Potential treatment options and future research to increase hepatitis C virus treatment response rate

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Abstract: Hepatitis C Virus (HCV) is a liver-tropic blood-borne pathogen that affects more than 170 million people worldwide. Although acute infections are usually asymptomatic, up to 90% of HCV infections persist with the possibility of long-term consequences such as liver fibrosis, cirrhosis, steatosis, insulin resistance, or hepatocellular carcinoma. As such, HCV-associated liver disease is a major public health concern. Although the currently available standard of care therapy of pegylated interferon α plus ribavirin successfully treats infection in a subset of patients, the development of more effective, less toxic HCV antivirals is a health care imperative. This review not only discusses the limitations of the current HCV standard of care but also evaluates upcoming HCV treatment options and how current research elucidating the viral life cycle is facilitating the development of HCV-specific therapeutics that promise to greatly improve treatment response rates both before and after liver transplantation.

Keywords: interferon, STAT-C, combination therapy, personalized therapy, emerging drug targets, viral life cycle

Hepatitis C virus (HCV) belongs to the Flaviviridae family, which includes other medically relevant viruses such as yellow fever, West Nile, dengue, and tickborne encephalitis viruses, but two characteristics that distinguish HCV are its hepatotropism and tendency to establish chronic infection. Although HCV persists in most infected individuals and is a leading cause of liver disease globally, the development of disease is highly variable, and the mechanisms that mediate disease are not understood. In the absence of a sufficient understanding of HCV infection, we have been left to rely clinically on suboptimal therapeutics leaving more than 170 million people worldwide infected. This review discusses not only the limitations of the current HCV standard of care (SOC), but more importantly the exciting advancements and ongoing research efforts that have the potential to revolutionize HCV clinical care in the foreseeable future.

HCV standard of care

With a worldwide prevalence of 2.2% and no vaccine available, interferon (IFN)-based therapy continues to be the primary intervention option available for the treatment of HCV. In 1991, IFN-α monotherapy was approved for HCV treatment; however, the sustained virological response (SVR) rate was below 20%.¹ In 1998, addition of ribavirin (RBV) was found to improve SVR rates to 34%–42%. In 2001, the development of stabilized pegylated-IFN (PegIFN) further increased the SVR rates to ~50%. Although combination PegIFN/RBV therapy achieves SVR rates of
~80% in individuals infected with genotype (GT) 2 or GT3 and 40%–50% in individuals infected with GT1 or GT4, these rates are still far from ideal, are affected by numerous host specific factors, and the therapy itself has a spectrum of adverse effects (reviewed in Glue et al). To maximize treatment effectiveness while minimizing toxicity, the optimal duration of IFN/RBV therapy is determined by viral GT and the decision to continue treatment depends on treatment response. Patients with GT1 and GT4 are generally treated for 48 weeks with PegIFN-α-2a (180 μg/wk) or PegIFN-α-2b (1.5 μg/kg body weight/wk) in combination with weight-based RBV dosing of 1,000–1,400 mg/d. Patients with GT2 and GT3 are treated for 24 weeks with PegIFN-α-2a or PegIFN-α-2b and RBV. Response parameters that determine whether treatment should be continued include early viral response (EVR) at week 12 and rapid viral response (RVR) at week 4 (reviewed in Poordad et al). EVR, defined as undetectable or $≥ 2-log_{10}$ decrease in serum HCV RNA by week 12 after the initiation of therapy, has long been considered a successful predictor of treatment failure (negative predictive value of 97%–100% for SVR); however, the positive predictive value of EVR to identify patients that will attain SVR is less accurate. As such, RVR, defined as undetectable serum HCV RNA by week 4 after the initiation of therapy, is also being adopted as an additional earlier predictor of treatment outcome.

**Limitations of HCV SOC**

Therapy-associated toxicity represents a major hurdle facing the current treatment. PegIFN/RBV treatment results in significant side effects, which necessitates dose reduction in 35%–42% of those treated, with one-third of this group requiring discontinuation of therapy. The major side effects include influenza-like symptoms (eg, fever, fatigue, headache, myalgias, and arthralgias), hematologic abnormalities (eg, anemia and neutropenia), and neuropsychiatric symptoms (eg, depression and anxiety). Suboptimal efficacy is also a major limitation of PegIFN/RBV SOC, with viral, host, and environmental factors having a significant impact on treatment success. For example, differences in SVR rates indicate that HCV GT1 and GT4 are more resistant to PegIFN/RBV treatment than GT2 and GT3. Independently, high viral load is also associated with poorer response to therapy. Host factors, such as male gender, older age, higher body mass index, coinfections, insulin resistance, and more advanced liver disease all contribute to poorer treatment response. Host ethnicity, in particular, appears to be a major determinant of IFN-based HCV treatment outcome. Most notably, African Americans have significantly lower response rates to PegIFN/RBV compared with other racial groups. Although not as disparate, Hispanics also have poorer response rates to PegIFN/RBV compared with non-Hispanic whites (34% vs 49%, $P < 0.001$) despite similar treatment adherence rates. Although the reasons for these host-related differences in response rates are not completely understood, they highlight that HCV patients may require individualized treatment and monitoring to reach SVR.

Many environmental factors that reduce HCV treatment response are related to adherence. In particular, self-administration of IFN injections and daily RBV dosing can make long-term adherence challenging, especially when the side effects associated with SOC severely impair a patient’s quality of life. Although individualized treatments in conjunction with management of side effects can help maintain adherence, doses often need to be reduced or terminated and failure to meet the assigned dose or duration is associated with lower SVR. Importantly, the variable efficacy of IFN treatment, which is dependent on host-specific immune responses, and other adherence-limiting issues could theoretically be reduced by the development of more specifically targeted HCV inhibitors and combination treatment strategies.

**Emerging treatments and combination therapies**

Currently, new potential HCV inhibitors are being tested clinically in combination with PegIFN and RBV. This not only suggests that IFN, likely in combination with RBV, will remain the backbone of HCV therapy for some time, but also reflects the consensus opinion that, like the highly active retroviral therapy used for the treatment of human immunodeficiency virus (HIV), the future of anti-HCV therapy will take a "cocktail" approach allowing for reduced dosing and treatment duration of toxic compounds while avoiding the viral escape that can occur during monotherapy with single inhibitors. Therefore, new emerging therapies include improved IFN-α formulations (to improve efficacy and ease of administration) and alternative RBV-like molecules (to reduce toxicity); however, the major focus has switched to more targeted inhibitors.

**Interferon α**

Several modified IFN-α products are under clinical development. Albinterferon α-2β (AlbIFN; Human Genome Sciences, Rockville, MD) is IFN covalently bound to albumin, yielding increased half-life. In Phase II clinical trials, AlbIFN dosed
every 2 weeks (BIW) or monthly with daily RBV yielded SVR similar to SOC (which requires weekly PegIFN dosing), with comparable side effects between groups. The Phase III ACHIEVE-1 trial compared AlbIFN BIW plus RBV with SOC in treatment-naive GT1 patients. Patients receiving 1,200 µg AlbIFN discontinued therapy because of pulmonary toxicity, but the patients receiving 900 µg AlbIFN achieved SVR rates comparable to that of SOC (48.7% vs 50.6%), with similar side effects. The Phase III ACHIEVE-2/3 trial compared 900 µg AlbIFN/RBV to SOC in GT2 or GT3 treatment-naive patients and showed comparable SVR in the non-Asian group (79.8% vs 80.5%). However, Asians exhibited higher SVR with SOC when compared with AlbIFN/RBV (95% vs 79%).

Locteron (BLX-883; Biolex Therapeutics, Pittsboro, NC) consists of recombinant IFN-α-2β in a biodegradable, polymeric drug delivery system that allows for biweekly dosing. The Phase IIa SELECT-1 trial evaluated Locteron plus RBV in 32 GT1 treatment-naive patients and found SVR and side effect profiles comparable to historical SOC data. A Phase II SELECT-2 trial is in progress.

Other IFN-α products in clinical development include Belerofon (Nautilus Biotech, Evry, France), which contains a mutation to lower protease sensitivity as a means of increasing stability, and oral IFN (Amarillo Biosciences, Amarillo, TX), which is administered as lozenges (http://www.amarbio.com/clinical-trials.html). In the DIRECT trial high daily dosing of one of the original unmodified IFNs, afacon-1 (Infergen, CIFN), was also recently tested in combination with RBV in patients who were previously nonresponders, but overall, patients showed SVR rates similar to that of other trials involving nonresponders, with only a selected subset of 10 patients with low fibrosis scores (F0–F2) and a > 2-log decrease in virus level achieving a higher SVR of 30% (3/10).

RBV analogs and alternatives

Clinically, the inosine monophosphate dehydrogenase inhibitors, mycophenolate mofetil and merimepodib (VX-497), had little HCV-antiviral efficacy as RBV alternatives; however, liver targeting of RBV is also being tested as a means to reduce RBV-associated toxicities. The RBV prodrug taribavirin (TVB; formerly viramidine; Valeant Pharm., Aliso Viejo, CA), which is activated only in the liver, was developed in an attempt to reduce RBV-induced anemia. Originally, the Phase III VISER-1 and VISER-2 trials reported that TVB was associated with less anemia than RBV and also significantly lower SVR; however, post hoc analysis revealed that patients receiving higher than 18 mg/kg TBV had SVR rates comparable to RBV. Consistent with this finding, a subsequent Phase IIb study of higher TBV doses (20, 25, or 30 mg/kg/d) vs weight-based RBV in 275 GT1 treatment-naive patients showed similar SVR compared with SOC. Notably, the 20 and 25 mg/kg/d TBV groups had less anemia than the RBV group (13.4% and 15.7% vs 35%), but had nearly twice the incidence of diarrhea.

Targeted treatments

As optimized versions of IFN and RBV promise less frequent, more convenient dosing with decreased toxicity, the largest improvements in treatment efficacy in the near future are expected to be achieved by increasing the complexity of the anti-HCV regimen to include more compounds, particularly compounds directly targeting the virus and/or critical virus–host interactions. Specifically targeted antiviral therapy for hepatitis C (STAT-C) is a term originally coined to describe inhibitors that target HCV proteins. Although issues of viral escape will have to be managed, these targeted therapies in theory should be less dependent on host-specific responses and avoid side effects associated with host immune stimulation. The two types of STAT-C molecules that have progressed furthest and are expected to be the first available clinically are NS3/4a protease and NS5B polymerase inhibitors.

NS3/4a protease inhibitors

The HCV NS3 serine protease and NS4a cofactor assemble to cleave the viral polyprotein into four nonstructural proteins (Figure 1); blocking this cleavage, therefore inhibits viral replication. Inhibition of NS3/4a also restores host innate immune signaling that is otherwise suppressed when NS3/4a cleaves the Toll-IL-1 receptor domain (TRIF) and IFN-β promoter stimulator (IPS-1, CARDIF, VISA), a mechanism by which HCV prevents dsRNA-induced IFN signaling. Initial proof of concept protease inhibitor clinical studies were conducted with BILN2061 (Boehringer Ingelheim, Ingelheim, Germany), which was halted due to cardiac toxicity in an animal model; however, two other protease inhibitors have progressed to Phase III trials.

Telaprevir (TVR; VX-950; Vertex Cambridge, MA) is a peptidomimetic NS3/4a protease inhibitor that covalently but reversibly binds NS3/4a. It is dosed orally every 8 hours. In Phase I, TVR monotherapy achieved a 4.4-log mean reduction in viral load over 14 days, but rapid selection of viral resistance at four different sites (T54, V36, R155, and A156) was observed. To minimize viral breakthrough, TVR has
since been studied in combination with PegIFN/RBV and three Phase II PROVE studies indicate that addition of TVR to SOC allows for shorter therapy duration in GT1 patients while increasing SVR ~20%–30% in treatment-naive patients and ~30% in previous nonresponders (Tables 1–3).

- PROVE 1 randomized treatment-naive GT1 patients in the United States into three arms of 12 weeks TVR plus varying durations of PegIFN/RBV (12, 24, or 48 weeks) compared with 48 weeks SOC (Table 1).44 Inferior SVR in the 12-week TVR/IFN/RBV arm showed the need for continued PegIFN/RBV treatment after the initial 12 weeks of triple therapy; however, TVR in combination with 24 or 48 weeks PegIFN/RBV improved SVR by 20% or more. Viral breakthrough occurred in 12/175 of patients receiving TVR who did not achieve viral suppression with breakthrough rates higher for GT1a than GT1b because the most common TVR resistance mutation (R155T/I) requires one nucleotide change in GT1a compared with two nucleotide changes in GT1b.45 Rash, gastrointestinal toxicity, and anemia were the main adverse effects associated with TVR requiring discontinuation of treatment in 18% of TVR subjects, most frequently due to severe rash (7%).45

- PROVE 2 studied treatment-naive GT1 patients receiving 12 weeks of TVR in combination with 12 weeks of PegIFN alone or PegIFN/RBV for 12 or 24 weeks (Table 2).45 Again, 12 weeks triple therapy followed by an additional 12 weeks of PegIFN/RBV resulted in more than a 20% increase in SVR compared to SOC (69% vs 46%). Overall discontinuation due to side effects was 11% in the TVR arms and 7% for SOC, again with severe rash in 5% of TVR patients. Notably, 24% viral breakthrough in patients taking TVR/PegIFN without RBV revealed the importance of RBV in the treatment regimen.

- PROVE 3 evaluated GT1 patients who previously failed SOC therapy (Table 3).46 Importantly, the addition of TVR to SOC increased SVR in these treatment experience patients. Again, viral breakthrough was highest in the group that did not receive RBV (21% vs 3%–11%) and severe rash occurred in 3%–5% of TVR patients.

Three Phase III trials (ADVANCE, ILLUMINATE, and REALIZE) with TVR in combination with various lengths of PegIFN/RBV therapy are in progress.

Boceprevir (BVR) (formerly SCH503034; Schering, Kenilworth, NJ) is another NS3/4a protease inhibitor administered orally every 8 hours that has entered Phase III clinical trials. In a Phase Ib trial with GT1 nonresponders, 400 mg BVR 3 times per day (TID) monotherapy resulted in a 1.61-log HCV reduction at day 14,47 but analogous to TVR, BVR monotherapy led to rapid selection of drug-resistant mutations including V36M/A, T54A/S, V55A, R155K/T, A156S, and V170A.48 BVR has
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since been studied in combination with PegIFN/RBV and has been found to improve SVR in GT1 treatment-naive patients, but might require 48 weeks of triple therapy for optimal results.

• The Phase II SPRINT-1 study assessed 800 mg TID BVR in combination with PegIFN/RBV in treatment-naive GT1 patients.49 Initial treatment arms were used to assess the use of a 4-week PegIFN/RBV lead-in for reducing viral load and hence resistance during subsequent triple BVR/PegIFN/RBV (Table 4). Although the 4-week PegIFN/RBV lead-in made no difference in SVR of patients in the 28-week therapy arms (56% and 55%), SVR was slightly higher in the 48-week therapy arm that received the PegIFN/RBV lead-in (74% vs 66%). More notably however, the lead-in arms showed higher RVR (64% vs 37%–40%) and fewer virological breakthroughs (4%–5% vs 7%–12%). In a subsequent branch of the study, low-dose RBV was associated with higher viral breakthrough and lower SVR rates (36%), demonstrating again the importance of RBV.

• In another Phase II trial in GT1 nonresponders, BVR/PegIFN/RBV or BVR/PegIFN alone yielded SVR of 7%–14% vs 2% for SOC; however, adjustments where made during the trial such as increasing BVR dosing and adding RBV to all groups, making it difficult to interpret the results (reviewed in Thompson and McHutchison).50

Two Phase III trials, SPRINT-2 and RESPOND-2, are underway in treatment-naive and nonresponders, respectively.

Many other HCV NS3/4a inhibitors are currently in preclinical and clinical development. Some inhibitors that have reached Phase II (Table 5) include the following:

• **MK7009** (Merck, Whitehouse Station, NJ) is a noncovalent competitive inhibitor of the HCV NS3/4a pro-

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**Table 1** Phase II clinical data from protease inhibitors in ongoing Phase III development: TVR PROVE 1

| Arms | n  | RVR (%) | SVR (%) | Relapse (%) | Halted (%) |
|------|----|---------|---------|-------------|------------|
| 260 Noncirrhotic treatment-naive GT1 (USA) |    |         |         |             |            |
| TVR/PR 12 wk | 17 | 59      | 35      | 33          | 18         |
| TVR/PR 12 wk + PR 12 wk (24 wk total) | 79 | 81      | 61      | 2           |            |
| TVR/PR 12 wk + PR 36 wk (48 wk total) | 79 | 81      | 67      | 6           |            |
| SOC 48 wk | 85 | 11      | 41      | 23          | 4          |

Notes: P = PegIFN; R = RBv; SOC = PegIFN/RBv; ”/” = in combination with; “+” = followed by.

**Table 2** Phase II clinical data from protease inhibitors in ongoing Phase III development: TVR PROVE 2

| Arms | n  | RVR (%) | SVR (%) | Relapse (%) | Halted (%) |
|------|----|---------|---------|-------------|------------|
| 334 Treatment-naive GT1 (Europe) |    |         |         |             |            |
| TVR/P 12 wk | 78 | 51      | 36      | 48          | 10         |
| TVR/PR 12 wk | 82 | 80      | 60      | 30          | 11         |
| TVR/PR 12 wk + PR 12 wk | 81 | 69      | 69      | 14          | 14         |
| SOC 48 wk | 93 | 13      | 46      | 22          | 7          |

Notes: P = PegIFN; R = RBv; SOC = PegIFN/RBv; ”/” = in combination with; “+” = followed by.

**Table 3** Phase II clinical data from protease inhibitors in ongoing Phase III development: TVR PROVE 3

| Arms | SVR (%) Total | Prior nonresponder | Prior relaper | Prior viral resistance |
|------|--------------|--------------------|--------------|-----------------------|
| 453 Treatment failure GT1 |                |                    |              |                       |
| TVR/PR 12 wk + 12 wk PR | 51            | 39                 | 69           | 57                    |
| TVR/PR 24 wk + 24 wk PR | 53            | 38                 | 76           | 63                    |
| TVR/P 24 wk | 24 | 11 | 42 | 36 |
| SOC 48 wk | 14 | 9 | 20 | 40 |

Notes: P = PegIFN; R = RBv; SOC = PegIFN/RBv; ”/” = in combination with; “+” = followed by.
tase that was shown to have antiviral activity when administered for 8 days as monotherapy. In Phase II trials combining various doses of MK7009 with PegIFN/RBV for 28 days, higher RVR was achieved in patients receiving MK7009 when compared to those in the SOC control group (70%–83% vs 5.6%), and all patients receiving 600 mg BID MK7009 were HCV RNA undetectable at day 42 of treatment. Virological failure in the 300 mg BID and 800 mg/d dosing groups were associated with HCV resistance mutations R155K and D168V. No serious adverse events were noted.

- **TMC435** (TMC435350-C201; Medivir/Tibotec, Stockholm, Sweden) monotherapy led to a 3.9-log mean reduction in previous treatment failures in Phase I trials. In the ongoing Phase II OPERA-1 trials of GT1 treatment-naive and treatment-experienced patients, arms include varying doses of TMC435 with PegIFN/RBV for 4 weeks followed by 24 or 48 weeks of PegIFN/RBV. At treatment week 4, TMC435/PegIFN/RBV arms have achieved mean HCV RNA reductions of 4.3–5.5 logs compared with 1.5 logs in the placebo/PegIFN/RBV arm with no increase in adverse reactions.

- **R7227** (formerly ITMN191; Intermune/Roche, Brisbane, CA) in multiple 14-day Phase I studies decreased HCV RNA levels 2.5–3.8 logs as monotherapy and 5.5 logs when administered 600 mg/d with PegIFN/RBV compared a 2-log reduction in the SOC only. In the combination study, no viral rebound occurred in any of the R7227 arms, and 13%–57% of R7227/PegIFN/RBV recipients had undetectable HCV RNA vs none with SOC. Adverse events were similar to SOC, except hyperbilirubinemia developed in two patients.

- **BI201335** (Boehringer Ingelheim) in a Phase I trial led to ~5 log reductions in viral load in treatment-experienced patients after 28 days of combination treatment of BI201335 (240 mg/d or BID) with PegIFN/RBV. Phase II studies are ongoing in treatment-naive patients and cirrhotic nonresponders.

### HCV polymerase inhibitors

The HCV RNA-dependant RNA polymerase, NS5B, synthesizes the viral RNA and thus plays a key role in viral replication. Although final triple therapy SVR results are still pending, current data indicate that HCV polymerase inhibitors tend to display lower potency during monotherapy than NS3/4a protease inhibitors (Table 6 vs Table 5), but show broader effectiveness against multiple GTs and can exhibit a higher barrier to viral resistance. Both nucleoside analog inhibitors (NIs) and nonnucleoside inhibitors (NNIs) are in development (Table 6). NIs target the highly conserved active site of NS5B by mimicking the polymerase substrate and hence tend to be more potent and have lower resistance rates than NNIs. NNIs interact with the NS5B polymerase outside of the catalytic site, causing allosteric changes that compromise NS5B complex formation and function. Five binding sights have been identified; however, rapid resistance can limit these targets.

Two oral NIs in Phase II development are R7128 (Roche/Pharmasset, Basel, Switzerland) and IDX184 (Idenix, Cambridge, MA). While a 2-week 1,500 mg BID R7128 monotherapy trial showed a 2.7-log HCV RNA reduction, a 4-week combination trial of R7128 (1,000 and 1,500 mg BID) with PegIFN/RBV in treatment-naive GT1 patients gave a 5-log mean reduction in HCV RNA vs a 2-log decrease in the SOC arm, resulting in higher RVR (88% and 85%) than SOC alone (19%). Likewise, in a Phase Ib study in GT2/3 relapsers and nonresponders, addition of R7128 was found to increase RVR (90%) compared with SOC alone (60%). No resistance was identified at 4 weeks. Phase II R7128 trials are currently enrolling patients. In a Phase I IDX184 study, treatment-naive GT1 patients receiving 25–100 mg/d IDX184 had mean HCV RNA reductions ranging from 0.47–0.74 log at 2 weeks, with no serious adverse events (Table 6). However, IDX184 is synergistic with PegIFN/RBV in vitro, and results from an ongoing Phase IIa trial indicate a 4-log mean HCV RNA reduction at day 14 after the combination therapy using IDX184 (50 mg BID or 100 mg/d) with PegIFN/RBV.

Several NNIs have also begun Phase II development (Table 6). In Phase I trials, ANA596 (Anadys Pharmaceuticals,}

| Arms                          | RVR (%) | SVR (%) | Breakthrough (%) | Halted (%) |
|-------------------------------|---------|---------|------------------|------------|
| 520 Treatment-naive GT1       |         |         |                  |            |
| 4 wk PR + BVR/PR 24 wk (28 wk total) | 64      | 56      | 4                | 26         |
| 4 wk PR + BVR/PR 44 wk (48 wk total) | 64      | 74      | 5                | 26         |
| BVR/PR 28 wk                  | 37      | 55      | 7                | 28         |
| BVR/PR 48 wk                  | 40      | 66      | 11               | 38         |
| SOC 48 wk                     | 8       | 38      | 0                | 15         |

**Notes:** P = PegIFN; R = RBV; SOC = PegIFN/RBV; “/” = in combination with; “+” = followed by.
Table 5  Phase 2 clinical data from NS3/4a inhibitors

| NS3/4a inhibitors in phase 2 | Arms | Log HCV reduction | RVR (%) |
|-----------------------------|------|-------------------|---------|
| MK7009 (Merck) | 25 mg BID MK7009 8 d | 1.9 | – |
| (29 GT1 treatment-naive and experienced) | 75 mg BID MK7009 8 d | 2.5 | – |
| 25 mg BID MK7009 8 d | 2.8 | – |
| 500 mg BID MK7009 8 d | 3.3 | – |
| 700 mg BID MK7009 8 d | 4.6 | – |
| 125 mg/d MK7009 8 d | 1.8 | – |
| 600 mg/d MK7009 8 d | 2.4 | – |
| Phase Iib; (85 GT1 treatment-naive) | 300 mg BID MK7009/PR 4 wk | 75 | – |
| 600 mg BID MK7009/PR 4 wk | 79 | – |
| 600 mg/d MK7009/PR 4 wk | 69 | – |
| 800 mg/d MK7009/PR. 4 wk | 82 | – |
| SOC 4 wk | 5.6 | – |
| TMC435 (Medivir/Tibotec) | 200 mg/d TMC 6 d | 3.9 | – |
| Phase I; (6 GT1 experienced) | 25 mg/d TMC/PR 4 wk + SOC 24/48 wk | 4.7 | – |
| (GT1 treatment-naive) | 75 mg/d TMC/PR 4 wk + SOC 24/48 wk | 5.5 | – |
| 200 mg/d TMC/PR 4 wk + SOC 24/48 wk | 5.4 | – |
| Phase Iib; (GT1 experienced) | 75 mg/d TMC/PR 4 wk + SOC 24/48 wk | 4.3 | – |
| 150 mg/d TMC/PR 4 wk + SOC 24/48 wk | 5.5 | – |
| 200 mg/d TMC/PR 4 wk + SOC 24/48 wk | 5.3 | – |
| SOC 24/48 wk | 1.5 | – |
| R7227 (Intermune/Roche) | 600 mg/d R7227 14 d | 3.8 | – |
| Phase I; (GT1 naive) | 600 mg/d R7227 14 d | 2.5 | – |
| Phase I; (GT1 experienced) | 100 mg R7227/PR 14 d | 4.7 | – |
| Phase I; (57 GT1 treatment-naive) | 600 R7227/PR 14 d | 5.7 | – |
| SOC 14 d | 2.0 | – |
| BI201335 (Boehringer Ingelheim) | 20 mg/d BI 14 d | 3.0 | – |
| Phase I; (34 GT1 treatment-naive) | 40 mg/d BI 14 d | 3.6 | – |
| 120 mg/d BI 14 d | 3.7 | – |
| 240 mg/d BI 14 d | 4.2 | – |
| Phase I; (19 GT1 experienced) | 48 mg/d BI/PR 4 wk | 5.0 | – |
| 120 mg/d BI/PR 4 wk | 5.2 | – |
| 240 mg/d BI/PR 4 wk | 5.3 | – |
| Phase Ib; (13 GT1 nonresponders) | 240 mg BI/PR 4 wk | 5.4 | – |
| Phase II; (GT1 treatment-naive/ongoing) | 240 mg/d BI/PR 28 wk | 92 | – |
| 240 mg BI 3 d + 240 mg/d BI/PR 28 wk | 84 | – |
| 120 mg BI 3 d + 240 mg/d BI/PR 28 wk | 90 | – |
| SOC 28 wk | 16 | – |

Notes: P = PegIFN; R = RBV; “/” = in combination with; “+” = followed by.

San Diego, CA) at 200, 400, or 800 mg BID for 3 days achieved log reductions of 2.4, 2.3, and 2.9, respectively, without viral rebound or serious adverse events. In an ongoing Phase II study, patients are receiving ANA569 (200 or 400 mg BID) with PegIFN/RBV for 12 weeks followed by 12 or 36 weeks of SOC. RVR rates of 42%–56% for the ANA596 arms compared with only 13% with SOC have been reported.65 Filibuvir (FBV; formerly PF868554; Pfizer, New York, NY) is another NNI. When given as monotherapy for 8 days (100–450 mg BID or 300 mg TID), FBV demonstrated dose-dependent inhibition of HCV in treatment-naive GT1 patients, with mean HCV RNA reductions of 0.97–2.13 log.65 A second Phase I study in which FBV (200, 300, or 500 mg BID) was given in combination with PegIFN/RBV for 4 weeks showed enhanced mean reductions of 3.62, 4.46, and 4.67 log, respectively, compared with 2.10 logs in the SOC-alone group. The Phase II FITNESS trial is currently enrolling patients to receive FBV/PegIFN/RBV for 24 weeks with or without an additional 24 weeks of PegIFN/RBV.

Protease and polymerase inhibitor combination therapy

Although data thus far indicate that IFN and RBV help prevent the emergence of viral resistance when used in
Table 6 Phase 2 clinical data from NS5B inhibitors

| NS5B inhibitors in phase 2 | Arms | Log HCV reduction | RVR (%) |
|---------------------------|------|-------------------|---------|
| **R7128** (Roche/Pharmasset) | **Phase I** | 1500 mg BID R7128 2 wk | 2.7 | – |
| (NI) | **Phase 1b; (81 GT1 treatment-naive)** | 1000 mg BID R7128/PR 4 wk | 5 | 88 |
| | | 1500 mg BID R7128/PR 4 wk | 5 | 85 |
| | | SOC 4 wk | 2 | 19 |
| | **Phase 1b; (25 GT2/3 treatment-experienced)** | 1500 mg BID R7128/PR 4 wk | 5 | 90 |
| | | SOC 4 wk | 3.7 | 60 |
| **IDX184** (Idenix) | **Phase 1; (12 GT1 treatment-naive)** | 25 mg/d IDX184 14 d | 0.47 | – |
| (NI) | | 50 mg/d IDX184 14 d | 0.69 | – |
| | | 75 mg/d IDX184 14 d | 0.70 | – |
| | | 100 mg/d IDX184 14 d | 0.74 | – |
| | **Phase IIa; (GT1 treatment-naive)** | 50 mg/d IDX184/PR 14 d + PR 14 d | 2.7 (14 d) | – |
| | | 100 mg/d IDX184/PR 14 d + PR 14 d | 4.2 (14 d) | – |
| **Filibuvir** (Pfizer) | **Phase I; (32 GT1 treatment-naive)** | 100 mg BID FVR 8 d | 0.68 | – |
| (NNI) | | 300 mg BID FVR 8 d | 1.26 | – |
| | | 450 mg BID FVR 8 d | 1.21 | – |
| | | 300 mg TID FVR 8 d | 1.95 | – |
| | **Phase Ib; (35 GT1 treatment-naive)** | 200 mg FVR/PR × 28 d + 44 wk PR | 3.6 | 60 |
| | | 300 mg FVR/PR × 28 d + 44 wk PR | 4.5 | 75 |
| | | 500 mg FVR/PR × 28 d + 44 wk PR | 4.7 | 63 |
| | | SOC 48 wk | 2.1 | 0 |
| **ANA 598** (Anadys) | **Phase I; (10 GT1 treatment-naive)** | 200 mg BID ANA 3 d | 2.4 | – |
| (NNI) | | 400 mg BID ANA 3 d | 2.3 | – |
| | | 800 mg BID ANA 3 d | 2.9 | – |
| | **Phase II; (GT1 treatment-naive)** | 200 mg BID ANA/PR 12 wk + PR 12/36 wk | – | 56 |
| | | 400 mg BID ANA/PR 12 wk + PR 12/36 wk | – | 42 |
| | | SOC 24 wk or 48 wk | – | 13 |
| **VCH 759** (Vertex) | **Phase I; (GT1 treatment-naive)** | 400 mg TID VCH 759 10 d | 1.0 | – |
| (NNI) | | 800 mg BID VCH 759 10 d | 1.0 | – |
| | | 800 mg TID VCH 759 10 d | 2.5 | – |
| **GS 9190** (Gilead) | **Phase I; (31 GT1 treatment-naive)** | 40 mg GS9190 8 d | 1.4 | – |
| (NNI) | | 120 mg GS9190 8 d | 1.7 | – |
| **BI 207127** (Boehringer Ingelheim) | **Phase I; (60 GT1 treatment-naive)** | 100 mg TID BI207127 5 d | 0.4 | – |
| (NNI) | | 200 mg TID BI207127 5 d | 0.8 | – |
| | | 400 mg TID BI207127 5 d | 1.3 | – |
| | | 800 mg TID BI207127 5 d | 3.8 | – |
| | | 1200 mg TID BI207127 5 d | 3.2 | – |

Notes: P = PegIFN; R = RBV; “/” = in combination with; “+” = followed by

combination with protease inhibitors, dual treatment of the protease inhibitor R7227 and the polymerase inhibitor R7128 was also found to suppress the emergence of R7227 resistance in the replicon system.67 Using this strategy, the INFORM-1 trial evaluated escalating doses of R7227 and R7128 in GT1 treatment-naive, treatment-experienced, and IFN-null responders.68 Overall, patients who received R7227/R7128 therapy had a mean 4.9-log HCV RNA reduction at day 14 without resistance or serious adverse effects. SVR data are pending. In the meantime, Phase II INFORM-2 trials are enrolling patients to receive 4 weeks of R7227/R7128 alone or in combination with PegIFN, RBV, or PegIFN/RBV. Additional studies testing other compound combinations are also being conducted (eg, VX-950 + VCH222, ClinicalTrials.gov Identifier: NCT00958152; BMS790052 + BMS650032, ClinicalTrials.gov Identifier: NCT00904059; NCT01012895).

Elucidation of the viral life cycle and identification of novel drug targets

Each step of the viral life cycle represents a potential drug target; however, progress in the development of specifically targeted HCV inhibitors has been hindered because the experimental cell culture and small animal models needed
to elucidate the HCV life cycle and host–virus interactions that determine infection outcome have not been available. Notably however, the development of HCV replicons,69–72 HCV pseudotyped particles (pp)73 and most recently the infectious HCV cell culture system74–76 have advanced our understanding of the viral life cycle (Figure 2) resulting in the identification of unprecedented number of putative inhibitors directed against both virus and host targets (Table 7). Of course, with these tools now available, each step of the HCV life cycle is being further studied to identify and test potentially more effective drug targets.

HCV entry

HCV infection begins with binding of the virus to cell surface receptors. Four cellular receptors – the tetraspanin protein CD81,73,76–79 the scavenger receptor class B member I (SR-B1),73,80–83 and the tight junction proteins claudin-184 and occludin85–87 have all been shown to be required for HCV entry (Figure 2, step 1). In addition, the low-density lipoprotein receptor,99,88–90 asialoglycoprotein receptor,91 and glycosaminoglycans (heparin sulfate) have been implicated, but their exact roles have not been determined. Following this multistep binding process, HCV enters the cell via clathrin-mediated endocytosis92,93 (Figure 2, step 2). Fusion between the viral envelope and endosomal membrane occurs in the acidified endosomal compartment77,94–98 via E1/E2-mediated class II fusion,99,100 ultimately resulting in the release of the viral genomic RNA into the cytoplasm (Figure 2, step 3). With the recent identification of several new HCV entry factors and progress identifying antibodies that neutralize the virus,101–104 we will likely see a wave of novel HCV entry inhibitors in clinical evaluation soon. Entry inhibitors in clinical trials now include the following:

- **Civacir** (Nabi Biopharmaceuticals, Rockville, MD) is a pool of fractionated and concentrated immunoglobulin from HCV antibody-positive human plasma that was shown to neutralize HCV in preclinical development. In a Phase I trial, 18 HCV-positive transplant recipients received a total of 17 infusions of 75 mg/kg, 200 mg/kg, or no treatment at the time of transplantation and for the following 14 weeks. The drug was well tolerated and associated with reduced aminotransferase (ALT) levels, but did not suppress hepatitis C viremia.105 The use of higher doses is being studied.

- **ITX5061** (iTherX Pharmaceuticals, San Diego, CA) is an SR-B1 inhibitor106 that blocks HCV entry in vitro and has demonstrated both preclinical and clinical safety.107 Phase Ib and IIa proof-of-concept studies in HCV patients have been initiated.

**Viral protein synthesis and processing**

After the ~9.6 kb viral RNA genome is released into the host cell cytoplasm, it serves as template for the translation of the viral proteins which are encoded in a single open-reading frame flanked by highly structured 5' and 3' untranslated regions (UTRs) (Figure 1). The 5' UTR contains an internal ribosome entry site (IRES) for cap-independent translation of the ~3,010-amino acid viral polyprotein (Figure 2, step 4), which is cleaved by cellular and viral proteases into individual structural and nonstructural (NS) proteins (Figure 2, step 5). Although past efforts targeting HCV IRES translation with antisense oligonucleotides, ribozyme-directed cleavage, and siRNAs have been halted due to the lack of response and/or toxicity, the positive clinical performance of NS3/4a...
protease inhibitors highlights viral protein production as an effective drug target.

**Viral replication**

Once expressed, the viral NS proteins (NS2-NS5B) assemble on and remodel endoplasmic reticulum membranes to form viral replication complexes (known as the membranous web; Figure 2, step 6). HCV RNA replication occurs within these membranous structures in a two-step process in which the viral NS5B polymerase uses the input genome as template to synthesize a negative-strand RNA intermediate, which then provides the template for ∼10-fold amplification of positive-strand genomic RNA (Figure 2, step 7). In addition to NS5B polymerase inhibitors, there is a growing list of compounds that block viral replication by inhibiting one of the other HCV NS proteins or by targeting a host cellular protein shown to be essential for viral replication. Some of those currently in clinical development include the following:

- **NS5A inhibitors**: The viral NS5A protein is essential for HCV replication, virus assembly, and also plays a role in

### Table 7 Specifically targeted HCV inhibitors (April 2010)

| Stage in viral life cycle | Viral/host target | Inhibitor class | Agent | Phase |
|---------------------------|-------------------|-----------------|-------|-------|
| Entry                     | E1/E2             | Neutralizing antibody | Civacir | 2     |
|                           | SR-B1             | SR-B1 inhibitor | ITX5061 | 2     |
| Protein synthesis         | NS3/4a            | Protease inhibitors | Telepravir | 3     |
|                           |                   |                 | Boceprevir | 3     |
|                           |                   |                 | TM 435 | 2     |
|                           |                   |                 | MK 7009 | 2     |
|                           |                   |                 | BI 201335 | 2     |
|                           |                   |                 | R 7227 (ITMN-191) | 2     |
|                           |                   |                 | SCH 900518 | 2     |
|                           |                   |                 | BMS-650032 | 2     |
|                           |                   |                 | VX 813 | 1     |
|                           |                   |                 | VX 985 | 1     |
|                           |                   |                 | ABT 450 | 1     |
|                           |                   |                 | VX 500 | 1     |
|                           |                   |                 | PHX 1766 | 1     |
|                           |                   |                 | ACH 1625 | 1     |
| Genome replication        | N55B              | Nucleoside analog | R 7128 | 2     |
|                           |                   |                 | IDX 184 | 2     |
|                           |                   |                 | PSI-7977 | 2     |
|                           |                   | Nonnucleoside analog | ANA 598 | 2     |
|                           |                   |                 | PF 868554 (Filibuvir) | 2     |
|                           |                   |                 | GS 9190 | 2     |
|                           |                   |                 | BI 207127 | 2     |
|                           |                   |                 | A 837093 | 1     |
|                           |                   |                 | VCH 759 | 1     |
|                           |                   |                 | ABT 333 | 1     |
|                           |                   |                 | VCH 916 | 1     |
|                           |                   |                 | VX 222 (VCH222) | 1     |
|                           |                   |                 | R0524048 | 1     |
|                           |                   |                 | IDX 375 | 1     |
|                           |                   |                 | ABT-072 | 1     |
|                           | N55A              |                 | BS 790052 | 2     |
|                           |                   |                 | A-832 | 2     |
|                           |                   |                 | A-689 | 1     |
|                           |                   |                 | PPI-461 | Pre   |
|                           | NS4B              |                 | Clemizole | 1     |
|                           | mR-122            |                 | SPC3649 | 1     |
|                           | Cyclophilins      |                 | Debio025 | 2     |
|                           |                   |                 | NIM811 | 2     |
|                           |                   |                 | SYC635 | 1     |
|                           | HMGCo-A           |                 | Statins | 2     |
| Assembly/secretion        | Glucosidases      |                 | Celgosivir | 2     |
|                           | LDL pathway       |                 | Naringenin | Pre   |
IFN resistance. As such, it represents a promising virus-specific drug target (Table 1). In a phase 1 study, a single dose of the NS5A inhibitor BMS-790052 (Bristol-Myers Squibb, New York, NY) was found to be well tolerated in HCV GT1 patients at 1, 10, and 100 mg doses with an average mean log reduction of 1.8 logs, 3.2 logs, and 3.3 logs, respectively, at 24 hours. Phase II studies comparing SOC ± BMS-790052 are underway.

- **NS4B inhibitors:** NS4B plays a major role in replication by inducing the membrane alteration required for formation of the HCV replication complexes. One NS4B inhibitor, Clemizole (Eiger BioPharmaceuticals, Palo Alto, CA), has recently entered Phase I trials.

- **HMG CoA reductase inhibitors:** In vitro studies indicate that HCV replication is dependent on interaction between FBL2 and NS5A. FBL2 is a host protein that requires anchoring by geranylgeranyl pyrophosphate (GGP), an intermediate in cholesterol synthesis. Because statins inhibit HMG CoA reductase lowering GGP levels, statins have anti-HCV activity in vitro. Although some in vivo studies have shown no antiviral effect, a recent post hoc analysis of the IDEAL study revealed a correlation between elevated cholesterol as well as statin use and SVR to PegIFN/RBV. Hence, targeting lipid metabolism as an additive treatment remains an area of interest.

- **Cyclophilin inhibitors:** Several laboratories have identified a number of cyclophilins to be involved in HCV replication (reviewed in Gallay and Gaither et al), and cyclophilin inhibitors have been shown to have anti-HCV activity in vitro. A Phase II trial evaluated the cyclophilin inhibitor DeBio-025 (DebioPharm, Lausanne, Switzerland) alone or in combination with PegIFN/RBV compared with SOC for 29 days. There was a 4.6-log reduction in HCV RNA with triple therapy (vs 2.5 logs in the SOC group, and 2.2 logs in the DeBio-025 monotherapy group). DeBio-025 was associated with reversible conjugated hyperbilirubinemia and hypertension. Phase II trials using combination therapy with PegIFN/RBV in treatment-naive GT1 patients are underway. Other cyclophilin inhibitors in clinical development are SCY-635 (ScyNexis, Research Triangle Park, NC) and NIM 811 (Novartis, Basel, Switzerland).

- **miR-122 inhibitors:** Multiple binding sites for the liver-specific microRNA miR-122 have been identified in the HCV genome, and sequestering miR-122 using antisense oligonucleotides results in the inhibition of HCV infection in cell culture. Importantly, preclinical analysis of an miR-122 inhibitor, LNA-antimiR/SPC3649 (Santaris Pharma, Copenhagen, Denmark), in HCV-infected chimpanzees was well tolerated, and HCV RNA levels were reduced 1.3–2.6 logs depending on the dose (1 and 5 mg/kg, respectively) without rebound (which occurred ~10 weeks after treatment conclusion).

### Progeny virus assembly and release

After positive-strand HCV RNA has been replicated in the cell, this RNA can serve as a template for additional translation and negative-strand synthesis or be packaged into progeny virus (Figure 2, step 8). Although the molecular details are not well defined, the viral capsid is believed to become enveloped when it buds into the endoplasmic reticulum and then exits the cell via the lipoprotein secretory pathway (Figure 2, step 9). With the availability of the cell culture HCV infection system, the viral proteins and virus–host interactions that mediate capsid assembly, envelopment, and virion maturation are now being elucidated, providing a plethora of promising drug targets (eg, viral proteins p7, NS2, and HCV core dimerization, as well as host targets related to lipoprotein secretion).

- **Lipoprotein synthesis inhibitors:** The body of evidence linking the HCV life cycle to lipid metabolism continues to grow, whether it is the need for HCV core to associate with lipid droplets, the dependence of particle morphogenesis on vLDL pathway components, or the presence of lipoproteins on the viral particle being involved in particle entry (reviewed in Popescu and Dubuisson). Along these lines, it has been shown in cell culture that the secretion of infectious HCV particles is blocked by the vLDL secretion inhibitor, naringenin, found in grapefruit.

- **α-Glucosidase inhibitors:** α-Glucosidases I and II are host enzymes involved in glycoprotein processing, but viruses are more sensitive to decreases in the activity of these enzymes resulting in misfolding of the viral glycoproteins and inhibition of viral assembly and release. Celgosivir (Mignex, Vancouver, Canada) is a prodrug of castanospermine that has been shown to inhibit α-glucosidase I (reviewed in Durantel). A Phase II trial in GT1 patients showed that 200 and 400 mg celgosivir once daily, and 200 mg BID were tolerable, but of the 35 patients who completed the 12-week treatment, only two had peak reductions of ≥1 log. Phase II combination trials of celgosivir + PegIFN/RBV are underway.

### Identification of new drug targets

To identify new drug targets on a more global scale, high-throughput screening (HTS) approaches using the infectious
HCV cell culture system are rapidly being developed. Yu et al\textsuperscript{136} recently developed a simple mix-and-measure fluorescence resonance energy transfer (FRET)-based in vitro HCV infection HTS capable of identifying inhibitors that act at any point in the viral life cycle. In addition, high-throughput assays are being used to screen siRNA libraries to functionally identify host factors involved in HCV infection.\textsuperscript{137–140} For example, Li et al\textsuperscript{140} reported a two-step screening approach in which imaging for HCV core protein in the initially inoculated culture was performed to identify siRNA-silenced genes that act early in infection, whereas imaging for HCV core protein in secondary cell cultures exposed to supernatant from the initial infection was used to identify genes that act late during infection. With these efforts ongoing, the list of targeted inhibitors is expected to grow.

**Therapeutics to boost HCV immune response and limit viral pathology**

In addition to elucidating the details of the viral life cycle, efforts to better understand the immunological basis of spontaneous HCV clearance and the mechanism of action of IFN and RBV are underway to provide insight into the host immune mechanisms that may be used to clear the virus. Other studies are focused on understanding the mechanistic basis of HCV-associated liver disease. Hence, alternative immunomodulators and antifibrotics are emerging as possible HCV therapeutics (Table 8).

**Immunomodulators**

Although specifically targeted HCV therapeutics are currently getting a great deal of attention, concerns about viral escape and past success with IFN are still driving the development of additional immunomodulators as potential therapeutics for HCV infection.

**Alternative IFNs**

Many different IFNs have been identified in humans and several are being considered as possible HCV therapeutics. Omega IFN is a type 1 IFN that shares 62% amino acid identity with IFN-\(\alpha\)-2\textsuperscript{143} and has been shown to have anti-HCV activity in vitro.\textsuperscript{142} In a Phase II study of GT1 treatment-naive patients, daily omega-IFN plus RBV led to a SVR of 37%.\textsuperscript{146}–\textsuperscript{148} The type III IFN, IFN-\(\lambda\) (IL-29), has demonstrated inhibitory activity against HCV in vitro\textsuperscript{144,145} and during a 4-week Phase I trial PegIFN-\(\lambda\) plus RBV achieved a 2-log HCV RNA reduction in GT1 treatment-naive patients and previous relapsers, most notably without the flu-like symptoms or hematological alterations associated with IFN-\(\alpha\).\textsuperscript{146–148} Phase II trials have begun in treatment-naive patients.

**Toll-like receptor agonists**

Toll-like receptors (TLRs) and RIG-I-like receptors (RLRs) are pattern recognition receptors that recognize structurally conserved molecules shared by pathogens that distinguish them from host molecules. Binding to TLRs and RLRs activate host cytokine signaling defenses, thus similar to IFN, TLR and RLR agonists stimulate host immune defenses. Although some TLR agonists have been halted due to side effects, other compounds in this class are under clinical evaluation. ANA 773 (Anadys Pharmaceuticals, San Diego, CA) induces IFN via the TLR7 pathway and in a Phase I trial ANA773 every other day (QOD) reduced HCV RNA 1.3 logs compared with a 0.3-log decline with placebo (\(P = 0.037\))\textsuperscript{149} with no serious adverse events. IMO2125 (Idera Pharmaceuticals) is a TLR9 ligand that has shown anti-HCV activity in HCV replicon studies and is now entering Phase I studies.\textsuperscript{150} Two other immunomodulatory dipeptides being tested are Oglufanide (formally IM862; Implicit Bioscience, Brisbane, Australia) and SCV07 (SciClone Pharmaceuticals, San Francisco, CA).

**Therapeutic vaccines**

Because chronic HCV infection results from a failure of the host immune response to clear infection, it is logical to hypothesize that an effective immune stimulant may allow for viral clearance. As such, therapeutic vaccine strategies are being pursued. One example is GI-5005 (Globeimmune, Louisville, CO), which uses modified HCV NS3 and core protein expressed by yeast to stimulate an anti-HCV T-cell response (reviewed in Habersetzer et al\textsuperscript{151}). Phase I trials showed that GI 5005 was tolerated and reduced HCV RNA levels.\textsuperscript{152} A Phase II trial evaluated GI-5005 + PegIFN/RBV compared with SOC alone in GT1 treatment-naive patients and previous nonresponders. GI-5005 was given for a 12-week lead-in period followed by 48 weeks of GI-5005 + PegIFN/RBV. In treatment-naive patients, the GI-5005 arm had a significantly higher end-of-treatment response (74% vs 59% with SOC) and a higher ALT normalization rate (55% vs 31%).\textsuperscript{153} Previous IFN nonresponders showed equivalent treatment response regardless of GI-5005 administration (31%); however, more ALT normalization was observed in patients receiving GI-5005 when compared with those receiving SOC alone (33% vs 20%). Several other therapeutic vaccines are in Phase I development including DNA-based vaccines, T-cell vaccines, and dendritic cell immunotherapy (Table 8).
Potential HCv treatment options and future research

PKR inducers
Protein kinase R (PKR) is part of the cellular antiviral defense system. Once activated, PKR induces phosphorylation of translation initiation factor 2α, resulting in the inhibition of host protein synthesis. Nitazoxanide (NTZ) is an approved antihelminthic agent, which was noted to reduce ALT levels in HCV/HIV coinfected patients being treated for cryptosporidium. Subsequent in vitro HCV replicon studies confirmed the anti-HCV activity of NTZ. The mechanism of action is not completely understood; however, one study provides evidence that NTZ induces the PKR antiviral pathway. The phase 2 STEALTH-C1 study evaluated HCV GT4 patients in Egypt. A 12-week NTZ lead-in followed by 36 weeks of NTZ + PegIFN/RBV or NTZ + PegIFN was compared with 48 weeks of SOC, and the SVR in patients receiving NTZ + PegIFN/RBV was higher than those receiving SOC alone (79% vs 50%). Adverse events were similar across the groups, with the exception of increased anemia in patients receiving RBV. The STEALTH C-2 study is evaluating lead-in NTZ + PegIFN/RBV for GT1 cirrhotic patients who were previous nonresponders, and preliminary results indicate an EVR of 38% vs 25% in patients receiving SOC. A STEALTH C-3 (ERAIS-C) trial is also underway in which treatment-naive GT1 subjects are receiving NTZ for 2 weeks, followed by NTZ + RBV for 2 weeks and then 12 weeks of NTZ/PegIFN/RBV. Initial results show a 12% RVR in the NTZ group vs 19% in the SOC group with EVRs of 80% vs 68%.

Antifibrotic therapy
Although SVR has been associated with HCV-associated fibrosis regression, there is interest in developing therapies to directly prevent or reverse fibrosis independent of SVR, and a variety of anti-inflammatory and antifibrotic drugs are in clinical development.

Caspase inhibitors
HCV-infected livers have higher than normal numbers of apoptotic cells, which promote inflammation and fibrogenesis. GS 9450 and PF-03491390 are caspase inhibitors, which block apoptosis. Although one concern is that antiapoptotic drugs might increase the risk of malignancy, GS-9450 was not associated with adverse effects with short-term administration in healthy adults and is now being studied in HCV patients. PF-03491390 (formerly IDN-6556; Pfizer) is an irreversible caspase inhibitor. Doses from 5 to 400 mg (given daily to TID) for 14 days in 80 HCV patients

Table 8 Other HCv inhibitors under evaluation

| Drug class            | Type                           | Name               | Phase |
|-----------------------|--------------------------------|--------------------|-------|
| Immunomodulators      | Alternative interferons         | Omega IFN          | 2     |
|                       |                                | PegIFN lambda      | 1     |
|                       | TLR agonist and immunostimulatory peptides | SCV 07              | 2     |
|                       | Therapeutic vaccines           | ANA 773            | 1     |
|                       |                                | SD-101             | 1     |
|                       |                                | IMO-2125           | 1     |
|                       |                                | Oguflanide         | 1     |
|                       | PKR activator                  | Gl 5005 (Tarmogen) | 2     |
|                       |                                | IC41               | 2     |
|                       |                                | CT 1011            | 1     |
|                       |                                | ChronVac C (DNA)   | 1     |
|                       |                                | TG4040 (MVA-HCV)   | 1     |
|                       |                                | PeviPROTM          | 1     |
|                       |                                | CSL123             | 1     |
|                       |                                | MBL-HCVI           | 1     |
|                       |                                | HuMax-HepC         | 1     |
|                       | Antifibrotic agents            | Caspase inhibitors | GS 9450 | 2     |
|                       |                                | PF03491390 (IDN-6556) | 2     |
|                       | Antioxidant                    | Mitoquinone        | 2     |
|                       |                                | Silybinin          | 2     |
|                       | Matrix metalloproteinase       | CTS 1027           | 2     |
|                       | PPAR γ agonist                 | G 1262570 (Farglitazar) | 2     |

for 2 weeks, followed by NTZ + RBV for 2 weeks and then 12 weeks of NTZ/PegIFN/RBV. Initial results show a 12% RVR in the NTZ group vs 19% in the SOC group with EVRs of 80% vs 68%.

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resulted in the reduction of ALT levels in all but the lowest dosing group; however, HCV RNA levels did not change significantly.164

**Matrix metalloproeinase inhibitors**

Matrix metalloproeinases (MMPs) are proteolytic enzymes that digest the scaffolding and architecture of the liver. Phase II studies of the MMP inhibitor CTS-1027 (Conatus, San Diego, CA) are underway.

**Antioxidants**

HCV infection is associated with oxidative stress, which leads to hepatic stellate cell activation, increasing collagen production and fibrogenesis. As such, antioxidants may help reduce HCV-induced fibrosis.

- **Mitoquinone** (Antipodean Pharmaceuticals, Auckland, New Zealand) is an antioxidant that covalently binds to Co-Q10 (known as ubiquinone), causing it to accumulate more than 100-fold in the mitochondria, and thus decrease oxidative stress.165 In a Phase II study, mitoquinone led to 26.4% (40 mg dose group) and 28% (80 mg dose group) reductions in ALT level in HCV patients, with no significant safety issues reported.

- **Silibinin** is the main component of milk thistle (silymarin) and has strong antioxidant and antifibrotic properties. A Phase I trial to assess the safety and efficacy of high oral doses of silymarin in previous nonresponders showed no adverse events and no antiviral efficacy.166 However, an intravenous (IV) administration of silbinin in nonresponders showed a dose-dependant decline in HCV RNA level.167 Specifically, patients received IV silbinin for 7 days before and 7 day after the initiation of PegIFN/RBV before being switched to oral silymarin. Silbinin monotherapy of 5, 10, 15 or 20 mg/kg for 7 days led to viral load decreases ranging from 0.55 to 3.02 logs. The addition of PegIFN/RBV at day 8 resulted in a further decline in viremia (giving a peak decline of 4.85 logs in the 20 mg/kg group), but some patients had slight rebound viremia after IV silbinin treatment was discontinued despite continued SOC.

**Future research challenges**

Although the future of HCV research and drug development looks promising, there are still many research challenges to overcome.

**Robust in vitro infection with other HCV GTs**

Except for the JFH-1 GT2a clone, robust infection of other HCV clones has not been achieved in cell culture. Attempts to propagate GT1a168 and GT1b169 virus in cell culture have achieved detectable infection, but low de novo virus production still impedes the use of these systems as experimental tools. Meanwhile, although the obstacles preventing the propagation of other HCV GTs in vitro are not completely understood, the 3’ end of the JFH-1 clone can confer replication permissive-ness, and thus chimeric HCV genomes expressing the necessary parts of the JFH-1 clone recombined with the structural proteins p7 and parts of the NS2 protein from GTs 1–6 have been developed as alternative systems.170-174

**More physiological hepatocyte culture systems**

To date, robust HCV infection has only been reported in Huh7-derived human hepatoma cells.175 Huh7 cells allow for the study of the viral life cycle, but they are transformed and only minimally mimic the state of hepatocytes in vivo176,177 limiting the ability to study how HCV alters hepatocyte function and induces HCV-associated liver disease. One approach to achieve more in vivo-like cell cultures has been to alter culture conditions to coax Huh7 cells to upregulate hepatocyte-specific genes, become competent for Phase I and Phase II drug metabolism, and increase expression and specific localization of tight junction, cell adhesion, and polarity markers.178-180 Although these systems may prove useful for elucidating how HCV interacts with polarized cells or disrupts specific aspects of hepatocyte physiology, Huh7 cells by nature remain transformed. Hence, numerous laboratories are trying to develop methods for maintaining primary human hepatocytes in culture without loss of differentiation. Recent publications reporting infection of primary human hepatocytes with patient serum containing HCV GTs 1, 2, 3, and 4 may represent a step forward in the use of primary human hepatocyte in HCV research.181-183

**Development of small animal models**

Efforts to develop small animal HCV infection models have included the attempts to transmit HCV to tree shrews,184,185 marmosets/tamarins,186-188 and other primates however, only the chimpanzee has proven to be a reliable HCV animal model. Although xenorepopulation approaches in which mice are repopulated with human hepatocytes have proven to be acceptable for studying HCV infection in vivo, serious technical challenges, such as hepatocyte availability, surgical technical expertise, variability in the degree of repopulation among mice, and the fragile nature of the repopulated mice themselves, all limit the widespread use of this humanized mouse model.190-192 To study the effects of HCV proteins
in vivo, several noninfectious, nonreplicating transgenic mouse lineages expressing one or more HCV proteins have been created.\textsuperscript{193–198} Despite variations in transgene expression levels and in resulting pathology, these mice have indicated that some HCV proteins can induce disturbances in lipid metabolism and possibly contribute to the development of HCC.

To create a more authentic HCV mouse model that includes active viral replication, which may be responsible for some of the clinically observed HCV-associated pathology, one goal is the development of a mouse permissive for HCV replication and/or infection. To this end, we and others have achieved HCV replication in cultured mouse cells, with JFH-1 GT2a RNA replicons replicating efficiently after transfection into mouse cells.\textsuperscript{199,200} Additionally, Ploss et al\textsuperscript{86} determined the species-specific determinants for HCV pseudoparticle (HCVpp) entry into mouse cells to be human occludin and human CD81. Hence, advancements toward HCV infection mouse models are being made.

The future of patient care

It is expected that personalized therapy, based on more efficacious treatment options and possibly more sophisticated viral and host genetic analysis, will reshape the future of HCV patient care.

Personalized therapy

In the past, treatment strategies were based on a one-size-fits-all approach; however, simple algorithms do not take into account complex viral host interactions and patient-specific factors. As such, personalized HCV therapy is emerging as the preferred management approach, whereby the duration, dose, and type of treatment are tailored to an individual. As the field of HCV mathematical modeling continues to grow,\textsuperscript{201–206} we will undoubtedly adopt more of these modeling approaches into our SOC not only in terms of predicting patient response, but also to monitor viral response, particularly the emergence of drug-resistant mutant, which may necessitate strategic changes in treatment.

Expanded treatment options

Likewise, as STAT-C and other novel therapeutics become available, the approach to HCV treatment will change. The ideal would be to have available a variety of drugs with distinct mechanisms of action that are effective across a range of GTs with shorter treatment duration and improved tolerability. However, even a moderate increase in treatment options (e.g. SOC plus 1 OR 2 STAT-C drugs) would likely have a great impact on SVR rates. Although we are rapidly approaching the STAT-C era, it will still be some time until these therapies are available for all patients and have been optimized to control viral drug resistance.

Viral and host genetic analysis

Within the realm of individualized patient care is the potential analysis of viral and host biomarkers as tools for predicting HCV patient response to therapy. In particular, specific host genetic polymorphisms in IL28 have been identified as determinants of response to PegIFN/RBV,\textsuperscript{207,208} and it is possible that ongoing research will reveal other genetic factors that could be analyzed. Likewise, analysis of viral factors, such as sequence variation, have proven efficacious in correlating HCV quasispecies complexity and treatment outcome.\textsuperscript{209–211} As such, the analysis of both viral and host markers together might significantly improve virological response predictions and aid in clinical treatment decisions.\textsuperscript{208,212}

Conclusion and Discussion

Despite substantial progress optimizing the current HCV SOC, there are still many patients who do not successfully respond to treatment and/or remain untreated due to the severe adverse effects associated with this IFN/RBV-based therapy. Improved versions of IFN and RBV hold promise of less frequent, more convenient dosing and decreased toxicity; however, the largest improvements in treatment efficacy in the near future will likely be achieved by increasing the complexity of the anti-HCV regimen to include more compounds, particularly compounds that are directly targeted against the virus and critical virus–host interactions. With the great diversity of HCV and its ability to rapidly mutate to escape targeted therapeutics, strategies to design the proper combination of inhibitors will be critical; however, the development of the HCV experimental systems needed to elucidate the molecular mechanisms that mediate HCV infection have greatly advanced our understanding of the viral life cycle, resulting in the identification of unprecedented number of putative HCV inhibitors. With many of these HCV antiviral compounds already in clinical development, it is clear that we are about to witness a great change in HCV patient care. In the immediate future, this is expected to include viral protease and/or polymerase inhibitors in combination with PegIFN and RBV, but in the long term, new therapeutics will ideally include a choice of multiple agents that block different aspects of infection while controlling the emergence...
of viral resistance ultimately allowing for efficacy across HCV GTs and in more patient populations, with decreased duration of treatment and improved tolerability.

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Disclosure

The authors report no conflicts of interest in this work.

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