Sertraline Pharmacokinetics in HIV-Infected and Uninfected Children, Adolescents, and Young Adults

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On behalf of the IMPAACT P1080 Protocol Team

Objective: Due to potential disease and drug interactions, the appropriate sertraline starting dose and titration range may require adjustment in pediatric patients living with HIV. This is the first report of sertraline pharmacokinetics in HIV-infected youth.

Methods: IMPAACT P1080 was a multicenter pilot study describing psychiatric medication pharmacokinetics in HIV-infected and uninfected youth. Participants were stable on sertraline, >6 to <25 years old, and (1) HIV-uninfected (HIV(–)), (2) HIV-infected taking efavirenz (EFV), or (3) HIV-infected taking boosting ritonavir/protease inhibitor (PI/r). Sampling occurred at pre-dose, 2, 4, 6, 12, and 24-h post-dose. Analyses were performed for sertraline and N-desmethylsertraline, and CYP2D6 phenotyping was completed with dextromethorphan.

Results: Thirty-one participants (16 HIV(-), 12 PI/r, and 3 EFV) had median (range) weight, age, and dose of 69.5 (31.5–118.2) kg, 21.8 (9.1–24.7) years, and 75.0 (12.5–150.0) mg once daily. Sertraline exposure was highest for HIV(–) and lowest for EFV cohorts; median dose-normalized $AUC_{0-24}$ was 1176 (HIV(–)), 791 (PI/r) and 473 (EFV) ng·hr/mL, and $C_{24}$ was 32.7 (HIV(–)), 20.1 (PI/r), and 12.8 (EFV) ng/mL. The urinary dextromethorphan/dextrorphan (DXM/DXO) ratio was higher in HIV(–) vs. PI/r cohorts ($p = 0.01$). Four HIV(–) participants were CYP2D6 poor metabolizers (ln(DXM/DXO) of > -0.5).

Conclusions: HIV(–) cohort had the highest sertraline exposure. Sertraline exposure was ~40% lower in the PI/r cohort than in HIV(–); the need to alter sertraline dose ranges for PI/r participants is not clear. The impact of efavirenz on sertraline needs further investigation due to limited numbers of EFV participants.

Keywords: sertraline, pharmacokinetics, HIV, pediatrics, antiretrovirals
INTRODUCTION

The lifetime prevalence of major depression in patients with human immunodeficiency virus (HIV) is 22–45% (1), far exceeding the general population. Youth living with HIV are four times as likely to be treated with anti-depressant medication than uninfected youth (2). Two meta-analyses demonstrated a strong correlation between severity of depression and non-adherence to antiretroviral therapy (3, 4). Therefore, standard practice is to treat patients living with HIV and comorbid psychiatric conditions with psychotropic medications, despite a lack of adequate evidence regarding the risks. Psychiatric medications are often titrated to effectiveness. However, the typical starting dose and titration range may need to be adjusted in special populations (e.g., HIV and pediatrics) to produce exposures that have shown efficacy and tolerability in standard patient populations lacking comorbid conditions and interacting drugs.

Sertraline, a commonly used antidepressant, has an elimination half-life of about 27 h in adults, children, and adolescents (5). Sertraline undergoes extensive metabolism by cytochrome P450 (CYP) enzymes, CYP2B6, CYP2C9, CYP2C19, CYP2D6, and CYP3A4 (6, 7). N-demethylation forms N-desmethylsertraline, the major inactive metabolite. N-desmethylsertraline has an elimination half-life of 62–104 h (5). Genetic differences in metabolic pathways can lead to interpatient variability, with higher exposures in poor metabolizers and lower exposures in extensive metabolizers. A rough estimate of CYP2D6 intrinsic activity can be determined by administering a single dose of dextromethorphan and measuring the extent of metabolite (dextrorphan) formation compared to parent compound (a CYP2D6 phenotype assessment). Based on concentration ratio results, patients can be classified as either extensive or poor metabolizers. Numerous antiretrovirals are metabolized by and modulators of many of these same biotransformation pathways.

Ritonavir, a protease inhibitor and pharmacokinetic booster of other antiretrovirals, inhibits CYP3A4, the efflux transporter P-glycoprotein, and to a lesser extent, CYP2D6 (8, 9). However, at typical ritonavir boosting doses (100 mg), CYP2D6 inhibition is not clinically relevant (8). Ritonavir also induces CYP1A2, 2B6, 2C9, and 2C19, and the phase II uridine diphosphateglucuronosyltransferase (UGT) enzymes (10–12).

Efavirenz, a common component of first-line antiretroviral therapy regimens, is a strong inducer of CYP3A4 and 2B6 (13). Efavirenz also induces CYP 2C19 in extensive and intermediate metabolizers, but not in CYP 2C9 poor metabolizers (14). Efavirenz decreases sertraline exposure (AUC₀–₂₄) by ~40% in adults. The induction of CYP 3A4 in particular, which has high concentrations in both hepatic and intestinal tissues, could contribute to decreased absorption through first-pass loss since sertraline is administered orally. Despite decreased sertraline exposure with efavirenz, no adjustment of starting dose is recommended (13).

In pediatric sertraline pharmacokinetic studies, sertraline exposure (AUC₀–₂₄ and Cmax) was ~22% lower in children and adolescents when plasma concentration was adjusted for weight (5, 9)/; other pharmacokinetic parameters were comparable to adults (15–19). In adolescents, sertraline kinetics are not linear; the half-life of sertraline shortens significantly (26.7–15.3 h) from single dose to steady-state (17). In addition, over a dose range of 50–200 mg, the half-life of sertraline at steady-state increases from 15.3–27.2 h (15, 17). The non-linearity of sertraline kinetics becomes increasingly important in youth with complex diseases and interacting medications. The objective of this study was to determine sertraline pharmacokinetics in youth without HIV, or with HIV and taking either an efavirenz-based or a ritonavir-boosted protease inhibitor-based regimen.

MATERIALS AND METHODS

International Maternal Pediatric Adolescent AIDS Clinical Trials Network (IMPAACT) P1080 was a multicenter, pilot study of psychiatric and antiretroviral medication concentrations in HIV-1-infected and uninfected youth. Target enrollment for the sertraline study arm was 45 participants >6 to <25 years of age. 45 participants had sufficient power to detect a 50% difference in apparent oral clearance (CL/F) among HIV-infected and uninfected cohorts, assuming a coefficient of variation of 30%. Participants were divided into three cohorts: (1) HIV-uninfected (HIV−), (2) HIV-infected taking concomitant efavirenz (EFV), and (3) HIV-infected taking boosting ritonavir with a protease inhibitor (PI/r): protease inhibitors could include atazanavir, darunavir, fosamprenavir, indinavir, lopinavir, saquinavir or tipranavir.

All participants gave signed informed consent or assent, and parents or guardians gave signed informed permission in accordance with the Declaration of Helsinki and local guidelines. The protocol was approved by human subjects’ protection committees or institutional review boards at each participating site (see Acknowledgments for a listing of participating sites). Participants took sertraline for clinical care for at least 2 weeks prior to enrollment. Participants who were pregnant or taking interacting illicit drugs were excluded.

Study procedures were standardized across multiple study arms. The single pharmacokinetic visit included a medication history, adherence survey, and a CYP2D6 phenotype assessment (the other psychotropic study medications were primarily metabolized by CYP 2D6). Participants with HIV had to be taking their antiretroviral medications consistently for at least 4 weeks prior to sampling. The visit was scheduled so that a witnessed dose of sertraline and antiretroviral medications occurred on time, according to the participant’s dosing regimen. Plasma samples were drawn at pre-dose and 2, 4, 6, 12, and 24-h post-dose.

A liquid chromatography-electrospray ionization-mass spectrometry method quantitated sertraline and N-desmethylsertraline. Isotope-labeled sertraline was used as an internal standard. Analytes and internal standard were extracted with 600 µL of acetonitrile from 200 µL of plasma, then eluted from a Gemini C18 column in under 4 min using acetonitrile/water/formic acid (85:15:0.1, v/v/v) as the mobile phase. Quantitation was performed with selective reaction monitoring of the transitions of m/z 306.2→159.1 for sertraline,
RESULTS

Thirty-one participants completed pharmacokinetic visits \((n = 16\text{ HIV}(–); n = 3\text{ EFV}; n = 12\text{ PI/r}; 5\text{ on atazanavir/ritonavir, 5\text{ on darunavir/ritonavir, and 2\text{ on lopinavir/ritonavir}.}\)) The median weight and height of participants on the day of sampling were 69.5 kg and 167.2 cm, respectively (Table 1). The median age was 21.8 years (range 9–24.7). Participants’ daily sertraline doses ranged from 12.5 to 150 mg. Median weight-normalized dose in HIV(–) (1.3 mg/kg) was higher than in both the PI/r and EFV groups (0.9 and 0.7 mg/kg; Table 1). A total of 181 plasma concentrations were measured. Two participants did not return for their 24-h time points, while three participants took their next dose of sertraline prior to the 24-h blood draw. Pharmacokinetics were estimated based on the pre-dose through 12 h post-dose concentrations for these participants.

Median normalized \(\text{AUC}_{0→24}\) values were 1176, 791, and 473 ng·h/mL for HIV(–), PI/r and EFV cohorts, respectively (Table 2 and Figure 1). Non-compartmental oral CL/F were not different in the HIV(–) and PI/r groups (1.4 vs. 1.6 L/h/kg, \(p = 0.59\)). However, CL/F was markedly higher in the EFV group (4.5 L/h/kg). Of C₀, Cmax, and C24, only C₀ was significantly higher in the HIV(–) compared to the PI/r cohorts (unadjusted and dose-normalized, \(p = 0.03\)).

Sertraline typical population pharmacokinetic values for CL/F, V/F, and \(k_{a}\) were estimated for the structural base model at 4.77 L/hr/kg\(^{0.75}\), 53.8 L/kg, and 0.45 hr\(^{-1}\) respectively (Table 3). The final population model (objective function decreased by 8.053 compared to the base model), which included the effect of age on clearance, resulted in estimated population values for CL/F, V/F, and \(k_{a}\) of 4.35 L/hr/kg\(^{0.75}\), 44.6 L/kg, and 0.42 hr\(^{-1}\).

Median normalized N-desmethylsertraline \(\text{AUC}_{0→24}\) in the PI/r group was significantly lower than in the HIV(–) group (899 vs. 1,533 ng·h/mL, \(p = 0.01\); Table 2 and Figure 1). Following normalization, median Cmax in HIV(–) was

### TABLE 1 | Participant demographics, Median (Interquartile Range)\(^{a}\).

| HIV(–) | PI/r | EFV |
|--------|------|-----|
| \(n = 16\) | \(n = 12\) | \(n = 3\) |
| Weight (kg) | 65 (58, 77) | 73 (69, 77) | 58 (45, 82) |
| Height (cm) | 166 (163, 172) | 169 (165, 175) | 152 (145, 165) |
| Weight Normalized Daily Dose (mg/kg) | 1.3 (0.9, 1.5) | 0.9 (0.6, 1.4) | 0.7 (0.6, 1.1) |
| Age (years) | 22.8 (18.2, 23.3) | 21.8 (20.9, 22.7) | 19.3 (14.2, 19.5) |
| SEX (%) | | | |
| Female | 10 (62.5) | 8 (66.7) | 2 (66.7) |
| Male | 6 (37.5) | 4 (33.3) | 1 (33.3) |
| RACE (%) | | | |
| American Indian | 1 (6.2) | 0 (0.0) | 0 (0.0) |
| Asian | 1 (6.2) | 0 (0.0) | 0 (0.0) |
| Black | 2 (12.5) | 11 (91.7) | 2 (66.7) |
| Unknown | 0 (0.0) | 1 (8.3) | 0 (0.0) |
| White | 12 (75.0) | 0 (0.0) | 1 (33.3) |

\(^{a}\)For the EFV cohort, median (minimum, maximum) are reported.

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292.2 → 275 for N-desmethylsertraline, and 309.2 → 159.1 for internal standard. The method was linear over the concentration range of 1–320 ng/mL for both parent and metabolite.

The ratio of dextromethorphan (DXM)/dextrophan (DXO) in urine was measured using a validated LC-MS/MS method following a single oral dose of dextromethorphan cough syrup (15 mg for participants ≥6 to < 12 years old; 30 mg for participants ≥12 to < 25 years old). Poor metabolizer phenotype was defined as urinary DXM/DXO molar ratios > 0.3 or log ratios > −0.5, while extensive metabolizer phenotype was defined as molar ratios ≤ 0.3 or log ratios ≤ −0.5.

Plasma concentrations of primary protease inhibitors, ritonavir and efavirenz were measured at the University of California San Diego Pediatric Pharmacology Laboratory by validated reverse phase high performance liquid chromatography or mass spectrometry methods. The interassay coefficients of variation were all <18%, and the mean recovery from the plasma ranged from 98 to 117%. The lower limits of quantitation were all validated reverse phase high performance liquid chromatography

California San Diego Pediatric Pharmacology Laboratory by validated reverse phase high performance liquid chromatography or mass spectrometry methods. The interassay coefficients of variation were all <18%, and the mean recovery from the plasma ranged from 98 to 117%. The lower limits of quantitation were all 0.047 µg/mL for atazanavir, 0.090 µg/mL for darunavir, 0.091 µg/mL for lopinavir, 0.094 µg/mL for ritonavir, and 0.039 µg/mL for efavirenz.

Plasma concentration at pre-dose (C₀), the maximum concentration (Cmax), the corresponding time (tmax), and the concentration at 24 h post-dose (C₂₄) were identified by direct inspection of concentration-time curves. The area under the concentration vs. time curve from time 0–24 h post dose (AUC₀→₂₄) was estimated using the trapezoidal rule. Apparent oral clearance (CL/F), where F is bioavailability, was calculated as dose divided by AUC₀→₂₄. The apparent volume of distribution (V/F) was determined as CL/F divided by k, where k is the terminal slope of the log plasma concentration-time curve. The half-life (t₁/₂) was calculated as 0.693/k. All concentration data were normalized to dose (100 mg) and to weight (70 kg). Non-compartmental parameters were estimated with Phoenix® WinNonlin® (Certara L.P., St. Louis, MO). Pharmacokinetic parameters were compared between HIV(–) and PI/r cohorts with Wilcoxon rank-sum tests, two-sided with significance set to 0.05. Statistical comparisons were not made with the Wilcoxon rank-sum tests, two-sided with significance set to 0.05. Statistical comparisons were not made with the Wilcoxon rank-sum tests, two-sided with significance set to 0.05. Statistical comparisons were not made with the Wilcoxon rank-sum tests, two-sided with significance set to 0.05.
TABLE 2 | Sertraline and N-desmethylsertraline pharmacokinetic parameters, median (Interquartile Range)a.

| Parameter                          | HIV(–) n = 16 | PI/r n = 12 | p-valueb | EFV n = 3 |
|------------------------------------|---------------|-------------|----------|-----------|
| **SERTRALINE**                     |               |             |          |           |
| AU(t24) (ng/hr/mL)                 | 652 (606, 1407) | 486 (415, 670) | 0.08     | 145 (138, 288) |
| Norm-AU(t24) (ng/hr/mL)c           | 1176 (713, 1880) | 791 (546, 961) | 0.12     | 473 (374, 614) |
| C0 (ng/mL)                         | 20.1 (12.6, 39.7) | 10.0 (7.5, 15.9) | 0.03     | 6.0 (3.0, 7.0) |
| Norm-C0 (ng/mL)c                   | 33.3 (17.6, 57.3) | 17.8 (9.9, 23.0) | 0.03     | 13.1 (9.2, 19.6) |
| Cmax (ng/mL)                       | 46.7 (36.5, 90.1) | 34.3 (23.6, 41.7) | 0.09     | 13.2 (8.8, 22.1) |
| Norm-Cmax (ng/mL)c                 | 78.3 (50.9, 110.7) | 46.9 (42.2, 68.7) | 0.06     | 28.8 (28.4, 58.7) |
| Tmax (hr)                          | 4 (4, 6)       | 4 (4, 6)     | 1.00     | 6 (4, 6)  |
| C24 (ng/mL)                        | 17.5 (14.3, 40.1) | 12.6 (8.6, 18.9) | 0.07     | 4.2 (2.9, 5.9) |
| Norm-C24 (ng/mL)c                  | 32.7 (17.6, 51.9) | 20.1 (11.8, 29.2) | 0.17     | 12.8 (7.6, 13.8) |
| CL/F (L/hr/kg)                     | 1.4 (0.8, 2.3) | 1.6 (1.2, 2.3) | 0.59     | 4.5 (1.6, 11.5) |
| T1/2 [hr]                          | 26.4 (14.1, 35.3) | 18.1 (12.5, 23.1) | 0.28     | 11.1 (10.2, 20.7) |
| AUC Ratio (DSRT/SRT)               | 1.4 (1.2, 1.7) | 1.3 (0.7, 1.6) | 0.13     | 2.2 (2.1, 2.6) |
| Ln(DXM/DXO)d                       | −2.3 (−3.0, −0.6) | −4.3 (−4.8, −3.8) | 0.01     | −2.35 |

| **N-DESMETHYLSERTRALINE**          |               |             |          |           |
| AU(t24) (ng/hr/mL)                 | 1063 (848, 1721) | 670 (230, 1244) | 0.09     | 376 (293, 624) |
| Norm-AU(t24) (ng/hr/mL)c           | 1533 (1053, 2123) | 899 (646, 1186) | 0.01     | 1223 (814, 1301) |
| C0 (ng/mL)                         | 41.7 (29.2, 65.8) | 17.8 (4.8, 43.1) | 0.05     | 15.5 (7.6, 25.5) |
| Norm-C0 (ng/mL)c                   | 50.6 (33.5, 100.9) | 23.8 (11.7, 45.0) | 0.01     | 33.7 (33.2, 50.3) |
| Cmax (ng/mL)                       | 56.3 (45.6, 92.0) | 34.8 (12.5, 57.6) | 0.07     | 17.5 (16.8, 34.1) |
| Norm-Cmax (ng/mL)c                 | 76.6 (54.1, 138.7) | 44.6 (33.1, 58.8) | 0.01     | 56.8 (44.5, 74.7) |
| C24 (ng/mL)                        | 40.8 (30.4, 68.3) | 20.5 (10.3, 45.1) | 0.07     | 15.2 (8.7, 20.8) |
| Norm-C24 (ng/mL)c                  | 59.4 (36.6, 82.0) | 33.6 (17.8, 56.4) | 0.03     | 38.7 (27.1, 49.3) |

a For the EFV cohort, median [minimum, maximum] are reported.
b P-values calculated using the Wilcoxon rank-sum test for HIV(–) and PI/r comparisons.
c Normalized to a 100 mg once daily dose and a weight of 70 kg.
d Urine DXM/DXO ratio was measured in 12 HIV(–), 6 PI/r, & 1 EFV.

Statistically higher than PI/r (76.6 vs. 44.6 ng/mL, p = 0.01). Following normalization, C0 and C24 in HIV(–) were statistically greater than PI/r (50.6 vs. 23.8 ng/mL, p = 0.01; 59.4 vs. 33.6 ng/mL, p = 0.03).

Median (interquartile range) ritonavir AUC0–12 and Cmax for PI/r participants were 4.9 (3.0, 9.4) µg·h/mL and 0.6 (0.4, 1.6) µg/mL. Median (range) efavirenz AUC0–24 and Cmax for the three EFV participants were 24.5 (24.4, 68.2) µg·h/mL and 1.3 (1.3, 3.4) µg/mL. No relationship was observed between ritonavir AUC0–12 and dose normalized sertraline AUC0–24 (r² = 0.005) or ritonavir AUC0–12 and the AUC0–24 ratio of N-desmethylsertraline/sertraline (r² = 0.09). No correlation was present between efavirenz AUC0–24 and dose-normalized sertraline AUC0–24 (r² = 0.01). A correlation was present between efavirenz AUC0–24 and the AUC0–24 ratio of N-desmethylsertraline/sertraline (r² = 0.97). However, with a small sample size (n = 3), these results should be interpreted with caution.

Metabolic phenotyping was performed in 19 of 31 participants (12 HIV(–), 6 PI/r, and 1 EFV). The log ratio of DXM to DXO in the HIV(–) group was significantly higher than the PI/r group (−2.3 vs. −4.3, p = 0.01). Four of twelve participants in the HIV(–) group were CYP2D6 poor metabolizers, compared to zero of six PI/r participants. CYP2D6 phenotype was tested as a covariate on the sertraline population pharmacokinetic model, but did not significantly affect CL/F or V/F.

DISCUSSION

This is the first report of sertraline pharmacokinetics among youth living with HIV. Sertraline and N-desmethylsertraline exposure parameters trended lower or were significantly lower in the PI/r compared to the HIV(–) groups. Sertraline exposure, corrected for weight and dose, was much lower in EFV participants, but the sample size was very small. Few patients with psychiatric conditions were taking efavirenz, likely due to clinician concerns about central nervous system side effects associated with efavirenz use in patients with psychotic disorders. The normalized sertraline AUC0–24 in the HIV(–) group (1.176 ng·hr/mL) was similar to that seen after multiple dosing in prior adult studies, normalized to a 100 mg dose (1038–1532 ng·hr/mL) (20). The normalized sertraline trough concentrations, C0 and C24, in the HIV(–) and PI/r groups of approximately 18–33 ng/mL were also similar to those in a therapeutic drug monitoring program, that reported a median and IQR sertraline trough with 100 mg daily doses of 20 ng/mL and 12–30 ng/mL (21). The observed troughs in the EFV group

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and UGT, a phase II metabolic pathway of sertraline (and CYP3A4, but is also a strong inducer of CYP2C19 and CYP3A4. Ritonavir is a strong inhibitor of CYP2D6.

FIGURE 1 (A) Median sertraline profiles by cohort. Sertraline concentrations were normalized to a dose of 100 mg and a patient weight of 70 kg. (B) Median N-desmethylsertraline profiles by cohort. Concentrations were normalized to a sertraline dose of 100 mg and a patient weight of 70 kg.

of 4.2 and 6 ng/mL were well below these typical trough values. The median weight-adjusted daily dose (mg/kg) in the HIV(−) group, while not statistically different, was higher than both groups of participants living with HIV. This suggests that clinicians are dosing sertraline more conservatively, possibly due to concerns about interacting antiretrovirals, in youth living with HIV. However, weight- and dose-normalized sertraline AUC0−24 suggests that youth taking common PI/r should be receiving equivalent, or potentially higher, weight-adjusted doses as their HIV-uninfected peers.

While we expected normalized sertraline exposures to be greater in HIV(−) vs. EFV participants, due to strong cytochrome P450 induction, we did not expect to also observe lower sertraline in PI/r participants. Sertraline is eliminated by multiple pathways including CYP2B6, CYP2D6, CYP2C9, CYP2C19, and CYP3A4. Ritonavir is a strong inhibitor of CYP2D6 and CYP3A4, but is also a strong inducer of CYP2C19 and UGT, a phase II metabolic pathway of sertraline (7). Involvement of numerous enzymes in sertraline metabolism make it unlikely for one agent to cause a clinically significant drug interaction (7, 20). Antiretroviral exposures in this study were similar to expected values, suggesting that participants were adherent to antiretrovirals. Our results, lower normalized (weight/dose) sertraline and N-desmethylsertraline AUC0−24 and the AUC ratio (N-desmethylsertraline/sertraline) in PI/r vs. HIV(−) participants, suggest that inhibition of various CYP enzymes by ritonavir does not increase sertraline exposure. Furthermore, decreased sertraline exposures in PI/r relative to HIV(−) were not a consequence of induction of enzymatic pathways, since ritonavir AUC0−24 did not correlate with the N-desmethylsertraline/sertraline AUC ratio. If induction was present, the AUC ratio would increase with increasing ritonavir. The most likely explanations for reduced sertraline exposures in the PI/r cohort are (1) decreased drug absorption in participants living with HIV and (2) lower sertraline doses prescribed (which can cause changes in sertraline pharmacokinetics that are not proportional to the difference in dose amount).

Another potential contributing factor to sertraline exposure differences could be underlying genetic polymorphisms in CYP enzymes. The Clinical Pharmacogenetics Implementation Consortium recommends a 50% decrease in sertraline dose in CYP 2C19 poor metabolizers due to increased adverse effects, but no change in dose for ultrarapid metabolizers. This recommendation is optional due to limited available evidence (22). CYP 2C19 genotypes of study participants were not determined.

A population pharmacokinetic study of geriatric patients (median age = 76 years) with Alzheimer’s Disease also determined a 1-compartment model with first-order absorption and elimination best described sertraline, and estimated a clearance of 83.1 L/h (23). Our study estimated clearance at 115 L/h (for a typical 70 kg patient); 38% higher. In our model, of the covariates tested on CL/F and V/F (age, gender, race, cohort, CYP2D6 phenotype, aspartate aminotransferase, alanine aminotransferase, serum creatinine, and AUC ratio (N-desmethylsertraline/sertraline)), only age (independent of size) significantly affected clearance. This finding contrasts with previous pediatric studies and the aforementioned geriatric study, which found no associations between age and sertraline pharmacokinetics. Our finding should be interpreted with caution due to the limited range of ages in this study, 14–23 years. Gender did not affect sertraline pharmacokinetics in our study, similar to prior pediatric trials, but different from adults where men had an elimination half-life approximately 1.5 times greater than women (19).

EFV participants had sertraline exposures ~50% lower than HIV(−) and PI/r groups. In previous pediatric studies (15, 17), AUC0−24 (normalized to 100 mg dose and 70 kg weight) ranged from 883 to 2,535 ng·h/mL, much higher than 473 ng·h/mL in the EFV group. The metabolite-to-parent AUC ratio was approximately 50% higher in EFV than other cohorts, and efavirenz AUC was correlated with metabolite-to-parent AUC ratio ($r^2 = 0.97$). These findings suggest that participants taking efavirenz have a much higher oral intrinsic clearance, possibly through induction of multiple CYP enzymes that contribute to sertraline metabolism, including 2B6, 2C19, and 3A4. For 3A4 in particular, the reduction in EFV exposure could be due to induction at the intestinal level along with hepatic induction,
resulting in both increased systemic clearance and decreased absorption due to first-pass loss upon oral administration. A larger sample size is needed to confirm this observation.

Participants in the HIV(−) group had sertraline weight-adjusted pharmacokinetic characteristics similar to adults. Ensuring dose modifications according to weight are therefore necessary for youth without HIV to achieve similar therapeutic exposures. Sertraline exposure in the PI/r cohort was ∼30–40% lower compared to HIV(−). Concomitant ritonavir did not increase sertraline exposure as it does with many other medications, and youth taking a ritonavir-boosted protease inhibitor should be receiving at least the same sertraline dose (mg/kg), or potentially even modestly higher doses, as uninfected youth and adults. Participants taking efavirenz had much lower sertraline exposures than the two other cohorts and adults. Even though psychiatric medications are often titrated to effect, these participants did not receive higher absolute or weight-adjusted doses than the other groups, and likely had sub-therapeutic exposures. Caution should be exercised when interpreting findings from the EFV cohort due to the small sample size (n = 3), but higher doses or therapeutic drug monitoring of sertraline in this population may be warranted.

**DATA AVAILABILITY STATEMENT**

The data cannot be made publicly available due the ethical restrictions in the study’s informed consent documents and in the International Maternal Pediatric Adolescent AIDS Clinical Trials Network (IMPAACT) Network’s approved human subjects protection plan; public availability may compromise participant confidentiality. However, data are available to all interested researchers upon request to the IMPAACT Statistical and Data Management Center’s data access committee (email address: sdc.data@fstrf.org) with the agreement of the IMPAACT Network.

**ETHICS STATEMENT**

All participants gave written informed consent, permission or assent in accordance with the Declaration of Helsinki and local guidelines. The protocol was approved by all relevant ethics or human subjects protection committees at all participating sites.

**AUTHOR CONTRIBUTIONS**

All co-authors reviewed, revised for content, and approved this article. MP, YH, SK, ES, GS, PB, EC, and BB: substantial contributions to the conception and design; NH, MP, BG, and BJ: acquisition of data; NH, MP, YH, SK, ES, GS, PB, EC, and BB: analysis or interpretation of data.

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**TABLE 4 | Antiretroviral exposures, median (Interquartile range).**

| ANALYTE     | n  | AUC0–tau (µg*hr/mL) | Cmax (µg/mL) | Clast (µg/mL) |
|-------------|----|---------------------|--------------|---------------|
| Atazanavir  | 5  | 27.2 (21.8, 38.7)   | 3.8 (3.1, 5.2) | 1.2 (0.9, 2.8) |
| Darunavir   | 5  | 60.1 (50.2, 66.5)   | 6.8 (6.2, 7.1) | 3.5 (3.2, 3.9) |
| Lopinavir   | 2  | 41.6, 62.0          | 11.9, 4.9    | 2.4, 3.5      |
| Ritonavir   | 12 | 4.9 (3.0, 9.4)      | 0.6 (0.4, 1.6) | 0.2 (0.1, 0.4) |
| Efavirenz   | 3  | 24.5 (24.4, 68.2)   | 1.2 (1.1, 3.4) | 1.3 (1.3, 3.4) |

*The range is reported for lopinavir and the median (range) is reported for efavirenz.
REFERENCES

1. Penzak SR, Reddy YS, Grimes SR. Depression in patients with HIV infection. Am J Health Syst Pharm. (2000) 57:376–86. doi: 10.1093/ajhp/57.4.376

2. Chernoff M, Nachman S, Williams P, Brouwers P, Heston J, Hodge J, et al. Mental health treatment patterns in perinatally HIV-infected youth and controls. Pediatrics (2009) 124:627–36. doi: 10.1542/peds.2008-2441

3. Gonzalez JS, Bacheleter AW, Psaros C, Safren SA. Depression and HIV/AIDS treatment nonadherence: a review and meta-analysis. J Acquir Immune Defic Syndr. (2011) 58:181–7. doi: 10.1097/QAI.0b013e31822d490a

4. Wagner GI, Goggins K, Remien RH, Rosen MI, Simonj J, Bangsberg DR, et al. A closer look at depression and its relationship to HIV antiretroviral adherence. Ann Behav Med. (2011) 42:352–60. doi: 10.1007/s12160-011-9295-8

5. Pfizer Canada I. Zoloft Product Monograph. Kirkland, QC: Pfizer Canada, Inc., (2014).

6. Kobayashi K, Ishizuka T, Shimada N, Yoshimura Y, Kajihara K. Sertraline N-demethylation is catalyzed by multiple isoforms of human cytochrome P-450 in vitro. Drug Metab Dispos. (1999) 27:763–6.

7. Obach RS, Cox LM, Tremaine LM. Sertraline is metabolized by multiple cytochrome P450 enzymes, monoamine oxidases, and glucuronil transferases in human: an in vitro study. Drug Metab Dispos. (2005) 33:262–70. doi: 10.1124/dmd.104.002428

8. KALETRA. Product Monograph. Kaletra (lopinavir/ritonavir). St Laurent, QC: AbbVie Corporation (2017).

9. Lee CG, Gottesman MM, Cardarelli CO, Ramachandra M, Jeang KT, Ambudkar SV, et al. HIV-1 protease inhibitors are substrates for the MDR1 multidrug transporter. Biochemistry (1998) 37:5394–01. doi: 10.1021/bi972709a.

10. Hogeland GW, Swiddells S, McNabb IC, Kashuba AD, Yee GC, Lindley CM. Lopinavir/ritonavir reduces bupropion plasma concentrations in healthy subjects. Clin Pharmacol Ther. (2007) 81:69–75. doi: 10.1038/sj.clpt.6100027

11. Hsu A, Granneman GR, Bertz RJ. Ritonavir. Clinical pharmacokinetics and interactions with other anti-HIV agents. Clin Pharmacokinet. (1998) 35:275–91. doi: 10.2165/00003088-199835040-00002

12. Yeh RF, Gaver VE, Patterson KB, Rezak NL, Baxter-Meheux F, Blake MJ, et al. Lopinavir/ritonavir induces the hepatic activity of cytochrome P450 enzymes CYP2C9, CYP2C19, and CYP1A2 but inhibits the hepatic and intestinal activity of CYP3A as measured by a phenotyping drug cocktail in healthy volunteers. J Acquir Immune Defic Syndr. (2006) 42:52–60. doi: 10.1097/01.qai.0000179774.20174.64

13. Sustiva. Sustiva (package insert). Princeton, NJ: Bristol-Myers Squibb Company (2017).

14. Michaud V, Kreutz Y, Skaar T, Ogburn E, Thong N, Flockhart D, et al. Efavirenz-mediated induction of omeprazole metabolism is CYP2C19 genotype-dependent. Pharmacogenomics J. (2014) 14:151–9. doi: 10.1038/pj.2013.17

15. Alderman J, Wolkow R, Chung M, Johnston HF. Sertraline treatment of children and adolescents with obsessive-compulsive disorder or depression: pharmacokinetics, tolerability, and efficacy. J Am Acad Child Adolesc Psychiatry (1998) 37:386–94. doi: 10.1097/00004583-199804000-00016

16. Alderman J, Wolkow R, Fogel IM. Drug concentration monitoring with tolerability and efficacy assessments during open-label, long-term sertraline treatment of children and adolescents. J Child Adolesc Psychopharmacol. (2006) 16:117–29. doi: 10.1089/cap.2006.16.117

17. Axelsson DA, Perel JM, Birnher B, Rudolph GR, Nuss S, Bridge J, et al. Sertraline pharmacokinetics and dynamics in adolescents. J Am Acad Child Adolesc Psychiatry (2002) 41:1037–44. doi: 10.1097/01.qai.00004583-200209000-00003

18. Taurines R, Burger R, Wewetzer C, Pfuhlmann B, Mehler-Wex C, Gerlach M, et al. The relation between dosage, serum concentrations, and clinical outcome in children and adolescents treated with sertraline: a naturalistic study. Ther Drug Monit. (2013) 35:84–91. doi: 10.1097/FTD.0b013e31827a1aad

19. Ronfeld RA, Tremaine LM, Wilner KD. Pharmacokinetics of sertraline and its N-demethyl metabolite in elderly and young male and female volunteers. Clin Pharmacokinet. (1997) 32(Suppl. 1):22–30. doi: 10.2165/00003088-199703021-00004

20. DeVane CL, Liston HL, Markowitz JS. Clinical pharmacokinetics of sertraline. Clin Pharmacokinet. (2002) 41:1247–66. doi: 10.2165/00003088-200241150-00002

21. Jons L, Margareta R, Finn B. Therapeutic drug monitoring of sertraline: variability factors as displayed in a clinical setting. Therap Drug Monitor (2000) 22:446–54. doi: 10.1097/00007691-200008000-00014

22. Hicks JK, Bishop JR, Sangkuhl D, Muller DJ, Ji Y, Leckband SG, et al. Clinical pharmacogenetics implementation consortium (CPIC) guideline for CYP 2D6 and CYP 2C19 genotypes and dosing of selective serotonin reuptake inhibitors. Clin Pharmacol and Ther. (2015) 98:127–34. doi: 10.1002/cpt.147

23. Li CH, Pollock BG, Lyketsos CG, Vaidya V, Drye LT, Kirchner M, et al. Population pharmacokinetic modeling of sertraline treatment in patients with Alzheimer disease: the DIADS-2 study. J Clin Pharmacol. (2013) 53:234–9. doi: 10.1177/0091270012445793

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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