A DNA Vaccine against Yellow Fever Virus: Development and Evaluation

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Abstract

Attenuated yellow fever (YF) virus 17D/17DD vaccines are the only available protection from YF infection, which remains a significant source of morbidity and mortality in the tropical areas of the world. The attenuated YF virus vaccine, which is used worldwide, generates both long-lasting neutralizing antibodies and strong T-cell responses. However, on rare occasions, this vaccine has toxic side effects that can be fatal. This study presents the design of two non-viral DNA-based antigen formulations and the characterization of their expression and immunological properties. The two antigen formulations consist of DNA encoding the full-length envelope protein (p/YFE) or the full-length envelope protein fused to the lysosomal-associated membrane protein signal, LAMP-1 (pL/YFE), aimed at diverting antigen processing/presentation through the major histocompatibility complex II precursor compartments. The immune responses triggered by these formulations were evaluated in H2b and H2d backgrounds, corresponding to the C57Bl/6 and BALB/c mice strains, respectively. Both DNA constructs were able to induce very strong T-cell responses of similar magnitude against almost all epitopes that are also generated by the YF 17DD vaccine. The pL/YFE formulation performed best overall. In addition to the T-cell response, it was also able to stimulate high titers of anti-YF neutralizing antibodies comparable to the levels elicited by the 17DD vaccine. More importantly, the pL/YFE vaccine conferred 100% protection against the YF virus in intracerebrally challenged mice. These results indicate that pL/YFE DNA is an excellent vaccine candidate and should be considered for further developmental studies.
**Author Summary**

DNA and other nucleic acid vaccine technologies are advancing quickly, and new potent delivery methods are demonstrating great potential in human clinical trials. In this manuscript, we report a highly protective DNA vaccine against the yellow fever virus. This vaccine was engineered with a molecular adjuvant technology to enhance the exposure of the vaccine antigens to the immune system, resulting in augmented CD4+ helper responses. We postulate that the robust CD4+ responses help the B cells and the CD8+ cells mature more efficiently and produce better antibodies and cytotoxic cells, respectively. Our results show that vaccination with this yellow fever DNA formulation elicited protective levels of neutralizing antibodies and very strong cellular responses at similar levels to the responses elicited by the live attenuated 17DD vaccine. In addition, these results also suggest a very important role for cellular responses in mediating protection against yellow fever virus. The results reported here are very promising and further studies may lead to a new yellow fever vaccine for human use.

**Introduction**

The yellow fever (YF) virus is considered the prototype member of the family Flaviviridae, which includes several other viruses of medical importance, such as the dengue, Japanese encephalitis, tick-borne encephalitis and West Nile viruses [1]. According to the World Health Organization (WHO), more than 200,000 cases of YF infection, including 30,000 deaths, occur annually, with 90% of cases occurring in Africa [2]. The safest strategy for preventing YF infection is still vaccination because there is currently no drug that is effective against YF virus infection. In the last 70 years, more than 500 million people around the world have been vaccinated with the YF 17D/17DD virus-attenuated vaccines with a remarkable record of safety and efficacy [3]. Attenuated YF virus vaccines generate both long-lasting neutralizing antibodies and T-cell responses [4, 5]. However, despite several improvements in the manufacturing process and quality control, severe side-effects resulting from vaccination continue to be reported [6–9]. In some cases, vaccination was associated with increased severity of symptoms [10] and on rare occasions with fatal reactions [11, 12]. In view of this, the development of alternative vaccination strategies, such as DNA-based vaccines encoding specific virus sequences, has been considered [13–16].

The YF virus genome consists of a single-stranded, positive-sense RNA molecule of ~10.8 kb, flanked by a 5’ cap and a 3’ non-polyadenylated terminal loop structure. It expresses three genes for structural proteins (capsid—C, pre-membrane/membrane—pM/M, and envelope—E) and seven genes that code for non-structural (NS) proteins (NS1, NS2a, NS2b, NS3, NS4a, NS4b, and NS5). Coexpression of flavivirus M and E genes in mammalian cells has been demonstrated to produce virus-like particles (VLPs) containing pM/M and E proteins [17–19]. The E protein is known to be the principal virus surface protein and the main target for neutralizing antibodies. pM/M and E coexpression, as a vaccination strategy, has been described as a way of triggering neutralizing antibodies against the Japanese encephalitis [19–21], West Nile [22] and dengue viruses [17,18,23,24].

DNA vaccines express endogenous cytoplasmic antigens, which are mostly introduced to the immune system through the major histocompatibility complex (MHC) class I molecules that are mostly associated with cellular cytotoxic responses and often fail to elicit a satisfactory humoral response, which is essential for efficient virus neutralization. Activation of CD4+ helper cells is important for the development of CD8+ responses, immunological memory [25],
antibody maturation, class switching and expansion of antigen-specific B cells [26]. Several strategies have been proposed for enhancing MHC class II presentation of antigens encoded by DNA vaccines. The targeting of the MHC II compartment with other flavivirus E antigens has been shown to enhance neutralizing antibody production in immunized mice [17, 22, 24] and in non-human primates (Raviprakash personal communication at 2004 ASTMH meeting, http://www.astmh.org/meeting_archives.htm).

One of the main strategies for targeting the MHC II compartment with DNA-encoded antigens is based on the expression of the antigen fused to the lysosomal-associated membrane protein 1 (LAMP-1), a protein primarily found in the outer membrane of lysosomes [27]. The chimeric antigens expressed by DNA formulations in the context of type I trans-membrane LAMP are directed to compartments rich in MHC II, called the MHC II compartment (MIIC), which is where the peptide-MHC II complexes are formed [28, 29]. Other LAMP/antigen chimeric strategies, such as LAMP/HIV Gag [25, 26, 30, 31] and LAMP/dengue virus 2 pM/M-E [17, 24] antigens, have been shown to target the MIIC and were found to elicit enhanced immune responses compared with vaccines encoding unmodified native antigens.

This study investigated T-cell and humoral immune responses to the envelope of YF virus in C57Bl/6 and BALB/c mice immunized with DNA formulations expressing the full-length YF envelope protein, either as a wild-type or fused to LAMP. Responses in the mice were compared with the results obtained with standard immunization using the YF 17DD vaccine. We also evaluated the ability of DNA vaccines to provide protection against a lethal challenge. We show that although the YF 17DD vaccine produced higher neutralizing antibody titers, both DNA vaccine constructs encoding the entire E protein were also able to protect the mice against lethal challenge.

Materials and Methods

Yellow fever vaccine, cells and antibodies

The attenuated 17DD human yellow fever vaccine was obtained from Bio-Manguinhos, a unit of the Oswaldo Cruz Foundation (FIOCRUZ, Rio de Janeiro, Brazil). The vaccine was reconstituted in chilled PBS, kept in an ice bath, and used for mouse immunizations within 4 hours of reconstitution. VERO and 293 cells were obtained from the ATCC (Rockville, MD, USA) and were grown according to the supplier’s instructions in a DMEM medium (Invitrogen) containing 10% fetal bovine serum (Gibco), 1% penicillin/streptomycin (Gibco) and 1% L-glutamine (Sigma). YF virus strain 17DD was propagated in Vero cells at 37°C in 5% CO₂ to a titer of 10⁶ plaque-forming units (PFUs) per ml. The polyclonal anti-YF hyperimmune serum used in immunofluorescence assays was obtained from mice immunized with the YF 17DD virus-attenuated vaccine in our laboratory. Secondary antibodies were purchased either from Jackson Immunoresearch Laboratories (Bar Harbor, ME, USA) or Molecular Probes (Seattle, WA, USA).

Peptides

A set of 120 peptides of 15 amino acids each (15-mers), overlapping by 11 amino acids (15x11) and comprising the entire length of the envelope protein of the YF 17DD virus (NCBI GenBank accession number U17066), was synthesized using Schafé-N (Copenhagen, Denmark). The peptides were HPLC-purified to 80% purity or greater, with the exception of a few peptides that could not be purified and were used as crude extracts. The identity of each peptide was confirmed via mass spectrometry, and the amount of purified peptide was precisely measured. Stock solutions of all peptides were prepared via dilution in water when possible, or in a solution of 10 to 100% DMSO, to a final concentration of 20 mg/mL and were stored at −20°C.
the ELISPOT assays, the peptides were used at a 10 μg/mL final concentration. The highest DMSO concentration in the ELISPOT experiments was 0.05%.

DNA constructs

The full YF genome, used as template to design primers for p/YFE and pL/YFE amplification, is deposited in NCBI’s GenBank under accession number NC 002031. The Kyte-Doolittle hydrophathy plot analyzed this sequence to identify the capsid ER translocation signal and the predicted envelope trans-membrane domain of the YF genome. The wild-type pM/M-E amplicon starts with the ER capsid signal and ends with the envelope trans-membrane domain. To generate the pL/YFE construct, we designed a reverse primer that hybridizes to the YF genome just upstream of the envelope trans-membrane domain to replace it with the human membrane anchor and cytoplasmic domains of LAMP (Fig 1). The DNA pM/M-E sequence was amplified from the YF 17DD infectious clone using specific primers that incorporated an ATG start site in the context of the Kozak sequence and a translational stop codon. PCR amplification was

![Diagram of DNA constructs](Fig 1. p/YFE and pL/YFE DNA vaccine construct design. The fragment pM/M-E (extending from nucleotides 392 to 2452, black arrows) was amplified using PCR and cloned into a p43.2 vector to generate the p/YFE vector. This construct starts with the ER capsid signal and ends with the envelope trans-membrane domain. To generate the pL/YFE construct, we designed a second forward primer that annealed just upstream of the envelope trans-membrane domain to amplify the fragment extending from nucleotides 392 to 2323. This fragment was fused to the C-terminal end of LAMP and cloned into the same vector.

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performed using the proofreading TGO DNA polymerase (Roche, Indianapolis, IN, USA) and 0.6 μM of each primer. The amplicon was inserted into the p43.2 vector between the XhoI and NsiI cleavage sites to generate the p/YFE construct. The pL/YFE construct, however, was obtained in two steps. First, the pM/M-E sequence was amplified using a reverse primer that hybridized upstream of the trans-membrane domain of the YF envelope protein. Then, the PCR product was inserted into the p43.2 vector between the NheI and XhoI sites, generating an intermediate construct (p/YFEINT), ready to receive the membrane anchor and cytoplasmic domains of LAMP. Second, LAMP was amplified from the p43.2-Gag/LAMP vector and was inserted into p/YFEINT, between the XhoI and XbaI sites, to generate the pL/YFE construct. Both the p/YFE and pL/YFE constructs were checked by sequencing; among the 2,061 nucleotides of the pM/M-E wild-type construction (p/YFE) that encode 687 amino acids, two nonsynonymous mutations were found. An alanine (A) was replaced with a valine (V) at position 250, and a serine (S) was replaced with an aspartic acid (D) at position 349. Given that both mutations were also found at the same locations in the 644-residue sequence of the pL/YFE construction, they are very likely present in the YF 17DD infectious clone that was used as a DNA template. Regardless of the source of the two mutations, both mutations were deemed to be irrelevant for our vaccine studies as the E protein has several B-cell and T-cell preserved epitopes distributed along its sequence.

Transfections, western blotting and fluorescence assays

293 cells were plated onto cover slips and transfected with p/YFE, pL/YFE or empty p43.2 vectors, using Lipofectamine 2000 (Invitrogen Life Technologies). For Western blotting, transfections were carried out in 6-well tissue culture plates with 10 μg of each plasmid and 40 μl of Lipofectamine 2000, whereas transfections for fluorescence assays were carried out in 24-well tissue culture plates with 2.5 μg of each plasmid and 10 μl of Lipofectamine 2000, both in accordance with the manufacturer’s instructions. Vero cell extracts infected with the YF 17DD virus strain were used as a positive control. After 48 hours, both transfected and infected cell extracts were processed.

For Western blot analysis, cell extracts were resuspended in 2x Laemmli denaturing protein sample buffer, fractionated in 12.5% SDS-PAGE and transferred to a polyvinylidene difluoride (PVDF) membrane. After blocking with 5% milk/0.05% PBS-Tween 20, membranes were incubated for 1 hour with the appropriate primary polyclonal antibodies (anti-YFV hyperimmune rabbit serum, previously produced in our laboratory) diluted 1:500 in 1% milk/0.1% PBS-Tween 20. Membranes were washed 3 times with 1x PBS for 10 minutes/wash and incubated for 1 hour with 1:5,000 goat anti-rabbit IgG antibody conjugated with horseradish peroxidase (Jackson Immunoresearch Laboratories). The Western blot reactions were detected using enhanced chemiluminescence (ECL) reactions (Millipore). For fluorescence assays, cell extracts were fixed in 100% methanol at—20°C for 5 minutes, blocked with 1% BSA/PBS solution for 30 minutes, and incubated with an anti-YFV hyperimmune mouse antibody diluted 1:200 for 1 hour, followed by a 1-hour incubation with secondary antibody diluted 1:500 (Alexa 488-conjugated goat anti-mouse, Molecular Probes, Seattle, WA, USA). The cover slips were then mounted on glass slides using ProLong Gold (Molecular Probes, Seattle, WA, USA) and observed through a confocal microscope. The images were acquired using a Leica SP II-AOBS confocal microscope (Leica Microsystems, Hm) with a 63× oil immersion objective NA 1.3. The Alexa 488 fluorochrome was excited using an ArKr laser at 488 nm. The digital image was acquired using Leica software in a 24-bit RGB format with a 1024 × 1024 pixel area. Fields were chosen for imaging based on the spread and morphology of the cells.
Immunological assays

Female BALB/c (H2d) and C57Bl/6 mice (H2b), aged 6 to 8 weeks (Charles River, Kingston, NY, USA), were used for the ELISPOT assays. They were housed in micro-isolator cages under specific pathogen-free conditions and handled in accordance with the Johns Hopkins Institutional Animal Care and Use Committee (IACUC) protocol number MO05M336. The animals were immunized at days zero and 21 and used for the experiments seven to ten days after the last immunization. For the neutralization and protection assays, three-week-old female BALB/c and C57Bl/6 mice were obtained from the Oswaldo Cruz Foundation Breeding Center (Rio de Janeiro, Brazil) and were housed at the Experimental Animal Laboratory (Oswaldo Cruz Foundation, Rio de Janeiro, Brazil) under specific pathogen-free conditions and handled in accordance with the Oswaldo Cruz Foundation Commission for Ethical Animal Use (CEAU) protocol number P0112-02. The animals were immunized at days zero, 30 and 45, and sera were collected via a cut in the tail vein a day before every immunization. For both protocols, the animals were immunized subcutaneously at the base of the tail with either the YF 17DD vaccine at $10^4$ PFUs/50 μl, the DNA constructs at 50 μg/50 μl, or 50 μl of PBS as a negative control.

ELISPOT assays were performed to quantify IFN-gamma spot-forming cells (SFCs) generated via DNA construct immunization. Seven to 10 days after the last immunization, the mice were sacrificed and their spleens were removed. Splenocytes were isolated using standard methods, and single-cell suspensions, depleted of red blood cells, were prepared from freshly isolated splenocytes in culture medium (RPMI 1640 medium supplemented with 10% v/v fetal bovine serum, 100 units/ml penicillin/streptomycin, 2 mM L-glutamine, 50 μM 2-mercaptoethanol and 1 M HEPES buffer). IFN-gamma ELISPOT assays were performed in accordance with the manufacturer’s instructions (BD-Biosciences, San Diego, CA, USA). First, the ELISPOT plates were coated with anti-IFN-gamma antibody at 5 μg/ml and incubated at 4°C overnight. The plates were blocked with RPMI 1640 medium containing 10% FBS for 2 h at room temperature, and total splenocytes (1×10^6 cells/well) from immunized mice were then added. The cells were cultured at 37°C in 5% CO2 with culture medium alone (RPMI 1640 medium supplemented with 5% v/v fetal calf serum, 100 units/ml penicillin/streptomycin, and 2 mM L-glutamine) or with culture medium in the presence of concanavalin A (2.5 μg/ml; Sigma), $10^9$ PFUs/mL of inactivated YF virus as a positive control (strain 17DD), or individual 15-mers from the envelope protein of the YF 17DD virus at 1 μg/ml. After 16 h of culture, the plates were washed and incubated with biotinylated anti-IFN-gamma for 2 h at room temperature, followed by HRP-conjugated avidin for 1 h at room temperature. Reactions were developed with AEC substrate (Calbiochem-Novabiochem Corporation, San Diego, CA, USA). The quantification of spot-forming cells (SFCs) was carried out using the Immunospot Series Analyzer ELISPOT reader (Cellular Technologies Ltd (CTL), Shaker Heights, OH, USA) with the aid of Immunospot software 3.0 (Cellular Technologies Ltd). The data are represented as the number of SFCs/10^6. The results were considered positive if the number of SFCs was greater than 20 and higher than the background (culture with medium alone) plus three standard deviations. The results are presented after subtraction of the background.

Viral neutralization assays

Plaque reduction neutralization tests (PRNT) were carried out using VERO cells seeded at a density of 62,500 cells/cm² in 96-well microplates, as previously described [32]. The PRNT tests for the detection of anti-YF nAb were performed after two-fold serial dilutions of serum (1/5 to 1/640) on microtiter plates and incubation with 30 PFUs of the YF 17DD challenge virus strain in each well. After incubation at 37°C in a 5% CO2 atmosphere for 1 h, 50 μl of
Vero cell suspensions (4×10⁴/well) in medium 199 (Invitrogen) was added, and the plates were incubated at 37°C for 3 h. The medium was then discarded and the cells were overlaid with 100 μl of medium containing 3.5% carboxymethylcellulose. After 6–7 days of incubation at 37°C in 5% CO₂, cell monolayers were fixed with formalin and stained with crystal violet so plaques could be counted. Standard sera of known antibody content in terms of International Units (IU) were included in each set of tests. The log₁₀ dilution of the test and standard sera, which reduced the plaque numbers by 50% relative to the virus control, were determined via interpolation. The mean antibody content at the 50% end point of the standard was then calculated and added to the log₁₀ end point for each sample to give log₁₀ mIU/ml. Plaque neutralization titers were calculated as the highest dilution of antibody able to reduce 50% of the plaques from input virus. The lower limit of detection of the assay was 84.5 mIU/mL.

Protection assays and statistical analysis

Groups of three-week-old BALB/c and C57Bl/6 mice immunized three times with either the YF 17DD vaccine or the DNA constructs were inoculated intra-cerebrally with 30 μl of M199 medium containing 10⁵ PFUs of the YF 17DD virus 15 days after the last immunization. The animals were monitored for 21 days and deaths were recorded. Moribund animals were sacrificed by exposure to CO₂.

Comparisons between ELISPOT and neutralization assay results were made using an unpaired T-test. The mean survival times in each group of mice were compared using a one-way analysis of variance (ANOVA) and the Kruskal-Wallis non-parametric test. Statistical tests and graphs were performed and produced using GraphPad Prism version 4.0 (GraphPad Software, San Diego, CA, USA; www.graphpad.com).

Results

The p/YFE and pL/YFE DNA constructs express yellow fever proteins in 293 cells

293 cells were transfected with the p/YFE and pL/YFE expression vectors encoding the pM/M-E and pM/M-E/LAMP proteins, respectively. Validation of plasmid protein expression and cellular steady-state localization was carried out using Western blot and immunofluorescence analyses. Both the E and E/LAMP proteins were stained using polyclonal anti-YF hyperimmune serum. Western blotting detected specific bands for the E (p/YFE) and E/LAMP (pL/YFE) proteins, as well as the wild-type (YF virus) E protein (Fig 2). Immunofluorescence assays showed the characteristic reticular membrane distribution (associated with the typical cellular trafficking of the viral envelope protein) in p/YFE-transfected 293 cells expressing the wild-type E protein (Fig 3A). By contrast, the E/LAMP chimeric protein from pL/YFE-transfected cells showed the typical punctuated lysosomal-like distribution of endogenous LAMP (Fig 3B). The figure represents several independent assays where the expression and cellular steady-state localization of both the E and E/LAMP proteins were considered to be invariable.

Characterization of the cellular response triggered by 17DD vaccination in C57Bl/6 mice

It is known that some mouse strains with distinct genetic backgrounds, when exposed to the same antigens, can polarize towards T-helper-1 (Th1) or T-helper-2 (Th2) responses. Some strains are more prone to produce Th1 responses, whereas others are more prone to polarize towards Th2. BALB/c and C57Bl/6 mice strains have been known to produce this type of distinct T-helper responses in several models and, thus, we selected these two strains to investigate
how they would respond to our vaccines. An optimized YF 17DD vaccine immunization protocol and IFN-gamma ELISPOT assay conditions, which were previously described for BALB/c (H2d, Dd, Kd, Ld, I-Ad, and I-Ed) mice [33], were used to characterize the T-cell responses to peptides of the YF 17DD virus proteins in C57Bl/6 (H2b, Db, Kb and I-Ab) mice. The first round of experiments with total splenocytes led to the identification of 11 antigenic 15-mer peptides from the YF envelope protein. The subsequent experiments were performed using splenocytes depleted of CD4+ or CD8+ lymphocytes, which lead to the identification of epitopes presented by MHC class I or II, respectively. The depletion typically removed >95% of the targeted population, as assessed via flow cytometry. The CD4-depleted splenocytes, which correspond to the CD8+ lymphocyte response, reacted to seven peptides, whereas the CD8-depleted splenocytes, which correspond to the CD4+ lymphocyte response, were able to respond.

Fig 2. p/YFE and pL/YFE DNA vaccine expression analysis via Western blotting. 293 cells were transfected with the p/YFE or pL/YFE DNA construct and incubated for 48 hours prior to total protein sample preparation. The p43.2 empty vector was used as a negative control, and cells infected with the YF 17DD virus strain were used as a positive control. Cell extracts were transferred to the PVDF membrane and incubated with anti-YFV hyperimmune antibody, followed by incubation with a secondary antibody conjugated with horseradish peroxidase that was revealed using an enhanced chemiluminescence (ECL) reaction. Both the E- and E-LAMP-encoded antigens were successfully expressed and were each the appropriate molecular size.

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Fig 3. p/YFE and pL/YFE DNA vaccine intracellular steady-state localization analysis using immunofluorescence assays. 293 cells were transfected with the p/YFE or pL/YFE DNA construct and incubated for 48 hours prior to methanol fixation. The p43.2 empty vector was used as a negative control, and cells infected with the YF 17DD virus strain were used as a positive control. Fixed cells were incubated with anti-YFV hyperimmune antibody, followed by incubation with a secondary antibody conjugated with Alexa Fluor-488 dye for microscopic analyses. The characteristic reticular membrane distribution of the viral envelope protein was detected in the p/YFE-transfected cell expressing the wild-type E protein (A), whereas the E/LAMP chimeric protein expressed by the pL/YFE-transfected cells showed the typical punctuated lysosomal-like distribution of endogenous LAMP (B).

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to five 15-mer peptides (Table 1). The results of quantitative (IFN-gamma SFCs/10^6) epitope mapping for the YF envelope protein in 17 DD immunized C57Bl/6 mice are shown in Table 2. Splenocytes from naïve animals did not react to any of the peptides tested.

| YF envelope peptide position | Peptide sequence | Restriction |
|-----------------------------|------------------|-------------|
| E1-15                       | AHCIGITDRDFIEGV  | CD8         |
| E201-215                    | ESWIVDROWAODTL   | CD4         |
| E229-243                    | HHLVEFEPHAATIR   | CD4         |
| E233-247                    | EFEPPHAATIRVLAL  | CD4         |
| E345-359                    | NKGILTVNPIASTN   | CD4/CD8     |
| E349-363                    | LVTVNPIASTNDDEV  | CD4         |
| E353-367                    | NPIASTNDDEVLIEV  | CD8         |
| E465-479                    | GINTRNMTSMSMIL   | CD4         |
| E473-487                    | MSMSMMSVIMMFL    | CD4         |
| E477-491                    | MILVGVIMMFLSLGV  | CD8         |
| E481-493                    | GVIMMFLSLGVGA    | CD8         |

Table 1. Position and restriction of the YF envelope epitopes in YF-17DD-immunized C57Bl/6 (H2\(^b\)) mice.

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p/YFE and pL/YFE DNA immunizations in C57Bl/6 (H2\(^b\)) and BALB/c (H2\(^d\)) mice elicit T-cell responses of similar magnitude and against the same repertoire of epitopes induced by the YF 17DD vaccine

The T-cell responses of H2\(^b\) and H2\(^d\) mouse strains induced by immunization with the p/YFE and pL/YFE plasmids were evaluated, and the results were compared with the responses observed for the YF 17DD vaccine immunization. Immunization with the p/YFE or pL/YFE plasmids generated a vigorous T-cell response in C57Bl/6 mice. Both plasmids, in addition to bringing about a T-cell response pattern similar to that produced by the YF 17DD vaccine, were able to elicit a significantly higher number of IFN-gamma SFCs (>200 IFN-gamma SFCs/10^6 splenocytes) for many immunogenic peptides of the YF envelope protein (Table 3; p<0.05) than was immunization with the attenuated virus vaccine.

Interestingly, p/YFE was able to generate a considerable response to the E\(_{413-427}\) and E\(_{417-431}\) peptides, which were not present after YF 17DD immunization and were very scarce after pL/YFE immunization (Table 3). Remarkably, p/YFE also brought about a stronger response to peptide E\(_1-15\) than both the YF 17DD and pL/YFE vaccines.

In BALB/c immunized mice, both the p/YFE and pL/YFE DNA constructs generated an immune response very similar to that obtained with the YF 17DD vaccine, eliciting almost the same immunogenic determinants (Table 3). The only considerable exception was the lack of response in YF-17DD-immunized mice to peptide E\(_{329-343}\), which contains a previously characterized MHC class I epitope (CD8\(^+\) response) (Table 4). Both the p/YFE and pL/YFE plasmids also produced a significantly higher number of T cells specific to the immunodominant E\(_{57-71}\) and E\(_{61-75}\) peptides (p<0.05), which contain MHC class I and class II epitopes for the H2\(^d\) mouse strain. The number of IFN-gamma SFCs was similar for all remaining positive peptides. Groups of mice from both strains immunized with either the empty plasmid or a plasmid expressing only the LAMP protein did not react to any of the peptides tested.
Evaluation of the humoral responses generated by the YF 17DD and the DNA construct immunizations

The protection provided by the YF vaccine is mainly attributed to the neutralizing antibody (nAb) response generated after vaccination. Because the presence of nAb is a hallmark of protection, we evaluated the humoral response of C57Bl/6 and BALB/c mice after immunization with the DNA constructs and compared them with the levels of nAb obtained after immunization with the YF 17DD vaccine. To investigate the kinetics of nAb responses, the animals were immunized at days zero, 30 and 45 and bled 15 days after each immunization.

The YF 17DD vaccine was able to produce very high levels of nAb in C57Bl/6 and BALB/c mice after the first immunization (day 15). nAb levels in C57Bl/6 mice seemed to reach a plateau after the second immunization (day 45) and increased slightly after the third immunization (day 60), whereas BALB/c mice showed increasing levels of nAb after the second (day 45) and third (day 60) immunization with the YF 17DD vaccine. The levels of nAb observed in C57Bl/6 mice (\(9,664.0\) mIU/mL; obtained at the highest dilution tested) were approximately 20% higher compared with the levels observed in BALB/c mice (\(7,500\pm780.1\) mIU/mL) (Fig 4). In C57Bl/6 mice, both plasmids expressing the whole envelope protein (p/YFE and pL/YFE) were able to produce significant levels of nAb (p<0.0039 and p<0.002, respectively) after three immunizations compared with the empty vector control. The DNA plasmid pL/YFE, expressing the chimeric E-LAMP protein, led the BALB/c mice to produce higher titers of nAb after the second immunization compared to the p/YFE plasmid. The levels of nAb titers increased after the third immunization and were significantly higher (p<0.045) than those of the control groups immunized with empty vector or PBS (Fig 4).

On average, the pL/YFE DNA immunization elicited nAb titers 7-fold greater than the p/YFE DNA immunization. Compared with the 17DD attenuated virus vaccine, the nAb titers produced by the pL/YFE DNA vaccine were approximately 3.5-fold lower. The fact that these DNA vaccines produced these levels of nAb may still be considered significant.

Table 2. Characterization of the cellular responses produced by 17DD vaccination in C57Bl/6 mice.

| # of IFN-γ SFCs/10^6 splenocytes | T-cell responses of C57Bl/6 (H2b) mice | YF envelope 15-mer peptides |
|----------------------------------|---------------------------------------|-----------------------------|
|                                  | Total splenocytes  | CD8-depleted cells  | CD4-depleted cells  |
| <100                              | 1–15                                  | 201–215                     | 353–367                     |
|                                  | 201–215                               | 345–359                     | 477–491                     |
|                                  | 335–367                               | 349–363                     | 481–493                     |
|                                  | 345–359                               | 473–487                     |                             |
|                                  | 349–363                               |                             |                             |
|                                  | 473–487                               |                             |                             |
|                                  | 477–491                               |                             |                             |
|                                  | 481–493                               |                             |                             |
| 100–200                           | 229–243                               | 229–243                     | 1–15                        |
|                                  | 465–479                               | 465–479                     |                             |
| >200                              | 233–247                               | 233–247                     | 345–359                     |

C57Bl/6 mice were immunized on day 0 and boosted on day 21 with 10^4 PFUs of the human YF 17DD vaccine, and the splenocytes were tested in IFN-γ ELISPOT assays 7–10 days after the boost. The peptides used for the in vitro stimulation were 15-mers overlapping by 11 amino acids and comprising the entire length of the YF envelope protein. The total splenocyte, CD8-depleted splenocyte and CD4-depleted splenocyte populations were analyzed. The SFC values represents the average of two to four experiments performed with pools of three to five mice each.

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Evaluation of the humoral responses generated by the YF 17DD and the DNA construct immunizations

The protection provided by the YF vaccine is mainly attributed to the neutralizing antibody (nAb) response generated after vaccination. Because the presence of nAb is a hallmark of protection, we evaluated the humoral response of C57Bl/6 and BALB/c mice after immunization with the DNA constructs and compared them with the levels of nAb obtained after immunization with the YF 17DD vaccine. To investigate the kinetics of nAb responses, the animals were immunized at days zero, 30 and 45 and bled 15 days after each immunization.

The YF 17DD vaccine was able to produce very high levels of nAb in C57Bl/6 and BALB/c mice after the first immunization (day 15). nAb levels in C57Bl/6 mice seemed to reach a plateau after the second immunization (day 45) and increased slightly after the third immunization (day 60), whereas BALB/c mice showed increasing levels of nAb after the second (day 45) and third (day 60) immunization with the YF 17DD vaccine. The levels of nAb observed in C57Bl/6 mice (\(9,664.0\) mIU/mL; obtained at the highest dilution tested) were approximately 20% higher compared with the levels observed in BALB/c mice (\(7,500\pm780.1\) mIU/mL) (Fig 4). In C57Bl/6 mice, both plasmids expressing the whole envelope protein (p/YFE and pL/YFE) were able to produce significant levels of nAb (p<0.0039 and p<0.002, respectively) after three immunizations compared with the empty vector control. The DNA plasmid pL/YFE, expressing the chimeric E-LAMP protein, led the BALB/c mice to produce higher titers of nAb after the second immunization compared to the p/YFE plasmid. The levels of nAb titers increased after the third immunization and were significantly higher (p<0.045) than those of the control groups immunized with empty vector or PBS (Fig 4).

On average, the pL/YFE DNA immunization elicited nAb titers 7-fold greater than the p/YFE DNA immunization. Compared with the 17DD attenuated virus vaccine, the nAb titers produced by the pL/YFE DNA vaccine were approximately 3.5-fold lower. The fact that these DNA vaccines produced these levels of nAb may still be considered significant.
Protection evaluation via challenge

Intra-cerebral challenge with the YF 17DD virus in mice is a useful model for evaluating the protection provided by vaccine candidates [34]. We evaluated our DNA constructs using the immunization/challenge model by injecting $10^5$ PFUs of the YF 17DD virus intra-cerebrally into DNA-immunized C57Bl/6 and BALB/c mice. The animals were immunized three times at days 0, 30 and 45 and were challenged 15 days after the last immunization. As previously reported [35], immunization with YF 17DD vaccine was able to protect both C57Bl/6 and BALB/c mouse strains against the intra-cerebral challenge. In a similar fashion, immunization with both DNA constructs expressing the full-length YF envelope protein was able to fully protect both mouse strains from the lethal challenge. The majority of mice immunized with PBS or empty vector died 10 to 14 days after the challenge assay (Table 5 and Fig 5).

### Table 3. Comparison of the C57Bl/6 and BALB/c T-cell responses produced by immunization with p/YFE, pL/YFE and the YF 17DD vaccine.

|                | C57Bl/6 (H2b)                      | BALB/c (H2b)                      |
|----------------|-----------------------------------|-----------------------------------|
| # of IFN-γ SFCs/10⁶ splenocytes | YF envelope 15-mer peptides (10 μg/mL) | YF envelope 15-mer peptides (10 μg/mL) |
|                | YF 17DD vaccine | p/YFE | pL/YFE | YF 17DD vaccine | p/YFE | pL/YFE |
| <200           | 1–15              | 5–19     | 169–183 | <200           | 25–39              | 65–79     | 21–35    |
| 201–215        | 201–215           | 169–183  | 201–215 | 129–143         | 129–143           | 25–39     |
| 229–243        | 201–215           | 349–363  | 349–363 | 133–147         | 137–151           | 129–143   |
| 233–247        | 225–239           | 385–389  | 385–389 | 157–171         | 201–215           | 201–215   |
| 345–359        | 317–331           | 413–427  | 317–331 | 201–215         | 213–227           | 213–227   |
| 349–363        | 345–359           | 417–431  | 345–359 | 213–227         | 233–247           | 233–247   |
| 465–479        | 349–363           | 473–487  | 473–487 | 221–235         | 237–251           | 237–251   |
| 473–487        | 353–367           | 477–491  | 385–389 | 233–247         | 425–439           | 437–451   |
| 477–491        | 385–389           | 237–251  | 477–491 | 329–343         | 461–475           | 477–491   |
| 200–400        | 229–243           | 1–15     | 200–400 | 61–75           | 133–147           | 61–75     |
| 200–400        | 417–431           | 229–243  | 200–400 | 133–147         | 133–147           | 61–75     |
| 473–487        | 233–247           | 233–247  | 473–487 | 465–479         | 465–479           | 477–491   |
|                | 465–479           | 477–491  | 477–491 | 465–479         | 465–479           | 477–491   |
| 401–600        | 233–247           | 401–600  | 57–71   | 57–71           | 57–71             | 61–75     |
| 413–427        | 57–71             | 57–71    | 61–75   | 57–71           | 57–71             | 57–71     |
| >600           | 1–15              | >600     | >600    | 1–15            | >600              | >600      |

C57Bl/6 and BALB/c mice were immunized on day 0 and boosted on day 21 with $10^4$ PFUs of the human YF 17DD vaccine or with 50 μg of either p/YFE or pL/YFE. Total splenocytes were harvested 7–10 days after the last immunization and assayed in vitro using an IFN-γ ELISPOT assay with 15-mers overlapping by 11 amino acids and comprising the entire length of the YF envelope protein. The SFC values represent the average of two to four experiments performed with pools of three to five mice each (p<0.05).

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Discussion

YF infection continues to be a worldwide problem, especially in tropical areas [11], but this may change as the world continues to be affected by climate change. Despite the high efficiency of commercially available YF vaccines, 17D and 17DD, there are a few reports of rare but fatal side-effects after vaccination [9, 11, 12]. Furthermore, these vaccines are not recommended for infants, pregnant women, immunodeficient subjects, or those allergic to the egg components present in the vaccine formulations [36]. In light of these factors, there is reason to pursue complementary or alternative YF vaccine strategies that could replace the use of the virus-attenuated vaccine version.

Although no DNA vaccines have yet been approved for human use, they represent potential candidates to replace live/attenuated vaccine formulations because they are considered safer. DNA formulations can be easily manipulated, do not require a cold-chain for distribution and eliminate the infectious nature of live/attenuated agents. They also allow the manipulation of immunogens to provide the immune system with the desired epitopes and signals while avoiding the use of unnecessary or potentially harmful antigens or epitopes [37, 38].

Previous studies have described the development of DNA vaccines against flaviviruses based on the expression of the pM/M-E virus sequence cloned in-frame with the LAMP sequence [17, 22, 24]. This approach showed that the chimeric protein, driven by the cytoplasmic sequence of LAMP, was targeted to LAMP-containing organelles, which also co-localized with MHC-class-II-rich intracellular compartments [25]. The immunofluorescence microscopy study of our plasmid expressing the YF pM/M-E in-frame with LAMP (pL/YFE) produced findings similar to those of previous studies and suggests that the presence of LAMP was indeed able to lead the chimeric protein to lysosomes; in contrast, the expression of pM/M-E without LAMP (p/YFE) resulted in a reticular membrane distribution. We also investigated the

| YF envelope peptide position | Peptide sequence   | Restriction  |
|-----------------------------|-------------------|-------------|
| E25-39                      | LEQDKCITYMAPDNP   | CD4         |
| E57-71                      | RKVCTCNVTHVKN     | CD8<sup>1</sup>/CD4 |
| E61-75                      | YNAVTHVKINDKCP    | CD8<sup>1</sup>/CD4 |
| E129-143                    | EVDDTQIKYVIRAQL   | CD4         |
| E133-147                    | TKIQVIRAOHVGA     | CD4<sup>2</sup> |
| E157-171                    | KTLKFDALGSEQVE    | CD4         |
| E201-215                    | ESIVDRGWAQDLTL    | CD4         |
| E213-227                    | LTLPWGSNGGGVWRE   | CD4         |
| E221-235                    | SGGVWREMHHLVFE    | CD4         |
| E233-247                    | EFEPPHAATIRVLAL   | CD4         |
| E237-251                    | PHAATIRVLALGNQE   | CD4         |
| E329-343                    | PCRIPIVADDLTA     | CD8         |
| E425-439                    | GFFTSVGKIHVTFG    | CD4         |
| E437-451                    | VFGSAFQGLFGLPNW   | CD4         |
| E461-475                    | LIWVGINTRNMMSM    | CD4         |
| E465-479                    | GINTRNMMSMSMIL    | CD4         |
| E473-487                    | MNSMALVGIMMFIL    | CD4         |
| E477-491                    | MILVGLIMFLGFLG    | CD4         |

<sup>1</sup> These peptides contained class I immunodominant epitopes and secondary class II epitopes.

<sup>2</sup> These peptides contained class II immunodominant epitopes (Maciel et al., 2008).
immunogenicity of these two plasmids as DNA vaccines against YF virus infection. The performance of our DNA constructs was compared with the successful human YF 17DD vaccine, which is a better positive control than the inactivated virus emulsified in CFA, for example [17, 22], which is used when an approved vaccine is not available.

We first carried out epitope mapping of the E protein, comparing the 17DD vaccine with the p/YFE and pL/YFE DNA constructs. We observed that the epitope profile repertoire

Fig 4. Kinetics of the neutralizing antibody levels in C57Bl/6 and BALB/c mice. C57Bl/6 and BALB/c mice were immunized with the YF 17DD vaccine, the DNA constructs or PBS on days 0, 30 and 45, and sera were obtained on days 15, 45 and 60. Sera were tested individually in neutralization assays and compared with a standard monkey serum having a known concentration of anti-YF neutralizing antibodies. The figures represent 1 of 3 experiments, which each had similar results. Bars represent the mean ± SE of 3–14 mice/group.

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recognized by the T cells of mice immunized with the DNA constructs was very similar to that of mice immunized with the standard YF vaccine. Moreover, the majority of the T-cell responses in the DNA-immunized mice showed higher numbers of IFN-gamma SFCs compared with the numbers observed for the 17DD vaccine. The immunization of C57Bl/6 mice with the DNA constructs expressing the whole envelope protein resulted in recognition of three extra peptides that were not produced by the YF 17DD vaccine. The T-cell response against the peptide E169–183 generated by p/YFE and pL/YFE was significant, and the responses to the peptides E413–427 and E417–431 were even higher in mice immunized with p/YFE compared with mice immunized with pL/YFE.

It seems that DNA immunization was able to lead to the presentation of some additional epitopes that are not normally induced by the YF 17DD vaccine. It is possible that different antigen-presenting cells (APCs) processed different epitopes, according to the source of the antigen, i.e., attenuated virus or DNA plasmids. However, the lack of response to the E329–343 15-mer observed in BALB/c mice immunized with DNA seemed to be partially because the same cells were able to respond to the minimum epitope within that sequence, as seen against the 9-mer E330–338. Our results also expanded the C57Bl/6 (H2b) epitope mapping for the envelope protein of the YF virus. Sequences E1–15 and E233–247 were previously described as containing CD8 and CD4 epitopes [39]; however, we were able to identify several new epitopes, six for CD4 and four for CD8 (Table 2).

The presence of anti-YF nAb is a recognized hallmark of protection against YF infection. A dose of 10⁴ PFUs of YF 17DD virus was potent enough to produce a high concentration of nAb after a single immunization in both mouse strains. The plasmid expressing the chimeric E-LAMP protein (pL/YFE) was able to produce significantly higher concentrations of anti-YF nAb after three immunizations in both mouse strains compared with the controls or p/YFE; however, the levels of nAb were considerably lower compared with the YF 17DD immunization. The p/YFE plasmid, expressing only the YF E protein, failed to generate high levels of anti-YF nAb in BALB/c mice and produced only a modest increase of anti-YF nAb in C57Bl/6 mice. These data are in accordance with previous reports demonstrating that the expression of chimeric proteins in-frame with LAMP lead to an improvement in B-cell responses [17, 22, 30]. Others have used extended immunization protocols to produce higher levels of antibody [17]. Although we have not tested this hypothesis here, it is interesting to speculate that extra DNA immunizations could further increase the levels of anti-YF nAb observed.

In addition to nAb, complement-fixing antibodies have also been described as a protective mechanism against YF [40, 41]. In fact, it has been shown in an animal model that expression of the YF NS1 protein in the vaccinia virus could partially protect mice from an intracranial
challenge [42], most likely through a mechanism involving complement-fixing antibodies. We cannot rule out the hypothesis that immunization with the YF envelope protein, as a DNA plasmid, could lead to the presentation of B-cell epitopes different from those found after
immunization with the attenuated YF vaccine. Moreover, these epitopes could be sites for neutralizing and complement-fixing antibodies. Thus, the characterization of the B-cell epitopes in the context of DNA vaccines could potentially become a relevant parameter for comparing DNA vaccines to their virus-attenuated counterparts.

To further explore the protection provided by DNA immunization, we challenged immunized mice with an intracranial injection of the 17DD virus. This in vivo protection assay enables the evaluation of how effectively a vaccine candidate can prevent the encephalitis caused by infection with the YF virus, and it has been extensively used [35, 41–44]. Both DNA constructs, p/YFE and pL/YFE, were able to protect both mouse strains from an intracranial challenge. These two plasmids were able to promote a very similar profile of T-cell epitope recognition compared to the YF 17DD vaccine. However, only the pL/YFE was able also to produce significant levels of anti-YF nAb. It is possible that the p/YFE plasmid, which did not raise appreciable levels of anti-YF nAb, led to the protection of the challenged mice through complementary mechanisms in the presence of low levels of nAb. It is also possible that strong T-cell responses were able to mediate protection in this system. T cells may play a role in protection from encephalitis caused by flaviviruses; it has been demonstrated that the depletion of CD4+ and/or CD8+ lymphocytes leads to a decrease in the protection offered by an experimental vaccine expressing the dengue envelope protein in the context of the YF virus [39]. The possibility that these DNA vaccines provide T-cell mediated protection is very interesting and we are planning to investigate this in more detail.

The results reported here are very encouraging, and we are confident that this vaccine candidate is worth further investigation in more relevant animal models, specifically in a non-human primate challenge model. It is interesting that even with lower neutralizing antibody titers the DNA vaccine was still capable of protection, suggesting an important role for T-cell mediated protection. In further studies, we plan to dissect in more details the mechanisms of protection provided by these DNA vaccines. Another critical point is the duration of the protection, and this also needs to be addressed in more relevant animal models. In summary, this research shows that expression of the envelope protein in-frame with the cytoplasmic targeting sequence of LAMP led to high levels of anti-YF nAb and produced a strong T-cell response. The possibility of generating a protective anti-YF response through a DNA vaccine may provide a safer alternative to the attenuated YF virus vaccine and should be further investigated.

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Author Contributions

Conceived and designed the experiments: MM RG ETAM RD. Performed the experiments: MM FdSPC MTC MAdM RdCCM RFCB RD RCBQdF. Analyzed the data: MM MTC MADM JTA ETAM RD RCBQdF. Contributed reagents/materials/analysis tools: RG MdSF JTA ETAM. Wrote the paper: MM KMSdMC JTA ETAM RD.

References

1. Julander JG. Experimental therapies for yellow fever. Antiviral Res. 2013; 97(2):169–79. doi: 10.1016/j.antiviral.2012.12.002 PMID: 23237991
2. World Health Organization, Division of Epidemiological Surveillance and Health Situation Trend Assessment. Global health situation and projections—estimates. Geneva, Switzerland: World Health Organization; 1992.

3. Monath TP, Cetron MS, McCarthy K, Nichols R, Archambault WT, Weld L, et al. Yellow fever 17D vaccine safety and immunogenicity in the elderly. Hum Vaccin 2005; 1(5): 207–214. PMID: 17012867

4. Poland JD, Calisher CH, Monath TP, Downs WG, Murphy K. Persistence of neutralizing antibody 30–35 years after immunization with 17D yellow fever vaccine. Bull World Health Organ 1981; 59(6): 895–900. PMID: 6978196

5. Reinhardt B, Jaspert R, Niedrig M, Kostner C, L’Age-Stehr J. Development of viremia and humoral and cellular parameters of immune activation after vaccination with yellow fever virus strain 17D: a model of human flavivirus infection. J Med Virol 1998; 56(2): 159–167. PMID: 9746073

6. Breugelmans JG, Lewis RF, Agbenu E, Veit O, Jackson D, Domingo C, et al. Adverse events following yellow fever preventive vaccination campaigns in eight African countries from 2007 to 2010. Vaccine. 2013; 31(14):1819–29. doi: 10.1016/j.vaccine.2013.01.054 PMID: 23395587

7. Miyaji KT, Luiz AM, Lara AN, do Socorro Souza Chaves T, Piorelli RD, Lopes MH, et al. Active assessment of adverse events following yellow fever vaccination of persons aged 60 years and more. Hum Vaccin Immunother. 2013; 9(2).

8. Sidibe M, Yactayo S, Kalle A, Sall AA, Sow S, Ndoutabe M, et al. Immunogenicity and safety of yellow fever vaccine among 115 HIV-infected patients after a preventive immunisation campaign in Mali. Trans R Soc Trop Med Hyg. 2012; 106(7):437–44. doi: 10.1016/j.trstmh.2012.04.002 PMID: 22627101

9. Thomas RE, Lorenzetti DL, Spragins W, Jackson D, Williamson T. Reporting rates of yellow fever vaccine 17D or 17DD-associated serious adverse events in pharmacovigilance data bases: systematic review. Curr Drug Saf. 2011; 6(3):145–54. PMID: 22122389

10. Monath TP, Nichols R, Archambault WT, Moore L, Marchesani R, Tian J, et al. Comparative safety and immunogenicity of two yellow fever 17D vaccines (ARILVAX and YF-VAX) in a phase III multicenter, double-blind clinical trial. Am J Trop Med Hyg 2002; 66(5): 533–541. PMID: 12201587

11. Lefeuvre A, Marianneau P, Deubel V. Current Assessment of Yellow Fever and Yellow Fever Vaccine. Curr Infect Dis Rep 2004; 6(2):96–104. PMID: 15023271

12. Vasconcelos PF, Luna EJ, Galler R, Silva LJ, Coimbra TL, Barros VL, et al. Brazilian Yellow Fever Vaccine Evaluation. Serious adverse events associated with yellow fever 17DD vaccine in Brazil: a report of two cases. Lancet 2001; 358(9276): 91–97. PMID: 11463409

13. Lewis PJ, Babiuk LA. DNA vaccines: a review. Adv Virus Res 1999; 54: 129–188. PMID: 10547676

14. Li L, Saade F, Petrovsky N. The future of human DNA vaccines. J Biotechnol. 2012; 162(2–3):171–82. doi: 10.1016/j.jbiotec.2012.09.018 PMID: 23036927

15. Robinson HL. DNA vaccines: basic mechanism and immune responses (Review). Int J Mol Med 1999; 4(5): 549–555. PMID: 10534580

16. Schultz J, Dollenmaier G, Molling K. Update on antiviral DNA vaccine research (1998–2000). Intervirology 2000; 43(4–6): 197–217. PMID: 11251381

17. Lu Y, Raviprakash K, Leao IC, Chikhlikar PR, Ewing D, Anwar A, et al. Dengue 2 PreM-E/LAMP chimera targeted to the MHC class II compartment elicits long-lasting neutralizing antibodies. Vaccine 2003; 21(17–18): 2178–2189. PMID: 12850357

18. Raviprakash K, Kochel TJ, Ewing D, Simmons M, Phillips I, Hayes CG, et al. Immunogenicity of dengue virus type 1 DNA vaccines expressing truncated and full length envelope protein. Vaccine 2000; 18(22): 2426–2434. PMID: 10738100

19. Konishi E, Terazawa A, Fuji A. Evidence for antigen production in muscles by dengue and Japanese encephalitis DNA vaccines and a relation to their immunogenicity in mice. Vaccine 2003; 21(25–26): 3713–3720.

20. Konishi E, Yamaoka M, Khin Sane W, Kurane I, Mason PW. Induction of protective immunity against Japanese encephalitis in mice by immunization with a plasmid encoding Japanese encephalitis virus premembrane and envelope genes. J Virol 1998; 72(6): 4925–4930. PMID: 95732560

21. Wu CJ, Li TL, Huang HW, Tao MH, Chan YL. Development of an effective Japanese encephalitis virus-specific DNA vaccine. Microbes Infect 2006; 8(11): 2578–2586. PMID: 16949850

22. Anwar A, Chandrasekaran A, Ng ML, Marques E, August JT. West Nile premembrane-envelope genspecific vaccine encoded as a chimera containing the transmembrane and cytoplasmic domains of a lysosome-associated membrane protein: increased cellular concentration of the transgene product, targeting to the MHC II compartment, and enhanced neutralizing antibody response. Virology 2005; 332(1): 66–77. PMID: 15661141
23. Konishi E, Yamaoka M, Kurane I, Mason PW. A DNA vaccine expressing dengue type 2 virus premembrane and envelope genes induces neutralizing antibody and memory B cells in mice. Vaccine 2000; 18(11–12): 1133–1139.

24. Raviprakash K, Marques E, Ewing D, Lu Y, Phillips I, Porter KR, et al. Synergistic neutralizing antibody response to a dengue virus type 2 DNA vaccine by incorporation of lysosome-associated membrane protein sequences and use of plasmid expressing GM-CSF. Virology 2001; 290(1): 74–82. PMID: 11883007

25. Marques ET Jr, Chikhlikar P, de Arruda LB, Leao IC, Lu Y, Wong J, et al. HIV-1 p55Gag encoded in the lysosome-associated membrane protein-1 as a DNA plasmid vaccine chimera is highly expressed, traffics to the major histocompatibility class II compartment, and elicits enhanced immune responses. J Biol Chem 2003; 278(39): 37926–37936. PMID: 12824194

26. de Arruda LB, Chikhlikar PR, August JT, Marques ET. DNA vaccine encoding human immunodeficiency virus-1 Gag, targeted to the major histocompatibility complex II compartment by lysosomal-associated membrane protein, elicits enhanced long-term memory response. Immunology 2004; 112(1): 126–133. PMID: 15129672

27. Lippincott-Schwartz J, Fambrough DM. Lysosomal membrane dynamics: structure and interorganellar movement of a major lysosomal membrane glycoprotein. J Cell Biol 1986; 102(5): 1593–1605. PMID: 2871029

28. Drake JR, Lewis TA, Condon KB, Mitchell RN, Webster P. Involvement of MiIC-like late endosomes in B cell receptor-mediated antigen processing in murine B cells. J Immunol 1999; 162(2): 1150–1155. PMID: 9916746

29. Kleijmeer MJ, Morkowski S, Griffith JM, Rudensky AY, Geuze HJ. Major histocompatibility complex class II compartments in human and mouse B lymphoblasts represent conventional endocytic compartments. J Cell Biol 1997; 139(3): 639–649. PMID: 9348281

30. Arruda LB, Sim D, Chikhlikar PR, Maciel M Jr, Akasaki K, August JT, et al. Dendritic cell-lysosomal-associated membrane protein (LAMP) and LAMP-1-HIV-1 gag chimeras have distinct cellular trafficking pathways and prime T and B cell responses to a diverse repertoire of epitopes. J Immunol 2006; 177(4): 2265–2275. PMID: 16897987

31. Chikhlikar P, Barros de Arruda L, Agrawal S, Byrne B, Guggino W, August JT, et al. Inverted terminal repeat sequences of adeno-associated virus enhance the antibody and CD8(+) responses to a HIV-1 p55Gag/LAMP DNA vaccine chimera. Virology 2004; 323(2): 220–232. PMID: 15139318

32. Stefano I, Sato HK, Panunuti CS, Omoto TM, Mann G, Freire MS et al. Recent immunization against dengue virus does not interfere with the sero-response to yellow fever vaccine. Vaccine 1999; 17(9–10):1042–6.

33. Maciel M Jr, Kellathur SN, Chikhlikar P, Dhalia R, Sidney J, Sette A et al. Comprehensive analysis of T cell epitope discovery strategies using 17DD yellow fever virus structural proteins and BALB/c (H2d) mice model. Virology 2008; 378(1):105–17. doi: 10.1016/j.virol.2008.04.043 PMID: 18579176

34. WHO. Requirements for the production and control of yellow fever vaccine. (Requirements for Biological Substances No. 3.) In: WHO Expert Committee on Biological Standardization. Forty-sixth report. Geneva, World Health Organization, 1998, Annex 2 (WHO Technical Report Series, No. 872).

35. Barrett AD, Gould EA. Comparison of neurovirulence of different strains of yellow fever virus in mice. J Gen Virol 1986; 67(4): 631–7.

36. Cetron MS, Marfin AA, Julian KG, Gubler DJ, Sharp DJ, Barwick RS, et al. Yellow fever vaccine. Recommendations of the Advisory Committee on Immunization Practices (ACIP), 2002. MMWR Recomm Rep 2002;51(RR-17): 1–11; quiz CE11-14.

37. Disis ML, Gralow JR, Bernhard H, Hand SL, Rubin WD, Cheever MA. Peptide-based, but not whole protein, vaccines elicit immunity to HER-2/neu, oncogenic self-protein. J Immunol 1996; 156(9): 3151–3158. PMID: 8617935

38. Sercazz EE, Lehmman PV, Ametani A, Benichou G, Miller A, Moudgil K. Dominance and crypticity of T cell antigenic determinants. Annu Rev Immunol 1003; 11: 729–766. PMID: 7682817

39. van Der Most RG, Murali-Krishna K, Ahmed R, Strauss JH. Chimeric yellow fever/dengue virus as a candidate dengue vaccine: quantitation of the dengue virus-specific CD8 T-cell response. J Virol 2000; 74(17):8094–101 PMID: 10993719

40. Brandriss MW, Schlesinger JJ, Walsh EE. Passive protection by monoclonal antibodies to the envelope proteins of 17D yellow fever and dengue 2 viruses. J Gen Virol 1986; 67(2):229–34.

41. Schlesinger JJ, Brandriss MW, Walsh EE. Protection against 17D yellow fever encephalitis in mice by passive transfer of monoclonal antibodies to the nonstructural glycoprotein gp48 and by active immunization with gp48. J Immunol 1985; 135(4):2805–9. PMID: 4031501
42. Putnak JR, Schlesinger JJ. Protection of mice against yellow fever virus encephalitis by immunization with a vaccinia virus recombinant encoding the yellow fever virus non-structural proteins, NS1, NS2a and NS2b. J Gen Virol 1990; 71 (Pt 8):1697–702.

43. Despres P, Dietrich J, Girard M, Bouloy M. Recombinant baculoviruses expressing yellow fever virus E and NS1 proteins elicit protective immunity in mice. J Gen Virol 1991; 72 (Pt 11):2811–6. PMID: 1834798

44. Pincus S, Mason PW, Konishi E, Fonseca BA, Shope RE, Rice CM et al. Recombinant vaccinia virus producing the prM and E proteins of yellow fever virus protects mice from lethal yellow fever encephalitis. Virology 1992; 187(1):290–7. PMID: 1736531