Hemophagocytic Lymphohistiocytosis Secondary to Human Immunodeficiency Virus-Associated Histoplasmosis

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Hemophagocytic lymphohistiocytosis (HLH) in immunocompromised hosts is a fulminant syndrome of immune activation with high rates of mortality that may be triggered by infections or immunodeficiency. Rapid diagnosis and treatment of the underlying disorder is necessary to prevent progression to multiorgan failure and death. We report a case of HLH in a patient with human immunodeficiency virus, disseminated histoplasmosis, Mycobacterium avium complex, and Escherichia coli bacteremia. We discuss management of acutely ill patients with HLH and treatment of the underlying infection versus initiation of HLH-specific chemotherapy.

Keywords. histoplasmosis; HIV; HLH.

CASE REPORT

A 32-year-old Mexican man presented to a community clinic with 2 months of low-grade fevers, night sweats, odynophagia, and 30 pound weight loss. He was diagnosed with HIV with a CD4 count of 3 cells/mm³ and HIV viral load of 354 800 copies/mL. On presentation to the HIV clinic, his family reported that he had become confused during the interval time. The patient had a temperature of 38.4°C and a heart rate of 115 beats per minute, with examination remarkable for oropharyngeal plaques, cachexia, and delirium without focal neurologic deficits. Laboratory test results were notable for aspartate aminotransferase 162 µ/L, alanine aminotransferase 56 µ/L, alkaline phosphatase 112 µ/L, total bilirubin 1.6 mg/dL, white blood cell count of 3.83 × 10³/mL, hemoglobin 10.4 g/dL, platelet count 145 × 10³/µL, and a lactate dehydrogenase of 1679 µ/L. A computed tomography scan of the chest, abdomen, and pelvis was remarkable for enlarged mediastinal lymph nodes up to 21 mm × 12 mm with borderline splenomegaly and no major abdominal or retroperitoneal lymphadenopathy, a nonspecific finding commonly seen in infection, malignancy, granulomatous diseases, and a number of other diseases (Figure 1). The mediastinal lymphadenopathy in the setting of HIV prompted concern for Hodgkin’s lymphoma, disseminated fungal infections, pulmonary tuberculosis, and multicentric Castleman’s disease. The patient was directly admitted to the hospital and started on fluconazole for oropharyngeal candidiasis and cefepime for fever and hypotension. Blood cultures grew Escherichia coli, susceptible to cefepime; however, daily fevers persisted. Epstein-Barr virus (EBV) studies showed a positive EBV immunoglobulin (Ig)G and nuclear antibody, negative IgM, and an EBV viral load of 830 IU/mL. Sepsis physiology persisted with worsening of liver function tests and pancytopenia. A bone marrow biopsy was performed that demonstrated hemophagocytosis (Figure 2), and the oncology consult service recommended initiation of dexamethasone and etoposide following the HLH-2004 protocol. This was deferred due to concern for underlying infection [3]. Further results included a serum ferritin of 64 410 ng/mL (reference range, 20–230 ng/mL), serum fibrinogen of serum triglyceride level of 234 mg/dL (reference range, 0–150 mg/dL), soluble CD25 of 2823 units/mL (reference range, 0 to ≤1033 pg/mL), and natural killer (NK) cell functional assay FC/LU30 = 7 (reference range, 7–125 LU30). With fever, pancytopenia, hemophagocytosis on bone marrow, elevated CD25, low NK cell activity, elevated triglycerides, and elevated ferritin, the patient met criteria for HLH. Wright-Giemsa stain from a blood culture showed small...
clusters of yeast-like organisms (Figure 1B) concerning for underlying histoplasmosis, and liposomal amphotericin B was initiated. Patient’s fever, hypotension, and hyperferritinemia resolved within the next 1–2 days. On hospital day (HD) 7, cultures from blood, bone marrow, and bronchoalveolar lavage all grew yeast, confirmed to be *Histoplasma capsulatum*; urine *Histoplasma* antigen was >19 ng/mL. Acid-fast bacilli cultures from blood and sputum grew *Mycobacterium avium complex* (MAC), and clarithromycin and ethambutol were started on HD-12. All subsequent blood and urine cultures were negative for *Histoplasma*, MAC, and *E coli*. After 2 weeks of clinical improvement on the liposomal amphotericin B, he was transitioned to oral itraconazole 200 mg twice daily, with plans to maintain treatment until CD4 >150 and a minimum of 12 months of therapy. We initiated antiretroviral therapy (ART) 3 weeks after starting *Histoplasma* treatment, balancing the evidence favoring immediate ART initiation during opportunistic infections with the risk of immune reconstitution inflammatory syndrome provoking recurrent HLH, which can be life-threatening [4]. Tenofovir/emtricitabine and raltegravir were chosen to avoid CYP3A4 interactions with itraconazole and MAC therapy; rifamycins for MAC were also avoided for the same reasons. The patient continues to do well 15 months later at the follow-up visit.

**DISCUSSION**

Hemophagocytic lymphohistiocytosis is a life-threatening syndrome of hyperinflammation leading to multiorgan injury. Patients often present with a sepsis-like syndrome, and high rates of mortality are observed with untreated disease. Hemophagocytic lymphohistiocytosis is diagnosed by identifying known genetic mutations or fulfilling a minimum of 5 of 8 of the following criteria: fever, splenomegaly, cytopenia in 2 cell lines, hypertriglyceridemia or hypofibrinogenemia, hemophagocytosis in bone marrow, spleen or lymph nodes, low or absent NK cell activity, ferritin >500 µg/L, and soluble CD25 >2400 U/mL [3]. The pathogenesis of HLH includes defective cytotoxic regulatory function of macrophages, and NK cells leads to unregulated cytokine release and cell damage. Natural killer cells are a subset of lymphocytes responsible for elimination of activated macrophages, and low levels of NK cell activity are part of the diagnostic criteria for HLH. As a result of this dysregulated activation, excessive cytokine release occurs with elevated levels of interferon-γ, tumor necrosis factor-α, interleukin (IL)-6, IL-10, and IL-12, and soluble interleukin-2 receptor (CD25). High levels of soluble CD25 are part of the diagnostic criteria for HLH [5]. Secondary HLH is postulated to be due to activation of the immune system in response to a trigger such as malignancy, infection, or autoimmune disease [1, 3]. The HLH-2004 guidelines recommend that primary HLH be treated with an induction phase of steroids, cyclosporine A, and etoposide for 8 weeks [3]. For secondary HLH, it is controversial when immunosuppressive therapy is indicated versus treatment of the underlying infection alone [3]. In contrast, EBV-associated HLH is successfully treated with immunosuppressive agents alone [3]. Management of HLH

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**Figure 1.** Chest computed tomography scan with contrast, showing mediastinal lymphadenopathy. Highlighted is one 12 × 21 mm enlarged lymph node in the right mediastinum.

**Figure 2.** Wright-Giemsa stains (A) illustrating 2 macrophages with red blood cells (arrows) and a lymphocyte (arrowhead) in the cytoplasm, consistent with hemophagocytosis. (B) Wright-Giemsa stains illustrate a macrophage with phagocytosed yeast forms surrounded by the clear zone characteristic of *Histoplasma capsulatum*.

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in the setting of disseminated histoplasmosis is less well defined (Table 1). Of 16 patients who received amphotericin B, 13 survived and had resolution of both disseminated histoplasmosis and HLH. In 1 case, initial treatment with immunotherapy resulted in symptom relapse and initiation of amphotericin B [6]. In another case, a patient responded to immunotherapy after worsening on amphotericin B treatment alone [7]. In our patient, treatment with amphotericin B was associated with marked clinical improvement without use of immunosuppressive therapy.

CONCLUSIONS

In conclusion, a thorough investigation should be done in an immunosuppressed patient with persistent fevers and suspicion for HLH, because prompt initiation of treatment for the specific trigger may improve outcomes and limit use of cytotoxic chemotherapy.

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