Research article

Selecting short-statured children needing growth hormone testing: Derivation and validation of a clinical decision rule

Laëtitia Duché1,2,3, Christine Trivin4, Wassim Chemaitilly3, Jean Claude Souberbielle4, Gérard Bréart2, Raja Brauner3 and Martin Chalumeau*1,2

Address: 1Clinical Epidemiological Unit-Department of Pediatrics, Saint-Vincent-de-Paul Hospital, AP-HP, Université Paris Descartes, 75014 Paris, France, 2INSERM U149, 75014 Paris, France, 3Université Paris Descartes and Assistance Publique Hôpitaux de Paris, Hôpital Bicêtre, Unité d’endocrinologie pédiatrique, 94270 Le Kremlin Bicêtre, France and 4Assistance Publique Hôpitaux de Paris, Necker-Enfants Malades Hospital, Service d’explorations fonctionnelles, 75743 Paris, France

Email: Laëtitia Duché - laetitia.duche@laposte.net; Christine Trivin - christine.trivin@nck.ap-hop-paris.fr; Wassim Chemaitilly - wchemaitilly@gmail.com; Jean Claude Souberbielle - jean-claude.souberbielle@nck.ap-hop-paris.fr; Gérard Bréart - breart@cochin.inserm.fr; Raja Brauner - raja.brauner@wanadoo.fr; Martin Chalumeau* - martin.chalumeau@svp.aphp.fr

* Corresponding author

Abstract

Background: Numerous short-statured children are evaluated for growth hormone (GH) deficiency (GHD). In most patients, GH provocative tests are normal and are thus in retrospect unnecessary.

Methods: A retrospective cohort study was conducted to identify predictors of growth hormone (GH) deficiency (GHD) in children seen for short stature, and to construct a very sensitive and fairly specific predictive tool to avoid unnecessary GH provocative tests. GHD was defined by the presence of 2 GH concentration peaks < 10 ng/ml. Certain GHD was defined as GHD and viewing pituitary stalk interruption syndrome on magnetic resonance imaging. Independent predictors were identified with uni- and multi-variate analyses and then combined in a decision rule that was validated in another population.

Results: The initial study included 167 patients, 36 (22%) of whom had GHD, including 5 (3%) with certain GHD. Independent predictors of GHD were: growth rate < -1 DS (adjusted odds ratio: 3.2; 95% confidence interval [1.3–7.9]), IGF-I concentration < -2 DS (2.8 [1.1–7.3]) and BMI z-score ≥ 0 (2.8 [1.2–6.5]). A clinical decision rule suggesting that patients be tested only if they had a growth rate < -1 DS and a IGF-I concentration < -2 DS achieved 100% sensitivity [48–100] for certain GHD and 63% [47–79] for GHD, and a specificity of 68% [60–76]. Applying this rule to the validation population (n = 40, including 13 patients with certain GHD), the sensitivity for certain GHD was 92% [76–100] and the specificity 70% [53–88].

Conclusion: We have derived and performed an internal validation of a highly sensitive decision rule that could safely help to avoid more than 2/3 of the unnecessary GH tests. External validation of this rule is needed before any application.

Published: 16 July 2008

BMC Pediatrics 2008, 8:29 doi:10.1186/1471-2431-8-29

Received: 13 January 2008
Accepted: 16 July 2008

This article is available from: http://www.biomedcentral.com/1471-2431/8/29

© 2008 Duché et al; licensee BioMed Central Ltd.
This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.
**Background**

Shortness or decreasing growth is a frequent reason for pediatric consultations. After ruling out other causes of short stature (intestinal malabsorption, chronic liver or kidney disease, hypothyroidism, etc), the possibility of growth hormone (GH) deficiency (GHD) is often considered. This deficiency is associated with excess mortality and substantial morbidity [1,2], and it can be treated. Many children are therefore referred by their physicians to specialist departments to test for GHD. Testing is based on the measurement of stimulated GH secretion [3,4]: the diagnosis is generally based on 2 GH peaks < 10 ng/mL (or 20 mU/l/mL).[4]. GHD cannot be considered certain unless there are also one or more of the following confirmatory markers: familial GHD, other deficiency of the hypothalamic-pituitary axis, micropenis, neonatal hypoglycemia, abnormalities of the median line and pituitary stalk interruption syndrome (PSIS) on magnetic resonance imaging (MRI) [5].

GH stimulation tests are invasive, expensive, and in view of the risk of severe hypoglycemia [6], potentially dangerous [7]. Moreover they are normal in most cases and thus retrospectively unnecessary. It would therefore be useful to be able to identify predictive factors of GHD to avoid these unnecessary tests. A selection strategy for GH stimulation tests, however, must offer sensitivity close to 100% for certain GHD, in view of the need to begin treatment rapidly [8]; it must also be sufficiently specific.

Clinical (height, growth rate, difference between height and midparental target height) [3] and laboratory (insulin-like growth factor-I IGF-I)] [3,9] criteria have been proposed to predict GHD. Used separately, these different criteria do not fulfill the objectives described above. It may therefore be useful to combine them. Earlier clinical decision rules have proposed combining clinical and laboratory variables [10,11] to avoid GH stimulation tests. One rule combined growth rate and IGF-I [10], and the other chronological age, bone age, body mass index (BMI) and IGF-I [11]. Nonetheless the results of these studies are limited by selection bias in patient recruitment [10], the absence of multivariate analyses despite the very probable correlations between variables [10], the complexity of the calculations necessary to apply the rule [11], and insufficient predictive performance [10]. This is probably why none of these tools has undergone internal or external validation.

The objective of this study was therefore to identify the predictive factors for GH deficiency in children consulting for short stature and/or decreased growth rate and to construct and validate internally a very sensitive and fairly specific predictive tool that is simple to use to avoid unnecessary tests.

**Methods**

**Patients**

This was a retrospective hospital-based cohort study. All patients were seen by a senior pediatric endocrinologist (RB) from January 1998 through June 2001 at Necker-Enfants Malades Hospital in Paris, France. The Ethical Review Committee (Comité de Protection des Personnes Ile de France III) stated that "this research was found to conform to generally accepted scientific principles and research ethical standards and to be in conformity with the laws and regulations of the country in which the research experiment was performed" (see Additional file 1). Written informed consent of the patients or their parents was not judged necessary for that kind of retrospective study.

The patients included were 1 to 16 years-old and had at least one of the principal auxological criteria for which the GH Research Society consensus conference guidelines require GH stimulation testing [3] (height ≤ -3 standard deviations (SD), growth rate ≤ -2 SD for chronological age, or height ≤ -2 SD, growth rate ≤ -1 SD, and a difference between current height and midparental target height > 1.5 SD). They had also had 2 tests assessing GH secretion: one of spontaneous secretion during sleep and one after pharmacological stimulation.

We excluded from this study the patients with conditions other than GHD that were responsible for their short stature (hypothyroidism, celiac disease, gastrointestinal inflammatory disease, cystic fibrosis, kidney failure, or Turner syndrome) as well as those for which GHD was due to a condition already known at the consultation (lesion, surgery and/or irradiation of the hypothalamic-pituitary region) and those with signs and findings highly suggestive of GHD: familial GHD, history or clinical picture suggesting pituitary deficiency (polyuric-polydipsic syndrome, severe hypoglycemia in the first months of life, micropenis, abnormalities of the median line). Indeed, for these high-risk patients, there is no need for a selective strategy. Patients who had had testosterone or estradiol priming and those with delayed puberty (defined by a Tanner stage of 1 for a girl older than 13 years or a boy older than 14) were also excluded.

**Predicted variable**

The variable to be predicted was GHD. Plasma GH (hGH immunoradiometric assay, Immunotech, Marseille, France) was measured for each patient from blood samples taken while sleeping (samples every 30 minutes from 22 h to 6 h) followed in the morning by a provocative test administering arginine and insulin sequentially (arginine 22 h to 6 h) followed in the morning by a provocative test. Patients who had had testosterone or estradiol priming and those with delayed puberty (defined by a Tanner stage of 1 for a girl older than 13 years or a boy older than 14) were also excluded.
gon (0.1 mg/kg intramuscular injection, 1 mg maximum, n = 30). During the study period, the treatment protocol called for MRI if the 2 GH peaks were less than 10 ng/mL, to look for PSIS (thin or interrupted stalk, ectopic posterior or hypoplastic anterior pituitary gland [12]).

Children were then classified in 2 groups as a function of the GH assay and MRI results: no GHD (1 GH peak ≥ 10 ng/mL) or GHD (2 GH peaks < 10 ng/mL). Within the GHD group, children with pituitary stalk interruption syndrome on MRI were considered to have certain GHD, and the other patients were considered to have uncertain GHD.

**Potential predictors**

The following potential clinical predictors were studied: chronological age expressed in years, height measured with a Harpenden stadiometer and expressed in SD, growth rate expressed in SD [13], BMI measured as weight in kilograms divided by the square of height in meters and expressed as a z-score compared with chronological age [14], difference in SD between height and the midparental target height, calculated from both parents’ height [15], and pubertal stage (breast or testes) [16,17]. Two potential non clinical predictors were also studied: plasma IGF-I (IGF-I-RIACT, Cis Bio, Gif sur Yvette, France) expressed in SD according to chronological age [18] and bone age delay (difference in years between chronological age and bone age) [19].

**Analysis**

STATA/SE 8 (Statacorp, College Station, TX, USA) software was used for the statistical analysis. We began by using the Mann-Whitney test to compare the distribution of the possibly predictive continuous variables as a function of GHD. Next, the continuous variables were dichotomized, either according to the standard cutoff point in the literature or according to their distribution in patients without GHD (median or one of the quartiles rounded to the nearest half point). For "pubertal stage", the last 4 Tanner stages were combined into one to obtain a reproducible variable (prepubertal versus pubertal children). We conducted a bivariate analysis to study the relation between GHD and the dichotomized variables and calculate odds ratios. Comparisons were tested with the Chi-2 test or Fisher’s exact test. Next, we used logistic regression to conduct a multivariate analysis.

**Decision rule derivation**

First, the discriminant power of the independent variables associated with GHD was studied by the calculation of their sensitivity, specificity, positive predictive value and negative predictive value for GHD and for certain GHD. To meet our objective of high sensitivity (close to 100%) for certain GHD with the best possible specificity (around 2/3), we varied the cutoff points of the independent predictors. Since no independent predictor used alone met these objectives, we then combined them by recursive partition to construct a decision rule, along the lines of previous rules for pediatric endocrinology [20,21]. To make the tool simple for clinicians to use, we chose only whole values close to the standard thresholds to dichotomize the variables.

**Decision rule validation**

The predictive tool was validated among 2 populations of consecutive patients meeting the inclusion criteria described above: a population of patients with certain GHD seen from 1990–1998 and 2001–2005 and a population of patients seen in 2002 with 1 GH peak ≥ 10 ng/mL and no cause for short stature found, and thus considered not GH-deficient. The data for the validation populations remained blinded during construction of the rule, and the rule was not modified after application to these populations.

**Results**

The analysis included 167 children. Their mean age was 8.2 years (range 1.1–15.5; interquartile range 5.1–11.3) and 49% were boys; 36 (22%) children diagnosed with GHD, including 5 (3%) with certain GHD.

**Predictive variables**

Patients with GHD (Table 1) had a lower growth rate, higher BMI, and lower IGF-I level than the patients without GHD (p < 0.05). No statistically significant (p > 0.10) difference was shown in the distribution of age, height, difference from midparental target height, weight or bone age delay between the two groups. After dichotomization (Table 2), there was a statistically significant association between GHD and growth rate < -1 SD (p = 0.005) as well as BMI z-score ≥ 0 (p = 0.006). A trend that did not reach statistical significance was seen between GHD and both a prepubertal Tanner stage (p = 0.09) and IGF-I < -2 SD (p = 0.09). No statistically significant association was observed with age < 5 years, height < -2.5 SD, height difference from midparental target height, weight or bone age delay between the two groups. After adjustments, GHD was not significantly (p > 0.05) associated with a prepubertal stage (Table 2), but was significantly and independently associated with growth rate < -1 SD, BMI z-score ≥ 0 and IGF-I < -2 SD.

**Decision rule**

None of the criteria used alone allowed us to reach the objectives we had set: sensitivity of 100% for certain GHD and specificity ≥ 2/3 (Table 3). The best combination of predictive independent variables was growth rate and IGF-I (Figure 1). A clinical decision rule suggesting that GH
stimulation testing was necessary only if these 2 indicators (growth rate < -1 SD and IGF-I < -2 SD) were both present yielded a specificity of 68% (95% CI [60–76]) with a sensitivity of 100% (95% CI [48–100]) for the certain GHD diagnosis. Adding BMI to this combination in a decision tree or composite score did not improve its predictiveness. Of the patients with uncertain GHD, 43% were not identified by the rule. These patients had a mean age of 8.5 years, a mean height of -2.3 SD, a mean growth rate of -0.8 SD and a mean IGF-I of -2.0 SD. None had panhypopituitarism and 84% had not had GH treatment.

For the periods 1990–1998 and 2001–2005, 13 patients who met the inclusion criteria had certain GHD. The sensitivity of the combination of growth rate < -1 SD and IGF-I level < -2 SD was 92% (95% CI [76–100]). The one patient with certain GHD who was not identified by the rule was a 13-year-old boy with a height -2.9 SD and a growth rate of -0.9 SD; he had no other pituitary deficiencies and was treated with GH. In 2002, 27 patients had at least one GH peak ≥ 10 ng/mL and met the inclusion criteria. The specificity of the predictive tool applied to this population was 70% (95% CI [53–88]).

Discussion

Three independent predictive factors were identified among the patients we studied: growth rate < -1 SD, IGF-I < -2 SD and BMI z-score ≥ 0. Growth rate is a classic predictor of GHD. Different cutoff points have been proposed in the literature [22-25] including the one we used here (< -1 SD). The predictive power of the IGF-I level has been studied repeatedly [5,9,25-29]. The results in terms of sensitivity and specificity vary widely, but this assay is very useful for the diagnosis of GHD [9]. The cutoff point we used (-2 SD) is that usually found in the literature.

In our study, a BMI z-score ≥ 0 was also an independent predictive factor of GHD. This criterion is most often considered a confounding factor instead [4]. That is, on the one hand, children with simple obesity have an abnormally low response to GH stimulation tests and on the other hand, some children with GHD have truncal obesity. Accordingly a predictive tool that uses this criterion might therefore be dangerous. Moreover, it does not improve the rule’s predictive power.

The clinical decision rule we propose here is that GH stimulation tests should be performed only on children with a growth rate < -1 SD and an IGF-I level < -2 SD. These variables were also included in the rule proposed by Cianfarani et al but with a different combination [10]. Our decision rule has good clinical applicability because it uses predictive variables at the rounded cutoff points already used by clinicians. Moreover, it is probably robust because it uses independent predictors identified by multivariate analysis. This rule should make it possible to avoid two thirds of the GH stimulation tests that are retrospectively unnecessary because normal, while missing in our series only one of 18 cases of certain GHD. This patient had an IGF-I level < -2 SD but a growth rate of -0.9 SD. It is probable that he would have reached -1 SD during his next follow-up, thus being “caught” by the rule. Moreover since he did not have panhypopituitarism, there was no immediate metabolic danger [1]. Our rule is a relatively insensitive predictive tool for the diagnosis of uncertain GHD: 43% of these patients were not identified. Compensating for this poor prediction is the fact that

### Table 1: Distribution of potential predictors among children with or without GHD.

| Variables                                      | Absence of GHD | GHD | Certain | p**     |
|------------------------------------------------|----------------|-----|---------|---------|
| Age (years)                                    | 131 8.3 3.8    | 31 8.1 3.6  | 5 5.2 2.4 | 0.4     |
| Height (SD)                                    | 131 2.2 0.6    | 31 2.4 0.7  | 5 2.3 0.5  | 0.2     |
| Weight (SD)                                    | 131 -1.7 0.8   | 31 -1.3 1.2 | 5 -1.3 1.4 | 0.07    |
| Growth rate (SD)                               | 127 -0.9 1.6   | 30 -1.5 1.0 | 5 -2.3 0.9  | 0.01    |
| Difference between height and MTH***           | 127 -2.1 0.9   | 29 -2.1 1.1 | 4 -3.1 0.3  | 0.7     |
| BMI (z-score)                                  | 131 -0.8 1.1   | 31 0.0 1.4  | 5 0.0 2.2  | 0.001   |
| Studies                                        | 131 -2.2 1.3   | 31 -2.7 1.1 | 5 -3.9 1.1  | 0.03    |
| IGF-I (SD)                                     | 125 2.1 1.1    | 29 2.0 0.9  | 5 2.2 0.9  | 0.9     |

* Standard deviation
** Non parametric Mann-Whitney U-test (between patients with and without GHD)
*** Midparental target height
these patients did not have panhypopituitarism, that 84% of them did not receive treatment, and that the abnormal character of uncertain GHD is currently the subject of much debate [30,31]. Indeed, GH secretion in most children with a subnormal GH response to GH stimulation tests but normal MRI becomes normal when they are retested at the completion of growth or even after few months. Thus, it is likely that many of the patients in the uncertain GHD group did not have GHD. Some of them have been retested (16%) and had normal GH secretion; they may be considered to have had transient GHD. Others reached a normal final height without treatment (7%). The other patients are still being closely monitored for their growth velocity.

Some of the patients (n = 5, 16%) with uncertain GHD, and a IGF-I concentration ≥ -2 SD but a growth rate < -1 SD, would not have been identified as requiring GH secretion evaluation according to the decision rule (Figure 1). This potential false-negative rate shows the need for follow-up of the growth of these patients not identified as requiring GH secretion evaluation.

There were two potential sources of bias in our study. First, patients came from a specialist pediatric endocrinology outpatient department and were very probably at higher risk of GHD than any other population. This bias is demonstrated by the very high prevalence (22%) of patients with GHD compared with other series [11]. We also excluded children with priming with testosterone or estradiol to avoid a confounding bias with IGF-I, because we were treating it as a potential predictive variable and priming increases both IGF-I and peak GH concentrations [32]. Second, there may have been classification errors for the predicted variable. That is, although stimulation tests must be used to evaluate short children for whom GHD is

Table 2: Relations between potential predictors and GHD, before and after adjustment.

| Variables                             | Absence of GHD (n = 131) | GHD (n = 36) | OR | CI | p  | AOR* | CI* | pAOR* |
|---------------------------------------|---------------------------|--------------|----|----|----|------|-----|-------|
| History and physical examination      |                           |              |    |    |    |      |     |       |
| Boys                                  | 62                        | 47           | 20 | 56 | 1.4 | 0.7-2.9 | 0.4 |
| Girls                                 | 69                        | 53           | 16 | 44 |     |     |     |       |
| Age < 5 years                         | 29                        | 22           | 10 | 28 | 1.4 | 0.6-3.1 | 0.5 |
| Age ≥ 5 years                         | 102                       | 78           | 26 | 72 |     |     |     |       |
| Height < -2.5 SD                      | 48                        | 37           | 16 | 44 | 1.4 | 0.7-2.9 | 0.4 |
| Height ≥ -2.5 SD                      | 83                        | 63           | 20 | 56 |     |     |     |       |
| Weight ≥ -2 SD                        | 21                        | 16           | 8  | 22 | 1.5 | 0.6-3.8 | 0.4 |
| Weight < -2 SD                        | 110                       | 84           | 28 | 78 |     |     |     |       |
| Growth rate < -1 SD                   | 64                        | 50           | 27 | 77 | 3.3 | 1.4-8.1 | 0.005 |
| Growth rate ≥ -1 DS                   | 63                        | 50           | 8  | 23 |     |     |     |       |
| Difference between height and MTH ≤ -3 SD | 15                      | 12           | 5  | 15 | 1.3 | 0.4-4.0 | 0.6 |
| Difference between height and MTH > -3 SD | 112                     | 88           | 28 | 85 |     |     |     |       |
| BMI ≥ 0 z-score                       | 31                        | 24           | 17 | 47 | 2.9 | 1.3-6.4 | 0.006 |
| BMI < 0 z-score                       | 100                       | 76           | 19 | 53 |     |     |     |       |
| Pre-pubertal children (Tanner = 1)    | 104                       | 79           | 33 | 92 | 2.9 | 0.8-10 | 0.09 |
| Pubertal children (Tanner > 1)        | 27                        | 21           | 3  | 8  |     |     |     |       |
| Studies                               |                           |              |    |    |    |      |     |       |
| IGF-I < -2 SD                         | 82                        | 63           | 28 | 78 | 2.1 | 0.9-5.0 | 0.09 |
| IGF-I ≥ -2 SD                         | 49                        | 37           | 8  | 22 |     |     |     |       |
| Bone age delay ≥ 1.5 years            | 88                        | 70           | 27 | 79 | 1.6 | 0.6-4.1 | 0.3 |
| Bone age delay < 1.5 years            | 37                        | 30           | 7  | 21 |     |     |     |       |

* OR adjusted for variables with a p < 0.1 in bivariate analysis (N = 162)
** Variable excluded from the model
*** Midparental target height
considered, their reproducibility is poor. Response varies substantially according to the stimulus used [33]. Moreover, test response also includes a component of intraindividual variability. In a study of 40 Israeli children, Zadik and coll. reported a moderate correlation (r = 0.52) between 2 tests conducted 6 weeks apart with the same pharmacological agent [34]. In our study, as in all studies that have used stimulation tests as the reference test, classification errors may intervene between the diagnosis of GHD (defined by 2 GH peaks < 10 ng/mL without a confirmation criterion) and no GHD. Errors for the certain cases are less plausible because the criterion of certain

Table 3: Predictive values of GHD predictors.

|              | Growth rate < -1 SD | BMI ≥ 0 z-score | IGF-I < -2 SD |              |              |              |              |
|--------------|---------------------|----------------|---------------|--------------|--------------|--------------|--------------|
|              | GHD                 |                |               |              |              |              |              |
|              | All                 | Certain        | All           | Certain      | All          | Certain      | All          |
|              | Sensitivity         | 77 [60–90]     | 100 [48–100]  | 47 [31–63]   | 60 [15–95]   | 78 [64–91]   | 100 [48–100] |
|              | Specificity         | 50 [41–58]     | Idem          | 76 [69–83]   | Idem         | 37 [29–46]   | Idem         |
|              | PPV***              | 30 [20–39]     | 7 [2–16]      | 35 [22–48]   | 9 [2–24]     | 25 [17–34]   | 6 [2–13]     |
|              | NPV**               | 89 [79–95]     | 100 [94–100]  | 84 [77–91]   | 98 [95–100]  | 86 [77–95]   | 100 [93–100] |
|              | Construction†       | 63 [47–79]     | 100 [48–100]  | 68 [60–76]   | Idem         | 70 [53–88]   |
|              | Validation‡         | Not applicable |               |              |              |              |              |

Values are expressed in % [95% CI]
* Positive predictive value
** Negative predictive value
*** To test patients with growth rate < -1 SD and IGF-I < -2 SD
† Construction population
‡ Validation population

Figure 1
Patients distribution according to their growth rate (SD) and IGF-I level (SD).
GHD (pituitary MRI) is more robust. Furthermore, when we used a stimulation test cut-off of 7 ng/ml instead of 10 ng/ml to define GHD, the decision rule’s sensitivity was 67% (95% CI [43–91]) vs 63% (95% CI [47–79]) for patients with GHD and did not change for patients with certain GHD (100% (95% CI [48–100])). The specificity was 64% (95% CI [56–71]) (vs 68% (95% CI [60–76])).

Conclusion
The suboptimal nature of a systematic strategy of stimulation tests and the intrinsic limitations of these tests make the construction of a predictive tool for GHD necessary. The tool we propose is very effective for certain GHD but far less so for uncertain GHD. The current debate about the abnormal character of uncertain GHD [30,31] highlights the interest of our tool. Nonetheless, in view of the limitations of our study and especially the low number of patients with certain GHD, these results should be validated at other centers, as other decision rules in pediatric endocrinology have been [21], before any widespread clinical application.

Abbreviations
GH – Growth hormone; GHD – Growth hormone deficiency; PSIS – Pituitary stalk interruption syndrome; MRI – Resonance magnetic imaging; IGF-I – Insulin-like growth factor-I; BMI – Body mass index; SD – Standard deviation.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
All the authors 1) have made substantial contributions to conception and design, or acquisition of data, or analysis and interpretation of data; 2) have been involved in drafting the manuscript or revising it critically for important intellectual content; and 3) have given final approval of the version to be published.

Additional material

Acknowledgements
LD received an unrestricted educational grant from Laboratoire France Lilly and the Fond d’Etude et de Recherche du Corps Médical de l’Assistance Publique-Hôpitaux de Paris.

References
1. Mills JL, Schonberger LB, Wysowski DK, Brown P, Durako SJ, Cox C, Kong F, Fradin JE: Long-term mortality in the United States cohort of pituitary-derived growth hormone recipients. J Pediatr 2004, 144:430–436.
2. Taback SP, Dean HJ: Mortality in Canadian children with growth hormone (GH) deficiency receiving GH therapy 1967–1992. The Canadian Growth Hormone Advisory Committee. J Clin Endocrinol Metab 1996, 81:1693–1696.
3. GH Research Society: Consensus guidelines for the diagnosis and treatment of growth hormone (GH) deficiency in childhood and adolescence: summary statement of the GH Research Society. J Clin Endocrinol Metab 2000, 85:3990–3993.
4. Sazonenko PC, Clayton PE, Cohen P, Hintz RL, Yanaka T, Laron Z: Diagnostic criteria and management of growth hormone deficiency in childhood and adolescence. Part I: diagnosis of growth hormone deficiency. Growth Horm IGF Res 2001, 11:137–165.
5. Adan L, Souberbielle JC, Brauner R: Diagnostic markers of permanent idiopathic growth hormone deficiency. J Clin Endocrinol Metab 1994, 78:353–358.
6. Shah A, Stanhope R, Matthew D: Hazards of pharmacological tests of growth hormone secretion in childhood. Br Med J 1992, 304:173–174.
7. Parkland J, Saenger P, Macgillivray M, Lafranchi S, Rosenfield R: Physician and clinic charges for diagnosing growth hormone deficiency. J Pediatr 1996, 128:561–62.
8. Guidelines for the use of growth hormone in children with short stature. In J Pediatr Volume 127. A report by the Drug and Therapeutics Committee of the Lawson Wilkins Pediatric Endocrine Society; 1995:857-867.
9. Ranke MB, Schweizer R, Lindberg A, Price DA, Reiter EO, Albertsson-Wikland K, Darendeliler F: Insulin-like growth factors as diagnostic tools in growth hormone deficiency during childhood and adolescence: the KIGS experience. Horm Res 2004, 62(Suppl 1):17-25.
10. Cianfarani S, Tondinelli T, Spadoni GL, Scire G, Boemi S, Boscherini B: Height velocity and IGF-I assessment in the diagnosis of childhood onset GH insufficiency: do we still need a second GH stimulation test? Clin Endocrinol (Oxf) 2002, 57:161–167.
11. Lee PD, Wilson DM, Rountree L, Hintz RL, Rosenfield RG: Efficacy of insulin-like growth factor 1 levels in predicting the response to provocative growth hormone testing. Pediatr Res 1990, 27:45–51.
12. Argyropoulou M, Perignon F, Brauner R, Brunelle F: Magnetic resonance imaging in the diagnosis of growth hormone deficiency. J Pediatr 1992, 120:886–91.
13. Sempe M, Pedron G, Roy-Pernot M: Auxologie, Méthodes et Séquences Paris: Theraplix; 1979.
14. Rolland-Cachera MF, Cole TJ, Sempe M, Tichet J, Rossignol C, Charraud A: Body Mass Index variations: centiles from birth to 87 years. Eur J Clin Nutr 1991, 45(1):13-21.
15. Tanner JM, Goldstein H, Whitehouse RH: Standards for children’s height at ages 2–9 years allowing for heights of parents. Arch Dis Child 1970, 45:752–762.
16. Marshall WA, Tanner JM: Variations in the pattern of pubertal changes in boys. Arch Dis Child 1970, 45:13-23.
17. Marshall WA, Tanner JM: Variations in pattern of pubertal changes in girls. Arch Dis Child 1969, 44:291–303.
18. Bussieres L, Souberbielle JC, Pinto G, Adan L, Noel M, Brauner R: The use of insulin-like growth factor I reference values for the diagnosis of growth hormone deficiency in prepubertal children. Clin Endocrinol (Oxf) 2000, 52:735–739.
19. Greulich W, Pyle S: Radiographic Atlas of Skeletal Development of the Hand and Wrist 2nd edition. Stanford University Press; 1959.
20. Chalumeau M, Chemaitilly W, Trivin C, Adan L, Breart G, Brauner R: Central precocious puberty in girls: an evidence-based diagnosis tree to predict central nervous system abnormalities. Pediatrics 2002, 109:61–67.
21. Chalumeau M, Hadjithanasiou CG, Ng SM, Cassio A, Mul D, Cisternino M, Partsch CJ, Theodoridis C, Didi M, Cacciari E, Oostdijk W, Borghesi A, Sippell WG, Breart G, Brauner R: Selecting girls with precocious puberty for brain imaging: validation of European evidence-based diagnosis rule. J Pediatr 2003, 143:445–450.
22. Hintz RL: The role of auxologic and growth factor measurements in the diagnosis of growth hormone deficiency. Pediatrics 1998, 102:524–526.
23. Rosenfeld RG: Is growth hormone deficiency a viable diagnosis? J Clin Endocrinol Metab 1997, 82:349-351.
24. Van Buuren S, Van Dommelen P, Zandwijk J, Grote F, Wit JM, Verkerk PH: Towards evidence based referral criteria for growth monitoring. Arch Dis Child 2004, 89:336-341.
25. Federico G, Street ME, Maghnie M, Caruso-Nicoletti M, Loche S, Bertelloni S, Cianfarani S: Assessment of serum IGF-I concentrations in the diagnosis of isolated childhood-onset GH deficiency: a proposal of the Italian Society for Pediatric Endocrinology and Diabetes (SIEDP/ISPED). J Endocrinol Invest 2006, 29:732-737.
26. Blum WF, Albertsson-Wikland K, Rosberg S, Ranke MB: Serum levels of insulin-like growth factor I (IGF-I) and IGF binding protein 3 reflect spontaneous growth hormone secretion. J Clin Endocrinol Metab 1993, 76:1610-1616.
27. Cianfarani S, Liguori A, Germani D: IGF-I and IGFBP-3 assessment in the management of childhood onset growth hormone deficiency. Endocr Dev 2005, 9:66-75.
28. Jensen RB, Jeppesen KA, Vlewwerth S, Michelsen KF, Main KM, Skakkebaek NE, Juul A: Insulin-like growth factor I (IGF-I) and IGF-binding protein 3 as diagnostic markers of growth hormone deficiency in infancy. Horm Res 2005, 63:15-21.
29. Nunez SB, Municchi G, Barnes KM, Rose SR: Insulin-like growth factor I (IGF-I) and IGF-binding protein-3 concentrations compared to stimulated and night growth hormone in the evaluation of short children – a clinical research center study. J Clin Endocrinol Metab 1996, 81:1927-1932.
30. Badr S, Wilson DM: Alternatives to growth hormone stimulation testing in children. Trends Endocrinol Metab 2004, 15:252-258.
31. Wilson DM, Fran e J: A brief review of the use and utility of growth hormone stimulation testing in the NCGS: do we need to do provocative GH testing? Growth Horm IGF Res 2005, 15(Suppl A):S21-25.
32. Adan L, Souberbielle JC, Brauner R: Management of the short stature due to pubertal delay in boys. J Clin Endocrinol Metab 1994, 78:478-482.
33. Siegel SF, Becker DJ, Lee PA, Gutai JP, Foley TP Jr. Drash AL: Comparison of physiologic and pharmacologic assessment of growth hormone secretion. Am J Dis Child 1984, 138:540-543.
34. Zadik Z, Chelew SA, Gilula Z, Kowarski AA: Reproducibility of growth hormone testing procedures: a comparison between 24-hour integrated concentration and pharmacological stimulation. J Clin Endocrinol Metab 1990, 71:1127-1130.

Pre-publication history
The pre-publication history for this paper can be accessed here:

http://www.biomedcentral.com/1471-2431/8/29/prepub