Data Article

Australian aquatic bio-optical dataset with applications for satellite calibration, algorithm development and validation

Nathan Drayson\textsuperscript{a,∗}, Janet Anstee\textsuperscript{a}, Hannelie Botha\textsuperscript{a}, Gemma Kerrisk\textsuperscript{a}, Phillip Ford\textsuperscript{a}, Bozena Wojtasiewicz\textsuperscript{b}, Lesley Clementson\textsuperscript{b}, James McLaughlin\textsuperscript{c}, Marlee Hutton\textsuperscript{c}

\textsuperscript{a}CSIRO, Oceans and Atmosphere, Black Mountain, Canberra, ACT 2601, Australia
\textsuperscript{b}CSIRO, Oceans and Atmosphere, Castray Esplanade, Battery Point TAS 7004, Australia
\textsuperscript{c}CSIRO, Oceans and Atmosphere, Crawley, WA 6009, Australia

A R T I C L E   I N F O

Article history:
Received 20 June 2022
Revised 14 July 2022
Accepted 20 July 2022
Available online 26 July 2022

Keywords:
Aquatic bio-optical properties
Inland water quality
Aquatic remote sensing reflectance
Aquatic remote sensing algorithm development

A B S T R A C T

The authors present bio-optical data spanning 316 sets of observations made at 34 inland waterbodies in Australia. The data was collected over the period 2013–2021 and comprise radiometric measurements of remote sensing reflectance (Rrs), diffuse attenuation extinction coefficient (Kd); optical backscattering; absorption of coloured dissolved organic matter (aCDOM), phytoplankton (aph) and non-algal particles (aNAP); HPLC analysis of algal pigments including chlorophyll-a (CHL-a); organic and inorganic total suspended solids (TSS); and total and dissolved organic carbon concentration. Data collection has been timed to coincide with either Landsat 8 or Sentinel-2 overpasses. The dataset covers a diverse range of optical water types and is suitable for algorithm development, satellite calibration and validation as well as machine learning applications.

© 2022 The Author(s). Published by Elsevier Inc. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/)

∗ Corresponding author.
E-mail address: Nathan.drayson@csiro.au (N. Drayson).

https://doi.org/10.1016/j.dib.2022.108489
2352-3409/© 2022 The Author(s). Published by Elsevier Inc. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/)
### Specifications Table

| Subject                      | Hydrology and Water quality          |
|------------------------------|--------------------------------------|
| Specific subject area        | Earth observation water quality      |
| Type of data                 | Table                                |
| How the data were acquired   | In situ instruments:                 |
|                              | Backscattering - WetLabs BB9, Wetlabs ECOTriplet BB2; |
|                              | Radiometry – RAMSES ACC-2 VIS planar irradiance sensor, RAMSES ACC-2 |
|                              | diffuse irradiance sensor, RAMSES ARC VIS radiance sensor |
|                              | Laboratory analysis of water samples: |
|                              | Absorption coefficients - dual-beam spectrophotometer (GBC Scientific Equipment Ltd., Cintra 404); software: Cintral ver. 2.2); |
|                              | Pigments – HPLC method described in [1] |
|                              | TSS – gravimetric determination on pre-rinsed, pre-weighed glass fibre filters. |
|                              | Total and dissolved organic carbon – Shimadzu Total Organic Carbon analyser after acidification and purging with high purity nitrogen. |
| Data format                  | Raw, analysed                        |
| Description of data collection | Data was collected using in situ instrumentation and laboratory analysis of water samples. |
| Data source location         | Institution: CSIRO                   |
|                              | City/Region: ACT                      |
|                              | Country: Australia                    |
| Data accessibility           | Repository name: CSIRO Data Access Portal |
|                              | Data identification number: [https://doi.org/10.25919/rtd7-j815](https://doi.org/10.25919/rtd7-j815) |
|                              | Direct URL to data: [https://data.csiro.au/collection/csiro:54602v1](https://data.csiro.au/collection/csiro:54602v1) |

### Value of the Data

- Unique description of a broad range of bio-optical properties for Australian inland waters.
- Basis for parameterisation and training for semi-analytical and machine learning inversion algorithms.
- Validation dataset for inversion algorithms.
- Examining relationships between inherent optical properties to validate machine learning training datasets.
- Determine relationship between apparent and inherent optical properties of lakes.

### 1. Data Description

Monitoring water pollution using remote sensing offers a greater understanding of spatial and temporal distribution of pollutants than traditional methods of in situ data collection [2,3]. The calibration and validation of models used to predict concentrations of optically active constituents (OACs) from remotely sensed data require representative samples from a diverse set of optical water types [2,4,5]. The dataset described in this article [6] provides a diverse set of bio-optical data collected from Australian inland waterbodies between 2013 and 2021.

The dataset described in this article contains ten datafiles. Each datafile contains observations for a specific modality of bio-optical measurement. Measurements from each modality can be merged using the observation_id column, which provides a unique identifier for each set of observations.

The observation data file contains metadata and site observations. This file defines the location, date, time, and name of waterbody for each observation set. Data on cloud cover, water conditions, temperature, Secchi depth and colour are also provided. Fig. 1 shows the locations of selected waterbodies described by this dataset. Column definitions of the observation data file are provided in Table 1.

Absorption coefficients and related parameters are presented in three data files, absorption_unfitted, absorption_fitted and absorption_slopes. Of these the unfitted datafile contains data obtained through direct measurement, while the absorption_fitted and absorption_slopes
Fig. 1. Locations of a selection of waterbodies.

Table 1
Data description of observations metadata file.

| Column name               | Unit    | Description                                                                 |
|---------------------------|---------|----------------------------------------------------------------------------|
| observation_id            | ID      | Unique identifier for each measurement set                                  |
| site                      | ID      | Unique identifier for each field site                                       |
| datetime_utc              | date    | date and time measurement acquisition commenced in UTC.                    |
|                           | time    | Datetime displayed as YYYY-MM-DD:HH:MM                                        |
| waterbody                 | string  | Name of waterbody                                                           |
| time_arrived_local        | date    | Local date time measurement acquisition commenced.                          |
|                           | time    | Local time displayed as YYYY-MM-DD:HH:MM                                     |
| local_time_zone           | string  | Designation of local time zone                                              |
| lat                       | float   | Latitude in decimal degrees obtained with EPSG:4326                         |
| lon                       | float   | Longitude in decimal degrees obtained with EPSG:4326                        |
| water_temp_degC           | float   | Surface temperature of the water in degrees Celsius                           |
| cloud_frac_pc             | int     | Percent fraction of cloud cover                                              |
| water_colour              | string  | Short description of water colour                                            |
| water_cond                | string  | Short description of water surface conditions                                |
| wind_speed_kn            | float   | Wind speed in knots                                                          |
| wind_direction            | string  | Cardinal wind direction                                                     |
| weather                   | string  | Short description of weather conditions                                      |
| secchi_depth_m            | float   | Secchi disc depth in meters                                                  |
| water_depth_m             | float   | Total water depth in meters                                                  |
| turbidity_ntu             | float   | Turbidity in Nephelometric Turbidity Unit (NTU)                              |

datafiles are derived from the unfitted data as described below (see 2.1). A total of 307 absorption measurements are available. The distribution of the absorption budget at 440 nm (Fig. 2) indicates that non-algal particles (NAP) and coloured dissolved organic matter (CDOM) are the primary absorption components for most observations in the dataset, with phytoplankton absorption (aph) dominating in comparatively few observations.

The pigments datafile contains three hundred and eight observations of 28 algal pigments obtained using the HPLC method described in Clementson [1]. Chlorophyll-a (chl-a) concentrations show an approximately log-normal distribution in the dataset (Fig. 3). The mean and median chl-a concentrations for the dataset are 22.2 and 7.0 μg L⁻¹, respectively.
Fig. 2. Absorption budget at 400 nm for samples in the absorption_fitted datafile.

Fig. 3. Distribution of chlorophyll-a concentrations.

The org_c datafile contains 174 observations of total and dissolved organic carbon. Concentrations of total organic carbon (TOC) range from 1.1 to 50 mg L^{-1} with dissolved organic carbon being the dominant form in the majority of samples.

The tss data file contains 309 measurements of total suspended solids (TSS). Observations of TSS range from 0.35 to 1786 mg L^{-1}. The contribution of inorganic and organic particles to TSS is evenly distributed with 51% of samples comprised primarily of inorganic particles.

Radiometric measurements are presented in two datafiles, radiometry and kd. One-hundred and twenty remote sensing reflectance (Rrs) observations are provided in the radiometry datafile while seventy-four observations of the irradiance attenuation coefficient (Kd) are provided in
the $kd$ data file. All radiometric data are provided in the spectral range of 350–900 nm at 1 nm resolution.

Two hundred and five particulate backscattering ($b_{bp}$) measurements are available in the $bb\_surface$ datafile. Particulate backscattering values at 555nm predominate in the range 0.007–1 m$^{-1}$ but values up to 4.76 m$^{-1}$ were measured (Fig. 5). Observations at the higher end of this range were obtained through serial dilution of surface samples (see methods). The spectral slopes ($\gamma_{bbp}$, Eq. (12)) of particulate backscattering are approximately normally distributed (Fig. 5). Some negative values of the spectral slope were obtained in low scattering waters and some waters with high concentrations of algal biomass.

2. Experimental Design, Materials and Methods

2.1. Absorption Measurements

The $absorption\_unfitted$ datafile contains absorption coefficients for coloured dissolved organic matter (aCDOM), particulate absorption (ap) and detrital non-algal particulates (ad).

CDOM absorption coefficients were measured from surface (10–20 cm depth) water samples collected in 1L acid washed Schott bottles using the method described in Clementson et al. [7]. 80 mL of water was vacuum filtered using a 0.2 μm Whatman Anodisc filter to separate particulate matter. Filtered water samples were covered with aluminium foil to prevent light degradation and preserved 0.5 mL of a 10% w/v solution of sodium azide (NaN$_3$). Samples were stored chilled and filtered within 24 h of collection and stored on ice for transport to the laboratory for analysis.

Particulate absorption samples were collected as for CDOM, using a clean 5 L polyethylene container. Samples were stored on ice and filtered using Whatman GF/F filters. Filters were stored flat in cryo-cages, covered in aluminium foil, and stored in liquid nitrogen until analysis.

CDOM samples were gradually warmed to room temperature and transferred to a 10 cm quartz cell with their absorbance spectra measured from 210 to 900 nm using a Cintra 404 UV/vis dual-beam spectrophotometer and Milli-Q water (Millipore) as a reference. Absorption coefficients were calculated using Eq. (1) where $A(\lambda)$ is the absorbance normalised to zero at 680 nm and $l$ is the cell pathlength in meters [7].

$$a(\lambda)_{CDOM} = 2.3[A(\lambda)/l]$$

(1)

$$a(\lambda) = a_{350}exp[-S(\lambda-350)] + b$$

(2)
Fig. 5. Characteristics of backscattering data, left particulate backscattering spectra, top right: distribution of particulate backscattering spectral slope, bottom right: particulate backscattering at 555 nm. Note backscattering spectra plotted using Eq. (12).
Absorbance scans for total particulate and non-algal particulate matter were obtained using a Cintra 404 UV/vis dual-beam spectrophotometer equipped with integrating sphere. Particulate optical density spectra were obtained using glass plates to hold sample and blank filters against an integrating sphere. Blank filters, from the same batch as the sample filters, were wetted with small volumes of filters sample and used as a reference. Optical density scans were made from 210 to 900 nm with a spectral resolution of 0.85 nm. A methanol extraction was then used to remove phytoplankton from the filter [8]. The filters were re-scanned to obtain non-algal particulate (NAP) optical density spectra.

Absorbance scans were corrected for path length amplification using the coefficients from Mitchell [9] to obtain absorption coefficients. Measured data are provided in the `absorption_unfitted` datafile.

Fitted and derived absorption values are provided in the `absorption_fitted` and `absorption_slopes` datafiles, respectively. The fitted datafile contains absorption coefficients for phytoplankton ($a_{ph}$), CDOM ($a_{cdom}$) and non-algal particulate matter ($a_{nap}$).

Fitted CDOM and NAP absorption data are obtained by fitting measured absorption coefficients to Eq. (2) where $a_{350}$ is the absorption coefficient at 350 nm, $S$ is the spectral slope, $\lambda$ is the wavelength in nm and $b$ is an offset used in baseline correction. Fitted data remove imperfections in the measured spectra such as residual phytoplankton absorption in the NAP measurement. Phytoplankton absorption spectra are calculated as the difference between particulate absorption and detrital absorption from the `absorption_unfitted` datafile. The $S$ parameter from Eq. (2) is included in the `absorption_slopes` datafile.

Total particulate absorption spectra were smoothed using a 10 nm running boxcar filter and the fitted NAP spectra subtracted to obtain phytoplankton absorption.

Quality control flags are provided for the absorption data to indicate data with missing spectral components, derived values, unusual shapes and suspected sampling or laboratory errors. Quality flags are provided to enable users to evaluate the data for their purposes.

Quality flag 1 indicates that some data have been provided with nominal values as the absorption component was below the detection limit. In these cases, a nominal absorption spectrum of $10^{-6}$ m$^{-1}$ at every wavelength was included to enable users to derive specific inherent optical properties from the data. On one field trip CDOM absorption spectra were negative at some sites. As the water body was small and relatively homogeneous non-negative CDOM spectra were averaged to provide estimated absorption coefficients. Tables 2 and 3 show the column definitions for the `absorption_unfitted` and `absorption_fitted` datafiles. Table 4 shows the quality flags and their definitions.

### 2.2. Pigments

The HPLC method described in Clementson [1] was used to obtain the concentration of 28 algal pigments including chlorophyll-a. Samples for pigment analysis were collected in clean 5L HDPE containers at a depth of 10–20 cm, cooled for storage and filtered within 24 h. Vacuum
filtration was performed using Whatman® glass microfiber filters, Grade GF/F, 0.7 μm until the filter paper was coloured by the sample. Typical filtration volumes ranged from 0.5 to 2 L depending on the concentration of chlorophyll and non-algal particulates present in the sample. Filter papers were folded in half and stored in liquid nitrogen until analysis. Prior to analysis filter papers are thawed and cut into 3–4 pieces. Pigments are extracted by centrifuge with 3 mL of 100% acetone. The samples are then chilled in an ice bath and then kept in the dark at 4°C for 15 h. Water is then added to the extraction mixture to make a 90:10 acetone:water solution and the sample is sonicated. The sample is centrifuged at 2500 rpm for five minutes at -2°C to separate the extract from the filter paper. The filtrate is then passed through a 0.2 μm Teflon syringe filter into a 2 mL amber HPLC vial. An auto sampler chilled to 4°C is used to apply an aqueous tetrabutyl ammonium acetate (TbAA) methanol solution immediately prior to sample injection. Following injection pigments are separated on a Zorbax Eclipse XDB-C8 stainless steel 150×4.6 mm chromatographic column and gradient eluted using a TbAA:methanol solvent [1]. Separated pigments are detected using a PDA detector and identified against standard spectra.

Pigments data are collected in the *pigments* datafile. All pigments are listed with concentration units of μg L⁻¹. The ‘tot_mv_chl_a’ column contains total chlorophyll-a concentrations consisting of the sum allomeric and epimeric forms of chlorophyll-a.

### 2.3. Organic Carbon

Total and dissolved organic carbon (TOC and DOC) are provided in a single data file, *org_c*. Organic carbon samples are collected as for the CDOM samples above. DOC samples are filtered in the same manner as CDOM, and the filtrate acidified with 0.5 mL of 50% H₂SO₄ solution and stored in acid washed glass containers wrapped in aluminium foil at 4°C for transport to the laboratory. Unfiltered TOC samples are treated identically to DOC samples. TOC and DOC are measured using a Shimadzu Total Organic Carbon Analyser (TOC-VCHS/CSN + TNM-1). Samples are acidified and purged with CO₂-free air, then combusted at 720°C and measured by a nondispersive infrared detector. *Table 5* provides column definitions for the organic carbon datafile.
Table 5
Data description of organic carbon dataset.

| Column name     | unit   | description                                      |
|-----------------|--------|--------------------------------------------------|
| observation_id  | ID     | Unique identifier for each measurement set       |
| DOC             | mg/L   | Dissolved organic carbon in mg/L                 |
| TOC             | mg/L   | Total organic carbon in mg/L                     |

Table 6
Data description of TSS dataset.

| Column name      | unit   | description                                                      |
|------------------|--------|------------------------------------------------------------------|
| observation_id   | ID     | Unique identifier for each measurement set                       |
| tss              | mg/L   | Average total suspended solids from three replicates              |
| tss_inorganic    | mg/L   | Average inorganic suspended solids from three replicates         |
| tss_organic      | mg/L   | Average organic suspended solids from three replicates            |
| tss_std          | mg/L   | Standard deviation of total suspended solids from three replicates|
| tss_inorganic_std| mg/L   | Standard deviation of inorganic suspended solids from three replicates|
| tss_organic_std  | mg/L   | Standard deviation of organic suspended solids from three replicates|

2.4. Total Suspended Solids

Total Suspended Solids (also referred to as Total Suspended Matter) (TSS) data are provided in a single datafile, tss (Fig. 4). TSS samples were collected from the upper 10 to 20 cm of the waterbody in 5 L high-density polyethylene containers. Triplicate TSS samples were subsamples from the 5 L container. A known volume of each subsample was filtered through a pre-ashed at 450°C and pre-weighed filter by vacuum filtration using Glass fibre filters (0.7 μm) prepared after Tilstone, et al. [10]. Following filtration, the filters were stored in the cool and dark while being transported to the laboratory for analysis. The filters were dried at 75°C with an initial drying period of 24 h. TSS is taken as the mass difference between the original filter weight minus the final weight divided by the volume of sample used. The contribution of organic and inorganic material to TSS was determined by the mass difference in TSS following combustion at 450°C for 3 h. TSS data are reported as averages and standard deviations made from triplicate measurements. Column definitions for the TSS datafile are given in Table 6.

2.5. Radiometry

Radiometry data are provided in the radiometry and Kd data files. The radiometry data file provides measurements of the remote sensing reflectance ($R_s$) (Eq. (3)) evaluated just above the water air interface where $E_d^{0+}($λ) is the spectral planar downwelling irradiance at the water surface and $L_w(λ,\theta,\phi)$ is the water leaving radianice at wavelength $\lambda$, at nadir viewing angle $\theta$ and azimuth angle $\phi$ just above the water surface as defined in Ruddick et al. [11]. For clarity the spectral component of radiometric quantities ($\lambda$) is omitted for equations following Eq. (3).

The radiometry data file identifies two methods of data collection, ‘interface’ and ‘kutser’. Each method is described separately below. All radiometric measurements are obtained using three Trios RAMSES sensors — a RAMSES ACC-2 VIS planar irradiance sensor ($E_s$), a RAMSES ACC-2 diffuse irradiance sensor ($E_d$) and a RAMSES ARC VIS radiance sensor.

Interface measurements follow the protocol for the ‘single-depth approach’ described in Zibordi and Talone [12].

Radiometers are positioned according to Fig. 6 to minimise shading from the instrument and vessel. Radiometric measurements are conducted in three phases shown in Fig. 7. The phases include above surface measurements (Fig. 7A), an interface stage (Fig. 7B) and a profiling stage (Fig. 7C). Radiometric measurements are made over a period of 20–30 minutes, during which time the light field is subject to change. To ensure all data are comparable, data from each mea-
measurements are normalised to the deck $E_s$ sensor \[ \text{as shown in Eq. (4), where } m_i \text{ is the measurement made at time } i \text{ with } E_s(t^i) \text{ and } E_s(t^0) \text{ as the deck } E_s \text{ measurements at time } i \text{ and time } = 0 \text{ respectively.}
\]

$$R_{rs}(\lambda, \theta, \phi) = \frac{L_w(\lambda, \theta, \phi)}{E_s^0(\lambda)} \quad \text{(3)}$$

$$m_{\text{norm}}(t^i) = m_i(t^i) \cdot \frac{E_s(t^0)}{E_s(t^i)} \quad \text{(4)}$$

The above surface phase (Fig. 7A) involved radiance and irradiance measurements made at approximately 40 cm above the water surface.

The interface phase (Fig. 7B) measures the upwelling radiance ($L_u(z)$) 5–10 cm below the water surface and calculates $R_{rs}$ according to the single depth approach (SDA) described in \[ \text{[12]. } L_u(z) \text{ is corrected for vertical attenuation between the measurement depth (} z_1 \text{) and the water/air interface (} L_u(0, z) \text{) according to Eq. (5). } L_u(z_1, t_1) \text{ is the radiance measurement made at depth } z_1 \text{ and at time } t_1 \text{ shown in Fig. 7B, with } K_{lu} \text{ being derived from the profiling phase (} \text{Fig. 7C). } K_{lu} \text{ is determined by examining the vertical attenuation of radiance measurements over the depth range } z_1-z_2 \text{ measured during the profiling phase (Fig. 7C). Typically, 5–10 measurements of the upwelling radiance are made during the profiling stage. } L_u(z_2, t_2) \text{ is selected from these measurements in order to minimise instrument noise but maximise the depth over which } K_{lu} \text{ is calculated. Typically } L_u(z_2, t_2) \text{ is taken as the second deepest measurement in the profile.}

$L_u(0)$ is then converted to the water leaving radiance ($L_w 0+$) by accounting for the spectral transmittance across the water-air interface using Eq. (7) \[ \text{[11] where } T_F \text{ is the Fresnel transmittance of radiance from water to air and } n_w \text{ is the refractive index of water and } L_u(0, z) \text{ is the upwelling irradiance just below the water surface.}

$$L_u(0, z) = L_u(z_1, t_1) \exp(K_{lu} \cdot z_1) \quad \text{(5)}$$

$$K_{lu} = \frac{1}{z_2 - z_1} \ln \left[ \frac{L_u(z_1, t_1)}{L_u(z_2, t_2)} \right] \quad \text{(6)}$$
Fig. 7. Radiometric measurement stages — (A) above surface, (B) interface and (C) profile. At the above surface stage, $L_T$ is the total radiance reaching the sensor, this quantity is composed of reflected light ($L_{\text{ref}}$) and the water leaving radiance ($L_w$). $L_w$ consists of the upward radiance ($L_u$) transmitted through the water surface.
\[ L_w(\lambda) = \frac{T_r}{n_w^2} L_d(0^-, \lambda) \]  

For flat seawater a spectrally independent value of 0.543 [14] for \( \frac{T_r}{n_w^2} \) is regularly used in the literature. In the current dataset the spectrally dependant transmittance is calculated after the procedure described in [15].

Following correction for transmittance across the water-air interface, \( R_{rs} \) is then calculated as in Eq. (3). To avoid interference from the vessel, the \( E_d \) obtained during above surface phase (Fig. 7A) is used in this calculation. This is the quantity reported with the ‘interface’ flags in the radiometry datafile.

At several locations the single-depth approach described above were not able to be made due to the contaminated nature of the waterbody or strong vertical attenuation generating noise in the radiance sensor at shallow depths (<0.5 m). At these sites the protocol described by Kutser et al. [16] was adopted. This method uses the measurement geometry shown in Fig. 7A. Kutser measurements are obtained by fitting a power law function to the ratio of \( L_f/E_d \) over the spectral range 350–380 and 890–900 nm. The function is then applied to the full spectral range and subtracted from \( L_f/E_d \) to give an estimate \( R_{rs} \) referred to as \( R_{rs(k)} \). This quantity is reported in the radiometry datafile identified by the ‘kutser’ flag in the measurement column.

\( K_d \) measurements are obtained by sequentially lowering the irradiance (\( E_d \)) sensor as shown in Fig. 7C. Measurements of downwelling irradiance \( E_d(z) \) are made at depths \( (z_i) \) to obtain the spectral diffuse attenuation coefficient \( (K_d) \). Irradiance measurements \( (E_d) \) are fit to Eq. (8), where \( E_d(0^-, \lambda) \) is the irradiance at wavelength \( \lambda \) measured just below the water surface and \( z \) is depth in meters.

\[ E_d(z, \lambda) = E_d(0^-, \lambda) \cdot \exp\left(-\int_0^z k_d(z, \lambda) \cdot dz\right) \]  

(8)

The number and sequence of depth intervals obtained were determined based on the rate that the optical signal diminished with depth and depth of the waterbody. Surface waves are known to introduce optical shading and amplification effects in \( E_d \) and to a lesser extent in \( L_d \) profiles [17]. To reduce this effect 10–20 measurements were made at each depth interval, from which a trimmed average was taken using the middle 50% of measurements. The middle 50% of measurements were determined through numerical integration of the spectrum to a zero baseline.

Radiometric measurements are provided in two data files described in Tables 7 and 8 below.

### 2.6. Backscattering

Backscattering data were collected using a Seabird ECO BB9 backscatter meter [18] and in some cases an ECO triplet (bb2) [19]. The data are provided in the \( bb \_surface \) data file. The bb9 and bb2 provide data of the total volume scattering coefficient \( (\beta_T(124^\circ, \lambda)) \) where \( \theta \) is the receiving angle of the instrument \( (124^\circ) \) and \( \lambda \) is the wavelength. Wavelengths available for the

**Table 7**

Data description for radiometry dataset.

| Column Name   | Unit    | Description                                      |
|---------------|---------|--------------------------------------------------|
| observation_id| ID      | Unique identifier for each measurement set.      |
| datetime_utc  | Date-time | Date-time for start of measurement. Format yyyy-mm-dd hh:mm:ssf in universal time coordinated time scale. |
| method        | categorical | Confirms interface or kutser methods. |
| Rs            | sr\(^{-1}\) | Remote sensing reflectance (Eq. (3)) |
| wls           | nm      | Wavelengths radiometric measurement. |


BB2 are 470, 532 nm while the BB9 measures are 412, 440, 488, 510, 532, 595, 650, 676, 715 nm.

Three methods designated, bucket, profile, and dilution, were used to obtain backscattering data. The bucket method involved sampling approximately 9L of water from the surface water. The water sample was then transfer to an opaque black vessel that was filled to enable the BB9 sensors to be submerged. Measurements were made over a two-minute period from which an average was taken for further processing. The vessel was covered during measurement to prevent the incursion of ambient light.

The profiling method involved attaching the BB9 to a metal cage suspended from a winch. A pressure sensor was used to determine depth as the cage was lowered through the water column. Back scattering measurements from the upper 1.5 m were averaged for use in further processing.

The BB9 was developed for use in oceanic and coastal waters and is subject to saturation in highly scattering inland waters. To provide comprehensive coverage of Australian bio-optical conditions the authors used a serial dilution technique to estimate the backscattering properties of highly turbid waterbodies. This technique involved obtaining ~20 L of sample water obtained from the first 50 cm of surface waters. Seven litres of water were transferred to an opaque black bucket vessel and the sample was then progressively diluted using 18 MΩ MilliQ water until all wavelengths were unsaturated. The sample water was then gently stirred to ensure particles remained suspended and a two-minute measurement made. This process was repeated 4–5 times to obtain measurements over a wide concentration range. Linear regression was then used to estimate the particulate backscattering at full concentration. Further details of this method and validation analysis are being prepared as part of a forthcoming publication. All observations are tagged to enable users to filter or select data based the measurement method and their requirements.

\[ \beta_T(124^0, \lambda) \] are corrected for absorption using Eq. (9), where \( L_p \) is the optical pathlength in meters and \( a_T \) is the total absorption (including water). The manufacturer recommends using 0.0391m for \( L_p \), however Monte Carlo simulations in highly absorbing waters have indicated a shorter path length of 0.01635 is a more appropriate value [20]. This value has been adopted for this dataset. All uncorrected values are provided in the dataset for the convenience of users. Total absorption (\( a_T \)) was calculated as the sum of CDOM, phytoplankton and NAP absorption from the absorption_fitted datafile. Absorption of water was obtained from Buiteveld et al. [21].

\( \beta_{\text{corrected}}(124^0, \lambda) \) coefficients are converted to particulate volume scattering coefficients (\( \beta_p(124^0, \lambda) \)) by subtracting the volume scattering function for water (\( \beta_{\text{water}}(124^0, \lambda) \)) after Eq. (10), where \( \lambda \) is the wavelength in nanometres and PSU is the salinity in practical salinity units.

The particulate backscattering coefficients (\( b_{bp}(\lambda) \)) are then estimated using Eq. (11).

\[
\beta_{\text{corrected}}(124^0, \lambda) = \beta_{\text{measured}}(124^0, \lambda) \cdot \exp(L_p \cdot a_T)
\]

\[
\beta_{\text{water}}(124^0, \lambda) = 1.38 \cdot \left( \frac{\lambda}{500} \right)^{-4.32} \cdot \left( 1 + 0.3 \cdot \frac{\text{PSU}}{37} \right) \cdot 10^{-4} \cdot \left( \frac{1 + \cos^2(124 \cdot 1-0.09)}{1-0.09} \right)
\]

\[
b_{bp}(\lambda) = 2\pi \cdot 1.1 \cdot \beta_p(124^0, \lambda)
\]
Table 9
Data description for the bb_surface datafile.

| Column Name | Unit | Description |
|-------------|------|-------------|
| observation_id | ID | Unique identifier for each measurement set |
| method | categorical | Method used, one of: |
| | | • surface_bb2 - measurement made at surface using bb2 |
| | | • dilution - obtained using serial dilutions using bb9 |
| | | • bucket - measurement made in bucket using surface water sample |
| | | • bb9_in_situ - obtained from in situ measurement using bb9 |
| beta_raw | m⁻¹ sr⁻¹ | Volume scattering function at 124 degrees at bbwls wavelengths. |
| beta_cor | m⁻¹ sr⁻¹ | Absorption corrected volume scattering function at 124 degrees at bbwls wavelengths. |
| bbp | m⁻¹ | Particulate backscattering coefficients at bbwls wavelengths. |
| bbwls | nm | Wavelengths of the backscattering meter in nanometres. |
| gamma_bbp | - | Particulate backscattering spectral slope obtained from the 7 wavelengths that best fit Eq. (12) |
| bbp_555 | m⁻¹ | Particulate backscattering at 555 nm |

The particulate backscattering spectral slope ($\gamma_{bbp}$) and backscattering at 555 nm (bbp_555) are calculated by fitting the bbp data to Eq. (12) using least squares regression on the natural log transformed data. Blue wavelengths are known to periodically deviate from the relationship described in Eq. (12). To account for this all combinations of seven of the nine wavelengths are fit to Eq. (12) with the combination that produces the smallest residuals being retained. Column definitions for the bb_surface datafile are shown in Table 9.

$$bbp(\lambda) = bbp_{555}\left(\frac{555}{\lambda}\right)^\gamma$$  \hspace{1cm} (12)

Ethics Statements

This work did not include human subjects, animal experimentation or data collection from social media platforms.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data Availability

Bio-optical data for Australian Inland Waters v.1 (Original data) (CSIRO Data Access Portal).

CRediT Author Statement

Nathan Drayson: Writing – original draft, Formal analysis, Investigation, Data curation, Software, Visualization; Janet Anstee: Conceptualization, Methodology, Project administration, Funding acquisition, Investigation, Supervision; Hannelie Botha: Conceptualization, Methodology, Investigation, Supervision; Gemma Kerrisk: Investigation, Resources; Phillip Ford: Investigation, Supervision; Bozena Wojtasiewicz: Investigation, Formal analysis, Methodology; Lesley Clementson: Investigation, Formal analysis, Methodology; James McLaughlin: Investigation.
Acknowledgments

The authors wish to acknowledge and thank the many CSIRO staff, past and present, involved in the acquisition of this data that underpins our science. Erin Hestir, Tim Malthus and Guy Byrne are acknowledged for their contributions to field data collection and planning. The DOC and TOC analyses were undertaken by John Gouzos and Claire Wright CSIRO L&W, Waite Campus. Mark Wilson (O&A, IOMRC Crawley) assisted with the collection of bio-optical data in the Perth area.

The authors would also like to pay respect to the Traditional Owners and their Nations of the lands on which we collected the data contained in this report. We acknowledge their deep cultural, social, environmental, spiritual and economic connection to their lands and waters. We would especially like acknowledge the generosity of the Barkindji-Maraura Elders Council for allowing us to access to Lake Victoria. We would also like to thank Clay Smith (SA Water) for his assistance with navigating and access to Lake Victoria.

We also gratefully acknowledge the contributions of Nick Crosbie (Melbourne Water) for assistance with navigating and accessing Western Wastewater Treatment Plant site and laboratories.

The data presented in this publication has been collected as part of many projects undertaken between 2013 and 2021.

The project IWQual has received funding from Geoscience Australia and CSIRO for the period July 2019 - March 2022 and we acknowledge the support of our Geoscience Australia colleagues, in particular, Dr Stephen Sagar, Assistant Director of Aquatic Product Development.

References

[1] L.A. Clementson, et al., The CSIRO method, in: S. Stanford, B. Hooker, et al., (Eds.), The Fifth SeaWiFS HPLC Analysis Round-Robin Experiment (SeaHARRE-5), Goddard Space Flight Center, Maryland: NASA, 2012.

[2] N. Pahlevan, B. Smith, J. Schalles, C. Binding, C. Cao, R. Ma, K. Alikas, K. Kangro, D. Gurlin, N. Ha, B. Matsushita, W. Moses, S. Greb, M. Lehmann, M. Ondrusek, N. Oppelt, R. Strumpf, Seamless retrievals of chlorophyll-a from Sentinel-2 (MSI) and Sentinel-3 (OLCI) in inland and coastal waters: a machine-learning approach, Remote Sens. Environ. 240 (2020) 111604.

[3] J. Kravitz, M. Matthews, L. Lain, S. Fawcett, S. Bernard, Potential for high fidelity global mapping of common inland water quality products at high spatial and temporal resolutions based on a synthetic data and machine learning approach. Front. Environ. Sci. 9 (2021) 587660.

[4] E. Botha, J. Anstee, S. Sagar, E. Lehmann, T. Medeiros, Classification of Australian Waterbodies across a wide range of optical water types, Remote Sens. 12 (2020) 3018–3042, doi: 10.3390/rs12183018.

[5] V.E. Brando, A.G. Dekker, Y.J. Park, T. Schroeder, Adaptive semi-analytical inversion of ocean color radiometry in optically complex waters, Appl. Opt. 51 (2012) 2808–2833.

[6] J. Anstee, N. Drayson, H. Botha, G. Kerrisk, S. Stephen, F. Phillip, B. Wojtasiewicz, L. Clementson, and G. Byrne. Bio-optical data for Australian inland waters v1, doi:10.25591/rtf7-j815.

[7] L.A. Clementson, J.S. Parslow, A.R. Turnbull, D.C. McKenzie, C.E. Rathbone, Optical properties of water in the Australian sector of the Southern Ocean, J. Geophys. Res. 106 (31) (2001) 611–631.

[8] M. Kishino, M. Takahashi, N. Okami, S. Ichimura, Estimation of the spectral absorption coefficients of phytoplankton in the sea, Bull. Mar. Sci. 37 (2) (1985) 634–642.

[9] B. Mitchel, Algorithms for determining the absorption coefficient for aquatic particulates using the quantitative filter technique, in: Proceeedings of SPIE, Ocean Optics X, 1990, pp. 137–148. vol.

[10] G.H. Tilstone, G.F. Moore, K. Sørensen, R. Doerrfer, R. Røttergs, K.G. Ruddick, R. Pasterkamp, P.V. Jørgensen, Protocols for the validation of MERIS products in Case 2 waters, in: Proceedings of the ENVISAT MAVT Conference, Frascati, Italy, 2003 20–24 October 2003.

[11] K. Ruddick, K. Voss, E. Boss, A. Castagna, R. Frouin, A. Gilerson, M. Hieronymi, B. Johnson, J. Kuusk, Z. Lee, M. Ondrusek, V. Vabson, R. Vendt, A review of protocols for fiducial reference measurements of water-leaving radiance for validation of satellite remote-sensing data over water, Remote Sens. 11 (2019) 2198–2236.

[12] C. Zbindi, M. Talone, On the equivalence of near-surface methods to determine the water-leaving radiance, Opt. Express 28 (3) (2020) 3200–3214.

[13] K. Ruddick, K. Voss, A. Banks, E. Boss, A. Castagna, R. Frouin, M. Hieronymi, C. Jamet, B. Johnson, J. Kuusk, Z. Lee, M. Ondrusek, V. Vabson, R. Vendt, A review of protocols for fiducial reference measurements of downwelling irradiance for the validation of satellite remote sensing data over water, Remote Sens. 11 (2019) 1742–1780.

[14] R. Austin, G. Halikas, The Index of Refraction of Seawater, Scripps Instition of Oceanography, UC San Diego, 1976 Technical Report.

[15] K. Voss, S. Flora, Spectral dependence of the seawater-air radiation transmission coefficient, J. Atmos. Ocean. Technol. 34 (2017) 1203–1205.
[16] T. Kutser, E. Vahtmae, B. Paavel, T. Kauer, Removing glint effects from field radiometry data measured in optically complex coastal and inland waters, Remote Sens. Environ. 133 (2013) 85–89.

[17] J. Mueller, Mueller, G. Fargionand, C. McClain (Eds.), In-water radiometric profile measurements and data analysis protocols, Ocean Optics Protocols for Satellite Ocean Color Sensor Validation, Revision 4, volume III: Radiometric Measurements and Data Analysis Protocols III (2003) 7–20 ch. 2.

[18] Sea Bird Scientific. “ECO Scattering Sensor.” https://www.seabird.com/scattering-sensors/eco-scattering-sensor/family-downloads?productCategoryId=54627869916 (accessed 01-04-22).

[19] Sea Bird Scientific. “ECO Triplet.” https://www.seabird.com/combination-sensors/eco-triplet/family?productCategoryId=54758054351 (accessed 01-04-22).

[20] D. Doxaran, E. Leymarie, B. Nechad, A. Dogliotti, K. Ruddick, P. Gernez, E. Knaeps, Improved correction methods for field measurements of particulate light backscattering in turbid waters, Opt. Express 24 (4) (2016) 3615–3637, doi:10.1364/OE.24.003615.

[21] H. Buiteveld, J. Hakvoort, M. Donze, Optical properties of pure water, in: Proceedings of the SPIE - The International Society for Optical Engineering Ocean Optics XII, 2258, 1994, pp. 174–183.