The first complete chloroplast genome of *Coptis chinensis* var. *brevisepala*, with implication for the phylogeny of Ranunculaceae

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**ABSTRACT**

*Coptis* is one of the most important medicinal plant genera in eastern Asia. To better understand the evolution of this genus, the complete chloroplast genome of *C. chinensis* var. *brevisepala* was obtained by next-generation sequencing. The plastome of *C. chinensis* var. *brevisepala* is 155,426 bp in length, and consists of large (LSC, 84,488 bp) and small (SSC, 17,402 bp) single-copy regions, separated by pair of inverted repeat regions (IRs, 26,768 bp). It harbours 111 unique genes, including 78 protein-coding genes, 29 transfer RNA genes, and four ribosomal RNA genes. *Rps19* and *ycf1* were pseudogenized due to incomplete duplication in IR regions. The nucleotide composition is asymmetric (30.5% A, 19.4% C, 18.7% G, and 31.3% T) with an overall G + C content of 38.2%. The phylogeny of Ranunculaceae based on 75 CDGs of 27 taxa showed that Ranunculaceae is paraphyletic and thus needs redefinition.

*Coptis* (Ranunculaceae) is one of the most important and well-known plant genera in eastern Asia. Dried rhizomes of *Coptis* plants are utilized for Rhizoma Coptidis (RC), a traditional Chinese medicine famous for its functions of clearing heat, dispelling dampness, and purging fire toxins (Chinese Pharmacopoeia Commission 2010). Like the other congeneric taxa, the populations of *Coptis chinensis* Franch. var. *brevisepala* W.T. Wang et Hsiao are heavily threatened or even on the verge of extinction in its natural range (Anhui, Fujian, Guangxi, Guangdong, Jiangxi, Zhejiang provinces), due to habitat destruction and long-term over-harvesting (Zhang and Zhang 2004, 2005). To better understand the evolution of this genus, we reported and characterized the first complete cp genome of *C. chinensis* var. *brevisepala*, and then presented a phylogenomic study of Ranunculaceae.

Leaf samples were collected from China, Zhejiang Province, Hangzhou City, Lin’an District, Qingliangfeng Botanical Garden. Voucher herbarium specimen (Pan Li LP161266) was deposited at the Herbarium of Zhejiang University (HZU). We extracted total DNA from ca. 3 mg of the silica-gel dried leaf tissue using DNA Plantzol Reagent (Invitrogen, Carlsbad, CA), following the manufacturer’s protocol. Then, next-generation sequencing was conducted on the Illumina Hiseq Platform (Illumina, San Diego, CA). The complete cp genome was assembled via NOVOPlasty (Dierckxsens et al. 2017) with the cp genome sequence of *Coptis chinensis* (GenBank accession number: NC_036485) as a reference. The annotation was performed using Geneious 11.0.4 (Biomatters Ltd., Auckland, New Zealand). After all, clean reads were re-mapped to the draft genome and yielded the cp genome sequence of *C. chinensis* var. *brevisepala*. The complete chloroplast genome sequence was deposited in GenBank (MH509384).

The complete plastome of *Coptis chinensis* var. *brevisepala* is 155,426 bp in length, including two single copy regions (LSC: 84,488 bp and SSC: 17,402 bp) and two inverted repeat regions (IRs: 26,768 bp). The nucleotide composition is asymmetric (30.5% A, 19.4% C, 18.7% G, and 31.3% T) with an overall G + C content of 38.2%. It contained 111 unique genes, including 78 protein-coding genes, 29 transfer RNA genes, and four ribosomal RNA genes. Sixteen genes are duplicated in the IR regions, including all four rRNA genes, seven tRNA genes, and five protein-coding genes. Each IR region contained seven tRNA genes (tmL-CAU, tmL-CAA, trnV-GAC, trnI-CAU, trnL-CAA, trnA-UGC, trnA-ACG, trnA-GUU), *rps19* and *ycf1* were pseudogenized due to incomplete duplication in IR regions.

The phylogeny of Ranunculaceae were reconstructed based on 75 CDVs of 26 Ranunculaceae species and one outgroup taxa (*Berberis amurensis*, Berberidaceae), using both maximum likelihood (ML) and Bayesian inference (BI) methods. ML analysis was implemented in RAxML-HPC v.8.2.10 (Stamatakis 2014) on CIPRES Science Gateway V. 3.3 (Miller et al. 2010), with the GTR + G model and 1000 bootstrap replicates. BI analysis was performed in MrBayes on XSEDE v3.2.6 (Ronquist and
Helsenbeck 2003) on CIPRES Science Gateway V. 3.3, with nst = mixed, rates = gamma, ngen = 10000000, samplefreq = 5000. ML and BI analyses generated the same tree topology (Figure 1). Subfamilies Hydrastidoideae, Coptidoideae, and Thalictroideae were found to be monophyletic, while subfamily Ranunculoideae was paraphyletic, because tribe Adonideae was sister to Thalictroideae. This result was consistent with the most recent phylogenetic studies on Ranunculaceae (Cossard et al. 2016; Zhang et al. 2018). Within Coptis, C. chinensis var. brevisepala is sister to C. chinensis. Overall, our data will largely enrich the genetic information of Coptis and facilitate future studies on its conservation genetics.

**Disclosure statement**

No potential conflict of interest was reported by the authors.

**Acknowledgements**

The authors are really grateful to the opened raw genome data from public database.

**Funding**

This research was supported by the National Natural Science Foundation of China (Grant No. 31760052) and the Science & Technology Basic Resources Investigation Program of China (Grant No. 2017FY100100).

**References**

Chinese Pharmacopoeia Commission. 2010. Pharmacopoeia of the People’s Republic of China. 1st ed. Beijing: China Medical Science Press.

Cossard G, Sannier J, Sauquet H, Damerval C, Craene LR, Jabbour F, Nadot S. 2016. Subfamilial and tribal relationships of Ranunculaceae: evidence from eight molecular markers. Plant Syst Evol. 302:419–431.

Dierckxsens N, Mardulyn P, Smits G. 2017. NOVOPlasty: de novo assembly and post-analysis of large phylogenies. Bioinformatics. 30:1312–1313.

Miller MA, Pfeiffer W, Schwartz T. 2010. Creating the CIPRES science gateway for inference of large phylogenetic trees. Gateway Computing Environments Workshop (GCE); New Orleans, LA. p. 1–8.

Ronquist F, Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics. 19:1572–1574.

Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics. 30:1312–1313.

Zhang L, Zhang XP. 2004. The investigation and protection of the endangered plant of Coptis chinensis var. brevisepala in Anhui Province. J Plant Resour Environ. 13:44–48.

Zhang L, Zhang XP. 2005. Population characteristics of Coptis chinensis var. brevisepala in Anhui Province and its endangering mechanism. J Appl Ecol. 16:1394–1398.

Zhang YY, Dong JX, Feng Y, Wang ZS, Li P. 2018. The first complete chloroplast genome of Coptis quinquesecta, a critically endangered medicinal plant in China. Mitochondrial DNA B: Resour. 3:370–372.