Targeting RNA-Mediated Toxicity in C9orf72 ALS and/or FTD by RNAi-Based Gene Therapy

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A hexanucleotide GGGGCC expansion in intron 1 of chromosome 9 open reading frame 72 (C9orf72) gene is the most frequent cause of amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD). The corresponding repeat-containing sense and antisense transcripts cause a gain of toxicity through the accumulation of RNA foci in the nucleus and deposition of dipeptide-repeat (DPR) proteins in the cytoplasm of the affected cells. We have previously reported on the potential of engineered artificial anti-C9orf72-targeting miRNAs (miC) targeting C9orf72 to reduce the gain of toxicity caused by the repeat-containing transcripts. In the current study, we tested the silencing efficacy of adeno-associated virus (AAV)-inspired miC in human-derived pluripotent stem cell (iPSC) neurons and in an ALS mouse model. We demonstrated that AAV5-miC transduces different types of neuronal cells and can reduce the accumulation of repeat-containing C9orf72 transcripts. Additionally, we demonstrated silencing of C9orf72 in both the nucleus and cytoplasm, which has an added value for the treatment of ALS and/or FTD patients. A proof of concept in an ALS mouse model demonstrated the significant reduction in repeat-containing C9orf72 transcripts and RNA foci after treatment. Taken together, these findings support the feasibility of a gene therapy for ALS and FTD based on the reduction in toxicity caused by the repeat-containing C9orf72 transcripts.

INTRODUCTION

Amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD) are two severe neurodegenerative diseases, with overlapping pathologic and genetic features but distinct clinical features. ALS is the most common adult-onset motor neuron degenerative disorder that affects mainly the upper and lower motor neurons in the brain and corticospinal tract.1,2 FTD is a presenile dementia characterized by the degeneration of neurons in the frontal and temporal lobes of the brain.3,4 A significant number of patients develop both diseases (ALS-FTD).4

The most common genetic mutation in both ALS and FTD is a hexanucleotide GGGGCC (G4C2) repeat expansion in the first intron of the chromosome 9 open reading frame 72 (C9orf72) gene.4–6 ALS and FTD patients display hundreds to a few thousand copies of the G4C2 repeat in the C9orf72 gene.6 The contribution of this mutation to the pathogenesis of both diseases has been debated for several years, with the loss of C9orf72 function (haploinsufficiency), a gain of toxicity, or a combination of both being implicated.7,8 Reduced C9orf72 mRNA levels in patients due to hypermethylation of the G4C2 repeat supports haploinsufficiency.1,9 On the other hand, a causal role for RNA-mediated toxicity is supported by accumulation of the repeat-containing transcripts that fold into stable structures, forming RNA foci enriched with RNA-binding proteins in the nucleus.5,10–13 RNA foci are detected in several repeat expansion diseases and can sequester RNA-binding proteins. RNA foci produced from both the sense (G4C2) and antisense (G2C4) repeat transcripts are detected in tissues and in induced pluripotent stem cell (iPSC)-derived neurons from ALS and FTD patients, proving that the repeat region is bidirectionally transcribed.12,14

Gain of toxicity is also supported by repeat-associated non-ATG (RAN) translation of the sense and antisense repeat transcripts, resulting in the accumulation of five aberrant dipeptide-repeat (DPR) proteins (poly(GA), poly(GR), poly(GP), poly(PA), and poly(PR)) in the cytoplasm, all with different toxicity profiles shown in vitro and in vivo.12,15,16 In addition, DPRs disrupt the nucleocytoplasmic transport system of cells.17,18 Furthermore, autopsy studies revealed that ~90% of C9orf72 ALS and ~50% of FTD patients have cytoplasmic aggregation of the transactive response DNA-binding protein of 43 kDa (TDP-43; encoded by TAR DNA binding protein [TARDBP]).5,19 Abnormal aggregation of P62 and ubiquitin have...
also been described in C9orf72-related ALS and FTD patients. A recently developed bacterial artificial chromosome (BAC) transgenic mouse model expressing the human C9orf72, including the expanded G4C2 repeat, showed gain-of-toxicity features such as RNA foci, DPRs, TDP-43, and p62 inclusions. These mice also develop neurodegeneration and ALS and/or FTD-like phenotypes, suggesting that these inclusions result in a gain of toxicity and contribute to the pathology observed in ALS and FTD.

Regardless of the contribution of either RNA-mediated toxicity or haploinsufficiency, a therapy reducing the repeat-containing transcripts could potentially translate into a reduction of RNA foci and DPR proteins, slowing down the disease progression. Silencing of C9orf72 transcripts by RNAi strategies, such as duplex and single-stranded small interfering RNAs (siRNAs), or by the RNase H-mediated antisense oligonucleotides (ASOs) indeed resulted in the reduction of RNA foci and DPR proteins in patient-derived iPSC neurons and in mouse models. Interestingly, targeting only the sense strand of C9orf72 with ASOs not only reduced RNA foci but also rescued the disrupted nucleocytoplasmic transport in patient-derived iPSC neurons.

Another promising strategy to achieve similar outcomes is by adeno-associated virus (AAV)-delivered artificial microRNAs (miRNAs) engineered to target C9orf72. miRNAs are short non-coding RNAs that bind to a complementary mRNA through specific base pairing, inducing its degradation and/or translational repression. As AAVs express stable extrachromosomal nuclear episomes, the primary miRNA transcripts can be continually produced, resulting in a longer-lasting therapeutic effect. The primary miRNA transcripts are processed into precursor miRNAs that are transported to the cytoplasm for further processing and incorporation into the RNA-induced silencing complex (RISC). Thus, one major challenge to overcome for a miRNA-based gene therapy approach is targeting of the repeat-containing C9orf72 transcripts within the cell nucleus. In one study, siRNAs, which are also processed by RISC, demonstrated the silencing of C9orf72 mRNA in patient-derived iPSC neurons, but the nuclear repeat-containing transcripts and RNA foci were unaffected, indicating a predominant efficacy in the cytoplasm.

We generated and characterized different human-derived iPSC neurons and astrocytes to validate the transduction of AAV5 in different cell types (Figure 1). iPSCs were induced into a neural progenitor state and differentiated into frontal brain-like neurons (FBNs) or astrocytes (Figure S1). In addition, commercially available mature dopaminergic neurons and motor neurons from a healthy person were obtained. Immunohistochemistry was performed and about 60% of FBNs were β-tubulin III positive and glial fibrillary acidic protein (GFAP) negative, implicating a successful differentiation rate of iPSCs into mature neurons. Similarly, mature astrocytes were ~90% GFAP positive, confirming a successful differentiation of iPSCs into astrocytes. Mature dopaminergic neurons were ~90% tyroxine hydroxylase (TH) positive, confirming successful differentiation. Mature motor neurons were also successfully differentiated as ~85% were choline acetyltransferase (CHAT) positive.

Following transduction with AAV5-GFP, ~90% of all the different cell types expressed GFP (Figure 1A). Immunohistochemistry for GFP combined with either β-tubulin III, TH, GFAP, or CHAT antibodies confirmed that all four cell types were transduced by AAV5 (Figure 1B). To compare the AAV transduction tropism of the different cell types, we isolated DNA and RNA of transduced cells and quantitated vector copies and GFP mRNA expression in the cells (Figures 1C and 1D). A similar dose-dependent transduction efficiency was observed in all cell types, and transduction correlated with GFP expression. Thus, AAV5 efficiently transduces different human CNS-specific cell types, including FBNs, dopaminergic neurons, motor neurons, and astrocytes, and, hence, it is a promising vector to deliver therapeutic genes to the CNS to treat neurodegenerative diseases such as ALS and FTD.
C9orf72 Expression Is Reduced in Neuronal Cells Derived from an FTD Patient

iPSCs from an FTD patient (ND42765) and a healthy non-diseased person (ND42245) were differentiated into FBNs (FTD-FBN) and astrocytes to compare the levels of C9orf72 mRNA and repeat-containing transcripts (Figure S1). qRT-PCR was performed 2 weeks after maturation for total C9orf72 mRNA (detecting all transcript variants) and the sense intronic transcripts (detecting sense transcripts containing the G4C2 repeat) to compare the expression levels in these cells. Primers amplifying a region spanning exon 2 to exon 4 were used to detect total C9orf72 mRNA (Figure 2A).10 The sense intronic transcripts were detected with primers amplifying a region in intron 1.10 The levels of total C9orf72 mRNA were significantly reduced in the FTD patient-derived cells: a reduction of ~60% was observed in FBNs and ~25% in astrocytes from the FTD patient as compared to healthy cells (Figure 2B). Interestingly, although at a low level, sense intronic transcript levels were increased by ~30% in FBNs and ~20% in astrocytes of the FTD patient as compared to healthy cells (Figure 2C). Thus, while total C9orf72 mRNA levels were reduced, sense intronic transcripts seem to accumulate in iPSC-derived FBNs and astrocytes from the FTD patient.

AAV5-miC Can Lower the Repeat-Containing Transcripts of C9orf72 in iPSC Neurons

Sequences on the human C9orf72 were previously selected to design miC.29 The miC sequences were embedded in the primary miR-101 and/or miR-451 scaffold by replacing the naturally expressed guide strand sequences. Four lead miC candidates were selected in the miR-101 scaffold based on their efficacy on reporter genes and their ability to reduce the endogenously expressed C9orf72 mRNA and sense intronic transcripts in cells.29 miC32 and miC46 were designed to target C9orf72 exon 2 and exon 11, respectively, targeting all sense C9orf72 transcripts (Figure 2A). miC2 and miC4 were designed in intron 1 to selectively silence the sense G4C2 sense intronic transcripts.

To determine whether miC delivered by AAV5 is functional in patient-derived cells, FTD-FBNs were transduced with AAV5-miC2, AAV5-miC4, AAV5-miC32, and AAV5-miC46. At 2 weeks following transduction, all four mature miC were expressed, suggesting a successful transduction by AAV5-miC and efficient processing into a mature miC (Figure 2D). Sense intronic transcript levels were reduced by ~40% in FBNs transduced with miC2 and miC4, while the C9orf72 mRNA levels were not affected (Figure 2E). Thus, both candidates exclusively target the sense intronic transcripts while preserving normal levels of C9orf72 mRNA. Of the candidates targeting the total C9orf72 mRNA, both miC32 and miC46 reduced the levels of C9orf72 mRNA (~50%) and the sense intronic transcript (~40%). Thus, the sense intronic transcripts could be also targeted by silencing total C9orf72 mRNA.
Additionally, we investigated silencing of C9orf72 in a healthy motor neuron cell line, as motor neurons are highly affected in ALS. The expression of C9orf72 was first evaluated in control (non-transduced) motor neurons, and both total and intronic C9orf72 were detected. However, the intronic C9orf72 expression in this cell line, which lacks the G4C2 expansion, was very low and slightly above the detection limit (data not shown). Having established that C9orf72 can be detected in healthy motor neurons, the cells were transduced with AAV5-miC32 and AAV5-miC46 for 2 weeks. We found expression of miC32 and miC46, confirming that AAV5 efficiently transduces human motor neurons (Figure 2F). Consistently, we observed an ~40% reduction in total C9orf72 mRNA by both miC candidates and a mild reduction in the intronic C9orf72 (~20%) (Figures 2G and 2H). Altogether, we demonstrated the reductions in total and intronic C9orf72 levels in FBNs and motor neurons, confirming that both neuronal cell types are transduced and that the miC candidates are effective at lowering C9orf72 in these cells.
Efficient Silencing of C9orf72 in the Nucleus of iPSC-Derived Neurons by AAV-miC

The accumulation of the G4C2 repeat-containing transcripts in the cell nucleus seems to highly contribute to the progression of both ALS and FTD. These transcripts form RNA foci in the cell nucleus that sequester RNA-binding proteins and inhibit their function or are transported to the cytoplasm for RAN translation into toxic DPRs. Thus, for a therapeutic approach, effector molecules that sequester RNA-binding proteins and inhibit their function or are transported to the cell nucleus seems to highly contribute to the progression of both ALS and FTD. The processing of miRNAs occurs through a multi-step process involving nuclear and cytoplasmic phases, but the mature miRNA product is produced in the cytoplasm. Therefore, miRNAs were initially thought to be predominantly expressed and active in the cytoplasm. We previously demonstrated that active mature miC is also detected in the nucleus of cells transfected with miC constructs, but at lower levels than in the cytoplasm. Here we evaluated whether the transduction of iPSC neurons by AAV5-miC is sufficient to express the mature miC and reduce C9orf72 levels in the nucleus (Figure 3).

FTD-FBNs were transduced with AAV5-miC32 and AAV5-miC46, and after a week RNA was isolated from nuclear and cytoplasmic fractions to calculate the percentage of RNA transcripts in both cellular compartments. In control FBNs, ~80% of total C9orf72 mRNA was detected in the nucleus and ~20% was measured in the cytoplasm, whereas sense intronic transcripts were predominantly (~95%) found in the nucleus of FTD-FBNs (Figure 3A). Thus, both C9orf72 mRNA and sense intronic transcript levels were significantly higher in the nucleus of FTD-FBNs. Next, the percentage of the mature miC and the silencing of C9orf72 were determined in nucleus and cytoplasm after transducing FTD-FBNs with AAV5-miC32 and AAV5-miC46. About 20% of the mature miC32 was detected in the nucleus while ~80% was measured in the cytoplasm (Figure 3B). In cells treated with AAV5-miC46, ~10% of the mature miC was expressed in the nucleus and ~90% in the cytoplasm. Interestingly, both AAV5-miC32 and AAV5-miC46 resulted in an ~30% reduction in C9orf72 mRNA in the nucleus and an ~40% reduction in the cytoplasm (Figures 3C and 3D). Consistently, an ~25% reduction in the sense intronic transcripts was observed in the nucleus (Figure 3E). Our data show that the mature miC32 and miC46 can both shuttle from the cytoplasm to the cell nucleus and can reduce levels of both C9orf72 mRNA and the sense intronic transcripts in the nucleus as well as in the cytoplasm.

AAV5-miC32 and AAV5-miC46 Can Both Reduce C9orf72 in Tg(C9orf72_3) Line 112 Mice

Having established the efficacy of AAV5-miCs in different human neuronal cell types, we next evaluated their efficacy in vivo in Tg(C9orf72_3) line 112 mice. This mouse model is based on several tandem copies of the human C9orf72, with repeat sizes ranging from 100–1,000 repeats. Although the progressive neurodegeneration seen in ALS and FTD patients is not observed in these mice, they do exhibit some of the pathological features seen in patients, such as RNA foci.
The 3-month-old mice were injected bilaterally in the striatum with AAV5-GFP, AAV5-miC32, and AAV5-miC46. Mice were sacrificed 6 weeks post-injection to determine the distribution of AAV5, mature miC expression, C9orf72 lowering, and the effect of miC on RNA foci formation. A widespread distribution of AAV5 to the cortex, striatum, and midbrain was observed after administration in the striatum (Figure 4A). A weak transduction of the cerebellum was observed while the spinal cord was not transduced. Consistent with the AAV5 distribution, small RNA TaqMan showed high expression of miC32 and miC46 in the cortex and striatum, which resulted in a 20%–40% lowering of C9orf72 mRNA and the sense intronic transcripts (Figures 4B–4D). Both AAV5-miC32 and AAV5-miC46 also target the mouse C9orf72 ortholog (310043O21 Rik), and indeed they lowered the target 310043O21 Rik. No behavioral and/or phenotypic changes were observed in mice treated with AAV5-miC32 or AAV5-miC46 (Figure S3).

AAV5-miC32 and AAV5-miC46 Are Processed Differently in the Mouse Brain

We further investigated the fidelity of miC processing in the mouse brain. Following transcription of the miC construct, the primary miR-101 is processed by Drosha cleavage and then by Dicer cleavage into a miRNA duplex. The miRNA duplex is then separated, and the guide strand is usually incorporated into the RISC while in most cases the passenger strand is degraded. The processing of the miC32 and miC46 was analyzed by small RNA sequencing to determine the ratio of guide and passenger strands that are produced. Small transcriptome analysis was performed on RNA isolated from the striatum of four mice that were injected with AAV5-miC32 or AAV5-miC46. For each sample, we obtained between 15 and 30 million small RNA reads that were subsequently adaptor trimmed and aligned to the mouse reference genome. No behavioral and/or phenotypic changes were observed in mice treated with AAV5-miC32 or AAV5-miC46 (Figure S3).

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AAV5-miC Reduces RNA Foci in Tg(C9orf72_3) Line 112 Mice

RNA foci formation by the repeat-containing transcripts is considered a hallmark of the RNA-mediated toxicity in ALS and/or FTD. Fluorescence in situ hybridization (FISH) using a TYE563-(C6G3)3 locked nucleic acid (LNA) probe showed that ~60%–80% of cells in cortex, hippocampus, and cerebellum of the Tg(C9orf72_3) line 112 mice contained RNA foci (Figure 4F). After confirming the presence of sense and antisense RNA foci in the cortex, hippocampus, and cerebellum, the efficacy of AAV5-miC32 and AAV5-miC46 to reduce RNA foci was determined. Both miC candidates caused a significant drop of sense RNA foci in cortex and hippocampus and the number of cells containing RNA foci, as well as in the total amount of RNA foci per cell (Figures 4G and 4H; Figure S2). AAV5-miC32 resulted in a 20% drop of cells containing RNA foci, and the number of cells containing 1–5 RNA foci and >5 RNA foci were reduced by 11% and 9%, respectively. Similarly, AAV5-miC46 treatment resulted in a 14% reduction in foci-containing cells, a 3% reduction in cells containing 1–5 foci, and an 11% reduction in cells containing >5 RNA foci. Hence, these data confirm that AAV5-delivered miC candidates against total C9orf72 mRNA are functional in reducing nuclear sense RNA foci in brain tissues of the Tg(C9orf72_3) line 112 mice.

Overall, we demonstrated that AAV5 can transduce different CNS cell types relevant for ALS and/or FTD treatment and that miC candidates targeting C9orf72 are successfully delivered and are functional in the mouse brain. Furthermore, we showed that total C9orf72 mRNA and sense intronic C9orf72 transcripts can be lowered in both the nucleus and cytoplasm of cells, increasing the potential for achieving therapeutic benefit in patients.

DISCUSSION

The intronic G4C2 repeat of C9orf72 produces repeat-containing transcripts that cause RNA foci and DPR proteins, which contribute to ALS and/or FTD pathology. Thus, a therapy reducing these gain-of-toxicity features could slow down disease progression in ALS.

Figure 4. Reduction of C9orf72 in C9BAC Mice

(A) Vector copy distribution of AAV5 upon intrastratial injection. The 3-month-old Tg(C9orf72_3) line 112 mice were injected with AAV5-GFP (5e10 GC), AAV5-miC32 (5e10 GC), and AAV5-miC46 (1e10 GC) bilaterally in the striatum. All mice were sacrificed 6 weeks after surgeries, and frontal cortex, striatum, midbrain, cerebellum, and spinal cord were collected. DNA was isolated from the tissues, and qPCR was performed with primers amplifying a 95-bp fragment from the CAG promoter region. The genome copies per tissue were calculated using a standard curve. (B) Expression of mature miC32 and miC46 guide strands in the cortex and striatum of Tg(C9orf72_3) line 112 mice. Performed as described in (A), total RNA was isolated from the cortex and striatum for small RNA TaqMan. MicroRNA input levels were normalized to U6 small nuclear RNA and set relative to AAV-GFP-treated mice. (C) Reduction of mature miC32 and miC46 guide strands in the cortex and striatum of Tg(C9orf72_3) line 112 mice. Performed as described in (A), total RNA was isolated from the cortex and striatum, and qRT-PCR was performed using primers for total C9orf72 mRNA and sense intronic transcripts. RNA input levels were normalized to GAPDH and set relative to AAV-GFP mice. (E) Detection of RNA foci in the cortex of Tg(C9orf72_3) line 112 mice. Mouse brain was frozen and sectioned in OCT. RNA FISH was performed using a TYE563-(C6G3)3 LNA probe to detect the sense foci. Sense foci (shown as white spots) were detected in Tg(C9orf72_3) line 112 mice (C9+), but not in control littermates (C9−). (G and H) Reduction of RNA foci in fronta cortex. Cells with 0, 1–5, or >5 foci were counted in control (AAV5-GFP) and treated groups (AAV5-miC32 and AAV5-miC46). Red arrows show the cells that contain RNA foci (C). The percentages of cells containing 0, 1–5, or >5 foci were calculated from 6 different images per treatment group (̄n = 3). Data were evaluated using a one-way ANOVA with Dunnett’s multiple comparison test (*p < 0.05, **p < 0.01, ***p < 0.001, and ****p < 0.0001).
and/or FTD patients. In this study, we provide evidence that AAV5-delivered miRNAs targeting C9orf72 reduce gain-of-toxicity features caused by the G\(_{4}C_{2}\) repeat in a murine model.

Initially described as a pure motor neuron disease, it is now thought that other cell types, including resident glial cells, are involved in ALS.\(^{41,42}\) Abnormal neuropsychological testing is observed in \(~50\%\) of ALS patients, indicating that, besides motor neurons, other neuronal cell types in the brain are affected.\(^{31}\) In addition, some patients develop both ALS and FTD, affecting different types of neurons in the brain and spinal cord. Delivery of therapeutics to the affected cell types is a major challenge for ALS and/or FTD therapy. We demonstrated that AAV5 can transduce various cell types of the CNS that are relevant to both diseases.

We observed a reduction of C9orf72 mRNA in astrocytes and FBNs derived from an FTD patient compared to healthy cells. This finding was consistent with several other studies that reported a reduction of C9orf72 mRNA and protein in iPSC neurons and in brain and spinal cord tissues from C9orf72-related ALS and/or FTD patients.\(^{6,9,43-46}\) This reduction is caused by methylation of the repeat region of C9orf72, which is located in or near the promoter region, leading to transcription inhibition.\(^{77,40}\) However, C9orf72 haploinsufficiency alone is most likely not sufficient to cause neurodegeneration. The reduction of C9orf72 in mice was tolerable, while complete elimination of C9orf72 caused splenomegally and enlarged lymph nodes, but not neurodegeneration or mis-localization of TDP-43.\(^{59-51}\)

Compared to C9orf72 mRNA levels, the expression of sense intrinsic transcripts was low in FTD and control cells. Yet, sense intrinsic transcripts were increased in FTD cells as compared to healthy cells, consistent with what has been reported by others.\(^{10}\) The elevated sense intrinsic transcript levels detected in the FTD cells seem to be caused by defective splicing of intron 1 due to the presence of the G\(_{4}C_{2}\) repeat.\(^{7,52}\) The corresponding transcripts containing the G\(_{4}C_{2}\) repeat may be protected from degradation, which allows them to accumulate in the cell.\(^{44,52-54}\) Despite a relatively low abundance as compared to C9orf72 mRNA, the intrinsic transcripts could still be sufficient to accumulate in RNA foci over time.\(^{28,47,52}\) Furthermore, it has been estimated that hundreds of protein products could be produced by a single miRNA, suggesting that even low levels of intrinsic transcripts are sufficient to cause the accumulation of toxic DPR proteins in the cell.\(^{5,55}\)

Several approaches, such as ASOs, duplex and single-stranded siRNAs, and small compounds, have been tested and proved promising to reduce gain-of-toxicity features caused by the G\(_{4}C_{2}\) repeat.\(^{44,26,56}\) In this study, we investigated the feasibility of a miRNA-based gene therapy to obtain long-term silencing of the repeat-containing transcripts of C9orf72. Four AAV5-miC candidates were tested on human-derived iPSC neurons, and all four resulted in sufficient transduction to express therapeutically relevant levels of the corresponding mature miC. Selective reduction of the sense intrinsic transcripts was achieved with the miC targeting intron 1, without affecting normal C9orf72 mRNA levels. This approach could prevent RNA-mediated toxicity without further reducing the C9orf72 protein. However, as sequence variations within intron 1 have been observed, genomic screening of patients for selection could be necessary when targeting this region.\(^{57}\) Thus, sequence conservation of miC2 and miC4 targets should be determined in larger cohort studies.

We previously used a publicly available RNA sequencing (RNA-seq) database of patients to investigate the conservation of miC2 and miC4 target sites in intron 1, but the intronic transcript levels detected were too low to determine their conservation.\(^{29,58}\) The target sequences of miC32 and miC46, targeting either exon 2 or exon 11, were well conserved between the patients, and both candidates also reduced levels of the sense intronic transcripts in iPSC neurons. The reduction of sense intrinsic transcripts by miC32 and miC46 supports previous findings that intron 1 is still present in the mature C9orf72 mRNA, suggesting defective splicing.\(^{32}\) Although miC32 and miC46 also reduced the levels of normal C9orf72 mRNA, its expression was not completely eliminated. Additionally, we observed transduction, mature miC expression, and the reduction of C9orf72 mRNA in a healthy motor neuron cell line. As expected, the expression of the sense intrinsic transcripts in this cell line was very low, as the repeat expansion is absent in healthy individuals.

Having established that miCs delivered by AAV5 are effective in human-derived iPSC neurons, we specifically studied their efficacy in the cell nucleus where C9orf72 mRNA and intrinsic transcripts are predominantly expressed. Although the primary and precursor miRNAs originate from the cell nucleus, their transport to the cytoplasm to exert post-transcriptional gene silencing via the RISC has been well described.\(^{26,38,39,59}\) Indeed, most studies have initially focused on post-transcriptional gene silencing of miRNAs in the cytoplasm. However, the discovery of several mature miRNA and RISC components enriched in the nucleus indicate that nuclear miRNAs do exist.\(^{60}\) Additionally, several proteins mediating nucleus-cytoplasm shuttling of small RNAs have been identified.\(^{60,61}\) We found ~5 times lower levels of mature miC in the nucleus compared to cytoplasm of transduced iPSC neurons, but the nuclear miC levels were still sufficient to reduce the levels of C9orf72 mRNA and the sense intrinsic transcripts in the nucleus. Thus, AAV-delivered miC can lower the repeat-containing transcripts that accumulate in the nucleus of ALS and/or FTD patients.

Moving forward to an in vivo proof-of-concept study, we tested the delivery and efficacy of AAV5-miC in the BAC transgenic Tg(C9orf72_3) line 112 mouse model.\(^{62}\) These mice exhibit pathologic features such as sense and antisense RNA foci and poly(GP) protein but no TDP-43 or P62 inclusions. These mice also do not develop the ALS and/or FTD-like phenotype in their lifespan, possibly due to the lack of 5′ and 3′ regulatory elements needed to control the sufficient expression of sense and antisense transcripts.\(^{23,30}\) Intracisternal injection of AAV5 resulted in a strong localized transduction on the injection site and surrounding areas, including frontal cortex and midbrain area, but it was not sufficient to transduce the spinal
in vivo. Moreover, a reduction in the mouse C9orf72 ortholog was also observed and was well tolerated in mice (3110043O21Rik). The processing of the miC in the mouse brain revealed that miC32 has a low amount of passenger strand, decreasing the risk for off-target effects in comparison to miC46. In this study, all mice were sacrificed at 5 months of age; future studies in older mice that have more accumulation of poly(GP) proteins would be needed to investigate the effect of miC on the DPR protein accumulation. However, based on the observed reduction of the sense intronic transcripts, a reduction of poly(GP) protein would be expected, as fewer repeat-containing transcripts are available to undergo translation.

Taken together, these data provide a proof of concept for the silencing of C9orf72 by AAV5-miC in relevant cell types as a potential treatment approach for ALS and/or FTD. miC32 offers the superior profile as a candidate to reduce gain of toxicity in ALS and/or FTD due to the sense intronic transcripts. We showed nuclear and cytoplasmic silencing, increasing the potential for a therapeutic effect of a miC32-based gene therapy that silences C9orf72.

**MATERIALS AND METHODS**

**Cell Culture**

Human control (ND42245) and Frontotemporal Dementia (ND42765) iPSCs derived from fibroblasts were ordered from Coriell Biorepository and cultured on Matrigel (Corning)-coated 6-well plates in mTeSR1 (STEMCELL Technologies). For embryoid body-based neural induction, iPSCs were seeded on AggreWell800 plates and cultured in STEMdiff Neural Induction Medium (STEMCELL Technologies) for 5 days with daily medium changes. Embryoid bodies were harvested and plated on 6-well plates coated with poly-D-lysine (Sigma-Aldrich) and laminin (Sigma-Aldrich) in STEMdiff Neural Induction Medium for 7 days with daily medium changes. Rosettes were harvested with STEMdiff Neural Rosette Selection Reagent (STEMCELL Technologies) and plated on poly-D-lysine- and laminin-coated 6-well plates in STEMdiff Neural Induction Medium for 24 h.

For differentiation into FBNs, STEMdiff Neural Induction Medium was replaced with STEMdiff Neuron Differentiation Medium (STEMCELL Technologies), and neuroprogenitor cells were differentiated for 5 days. For differentiation into astrocytes, neuroprogenitor cells were differentiated in STEMdiff Astrocyte Differentiation Medium (STEMCELL Technologies). The neuroprogenitor cells were then plated on poly-D-lysine- and laminin-coated plates in STEMdiff Neuron Maturation Medium (STEMCELL Technologies) for 1 week or STEMdiff Astrocyte Maturation Medium for 3 weeks. The mature FBNs and astrocytes were stored in liquid nitrogen in Neuroprogenitor Freezing Medium (STEMCELL Technologies).

Cryopreserved non-diseased mature dopaminergic neurons (iCell Dopaneurons, 01279, C1028, lot 102477) were ordered at FUJIFILM Cellular Dynamics. Cryopreserved non-diseased mature motor neurons (40HU-005, lot 400089) were ordered at iXCells Biotechnologies.

**Generation of AAV5-miC Vectors and Transductions**

The design and cloning of the miC constructs were performed as described previously.29,62 The miC constructs were all expressed by the synthetic cytomegalovirus (CMV) early enhancer and chicken β-actin (CAG) promoter. To produce AAV5, the CAG-miC2, CAG-miC4, CAG-miC32, and CAG-miC46 cassettes were obtained by digestion with restriction enzymes HindIII and PvuI and cloned in a uniQure transfer plasmid in order to generate an entry plasmid. The presence of the two inverted terminal repeats (ITRs) was confirmed by restriction digestion with SmaI. The ITR-CAG-miC cassettes were inserted into a recombinant baculovirus vector by homologous recombination in Spodoptera frugiperda Sf9 cells, and clones were selected by plaque purification and insert PCR. The recombinant baculovirus containing the ITR-CAG-miC was further amplified until passage 6 (P6) in Sf+ cells and screened for the best production and stability by PCR and qRT-PCR. To generate AAV5, Sf+ cells were triple infected with three different recombinant baculoviruses expressing the ITRs-CAG-miC, the replicon enzyme, and the capsid protein. The cells were lysed 72 h after the triple infection, and the crude lysate was treated with 50 U/mL Benzonase (Merck, Darmstadt, Germany) for 1 h at 37°C. AAV5 was purified on an AVB Sepharose column (GE Healthcare, Little Chalfont, UK) and eluted in a formulation buffer consisting of 1× PBS and 4% sucrose. The final titer was determined by qRT-PCR with primers amplifying a 95-bp fragment from the CAG promoter region.

For transductions with AAV, FBNs, dopaminergic neurons (DPNs), and motor neurons (MNs) were plated in 24-well plates at 0.3*10^6 cells/well. Astrocytes were plated at 0.1*10^6 cells/well in STEMdiff Astrocyte Maturation Medium (STEMCELL Technologies) on Matrigel-coated plates. FBNs were plated in STEMdiff Neuron Maturation Medium (STEMCELL Technologies) on poly-D-lysine- and laminin-coated plates. Dopaminergic neurons were plated in iPSC Neural Base Medium (FUJIFILM Cellular Dynamics), according to the manufacturer’s description on poly-D-lysine- and laminin-coated plates. Motor neurons were plated in Motor Neuron Maintenance Medium, according to the manufacturer’s description on Matrigel-coated plates. After 1 week of acclimation, cells were transduced with AAV5 for 1–2 weeks.

**RNA and DNA Isolation**

For RNA, plated cells and tissues were lysed in 300 µL TRizol. RNA was isolated from TRizol using the DIRECT-ZOL miniprep kit (R2050, Zymo Research). DNA was isolated using the DNeasy
Next-Generation Sequencing
Small RNA sequencing libraries for the Illumina sequencing platform were generated using high-quality total RNA as input and the NEXTflex Small RNA Sequencing kit (Bioo Scientific, Austin, TX, USA). Briefly, the small RNA species were subjected to ligation with 3′ and 5′ RNA adapters, first-strand reverse transcription, and PCR amplification. Sample-specific barcodes were introduced in the PCR step. The PCR products were separated on Tris/Borate/EDTA (TBE)-PAGE, and the expected band around 30 bp was obtained. The raw sequencing data were processed, removing the sequence reads that were of too low quality (only “passing filter” reads were selected). In total, we generated between 15 and 35 million reads for each sample. The resulting sequencing libraries were quantified on a BioAnalyzer (Agilent Technologies, Santa Clara, CA). The libraries were multiplexed, clustered, and sequenced on an Illumina HiSeq 2000 (TruSeq version [v.]3 chemistry), with a single-read 36-cycle sequencing protocol and indexing. The sequencing run was analyzed with the Illumina CASAVA pipeline (v.1.8.2), with demultiplexing based on sample-specific barcodes. The raw sequencing data produced were processed, recovering 30 bp was recorded for each sample. The resulting sequencing libraries were quantified on a BioAnalyzer (Agilent Technologies, Santa Clara, CA). The libraries were multiplexed, clustered, and sequenced on an Illumina HiSeq 2000 (TruSeq version [v.]3 chemistry), with a single-read 36-cycle sequencing protocol and indexing. The sequencing run was analyzed with the Illumina CASAVA pipeline (v.1.8.2), with demultiplexing based on sample-specific barcodes. The raw sequencing data produced were processed, removing the sequence reads that were of too low quality (only “passing filter” reads were selected). In total, we generated between 15 and 35 million reads per sample.

NGS Data Analysis
Next-generation sequencing (NGS) small RNA raw datasets were analyzed using the CLC Genomics Workbench 8 (QIAGEN). The obtained reads were adaptor trimmed, which decreased the average read size from ~50 to ~25 bp. All reads containing ambiguity N symbols, reads shorter than 10 nt, longer than 45 nt, and reads representing less than 10 times were discarded. Next, the obtained unique small RNA reads were aligned to the reference sequences of the pre-miC9 constructs, with a maximum of 3 nt mismatches allowed. The percentages of reads based on the total number of reads matching the reference sequence were calculated (Table S1).

qRT-PCR and miRNA TaqMan Assay
To determine C9orf72 mRNA knockdown in cells, RNA was isolated, and first-strand complementary DNA was reverse transcribed using random hexamer primers with the Dynamo kit (Finnzymes, Espoo, Finland). Real-time PCR amplification was performed with primers to detect total C9orf72 mRNA and the sense intronic transcripts of human. Total C9orf72 mRNA was detected with the following primers: mFP 5′-CGGAAAGGAAGATGATGC-3′, mRP 5′-CCATTA CAGGAATATCCTTCCA-3′, and probe mPRB 5′-AGCATGG AATAACTCTCTCCCTGATCTTC-3′. The sense intronic transcripts were detected with the following primers: SPF 5′-AAGCCTG CACAATTTCACGCCCA-3′, SRF 5′-CAAGTCTGTCATTGCATCTCG GAGCTG-3′, and probe SPRB 5′-TGAGGCGAATGCAATGCTTGGTGG-3′. The pri-mRNA expression levels were normalized to human glyceraldehyde 3-phosphate dehydrogenase (GAPDH) (forward 5′-GAAGATGGTGGATCTGAC-3′, reverse 5′-GAAGATGCCTGA TGGGATTTC-3′, and probe 5′-CAAGCTCCGTTCTCAGGC-3′) as an internal control.

PCR reaction conditions were as follows: 95°C for 10 min, followed by 40 cycles of 15 s at 95°C and 1 min at 60°C. The assays were performed on an ABI 7000 or ABI 7500 (Applied Biosystems, Foster City, CA, USA). Gene expression levels were normalized to GAPDH as an internal control, and the level of gene expression was calculated relative to control cells. To determine the expressions of miC2, miC5, miC6, and miC46, Custom TaqMan Small RNA Assay (Thermo Fisher Scientific) was used (miC2, assay ID CTETPR3R; miC4, assay ID CTFVKNN; miC32, assay ID CSGIPRB; and miC46, assay ID CSHSNXJ). The RT reaction and TaqMan were performed according to the manufacturer’s protocol.

Animals
Two breeding couples were ordered at The Jackson Laboratory (stock number 023099) and were kept and bred at the Neurosciences Division, Center for Applied Medical Research, CIMA, University of Navarra, Pamplona, Spain. The animals were housed 4–5 per cage with ad libitum access to food and water and maintained in a temperature-controlled environment on a 12-h dark-light cycle. All procedures were carried out in accordance with the current European and Spanish regulations (86/609/EEC; RD1201/2005). This study was approved by the Ethical Committee of the University of Navarra (137/010).

Intrastriatal Injection in Tg[C9orf72_3] Line 112 mice
Surgery was performed as described previously.63 In brief, 3-month-old mice were anesthetized with ketamine and xylazine (80/10 mg/kg, intraperitoneally [i.p.]), placed in a stereotactic frame. The scalp was shaved, and a longitudinal incision was made along the midline of the skull. The dorsal surface of the skull was then exposed, and two burr holes were drilled above the infusion sites. 2 or 5 µl virus suspension or PBS solution (sham mice) was infused bilaterally to striatum (+0.8 mm anterior-posterior [AP], ±2 mm medial-lateral [ML], −4.0 mm dorsal-ventral [DV] to bregma), according to the Paxinos and Watson.64 A 5-µl Hamilton syringe (or 10 for ventricle surgeries) was used for the infusion (Hamilton, Reno, NV, USA). The infusion rate was 0.2 µl/min, and the needle remained in place for 5 min after the infusion for vector absorption. Finally, the site was stitched closed.

RNA Foci FISH
RNA FISH was performed as described previously with some adjustments.23,30,56 In brief, whole mouse brains were fixed in 4% paraformaldehyde (PFA) for 1 week at 4°C. Brains were transferred into 15-mL tubes containing 10 mL 30% PBS and sucrose and left at 4°C until brains sunk to the bottom. The brains were then frozen in optimal cutting temperature (OCT) compound and 25-µM-thick cryostat sections were prepared. Brain sections were permeabilized in 0.2% Triton and 1× PBS for 10 min and incubated for 1 h in hybridization buffer (50% formamide, 10% dextran sulfate, 0.1 mg/mL yeast tRNA, 2× saline-sodium citrate (SSC), and 50 mM sodium phosphate) at 55°C. Hybridization was performed overnight with 40 nm TYE563-(C4G2)3 LNA probe in hybridization buffer at 55°C. Brain sections were then washed once with 40% formamide and 1× SSC for 30 min at 55°C, twice in 2× SSC and 0.1% Tween-20 at room temperature.
temperature for 5 min, and 3 times in 0.1 × SSC for 10 min at room temperature. The slides were mounted with ProLong Gold Antifade Mountant with DAPI (Invitrogen) and visualized using a Leica DM2500 fluorescence microscope.

**Statistical Analysis**
Data were analyzed using Student’s t test or ordinary one-way ANOVA to determine statistical significances. The p values are represented by the following: *p < 0.05, **p < 0.01, ***p < 0.001, and ****p < 0.0001.

**SUPPLEMENTAL INFORMATION**
Supplemental Information includes three figures and one table and can be found with this article online at https://doi.org/10.1016/j.omtn.2019.02.001.

**AUTHOR CONTRIBUTIONS**
Conceptualization, P.K and R.M.; Investigation, R.M., J.M.L., A.G.-O., M.C.-T., M.E., and S.U.; Supervision, P.K and R.M.; Formal Analysis, Visualization, and Writing – Initial Draft, P.K., R.M., M.M.E., and S.J.v.D.; Project Administration and Writing – Review and Editing, P.K., S.J.v.D., and H.P.; Funding Acquisition, P.K.

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