Combination of automated brain volumetry on MRI and quantitative tau deposition on THK-5351 PET to support diagnosis of Alzheimer’s disease

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Imaging biomarkers support the diagnosis of Alzheimer’s disease (AD). We aimed to determine whether combining automated brain volumetry on MRI and quantitative measurement of tau deposition on [18F] THK-5351 PET can aid discrimination of AD spectrum. From a prospective database in an IRB-approved multicenter study (NCT02656498), 113 subjects (32 healthy control, 55 mild cognitive impairment, and 26 Alzheimer disease) with baseline structural MRI and [18F] THK-5351 PET were included. Cortical volumes were quantified from FDA-approved software for automated volumetric MRI analysis (NeuroQuant). Standardized uptake value ratio (SUVR) was calculated from tau PET images for 6 composite FreeSurfer-derived regions-of-interests approximating in vivo Braak stage (Braak ROIs). On volumetric MRI analysis, stepwise logistic regression analyses identified the cingulate isthmus and inferior parietal lobule as significant regions in discriminating AD from HC and MCI. The combined model incorporating automated volumes of selected brain regions on MRI (cingulate isthmus, inferior parietal lobule, hippocampus) and SUVRs of Braak ROIs on [18F] THK-5351 PET showed higher performance than SUVRs of Braak ROIs on [18F] THK-5351 PET in discriminating AD from HC (0.98 vs 0.88, \( P = 0.033 \)) but not in discriminating AD from MCI (0.85 vs 0.79, \( P = 0.178 \)). The combined model showed comparable performance to automated volumes of selected brain regions on MRI in discriminating AD from HC (0.98 vs 0.94, \( P = 0.094 \)) and MCI (0.85 vs 0.78; \( P = 0.065 \)).

Abbreviations
AD  Alzheimer’s disease
HC  Healthy control
MCI  Mild cognitive impairment
AUROC  Area under the receiver operating characteristics
MAO-B  Monoamine oxidase B
SUVRs  Standard uptake value ratios
ANOVA  Analysis of variance
FDR  False discovery rate

Imaging biomarkers play an important role in supporting the diagnosis of Alzheimer’s disease (AD), not only in the research field but also in clinical practice. With the advent of amyloid and tau PET ligands, there has been a huge advance in the understanding of pathophysiologic mechanism of AD, and early diagnosis of AD can be made even in the preclinical or prodromal stage⁴. Amyloid and tau PET enable in vivo visualization of the

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underlying pathophysiologic culprits of AD, namely senile plaques and neurofibrillary tangles, but their clinical use may be affected by its availability, cost and consideration of ionizing radiation exposure. MRI is used for standard practice to support the diagnosis of AD and to exclude other causes of cognitive impairment such as stroke, vascular dementia, normal pressure hydrocephalus, or inflammatory or neoplastic conditions. Atrophy of the medial temporal lobe, particularly the hippocampus and entorhinal cortex, was shown to be important in patients on the AD spectrum and these regions were identified as predictors of time to progression from mild cognitive impairment (MCI) to AD. Qualitative visual assessment of atrophy on MRI suffers from poor inter-observer agreement, which is a major limitation for implementing it in a clinical setting. This limitation can be addressed by quantitative volumetry analysis, and automated software programs for quantitative volumetric analysis have been developed. Extensive validation of these software packages has been done for clinical use with comparison to manually segmented volumetric measurements, and commercially available automated brain volumetric tool showed high correlation in brain volume measurement with tools extensively used in research settings with moderate to high sensitivity (63.3–83%) and high specificity (93–100%) in differentiating AD from healthy controls (HC).

[18F] THK-5351 is one of the first generation of tau PET tracers and a close correspondence between the Braak staging of tau pathology and retention of the first generation tau PET tracers has been demonstrated in AD. Braak stages of tau pathology, derived from cross-sectional data, proposed how AD-related tau pathology begin in medial temporal structures extending to limbic areas, posterior cingulate cortex and then widely to isocortical brain areas. While tracer specificity related to off-target binding of [18F] THK5351 to monoamine oxidase B (MAO-B) remains as a possible limitation, previous studies reported that binding of [18F] THK5351 in the hippocampus was not influenced by off-target binding in the choroid plexus unlike [18F] AV-1451. Moreover, longitudinal analysis of tau PET demonstrated that measurement of changes in the tau PET SUVR can be used as an efficient outcome measure in disease modifying clinical trials.

While tau PET reflects the underlying pathophysiologic hallmark of AD, MRI demonstrates regional atrophy with automated brain volumetry extensively validated for clinical use in AD. To our knowledge, no direct comparison has been performed between commercially available automated brain volumetry on MRI and [18F] THK-5351 PET in discrimination of AD spectrum. The purpose of this study was to compare commercially available automated brain volumetry on MRI and [18F] THK-5351 PET, and determine if the combination of these tools can aid discrimination of AD spectrum in a clinically feasible setting.

Methods
All the experimental protocols were approved by institutional review boards of Asan Medical Center and Samsung Medical Center, Seoul, South Korea. All relevant study protocols for involving humans were in accordance with guidelines of institutional ethics committees.

Participants. From a prospective cohort of a multicenter clinical trial (NCT02656498), 113 participants who had both baseline structural MRI and [18F] THK-5351 PET were included. All participants or their appropriate representatives provided informed consent, and all participants were examined under the protocols approved by the institutional review board of the two tertiary medical centers (Asan Medical Center and Samsung Medical Center). A flow diagram of participant inclusion is shown in Fig. 1. Participants with MCI and AD underwent structural MRI and [18F] THK-5351 PET at the time of initial diagnosis and the maximum time interval between the structural MRI and [18F] THK-5351 PET was 7 months. All participants underwent standard dementia screening, including recording of medical history, physical examination, brain MRI, [18F]...
THK-5351 PET, and neuropsychological testing. Clinical diagnosis was made according to standard research criteria for probable AD and MCI, applied across a multidisciplinary team. Participants assigned as HC had a Korean version of Mini-Mental State Examination (K-MMSE) score > 1.5 standard deviation of the normative means and without current clinical depression (Geriatric Depression Scale < 11). Participants were excluded if they had a history of stroke, head trauma, or alcoholism. The 113 finally enrolled participants included 32 HC, 55 participants with MCI, and 26 participants with AD. The demographics of the study participants are provided in Table 1.

### MRI acquisition.
A 3.0-T system MRI (Achieva; Philips Medical Systems, Best, The Netherlands) with an eight-channel sensitivity-encoding head coil was used. A high-resolution anatomical three-dimensional (3D) volume image was obtained using a 3D gradient-echo T1-weighted sequence. The following parameters were used: repetition time/echo time, 9.9/4.6 ms; flip angle, 8°; field of view, 224 × 224 mm; matrix, 224 × 224; and slice thickness, 1 mm with no gap.

### [18F] THK-5351 PET imaging acquisition.
All PET images were acquired using Discovery 690, 710, and 690 Elite PET/CT scanners (GE Healthcare; Milwaukee, WI, USA) at Asan Medical Center and a Discovery STE PET/CT scanner (GE Healthcare) at Samsung Medical Center. The identical imaging and reconstruction protocol was used in both centers. [18F] THK-5351 PET images were obtained for 20 min, starting 50 min after injection of 185 ± 18.5 MBq of [18F] THK-5351, which binds to the aggregated tau in paired helical filaments.

Prior to the PET scan, a head holder was applied to minimize head motion and brain CT images were obtained for attenuation correction. Using the ordered-subsets expectation maximization algorithm (iteration = 4, subset = 16), 3D PET images were reconstructed with a voxel size of 2.0 × 2.0 × 3.27 mm³. To increase data uniformity between different PET scanners, a 3D Hoffman phantom-based PET harmonization method was applied based on the results of correction factors obtained from phantom studies using 3D Hoffman brain phantom.

### Quantitative image analysis.
**MRI volumetry.** Automated MRI-volumetry analyses using NeuroQuant software package (CorTechs Labs, La Jolla, CA, USA) were performed via the standard processing pipeline. The details of this procedure are previously described elsewhere. DICOM files were uploaded to the servers for processing. Briefly, the protocol entails a quality check, adjustment for gradient non-linearity/B1 field inhomogeneity, and skull stripping followed by a discrete cosine transformation and registration onto a dynamic probabilistic atlas. An anatomic label is designated to each voxel based on approximations from the dynamic probabilistic atlas that is structure-wise similar to the FreeSurfer but uses an independent code-based method for intensity normalization and gradient distortion correction to accommodate scanner-specific acquisition-level differences and to better represent the aged population. The MRI analysis provides a report that includes raw and corrected volumes (% intracranial volume) for 66 brain regions. All reports were reviewed to ensure adequate quality.

**Tau PET.** [18F] THK-5351 PET images were segmented by using the default automated gyral-based parcelation method of FreeSurfer (version 5.3.0; http://surfer.nmr.mgh.harvard.edu)~19,20. The SUVr of the cerebral cortex was calculated using the SUV of the cerebellar cortex as a reference region. Bilateral Braak stage region of interest (ROI) were created by combining FreeSurfer ROIs into non-overlapping Braak regions, using the Desikan-Killiany cortical atlas. Further analyses from the automatic FreeSurfer segmentation were utilized for the left and right hippocampus. The values for the left and right hemispheres were then averaged.

The ROIs of the in vivo Braak staging were expressed as Braak I/II, Braak III/IV and Braak V/VI, reflecting the propagation of tau pathology, which begins in the medial temporal structures, extends to limbic areas and then widely spreads into isocortical brain areas. The mean SUVr of ROIs belonging to Braak I/II, Braak III/IV and Braak V/VI were calculated and used for the analysis. Braak I/II include entorhinal cortex and hippocampus. Braak III/IV include parahippocampal, fusiform, lingual, middle temporal, caudal anterior cingulate, rostral anterior cingulate, posterior cingulate, cingulate isthmus, insula, inferior temporal and temporal pole. Braak V/VI include superior frontal, lateral orbitofrontal, medial orbitofrontal, frontal pole, caudal middle frontal, superior parietal and precuneus.

### Table 1. Demographics of the study participants. HC, healthy controls; MCI, mild cognitive impairment; AD, Alzheimer disease; K-MMSE, Korean version of the Mini-Mental State Examination. *0/1/2 indicates non-carriers/heterozygotes/homozygotes for APOE ε4 respectively.

|                  | HC     | MCI    | AD     | P value |
|------------------|--------|--------|--------|---------|
| N                | 32     | 55     | 26     |         |
| Age (years)      | 69.4 ± 6.0 | 70.8 ± 7.1 | 63.6 ± 10.6 | 0.001   |
| Sex (M/F)        | 11/21  | 20/35  | 11/15  | 0.816   |
| Education (years)| 10.6 ± 4.9 | 11.1 ± 4.6 | 10.6 ± 4.7 | 0.846   |
| APOE ε4 (0/1/2)  | (24/7/0) (NA 1) | (28/17/5) (NA 5) | (14/6/2) (NA 4) | 0.082   |
| K-MMSE score     | 28.6 ± 1.2 | 25.0 ± 3.2 | 19.9 ± 5.0 | <0.001  |
| Days between MRI and [18F] THK-5351 scan | 47.5 ± 35.6 | 57.1 ± 50.1 | 53.6 ± 47.0 | 0.659   |
rostral middle frontal, pars opercularis, pars orbitalis, pars triangularis, lateral occipital, supramarginal, inferior parietal, superior parietal, precuneus, superior temporal, transverse temporal, pericalcarine, postcentral, cuneus, precentral and paracentral. We excluded basal ganglia and thalamus from the Braak ROIs as these regions are likely to reflect extensive off-target binding.\(^4\)

**Statistical analysis.** Comparison of demographics and quantitative values. The clinical characteristics of the participants in each group (HC, MCI, and AD) were compared. Chi-square tests were used for categorical variables and analysis of variance (ANOVA) was used for continuous variables with post hoc pairwise comparisons where relevant.

The cortical volumes and SUVRs of Braak ROIs were subsequently compared between groups using ANOVA, and baseline age was used as a covariate. Post-hoc analysis between groups was also conducted. In order to account for multiple comparisons across the multiple cortical regions, false discovery rate (FDR)-corrected \(P\)-value < 0.05 was used. \(P\) values for SUVRs were Bonferroni-corrected. Normality of all ROIs and Braak staging was verified using Kolmogorov-Smirnov and Shapiro–Wilk tests.

Construction of a diagnostic model. In order to identify significant brain regions in differentiating AD from HC and MCI, univariate and multivariate logistic regression analysis were performed using a general linear model with baseline age as a covariate. Stepwise logistic regression analyses were performed, and regions identified as significant regions upon univariate analysis underwent multivariate fitting to determine the best combination of regions for discriminating AD from HC and MCI. Variance inflation factor was considered to check for multicollinearity of the variables included in the regression equations and variables with variance inflation factor greater than 5 were excluded. The volume of the hippocampus was included as selected brain regions as the hippocampus is a well-established brain region that is known to be affected in AD and its atrophy is one of the most commonly used imaging criteria used for diagnosing AD.\(^2,\,6,\,43\). The combined model was constructed using volumes of selected brain regions and SUVRs of Braak ROIs.

Performance measurement. Performance of discriminating AD from HC and MCI was quantified by area under the receiver operating characteristics (AUROC). The performances of selected brain regions, SUVRs of Braak ROIs and a combined model were calculated. The performance of the hippocampus alone was also evaluated.

Additional analysis. In order to provide the same ROI categorization for automated volumes on MRI and SUVRs on [\(^{18}\)F] THK-5351 PET, automated volumes on MRI were grouped into Braak I/II, Braak III/IV and Braak V/VI as previously described, and univariate and multivariate logistic regression analysis on SUVRs from [\(^{18}\)F] THK-5351 PET was performed as previously described. Performance of automated volumes on MRI grouped into Braak ROIs and SUVR of selected brain region was also evaluated.

Statistical analyses were performed using R statistical software (version 3.3.3, R Core Team, Vienna, Austria). Figure 2 shows the overall process for image and statistical analysis.

**Results**

Participants. The demographics of the study participants are provided in Table 1. Of note, the mean age in the AD group was lower than in the HC and MCI groups \((P = 0.001)\). The sex ratio, education level, and APOE allele variations were comparable across the three groups. In particular, there was no difference in the proportions of participants who are either homozygotes or heterozygotes for APOE allele variations between the AD and MCI groups \([31\% (8/26) \text{ vs } 40\% (22/55); P = 0.437]\). As expected, the K-MMSE score was lower in the AD group than in the other groups \((P < 0.001)\). The mean interval between [\(^{18}\)F] THK-5351 PET and MRI was 53.6 days (range, 0–202 days; standard deviation, 47 days) with no significant difference among the three groups.

**Comparison of automated cortical volumes and SUVRs of Braak ROIs in AD vs. HC and AD vs. MCI.** Automated volumes on MRI were compared and post-hoc analysis was performed between groups (Supplementary Table 1). Volumes of the following brain regions were different amongst the AD, MCI and HC groups on the ANOVA analysis: hippocampus \((P = 0.015)\), entorhinal cortex \((P = 0.001)\), superior temporal \((P = 0.008)\), middle temporal \((P = 0.001)\), basal ganglia \((P = 0.011)\), amygdala \((P = 0.011)\), anterior cingulate \((P = 0.002)\), posterior cingulate \((P = < 0.001)\), cingulate isthmus \((P = < 0.001)\), anterior middle frontal \((P = 0.004)\), inferior parietal lobule \((P = < 0.001)\) and supramarginal \((P = < 0.001)\). Upon post-hoc analysis, the AD group showed lower volume compared to the HC group in the hippocampus \((P = < 0.001)\), entorhinal cortex \((P = 0.001)\), superior temporal \((P = < 0.001)\), middle temporal \((P = < 0.001)\), basal ganglia \((P = < 0.001)\), amygdala, \((P = < 0.001)\), anterior cingulate \((P = 0.006)\), posterior cingulate \((P = 0.041)\), cingulate isthmus \((P = 0.001)\), anterior middle frontal \((P = < 0.001)\), inferior parietal lobule \((P = < 0.001)\) and supramarginal \((P = < 0.001)\). Compared to the MCI group, the AD group showed lower volume in the hippocampus \((P = 0.037)\), entorhinal cortex \((P = 0.001)\), superior temporal \((P = 0.001)\), middle temporal \((P = < 0.001)\), basal ganglia \((P = 0.006)\), amygdala \((P = 0.003)\), anterior cingulate \((P = 0.013)\), posterior cingulate \((P = 0.011)\), cingulate isthmus \((P = 0.006)\), anterior middle frontal \((P = 0.007)\), inferior parietal lobule \((P = 0.005)\) and supramarginal \((P = 0.021)\).

SUVRs of Braak ROIs were also compared and post-hoc analysis was performed between groups (Supplementary Table 1). The SUVRs were different amongst the HC, MCI and AD groups in Braak I/II \((P = 0.011)\), Braak III/IV \((P < 0.001)\) and Braak V/VI \((P < 0.001)\). SUVRs of Braak V/VI was different in both AD vs. HC \((P = < 0.001)\)
and AD vs. MCI ($P = 0.002$). SUVRs of Braak III/IV was different in AD vs. HC ($P < 0.001$) but not in AD vs. MCI ($P = 0.231$). SUVRs of Braak I/II was different in MCI vs. HC ($P = 0.031$).

**Significant brain regions among automated volumes on MRI for discriminating AD from HC and MCI.** For discriminating AD from HC, the univariate analysis (Table 2) identified 9 brain regions (hippocampus, $P < 0.001$; entorhinal cortex, $P = 0.012$; middle temporal, $P = 0.002$; anterior cingulate, $P = 0.013$; posterior cingulate, $P = 0.006$; cingulate isthmus, $P < 0.001$; anterior middle frontal, $P = 0.011$; inferior parietal lobule, $P < 0.001$; supramarginal, $P < 0.001$) as significant regions. Upon the multivariate analysis (Table 2), the cingulate isthmus ($P = 0.009$) and inferior parietal lobule ($P = 0.022$) were identified as significant regions for discriminating AD from HC.

For discriminating AD from MCI, the univariate analysis (Table 2) identified 4 brain regions (anterior cingulate, $P = 0.006$; posterior cingulate, $P = 0.001$; cingulate isthmus, $P = 0.002$; inferior parietal lobule, $P = 0.001$) as significant regions. Upon the multivariate analysis (Table 2), the cingulate isthmus ($P = 0.041$) and inferior parietal lobule ($P = 0.012$) were identified as significant regions for discriminating AD from MCI.

**Combined and individual performance of volumes of selected brain regions, SUVRs of Braak ROIs and volume of the hippocampus.** Table 3 compares the combined and individual performance of volumes of selected brain regions (cingulate isthmus, inferior parietal lobule, hippocampus), SUVRs of Braak ROIs and volume of the hippocampus in discriminating AD from HC and MCI. In discriminating AD from HC, the combined model incorporating volumes of selected brain regions and SUVRs of Braak ROIs (AUROC 0.98, 0.95–1.00) showed comparable performance to volumes of selected brain regions (AUROC 0.94, 0.88–1.00; $P = 0.094$) but higher than SUVRs of Braak ROIs (AUROC 0.88, 0.79–0.98; $P = 0.033$) and volume of the hippocampus (AUROC 0.78, 0.65–0.91; $P = 0.003$).

In discriminating AD from MCI, the combined model incorporating volumes of selected brain regions and SUVRs of Braak ROIs (AUROC 0.85, 0.75–0.94) showed comparable performance to volumes of selected brain regions (AUROC 0.78, 0.68–0.89; $P = 0.065$) and SUVRs of Braak ROIs (AUROC 0.79, 0.62–0.92; $P = 0.178$). The combined model showed higher performance than volume of the hippocampus (AUROC 0.48, 0.34–0.63;
P < 0.001). Figure 3 shows AUROC of the combined model and individual parameters in discriminating AD from HC and MCI.

**Additional analysis.** The results of univariate and multivariate analysis of SUVRs from [18F] THK-5351 PET are provided in Supplementary Table 2. The supramarginal gyrus was identified as the significant region for discriminating AD from HC and MCI. The performance of SUVR of selected brain region and automated volumes on MRI grouped into Braak ROIs is shown in Supplementary Table 3.

**Discussion**

In this study, performances of automated brain volumetry on MRI and quantitative measurement of tau deposition on [18F] THK-5351 PET were evaluated in discriminating AD on the AD spectrum. The combined model incorporating automated volumes of selected brain regions on MRI (cingulate isthmus, inferior parietal lobule, hippocampus) and SUVRs of Braak ROIs on [18F] THK-5351 PET showed higher performance than SUVRs...
of Braak ROIs on [18F] THK-5351 PET in discriminating AD from HC (0.98 vs 0.88, $P = 0.033$) but not in discriminating AD from MCI (0.85 vs 0.79, $P = 0.178$). The combined model showed comparable performance to automated volumes of selected brain regions on MRI in discriminating AD from HC (0.98 vs 0.94, $P = 0.094$) and MCI (0.85 vs 0.78, $P = 0.065$). The combined model showed higher performance than volume of the hippocampus in discriminating AD from HC (0.98 vs 0.78; $P = 0.003$) and MCI (0.85 vs 0.48; $P < 0.001$). This underscores the robust performance of automated brain volumetry on MRI and quantitative measurement of tau deposition on [18F] THK-5351 PET in discrimination of AD spectrum with added value of automated brain volumetry in discriminating AD from HC.

While tau PET reflects the underlying pathophysiologic mechanism of AD, MRI demonstrates regional atrophy for which automatic volumetric assessment tool is available. In our study, commercially available, FDA-approved automated brain volumetry tool was evaluated that is being increasingly used in clinical settings as it is easily accessible providing fully automated service in a clinically acceptable time-period and feasible setting. Diagnostic efficacy of the automated volumetric assessment tool has previously been demonstrated in patients on the AD spectrum with high sensitivity (63.3–83%) and high specificity (93–100%) in differentiating AD from HC, which is comparable to the performance demonstrated in our study (AUROC, 0.94; sensitivity, 88.5%; specificity, 90.6%). Previous studies used volume of the medial temporal lobe in the discrimination of AD while our study identified volumes of the cingulate isthmus and inferior parietal lobule by performing stepwise logistic regression analyses. The value of automated volumetric measurement of both the cingulate isthmus and inferior parietal lobule has been previously reported, and the combination of automated volumetry of the entorhinal cortex and inferior parietal lobule was the best predictor of time to progress from MCI to AD. Preferential atrophy of specific laminae in the inferior parietal lobule has been demonstrated in the early stages of AD on pathologic studies. In addition, projections from the inferior parietal lobule include subfields within the medial temporal lobe reflecting spread of AD pathology from the temporal lobe to an interconnected region in the parietal lobe.

While automated brain volumetry on MRI may be easily accessible in daily clinical practice, most studies have been performed in a research setting for tau PET. Currently there is no consensus for quantifying tau deposition on tau PET, and multiple tau-PET quantification methods (in vivo Braak staging, regional uptake in Braak composite regions, several whole-brain measures of tracer uptake, regional uptake in AD-vulnerable voxels and uptake in a priori defined regions) showed that all methods were related to amyloid and global cognition but regional measures covering AD-vulnerable regions increased sensitivity to early tau PET signal, atrophy and memory decline. We based our analysis of [18F] THK-5351 PET according to the methods described by Scholl et al. who used weighted bilateral composite FreeSurfer-derived regions of interests approximating the anatomical definitions of transentorhinal (Braak stage I/II), limbic (III/IV) and isocortical (V/VI) Braak stages. This reflects the propagation of tau accumulation through the course of the disease as supported by neuropathological data by Braak and Braak. Off-target binding remains as an important issue with [18F] THK-5351 in relation...
to MAO-B with greatest reductions seen in SUVs in the basal ganglia and thalamus when MAO-B inhibitors were used. While the heterogeneous MAO-B availability across the cortex may limit the interpretation of [18F] THK-5351, studies have demonstrated that no statistically significant reductions were seen using standard reference region-based approach. In addition, significant tau tracer retention in the temporal lobe as well as extra-temporal regions has been reported even in cognitively normal elderly population supporting the primary age-dependent tauopathy despite some controversy regarding this entity.

There were several limitations in our study. First, there were relatively small number of patients with AD (23%, 26/113) and overrepresentation of MCI (49%, 55/113). Second, age correction was not performed for automated volumetric analysis was not tested. In conclusion, this study demonstrated the robust performance of automated brain volumetry on MRI and quantitative measurement of tau deposition on [18F] THK-5351 PET in discrimination of AD spectrum with added value of automated brain volumetry in discriminating AD from HC. The cingulate isthmus and inferior parietal lobule were identified as significant brain regions in discriminating AD on the AD spectrum, and may provide new insights into the early diagnosis of AD.

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References
1. Ishii, K. PET approaches for diagnosis of dementia. Am. J. Neuroradiol. 35, 2030–2038. https://doi.org/10.3174/ajnr.A3695 (2014).
2. Jack, C. R. Jr. et al. Medial temporal atrophy on MRI in normal aging and very mild Alzheimer’s disease. Neurology 49, 786–794 (1997).
3. Dickerson, B. C. et al. Differential effects of aging and Alzheimer’s disease on medial temporal lobe cortical thickness and surface area. Neurobiol. Aging 30, 432–440. https://doi.org/10.1016/j.neurobiolaging.2007.07.022 (2009).
4. Fennema-Notestine, C. et al. Structural MRI biomarkers for preclinical and mild Alzheimer’s disease. Hum. Brain Mapp. 30, 3238–3253. https://doi.org/10.1002/hbm.20744 (2009).
5. McEvoy, L. K. et al. Mild cognitive impairment: Baseline and longitudinal structural MR imaging measures improve predictive performance. Radiology 259, 834–843. https://doi.org/10.1148/radiol.11101975 (2011).
6. Desikan, R. S. et al. Temporoparietal MR imaging measures of atrophy in subjects with mild cognitive impairment that predict subsequent diagnosis of Alzheimer disease. Am. J. Neuroradiol. 30, 532–538. https://doi.org/10.3174/ajnr.A11397 (2009).
7. Scheltens, P., Launer, L. I., Barkhof, F., Weinstein, H. C. G. & van Gool, W. A. Visual assessment of medial temporal lobe atrophy on magnetic resonance imaging: Interobserver reliability. J. Neurol. 242, 557–560. https://doi.org/10.1007/s004150050807 (1995).
8. Scheltens, P., Pasquier, F., Weerts, J. G., Barkhof, F. & Leys, D. Qualitative assessment of cerebral atrophy on MRI: Inter- and intraobserver reproducibility in dementia and normal aging. Eur. Neurol. 37, 95–99. https://doi.org/10.1159/000117417 (1997).
9. Ratnakrishnan, B. G., Doraiswamy, P. M. & Petrella, J. R. Science to practice: Translating automated brain MRI volumetry in discriminating AD from HC. The cingulate isthmus and inferior parietal lobule were identified as significant brain regions in discriminating AD on the AD spectrum, and may provide new insights into the early diagnosis of AD.

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Braak, H. & Braak, E. Neuropathological staging of Alzheimer-related changes. *Acta Neuropathol.* **82**, 239–259 (1991).

Hulette, C. M. et al. Neuropathological and neuropsychological changes in "normal" aging: Evidence for preclinical Alzheimer disease in cognitively normal individuals. *J. Neuropathol. Exp. Neurol.* **57**, 1168–1174 (1998).

Scholl, M. et al. PET imaging of tau deposition in the aging human brain. *Neuron* **89**, 971–982. https://doi.org/10.1016/j.neuron.2016.01.028 (2016).

Lemoine, L., Leuzy, A., Chiotis, K., Rodriguez-Viteze, E. & Nordberg, A. Tau positron emission tomography imaging in tauropathies: the added hurdle of off-target binding. *Alzheimer's Dement. Diagn. Assess. Dis. Mo.** **10**, 232–236. https://doi.org/10.1016/j.jadm.2018.01.007 (2018).

Lockhart, S. N. et al. Dynamic PET measures of tau accumulation in cognitively normal older adults and Alzheimer's disease patients measured using [18F] THK-5351. *PLoS ONE* **11**, e0158460. https://doi.org/10.1371/journal.pone.0158460 (2016).

Saint-Aubert, L. et al. Tau PET imaging: Present and future directions. *Mol. Neurodegener.* **12**, 19. https://doi.org/10.1186/s13024-017-0162-3 (2017).

Chiotis, K. et al. Imaging in vivo tau pathology in Alzheimer's disease with THK5317 PET in a multimodal paradigm. *Eur. J. Nucl. Med. Mol. Imaging* **43**, 1686–1699. https://doi.org/10.1007/s0033-016-3363-2 (2016).

Ossenkoppele, R. et al. Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. *Brain* **139**, 1551–1567. https://doi.org/10.1093/brain/aww027 (2016).

Jack, J. C. R. et al. Longitudinal tau PET in aging and Alzheimer's disease. *Brain* **141**, 1517–1528. https://doi.org/10.1093/brain/awy059 (2018).

Chiotis, K. et al. Longitudinal changes of tau PET imaging in relation to hypometabolism in prodromal and Alzheimer's disease dementia. *Mol. Psychiatry* **23**, 1666–1673. https://doi.org/10.1038/mp.2017.108 (2018).

McKhann, G. M. et al. The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement.** **7**, 263–269. https://doi.org/10.1016/j.jalz.2011.03.005 (2011).

Albert, M. S. et al. The diagnosis of mild cognitive impairment due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement.* **7**, 270–279. https://doi.org/10.1016/j.jalz.2011.03.008 (2011).

Song, M., Lee, S. H., Yu, K.-H. & Kang, Y. Development and validation of the full version of story memory in the Korean-Mini Mental State Examination, 2nd Edition: Expanded version (K-MMSE-2: EV). *Dement. Neurocogn. Disord.* **18**, 96–104. https://doi.org/10.1270/dd.dnd.2019.18.3.96 (2019).

Ye, B. S. et al. Effects of education on the progression of early- versus late-stage mild cognitive impairment. *Int. Psychogeriatr.* **25**, 597–606. https://doi.org/10.1017/S1041610212002001 (2013).

Park, J. E. et al. Intra-individual correlations between quantitative THK-5351 PET and MRI-derived cortical volume in Alzheimer's disease differ according to disease severity and amyloid positivity. *PLoS ONE* **14**, e0226265. https://doi.org/10.1371/journal.pone.0226265 (2019).

Jang, Y. K. et al. Head to head comparison of [(18)F] AV-1451 and [(18)F] THK5351 for tau imaging in Alzheimer's disease and frontotemporal dementia. *Eur. J. Nucl. Med. Mol. Imaging* **45**, 432–442. https://doi.org/10.1007/s0033-017-3878-0 (2018).

Joshi, A., Koeppe, R. A. & Fuselli, J. A. Reducing between scanner differences in multi-center PET studies. *Neuroimage* **46**, 154–159. https://doi.org/10.1016/j.neuroimage.2009.01.057 (2009).

Hoffman, E. J., Cutler, P. D., Digby, W. M. & Mazzotti, J. C. 3-D phantom to simulate cerebral blood flow and metabolic images for PET. *IEEE Trans. Nucl. Sci.* **37**, 616–620. https://doi.org/10.1109/23.1066868 (1990).

England, H. B., Gillis, M. M. & Hampstead, B. M. RBANS memory indices are related to medial temporal lobe volumetrics in healthy older adults and those with mild cognitive impairment. *Arch. Clin. Neuropsychol.* **29**, 322–328. https://doi.org/10.1093/arclin/acu012 (2014).

Desikan, R. S. et al. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *Neuroimage* **31**, 968–980. https://doi.org/10.1016/j.neuroimage.2006.01.021 (2006).

Ng, K. P. et al. Monoamine oxidase B inhibitor, selegiline, reduces 18F-THK5351 uptake in the human brain. *Alzheimer's Res. Ther.* **12**, 4. https://doi.org/10.1186/s13075-019-1349-x (2020).

Petrella, J. R., Coleman, R. E. & Doraiswamy, P. M. Neuroimaging and early diagnosis of Alzheimer disease: A look to the future. *Radiology* **226**, 315–336. https://doi.org/10.1148/radiol.2262011600 (2003).

Pearson, R. C., Esiri, M. M., Horns, R. W., Wilcock, G. K. & Powell, T. P. Anatomical correlates of the distribution of the pathological changes in the neocortex in Alzheimer disease. *Proc. Natl. Acad. Sci.* **82**, 4531–4534. https://doi.org/10.1073/pnas.82.13.4531 (1985).

Hof, P. R., Bouras, C. & Morrison, J. H. In *Cerebral Cortex: Neurodegenerative and Age-Related Changes in Structure and Function of Cerebral Cortex* (eds Peters, A. & Morrison, J. H.) 175–311 (Springer US, 1999).

Ding, S.-L., Van Hoesen, G. & Rockland, K. S. Inferior parietal lobule projections to the presubiculum and neighboring ventromedial cortical areas. *J. Comp. Neuro.* **425**, 510–530. https://doi.org/10.1002/0960-9998(20001002)425:4<510::AID-cne3>3.0.CO;2-e (2000).

Massa, A. et al. Comparison of multiple tau-PET measures as biomarkers in aging and Alzheimer's disease. *Neuroimage* **157**, 448–463. https://doi.org/10.1016/j.neuroimage.2017.05.055 (2017).

Lowe, V. J. et al. Widespread brain tau and its association with ageing, Braak stage and Alzheimer's dementia. *Brain* **141**, 271–287. https://doi.org/10.1093/brain/awx320 (2018).

Cravy, J. E. et al. Primary age-related tauopathy (PART): A common pathology associated with human aging. *Acta Neuropathol.* **128**, 755–766. https://doi.org/10.1007/s00401-014-1349-4 (2014).

Isac, Z. et al. Test–retest reliability of freesurfer measurements within and between sites: Effects of visual approval process. *Hum. Brain Mapp.* **36**, 3472–3485. https://doi.org/10.1002/hbm.22856 (2015).

**Author contributions**

M.K., S.J.K. and J.S.K. have made substantial contributions to the conception or design of the work, acquisition, analysis, or interpretation of data, drafted the work or substantively revised it. J.E.F., J.Y., W.H.S., J.S.O., M.O., J.H.R., S.W.S. and S.J.O. have made substantial contributions to substantively revise it.

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