Article

Graphene Oxide/Ferrocene-Containing Polymer/Gold Nanoparticle Triple Nanocomposite

Wenhao Qian 1,*, Tao Song 1, Mao Ye 1, Haiyan Zhang 1, Chun Feng 2, Guolin Lu 2 and Xiaoyu Huang 2,*

1 Department of Stomatology, Shanghai Xuhui District Dental Center, 685 Zhaojiabang Road, Shanghai 200032, China; songtao21@139.com (T.S.); doreye@139.com (M.Y.); 13641956377@163.com (H.Z.)
2 Key Laboratory of Synthetic and Self-Assembly Chemistry for Organic Functional Molecules, Center for Excellence in Molecular Synthesis, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, 345 Lingling Road, Shanghai 200032, China; cfeng@sioc.ac.cn (C.F.); luguolin@sioc.ac.cn (G.L.)
* Correspondence: pingyanlaoto@163.com (W.Q.); xyhuang@sioc.ac.cn (X.H.); Tel.: +86-21-64045643 (W.Q.); +86-21-54925310 (X.H.)

Received: 24 January 2019; Accepted: 20 February 2019; Published: 25 February 2019

Abstract: A facile strategy to prepare GO-based nanocomposites with both gold nanoparticles (AuNPs) and ferrocene (Fc) moieties was developed. The surface of GO was modified with PFcMAss homopolymer by surface-initiated atom transfer radical polymerization of a new methacrylate monomer of 2-((2-(methacryloyloxy)ethyl)disulfanyl)ethyl ferrocene-carboxylate (FcMAss), consisting of disulfide as an anchoring group for stabilizing AuNPs and Fc group as an additional functionality. AuNPs with an average diameter of about 4.1 nm were formed in situ on the surface of PFcMAss-decorated GO (GO-PFcMAss) via Brust-Schiffrin method to give GO-PFcMAss-AuNPs multifunctional nanocomposites bearing GO, AuNPs and Fc groups. The obtained nanocomposites were characterized by X-ray photoelectron spectroscopy (XPS), X-ray diffraction (XRD), transmission electron microscopy (TEM) and atomic force microscopy (AFM). Since disulfide-containing polymers, rather than the commonly used thiol-containing compounds, were employed as ligands to stabilize AuNPs, much more stabilizing groups were attached onto the surface of GO, and thus more AuNPs were able to be introduced onto the surface of GO. Besides, polymeric chains on the surface of GO endowed GO-PFcMAss-AuNPs nanocomposites with excellent colloidal stability, and the usage of a disulfide group provides possibility to efficiently incorporate additional functionalities by easily modifying structure of disulfide-based monomer.

Keywords: graphene oxide; ferrocene; ATRP; disulfide bond; gold nanoparticle

1. Introduction

Graphene, a fascinating two-dimensional graphitic carbon system, has quickly sparked tremendous interests across many fields including biosensors, electrochemical energy storage and electronics due to its extraordinary properties of large surface area, good mechanical properties and extremely high electronic conductivity properties [1–6]. Graphene oxide (GO), a main derivative of graphene, offers great potential for large-scale production of graphene-based materials by taking advantage of carboxyl, hydroxyl and epoxy groups on the surface of GO. As GO can be prepared efficiently in large scale and low-cost fashion from cheap graphite by chemical methods [7–10], it is considered as a star building motif for the construction of graphene-based composite materials. Up to now, a large variety of functional moieties, including inorganic particles, polymers, metal-organic frameworks (MOFs), carbon nanotubes (CNTs) and so on, have been incorporated onto the surface of GO to improve inherent properties of GO or endow GO-based hybrid materials with new
functionalities [11–23]. Although significant advances have been achieved in the preparation of GO-based hybrid materials, increasing attention has been still paid on exploring more facile and robust strategies to efficiently introduce diverse functionalities onto the surface of GO for the fabrication of hierarchically multi-functional GO-based composite materials.

Gold nanoparticles (AuNPs), one of the most attractive inorganic nanoparticles, have attracted intense interest for applications in photonics, information storage, electronic and optical detection system, therapeutics, diagnostics, photovoltaics and catalysis because of their unique properties, such as high surface free energy, strong adsorption ability, distinctive optical and electrical properties, and excellent biocompatibility [24–31]. Therefore, various composites consisting of AuNPs and GO have been fabricated to prepare desired multi-functional materials, in which functionalities of AuNPs and GO were combined together. The synergetic effect would not only lead to the improvement of performance of composites, but also endow the composites with some new functionalities, which cannot be gained by either GO or AuNPs separately [32–38]. In previous reports, AuNPs were anchored onto the surface of GO via either noncovalent interactions including van der Waals interaction, hydrogen bonding, π-π stacking, electrostatic interactions, or covalent bonds [32–38]. Since the noncovalent interactions can be easily weakened by temperature, salt, pH or some specific compounds, the limited stability of GO/AuNPs composites would hinder the application of these composites. Although the stability can be significantly improved by using covalent bonds instead of noncovalent interactions, only a few approaches were developed [39–43]. For these approaches, the linkers between AuNPs and GO were small molecules with a thiol group, where theoretically one reactive site of GO only can be used to install one thiol group. But a large number of thiol-based ligands are generally necessary to stabilize one gold nanoparticle. Thus, the accessibility of GO surface to AuNPs was highly retarded. In addition, these methods were not efficient for the incorporation of additional functional moieties.

Ferrocene (Fc) is an interesting metal-containing compound with unique redox, preceramic, etch-resistant and catalytic properties [44–46]. Thus, Fc and its derivatives are considered to be attractive building blocks for the preparation of functional materials. For example, Fc has been widely used to construct electrochemical biosensors due to its reversibility, regeneration at low potential and generation of stable redox states [44–46]. Ferrocene and its derivatives, as excellent redox mediators, have been well studied in electrochemical sensing. However, ferrocene cannot form stable adsorption layers on electrodes so that enormous efforts have been performed to immobilize ferrocene and its derivatives on electrode. As it is well known, various nanomaterials, such as AuNPs, carbon nanotubes and graphene, have been used as excellent carriers to enhance the sensitivity of sensors. Herein, we reported a facile strategy to prepare GO-based nanocomposites with both AuNPs and Fc moieties as shown in Scheme 1. We firstly designed and synthesized a new methacrylate monomer of 2-((2-(methacryloyloxy)ethyl)disulfanyl)ethyl ferrocenecarboxylate (FcMAss) with disulfide and Fc groups, in which disulfide group can serve as the anchoring group for AuNPs and Fc group can act as an additional functional unit. Subsequently, poly(2-((2-(methacryloyloxy)ethyl)disulfanyl)ethyl ferrocenecarboxylate) (PFcMAss) was introduced onto the surface of GO by surface-initiated atom transfer radical polymerization (SI-ATRP) [47,48]. Then, by the virtue of the strong coordinating ability of disulfide moieties toward gold nanoparticles, gold nanoparticles with an average diameter of 4.1 nm were formed in situ on the surface of PFcMAss-decorated GO (GO-PFcMAss) via Brust-Schiffrin method [49] to give a multi-functional nanocomposite of GO-PFcMAss-AuNPs consisting of GO, AuNPs and Fc groups. In the current case, disulfide-containing polymers covalently connected to GO can be employed as ligands to stabilize AuNPs and simultaneously introduce Fc groups onto the surface of AuNPs. Therefore, more AuNPs and Fc groups are able to be immobilized onto the surface of GO, which allows the amplification of detection signal. Thus, the obtained nanomaterials combining ferrocene, gold nanoparticles and GO together, may allow the development of a signal amplification detection platform for electro-active molecules. The obtained GO-PFcMAss-AuNPs nanocomposite was characterized by X-ray photoelectron spectroscopy (XPS), X-ray diffraction (XRD), transmission electron microscopy (TEM) and atomic force microscopy (AFM).
Scheme 1. Preparation of GO-PFcMAss-AuNPs nanocomposite.

2. Experimental

2.1. Characterization

FT-IR spectra were recorded on a Nicolet AVATAR-360 FT-IR spectrophotometer (Waltham, MA, USA) with a resolution of 4 cm⁻¹. ¹H-NMR measurements were performed on a JEOL resonance ECZ 400S s (400 MHz) pectrometer (Tokyo, Japan) in CDCl₃ using tetramethylsilane (TMS) as an internal standard. Electrospray ionization mass spectrometry (ESI-MS) was measured by an Agilent FTMS-7.0 Fourier transformation mass spectrometer (Santa Clara, CA, USA). Relative molecular weight and molecular weight distribution of PFcMAss homopolymer were measured by a conventional gel permeation chromatography (GPC) system equipped with a Waters 1515 Isocratic HPLC pump (Milford, MA, USA), a Waters 2414 refractive index detector (Milford, MA, USA) and a set of Waters Styragel columns (HR3 (500–30,000), HR4 (5,000–600,000) and HR5 (50,000–4,000,000), 7.8 × 300 mm, particle size: 5 μm). GPC measurement was carried out at 35 °C using THF as eluent with a flow rate of 1.0 mL/min. The system was calibrated with linear poly(methyl methacrylate) standards. Powder X-ray diffraction (XRD) measurements were run by a Philips X’Pert PRO X-ray powder diffractometer (Amsterdam, Netherlands) with Cu Kα (1.541 Å) radiation (40 kV, 40 mA), the samples were exposed at a scan rate of 3° = 0.0424°/s in the range of 5° and 60°. X-ray photoelectron spectroscopy (XPS) was recorded on a PHI 5000c ESCA photoelectron spectrometer (Waltham, IN, USA). Transmission electron microscopy (TEM) images were obtained by a JEOL JEM 1230 instrument (Tokyo, Japan) operated at 80 kV. Atomic force microscopy (AFM) images were taken by a Veeco DI MultiMode SPM (Plainview, TX, USA) in the tapping mode of dropping the sample solution onto the freshly exfoliated mica substrate.

2.2. Preparation of 2-((2-(Methacryloyloxy)ethyl)disulfanyl)ethyl Ferrocene-carboxylate (FcMAss)

Dichloromethane (DCM) was dried over CaH₂ and distilled over CaH₂ under N₂ prior to use. Triethylamine (Et₃N, Aldrich, 99.5%) was dried over KOH and distilled over CaH₂ under N₂ prior to use. As shown in Scheme 2, ferrocenyl chloride obtained by reacting ferrocenecarboxylic acid (Alfa Aesar (Shanghai, China), 98%) with oxalyl chloride (J&K, Beijing, China, 98%) was treated with bis(2-hydroxyethyl) disulfide (Alfa Aesar, 90%) to afford the monoesterification product, which reacted with methacryloyl chloride (Aldrich, St. Louis, MO, USA, 90%) in the presence of Et₃N to produce the target monomer, FcMAss (total yield: 53.4%). FI-IR (film): ν (cm⁻¹): 2960, 2858, 1716, 1637, 1460, 1380, 1273, 1218, 1132, 1068, 941, 912, 818, 773. ¹H-NMR (400 MHz, CDCl₃): δ (ppm): 1.95 (3H, CH₃C), 3.02 (4H, CO₂CH₂CH₂C=CCH₂CH₂O₂C), 4.21, 4.40, 4.81 (9H, ferrocenyl), 4.45 (4H, CO₂CH₂CH₂S-SCH₂CH₂O₂C), 5.85, 6.14 (2H, CH₂=C).

2.3. Preparation of PFcMAss via ATRP of FcMAss

Toluene (Aldrich, 99%) were dried over CaH₂ and distilled from sodium and benzophenone under N₂ prior to use. Copper(I) bromide (CuBr, Aldrich, 98%) was purified by stirring over CH₃COOH overnight at room temperature, followed by washing with ethanol, diethyl ether and acetone prior to drying at 40 °C in vacuo for one day. As shown in Scheme 3, CuBr (5.8 mg, 0.04 mmol) and FcMAss (520.8 mg, 1.2 mmol) were first added to a 25 mL Schlenk flask (flame-dried
under vacuum prior to use) sealed with a rubber septum. After evacuating and backfilling with Ar, pentamethyldiethylenetriamine (PMDETA, Aldrich, 99%, 8.5 μL, 0.04 mmol), ethyl α-bromoisobutyrate (EBiB, Aldrich, 98%, 6 μL, 0.04 mmol) and dry toluene (3 mL) were introduced via a gastight syringe followed by three cycles of freezing-pumping-thawing. The flask was immersed into an oil bath set at 50 °C. After 48 h, the polymerization was terminated by immersing the flask into liquid N2. The reaction mixture was diluted with THF and filtered through a short Al2O3 column to remove the residual copper complex. THF was rotary evaporated and the crude product was purified by dissolving in THF and precipitating in cold n-hexane/diethyl ether (v:v = 1:1) three times followed by drying in vacuo overnight to give 230.8 mg of orange yellow powder, poly(2-((2-(methacyryloyloxy)ethyl)disulfanyl)ethyl ferrocene-carboxylate) (PFCMAss).

![Scheme 2. Synthesis of FcMAss monomer.](image)

**Scheme 2.** Synthesis of FcMAss monomer.

**Scheme 3.** ATRP of FcMAss monomer.

2.4. Preparation of PFCMAss-grafted Graphene Oxide Sheets (GO-PFCMAss)

GO-based initiator containing ATRP initiation groups (–COC(CH3)2Br), TRIS-GO-Ini, was obtained by treating tris(hydroxymethyl) aminomethane (TRIS)-modified GO with 2-bromo-2-methylpropionyl bromide according to previous reports [47,48]. CuBr (3.0 mg, 0.02 mmol), TRIS-GO-Ini (65.8 mg) and FcMAss (625.2 mg, 1 mmol) were added to a 25 mL Schlenk flask (flame-dried under vacuum prior to use) sealed with a rubber septum. After three cycles of evacuating and purging with N2, 5 mL of dried toluene was charged via a gastight syringe and the mixture was sonicated for 30 min in a thermostatic water bath at 25 °C. After two cycles of freezing-pumping-thawing, PMDETA (4.3 μL, 0.02 mmol) and EBiB (3.3 μL, 0.02 mmol) were added and the flask was degassed by three cycles of freezing-pumping-thawing, followed by immersing the flask into an oil bath set at 50 °C. The polymerization lasted 2 days and it was terminated by putting the flask into liquid N2. After diluting with toluene, the reaction mixture was filtered through a 0.22 μm filter and washed exhaustively with toluene and deionized water until the filtrate turned clear.

The obtained black solid was dried overnight at 45 °C in vacuo so as to afford 55.2 mg of GO-PFCMAss. Meanwhile, the filtrate was passed through a short Al2O3 column to remove the residual copper complex and precipitated into cold n-hexane/diethyl ether (v:v = 1:1) three times. Subsequently, free PFCMAss homopolymer was obtained by drying in vacuo overnight at 45 °C. GPC: \(M_n = 13,600 \text{ g/mol}, M_w/M_n = 1.48\). GO-PFCMAss: XPS (molecular molar ratio): C, 72.71%; N, 1.31%; O, 23.26%, Fe, 1.01%, S, 1.60%. FT-IR: \(\nu\) (cm\(^{-1}\)): 2958, 2852, 1708, 1639, 1593, 1459, 1400, 1271, 1138, 817.
2.5. Fabrication of PFcMAss-AuNPs

Nanoparticles were prepared by Brust-Schiffrin protocol with some modifications [49]. A solution containing tetraoctylammonium bromide (TOAB, J&K, 98%, 13.7 mg, 0.025 mmol) in 10 mL of toluene was mixed with a solution containing tetrachloroauric acid (HAuCl₄·4H₂O, Sinopharm, AR, 10.3 mg, 0.025 mmol) in 0.21 mL of deionized water. After stirring at room temperature for 1 h, two layers were observed: the upper organic phase was orange-yellow, while the lower aqueous phase was colorless. Next, a solution containing sodium borohydride (NaBH₄, Alfa Aesar, 98%, 9.5 mg, 0.025 mmol) in 2 mL of deionized water was added dropwise, resulting in a color change of the upper organic phase to wine red. After the mixture was stirred for another 2 h, a solution containing PFcMAss homopolymer (16.4 mg, amount of –S–S– disulfide bond: 0.0375 mmol) in 2 mL of toluene was added to the separated organic phase followed by stirring overnight at room temperature. The organic phase was concentrated and then dispersed in a toluene/ethanol mixture (v/v = 2:3). The resultant was sonicated for 1 min and the precipitant was collected. The crude product was purified by centrifugation to give a wine-red solid of PFcMAss-AuNPs.

2.6. Fabrication of GO-PFcMAss-AuNPs Nanocomposite

Nanoparticles were prepared by Brust-Schiffrin protocol with some modifications [49]. A solution containing TOAB (13.7 mg, 0.025 mmol) in 10 mL of toluene was mixed with a solution containing HAuCl₄·4H₂O (10.3 mg, 0.025 mmol) in 0.21 mL of deionized water. After stirring at room temperature for 1 h, two layers were observed: the upper organic phase was orange-yellow, while the lower aqueous phase was colorless. Next, a solution containing NaBH₄ (9.5 mg, 0.025 mmol) in 2 mL of deionized water was added dropwise, resulting in a color change of the upper organic phase to wine-red. After the mixture was stirred for another 2 h, the upper organic phase was separated. GO-PFcMAss (10 mg) was dispersed in 2 mL of toluene followed by sonication for 10 min. The dispersion was added to the separated organic phase followed by stirring overnight at room temperature. Afterward, the organic phase was diluted with toluene and then centrifugated (6,000 rpm) to remove the unreacted AuNPs. The filtrate was concentrated and diluted with deionized water followed by centrifugation (6,000 rpm) to remove the residual TOAB. The residual mixture was lyophilized to give 31.5 mg of wine-red solid, i.e. GO-PFcMAss-AuNPs.

3. Results and Discussion

3.1. Synthesis of FcMAss Monomer and Preparation of FcMAss Homopolymer

Although thiol-based compounds have been widely used as anchoring groups via Au–S bond, the thiol group, which can serve as a chain transfer agent, will interfere with smooth process of living/controlled radical polymerization [50–52]. Therefore, thiol-containing monomer generally cannot be directly polymerized by these methods. Satisfyingly, Tong and co-workers reported that dialkyl disulfide also can serve as anchoring group to stabilized AuNPs, similar to thiol [53]. Therefore, a new methacrylate monomer of 2-((2-(methacryloyloxy)ethyl)disulfanyl)ethyl ferrocene-carboxylate (FcMAss) containing disulfide and Fc groups was designed and synthesized via the estification reaction between methacryloyl chloride and the intermediate which was obtained by the reaction of bis(2-hydroxyethyl) disulfide with ferrocenyl chloride (Scheme 2). For this new monomer, disulfide serves as an anchoring group to stabilize AuNPs and Fc is used as a model moiety to demonstrate that disulfide group can serve as a linker to introduce some desired functional groups. 

The chemical structure of FcMAss monomer was examined by FT-IR, ¹H-NMR and MS. In Figure 1A, the proton resonance signals of double bond located at 6.14 (peak ‘a’) and 5.59 (peak ‘a’) ppm and 9 protons of Fc unit located at 4.81 (peak ‘g’), 4.40 (peak ‘h’) and 4.21 (peak ‘i’) ppm were found, and the ESI-MS result (434) was also consistent with the theoretical value. All these results confirmed the successful synthesis of FcMAss monomer.
Surface-modification for various materials can not only transform those materials with have excellent bulk physical and chemical properties into valuable materials with suitable surface properties required for specific applications, but also combine different functionalities of various materials to provide desired materials with multi-functionality or improved functionality. To extend the application of carbon nanomaterials, great efforts on the surface modification of carbon nanomaterials including fullerenes, carbon nanotubes, graphene and GO, have been made to improve the properties or introduce new functionality [54–58]. Among these methods, surface modification through grafting polymers is one of the most efficient ways to adjust surface functionality and properties. In the present work, SI-ATRP, which is considered to be one of the most versatile techniques to introduce functional polymeric chains onto the surface of matrices to prepare hybrid organic/inorganic materials [59–62], was used for preparation of PFcMAss-grafted graphene sheets. Before the surface functionalization of GO with PFcMAss by SI-ATRP, we firstly ran bulk ATRP of FcMAss by using ethyl α-bromoisobutyrate (EBiB) as initiator and CuBr/PMDETA as catalytic system at 50 °C to explore suitable polymerization conditions for SI-ATRP (Scheme 3). After polymerization for 2 days, GPC curve of the resulting PFcMAss homopolymer (Figure S1) shows only a unimodal and symmetrical eluent peak with a relatively narrow molecular weight distribution ($M_w/M_n = 1.33$). Moreover, typical proton resonance signals of double bond also disappeared while characteristic peaks attributed to the protons of polymethacrylate backbone appeared at 0.75–1.26 and 1.87 ppm (peak ‘a’ and ‘b’) in $^1$H NMR spectrum after ATRP (Figure 1B). The preserving of signals attributed to Fc (peak ‘g’, ‘h’ and ‘i’) and methylene (peak ‘c’, ‘d’, ‘e’ and ‘i’) groups of FcMAss repeat unit illustrated that the structure of monomer remained intact during the polymerization. In addition, the appearance of a minor peak at 3.75 ppm (peak ‘l’) assigned to two protons of methylene moiety in the initiating group verified the mechanism of ATRP. The molecular weight of obtained PFcMAss homopolymer was estimated to be about 8,400 g/mol on the basis of integrations of peak ‘l’ and ‘g’ ($M_n,NMR = M_{ini} + M^* S_G / S_l$), $M_{ini}$ and $M$ are molecular weights of EBiB initiator and FcMAss monomer, respectively; $S_l$ and $S_g$ are integrations of peak ‘l’ and ‘g’, respectively), which is very closed to the value obtained by GPC ($M_{n,GPC} = 8,600$ g/mol). Thus, it is clear that FcMAss monomer could be successfully polymerized by ATRP using CuBr/PMDETA as catalytic system and EBiB as initiator at 50 °C.

Figure 1. $^1$H-NMR spectra of (A) FcMAss monomer and (B) PFcMAss homopolymer.

3.2. Preparation of PFcMAss-Grafted Graphene Sheets (GO-PFcMAss)

According to our previous reports [47,48], ATRP initiating groups were introduced onto the surface of GO by treating GO with tris(hydroxymethyl) aminomethane (TRIS), followed by reacting with BrCOC(CH$_3$)$_2$Br. SI-ATRP of FcMAss was then conducted at 50 °C in toluene using Br-decorated
GO as initiator and CuBr/PMDETA as catalytic system. EBIB was also added into the polymerization system as a sacrificial initiator for subsequent determination of molecular weight of PFcMAss chains grafted from GO, assuming that free polymers formed in the solution have the similar molecular weight as those formed on the surface [62–64].

The as-prepared GO-PFcMAss samples showed good dispersible properties in solvents such as acetone, chloroform, THF, DMF and toluene (Figure 2). However, they showed poor dispersibility in water and methanol, which are good solvents for GO, but poor solvents for PFcMAss (Figure 2A). From FT-IR spectra of GO before and after SI-ATRP (Figure 3), one can notice that typical signals of Fc group at 2958, 1459 and 817 cm\(^{-1}\) and a new peak at 2852 cm\(^{-1}\) attributed to stretching vibration of methylene group appeared, along with an obvious increase of the intensity of characteristic peak of carbonyl at 1708 cm\(^{-1}\). In addition, a characteristic disulphide mode at 527 cm\(^{-1}\) was observed, indicating the presence of disulfide bonds in the obtained GO-PFcMAss. From XPS spectra of pristine GO and functionalized GO after SI-ATRP (Figure 4A), two peaks at 709 and 71 eV were attributed to Fe 2p and Fe 3p3, along with two peaks at 229 and 165 eV originating from S 2s and S 2p3. From the elemental composition of obtained GO-PFcMAss obtained from XPS (Table S1), one can see that the content of Fe (mol%) is about 1.1 mol\%, which meant that the weight content of PFcMAss in GO-PFcMAss composite is about 44.2 wt\%. Moreover, GPC analysis showed that free PFcMAss homopolymer had a \(M_n\) of 13,600 g/mol with a \(M_w/M_n\) of 1.48 (Figure S1). All of these observations indicated that PFcMAss chains with a \(M_n\) of 13,600 g/mol were introduced onto the surface of GO by SI-ATRP.

![Figure 2. Digital photo of GO-PFcMAss in different organic solvents and water.](image)

![Figure 3. FT-IR spectra of GO, PFcMAss and GO-PFcMAss.](image)
All these evidences demonstrated that PFcMAss chains were attached on the surface of AuNPs. 

\[ \text{Au(III)} \text{ of AuCl}_4^{-} \text{ crystal planes of AuNPs (Figure 3B), consistent with XRD results.}\]

To prepare AuNPs by Brust-Schiffrin protocol with some modifications as outlined in Scheme S1. After preparing thiolate-protected gold nanoparticles using thiol-based ligands with extraordinary stability, the core size of 5.1 nm exhibits five concentric diffraction rings from inside to outside corresponding to the Scherrer formula. A selected area electron diffraction (SAED) pattern of obtained AuNPs with a uniform in size with an average diameter of 4.7 nm, which is consistent with the result calculated from Scherrer formula. Based on XRD results, the average diameter of AuNPs was estimated to be about 4.5 nm according to Scherrer factor, respectively.

\[ hkl = k\lambda / \beta \cos \theta, \text{ where } L_{hkl}, \lambda, \beta, \theta, \text{ and } k \text{ are the size of microcrystals perpendicular to face (hkl), wavenumber of X-ray, Bragg angle, diffraction line width and Scherrer factor, respectively}.\]

As shown in TEM image of PFcMAss-AuNPs (Figure 6A), the obtained AuNPs are spherical and uniform in size with an average diameter of 4.7 nm, which is consistent with the result calculated from the Scherrer formula. A selected area electron diffraction (SAED) pattern of obtained AuNPs with a core size of 5.1 nm exhibits five concentric diffraction rings from inside to outside corresponding to crystal planes of AuNPs (Figure 3B), consistent with XRD results. In addition, the spacing value was found to be 0.238 nm from HRTEM image (Figure 3C), in good agreement with interplanar spacing of Au (111) [65]. Since TEM measurement generally only shows interactions, the surface of AuNPs would be coated by PFcMAss via ligand exchange since Au-S bond was stronger than the interaction between Au and N of tetraoctylammonium bromide (TOAB).

The as-prepared PFcMAss-AuNPs can be dispersed in chloroform, toluene, THF and ethanol to give transparent colloidal solution with a wine-red color. These solutions still remained stable over several months. On the contrary, purple aggregates of gold particles were formed in a control experiment, where PFcMAss homopolymer was not added (Figure S2). From 1H-NMR spectrum of PFcMAss-AuNPs powder obtained by centrifugation (Figure S3), one can notice the typical signals at 4.22, 4.41 and 4.82 ppm attributed to Fc group and peaks at about 2.85 and 3.12 ppm originating from the protons in the methylene spacer group. Additionally, the characteristic signals originating from Fc units (2958, 1459 and 817 cm\(^{-1}\)) and methylene group (2923 and 2852 cm\(^{-1}\)) of PFcMAss repeat unit were found in FT-IR spectrum (Figure S4) of the obtained PFcMAss-AuNPs powder. All these evidences demonstrated that PFcMAss chains were attached on the surface of AuNPs.

3.3. Fabrication of GO-PFcMAss-AuNPs Nanocomposite

Brust-Schiffrin method has been considered to be one of the most attractive approaches for preparing thiolate-protected gold nanoparticles using thiol-based ligands with extraordinary stability and potentials for further surface functionalization [49]. We then tested whether we could use PFcMAss with disulfide groups, instead of thiol-containing species, in the preparation of AuNPs via Brust-Schiffrin protocol [49]. In the present work, we also employed PFcMAss as stabilizer to prepare AuNPs by Brust-Schiffrin protocol with some modifications as outlined in Scheme S1. After Au(III) of AuCl\(_4^{-}\) ions were reduced to Au(0) by NaBH\(_4\), the organic phase was firstly separated before the addition of PFcMAss so as to avoid the decomposition of PFcMAss caused by NaBH\(_4\) in aqueous phase. It is supposed that the surface of AuNPs would be coated by PFcMAss via ligand exchange since Au-S bond was stronger than the interaction between Au and N of tetraoctylammonium bromide (TOAB).

The as-prepared PFcMAss-AuNPs can be dispersed in chloroform, toluene, THF and ethanol to give transparent colloidal solution with a wine-red color. These solutions still remained stable over several months. On the contrary, purple aggregates of gold particles were formed in a control experiment, where PFcMAss homopolymer was not added (Figure S2). From 1H-NMR spectrum of PFcMAss-AuNPs powder obtained by centrifugation (Figure S3), one can notice the typical signals at 4.22, 4.41 and 4.82 ppm attributed to Fc group and peaks at about 2.85 and 3.12 ppm originating from the protons in the methylene spacer group. Additionally, the characteristic signals originating from Fc units (2958, 1459 and 817 cm\(^{-1}\)) and methylene group (2923 and 2852 cm\(^{-1}\)) of PFcMAss repeat unit were found in FT-IR spectrum (Figure S4) of the obtained PFcMAss-AuNPs powder. All these evidences demonstrated that PFcMAss chains were attached on the surface of AuNPs.

From XRD pattern of PFcMAss-AuNPs powder obtained by centrifugation (Figure 5A), one can notice five diffraction peaks at scattering angles of 38.4°, 44.7°, 65.1°, 77.8° and 81.7°, corresponding to (111), (200), (220), (311) and (222) crystal planes of the face center cubic structure of AuNPs [46]. Based on XRD results, the average diameter of AuNPs was estimated to be about 4.5 nm according to Scherrer formula (\(L_{hkl} = \frac{k\lambda}{\beta \cos \theta}\), where \(L_{hkl}\), \(\lambda\), \(\beta\), \(\theta\), and \(k\) are the size of microcrystals perpendicular to face (hkl), wavenumber of X-ray, Bragg angle, diffraction line width and Scherrer factor, respectively).

As shown in TEM image of PFcMAss-AuNPs (Figure 6A), the obtained AuNPs are spherical and uniform in size with an average diameter of 4.7 nm, which is consistent with the result calculated from the Scherrer formula. A selected area electron diffraction (SAED) pattern of obtained AuNPs with a core size of 5.1 nm exhibits five concentric diffraction rings from inside to outside corresponding to (111), (200), (220), (311) and (222) crystal planes of AuNPs (Figure 3B), consistent with XRD results. In addition, the spacing value was found to be 0.238 nm from HRTEM image (Figure 3C), in good agreement with interplanar spacing of Au (111) [65]. Since TEM measurement generally only shows interactions, the surface of AuNPs would be coated by PFcMAss via ligand exchange since Au-S bond was stronger than the interaction between Au and N of tetraoctylammonium bromide (TOAB).

The as-prepared PFcMAss-AuNPs can be dispersed in chloroform, toluene, THF and ethanol to give transparent colloidal solution with a wine-red color. These solutions still remained stable over several months. On the contrary, purple aggregates of gold particles were formed in a control experiment, where PFcMAss homopolymer was not added (Figure S2). From 1H-NMR spectrum of PFcMAss-AuNPs powder obtained by centrifugation (Figure S3), one can notice the typical signals at 4.22, 4.41 and 4.82 ppm attributed to Fc group and peaks at about 2.85 and 3.12 ppm originating from the protons in the methylene spacer group. Additionally, the characteristic signals originating from Fc units (2958, 1459 and 817 cm\(^{-1}\)) and methylene group (2923 and 2852 cm\(^{-1}\)) of PFcMAss repeat unit were found in FT-IR spectrum (Figure S4) of the obtained PFcMAss-AuNPs powder. All these evidences demonstrated that PFcMAss chains were attached on the surface of AuNPs.

From XRD pattern of PFcMAss-AuNPs powder obtained by centrifugation (Figure 5A), one can notice five diffraction peaks at scattering angles of 38.4°, 44.7°, 65.1°, 77.8° and 81.7°, corresponding to (111), (200), (220), (311) and (222) crystal planes of the face center cubic structure of AuNPs [46]. Based on XRD results, the average diameter of AuNPs was estimated to be about 4.5 nm according to Scherrer formula (\(L_{hkl} = \frac{k\lambda}{\beta \cos \theta}\), where \(L_{hkl}\), \(\lambda\), \(\beta\), \(\theta\), and \(k\) are the size of microcrystals perpendicular to face (hkl), wavenumber of X-ray, Bragg angle, diffraction line width and Scherrer factor, respectively).

As shown in TEM image of PFcMAss-AuNPs (Figure 6A), the obtained AuNPs are spherical and uniform in size with an average diameter of 4.7 nm, which is consistent with the result calculated from the Scherrer formula. A selected area electron diffraction (SAED) pattern of obtained AuNPs with a core size of 5.1 nm exhibits five concentric diffraction rings from inside to outside corresponding to (111), (200), (220), (311) and (222) crystal planes of AuNPs (Figure 3B), consistent with XRD results. In addition, the spacing value was found to be 0.238 nm from HRTEM image (Figure 3C), in good agreement with interplanar spacing of Au (111) [65]. Since TEM measurement generally only shows interactions, the surface of AuNPs would be coated by PFcMAss via ligand exchange since Au-S bond was stronger than the interaction between Au and N of tetraoctylammonium bromide (TOAB).
the morphology of Au nanoparticles, and the organic or polymeric ligands on the surface of gold nanoparticles are not able to be visualized due to their much lower electron density than that of gold nanoparticles [66]. However, both gold nanoparticle and ligand can be detected by AFM since AFM measurement mainly reflects the height of sample, regardless of electron density of the object. It can be found that the average diameter of spherical particles in AFM image was about 16 nm (Figure S5), much larger than the size obtained by TEM (4.7 nm). These results further made sure that AuNPs were coated with PFcMAss polymers. Thus, it can be concluded that PFcMAss homopolymer could serve as excellent stabilizer for the preparation of AuNPs with high quality by the optimized Brust-Schiffrin method.

Figure 5. XRD pattern of PFcMAss-functionalized AuNPs (A) and GO-PFcMAss-AuNPs (B).

Figure 6. (A) TEM image, (B) selected area electron diffraction pattern and (C) HRTEM image and of PFcMAss-AuNPs.

On the basis of these results, we then attempted to functionalize GO-PFcMAss with AuNPs through depositing AuNPs onto the surface of GO-PFcMAss in situ via the modified Brust-Schiffrin strategy using GO-PFcMAss as stabilizer instead of PFcMAss homopolymer. Figure 5B displays XRD pattern of GO-PFcMAss decorated with AuNPs (GO-PFcMAss-AuNPs nanocomposites). Five obvious diffraction peaks appeared at scattering angles of 38.4°, 44.7°, 65.1°, 77.8° and 81.7°, corresponding to (111), (200), (220), (311) and (222) crystal planes, respectively, which is indicative of the presence of AuNPs in the as-prepared nanocomposites. The average diameter of formed AuNPs was calculated to be about 3.8 nm on the basis of XRD results and Scherrer formula. In the XPS spectrum of GO-PFcMAss-AuNPs nanocomposite (Figure 4A), the Fe3p3 peak at a binding energy value of 71 eV was observed, suggesting the presence of ferrocene unit on GO sheets. It can be seen that the C1s peak attributed to GO appeared at a binding energy value of 285.7 eV. The appearing of Au 4f doublet (86 and 90 eV, in Figure 4B) was consistent with the Au⁰ state, revealing the presence of AuNPs on graphene sheets. Furthermore, the element contents of TRIS-GO-Ini, GO-PFcMAss and GO-PFcMAss-AuNPs based on XPS results were obtained as outline in Table S1, indicating the existence of C, N, O, Fe, S and Au elements in GO-PFcMAss-AuNPs. According to previous literatures [67,68], there are two sulfur
species in organic layer absorbed on the surface of AuNPs. The weight content of Au in the obtained GO-PFcMAss-AuNPs nanocomposite was estimated to be 1.1 wt% based on the elemental composition of GO-PFcMAss-AuNPs (Table S1). In addition, the contents of Fe and S decreased from 1.01 mol% and 1.60 mol% to 0.04 mol% and 0.38 mol%, respectively, after the introduction of AuNPs. The decrease of contents of Fe and S, especially the decrease of ratio of content of Fe to content of S, indicated that part of disulfide groups might be broken during the formation of AuNPs to give FcCH₂CH₂S-moieties and part of produced FcCH₂CH₂S-species were lost during the preparation of GO-PFcMAss-AuNPs nanocomposite. However, there was about 1.0 wt% of Fe units in GO-PFcMAss-AuNPs nanocomposite.

From TEM image (Figure 7A), one can notice that there were a large number of AuNPs located on the surface of GO and almost no free AuNPs can be found. These observations indicated that PFcMAss of GO-PFcMAss had a high affinity to capture any free AuNPs formed in the solution via strong S-Au interactions and PFcMAss domains on the surface of GO served as excellent nucleation sites for the growth of AuNPs in situ. The size of formed AuNPs was about 4.1 nm, which was close to the value estimated from XRD result (3.8 nm). The spacing value of one gold nanoparticle was estimated to be about 0.235 nm (Figure 7C), in good agreement with interplanar spacing of Au (111) [65], which further verified the formation of AuNPs on the surface of GO-PFcMAss. Therefore, all these results indicated that GO-PFcMAss-AuNPs nanocomposite can be prepared by using GO-PFcMAss as stabilizer, instead of organic ligand, via modified Brust-Schiffrin method.

![Figure 7. (A) TEM, (B) and (C) HRTEM images of GO-PFcMAss-AuNPs nanocomposite.](image)

**Figure 7.** (A) TEM, (B) and (C) HRTEM images of GO-PFcMAss-AuNPs nanocomposite.

### 4. Conclusions

In summary, we developed a facile and versatile strategy to prepare multi-functional GO-based nanocomposites containing AuNPs and Fc groups. A new methacrylate monomer of FcMAss containing both disulfide and Fc groups was designed and synthesized. This monomer can be polymerized by ATRP using CuBr/PMDETA as catalytic system and EBiB as initiator at 50 °C. PFcMAss polymer chains were then efficiently introduced onto the surface of GO by SI-ATRP using Br-containing GO as macroinitiator. By taking advantage of strong coordinating ability of disulfide moieties toward AuNPs, a large number of AuNPs with an average diameter of about 4.1 nm were formed in situ on the surface of PFcMAss-decorated GO (GO-PFcMAss) via modified Brust-Schiffrin strategy to give multifunctional GO-PFcMAss-AuNPs nanocomposite consisting of GO, AuNPs and Fc groups. The usage of disulfide-containing polymers, instead of commonly used thiol-based compounds, not only endowed GO-PFcMAss-AuNPs nanocomposites with excellent dispersibility and colloidal stability, but also facilitated the introduction of much more AuNPs onto the surface of GO due to a high local density of disulfide groups. The polymeric chains on the surface of GO endowed GO-PFcMAss-AuNPs nanocomposite with excellent colloidal stability. Additionally, the disulfide group of PFcMAss provided potential sites to efficiently incorporate additional functionalities by easily modifying structure of disulfide-based monomer. Significantly, given attractive properties and broad applications of GO-, AuNPs- and Fc-based materials, this kind of nanocomposite undoubtedly shows promising applications in the fabrication of multi-functional materials.
Supplementary Materials: The following are available online at http://www.mdpi.com/2079-4991/9/2/310/s1, Figure S1: GPC curves of PFcMAss homopolymer and free PFcMAss polymer during surface-initiated ATRP, Table S1: Elemental analysis of TRIS-GO-Ini, GO-PFcMAss, and GO-PFeMAss-AuNPs obtained from XPS, Figure S2: Photos of solutions of AuNPs prepared by Brust-Schiffrin protocol with (A) and without (B) the addition of TRIS-GO-PFcMAss, Figure S3: 1H NMR spectra of PFcMAss and PFcMAss-AuNPs in CDCl3 and the photo (inset) of PFcMAss in toluene (0.1 mg/mL), Figure S4: FT-IR spectra of PFcMAss and PFcMAss-AuNPs, Figure S5: Tapping mode AFM images with section analysis of PFcMAss-functionalized AuNPs.

Author Contributions: Conceptualization, W.H.Q. and X.Y.H.; methodology, W.H.Q. and X.Y.H.; software, T.S. and G.L.L.; validation, M.Y., H.Y.Z. and C.F.; formal analysis, W.H.Q.; investigation, W.H.Q. and X.Y.H.; visualization, W.H.Q.; supervision, W.H.Q. and X.Y.H.; project administration, W.H.Q. and X.Y.H.; funding acquisition, W.H.Q. and X.Y.H.

Funding: This research was funded by National Science Foundation for Distinguished Young Scholars (51825304), National Natural Science Foundation of China (21632009 and 21674124), Strategic Priority Research Program of CAS (XDB20000000), Shanghai Scientific and Technological Innovation Project (16JC1402500, 17ZR1426700, 17DZ1205402, 18JC1415500 and 18ZR1448600), Scientific Research Project of Science and Technology Commission of Xuhui Municipality (SHXH201613) and Scientific Research Project of Xuhui Provincial Commission of Health and Family Planning (SHXH201706).

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Novoselov, K.S.; Geim, A.K.; Morozov, S.V.; Jiang, D.; Zhang, Y.; Dubonos, S.V.; Grigorieva, I.V.; Firsov, A.A. Electric field in atomically thin carbon films. Science 2004, 306, 666–669. [CrossRef] [PubMed]
2. Zhang, Y.B.; Tan, Y.W.; Stormer, H.L.; Kim, P. Experimental observation of the quantum Hall effect and Berry’s phase in graphene. Nature 2005, 438, 201–204. [CrossRef] [PubMed]
3. Geim, A.K.; Novoselov, K.S. The rise of graphene. Nat. Mater. 2007, 6, 183–191. [CrossRef] [PubMed]
4. Lee, C.; Wei, X.D.; Kysar, J.W.; Hone, J. Measurement of the elastic properties and intrinsic strength of monolayer graphene. Science 2008, 321, 385–388. [CrossRef] [PubMed]
5. Geim, A.K. Graphene: Status and prospects. Science 2009, 324, 1530–1534. [CrossRef] [PubMed]
6. Zhang, L.S.; Viola, W.; Andrew, T.L. High energy density, super-deformable, garment-integrated micro-supercapacitors for powering wearable electronics. ACS Appl. Mater. Interfaces 2018, 10, 36834–36840. [CrossRef] [PubMed]
7. Stankovich, S.; Dikin, D.A.; Piner, R.D.; Kohlhaas, K.A.; Kleinhammes, A.; Jia, Y.; Wu, Y.; Nguyen, S.T.; Ruoff, R.S. Synthesis of graphene-based nanosheets via chemical reduction of exfoliated graphite oxide. Carbon 2007, 45, 1558–1565. [CrossRef]
8. Shin, H.J.; Kim, K.K.; Benayad, A.; Yoon, S.; Park, H.K.; Jung, I.; Jin, M.H.; Jeong, H.; Kim, J.M.; Choi, J.; et al. Efficient reduction of graphite oxide by sodium borohydride and its effect on electrical conductance. Adv. Funct. Mater. 2009, 19, 1987–1992. [CrossRef]
9. Moon, I.K.; Lee, J.; Ruoff, R.S.; Lee, H. Reduced graphene oxide by chemical graphitization. Nat. Commun. 2010, 1, 1–6. [CrossRef] [PubMed]
10. Dong, L.; Yang, J.; Chhowalla, M.; Loh, K.P. Synthesis and reduction of large sized graphene oxide sheets. Chem. Soc. Rev. 2017, 46, 7306–7316. [CrossRef] [PubMed]
11. Loh, K.P.; Bao, Q.; Ang, P.K.; Yang, J.X. The chemistry of graphene. J. Mater. Chem. 2010, 20, 2277–2289. [CrossRef]
12. Pan, Y.Z.; Bao, H.Q.; Sahoo, N.G.; Wu, T.F.; Li, L. Water-soluble poly(N-isopropylacrylamide)-graphene sheets synthesized via click chemistry for drug delivery. Adv. Funct. Mater. 2011, 21, 2754–2763. [CrossRef] [PubMed]
13. Huang, X.; Qi, X.; Boey, F.; Zhang, H. Graphene-based composites. Chem. Soc. Rev. 2012, 41, 666–686. [CrossRef] [PubMed]
14. Zhang, B.; Zhang, Y.; Peng, C.; Yu, M.; Li, L.; Deng, B.; Hu, P.; Fan, C.H.; Li, J.; Huang, Q. Preparation of polymer decorated graphene oxide by γ-ray induced graft polymerization. Nanoscale 2012, 4, 1742–1748. [CrossRef] [PubMed]
15. Kuila, T.; Bose, S.; Mishra, A.K.; Khanna, P.; Kim, N.H.; Lee, J.H. Chemical functionalization of graphene and its applications. Prog. Mater. Sci. 2012, 57, 1061–1105. [CrossRef]
16. Tran, T.H.; Nguyen, H.T.; Pham, T.T.; Choi, J.Y.; Choi, H.G.; Yong, C.S.; Kim, J.O. Development of a graphene oxide nanocarrier for dual-drug chemo-phototherapy to overcome drug resistance in cancer. ACS Appl. Mater. Interfaces 2015, 7, 28647–28655. [CrossRef] [PubMed]

17. Nia, A.S.; Binder, W.H. Graphene as initiator/catalyst in polymerization chemistry. Prog. Polym. Sci. 2017, 67, 48–76.

18. Liu, Z.; Robinson, J.T.; Sun, X.; Dai, H.J. PEGylated nanographene oxide for delivery of water-insoluble cancer drugs. J. Am. Chem. Soc. 2008, 130, 10876–10877. [CrossRef] [PubMed]

19. Zhang, X.; Yin, J.; Peng, C.; Hu, W.; Zhu, Z.; Li, W.; Fan, C.H.; Huang, Q. Distribution and biocompatibility studies of graphene oxide in mice after intravenous administration. Carbon 2011, 49, 986–995. [CrossRef]

20. Bitounis, D.; Ali-Boucetta, H.; Hong, B.H.; Min, D.-H.; Kostarelos, K. Prospects and challenges of graphene in biomedical applications. Adv. Mater. 2013, 25, 2258–2268. [CrossRef] [PubMed]

21. Georgakilas, V.; Tiwari, J.N.; Kemp, K.C.; Perman, J.A.; Bourlinos, A.B.; Kim, K.S.; Zboril, R. Noncovalent functionalization of graphene and graphene oxide for energy materials, biosensing, catalytic, and biomedical applications. Chem. Rev. 2016, 116, 5464–5519. [CrossRef] [PubMed]

22. Pu, J.; Li, J.C.; Shen, Z.H.; Zhong, C.L.; Liu, J.Y.; Ma, H.X.; Zhu, J.; Zhang, H.G.; Braun, P.V. Interlayer lithium plating in Au nanoparticles pillared reduced graphene oxide for lithium metal anodes. Adv. Funct. Mater. 2018, 48, 1804133. [CrossRef]

23. Liu, S.; Cao, S.T.; Guo, J.Y.; Luo, L.Q.; Zhou, Y.; Lin, C.L.; Shi, J.Y.; Fan, C.H.; Lv, M.; Wang, L.H. Graphene oxide-silver nanocomposites modulate biofilm formation and extracellular polymeric substance (EPS) production. Nanoscale 2018, 10, 19603–19611. [CrossRef] [PubMed]

24. Alivisatos, P. The use of nanocrystals in biological detection. Nat. Biotechnol. 2004, 22, 47–52. [CrossRef] [PubMed]

25. Daniel, M.C.; Astruc, D. Gold nanoparticles: Assembly, supramolecular chemistry, quantum-size-related properties, and applications toward biology, catalysis, and nanotechnology. Chem. Rev. 2004, 104, 293–346. [CrossRef] [PubMed]

26. Okazaki, K.; Ichikawa, S.; Maeda, Y.; Haruta, M.; Kohyama, M. Electronic structures of Au supported on TiO2. Appl. Catal. A 2005, 291, 45–54. [CrossRef] [PubMed]

27. De, M.; Ghosh, P.S.; Rotello, V.M. Applications of nanoparticles in biology. Adv. Mater. 2008, 20, 4225–4241. [CrossRef]

28. Fujitani, T.; Nakamura, I.; Akita, T.; Okumura, M.; Haruta, M. Hydrogen dissociation by gold cluster. Angew. Chem. Int. Ed. 2009, 48, 9515–9518. [CrossRef] [PubMed]

29. Giljohann, D.A.; Seferos, D.S.; Daniel, W.L.; Massich, M.D.; Patel, P.C.; Mirkin, C.A. Gold nanoparticles for biology and medicine. Angew. Chem. Int. Ed. 2010, 49, 3280–3294. [CrossRef] [PubMed]

30. Dykman, I.; Khlebtsov, N. Gold nanoparticles in biomedical applications: Recent advances and perspectives. Chem. Soc. Rev. 2012, 41, 2256–2282. [CrossRef] [PubMed]

31. Lyu, Z.L.; Wang, H.W.; Wang, Y.Y.; Ding, K.G.; Liu, H.; Yuan, L.; Shi, X.J.; Wang, M.M.; Wang, Y.W.; Chen, H. Maintenance of the pluripotency of mouse embryonic stem cells on gold nanoparticle layers with nanoscale but not microscale surface roughness. Nanoscale 2014, 6, 6959–6969. [CrossRef] [PubMed]

32. Jasuja, K.; Berry, V. Implantation and growth of dendritic gold nanostructures on graphene derivatives: Electrical property tailoring and Raman enhancement. ACS Nano 2009, 3, 2358–2366. [CrossRef] [PubMed]

33. Huang, J.; Zhang, L.; Chen, B.; Ji, N.; Chen, F.; Zhang, Y.; Zhang, Z. Nanocomposites of size-controlled gold nanoparticles and graphene oxide: Formation and applications in SERS and catalysis. Nanoscale 2010, 2, 2733–2738. [CrossRef] [PubMed]

34. Zhu, Z.; Ma, L.; Su, M.; Liu, D.; Wang, Z. Preparation and application of thionin-bridged graphene-gold nanoparticle nanohybrids. J. Mater. Chem. B 2013, 1, 1432–1438. [CrossRef]

35. Han, S.T.; Zhou, Y.; Wang, C.; He, L.; Zhang, W.; Roy, V.A.L. Layer-by-layer-assembled reduced graphene oxide/gold nanoparticle hybrid double-floating-gate structure for low-voltage flexible flash memory. Adv. Mater. 2013, 25, 872–877. [CrossRef] [PubMed]

36. Govindhan, M.; Amiri, M.; Chen, A. Au nanoparticle/graphene nanocomposite as a platform for the sensitive detection of NADH in human urine. Biosens. Bioelectron. 2015, 66, 474–480. [CrossRef] [PubMed]
37. Kumarasamy, J.; Camarada, M.B.; Venkatraman, D.; Ju, H.X.; Dey, R.S.; Wen, Y.P. One-step coelectrodeposition-assisted layer-by-layer assembly of gold nanoparticles and reduced graphene oxide and its self-healing three-dimensional nanohybrid for an ultrasensitive DNA sensor. *Nanoscale* **2018**, *10*, 1196–1206. [CrossRef] [PubMed]

38. Ensafi, A.A.; Akbarian, F.; Heydari-Soureshjani, E.; Rezaei, B. A novel aptasensor based on 3D-reduced graphene oxide modified gold nanoparticles for determination of arsenite. *Biosens. Bioelectron.* **2018**, *10*, 1196–1206. [CrossRef] [PubMed]

39. Goncalves, G.; Marques, P.A.P.; Granadeiro, C.M.; Nogueira, H.I.S.; Singh, M.K.; Gracio, J. Surface modification of graphene nanosheets with gold nanoparticles: Theroile of oxygen moieties at graphene surface on gold nucleation and growth. *Chem. Mater.* **2009**, *21*, 4796–4802. [CrossRef]

40. Ismaili, H.; Geng, D.; Sun, A.X.; Kantzas, T.T.; Workentin, M.S. Light-activated covalent formation of gold nanoparticle-graphene and gold nanoparticle-glass composites. *Langmuir* **2011**, *27*, 13261–13268. [CrossRef] [PubMed]

41. Pham, T.A.; Choi, B.C.; Lim, K.T.; Jeong, Y.T. A simple approach for immobilization of gold nanoparticles on graphene oxide sheets by covalent bonding. *Appl. Surf. Sci.* **2011**, *257*, 3350–3357. [CrossRef]

42. Leea, J.U.; Leea, W.; Yoona, S.S.; Kimb, J.; Byun, J.H. Site-selective immobilization of gold nanoparticles on graphene sheets and its electrochemical properties. *Appl. Surf. Sci.* **2014**, *315*, 73–80. [CrossRef]

43. Liu, G.; Qi, M.; Zhang, Y.; Cao, C.; Goldys, E.M. Nanocomposites of gold nanoparticles and graphene oxide towards an stable label-free electrochemical immunosensor for detection of cardiac marker troponin-I. *Anal. Chim. Acta* **2016**, *909*, 1–8. [CrossRef] [PubMed]

44. Garcia, A.; Peniche-Covas, C.; Chico, B.; Simpson, B.K.; Villalonga, R. Ferrocene branched chitosan for the construction of a reagentless amperometric hydrogen peroxide biosensor. *Macromol. Biosci.* **2007**, *7*, 435–439. [CrossRef] [PubMed]

45. Bai, Z.H.; Li, G.Y.; Liang, J.T.; Su, J.; Zhang, Y.; Chen, H.Z.; Huang, Y.; Sui, W.G.; Zhao, Y.X. Non-enzymatic electrochemical biosensor based on Pt NPs/RGO-Cs-Fc nano-hybrids for the detection of hydrogen peroxide in living cells. *Biosens. Bioelectron.* **2016**, *82*, 185–194. [CrossRef] [PubMed]

46. Li, Y.; Li, Y.; Wu, Y.; Lu, F.; Chen, Y.; Gao, W. An electrochemiluminescence biosensor for endonuclease EcoRI detection. *Biosens. Bioelectron.* **2017**, *89*, 585–591. [CrossRef] [PubMed]

47. Deng, Y.; Liu, Y.J.; Dai, J.; Lang, M.D.; Huang, X.Y. Functionalization of graphene oxide towards thermo-sensitive nanocomposites via moderate in situ SET-LRP. *J. Polym. Sci. Polym. Chem.* **2011**, *49*, 4747–4755. [CrossRef]

48. Deng, Y.; Zhang, J.Z.; Li, Y.J.; Hu, J.H.; Yang, D.; Huang, X.Y. Thermoresponsive graphene oxide-PNIPAM nanocomposites with controllable grafting polymer chains via moderate in situ SET-LRP. *J. Polym. Sci. Polym. Chem.* **2012**, *50*, 4451–4458. [CrossRef]

49. Brust, M.; Fink, J.; Bethell, D.; Schiffrin, D.J.; Kiely, C. Synthesis and reactions of functionalised gold nanoparticles. *J. Chem. Soc. Chem. Commun.* **1995**, *1655–1656*. [CrossRef]

50. Oh, E.; Susumu, K.; Makinen, A.J.; Deschamps, J.R.; Huston, A.L.; Medintz, I.L. Colloidal stability of gold nanoparticles coated with multithiol-poly(ethylene glycol) ligands: Importance of structural constraints of the sulfur anchoring group. *J. Phys. Chem. C* **2013**, *117*, 18947–18956. [CrossRef]

51. Mei, B.C.; Oh, E.; Susumu, K.; Farrell, D.; Mountzias, T.J.; Mattoussi, H. Effects of ligand coordination number and surface curvature on the stability of gold nanoparticles in aqueous solutionss. *Langmuir* **2009**, *25*, 10604–10611. [CrossRef] [PubMed]

52. Zaluzhna, O.; Li, Y.; Allison, T.C.; Tong, Y.J. Inverse-micelle-encapsulated water-enabled bond breaking of dialkyl diselenide/disulfide: A critical step for synthesizing high-quality gold nanoparticles. *J. Am. Chem. Soc.* **2012**, *134*, 17991–17996. [CrossRef] [PubMed]

53. Vukicevic, R.; Beuermann, S. Fullerenes decorated with poly(vinylidene fluoride). *Macromolecules* **2011**, *44*, 2597–2603. [CrossRef]

54. Megiel, E. Surface modification using TEMPO and its derivatives. *Adv. Colloid Inter. Sci.* **2017**, *250*, 158–184. [CrossRef] [PubMed]
56. Sadegh, H.; Ali, G.A.M.; Gupta, V.K.; Makhlouf, A.S.H.; Shahryari-ghoshekandi, R.; Nadagouda, M.N.; Sillanpää, M.; Megiel, E. The role of nanomaterials as effective adsorbents and their applications in waste water treatment. *J. Nanostruct. Chem.* 2017, 7, 1–14. [CrossRef]  
57. Sangermano, M.; Rodriguez, D.; Gonzalez, M.C.; Laurenti, E.; Yagci, Y. Visible light induced cationic polymerization of epoxides by using multiwalled carbon nanotubes. *Macromol. Rapid Commun.* 2018, 39, 1800250. [CrossRef] [PubMed]  
58. Jiang, X.; Lu, G.L.; Huang, X.Y.; Li, Y.; Cao, F.Q.; Chen, H.; Liu, W.B. Thermo-responsive graphene oxide/poly(ethyl ethylene phosphate) nanocomposite via ring opening polymerization. *Nanomaterials* 2019, 9, 207. [CrossRef] [PubMed]  
59. Duan, H.W.; Kuang, M.; Wang, D.Y.; Kurth, D.G.; Mohwald, H. Colloidal amphiphilic nanocrystals derived from poly(2-(dimethylamino)ethyl methacrylate) capping. *Angew. Chem. Int. Ed.* 2005, 44, 1717–1720. [CrossRef]  
60. Yan, J.J.; Pan, X.C.; Wang, Z.Y.; Zhang, J.; Matyjaszewski, K. Influence of spacers in tetherable initiators on surface-initiated atom transfer radical polymerization (SI-ATRP). *Macromolecules* 2016, 49, 9283–9286. [CrossRef]  
61. Matyjaszewski, K. Advanced materials by atom transfer radical polymerization. *Adv. Mater.* 2018, 30, 1706441. [CrossRef] [PubMed]  
62. Ejaz, M.; Tsuji, Y.; Fukuda, T. Controlled grafting of a well-defined polymer on a porous glass filter by surface-initiated atom transfer radical polymerization. *Polymer* 2001, 42, 6811–6815. [CrossRef]  
63. Husseman, M.; Malmstroem, E.E.; McNamara, M.; Mate, M.; Mecerreyes, D.; Benoit, D.G.; Hedrick, J.L.; Mansky, P.; Huang, E.; Russell, T.P.; et al. Controlled synthesis of polymer brushes by "living" free radical polymerization techniques. *Macromolecules* 1999, 32, 1424–1431. [CrossRef]  
64. Fang, M.; Wang, K.; Lu, H.; Yang, Y.; Nutt, S. Single-layer graphene nanosheets with controlled grafting of polymer chains. *J. Mater. Chem.* 2010, 20, 1982–1992. [CrossRef]  
65. Xu, P.; Yu, H.; Li, X. In situ growth of noble metal nanoparticles on graphene oxide sheets and direct construction of functionalized porous-layered structure on gravimetric microsensors for chemical detection. *Chem. Commun.* 2012, 48, 10784–10786. [CrossRef] [PubMed]  
66. Feng, C.; Gu, L.N.; Yang, D.; Hu, J.H.; Lu, G.L.; Huang, X.Y. Size-controllable gold nanoparticles stabilized by PDEAEMA-based double hydrophilic graft copolymer. *Polymer* 2009, 50, 3990–3996. [CrossRef]  
67. Kaim, A.; Szydłowska, J.; Piotrowski, P.; Megiel, E. One-pot synthesis of gold nanoparticles densely coated with nitrooxide spins. *Polyhedron* 2012, 46, 119–123. [CrossRef]  
68. Gozdzielowska, M.; Cichowicz, G.; Markowska, K.; Zawada, K.; Megiel, E. Nitrooxide-coated silver nanoparticles: Synthesis, surface physicochemistry and antibacterial activity. *RSC Adv.* 2015, 5, 58403–58415. [CrossRef]