The Phosphatase PTP-PEST/PTPN12 Regulates Endothelial Cell Migration and Adhesion, but Not Permeability, and Controls Vascular Development and Embryonic Viability*

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PROTEIN-TYROSINE PHOSPHATASE (PTP)-PEST is a ubiquitously expressed cytosolic PTP with an amino (N)-terminal region involved in protein-protein interactions. It is essential for normal embryonic development and embryonic viability in mice. Herein we addressed the involvement of PTP-PEST in endothelial cell functions using a combination of genetic and biochemical approaches. By generating primary endothelial cells from an inducible PTP-PEST-deficient mouse, we found that PTP-PEST is not needed for endothelial cell differentiation and proliferation or for the control of endothelial cell permeability. Nevertheless, it is required for integrin-mediated adhesion and migration of endothelial cells. PTP-PEST-deficient endothelial cells displayed increased tyrosine phosphorylation of Cas, paxillin, and Pyk2, which were previously also implicated in integrin functions. By eliminating PTP-PEST in endothelial cells in vivo, we obtained evidence that expression of PTP-PEST in endothelial cells is required for normal vascular development and embryonic viability. Therefore, PTP-PEST is a key regulator of integrin-mediated functions in endothelial cells seemingly through its capacity to control Cas, paxillin, and Pyk2. This function explains at least in part the essential role of PTP-PEST in embryonic development and viability.

Protein-tyrosine phosphatase (PTP)-PEST (or PTPN12) is ubiquitously expressed. It is essential for normal embryonic development and embryonic viability in mice. Herein we addressed the involvement of PTP-PEST in endothelial cell functions using a combination of genetic and biochemical approaches. By generating primary endothelial cells from an inducible PTP-PEST-deficient mouse, we found that PTP-PEST is not needed for endothelial cell differentiation and proliferation or for the control of endothelial cell permeability. Nevertheless, it is required for integrin-mediated adhesion and migration of endothelial cells. PTP-PEST-deficient endothelial cells displayed increased tyrosine phosphorylation of Cas, paxillin, and Pyk2, which were previously also implicated in integrin functions. By eliminating PTP-PEST in endothelial cells in vivo, we obtained evidence that expression of PTP-PEST in endothelial cells is required for normal vascular development and embryonic viability. Therefore, PTP-PEST is a key regulator of integrin-mediated functions in endothelial cells seemingly through its capacity to control Cas, paxillin, and Pyk2. This function explains at least in part the essential role of PTP-PEST in embryonic development and viability.

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Background: PTP-PEST is a phosphatase essential for embryonic viability.

Results: PTP-PEST is critical for adhesion and migration of endothelial cells. Its absence in endothelial cells results in mouse embryonic lethality.

Conclusion: The embryonic viability seen in constitutive PTP-PEST-deficient mice is due to a defect in endothelial cell functions.

Significance: PTP-PEST is a key regulator of endothelial cell functions in vitro and in vivo.

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blood vessel formation from pluripotent mesenchymal cells, whereas angiogenesis implicates blood vessel formation from existing vessels. Both require extensive endothelial cell proliferation and migration.

Blood vessel integrity is maintained by tight intercellular junctions between endothelial cells as well as by interactions between endothelial cells and surrounding basement membranes, stromal cells, and smooth muscle cells (21–24). Adhesion receptors, especially vascular endothelial (VE)-cadherin and integrins, mediate these various contacts. The VE-cadherin pathway controls vascular permeability (20, 25, 26). It is tightly regulated by PTKs such as Tie2, which dynamically modulates the function of VE-cadherin (25, 26). A PTP, VE-PTP, influences VE-cadherin function through its ability to regulate Tie2 (27, 28). In contrast, the integrin pathway influences adhesion and migration (29–32). The PTK FAK was shown to be a central regulator of the integrin pathway in endothelial cells (29–32). However, the identity of the PTP(s) influencing this pathway in endothelial cells has not been determined.

Here we examined the role of PTP-PEST in endothelial cells. Using primary endothelial cells from an inducible PTP-PEST-deficient mouse, we found that PTP-PEST is required for migration and adhesion of endothelial cells, but not for proliferation, differentiation, and modulation of permeability to macromolecules by these cells. This function correlates with the capacity of PTP-PEST to dephosphorylate Cas, pyk2, and Pyk2. Studies of a mouse in which PTP-PEST was specifically eliminated in endothelial cells provided evidence that the function of PTP-PEST in endothelial cells is critical for normal vascular development and embryonic viability.

**Experimental Procedures**

*Mice—*Mice expressing a tamoxifen-inducible Cre (Cre-ERT2) under the control of the ubiquitin C promoter (UBC-Cre-ERT2) or the Cre recombinase under the control of the Tie2 promoter (Tie2-Cre), and ROSA26EYFP mice were obtained from The Jackson Laboratory (Bar Harbor, ME) (33–35). They were then bred with mice expressing a conditionally deleted allele of the PTP-PEST-encoding gene, Ptpn12fl/fl, backcrossed for at least 15 generations to the C57BL/6 background (11). A constitutive PTP-PEST-deficient mouse (Ptpn12−/−) in which PTP-PEST expression was constitutively lacking in all cells was generated by breeding Ptpn12+/+ mice with mice expressing a ubiquitous Cre transgene (11). Timed matings were set up between male Ptpn12fl/+;Tie2-Cre+ and female Ptpn12fl/fl mice. Day 0.5 of embryonic development (E0.5) was defined as the morning when the vaginal plug was observed. All mice used in this study were bred under specific pathogen-free conditions. Animal experimentation was approved by the Clinical Research Institute of Montreal Animal Care Committee in accordance with the regulations of the Canadian Council for Animal Care.

*Inducible Deletion of PTP-PEST in Adult Mice—*Female Ptpn12fl/fl mice were bred with male Ptpn12fl/fl;UBC-Cre-ERT2 mice. For tamoxifen-induced deletion of PTP-PEST, mice were fed for 5 consecutive days with tamoxifen (200 µg/g of weight/day in corn oil; Sigma-Aldrich). After 10 days, mice were sacrificed and used for endothelial cell isolation.

**Isolation of Lung-derived Endothelial Cells—**Primary lung-derived endothelial cells were extracted from lung tissue of 8–10-week-old mice as described below for embryonic endothelial cells. Cells were then cultured in low glucose DMEM/F-12 medium (1:1 mixture; Invitrogen) supplemented with 20% fetal bovine serum (FBS; Hyclone, Logan, UT), endothelial growth supplement (5 µg/ml; BD Biosciences), and heparin (100 µg/ml; Sigma-Aldrich) using gelatin-coated tissue culture dishes. Two successive rounds of endothelial cell purification were then performed using rat anti-mouse ICAM-2 antibody (mAb 3C4, BD Biosciences) coupled to sheep anti-rat IgG-coated magnetic beads (Dynabeads, Invitrogen). The purity of endothelial cells was confirmed by flow cytometry analysis using antibodies directed against the endothelial cell markers CD31 (rat anti-mouse CD31 mAb 390, eBioscience, San Diego, CA) and ICAM-2 (rat anti-mouse mAb 3C4, Biolegend, San Diego, CA). After 16 days in culture, cells were used for experimentation. At that time, >95% of cells expressed the endothelial cell-specific markers.

**Microscopy Analyses of Mouse Embryos—**Embryos and yolk sacs were harvested from 8.5- to 9.5-day-old pregnant dams, washed, minced, and digested at 37 °C for 1 h with collagenase IV (1 mg/ml; Sigma-Aldrich). The cell suspension was washed through a 70 µm cell strainer and then plated in growth medium (F-12 medium and 20% FBS). After 16 days in culture, cells were treated with 0.5% Triton X-100 and stained with Alexa Fluor 488-conjugated rat anti-mouse ICAM-2 antibody. Alexa Fluor 488-conjugated rat anti-mouse ICAM-3 antibody (mAb 1H10, Biolegend, San Diego, CA) was used as a vascular endothelial (VE)-cadherin marker. Stained cells were imaged using a confocal microscope (Zeiss). For cross-section analyses, fixed embryos were incubated overnight at 4 °C in a PBS solution containing 2% formaldehyde (Canemco, Québec, Canada). After blocking for 2 h at room temperature in a PBS solution containing 2% skim milk and 2.5% Triton X-100, embryos were incubated overnight with purified rat anti-mouse CD31 monoclonal antibody (mAb MEC13.3, BD Biosciences). After several washes, embryos were incubated overnight at 4 °C in the presence of a 1:50 dilution of Alexa Fluor 488-conjugated goat anti-rat IgG (Invitrogen). After several washes, images were obtained by confocal microscopy using an LSM710 microscope (Zeiss). For cross-section analyses, fixed embryos were embedded in Epon, sectioned (0.5-µm thickness), stained with toluidine blue, and visualized by microscopy (Axioskop, Zeiss) as described above.

**Isolation of Embryonic Endothelial Cells—**Male Ptpn12fl/fl; Rosa26EYFP+/Tie2-Cre+ mice were bred with female Ptpn12fl/fl; ROSA26EYFP+ mice. E10.5 embryos were harvested from timed pregnancies, washed, minced, and digested at 37 °C for 1 h with collagenase A (1 mg/ml; Roche Applied Science). To remove undigested material, the cell suspension was washed through a cell strainer (BD Biosciences). Isolated cells were then incubated on ice for 1 h with the antibodies described below. Cells were washed, and enhanced yellow fluorescent protein-positive (EYFP+) cells were analyzed by flow cytometry (FACSCalibur, BD Biosciences).

**Flow Cytometry—**Embryonic cells or primary lung-derived endothelial cells were stained and analyzed by flow cytometry using the following antibodies (from Biolegend, BD Biosciences, or eBioscience): Alexa Fluor 647-conjugated rat anti-mouse CD31 (mAb 390), Alexa Fluor 647-conjugated rat anti-mouse ICAM-1 (mAb YN1/1.7.4), Alexa Fluor 647-conjugated rat anti-mouse ICAM-2 (mAb 3C4), Alexa Fluor 647-conjugated rat anti-
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mouse VE-cadherin (mAb VECD1), phycoerythrin-coupled hamster anti-mouse β1 integrin (CD29; mAb HMB1-1), fluorescein isothiocyanate (FITC)-coupled rat anti-mouse β2 integrin (CD18; mAb M18/2), and rat anti-mouse isotype control (mAb RTK2758).

Proliferation Assays—Primary lung-derived endothelial cells (4 × 10^4 cells) were seeded in 24-well, gelatin-coated tissue culture dishes. At the desired time point, cells were harvested by trypsin (Invitrogen) and counted.

Adhesion Assays—96-well flat-bottom Immuno plates (Nunc, Thermoscientific, Rochester, NY) were coated overnight with fibronectin (10 μg/ml; BD Biosciences), type I collagen (10 μg/ml; BD Biosciences), or bovine serum albumin (BSA; 2 mg/ml; Sigma-Aldrich). After washing, plates were blocked with BSA, and 10^4 endothelial cells were allowed to adhere for 30 min at 37°C. After washing away non-adherent cells, adherent cells were fixed and stained with crystal violet (Fisher Scientific). Cell adhesion was estimated by measuring the absorbance at 562 nm in a microplate scanning spectrophotometer (Power Wave X, Bio-Tek Instruments, Inc., Winooski, VT). Photographs were also obtained using an inverted microscope (Model Axiovert S100TV, Zeiss) with a QImaging camera and Volocity image analysis software (PerkinElmer Life Sciences).

Spreading Assays—Coverslips were coated at 4°C overnight with fibronectin or collagen as above. Then coverslips were washed with PBS and placed in 6-well dishes. Endothelial cells were harvested and seeded (1 × 10^5 cells) on the coverslips. After 90 min at 37°C, unbound cells were washed, and the remaining cells were fixed with 2% paraformaldehyde and mounted on a glass slide for examination by contrast microscopy. Data from 10 independent fields were acquired.

Migration Assays—For Transwell migration assays, endothelial cells were deprived of growth supplement and FBS overnight. Then 10^5 cells were loaded in the upper chamber of a Transwell insert (8-μm pores; Costar, Corning, Lowell, MA) previously coated with fibronectin (5 μg/ml). The medium in the lower chamber was then replaced by medium containing 20% FBS, endothelial growth supplement (5 μg/ml; BD Biosciences), and heparin (100 μg/ml; Sigma-Aldrich). Cells were allowed to migrate for 24 h. Cells having migrated in the lower chamber were counted using Volocity image analysis software. For wound healing assays, endothelial cells were cultured in gelatin-coated 6-well tissue culture plates (Nunc). Once the monolayer was confluent, a scratch was performed using a 200-μl pipette tip. Wound closure was monitored with a time lapse microscope (DM IRE2, Leica), capturing images every 10 min. The speed of migration and persistence index, which is calculated as the ratio of the distance between the first and the last points of the tracked cells over the total distance traveled, were assessed after labeling cells with Hoechst using Volocity image analysis software. Image analyses and measurements were performed using Matlab software (MathWorks Inc., Torrance, CA).

Permeability Assays—Permeability was measured using type I collagen-coated Transwell units (0.4-μm pores; Costar). The indicated numbers of endothelial cells were seeded in the upper chamber and allowed to form a monolayer. Once a monolayer was formed, cells were deprived of serum and endothelial growth supplement for 1 or in some cases 4 h. After deprivation, FITC-dextran (molecular weight, 40,000; 1 mg/ml; Invitrogen) was added to the upper chamber and allowed to diffuse into the lower chamber for 30 or 60 min at 37°C in the absence or presence of vascular endothelial growth factor (VEGF; 75 ng/ml) (36). Endothelial permeability was determined by measuring the fluorescence in the lower chamber at 520 nm with an excitation wavelength of 492 nm. Triplicates measurements were taken.

Immunoprecipitation and Immunoblots—Cells were lysed directly on the plate with TNE buffer (50 mM Tris, pH 8.0, 1% Nonidet P-40, 2 mM EDTA) supplemented with protease and phosphatase inhibitors as described (37). Immunoprecipitation was accomplished by incubating 400 μg of total cell lysates with a protein-specific antibody for 45 min at 4°C. To collect immune complexes, Pansorbin (Calbiochem, EMD Millipore) was added for an additional 45 min. Unbound proteins were washed away, and the precipitated protein was loaded for 8% SDS-PAGE. Immunoblots were performed as outlined elsewhere (37). The following antibodies were used for immunoprecipitation: Cas (sc-860, Santa Cruz Biotechnology, Santa Cruz, CA), paxillin (610272, BD Biosciences), and FAK (sc-558, Santa Cruz Biotechnology). Phosphospecific antibodies against FAK (Tyr(P)-397 (3483) and Tyr(P)-576/577 (3281)) and antibodies recognizing phospho-extracellular signal-regulated kinase (ERK) (mAb E10) were from Cell Signaling Technology (Danvers, MA). The following antibodies were used for reprobing the membrane: Cas (sc-860 (Santa Cruz Technology, Santa Cruz, CA)), paxillin (610272, BD Biosciences), FAK (610088, BD Biosciences), and ERK (06-182, Millipore, Burlington, Ontario, Canada). Rabbit antibodies directed against Pyk2, PTP-PEST, and Csk were described elsewhere (12, 38). To evaluate VEGF-induced signals, endothelial cells were deprived of serum and endothelial growth supplement for 2–4 h. They were then stimulated with VEGF (50 ng/ml) for the indicated periods of time, and total cell lysates were probed by immunoblotting with antibodies recognizing phospho-ERK.

Statistical Analyses—t test analyses were performed using GraphPad version 5.00 for Mac (GraphPad Software, San Diego, CA).

RESULTS

PTP-PEST Is Not Required for Endothelial Cell Proliferation and Differentiation in Vitro—The midgestation embryonic lethality previously observed in a constitutive PTP-PEST-deficient mouse prevented establishing primary cultures of PTP-PEST-deficient endothelial cells to address the possible role of PTP-PEST in endothelial cells (13). To resolve this issue, we bred mice carrying a conditional allele of the PTP-PEST-encoding gene (Ptpn12^[fl/fl]) with mice expressing an estrogen-responsive Cre (Cre-ERT2), under the control of the ubiquitin C (UBC) promoter (Fig. 1A) (34). In the absence of tamoxifen, Cre-ERT2 is cytosolic and not functional. However, when mice are fed tamoxifen, Cre-ERT2 translocates to the nucleus where it can mediate deletion of the conditional allele. This enables deletion of the Ptpn12 gene in adult mouse tissues.
PTP-PEST Regulates Integrin-mediated Adhesion and Spreading in Endothelial Cells—Integrins are adhesion molecules involved in cell-cell and cell-substratum interactions (23, 29, 32). Compared with control cells, endothelial cells lacking PTP-PEST had more irregular sizes and shapes (Fig. 2A). To address the possibility that this alteration was the result of abnormal integrin function, the ability of endothelial cells to adhere in response to the integrin ligands fibronectin and collagen was tested (Fig. 2B). Fibronectin binds α5β1 and αvβ3 integrins, whereas collagen recognizes α1β1 and α2β1 integrins on endothelial cells. After 30 min of adhesion to integrin ligand-coated surfaces, cells were washed, and adherent cells were detected by staining with crystal violet. In comparison with cells expressing PTP-PEST, cells lacking PTP-PEST had a decrease in adhesion to fibronectin and collagen (Fig. 2B).

Once cells adhere to an integrin ligand-coated surface, they undergo integrin-dependent signaling events that lead to cell spreading. In light of this, the ability of endothelial cells to spread in response to integrin ligands was evaluated (Fig. 2C). Cells were added to fibronectin- or collagen-coated glass cover slips. After 90 min, non-adherent cells were washed away, and spreading of the remaining adherent cells was assessed. Whereas PTP-PEST-deficient cells exhibited less frequent adhesion than control cells, they underwent more frequent spreading (Fig. 2C).

Therefore, PTP-PEST was critical for integrin-mediated adhesion of endothelial cells. It also regulated endothelial cell spreading.

PTP-PEST Is Not Required for Integrity of Endothelial Cell-Cell Junctions—Endothelial wall integrity in the embryo and postnatal life is maintained by close contacts between endothelial cells (20–22). This is mediated in large part by VE-cadherin, a self-binding endothelial cell-specific member of the cadherin family of adhesion molecules. Given the impact of PTP-PEST on integrin-mediated adhesion noted above, we also examined the role of PTP-PEST in endothelial cell permeability. To this end, we first stained primary endothelial cells expressing or not expressing PTP-PEST with antibodies against VE-cadherin, which identifies cell-cell junctions (Fig. 3A). Staining was then...
detected by confocal microscopy. As was the case for control cells, PTP-PEST-deficient cells exhibited staining with anti-VE-cadherin antibodies at the cell surface. Although cell sizes and shapes were distinct between the two cell populations, the anti-VE-cadherin antibody-decorated intercellular junctions did not seem to be appreciably affected by PTP-PEST deficiency.

Next, endothelial cell integrity was examined using a vascular permeability assay (Fig. 3, B and C). In this assay, we monitored the ability of a small molecule, FITC-dextran, to diffuse across endothelial cells plated on a membrane with small pores (0.4-μm pores). First, we assessed basal permeability (Fig. 3B). By plating progressively higher numbers of endothelial cells, we found that lack of PTP-PEST had no appreciable effect on the ability of endothelial cells to block transfer of FITC-dextran. Second, we examined the ability of the growth factor VEGF to induce an increase in vascular permeability (Fig. 3C). In three independent experiments, the absence of PTP-PEST had no impact on the capacity to augment transfer of FITC-dextran.

These findings indicated that PTP-PEST did not influence basal or VEGF-induced permeability of endothelial cells. They also implied that PTP-PEST had no effect on VE-cadherin function.

PTP-PEST Is Required for Endothelial Cell Migration—Next, we ascertained the impact of PTP-PEST on the ability of endothelial cells to migrate, which is required for vascular development (Fig. 4). Migration was first analyzed using a Transwell migration assay (Fig. 4A). Endothelial cells were put in the upper chamber of a Transwell apparatus (4-μm pores), and their migration toward the bottom chamber, which contained endothelial growth supplement and serum, was analyzed. Lack of PTP-PEST significantly reduced (by ~60%) the ability of endothelial cells to migrate in this assay.

Migration was also examined in a wound healing assay (Fig. 4, B–E). Primary endothelial cells were plated and, when confluent, inflicted with a scratch wound. Cell migration was then monitored over 24 h by time lapse microscopy and still photography (Fig. 4B). Endothelial cells from control mice migrated toward the wound and enabled its closure after 18 h. Although PTP-PEST-deficient endothelial cells also migrated toward the wound, they failed to close the wound after 24 h. Quantitation of the speed of wound closure (Fig. 4C) and the area repaired...
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We also analyzed the impact of PTP-PEST deficiency on VEGF-induced signals (Fig. 5D). Cells were stimulated or not for various periods of time with VEGF, and activation of ERK was monitored by immunoblotting total cell lysates with antibodies recognizing activated phospho-ERK. The lack of PTP-PEST had little or no effect on VEGF-induced ERK activation in keeping with the lack of effect on vascular permeability noted earlier.

Thus, the lack of PTP-PEST in endothelial cells resulted in augmented tyrosine phosphorylation of Cas, paxillin, and, to a lesser extent, Pyk2. All three substrates are known regulators or effectors of the integrin pathway. However, it had no obvious effect of VEGF-induced signals.

PTP-PEST Expression in Endothelial Cells Is Necessary for Vascular Development and Embryonic Viability—Previous analyses of a constitutive PTP-PEST-deficient mouse in which PTP-PEST was missing from all cells showed that PTP-PEST is critical for embryonic development and viability (13). To examine whether this was due to a role of PTP-PEST in endothelial cells, Ptpn12fl/fl mice were crossed with mice expressing Cre under the control of the Tie2 (or Tek) promoter (Tables 1 and 2 and Fig. 6) (29, 32, 33). Tie2-Cre is active in all endothelial cells and a subset of hematopoietic cells and is widely used to delete conditional alleles in endothelial cells. Heterozygous Ptpn12+/−; Tie2-Cre+ mice were then bred with Ptpn12fl/fl mice. Strikingly, genotype analyses of a total of 339 mice born from this cross failed to identify any Ptpn12fl/fl; Tie2-Cre+ pups (Table 1). Normally, ~25% of live pups would be expected to be Ptpn12fl/fl; Tie2-Cre+. This finding suggested that, as reported for consti-
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**FIGURE 4. Reduced migration by PTP-PEST-deficient endothelial cells.** A, the migration of primary endothelial cells from tamoxifen-fed Ptpn12fl/fl;UBC-Cre-ERT2+ (control (CTRL)) or tamoxifen-fed Ptpn12fl/fl;UBC-Cre-ERT2+ (KO) mice was examined in a Transwell migration assay. Data from three independent experiments with average values and error bars representing standard deviations are graphically presented. B–F, wound healing assay. A scratch wound was inflicted in monolayers of endothelial cells. The closure of the wound was monitored by time lapse microscopy. Still images taken at various times are shown in B, and the speed of migration is presented in C. Error bars representing standard deviations of triplicates with p values and mean values are shown. The surface of the wound covered by cells (depicted as pixels covered) was also estimated over time (D). Lastly, individual cells (stained with Hoechst) were tracked over a period of 6 h, and the direction and extent of their migration were analyzed (E). In the latter analysis, plotting the angle with respect to the initial position of the cell in a rose plot diagram depicts directionality of cell migration. The area of each bin represents the number of cells moving in the same direction of migration. For simplification, all cells in (E) were analyzed. 

Error bars representing standard deviations are shown. Image analyses and measurements were performed using Velocity software (A and C) or Matlab software (D–F). Data are representative of 15 (B and D), seven (C), or three (E and F) experiments.

We also performed a side-by-side comparison of anti-CD31-stained Ptpn12fl/fl;Tie2-Cre+ embryos with constitutive Ptpn12+− embryos (Fig. 6B). As reported elsewhere (13), constitutive PTP-PEST-deficient embryos died at ~E9.5–10.5, that is ~1 day earlier than Ptpn12fl/fl;Tie2-Cre+ embryos. Moreover, at E9.5 and E10.5, they were smaller and more distorted and showed more extensive loss of vascular sprouting in comparison with Ptpn12fl/fl;Tie2-Cre+ embryos. Possible explanations for the distinctions between Ptpn12fl/fl;Tie2-Cre+ and Ptpn12+− embryos will be provided under Discussion.

To assess whether lack of PTP-PEST affected endothelial cell differentiation in vivo, Ptpn12fl/fl;Tie2-Cre+ mice were bred with ROSA26-EYFP mice in which a stop sequence flanked by loxP sites is inserted upstream of the EYFP gene to prevent expression of EYFP (Fig. 6D). In the presence of Cre, however, the stop sequence is removed, thereby enabling expression of EYFP and detection of Ptpn12-deleted cells (35). Flow cytometry analyses of EYFP+ cells from disaggregated embryos revealed that large proportions of cells from PTP-PEST-deficient mice expressed CD31, ICAM-1, and ICAM-2. However, the proportions of cells expressing higher levels of these mark-
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To generate PTP-PEST-deficient endothelial cells, Ptpn12fl/fl mice were bred with UBC-Cre-ERT2 transgenic mice. Adult Ptpn12fl/fl,UBC-Cre-ERT2− mice were then fed tamoxifen, and primary cultures of lung-derived endothelial cells were established. These cells exhibited a total absence of PTP-PEST expression. Despite this, they proliferated normally in vitro and displayed normal expression of endothelial cell markers. The lack of effect of PTP-PEST deficiency on proliferation or differentiation of endothelial cells was consistent with similar findings made in other cell types, namely fibroblasts and T cells in which lack of PTP-PEST had no appreciable impact on these functions (7, 11).

Nonetheless, PTP-PEST deficiency resulted in alterations of the morphology of endothelial cells. Moreover, although integrin expression was not altered, PTP-PEST-deficient endothelial cells had markedly reduced adhesion to fibronectin- or collagen-coated surfaces. Once adhered to these surfaces, they also exhibited enhanced spreading. In addition, PTP-PEST-deficient endothelial cells exhibited a pronounced defect in migration. This effect was noted in Transwell migration and wound healing assays. In the latter assay, lack of PTP-PEST reduced the speed of migration but had no effect on directional movement. From this observation, it was inferred that PTP-PEST is not required for sensing environmental cues that trigger migration but rather is needed for the migration process itself.

Although PTP-PEST is necessary for homotypic adhesion between activated T cells (11), it was not needed for homotypic interactions between endothelial cells (this report). In support of the latter, staining with anti-VE-cadherin antibodies showed that the intercellular junctions between endothelial cells were not detectably affected by lack of PTP-PEST. Additionally, in a vascular permeability assay using the small molecule FITC-dextran, endothelial cell integrity was not compromised in the absence or presence of VEGF. Although this distinction between T cells and endothelial cells might seem surprising, it likely reflects differences in the molecular basis of the homotypic interactions between these cells. Interactions between T cells are principally mediated by integrins, whereas interactions...
between endothelial cells are primarily due to VE-cadherin. Seemingly, PTP-PEST influences the function of integrins but not that of VE-cadherin. It is probable that dysregulated tyrosine phosphorylation of Cas, paxillin, and Pyk2 explains the adhesion and migration defects seen in PTP-PEST-deficient endothelial cells. All three molecules are known regulators of the integrin pathway (23, 24). More importantly, previous data showed that Cas plays a critical role in integrin-mediated endothelial cell migration and vascular development (39, 40). Likewise, Pyk2 was shown to be involved in the control of angiogenesis (41). At this time, however, we cannot rule out that additional, as yet unidentified, substrates also contributed to the adhesion and migration defects seen in endothelial cells lacking PTP-PEST.

To examine the role of PTP-PEST expression in endothelial cells in vivo, we crossed Ptpn12fl/fl mice with Tie2-Cre mice. Genotype analyses of embryos and born pups showed that all Ptpn12fl/fl;Tie2-Cre+/H11001 embryos died in utero between E10.5 and E11.5. Furthermore, prior to death, these embryos showed markedly retarded growth. By staining embryos for CD31, it

**TABLE 2**

Distribution of genotypes in embryos from cross between Ptpn12fl/fl;Tie2-Cre+ and Ptpn12fl/fl

| Genotype         | E9.5   | E10.5  | E11.5  | E12.5  | E13.5  |
|------------------|--------|--------|--------|--------|--------|
| Ptpn12fl/fl;Tie2-Cre+ | 28 (23.0%) | 39 (25.8%) | 0 (0%) | 0 (0%) | 0 (0%) |
| Ptpn12fl/fl;Tie2-Cre−  | 29 (23.8%) | 39 (25.8%) | 38 (33.9%) | 35 (33.0%) | 38 (33.0%) |
| Ptpn12fl/fl;Tie2-Cre+  | 32 (26.2%) | 36 (23.8%) | 36 (32.2%) | 35 (33.0%) | 39 (33.9%) |
| Ptpn12fl/fl;Tie2-Cre−  | 33 (27.0%) | 37 (24.6%) | 38 (33.9%) | 36 (34.0%) | 38 (33.1%) |
| Total            | 112 (100%) | 151 (100%) | 112 (100%) | 106 (100%) | 115 (100%) |

**FIGURE 6. Analyses of Ptpn12fl/fl;Tie2-Cre+ embryos.** A, E9.5 and E10.5 embryos from Ptpn12fl/fl;Tie2-Cre+ (KO) and Ptpn12+/−;Tie2-Cre+ (control (CTRL)) mice were photographed. Data are representative of >9 experiments. B, whole-mount staining with anti-CD31 antibodies of E9.5 (panels a–i) and E10.5 (panels j–r) embryos from KO and control mice. Embryos from a constitutive PTP-PEST-deficient mouse (TOTAL KO) are also shown for comparison. Regions defined in the blue-bordered insets in panels a–c and j–l were analyzed at greater magnification in panels d–i and m–r. Scale bars for panels A–C represent 800 μm, whereas those for panels D–L represent 1000 μm. Data are representative of two (E9.5) or three (E10.5) experiments. C, toluidine blue-stained cross-sections of E10.5 embryos from control and KO mice. For each mouse, the region defined in the inset in the left-hand panel was analyzed at greater magnification in the right-hand panel. Arrows show the location of the dorsal aorta. D, flow cytometry analyses of cells from disaggregated E10.5 embryos from control and KO mice. Analyses were gated on EYFP-positive cells. The percentages of cells positive for the various markers are shown in the upper right quadrant. Data are representative of three experiments.
was found that Ptpn12 deletion resulted in compromised vascular development, in particular angiogenesis. This was likely due to a defect in endothelial cell adhesion and migration as both processes are critical for normal vascular development (20, 24). The time at which embryonic lethality occurred was also consistent with a vascular defect. Indeed, several other gene mutations affecting the vasculature result in embryonic lethality at ~E9.5–10.5 (20, 25–27, 29–32, 39).

Of note, however, the embryonic lethality observed in Ptpn12fl/fl;Tie2-Cre* mice arose approximately 1 day later than that previously noted in constitutive PTP-PEST-deficient mice (13). Moreover, a side-by-side comparison showed that Ptpn12−/− embryos showed more extensive loss of vascular sprouting in comparison with Ptpn12fl/fl;Tie2-Cre embryos. These disparities might be explained by the inducible nature of conditional Cre-mediated deletion. Although Tie2-Cre is expressed as early as E6.5, deletion of both copies of the conditional allele and degradation of all PTP-PEST RNA and protein (the half-life of the PTP-PEST protein has been estimated to be at least 4 h (42)) are needed for complete PTP-PEST deficiency. This might require a few days and likely enabled partial expression of PTP-PEST between E6.5 and E10.5. Alternatively, it might reflect a role of PTP-PEST in additional cell types that are not targeted by Tie2-Cre.

Adhesion receptors play critical roles in vascular functions during embryogenesis and adult life. The VE-cadherin pathway is a key regulator of vascular permeability. Previous data showed that Tie2, a PTK, and VE-PTP, a PTP, regulate the activity of VE-cadherin and that both are essential for vascular development and embryogenesis (20, 25–27). By opposition, the integrin pathway is important for the control of endothelial cell adhesion and migration. Earlier results, in addition to the findings herein, revealed that another PTK, FAK, and another PTP, PTP-PEST, control the activity of integrins. Both also have pivotal roles in vascular and embryonic development (29–32). In this manner, unique pairs of PTKs and PTPs regulate the two major adhesion receptor families controlling vascular functions.

Together, the findings reported herein show that PTP-PEST is a key regulator of integrin-mediated adhesion and migration of endothelial cells in vivo and in vitro. This function is likely mediated by the ability of PTP-PEST to dephosphorylate the cytoskeleton regulators Cas, paxillin, and Pyk2. Moreover, such an activity likely explains the critical role of PTP-PEST in vascular development and embryonic viability. Given the continuing importance of angiogenesis after birth, it is probable that PTP-PEST also plays a key function in postnatal processes involving endothelial cells. These processes may include wound healing and tissue repair and pathological situations such as tumor formation, inflammation, and diabetic retinopathy. The possible involvement of PTP-PEST in pathological angiogenesis raises the possibility that pharmacological inhibitors of PTP-PEST or of PTP-PEST-regulated pathways may have therapeutic benefit in these conditions.

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