Introduction

Cassava (*Manihot esculenta crantz*) is a perennial shrub grown principally for its starchy roots which are used as food, animal feed and as a source of starch. Cassava tends to be grown on the poorer agricultural lands, without irrigation and with limited application of purchased inputs. It is naturally well adapted to these conditions [1]. Nevertheless, the tremendous variation in conditions to
which it is subjected means that cassava production technology needs to be adapted to the varying natural conditions, rather than using costly modifications of the environment to suit a particular production system [2]. An understanding of the manner in which the crop responds to varying environmental conditions is an essential component of designing improved low-input technologies like biofertilizer, well adapted to the particular conditions where individual farmers grow their crops. The cassava crop is essentially grown between 300 S and 300 N latitude of the globe. As the crop moves further north or south of the equator the maximum altitude at which it grows and produces will decrease. The crop is generally not found in areas where the mean average temperature is less than about 20 °C, although in areas near the equator where seasonal temperature fluctuations are small it can be found growing in areas with a mean temperature as low as 17 °C [3].

To increase the yield potential of cassava, the crop has been reported to respond to good soil fertility and adequate fertilizer [4]. The major nutrients required by cassava for optimum top growth and tuber yields are nitrogen (N) and potassium (K). Soils that have low N (<0.10% total N) and K (<0.15 meg/100 g) will require an additional fertilizer for optimum tuber yield [5]. Adequate K levels in the soil stimulate the response to N fertilizers, but excess amount of both nutrient leads to luxuriant growth at the expense of tuber formation [6]. Cropping systems influence fertilizer requirements of cassava; for example, the continuous cropping of cassava leads to fast depletion of major nutrients, especially N and K and will require fertilizer supplement to give stable yield. Cassava removes about 55 kg/ha N, 132 kg/ha P and 112 kg/ha K reported by [7].

Cassava Improvement and Biotechnology

However, farmers rarely use chemical fertilizer due to scarcity and cost, hence the dependence on cheap organic sources of nutrients. These reasons necessitate research on increasing effectiveness of organic manures and suitable rate of application. The effect of digester effluent was compared with pig and cattle manure [8] and it was found that bio-digester effluent gave higher biomas, yield and protein content of cassava. These necessitate the biofertilizer research production using agriculture wastes inoculated with broad spectrum microorganism accelerated composed in an anaerobic bio digester. One of the factors responsible for low yield is declining soil fertility. In the past, soil fertility has been sustained through long fallowing [9]. When biofertilizers applied as soil inoculants, they multiply and participate in nutrient cycling and benefit crop productivity [10]. OB-Digester composted using anaerobic bio-digester technology (https://www.youtube.com/watch?v=Hi_OpgVcFcg biofertilizer) from bio-waste in anaerobic digester inoculated with beneficial microbes that exhibit differing metabolic capabilities.

Biotechnology tools have been adapted to cassava and are currently incorporated in different projects for its genetic improvement. A molecular map has been developed [11,12] and marker-assisted selection is currently used for key traits [13]. Molecular markers that allow selection of segregating progenies that carry the resistance of Cassava Mosaic Disease (CMD) have been successfully utilized [13]. A molecular genetic map has been developed for cassava [12] where cassava genetic improvement can be made more efficient through the use of easily assayable molecular genetic or DNA markers (MAS) that enable the precise identification of genotype without the confounding effect of the environment; in other words, increasing heritability.

Genetic transformation protocols are available and have been used successfully for the incorporation of different genes [14]. As the crop has evolved and new improved varieties that satisfy the most important needs have been released and adopted by farmers, new challenges and opportunities arise. An important need is to introduce herbicide tolerance in the crop. Several approaches can be taken from genetic transformation [15], to screening for the natural occurrence of tolerance to certain herbicides, to the induction of mutations as already demonstrated for different herbicides and different crops [16]. The lack of genetic variability for overcoming the problem of post-harvest physiological deterioration remains a major bottleneck for cassava utilization and commercialization, although significant breakthroughs have been achieved recently.

Systemic Approach for Sustainable Agriculture Delivering food security and improving food quality to sustain population growth without compromising environmental safety is global benchmark for green revolution [17]. The beneficial effects and mechanisms of microbes on plant health and fitness and their utilization in agriculture are widely studied and documented [18-23]. The rhizosphere is the interface between roots and the soil where nutrient absorption for plant growth in agroecosystems is facilitated. Carbon flows from the plant to the soil ecosystem as simple organic compounds providing the necessary food basis for the corresponding microbiological processes that are vital for soil ecosystem functioning [24]. Both plant beneficial microorganisms (plant growth promoters and biocontrol agents) and pests (root pathogens and root feeding insects) are common inhabitants of the rhizosphere [25], all affecting C, N and P biogeochemical processes in the soil. Functional traits of the beneficial rhizosphere microbiome in relation to plant nutrition and health include organic matter decomposition, P solubilization and transport, N fixation and biocontrol of root pests [26]. Biological indicators in the past stressed management effects on biodiversity, e.g., were conservation oriented, but recent developments emphasize methods indicating soil functions and general soil health [27-30]. These different approaches reflect the obvious interrelations between physical, chemical and biological agents in the soil systems.

Biofertilizer

A key merit of microorganisms is to assimilate phosphorus for their own requirement, which in turn available as its soluble form in sufficient quantities in soil. Pseudomonas, Bacillus, Micrococcus, Flavobacterium, Fusarium, Sclerotium, Aspergillus and Penicillium have been reported to be active in the solubilization process [31].

A phosphate-solubilizing bacterial strain Micrococcus sp. has polyvalent properties including phosphate solubilization

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and siderophore production [32]. Similarly, two fungi Aspergillus fumigatus and Aspergillus Niger were isolated from decaying cassava peels were found to convert cassava wastes by the semi-

solid fermentation technique to phosphate biofertilizers [33] Burkholderia vietnamiensis, stress tolerant bacteria, produces gluconic and 2-ketogluconic acids, which involved in phosphate solubilization. Potassium solubilizing microorganisms (KSM) such as genus Aspergillus, Bacillus and Clostridium are found to be efficient in potassium solubilization in the soil and mobilize in different crops [34]. Mycorrhizal mutualistic symbiosis with plant roots satisfies the plant nutrients demand [35] which leads to the enhancement plant growth and ’development and protect plants from pathogens attack and environmental stress Figure 1. Pseudomonas aeruginosa has been shown to withstand biotic and abiotic stresses [31], Paul and Nair [36] found that P. fluorescens MSP-393 produces osmolytes and salt-stress induced proteins that overcome the negative effects of salt. Microbial inoculants genera in the OBD-Biofertilizer are isolated using the growth media in Table 1 from different agro biowaste and inoculated into the composted biofertilizer (Plant Growth-Promoting Rhizobacteria. Microorganisms, PGPRs) Table 2.

Figure 1: Endophytic bacterial colonization in cassava plants modelled. Bacteria can enter a plant at several root zones as indicated above. Endophytes can either remain at the site of entry (indicated in blue) or move deeper inside or occupy the intercellular space of the cortex and xylem vessels (indicated in green). Red and yellow represent rhizospheric bacteria which are unable to colonize inner plant tissues adapted from Maheshwari & Annapurna [138].

Table 1: Microbial culture techniques of beneficial microorganisms presents in OBD-Biofertilizer.

| S.N. | Growth Media | Microbes | References |
|------|--------------|----------|------------|
| 1    | Ammonium mineral salt | Methylotrophs | Holland et al. (2000) |
| 2    | Congo red yeast manitol | Rhizobium | Yumoto et al. (2002) |
| 3    | DSMZ-97, DSMZ-823, DSMZ-1104; OS | Halophilic archaea | Yadav et al. (2015c) |
| 4    | Jensen agar | N2-fixing bacteria | Jensen (1965) |
| 5    | King’s Bagar | Pseudomonas sp. | Mishra et al. (2009) |
| 6    | Luria Bertani agar | Endophytic bacteria | Ventosa et al. (1982) |
| 7    | Nutrient agar | Heterotrophic bacteria | Ramesh and Lonsane (1987) |
| 8    | Potato dextrose agar | Fungus | Sehgal and Gibbons (1960) |
| 9    | Soil extract agar | Soil-specific microbes | Shivaji et al. (1988) |
| 10   | Tryptic soy agar | Arthrobacter | Shivaji et al. (1989) |

Table 2: PGPRs Biosurfactants presents in OTAI AG® Inocula with literature references.

| N/S | Biosurfactants | Microorganisms | Economic Importance | References |
|-----|----------------|----------------|---------------------|------------|
| 1   | Rhamnolipids   | Pseudomonas aeruginosa | Antimicrobial | Jadhav, et al. [128] |
|     |                |                 | biocontrol properties |           |
| 2   | Viscosin       | Pseudomonas fluorescens | Lipopeptides | Banat, et al. [114] |
| 3   | Ornithine lipids | Agrobacterium sp. | Bio-emulsifiers | Desai & Banat [120] |
|     |                | Pseudomonas sp. | Bio-emulsifiers |           |
Manipulation of the plant microbiome has great potential in reducing the incidence of pests and diseases [37,38], promoting plant growth and plant fitness, and increasing productivity [39,40]. Single strains or mixed inoculum treatments induced resistance to multiple plant diseases [41]. In recent years, several microbial biofertilizers and inoculants were formulated, produced, marketed, and successfully used by farmers worldwide. Many processes in the rhizosphere lead to interactions been roots, microbes, water, and nutrients. For example, plant roots and microbes compete for nitrogen, and most likely other nutrients. The rhizosphere has been defined in terms of the effects of roots on soil microorganisms, the depletion of water [120], changes in pH [43], adhering soil [44], Hiltner [45] defined the rhizosphere as the soil influenced by roots. The rhizosphere is not a static place, but rather a dynamic system of processes and increasing the spatiotemporal resolution of rhizosphere measurements will lead to new insights and allows it to be considered as an extended phenotype [46], or an external manifestation of a plant’s genetics.

**Rationale and Significance**

The cultivation of the resistance of Cassava Mosaic Disease (CMD) in response to chemical and biofertilizer using recommended application to study the crop yield and crops components was the object of the research. Planting one stake of cassava per hill or stand using 1m x 1m spacing, by the National Root Crops Research Institute (NRCRI) Umudike, Nigeria, was a major breakthrough in cassava-based farming system practices results to the production of larger roots, high yield per stand and makes other farm operations such as weeding and fertilizer application easy. The reasons advanced for the low adoption was low cassava population, low yield per unit area and weed growth in farms. Therefore, farmers are demanding for an increase in the number of cassavas to be planted per hill or with little or no chemical pesticides applied during cultivation. Cassava yields are compromised by pests such as whiteflies, mites, and weevils, which cause significant crop losses through the spread of viral disease and direct damage to plants which can reduce yields by up to 40% [47].

Studies shows the enhanced by the production of bioactive substances having similar effects as that of growth regulators besides nitrogen fixation through biofertilizer leading to greater dry matter production was reported by Ramanandam et al. [48]. The higher dry matter production is attributed to the cumulative effect of progressive increase in the growth attributes, viz., plant height, stem girth and number of leaves per plant [49]. Cassava is generally weeded by hand (hoe) 2-3 times during the first 3-4 months, but herbicides across the globe, Figure 3. The excitation (biofertilizer application) to the cassava cultivation soil system is functional characteristics of the impacts on rhizosphere considered for diagnostic tests of ‘Biofertilizer rhizosphere holistic soil function’. With the narratives in Figure 2.

| Microorganism | Bio-emulsifiers | Reference |
|---------------|----------------|-----------|
| *Thiobacillus thiooxidans* | *P.fluorescens* | Nerurka, et al. [140] |
| Protein PA | *Paeruginosa* | Hisatsuka et al., 1999 |
| Whole cell | *Cyanobacteria* | Levy, et al. [136] |
| Surfactin/Iiturin | *B. subtilis*, Antimicrobial | Arguella et al., 2009 |
| Subtilisin | *B. subtilis*, Antimicrobial properties | Sutiyak, et al. [152] |
| Aminoacids lipids | *Bacillus sp*, Antimicrobial properties | Cotter, et al. [117] |
| Lichenysin | Enhance oil recovery | Yakimov et al., 2009 |

**Figure 2:** The excitation (biofertilizer application) to the cassava cultivation soil system is functional characteristics of the impacts on rhizosphere considered for diagnostic tests of ‘Biofertilizer rhizosphere holistic soil function’. Where indicated by grey (Paper 1) circles while the connecting processes are in white (Paper 2) and Paper 3 (sustainable soil management).

**Source:** Adapted from Vogel, Hans-Jörg et al. [124].
Optimizing cocktails of microorganisms inoculated in the biofertilizer production will produce microbial enzymes and metabolites, which mimic the multiplicity of bio-control mechanisms, set-up by microorganisms [50]. This category includes microbial secondary metabolites and hydrolytic enzymes as glucanases, proteases, lipases, and chitinases. These molecules can be used alone or, better, in combination; they can also be exploited in addition to chemical pesticides with the scope to favor their action, thus reducing the introduction and impact of synthetic pesticides on ecosystems. Yadav et al. [51] obtained consistently higher crop yield with NPK fertilizer mixed with organic manure over NPK inorganic fertilizer alone, Ano and Emehute [52] also obtained higher ginger rhizome yield with organic manure mixed with inorganic fertilizer over inorganic fertilizer alone. Complementary use of organic manure and inorganic fertilizer improves the soil resource base. The effect of biofertilizer on cassava microbiome and phytobiome is unknown or under investigation today. This research article series reports results application of accelerated OBD-Biofertilizer applied alone and in combination with inorganic fertilizer NPK (15:15:15).

The objectives of this paper are “biofertilizer rhizosphere diagnostic tests” to directly evaluate the dynamics of soil in response to targeted forcing (application of bio-fertilizer) in cassava cultivation in the tropic (eastern Nigeria). The observable dynamics provide information on the internal pattern of interacting processes of cassava-microbes interaction as an integral function of the impact of soil management and affirmed by Kibblewhite et al. [28] criticize the “reductionist” approach of using simple indicators describing some fixed state of the soil. In recent years, several microbial biofertilizers and inoculants were formulated, produced, marketed, and successfully used by farmers worldwide. The relationship between rhizodeposition and plant nutrient status is highlighted by the rhizosphere priming effect where N mineralization is increased near roots due to microbial activity [53]. The mapping of microbes in soil has identified microbial hotspots in the rhizosphere [54].

Better understanding of interactions between roots and rhizosphere processes promise to lead to new knowledge and mechanistic insights of the functions of rhizosphere microorganisms on cassava crop cultivation and development:

**Paper 1:** (Biofertilizer impacts on cassava (*Manihot esculenta crantz*) Rhizosphere: Crop yield and Growth components, Igbariam, Nigeria).

**Paper 2:** (Biofertilizer impacts on cassava (*Manihot esculenta crantz*) rhizosphere: Soil health and Quality, Igbariam, Nigeria).

**Paper 3:** (Biofertilizer Impacts on Cassava (*Manihot Esculenta Crantz*) Rhizosphere: Sustainable soil management, Igbariam, Nigeria).

These papers construct was affirmed by similar work of Cheng et al., [55], where the strategy of one organism depended on the strategies of others, demonstrated that rhizosphere priming could develop as a mutualism between plants and microbes in some limited ecological conditions. Such a systemic approach ~ (Figure 3), providing a clear perspective on how soil functions emerge from small-scale process interactions, is a prerequisite to actually understanding the basic controls and to developing science-based strategies towards sustainable soil management [56].

**Materials and Method**

**Anaerobic digestion**

During the anaerobic digestion process, organic compounds are broken down, firstly via lactogenic bacteria to methane precursors, largely volatile fatty acids (VFAs) and then to methane and other products via methanogenic bacteria. OBD-Biofertilizer composted using anaerobic bio-digester technology (https://www.youtube.com/watch?v=Hi_OpgVCfg biofertilizer) from bio-waste in anaerobic digester inoculated with beneficial microbes that exhibit differing metabolic capabilities. Under anaerobic conditions, organic forms of nitrogen (N) are converted into Ammonium-N (NH-N), i.e. readily available nitrogen.

The readily available nitrogen (RAN) content of cattle slurry is typically 50% and pig slurry 60% of Total-N [57]. It might be anticipated that a measurable increase in the proportion of readily available nitrogen (N) would be achieved by applying biofertilizer.
available N would occur in these materials, as a result of the digestion process. Propagation of beneficial microbes. The isolated microbe was propagated using enriching non-selective medium (CPMA). The strains of each group were incubated separately for 7 days at 25 °C and then in mixture for 72 hours at 37 °C under agitation at 75 rpm. The bacterial isolates were then subjected to a series Gram staining and colony count [58]. The comparison was made against the database containing identification patterns for Gram positive and Gram-negative bacteria species.

Bio-waste Recycling to Biofertilizer

Agriculture bio-waste materials composted by anaerobic digester (AD) and inoculated with broad spectrum inoculants OTAI AG® (Table 2) and Oso Bio-Degradar (OBD-Plus®) called microbial inoculants. Inoculated beneficial microbe’s direct analysis of metabolites in situ has been achieved for antibiotic lipopeptides from several Bacillus subtilis and for pyrroline, 2,4-diacylphloroglucinol and phenazene-1-carboxylic acid from *Pseudomonas fluorescens* strains. During the anaerobic digestion process (Figure 4, Plates 1&2) available for the biofertilizer production, Abeokuta, Nigeria.

YOU Tube: https://www.youtube.com/watch?v=pG20DAx3ICY. OTAI AG® is PGPR (Table 2) and beneficial microbial inoculate an easy-to-use applied to the biowaste carrier material (composted) biofertilizer production with industrial standardized process of production and similar to the report by Schmidt [31]. Biofertilizers price is not the same as composts and have been tested as growth media for PGPR [31]. PGPR and/or arbuscular mycorrhiza fungi (AMF) [59] combine inoculation often resulted in increased growth and yield, compared to single inoculation through improved nutrient uptake [60] and resultant interaction of bacteria and AM fungi have beneficial functions related to nutrient uptake, particularly when PGPR [61] and N2-fixing bacteria are involved. Survival of the PGPR is important both during the storage period of the bioproduct and after being introduced into the soil for solid carriers, powder or granules. Standard sizes of the powder material may vary from 75 µm to 0.25 mm [62] and application methods depend on the kind of crop concerned can be inoculated by broadcasting the inoculum over the soil surface, alone or together with seeds, or by in-furrow application, seed dressing, or coating; tree crops can be initially inoculated by root dipping or seedling inoculation [63].

**Chemical analyses:** Total Nitrogen Kjeldahl procedure [64]; Available P Olsen’s method [65]; Available K by Flame photometric method [66]; pH [67]; Electrical Conductivity by Walkley Black method [68]; Micronutrients (Zinc, Iron, Copper, Manganese) ppm Atomic absorption Spectrophotometric method using DTPA (Diethyl Triamine Penta Acetic Acid) by Lindsay and Norvell [69].

**Results and Discussion**

**Biofertilizer physico-chemical properties**

Bio-fertilizers contained 3.5% - 4% nitrogen, 2% - 2.5% phosphorus and 1.5% potassium. The nutrient content of the OBD-Biofertilizer indicated the followings:

- **Macroelements:** pH in water (5.8); %N (0.95); %P (3.1); % Ca (13.05); %Mg (0.79); %K (0.35); ppm Na (40.74); ppm Mn (456.78); ppm Fe (460.01); ppm Cu (17.47); ppm Zn (95.5); %Carbon (35.68) and C/N ratio (35.56).

- **Micronutrients:** Molybdenum (Mo) 20 ppm; Boron (B) 30ppm; Copper (Cu) 17.47ppm; Manganese (Mn) 456.78 ppm; Zinc (Zn) 95.5 ppm; Iron (Fe) 460.01 ppm; Sodium (Na) 40.74ppm.

- **Essential Nutrients:** Humic acid 2.1. Beneficial microorganisms presents are in the OBD-Biofertilizer: Azotobacter spp, Clostridium spp, *Bacillus spp*, Escherichia, *Rhizobium* spp. Fe (iii) reducing Bacteria (*Shewanella putrefaciens*), Phosphate Solubilizing Fungi (*Actinomycetes*), Potassium Solubilizing Bacteria (*Bacillus mucilaginosus*), Phosphate Solubilizing Bacteria (*Rhizobium, Agrobacterium*), Sulphur Oxidising Bacteria (*Thiobacillus thioxidans*).

- **Materials:** Shea cake and Poultry waste (SPW), Swine waste (SW), Wood ash (WA)

- **Ratios:** SPW 12: SW 12.5: WA 1.

- **Percentages:** PW 47.06; SW 49.02; WA 3.92

- **Microbial Inoculants:** OBD-Plus® and OTAI AG®

The trial was conducted at the National Root Crops Research Institute’s, substation in Igbariam in 2012/2013 cropping seasons, Anambra state, Nigeria. The soil was an Ultisol and had a pH in water of 5.0, 2.06% organic matter, 0.14% total nitrogen, 5.8 mg/kg Bray P , exchangeable calcium, Mg and K of 4.60 cmol/kg, 2.50 cmol/kg of 5.0, 2.06% organic matter, 0.14% total nitrogen, 5.8 mg/kg Bray 2 P, exchangeable calcium, Mg and K of 4.60 cmol/kg, 2.50 cmol/kg and 0.12 cmol/kg respectively. The field was slashed ploughed, harrowed and ridged, thereafter plots each measuring 5m x 5m were marked out. Cassava stakes each measuring about 25cm were then planted at 1m x 1m spacing on top of the ridges on 16th November 2012. Cassava Mosaic Disease (CMD) resistant varieties were marked out. Cassava stakes each measuring about 25cm were then planted at 1m x 1m spacing on top of the ridges on 16th November 2012. Cassava Mosaic Disease (CMD) resistant varieties used was from the International Institute of Tropical Agriculture’s (IITA) fields. The treatment was arranged in randomized complete block design replicated three times. The treatments were as follow:

- 300 kg/ha NPK 15:15:15 + 1.0 t/ha
  - OBD- Biofertilizer
- 300 kg/ha NPK 15:15:15 + 2.0 t/ha
  - OBD-Biofertilizer
- 300 kg/ha NPK 15:15:15 + 3.0 t/ha
  - OBD-Biofertilizer
- 300 kg/ha NPK 15:15:15 + 4.0 t/ha
  - OBD-Biofertilizer
- 2.0 t/ha OBD-Biofertilizer
- 600 kg/ha NPK (15:15:15)
The treatments were applied on 9th May 2013. Harvesting of cassava was carried out in November 2013 and the fresh root yield was measured. The data obtained was subjected to analysis of variance. Significant treatment means were separated using Fischer’s least significant difference (F-LSD) at 5% probability and see Plates 1 and 2 below.

Table 3: Effect of OBD-Biofertilizer applied alone and in combination with inorganic fertilizer on cassava root yield.

| S/N | Treatments                                      | Cassava Root Yield (t/ha) |
|-----|------------------------------------------------|---------------------------|
| 1   | 300 kg/ha NPK 15:15:15 + 1.0 t/ha OBD-Biofertilizer | 20.5                      |
| 2   | 300 kg/ha NPK 15:15:15 + 2.0 t/ha OBD-Biofertilizer  | 22                        |
| 3   | 300 kg/ha NPK 15:15:15 + 3.0 t/ha OBD-Biofertilizer  | 30                        |
| 4   | 300 kg/ha NPK 15:15:15 + 4.0 t/ha OBD-Biofertilizer  | 31.2                      |
| 5   | 5.0 t/ha OBD-Biofertilizer                       | 16                        |
| 6   | 600 kg/ha NPK (15:15:15)                         | 35.6                      |
| 7   | Control (no application)                         | 12                        |
|     | F-LSD (0.05)                                     | 3                         |

OBD-Biofertilizer applied at the rate of 4 t/ha mixed with 300 kg NPK (15:15:15) / ha gave the highest cassava root yield of 31.2 t/ha which was however not significantly higher than 30.0 t/ha obtained with OBD-Biofertilizer applied at the rate of 3 t/ha mixed with 300 kg NPK (15:15:15) / ha. Complementary use of OBD-Biofertilizer and inorganic fertilizer is therefore more beneficial than OBD-Biofertilizer alone in cassava production. This result is in agreement with Ano & Ikwelle [70] and Mokwunye,[71]. Soils of the experimental site and indeed most Nigerian soils are highly weathered and have low activity clays [72] and therefore require application of soil amendment for high crop yield to be obtained. This explains why inorganic fertilizer NPK 15:15:15 applied at the recommended rate of 600 kg/ha or inorganic fertilizer applied at 300 kg/ha mixed with OBD-Biofertilizer gave significantly higher yield than the control, Figure 4.

Agronomic Practices

Enhancing the adoption of soil conservation practices

Cassava is well adapted to low fertility soils that predominate in large areas of the tropics. It is frequently cultivated on the highly weathered and leached Oxisols, Ultisols and Alfisols, with smaller areas found on Inceptisols (particularly in India) and Entisols. Cassava is extremely tolerant of low soil pH and high levels of aluminum saturation that often accompany them. It can be found producing moderate yields where many crops simply fail due to the low pH and high levels of aluminum [3]. The highest yields were obtained using the full land preparation methods, including twice plowing and disking followed by contour ridging? To reduce erosion peanut intercrop, with cassava planted on contour ridges, with or without fertilizer application and planting on contour ridges resulted in a decrease in soil loss of almost 40% [73]. At planting time, there should be enough soil moisture to get at least 80-90% germination, while soils should not be too wet to prevent adequate aeration and root formation [74].
Growth, Yield and Root Quality

Biofertilizer facilitate the below-ground

| S/N | Treatments                                      | Cassava Root Yield (t/ha) |
|-----|------------------------------------------------|--------------------------|
| 1   | 300 kg/ha NPK 15:15:15 + 1.0 t/ha OBD-Biofertilizer | 20.5                     |
| 2   | 300 kg/ha NPK 15:15:15 + 2.0 t/ha OBD-Biofertilizer | 22                       |
| 3   | 300 kg/ha NPK 15:15:15 + 3.0 t/ha OBD-Biofertilizer | 30                       |
| 4   | 300 kg/ha NPK 15:15:15 + 4.0 t/ha OBD-Biofertilizer | 31.2                     |
| 5   | 5.0 t/ha OBD-Biofertilizer                      | 16                       |
| 6   | 600 kg/ha NPK (15:15:15)                        | 35.6                     |
| 7   | Control (no application)                        | 12                       |
|     | F-LSD (0.05)                                    | 3                        |

Biological activity of earthworms, bacteria and fungi, and supply a wide range of nutrients, including secondary and micro-nutrients (Table 4). Amanullah et al. [49] observed that the number of roots per plant was significantly influenced by organic fertilizer treatment. Adjei-Nsiah & Issaka [76] observed that average fresh root yield increase from 13.7 t/ha without amendment to 23.7 t/ha with application of 4 t/ha poultry manure and compared with Table 4 where biofertilizer application at 5t/ha yield 16 t/ha and control yield 12 t/ha (Figure 5) due to the impacts of the beneficial microorganisms. Organic fertilizer promotes the growth of stems and leaves of cassava, increase the chlorophyll content and the photosynthesis of leaves and improve the physiological metabolism of cassava [77]. The period of maximum rate of dry matter partitioning depends on genotype-by-environmental interaction [9].

Amanullah et al. [49] observed that the number of roots per plant was significantly influenced by organic fertilizer treatment. Adjei-Nsiah & Issaka [76] observed a significant increase in number of roots per plant, and tuber diameter having a positive correlation with fertilizer treatment. Manure application has resulted in higher root yields of cassava [80]. Manure application enhances the cooking quality (mealiness) of cassava [75]. Various observations have been made of a positive correlation between dry matter content and cooking quality of cassava [81].

The delicate balance between top growth and root growth is the key to understanding the growth and development of cassava [82]. The photosynthetic rate of plants obtained from mother plants that have not been fertilized is similar to that of fertilized mother plants. The status of the nutrient reserves is important for the establishment is important in determining final yield [83] and that confirmed the properties of the biofertilizer in root crops development. The number of thickened roots is determined early in the growth cycle, normally in the first three months [84]. The fibrous feeder roots of cassava were described as being rather sparse by Connor et al. [85], reaching a maximum level of 1 km/m² and depths of at least 2.6m.

Biofertilizer and Inorganic Fertilizer

| N/B | Treatment Materials | Control Yield/Ton | Treatment Yield/Ton | Impacts on Crop Component | References |
|-----|---------------------|-------------------|--------------------|---------------------------|------------|
| 1   | Poultry manure      | 13.7 t/ha         | 23.7 t/ha          | Increase the chlorophyll content in stem/leaves and the photosynthesis and physiological metabolism | Adjei-Nsiah & Issaka [151]; Luo, et al. [77]; Fregence, et al. [9]; Lebot, 2009 |
|     |                     |                   |                    | Higher root yields of cassava | Wilson & Dufour, 2002; Agbaje & Akinlosotu [80]; ssaka, et al 2007; Ojeniyi, et al. [87]; |
|     | 4 t/ha              | 13.7 t/ha         | 23.7 t/ha          | Enhances the cooking quality (mealiness) | Amanullah et al. 2019; |
|     |                     |                   |                    | Positive correlation between dry matter content and cooking quality | Safo-Kantanka & Asare [81]; Safo-Kantanka & Owusu Nipa [62] |

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The use of mineral fertilizer in combination with poultry manure has shown an increase yield as much as 60 t/ha of cassava roots [68]. The fertilizers supplied the bulk of the macronutrients needed by the plants, while the organic sources provide secondary and micronutrients which are only needed in very small quantities and improve the soil’s physical conditions [86,87] as confirmed in Table 5 and Figure 4 where treatments with 600 kg/ha NPK had a yield of 30 t/ha using Fischer’s least significant difference (F-LSD) at 5% probability due to the impacts of microbial metabolic processes related to plant nutrition in the biofertilizer. Nutrients contained in organic manures are released more slowly and are stored for a longer time in the soil, thereby ensuring a long residual effect [88]. A combined use will increase synchrony and reduce losses by converting inorganic N into the environmental problems that may arise from the use of sole manure and chemical fertilizer.

### Impacts of photosynthesis

The C₄ plants tend not to light saturate, have low photorespiration, high photosynthetic rates on a per unit leaf area basis and hence are also nitrogen and water use efficient. Cassava has normally been considered to be a typical C₄ plant [91,92]. The photosynthetic rates reported by El Sharkawy [93] for cassava in field grown plants (40 μmol CO₂/m²/s) are high for a C₄ plant. Furthermore, wild species of Manihot have photosynthetic rates as high as 50 μmol CO₂/m²/s [93]. Work at CIAT suggested that cassava might be a C₃-C₄ intermediate [94]. Connor & Palta [95] found that cassava stomata closed in well-watered and stressed plants in the field at midday. Cassava stomata are extremely sensitive to the Vapor Pressure Deficit (VPD) between the leaf and the air [39]. A
whole series of trials have shown a relation between photosynthetic rate of individual leaves and the root yield of cassava under stressed and unstressed conditions. El Sharkawy et al. [96] found that photosynthetic rate measured in preliminary yield trials was correlated with root yield in subsequent independent yield trials.

Increased yield associated with increased photosynthetic rate, as expected, increases the nitrogen use efficiency [96], and also, presumably, water use efficiency. Furthermore, recently it has been shown that activity of PEP carboxylase, an enzyme associated with C4 photosynthesis, is correlated with photosynthetic rate and yield. Thus, it might be easier to screen parent materials for crosses for their PEP carboxylase activity in breeding programs. The dry matter content of cassava roots ranges from about 25% to up to 40%. Hence, it is more cost effective to produce high dry matter products [97]. The dry matter content of cassava roots is a varietal characteristic. The drop in dry matter content is probably due to mobilization of starch reserves in the roots to support the flush of new leaves [98]. Consequently, it is not uncommon for stressed plants of vigorous varieties to produce more roots than unstressed plants [95]. Cassava is capable of exploiting the available water to a depth of 2m [99]. In a fairly typical soil, the cassava plant can extract the equivalent of 160 mm of soil water during a drought period [93]. Rather than growing continuously and ending up with low levels of nutrients in plant tissue, the cassava plant tends to reduce its growth according to the available nutrients (Plates 3 & 4). This is particularly true in the case of nitrogen [100] but appears to be less so in the case of phosphorous [101] and potassium. Nevertheless, on extremely low phosphorous soils, well managed to maintain effective strains of mycorrhiza, cassava performs very well [102]. Since nutrient removal is mainly a function of yield, it is more practical to calculate nutrient removal per ton of fresh roots harvested. Howeler [73] at harvest reported only the crop removes mainly K, less N and very little P.

Plate 3: Control field Cassava experiment.

Plate 4: Treated field cassava experiment using Biofertilizer.

Cassava Products and Markets

Cassava roots and leaves (or young plant tops) have multiple end-uses, including for direct human consumption of fresh roots and leaves (after boiling), on-farm animal feeding, commercial production of animal feed, and production of starch or starch derivatives, Figure 5. Most of the chips and pellets are used for production of alcohol and animal feed, respectively. Besides dry chips, China also imported nearly 1.2 million tons of cassava starch, much of which is being converted to modified starch within China. China itself was producing about 4.5 million tons of fresh roots in 2009 [103], while importing the equivalent of 18.7 million tons of fresh roots. In China, cassava starch production was about 900,000 tons in 2007, while an additional 500,000 tons of cassava starch were imported [104].

Starch-based sweeteners and Hydrogenated sweeteners

Cassava starch can be used for the production of many types of sweeteners after hydrolyzation by either acids or enzymes, or both. These sweeteners include maltose, glucose syrup, glucose and fructose, which can be further processed into various oligosaccharides [105]. Hydrogenated sweeteners are produced by treating starch with hydrogen gas in high-pressure tanks, using a special catalyst and ion-exchange resins. These include sorbitol,
mannitol and maltol. Sorbitol is used mainly for the production of vitamin C and as a moisture conditioner in toothpastes [106].

**Organic acids and Monosodium glutamate (MSG) and lysine**

Organic acids made from cassava starch include citric acid, acetic acid, lactic acid and citric acid, which are used in the food industry as well as for the production of plastics, synthetic resins, rubber products etc. Lactic acid is produced by the fermentation of starch with Lactobacillus amylovorus [107]. MSG is a well-known flavor-enhancing agent used in many Asian kitchens. It is made through the microbial fermentation of starch or sugar (molasses) in the presence of ammonium salts. Lysine is an important amino acid used as a supplement in animal feed, especially for pigs.

**Degradable plastics and Ethanol**

Various types of starches are being used for the production of bio- or photo-degradable plastics, either by mixing starch or modified starch with polyvinyl hydrocarbons, or by polymerization of starch, which is then blended with various other polymers [108]. The use of cassava starch for these processes still requires much research. Cassava as the feedstock for the production of ethanol; this includes drinking alcohol, industrial alcohol and fuel-alcohol. The latter is also called "dehydrated" or "anhydrous" ethanol and must be 99.5% pure ethanol.

**Table 6: Combine Organic/Chemical Fertilizers Application to Cassava Crop.**

| N/B | Treatment Materials | Control Yield/Ton | Treatment/ Yield/ton | Impacts on Crop Component | References |
|-----|---------------------|-------------------|----------------------|---------------------------|------------|
| 1   | Mineral fertilizer/Poultry manure | 60 t/ha | | Chemical fertilizers supplied the bulk of the macronutrients | CSIR-AGRA [68] |
| 2   | OBD-Biofertilizer | | | | |
|     | Urea/NPK 15:15:15 /OBD-Biofertilizer | | | | |
|     | 300 kg/ha NPK 15:15:15 + 1.0 t/ha OBD-Biofertilizer | 12 t/ha | 20.5 t/ha | Reduce losses by converting inorganic N into organic forms | Kramer, et al. [89] |
|     | 300 kg/ha NPK 15:15:15 + 2.0 t/ha OBD-Biofertilizer | 22 t/ha | | Improves the microbial properties of the soil | Belay et al., 2001 |
|     | 300 kg/ha NPK 15:15:15 + 3.0 t/ha OBD-Biofertilizer | 30 t/ha | | Sustain maximum crop productivity and profitability | Ayeni [90] |
|     | 300 kg/ha NPK 15:15:15 + 4.0 t/ha OBD-Biofertilizer | 31.2 t/ha | | Minimizing environmental impact from nutrient use | |
|     | 600 kg/ha NPK (15:15:15) | 35.6 t/ha | | | |

In Figure 2, excitation (biofertilizer application) to the cassava cultivation soil system is functional characteristics of the impacts on rhizosphere considered for diagnostic tests of ‘Biofertilizer rhizosphere holistic soil function’. Where indicated by grey (Paper 1) circles while the connecting processes are in white (Paper 2) and Paper 3 (sustainable soil management) [143-164].

**Conclusion**

Cassava production has continued to expand throughout the lowland tropics, mainly on the less-fertile, poor-quality agricultural lands. The quality of farm-inputs affects the quality of cassava yield and soil health as confirmed in the report of Yomeni et al. [109]. Poor quality planting material is often associated with marginal growth and productivity of cassava. Microbial inoculants have paramount significance in integrated nutrient management systems to sustain agricultural productivity and healthy environment [110]. Application of 300 kg/ha NPK 15:15:15 + 3t/ha OBD-Biofertilizer gave high cassava root yield and is recommended for cassava production on soils having similar characteristics as the soil of the field experimental site (Table 5). Environmental stresses are becoming a major problem and productivity is declining at an unprecedented rate. Biofertilizers can help solve the problem of feeding an increasing global population at a time when agriculture is facing various environmental stresses [111-128]. It is important to realize the useful aspects of biofertilizers and implement its application to modern agricultural practices (Table 6). The use of farmer participatory research (FPR) and extension (FPE) methodologies, in which farmers become directly involved in the testing, selection and dissemination of new technologies will enhance crop development [129-142].

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Conflict of Interest

No conflict of interest.

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