai, L. borgpeterseni, and L. weilii (Fig. 2A). No such protein was detected in the non-pathogenic species L. biflexa.
Figure 1. Relationships between Len proteins and genes. (A) Schematic of Len proteins, with individual Len-motifs indicated by grey rectangles. LenC, LenD, LenE and LenF each consist of 2 Len-motifs, bridged by proline-rich linkers. (B) Alignment of predicted amino acid sequences of representative proteins: serovar Lai LenA (LenA, Lai), serovar Bratislava LenB (LenB, Brat), serovar Bratislava LenC-1 (LenC, Brat), serovar Canicola LenC-2 (LenC, Can), serovar Pomona LenD (LenD, Pom), serovar Grippotyphosa LenE (LenE, Gripp), serovar Pomona LenF-1 (LenF, Pom), and serovar Hardjo LenF-1 (LenF, Har). Sequences of the proteins possessing two Len-motifs (LenC, LenD, LenE and LenF) were divided in the middle, after the well-conserved internal CVEQ sequence, to permit alignment of each Len-motif, and the amino- and carboxy-terminal Len-motifs are indicated by “-N” and “-C”, respectively. An alignment of these same proteins, undivided, is presented in Figure S1. Identical amino acids found in the majority of proteins are boxed and shaded. Cysteine residues that may serve as amino-terminal lipidation sites are circled. (C) Unrooted phylogenetic tree of predicted amino acid sequences of each identified Len protein. Bootstrap values of each major node are indicated. (D) Alignment of sequences located 5’ of lenA and lenB genes. Identical nucleotides are boxed and shaded.

doi:10.1371/journal.pone.0001188.g001
Table 1. ORF numbers of len genes contained in completed genomes of L. interrogans.

| Gene name | L. interrogans serovar Lai strain 56601 (GenBank/ TIGR) | L. interrogans serovar Copenhageni strain Fiocruz L1-130 |
|-----------|------------------------------------------------------|-----------------------------------------------------------|
| lenA      | LA0695/LA0695                                       | LIC12906                                                  |
| lenB      | LA3103/LA3102                                       | LIC10997                                                  |
| lenC      | LA0563/LA0563                                       | LIC13006                                                  |
| lenD      | LA1433/LA1433                                       | LIC12315                                                  |
| lenE      | LA4324/LA4323                                       | LIC13457                                                  |
| lenF      | LA4073/LA4072                                       | LIC13248                                                  |

*GenBank and TIGR assigned different identifying numbers to many ORFs of L. interrogans serovar Lai strain 56601. doi:10.1371/journal.pone.0001188.t001

Cellular localization and biophysical characterization of Len proteins

LenA is an outer membrane protein [2,12]. All Len proteins bear at least some hallmarks of being lipoproteins, with LenA, LenB, LenC, LenD and LenF having high probabilities of being so according to analyses using the spirochete-specific lipoprotein algorithm SpLip [14,15]. Using the above-described anti-LenD antiserum, cellular localization of that protein was assessed by Triton X-114 detergent solubilization and phase partitioning of live leptospires. The reliability of this technique has been validated by comparisons with results obtained by sucrose density gradient isolation of outer membrane vesicles [19]. This method yields three fractions: a detergent fraction consisting of outer membrane components, an aqueous fraction consisting of periplasmic contents, and a pellet consisting of inner membrane-associated components, cytoplasmic contents, and undisrupted cells [20]. LenD was found in the detergent fraction (Fig. 2B). Control analysis of the endolagellar sheath protein FlaA1 (which is attached to the inner membrane) confirmed that the detergent phase was not contaminated with inner membrane components (Fig. 2B). Presence of LenD in the pellet fraction is typical of leptospiral outer membrane proteins [21–23], and is indicative of incomplete Triton X-114 extraction. These results indicate that LenD is also an outer membrane protein.

The LenC, LenD, LenE and LenF proteins appear to be fused dimers of LenA/LenB-like proteins. This suggested to us that LenA and LenB might function as dimers, with each dimer being the equivalent of a single LenC, LenD, LenE or LenF protein. To explore that possibility, we examined whether or not recombinant LenA forms dimers. However, HPLC through a size-exclusion column yielded a single peak, with a calculated molecular mass of 24.2 kDa (data not shown), comparable to the calculated molecular mass of 24.4 kDa for the recombinant LenA monomer.

Circular dichroism (CD) analysis of recombinant LenA indicated that it is composed of 36% β-sheet, 23% turns, and 41% unstructured/other, with no evidence of any α-helices (Fig. 3A).

These data are in line with previous CD analyses indicating that LenA contains β-sheets [2]. Recombinant LenA was found to be a relatively stable protein, with a melting point of 53°C (+2°C) (Fig. 3B). Due to the limited solubility of the other recombinant Len paralogs, those proteins could not be analyzed by these biophysical techniques. PHYRE modeling of the predicted amino acid sequences of all Len proteins indicated moderate to strong probabilities (ranging between 25 and 60% estimated precision) for each Len-motif folding into a structure similar to those of mammalian endostatins, which are derived from the carboxy-termini of collagens XVIII and XV [24–26]. Among other functions, endostatins bind various ECM components, including laminin, [25,26]. We did not detect sequence or predicted structural similarities between Len proteins and any other known adhesins.

Functional characterization of Len paralogs

Through use of both affinity blot analyses and surface plasmon resonance, members of our laboratories previously demonstrated that LenA binds host complement factor H [12]. Ligand affinity blot analyses of LenB indicated that it, too, can bind human factor H (Fig. 4). However, none of the other Len proteins exhibited binding of factor H, as assessed by ligand affinity blot.

A previous study indicated that recombinant LenA (Lsa24) bound laminin [2]. We therefore examined whether or not the five newly-identified L. interrogans proteins shared that property. Each recombinant protein was solubilized in SDS buffer, subjected to SDS-PAGE and transferred to nitrocellulose membranes, then examined for ability to bind soluble laminin. All the recombinant Len proteins bound laminin, although LenC-1, LenC-2, LenE, LenF-1 and LenF-2 consistently yielded the strongest affinity blot signals, with LenD, LenB and then LenA exhibiting progressively weaker relative binding of laminin (Fig. 5). No laminin binding by control protein BSA was detected, even with extended film exposure times, demonstrating that binding of laminin by the Len proteins was specific.
Binding by soluble LenA and LenB was examined further by ELISA, using laminin immobilized in microtiter wells as the target ligand. LenB showed saturable binding ($K_d = 18.8\pm2.3$ nM, mean and standard deviation from four independent experiments), while LenA exhibited significantly weaker activity (Fig. 6A). The other recombinant Len proteins were insoluble in buffers compatible with ELISA, preventing their characterization using that technique.

Ligand affinity blot analyses showed adverse effects on laminin binding by all recombinant Len proteins when the ionic strength of the TBS-T buffer was increased (data not shown). The dependence on ionic interactions for laminin binding by LenA and LenB was examined by ELISA in the presence of increasing concentrations of NaCl. When compared to laminin-binding activity in buffer containing 150 mM NaCl, binding to LenA was reduced 49 and 60% by 200 mM and 500 mM NaCl, respectively, while binding to LenB was inhibited 40 and 79% by 200 mM and 500 mM NaCl, respectively (Fig. 6B).

Since laminin may interact with charged moieties through its "heparin-binding" sites [27], we examined the ability of heparin to compete with Len proteins for binding to laminin. One μM heparin reduced the ability of 1 μM LenA to bind laminin to 26% of the no-heparin control (Fig. 6C). One μM and 4 μM heparin inhibited the laminin-binding activity of 1 μM LenB to 75% and 55% of the no-heparin control (data not shown). A more pronounced inhibition, to only 25% of the no-heparin control, was observed when immobilized laminin was preincubated with 50 μM heparin followed by ELISA using 0.25 μM LenB in the presence of 50 μM heparin (Fig. 6C).

LenA was previously reported to bind weakly to fibronectin [2]. Ligand affinity blot analyses were therefore used to explore the abilities of the other Len proteins to bind soluble fibronectin. Strong binding signals were obtained for LenC-1, LenC-2, LenE, LenF-1, and LenF-2 (Fig. 7). Weaker signals from LenB and LenD were visible following prolonged film exposure times (see Fig. S2), suggesting lower avidities of those two proteins relative to the other five leptospiral proteins. By this technique, no signals were detected from LenA or from the negative control protein, BSA.

Fibronectin binding was further examined by ELISA using soluble recombinant LenA and LenB. LenB exhibited strong, saturable binding ($K_d = 106.2\pm8$ nM, from three experiments, of the no-heparin control (Fig. 6C). One μM and 4 μM heparin inhibited the laminin-binding activity of 1 μM LenB to 75% and 55% of the no-heparin control (data not shown). A more pronounced inhibition, to only 25% of the no-heparin control, was observed when immobilized laminin was preincubated with 50 μM heparin followed by ELISA using 0.25 μM LenB in the presence of 50 μM heparin (Fig. 6C).

LenA was previously reported to bind weakly to fibronectin [2]. Ligand affinity blot analyses were therefore used to explore the abilities of the other Len proteins to bind soluble fibronectin. Strong binding signals were obtained for LenC-1, LenC-2, LenE, LenF-1, and LenF-2 (Fig. 7). Weaker signals from LenB and LenD were visible following prolonged film exposure times (see Fig. S2), suggesting lower avidities of those two proteins relative to the other five leptospiral proteins. By this technique, no signals were detected from LenA or from the negative control protein, BSA.

Fibronectin binding was further examined by ELISA using soluble recombinant LenA and LenB. LenB exhibited strong, saturable binding ($K_d = 106.2\pm8$ nM, from three experiments,
Our analyses indicated relatively weak interactions between LenA and fibronectin, much as was previously described [2], and was not studied further. In contrast to laminin binding, heparin did not detectably affect LenB-fibronectin interaction (Fig. 8B). The binding activity of 0.25 mM LenB remained intact even in the presence of 50 mM heparin (data not shown). Assays with proteolytic fragments of fibronectin indicated that the N-terminal 70 kDa could account for all of the binding by LenB observed with intact fibronectin (Fig. 8C). This fragment comprises the type I repeat modules in fibronectin, which can be divided into two functional domains, the N-terminal domain (NTD; available as a 30kDa fragment) and the adjacent gelatin-binding domain (GBD; a 45 kDa proteolytic fragment) [28]. LenB interacted only with the 30 kDa NTD, with $K_d = 69.5 \pm 0.7$ nM (Fig. 8D).

$L. interrogans$ LigB, which binds both the NTD and the GBD [3], served as a positive control for GBD binding (data not shown).

**DISCUSSION**

$L. interrogans$ is an invasive extracellular pathogen, capable of disseminating through its hosts’ bloodstream to the kidneys and other organs, then colonizing those tissues. To do so requires that the bacterium evade complement-mediated killing and adhere to host cell surfaces and/or extracellular matrices, especially epithelial and endothelial basement membranes. We have extended upon earlier studies by demonstrating that infectious strains of $L. interrogans$ encode up to six distinct paralogous proteins with affinities for host fibronectin, laminin, factor H and/or FHR-1.

Differences in ligand binding were apparent among the Len paralogs: recombinant LenC, LenE and LenF exhibited appreciably greater affinities for laminin and fibronectin than did the other paralogs, LenB bound both those ligands more tightly than did LenA, and only LenA and LenB were demonstrated to bind host factor H. Such diversification of function is frequently observed in other organisms following gene duplication events [29].

The 5′ noncoding regions of the intact *lenA* and *lenB* genes showed extensive similarities, but that pair of loci and the *lenC*, *lenD*, *lenE* and *lenF* loci all differed considerably in their 5′ noncoding regions, suggesting that transcription of each is likely to be governed by a distinct regulatory mechanism. This study and previous array studies support of that hypothesis, with culture temperature having opposite effects upon transcription of *lenD* and *lenE* [17], osmolarity of culture medium significantly affecting only *lenD* [30], and only *LenD* being produced at detectable levels by *L. interrogans* serovar Copenhageni Fiocruz L1-130 when cultured in EMJH medium (this work). Many other leptospiral genes also exhibit differences in expression during mammalian infection, growth in the external environment, or when cultured in media of various compositions or temperatures [17,18,30–33]. Diversification of gene regulatory elements is also a frequent occurrence among paralogous gene families [34].
The roles of Len protein interactions with host proteins during infection processes remain to be determined. Laminins are important components of basement membranes, and fibronectin is a major component of ECM and serum, so binding those host proteins could facilitate interactions directly with ECM or serve as a bridge for adherence to cell surfaces [27,28,35]. Factor H is a regulator of the alternative pathway of complement activation, and adheres to eukaryotic cells through a variety of specific and non-specific receptors [13,36–39]. Binding of factor H by L. interrogans may therefore help protect the bacterium from killing by complement and/or serve as a bridge to facilitate adherence to host tissues [40]. The functions of FHR-1 and other factor H-related proteins are poorly understood, but appear to include control of complement activation, cellular adherence and other functions [13,41–45]. One feature held in common by all the identified host ligands of Len proteins is their affinities for heparin/heparan sulfate. In our studies, binding of both LenA and LenB to laminin were inhibited by heparin. LenB was determined to bind the amino-terminal domain of fibronectin, which contains heparin-binding sites. However, the inability of heparin to inhibit fibronectin-binding by LenB indicates that either fibronectin has a much higher affinity for LenB than for heparin or LenB contains one binding site for laminin and a second site that binds fibronectin. LenA binds both factor H and FHR-1, which contain highly similar heparin-binding regions, but LenA does not bind FHL-1, which lacks that heparin binding domain [12,13]. Many additional vertebrate proteins are known to bind heparin/heparan sulfate [46], and we are continuing to investigate whether such proteins may also be ligands for leptospiral Len proteins.

Gene duplication events probably gave rise to the six len paralogous genes and the paired len-motifs of the larger genes. A different type of recombination event led to the lenF-1 clade, with len-motifs of the lenC lineage replacing the homologous sequences of a primordial lenF gene. Intriguingly, the new lenF-1 lineage retained its ancestral lenF-like leader polypeptide and the flanking non-coding DNA sequences. Phylogenetic mosaicism has pre-

![Figure 8. ELISA results of LenA and LenB interactions with fibronectin or its proteolytic fragments.](image)

Table 2. Oligonucleotide primers used to amplify and clone len loci.

| Locus | Oligonucleotide name | Sequence relative to amplified locus | Sequence (5’ to 3’) |
|-------|----------------------|------------------------------------|-------------------|
| lenA  | LFH-2                | 5’                                 | TTA GTC GGT AAT AGA GTT TTA GCG |
|       | LFH-11               | 3’                                 | ACA ATC TTC CAA AGA TCC TAA CG |
| lenB  | 3102-1               | 5’                                 | TTT TTG ATG GCT GCA GAA ATG GGG |
|       | 3102-2               | 3’                                 | AAC TTA CTG TTC TAC ACA GAG TAG |
|       | 3102-4               | 3’                                 | TTC TAC TAT TAG CCT GAA AGC CTG |
| lenC  | 563-1                | 5’                                 | ATT AGC CCA AAC TAA CGT TAA TCG |
|       | 563-4                | 3’                                 | TTA CTC GTC ATT GAA AAA AGG TTA |
| lenD  | 1433-1               | 5’                                 | AAA TAT CTA AGT TAC CGT CGC TCG |
|       | 1433-2               | 3’                                 | TCA TCA TCT AGG CAA AGA ATT GCG |
| lenE  | 4323-1               | 5’                                 | ACA GAA GTC TAT CTT CAG AAT GAG |
|       | 4323-2               | 3’                                 | ATG AGA TTC AAA ATA ATC GAT CCG |
| lenF  | 4072-1               | 5’                                 | TTT CAG AAG GGC CCT TAA AGG ATT GAG |
|       | 4072-4               | 3’                                 | TTA CTC GTC ATT GAA AAA AGG TTA |
viously observed for another leptomastigote gene, which encodes the OmpL1 porin [47].

In conclusion, infectious *L. interrogans* encode six paralogous Len proteins that can interact with host laminin, fibronectin, and/or complement regulatory proteins. All six members of the Len family share structural and functional characteristics with mammalian endostatins, fragments of collagens XVIII and XV which bind laminin and other cell surface and tissue proteins. The apparently widespread distribution of *len* genes among virulent leptospires, their presence in multiple copies in *L. interrogans* genomes, and their absence from non-pathogenic *Leptospira* species, suggest that Len proteins perform important roles during pathogenesis and have provided a selective advantage during mammalian infection. Generation of *len* sequence diversity occurred early during leptomastigote evolution through genetic drift and recombination between *len* genes, prior to the development of distinct antigenic serovars. Diversity within the paralogous Len family appears to have resulted in functional differences, which may facilitate colonization of multiple niches and hosts. While site-specific genetic manipulation of *L. interrogans* is currently impossible, further in vitro studies on functions of the Len proteins and analyses of their expression during infection will continue to increase our understanding of the mechanisms of host tissue interactions and complement evasion employed by this pathogen.

**MATERIALS AND METHODS**

**Bacterial strains and culture conditions**

Infectious *L. interrogans* serovar Copenagheni strain Fiocruz L1-130 [48] was obtained from Albert Ko (Gonçalo Moniz Research Institute for Genomic Research (TIGR) Comprehensive Microbial Resource database (http://tigrblast.tigr.org/cmr-blast). Analyses of GenBank (http://www.ncbi.nlm.nih.gov/blast) and the Institute for Genomic Research (TIGR) Comprehensive Microbial Resource database (http://tigrblast.tigr.org/cmr-blast).

Genomic DNA from *L. interrogans* strains was isolated from 5 ml cultures, as previously described [52]. DNA segments that included each *len* locus were PCR amplified using *r*Taq DNA polymerase (Takara, Otsu, Japan) and 25 cycles of 94°C for 1 min, 55°C for 1 min and 72°C for 2 min. Oligonucleotide primers utilized (Table 2) were complementary to conserved sequences located 5’ and 3’ of the *len* genes of serovar Pomona strain, JEN4, serovar Canicola strain Fiocruz L1-130, and serovar Lai strain 56601 [12, 48, 49]. Amplicons were cloned into pCR2.1 (Invitrogen, Carlsbad, CA) and both strands sequenced completely (Davis Sequencing, Davis, CA).

DNA and protein alignments were performed using Clustal X [53]. Phylogenies were reconstructed both by the neighbor joining method under default settings of amino acid substitution using PAUP* version 4.0b10 software [54] and by Bayesian inference under the Hasegawa, Kishino, Yano nucleotide substitution model [55] and a relaxed molecular clock [56] using BEAST 1.4 software. In the former case, bootstrapping provided measures of clad credibility [57]. To estimate the Bayes factor in favor of a recombination event [58] giving rise to the *lenF-1* group over the alternative hypothesis in which *lenF-1* and *lenF-2* sharing a common history, an unconstrained tree topology model and a model in which *lenF-1* and *lenF-2* sequences are constrained to be monophyletic were fit. From each model, the marginal likelihood was estimated using the harmonic mean estimator [59] and the Bayes factor found by taking the ratio. If two hypotheses are equally likely a priori, then the Bayes factor is the relative posterior probabilities of the competing hypotheses; a log10 Bayes factor >2 is generally taken as strong evidence [16].

The spirochete-specific lipoprotein algorithm SpLip [15] was used to determine the probabilities that each *len* gene encodes a lipoprotein. This algorithm is a hybrid weight matrix approach using 28 experimentally verified spirochetal lipoproteins in the training set. All lipoproteins contain a hydrophobic amino-terminal leader polypeptide, followed by variable 3–4 amino acid “lipobox”, then a cysteine residue [14, 60]. During processing of the pre-protein to the mature lipoprotein, the leader polypeptide is removed and lipid moieties attached to the cysteine. The ~1 position relative to the cysteine is the most constrained position in the lipobox, and is typically populated by a small, uncharged amino acid. The four most frequently-occurring residues at the ~1 position in leptomastigote lipoproteins are serine, asparagine, glycine and alanine (listed in descending order of frequency).

**Recombinant Len proteins and antibodies**

A polyhistidine-tagged recombinant LenA protein, based upon the *lenA* gene of serovar Lai, was previously described [12]. Additional polyhistidine-tagged recombinant proteins were produced using pET200 (Invitrogen). Recombinant LenB was produced from the gene of serovar Bratislava, one of the two serovars identified as having a complete *lenB* ORF. Recombinant proteins LenC-1 and LenC-2 were produced from the genes of serovars Bratislava and Canicola, respectively, representatives of the two *lenC* allele groups. As the *lenD* and *lenE* genes each form a tight phylogenetic cluster (Fig. 1C), serovars Pomona and Grippotyphosa were chosen at random for production of recombinant proteins LenD and LenE, respectively. The *lenF* genes of serovars Pomona and Hardjo served as templates for recombinant proteins LenF-1 and LenF-2, respectively, representatives of the two allele groups of that gene. Recombinant proteins formed insoluble inclusion bodies when produced in *Escherichia coli*, and so were purified in the presence of 8M urea using MagneHis nickel conjugated magnetic beads (Promega, Madison, WI). As a final step in purification, recombinant proteins were dialyzed against PBS using 10 kDa Slide-a-Lyzer cassettes (Pierce, Rockford, IL). Each of the new recombinant proteins precipitated out of solution during dialysis, and all except LenB remained insoluble unless 8M
urea was included in solvent. LenB dissolved into PBS after 2–3 days incubation at 4°C. Concentrations of the insoluble LenC-1, LenC-2, LenD, LenE, LenF-1 and LenF-2 recombinant proteins were determined by SDS-PAGE of homogeneous suspensions and Coomassie brilliant blue staining alongside protein standards of known concentration.

Polyclonal rabbit antisera directed against each Len protein were produced by Animal Pharm Services (Healdsburg, CA), using one round of their standard vaccination procedure (www.animalpharmservices.com). Briefly, approximately 2 mg of recombinant protein was suspended into PBS by vigorous mixing, then divided into 6 equal aliquots. A New Zealand White rabbit was injected 6 times over a 6 week period, then serum collected 1 week after the final injection.

Triton X-114 extraction

 Cultures of L. interrogans serovar Copenhageni strain LI-130 (approximately 10^6 cells/ml) were fractionated using Triton X-114 [20]. Bacteria were pelleted by centrifugation, washed in PBS containing 5 mM MgCl_2, then extracted with 0.5% protein-grade Triton X-114 (Calbiochem), 150 mM NaCl, 10 mM Tris, pH 8, and 2 mM EDTA, at 4°C. Insoluble material was pelleted by centrifugation at 16000 x g for 10 min. After centrifugation, 1 M CaCl_2 was added to the supernatant, to a final concentration of 20 mM. Phase separation was performed by warming the supernatant to 37°C, and subjecting it to centrifugation for 10 min at 20000 x g. The detergent and aqueous phases were separated, and proteins precipitated with acetone. Proteins were then separated by SDS-PAGE, blotted onto PVDF membranes and probed with polyclonal sera raised against leptospiral Len proteins or the flagellar component FlaA1 [61]. A 200 μl of 1 mg/ml blue dextran in elution buffer with 5% glycerol. The column was run with a flow rate of 0.20 ml per min. The elution of each standard was determined by monitoring A280. A calibration curve was created using an MW-GF-70 low molecular weight calibration kit (Sigma-Aldrich), and the void volume, V_0, was determined by injection of 200 mM NaCl, 50 mM Tris-HCl (pH = 7.5), 1% (vol/vol) glycerol. The column was run with a flow rate of 0.20 ml per min. The elution of each standard was determined by monitoring A280. A calibration curve was created using an MW-GF-70 low molecular weight calibration kit (Sigma-Aldrich), and the void volume, V_0, was determined by injection of 200 μl of 1 mg/ml blue dextran in elution buffer with 5% glycerol. The remaining protein standards: bovine lung aprotinin (6.5 kDa), horse heart cytochrome C (12.4 kDa), bovine carbonic anhydrase (29 kDa), and bovine serum albumin (66 kDa), were individually prepared in elution buffer with 5% glycerol to total concentrations of 0.3 mg/ml. The molecular mass calibration curve was generated by plotting the log (molecular mass) vs. V/V_0 [62]. A 200 μl sample of recombinant LenA (approximately 0.2 mg/ml) was then injected and its elution compared to the established curve.

Protein structure analyses

Circular dichroism (CD) spectra were collected using a J-810 spectropolarimeter equipped with a Peltier heating block (Jasco, Easton, MD). A 1 mm path length cuvette was employed, with reported spectra being the average of four scans taken at scan rates of 20 nm/min. Melting scans were performed at a scan rate of 1°C/min, recording the ellipticity at 202 nm which is the wavelength at which the largest change in ellipticity was observed. Protein concentrations were determined using the method of Brands and Kaplan [63]. Absorbance was measured in a 1.0 cm path length cuvette in a DU 640B spectrophotometer (Beckman-Coulter, Fullerton, CA). Secondary structure analysis of the CD spectra was performed using Dichroweb [http://www.cryst.biobk.ac.kr/dichweb/html/home.html] [64]. Reported secondary structure contents are averages of those calculated using the SELCON3 [65,66], ContinarL [67,68], and CDSSSTR [69,70] analysis programs.

Ligand-binding assays

Aliquots (1 μg) of each recombinant Len protein and negative control protein BSA were subjected to SDS-PAGE, then transferred to nitrocellulose membranes. Care was taken not to overheat recombinant proteins prior to gel loading, as incubation in boiling water for longer than 15 s tended to irreversibly interfere with ligand binding. Interactions between Len proteins and purified human factor H were examined as previously described [12]. For analyses of laminin and fibronectin binding, membranes were blocked with 5% BSA in Tris-buffered saline-Tween 20 (TBS-T; 20 mM Tris, 150 mM NaCl, 0.05% Tween 20 [pH 7.5]), then incubated for 1 h at room temperature with either murine laminin or human fibronectin (both from Sigma-Aldrich) at concentrations of 20 μg/ml in TBS-T. Following extensive washing with TBS-T, membranes were incubated for 1 h at room temperature with rabbit polyclonal antibodies specific for either murine laminin (diluted 1:5000) or human fibronectin (diluted 1:1000) (both from Sigma-Aldrich). Finally, the membranes were washed with TBS-T and incubated for 1 h at room temperature with horseradish peroxidase-conjugated protein A (GE Healthcare). Bound antibodies were detected using SuperSignal West Pico enhanced chemiluminescence substrate (Pierce).

Only recombinant LenA and LenB were soluble in the absence of urea (see above). Binding of host proteins by these soluble recombinant Len proteins was further measured using ELISA-based techniques, as described previously [3]. Immobilized target ligands included murine laminin, human plasma fibronectin, proteolytic fragments of fibronectin (70-kDa N-terminal fragment, the 30-kDa amino-terminal domain [NTD], and the 45-kDa gelatin-binding domain [GBD]) (all from Sigma-Aldrich), and human factor H (Calbiochem). In experiments testing the effect of ionic strength on Len-laminin interactions, additional NaCl was included in the PBS-based binding buffer. For heparin-competition assays, porcine heparin (Sigma-Aldrich) was added to the binding buffer along with the tested recombinant Len protein. In some experiments, heparin was also added to the laminin- or fibronectin-coated wells 1 h prior to the Len proteins. Results were reported as absorbance at 450 nm for the activity of horseradish peroxidase conjugated to a goat antibody (Novagen) against a monoclonal antibody for polyhistidine (Novagen). K_d values were calculated as the concentration of Len protein giving half-maximal binding. Means of independent experiments with equal variance were compared with Student’s t-test and alpha at 0.05.

Accession numbers

The new L. interrogans DNA sequences described in this work have been deposited in GenBank and given the following accession numbers. Serovar Pomona type kennewicki strain JEN4: lenB, lenC, lenD, lenE and lenF: EF606688 through EF606892; serovar Pomona strain Pomona lenA: EF606893; serovar Copenhageni strain M 20: lenA, lenB, lenC, lenD, lenE and lenF: EF606894 through...
EF606899; serovar Bratislava lenA, lenB, lenC, lenD, lenE, and lenF: EF632554 through EF632558; serovar Canicola strain Hond Utrecht IV lenA, lenB, lenC, lenD, lenE, and lenF: EF611235 through EF611240; serovar Grippotyphosa strain Andaman lenA, lenB, lenC, lenD, lenE, and lenF: EF999884 through EF999889; and serovar Hardjo strain Hardjo-pajroman lenA, lenB, lenC, lenD, lenE, and lenF: EF999890 through EF999895.

**SUPPORTING INFORMATION**

**Figure S1** Alignment of predicted amino acid sequences of representative proteins: serovar Lai LenA (LenA, Lai), serovar Bratislava LenB (LenB, Brat), serovar Bratislava LenC-1 (LenC, Brat), serovar Canicola LenC-2 (LenC, Can), serovar Pomona LenD (LenD, Pom), serovar Grippotyphosa LenE (LenE, Gripp), serovar Pomona LenF-1 (LenF, Pom), and serovar Hardjo LenF-1 (LenF, Har).

An alignment of these same proteins, with the proteins having two Len-motifs divided after the well-conserved internal CVEQ sequence, is presented in Figure 1. Identical amino acids found in the majority of proteins are boxed and shaded.

Cysteine residues that may serve as amino-terminal lipidation sites are circled. Found at: doi:10.1371/journal.pone.0001188.s001 (133.57 MB TIF)

**REFERENCES**

1. Bharri AR, Nally JE, Ricaldi JN, Mathias MA, Díaz MM, et al. (2003) Leptospirosis: a zoonotic disease of global importance. Lancet Infect Dis 3: 757–771.
2. Barbosa AS, Abreu PAE, Neves FO, Atzingen MV, Watanabe MM, et al. (2006) A newly identified leptospiral adhesin mediates attachment to laminin. Infect Immun 74: 6536–6546.
3. Choy HA, Kelley MM, Chen TL, Moller AK, Matsunaga J, et al. (2007) Physiological osmotic induction of Leptospira interrogans adhesion. LigA and LigB bind extracellular matrix proteins and fibrinogen. Infect Immun 75: 2541–2549.
4. Cinco M, Cini B, Perticarari S, Presani G (2002) Leptospira interrogans binds to the CR3 receptor on mammalian cells. Microb Pathog 33: 299–305.
5. Ito T, Yanagawa R (1987) Leptospiral attachment to four structural components of extracellular matrix. Jpn J Vet Sci 49: 875–882.
6. Merien F, Truccolo J, Baranton G, Perolat P (2000) Identification of a 36-kDa fibronectin-binding protein expressed by a virulent variant of Leptospira interrogans serovar australis/humabrae. FEMS Microbiol Lett 183: 17–22.
7. Cinco M, Banfi E (1993) Activation of complement by leptospires and in bacteriocidial activity. Zbl Bakter Hyg A 254: 261–265.
8. Johnson RC, Muschel LH (1985) Anti-leptospiral activity of normal serum. J Bacteriol 191: 265–267.
9. Johnson RC, Muschel LH (1986) Anti-leptospiral activity of serum: normal and immune serum. J Bacteriol 191: 1403–1409.
10. Johnson RC, Harris VG (1967) Anti-leptospiral activity of serum: leptospiral virulence factor. J Bacteriol 93: 513–519.
11. Meri T, Murgia R, Stefanel P, Meri S, Cinco M (2005) Regulation of complement activation at the C3b-level by serum resistant leptospires. Microbiol Pathog 39: 139–147.
12. Verma A, Helgason I, Artis LH, Helgason A, et al. (2004) Osmotic regulation of expression of two extracellular matrix-binding proteins and a haemolysin of Leptospira interrogans: differential effects on LigA and Spih extracellular release. Microbiology 150: 3377–3379.
13. Hohenester E, Sasaki T, Olsen BJ, Timpl R (1990) Crystal structure of the angiogenesis inhibitor endostatin at 1.5 Å resolution. EMBO J 17: 1566–15664.
14. Iozzo RV (2005) Basement membrane proteoglycans: from cellar to ceiling. Nat Rev Mol Cell Biol 6: 646–656.
15. Mar钥匙os RA, Sullivan DR (2000) Form and function: the laminin family of heterotrimers. Dev Dynamics 218: 213–234.
16. Pankov R, Yamada KM (2002) Fibronectin binding protein expressed by a virulent variant of Leptospira interrogans serovar pomona as a novel surface-exposed lipoprotein expressed during leptospiral dissemination in the mammalian host. Microbiology 152: 3757–3279.
17. Matsunaga J, Notsu M, Sanchez Y, Wernus KF, Ko AI (2007) Osmotic regulation of expression of two extracellular matrix-binding proteins and an endostatin. FASEB J 17: 1566–15664.
18. Haake DA, Pomona LenA (LenA, Pom), an intracellular release. Microbiology 153: 3390–3398.
19. Matsunaga J, Wernus KF, Zuerer RL, Frank A, Haake DA (2006) LipL41 is a novel surface-exposed lipoprotein expressed during leptospiral dissemination in the mammalian host. Microbiology 152: 3777–3786.
20. Sanyal J, Timoney JF, Stevenson B (2001) Temperature-regulated protein synthesis by Leptospira interrogans. Infect Immun 69: 400–408.
21. Matsunaga K, Sanyal J, Wernus KF, Ko AI (2007) Characterization of a leptospiral outer membrane lipoprotein LipL36: down-regulation associated with late-log-phase growth and mammalian infection. Infect Immun 66: 1579–1587.
22. Nally JE, Timoney JF, Stevenson B (2001) Temperature-regulated protein synthesis by Leptospira interrogans. Infect Immun 69: 400–408.
23. Matsunaga K, Sanyal J, Wernus KF, Ko AI (2007) Characterization of a leptospiral outer membrane lipoprotein LipL36: down-regulation associated with late-log-phase growth and mammalian infection. Infect Immun 66: 1579–1587.
24. Haake DA, Pomona LenA (LenA, Pom), an intracellular release. Microbiology 153: 3390–3398.
25. Matsunaga J, Notsu M, Sanchez Y, Wernus KF, Ko AI (2007) Osmotic regulation of expression of two extracellular matrix-binding proteins and an endostatin. FASEB J 17: 1566–15664.
26. Iozzo RV (2005) Basement membrane proteoglycans: from cellar to ceiling. Nat Rev Mol Cell Biol 6: 646–656.
27. Mar钥匙os RA, Sullivan DR (2000) Form and function: the laminin family of heterotrimers. Dev Dynamics 218: 213–234.
28. Pankov R, Yamada KM (2002) Fibronectin binding protein expressed by a virulent variant of Leptospira interrogans serovar pomona as a novel surface-exposed lipoprotein expressed during leptospiral dissemination in the mammalian host. Microbiology 152: 3777–3786.
29. Hohenester E, Sasaki T, Olsen BJ, Timpl R (1990) Crystal structure of the angiogenesis inhibitor endostatin at 1.5 Å resolution. EMBO J 17: 1566–15664.
30. Iozzo RV (2005) Basement membrane proteoglycans: from cellar to ceiling. Nat Rev Mol Cell Biol 6: 646–656.
31. Mar钥匙os RA, Sullivan DR (2000) Form and function: the laminin family of heterotrimers. Dev Dynamics 218: 213–234.
32. Pankov R, Yamada KM (2002) Fibronectin binding protein expressed by a virulent variant of Leptospira interrogans serovar pomona as a novel surface-exposed lipoprotein expressed during leptospiral dissemination in the mammalian host. Microbiology 152: 3777–3786.
33. Matsunaga J, Wernus KF, Zuerer RL, Frank A, Haake DA (2006) LipL41 is a novel surface-exposed lipoprotein expressed during leptospiral dissemination in the mammalian host. Microbiology 152: 3777–3786.
34. Force A, Lynch M, Pickett FB, Amores A, Yan Y, et al. (1999) Preservation of duplicate genes by complementary, degenerative mutations. Genetics 151: 1531–1545.
35. van Putten JPM, Duensing TD, Cole RL (1998) Entry of Opa+ gonococci into HEP-2 cells requires concerted action of glycosaminoglycans, fibrinectin and integrin receptors. Mol Microbiol 29: 369–379.
36. DiScipio RG, Dalfen PJ, Schraufstatter IU, Srinaroo P (1998) Human polymorphonuclear leukocytes adhere to complement factor H through an interaction that involves $\alpha_3\beta_1$ (CD11b/CD18). J Immunol 160: 4057–4066.

**ACKNOWLEDGMENTS**

We thank Michael Donahue, Albert Ko and Mathieu Picardeau for providing bacterial strains; Kelly Babh, Sara Bair, Logan Burns, Tomasz Bykowski, Sarah Kearns, James Matsunaga, Natalie Mickelson, Sean Riley, John Timoney, and Michael Woodman for assistance and helpful comments on this work.

**Author Contributions**

Conceived and designed the experiments: MS DH BS HC AV. Performed the experiments: TC BS HC MP MM PK AC CB AV. Analyzed the data: TC MS DH BS HC ED AV. Contributed reagents/materials/analysis tools: TC MS DH ED PK. Wrote the paper: DH BS.
37. Jokiranta TS, Cheng ZZ, Seeberger H, Jozsi M, Heinen S, et al. (2005) Binding of complement factor H to endothelial cells is mediated by the carboxy-terminal glycosaminoglycan binding site. Am J Pathol 167: 1173–1181.

38. Malhotra R, Ward M, Sim RB, Bird MI (1999) Identification of human complement factor H as a ligand for L-selectin. Biochem J 341: 61–69.

39. Vaziri-Sani F, Hellwage J, Zipfel PF, Sjoholm AG, Iancu R, et al. (2005) Factor V has a functional interaction with the antiplatelet glycoprotein GPIIb-IIIa. Thromb Haemostasis 93: 154–162.

40. Hammerschmidt S, Agarwal V, Kunert A, Haelbich H, Skerka C, et al. (2007) The host immune regulator factor H interacts via two contact sites with the PspC protein of Streptococcus pneumoniae and mediates adhesion to epithelial cells. J Immunol 178: 5848–5858.

41. Hellwage J, Jokiranta TS, Koistinen V, Vaarala O, Meri S, et al. (1999) Functional properties of complement factor H-related proteins FHR-3 and FHR-4 binding to the C3d region of C3b and differential regulation by heparin. FEBS Lett 462: 345–352.

42. Hellwage J, Eberle F, Babuke T, Seeberger H, Richter H, et al. (2006) Two factor H-related proteins from the mouse: expression, analysis and functional characterization. Immunogenetics 58: 883–893.

43. McRae JL, Dubby TG, Griggs KM, Ormsby RJ, Cowan PJ, et al. (2005) Human factor H-related protein 3 has cofactor activity, inhibits C3 convertase activity, binds heparin and C-reactive protein. J Immunol 174: 6250–6256.

44. Park CT, Wright SD (1996) Plasma lipopolysaccharide-binding protein is found associated with a particle containing apolipoprotein A-I, phospholipid, and factor H-related proteins. J Biol Chem 271: 18054–18060.

45. Zipfel PF, Ezley M, Heinen S, Jozsi M, Richter H, et al. (2007) Deletion of complement factor H-related genes CFHR1 and CFHR3 is associated with atypical hemolytic uremic syndrome. PLoS Genet 3: e41.

46. Saito A, Munakata H (2007) Analysis of plasma proteins that bind to Leptospira interrogans. FEBS Lett 586: 345–352.

47. Haake DA, Dundoo M, Cader R, Kubak BM, Hartskeerl RA, et al. (2002) Molecular evolution and mosaicism of leptospiral outer membrane proteins involves horizontal DNA transfer. J Bacteriol 184: 2818–2828.

48. McRae JL, Dubby TG, Griggs KM, Ormsby RJ, Cowan PJ, et al. (2005) Human factor H-related protein 3 has cofactor activity, inhibits C3 convertase activity, binds heparin and C-reactive protein. J Immunol 174: 6250–6256.

49. Ren S-X, Fu G, Jiang X-G, Zeng R, Miao Y-G, et al. (2003) Unique glycosaminoglycan binding site. Am J Pathol 167: 1173–1181.

50. Haake DA, Suchard MA, Kelley MM, Dundoo M, Ah DP, et al. (2004) The host immune regulator factor H interacts via two contact sites with the PspC protein of Streptococcus pneumoniae and mediates adhesion to epithelial cells. J Immunol 178: 5848–5858.

51. Johnson RC, Harris VG (1967) Differentiation of pathogenic and saprophytic disease spirochetes. Infect Immun 72: 742–749.

52. Artushin S, Timoney JF, Nally J, Verma A (2004) Host-inducible immunogenic sphingomyelinase-like protein, Lk73.5, of Leptospira interrogans. Infect Immun 72: 4853–4863.

53. Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG (1997) The Clustal X window interface: flexible strategies for multiple sequence alignment aided by quality analyses tools. Nucleic Acids Res 24: 4876–4882.

54. Swofford DL (2000) PAUP*: phylogenetic analysis using parsimony (*and other methods), version 4. Sunderland, MA: Sinauer Associates.

55. Hasegawa M, Kishino H, Yano T (1985) Dating the human-ape splitting by a molecular clock of mitochondrial DNA. J Mol Evol 22: 160–174.

56. Drummond AJ, Ho SYW, Phillips MJ, Rambaut A (2006) Relaxed phylogenetics and dating with confidence. PLoS Biology 4: e88.

57. Efron B, Halloran E, Holmes S (1996) Bootstrap confidence levels for phylogenetic trees. Proc Natl Acad Sci USA 93: 13429–13434.

58. Suchard MA, Weiss RE, Dorman KS, Sinsheimer JS (2002) Oh brother, where art thou? A Bayes factor test for recombination with uncertain heritage. Systematic Biol 51: 715–728.

59. Suchard MA (2005) Stochastic models for horizontal gene transfer: taking a random walk in tree-space. Genetics 170: 419–431.

60. Haake DA (2000) Spirochetal lipoproteins and pathogenesis. Microbiology 146: 1491–1504.

61. Cullen PA, Xu X, Matsunaga J, Sanchez Y, Ko AI, et al. (2005) Surfaceome of Leptospira spp. Infect Immun 73: 4853–4863.

62. Andrews P (1964) Estimation of the molecular weights of proteins by Sephadex gel-filtration. Biochem J 91: 222–233.

63. Brands JF, Kaplan LJT (1973) Derivative spectroscopy applied to tyrosyl chromophores. Studies on ribonuclease, lima bean inhibitors, insulin, and pancreatic trypsin inhibitor. Biochemistry 12: 2011–2024.

64. Lobley A, Whitmore I, Wallace BA (2002) DICROWEB: an interactive website for the analysis of protein secondary structure from circular dichroism spectra. Bioinformatics 18: 211–212.

65. Sreerama N, Venyaminov SY, Woody RW (1999) Estimation of the number of alpha-helical and beta-strand segments in proteins using circular dichroism spectroscopy. Protein Sci 8: 370–380.

66. Sreerama N, Woody RW (1993) A self-consistent method for the analysis of protein secondary structure from circular dichroism. Anal Biochem 209: 32–44.

67. Provencher SW, Glowacki J (1981) Estimation of globular protein secondary structure from circular dichroism. Biochemistry 20: 35–57.

68. van Stokkum IH, Spoelder HJ, Bloemendal M, van Grondelle R, Groen FC (1990) Estimation of protein secondary structure and error analysis from circular dichroism spectra. Anal Biochem 191: 110–118.

69. Severson JL, Williams SR (1994) Discussion of pathogenic and saprophytic leptospires: I. Growth at low temperatures. J Bacteriol 176: 2164–2172.

70. Ren S-X, Fu G, Jiang X-G, Zeng R, Miao Y-G, et al. (2003) Unique physiological and pathogenic features of Leptospira interrogans revealed by whole-genome sequencing. Nature 422: 888–893.

71. Johnson RC, Harris VG (1967) Differentiation of pathogenic and saprophytic leptospires: I. Growth at low temperatures. J Bacteriol 94: 27–31.

72. Amundson S, Timoney JF, Nally J, Verma A (2004) Host-inducible immunogenic sphingomyelinase-like protein, Lk73.5, of Leptospira interrogans. Infect Immun 72: 742–749.