Clinical Observation of COVID-19 in a Patient With an Acquired Humoral Deficiency Secondary to Chemotherapeutic Agents

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Abstract

Introduction: The coronavirus disease 2019 (COVID-19) pandemic due to the novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) causes worldwide devastation. We describe the course of a patient with COVID-19 in the setting of an acquired humoral deficiency as a result of chemotherapeutic treatment for rheumatologic conditions.

Case Report: A 49-year-old Caucasian male presented with non-relieving fever, hypoxemia, and persistent diarrhea after seven days following a positive SARS-CoV-2 polymerase chain reaction (PCR) assay. The patient’s past medical history was significant for mixed connective tissue disease, rheumatoid arthritis, and systemic lupus erythematosus treated with methotrexate and rituximab since 2008. He was diagnosed with acquired humoral deficiency in 2017 managed by intravenous immunoglobulin (IVIG) infusion every three weeks. The patient’s course of hospitalization was complicated by acute respiratory distress which necessitated intensive unit care and required up to 20 L/min oxygen supplementation via a humidified high flow generator. He was treated with hydroxychloroquine and azithromycin and received an IVIG transfusion. The patient was discharged to home after forty-two days of hospitalization with oxygen supplementation only during ambulation and a complete resolution of diarrhea.

Discussion: According to current limited data, patients with immunodeficiency have longer length of hospitalization compared to immunocompetent individuals. Our patient demonstrated a form of immunodeficiency as the result of a chemotherapeutic agent, and his clinical course appeared to be more severe.

Conclusion: More studies are necessary to shed light on the immunological response to SARS-CoV-2 and its impact on immunocompromised and immunocompetent and individuals. We describe the course of a patient with COVID-19 in the setting of an acquired humoral deficiency.

Keywords
COVID-19, immunodeficiency, rituximab, SARS-CoV-2, chemotherapy

Introduction

The coronavirus disease 2019 (COVID-19) caused by the novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has created a multifaceted global devastation.1 There is an existing need for information on the clinical course of COVID-19 in patients with immunodeficiency. We describe the course of a patient with COVID-19 in the setting of an acquired humoral deficiency secondary to chemotherapeutic agents.
deficiency as a result of chemotherapeutic treatment for rheumatologic conditions.

Case Report

A 49-year-old Caucasian male presented with a two-week history of sinusitis, nonproductive cough, and sore throat, which progressed to a fever up to 38.9°C unrelieved by acetaminophen. The patient also experienced generalized myalgia, and had a two-day history of decreased appetite, diarrhea and frontal headache. He tested positive for SARS-CoV-2 by polymerase chain reaction (PCR) assay and was quarantined and medically managed at home for seven days prior to hospital admission. He was admitted due to hypoxemia with oxygen saturation in the low 80s.

The patient's past medical history was significant for mixed connective tissue disease, rheumatoid arthritis, and systemic lupus erythematosus. The patients' rheumatological symptoms were controlled by a combination of methotrexate weekly and rituximab every six months since 2008. However, the prolonged course of rituximab resulted in chronic pansinusitis which led to a diagnosis of acquired humoral deficiency. The patient was referred by an otolaryngologist for an immunological evaluation because his chronic pansinusitis did not resolve with septoplasty and bilateral sinus surgery followed by several rounds of clarithromycin. His laboratory results revealed poor humoral function as demonstrated by peripheral lymphopenia with zero absolute CD19 count with an absence of IgA and IgM, but a normal level of IgG (Table 1). The IgG serology showed no titer response following a pneumococcal vaccination. Because rituximab was the key in management of his rheumatological conditions, intravenous immunoglobulin (IVIG) was initiated. Initially, the IVIG treatment and dosage were targeted at 400 mg/kg per dose every 4 weeks, increased to 600 mg/kg per dose every 4 weeks, then eventually to 600 mg/kg per dose every 3 weeks along with prophylactic azithromycin daily to effectively manage his chronic sinus infections. The patient’s last IVIG and last dose of rituximab was seventeen days prior to hospitalization and two days prior to the start of a non-productive cough.

On admission, the patient presented with tachypnea and visible respiratory distress. His pulse oximetry was 96% on 2 L/min of oxygen via nasal cannula. The vital sign measurements were as follows: temperature of 37.6°C, blood pressure of 119/87 mmHg, heart rate at 82 beats per minute, and respiratory rate at 20 breaths per minute. The auscultation of the lungs revealed fine bibasilar inspiratory crackles with symmetric chest expansion. The initial chest X-ray revealed cardiome diastinal silhouette enlargement with increased pulmonary vascularity and mild bibasilar atelectasis. The complete blood count demonstrated leukopenia and lymphopenia, which were both normal in January 2020 (Table 2). At this time, the patient was experimentally given hydroxychloroquine and azithromycin, but the lack of pulmonary infiltrate on the initial chest X-ray did not qualify him for the remdesivir trial.

On hospital day four, the patient was transferred to the intensive care unit due to worsening hypoxemia. The laboratory results in Table 2 demonstrated persistent leukopenia and lymphopenia with a new elevation of IL-6, C reactive protein, D-dimer, ferritin, glucose, alanine aminotransferase, aspartate transaminase, and lactate dehydrogenase levels. On day five, IVIG was given at 600 mg/kg followed by another 600 mg/kg dose two weeks later. Supplemental oxygen via high flow nasal cannula was initiated at 15 L/min and titrated up to 20 L/min by day twelve. A repeat chest X-ray revealed multifocal ground-glass consolidative opacity that was more significant in the left lower lung with worse aeration compared to prior study.

His respiratory condition was complicated by abdominal distention with diarrhea and ileus, necessitating a nasogastric tube placement. An abdominal computed tomography study was completed on day twenty-six.

| Test name                             | Result       |
|---------------------------------------|--------------|
| IgA, serum (mg/dL)                    | <5 L         |
| IgG, serum (mg/dL)                    | 739          |
| IgM, serum (mg/dL)                    | <5 L         |
| IgG subclass 1 (mg/dL)                | 599          |
| IgG subclass 2 (mg/dL)                | 89 L         |
| IgG subclass 3 (mg/dL)                | 34           |
| IgG subclass 4 (mg/dL)                | 13           |
| CD3 absolute (×10^3/L)                | 0.706 L      |
| CD3+CD4+ absolute (×10^3/L)           | 0.540        |
| CD3+CD8+ absolute (×10^3/L)           | 0.174        |
| CD19 absolute (×10^3/L)               | 0 L          |
| CD3 %                                 | 85           |
| CD3+CD4+ %                            | 65 H         |
| CD3+CD8+ %                            | 21           |
| CD4/CD8 ratio                         | 3.10         |
| CD3+CD4–CD8– %                        | 1            |
| CD45 %                                | 100          |
| CD3+CD16+CD56+ %                      | 14           |
| CD3+CD16+CD56+ absolute (×10^3/L)     | 0.116        |
| CD19 %                                | 0 L          |
| Mannose binding lectin                | 967          |
| C3 complement                          | 140          |
| C4 complement                          | 21           |

L = low; H = high.

Note. The boldface values signify abnormal laboratory values (higher or lower than normal reference range).

Table 1. Humoral Panel at Diagnosis of Acquired Humoral Deficiency in 2017.
revealing thickening of the colon. On day twenty-seven, he was discharged from the intensive care unit. Cultures of Histoplasma, Pneumocystis jiroveci, Staphylococcus aureus, Rhinovirus, Metapneumovirus, Adenovirus, and Clostridium difficile were found negative for respiratory and gastrointestinal symptoms; however, SARS-CoV-2 remained positive via PCR assay. A computed tomography of the chest was performed on day thirty-three showing diffuse patchy infiltrates in both lungs, worse in the right upper and middle lobes as shown in Figure 1. The patient was discharged to home after forty-one days of hospitalization with oxygen supplementation only with ambulation and a complete resolution of diarrhea despite still being tested positive for SARS-CoV-2. The patient eventually was cleared of SARS-CoV-2 after three different PCR assays at three weeks post hospital discharge.

Discussion

Individuals diagnosed with primary or acquired immunodeficiencies are at high risk for fatal outcomes with COVID-19. Guan et al. found that the mean hospitalization stay for an immunocompetent patient was 12.8 days compared to the forty-two-day length of stay for our patient. COVID-19 pneumonia was described in previously reported cases with primary immunodeficient diseases showing similar respiratory decline but followed a shorter course of hospitalization (14–25 days) when compared to our patient’s length of stay. According to current data, patients with immunodeficiency have longer lengths of hospitalization compared to immunocompetent individuals.

The “cytokine release syndrome” has been shown to be associate with severe COVID-19 conditions. In the setting of lymphocytopenia observed in severe COVID-19, the cytokine storm release has been suggested to be facilitated by leukocytes other than T lymphocytes. The possible role of B lymphocytes in the cytokine storm in COVID-19 was discussed by Quinti et al., evidenced by a milder presentation of COVID-19 in patients with agammaglobulinemia when compared to a more severe course in patients with common variable immunodeficiency.

| Timeline                | 3 months prior to COVID-19 | At hospital admission | In ICU | At discharge |
|-------------------------|----------------------------|-----------------------|--------|--------------|
| WBC (cells/L)           | 4.5 x 10^9                 | 3.2 x 10^9 L          | 3.2 x 10^9 L | 4.1 x 10^9 L |
| Absolute lymphocytes (cells/µL) | 1157                      | 510 L                 | 330 L       | 1550         |
| C-reactive protein (mg/L) | 2.5 H                     | 5.5 H                 | 10.83 H    | 3.34 H       |
| Aspartate transaminase, serum (U/L) | 27                        | 33                    | 122 H      | 49 H         |
| Alanine aminotransferase, serum (U/L) | 37                        | 25                    | 89 H       | 70 H         |
| PT (sec)                | 1.3 H                      | 1.2 H                 | 14.6 H     | 12.9 H       |
| PTT (sec)               | 23 L                       | 32                    | 23 L       | 32           |
| D-dimer (ng/mL)         | 403                        | 887 H                 | 433       | 433          |
| Fibrinogen (mg/dL)      | 673 H                      | 0.04 H                | 3.02 H     |
| Troponin I (ng/mL)      | <0.02                      | 0.04 H                | <0.02      |
| Ferritin, serum (ng/mL) | 110                        | 685 H                 | 87        |
| Glucose, serum (mg/dL)  | 135 H                      | 75                    | 87        |
| Ca (mg/dL)              | 8.4 L                      | 7.7 L                 | 8.7       |
| Na (mmol/L)             | 137                        | 131 L                 | 141       |
| Cl (mmol/L)             | 101                        | 94 L                  | 103       |
| BUN (mg/dL)             | 11                         | 31 H                  | 7         |
| Creatinine (mg/dL)      | 0.79                       | 0.9                   | 0.51      |
| Albumin (g/dL)          | 3.6                        | 2.4 L                 | 3 L       |
| Lactate dehydrogenase (U/L) | 244                      | 592 H                 | 257 H     |
| pH, arterial            |                            | 7.53 H                |           |
| pCO2, arterial          |                            | 35 L                  |           |
| pO2, arterial           |                            | 53 L                  |           |
| Bicarbonate, arterial (mmol/L) | 30                        | 26                    | 30        |
| Anion gap (mEq/L)       | 14                         | 9 L                   | 13        |
| IL-6 (pg/mL)            | 22 H                       | <5 L                  | <5 L      |
| IgA, serum (mg/dL)      | <5 L                       | <5 L                  | <5 L      |
| IgG, serum (mg/dL)      | 1234                       | 1100                  |           |
| IgM, serum (mg/dL)      | <5 L                       | <5 L                  |           |

L = low; H = high.
Note. The boldface values signify abnormal laboratory values (higher or lower than normal reference range).

Table 2. Laboratory Results Prior and During COVID-19.
Similar observations were reported for two patients with X-linked agammaglobulinemia in Italy infected with SARS-CoV-2 who were able to recover without requiring intensive level of care.6 Our patient demonstrated a form of immunodeficiency as the result of a chemotherapeutic agent, and his clinical course appeared to be more severe. Rituximab is known to modulate autoimmune responses by suppressing the humoral function and decreasing the body’s ability to respond to infection independent of COVID-19.9 In our case, while providing an essential role in managing the rheumatologic conditions, rituximab resulted in an acquired humoral deficiency requiring IVIG to relieve chronic sinus infections prior to COVID-19. The continuation of IVIG regimen during the course of COVID-19 conferred the importance of passive immunity in helping our patient to recover from COVID-19 pneumonia without the presence of superimposed infections. The highlighted role of IVIG in immunocompromised patients6,7 further supported the exploration of the effectiveness of convalescent plasma therapy for COVID-19. Anti-IL-6 monoclonal antibodies were experimentally given to primary immunodeficient patients in Italy to manage the cytokine storm,5 but they are currently under phase 2 clinical trials for immunocompetent patients with COVID-19 (Clinicaltrials.gov identifier: NCT04322773).10

The inflammatory response to SARS-CoV-2 may be milder in patients with agammaglobulinemia due to the lack of immunoglobulin mediated inflammatory process. It is possible that the pathway which SARS-CoV-2 amplifies inflammatory response is either absent or diminished in patients with agammaglobulinemia, which results in milder course of COVID-19. On the other hand, CVID or acquired humoral deficiencies may have some intact inflammatory mediated pathways that allow SARS-CoV-2 to induce the cytokine storm, which results in a worse clinical presentation of COVID-19. More studies are necessary to shed light on the immunological response to SARS-CoV-2 and its impact on immunocompromised and immunocompetent individuals. We describe the course of a patient with COVID-19 in the setting of an acquired humoral deficiency as a result of chemotherapeutic treatment for rheumatologic conditions.
Author Contributions
Phuong Daniels DPT, OMS-II, Jason Schend D.O., Neha Sanan D.O., Haig Tcheurekdjian M.D., and Robert Hostoffer D.O.: conception and design of the study, data generation, analysis and interpretation of the data, and preparation and clinical revision of the manuscript.

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