Loss of function of a rice TPR-domain RNA-binding protein confers broad-spectrum disease resistance

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Crops carrying broad-spectrum resistance loci provide an effective strategy for controlling infectious disease because these loci typically confer resistance to diverse races of a pathogen or even multiple species of pathogens. Despite their importance, only a few crop broad-spectrum resistance loci have been reported. Here, we report the identification and characterization of the rice bsr-k1 (broad-spectrum resistance Kitaake-1) mutant, which confers broad-spectrum resistance against Magnaporthe oryzae and Xanthomonas oryzae pv oryzae with no major penalty on key agronomic traits. Map-based cloning reveals that Bsr-k1 encodes a tetratricopeptide repeat (TPR)-containing protein, which binds to mRNAs of multiple OsPAL (OsPAL1–7) genes and promotes their turnover. Loss of function of the Bsr-k1 gene leads to accumulation of OsPAL1–7 mRNAs in the bsr-k1 mutant. Furthermore, overexpression of OsPAL1 in wild-type rice TP309 confers resistance to M. oryzae, supporting the role of OsPAL1. Our discovery of the bsr-k1 allele constitutes a significant conceptual advancement and provides a valuable tool for breeding broad-spectrum resistant rice.

Significance

Crops carrying broad-spectrum resistance loci provide an effective strategy for controlling infectious disease. Despite their importance, few broad-spectrum resistance loci have been reported, and the underlying mechanisms controlling the trait remain largely unknown. This report describes the identification of a gene, called “bsr-k1,” conferring broad-spectrum resistance and demonstrates that the encoded protein regulates immunity-related genes. Loss of function of BSR-K1 in rice leads to enhanced broad-spectrum resistance to two serious rice diseases with no major penalty on yield. This report provides insights into broad-spectrum resistance and offers an efficient strategy to breeding durably resistant rice.

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resistance to diverse isolates of *M. oryzae* and *Xoo*. The BSR-K1 protein binds to mRNAs of the defense-related *OsPAL* genes, promoting their turnover. Loss of function of BSR-K1 decreases mRNA turnover, resulting in *OsPAL* mRNA accumulation in the *bsr-k1* mutant. Collectively, our study reveals the mechanism by which BSR-K1 regulates the immune response and provides a tool for breeding rice with broad-spectrum resistance.

**Results**

The Rice *bsr-k1* Mutant Confers Enhanced Resistance Against Diverse Races of *M. oryzae* and *Xoo*. To identify genes involved in rice blast resistance, we generated a mutant population containing ~50,000 lines in the model rice Kitaake cultivar through ethylmethane sulfonate (EMS)-mediated mutagenesis. We then inoculated these lines with mixed spores derived from seven Kitaake-compatible *M. oryzae* isolates (ZB25, Zhong1, Tetep, 99-20-2, 97-27-2, 0755-1-1, and ZE-1) in the field. We obtained five mutant lines showing enhanced disease resistance compared with wild-type Kitaake. Of these, the *bsr-k1* mutant exhibits significantly smaller lesion length and fewer lesions than Kitaake (Fig. 1A and B). We validated the smaller lesion size and lower fungal biomass accumulation in *bsr-k1* (Fig. S1A) using a punch inoculation method (21). We then tested the seven Kitaake-compatible blast isolates individually in field inoculation. We found that the lesion numbers were dramatically reduced in *bsr-k1* with all isolates (Fig. 1C). For example, *bsr-k1* yields 1.6 ± 0.8 lesions, while Kitaake has 4.4 ± 1.2, when inoculated with isolate ZB25 (Fig. 1C). To assess if *bsr-k1* enhances resistance to other pathogens, we inoculated *bsr-k1* and Kitaake with 10 *Xoo* isolates (P2–P7, P9, P10, 8248, and *Xoo*–4) that are commonly used to test for broad-spectrum resistance to rice bacterial blight (22). We found that *bsr-k1* confers enhanced resistance to all 10 *Xoo* isolates (Fig. 1D).

Because the immune response in plants is usually accompanied by the induction of defense-related genes, such as *OsPR1a* (*Os07g0129200*), *OsPR10* (*Os12g0555200*), and *PBZ1* (*Os12g0555500*) (21), we measured the expression of these three genes in *bsr-k1* and Kitaake plants using qRT-PCR. We observed that expression levels of these three defense-related genes are moderately elevated (two- to threefold) in *bsr-k1* compared with Kitaake plants (Fig. 1E). Together, these results demonstrate that the *bsr-k1* mutant confers broad-spectrum resistance to *M. oryzae* and *Xoo* compared with Kitaake.

The *bsr-k1* Mutant Maintains Key Agronomic Traits. We next monitored several key agronomic traits in the *bsr-k1* mutant to assess its application potential in rice breeding. We examined *bsr-k1* and Kitaake plants for their yield-related traits (in Lingshui, Hainan, China), including thousand-grain weight, grain length and width, panicle length, grain numbers per panicle, grain yield per plant, and other traits, including tiller numbers and heading date. Few differences were observed between *bsr-k1* and Kitaake (Fig. 1F and Fig. S1B). For example, the thousand-grain weights of *bsr-k1* (28.52 ± 0.38 g) and Kitaake (28.89 ± 0.23 g) are similar, and the average heading dates of *bsr-k1* (55.9 ± 1.1 d) and Kitaake (56.1 ± 1.4 d) are comparable (Fig. 1F). These results suggest that the *bsr-k1* allele does not penalize major agronomic traits.

Map-Based Cloning Identified the Causative Mutation of *bsr-k1*. To identify the *bsr-k1* locus, we generated three *F2* populations by crossing the resistant *bsr-k1* mutant with susceptible cultivars Kitaake and Joden, separately. Genetic analyses showed that the *bsr-k1* resistance phenotype was controlled by a single recessive locus (Dataset S1). We selected 995 *F2* plants from Joden × *bsr-k1* that exhibited high-level resistance against blast isolate 99-20-2 and determined the resistance level in each *F3* progeny (Dataset S1). By using 88 resistant *F2* individuals, we first mapped the *bsr-k1* locus to a 1.37-Mb interval between markers RM25789 and RM333 on chromosome 10 (Fig. 2A). Analysis of another 907 resistant *F2* individuals located it to a 50-kb region between insertion–deletion markers ID13 and ID16 (Fig. 2A) containing 11 predicted genes (Dataset S2). We then sequenced the genomic DNA sample bulked from 25 resistant BC1F4 individuals and Kitaake control DNA separately using whole-genome resequencing. In the 50-kb region, we identified a single nucleotide substitution (G2447A) located at the 3′ splice site of intron 8 in gene *Os10g0548200* (Fig. 2B and Fig. S2A). Sequencing of cDNA confirms that *bsr-k1* produces a variant *Os10g0548200* transcript with a 71-bp insertion derived from intron 8, causing a premature termination (Fig. S2B–D).
To confirm that the *bsr-k1* resistance phenotype was caused by this mutation, we introduced the *Os10g0548200* genomic DNA (9.6 kb) into *bsr-k1*. All 15 independent, complemented lines carrying wild-type *Os10g0548200* reverted to susceptibility to all seven blast isolates. We chose four of the complemented lines for further investigation. Punch inoculation with *M. oryzae* isolates 99-20-2 and ZE-1 revealed that the lesions in these complemented lines were similar in size to those in Kitaake whereas the *bsr-k1* mutant was resistant (Fig. 2 C–F). These complemented lines also show susceptibility to *Xoo* similar to Kitaake (Fig. S3A). We then generated two independent *Os10g0548200*-KO lines in Kitaake using the CRISPR/Cas9 approach (Fig. S3 B and C) and performed inoculation with *M. oryzae* and *Xoo*. These KO lines show enhanced resistance, displaying smaller lesion size, less *M. oryzae* DNA accumulation, and lower *Xoo* population than Kitaake (Fig. S3 D–F). In addition, plants overexpressing *Os10g0548200* show much larger lesions compared with Kitaake (Fig. S3 G–J). Taken together, these results demonstrate that the

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Fig. 2. Positional cloning of *bsr-k1*. (A) Fine mapping of the *bsr-k1* locus. The molecular markers and the number of recombinants are indicated. (B) Structure of the *Bsr-k1* gene and the mutation in *bsr-k1*. Filled boxes indicate exons (numbered 1–22) of *Os10g0548200*. The change from G to A in *Os10g0548200* in *bsr-k1* is indicated in the open box above exon 9. Nucleotide sequences between exon 8 and exon 9 in Kitaake and *bsr-k1* with deduced amino acid sequences are shown below. The substitution in *bsr-k1* (in red) abolishes the 3′ splice site of intron 8, resulting in premature termination of translation. (C–F) Blast-resistance test of *Bsr-k1*–complemented lines. (C and E) Photographs of four independent representative lines at 7 dpi with blast fungal isolates 99-20-2 (C) and ZE-1 (E). (D and F) Lesion lengths (mean ± SD, n > 10) of the complemented lines, *bsr-k1*, and Kitaake inoculated with blast isolates 99-20-2 (D) and ZE-1 (F). Different letters above bars indicate significant differences (P < 0.05, Tukey’s test). This punch inoculation was repeated twice with similar results.

Fig. 3. Expression pattern of *Bsr-k1* and subcellular localization of BSR-K1. (A) *Bsr-k1* RNA expression pattern. (B) *Bsr-k1* RNA levels (mean ± SEM, n = 3) post *M. oryzae* or mock inoculation. This experiment was repeated twice with similar results. (C) Schematic structure of BSR-K1. The nucleotide numbers above the cDNA indicate the locations of the five TPR motifs. The TPR motifs are represented as red boxes. (D) Subcellular localization of BSR-K1 in rice root tips. Photographs of representative root tips were taken under confocal laser-scanning microscopy. N, nucleus. (Scale bars: 20 μm.) (E) Subcellular localization of BSR-K1 in rice protoplasts. The white square in the merged image is enlarged and shown at the right. CM, mCherry-cytomembrane. (Scale bars: 5 μm.)
To investigate the expression pattern of RNA in the inoculation and found that \( Bsr-k1 \) is Constitutively Expressed and Encodes a Cytoplasm-Localized TPR, the \( Os11g0671000 \) mutation where a single nucleotide change results in enhanced resistance. The \( Bsr-k1 \) ORF encodes a protein of 1,196 amino acids containing five tandem TPRs (Fig. 3C). To determine the subcellular localization of the \( BSR-K1 \) protein, we used confocal laser-scanning microscopy to analyze the roots of plants carrying a functional GFP-tagged \( BSR-K1 \) protein, which complemented the \( bsr-k1 \) mutant (Fig. S3 G–J), and found that its GFP signal is excluded from the nucleus, whereas the GFP protein alone is present in both the cytoplasm and nucleus (Fig. 3D). We validated this result by expressing the GFP-BSR-K1 protein in \( N. benthamiana \) (Fig. S4) and in rice protoplasts, where the GFP-BSR-K1 protein was detected only in the cytoplasm but not in the nucleus or cytomembrane (visualized by the mCherry-cytomembrane marker, RFP-CM, in Fig. 3E). Together, these results suggest that \( BSR-K1 \) functions in the cytoplasm.

\( BSR-K1 \) Binds to \( OsPAL1 \) mRNA in Rice. TPR-containing proteins have been reported to regulate mRNA metabolism (23–25). We hypothesized that \( BSR-K1 \), containing TPRs, may associate selectively with specific RNA transcripts. To address this, we performed two independent experiments of in vitro RNA immunoprecipitation (RIP) assays by separately co-incubating GST and recombinant GST-BSR-K1 protein with rice total RNA (Fig. S5 A and B) and subsequently purifying GST, followed by high-throughput sequencing to identify \( BSR-K1 \)–associated mRNAs. RNA electrophoresis analysis revealed that GST-BSR-K1, but not GST alone, is able to bind to RNA in vitro (Fig. S5C). By comparing the RNAs coprecipitated with GST-BSR-K1 (RIP) and GST (mock), we identified 82 and 110 RNA fragments from these two RIP assays, respectively, that were specifically and significantly coprecipitated by the GST-BSR-K1 protein (Dataset S3), the truncated BSR-K1 protein in the \( bsr-k1 \) mutant, whereas the GFP protein alone is present in both the cytoplasm and nucleus (Fig. 3D). We validated this result by expressing the GFP-BSR-K1 protein in \( N. benthamiana \) (Fig. S4) and in rice protoplasts, where the GFP-BSR-K1 protein was detected only in the cytoplasm but not in the nucleus or cytomembrane (visualized by the mCherry-cytomembrane marker, RFP-CM, in Fig. 3E). Together, these results suggest that \( BSR-K1 \) functions in the cytoplasm.

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RIP precipitates (Fig. 4A). Further analysis showed that biotinylated OsPAL1 RNA was able to pull down the full-length BSR-K1 (GST-BSR-K1<sup>1-325</sup> aa) protein but not two truncated, non-functional BSR-K1 proteins, GST-BSR-K1<sup>1-165</sup> aa and GST-BSR-K1<sup>1-285</sup> aa (Fig. 4B). Together, these results demonstrate direct binding of the BSR-K1 protein to OsPAL1 mRNA in vitro.

We then generated transgenic plants carrying the native promoter-3xHA-Bsr-k1, which encodes HA-tagged BSR-K1 protein and complements the bsr-k1 mutant (Fig. S5D). We detected and immunoprecipitated the 3xHA-BSR-K1 fusion protein using anti-HA antibody (Fig. S5 E and F). We then performed an in vivo RIP assay using transgenic and Kitaake plants. qRT-PCR analysis showed that the OsPAL1 mRNA was pulled down by the 3xHA-BSR-K1 protein (Fig. 4C), confirming the binding of the BSR-K1 protein to OsPAL1 mRNA in vivo.

**BSR-K1 Regulates the OsPAL1 mRNA Level Through Modulating the OsPAL1 mRNA Turnover.** We next tested whether the OsPAL1 mRNA level is altered in bsr-k1. The results showed that OsPAL1 mRNA accumulated to a higher level (~threefold) in bsr-k1 than in Kitaake plants (Fig. 4D), indicating that BSR-K1 suppresses OsPAL1 mRNA accumulation in Kitaake plants.

We then compared the turnover rate of OsPAL1 mRNA in bsr-k1 and Kitaake. We designed primers for qPCR to detect the 5′ and 3′ end of OsPAL1 mRNA, respectively (Fig. S6A), because mRNA degradation generally starts from the 3′ end, and thus the 5′-end/3′-end ratio represents its turnover rate (28). The reverse primers for each fragment were used to prime separate cDNA synthesis reactions (28). We found that Kitaake has a 5′-end/3′-end ratio approximately twice as high as bsr-k1 (Fig. 4E), suggesting that the turnover rate of OsPAL1 mRNA is compromised in bsr-k1. Together, these results indicate that BSR-K1 binds OsPAL1 mRNA and promotes its turnover in Kitaake plants.

To determine whether the elevated OsPAL1 mRNA level contributes to resistance to rice blast, we overexpressed the OsPAL1 gene in a susceptible rice variety, TP309 (Fig. 4F), and performed blast inoculation. We found that the OsPAL1 overexpression lines showed enhanced resistance to blast with lesion lengths significantly (three- to sevenfold) shorter than those observed for TP309 (Fig. 4G and H), supporting the role of OsPAL1 in bsr-k1-mediated resistance. However, KO of OsPAL1 in bsr-k1 or Kitaake plants did not compromise resistance to blast (Fig. 4I and J and Fig. S6 B–E), suggesting that other OsPAL1 members may function redundantly and compensate for the loss of OsPAL1.

In support, we also find that bsr-k1 accumulates higher levels of OsPAL1–OsPAL4 (especially OsPAL1–OsPAL4) mRNAs than Kitaake (Fig. 4K) and that BSR-K1 binds OsPAL1–OsPAL7 mRNAs and facilitates their turnover in vivo (Fig. S6 F–H). For comparison, we also analyzed other defense-related genes, Os11g0703900, Os05g0343400 (WRKY53), and Os05g0293500, detected in RIP sequencing (RIP-seq) (Dataset S3C). BSR-K1 showed very weak or no binding to these genes (Fig. S6G). Consistently, none of their mRNA turnover was remarkably repressed (Fig. S6H); their mRNAs did not significantly accumulate compared with OsPAL1–7 in the bsr-k1 mutant (Fig. 4K). Together, these results indicate that BSR-K1 preferentially promotes OsPAL1 mRNA turnover in rice plants and that loss of function of BSR-K1 (bsr-k1) results in mRNA accumulation of multiple OsPAL genes, leading to enhanced resistance in bsr-k1.

**Plants Carrying Homozygous bsr-k1 Alleles Exhibit Enhanced Resistance with No Major Yield Penalty.** To assess the usefulness of the bsr-k1 allele for rice blast control, we performed field trials in two rice-growing regions. Field one (Pujiang, Sichuan, China) was a field with high blast infection (29); as a control, field two (Lingshui, Hainan, China) had little or no M. oryzae present. We found that the bsr-k1 plants displayed high levels of leaf and panicle resistance to M. oryzae in field one, whereas the Kitaake control plants succumbed to infection (Fig. 5A and Fig. S7).

We then evaluated the grain yield of bsr-k1 plants and Kitaake control plants in the two fields. In field two, where no obvious blast infection occurred, the bsr-k1 plants yielded a similar amount of grain as Kitaake plants (1.02 vs. 1.10 kg) (Fig. 5B). In contrast, in field one, where heavy blast infection occurred, the yield of bsr-k1 plants was significantly higher than that of Kitaake plants (0.72 vs. 0.49 kg) (Fig. 5B). Furthermore, in three field tests at two locations, elite rice cultivars carrying bsr-k1, including Shuhui527<sup>b</sup>~<sup>bsr-k1/bsr-k1</sup>, Minghui63<sup>b</sup>~<sup>bsr-k1/bsr-k1</sup>, and Mianhui725<sup>b</sup>~<sup>bsr-k1/bsr-k1</sup> (Fig. S8), showed agronomic traits and yields similar to those of their parents, except for slightly lower yields of Shuhui527<sup>b</sup>~<sup>bsr-k1/bsr-k1</sup> and Minghui63<sup>b</sup>~<sup>bsr-k1/bsr-k1</sup> in one test (in Pujiang, 2016) (Dataset S4).

**Discussion**

We have demonstrated that loss of function of the Bsr-k1 gene encoding a TPR-domain protein leads to enhanced resistance to all tested isolates of M. oryzae and Xoo. We conclude that the bsr-k1 mutant clearly confers broad-spectrum disease resistance. None of the previously reported genes (7–15) conferring broad-spectrum resistance encodes proteins with RNA-binding activity. Thus, BSR-K1 represents a class of protein regulating a pathway leading to broad-spectrum resistance. Loss of functional BSR-K1 (bsr-k1) results in mRNA accumulation of multiple OsPAL (OsPAL1–OsPAL7) genes (Fig. 4K). We hypothesize that BSR-K1 promotes mRNA degradation of these OsPAL genes and that loss of function of BSR-K1 results in lower turnover of their mRNAs, leading to mRNA accumulation.

Previous studies have shown that the expression of PAL1 genes is required for the biosynthesis of secondary metabolites, including lignin and SA, a phytohormone associated with defense against biotrophic pathogens (27). Our results show that bsr-k1 accumulates more lignin, but not total SA (Fig. S9 A and B), despite the elevation of defense-related genes such as OsPR1a in bsr-k1 (Fig. 1E). Consistently, the lignin content also accumulates in OsPAL1 overexpression plants (Fig. 4K), which exhibit enhanced resistance to rice blast (Fig. 4F–H), but the SA content remains unchanged (Fig. S9D). These results indicate that higher expression levels of OsPAL genes increase the biosynthesis of lignin, instead of SA, and resistance to pathogens in the bsr-k1 mutant. However, how the elevation of lignin content is associated with the activation of defense genes remains unknown.

TPR-containing proteins are involved in the regulation of RNA metabolism in various organisms (23–25). Our results demonstrate that the BSR-K1 protein regulates mRNA turnover of multiple OsPAL genes (Fig. 4E and Fig. S6F). Thus, BSR-K1 regulates plant immunity through a mechanism different from those of
previously identified RNA-binding proteins (30–32). For example, Arabidopsis MOS2 is required for proper splicing of SNC1 (30). Bean PRP-BP participates in the elicitor-induced destabilization of PPrP1 mRNA, which encodes a cell wall-strengthening protein, implicating PRP-BP in plant defense (31). Human HuR regulates the splicing of Ditr mRNA, encoding a subunit of the cytosolic dehydrogenase (a-KGDH) complex; absence of HuR results in large amounts of reactive oxygen species and B cell death (32).

In rice, it has been reported that TPR proteins mainly act as interaction scaffolds in the formation of multiprotein complexes involved in the immune response, such as in the defense of chitin-induced immunity (33). This defense complex includes two TPR proteins, OsSST1 and OsSt1a, which associate with the HSP90 immune complex to regulate chitin-triggered immunity and resistance against rice blast fungus (33). In contrast, BSR-K1 does not directly interact with the rice defense complex (Fig. S104). Thus, the action of the BSR-K1 TPR protein represents a mechanism leading to immunity regulated through interaction with mRNA. Because the TPR motifs are highly conserved in diverse organisms (34), we hypothesize that TPR proteins, especially those sharing high homologies to BSR-K1, including Arabidopsis AtSK3 (Fig. S106), may also be involved in regulating immune responses via similar mechanisms.

In practice, most breeding programs rely on dominant resistance (R) genes (35), which often confer race-specific resistance and are overcome by the evolution of new pathogen races within a few years (36). Although a number of genes or QTLs for broad-spectrum and durable resistance have been identified (7–15), the tight linkage to undesirable agronomic traits has often hindered breeding. The bsr-k1 mutant confers broad-spectrum resistance with no major penalty on main agronomic traits (Fig. 1F and Fig. S118). The moderately elevated activation of defense-related genes in bsr-k1 (Fig. 1E) is sufficient to suppress pathogen attack, similar to the putative characteristics of p21-mediated resistance (10). Indeed, the bsr-k1 allele strikingly improves rice resistance in Kitaake in a naturally blast-diseased field compared with the wild-type bsr-k1 allele (Fig. S4 and Fig. S7). Three field tests of elite rice cultivars suggest that the bsr-k1 allele does not carry a major penalty affecting main agronomic traits or yield (Dataset S4), making it a good candidate for rice breeding.

Materials and Methods
Rice blast and Xoo infections and disease assessments were performed as described (21, 22, 29). Purified recombinant proteins from Escherichia coli were used for in vitro RIP assays and RNA pulldown assays. Immunoprecipitated recombinant proteins from transgenic rice were used for in vivo RIP assays. In vitro RIP assays and in vivo RIP assays were performed as described (37). The RNA pulldown assay was performed using the Pierce Magnetic RNA-Protein Pull-Down Kit following the manufacturer’s instructions (Thermo Scientific). Determination of mRNA turnover was performed as described (28). The blast isolates used to identify Kitaake-compatible blast isolates are listed in Dataset S5. The primers used for qRT-PCR are listed in Dataset S6. All other materials and details of experimental methods can be found in SI Materials and Methods.

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