Building and identifying highly active oxygenated groups in carbon materials for oxygen reduction to H$_2$O$_2$

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The one-step electrochemical synthesis of H$_2$O$_2$ is an on-site method that reduces dependence on the energy-intensive anthraquinone process. Oxidized carbon materials have proven to be promising catalysts due to their low cost and facile synthetic procedures. However, the nature of the active sites is still controversial, and direct experimental evidence is presently lacking. Here, we activate a carbon material with dangling edge sites and then decorate them with targeted functional groups. We show that quinone-enriched samples exhibit high selectivity and activity with a H$_2$O$_2$ yield ratio of up to 97.8% at 0.75 V vs. RHE. Using density functional theory calculations, we identify the activity trends of different possible quinone functional groups in the edge and basal plane of the carbon nanostructure and determine the most active motif. Our findings provide guidelines for designing carbon-based catalysts, which have simultaneous high selectivity and activity for H$_2$O$_2$ synthesis.
Hydrogen peroxide is widely used as a green oxidant in disinfectants, bleaching agents, sanitizing agents, chemical synthesis, and even as a potential energy carrier.\textsuperscript{1-4} According to a report by Global Industry Analysts, Inc.\textsuperscript{5}, the global consumption of H₂O₂ is projected to reach 6.0 million metric tons by 2024. Approximately 99% of all H₂O₂ is currently produced using the multi-step anthraquinone process. However, this process is energy intensive, and can only be performed in a centralized factory.\textsuperscript{1,5,10} The inherent complexity of the anthraquinone process has driven many researchers to investigate one-step methods, which can continuously produce H₂O₂ on-site at small scale using a simple device.\textsuperscript{15} Among these, the electrochemical synthesis approach is one of the best choices to meet the above demands.

Metal-free oxidized carbon materials have received particular attention for this application because they offer a simple synthetic procedure, cost-effectiveness, as well as high activity and selectivity.\textsuperscript{18-29} They allow the incorporation of a variety of oxygen functional groups, with the potential to widely tune performance, and to optimize active sites. For example, Lu et al. recently used nitric acid to oxidize carbon nanotubes, and demonstrated that the resulting oxidized carbon material was highly active for oxygen reduction to hydrogen peroxide (ORHP)\textsuperscript{16}. They considered the active sites to be the carbon atoms adjacent to carboxylic acid and etheric groups (–COOH and –O–C). Kim et al. used few-layer mildly reduced graphene oxide electrodes, by partially removing oxygen from graphene oxide using hydrothermal heating without a reducing agent, and showed that SP²-hybridized carbon near ring ethers along the sheet edges were the most active sites for ORHP\textsuperscript{17}.

All of these pioneering studies apply harsh oxidation conditions to cleave the strong SP² C–C bonds\textsuperscript{16,30} and incorporate oxygen functional groups, and this in turn makes it difficult to functionalize the carbon materials with targeted groups. As a result, the oxygen functional groups inevitably saturate the dangling bonds on the edges with sophisticated multi-components, which hinders systematic experimental study. It becomes inherently difficult to accurately identify the most active functional groups, because the resolution of ordinary characterization methods is not high enough to distinguish between similar groups.

From the industrial anthraquinone process, we know that quinones are the champion catalytically active oxygen functional groups towards H₂O₂ synthesis\textsuperscript{10,31}. Herein we demonstrate a facile synthesis method to incorporate quinone functional groups in carbon nanostructures. For comparison, we also employ a pre-activated method to build carbonyl-enriched and etheric ring-enriched graphitic nanoplatelets (denoted as GNP\textsubscript{C–O}) and GNP\textsubscript{C–O,C} respectively. Using fine characterization methods, such as soft XANES, XPS, FTIR, and CV, we show that each of our samples have targeted and desired functional groups (etheric ring, carboxyl and quinone). Electrochemical measurements reveal that the sample with abundant quinone functional group (GNP\textsubscript{C–O,1}) exhibits high selectivity, with a H₂O₂ yield ratio of 97.8% at 0.75 V, which is superior to previously reported etheric ring and carboxylic acid groups.

We further demonstrate the activity of the quinone functional groups by examining standalone quinone molecular systems. Finally, we use density functional theory (DFT) calculations to model different possible quinone functional groups and examine their activity toward ORHP. We find that the quinone groups exhibit very high activity with negligible overpotential.

The free edge sites were first created by simultaneously crushing and exfoliating graphite mechanochemically. Then, the reactive edge sites were saturated with mild oxidants of CO₂ or diluted O₂. We found that when CO₂ was applied as an oxidant, carbonyl-related groups were easily formed. In this case, the as-prepared product is designated GNP\textsubscript{C–O}. Even dilute O₂ could also be selected as an oxidant. Because the electronic ground state of O₂ and carbon are different, the triplet O₂ cannot react with singlet defect-free graphitic carbon spontaneously, due to the energy barrier present. This allows selective oxidation of the reactive edge sites with a high degree of controllability. Etheric ring (–C–O–C)-functionalized graphitic nanoplatelets (GNP\textsubscript{C–O,C}) were obtained in dilute O₂ atmosphere.

We first characterized the morphology of the above samples using field emission scanning electron microscopy (FESEM, Supplementary Fig. 1) and transmission electron microscopy (TEM, Supplementary Fig. 2). This analysis showed that GNP\textsubscript{C–O} and GNP\textsubscript{C–O,C} had morphologies typical of graphitic nanoplatelets and nanoparticles, respectively. The successful nanosizing and functionalization were further verified by Raman spectra and X-ray powder diffraction (XRD) patterns. As shown in Supplementary Fig. 3, the strong D band (usually as an indicator of defective edges) and broad (002) facets (derived according to the Scherrer equation), together indicate the existence of plenty of edge sites in the nanosized GNP. The Brunauer–Emmett–Teller (BET, Supplementary Fig. 4) analysis determined that the specific surface areas of GNP\textsubscript{C–O,0}, GNP\textsubscript{C–O,2}, and GNP\textsubscript{C–O,C} are 450, 753, and 657 m² g⁻¹, respectively.

Next, we measured sample oxygen content using three different methods, elemental analysis (EA), energy dispersive spectroscopy (EDS), and XPS. The results (summarized in Supplementary Table 1 and Figs. 5–7) agreed with each other well. Here, we highlight the EA results because it has higher accuracy than the other two methods. The amount of oxygen content was calculated to be 10.5 at% for GNP\textsubscript{C–O,1}, 20.6 at% for GNP\textsubscript{C–O,2}, and 10.0 at% for GNP\textsubscript{C–O,C}. A possible iron remnant was checked by time-of-flight secondary ion mass spectrometry (TOF-SIMS), which has a detection limit as low as ppm. The results (Supplementary Fig. 6) showed that there was no detectable iron in the samples.

Finally, to carefully unravel the nature of the oxygen functional groups we used a combination of techniques, including soft XANES, XPS, FTIR, and CV. Among them, soft XANES is one of the most powerful tools for characterizing graphitic materials, and can provide important information about bonding configurations with high resolution.

As shown in Fig. 1a, the C K-edge includes unoccupied π* (peak A–C) and excited σ* (peak D, E) states. Here, we only discuss the fingerprint region of π* in detail. The peak A around 284.5 eV is assigned to 1s – π* from sp² C–C\textsuperscript{32-36} . The peak B results from the charge transfer induced by the O in the etheric ring. The GNP\textsubscript{C–O,C} and partially reduced graphene oxide (pRGO, Supplementary Methods) exhibit a minor shoulder at 287.2 eV (Peak B₁), which corresponds to the out-of-plane etheric ring (C–O–C)\textsuperscript{32,33,35}. Since the in-plane C–O bonding of C–O–C is much stronger than the out-of-plane, the peak at 288.2 eV is supposed to be in-plane C–O–C at the edge of the GNP. Thus, the GNP\textsubscript{C–O,C} is mainly composed of in-plane C–O–C, and partly by out-of-plane C–O–C.

In pRGO there is only out-of-plane C–O–C. The peak C around 288.6 eV is attributed to π*(C=O)\textsuperscript{32,35}, which should be determined in combination with other characterizations.

The soft O K-edge XANES (Fig. 2b) provides further information about the groups. Here, the pRGO shows an exclusive A₁ peak (530.1 eV), which is assigned to the π*(C=O) of the organic carbonate. The peak A₂ (531.0 eV) is assigned to quinone contributions\textsuperscript{33}. The results of the O K-edge demonstrate that the GNP\textsubscript{C–O,1} is enriched with quinone. The presence

Results

Synthesis and structure characterization. We adopted a two-step method to prepare edge-oxygenated graphitic nanoplatelets.
of a peak A2 shoulder indicates the GNPC C=O and pRGO also contain less quinone groups. Peak A3 (531.8 eV) may be composed of ketones or carboxylic acid. Fortunately, peak A3 can be identified by peak C2 (539.6 eV), which originates from the σ*(C–O) in carboxylic acid (COOH)32–34,36. Peaks A3 and C2 are dominant in GNPC=O,2 and pRGO, which indicates that they are mainly composed of COOH.

We can draw the conclusion that the quinone was easily formed in the short reaction time (GNPC=O,1). However, the carbonyl group was the main after the prolonged reaction time (GNPC=O,2). The contribution of π* peak B is complicated. It originates from the resonance transition of quinone, or the charge transfer due to the influence of the neighboring O in the etheric ring33. The GNPC=O,C gives rise to a σ* resonance at 536.3 eV (peak C3), which is attributed to C–O–C34. Here, we noticed that GNPC=O,C contains peak A3, and a small shoulder of π*(C=O) in the C K-edge, which indicates that some carbonyl-related groups are present in the GNPC=O,C.

FTIR is another powerful tool for characterizing functional groups, and was employed here. As shown in Fig. 2a, all of the spectra exhibit a broad band at around 3400 cm−1, which is mainly assigned to the stretching vibration (υ OH) of the adsorbed moisture in KBr. Another common band, located at 1610 cm−1, is attributed to the asymmetric stretching of sp2-hybridized C–C bonds (υ C–C). Its shifting and intensity are intimately related to the oxygenated groups37,38.

The fingerprint region, ranging from 1750 to 1000 cm−1, was scrutinized to identify the functional groups present. The well-documented pRGO was analyzed first. Consistent with other reports38–42, the functional groups in the pRGO were assigned to ketones (υ C=O, the shoulder at 1654 cm−1), carbonylic acid (υ C=O at 1715 cm−1, δ OH at 1402 cm−1, v C–OH at 1214 cm−1), organic carbonate (υ C=O at 1715 cm−1, δ OH at 1384 cm−1), ethers (υ C=O at 1084 cm−1), and aromatic hydrocarbons (δ C–H at 740 cm−1)38–42.

Both GNPC=O,1 and GNPC=O,2 produced a typical fingerprint band of carboxylic acid (υ C=O in COOH, 1716 cm−1). The COOH is formed via hydrolysis in acid33,44. GNPC=O,1 and GNPC=O,2 also displayed a peak and a shoulder, respectively, at 1632 cm−1, which was thought to be the quinone based on the XANES results. The GNPC=O,C displayed a broad band from 1380 to 1050 cm−1, which was attributed to the asymmetric C–O–C stretching vibration (in-plane υ C–O–C) of the etheric rings. This is caused by an unusual absorption mechanism40.

The XPS was then performed (Supplementary Fig. 7). It is known that the C 1s typically exhibits a pronounced
asymmetric tail at higher binding energy\textsuperscript{45}, making it difficult to obtain accurate deconvolution results, especially considering the controversial assignment. Here, we only focused on the O 1s spectroscopy.

The high-resolution O 1s are shown in Fig. 2b. After careful deconvolution with the same standard rules, four distinct regions were identified. To easily distinguish them, we labelled them O\textsubscript{1} (531.35 ± 0.05 eV, C=C-O related groups)\textsuperscript{6,45}, O\textsubscript{3} (532.8 eV, C-O=C or COOH)\textsuperscript{6,47}, O\textsubscript{II} (534.6 eV, C-O(H))\textsuperscript{40}, and O\textsubscript{IV} (535.6 eV, physically adsorbed H\textsubscript{2}O and O\textsubscript{3})\textsuperscript{41,42}. Because the different groups overlap, it was difficult to determine the contents of the groups by O 1s alone. However, we can still draw some useful information. All samples had a very small O\textsubscript{III} region, which indicates that very little phenolic -OH is present. Based on the XANES and FTIR results, it was determined that the O\textsubscript{II} regions in GNPC\textsubscript{O-C} and GNPC\textsubscript{C-O} are the same and related to the -OH peak at 531.35 eV. The O\textsubscript{IV} region still could not be distinguished.

Fortunately, quinone is sensitive to cyclic voltammetry (CV). Since the quinone redox couple is the only reaction that can be detected by CV, we used CV to characterize the content of quinone in the samples (Supplementary Fig. 8)\textsuperscript{10,31}. As shown in Fig. 2c, the quinone content was 4.2%, 7.4%, 0.0%, and 2.3% for GNPC\textsubscript{C-O} (C'O,1), GNPC\textsubscript{O-C} (C'O,2), GNPC\textsubscript{C-O} (C'O,3), and pRGO, respectively. The absence of a quinone redox couple indicates the presence of ketone. The reduced quinone redox couple in pRGO demonstrates that most of the O\textsubscript{I} region is composed of COOH. The functional groups of the samples are summarized in Supplementary Table 2.

Oxygen reduction to hydrogen peroxide. The oxygen reduction to hydrogen peroxide (ORHP) performance of the synthesized samples with different oxygen groups was evaluated using a rotating ring–disk electrode (RRDE). Since H\textsubscrip{2}O\textsubscript{2} can upshift the potential of the reference electrode because of its strong oxidizing nature (Supplementary Fig. 9), a salt bridge must be used to separate the electrochemical cell and reference electrode (Supplementary Fig. 10). For a fair comparison of activity, all of the tested samples were tuned to the same capacitance by changing the diameter of drop-cast ink. We used a reversible hydrogen electrode (RHE, Supplementary Fig. 11) for all our analyses.

The polarization curves were measured in O\textsubscript{2}-saturated 0.1 M KOH solution at a scan rate of 1600 rpm. To eliminate the contributions of capacitance, we averaged the current of the forward and backward scans. The H\textsubscript{2}O\textsubscript{2} current (J\textsubscript{H2O2}) was collected at the ring electrode (J\textsubscript{R}) with an applied potential of 1.15 V, and the collection efficiency was 37% (Supplementary Fig. 12). The four-electron byproducts of H\textsubscript{2}O\textsubscript{2} were calculated using the relationship: J\textsubscript{H2O2} = J\textsubscript{R} - J\textsubscript{K}.

As shown in Fig. 3a and Supplementary Table 3, the carbonyl-enriched samples, GNPC\textsubscript{C-O} (0.826 V), GNPC\textsubscript{C-O} (0.815 V), and pRGO (0.810 V), exhibited a higher onset potential than GNPC\textsubscript{C-O} (0.805 V). Here, the onset potential is defined as the potential measured at a current density of 0.15 mA cm\textsuperscript{-2} (5% of the theoretical limiting current) for ORHP. The result indicates that the sample with the in-plane etheric ring had lower activity than the carbonyl-enriched samples.

It is noteworthy that all of the onset potentials were even higher than the thermodynamic equilibrium potential (O\textsubscript{2} + H\textsubscript{2}O + 2 e\textsuperscript{-} → H\textsubscript{2}O\textsubscript{2} + OH\textsuperscript{-}, 0.75 V vs RHE). This counterintuitive phenomenon has been widely reported\textsuperscript{16,48}. Bao and his colleagues attributed this to a Nernst-related potential shift due to the low concentration of H\textsubscript{2}O\textsubscript{2} in the electrolyte, and/or a possible pH-related change\textsuperscript{48}. Although H\textsubscript{2}O\textsubscript{2} can upshift the potential of the reference electrode, its influence can be easily ruled out by employing a salt bridge and refreshing the electrolyte for every measurement. This abnormal phenomenon did not exist with a planar electrode, which further confirms that the potential shifting is unrelated to the yield of H\textsubscript{2}O\textsubscript{2}. We suggest this abnormal phenomenon only results from the localized pH-related change in the mesoporous electrode due to the constraint of mass transport.

The kinetic current of H\textsubscript{2}O\textsubscript{2} (I\textsubscript{K,H2O2}) was calculated according to the Koutecky-Levich equation: I\textsubscript{K,H2O2} = I\textsubscript{L,H2O2} + I\textsubscript{f,H2O2}, where I\textsubscript{K,H2O2} is the measured current of the H\textsubscript{2}O\textsubscript{2} yield, I\textsubscript{L,H2O2} is the theoretical limiting current of ORHP\textsuperscript{23,49}, and I\textsubscript{f,H2O2} was obtained from the Levich equation and determined to be 2.9 mA cm\textsuperscript{-2}. The calculated results are shown in the Tafel plot in Fig. 3b. GNPC\textsubscript{C-O}, and GNPC\textsubscript{C-O} showed the smallest Tafel slope (48 mV dec\textsuperscript{-1}), which was superior to the C-O-C-enriched GNPC\textsubscript{C-O} (55 mV dec\textsuperscript{-1}). The same Tafel slope indicates similar active sites. Although the pRGO was also carbonyl-enriched, its Tafel slope was only 63 mV dec\textsuperscript{-1}. This is probably due to the considerable N content, which is inevitably introduced during the preparation of pRGO. The doped N is known to enhance the yield of 4e product H\textsubscript{2}O\textsubscript{2}\textsuperscript{50}.

The I\textsubscript{K,H2O2} at 0.65 and 0.75 V were selected to compare activity (Fig. 3c). We note that GNPC\textsubscript{C-O} had the highest activity (25.1 mA cm\textsuperscript{-2}) at 0.65 V. However, the most active sample was GNPC\textsubscript{C-O} (3.6 mA cm\textsuperscript{-2}) at a higher potential of 0.75 V. This phenomenon is caused by the samples’ different selectivity (the H\textsubscript{2}O\textsubscript{2} yield ratio, Fig. 3d), which plays a major role in the low potential region. The GNPC\textsubscript{C-O} exhibited higher selectivity (the H\textsubscript{2}O\textsubscript{2} yield ratio, 97.8% at 0.75 V) with an electron transfer number of nearly 2 (Supplementary Fig. 13), which is the highest ratio reported so far (Supplementary Table 3)\textsuperscript{16,18,19,21,22,25,29,48}. The pRGO exhibited the poorest I\textsubscript{K,H2O2} (1.2 mA cm\textsuperscript{-2} at 0.75 V) and the lowest H\textsubscript{2}O\textsubscript{2} yield ratio (83.4% at 0.75 V) because of the presence of graphitic N (Supplementary Fig. 14), which is reported to be the active sites for 4e ORR\textsuperscript{50}.

The H\textsubscript{2}O\textsubscript{2} yield ratio of COOH-enriched GNPC\textsubscript{C-O} decreased relative to the quinone-enriched GNPC\textsubscript{C-O}. The possible reason is that COOH has a negative impact on selectivity. The H\textsubscript{2}O\textsubscript{2} yield ratio of all of the carbonyl-enriched samples decreased as we lowered the potential. This may have occurred due to the reduction of quinone to hydroquinone at low potential. Distinctively, the GNPC\textsubscript{C-O} displayed a tendency opposite to the others, due to the absence of quinone.

The ORHP performance was also evaluated in neutral (0.05 M Na\textsubscript{2}SO\textsubscript{4}) and acidic (0.1 M HClO\textsubscript{4}) media. The results are shown in Supplementary Fig. 15. Both results in neutral and acidic media demonstrated that the quinone-enriched sample (GNPC\textsubscript{C-O}) had the highest selectivities, which were 95% (0.6 V in neutral) and 85% (0.2 V in acid), respectively. In the neutral medium, the onset potential is the same with the thermodynamic equilibrium potential (0.70 V), which indicates the absence of potential shift, which is caused by the localized pH-related change. In the acidic medium, the activity toward ORHP is poor. The result agrees well with the previous reports\textsuperscript{16–18}.

Besides selectivity and activity, stability is also one of the three indispensable factors for catalysts, and it affects their economical durability. The samples’ stability was measured in an H-type cell, which was separated by a Nafion 115 membrane (Supplementary Fig. 16). It is known that the alkaline H\textsubscript{2}O\textsubscript{2} solution is unstable, and spontaneous decomposition will occur\textsuperscript{31,32}. Approximately 30% of H\textsubscript{2}O\textsubscript{2} was decomposed during a 30 h stability test (Supplementary Fig. 17). Therefore, MgSO\textsubscript{4}, a popular stabilizer in alkaline solution, was selected to suppress the decomposition\textsuperscript{31}. 

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As shown in Fig. 3e, the current exhibited a slow decline as the time increased. The possible reason is that the increasing H$_2$O$_2$ concentration enhances electrolyte viscosity. The increasingly sluggish diffusion of O$_2$ in the H$_2$O$_2$ solution subsequently deteriorates the ORHP current. This speculation was further verified by the observation that there was no current drop after changing to a fresh electrolyte. Even after 120 h, there was no evident decline, which demonstrates that the catalyst has good stability.

The concentration of generated H$_2$O$_2$ was determined using the KMnO$_4$ titration method (Supplementary Movie 1). With an applied potential of 0.65 V in 90 mL electrolyte, a typical concentration of H$_2$O$_2$ after 30 h reaction is about 6.1 mM, with a Faraday efficiency of 95%.

**Determination of active sites.** So far, we have shown that the carbonyl-enriched groups possess higher activity than the etheric ring groups. However, the nature of the active carbonyl groups remains elusive. To elaborate the nature of the active carbonyl groups, we designed a special experiment.

We tuned the content of quinone and carboxylic acid by leaching the sample of GNPC=O,2 in an acidified concentrated H$_2$O$_2$ solution. The leached sample was labelled GNPC=O,3. As shown in Fig. 4a, FTIR determined that the carboxylic acid content evidently increased after leaching. However, the quinone content was reduced from 7.4% to 4.5% (Fig. 4b). This change in groups was also recorded by XPS (Supplementary Fig. 18). The active sites can be easily identified by comparing the ORHP performance of GNPC=O,2 and GNPC=O,3. The polarization curves are shown in Fig. 4c. The GNPC=O,3 had both a lower onset potential of 14 mV and $J_{K,H_2O_2}$ (3.1 mA cm$^{-2}$, Supplementary Fig. 19) than GNPC=O,2. The relation of $J_{K,H_2O_2}$ and quinone content was further checked, and the results are shown in Fig. 4d. $J_{K,H_2O_2}$ increased as the quinone content increased. Because of the influence of other groups, it did not follow a linear relation. These results confirmed that the active sites were quinone.
To further verify these results, we investigated several standalone molecules with quinone, carboxylic acid, and etheric ring groups, such as phenanthrenequinone, anthraquinone, naphthalenetetracarboxylic dianhydride, perylenetetracarboxylic dianhydride, dibenzodioxin, and dibenzofuran. The polarization curves are shown in Fig. 5. Except for phenanthrenequinone and anthraquinone, the other four molecules did not show activity towards ORHP; the activity was inferior to blank glass carbon (GC). The phenanthrenequinone was superior to anthraquinone in both the $J_{\text{H}_2\text{O}_2}(0.7 \text{ vs } 0.5 \text{ mA cm}^{-2} \text{ at } 0.65 \text{ V})$ and Tafel slope results (45 vs 48 mV dec$^{-1}$, Supplementary Fig. 20). These molecular chemistry results further confirm that the quinones are the active sites.

**Theoretical investigation.** To gain atomistic insights about the nature of the active quinone motifs, we next used density functional theory (DFT) calculations. We examined a variety of model structures (Fig. 6a) to study the different possible quinone groups on the edge and basal planes. These model structures were used to model the ORHP reaction pathway (Eqs. 1 and 2)$^{5,16}$:

$$\text{O}_2 + \text{H}_2\text{O} + e^- + * \rightarrow \text{OOH}* + \text{OH}^- \quad (1)$$

$$\text{OOH}* + e^- \rightarrow \text{HO}_2^* + * \quad (2)$$

where the O$_2$ molecule adsorbs at the carbon surface and is reduced through the first proton-electron transfer to form OOH$^*$. Theoretical investigation. To gain atomistic insights about the nature of the active quinone motifs, we next used density functional theory (DFT) calculations. We examined a variety of model structures (Fig. 6a) to study the different possible quinone groups on the edge and basal planes. These model structures were used to model the ORHP reaction pathway (Eqs. 1 and 2)$^{5,16}$:

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Fig. 4 Determination of active sites. a FTIR. The COOH is increased. b Cyclic voltammetry, which was measured in Ar-saturated 0.5 M H$_2$SO$_4$ at a scan rate of 50 mV s$^{-1}$. The quinone is decreased. c The polarization curves of $J_{\text{H}_2\text{O}_2}$, $J_{\text{D}}$, and $J_{\text{H}_2\text{O}}$. The curves were measured in O$_2$-saturated 0.1 M KOH solution at a scan rate of 10 mV s$^{-1}$ by RRDE with a rotation speed of 1600 rpm. The current was the average of the forward and backward scans. d The $J_{\text{K}, \text{H}_2\text{O}_2}$ as a function of quinone content.

![Fig. 4 Determination of active sites. a FTIR. The COOH is increased. b Cyclic voltammetry, which was measured in Ar-saturated 0.5 M H$_2$SO$_4$ at a scan rate of 50 mV s$^{-1}$. The quinone is decreased. c The polarization curves of $J_{\text{H}_2\text{O}_2}$, $J_{\text{D}}$, and $J_{\text{H}_2\text{O}}$. The curves were measured in O$_2$-saturated 0.1 M KOH solution at a scan rate of 10 mV s$^{-1}$ by RRDE with a rotation speed of 1600 rpm. The current was the average of the forward and backward scans. d The $J_{\text{K}, \text{H}_2\text{O}_2}$ as a function of quinone content.](image1)

![Fig. 5 The ORHP performance of standalone molecules. Phenanthrenequinone, anthraquinone, naphthalenetetracarboxylic dianhydride, perylenetetracarboxylic dianhydride, dibenzodioxin, and dibenzofuran, and blank glass carbon (GC) were compared. The curves were measured in O$_2$-saturated 0.1 M KOH solution at a scan rate of 10 mV s$^{-1}$ by RRDE with a rotation speed of 1600 rpm. The applied potential of the ring was 1.15 V. The current was the average of the forward and backward scans.](image2)
groups on the edge seems more feasible than in the basal area, because the formation of Q-Basal groups significantly interrupts the \( sp^2 \) network and requires a lot of energy input. However, the edge-located structures of Q-Edge 4 and 5 are easily formed, because the \( sp^2 \) C–C bond breaking is lower. Thus, the Q-Edges are the most likely sites.

Furthermore, a different mechanism similar to industrial anthraquinone process was considered. The results of the anthraquinone mechanism shown in Supplementary Figs. 21 and 22 reveals that the formation of anthrahydroquinone (AHQ) is uphill by 3.64 eV. In addition, the next step, which is transferring proton from of AHQ to \( O_2 \) molecule and forming radical OOH, is even more exergonic around 4.93 eV. Both of them are far beyond the energy capacity for the two-electron ORR. Therefore, the anthraquinone mechanism is not a possible competitive pathway for our catalyst system.

**Discussion**

In summary, we adopted a pre-activated method to decorate the dangled edges of graphitic materials with targeted groups (ether, carboxyl and quinone). The functional groups were then characterized by a combination of soft XANES, XPS, FTIR and CV. Our results confirmed a new class of quinone-edged groups, which exhibited higher selectivity than previously reported oxygenated groups with similar onset potential. The quinone-enriched samples (GNPC\(_{O-C} \)) exhibited a \( H_2O_2 \) yield ratio of 97.8% at 0.75 V. The results were further verified using stand-alone molecular chemistry and theoretical analysis. These findings will be beneficial for understanding active sites in ORHP, and will be a guide to designing high ORHP catalysts.

**Methods**

**Preparation of graphitic nanoplatelets (GNP).** We first cleaved graphite using the mechanochemical method. The graphite crushing and exfoliation into nanosized particles were accomplished at the same time. The freshly broken edges are free and reactive. Then, the activated edges were reacted with target molecules, such as \( CO_2 \) and \( O_2 \). The groups were edge-enriched with \( CO_2 \) and \( O_2 \), and the resulting as-prepared GPNs were designated as GNPC\(_{O-C} \) respectively.

**GNPC\(_{O-C} \):** the mechanochemical ball-milling method was employed to activate the surface and saturate the graphite at the same time. The preparation was conducted on a planetary ball-milling device (Pulverisette 6, Fritsch GmbH) in a rotation speed of 500 rotation per minutes (rpm). In brief, graphite (15 g, Alfarcens, 100 mesh, 99.9995%, product number: 14735), dry ice (100 g, Hanyu Chemical Inc.), and hardened steel balls with a diameter of 3 mm (500 g) were placed into a ball-mill container (250 mL). Then, the air in the container was completely pumped out by five repeated argon (Ar) charging/discharging cycles for 15 min and ball-milled under reduced pressure for 15 h. Finally, the as-prepared GNPC\(_{O-C} \) was leached in 1 M aq. \( H_2SO_4 \) solution for 24 h to completely remove possible contamination from unbound Fe debris, followed by rinsing with ultra-pure water (18.2 M cm, Direct-Q 3 UV, Millipore Corporation) more than 6 times and freeze-drying in tert-butyl alcohol. Finally, the samples were further dried in vacuum oven at 80 °C for 10 h.

**GNPC\(_{O-C} \):** the amount of carboxyl-related group loading was controlled by varying the graphite loading amounts and ball-milling conditions. GNPC\(_{O-C} \) was prepared with more carboxyl-related groups by ball-milling graphite (10 g) and dry ice (100 g) with hardened steel balls (500 g, \( \Phi = 5 \) mm) for 40 h.

**GNPC\(_{O-C} \):** the different contents of carboxyl-related groups was obtained by \( H_2O_2 \) leaching in acid. The leaching process was conducted by immersing GNPC\(_{O-C} \) in 20 mL 3.5 M aq. \( H_2SO_4 \) and 1.0 M acetic acid mixture for 12 h. Here, acetic acid acted as a stabilizer to suppress the spontaneous decomposition of \( H_2O_2 \). Finally, the samples were further dried in a vacuum oven at 80 °C for 10 h before characterizations.

**GNPC\(_{O-C} \):** the mechanochemical ball-milling method was first applied for activation. The experiment procedures were conducted in a planetary ball-milling device (Pulverisette 6, Fritsch GmbH) at a rotation speed of 500 rpm with the protection of 5 bar Ar (UHP, 99.999%, N50, KOSEM, Korea). In brief, graphite (15 g) and hardened steel balls (500 g, \( \Phi = 3 \) mm) were charged in a ball-mill container (250 mL). Then, the container was filled with argon gas (5 bar), after five purging cycles with the aid of a vacuum pump to remove residual air.

After cooling to room temperature, the container was filled with an \( O_2/Ar \) mixture (10 vol%) for 6 h at a flow rate of 250 standard cubic centimeters per
minute (scm). SAFETY NOTE: The concentration of O₂ in the gas mixture should be lower than the burn-off point to avoid fire, which can be caused by violent oxidation.

Since the dangling edges activated by unzipping the graphene framework tend to reconstruct spontaneously to reduce their surface energy, the cleavage process was exponentially reduced as the ball-milling was prolonged. Gas oxidation in the O₂/Ar mixture was divided into 7 periods. The ball-milling time for each period was 20 min, 20 min, 20 min, 30 min, 30 min, 60 min, and 120 min, respectively. The total ball-milling time was 5 h.

Finally, to completely remove unbound Fe debris, the as-prepared GNPc₁₋₀₋₀ was washed in 1 M aq. H₂SO₄ solution for 24 h, followed by rinsing with ultra-pure water more than six times and freeze-drying in tert-butyl alcohol. Finally, the samples were further dried in a vacuum oven at 80 °C for 10 h.

**Structural characterization.** The microstructures were characterized on a JEM-2100 transmission electron microscope (TEM) at an accelerating voltage of 200 kV, and by field emission scanning electron microscopy (FESEM, Nova NanoSEM, FEI), equipped with energy disperse spectroscopy (EDS, EDAX, AMETEK). X-ray Diffraction (XRD) patterns were recorded on a D/Max2500V (Rigaku, Japan) using Cu-Kα radiation (40 kV, 100 mA, λ = 1.5418 Å) in a 2θ range of 3°–60° at a scan rate of 4° min⁻¹. The specific surface area was analyzed on a Micromeritics ASAP 2504 N by nitrogen adsorption-desorption isotherms using the Brunauer-Emmet-Teller (BET) method. The pore distributions were calculated by the non-local density functional theory (NLDFT) method.

Fourier transform infrared spectra (FTIR) were collected on a Perkin-Elmer Spectrum 100 with a resolution of ~1 cm⁻¹, and the samples were tableted with KBr as support. The Raman spectra were characterized on a WITec Alpha300R with a laser wavelength of 532 nm.

The elemental analysis (EA) was conducted using a Flash 2000 CHNS/O Analyzers (Thermo Scientific). All samples were measured at least three times. The element of iron was determined by time-of-flight secondary ion mass spectrometry (TOF-SIMS, TOFSIMS®, IONTOF GmbH, Germany) with a resolution of (ppm). The primary ion species was Bi with a dose of 2.0 × 10¹⁰ and the raster area was about 400 × 400 μm². The X-ray photoelectron spectra (XPS) was recorded on a Thermo Fisher XPS spectrometer (K-alpha), which employed monochromatic Al Kα radiation as the X-ray source. The soft X-ray absorption near edge structure (XANES) experiments were performed at the BL12R-A beamline in the National Synchrotron Radiation Laboratory (NSRL), University of Science and Technology of China (USTC), Hefei, P. R. China.

**Electrochemical measurements.** The electrochemical measurements were conducted in a three-electrode electrochemical cell on a workstation of CompactStat (Ivium Technologies B.V., Netherlands). A graphite rod (Alfa Aesar, Ultra purity, 99.9995 %) and an Ag/AgCl electrode were selected as the counter electrode and the reference electrode, respectively. As-prepared GNPc₁₋₀₋₀, GNPc₁₋₀₋₀, GNPc₁₋₀₋₀, and rGO inks were drop-cast on glassy carbon (GC, 0.247 cm²) with a vacuum of 20 Å to decouple the periodic replicas. To model the quinone element of iron was detected by time-of-flight secondary ion mass spectrometry (TOF-SIMS) and by light secondary ion mass spectrometry (SIMS, FEI), equipped with energy dispersive spectroscopy (EDS, EDAX, AMETEK). X-ray photoelectron spectroscopy (XPS) was recorded on a Thermo Fisher XPS spectrometer (K-alpha), which employed monochromatic Al Kα radiation as the X-ray source. The soft X-ray absorption near edge structure (XANES) experiments were performed at the BL12R-A beamline in the National Synchrotron Radiation Laboratory (NSRL), University of Science and Technology of China (USTC), Hefei, P. R. China.

To obtain reliable and reproducible measurements, the cleanliness of the GC supports is particularly important. Before each measurement, the cleanliness of blank GC was verified by scanning CV at a scan rate of 50 mV s⁻¹. The current curves should only exhibit the shape of capacitance with a current density less than an order of magnitude below 10⁻⁷ A. If it was not, the GC support was polished with alumina (0.05 μm), and then ultrasonically cleaned in ethanol and ultra-pure water.

**Computational method.** We used the Atomic Simulation Environment (ASE)³⁴ to handle the simulation and the QUANTUM ESPRESSO³⁵ program package to perform electronic structure calculations. The electronic wavefunctions were expanded in plane waves up to a cutoff energy of 500 eV, while the electron density is represented on a grid with an energy cutoff of 5000 eV. Core electrons were approximated using ultrasoft pseudopotentials. To describe chemisorption properties on graphene structures, we used the PBE exchange-correlation functional with dispersion correction. Graphene structures were modeled as one layer with a vacuum of 20 Å to decouple the periodic replicas. To model the quinone functional groups in the basal plane, we use a 5 × 5 super cell lateral size, and the Brillouin zone was sampled with (4 × 4 × 1) Monkhorst-Pack k-points. For the oxygen functional groups in the edge, we used a super cell with a lateral size 5 × 6 and the Brillouin zone was sampled with a (1 × 4 × 1) Monkhorst-Pack k-points.

**Data availability**

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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Author contributions
J-B.B. conceived the project and oversaw all the research phases. J-B.B. and G-F.H. designed the project. G-F.H. synthesized and characterized the samples. F.L., Z.F., Y.L., and W.Z. measured the soft XANES S.S. and M.K. conducted the theoretical calculations. S-W.K. and Y.B. built the two-electrode device. S-K.K. and J-P.J. performed BET measurements. Data collection and analysis were conducted by J-B.B., S.S., and G-F.H. All the authors contributed to and commented on this paper.

Competing interests
The authors declare no competing interests.

Additional information
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