Research Article

A Post–Mortem Examination of COVID–19 Pulmonary Pathology in 9 Cases

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Abstract

A new novel virus called SARS-CoV-2 has expanded into a pandemic in the past several months. The virus is an acute respiratory RNA virus that has symptoms in three clinical groups: asymptomatic, suspicious, and COVID-19 positive. The clinical lab tests used for diagnosis are Nasalpharyngeal swabs, with further testing done with sputum or BAL samples. Serological samples are collected for diagnosis in deceased patients using RT-PCR. The clinical symptoms usually occur 2 to 14 days after exposure and include fever, dry cough, and fatigue. In some cases, symptoms can progress and cause multiple organ failure due to adult respiratory distress syndrome (ARDS). The virus is in the family of coronaviruses and has also been identified in lung tissue using transmission electron microscopy. Gross and microscopic lung pathology was examined in five positive cases and four negative cases by hematoxylin and eosin (H&E) and Masson’s Trichrome stains. Of the collected, the age range was 28 to 76. The ethnicities were six Caucasians and three minorities, with a male to female ratio of 7:2. The salient histology features seen in the study were multifocal to diffuse alveolar necrosis, bronchiolar epithelial necrosis, and interstitial mononuclear lymphocytic infiltrates. Other features were perivascular and peribronchiolar lymphoid infiltrates and marked congestion. Scattered fibroplasia was found in the damaged alveoli and the alveolar septae in the more severe cases. These pathologic features are similar to other coronaviruses.

Introduction

Since December 2019, the infectious disease agent SARS–CoV–2 has spread across the globe affecting people on every continent except Antarctica. In general, the virus can cause acute respiratory disease with severe symptoms that can lead to death [1–3]. The clinical lab tests used for diagnosis are Nasalpharyngeal swabs, with further testing done with sputum or BAL samples [4–8]. As of now, COVID–19 has infected over 4.4 million people worldwide, but this number is increasing constantly. The death rate is approximately 2% [1]. Severe symptoms causing death is due to severe alveolar and small bronchiolar damage with subacute to chronic inflammation [1–3,9]. Recent studies have shown SARS–CoV–2 to have a high affinity for the ACE2 receptor found in ciliated bronchial epithelium [10]. The ACE2 receptor triggers a protective effect in the bronchial epithelium against ARDS [11]. With this pathway compromised by SARS–CoV–2, there is severe damage to the bronchiolar epithelium and an increase of ARDS (Figure 1).

Figure 1: A) Focal interstitial pneumonia characterized by alveolar wall thickening. B) Bronchiole epithelial necrosis with hyaline membrane disease. C) Bronchiole epithelial necrosis surrounded by peribronchiolar lymphocytic infiltration and marked congestion. D) Large area of marked fibroplasia with focal lymphocytic infiltration and bronchiolar necrosis.
In this paper, the lung pathology of 5 patients who had died from severe COVID-19 symptoms were analyzed along with 4 negative COVID-19 cases (Table 1). These samples were collected post-mortem, and patients were not seen in the clinical setting. The pathology findings from these cases will add to the pathogenesis of this novel infectious viral disease.

Materials and methods

Informed consent and IRB approval was obtained for this decedent study. All lung and blood specimens were collected from the Marion County Coroner’s Office and private funeral homes. These were collected from partial autopsies over the period of April 9th to May 7th. The cases collected were deemed COVID-19 suspicious or COVID-19 positive based on the medical forensic coroner’s decision at the scene of death. All cases were then tested by the Indiana Department of Health Lab using a CDC approved RT-PCR method and then were classified into COVID-19 positive and COVID-19 negative. Of the collected, the age range was 28 to 76. The ethnicities were six Caucasians and three minorities, with a male to female ratio of 7:2.

Most COVID-19 patients that died in hospitals in the central Indiana area were sent directly to funeral homes. The collected cases were found dead in their homes, with one in the nursing home. Cases are designated as a coroner’s case based on dying at home. Cases are designated as a coroner’s case based on dying in their homes and not in a hospital setting. Therefore, medical history is limited and only fragments can be collected from next of kin.

The blood was drawn from the heart using an autopsy needle and sent to the Indiana Department of Health Laboratories. This testing is required by the state on COVID-19 suspicious cases, and the test results were used to classify each patient as COVID-19 positive or COVID-19 negative. Next, a four centimeter incision was made in the chest between the 4th and 5th ribs on the right side. A section of lung approximately 3 cm by 2 cm was removed and placed in 10% NBF.

The fixation time in 10% NBF was extended to 48 hours due to the infectious nature of the disease. Usual fixation time for most tissues is approximately 24 hours. When the tissues were fixed, they were trimmed and placed into tissue cassettes. Fourteen tissue cassettes were prepared for each case. The tissues were then transferred to 70% ethanol before processing into a paraffin block. Five micron sections were microtomed and stained with H&E and Masson’s Trichrome stains.

Results

Histology from the 5 positive cases revealed multifocal to diffuse alveolar necrosis and bronchioral respiratory epithelial necrosis. Interstitial mononuclear inflammatory infiltrates, mainly lymphocytes, were seen in a multifocal pattern throughout the biopsies. Perivascular and peribronchioral lymphoid infiltrates were also seen throughout the biopsies, along with marked congestion. Scattered fibroplasia can also be seen in the severe cases, extending into alveolar spaces and thickening the alveolar septum (Figure 1a,b,c,d). In one case, there was mild hyaline membrane formation and slight microthrombi formations in small pulmonary vasculature. Another case was found to have aspiration pneumonia. This was characterized by plant fiber and other foreign material, along with numerous neutrophils in the lumen of the bronchioles.

The 4 negative cases were characterized by diffuse pulmonary edema and marked congestion. There was no evidence of pneumonia or inflammation in the airways or alveoli.

Discussion

The lung pathological features of COVID-19 found in this paper are similar to SARS, MERS, and a recent published COVID-19 clinical case with pathology [1,8,12,13]. Based on the consistent findings in these cases, the main pathology change seen in all diseases listed above was diffuse alveolar damage [12,13]. In addition, this study found a complete loss of the tertiary bronchioral epithelial layers also associated with peribronchioral lymphoid hyperplasia in severe cases. Recent studies have shown SARS–CoV–2 to have a high affinity for the ACE2 receptor found in ciliated bronchial epithelium, and when induced by SARS–CoV–2 there is an increase of tissue damage [10,11].

Table 1: Patient demographics and COVID-19 test results for the 9 cases analyzed.

| Patient ID | Age | Decedent Sex | Medical History | Race | Smoking Status | COVID-19 Status | Hospital Admission, length of stay |
|------------|-----|--------------|----------------|------|----------------|----------------|-------------------------------------|
| 1          | 56  | Male         | Diabetes/Fever | Black| Yes            | Confirmed Covid19 | N/A                                |
| 2          | 46  | Male         | Diabetes/Coughing | White| Unknown       | Negative Covid19 | N/A                                |
| 3          | 28  | Male         | Hypertension/Seizures | White| Marijuana | Confirmed Covid19 | Discharged From hospital 4/10/2020 |
| 4          | 54  | Male         | Shortness of Breath | Black| No           | Negative Covid19   | N/A                                |
| 5          | 34  | Male         | Diabetes/Vomiting | White| Unknown | Negative Covid19 | N/A                                |
| 6          | 42  | Female       | Asthma/High Blood Pressure | White| Unknown | Negative Covid19 | In ER approximately 10 min       |
| 7          | 76  | Female       | N/A             | N/A  | Unknown       | Confirmed Covid19 | N/A                                |
| 8          | 67  | Male         | Shortness of Breath | White| Unknown | Confirmed Covid19 | Nursing Home                       |
| 9          | 53  | Male         | Flu Symptoms    | White| Unknown | Confirmed Covid19 | N/A                                |

References

1. Zangrillo A, Beretta L, Scandroglio AM, Monti G, Fominskiy E, et al. (2020) Pathological findings in COVID-19 associated with acute respiratory distress syndrome. Lancet Respir Med 8: 420-422. Link: https://bit.ly/3dKj1kO

2. Gattinoni L, Coppola S, Cressoni M, Busana M, Rossi S, et al. (2020) Covid-19 Does Not Lead to a “Typical” Acute Respiratory Disease Syndrome. Am J Respir Crit Care Med 201: 1299-1300. Link: https://bit.ly/2BKRAZZ

3. Xu Z, Shi L, Wang Y, Zhang J, Huang L, et al. (2020) Pathological findings of COVID-19 associated with acute respiratory distress syndrome. Lancet Respir Med 8: 420-422. Link: https://bit.ly/3dKj1kO

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Patients with Lung Cancer. J Thoracic Oncology 15: 700-704. Link: https://bit.ly/2Yb4TsA

10. Alanagreh L, Alzoughool F, Atoum M (2020) The Human Coronavirus Disease COVID-19: Its Origin, Characteristics, and Insights into Potential Drugs and Its Mechanisms. Pathogens 9: 331. Link: https://bit.ly/3dYCN5

11. Bourgonje A, Abdulle AE, Timens W, Hillebrands JL, Navis GJ, et al. (2020) Angiotensin-converting enzyme-2 (ACE2), SARS-CoV-2 and pathophysiology of coronavirus disease 2019 (COVID-19). J Pathol Link: https://bit.ly/37dkjIX

12. Ding Y, Wang H, Shen H, Li Z, Geng J, et al. (2003) The clinical pathology of severe acute respiratory syndrome (SARS): a report from China. J Pathol 200: 282-289. Link: https://bit.ly/2ASxRhU

13. Ng DL, Al Hosani F, Keating MK, Gerber SI, Jones TL, et al. (2014) Clinicopathologic, immunohistochemical, and ultrastructural findings of a fatal case of Middle East respiratory syndrome coronavirus infection in the United Arab Emirates. Am J Pathol 186: 652-658. Link: https://bit.ly/2BLSLbf

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