Earthworm (*Pheretima aspergillum*) extract stimulates osteoblast activity and inhibits osteoclast differentiation

Yuan-Tsung Fu1,2†, Kuo-Yu Chen3†, Yueh-Sheng Chen1,4 and Chun-Hsu Yao1,4,5*

Abstract

**Background:** The potential benefits of earthworm (*Pheretima aspergillum*) for healing have received considerable attention recently. Osteoblast and osteoclast activities are very important in bone remodeling, which is crucial to repair bone injuries. This study investigated the effects of earthworm extract on bone cell activities.

**Methods:** Osteoblast-like MG-63 cells and RAW 264.7 macrophage cells were used for identifying the cellular effects of different concentrations of earthworm extract on osteoblasts and osteoclasts, respectively. The optimal concentration of earthworm extract was determined by mitochondrial colorimetric assay, alkaline phosphatase activity, matrix calcium deposition, Western blotting and tartrate-resistant acid phosphatase activity.

**Results:** Earthworm extract had a dose-dependent effect on bone cell activities. The most effective concentration of earthworm extract was 3 mg/ml, significantly increasing osteoblast proliferation and differentiation, matrix calcium deposition and the expression levels of alkaline phosphatase, osteopontin and osteocalcin. Conversely, 3 mg/ml earthworm extract significantly reduced the tartrate-resistant acid phosphatase activity of osteoclasts without altering cell viability.

**Conclusions:** Earthworm extract has beneficial effects on bone cell cultures, indicating that earthworm extract is a potential agent for use in bone regeneration.

**Keywords:** Earthworm, Osteoblasts, Osteoclasts
osteoclast activities via in vitro cell culture. To the best of our knowledge, this study is the first to evaluate the effects of earthworm extract on the bone cells activities. Bone cell activities were assessed using the tetrazolium bromide colorimetric assay, alkaline phosphatase (ALP) activity assay, matrix calcium deposition, Western blotting assay and the tartrate-resistant acid phosphatase (TRAP) activity assay.

Methods
Preparation of earthworm extract
Dried earthworms were obtained from the Chung Song Zong Pharmaceutical Co. (Kaohsiung, Taiwan). Their identity was confirmed by experts in pharmacognosy. Earthworm extract was prepared using a method described previously [18]. Briefly, 600 g dried earthworm was added to 7.2 l deionized water and boiled at 100°C for 1 h. The residue was volatilized at room temperature. Another extract was obtained from the pretreated earthworm by boiling deionized water for two times [water-to-earthworm ratios (ml/g) at 10:1 and 8:1] for 1 h. The aqueous extracts were filtered to remove insoluble debris and concentrated at 65°C via vacuum evaporation to 2.3 g/ml (equivalent to the dry weight of earthworms) and stored at 4°C until in vitro assays.

Osteoblast culture
The cellular effects of earthworm extract on osteoblasts were assessed using the human osteoblast-like cell line MG-63 (BCRC no. 60279, Food Industry Research and Development Institute, Hsinchu, Taiwan). Cells were cultured in a growth medium consisting of Dulbecco’s modified Eagle’s medium (DMEM; Gibco, Grand Island, NY, USA), 10% fetal bovine serum (FBS; Gibco) and 1% penicillin/streptomycin (Gibco) at 37°C under 5% CO₂ in humidified air. The medium was changed every 2 days. Adherent cells were allowed to reach roughly 80% confluence. The cells were then passaged in the culture. Passage 2 culture was used for all in vitro assays. Cultured MG-63 cells were seeded in growth medium in 96-well plates at a density of 1 × 10⁵ cells/well. After 1 day of incubation, the culture medium was replaced with mixed solutions of new culture medium and various concentrations of earthworm extract at a ratio of 9:1 (v/v). In the control group, the culture medium was replaced with PBS at a ratio of 9:1 (v/v) for cell cultures. After 2 days of culture, osteoclast proliferation and differentiation were determined by MTT assay and TRAP activity assay, respectively.

RAW 264.7 culture
The effects of earthworm extract on osteoclast proliferation and differentiation were evaluated using murine monocyte/macrophage RAW 264.7 cells (BCRC No. 60001, Food Industry Research and Development Institute). 2 × 10⁵ cells/well RAW 264.7 cells were seeded in individual wells of a 48-well tissue culture plate in a growth medium. After 1 day of incubation, osteoclast differentiation from RAW 264.7 cells was induced with 50 ng/ml RANKL (Enzo Life Sciences, Lausen, Switzerland) in α-minimal essential medium (α-MEM; Gibco) with 10% PBS for 6 days because the formation of mature osteoclasts requires 5–6 days [21-23]. The culture medium was refreshed every 2 days. After 6 days culturing, the culture medium was replaced with mixed solutions of new culture medium and various concentrations of earthworm extract at a ratio of 9:1 (v/v). In the control group, the culture medium was mixed with PBS at a ratio of 9:1 (v/v) for cell cultures. After 2 days of culture, osteoclast proliferation and differentiation were determined by MTT assay and TRAP activity assay, respectively.

MTT assay for cell proliferation
After incubation for 2 days, the supernatant was removed. Then, 10 μl MTT solution (5 mg/ml) and 100 μl culture medium were added to each well. The plate was incubated at 37°C for 4 h to facilitate the formation of insoluble formazan crystals. The solution was then removed and 100 μl/well acidic isopropyl alcohol (0.04 M HCl in isopropyl alcohol) was added to dissolve the dark-blue formazan crystals. After shaking for a few minutes, the concentration of intracellular formazan crystals was determined at a test wavelength of 570 nm against a reference wavelength of 650 nm by an enzyme-linked immunosorbent assay (ELISA) reader (uQuant; Bio-Tek Instruments, Inc., Sunnyvale, CA, USA). The number of viable cells in each well was calculated by transforming the optical density values of the MTT assay into numbers of cells/well, based on a standard curve. All experiments were performed in triplicate.

Liu’s stain for morphological observation and osteoblast proliferation
Liu’s stain was applied to observe osteoblast morphology and proliferation [24]. After culturing for 2 days, the cell layers were rinsed three times with PBS, fixed in 2% glutaraldehyde (Acros, Geel, Belgium) for 30 min, and stained with Liu’s stain solution (30 s in solution A, followed by 90 s in solution B) (Chin Pao Co., Ltd., Taipei, Taiwan). After washing three times with deionized water to remove the remaining stain, the cell layers were observed using an inverted optical microscope (Axiovert 25; Carl Zeiss, Inc., Goettingen, Germany).
Analysis of alkaline phosphatase for osteoblast differentiation
After 2 days of culturing, the medium was replaced with 20 μl/well 0.1% Triton X-100 (Sigma) and incubated at room temperature for 5 min for cell lysis. 100 μl/well of the commercially available ALP assay kit (Procedure No. DG1245-K; Sigma) was then added within 1 min. Absorbance at 405 nm caused by p-nitrophenol production was assessed for 30 min at room temperature. The change in rate of absorbance was directly proportional to ALP activity. Each experimental condition was repeated three times.

Analysis of matrix calcium deposition
Von Kossa stain was utilized to examine the formation of the mineralized matrix [25]. Briefly, 5 × 10⁴ cells/well cultured MG-63 cells were added to the osteogenic medium (growth medium supplemented with 50 μg/ml L-ascorbic acid (Sigma), 10 mM β-glycerol phosphate (Sigma) and 10 nM dexamethasone (Sigma) in a 35-mm culture dish. The medium was mixed with various concentrations of earthworm extract at a ratio of 9:1 (v/v). In the control group, the medium was mixed with PBS at a ratio of 9:1 (v/v) for cell cultures. The medium was changed every 3 days. After culturing for 14 days, cultures were washed with PBS twice and fixed in 2% glutaraldehyde for 20 min. After incubation for 30 min with 5% silver nitrate (Union Chemical Works, Ltd., Hsinchu, Taiwan) in darkness at room temperature, cells were rinsed twice in deionized water. After drying in air, cells were exposed to ultraviolet light for 1 h until color development was complete. The cells were then immersed in 5% sodium thiosulfate (Union Chemical Works, Ltd.) for 2 min. Finally, calcification was visualized by counterstaining cells with 0.1% nuclear fast red (Sigma) dissolved in 5% aluminum sulfate (JT Baker, Phillipsburg, NJ, USA) for 5 min. Following rinsing in deionized water, matrix calcium deposition was observed with an optical microscope.

Western blot analysis
Western blot analysis was performed to study the expression of ALP, osteopontin and osteocalcin of osteoblasts while interacting with earthworm extract. Briefly, 4 × 10⁵ cells/well cultured MG-63 cells were seeded to blasts while interacting with earthworm extract. Briefly, expression of ALP, osteopontin and osteocalcin of osteoblasts while interacting with earthworm extract. Briefly, expression of ALP, osteopontin and osteocalcin of osteoblasts while interacting with earthworm extract.

Analysis of TRAP for osteoclast differentiation
After 2 days of culturing, TRAP activity was assessed by measuring the amount of acid phosphatase (ACP) released from cells into the medium using a commercially available kit (Procedure No. 435, Sigma). Briefly, 30 μl culture media was mixed with 100 μl acid phosphatase reagent. Absorbance at 405 nm caused by p-nitrophenol production was observed for 30 min at room temperature. The change in rate of absorbance was directly proportional to TRAP activity. Each experimental condition was repeated three times.

Statistical analysis
Numerical data are presented as mean ± SD. Statistical analysis was performed using one-way analysis of variance (ANOVA), followed by post hoc Fisher’s LSD multiple comparison test. Statistical significance was p < 0.05.

Results
Earthworm extract enhances osteoblast proliferation
Figure 1 shows the effects of different concentrations of earthworm extract on osteoblast proliferation measured by MTT assay after 2 day of culture. Osteoblast proliferation was dose-dependent for high concentrations of earthworm extract. Treatment with earthworm extract at concentrations from 500 μg/ml to 6 mg/ml significantly enhanced osteoblast proliferation (p < 0.05). Particularly, 3 mg/ml earthworm extract increased cell proliferation most (p < 0.01), in which the number of osteoblasts was 54% higher than that in the control group. However, the number of osteoblasts decreased significantly when the earthworm extract concentration was > 6 mg/ml (p < 0.05). Notably, 12 mg/ml earthworm extract significantly inhibited osteoblast proliferation (p < 0.001), in which the number of osteoblasts was 43% lower than that in the control group. It indicated that 12 mg/ml earthworm extract was likely toxic to osteoblasts. These analytical results were further confirmed by staining results with Liu’s stain.
Figure 2 shows the morphology of osteoblasts cultured with various concentrations of earthworm extract for 2 days. The osteoblasts displayed spindle-shaped. The morphology of osteoblasts did not obviously differ between the control group and experimental groups. However, the number of osteoblasts increased as the concentration of earthworm extract increased from 0.5 μg/ml to 3 mg/ml, and then decreased as the concentration was raised further. Particularly, treatment with 12 mg/ml earthworm extract reduced the osteoblast population as compared with that of the control group. Similar findings were obtained by MTT analysis.

**Earthworm extract stimulates osteoblast differentiation**

ALP, a membrane-bound enzyme, is a differentiation marker of early osteoblasts. Differentiated osteoblasts exhibit elevated ALP activity, which correlates with high levels of enzyme expression [26]. Treatment of MG-63 cells with earthworm extract for 2 days stimulated the ALP activity of osteoblasts in a dose-dependent manner (Figure 3). Compared with the control group, the ALP activity was significantly increased when the concentration of earthworm extract was between 500 ng/ml and 3 mg/ml ($p < 0.05$). Notably, at higher concentrations (1 and 3 mg/ml), ALP activity increased significantly by about 1.4-fold ($p < 0.001$). However, at 6 mg/ml, ALP activity of osteoblasts did not differ significantly from that of the control group ($p > 0.05$). Moreover, at 12 mg/ml, ALP activity was significantly lower than that of the control group ($p < 0.05$).

**Earthworm extract induces matrix calcium deposition**

Von Kossa staining is specific for the calcified extracellular matrix [27]. Dark staining of nodules demonstrates deposition of the mineralized matrix of bone formation. Figure 4 shows the effect of various concentrations of earthworm extract on matrix calcium deposition after 14 days of incubation. Osteoblasts cultured in medium containing 500 μg/ml to 3 mg/ml earthworm extract had high matrix mineralization. However, mineralization of osteoblasts treated with 12 mg/ml earthworm extract was not observed after culturing for 14 days.

**Earthworm extract stimulates the expression of osteogenic-related proteins**

Western blot analysis was applied to observe the expression of osteogenic-related proteins after treatment with different concentrations of earthworm extract for 7 days. All ALP, osteopontin and osteocalcin expression levels on earthworm extract-treated osteoblasts were higher than those of the control group (Figure 5). Cells cultured with medium containing 500 μg/ml to 6 mg/ml, 5 μg/ml to 6 mg/ml and 500 μg/ml to 6 mg/ml earthworm extract had higher upregulation of ALP, osteopontin and osteocalcin expression, respectively. Particularly, osteoblasts treated with 3 mg/ml earthworm extract had the highest ALP expression.

**Earthworm extract has no effect on the proliferation of osteoclasts**

The RAW 264.7 cells were used to investigate the osteoclastogenic effect of earthworm extract. Figure 6 displays...
Figure 2 The morphology and proliferation of osteoblasts treated with various concentrations of earthworm extract. MG-63 cells were treated with PBS as a control or different concentrations of earthworm extract. After 2 day of culture, the cell layers were stained with Liu's stain solution and observed using an inverted optical microscope. Arrows point to osteoblasts.

Figure 3 Earthworm induces the differentiation of osteoblasts. MG-63 cells were cultured with PBS as a control or different concentrations of earthworm extract. After culturing for 2 days, osteoblast differentiation was assessed by ALP activity assay. Results are expressed as percentage of control (*p < 0.05, **p < 0.01 and ***p < 0.001 vs. control; ****p < 0.001 vs. 3 mg/ml).
the effect of earthworm extract on the proliferation of osteoclasts assessed by MTT assay after treatment with various concentrations of earthworm extract for 2 days. No significant difference in number of osteoclasts existed between the control group and experimental groups ($p > 0.05$), indicating that all test dosages (500 ng/ml to 6 mg/ml) of earthworm extract had no cytotoxic effects on osteoclasts.

Earthworm extract inhibits the differentiation of osteoclasts

The effect of earthworm extract on osteoclast differentiation was evaluated by TRAP, a marker of osteoclast differentiation [28]. Figure 7 shows TRAP activity assay results for osteoclasts cultured with various concentrations of earthworm extract for 2 days. A dose-dependent response for TRAP activity existed. Earthworm extract at concentrations < 3 mg/ml markedly decreased the TRAP activity of osteoclasts as compared with that of the control group ($p < 0.05$). Inhibition of TRAP activity was stronger when the earthworm extract concentration was < 500 µg/ml ($p < 0.05$). However, 6 mg/ml earthworm extract had no significant effect on TRAP activity ($p > 0.05$).

Discussion

Bone remodeling, an important process in the renewal and repair of damaged bone, is initiated by osteoclastic bone resorption and subsequent osteoblastic bone formation. Previous studies have shown that several Chinese medicines significantly affect bone cell activities and
enhance bone regeneration [29-33]. The earthworm has been used as a crude drug for thousands of years in China and the wider Asian region [10]. Earthworm extract can promote the differentiation of neurite-bearing cells and the proliferation and migration ability of RSC96 Schwann cells [18,34,35]. Chen et al. [18] found that earthworm extract markedly enhanced the nerve growth factor-mediated neurite outgrowth from PC 12 cells. Chang et al. [35] reported that treatment with 125 μg/ml earthworm extract for 24 h induced RSC96 cells proliferation. However, toxicity occurred at high concentrations (250–1000 μg/ml).

Moreover, Grdiša et al. [12] found that glycolipoprotein extract markedly enhanced the nerve growth factor-mediated neurite outgrowth from PC 12 cells. Chang et al. [35] reported that treatment with 125 μg/ml earthworm extract for 24 h induced RSC96 cells proliferation. However, toxicity occurred at high concentrations (250–1000 μg/ml).

Figure 6 Earthworm extract does not affect the proliferation of osteoclasts. RAW 264.7 cells were seeded in 48-well plates and allowed to adhere for 1 day. Osteoclast differentiation from RAW 264.7 cells was induced with 50 ng/ml RANKL in α-MEM for 6 days. Osteoclasts were then cultured with PBS as a control or different concentrations of earthworm extract for 2 days. Osteoclast viability was measured by MTT assay.

Figure 7 Earthworm extract inhibits the differentiation of osteoclasts. RAW 264.7 cells were seeded in 48-well plates and allowed to adhere for 1 day. Osteoclast differentiation from RAW 264.7 cells was induced with 50 ng/ml RANKL in α-MEM for 6 days. Osteoclasts were then cultured with PBS as a control or different concentrations of earthworm extract for 2 days. Osteoclast differentiation was determined by TRAP activity assay. Results are expressed as percentage of control (*p < 0.05 and **p < 0.01 vs. control).
extract (G-90) from the earthworm had a protective effect against H₂O₂ toxicity and stimulated the growth of human fibroblasts and epithelial cells. Additionally, Grđiša et al. [15] found that earthworm extract stimulated the synthesis of epidermal growth factor and fibroblast growth factor during wound healing on mice skin. Additionally, lumbrokinase is an important protease derived from earthworms [36]. Recent investigations have found that lumbrokinase has protective effects on hippocampus apoptosis, therapeutic potential in diabetic nephropathy and anti-ischemic action in brain [37-39].

This study evaluated for the first time the effects of earthworm extract on bone cell activities. Earthworm extract significantly promoted the proliferation, differentiation and matrix calcium deposition of osteoblasts. The most effective concentration of earthworm extract in enhancing osteoblasts proliferation and differentiation was 3 mg/ml. However, they were inhibited by high concentration (12 mg/ml), indicating that excessive earthworm extract can have an adverse effect on osteoblasts.

Osteoblasts produce ALP, osteopontin and osteocalcin, which are bone-specific markers [40]. Among these proteins, ALP and osteopontin are early markers of osteoblastic differentiation. Osteocalcin directly participates in the mineralization process and is a marker for mature osteoblasts that is expressed in the late stages of osteogenic differentiation. Moreover, osteopontin and osteocalcin are extracellular matrix proteins and crucial for bone tissue formation. The Western blot analysis results show that 3 mg/ml earthworm extract-treated osteoblasts expressed the highest level of ALP with higher levels of osteopontin and osteocalcin. It indicates that earthworm extract stimulated early differentiation of osteoblasts and is favorable for osteoblast mineralization. Calcium deposition results by von Kossa staining were in agreement with these analytical results. The most effective concentration of earthworm extract for matrix calcium deposition was between 500 µg/ml and 3 mg/ml. These results reveal that the capacity to form matrix mineralization was positively correlated with osteoblast proliferation and differentiation.

The experiments also show that 3 mg/ml earthworm extract did not significantly affect osteoclast proliferation. However, 3 mg/ml earthworm extract reduced the amount of TRAP, indicating that it inhibited osteoclast activity. These analytical results demonstrate that 3 mg/ml earthworm extract can be used for bone repair because it can promote osteoblast proliferation and differentiation as well as inhibit osteoclast activity.

Conclusions

Our findings show that an appropriate dose of earthworm extract has potential effects on bone cell cultures. Therefore, earthworm extract might be a promising agent for bone tissue regeneration. Nevertheless, this research is only preliminary in evaluating the effects of earthworm extract on bone cell activities. Further studies are needed to identify the specific interactions between different earthworm components and bone cells and to determine the exact bioactive compounds in earthworm extract that are responsible for bone cell activities.

Competing interests

The authors declare that there are no competing interests.

Authors’ contributions

YTF and KYC planned and carried out the experiments, analyzed the data and drafted the manuscript. YSC participated in the design of the study and helped in the interpretation of the results. CHY conceived of the study, and participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

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Author details

1 School of Chinese Medicine, China Medical University, 91 Hsueh-Shih Road, Taichung 40402, Taiwan. 2 Department of Chinese Medicine, Taichung Tzu Chi Hospital, The Buddhist Tzu Chi Medical Foundation, Taichung 40427, Taiwan. 3 Department of Chemical and Materials Engineering, National Yunlin University of Science and Technology, Yunlin 64002, Taiwan. 4 Department of Biomedical Imaging and Radiological Science, China Medical University, Taichung 40402, Taiwan. 5 Department of Biomedical Informatics, Asia University, Taichung 41354, Taiwan.

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