Amorphous aggregation of tau in the presence of titanium dioxide nanoparticles: biophysical, computational, and cellular studies

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\textbf{Background:} Nanoparticles (NPs) when injected into the body can reach target tissues like nervous system and interact with tau proteins and neurons. This can trigger conformational changes of tau and may affect NP toxicity.

\textbf{Methods:} In this study, we used several biophysical techniques (extrinsic and intrinsic fluorescence spectroscopy, circular dichroism (CD) spectroscopy, ultraviolet (UV)-visible spectroscopy), transmission electron microscopy (TEM) investigations, molecular docking and molecular dynamics studies, and cellular assays [3-(4,5-Dimethylthiazol-2-Yl)-2,5-Diphenyltetrazolium Bromide (MTT) and flow cytometry] to reveal how structural changes of tau protein can change the cytotoxicity of titanium dioxide (TiO\textsubscript{2}) NPs against neuron-like cells (SH-SY5Y) cells.

\textbf{Results:} It was shown that TiO\textsubscript{2} NPs result in hydrophilic interactions, secondary and tertiary structural changes, and the formation of amorphous tau aggregates. Conformational changes of tau increased the induced cytotoxicity by TiO\textsubscript{2} NPs. These data revealed that the denatured adsorbed protein on the NP surface may enhance NP cytotoxicity.

\textbf{Conclusion:} Therefore, this study provides useful insights on the NP–protein interactions and discusses how the protein corona can increase cytotoxicity to determine the efficacy of targeted delivery of nanosystems.

\textbf{Keywords:} titanium oxide nanoparticle, tau, amorphous aggregation, cytotoxicity, neuron-like cells

\section*{Introduction}

Advancements in nanobiotechnology have resulted in the progress of a new area of nanomedicine, which incorporates the implementation of nanomaterials in diagnostic,\textsuperscript{1} imaging,\textsuperscript{2} and therapeutic settings.\textsuperscript{3} Although, nano-based agents are presently being extensively utilized in biological and medicinal approaches, there is a momentous concern regarding human\textsuperscript{4} and environmental risk\textsuperscript{5} upon implications of fabricated nanomaterials. Recent in vivo\textsuperscript{6} and in vitro\textsuperscript{6,7} investigations have suggested that utilization of some nanoparticles (NPs) may induce serious toxic health impacts and the application of nanomaterials-based medical agents may result in chronic or acute health problems.\textsuperscript{8} Thus, along with the development of unique NPs, simultaneous investigation of the impacts of NPs on the biological systems should be carried out. Once NPs interact with a biological system, they are exposed to corona formation around them. The biological responses of NPs coated with protein corona are totally different compared to bare NPs.\textsuperscript{9} The first investigation on the interactions between NPs and proteins was carried out in 1996.\textsuperscript{10} Studies are now based on NP–protein corona complexes as the working NPs in the biological fluids.\textsuperscript{7} During the interaction of
NPs and biomolecules, proteins are subjected to pronounced alterations in their structure and may change the fate of NPs.\(^9\)

Therefore, characterization of the proteins in the presence of NPs and cytotoxicity of NP–protein corona complexes have become crucial in experimental settings involving the application of NPs. Several studies have been recently devoted to extrapolate general rules to investigate the biological effect of the NP–protein corona on human cells.\(^11–13\)

Among different fabricated metal oxide NPs, titanium dioxide (TiO\(_2\)) NPs have been extensively utilized for biological and medical applications like cancer ablation\(^14–17\) and as antibacterial agents.\(^18\) However, the fundamental demands for the potential application of NPs in biomedical settings include their good colloidal stability, marginal structural changes of protein coronas, and low adverse effects.

Application of NPs results in their interaction with tissues components like the nervous system and inducing tau aggregation and neuronal death accompanied by neurodegenerative diseases. The two major neurodegenerative diseases affecting the world population include Alzheimer and Parkinson: the major tauopathies and alpha-synucleopathies, respectively. Alzheimer’s disease has been described in populations highly exposed to NPs. In children and young adults, NPs are found to be associated with major abnormalities in subcellular organelles.\(^19–21\)

Therefore, studies should be oriented toward investigating the interaction of TiO\(_2\) NPs with some biological systems such as the nervous system. Here, we tried to explore the interaction of TiO\(_2\) NPs with tau and the cytotoxicity of TiO\(_2\) NPs/tau complex against SH-SY5Y as models of nervous systems in vitro by biophysical, bioinformatic, and cellular assays.

**Materials and methods**

**Materials**

Tau-441 human recombinant, expressed in *Escherichia coli*, thioflavin T (ThT), Nile red, 8-anilinonaphthalene-1-sulfonic acid (ANS), DMEM and Ham’s F-12 nutrient mixture (DMEM-F12), FBS, penicillin, and streptomycin were purchased from Sigma-Aldrich Co (St Louis, MO, USA). The SH-SY5Y human neuroblastoma cell line was purchased from Pasteur Institute, Tehran, Iran. All other chemicals were of reagent grade.

**Sample preparation**

Tau solution was prepared in phosphate buffer, with pH 7.4 and ionic strength of 10 mM. Tau concentration was estimated spectrophotometrically at 280 nm, using an extinction coefficient of 7,450 cm\(^{-1}\) M\(^{-1}\). Stock solution of TiO\(_2\) NP was freshly prepared at 10 mg/mL, using ethanol (0.5%) as a solvent.

**Tryptophan fluorescence assay**

Aliquots of the tau solutions (0.1 µg/mL) were incubated without or with different concentrations of TiO\(_2\) NPs (0.5, 1, 5, 10, 20, 30, and 40 µg/mL) for 2 minutes. Fluorescence assays were then carried out by a Cary Eclipse VARIAN fluorescence spectrophotometer (Agilent, Mulgrave, VIC, Australia). Samples were excited at 270 nm, and the band intensity was determined between 275 and 400 nm, with 10-nm slit width for both excitation and emission wavelengths.

**ANS fluorescence assay**

The tertiary structural changes of tau were revealed by changes in the ANS fluorescence intensity of a mixture with 0.1 µg/mL tau solutions and 10 µM ANS in the absence and presence of varying concentrations of TiO\(_2\) NPs (0.5, 1, 5, 10, 20, 30, and 40 µg/mL) with the excitation wavelength fixed at 350 nm. Excitation and emission slit widths were both set to 5 nm. ANS fluorescence measurement was corrected against buffer and NP solutions.

**ThT fluorescence assay**

The formation of tau fibrils was detected by changes in the ThT fluorescence intensity of a mixture including 0.1 µg/mL tau solutions and 10 µM ThT in the presence of varying concentrations of TiO\(_2\) NPs (0.5, 1, 5, 10, 20, 30, and 40 µg/mL). The excitation and emission wavelengths were set to 440 and 485 nm, respectively. The other fluorescence spectroscopy parameters were similar with the ANS and tryptophan fluorescence assay sections.

**Ultraviolet (UV)-visible assay**

Tau solution (0.5 µg/mL) was incubated with various concentrations of TiO\(_2\) NPs (0.5, 1, 5, 10, 20, 30, and 40 µg/mL). After 2 minutes of incubation at room temperature, absorbance spectra were read at 360 nm. UV-visible assays were performed by a Cary Eclipse VARIAN spectrophotometer. In this assay, the NP absorbance was subtracted from the protein signal and the calculated absorbance was also corrected against dilution.

**Far-UV circular dichroism (CD) measurement**

CD spectra of tau samples (0.2 µg/mL) in the absence and presence of the highest studied concentration of TiO\(_2\) NPs (40 µg/mL) were recorded by an Aviv 215 spectropolarimeter (Aviv Biomedical Inc, Lakewood, NJ, USA). After 2 minutes of incubation, the spectra were recorded in the range of 190–260 nm. The NP ellipticity was subtracted from
the protein ellipticity and the calculated ellipticity was also corrected against dilution.

Transmission electron microscopy (TEM) analysis
Tau solution with a concentration of 2 μM was incubated with the highest studied concentration of TiO₂ NPs (40 μg/mL). After 2 minutes of incubation, the sample was prepared for TEM analysis. The morphology of the aggregated species was then determined using a TEM microscope (EM10C-100 KV; Carl Zeiss Meditec AG, Jena, Germany).

Computational study
(TiO₂)ₐ and (TiO₂)ₐ clusters were optimized as models of TiO₂ NPs. The Cartesian coordinate of atoms was obtained from the study of Auvinen et al. HEX 6.3 software (http://hex.loria.fr/dist63/) was used to run the molecular docking study. The molecular dynamics simulations were performed using the Forcite code and universal force field.

Viability assay
SH-SY5Y cells were cultured in DMEM-F12 supplemented with 12% (V/V) FBS, 100 U/mL penicillin, and 100 mg/mL streptomycin and incubated at 37°C with 95% humidified air and 5% CO₂. The cells (1×10⁴ per well) were then cultured in 96-well plates, and the TiO₂ NP-induced cell viability was calculated by the MTT assay. Briefly, after 24-hour exposure of different doses (0.01, 0.1, 1, 10, 50, and 100 μg/mL) of TiO₂ NPs and a single dose of TiO₂ NP/tau complex sample (100 μg/mL), 40 μL of MTT (2 mg/mL in PBS) was added to each sample for 4 hours. The supernatants were then gently removed and 100 μL of dimethyl sulfoxide was added to each well. The absorbance was then read at 570 nm using a microplate reader (model 3550; Bio-Rad Laboratories Inc, Hercules, CA, USA).

Quantification of apoptosis
Cells with a density of 3×10⁴ cells per well were seeded into 6-well plates. Following the incubation of cells with 100 μg/mL of TiO₂ NPs and 100 μg/mL of TiO₂ NPs/tau complex for 24 hours, the cells were washed, centrifuged (67×g for 5 minutes), washed again, resuspended in 5 μL propidium iodide and 5 μL Annexin V, and incubated for 30 minutes in the dark. The cells were then assayed by flow cytometry (BD FACSCalibur™; BD Biosciences, San Jose, CA, USA).

Statistical analyses
All assays were done in triplicate. Data are presented as mean ± SE of three independent experiments. Statistical significance was estimated by one-way ANOVA followed by Dunnett’s post hoc test. Values of *P<0.05 were reported to be significant.

Results
Characterization of NPs
TiO₂ NPs were synthesized by the sol-gel route and were fully characterized by dynamic light scattering, TEM, and X-ray diffraction techniques. It was observed that fabricated TiO₂ NPs have a narrow nano-sized diameter of around 30 nm and good colloidal stability. The X-ray diffraction pattern of the fabricated NPs also confirmed that the synthesized TiO₂ NPs are in the anatase phase.

Tryptophan fluorescence assay
Intrinsic tau fluorescence originated from tyrosine residues, which can be selectively excited at 270 nm. Changes in the emission signal of tyrosine residues are due to the protein structural changes induced by NPs influencing local micro-environment changes surrounding the indole ring. In this assay, the intrinsic fluorescence of tyrosine was read following excitation at 270 nm in the presence of increasing concentration of TiO₂ NPs by a spectrofluorometer. Figure 1 shows the TiO₂ NPs-induced fluorescence quenching of tyrosine residues after incubation with tau for 2 minutes. The effect of reducing the intensity of tyrosine emission spectra in the presence of increasing doses of TiO₂ NPs can be deduced by the interaction of NPs/drugs with the native proteins.

Stern–Volmer equation was used to calculate the Stern–Volmer constant (Ksv) in the presence (F) and absence (F₀) of NPs based on the following equation:

$$F₀/F = K_{SV} [NP] + 1$$

Figure 1 Intrinsic fluorescence quenching of tau in the absence and presence of increasing doses of TiO₂ NPs at room temperature.

Abbreviations: NPs, nanoparticles; TiO₂, titanium dioxide.
Conformational changes of proteins in the presence of ligands like NPs can result in protein aggregation. To monitor the morphology of the aggregated species of tau in the presence of TiO$_2$ NPs, ThT fluorescence assay was carried out. Figure 4 shows that by adding varying concentrations of TiO$_2$ NPs, the ThT fluorescence intensities did not significantly change compared to the negative control sample. The ThT fluorescence intensities of tau samples were $19 \pm 0.63$, $22 \pm 0.73$, $21 \pm 0.70$, $23 \pm 0.76$, $22 \pm 0.86$, $24 \pm 1.14$, $23 \pm 1.12$, and $25 \pm 1.54$ in the presence of $0$, $0.5$, $1$, $5$, $10$, $20$, $30$, and $40 \mu$g/mL of TiO$_2$ NPs, respectively. Therefore, this assay confirms that incubation of tau protein with TiO$_2$ NPs for 2 minutes does not induce amyloid formation of tau proteins.

**UV-visible assay**

UV spectroscopy at 360 nm is widely used to detect the induction of protein aggregation by ligands. Figure 5 depicts that the absorbance of tau samples increases at 360 nm as the concentrations of NPs rise. The absorbance intensities of the tau sample were $0.09 \pm 0.01$, $0.11 \pm 0.01$, $0.15 \pm 0.02$, $0.19 \pm 0.03$, $0.27 \pm 0.03$, $0.33 \pm 0.03$, $0.42 \pm 0.05$, and $0.47 \pm 0.06$ in the presence of $0$, $0.5$, $1$, $5$, $10$, $20$, $30$, and $40 \mu$g/mL of TiO$_2$ NPs, respectively. Therefore, these data indicate the induction of tau aggregate formation by TiO$_2$ NPs after 2 minutes of incubation.

**Circular dichroism assay**

We speculated that TiO$_2$ NPs might drive early structural changes in tau to initiate subsequent aggregation. To study this hypothesis, we carried out CD spectroscopy. Figure 6 shows the CD spectra before and after incubation with TiO$_2$ NPs for 2 minutes. It was observed that the tau structure was largely unstructured in the presence of TiO$_2$ NPs (40 $\mu$g/mL) and did not adopt a random coil structure similar to the negative control tau. Therefore, it may be concluded that the natively disordered structure of tau shifted toward a more folded conformation in the presence of TiO$_2$ NPs.

**TEM observation**

To visualize amorphous tau aggregate species, we carried out TEM on TiO$_2$ NPs samples incubated without (Figure 7A) or...
with (Figure 7B) tau. We observed that tau forms amorphous aggregates (Figure 7B) when incubated with TiO$_2$ NPs. It can be concluded that incubation of tau with NPs for 2 minutes results in the formation of amorphous tau aggregates. These amorphous aggregates then should be examined to explore if they act as a nucleus to initiate the formation and elongation of amyloid aggregates over time.

Molecular docking study

Molecular docking is considered to be a leading tool in biophysical and pharmaceutical sciences.$^{31,32}$ The aim of NP–protein docking is to complete the experimental data and estimate the predominant binding site(s) of an NP to a protein. Molecular docking was performed with two clusters of TiO$_2$ NPs with different sizes (Figure 8). The crystallographic fragments of human tau protein (PDB IDs: 2MZ7, 4E0N, 5O3T) were downloaded from the online Protein Data Bank, RCSB PDB (http://www.pdb.org). Visualization of the docked sites was carried out by Chimera (www.cgl.ucsf.edu/chimera) and PyMOL (http://pymol.sourceforge.net/) tools. The docked complexes of (TiO$_2$)$_{22}$ cluster-tau segments and (TiO$_2$)$_{38}$-tau segments are shown in Figures 9 and 10, respectively. The TiO$_2$ cluster with the closest residues (4 Å) is demonstrated in Figure 11. The resulting binding energies and the interacting residues for (TiO$_2$)$_{22}$ cluster-tau segments and (TiO$_2$)$_{38}$-tau segments are summarized in Tables 2 and 3, respectively.

Data clearly indicated that the applications of different clusters with different sizes resulted in changing the binding site location on tau segments. Data also showed that the (TiO$_2$)$_{38}$ cluster provides more affinity to interact with tau segments than the (TiO$_2$)$_{22}$ cluster. The affinity of the larger cluster to 5O3T segments (aggregated segment) was highest among other segments (with the lowest binding energy of $-451.53$).

The important interactive residues of 5O3T segment with (TiO$_2$)$_{38}$ cluster were Arg-349.J, Arg-349.H, Gln-351.J, Gln-351.F, Lys-369.D, Ile-371.H, Ile-371.F, Ile-371.D, Ile-371.B, Thr-373.B, Thr-373.H, Thr-373.F, and Thr-373.D. It can be suggested that hydrophilic residues on the 5O3T segment are predominant active sites interacting with the (TiO$_2$)$_{38}$ cluster.

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**Figure 3** ANS fluorescence of tau in the absence and presence of increasing doses of TiO$_2$ NPs at room temperature.

**Abbreviations:** ANS, 8-anilinonaphthalene-1-sulfonic acid; NPs, nanoparticles; TiO$_2$, titanium dioxide.

**Figure 4** ThT fluorescence intensity of tau in the absence and presence of increasing doses of TiO$_2$ NPs at room temperature.

**Abbreviations:** NPs, nanoparticles; ThT, thioflavin T; TiO$_2$, titanium dioxide.

**Figure 5** The absorbance intensity of tau at 360 nm in the absence and presence of increasing doses of TiO$_2$ NPs at room temperature.

**Note:** *P* < 0.05, **P** < 0.01, and ***P** < 0.001 vs control group.

**Abbreviations:** NPs, nanoparticles; TiO$_2$, titanium dioxide.

**Figure 6** CD signals of tau in the absence and presence of TiO$_2$ NPs at room temperature.

**Abbreviations:** NPs, nanoparticles; TiO$_2$, titanium dioxide.
Molecular simulation

For molecular dynamics simulations, the NP cluster and 2MZ7 segment were surrounded by 500 water molecules. Also, the annealing process was set to the equilibrium temperature of 298 K. The NVE ensemble with a time step of 1 fs, and a simulation time of 300 ps, was run. The structural changes of the 2MZ7 segment in the presence (Figure 12A) and absence (Figure 12B) of the TiO$_2$ NP cluster after 300 ps evolution are shown in Figure 12. As shown in Figure 12, the adsorbed protein tends to shift from a disordered structure toward a more folded structure.

MTT assay

MTT assay was performed to explore the TiO$_2$ NP- and TiO$_2$ NP/tau complex-induced cytotoxicity against SH-SY5Y cells. Figure 13 shows that after addition of different concentrations of TiO$_2$ NP, the cell viability reduced in a dose-dependent manner. In the next phase, cells were incubated with TiO$_2$ NP (100 µg/mL)/tau (100 µg/mL) complex for 24 hours to assess the effect of amorphous tau aggregates on the TiO$_2$ NPs-induced cytotoxicity. As shown in Figure 13, a significant difference was observed between TiO$_2$ NPs and TiO$_2$ NPs/tau samples in the induced cytotoxicity against SH-SY5Y cells ($P<0.05$). It can be suggested that incubation of TiO$_2$ NPs with tau and formation of amorphous tau aggregates on the NP surface may increase the induced cytotoxicity by TiO$_2$ NPs.
Quantification of apoptosis
Flow cytometry analysis was also employed to quantify the induction of apoptosis by TiO$_2$ NPs and TiO$_2$ NPs/tau samples. Figure 14 shows that 94% of the control cells are in the viable cells quadrant (Q1) (Figure 14A). However, incubation of cells with 100 µg/mL of TiO$_2$ NPs (Figure 14B) and 100 µg/mL of TiO$_2$ NPs/tau (Figure 14C) samples resulted in 55.66% (**P<0.01) and 95.42% (**P<0.01) apoptosis.

Table 2 Docking results of (TiO$_2$)$_{22}$ with tau segments

| NP      | PDB ID of receptor | Docking score (E-value) | Residue interacted                                                                 |
|---------|---------------------|-------------------------|-----------------------------------------------------------------------------------|
| (TiO$_2$)$_{22}$ | 2MZ7               | –333.52                 | Cys-291, Lys-290, Ser-289, Gln-286, Gln-269, His-268                              |
| (TiO$_2$)$_{22}$ | 4E0N               | –364.70                 | Ser-1, Gln-3, Lys-13, Val-5                                                       |
| (TiO$_2$)$_{22}$ | 5O3T               | –335.66                 | Gln-307.J, Gln-307.H, Gln-307.F, Val-309.J, Val-309.H, Val-309.F, Lys-311.J, Lys-311.H, Lys-311.F, Lys-311.D, Asn-327.G, Gly-326.G, Asn-327.E, Gly-326.E, Asn-327.C, Asn-327.A |

Abbreviations: NP, nanoparticle; PDB, Protein Data Bank; TiO$_2$, titanium dioxide.
Table 3 Docking results of (TiO$_2$)$_38$ with tau segments

| NP       | PDB ID of receptor | Docking score (E-value) | Residue interacted                                                                 |
|----------|--------------------|-------------------------|------------------------------------------------------------------------------------|
| (TiO$_2$)$_38$ | 2MZ7               | -377.22                 | Pro-270, Gln-269, His-268, Gly-271, Gly-272, Asp-283, Ser-285, Asn-286, Gln-288, Ser-289, Lys-290, Cys-291 |
| (TiO$_2$)$_38$ | 4E0N               | -434.02                 | Gln-3, Val-5, Lys-13, Lys-7                                                        |
| (TiO$_2$)$_38$ | 5O3T               | -451.53                 | Arg-349.J, Arg-349.H, Gln-351.J, Gln-351.F, Lys-369.D, Ile-371.H, Ile-371.L, Ile-371.D, Ile-371.B, Thr-373.B, Thr-373.H, Thr-373.F, Thr-373.D |

Abbreviations: NP, nanoparticle; PDB, Protein Data Bank; TiO$_2$, titanium dioxide.

induction in SH-SY5Y cells. It was also observed that the induction of apoptosis was more pronounced ($^{a*}P<0.01$) for TiO$_2$ NPs/tau than TiO$_2$ NPs sample (Figure 14D). Therefore, it may be concluded that the TiO$_2$ NPs induced the formation of amorphous tau aggregates and these species can enhance the derived cytotoxicity of TiO$_2$ NPs.

Discussion

Neurodegenerative diseases are characterized by tau aggregation.$^{33}$ While several studies have focused on explaining the mechanisms of tau polymerization and a large body of data is available with information regarding the toxic effect of tau oligomers in the pathology of neurodegenerative diseases,$^{34,35}$ studies exploring the aggregation of tau in the presence of NPs and induced cytotoxicity of NP/protein complex have been less numerous. Therefore, the aim of this study was to explore more detailed data regarding the structural changes of tau in the presence of TiO$_2$ NPs and the subsequent cytotoxicity of the TiO$_2$ NP/tau complex.

Intrinsic fluorescence spectroscopy revealed that TiO$_2$ NPs quench the tyrosine fluorescence signal of protein and may form a complex with the protein. ANS fluorescence spectroscopy and CD spectroscopy investigations showed that the tertiary and secondary structure of tau was changed, respectively, in the presence of TiO$_2$ NPs toward a more packed structure. UV-visible spectroscopy data indicated that increasing doses of TiO$_2$ NPs accelerate the formation of amorphous tau aggregates. We found that TiO$_2$ NPs strongly increase the formation of amorphous tau aggregates only after 2 minutes of incubation, suggesting that the NPs may induce an aggregation nucleus that appears early in the amyloid formation pathway. ThT fluorescence was almost constant after addition of increasing concentrations of TiO$_2$ NPs which could be a reliable marker for the formation of amorphous aggregated structures.$^{29}$ In the presence of TiO$_2$ NPs, tau aggregation was oriented toward a distinct aggregate species that shows no affinity for ThT.

It has been also reported that copolymer NPs, cerium oxide NPs, quantum dots, and carbon nanotubes may increase the probability of induction of an early nucleus for protein aggregation.$^{36}$ The nucleation phase is strongly modulated by the physicochemical properties of NPs. NPs may change the structure of proteins and extend the formation of aggregated species. It was also reported that Aβ fibril formation increases in the presence of TiO$_2$ NPs.$^{37}$ NPs due to their unique properties could shorten the lag phase, which is the

![Figure 12](image-url)  
**Figure 12** The molecular dynamic study of 2MZ7 segment in the presence (A) and absence (B) of TiO$_2$ cluster after 300 ps evolution.  
**Abbreviation:** TiO$_2$, titanium dioxide.

![Figure 13](image-url)  
**Figure 13** Effect of tau (100 µg/mL), TiO$_2$ NPs (0.01–100 µg/mL), and TiO$_2$ NPs/tau complex (100 µg/mL/100 µg/mL) on viability of SH-SY5Y cells.  
**Notes:** Cell viability was studied using the MTT assay. Data are shown as mean ± SE of three independent experiments. $^{*}P<0.05$ and $^{**}P<0.01$ vs negative control group (0 µg/mL of TiO$_2$ NP). $^{*}P<0.05$ vs TiO$_2$ NP (100 µg/mL of TiO$_2$ NP) group.  
**Abbreviations:** NPs, nanoparticles; SE, standard error of mean; TiO$_2$, titanium dioxide.
Figure 14 Probing the apoptosis induction of TiO$_2$ NPs (0 µg/ml) (A), TiO$_2$ NPs (100 µg/ml) (B), and TiO$_2$ NPs/tau complex (100 µg/mL/100 µg/mL) (C) in SH-SY5Y cells by flow cytometry assay; the column graph represents the statistical analysis of apoptosis induction in SH-SY5Y cells by TiO$_2$ NPs (D).

Notes: Data are shown as mean ± SE of three independent experiments. *P<0.01 and **P<0.001 vs negative control group (0 µg/mL of TiO$_2$ NP). ##P<0.01 vs TiO$_2$ NP (100 µg/mL of TiO$_2$ NP) group.

Abbreviations: NPs, nanoparticles; PI, propidium iodide; SE, standard error of mean; TiO$_2$, titanium dioxide.

key path of aggregation. However, some conflicting results regarding NPs and aggregation have been reported. NPs have been investigated in several studies as a drug candidate that reduces the aggregation of proteins involved in neurodegenerative disorders. For example, an NP–chelator conjugate was developed as a therapeutic approach for Alzheimer disease to reduce the formation of fibril and neurotoxicity. Liao et al also reported that negatively charged gold NPs reduce the induction of amyloid β fibrillation and decrease neurotoxicity. Mahmoudi et al showed that physiochemical features of superparamagnetic iron oxide NPs can change the amyloid β fibrillation in solution. Concentration, surface groups, and diameter of NPs can provide a “dual” effect on protein fibrillation. Therefore, the impact of NP on protein aggregation or disaggregation should be well investigated.

Cellular assays revealed that the TiO$_2$ NP/tau sample is more toxic than free TiO$_2$ NPs. As the NP–protein interaction results in denaturation of proteins, it is crucial to explore the NP–protein complex interaction with cells. Here, we showed that the TiO$_2$ NPs folded the tertiary structure of tau which in turn increases the cytotoxicity of TiO$_2$ NPs.

The formed hydrophobic patches of tau in the presence of TiO$_2$ NPs can result in increased cytotoxicity of the TiO$_2$ NPs/tau complex compared to pure TiO$_2$ NPs. Rapid corona formation and the structural changes of proteins are revealed to influence cell death at a fixed exposure time.

TiO$_2$ NPs and gold NPs were demonstrated to trigger structural changes of tubulin and serum albumin, respectively. It was hypothesized that the mechanism of cytotoxicity is the increased immunogenicity induced by exposing protein hydrophobic cores on their surface in a denatured conformation. For example, it was shown that the presence of some moieties on the surface of gold NPs caused unfolding of fibrinogen that, in turn, resulted in an interaction with the leukocyte receptor, thereby inducing inflammation. Disaggregation of proteins may result in mitigating the aggregated species of protein-induced cytotoxicity. For example, Meng et al revealed that polyphenols inhibit
amyloid formation, disaggregate protein aggregation, and protect cells against amyloid polypeptide-induced cytotoxicity.45 Sheynis et al also reported that aggregation modulators can interfere with cellular interactions of aggregated species of proteins.46 Also, it was suggested that the surface charge,47 NP distribution,48 adsorbed proteins,49 and physicochemical properties of NPs50 may affect the NP-induced cytotoxicity. Protein adsorption on the NP surface may influence all these features and result in increasing or decreasing the cytotoxicity of NPs.

Conclusion
Biomacromolecules like proteins can be denatured in the presence of NPs and subsequently may change the fate of NPs in biological systems. Since, protein corona modifies the NP surface, protein structural changes thus affecting the adverse effects of NPs. In this study, we showed that TiO$_2$ led to the formation of hydrophilic interactions with TiO$_2$ NPs and altered the secondary and tertiary structure of the tau protein. These structural changes finally resulted in the formation of amorphous tau aggregates. Afterwards, it was shown that the conformational changes of tau enhance the derived cytotoxicity from TiO$_2$ NPs. These data determined the adverse effects of NPs on protein structure and the role of the protein corona in NP-induced cytotoxicity. Therefore, this study can be used in the development of new approaches to modulate adsorbed proteins to more efficiently apply NPs for medical settings.

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Disclosure
The authors report no conflicts of interest in this work.

References
1. Tian X, Zhang L, Yang M, et al. Functional magnetic hybrid nanomaterials for biomedical diagnosis and treatment. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2018;10(1):e1476.
2. Smith BR, Gambhir SS. Nanomaterials for in vivo imaging. Chem Rev. 2017;117(3):901–986.
3. Mokhtarzadeh A, Alibakhshi A, Hashemi M, et al. Biodegradable nano-polymer as delivery vehicles for therapeutic small non-coding ribonucleic acids. J Control Release. 2017;245:116–126.
4. Nasrollahi SA, Hassanzade H, Moradi A, et al. Safety assessment of tretinoin loaded nano emulsion and nanostructured lipid carriers: a non-invasive trial on human volunteers. Curr Drug Deliv. 2017;14(4):575–580.
5. Hjorth R, Couturis C, Nguyen NHA, et al. Ecotoxicity testing and environmental risk assessment of iron nanomaterials for sub-surface remediation – Recommendations from the FP7 project NanoRem. Chemosphere. 2017;182:525–531.
6. Ma Z, Garrido-Maestu A, Lee C, et al. Comprehensive in vitro and in vivo risk assessments of chitosan microparticles using human epithelial cells and Caenorhabditis elegans. J Hazard Mater. 2018;341:248–256.
7. Laloux L, Polet M, Schneider YJ. Interaction between ingested-engineered nanomaterials and the gastrointestinal tract: in vitro toxicity aspects. Nanotechnology in Agriculture and Food Science. Weinheim: Wiley-VCH; 2017.
8. Kornberg T, Stueckle T, Antonini J, et al. Potential toxicity and underlying mechanisms associated with pulmonary exposure to iron oxide nanoparticles: conflicting literature and unclear risk. Nanomaterials. 2017;7(10):307.
9. Corbo C, Molinaro R, Parodi A, Toledano Furman NE, Salvatore F, Tasciotti E. The impact of nanoparticle corona on cytotoxicity, immunotoxicity and target drug delivery. Nanomedicine. 2016;11(1):81–100.
10. Diederichs JE. Plasma protein adsorption patterns on liposomes: establishment of analytical procedure. Electrophoresis. 1996;17(3):607–611.
11. Monteiro-Riviere NA, Inman AO. Challenges for assessing carbon nanomaterial toxicity to the skin. Carbon. 2006;44(6):1070–1078.
12. Balbus JM, Maynard AD, Colvin VL, et al. Meeting report: hazard assessment for nanoparticles – report from an interdisciplinary workshop. Environ Health Perspect. 2007;115(11):1654–1659.
13. Monteiro-Riviere NA, Inman AO, Zhang LW. Limitations and relative utility of screening assays to assess engineered nanoparticle toxicity in a human cell line. Toxicol Appl Pharmacol. 2009;234(2):222–235.
14. Srivastava RK, Rahman Q, Kashyap MP, et al. Nano-titanium dioxide induces genotoxicity and apoptosis in human lung cancer cell line, A549. Hum Exp Toxicol. 2013;32(2):153–166.
15. Lagopati N, Kistsiov PV, Kontos AI, et al. Photo-induced treatment of breast epithelial cancer cells using nanostructured titanium dioxide solution. J Photochem Photobiol A Chem. 2010;214(2–3):215–223.
16. Wu X, Jiang H, Zhou Y, et al. Selective determination of drug resistant cancer cells on indium tin oxide electrode modified with nano titanium dioxide. Electrochem Commun. 2010;12(7):962–965.
17. Yin ZF, Wu L, Yang HG, Su YH, Fei Yin Z, Gui Yang H. Recent progress in biomedical applications of titanium dioxide. Phys Chem Chem Phys. 2013;15(14):4844–4858.
18. Kaviyarasu K, Geetha N, Kanimozhi K, et al. In vitro cytotoxicity effect and antibacterial performance of human lung epithelial cells A549 activity of Zinc oxide doped TiO$_2$ nanocrystals: Investigation of bio-medical application by chemical method. Mater Sci Eng C Mater Biol Appl. 2017;74:325–333.
19. Calderón-Garcidueñas L, Reynoso-Robles R, Pérez-Guillé B, Mukherjee PS, González-Maciel A. Combustion-derived nanoparticles, the neuroenteric system, cerebral vagus, hyperphosphorylated alpha synuclein and tau in young Mexico City residents. Environ Res. 2017;159:186–201.
20. González-Maciel A, Reynoso-Robles R, Torres-Jardon R, Mukherjee PS, Calderón-Garcidueñas L. Combustion-Derived nanoparticles in key brain target cells and organelles in young Urbanites: culprit hidden in plain sight in Alzheimer’s disease development. J Alzheimers Dis. 2017;59(1):189–208.
21. Maher BA, Ahmed IAM, Karloukovski V, et al. Magnetite pollution nanoparticles in the human brain. Proc Natl Acad Sci U S A. 2016;113(39):10797–10801.
22. Avvinen S, Lahti M, Alatalo M. Unoccupied titanium 3 d states due to subcluster formation in stoichiometric TiO$_2$ nanoparticles. Int J Quantum Chem. 2015;115(17):1175–1180.
23. Ritchie DW, Venkatraman V. Ultra-fast FFT protein docking on graph subcluster formation in stoichiometric TiO$_2$ nanoparticles. J Photochem Photobiol A Chem. 2015;115(17):1175–1180.
24. Rappé AK, Casewit CJ, Colwell KS, Goddard WA, Skiff WM. UFF, a full periodic table force field for molecular mechanics and molecular dynamics simulations. J Am Chem Soc. 1992;114(25):10024–10035.
25. Abdolmajid E, Kharazi H, Chalaki M, et al. Titanium oxide nanoparticles fabrication, hemoglobin interaction, white blood cells cytotoxicity, and antibacterial studies. J Biometall Struct Dyn. 2018:1–38.
26. Rahmani S, Moghazalizadeh L, Attar F, Rezayat SM, Mousavi SE, Falahati M. Probing the interaction of silver nanoparticles with tau protein and neuroblastoma cell line as nervous system models. J Biometall Struct Dyn. 2017;17(1):1–15.
27. Zeinabad HA, Zarrabian A, Saboury AA, Alizadeh AM, Falahati M. Interaction of single and multi wall carbon nanotubes with the biological systems: Tau protein and PC12 cells as targets. Sci Rep. 2016;6(1):26508.
28. Falahati M, Ma’mani L, Saboury AA, Shafiee A, Foroumadi A, Badiei AR. Aminopropl-functionalized cubic Ia3d mesoporous silica nanoparticle as an efficient support for immobilization of superoxide dismutase. Biochim Biophys Acta. 2011;1814(9):1195–1202.
29. Mahdavinmez M, Meratan AA, Ghobeh M, Ghasemi A, Saboury AA, Nemat-Gorgani M. Inhibition of HEWL fibril formation by taxifolin: mechanism of action. PLoS One. 2017;12(11):e0187841.
30. Wang SS, Wen WS. Examining the influence of ultraviolet C irradiation on recombinant human yD-crystallin. Mol Vis. 2010;16:2777–2790.
31. Shokohi-Pour Z, Chiniforoshan H, Sabzalian MR, Esmaeil SA, Monttazi-Borjeni AA. Cobalt (II) complex with novel unsymmetrical tetratadentate Schiff base (ON) ligand: in vitro cytotoxicity studies of complex, interaction with DNA/protein, molecular docking studies, and antibacterial activity. J Biomol Struct Dyn. 2018;36(2):532–549.
32. Hunter NH, Bakula BC, Bruce CD. Molecular dynamics simulations of apo and holo forms of fatty acid binding protein 5 and cellular retinoic acid binding protein II reveal highly mobile protein, retinoic acid ligand, and water molecules. J Biomol Struct Dyn. 2018;36(7):1893–1907.
33. Goedert M, Masuda-Suzukake M, Falcon B. Like prions: the propagate mechanism of action. Acta Biochim Biophys Sin. 2008;40(9):777–782.
34. Wang S, Xiao X, Yang Y. Tio2 NPs promote aggregation-engendered toxicity. Acta Biochim Biophys Sin. 2008;40(9):777–782.
35. Mohamoudi M, Quinlan-Pluck F, Monopoli MP, et al. Influence of the physicochemical properties of superparamagnetic iron oxide nanoparticles on amyloid β protein fibrillation in solution. ACS Chem Neurosci. 2013;4(3):475–485.
36. Darghali ZN, Riazzi GH, Ahmadian S, Ghafar M, Mahinpour R. Toxicity and interaction of titanium dioxide nanoparticles with microtubule protein. Acta Biochim Biophys Sin. 2008;40(9):777–782.
37. Wang S, Suri CR, Shekhawat G. Interaction of gold nanoparticles with protein: a spectroscopic study to monitor protein conformational changes. Appl Phys Lett. 2008;92(13):133104.
38. Sheynis T, Friediger A, Xue WF, et al. Aggregation modulators interfere with membrane interactions at the nano-bio interface. Nat Mater. 2009;8(7):543–557.
39. Deng ZJ, Liang M, Monteiro M, Toth I, Minchin RF. Nanoparticle-induced unfolding of fibrinogen promotes Mac-1 receptor activation and inflammation. Nat Nanotechnol. 2011;6(1):39–44.
40. Frenkel-Pinter M, Tal S, Scherzer-Attali R, et al. C-NQTrp alleviates uptake and cytotoxicity of carbon nanoparticles. Biomaterials. 2009;30(6):8127–8133.
41. Sheynis T, Friediger A, Xue WF, et al. Aggregation modulators interfere with membrane interactions of β2-microglobulin fibrils. Biophys J. 2013;105(3):745–755.
42. Fröhlich E. The role of surface charge in cellular uptake and cytotoxicity of medical nanoparticles. Int J Nanomedicine. 2012;7:5577.
43. Aggarwal P, Hall JB, Mcleland CB, Dobrovolskaia MA, Mcneil SE. Nanoparticle interaction with plasma proteins as it relates to particle biodistribution, biocompatibility and therapeutic efficacy. Adv Drug Deliv Rev. 2009;61(6):428–437.
44. Zhu Y, Li W, Li Q, et al. Effects of serum proteins on intracellular uptake and cytotoxicity of carbon nanoparticles. Carbon. 2009;47(5):1351–1358.
45. Rivera-Gil P, Jimenez de Aberasturi D, Wulf V, et al. The challenge to relate the physicochemical properties of colloid nanoparticles to their cytotoxicity. Acc Chem Res. 2013;46(3):743–749.