A case of monoclonal gammopathy of renal significance presenting as atypical amyloidosis with IgA lambda paraproteinemia

Chankyung Kim¹, John Brealey¹, Anjelo Jobert², James Nolan¹

¹Department of Anatomical Pathology, SA Pathology, Adelaide; ²Central and Northern Adelaide Renal Transplantation Services, Royal Adelaide Hospital, Adelaide, Australia

Monoclonal gammopathy of renal significance is defined as any B cell or plasma cell clonal lymphoproliferation which neither causes tumor complications nor meets any current hematological criteria for specific therapy, with one or more kidney lesions related to the produced monoclonal immunoglobulin, such as amyloidosis. A 50-year-old male presented with heavy proteinuria and blood tests showing IgA and Lambda paraproteinemia. Light microscopy showed mesangial eosinophilic ground substance extending into the capillary loops, and positive staining within the glomeruli and vessel walls for amyloid P immunohistochemistry was also noted. Immunofluorescence showed positive staining for IgA and Lambda in the mesangia and capillary loops. Electron microscopy exhibited organized fibrils measuring 4–5 nm in diameter in the mesangia, glomerular basement membranes and vessel walls. We interpreted the overall findings as atypical renal amyloidosis with IgA and Lambda deposition on immunofluorescence. Further amyloid typing using laser microdissection-liquid chromatography and mass spectrometry will be useful.

Key Words: Monoclonal gammopathy of renal significance; AHL amyloidosis; Immunoglobulin A; Atypical amyloidosis

CASE REPORT

A 50-year-old male was evaluated by a renal physician due to the clinical features of nephrotic syndrome including heavy proteinuria and edema. A native kidney biopsy was sent to our laboratory for light microscopy, fresh frozen immunofluorescence and electron microscopy. Blood tests showed albumin of 17 g/L (normal range, 34 to 48 g/L) and creatinine of 99 μmol/L (normal range, 60 to 110 μmol/L), and the 24-hour urine protein was 8,596 mg/24 hr. The serum free kappa/lambda light chain ratio measured 0.09 (kappa light chain = 24.03 mg/L [normal range, 3.00 to 19.00 mg/L], lambda light chain = 276.87 mg/L [normal range, 6.00 to 26.00 mg/L]). The serum electrophoresis detected 5 g/L of paraprotein of IgA lambda and free lambda types.

A bone marrow biopsy showed the following: mildly increased plasma cells, less than 5%; phenotype: CD38⁺, CD138⁺, CD19⁻, known as MGRS-associated lesions, conditions or disorders [3].

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CD20−, CD56−, CD28+, CD117−; and cytoplasmic lambda light chain restriction. This was consistent with monoclonal gammopathy of undetermined significance (MGUS).

A percutaneous core renal biopsy was performed, and the following tissues were submitted: formalin-fixed tissue for histology, fresh tissue for immunofluorescence and glutaraldehyde-fixed tissue for electron microscopy.

Light microscopy showed mesangial eosinophilic ground substance extending into the capillary loops (Fig. 1A). Positive staining within the glomeruli and vessel walls was identified for amyloid P immunohistochemistry (Fig. 1B). The Congo red stain demonstrated faint non-specific staining of the glomeruli in which no green birefringence was seen on polarized light microscopy.

Immunofluorescence showed positive staining for IgA (Fig. 2A) and Lambda (Fig. 2B) in the mesangia and capillary loops. Electron microscopy exhibited fibrils measuring 4–5 nm in diameter in the mesangia, glomerular basement membranes and vessel walls (Fig. 2C).

DISCUSSION

We interpret this case as an unusual form of amyloidosis in a patient with a clinical presentation of heavy proteinuria in the setting of MGUS. The overall findings fit with a recently described diagnostic entity known as MGRS. Deposition of fibrillar eosinophilic material that show positive labelling with amyloid P im-

Fig. 1. Hematoxylin and eosin stain and Amyloid P immunohistochemistry. (A) The glomerulus shows mesangial eosinophilic ground substance which extends into the capillary loops. (B) Positive staining is seen within the glomerulus with amyloid P immunohistochemistry.

Fig. 2. Immunofluorescence and electron microscopy. (A) Positive mesangial and capillary loop staining with IgA on immunofluorescence. (B) Positive mesangial and capillary loops staining with Lambda on immunofluorescence. (C) The fine fibrillar material (F) is present adjacent to the mesangial cell (M) (×40,000). The inset shows amyloid-like fibrils measuring approximately 5 nm in diameter (×120,000).
munohistochemistry in the mesangia, glomerular basement membranes and blood vessels supports the diagnosis of amyloidosis. On the other hand, there were two factors that are unusual for typical amyloidosis. First, the fibrils in this case only measured 4–5 nm in thickness, whereas amyloid fibrils typically measure 7–12 nm [4]. The absence of green birefringence or anomalous colors on polarized microscopy was the second unusual finding.

The weak non-specific staining of glomeruli with Congo red without green birefringence on polarized microscopy observed in this case could be secondary to the properties of the amyloid protein. A study by Suhr et al. [5] revealed that different types of transthyretin (TTR) amyloid fibrils can exhibit altered affinity for Congo red stain and birefringence. For example, type A, which consists of carboxy-terminal TTR-related amyloidosis (ATTR) fragments and full-length TTR, displayed strong affinity for Congo red stain, whereas type B, which consists only of full-length TTR, showed weak affinity for Congo red stain and absence of glittering birefringence. Thus, amyloids may demonstrate a variable affinity for Congo red and variable birefringence.

The immunofluorescence staining suggests that the amyloid is of IgA lambda origin, and hence, a diagnosis of IgA-lambda type renal heavy- and light-chain amyloidosis (AHL amyloidosis) is being considered. However, there is a case report from Japan which described AL amyloidosis in a patient with lymphoplasmacytic lymphoma secreting IgM-kappa chains [6]. In this case, the origin of the amyloid was initially thought to be from both IgM heavy and kappa light chain as suggested by renal immunofluorescence and laser microdissection–liquid chromatography and mass spectrometry (LMD-LC-MS). However, LMD-LC-MS of the amyloid fibrils after the amyloid purification method suggested that the heavy chain identified was actually from the non-specific co-deposition of monoclonal IgM-kappa, which demonstrated that the amyloid was only of light chain origin. Therefore, it would be imprudent to diagnose the present case as AHL amyloidosis until we obtain the LMD-LC-MS results.

According to the study published by Nasr et al. [7], among 16 heavy-chain (AH) and heavy- and light-chain renal amyloidosis cases, five showed detection of IgA, and one showed both IgA and lambda positivity on laser microdissection and mass spectrometry (LMD-MS). There were 202 cases of AL renal amyloidosis during the same time period, with AH/AHL renal amyloidosis cases accounting for 7.3% of the entire cohort and IgA and lambda positive AH/AHL occupying only less than 0.01% of the entire cohort. Interestingly, there was a case in which immunofluorescence was positive for both IgA and lambda, but only IgA was found on LMD-MS. This again demonstrates the difficulty in accurate typing of AH/AHL solely by immunofluorescence.

Given the unusual findings for typical amyloidosis in the present case, other possible diagnoses were considered. Negative staining with Congo red suggested monoclonal immunoglobulin deposition disease (MIDD) as an alternate diagnosis, especially when amyloid P immunohistochemistry can sometimes show non-specific staining (from our experiences). However, electron microscopy showed organized fibrils rather than the non-fibrillar “powdery” electron dense deposits which are typically seen in MIDD [8]. This made the diagnosis of MIDD less likely. Positive IgA staining on immunofluorescence is unusual for amyloidosis and raised the possibility of IgA nephropathy, but no mesangial or endocapillary hypercellularity was present, C3 was negative, and most importantly, the clinical presentation did not fit with IgA nephropathy. Hence, this was disregarded from our diagnostic work-up.

As mentioned, further renal amyloid typing can be performed by LMD-LC-MS [3]. Vrana et al. [9] reported that this method showed 100% specificity and 98% sensitivity in the identification of the types of amyloid protein. This may be indicated in our case due to the questionable Congo red staining and bright staining of IgA by immunofluorescence. Identification of the amyloid type is incredibly important as the disease management depends on the underlying cause of amyloidosis. In our case, further amyloid typing is being considered, and meanwhile, the patient will undergo autologous bone marrow transplantation for treatment.

Ethics Statement
Informed consent was obtained from the patient, and the study was approved by the Central Adelaide Local Health Network Human Research Ethics Committee (CALHN reference number: 13298, approved on 01/06/2020).

ORCID
Chanlyung Kim https://orcid.org/0000-0001-6054-0605
James Nolan https://orcid.org/0000-0002-6034-7159

Author Contributions
Conceptualization: CK, JN. Data curation: CK. Formal analysis: CK, JB, JN. Investigation: CK, JN. Methodology: CK, JB, AJ, JN. Resources: JB, AJ, JN. Supervision: JN. Validation: JB, AJ, JN. Visualization: CK. Writing—original draft: CK. Writing—review & editing: CK, JB, AJ, JN. Approval of final manuscript: all authors.

Conflicts of Interest
The authors declare that they have no potential conflicts of interest.

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References
1. Westermark P, Benson MD, Buxbaum JN, et al. A primer of amyloid nomenclature. Amyloid 2007; 14: 179-83.
2. Holanda DG, Acharya VK, Dogan A, Racusen LC, Atta MG. Atypical presentation of atypical amyloid. Nephrol Dial Transplant 2011; 26: 373-6.
3. Leung N, Bridoux F, Batuman V, et al. The evaluation of monoclonal gammopathy of renal significance: a consensus report of the International Kidney and Monoclonal Gammopathy Research Group. Nat Rev Nephrol 2019; 15: 45-59.
4. Leung N, Drosou ME, Nasr SH. Dysproteinemias and glomerular disease. Clin J Am Soc Nephrol 2018; 13: 128-39.
5. Suhr OB, Lundgren E, Westermark P. One mutation, two distinct disease variants: unravelling the impact of transthyretin amyloid fibril composition. J Intern Med 2017; 281: 137-47.
6. Manabe S, Iwasaki C, Hatano M, et al. AL amyloidosis with non-amyloid forming monoclonal immunoglobulin deposition: a case mimicking AH1 amyloidosis. BMC Nephrol 2018; 19: 337.
7. Nasr SH, Said SM, Valeri AM, et al. The diagnosis and characteristics of renal heavy-chain and heavy/light-chain amyloidosis and their comparison with renal light-chain amyloidosis. Kidney Int 2013; 83: 463-70.
8. Nasr SH, Valeri AM, Cornell LD, et al. Renal monoclonal immunoglobulin deposition disease: a report of 64 patients from a single institution. Clin J Am Soc Nephrol 2012; 7: 231-9.
9. Vrana JA, Gamez JD, Madden BJ, Theis JD, Bergen HR 3rd, Dogan A. Classification of amyloidosis by laser microdissection and mass spectrometry-based proteomic analysis in clinical biopsy specimens. Blood 2009; 114: 4957-9.