Intratumoral treatment with radioactive beta-emitting microparticles: a systematic review

Robbert C. Bakker1,2 · Marnix G.E.H. Lam1 · Sebastiaan A. van Nimwegen3 · Antoine J.W.P. Rosenberg2 · Robert J.J. van Es4 · J. Frank W. Nijsen1

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Abstract
Purpose The purpose of this study was to review the role of radioactive microparticles (1–100 μm) for the treatment of solid tumors and provide a comprehensive overview of the feasibility, safety, and efficacy.
Methods A systematic search was performed in MEDLINE, EMBASE, and The Cochrane Library (January 2017) by combining synonyms for the determinants “tumor,” “injection,” and “radionuclide.” Data on injection technique, toxicity, tumor response, and survival were collected.
Results The search yielded 7271 studies, and 37 were included for analysis. Twelve studies were performed in human patients and 25 animal studies. The studies were heterogeneous in patient population, tumors, follow-up time, and treatment characteristics. The direct intratumoral injection of radioactive microparticles resulted in a response rate of 71% in a variety of tumors and uncomplicated procedures with high cumulative doses of >19,000 Gy were reported.
Conclusion The large variety of particles, techniques, and treated tumors in the studies provided an important insight into issues concerning efficacy, safety, particle and isotope choice, and other concepts for future research. Animal studies showed efficacy and a dose response. Most studies in humans concluded that intratumoral treatment with radioactive beta-emitting microparticles is relatively safe and effective. Conflicting evidence about safety and efficacy might be explained by the considerable variation in the treatment characteristics. Larger particles had a better retention which resulted in higher anti-tumor effect. Leakage seems to follow the path of least resistance depending on anatomical structures. Subsequently, a grid-like injection procedure with small volume depots is advised over a single large infusion. Controlled image-guided treatment is necessary because inadequate local delivery and inhomogeneous dose distribution result in reduced treatment efficacy and in potential complications.

Keywords Microbrachytherapy · Injection · Microspheres · Particles · Brachytherapy · Selective internal radiation therapy

Introduction

Interventional oncology is an emerging field in cancer care that has the potential to complement existing treatment modalities. Today, various image-guided interventions have an active role in the palliative cancer treatment setting [1–3]. Driven by technical innovation, new image-guided treatment solutions are continuously developing. Interventional oncology techniques, using microspheres or “microbrachytherapy,” have potential benefits, including minimal invasive delivery, outpatient treatment, and improved (progression-free) survival and quality of life [4, 5]. The high-absorbed dose of beta-radiation enables a local tumor-ablative effect while the...
The aim of this literature study was to review the potential role of beta-emitting microparticles for intratumoral (IT) treatment of solid malignant neoplasms. A comprehensive overview of the technical aspects and the characteristics of commonly used radionuclides are provided. Finally, recommendations for further investigation are formulated.

### Methods

#### Protocol and registration

Methods of the analysis and inclusion criteria were specified in advance and documented in a protocol registered in an international prospective register of systematic reviews (PROSPERO) [6].

#### Eligibility criteria

Type of studies: There were no restrictions based on study design, setting, timing, and publication date or publication status. Only full-text articles reported in the English language were included. Studies that examined human or veterinary patients or animal models with solid tumors were included. There were no restrictions on tumor size, type, or location. The administration of the radioactive microparticles had to be performed directly into the tumor. Particles sized between 1 and 100 μm fulfilled our definition of microparticles (Fig. 1), and the particles had to emit beta-radiation. Combined treatment regimens with external beam radiotherapy (EBRT) or chemotherapy were also included. Local treatments after incomplete tumor resections were excluded.

Endpoints included technical details (particle size, injection method, and the amount of injection fluid), biodistribution (retention, IT distribution, and leakage of activity), safety (local and systemic adverse events), and efficacy.

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**Table 1** Characteristics of radionuclides in microparticles

| Radionuclide | Half-life (days) | Beta energy (MeV) | Tissue penetration (mm) | Gamma energy | Production method |
|--------------|-----------------|------------------|------------------------|--------------|------------------|
|              | Mean | Maximum | Mean | Maximum | keV | % Decay |
| Phosphorus-32 $^{32}$P | 14 | 0.695 | 1710.6 | 2.9 | 8 | – | – | Reactor |
| Yttrium-90 $^{90}$Y | 2.7 | 0.935 | 2280.1 | 3.9 | 11 | – | – | Reactor or strontium-90/yttrium-90 generator |
| Iodine-131 $^{131}$I | 8.0 | 0.182 | 806.9 | 0.9 | 5 | 365 | 82% | Reactor |
| Holmium-166 $^{166}$Ho | 1.1 | 0.666 | 1854.9 | 3.2 | 9 | 81 | 6.7% | Reactor |
| Rhenium-186 $^{186}$Re | 3.8 | 0.362 | 1069.5 | 1.8 | 7 | 137 | 9.8% | Reactor |
| Rhenium-188 $^{188}$Re | 0.71 | 0.764 | 2120.4 | 3.5 | 10 | 155 | 15.6% | Tungsten-188/rhenium-188 generator |

MeV mega electron volt

limited penetration depth of maximum 2–11 mm (Table 1) minimizes side effects.

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**Fig. 1** Illustration of particle size.

Particles sized between 1 and 100 μm fulfilled our definition of microparticles, as compared to smaller carriers like antibodies for radioimmunotherapy or nanoparticles, and larger particles or seeds for conventional brachytherapy.
Search

For this review, the electronic databases MEDLINE, EMBASE, and Cochrane Library were searched from dates of inception until January 1, 2017. To ensure literature saturation, the reference lists and citing articles of included studies or relevant reviews identified through the search were scanned.

The full search strategy is listed in the Appendix.

Study selection

The records derived from the search were assessed for eligibility by the author (R.B.) on the titles and abstracts. Full-text manuscripts were screened for all titles that met the inclusion criteria. The reasons for exclusion were recorded. The risk of bias was assessed according to the Newcastle-Ottawa scale to ascertain the validity of eligible trials [7].

Data extraction

The data included (1) methodology, (2) participant details, (3) intervention details, and (4) treatment effect and side effects.

Results

After the removal of duplicates, 7271 records remained out of 10,247 initial records. Of these, 7151 publications did not meet the criteria after reviewing the title and abstract. Subsequently, 22 of the 120 publications were discarded because full text was not available (n = 2), not in the English language (n = 9), or conference abstract or poster (n = 11). The full texts of the remaining 98 studies revealed another 68 studies that did not meet the inclusion criteria. Two additional studies were excluded because of preliminary data and double publication. Cross-referencing identified nine additional studies that fulfilled the inclusion criteria. A total of 37 studies (performed between 1962 and 2014) were included in this review (Fig. 2).

Characteristics and quality of included studies

Twelve studies described the use of beta-emitting microparticles in humans, 23 studies a single animal model, a single study two species, and a single study was performed in veterinary patients. In humans, only one randomized clinical trial was conducted, six cohort studies and five case series. In total, 183 human patients were treated, including a large variety of malignancies, all refractory to other treatments. The used
animals in the tumor model studies were mice (13/24), rats (10/24), and rabbits (2/24). Microbrachytherapy in animals was performed in relatively small tumors (±1 cm), as larger tumors are considered not ethically feasible in small rodent models. The tumors were implanted subcutaneously \((n = 15)\) or in the organ of origin (orthotopically) \((n = 7)\), or were chemically induced \((n = 2)\) [8, 9]. Furthermore, a case series of three feline veterinary patients with a large spontaneous tumor in the liver were treated [5]. The quality of evidence was poor, primarily by design and number of participants. Furthermore, the large variety of microparticles, treatment methods, tumor type, and location made a proper systematic comparison impossible. Therefore, a more descriptive approach was necessary.

**Type of microparticles**

Microspheres (MS) [10–14] and chronic phosphate particles (CPP) were used in five and seven human studies, respectively [15–21] (Table 2). In the 25 animal studies including the study in veterinary patients, a similar division between MS [5, 22–35] \((n = 15)\) and CPP [8, 9, 36–40] \((n = 7)\) was made, with some additional microparticles like \(^{188}\text{Re}\) sulfide particles \((n = 2)\) [41, 42] and labeled MAA particles [43] (Table 3).

The MS were initially made of inert materials such as ceramics/glass, acetylatedonate [5, 31, 32], resin [24, 29], and plastics [10–13]. Nowadays, a large variety of biodegradable MS exists made of biosilicon [14, 28] and gelatin [33, 34]. The currently used MS are often chemically stable for at least the time that they remain radioactive, about 5–10 times the halflife of the incorporated isotope [14]. Thus, the minimum stability depends on the radioisotope; e.g., \(^{90}\text{Y}\) MS with a half-life of 2.6 days must be stable for at least 13 days and \(^{166}\text{Ho}\) with a half-life of 1.1 days at least 5.5 days. In most studies, stability was much longer than minimally required [31, 34].

The second group consisted of CPP with phosphorus-32 \(^{32}\text{P}\). These particles were mostly used in the treatment of hemophilic arthropathy or natural cavities with malignant effusion. The main reason for a direct IT approach with these particles was the inability to deliver sufficient absorbed doses with systemic radioimmunotherapy [40]. In addition to the \(^{32}\text{P}\) CPP, \(^{188}\text{Re}\) sulfide particles were fabricated, with the advantage of easier production by generator and the possibility of SPECT imaging of the gamma radiation [42].

**Particle size**

In the included studies (see Tables 1 and 2), different particle sizes were used. Only one study investigated the preferred microparticle size for IT ablation [41]. In that study, two suspensions of \(^{188}\text{Re}\) sulfide particles with a particle size distribution of 70.1% of 1–5 μm and 19.8% of 5–10 μm particles compared to 86.6 and 10.9%, respectively, were injected in a sarcoma model with a diameter of 1 cm in Kunming mice [41]. The IT retention was higher for the larger particles at various time points (Fig. 3). A similar trend was observed in other studies that investigated the kinetics of IT injected microparticles compared to sub-micron [35], nanoparticles [9], or the effect of the addition of larger particles [40].

The effect of particle size on distribution, retention, elimination, and efficacy was clearly displayed in a study of five different sized phosphorus-32 \(^{32}\text{P}\) compounds in 89 Sprague-Dawley rats with chemically induced breast tumors [9]. Thirty-two days after injection, an IT retention of radioactivity was found of 2.51 ± 0.39% for molecular \(^{32}\text{P}\) sodium orthophosphate (<1 nm), while 10–30 nm CPP had a retention of 28.93 ± 1.30%. The retention further increased for 30–70 nm (49.82 ± 5.41%) and 0.6–1.3 μm (51.61 ± 5.82%) sized particles. Larger charcoal CPP of 2.5–4.0 μm had the best retention of 84.50 ± 2.50% after 32 days. The elimination was primarily through urine and feces and had an inverse relationship with particle size ranging from 85.90 to 12.70% of the injected dose, respectively. The anti-tumor efficacy improved with higher retention because the tumor size ratios (tumor diameter after 32 days/tumor diameter at the start) after 32 days were 4.9 in non-treated controls and 4.5, 1.4, 1.1, 0.9, and 0.6 for the treated tumors in increasing order of particle size.

**Beta-emitting isotopes**

Eight human studies used \(^{32}\text{P}\) and four used \(^{90}\text{Y}\). These isotopes were often considered ideal by the authors because of their pure beta-emission. In animal studies, \(^{131}\text{I}\), \(^{166}\text{Ho}\), \(^{186}\text{Re}\), and \(^{188}\text{Re}\) were used [44]. These isotopes also emit gamma radiation, which can be used for particle localization and quantitative imaging. None of the reviewed studies compared safety and efficacy between different radionuclides. Experience, production, biodistribution, imaging possibilities, pharmacokinetics, and clearance mostly defined preference [33, 42]. Besides these differences, relatively small differences in the energy spectrum, penetration depth, and half-life time exist. See Table 1.

**Technique**

Due to the experimental nature of IT microbrachytherapy, no generally accepted standard administration method exists. Furthermore, most research was performed in small rodent tumor models, which are less informative for translation of the administration technique to the human
| Study               | Number of patients | Tumor type                                         | Isotope | Particle | Particle size (μm) | Multiple injections/ single infusion | Imaging | Needle gauge | Tumor size[^a] | Amount of fluid Injected activity (MBq)/ absorbed dose (Gy) |
|---------------------|--------------------|---------------------------------------------------|---------|----------|-------------------|--------------------------------------|---------|--------------|----------------|-----------------------------------------------------------|
| Kim 1962 [10]       | 10                 | Breast, bladder, brain, prostate, lung, metastasis | ^90^Y   | Ceramic MS | 60 ± 5            | Multiple injections                  |         |              |                |                                                           |
| Blanchard 1965 [11]| 12                 | Bladder, prostate, breast, lung, metastasis       | ^90^Y   | Ceramic MS | 40–60             | Multiple injections                  |         |              |                |                                                           |
| Ariel 1978 [12]     | 1                  | Rhabdomyosarcoma                                  | ^90^Y   | Ceramic MS | 15 ± 10            | Multiple injections                  |         |              |                |                                                           |
| Order 1996 [15]     | 47                 | Pancreas carcinoma                                | ^32^P   | CPP + MAA  | 0.6–1.3 + 10–90    | Single infusion                      | CT      | T1–T3        | <4.5 ml        | 148–1110 MBq                                               |
| Tian 1996 [13]      | 33                 | 27 HCC 6 liver metastasis                         | ^90^Y   | Glass MS   | 0.6–1.3 + 10–90    | Multiple injections                  | US      | 1.8–10.7 cm  | 0.1–0.3 ml     | 370/4440 MBq                                               |
| Westlin 1997 [16]   | 17                 | Pancreas carcinoma                                | ^32^P   | CPP + MAA  | 0.6–1.3 + 10–90    | Single infusion                      | US      | 13 cm³ (3.1–37.5) | Max 25% tumor volume | 1390–9000 Gy                                               |
| DeNittes 1999 [17]  | 5                  | Pancreas carcinoma                                | ^32^P   | CPP + MAA  | 0.6–1.3 + 10–90    | Single infusion                      | US      | 10–290 cm³  | 3–4.5 ml       | 110 MBq                                                    |
| Firusian 1999 [18]  | 17                 | Various solid malignancies                        | ^32^P   | CPP       | 0.6–2             | Single infusion                      | US      |              | 5–15 ml        |                                                           |
| Montijo 2003 [19]   | 1                  | Pancreas carcinoma                                | ^32^P   | CPP + MAA  | 0.6–1.3            | Single infusion                      |         |              |                | 544 MBq                                                   |
| Alimi 2007 [20]     | 14                 | Secondary resistant H&N tumors                    | ^32^P   | CPP       | 0.6–1.3            | Single infusion                      | US      |              | 5–15 ml        | <20 cm³: 74 MBq; 20–40 cm³: 148 MBq; 50–100 cm³: 222 MBq; >100 cm³: 2–3 sessions; 100–200 cm³: 370 MBq; 200–300 cm³: 555 MBq |
| Goh 2007 [14]       | 8                  | HCC                                               | ^32^P   | BioSilicon MS | 30                 | Multiple injections                  | US/CT   | 18 outer, 22 inner | 7% of tumor volume | 4 MBq/cm² of tumor                                        |
| Rosemurgery 2008 [21]| 30                | Pancreas carcinoma                                | ^32^P   | CPP       | 0.6–1.3            | Multiple injections                  | CT      | 19.7 ± 10.5 cm³ | 25% of tumor volume | (18.5 MBq/g tissue max + 740 MBq) Median dose 1255.34 Gy |

[^a]Tumor size presented as TNM stage, mean ± SD, or median and/or range, cm: diameter, cm²: tumor cross-sectional area, cm³: volume

**HCC** hepatocellular carcinoma, ^90^Y yttrium-90, ^32^P phosphorus-32, **CPP** chromic phosphate particles, **MAA** macroaggregated albumin, **MS** microspheres, **US** ultrasound, **CT** computed tomography, **MBq** megabecquerel, **Gy** Gray
| Study            | Number and type of animals | Tumor type                                                                 | Location SC/orthotopic/spontaneous | Isotope/Particle | Particle size (μm) | Number of injections | Needle gauge | Tumor size | Amount of fluid | Amount of activity |
|------------------|---------------------------|---------------------------------------------------------------------------|-----------------------------------|------------------|-------------------|---------------------|--------------|------------|----------------|-------------------|
| Nakhgevany 1988 | 150 Lewis Wistar rats    | Rat mammary carcinoma: AC33                                               | SC                                | $^{90}$ Y MS     | 18                | 1                   | 0.73         | 0.5 ml    | 37             |                   |
|                  |                           | Human mammary carcinoma: BT-20                                             | SC                                | $^{166}$ Ho       | Glass fragments   | 2–5 Irregular       | 19 (7–37) mm$^3$ | 0.04 ml    | 7.4            |                   |
| Osler 1994       | 27 Male ACI rats          | Rat hepatoma: H4:E                                                          | SC                                | $^{32}$ P CPP + MAA | 0.6–1.3 + 10–90   | 1                   | 0.5–1.5 mm    | 3.7         |                |                   |
| Loo 1997         | C3Hf/HeJ mice             | Murine fibrosarcoma: FsaII                                                  | SC                                | $^{32}$ P CPP + MAA | 1                 | FsAl/LS174t 0.5 cm$^3$ | 0.01 ml     | HBSS/MAA 3.7, 7.4, 14.8 |          |
| Nguyen 1997      | Nude mice                 | Human melanoma: HBL, Human head and neck squamous cell carcinoma: SCC1    | SC                                | $^{32}$ P CPP + MAA | 0.6–1.3 + 10–90   | 1                   | 500 mm$^3$    | 0.1 ml     | 1.85           |                   |
| Watanabe 1997    | Balb/c nude mice          | Human neuroblastoma cell line: SK-N-MC                                     | SC                                | $^{90}$ Y MAA     | 1                 | Fine                             | 1.0 cm$^3$ | 0.05 ml    | 18.5           |                   |
| Zubillaga 1997   | 89 SD rats                | NMU-induced breast carcinoma                                               | Orthotopic                        | $^{32}$ P CPP     | 2.5–4             | 1                   | 2 cm         | 0.1 ml     | 7.4, 37        |                   |
| Wang 1998        | 42 SD rats                | Rat hepatoma: N1S1                                                          | Orthotopic                        | $^{186}$ Re Resin-MS | 15 ± 2          | 1                   | 2 cm         | 0.05 ml    | 18.5           |                   |
| Zubillaga 1998   | 70 SD rats                | Human breast                                                             | Orthotopic                        | $^{32}$ P CPP with charcoal | 2.5–4           | 1                   | 500 mm$^3$    | 0.1 ml     | 17.02 + 6 days | 23.31            |
| Junfeng 1999     | Kunming mice              | Human pancreatic carcinoma: AsPC-1                                         | SC                                | $^{32}$ P CPP + MAA | 0.6–1.3 + 10–60   | 1                   | 500 mm$^3$    | 0.1 ml     |                |                   |
| Lee 1999         | Nude mice                 | Human pancreatic carcinoma: AsPC-1                                         | SC                                | $^{32}$ P CPP + MAA | 0.6–4             | 1                   | 500 mm$^3$    | 0.1 ml     |                |                   |
| Liu L 1999       | Balb/c mice               | Human liver cancer: H-CS                                                  | SC                                | $^{32}$ P Glass MS | 46–76             | 1                   | 0.7–1.0 cm | 0.9–1.2 cm | 0.1 ml | 183–7220 Gy |                   |
| Liu Junfeng 2000 | Athymic nude mice         | Human liver cancer: HepG2 human liver carcinoma: L172                       | SC                                | $^{32}$ P Sulfide suspension | 1–10             | 1                   | 1.0 cm         | 0.05 ml | 18.5, 29.6 |                   |
| Lin 2000         | SD rats                   | Rat hepatoma: N1S1                                                         | Orthotopic                        | $^{90}$ Y Glass MS | 20–30             | 1                   | 2 cm         | 0.1 ml     | 7.4           |                   |
| Chen 2001        | SD rats                   | Rat hepatoma: N1S1                                                         | Orthotopic                        | $^{90}$ Y Glass Theraspheres$^\text{TM}$ | 20–30           | 1                   | 2 cm         | 0.1 ml     | 7.4           |                   |
| Lin 2005         | NZW rabbits               | Rabbit SCC: VX2                                                           | Orthotopic                        | $^{188}$ Re Resin MS | 15 ± 2          | 1                   | 22           | 2 cm 50 µl | 370           |                   |
| Zhang 2005       | BALB/c mice               | Human liver carcinoma: HepG2 human liver carcinoma: L172                   | Orthotopic                        | $^{32}$ P BioSilicon MS | 20               | 1                   | 65.3–88.9 mm$^3$ | 0.5, 1.2 |                |                   |
| Hafeli 2007      | SD rats                   | Rat gliosarcoma: 9L                                                       | Orthotopic                        | $^{186}$ Re Glass MS | 25–35            | 1                   | Identical location | 2 x 10 µl | 2.3 cm | 1.85 186Re/188Re | (ratio 3:1) |
| Lubo 2009        | Wistar rats               | Rat mammary carcinoma: Walker carcinoma 256                               | SC                                | $^{188}$ Re Colloids | 0.3              | 1                   | 10–15 mm    | 2 µl        |                |                   |
| Bult 2012        | NZW rabbits               | Rabbit SCC: VX2                                                           | Orthotopic                        | $^{166}$ Ho Acetyllactonate MS | 15               | 1                   | 2 cm         | 0.1 ml     | 50             |                   |
| Bult 2013        | Balb/c mice               | Mouse renal cell carcinoma                                                | Orthotopic                        | $^{166}$ Ho Acetyllactonate MS | 10–15            | 1                   | 29           | 0.01 ml    | 5              |                   |
| Bult 2013        | DS cats                   | Various                                                                  | Orthotopic                        | $^{166}$ Ho Acetyllactonate MS | 8 ± 2            | 1                   | 29           | 5.6 ± 1.6 mm | 550–2170       |
situation. Therefore, the differences and similarities of the 12 reviewed human studies and the one treatment in veterinary patients will be described in the following paragraphs. These include differences in administration method, the amount of injection, and imaging during and after administration.

**Administration methods**

The larger MS were most often injected using multiple manual injection locations (i.e., sub-milliliter volumes) in a grid-like pattern [13, 21]. The smaller CPP were also administered through a single infusion technique in which a larger volume up to 4.5 ml of $^{32}$P CPP was administered in the tumor center, assuming that the pressure force would distribute the particles throughout the tumor [15]. Empirically, 5 cm was the largest tissue diameter satisfactorily covered by microparticles after a single infusion.

The specific characteristics of the needle used for IT injection were not frequently described. Needle sizes between 18 and 22G (outer diameter 1.2–0.7 mm) were commonly used percutaneously. In addition, an endoscopic ultrasound approach with a 22G needle was utilized in a published abstract [45] and unpublished trial (clinicaltrial.gov NCT00346281). Only a single study in eight humans with pancreatic tumors describes the use of a gel foam, which was injected through the 18G introducer needle to minimize back leakage and seal the needle tract [14].

**Volume of injection**

No studies related the injected volume of fluid-suspended microparticles to the amount of leakage or distribution. The ideal amount of injected fluid to obtain the desired IT distribution is unknown. However, some suggested that larger fluid volumes might result in leakage of microparticles out of the tumor.
With a higher amount of volume (i.e., 4.5 ml), high resistance with a sudden release of syringe pressure was often felt during infusion [15]. Subsequently, radioactivity was detected outside the tumor, presumably due to tissue destruction and leakage to surrounding tissues [15].

The amount of injection volume varied from 7 to 25% of the tumor volume in the most recent studies in pancreatic cancer [14, 21]. In liver tumors with a diameter of 2 to 8.8 cm, small (0.1–0.3 ml) volumes were used per location with a total maximum of 1.0–1.5 ml per treatment session [13]. Results obtained from non-particle intratumoral radionuclide therapies showed that larger volumes were associated with more side effects [46, 47]. In prostate cancer, 20–50 ml (equal to the prostate volume) was injected, which resulted in 55 adverse events like strictures, fistulas, and ulcers in the first 100 patients. In the subsequent 87 patients, only 2–3 ml injection fluid was used, and only eight events occurred. [46].

Amount of activity/absorbed dose

The absorbed dose in tissue (Gray) varied from 120 to 19,300 Gy. A proper rationale for the injected activity or the desired absorbed dose was often missing. In a phase I study of $^{32}$P CPP in 28 patients with unresectable pancreas tumors, a maximum of 1110 MBq for a single infusion was decided [15]. This empirically determined maximum was based on the expected limitation of the injection volume of 4.5 ml. This approach resulted in a maximum cumulative absorbed dose of 17,000 Gy. A more accurate dosing approach was applied in recent studies on pancreas cancer patients. However, the injected activity per gram (or cm$^3$) in the RCT with $^{32}$P CPP and the cohort study with $^{32}$P BioSilicon MS still varied with a factor of 4.6 (4 vs. 18.5 MBq/cm$^3$ tumor) [14, 21].

Image-guided administration

The administration procedure was image guided in nine human studies. In most animal studies ($n = 22$), no imaging was used during the administration. In the veterinary patients and rabbit studies with liver tumors, ultrasound guidance ($n = 2$) was used, and a stereotactic frame was used in glioma-bearing rats. With CT or ultrasound, the tip of the needle was positioned at the desired location before administration. Some authors preferred ultrasound because this modality provided easy and real-time imaging during the actual injection [13]. During the injections of MS, echogenic spots were sometimes seen “flowing” in some narrow, vessel-like gaps and sometimes even out of the tumor boundaries, especially after a fast, forceful injection [13]. Subsequently, shaking of the vial before administration, resulting in air bubbles, was used to visualize any major unexpected leakage outside the tumor during the injection on ultrasound [16].

Outcomes

The primary outcomes of IT microbrachytherapy were safety and efficacy. However, a more fundamental understanding of this treatment is necessary, especially because the outcomes of safety and efficacy probably mostly depend on the distribution of the activity. Therefore, distributional data will be described first.

Distribution

Insufficient retention of radioactive microparticles leads to an insufficiently absorbed dose and therefore an ineffective treatment. However, apart from total absorbed dose, the IT distribution of activity throughout the tumor is crucial, as “missed” parts of the tumor will result in residual vital tumor. Leakage of activity, on the other hand, may lead to an unintended absorbed dose to healthy tissue and could potentially result in side effects.

Leakage

Several potential routes of leakage were identified, and a distinction was made between external leakage and internal leakage. External leakage from the syringe occurred twice during treatment of pancreatic tumors with $^{32}$P CPP infusion due to high resistance in the tumor [16]. The authors experienced in an experiment, with $^{166}$Holmium microsphere injections in ex vivo tissues, a needle disconnection from a Luer lock syringe after exerting high pressure to overcome tissue resistance. Another possible route of external leakage is injection canal leakage. Injection canal leakage was not described in the human or animal studies. However, in the study of eight humans with pancreatic tumors, the authors describe the use of a gel foam pledget/slurry which was injected through the introducer needle to minimize back leakage and seal the needle tract [14].

Internal leakage to non-target tissues was divided in hematogenous or intravenous and intraductal leakage. In the majority of human [13, 15, 16] and animal [8, 9, 24, 35, 39, 43] studies, some degree of intravenous leakage or shunting of particles through the capillary bed was described. After the improved retention of CCP particles with an additional injection of larger 10–90 μm MAA particles, 56 vs. 90%, respectively, and the hypothesis of a vascular blockade, vascularity became an important variable for leakage. $^{32}$P CCP 0.6–1.3 μm was injected in nude mice with human pigmented melanoma cell line (HBL) and a human squamous cell carcinoma cell line (SCC1); three to four times higher organ counting was found in SCC1 [39]. This phenomenon is probably explained by the difference in vascularity between HBL and SCC1 tumors, which contained 5.7 vs. 21.4 blood vessels/mm$^2$, respectively.
In Wistar rats, $^{188}$Re MS (25 μm) and small $^{186}$Re sulfide particles (0.3 μm) were injected in hypervascularized Walker 256 carcinomas and hypovascularized Yoshida sarcomas. This study revealed a bi-phase drainage of the injected particles out of the tumor. A fast wash-out phase, where the IT activity decreases to approximately 70% within 10 min, was followed by a slow decline in which IT activity falls to 60% of the initially injected activity at 48 h. The fast leakage was more pronounced in hypervascularized tumors with smaller particles, whereas the slow decline was independent of particle size and vascularity [35].

In addition, the distribution of activity after IV leakage depends on the tumor location and particle size. In 33 liver cancer patients treated with $^{90}$Y MS, a lung shunt of 9–20% of the injected activity was observed in six patients [13]. Similar shunts were observed after IT injections with $^{188}$Re MS in rats with subcutaneous and liver tumors which resulted in trapped MS in the pulmonary capillary bed [24, 35]. Detected activity in the liver, after the treatment of the pancreas, was probably caused by venous shunting of CCP + MAA [15, 16]. However, small particles such as CCP (±1 μm) and $^{186}$Re-sulfide particles (0.3 μm) can probably also pass through the capillary bed of the tumor and phagocytized in the reticuloendothelial system and therefore detected in the liver.

During the treatment of malignancies in the pancreas and liver, intraductal leakage and activity in the gastrointestinal tract were described. In the 48 patients with $^{32}$P CCP infusion in pancreatic cancer, accidental needle placement and injection into the pancreatic duct occurred. Forty-eight hours after injection, all intestinal activity was excreted without gastrointestinal toxicity [15]. During the treatment of 33 patients with liver malignancies with $^{90}$Y MS, a similar leakage was found in the intestines in four patients that disappeared within 1–2 days [13].

Lymphatic drainage is an additional potential route which was however not observed in the reviewed studies. This well-known route of tumor drainage is commonly used in the sentinel node procedure. The microparticles were presumably too large for drainage of significant amounts of radioactivity to the draining lymph nodes.

**Safety**

The safety and toxicity were closely related to the distribution. The safety or clinical complications were divided into local and systemic side effects. The experimental treatment was often performed in progressively ill patients [10]. The probability of a causal relationship between an event and treatment was therefore often difficult to determine. However, in general, the authors of both animal and human studies concluded that the treatment was safe.

A safety concern, which was not described in the clinical studies, was needle tract metastasis. This complication might have occurred in one animal study. After three thallium-201 injections in eight Fischer 344 rats with an orthotopic glioma model, five metastases occurred of which three were along the needle tract [48]. Whether this was due to disruption of natural barriers or by dragging cells into the needle tract was ambiguous.

**Local side effects**

A reported local side effect in eight pancreatic tumor patients treated with $^{32}$P BioSilicon MS was pain at the injection site ($n$ = 3) and the treated region ($n$ = 1) which resolved within 1 or 2 days [14]. Similar results were found with $^{32}$P CPP in the pancreas. The injection of $^{90}$Y MS in the liver was not painful, in contrast to ethanol injections [13]. Another mild effect that was observed twice was transient erythema after microbrachytherapy of superficial cervical lymph node metastasis of H&N tumors with $^{32}$P CPP [18]. In the 23 patients from the three case series treated with $^{90}$Y MS, the following four local complications were reported: a rectovesical fistula in prostate cancer, a lung abscess and localized radiation fibrosis in bronchial cancer, and a skin defect in a rhabdomyosarcoma of the nose [10–12, 14]. In addition, after treatment of pancreas cancer with $^{32}$P, some patients had increased serum amylase as a sign of local damage [14, 15, 21].

In the randomized trial of 30 patients with pancreas carcinoma treated with a combination of 5FU, 60 Gy EBRT, and gemcitabine [21], 18 patients were additionally treated with $^{32}$P therapy. A gastrointestinal bleeding was experienced in 15 patients of whom 13 were treated with $^{32}$P. In eight patients, this complication seemed attributable to pancreatic tumor eroding into the duodenum. This complication was described in two other pancreas carcinoma patients treated with $^{32}$P CPP [16, 19] (Table 4).

**Systemic side effects**

Hematological abnormalities were a frequently described side effect. This could result from treatment of blood-pooled organs, lacking of activity from microspheres, or disintegration of microspheres into smaller particles. Most of the used radioactive isotopes do have an increased accumulation in bone after leakage, which may result in bone marrow suppression. Pancycopenia was described in 1965 in a patient in whom 10% of the activity leaked from an inadequate batch of $^{90}$Y MS. In the cohort of 48 pancreas carcinoma patients, grade 3 leukopenia and grade 3/4 thrombocytopenia were observed in three and five patients, respectively [15]. Additionally, after treatment of the liver with $^{90}$Y MS, leukopenia was observed in 2 out of 33 patients [13]. However, since the amounts of activity were low (venous samples <11 Bq/ml) [18], the leakage often did not result in clinical toxicity.
Efficacy

The tumoricidal efficacy of intratumoral treatment with radioactive beta-emitting microparticles was shown in animal models. Forty nude mice with subcutaneous liver tumors were treated with \( ^{32}\)P glass MS. This study did not only show that \( ^{32}\)P glass MS were effective in the treatment of a subcutaneous liver tumor model; it additionally showed a dose-response relation [25]. The tumor-inhibiting rate improved from the lowest dose of 183 Gy to the highest dose of 7320 Gy, from 59.7 to 93.6\%, respectively. These results were confirmed in another liver carcinoma line in nude mice with \( ^{188}\)Re [42] (Table 5).

The efficacy in the human studies was more difficult to interpret as 11/12 were non-comparative studies (Table 4). However, the results of eight patients with pancreas carcinomas treated with \( ^{32}\)P BioSilicon MS were promising, with two complete responses, two partial responses, and four patients with stable disease after 12 weeks [14]. Furthermore, a survival benefit was found in the responders as compared to the non-responders for 14 head and neck cancer patients treated with \( ^{32}\)P CPP. On the other hand, a survival benefit was not found in the RCT in 30 pancreas cancer patients with a treatment history of 5-FU, EBRT, and gemcitabine. Patients receiving \( ^{32}\)P CPP in addition to standard therapy survived a median of 5.2 months, whereas patients receiving standard therapy alone survived 12.2 months, \( p = 0.16\). A decrease in radiologic tumor size was not detected on CT because cancer persisted along the periphery of the injection sites [21].

Discussion

In this study, all currently available literature on the potential role of beta-emitting microparticles for IT treatment of solid malignant neoplasms was reviewed. The results of 12 human and 25 animal studies were included. The large variety of particles, techniques, and treated tumors in the studies provided an important insight into issues concerning efficacy, safety, particle and isotope choice, and other concepts for future research.

Is microbrachytherapy effective? Based on the reviewed data, it can be concluded that beta-emitting microparticles seem to be an effective tumoricidal agent. The majority of the studies showed promising results in both humans and animals with complete responses and long-term survival [14]. However, a direct IT injection with tumoricidal particles does not automatically lead to an effective tumor treatment [21]. Obtaining a sufficient dose coverage of all tumor tissue requires the challenging design of an optimal treatment modality with regard to biological stability, injection techniques, dosimetry, biodistribution, etc.

Is microbrachytherapy safe? In the only performed RCT, concerns were raised about the safety of additional IT treatment with small \( ^{32}\)P CPP in pancreas cancer patients treated with 5-fluorouracil, EBRT, and gemcitabine [21]. More patients experienced gastrointestinal bleeding compared to the standard therapy alone. Bleedings were not observed in studies with other particles and other tumors. Other local side effects included manageable discomfort at the injection site. Except for manageable hematological abnormalities, other systemic adverse events were not encountered. Therefore, apart from pancreas tumors, IT treatment seems to be a reasonably safe alternative.

Can we predict complications? Leakage appears to follow the path of least resistance. An easy route of leakage after IT administration is injection canal leakage. The use of a small needle can reduce this. However, care should be taken to prevent premature settling and clotting of microparticles inside the syringe and blocking the needle [5, 31, 32]. A 21G needle seems to be the preferred needle to use. Additional measures to reduce leakage may include slow injection and withdrawal of the needle with slight pressure or injection of obstructing pledget/foam. Other routes of leakage (i.e., intravascular or intraductal) may be caused by injection position, excessive volume, or pressure. Increased permeability of tumor neovascularization may be considered a risk factor for hematogenous leakage. Leakage of an entire dose may happen when a single infusion technique is used [15]. A grid-like injection procedure with larger MS in small volume depots may, therefore, be preferred over the infusion of smaller particles.

How much fluid should be injected during microbrachytherapy? Theoretically, more fluid results in more propelling force and a more homogeneous distribution of microparticles in the target tissue. This should be balanced against the chances of more side effects [46, 47]. The injected volume should probably range between 7 and 30\% of the tumor volume [14, 16] as excessive volume or pressure may result in leakage [15, 16]. In addition, intratumoral pressure depends on tumor characteristics and location and should be taken into account [36–38]. A more viscous fluid may be used to obtain even more control [6, 12]. For example, 25\% glucose, fibrin glue, and other formulas were used to improve the injection procedure [14, 30], or hydrogels such as chitosan [49].

Which particles should be used? There is a relation between particle size and retention: the larger the particle, the higher the retention. Subsequently, preferences for the larger MS exist. On the other hand, particles must be small enough to distribute evenly throughout the tumor to deliver an adequate homogeneously absorbed dose. The optimal number of particles was not mentioned in the studies, but it is likely to influence biodistribution, safety, and efficacy too, and must be investigated to result in a better understanding of IT injection.

What are the ideal radionuclide characteristics? \( ^{90}\)Y is often considered the ideal isotope, with a high energy, pure beta-emitter for easy radiation protection, and an intermediate half-life of 64 h. However, because of the questions related to both IT distribution and retention of microparticles, isotopes with
## Table 4  Outcomes of distribution, efficacy, and safety of human studies

| Study        | Number of patients | Tumor type                          | Isotope | Retention | Leakage     | Toxicity                          | Efficacy                        |
|--------------|--------------------|-------------------------------------|---------|-----------|-------------|-----------------------------------|---------------------------------|
| Kim 1962 [10] | 10                 | Breast, bladder, brain prostate, lung, metastasis | $^{90}$Y |           | N = 4       | 1 localized radiation fibrosis    | N = 4 regression                |
| Blanchard 1965 [11] | 12               | Bladder, prostate, breast, lung, and metastasis   | $^{90}$Y |           | N = 11      | 1 lung abscess                    | N = 1 marked regression         |
| Ariel 1978 [12] | 1                 | Rhabdomyosarcoma                     | $^{90}$Y | Patients without metastasis | Blood 1.85–3552 Bq/ml N = 12 | 1 skin defect                     | N = 1 complete response         |
| Order 1996 [15] | 47                | Pancreas carcinoma                   | $^{32}$P | Without shunting | N = 12       | Without metastases                | N = 7 complete response         |
| Tian 1996 [13]  | 33                | HCC 27 liver metastasis              | $^{90}$Y | Biological T1/2 = 57.6 ± 1.02 h Physical T1/2 = 66 h | N = 6 lung 8.8–20.8% | 1 acute myocardial infarction day 0 of 2nd treatment | N = 12 ≤50% or more |
| Westlin 1997 [16] | 17                | Pancreas carcinoma                   | $^{32}$P |               | N = 2 intestines | 1 arterial bleeding                | N = 5 complete response         |
| DeNittes 1999 [17] | 5                 | Pancreas carcinoma                   | $^{32}$P |               | N = 2 liver       | 2 slightly decreased blood counts | N = 7 stable disease             |
| Firusian 1999 [18] | 17                | Various solid malignancies           | $^{32}$P |               | N = 20%       | No significant toxicity           | N = 2 complete response         |
| Montijo 2003 [19] | 1                 | Pancreas carcinoma                   | $^{32}$P | Biologic T1/2 = physical T1/2 | Blood <11 Bq/ml | 1 Gr IV thrombocytopenia           | N = 7 complete response         |
| Alimi 2007 [20]  | 14                | Secondary resistant H&N tumors       | $^{32}$P |               | N = 5          | 1 Gr IV thrombocytopenia          | N = 5 complete response         |
| Goh 2007 [14]   | 8                 | HCC                                 | $^{32}$P |               | N = 20%       | 1 Gr II thrombocytopenia          | N = 8 partial response          |
|                |                   |                                     |         |              |             | 1 Gr I III metastases             | N = 6 no response                |
|                |                   |                                     |         |              |             | 2 transient erythema              |                                |
|                |                   |                                     |         |              |             | 3 injection site pain             |                                |
|                |                   |                                     |         |              |             | 2 fatigue                         |                                |
|                |                   |                                     |         |              |             | 2 portal hypertension             |                                |
|                |                   |                                     |         |              |             | 1 abdominal pain                   |                                |
|                |                   |                                     |         |              |             | 1 rigors                          |                                |
|                |                   |                                     |         |              |             | 1 vomiting                        |                                |
|                |                   |                                     |         |              |             | 1 Gr IV diabetes mellitus          |                                |
|                |                   |                                     |         |              |             | 1 Gr III neutropenia               |                                |
|                |                   |                                     |         |              |             | 1 Gr III pancytopenia              |                                |

Note: The data for toxicity and efficacy are summarized based on the published studies without providing detailed patient-specific outcomes.
better imaging properties are more suitable for imaging-guided monitoring of IT particle distribution and dosimetry. For leakage to other organs, low-resolution bremsstrahlung scintigraphy is sufficient. However, the resolution of this technique is insufficient for local tumor dose distribution monitoring. SPECT imaging can greatly improve particle distribution measurements for $^{186}$Re, $^{188}$Re, and $^{166}$Ho because of the associated gamma-radiation of 80–200 keV. Furthermore, $^{166}$Ho can be visualized and quantified with CT and MRI [32]. There are several developments in imaging of these isotopes. $^{90}$Y PET/CT is also quantitative but requires long acquisition times due to the low number of positrons. Another relative new imaging opportunity is Cerenkov luminescence imaging (CLI) [50]. CLI could provide quantitative high-resolution imaging and image-based dosimetry for a large variety of isotopes [50–52]. The main limitation of CLI is the limited penetration depth of light of approximately 10 mm into tissue, however very promising, in small animal models [51, 53].

In addition to imaging characteristics, half-life and beta energy should be considered in relation to efficacy and safety, but also logistics, like production and cost. In this respect, a generator like the tungsten-188/rhenium-188 generator may be beneficial. In theory, a high dose rate (i.e., short half-life) will prevent the recovery of radiation damaged tumor cells and may lead to higher efficacy. In terms of logistics, a short half-life may lead to production and logistic challenges on the one hand, but a shorter hospital stay with fewer restrictions after discharge on the other hand.

IT injections can be performed in a variety of tumor types and organs. Based on the postulated methods of leakage, potential risks of side effects, and more challenging administration, the pancreas seems to be a difficult-to-treat organ. Superficial tumors, such as lymph node metastases of the head and neck region, and liver tumors are better accessible, show minimal leakage, and have minimal side effects. With increasing knowledge, microbrachytherapy may be adjusted to tumor characteristics, for example, the addition of a vasoconstrictive drug in hypervascular tumors.

**Conclusion**

Intratumoral treatment with radioactive beta-emitting microparticles, microbrachytherapy, in solid malignant neoplasms may have additional value for patients with tumors at various locations. The uncomplicated treatments with high cumulative doses of up to 19,000 Gy suggest that microbrachytherapy is relatively safe. Larger particles resulted in a higher retention and tumor-inhibiting efficacy of >90% with an intratumoral absorbed dose of 7320 Gy. A small injected volume of 7–30% of the tumor volume divided in small volume deposits, 0.1–0.3 ml, administered in a grid-like injection procedure is preferred. With accurate administration and high-resolution imaging, the efficacy may be
Table 5  Main outcomes of animal studies

| Study          | Animal | Tumor type                  | Isotope | Retention | Toxicity | Efficacy | Survival |
|----------------|--------|----------------------------|---------|-----------|----------|----------|----------|
| Nakhgevan y 1988(22) | Lewis Wistar | Rat mammary carcinoma: AC33 | $^{90}$Y | No histological evidence of radiation damage of liver, bone marrow, kidney |
| Brown 1991(23) | BALB/c mice | Human mammary carcinoma: BT-20 | $^{153}$Ho | | | |
| Order 1994(40) | Male ACI rats | Rat hepatoma: H4-E | $^{32}$P | | | |
| Lee 1997(38) | Rats | Rat hepatoma: H4-E | $^{32}$P | | | |
| Nguen 1997(39) | Nude mice | Human melanoma: HBL Human head and neck squamous cell carcinoma: SCC1 | $^{32}$P | | | |
| Watanabe 1997(43) | Balb/c nude mice | Human Neuroblastoma Cell Line: SK-N-MC | $^{188}$Re | 90% | | |
| Zubillaga 1997(9) | 89 SD rats | NMU-induced breast carcinoma | $^{90}$Y | | | |
| Wang 1997(44) | 42 SD | Rat hepatoma: NIS1 | $^{103}$Re | | | |

| Study | Tumor volume (mm$^3$) | Control | Treated | Survival |
|-------|----------------------|---------|---------|----------|
| Nakhgevan y 1988(22) | 6.3 (1.9-7.0) | 17.4 | | |
| Brown 1991(23) | 10.5±5.8 | 20.7±4.6 | 15.6±3.84 | |
| Order 1994(40) | 25.0±10.5 | 15.6±3.84 | |
| Lee 1997(38) | 30.0±15.0 | 15.6±3.84 | |
| Nguen 1997(39) | 45.0±20.0 | 20.7±14.6 | |
| Watanabe 1997(43) | 60.0±30.0 | 15.6±3.84 | |
| Zubillaga 1997(9) | 75.0±37.5 | 15.6±3.84 | |
| Wang 1997(44) | 90.0±45.0 | 15.6±3.84 | | |

| Study | Tumor volume (mm$^3$) | Control | Treated | Mean |
|-------|----------------------|---------|---------|------|
| Nakhgevan y 1988(22) | 6.3 (1.9-7.0) | 17.4 | | 30.8 |
| Brown 1991(23) | 10.5±5.8 | 20.7±4.6 | | |
| Order 1994(40) | 25.0±10.5 | 15.6±3.84 | | |
| Lee 1997(38) | 30.0±15.0 | 15.6±3.84 | | |
| Nguen 1997(39) | 45.0±20.0 | 20.7±14.6 | | |
| Watanabe 1997(43) | 60.0±30.0 | 15.6±3.84 | | |
| Zubillaga 1997(9) | 75.0±37.5 | 15.6±3.84 | | |
| Wang 1997(44) | 90.0±45.0 | 15.6±3.84 | | |
| Study      | Year | Species | Disease Model | Activity | Tumor Size Reduction | Survival |
|------------|------|---------|---------------|----------|----------------------|----------|
| Zubillaga  | 1998 | 70 SD rats | NMU-induced breast carcinoma | 32P | 80% (12/15) | 80% (12/15) |
| Junfeng    | 1999 | Kunming mice | Mice sarcoma: S180 | 188Re | 77.0 | 78.3 |
| Lee        | 1999 | Nude mice | Human pancreatic carcinoma: AsPC-1 | 32P | 78.8 ± 3.0% | 78.8 ± 3.0% |
| Lee        | 1999 | Nude mice | Human pancreatic carcinoma: AsPC-1 | 32P | 80.2 ± 3.8% | 80.2 ± 3.8% |
| Liu        | 1999 | Balb/c | Human liver cancer: H-CS | 32P | 88.3 ± 10.9% | 88.3 ± 10.9% |
| Junfeng    | 2000 | Athymic nude mice | Human liver cancer: SMMC 7721 | 188Re | 50.7 ± 6.8% | 50.7 ± 6.8% |
| Lin        | 2000 | SD rats | Rat hepatoma: N1S1 | 90Y | 32.1 ± 3.4% | 32.1 ± 3.4% |
| Chen       | 2001 | SD rats | Rat hepatoma: N1S1 | 90Y | 22.3 ± 3.8% | 22.3 ± 3.8% |
| Table 5 (continued) |
|---------------------|
| Lin 2005(29) NZW rabbits Rabbit SCC: VX2 | 186Re | 72h 20.91±6.75 | Tumor volume cm³ | Survival (mean± SD) |
| MS 6.82±3.93 | 11.82±7.27 | 14 days | 65±9.8 |
| Ethanol 8.96±3.99 | 31.04±16.23 | Control 13.16±8.93 | 47.36±21.66 | 38.8±6.8 |
| Zhang 2005(28) BALB/c mice Human liver carcinoma: HepG2 Human liver carcinoma: 2119 | ³²P | | 8 weeks 67% complete response treated with 2MBq 200Gy |
| Hafezi 2007(30) SD rats Rat gliosarcoma: 9L | 186Re 188Re | | Survival at 36 days |
| Lubolt 2009(35) Wistar Rats Rat mammary carcinoma: Walker Carcinoma 256 Rat Yoshida Sarcoma | 186Re 188Re | 10 min 70% retention. 48 h 60% retention. Fast leakage depends on particle size and vascularization. Slow decline is independent. Adrenail reduced the washout of the small particles. MS accumulated mainly in the lungs, smaller colloids in the liver. |
| Bult 2012(31) NZW rabbits Rabbit SCC: VX2 | 166Ho | No holmium was detected in the feces, urine, femur and blood. |
| Bult 2013(32) 24 Balb/C mice Mice Renal cell carcinoma | 166Ho | Tumor volume cm³ |
| 3 days 0.14±0.01 | 0.12±0.03 |
| 14 days 0.10±0.01 | 0.11±0.03 |
| | 0% (0/0) |
| | 60% (3/5) |
| Bult 2013(35) 3 DS cats Various | 166Ho | Body weight, alertness, mobility, and coat condition of the animals improved markedly. |
| Li 2014(33) Nude BABL/c mice Human breast: MCF-7 | ¹³¹I | Time | ¹³¹I MS | Na¹³¹I |
| 1 h 1.83 ± 0.46 |
| 6 h 0.60 ± 0.29 |
| 24 h 43.29±5.27 0.45±0.07 |
| 48 h 0.14±0.02 |
| 72 h 0.06±0.01 |
| 4d 29.2±3.72 |
| 8d 24.7±7.28 |
| 16d 19.9±5.24 |
| | Tumor volume cm³ |
| 21d 0.63±0.39 | 0.70±0.39 |
| 4d 29.28±3.72 |
| 8d 24.7±7.28 |
| 16d 19.9±5.24 |
| Chi 2014(34) Nude BABL/c mice Human HCC: HepG2 | ¹³¹I | 1 day 39.06±63.78 | Growth rate (cm³/week) |
| 4 days 26.43±60.24 |
| 8 days 23.28±61.06 |
| 16 days 21.55±63.64 |
| 24 days 19.55±64.29 |
| | Survival 64 days |
| | 0.037±0.04 |
| | 73.3% (11/15) |
| | 0.68±0.39 |
| | 13.3% (2/15) |
improved while the risk of side effects will be reduced. Particles that emit a small amount of gamma-radiation and can be visualized with high-resolution imaging are preferred at this stage. Experiments should be performed in larger tumor models to obtain better clinical relevant data on the IT distribution. Subsequently, the threshold absorbed dose to successfully treat the tumor should be investigated. Furthermore, accurate administration requires skilled physicians and controlled injection, and will be time consuming. In the near future, with advanced technologies such as controllable needle placement and injection systems, the procedure could be performed easily, quickly, and safely for patients and personnel.

Compliance with ethical standards

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Conflict of interest  Author R.C.B. declares that he has no conflict of interest. M.G.E.H.L. is consultant for Sirtex, BTG, Mirada, and Bayer Healthcare. Author S.A.V.N. declares that he has no conflict of interest. Author A.J.W.P.R. declares that he has no conflict of interest. Author R.J.J.v.E. declares that he has no conflict of interest. Author J.F.W.N. is co-founder and scientific director of Quirem Medical and has a minority share in the company Quirem Medical. Furthermore, J.F.W.N. is inventor on the patents related to the $^{166}$Ho-PLLA-microspheres that are assigned to University Medical Center Utrecht Holding BV (patent numbers WO2012060707 A1 and US 2005/0201940 A1). The Department of Radiology and Nuclear Medicine of the UMC Utrecht receives royalties from Quirem Medical BV.

Ethical approval  This article does not contain any studies with human participants or animals performed by any of the authors.

Informed consent  For this type of study, formal consent is not required.

Appendix. Search strategy

Medline

#1. Humans [Mesh]
#2. Animals [Mesh]
#3. Animal [Title/Abstract]
#4. Human [Title/Abstract]
#5. #1 OR #2 OR #3 OR #4
#6. Neoplasms [Mesh]
#7. Tumor*[Title/Abstract]
#8. Tumour*[Title/Abstract]
#9. Cancer*[Title/Abstract]
#10. #6 OR #7 OR #8 OR #9
#11. Intratumor*[Title/Abstract]
#12. Intra-tumor*[Title/Abstract]
#13. Intratumour*[Title/Abstract]
#14. Intra-tumour*[Title/Abstract]
#15. Intralesion*[Title/Abstract]
#16. Intra-lesion*[Title/Abstract]
#17. Interstitial*[Title/Abstract]
#18. #10 OR #11 OR #12 OR #13 OR #14 OR #15 OR #16 OR 17#
#19. Radiotherapy [Mesh Terms]
#20. Radiotherapy [Title/Abstract]
#21. Radioisotopes [Mesh]
#22. Isotopes [MeSH]
#23. Gold [Mesh]
#24. Gold [Title/Abstract]
#25. Lutetium [Mesh]
#26. Lutetium [Title/Abstract]
#27. Rhenium [Mesh]
#28. Rhenium [Title/Abstract]
#29. Holmium [Mesh]
#30. Holmium [Title/Abstract]
#31. Iodine [Mesh]
#32. Iodine [Title/Abstract]
#33. Yttrium [Mesh]
#34. Yttrium [Title/Abstract]
#35. Phosphorus [Mesh]
#36. Phosphorus [Title/Abstract]
#37. 32P[Title/Abstract]
#38. P32 [Title/Abstract]
#39. 32-P [Title/Abstract]
#40. P-32 [Title/Abstract]
#41. #19 OR #20 OR #21 OR #22 OR #23 OR #24 OR #25 OR #26 OR #27 OR #28 OR #29 OR #30 OR #31 OR #32 OR #33 OR #34 OR #35 OR #36 OR #37 OR #38 OR #39 OR #40 OR #41
#42. "Treatment Outcome"[Mesh]
#43. Survival [Title/Abstract]
#44. "Tissue Distribution"[Mesh]
#45. Distribut*[Title/Abstract]
#46. Safe*[Title/Abstract]
#47. Toxicity [Subheading]
#48. Toxic*[Title/Abstract]
#49. Effic*[Title/Abstract]
#50. Effec*[Title/Abstract]
#51. #42 OR #43 OR #44 OR #45 OR #46 OR #47 OR #48 OR #49 OR #50
#52. #5 AND #10 AND #18 AND #41 AND #51

EMBASE

#1. ‘in vivo study’/exp
#2. ‘human’/exp
#3. ‘animal’/exp
#4. #1 OR #2 OR #3
#5. ‘neoplasm’/exp
#6. cancer:ab,ti
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