Chemical composition and biological activities of essential oils of two new chemotypes of *Glebionis* Cass.

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Abstract: This paper includes the result of the first study of the chemical composition, antioxidant, antiinflammatory, and anti-diabetic activities of the essential oils of *Glebionis coronaria* (L.) Cass. ex Spach and *Glebionis segetum* (L.) Fourr. from Turkey. In the current study, nine and twenty-eight constituents were determined in the essential oils of aerial parts of *G. coronaria* (GCE) (92.1%) and *G. segetum* (GSE) (90.0%), respectively. The main components were capillin (65.9%) in GCE, capillene (53.4%) in GSE. The essential oil compositions were evaluated and compared with previous researches. In the current study, the plants are classified as chemotypes of *Glebionis* species. GCE and GSE showed poor and very poor DPPH radical scavenging activity, respectively. GCE and GSE exhibited significant and strong antiinflammatory activity against the 5-lipoxygenase enzyme, respectively. Also, GCE and GSE displayed moderate and weak anti-diabetic activity against the α-glucosidase enzyme, respectively. Polycytylenes were determined as the main class of compounds in GCE and GSE and had a notable anti-inflammatory activity.

Key words: *Glebionis coronaria*, *Glebionis segetum*, essential oil, chemotype, anti-inflammatory activity

1. Introduction

The genus *Glebionis* Cass. is an annual herb and belongs to the Asteraceae family. *Glebionis* is extended in the Mediterranean, North Africa, Europe, and Asia. *Glebionis* is represented in Turkey by two species namely *Glebionis coronaria* (L.) Cass. ex Spach (syn.: *Chrysanthemum coronarium* L.) and *Glebionis segetum* (L.) Fourr. (syn.: *Chrysanthemum segetum* L.). *G. coronaria* leaves are 2-3-pinnatisect. The plant grows on banks, roadsides, and fields near the sea of East Thrace, West and South Anatolia region of Turkey. *G. segetum* leaves are 1-pinnatisect (or upper entire). The plant grows on fallow fields, cornfields, and roadsides of West and South Anatolia and North-West region of Turkey [1]. *G. coronaria* and *G. segetum* called as ‘Dağlama, Sari papatya, Krizantem, Ale gömeci or Kasımpatı’ in Anatolia are traditionally used for the relief of abdominal pain and sore throat, treat shortness of breath and against hair loss by human [2]. They have antimicrobial, antioxidant, antiviral, antimycotic, cytotoxic activities [3–6]. *G. coronaria* is used as an edible plant in Japan and China. The leaves are used for the suppression of the fishy odors in food in Japan. Moreover, the leaves and stems of the Japanese sample have β-carotene, minerals, and vitamins in a high amount [7]. The aerial parts of both species are used as salad and food in the southern part of Turkey as well [8]. The essential oil constituents of *G. coronaria* have also been widely studied, whereas there is less report on the essential oil composition of *G. segetum*. The essential oils of *G. coronaria* displayed variation in their major compounds due to distinct geographical places. Camphor, α-pinene, l-yratyl acetate, trans-chrysanthenyl acetate, trans-chrysanthenyl isovalerate, cis-chrysanthenyl acetate, trans-tonghaosu, α-humulene, γ-curcumene, (Z)-ocimene, myrcene, bornyl acetate, α-bisabolol, chrysanthenyl acetate, chrysanthemol, santolinatriene, and yomogi alcohol were found as main components of *G. coronaria* grown in Italy, Spain, Greece, Tunisia, Chile, South Korea, Ukraine, Jordan, and Cyprus [3,4,9–18]. (E,E)-α-farnesene, α-humulene, and tonghaosu were determined as the main compounds of *G. segetum* grown in Italy and China [4,19]. The previous studies displayed high variability in the essential oil ingredients and main compounds. In the current study, capillin and capillin were determined as the main compounds of *G. segetum* and *G. coronaria* collected from Istanbul, respectively. However, capillin and capillene were not determined in the essential oils of *G. coronaria* and *G. segetum* in previous studies [3,4,9–19]. There are no reports on the essential oil composition of *G. coronaria* and *G. segetum* from Turkey as well as on the α-glucosidase and 5-lipoxygenase inhibitory activity of GCE and GSE. This study aimed to specify the chemical composition and biological activities of the essential oils of two new chemotypes of *Glebionis*.

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2. Materials and methods

2.1. Plant material

*G. coronaria* and *G. segetum* were collected in İkitelli-Başakşehir (41° 04’ 34.9” N; 28° 47’ 32.2” E) and Kayaşehir- Başakşehir (41° 7’ 36” N; 28° 46’ 53” N), İstanbul, Turkey in May 2017 and 2019 by Asst. Prof. Dr. Huseyin Servi and identified by Dr. Ahmet Dogan, respectively. Herbarium specimens of *G. coronaria* and *G. segetum* were deposited in the Marmara University Herbarium (Herbarium numbers: MARE20235 for *G. coronaria* and MARE22152 for *G. segetum*).

2.2. Distillation

The dried aerial parts of GCE (339 g) and GSE (420 g) were subjected to hydrodistillation for 3 h, using a Clevenger apparatus.

2.3. Chromatographic analyses

The GC-MS analysis and determination of essential oils components were performed as described by Abu Zarga et al. [20]. In GC-MS analyses, the DP-5 (5% diphenyl, 95% dimethyl polysiloxane) capillary column (30m × 0.25 mm, 0.25 m film thickness) and helium as carrier gas (0.9 mL/min) were used. The determination of the components was done by relative retention indices comparison of n-alkane series to the literature and with mass spectra comparison (Wiley 8th Ed./NIST 05 Mass Spectra library) [21].

The GC analyses were performed as described by Abu Zarga et al. [20]. The temperature of the FID detector was maintained at 300 °C. The same operating conditions and column were utilized in the GC-MS analyses.

2.4. DPPH radical scavenging activity

The free radical scavenging activity of the essential oils, based on the scavenging activity of the stable 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical was determined by the method described by Zou et al. [22]. Ascorbic acid (100-0.02 µg/mL) was used as a standard. In short, 10 µL of oils (250–0.49 µg/mL) or ascorbic acid dissolved in dimethylsulfoxide (DMSO) at different concentrations. In a 96-well plate, the oils and standard were mixed with 190 µL of 0.1 mM DPPH solution in MeOH. The reaction mixture was left in the dark at RT for 30 min. The absorbance of the mixture was measured spectrophotometrically at 517 nm. Each experiment was performed in triplicate. The percent radical scavenging activity of oils and standard against DPPH was calculated according to the following:

\[
\text{DPPH radical-scavenging activity (\%) = \left[ \left( A_0 - A_1 \right) / A_0 \right] \times 100 }
\]

where \( A_0 \) is the absorbance of the control (containing all reagents except the test compounds), and \( A_1 \) is the absorbance of the oils/standard. Oil concentration providing 50% inhibition (IC\(_{50}\)) was calculated from the graph plotting inhibition percentage against oil concentration.

2.5. In vitro antiinflammatory activity

5-lipoxygenase inhibition activity was analyzed by the method of Phosrithong and Nuchtavorn [23] with slight modifications described by Yildirim et al. [24]. Indomethacin (100–0.02 µg/mL) was used as a standard. 10 µL of oils (250–0.49 µg/mL) or indomethacin were added to ethanol (20 µL), pure water (20 µL), sodium borate buffer solution (25 µL, 0.1 M, pH 9) and type V soybean lipoygenase solution (25 µL, 20.000 units/mL) in the buffer (pH 9). The reaction mixture was preincubated at 25 °C for 5 min. Then, linoleic acid solution (100 µL of 0.6 mM) was added to solutions, mixed well, and the change in absorbance at 234 nm was followed for 6 min. Each reaction was run in triplicate. The percent inhibition was calculated from the following equation:

\[
\% \text{ inhibition} = \left[ \left( A_{\text{control}} - A_{\text{sample}} \right) / A_{\text{control}} \right] \times 100
\]

A dose-response curve was plotted to determine the IC\(_{50}\) values. IC\(_{50}\) is defined as the concentration sufficient to obtain 50% of maximum antiinflammatory activity.

2.6. In vitro antidiabetic activity

The inhibition assay for α-glucosidase activity was conducted as described by Ramakrishna et al. [25] with slight modifications described by Sen et al. [26]. Acarbose (100–0.02 µg/mL) was used as a standard. In a 96-well plate, 10 µL of essential oils (250–0.49 µg/mL) or acarbose, 40 µL of 0.1 M sodium phosphate buffer (pH 6.9), and 100 µL of α-glucosidase (obtained from *Saccharomyces cerevisiae*) solution (1 unit/mL) were mixed. After preincubation (at 25 °C for 10 min), p-nitrophenyl-α-D-glucopyranoside (pNPG) (50 µL of 5 mM) to the solutions was added and reincubated at 25 °C for 5 min. The absorbance reading was taken before and after incubation at 405 nm using a microplate reader. Tests were performed in triplicate. The percentage inhibitory activity of the oils and standard against α-glucosidase enzyme were calculated according to the following:

\[
\text{α-glucosidase inhibitor activity (\%) = \left[ \left( A_0 - A_1 \right) / A_0 \right] \times 100 }
\]

where \( A_0 \) is the absorbance of the control (containing all reagents except the test compounds), and \( A_1 \) is the absorbance of the oils/standard. Essential oils or standard concentration providing 50% inhibition (IC\(_{50}\)) was calculated from the graph plotting inhibition percentage against oil or standard concentration.
2.7. Statistical analysis
The results were indicated with ± standard deviation. The statistical analysis was done by Tukey’s Multiple Comparison Test with a confidence interval (CI) of 95% for each using the GraphPad Prism 5. Differences between means at p<0.05 levels were regarded as important.

3. Results and discussion
The yields of GCE and GSE were 0.12 and 0.30 (v/w), respectively. Nine and twenty-eight constituents were determined in GCE (92.1%) and GSE (90.0%), respectively. The main components were determined as capillin 65.9% in GCE (given in Figure 1 and Table 1), and as capillene 53.4% in GSE (given in Figure 2 and Table 1). Other major compounds were capillene 11.9% and caryophyllene oxide 6.8% in the GCE, and caryophyllene oxide 9.2%, 1-phenyl-penta-2,4-diyne 7.3%, and capillin 7.2% in the GSE (Table 1). The GCE and the GSE were dominated by the presence of polyacetylenes (79.0% and 68.1%, respectively).

The essential oils of G. coronaria and G. segetum possessed remarkable diversities in the main compounds. These diversities could be related to morphological characteristics and different geographical origins. The main compounds of the essential oil from flowers of G. coronaria grown in Sardinia-Italy were α-humulene, camphor, and γ-curcumene [4]; camphor and cis-chrysanthenyl acetate from a population in Tuscany-Italy [13]; trans-tonghaosu from populations in Sicily and Campania-Italy [16]; camphor, α-pinene and lyratyl acetate from a population Murcia-Spain [10]; trans-chrysanthenyl acetate and trans-chrysanthenyl isovalerate from a population in Korinthos-Greece [12]; camphor from a population in Attiki-Greece [12]; cis-chrysanthenyl acetate and trans-chrysanthenyl acetate from a population in Zanghouan-Tunisia [14]; camphor, cis-chrysanthenyl acetate and bornyl acetate from a population in Chile [15]; chrysanthenyl acetate and chrysanthemol from a population in Ukraine [3]; camphor, perilla aldehyde and cis-chrysanthenyl acetate from a population in Jordan [17]. The main compounds of essential oil from stems and leaves of G. coronaria were myrcene, α-bisabolol and (E,E)-α-farnesene from a population in Namyamju-South Korea [18]; (Z)-ocimene and myrcene from a...

**Figure 1.** GC-MS chromatogram of the GCE (1: Capillene; 2: Capillin)
population in Sicily-Italy [16]. The main compounds of the essential oil from aerial parts of *G. coronaria* were myrcene and (E)-β-farnesene from a population in Jordan [17]; camphor, santolinatriene, yomogi alcohol, *cis*-chrysanthenyl acetate, and bornyl acetate from a population in Cyprus [9]. The main compounds of the essential oil from flowers of *G. segetum* were (E,E)-α-farnesene and α-humulene from a population in Sardinia-Italy [4]; tonghaosu from a population in China.

Table 1. The essential oil compositions of the aerial parts of *G. coronaria* and *G. segetum.*

| RRI Exp. | RRI Lit. | Compounds | *G. coronaria (%)* | *G. segetum (%)* |
|---------|----------|-----------|--------------------|-----------------|
| 932     | 939      | α-Pinene  | -                  | 0.2             |
| 958     | 963      | Benzaldehyde | 2.6               | 0.6             |
| 1027    | 1029     | Limonene  | -                  | 0.2             |
| 1284    | 1280     | 1-Phenyl-penta-2,4-diyne | 0.4               | 7.3             |
| 1375    | 1377     | α-Copaene | -                  | 0.1             |
| 1419    | 1419     | β-Caryophyllene | -                  | 1.1             |
| 1453    | 1455     | α-Humulene | -                  | 0.2             |
| 1458    | 1458     | (E)-β-farnesene | -                  | 4.8             |
| 1496    | 1490     | Capillene | **11.9**           | **53.4**        |
| 1537    | 1546     | 1-(4-Methoxyphenyl)-2,4-pentadiyne | 0.8               | 0.2             |
| 1552    | 1555     | Isocaryophyllene oxide | -                  | 0.2             |
| 1565    | 1563     | (E)-Nerolidol      | -                  | 0.5             |
| 1571    | 1567     | (Z)-3-hexenyl benzoate | -                  | 1.0             |
| 1574    | 1574     | Dendrolasen | -                  | 0.3             |
| 1578    | 1574     | Hexyl benzoate | -                  | 0.6             |
| 1582    | 1581     | Caryophyllene oxide | 6.8               | 9.2             |
| 1585    | 1579     | Isoaromadendrene epoxide | -                  | 0.1             |
| 1593    | 1595     | Salvial-4(14)-en-1-one | -                  | 0.2             |
| 1609    | 1606     | Humulene oxide II | -                  | 0.5             |
| **1642**| **1637** | Capillin  | **65.9**           | 7.2             |
| 1668    | 1689     | 2,3-dihydro farnesol | -                  | 0.1             |
| 1838    | 1837     | Neophytadiene      | -                  | 0.3             |
| 1845    | 1847     | Hexahydrofarnesyl acetone | 0.9               | -               |
| 1867    | 1868     | Isobutyl-o-phthalate | -                  | 0.3             |
| 2099    | 2086     | 9,12-15-octadecatrienoic acid methyl ester | -                  | 0.2             |
| 2111    | 2116     | Phytol             | -                  | 0.2             |
| 2298    | 2300     | Tricosane          | 1.3               | 0.5             |
| 2498    | 2500     | Pentacosane        | 1.5               | 0.3             |
|         |          | Phenyl alkynes     | 79.0              | 68.1            |
|         |          | Monoterpens        | -                  | 0.4             |
|         |          | Sesquiterpenes     | 7.7               | 17.0            |
|         |          | Diterpenes         | -                  | 0.2             |
|         |          | Fatty acid derivatives | 2.8               | 1.1             |
|         |          | Others             | 2.6               | 3.1             |
|         |          | **Total identified compounds** | **92.1**           | **90.0**        |

1RRI Exp.: Relative retention indices calculated against *n*-alkanes (C5-C30); 2RRI Lit: Relative retention indices given in the literature for the compound in similar columns and analysis conditions.
These main compounds were found either in low concentration or not determined in the oils of the present study. The absence of these main compounds in the present study indicates the different chemovarieties of the plants. The previous reports displayed that *G. coronaria* and *G. segetum* collected from Italy, Spain, Greece, Tunisia, Chile, South Korea, Ukraine, Jordan, China, and Cyprus had monoterpenes, oxygenated monoterpenes, sesquiterpenes, oxygenated sesquiterpenes, and acetylenes as main groups. In the present research, GCE and GSE have polyacetylenes as the main group and displayed dissimilar chemical profiles from the previous studies.

The variations of the essential oil ingredients and composition may be connected to factors such as plant parts used, geographical regions, genotype, ecotype, chemotype, phenophases, and the environmental factors which can be temperature differences, relative humidity, irradiance, and photoperiod. The quantitative composition of the volatile oils of numerous aromatic plants is significantly influenced by the harvesting time, plant age, and product density [27].

Capillene and capillin were mainly determined from the essential oil of various *Artemisia* species including *A. campestris* [28], *A. campestris* var. *glutinosa* [29], *Artemisia capillaris* [30], *A. dracunculus* [31], *A. glauca* [32], *A. lehmanniana* [31], *A. monosperma* [33], *A. ordosica* [34], *A. scoparia* [35], *A. stricta* [36], and *Santolina rosmarinifolia* L. ssp. *rosmarinifolia* [37].

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**Table 2.** Antioxidant, antiinflammatory, and antidiabetic activities of the GCE and GSE.

| Essential oils and standards | DPPH radical scavenging activity (IC_{50} mg/mL) | Antilipoxygenase activity | α-glucosidase inhibitory activity |
|-----------------------------|---------------------------|---------------------------|----------------------------------|
| GCE*                        | 1.020 ± 0.004b            | 0.151 ± 0.001c            | 0.499 ± 0.006b                   |
| GSE*                        | 2.525 ± 0.017c            | 0.017 ± 0.003a            | 0.967 ± 0.006c                   |
| Ascorbic acid               | 0.018 ± 0.000a            |                           |                                  |
| Indomethacin                |                           | 0.022 ± 0.000a            |                                  |
| Acarbose                    |                           |                           | 0.040 ± 0.002a                   |

*Abbreviations: GCE and GSE show essential oils of aerial parts of *Glebionis coronaria* and *Glebionis segetum*, respectively. **Each value in the table is represented as mean ± SD (n = 3). Different letter superscripts in the same column indicate significant differences (p < 0.05).*
Capillene and capillin are polyacetylenes that are a group of bioactive secondary metabolites. Polyacetylenes display toxic features that possess strong skin sensitization, irritant, and allergic contact dermatitis, and neurotoxicity at high concentrations. Despite the toxic properties of polyacetylenes, many studies reported that polyacetylenes have various biological activities such as antidiabetic, antitumor, antiinflammatory, and antimicrobial activities [38-41]. Additionally, capillene and capillin displayed significant antimicrobial, antiapoptotic, cytotoxic, anticancer, antifeedant, and antiinflammatory activities [42-44].

There are studies on DPPH radical scavenging activity of essential oil of Chrysanthemum coronarium (Glebionis coronaria) in the literature. In a study, Polatoglu et al. reported that C. coronarium (G. coronaria) essential oil did not have significant DPPH radical scavenging activity [9]. In another study, Hosni et al. found that hydro-distilled essential oil obtained from the C. coronarium flowerheads had DPPH radical inhibition lesser than 15% at a concentration of 200 µg/mL [14]. However, there is no study on the DPPH radical scavenging activity of the essential oil of Glebionis segetum. In the current study, GCE and GSE showed poor and very poor radical scavenging activity with IC\textsub{50} values of 1.020 and 2.525 mg/mL against DPPH radical, respectively (given in Table 2).

GCE and GSE displayed moderate and weak antidiabetic activity with IC\textsub{50} values of 0.499 and 0.967 mg/mL against the α-glucosidase enzyme, respectively (given in Table 2). There is no study on the α-glucosidase inhibitory activity of essential oils of GCE and GSE in the literature. However, there is a study on the α-glucosidase inhibitory activity of capillin obtained from CH\textsub{2}Cl\textsub{2} fraction of MeOH extract of A. capillaris. Capillin showed strong α-glucosidase inhibitory activity [38]. Thus, capillin, which was the main compound of GCE may be responsible for the antidiabetic activity of the oil.

Also, the GCE and GSE exhibited significant and strong antiinflammatory activity with IC\textsub{50} values of 0.151 and 0.017 mg/mL against 5-lipoxygenase enzyme, respectively (given in Table 2). Although there is no report on the 5-lipoxygenase enzyme inhibitory or antiinflammatory activity of GCE and GSE, there are studies on the antiinflammatory activity of the extracts of these species. In a study, Strzelecka et al. notified that ethanol extract of C. coronarium (G. coronaria) decreased cytokine or LPS-stimulated iNOS mRNA levels in MBE (Murine brain microvascular endothelial cells) and P388D1 (Murine monocyte/macrophage-like cell line cells) [45]. In another study by Mascolo et al. found the ethanol extract of C. segetum (G. segetum) inhibited carrageenin foot edema by 11% in rats (100 mg/kg p.o.) [46]. Capillin and capillene have been notified to have antiinflammatory features in previous studies [47,48]. Thus, capillin and capillene, determined as main compounds in the GCE and GSE may be responsible for the antiinflammatory activity of the oils.

4. Conclusion
In general, the essential oil composition of the current study showed differences in quality and quantity from the previous research. These differences of the current study may be considered as chemotypes, which can be named capillin and capillene chemotypes. As a result, polyacetylenes were the main group of the GCE and GSE and had a notable antiinflammatory activity. However, in vivo studies are required to verify our findings.

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