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Colon cancer-associated B2 *Escherichia coli* colonize gut mucosa and promote cell proliferation

Jennifer Raisch, Emmanuel Buc, Mathilde Bonnet, Pierre Sauvanet, Emilie Vazeille, Amélie de Vallée, Pierre Déchelotte, Claude Darcha, Denis Pezet, Richard Bonnet, Marie-Agnès Bringer, Arlette Darfeuille-Michaud

**Abstract**

**AIM:** To provide further insight into the characterization of mucosa-associated *Escherichia coli* (*E. coli*) isolated from the colonic mucosa of cancer patients.

**METHODS:** Phylogroups and the presence of cyclo-modulin-encoding genes of mucosa-associated *E. coli* from colon cancer and diverticulosis specimens were determined by PCR. Adhesion and invasion experiments were performed with I-407 intestinal epithelial cells using gentamicin protection assay. Carcinomembranic antigen-related cell adhesion molecule 6 (CEACAM6) expression in T84 intestinal epithelial cells was measured by enzyme-linked immunosorbent assay and by Western Blot. Gut colonization, inflammation and pro-carcinogenic potential were assessed in a chronic infection model using CEABAC10 transgenic mice. Cell proliferation was analyzed by real-time mRNA quantification of PCNA and immunohistochemistry staining of Ki67.

**RESULTS:** Analysis of mucosa-associated *E. coli* from colon cancer and diverticulosis specimens showed that whatever the origin of the *E. coli* strains, 86% of cyclo-modulin-positive *E. coli* belonged to B2 phylogroup and most harbored polyketide synthase (*pks*) island, which encodes colibactin, and/or cytotoxic necrotizing factor (*cnf*) genes. In vitro assays using I-407 intestinal epithelial cells revealed that mucosa-associated B2 *E. coli* strains were poorly adherent and invasive. However, mucosa-associated B2 *E. coli* similarly to Crohn’s disease-associated *E. coli* are able to induce CEACAM6 expression in T84 intestinal epithelial cells. In addition, in vivo experiments using a chronic infection model of CEACAM6 expressing mice showed that B2 *E. coli* strain 11G5 isolated from colon cancer is able to highly persist in the gut, and to induce colon inflammation, epithelial damages and cell proliferation.

**CONCLUSION:** In conclusion, these data bring new insights into the ability of *E. coli* isolated from patients with colon cancer to establish persistent colonization, exacerbate inflammation and trigger carcinogenesis.

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**Key words:** B2 *Escherichia coli*; Carcinomembranic antigen-related cell adhesion molecule 6; Cell proliferation;
Colon cancer; Polyketide synthase genomic island

Core tip: Tumors and mucosa of patients with colon cancer are abnormally colonized by Escherichia coli (E. coli) belonging to B2 phylogroup. The aim of the present study was to provide further insight into the characterization of colon cancer-associated E. coli. Despite their poor ability to adhere to and to invade intestinal epithelial cells in vitro, we showed that colon cancer-associated E. coli induce carcinoembryonic antigen-related cell adhesion molecule 6 (CEACAM6) expression, a receptor involved in adhesion of pathogenic E. coli. These bacteria were also able to persist and promote low grade inflammation and cell proliferation, in a chronic infection model of CEACAM6 expressing mice, highlighting their oncogenic potential.

Patients with IBD have an increased risk of colon cancer and small bowel adenocarcinoma[6,17]. As in colon cancer patients, dysbiosis toward selected micro-organisms and decreased complexity of commensal bacteria have been observed in patients with Crohn’s disease (CD) and ulcerative colitis (UC), but it is not clear whether dysbiosis contributes to the development of IBD or is instead a consequence of the disease. Patients with IBD, compared to healthy controls, have fewer bacteria with anti-inflammatory properties and/or more bacteria with pro-inflammatory properties. Several metagenomic-based studies reported that members of the phyla Bacteroidetes and Firmicutes were reduced in patients with CD or UC[18-20]. Among the Firmicutes, Faecalibacterium prausnitzii has anti-inflammatory properties; its numbers are reduced in patients with CD and associated with a risk of post-resection recurrence of ileal CD[28]. In contrast, a greater relative abundance of Enterobacteriaceae, mostly E. coli belonging to the B2 phylogenetic group, has been reported in CD patients more notably on mucosa-associated microbiota than in fecal samples[28,29,30,21-24]. Intestinal colonization by E. coli correlates with bacterial adhesion of CD-associated E. coli strains to intestinal epithelial cells[10,25]. CD-associated E. coli share abilities to adhere to and to invade intestinal epithelial cells and to survive within macrophages[26,27] and they are termed accordingly adherent-invasive E. coli (AIEC). The abnormal colonization of CD mucosa by AIEC involves abnormal expression of a host receptor, the carcinoembryonic antigen-related cell adhesion molecule 6 (CEACAM6) [28]. Interestingly, CEACAM6 is not only abnormally expressed in the ileum of patients with CD but expression of this molecule is also up-regulated in proliferating cells in adenomas and colorectal tumors[25,28]. However, the origin

INTRODUCTION

Colorectal cancer (CRC) is the fourth leading cause of cancer death and is responsible for about 610000 deaths per year worldwide[1]. Although many etiologic genetic changes are associated with progression from adenomatous lesions to invasive carcinoma[2], the specific causative factors in the development of sporadic CRC remain unclear. Accumulating evidence supports that inflammation and gut microbial communities influence the development of colorectal carcinoma[1-3]. Two theories have emerged to explain the contribution of bacteria in CRC: (1) the “alpha bug” concept, wherein select members of a microbial community with virulence and pro-carcinogenic features are capable of remodeling the microbiome as a whole to drive pro-inflammatory immune responses and colonic epithelial cell transformation leading to cancer[8]; and (2) the “driver-passenger” concept, wherein certain indigenous intestinal bacteria, termed “bacteria drivers”, initiate CRC by inducing epithelial DNA damages: the resulting tumorigenesis induces intestinal niche alterations that promote the proliferation of opportunistic opportunistic bacteria with a growth advantage in the tumour microenvironment[9].

Dysbiosis of the intestinal microbiota has been observed in CRC patients. Recent pyrosequencing data of CRC-associated bacterial microbiota have revealed, in particular, over-representation of some bacteria such as Bacteroides/Prevotella, Faecalibacterium and Fusobacterium[8,9]. In addition, independent studies show that colonic adenomas, carcinomas and the mucosa of CRC patients are abnormally colonized by high numbers of adherent Escherichia coli (E. coli) compared to controls[10-12]. It has been suggested that the role of E. coli in CRC promotion and development is related to chronic inflammation. Inflammation can result from bacterial infection, via its effects on both the host and the microbiota, in particular by promoting the expansion of E. coli, which actively contribute to the accumulation of mutations resulting from DNA damages induced by genotoxins, or by downregulating host DNA mismatch repair proteins[1,3,13]. In particular, E. coli strains harboring the polyketide synthase (pks) genotoxic island, which are found in a significantly high percentage of inflammatory bowel disease (IBD) and CRC patients, can promote invasive carcinoma in monocolonized azoxymethane (AOM)-treated Il10−/− mice[3]. In addition, certain pathogenic bacteria can also be involved in cancer development, like, for example enterotoxigenic Bacteroides fragilis (ETBF), a common human commensal bacterium that is associated with colon cancer[14]. ETBF-induced chronic inflammation and tumorigenesis in Apo-luciferase mice (a mouse model of familial adenomatous polyposis) involve the induction of the polylamine catabolic enzyme spermine oxidase, which causes DNA damages and uncontrolled cell proliferation in intestinal epithelial cells[15].

Patients with IBD have an increased risk of colon cancer and small bowel adenocarcinoma[6,17]. As in colon cancer patients, dysbiosis toward selected micro-organisms and decreased complexity of commensal bacteria have been observed in patients with Crohn’s disease (CD) and ulcerative colitis (UC), but it is not clear whether dysbiosis contributes to the development of IBD or is instead a consequence of the disease. Patients with IBD, compared to healthy controls, have fewer bacteria with anti-inflammatory properties and/or more bacteria with pro-inflammatory properties. Several metagenomic-based studies reported that members of the phyla Bacteroidetes and Firmicutes were reduced in patients with CD or UC[18-20]. Among the Firmicutes, Faecalibacterium prausnitzii has anti-inflammatory properties; its numbers are reduced in patients with CD and associated with a risk of post-resection recurrence of ileal CD[28]. In contrast, a greater relative abundance of Enterobacteriaceae, mostly E. coli belonging to the B2 phylogenetic group, has been reported in CD patients more notably on mucosa-associated microbiota than in fecal samples[28,29,30,21-24]. Intestinal colonization by E. coli correlates with bacterial adhesion of CD-associated E. coli strains to intestinal epithelial cells[10,25]. CD-associated E. coli share abilities to adhere to and to invade intestinal epithelial cells and to survive within macrophages[26,27] and they are termed accordingly adherent-invasive E. coli (AIEC). The abnormal colonization of CD mucosa by AIEC involves abnormal expression of a host receptor, the carcinoembryonic antigen-related cell adhesion molecule 6 (CEACAM6) [28]. Interestingly, CEACAM6 is not only abnormally expressed in the ileum of patients with CD but expression of this molecule is also up-regulated in proliferating cells in adenomas and colorectal tumors[25,28]. However, the origin
of CEACAM6 overexpression in colon cancer is not yet clearly understood.

The aim of the present study was to provide further insight into the characterization of mucosa-associated E. coli isolated from the colonic mucosa of cancer patients. We determined their ability to interact with intestinal epithelial cells, with a particular focus on biofilm formation and the presence of cyclomodulin-encoding genes, and to induce CEACAM6 expression in intestinal epithelial cells. Finally, using CEABAC10 transgenic mice expressing human CEACAMs, we assessed the effects of long-term chronic infection by the colon cancer-associated E. coli strain 11G5 for its ability to colonize the gut, to potentiate inflammation and to induce cell proliferation.

MATERIALS AND METHODS

Ethical considerations

Ethical approval for the study was granted by the Clermont-Ferrand research ethics committee. This IRB allowed for the waiver of written consent from potential subjects, because the research involved no procedures for which written consent is normally required outside of the research context and presents no risk of harm to subjects. The biological samples were collected from colorectal resections, which were required for the treatment of patients. The investigators explained the study to the potential subject verbally, providing all pertinent information such as purpose, procedures and putative risks. Following this verbal explanation, the potential subject was provided with a study information sheet. After allowing the potential subject time to read the study information sheet, the investigators answered any additional questions the subject may have. A verbal agreement to participate in the research was obtained for all patients included in the study. The dates of verbal consent were tracked in a non-identifiable manner.

Patients

Eighty-one patients were studied between March 2007 and July 2010 at the University of Clermont-Ferrand, France, 48 with colon cancer (adenocarcinoma), and 33 with diverticulosis. For ethical considerations no surgical specimens from healthy patients were included and diverticulosis specimens were used as non-neoplastic controls. Among patients with diverticulosis, we excluded one representative isolate was subsequently analysed and confirmed with the automated Vitek II® (bioMérieux) system. When possible a maximum of 96 E. coli isolates per sample were collected for molecular typing. The bacteria were subcultured for 24 h at 37 °C in 96-well plates in Luria Bertani medium, supplemented with 15% glycerol and then stored at -80 °C.

Molecular phylogenetic grouping and PCR assay for cyclomodulin and adhesin-encoding genes

Ten isolates per sample were typed with molecular methods to identify the E. coli isolates (E. coli genotypes) colonizing the samples. Two genotyping methods were used: an “Enterobacterial Repetitive Intergenic Consensus” sequence (ERIC)-PCR using primer ERIC2 (5’ AAGTAAAGTGCATGGGTGAGCG 3’) and a “Random Amplified Polymorphic DNA” (RAPD)-PCR using primer 1283 (5’ GCGATCCCC 3’), or intimin (eae), adhesin (afa, afa/dr and aaf), or cyclomodulin (cycA) encoding genes, PCR assays were performed using primers listed in Table 1.

Cell culture

The intestinal epithelial cell lines T84 (ATCC, CCL-248) and Intestine-407 (I-407; ATCC, CCL-6) were maintained in an atmosphere containing 5% CO₂ at 37 °C in the culture medium recommended by ATCC. For infection assays, cells were seeded in 24-well plates at a density of 2 × 10⁵/cm².
Table 1  List of primers used for PCR assays

| Primer name | Sequence (5’-3’) | Region specific for |
|-------------|-----------------|---------------------|
| afa-f       | CGGCTTTTCTGCTGAACTGGACGAGCC | afaC [49] |
| afa-r       | CGGCTTCTCCAGTAGACTGGAATGAAGCG | afaC |
| afa1        | CTTGGGCAGCAGAACATGATATACTTCC | afaBC |
| afa2        | GTTCAGCCTGCTGTTGCTGCCCGC | afaE1 |
| afa3        | TTAGAGGTCACGTCTGGGTTGCC | afaE2 |
| dae-r       | CCCTATTTGTTGTCGCTGATCAGGAA | daeE |
| afa5-f      | TCAACTACCACTGAGATGCGGAC | afaE5 |
| afa5-r      | AGCAAGGTTGACCCACGGTACG | afaE5 |
| afa7-f      | GTCTTTTTCATGCTGGCAGGACCAT | afaE7 |
| afa7-r      | GGACATACCAATAGGCGGAATTA | afaE7 |
| afa8-f      | CTAAGGCATGTGACAGGATCGTA | afaE8 |
| afa8-r      | TGATCCTTCTGAGTGTGATACT | afaE8 |
| aggR        | CTGAGATTCACGGATCGTCA | aggR [50] |
| pksorf9-10  | ATCGTATAGCGTCCACCAAC | cibK-cibl [51] |
| pksorf9-10  | GGATGCTTCAAGTGGATCAGTG | cibK-cibl |
| CNF-1s      | GGAGAAGATGACAGAAGATTA | cfnf1 [51] |
| CNF-1as     | TGGCCGCTACTCTCAGCATG | cfnf1 |
| CNF-2s      | TATCCTAGCGGAGGAGAACGCC | cfnf2 |
| CNF-2as     | GTGCACATTACAGAATATTTCGAC | cfnf2 |
| CNF-3-3d    | TAAAGCTTATAGCAAAGAAGA | cfnf3 |
| CNF-3as     | GGCTTACATCTACAGTGTA | cfnf3 |
| cdt B-s     | AAATTCTCCTGCAATCATCCAGTTA | cdtB-s1, cdtB-s2 |
| cdt B-r     | AAATTCTCCTGCAATCATCCAGTTA | cdtB-s1, cdtB-s2 |
| cdt C-s     | TTTCCTTCTGAGTGTGATACT | cdtB-s1 |
| cdt C-r     | TTGCAGATTCACGGATCGTCA | cdtB-s1 |
| cdt D-s     | AAATTCTCCTGCAATCATCCAGTTA | cdtB-s1 |
| cdt D-r     | AAATTCTCCTGCAATCATCCAGTTA | cdtB-s1 |
| cdt E-s     | AAATTCTCCTGCAATCATCCAGTTA | cdtB-s1 |
| cdt E-r     | AAATTCTCCTGCAATCATCCAGTTA | cdtB-s1 |
| cif-f       | AAAGATGAGCAAGACAGCAG | cif [51] |
| cif-r       | AAAGATGAGCAAGACAGCAG | cif |
| cibQ-F      | GCCTAGCTTTTACCTTTCTC | This study |
| cibQ-R      | GCCTAGCTTTTACCTTTCTC | This study |

1The afa-f and afa-r primers detect all afa-positive strains, irrespective of the afa-f subtype and the binding properties of the adhesins (Afa/Dr+ and Afa/Dr-). Specific detection of strains encoding Afa/Dr+ adhesins was performed using afa-1 and afa-2 primers; 2Primers used with CDT-s1 primer.

**Adhesion and invasion assays**

1–407 cells were infected at a multiplicity of infection (MOI) of 10 bacteria per cell. Adhesion and invasion assays were performed as previously described[34]. For adhesion assays, monolayers were washed five times in PBS after 3 h of incubation at 37 °C. To determine the numbers of intracellular bacteria (invasion assay), cell culture medium containing gentamicin at a concentration of 200 μg/mL was added for 1 h to kill extracellular bacteria. The epithelial cells were then lysed with 1% Triton X-100 in deionized water. This concentration of Triton X-100 had no effect on bacterial viability for at least 30 min. Samples were diluted and plated onto LB agar plates to determine the number of CFU.

**Biofilm formation assays**

Biofilm formation assays on abiotic surface were performed using a previously described method[33]. Biofilm measurements were calculated using the formula SBF = (AB-CW)/G, in which SBF is the specific biofilm formation, AB the OD570nm of the attached and stained bacteria, CW the OD570nm of the control wells containing only bacteria-free medium (to eliminate unspecific or abiotic OD values), and G is the OD570nm of cell growth in broth. Assays were performed in triplicate.

Biofilm formation assays were also performed using PFA-fixed 1–407 cells[33]. Briefly, confluent 1–407 monolayers were fixed for 15 min in 3.7% PFA-PBS. The fixed cells were washed and infected with bacteria in M63 minimal medium and incubated overnight at 30 °C without shaking. For visualization, infected epithelial cells were fixed for 15 min in 3.7% PFA-PBS and permeabilized in PBS-0.1% Triton X-100. Coverslides were incubated with goat anti-E. coli polyclonal antibodies (dilution 1/100, AbD serotec) and Alexa 488-labeled anti-goat antibodies (dilution 1/300, Invitrogen). Actin cytoskeleton was stained using TRITC-labelled-phalloidin (Sigma). The slides were examined with a Zeiss LSM 510 Meta confocal microscope (IHCF platform, Clermont-Ferrand, France).

**Mouse model infection**

CEBAC10 transgenic mice (heterozygote[33]) were housed in specific pathogen-free conditions in the animal care facility at Université d’Auvergne, Clermont-Ferrand, France). Mice from the same generation were used for experimentation. Animal protocols were approved by the Committee for Research and Ethical Issues of the International Association for the Study of Pain. A total of 22 female 10 wk-old CEBAC10 mice were divided into three groups: non-infected control group (n = 6), 11G5-infected group (n = 7) and AIEC LF82-infected group (n = 9). The animals were pretreated once before the first infection cycle by oral administration of the broad-spectrum antibiotic streptomycin (20 mg intragastric per mouse) to disrupt normal resident bacterial flora in the intestinal tract and received a dose of 0.25%...
of dextran sulfate sodium (DSS; molecular mass = 36000-50000 daltons; MP Biomedicals) in drinking water 3 d before infection to increase the accessibility of bacteria to the surface of the epithelial layer. The administration of 0.25% DSS did not affect the body weight of mice and did not induce clinical symptoms of colitis [36].

The mice were subjected to 8 consecutive cycles of infection. For each infection cycle, they were orally challenged twice a week by intra-gastric gavage with $2 \times 10^8$ bacteria for a 3-wk period. This infection period was followed by a 1-wk recovery period without infection. For each cycle, 5 d after the last oral bacterial infection, fresh fecal pellets (100-200 mg) were collected and suspended in PBS to evaluate colonization. After serial dilution, bacteria were enumerated by plating on LB agar medium containing 50 µg/mL of kanamycin and 50 µg/mL of ampicillin isolate 11G5 bacteria or 100 µg/mL of ampicillin and 20 µg/mL of erythromycin to isolate LF82 bacteria, and incubated at 37 °C overnight.

**Histological grading of intestinal inflammation and epithelial damages**

After mouse sacrifice, the entire colon was excised and rolls of the proximal colon were fixed in buffered 4% formalin, paraffin-embedded, cut into 5-µm slices, and stained with hematoxylin/eosin/safranin. The histological severity of colitis was graded in a blinded fashion by a GI pathologist. The tissue samples were assessed for the extent and depth of inflammation and the extent of epithelial damages, as presented in Table 2. The histology score corresponds to the sum of each item.

**Immunohistochemistry**

For immunohistochemical staining of mouse Ki-67, heat-induced epitope retrieval was performed using sodium citrate buffer (pH 6.0). Ki-67 antigen was detected using anti-mouse Ki-67 polyclonal antibodies (Leica) and revealed with Vectastain ABC kit (Vector) and DAB detection kit (Invitrogen). The sections were counterstained using Gill’s hematoxylin (Vector).

**Real-time mRNA quantification**

Total RNAs were extracted from tissue using a Nucleospin® RNA/Protein extraction kit (Macherey-Nagel GmbH & Co). RNA samples were subjected to reverse transcription using High-Capacity cDNA Reverse Transcription Kit and non-specific random hexamer primers (Applied Biosystems) and quantification was performed using FastStart SYBR® Green Master kit (Roche Applied Science). The primer sequences used are given in Table 3. Gene expression values were calculated based on the $\Delta \Delta Ct$ method.

**Enzyme-linked immunosorbent assay**

T84 colon epithelial cells were infected with bacteria for 6 h at a MOI of 100 bacteria per cell. The amount of CEACAM6 on whole cell protein extracts was determined by enzyme-linked immunosorbent assay (ELISA) according to manufacturer’s instructions (R and D systems).

**Western immunoblotting**

T84 colon epithelial cells were infected for 6 h at a MOI of 100 bacteria per cell. Whole-cell protein extracts were prepared by adding NP-40 lysis buffer. Protein concent-

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**Table 2 Histological grading of intestinal inflammation**

| Symptoms (wt/vol) of dextran sulfate sodium (DSS; molecular mass = 36000-50000 daltons; MP Biomedicals) in drinking water 3 d before infection to increase the accessibility of bacteria to the surface of the epithelial layer. The administration of 0.25% DSS did not affect the body weight of mice and did not induce clinical symptoms of colitis [36]. The mice were subjected to 8 consecutive cycles of infection. For each infection cycle, they were orally challenged twice a week by intra-gastric gavage with $2 \times 10^8$ bacteria for a 3-wk period. This infection period was followed by a 1-wk recovery period without infection. For each cycle, 5 d after the last oral bacterial infection, fresh fecal pellets (100-200 mg) were collected and suspended in PBS to evaluate colonization. After serial dilution, bacteria were enumerated by plating on LB agar medium containing 50 µg/mL of kanamycin and 50 µg/mL of ampicillin isolate 11G5 bacteria or 100 µg/mL of ampicillin and 20 µg/mL of erythromycin to isolate LF82 bacteria, and incubated at 37 °C overnight.

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**Western immunoblotting**

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**Table 3 List of primers used for RT qPCR assays**

| Primer name | Sequence (5’-3’) | Region specific for | Ref. |
|-------------|-----------------|---------------------|------|
| mmu-26s-FW  | TGTACATCGGAAACATTGTAG | S26 | This study |
| mmu-26s-RV | GCCTTTGTGAGGTC | mmu-PCNA-FW | CCACATTGGAGATGCTGTG | PCNA |
| mmu-PCNA-RV | CATGTGAGTGGCATTGG | Raisch J et al. B2 E. coli and colon cancer |
trations were determined by Bradford assay. Total proteins were subjected to SDS-PAGE on 12% gels.

Proteins were electroblotted onto nitrocellulose membranes (Amersham International), and the membranes were immunoblotted for CEACAM6 (mouse anti-CEACAM6, dilution 1/1,000, Genovac) and GAPDH (rabbit anti-GAPDH; dilution 1/1,000, Cell Signaling). Immunoreactants were detected using horseradish peroxidase-conjugated anti-rabbit or antimouse immunoglobulin G antibodies, ECL reagents (Amersham Biosciences) and autoradiography.

**Statistical analysis**

Where appropriate, nonparametric data were expressed by median value (range). Normally distributed data were expressed as means. Error bars represent SEM. Statistical analysis was done using ANOVA (Histopathological score and RT-qPCR), Mann Whitney (adhesion, invasion and biofilm assays) using GraphPad prism5 software. A P value of 0.05 was considered significant.

**RESULTS**

**Most cyclomodulin-producing *E. coli* strains associated with colon cancer and diverticulosis belong to B2 phylogroup**

The analysis of the presence of cyclomodulin-encoding genes, *pks* island coding for colibactin, and/or *cnf* and/or *cdt* and/or *cif* genes coding for cytotoxic necrotizing factor (CNF), cytotoxic distending toxin (CDT), and cycle-inhibiting factor (Cif) indicated that, whatever the origin of the *E. coli* strains, either from colon cancer or diverticulosis samples, 86% of cyclomodulin-positive *E. coli* belonged to B2 phylogroup (Tables 4 and 5). Among *E. coli* strains isolated from colon cancer specimens, B2 *E. coli* strains harboring *pks, cnf* and *cdt* genes represented 26%, 18% and 11% of the total strains isolated, respectively (Tables 4 and 6). Among *E. coli* strains isolated from patients with diverticulosis, *pks*-positive B2 *E. coli* strains represented 13%, and *cnf*-positive B2 *E. coli* strains 9% of the total strains isolated. Although a higher prevalence of B2 *E. coli* strains harboring *pks* or *cnf* genes was observed in colon cancer patients than in patients with diverticulosis, this was not significant (P = 0.06 for both *pks* and *cnf* genes). Of interest, all but two *cnf*-positive strains also harbored *pks* and all *cnf*- and *pks*-positive *E. coli* strains belonged to the B2 phylogroup.

**Low level of adhesion and invasion but high ability to form biofilm of B2 *E. coli* strains isolated from colon cancer or diverticulosis patients**

The analysis of the ability of *E. coli* strains to adhere to and to invade intestinal epithelial cells was restricted to B2 *E. coli*, which were the main cyclomodulin producers in our study. Of note, due to cytolytic activity of hemolysin on cultured cells, hemolysin-positive *E. coli* strains were not tested. Results showed that B2 phylogroup *E. coli* strains isolated from colon cancer and from diverticulosis displayed low levels of adhesion to I -407 intestinal epithelial cells (Figure 1A). Compared to the adhesion level of the AIEC reference strain LF82, for which a mean adhesion index of 53.23 ± 6.63 was observed, the adhesion levels of all *E. coli* strains isolated from colon cancer (except *E. coli* strain 14H4, which had a mean adhesion level of 25.76 ± 5.06) or from diverticulosis ranged from 0.15 ± 0.02 to 4.04 ± 1.24 or from 0.10 ± 0.04 to 9.17 ± 3.40, respectively. Microscopy examination after Giemsa staining showed a diffuse adhesion pattern (data not shown), and we therefore searched for adhesive factor-encoding genes associated with diffusely adhering

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**Table 4** Distribution of *Escherichia coli* strains producing various cyclomodulins according to phylogroups and specimen origins n (%)  

| Phylogroups | E. coli strains exhibiting cyclomodulin-encoding genes |
|-------------|-----------------------------------------------------|
|             | pks cnf cdt cif                                      |
| Colon cancer-associated *E. coli* strains (n = 88) | A (n = 20) 0 (0) 0 (0) 0 (0) 2 (2) |
| B1 (n = 14) | 0 (0) 1 (1) 1 (1) 1 (1)                              |
| B2 (n = 38) | 23 (26) 16 (18) 4 (11) 0 (0)                         |
| D (n = 16)  | 0 (0) 1 (1) 1 (1) 0 (0)                              |
| Diverticulosis-associated *E. coli* strains (n = 46) | A (n = 17) 0 (0) 0 (0) 0 (0) 0 (0) |
| B1 (n = 3)  | 0 (0) 0 (0) 0 (0) 0 (0)                              |
| B2 (n = 15) | 6 (13) 4 (9) 0 (0) 0 (0)                             |
| D (n = 11)  | 0 (0) 0 (0) 0 (0) 0 (0)                              |

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1Isolated from 48 colon cancer patients; 2Isolated from 35 patients with diverticulosis. *E. coli: Escherichia coli*.
E. coli (DAEC) strains (i.e., Afa and Afa/Dr adhesin-encoding genes). None of the B2 E. coli strains tested was positive for afa or afa/dr genes except the highly adherent E. coli strain 14H4 isolated from colon cancer (Table 6). Of note, none of the B2 E. coli strains tested was positive for eae gene coding for intimin of enteropathogenic E. coli or for aaf gene coding for the adhesive factor AAF of enteroaggregative E. coli, indicating that B2 E. coli strains studied do not belong to these E. coli pathovars. Analysis of the ability of bacteria to invade I-407 cells showed

### Table 6 Hemolysin expression and presence of cyclomodulin- and adhesin-encoding genes in B2 phylogroup Escherichia coli strains

| E. coli strains | Haemolytic phenotype | Cyclomodulin-encoding genes | Adhesin-encoding genes |
|-----------------|----------------------|-----------------------------|------------------------|
|                 | pks                  | cnf                         | cdtB                   |
| Colon cancer    |                      |                             |                        |
| 1C12            | +                    | cnf1                        | -                      |
| 1D2             | -                    | -                           | -                      |
| 1F8             | -                    | -                           | -                      |
| 2D5             | -                    | -                           | -                      |
| 2F8             | +                    | cnf1                        | -                      |
| 2G2             | +                    | -                           | -                      |
| 4A9             | -                    | -                           | -                      |
| 6A8             | +                    | cnf1                        | -                      |
| 6G8             | +                    | cnf1                        | -                      |
| 6G10            | -                    | -                           | -                      |
| 6G11            | -                    | -                           | -                      |
| 7G1             | -                    | -                           | -                      |
| 7G2             | +                    | cnf1                        | -                      |
| 8A9             | +                    | -                           | -                      |
| 8A10            | -                    | -                           | -                      |
| 8F1             | +                    | cnf1                        | -                      |
| 8G8             | -                    | -                           | -                      |
| 9C5             | +                    | cnf1                        | -                      |
| 10D12           | -                    | -                           | -                      |
| 10E9            | +                    | cnf1                        | -                      |
| 11F1            | -                    | -                           | -                      |
| 11G5            | -                    | +                           | -                      |
| 12B1            | +                    | cnf1                        | -                      |
| 13H2            | +                    | cnf1                        | -                      |
| 14H4            | +                    | -                           | -                      |
| 15D1            | +                    | -                           | -                      |
| 15D3            | +                    | cnf1                        | -                      |
| 16C1            | -                    | +                           | -                      |
| 17G3            | +                    | -                           | -                      |
| 18C3            | +                    | cnf1                        | -                      |
| 18C5            | -                    | -                           | -                      |
| 18H5            | -                    | +                           | -                      |
| 19D12           | -                    | +                           | -                      |
| 19G1            | -                    | +                           | -                      |
| 19H2            | +                    | cnf1                        | -                      |
| 20B6            | +                    | cnf1                        | -                      |
| 20C3            | -                    | +                           | -                      |
| 20D5            | -                    | -                           | -                      |
| Diverticulosis  |                      |                             |                        |
| 1D5             | -                    | -                           | -                      |
| 4D5             | +                    | cnf1                        | -                      |
| 9D7             | -                    | -                           | -                      |
| 9F1             | -                    | -                           | -                      |
| 9F4             | -                    | -                           | -                      |
| 11D9            | +                    | cnf1                        | -                      |
| 12H1            | -                    | +                           | -                      |
| 13D1            | +                    | cnf1                        | -                      |
| 15C1            | -                    | +                           | -                      |
| 16A4            | -                    | +                           | -                      |
| 16A8            | -                    | -                           | -                      |
| 17C1            | -                    | -                           | -                      |
| 17F2            | -                    | -                           | -                      |
| 17E1            | -                    | -                           | -                      |
| 18E6            | +                    | -                           | -                      |

*Alpha-hemolysin expression was analysed after over-night growth of bacteria on agar plates containing sheep blood. cnf: Cytotoxic necrotizing factor; cdt: Cytolethal distending toxins; cif: Cycle-inhibiting factor; afa: Afimbrial adhesion. E. coli: Escherichia coli.*
that whatever the origin of the B2 *E. coli* strains their invasion levels were very low, ranging from 0.02% to 1.49%, except strain 14H4, for which invasion level was similar to that of the AIEC strain LF82 (Figure 1B).

We also investigated the ability of B2 *E. coli* isolated from colon cancer to induce CEACAM6 expression as abnormal CEACAM6 expression was shown to promote gut colonization by AIEC \(^3^8\) and AIEC bacteria were reported to be able to induce increased CEACAM6 expression in intestinal epithelial cells \(^3^9\). A quantitative analysis of the level of CEACAM6 expression by T84 intestinal epithelial cells in response to B2 *E. coli* infection was determined by ELISA (Figure 1C) and Western blot (Figure 1D). Interestingly, we observed that most of B2 *E. coli* strains isolated from colon cancer induced increased expression of CEACAM6 to a level similar to that of AIEC strain LF82. Of note, B2 *E. coli* strains isolated from diverticulosis induced no or very low expression of CEACAM6 in T84 cells.

Another important bacterial trait involved in the colonization of the intestinal mucosa by gut resident bacteria is their ability to form biofilm. This property was investigated both on abiotic and on fixed intestinal epithelial cells. The level of biofilm formation on abiotic surface was evaluated by calculating the specific biofilm formation index (SBF). An SBF index of 3.13 ± 0.23 was obtained for AIEC strain LF82 compared to 0.99 ± 0.22 for the non-pathogenic K-12 *E. coli* strain C600 (Figure 2A).

We observed that 7/19 (37%) B2 *E. coli* strains isolated from colon cancer and 2/8 (25%) B2 *E. coli* strains isolated from diverticulosis harbored SBF index similar to that of the biofilm producer AIEC strain LF82 (P ≤ 0.05). Biofilm formation on fixed I-407 intestinal epithelial cells was evaluated by confocal microscopy (Figure 2B), which confirmed that 6/9 B2 *E. coli* strains having a high SBF index on abiotic surface were able to form a strong biofilm on fixed I-407 cultured cells. Combining the two methods of biofilm formation assessment, 16/27 B2 *E. coli* strains tested were able to form biofilm. This shows that even B2 *E. coli* strains have a low ability to adhere to intestinal epithelial cells, at least half of them were able to form biofilm to a level similar to that of CD-associated *E. coli* strain LF82 known to form a strong biofilm and no difference was observed between B2 *E. coli* strains isolated from colon cancer patients or from patients with diverticulosis.

**Colonization of colon mucosa in CEACAM-expressing mice by colon cancer-associated B2 *E. coli* strain 11G5:** induction of inflammation and enhanced epithelial intestinal cell proliferation

CEABAC10 mice harboring a bacterial artificial chromosome that contains part of the human CEA family gene cluster including the *CEACAM6* gene were infected with AIEC reference strain LF82 or B2 *E. coli* strain 11G5 isolated from colon cancer patient. To assess bacterial colonization, the levels of bacteria in the stools were determined 5 d after the last infection of each cycle, over the...
E. coli - Colon cancer - 12H1 strain were compared to that of the non-pathogenic K-12 harbors genes. Escherichia coli strains belonged to P strains belonging to B2 phylogroup have a with AIEC strain LF82 and inflammation and epithelial damages for mice infected analysis showed a similar colonic histological score for colon cancer-associated 11G5 the colon of mice infected with AIEC strain LF82 or development such as neoplasia or polyp was observed in 11G5 bacteria (Figure 3A). Infected with AIEC LF82 and colon cancer-associated revealed similar ( between infection with B2 E. coli strain 11G5 associated with colon cancer.

DISCUSSION

Accumulating evidence supports the involvement of infectious agents in the development of cancer, especially in organs that are continuously exposed to microorganisms such as the colon. Remodeling of the colonic microbiota due to environmental changes is thought to contribute to the pathogenesis of colon cancer by suppressing the growth of cancer-protective bacterial species and allowing the emergence or expansion of bacterial species with oncogenic potential. It has been suggested that the role of E. coli in CRC promotion and development is linked to chronic inflammation, which can result from bacterial infection via its effects on both the host and the microbiota, in particular that of promoting the expansion of certain bacteria, such as pro-inflammatory E. coli[37] or ETBF[38,39]. In parallel, two different studies have reported that between 71% and 82% of patients with colonic adenoma or carcinoma[10,12] are highly colonized by mucosa-associated E. coli compared to controls. The aim of the present study was to provide further insight into the characterization of the E. coli colonizing the mucosa of colon cancer patients.

It is well documented that B2 E. coli harbors genes coding for cyclomodulins such as colibactin, which is encoded by the pks genomic island, CDT, CNF or Cif, which can act as genotoxic agents and/or can modulate cellular differentiation, apoptosis, and proliferation[13,40,41]. In the present study, we observed that 86% of cyclomodulin-positive E. coli isolated from colon cancer and diverticulosis specimens belonged to B2 phylogroup. Of interest, all but two coly-positive strains also harbored pks and all coly- and pks-positive E. coli strains belonged to the B2 phylogroup. Our results are in good agreement with those reported by Arthur et al[9], who observed that 66.7% of patients with CRC and 20.8% of controls harbored pks-positive E. coli.

E. coli strains belonging to B2 phylogroup have a

Figure 2  Ability of B2 Escherichia coli strains to form biofilm. Biofilm formation of colon cancer-associated and diverticulosis-associated B2 Escherichia coli (E. coli) were compared to that of the non-pathogenic K-12 E. coli strain C600 and the biofilm producer AIEC strain LF82. A: Biofilm formation on abiotic surface. Results are expressed as specific biofilm formation (SBF) index; B: Biofilm formation on human I-407 intestinal epithelial cells. E. coli strain 11G5 was isolated from a patient with colon cancer and E. coli strain 12H1 from a patient with diverticulosis. Bacteria were stained using goat anti-E. coli polyclonal antibodies (green) and I-407 cells were labeled for actin cytoskeleton using TRITC-labeled phalloidin (red). Y- and Z-stack projections are presented.

8 consecutive cycles of infection. Analysis of the number of bacteria recovered in the stools at cycle 4 and cycle 8 revealed similar (P = 0.78) colonization levels for mice infected with AIEC LF82 and colon cancer-associated 11G5 bacteria (Figure 3A).

On macroscopic examination, no sign of tumor development such as neoplasia or polyp was observed in the colon of mice infected with AIEC strain LF82 or colon cancer-associated 11G5 E. coli strain. Histological analysis showed a similar colonic histological score for inflammation and epithelial damages for mice infected with AIEC strain LF82 and E. coli strain 11G5 (P ≥ 0.05)
greater ability to colonize the human gut, due, at least in part, to accumulation of genes encoding fitness factors such as pili and adhesins[42,43]. In addition, an increased proportion of mucosa-associated \textit{E. coli} expressing hemagglutinins was observed in CD patients (39%) and colon cancer patients (38%) compared to controls (4%), in correlation with the ability of bacteria to adhere to I-407 and HT-29 intestinal epithelial cells[10]. However in our study, analysis of the adhesive abilities of B2 \textit{E. coli} isolated from colon cancer or diverticulosis revealed that the strains were poorly adherent to I-407, even if the majority of them were able to form biofilm. However some B2 \textit{E. coli} strains isolated from colon cancer induced increased expression of CEACAM6 to a level similar to that of AIEC strain LF82 associated with CD, indicating that colon cancer-associated \textit{E. coli} could influence carcinogenesis, since CEACAM6 has been implicated in cellular adhesiveness, invasiveness, and metastatic behavior of tumor cells[30,44]. In addition, this result indicates that, in agreement with what we previously reported for AIEC strains isolated from CD patients[28], colon cancer-associated \textit{E. coli} strains could have the ability to promote their own colonisation since CEACAM6 serves as a receptor for mediating adherence and/or cell entry of pathogenic bacteria such as \textit{Neisseria} bacteria[45], diffusely-adhering \textit{E. coli} (DAEC)[46] or AIEC[28].

Figure 3 Impact of CEABAC10 mice colonization by B2 \textit{Escherichia coli} strain on inflammation and cell proliferation. CEABAC10 mice transgenic for human CEACAMs, including CEACAM6, were subjected to 8 consecutive cycles of infection with AIEC LF82 or B2 phylogroup \textit{Escherichia coli} (E. coli) strain 11G5. Control mice received PBS. A: Quantification of the number of bacteria in the faeces of mice at cycle 4 and cycle 8; B: Histopathological scoring for several parameters of inflammation and epithelial damages (see Table 2) was performed at the end of the 8th cycle. \(P < 0.05\) vs control; \(P = \text{NS}\) vs LF82; C: Hematoxylin/eosin/safran (HES) staining of colonic tissue sections; D: Total RNAs from colons were extracted at the end of the 8th cycle. PCNA and S26 mRNA levels were measured by RT-qPCR. PCNA amount relative to S26 is presented. \(P < 0.05\) vs control; \(P = \text{NS}\) vs LF82; E: Immunohistochemistry examination of Ki67 on colonic tissue sections. NS: Not significant.

Experiments of long-term colonization of CEABAC10 mice revealed that an \textit{E. coli} strain isolated from colon cancer (strain 11G5) was able to persist in the gut of CEABAC10 transgenic mice expressing human CEACAMs, including CEACAM6 and to exacerbate colonic inflammation. Whether colonisation of the intestinal mucosa of colon cancer patients by B2 \textit{E. coli} is a cause or a consequence of malignant transformation is a ques-
tion that has yet to be addressed. We show here that B2 phylogroup *E. coli* isolated from colon cancer increased the proliferative index of epithelial cells in crypts in the chronic infection model of CEABAC10 mice. This indicates that colonic mucosa cells undergo accelerated proliferation in response to infection by B2 *E. coli*. The ability to induce cell proliferation is a common trait of various pathogens involved in carcinogenesis. Indeed, *Bacteroides fragilis* enterotoxin induces c-myc transcription and translation and persistent cellular proliferation ensues, mediated in part by β-catenin/T-cell factor-dependent transcriptional activation[43]. Another example is *Helicobacter pylori* (*H. pylori*), which increases the proliferation of gastric cancer cells. This process is dependent on the LPS-TLR4 pathway since *H. pylori* LPS induces the proliferation of gastric cancer cells and the use of neutralizing antibody against TLR4 almost completely abrogates the proliferative activities of cancer cells[44]. Some cyclomodulins, such as CNF, which are mostly produced by B2 *E. coli*, induce epithelial cell proliferation[45]. In our study the B2 *E. coli* strain 11G5 did not harbor the *cnf* genes and was able to promote cell proliferation as observed in infected CEABAC10 mice. This effect could be related to *E. coli*-derived LPS, which was previously reported to have a more remarkable cancer proliferative activity than *H. pylori*-derived LPS[46]. Because *E. coli* inhabits the host colon as normal intestinal flora, owing to host tolerance toward *E. coli*, it is likely that *E. coli* LPS stimulates the host cellular immune response to prevent cancer progression. However, we can hypothesize that when too great a load of *E. coli* colonize the colonic mucosa, as observed in 11G5-infected CEABAC10 mice, potent tumor proliferative activity is no longer effectively repressed. The cell proliferation observed in 11G5-infected CEABAC10 mice could also result from the presence of colibactin. Colibactin with its genotoxic activity promotes DNA damages, which leads to carcinogenesis and cell proliferation.

In conclusion, B2 *E. coli* abnormally colonized the mucosa of colon cancer patients, indicating that microbiota remodeling had occurred promoting their expansion. Together with previous findings reported by Arthur et al[47], this study on a larger cohort of patients confirms the high prevalence of B2 *pks*-positive or *pks-cnf*-positive *E. coli* in colon cancer patients. The study also indicates that, these bacteria can promote low grade inflammation and cell proliferation, as shown in the CEABAC10 infected mouse model. Analyses to determine whether these bacteria take advantage of the tumor microenvironment to colonize the gut or promote their own colonization may be an important step in understanding their role in carcinogenesis and in the development of therapeutic strategies.

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**REFERENCES**

1. **World Health Organization World.** WHO Global Malaria Programme. Switzerland: World Health Organization, 2011. Available from: URL: http://www.who.int/malaria/world_malaria_report_2011/9789241564403_eng.pdf
2. **Vogelstein B, Fearon ER, Hamilton SR, Kern SE, Preisinger AC, Leppert M, Nakamura Y, White R, Smits AM, Bos JL.** Genetic alterations during colorectal-tumor development. *N Engl J Med* 1988; 319: 525-532 [PMID: 2841597 DOI: 10.1056/NEJM198809013190901]
3. **Arthur JC, Perez-Chanona E, Mühlbauer M, Tomovick SJ, Uronis JM, Fan TJ, Campbell BJ, Abujamel T, Dogan B, Rogers AB, Rhodes JM, Stintzi A, Simpson KW, Hansen JJ, Keku TO, Fodor AA, Jobin C.** Intestinal inflammation targets antigen-related cell adhesion molecule 6 (CEACAM6) receptor in intestinal epithelial cells, and that these bacteria were able to persist in a chronic infection model of CEACAM6 expressing mice and had oncogetic potential.

**COMMENTS**

**Background**

Colorectal cancer (CRC) is one of the most prevalent cancers worldwide, and is the fourth leading cause of cancer death worldwide. Inflammation and changes in composition and function of gut microbial communities are suspected to be causative factors in the development of sporadic CRC.

**Research frontiers**

The authors and other researchers have reported abnormal colonization of tumors and mucosa of colon cancer patients by *Escherichia coli* (*E. coli*) belonging to B2 phylogroup.

**Innovations and breakthroughs**

To date, there has been a limited number of studies analyzing interaction of colon cancer-associated *E. coli* to intestinal epithelial cells. The authors showed that colon cancer-associated *E. coli* induce expression of the carcinoembryonic antigen-related cell adhesion molecule 6 (CEACAM6) receptor in intestinal epithelial cells, and that these bacteria were able to persist in a chronic infection model of CEACAM6 expressing mice and had oncogenic potential.

**Applications**

The authors have analyzed the ability of colon cancer-associated *E. coli* to colonize gut mucosa and influence carcinogenesis. Analyses to determine whether these bacteria take advantage of the tumor microenvironment to colonize the gut or promote their own colonization may be an important step in understanding their role in carcinogenesis and in the development of therapeutic strategies.

**Terminology**

CEACAM6 molecule serves as a receptor for mediating mucosa colonization by pathogenic bacteria.

**Peer review**

This study provides evidence supporting the hypothesis that colon cancer-associated *E. coli* are able to colonize gut mucosa and to induce cell proliferation in a mouse model with overexpression of human CEACAM6.

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Mucosal flora in inflammatory bowel diseases. *Inflamm Bowel Dis* 2005; 11: 481-487 [PMID: 15867888]

23 Baumgart M, Dogan B, Rishniw M, Weitzman G, Bosworth B, Yantis R, Orsi RH, Wiedmann M, McDonough P, Kim SG, Berg D, Schukken Y, Schell E, Simpson KW. Culture independent analysis of ileal mucosa reveals a selective increase in invasive Escherichia coli of novel phylogeny relative to depletion of Clostridiales in Crohn’s disease involving the ileum. *ISME J* 2007; 1: 403-418 [PMID: 18043660 DOI: 10.1038/ismej.2007.52]

24 Kotlowski R, Bernstein CN, Sepehr S, Krause DO. High prevalence of Escherichia coli belonging to the B2-D phylogenetic group in inflammatory bowel disease. *Gut* 2007; 56: 669-675 [PMID: 17028128 DOI: 10.1136/gut.2006.099796]

25 Darfeuille-Michaud A, Neut C, Barnich N, Lederman E, Di Martino P, Desreumaux P, Gambiez L, Joly B, Cortot A, Colombel JF. Presence of adherent Escherichia coli strains in ileal mucosa of patients with Crohn’s disease. *Gastroenterology* 1998; 115: 1405-1413 [PMID: 9834288]

26 Boureau J, Glasser AL, Masseret E, Joly B, Darfeuille-Michaud A. Invasive ability of an Escherichia coli strain isolated from the ileal mucosa of a patient with Crohn’s disease. *Infect Immun* 1999; 67: 4409-4509 [PMID: 10456892]

27 Darfeuille-Michaud A, Boureau J, Bulois P, Neut C, Glasser AL, Barnich N, Bringer MA, Swidsinski A, Beaugerie L, Colombel JF. High prevalence of adherent-invasive Escherichia coli associated with ileal mucosa in Crohn’s disease. *Gastroenterology* 2004; 127: 412-421 [PMID: 15300573]

28 Barnich N, Carvalho FA, Glasser AL, Darcha C, Jantscheff P, Allez M, Peeters H, Bommelra G, Desreumaux P, Colombel JF, Darfeuille-Michaud A. CEACAM6 acts as a receptor for adherent-invasive E. coli, supporting ileal mucosa colonization in Crohn disease. *J Clin Invest* 2007; 117: 1566-1574 [PMID: 17528500 DOI: 10.1172/JCI30504]

29 Schöbel S, Zimmermann W, Schwarzkopf G, Grunert F, Rogaczewski B, Thompson J. Carcinoembryonic antigen family members CEACAM6 and CEACAM7 are differentially expressed in normal tissues and oppositely deregulated in hyperplastic colorectal polyps and early adenomas. *Am J Pathol* 2000; 156: 599-605 [PMID: 10666389 DOI: 10.1016/S0002-9440(10)64764-5]

30 Kim KS, Kim JT, Lee SJ, Kang MA, Choe IS, Kang YH, Kim SY, Yeom YI, Lee YH, Kim JH, Kim KH, Kim CN, Kim JW, Nam MS, Lee HG. Overexpression and clinical significance of carcinoembryonic antigen-related cell adhesion molecule 6 in colorectal cancer. *Clin Chim Acta* 2013; 415: 12-19 [PMID: 22975528 DOI: 10.1016/j.cca.2012.09.003]

31 Versalovic J, Koeuth T, Lupski JR. Distribution of repetitive DNA sequences in eubacteria and application to fingerprinting of bacterial genomes. *Nucleic Acids Res* 1991; 19: 6823-6831 [PMID: 1762913]

32 Wang G, Whitman TS, Berg CM, Berg DE. RAPD (arbitrary primer) PCR is more sensitive than multilocus enzyme electrophoresis for distinguishing related bacterial strains. *Nucleic Acids Res* 1993; 21: 5930-5933 [PMID: 8293054]

33 Bidet P, Metais A, Mahjoub-Messai F, Durand L, Dehem M, Aujard Y, Bingen E, Nassiri X, Bonacorsi S. Detection and identification by PCR of a highly virulent phylogenetic subgroup among extraintestinal pathogenic Escherichia coli B2 strains. *Appl Environ Microbiol* 2007; 73: 2373-2377 [PMID: 17295307 DOI: 10.1128/AEM.02341-06]

34 Chassaing B, Darfeuille-Michaud A. The O2 pathway is involved in biofilm formation by Crohn’s disease-associated adherent-invasive Escherichia coli. *J Bacteriol* 2013; 195: 76-84 [PMID: 23104802 DOI: 10.1128/JB.01079-12]

35 Chan CH, Stanners CP. Novel mouse model for carcinoembryonic antigen-based therapy. *Mol Ther* 2004; 9: 775-785 [PMID: 15194045 DOI: 10.1038/jmt.2004.3.009]

36 Carvalho FA, Barnich N, Sivignon A, Darcha C, Chan CH, Stanners CP, Darfeuille-Michaud A. Crohn’s disease ad-
herent-invasive Escherichia coli colonize and induce strong gut inflammation in transgenic mice expressing human CEACAM. J Exp Med 2009; 206: 2179-2189 [PMID: 19737864 DOI: 10.1084/jem.20090741]

Mukhopadhyay I, Hansen R, El-Omar EM, Hold GL. IBD-2010; 2003; 1997; 2001; Hogan AM, Winter DC. Microbial and viral enterotoxigenic Bacteroides fragilis: a rogue family member CEACAM6 disrupts tissue architecture and persistent expression of the human tumor marker CEA and CEA Ilantzis C [PMID: 16483819 DOI: 10.1016/j.micinf.2005.10.011]

Raisch J et al. B2 E. coli and colon cancer

What role do Proteobacteria play? Nat Rev Gastroenterol Hepatol 2012; 9: 219-230 [PMID: 22349170 DOI: 10.1038/n gastro.2012.14]

Sears CL. Enterotoxigenic Bacteroides fragilis: a rogue among symbiotes. Clin Microbiol Rev 2009; 22: 349-369, Table of Contents [PMID: 19366918 DOI: 10.1128/CMR.00053-08]

Toprak NU, Yagci A, Gulluoglu BM, Akin ML, Demirkalem P, Celenk T, Soyiletir G. A possible role of Bacteroides fragilis enterotoxin in the aetiology of colorectal cancer. Clin Microbiol Infect 2006; 12: 782-786 [PMID: 16842574 DOI: 10.1111/j.1469-0691.2006.01494.x]

Nougayrède JP, Taieb F, De Rycke J, Oswald E. Cyclomodulins: bacterial effectors that modulate the eukaryotic cell cycle. Trends Microbiol 2005; 13: 103-110 [PMID: 15737728 DOI: 10.1016/j.tim.2005.01.002]

Collins D, Hogan AM, Winter DC. Microbial and viral pathogens in colorectal cancer. Lancet Oncol 2011; 12: 504-512 [PMID: 21067973 DOI: 10.1016/S1470-2045(10)70186-8]

Picard B, Garcia JS, Gouriou S, Duriez P, Brahimi N, Bingen E, Elion J, Denamur E. The link between phylogeny and virulence in Escherichia coli extraintestinal infection. J Clin Microbiol 1999; 37: 546-553 [PMID: 9916687]

Nowrouzian FL, Adlerberth I, Wold AE. Enhanced persistence in the colon microbiota of Escherichia coli strains belonging to phylogenetic group B2: role of virulence factors and adherence to colonic cells. Microbes Infect 2006; 8: 834-840 [PMID: 16483819 DOI: 10.1016/j.micinf.2005.10.011]

Ilantzis C, DeMartel L, Screaton RA, Stanners CP. Deregulated expression of the human tumor marker CEA and CEA family member CEACAM6 disrupts tissue architecture and blocks colonocyte differentiation. Neoplasia 2002; 4: 151-163 [PMID: 11896570 DOI: 10.1088/aj/neo/7900201]

Bos MF, Grunert F, Belland RJ. Differential recognition of members of the carcinoembryonic antigen family by Opa variants of Neisseria gonorrhoeae. Infect Immun 1997; 65: 2353-2361 [PMID: 9169774]

Guignot J, Hudault S, Kansau J, Chau I, Servin AL. Human decay-accelerating factor and CEACAM receptor-mediated internalization and intracellular lifestyle of Afa/Dr diffusely adhering Escherichia coli in epithelial cells. Infect Immun 2009; 77: 517-531 [PMID: 19015254 DOI: 10.1128/IAI.00695-08]

Wu S, Morin PJ, Maouyo D, Sears CL. Bacteroides fragilis enterotoxin induces C-Myc expression and cellular proliferation. Gastroenterology 2003; 124: 392-400 [PMID: 12557145 DOI: 10.1053/gast.2003.50047]

Chochi K, Ichikura T, Kinoshita M, Majima T, Shinomiya N, Tsujimoto H, Kawabata T, Sugasawa H, Ono S, Seki S, Mohizuki H. Helicobacter pylori augments growth of gastric cancers via the lipopolysaccharide-toll-like receptor 4 pathway whereas its lipopolysaccharide attenuates antitumor activities of human mononuclear cells. Clin Cancer Res 2008; 14: 2909-2917 [PMID: 18483357 DOI: 10.1158/1078-0432.CCR-07-4467]

Le Bouguéene C, Lalouli L, du Merle L, Jouve M, Courcoux P, Bouzari S, Selvarangan R, Nowicki BJ, Germani Y, Andremont A, Gounon P, Garcia MI. Characterization of AfaE adhesins produced by extraintestinal and intestinal human Escherichia coli isolates: PCR assays for detection of Afa adhesins that do or do not recognize Dr blood group antigens. J Clin Microbiol 2001; 39: 1738-1745 [PMID: 11325983 DOI: 10.1128/JCM.39.5.1738-1745.2001]

Bernier C, Gounon P, Le Bouguéné C. Identification of an aggregative adhesion fimbria (AAF) type III-encoding operon in enteropathogenic Escherichia coli as a sensitive probe for detecting the AAF-encoding operon family. Infect Immun 2002; 70: 4302-4311 [PMID: 12117939]

Dubois D, Delmas J, Cady A, Robin F, Sivignon A, Oswald E, Bonnet R. Cyclomodulins in urosepsis strains of Escherichia coli. J Clin Microbiol 2010; 48: 2122-2129 [PMID: 20375237 DOI: 10.1128/JCM.02365-09]
