Expression of miR-146a and miR-155 in Egyptian patients with Behçet’s disease: clinical significance and relationship with disease activity

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Abstract

Background: Behçet’s disease (BD) is a systemic vasculitis disorder with multifactorial immunopathogenesis and associated with significant morbidity and mortality. MicroRNAs (miRNAs) are involved in the pathogenesis of inflammatory diseases. MiR-146 and miR-155 are known key regulators of immune response. This study was conducted to determine the expression of miRNA-146a and miRNA-155 in patients with BD and to link their possible association with the clinical manifestations and activity of this disease to evaluate their role as diagnostic or prognostic markers. A total of 60 patients with BD and 25 age- and gender-matched healthy controls were examined in a case-control study from October 2017 to September 2018 for the expression levels of miR-146a and miR-155 using singleplexTaqMan two-step stem loop quantitative reverse transcription real-time polymerase chain reaction (qRT-PCR).

Results: Patients with BD had significantly lower miR-146a levels than control subjects ($P < 0.001$). Regarding the miR-155 expression level, no statistically significant differences were detected between patients and healthy controls ($P = 0.736$). The expression level of miR-146a showed no significant association with the different clinical manifestations of patients with BD.

Conclusion: This study suggests the possibility that miR-146a expression in patients with BD is involved in the pathogenesis of disease. Furthermore, it can be used as a diagnostic biomarker and a therapeutic target for BD in the future.

Keywords: Behçet’s disease, Behçet’s Disease Current Activity Form, MicroRNA-146a, MicroRNA-155, Quantitative reverse transcription real-time polymerase chain reaction

Background

Behçet’s disease (BD) is a chronic, relapsing, inflammatory disease affecting almost all organ systems with characteristic exacerbations and remissions [1] and significant organ-threatening morbidity and mortality [2] that may result from widespread inflammatory effects on both arteries and veins of all sizes. Egypt is known to have a high incidence of BD with a higher male-to-female ratio and a higher incidence of neurological (34.9%) and vascular (57.1%) lesions than other countries [3].

The pathogenesis of BD might involve genetic susceptibility, environmental factors, as well as inflammatory response abnormalities and abnormal immune responses [4]. Typical presentations include the triad of oral ulcers, genital ulcers, and uveitis. Nevertheless, the skin, gastrointestinal tract, kidneys, lungs, heart, blood vessels, and the central nervous system are also commonly affected [5].
Due to the lack of pathognomonic signs or specific laboratory, radiologic, or histologic findings for BD [6], its diagnosis depends on the presence of a number of clinical findings defined by a set of diagnostic criteria, which are known as the International Criteria of Behçet’s Disease (ICBD), which define that seven clinical items with a total score of 4 points is diagnostic of BD [7].

MicroRNAs (miRNAs), one of the noncoding RNA families, play a significant role in mediating the posttranscriptional regulation of gene expression [8] and controlling biological events, including development, proliferation, differentiation, cell fate determination, apoptosis, signal transduction, and organ development [9, 10]. Dysregulations in miRNAs have also been implicated in a wide range of diseases, including diabetes, cancer, and cardiovascular, renal, and autoimmune disorders [11]. Among the various miRNAs playing a possible role in regulating the immune system, miR-146 (miR-146a and miR-146b) and miR-155 have emerged as key regulators of the immune response [12].

MiR-146 is a multifunctional miRNA that enhances the apoptosis of dendritic cells (DCs), suppresses the production of proinflammatory cytokines such as TNF-α, suppresses Th1 and Th17 cells, and activates T regulatory lymphocytes (Tregs) [13]. MiR-146 deficiency results in an excessive production of IL6 and TNF-α, myeloproliferative diseases, and inflammatory diseases [14]. On the other hand, miR-155 activates DCs and increases the production of inflammatory cytokines; stimulates the differentiation of Th1 and Th17 cells, proliferation of B cells, and antibody production; and is required for maturation and effector functions of natural killer and CD8 cells [12].

BD activity correlates with a proinflammatory shift in peripheral T cell populations characterized by the expansion of both Th17 and Th22 subsets and a reduction in Tregs. Dysregulation in miRNAs mediating the immune response pathways and the effector cell development may be involved in these population shifts [15]. Regarding BD, there is limited information about the expression of miRNAs, and furthermore, no high-throughput miRNA expression studies have been conducted to identify miRNAs specifically associated with this disease [16].

The objective of this study was to determine the expression levels of miRNA-146a and miRNA-155 in Egyptian patients with BD and analyze their possible association with various manifestations of this disease.

**Methods**

This study was conducted on 85 participants who were divided into two groups. The first group consisted of 60 Egyptian patients with BD diagnosed according to the ICBD [7]. There were 6 female and 54 male patients aged 23–65 years (mean 36.33 ± 8.32 years). These patients were recruited from the Rheumatology Department, Cairo University, during October 2017 to September 2018. The second group, i.e., the control group, consisted of 25 age- and sex-matched healthy subjects, including 6 females and 19 males, with an age range of 21–60 years (mean 36.2 ± 10.4 years).

All measurements were performed according to the ethical standards of Kasr-Alainy Cairo University Hospitals research committee and the ethical standards of the 1964 Declaration of Helsinki. Informed consent was obtained directly from each subject before data collection and after the explanation of the study objectives. Patients with BD suffering from severe infections, malignant tumors, or other autoimmune diseases were excluded.

All patients with BD were subjected to full history-taking and clinical examination, including disease activity assessment using the Behçet’s Disease Current Activity Form (BDCAF) [17]. The severity levels of BD were graded as mild, moderate, and severe [18]. Routine laboratory investigations consisting of ESR, qualitative CRP, complete blood count, liver function tests (ALT and AST), and kidney function tests (creatinine and urea) were performed for all patients and control subjects.

**Determination of serum levels of two mature miRNAs (miR-146a and miR-155)**

Peripheral blood samples (3 mL) were collected in serum vacutainer tubes and centrifuged at 3000 rpm for 10 min at room temperature. The supernatant was transferred to Eppendorf tubes. These samples were again centrifuged at 15,000 rpm for 10 min to precipitate the cell debris, and the resulting supernatants were stored at −80 °C until RNA extraction. RNA for miRNA expression profiling was isolated from the serum using the Qiagen® miRNeasy Serum/Plasma Kit (Catalog no. 217184) according to the manufacturer’s instructions. miR-146a and miR-155 were detected using the TaqMan® miRNA assay. U6 snRNA was used as endogenous control for normalization. Sequences of mature miRNAs and endogenous control were identified using the miRbase shown in Table 1.

RNA was reverse-transcribed into cDNA using the TaqMan® MicroRNA RT Kit (Applied Biosystems, USA

| Table 1 Target MiRNA and endogenous control sequences to be studied and amplified by RT-qPCR |
|----------------------------------------|
| **MiRNA** | **miRbase No.** | **Mature sequence** |
| U6snRNA | NR_004394 | GTGCTCGGCTTGGCAGCACATATACAAAA |
| | | TTGGAAGATACAGAGATACAGTGG |
| | | CCCCTTGGGCAAGCGATAGCAGCAATACTCG |
| | | TGAAGGCTGCTCATATTT |
| MiR-146a | MIMAT0000449 | UGAGAACUGAAUCCUUGG |
| MiR-155 | MIMAT0000646 | UUAUUGCUAAUCCUGAUAGG |
| | | UGAGAACUGAAUCCUUGG |
| | | UUAUUGCUAAUCCUGAUAGG |
Part No. 4366596), in combination with the stem loop-TaqMan® MicroRNA and control RT primer 5× (Applied Biosystems, USA, part No. 4427975), allowing the simultaneous reverse transcription of target miRNAs and control according to the manufacturer’s instructions.

Real-time polymerase chain reaction amplification and analysis
Real-time polymerase chain reaction (PCR) was performed using TaqMan microRNA assays specific to the corresponding mature sequence analyzed. Quantitative PCR (qPCR) steps were run in the StepOne real-time PCR system (Applied Biosystems) using TaqMan® MicroRNA assays 20× (Applied Biosystems, USA, part No.4427975), TaqMan® Endogenous Control Assay 20× (Applied Biosystems, USA, part No.4427975), and TaqMan® Universal Master Mix II, no UNG 1 × 1 mL (Applied Biosystems, USA, part No.4440043).

Data analysis: comparative CT method ($\Delta \Delta C_T$)
The miRNA expression level was measured using the C_T method. The expression for each miRNA is represented by the difference between its C_T value and the average C_T value of reference genes, per sample, within a given sample set [19]. We used U6 snRNA as the reference gene. The relative expression (fold change) for each candidate miRNA within each group was then calculated using the equation $2^{-\Delta \Delta C_T}$ [20]. The $\Delta C_T$ for each miRNA in each sample was calculated as follows: $\Delta C_T$sample = $C_T$targetmiRNA $- C_{TU6snRNA}$. Then, $\Delta \Delta C_T$ was calculated as follows: $\Delta \Delta C_T = (C_{TmiRNA} - C_{TU6snRNA})$ patients $- (C_{TmiRNA} - C_{TU6snRNA})$ controls.

Statistical analysis
Statistical analysis was conducted using IBM SPSS® Statistics version 22 (IBM® Corp., Armonk, NY, USA). Numerical data were expressed as mean and standard deviation or median and range as appropriate. Qualitative data were expressed as frequency and percentage. Pearson’s chi-square test or Fisher’s exact test was used to determine the relationship between qualitative variables.

The receiver-operating characteristic (ROC) curve was used to evaluate the area under the curve (AUC) and its 95% confidence interval and for predicting the cut-off values. The diagnostic accuracy of miRNAs was evaluated by calculating the sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), and total accuracy. All tests were two-tailed. A P value < 0.05 was considered as statistically significant.

Results
This study was conducted on 60 Egyptian patients with BD and 25 age- and sex-matched healthy subjects (control group).

In the patient group, there were 54 (90%) males and 6 (10%) females with a mean age of 36.3 ± 8.3 years (range 23–65 years). The control group consisted of 20 (80%) males and 5 (20%) females with a mean age of 36.2 ± 10.4 years (range 21–60 years). The demographic data of the patients and the control group are presented in Table 2.

Clinical features of patients with BD
All patients were subjected to clinical assessments. The age at onset of BD ranged from 11 to 45 years with a mean of 27.25 ± 7.61 years, and the disease duration ranged from 1 to 37 years with a mean of 9.05 ± 7.56 years. Table 3 shows the clinical characteristics of the patients with BD.

Statistical analysis revealed a significantly earlier onset of disease in patients with skin lesions (mean 24.7 ± 7.3 years) than in patients with non-skin lesions (mean 29.2 ± 7.4 years) ($P = 0.024$). There was a trend toward an earlier onset of disease in patients with ocular lesions (mean 25.9 ± 7.1 years) than in patients with non-ocular lesions (mean 29.5 ± 8.0 years) ($P = 0.074$). The proportion of vascular and neurological manifestations was greater in the late-onset patient group, but not reaching any statistical significance.

Results of BDCAF (activity score) and severity score in patients with BD
The BD activity in all patients was evaluated using the BDCAF. The results revealed eye involvement in 65% of patients, followed by CNS manifestations in 45% and joint arthralgia in 33.3% of patients. Table 4 shows the BDCAF results of patients with BD. The disease severity score was also evaluated for the patients, which revealed mild severity in 4 patients (6.7%), moderate severity in 9 patients (15%), and high severity in 46 patients (78.3%).

Table 2 Comparison between demographic features of BD patients and controls

|                  | Patients (total no. = 60) | Controls (total no. = 25) | $P$ value |
|------------------|--------------------------|---------------------------|----------|
| Age (years)      | 36.3 ± 8.3               | 36.2 ± 10.4               | 0.954    |
| Sex no. (%)      |                          |                           |          |
| Males            | 54 (90%)                 | 19 (76%)                  | 0.168    |
| Females          | 6 (10%)                  | 6 (24%)                   |          |
Association between BD severity score and clinical manifestations of patients with BD

We compared the clinical manifestations between patients with mild/moderate (n = 13) and those with severe (n = 46) BD and found no statistically significant differences, except pulmonary vascular involvement and BD severity score that demonstrated a significant association. We observed that 38.4% of patients with mild/moderate BD and 10.6% of patients with severe BD manifested pulmonary vascular involvement (P = 0.017) and arterial vascular involvement that was almost but not quite significant (P = 0.068) (Table 5).

Medical treatment of patients with BD

All the study patients were under medical treatment to control various disease manifestations and other comorbidities. The majority of them received corticosteroid therapy throughout the course of their disease illness. Almost all patients were treated with corticosteroid therapy in the form of either oral corticosteroid (prednisone) (93.3%) or pulse methylprednisolone (0.5–1 g) (41.6%).

Almost all patients with BD received immunosuppressive therapy to control various active manifestations of the disease. Biologic disease-modifying antirheumatic drugs primarily in the form of TNF-α blockers were used in 28.3% of patients, which comprised intravenous infusion of the drug infliximab (Remicade). Anticoagulant therapy was applied in 58.3% of patients. A significant association was observed between the severity of BD and pulse IV cyclophosphamide therapy (P = 0.004).

MiRNA-146a and miR-155 expression in patients with BD and control subjects

Patients with BD showed a median 0.217 fold change in miR-146a expression levels (range 0.001–7.831), whereas this value was 1.658 (range 0.099–6.361) in the control group, indicating a statistically significantly lower expression of miR-146a in the patients than in control subjects (P < 0.001).

Regarding miR-155 expression, patients with BD showed a median 0.996 fold change (range 0.037–11.791), whereas the control subjects showed a median 1.133 fold change (range 0.011–10.265), with no statistically significant difference being found between both groups (P = 0.736) (Table 6).

Correlation analysis between miR-146a and miR-155 expression in patients with BD and between miR-146a/miR-155, demographic data, and duration of BD

In patients with BD, a statistically significant positive correlation was found between miR-146a and miR-155 expression (P < 0.001, r = 0.780) (Fig. 1). However, no statistically significant correlation was found between miR-146a/miR-155 expression and the age of patients with BD, age at onset, and disease duration (Table 7).

Association between miR-146a expression and clinical manifestations and laboratory parameters in patients with BD

There was no significant association between miR-146a expression and the different clinical manifestations of patients with BD, including ocular versus non-ocular lesions, vascular versus nonvascular manifestations, and skin versus non-skin lesions (all P = 1.000) and neuro-BD versus non-neuro-BD (P = 0.793) (Table 8). There was also no significant association between miR-146a expression and different laboratory parameters.

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**Table 3** Clinical characteristics of the studied BD patients

|               | No. of pt. (total no. = 60) | %     |
|---------------|----------------------------|-------|
| Oral ulcer    | 59                         | 98.3  |
| Genital ulcer | 54                         | 90    |
| Ocular lesion | 37                         | 61.6  |
| Skin lesion   | 26                         | 43.3  |
| Erythema nodosum | 7 (of total 26) | 26.9  |
| Folliculitis  | 6 (of total 26)            | 23    |
| Papule        | 1 (of total 26)            | 3.8   |
| Papulopustular lesions | 2 (of total 26) | 7.6   |
| Pustule       | 1 (of total 26)            | 3.8   |
| Arthritis     | 8                          | 13.3  |
| Arthralgia    | 8                          | 13.3  |
| Neurological manifestations | 10 | 16.6  |
| Vascular manifestations | 31 | 51.6  |

**Table 4** Results of BDCAF in the studied BD patients

|                  | No. of pt. (total no. = 60) | %     |
|------------------|----------------------------|-------|
| Headache         | 18                         | 30    |
| Mouth ulceration | 18                         | 30    |
| Genital ulceration | 8                      | 13.3  |
| Erythema         | 6                          | 10    |
| Skin pustules    | 15                         | 25    |
| Joint arthalia   | 20                         | 33.3  |
| Joint arthritis  | 19                         | 31.6  |
| Nausea/vomiting/abdominal pain | 9 | 15    |
| Diarrhea & bleeding per rectum | 6 | 10    |
| Eye involvement  | 39                         | 65    |
| Nervous system involvement | 27 | 45    |
| Major vessel involvement | 17 | 28.3  |
| Mean ± standard deviation | Range  |        |
| Total current activity from 2006  | 2.35 ± 2.25 | 0–9   |
ROC analysis

ROC curve analysis was performed to analyze the expression of miR-146a as a potential biomarker for diagnosing BD. The expression level of miR-146a at a cut-off value of 0.650 demonstrated a sensitivity of 80%, a specificity of 64%, a PPV of 84.2%, an NPV of 57.1%, and an accuracy of 75.3% with an AUC of 0.786 (95% confidence interval 0.686–0.887) (Fig. 2).

Association between the expression of miR-146a and miR-155 and the BD severity score and BDCAF

No statistically significant difference was observed between the expression of miR-146a and miR-155 and the BD severity level. There was also no significant correlation between BDCAF and both the miRNAs (Table 9).

Discussion

The pathogenic process of autoimmune disorders, especially those affecting multi-system organs in the body, is always multifactorial and complicated. The immune dysfunctions may be caused due to several genetic or environmental factors that may intersect in the causation of the disease [21].

In the present study, the expression levels of two miRNAs in patients with BD were investigated in an attempt to disclose the possible regulatory effects exerted by these miRNAs on the molecular pathways affecting the disease pathogenesis and to understand their immunologic effects on different clinical presentations, disease activity, and severity of symptoms. Therefore, the expression levels of two selected miRNAs (miR-146a and miR-155) were examined using quantitative real-time qRT-PCR in the serum of 60 adult patients with BD, diagnosed according to the International Study Group for Behçet’s disease criteria, and 25 healthy controls. We observed that miR-146a expression was significantly lower in patients with BD than in the control group, whereas miR-155 expression showed no statistically significant difference between both groups. There was also no significant association between the expression of miR-146a or miR-155 and the BDCAF.

These results collectively suggest that a downregulated miR-146a expression is involved in the pathogenesis of BD.

MiRNAs are known to be involved in several physiological and pathological processes by targeting multiple functionally related proteins or key regulators of the immune system or the inflammatory response and by modulating the differentiation and response of various immune cells [22–25].

MiR-146a could exert its role in BD pathogenesis possibly through its anti-inflammatory role via a negative feedback regulation loop involving the downregulation of NF-κB to regulate excessive inflammation in response to proinflammatory stimuli, and its altered expression has been related to pathological conditions of the immune system [26].

Results of the present study are in general different from earlier reports. Kolahi et al. [27] showed that there was no significant difference in the miR-146a expression level between patients with BD and controls. Hou et al. [8] also reported that miR-146a did not confer susceptibility to BD. In contrast to our results, Ibrahim et al. [28] found that miRNA-146a expression was significantly higher in patients with BD than in controls.

Interestingly, previous research showed that overexpression of miR-155 could significantly inhibit intracellular IL-
17 production by CD4T cells, whereas a downregulated miR-155 expression promoted intracellular IL-17 production by CD4-T cells. Collectively, these results suggest that a decreased expression of miR-155 can lead to the production of certain proinflammatory cytokines and the downregulated expression of IL-10, an important anti-inflammatory cytokine, thus contributing to BD development [22]. Therefore, in the present study, we attempted to delineate the role of miR-155 in BD pathogenesis by measuring its expression level in patients with BD, but we did not find any statistically significant difference in its expression level between the patients and control subjects. This result is in contrast to that reported by Kolahi et al. [27], who found that miRNA-155 expression was significantly increased in patients with compared to that in healthy controls. This difference could be attributed to the fact that all the examined patients were either under corticosteroid treatment or under other immunosuppressive therapy.

We also found no statistically significant correlation between miR-146a or miR-155 expression and the gender of patients with BD. Similarly, Ibrahim et al. [28] also reported the lack of a statistically significant difference between males and females in terms of miRNA-146a expression.

In the present study, no significant correlation was detected between BDCAF and the examined miRNAs. Consistent with our result, Kolahi et al. [27] reported no significant association between the expressions of miR-155 and miR-146a and BD activity. There was also no significant association between miR-146a expression and the different clinical presentations of patients with BD.

Table 7 Correlation between miR146a/miR155 and the age of BD patients, age at disease onset, and duration of BD

|       | miR146a |       | miR155 |       |
|-------|---------|-------|--------|-------|
|       | r       | P value | r     | P value |
| Age   | − 0.149 | 0.083  | 0.070  | 0.528  |
| Age at disease onset | 0.139 | 0.298  | 0.229  | 0.079  |
| Duration of disease   | − 0.227 | 0.081  | − 0.192 | 0.142  |

We also found no statistically significant correlation between miR-146a or miR-155 expression and the gender of patients with BD. Similarly, Ibrahim et al. [28] also reported the lack of a statistically significant difference between males and females in terms of miRNA-146a expression.

In the present study, no significant correlation was detected between BDCAF and the examined miRNAs. Consistent with our result, Kolahi et al. [27] reported no significant association between the expressions of miR-155 and miR-146a and BD activity. There was also no significant association between miR-146a expression and the different clinical presentations of patients with BD.

Table 8 Association of age at disease onset with different clinical manifestations of BD

| Clinical manifestations of BD | Age at disease onset |
|------------------------------|----------------------|
| Occular lesions              | 25.9 ± 7.1           |
| Non-occular lesions          | 29.5 ± 8.0           |
| Vascular manif.              | 27.5 ± 7.8           |
| Non-vascular manif.          | 26.9 ± 7.5           |
| Skin lesions                 | 24.7 ± 7.3           |
| Non-skin lesions             | 29.2 ± 7.4           |
| Neurobehcet                  | 29.8 ± 7.3           |
| Non-neurobehcet              | 26.7 ± 7.6           |

r = 0.780 p = <0.001
However, in contrast to our results, Ibrahim et al. [28] reported a significant association between miRNA-146a expression and both eye activity and vascular activity in patients with BD. In addition, Hassouna et al. [29] reported that miRNA-155 expression was significantly decreased with the increase in the BD activity assessed by BDCAF. Furthermore, Tili et al. [30] found that miRNA-155 expression was increased with BD activity.

A statistically significant positive correlation was found between miR-146a and miR-155 expression in patients with BD.

The findings of some previous studies are either consistent or inconsistent with our study results, which may be due to different ethnicities, different clinical manifestations, and disease severity and activity and also due to environmental factors among patients in the different studies.

The different miRNA expressions may be related to the effect and duration of the immunosuppressive treatment; however, there is a limited number of previous comparative studies.

It is important to mention that there are some limitations in our study. All our patients were on regular treatment with immunosuppressive agents. Our study results demonstrated a decreased expression of miR-146a in patients with BD, and no difference was found in the miR-155 expression level compared with control subjects. However, it is not clear whether these levels were affected by therapy as it was not possible to include treatment-naive patients with BD. It is also not clear whether the effect of miR-155 expression may be exerted by other immune cells or cytokine levels or a combination of both.

**Conclusion**

The present study showed that miR-146a expression was downregulated in patients with BD in comparison with healthy controls. The downregulated miR-146a expression could be involved in the pathogenesis of BD.
through a loss of control concerning the production of proinflammatory cytokines by DCs and CD4-T cells. Therefore, miR-146a may be used as a potential biomarker for the diagnosis of BD and as a therapeutic target in the future. However, our study did not reveal any significant difference in the miR-155 expression level between patients with BD and control subjects. There was also no association between BD activity and miR-146a and miR-155 expression levels.

**Future directions**

A study with a larger sample size is recommended to confirm the utility of miR-146a in the diagnosis and follow-up of patients with BD and to evaluate its potential role as a therapeutic target. Further studies are also required to focus on miR expression and its effect on BD activity and severity level, to link the expression of more miRs to pathological conditions of the immune system, and to correlate miR expression with immunosuppressive medications.

**Abbreviations**

AUC: Area under the curve; BD: Behçet’s disease; BDCAF: Behçet’s Disease Current Activity Form; ICBD: International Criteria of Behçet’s Disease; miRNAs: MicroRNAs; NPV: Negative predictive value; PPV: Positive predictive value; qRT-PCR: Quantitative reverse transcription real-time polymerase chain reaction; ROC: Receiver-operating characteristic; Th17: T-helper 17; Th22: T-helper 22; Treg: Regulatory lymphocytes

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**Authors’ contributions**

E E has made a major role in interpretation of data and writing the manuscript. G R analyzed and interpreted the patient data regarding the rheumatological disease and the activity. N A has made substantial contributions to the conception and design of the work. E A performed the molecular acquisition, analysis, and interpretation of data. A W was a major contributor in writing and publishing the manuscript. The authors read and approved the final manuscript.

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**Availability of data and materials**

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

**Ethics approval and consent to participate**

All measures performed were in accordance with the ethical standards of Kar-AIandy Cairo University Hospitals and with the 1964 Helsinki Declaration ethical standards. Research ethics committee of Cairo University Hospitals approved the study protocol No: L230317. Written informed consent was obtained directly from each subject before data collection and after explanation of the study objectives.

**Consent for publication**

The authors agree for the publication of the article.

**Competing interests**

The authors declare that they have no competing interests.

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**References**

1. Alpsoy E (2016 Jun) Behçet’s disease: a comprehensive review with a focus on epidemiology, etiology and clinical features, and management of mucocutaneous lesions. The journal of dermatology. 43(6):620–632
2. Nar JR, Moots RJ (2017 Feb) Behçet’s disease. Clinical medicine. 17(1):71
3. El Menyawi MM, Raslan HM, Edrees A (2009 Apr) Clinical features of Behçet’s disease in Egypt. Rheumatology international. 29(6):641–646
4. Dabbagh F, Haghhighi AB, Gharemy Y (2014 Apr) Behçet’s disease: from heat shock proteins to infections. Asian biomedical. 8(2):139–155
5. Alkhachroum AM, Saeed S, Kaur J, Shams T, De Georgia MA (2016 Mar) A case of neuro-Behçet’s disease presenting with central neurogenic hyperventilation. The American journal of case reports. 17:154
6. Hatemi G, Seyahi E, Fresko I, Talarico R, Hamuryudan V (2015 Dec) Behçet’s syndrome: a critical digest of the 2014-2015 literature. Clinical and experimental rheumatology. 33(6 Suppl 94):S3–S14
7. Davatchi F, Assaad-Khallal S, Calamia KT, Crock EE, Sadeghi-Abdollahi B, Schirmer M, Tzéllos T, Zouboulis CC, Alkñahi M, Al-Dalān A, Alekberova ZS (2014 Mar) International Team for the Revision of the International Criteria for Behçet’s Disease (ITR-ICBD) Collaborative Study Group. The International Criteria for Behçet’s Disease (ICBD): a collaborative study of 27 countries on the sensitivity and specificity of the new criteria. Journal of the European academy of dermatology and venereology. 28(3):338–347
8. Hou S, Ye Z, Liao D, Bai L, Liu Y, Zhang J, Klijstara A, Yang P (2016 Jan) miR-23a, miR-146a and miR-301a confer predisposition to Vogt-Koyanagi-Harada syndrome but not to Behçet’s disease. Scientific reports. 6(1):1–9
9. Guarnieri DJ, DiLeone RJ (2008 Jan) MicroRNAs: a new class of gene regulators. Annals of medicine. 40(3):197–208
10. Bartel DP (2009 Jun) MicroRNAs: target recognition and regulatory functions. Cell. 136(2):215–233
11. Paul P, Chakraborthy A, Sarkar D, Langthasa M, Rahman M, Bari M, Singha RS, Malakar AK, Chakraborty S (2018 Mar) Interplay between miRNAs and human diseases. Journal of cellular physiology. 233(3):2007–2018
12. Testa U, Pelosi E, Castelli G, Llabaye C, miR-146 and miR-155: two key modulators of immune response and tumor development. Non-coding RNA. 2017 Sep;3(3):22.
13. Chen HF, Hu TT, Zheng XY, Li MQ, Luo MH, Yao YX, Chen Q, Yu SY (2013 Jun) Association between miR-146a rs2910164 polymorphism and autoimmune diseases susceptibility: a meta-analysis. Gene. 521(2):259–264
14. Boldin MP, Taganov KD, Rao DS, Yang L, Zhao JL, Kalwani M, Garcia-Flores Y, Luong M, Devirekkit A, Xui J, Sun G (2011 Jun) miR-146a is a significant brake on autoimmunity, myeloproliferation, and cancer in mice. The journal of experimental medicine. 208(6):1189–1201
15. Muhammad JS, Ishaq M, Ahmed K (2019 Feb) Genetics and epigenetics mechanism in the pathogenesis of Behçet’s disease. Current rheumatology reviews. 15(1):7–13
16. Puccetti A, Pelosi A, Fiore PF, Patuzzo G, Lunardi C, Dolcino M (2018 Oct) MicroRNA expression profiling in Behçet’s disease. Journal of immunology research. 2018
17. Gul FC, Nazik H, Cicek D, Demir B (2017 May) Activity criteria in Behçet’s disease. Behçet’s Disease. 97
18. Krause I, Uziel Y, Guedj D, Mukanrel M, Harel L, Molad Y, Weinberger A (1999 May) Childhood Behçet’s disease: clinical features and comparison with adult-onset disease. Rheumatology. 38(5):457–462
19. Mestdagh P, Van Vlierberghve P, De Weer A, Muth D, Westermann F, Speelman F, Vandenbossche J (2009 Jun) A novel and universal method for microRNA RT-qPCR data normalization. Genome biology. 10(6):R64
20. Schmittgen TD, Livak KJ (2008 Jun) Analyzing real-time PCR data by the comparative C T method. Nature protocols. 3(6):1101
in activated human monocyte-derived dendritic cells. Proceedings of the national academy of sciences. 106(8):2735–2740

23. Xiao C, Rajewsky K (2009 Jan) MicroRNA control in the immune system: basic principles. Cell. 136(1):26–36

24. Lin YC, Kuo MW, Yu J, Kuo HH, Lin RJ, Lo WL, Yu AL (2008 Oct) c-Myb is an evolutionary conserved miR-150 target and miR-150/c-Myb interaction is important for embryonic development. Molecular biology and evolution. 25(10):2189–2198

25. Xiao C, Calado DP, Galler G, Thai TH, Patterson HC, Wang J, Rajewsky N, Bender TP, Rajewsky K (2007 Oct) MiR-150 controls B cell differentiation by targeting the transcription factor c-Myb. Cell. 131(1):146–159

26. Tahamtan A, Teymoori-Rad M, Nakstad B, Salimi V (2018 Jun) Anti-inflammatory microRNAs and their potential for inflammatory diseases treatment. Frontiers in immunology. 9:1377

27. Kolahi S, Farajzadeh MJ, Alipour S, Abhari A, Farhadi J, Bahavarnia N, Mahdavi AM, Khabbazi A, Sakhinia E (2018 Dec) Determination of mir-155 and mir-146a expression rates and its association with expression level of TNF-α and CTLA4 genes in patients with Behcet's disease. Immunology letters. 204:55–59

28. Ibrahim W, Sakr BR, Obaya E, Ghonem H (2019 Feb) MicroRNA-146a expression and microRNA-146a rs2910164 polymorphism in Behcet's disease patients. Clinical rheumatology. 38(2):397–402

29. Hassouna SS, Tayel MY, Elkaffash DM, Abdelhady AM, Elayed EH (2018) MicroRNA155 expression in relation to BDCAF scored Behçet’s disease in an Egyptian patients’ sample. The open rheumatology journal. 12:115

30. Tili E, Michaille JJ, Cimino A, Castinean S, Dumitrut CD, Adair B, Fabbi M, Alder H, Liu CG, Calin GA, Croce CM (2007 Oct) Modulation of mir-155 and mir-125b levels following lipopolysaccharide/TNF-α stimulation and their possible roles in regulating the response to endotoxin shock. The journal of immunology. 179(8):5082–5089

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