Neuroimaging

White matter hyperintensities are more highly associated with preclinical Alzheimer’s disease than imaging and cognitive markers of neurodegeneration

Benjamin M. Kandel a,*, Brian B. Avants b, James C. Gee b, Corey T. McMillan c, Guray Erus d, Jimit Doshi d, Christos Davatzikos d, David A. Wolk c, for the Alzheimer’s Disease Neuroimaging Initiative

aPenn Image Computing and Science Laboratory and Department of Bioengineering, University of Pennsylvania, Philadelphia, PA, USA
bPenn Image Computing and Science Laboratory and Department of Radiology, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, USA
cDepartment of Neurology, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, USA
dCenter for Biomedical Image Computing and Analytics, University of Pennsylvania, Philadelphia, PA, USA

Abstract

Introduction: Cognitive tests and nonamyloid imaging biomarkers do not consistently identify preclinical AD. The objective of this study was to evaluate whether white matter hyperintensity (WMH) volume, a cerebrovascular disease marker, is more associated with preclinical AD than conventional AD biomarkers and cognitive tests.

Methods: Elderly controls enrolled in the Alzheimer’s Disease Neuroimaging Initiative (ADNI, n = 158) underwent florbetapir-PET scans, psychometric testing, neuroimaging with MRI and PET, and APOE genetic testing. Elderly controls the Parkinson’s progression markers initiative (PPMI, n = 58) had WMH volume, cerebrospinal fluid (CSF) Aβ1-42, and APOE status measured.

Results: In the ADNI cohort, only WMH volume and APOE ε4 status were associated with cerebral Aβ (standardized β = 0.44 and 1.25, P = .03 and .002). The association between WMH volume and APOE ε4 status with cerebral Aβ (standardized β = 1.12 and 0.26, P = .048 and .045) was confirmed in the PPMI cohort.

Discussion: WMH volume is more highly associated with preclinical AD than other AD biomarkers.

© 2016 The Authors. Published by Elsevier Inc. on behalf of the Alzheimer’s Association. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Keywords: Alzheimer’s disease; Aging; MRI; PET; White matter; Leukoaraiosis; Preclinical Alzheimer’s disease; Amyloid

1. Introduction

Owing to the recent failures of several clinical trials in treating symptomatic Alzheimer’s disease (AD) [1], focus in therapeutic trials is shifting from reversing the effects of AD to preventing cognitive decline due to AD at the preclinical stage, before any noticeable cognitive change has occurred [2]. Preclinical AD is defined based on the presence of cerebral amyloidosis, detected by either amyloid PET or measurement of cerebrospinal Aβ1-42 [3]. We focus here on preclinical AD, which is simply defined as presence of cerebral Aβ [3]. Presence of preclinical AD does not
necessarily imply that clinical AD will result but does appear to come with a higher risk of developing clinical AD [4]. Because of the importance of preclinical AD, an accurate and thorough understanding of the cognitive and brain changes at this stage is critical. Furthermore, predictors of preclinical AD are potentially valuable in the context of clinical trials to enrich populations before the use of more expensive or invasive amyloid measurement.

Within cognitively normal older adults, two predictors of amyloid status have already been relatively well established: age and apolipoprotein E (APOE) status [5,6]. Beyond these risks, it is possible that other neurodegenerative biomarkers and cognitive changes that presumptively represent the downstream effect of the presence of cerebral Aβ, such as hippocampal atrophy, hypometabolism, and subjective cognitive impairment, may also be sensitive to preclinical AD [7–9]. Although these markers clearly predict conversion from mild cognitive impairment (MCI) to probable AD [10–14] and the presence of cerebral amyloid in MCI to varying degrees [15], their value in preclinical disease is less well established.

One neuroimaging measure that has received less, but growing, attention in relationship to AD is the presence of white matter hyperintensity (WMH) volume. WMH volume has been associated with clinical AD [16,17], cognitive ability [18], cortical atrophy [19], and AD pathology in cognitively normal populations [20], but no study has examined the association of WMH volume with preclinical AD in the context of more established imaging and cognitive AD biomarkers. Here, we compare the association of a variety of biomarkers, including neurodegenerative, genetic, functional, and cognitive biomarkers, as well as WMH volume, with preclinical AD. This comparison sheds light on the pathogenesis of AD and can inform subsequent studies on longitudinal trajectories of AD biomarkers.

2. Methods

2.1. Clinical data

2.1.1. Subjects

Data used in the preparation of this article were obtained from two publicly available data repositories: the Alzheimer’s Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu) and the Parkinson’s Progression Markers Initiative (PPMI) database (www.ppmi-info.org/data). The ADNI was launched in 2003 by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, the Food and Drug Administration, private pharmaceutical companies and nonprofit organizations, as a $60 million, 5-year public-private partnership. The primary goal of ADNI has been to test whether serial magnetic resonance imaging (MRI), positron emission tomography (PET), other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of MCI and early Alzheimer’s disease (AD). For up-to-date information on the PPMI study, visit www.ppmi-info.org.

Data used in this article were downloaded from the ADNI website in November 2014. We included all cognitively normal subjects from ADNI2 and ADNI-GO who had undergone flortetapir-PET scans to obtain a measure of cerebral amyloidosis, APOE genotyping, FDG-PET, structural magnetic resonance (MR) imaging, and all cognitive tests examined. Only subjects with Freesurfer cortical and hippocampal segmentations judged acceptable by the structural MR processing core were included. Inclusion criteria for the study and diagnostic criteria for establishing disease state were as previously reported [21]. For up-to-date information on specific inclusion and exclusion criteria, please see www.adni-info.org. Data were also downloaded from the PPMI website, October 2014. Inclusion criteria for these study data included a baseline diagnosis of cognitively normal, a T1-weighted and Flair MRI, CSF analysis of AD biomarkers, and APOE genotyping. For up-to-date information on the PPMI study, visit www.ppmi-info.org.

2.2. Psychometric testing

The following measures were included in the analysis: the mini-mental state examination [22], Rey Auditory Verbal Learning Test [23], immediate and delayed recall of the Logical Memory Test [24], the Trail Making Test [trails A and trails B] [25], category fluency [animals [26]], and Boston Naming Test [27]. Given the importance of memory in prodromal AD, we examined several of the AVLT measures, which depend on differential aspects of episodic and working memory [28]. For the present study, we analyzed performance on the fifth immediate memory trial (AVLT Trial 5 Recall), 5-minute and 30-minute delayed recall (AVLT 5-min Recall, AVLT 30-min Recall), and recognition memory discrimination (AVLT recognition discrimination). To account for false alarms to nonstudied items, we calculated a measure of discriminability, d-prime (d’), in a standard fashion [29].

In addition to psychometric measures, we also examined a measure of cognitive complaints via the Everyday Cognition (ECog) questionnaire [30,31], using both informant-report and self-report data. Informants and participants are separately queried as to the degree to which particular everyday functioning has changed compared to 10 years earlier. Responses for ADNI were obtained on a five-point scale, with increasing values indicating more complaints and 5 indicating “do not know”. The global scores were averaged separately over informant-rated and self-rated scales, excluding values of 5.

2.3. Determination of amyloid status

Florbetapir-PET was administered in accordance with the ADNI PET protocols available online (http://adni.loni.usc.edu/data-samples/pet), and image processing...
was performed by the ADNI core laboratory as described previously [32]. Briefly, a PET scan was acquired 50–70 minutes after injection of florbetapir. Images were smoothed and aligned to an MPAGE anatomic image to obtain a cortical segmentation. Mean florbetapir uptake in lateral and medial frontal, anterior and posterior cingulate, lateral parietal, and lateral temporal regions was normalized to uptake in the cerebellum to obtain a mean cortical standardized uptake value ratio (SUVr). Cortical florbetapir uptake of $\geq 1.11$ was considered “positive” for cortical Aβ [32]. APOE genotyping was performed as described on the ADNI website (http://adni.loni.usc.edu/data-samples/genetic-data/).

2.4. Neuroimaging biomarkers

Processing of neuroimaging data was performed by ADNI cores and made publicly available. FDG-PET scans were acquired and analyzed in accordance with a standard protocol [33]. Mean FDG uptake was averaged >5 ROI’s that are sensitive to AD-related changes in metabolism, including right and left angular gyrus, right and left inferior temporal regions, and bilateral posterior cingulate. Cortical thickness and hippocampal volume measurement based on MRI scans were performed according to the standard ADNI Freesurfer [34] processing pipeline and downloaded from the ADNI website. Only images that passed ADNI quality control for the temporal, occipital, and parietal lobe were included. Cortical thickness in the caudal portion of the middle frontal gyrus, medial portion of the orbital frontal cortex, inferior parietal lobule, lateral portion of the occipital cortex, inferior temporal gyrus, entorhinal cortex, temporal pole, and the isthmus of the cingulate cortex were averaged to form a meta-ROI thought sensitive to early AD-related neurodegeneration, as previously suggested [35]. WMH volumes were computed by the ADNI core laboratory in accordance with previously published protocols [36]. Briefly, FLAIR MR images were corrected for inhomogeneity and warped to T1 images to provide a segmentation. WMHs are seeded at points that are $>3.5$ standard deviations from the mean signal in white matter, and final segmentation is based on a Bayesian approach, combining spatial priors and tissue class constraints. The WMH segmentation also included segmentations of white matter, gray matter, and CSF; the sum of the tissue volumes was used as a surrogate for intracranial volume. For analysis, WMH volumes were normalized to intracranial volume and transformed using the natural logarithm.

2.5. PPMI analysis

To analyze evidence for the presence of evidence for cerebral amyloidosis, we evaluated CSF amyloid-beta (Aβ1–42). PPMI has completed two CSF analyses, Project 101 and Project 103, and overall Aβ1–42 were significantly elevated in the latter. To maximize sample sizes, we adapted a linear regression transformation method for CSF Aβ1–42 to transform the elevated values from Project 103 to match values from Project 101, as previously reported [37]: Transformed $Aβ_{1-42} = 1.82994 + (Aβ_{1-42} \times 0.61562)$. In our transformed CSF series, we observed that a $Aβ_{1-42}$ CSF cutoff of 198 pg/mL marked the first point of deviation from the normal distribution of $Aβ_{1-42}$ CSF values, so we selected this cutoff to classify participants as being positive or negative for CSF Aβ.

We applied an automated MR image processing pipeline for quantifying WMH volume in the PPMI cohort. The T1-weighted scan of each subject was first preprocessed for correction of intensity inhomogeneities [38]. A multiatlas skull-stripping algorithm was applied using study-specific atlases for the extraction of the brain tissue [39]. Images with quality issues, such as low T1 resolution, were excluded from the analysis and brain masks with errors were manually corrected. A multi-atlas label-fusion method, which uses nonlinear registration for transferring atlas labels to subject space, was applied to form the basis of the white and gray matter segmentations [40,41]. Regions of WMH were segmented using a multimodal segmentation method, white matter lesion segmentation (WMLS), using T1-weighted and fluid-attenuated inversion recovery (FLAIR) images [42]. WMLS is a supervised learning method that trains on lesions manually delineated by an expert radiologist. The lesion segmentation involves data preprocessing via histogram standardization and co-registration, feature extraction, training a voxelwise discriminative model, voxelwise label assignment, and false-positive elimination. Quality control was performed on final volumetric data by overlaying each subject’s lesion map on the FLAIR image. None of the lesion masks had errors that would require exclusion. There were minor errors, particularly in the determination of the boundaries of large lesions. However, we did not prefer to correct them manually, as the intra-rater and inter-rater variability associated with manual delineations could potentially bias the results.

2.6. Statistical analysis

All statistical analyses were performed using the R programming language, version 3.1.0. Two-tailed two-sample t tests with unequal variances (Welch’s t test) were used to assess differences in demographic characteristics between WMH positive and WMH negative subjects. Logistic regression using a logit link function was used to assess the relationship between white matter hyperintensities and presence of cerebral Aβ. Stepwise forward regression was performed to generate an ideal multivariate linear model, using the Bayesian Information Criterion to regularize the model [43]. For all analyses, patient age, gender, and education were used as covariates. For hippocampal volume, intracranial vault volume (ICV) was used as an additional covariate.
3. Results

3.1. Subject demographics

A total of 184 cognitively normal subjects with florbetapir-PET were identified from the ADNI database. Of these, 155 subjects had complete psychometric and imaging variables as described in Methods, including acceptable cortical and hippocampal segmentations. A summary of the demographics of the study population, including the psychometric and imaging information, is given in Table 1. Aβ+ subjects were slightly older (M = 75.1, standard deviation (SD) = 5.7) than Aβ− subjects (M = 72.5, SD = 6.1), t(100) = 2.4, P = .02 and tended toward having slightly less education (for Aβ+ subjects, M = 16 and SD = 2.4 years; for Aβ− subjects, M = 17 and SD = 2.5 years; t(95) = −2, P = .05).

3.1.1. Associative models

We computed a logistic regression relating each psychometric test and modality with Aβ status, while covarying for age, gender, and education (Table 2). The logistic regression results indicated that the best univariate predictor of cerebral Aβ was APOE ε4 status, followed by white matter hyperintensity (WMH) volume. All other imaging and cognitive measures, including FDG-PET, hippocampal volume, ECOG, and any AVLT measure, were not significant predictors of Aβ status. WMH volume was not increased in APOE ε4-carrying subjects (t(85.8) = −0.018, P = .99). The independence between WMH volume and Aβ status was confirmed by running a stepwise forward multivariate regression model, which selected only WMH volume and APOE status as independent predictors. A boxplot comparing WMH volumes in Aβ+ and Aβ− subjects is shown in Fig. 1. There was no significant association between WMH and either τ or τ/Aβ ratio (P = .56 and .09, n.s.).

3.1.2. Replication in PPMI data

Owing to conflicting results in prior studies about the link between WMH volume and Aβ pathology, we sought to replicate correlation between WMH volume and Aβ pathology with elderly controls from the Parkinson’s progression markers initiative. We identified 240 subjects with FLAIR and T1 images. A total of 207 images passed manual quality control; of these, 58 were control subjects. Log WMH volume normalized to ICV was not associated with age (ρ = 0.12, P = .37) or gender (t(38) = −1.6, P = .12). A summary of the demographics of the study population is given in Table 3. In contrast to the ADNI cohort, WMH volume was significantly increased in APOE ε4 carriers (t(24) = 2.3, P = .03). After correcting for age and gender, WMH volume was significantly predictive of Aβ status (β estimate 1.12 ± 0.57, z = 1.97, P = .048), even given the limited sample size. A boxplot showing WMH volumes in the PPMI cohort is shown in Fig. 2. However, in the PPMI cohort, a stepwise forward regression model included only APOE ε4 status as a predictor of Aβ and did not include WMH.

4. Discussion

This study represents the first comprehensive comparative evaluation of a variety of biomarkers to predict...
The presence of preclinical AD pathology in a cognitively normal population. In particular, the surprising finding that WMHs are more highly correlated with AD pathology than any of the standard AD imaging biomarkers or cognitive tests suggests that in the earliest stages of AD, vascular disease, as reflected by WMH, may play a significant role in the development of cerebral amyloidosis or be an early downstream effect of this molecular pathology. The replication of this result in an independent data set, using different image processing techniques, points to the robustness of the finding. The salient association of WMH with preclinical AD supports earlier studies [44–46] that have demonstrated a link between WMH and cognitive decline at this stage [44]. The strength of this association is in contrast to conventional thinking about this disease stage which assumes that biomarkers and, perhaps, subtle cognitive symptoms traditionally used to characterize MCI due to AD and probable AD are the same ones that would characterize preclinical AD [3,47]. Indeed, AD biomarker cascade models do not generally include a measure of white matter integrity.

4.1. White matter hyperintensities and amyloid

The degree of association between WMH volume and amyloid deposition in nondemented control subjects is controversial. Some studies have not found a link between WMH volume or other vascular disease markers and amyloid [48–52], whereas other studies, including a large (n = 337) Amsterdam study [20,53,54], have reported such a correlation. It is possible that differences in WMH calculation, such as WM histogram normalization, thresholds for defining WMH, and the incorporation of priors for segmenting WMHs, may at least partially account for these discrepancies. Standardization and rigorous comparison of competing methods for evaluating WMH volume may help to reduce the variation in results. Differential involvement of periventricular and subcortical WMH may also contribute to differing study results [55], although the relative importance of WMH anatomic distribution as compared to total WMH volume is still a matter of debate [36].

The relationship of cerebrovascular disease to clinical AD and amyloid pathology is likely complex, and an evolving understanding is emerging [56,57], but there are clear links between the severity of cerebrovascular disease and the risk of clinical dementia associated with AD pathology [58]. There are a number of overlapping risk factors for cerebrovascular disease and WMH with AD,

Table 2
Summary of univariate logistic regressions predicting Aβ status from each psychometric test and imaging biomarker for ADNI subjects.

| Variable                        | Standardized \( \beta \) estimate | Standard error | \( z \) value | \( P \) value |
|---------------------------------|-----------------------------------|----------------|---------------|--------------|
| AVLT trial 5 recall            | -0.09                             | 0.17           | -0.53         | .60          |
| AVLT 5-min recall              | -0.29                             | 0.18           | -1.63         | .10          |
| AVLT 30-min recall             | -0.17                             | 0.17           | -0.95         | .34          |
| Trail making test A            | 0.19                              | 0.17           | 1.14          | .26          |
| Trail making test B            | 0.04                              | 0.17           | 0.21          | .83          |
| Boston naming test             | 0.05                              | 0.17           | 0.29          | .77          |
| Category fluency (animals)     | 0.24                              | 0.18           | 1.34          | .18          |
| Mini-mental status examination | 0.08                              | 0.18           | 0.47          | .64          |
| Discrimination                 | -0.01                             | 0.17           | -0.08         | .94          |
| Logical memory                 | -0.02                             | 0.17           | -0.11         | .91          |
| Logical memory, delayed        | -0.13                             | 0.17           | -0.74         | .46          |
| Subject-reported ECOG score    | -0.10                             | 0.18           | -0.58         | .56          |
| Log white-matter hyperintensity volume | 0.44                             | 0.20           | 2.19          | .028*        |
| Mean FDG-PET SUVR of AD meta-ROI | -0.18                         | 0.17           | -0.77         | .44          |
| Hippocampal volume             | -0.13                             | 0.17           | -0.77         | .44          |
| Mean cortical thickness of AD meta-ROI | 0.01                             | 0.17           | 0.07          | .94          |
| APOE ε4 status                 | 1.25                              | 0.40           | 3.15          | .002**       |

Age, gender, and education level (in years) were included as covariates. All data were scaled before regression to facilitate inspection of regression coefficients. The only variables significant at the \( P = .05 \) level were APOE status and log white-matter hyperintensity volume.

\* \( P < .05 \).
** \( P < .005 \).

Fig. 1. Boxplots comparing white matter hyperintensity volumes for the ADNI cohort, normalized to intracranial volume, and log transformed. Aβ+ subjects had significantly higher WMH volumes than Aβ− subjects.
including hypertension, diabetes, obesity, and tobacco use. For example, poorly controlled hypertension has been found to correlate with amyloid plaque pathology [59], and circle of Willis atherosclerosis is more highly related to AD pathology than other common proteinopathies [60,61]. Other work has related blood pressure and systemic arterial stiffness, also associated with WMH, to amyloid burden measured by amyloid imaging [62,63]. We did not observe statistically significant differences between mean arterial pressure, systolic, and diastolic blood pressure, body mass index, or history of cardiovascular disease, hypertension, or smoking between Aβ+ and Aβ− subjects in the ADNI cohort. Although we did not have complete medical histories of all subjects in the PPMI cohort, we did not observe differences in blood pressure between Aβ+ and Aβ− subjects in the PPMI cohort either.

Whether these are common risk factors with dissociable associations or are more directly mechanistically related is unclear. WMH volume may be a marker of cerebral amyloid due to the increased likelihood of deposition of amyloid in vessel walls leading to development of cerebral amyloid angiopathy [64]. Alternatively, there are a number of potential mechanisms by which cerebrovascular changes may directly relate to deposition of cerebral amyloid. For example, Aβ clearance may depend of the perivascular “glymphatic” system which is diminished by reduced arterial pulsatility or stiffness [65]. Furthermore, a number of factors associated with cerebrovascular alterations, including hypoperfusion, may accelerate Aβ production [66]. An enhanced understanding of these linkages may provide vascular specific therapeutic options at presymptomatic stages.

4.2. Independent association of white matter hyperintensities and APOE status with cerebral amyloid

The finding that WMH and APOE ε4 carrier status are both independently associated with amyloid, and that WMH volume is not associated with APOE implies that vascular burden increases the risk for cerebral amyloid over and above this highly significant genetic risk. This finding is in consonance with other studies that have shown that WMH volume is an independent risk factor for incident dementia [67], although not all studies have supported this result [52]. The independence of these factors is particularly interesting given the relationship of APOE with cardiovascular risk [68,69]. Nonetheless, it is worth noting that we did not observe the same dissociation in the PPMI data set, although this could be due in part to an issue of power.

4.3. Lack of relationship with traditional AD neurodegenerative biomarkers and cognitive measures

The current findings did not support the role of neuroimaging biomarkers that have more traditionally been associated with AD in the prodromal and dementia stages of disease, including hippocampal volumes, cortical thickness, and FDG-PET. Although some prior studies have found associations between cortical thinning or volume changes and amyloid status in cognitively normal individuals [70–74], this has not been a consistent finding [75]. The differences in results may be due to methodologic differences, with some studies using rate of atrophy and others cross-sectional measures. The observed inconsistency may also be due to the heterogeneity among the proposed “stages” of preclinical AD, which include a spectrum of neurodegeneration and cognitive

Table 3
Summary of PPMI cohort demographics

| Characteristic                  | All subjects (mean ± standard deviation) | Aβ+ | Aβ− |
|--------------------------------|-----------------------------------------|-----|-----|
| Number of subjects             | 58                                      | 12  | 46  |
| Number of males                | 35                                      | 7   | 28  |
| Age (y)                        | 60 ± 13                                 | 62 ± 18 | 60 ± 11 |
| Number of APOE ε4 carriers     | 12                                      | 5   | 7   |
| White-matter lesion volume (mm³) | 2654 ± 5757                             | 4757 ± 7509 | 2105 ± 5168 |

Fig. 2. Boxplots comparing log white matter hyperintensity volumes for the PPMI cohort, normalized to intracranial volume and log transformed. As in the ADNI cohort, Aβ+ subjects had significantly higher WMH volumes than Aβ− subjects.
change [3]. This also may explain why in some instances cognitive measures have also been reported to be associated with preclinical AD [8,76,77]. It is worth pointing out that while not significant, hippocampal volume was smaller and 5-minute delayed recall on the AVLT was poorer in the group with evidence of cerebral amyloid. Finally, despite work suggesting a link between subjective complaints and amyloid status [7], we did not find either the patient or informant-based ECog of value for predicting preclinical AD; this finding resonates with results from a recent meta-analysis of amyloid status in cognitively normal adults with and without subjective symptoms [5].

Although small vessel disease is typically associated with executive dysfunction, we did not observe an association between executive function and WMH volume. This lack of association may be because we studied only cognitively normal subjects, so the variance in cognitive ability is relatively limited. It also may be the case that white matter findings associated with cerebral amyloid do not have the same impact on executive function as those unassociated with amyloid and instead accompanied by more pervasive cerebrovascular disease.

4.4. Limitations

Our study is limited by the cross-sectional nature of the data, and a longitudinal study looking at the relative timing of the development of WMH and cerebral amyloid would be informative of the direction of causality in this relationship. The characteristics of the ADNI cohort may impact the generalizability of the findings. The ADNI cohort is racially and socioeconomically homogeneous and is relatively free of cardiovascular disease and other comorbidities. It is uncertain whether the relationship between WMH and Aβ would be strengthened or weakened in a more heterogeneous cohort with more prevalent cardiovascular pathology. In addition, the lack of standardization both of FLAIR imaging methods, including resolution and WMH quantification techniques makes comparisons of different populations difficult. The clinical applicability of WMH quantification would be greatly enhanced by a standardized measurement method. Additional replication in other cohorts would bolster confidence in the observed association. Although manual quality control was performed on all hippocampal and WMH segmentations to avoid major failures, it is possible that minor errors affected the results. Finally, although cerebral Aβ has been adopted as the defining feature of preclinical AD, the lack of complete longitudinal data on ADNI 2/GO precludes any definitive conclusion on the impact of WMH on developing clinical AD in the future.

5. Conclusion

In our samples of cognitively normal controls, white matter hyperintensities (WMHs) are more highly associated with biomarker evidence of cerebral Aβ than any other recognized biomarker or cognitive test. This finding challenges the assumption that biomarkers of neurodegeneration, which are well established in later stages of disease, are reliably sensitive at the preclinical stage. As the preclinical stage of AD is increasingly recognized as a period of primary importance for preventing and ameliorating incipient neurodegeneration, the importance of accurately understanding correlates of AD pathology at this stage similarly increases. At the same time, the biological mechanism of the correlation between Aβ and WMH is not clear. A more thorough understanding of the nature of the relationship between Aβ and WMH is necessary to establish potential targets for disease-modifying interventions. Nevertheless, it is clear that WMH should be considered as a potential biomarker for preclinical AD in addition to more widely used cognitive tests and imaging biomarkers.

Acknowledgments

Author contributions: B.M.K. and D.A.W. had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. D.A.W. and B.M.K. contributed to study concept and design. B.M.K., D.A.W., B.B.A., J.C.G., C.D., C.M., G.E., and J.D. performed the analysis and interpretation of data. B.M.K. and D.A.W. drafted the article. B.M.K., D.A.W., B.B.A., J.C.G., and C.D. performed the critical revision of the article for important intellectual content. B.M.K. and D.A.W. performed the statistical analysis of data. D.A.W., J.C.G., and C.D. obtained funding. D.A.W. and J.C.G. provided the administrative, technical, or material support. D.A.W. supervised the study.

Study funding: This research was supported by NIH grants T32-EB009384, K01-ES025432-01, R01-AG14971, R01-NS061906, AG010124, AG043503, AG040271, and the Berkman Charitable Trust. Joint funding was provided by the Michael J. Fox Foundation for Parkinson’s Research, Alzheimer’s Association, and Weston Brain Institute (BAND-9665). D.A.W. has performed consulting for GE Healthcare, Inc. B.M.K., B.B.A., J.C.G., C.T.M., J.M., C.D., and G.E. have no disclosures to report. The funding agencies had no say in the design and conduct of the study; collection, management, analysis, and interpretation of the data; or preparation, review, and approval of the article.

Data collection and sharing for this project was funded by the Alzheimer’s Disease Neuroimaging Initiative (ADNI) (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: Alzheimer’s Association; Alzheimer’s Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen Idec Inc.; Bristol-Myers Squibb Company; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd; and Medivation, Inc., Novartis Pharmaceuticals Corp.; Pfizer Inc.; Servier; Takeda; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). Public private partnership funds are provided by the Federal Bureau of Investigation’s Scientific Research and Development Fund.
and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Medpace, Inc.; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Synarc Inc.; and Takeda Pharmaceutical Company. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer’s Disease Cooperative Study at the University of California, San Diego. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California.

PPMI, a public-private partnership, is funded by the Michael J. Fox Foundation for Parkinson’s Research and funding partners, including Abbvie, Avid, Biogen, Bristol-Myers Squibb, Covance, GE Healthcare, Genentech, GlaxoSmitKline, Lilly, Lundbeck, Merck, Meso Scale Discovery, Pfizer, Piramal, Roche, Servier, and UCB.

**RESEARCH IN CONTEXT**

1. Systematic review: We reviewed the literature using PubMed and Google Scholar searches, as well as meeting abstracts and presentations. There has been an increasing interest in the contribution of vascular disease to Alzheimer’s Disease in general, and several recent studies have specifically looked at white matter hyperintensity (WMH) volume in the context of preclinical Alzheimer’s disease. These studies are referenced.

2. Interpretation: We found that WMH volume is more highly associated with preclinical Alzheimer’s disease than any conventional biomarker. This finding supports the notion that vascular disease, as marked by WMH, is associated with cerebral amyloid deposition at early disease phases, perhaps preceding other downstream neurodegenerative changes.

3. Future directions: This work raises several questions for future research. First, although WMH volume is an important marker of small vessel disease, there may be other markers of vascular health that are more sensitive to preclinical Alzheimer’s. Second, and most importantly, the direction of causality between cerebrovascular disease and amyloid deposition is uncertain and deserves intensive study given implications for potential preventative interventions.

**References**

[1] Sperling RA, Jack CR, Aisen PS. Testing the Right Target and Right Drug at the Right Stage. Sci Transl Med 2011;3:111cm33.
[2] Sperling RA, Rentz DM, Johnson KA, Karlwash J, Donohue M, Salmon DP, et al. The A4 Study: Stopping AD before symptoms Begin? Sci Transl Med 2014;6:228613.
[3] Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, et al. Toward defining the preclinical stages of Alzheimer’s disease: recommendations from the National Institute on Aging-Alzheimer’s Association workgroups on diagnostic guidelines for Alzheimer’s disease. Alzheimers Dement 2011;7:280–92.
[4] Vos SJ, Xiong C, Visser PJ, Jasielec MS, Hassenstab J, Grant EA, et al. Preclinical Alzheimer’s disease and its outcome: a longitudinal cohort study. Lancet Neurol 2013;12:957–65.
[5] Jansen WJ, Ossenkoppele R, Knol DL, Tijms BM, Scheltens P, Verhey FR, et al. Prevalence of cerebral amyloid pathology in persons without dementia: A meta-analysis. JAMA 2015;313:1924–38.
[6] Morris JC, Roe CM, Xiong C, Fagan AM, Goate AM, Holtzman DM, et al. APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. Ann Neurol 2010;67:122–31.
[7] Amariglio RE, Becker JA, Carmasins J, Wadsworth LP, Lorias N, Sullivan C, et al. Subjective cognitive complaints and amyloid burden in cognitively normal older individuals. Neuropsychologia 2012;50:2880–6.
[8] Wolk DA, Mancuso L, Kliot D, Arnold SE, Dickerson BC. Familiarity-based memory as an early cognitive marker of preclinical and prodromal AD. Neuropsychologia 2013;51:1094–102.
[9] Papp KV, Amariglio RE, Mormino EC, Hedden T, Dekhytar M, Johnson KA, et al. Free and cued memory in relation to biomarker—based marker of Alzheimer’s disease. Neuropsychologia 2015;73:169–75.
[10] Killiany RJ, Gomez-Isa I, Moss M, Kikinis R, Sandor T, Jolesz F, et al. Use of structural magnetic resonance imaging to predict who will get Alzheimer’s disease. Ann Neurol 2000;47:430–9.
[11] Davatzikos C, Bhattacharyya S, Batmanghelich KN, Trojanowski JQ. Prediction of MCI to AD conversion, via MRI, CSF biomarkers, and pattern classification. Neurobiol Aging 2011;32:2322.e19–27.
[12] Eckerström C, Olsson E, Bjerke M, Malmgren H, Edman A, Wallin A, et al. A combination of neuropsychological, neuroimaging, and cerebrospinal fluid markers predicts conversion from mild cognitive impairment to dementia. J Alzheimers Dis 2013;36:421–31.
[13] Ewers M, Walsh C, Trojanowski JQ, Shaw LM, Petersen RC, Jack CR Jr, et al. Prediction of conversion from mild cognitive impairment to Alzheimer’s dementia based upon biomarkers and neuropsychological test performance. Neurobiol Aging 2012;33:1203–1214.
[14] Gomar JJ, Bubbs-Bascaran MT, Conejero-Goldberg C, Davies P, Goldberg TE, Alzheimer’s Disease Neuroimaging Initiative. UTility of combinations of biomarkers, cognitive markers, and risk factors to predict conversion from mild cognitive impairment to Alzheimer disease in patients in the Alzheimer’s Disease Neuroimaging Initiative. Arch Gen Psychiatry 2011;68:961–9.
[15] Kandel BM, Avants BB, Gee JC, Arnold SE, Wolk DA. Neuropsychological testing predicts cerebrospinal fluid amyloid-β in mild cognitive impairment. J Alzheimers Dis 2015;46:901–12.
[16] Yoshita M, Fletcher E, Harvey D, Ortega M, Martinez O, Mungas DM, et al. Extent and distribution of white matter hyperintensities in normal aging, MCI, and AD. Neurology 2006;67:2192–8.
[17] Provenzano FA, Muraskin J, Tosto G, Narkhede A, Wasserman BT, Griffith EY, et al. White matter hyperintensities and cerebral amyloidosis: Necessary and sufficient for clinical expression of Alzheimer disease? JAMA Neurol 2013;70:455–61.
[18] Son SJ, Lee KS, Lee Y, Baek JH, Choi SH, Na DL, et al. Association between white matter hyperintensity severity and cognitive impairment according to the presence of the apolipoprotein E (APOE) e4 allele in the elderly: retrospective analysis of data from the CREDO study. J Clin Psychiatry 2012;73:1555–62.
Barnes J, Carmichael OT, Leung KK, Schwarz C, Ridgway GR, Bartlett JW, et al. Vascular and Alzheimer’s disease markers independently predict brain atrophy rate in Alzheimer’s Disease Neuroimaging Initiative controls. Neurobiol Aging 2013;34:1996–2002.

Kester ML, Goos JD, Teunissen CE, Benedictus MR, Bouwman FH, Wattjes MP, et al. Associations between cerebral small-vessel disease and Alzheimer disease pathology as measured by cerebrospinal fluid biomarkers. JAMA Neurology 2014;71:855–62.

Petersen RC, Aisen PS, Beckett LA, Donohue MC, Gamst AC, Harvey DJ, et al. Alzheimer’s Disease Neuroimaging Initiative (ADNI): clinical characterization. Neurology 2010;74:201–9.

Folstein MF, Folstein SE, McHugh PR. “Mini-mental state”: A practical method for grading the cognitive state of patients for the clinician. J Psychiatr Res 1975;12:189–98.

Rey A. L’examen clinique en psychologie. Paris: Presses universitaires de France; 1964.

Wechsler D. Wechsler Memory Scale — Revised Manual. New York: The Psychological Corporation, Harcourt Brace Jovanovich, Inc.; 1987.

Reitan RM. Validity of the trail making test as an indicator of organic brain damage. Percept Mot Skills 1958;8:271–6.

Butters N, Granholm E, Salmon DP, Grant I, Wolfe J. Episodic and semantic memory: A comparison of amnestic and demented patients. J Clin Exp Neuropsychol 1987;9:479–97.

Kaplan E, Goodglass H, Weintraub S, Goodglass H. Boston Naming Test. Philadelphia: Lea & Febiger; 1983.

Wolk DA, Dickerson BC. Fractionating verbal episodic memory in Alzheimer’s disease. Neuroimage 2011;54:1530–9.

Snodgrass JG, Corwin J. Pragmatics of measuring recognition memory: Applications to dementia and amnesia. J Exp Psychol Gen 1988;117:34–50.

Farias ST, Mungas D, Reed BR, Cahn-Weiner D, Jagust W, Baynes K, et al. The measurement of everyday cognition (ECog): Scale development and psychometric properties. Neuropsychology 2008;22:531–44.

Edmonds EC, Delano-Wood L, Galasko DR, Salmon DP, Bondi MW. Subjective Cognitive Complaints Contribute to Misdiagnosis of Mild Cognitive Impairment. J Int Neuropsychol Soc 2014;20:836–47.

Landau SM, Mintun MA, Joshi AD, Koeppe RA, Petersen RC, Aisen PS, et al. Amyloid deposition, hypometabolism, and longitudinal cognitive decline. Ann Neurol 2012;72:587–86.

Landau SM, Harvey D, Madison CM, Koeppe RA, Reiman EM, Foster NL, et al. Associations between cognitive, functional, and FDG-PET measures of decline in AD and MCI. Neurobiol Aging 2011;32:1207–18.

Dale AM, Fischl B, Sereno MI. Cortical surface-based analysis. I. Segmentation and surface reconstruction. Neuroimage 1999;9:179–94.

Desikan RS, Sabuncu MR, Schmansky NJ, Reuter M, Cabral HJ, Hess CP, et al. Vascular burden and Alzheimer disease pathologic progression. Neurology 2011;79:1349–55.

Thamprasertskul S, Martinez-Ramirez S, Pontes-Neto OM, Ni J, Ayres A, Reed A, et al. Posterior white matter disease distribution as a predictor of amyloid angiopathy. Neurology 2014;83:794–800.

Lo KY, Sotiras A, Paragios N, Davatzikos C. DRAMMS: Deformable registration via attribute matching and mutual-saliency weighting. Med Image Anal 2011;15:622–39.

Zacharakis EI, Kanterakis S, Bryan RN, Davatzikos C. Measuring brain lesion progression with a supervised tissue classification system. Med Image Comput Comput Assist Interv 2008;11:620–7.

Schwarz G. Estimating the dimension of a model. Ann Stat 1978;6:461–4.

Silbert LC, Dodge HH, Perkins LG, Sherbakov L, Lahna D, Ertens-Lyons D, et al. Trajectory of white matter hyperintensity burden preceding mild cognitive impairment. Neurology 2012;79:741–7.

Zhuang L, Sachdev PS, Trolle JN, Kochan NA, Reppermund S, Brodaty H, et al. Microstructural white matter changes in cognitively normal individuals at risk of amnestic MCI. Neurology 2012;79:748–54.

Carmichael O, Mungas D, Beckett L, Harvey D, Farias ST, Reed B, et al. MRI predictors of cognitive change in a diverse and carefully characterized elderly population. Neurobiol Aging 2012;33:83–95.

Jack CR, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, et al. Tracking pathophysiological processes in Alzheimer’s disease: an updated hypothetical model of dynamic biomarkers. Lancet Neurol 2013;12:207–16.

Hedden T, Mormino EC, Amariglio RE, Younger AP, Schultz AP, Becker JA, et al. Cognitive profile of amyloid burden and white matter hyperintensities in cognitively normal older adults. J Neurosci 2012;32:16233–42.

Marchant NL, Reed BR, DeCarli CS, Madison CM, Weiner MW, Chui HC, et al. Cerebrovascular disease, beta-amyloid, and cognition in aging. Neurobiol Aging 2012;33:1006.e25–36.

Rutten-Jacobs LC, de Leeuw FE, Geurts-van Bon L, Gordinou de Gouville MC, Schepens-Franke AN, Dederen PJ, et al. White Matter Lesions Are Not Related to Beta-Amyloid Deposition in an Autopsies-Based Study. Curr Gerontol Geriatr Res 2011;2011:e826862.

Tanskanen M, Kalaria RN, Notkola IL, Mäkelä M, Polvikoski T, Mällykangas L, et al. Relationships between white matter hyperintensities, cerebral amyloid angiopathy and dementia in a population-based sample of the oldest old. Curr Alzheimer Res 2013;10:1090–7.

Lo KY, Jagust WJ, Alzheimer’s Disease Neuroimaging Initiative. Vascular burden and Alzheimer disease pathologic progression. Neurology 2012;79:1349–55.

Schnakenberg S, Martinez-Ramirez S, Pontes-Neto OM, Ni J, Ayres A, Reed A, et al. Posterior white matter disease distribution as a predictor of amyloid angiopathy. Neurology 2014;83:794–800.

Kling MA, Trojanowski JQ, Wolk DA, Lee VM, Arnold SE. Vascular burden and Alzheimer disease pathologic progression. Neurology 2011;77:759–65.

McMillan C, Wolklow CSF, et al. Selective Disruption of the Cerebral Neocortex in Hypertensive than nonhypertensive persons. Neurology 2009;72:1148–55.

Klunk WE, Cohen AD, et al. Arterial stiffness and beta-amyloid progression and surface reconstruction. Neuroimage 2011;53:2592–97.

Steussert V, Hofoss D, Johnson L, Berstad AE, Seguard A, Skinningsrud A, et al. White matter lesion load increases the risk of low CSF Aβ42 in alipoprotein E4 carriers attending a memory clinic. J Neuroimaging 2011;21:78–82.

de Leeuw FE, Richard F, de Groot JC, van Duijn CM, Hofman A, Van Gijn J, et al. Interaction between hypertension, apoE, and cerebral white matter lesions. Stroke 2004;35:1057–60.

Kling MA, Trojanowski QJ, Wolk DA, Lee VM, Arnold SE. Vascular disease and dementia: paradigm shifts to drive research in new directions. Alzheimers Dement 2013;9:76–92.

Hughes TM, Kuller LH, Barinas-Mitchell EM, McDade EM, Kunkle WE, Cohen AD, et al. Arterial stiffness and β-amyloid progression in nondemented elderly adults. JAMA Neurol 2014;71:562–8.

Schneider JA, Wilson RS, Bienias JL, Evans DA, Bennett DA. Cerebral infarctions and the likelihood of dementia from Alzheimer disease pathology. Neurology 2004;62:1148–55.

Hoffman LB, Schneider J, Lesser GT, Beeri MS, Purohit DP, Grossman HT, et al. Less Alzheimer disease neuropathology in medicated hypertensive than nonhypertensive persons. Neurology 2009;72:1720–6.

Beach TG, Wilson JR, Sue LJ, Newell A, Poston M, Cisneros R, et al. Circle of Willis atherosclerosis: association with Alzheimer’s disease, neuritic plaques and neurofibrillary tangles. Acta Neuropathol 2007;113:13–21.
[61] Yarchoan M, Xie SX, Kling MA, Toledo JB, Wolk DA, Lee EB, et al. Cerebrovascular atherosclerosis correlates with Alzheimer pathology in neurodegenerative dementias. Brain 2012;135:3749–56.

[62] Toledo JB, Toledo E, Weiner MW, Jack CR Jr, Jagust W, Lee VM, et al. Cardiovascular risk factors, cortisol, and amyloid-β deposition in Alzheimer’s Disease Neuroimaging Initiative. Alzheimers Dement 2012;8:483–9.

[63] Hughes TM, Kuller LH, Barinas-Mitchell EJ, Mackey RH, McDade EM, Klunk WE, et al. Pulse wave velocity is associated with -amyloid deposition in the brains of very elderly adults. Neurology 2013;81:1711–8.

[64] Gurol ME, Viswanathan A, Gidicsin C, Hedden T, Martinez-Ramirez S, Dumas A, et al. Cerebral amyloid angiopathy burden associated with leukoaraiosis: A positron emission tomography/magnetic resonance imaging study. Ann Neurol 2013;73:529–36.

[65] Iliff JJ, Wang M, Liao Y, Plogg BA, Peng W, Gundersen GA, et al. A Paravascular Pathway Facilitates CSF Flow Through the Brain Parenchyma and the Clearance of Interstitial Solutes, Including Amyloid β. Sci Transl Med 2012;4:147ra111.

[66] Iadecola C. The Pathobiology of Vascular Dementia. Neuron 2013; 80:844–66.

[67] Debette S, Beiser A, DeCarli C, Au R, Himali JJ, Kelly-Hayes M, et al. Association of MRI Markers of Vascular Brain Injury With Incident Stroke, Mild Cognitive Impairment, Dementia, and Mortality The Framingham Offspring Study. Stroke 2010;41:600–6.

[68] Gungor Z, Anuurad E, Enkhmaa B, Zhang W, Kim K, Berglund L, Apo E4 and lipoprotein-associated phospholipase A2 synergistically increase cardiovascular risk. Atherosclerosis 2012;223:230–4.

[69] Rodrigue KM, Rieck JR, Kennedy KM, Devous MD Sr, Diaz-Arrastia R, Park DC. Risk factors for β-amyloid deposition in healthy aging: Vascular and genetic effects. JAMA Neurol 2013;70:600–6.

[70] Dickerson BC, Stoub TR, Shah RC, Sperling RA, Killiany RJ, Albert MS, et al. Alzheimer-signature MRI biomarker predicts AD dementia in cognitively normal adults. Neurology 2011;76:1395–402.

[71] Dickerson BC, Wolk DA. MRI cortical thickness biomarker predicts AD-like CSF and cognitive decline in normal adults. Neurology 2012;78:84–90.

[72] Oh H, Mormino EC, Madison C, Hayenga A, Smilic A, Jagust WJ. β-Amyloid affects frontal and posterior brain networks in normal aging. Neuroimage 2011;54:1887–95.

[73] Mormino EC, Betensky RA, Hedden T, Schultz AP, Amariglio RE, Rentz DM, et al. Synergistic effect of β-amyloid and neurodegeneration on Cognitive Decline in Clinically Normal Individuals. JAMA Neurol 2014;71:1379–85.

[74] Schott JM, Bartlett JW, Fox NC, Barnes J, for the Alzheimer’s Disease Neuroimaging Initiative Investigators. Increased brain atrophy rates in cognitively normal older adults with low cerebrospinal fluid Aβ1-42. Ann Neurol 2010;68:825–34.

[75] Whitwell JL, Tosakulwong N, Weigand SD, Senjem ML, Lowe VJ, Gunter JL, et al. Does amyloid deposition produce a specific atrophic signature in cognitively normal subjects? Neuroimage Clin 2013;2:249–57.

[76] Sperling RA, Johnson KA, Doraiswamy PM, Reiman EM, Fleisher AS, Sabbagh MN, et al. Amyloid deposition detected with florbetapir F 18 (18F-AV-45) is related to lower episodic memory performance in clinically normal older individuals. Neurobiol Aging 2013;34:822–31.

[77] Rentz DM, Parra Rodriguez MA, Amariglio R, Stern Y, Sperling R, Ferris S. Promising developments in neuropsychological approaches for the detection of preclinical Alzheimer’s disease: a selective review. Alzheimers Res Ther 2013;5:58.