DNA repair and aging: the impact of the p53 family

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Abstract: Cells are constantly exposed to endogenous and exogenous factors that threaten the integrity of their DNA. The maintenance of genome stability is of paramount importance in the prevention of both cancer and aging processes. To deal with DNA damage, cells put into operation a sophisticated and coordinated mechanism, collectively known as DNA damage response (DDR). The DDR orchestrates different cellular processes, such as DNA repair, senescence and apoptosis. Among the key factors of the DDR, the related proteins p53, p63 and p73, all belonging to the same family of transcription factors, play multiple relevant roles. Indeed, the members of this family are directly involved in the induction of cell cycle arrest that is necessary to allow the cells to repair. Alternatively, they can promote cell death in case of prolonged or irreparable DNA damage. They also take part in a more direct task by modulating the expression of core factors involved in the process of DNA repair or by directly interacting with them. In this review we will analyze the fundamental roles of the p53 family in the aging process through their multifaceted function in DDR.

The aging process

Aging can be defined as the gradual biological impairment of normal body functions accompanied by a decreased ability to respond to stress and by a greatly increased risk of morbidity and mortality [1]. This complex and multi-factorial process is characterized by a progressive failure in maintaining tissue homeostasis with a consequent direct impact on the functional ability of organs and eventually of the entire organism that causes a significant loss of fitness. The resulting deterioration represents the main risk factor for the age-related pathologies, including cancer, cardiovascular diseases and neurodegenerative disorders that ultimately lead aged organisms to death.

The full understanding of aging is far from being achieved, due to the multiplicity of mechanisms involved in this process. However, in the last decades, the ever-growing number of analytical techniques has allowed a better knowledge of the pathways underlying this process and a deeper understanding of the molecular basis of aging with the far-reaching goal of extending the human life span. Despite the existence of different but not mutually exclusive theories on aging (see [2] for an overview), it is now widely accepted that the main cause of this process is the gradual, life-long accumulation of molecular and cellular damage [1,3,4].

Recently, Lopez-Otín and co-workers have exhaustively reviewed the cellular and molecular hallmarks of aging [5], precisely defining the so-called “aging phenotype”. In particular, they classified these hallmarks into three different categories: 1- primary hallmarks, such as DNA damage, telomere loss and epigenetic alterations that initially trigger the aging process; 2- antagonistic hallmarks, as senescence or Reactive Oxygen Species...
ROS), whose effects initially protect the organism from damage, but become progressively negative throughout the process promoted by the primary hallmarks; 3-integrative hallmarks, like inflammation or stem cell exhaustion, that directly impair homeostasis when the process that leads to accumulation of damage becomes irreversible.

Genomic instability is considered one of the main drivers of the aging process [6]. Indeed, during the cell lifetime, the genomic DNA is continuously exposed to different hazards that undermine its integrity and functionality. In fact, DNA can be either challenged by exogenous environmental factors, such as oxidative stress, genotoxic drugs and ionizing radiation [7-11] or by endogenous chemicals, the most relevant of which are ROS, by-products of the normal mitochondrial metabolism [12-14] that can also cause cellular apoptosis by p53 activation [15]. Furthermore, replication errors and spontaneous hydrolysis reactions represent additional endogenous sources of DNA damage [16].

Different cellular outcomes can follow DNA injury: if DNA lesions are not properly repaired or are fixed by replication, they are converted into permanent mutations that significantly increase the risk of cancer; alternatively, DNA damage could also cause replication arrest leading to cellular senescence or cell death, thus contributing to the onset of the aging process. Therefore, cancer and aging, both arising as the consequence of irreparable DNA damage, can be considered as the two sides of the same coin [13]. Taking into account these considerations, little doubt remains about the causative role of DNA damage in tumorigenesis as well as in aging [17]. In particular, in aging the increase of DNA damage rate is the overall result of the imbalance between generation and disposal of by-products deriving from cellular metabolism [18], and the functional decline of DNA repair efficiency [19-21] (Figure 1).

Figure 1. Schematic representation of the factors and the pathways that modulate the aging process. The accumulation of DNA damage plays a pivotal role in triggering the aging process. The progressive failure of the efficiency of the DNA repair mechanisms induces a feedback loop that enforces the aging process.
Figure 2. DNA repair systems in mammals. (Left) SSBs repair mechanisms: BER: Different damage-specific glycosylases (such as OGG1 or UNG) recognize and excise the damaged base (here shown in green). The resulting AP site is target of the endonuclease APE1 that physically creates the SSB. In the short-patch sub-pathway, only one nucleotide is replaced by Polβ while the gap is sealed by XRCC1-ligase IIIa. On the other hand, during the long-patch sub-pathway, Polδ/ε synthesizes 2-8 nucleotides and FEN1 removes the 5’ flap DNA, whereas the ligation step is carried out by the complex PCNA-ligase I; NER: The initial step of lesion detection (here shown in yellow) is the only difference between GG-NER and TC-NER and it is executed by XPC/XPE or by CSB/CSA, respectively. After this step, the DNA helix is locally opened by the helicases subunits of TFIIH that allow the damage verification by XPA. The endonucleases XPF and XPG finally perform a dual incision flanking the lesion, thus releasing a 25-30 nucleotides oligomer.

(Left) DNA repair systems in mammals. (Right) DSBs repair mechanisms: HR: The ssDNA coating is target of the endonucleases XPF and XPG that perform a dual incision to create the DSBs; NHEJ: The recognition of DSBs is carried out by the Ku70/Ku86 heterodimer that activates the DNA-PK complex and synapsis; gap filling

MRN complex
Rad51/RPA nucleofilament
BRCA1/BRCA2/Rad52

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The single-strand gap is filled by the polymerases δ and ε, while the final nick is sealed by DNA ligase I and the complex XRCC1-ligase IIIα.

(Right) DSBs repair mechanisms: HR: This process is initiated by the resection of the DSB by the nuclease activity of the MRN complex, in order to generate 3’ ssDNA tails. The protruding DNA is rapidly coated by RPA protein to keep it unwound; then Rad51, supported by several other factors such as BRCA1, BRCA2 and Rad52, drives the formation of the nucleoprotein filament on the ssDNA coated with RPA. RAD51 also mediates the strand invasion and the search for homologous sequences of the nucleoprotein complex. The process ends with the resolution of Holliday junction; NHEJ: The heterodimer Ku70/Ku86 rapidly senses the presence and binds to free dsDNA ends, thus recruiting and activating the DNA-PK catalytic subunit. The kinase activity of DNA-PK is required to activate the nuclease activity of Artemis, necessary to process ends before joining. The Ligase IV and its cofactor XRCC4 finally perform the ligation step. XLF and XLS are recently established members of this pathway whose functions are still under debate.

The multi-factorial nature of aging has somehow hampered the study of this process in complex organisms in part because of the lack of genetically-modified animal models carrying multiple mutations that mimic the progressive functional deterioration typical of the aged organisms. Some age-related pathologies such as Alzheimer’s disease give some hints to understand the multifactorial nature of the aging process [22]. Furthermore, the pivotal role of the maintenance of genome stability in preventing the aging process through the activation of different DNA repair mechanisms is suggested by progeroid syndromes such as Werner syndrome, Hutchinson–Gilford progeria syndrome and Cockayne syndrome [23,24]. These well-defined diseases share, in fact, defects in some of the key components of the cellular response to DNA damage and are characterized by an early onset of aging-related features in affected individuals. Thus, progeroid syndromes provide important clues to understand the molecular mechanisms underlying human aging and, despite their “segmental” nature, represent a useful tool to improve our knowledge on still unknown aspects of the aging process. In addition, although unsuitable for representing complete in vivo models, single gene knock-outs have helped in clarifying the aging-related role of individual genes, for instance of those belonging to the p53 family [25-32]. In the future, recently acquired techniques such as CRISPR/Cas9 that allows the ablation of multiple genes in a single step [33,34], will likely have an impact in this field by providing more faithful experimental replica of aging in which the role of individual players of the process can be identified.

The DNA repair mechanisms in mammals

To cope with DNA damage, cells have evolved the DNA Damage Response (DDR), an intricate and finely regulated genomic maintenance apparatus that senses and signals the presence of lesions to finally promote their repair. The DDR includes various checkpoints, signal-transduction cascades and effectors systems that affect important cellular processes, such as replication, transcription, cell cycle progression, chromatin remodeling, differentiation or apoptosis [35-38]. Furthermore, the DDR triggers distinct repair pathways, each specific for a particular class of DNA lesion aimed at restoring the integrity of the chemical structure of DNA molecule [39]. In Figure 2 the main repair mechanisms for Single Strand Breaks (SSBs) and Double Strand Breaks (DSBs) are schematized, including their key components. Taking into account all its properties, this complex network of DNA repair systems each one dedicated to a specific type of damage, is one of the most powerful determinant of the cell’s choice among survival, replicative senescence or death [40,41].

SSBs repair

Cells repair DNA lesions affecting only one strand of the double helix through excision mechanisms, divided into Base Excision Repair (BER) and Nucleotide Excision Repair (NER) (Figure 2 left). The mechanism of action shared by these two repair systems is based on the informational redundancy of two complementary strands in the genomic DNA. Hence, these “cut and patch” repair mechanisms recognize the damaged nucleotide, remove the damaged base and, by using the opposite strand as template, fill the gap and finally ligate the pieces. Either BER or NER are complex multi-step processes involving different sets of repair enzymes. In particular, the core complex of BER, mainly involved in repairing small DNA alterations such as the oxidative damage induced by ROS, is composed of some glycosylases that leave apurinic/apyrimidinic site (AP site), the endonuclease APE1, the polymerases β, δ and ε, the DNA ligase I and the complex XRCC1-ligase IIIα [42-44]. This mechanism is not directly linked to the aging process, probably because hereditary defects in the essential BER components are embryonically lethal [45,46]; nevertheless, multiple evidences demonstrate an age-related decline in BER efficiency that is most likely associated with the accumulation of oxidative lesions with aging [47-50].
On the contrary, NER, implicated in resolving most of the helix-distorting lesions such as the bulky-chemical adducts or the UV-induced photoproducts and the ionizing radiation-induced SSB [51], is divided into two sub-pathways: the global genome NER (GG-NER) and the transcription-coupled NER (TC-NER) that differ for the set of recognition proteins utilized (i.e. XPC and XPE for GG-NER; CSB and CSA for TC-NER) [52-55]. Following the initial recognition step, the two sub-pathways share the same core complex that is basically composed by the helicases subunits of TFIH (XPB and XPD), the endonucleases ERCC1-XPF and XPG, the polymerases δ and ε, the DNA ligase I and the complex XRCC1-ligase IIIα [54,56]. NER is strictly connected with premature aging, as demonstrated by the association of deficiencies in this pathway with the human heritable disorders xeroderma pigmentosum (XP), trichothiodystrophy (TTD) and Cockayne syndrome (CS) [57]; moreover, several studies showed that NER efficiency and accuracy dramatically decline with age [58-60].

**DSBs repair**

Breaks in both strands of DNA, as those caused by ionizing radiations, are the most lethal lesions that cells have to cope with: in fact, if left unrepaired, DSBs can lead to genomic rearrangements and eventually to cell transformation or to cell death. The two main cellular mechanisms to repair DSBs are the Homologous Recombination (HR) and the Non-Homologous End Joining (NHEJ) [61-63] (Figure 2 right). The HR pathway [64] provides an accurate repair of DSBs by using the sister chromatid as template. However, this mechanism is restricted to late S and G2 phases of the cell cycle, when an extra-copy of each chromosome is available, while its use seems to be inhibited in G1 phase, likely to prevent loss of heterozygosity [65-67]. The key factor of HR is the recombinase Rad51 that form the nucleoprotein filament implied in strand invasion and branch migration. Additional proteins involved in this process comprise the MRN complex (composed of its subunits Mre11, Rad50 and Nbs1), RPA, Rad52, BRCA1 and BRCA2 and other components whose role is still under scrutiny. Recently it has been demonstrated a decline in the expression levels of some HR proteins (such as Mre11, Rad51 and BRCA1 but not BRCA2) in aged murine and human oocytes that underlines the importance of active DNA repair in maintaining ovarian reserve [68,69].

The other main mechanism of DSBs repair, NHEJ, predominates in G0/G1 cells even if it works in all phases of cell cycle, thus representing the most utilized repair mechanism in eukaryotic cells [70]. This mechanism simply joins two broken ends by using extremely limited or none sequence homology [71,72]: This often results in little deletions or insertions that render NHEJ an error-prone process contributing to accumulation of mutations and aging [73]. The core protein components of NHEJ include the kinase DNA-PKcs, the Ku70-Ku86 heterodimer [74], the nuclease Artemis, the polymerases μ and λ, the DNA ligase IV and the scaffolding proteins XRCC4, XLF [75-77] and the newly characterized PAXX4/XLS/c9orf142 [78-80]. Depletion of PAXX4/XLS/c9orf142 in cells impaired DSB repair consistent with a defect in NHEJ [78]. During aging NHEJ becomes more error-prone and less efficient, as demonstrated in human senescent fibroblasts [81]; this decline is likely correlated with the altered regulation and the reduced availability of Ku proteins [82]. Furthermore, deficiency for other components of NHEJ pathway (such as XRCC4 and Ligase IV) leads to premature senescence and to a high degree of apoptosis; of note, these phenotypes are relieved in a p53+/- background [83-86].

Finally, recent evidences revealed the existence of a third, less well-characterized DSBs repair pathway: the so-called microhomology-mediated end-joining (MMEJ) [87,88]. This mechanism can be considered as a sort of a hybrid between HR and NHEJ, as it joins broken ends with microhomology sequences of 5-25 base pairs in a Ku-independent manner. Even if it is an error-prone process, it could represent a backup mechanism that acts when other repair pathways fail. The exact number of the components of each repair pathway is not known yet. Broadly, proteins that participate in the repair mechanisms can be subdivided in core proteins (i.e. those that are necessary to the execution of the process such as glycosylases, polymerases, ligases, etc.) and accessory proteins (i.e. those that facilitate and optimize the process but, whose presence is not indispensable [88]). Examples of accessory proteins are NR4A nuclear orphan receptors that interact with the DNA-PK catalytic subunit and, upon exposure to DNA damage, translocate to DSB foci by a mechanism dependent on the activity of poly(ADP-ribose) polymerase-1 (PARP-1) [89,90], APLF that functions together with PARP-3 to accelerate NHEJ [91] and Nucleolin which mediates nucleosome disruption critical for DNA double strand breaks repair [92].

A role apart among the accessory proteins is played by transcription factors that control the expression of essential components of the repair pathways. In their quality of transcriptional regulators, these proteins are not directly involved in the repair mechanisms but, they can greatly affect their execution by determining the
availability of core components of the repair machinery [93]. In this regard, our laboratory has recently demonstrated the ability of the zinc finger factor ZNF281 in promoting the expression of XRCC2 and XRCC4 two proteins involved in HR and NHEJ, respectively [94].

Of note, recent work highlighted the presence of many transcription factors at the site of damaged DNA [95]. Interestingly, most of them were PARP-dependent for localization to sites of DNA damage. These latter findings expand the role of transcription factors in DNA repair since one of the most likely function of transcription factors in the proximity of damaged DNA is the recruitment of chromatin remodeling complexes, which can modulate the accessibility of core components of the DNA repair machinery.

In the third part of this review we will discuss the role that the transcription factors of the p53 family have in DNA repair and how this could be linked to the process of aging.

**The p53 family**

The p53 family is composed of a group of transcription factors, the well-characterized tumor suppressor p53, and its homologs p63 and p73 whose overall protein architecture is highly conserved from *Drosophila melanogaster* to humans [96]. The common gene framework of the p53 family is constituted by three main domains: an N-terminal transactivation domain (TAD), a central DNA-binding domain (DBD) and an oligomerization domain (OD). The sterile α-motif domain (SAM) involved in protein-protein interactions is distinctive of p63 and p73 and the C-terminal transcription inhibitory domain (TID) is only present in p63 [96]. Moreover, in p63 and p73 the usage of an internal promoter leads to the expression of two different types of proteins: the TA-isoforms, containing the whole N-terminal transactivation domain, and the truncated ∆N-isoforms, lacking almost all this domain. The ∆N-isoforms bound to their target promoters can behave as dominant-negative varieties of the TA-isoforms but, ∆Np63 can also affect the transcription of its target genes through the usage of alternative transcription activation domains [97]. It should also be mentioned that p53 is transcribed by an alternative p53 promoter in intron 4 and encodes N-terminal truncated forms whose function is to antagonize p53-dependent apoptosis or to inhibit cell replicative senescence [98,99]. In addition, alternative splicing mechanisms generate a great number of protein variants containing different C- and N-termini [96, 97, 100, 101] (Figure 3).

The high sequence homology among p53 family members allows binding and transactivation of the same promoters, suggesting a partial redundancy of their effect [96]. Nevertheless, recent data demonstrated that p53 SNP variants possess distinct transcriptional and DNA-binding properties [102]. The transactivation potential of p63 and p73 proteins also depends on their oligomeric state. Indeed, TAp63α forms an inactive, dimeric and compact conformation in resting oocytes, while the detection of DNA damage leads to the formation of an active, tetrameric and open conformation [103]. In contrast to p63, TAp73α is a constitutive open tetramer [103]. Each member of the family can act on unique targets, strongly suggesting that these proteins play not completely overlapping roles in the cell. The latter statement is further confirmed by the knock-out mice models. In particular, p53−/− mice develop normally but have a very high predisposition to tumor transformation [104]; indeed, p53 somatic mutations can be found in almost two thirds of human cancers [105,106]. Skin-specific p53-null background resulted in accelerated formation of spontaneous tumors and enhanced metastasis [107]. On the other hand, p63−/− mice show limb truncations, craniofacial abnormalities and die early after birth as a consequence of dehydration: these mice fail in fact to develop skin and other epithelial tissues, demonstrating the pivotal role played by p63 in epidermal morphogenesis [108-110]. In addition, loss of p63 can affect reprogramming via several mechanisms such as reduced expression of mesenchymal-epithelial transition and pluripotency genes [111]. Interestingly, p63 heterozygous mice have a shortened life-span and show features of accelerating aging [29]. As for the specific role of p63 in apoptosis, recent data highlighted the contribution of p63 pathway in damaged oocyte elimination in adulthood [112]. Finally, p73−/− mice are viable but show severe neurological defects, including hydrocephalus, hippocampal dysgenesis, as well as abnormalities in pheromone sensory pathways and chronic inflammations [113,114]. Although p73 is dispensable for commitment to neural stem cell fate, it has been demonstrated to be essential for neural stem cell maintenance and for blocking premature differentiation [115-118]. Another unique role of TAp73 is in macrophage-mediated innate immunity and in the resolution of inflammatory responses [119]. In contrast to p53, both p63 and p73 are rarely mutated in cancers but they frequently display an altered ratio between TA- and ∆N-isoforms [120-122]. In the next paragraphs we will analyze the roles that p53 family plays in the cell and the connection between p53 family, DNA damage and aging [123].
Upon DNA damage induction, p53 family members are stabilized through different mechanisms. Indeed, post-translational modifications play a key role in modulating the activation, stabilization or degradation of these proteins [124]. For example, in unstressed conditions p53 is maintained at low levels by the

Figure 3. Structural motifs of p53 family members. Gene and protein structures of p53 (top), p63 (middle) and p73 (bottom). Colors indicate protein domains encoded by the exons. The different transcription start sites P1 and P2 (indicated by blue arrows) give rise, respectively, to the TA and ΔN isoforms in both p63 and p73. In p53 gene the existence of an alternative translation start site (shown in red) generates the Δ40 isoform, while the Δ133 is transcribed by the P2 promoter. The multiple alternative splicing events leading to the different protein isoforms are represented by black dotted lines. Black solid lines represent splicing events leading to the formation of α proteins, the longest isoforms of each family member. Splicing of consecutive exons is omitted for simplicity. TAD, transactivation domain; DBD, DNA-binding domain; OD, oligomerization domain; SAM, sterile α-motif domain; TID, transcription inhibitory domain.

p53 family in DNA damage

Upon DNA damage induction, p53 family members are stabilized through different mechanisms. Indeed, post-translational modifications play a key role in modulating the activation, stabilization or degradation of these proteins [124]. For example, in unstressed conditions p53 is maintained at low levels by the
MDM2-mediated degradation, while this regulatory mechanism is released following p53 phosphorylation by the master kinases of the DDR, such as ATM, ATR, DNA-PK and Chk2 [125-128]. In addition to prevent its degradation, these and other post-translational modifications (such as ubiquitination, acetylation or methylation) serve also to increase the transcriptional activity of p53 protein [129,130]. Finally, it should be mentioned that in its function of transcriptional regulator, p53 is directly involved in promoting the transcription of genes of the DNA damage response such as MSH2 [131,132] and FANCC [133]. With regard to p63, the DNA damage-induced phosphorylation could exert opposite roles, depending on the p63 isoforms involved. In particular, c-Abl-mediated phosphorylation can promote TAp63α switch from the inactive dimer to the active tetramer in mammalian oocytes, to protect the female germ line during meiotic arrest [134-136]. On the contrary, ΔNp63α is rapidly phosphorylated by ATM and other kinases in head and neck squamous cell carcinomas (HNSCC) upon different treatments: this modification primes ΔNp63α for degradation to promote apoptosis of the damaged cells [137]. Likewise, p73 is a target of different protein modifiers, such as the kinases c-Abl, Chk1 and Chk2 or the acetyltransferase p300 that, after DNA damage induction, activate p73 protein [138-140]. Similarly to p63, the ΔN-isoform of p73 has to be degraded to allow apoptosis induction [139,141]. Of note, it has also been demonstrated that TAp73 knockout mice show an increase of DNA damage rate in spermatogonia that finally leads to male infertility. Thus, p73 has been proposed as the guardian of male germ line [142].

**p53 family in the control of cellular senescence**

Consistent with their accumulation after genotoxic stress, p53-family proteins are functionally involved in the induction of different DNA damage responses that determine cellular fate. In particular, they can promote transient cell cycle arrest to allow proper DNA repair and to prevent potentially tumorigenic cells from dividing. However, in case of continuous or irreparable DNA damage, the exit from cell cycle becomes permanent, leading to the process known as cellular senescence. Cells that have undergone senescence appear enlarged and flattened, become refractory to mitogens or growth factors and show dramatic changes in gene expression [143] and chromatin structure [144, 145]. Interestingly, progressive accumulation of senescent cells has been associated to aging in vivo [146]. Mechanistically, senescence is driven by two main interconnected still independent pathways that halt cell proliferation: the p53-p21 and the p16-Rb. Both p21 and p16 are cyclin-dependent kinase inhibitors (CDK1) that, among other activities, prevent phosphorylation, hence inactivation, of Rb [144, 145]. The overall effect is the suppression of E2F activity, a transcription factor that promotes the expression of genes required for cell cycle-progression [147, 148]. With reference to the involvement of p53 family members in this program, the direct induction of p21 promoter by p53 [149], TAp63 [150] or TAp73 [151] isoforms in response to genotoxic stress has been demonstrated. ROS production is well known to provoke senescence and genes that are induced by p53 may also play a relevant role in senescence induction [152]. Nevertheless, p53 also promotes the expression of several antioxidant genes which account for p53 ability to reduce the oxidative stress [153]. Likely, p53’s role in increasing and decreasing oxidative stress contributes to its dual effect on senescence [154]. The role of p53 family in ROS control is further highlighted by the recent finding that TAp63 controls the expression of GLS2 which, in turn, is important in the cellular antioxidant pathway [155]. As for p73, studies on TAp73-null mice demonstrated that TAp73 protects against aging by regulating mitochondrial activity and preventing ROS accumulation [156].

An additional layer of complexity in the involvement of p53 family members in the regulation of senescence was added by experimental evidence demonstrating that ΔNp63α inhibits the senescence-inducing miR-138, -181a, -181b, and -130b expression by binding directly to p63-responsive elements located in close proximity to the genomic loci of these miRNAs in primary keratinocytes [157]. Thus, suppression of miR-138, -181a, -181b, and -130b expression by ΔNp63α can be regarded as a mechanism to fine-tune the balance between cellular proliferation and senescence in epidermal proliferating cells. In addition, TAp73 induces the activation of anabolic metabolism, with enhanced pentose phosphate shunt (PPP) and nucleotide biosynthesis [158]. The metabolic effect of p73 can be interpreted as a way to counteract cellular senescence rather than to support proliferation. Furthermore, p53, p63 and p73 can work together to regulate the balance between survival, cell death, and senescence [159]. In conclusion, the pro- and anti-senescent roles of the members of p53 family depend on the presence of other regulators and more in general on the cellular context.

**p53 family in apoptosis**

The most investigated biological function of p53 and its family members is doubtlessly apoptosis control [160]. Although the involvement of p53 family in apoptosis is not the focus of this review, it is worth to remind that
p53 is the main hub in the control of cellular fate after damage. Nevertheless, in some contexts the pro-apoptotic function of p53 depends on the presence of other transcription factors such as c-Myc that is necessary for p53-induced apoptosis in response to DNA damage in vivo [161]. Among the many ways in which p53 can trigger apoptosis, one of the most relevant is through the activation of PUMA (p53-upregulated modulator of apoptosis), a BH3-only protein that induces apoptosis through the mitochondrial pathway [162-164]. In addition, p53 can induce apoptosis by promoting transcription of other pro-apoptotic proteins [165] in association with other regulators. For instance, NOXA is transcriptionally induced by binding of p53 in complex with p18/Hamlet to its promoter [166]. Intriguingly, not all p53 targets are downstream executors of apoptosis. Slug (SNAI2), a central regulator of the epithelial mesenchymal transition (EMT) process that is important for tumor metastasis, is a transcriptional target of p53 but, in turn, it is a repressor of PUMA thus preventing the pro-apoptotic activity of the latter gene [167]. Finally, it should be mentioned that p53-independent apoptotic pathways exist in normal and cancer cells [168]. The role of p63 and p73 in apoptosis was, at least at the beginning, largely interpreted as a functional overlapping of the pro-apoptotic function of p53. Nevertheless, there are cellular contexts in which the activity of p63 and p73 is required for p53-dependent apoptosis. Thus, p63 and p73 are required for p53-induced apoptosis after DNA damage. In fact, it has been demonstrated that the combined loss of p63 and p73 results in the failure of cells containing functional p53 to undergo apoptosis in response to DNA damage [169]. In addition, the distinct role of the p53 paralogs is evident in the case of male gonocytes where p53 is not expressed. The lack of p63 in gonocytes sharply decreases the rate of physiological apoptosis causing abnormal morphology of the germ cells [170]. Enhanced resistance to chemotherapy has been correlated with high levels of ΔNp73 [171]. Recent data demonstrated that acetylpolymamine oxidase (PAOX) upregulates ΔNp73 levels by suppressing its degradation [171]. The deficiency of the TAp73 isoform results in male infertility because of severe impairment of spermatogenesis. Accordingly, mice lacking TAp73 exhibited increased DNA damage and cell death in spermatogonia, disorganized apical ectoplasmic specialization, malformed spermatids, and marked hyperspermaria [142]. Furthermore, the pro-apoptotic activity of p53 paralogs can be regulated by cross-talk with other factors. p73 induces apoptosis in T cells by promoting the transcription of Bim. In activated T cells NF-κB induces Mdm2 that in turn, forms a complex with p73 thus inhibiting p73-dependent activation of Bim and the resulting apoptosis [172]. Noteworthy, although apoptosis is the most common form of cell demise induced by p53 family members, there are cases in which p53 promotes other types of cellular death [173].

**p53 family in the regulation of the aging process**

A less obvious, nonetheless relevant function of the p53 family members is in cellular aging through their activity on DNA repair mechanisms. p53, as well as its paralogs, can influence the aging process by regulating core DNA repair proteins expression, by directly interacting with repair factors or by inducing apoptosis in response to DNA damage. A paradigmatic example of the link p53-DNA repair-aging is the role of p53 in the progeroid Cockayne syndrome [174]. Complementation group B (CSB) protein is an ATP-dependent chromatin remodeler with an essential function in transcription-coupled DNA repair. Mutations in the CSB gene are frequent in Cockayne syndrome. Normally, p53 tumor protein interacts with CSB, and the chromatin association of CSB and p53 is inversely related. CSB facilitates the sequence-independent association of p53 with chromatin when p53 concentrations are low and p53 prevents CSB from binding to nucleosomes when p53 concentrations are elevated. These results suggest that the reciprocal regulation of chromatin access by CSB and p53 could be part of a mechanism by which these two proteins coordinate their activities to regulate DNA repair, cell survival and aging.

Genotoxic stresses such as UV irradiation and exposure to environmental pollutants cause damage to cellular DNA, a powerful driver of senescence and aging. Besides acting directly in the control of genes involved in the UV-induced DNA damage response, p53 promotes the transcription of genes that block the cell cycle progression (i.e. p21) and regulates the expression of different cyclin-dependent kinase inhibitors (CDKIs) thus contributing to the proliferative stop necessary to allow the execution of DNA repair [175]. In this context, the activity of p53 can be regarded as a shielding mechanism to maintain the genome stability necessary to prevent the aging process [175].

The mechanisms that drive neuronal impairment in the central nervous system (CNS) are of paramount importance to understand the functional decline that occurs in aging brains. In an animal model that utilized hydroxyurea (HU) treatment, the phenotypic features of cellular senescence [176] such as senescence-associated-β-galactosidase (SA-β-gal) staining, decreased proliferation and differentiation capacity,
increased G0/G1 cell cycle arrest, elevated ROS level and diminished apoptosis, were accompanied by a marked increase of p53 expression, as well as a decreased expression of key proteins in various DNA repair pathways such as XRCC2, XRCC3 and Ku70 [177]. Here, p53 activity is necessary for G1/S cell cycle arrest and to mitigate the devastating consequences induced by DNA damage and allow time to repair. Nevertheless, p53-mediated G1 arrest can alternatively coax cells to enter a senescence state that precedes aging. In this system, p53 acts as a modulator of the biological response by directing cells to DNA repair or towards aging depending on the extent of the damage.

As for the p53 paralogs, it is well known that they have distinctive functions in the DNA damage response. In fact, TAp73 and TAp63 (but not p53) promote the transcription of BRCA1, RAD50 and MRE11 which are involved in HR [93]. Nevertheless, their link to the aging process has just begun to be investigated in detail. The first experimental evidence of a role of TAp73 in aging comes from an in vivo study in which it has been demonstrated that TAp73-null mice show more pronounced aging with increased oxidative damage and senescence due to metabolic dysregulation [156]. ∆Np73 localizes directly to the site of DNA damage, can interact with the DNA damage sensor protein 53BP1, and inhibits ATM activation and subsequent p53 phosphorylation [178]. This finding is particularly interesting in light of the recent discovery of a considerable number of transcription factors localized in the vicinity of the sites of DNA damage [95]. In addition, E1A- and RASV12-transformed ∆Np73−/− MEFs display a significant delay in tumor initiation, as well as a decrease in tumor size compared with wild-type controls [179]. Intriguingly, the reduced tumorigenic ability of ∆Np73−/− MEFs to form tumors was not due to a higher level of apoptosis, but instead to enhanced cellular senescence, as demonstrated by increased expression of the senescence markers p16Ink4a, senescence-associated β-galactosidase (SA-β-gal), and DcR2 [179]. Since ∆Np73−/− mice show signs of neurodegeneration in their brain [178], it is feasible that the lack of ∆Np73 contributes to a process of senescence (and subsequent aging) in neuronal tissues. In line with an anti-aging role of p73, it has been demonstrated that TAp73 is a major regulator of autophagy by promoting the expression of ATG5 thus preventing aging by maintaining a homeostatic control [180].

![Figure 4. The role of p53 family members in the prevention of aging.](image)

Figure 4. The role of p53 family members in the prevention of aging. p53 family proteins are involved in different mechanisms after the induction of a DNA damage. A transient block of cell cycle progression gives time to the cells to repair the DNA. Nevertheless, p53 family proteins can also directly affect the DNA repair mechanisms (see text for details). However, if the damage is too extensive or cannot be repaired, p53 family members could trigger either a permanent exit from the cell cycle (senescence) or a cell death program (apoptosis) as mechanisms of defense. The overall result can be read as a check of the genomic stability that protects cells from aging.
The development of inducible TAp63-KO mice has contributed to the understanding of the role of p63 in senescence and aging. In the epithelial compartment, the lack of p63 accelerates aging as indicated by the accumulation of senescence markers [29]. Furthermore, TAp63 isoform specific KO highlighted skin ulceration, premature aging and reduced lifespan that are associated with genomic instability [181, 182] which may be due to a still incompletely defined role of p63 in DNA repair processes. Although the above reported results highlighted an anti-aging function of p63, it should be mentioned that TAp63 mediates the induction of oncogene induced senescence (OIS) in keratinocytes [183]. Thus it is likely that similarly to p53, p63 can promote senescence upon oncogenic stress but, in normal conditions, its absence accelerates aging. A complete understanding of the role of p63 in the aging process is furthermore complicated by the existence of N-terminal truncated isoforms (ΔNp63) [184] that can have distinct functions.

**Concluding remarks**

Genomic instability is tightly linked to the aging process. As the integrity of DNA is continuously challenged by endogenous and exogenous factors, the mechanisms that safeguard genome maintenance can be considered to all intents and purposes as anti-aging processes. Among these, the different and specific repair mechanisms that cells have developed to restore the chemical structure of the DNA molecule should be unquestionably included. However, in addition to the core proteins, that are instrumental for the execution of the repair processes, there is an ever growing number of accessory proteins involved in these mechanisms that indirectly help the DNA repair. In particular, transcription factors can support DNA repair by regulating the expression of key factors of the DNA repair machinery. Nevertheless, they can also recruit different chromatin remodeling complexes to the DNA damage sites, thus mediating the accessibility of the DNA that has to be repaired. Furthermore, they are also usually involved in the regulation of the other processes strictly linked to the repair mechanisms that occur during the DNA damage response. In this context, the p53 family plays multiple important roles in modulating the aging process (Figure 4). Following a DNA damage induction, the members of this family are induced or stabilized, usually through post-translational modifications of the proteins. In their active form, they can exert partially overlapped roles in regulating processes that determine the fate of the damaged cells. Depending on the extent of the damage, their role can vary from (i) the induction of a transient cell cycle arrest to allow DNA repair, to (ii) the induction of a permanent blockade of proliferation resulting in senescence, to (iii) the stimulation of apoptosis to eliminate damaged cells. In their quality of transcriptional regulators p53, p63 and p73 can also influence the efficiency of repair mechanisms by regulating the expression of key factors or by directly interacting with them as we saw in the previous paragraphs. In light of these evidences, there is no doubt that the p53 family plays a central role in maintaining the genome stability. As a direct consequence of this, we can conclude that their function strongly impacts on the complex regulation of the aging process.

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**Conflict of interest statement**

The authors declare no conflict of interest.

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The image contains a page of text with multiple references to scientific articles and research. The text includes terms such as "p53", "DNA", "apoptotic", "transcriptional", and "cancer". The page appears to be discussing various aspects of p53, its variants, and its role in cancer and apoptosis. The references are numbered from 91 to 108, indicating a comprehensive review or discussion on the topic. The text seems to be part of a scientific or academic discussion, possibly from a journal or a research paper.
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