Effects of Living Mulch and Fertilizer on the Performance of Broccoli in Plasticulture

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Abstract. Living mulch systems allow cover crops to be grown during periods of cash crop production, thereby extending the duration of cover crop growth and associated beneficial agroecosystem services. However, living mulches may also result in agroecosystem disservices such as reduced cash crop yields if the living mulch competes with the crop for limiting resources. We examined whether the effects of an Italian ryegrass \textit{(Lolium multiflorum} (Lam.) Husnot\textit{)–white clover \textit{(Trifolium repens} L., cv. New Zealand\textit{)} living mulch on broccoli \textit{(Brassica oleracea} L. var. \textit{italica}) yield and yield components were dependent on fertilizer rate in field experiments conducted in Durham, NH, in 2011 (Expt. 1) and 2012 (Expt. 2). Drip-irrigated broccoli was grown under a range of organic fertilizer application rates in beds covered with plastic, with and without a living mulch growing in the uncovered, interbed space. Broccoli yields were similar in the living mulch and bare soil controls under the highest rates of fertilizer application in Expt. 1. In Expt. 2, living mulch reduced broccoli yields from 28% to 63%, depending on fertilizer rate. Differences in leaf SPAD values suggest that yield reductions were attributable, in part, to competition for nitrogen; however, other factors likely played a role in determining living mulch effects. Despite yield reductions, the living mulch reduced the prevalence of hollow stem in broccoli in Expt. 1. Organic fertilizer may have inconsistent effects on broccoli yields in living mulch systems.

Cover crops provide many beneficial eco-system services to agricultural production systems, including soil and nutrient retention, resources and habitat for beneficial organisms, and weed suppression (Hartwig and Ammon, 2002). In regions such as northern New England, however, short growing seasons can limit opportunities to establish productive cover crops between cash crop growing periods, particularly in vegetable crop rotations (Snapp et al., 2005). In these cases, living mulch (LM) systems, a form of intercropping that involves growing a cover crop or cover crop mixture simultaneously with a cash crop for part or all of the cropping season, may provide an opportunity to establish cover crops earlier in the growing season and thereby increase the duration of cover crop growth (Teasdale, 1996).

Annual green heading broccoli (\textit{Brassica oleracea} L. var. \textit{italica}) is an important fresh-market vegetable crop in the United States with annual production valued at an estimated \$742.6 million in 2011 (USDA NASS, 2012). In New Hampshire and other northern New England states, broccoli is often planted in the summer in raised beds and harvested in the fall. This production schedule means that it can be especially challenging to establish fall-sown cover crops in these systems. Furthermore, although the beds on which broccoli is planted are often covered with plastic mulch, the spaces between beds are frequently managed as bare soil through the use of cultivation or herbicides. Thus, growing cover crops as LM systems concurrently with broccoli in a plasticulture system may both provide an opportunity to include or expand the use of cover crops in broccoli production systems in short-season regions and reduce the need for soil disturbance in the spaces between beds.

A potential tradeoff of using LMs in broccoli production systems is the possibility of yield reduction resulting from competition from the LM. Reductions in crop yields resulting from LM have been reported for a number of other crops including corn (Faget et al., 2012; Liedgens et al., 2004a, 2004b; Martin et al., 1999), zucchini squash (Walters and Young, 2008), strawberries (Neuweiler et al., 2003), and asparagus (Brainard et al., 2012; Paine et al., 1995). In broccoli LM systems specifically, previous research indicates that the effects of LM on broccoli yields tend to range from neutral to negative. Of the 12 previous broccoli LM studies that we are aware of, only one reported that LM increased mean broccoli head weight (Costello, 1994) and overall marketable yield, although not total yield (Costello and Altieri, 1994).

One study found a consistent reduction in broccoli yield resulting from LM (Chase and Mbuya, 2008), and the other studies reported mixed or no effects of LM on broccoli yields (Brainard and Bellinder, 2004; Ellis et al., 2000; Hooks and Johnson, 2001, 2004; Infante and Morse, 1996; Ponti et al., 2007; Theriault et al., 2009). In contrast, most of the studies reported beneficial effects of the LM on other aspects of agroecosystem performance, including suppression of weeds (Brainard and Bellinder, 2004; Chase and Mbuya, 2008; Infante and Morse, 1996) or insect pests (Bhan et al., 2010; Costello, 1994; Hooks and Johnson, 2001, 2004, 2006; Kloen and Altieri, 1990; Ponti et al., 2007). Given these beneficial effects, there is a clear need to identify factors that could improve broccoli yields in LM systems so as to make them more valuable to growers.

Previous broccoli LM studies have examined the effects of LM species, planting date, tillage system, and fertility source; however, we are not aware of any studies that have included multiple rates of fertilizer application as an explicit treatment factor as has been done with some other cruciferous crop LM systems (e.g., Brainard and Bellinder, 2004). Therefore, it remains unknown whether the effects of LM on broccoli yield depend on fertilizer rate and whether competition from the LM could be alleviated through fertility management. Additionally, no research has been conducted on broccoli LM systems under the soil and climate conditions specific to northern New England.

The objective of this study was to assess the performance of irrigated, summer-sown broccoli grown in plasticulture with and without LM under different rates of organic fertilizer application. We hypothesized that LM would reduce broccoli yield in the absence of supplemental fertilization and that competition from LM could be reduced or eliminated by adjusting fertility rates.

Materials and Methods

Site description. Two experiments were conducted at the University of New Hampshire (UNH) Woodman Horticultural Research Farm, in Durham, NH (lat. 43°15’ N, long. 70°49’ W) in 2011 (Expt. 1) and 2012 (Expt. 2). The two experiments were conducted in separate 0.045-ha fields to avoid additive effects of LM and broccoli-specific
pathogens. In both fields, the soil was a Charlton fine sandy loam (Coarse-loamy, mixed, active, mesic Typic Dystrudepts) (NRCS, Soil Survey Staff, 2013). The field used in 2011 had a history of mixed vegetable production and was planted to a cover crop of buckwheat (*Fagopyrum esculentum* Moench) before the initiation of the study. The field used in 2012 was adjacent to the first field, had a similar cropping history, and had been planted with a cover crop of winter rye (*Secale cereale* L.) before establishing the experiment.

**Experimental design and field management.** The experimental sites were rototilled in the spring, 7 (Expt. 1) and 14 (Expt. 2) d before establishment so as to account for N contributed by in situ soil organic matter and the previous season’s cover crop. In Expt. 1, the contribution of N from organic matter was estimated at 62 kg N/ha. Thus, to achieve a target 1X rate of 106 kg N/ha, we had to apply 84 kg N/ha in Expt. 1, but only 44 kg N/ha in Expt. 2 (Table 1). Both experiments included a 0X, 1X, and 1.5X rate, and Expt. 2 included two additional higher fertility rates (2X and 2.5X), whereas Expt. 1 included a 0.5X rate. The higher target fertility rates were added to Expt. 2 because of the higher in situ N levels in the field used for that experiment and were intended to achieve applied N rates that were comparable to and higher than those used in the 1.5X rate in Expt. 1 (Table 1).

Subplots were bed rows within each LM and BS whole plot and incorporated 17 and 14 broccoli plants in Expts. 1 and 2, respectively. Fertilizer rate treatments were randomly assigned to each subplot within each individual whole-plot replicate (Fig. 1). In both experiments, we used granular PRO-GRO (5N:3P:4K; North Country Organics, Bradford, VT) to create our fertilizer rate treatment levels. In Expt. 1, all subplots except those assigned to the 0X treatment

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Table 1. Fertility treatment levels in Expts. 1 and 2.

| Expt. | Treatment | Estimated soil in situ N | Applied N | Total N |
|-------|-----------|--------------------------|-----------|---------|
| 1     | 0X        | 22                       | 0         | 22      |
|       | 0.5X      | 22                       | 27        | 49      |
|       | 1.0X      | 22                       | 84        | 106     |
|       | 1.5X      | 22                       | 141       | 164     |
|       | 2.0X      | 62                       | 0         | 62      |
|       | 2.5X      | 62                       | 44        | 106     |
|       | 1.0X      | 62                       | 102       | 164     |
|       | 2.0X      | 62                       | 150       | 212     |
|       | 2.5X      | 62                       | 202       | 263     |

* Rates of applied N from commercial organic fertilizer necessary to achieve total N rates were based on estimated N available in in situ soil organic matter.

*N = nitrogen.*
received 24 kg N/ha equivalent of de-hulled soybean meal (6N:1P:1K; Blue Seal, Muscatine, IA), which was banded along bed rows before bed formation; the remainder of the fertility was side-dressed at each plant. Applications of PRO-GRO were evenly split between an application at the time of broccoli transplanting and a side-dressing application that occurred at 3 weeks after transplant in Expt. 1 and 4 weeks after transplant in Expt. 2. At the time of each application, PRO-GRO was applied to each plant individually by incorporating the fertilizer through the holes in the plastic mulch that were created at the time of transplanting. This approach, although time-consuming, and not practical for commercial production systems, ensured that our target rates were consistent and confined to the broccoli root zones.

Organic pesticides were used when necessary to control insect pests over the course of the experiments. Imported cabbage worm (Pieris rapae L.) caused minor damage to broccoli plants in both years and were treated with applications of Bacillus thuringiensis (DiPel; Valent BioSciences, Libertyville, IL). All bed rows were irrigated as needed when soil under the plastic became dry to the touch.

**Broccoli yield.** Broccoli yield was measured by harvesting mature heads by hand from the inner 15 (Expt. 1) and 12 (Expt. 2) plants in each subplot row. Broccoli were harvested at the ends of each subplot, served as buffers and were not measured. At each harvest, stems were trimmed to 2.5 cm below the bottom branch of the head and fresh weight was recorded for each individual plant. Floret size and density were evaluated to determine marketable maturity using a modified grading table (Sorenson and Grevesen, 1994; Theriault et al., 2009). Once the first heads reached a marketable stage, we harvested daily or every other day as broccoli matured and continued until all heads were harvested. The total number of marketable heads, excluding heads that failed to mature or exhibited defects severe enough to prevent marketability, was measured for each plot. Maturity rates were calculated for each harvested broccoli head based on the number of days after transplanting (DAT).

In addition to broccoli yield, we also measured incidence of hollow stem. Hollow stem is a common broccoli physiological disorder characterized by a gap that develops in the center of the stem, which may become discolored thereby reducing quality (Tremblay, 2009). We used a handheld chlorophyll meter (SPAD 502 Plus; Konica Minolta, Spectrum Technologies, Inc. Aurora, IL) to determine leaf chlorophyll values of broccoli in each treatment. SPAD values are useful for comparing relative differences in N status between treatments; however, without corresponding tissue analysis (which was not performed in this study), they cannot be used as an absolute proxy for N content. SPAD values were measured on 26 Aug. 2011 (52 DAT) in Expt. 1 by taking two measurements on the newest mature leaf from five individual plants in each subplot. In Expt. 2, SPAD measurements were taken on 12 July (36 DAT) and 17 Aug. (72 DAT) 2012 by taking six measurements on the fifth newest leaf on four individual plants in each subplot.

Weather data. For each growing season (6 July to 19 Sept. 2011 and 7 June to 16 Aug. 2012) were obtained from UNH’s weather station in Durham, NH (except for 3 to 25 Aug. 2011, which were obtained from Weather Underground Inc.). Temperature data were used to calculate growing degree-days (GDD) over the two seasons. The base temperature of 7.2 °C was selected because it was reported to be the least variable predictor of broccoli maturity rates (Dufault, 1997). Season GDD totals were obtained by summing the daily values, which were calculated using the following formula: GDD = (average daily temperature) – (base temperature 7.2 °C).

**Statistical analyses.** All yield analyses were conducted for each experiment separately by using a model appropriate for a split-plot experimental design. Yield and SPAD data were analyzed with analysis of variance (ANOVA) using a standard least squares regression model with blocking, LM treatment, and fertility as fixed effects. Also included were the interactions between blocking and LM (considered random) and LM and fertility (fixed). If significant effects were detected (P < 0.05), Tukey’s honestly significant difference test was used for mean comparisons (P < 0.05). We assessed the effects of the LM and fertility treatments on plot-to-plot uniformity of broccoli head weight by calculating the CV for each subplot (n = 4). The CV was calculated using the sd of mean marketable head weight within a subplot divided by the overall mean of the subplot. For hollow stem count data, we used a generalized linear model and a Poisson distribution to fit the main effects of blocking, LM, and fertility. Soil moisture data were analyzed with ANOVA for each time point by block and LM treatment.

Analyses were conducted with JMP® Pro Version 10.0.0 (SAS Institute Inc., Cary, NC). Data were checked to ensure that they met the assumptions of each test and no transformations were necessary.

**Results.** The mean daily temperature was 21 °C during each of the two experiments. The total number of GDD from transplant to final harvest was slightly higher in Expt. 1 than in Expt. 2 (+30 GDD) (Table 2). Cumulative precipitation between the dates of broccoli transplant to the final harvest was greater in Expt. 1, totaling 241 mm compared with 174 mm in Expt. 2.

**Broccoli yield and yield components.** Total marketable broccoli yields across all treatments averaged 3094 kg·ha⁻¹ in Expt. 1 and 2543 kg·ha⁻¹ in Expt. 2. In Expt. 1, there was a significant fertility effect and fertility × LM treatment interaction (fertility: F₁,₁₈ = 3.81, P = 0.028; interaction: F₃,₁₈ = 4.49, P = 0.016) with marketable yields converging at higher fertility levels in the LM and BS treatments. The effect of fertility on marketable yield was not linear; the highest yields were observed below the highest (1.5x) fertility level (Fig. 2). In Expt. 2, marketable yields were higher in BS compared with LM (F₃,₉₁ = 51.96, P = 0.006). The fertility treatment also affected yields (fertility: F₄,₂₄ = 8.98, P < 0.001); however, in contrast to Expt. 1, there was no interaction between fertility and LM (P = 0.237) (Fig. 2).

In Expt. 1, there was a LM × fertility interaction for the number of marketable broccoli heads (interaction: F₁,₁₈ = 3.26, P = 0.046) with LM having 21% fewer marketable heads compared with BS at 0x and 29% more than BS at 1.5x. In contrast, the number of marketable heads did not differ among treatments in Expt. 2 (P > 0.05).

Mean marketable fresh head weight was unaffected by LM in Expt. 1 (P = 0.123); however, head weights did increase with increasing fertility and were highest in 1.5x (fertility: F₁,₁₈ = 23.08, P < 0.0001). In Expt. 2, LM reduced mean head weight by an average of 46% compared with BS (F₁,₂₄ = 478.01, P = 0.0002). Similar to Expt. 1, head weights in Expt. 2 increased with increasing fertility rate (fertility: F₄,₂₄ = 11.15, P < 0.0001) (Fig. 3). There was no LM × fertility interaction effect on mean head weight in either experiment (P > 0.05).

The time until harvestable maturity was unaffected by LM in either experiment.
In Expts. 1 and 2, broccoli matured 6.3% and 3.8% more slowly, respectively, at the 0· rate compared with the other fertility rates. Interactions between LM and fertility were not significant in either experiment (P > 0.05).

Within-plot variability in mean marketable fresh head weight was unaffected by LM in either experiment (P > 0.05) but was affected by fertility (Expt. 1: F_{3,18} = 3.26, P = 0.047; Expt. 2: F_{4,24} = 5.10, P = 0.0041). In both experiments, head weights tended to be more variable (higher cv) at the highest rates of fertility. Interactions between LM and fertility were not significant in either experiment (P > 0.05).

In Expt. 1, higher frequencies of hollow stem were observed in BS (mean frequency 0.35 ± 0.12) compared with LM (mean frequency 0.04 ± 0.12) (χ^2 4.8, df = 1, P = 0.027). Very little hollow stem was observed in 2012 (less than 2.5% in BS plots and none in LM) and there were no significant differences between fertility treatments.

Resource availability. Given that bed rows were irrigated with drip tape, we did not expect our LM treatments to lead to moisture limitation. Indeed, for each measurement period, we observed few differences in soil moisture in either the bed row or between rows in LM and BS treatments or as a function of fertility rate in either experiment. In the few instances in which moisture differences did occur, soil moisture was actually higher in the LM treatments (Table 3).

The leaf chlorophyll index did not differ between treatments in Expt. 1 at 52 DAT. In Expt. 2, however, we observed higher SPAD values in the highest fertility levels compared with the lower fertility levels at 35 (F_{4,24} = 4.85,
Table 3. Soil moisture in living mulch (LM) and bare soil (BS) treatments measured between bed rows (interrow) or within bed rows (before irrigation) in Expt. 1 (2011) and Expt. 2 (2012).\(^{a}\)

| Date/position | Living mulch | Bare soil | \(P\) value |
|---------------|--------------|-----------|-------------|
| Interrow      |              |           |             |
| 29 July 2011  | 15.2         | 14.7      | 0.6251      |
| 1 Sept. 2011  | 16.8         | 14.1      | 0.0405      |
| 18 June 2012  | 15.5         | 12.0      | 0.0900      |
| 12 July 2012  | 12.6         | 13.2      | 0.6292      |
| 9 Aug. 2012   | 11.2         | 6.8       | 0.0029      |
| 20 Aug. 2012  | 18.8         | 11.6      | 0.0035      |
| Bed row       |              |           |             |
| 9 Aug. 2012   | 4.0          | 3.6       | 0.4001      |
| 20 Aug. 2012  | 8.7          | 7.0       | 0.1017      |

\(^{a}\)Data are means; \(n = 4\).

\(^{b}\)Percent volumetric water content.

Discussion

The hypothesis that LM reduces broccoli marketable yield in the absence of fertilizer addition (i.e., at the 0x fertilizer rate) was supported in both experiments (Figs. 2 and 3). These results are consistent with previous research demonstrating yield reductions in cash crops grown with living mulch intercrops, including in broccoli production systems (Chase and Mbuya, 2008). Not all studies have reported yield reductions resulting from LM, however, and this inconsistency in LM effects among studies likely reflects differences in experimental treatments, LM and cash crop species, and a myriad of other environmental and edaphic factors that vary across cropping systems and sites (Costello, 1994; Hartwig and Ammon, 2002; Kloe and Altieri, 1990; Zenenchik et al., 2000).

The hypothesis that the observed broccoli yield reductions resulting from LM were a consequence of competition for N was only partially supported by this study. Only Expt. 1 was consistent with broccoli yield reductions in LM being the result of N limitation, as evidenced by the fact that LM yield reductions were ameliorated by the addition of fertilizer N (Fig. 2). No such patterns were observed in Expt. 2, where LM reduced broccoli yield even at rates that were equivalent to applying 2.5x higher N than recommended (Howell and Hazzard, 2012; Vagen et al., 2004; Table 1). Interestingly, chlorophyll content of the broccoli leaves, which we used as a nondestructive measure of plant N status, differed between the LM and BS treatments only in Expt. 2 and only after harvest (i.e., 72 DAT), although a postharvest SPAD measurement was not taken in Expt. 1. Thus, our indirect measure of plant nutrient status did not reflect the patterns of yield reduction that we observed in the LM treatments in Expt. 1 or in Expt. 2 until after the final harvest had already occurred. Taken together, these results provide little support for the hypothesis that broccoli yield reductions resulting from LM can be attributed solely to competition for N or ameliorated by increasing organic fertilizer N rates. Possible explanations for this finding and the relative degree of support for each are discussed below.

The LM may have reduced the availability of resources other than N. In addition to soil N, a LM may compete with a cash crop for soil moisture, light, and other soil nutrients (Teasdale, 1996). In our experiments, we attempted to minimize the potential for water and light competition by growing the broccoli on plastic-covered raised beds that were irrigated and by mowing the LM periodically during the growing season. In contrast to what we would expect if the LM was competing for soil moisture, our periodic measurements of volumetric water content within the bed row and directly in the space between rows managed as either LM or BS indicated that the LM did not reduce soil moisture relative to the BS treatment (Table 3). The LM canopy was also separated from the broccoli canopy by a distance of \(\approx 0.25\) m (Fig. 1) and was elevated relative to the soil surface between rows. Additionally, the height of the LM was never allowed to exceed 20 cm so as to restrict the possibility of shading by the LM. Thus, it is unlikely that direct competition by the LM for water and/or light occurred to a degree necessary to explain the patterns observed in this study.

Although lower leaf chlorophyll concentrations are consistent with N limitation in broccoli (Bowen et al., 1999), in other crops they have also been associated with limitations in other essential soil nutrients such as phosphorus (P) (Sanchez-Rodriguez et al., 2013). Concentrations of P and other nutrients in the soil and broccoli tissue were not measured after treatment establishment; thus, we cannot discount the possibility that the LM reduced the availability of these essential soil nutrients.

The LM may have altered light quality reaching the broccoli, specifically the ratio of red to far-red radiation. Neighboring plants such as those grown as LM can illicit shade avoidance responses in crop plants, well before resource competition occurs (Ballare et al., 1990; Rajcan and Swanton, 2001). These shade avoidance responses, which occur when far-red (FR) light is reflected off the canopy of neighboring plants, may contribute to yield losses by reducing crop growth rate and allocation of resources to reproductive and other structures (Page et al., 2009). For example, Yang et al. (2014) found...
that soybean planted adjacent to corn was exposed to light with a lower red (R)/FR ratio than soybean grown in monoculture and this led to reduced soybean seedling above-ground biomass and total root biomass. In our study, the LM likely reflected light with a lower R/FR ratio signal than the bare soil, and this could have led to reduced broccoli yields in the LM treatment that may have been independent of or interacted with fertilizer rate. We did not measure the quality of light reaching the broccoli plants in the LM treatment and therefore cannot rule out this possibility as an explanation for our results. Given that the LM was spatially separated from the broccoli as a result of the raised nature of the beds and the use of plastic mulch (Fig. 1), the magnitude of yield loss attributable to the LM is difficult to explain without invoking some form of noncompetitive interference such as changes in reflected light quality.

Effects of LM and fertilizer on broccoli yield may be strongly context-dependent. We observed a LM × fertilizer interaction in Expt. 1 but not Expt. 2. This result is difficult to explain. Several factors differed between Expts. 1 and 2, including the cover crop that was present before the initiation of the study, LM seeding rate, quantity of in situ soil organic matter, broccoli transplanting date, and precipitation. Each of these factors could individually (Gaskell and Smith, 2007; Peek et al., 2012; Schonbeck et al., 1993; Sorensen and Greven, 1994; Tan et al., 2000) or in combination (Ateh and Doll, 1996; Brainard et al., 2012; Brainard and Bellinder, 2004; Ilinicki and Enache, 1992; Newenhouse and Dana, 1989; Paine et al., 1995; Teasdale and Daughtry, 1993). Reducing tillage can result in improved soil physical and biological properties (Franzleuebbers et al., 1999), possibly contributing to improvements in longer-term crop productivity. It is possible that improvements in soil quality associated with reducing tillage, coupled with organic inputs from the LM itself, may reduce the competitive effects often observed between LMs and cash crops over time (Smith et al., 2010).

Our study considered a specific LM, a mixture of Italian ryegrass and clover, and a specific commercial organic fertilizer; therefore, the results reported here may not extend to other species of LM, other LM cash crop systems, or other types of fertilizer. Additional research will be necessary to determine the longer-term effects of LM in broccoli production systems and the role that changes in reflected light quality may play in mediating broccoli yield responses in LM systems.

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