Biological active compounds and antioxidant activity of plants from the collection of Central Siberian Botanical Garden. II. Lamiaceae

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Abstract. The contents of the major groups of biologically active compounds (catechins, flavonoids, tannins, saponins, pectin, and carotenoids) and total antioxidants (TA) in the leaves and the flowers of Agastahe rugosa (Fisch. & C.A. Mey.) Kuntze, Betonica officinalis L., Dracocephalum nutans L., Mentha caucasica Gand., Nepeta grandiflora M. Bieb., Origanum vulgare L., Scutellaria baicalensis Georgi (Lamiaceae), collected during the flowering period of 2019 in "Collections of living plants indoors and outdoors" USU 440534 of Central Siberian Botanical Garden SB RAS, Novosibirsk, were studied. The concentrations of constituent majority were higher in leaves compared to flowers. In the leaves, maximums of catechins (15.60 mg·g⁻¹, B. officinalis), flavonoids (61.5 mg·g⁻¹, S. baicalensis), saponins (178.5 mg·g⁻¹, D. nutans), and propectin (106.0 mg·g⁻¹, B. officinalis) were revealed. In the flowers, maximums of tannins (247.4 mg·g⁻¹, O. vulgare) and pectin (16.31 mg·g⁻¹, S. baicalensis) were found. Maximum of total antioxidant contents (TAC) (6.21 mg·g⁻¹) was detected in the leaves of O. vulgare. Positive correlations between TAC and the content of tannins and flavonoids were revealed. The results confirm health benefits of the studied species and contribute to the knowledge of the distribution of saponins and pectins in the Lamiaceae.

1 Introduction

Lamiaceae Lindl. is a large plant family of mostly shrubs and herbs, which are popular due to various biological effects on human health, including antioxidant and antimicrobial properties closely associated with a variety of secondary metabolites [1]. Numerous members of the Lamiaceae family have been used in folk medicine for many years. Currently, antitussive, diuretic, anti-asthmatic, antiseptic, antispasmodic, and febrifuge activities of the herbs were revealed [2]. Some species displayed antiviral properties [3]. Chemical composition of Lamiaceae members combines a variety of terpenoids with iridoid glycosides, flavonoids and other phenolic compounds [4]. Carotenoids and flavonoids were reported to be crucial determinants of antioxidant properties of plants [5]. Lamiaceae species were also examined for pectic substances [6]. Pectin was proved to have diverse biological activities including lipid and cholesterol level lowering effects, serum glucose

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content lowering effects, and anti-cancer activities [7]. In addition, polysaccharides of pectin possess antioxidant activity and may provide auxiliary functions for other active antioxidant components [8]. Meanwhile, variation of health promoting compounds through locations and plant body parts are in focus of current investigations with particular emphasis on the constituents with high antioxidant activity [9].

The collection of Central Siberian Botanical Garden of Siberian Branch of Russian Academy of Sciences (CSBG SB RAS) USU 440534 in Novosibirsk contains the members of several Lamiaceae genera known for their positive health effects. Most of them belong to the Nepetoideae Burnett subfamily. It is the most numerous in the Lamiaceae family and it is considered a source of antioxidants [1]. Betonica officinalis L. (Lamiioideae Harley) and Scutellaria baicalensis (Scutellarioideae Prantl) were documented to have medical properties [2].

The aim of this study was to determine the profiles of biological active compounds (in terms of quantities of selected major groups of compounds in the leaves and flowers) of 7 Lamiaceae species and to compare their antioxidant properties.

2 Material and methods

The leaves and flowers of Agastahe rugosa (Fisch. & C.A. Mey.) Kuntze, Betonica officinalis L., Dracecephalum nutans L., Mentha caucasica (Gand.) Briq., Nepeta grandiflora M. Bieb., Origanum vulgare L., Scutellaria baicalensis Georgi were collected during the flowering period of 2019 from “Collections of living plants indoors and outdoors” USU 440534 of CSBG SB RAS in the garden plot located in a forest-steppe region with gray forest soil and the mean yearly temperature/precipitation of 1.8 °C/448 mm. Air-dried plant material were ground up using a household mill. Weighted samples were extracted according to the proper methods. Dry matter concentrations in the samples were calculated by the gravimetric method. Total catechin contents (C) were determined in the ethanol extracts by the vanillin-HCl method [10]. Total flavonoid contents (F) determination was performed by spectrophotometry at 415 nm after a reaction of complexation of flavone and flavonols with aluminium chloride in the aqueous ethanol extract. The flavonoid concentrations were calculated as rutin equivalents [11]. For hydrolysable tannin (T) estimation, spectrophotometric measurement of intensity of reaction of aqueous extract of the leaves and flowers with a 2% aqueous solution of ammonium molybdate at 420 nm was carried out. The state standard of tannin was used as outside standard [12]. Total saponins contents as crude saponin (S) were determined by the gravimetric method. Powdered sample was extracted with chloroform using Soxhlet extractor to remove lipids and resins. Then, the residue was extracted by 50, 60, 96% ethanol sequentially (twice each concentration for 30 min) at T=70 °C. The combined extract was evaporated to 5 ml, and a 7-fold volume of acetone was added. After 18 hours, the resulting precipitate was filtered out, dried at 70°C, weighed, and the content of crude saponin was calculated [13]. The contents of pectins (P) and propectins (PP) (after hydrolysis) were determined by spectrophotometric method (480 nm) based on obtaining a specific yellow-orange staining of uronic acids with thymol in a sulfuric acid medium after removal sugars from the samples. The percentage of pectic substances was calculated using a calibration curve for galacturonic acid from Sigma (St. Louis, MO, USA) [14]. The total carotenoids (Car) was determined spectrophotometrically after extraction by acetone and ethanol consecutively. The absorption of the solution was measured at wavelengths corresponding to absorption maxima of chlorophyll a (Cₐ) (662 nm), chlorophyll b (C₈) (644 nm), and carotenoids (440.5 nm) [15]. All above tests were performed in triplicate, and the biochemical parameters were calculated in mg per g of absolutely dry weight (mg·g⁻¹ DW).
Total antioxidant contents (TAC) was determined in aqueous ethanol extracts (70 % v/v) by amperometric method using flow injection system with amperometric detection “Tsvet Yauza-01-AA” and calculated as gallic acid (GA) equivalents [16]. These tests were performed in five replication. All the data were processed in the Statistica 10.0 software (Statsoft Inc., Tulsa, OK, USA), were reported as mean ± standard error of three (or five) replicates, and were compared by Duncan’s multiple range test. Differences between the means were considered statistically significant at the 5% level (p < 0.05). Linear regression was calculated by means of Multiple Regression analysis. Reliability of the influence of the factors (‘species’ and ‘plant organ’) was evaluated by two-way ANOVA.

### 3 Results

The results of the tests showed a significant variation in all parameters by the species (Table). The table shows the parameters of the species that are most rich in biological active compounds.

**Table.** Concentrations of biological active compounds (mg·g⁻¹ DW) and antioxidant activities (mg gallic acid·g⁻¹ DW) of the Lamiaceae species

|                  | Betonica officinalis | Dracocephalum nutans | Nepeta grandiflora | Origanum vulgare | Scutellaria baicalensis |
|------------------|----------------------|----------------------|--------------------|------------------|------------------------|
| **C**            | 15.60 ± 0.02²       | 1.04 ± 0.01²         | 0.97 ± 0.01²       | 2.09 ± 0.01²     | 1.39 ± 0.01²           |
| 3.07 ± 0.01³     | 0.94 ± 0.00²        | 0.66 ± 0.01²         | 1.45 ± 0.00³      | 1.11 ± 0.01³      |
| **T**            | 125.3 ± 1.8³        | 202.8 ± 1.8³         | 149.8 ± 1.3³      | 242.4 ± 4.7³     | 247.4 ± 2.1³           |
| 89.0 ± 1.3³      | 205.9 ± 1.8³        | 79.5 ± 1.5³          | 205.2 ± 3.7³     | 90.8 ± 1.5³       |
| **F**            | 21.9 ± 0.6³         | 60.2 ± 0.8³          | 19.8 ± 0.4³       | 48.4 ± 0.9³      | 61.5 ± 1.1³            |
| 9.2 ± 0.2³       | 47.0 ± 0.6³         | 7.6 ± 0.1³           | 43.0 ± 0.4³      | 39.7 ± 0.5³       |
| **P**            | 4.96 ± 0.07²        | 5.28 ± 0.05²         | 8.99 ± 0.00³     | 5.21 ± 0.10³     | 14.62 ± 0.18³          |
| 7.27 ± 0.16²     | 6.15 ± 0.10²        | 7.94 ± 0.08³         | 4.06 ± 0.03³     | 16.31 ± 0.43³     |
| **PP**           | 106.0 ± 0.8²        | 68.2 ± 0.2³          | 103.6 ± 1.9²     | 81.7 ± 0.8²      | 97.7 ± 0.5²            |
| 59.2 ± 0.5³      | 78.3 ± 0.1³         | 67.5 ± 1.00³         | 86.2 ± 0.5³      | 59.5 ± 0.8³       |
| **Car**          | 0.80 ± 0.01³        | 0.39 ± 0.01³         | 0.64 ± 0.01³     | 0.72 ± 0.01³     | 0.91 ± 0.01³           |
| 0.13 ± 0.00³     | 0.19 ± 0.00³        | 0.12 ± 0.00³         | 0.33 ± 0.00³     | 0.06 ± 0.00³     |
| **S**            | 70.6 ± 3.0²         | 178.5 ± 4.8³         | 115.2 ± 5.8³     | 177.3 ± 5.5³     | 115.2 ± 5.6³           |
| 57.2 ± 3.7³      | 94.5 ± 6.3³         | 147.9 ± 7.2³        | 126.7 ± 5.2³     | 149.5 ± 5.8³     |
| **TAC**          | 0.30 ± 0.02²        | 2.04 ± 0.03³         | 0.66 ± 0.02³     | 6.21 ± 0.01³     | 1.75 ± 0.01³           |
| 0.43 ± 0.01³     | 1.54 ± 0.00³        | 0.46 ± 0.01³         | 7.32 ± 0.03³     | 1.30 ± 0.00³     |

Parameters of the leaves are in numerator, and parameters of the flowers are in denominator; means followed by the same letter are not significantly different according to Duncan’s test (p < 0.05)

Total catechin content and TAC varied most significantly. Meanwhile, ranges of variability of these parameters in the leaves were more widely compared to these in the flowers. Protopectin and tannin were the least variable. Ranges of variability of protocatechin in the leaves and in the flowers were identical (1.5 times), and tannins in the flowers varied greater (3.1 times) than in the leaves (1.6 times). The rest of the studied parameters varied 3-6 times, and ranges of their variability in the flowers were the same or wider compared to the leaves.

The concentrations of majority of constituents in the leaves were higher compared to the flowers. In the leaves, a high contents of catechins (15.60 mg·g⁻¹ in _B. officinalis_), flavonoids (61.5  and 60.2 mg·g⁻¹ in _S. baicalensis_ and _D. nutans_, respectively), tannins (242.4, 205.2, and 202.8 mg·g⁻¹ in _O. vulgare_, _S. baicalensis_, and _D. nutans_), saponins (178.5 and 177.3 mg·g⁻¹ in _D. nutans_ and _O. vulgare_), pectin (14.62 mg·g⁻¹ in _S. baicalensis_), and protocatechin (106.0 and 103.6 mg·g⁻¹ in _B. officinalis_ and _N. grandiflora_) were noted. In the flowers, a high contents of tannins (247.4 and 205.9 mg·g⁻¹ in _O. vulgare_...
and *D. nutans* and maximal content of pectin (16.31 mg·g⁻¹ in *S. baicalensis*) were determined.

Maximum of TAC (6.21 mg·g⁻¹ GA) was found in the leaves of *O. vulgare*. *S. baicalensis* thus had the largest number of high (tannins) and maximal (flavonoids, pectin, carotenoids) biochemical parameters, but its TAC was low (1.75 mg·g⁻¹ GA). Meanwhile, *O. vulgare* with maximal TAC possessed highest tannin content. The correlation between TAC and total tannin content of the samples was revealed (Fig. 1), as well as between TAC and total flavonoid content.

The obtained results contribute to knowledge of distribution of these multifunctional bioactive compounds in representatives of Lamiaceae. Pectin content in the studied samples varied significantly, and maximal levels of pectin in the leaves and flowers of *S. baicalensis* correspond with high levels of pectin in some species of this family [18]. The found correlation between total flavonoid contents and total tannin contents, and TAC corresponds with similar correlations revealed previously [19-21].

### 4 Conclusions

The results adds an information to the data on the patterns of biological active compounds of the species *Agastahe rugosa*, *Betonica officinalis*, *Dracocephalum nutans*, *Mentha caucasica*, *Nepeta grandiflora*, *Origanum vulgare*, and *Scutellaria baicalensis*. High levels of flavonoids (including catechins), tannins and pectins coupled with carotenoids and saponins provide antioxidant properties and point to a good therapeutic potential of these species.
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References

1. K. Carović-Stanko, M. Petek, M. Grdiša, J. Pintar, D. Bedeković, M. Herak Ćustić, Z. Šatović, Czech J. Food Sci. 34, 377 (2016)
2. M. Uritu, C.T. Mihai, G. Stanciu, G. Dodi, T. Alexa-Stratulat, A. Luca, M. Leon-Constantin, R. Stefanescu, V. Bild, S. Melnic, B.I. Tamba, Pain Res. Manag. 2018, 7801543 (2018)
3. I.E. Lobanova, E.I. Filippova, G.I. Vysochina, N.A. Mazurkova, Plant Life of Asian Russia 2, 64 (2016)
4. F. Naghibi, M. Mosaddegh, S. Mohammadi, A. Ghorbani, Iran J. Pharm. Res. 2, 63 (2005)
5. D.P. Xu, Y. Li, X. Meng, T. Zhou, Y. Zhou, J. Zheng, J.J. Zhang, H.B. Li, Int. J. Mol. Sci. 18, 96 (2017)
6. E.P. Kukhta, V.Y. Chirva, M.R. Borovskii, M.A. Tolmacheva, I.V. Eroshenko, Chem. Nat. Compd. 18, 251 (1982)
7. A. Wikiera, M. Irla, M. Mika, Postepy Hig. Med. Dosw. 68, 590 (2014)
8. S.T. Minzanova, V.F. Mironov, D.M. Arkhipova, A.V. Khabibullina, L.G. Mironova, Y.M. Zakirova, V.A. Milyukov, Polymers (Basel) 10, 1407 (2018)
9. I.B.H. Yahia, R. Jaouadi, R. Trimech, M. Boussaid, Y. Zaouali, Biochem. Syst. Ecol. 84, 8 (2019)
10. T.A. Kukushkina, A.A. Zykov, L.A. Obukhova Common cuff (Alchemilla vulgaris L.) as a source of medicines, in Proceedings of the VII International Congress on Actual Problems of Creating New Drugs of Natural Origin, 3-5 July 2003, St. Petersburg, Russia (2003)
11. V.V. Belikov, M.S. Shreiber, Farmatsiya 1, 66 (1970)
12. L.M. Fedoseeva, Chemistry of Plant Row Material, 3, 45 (2005)
13. A.V. Kiseleva, T.A. Volkhnoskaya, V.E. Kiselev, Biologically Active Substances of Medicinal Plants of Southern Siberia (Nauka, Novosibirsk, 1991)
14. V.I. Kriventsov, Proceedings of State Nikitsky Botanical Garden 109, 128 (1989)
15. Methods of biochemical research of plants, ed. by A.I. Yermakov (Leningrad, 1987)
16. P.A. Fedina, A.Ya., N.I. Yashin, Chemistry of Plant Row Material, 2, 91 (2010)
17. M. Shanaida, O. Golembiovska, N. Hudz, P. Wieczorek, Curr. Issues Pharm. Med. Sci. 31, 194 (2018)
18. J. Lu, J. Li, R. Jin, S. Li, J. Yi, J. Huang, Int. J. Biol. Macromol. 131, 323 (2019)
19. R. Amarowicz, A. Troszyńska, N. Barylko–Pikielna, J. Food Lipids 11, 278 (2004)
20. V. Katalinic, M. Milos, T. Kulisic, M. Jukic, Food Chem. 94, 550 (2006)
21. R. Farhoudi, Free Radical Bio. Med. 108, 24 (2017)