Leucine-Rich Alpha-2-Glycoprotein (LRG-1) as a Potential Kidney Injury Marker in Kidney Transplant Recipients

Background: Kidney transplantation is the treatment of choice for most patients with end-stage renal disease. To improve patient and transplant survival, non-invasive diagnostic methods for different pathologies are important. Leucine-rich alpha-2-glycoprotein (LRG-1) is an innovative biomarker that is elevated in cases of angiogenesis, inflammation, and kidney injury. However, there are limited data about the diagnostic role of LRG-1 in kidney transplant recipients. The aim of this study was to evaluate the association between serum LRG-1, urine LRG-1, and kidney transplant function and injury.

Material/Methods: We enrolled 35 kidney transplant recipients in the study. LRG-1 in the serum and urine was detected using ELISA. We evaluated the correlation of serum and urine LRG-1 with traditional serum and urine kidney injury markers.

Results: A higher level of serum LRG-1 correlates with a higher level of urine LRG-1. Serum LRG-1 has a positive correlation with transplant age, serum urea, serum creatinine, serum cystatin C, proteinuria, and fractional excretion of sodium (FENa) and a negative correlation with hemoglobin and estimated glomerular filtration rate (eGFR). Urine LRG-1 has a positive correlation with serum cystatin C, proteinuria, and urine neutrophil gelatinase-associated lipocalin (NGAL).

Conclusions: Higher levels of serum and urine LRG-1 are associated with kidney transplant injury and functional deterioration. Thus, LRG-1 might be also as a biomarker for tubular dysfunction in patients after kidney transplantation.

Keywords: Kidney Function Tests • Kidney Transplantation • LRG1 Protein, Human

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Background

For most patients with end-stage renal disease, kidney transplantation is the treatment of choice because it significantly improves survival and quality of life compared to dialysis [1]. Leucine-rich alpha-2-glycoprotein-1 (LRG-1) could be a potential non-invasive diagnostic tool for kidney transplant recipients. Improving the health of recipients and maintaining graft function not only improves the quality of patient’s lives and survival but also reduces the need for re-transplantation and increases the number of organs available for transplantation [2].

LRG-1, which was first isolated from human serum by Haupt and Baudner in 1977 [3], is a protein that is expressed during blood cell-granulocyte differentiation [4]. LRG-1 consists of 347 amino acids [5,6] and has a molecular weight of 50 kDa [5]. Based on mRNA analysis, LRG-1 is thought to be normally produced in liver cells and neutrophils [4], but it may be expressed in a variety of cells in the presence of inflammatory cytokines (interleukin-6, interleukin-1b), lipopolysaccharides and tumor necrosis factor-alpha (TNF-alpha) [3,6]. LRG-1 levels increase in inflammatory processes and infectious diseases [3,7-10], malignancy [11-16], autoimmune diseases such as rheumatoid arthritis [10], inflammatory bowel disease [16], systemic lupus erythematosus [17], and the pathogenesis of diabetic nephropathy [5].

LRG-1 has been described as a biomarker of acute kidney injury [6] and tubular dysfunction [5,6]. Terada et al showed that serum and urine LRG-1 levels were increased in a mouse model of acute kidney injury [6]. In that study, immunohistological examination showed that LRG was mainly expressed in renal tubular cells [6]. Lee and co-authors also studied a mouse model and observed that urine LRG-1 was expressed in proximal, distal tubules and collecting ducts. Data suggest that LRG-1 could be used as a marker of tubular damage in various kidney diseases [5].

There are few studies on the role of LRG-1 in patients undergoing renal replacement therapy [18,19]. In a study by Glorieux and co-authors, plasma LRG-1 levels were 2-fold higher in patients with end-stage renal disease compared with chronic kidney disease (CKD) stages 2 and 3 [18]. Serum LRG-1 has been shown to have a negative correlation with GFR [18]. Thus, LRG-1 could be a potential marker for CKD [18].

Banon-Maneus and co-authors studied the presence of various proteomes in urine and concluded that urine LRG-1 can be used as a biomarker to assess the severity of interstitial fibrosis and tubular atrophy in patients after kidney transplantation [19].

Few studies have described the diagnostic role of LRG-1 in kidney transplant recipients. Therefore, the aim of the present study was to evaluate the association between this biomarker and the function and injury of kidney transplants.

Material and Methods

This study was performed according to the ethical principles of the Declaration of Helsinki and approved by the Clinical Research Ethics Committee of the Development Society of Pauls Stradiņš Clinical University Hospital, approval number 280520-18L. The patients provided written informed consent prior to study participation.

The study, conducted in spring 2020 at Pauls Stradiņš Clinical University Hospital, included 35 patients over 18 years old with a stable stage functioning kidney transplant. The study did not include patients with active infections, autoimmune diseases or diabetes mellitus, patients who had had a kidney transplantation in the last 12 months, or pregnant women. Following criteria despite kidney function may influence LRG-1 levels.

Baseline demographics and clinical characteristics were obtained from patient and medical records. We collected peripheral blood and urine samples. Urine and blood examination was performed 7.7 (±6.9) years after transplantation (median time – 6 years [1 to 25]). To assess the association between LRG-1 and kidney function, we used the correlation with serum creatinine, eGFR and cystatin C. Clinical chemistry tests (eg, creatinine, urea, sodium) were evaluated on a Siemens Atellica analyzer. GFR was estimated according to the MDRD (Modification of Diet in Renal Disease) formula. NGAL was measured using chemiluminescent microparticle immunoassay. Correlation with urine NGAL and FENa was performed to obtain association between LRG-1 and tubular dysfunction. Proteinuria was detected using the protein-to-creatinine ratio.

Human Leucine-rich alpha-2-glycoprotein 1 ELISA kit (Catalog No. NBP2-60577, Novus Biologicals, USA) was used according to the manufacturer’s instructions to detect LRG-1 in blood serum and urine. This ELISA test is a quantitative sandwich-type enzyme immunoabsorbent technique. The obtained result was converted from ng/mL to μg/mL for further statistical analysis.

The obtained data were processed with Microsoft Office Excel, IBM SPSS Statistics and R-Studio programs. Parameters are reported as median values with interquartile ranges, and the frequency of results as a percentage. Given that the obtained data did not correspond to the normal distribution, the Spearman method was used to determine correlations between variables. The correlation is considered weak if the correlation coefficient (r) is less than 0.25, moderate if r=0.25-0.75, and strong if r >0.75. Age adjustment was used for data analysis. To estimate urine LRG-1 as a biomarker for proteinuria detection, we used receiver operating characteristic (ROC) curve.
Results

Patient Demographic and Clinical Characteristics

There were 35 patients with a functioning kidney graft enrolled in this study. Their median age was 57 years, and 60% of patients were male. The median transplant age was 6.0 (IQR 2.0-10.0) years. The patient median serum creatinine was 123 (IQR 108-176) µmol/L, and the eGFR was 47.1 (IQR 33.7-60.0) mL/min/1.73 m². The most common maintenance immunosuppressive regimen was therapy with tacrolimus, mycophenolate, and prednisolone (48.6% of patients). Patient characteristics are listed in Table 1.

Serum and Urine LRG-1 Levels in Kidney Transplant Recipients

The mean concentration of serum LRG-1 was 51.07±19.81 µg/ml, median – 47.1 µg/mL, and the mean concentration of urine LRG-1 was 0.17±0.19 µg/ml, median – 0.09 µg/mL. After adjustment for patient’s age, there was a positive correlation between serum and urine LRG-1 (r=0.41; 95% CI: 0.09-0.66; p=0.01). Linear regression analysis was conducted with urine LRG-1 as dependent variable. Every 1 ug/ml increase in urine LRG-1 was associated with a 53-units increase in serum LRG-1. Linear regression accounted for R²=25% of the variance of the serum LRG-1, as shown in Figure 1.

Serum and Urine LRG-1 Correlation with Other Quantitative Data

After adjustment for patient’s age, serum LRG-1 had a positive correlation with transplant age (r=0.52; 95% CI: 0.26-0.71; p<0.01), serum urea (r=0.59; 95% CI: 0.23-0.68; p<0.01), serum creatinine (r=0.33; 95% CI: 0.01-0.59; p=0.04), serum creatatin C (r=0.42; 95% CI: 0.08-0.67; p=0.01), serum phosphorus (r=0.48; 95% CI: 0.14-0.72; p<0.01), proteinuria (r=0.35; 95% CI: 0.02-0.61; p=0.03), and FENa (r=0.35; 95% CI: 0.01-0.62; p=0.04). Serum LRG-1 had a negative correlation with

| Characteristic of patients                  | Value |
|--------------------------------------------|-------|
| Male, no.                                  | 21 (60%) |
| Median patient age, years (Q1; Q3)         | 57 (48; 64) |
| Primary diagnosis, no.                     | 11 (31%) |
| Glomerulonephritis                         | 8 (23%) |
| Autosomal dominant polycystic kidney disease | 6 (17%) |
| Hypertensive nephropathy                   | 2 (6%) |
| Other                                      | 8 (23%) |
| Donor, no.                                 | 29 (83%) |
| Living                                     | 6 (17%) |
| Cadaveric                                  | 2 (6%) |

Kidney transplant, no.

| Characteristic of patients                  | Value |
|--------------------------------------------|-------|
| Primary                                    | 29 (83%) |
| Re-transplant                              | 6 (17%) |

Maintenance immunosuppression, no.

| Characteristic of patients                  | Value |
|--------------------------------------------|-------|
| Glucocorticoids                            | 24 (69%) |
| Tacrolimus                                 | 26 (74%) |
| Cyclosporine                               | 5 (14%) |
| Mycophenolate mofetil                      | 34 (97%) |
| Sirolimus                                  | 3 (9%) |
| Azathioprine                               | 1 (3%) |

Kidney replacement therapy modality before transplantation, no.

| Characteristic of patients                  | Value |
|--------------------------------------------|-------|
| Hemodialysis                               | 21 (60%) |
| Peritoneal dialysis                        | 11 (31%) |
| Combined (hemodialysis and peritoneal dialysis) | 2 (6%) |
| Preemptive transplantation                 | 1 (3%) |
hemoglobin ($r=-0.37$; 95% CI: -0.66 - -0.01; $p=0.01$) and eGFR ($r=-0.35$; 95% CI: -0.01 - -0.61; $p=0.03$). The results are shown in Table 2.

Urine LRG-1 had a positive correlation with serum cystatin C ($r=0.39$; 95% CI: 0.02-0.67; $p=0.03$), parathyroid hormone (PTH) ($r=0.5$; 95% CI: 0.19-0.72; $p<0.01$), proteinuria ($r=0.59$; 95% CI: 0.01-0.3; $p<0.01$) and urine NGAL ($r=0.44$; 95% CI: 0.13-0.67; $p<0.01$) (shown in Table 2).

The chance line on the ROC curve means the same probability of detecting healthy/sick using the urine LRG-1 biomarker as the area of the ROC curve AUC=0.5. The diagnostic accuracy of urine LRG-1 for the diagnosis of proteinuria >0.5 mg/g was moderate (AUC, 0.77; 95% CI: 0.60-0.94) (Figure 2). This means there is a 77% chance by increase of LRG-1 to distinguish between patients with or without proteinuria.

The ROC curve-derived optimal cut-off values for urine LRG-1 was 0.1 ug/ml. The use of these cut-off values resulted in sensitivity (Se), specificity (Sp), positive predictive value (PPV), and negative predictive value (NPV) of 79%, 70%, 65%, and 82%, respectively, for urine LRG-1 (Table 3). This refers to the higher probability of a positive test (79%) than the probability of a negative test (70%).

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### Table 2. Correlation between patient serum LRG-1, urine LRG-1, transplant age, and laboratory analysis results after adjustment for patient age.

| Parameter                | Reference interval | Median value, (Q1, Q3) | Correlation with serum LRG-1 (95% CI) | Correlation with urine LRG-1 (95% CI) |
|--------------------------|--------------------|------------------------|--------------------------------------|--------------------------------------|
| Transplant age (years)   | 6.0 (2; 10)        |                        | 0.52 (0.26-0.71) <0.01*               | 0.26 (0.53-0.06) 0.11                 |
| Hemoglobin (g/L)         | 120-160            | 127 (118; 140)         | -0.37 (-0.66-0.01) 0.01*              | -0.15 (-0.49-0.22) 0.43               |
| Serum urea (mmol/L)      | 3.2-8.2            | 9.7 (7.55; 12.7)       | 0.59 (0.23-0.68) <0.01*               | 0.31 (-0.01-0.58) 0.058               |
| Serum creatinine (µmol/L)| 49-90              | 123 (108; 176)         | 0.33 (0.01-0.59) 0.04*                | 0.33 (-0.002-0.59) 0.051               |
| eGFR (mL/min/1.73 m²)    | >90                | 47.1 (33.7; 60)        | -0.35 (-0.01 - -0.61) 0.03*           | -0.31 (0.01 - 0.57) 0.057              |
| Serum cystatin C (mg/L)  | 0.61-0.95          | 1.78 (1.49; 2.37)      | 0.42 (0.08-0.67) 0.01*                | 0.39 (0.02-0.67) 0.03*                 |
| Serum calcium (mmol/L)   | 2.08-2.65          | 2.33 (2.22; 2.41)      | -0.27 (-0.57 - -0.07) 0.12            | -0.34 (-0.64 - -0.04) 0.08             |
| Serum phosphorus (mmol/L)| 0.78-1.65         | 1.10 (1.01; 1.25)      | 0.48 (0.14-0.72) <0.01*               | 0.10 (-0.27-0.46) 0.58                 |
| Serum PTH (ng/ml)        | 15-68.3            | 119 (72.5; 180)        | 0.29 (-0.08-0.60) 0.12                | 0.50 (0.19-0.72) 0.002*                |
| Serum albumin (g/L)      | 35-52              | 44 (43; 46.8)          | -0.28 (-0.58 - -0.08) 0.13            | 0.38 (-0.67 - -0.01) 0.059             |
| Ferritin (ng/mL)         | 10-291             | 87.6 (59.5; 135)       | 0.21 (-0.16-0.53) 0.27                | 0.01 (-0.36-0.38) 0.94                 |
| Proteinuria (mg/g)       | 0-0.5              | 0.235 (0.1; 0.723)     | 0.35 (0.02-0.61) 0.03*                | 0.59 (0.01-0.30) <0.01*                |
| FENa (%)                 | <1%                | 1.0 (0.75; 1.6)        | 0.35 (0.01-0.62) 0.04*                | 0.11 (-0.21-0.42) 0.48                 |
| Urine NGAL (ng/mL)       | 0-132              | 28.9 (8.85; 52.9)      | 0.29 (-0.03-0.56) 0.08                | 0.44 (0.13-0.67) <0.01*                |

*p value<0.05. eGFR – estimated glomerular filtration rate; PTH – parathyroid hormone; FENa – fractional excretion of sodium; NGAL – neutrophil gelatinase-associated lipocalin.

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This study is one of the few to explore LRG-1 as a potential biomarker for kidney injury, particularly in a kidney transplant recipient population.

There is no defined reference interval for serum and urine LRG-1. Weivoda et al analyzed patients with different infections and autoimmune diseases, and the normal serum LRG-1 level was defined as up to 50 µg/mL based on a control group of 20 patients [20]. In a study of systemic lupus erythematosus, Ahn et al found that a serum LRG-1 level of more than 45.7 ng/mL could indicate disease activity [17]. The reported norms differ significantly since the 2 studies used different units of measurement. In this study, mean serum LRG-1 was 51.07±19.81 µg/mL. In comparison, in a study by Yang et al, which included 169 hemodialysis patients, the mean serum LRG-1 was higher (67.73±15.10 µg/mL) [21]. This result indirectly suggests that patients with lower kidney function have higher serum LRG-1.

There was a positive correlation between serum and urine LRG-1. Similar data have been described in type 2 diabetes mellitus patients who were studied over a 3-year period to investigate the correlation of LRG-1 with albuminuria progression. The researchers found that higher serum LRG-1 correlated with higher urine LRG-1 and indicated the progression of albuminuria [22].

We determined that a higher serum LRG-1 level had a moderate positive correlation with the level of serum urea, serum creatinine, and serum cystatin C and a moderate negative correlation with eGFR. These correlations show that serum LRG-1 is associated with decreased kidney function of the renal transplant. Glorieux et al also found a negative correlation between serum LRG-1 and eGFR [18].

In our study, serum LRG-1 had a negative correlation with the level of hemoglobin, which may indirectly indicate worse kidney transplant function because decreased renal function is associated with erythropoietin deficiency and secondary anemia [23]. In addition, serum LRG-1 had a positive correlation with a higher level of phosphorus. This might be related to a decreased transplant function, tubular injury, and secondary hyperparathyroidism [23]. Yang et al found a negative correlation between serum LRG-1 and hemoglobin in hemodialysis patients, but they did not find a correlation between serum LRG-1 and bone mineral disorders [21].

Table 3. Diagnostic performance of urine LRG-1.

| Parameter | Se (95% CI) | Sp (95% CI) | PPV (95% CI) | NPV (95% CI) | LR+ (95% CI) | LR- (95% CI) |
|-----------|-------------|-------------|--------------|--------------|--------------|--------------|
| Urine LRG-1 >0.1 ug/ml | 0.79 (0.49-0.95) | 0.70 (0.46-0.88) | 0.65 (0.38-0.86) | 0.82 (0.57-0.96) | 2.62 (1.27-5.40) | 0.31 (0.11-0.87) |

Se = sensitivity; Sp = specificity; PPV = positive predictive value; NPV = negative predictive value; LR+ = positive likelihood ratio; LR- = negative likelihood ratio.
We found a positive correlation between serum LRG-1 and transplant age. A longer time after kidney transplantation is associated with decreased GFR levels due to chronic transplant dysfunction [24]. Banon-Maneus et al studied different proteomes in urine. They found that urine LRG-1 may be used as a marker in diagnosing chronic graft dysfunction because it correlates with the severity of interstitial fibrosis and tubular atrophy [19].

In our study, urine LRG-1 had a positive correlation with proteinuria and urine NGAL. Other studies suggest that plasma and urine LRG-1 levels are increased in cases of proteinuria and tubular damage [5,22]. In a study on a mouse model of induced acute kidney injury, Lee et al found that urine LRG-1 was expressed in the renal tubular epithelium and could be used as a tubular damage marker [5]. Urine NGAL is produced by tubular epithelial cells during renal damage [25], and the correlation of these 2 markers (urine LRG-1 and urine NGAL) in our study indirectly suggests tubular damage.

This is a pilot study with the relatively small study cohort. Similar research needs to be performed in a larger cohort to obtain more precise correlations.

Conclusions

Our study demonstrates the potential role of serum and urine LRG-1 as a kidney injury marker in kidney transplant recipients. It correlates with other tubular injury markers (NGAL) and functional deterioration. Further research regarding LRG-1 in a larger post-transplant patient cohort with subsequent transplant biopsies is warranted.

Data Availability Statement

The data that support the findings of this study are not publicly available because they contain information that could compromise the privacy of research participants.

Declaration of Figures’ Authenticity

All figures submitted have been created by the authors, who confirm that the images are original with no duplication and have not been previously published in whole or in part.

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