Ecophysiological Studies of a Cereal Crop *Oryza sativa* L. with Zinc Stress

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### Abstract

Plants exposed to zinc stress at high concentration inhibit seed germination, seedling growth and development. The germination data showed that there is a negative impact of concentration of zinc sulfate on the germination of seeds. Root and shoot growth of seedlings was worse affected when exposed to higher concentration on ZnSO₄. Shoots were more affected than roots. Morphologically they look different from normal roots by their shape and size. Zn toxicity inhibits the chlorophyll formation and with the increase in concentration of Zn, the shoots showed a decline in chlorophyll-a (chl-a) and chlorophyll-b (chl-b) and total chlorophyll pigment content. Conclusively, the result shows that zinc sulphate at high concentration decreases seed germination, chlorophyll-a, chl-b and total chlorophyll content and also decreases root and shoot growth of plants.

### 1. Introduction

Pesticides treated to seeds have tendency to penetrate into tissue of the plant. There, they transformed into metabolites, which are physiologically more active than the parent compounds and finally affects the seed health and quality. Herbicides treatment was found to be inhibiting the uptake of potassium ions and rice seedling growth. In order to accumulate Zn in grains, rice plants have to uptake, mobilize, and transport Zn from soil to grain, which involves many complex physiological processes at different levels within the rice plant, provided there is an adequate supply of Zn in the soil. In general there are three major rate of limiting steps or barriers for efficient Zn accumulation in rice grain: 1) soil-to-root barriers; 2) root-to-shoot barriers; and 3) barriers in loading Zn into grains. Root uptake is the first step towards the accumulation of Zn in rice grains.

The availability of soil Zn for rice from flooded (anaerobic) soil is affected by an additional set of factors such as soil redox potential, total sulfur content, and soluble bicarbonate. Thus, a combination of agronomic management practices and genetic approaches are essential to improve the soil health conditions to enhance the root uptake of Zn. In rice, direct root uptake, remobilizations from vegetative tissues or combination of both of these two approaches are the main source of Zn in grains [1]. There is a continuous xylem flow from root to grain mediated by transpiration system, which can directly transport Zn to grains [2]. However Zn movement is restricted by the presence of barriers for root-to-shoot transfer and for internal allocation and reallocation of Zn within and between vegetative and reproductive tissues, which leads to reduced accumulation of Zn in grains [3].

Plant factors that affect root Zn uptake include root architecture, root hairs, crown root development, root surface area, root anatomical structures and modification of rhizosphere chemistry through exudation of protons, which can change soil pH, thereby improve the solubility of Zn in the soil and facilitate its diffusion to the root surface. Soil factors that affect the plant-availability of Zn for all crops include soil pH, texture, organic matter content, mineralogy, and microbial populations [4].

Even though there are huge amounts of Zn in vegetative tissues of rice plants, remobilization of Zn from vegetative tissues to reproductive tissues and finally to grains is limited due to selective phloem transport of Zn from old tissues to new tissues and to the grains [1, 5]. Flag leaf, which plays an important role in photosynthesis and grain yield, were found to have a little contribution to grain Zn [6]. Wu, et al. reported significant translocation of Zn from flag leaf to the grain. A continuous supply of Zn to different tissues throughout the life cycle by translocation and phloem remobilization to grains is an important feature of Zn efficient rice genotypes [7]. It is also interesting to note that at lower tissue Zn concentrations, most of the Zn was found in leaf and reproductive tissues, while at higher Zn levels, stem and roots showed increased Zn. Also, the increased root uptake of Zn and root to shoot transfer could not proportionately increase the grain Zn concentrations indicating that internal translocation/retranslocation of Zn from vegetative tissues to grains is the major bottleneck for improving grain Zn concentrations [7, 9].

Though, a number of physiological studies have been published about Zn-efficient rice, little is known on how Zn is redistributed and remobilized from vegetative tissues to the grains [9].

A better understanding of the mechanisms involved in loading of Zn into the endosperm of rice and identification of rice genotypes with better Zn remobilization capacity without having any adverse effect on yield will be highly useful for Zn biofortification of rice [5, 10]. Rice has also been found to show different levels and patterns of Zn accumulation under high or low Zn conditions and in different rice ecosystems [11-13].

### 2. Experimental Methods

#### 2.1 Selection of Rice Cultivars (Experimental Plant)

*Oryza sativa* L. is a common cereal crop in Odisha and widely cultivated *Oryza sativa* L. Pooja is widely cultivated cereal crop by the local farmers in Odisha which are likely to expose heavy metal stress and there is a need to study the effects of different concentration of heavy metal. The poonadan (IR74371-70-1-1) is a drought tolerant variety developed at IRRI, New Delhi.

#### 2.2 Seed Selection

Pure line of *Oryza sativa* L. Pooja is procured from OUAT Ratnapur, Berhampur, is used for experiment. These certified seeds are examined under preliminary selection for uniformity of size, shape and colour. The healthy and uniform seeds were sorted out by sieving and sorting before experiment.

#### 2.3 Test Chemical

Zinc sulphate (ZnSO₄) was used as the test chemical. First stock solution of 1000 mg/L was prepared by dissolving 1g of test chemical in 1 L of distilled water, different concentration of solution of 25, 50, 75, 100 mg/L.

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prepared by proportional dilution with distilled water which is used for various treatments.

2.4 Study of Seed Viability

The seeds were soaked with distilled water or different concentration of test solution from 12-14 h. After scheduled period of exposure, the seeds were washed with tap water, followed by distilled water and their viability was tested according and the result was expressed as % of mortality of seed grains.

2.5 Germination Studies

The seeds of *Oryza sativa* L. Poja showed 90% germination during September-December. For germination studies sterilized plastic bowls were used for the study. Sterilized absorbent cotton clothes were spread on the bowls where test chemicals or distilled water were added to it. Seeds were soaked for 24 hr in the desired concentration of water, 15 numbers of seeds were sown in each bowl at uniform distance in all the sets. The cotton clothes and soil were made to wet at regular interval with distilled water and respective concentration of the test chemical (25, 50, 75, 100 mg/L). The bowls were placed under mild sunlight. The emergence of radical or plumule was considered as an index of germination. Seeds were allowed to germinate. Better sprouted and healthy seedling of 10 days old were used as experimental material. Care was taken to avoid drying and over flooding of test chemical in the bowls.

2.6 Selection of Effective Concentration

Pilot experiment indicates that the metal exerts both promoting and retarding effect on the germination and seedling growth of the plant and it was essential to select a limited no. of concentration for further experiments. Thus four concentration of ZnSO₄, 25, 50, 75, 100 mg/L were selected. The data obtained from the germination and seedling growth experiments are used for screening.

2.7 Morphological Studies

The growth of plant was evaluated by measuring the shoot and root length of seedlings on 11th day. 15 cm scale was used for the measurement of the shoot and root length.

2.8 Biochemical Studies

Biochemical studies were made using 10 day’s old seedling. For this purpose seedlings were grown as described as before. However, in this case in addition to the control, 4 concentration (mg/L) of the metal (25, 50, 75, 100 mg/L) were selected by a preliminary screening were used for the growth of the plants. The root and shoot portion of the 10 days old seedling were separated, weighted and analyzed for various biochemical components.

2.9 Estimation of Chlorophyll

The fresh samples of shoot materials of the 10 days old seedlings were collected. Care was taken for separation of each control and treated samples. A known quantity of about 100mg of samples of weighted shoot material was taken in a mortar and pestle and macerated the paste by adding 80% acetone was added, stirred thoroughly and again centrifuged (20 min). Both supernatants were mixed together and the volume was made to 10 mL. The residue was kept for protein estimation. The absorbance of each extract was determined in a spectrophotometer at a wavelength of 610 and 700 nm of wavelength.

The total chlorophyll (chl), chl-a, chl-b content was measured by recording the absorbance of each extract at 610 and 700 nm wavelength and the values were calculated by using the formula given by Arnon (1949, 1959) [14].

3. Results and Discussion

The changes in root and shoot length of 10 days old seedling were shown in table and figure. The radical emerged with in 48 hrs. Incubation in the dark room at temperature. The emergence of radical or plumule was considered as index of germination. The seeds are exposed to light after germination. In the next day after exposing the seeds to light there was rapid elongation of radicle and plumule. At the end of 92 hrs, the most of seeds showed radicle emergence and there was rapid elongation of primary root and shoot splits from coleoptiles and emerged as primary leaf. At the end of 10th day, the seedling showed primary root with adventitious roots (nodal roots), a number of rootlets and fully expanded 1st and 2nd leaf in control.

![Fig. 1](https://doi.org/10.30799/sepfr.147.18040402)

*Fig. 1* Photo showing the growth of Rice seedlings (up to 10 days) after zinc stress at varied concentrations of ZnSO₄

The root length and shoot length were measured in the seedling after treatment with different concentration of zinc sulphate for 10 days (Figs. 1-5). From the table and figures, data clearly indicates a sharp decline in root and shoot of seedlings. As we increased the concentration of zinc sulphate the root and shoot length decreased. On the whole there was an inverse relationship between zinc concentration and seedling growth. The ratio of root to shoot length was also shown in table. The ratio also decreased constantly from control (1.73) to high concentration of ZnSO₄ at 100 mg/L (1.95).

![Fig. 2](https://doi.org/10.30799/sepfr.147.18040402)

*Fig. 2* Percentage germination of rice seeds in response to Zn stress

![Fig. 3](https://doi.org/10.30799/sepfr.147.18040402)

*Fig. 3* Percentage of leaf emergence (168 h) in rice seedlings after Zn stress

![Fig. 4](https://doi.org/10.30799/sepfr.147.18040402)

*Fig. 4* Changes in root length (Series 1), shoot length (Series 2) and R/S ratio (Series 3) 10 days old in Rice seedlings after Zn stress

![Fig. 5](https://doi.org/10.30799/sepfr.147.18040402)

*Fig. 5* Pigment concentration in rice seedlings (10 days old) after Znic stress
There were visible changes seen in the roots which appeared different from that of the control. There was marked difference in the development of primary roots and adventitious roots. On exposure to high concentration beyond 50 mg/L, there was a reduction in the primary roots, no lateral roots developed, and blackening or browning of root tip. Shoot elongation affected as we increased the concentration of test chemical. Chlorosis of leaves, patchy brown spots on leaves, necrotic lesions on leaves and eventually senescence of leaves starts at higher concentration. Effect of the test chemical on the chlorophyll contents against the control and percentage in the pigment are presented. Slightly decreased in chlorophyll pigment were observed in increase in the concentration of ZnSO₄. The percent change in the amount of chlorophyll content and total chlorophyll of the exposed seedlings of 1000mg/l showed a fall. Zinc efficiency, which is used synonymously with Zn deficiency tolerance, reflects the ability of a plant to grow and yield well under Zn-deficient conditions. The proposed physiological mechanisms for Zn efficiency in rice at the early vegetative growth stage can be grouped into two categories: those related to increased root uptake of Zn and those related to internal Zn distribution. Mechanisms for increased root uptake in rice include proliferation of crown root growth, exudation of organic acids or phytosiderophores, and increased tolerance of radical oxygen stress.

The present study, exposure of ZnSO₄ stress and its responses at different parameters of Pooja variety of rice seedling: germination studies, seedling growth, chlorophyll content and total chlorophyll content.

In the present study on exposure of rice seedling to high concentration of ZnSO₄ decreases the chlorophyll, total chlorophyll. In general, decrease in germination has been one of the important manifestations of metal toxicity [15]. The phytotoxicity ZnSO₄ indicated by decrease in growth and development and metabolism an induction of oxidative damage in various plant species. ZnSO₄ toxicity in plants limited the growth of both root and shoot. Despite being an essential micronutrient for normal growth and development Zinc becomes phytotoxic and inhibit plant growth when in higher concentration. ZnSO₄ may cause cytological disorder in plants, though much higher concentration are needed for affecting both patterns [16].

A number reports were given by different workers for studies of the effect on ZnSO₄ on seedling growth of various plants gave a report on a drastic reduction of germination rice seedling. Their studies gave a clear picture roots are more responsive to metal compare to which support our present that on seedling growth.

We obtained almost similar results when we compare the effect of ZnSO₄ growth of Oryza sativa L. seedlings with available literature. The inhibitory effect achieved in the present study may be due to higher dose than dose required by plant for its growth.

Under high ZnSO₄, the chloroplast swells and grana being destroy in Oryza Sativa L. seedling. General symptoms of zinc toxicity are turgor loss, chlorosis and necrosis of leaves [17]. The excess ZnSO₄ treatment bought about a marked depression in photosynthetic pigment like chl-a and chl-b in plants. It might be due to excess supply of ZnSO₄ in interference with synthesis of chlorophyll.

In this above view chl-a, chl-b and total chlorophyll content in exposed leaves slightly declined with increase in concentration of ZnSO₄ which was measured in shoot of 10 day seedling. The findings also indicate that at higher concentration of metals cause leaves chlorosis and necrosis.

5. Conclusion

The present investigation of experimental induction of Zn stress to rice seedlings, showed decrease in seed germination, chl-a, chl-b and total chlorophyll content and also decreases root and shoot growth of plants. It has also been observed that, the increase in Zn concentration has decreased the observed parameters significantly.

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References

[1] S.M. Impa, M.J. Morete, A.M. Ismail, R. Schulin, S.E. Johnson-Beebout, Zn uptake translocation and grain Zn loading in rice (Oryza sativa L.) genotypes selected for Zn-deficiency tolerance and high grain Zn, J. Exp. Bot. 64 (2013) 2739-2751.
[2] S. Krishnan, P. Dayanandan, Structural and histochemical studies on grain-filling in the caryopsis of rice (Oryza sativa L.), J. Biosci. 28 (2003) 455-469.
[3] W. Jiang, P.C. Strulk, H. Van Keulen, M. Zhao, L.N. Jin, T.J. Stomph, Does increased zinc uptake enhance grain zinc mass concentration in rice?, Ann. Appl. Biol. 153 (2008) 135-147.
[4] G. Hacisalihoglu, L.V. Kochian, How do some plants tolerate low levels of soil zinc? Mechanisms of zinc efficiency in crop plants, New Physiol. 159 (2003) 341-350.
[5] C.Y. Wu, L.L. Lu, X.E. Yang, Y. Geng, Y.Y. Wei, et al., Uptake translocation and remobilization Zinc absorbed at different growth stages by rice genotypes of different Zn densities, J. Agric. Food Chem. 58 (2010) 6767-6773.
[6] R.A. Sperotto, P.K. Ricchadenevsky, V. Waldow, A.L.H. Muller, V.L. Dressier, J.P. Fett, Rice grain Fe, Mn and Zn accumulation: How important are flag leaves and seed number?, Plant Soil Environ. 59 (2013) 262-266.
[7] H.J. Yin, X.P. Gao, T. Stomph, L. Li, F. Zhang, C.Q. Zou, Zinc concentration in rice (Oryza sativa L.) grains and allocation in plants as affected by different zinc fertilization strategies, Commun. Soil Sci. Plant Anal. 47 (2016) 761-768.
[8] T.J. Stomph, W. Jiang, P.E. Van Der Putten, P.C. Strulk, Zinc allocation and re-allocation in rice, Front Plant Sci. 5(8) (2014) 1-12.
[9] X.L. Ren, Q.L. Liu, D.X. Wu, Q.Y. Shu, Variations in concentration and distribution of health-related elements affected by environmental and genotypic differences in rice grains, Rice Sci. 13 (2006) 170-178.
[10] S.I. Jiang, J.G. Wu, Y. Feng, X.E. Yang, C.H. Shi, Correlation analysis of mineral element contents and quality traits in milled rice (Oryza sativa L.), J. Agril. Food Chem. 55 (2007) 9608-9613.
[11] M. Wixuxua, A.M. Ismail, S. Vanaghara, Effects of zinc deficiency on rice growth and genetic factors contributing to tolerance, Plant Physiol. 142 (2006) 731-741.
[12] R.L. Mabesa, S.M. Impa, D. Grewal, S.E. Johnson-Beebout, Contrastng grain-Zn response of biofortification rice (Oryza sativa L) breeding lines to foliar Zn application, Field Crop Res. 149 (2013) 223-233.
[13] S.M. Impa, A.S. Gramlich, S. Tandy, R. Schulin, E. Frossard, S.E. Johnson-Beebout, Internal Zn allocation influences Zn deficiency tolerance and grain Zn loading in rice (Oryza sativa L), Front Plant Sci. 4(534) (2013) 1-10.
[14] D.J. Arnon, Copper enzymes in isolated chloroplast polyphenol oxidase in Beta vulgaris, Plant Physiol. 24 (1949) 1-15.
[15] W. Bergmann, Nutritional disorders of plants, Gustav Fischer Verlag, GERMANY, 1992.
[16] A.M.B. Pahlsson, Toxicity of heavy metals (Zn, Cu, Cd, Pb) to vascular plants: A literature review, Water, Air Soil Pollut. 47 (1989) 267-319.
[17] S. Umar, Moinuddin, M. Iqbal, G.M. Vidyasagar, D. Koteshra, et al., Role of endosulfan in mediating stress responses in Sorghum bicolor (L) Moench, J. Environ. Biol. 30(2) (2009) 217-220.