Ubiquitin–proteasome system (UPS) as a target for anticancer treatment

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Abstract The ubiquitin–proteasome system (UPS) plays an important role in the cellular processes for protein quality control and homeostasis. Dysregulation of the UPS has been implicated in numerous diseases, including cancer. Indeed, components of UPS are frequently mutated or abnormally expressed in various cancers. Since Bortezomib, a proteasome inhibitor, received FDA approval for the treatment of multiple myeloma and mantle cell lymphoma, increasing numbers of researchers have been seeking drugs targeting the UPS as a cancer therapeutic strategy. Here, we introduce the essential component of UPS, including ubiquitinating enzymes, deubiquitinating enzymes and 26S proteasome, and we summarize their targets and mechanisms that are crucial for tumorigenesis. In addition, we briefly discuss some UPS inhibitors, which are currently in clinical trials as cancer therapeutics.

Keywords Ubiquitin–proteasome system (UPS) · E3 ligase · Deubiquitinating enzymes (DUBs) · Proteasome · Cancer · Small molecule inhibitors

Introduction

The ubiquitin–proteasome system (UPS) is the major proteolytic system that controls protein degradation and it regulates many cellular processes in eukaryotic cells, such as DNA repair, stress responses and cell proliferation. The UPS consists of specific enzymes that modify protein substrates using ubiquitin, and 26S proteasomes responsible for proteolysis of ubiquitin-tagging substrates (Mata-Cantero et al. 2015). This ubiquitin conjugation to the substrate is carried out by a multistep cascade reaction consisting of the E1, E2, and E3 enzymes. In brief, ubiquitin-activating enzymes (E1s) use energy from ATP hydrolysis to generate thioester bond between the C-terminal of ubiquitin and a Cys residue in the active catalytic site of E1 enzymes. This activated ubiquitin is then transferred to the ubiquitin-conjugating enzymes (E2s), which forms a thioester bond between E2 enzymes and the ubiquitin. Finally, the charged E2 enzymes cooperate with one of hundreds of ubiquitin ligases (E3s) to transfer the activated ubiquitin to a target substrate (Fig. 1) (Eldridge and O’Brien 2010; Yuan et al. 2018).

E1, E2, E3 ubiquitinating enzymes

The human genome contains two E1 genes, UBA1 and UBA6, about fifty E2 genes and genes encoding around seven hundred E3 enzymes. These E3 enzymes are generally classified into three groups based on the mechanism that they use to transfer ubiquitin from the E2 enzyme to the substrate: the ‘really interesting new gene’ (RING) class, the ‘homologous to E6-AP carboxyl terminus’ (HECT) class, and the ‘RING-between-RING’ (RBR) class (Gupta et al. 2018; Zhang et al. 2020). The RING class, the largest class of E3 enzymes, helps transfer the ubiquitin attached to E2 directly to the substrate without forming thioester bonds with ubiquitin (Lorick et al. 1999; Mani and Gelmann 2005). The HECT class forms a catalytic Cys-dependent intermediate with ubiquitin attached to E2, and then transfers it to a target substrate (Huibregtse...
Ubiquitin–proteasome system (UPS) as a target for anticancer treatment

et al. 1995; Scheffner et al. 1995). The RBR class has two canonical RING domains (RING1 and RING2) linking an ‘in-between RING’ (IBR) domain (Wenzel et al. 2011). The RING1 domain initially recognizes the ubiquitin-attached E2 (RING-like), but then the RING2 domain offers a Cys residue to the active site of E2 (HECT-like) which forms a thioester intermediate with the ubiquitin attached to the E2 (Gupta et al. 2018).

Ubiquitin contains seven Lys residues including Lys6, 11, 27, 29, 33, 48, and 63, and all of them can covalently attach to other ubiquitins; thus, various linear or branched ubiquitin chains are formed (Pohl and Dikic 2019). Among these polyubiquitin chains, Lys48 and Lys11-linked polyubiquitin chains serve as the most potent signals for degradation by the proteasome, whereas Lys63-linked chains are more typically associated with non-proteasomal signaling, including endocytic trafficking, DNA replication and signal transduction. However, McKeon et al. reported that Lys63-ubiquitination could also induce substrate degradation via the autophagy-lysosome pathway (McKeon et al. 2015). Other linkage types are less well-characterized, even though some reports have shown that Lys6, 27, 29, and 33-linked polyubiquitin chains are involved in proteasomal degradation (Xu et al. 2009). Recently, systemic analysis has revealed that monoubiquitination can also target some small proteins, particularly proteins of 20 to 150 residues, for degradation by the proteasome (Shabek et al. 2007, 2009, 2012; Nakagawa and Nakayama 2015; Braten et al. 2016).

Deubiquitinating enzymes (DUBs)

Ubiquitination is a dynamic and highly reversible process. Deubiquitinating enzymes (DUBs) can counteract the activity of E3 ligases by removing the ubiquitin chain from the target proteins, which prevents their degradation and reverses other functional changes caused by the ubiquitination. In addition, DUBs have crucial roles in maintaining ubiquitin homeostasis via their involvement in ubiquitin maturation, editing and recycling (Komander et al. 2009). The human genome encodes more than one hundred DUBs, and they are classified into eight families: ubiquitin-specific proteases (USPs), ovarian tumor proteases (OTUs), ubiquitin C-terminal hydrolases (UCHs), Machado-Joseph disease protein domain proteases (MJDs), JAP1/MPN/Mov34 metallopeptidases (JAMMs), the motif interacting with ubiquitin-containing novel DUB family (MINDY), the monocyte chemotactic protein-induced protein (MCPIP), and zinc-finger and UFM1-specific peptidases (ZUFSPs) (Reyes-Turcu et al. 2009; Liang et al. 2010; Abdul Rehman et al. 2016; Hermanns et al. 2018). In addition to reversing ubiquitination, DUBs control multiple cellular pathways such as protein trafficking, chromatin remodeling, cell cycle regulation...
and apoptosis. Therefore, DUBs play roles in a variety of clinical diseases.

26S proteasome

The 26S proteasome, composed of one 20S core particle (20S CP) and one or two 19S regulatory particles (19S RP), is an ATP-dependent multi-subunit complex responsible for hydrolyzing the protein into small peptides. The 20S CP is a barrel-shaped proteolytic core that contains active catalytic sites and is where proteolysis finally occurs, while the 19S RPs serves to selectively and effectively transfer ubiquitinated proteins to the 20S CP by mediating deubiquitination (Groll et al. 1997; Glickman et al. 1998; Bedford et al. 2010; Selvaraju et al. 2015). The 20S CP consists of four stacked heptameric rings (7α, 7β, 7β and 7α) around a central cavity. The outer two α rings allow the interaction with the 19S RPs, while the inner two β rings consist of a total of seven β subunits, of which three β subunits (β1, β2 and β5) are responsible for proteolytic activity (Arendt and Hochstrasser 1999; Voges et al. 1999; Nguyen et al. 2013). The 19S RP has at least 19 subunits and is composed of a lid and a base. Ubiquitinated proteins are captured by specific receptors on the 19S RP, and then they are deubiquitinated by proteasome-associated DUBs (Arendt and Hochstrasser 1999; Voges et al. 1999; Lee et al. 2011). Six different ATPase subunits in the base of the 19S RP supply energy to open the lid of the proteasome channel (Ciechanover 2005).

E3 ubiquitin ligases in cancer

Several E3 ligases have been reported to be mutated or overexpressed in various cancers. The resulting dysfunction of E3 ligases affects DNA damage repair, cell cycle regulation, gene expression, and signaling transduction. Here, we introduce some ubiquitinating enzymes involved in oncogenic signaling and cancer progression (Fig. 2a).

Anaphase-promoting complex (APC/C)

APC/C is a representative E3 ligase essential for mitosis progression. Although APC/C itself is rarely mutated in cancer, it has been reported that deregulation of its co-activators cell division cycle 20 (CDC20) or CDC20-like protein 1 (CDH1) is associated with cancers (Senft et al. 2018).

CDH1-deficient old mice exhibit increased rates of spontaneous epithelial tumors in various organs and downregulation of CDH1 induces premature S-phase entry and genomic instability, indicating that CDH1 acts as a tumor suppressor (Garcia-Higuera et al. 2008; Greil et al. 2016). CDH1 promotes ubiquitination-mediated proteolysis of BRAF, the downstream effector of RAS that drives tumorogenesis, in both APC-dependent and -independent manners. However, hyperactivated ERK and CDK4 phosphorylate CDH1 and disrupt the formation of the APC-CDH1 complex, leading to BRAF dimerization and activation in various cancer cells (Wan et al. 2017). Furthermore, CDH1 is involved in the proliferation of breast cancer cells by inhibiting the kinase activity of c-Src, a proto-oncogene, in an APC-independent manner. Conversely, c-Src suppresses CDH1 activity by phosphorylating it (Han et al. 2019). Besides, CDH1 restrains oncogenic signals such as polo-like kinase (PLK), mitotic cyclins, and Skp2 (Fujita et al. 2009; Zhang et al. 2014). In contrast to CDH1, CDC20 is often overexpressed in a variety of cancers. Moreover, inhibition of CDC20 blocks mitotic exit and induces apoptosis, suggesting that CDC20 has oncogenic properties (Huang et al. 2009; Manchado et al. 2010). The depletion of CDC20 inhibits Wnt signaling and subsequently attenuates cell proliferation in colorectal cancer cells (Kidokoro et al. 2008; Hadjihannas et al. 2012).

S-phase kinase-associated protein 1 (SKP1)-cullin1-F-box protein (SCF) complex

The SCF E3 ligase complex, a multimeric ubiquitination complex consisting of cullin1, SKP1 and one of about 70 F-box proteins, plays multiple roles in cellular processes. The main scaffold of the SCF complex, cullin1 directly binds to SKP1, which then binds an F-box protein that directly binds to their substrates.

F-box and WD repeat domain-containing 7 (FBXW7) is a key component of the SCF E3 ligase complex that regulates the stability of cell cycle regulators (e.g., cyclin E and Aurora A), oncogenic transcription factors (e.g., c-Myc), cell surface receptors (e.g., Notch1), and signaling molecules (e.g., mTOR) (Fujii et al. 2006; Yang et al. 2015). Therefore, impaired SCF-FBXW7 function leads to sustained proliferation and survival, genomic instability, and signaling programs that affect cancer invasion and metastasis (Mao et al. 2004; Rajagopalan et al. 2004; Grim et al. 2012). For example, decreased FBXW7 promotes mTOR activity that support the metastatic potential of cholangiocarcinoma (CCA) to the liver and lung (Yang et al. 2015). In addition, the deletion of FBXW7 increases cyclin E expression and Aurora A deregulation, resulting in chromosomal instability in mice (Koepf et al. 2001; Moberg et al. 2001; Strohmaier et al. 2001; Mao et al. 2004). Indeed, FBXW7 is downregulated in breast, colorectal, and gastric cancers. Its loss is correlated with a poor prognosis and survival, and increased invasion and metastasis (Iwatsuki et al. 2010; Ibusuki et al. 2011; Yang et al. 2015).

SKP2 is an F-box protein of the SCF E3 ligase complex and it is responsible for the degradation of several cyclin-dependent kinase (CDK) inhibitors, such as p21\(^{Cip1}\), p27\(^{Kip1}\), and p18\(^{Kip1}\).
1147 Ubiquitin–proteasome system (UPS) as a target for anticancer treatment

13 Among them, the best studied is the tumor suppressor p27Kip1. SKP2 overexpression induces p27Kip1 degradation, promoting cytokine-stimulated cell cycle progression and the survival of T-cell acute lymphoblastic leukemia (T-ALL) cells (Dohda et al. 2007). Indeed, several studies have shown that SKP2 expression is negatively correlated with p27 levels in different types of tumors (Hershko and Shapira 2006). It has been reported that oncogenic Notch upregulates SKP2 expression, consequently enhancing p27Kip1 degradation in T-ALL cells (Dohda et al. 2007). SKP2 inhibitor C1 (SKPin C1) prevents SKP2-mediated p27Kip1 ubiquitination and degradation, leading to apoptosis by p27 accumulation and cell cycle arrest in uveal melanoma and multiple myeloma (MM) cell lines (Asmamaw et al. 2020). In addition, SKP2 promotes programmed cell death protein 4 (PDCD4) degradation by phosphorylation and ubiquitination, leading to increased cell proliferation and radiation tolerance in breast cancer (Li et al. 2019). Also, SKP2 ubiquitinates FOXO1, which is phosphorylated by AKT, and inhibits its proteasomal degradation, inhibiting apoptosis in prostate cancer cells (Huang et al. 2005).

Neuronally expressed developmentally downregulated 4-1 (NEDD4-1)

NEDD4-1 has been demonstrated to play a critical role in regulating cancer progression. Overexpression of NEDD4-1 promotes tumorigenesis through ubiquitination and inactivation of phosphatase and tensin homolog (PTEN) in non-small cell lung carcinomas (NSCLC) and prostate cancer cells (Wang et al. 2007, 2008; Amodio et al. 2010). Also, NEDD4-1 stabilizes MDM2 by Lys63-linked polyubiquitination, resulting in the promotion of p53 degradation and consequently cell proliferation (Xu et al. 2015). Moreover, NEDD4-1 binds to N-Myc and negatively regulates it by increasing its polyubiquitination.

Fig. 2 Regulation of tumor progression by E3 ligases and DUBs. a Many E3 ligases and b DUBs are involved in cancer progression, and they have several substrates that participate in various biological signaling pathway. For example, USP7 stabilizes MDM2 and DNMT1, resulting in inhibiting cell apoptosis. Also, USP7 regulates PTEN localization and eventually inducing tumorigenesis. Some E3 ligases or DUBs can act as both oncogenic protein and tumor suppressor, depending on the target and type of cancer.
promoting cell proliferation in neuroblastoma cancer cells (Liu et al. 2013). Depletion of NEDD4-1 reduces the phosphorylation of AKT and increases PTEN levels, inhibiting cell growth and migration in hepatocellular carcinoma (HCC) cells (Huang et al. 2017). Meanwhile, NEDD4-1 suppresses tumorigenesis by inducing RAS degradation. RAS genes are frequently mutated in cancers and mutated RAS functions as an oncogene. Zeng et al. reported that RAS signaling stimulates the expression of NEDD4-1, which negatively regulates RAS levels. However, oncogenic-mutated RAS promotes the RAS-induced overexpression of NEDD4-1 and the degradation of PTEN by preventing NEDD4-1-mediated RAS ubiquitination, leading to tumorigenesis in RAS-driven tumors (Zeng et al. 2014). In addition, NEDD4-1 ubiquitinates phosphorylated AKT-Ser473 and induces its proteasomal degradation, inhibiting AKT signaling and promoting apoptosis of MM cells (Huang et al. 2020).

MDM2

MDM2 is an essential regulator of p53, which ubiquitinates p53 and induces its proteasomal degradation. Tumor suppressor p53 is an important transcription factor regulating cellular signaling that affects processes such as apoptosis, cell cycle, DNA repair and senescence. It is functionally impaired by mutation or deletion in about 50% of cancers. In contrast, MDM2 is overexpressed in many malignancies (Shangary and Wang 2008). Overexpressed MDM2 and mutated or downregulated p53 induces cisplatin resistance in several cancers (Kondo et al. 1995; Hayashi et al. 2006). Cisplatin is a drug that causes apoptosis in cancer cells by phosphorylating and activating p53. In addition, resistance to other chemotherapeutic drugs including temozolomide (Sato et al. 2011), doxorubicin (Suzuki et al. 1998) gemcitabine (Sheng et al. 2017), and fluorouracil (5-FU) (Han et al. 2018) is associated with MDM2-p53 negative feedback in many cancers.

DUBs in cancer

As mentioned above, the DUB do not just reverse ubiquitination, but they also have many diverse functions, such as protein trafficking, apoptosis, chromatin remodeling, DNA damage repair and cell cycle regulation, and are particularly involved in cell signaling related to cancers (Antao et al. 2020). Below, we describe some DUBs that act as tumor suppressors or oncogenic proteins in relation to cancers (Fig. 2b).

USP2a

USP2a regulates cell proliferation by deubiquitinating and stabilizing MDM2 and MDM4. Overexpression of USP2a causes an increase in MDM2 accumulation and promotes proteasomal degradation of p53, and thereby it has oncogenic properties in prostate cancer (Priolo et al. 2006; Stevenson et al. 2007). On the contrary, in glioma cells, overexpression of USP2a stabilizes MDM4 and facilitates the p53-dependent intrinsic apoptotic pathway in glioma cells (Wang et al. 2014a). It also deubiquititates fatty acid synthase (FASN), resulting in increased FASN levels, thus enhancing cell proliferation in prostate cancer cells (Graner et al. 2004). Furthermore, USP2a directly interacts with cyclin D1 and promotes its stabilization. Cyclin D1 is a crucial regulator of the G1-S phase transition and it can function as a proto-oncoprotein that is overexpressed in various types of cancers. The accumulation of cyclin D1 by USP2a accelerates the G1 to S phase progression and cell growth in cancer cells (Shan et al. 2009). Finally, the upregulation of USP2a has been reported to induce cell proliferation, migration, invasion and resistance to chemotherapeutic agents in bladder cancer (Kim et al. 2012), and it is associated with a poor prognosis in oral squamous cell carcinomas (da Silva et al. 2009).

USP7

USP7 is a well-known oncogenic protein. It has been reported that an excess amount of USP7 is directly correlated with the development of multiple cancers, such as prostate, colon, liver, lung, breast, glioma, and ovarian cancer (Tavana and Gu 2017; Bhattacharya et al. 2018). Typically, USP7 could stabilize MDM2 by deubiquitination and subsequently promotes the degradation of p53, causing the inhibition of apoptosis in cancer cells (Bhattacharya et al. 2018). In addition, USP7 has several other targets that are associated with tumor progression. For example, USP7 can deubiquitinate PTEN and regulate its localization. Nuclear PTEN is essential for tumor suppression, but when monoubiquitination of PTEN is reduced by USP7, PTEN migrates from the nucleus to the cytoplasm and eventually promotes tumorigenesis. Indeed, USP7 is overexpressed in prostate cancer, and high levels of USP7 are directly correlated with the nuclear exclusion of PTEN and tumor aggressiveness (Song et al. 2008). Also, USP7 deubiquitinates and stabilizes DNA methyltransferase 1 (DNMT1). Because fine-tuning of DNMT1 abundance through posttranslational modifications (PTMs) is important for tumor suppression, increased amounts of USP7 correlate with increased amounts of DNMT1 in colon cancer, consequently promoting tumor progression (Du et al. 2010). Moreover, USP7 promotes tumor progression by deubiquitinating and stabilizing the histone demethylase
PHF8 and by upregulating cyclin A2 in breast cancer (Wang et al. 2016). USP7 also activates Wnt signaling in adenomatous polyposis coli gene (APC)-mutated colorectal cancer through β-catenin deubiquitination (Novellasdemunt et al. 2017). Conversely, the depletion of USP7 reduces the stability of the checkpoint with forhead and ring-finger (CHRF), a mitotic E3 ligase, leading to the accumulation of Aurora A and cyclin B, and increased mitotic defects in a p53-independent manner. This mechanism may explain why breast cancer patients with low levels of USP7 are resistant to the Taxanes, a group of anticancer drugs including Taxol and Taxotere (Giovinazzi et al. 2013). Recently, He et al. reported that USP7 is associated with progression of MM. USP7 promotes MM cell survival by deubiquitinating Maf proteins, including c-Maf, MafA, and MafB, which are critical transcription factors in myelomagenesis. Indeed, USP7 is up-regulated in MM cell line and MM patients with high USP7 expression exhibit lower survival rate (He et al. 2020).

Cylindromatosis (CYLD)

CYLD was originally identified as a germline mutated gene in familial cylindromatosis (FC) (Bignell et al. 2000). CYLD mainly acts as a tumor suppressor in several cancers such as melanoma, human salivary gland (HSG) tumor, colon cancer, HCC, and breast cancer (Park et al. 2019). Wnt-mediated nuclear translocation of β-catenin enhances cylindroma cell proliferation. Downregulation of CYLD in cylindroma cells promotes β-catenin accumulation in the nucleus through Lys63-ubiquitination of Dishevelled (Dvl) (Tauriello et al. 2010). In addition, the expression of CYLD is reduced in melanoma. Repression of CYLD expression increases the ubiquitination of the proto-oncogene Bcl-3 and induces its translocation to the nucleus, resulting in increased expression of cyclin D1 and N-cadherin; thus, melanoma cell proliferation and invasion is enhanced (Massoumi et al. 2009).

CYLD is a critical negative regulator of NF-κB signaling (Park et al. 2019). Sustained NF-κB activity is associated with decreased CYLD expression in HSG tumors and HCC, leading to increased cell survival, tumor progression and metastasis (Fukuda et al. 2006, 2008; Hellerbrand et al. 2007). CYLD removes Lys63-polyubiquitin chains from TRAF-2, TRAF-6, and NEMO, which are intermediates of the NF-κB pathway, resulting in the attenuation of NF-κB activity. However, in breast cancer cells, when CYLD is phosphorylated by overexpressed IkB kinases (IKK-α, β, ε), it is dissociated from the NF-κB intermediates, increasing their ubiquitination, and eventually promoting cell proliferation (Reiley et al. 2005; Hayden and Ghosh 2008; Hutti et al. 2009). Meanwhile, BRG1- and hBRM-associated factor (BAF57) induce cell cycle arrest and apoptosis by stimulating CYLD expression in breast cancer cells (Wang et al. 2005).

A20

Like CYLD, A20 suppresses cell proliferation and promotes apoptosis as a negative regulator of the NF-κB pathway, and it is well known to play an important role in inflammation and immunity (Dixit et al. 1990; Hoesel and Schmid 2013). Recently, many studies have proposed that A20 is highly involved in tumorigenesis. For example, A20 is frequently inactivated by somatic mutations or deletions in lymphomas, and its inactivation is associated with cancer progression (Tavares et al. 2010; Chu et al. 2011; Hovelmeyer et al. 2011). Meanwhile, A20 is highly expressed in diverse solid tumors; in these cases, the overexpression of A20 affects chemotherapy and radiation resistance (Guo et al. 2009; Chen et al. 2015; Yang et al. 2018).

A20 is unique in that it has both DUB and E3 ligase activities. Lee et al. reported that A20 facilitates TGF-β1-induced epithelial-mesenchymal transition (EMT) and metastasis by increasing multi-monoubiquitination of Snail1 in basal-like breast cancers (Lee et al. 2017).

Proteasome-associated DUBs in cancer

Three DUBs are associated with the 19S RP in mammalian cells. USP14 (also known as Ubp6) and UCH37 (also known as UCHL5) are cysteine proteases and are located in the lid. They mediate a stepwise removal of ubiquitin from the substrate by trimming the distal tip of the ubiquitin chain (Patel et al. 2018). The third DUB, RPN11 (also known as POH1) is a metalloprotease and it cleaves at the base of the ubiquitin chain where it is linked to the substrate (Lee et al. 2011). In particular, RPN11-mediated deubiquitination promotes substrates degradation, but USP14 and UCH37 antagonize degradation (Verma et al. 2002). Proteasome-associated DUBs play an important role in maintaining ubiquitin homeostasis through the removal of ubiquitin from the substrate and its recycling, and they regulate protein degradation by rescuing poorly ubiquitinated proteins or enhancing the activity of the proteasome (Lee et al. 2011).

USP14

USP14 is closely related to the progression of multiple tumors (Peth et al. 2009). It deubiquitinates and stabilizes the androgen receptor (AR), which contributes to the progression and development of prostate and breast cancers (Liao et al. 2017, 2018). Furthermore, USP14 promotes cell growth, migration and invasion in gastric cancer by targeting vimentin, which is involved in EMT (Zhu et al. 2017).
Wnt signaling regulates various homeostatic mechanisms; hence, its aberrant activation is observed in diverse cancers (Reya and Clevers 2005). USP14 is involved in Wnt signaling activation and β-catenin accumulation by regulating the ubiquitination of Dvl and its subsequent phosphorylation (Jung et al. 2013). In addition, a strong correlation has been observed between USP14 and β-catenin levels in colorectal cancer cells, and overexpressed USP14 could promote cell proliferation through the accumulation of β-catenin in NSCLC cells (Jung et al. 2013; Wu et al. 2013).

**UCH37**

Overexpression of UCH37 has been reported to be associated with a lower survival rate and an increased risk of cancer recurrence in cancer patients (Wang et al. 2014b). In endometrial cancer (EC), Wnt signaling is involved in cell survival, cell cycle, proliferation and metastasis. UCH37 activates Wnt signaling and affects the expression of its target genes, such as β-catenin, cyclin D1 and c-Myc, thereby increasing the cell growth of EC (Liu et al. 2020). It also regulates TGF-β signaling, and aberrant TGF-β signaling is closely related to the development of cancer. UCH37 deubiquitinates and stabilizes type I TGF-β receptors by directly binding with SMAD7, which forms a complex with Smurf E3 ligase and induces degradation of the TGF-β receptor (Wicks et al. 2005). The specific USP14 and UCH37 inhibitor b-AP15 suppresses cell growth via TGF-β signaling and induces apoptosis in ovarian cancer cells (Fukui et al. 2019). UCH37 interacts with COP5 to induce the ubiquitination and degradation of p53 and p21, promoting cell proliferation. Treatment with b-AP15 downregulates COP5 levels and upregulates p53 levels, resulting in durable tumor regressions in p53-deficient mice (Ma et al. 2020). Furthermore, UCH37 knockdown results in reduced cell survival, increased caspase activity and a higher ratio of apoptotic regulator Bax/anti-apoptotic factor Bcl-2 in NSCLC cells (Chen et al. 2011b).

**RPN11**

Previous studies have shown that RPN11 expression is upregulated in various cancers such as breast cancer, MM and hepatocellular carcinoma (Luo et al. 2017). Downregulation of RPN11 induces apoptosis causing cell arrest in the G0-G1 phase, and ultimately leading to senescence by regulating the phosphorylation of cyclin-dependent kinases (CDKs) and retinoblastoma protein (Rb) in cancer cells (Byrne et al. 2010). Conversely, overexpression of RPN11 promotes cancer progression by stabilizing the E2F transcription factor 1 (E2F1) and promoting the expression of its target genes in hepatocellular carcinoma cells (Wang et al. 2015). Furthermore, RPN11 deubiquitinates ErbB2, leading to its accumulation, resulting in the promotion of cell growth and the inhibition of apoptosis of ErbB2-positive breast cancer cells (Liu et al. 2009; Song et al. 2017).

## Inhibitors of ubiquitinating enzymes-clinical trials

As mentioned above, several E3 ligases have been considered as targets for novel anticancer drugs. However, clinical trials of inhibitors targeting E3 ligases are limited, because E3 ligases can act as both tumor suppressors and oncogenic proteins and their underlying mechanisms of regulating cellular processes are very complex (Zhang et al. 2020). Here, we introduce the inhibitors of ubiquitinating enzymes, especially E3 ligases that are currently in clinical studies.

### MLN4924 (pevonedistat)

MLN4924 is a small molecule inhibitor of the NEDD8-activating enzyme (NAE) E1 enzyme (Soucy et al. 2009; Luo et al. 2012). This small molecule is an adenosine sulfamate analog that covalently binds to NAE to create a NEDD8-MLN4924 adduct, consequently blocking neddylation of all cullin-RING ligases (CRLs), which regulate the destruction of many intracellular proteins. Inhibition of CRL neddylation suppresses cancer cell growth by increasing the levels of their substrates, which in turn triggers the DNA damage response, cell-cycle arrest, apoptosis and autophagy (Aubry et al. 2020). MLN4924 has been applied in several phase I clinical trials for certain solid tumors and hematologic malignancies and showed clinical activity in a phase I clinical trial of acute myelogenous leukemia (AML) (Swords et al. 2010). It is also known to inhibit angiogenesis during tumor development (Yao et al. 2014).

### MDM2 inhibitors

MDM2 negatively regulates p53 activity, increasing cell survival in various cancers. The first small molecule inhibitor of MDM2 is Nutlin-3a, which inhibits the interaction between MDM2 and p53 (Vassilev et al. 2004). After optimization of Nutlin-3a, several derivatives were generated. Among them, RO-5503781 (Idasanutlin) is developed by Hoffmann-La Roche (Ding et al. 2013). This orally available inhibitor is proposed to treat glioblastoma (in preclinical trials), diffuse large B cell lymphoma, MM, prostate cancer (in a phase I/II clinical trial), and AML (in a phase III clinical trial combined with chemotherapy) (adisinsight.springer.com). However, this inhibitor was terminated in phase II clinical trials. In addition, MK-8242 developed by Merch & Co was terminated in phase I clinical trials for patients with advanced solid tumors and recurrent AML and liposarcomas. Besides that, SAR-405838, developed by Ascenta
Therapeutics and Sanofi, CGM097 developed by Novartis, and DS3032b developed by Daichi Sankyo are all targeting the interaction of MDM2 and p53, and they are currently in phase I clinical trials for patients with advanced solid tumors, either alone or in combination with chemotherapy.

**Inhibitors of the proteasome-clinical trials**

Since the first proteasome inhibitor bortezomib was approved by the FDA for treating relapsed and refractory MM in 2003, the development of anticancer drugs targeting the proteasome among the components of the UPS has been the most active research field. All of the proteasome inhibitors developed so far play a role in inhibiting proteolytic activity by binding to the inner β-subunit of the 19S RP of the proteasome.

**Bortezomib (BTZ, velcade)**

Bortezomib is a boronic acid-containing dipeptide, the first proteasome inhibitor introduced for the treatment of MM. This is a unique and reversible inhibitor of the 26S proteasome that binds to the β5 subunit of 19S RP and inhibits its chymotrypsin-like enzymatic activity, thus leading to the accumulation of polyubiquitinated proteins (Zhang et al. 2020). Incomplete and transient inhibition of the 26S proteasome by BTZ causes apoptosis of myeloma cells by activation of caspases-8/9 (Hu et al. 2013). BTZ is currently used for the treatment of various cancers including MM, relapsed mantle cell lymphoma, diffuse large B-cell lymphoma, colorectal cancer, and thyroid carcinoma. However, BTZ is dose-limiting toxic and has been reported to cause multiple side effects, including pain, fatigue, peripheral neuropathy, myelosuppression, and cardiotoxicity. Additionally, about 60% of patients treated with BTZ developed resistance to this drug. To overcome BTZ resistance, the FDA approved the use of BTZ in combination with chemotherapeutic agents, such as Doxorubicin, Adriamycin, and Dexamethasone in clinical trials (Mata-Cantero et al. 2015; Zhang et al. 2020).

**Carfilzomib (CFZ, kyprolis)**

Carfilzomib is an epoxyketone peptide and is the second proteasome inhibitor approved by the FDA in 2012 for the treatment of patients with MM who have shown resistance to BTZ (Kuhn et al. 2007). CFZ inhibits the chymotrypsin-like activity of the proteasome more strongly than BTZ by forming a covalent and irreversible bond with catalytic subunits, predominantly β5. Furthermore, it has a more selective mechanism of action and high chemical stability, resulting in fewer side effects compared to BTZ (Arastu-Kapur et al. 2011; Stewart et al. 2015). However, CFZ also has a problem: it does not dissolve well in water. For this reason, CFZ cannot be administered orally and a large amount of cyclodextrin is required for intravenous injection formulations (Zhang et al. 2020).

**Ixazomib (IXZ, ninlaro)**

In 2015, ixazomib received FDA approval as the first orally bioavailable proteasome inhibitor for the treatment of MM (Chauhan et al. 2011). It also binds to catalytic β-subunits and inhibits proteasome activity. Because of its great advantage that IXZ can be administered orally, its effectiveness is being studied as either a single or combined chemotherapeutic agents for a number of cancers. Indeed, it is known that IXZ does not affect the mitochondrial serine protease Htr/Omi, which is an off-target of BTZ related to neuropathy, which is one of the side effects of BTZ (Zhang et al. 2020).

**Inhibitors of DUB—clinical trials**

DUBs have a high level of substrate specificity and a catalytic domain with properties similar to that of which it is attracting attention as a promising therapeutic target for cancers. (Eletr and Wilkinson 2014). Over the last few years, a large of small molecule inhibitors against DUBs have been reported, but only a few of them have entered into clinical trials for certain cancers.

**Pimozide**

Pimozide is a reversible inhibitor of the enzymatic activity of the USP1/UAF1 complex. It was originally discovered as an antipsychotic drug and approved by the FDA for the treatment of Tourette’s syndrome. However, it has been revealed through HTS that pimozide inhibits USP1/UAF1, which acts synergistically with cisplatin to inhibit cell proliferation in cisplatin-resistant NSCLC cells (Chen et al. 2011a). This inhibitor is in a phase II clinical trial in patients with amyotrophic lateral sclerosis (Antao et al. 2020).

**Mitozantrone (novantrone)**

Mitozantrone was discovered through HTS of a large panel of FDA-approved compounds for inhibition of USP11 catalytic activity. This drug inhibits the DNA-damage repair (DDR) system induced by USP11, and it impacts pancreatic ductal adenocarcinoma (PDA) cell survival (Burkhart et al. 2013). In addition, mitozantrone inhibits TGF-β1-induced phosphorylation of SMAD2/3 and the type I TGF-β receptor through the reduction of type II TGF-β receptor stability maintained by USP11 in lung fibroblast cells (Jacko et al. 2016). Currently, numerous phase I–IV clinical trials are
under way targeting T-cell lymphoma, AML, multiple sclerosis, and breast cancer using mitozantrone (Antao et al. 2020).

### Papain-like cysteine protease (PLpro) inhibitors

6-mercaptopurine (6MP) and 6-thioguaninie (6TG) are specific inhibitors of PLpro, a protease of the severe acute respiratory syndrome coronavirus (SARS-CoV). PLpro

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**Table 1** Inhibitors targeting UPS components in clinical trials

| Name             | Target          | Functions                                                                 | Clinical status | Cancer types                                      | Structure | Ref                                |
|------------------|-----------------|---------------------------------------------------------------------------|-----------------|-------------------------------------------------|-----------|-----------------------------------|
| MLN4924          | NAE (E1)        | Blocks the neddylation of all CRLs, leading to apoptosis in cancer cells | Phase I         | AML, LSC/LC, Hepatocellular carcinoma, mesothelioma, Advanced solid tumor, MM, Lymphoma, AML, ALL, Melanoma | ![Structure](image1) | www.clinicaltrials.gov (Aubry et al. 2020) |
| NSC697923        | Ubc13- Uev1A (E2) | Blocks the formation of the E2-Ub thioester conjugate and inhibits NF-κB signaling | Pre-clinical    | Melanoma, B-cell lymphoma, Neuroblastoma, Colorectal cancer | ![Structure](image2) | (Pulvino et al. 2012; Zhang et al. 2020) |
| SAR-405838       | HDM2-p53 interaction | Selective spirooxindole derivative antagonist of HDM2, Leads to p53 activation | Phase I         | Solid tumor                                      | ![Structure](image3) | www.clinicaltrials.gov (de Weger et al. 2019) |
| CGM-097          | HDM2-p53 interaction | Orally bioavailable HDM2 antagonist, Leads to p53 activation               | Phase I         | Advanced solid tumor with p53 wild type status   | ![Structure](image4) | www.clinicaltrials.gov            |
| DS-3032          | MDM2-p53 interaction | Orally bioavailable HDM2 antagonist, Leads to p53 activation               | Phase I/II      | AML                                            | ![Structure](image5) | www.clinicaltrials.gov            |
| Al-406           | XIAP, cIAP1/2    | Orally active antagonist of IAPs, Activates apoptosis                     | Phase I         | Lymphoma, Advanced solid tumor                  | ![Structure](image6) | www.clinicaltrials.gov            |
Table 1 (continued)

| Name          | Target | Functions                                      | Clinical status | Cancer types                                      | Structure | Ref                                            |
|---------------|--------|-----------------------------------------------|-----------------|--------------------------------------------------|-----------|------------------------------------------------|
| HGS-1029      | cIAP2  |                                               | Phase I         | Advanced solid tumor                              |           | [www.clinicaltrials.gov](http://www.clinicaltrials.gov) |
| Compound #25  | SKP2   | Suppresses SKP2 E3 ligase activity            | Preclinical     | Prostate cancer                                   |           | (Chan et al. 2013)                              |
| NAHA          | CDC20  | Decreases CDC20 expression and inhibits tumor proliferation | Preclinical     | Breast cancer                                     | 2-[Benzyl-(2-nitro-benzenesulfonyl)-amino]-N-hydroxy-3-methyl-N-propyl-butyramide | (Wang et al. 2013) |
| Bortezomib    | Proteasome (β5>β1) | Inhibits the chymotrypsin-like activity of the proteasome by reversible binding to the β5 subunit and leads to accumulation of polyubiquitinated proteins. | Launched MM     | Non-Hodkin’s lymphoma Other lymphoma Leukemia NSCLC Kidney cancer Mesothelioma Soft tissue sarcoma Glioma Neuroblastoma Head and neck cancer Prostate cancer Brain tumor Breast cancer Colorectal cancer Ovarian cancer |           | [www.clinicaltrials.gov](http://www.clinicaltrials.gov) (Mata-Cantero et al. 2015; Zhang et al. 2020) |
| Carfilzomib   | Proteasome (β5) | Inhibits chymotrypsin-like activity of proteasome more strongly than BTZ by forming a covalent and an irreversible bond with catalytic subunits, predominantly β5 | Launched MM     | Non-Hodkin’s lymphoma Kidney cancer Prostate cancer Lung cancer Ovarian cancer Hodkin’s lymphoma |           | [www.clinicaltrials.gov](http://www.clinicaltrials.gov) (Arastu-Kapur et al. 2011; Stewart et al. 2015) |
| Ixazomib      | Proteasome (β5>β1) | First orally bioavailable proteasome inhibitor binds to catalytic β-subunits and inhibits proteasome activity | Phase III Phase II | MM Lymphoma Advanced solid tumors ALL |           | [www.clinicaltrials.gov](http://www.clinicaltrials.gov) (Chauhan et al. 2011) |
| Delanzomib    | Proteasome (β5, β1, β2) | Inhibits chymotrypsin-like activity of proteasome and represses the proteasomal degradation of proteins, including IκB-α | Phase I | Non-Hodkin’s lymphoma |           | [www.clinicaltrials.gov](http://www.clinicaltrials.gov) (www.cancer.gov) |
Table 1 (continued)

| Name                  | Target     | Functions                                                                 | Clinical status | Cancer types                                      | Structure                          | Ref                                      |
|-----------------------|------------|---------------------------------------------------------------------------|-----------------|--------------------------------------------------|-------------------------------------|-----------------------------------------|
| **Proteasome inhibitors** |            |                                                                           |                 |                                                  |                                     |                                         |
| Oprozomib             | Proteasome (β5-β1) | Structural homologue of CFZ and orally bioavailable inhibitor Inhibits activity of proteasome and induces apoptosis | Phase II, Phase I | MM, Advanced solid tumor                          | ![Structure](image1)                  | [www.clinicaltrials.gov](www.clinicaltrials.gov) [www.cancer.gov](www.cancer.gov) |
| Marizomib             | Proteasome (20S CS) | Irreversibly binds to and inhibits the 20S catalytic core subunit of the proteasome by covalently modifying its active site threonine residues | Phase III, Phase II, Phase I | Glioblastoma, MM, Ependymoma, Glioma, NSCLC, Pancreatic cancer, Melanoma, Lymphoma | ![Structure](image2)                  | [www.clinicaltrials.gov](www.clinicaltrials.gov) [www.cancer.gov](www.cancer.gov) |
| **DUB inhibitors**    |            |                                                                           |                 |                                                  |                                     |                                         |
| Pimozide              | USP11, Pan-DUBs | Reversible, non-competitive inhibition of USP1                            | Pre-clinical    | Melanoma, Breast cancer, LSCLC                    | ![Structure](image3)                  | (Chen et al. 2011a)                      |
| Mitoxantrone          | USP11      | Unknown                                                                   | Phase IV, Phase III, Phase I/II | APL, AML, ALL, Lymphoma, Breast cancer, Leukemia, Prostate cancer, Breast cancer, Ovarian cancer | ![Structure](image4)                  | [www.clinicaltrials.gov](www.clinicaltrials.gov) (Burkhart et al. 2013) |
| 6MP, 6TG              | PLpro, USP14 | Host antiviral immune response activation                                | Launched        | ALL, Lymphoma, Leukemia acute lymphocytic, LAM, Lymphoma, Colorectal cancer, Breast cancer, Glioblastoma | ![Structure](image5)                  | [www.clinicaltrials.gov](www.clinicaltrials.gov) (Chen et al. 2009; Kemp 2016) |
| b-AP15                | USP14, UCHL5 | Inhibition of 19S RP deubiquitinating activity                            | Pre-clinical    | MM, AML, Large B-cell lymphoma, Neuroblastoma, Prostate cancer, Ovarian cancer, Colorectal cancer, Lung cancer | ![Structure](image6)                  | (Zhang et al. 2020)                      |
| IU1                   | USP14      | Inhibition of ubiquitin chain trimming                                   | Pre-clinical    | Breast cancer                                     | ![Structure](image7)                  | (Xia et al. 2019)                       |
| Ortho-phenanthroline (OPA) | RPN11          | Inhibits RPN11 activity, but does not affect proteasome activity         | Pre-clinical    | MM, Hepatocellular carcinoma, Cervical cancer, Breast cancer | ![Structure](image8)                  | (Guterman and Glickman 2004; Song et al. 2017) |
blocks IRF-dependent host antiviral immune responses by acting as a DUB that removes ubiquitin or ISG15 (another ubiquitin-like peptide) from host cell proteins (Kemp 2016). They have been used for the treatment of acute lymphatic leukemia and Non-Hodgkin’s lymphoma (FDA-approved drugs). Also, they are currently in phase II–IV clinical trials in patients with leukemia, lymphoma, colorectal and breast cancers (Mata-Cantero et al. 2015). In addition, 6MP and 6TG show favorable binding energy to USP14, suggesting that they are potential inhibitors of USP14 (Chen et al. 2009).

Conclusion

Since the 2000s, the successful development of the proteasome inhibitor bortezomib as a MM treatment has marked the importance of the UPS in tumorigenesis, and numerous studies related to this are still ongoing. Not only the proteasome, but also the various components of the UPS play an important role in cancer progression, and it has been consistently demonstrated that small molecule inhibitors against them have an excellent effect on tumor suppression (Table 1). However, only a few UPS inhibitors are still in use in clinical practice, and many challenges including drug resistance, several side effects and limited efficacy in the treatment of solid tumors, are preventing them from being developed as successful cancer treatments. If these challenges can be overcome, the UPS will certainly become an attractive target for cancer therapeutics.

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Compliance with ethical standards

Conflict of interest All authors declares that they have no conflict of interest.

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**Ubiquitin–proteasome system (UPS) as a target for anticancer treatment**

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