Coadministration of FTY720 and rt-PA in an experimental model of large hemispheric stroke—no influence on functional outcome and blood–brain barrier disruption

Aijia Cai1, Frieder Schlunk1, Ferdinand Bohmann1, Sepide Kashefiolasl3, Robert Brunkhorst1,2, Christian Foerch1 and Waltraud Pfeilschifter1,2*

Abstract

Background: Systemic thrombolysis with recombinant tissue plasminogen activator (rt-PA) is the standard of acute stroke care. Its potential to increase the risk of secondary intracerebral hemorrhage, especially if administered late, has been ascribed to its proteolytic activity that has detrimental effects on blood–brain barrier (BBB) integrity after stroke. FTY720 has been shown to protect endothelial barriers in several disease models such as endotoxin-induced pulmonary edema and therefore is a promising candidate to counteract the deleterious effects of rt-PA. Besides that, every putative neuroprotectant that will be eventually forwarded into clinical trials should be tested in conjunction with rt-PA.

Methods: We subjected C57Bl/6 mice to 3 h filament-induced tMCAO and postoperatively randomized them into four groups (n = 18/group) who received the following treatments directly prior to reperfusion: 1) vehicle-treatment, 2) FTY720 1 mg/kg i.p., 3) rt-PA 10 mg/kg i.v. or 4) FTY720 and rt-PA as a combination therapy. We measured functional neurological outcome, BBB disruption by quantification of EB extravasation and MMP-9 activity by gelatin zymography.

Results: We observed a noticeable increase in mortality in the rt-PA/FTY720 cotreatment group (61%) as compared to the vehicle (33%), the FTY720 (39%) and the rt-PA group (44%). Overall, functional neurological outcome did not differ significantly between groups and FTY720 had no effect on rt-PA- and stroke-induced BBB disruption and MMP-9 activation.

Conclusions: Our data show that FTY720 does not improve functional outcome and BBB integrity in large hemispheric infarctions, neither alone nor in conjunction with rt-PA. These findings stand in contrast to a recently published study that showed beneficial effects of FTY720 in combination with thrombolysis in a thrombotic model of MCAO leading to circumscribed cortical infarctions. They might therefore represent a caveat that the coadministration of these two drugs might lead to excess mortality in the setting of a severe stroke.

Keywords: Stroke, MCAO, Thrombolysis, FTY720, Fingolimod, Sphingolipid, S1P1 receptor, Brain, Hemorrhage, Blood–brain barrier.
Background

Thrombolysis with recombinant human tissue plasminogen activator (rt-PA) is the only approved and evidence-based medical therapy for ischemic stroke. Within the narrow therapeutic time window of 4.5 h, it confers a clear net benefit to all stroke patients who receive thrombolysis which comes at the cost of an increased risk of secondary intracerebral hemorrhage for the individual patient, mostly caused by hemorrhagic transformation (HT) of the infarcted brain tissue [1].

Magnetic resonance imaging studies in acute stroke patients have shown that blood–brain barrier (BBB) disruption is significantly more prevalent in stroke patients who received thrombolysis than in untreated stroke patients [2]. Based on the description of HARM (hyperintense acute reperfusion marker) which was defined as the presence of gadolinium extravasation into the cerebrospinal fluid (CSF) space adjacent to the infarction on the fluid-attenuated inversion recovery (FLAIR) sequence of a follow up scan after gadolinium injection for a scan that took place a few hours earlier, it became evident that preceding BBB disruption was present in 73% of patients who developed hemorrhagic transformation within the next hours [3].

These observations were supported by animal studies showing that cerebral ischemia leads to an increase of matrix-metalloproteinase (MMP) activity, especially of MMP-9 which follows the same time course as BBB disruption after experimental stroke and that both are aggravated by treatment with rt-PA [4] and correlate with HT [5]. Therefore, combination therapies with drugs that protect endothelial barrier function seem to be a reasonable approach to limit the risks of rt-PA treatment.

One of these substances is the sphingosine 1-phosphate analog FTY720, an immunomodulator that has been marketed in 2010 for the treatment of relapsing–remitting multiple sclerosis (MS). The presumed mechanism of this drug is the induction of peripheral lymphocytopenia that is caused by agonist-induced internalization of the S1P1 receptor of B and T lymphocytes which limits lymphocyte egress from primary lymphoid organs [6]. But beside its effects on immune cell trafficking, FTY720 also has effects on neurons, glia, and progenitor cells in the brain and an effect on the BBB can be presumed [7]. Interestingly, FTY720 treatment led to a downregulation of inflammatory genes including MMP-9 and to an increased expression of its counterpart tissue inhibitor of metalloproteinase (TIMP) in experimental autoimmune encephalitis (EAE) [8]. Given as a rescue therapy, FTY720 reduced BBB leakage after disease onset [8]. Studies in other vascular beds have shown that FTY720 can protect endothelial cells from apoptotic cell death [9]. However, there are conflicting data on the net effect of S1P receptor agonism on endothelial barrier function, with FTY720 being protective in a model of pulmonary edema caused by systemic LPS injection [10] while it has deleterious effects in a model of bleomycin-induced pulmonary fibrosis [11].

A neuroprotective effect of FTY720 in the acute phase of stroke has been shown in several experimental studies [12-14]. From a translational point of view, every neuroprotectant that is aimed at ameliorating acute brain damage in the first hours within stroke onset should be tested in conjunction with rt-PA. Therefore, the aim of this study was to assess the effect of FTY720 in conjunction with rt-PA treatment in an experimental model of stroke and its effect of stroke- and rt-PA-induced BBB disruption and matrix metalloproteinase expression.

Methods

Experimental model of middle cerebral artery occlusion

C57Bl/6 mice (Charles River Laboratories, Sulzfeld, Germany) were used at 10–12 weeks of age. All experiments were approved by the local governmental authorities (Regierungspräsidium Darmstadt, Germany, approval number F143/51) and conducted in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals. Mice were subjected to transient middle cerebral artery occlusion (MCAO) as described previously [15]. Briefly, mice were anesthetized with 1.5–2.5% isoflurane (Forene; Abbott, Wiesbaden, Germany) and 0.1 mg/kg buprenorphine (Temgesic®; Essex Pharma, Munich, Germany) under spontaneous respiration. Focal cerebral ischemia was induced by inserting a custom made filament with a tip diameter of 0.23 mm (Doccol, Sharon, USA) into the middle cerebral artery (MCA). Regional cerebral blood flow was monitored by laser Doppler flowmetry (PF5010, Perimed; Stockholm, Sweden) to confirm vessel occlusion. After an occlusion time of 3 h, the filament was withdrawn to initiate reperfusion. After the operation, mice were allowed to recover from anesthesia with free access to food and water. At the end of a 24 h observation period, mice were lethally anaesthetized and perfused transcardially with PBS. Brains were removed quickly and divided into ischemic and non-ischemic hemispheres before they were frozen and stored at –80°C to await further procedures. Brains from mice that had died within the 24 h observation period were harvested without prior transcardial perfusion.

Sample size calculation, experimental groups and substance application

We based our sample size calculation on the quantifiable parameters Evans Blue extravasation and matrix metalloproteinase-9 activity. Assuming an increase of 25% in these two parameters between the ischemic hemispheres of the vehicle-treated and rt-PA-treated mice that has been shown in previous studies and a standard deviation of 33% of the respective mean values (Cohen’s D 0.85), a group size of 18 animals was necessary to show
this effect with a power of 0.8 and a probability of a type I error of < 0.5 [16]. After the MCAO operation and prior to reperfusion, we randomized 18 mice per group into the four treatment groups: 1) vehicle-treatment, 2) 1 FTY720 1 mg/kg 3) rt-PA 10 mg/kg, or 4) FTY720 and rt-PA as a combination therapy. The mice received either FTY720 (1 mg/kg, dissolved in 0.9% NaCl; Fingolimod; Cayman Chemicals, Ann Arbor, USA) or saline i.p. in combination with an i.v. bolus of either rt-PA (10 mg/kg; Actilyse®; Boehringer Ingelheim, Ingelheim, Germany) or aqua ad injectabilia. The operator was blinded to the pharmaceutical treatment during the whole study. Additionally, we assessed whether preconditioning of the rt-PA-solution by exposure to a fibrin-containing clot would enhance its deleterious effects on the blood–brain barrier. To this end, blood was drawn from a donor mouse to generate a spontaneously-formed blood clot. Rt-PA was incubated with this blood clot for 30 min under gentle shaking, generating “activated” rt-PA (Art-PA). We performed 3 h of MCAO in two sets of mice that received an i.v. bolus of either rt-PA (10 mg/kg) or Art-PA (10 mg/kg).

**FACS analysis of lymphocyte counts**

500 µl of blood were drawn into EDTA-coated tubes from 3 animals per group (1 mg/kg FTY720 vs. saline) 2 h after the injection of FTY720 or vehicle, cells were spun down and erythrocytes were lysed with red cell lysis buffer (8.3 g NH₄Cl in 0.01 mol/L Tris–HCl, pH 7.4). After washing with RPMI, samples were incubated with FACS buffer (1% FCS in PBS/0.01% NaN₃) and the respective antibodies (Rat anti-mouse CD4 [APC-coupled, Southern Biotech, 1540–11], rat anti-mouse CD8 [PerCP-CTM5.5-coupled, BD Pharmingen, 551162]) for 30 minutes before overnight fixation in 4% PFA in PBS at 4°C. Samples were analyzed on a FACSCalibur with the CellQuestPro Software (BD Biosciences).

**Evaluation of neurological function**

Neurological function was evaluated at 3 h directly prior to reperfusion and at 24 h post-MCAO on a 14 point scale modified from Chen et al. [17] testing hemiparesis, gait, coordination and sensory functions (Additional file 1: Table S1).

**Determination of blood–brain barrier leakage**

To assess blood–brain barrier leakage, the extravasation of the autofluorescent dye Evans Blue (EB) that binds to plasma albumin was quantified from brain hemispheres as described before [5]. 23 h after MCAO, 200 µl of 2% Evans Blue in 0.9% NaCl were injected into the retrobulbar venous plexus and allowed to circulate for 1 h prior to transcardial perfusion and brain removal. Brain samples were homogenized in lysis buffer as described above and additionally with ultrasound. Protein precipitation was obtained by adding 50% trichloroacetic acid. The supernatant was diluted 4-fold with ethanol. The amount of Evans blue dye was measured by a microplate fluorescence reader (excitation 600 nm, emission 650 nm; SpectraMax M5; Molecular Devices, Sunnyvale, USA).

**MMP-9 activity**

Gelatin zymography was used to measure the levels of MMP-9 activity in the brain samples as described previously [5,18]. We analyzed the brains of all mice, including those that did not survive the observation period. Brain hemispheres were homogenized in ice-cold lysis buffer containing protease inhibitors. After centrifugation, the supernatant was collected and total protein concentration of each sample was determined by the Bradford assay (Nanoquant, Roth, Karlsruhe, Germany). Equal volumes of total protein extracts in sample buffer (4% SDS, 0.005% bromphenol blue, and 20% glycerol) were loaded onto 10% polyacrylamide gels containing 0.1% gelatin as a substrate. After electrophoresis, the gels were incubated in 2.5% Triton X-100 at room temperature for 30 min with gentle agitation and stained with 0.5% Coomassie Blue G250. For densitometry, gels were scanned, inverted and integrated density of the bands was quantified with NIH image J 1.44p.

**MMP-9 protein expression**

We evaluated MMP-9 expression on the protein level by Western Blotting. Samples of equal total protein content were loaded onto polyacrylamide gels. After migration, proteins were electrotransferred onto a nitrocellulose blotting membrane. To confirm successful transfer, the membrane was stained with Ponceau S. It was then blocked in 5% dry milk and 0.05% Tween-20 for 1 h at room temperature under gentle agitation. After extensive washing, the membrane was incubated with the primary antibody (Anti-MMP-9 rabbit polyclonal; Millipore, Billerica, USA) overnight. After exposition to a second antibody (goat anti-rabbit antibody; Bio-Rad, Munich, Germany), the blots were developed with a chemiluminescence reagent on hyperfilm.

**Statistical analysis**

Graph Pad Prism 4 (Graph Pad Software Inc., La Jolla, CA, USA) was used for statistical analysis. Results are expressed as means +/- standard deviation. Statistical significance of the differences between groups was evaluated with One-way ANOVA with Bonferroni’s correction for parametric values or the Kruskal-Wallis test with Dunn’s correction for nonparametric values.
Results

Intraperitoneal injection of FTY720 leads to a rapid decrease of blood lymphocyte counts in mice

To verify efficient uptake and potency of FTY720 via the route of administration that we chose for our experiment, we sampled blood 2 h after i.p. injection of 1 mg/kg FTY720. Fluorescence-activated cell sorting (FACS) analysis of CD4+ and CD8+ T cell populations showed a significant decrease in circulating T lymphocytes to 22.5% for CD8+ T cells and 4.3% for CD4+ T cells as compared to vehicle treated mice (Figure 1).

FTY720 in conjunction with rt-PA does not improve survival or functional neurological outcome in large hemispheric infarctions

Mice were randomized into four groups (n = 18/group) to receive either 10 mg/kg rt-PA or vehicle i.v. at the end of the 3 h MCAO period in combination with either 1 mg/kg FTY720 or vehicle i.p. The vehicle only-treated group showed a mortality of 33%. Interestingly, the group who received FTY720 in conjunction with rt-PA showed a considerably higher mortality of 61% (Figure 2A). There were no significant differences between groups in the functional neurological examination with the 14-point neuroscore mNSS (Figure 2B).

FTY720 does not enhance blood–brain barrier integrity in large hemispheric infarcts alone or in combination with rt-PA

Brain weight of the ischemic hemisphere was increased by 30–40% in comparison to the non-ischemic hemisphere. Neither rt-PA nor FTY720 treatment alone had an influence on this very crude measure of brain swelling and also rt-PA in combination with FTY720 did not lead to significant changes in wet brain weight. We only included mice whose ischemic hemisphere showed at least a weight increase by 10% to exclude all mice who showed postmortem global brain swelling (Figure 3A) after being found dead during the observation period. All mice who survived the 24 h observation period received 200 μl 2% Evans Blue (EB) i.v. one h prior to sacrifice to assess BBB permeability for macromolecules such as albumin. Fluorometric EB
quantification from brain homogenates of transcardially perfused mice did also not show an alteration of stroke-induced EB extravasation by rt-PA, FTY720 or the combination of both substances (Figure 3B).

Neither rt-PA nor FTY720 induce significant changes in MMP-9 activity in brain homogenates 24 h after MCAO. Matrix metalloproteinase-9 (MMP-9) is a serine protease that becomes activated in case of ischemic brain injury and has been shown to be a key mediator of blood–brain barrier breakdown in experimental stroke [5]. We performed gelatin zymography to assess MMP-9 activity in the infarcted and non-infarcted hemisphere. While there was a clear, approximately five-fold increase in the infarcted hemisphere, much to our surprise, the administration of FTY720 or rt-PA or the combination of both substances at the end of the 3 h MCAO period did not lead to significant changes in MMP-9 activity (Figure 4A). We performed exemplary Western Blots of MMP-9 and found that MMP-9 activity as assessed by gelatin zymography was highly correlated with MMP-9 protein expression (Figure 4B).

Activated rt-PA (Art-PA) does not lead to a further increase in blood–brain barrier breakdown after cerebral ischemia.

We aimed to clarify the lack of a clear detrimental effect of rt-PA on BBB integrity and MMP-9 activity in our experimental setting. Since it is conceivable that rt-PA only develops its proteolytic effect when it is activated by a fibrin rich clot of adequate surface, we preincubated rt-PA with a spontaneously-formed autologous blood clot for 30 min at room temperature prior to injection after MCAO to generate activated rt-PA (Art-PA). This preconditioning of rt-PA, however, also did not lead to the 2–4-fold increase of the cerebral ischemia-induced MMP-9 activity in the ischemic hemisphere that has been reported elsewhere for the filament occlusion model [4] (Figure 5).

**Discussion**

We found no relevant protective effect of FTY720 when applied in conjunction with rt-PA in an experimental model of large hemispheric strokes, neither on functional neurological outcome nor on markers of BBB disruption. In contrast, our data rather point towards safety concerns against the coadministration of these two drugs in patients with severe strokes.

Analyzing mortality, we found a noticeable difference between the vehicle group (33%) and the groups with administration of FTY720 alone (39%) or rt-PA alone (44%) on the one hand and the cotreatment group who received rt-PA in conjunction with FTY720 (61%) on the other hand. When evaluating functional outcome after 24 h on the 14 point neuroscore including dead animals which were assigned the maximal score of 14, the differences between groups were not statistically significant.

These data are in some aspects remindful of the results of the multicentric clinical phase II/III trial of erythropoietin (EPO) published in 2009, which assessed its safety and efficacy in acute stroke [19]. This clinical trial was preceded by promising experimental studies that had shown a robust neuroprotective effects and a clinical phase I trial of EPO as a monotherapy in the acute phase of stroke [20] that had demonstrated adequate safety. In contrast, the phase II/III trial, which for the first time allowed systemic thrombolysis in conjunction with EPO treatment, failed to show efficacy. Characteristics of this trial population were...
rather severe strokes (mean NIHSS: 13) and the coadministration of systemic thrombolysis in 60% of patients [19]. There was an excess of mortality in the group of erythropoietin-treated patients compared to the placebo group and the patients who received erythropoietin as a cotreatment together with thrombolysis fared distinctly worse [19]. Therefore, we interpret our data as a caveat that FTY720, which has shown a beneficial effect on outcome and infarct size in several different stroke models, might not be effective if used as a treatment for severe strokes, especially in conjunction with systemic thrombolysis.

Regrettably, our data do not allow a functional explanation of the excess mortality of rt-PA treatment in conjunction with FTY720. Focusing on BBB analyses, we did not perform a quantification of hemorrhagic transformation, e.g. with brain imaging or a hemoglobin assay. Therefore, we have no information on whether the combination therapy led to an increase in hemorrhagic transformation. Concerning alternative extracranial causes of mortality, FTY720 has been shown to induce bradycardia [21], bronchoconstriction and mild pulmonary edema [22] in mice and humans. Besides that, the paucity of circulating lymphocytes could in principle entrain an increased susceptibility towards infections, even though we showed previously, that FTY720 does not increase the rate of stroke associated pneumonia in mice [23] and similar findings have been shown for equally specific immunomodulatory interventions in experimental stroke [24]. However, these symptoms are no common side effects of rt-PA treatment and should have occurred to the same extent in the group receiving FTY720 alone if they were to explain this excess mortality.

Much to our surprise, we did not detect a deleterious effect of rt-PA alone on stroke induced BBB dysfunction. This is at odds with many experimental studies describing an aggravating effect of rt-PA on BBB disruption after stroke [4,5]. One possible explanation could have been that rt-PA was not biologically active in our experimental
system (given as an i.v. bolus injection of 10 mg/kg). We cannot directly prove an effect of rt-PA, but previous studies from our group using the same injection technique showed clear effects of rt-PA on HT after stroke [25]. Interestingly, the increase of BBB disruption after stroke has been shown to be greater in embolic than in mechanical models of stroke as the filament-occlusion used in the present study [4]. There are even reports that the administration of rt-PA in the filament-model per se does not increase BBB damage unless it is “activated” by preincubation with a clot [26,27]. However, in our hands, even after preincubation of the rt-PA solution with an autologous blood clot, we did not find a relevant increase in the ischemia-induced MMP-9 activity by activated rt-PA. Therefore, use of preactivated rt-PA did not serve to improve our experimental model.

Against the backdrop of our own observations and the published studies on FTY720 in experimental stroke studies, it is somewhat surprising that we did not find a beneficial effect of FTY720 on functional outcome in this experiment. We ascribe this discrepancy to the size of the infarction produced by the respective experimental model which might have led to a ceiling effect in our case, where a modest protective effect is no longer discernible. We chose large hemispheric infarctions to induce severe BBB damage in order to increase the aggravation of BBB injury by rt-PA.

From a translational point of view, our data point towards the issue that the protective effect of FTY720 in acute stroke may be lost in large hemispheric infarctions. They represent a caveat that the combination therapy of FTY720 and rt-PA might lead to an excess mortality.

Additional file

Additional file 1: Table S1. Neurological Deficit Score.

Competing interests
WP receives project-specific funding from Novartis Pharma Nürnberg, Germany for another project not related to the data presented in this manuscript.

Authors’ contributions
AC participated in designing the study, conducted the experiments and contributed to the manuscript, FS conducted the blinded evaluation of endpoints, FB participated in designing the study and contributed important intellectual content, SK conducted experiments and contributed important intellectual content, CF participated in designing the study and writing the manuscript, WP designed the study, performed statistical evaluation of the results and wrote the manuscript. All authors read and approved the final manuscript.

Author details
1Department of Neurology, University Hospital, Goethe University Frankfurt, Frankfurt, Germany. 2Department of General Pharmacology and Toxicology, University Hospital, Goethe University Frankfurt, Frankfurt, Germany.
3Department of Neurosurgery, University Hospital, Goethe University Frankfurt, Frankfurt, Germany.

Received: 20 September 2013 Accepted: 14 October 2013
Published: 28 October 2013
References

1. Lees KR, Bluhmki E, von Kummer R, Brott TG, Toni D, Grotta JC, Albens GV, Kaste M, Marler JR, Hamilton SA, Tilley DC, Bassi MS, Donnan GA, Hacket W, ECASS, ATLANTIS, NINDS and EPITHET rt-PA Study Group, Allen K, Mau J, Meier D, del Zoppo G, De Silva DA, Butcher KS, Parsons MW, Barber PA, Levy C, Bladin C, Byrne G: Time to treatment with intravenous alteplase and outcome in stroke: an updated pooled analysis of ECASS, ATLANTIS, NINDS, and EPITHET trials. Lancet 2010, 375(9727):1695–1703.

2. Latour LL, Kang DW, Ezeddaime MA, Chalela JA, Warach S: Early blood–brain barrier disruption in human focal brain ischemia. Ann Neurol 2004, 56:468–477.

3. Warach S, Latour LL: Evidence of reperfusion injury, exacerbated by thrombolytic therapy, in human focal brain ischemia using a novel imaging marker of early blood–brain barrier disruption. Stroke 2004, 35:2659–2661.

4. Aoki T, Sumii T, Mori T, Wang X, Lo EH: Blood–brain barrier disruption and matrix metalloproteinase-9 expression during reperfusion injury: mechanical versus ertibolic focal ischemia in spontaneously hypertensive rats. Stroke 2002, 33:831–836.

5. Sumii T, Lo EH: Involvement of matrix metalloproteinase in thrombolysis-associated hemorrhagic transformation after ertibolic focal ischemia in rats. Stroke 2002, 33:831–836.

6. Brinkmann V: FTY720 (fingolimod) in Multiple Sclerosis: therapeutic effects in the immune and the central nervous system. Br J Pharmacol 2009, 158:1173–1182.

7. Hla T, Brinkmann V: Sphingosine 1-phosphate (S1P): physiology and the effects of S1P receptor modulation. Neurology 2011, 76(Suppl 3):S53–S58.

8. Foster CA, Mechtcheriakova D, Storch MK, Balatoni B, Howard LM, Borrmian F, Wlachos A, Sobanov J, Kinnunen A, Baumruker T: FTY720 rescue therapy in the dark agouti rat model of experimental autoimmune encephalomyelitis: expression of central nervous system genes and reversal of blood–brain-barrier damage. Brain Pathol 2009, 19(2):254–266.

9. Sanchez T, Estrada-Hernandez T, Palk JH, Wu MT, Venkataraman K, Brinkmann V, Caffey K, Hla T: Phosphorylation and action of the immunomodulator FTY720 inhibits vascular endothelial cell growth factor-induced vascular permeability. J Biol Chem 2007, 282(47):47281–47290.

10. Peng X, Hassan PM, Sammann S, McVerry BJ, Bume MU, Rabb H, Pearse D, Tuder RM, Garcia JG: Protective effects of sphingosine 1-phosphate in murine endotoxin-induced inflammatory lung injury. Am J Respir Crit Care Med 2004, 169(1):1245–1251.

11. Shea BS, Brooks SF, Fontaine BA, Chun J, Luster AD, Tager AM: Prolonged S1P1 agonist exposure exacerbates vascular leak, fibrosis, and mortality after lung injury. Am J Respi Crit Care Med 2010, 83(4):662–673.

12. Czech B, Pfleisch fter W, Mazaheri-Ommari N, Strobel MA, Kahles T, Neumann-Haefelin T, Rami A, Huwiler A, Pfleisch fter J: The immunomodulatory sphingosine 1-phosphate analog FTY720 reduces lesion size and improves neurological outcome in a mouse model of cerebral ischemia. Biochem Biophys Res Commun 2009, 389(2):251–256.

13. Wei Y, Yermisci M, Kim HH, Yung LM, Shin HK, Hoang SK, Guo S, Qin T, Alsharif N, Brinkmann V, Liao JK, Lo EH, Waeb er C: Fingolimod provides long-term protection in rodent models of cerebral ischemia. Ann Neurol 2011, 69(1):119–129.

14. Kraft P, Gob E, Schumann MK, Gobel K, Deppermann C, Thielmann I, Herrmann AM, Lorenz K, Brede M, Stoll G, Meuth SG, Nienswandt B, Pfleisch fter W, Kleinschmidt K: FTY720 ameliorates acute ischemic stroke in mice by reducing thrombo-inflammation but not by direct neuroprotection. Stroke 2013, epub ahead of print, DOI:10.1161/STROKEAHA.113.002880.

15. Bohmnn F, Menas a K, Pfleisch fter J, Lindhoff-Last E, Steinitz H, Foerch C, Pfleisch fter W: No influence of dabigatran anticoagulation on hemorrhagic transformation in an experimental model of ischemic stroke. PLoS One 2012, 7(7):e40804.

16. Liu R, Liu Q, He S, Simpkins JW, Yang SH: Combination therapy of 17beta-estradiol and recombinant tissue plasminogen activator for experimental ischemic stroke. J Pharmacol Exp Ther 2010, 332(3):1000–1012.

17. Chen J, Sanberg PR, Li Y, Wang L, Lu M, Willing AE, Sanchez-Ramos J, Chopp M: Intravenous administration of human umbilical cord blood reduces behavioral deficits after stroke in rats. Stroke 2001, 32(11):2682–2688.

18. Eberhardt W, Begg T, Beck KF, Walpen S, Gauer S, BoBes H, Pfleisch fter J: Nitric oxide modulates expression of matrix metalloproteinase-9 in rat mesangial cells. Kidney Int 2000, 57(1):59–69.

19. Ehrenreich H, Weissborn K, Prange H, Schneider D, Weimar C, Wartenberg K, Schellinger PD, Bohn M, Becker H, Wegryn M, Jahnig P, Herrmann M, Knauth M, Bahr M, Heide W, Wagner A, Schwab S, Reichmann H, Schwendemann G, Dengler R, Kastrup A, Bartels C, EPO Stroke Trial Group: Recombinant human erythropoetin in the treatment of acute ischemic stroke. Stroke 2009, 40(12):e647–e656.

20. Byrs N, Siren AL: Erythropoietin: a multimodal neuroprotective agent. Exp Transl Stroke Med 2009, 21:14.

21. Kapoos L, radue EW, O’Connor P, Polman C, Hohlfeld R, Calabresi P, Selmay M, Agoropoulou C, Leyk M, Zhang-Auberson L, Burtn P, FREDOMS Study Group: A placebo-controlled trial of oral fingolimod in relapsing multiple sclerosis. N Engl J Med 2010, 362(9):387–401.

22. Triflief A, Fozard JR: Sphingosine-1-phosphate-induced airway hyper-reactivity in rodents is mediated by the sphingosine-1-phosphate type 3 receptor. J Pharmacol Exp Ther 2012, 342(2):399–406.

23. Pfleisch fter W, Czech-Zechmeister B, Sujak M, Foerch C, Wichelhaus TA, Pfleisch fter J: Treatment with the immunomodulator FTY720 does not promote spontaneous bacterial infections after experimental stroke in mice. Exp Transl Stroke Med 2011, 3:33.

24. Liesl A, Zhou W, Mracsko C, Karcher S, Bauer H, Schwarting S, Sun L, Bruder D, Stegemann S, Cervenka A, Sommer C, Dalpke AH, Veltkamp R: Inhibition of lymphocyte trafficking shields the brain against deleterious neuroinflammation after stroke. Brain 2011, 134(Pt 3):704–720.

25. Pfleisch fter W, Spitzer D, Czech-Zechmeister B, Steinmetz H, Foerch C: Increased risk of hemorrhagic transformation in ischemic stroke occurring during warfarin anticoagulation: an experimental study in mice. Stroke 2011, 42(9):1116–1121.

26. Kahles T, Foerch C, Sieker M, Schroeter M, Steinmetz H, Rami A, Neumann-Haefelin T: Tissue plasminogen activator mediated blood–brain barrier damage in transient focal cerebral ischemia in rats: relevance of interactions between thrombotic material and thrombolytic agent. Vascul Pharmacol 2005, 43(6):254–259.

27. Gau tier S, Petruatt O, Gele P, Lapra i M, Bastide M, Bauters A, Deplanque D, Jude B, Caron J, Boret R: Involvement of thrombolysis in recombinant tissue plasminogen activator-induced cerebral hemorrhages and effect on infarct volume and postischemic endothelial function. Stroke 2003, 34(12):2975–2979.

28. Campos F, Qin T, Castillo J, Seo JH, Arai K, Lo EH, Waeb er C: Fingolimod reduces hemorrhagic transformation associated with delayed tissue plasminogen activator treatment in a mouse thromboembolic model. Stroke 2013, 44(2):505–511.