Abstract: Angiogenesis is a process of generation of de-novo blood vessels from already existing vasculature. It has a crucial role in different physiological process including wound healing, embryonic development, and tumor growth. The methods by which therapeutic drugs inhibit tumor angiogenesis are termed as anti-angiogenesis cancer therapy. Developments of angiogenic inhibiting drugs have various limitations causing a barrier for successful treatment of cancer, where angiogenesis plays an important role. In this context, investigators developed novel strategies using nanotechnological approaches that have demonstrated inherent antiangiogenic properties or used for the delivery of antiangiogenic agents in a targeted manner. In this present article, we decisively highlight the recent developments of various nanoparticles (NPs) including liposomes, lipid NPs, protein NPs, polymer NPs, inorganic NPs, viral and bio-inspired NPs for potential application in antiangiogenic cancer therapy. Additionally, the clinical perspectives, challenges of nanomedicine, and future perspectives are briefly analyzed.

Keywords: angiogenesis; anti-angiogenesis; nanomedicine; cancer; theranostics

1. Introduction

Similar to normal tissues, tumors need nourishments by means of food and oxygen as well as a capacity to remove metabolic excretes and carbon dioxide. Diverse patterns of tumor-associated neovascularization, obtained by angiogenesis, cope with these demands. Angiogenesis, sprouting of new vessels from existing quiescent ones, remains almost always turned on during the process of tumorigenesis for sustenance of neoplastic expansions [1–3]. A riveting account of studies corroborates that the “angiogenic switch” is regulated by counterbalancing factors like signaling proteins for induction (vascular endothelial growth factor (VEGF)-signaling via three receptor tyrosine kinases, etc.) or inhibition (thrombospondin-1, etc.) of angiogenesis and gets inclined towards angiogenesis when stimulated by hypoxia or inflammation [4–12]. Other pro-angiogenic signals fibroblast growth factor (FGF) family members, transforming growth factor-β, etc.), when persistently upregulated, also contribute to sustain tumor angiogenesis [5,13]. Amid all plausible ways to treat cancer, anti-angiogenic therapy—targeting tumor vasculature to prevent aberrant capillary sprouting—has received an astounding outpouring of research in the last few decades [14,15].
The idea to decouple tumors from surrounding blood vessels has led to discovery and clinical approval of several anti-angiogenic drugs, namely, monoclonal antibody inhibitors (Bevacizumab, IMC-1121B, 2C3), receptor tyrosine kinase inhibitors (sorafenib, sunitinib, pazopanib), soluble receptor chimeric protein (VEGF-Trap), inhibitors of endothelial cell proliferation (thalidomide, angiostatin), inhibitors of integrin’s proangiogenic activity (Cilengitide, medi-522), matrix metalloproteinase inhibitors (Neovastat, Prinomastat, Marimastat), vascular targeting drug (combretastatin), etc. [16–19]. However, many of these meteoric developments, alone or in combination with chemotherapy or radiotherapy, were vitiated by sparse clinical efficacy to combat tissue invasion and metastases. Drug resistance, upregulation of various proangiogenic signals, hypoxia resistance, delayed response to radiotherapy, toxicity issues, etc., have hindered the preponderance of antiangiogenic therapy [8,20–23].

To this end, nanotechnology offers an attractive biomedical platform, involving smart design of vehicles with unique physicochemical properties for targeted delivery and sustained release of therapeutics at the site of action along with their tracking details, which holds the promise to circumvent the existing limitations [13,22,24–27]. In the present review, we shall elaborately discuss the limitations of current therapies, advent of nanomedicine as an alternative modality in antiangiogenic cancer therapy including lipid-based and polymer-based nanoparticles, inorganic nanoparticles, protein and viral based nanocarriers, their safety, challenges, clinical outlook, and future perspectives.

2. Cancer, Statistics, Conventional Therapy, Challenges

Cancer is the leading cause of death throughout the world and is second to cardiovascular diseases and causes enormous health and economic burden. As per the World Health Organization (WHO) and an approximate of 9.6 million people died worldwide due to cancer in 2018 only. In 2019 alone, an estimated 1,762,450 new cancer cases were reported and approximately 606,880 cancer deaths happened in the United States. The data presented by the National Center for Health Statistics (NCHS) is promising as it shows an overall decrease in cancer date rate by 27% during the period of 1991 to 2016. Conventional cancer therapies have several limitations and associated side effects. The data presented by NCHS, WHO, and NCI suggests that early detection, and diagnosis coupled with better treatment strategies can reduce the burden of cancer [28,29]. Angiogenesis promotes tumor progression and metastasis. Metastatic disease is overwhelmingly the predominant cause of cancer death [30]. Several data suggest that angiogenesis is the prerequisite for the dissemination and establishment of metastatic tumor cells to distant organs. Hence combination therapies with emphasis on anti-angiogenic and antilymphangiogenic treatments to prevent the spread of cancer [31]. Understanding the morphology and molecular differences of the newly formed angiogenic blood vessels is critical in designing antiangiogenic therapies. Considering the success of antiangiogenic treatment and advances made in nanoparticles-based specific delivery of therapeutics to the newly formed blood vessels during neo-angiogenesis and tumor infiltration may reduce the cancer burden significantly. Considering, the recent success in immunotherapy and cancer vaccination and the close link between immune microenvironment and angiogenesis is unique. Combinations should be used to reduce the cancer burden [21]. Nanobiotechnology may play a key role in effective detection, targeting, and delivery of immune-therapeutics and antiangiogenic therapeutics at the tumor site. Nanotechnology and nanoparticles offer several advantages like increased half-life, reduced toxicity, specific and selective delivery over the free drugs/therapeutics [13,22,26]. Nanovectors have also opened up new avenues for noninvasive imaging and can be successfully used to create angiogenic maps for patient specific therapeutic planning [21].

3. Angiogenesis and Cancer

Angiogenesis is the process of formation of new blood vessels orchestrated by proangiogenic and antiangiogenic factors during development, reproduction, and repair. Of utmost importance is pathological angiogenesis, especially in the context of neoplastic disease. Tumor cells rapidly proliferate and need a continuous supply of oxygen and nutrients and thereby requires steady requirement
for infiltrating blood vessels. Angiogenesis is one of the hallmarks of cancer [32]. Targeting the angiogenic process is regarded as a logical approach to the treatment of various malignancies and various antiangiogenic treatment agents have been developed and tested in clinical trials. The concept of antiangiogenic therapy came into being while Dr. Folkman discovered that tumors require steady oxygen and nutrient supply, and the neogenesis of tumor-infiltrating vessels helps sustain the continuous growth of the tumor. The inhibition of angiogenesis can lead to tumor starvation and tumor cell death [33,34]. Angiogenesis or the formation of new blood vessels from pre-existing vasculature is the common mechanism of new angiogenesis [35]. Angiogenic blood vessel formation may occur by three major processes, sprouting angiogenesis, vasculogenesis, and intussusception. Sprouting angiogenesis is considered the most critical process of blood vessel formation in the tumor but evidence suggests that a non-angiogenic process called vessel co-option is also essential. Tumor cells opt for vessel co-option at different anatomical sites by hijacking pre-existing blood vessels from the normal neighboring issue [36,37].

Hypoxia is the key factor that regulates tumor angiogenesis. Tumor cells grow rapidly and are hypoxic in nature, and secrete vascular endothelial growth factor A (VEGFA). Not only the tumor cells but also the tumor associated stromal cells produce proangiogenic factors. The endothelial cells in the microenvironment express VEGF receptor (VEGFR2). The gradient of VEGF can be sensed by the endothelial cell based on the VEGF–VEGFR interaction; this leads to the formation of new endothelial sprout towards the tumor cell [35,38].

Tumor cell induces a complex cascade of angiogenic signaling and activates downstream cellular events in multiple cell types, especially the endothelial cells, leading to angiogenesis. Proangiogenic factors include fibroblast growth factor (FGF) families, vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), transforming growth factors-alpha/beta (TGF-α/β), and angiopoietin (Ang 1,2) and the associated receptors. The detachment of the perivascular cells from the mature blood vessels initiates vessel remodeling and endothelial cell proliferation. Platelets become activated and recruited to the sites of the exposed basement membrane. Tumor-associated macrophages (TAM) produce angiogenic factors such as VEGF, Matrix metalloproteinases (MMPs), and urokinase-type plasminogen activator (uPA). On the other hand, precursor endothelial cells move to the perceived wound site and release angiogenic factors. The activated endothelial cells release proteases and lead to extracellular matrix (ECM) remodeling, followed by directional sprouting. The signaling cascades activate tube formation and branching, followed by vessel arterio-venous patterning and maturation. Cancer progenitor/stem cells can differentiate to endothelial cells, and thereby participates directly in angiogenesis.

The process of vessels sprouting is a coordinated process including the tip/stalk cell selection, followed by tip cell directed migration, stalk cell proliferation, branching coordination, elongation of stalk, lumen formation, and vessel maturation [39]. Additional signaling molecules like delta-ligand-like 4 (DLL4) and angiopoietin 2 (ANGPT2) also play an important role in the process of angiogenesis. Until proangiogenic signals fade away, the establishment of the basement membrane, the sprouting and branching continues on, followed by vessel maturation. In cancer, however, the angiogenesis process is rapid but considerable variation exists in the vessel morphology, functionality, and integrity [40]. Figure 1 is an overall presentation of angiogenic signaling pathway and angiogenesis.
Figure 1. Angiogenic signaling pathway and angiogenesis. Tumor cell induces a complex cascade of angiogenic signaling and activates downstream cellular events in multiple cell types, especially the endothelial cells, leading to angiogenesis. Pro-angiogenic factors include fibroblast growth factor (FGF) families, vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), transforming growth factors-alpha/beta (TGF-α/β), and angiopoietin (Ang 1,2) and the associated receptors. The detachment of the perivascular cells from the mature blood vessels initiates vessel remodeling and endothelial cell proliferation. Platelets become activated and recruited to the sites of the exposed basement membrane. Tumor-associated macrophages (TAM) produce angiogenic factors such as VEGF, MMPs and urokinase-type plasminogen activator (uPA). On the other hand, precursor endothelial cells move to the perceived wound site and release angiogenic factors. The activated endothelial cells release proteases and lead to extracellular matrix (ECM) remodeling, followed by directional sprouting. The signaling cascades activate tube formation and branching, followed by vessel arterio-venous patterning and maturation. Cancer progenitor/stem cells can differentiate to endothelial cells, and thereby participates directly in angiogenesis.

4. Current Antiangiogenic Therapies in Cancer and Their Limitations

Angiogenesis inhibitors are designed to target existing tumor infiltrating blood vessels and inhibit the formation of newly formed blood vessels and thereby halt the tumor metabolism and growth. Based on the inherent nature of the neo-angiogenesis in primary tumors, angiogenesis inhibitors might activate the destruction of immature angiogenic vessels or stop the angiogenic switch thereby preventing vascular metastasis [13]. Angiogenic inhibitors in conjunction with combination chemotherapy, may increase the antitumor potency to a significant level as these agents can halt the tumor progression but not eliminate them. There are two types of angiogenesis inhibitors; the first group includes direct inhibitors, agents that block main angiogenesis proteins. The second group comprises indirect inhibitors that target the tumor cell or stromal cells and modulate angiogenesis regulators [41]. The direct angiogenesis inhibitors exert their antiangiogenic effect by binding with angiogenesis inducers like VEGF, bFGF, and PDGF. Bevacizumab was the first antiangiogenic therapy that was approved by the FDA in 2004 for the treatment of colorectal cancer. Antiangiogenic treatment is the fourth modality of cancer therapy, as the treatment targets microvascular endothelial cells in the tumor microenvironment and not the tumor directly [13,42]. Fast-growing tumors need greater microvasculature and are more susceptible to antiangiogenic therapy. Table 1 shows the balance between pro and antiangiogenic factors is the key to antiangiogenic treatment. Table 1 shows the chemical structure and the mechanism of action of a list of FDA approved angiogenesis inhibitors for different cancers. Several clinical trials are in progress that can induce modest disease-free survival advantages. Table 1 also shows a list of potential drugs that have demonstrated antiangiogenic properties in preclinical trials.
Table 1. FDA approved angiogenic inhibitors, trade name, chemical structures, target, and FDA and approved treatments.

| Drug (Trade Name) | Structure | Chemical Name, Target, and FDA Approved to Treat Patients with | Ref. |
|------------------|-----------|---------------------------------------------------------------|------|
| Bevacizumab (Avastin®) | Anti-VEGF monoclonal antibody | Anti-VEGF monoclonal antibody Cervical cancer: Nonresponsive to other treatment/metastatic/recurrent. Colorectal cancer: metastatic. Glioblastoma: Nonresponsive to other treatment/recurrent. Nonsquamous non-small cell lung cancer: locally advanced, nonresectable/metastatic/recurrent. Ovarian epithelial, fallopian tube, or primary peritoneal cancer: stage III/stage IV/recurrent. Renal cell carcinoma: metastatic. | [13,43–45] |
| Thalidomide (Synovir, Thalomid®) | (±)-2-(2,6-Dioxo-3-piperidinyl)-1H-isoindole-1,3(2H)-dione | Immune modulator and inhibits VEGF and bFGF production Multiple myeloma: newly diagnosed. | [46] |
| Lenalidomide (Revlimid®) | 1-oxo-2-(2,6-dioxopiperidin-3-yl)-4-aminoisoindoline | VEGF-induced PI3K-Akt pathway signaling and HIF-1α expression Anemia associated with certain types of myelodysplastic syndromes. Follicular lymphoma: Nonresponsive to other treatment. Mantle cell lymphoma: Nonresponsive to other treatment/recurrent. Marginal zone lymphoma: Nonresponsive to other treatment. Multiple myeloma and as maintenance therapy | [13,42,47] |
| Sorafenib (Nexavar®) | 4-[4-[[4-chloro-3-(trifluoromethyl)phenyl]carbamoyl]amino phenoxyl]-N-methylpyridine-2-carboxamide | Small molecule inhibitors of the VEGFR-2 tyrosine kinase activity Hepatocellular carcinoma: Nonresectable. Renal cell carcinoma: Advanced. Thyroid cancer: Progressive/metastatic/recurrent. | [41,48] |
| Drug (Trade Name)         | Structure | Chemical Name, Target, and FDA Approved to Treat Patients with                                                                 | Ref.          |
|--------------------------|-----------|----------------------------------------------------------------------------------------------------------------------------------|--------------|
| Sunitinib (Sutent®)      | ![Structure](image1) | (Z)-N-(2-(diethylamino)ethyl)-5-((5-fluoro-2-oxoindolin-3-ylidene)methyl)-2,4-dimethyl-1H-pyrrole-3-carboxamide Small molecule inhibitors of the VEGFR-2 tyrosine kinase Gastrointestinal stromal tumor: nonresponsive to imatinibmesylate. Pancreatic cancer: progressive neuroendocrine tumors that are nonresectable/metastatic. Renal cell carcinoma: advanced disease. | [13,41,42,49] |
| Temsirolimus             | ![Structure](image2) | 42-[3-Hydroxy-2-(hydroxymethyl)-2-methylpropanoate]-rapamycin Reduces synthesis of VEGF and targets the mammalian target of rapamycin (mTOR) Retinoblastoma. Renal cell carcinoma: advanced disease. | [50]         |
| Axitinib (Inlyta®)       | ![Structure](image3) | N-Methyl-2-((3-((1E)-2-(pyridin-2-yl)ethenyl)-1H-indazol-6-yl)sulfanyl)benzamide Inhibitor of VEGF-1, -2, and -3 Renal cell carcinoma: Advanced/nonresponsive to other treatment. | [13,42,51]   |
Table 1. Cont.

| Drug (Trade Name) | Structure | Chemical Name, Target, and FDA Approved to Treat Patients with | Ref. |
|-------------------|-----------|---------------------------------------------------------------|------|
| **Pazopanib (Votrient®)** | ![Structure](image1.png) | 5-(((2,3-dimethyl-2H-indazol-6-yl)(methyl)amino)pyrimidin-2-yl)(methyl)amino)-2-methylbenzenesulfonamide  
Small molecule multi-targeted receptor tyrosine kinase inhibitor  
Renal cell carcinoma: Advanced. Soft tissue sarcoma: Advanced. Nonresponsive to other treatment. | [13,52] |
| **Lenvatinibmesylate (Lenvima®)** | ![Structure](image2.png) | 4-(3-chloro-4-(3-cyclopropylureido)phenoxy)-7-methoxyquinoline-6-carboxamide methane sulfonate  
Lenvatinib inhibits tyrosine kinase activity of VEGF1, 2 and 3, fibroblast growth factor receptors (FGFRs) 1–4  
Endometrial carcinoma: Advanced/nonresponsive to other treatment.  
Hepatocellular carcinoma: first-line treatment in nonresectable tumor.  
Renal cell carcinoma: Advanced. Thyroid cancer: Progressive/recurrent/metastatic/nonresponsive to radioactive iodine treatment. | [53] |
| **Cabozantinib (Cometriq®)** | ![Structure](image3.png) | 1,1-cyclopropanedicarboxamide,  
n'-[4-[[6,7-dimethoxy-4-quinolinyl]oxy]phenyl]-n-(4-fluorophenyl)-c-MET and VEGFR2 Inhibitor  
Hepatocellular carcinoma: already been treated with sorafenib. Medullary thyroid cancer: Progressive/metastatic. Renal cell carcinoma: Advanced. | [41,54] |
| Drug (Trade Name)          | Structure | Chemical Name, Target, and FDA Approved to Treat Patients with                                                                                   | Ref. |
|---------------------------|-----------|-------------------------------------------------------------------------------------------------------------------------------------------------|------|
| Everolimus (Afinitor®)    | ![Structure of Everolimus](image1) | (1R,9S,12S,15R,16E,18R,19R,21R,23S,24E,26E,28E,30S,35R)-1,18-dihydroxy-12-((2R)-1-[(1S,3R,4R)-4-(2-hydroxyethoxy)-3-methoxycyclohexyl]propan-2-yl)-19,30-dimethoxy-15,17,21,23,29,35-hexamethyl-11,36-dioxo-4-azatricyclo[3.0.3.0{4,9}]hexatriaconta-16,24,26,28-tetraene-2,3,10,14,20-pentone40-O-(2-hydroxyethyl)-rapamycin Immunosuppression and targets the mTOR pathway Breast cancer: Advanced hormone receptor–positive (HR+) breast cancer that is also HER2 negative. Pancreatic cancer, gastrointestinal cancer, and lung cancer: Neuroendocrine tumors/nonresectable/metastatic. Renal cell carcinoma: Advanced. Subependymal giant cell astrocytoma: Nonresectable. | [13,41,55] |
| Vandetanib (Caprelsa®)    | ![Structure of Vandetanib](image2) | (4-Bromo-2-fluoro-phenyl)-[6-methoxy-7-(1-methyl-piperidin-4-ylmethoxy)-quinazolin-4-yl]-amine Dual Inhibitor of VEGFR and Epidermal Growth Factor Receptor (EGFR) Tyrosine Kinases and also inhibits the mTOR–HIF-1 alpha–VEGF signaling axis Medullary thyroid cancer: Nonresectable/metastatic. | [13,41,42,56] |
| Ramucirumab (Cyramza®)    | ![Structure of Ramucirumab](image3) | Anti-VEGFR2 monoclonal antibody Colorectal cancer: Metastatic/nonresponsive to other treatment like bevacizumab, oxaliplatin, and fluoropyrimidine. Hepatocellular carcinoma: Nonresponsive to sorafenib. Non-small cell lung cancer: Metastatic/aggravated after platinum chemotherapy/with a mutation in the EGFR gene or ALK gene. Stomach adenocarcinoma or gastroesophageal junction adenocarcinoma: Advanced/metastatic | [41,57] |
Table 1. Cont.

| Drug (Trade Name)          | Structure | Chemical Name, Target, and FDA Approved to Treat Patients with                                                                 | Ref. |
|----------------------------|-----------|---------------------------------------------------------------------------------------------------------------------------------|------|
| Regorafenib (Stivarga®)    | ![Structural formula](image) | 4-(4-(3-(4-Chloro-3-(trifluoromethyl)phenyl)ureido)-3-fluorophenoxy)-N-methylpicolinamide. Dual targeted VEGFR2-TIE2 tyrosine kinase inhibition. Colorectal cancer: Metastatic/nonresponsive to other treatment. Gastrointestinal stromal tumor: Advanced/nonresectable/metastatic/nonresponsive to imatinibmesylate and sunitinib malate. Hepatocellular carcinoma: Nonresponsive to sorafenib. | [41,58] |
| Ziv-aflibercept (Zaltrap®) | ![Structural formula](image) | A recombinant fusion protein comprising the extracellular domains of human VEGF receptors 1 and 2 Inhibitor of VEGF Colorectal cancer: Metastatic/nonresponsive to other treatment. | [41,59] |
Although the concept of targeting tumor angiogenesis has opened up new possibilities to treat cancer, many limitations need to be addressed to make this therapy successful. Toxicity is one of the main drawbacks of antiangiogenic treatment, the main side effects being hemorrhage, hypertension proteinuria, thrombosis, and poor wound healing. Preclinical data have suggested that the medication affects not only the tumor vasculature but also the vasculature in multiple organs, especially vasculature rich organs. Optimal dose calculation, especially in combination chemotherapy, is very challenging. Current decisions in chemotherapy management do not consider inter-tumoral vessel vasculature to decide the dose and type of antiangiogenic therapy [60,61]. Another drawback is the development of acquired resistance to antiangiogenic therapy, causing transient disease stabilization and activation of alternative pathways, vessel co-option, vessel mimicry, and enhanced metastasis. Cancer cells can develop resistance to antiangiogenic treatment by multiple mechanisms [61]. Cancer cells upregulate proangiogenic factors like angiopoietins (Ang), epidermal growth factor (EGF), fibroblast growth factor (FGF), interleukin 8 (IL-8) etc., to activate compensatory pathways to stimulate blood vessel formation [60,61]. Activation of hypoxia inducing factor (Hif1) and downstream ZEB2 are the alternative angiogenic pathway that can be targeted.

Cancer cells also secrete several growth factors that recruit cells that ultimately cause resistance to therapy. Bone marrow-derived cells, monocytes, macrophages, endothelial precursor cells, myeloid-derived suppressor cells, and cancer-associated fibroblasts can infiltrate and produce proangiogenic factors which might lead to VEGF-independent new blood vessel formation. The association between the immune system and angiogenesis is not well understood and is a bottleneck in antiangiogenic therapy. Vascular heterogeneity, especially in tumor vessels, is quite remarkable, leading to a variable degree of pericyte coverage and VEGF expression; vessel leakage can thereby lead to altered anti-VEGF response [62,63]. Non-angiogenic cancers exploit the process of vessel co-option (by hijacking pre-existing vessels) and thereby cause resistance to antiangiogenic therapy as evident from the results of several clinical trials [63]. The other potential mechanism operations can be extracellular vesicles, post-translational modification, genetic polymorphism. Therefore, the knowledge of acquired resistance will help design theranostic drugs.

5. Alternative Therapy: Nanomedicine

In this context, nanomedicine plays an important role to overcome the existing limitations of present antiangiogenic therapy due to their interesting physicochemical properties (small size and high surface area) at nanoscale. Recently, many investigators as well as our group exhibited various, multifunctional theranostics applications of nanomedicine in different diseases including cancer, diabetes, neurodegenerative disease, cardiovascular diseases, antibacterial, spinal cord injury, etc. [23,25,64–66]. Nanoparticles conjugated to various targeting ligands can be employed to utilize active targeting of antiangiogenic drugs for better therapeutic efficacy. Additionally, various reports showed the anti and proangiogenic properties of several inorganic nanoparticles (NPs) including silver NPs (AgNPs), gold NPs (AuNPs), copper nanoparticles (CuNPs), carbon nanotubes (CNT), europium hydroxide nanorods (EHNs), graphene oxides (GO), zinc oxide nanoflowers, and cerium oxide nanoparticles (NCe) [13,22,67,68]. Moreover, various other nanomaterials including liposomes, lipid NPs, protein NPs, polymer NPs, viral and bio-inspires NPs are utilized for the targeted delivery of antiangiogenic agents to tumors for the suppression of tumor angiogenesis [26,69–72] (Figure 2). Active targeting of antiangiogenic drugs also help to reduce the unwanted side effects and toxicity.
Figure 2. Schematic representation of multiple nanomedicine approaches that may be used for the diagnosis and treatment of angiogenesis.

6. Lipid-Based Nanoparticles for Antiangiogenic Therapy

Amid a variety of nanocarriers, lipid-based systems such as liposomes, solid lipid NPs, lipid-polymer hybrid NPs, and nanostructured lipid carriers are most widely explored for cancer therapeutics including antiangiogenic therapeutics, as lipid-based vehicles have advantages to solubilize insoluble drugs, encapsulate multiple hydrophobic and hydrophilic drugs, and deliver them at specific tissue sites to accomplish enhanced bioavailability while avoiding off-target side-effects. In this section, we will highlight the recent advancements in lipid-based NPs in antiangiogenic therapy. As discussed earlier, VEGF and its receptors (VEGFRs) play a crucial role in angiogenesis and proliferation of many type of cancer cells, including melanoma, breast, lung, and brain cancer [73]. Downregulation of VEGF expression or inhibiting its receptors activity through various methods, thus, was proposed to efficiently suppress tumor angiogenesis with simultaneous tumor growth inhibition (in combination with some other chemotherapeutics). Recently, Prof. Leaf Huang’s laboratory developed a polymetformin (PolyMet) containing hyaluronic acid decorated lipid NPs for systemic gene delivery. PolyMet NPs were shown to be highly capable of VEGF siRNA delivery for VEGF knockdown in a human lung cancer xenograft, leading to enhanced tumor activity by inhibiting angiogenesis [74] (Figure 3). The same group also developed lipid-based dual functionalized NPs to VEGF siRNA in vivo [75]. Recently, Yang and his colleagues developed a low-density lipoprotein receptor-related protein receptor (Angiopep-2) and neuropilin-1 receptor (tLyP-1) targeting cationic liposomes for delivery of siRNA and docetaxel to gliomas. The dual peptide-decorated liposomes showed efficacious antiangiogenic activity by knocking down VEGF siRNA along with antiproliferative apoptotic effects exerted by docetaxel in U87 MG tumors [76]. Dr. Wang’s laboratory developed a miRNA loaded cRGD-functionalized lipid NPs for antiangiogenic therapy [77]. Another report showed promising VEGF inhibition results by
using dual receptor targeting liposomes [78]. Guo and his team demonstrated that Lcn2 siRNA loaded ICAM-1-targeting liposomes showed a potent antiangiogenic effect in triple-negative breast cancer (TNBC) [79]. Several other research groups also developed similar lipid-based NPs for gene-mediated angiogenesis inhibition [80–87]. Sorafenib inhibits angiogenesis and proliferation via binding to VEGFR-2, VEGFR-3, and PDGFR-b tyrosine kinases. The clinical trials of sorafenib are suggesting that it displays a high antiangiogenic efficacy in several cancers and it also strengthens the efficacy of other chemotherapeutics. However, very poor water solubility and off-target side effects of sorafenib limit its clinical usage. Scientists are using nanotechnology tools to overcome these issues. For example, Menei and his team developed sorafenib-encapsulated lipid NPs for the treatment of glioblastoma, which showed high efficiency in the suppression of angiogenesis by inhibiting CD31 [88]. More recently, Zang and his group demonstrated that co-delivery of VEGF-siRNA and Sorafenib through pH-sensitive liposomes showed a synergistic effect in hepatocellular carcinoma [89]. Several other lipid-based NPs, along with other chemotherapeutics, are also being used for better antiangiogenic and antitumor therapy [90,91]. mTOR inhibitor like rapamycin and its analogs also proved their antiangiogenic capability. Many studies confirmed that these analogs inhibit the expression of VEGF in tumor cells. Recently, several studies elicited that rapamycin and its analogs, loaded as single and in combination with other drugs onto lipid-based NPs, could be used for effective antiangiogenic therapy [92–95].

**Figure 3.** Lipid nanoparticles made up of PolyMet can systemically deliver vascular endothelial growth factor (VEGF)silRNA to the tumor site and inhibit tumor growth. (a) H460 tumor-bearing mice were injected i.v. every other day and tumor volumes were measured every day. (b) H460 tumor VEGF protein levels after two injections were measured by Western blot analysis. The bar chart in (b) represents the quantitative analysis of relative normalized VEGF band intensity (Image J). Data are mean ± s.e.m. (n = 5 per group) analyzed by two-way analysis of variance with Tukey’s correction. Data are representative of (b) or combined from (a) three independent experiments. NS, not significant; * p < 0.05, ** p < 0.01, *** p < 0.005. Reproduced with permission from [74]. Copyright, 2016, NPG.

There is a piece of evidence that somatostatin receptors (SSTRs), mainly subtype 2 (SSTR2), are significantly expressed in both glioma and glioma vasculature endothelial cells. Recently, Misra’s lab developed paclitaxel (PTX) loaded solid lipid NPs (SLN) functionalized with Tyr-3-octreotide (ligand for SSTR2) to facilitate dual-targeted chemotherapy by targeting both brain tumor and tumor neovasculature cells. The study demonstrated excellent tumor growth inhibition and enhanced survival by an antiangiogenic (CD31 inhibition) and antitumor effect of PTX in orthotopic glioma-bearing...
rats. Additionally, the authors studied tumor vasculature and tumor targeting efficiency of NPs by conjugating $^{99}$mTc [96]. In another recent study, the authors demonstrated significant suppression of angiogenesis by targeting oxaliplatin loaded PEGylated cationic liposomes in a dorsal air sac mouse model [97]. Earlier this century, Sengupta et al. [98] and Ebos et al. [20] developed polymer lipid hybrid nanocarriers for delivery of combretastatin (an anti-angiogenesis drug) along with doxorubicin as a chemotherapeutic. In summary, there is an enormous amount of progress observed in lipid-based antiangiogenics.

7. Polymeric Nanomedicine

Among all the commonly used biodegradable materials, polymers offer a superior advantage in the drug delivery field for tumor angiogenesis. Poly (lactic-co-glycolic acid) (PLGA) is a widely used, FDA approved biocompatible polymer, which offers a versatile platform to load multiple hydrophobic and hydrophilic small molecule drugs or in combination using various emulsion procedures [99,100].

After Judah Folkman unequivocally enunciated the “angiogenic switch” hypothesis for tumor progression in 1991, angiogenesis has become an important component of tumor growth and development and there has been an incredible rush in targeting angiogenesis for cancer therapeutics [101]. Therefore, there is an urgent need for efficient angiogenesis inhibitors development. O-(chloracetyl-carbamoyl) fumagillol (TNP-470, angiogenesis inhibitor) reduced tumor growth in patients with metastatic cancer. However, at required higher doses, many patients experienced neurotoxicity. To overcome this, Folkman and his team developed a water-soluble TNP-470 conjugated 2-Hydroxypropyl methacrylamide (HPMA) copolymer and nanopolymeric micelles (Lodamin). These formulations demonstrated beneficial drug delivery features, such as prolonged systemic circulation half-life, targeting capabilities, controlled drug release, and used as oral nontoxic antiangiogenic drugs [102,103].

Importantly, as shown in Figure 4, TNP-470 conjugated HPMA copolymer significantly inhibited A2058 human melanoma and Lewis lung carcinoma (LLC) tumor growth which suggesting compelling future antiangiogenic and anticancer treatment options for patients [102]. In another study, Harfouche et al. described LY294002 loaded PLGA nanoparticles, which can efficiently inhibit melanoma tumor growth by inducing apoptosis in zebrafish tumors [104]. A combination of chemo- and anti-angiogenesis therapy holds immense potential for effective tumor growth inhibition. For example, Yao and his group developed heparin–gambogic acid-containing and c(RGDyK)-functionalized self-assembled polymeric amphiphilic nanosystem. This formulation showed considerable inhibition of VEGF, hypoxia inducible factor-1 alpha, and CD31 expression with significant downregulation of pVEGFR2. These results offer a versatile nanoplatform for efficient combinatorial tumor therapy [105]. In a similar study, nanopolymer was developed for targeted co-delivery of multiple anticancer and antiangiogenic agents using LyP-1 peptide as a targeting ligand [106]. Later on, several other hybrid polymers have been developed for antiangiogenic therapy; for example, mitomycin C and doxorubicin co-encapsulated polymeric.
Nanoparticles exhibited superior anti-angiogenesis and antitumor activity with minimal systemic toxicity in both sensitive and drug-resistant orthotopic xenografts of breast cancer [107]. Lung metastasis is one of the primary causes of mortality with no cure available currently. The dual-treatment options, such as, targeting anticancer and anti-angiogenesis agents may offer some advantages. Recently the same group developed a similar approach using RGD peptide as a targeting moiety and demonstrated significant inhibition of the lung metastasis progression and extended median survival [108]. As shown in Figure 5, Chen and coworkers developed a poly(L-glutamic acid)-CA4 containing polymeric NPs for selective disruption of unusual tumor vasculature, in addition to elevating the hypoxia level of the tumor microenvironment to further boost up the antitumor ability of Tirapazamine in metastatic tumors [109]. Additionally, developments in gene therapy for antiangiogenic cancer therapy has become more attractive [110]. For example, therapeutic gene combinations of siMyc, siVEGF, and siBcl-2 and an imaging agent containing poly(D,L-lactide-co-glycolide) (PLG) multi-model nanomaterials functionalized with rabies virus glycoprotein (RVG) peptide for neuroblastoma-targeting delivery showed potential antitumor efficacy in a neuroblastoma mouse model [111]. A similar study was performed using block catiomer of poly (ethylene glycol)
(PEG)-b-poly[N'-(2-[aminoethyl]-2-aminoethyl]aspartamide]-cholesteryl conjugated with RGD and an antiangiogenic gene which demonstrated promising results [112]. For interested readers, we provide a few more reviews describing recent advances of nanotherapeutic-based cancer starvation therapy, challenges, and future prospects of these anticancer strategies [21,71,113–115].

Figure 5. Schematic illustration of hypoxia-inducing vascular disrupting agents (VDA) nanodrug combined with hypoxia-activated prodrug for cancer therapy (A). Tumor volume changes of BALB/c mice bearing 4T1 tumors with both moderate sizes (≈180 mm$^3$) ($n=6$) (B) and large sizes (≈500 mm$^3$) ($n=6$). (C). All data points are presented as mean ± standard deviation (s.d.) (*$p<0.05$, **$p<0.01$, ***$p<0.001$). Reproduced with permission from [109]. Copyright, 2019, Wiley-VCH.

8. Inorganic Nanoparticles

Inorganic nanoparticles including AuNPs, AgNPs, CNTs, PtNPs, CuNPs, ZnO, and Fe$_3$O$_4$, etc., have gained immense attention due to their multifunctional properties, easy synthesis, easy functionalization, and inherent pro and/or antiangiogenic properties [22,25,26]. Several investigations, including ours, demonstrated the antiangiogenic properties of various inorganic nanomaterials in cancer therapy in recent times [13,22,116,117].

Mukherjee et al. demonstrated that 5 nm sized AuNPs inhibited the function of VEGF165 (HBGF) demonstrating anti-angiogenesis [67]. Balakrishnan and co-workers reported the use of chemically synthesized gold nanoparticle conjugated with naturally available photochemical quercetin
for the inhibition of tumor angiogenesis, epithelial–mesenchymal transition, and tumor metastasis via EGFR/VEGFR-2-controlled pathway in vitro and in vivo breast cancer (Figure 6) [118]. Pan et al. showed the antiangiogenesis and tumor inhibition of AuNPs by inhibiting VEGF165-influenced VEGFR2 and phosphorylation of AKT pathways in mouse xenograft models [119]. Gurunathan et al. exhibited the antiangiogenic properties of biosynthesized silver nanoparticles using Bacillus licheniformis [120] observed by several in vitro (cell proliferation, tube formation, migration etc.) and in vivo (Matrigel plug assay) models. Song et al. showed the inhibition of angiogenesis using CuNPs causing inhibition of HUVEC migration, tube formation, and cell cycle arrest at various doses of treatment [121]. Giri et al. reported the use of nanoceria (NCe) nanoparticles for the inhibition of ovarian cancer growth in an in vivo mouse model that caused activation of MMP2 apart from inhibition of endothelial cell proliferation and migration [122]. In another recent report, Setyawati et al. demonstrated the antiangiogenic and antitumor activities of mesoporous silica nanoparticles in a size-dependent manner causing generation of ROS further activating the tumor suppressing p53 signaling pathways [123].

Figure 6. Effect of free Qu and AuNPs-Qu-5 on DMBA-induced mammary carcinoma in Sprague-Dawley rats. AuNPs-Qu-5 inhibited the DMBA-induced tumor growth in Sprague-Dawley rats:
(a) representative photographs of breast tumors in each group; (b) weight of breast tumors in each group; (c) body weight of all the animals in each group. “a” DMBA alone vs. others; “b” DMBA induced animals + free AuNPs vs. others; and “c” DMBA induced animals + free Qu. (d) Effect of free Qu and AuNPs-Qu-5 on histopathological examination. (d) Histopathological examination of DMBA-induced breast cancer in Sprague-Dawley rats. Histopathological changes in the mammary tissues of cancer-induced vehicle and experimental animals (hematoxylin and eosin, 10×). L1: Cancer-induced breast cancer animals show the extensive solid areas and several neoplastic cells lobular structural disruption; L2: CI + free AuNPs show extensive solid tumors; L3: free quercetin-treated animal shows a small amount of neoplastic structure; L4: AuNPs-Qu-5-treated animal shows normal mammary epithelial cells appearance.*Represents neoplastic cells.

Mukherjee et al. showed the dose-dependent manipulation of anti-angiogenesis and angiogenesis using the treatment of graphene oxides by the modulation of ROS in in vitro (endothelial cell proliferation assay, scratching assay, and tube formation assay) and ex vivo models (chicken embryonic angiogenesis assay) [72].

9. Protein Based Nanoparticles

Protein-based NPs have also attracted substantial attention owing to their high biodegradability, highly symmetrically organized edifices, ideal size for delivery, ability for different interfaces functionalization, etc. [124–127]. For example, as shown in Figure 7, Lin et al. developed albumin based NP encapsulating paclitaxel and 4-HPR (angioprevention vitamin A analog) functionalized with blood brain barrier (BBB) crossing targeting peptide. These dual drug-loaded NPs showed excellent anti-glioma efficacy on the subcutaneous glioma mouse model by inhibiting angiogenesis, regulating tumor immune microenvironment, and inducing apoptosis [128]. Itraconazole (ITA) is originally an antifungal drug, but in recent years, it is being used in cancer as a multitarget anti-angiogenesis agent [129]. ITA inhibits several angiogenic pathways, including VEGF, VEGFR-2, FGF, etc. [129]. However, its poor water solubility is obstructing its usage as an effective antiangiogenic drug. Recently, Zhang and his team developed an ITA encapsulated BSA-NP for effective antiangiogenic and antitumor therapy on a patient-derived xenograft NSCLC model [129]. Rapamycin is one of the best angiogenic inhibitors that blocks angiogenesis by inhibiting downstream signals such as mTOR. Nevertheless, its reduced chemical stability, poor water solubility, and significant side effects limit its usage. Desai and his team employed albumin-based NP to overcome all issues related to rapamycin and demonstrated superior antiangiogenic and antitumor efficacy using nab-rapamycin in combination with nab-paclitaxel in human colon and breast cancer xenograft models. In another study, researchers developed PEG-modified gelatin-based nanovectors for sFlt-1 plasmid DNA delivery to solid tumor xenograft of breast cancer [130]. In summary, protein-based NPs provide a large contribution to antiangiogenic cancer therapy.
Figure 7. Antiglioma efficacy on the subcutaneous glioma mouse model. (A) Tumor growth curve. (B) Tumor weight and tumor inhibition rate. (C) Representative tumor tissues. (D) Bodyweight variations in the treatment course. (E) Organ coefficients (* \( p < 0.05 \), ** \( p < 0.01 \)). Reproduced with permission from [128]. Copyright, 2016, ACS.

10. Viral and Other Bio-Inspired Nanoparticles

During prior decades, the understanding of viral-based nanotechnology has improved, simultaneous with development in their design and production [131,132]. Among various virus-based nanoparticles (VNPs), plant-derived VNPs are considered safe from a human health perspective because they are not pathogenic in mammals and proteinaceous plant VNPs have appeared as a key platform for engineering with multiple drugs, imaging molecules, and targeting ligands [133,134]. VNPs emanated in various sizes, shapes, and each virus species is highly symmetrical and monodisperse. Numerous VNPs including plant viruses including Cowpea mosaic virus (CPMV), the Human papillomavirus (HPV), Brome mosaic virus (BMV), Cowpea chlorotic mottle virus (CCMV), Red clover necrotic mottle virus (RCNMV) and Hibiscus chlorotic ringspot virus (HCSR) are materialized for various nanomedical applications [134,135]. Recently, VNPs are also being used in antiangiogenic therapy. For example, in 2019, Gamper et al. developed a nanocarrier containing coat protein (CP) of Tobacco mosaic virus (TMV) fused with a highly hydrophobic, insoluble peptide that targets the transmembrane domain of Neuropilin-1 (NRPI) receptor in cancer cells. The virus conjugated nanopeptide inhibited angiogenesis and cell migration by disrupting the NRPI-PlexA1 complex and downstream [136]. In another study, Dawson and co-workers developed Cowpea mosaic virus (CPMV) NPs fused with a fluorescent PEGylated peptide and VEGFR-1 ligand for tumor targeting and imaging. These NPs
showed a high affinity towards VEGFR-1 on endothelial cell lines and VEGFR1-expressing tumor xenografts in mice [137]. The vimentin overexpression in tumor endothelium shows a relationship with the CPMV uptake in tumor endothelial cells, as revealed in studies using the chick chorioallantoic membrane tumor model. The fluorescent CPMV sensors allowed for the visualization of the flow of blood and further exploited in tumor angiogenesis imaging [138]. The use of CPMV as a natural endothelial probe in imaging vascular disease may provide novel insights into the expression pattern of surface vimentin [138,139]. Similarly, a fluorescent plant VNPs with specific targeting ligands TMV-BF3 were developed for intravital imaging of the mouse brain vasculature (Figure 8). Further, these VNPs were used for delivery of therapeutic agents such as drugs or peptides and may lead to the development of novel cost-effective tools for in vivo theranostics [140].

Regardless of the enormous development in VEGF mediated antiangiogenic therapies available for therapeutic use, clinical evidence is escalating to recommend that targeting only VEGF may not be effective in inhibiting tumor angiogenesis. The epidermal growth factor-like domain 7 protein (EGFL7, 30 kDa) is only expressed by vascular remodeling endothelial cells and identified as a key controller of various angiogenic pathways [141]. Recently, researchers developed EGFL7 ligand decorated CPMV viral nanoparticles for intravital imaging of tumor neovasculation [141]. They further used $^{68}$Ga-labeled E7p72 radiotracer for in vivo targeting PET imaging. These studies suggest that using EGFL7 expression as a biomarker for tumor angiogenesis would be beneficial since it is expressed in cells associated with tumor blood cell remodeling and not by mature blood vessels [134,141]. Several other nanoparticles for antiangiogenic therapy were discussed in detail in the recent literature cited [13,21,22,142–146].

We have tabulated the recent examples of various nanomaterials used for the antiangiogenic therapy (Table 2).
Table 2. Various nanomaterials and their anti-angiogenesis applications.

| S.No | Nanoparticle          | Therapeutics                        | Application                                                                                           | Ref        |
|------|-----------------------|-------------------------------------|-------------------------------------------------------------------------------------------------------|------------|
| 1    | Liposomes             | Honokiol (potent anti-angiogenesis agent) | Liposomal honokiol improved efficacy of radiotherapy and chemotherapy in lung andovarian tumors.       | [147–149] |
| 2    | Liposomes             | Gd-DTPA Rhodamine PE                | Gd-RGD-liposomes for target-specific MRI imaging and therapy of tumor angiogenesis.                   | [150]      |
| 3    | Liposomes             | Anginex-peptide                     | Anginex-liposomes used imaging for the angiogenesis-dependent disease.                                 | [151]      |
| 4    | Liposomes             | Everolimus (mTOR) EG00229 (VEGF/NRP1) | Showed effective tumor growth inhibition in a highly aggressive syngeneic immune-competent mouse model. | [152]      |
| 5    | Solid-lipid nanoparticle | Bevacizumab | BSLNPs showed highly more effective than the parent in glioblastoma.                                 | [153]      |
| 6    | Liposomes             | Fenretinide                          | Fenretinide–liposomes showed enhanced antiangiogenic and antitumor activity on human neuroblastoma.    | [154]      |
| 7    | Liposomes             | ALK-siRNA                            | ALKsiRNA loaded liposomes induce apoptosis and inhibit angiogenesis.                                   | [155]      |
| 8    | Liposomes             | Clodronate                           | Clo-liposomes efficiently deplete tumor-associated macrophages and showed antiangiogenic and antitumor effects in primary and metastatic melanoma. | [156]      |
| 9    | Gold nanoparticles    | Recombinant human endostatin (antiangiogenic agent) | Endostatin-gold nanoparticles normalized vessels in metastatic colorectal cancer.                   | [116]      |
| 10   | Gold nanoparticles    | GNP s                               | Gold nanoparticles inhibit subsequent angiogenesis-related signaling events.                          | [67]       |
| 11   | Gold nanoparticles    | Quercetin                            | Quercetin-GNP s inhibits EMT, angiogenesis and invasiveness in cancer.                                | [118,157–160] |
| 12   | Gold NPs              | Peptides                             | Inhibit angiogenesis.                                                                                  | [161–163] |
| 13   | Nanoparticles         | Small molecules                      | Inhibits tumor angiogenesis and tumor growth.                                                         | [99,142,143,164,165] |
| 14   | Lipid conjugates      | PTX/LGC                             | IRGD-nanoconjugates improve tumor vessel normalization to achieve optimal chemo drug delivery into solid tumors. | [144]      |
| 15   | PLA -NPs              | Delta-like ligand 4 (DII4-GD16-PTX) | GD16-PTX-NP demonstrated significant antiangiogenic and anticancer activity.                          | [166]      |
| 16   | Cerium oxide-NPs      | Nanoceria (NCe)                      | NCe-FA demonstrated excellent antiangiogenic effect in ovarian cancer.                                | [167,168] |
| 17   | Tetrac-NP             | Tetraiodothyroacetic acid            | Tetrac-NP significantly suppressed tumor growth and angiogenesis in murine xenograft models.         | [169]      |
| 18   | Polymeric Nanoparticle | Diamino Propane Tetraiodothyroacetic Acid | NPs showed excellent pharmacokinetics, biodistribution, and antiangiogenesis properties.       | [170]      |
| 19   | Carbon-NPs            | Angiogenesis inhibitors              | Inhibits tumor angiogenesis and tumor growth.                                                         | [146,171] |
| 20   | Silver nanoparticles (Ag-NPs) | Ag-NPs     | Ag-NPs inhibit vascular endothelial growth factor (VEGF) and the formation of new blood microvessels. | [120,172] |
| 21   | Chitosan nanoparticles (CNP) | Alphastatin/CNPs | AsCs-NPs inhibited the SphK1-S1P signaling pathway and enhanced the antiangiogenic effect of Alphastatin both in vitro and in vivo. | [173,174] |
Table 2. Cont.

| S.No | Nanoparticle                   | Therapeutics       | Application                                                                 | Ref    |
|------|--------------------------------|--------------------|-----------------------------------------------------------------------------|--------|
| 22   | Graphene-NPs                   | Graphite(G), rGO and nGO | Graphite nanoparticles and graphene oxide nanoplatelets showed potential antiangiogenic effects. | [175,176] |
| 23   | Chitosan-derived micelles      | Apatinib           | Apatinib-micelles showed effective anti-angiogenesis cancer therapy.        | [177]   |
| 24   | Cationic PEGylated liposomes   | Gambogic acid      | GAL significantly inhibited angiogenesis against TNBC.                      | [178]   |
| 25   | PLGA copolymer                 | Osseltamivirphosphate (OP) | PLGA-OP actively impedes tumor neovascularization, growth, and metastasis in a mouse model of human pancreatic carcinoma. | [179]   |
| 26   | Lipid-PA nanoparticles         | Rapamycin and DiR   | RDLPNPs showed an excellent antitumor effect with the enhanced photothermal and antiangiogenic effect. | [180]   |
| 27   | Selenium nanoparticles         | VEGF siRNA         | Showed enhanced in vivo VEGF-siRNA silencing and fluorescence imaging efficacy. | [113]   |
| 28   | Mesoporous silica nanoparticle | Combretastatin A4 doxorubicin | Tumor vascular-targeted co-delivery iRGD-NPs presented excellent anti-angiogenesis and antitumor activity. | [181]   |
| 29   | pH-sensitive polymeric nanoparticles | Doxorubicin curcumin | Displaced enhanced proapoptotic and antiangiogenic activities. | [182]   |
| 30   | Multifunctional nanodrugs      | LMWH and ursolic acid | Demonstrated excellent anti-angiogenesis and antitumor activity.          | [183]   |

11. Challenges of Nanomedicine, Conclusion, and Future Perspective

Various nanomaterials were comprehensively utilized for different biomedical applications including cancer theranostics and anti-angiogenic cancer therapy. Nevertheless, some of these nanomaterials occasionally show significant toxicity and other side effects causing potential challenges for successful clinical application. Hence, it is crucial to test their biosafety, long term metabolic activity, degradation property, pharmacokinetics and pharmacodynamics, interaction with immune cells, sustainable circulation in the body, etc., before their use in humans [25]. The future challenge is to develop robust cancer targeted anti-angiogenic nanomedicine with minimum side effects. Additionally, combination of FDA approved anti-angiogenic drugs to these nanomaterials can potentially increase the therapeutic effects and decrease the dose of treatment and subsequent toxicity. Additionally, vital information about NPs interaction, uptake, circulation, retention, and excretion are necessary and need to be thoroughly investigated. Apart from that, more studies are required to reduce the cost of industrial scale production, develop better technologies to synthesize multifunctional nanomaterials, and to identify the best route of drug administration [26].

The erratic structure and aberrant functionalities of tumor-associated neo-vessels provide much needed information to fight and evade the challenges associated with anticancer therapies. It is quite evident that these therapeutic modalities exhibited an efficacious management of cancer under certain settings, yet a substantial range of benefits is to be realized for many patients in the form of metastasis as well as overall survival. Moreover, such approaches and associated with untoward toxicity to normal cells along with many adverse side effects. The primary future challenge, thus, boils down to designing novel targeted nanomedicine for anti-angiogenic therapeutics. This should be accompanied by combinatorial strategy with chemotherapeutics to enhance the anticancer efficacy along with antiangiogenic tumor growth inhibition. Discovery of novel biomarkers is also urgently needed to accomplish efficacious drug treatment and their safety. Formulating new strategies to overcome drug resistance is a must as cancer cells routinely become impervious to drugs. For sure, pharmacokinetics
and pharmacodynamics parameters of the nanomaterials should be assessed properly, biodegradability and clearance should be determined meticulously, and route of administration and dose regimen should be optimized accurately. Obviously, the future cancer nanomedicine will demand a more intense understanding of metabolic aspects of cancers, possibility of immune destruction, circumventing various physiological barriers, exploring material properties, etc., to achieve exceedingly efficient nanocarriers for anti-angiogenic cancer therapy.

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**References**

1. Hanahan, D.; Folkman, J. Patterns and Emerging Mechanisms of the Angiogenic Switch during Tumorigenesis. *Cell* 1996, 86, 353–364. [CrossRef]
2. Dvorak, H.F. Angiogenesis: Update 2005. J. Thromb. Haemost. 2005, 3, 1835–1842. [CrossRef]
3. Folkman, J. Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat. Med.* 1995, 1, 27–31. [CrossRef]
4. Petrillo, M.; Patella, F.; Pesapane, F.; Suter, M.B.; Ierardi, A.M.; Angileri, S.A.; Floridi, C.; de Filippo, M.; Carrafiello, G. Hypoxia and tumor angiogenesis in the era of hepatocellular carcinoma transarterial loco-regional treatments. *Future Oncol.* 2018, 14, 2957–2967. [CrossRef]
5. Baeriswyl, V.; Christofori, G. The angiogenic switch in carcinogenesis. *Semin. Cancer Biol.* 2009, 19, 329–337. [CrossRef]
6. Ferrara, N. Vascular Endothelial Growth Factor. *Arterioscler. Thromb. Vasc. Biol.* 2009, 29, 789–791. [CrossRef]
7. Ferrara, N. Pathways mediating VEGF-independent tumor angiogenesis. *Cytokine Growth Factor Rev.* 2010, 21, 21–26. [CrossRef]
8. Ferrara, N.; Adamis, A.P. Ten years of anti-vascular endothelial growth factor therapy. *Nat. Rev. Drug Discov.* 2016, 15, 385–403. [CrossRef]
9. Bergers, G.; Benjamin, L.E. Tumorigenesis and the angiogenic switch. *Nat. Rev. Cancer* 2003, 3, 401–410. [CrossRef]
10. Bergers, G.; Hanahan, D. Modes of resistance to anti-angiogenic therapy. *Nat. Rev. Cancer* 2008, 8, 592–603. [CrossRef] [PubMed]
11. Lv, X.; Li, J.; Zhang, C.; Hu, T.; Li, S.; He, S.; Yan, H.; Tan, Y.; Lei, M.; Wen, M.; et al. The role of hypoxia-inducible factors in tumor angiogenesis and cell metabolism. *Genes Dis.* 2017, 4, 19–24. [CrossRef]
12. Wang, J.C.; Li, G.Y.; Li, P.P.; Sun, X.; Li, W.M.; Li, Y.; Lu, S.Y.; Liu, P. Suppression of hypoxia-induced excessive angiogenesis by metformin via elevating tumor blood perfusion. *Oncotarget* 2017, 8, 73892–73904. [CrossRef]
13. Mukherjee, S.; Patra, C.R. Therapeutic application of anti-angiogenic nanomaterials in cancers. *Nanoscale* 2016, 8, 12444–12470. [CrossRef]
14. Carmeliet, P.; Jain, R.K. Principles and mechanisms of vessel normalization for cancer and other angiogenic diseases. *Nat. Rev. Drug Discov.* 2011, 10, 417–427. [CrossRef]
15. Heath, V.L.; Bicknell, R. Anticancer strategies involving the vasculature. *Nat. Rev. Clin. Oncol.* 2009, 6, 395–404. [CrossRef]
41. Rajabi, M.; Mousa, S.A. The Role of Angiogenesis in Cancer Treatment. *Biomedicines* 2017, 5, 34. [CrossRef] [PubMed]
42. Folkman, J. Angiogenesis: An organizing principle for drug discovery? *Nat. Rev. Drug Discov.* 2007, 6, 273–286. [CrossRef] [PubMed]
43. Minion, L.E.; Tewari, K.S. The safety and efficacy of bevacizumab in the treatment of patients with recurrent or metastatic cervical cancer. *Expert Rev. Anticancer Ther.* 2017, 17, 191–198. [CrossRef] [PubMed]
44. Cloughesy, T.F.; Brenner, A.; de Groot, J.F.; Butowski, N.A.; Zach, L.; Campman, J.L.; Elllionson, B.M.; Freedman, L.S.; Cohen, Y.C.; Lowenton-Spier, N.; et al. A randomized controlled phase III study of VB-111 combined with bevacizumab vs. bevacizumab monotherapy in patients with recurrent glioblastoma (GLOBE). *Neuro Oncol.* 2019. [CrossRef]
45. Falcon, B.L.; Chinthalapalli, S.; Uhlik, M.T.; Pytowski, B. Antagonist antibodies to vascular endothelial growth factor receptor 2 (VEGFR-2) as anti-angiogenic agents. *Pharmacol. Ther.* 2016, 164, 204–225. [CrossRef]
46. Ribatti, D.; Vacca, A. New Insights in Anti-Angiogenesis in Multiple Myeloma. *Int. J. Mol. Sci.* 2018, 19, 2031. [CrossRef]
47. Montoro, J.; Yerlikaya, A.; Ali, A.; Raza, A. Improving Treatment for Myelodysplastic Syndromes Patients. *Curr. Treat. Options Oncol.* 2018, 19, 66. [CrossRef]
48. Cheng, C.C.; Chao, W.T.; Liao, C.C.; Hsu, Y.H.; Liu, Y.H. The Roles Of Angiogenesis And Cancer Stem Cells In Sorafenib Drug Resistance In Hepatocellular Carcinoma. *Onco Targets Ther.* 2019, 12, 8217–8227. [CrossRef]
49. Hao, Z.; Sadek, I. Sunitinib: The antiangiogenic effects and beyond. *Onco Targets Ther.* 2016, 9, 5495–5505. [CrossRef]
50. Chen, Z.; Yang, H.; Li, Z.; Xia, Q.; Nie, Y. Temsirolimus as a dual inhibitor of retinoblastoma and angiogenesis via targeting mTOR signalling. *Biochem. Biophys. Res. Commun.* 2019, 516, 726–732. [CrossRef]
51. Kamli, H.; Li, L.; Gobe, G.C. Limitations to the Therapeutic Potential of Tyrosine Kinase Inhibitors and Alternative Therapies for Kidney Cancer. *Ochsner J.* 2019, 19, 138–151. [CrossRef]
52. Chellappan, D.K.; Chellian, J.; Ng, Z.Y.; Sim, Y.J.; Theng, C.W.; Ling, J.; Wong, M.; Foo, J.H.; Yang, G.J.; Hang, L.Y.; et al. The role of pazopanib on tumour angiogenesis and in the management of cancers: A review. *Biomed. Pharmacother.* 2017, 96, 768–781. [CrossRef]
53. Matsuki, M.; Hoshi, T.; Yamamoto, Y.; Ikemori-Kawada, M.; Minoshima, Y.; Funahashi, Y.; Matsui, J. Lenvatinib inhibits angiogenesis and tumor fibroblast growth factor signaling pathways in human hepatocellular carcinoma models. *Cancer Med.* 2018, 7, 2641–2653. [CrossRef]
54. Abdelaziz, A.; Vaishampayan, U. Cabozantinib for the treatment of kidney cancer. *Expert Rev. Anticancer Ther.* 2017, 17, 577–584. [CrossRef]
55. Lee, L.; Ito, T.; Jensen, R.T. Everolimus in the treatment of neuroendocrine tumors: Efficacy, side-effects, resistance, and factors affecting its place in the treatment sequence. *Expert Opin. Pharmacother.* 2018, 19, 909–928. [CrossRef]
56. Morabito, A.; Piccirillo, M.C.; Falasconi, F.; De Feo, G.; Del Giudice, A.; Bryce, J.; Di Maio, M.; De Maio, E.; Normanno, N.; Perrone, F. Vandetanib (ZD6474), a dual inhibitor of vascular endothelial growth factor receptor (VEGFR) and epidermal growth factor receptor (EGFR) tyrosine kinases: Current status and future directions. *Oncologist* 2009, 14, 378–390. [CrossRef]
57. Javle, M.; Smyth, E.C.; Chau, I. Ramucirumab: Successfully targeting angiogenesis in gastric cancer. *Clin. Cancer Res.* 2014, 20, 5875–5881. [CrossRef]
58. Rey, J.B.; Launay-Vacher, V.; Tourmigand, C. Regorafenib as a single-agent in the treatment of patients with gastrointestinal tumors: An overview for pharmacists. *Target. Oncol.* 2015, 10, 199–213. [CrossRef]
59. Ivanova, J.I.; Saverno, K.R.; Sung, J.; Duh, M.S.; Zhao, C.; Cai, S.; Vekeman, F.; Peeyh house, A.; Dhawan, R.; Fuchs, C.S. Real-world treatment patterns and effectiveness among patients with metastatic colorectal cancer treated with ziv-aflibercept in community oncology practices in the USA. *Med. Oncol.* 2017, 34, 193. [CrossRef]
60. Ribatti, D. Tumor refractoriness to anti-VEGF therapy. *Oncotarget* 2016, 7, 46668-46677. [CrossRef]
61. Gacche, R.N.; Assaraf, Y.G. Redundant angiogenic signaling and tumor drug resistance. *Drug Resist. Updates* 2018, 36, 47–76. [CrossRef] [PubMed]
62. van Beijnum, J.R.; Nowak-Sliwinska, P.; Huijbers, E.J.; Thijsen, V.L.; Griffioen, A.W. The great escape; the hallmarks of resistance to antiangiogenic therapy. *Pharmacol. Rev.* 2015, 67, 441–461. [CrossRef] [PubMed]
Hida, K.; Maishi, N.; Sakurai, Y.; Hida, Y.; Harashima, H. Heterogeneity of tumor endothelial cells and drug delivery. *Adv. Drug Deliv. Rev.* 2016, 99 Pt B, 140–147. [CrossRef] [PubMed]

Meka, R.R.; Mukherjee, S.; Patra, C.R.; Chaudhuri, A. Shikimoyl-ligand decorated gold nanoparticles for use in ex vivo engineered dendritic cell based DNA vaccination. *Nanoscale* 2019, 11, 7931–7943. [CrossRef] [PubMed]

Muthuraj, B.; Mukherjee, S.; Patra, C.R.; Iyer, P.K. Amplified Fluorescence from Polyfluorene Nanoparticles with Dual State Emission and Aggregation Caused Red Shifted Emission for Live Cell Imaging and Cancer Theranostics. *ACS Appl. Mater. Interfaces* 2016, 8, 32220–32229. [CrossRef] [PubMed]

Gaddam, R.R.; Mukherjee, S.; Punugputi, N.; Vasudevan, D.; Patra, C.R.; Narayan, R.; Vsn Kothapalli, R. Facile synthesis of carbon dot and residual carbon nanobeads: Implications for ion sensing, medicinal and biological applications. *Mater. Sci. Eng. C Mater. Biol. Appl.* 2017, 73, 643–652. [CrossRef]

Mukherjee, P.; Bhattacharya, R.; Wang, P.; Wang, L.; Basu, S.; Nagy, J.A.; Atala, A.; Mukhopadhyay, D.; Soker, S. Antiangiogenic Properties of Gold Nanoparticles. *Clin. Cancer Res.* 2005, 11, 3530. [CrossRef] [PubMed]

Barui, A.K.; Veeriah, V.; Mukherjee, S.; Manna, J.; Patel, A.K.; Patra, S.; Pal, K.; Murali, S.; Rana, R.K.; Chatterjee, S.; et al. Zinc oxide nanoflowers make new blood vessels. *Nanoscale* 2012, 4, 7861–7869. [CrossRef]

Afsharzadeh, M.; Abnous, K.; Yazdian-Robati, R.; Ataranzadeh, A.; Ramezani, M.; Hashemi, M. Formulation and evaluation of anticancer and antiangiogenesis efficiency of PLA-PEG nanoparticles loaded with galbamic acid in C26 colon carcinoma, in vitro and in vivo. *J. Cell. Physiol.* 2019, 234, 6099–6107. [CrossRef]

Min, H.; Wang, J.; Qi, Y.; Zhang, Y.; Han, X.; Xu, Y.; Xu, J.; Li, Y.; Chen, L.; Cheng, K.; et al. Biomimetic Metal-Organic Framework Nanoparticles for Cooperative Combination of Antiangiogenesis and Photodynamic Therapy for Enhanced Efficacy. *Adv. Mater.* 2019, 31, e1808200. [CrossRef] [PubMed]

Yu, M.; Su, D.; Yang, Y.; Qin, L.; Hu, C.; Liu, R.; Zhou, Y.; Yang, C.; Yang, X.; Wang, G.; et al. D-T7 Peptide-Modified PEGylated Bilirubin Nanoparticles Loaded with Cediranib and Paclitaxel for Antiangiogenesis and Chemotherapy of Glioma. *ACS Appl. Mater. Interfaces* 2019, 11, 176–186. [CrossRef] [PubMed]

Mukherjee, S.; Sriram, P.; Barui, A.K.; Nethi, S.K.; Veeriah, V.; Chatterjee, S.; Suresh, K.I.; Patra, C.R. Graphene Oxides Show Antiangiogenic Properties. *Adv. Healthc. Mater.* 2015, 4, 1722–1732. [CrossRef] [PubMed]

Shibuya, M. Vascular Endothelial Growth Factor (VEGF) and Its Receptor (VEGFR) Signaling in Angiogenesis: A Crucial Target for Anti- and Pro-Angiogenic Therapies. *Genes Cancer* 2011, 2, 1097–1105. [CrossRef] [PubMed]

Zhao, Y.; Wang, W.; Guo, S.; Wang, Y.; Miao, L.; Xiong, Y.; Huang, L. PolyMetformin combines carrier and anticancer activities for in vivo siRNA delivery. *Nat. Commun.* 2016, 7, 11822. [CrossRef] [PubMed]

Shi, K.; Zhao, Y.; Miao, L.; Satterlee, A.; Haynes, M.; Luo, C.; Musetti, S.; Huang, L. Dual Functional LipoMET Mediates Envelope-type Nanoparticles to Combinational Oncogene Silencing and Tumor Growth Inhibition. *Mol. Ther.* 2017, 25, 1567–1579. [CrossRef]

Yang, Z.-Z.; Li, J.-Q.; Wang, Z.-Z.; Dong, D.-W.; Qi, X.-R. Tumor-targeting dual peptides-modified cationic liposomes for delivery of siRNA and docetaxel to gliomas. *Biomaterials* 2014, 35, 5222–5239. [CrossRef] [PubMed]

Liu, X.-Q.; Song, W.-J.; Sun, T.-M.; Zhang, P.-Z.; Wang, J. Targeted Delivery of Antisense Inhibitor of miRNA for Antiangiogenesis Therapy Using cRGD-Functionalyzed Nanoparticles. *Mol. Pharm.* 2010, 8, 250–259. [CrossRef] [PubMed]

Yang, Z.; Xiang, B.; Dong, D.; Wang, Z.; Li, J.; Qi, X. Dual Receptor-Specific Peptides Modified Liposomes as VEGF siRNA Vector for Tumor-Targeting Therapy. *Curr. Gene Ther.* 2014, 14, 289–299. [CrossRef]

Guo, P.; Yang, J.; Jia, D.; Moses, M.A.; Auguste, D.T. ICAM-1-Targeted, Lcn2 siRNA-Encapsulating Liposomes are Potent Anti-angiogenic Agents for Triple Negative Breast Cancer. *Theranostics* 2016, 6, 1–13. [CrossRef]

Chen, J.; Sun, X.; Shao, R.; Xu, Y.; Gao, J.-Q.; Liang, W.-Q. VEGF siRNA delivered by polycation liposome-encapsulated calcium phosphate nanoparticles for tumor angiogenesis inhibition in breast cancer. *Int. J. Nanomed.* 2017, 12, 6075–6088. [CrossRef] [PubMed]

Xia, Y.; Tian, J.; Chen, X. Effect of surface properties on liposomal siRNA delivery. *Biomaterials* 2016, 79, 56–68. [CrossRef] [PubMed]

Singh, A.; Trivedi, P.; Jain, N.K. Advances in siRNA delivery in cancer therapy. *Artif. Cells Nanomed. Biotechnol.* 2017, 46, 274–283. [CrossRef] [PubMed]
83. Li, F.; Wang, Y.; Chen, W.-L.; Wang, D.-D.; Zhou, Y.-J.; You, B.-G.; Liu, Y.; Qu, C.-X.; Yang, S.-D.; Chen, M.-T.; et al. Co-delivery of VEGF siRNA and Etoposide for Enhanced Anti-angiogenesis and Anti-proliferation Effect via Multi-functional Nanoparticles for Orthotopic Non- Small Cell Lung Cancer Treatment. *Theranostics* 2019, 9, 5886–5898. [CrossRef] [PubMed]

84. Nagase, K.; Hasegawa, M.; Ayano, E.; Maitani, Y.; Kanazawa, H. Effect of Polymer Phase Transition Behavior on Temperature-Responsive Polymer-Modified Liposomes for siRNA Transfection. *Int. J. Mol. Sci.* 2019, 20, 430. [CrossRef]

85. Nagase, K.; Hasegawa, M.; Ayano, E.; Maitani, Y.; Kanazawa, H. Efficacy of PEGylated cationic liposomes for suppression of tumor formation and metastasis in a rat model of lung cancer. *Int. J. Biol. Pharm. Biochem. Pharmacol.* 2012, 6, 1756–1765. [PubMed]

86. Lee, S.J.; Yook, S.; Yhee, J.Y.; Yoon, H.Y.; Kim, M.-G.; Ku, S.H.; Kim, S.H.; Park, J.H.; Jeong, J.H.; Kwon, I.C.; et al. Co-delivery of VEGF and Bcl-2 dual-targeted siRNA polymer using a single nanoparticle for synergistic anti-cancer effects in vivo. *J. Control. Release* 2015, 220, 631–641. [CrossRef]

87. Hashemi Goradel, N.; Ghiyami-Hour, F.; Jahangiri, S.; Negahdari, B.; Sahebkar, A.; Masoudifar, A.; Mirzaei, H. Characterization of sorafenib-loaded calixarene nanoparticles for cancer therapy. *Int. J. Pharm.* 2015, 485, 103–115. [CrossRef]

88. Kim, Y.; Lim, J.; Lee, S.J.; Yook, S.; Kim, D.D.; Kim, H.; Lee, J.H.; Jeong, J.H.; Kwon, I.C.; et al. Co-delivery of sorafenib and VEGF-siRNA via pH-sensitive liposomes for synergistic anti-cancer effects in vivo. *J. Control. Release* 2015, 220, 631–641. [CrossRef]

89. Yao, Y.; Wang, T.; Liu, Y.; Zhang, N. Co-delivery of sorafenib and VEGF-siRNA via pH-sensitive liposomes for the synergistic treatment of hepatocellular carcinoma. *Artif. Cells Nanomed. Biotechnol.* 2019, 47, 1374–1383. [CrossRef] [PubMed]

90. Sun, W.; Wang, Y.; Cai, M.; Lin, L.; Chen, X.; Cao, Z.; Zhu, K.; Shuai, X. Codelivery of sorafenib and GPC3 siRNA with PEI-modified liposomes for hepatoma therapy. *Biomater. Sci.* 2017, 5, 2468–2479. [CrossRef]

91. Chen, Y.; Liu, Y.-C.; Sung, Y.-C.; Ramjiawan, R.R.; Lin, T.-T.; Chang, C.-C.; Jeng, K.-S.; Chang, C.-F.; Liu, C.-H.; Gao, D.-Y.; et al. Overcoming sorafenib evasion in hepatocellular carcinoma using CXCR4-targeted nanoparticles to co-deliver MEK-inhibitors. *Sci. Rep.* 2017, 7, 44123. [CrossRef] [PubMed]

92. Pal, K.; Madamsetty, V.S.; Dutta, S.K.; Mukhopadhyay, D. Co-delivery of everolimus and vinorelbine via a tumor-targeted liposomal formulation inhibits tumor growth and metastasis in RCC. *Int. J. Nanomed.* 2019, 14, 5109–5123. [CrossRef] [PubMed]

93. Abud, M.B.; Louzada, R.N.; Issac, D.L.C.; Souza, L.G.; dos Reis, R.G.; Lima, E.M.; de Ávila, M.P. In vivo and in vitro toxicity evaluation of liposome-encapsulated sirolimus. *Int. J. Retin. Vitr.* 2019, 5, 35. [CrossRef] [PubMed]

94. Yoon, H.Y.; Chang, I.H.; Goo, Y.T.; Kim, C.H.; Kang, T.H.; Kim, S.-Y.; Lee, S.J.; Song, S.H.; Whang, Y.M.; Choi, Y.W. Intravesical delivery of rapamycin via folate-modified liposomes dispersed in thermo-reversible hydrogel. *Int. J. Nanomed.* 2019, 14, 6249–6268. [CrossRef]

95. Iwase, Y.; Maitani, Y. Preparation and in Vivo Evaluation of Liposomal Everolimus for Lung Carcinoma and Thyroid Carcinoma. *Biol. Pharm. Bull.* 2012, 35, 975–979. [CrossRef]

96. Banerjee, I.; De, K.; Mukherjee, D.; Dey, G.; Chattopadhyay, S.; Mukherjee, M.; Mandal, M.; Bandyopadhyay, A.K.; Gupta, A.; Ganguly, S.; et al. Paclitaxel-loaded solid lipid nanoparticles modified with Tyr-3-octreotide for enhanced anti-angiogenic and anti-glioma therapy. *Acta Biomater.* 2016, 38, 69–81. [CrossRef]

97. Abu-Lila, A.; Suzuki, T.; Doi, Y.; Ishida, T.; Kiwada, H. Oxaliplatin targeting to angiogenic vessels by PEGylated cationic liposomes suppresses the angiogenesis in a dorsal air sac mouse model. *J. Control. Release* 2009, 134, 18–25. [CrossRef]

98. Sengupta, S.; Eavarone, D.; Capila, I.; Zhao, G.; Watson, N.; Kiziltepe, T.; Sasidekharan, R. Temporal targeting of tumour cells and neovasculature with a nanoscale delivery system. *Nature* 2005, 436, 568–572. [CrossRef]

99. Bhattacharai, P.; Hameed, S.; Dai, Z. Recent advances in anti-angiogenic nanomedicines for cancer therapy. *Nanoscale* 2018, 10, 5393–5423. [CrossRef]

100. Duncan, R. Polymer conjugates as anticancer nanomedicines. *Nat. Rev. Cancer* 2006, 6, 688–701. [CrossRef] [PubMed]

101. Weidner, N.; Semple, J.P.; Welch, W.R.; Folkman, J. Tumor Angiogenesis and Metastasis—Correlation in Invasive Breast Carcinoma. *N. Engl. J. Med.* 1991, 324, 1–8. [CrossRef] [PubMed]
102. Satchi-Fainaro, R.; Puder, M.; Davies, J.W.; Tran, H.T.; Sampson, D.A.; Greene, A.K.; Corfas, G.; Folkman, J. Targeting angiogenesis with a conjugate of HPMA copolymer and TNP-470. Nat. Med. 2004, 10, 255–261. [CrossRef] [PubMed]

103. Benny, O.; Fainaru, O.; Adini, A.; Cassiola, F.; Bazinet, L.; Adini, I.; Pravda, E.; Nahmias, Y.; Koirala, S.; Corfas, G.; et al. An orally delivered small-molecule formulation with antiangiogenic and anticancer activity. Nat. Biotechnol. 2008, 26, 799–807. [CrossRef] [PubMed]

104. Harfouche, R.; Basu, S.; Soni, S.; Hentschel, D.M.; Mashelkar, R.A.; Sengupta, S. Nanoparticle-mediated targeting of phosphatidylinositol-3-kinase signaling inhibits angiogenesis. Angiogenesis 2009, 12, 325–338. [CrossRef]

105. Dahmani, F.Z.; Xiao, Y.; Zhang, J.; Yu, Y.; Zhou, J.; Yao, J. Multifunctional Polymeric Nanosystems for Dual-Targeted Combinatorial Chemo/Antiangiogenesis Therapy of Tumors. Adv. Healthc. Mater. 2016, 5, 1447–1461.

106. Tian, F.; Dahmani, F.Z.; Qiao, J.; Ni, J.; Xiong, H.; Liu, T.; Zhou, J.; Yao, J. A targeted nanoplatform co-delivering chemotherapeutic and antiangiogenic drugs as a tool to reverse multidrug resistance in breast cancer. Acta Biomater. 2018, 75, 398–412. [CrossRef]

107. Prasad, P.; Shuhendler, A.; Cai, P.; Rauth, A.M.; Wu, X.Y. Doxorubicin and mitomycin C co-loaded polymer-lipid hybrid nanoparticles inhibit growth of sensitive and multidrug resistant human mammary tumor xenografts. Cancer Lett. 2013, 334, 263–273. [CrossRef]

108. Zhang, T.; Prasad, P.; Cai, P.; He, C.; Shan, D.; Rauth, A.M.; Wu, X.Y. Dual-targeted hybrid nanoparticles of synergistic drugs for treating lung metastases of triple negative breast cancer in mice. Acta Pharmacol. Sin. 2017, 38, 835–847. [CrossRef]

109. Yang, S.; Tang, Z.; Hu, C.; Zhang, D.; Shen, N.; Yu, H.; Chen, X. Selectively Potentiating Hypoxia Levels by Combretastatin A4 Nanomedicine: Toward Highly Enhanced Hypoxia-Activated Prodrug Tirapazamine Therapy for Metastatic Tumors. Adv. Mater. 2019, 31, 1805955. [CrossRef]

110. Tandle, A.; Blazer, D.G.; Libutti, S.K. Antiangiogenic gene therapy of cancer: Recent developments. J. Transl. Med. 2004, 2, 22. [CrossRef]

111. Lee, J.; Jeong, E.J.; Lee, Y.K.; Kim, K.; Kwon, I.C.; Lee, K.Y. Optical Imaging and Gene Therapy with Neuroblastoma-Targeting Polymeric Nanoparticles for Potential Theranostic Applications. Small 2016, 12, 1201–1211. [CrossRef] [PubMed]

112. Chen, Q.; Osada, K.; Ge, Z.; Uchida, S.; Tockary, T.A.; Dirisala, A.; Matsui, A.; Toh, K.; Takeda, K.M.; Liu, X.; et al. Polyplex micelle installing intracellular self-processing functionalities without free catiomers for safe and efficient systemic gene therapy through tumor vasculature targeting. Biomaterials 2017, 113, 253–265. [CrossRef] [PubMed]

113. Yu, Q.; Liu, Y.; Cao, C.; Le, F.; Qin, X.; Sun, D.; Liu, J. The use of pH-sensitive functional selenium nanoparticles shows enhanced in vivo VEGF-siRNA silencing and fluorescence imaging. Nanoscale 2014, 6, 9279–9292. [CrossRef] [PubMed]

114. Yang, S.; Tang, Z.; Hu, C.; Zhang, D.; Shen, N.; Yu, H.; Chen, X. Selectively Potentiating Hypoxia Levels by Combretastatin A4 Nanomedicine: Toward Highly Enhanced Hypoxia-Activated Prodrug Tirapazamine Therapy for Metastatic Tumors. Adv. Mater. 2019, 31, 1805955. [CrossRef]

115. Sun, H.; Dong, Y.; Feijen, J.; Zhong, Z. Peptide-decorated polymeric nanomedicines for precision cancer therapy. J. Control. Release 2018, 290, 11–27. [CrossRef]

116. Pan, F.; Yang, W.; Li, W.; Yang, X.-Y.; Liu, S.; Li, X.; Zhao, X.; Ding, H.; Qin, L.; Pan, Y. Conjugation of gold nanoparticles and recombinant human endostatin modulates vascular normalization via interruption of anterior gradient 2–mediated angiogenesis. Tumor Biol. 2017, 39. [CrossRef]

117. Shin, S.W.; Jung, W.; Choi, C.; Kim, S.Y.; Son, A.; Kim, H.; Lee, N.; Park, H.C. Fucoidan-Manganese Dioxide Nanoparticles Potentiate Radiation Therapy by Co-Targeting Tumor Hypoxia and Angiogenesis. Mar. Drugs 2018, 16, 510. [CrossRef]

118. Balakrishnan, S.; Bhat, F.A.; Raja Singh, P.; Mukherjee, S.; Elumalai, P.; Das, S.; Patra, C.R.; Arunakaran, J. Gold nanoparticle-conjugated quercetin inhibits epithelial-mesenchymal transition, angiogenesis and invasiveness viaEGFR/VEGFR-2-mediated pathway in breast cancer. Cell Prolif. 2016, 49, 678–697. [CrossRef]

119. Pan, Y.; Ding, H.; Qin, L.; Zhao, X.; Cai, J.; Du, B. Gold nanoparticles induce nanostructural reorganization of VEGFR2 to repress angiogenesis. J. Biomed. Nanotechnol. 2013, 9, 1746–1756. [CrossRef]

120. Gurunathan, S.; Lee, K.-J.; Kalishwaralal, K.; Sheikpranbabu, S.; Vaidyanathan, R.; Eom, S.H. Antiangiogenic properties of silver nanoparticles. Biomaterials 2009, 30, 6341–6350. [CrossRef]
121. Song, H.; Wang, W.; Zhao, P.; Qi, Z.; Zhao, S. Cuprous oxide nanoparticles inhibit angiogenesis via down regulation of VEGFR2 expression. *Nanoscale* **2014**, *6*, 3206–3216. [CrossRef] [PubMed]

122. Giri, S.; Karakoti, A.; Graham, R.P.; Maguire, J.L.; Reilly, C.M.; Seal, S.; Rattan, R.; Shridhar, V. Nanoceria: A rare-earth nanoparticle as a novel anti-angiogenic therapeutic agent in ovarian cancer. *PloS ONE* **2013**, *8*, e54578. [CrossRef] [PubMed]

123. Setyawati, M.I.; Leong, D.T. Mesoporous Silica Nanoparticles as an Antitumoral-Angiogenesis Strategy. *ACS Appl. Mater. Interfaces* **2017**, *9*, 6690–6703. [CrossRef] [PubMed]

124. Neek, M.; Kim, T.I.; Wang, S.-W. Protein-based nanoparticles in cancer vaccine development. *Nanomed. Nanotechnol. Biol. Med.* **2019**, *15*, 164–174. [CrossRef]

125. Diaz, D.; Care, A.; Sunna, A. Bioengineering Strategies for Protein-Based Nanoparticles. *Genes* **2018**, *9*, 370. [CrossRef] [PubMed]

126. Lee, E.J.; Lee, N.K.; Kim, I.-S. Bioengineered protein-based nanocage for drug delivery. *Adv. Drug Deliv. Rev.* **2016**, *106*, 157–171. [CrossRef] [PubMed]

127. Sandra, F.; Khaliq, N.U.; Sunna, A.; Care, A. Developing Protein-Based Nanoparticles as Versatile Delivery Systems for Cancer Therapy and Imaging. *Nanomaterials* **2019**, *9*, 1329. [CrossRef] [PubMed]

128. Lin, T.; Zhao, P.; Jiang, Y.; Tang, Y.; Jin, H.; Pan, Z.; He, H.; Yang, V.C.; Huang, Y. Blood–Brain-Barrier-Penetrating Albumin Nanoparticles for Biomimetic Drug Delivery via Albumin-Binding Protein Pathways for Antiglioma Therapy. *ACS Nano* **2016**, *10*, 9999–10012. [CrossRef]

129. Zhang, L.; Liu, Z.; Yang, K.; Kong, C.; Liu, C.; Chen, H.; Huang, J.; Qian, F. Tumor Progression of Non-Small Cell Lung Cancer Controlled by Albumin and Micellar Nanoparticles of Itraconazole, a Multitarget Angiogenesis Inhibitor. *Mol. Pharm.* **2017**, *14*, 4705–4713. [CrossRef]

130. Kommareddy, S.; Amiji, M. Antiangiogenic gene therapy with systemically administered sFlt-1 plasmid DNA in engineered gelatin-based nanovectors. *Cancer Gene Ther.* **2007**, *14*, 488–498. [CrossRef] [PubMed]

131. Lizotte, P.; Wen, A.M.; Sheen, M.R.; Fields, J.; Reilly, C.M.; Seal, S.; Rattan, R.; Shridhar, V. Nanoceria: A rare-earth nanoparticle as a novel anti-angiogenic therapeutic agent in ovarian cancer. *PloS ONE* **2013**, *8*, e54578. [CrossRef] [PubMed]

132. Giri, S.; Karakoti, A.; Graham, R.P.; Maguire, J.L.; Reilly, C.M.; Seal, S.; Rattan, R.; Shridhar, V. Nanoceria: A rare-earth nanoparticle as a novel anti-angiogenic therapeutic agent in ovarian cancer. *PloS ONE* **2013**, *8*, e54578. [CrossRef] [PubMed]

133. Wang, C.; Beiss, V.; Steinmetz, N.F.; Simon, A.E. Cowpea Mosaic Virus Nanoparticles and Empty Virus-Like Particles Show Distinct but Overlapping Immunostimulatory Properties. *J. Virol.* **2019**, *93*, e00129-19. [CrossRef] [PubMed]

134. Beatty, P.H.; Lewis, J.D. Cowpea mosaic virus nanoparticles for cancer imaging and therapy. *Adv. Drug Deliv. Rev.* **2019**, *145*, 130–144. [CrossRef]

135. Czapar, A.E.; Steinmetz, N.F. Plant viruses and bacteriophages for drug delivery in medicine and biotechnology. *Curr. Opin. Chem. Biol.* **2017**, *38*, 108–116. [CrossRef]

136. Gamper, C.; Spenlè, C.; Boscà, S.; van der Heyden, M.; Erhardt, M.; Orend, G.; Bagnard, D.; Heinlein, M. Functionalized Tobacco Mosaic Virus Coat Protein Monomers and Oligomers as Nanocarriers for Anti-Cancer Peptides. *Cancers* **2019**, *11*, 1609. [CrossRef]

137. Brunel, F.M.; Lewis, J.D.; Destito, G.; Steinmetz, N.F.; Manchester, M.; Stuhlmann, H.; Dawson, P.E. Hydrazone Ligation Strategy to Assemble Multifunctional Viral Nanoparticles for Cell Imaging and Tumor Targeting. *Nano Lett.* **2010**, *10*, 1093–1097. [CrossRef]

138. Lewis, J.D.; Destito, G.; Zijlstra, A.; Gonzalez, M.J.; Quigley, J.P.; Manchester, M.; Stuhlmann, H. Viral nanoparticles as tools for intravital vascular imaging. *Nat. Med.* **2006**, *12*, 354–360. [CrossRef]

139. Steinmetz, N.F. Viral nanoparticles as platforms for next-generation therapeutics and imaging devices. *Nanomed. Nanotechnol. Biol. Med.* **2010**, *6*, 634–641. [CrossRef]

140. Niehl, A.; Appaix, F.; Boscà, S.; van der Sanden, B.; Nicouz, J.-F.; Bolze, F.; Heinlein, M. Fluorescent Tobacco mosaic virus-Derived Bio-Nanoparticles for Intravital Two-Photon Imaging. *Front. PlantSci.* **2016**, *6*, 1244.

141. Cho, C.-F.; Yu, L.; Nsiama, T.K.; Kadam, A.N.; Raturi, A.; Shukla, S.; Amadei, G.A.; Steinmetz, N.F.; Luyt, L.G.; Lewis, J.D. Viral nanoparticles decorated with novel EGFl7 ligands enable intravital imaging of tumor neovasculature. *Nanoscale* **2017**, *9*, 12096–12109. [CrossRef] [PubMed]

142. Saeed, B.A.; Lim, V.; Yusof, N.A.; Khor, K.Z.; Rahman, H.S.; Abdul Samad, N. Antiangiogenic properties of nanoparticles: A systematic review. *Int. J. Nanomed.* **2019**, *14*, 5135–5146. [CrossRef] [PubMed]
143. Jiang, W.; Huang, Y.; An, Y.; Kim, B.Y.S. Remodeling Tumor Vasculature to Enhance Delivery of Intermediate-Sized Nanoparticles. *ACS Nano* 2015, 9, 8689–8696. [CrossRef] [PubMed]

144. Du, S.; Xiong, H.; Xu, C.; Lu, Y.; Yao, J. Attempts to strengthen and simplify the tumor vascular normalization strategy using tumor vessel normalization promoting nanomedicines. *Biomater. Sci.* 2019, 7, 1147–1160. [CrossRef] [PubMed]

145. Lu, L.; Chen, H.; Hao, D.; Zhang, X.; Wang, F. The functions and applications of A7R in anti-angiogenic therapy, imaging and drug delivery systems. *Asian J. Pharm. Sci.* 2019, 14, 595–608. [CrossRef]

146. Madamsetty, V.S.; Pal, K.; Keshavan, S.; Caulfield, T.R.; Dutta, S.K.; Wang, E.; Fadeel, B.; Mukhopadhyay, D. Development of multi-drug loaded PEGylated nanodiamonds to inhibit tumor growth and metastasis in genetically engineered mouse models of pancreatic cancer. *Nanoscale* 2019, 11, 22006–22018. [CrossRef] [PubMed]

147. Hu, J.; Chen, L.-J.; Liu, L.; Chen, X.; Chen, P.; Yang, G.-L.; Hou, W.-L.; Tang, M.-H.; Zhang, F.; Wang, X.-H.; et al. Liposomal honokiol, a potent anti-angiogenesis agent, in combination with radiotherapy produces a synergistic antitumor efficacy without increasing toxicity. *Exp. Mol. Med.* 2008, 40, 617. [CrossRef]

148. Jiang, Q.-Q.; Fan, L.-Y.; Yang, G.-L.; Guo, W.-H.; Hou, W.-L.; Chen, L.-J.; Wei, Y.-Q. Improved therapeutic effectiveness by combining liposomal honokiol with cisplatin in lung cancer model. *BMC Cancer* 2008, 8, 242. [CrossRef]

149. Liu, Y.; Chen, L.; He, X.; Fan, L.; Yang, G.; Chen, X.; Lin, X.; Du, L.; Li, Z.; Ye, H.; et al. Enhancement of therapeutic effectiveness by combining liposomal honokiol with cisplatin in ovarian carcinoma. *Int. J. Gynecol. Cancer* 2008, 18, 652. [CrossRef]

150. Strijkers, G.J.; Kluz, E.; Van Tilborg, G.A.F.; van der Schaft, D.W.J.; Griffioen, A.W.; Mulder, W.J.M.; Nicolay, K. Paramagnetic and fluorescent liposomes for target-specific imaging and therapy of tumor angiogenesis. *Angiogenesis* 2010, 13, 161–173. [CrossRef]

151. Brandwijk, R.J.M.G.E.; Mulder, W.J.M.; Nicolay, K.; Mayo, K.H.; Thijsen, V.L.J.L.; Griffioen, A.W. Anginex-Conjugated Liposomes for Targeting of Angiogenic Endothelial Cells. *Bioconjugate Chem.* 2007, 18, 785–790. [CrossRef] [PubMed]

152. Pal, K.; Madamsetty, V.S.; Dutta, S.K.; Wang, E.; Angom, R.S.; Mukhopadhyay, D. Synchronous inhibition of mTOR and VEGF/NRP1 axis impedes tumor growth and metastasis in renal cancer. *NPJ Precis. Oncol.* 2019, 3, 1–11. [CrossRef]

153. Battaglia, L.; Gallarate, M.; Peira, E.; Chirio, D.; Solazzi, I.; Giordano, S.M.A.; Gigliotti, C.L.; Riganti, C.; Dianzani, C. Bevacizumab loaded solid lipid nanoparticles prepared by the coacervation technique: Preliminary in vitro studies. *Nanotechnology* 2015, 26, 255102. [CrossRef] [PubMed]

154. Di Paolo, D.; Pastorino, F.; Zucatti, G.; Caffa, I.; Loi, M.; Marimpietri, D.; Brignole, C.; Perri, P.; Cilli, M.; Nico, B.; et al. Enhanced anti-tumor and anti-angiogenic efficacy of a novel liposomal fenretinide on human neuroblastoma. *J. Control. Release* 2013, 170, 445–451. [CrossRef] [PubMed]

155. Di Paolo, D.; Ambrogio, C.; Pastorino, F.; Brignole, C.; Martinengo, C.; Carosio, R.; Loi, M.; Pagnan, G.; Emionite, L.; Cilli, M.; et al. Selective Therapeutic Targeting of the Anaplastic Lymphoma Kinase With Liposomal siRNA Induces Apoptosis and Inhibits Angiogenesis in Neuroblastoma. *Mol. Ther.* 2011, 19, 2201–2212. [CrossRef]

156. Piaggio, F.; Kondylis, V.; Pastorino, F.; Di Paolo, D.; Perri, P.; Cossu, I.; Schorn, F.; Marinaccio, C.; Murgia, D.; Daga, A.; et al. A novel liposomal Clodronate depletes tumor-associated macrophages in primary and metastatic melanoma: Anti-angiogenic and anti-tumor effects. *J. Control. Release* 2016, 223, 165–177. [CrossRef]

157. Balakrishnan, S.; Mukherjee, S.; Das, S.; Bhat, F.A.; Raja Singh, P.; Patra, C.R.; Arunakaran, J. Gold nanoparticles-conjugated quercetin induces apoptosis via inhibition of EGFR/P38/akt-mediated pathway in breast cancer cell lines (MCF-7 and MDA-MB-231). *Cell Biochem. Funct.* 2017, 35, 217–231. [CrossRef]

158. Bhat, F.A.; Sharmila, G.; Balakrishnan, S.; Arunkumar, R.; Elumalai, P.; Suganya, S.; Raja Singh, P.; Srinivasan, N.; Arunakaran, J. Quercetin reverses EGFR-induced epithelial to mesenchymal transition and invasiveness in prostate cancer (PC-3) cell line via EGFR/P38/Akt pathway. *J. Nutr. Biochem.* 2014, 25, 1132–1139. [CrossRef]

159. Gautam, S.; Praheeshkumar, P.; Budhraja, A.; Son, Y.-O.; Wang, X.; Zhang, Z.; Ding, S.; Wang, L.; Hitron, A.; Lee, J.-C.; et al. Quercetin Inhibits Angiogenesis Mediated Human Prostate Tumor Growth by Targeting VEGFR-2 Regulated AKT/mTOR/P70S6K Signaling Pathways. *PLoS ONE* 2012, 7, e47516.
160. Darveesh, R.S.; Ayoub, N.M.; Nazzal, S. Gold nanoparticles and angiogenesis: Molecular mechanisms and biomedical applications. *Int. J. Nanomed.* **2019**, *14*, 7643–7663. [CrossRef]

161. Seo, S.J.; Lee, S.H.; Kim, K.H.; Kim, J.K. Anti-Fli-IT peptide and cyanine-conjugated gold nanoparticles for the concurrent antiangiogenic and endothelial cell proton treatment. *J. Biomed. Mater. Res. Part B Appl. Biomater.* **2019**, *107*, 1272–1283. [CrossRef] [PubMed]

162. Roma-Rodrigues, C.; Heuer-Jungemann, A.; Fernandes, A.R.; Kanaras, A.G.; Baptista, P.V. Peptide-coated gold nanoparticles for modulation of angiogenesis in vivo. *Int. J. Nanomed.* **2016**, *11*, 2633–2639.

163. Bartczak, D.; Muskens, O.L.; Sanchez-Elsner, T.; Kanaras, A.G.; Millar, T.M. Manipulation of in vitro angiogenesis using peptide-coated gold nanoparticles. *ACS Nano* **2013**, *7*, 5628–5636. [CrossRef]

164. Banerjee, D.; Harfouche, R.; Sengupta, S. Nanotechnology-mediated targeting of tumor angiogenesis. *Vasc. Cell* **2011**, *3*, 3. [CrossRef]

165. Bhattacharyya, S.; Kudqus, R.A.; Bhattacharya, R.; Mukherjee, P. Inorganic Nanoparticles in Cancer Therapy. *Pharm. Res.* **2010**, *28*, 237–259. [CrossRef] [PubMed]

166. Liu, Y.-R.; Guan, Y.-Y.; Luan, X.; Lu, Q.; Wang, C.; Liu, H.-J.; Gao, Y.-G.; Yang, S.-C.; Dong, X.; Chen, H.-Z.; et al. Delta-like ligand 4-targeted nanomedicine for antiangiogenic cancer therapy. *Biomaterials* **2015**, *42*, 161–171. [CrossRef] [PubMed]

167. Hijaz, M.; Das, S.; Mert, I.; Gupta, A.; Al-Wahab, Z.; Tebbe, C.; Dar, S.; Chhina, J.; Giri, S.; Munkarah, A.; et al. Folic acid taggered nanoceria as a novel therapeutic agent in ovarian cancer. *BMC Cancer* **2016**, *16*, 220. [CrossRef]

168. Vassie, J.A.; Whitelock, J.M.; Lord, M.S. Targeted Delivery and Redox Activity of Folic Acid-Functionalized Nanoceria in Tumor Cells. *Mol. Pharm.* **2018**, *15*, 994–1004. [CrossRef]

169. Mousa, S.A.; Yalcin, M.; Bharali, D.J.; Meng, R.; Tang, H.-Y.; Lin, H.-Y.; Davis, F.B.; Davis, P.J. Tetraiodothyroacetic acid and its nanoformulation inhibit thyroid hormone stimulation of non-small cell lung cancer cells in vitro and its growth in xenografts. *Lung Cancer* **2012**, *76*, 39–45. [CrossRef]

170. Li, W.; Yalcin, M.; Bharali, D.J.; Lin, Q.; Godugu, K.; Fujioka, K.; Keating, K.A.; Mousa, S.A. Pharmacokinetics, Biodistribution, and Anti-Angiogenesis Efficacy of Diamino Propane Tetraiodothyroacetic Acid-conjugated Biodegradable Polymeric Nanoparticle. *Sci. Rep.* **2019**, *9*, 9006. [CrossRef]

171. Chwalibog, A.; Hotowy, A.; Grodzik, M.; Wierzbicki, M.; Mitura, K.; Orlowski, P.; Sawosz, E.; Szmidt, M.; Niemiec, T. Nanoparticles of carbon allotropes inhibit glioblastoma multiforme angiogenesis in ovo. *Int. J. Nanomed.* **2011**, *6*, 3041. [CrossRef] [PubMed]

172. Yang, T.; Yao, Q.; Cao, F.; Liu, Q.; Liu, B.; Wang, X. Silver nanoparticles inhibit the function of hypoxia-inducible factor-1 and target genes: Insight into the cytotoxicity and antiangiogenesis. *Int. J. Nanomed.* **2016**, *11*, 6679–6692. [CrossRef] [PubMed]

173. Zhang, L.; Hu, Y. Alphastatin-Loaded Chitosan Nanoparticle Preparation and Its Antiangiogenic Effect on Lung Carcinoma. *Int. J. Polym. Sci.* **2019**, *2019*, 1–9. [CrossRef] [PubMed]

174. Adhikari, H.S.; Yadav, P.N. Anticancer Activity of Chitosan, Chitosan Derivatives, and Their Mechanism of Action. *Int. J. Biomater.* **2018**, *2018*, 1–29. [CrossRef]

175. Wierzbicki, M.; Sawosz, E.; Strojny, B.; Jaworski, S.; Grodzik, M.; Chwalibog, A. NF-κB-related decrease of glioma angiogenic potential by graphite nanoparticles and graphene oxide nanoplatelets. *Sci. Rep.* **2018**, *8*, 14733. [CrossRef]

176. Chakraborty, S.; Ponrasu, T.; Chandel, S.; Dixit, M.; Muthuvijayan, V. Reduced graphene oxide-loaded nanocomposite scaffolds for enhancing angiogenesis in tissue engineering applications. *R. Soc. Open Sci.* **2018**, *5*, 172017. [CrossRef]

177. Dai, Y.; Wang, S.; Shi, W.; Lang, M. pH-responsive carboxymethyl chitosan-derived micelles as apatinib carriers for effective anti-angiogenesis activity: Preparation and in vitro evaluation. *Carbohydr. Polym.* **2017**, *176*, 107–116. [CrossRef]

178. Doddapaneni, R.; Patel, K.; Owaid, I.H.; Singh, M. Tumor neovascularature-targeted cationic PEGylated liposomes of gambogic acid for the treatment of triple-negative breast cancer. *Drug Deliv.* **2015**, *23*, 1232–1241. [CrossRef] [PubMed]

179. Hrynyk, M.; Ellis, J.P.; Haxho, F.; Allison, S.; Steele, J.A.; Abdulkhalak, S.; Neufeld, R.J.; Szewczuk, M.R. Therapeutic designed poly (lactic-co-glycolic acid) cylindrical oseltamivir phosphate-loaded implants impede tumor neovascularization, growth and metastasis in mouse model of human pancreatic carcinoma. *Drug Des. Dev. Ther.* **2015**, *9*, 4573–4586.
180. Wang, J.; Guo, F.; Yu, M.; Liu, L.; Tan, F.; Yan, R.; Li, N. Rapamycin/DiR loaded lipid-polyaniline nanoparticles for dual-modal imaging guided enhanced photothermal and antiangiogenic combination therapy. *J. Control. Release* 2016, 237, 23–34. [CrossRef] [PubMed]

181. Li, X.; Wu, M.; Pan, L.; Shi, J. Tumor vascular-targeted co-delivery of anti-angiogenesis and chemotherapeutic agents by mesoporous silica nanoparticle-based drug delivery system for synergetic therapy of tumor. *Int. J. Nanomed.* 2015, 11, 93–105. [CrossRef] [PubMed]

182. Zhang, J.; Li, J.; Shi, Z.; Yang, Y.; Xie, X.; Lee, S.M.; Wang, Y.; Leong, K.W.; Chen, M. pH-sensitive polymeric nanoparticles for co-delivery of doxorubicin and curcumin to treat cancer via enhanced pro-apoptotic and anti-angiogenic activities. *Acta Biomater.* 2017, 58, 349–364. [CrossRef] [PubMed]

183. Li, Y.; Wu, Y.; Huang, L.; Miao, L.; Zhou, J.; Satterlee, A.B.; Yao, J. Sigma receptor-mediated targeted delivery of anti-angiogenic multifunctional nanodrugs for combination tumor therapy. *J. Control. Release* 2016, 228, 107–119. [CrossRef] [PubMed]

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