Assessment of Trace Elements Supply in Canned Tuna Fish Commercialized for Human Consumption in Brazil

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Abstract: This study evaluates the elemental content in 4 types of canned tuna fish groups, each with 4 brands that are commercialized for human consumption in Brazil. The results are based on trace elements in canned tuna fish quantified by ICP OES and a comparison to limit levels set by the FAO/WHO. We also checked the carcinogenic risk (CR), non-carcinogenic risk (Hazard Index (HI) and Hazard Quotient (HQ)), and pollution index (PI) for the studied canned tuna samples. As and Se concentrations in all groups are above the intake values set by FAO/WHO considering specific groups. The carcinogenic risk values for arsenic (As) in groups are considerably unacceptable (≥10⁻⁴). Hazard quotients (HQ) were >1 for As in all groups, while no sample was below 1 for HI. The pollution index (PI) results show that the main canned tuna fish contaminant is aluminum, then selenium and arsenic, respectively. Only half of the samples did not present elemental contaminant levels. All studied brands of canned tuna presented elemental concentrations that could pose a health risk to human consumption, that could be from CR, HQ, HI, or PI. The contaminant levels are alarming and should raise a red flag for the intake of these products, especially a long-term one. These results urge the authorities to supervise and enforce better practices for this type of food, protecting their population from health hazards.

Keywords: metalloid; (non)metal; minerals; macro- and microelements; health risk; pollutant; processed fish

1. Introduction

Worldwide increasing natural resources usage, including land, leads to the spread of several heavy metals and metalloids from modern agriculture processes and motorized vehicle pollution [1]. The contamination of the soil, water, and river basin is highly associated with the massive food production in agriculture and the increased industry process of pesticides and fertilizers production and use; plus, residential source sewage/sludge pollution, intensive mining industries, and natural sources of heavy metals in soil and food crops for animal and human intake [2–5].

Wind and water flow carry several chemical elements to lakes, streams, and rivers during drought and rain season, including heavy metals, metals, and metalloids [6]. These watercourses and winds are directly linked to seas and oceans metals and metalloids accumulation, becoming pollutants and contaminant matter to plankton and animals, and fishes used for human food [7].

Fish consumption is recommended worldwide, as fish are a source of macro- and micro-elements, vitamins B12 and D [8], protein [9], and omega-3 polyunsaturated fatty acids (eicosapentaenoic and docosahexaenoic acids), which increase the health benefits for...
humans, lowering obesity, weight gain, body mass index, insulin resistance, type 2 diabetes mellitus, inflammatory bowel diseases, and maintain harmony gut microbiota balance [10]. The mentioned benefits boosted worldwide fish consumption to 21 kg/person/year [11].

Although the canning procedure is suitable for preserving food, this does not mean that such food is not subject to chemical elements contamination. Some countries have conducted several studies on canned fish commercialized in local markets, quantifying and monitoring heavy metals and metalloids concentration, and guaranteeing food safety and public health for consumers [12–23]. Studies have shown that heavy metals, metals, and metalloids in some canned fish samples [12–18,20–24] are a factor of concern; some chemical elements are toxic and can harm health. Arsenic exposure is associated with liver, lung, prostate, and bladder cancers [25]. Other chemical elements such as Al are toxic [26] and are related to diseases such as autism spectrum disorder and multiple sclerosis [27]. Zinc plays a role in numerous biochemical processes in humans and animals; however, overexposure to zinc is related to toxic effects [28]. Excessive iron intake can lead to free radicals linked to oxidative stress, mood disorders, and other diseases [29]. Selenium is nutritionally essential for humans, while it becomes toxic in high doses [30]. An excess of barium can occur, provoking kidney diseases, neurological impairments, cardiovascular, mental, and metabolic disorders [31]. The presence of trace metals in marine waters may happen naturally (from the erosion course of rocks and spoils) [32], or by human activities, from the industrialization and urbanization process [33].

Thus, elemental content surveillance of canned fish is necessary, and should be carried out periodically in several countries.

Based on data published by Lima et al. [24], the purpose of this study is to conduct a human health risk assessment due to the ingestion of chemical elements such as aluminum (Al), arsenic (As), barium (Ba), calcium (Ca), copper (Cu), iron (Fe), selenium (Se) and zinc (Zn) in 4 types of canned tuna fish: (i) natural grated tuna (NGT), (ii) oil grated tuna (OGT), (iii) solid natural tuna (SNT), and (iv) solid tuna in oil (STO). There are four brands of the company that sell these 4 types of condiments in canned tuna in Brazil.

The study showed that carcinogenic risk values for As are above acceptable values. In addition, Al, Se, and As are the principal pollutants with samples achieving pollution index (PI) values above 1.

2. Materials and Methods

2.1. Tuna Fish Samples Acquisition and Preparation

A total of 48 samples of canned tuna fish of different brands were purchased in supermarkets in Campo Grande, Mato Grosso do Sul, Brazil. In this study, two main types of canned tuna fish were considered: grated and solid. Each of the four brands (G, C, O, and P) sells 4 types of canned tuna fish: (i) natural grated tuna (NGT), (ii) oil grated tuna (OGT), (iii) solid natural tuna (SNT), and (iv) solid tuna in oil (STO). The natural samples are respective to those in brine rather than oil. A detailed description of analyzed brands of companies and types of canned tuna fish is presented in the work of Lima et al. (2021) [24].

2.2. Microwave-Assisted Digestion Procedure, Inductively Coupled Plasma–Optical Emission Spectrometry (ICP OES) Elemental Analysis, and Calibration Curves

Procedures were taken as described by Lima et al. [24]. About 400 mg of the canned tuna samples of each type and from different companies were accurately weighed in a Teflon digestion vessel. Next, 1 mL of nitric acid and 3 mL of hydrogen peroxide were added. Digestion of samples was carried out in a microwave digestion system. All the digestion analyses steps were conducted in triplicate.

The procedure for quantifying Al, As, Ba, Ca, Cu, Fe, Se, and Zn in canned tuna using ICP OES, analytical calibration curve, the limit of detection (LOD), limit of quantification (LOQ), and correlation coefficient (R²) are described by Lima et al. (2021) [24]. The LOD was
calculated as three times the standard deviation of the blank signal (B) expressed in concentration divided by the slope of the analytical curve (AC): LOD = 3*B/AC, and the LOQ was obtained as ten times the standard deviation of the blank divided by the slope of the analytical curve: LOQ = 10*B/AC [24,40,41].

An addition/recovery test for the elements under study was conducted in a tuna fish sample by spiking 0.5 mg/L of each analyte. The method had a recovery interval of 80–110% for the spike 0.5 mg/L, which is between 80–120% of the previously established limit proposed by the Union of Pure and Applied Chemistry (IUPAC) and Association of Official Analytical Chemists (AOAC) [40,42–44].

2.3. Human Health Risk Assessment and Pollution Index

Carcinogenic risk estimates represent the probability that an individual will develop cancer over a lifetime due to a specific exposure to a carcinogenic chemical; that is, exposure to daily doses over the years of life. Carcinogenic risk (CR) is calculated by the following equation:

\[ \text{Carcinogenic Risk} = \text{CDI} \times \text{SF} \]  

(1)

CDI is the chronic daily intake dose of carcinogenic elements (mg/kg/day), and carcinogenic risk (CR) is quantified by the chemical element cancer slope factor (SF). The SF results from the application of a low-dose extrapolation procedure, presented as “mg/kg/day” [45]. Their units are the inverse of the lifetime average daily dose CDI, because the ratio is a probability (i.e., unitless). The SF of As is 1.5/mg/kg/day. The cancer risk is a sum of individual carcinogenic elements in different exposure pathways within total cancer (R). According to the United States Environmental Protection Agency (US EPA) [46], the value of acceptable cancer risk ranges from \(10^{-6}\) to \(10^{-4}\), while values \(>10^{-4}\) are considered unacceptable.

The human health risk of heavy metal intake was evaluated based on the chronic daily intake dose (CDI) for a chemical contaminant in the tuna fish over the exposure period and the fish intake quantity. CDI (mg/kg/day) was calculated using the following Equation (2):

\[ \text{CDI} = \frac{C_{\text{tuna fish}} \times IR_{\text{tuna fish}} \times EF \times ED}{BW \times AT} \]  

(2)

where CDI is the chronic daily tuna fish intake dose; \(C_{\text{tuna fish}}\) is the concentration of the chemical elements present in samples (mg/kg) sold by companies (G, C, O, and P) in different types of canned fish (NGT, OGT, SNT, and STO) [24]; \(IR_{\text{tuna fish}}\) is the ingestion rate (130 g/day); EF is the exposure frequency (3 times per week = 156 days/year) as recommended by FDA and EPA [42,45]; ED is the exposure duration (life exposure = 8, 18 and 30 years); BW is the body weight (kg), and we considered 26 kg for an 8-year-old; 62 kg for an 18-year-old, and 70 kg for a 30-year-old. The AT is the average time (AT = ED × 365 days/year). The average daily fish consumption was set as 130 g/day, which is near the recommended amount [47,48], and it is the portion of choice once it is the content of one canned tuna.

The non-carcinogenic health risk to humans by the intake of heavy metal-contaminated fish was obtained using a hazard quotient (HQ), which is a ratio of CDI and chronic oral reference dose (RfD), determined by the following Equation (3):

\[ HQ = \frac{\text{CDI}}{\text{RfD}} \]  

(3)

The RfD values for the risk calculation were established by the Joint Food and Agriculture Organization/World Health Organization (FAO/WHO) Expert Committee on Food Additives “Food safety and quality: Summary reports”, [37] and the regional screening levels for use by risk assessors in site screening for chemical contaminants for the assessment of human health, which the RfD values for the elements were established: Al = 1.0 mg/kg/day; As = 0.0003 mg/kg/day; Ba = 0.2 mg/kg/day; Ca = not available; Cu = 0.04 mg/kg/day; Fe = 0.7 mg/kg/day; Se = 0.005 mg/kg/day; Zn = 0.3 mg/kg/day [49,50]. As show in
Equation (3), a toxic risk is considered to occur if HQ > 1, whereas HQ < 1 represents a negligible hazard (adverse non-carcinogenic effects) [51].

Another critical concept related to the HQ is the HI. It is the sum of the risk quotients for simultaneous exposure to two or more metals; that is, 

\[ HI = HQ_{Al} + HQ_{As} + HQ_{Ba} + HQ_{Cu} + HQ_{Fe} + HQ_{Se}. \]

If HI < 1, canned tuna fish consumption is safe, while in the case of HI > 1, canned tuna fish consumption may pose a health risk [51].

The pollution index (PI) was calculated following Equation (4) adapted by Adebiyi et al. [47]. For PI > 1 values, there is an assumption of contaminated samples, whereas PI < 1 stand for non-contaminated samples.

\[ PI = \frac{Cn}{AC} \]  

(4)

where Cn—chemical element concentration in tuna fish and AC—acceptable values limit in food: Al = 1 μg/g [52]; Fe = 43 μg/g in food [53]; Zn = 50 μg/g [53]; As = 2 μg/g [54]; Cu = 40 μg/g [53], and Pb = 0.4 μg/g [55]. For Se, we adopted the Tolerable Upper Intake Level (UL), since there is currently no permissible limit for the element in fish.

2.4. Statistical Analysis

The data were analyzed by two-way ANOVA using the GraphPad Prism 8 software version 8.0 for Windows (GraphPad Software, San Diego, CA, USA). The considered sources of variations were sample brands, and structure (grated or solid), and solvent. The significance of the differences between the means for the individual trace element was considered at \( p < 0.05 \).

3. Results

The results for all canned tuna fish were represented by two sub-sections. Section 3.1 presents data on the concentration of the trace elements quantified in canned tuna fish purchased in Brazil. Section 3.2 describes the results of the CR, HQ, HI, and PI of the trace elements, based on ingestion of 130 g/day of canned tuna fish for individuals aged 8, 18, and 30 years old.

3.1. Canned Tuna Content and Intake Limits

In this study, the concentrations of Al, As, Ba, Ca, Cu, Fe, Se, and Zn in NGT, OGT, SNT, and STO samples in units of mg/kg were converted to mg per 130 g, once this is the canned tuna net weight sold in Brazilian markets. Table 1 presents the elemental concentration for each type of canned tuna and its brand. Al, Ba, Ca, Se, and Cu contents in some canned foods are below the detection limit (<LOD), while Cd and Pb were below the LOD for all samples.

The level of Al in NGT, SNT, and STO groups ranged from 0.00065 ± 1.3 × 10^{-5} to 6.1 ± 0.4 mg/130 g (Table 1). The structure and solvent (how the tuna is presented in the can: either solid or grated; and in which solvent it is stored: either brine or oil) contributed 79.08% to the total aluminum variation among samples (\( p < 0.0001 \)).

The arsenic (As) concentration variations in NGT, OGT, SNT, and STO groups were between 0.2 ± 0.003 and 0.3 ± 0.01 mg/130 g (Table 1), and did not statistically differ among samples.

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The interaction between these factors represented 38.38% of the variation in barium content (\( p = 0.0029 \)).
Table 1. Concentrations of metals and metalloids in canned tuna (NGT, OGT, SNT, and STO) of the company’s four brands (G, C, O, and P) compared with FAO/WHO limits.

| Element | Natural Grated Tuna—NGT (mg/130 g) | Oil Grated Tuna—OGT (mg/130 g) | Solid Natural Tuna—SNT (mg/130 g) | Solid Oil Tuna—STO (mg/130 g) | Reference for 70 kg Adults (mg/day) |
|---------|----------------------------------|---------------------------------|----------------------------------|-----------------------------|-----------------------------------|
|         | NGT-G | NGT-C | NGT-O | NGT-P | OGT-G | OGT-C | OGT-O | OGT-P | SNT-G | SNT-C | SNT-O | SNT-P | STO-G | STO-C | STO-O | STO-P | |
| Al      | 1.9 ± 0.4 | 1.3 ± 0.003 | 2.4 ± 0.4 | 1.7 ± 0.3 | <LOD | <LOD | <LOD | <LOD | 2.6 ± 0.03 | 2.7 ± 0.4 | 6.1 ± 0.4 | 3.3 ± 0.5 | <LOD | <LOD | <LOD | <LOD | 0.00065 ± 1.3 × 10⁻⁶ | 19.95 [37] |
| As      | 0.3 ± 0.01 | 0.2 ± 0.007 | 0.2 ± 0.01 | 0.2 ± 0.01 | 0.2 ± 0.006 | 0.2 ± 0.03 | 0.2 ± 0.002 | 0.2 ± 0.01 | 0.2 ± 0.003 | 0.2 ± 0.006 | 0.2 ± 0.001 | 0.2 ± 0.01 | 0.2 ± 0.001 | 0.2 ± 0.002 | 0.147 [39] |
| Ba      | <LOD | <LOD | <LOD | <LOD | <LOD | <LOD | 0.004 ± 0.003 | <LOD | 15 ± 4 | 10 ± 4.9 | <LOD | 0.008 ± 0.003 | 0.03 ± 0.007 | 0.08 ± 0.03 | 0.02 ± 0.004 | 1.4 [38] |
| Ca      | 0.07 ± 0.03 | 0.01 ± 0.003 | 0.03 ± 0.003 | 0.03 ± 0.006 | <LOD | <LOD | <LOD | <LOD | 5.4 ± 0.2 | 5.2 ± 0.3 | 5.8 ± 0.1 | 6.2 ± 0.04 | 5 ± 0.2 | 8.2 ± 0.4 | 9.2 ± 2.6 | 8.9 ± 0.4 | 2000–2500 [36] |
| Cu      | 0.08 ± 0.004 | 0.07 ± 0.003 | 0.09 ± 0.01 | 0.04 ± 0.01 | 0.06 ± 0.002 | 0.03 ± 0.005 | 0.04 ± 0.006 | 0.01 ± 0.013 | <LOD | 0.008 ± 0.002 | <LOD | 0.02 ± 0.004 | 0.04 ± 0.004 | 0.03 ± 0.005 | 0.03 ± 0.004 | 3 [35] |
| Fe      | 3.9 ± 0.5 | 3.3 ± 0.03 | 3.3 ± 0.06 | 4 ± 0.2 | 2.3 ± 0.6 | 1.4 ± 0.1 | 1.8 ± 0.7 | 2.7 ± 0.4 | 1 ± 0.059 | 2 ± 0.04 | 1 ± 0.06 | 2.2 ± 0.2 | 1.2 ± 0.02 | 1.1 ± 0.04 | 1 ± 0.009 | 1.4 ± 0.106 | 17 [34] |
| Se      | 0.3 ± 0.02 | 0.2 ± 0.003 | 0.3 ± 0.005 | 0.2 ± 0.01 | 0.2 ± 0.01 | 0.3 ± 0.01 | 0.3 ± 0.005 | 0.3 ± 0.03 | 0.2 ± 0.001 | 0.2 ± 0.01 | 0.2 ± 0.001 | 0.2 ± 0.005 | 0.2 ± 0.01 | 0.2 ± 0.005 | 0.26 ± 0.01 | 0.3 ± 0.02 | 0.04 [36] |
| Zn      | 0.05 ± 0.005 | 0.02 ± 0.0003 | 0.04 ± 0.001 | 0.03 ± 0.001 | 0.03 ± 0.001 | 0.03 ± 0.004 | 0.05 ± 0.006 | 4 × 10⁻⁶ | 0.03 | 0.003 | 0.01 | 0.02 | 0.01 | 0.02 | 0.002 | 0.013 | 70 [35] |

Note: <LOD—analyte concentration below the limit of detection; ND = not determined.
Calcium contents ranged from 0.01 ± 0.003 to 9.2 ± 2.6 mg/130 g (Table 1). Structure and solvent were the main sources (95.79%; p < 0.0001) of calcium variations in samples, with high calcium content in solid samples only.

Copper concentrations in samples varied from 0.008 ± 0.002 to 0.1 ± 0.01 mg/130 g (Table 1). Solvent and structure were critical to copper variation in samples, accounting for 61.4% (p < 0.0001), followed by the interaction with the brands (28.74%; p = 0.0001); grated samples and samples in oil registered higher copper amounts. The brand alone was responsible for only 3.88% of variations (p = 0.04).

Iron concentrations in canned tuna samples were between 1 ± 0.06 and 4 ± 0.2 mg/130 g (Table 1). The two-way ANOVA identified that variations in iron were due to structure and solvent (81.94%; p < 0.0001), brand (8.045%; p < 0.0001), and the interaction between these factors (7.525%; p = 0.018).

The concentration of Se in canned tuna samples ranged from 0.2 ± 0.005 to 0.3 ± 0.02 mg/130 g (Table 1). The variations in the amount of selenium mainly depend on the structure and solvent (53.01%; p < 0.0001), while the brand represents just 8.064% of the difference, and the interaction between factors is 37.17%.

Zinc (Zn) concentrations varied between 0.01 and 0.05 mg/130 g (Table 1). Zinc quantities varied according to structure and solvent (52.04%; p < 0.0001), followed by the interaction between factors (33.70%, p < 0.0001) and brand (13.43%; p < 0.0001).

Elements that are not essential displayed a considerable detectable amount in the samples (Figure 1).

The natural samples (brine—NGT and SNT), aluminum detection presented a substantial amount opposite to those in oil. In the same way, all samples showed the presence of arsenic, which is not only not essential, but also toxic.

### 3.2. Health Risk Assessment

Table 2 shows the CR calculated using Equation (1) for As quantified in NGT, OGT, SNT, and STO canned foods marketed by the four Brazilian companies (G, C, O, and P).

In the CDI, we considered life exposure, ED = 8, 18, 30 years, and the ingestion of canned tuna of 130 g/day. The carcinogenic risk values were as follows: NGT = 1.8 × 10^{-3}–6.2 × 10^{-3}, OGT = 1.3 × 10^{-3}–5.7 × 10^{-3}, SNT = 1.2 × 10^{-3}–5.4 × 10^{-3} and STO = 2.0 × 10^{-3}–6.0 × 10^{-3} for As. The acceptable values are from 10^{-6}–10^{-4}.

Hazard quotients and hazard index (HI) for Al, As, Ba, Cu, Fe, Se, and Zn for canned tuna fish consumption for males, females, and children are shown in Table 3.
Table 2. Life exposure (Age = 8, 18, 30 years), values of carcinogenic risk (CR) due to exposure of As in canned NGT, OGT, SNT, and STO from the four companies (G, C, O, and P), considering a daily intake of 130 g/day.

| Sample | Cancer Risk Arsenic (As) |
|--------|--------------------------|
|        | 8 Years Old  | 18 Years Old  | 30 Years Old  |
| NGT-G  | 0.0062       | 0.0026       | 0.0023       |
| NGT-C  | 0.0051       | 0.0022       | 0.0019       |
| NGT-O  | 0.0053       | 0.0022       | 0.0020       |
| NGT-P  | 0.0047       | 0.0020       | 0.0018       |
| OGT-G  | 0.0057       | 0.0024       | 0.0013       |
| OGT-C  | 0.0054       | 0.0023       | 0.0020       |
| OGT-O  | 0.0044       | 0.0019       | 0.0016       |
| OGT-P  | 0.0052       | 0.0022       | 0.0019       |
| SNT-G  | 0.0043       | 0.0018       | 0.0016       |
| SNT-C  | 0.0049       | 0.0021       | 0.0018       |
| SNT-O  | 0.0041       | 0.0017       | 0.0015       |
| SNT-P  | 0.0054       | 0.0022       | 0.0012       |
| STO-G  | 0.0060       | 0.0025       | 0.0022       |
| STO-C  | 0.0054       | 0.0023       | 0.0020       |
| STO-O  | 0.0056       | 0.0024       | 0.0021       |
| STO-P  | 0.0060       | 0.0025       | 0.0022       |

In Table 3, the hazard quotients for Se in canned tuna fish NGT-C and Ba in SNT-C are above one for 8-year-olds. All four brands’ hazard quotients (HQ) for arsenic surpass 1 for all studied population groups. When HQ > 1, there is a toxic risk to be considered.

According to the calculated non-carcinogenic hazard index (HI), which is the sum of the risk quotients for simultaneous exposure to metals, that is, HI = HQ_{Al} + HQ_{As} + HQ_{Ba} + HQ_{Cu} + HQ_{Fe} + HQ_{Se} + HQ_{Zn} in each sample, there is a HI value superior to 1 for all studied population groups.

The pollution index (PI) of the trace elements calculated in this study is presented in Figure 2.
Table 3. Hazard quotient (HQ) and Hazard index (HI) due to the ingestion of canned tuna fish commercialized in Brazil for individuals aged 8, 18, and 30 years old.

| Age (Years) | Samples | HQ | HI | Samples | HQ | HI |
|-------------|---------|----|----|---------|----|----|
|             |         | Al | As | Ba | Cu | Fe | Se | Zn |         | Al | As | Ba | Cu | Fe | Se | Zn |         |
| 8           | NGT-G   | 0.03 | 13.84 | 0.00 | 0.03 | 0.09 | 1.11 | 0.003 | 15.11 | SNT-G | 0.04 | 9.47 | 0.00 | 0.00 | 0.02 | 0.54 | 0.0007 | 10.08 |
|             | NGT-C   | 0.02 | 11.48 | 0.00 | 0.03 | 0.08 | 0.74 | 0.001 | 12.35 | SNT-C | 0.04 | 10.96 | 1.25 | 0.003 | 0.05 | 0.62 | 0.001 | 11.91 |
|             | NGT-O   | 0.04 | 11.70 | 0.00 | 0.04 | 0.08 | 0.85 | 0.002 | 12.75 | SNT-O | 0.10 | 9.10 | 0.80 | 0.00 | 0.03 | 0.63 | 0.0006 | 10.64 |
|             | NGT-P   | 0.03 | 10.50 | 0.00 | 0.02 | 0.09 | 0.81 | 0.002 | 11.44 | SNT-P | 0.05 | 11.90 | 0.00 | 0.007 | 0.05 | 0.54 | 0.0009 | 12.54 |
|             | OGT-G   | 0.00 | 12.70 | 0.00 | 0.02 | 0.05 | 0.74 | 0.002 | 13.52 | STO-G | 0.00 | 13.36 | 0.0006 | 0.018 | 0.03 | 0.66 | 0.001 | 14.05 |
|             | OGT-C   | 0.00 | 12.06 | 0.00 | 0.01 | 0.03 | 0.83 | 0.0008 | 12.93 | STO-C | 0.00 | 11.99 | 0.003 | 0.01 | 0.03 | 0.85 | 0.0008 | 12.88 |
|             | OGT-O   | 0.00 | 9.82 | 0.0003 | 0.02 | 0.04 | 0.87 | 0.001 | 10.75 | STO-O | 0.00 | 12.51 | 0.001 | 0.01 | 0.03 | 0.85 | 0.0009 | 13.40 |
|             | OGT-P   | 0.00 | 11.47 | 0.0001 | 0.04 | 0.07 | 0.84 | 0.003 | 12.41 | STO-P | 0.00001 | 13.23 | 0.001 | 0.01 | 0.03 | 0.88 | 0.0007 | 14.19 |
| 18          | NGT-G   | 0.01 | 5.80 | 0.00 | 0.01 | 0.04 | 0.47 | 0.001 | 6.33 | SNT-G | 0.02 | 3.97 | 0.00 | 0.00 | 0.01 | 0.23 | 0.0003 | 4.23 |
|             | NGT-C   | 0.01 | 4.82 | 0.00 | 0.01 | 0.03 | 0.31 | 0.0005 | 5.18 | SNT-C | 0.02 | 4.60 | 0.52 | 0.001 | 0.02 | 0.26 | 0.0005 | 5.41 |
|             | NGT-O   | 0.02 | 4.90 | 0.00 | 0.01 | 0.03 | 0.36 | 0.0008 | 5.30 | SNT-O | 0.04 | 3.82 | 0.33 | 0.00 | 0.01 | 0.26 | 0.0003 | 4.46 |
|             | NGT-P   | 0.01 | 4.40 | 0.00 | 0.006 | 0.04 | 0.34 | 0.0007 | 4.80 | SNT-P | 0.02 | 4.99 | 0.00 | 0.003 | 0.021 | 0.23 | 0.0004 | 5.26 |
| 30          | OGT-G   | 0.00 | 5.33 | 0.00 | 0.01 | 0.02 | 0.31 | 0.0007 | 5.67 | STO-G | 0.00 | 5.60 | 0.0003 | 0.008 | 0.012 | 0.28 | 0.0005 | 5.90 |
|             | OGT-C   | 0.00 | 5.06 | 0.00 | 0.01 | 0.01 | 0.35 | 0.0003 | 5.42 | STO-C | 0.00 | 5.03 | 0.0010 | 0.004 | 0.01 | 0.36 | 0.0003 | 5.40 |
|             | OGT-O   | 0.00 | 4.12 | 0.0001 | 0.01 | 0.02 | 0.37 | 0.0006 | 4.50 | STO-O | 0.00 | 5.25 | 0.0006 | 0.004 | 0.01 | 0.36 | 0.0004 | 5.60 |
|             | OGT-P   | 0.00 | 4.81 | 0.00006 | 0.02 | 0.027 | 0.35 | 0.001 | 5.21 | STO-P | 0.000004 | 5.87 | 0.0006 | 0.0047 | 0.0140 | 0.37 | 0.0003 | 5.95 |
|             | NGT-G   | 0.01 | 5.14 | 0.00 | 0.01 | 0.03 | 0.41 | 0.0011 | 5.61 | SNT-G | 0.01 | 3.51 | 0.00 | 0.00 | 0.01 | 0.20 | 0.0003 | 3.74 |
|             | NGT-C   | 0.01 | 4.27 | 0.00 | 0.01 | 0.03 | 0.27 | 0.0004 | 4.38 | SNT-C | 0.02 | 4.07 | 0.46 | 0.0013 | 0.02 | 0.23 | 0.0004 | 4.80 |
|             | NGT-O   | 0.01 | 4.34 | 0.00 | 0.01 | 0.03 | 0.31 | 0.0007 | 4.71 | SNT-O | 0.04 | 3.38 | 0.29 | 0.00 | 0.01 | 0.23 | 0.0002 | 3.96 |
|             | NGT-P   | 0.01 | 3.90 | 0.00 | 0.01 | 0.03 | 0.31 | 0.0007 | 4.24 | SNT-P | 0.01 | 2.65 | 0.00 | 0.002 | 0.01 | 0.12 | 0.0002 | 2.79 |
|             | OGT-G   | 0.00 | 2.83 | 0.00 | 0.01 | 0.01 | 0.17 | 0.0004 | 3.01 | STO-G | 0.00 | 4.96 | 0.0002 | 0.007 | 0.01 | 0.24 | 0.0004 | 5.22 |
|             | OGT-C   | 0.00 | 4.48 | 0.0004 | 0.01 | 0.03 | 0.31 | 0.0003 | 4.80 | STO-C | 0.00 | 4.45 | 0.0009 | 0.003 | 0.01 | 0.32 | 0.0003 | 4.78 |
|             | OGT-O   | 0.00 | 3.65 | 0.00012 | 0.006 | 0.02 | 0.32 | 0.0005 | 3.99 | STO-O | 0.00 | 4.65 | 0.0005 | 0.004 | 0.01 | 0.32 | 0.0003 | 4.98 |
|             | OGT-P   | 0.00 | 4.26 | 0.00005 | 0.01 | 0.02 | 0.31 | 0.001 | 4.61 | STO-P | 0.000004 | 4.93 | 0.0005 | 0.004 | 0.01 | 0.32 | 0.0003 | 4.65 |
Figure 2 shows that aluminum is the principal pollutant element, most quantified in canned tuna fish, with a PI value reaching 47.

Other elements such as selenium and arsenic presented PI > 1 for a few samples (Figure 2). Six samples—OGT-G, OGT-C, OGT-P, STO-G, STO-C, STO-O—were not contaminated by chemical elements.

4. Discussion

4.1. Canned Tuna Content and Intake Limits

The Al concentrations (Table 1) are higher than the content reported for canned tuna fish in Lebanon (0.62 mg/130 g) [13] and Turkey (0.70 mg/130 g) [56].

The aluminum consumption ranges from 21–69 mg/week for children (30 kg) and 14–105 mg/week for adults (70 kg) [57]. Aluminum exposure from foods can pose a higher risk to children, considering their body weight and the threat of achieving the threshold set by WHO of 2 mg/kg/week [53].

Aluminum is not an essential element for life, and is commonly considered toxic to humans; however, its toxicity depends on the route of exposure and solubility. This element accumulates in various body parts like tissues, such as the brain, bones, kidneys, and liver. Long-term exposure to low Al levels leads to toxic effects (Klotz et al., 2017 [26]), as well as prolonged exposure to low levels of aluminum leading to changes associated with brain aging and neurodegeneration [58]. In fact, the risk of consuming food with a high amount of Al is associated with Alzheimer’s diseases, Parkinson’s disease [27], bone disorder (competing with calcium and phosphate), kidney dysfunctions, anemia, gut dysfunctions, cytotoxic and neurotoxic, and others [57,59].

The arsenic (As) concentrations found in our results indicate that the arsenic is close to the levels reported for canned tuna in Iran (0.18 mg/130 g) [18] and Galicia in Spain (0.14–0.3 mg/130 g) [60]. On the other hand, As values in Table 1 are lower than the tuna commercialized in São Paulo in Brazil (0.57–1.53 mg/130 g) [17]. The only element that did not differ among several samples was arsenic (p > 0.05), present in similar amounts.

Safe daily provisions determined by the UL for men, women, pregnant women, and children have not yet been established. However, there are no safe levels for arsenic intake once this value was withdrawn [53]; therefore, we used the previous determination of a weekly limit consumption of 0.015 mg/kg, according to other studies [60–62]. In this instance, the concentrations of arsenic in Table 1 are above the tolerable daily intake limit values for this element set by FAO/WHO for foods (0.0021 mg/kg/day, equivalent to 0.147 mg/day for 70 kg adults). According to the Nationwide Food Consumption Survey (NFCS) in the US, the estimated daily arsenic intake is 20 mg/day for 6-year-old children, 47 mg/day for 40–45-year-old men, and 37 mg/day for 40–45-year-old women. Seafood contributes to around 90% of arsenic consumption for children (2-year-old), while other essential food sources such as rice represent only about 4% of arsenic intake; thus, the values of arsenic in tuna (Table 1) are below the values set by NFCS for As in food [63].

Long-term exposure to arsenic from drinking water and food can cause cancer and bladder cancers [64]. In fact, the International Agency for Research on Cancer (IARC) has classified arsenic and arsenic compounds as carcinogenic to humans. This is based on sufficient evidence in humans that these compounds can cause: respiratory dysfunctions, gastrointestinal and neuro-cardiovascular diseases, anemia disorder, liver disorder, leucopenia, and thrombocytopenia, diabetes, cytotoxic, and genotoxicity effects [64–66].

Barium showed high concentrations only in samples of natural solid tuna in two brands (Figure 1). The Ba contents in OGT and STO are near the reported values for canned tuna fish (0.06 mg/130 g) purchased in Jordanian markets. The SNT samples had a higher barium content than those reported in this same study [12], as well as from samples from New Zealand and the United Kingdom (0.52–17.03 mg/130 g) [67].

So far, there is no UL set for barium. On the other hand, barium’s tolerable daily intake limit values are determined by FAO/WHO [38] 0.02 mg/kg/day in drinking water (equivalent to 1.4 mg/day for 70 kg adults). Besides, Montanari (2015) [68] described...
barium as a part of metal contaminants, where barium sulfate is used to manufacture cans and lids as an inorganic charge. Considering the threshold proposed by the Commission Regulation 10/2011 of 1 mg/kg for specific migration from packaging compounds to food [69], we can say that the values of Ba in NGT, STO, and OGT are below these limits, while SNT-C and SNT-O are above it.

Barium accumulation can occur from an occupational hazard or from the consumption of contaminated water and food. The average amount of ingestion worldwide and its geographic variation are unknown, due to a lack of research attention. The presence of the element can produce different effects, especially in cases of exposure, either in low or moderate doses. Information on the potential health effects of barium exposure is primarily from animal studies and reported to encompass renal, neurological, cardiovascular, mental, and metabolic diseases [31].

The quantified calcium levels in SNT and STO are near the concentrations reported for raw and steamed tuna fish (4%) in Spain [70], while NGT and OGT presented a lower calcium concentration. Structure and solvent were the main sources (95.79%; \( p < 0.0001 \)) of calcium variations in samples, with high calcium content in solid samples only.

Calcium is an essential element, and its adequate consumption is related to a minor risk of hypertensive disorders, lower blood pressure, lower cholesterol values, osteoporosis reduction, bone resorption, and others, while in a higher concentration, it is associated with renal stones formation and myocardial infarction in older humans [70].

The permissible limit of Ca set by UL is 2500 mg/day. Comparison between detected values and UL set value considers calcium concentration in the samples below the proposed limits, not posing a risk from this element intake [36].

As for copper concentrations in the samples, the OGT-P sample showed the highest concentration with an average copper concentration of 0.10 ± 0.01 mg/130 g; these values are close to the tuna obtained in Egypt (0.16 mg/130 g) [71] and higher than those found in Turkey (0.0026 mg/130 g) [20]. Solvent and structure were critical to copper variation in samples, accounting for 61.4% \( (p < 0.0001) \), followed by the interaction with the brands (28.74%; \( p = 0.0001 \)); grated samples and samples in oil registered higher copper amounts. The brand alone was responsible for only 3.88% of variations \( (p = 0.04) \).

There is no established UL for Cu at the moment. However, all Cu concentrations in canned tuna are below the tolerable daily intake limit value set by FAO/WHO for Cu (0.5 mg/kg/day, equivalent to 35 mg/day for 70 kg adults). Therefore, the ingestion of these canned tuna samples should be safe for consumption regarding copper content. Adequate copper consumption promotes health benefits, correlating with good functionality of the cardiovascular system, lower blood glucose, cholesterol, and lipid levels [72], cognitive, and is not associated with arthritis or cancer, cofactor, antioxidant effects, oxidative activity, absorption, and others [73]; whereas, elevated copper intake is related to mitochondrial dysfunction [74], liver damage, and Alzheimer’s disease [75]. Disturbances in the copper metabolism due to genetic conditions can result in copper deficiency (Menkes syndrome) and toxicity (Wilson’s disease) [76].

The iron content observed in this study agrees with the reported systematic review studies of canned tuna fish (1.71 mg/130 g) in Iran [77], and it was consistent with Fe content found in Mediterranean wild Atlantic Bluefin tuna (about 1.7 mg/130 g) [78]. The two-way ANOVA identified that variations in iron were due to structure and solvent (81.94%; \( p < 0.0001 \)), brand (8.045%; \( p < 0.0001 \)), and the interaction between these factors (7.525%, \( p = 0.018 \)).

The UL for males, females, and pregnancy is 45 mg/day of iron, while for children, it is 40 mg/day. In addition, the tolerable daily intake limit value established by the FAO/WHO [34] is 0.8 mg/kg/day, equivalent to 56 mg/day for 70 kg adults. Thus, the concentration of iron in canned tuna samples is unlikely to cause adverse health effects. Iron levels adequacy is correlated with maximal oxygen respiration and exercise performance, electron transport, hemoglobin synthesis, immunity, anemia prevention, pregnancy
development, deoxyribonucleic acid synthesis, gut microbiota health modulation, neurodevelopment, and others [79,80].

The Se contents observed in our study are according to those reported (0.0169–0.5850 mg/130 g) for canned fish marked in Iran [22], and it is consistent with Se content found in Atlantic Bluefin tuna from the Mediterranean Sea (about 0.143 mg/130 g) [81]. The variations in selenium amount mainly depend on the structure and solvent (53.01%; \( p < 0.0001 \)), while brand represents just 8.064% of the difference, and the interaction between factors is 37.17%.

Selenium detections are below the UL for the consumption of Se in male/female (0.4 mg/day) and children (0.15 mg/day). However, these values are higher than the value established by FAO/WHO for pregnant women (0.06 mg/day) [82]. Food toxicity of selenium in humans is rare; however, selenium (IV) is generally more toxic than selenium (VI). Excessive selenium intake can lead to selenosis, dermatitis, alopecia, elevated mortality rate, an enhanced risk for prostate cancer, and non-melanoma skin cancer [83]. On the other hand, sufficient selenium intake is associated with preventing and decreasing diabetes mellitus, cancers [22,84], improving male fertility, human neuropathies, and hepatic steatosis [85–87].

The concentration of Zn in all samples in Table 1 is below the study reported on the levels of selected heavy metals in canned tuna fish produced in Turkey for Zn (1.066–1.482 mg/130 g for Zn) [20]. Zinc quantities varied according to structure and solvent (52.04%; \( p < 0.0001 \)), followed by the interaction between factors (33.70%, \( p < 0.0001 \)) and brand (13.43%, \( p < 0.0001 \)).

The values of Zn set by UL for male/female and pregnant woman are 40 mg/day, and children 5 mg/day, while the values established by FAO/WHO [35] are 1 mg/kg/day, equivalent to 70 mg/day for 70 kg adults. All zinc values in samples are below the values set by UL and FAO/WHO. Thus, they should be safe for human consumption regarding this element. Zinc has a critical effect on homeostasis; immune function in oxidative stress. However, high doses of this element have toxic effects, making acute zinc intoxication a rare event [88].

Regarding the content of non-essential elements (Figure 1), a great difference was noted between the aluminum contents in samples preserved in brine (NGT and SNT) and in oil. In the natural samples (brine–NGT and SNT), aluminum detection presented a substantial amount opposite to those in oil. Aluminum can migrate from the cans to the food, and some conditions may facilitate this transference. Our results agree with the Stahl et al. (2017) findings [59], where aluminum migrated in a more critical matter in acidic water, water, and then oils.

Another significant point is the presence of arsenic in all samples, which is toxic and non-essential. The previous limit of consumption was withdrawn once they could no longer be considered safe [37]. Therefore, while the excess of some elements can pose a hazard for human intake, the mere presence of others can already raise a red flag.

In the following subsection, we will discuss the potential risks of the elemental content in canned samples. The potential risks of arsenic content in canned tuna were verified using the carcinogenic risk equation.

### 4.2. Health Risk Assessment

In the calculation of CDI, we considered life exposure, \( ED = 8, 18, 30 \) years, and ingestion of canned tuna of 130 g/day. The carcinogenic risk (CR) values obtained for 8-year-old children were higher in the NGT-G sample when compared to other companies (C, O, and P) and types of canned tuna fish (OGT, SNT, and STO). The carcinogenic risk values for NGT (1.8 \times 10^{-3}–6.2 \times 10^{-3}), OGT (1.3 \times 10^{-3}–5.7 \times 10^{-3}), SNT (1.2 \times 10^{-3}–5.4 \times 10^{-3}), and STO (2 \times 10^{-3}–6 \times 10^{-3}) for As are higher than the acceptable values \((10^{-6}–10^{-4})\); that is, all CR for arsenic are considerable unacceptable in these canned tuna samples. Arsenic is the main contaminant trace element that can be correlated with several cancer incidences among all heavy metals in the canned tuna fish samples. Furthermore, the total cancer risk
incidence can increase for those who consume the recommended 150 g/day [11] of canned tuna fish of types NGT, OGT, SNT, and STO, with an elevated risk for the youngest ones.

The risk assessment can provide information on non-cancerous health risks through the HQ factors (hazard quotient). The risk quotient for Se in NGT-C canned tuna and Ba in SNT-C (Table 3) is greater than one for 8-year-olds. The risk quotients (HQ) of all four brands for arsenic exceed 1 for all groups studied.

When HQ > 1, there is a toxic risk to be considered. All HQ values for Al, Fe, Cu, and Zn for all studied populations are below 1, indicating no potential health risk through canned tuna consumption for each element [51].

According to the calculated non-carcinogenic hazard index (HI), which is the sum of the risk quotients for simultaneous exposure to metals, that is, HI = HQ_{Al} + HQ_{As} + HQ_{Ba} + HQ_{Cu} + HQ_{Fe} + HQ_{Se} + HQ_{Zn} in each sample, there is a HI value superior to 1 for all studied population groups, indicating that canned tuna consumption from local markets can pose a risk for human health regarding metal and metalloid content. The HI in the Table 3 was higher for children than adults. The HI values in Table 3 are higher than those obtained in a study carried out in China with marine fish, with HI = 0.945 for adults. However, the results for children (HI = 8.556) published by Han et al. (2021) are within the values obtained for children in Table 3 [89]. Arsenic was the element that contributed the most to an elevated HI in most canned tuna samples (Table 3) and marine fish [89].

In general, the presence of the chemical elements Al, As, Se, Cu, Fe, and Ba in all types of canned fish can be explained by a higher occurrence of these metals and metalloids in various kinds of water springs, lakes, streams, rivers, seas, and oceans, mainly linked to anthropogenic activities [2–4]. In addition, Fe and Al in canned tuna samples may be explained by the migration of these elements from the can package into the fish [13]. Besides, tuna was recognized as a predator able to concentrate large amounts of heavy metals [90]. Therefore, with the consumption of 130 g/day of canned tuna fish, these chemical elements can be a non-carcinogenic hazard to human health.

The concentration of metals in fish relates to environmental pollution; since fish can accumulate pollutants from the surrounding area [91]. While fatty tissues like the liver accumulate in most metals, muscle tissue has lower contents [92,93].

The pollution index values of the samples are represented in Figure 2. For the interpretation of the pollution index results, it is known that a PI above 1 indicates sample contamination, and can be considered toxic [47]. Aluminum is the main pollutant, the most quantified in canned tuna, with a PI value of 47.

According to Hydes, besides anthropogenic factors, those responsible for the high concentration of aluminum in the sea are the clayey sediments that probably arise from biological activities [94], originating from the aluminum of atmospheric particles and by the balance in the sediment through the silicon generated by the death of aquatic organisms [95]. On the other hand, a high amount of Al in canned fish may be correlated with the interaction between aluminum foil particles in sauced food, which is potentially hazardous to several metals, skeleton diseases, cancers, and so on [58]. The use of internal coatings reduced the metal migration to the food (Al, Fe, Cd, Sn, and Pb) [96], but it still happens, mainly due to the discontinuous or not compact coating of the materials [97].

Other elements such as selenium and arsenic presented PI > 1 for a few samples (Figure 2). Sediments can be a significant source of selenium in fish and invertebrates. Toxic effect threshold levels for selenium in fish have been reported as 4 mg/kg (for whole fish) [98]. Therefore, high content calculated to Se in canned tuna fish can be explained by bioaccumulation in its several tissues from the water of environment, plankton, and other food types in the chain consumption [99]. In addition to anthropogenic factors, marine algae release arsenite into the seawater, which is toxic to marine phytoplankton, marine invertebrates, and fish. Tissues of marine invertebrates and fish contain high concentrations of arsenic. Therefore, marine arsenic represents a low risk to human consumers of fishery products [100]. The samples that did not indicate contamination by chemical elements were: OGT-G, OGT-C, OGT-P, STO-G, STO-C, STO-O.
Tuna fish and other fish species are critical in the human diet and represent a source of protein, but they may accumulate potentially toxic metals. According to the First World Ocean Assessment released in 2015, many ocean parts had been seriously degraded. The results obtained with Brazilian tuna, tuna marketed in Iran, Egypt, Turkey, Thailand, and other studies indirectly show that ocean degradation still remains and has increased over the years, affecting some fish species. As an alternative, the United Nations has proclaimed the Decade of Ocean Science for Sustainable Development (2021–2030) to support efforts to reverse the cycle of decline in ocean health [101].

5. Conclusions

All the canned tuna fish samples that we studied accumulated heavy metals. The majority of elements (Al, Fe, Ba, Ca, Cu, and Zn) in Brazilian canned tuna fish complied with the permissible limits by UL and FAO/WHO, while As in all samples, and Ba in SNT, is above these thresholds. The Se levels in our canned tuna fish are elevated for pregnant women consumption by the limit established by FAO/WHO.

The carcinogenic risk (CR) values due to the ingestion of Brazilian canned tuna fish obtained for 8-year-old children are higher than for adolescents and adults, related to the expected weight. The carcinogenic risk values for As are above acceptable values set by US EPA ($\geq 10^{-4}$) in all samples, and a potential hazard.

The primary contaminant in the samples was aluminum, in large amounts in samples in brine. While aluminum presented higher quantities, despite lower arsenic concentrations, they proved unacceptable, contributing to the general Hazard Index in all samples.

Considering the pollution index, Al, Se, and As are the principal pollutants, with samples achieving PI values above 1. Since the canned tuna samples had a high concentration of heavy metals, they could serve as bioindicators of seas and oceans pollution that may be contaminated with various heavy metals. Yet, it is not possible to disregard the role of packaging contaminations for some elements, such as aluminum.

With the high concentration of heavy metals in our samples and other studies, it is not safe to maintain the annual intake established by the FAO/FDA for fish consumption when canned samples are the only source, considering that the average serving amount of canned tuna sold in Brazil is 130 g, which can be harmful from an elementary point of view. Lower amounts may be more adequate. This statement does not mean that fresh fish is free from contamination, and further studies should be carried out.

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References

1. Machate, D.J.; Melo, E.S.d.P.; Arakaki, D.G.; Guimarães, R.d.C.A.; Hiane, P.A.; Bogo, D.; Pott, A.; Nascimento, V.A.d. High Concentration of Heavy Metal and Metalloid Levels in Edible Camponemasia Adamantium Pulp from Anthropic Areas. *Int. J. Environ. Res. Public Health* 2021, 18, 5503. [CrossRef]

2. Palma-Lara, I.; Martinez-Castillo, M.; Quintana-Pérez, J.C.; Arellano-Mendoza, M.G.; Tamay-Cach, F.; Valenzuela-Limón, O.L.; García-Montalvo, E.A.; Hernández-Zavala, A. Arsenic Exposure: A Public Health Problem Leading to Several Cancers. *Regul. Toxicol. Pharmacol.* 2020, 110, 104539. [CrossRef] [PubMed]

3. Rai, P.K.; Lee, S.S.; Zhang, M.; Tsang, Y.F.; Kim, K.-H. Heavy Metals in Food Crops: Health Risks, Fate, Mechanisms, and Management. *Environ. Int.* 2019, 125, 365–385. [CrossRef] [PubMed]

4. Srivastava, V.; Sarkar, A.; Singh, S.; Singh, P.; de Araújo, A.S.F.; Singh, R.P. Agroecological Responses of Heavy Metal Pollution with Special Emphasis on Soil Health and Plant Performances. *Front. Environ. Sci.* 2017, 5, 64. [CrossRef]

5. Withanachchi, S.S.; Ghambashidze, G.; Kunchulia, I.; Urushadze, T.; Ploeger, A. Water Quality in Surface Water: A Preliminary Assessment of Heavy Metal Contamination of the Mashavera River, Georgia. *Int. J. Environ. Res. Public Health* 2018, 15, 621. [CrossRef]

6. Custodio, M.; Cuadrado, W.; Peñaloza, R.; Montalvo, R.; Ochoa, S.; Quispe, J. Human Risk from Exposure to Heavy Metals and Arsenic in Water from Rivers with Mining Influence in the Central Andes of Peru. *Water* 2020, 12, 1946. [CrossRef]

7. Landrigan, P.J.; Stegeman, J.J.; Fleming, L.E.; Allemand, D.; Anderson, D.M.; Backer, L.C.; Brucker-Davis, F.; Chevalier, N.; Corra, L.; Czerucka, D.; et al. Human Health and Ocean Pollution. *Ann. Glob. Health* 2020, 86, 151. [CrossRef]

8. Fernandes, A.C.; Medeiros, C.O.; Bernardo, G.L.; Ebene, M.V.; Di Pietro, P.F.; de Assis, M.A.A.; Vasconcelos, F.d.A.G.d. Benefits and Risks of Fish Consumption for the Human Health. *Rev. Nutr.* 2012, 25, 283–295. [CrossRef]

9. Mei, J.; Ma, X.; Xie, J. Review on Natural Preservatives for Extending Fish Shelf Life. *Foods* 2019, 8, 490. [CrossRef]

10. Machate, D.J.; Figueiredo, P.S.; Marcelino, G.; Guimarães, R.d.C.A.; Hiane, P.A.; Bogo, D.; Pinheiro, V.A.Z.; de Oliveira, L.C.S.; Pott, A. Fatty Acid Diets: Regulation of Gut Microbiota Composition and Obesity and Its Related Metabolic Dysbiosis. *Int. J. Mol. Sci.* 2020, 21, 4093. [CrossRef]

11. FAO. The State of World Fisheries and Aquaculture 2020. In *Brief: Sustainability in Action*; FAO: Rome, Italy, 2020; ISBN 978-92-5-132777-9.

12. Ababneh, F.A.; Al-Momani, I.F. Levels of Mercury, Cadmium, Lead and Other Selected Elements in Canned Tuna Fish Commercialised in Jordan. *Int. J. Environ. Anal. Chem.* 2013, 93, 755–766. [CrossRef]

13. Al Ghoul, L.; Abiad, M.G.; Jammoul, A.; Matta, J.; El Darra, N. Zinc, Aluminium, Tin and Bis-Phenol a in Canned Tuna Fish Commercialized within the Specialized in Jordan. *Int. J. Environ. Anal. Chem.* 2013, 93, 755–766. [CrossRef]

14. Alva, C.V.; Mazaric, E.T.; Ribiero, R.d.O.R.; da Carneiro, C.S.; Simões, J.S.; da Ferreira, M.S. Concentrations and Health Risk Assessment of Total Mercury in Canned Tuna Marketed in Southeast Brazil. *J. Food Compos. Anal.* 2020, 88, 103357. [CrossRef]

15. Andyahren, S.; Hadiani, M.R.; Moussavi, Z.; Shoeibi, S. Lead, Cadmium, Arsenic and Mercury in Canned Tuna Fish Marketed in Tehran, Iran. *Food Addit. Contam. Part B* 2015, 8, 93–98. [CrossRef]

16. Boufleur, L.A.; dos Santos, C.E.I.; Debastiani, R.; Yoneama, M.L.; Amaral, L.; Dias, J.F. Elemental Characterization of Brazilian Canned Tuna Fish Using Particle Induced X-ray Emission (PIXE). *J. Food Compos. Anal.* 2013, 30, 19–25. [CrossRef]

17. de Paiva, E.L.; Morgano, M.A.; Milani, R.F. Cadmium, Lead, Tin, Total Mercury, and Methylmercury in Canned Tuna Commercialized in São Paulo, Brazil. *Food Addit. Contam. Part B* 2017, 10, 185–191. [CrossRef]

18. Idriss, A.A.; Ahmad, A.K. Heavy Metal Concentrations in Fishes from Juru River, Estimation of the Health Risk. *Bull. Environ. Contam. Toxicol.* 2015, 94, 204–208. [CrossRef]

19. Ikem, A.; Egibeor, N.O. Assessment of Trace Elements in Canned Fishes (Mackerel, Tuna, Salmon, Sardines and Herrings) Marketed in Georgia and Alabama (United States of America). *J. Food Compos. Anal.* 2005, 18, 771–787. [CrossRef]

20. Mol, S. Levels of Selected Trace Metals in Canned Tuna Fish Produced in Turkey. *J. Food Compos. Anal.* 2011, 24, 66–69. [CrossRef]

21. Russo, R.; Lo Voi, A.; De Simone, A.; Serpe, F.P.; Anastasio, A.; Pepe, T.; Cacace, D.; Severino, L. Heavy Metals in Canned Tuna from Italian Markets. *J. Food Prot.* 2013, 76, 355–359. [CrossRef]

22. de Paiva, E.L.; Morgano, M.A.; Milani, R.F. Cadmium, Lead, Tin, Total Mercury, and Methylmercury in Canned Tuna Commercialized in São Paulo, Brazil. *Food Addit. Contam. Part B* 2017, 10, 185–191. [CrossRef]

23. Hosseini, S.V.; Sobhanardakani, S.; Tayebi, L. Heavy Metals Contamination of Canned Fish and Related Health Implications in Iran. *Turk. J. Fish. Aquat. Sci.* 2018, 18, 951–957.

24. Tuzen, M.; Soyak, M. Determination of Trace Metals in Canned Fish Marketed in Turkey. *Food Chem.* 2007, 101, 1378–1382. [CrossRef]

25. De Lima, N.V.; Melo, E.S.d.P.; Arakaki, D.G.; Tschinkel, P.F.S.; de Souza, I.D.; de Oliveira Ulbrecht, M.O.; Mendes dos Reis, F.J.; Rosa, A.C.G.; Rosa, R.H.; Aragão do Nascimento, V. Data on Metals, Nonmetal, and Metalloid in the Samples of the Canned Tuna and Canned Sardines Sold in Brazil. *Data Brief* 2021, 35, 108656. [CrossRef]

26. Hong, Y.-S.; Song, K.-H.; Chung, J.-Y. Health Effects of Chronic Arsenic Exposure. *J. Prev. Med. Public Health* 2014, 47, 245–252. [CrossRef] [PubMed]

27. Klotz, K.; Weistenhöfer, W.; Neff, F.; Hartwig, A.; van Thriel, C.; Drexler, H. The Health Effects of Aluminum Exposure. *Dtsch. Arztebl. Int.* 2017, 114, 653–659. [CrossRef]

28. Inan-Eroglu, E.; Ayaz, A. Is Aluminum Exposure a Risk Factor for Neurological Disorders? *J. Res. Med. Sci* 2018, 23, 51. [CrossRef] [PubMed]

29. Roney, N. Toxicological Profile for Zinc; Agency for Toxic Substances and Disease Registry: Atlanta, GA, USA, 2005.
29. Wessling-Resnick, M. Excess Iron: Considerations Related to Development and Early Growth. *Am. J. Clin. Nutr.* 2017, 106, 1605S–1608S. [CrossRef]

30. Petrović, M. Selenium: Widespread yet Scarce, Essential yet Toxic. *ChemTexts* 2021, 7, 11. [CrossRef]

31. Peana, M.; Medici, S.; Dadar, M.; Zoroddu, M.A.; Pelucelli, A.; Chasapis, C.T.; Bjørklund, G. Environmental Barium: Potential Exposure and Health-Hazards. *Arch. Toxicol.* 2021, 95, 2605–2612. [CrossRef]

32. Song, W.; Qi, R.; Zhao, L.; Xue, N.; Wang, L.; Yang, Y. Bacterial Community Rather than Metals Shaping Metal Resistance Genes in Water, Sediment and Biofilm in Lakes from Arid Northwestern China. *Environ. Pollut.* 2019, 245, 113041. [CrossRef]

33. Qu, B.; Song, J.; Yuan, H.; Li, X.; Li, N.; Duan, L.; Liang, X. Historical Evolutions of Sediment Quality in Bays under Serious Anthropogenic Influences in China, Basing on Fuzzy Comprehensive Assessment of Heavy Metals. *Environ. Sci. Pollut. Res.* 2020, 27, 25933–25942. [CrossRef] [PubMed]

34. Joint FAO/WHO Expert Committee on Food Additives; World Health Organization. *Evaluation of Certain Food Additives and Contaminants: Fifty-Third Report of the Joint FAO/WHO Expert Committee on Food Additives; World Health Organization*: Geneva, Switzerland, 2000.

35. Joint FAO/WHO Expert Committee on Food Additives; World Health Organization. *Evaluation of Certain Food Additives and Contaminants: Sixty-First Report of the Joint FAO/WHO Expert Committee on Food Additives; World Health Organization*: Geneva, Switzerland, 2004; Volume 61, ISBN 92-4-120922-4.

36. National Academies of Sciences, E.; Oria, M.; Harrison, M.; Stallings, V.A. Dietary Reference Intakes (DRIs): Tolerable Upper Intake Levels, Elements, Food and Nutrition Board, National Academies. Available online: https://www.ncbi.nlm.nih.gov/books/NBK545442/table/app_tab9/ (accessed on 5 October 2020).

37. WHO; JECFA Evaluations of the Joint FAO/WHO Expert Committee on Food Additives (JECFA). Available online: https://apps.who.int/food-additives-contaminants-jecfa-database/chemical.aspx?chemID=298 (accessed on 6 October 2020).

38. World Health Organization. *Barium in Drinking-Water. Background Document for Preparation of WHO Guidelines for Drinking-Water Quality*: WHO, Geneva, Switzerland, 2004; Available online: https://www.who.int/water_sanitation_health/dwq/GDWQ2004web.pdf (accessed on 20 September 2021).

39. World Health Organization. *Principles and Methods for the Risk Assessment of Chemicals in Food; World Health Organization*: Geneva, Switzerland, 2009; ISBN 92-4-157240-X.

40. AOAC. *Guidelines for Single Laboratory Validation of Chemical Methods for Dietary Supplements and Botanicals*: AOAC: Arlington, VA, USA, 2002.

41. Mermet, J.-M.; Poussel, E. ICP Emission Spectrometers: 1995 Analytical Figures of Merit. *Appl. Spectrosc.* 1995, 49, 12A–18A. [CrossRef]

42. FDA, E. 06/09/2014: FDA and EPA Issue Updated Draft Advice for Fish Consumption/Advice Encourages Pregnant Women and Breastfeeding Mothers to Eat More Fish That Are Lower in Mercury. Available online: https://archive.epa.gov/epapages/newsroom_archive/newsreleases/b8edc480d8fe29b85257cf20065f826.html (accessed on 17 August 2021).

43. Means, B. *Risk-Assessment Guidance for Superfund. Volume I. Human Health Evaluation Manual. Part A. Interim Report (Final)*; Environmental Protection Agency: Washington, DC, USA, 1989.

44. Thompson, M.; Ellison, S.L.R.; Wood, R. Harmonized guidelines for single-laboratory validation of methods of analysis (IUPAC Technical Report). *Pure Appl. Chem.* 2002, 74, 835–855. [CrossRef]

45. USEPA. *Integrated Risk Information System*. Available online: https://www.epa.gov/iris (accessed on 13 February 2021).

46. USEPA. *Guidance Manual for Assessing Human Health Risks from Chemically Contaminated, Fish and Shellfish*; USEPA: Washington, DC, USA, 1989.

47. Adebiyi, F.M.; Ore, O.T.; Ogunjimi, I.O. Evaluation of Human Health Risk Assessment of Potential Toxic Metals in Commonly Consumed Crayfish (Palaemon Hastatus) in Nigeria. *Heliyon* 2020, 6, e03092. [CrossRef] [PubMed]

48. Hosomi, R.; Yoshida, M.; Fukunaga, K. Seafood Consumption and Components for Health. *Glob. J. Health Sci.* 2012, 4, 72–86. [CrossRef] [PubMed]

49. Food and Agriculture Organization of the United Nations. *Codex Nutrient Reference Values: Especially for Vitamins, Minerals and Protein*; Food & Agriculture ORG: Rome, Italy, 2019; ISBN 978-92-5-131957-4.

50. USEPA. Program Information about the Integrated Risk Information System: Chronic Oral Reference Dose (RfD). Available online: https://www.epa.gov/iris/iris-reference-dose-rfd-description-and-use-health-risk-assessments (accessed on 11 September 2021).

51. Liang, Y.; Yi, X.; Dang, Z.; Wang, Q.; Luo, H.; Tang, J. Heavy Metal Contamination and Health Risk Assessment in the Vicinity of a Tailing Pond in Guangdong, China. *Int. J. Environ. Res. Public Health* 2017, 14, 1557. [CrossRef]

52. ANVISA. Resolução RDC No. 42 de 29 de Agosto de 2013, Dispõe Sobre o Regulamento Técnico MERCOSUL Sobre Limites Máximos de Contaminantes Inorgânicos Em Alimentos. 2013. Available online: https://bvsms.saude.gov.br/bvs/saudelegis/anvisa/2013/rdc0042_29_08_2013.html#:~:text=Disp%C3%A9%20sobre%20Regulamento%20T%C3%A9cnico%20e%20RDC%20%20art%20II%20%200%20art (accessed on 19 August 2021).

53. Joint FAO/WHO. *WHO Food Standards Programme Codex Committee on Contaminants in Foods*. 2011. Available online: https://www.fao.org/who-codexalimentarius/committees/committee/en/?committee=CCCF (accessed on 20 September 2021).
54. NRS National Residue Survey (NRS) Annual Report 2010–11. Available online: https://nla.gov.au/nla.obj-763108559 (accessed on 30 August 2021).

55. European Commission. Commission Regulation (EC) No 1881/2006. Off. J. Eur. Union 2006, 49, 5–24. Available online: https://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2006:364:0005:0024:EN:PDF (accessed on 20 September 2021).

56. Türkmen, A.; Türkmen, M.; Tepe, Y.; Akyurt, I. Heavy Metals in Three Commercially Valuable Fish Species from Iskenderun Bay, Northern East Mediterranean Sea. Turkey. Food Chem. 2005, 91, 167–172. [CrossRef]

57. Dordevic, D.; Buchtova, H.; Janickova, S.; Macharackova, B.; Jarosova, M.; Vitez, T.; Kushkevych, I. Aluminum Contamination of Food during Culinary Preparation: Case Study with Aluminum Foil and Consumers’ Preferences. Food Sci. Nutr. 2019, 7, 3349–3360. [CrossRef]

58. Bondy, S.C. Prolonged Exposure to Low Levels of Aluminum Leads to Changes Associated with Brain Aging and Neurodegeneration. Toxicology 2014, 315, 1–7. [CrossRef] [PubMed]

59. Stahl, T.; Falk, S.; Rohrbeck, A.; Georgii, S.; Herzog, C.; Wiegand, A.; Hotz, S.; Bosche, B.; Zorn, H.; Brunn, H. Migration of Aluminum from Food Contact Materials to Food—A Health Risk for Consumers? Part I of III: Exposure to Aluminum, Release of Aluminum, Tolerable Weekly Intake (TWI), Toxicological Effects of Aluminum, Study Design, and Methods. Environ. Sci. Eur. 2017, 29, 19. [CrossRef]

60. Ciminelli, V.S.T.; Gasparon, M.; Ng, J.C.; Silva, G.C.; Caldeira, C.L. Dietary Arsenic Exposure in Brazil: The Contribution of Rice and Beans. Chemosphere 2017, 168, 996–1003. [CrossRef] [PubMed]

61. Lee, S.G.; Kim, D.H.; Lee, Y.S.; Cho, S.-Y.; Chung, M.-S.; Cho, M.; Kang, Y.; Kim, H.; Kim, D.; Lee, K.-W. Monitoring of Arsenic Contents in Domestic Rice and Human Risk Assessment for Daily Intake of Inorganic Arsenic in Korea. J. Food Compos. Anal. 2018, 69, 25–32. [CrossRef]

62. Roya, A.Q.; Ali, M.S. Heavy Metals in Rice Samples on the Torbat-Heidarieh Market, Iran. Food Addit. Contam. Part B 2017, 10, 59–63. [CrossRef] [PubMed]

63. Tao, S.; Bolger, P.M. Dietary Arsenic Intakes in the United States: FDA Total Diet Study, September 1991–December 1996. Food Addit. Contam. 2010, 16, 465–472. [CrossRef]

64. Huq, S.M.I.; Joardar, J.C.; Parvin, S.; Correll, R.; Naidu, R. Arsenic Contamination in Food-Chain: Transfer of Arsenic into Food Materials through Groundwater Irrigation. J. Health Popul. Nutr. 2006, 24, 305–316.

65. Santra, S.C.; Samal, A.C.; Bhattacharya, P.; Banerjee, S.; Biswas, A.; Majumdar, J. Arsenic in Foodchain and Community Health Risk: A Study in Gangetic West Bengal. Procedia Environ. Sci. 2013, 18, 2–13. [CrossRef]

66. Shankar, S.; Shanker, U. Shikha Arsenic Contamination of Groundwater: A Review of Sources, Prevalence, Health Risks, and Strategies for Mitigation. Sci. World J. 2014, 2014, e304524. [CrossRef]

67. Pearson, A.J.; Ashmore, E. Risk Assessment of Antimony, Barium, Beryllium, Boron, Bromine, Lithium, Nickel, Strontium, Thallium and Uranium Concentrations in the New Zealand Diet. Food Addit. Contam. Part A 2020, 37, 451–464. [CrossRef]

68. Montanari, A. Inorganic Contaminants of Food as a Function of Packaging Features. In Food Packaging Hygiene; Barone, C., Bolzoni, L., Caruso, G., Montanari, A., Parisi, S., Steinka, I., Eds.; SpringerBriefs in Molecular Science; Springer International Publishing: Cham, Switzerland, 2015; pp. 17–41. ISBN 978-3-319-14827-4.

69. European Commission. Commission Regulation (EU) No. 10/2011 of 14 January 2011 on Plastic Materials and Articles Intended to Come into Contact with Food. Off. J. Eur. Union 2011, L12, 1–89. Available online: https://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2011:012:0001:0089:en:PDF (accessed on 20 September 2021).

70. Cormick, G.; Belizáin, J.M. Calcium Intake and Health. Nutrients 2019, 11, 1606. [CrossRef] [PubMed]

71. Hussein, A.; Khaled, A. Determination of Metals in Tuna Species and Bivalves from Alexandria, Egypt. Egypt J. Aquat. Res. 2014, 40, 9–17. [CrossRef]

72. Gonoodi, K.; Moslem, A.; Darroudi, S.; Ahmadnezhad, M.; Mazloum, Z.; Tayefi, M.; Zadeh, S.A.T.; Esfami, S.; Shafiee, M.; Khayatarmanes, Z.; et al. Serum and Dietary Zinc and Copper in Iranian Girls. Clin. Biochem. 2018, 54, 25–31. [CrossRef] [PubMed]

73. Bost, M.; Houdart, S.; Oberli, M.; Kalonji, E.; Huneau, J.-F.; Margaritis, I. Dietary Copper and Human Health: Current Evidence and Unresolved Issues. J. Trace Elem. Med. Biol. 2016, 35, 107–115. [CrossRef] [PubMed]

74. Mehta, R.; Templeton, D.M.; O’Brien, P.J. Mitochondrial Involvement in Genetically Determined Transition Metal Toxicity: II. Copper Toxicity. Chem. Biol. Interact. 2006, 163, 77–85. [CrossRef]

75. Brewer, G.J. Copper-2 Ingestion, Plus Increased Meat Eating Leading to Increased Copper Absorption, Are Major Factors Behind the Current Epidemic of Alzheimer’s Disease. Nutrients 2015, 7, 10053–10064. [CrossRef]

76. De Romana, D.L.; Olivares, M.; Uaay, R.; Araya, M. Risks and Benefits of Copper in Light of New Insights of Copper Homeostasis. J. Trace Elem. Med. Biol. 2011, 25, 3–13. [CrossRef]

77. Mehta, R.; Templeton, D.M.; O’Brien, P.J. Mitochondrial Involvement in Genetically Determined Transition Metal Toxicity: II. Copper Toxicity. Chem. Biol. Interact. 2006, 163, 77–85. [CrossRef]

78. Rahmani, J.; Fakhrri, Y.; Shahsavani, A.; Rahmani, Z.; Urbina, M.A.; Chirumbolo, S.; Keramati, H.; Moradi, B.; Bay, A.; Bjerklund, G. A Systematic Review and Meta-Analysis of Metal Concentrations in Canned Tuna Fish in Iran and Human Health Risk Assessment. Food Chem. Toxicol. 2018, 118, 753–765. [CrossRef]

79. Girolametti, F.; Annibaldi, A.; Carnevali, O.; Pignalosa, P.; Illuminati, S.; Truzzi, C. Potential Toxic Elements (PTEs) in Wild and Farmed Atlantic Bluefin Tuna (Thunnus Thynnus) from Mediterranean Sea: Risks and Benefits for Human Consumption. Food Control 2021, 125, 108012. [CrossRef]

80. Abbaspour, N.; Hurrell, R.; Kelishadi, R. Review on Iron and Its Importance for Human Health. J. Res. Med. Sci. 2014, 19, 164–174.
80. Pasricha, S.-R.; Low, M.; Thompson, J.; Farrell, A.; De-Regil, L.-M. Iron Supplementation Benefits Physical Performance in Women of Reproductive Age: A Systematic Review and Meta-Analysis. *J. Nutr.* 2014, 144, 906–914. [CrossRef] [PubMed]

81. Annibaldi, A.; Truzzi, C.; Carnevali, O.; Pignalosa, P.; Api, M.; Scarponi, G.; Illuminati, S. Determination of Hg in Farmed and Wild Atlantic Bluefin Tuna (*Thunnus thynnus* L.) Muscle. *Molecules* 2019, 24, 1273. [CrossRef] [PubMed]

82. WHO. *Trace Elements in Human Nutrition and Health*; World Health Organization: Geneva, Switzerland, 1996; ISBN 92-4-156173-4.

83. Rayman, M.P. Selenium Intake, Status, and Health: A Complex Relationship. *Hormones* 2020, 19, 9–14. [CrossRef] [PubMed]

84. Stranges, S.; Marshall, J.; Natarajan, R.; Trevisan, M.; Combs, G.; Cappuccio, F.; Ceriello, A.; Reid, M. Effects of Long-Term Selenium Supplementation on the Incidence of Type 2 Diabetes: A Randomized Trial. *Ann. Intern. Med.* 2007, 147, 217–223. [CrossRef] [PubMed]

85. Behne, D.; Weiler, H.; Kyriakopoulos, A. Effects of Selenium Deficiency on Testicular Morphology and Function in Rats. *Reproduction* 1996, 106, 291–297. [CrossRef]

86. Hill, K.E.; Zhou, J.; McMahan, W.J.; Motley, A.K.; Burk, R.F. Neurological Dysfunction Occurs in Mice with Targeted Deletion of the Selenoprotein P Gene. *J. Nutr.* 2004, 134, 157–161. [CrossRef]

87. Vézina, D.; Mauffette, F.; Roberts, K.D.; Bleau, G. Selenium-Vitamin E Supplementation in Infertile Men. *Biol. Trace Elem. Res.* 1996, 53, 65–83. [CrossRef]

88. Plum, L.M.; Rink, L.; Haase, H. The Essential Toxin: Impact of Zinc on Human Health. *Int. J. Environ. Res. Public Health* 2010, 7, 1342–1365. [CrossRef]

89. Han, J.-L.; Pan, X.-D.; Chen, Q.; Huang, B.-F. Health Risk Assessment of Heavy Metals in Marine Fish to the Population in Zhejiang, China. *Sci. Rep.* 2021, 11, 11079. [CrossRef]

90. Ashraf, W. Levels of Selected Heavy Metals in Tuna Fish. *Arab. J. Sci. Eng.* 2006, 31, 89.

91. Eriksson, B.K. Long-Term Changes in Macroalgal Vegetation on the Swedish Coast: An Evaluation of Eutrophication Effects with Special Emphasis on Increased Organic Sedimentation. Ph.D. Thesis, Acta Universitatis Upsaliensis, Uppsala, Sweden, 2002.

92. Kojadinovic, J.; Potier, M.; Le Corre, M.; Cosson, R.P.; Bustamante, P. Bioaccumulation of Trace Elements in Pelagic Fish from the Western Indian Ocean. *Environ. Pollut.* 2007, 146, 548–566. [CrossRef] [PubMed]

93. Omar, W.A.; Saleh, Y.S.; Marie, M.-A.S. Integrating Multiple Fish Biomarkers and Risk Assessment as Indicators of Metal Pollution along the Red Sea Coast of Hodeida, Yemen Republic. *Ecotoxicol. Environ. Saf.* 2014, 110, 221–231. [CrossRef] [PubMed]

94. Hydes, D.J. Dissolved Aluminium Concentration in Sea Water. *Nature* 1977, 268, 136–137. [CrossRef]

95. Hydes, D.J. Aluminum in Seawater: Control by Inorganic Processes. *Science* 1979, 205, 1260–1262. [CrossRef] [PubMed]

96. Simal-Gándara, J. Selection of Can Coatings for Different Applications. *Food Rev. Int.* 1999, 15, 121–137. [CrossRef]

97. Ninčević Grassino, A.; Grabarić, Z.; Pezzani, A.; Squitieri, G.; Fasanaro, G.; Impembo, M. Corrosion Behaviour of Tinplate Cans in Contact with Tomato Purée and Protective (Inhibiting) Substances. *Food Addit. Contam. Part A* 2009, 26, 1488–1494. [CrossRef] [PubMed]

98. Lemly, A.D. Guidelines for Evaluating Selenium Data from Aquatic Monitoring and Assessment Studies. *Environ. Monit. Assess.* 1993, 28, 83–100. [CrossRef]

99. Hamilton, S.J. Review of Selenium Toxicity in the Aquatic Food Chain. *Sci. Total Environ.* 2004, 326, 1–31. [CrossRef]

100. Neff, J.M. Ecotoxicology of Arsenic in the Marine Environment. *Environ. Toxicol. Chem.* 1997, 16, 917–927. [CrossRef]

101. UN. The Second World Ocean Assessment. Available online: https://www.un.org/regularprocess/woa2launch (accessed on 2 September 2021).