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Optimization of interstrand interactions enables burn detection with a collagen-mimetic peptide†

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Collagen is an abundant component of the extracellular matrix and connective tissues. Some collagen-mimetic peptides (CMPs) that do not form homotrimers can anneal to damaged tissue. Here, through a computational screen, we identify (flpHypGly) 7 as an optimal monomeric CMP for heterotrimer formation. We find that (flpHypGly) 7 forms stable triple helices with (ProProGly) but not with itself. The nonnatural amino acid HflpOH, which is (2S,4R)-4-fluoroproline, is not toxic to human fibroblasts or keratinocytes. Conjugation of (flpHypGly) 7 to a fluorescent dye enables the facile detection of burned collagenous tissue with high specificity. The ubiquity of collagen and the prevalence of injuries and diseases that disrupt endogenous collagen suggests widespread utility for this approach.

Introduction

Collagen—the basic building block of the extracellular matrix (ECM)—is the most abundant protein in vertebrates. Comprising a third of the total protein and three quarters of the dry weight of skin in humans, collagen is an essential component of all connective tissues. Collagen also plays a crucial role in maintaining the biological and structural integrity of the ECM. Its continual remodeling impacts and controls cell differentiation and proliferation, and atypical organization of the collagen matrix is a biomarker for various diseases, including fibrosis, connective tissue disorders, and cancer.3,7,11–14

Natural collagen is comprised of three left-handed, polyproline II-type (PPII) helices aligned in a parallel manner and coiled with each other to form a right-handed triple helix.1,8 Each of the peptide strands is constituted by a continuous repeat of Xaa-Yaa-Gly units. Even though the Xaa and Yaa positions can accommodate any amino acid, (2S)-proline (Pro; 28% abundance) and (2S,4R)-4-hydroxyproline (Hyp; 38% abundance) are the most prevalent, respectively. As a result, ProHypGly is the most common triplet in collagen (10.5%).9 These natural preferences at the Xaa and Yaa positions promote the conformational stability of the collagen triple helix.1

Collagen stability is governed by stereoelectronic effects that preorganize its prevalent pyrrolidine rings.1 A Cγ-endo and Cδ-exo pyrrolidine-ring pucker is preferred at the Xaa and Yaa position, respectively, in stable triple-helical structures. Substituents at the 4-position of a proline residue can enforce the preferred pyrrolidine-ring pucker via a gauche10 or steric11 effect, lowering the entropic cost for forming a triple helix and increasing its conformational stability (Table 1).3 For example, the stereospecific installation of a fluoro group generates a gauche effect that leads to a Cγ-endo pucker in (2S,4R)-4-fluoroproline (flp) and supports the Cδ-exo pucker in (2S,4R)-4-fluoroproline (Flp) to a greater extent than does the 4R-hydroxy group of Hyp.12–14 The impact of such substitutions are apparent in the thermostability of collagen-mimetic peptides (CMPs). The value of Tm, which is the temperature at the midpoint of the thermal transition between a triple helix and its component strands, tracks the strength of the inductive effect induced by the substitutions.15 For example, replacing Hyp with Flp at the Yaa position in (ProYaaGly) leads to ΔTm = 9 °C (Table 1).

Not all Xaa and Yaa pairings are compatible. The register of the strands in a triple helix is staggered such that each cross-section contains an Xaa, Yaa, and Gly residue.1 Even though
(flpProGly)7 and (ProFlpGly)7 form highly stable triple helices (Table 1).16 (flpFlpGly)7 is a monomer in solution due to unfavorable steric interactions between neighboring flp and Flp residues (Fig. 1).17 Still, mixtures of (flpFlpGly)7 and (ProProGly)7 produce stable triple helices because of the preorganization of (flpFlpGly)7 and the minimal steric clashes between flpFlp and neighboring Pro residues.17

Because (flpFlpGly)7 and (ProProGly)7 form only heterotrimeric triple helices,‡ they can anneal readily to exposed strands on denatured collagen (Fig. 2). Such annealing could allow for the development of diagnostic or therapeutic agents conjugated to a monomeric CMP.19–22 Previously, we have used CMP-conjugates to deliver fluorescent dyes,23 a sunscreen,24 and growth factors25 to damaged collagen. Although (flpFlpGly)7 seemed to outperform (ProProGly)7, as an annealing strand,23 its exceptional propensity was hindered by its poor solubility, which can complicate formulation and dosing, and diminish bioavailability.26 The highly soluble (ProHypGly)7 peptide can be successful in this context, but its homotrimerization necessitates a denaturation step prior to application,27,28 complicating its use. Another approach invokes protecting groups to deter triple-helix formation,29,30 but their removal introduces operational complexity.

Here, we conduct a computational survey of proline analogs to identify (XaaYaaGly)7 variants that are monomeric in solution, but have high ability to anneal with damaged collagen. We confirm the association characteristics of our design in vitro and demonstrate its specificity for damaged collagen by identifying a burned region on a rat-tail tendon ex vivo.

| ⋆Xaa···Yaa interaction energies (kcal mol⁻¹) of 4-substituted proline residues in a cross-section of a collagen triple helix§ |
|---|---|---|---|---|---|---|---|
| Xaa (R¹,R²) | Yaa (R³,R⁴) | Flp (F,H) | Mep (OMe,H) | Hyp (OH,H) | Mep (H,CH₃) | Clp (Cl,H) | Pro (H,H) |
| flp (H,F) | 2.8 | 1.6 | 3.8 | 1.1 | 5.6 | 1.3 |
| mep (CH₃,H) | −0.4 | −0.3 | −0.1 | −0.6 | 2.4 | −0.4 |
| Pro (H,H) | −0.1 | −0.6 | 0.3 | −0.2 | 1.2 | 0.0 |

| (XaaProGly)7 | (XaaProGly)10 |
|---|---|
| Tm (°C) | NH |
| [ProProGly]7 | 33 |
| [ProProGly]10 | 58 |
| ND |
| NH |

‡ Neither (flpFlpGly)7 nor (ProProGly)7 forms a stable homotrimeric triple helix, though for different reasons. (flpFlpGly)7 strands are highly preorganized for triple-helix formation, but experience strong internstrand FLF clashes in a triple helix.17 (ProProGly)7 strands do not engender steric clashes upon association, but are not especially well-organized for triple-helix formation.18

Results and discussion

(flpHypGly)7 is an optimal CMP

An optimal CMP is a highly soluble monomer under physiological conditions, but forms strong triple helices with natural collagen strands. To search for such (XaaYaaGly)7 variants, we defined two sets of proline analogs that are compatible with the steric and electronic requirements at the Xaa and Yaa positions (Table 1).† The resulting 18 combinatorial Xaa···Yaa pairs represent interactions between Xaa and Yaa residues on the neighboring strands of an (XaaYaaGly)7 homotrimer. To determine the extent of steric repulsion in each homotrimer, we applied molecular mechanics calculations to models derived from a high-resolution structure of a (ProProGly)10.
triple helix (PDB entry 1kf6). Briefly, Xaa and Yaa residues were positioned on neighboring positions on the triple helix. Following energy minimization, the interaction between the Xaa⋯Yaa pair was calculated as the energy difference between the homotrimeric helix and the free strands. Three known examples of monomeric CMPs—(ProProGly)₇, (flp Flp Gly)₇, and (clp Clp Gly)₇—served as experimental benchmarks for data analysis. Interaction energies (IE) revealed that four combinations—Xaa flp⋯Flp, flp⋯Mop, flp⋯Hyp, and flp⋯Clp—engender unfavorable interactions (Table 1). Among these four, a flp⋯Flp pair of Xaa⋯Yaa residues (IE = 2.8 kcal mol⁻¹) merits special consideration. flp in the Xaa position and Flp in the Yaa position are individually the most stabilizing 4-substituted proline analogs. Yet, (flp Flp Gly)₇ does not form a stable triple helix, suggesting that Xaa⋯Yaa clashes of IE ≥ 2.8 kcal mol⁻¹ are likely to produce monomeric peptides. Peptides that satisfied this criterion include (flp Mop Gly)₇, (flp Hyp Gly)₇, and (flp Clp Gly)₇ (IE = 3.4, 3.0, and 5.6 kcal mol⁻¹, respectively). The higher solubility bestowed by Hyp (hydroxy group) compared to Mop (methoxy group) or Clp (chloro group) led to explore (flp Hyp Gly)₇ experimentally. Notably, a stereospecific H→F substitution is the only difference between a flp-Hyp-Gly unit and the isologous Pro-Hyp-Gly unit of natural collagen. Moreover, deleterious O)(F steric clashes engendered by (flp Hyp Gly)₇ will not preclude annealing to strands of human collagen, which lack flp entirely and have Hyp in only 38% of its Xaa-Yaa-Gly units. Finally, the HflpOH monomer is readily accessible from natural HHypOH by an SN₂ reaction. We note that CMPs containing flp-Hyp-Gly units have been examined previously, though in contexts distinct from those described herein. For example, (flp Hyp Gly)₉ was found to form stable triple helices.

§ We also tested clp (R¹ = H, R² = Cl) as a candidate at the Xaa position. Its use, however, produces extremely unfavorable interactions, even when its neighbor is Pro (IE = 13.8 kcal mol⁻¹). This incompatibility is consistent with the low thermostability of triple helices with clp Pro Gly units, and disqualifies the use of clp at the Xaa position of an annealing strand.

¶ Log P values are HHypOH, −2.64; HMopOH, −2.02; HProOH, −1.72; HFlpOH, −1.71; and HClpOH, −1.40; as calculated with software from Molinspiration (Slovenský Grob, Slovak Republic).
a stable triple helix at ambient temperature, complicating its use in biological contexts.\textsuperscript{27} Hence, we examined a shorter peptide: (flpHypGly)$_7$.

**(flpHypGly)$_7$ is structured only as a heterotrimer**

The (flpHypGly)$_7$ peptide and its variants were synthesized by microwave-assisted solid-phase peptide synthesis and purified by reversed-phase HPLC. Samples prepared in 50 mM HOAc were heated to 65 °C, cooled to 4 °C, and equilibrated at 4 °C for ≥24 h prior to analysis. The structure of their peptides and their mixtures was assessed with circular dichroism (CD) spectroscopy, following the characteristic CD signature for the collagen triple helix and PPII helix at ~225 nm.

CD spectra for (flpHypGly)$_7$ and (ProProGly)$_7$ reveal shallow peaks near 225 nm (Fig. 3A). These samples exhibit no cooperative denaturation with increasing temperature, indicating the absence of triple helices in solution (Fig. 3B). This behavior is typical for PPII structures that do not associate into triple helices.\textsuperscript{36-38} In marked contrast, a 1:2 (flpHypGly)$_7$/ (ProProGly)$_7$ solution exhibits a strong collagen signature in its CD spectrum (Fig. 3A), as well as cooperative denaturation with $T_m = 28 \pm 2$ °C (Fig. 3B and S2A). A 2:1 (flpHypGly)$_7$/ (ProProGly)$_7$ solution exhibits similar thermostability (Fig. S1A, S1B, and S2B†). The results of these mixing experiments indicate that (flpHypGly)$_7$ can form a stable triple helix with (ProProGly)$_7$ strands, but not with itself.

**(HypflpGly)$_7$ does not form a triple helix**

Next, we sought to use a (flpHypGly)$_7$–dye conjugate in a physiological context, but we needed a negative control. A free (that is, unconjugated) dye could serve as a negative control for such experiments. The disparity between the physicochemical attributes of a CMP–dye conjugate and the dye alone could, however, compromise data-interpretation.

To mimic the attributes of the (flpHypGly)$_7$ peptide, we synthesized (HypflpGly)$_7$, which is a compositional isomer (CI). Swapping the flp and Hyp residues generates a sequence that violates the conformational restrictions of the Xaa and Yaa positions (Table 1), and should preclude triple-helix formation.\textsuperscript{12,39} Indeed, the CD spectrum of (HypflpGly)$_7$ mimics closely that of (flpHypGly)$_7$ (Fig. 3C), and does not exhibit the cooperative transition of triple-helix denaturation (Fig. 3D). Further, the CD spectrum of a 1:2 or 2:1 (HypflpGly)$_7$/ (ProProGly)$_7$ mixture is a linear combination of the individual (ProProGly)$_7$ and (HypflpGly)$_7$ spectra and exhibits no evidence of a triple helix (Fig. 3C and S1C†). These spectra, together with the absence of a cooperative transition (Fig. 3D and S1D†), provide strong evidence that (HypflpGly)$_7$ does not interact with (ProProGly)$_7$. Because (HypflpGly)$_7$ has the same composition as does (flpHypGly)$_7$, but lacks its annealing properties, (HypflpGly)$_7$ serves as an ideal negative control for our physiological experiments.

**HflpOH is not cytotoxic**

The proteolysis of (flpHypGly)$_7$ would generate three amino acids: HflpOH, HHypOH, and HGlyOH. Two of these amino acids, HHypOH and HGlyOH, are prevalent in human collagen; HflpOH, though, is nonnatural. As 1/5 of FDA-approved drugs contain fluorine,\textsuperscript{40} we expected HflpOH to be well-tolerated in humans. Nevertheless, HflpOH itself is not a component of any approved drug. Accordingly, we tested the tolerance of two relevant cell types for this amino acid: human fibroblasts and primary human epidermal keratinocytes. Fibroblasts are the most common cells in human connective tissue, and maintain its health and integrity. Keratinocytes constitute 90% of the cells in the epidermis, which is the barrier against environmental damage. We found that HflpOH is not detectably toxic to either fibroblasts or keratinocytes at concentrations up to 1 mM (Fig. 4).

**(flpHypGly)$_7$ anneals to damaged collagen ex vivo**

To evaluate the binding of a (flpHypGly)$_7$–dye conjugate to fibrillar collagen, we used the rat-tail tendon as a testbed, as it is comprised primarily of fibrillar collagen.\textsuperscript{41} Type I collagen from rat-tail tendons has been studied extensively, and is accessible by well-developed procedures.\textsuperscript{42} For the treatment of rat-tail tendon, we prepared variants of (flpHypGly)$_7$ and (HypflpGly)$_7$ that are conjugated via their N-terminal amino group to a cyanine 5 (Cy5) dye, obtaining Cy5CMP and Cy5CI, respectively (Fig. 5A). We choose Cy5 because of the far-red wavelength of its fluorescence excitation ($\lambda_{\text{max}} = 652$ nm) and emission ($\lambda_{\text{em}} = 672$ nm), which allows for detection in tissue, as well as its high brightness ($\varepsilon \times \Phi = 0.18$).\textsuperscript{43} To test whether...
(flpHypGly)$_7$ can distinguish damaged collagen from healthy collagen, we used a 350 °C soldering iron to burn a rat-tail tendon through glass (Fig. 5B). We then treated the tendon with Cy5CMP and Cy5CI, incubated for 1 h at 37 °C, and washed extensively with PBS.

Second harmonic generation (SHG) microscopy is a nonlinear optical method that is used extensively to identify and visualize fibrillar collagens in tissues.$^{45-47}$ Using a multiphoton laser-scanning microscope, data can be acquired from both SHG and Cy5 channels, revealing the distinction between damaged and healthy collagen (SHG) and its detection with our probe (Cy5). The fibrils exposed to 350 °C show little or no signal through the SHG channel, confirming the absence of intact collagen structure in the burn area (Fig. S3†). Upon treatment with Cy5CMP, the regions that fail to show a strong SHG signal exhibit a strong Cy5 fluorescence (Fig. 5C), indicating the association Cy5CMP with damaged collagen. Importantly, the same intensity is not observed in intact regions. In contrast, the distinction between damaged and healthy collagen is not apparent with the negative control, Cy5CI (Fig. 5D).

Quantification of Cy5 fluorescence reveals that the localization of Cy5CMP on burned collagen is greater by 25-fold (i.e., 148/6) than its fluorescence on intact collagen (Fig. 5E). Similarly, the labeling of these regions by Cy5CMP is greater by 5.3-fold (i.e., 148/28) than that by the Cy5CI peptide. These results confirm annealing by (flpHypGly)$_7$ to regions of collagen damage. Finally, we note that Cy5CMP is highly soluble in water, showing no evidence of precipitation despite its pendant hydrophobic dye.

**Conclusions**

An optimized CMP can act as a vehicle for the detection of mammalian collagen that has suffered burn damage. Burn injuries affect more than 7 million individuals worldwide and result in significant morbidity and mortality.$^{49}$ Current methods to diagnose burns face challenges, such as the disparity between the clinical identification of burn depth and the histopathologic determination of cellular and ECM damage.$^{50}$ Further, only 70% of visual assessments of full-thickness burn injuries are accurate.$^{51}$ Our approach could enable the reliable detection of thermally damaged collagen in a clinical setting. The ubiquity of collagen as well as the disruptive effects of fibrosis, connective-tissue disorders, and cancer on collagen in the ECM suggest its utility in other regimens as well.
Conflicts of interest

There are no conflicts to declare.

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