SIRT1 (rs3740051) role in pituitary adenoma development

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Abstract
Background: Our purpose was to determine if SIRT1 (rs4746720, rs3740051) genotypes have an influence on the development of pituitary adenoma (PA).
Methods: The study group included 142 patients with pituitary adenoma (PA) and the control group consisted of 826 healthy people. The genotyping of SIRT1 (rs4746720, rs3740051) was carried out using the real-time polymerase chain reaction method.
Results: Statistically significant results were obtained in the analysis of SIRT1 rs3740051. Significant differences in genotype (G/G, G/A, A/A) distribution were obtained comparing patients with PA without recurrence and PA with recurrence (0, 17.9, 82.1% vs. 6.7, 6.7, 86.7%, respectively, \( p = 0.022 \)). Also, statistically significant differences were observed when comparing the genotype (G/G, G/A, A/A) distribution in the non-invasive PA group and the invasive PA group (3.4, 25.9, 70.7% vs. 0, 8.3, 91.7%, respectively, \( p = 0.003 \)), and allele G was less frequently observed in invasive PA, than in non-invasive PA (4.2% vs. 16.4%, \( p < 0.001 \)). Further analysis revealed that G/A (OR = 0.261; 95% CI:0.099–0.689; \( p = 0.007 \)) and each allele A (OR = 0.229; 95% CI:0.091–0.575; \( p = 0.002 \)) were associated with lower odds of occurring an invasive PA.
Conclusions: Our study revealed that SIRT1 rs3740051 is associated with PA recurrence and invasiveness. The haplotype containing alleles C-A in rs12778366-rs3740051 was found to be associated with increased odds of PA development as well.
Keywords: Pituitary adenoma, SIRT1 (rs4746720, rs3740051) gene polymorphisms, Invasiveness

Background
Pituitary adenoma (PA), a benign pituitary tumour, originates in the adenohypophyseal cells of the anterior lobe of the pituitary gland [1]. Usually, there is no capsule to isolate this soft tumour from microglia and the surrounding structures can be infiltrated by the growing tumour [2]. According to their size PAs can be classified as microadenomas and macroadenomas [2]. Women are at a twice higher risk of developing PA in comparison with men [1]. Meta-analysis showed that pituitary tumours are common and occur in almost 20% of the population [3]; they amount to about 10 to 15% of surgically removed intracranial tumours [4–6]. A study by Daly et al. revealed that the prevalence of PAs is one case in 1064 individuals [7]. Few studies have found age-adjusted incidence rate of PA is 2.7–2.87 cases per 100,000 [8, 9]. Hemminki and colleagues have analyzed familial risks for PAs and revealed that the risk of PA was significantly increased in individuals whose siblings were diagnosed with colorectal cancer [10].
However, the mechanisms of PA development are still not clear, as sporadic pituitary tumours rarely involve mutations of classical oncogenes or tumour suppressor genes [4, 5]. Despite scarcely being malignant, pituitary adenomas may display various invasive behaviors that have been related to the particular pathological subtype [11] and to different degrees of morbidity [3]. The tumour invasiveness affects the management and prognosis of PA and needs to be investigated [12]. The present study aimed to identify a molecular marker possibly involved in PA tumorigenesis, which could be used as a potential diagnostic and prognostic tool. In our study, two SIRT1 gene polymorphisms were selected for further investigation.

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Sirtuins (SIRT) are a highly conserved family of NAD-dependent class III deacetylases that help to regulate the longevity of various organisms [13]. In mammals, seven human Sir2 homologues (sirtuins) designated as SIRT1 to SIRT7 have been identified to date. These are associated with calorie restriction, aging, metabolism, stress response, inflammation, cancer, transcriptional silencing, chromosomal stability, cell differentiation, apoptosis, and DNA repair. Generally, sirtuins are reported to have key roles in cellular senescence, cell differentiation, and inflammation [14–17]. It has been determined that levels of SIRT1 increased significantly in hepatocellular carcinoma [18], breast cancer [19], glioblastoma [20], lymphoma [21], and other types of cancer development and invasion [22–25]. The SIRT1 rs12778366 polymorphism was found to have a relation with breast cancer [26]. In our previous study, analyzing the SIRT1 rs12778366 polymorphism in the overall group, we revealed different genotype distributions in the PA and control groups [27]. That is why we aimed to investigate whether there was an association between pituitary adenoma and two other polymorphisms (rs4746720, rs3740051) of the SIRT1 gene.

Methods
Patients and selection
The study was carried out in the Department of Ophthalmology, Department of Pathology and in the Department of Neurosurgery, Lithuanian University of Health Sciences (LUHS).

Our study consisted of 142 patients with pituitary adenoma (PA) group and controls who were involved by the inclusion and exclusion criteria [28].

Activeness and recurrence evaluation
The analysis of all pituitary adenomas was based on histopathological findings of PA and hormone levels in the blood serum before surgery. All 142 subjects were categorized into two groups – active and inactive PA. The active PA group was not divided into smaller subgroups by increase of specific hormone because dominant tumours were prolactinomas, while others would not make sufficient numbers for the study. Since some of the 142 subjects had already had surgery in recent years, we categorized them by recurrence of pituitary adenoma into two groups – PA with and without recurrence.

Patients before medical or surgical treatment who were newly diagnosed with PA or came for follow up with PAs diagnosis to the Department of Neurosurgery were included in the study which lasted for 5 years and was financed by the Research Council of Lithuania (grant no. MIP 008/2014). Pituitary adenoma recurrence was diagnosed when enlargement of a residual tumor or a new growth was documented on follow-up magnetic resonance imaging (MRI) after surgical resection during the period of this study. The residual tumor was considered stable if there were no signs of tumor progression on follow-up MRI. Most of prolactinomas were surgically treated because of remaining pressure effects of surrounding structures or because of ineffective medical treatment.

Invasiveness evaluation, Ki-67 labelling index, DNA extraction and genotyping
The analysis of pituitary adenoma invasiveness and method for Ki-67 labelling index has been widely described in our previous article [28], as well as DNA extraction and genotyping based on real-time polymerase chain reaction (RT-PCR) methods [29].

Statistical analysis
Deviation from Hardy-Weinberg equilibrium (HWE) of SIRT1 (rs3740051) alleles was calculated with the Pearson’s χ² statistical test in both the case and the control subjects.

Statistical analysis was performed using SPSS 20.0 software (IBM SPSS, Armonk, NY, USA).

Frequencies of SIRT1 (rs3740051) genotypes and alleles, and other categorical variables were expressed as absolute numbers with percentages in brackets compared using the Pearson’s χ² and Fisher’s exact test (when N is less than 50) in study groups. The computational of statistical power was performed using Bioinformatics Institute’s Online Sample Size Estimator (OSSE) (http://osse.bii.a-star.edu.sg/calculation2.php). The age was presented as median and interquartile range (IQR) and compared between PA and control groups using nonparametric Mann-Whitney U test. Binomial logistic regression analysis was performed to estimate the impact of genotypes on PA development and expressed as genetic models (codominant: heterozygotes versus wild type homozygotes and minor allele homozygotes versus wild type homozygotes; dominant: minor allele homozygotes and heterozygotes versus wild type homozygotes; recessive: minor allele homozygotes versus wild type homozygotes and heterozygotes; overdominant: heterozygotes versus wild type homozygotes and minor allele homozygotes; additive model was used to evaluate the impact of each minor allele on PA development). Odds ratios (ORs) with 95% confidence intervals (CIs) are presented. Akaike information criterion (AIC) with the lowest AIC values showed the best genetic models. Haplotypes were constructed of two single nucleotide polymorphisms (SNPs) located on the same chromosome (rs12778366-rs3740051). Estimation of haplotype association tests was carried out using PLINK software version 1.07 [30] as reported previously [29]. Statistical significance was indicated when p < 0.05.
Results
The study group involved 142 patients with pituitary adenoma (PA): 55 males and 87 females. The median age of the groups was 53.5 years. The control group consisted of 826 subjects: 320 males and 506 females; the median age was 53 years as well and did not differ statistically significantly from PA group (Table 1). The distributions of PA recurrence, invasiveness, hormonal activity, and Ki-67 LI are presented in Table 1.

The frequency of genotypes and alleles of rs3740051 in patients with PA and control subjects
While the SIRT1 rs4746720 polymorphism was not involved in statistical analysis because genotyping results showed only wild type genotype for all study subjects, the genotyping of SIRT1 rs3740051 was performed successfully and the genotype distribution did not deviate from Hardy–Weinberg equilibrium (HWE) ($p > 0.05$).

Analysis of genotype (G/G, G/A and A/A) and allele (A and G) distributions in PA and control groups did not reveal any statistically significant differences between these groups (83.1, 15.5 and 1.4% vs. 86.6, 12.5 and 1%, $p = 0.534$; 90.8 and 9.2% vs. 92.8 and 7.2%, $p = 0.248$, respectively) (Table 2).

Binary logistic regression was performed to assess the rs3740051 impact on PA development, but the results did not show any significance (Additional file 1).

Table 1 Demographic characteristics

| Characteristic          | Group               | $p$ value |
|-------------------------|---------------------|-----------|
|                         | PA, N (%) (n = 142) | Control, N (%) (n = 826) |   |
| Gender                  |                     |           |   |
| Females                 | 87 (61.3)           | 506 (61.3) | 0.998*   |
| Males                   | 55 (38.7)           | 320 (38.7) |           |
| Age, median (IQR)       | 53.5 (22)           | 53 (11)   | 0.764**  |
| Recurrence              |                     |           |   |
| Yes                     | 30 (21.1)           | –         | –        |
| No                      | 112 (78.9)          | –         | –        |
| Invasiveness            |                     |           |   |
| Yes                     | 84 (59.2)           | –         | –        |
| No                      | 58 (40.8)           | –         | –        |
| Hormonal activity       |                     |           |   |
| Yes                     | 80 (56.3)           | –         | –        |
| No                      | 62 (43.7)           | –         | –        |
| Ki-67 LI                |                     |           |   |
| < 1%                    | 41 (74.6)           | –         | –        |
| 1%                      | 8 (14.5)            | –         | –        |
| > 1%                    | 6 (10.9)            | –         | –        |

IQR interquartile range
*Pearson’s $\chi^2$ test
**Mann–Whitney U test

The frequency of genotypes and alleles of rs3740051 in patients with PA and control subjects by gender
Statistical analysis was performed in patients and control subjects by gender as well. Unfortunately, the results did not reveal any associations of the rs3740051 with PA neither in males nor in females (Additional files 2, 3).

Looking for the possible cause of PA development, we performed a detailed statistical analysis of the frequency of genotypes and alleles of rs3740051 by PA recurrence, invasiveness, and hormonal activity.

The frequency of genotypes and alleles of rs3740051 in patients with PA by recurrence
Statistical analysis revealed significant differences in genotype (G/G, G/A, A/A) distribution only between the PA without recurrence and PA with recurrence groups (0, 17.9, 82.1% vs. 6.7, 6.7, 86.7%, $p = 0.022$ with the lack of power (3.4%). Logistic regression analysis did not show any significant results (Table 3).

The frequency of genotypes and alleles of rs3740051 in patients with PA by PA invasiveness
The next part of our study was to evaluate the impact of rs3740051 on PA development by tumour invasiveness. Statistically significant differences were observed when comparing the genotype (G/G, G/A, A/A) distribution in the non-invasive PA group and the invasive PA group (3.4, 25.9, 70.7% vs. 0, 8.3, 91.7%, respectively, $p = 0.003$) (Table 4). Also, allele G was less frequently observed in invasive PA, than in non-invasive PA (4.2% vs. 16.4%, $p < 0.001$) with the statistical power of 65.6%. Logistic regression analysis showed that genotype G/A (OR = 0.261; 95% CI:0.099–0.689; $p = 0.007$) and each allele A (OR = 0.229; 95% CI:0.091–0.575; $p = 0.002$) were associated with lower odds of occurring an invasive PA (Table 4).

The frequency of genotypes and alleles of rs3740051 in patients with PA by the PA hormonal activity
The distribution of genotypes and alleles of rs3740051 was evaluated in patients with active PA and non-active PA. Unfortunately, we found no associations between the rs3740051 and active PA or non-active PA development (Additional file 4). Also, we compared the distribution of genotypes and alleles of rs3740051 between patients with prolactinomas as those were the dominant tumours and control group, but no statistically significant differences were found (Additional file 5). Logistic regression analysis did not show any significant results as well (Additional file 6).

Ki-67 labeling index and SIRT1 rs3740051 association in patients with PA
Ki-67 LI was evaluated in 55 patients with PA (Table 5). The analysis of SIRT1 rs3740051 genotypes was
performed in patients with PA by Ki-67 LI. According to our results, the rs3740051 G/G genotype was not observed in this group of patients. Also, we compared the distribution of A/A and G/A genotypes in all three Ki-67 LI groups, but results did not show statistically significant differences (Table 5).

Table 2 The frequency of genotypes and alleles of rs3740051 in patients with PA and control subjects

| Genotype/allele | Frequency (%) | Control group, N (%) (n = 826) | PA group, N (%) (n = 142) | HWE p value | p value* |
|-----------------|---------------|-------------------------------|--------------------------|-------------|----------|
| Genotype        |               |                               |                          |             |          |
| G/G             | 8 (1.0)       | 2 (1.4)                       | 0.053                    | 0.414       | 0.534    |
| G/A             | 103 (12.5)    | 22 (15.5)                     | 2.014                    | 0.148       |          |
| A/A             | 715 (86.6)    | 118 (83.1)                    |                          |             |          |
| Allele          |               |                               |                          |             |          |
| G               | 119 (7.2)     | 26 (9.2)                      | 2.014                    | 0.148       |          |
| A               | 1533 (92.8)   | 258 (90.8)                    |                          |             |          |

PA pituitary adenoma, p value significance level, HWE p value Hardy-Weinberg significance level  
*Pearson’s χ² test

Haplotype associations with PA development

In this section, we performed an association analysis between the risk of PA and the haplotype of rs12778366-rs3740051. This haplotype was constructed from one previously published SNP (rs12778366) [27] and one SNP analysed in our study (rs3740051). We evaluated the linkage disequilibrium between the two rs12778366-rs3740051 and obtained a D’ value of 0.196. Only the haplotype containing alleles C-A in rs12778366-rs3740051 was significantly associated with the increased risk of PA development (17.56% in PAs vs. 11.22% in control subjects, χ² = 8.984, p = 0.003).

Discussion

SIRT1 is a conserved nicotinamide adenine dinucleotide-dependent protein deacetylase that acts as a longevity regulator [31]. However, SIRT1 is also associated with cancer cell growth, apoptosis, and tumorigenesis [32]. It has been hypothesized that SIRT1 inactivates the Akt pathway in a SIRT1 deacetylase-dependent manner; thus, SIRT1 is responsible for the deacetylation of the tumour suppressor PTEN [33], a known negative regulator for the phosphatidylinositol 3-kinase (PI3K)/Akt pathway, which is a key oncogenic pathway that promotes cell growth and survival. It has been suggested that modulation of mammalian sirtuins like SIRT1 may thus be a right approach in slowing the trajectory of aging-related degenerative changes in the central nervous system and neurologic disorders [34]. SIRT1 was first found as a nuclear protein [35] that may also shuttle to the cytoplasm during neuronal differentiation and neurite outgrowth [36–38], tumour progression [39, 40] and apoptosis [41]. Roth and Chen [42] suggested that

Table 3 The frequency of genotypes and alleles of rs3740051 in patients with PA without recurrence and PA with recurrence

| Genotype/allele | Frequency (%) | PA without recurrence group, N (%) (n = 112) | PA with recurrence group, N (%) (n = 30) | OR*** (CI) | p value* |
|-----------------|---------------|-----------------------------------------------|----------------------------------------|-----------|----------|
| Genotype        |               |                                               |                                        |           |          |
| G/G             | 0 (0)         | 2 (6.7)                                       |                                        |           |          |
| G/A             | 20 (17.9)     | 2 (6.7)                                       |                                        |           |          |
| A/A             | 92 (82.1)     | 26 (86.7)                                     |                                        |           |          |
| Allele          |               |                                               |                                        |           |          |
| G               | 20 (8.9)      | 6 (9.7)                                       |                                        |           |          |
| A               | 204 (91.1)    | 56 (90.3)                                     |                                        |           |          |

PA pituitary adenoma, p value significance level, OR odds ratio, CI confidence interval  
*Pearson’s χ² test
**Fisher’s exact test
***Odds ratios were evaluated under the codominant and additive genetic models  
Bold entries have significant values
SIRT1 may play a protective role in normal cell cycle by suppressing the tumorigenesis. While it was proposed that tumor cells act through SIRT1-regulated pathways by preventing aberrant life cycle, the main role of SIRT1 remains unclear.

Several studies have analysed SIRT1 gene polymorphisms and expression in association with various types of tumours. Rizk et al. [26] have investigated SIRT1 (SNPs) rs3758391, rs3740051, and rs12778366 gene polymorphisms in patients with breast cancer and proved that two of those (rs3758391 and rs12778366) were associated with breast cancer development in the Egyptian population. Also, their data implicate that SIRT1 rs3740051 can be a possible contributor to breast tumorigenesis and suggest that the G allele may play a role in the increased expression of SIRT1 and higher susceptibility to breast cancer [26].

Immunohistochemical analysis has shown that SIRT1 is significantly elevated in human prostate cancer [19], acute myeloid leukemia [43], primary colon [44], and different types of skin cancer [45], but Wang et al. have found reduced SIRT1 expression in several types of cancer (glioblastoma, bladder carcinoma, prostate carcinoma, ovarian cancer, and hepatic carcinoma), compared to the corresponding normal tissues [46]. Chu-Xia Deng also concluded that the SIRT1 acts as a tumour suppressor rather than a promoter [47].

Our previous study analyzed other SIRT1 SNP and demonstrated that SIRT1 rs12778366 could be associated with PA development. We previously found that the carriers of the minor allele C at SIRT1 rs12778366 had an increased risk of PA development [27]. Our current study revealed that the SIRT1 rs3740051 A/A genotype decreases odds of recurrent PA development, and the G/A and G/G genotypes and each G allele increases the odds of non-invasive PA development. To our knowledge, it is the first study analysing the SIRT1 rs3740051 gene polymorphism association with PA. We disagree with the Rizk et al. [26] study, which states that the G allele is associated with the increased expression of SIRT1 and higher susceptibility to breast cancer [26], as we believe that the G allele increases the odds of non-invasive PA development. Also, we found that the haplotype containing alleles C-A in rs12778366-rs3740051 was significantly (p = 0.003) associated with increased risk of PA development in our study.

To our knowledge, the frequency of pituitary adenomas varies greatly according to age and sex. The various adenoma types have their peak occurrence in distinctly different age groups and differ greatly in their female-to-male ratios [48]. The biology and the clinical course of clinically non-functioning pituitary adenoma seem to differ in women and men [49]. Unfortunately, our study results did not reveal any associations of rs3740051 with PA neither in males nor in females. In an analysis of males and females separately, we expected to find differences of SNPs between patients with PA and controls, and a possible link to PA in females. In our previous study [27] significant differences comparing SIRT1 rs12778366 genotype distribution in females with PA with healthy females were revealed. The T/C genotype was less frequently present in females with PA compared with the healthy control females (0 vs. 17.5%, respectively; p < 0.001) and C/C was more frequent in PA females.

### Table 4 The frequency of genotypes and alleles of rs3740051 in patients with non-invasive PA and invasive PA

| Genotype/allele | Frequency (%) | OR** (CI) | p value* |
|-----------------|--------------|-----------|----------|
| Non-invasive PA group, N (%) (n = 58) | Invasive PA group, N (%) (n = 84) |          |
| **Genotype**    |              |           |          |
| G/G             | 2 (3.4)      | 0 (0)     |          |
| G/A             | 15 (25.9)    | 7 (8.3)   | 0.003*   |
| A/A             | 41 (70.7)    | 77 (91.7) |          |
| **Allele**      |              |           |          |
| G               | 19 (16.4)    | 7 (4.2)   | < 0.001* |
| A               | 97 (83.6)    | 161 (95.8)|          |

PA pituitary adenoma, p value significance level, OR odds ratio, CI confidence interval.

*Pearson’s χ² test

**Odds ratios were evaluated under the codominant and additive genetic models

### Table 5 Association of Ki-67 LI and rs3740051 in patients with PA

| Genotype of rs3740051 | Ki-67 LI |
|-----------------------|----------|
|                       | <1%      | 1%      | >1%      |
| G/G, n (%)            | 0 (0)    | 0 (0)   | 0 (0)    | 0.534   |
| G/A, n (%)            | 5 (12.2) | 0 (0)   | 1 (16.7) |
| A/A, n (%)            | 36 (87.8)| 8 (100) | 5 (83.3) |

p value significance level

*Pearson's χ² test
compared with healthy females (19.3 vs. 2.7%, respectively; \( p < 0.001 \)). When analyzing genotype distribution in males, T/C genotype was not observed in males with PA, only in healthy control males (0 vs. 17.4%, respectively; \( p < 0.001 \)), while the C/C genotype was more frequent in males with PA compared with the controls (18.2 vs. 2.0%, respectively; \( p < 0.001 \)). Our present study showed that \( SIRT1 \) rs3740051 SNP decreases the odds of recurrence and increases the odds of non-invasive PA development as well.

### Conclusions

Our study revealed that \( SIRT1 \) rs3740051 is associated with PA recurrence and invasiveness. The haplotype containing alleles C-A in rs12778366-rs3740051 was found to be associated with increased odds of PA development as well.

### Supplementary information

Our study revealed that \( SIRT1 \) rs3740051 is associated with PA recurrence and invasiveness. The haplotype containing alleles C-A in rs12778366-rs3740051 was found to be associated with increased odds of PA development as well.

**Additional file 1.** The impact of rs3740051 on PA development. Logistic regression analysis was performed to evaluate the impact of rs3740051 on PA development under genetic models.

**Additional file 2.** The frequency of genotypes and alleles of rs3740051 in patients with PA and control subjects by gender. Frequency of genotypes and alleles of rs3740051 were estimated to compare differences between patients with PA and control subjects by gender.

**Additional file 3.** The impact of rs3740051 on PA development by gender. Logistic regression analysis was performed to evaluate the impact of rs3740051 on PA development under genetic models in female and male groups.

**Additional file 4.** The frequency of genotypes and alleles of rs3740051 in patients with active PA and inactive PA. Frequency of genotypes and alleles of rs3740051 were estimated to compare differences between patients with active PA and inactive PA.

**Additional file 5.** The frequency of genotypes and alleles of rs3740051 in patients with prolactinomas and control subjects. Frequency of genotypes and alleles of rs3740051 were estimated to compare differences between patients with prolactinomas and control subjects.

**Additional file 6.** The impact of rs3740051 on development of prolactinomas. Logistic regression analysis was performed to evaluate the impact of rs3740051 on prolactinomas development under genetic models.

### Abbreviations

- A: Adenine
- AL: Alkaline phosphatase
- ALT: Alanine aminotransferase
- C: Cytosine
- Cl: Confidence intervals
- CRH: Corticotropin-releasing hormone
- DNA: Deoxyribonucleic acid
- G: Guanine
- HWE: Hardy-Weinberg equilibrium
- IQR: Interquartile range
- K: Kinetic analysis
- Ki-67: Ki-67 index
- LD: Linkage disequilibrium
- LHR: Luteinizing hormone releasing hormone
- Mg2+: Magnesium ions
- MIB-1: Malignancy index
- MIF: Macrophage migration inhibitory factor
- MRI: Magnetic resonance imaging
- NAD: Nicotinamide adenine dinucleotide
- OR: Odds ratios
- OSSE: Online sample size estimator
- P: Probability
- PA: Pituitary adenoma
- PCR: Polymerase chain reaction
- PI3K: Phosphatidylinositol 3-kinase
- PTEN: Phosphatase and tensin homologue
- qPCR: Quantitative polymerase chain reaction
- SNP: Single nucleotide polymorphism
- T: Thymine
- T/E: Thymine/ethylnitrosourea
- TNF: Tumor necrosis factor
- TNFAIP3: Tumor necrosis factor alpha-induced protein 3
- WT: Wild-type

### Authors’ contributions

RL, UK and BG performed the Ophthalmological evaluation. GM carried out the genotyping. AV carried out a part of genotyping and performed the statistical analysis. GM drafted the manuscript. BG participated in the design of the study and RL conceived the study, participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

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### Availability of data and materials

The datasets used and analysed during the current study are available from the corresponding author on reasonable request.

### Ethics approval and consent to participate

The study was approved by the Ethics Committee for Biomedical Research at Lithuanian University of Health Sciences (LUHS) (Number – BE-2/-13). All subjects provided written informed consent in accordance with the Declaration of Helsinki.

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

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14. Broquier C, Dennis G, Rivieccio MA, et al. Specific acetylation of p53 by HDAC inhibition prevents DNA damage-induced apoptosis in neurons. J Neurosci. 2013;33(20):8621–32.

15. Kaebelerin M, McVey L, Guarente L. The SIRT2/34 complex and SIRT2 alone promote longevity in Saccharomyces cerevisiae by two different mechanisms. Genes Dev. 1999;13(19):2570–80.

16. Wojcik M, Mac-Marjanek K, Wozniak LA. Physiological and pathophysiological functions of SIRT1. Mini-Rev Med Chem. 2009;9(3):386–94.

17. Han J, Hubbard BP, Lee J, et al. Analysis of 41 cancer cell lines reveals excessive allelic loss and novel mutations in the SIRT1 gene. Cell Cycle. 2013;12(2):263–70.

18. Chen HC, Jeng YM, Yuan RH, Hsu HC, Chen YL. SIRT1 promotes tumorigenesis and resistance to chemotherapy in hepatocellular carcinoma and its expression predicts poor prognosis. Ann Surg Oncol. 2012;19:2011–9.

19. Jin X, Wei Y, Xu F, Zhao M, Dai K, Shen R, Yang S, Zhang N. SIRT1 promotes formation of breast cancer through modulating Akt activity. J Cancer. 2018 Apr 30(11):2012–23.

20. Liu G, Yuan X, Zeng Z, Tunic P, Ng H, Abdulkadir IR, Lu L, Irvin D, Black KL, Yu JS. Analysis of gene expression and chromoeexpression of CD133+ cancer stem cells in glioblastoma. Mol Cancer. 2006;5:67.

21. Jang KY, Hwang SH, Kwon KS, Kim KR, Choi HN, Lee NR, Kwak JY, Park BH, Park HS, Chung MJ, et al. SIRT1 expression is associated with poor prognosis of diffuse large B-cell lymphoma. Ann J Surg Pathol. 2008;32:1523–31.

22. Huffman DM, Grizzle WE, Bamman MM, Kim JS, Eltoum IA, Elgavish A, Nagy TR. SIRT1 is significantly elevated in mouse and human prostate cancer. Cancer Res. 2007;67:6612–8.

23. Jang KY, Kim KS, Hwang SH, Kwon KS, Kim KR, Park HS, Park BH, Chung MJ, Kang MJ, Lee DG, Moon WS. Expression and prognostic significance of SIRT1 in ovarian epithelial tumours. Pathology. 2009;41:366–71.

24. Cha EJ, Noh SJ, Kwon KS, Kim KR, Park HS, Park BH, Chung MJ, Kang MJ, Lee DG, et al. Expression of DBC1 and SIRT1 is associated with poor prognosis of gastric carcinoma. Clin Cancer Res. 2009;15:4453–45.

25. Wood J. Function of the SIRT1 protein deacetylase in cancer. Biotechnol J. 2007;2:1360–8.

26. Ritz SM, Shahin NN, Shaker OG. Association between SIRT1 gene polymorphisms and breast cancer in Egyptians. PLoS One. 2016;11:e0151901.

27. Glebauskienė B, Vilkevičiute A, Liukstienė R, Jagstienė S, Kreivienė P, Žemaitienė R, Zaliuniene D. Association of FGFR2 rs2981382, SIRT1 rs12738366, STAT3 rs744166 gene polymorphisms with pituitary adenoma. Oncol Lett. 2017;13:3087–99.

28. Sidaraitė A, Liukstienė R, Glebauskienė B, Vilkevičiute A, Kreivienė P, Žemaitienė R, Zaliuniene D, et al. Associations of cholesteryl ester transfer protein (CETP) gene variants with pituitary adenoma. Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub. 2019. https://doi.org/10.53507/bp.2019.016. Epub ahead of print.

29. Gedvilaite G, Vilkeviciute A, Kriauciuniene L, Asmoniene V, Liukstienė R. Does CETP rs5882, rs708272, SIRT1 rs12738366, FGFR2 rs2981382, STAT3 rs744166 gene polymorphisms play a role in optic neuritis development? Ophthalmic Genet. 2019;40(3):219–26.

30. Purcell S, Neale B, Todd-Brown K, et al. PLINK: a tool set for whole-genome association and population-based linkage analysis. Am J Hum Genet. 2007;81:559–75.

31. Ford J, Jiang M, Miler J. Cancer-specific functions of SIRT1 enable human epithelial cancer cell growth and survival. Cancer Res. 2005;65:10457–63.

32. Saunders LR, Verdin E. Sirtuins: critical regulators at the crossroads between cancer and aging. Oncogene. 2007;26:5489–504.

33. Ikemou T, Inoki K, Zhao B, Guan KL. PTEN acetylation modulates its function through the mammalian target of rapamycin signaling. Cell. 2011;144:430–41.

34. Hishara S, Chiba S, Matsumoto H, Tanno M, Yagi H, Shimohama S, Sato M, Hori Y. Histone deacetylase SIRT1 modifies neuronal differentiation by its nuclear translocation. Proc Natl Acad Sci U S A. 2008;105:15599–604.

35. Sugino T, Manuyama M, Tanno M, Kuno A, Houkin K, Hori Y. Protein deacetylase SIRT1 in the cytoplasm promotes nerve growth factor-induced neurite outgrowth in PC12 cells. FEBS Lett. 2010;584:2821–6.