The Pharmacokinetics, Metabolism, and Clearance Mechanisms of Abrocitinib, a Selective Janus Kinase Inhibitor, in Humans

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ABSTRACT
Abrocitinib is an oral once-daily Janus kinase 1 selective inhibitor being developed for the treatment of moderate-to-severe atopic dermatitis. This study examined the disposition of abrocitinib in male participants following oral and intravenous administration using accelerator mass spectrometry methodology to estimate pharmacokinetic parameters and characterize metabolite (M) profiles. The results indicated abrocitinib had a systemic clearance of 64.2 L/h, a steady-state volume of distribution of 100 L, extent of absorption >90%, time to maximum plasma concentration of ~0.5 hours, and absolute oral bioavailability of 60%. The half-life of both abrocitinib and total radioactivity was similar, with no indication of metabolite accumulation. Abrocitinib was the main circulating drug species in plasma (~26%), with 3 major monohydroxylated metabolites (M1, M2, and M4) at >10%. Oxidative metabolism was the primary route of elimination for abrocitinib, with the greatest disposition of radioactivity shown in the urine (~85%). In vitro phenotyping indicated abrocitinib cytochrome P450 fraction of metabolism assignments of 0.53 for CYP2C19, 0.30 for CYP2C9, 0.11 for CYP3A4, and ~0.06 for CYP2B6. The principal systemic metabolites M1, M2, and M4 were primarily cleared renally. Abrocitinib, M1, and M2 showed pharmacology with similar Janus kinase 1 selectivity, whereas M4 was inactive.

SIGNIFICANCE STATEMENT
This study provides a detailed understanding of the disposition and metabolism of abrocitinib, a Janus kinase inhibitor for atopic dermatitis, in humans, as well as characterization of clearance pathways and pharmacokinetics of abrocitinib and its metabolites.

Introduction
Abrocitinib (PF-04965842) is a selective small molecule Janus kinase (JAK)-1 inhibitor under development at daily doses of 200 and 200 mg to treat moderate-to-severe atopic dermatitis (Gooderham et al., 2019; Silverberg et al., 2020; Simpson et al., 2020; Bieber et al., 2021). Selective inhibition of cytokines by abrocitinib that signal through JAK1 dependent pairs (i.e., JAK1/JAK2, JAK1/JAK3, and JAK1/tyrosine kinase (TYK) 2) includes interleukin (IL)-4, IL-13, and other cytokines (e.g., IL-31, IL-22, and thymic stromal lymphopoietin) involved in the pathogenesis of atopic dermatitis and pruritus (Trier and Kim, 2018) while sparing non-JAK1 pairs such as JAK2/JAK2 inhibition and thereby minimizing the risk for neutropenia and anemia (Akada et al., 2014).

Abrocitinib is a weak base of molecular mass 323.4 with measured logP of 1.7 and pKa of 5.2 (data on file). Abrocitinib showed high in vitro permeability in Madin-Darby canine kidney cells at 16 × 10⁻⁶ cm/s. Aqueous solubility was pH dependent at ~8 mg/mL at pH ~1 and ~35 μg/mL at pH ~8. Abrocitinib pharmacokinetics have been investigated in healthy volunteers (Peeva et al., 2018). Abrocitinib was absorbed rapidly following single doses of 3–200 mg [the median time that plasma Cmax occurred (Tmax) was <1 hour] and more slowly at higher doses of 400 and 800 mg (median Tmax 1.5–4 hours), indicating some delayed absorption at higher doses. Plasma Cmax increased proportionally from 3 to 800 mg: the area under concentration-time curve extrapolated to infinity (AUCinf) was greater than proportional with doses of 400 and 800 mg. Multiple dose pharmacokinetics
showed steady state was reached by day 4 with an observed accumulation ratio of 1.3:1.5 with once-daily dosing between 30 and 200 mg, and plasma half-life ranged between approximately 2 and 4 hours. Renal clearance of abrocitinib was estimated to be between 0.59 to 0.67 L/h (Peeva et al., 2018).

Examining human radiolabeled absorption, distribution, metabolism, and excretion (ADME) provides a comprehensive profile of the disposition of a drug. Recent application of accelerator mass spectrometry (AMS) analytical methodology to human ADME studies has been an important advancement in study design as this enables administration of up to 1000-fold lower doses of carbon-14 compared with traditional methods of study (Spracklin et al., 2020). Using microtracer of carbon-14 (0.1–1 µCi administration), more informative ADME studies that include two-period fixed sequence oral (period A) and intravenous (period B) designs can be performed; these studies allow the determination of additional drug parameters including systemic clearance, volume of distribution, gut availability, and absolute bioavailability. In addition, the low level of radioactivity eliminates the need for the prerequisite quantitative whole-body autoradiography rat study to estimate dosimetry or the need to run nonclinical safety studies with an intravenous formulation since the microtracer is adequately supported by oral toxicology studies. An important design element of the microtracer intravenous period is the administration of a clinically relevant nonlabeled dose of the drug, followed by the administration of the intravenous microtracer of 14C-drug at approximately T_max to reflect an appropriate total mass of drug in the body and allow scaling of the intravenous pharmacokinetic parameters to a therapeutic dose (Spracklin et al., 2020).

The objectives of the present fixed-dose crossover oral and intravenous study were to investigate the disposition, mass balance, and metabolic profiles of a therapeutic oral dose of 200 mg abrocitinib in healthy participants as well as determine its systemic clearance, volume of distribution, gut availability, and absolute bioavailability. In addition, metabolic clearance mechanisms of abrocitinib were investigated with in vitro hepatic systems, and in vitro pharmacology of both abrocitinib and primary circulating metabolites was assessed in human whole-cell assays.

**Materials and Methods**

**Materials and Reagents**

Abrocitinib (PF-04965842), its deuterated standard (PF-06651703), and metabolites (M)-1 (PF-06471658), M2 (PF-07055087), M3 (PF-07055090), M4 (PF-07054874), M5 (PF-07054926), M6 (PF-07054627), M7 (PF-06737821), and 2-phenyl-2-(1-piperidinyl) propane (PPP) were synthesized at Pﬁzer (Groton, CT). ^14C-abrocitinib (^14C label on the 2-position of the carbon of the pyrimidine ring) was synthesized by Perkin Elmer Inc. (Boston, MA), with radiochemical purity of 98.3% and specific radioactivity of 6.31 µCi/mg. Analytical chemicals of analytical grade or better were obtained from Fisher Scientific (Waltham, MA). MgCl₂, NaDHP, dicrofenac, furafylline, tienilic acid, esomeprazole, quinidine, aminobenzotriazole, 1 M potassium phosphate dibasic solution, 1 M potassium phosphate monobasic solution, sodium bicarbonate, and DMSO were purchased from Sigma Aldrich (St. Louis, MO). Troleandomycin was ordered from Fisher Scientific (Waltham, MA), and gemfibrozil acyl glucuronide was synthesized at Wuxi App Tecx (Shanghai, China). HPLC solvents included ammonium acetate, acetonitrile, methanol, and formic acid from Biosolve (Valkenswaard, Netherlands); trifluoroacetic acid from Sigma Aldrich (Amsterdam, Netherlands); isopropyl amine from iPAm, Merck (Amsterdam, Netherlands); 2 mM ammonium acetate/methanol/acetonitrile (95:2.5:2.5 % v/v) from Brand-Nu Laboratories (Meriden, CT); and methanol, acetonitrile, and water from J.T. Baker (Avantor, Radnor, PA). Cell culture medium included Williams E media (GIBCO-BRL custom formula Lot# 1081513) and HEPES (Lonza, Switzerland). Pooled human liver microsomes (HLM) [HLM103, 20.0 mg of protein per ml, 0.31 nmol of cytochrome P450 (P450) per mg of protein] were prepared and characterized by Sekisui Xenotech (Kansas City, KS) from 50 individual human donors of both sexes. Cryopreserved human hepatocytes (lot DCM) were prepared and characterized by Bioreclamation IVT (Baltimore, MD) from 10 individual donors of both sexes.

**Clinical Study and Sample Collection**

This study was a phase I, open-label, nonrandomized, 2-period, fixed sequence study conducted at PRA-EDS International (Groningen, Netherlands). Six healthy male participants (1 Hispanic/5 non-Hispanic), 22–28 years of age, with a mean body weight of 83.1 kg and body mass index of 24.4 kg/m² were enrolled. Participants were genotyped for CYP2C19 and 2C9 with commercially available TaqMan assays and analyzed on a QuantStudio 12K Flex Real-Time PCR System. Each participant received 2 dose regimens in periods A and B (14 days following period A) following an overnight fast. Period A was an oral solution dose of 200 mg abrocitinib and 80 µg ^14C-abrocitinib. Plasma samples were collected before dosing and from 0.25 to 96 hours after dose. Urine and feces were collected prior to dose and up to 240 hours following dose (excreta collection continued until at least 90% of administered dose was recovered or less than 1% was recovered from excreta from 2 consecutive days). Period A samples were analyzed for total ^14C and metabolite profiling. Period B was an oral solution dose of 200 mg abrocitinib followed by a 100 µg ^14C-abrocitinib intravenous approximately 1 hour later (T_max). Plasma samples were collected before dose and at various times postdose and analyzed for total ^14C, ^14C-abrocitinib (0.083–95 hours postintravenous dose), and abrocitinib (0.25–96 hours following oral dose) concentrations. Urine was collected before dose and up to 143 hours postintravenous dose and analyzed for total ^14C radioactivity. Feces were not collected in period B based on the observation that <10% of dose was recovered in feces in period A. The final protocol and informed consent documentation were reviewed and approved by the Independent Ethics Committee at the investigational center participating in the study. This study was conducted in compliance with the ethical principles originating in or derived from the Declaration of Helsinki and in compliance with all International Council for Harmonization Good Clinical Practice Guidelines.

**Radiochemical Analysis**

Analyses of total ^14C radioactivity in plasma, urine, and feces homogenate were conducted at TNO (Microbiology and Systems Biology, Zeist, Netherlands) using ^14C detection by AMS. AMS analysis was performed on a 1MV multielement AMS, model 4110 Bo, High Voltage Engineering (software: AMS 155). Low-level scintillation counting was performed on a Quantulus 1220 (software WinQ version 1.2). Aliquots of plasma (5 µL), urine (15 µL, diluted or undiluted) or feces homogenate (homogenized at the PRA Health Sciences Bioanalytical Laboratory (Drenthe, Netherlands), 30 mg, diluted or undiluted) was transferred to tin foil cups. The samples were dried under a stream of nitrogen and subsequently placed in the elemental analyzer (Vario Micro; Elementar, Langenselbold, Germany), which acted as autosampler and CO₂ combustion device for the AMS (van Duijn et al., 2014). The natural background ^14C/C level, measured in predose samples of each matrix, was subtracted to obtain the amount of ^14C abrocitinib-related ^14C. ^14C/C isotope ratios of the samples were converted to mBq/mL for plasma and urine samples, whereas the homogenized feces samples data were converted into mBq/g. The total ^14C radioactivity concentrations in urine and feces were further converted to percentage of dose recovered in urine and feces based upon the radioactivity dose administered to each participant. A reference standard ANU sucrose-8542 with a certified ^14C/^{12}C isotope ratio was used as system suitability sample. In each batch, the measured ^14C/^{12}C ratio deviated <15% from the certified ratio. Plasma standard samples were included in each analysis as AMS quality control (QC) samples with a minimum of 3 replicates. The samples were prepared by spiking blank pooled plasma with a known amount of ^14C (from ^14C-paracetamol). The ^14C/^{12}C ratios were determined by AMS and found to deviate no more than 15% from the nominal value. In addition, the CV was also <15%.

**Metabolite Profiling in Plasma, Urine, and Feces**

**Sample Preparation.** To profile circulating metabolites, plasma samples from each individual at 0–12 hours postdose, which covered >97% of the total radioactivity area under the curve (AUC), were pooled according to the method of Hamilton et al. (1981). The pre- and postdose pools were prepared using equal volumes from each participant. The pooled plasma (100 µL) was diluted with acetonitrile (500 µL), vortexed for 1 minute, and centrifuged (14,000g, 5 minutes,
homogenate sample, 500 μL supernatant was removed and dried under a stream of nitrogen. The residue was dissolved in 2 mM ammonium acetate/acetoni­terile/methanol (80%-10%:10% v:v:v), vortexed for 30 seconds, and centrifuged (1000g, 1 minute, room temperature). Urine samples collected from each participant from 0 to 12 hours following dosing represented 96% of the excreted radioactivity. A predose pool was prepared by mixing equal volumes of each predose sample. An equal volume from each individual was pooled, 50 μL of urine sample, 700 μL 2 mM ammonium acetate/acetoni­terile/methanol (80%-10%:10% v:v:v) was added, vortexed for 1 minute, and centrifuged (1000g, 1 minute, room temperature). Individual fecal homogenate pools accounting for 97% of the total dose collected from feces were prepared for each participant by combining proportional amounts of the sample collected over each time period (48–144 hours, 24–96 hours, 24–72 hours, 48–168 hours, 0–48 hours, and 0–72 hours, respectively). An equal amount from each individual participant pool was then combined into a master pool. A pooled predose sample was prepared by taking equal aliquots per participant. To a 100 mg of feces homogenate sample, 500 μL acetoni­terile was added, equilibrated for 5 minutes, and vortexed for 1 minute. The sample was centrifuged (14,000g, 5 minutes, 10°C), 500 μL supernatant was removed, and a second extraction was performed using the same methods. The residue was dissolved in 2 mM ammonium acetate/acetoni­terile/methanol (80%-10%:10% v:v:v), vortexed for 30 seconds, and centrifuged (100g, 1 minute, room temperature). Sample extraction and liquid chromatography–mass spectrometry (LC-MS) profiling column recoveries were all greater than 89%.

Metabolite Profiling Ultra-High-Performance Liquid Chromatography Methods. Pooled urine and extracts from pooled plasma and homogenized feces were profiled using a high-resolution liquid chromatography–tandem mass spectrometry (LC-MS/MS) method described in Supplemental Table 1. After the column, the eluent was split: 1) part of the flow was used to generate on-line high-resolution mass spectrometry (MS) [and the data-dependent mass spectrometry (MS/MS) spectrum], and 2) the other part of the flow was collected for off-line AMS analysis. For each chromatogram, a total of 154 fractions were collected (0–14 minutes, 6-second fractions; 14–28 minutes, 1-minute fractions).

AMS Analysis. To each fraction from 0–14 minutes, additional carbon-12 was added (paracetamol in methanol), and the whole fraction was transferred to a tinfoil cup, dried under a stream of nitrogen, and subsequently placed in the elemental analyzer, which acted as autosampler and combustion device for the AMS. To each fraction from 14–28 minutes, additional carbon-12 was added to obtain a suitable amount of carbon for AMS analysis. An aliquot of 60 μL was transferred to a tinfoil cup, dried under a stream of nitrogen, and subsequently placed in the elemental analyzer, which acted as autosampler and combustion device for the AMS.

Metabolite Identification. The Q Exactive mass spectrometer operated in the positive electrospray ionization mode. The heated electrospray ion source voltage was 3.0 kV. The heated capillary temperature was 350°C. Auxiliary and spare gases were set to 20 and 0 units, respectively. The scan-event cycle consisted of a full-scan mass spectrum (100–1000 m/z) at a resolving power of 35,000, and the corresponding data-dependent tandem mass spectrometry scans were acquired at a resolving power of 35,000. Accurate mass measurements were performed using external calibration. A reference list of known metabolites of abrocitinib and their expected molecular ions was added to an inclusion list to prioritize those ions during data acquisition. Postacquisition searching of LC-MS data for molecular ions representing possible metabolites of 14C abrocitinib was performed manually. In addition, the LC-MS data associated with every radiochromatographic peak was also interrogated. LC-MS peaks identified to be possible metabolites, based on the molecular ions, were compared against blank samples. In cases where the signal strength was of sufficient intensity to trigger data-dependent tandem mass spectrometry, the subsequent fragment spectra were examined to further confirm the identity and structure of possible metabolites. In addition, LC-MS chromatograms were compared directly against the appropriate predose matrix chromatogram to identify additional metabolite peaks.

Chiral Separation of Metabolites M2 and M3. Plasma, urine, and feces homogenate extracts were injected in triplicate using the UHPLC system described above (Supplemental Table 1), and the fraction containing both enantiomers was collected. For each matrix, these 3 fractions (from the triplicate injections per matrix) were pooled and dried under a stream of nitrogen. The dried fractions were reconstituted in 200 μL mobile phase (0.1% trifluoroacetic acid, 0.1% isoproplamine in acetonitrile/methanol (90%-10% v:v:v)). The reconstituted fractions were injected on the UHPLC where the chromatography was performed using a ChiralPAK ZWIX(R) (4 × 150 mm, 3 μm, Diacel, Chiral Technologies, Inc., West Chester, PA). An isotropic flow rate of 0.25 mL/min was maintained throughout the analysis. For each matrix, a duplicate injection of 25 μL was performed. After the column, the eluent was fraction collected for off-line AMS analysis.

Isolation of Metabolites M6 and M7. Approximately 30 mL of 0- to 24-hour-pooled human urine was centrifuged at 1800g for 20 minutes and the supernatant applied to a Bond Elut C-18 SPE cartridge (10 gm, Agilent Technologies, Santa Clara, CA) equilibrated in 5% methanol in water at a flow rate of 2.0 mL/min. The cartridge was successively eluted with 50 mL of 10 mM ammonium acetate in water containing varying percentages of 1% methanol/acetoni­terile (2.5%, 5%, 10%, 15%, 20%, 25%, 30%, and 2.5%). Fifty microliters of the flow through and each eluate were analyzed by LC-MS/MS (Supplemental Table 2) to determine which fractions contained metabolites M6 and M7. Eluates rich in M6 and M7 were evaporated to near dryness in a vacuum centrifuge at room temperature. Residues were redissolved in 0.1% formic acid in 1.1 mM methanol/aceto­nitirile (97.5%-2.5% v:v), and 5-μL aliquots were fractionated using the LC-MS/MS method shown in Supplemental Table 3. One-minute fractions were collected, and 10 μL of fractions between 9.0 and 36 minutes were analyzed by LC-MS/MS for presence of M6 and M7 metabolites (Supplemental Table 4). Fractions rich in M6 and M7 were pooled and evaporated to dryness in a vacuum centrifuge at room temperature. The residues were redissolved in 1.0 mL 10 mM ammonium acetate/methanol/acetoni­terile (97.5%-1:2.5%-2.5% v:v:v) and purified using the LC-MS/MS method shown in Supplemental Table 5. One-minute fractions were collected, and 10 μL of fractions between 4.0 minutes and 32 minutes were analyzed by LC-MS/MS (Supplemental Table 4) for presence of m/z 354 (M7) and m/z 370 (M6) metabolites. Fractions containing metabolites were then pooled and evaporated to dryness for NMR analysis.

Nuclear Magnetic Resonance Characterization of Metabolite M6 and M7. All samples were dissolved in 0.05 mL of DMSO-d6 (~100% (Cambridge Isotope Laboratories, Andover, MA) and placed in a 1.7 mm NMR tube in a dry argon atmosphere. 1H and 13C spectra were referenced using residual DMSO-d6 (2.50 ppm relative to tetramethylsilane, δ = 0.00, 13C δ = 39.50 ppm relative to tetramethylsilane, δ = 0.00). NMR spectra were recorded on a Bruker Avance 600 MHz (Bruker BioSpin Corporation, Billerica, MA) controlled by Topspin V3.1 and equipped with a 1.7 mm TCI Cryo probe. One-dimensional spectra were recorded using a sweep width of 10,000 Hz and a total recycle time of 7 second. The resulting time-averaged free induction decays were transformed using an exponential line broadening of 1.0 Hz to enhance signal to noise. The two dimensional data were recorded using the standard pulse sequences provided by Bruker. At minimum, a 1K × 128 data matrix was acquired using a minimum of 2 scans and 16 dummy scans with a spectral width of 10,000 Hz in the j2 dimension. The data were zero-filled to at least 1K data point.

Abrocitinib Quantification. Plasma samples from period B were analyzed for abrocitinib concentration at WuXi AppTec (Shanghai, China) with a validated LC-MS/MS method. Abrocitinib and internal standard PF-06651703 (deuterated abrocitinib) were extracted from 75 μL human plasma using liquid-liquid extraction with ethyl acetate, and extracts were analyzed with the LC-MS/MS method shown in Supplemental Table 6. The calibration range was 1.00–2000 ng/mL, and the QC concentrations were 3.00, 60.0, 1000, and 1600 ng/mL. The interday assay accuracy ranged from 4.8% to 0.0%, and the between-day precision was ≤7.8%. Plasma samples from period B were analyzed for 14C-abrocitinib concentration at TNO (Microbiology and Systems Biology, Ziest, Netherlands) using a qualified analytical method. Protein precipitation of 100 μL plasma was performed using 3:1 acetoni­terile/plasma followed by centrifugation, drying under a stream of nitrogen, and reconstitution with 80:10:10 2 mM ammonium acetate/acetoni­terile/methanol. Samples were analyzed by LC-MS/MS methods (Supplemental Table 1) with AMS detection (1MV multielement AMS model 4110B, High Voltage Engineering) The calibration range was 0.50–100 mBq/mL, and QC concentrations were 1.50, 20.0, 75.0, and 200 mBq/mL. The interday assay accuracy ranged from −10.9% to −3.0%, and the between-day precision was ±3.7%.
In Vitro P450 Assignment

The experimental conditions used in vitro were confirmed to yield linear reaction velocities as determined from preliminary range-finding experiments.

In Vitro Hepatocyte Fractional Metabolism.

The reaction phenotyping experiment was conducted using $^{13}$C abrocitinib (1 μM) incubated in pooled human cryopreserved hepatocytes at 0.75 million cells per mL in Williams’ E Medium at 37°C in 5% CO$_2$/95% air and 85% relative humidity for 30 minutes. Aliquots were quenched into acetonitrile (1:4, v/v) and centrifuged (1800g) for 5 minutes, and the supernatant was transferred to clean 15 mL conical glass tubes. The supernatants were dried in a Genevac (Genevac Inc, Valley Cottage, NY) evaporative centrifuge at 37°C, and the resulting residues were reconstituted in 100 μL of 2 mM ammonium acetate/acetonitrile/methanol (95%:2.5%:2.5% v/v/v/v).

Sample bioanalysis was performed using the LC-MS/MS method shown in Supplemental Table 7. Postcolumn, the eluent was split: 1) part of the flow was used to generate on-line high-resolution MS (and/or data-dependent MS/MS) spectrum, and 2) the other part of the flow was diverted to the fraction collector. Fractions were collected in 6-second intervals into Lumnaplate-96 well plates (PerkinElmer, Waltham, MA) and dried in a Genevac evaporative centrifuge at 37°C. Plates were then placed in a PerkinElmer MicroBeta2 counter and each well counted for total $^{13}$C for 5 minutes. The results were used to generate a reconstructed radiochromatogram.

Enzyme Kinetics. Experiments for the determination of enzyme kinetic parameters were conducted in human liver microsomes. Abrocitinib (0.1–1000 μM) was incubated in 100 mM potassium phosphate buffer (pH 7.4) containing 3 mM MgCl$_2$ and 0.3 mg/mL microsomal protein HLM-103 at 37°C. Reactions were initiated with NADPH (1.2 mM) immediately followed by substrate and terminated after 15 minutes by quenching aliquots of the incubation mixture with acetonitrile-containing internal standard (Diclofenac, 10 ng/mL). Samples were centrifuged (1700g) for 10 minutes, and 100 μL of supernatant was transferred to a 96 deep-well plate. The supernatants were dried down under a stream of nitrogen gas and reconstituted in 100 μL of 90% water/10% acetonitrile. Incubations for enzyme kinetic determination were conducted in triplicate.

P450 Chemical Inhibition. The P450-selective chemical inhibition experiment was conducted using abrocitinib (10 μM), pooled human hepatocytes, and chemical inactivators of P450s 1A2, 2B6, 2C8, 2C9, 2C19, 2D6, 3A, and pan-P450 in a 96-well plate. Hepatocytes (120 μL) at 0.75 million cells per mL in Williams E media were preincubated with inhibitors for 15 or 30 minutes prior to initiating the reactions by the addition of abrocitinib. Incubations containing 100 μM gemfibrozil glucuronide (CYP2C8), 15 μM tienilic acid (CYP2C9), and 1 mM 1-aminobenzotriazole (pan-P450) were preincubated for 30 minutes, and 10 μM furafylline (CYP1A2), 5 μM PFP (CYP2B6), 5 μM esomeprazole (CYP2C19), 2 μM CYP51cide (CYP3A4), and 25 μM troleandomycin (CYP3A4/5) were preincubated for 15 minutes with human hepatocytes to achieve complete inactivation of respective P450 isoforms (Walsky and Obach, 2007; Yang et al., 2016). Quinidine (10 μM) was used as a competitive inhibitor (CYP2D6) without preincubation prior to the addition of abrocitinib. Incubations of abrocitinib were conducted at 37°C in 5% CO$_2$/95% air and 75% relative humidity for 30 minutes. Aliquots were collected into acetonitrile-containing internal standards to quench the reaction. Quenched samples were centrifuged (1700g) for 10 minutes, and 150 μL of supernatant was transferred to a 96 deep-well plate. The supernatants were dried under a stream of nitrogen gas and reconstituted in 100 μL of 10% acetone/90% water. Selective chemical inhibition experiments were conducted in triplicate.

Enzyme Kinetics and Chemical Inhibition Bioanalysis. Sample bioanalysis was performed using the LC-MS/MS method shown in Supplemental Table 8. Integration and quantitation of metabolites and internal standard molecule peak areas were performed using Analyst version 1.6.2 (AB Sciex) to derive the analyte to internal standard peak area ratios. Standard curves were prepared from plots of area ratio versus concentration and analyzed using a linear regression with either 1/x or 1/x$^2$ weighting.

Fractional Clearance in Human Hepatocytes. To determine the fractional contributions of metabolic pathways of $^{13}$C-abrocitinib, radioactivity counts for fractions were plotted against time to reconstruct liquid chromatography chromatograms, which were then aligned and assessed against MS spectral data for agreement. The sum of radioactive counts associated with an individual chromatographic peak was determined and compared with the total radioactivity recovered to derive the percentage of total radioactivity using the 30-minute time point. The contribution of a single metabolic pathway (represented by an individual metabolite plus any secondary metabolites deriving from it) was calculated as the response of the individual metabolite divided by the total metabolism of abrocitinib.

Enzyme Kinetics in Human Liver Microsomes. Substrate concentration (S) and velocity (V) data were fitted to the appropriate enzyme kinetic model by nonlinear least-squares regression analysis (SigmaPlot version 13; Systat Software, Inc., San Jose, CA or GraphPad version 6.03; GraphPad Software Inc., La Jolla, CA) to derive the apparent enzyme kinetic parameters using the Michaelis-Menten model (eq. 1) and the substrate inhibition model (eq. 2).

\[
V = \frac{V_{\text{max}} \times S}{K_m + S} \quad (1)
\]

\[
V = \frac{V_{\text{max}} \times S}{K_m + S + (1 + S/K_i)} \quad (2)
\]

where $V_{\text{max}}$ is the maximal velocity, $K_m$ or $S_{50}$ is the substrate concentration at half-maximal velocity, and $K_i$ is an inhibition constant. The best fit was based on a number of criteria, including visual inspection of the data plots (Michaelis-Menten and Eadie-Hofstee), distribution of the residuals, size of the sum of the squared residuals, and the standard error of the estimates. Selection of models other than Michaelis-Menten was based on the F-test ($P < 0.05$) or the Akaike Information Criterion. In vitro apparent intrinsic clearance ($CL_{\text{int}}$) values were calculated using eq. 3:

\[
CL_{\text{int}} = \frac{V_{\text{max}}}{K_m} \quad (3)
\]

$CL_{\text{int}}$ data were also calculated from rate of product formation (V) obtained at S below the apparent $K_m$ ([$S_{50}$], in which cases eq. 1 was rearranged to $CL_{\text{int}} = V/[S]$.

In vitro apparent intrinsic clearance ($CL_{\text{int(app)}}$) and scaled in vivo apparent intrinsic clearance ($CL_{\text{int(app,s)}}$) Values for M1, M2/M3, and M4 were calculated using the following equations:

\[
CL_{\text{int(app)}}(\text{HLM}) = CL_{\text{int(app)}}(\text{HLM}) \times 45 \text{ mg protein/gram liver} \times 21.2 \text{ kg body weight} \quad (4)
\]

Selective P450 Chemical Inhibition in Human Hepatocytes. To determine the contribution of each P450 isoform to abrocitinib metabolism in human hepatocytes, the metabolite formation rates were calculated by dividing the metabolite concentration by incubation time and protein concentration. The effect of P450 selective chemical inhibitors on the formation rate of each metabolite was calculated using the following formula:

\[
\%\text{Inhibition}(\text{unscaled}) = \left( 1 - \frac{V_{\text{inh}}}{V_{\text{ctrl}}} \right) \times 100 \quad (5)
\]

wherein $V_{\text{inh}}$ is the rate of formation in the presence of inhibitor and $V_{\text{ctrl}}$ is the rate of metabolite formation in the control human hepatocyte incubation. To determine the contribution of each P450 isoform for the metabolites without standard, the analyte-to-internal-standard-area ratio was used. When the combined inhibitor effects on a metabolic pathway totaled more than 100%, the percentage inhibition values for that pathway were normalized to 100%. The fraction of abrocitinib metabolism represented by a single metabolite $x$ and specific P450 isoform $z$ ($f_{\text{M450}(x,P450)}$) was calculated using the following equation:

\[
f_{\text{M450}(x,P450)} = \frac{\%\text{Inhibition}}{100} \times f_{\text{CL}} \quad (6)
\]

where $\%\text{Inhibition}$ is the scaled inhibition of metabolite $x$ by a P450-specific inhibitor $z$, and $f_{\text{CL}}$ is the fraction of abrocitinib clearance represented by metabolite $x$ determined in radiolabeled human hepatocyte experiments.

A statistical analysis was conducted comparing the metabolite formation rates in the presence of inhibitor versus control incubations to confirm
significance. A one-way analysis of variance analysis with a Dunnett’s Test for multiple comparisons was applied to the triplicate values from inhibited and control conditions.

In Vitro Pharmacology

The potency of abrocitinib and its metabolites against the four JAK iso-
forms, JAK1, JAK2, JAK3, and TYK2, was measured in terms of IC50 as previously reported (Dowty et al., 2019). In addition, the potency of abrocitinib and its metabolites to inhibit the ability of various cytokines to induce signal transducer and activator of transcription (STAT) phosphory-
lation in human whole blood, human keratinocytes, or human acute mono-
cytic leukemia (THP-1) cells was assessed as previously reported (Dowty et al., 2019). More methodology details are provided in the Supplemental Methods.

Plasma Protein Binding

An equilibrium dialysis method was used to determine plasma fraction un-
bound (fu) values as described previously (Riccardi et al., 2017). Briefly, dialysis membranes (MWCO 12-14K) and 96-well dialysis devices were assembled fol-
lowing the manufacturer’s instructions (HTDialysis, LLC, Gales Ferry, CT). Human plasma samples (pooled mixed sex; BioIVT, www.bioivt.com) containing 1 μM test compounds with 1% DMSO were dialyzed against PBS for 6 hours in a humidified incubator (75% relative humidity; 5% CO2/95% air) at 37°C with shaking at 450 RPM. Quadruplicates of binding were measured for each compound. Samples were matrix-matched and quenched with cold acetonitrile-containing internal standard(s). The solutions were centrifuged (3600g, 30 minutes, 4°C), and the supernatant was analyzed using a generic LC-MS/MS method (Supplemental Tables 9–11).

Blood-to-Plasma Ratio

Human blood-to-plasma ratio was measured by Unilabs York Bioanalytical Services (York, UK). Test compounds were incubated in quadruplicate with fresh human blood (mixed sex, at least 1 sample per sex, Clinical Trials Labora-
tory Services Ltd, London, UK) at 1 μM in a humidified incubator (95% relative humidity; 5% CO2/95% air) at 37°C with shaking at 450 RPM. Following incubation, plasma samples were obtained by centrifuging blood samples (3000g; 7 minutes). Both plasma and blood samples were matrix-
matched with each other and quenched with acetonitrile-containing internal standard. The solutions were centrifuged, and the supernatant was analyzed using a generic LC-MS/MS method (Supplemental Tables 9–11). Peak area ratios were used to calculate blood-to-plasma ratio.

Results

Clinical Safety. All 6 participants completed the study. Oral and intravenous doses of abrocitinib in period A and B were safe and well tolerat-
ed, and no serious or drug-related adverse events were reported. All reported adverse events were mild in severity (Supplemental Table 12), and there were no clinically relevant changes in vital signs, clinical laboratory test results, or electrocardiogram measurements throughout the study period.

Pharmacokinetics. Plasma concentration-time profiles and pharma-
koetic parameters of abrocitinib are summarized in Fig. 1 and Table 1,
respectively. Abrocitinib showed rapid absorption following oral administration, with a $T_{max}$ of 0.5 hour. Systemic clearance and volume of distribution of abrocitinib were estimated at 64.2 L/h and 100 L, respectively. The absorption and elimination profiles of total radioactivity were comparable to those of abrocitinib, with >97% of circulating radioactivity measured within the 0- to 12-hour collection pool. There were some differences in P450 genotypes, with 3 participants with 2C19*1/*1 [extensive metabolizer (EM)] and 2C9*1/*1 (EM); 2 participants with 2C19*1/*17 (rapid metabolizer) and 2C9*1/*12 (intermediate metabolizer); and 1 participant with 2C19*1/*17 (rapid metabolizer) and 2C9*1/*1 (EM). However, these differences did not translate into significant differences in systemic clearance as percentage coefficient of variation was only 14%.

**Disposition and Metabolite Profiles.** After a single oral dose of $^{14}$C-abrocitinib in male participants, radioactivity was excreted predominantly in the urine (Fig. 2). At 240 hours after the dose, the mean (plus or minus standard deviation) cumulative excretion in the urine was 85.0% ± 6.0% and in the feces was 9.5% ± 0.5% (Table 2). In total, 94.5% ± 5.9% of the radioactive dose was recovered in urine and feces, with a major portion of excreted radioactivity recovered during the first 24 hours after dosing [urine 84.0% ± 6.4%, feces 17% ± 3.0%, total 85.7% ± 8.7%]. Collectively, 13 metabolites were identified in urine and feces (Table 2). Oxidative metabolites M6, M1, M2, and M4 were the most abundant in urine at 12.2%, 16.2%, 13.5%, and 15.4% of the administered dose, respectively. All other metabolites comprised less than 10% of the dose. Representative HPLC radiochromatograms of plasma (0- to 12-hour pool after dose, representing >97% radioactivity AUC), urine (0- to 12-hour pool after dose, representing 96% of urine total recovery), and fecal (pools representing 97% of the feces total recovery) profiling are shown in Fig. 3. Unchanged abrocitinib accounted for the majority (25.8%) of the total circulating radioactivity (Table 2), with M1, M2, and M4 contributing 11.3%, 12.4%, and 13.8%, respectively. All remaining circulating metabolites made up less than 10% each of total circulating radioactivity.

**Metabolite Identification.** Structural determination was performed on 13 metabolites of abrocitinib. Metabolites were identified by comparing retention times on chromatograms and mass spectra of authentic standards or elucidated based on mass fragmentation patterns (Table 3; Supplemental Table 13). The proposed metabolic pathways of abrocitinib are shown in Fig. 4.

The parent compound, abrocitinib, had a retention time of 11.5 minutes using the HPLC conditions described. Abrocitinib has a protonated molecular ion at m/z 324.1490 (molecular formula C14H21N5O2S, theoretical mass m/z 324.1489). Its MS/MS spectrum showed diagnostic fragment ions at m/z 201, 176, 175, 149, and 70. The ion at

![Fig. 2. Cumulative mean (plus or minus standard deviation) recovery of administered radioactivity in urine (85.0% ± 6.0%) and feces (9.5% ± 0.5%) following a single oral dose of 200 mg $^{14}$C-abrocitinib (n = 6). Total recovery was 94.5% ± 5.9%.](image)

### Table 2

| Analyte | Plasma (% of Total Radioactivity) | Urine $^b$ (% of Dose) | Feces $^b$ (% of Dose) | Total Urine/Feces (% of Dose) |
|---------|----------------------------------|------------------------|-----------------------|-------------------------------|
| Abrocitinib | 25.8 | 0.6 | 0.3 | 0.9 |
| M4 | 13.8 | 15.4 | 0.3 | 15.7 |
| M2$^a$ | 12.4 | 13.5 | 0.5 | 14.0 |
| M1 | 11.3 | 16.2 | 1.7 | 17.9 |
| M3$^a$ | 4.8 | 4.5 | 0.3 | 4.8 |
| M7 | 4.6 | 5.6 | 1.5 | 7.1 |
| M6 | 3.4 | 12.2 | 0.8 | 13.0 |
| 372-1 | 2.5 | 0.7 | 0.2 | 0.9 |
| 358-1 | 2.0 | 1.1 | 0.1 | 1.2 |
| 356-1a | 1.5 | 1.7 | 0.2 | 1.9 |
| 356-2 | 1.5 | 3.3 | 0.6 | 3.9 |
| M8 | 1.4 | 1.3 | nd | 1.3 |
| 340-5 | 1.0 | 0.4 | 0.2 | 0.6 |
| M5 | 0.4 | 0.3 | nd | 0.3 |

Identified analytes ordered in relationship to prevalence in plasma. $^a$M2 and M3 are stereoisomers. $^b$Only percentages of identified metabolites are reported.
m/z 201 resulted from the neutral loss of the propane-1-sulfonamide. Fragment m/z 176 represented the N-cyclobutylpropane-1-sulfonamide. Fragmentation through the cyclobutane ring yielded m/z 175. Fragment m/z 149, N-methyl-7H-pyrrolo[2,3-d]pyrimidin-4-amine, resulted from the neutral loss of N-cyclobutylpropane-1-sulfonamide, whereas fragment m/z 70 represented the cyclobutanamine ion.

**Metabolite M6.** M6 is a minor metabolite in plasma (3.4% of the radioactivity in the circulation) and was identified in urine (14.7%) and in fecal homogenate (8.3%) from profiled radioactivity. The amount of dose excreted as M6 in urine and fecal homogenate was 12.2% and 0.8%, respectively. M6 (molecular formula C14H19N5O5S, theoretical mass m/z 370.1180, observed mass m/z 370.1179) was 46 mass units

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**Fig. 3.** Representative HPLC radiochromatograms of plasma (A), urine (B), and feces (C) following a single 200 mg dose of 14C-abrocitinib.
greater than parent and had a retention time of 3.4 minutes on HPLC. Fragmentation of the molecular ion yielded m/z 217, 165, and 70. Fragment m/z 217 represented the neutral loss of propan-1-ol and the addition of 16 mass units to the N-cyclobutyl-N-methyl-7H-pyrrolo[2,3-d]pyrimidin-4-amine. Fragment m/z 70, cyclobutanamine, remained intact. The remaining increase of 30 mass units was suggested to occur on the propane through di-oxidation and reduction.

NMR analysis of M6 (Supplemental Fig. 1) isolated from pooled 0- to 12-hour human urine supported formation of a carboxylic acid on the methyl group of the propyl side chain; it also supported formation of an amide carbonyl and a methylene at the 6′ and 5′ positions of the pyrrolo pyrimidine ring, respectively. The methyl group was absent in the 1H spectrum of M6; in the homonuclear correlation spectroscopy (COSY) spectrum, cross peaks were observed between positions H18 (δ = 3.07 ppm) and H21 (δ = 2.31 ppm). Formation of the acid was further confirmed in the Heteronuclear Multiple Bond Correlation (HMBC) spectrum, which revealed cross peaks from both H18 and H21 into a carbon with a chemical shift value of 172.5 ppm, likely corresponding to the acid carbonyl (C22). Methines corresponding to the 6′ and 5′ positions of the pyrrolo pyrimidine ring were both absent in the 1H spectrum of M6, and a new methylene at the 5′ position (1H/13C δ = 3.8/35.6 ppm) was observed. The HMBC spectrum revealed a cross peak between the newly formed methylene at the 5′ position (H9) into a carbon resonance at 175.2 ppm, suggesting the presence of an amide carbonyl at the 6′ position (C8). Additional cross peaks in both COSY and HMBC spectra as well as resonances observed in the heteronuclear single quantum coherence spectrum (data not shown) indicated the rest of the molecule remained unmodified with respect to parent drug.

**Metabolite 372-1.** 372-1 is a minor metabolite in plasma (2.5% of the radioactivity in the circulation). The amount of dose excreted as 372-1 in urine and fecal homogenate was 0.7% and 0.2%, respectively. The tri-hydroxylated metabolite, 372-1 (molecular formula C13H17N5O4S, theoretical mass m/z 372.1336, observed mass m/z 372.1334), was 48 mass units greater than parent and had a retention time of 5.0 minutes on HPLC. Fragmentation of the molecular ion yielded m/z 176 and 70. Fragment m/z 176 represented the intact N-cyclobutylpropane-1-sulfonamide, indicating that each oxidation was taking place on the N-methyl-7H-pyrrolo[2,3-d]pyrimidin-4-amine.

**Metabolite M7.** M7 was defined as a minor metabolite in plasma (4.6% of the radioactivity in the circulation). The amount of dose excreted as M7 in urine and fecal homogenate was 5.6% and 1.5%, respectively. The carboxylic metabolite, M7 (PF-07057821, 354-1) (molecular formula C14H21N5O4S, theoretical mass m/z 354.1230, observed mass m/z 354.1230), was 30 mass units greater than parent and had a retention time of 5.2 minutes on HPLC. Fragmentation of the molecular ion yielded m/z 218, 201, 175, 149, and 70. Fragment m/z 201 represented unchanged N-cyclobutyl-N-methyl-7H-pyrrolo[2,3-d]pyrimidin-4-amine.

NMR analysis of M7 (Supplemental Fig. 2) isolated from pooled 0- to 12-hour human urine supported formation of a carboxylic acid on the methyl group of the propyl side chain. The methyl group was absent in the 1H spectrum of 354-1, whereas in the COSY spectrum cross peaks were observed between positions H18 (δ = 3.12 ppm) and H21 (δ = 2.40 ppm). Formation of the acid was further confirmed in the HMBC spectrum, which revealed cross peaks from both H18 and H21 into a carbon with a chemical shift value of 172.5 ppm, likely corresponding to the acid carbonyl (C22). Additional cross peaks in both COSY and HMBC spectra as well as resonances observed in the heteronuclear single quantum coherence spectrum (data not shown) indicated the rest of the molecule remained unmodified with respect to parent drug.

**Metabolite M8.** M8 is a minor metabolite in plasma (1.4% of the radioactivity in the circulation). The amount of dose excreted as M8 in urine was 1.3%. The di-hydroxylated metabolite, M8 (PF-067055039, 356-1) (molecular formula C15H23N5O5S, theoretical mass m/z 356.1387, observed mass m/z 356.1387), was 32 mass units greater than parent and had a retention time of 5.8 minutes on HPLC. Fragmentation of the molecular ion yielded m/z 217, 165, and 70, which represented the proposed hydroxylation of the 7H-pyrrolo[2,3-d]pyrimidin-4-yl group. The remaining hydroxylation was proposed to be located on the propyl group. Fragmentation was identical to the synthetic reference standard.

**TABLE 3**

Molecular ions and characteristic fragment ions of abrocitinib and metabolites detected in plasma and excreta

| ID     | Standard ID    | Retention Time (min) | [M+H]<sup>+</sup> m/z | Parent Fragments m/z | Diagnostic Fragments m/z |
|--------|----------------|----------------------|------------------------|----------------------|-------------------------|
| M6     | PF-07059462    | 3.4                  | 370.1179               | 70                   | 217, 165                |
| 340-5  | 4.9            | 370.1074             | 201, 176, 175, 149, 70 | 296                  |
| 372-1  | 5.0            | 372.1334             | 176, 70                |                      |
| M7     | PF-06737821    | 5.2                  | 354.1230               | 201, 175, 149, 70    | 218                     |
| M8     | PF-07055039    | 5.8                  | 356.1387               | 70                   | 217, 165                |
| 358-1  | 6.1            | 358.1543             | 176, 70                |                      | 235, 217, 183, 165      |
| 356-1a | 6.4            | 356.1386             | 70                     |                      | 217, 192, 191, 165      |
| 356-2  | 6.5            | 356.1384             | 176, 70                |                      | 233, 207, 181           |
| M1     | PF-06471658    | 7.6                  | 340.1438               | 201, 175, 149, 70    | 218                     |
| M2     | PF-07055087    | 8.5<sup>a</sup>      | 340.1438               | 201, 175, 149, 70    | 218                     |
| M3     | PF-07055090    | 8.5<sup>a</sup>      | 340.1438               | 201, 175, 149, 70    | 218                     |
| M4     | PF-07054874    | 10.0<sup>a</sup>     | 340.1438               | 176, 70              | 234, 217, 191, 165      |
| M5     | PF-07054926    | 10.0<sup>a</sup>     | 310.1332               |                      | 204, 187, 161, 135      |
| Abrocitinib | PF-04965842 | 11.5                | 324.1490               | 201, 176, 149, 70    |                      |

<sup>a</sup>Coeluting enantiomers followed by a second chromatographic separation (see Methods).
<sup>b</sup>Coeluting peaks with subsequent identification from urine isolation and nuclear magnetic resonance analyses (see Methods).
Fig. 4. Proposed metabolic pathways of abrocitinib following oral administration.
Metabolite 358-1. 358-1 was defined as a minor metabolite in plasma (2.0% of the radioactivity in the circulation). The amount of dose excreted as 358-1 in urine and fecal homogenate was 1.1% and 0.1%, respectively. 358-1 (molecular formula C14H22N4O4S, theoretical mass m/z 358.1544, observed mass m/z 358.1543) was 34 mass units greater than parent, which represented the result of hydroxylation and hydrolysis. It had a retention time of 6.1 minutes on HPLC. Fragmentation of the molecular ion yielded m/z 217, 176, 165, and 70. Fragment m/z 176 represented the intact N-cyclobutylpropane-1-sulfonamide group. Fragment m/z 183 represented the N-methyl-7Hpyrrolo[2,3-d]pyrimidin-4-amine with the addition of 34 mass units. The ions m/z 217 and 165 represented the loss of water and dehydration with the intact hydroxy group on the 7H-pyrrolo[2,3-d]pyrimidin-4-yl group. The ion m/z 192 represented the proposed hydroxylation of the 7H-pyrrolo[2,3-d]pyrimidin-4-amine group. Fragmentation of the molecular ion yielded ions m/z 217, 191, 165, and 70. The ion m/z 165 represented the proposed hydroxylation of the 7H-pyrrolo[2,3-d]pyrimidin-4-yl group. The ion m/z 192 represented the proposed hydroxylation of the N-cyclobutylpropane-1-sulfonamide group.

Metabolite 356-1a. 356-1a was defined as a minor metabolite in plasma (1.5% of the radioactivity in the circulation). The amount of dose excreted as 356-1a in urine and fecal homogenate was 1.7% and 0.2%, respectively. The di-hydroxylated metabolite, 356-1a (molecular formula C14H21N5O3S, theoretical mass m/z 356.1386, observed mass m/z 356.1386), was 32 mass units greater than parent and had a retention time of 6.4 minutes on HPLC. Fragmentation of the molecular ion yielded m/z 217, 191, 165, and 70. Fragment m/z 165 represented the proposed hydroxylation of the 7H-pyrrolo[2,3-d]pyrimidin-4-yl group. The ion m/z 192 represented the proposed hydroxylation of the N-cyclobutylpropane-1-sulfonamide group.

Metabolite 356-2. 356-2 was defined as a minor metabolite in plasma (1.5% of the radioactivity in the circulation). The amount of dose excreted as 356-2 in urine and fecal homogenate was 3.3% and 0.6%, respectively. The di-hydroxylated metabolite, 356-2 (molecular formula C14H21N5O3S, theoretical mass m/z 356.1387, observed mass m/z 356.1386), was 32 mass units greater than parent and had a retention time of 6.5 minutes on HPLC. Fragmentation of the molecular ion yielded m/z 217, 191, 165, and 70. The ion m/z 165 represented the proposed hydroxylation of the 7H-pyrrolo[2,3-d]pyrimidin-4-yl group. The ion m/z 192 represented the proposed hydroxylation of the N-cyclobutylpropane-1-sulfonamide group.

Metabolite M1. M1 was defined as a major metabolite in plasma (11.3% of the radioactivity in the circulation). The amount of dose excreted as M1 in urine and fecal homogenate was 16.2% and 17.7%, respectively. The hydroxylated metabolite, M1 (molecular formula C14H22N4O4S, theoretical mass m/z 340.1438, observed mass m/z 340.1438), was 16 mass units greater than parent and had a retention time of 7.6 minutes on HPLC. Fragmentation of the molecular ion yielded ions m/z 218, 175, 149, and 70, all identical to the parent molecule. Fragmentation was identical to the synthetic reference standard.

Metabolite M2. Both M2 and M3 are coeluting enantiomers. The abundance of the individual enantiomers was determined using both UHPLC and chiral chromatography. M2 was defined as a major metabolite in plasma (12.4% of the radioactivity in the circulation). The amount of dose excreted as M2 in urine and fecal homogenate was 13.5% and 0.5%, respectively. Hydroxylated M2 (molecular formula C14H21N4O3S, theoretical mass m/z 340.1438, observed mass m/z 340.1438) and M3 (molecular formula C14H21N5O3S, theoretical mass m/z 340.1438) are 16 mass units greater than parent. M2 had a retention time of 8.5 minutes on HPLC. Fragmentation of the molecular ion yielded ions m/z 201, 175, 149, and 70, which are all identical to the parent molecule. Fragmentation was identical to the synthetic reference standard.

Metabolite M3. M3 was defined as a minor metabolite in plasma (4.8% of the radioactivity in the circulation). The amount of dose excreted as M3 in urine and fecal homogenate was 4.5% and 0.3%, respectively. Fragmentation of the molecular ion yielded ions m/z 201, 175, 149, and 70, all identical to the parent molecule. Fragmentation was identical to the synthetic reference standard.

Metabolite M4. M4 was defined as a major metabolite in plasma (13.8% of the radioactivity in the circulation). The amount of dose excreted as M4 in urine and fecal homogenate was 15.4% and 0.3%, respectively. The oxidized metabolite, M4 (PF-07054874, 340-4), (molecular formula C14H21N5O3S, theoretical mass m/z 340.1438, observed mass m/z 340.1438) was 16 mass units greater than parent. This metabolite coeluted with M5. It had a retention time of 10.0 minutes on HPLC. Fragmentation of the molecular ion yielded m/z 217, 191, 176, 165, and 70. Fragmentation was identical to the synthetic reference standard.

Metabolite M5. M5 was defined as a minor metabolite in plasma (0.4% of the radioactivity in the circulation). The amount of dose excreted as M5 in urine was 0.3% and was not detected in fecal homogenate. The N-desmethylated metabolite, M5, (molecular formula C13H19N5O2S, theoretical mass m/z 310.1332, observed mass m/z 310.1332) was 14 mass units less than parent and had a retention time of 10.0 minutes on HPLC. This metabolite coeluted with M4. Fragmentation of the molecular ion yielded m/z 187, 161, and 135 (Table 3). Fragmentation was identical to the synthetic reference standard. The abundance of coeluting M4 and M5 were determined using the mass spectrometry, peak areas of reference standards at a single concentration. These peak

### Table 4

| Parameter          | M1   | M7   | M2/M3 | M4   | 358-1 | 372-1 | 149 | Abrocitinib |
|--------------------|------|------|-------|------|-------|-------|-----|-------------|
| % radioactivity    | 5.6  | 0.35 | 4.2   | 3.6  | 0.16  | 0.16  | 0.42| 85          |
| Metabolite fCL     | 0.39 | 0.024| 0.29  | 0.25 | 0.011 | 0.011 | 0.029| —           |
| Final fCL          | 0.41 | 0.29 | 0.27  | 0.029| 0.029 | 0.029 | —   | —           |

### Table 5

| Kinetic Parameter | M1                  | M2/M3               | M4                  | Total |
|------------------|---------------------|---------------------|---------------------|-------|
| K<sub>m</sub>, µM | 143 ± 20            | 251 ± 26            | 519 ± 100           | —     |
| V<sub>max</sub>, pmol/min per mg | 26.7 ± 2.2 | 73.8 ± 5.0 | 881 ± 125 | —     |
| K<sub>i</sub>, µM | 1480 ± 316 | 1680 ± 320 | 1340 ± 430 | —     |
| CL<sub>mut</sub>, µL/min per mg | 0.187 | 0.294 | 1.7 | 2.18  |
| CL<sub>mut</sub>, mL/min per kg | 0.176 | 0.278 | 1.6 | 2.05  |
| Fractional proportion CL<sub>mut</sub>, % | 0.086 | 0.136 | 0.780 | —     |

Data expressed as the mean plus or minus standard error of mean from triplicate replicates; unbound fraction (f<sub>inc</sub>) of abrocitinib in human liver microsomes (0.3 mg/mL) was 1. f<sub>CL</sub>, unbound fraction; K<sub>i</sub>, inhibition constant for substrates exhibiting substrate inhibition kinetics; V<sub>max</sub>, maximum rate of metabolism.
areas were used to calibrate the abundance of M4 and M5 in the pooled plasma, urine, and feces sample extract profiles. The estimated mass ratio of M4 to M5 was determined to be 38.4, 49.4, and 13.3 for pooled plasma, urine, and fecal homogenate extracts, respectively. The combined [14C] peak area for the pooled plasma extracts, pooled urine, and pooled fecal homogenate extracts was 14.2, 19.2, and 3.6%, respectively. The calculated individual contribution of [14C] peak area for each metabolite (M4/M5) was 13.8/0.4% in pooled plasma extracts, 18.8/0.4% in pooled urine, and 3.3/0.3% in pooled fecal homogenate extracts.

**P450 Phenotyping.** The primary metabolites of abrocitinib were quantitated following a 30-minute incubation of 14C-abrocitinib with human hepatocytes for determination of fractional contribution to total clearance. The resulting radiochromatogram is presented in Supplemental Fig. 3, and the percentage of total radioactivity associated with the formation of metabolites is presented in Table 4. Based on the percentage of total radioactivity observed at the 30-minute time point, the respective fCL values were 0.390 for M1, 0.288 for M2/M3, 0.247 for M4, 0.0240 for M7, 0.011 for 358-1, 0.011 for 372-1, and 0.0291 for 149. Since M7 is likely a secondary metabolite of M1 (Fig. 4), the contribution of M7 metabolism was combined into the M1 route. Metabolites 358-1 and 372-1 are likely secondary metabolites of M4 (Fig. 4); therefore, the contributions were combined into the M4 route. The final fCL values for the 4 metabolic pathways characterized were 0.414 for M1, 0.288 for M2/M3, 0.269 for M4, and 0.0291 for 149. The proportion of metabolism attributed to P450 activity from these 4 pathways totaled 1.0.

Enzyme kinetic parameters and intrinsic clearance values determined for the formation of M1, M2/M3, and M4 from abrocitinib metabolism in human liver microsomes are listed in Table 5, and associated data are presented graphically in Supplemental Fig. 4. Kinetic parameters for metabolite 149 could not be determined. Visual inspection of the Eadie-Hofstee plots for the M1, M2/M3, and M4 reaction data demonstrated nonlinear profiles consistent with substrate inhibition. For all 3 metabolites, a Michaelis-Menten equation with substrate inhibition was selected for the determination of enzyme kinetic parameters. Based on these kinetic parameters, the respective M1, M2/M3, and M4 CLint,app values were 0.187, 0.294, and 1.70 μL/min per mg and CLint,app,sc values were 0.176, 0.278, and 1.60 mL/min per kg. The total CLint,app,sc of three major metabolites for abrocitinib was 2.05 mL/min per kg.

The results of P450 selective chemical inhibition on the metabolism of abrocitinib in human hepatocytes are presented in Table 6. The metabolic formation rates of M1, M2/M3, and M4 were primarily inhibited in the presence of PPP (2B6), tiensilic acid (2C9), esomeprazole (2C19), and troleandomycin (3A). After scaling, formation of metabolite M1 was inhibited 16% by PPP (2B6), 24% by tiensilic acid, 52% by esomeprazole, and 8.0% by troleandomycin. Formation of metabolite M2/M3 was inhibited 45% by tiensilic acid and 55% by esomeprazole. M4 was inhibited 25% by tiensilic acid, 57% by esomeprazole, and 18% by troleandomycin. A summary of the abrocitinib P450 assignment is presented in Table 7. After normalizing for fCL, for each metabolic pathway, the fraction of metabolic clearance of abrocitinib catalyzed by CYP2C19, 2C9, 3A4, and 2B6 were 0.53, 0.30, 0.11, and 0.066, respectively.

**In Vitro Pharmacology.** The enzyme potency of abrocitinib and the oxidative metabolites M1, M2, and M4 was assessed against four JAK isoforms using enzymatic assays at 1 mM adenosine triphosphate (Table 7). Abrocitinib was more potent against JAK1 versus JAK2 or TYK2 and demonstrated little potency for JAK3 (IC50 >10,000 nM). M1 and M2 exhibited a similar potency and selectivity profile as abrocitinib. The IC50 values for M4 with all 4 JAK isoforms were >10,000 nM. Therefore, the circulating metabolites M1 and M2 are active metabolites that may contribute to the overall pharmacology of abrocitinib, whereas M4 is pharmacologically inactive.

The cellular potency of abrocitinib, M1, and M2 was evaluated by measuring the inhibition of phosphorylation of STATs following stimulation with various cytokines in human whole blood (represented as total and unbound IC50), human keratinocytes, and THP-1 cells (IC50 values considered unbound) (Table 8). Abrocitinib was much more potent against cytokines that transduce their signals via JAK1-dependent pathways, with IC50 values ranging from 40.0 nM for IL-31–induced STAT3 phosphorylation to >16,500 nM (unbound) (Table 8). Abrocitinib was much more potent against cytokines that transduce their signals via JAK1-dependent pathways, with IC50 values ranging from 40.0 nM for IL-31–induced STAT3 phosphorylation to >16,500 nM (unbound) (Table 8). Abrocitinib was much more potent against cytokines at IC50 values of 0.36 for abrocitinib, 0.63 for M1, and 0.71 for M2. The

### TABLE 6

| Metabolite   | fCL (Scaled) | 2B6 | 2C9 | 2C19 | 3A4 |
|--------------|--------------|-----|-----|------|-----|
| M1 (M7)      | 0.41         | 16  | 24  | 52   | 8.0 |
| M2/M3        | 0.29         | 44  | 55  |      |     |
| M4 (358-1, 372-1) | 0.27       | 25  | 57  |      |     |
| 149          | 0.029        | 100 |     |      |     |
| Overall fCL  |              |    |     |      |     |

Nonstatistical contributions by the remaining P450 isoforms for M1 (1A2, 2C8, 2D6), M2/M3 (1A2, 2B6, 2C8, 2D6, 3A), M4 (1A2, 2B6, 2C8, 2D6), and 149 (1A2, 2B6, 2C8, 2C9, 2C19, 2D6) were observed and excluded from fraction metabolized determinations. Fraction metabolized was calculated as the product of fCL and fraction inhibited (scaled) as described in eq. 7.

### TABLE 7

| JAK Assay | Abrocitinib | M1 | M2 | M4 |
|-----------|------------|----|----|----|
| JAK1      | 29.2       | 43.4| 17.9| >10,000 |
| JAK2      | 803        | 1140| 886| >10,000 |
| JAK3      | >10,000    | >10,000| >10,000| >10,000 |
| TYK2      | 1250       | 3190| 1210| >10,000 |

*IC50 values obtained at 1 nM adenosine triphosphate.*
blood/plasma ratios for each drug species were determined to be 1.07 for abrocitinib, 1.13 for M1, and 1.27 for M2. Blood binding parameters were used to convert whole-blood pharmacology potency values into unbound values (Table 8).

**Discussion**

In this report, the pharmacokinetics and metabolism of abrocitinib were evaluated in healthy human participants. Recovery of the radiolabeled dose was 95%, indicating that the disposition of abrocitinib was well characterized. Oral absorption of abrocitinib was >90% and rapid, with peak concentrations observed at 0.5 hour, despite being identified as an MDR1 and BCRP efflux substrate. Absolute oral bioavailability was determined to be 60% and elimination half-life (t1/2) 2 hour. Systemic clearance was observed to be 64.2 L/h, and steady-state volume of distribution was 100 L. Clinical experience with abrocitinib has shown dose-proportional oral pharmacokinetics over a range of 30–400 mg, which supports linearity of parent and metabolite profiles. The t1/2 of both abrocitinib and total radioactivity profiles was similar, indicating formation rate limited kinetics of metabolites. Following a single oral dose of abrocitinib, the major circulating component of drug-related material in plasma was abrocitinib (~26%), followed by three monohydroxylated metabolites (M1, M2, and M4) at >10%. However, following multiple daily dosing of abrocitinib to steady state, analytical measurement of metabolites indicated that M2 and M4 were observed at >10%, with M1 being considered minor at <10% (data on file).

Disposition profiles indicated oxidative metabolism as the major pathway for clearance of abrocitinib, whereas the renal route of elimination was minor at <1%. All the metabolites of abrocitinib were consistent with oxidative P450 metabolism, with no observed evidence of direct conjugative mechanisms. In vitro phenotyping indicated multiple P450 enzymes involved in the metabolism of abrocitinib, with hepatic fraction of metabolism (fm) assignments of 0.53 for CYP2C19, 0.30 for CYP2C9, 0.11 for CYP3A4, and 0.06 for CYP2B6.

**TABLE 8**

| Cytokine | JAK Pair | pSTAT Assay | Cell Assay | Abrocitinib | M1 | M2 |
|----------|----------|-------------|------------|-------------|----|----|
| IL-4     | JAK1/JAK2 | pSTAT6      | Human keratinocytes | 77.0 | 433 | 134 |
| IL-13    | JAK1/JAK2 | pSTAT6      | Human keratinocytes | 81.9 | 236 | 84.1 |
| IL-22    | JAK1/JTYK2 | pSTAT3     | Human keratinocytes | 420  | 703 | 198 |
| IL-31    | JAK1/JAK2 | IFNγ-primed THP-1 | 40.0 | 79.6 | 56.0 |
| TSLP     | JAK1/JAK2 | pSTAT3      | hWB (CD3+ cells) | 1020/343 | 785/438 | 27/1152 |
| IFNγ     | JAK1/JAK2 | pSTAT3      | hWB (lymphocytes) | 174/58.5 | 296/165 | 90.5/50.6 |
| IFNγ     | JAK1/JAK2 | pSTAT1      | hWB (CD14+ cells) | 1690/569 | 1950/1090 | 2160/1210 |
| IL-6     | JAK1/JAK2 | pSTAT1      | hWB (CD3+ cells) | 343/115 | 171/95.3 | 136/76.0 |
| IL-12    | JAK2/JTYK2 | pSTAT4     | hWB (lymphocytes) | 9730/3270 | 33,400/18,600 | 5170/2890 |
| IL-15    | JAK1/JAK3 | pSTAT5      | hWB (lymphocytes) | 537/181 | 558/311 | 353/197 |
| IL-21    | JAK1/JAK3 | pSTAT3      | hWB (lymphocytes) | 516/174 | 844/471 | 487/272 |
| IL-23    | JAK2/JTYK2 | pSTAT3     | hWB (lymphocytes) | >16,500>5550 | 26,200/14,600 | 6210/3470 |
| IL-27    | JAK1/JAK2 | pSTAT3      | hWB (lymphocytes) | 228/76.7 | 382/213 | 23/131 |
| EPO      | JAK2/JAK2 | pSTAT5      | hWB (spiked CD34+ cells) | 7180/2420 | 9750/5440 | 9470/5290 |

B/P, blood/plasma ratio; EPO, erythropoietin; hWB, human whole blood; IC50,u, unbound IC50; pSTAT, phospho-signal transducer and activator of transcription; TSLP, thymic stromal lymphopoietin.

**Fig. 5.** Proposed mass balance model for abrocitinib following oral administration. CLp, plasma clearance; CLr, renal clearance; F, bioavailability; fafg, fraction absorbed × fraction surviving gut metabolism; Vss, steady state volume of distribution.
CYP2C9, 0.11 for CYP3A4, and ~0.06 for CYP2B6. These in vitro results are in agreement with the impact of strong CYP2C19 inhibition by fluvoxamine and flunoxazone on the exposure of coadministered abrocitinib (Wang et al., 2022b). Collectively, a mass balance model was assembled for abrocitinib and is shown in Fig. 5.

In contrast to abrocitinib, renal elimination was observed as the major pathway of excretion for metabolites. The fR of renal elimination was estimated to be 0.74 for M1, 0.94 for M2, and 0.93 for M4, based on clinical observation (Wang et al., 2022a). Active renal clearance was predicted to be >80% for each of the metabolites, with the balance from passive filtration (fP × glomerular filtration rate), based on estimates of metabolite exposure in plasma and total amount excreted in urine (total renal clearance = mass in urine/AUC). In vitro profiling showed that each of the primary metabolites was a substrate for the OAT3 transporter (internal data). A clinical study with the OAT3 inhibitor probenecid each of the primary metabolites was a substrate for the OAT3 trans-renal clearance.

Passive metabolite exposure in plasma and total amount excreted in urine (total filtration rate), based on estimates of metabolite exposure in plasma and total amount excreted in urine (total renal clearance = mass in urine/AUC). In vitro profiling showed that each of the primary metabolites was a substrate for the OAT3 transporter (internal data). A clinical study with the OAT3 inhibitor probenecid showed an AUCpass increase of approximately twofold for each of the metabolites in plasma (M1, 1.8-fold; M2, 2.2-fold; M4, 2.2-fold) (Wang et al., 2022b).

Abrocitinib and its metabolites M1 and M2 were pharmacologically active at inhibiting cytokine signaling through JAK1 homodimer pairs (i.e., JAK1/JAK2, JAK1/TYK2, and JAK1/JAK3), whereas M4 was inactive. The relative JAK selectivity was similar among abrocitinib, M1, and M2. Clinical assessment of the contribution of abrocitinib, M1, and M2 to circulating plasma pharmacology based on steady-state AUC and relative potency (Leclercq et al., 2009) indicated respective percentages of ~60%, ~10%, and ~30% (data on file). Dose adjustment of abrocitinib in the context of drug-drug interaction and in special populations (renal/hepatic impairment) will be based on the total contribution of abrocitinib, M1, and M2 pharmacology (i.e., total active moiety) (Wang et al., 2021; Wang et al., 2022a; Wang et al., 2022b).

In summary, these study results provide an important and comprehensive characterization of abrocitinib pharmacokinetics, metabolism, and clearance mechanisms as well as disposition and pharmacology of major metabolites. Furthermore, the disposition characterization of abrocitinib and its major metabolites has been consistent with clinical experience in drug-drug interaction studies and special populations.

Upon request, and subject to review, Pfizer will provide the data that support the findings of this study. Subject to certain criteria, conditions, and exceptions, Pfizer may also provide access to the related individual deidentified participant data. See https://www.pfizer.com/science/clinical-trials/trial-data-and-results for more information.

Clinicaltrials.gov identifier: NCT03250039.

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Authorship Contributions

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The Pharmacokinetics, Metabolism, and Clearance Mechanisms of Abrocitinib, a Selective Janus Kinase Inhibitor, in Humans

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Drug Metabolism and Disposition

DMD-AR-2022-000829
Supplemental Methods

Cytokine-induced STAT Phosphorylation Assays. A total of 14 cytokines was used to induce signal transducer and activator of transcription (STAT) phosphorylation in human whole blood, human keratinocytes, or human acute monocytic leukemia (THP-1) cells. The method of cytokine-induced STAT phosphorylation in human whole blood was previously described in detail by Dowty et al (2019). Human primary keratinocytes were used in the IL-4, IL-12, and IL-22-induced STAT phosphorylation assays. Human primary keratinocytes were cultured in DermaLife medium with supplement kit (Lifeline Cell Technology) to expand cell populations. Cell passages 2 to 5 were used in this study. Keratinocytes were suspended in warm DermaLife medium and aliquoted in 96-well, deep-well, V-bottom plates. Cells were treated with compounds (0.0003 to 20 µM) for 60 minutes and followed by stimulation with IL-4 (2 ng/mL) or IL-13 (20 ng/mL) for 15 minutes. For the IL-22 assay, keratinocytes were seeded in 24-well plates and cultured overnight. Cells were switched to DermaLife medium, treated with compounds (0.0003 to 20 µM) for 60 minutes, and then stimulated with IL-22 for 30 minutes. Cells were detached by the treatment of 0.25% trypsin/EDTA. Cytokine stimulated cells were fixed by 2% paraformaldehyde and permeabilized by 90% methanol. Fixed and permeabilized cells were stained with AlexaFluor647 labeled anti-pSTAT6 antibody for IL-4- and IL-13-treated cells, and AlexaFluor647 labeled anti-pSTAT3 antibody for IL-22-treated cells. THP-1 cells were maintained in RPMI 1640 medium containing 10% fetal bovine serum, 50 mM 2-mercaptoethanol, 50 U/mL penicillin, 50 mg/mL streptomycin, and 2 mM L-glutamine. THP-1 cells were treated with interferon (IFN)γ (20 ng/mL) for 18 hours. IFNγ-primed THP-1 cells were resuspended in fresh RPMI 1640 medium, treated with compounds (0.0003-20 µM) for 60 minutes, followed by stimulation with IL-31(1 mg/mL) for additional 10 minutes. Cells were
then fixed by 2% paraformaldehyde and permeabilized by 90% methanol. Fixed and permeabilized cells were stained with AlexaFluor647 labeled anti-pSTAT3 antibody. Fluorescence was analyzed with a LSRFortessa equipped with a plate-based autosampler. Data was analyzed using FACSDiva version 8.0.
Table S1. LC-MS/MS Conditions for Metabolite Profiling and $^{14}$C-Abrocitinib Quantitation

| Parameter    | Description                                                                 |
|--------------|-----------------------------------------------------------------------------|
| Mass spectrometer | Q Exactive™ Hybrid Quadrupole-Orbitrap (Thermo Fisher Scientific, Waltham, MA) |
| UHPLC        | Acquity UHPLC system (Waters, Milford, MA)                                  |
| Detection mode | Electrospray ionization (ESI), Positive Mode                                |
| Column       | Waters Acquity UHPLC-HSS C18 2.1x100 1.8 μm, (Waters Corporation) maintained at 40°C. |
| Mobile phase A | 2 mM ammonium acetate:acetonitrile:methanol (95:2.5:2.5 % v/v)             |
| Mobile phase B | 2 mM ammonium acetate:acetonitrile:methanol (10:45:45 % v/v)               |
| Gradient     | Time (min) | Flow Rate (µL/min) | Mobile Phase A (%) | Mobile Phase B (%) |
|              | 0.0        | 400               | 95                | 5                |
|              | 1.25       | 400               | 95                | 5                |
|              | 5.0        | 400               | 80                | 20               |
|              | 7.5        | 400               | 80                | 20               |
|              | 8.75       | 400               | 65                | 35               |
|              | 12.5       | 400               | 55                | 45               |
|              | 13.75      | 400               | 0                 | 100              |
|              | 15.65      | 400               | 0                 | 100              |
|              | 16.0       | 400               | 95                | 5                |
|              | 28.50      | 400               | 95                | 5                |
| Software     | Xcalibur v2.2 sp1.48 (Thermo Fisher Scientific)                             |
**Table S2.** LC-MS/MS Conditions for Isolation of Metabolites M6 and M7 - Initial Assessment

| Parameter       | Description                                                                 |
|-----------------|-----------------------------------------------------------------------------|
| Mass spectrometer | LTQ Orbitrap XL™ Hybrid Ion Trap (Thermo Fisher Scientific)                  |
| HPLC            | Surveyor HPLC System (Thermo Fisher Scientific)                             |
| Detection mode  | Electrospray ionization (ESI), Positive Mode                                |
| Column          | Luna C18 150 × 4.6 mm column (Agilent, Santa Clara, CA)                     |
| Mobile phase    | C 10 mM ammonium acetate                                                    |
| Mobile phase    | D methanol:acetonitrile (50:50 % v/v)                                       |

**Gradient**

| Time (min) | Flow Rate (µL/min) | Mobile Phase C (%) | Mobile Phase D (%) |
|------------|--------------------|--------------------|--------------------|
| 0.0        | 500                | 95                 | 5                  |
| 5.0        | 500                | 95                 | 5                  |
| 20         | 500                | 5                  | 95                 |
| 25         | 500                | 5                  | 95                 |
| 26         | 500                | 95                 | 5                  |
| 30         | 500                | 95                 | 5                  |

Software: Xcalibur v2.2 sp1.48
Table S3. LC-MS/MS Conditions for Isolation of Metabolites M6 and M7 - Initial Fractionation

| Parameter                  | Description                                                                 |
|----------------------------|------------------------------------------------------------------------------|
| Mass spectrometer          | LTQ Orbitrap XL™ Hybrid Ion Trap Mass Spectrometer                           |
| HPLC                       | Surveyor HPLC System                                                         |
| Fraction Collector         | Gilson FC 203B                                                               |
| Detection mode             | Electrospray ionization (ESI), Positive Mode                                 |
| Column                     | YMC Ph-Pack 250 × 4.6 mm column (YMC America, Devens, MA)                   |
| Mobile phase A             | 0.1% formic acid in water                                                    |
| Mobile phase D             | Methanol:acetonitrile (50:50 % v/v)                                          |
| Gradient                   | Time (min) Flow Rate Mobile Phase Mobile Phase (μL/min A % D %)              |
| 0.0                        | 1000 95 5                                                                  |
| 5                          | 1000 95 5                                                                  |
| 45                         | 1000 60 40                                                                 |
| 50                         | 1000 5 95                                                                 |
| 54                         | 1000 5 95                                                                 |
| 55                         | 1000 95 5                                                                  |
| 60                         | 1000 95 5                                                                  |
| Software                   | Xcalibur v2.2 sp1.48                                                         |
**Table S4.** LC-MS/MS Conditions for Isolation of Metabolites M6 and M7 - Analysis of Fractions

| Parameter         | Description                                                                 |
|-------------------|-----------------------------------------------------------------------------|
| Mass spectrometer | LTQ Orbitrap XL™ Hybrid Ion Trap (Thermo Fisher Scientific)                  |
| HPLC              | Surveyor HPLC System (Thermo Fisher Scientific)                             |
| Detection mode    | Electrospray ionization (ESI), Positive Mode                                |
| Column            | Zorbax SB-Phenyl 150 × 4.6 mm column (Agilent)                              |
| Mobile phase A    | 0.1% formic acid in water                                                   |
| Mobile phase D    | Methanol:acetonitrile (50:50 % v/v)                                         |
| Gradient          |                                                                             |
| Time (min)        | Flow Rate Mobile Phase Mobile Phase                                       |
|                   | (µL/min) A (%) D (%)                                                        |
| 0.0               | 500 95 5                                                                 |
| 1                 | 500 95 5                                                                 |
| 12                | 500 5 95                                                                  |
| 15                | 500 5 95                                                                  |
| 16                | 500 95 5                                                                 |
| 20                | 500 95 5                                                                 |
| Software          | Xcalibur v2.2 sp1.48                                                       |
**Table S5.** LC-MS/MS Conditions for Isolation of Metabolites M6 and M7 – Final Fractionation

| Parameter                  | Description                                                                 |
|----------------------------|------------------------------------------------------------------------------|
| Mass spectrometer          | LTQ Orbitrap XL™ Hybrid Ion Trap Mass Spectrometer                           |
| HPLC                       | Surveyor HPLC System                                                         |
| Fraction Collector         | Gilson FC 203B                                                               |
| Detection mode             | Electrospray ionization (ESI), Positive Mode                                |
| Column                     | Zorbax SB-Pheny1 150 × 4.6 mm column (Agilent)                               |
| Mobile phase C             | 10 mM ammonium acetate                                                      |
| Mobile phase D             | Methanol:acetonitrile (50:50 % v/v)                                          |
| Gradient                   | Time (min)  | Flow Rate  | Mobile Phase C (%) | Mobile Phase D (%) |
|                            | 0.0        | 1000       | 98                | 2                 |
|                            | 5          | 1000       | 98                | 2                 |
|                            | 45         | 1000       | 80                | 20                |
|                            | 50         | 1000       | 5                 | 95                |
|                            | 54         | 1000       | 5                 | 95                |
|                            | 55         | 1000       | 98                | 2                 |
|                            | 60         | 1000       | 98                | 2                 |
| Software                   | Xcalibur v2.2 sp1.48                                                         |
Table S6. LC-MS/MS Conditions for Abrocitinib in Plasma

| Parameter          | Description                                                                 |
|--------------------|-----------------------------------------------------------------------------|
| Mass spectrometer  | SCIEX Triple Quad™ API-5500 (SCIEX, Framingham, MA)                         |
| HPLC               | Acquity UHPLC system                                                         |
| Detection mode     | Electrospray ionization (ESI), Positive Mode                                |
| Column             | Aquasil C18 2.1 × 50 mm, 3 μm (Thermo Fisher Scientific)                    |
| Mobile phase A     | A 0.1% formic acid and 10 mM ammonium acetate in acetonitrile:water (10:90 % v/v) |
| Mobile phase B     | B 0.1% formic acid and 10 mM ammonium acetate in acetonitrile:water (90:10 % v/v) |
| Gradient           | Time (min)  | Flow Rate (μL/min) | Mobile Phase A (%) | Mobile Phase D (%) |
|                    | 0.0        | 1000              | 85                  | 15                 |
|                    | 0.99       | 1000              | 85                  | 15                 |
|                    | 1.0        | 1000              | 0                   | 100                |
|                    | 2.0        | 1000              | 0                   | 100                |
|                    | 2.01       | 1000              | 85                  | 15                 |
|                    | 3.0        | 1000              | 85                  | 15                 |
| Software           | Analyst TM version 1.4.2 (SCIEX)                                            |
|                    | Watson LIMS 7.2.0.02 (Thermo Fisher Scientific)                             |
| MRM transitions | Analyte                        | Q1   | Q3   |
|-----------------|--------------------------------|------|------|
|                 | Abrocitinib (PF-04965842)      | 324.4| 134.2|
|                 | d2-Abrocitinib (PF-06651703)   | 327.2| 178.2|
| Parameter               | Description                                                                                                                                 |
|------------------------|---------------------------------------------------------------------------------------------------------------------------------------------|
| Mass spectrometer      | SCIEX TripleTOF® 6600 quadrupole time-of-flight (QTOF) mass analyzer (SCIEX)                                                                |
| HPLC                   | Agilent 1290 HPLC System (Agilent)                                                                                                          |
| Fraction Collector     | PAS HTS-xt fraction collector (Leap Technologies, Morrisville, NC)                                                                                |
| Detection mode         | Electrospray ionization (ESI), Positive Mode                                                                                               |
| Column                 | Acquity UHPLC-HSS C18 (2.1 × 100 mm, 1.7 μm) maintained at 40°C                                                                               |
| Mobile phase A         | 2 mM ammonium acetate:acetonitrile:methanol (95:2.5:2.5 % v/v)                                                                               |
| Mobile phase B         | 2 mM ammonium acetate:acetonitrile:methanol (10:45:45 % v/v)                                                                               |
| Gradient               |                                                                                                                                            |
| Time (min)             | Flow Rate | Mobile Phase A (%) | Mobile Phase D (%) |
|                        | (μL/min)  |                  |                   |
| 0.0                    | 400       | 95                | 5                 |
| 1.25                   | 500       | 95                | 5                 |
| 5.0                    | 400       | 80                | 20                |
| 7.5                    | 500       | 80                | 20                |
| 8.75                   | 400       | 65                | 35                |
| 12.5                   | 400       | 55                | 45                |
| 13.75                  | 400       | 0                 | 100               |
| 15.65                  | 400       | 0                 | 100               |
| 16.0                   | 400       | 95                | 5                 |
| 28.0                   | 400       | 95                | 5                 |
| Parameter  | Description              |
|------------|--------------------------|
| Software   | Analyst TF 1.7.1 (SCIEX) |
Table S8. LC-MS/MS conditions for in vitro CYP450 assignment (Enzyme kinetics and chemical inhibition)

| Parameter               | Description                                                                 |
|-------------------------|-----------------------------------------------------------------------------|
| Mass spectrometer       | SCIEX Triple Quad™ API-5500                                                 |
| HPLC pump               | Shimadzu LC-30AD (Shimadzu, Columbia, MD)                                   |
| Autosampler             | Leap CTC HTS PAL (Leap Technologies)                                        |
| Detection mode          | Electrospray ionization (ESI), Positive Mode                               |
| Column                  | ACQUITY UHPLC BEH C18 column, 130Å, 1.7 µm, 2.1 mm × 50 mm (Waters)         |
| Injection volume        | 3 µL                                                                        |
| Mobile phase A          | 0.1% formic acid in water                                                   |
| Mobile phase B          | 0.1% formic acid in acetonitrile                                            |
| Flow rate (mL/min)      | 0.5                                                                         |
| Gradient                |                                                                             |
|                         | Time  | Flow Rate (µL/min) | Mobile Phase A (%) | Mobile Phase B (%) |
|                         | (min) |                  |                  |                  |
| 0.01                    | 400   | 95                | 5                |
| 0.16                    | 500   | 95                | 5                |
| 0.63                    | 400   | 80                | 20               |
| 0.94                    | 500   | 80                | 20               |
| 1.09                    | 400   | 65                | 35               |
| 1.56                    | 400   | 55                | 45               |
| 1.72                    | 400   | 0                 | 100              |
| 3                       | 400   | 0                 | 100              |
| Parameter | Description |
|-----------|-------------|
| 3.01      | 400         | 95 | 5 |
| 4         | 400         | 95 | 5 |
| MRM transitions | | |
| Analyte  | Q1 | Q3 | Polarity |
| Abrocitinib (PF-04965842) | 324.2 | 149.1 | Positive |
| M1 (PF-06471658) | 340.1 | 149.1 | Positive |
| M2/M3 (PF-07055087/PF-07055090) | 340.1 | 149.1 | Positive |
| M4 (PF-07058474) | 340.1 | 217.1 | Positive |
| Diclofenac (IS) | 296.0 | 215.0 | Positive |
| 372-1 | 372.2 | 328.1 | Positive |
| 358-1 | 358.3 | 340.0 | Positive |
| 356-1 | 356.3 | 181.0 | Positive |
| M7 (PF-067377821) | 354.3 | 149.0 | Positive |

**Analyte concentration range** 1.0-2000 nM

**Software** Analyst 1.6

Q1, first quadrupole; Q3, third quadrupole.
Table S9. LC-MS/MS conditions for abrocitinib in plasma protein binding and blood-to-plasma ratio studies

| Mass Spectrometer and Source Type | AB Sciex API 4000 Triple Quadrupole Electrospray |
|----------------------------------|--------------------------------------------------|
| HPLC pumps                       | Agilent 1290                                     |
| Autosampler                      | CTC HTS PAL                                      |
| Injection volume                 | 25 μL                                            |
| Loading column                   | Halo Fused-core 2.7μ C18 100Å (5 x 4.6 mm ID)    |
| Loading solvent                  | 90% Water, 10% Methanol (containing 0.1% Trifluoroacetic Acid) |
| Mobile phase A                   | 5 mM Ammonium Acetate (containing 0.075% Formic Acid) |
| Mobile phase B                   | Acetonitrile                                     |
| Flowrate (mL/min)                | 1.0                                              |

| Gradient | Time (min) | Mobile Phase A (%) | Mobile Phase B (%) |
|----------|------------|--------------------|--------------------|
|          | 0.00       | 90                 | 10                 |
|          | 0.40       | 90                 | 10                 |
|          | 1.20       | 10                 | 90                 |
|          | 2.20       | 10                 | 90                 |
|          | 2.25       | 90                 | 10                 |
|          | 2.80       | 90                 | 10                 |

| Column | Kinetex XB 2.6μ C18 100Å (50 x 3 mm ID) |
|--------|----------------------------------------|
| Detection mode | Positive ion MRM |
| Data collection software/version | Analyst 1.6.2 |
| Data analysis software/version | Analyst 1.6.2 |

| MRM transitions | Compound | Q1     | Q3     |
|-----------------|----------|--------|--------|
|                 | Abrocitinib | 324.0  | 149.1  |
|                 | Chloroquine (PF-00345351) | 320.1  | 247.0  |
|                 | Internal standard (Tolbutamide) | 271.2  | 172.0  |

HPLC, high-performance liquid chromatography; MRM, multiple reaction monitoring.
**Table S10.** LC-MS/MS conditions for M1 in plasma protein binding and blood-to-plasma ratio studies

| Mass Spectrometer and Source Type | AB Sciex API 4000 Triple Quadrupole Electrospray |
|----------------------------------|--------------------------------------------------|
| HPLC pumps                       | Shimadzu LC-20AB Binary Pumps                    |
| Autosampler                      | CTC HTS PAL                                      |
| Injection volume                 | 50 μL                                            |
| Loading column                   | Halo Fused-core 2.7μ C18 100Å (5 x 4.6 mm ID)    |
| Loading solvent                  | 90% Water, 10% Methanol (containing 0.1% Trifluoroacetic Acid) |
| Mobile phase A                   | 5 mM Ammonium Acetate (containing 0.075% Formic Acid) |
| Mobile phase B                   | Acetonitrile                                     |
| Flowrate (mL/min)                | 1.0                                              |

| Gradient | Time (min) | Mobile Phase A (%) | Mobile Phase B (%) |
|----------|------------|--------------------|--------------------|
| 0.00     | 90         | 10                 |
| 0.30     | 90         | 10                 |
| 1.00     | 10         | 90                 |
| 2.00     | 10         | 90                 |
| 2.05     | 90         | 10                 |
| 3.00     | 90         | 10                 |

| Column | Kinetex XB 2.6μ C18 100Å (50 x 3 mm ID) |
|--------|----------------------------------------|
| Detection mode | Positive ion MRM |
| Data collection software/version | Analyst 1.6.2 |
| Data analysis software/version | Analyst 1.6.2 |

| MRM transitions | Compound | Q1     | Q3     |
|-----------------|----------|--------|--------|
| M1              | 340.2    | 149.1  |
| Chloroquine (PF-00345351) | 320.2 | 247.0  |
| Internal standard (Tolbutamide) | 271.2 | 172.0  |

HPLC, high-performance liquid chromatography; MRM, multiple reaction monitoring.
**Table S11.** LC-MS/MS conditions for M2 in plasma protein binding and blood-to-plasma ratio studies

| Mass Spectrometer and Source Type | AB Sciex API 4000 Triple Quadrupole Electrospray |
|-----------------------------------|-----------------------------------------------|
| HPLC pumps                        | Shimadzu LC-20AB Binary Pumps                 |
| Autosampler                       | CTC HTS PAL                                   |
| Injection volume                  | 50 μL                                         |
| Loading column                    | Halo Fused-core 2.7μ C18 100Å (5 x 4.6 mm ID) |
| Loading solvent                   | 90% Water, 10% Methanol (containing 0.1% Trifluoroacetic Acid) |
| Mobile phase A                    | 5 mM Ammonium Acetate (containing 0.075% Formic Acid) |
| Mobile phase B                    | Acetonitrile                                  |
| Flowrate (mL/min)                 | 1.0                                           |

| Gradient | Time (min) | Mobile Phase A (%) | Mobile Phase B (%) |
|----------|------------|--------------------|--------------------|
|          | 0.00       | 90                 | 10                 |
|          | 0.30       | 90                 | 10                 |
|          | 1.00       | 10                 | 90                 |
|          | 2.00       | 10                 | 90                 |
|          | 2.05       | 90                 | 10                 |
|          | 3.00       | 90                 | 10                 |

| Column | Kinetex XB 2.6μ C18 100Å (50 x 3 mm ID) |
|---|---|
| Detection mode | Positive ion MRM |
| Data collection software/version | Analyst 1.6.2 |
| Data analysis software/version | Analyst 1.6.2 |

| MRM transitions | Compound               | Q1    | Q3    |
|-----------------|------------------------|-------|-------|
|                 | M2                     | 340.1 | 149.1 |
|                 | Chloroquine (PF-00345351) | 320.2 | 247.0 |
|                 | Internal standard (Tolbutamide) | 271.2 | 172.0 |

HPLC, high-performance liquid chromatography; MRM, multiple reaction monitoring.
Table S12. Incidence of treatment-emergent mild adverse events

| Number of Subjects with AE by MedRA Preferred Term | Period A (N=6) | Period B (N=5) |
|---------------------------------------------------|---------------|---------------|
| Dizziness                                         | 3             | 2             |
| Headache                                          | 1             | 0             |
| Nausea                                            | 2             | 2             |
| Dyspepsia                                         | 1             | 0             |
| Oral paresthesia                                  | 1             | 0             |
Table S13. Diagnostic MS product ions used for abrocitinib and proposed metabolite structures

| ID  | Structure and Product Ions | Ion m/z  | Diagnostic Fragment Ions               |
|-----|-----------------------------|----------|----------------------------------------|
| Abrocitinib | ![Structure](image) | 324.1490 | 201, 176, 175, 149, 70                  |
| M6   | ![Structure](image)         | 370.1179 | 217, 165                                |
| 340-5 | ![Structure](image)        | 340.1074 | 296, 218, 201, 175, 149                 |
| 372-1 | ![Structure](image)        | 372.1334 | 176, 70                                 |
| M7   | ![Structure](image)         | 354.1230 | 218, 201, 175, 149, 70                  |
M2

340.1438  218, 201, 175, 149, 70

M3

340.1438  218, 201, 175, 149, 70

M4

340.1438  234, 217, 191, 176, 165, 70

M5

310.1332  204, 187, 176, 161, 135, 70
**Figure S1.** $^{1}H-^{13}C$ HMBC spectrum (left) and $^{1}H-^{1}H$ COSY spectrum (right) of M6 (PF-07095462, 370, m/z 370) metabolite isolated from pooled human urine.

COSY, homonuclear correlation spectroscopy; HMBC, heteronuclear multiple bond correlation spectroscopy.
**Figure S2.** $^1$H-$^{13}$C HMBC spectrum (left) and $^1$H-$^1$H COSY spectrum (right) of M7 (PF-06737821, 354-1, m/z 354) metabolite isolated from pooled human urine.

COSY, homonuclear correlation spectroscopy; HMBC, heteronuclear multiple bond correlation spectroscopy.
Figure S3. Radiochromatogram following 30-minute incubation of 1 µM $^{14}$C-abroctinib (PF-04965842) in human hepatocytes (0.75 million cells/mL)
**Figure S4.** Enzyme kinetic plots for primary metabolites M1 (A), M2/M3 (B), M4 (C), and 149 (D) and secondary metabolites M7 (E), 358-1 (F), and 372-1 (G) from abrocitinib metabolism

A. Enzyme kinetics of primary metabolite M1 (PF-06471658) formation following incubation of abrocitinib (PF-04965842) with human liver microsomes

B. Enzyme kinetics of primary metabolite M2/M3 (PF-07055087/PF-07055090) formation following incubation of abrocitinib (PF-04965842) with human liver microsomes
C. Enzyme kinetics of primary metabolite M4 (PF-07054874) formation following incubation of PF-04965842 with human liver microsomes

![PF-04965842 Michaelis-Menten Plot](image1)

![PF-04965842 Eadie-Hofstee Plot](image2)

D. Enzyme kinetics of primary metabolite m/z 149 formation following incubation of abrocitinib (PF-04965842) with human liver microsomes

![PF-04965842 Michaelis-Menten Plot](image3)

![PF-04965842 Eadie-Hofstee Plot](image4)
E. Enzyme kinetics of secondary metabolite M7 (PF-06737821) formation following incubation of abrocitinib (PF-04965842) with human liver microsomes

![Graph depicting enzyme kinetics of M7 formation](image1)

F. Enzyme kinetics of secondary metabolite 358-1 formation following incubation of abrocitinib (PF-04965842) with human liver microsomes

![Graph depicting enzyme kinetics of 358-1 formation](image2)
G. Enzyme kinetics of secondary metabolite 372-1 formation following incubation of abrocitinib (PF-04965842) with human liver microsomes