Abstract. Hypoxia promotes drug resistance and induces the expression of hypoxia inducible factor (HIF)-1α in liver cancer cells. However, to date, no selective HIF-1α inhibitor has been clinically approved. The aim of this study is to investigate a drug-targetable molecule that can regulate HIF-1α under hypoxia. The present study demonstrated that hyperactivation of dual-specificity tyrosine-phosphorylation-regulated kinase 1A (DYRK1A)/HIF-1α signaling was associated with an increased risk of liver cancer. In addition, DYRK1A knockdown using small interfering RNA transfection or treatment with harmine, a natural alkaloid, significantly reduced the protein expression levels of HIF-1α in liver cancer cells under hypoxic conditions in vitro. Conversely, DYRK1A overexpression-vector transfection in liver cancer cell lines notably induced HIF-1α expression under the same conditions. Furthermore, DYRK1A was shown to interact and activate STAT3 under hypoxia to regulate HIF-1α expression. These findings indicated that DYRK1A may be a potential upstream activator of HIF-1α and positively regulate HIF-1α via the STAT3 signaling pathway in liver cancer cells. Additionally, treatment with harmine attenuated the proliferative ability of liver cancer cells under hypoxic conditions using sulforhodamine B and colony formation assay. Furthermore, DYRK1A knockdown could significantly enhance the anti-liver cancer effects of regorafenib and sorafenib under hypoxia. Co-treatment with harmine and either regorafenib or sorafenib also promoted cell death via the STAT3/HIF-1α/AKT signaling pathway under hypoxia using PI staining and western blotting. Overall, the results from the present study suggested that DYRK1A/HIF-1α signaling may be considered a novel pathway involved in chemoresistance, thus providing a potentially effective therapeutic regimen for treating liver cancer.

Introduction

Liver cancer is the sixth most commonly diagnosed cancer; it can be resistant to the majority of anticancer therapies and is the third leading cause of cancer-related death worldwide (1). Liver cancer is one of the most hypoxic tumors, with a median oxygen level of 0.8%. Therefore, the hypoxic microenvironment is a typical characteristic of liver cancer (2). It has been reported that hypoxia induces hypoxia inducible factor (HIF-1α) upregulation, promotes drug resistance and metastasis, and enhances the aggressiveness of liver cancer cells (3). Therefore, targeting tumor hypoxia/HIF-1α could be considered as an attractive strategy for developing novel anti-liver cancer drugs and enhancing the chemosensitivity of liver cancer cells (4,5). Although numerous HIF-1α inhibitors have been identified, no selective HIF-1α inhibitor has been clinically approved. However, several clinically approved drugs, such as vorinostat and digitoxin, have been shown to directly or indirectly inhibit HIF-1α expression (6). Therefore, alternative strategies for implementing HIF-1α suppression are urgently needed.
Dual-specificity tyrosine-phosphorylation-regulated kinase 1A (DYRK1A) is a serine/threonine kinase located on human chromosome 21. Previous studies have suggested that DYRK1A overexpression is involved in the development of neurodegenerative diseases, including Down's syndrome, Alzheimer's, Parkinson's and Huntington's disease (7,8). In addition to its role in brain development, DYRK1A contributes to the progression and development of a number of malignancies, such as glioblastoma multiform and pancreatic ductal adenocarcinoma (9,10). Recent studies have shown that DYRK1A could serve a significant role in drug resistance, since DYRK1A silencing could enhance the anticancer effects of AZD9291 or Bcl-2 inhibitors (11,12). Nevertheless, signaling events associated with DYRK1A-regulated drug resistance remain to be further exploited. A previous study revealed that DYRK1 could phosphorylate inhibitor of DNA binding 2 (ID2) on Thr27 and promote the ubiquitylation of HIF-2α under normoxic conditions, whereas under hypoxic conditions, DYRK1 inactivation attenuated phosphorylation in glioblastoma cells (13). The present study aimed to investigate whether DYRK1A could affect HIF-1α expression and/or regulate HIF-1α expression through STAT3 in liver cancer cells under hypoxic conditions.

Sorafenib and regorafenib are commonly used to treat patients with liver cancer; however, this is often with no satisfactory benefits (14). The activity of sorafenib and regorafenib is limited owing to primary and acquired resistance mechanisms. In addition, the resistance to these drugs is associated with the increased expression levels of multidrug resistance protein 1, glucose transporter 1 and VEGF, owing to the stabilization of HIF-1α protein (15,16). Therefore, the present study aimed to investigate whether DYRK1A downregulation could affect the anti-liver cancer effect of sorafenib and sorafenib through the regulation of HIF-1α, thus providing a novel strategy for improving the clinical efficacy of sorafenib and sorafenib in the treatment of patients with liver cancer.

Materials and methods

Materials. Regorafenib (cat. no. A8236; 98.24% purity) and sorafenib (cat. no. A3009; 99.87% purity) were purchased from APeX BIO Technology LLC. Harmine (cat. no. HY-N0737A; ≥95% purity) was provided by MedChemExpress. The chemical structures of harmine, regorafenib and sorafenib were shown in Fig. 1. Cycloheximide (CHX, cat. no. 01810; ≥95% purity) was obtained from Sigma-Aldrich (Merck KGaA), and 20 μg/ml CHX was used to prevent the synthesis of new proteins. DMSO was obtained from Hangzhou Keyi Biotechnology Co., Ltd., and the concentrations of DMSO used in cells under hypoxic conditions.

Sorafenib in the treatment of patients with liver cancer.

Materials and methods

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Cell culture. 293T, HepG2, Hep3B and HuH7 were purchased from Shanghai Institute of Biochemistry and Cell Biology. Hep3B cells were maintained in Minimum Essential Medium (cat. no. A1599001; Lonza) supplemented with 10% FBS (cat. no. P30-3302, PAN-Biotech). HepG2 and HuH7 cells were maintained in DMEM (cat. no. C0006; Hangzhou Keyi Biotechnology Co., Ltd.) supplemented with 10% FBS. 293T cells were cultured in 90% DMEM supplemented with 10% FBS. The hypoxic condition was created by placing liver cancer cells in a sealed hypoxic chamber and equilibrating it with a gas mixture of 1% O2, 5% CO2 and 94% N2. To prevent mycoplasma contamination, mycoplasma testing was performed for the cell lines used. Mycoplasma testing was performed on cell lines, which were then authenticated for genotypes using short tandem repeat DNA fingerprinting; the cells were passaged for <6 months (17).

Sulfurhexamine B (SRB) assay. HepG2, Hep3B and HuH7 cells were seeded into a 96-well plate at a density of 8x103 cells/well. At 30% confluence, cells were incubated with 4-32 μM harmine, 0.25-8 μM regorafenib and/or 2.5-20 μM at 37°C. Following incubation for 72 h, cells were fixed with 10% trichloroacetic acid for 4 h at 4°C, after which the 96-well plate was washed with tap water. After the wells had dried, cells were stained with 0.4% SRB solution for 30 min at room temperature. Subsequently, cells were washed with 1% acetic acid. After the plates were dried up, 50 μl SRB dye was solubilized in Tris-based solution (10 mM) at room temperature for 1 min. Finally, to determine cell cytotoxicity, the absorbance at a wavelength of 515 nm was measured in each well (18). Cell viability was calculated according to the following equation: Cell viability=cell count of treated sample/cell count of control.

Colony formation assay. HepG2 and Hep3B cells were plated into culture dishes at a density of 6x104 cells/dish and were then treated with the 5, 10, 20 or 40 μM harmine, 4 μM regorafenib and 2 μM sorafenib for 14 days at 37°C. The medium containing the compounds was replaced every 3 days. Following incubation for 14 days at 37°C, cells were incubated with 0.1% crystal violet containing 1% methanol for 30 min at room temperature cells were then washed with PBS, as previously described (19). Colonies containing >50 cells were viewed by light microscope (x40 magnification) and counted using ImageJ software (National Institutes of Health; version v1.8.0).

Cell death detection. HepG2 and Hep3B cells were seeded into a six-well plate at a density of 2x104 cells/well. When cell confluence reached ~30%, as determined by light microscopy, they were treated with the 5 or 8 μM harmine and/or 14 or 16 μM regorafenib for 48 h at 37°C. Subsequently, cells were collected at 448 x g for 5 min at room temperature and...
then performed using an iQ SYBR-Green Supermix (cat. no. 1708880; Bio-Rad Laboratories, Inc.) according to the manufacturer's protocol. Thermocycling conditions were as follows: Initial denaturation of 15 min at 95˚C; followed by 40 cycles of denaturation at 94˚C for 30 sec, annealing at 56˚C for 30 sec and extension at 72˚C for 30 sec; with a final extension at 72˚C for 5 min. The 2^(-ΔΔCq) method was used to analyzed the mRNA levels of indicated genes. The primers were as follows: DYRK1A forward, 5'-TCTGGGATTCCACCTGTC-3' and reverse, 5'-GTCCCTCTGTTTTCACCTCA-3'; HIF-1α forward, 5'-ATCCATGTACCATGAGGAAGAAG TG-3' and reverse, 5'-TCCGGCTAAGATTAGGATACCTC-3'; and GAPDH forward, 5'-GAGTCAACGGATTTGGTCGT-3' and reverse, 5'-TTGATTTGGAGGGATCTCG-3'.

Co-immunoprecipitation (Co-IP) assay. For Co-IP assays, 293T cells overexpressing DYRK1A were lysed with IP buffer (cat. no. P0013; Beyotime Institute of Biotechnology) followed by centrifugation at 10,000 x g for 30 min at 4˚C. Subsequently, the cell lysates were incubated with 20 µl protein G magnetic beads at 4˚C for 1 h. Each Co-IP reaction was set up with equal quantities of lysate (1,000 µg). The lysates were incubated with the anti-STAT3 antibody (cat. no. sc-7148; 1:200 dilution; Santa Cruz Biotechnology, Inc.) and overnight with gentle shaking at 4˚C. Following incubation, the lysates were centrifuged at 3,000 x g for 5 min at 4˚C. Subsequently, the supernatant was carefully aspirated and the protein G magnetic beads were washed 3-4 times with lysis buffer, supplemented with SDS loading buffer and subjected to SDS-PAGE, as previously described (20).

Plasmid and small interfering RNA (siRNA) transfection. HepG2, Hep3B and HuH7 cells were seeded into six-well plates at a density of 3x10^4 cells/well. When reaching 30% confluency, cells were transfected with overexpression plasmids or siRNA using the jetPRIME reagent (Polyplus-transfection SA) according to the manufacturer's instructions. siRNAs against human DYRK1A and HIF-1α were synthesized by Shanghai GenePharma Co., Ltd. (20). The sense sequences of human siRNAs were as follows: siDYRK1A-1, 5'-AUGGAGGCUAUGGACGUAATT-3'; siDYRK1A-2, 5'-AAACUCCGAAUUCACCUATT-3'; siHIF-1α-1, 5'-GCUGAUGACCCAGAACCUCUGATT-3'; siHIF-1α-2, 5'-AUCCAGAGUCACUGGCAACUTT-3'; and negative control siRNA (siNC), 5'-UCUCCGAACGUGACACAGUTT-3'. The plasmid pcDNA3.1 containing the insert DYRK1A and empty vector pcDNA3.1 were purchased from GenScript. After 24 h siRNA transfection, the cells were treated with 2.5-20 µM sorafenib and/or 0.25-8 µM regorafenib (ranging from 0.25-8 µM) at 37˚C for 72 h. SRB was used to detect the cytotoxicity of cancer cells.

HIF-1α accumulation detection. HepG2 cells were pretreated with 20 µg/ml CHX under normoxic conditions for 4 h to inhibit synthesis of new proteins and HIF-1α degradation. The CHX-pretreated cells were then cultivated in fresh medium and cultured under hypoxic conditions to detect the accumulation of HIF-1α using western blot.

Bioinformatics analysis. The co-expressed genes of DYRK1A in the TCGA database were examined by cbioportal (www.cbioportal.org). The study ‘Liver Hepatocellular Carcinoma fixed with pre-cold 75% ethanol at 4˚C for 2 h followed by treatment with 400 µl PBS containing 50 µg/ml RNase A for 30 min at 37˚C. Subsequently, liver cancer cells were incubated with 5 µl PI solution for 15 min at room temperature and were then analyzed using FACSCalibur flow cytometer, as previously described (17).

Western blot analysis. Liver cancer cells were collected, then incubated with lysis buffer (cat. no. P0013; Beyotime Institute of Biotechnology) on ice for 30 min. The protein concentrations were determined by the BCA method using Enhanced BCA Protein Assay kit (cat. no. P0009; Beyotime Institute of Biotechnology). Proteins (40 µg) were separated by SDS-PAGE on 8-12% Tris-glycine gels. Following electrophoresis, the proteins were transferred to PVDF membranes, which were then blocked with 5% skimmed milk for 2 h at room temperature followed by washing with TBS + 0.1% Tween-20 (TBST) three times. Following incubation with primary antibodies overnight at 4˚C, the membranes were washed thrice with TBST and then incubated with the corresponding secondary antibodies (DyLight™ 800 4X PEG-conjugated anti-rabbit-IgG; DyLight™ 800 4X PEG-conjugated anti-mouse-IgG; 1:50,000; Cell Signaling Technology) for 2 h at room temperature. Finally, after washing three times with TBST, 5 min each, the bands were visualized using an LiCor Odyssey CLx imager and Odyssey CLX Image Studio software version 5.0.21 (LI-COR Biosciences) (17). ImageJ (version: 1.52a; National Institutes of Health) was used to semi-quantitatively analyze protein expression levels.

Reverse transcription-quantitative PCR (RT-qPCR). Total RNA was extracted from liver cancer cells using TRizol® reagent (Invitrogen; Thermo Fisher Scientific, Inc.), precipitated with isopropl alcohol and rinsed with 70% ethanol. Subsequently, total RNA was reverse transcribed into cDNA using the iScript cDNA Synthesis kit according to the manufacturer's protocol (Bio-Rad Laboratories, Inc.). qPCR was then performed using an iQ SYBR-Green Supermix (cat. no. 1708880; Bio-Rad Laboratories, Inc.) according to the manufacturer’s protocol. Thermocycling conditions were as follows: Initial denaturation of 15 min at 95˚C; followed by 40 cycles of denaturation at 94˚C for 30 sec, annealing at 56˚C for 30 sec and extension at 72˚C for 30 sec; with a final extension at 72˚C for 5 min. The 2^(-ΔΔCq) method was used to analyze the mRNA levels of indicated genes. The primers were as follows: DYRK1A forward, 5'-TCTGGGATTCCACCTGTC-3' and reverse, 5'-GTCCCTCTGTTTTCACCTCA-3'; HIF-1α forward, 5'-ATCCATGTACCATGAGGAAGAAG TG-3' and reverse, 5'-TCCGGCTAAGATTAGGATACCTC-3'; and GAPDH forward, 5'-GAGTCAACGGATTTGGTCGT-3' and reverse, 5'-TTGATTTGGAGGGATCTCG-3'.

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(TCGA, Provisional) (n=442) was selected to evaluate Pearson's correlation coefficient between the expression of DYRK1A and HIF-1α or DYRK1A and HIF-2α in liver cancer (17,18).

Statistical analysis. All experiments were conducted at least three times. All data are presented as the mean ± SD. A two-tailed unpaired Student's t-test was used to determine the differences between two groups. One-way ANOVA followed by Tukey's post hoc test was used to examine the differences among multiple groups. Statistical analyses were performed using GraphPad Prism version 7 (GraphPad Software, Inc.). P<0.05 was considered to indicate a statistically significant difference. The combination index (CI) was calculated using CalcuSyn software (version 2.0; Biosoft). The CI value indicated the interaction of the treatment combination as follows: CI <0.9, synergism; 0.9‑1.10, additive; and >1.10, antagonism.

Results

DYRK1A positively regulates HIF-1α in liver cancer cells under hypoxic conditions. As shown in Fig. 2A, the expression levels of DYRK1A in liver cancer were positively correlated with those of HIF-1α (Pearson's correlation test, r=0.50;
Figure 3. DYRK1A regulates the expression of HIF-1α via STAT3 signaling in liver cancer cells. (A) Western blot analysis of the protein expression levels of HIF-1α in HepG2 and Hep3B cell lines treated with 20 µg/ml CHX with or without 10 µM harmine for 30, 90 or 180 min. A two-tailed unpaired Student's t-test was used to determine the differences between DMSO-treated group and harmine-treated group. (B) HepG2 cells pretreated with CHX for 4 h were co-treated with the indicated siRNA clones for 48 h under normoxic conditions. Subsequently, cells were cultured in new medium and treated with or without harmine for the indicated time points under hypoxic conditions. Western blot analysis was used to detect the protein expression levels of DYRK1A and HIF-1α. (C) Reverse transcription-quantitative PCR analysis of DYRK1A and HIF-1α mRNA expression levels in HepG2 cells transfected with the indicated siRNA clones for 48 h. A two-tailed unpaired Student's t-test was used to determine the differences between DYRK1A siRNA and siNC. **P<0.01 and ***P<0.001. (D) Co-immunoprecipitation analysis of the interaction between DYRK1A and STAT3 in 293T cells transfected with DYRK1A overexpression plasmid for 48 h. (E) Western blot analysis of the protein expression levels of STAT3 and p-STAT3 (Tyr705) in HepG2 and Hep3B cell lines treated with the indicated siRNA clones for 48 h and then cultured under normoxic or hypoxic conditions for 6 h. (F) Following transfection for 48 h, liver cancer cells were treated with or without 20 µg/ml IL-6 for 1 h. The expression levels of the associated proteins, including STAT3, p-STAT3 (Tyr705), AKT, p-AKT(Ser473), and HIF-1α, were detected by western blot analysis. CHX, cycloheximide; DYRK1A, dual-specificity tyrosine-phosphorylation-regulated kinase 1A; HIF-1α, hypoxia inducible factor-1α; NS, not significant; p, phosphorylated; si, small interfering RNA.
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P=6.63x10⁻²⁴) as well as HIF-2α (Pearson’s correlation test, r=0.30 and P=5.74x10⁻⁹) (17). These results were collected from cBioPortal and suggested that DYRK1A expression exhibited a stronger positive correlation with HIF-1α expression compared the correlation between DYRK1A and HIF-2α in liver cancer, indicated that DYRK1A co-expressed with HIF-1α in liver cancer patients.

Furthermore, the in vitro transfection results revealed that HIF-1α protein expression was markedly increased in the siNC group under hypoxia compared with normoxia. However, the protein expression levels of HIF-1α were notably reduced in the DYRK1A knockdown group compared with siNC group under hypoxia (Fig. 2B). Additionally, cell treatment with the DYRK1A inhibitor harmine downregulated HIF-1α in liver cancer cells (Fig. 2C). Conversely, DYRK1A overexpression in liver cancer cells notably upregulated HIF-1α compared with empty vector group under hypoxia (Fig. 2D). However, HIF-1α silencing did not affect the expression of DYRK1A in liver cancer cells (Fig. 2E). Furthermore, liver cancer cell lines overexpressing DYRK1A exhibited upregulated expression of HIF-1α under hypoxia (Fig. 2F). The aforementioned findings indicated that DYRK1A could be a potential upstream activator of HIF-1α in liver cancer cells.

DYRK1A regulates the expression of HIF-1α via STAT3 signaling. STAT3 directly binds to the HIF-1α gene promoter to transcriptionally regulate its expression (20). Co-IP results indicated that DYRK1A may directly interact with STAT3 (Fig. 3D). Furthermore, DYRK1A knockdown reversed the hypoxia-induced activation of STAT3 compared with control siRNA group (Fig. 3E). Additionally, the IL-6-induced phosphorylation of STAT3 and AKT expression was attenuated by DYRK1A depletion (Fig. 3F). These findings indicated that DYRK1A may regulate the expression of HIF-1α via the STAT3 signaling pathway.

DYRK1A knockdown inhibits the regorafenib-mediated liver cancer cell viability under hypoxia. We hypothesized that suppressing DYRK1A expression may be a novel strategy for overcoming drug resistance in liver cancer treatment. As shown in Fig. 4A and B, harmine inhibited the viability of liver cancer cells in a dose-dependent manner under hypoxic conditions. Sorafenib and regorafenib are the recommended first- and second-line therapies for patients with advanced liver cancer, respectively, and the Food and Drug Administration (FDA) has approved regorafenib for treating patients with liver cancer who progress after sorafenib failure (21,22). We hypothesized that DYRK1A may regulate the expression of HIF-1α and
be associated with resistance to sorafenib and regorafenib under hypoxic conditions. Under hypoxia, DYRK1A silencing significantly inhibited the viability of regorafenib-treated liver cancer cells (Fig. 5A). Furthermore, co-treatment of liver cancer cells with harmine and regorafenib exhibited synergistic anti-proliferative effects on liver cancer cells under hypoxia (Fig. 5B and C).

Harmine enhances the apoptotic effect of regorafenib on liver cancer cells under hypoxic conditions. To further verify the synergistic anti-liver cancer effect of harmine and regorafenib, PI staining was carried out to evaluate cell death. As demonstrated in Fig. 6A and B, under hypoxia, harmine plus regorafenib enhanced the apoptotic effect compared with harmine or regorafenib alone treatment on HepG2 and Hep3B cells. In addition, western blot analysis was performed to detect the expression levels of apoptosis-related proteins, cleaved caspase 3 and cleaved PARP, and the results confirmed that under hypoxic conditions harmine treatment in combination with regorafenib notably promoted liver cancer cell apoptosis compared with harmine or regorafenib single agent treatment (Fig. 6C). Furthermore, harmine plus regorafenib treatment markedly downregulated Mcl-1 protein expression levels compared with cell treatment with each agent alone, suggesting that the combination of harmine and regorafenib could induce apoptosis via the mitochondrial pathway. In addition, harmine combined with regorafenib markedly inhibited the protein expression levels HIF-1α and the phosphorylation of STAT3 and AKT compared with harmine or regorafenib single agent treatment (Fig. 6D). Overall, these results indicated that harmine plus regorafenib may enhance liver cancer cell apoptosis under hypoxic conditions.

DYRK1A downregulation enhances the anti-liver cancer activity of sorafenib under hypoxia. As shown in Fig. 7A, DYRK1A knockdown decreased the viability of sorafenib-treated liver cancer cells compared with control siRNA group under hypoxic conditions. Furthermore, cell
co-treatment with harmine and sorafenib for 72 h significantly reduced the viability of liver cancer cells compared with cells treated with each agent alone (Fig. 7B). CI value analysis revealed that harmine may work synergistically with sorafenib under hypoxic culture conditions (CI<0.9; Fig. 7C). Furthermore, colony formation assays showed that treatment with harmine plus regorafenib enhanced the inhibition of liver cancer cell proliferation compared with harmine or regorafenib single agent alone under hypoxic conditions, the same phenomenon was also observed in harmine plus sorafenib combination treatment (Fig. 7D). In addition, co-treatment with harmine and sorafenib further promoted cell death in...
HepG2 cells, as indicated by the upregulated expression levels of cleaved PARP and the downregulated expression of caspase 3 (Fig. 7E). Collectively, these findings suggested that targeting DYRK1A could be an effective approach for preventing hypoxia-induced liver cancer progression and therapy resistance.

**Discussion**

DYRK1A serves a crucial role in cell growth and tumorigenesis; it exhibits both tumor suppressive and oncogenic activities, depending on the tissue (23). More specifically, DYRK1A acts as an oncogenic protein in liver cancer. For example, a previous study demonstrated that DYRK1A could interact with trophinin-associated protein and promote cell growth by activating AKT/glycogen synthase kinase-3β signaling (24), which suggested that targeting DYRKA may be a promising therapeutic strategy for treating patients with liver cancer. Furthermore, DYRK1A is a serine/threonine kinase that phosphorylates multiple substrates associated with resistance to cancer chemotherapy, such as Met and Mcl-1 (10,12). It has been reported that DYRK1 phosphorylates ID2 on Thr27 to destabilize HIF-2α, but not HIF-1α, in a transcription-independent manner in glioblastoma under normoxic conditions (13). However, the role of DYRK1A in drug resistance under hypoxic conditions remains unclear. To the best of our knowledge, the present study demonstrated for the first time that DYRK1A could positively regulate HIF-1α expression in liver cancer cells. Additionally, DYRK1A expression showed a more potent positive correlation with HIF-1α compared with HIF-2α in tumor tissue samples derived from patients with liver cancer from the cBioPortal database. DYRK1A and HIF-1α co-expression was associated with a poor prognosis in liver cancer. The results of the present study further revealed that DYRK1A-mediated HIF-1α expression could promote liver cancer chemoresistance in a hypoxic tumor microenvironment. Although DYRK1A regulated the expression of HIF-1α, HIF-1α still could be regulated by traditional signal pathway under hypoxia, such as PI3K and MAPK (25,26). Lee et al (13) and the present study demonstrated that DYRK1A is inhibited by hypoxia; it was hypothesized that it may act as a feedback loop when HIF-1α is activated by PI3K or MAPK pathway under hypoxia in cancer cells. However, the hypothesis needs further investigation.
Although sorafenib and regorafenib have been approved by the FDA to treat liver cancer, numerous resistance mechanisms including hypoxia, fibrosis, autophagy, inflammation, angiogenesis, oxidative stress and epithelial-mesenchymal transition impair the efficacy of these drugs, which still need to be resolved (27-29). Therefore, the identification of new compounds is urgently needed to overcome sorafenib and regorafenib resistance (30). Harmine, isolated from the seeds of the medical plant *Peganum harmala L.*, is a high-affinity ATP-competitive inhibitor of DYRK1A (31,32). The present data demonstrated that under hypoxic conditions, harmine reduced proliferation of liver cancer cells by regulating HIF-1α expression. However, the regulation of DYRK1A was not obvious after 48 h treatment of harmine in liver cancer cell, thus we hypothesized that long exposure of hypoxia may enhance the accumulation of DYRK1A and HIF-1α. Furthermore, DYRK1A suppression increased the anti-proliferative and apoptosis induction activities of sorafenib and regorafenib by regulating HIF-1α in liver cancer cells. Thus, the present findings indicated a new strategy to combat chemoresistance against liver cancer under hypoxia, but the efficacy of combining harmine with sorafenib or regorafenib in vivo may need further investigation.

DYRK1A phosphorylates the Ser-727 residue of STAT3, which is critical for B cell acute lymphoblastic leukemia development (33,34). STAT3, along with HIF1, activates HIF1 target genes and drives HIF1-dependent tumorigenesis under hypoxic conditions (35). Recently, a study reported that DYRK1A can modulate the STAT3/EGFR/Met signaling pathway and that DYRK1A inhibition can facilitate the anticancer effect of AZD9291, a third-generation epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor, through the STAT3 axis in EGFR wild-type NSCLC cells (11). The present findings indicated that DYRK1A directly interacts with STAT3 and may regulate the expression of HIF-1α via STAT3 signaling in liver cancer cells. However, the molecular mechanism of DYRK1A in regulating STAT3 and HIF-1α may need further investigation. In addition, DYRK1A suppression may sensitize liver cancer cells to sorafenib and regorafenib treatment through the STAT3/HIF-1α/AKT axis. The data of the present study may thus explain the mechanism by which DYRK1A promotes resistance. Moreover, the results indicated that the DYRK1A inhibitor may be a chemosensitizing therapeutic agent in combination with sorafenib or regorafenib treatment.

In conclusion, the present study indicated that DYRK1A regulated the expression of HIF-1α through STAT3. DYRK1A suppression may sensitize liver cancer cells to sorafenib and regorafenib treatment through the STAT3/HIF-1α/AKT axis. The present data identified a new DYRK1A/HIF-1α pathway in chemoresistance, and DYRK1A inhibitor plus sorafenib or regorafenib might be efficacious regimens to treat liver cancer.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors’ contributions

CZ, LHZ and YLL conceptualized the study. LWW, ZDL, MMZ and FHD curated the data. LWW, ZDL, FHD, MMZ and JW formally analyzed the data. CZ, LHZ and YLL supervised the study. CZ wrote the original draft. CZ, ZDL, LHZ, MMZ and YLL wrote, revised and edited the manuscript. YLL, ZDL, LWW, MMZ, and CZ confirm the authenticity of all the raw data. All authors read and approved the final version of the manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

1. Chen C and Lou T: Hypoxia inducible factors in hepatocellular carcinoma. Oncotarget 8: 46691-46703, 2017.
2. McKeown SR: Defining normoxia, physioxia and hypoxia in tumours-implications for treatment response. Br J Radiol 87: 20130676, 2014.
3. Liu Z, Tu K, Wang Y, Yao B, Li Q, Wang L, Dou C, Liu Q and Zheng X: Hypoxia accelerates aggressiveness of hepatocellular carcinoma cells involving oxidative stress, epithelial-mesenchymal transition and non-canonical hedgehog signaling. Cell Physiol Biochem 44: 1856-1868, 2017.
4. Lau CK, Yang ZF, Ho DW, Ng MN, Yeoh GC, Poon RT and Fan ST: An Akt/hypoxia-inducible factor-1alpha/platelet-derived growth factor-BB autocrine loop mediates hypoxia-induced chemoresistance in liver cancer cells and tumorigenic hepatic progenitor cells. Clin Cancer Res 15: 3462-3471, 2009.
5. Koh MY, Spivak-Kroizman TR and Powis G: HIF-1alpha and cancer therapy. Recent Results Cancer Res 180: 15-34, 2010.
6. Tang W and Zhao G: Small molecules targeting HIF-1alpha pathway for cancer therapy in recent years. Biogor Med Chem 28: 115235, 2020.
7. Kargbo RB: Selective DYRK1A inhibitor for the treatment of neurodegenerative diseases: Alzheimer, parkinson, huntington, and down syndrome. ACS Med Chem Lett 11: 1795-1796, 2020.
8. Wegiel J, Gong CX and Hwang YW: The role of DYRK1A in neurodegenerative diseases. FEBS J 278: 236-245, 2011.
9. Laham AJ, Saber-Ayad M and El-Awady R: DYRK1A: A down syndrome-related dual protein kinase with a versatile role in tumorigenesis. Cell Mol Life Sci 78: 603-619, 2021.

10. Luna J, Boni J, Cuatrecasas M, Botíll-De Ros X, Núñez-Manchón E, Gironella M, Vaquero EC, Arbones ML, de la Luna S and Fillat C: DYRK1A modulates e-MET in pancreatic ductal adenocarcinoma to drive tumour growth. Gut 68: 1465-1476, 2019.

11. Li YL, Ding K, Hu X, Wu LW, Zhou DM, Rao MJ, Lin NM and Zhang C: DYRK1A inhibition suppresses STAT3/EGFR/Met signalling and sensitizes EGFR wild-type NSCLC cells to AZD9291. J Cell Mol Med 23: 7427-7437, 2019.

12. Li Y, Zhou D, Xu S, Rao M, Zhang Z, Wu L, Zhang C and Lin N: DYRK1A suppression restrains McI-1 expression and sensitizes NSCLC cells to Bcl-2 inhibitors. Cancer Biol Med 17: 387-400, 2020.

13. Lee SB, Frattini V, Bansal M, Castano AM, Sherman D, Hutchinson K, Bruce JN, Calafano A, Liu G, Cardozo T, et al: An ID2-dependent mechanism for VHL inactivation in cancer. Nature 529: 172-177, 2016.

14. Dituri F, Scialpi R, Schmidt RA, Frusciante M, Mancarella S, Dituri F, Scialpi R, Schmidt TA, Stefanovic M, Boix L, Cucarull B, Zamora A, Méndez-Blanco C, Fondevila F, García-Palomo A, González J and Muntané J: Differential effectiveness of tyrosine kinase inhibitors in 2D/3D culture according to cell differentiation, p53 status and mitochondrial respiration in liver cancer cells. Cell Death Dis 11: 339, 2020.

15. Dituri F, Scialpi R, Schmidt RA, Frusciante M, Mancarella S, Dituri F, Scialpi R, Schmidt TA, Stefanovic M, Boix L, Cucarull B, Zamora A, Méndez-Blanco C, Fondevila F, García-Palomo A, González J and Muntané J: Differential effectiveness of tyrosine kinase inhibitors in 2D/3D culture according to cell differentiation, p53 status and mitochondrial respiration in liver cancer cells. Cell Death Dis 11: 339, 2020.

16. Tutusaus A, Stefanovic M, Boix L, Cucarull B, Zamora A, Blasco L, de Frutos PG, Reig M, Fernandez-Checa JC, Mari M, et al: Antiapoptotic BCL-2 proteins determine sorafenib/regorafenib resistance and BH3-mimetic efficacy in hepatocellular carcinoma. Oncotarget 9: 16701-16717, 2018.

17. Gao J, Ajkoy BA, Dogrusoz U, Dresdner G, Gross BE, Sumer SO, Jacobsen A, Byrne CJ, Heuer ML, Larsson E, et al: Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal 6: pl1, 2013.

18. Cerami E, Gao J, Dogrusoz U, Gross B, Sumer SO, Ajkoy BA, Jacobsen A, Byrne CJ, Heuer ML, Larsson E, et al: The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov 2: 401-404, 2012.

19. Aguirre-Gamboa R, Gomez-Rueda H, Martinez-Ledesma E, Martinez-Torteya A, Chacolla-Huaringa R, Rodriguez-Barrientos A, Tamez-Peña JG and Treviño V: SurvExpress: an online biomarker validation tool and database for cancer gene expression data using survival analysis. PLoS One 8: e74250, 2013.

20. Carpenter RL and Lo HW: STAT3 target genes relevant to human cancers. Cancers (Basel) 6: 897-925, 2014.

21. Rodríguez-Hernández MA, Chapresto-Garzón R, Cadenas M, Navarro-Villarán E, Negrete M, Gómez-Bravo MA, Victó VM, Padillo FJ and Muntané J: Differential effectiveness of tyrosine kinase inhibitors in 2D/3D culture according to cell differentiation, p53 status and mitochondrial respiration in liver cancer cells. Cell Death Dis 11: 339, 2020.

22. Wu LW, Zhou DM, Zhang ZY, Zhang JK, Zhu HJ, Lin NM and Zhang C: Suppression of LSD1 enhances the cytotoxic and apoptotic effects of regorafenib in hepatocellular carcinoma cells. Biochem Biophys Res Commun 512: 852-858, 2019.

23. Fernandez-Martinez P, Zahnoro C and Sanchez-Gomez P: DYRK1A: The double-edged kinase as a protagonist in cell growth and tumorigenesis. Mol Cell Oncol 2: e970048, 2015.

24. Li L, Wei JR, Song Y, Fang S, Du Y, Li Z, Zeng TT, Zhu YH, Li Y and Guan XY: TROAP switches DYRK1 activity to drive hepatocellular carcinoma progression. Cell Death Dis 12: 125, 2021.

25. Zheng Z, Yao L, Yang J, Wang Z and Du G: PI3K/Akt and HIF1 signaling pathway in hypoxia-ischemia (review). Mol Med Rep 18: 3547-3554, 2018.

26. Kim HS, Kang YH, Lee J, Han SR, Kim DB, Ko H, Park S and Lee MS: Biphasic regulation of mitogen-activated protein kinase phosphorylation in hypoxic colon cancer cells. Mol Cells 44: 710-722, 2021.

27. Juengpanich S, Topatana W, Lu C, Staiculescu D, Li S, Cao J, Lin J, Hu J, Chen M, Chen J and Cai X: Role of cellular, molecular and tumor microenvironment in hepatocellular carcinoma: Possible targets and future directions in the regorafenib era. Int J Cancer 147: 1778-1792, 2020.

28. Liang C, Dong Z, Cai X, Shen J, Xu Y, Zhang M, Li H, Yu W and Chen W: Hypoxia induces sorafenib resistance mediated by autophagy via activating FOXO3a in hepatocellular carcinoma. Cell Death Dis 11: 1017, 2020.

29. Tomida C, Aibara K, Yamagishi Y, Nishio H, Tanaka R, Abe T, Ohno A, Hirasaka K, Nakawa K and Teshima-Kondo S: The malignant progression effects of regorafenib in human colon cancer cells. J Med Invest 62: 195-198, 2015.

30. Suk FM, Liu CL, Hsu MH, Chuang YT, Wang JP and Liao YJ: Treatment with a new benzimidazole derivative bearing a pyrrolidine side chain overcomes sorafenib resistance in hepatocellular carcinoma. Sci Rep 9: 17259, 2019.

31. Rüben K, Wurlzbauer A, Walte A, Sippl W, Bracher F and Ohno A, Hirasaka K, Yamagishi Y, Nishio H, Tanaka R, Abe T, Ohno A, Hirasaka K, Nakawa K and Teshima-Kondo S: The malignant progression effects of regorafenib in human colon cancer cells. J Med Invest 62: 195-198, 2015.

32. Suk FM, Liu CL, Hsu MH, Chuang YT, Wang JP and Liao YJ: Treatment with a new benzimidazole derivative bearing a pyrrolidine side chain overcomes sorafenib resistance in hepatocellular carcinoma. Sci Rep 9: 17259, 2019.

33. Bhansali RS, Rammohan M, Lee P, Laurent AP, Wen Q, Suraneni P, Yip BH, Tsai YC, Jenni S, Bornhauser B, et al: DYRK1A regulates B cell acute lymphoblastic leukemia through phosphorylation of FOXO1 and STAT3. J Clin Invest 131: e135937, 2021.

34. Wiechmann S, Czajkowska H, de Graaf K, Grotzinger J, Göckler N, Jofre G, Heuer ML, Larsson E, et al: Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal 6: pl1, 2013.