The development and application of geochemical techniques to identify redox conditions in modern and ancient aquatic environments has intensified over recent years. Iron (Fe) speciation has emerged as one of the most widely used procedures to distinguish different redox regimes in both the water column and sediments, and is the main technique used to identify oxic, ferruginous (anoxic, Fe(II) containing) and euxinic (anoxic, sulfidic) water column conditions. However, an international sediment reference material has never been developed. This has led to concern over the consistency of results published by the many laboratories that now utilise the technique. Here, we report an interlaboratory comparison of four Fe speciation reference materials for palaeoredox analysis, which span a range of compositions and reflect deposition under different redox conditions. We provide an update of extraction techniques used in Fe speciation and assess the effects of both test portion mass, and the use of different analytical procedures, on the quantification of different Fe fractions in sedimentary rocks. While atomic absorption spectroscopy and inductively coupled plasma-optical emission spectrometry produced comparable Fe measurements for all extraction stages, the use of ferrozine consistently underestimated Fe in the extraction step targeting mixed ferrous–ferric minerals such as magnetite. We therefore suggest that the use of ferrozine is discontinued for this Fe pool. Finally, we report the combined data of four independent Fe speciation laboratories to characterise the Fe speciation composition of the reference materials. These reference materials are available to the community to provide an essential validation of in-house Fe speciation measurements.

Keywords: iron speciation, sequential extraction, reference materials, water column redox, total iron, ancient sediments.

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FeHR enrichments arise due to precipitation of non-sulfidised Fe minerals such as Fe-(oxyhydr)oxides (e.g., Sun et al. 2015), green rust and magnetite (Zegeye et al. 2012). Fe carbonate minerals (e.g., Jiang and Tosca 2019) or potentially Fe silicates (e.g., Rasmussen et al. 2015). Recognising that magnetite and Fe carbonate minerals such as siderite were not extracted by existing techniques, Poulton and Canfield (2005) further refined the iron speciation methodology. This resulted in the development of a sequential extraction procedure to determine magnetite (Fe₃mag) and iron-carbonate (Fe₃cₗke) minerals, in addition to the previously identified Fe₃oxy, Fe₃py and Fe₃gr pools (with Fe₃IR calculated as the sum of Fe₃cₗke, Fe₃mag and Fe₃py).

Based on observations from the Black Sea (Anderson and Raiswell 2004), Poulton et al. (2004a) developed the utility of Fe speciation further, by utilising the Fe₃py/Fe₃oxy ratio to distinguish euxinic (sulfidic) and ferruginous (containing dissolved Fe²⁺) depositional conditions. In addition, noting that rapid deposition of terrigenous sediment and/or transfer of Fe₃oxy to Fe₃py, under anoxic non-sulfidic conditions can both decrease depositional Fe₃oxy/Fe₃py ratios (to potentially give a false oxic signal under anoxic depositional conditions), Poulton and Canfield (2011) revised the calibration boundaries. Thus, oxic depositional conditions are now commonly recognised by Fe₃oxy/Fe₃py < 0.22, ferruginous conditions are characterised by Fe₃oxy/Fe₃py > 0.38 and Fe₃py/Fe₃oxy < 0.7–0.8, and euxinic conditions are characterised by Fe₃oxy/Fe₃py > 0.38 and Fe₃py/Fe₃oxy > 0.7–0.8. When Fe₃oxy/Fe₃py ratios are between 0.22 and 0.38, an “equivocal” zone is recognised, where additional consideration is required to evaluate water column redox conditions. In particular, Fe₃oxy concentrations and Fe₃py/Fe₃oxy ratios (Poulton et al. 2010, Cumming et al. 2013, Doyle et al. 2018), and Fe₃/Al ratios (Lyons and Severmann et al. 2006, Clarkson et al. 2014) may be used to identify whether transfer of Fe₃oxy to Fe₃py has lowered initial depositional Fe₃oxy concentrations.

As a consequence of these developments, the iron speciation scheme of Poulton and Canfield (2005) has become widely used for evaluating palaeoredox depositional conditions. However, while individual laboratories commonly use their own in-house reference materials as a procedural check, there is concern that discrepancies in operational procedures across different laboratories may be producing inconsistent results. Consequently, there is a clear requirement for a set of international reference materials. Here, we report the development of four reference materials for assessing ancient water column redox conditions via Fe speciation. This is based on the results of four independent laboratories, including the laboratories of the authors who developed and calibrated...
the Fe speciation technique that is now widely used (Poulton and Canfield 2005). We additionally present details of the methodology applied and discuss operational issues related to the technique.

**Experimental procedure**

**Samples**

Four marine shale samples (WHIT, KL133, KL134 and BHW) were selected to encompass a range of iron phase compositions, depositional settings and periods of Earth history. WHIT was collected from the Mulgrave Shale Member of the Whitby Mudstone Formation at Saltwick Bay, Whitby, UK (Simms et al. 2004). The sample is early Jurassic (Toarcian; ~ 183 Ma) in age (Simms et al. 2004) and is a fine-grained, laminated, organic carbon-rich mudstone thought to have been deposited in an anoxic water column (Wignall et al. 2005). KL133 and KL134 were collected from well-preserved drill core (borehole KL1/65) at the National Core Library, Donkerhoek, South Africa. These two Late Permian (Catuneanu et al. 2005, Branch et al. 2007) samples are from below and above the occurrence of the Upper Ecca microfloras of the Ecca and Beaufort Groups (Linol et al. 2016, Chere et al. 2017). KL133 (1025 m depth in core KL1/65) is from just beneath the Upper Ecca microflora and is comprised of grey-black silty shale (Linol et al. 2016). KL134 (104 m depth in core KL1/65) is a light-grey siltstone from above the microfloras (Linol et al. 2016). While there is ongoing debate as to the absolute ages of the Ecca and Beaufort Groups, the two samples were deposited at ~265 Ma (e.g., McKay et al. 2015, Linol et al. 2016). BHW is a partially silicified, dolomitic black shale of the Archaean (~2.6 Ga) Black Reef Quartzite Formation, Transvaal Super Group. The sample was taken from well-preserved drill core (62.5 m depth in core BHW-289) stored at the National Core Library, Donkerhoek, South Africa.

**Sample preparation and storage**

Post-collection, weathered surfaces were removed and rocks were crushed at the University of Leeds using an agate TEMA pulverising mill, to obtain powder with the consistency of flour and without any larger isolated mineral grains. Initial attempts to sieve several of the samples were found to be problematic, due to coagulation of clay minerals during the procedure, which prevented adequate sieving and altered the nature of the sieved sediment. Thus, to ensure homogeneity of each entire bulk sample, powders were well-mixed via the repetitive use of a v-splitter, before decantation into acid-clean jars containing ~100 g of rock powder. For longer term storage, samples are preserved under a nitrogen atmosphere at a constant temperature of 20 °C to prevent sample oxidation. For short-term storage, we recommend that samples are kept in a desiccator, either under vacuum or under an anaerobic atmosphere, to minimise potential oxidation and to retain a low moisture content (Kane and Potts 2007).

**Organic carbon analyses**

Total organic carbon (TOC) was determined at the University of Leeds. Samples (n = 12 for each reference material) of approximately 0.5 g were initially decarbonated with 10 ml of 20% v/v HCl for one hour. This was performed in 15 ml centrifuge tubes, which were left open to allow for CO₂ degassing. After centrifugation, the supernatant was decanted, and samples were then treated with a further 10 ml of 20% v/v HCl, followed by constant shaking at room temperature for 16 h. Following this, the supernatant was decanted, and 10 ml of high-purity water (from a Milli-Q® system, Molsheim, France) was added to the samples and agitated for 30 min. The samples were then repeatedly washed with high-purity water until the supernatant reached pH > 4. The samples were then left to dry overnight, and TOC was measured using a LECO carbon–sulphur analyser, with LECO's certified carbon soil used as an internal reference material. This internal reference material had a recovery of 101.03% TOC and a reproducibility (RSD) of 1.60% (n = 8).

**Major element determinations**

Major element determinations were performed at the University of Oldenburg and the University of Leeds using wavelength dispersive X-ray fluorescence spectrometry. At ICBM, borate glass beads were produced by fusing 0.7 g of sample with 4.2 g of Li₂B₄O₇, following a peroxidation procedure with 1.0 g of (NH₄)₂NO₃ in a platinum crucible. Samples were then analysed using a Panalytical AxiosMAX spectrometer. At the University of Leeds, glass beads were produced by fusing 0.4 g of sample with 4 g of flux (66% Li₂B₄O₇ + 34% UBO₃) and two drops of lithium iodide solution (250 g l⁻¹) in a platinum crucible, and samples were measured using a Rigaku ZSX Primus II spectrometer. Calibration, including line overlap correction and matrix correction, was based on international reference samples (66, ICBM; 70, University of Leeds). Accuracy was checked by international and in-house reference materials not included in the calibration, with an error for major elements of <6% at ICBM and <3% at the University of Leeds. Measurement precision was <1% for major elements.
Iron extractions

All iron extractions were conducted under oxic conditions using Analytical Reagent grade chemicals, and each analyst performed a batch of eight replicates of each extraction. The sequential extractions and pyrite dissolutions were performed at four independent Fe speciation laboratories, including the Cohen Laboratory at the University of Leeds (three different analysts), the NordCEE Laboratory at the University of Southern Denmark, the Sediment and Aqueous Geochemistry Laboratory at the University of Copenhagen, and the Marine Geosystems Laboratory at the GEOMAR Helmholtz Centre for Ocean Research. The broad target phases for each extraction are reported in Table 1. However, it should be noted that these are operationally defined extractions, and the Fe speciation technique for palaeoredox analysis is predicated on the reactivity of different Fe pools towards dissolved sulfide, rather than the quantification of specific Fe minerals, which is a common misconception. Thus, the precise minerals extracted in each step (and the extent of their dissolution) will vary dependent on mineral crystallinity (see Raiswell et al. 1994) and a host of other factors, including impurities within the structure (see Poulton et al. 2004b). However, the Fe dissolved in each extraction can be considered to comprise an iron pool of similar reactivity towards dissolved sulfide, and it is this factor that has been calibrated in the use of Fe speciation as a palaeoredox indicator.

The sequential iron extractions (steps a–c; Table 1) were performed with a standard test portion mass of 60 ± 10 mg (accurately weighed), but tests were also performed using a test portion of up to 100 mg. Extraction solutions were prepared at room temperature, and the extractant volume for each step was 10 ml. During extractions, samples were constantly agitated (either horizontally on a shaking table or via an overhead shaker) in 15 ml centrifuge tubes as occasional shaking was found to result in incomplete extraction of the Fe phases. For step (a) at 50 °C, samples were shaken either on a heated shaking table, or on a conventional shaking table placed in an oven. Between extraction steps, samples were centrifuged prior to decanting and analysis.

(a) Sodium acetate: Samples were subjected to 10 ml of a 1 mol l⁻¹ sodium acetate solution buffered with acetic acid (pH = 4.5) for 48 h, at a constant temperature of 50 °C. Carbonate-poor samples were degassed 1 h after the addition of sodium acetate and again after 6 h. Carbonate-rich samples were degassed 1, 2, 6 and 24 h after sodium acetate addition. This first step (Fe_{sub}) primarily targets iron associated with carbonate phases (Table 1).

(b) Sodium dithionite: The sample was then treated with 10 ml of a sodium dithionite solution (50 g l⁻¹ sodium dithionite, 58.82 g l⁻¹ tri-sodium citrate, 20 ml l⁻¹ acetic acid) for 2 h. The sodium dithionite solution was always prepared immediately prior to use, to avoid oxidation of the solution and hence a lower extraction potential. This step (Fe_{ox}) primarily targets ferric oxide minerals (Table 1).

(c) Ammonium oxalate: The final step of the sequential iron extraction targets mixed ferric/ferrous oxides, such as magnetite (Fe_{mag}). This was achieved with 10 ml of a 0.2 mol l⁻¹ ammonium oxalate/0.17 mol l⁻¹ oxalic acid solution, with a treatment time of 6 h.

(d) Hot chromous chloride distillation: This method dissolves sulfide minerals, primarily comprising acid-volatile sulfides (AVS) and pyrite (Fe_{pyr}, Canfield et al. 1986). The amount of sample required for this method depends on the amount of sulfide present. For the WHIT, reference sample ~ 0.2 g was used for the extraction, for KL133 and KL134 ~ 2.5 g was used, and for BHW ~ 1.75 g was used. These sample masses ensure a sufficient amount of Ag₂S precipitation for later quantification. The samples were initially treated with near-boiling 50% v/v HCl (6 ml) under a nitrogen atmosphere to test for the presence of AVS (Canfield et al. 1986). However, no AVS was detected in any of the samples, and thus, after addition of the HCl, 16 ml of chromous chloride was added. This solution was boiled for 1 h, also under a nitrogen atmosphere, and the released hydrogen sulfide was trapped (as Ag₂S) in a 1 mol l⁻¹ AgNO₃ solution (with additional AgNO₃ added where appropriate to avoid saturation of the trap with sulfide). The
Ag₂S precipitates were then filtered, dried and weighed, and the concentration of pyrite Fe was determined stoichiometrically. In one of the laboratories, Zn acetate was used to trap the released H₂S instead of AgNO₃, and sulfide analysis was performed by spectrophotometry using diamine reagent (Cline 1969).

(e) Concentrated HCl: To determine Feᵦ, approximately 100 mg of sample was weighed into a glass test tube. Concentrated HCl (5 ml) was added, and the sample was immediately gently heated for 60 s to bring to the boil. The sample was then boiled more aggressively for a further 60 s (Berner 1970, Raiswell et al. 1994). Samples were then immediately quenched with high-purity water and transferred quantitatively to 100 ml volumetric flasks and made up to volume. The difference between Feᵦ and the sum of Feₚₕₛ, Feₕₓ and Feₚₖₕₖₖ gives Feᵦ. Note, however, that this extraction may also be performed sequentially after steps la–lc (Table 1), which gives a direct measurement of Feᵦ without the need to subtract Feₚₕₛ, Feₕₓ and Feₚₖₖₖ (Poulton and Canfield 2005).

Analysis of Fe solutions

Three commonly used techniques were compared for the analysis of the Fe solutions from steps a–c and e. Atomic absorption spectrometry (AAS) was the primary technique used by three of the laboratories. In this case, for the sequential extraction steps a–c, the supernatant was subjected to a twenty times dilution with high-purity water prior to analysis relative to matrix-matched reference materials. The same procedure was used for boiling HCl extractions, but with a five times dilution of the initial 100 ml solution. The fourth laboratory determined dissolved Fe via inductively coupled plasma-optical emission spectroscopy (ICP-OES). Here, solutions were diluted forty-fold with 1% v/v HNO₃. The dilution acid contained 10 µg g⁻¹ yttrium as an internal standard element, which was monitored to compensate for matrix-related signal fluctuation.

Finally, we tested the utility of the ferrozine method (Stookey 1970). Here, we used the approach of Sperling et al. (2013), whereby 100 µl of extract was added to 4 ml of solution (prepared immediately prior to analysis) containing 12 g l⁻¹ HEPES buffer, 0.2 g l⁻¹ ferrozine reagent and 10 g l⁻¹ hydroxylamine hydrochloride (which reduces Fe(III) to Fe(II)).

![Figure 1. Recovery of Fe for KL133 (solid line ± 1s) in the Feₚₕₛ and Feₕₓ fractions, as determined by the spectrophotometric ferrozine method, relative to the mean combined value determined via AAS and ICP-OES (dashed line ± 1s).](image)

| Table 2. Mean (± 1s) major element mass fractions of reference materials (expressed as % m/m), measured by two independent XRF laboratories |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| RM              | Si   | Ti   | Al   | Fe   | Mn   | Mg   | Ca   | Na   | K   | P   | Mean 1s | Mean 1s | Mean 1s | Mean 1s | Mean 1s | Mean 1s | Mean 1s | Mean 1s | Mean 1s | Mean 1s | Mean 1s |
| WHIT            | 22.74 | 0.24 | 0.553 | 0.009 | 11.9 | 0.16 | 4.54 | 0.02 | 0.019 | 0.001 | 1.10 | 0.02 | 1.03 | 0.02 | 0.378 | 0.009 | 2.65 | 0.02 | 0.070 | 0.003 |
| KL133           | 31.06 | 0.19 | 0.371 | 0.007 | 7.81 | 0.03 | 3.19 | 0.03 | 0.065 | 0.001 | 1.44 | 0.04 | 0.970 | 0.016 | 1.18 | 0.03 | 3.13 | 0.04 | 0.883 | 0.002 |
| KL134           | 28.49 | 0.83 | 0.385 | 0.007 | 8.87 | 0.25 | 5.03 | 0.07 | 0.087 | 0.002 | 1.00 | 0.04 | 0.652 | 0.006 | 1.60 | 0.08 | 2.66 | 0.04 | 0.092 | 0.003 |
| BHM             | 32.75 | 0.33 | 0.236 | 0.009 | 7.48 | 0.06 | 1.62 | 0.02 | 0.017 | 0.001 | 1.93 | 0.03 | 1.000 | 0.010 | 0.656 | 0.004 | 6.88 | 0.09 | 0.018 | 0.003 |
to Fe(II), to allow measurement by ferrozine). At the same time, a range of matrix-matched reference materials was prepared. For these reference materials, we compared the results of using both an Fe(III) stock solution (1000 µg ml⁻¹ Fe(NO₃)₃ in 0.5 mol l⁻¹ HNO₃) and an Fe(II) stock solution (Mohr’s salt, (NH₄)₂Fe(SO₄)₂.6H₂O). For the Fecarb and Feox extractions, similar results were obtained using both stock Fe solutions. However, for the Femag extraction, a precipitate formed when using the Fe(II) stock solution, and thus, all of our results are reported relative to reference materials prepared with the Fe(III) stock solution. Sperling et al. (2013) left samples overnight to allow colour development, followed by analysis by spectrophotometer. To further test this technique, we performed regular repeat measurements of the solutions and reference materials (on a Genesys 6 spectrophotometer, Thermo Scientific, Massachusetts, USA) for up to 16 days after preparation.

Results and discussion

Table 3. Mean mass fractions (± 1 s) of Fe in each sequential Fe extraction (% m/m), as determined by AAS, ICP-OES and spectrophotometry

| ID     | Fe₉₉,carb | 1s | % RSD | Fe₉₉,ox | 1s | % RSD | Fe₉₉,mag | 1s | % RSD |
|--------|-----------|----|-------|---------|----|-------|----------|----|-------|
| WHIT   |           |    |       |         |    |       |          |    |       |
| AAS    | 0.581     | 0.033 | 5.7 | 0.063 | 0.010 | 15.9 | 0.106 | 0.012 | 11.3 |
| ICP-OES | 0.593 | 0.009 | 1.5 | 0.058 | 0.002 | 3.5 | 0.081 | 0.002 | 2.5 |
| Spec.  | 0.616 | 0.008 | 1.3 | 0.084 | 0.001 | 1.6 | 0.106 | 0.003 | 2.8 |
| KL133  |           |    |       |         |    |       |          |    |       |
| AAS    | 0.139     | 0.006 | 4.3 | 0.046 | 0.004 | 8.7 | 0.169 | 0.011 | 6.5 |
| ICP-OES | 0.139 | 0.004 | 2.9 | 0.046 | 0.002 | 4.3 | 0.157 | 0.010 | 6.4 |
| Spec.  | 0.145 | 0.002 | 1.4 | 0.049 | 0.001 | 2.0 | 0.149 | 0.003 | 2.0 |
| KL134  |           |    |       |         |    |       |          |    |       |
| AAS    | 0.711     | 0.041 | 5.8 | 0.095 | 0.010 | 10.5 | 0.584 | 0.028 | 4.8 |
| ICP-OES | 0.680 | 0.020 | 2.9 | 0.088 | 0.003 | 3.4 | 0.532 | 0.016 | 3.0 |
| Spec.  | 0.781 | 0.005 | 0.6 | 0.110 | 0.001 | 0.9 | 0.538 | 0.012 | 2.2 |
| BHW    |           |    |       |         |    |       |          |    |       |
| AAS    | 0.044     | 0.006 | 13.6 | 0.016 | 0.003 | 18.8 | 0.024 | 0.002 | 8.3 |
| ICP-OES | 0.044 | 0.002 | 4.5 | 0.018 | 0.003 | 16.7 | 0.023 | 0.005 | 21.7 |
| Spec.  | 0.045 | 0.001 | 2.2 | 0.015 | 0.001 | 6.7 | 0.020 | 0.001 | 5.0 |

Spectrophotometry data are reported after 24 h for Fecarb and Feox, and after 168 h for Femag (see text for further details).

Comparison of iron determinations in the extraction solutions by different techniques

A comparison of iron determinations by AAS, ICP-OES and spectrophotometry is presented in Table 3. The RSD for each measurement was generally within ~ 6%, with the exception of fractions with very low Fe contents where, as expected, RSDs are commonly higher. Nevertheless, despite these higher RSDs, the magnitude of the measured standard deviation is relatively small for the low Fe fractions (Table 3), and this degree of variability has little impact in terms of quantifying FeHR/FeT and Fepy/FeHR ratios (see below). The RSD for AAS analyses is often higher than for the other measurement techniques, which likely reflects the fact that extractions for the solutions measured by AAS were performed by multiple users across three different laboratories, whereas extractions measured by ICP-OES and spectrophotometry were performed by one user in one laboratory.

In general, there is good agreement (within error) between the measurements by AAS and ICP-OES for all
extracted phases (Table 3), highlighting that both techniques are suitable for measuring the sequential Fe solutions, despite the potential for strong matrix effects. In addition, we found that the ferrozine technique generally produced comparable results for Fe\textsubscript{carb} and Fe\textsubscript{ox} (Table 3), and this was a consistent feature across the 11 days over which these analyses were performed (Figure 1). However, while there is reasonable agreement between both the Fe\textsubscript{mag} AAS/ICP-OES analyses and the spectrophotometric analyses after around 7 days of ferrozine reaction (Table 3), after 24 h of reaction (the current standard technique is to leave solutions overnight prior to analysis) only ~ 60–85% of the Fe\textsubscript{mag} pool was measured by spectrophotometer (Figure 2). This suggests that the extracted Fe may be strongly complexed by the reagents in the oxalate extraction, such that considerable time is required for the reaction with ferrozine to proceed to completion. Furthermore, the mean spectrophotometric Fe\textsubscript{mag} results for KL133, KL134 and BHW were always lower than the mean value determined by AAS, with a distinct decrease to even lower values after ~ 11 days. We thus conclude that the ferrozine spectrophotometric technique is not suitable for the measurement of Fe\textsubscript{mag}.

**Effect of test portion mass on sequential extraction efficiency**

During the course of our analyses, we found that the initial test portion sample mass to extractant ratio may affect the quantity of Fe dissolved. To test this, we performed replicate extractions \((n = 4–8)\) for all four reference materials using three initial masses: 50, 70 and 100 mg, with Fe determined by ICP-OES. We found that lower concentrations were consistently obtained for Fe\textsubscript{carb} as sample mass increased (Table 4). By contrast, the subsequent Fe\textsubscript{ox} and Fe\textsubscript{mag} extractions showed no consistent trends as sample mass increased, and thus, the total amount of iron extracted during the three sequential phases decreased at higher sample masses (Table 1). We observed no consistent trend in the RSD of analyses over the range of sample masses used in our tests, even though in general, the relative
standard deviation would be expected to increase at lower sample masses. Based on these considerations, we propose an optimal sample mass for the sequential extractions of 60 mg for 10 ml of extractant.

**Development of iron speciation reference materials**

We utilise the replicate extractions (as measured by AAS and ICP-OES) of the six users from four independent laboratories to determine the Fe speciation characteristics of the four reference materials (Table 5). Concentrations of Fe$_{ox}$ are relatively low for all four samples, as might be expected for sediments that have experienced anaerobic conditions during early diagenericity (whereby dissimilatory iron reduction and reaction with dissolved sulfide both result in the reductive dissolution of Fe-(oxyhydr)oxides), but the remaining Fe pools show considerable variability. In addition, mean poorly reactive sheet silicate Fe mass fractions (calculated as Fe$_{prs}$ = Fe$_{R}$ - (Fe$_{carb}$ + Fe$_{ox}$ + Fe$_{mag}$)) are 0.219\% m/m for WHIT, 1.399\% m/m for KL133, 2.283\% m/m for KL134, and 0.462\% m/m for BHW. This gives Fe$_{prs}$/Fe$_{T}$ ratios of 0.05 for WHIT, 0.44 for KL133, 0.45 for KL134, and 0.29 for BHW. The Fe$_{prs}$/Fe$_{T}$ ratio for WHIT is low compared with the mean ratio for Phanerozoic shales (0.39 ± 0.11; Raiswell et al., 2008), but the remaining samples fall close to this mean.

In Figure 3, we plot Fe$_{HR}$/Fe$_{T}$ and Fe$_{py}$/Fe$_{HR}$ ratios for each reference material (see Table 5), and we show the standard deviations that are obtained as a result of propagating the precision of measurements for each Fe pool through to the calculation of Fe speciation ratios. This demonstrates that the determination of Fe speciation ratios is highly reproducible, with the largest degree of variability occurring for the BHW Fe$_{py}$/Fe$_{HR}$ ratio, which arises due to

| Table 4. The effect of test portion mass on extraction efficiency. The sum of the three sequential extraction phases is also shown (Fe$_{carb}$ + Fe$_{ox}$ + Fe$_{mag}$) |
|-----------------|--------------|--------------|--------------|--------------|-----------------|--------------|--------------|--------------|--------------|
| ID | Fe$_{carb}$ (% m/m) | Fe$_{ox}$ (% m/m) | Fe$_{mag}$ (% m/m) | Sum (% m/m) |
| Mean | 1s | % RSD | Mean | 1s | % RSD | Mean | 1s | % RSD | Mean | 1s | % RSD |
| WHIT 50 mg | 0.618 | 0.003 | 0.5 | 0.056 | 0.001 | 1.8 | 0.073 | 0.001 | 1.4 | 0.748 | 0.003 | 0.4 |
| 70 mg | 0.593 | 0.008 | 1.3 | 0.058 | 0.002 | 3.4 | 0.078 | 0.003 | 3.9 | 0.729 | 0.010 | 1.4 |
| 100 mg | 0.549 | 0.008 | 1.4 | 0.056 | 0.002 | 3.6 | 0.078 | 0.003 | 3.9 | 0.683 | 0.011 | 1.6 |
| KL133 50 mg | 0.151 | 0.005 | 3.3 | 0.048 | 0.004 | 8.3 | 0.149 | 0.001 | 0.7 | 0.348 | 0.009 | 2.6 |
| 70 mg | 0.137 | 0.006 | 4.4 | 0.046 | 0.002 | 4.3 | 0.155 | 0.009 | 5.8 | 0.338 | 0.012 | 3.6 |
| 100 mg | 0.133 | 0.005 | 3.8 | 0.045 | 0.002 | 4.4 | 0.145 | 0.009 | 6.2 | 0.323 | 0.014 | 4.3 |
| KL134 50 mg | 0.734 | 0.026 | 3.5 | 0.091 | 0.004 | 4.4 | 0.507 | 0.012 | 2.4 | 1.332 | 0.026 | 2.0 |
| 70 mg | 0.689 | 0.019 | 2.8 | 0.087 | 0.003 | 3.4 | 0.512 | 0.023 | 4.5 | 1.289 | 0.016 | 1.2 |
| 100 mg | 0.651 | 0.019 | 1.4 | 0.088 | 0.003 | 3.4 | 0.513 | 0.012 | 2.3 | 1.251 | 0.021 | 1.7 |
| BHW 50 mg | 0.048 | 0.003 | 6.3 | 0.018 | 0.002 | 11.1 | 0.02 | 0.002 | 10.0 | 0.087 | 0.007 | 8.0 |
| 70 mg | 0.044 | 0.002 | 4.5 | 0.019 | 0.003 | 15.8 | 0.021 | 0.004 | 19.0 | 0.084 | 0.007 | 8.3 |
| 100 mg | 0.038 | 0.001 | 2.6 | 0.015 | 0.001 | 6.7 | 0.018 | 0.005 | 27.8 | 0.071 | 0.005 | 7.0 |

| Table 5. Mean mass fractions (% m/m ± 1s) of Fe in each fraction, and Fe$_{HR}$/Fe$_{T}$ and Fe$_{py}$/Fe$_{HR}$ ratios (± 1s) for each reference material |
|-----------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|--------------|
| ID | Fe$_{carb}$ | Fe$_{ox}$ | Fe$_{mag}$ | Fe$_{py}$ | Fe$_{HR}$ | Fe$_{HR}$/Fe$_{T}$ | Fe$_{py}$/Fe$_{HR}$ |
| Mean | 1s | Mean | 1s | Mean | 1s | Mean | 1s | Mean | 1s |
| WHIT | 0.583 | 0.030 | 0.062 | 0.009 | 0.103 | 0.014 | 1.970 | 0.087 | 0.967 | 0.097 | 0.60 | 0.02 | 0.73 | 0.04 |
| KL133 | 0.139 | 0.005 | 0.046 | 0.004 | 0.167 | 0.011 | 0.011 | 0.002 | 1.751 | 0.121 | 0.11 | 0.00 | 0.03 | 0.01 |
| KL134 | 0.705 | 0.039 | 0.094 | 0.009 | 0.578 | 0.032 | 0.011 | 0.002 | 3.660 | 0.124 | 0.28 | 0.01 | 0.01 | 0.00 |
| BHW | 0.044 | 0.005 | 0.017 | 0.003 | 0.023 | 0.003 | 0.068 | 0.008 | 0.546 | 0.077 | 0.09 | 0.01 | 0.45 | 0.06 |
the low concentration of each Fe fraction in this sample (Table 5). The reference materials also document a range of redox conditions, including oxic (KL133, BHW), equivocal (KL134) and anoxic (WHIT). $\text{Fepy/Fe}_{\text{HR}}$ ratios are only relevant as a water column redox indicator for samples that show clear evidence of deposition under anoxic water column conditions (Poulton and Canfield 2011), and the anoxic WHIT sample plots close to the threshold for identifying euxinia. The remaining reference materials have variable $\text{Fepy/Fe}_{\text{HR}}$ ratios, which likely reflect different levels of sulfide production during diagenesis. Taken together, the variable speciation characteristics, combined with the wide range of mass fractions evident across the Fe fractions, suggest that these four samples are ideal as international reference materials.

Conclusions

We have developed four reference materials that may be used by researchers conducting Fe speciation analyses of palaeodepositional redox conditions. In the process of creating these reference materials, we have refined ‘best practice’ techniques for Fe speciation analyses, including detailed evaluation of the commonly employed techniques for determining Fe mass fractions in extracted solutions. The amount of iron dissolved in each extraction step is sensitive to both sample agitation and test portion mass, and we recommend that extractions are performed with a test portion mass of $60 \pm 10$ mg for 10 ml of extractant. In addition, we recommend that the spectrophotometric determination of Fe$_{\text{mag}}$ by ferrozine is discontinued.

The reference materials comprise a range of Fe fraction mass fractions and document a range of depositional redox conditions. In addition, Fe mass fractions and speciation ratios are generally highly reproducible, with a greater degree of uncertainty being limited to those fractions containing very low mass fractions of Fe. These characteristics confirm the wide-ranging suitability of the samples as international reference materials for iron speciation analyses. The samples are stored under controlled conditions, where oxygen, light and moisture are eliminated, making them suitable as long-term reference materials for the community.

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Data availability statement

Data supporting this study are available on request from the authors.

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