Histological evaluation of capsules formed by texturized silicone implants with and without polyester mesh coverage (Parietex®). A study on female rats

Ralf Berger1,*, Jurandir Marcondes Ribas Filho2, Osvaldo Malafaia3, Paulo Afonso Nunes Nassif2, Eduardo Nascimento Silva4, Alfredo Benjamin Duarte da Silva5, Milka Takejima6, Marcelo Augusto de Souza7, Pedro Henrique de Paula7, Mário Rodrigues Montemor Netto8, Lucia de Noronha9

1. Fellow Master degree. Postgraduate Program in Principles of Surgery - Mackenzie Evangelical School of Medicine – Curitiba (PR), Brazil.
2. PhD, Associate Professor. Postgraduate Program in Principles of Surgery - Mackenzie Evangelical School of Medicine – Curitiba (PR), Brazil.
3. PhD, Full Professor. Postgraduate Program in Principles of Surgery - Mackenzie Evangelical School of Medicine – Curitiba (PR), Brazil.
4. PhD. General Surgery – Universidade Estadual de Ponta Grossa – Ponta Grossa (PR), Brazil.
5. MD. Plastic Surgery Department - Hospital Erasto Gaertner – Curitiba (PR), Brazil.
6. MD. Postgraduate Program in Plastic Surgery - Hospital Universitário Evangélico Mackenzie, and Instituto de Pesquisa Médica – Curitiba (PR), Brazil.
7. Graduate Student. Universidade Estadual de Ponta Grossa - Ponta Grossa (PR), Brazil.
8. MSc. Clinical Surgery – Universidade Federal do Paraná – Curitiba (PR). Assistant Professor. Anatomical Pathology – Universidade Estadual de Ponta Grossa - Ponta Grossa (PR). Head. Department of Anatomical Pathology - Santa Casa de Misericórdia - Ponta Grossa (PR), Brazil.
9. PhD. School of Medicine – Pontifícia Universidade Católica do Paraná – Curitiba (PR), Brazil.

ABSTRACT

Purpose: To evaluate capsules formed by microtextured silicone implants with and without Parietex® mesh coverage histologically. Methods: Sixty Wistar rats were divided in two groups (meshed and unmeshed). Each group was, then, divided into two subgroups for evaluation at 30 and 90 days. Capsules were analyzed based on hematoxylin and eosin (HE) and picrosirius staining. Results: The number of fibroblasts, neutrophils and macrophages was similar among all subgroups. There was a higher lymphocyte reaction in the 30-day meshed group (p = 0.003). Giant cell reaction, granulation tissue and neoangiogenesis were similar among the subgroups. Synovial metaplasia was milder at 90-day in the unmeshed (p = 0.002) and meshed group (p < 0.001). Capsular thickness was significantly greater in the meshed samples (30-day p < 0.001 and 90-day p < 0.001). There was a similar amount of collagen types I and III in both groups. Conclusions: The mesh-covered implants produced capsules similar to the microtextured ones when analyzing inflammatory variables. Synovial metaplasia was milder at 90 than at 30 days, and the capsular thickness was significantly greater in the meshed group. A similar amount of collagen types I and III was observed. Due to these characteristics, the mesh coverage did not seem to significantly affect the local inflammatory activity.

Keywords: Breast Implants. Prostheses and Implants. Mammaplasty. Rats.

*Corresponding author: ralfberger_@hotmail.com | (55 42)99151-6496
Received: Jan 09, 2021 | Review: Mar 11, 2021 | Accepted: Apr 13, 2021
Conflict of interest: Nothing to declare.
Research performed at Postgraduate Program in Principles of Surgery, Mackenzie Evangelical School of Medicine, Curitiba (PR), Brazil. Part of Master Degree, Postgraduate Program in Principles of Surgery. Tutor: Prof. Dr. Jurandir Marcondes Ribas Filho.
Introduction

Breast reconstruction can be performed with autologous techniques, using the patient’s own tissues, which is generally cited as the standard procedure. Autologous reconstruction, however, may not be possible in some patients. For example, thin women may not have enough abdominal tissue to enable the reconstruction with rectus abdominis muscle flap. In addition, some women might not be willing to accept the donor site morbidity, extended operative and recovery time, inherent to autologous reconstruction. The presence of comorbidities can also limit the options for reconstruction.

The alternative to autologous reconstruction is the implant-based surgery, in which an important restriction is the inadequate soft tissue coverage, which can lead to skin damage, implant exposure, poor aesthetic results and asymmetry.

An alternative to provide tissue coverage is the acellular dermal matrix, which provides an extra layer and support for the lower pole of the reconstructed breast. The acellular dermal matrix has also reduced complications such as visibility of implant ripples, unstable position and capsular contracture.

Although well established in the literature, the use of acellular dermal matrix is expensive, often prohibitive in Brazil. Therefore, the use of synthetic meshes may be a low-cost option.

The use of meshes made by different materials has been increasingly applied during immediate breast reconstruction with silicone implants. The complication rates when using polypropylene and titanium meshes on silicone implants seem to be similar to those observed in pure silicone implants. However, the use of synthetic meshes entails new scenarios and demand for surgeons to recognize new complications and their histological behavior, since there is lack of knowledge regarding inflammatory alterations on meshes associated with silicone implants.

According to some authors, a capsular contracture with clinical symptoms is related to local inflammatory activity. Several studies have successfully evaluated the use of meshes during breast reconstruction with implants. However, the histological behavior of Parietex Composite® (Covidien, Boulder, United States) associated with silicone implants is not known.

The aim of this study was to evaluate the capsules formed around silicone implants with and without a Parietex Composite® coverage histologically, assessing the mesh effect on inflammatory variables, synovial metaplasia, capsular thickness and collagen types I and III.

Methods

This study was carried out in the vivarium and in the Laboratory of Operative Technique and Experimental Surgery at Universidade Estadual de Ponta Grossa (protocol numbers 13,252/2018 and 3,973/2018), after being approved by the Ethics Committee on the Use of Animals (CEUA), process number 032/2018.

This is a primary interventional prospective non-randomized study. No calculations were performed for the sample size, obtaining a smaller sample based on already published studies similar to this one, facilitating the process of acceptance by the CEUA.

Sixty albino rats (Rattus norvegicus) weighing between 200 and 300 grams, 100 days old, of Wistar strain, were used. The 60 animals were distributed in two groups of 30 rats each (implants with and without mesh coverage), and each group was divided into two subgroups, to be evaluated at 30 and 90 days. Four rats were allocated per 450- cm³ acrylic box, lined with wood shavings. They had free access to water and a specific diet for the species, ad libitum, in addition to alternating light in 12-hour cycles at room temperature.

By the date of the first euthanasia, with 30 days, eight animals in the unmeshed group and five in the meshed group died. One animal from each group was excluded due to the lack of quality of the piece, and two animals from the meshed group by rotation of the mesh-implant set. After that, the following distribution was made (Table 1):

| Groups | Subgroups | 30 days | 90 days |
|--------|-----------|---------|---------|
| Unmeshed | 10 animals | 11 animals |
| Meshed  | 10 animals | 12 animals |

Implanted materials

LifeSil® (Curitiba, PR, Brazil) implants were used, which have the same characteristics as micro-texture implants, except that they are not filled with silicone, constituted only by the 20-mm-diameter microtextured implant cover.

The Parietex Composite® mesh, used to cover the outer surface of the implants in one of the groups, consisted of three-dimensional multifilament polyester with an absorbable, continuous and hydrophilic film on one side. The film consists of porcine collagen, polyethylene glycol and glycerol.
**Surgical procedure**

The animals were anesthetized with intraperitoneal injection of ketamine 10%, 80 mg/kg, and xylazine 2%, 10 mg/kg. No fasting was performed, and they were placed in prone position after trichotomy.

A 1.5-cm-long incision was made in the posteroinferior costal margin, in the midline. The implant pocket was round, with a 5-mm margin from the implants.

The implants were positioned 5 mm from the incision. On the meshed implants, the matrix was positioned on the dorsal side. The suture was performed with four stitches, Prolene® 5.0 (Ethicon, Somerville, New Jersey, United States), and there were no dressings.

Postoperative analgesia was performed with two subcutaneous doses of ketoprofen 5 mg/kg, with an administration interval of 24 hours.

Euthanasia was performed with triple the therapeutic dose of Cetamin®/240–270 mg/kg and Xilazin®/30–40 mg/kg intraperitoneally, followed by cervical dislocation.

**Histological evaluation**

**Hematoxylin and eosin staining**

The procedure was used for the evaluation of inflammatory variables, synovial metaplasia, and capsular thickness.

**Picrosirius coloring**

This technique was used to assess the amount of collagen types I and III. The software AxioVision® 4.9.1.0 (Zeiss, Oberkochen, Germany) was used to obtain the images. The percentage of collagen types I and III was measured using semi-automatic segmentation, in the Image Proplus® 4.5 morphometry program (Media Cybernetics, Rockville, MD, United States).

**Statistical analysis**

For each of the variables, the groups with and without mesh coverage were compared, in the 30 and 90-day subgroups. Then, the subgroups were compared with one another.

The results were described by averages, standard deviations, medians, minimum and maximum values (quantitative variables) or by frequencies and percentages (categorical variables). Fisher’s exact test was used for inflammatory variables, the Mann-Whitney non-parametric test for capsular thickness and Student’s t test for comparison in relation to the percentage of collagen. The significance level of 0.05 was adjusted by applying the Bonferroni correction (p < 0.012). The data were analyzed with the Stata/SE® v. 14.1 (Stata Corporation LLC, College Station, TX, United States) software.

**Results**

**Hematoxylin and eosin staining**

Only the variables with statistical significance were highlighted in the pictures. Table 2 shows the percentage of rats that had each characteristic evaluated as moderate or accentuated.

### Table 2 - Percentage of cases with moderate/intense classification according to the group (meshed or unmeshed) and subgroup (30 days or 90 days).

| Variable          | Subgroup | Group                                                                 |
|-------------------|----------|----------------------------------------------------------------------|
|                   |          | Unmeshed | Meshed | p* (unmeshed × meshed) |
| **Fibroblasts**   |          |          |        |                          |
| 30 days           | 20%      | 70%      |        | 0.070                    |
| 90 days           | 27.3%    | 16.7%    |        | 0.640                    |
| **p* (30d × 90d)**|          | 1        | 0.030  |                          |
| **Neutrophils**   |          |          |        |                          |
| 30 days           | 20%      | 0%       |        | 0.474                    |
| 90 days           | 0%       | 0%       |        | 1                        |
| **p* (30d × 90d)**|          | 0.213    | 1      |                          |
| **Macrophages**   |          |          |        |                          |
| 30 days           | 0%       | 0%       |        |                          |
| 90 days           | 0%       | 0%       |        |                          |
| **p* (30d × 90d)**|          |          |        |                          |
| **Lymphocytes**   |          |          |        |                          |
| 30 days           | 30%      | 100%     |        | 0.003                    |
| 90 days           | 45.4%    | 91.7%    |        | 0.027                    |
| **p* (30d × 90d)**|          | 0.659    | 1      |                          |
| **Granulation tissue** |          |          |        |                          |
| 30 days           | 10%      | 0%       |        | 1                        |
| 90 days           | 0%       | 0%       |        | 1                        |
| **p* (30d × 90d)**|          | 0.472    | 1      |                          |
| **Neoangiogenesis** |          |          |        |                          |
| 30 days           | 10%      | 10%      |        | 1                        |
| 90 days           | 0%       | 16.7%    |        | 0.478                    |
| **p* (30d × 90d)**|          | 0.476    | 1      |                          |
| **Synovial metaplasia** |          |          |        |                          |
| 30 days           | 80%      | 90%      |        | 1                        |
| 90 days           | 9.1%     | 8.3%     |        | 1                        |
| **p* (30d × 90d)**|          | 0.002    | <0.001 |                          |
Giant cell reaction was analyzed only as present or absent. All animals of all groups had the presence of this variable.

Fibroblasts

In the unmeshed group, in both subgroups (30 and 90 days), most animals had a mild presence. In the meshed group, the majority had a moderate presence at 30 days and a mild presence at 90 days. Although the 30 and 90-day unmeshed subgroups and the 90-day meshed subgroup showed a mild presence, no statistical significance was obtained.

Neutrophils

The majority of the animals, in both groups, had a mild presence. No significant differences were found between the two groups in the different subgroups.

Macrophages

This variable had a mild presence in both groups, in all analyzed animals. Thus, there was no statistical comparison.

Lymphocytes

In the unmeshed group, the presence was mild in the 30-day subgroup, whereas in the meshed group the majority of the animals exhibited a moderate or intense presence of this variable in both subgroups.

When comparing the 30-day meshed and unmeshed groups, statistical significance was obtained (p = 0.003) (Fig. 1).

**Figure 1** - Photomicrography of microtextured implant (a) and meshed implant (b), showing lymphocytes.

**Giant cell reaction**

This reaction was only analyzed as absent or present, and all animals in the four subgroups had this characteristic. Thus, there was no statistical comparison.

**Granulation tissue**

The vast majority of animals had a mild presence of this variable. When the groups and subgroups were compared, there was no statistical significance.

**Neoangiogenesis**

In all subgroups, the majority of the rats had a mild presence of the variable. When the groups and subgroups were compared with one another, there was no statistical significance.

**Synovial metaplasia**

In the 30-day subgroups, a moderate or intense presence of this variable was found in most animals, while in the 90-day subgroups most of them had a mild presence. In both groups, when comparing 30 and 90-day subgroups, there was statistically significant difference (unmeshed p = 0.002/meshed p<0.001) (Fig. 2).

**Figure 2** - Photomicrography of microtextured implant (a) and meshed implant (b), showing synovial metaplasia.

**Capsule thickness**

This finding was lower in the unmeshed compared to the meshed group, with statistical significance (30 days p < 0.001 / 90 days p < 0.001) (Fig. 3).
Figure 3 - Photomicrography of microtextured implant (a) and meshed implant (b), showing capsular thickness (magnification x20).

Table 3 contains the median with the minimum and maximum values of capsular thickness. Table 4 contains p-values.

Table 3 - Median, minimum and maximum values of the capsule thickness (μm) according to the group (meshed or unmeshed) and the subgroup (30 and 90 days).

| Variable | Subgroup     | Group    | Unmeshed | Meshed |
|----------|--------------|----------|----------|--------|
|          |              |          | 30 days  | 90 days |
| Thickness| 30 days      | 70.4     | (35.3–144) | 683.3  | (566.7–766.7) |
|          | 90 days      | 56.7     | (32.3–93.7) | 633.3  | (500–700) |

Table 4 – Compared groups and subgroups in relation to capsular thickness with p-value.

|                  | Collagen I (%) |
|------------------|----------------|
|                  | Avg. | Median | Min. | Max. | SD  |
| Unmeshed 30 days | 63.2 | 71.6   | 24.4 | 82   | 20.7|
| Meshed 30 days   | 53.5 | 51     | 40.2 | 75.9 | 10.9|
| Unmeshed 90 days | 64.6 | 68     | 46.8 | 88   | 12.2|
| Meshed 90 days   | 50   | 49.9   | 26.2 | 69   | 13.7|

Mann-Whitney non-parametric test, p < 0.012 (Bonferroni correction).

Picrosirius staining

Collagen types I and III

The Fig. 4 shows type I collagen in reddish color and type III collagen in greenish color. In the meshed group, the matrix is exhibited by the bluish color.

Figure 4 - Photomicrography evidencing collagen fibers. (a) 30-day unmeshed group, (b) 90-day unmeshed group, (c) 30-day meshed group, (d) 90-day meshed group. Note: Red: type I collagen; Green: type III collagen; Blue: mesh (picrosirius staining, magnification x400, polarized light).

In the unmeshed group, in both subgroups, the averages were slightly higher for collagen type I. However, when the groups and subgroups were compared with one another, no statistical significance was found (Tables 5 and 6).

Table 5 - Descriptive statistics of collagen type I according to the subgroups.

| Group | Days | n  | Collagen I (%) | Avg. | Median | Min. | Max. | SD  |
|-------|------|----|----------------|------|--------|------|------|-----|
| Unmeshed | 30   | 10 | 63.2           | 71.6 | 24.4   | 82   | 20.7 |
| Meshed | 10   | 10 | 53.5           | 51   | 40.2   | 75.9 | 10.9 |
| Unmeshed | 90   | 11 | 64.6           | 68   | 46.8   | 88   | 12.2 |
| Meshed | 12   | 12 | 50            | 49.9 | 26.2   | 69   | 13.7 |

Avg: Average; Min: Minimum; Max: Maximum; SD: Standard Deviation
Histological evaluation of capsules formed by texturized silicone implants with and without polyester mesh coverage (Parietex®). A study on female rats

Table 6 – Compared groups and subgroups in relation to collagen type I with p-value.

|       | 30 days | 90 days | Unmeshed | Meshed |
|-------|---------|---------|----------|--------|
| unmeshed × meshed | 0.209 | 0.012 |
| 30 days × 90 days | 0.843 | 0.519 |

Table 8 – Compared groups and subgroups in relation to collagen type III with p-value.

|       | 30 days | 90 days | Unmeshed | Meshed |
|-------|---------|---------|----------|--------|
| unmeshed × meshed | 0.209 | 0.012 |
| 30 days × 90 days | 0.843 | 0.519 |

The amount of collagen type III was similar between groups and subgroups. When the groups and subgroups were compared, there was no statistical significance (Tables 7 and 8).

Table 7 - Descriptive statistics of collagen type III according to the subgroups.

| Group     | Days | n  | Collagen III (%) | Avg | Median | Min. | Max. | SD |
|-----------|------|----|------------------|-----|--------|------|------|----|
| Unmeshed  | 30   | 10 |                  | 36.8| 28.4   | 18   | 75.6 | 20.7|
| Meshed    | 30   | 10 |                  | 46.5| 49     | 24.1 | 59.8 | 10.9|
| Unmeshed  | 90   | 11 |                  | 35.4| 32     | 12   | 53.2 | 12.2|
| Meshed    | 90   | 12 |                  | 50  | 50.1   | 31   | 73.8 | 13.7|

Avg: Average; Min: Minimum; Max: Maximum; SD: Standard Deviation

Figure 5 - Average, standard errors and standard deviations of area percentages with collagen type I in each subgroup.

Figure 6 - Average, standard errors and standard deviations of area percentages with type III collagen in each subgroup.

Discussion

The rat (Rattus norvegicus albinus) chosen by the authors is the most used animal in capsular contracture studies, for presenting easy reproducibility of results and resistance to surgical procedures.25-27

Due to the difficulty in obtaining large animal samples for research in our institution, we have based our sample size on already published studies similar to this, which also used animals for experimentation. Thus, no calculations were performed for the sample size, obtaining a smaller sample, facilitating the process of acceptance by the CEUA. Since it’s a small sample, there may have been a loss of statistical power in the analysis of some variables. Despite differing percentages in their values, we need to rely on the stipulated significance range (p<0.012 with Bonferroni correction).
E-covered implants. This study disagrees with Bergmann to implants that used other types of coverage, namely: capsular thickness in textured implants when compared the unmeshed group. Other authors also found smaller inflammatory variables when studying the Parietex® mesh.

In the capsular thickness, the alignment of collagenous fibers, the presence of contractile microfibroblasts, and greater alpha-SMA expression.

Due to this association between inflammation, capsular thickness and contracture, we opted for analyzing the greater alpha-SMA expression.

Haddad Filho et al.35 found a higher number of neutrophils in the mesh-covered group at 30 days, unlike this study, that did not find differences in the neutrophil count.

All capsules under analysis presented similar number of macrophages, which is in agreement with other studies23,35.

In the meshed group, the number of lymphocytes was higher at 30 days, contradicting Haddad Filho et al.35, who found similar numbers between the textures and PTFE-E covered groups.

Giant cell reaction was observed in all samples, which is in agreement with other studies that compared textured implants to implants using different types of coverage26-28,33,35. Unlike Silva et al.28, who found intense formation of granulation tissue in polyurethane implants.

Neovascular formation was essentially mild in all subgroups, corroborating findings by Silva et al.28 in subgroups from the same evaluation period. This result opposes to the one by Haddad Filho et al.35, who found, in the unmeshed subgroup, greater intensity of vascular formation in the 90-day subgroups when compared to the 30-day subgroups. Those authors also reported higher neoangiogenesis in the PTFE-E group at 30 days. Other authors also found more intense neovascularization in the presence of another coverage in addition to the textured one27,34.

Bergmann et al.25, however, reported intense neovascular formation in the textured group when compared to the titanium-covered mesh group.

This study partially agrees with Prantl et al.9, who evaluated implant capsules in humans and found the presence of synovial metaplasia in most of them, whereas in this study this variable was present in all animals of both groups.

Unlike Bassetto et al.26, who found similar synovial metaplasia between the subgroups, this study showed a more pronounced presence of this characteristic in the 30-day subgroups.

The moderate and accentuated presence of synovial metaplasia at 30 days and mild at 90 days differs from the findings by Silva et al.28, who detected an absent or mild
presence throughout the evaluation period, despite the fact that those researchers compared textured implants with polyurethane implants.

The findings of this study are close to those found by Hansson et al.\textsuperscript{24}, who compared the use of biological and synthetic meshes and reported the presence of synovial metaplasia in most cases, since in this research this variable was observed in all cases.

Alterations in collagenous fibers might be present in capsular contracture cases\textsuperscript{31,32,37}. Therefore, they were analyzed in this study. Brazin et al.\textsuperscript{38} studied patients with grade IV Baker contracture and concluded that the capsule collagenous production by fibroblasts is mediated by mastocytes.

In agreement with Minami et al., who observed a slight increase in collagen type III in the textured implant group, in this study the results in the unmeshed group (microtextured implant) in the 30 and 90-day subgroups were similar. Those authors also found a slight decrease in collagen type III in the textured group from 30 to 90 days. Similar results were found in this study in the 30 and 90-day subgroups.

This study disagrees with Balderrama et al.\textsuperscript{33}, who found a significant decrease in the amount of type III collagen in the textured group in the 30 to 60-day subgroups, because in this study type III collagen remained similar in the unmeshed group. Those authors also found a significant increase in the amount of type I collagen in the subgroup from 30 to 60 days, whereas in this study, in the unmeshed group, a similar amount of type I collagen was found in the subgroups analyzed.

Differing from Silva et al.\textsuperscript{28}, the percentage of collagen types I and III was similar between textured implants and those with additional coating in all subgroups analyzed, despite the fact that those researchers used polyurethane implants for comparison. Due to these characteristics, the mesh coverage did not seem to significantly affect the local inflammatory activity.

## Conclusions

The implants covered by Parietex Composite® mesh produced capsules similar to those ones found in textured implants when analyzing inflammatory variables. Synovial metaplasia was milder at 90 than at 30 days, and the capsular thickness was significantly greater with the mesh coating. A similar amount of collagen types I and III was formed in the meshed and unmeshed implant capsules. Due to these characteristics, the mesh coating did not seem to significantly affect the local inflammatory activity.

## Author’s contribution

Design the study: Berger R; Acquisition of data: Souza MA; Acquisition and analysis of data: Berger R; Interpretation of data: Montemor Netto MR, Noronha L and Paula PH; Technical procedures: Souza MA; Histopathological examinations: Noronha L; Manuscript writing: Berger R and Paula PH; Critical revision: Ribas Filho JM, Malafaia O, Silva EM, Silva ABD, Takejima M and Montemor Netto MR.

## Data availability statement

Data will be available upon request.

## Funding

Coordenação de Aperfeiçoamento de Pessoal de Nível Superior [https://doi.org/10.13039/501100002322] Grant 001

## Acknowledgments

Not applicable.

## References

1. Recht, A, Edge SB, Solin LJ, Robinson DS, Estabrook A, Fine RE, Fleming GF, Formenti S, Hudis C, Kirshner JJ, Krause DA, Kuske RR, Langer AS, Langer RR, Miettinen OA, Pfister DG, American Society of Clinical Oncology. Postmastectomy radiotherapy: clinical practice guidelines of the American Society of Clinical Oncology. J Am Soc Clin Oncol. 2001;19(5):1539–69. https://doi.org/10.1200/JCO.2001.19.5.1539

2. Wang HY, Ali RS, Chen SC, Chao TC, Cheng MH. One-stage immediate breast reconstruction with implant following skin-sparing mastectomy in Asian patients. Ann Plast Surg. 2008;60(4):362–6. https://doi.org/10.1097/SAP.0b013e318063ef70

3. Malata CM, McIntosh SA, Purushotham AD. Immediate breast reconstruction after mastectomy for cancer. Br J Surg. 2000;87(11):1455-72. https://doi.org/10.1046/j.1365-2168.2000.01593.x

4. Agha-Mohammadi S, De La Cruz C, Huvitz DJ. Breast reconstruction with alloplastic implants. J Surg Oncol. 2006;94(6):471-8. https://doi.org/10.1002/jso.20484

5. Breuing KH, Warren SM. Immediate bilateral breast reconstruction with implants and inferolateral AlloDerm slings. Ann Plast Surg. 2005;55(3):232–9. https://doi.org/10.1097/01.sap.0000168527.52472.3c

6. Liu J, Hou J, Li Z, Wang B, Sun J. Efficacy of acellular dermal matrix in capsular contracture of implant-based breast reconstruction. J Plast Reconstr Aesthet Surg. 2018;71(3):459-65. https://doi.org/10.1016/j.bjps.2017.11.009
15. Prantl L, Schreml S, Fichtner-Feigl S, Pöppl N, Eisenmann-
Klein, M. Serologic and histologic findings in patients
with capsular contracture after breast augmentation with
smooth silicone gel implants: is serum hyaluronan a
potential predictor? Aesthetic Plast Surg. 2005;29(6):510–
8. https://doi.org/10.1007/s00266-005-5049-y

9. Poeppl N, Schreml S, Lichtenegger F, Lenich A, Eisenmann-
Klein M, Prantl L. Does the surface structure of implants have an impact on the formation of a capsular contracture? Aesthetic Plast Surg. 2007;31(2):133-9.
https://doi.org/10.1007/s00266-006-0091-y

10. Haynes DF, Kreithen JC. Vicryl mesh in expander/implant
breast reconstruction: long-term follow-up in 38 patients. Plast Reconstr Surg. 2014;134(5):892-9. https://doi.org/10.1097/PR.0000000000000610

11. Dieterich M, Angres J, Stubert J, Stachs A, Reimer T, Gerber B. Patient-reported outcomes in implant-based breast reconstruction alone or in combination with a titanium-coated polypropylene mesh - A detailed analysis of the BREAST-Q and overview of the literature. Geburtshilfe Frauenheilkd. 2015;75(7):692-701. https://doi.org/10.1055/s-0035-1546218

12. Baldelli I, Cardoni G, Franchelli S, Fregatti P, Friedman D, Pesce M, Ponte E, Santori G, Santi P. Implant-based breast reconstruction using a polyester mesh (Surgimesh-
PET): a retrospective single-center study. Plast Reconstr Surg. 2016;137(6):931e-9. https://doi.org/10.1097/PRAS.0000264398.85652.9a

13. Caputo GG, Marchetti A, Dalla Pozza E, Vigato E, Domenici L, Cigna E, Governi M. Skin-reduction breast reconstructions with prepectoral implant. Plast Reconstr Surg. 2016;137(6):1702-5. https://doi.org/10.1097/PRAS.000000000002227

15. Gschwantler-Kaulich D, Schrenk P, Bjelic-Radisic V, Unterrieder K, Leser C, Fink-Retter A, Salama M, Singer C. Mesh versus acellular dermal matrix in immediate implant-based breast reconstruction - A prospective randomized trial. Eur J Surg Oncol. 2016;42(5):665-71. https://doi.org/10.1016/j.ejso.2016.02.007

16. Pukancik D, Kelemen P, Gulyás G, Újhelyi M, Kovács E, Éles K, Mészáros N, Kencesyi I, Pálházi P, Kocsis T, Káslér M, Márton Z. Clinical experiences with the use of ULTRAPRO® mesh in single-stage direct-to-implant immediate postmastectomy breast reconstruction in 102 patients: a retrospective cohort study. Eur J Surg Oncol. 2017;43(7):1244-51. https://doi.org/10.1016/j.
ejso.2017.01.236

17. Zenn M, Venturi M, Pittman T, Spear S, Gurtner G, Robb G, Mesbah A, Dayan J. Optimizing outcomes of postmastectomy breast reconstruction with acellular dermal matrix: a review of recent clinical data. Eplasty. 2017;17:e18.

18. Momeni A, Kanchwala SK. Improved pocket control in immediate microsurgical breast reconstruction with simultaneous implant placement through the use of mesh. Microsurgery. 2018;38(5):450-7. https://doi.org/10.1002/micr.30123

19. Pompei S, Evangelidou D, Arelli F, Ferrante G. The use of TIGR matrix in breast aesthetic and reconstructive surgery: is a resorbable synthetic mesh a viable alternative to acellular dermal matrices? Clin Plast Surg. 2018;45(1):65-73. https://doi.org/10.1016/j.cps.2017.08.005

20. Gfrerer L, Liao EC. Technique refinement in prepectoral implant breast reconstruction with vicryl mesh pocket and acellular dermal matrix support. Plast Reconstr Surg Glob Open. 2018;6(4):e1749. https://doi.org/10.1097/GOX.0000000000001749

21. Potter S, MacKenzie M, Blazeby JM. Does the addition of mesh improve outcomes in implant based breast reconstruction after mastectomy for breast cancer? BMJ. 2018;362:k2607. https://doi.org/10.1136/bmj.k2607

22. Bonomi S, Sala L, Gennaro M, Ricci C, Cortinovis U. Skin-reducing mastectomy and direct-to-implant breast reconstruction with submuscular-dermal-mesh pocket. Ann Plast Surg. 2019;82(1):19-27. https://doi.org/10.1097/SAP.0000000000001614

23. Casella D, Di Taranto G, Marcasciano M, Sordi S, Kothari A, Kovacs T, Lo Torto F, Cigna E, Calabrese C, Ribuffo D. Evaluation of prepectoral implant placement and complete coverage with TiLoop Bra mesh for breast reconstruction: a prospective study on long-term and patient-reported BREAST-Q outcomes. Plast Reconstr Surg. 2019;143(1):1e-9e. https://doi.org/10.1097/PRS.0000000000005078

24. Hansson E, Burian P, Hallberg H. Comparison of inflammatory response and synovial metaplasia in immediate breast reconstruction with a synthetic and a biological mesh: a randomized controlled clinical trial. J Plast Surg Hand Surg. 2020;54(3):131–6. https://doi.org/10.1080/2000656X.2019.1707466.

25. Bergmann PA, Tamouridis G, Lohmeyer JA, Mauss KL, Becker B, Knobloch J, Mailänder P, Siemers F. The effect of a bacterial contamination on the formation of capsular contracture with polyurethane breast implants
in comparison with textured silicone implants: an animal study. J Plast Reconstr Aesthetic Surg. 2014;67(10):1364–70. http://doi.org/10.1016/j.bjps.2014.05.04.

26. Mendes PRDS, Bins-Ely J, Lima EADS, De Vasconcellos ZAA, D’Acampora AJ, Neves RDE. Histological study on acute inflammatory reaction to polyurethane-coated silicone implants in rats. Acta Cir Bras. 2008;23(1):93–101. http://doi.org/10.1590/S0102-86502008000100015

27. Wagenführ-Júnior J, Ribas Filho JM, Nascimento MM do, Ribas FM, Wanka MV, Godoi A by L. Histopathological reaction over prosthesis surface covered with silicone and polyurethane foam implanted in rats. Acta Cir Bras. 2012;27(12):866–73. http://dx.doi.org/10.1590/S0102-86502012001200007

28. Silva EN, Ribas-Filho JM, Czeczko NG, Pachnicki JPA, Netto MRM, Lipinski LC, Noronha L, Colman J, Zeni JO, Carvalho CA. Histological evaluation of capsules formed by silicone implants coated with polyurethane foam and with a textured surface in rats. Acta Cir Bras. 2016;31(12):774–82. https://doi.org/10.1590/s0102-86502016012000001

29. Silva EN, Ribas-Filho JM, Tabushi FI, Silva MAP, Siqueira EBD, de Noronha L, da Silva ABD, Lipinski LC, Guth I, Vosgerau LM. Smooth muscle alpha actin immunoeexpression (α-Sma) and CD-117 antibody (C-Kit) in capsules formed by polyurethane foam-coated silicone implants and with textured surface: a study on rats. Aesthetic Plast Surg. 2019;43(1):233-42. https://doi.org/10.1007/s00266-018-1238-3

30. Biondo-Simões MLP, Sichiciopi AA, Ioshii SO, Robes RR, Biondo-Simões R. Comparative study of fibrosis induced by Marlex®, Parietex Composite®, Vicryl® and Ultrapro® meshes. Acta Cir Bras. 2018;33(9):792-8. https://doi.org/10.1590/s0102-86502018009000007

31. Minami E, Koh IJH, Ferreira JCR, Wartzberg AFI, Chifferi V, Rosewick TF, Pereira MD, Saldiva PHN, de Figueiredo LFP. The composition and behavior of capsules around smooth and textured breast implants in pigs. Plast Reconstr Surg. 2006;118(4):874–84. https://doi.org/10.1097/01.pr.s.0000240878.24213.b7

32. Bui JM, Perry TA, Ren CD, Nofrey B, Teitelbaum S, van Epps DE. Histological characterization of human breast implant capsules. Aesthetic Plast Surg. 2015;39(3):306–15. http://doi.org/10.1007/s00266-014-0439-7

33. Balderrama CMR, li JMR, li OM, Gregori N, li C. Healing reaction to mammary prostheses covered by textured silicone and silicone. Acta Cir Bras. 2009;24(5):367–76. http://doi.org/10.1590/S0102-86502009000500006

34. Vieira VJ, d'Acampora AJ, Marcos ABW, Di Giunta G, de Vasconcellos ZAA, Bins-Ely J, d'Eça Neves R, Figueiredo CP. Vascular endothelial growth factor overexpression positively modulates the characteristics of periprosthetic tissue of polyurethane-coated silicone breast implant in rats. Plast Reconstr Surg. 2010;126(6):1899–910. https://doi.org/10.1097/PRS.0b013e3181f446d5

35. Haddad Filho D, Zveibel DK, Alonso N, Gernaerli R. Comparison between textured silicone implants and those bonded with expanded polytetrafluoroethylene in rats. Acta Cir Bras. 2007;22(3):187–94. http://doi.org/10.1590/S0102-86502007000300006

36. Bassetto F, Scarpa C, Caccialanza E, Montesco MC, Magnani P. Histological features of periprosthetic mammmary capsules: silicone vs. polyurethane. Aesthetic Plast Surg. 2010;34(4):481–5. https://doi.org/10.1007/s00266-010-9483-0

37. Moyer KE, Ehrlich HP. Capsular contracture after breast reconstruction: Collagen fiber orientation and organization. Plast Reconstr Surg. 2013;131(4):680–5. https://doi.org/10.1097/PRS.0b013e31828189d0

38. Brazin J, Malliaris S, Groh B, Mehrara B, Hidalgo D, Otterburn D, Silver RB, Spector JA. Mast cells in the periprosthetic breast capsule. Aesthetic Plast Surg. 2014;38(3):592–601. https://doi.org/10.1007/s00266-014-0318-2