**Pax6 Inactivation in the Adult Pancreas Reveals Ghrelin as Endocrine Cell Maturation Marker**

Zeeshan Ahmad1 *, Maria Rafeeq1, Patrick Collombat2,3,4,5, Ahmed Mansouri1,4,6 *

1 Max Planck Institute for Biophysical Chemistry, Department of Molecular Developmental Biology, RG Molecular Cell Differentiation, Goettingen, Germany, 2 Université de Nice Sophia Antipolis, Nice, France, 3 Inserm U1091, IBV, Diabetes Genetics Team, Nice, France, 4 JDRF, New York, NY, United States of America, 5 Genome and Stem Cell Center, GENKOK, Erciyes University, Kayseri, Turkey, 6 University of Goettingen, Department of Clinical Neurophysiology, Goettingen, Germany

* amanou@gwdg.de (AM); zahmad@gwdg.de (ZA)

**Abstract**

The transcription factor Pax6 is an important regulator of development and cell differentiation in various organs. Thus, Pax6 was shown to promote neural development in the cerebral cortex and spinal cord, and to control pancreatic endocrine cell genesis. However, the role of Pax6 in distinct endocrine cells of the adult pancreas has not been addressed. We report the conditional inactivation of Pax6 in insulin and glucagon producing cells of the adult mouse pancreas. In the absence of Pax6, beta- and alpha-cells lose their molecular maturation characteristics. Our findings provide strong evidence that Pax6 is responsible for the maturation of beta-, and alpha-cells, but not of delta-, and PP-cells. Moreover, lineage-tracing experiments demonstrate that Pax6-deficient beta- and alpha-cells are shunted towards ghrelin marked cells, sustaining the idea that ghrelin may represent a marker for endocrine cell maturation.

**Introduction**

The pancreas develops from the endoderm at the foregut/midgut junction to give rise to two main cell compartments, comprising exocrine and endocrine cells. The major compartment of the pancreas is the exocrine tissue (acinar and duct cells) producing digestive enzymes that reach the duodenum through an intricate ductal system. The endocrine counterpart regroups hormone-producing cells into functional units called islets of Langerhans, consisting of beta-, alpha-, delta-, PP-, and epsilon-cells secreting insulin, glucagon, somatostatin, pancreatic polypeptide, and ghrelin, respectively, to control glucose homeostasis. The development and maturation of the endocrine pancreas is under the control of a variety of transcription factors (for review see [1, 2]). Among these, members of the Pax gene family, Pax4 and Pax6, were shown to play a crucial role in endocrine pancreas genesis [3–8]. Accordingly, Pax4 inactivation resulted in the loss of beta- and delta-cells, concomitant with a proportional increase of glucagon-labeled cells [3]. Moreover, the forced expression of Pax4 in glucagon-producing alpha-cells of
transgenic mice provoked their conversion into beta-like cells that counter chemically induced diabetes [9–12]. In contrast, the loss of Pax6 gene activity was accompanied with a highly reduced number of all endocrine cells, and where glucagon-expressing cells were predominantly affected. Mice with conditional inactivation of Pax6 in Pdx1 or Pax6 expression domains died few days postpartum and suffered from diabetes [5]. Of interest is the maintained expression of some pancreatic transcription factors such as Nkx2.2, Isl1, and Pdx1 in these Pax6 mutant pancreata. This suggested that Pax6 is required to control the expression of genes involved in pancreatic endocrine function such as insulin, Glut2, and glucagon [5]. This is consistent with a recent report using global conditional inactivation of Pax6 in 6 months old mice [7]. Several in vitro studies clearly confirmed that Pax6 is an important regulator of endocrine cell development and function. Accordingly, Pax6 was found to control the transcription of factors necessary for alpha-cell differentiation, such as proglucagon and the processing enzyme PC2 [13–15]. Along the same line of evidence Pax6 directly activates the transcription of MafB, cMaf, and Beta2/NeuroD, and Pax6 binding sites are found on their respective promoter [13–15]. However, Pax6 is not necessary for alpha-cell specification [16]. Interestingly, Pax6 also promotes proper beta-cell function, since it acts upstream of MafB which triggers the expression of the beta-cell maturation factors Pdx1 and MafA [13]. Similarly, Pax6 was shown to control the transcription of several factors that are crucial for beta-cell function, such as Glut2, PC1/3, Nkx6.1, and GLP1R [17]. Moreover, the knockdown of Pax6 function in primary beta-cells was accompanied by alterations in insulin synthesis and secretion, as well as decreased GLP-1 activity [17]. In addition, in zebrafish and in mice, pancreas specific enhancers that control Pax6 gene activity were identified and were found to be dependent on Pbx and Meis homeoproteins [18, 19].

Notwithstanding, Pax6 function in distinct adult endocrine cells is still not clear. Here we report the conditional inactivation of Pax6 in insulin- and glucagon-producing beta- and alpha-cells of adult mice, respectively. Using lineage tracing and endocrine hormone examination, we could show that beta- and alpha-cells lacking Pax6 function are shunted towards a ghrelin-labeled cell fate. Interestingly, beta- or alpha-cells with depleted Pax6 gene activity fail to activate the expression of the maturation factors MafA or MafB, respectively, but maintained their respective pancreatic endocrine cell subtype destiny. While Pax6-/ beta-cells continue to express the transcription factor Pdx1, Pax6-deficient alpha-cells sustain the expression of the alpha-cell factor Arx. We conclude that Pax6 is necessary for the maintenance of molecular maturation characteristics of pancreatic endocrine cells.

Materials and Methods

Animals and treatments

Pax6fl/fl;RIP-CreER and Pax6fl/fl;Glu-Cre mice were generated by crossing Pax6fl/fl mice [20] with RIP-CreERT [21] and Glucagon-Cre mice [22], respectively. Pax6fl/fl;TetO-Cre;Glu-rtTA mice were generated by crossing Pax6fl/fl mice with TetO-Cre [23] and Glucagon-rtTA mice [11]. Pax6fl/fl;RIP-CreER, Pax6fl/fl;Glu-Cre, and Pax6fl/fl;TetO-Cre;Glu-rtTA mice were further crossed with Rosa26-YFP reporter mice [24] to incorporate the YFP reporter transgene for lineage tracing purpose. Lastly, Pax6fl/fl;CG-CreER mice were generated by crossing Pax6fl/fl mice with CAG-CreERT mice [25]. Genotyping was performed by means of PCR using genomic DNA isolated from tail-tip or ear biopsies. Primer sequences and PCR conditions are available on request.

Tamoxifen (Sigma-Aldrich) prepared in corn oil (at 20 mg/ml), was administered by intraperitoneal injection. In case of RIP-CreERT mice, 3–4 week old mice received 1 mg of tamoxifen per day for 3 days every other day, and 1.5–2.5 month old mice received 2 mg of tamoxifen per day for 4 days every other day. Age at tamoxifen-induction and pancreas analyses time points used for beta-cell-specific Pax6 ablation mice are shown in Fig 1 along with the respective figure.
numbers related to that specific setting. In case of CAG-CreERT mice, 2 month old mice were injected with 2 mg of tamoxifen per day for 5 consecutive days. Doxycycline (Sigma-Aldrich) was given to the mice in drinking water (at 2 g/l with 1% sucrose) for a period of 6 weeks. Doxycycline containing water was protected from light and replaced every week.

All animal experiments were conducted according to the German animal welfare law (LAVES Niedersachsen: Nds. Landesamt für Verbraucherschutz und Lebensmittelsicherheit (LAVES), Dezernat 33, 26029 Oldenburg). Hence, the experiments were performed according to the permission from Laves under: Vorhaben 33.9-42502-04-11/0402 (Kurztitel „Transgene01“) und 33.9-42502-04-11/0622 (Kurztitel „Transgene02“).

**Blood glucose measurements**

Blood from the tail tip was used to measure the non-fasting blood glucose levels with the help of Accu-Chek Aviva (Roche) blood glucose meter.

**Immunohistochemistry**

Mice were killed by CO2 inhalation and/or cervical dislocation. Pancreata were removed, washed in ice-cold PBS, and fixed in 4% paraformaldehyde (in PBS) for 1 hour at 4°C. Fixed pancreata were then washed in PBS (4x20 minutes) and dehydrated in 25% sucrose (in PBS) for overnight at 4°C. Finally, they were embedded in the Jung Tissue Freezing Medium™ (Leica Microsystems). For immunofluorescence staining, 8 μm thick cryosections were used. Blocking was performed in 10% fetal calf serum (in PBS containing 0.1% Triton X-100) for 1 hour at room temperature. For some nuclear antigens, the signal was improved by an additional permeabilization with 0.2% Triton X-100 (in PBS) for 30 minutes at room temperature before the blocking. After blocking, the sections were incubated with primary antibodies (in blocking solution) for overnight at 4°C. This was followed by, washing in PBS, and incubation with appropriate secondary antibodies (in blocking solution) for 1 hour at room temperature. At the end, sections were washed again in PBS and mounted with Vectashield® mounting medium containing DAPI as a nuclear counterstain.

The following primary antibodies were used in this study: guinea pig anti-insulin 1/1000, rabbit anti-somatostatin 1/600, and rat anti-Ki67 1/100 (Dako), mouse anti-insulin 1/500 (Sigma-Aldrich), mouse anti-glucagon 1/1000, rabbit anti-glucagon 1/50, rabbit anti-7B2 1/5000, and chicken anti-GFP 1/1000 (Abcam), mouse anti-insulin 1/200, rabbit anti-pancreatic polypeptide (PP) 1/1000, rabbit anti-Rfx6 1/2000, rabbit anti-Arx 1/200, rabbit anti-Glut2 1/1000, rabbit anti-PC1/3 1/500, and rabbit anti-PC2 1/200 (Millipore), rabbit anti-MafA 1/500 and rabbit anti-MafB 1/1000 (Bethyl Labs), rabbit anti-C-peptide 1/100 (Cell Signaling), rabbit anti-IAPP 1/500 (Phoenix Pharma), goat anti-ghrelin 1/50 (Santa Cruz), rabbit anti-Pax6 1/300 (Covance), mouse anti-Nkx6.1 1/50 (Developmental Studies Hybridoma Bank), rabbit anti-Pdx1 1/2000 (kindly provided by C. Wright, Vanderbilt University, Nashville, TN), mouse anti-ghrelin 1/1000 (kindly provided by C. Tomasetto, Université Louis Pasteur, Strasbourg), rabbit anti-GLP-1 receptor 1/4000 (kindly provided by J.F. Habener, Harvard Medical School, MA), rabbit anti-7B2 1/200 and rabbit anti-ProSAAS 1/50 (kindly provided by I. Lindberg, University of Maryland-Baltimore, Maryland [26, 27]).

All of the secondary antibodies were obtained from Invitrogen (diluted at 1/1000) with the exception of sheep Cy5 anti-mouse (diluted at 1/300) that was obtained from Jackson Immunoresearch Laboratories. Images were acquired using either an Olympus BX60 fluorescent microscope or a Leica TCS SP5 confocal laser-scanning microscope. In case of triple immunofluorescence stainings, Cy5 channel was pseudo-colored in magenta, white, or green as required to show a better colocalization with other channels.
Fig 1. Different experimental time lines used for the beta-cell-specific Pax6 ablation. Age at tamoxifen induction and analysis time point post-induction is indicated. After each timeline, the respective figure number is also mentioned.

doi:10.1371/journal.pone.0144597.g001
Quantification of islet cell numbers and statistical analysis

For each quantification category, at least 3 mice per genotype per condition were used. The entire pancreata were serially cut and stained cells on every 20th section were manually counted to obtain an average number per islet section. The values are depicted as mean±Standard error of mean (SEM). Statistical analysis was performed using student’s two-tailed t-test and p<0.05 was considered as significant.

Results

Pax6 ablation in insulin-producing beta-cells of the adult pancreas

Pancreatic endocrine cell function was shown to depend on Pax6 gene activity. In the absence of Pax6 the number of the cells expressing insulin and glucagon was altered [4, 7, 8]. However, the role of Pax6 in specific endocrine cell types in the adult pancreas is still obscure. We therefore used Pax6^{−/−} mice and RIP-CreERT transgenic mice to generate animals where Pax6 is specifically inactivated in beta-cells upon tamoxifen induction (RIP-CreERT;Pax6^{fl/fl}). Furthermore, we used lineage tracing to follow the fate of Pax6-deficient beta-cells over an extended time period. Following tamoxifen administration, the successful depletion of Pax6 in beta-cells was confirmed by immunohistochemical analyses showing 95% KO efficiency (Fig 2).

Previous studies revealed that Pax6 loss-of-function in mouse embryonic, as well as adult, pancreas leads to hyperglycemia [5, 7]. Similarly, our adult beta-cell-specific Pax6 knockout mice developed hyperglycemia with blood glucose levels reaching as high as 450 mg/dl (see later). This augmentation in blood glucose level was associated with a decrease in insulin positive cell numbers in the pancreas of mutant mice (see below).

Depletion of Pax6 gene activity provokes the loss of mature beta-cell molecular characteristics and the activation of ghrelin expression

Following two weeks of tamoxifen induction, insulin-producing beta-cells lost the expression of MafA and Glut2, two important markers of mature beta-cells (Fig 3). However, while the expression of Pdx1 remained unaffected, that of Nkx6.1 first appeared normal few days past-tamoxifen treatment, and gradually decreased at later time points (Fig 3d, 3h, 3i and 3j). Supporting this result, a qRT-PCR performed at 3 months post-tamoxifen induction also showed decrease in the expression of MafA, Glut2, and Nkx6.1. However, the expression of Pdx1 as well as that of Nkx2.2 was not significantly changed (S1 Fig). In addition, when followed over a longer time-period, it was possible to notice a decrease in the number of insulin positive cells in the KO pancreata with a concomitant augmentation in the content of ghrelin labeled cells (Figs 4 and 5). Of interest was the detection of some ghrelin/insulin co-positive cells (Fig 6). A decrease in the expression of insulin and augmentation in the expression of ghrelin was also evident by qRT-PCR (S1 Fig). Collectively, these observations are consistent with the idea that, following Pax6 inactivation, beta-cells lose insulin expression and start to express ghrelin instead. This was supported by the fact that these ghrelin expressing cells were positive for beta-cell-specific transcription factors, such as Pdx1 and Nkx6.1 (Fig 7). Additionally, they also expressed PC1/3 and iAPP, two other beta-cell-related factors (Fig 6). It is important to notice that iAPP production was never affected in Pax6-deficient beta-cells. As a consequence, the normal iAPP/insulin ratio is altered and might contribute to further deterioration of beta-cell function due to the formation of amyloid deposits [28]. Moreover, in the islets of beta-cell-specific Pax6 knockout mice the numbers of glucagon*, somatostatin*, and PP* cells were also increased (S2 Fig).
Fig 2. Beta-cell-specific ablation of Pax6. Double immunofluorescence staining of pancreatic cryosections from 2 month old mice at 4 days (a-d) or 4 weeks (g-j) after tamoxifen induction. Pax6 expression is lost from a majority of insulin+ cells (c, d). Ablation of Pax6 is specific to YFP labeled cells (i, j) as YFP+ insulin+ cells continue to express Pax6 (arrows i, j). Quantification of Pax6+ insulin+ (e) and YFP+ insulin+ (f) cells in the pancreata of 2 month old mice at 4 days after tamoxifen induction (n = 3). Error bars represent SEM; *p<0.05.

doi:10.1371/journal.pone.0144597.g002
Fig 3. Expression of beta-cell-related factors in the islets of beta-cell-specific Pax6 KO mice. Double immunofluorescence staining of pancreatic cryosections from 2.5 month old mice at 2 weeks after tamoxifen induction (a–h) or 3.5 month old mice at 6 weeks after tamoxifen induction (i, j). Expression of MafA and Glut2 is lost (e, f), expression of Pdx1 is maintained (g), and that of Nkx6.1 is maintained at 2 weeks (h) but sharply reduced at 6 weeks (j) post-tamoxifen induction in the islets of beta-cell-specific Pax6 KO pancreata.

doi:10.1371/journal.pone.0144597.g003
Lineage tracing revealed that $Pax6^{-/-}$ beta-cells are shunted towards ghrelin-labeled cells

Lineage tracing with YFP reporter clearly demonstrated the conversion of beta-cells into ghrelin-expressing cells. Indeed, the analysis of KO pancreata ($RIP$-$CreER;Rosa26$-$YFP;Pax6^{fl/fl}$) at different time points following tamoxifen induction revealed a progressive conversion of reporter-labeled insulin positive cells into YFP-marked ghrelin positive cells (Figs 5 and 8). Additionally, some of the reporter-labeled ghrelin positive cells lose the expression of ghrelin over time, as evidenced by the presence of reporter labeled insulin and ghrelin negative (YFP$^+$ insulin$^-$ ghrelin$^-$) cells. Of interest was the observation that the YFP-labeled cells maintain the expression of Rfx6 and Pdx1 even at 4.5 month after tamoxifen induction. We conclude that in the absence of Pax6 activity adult beta-cells maintain their insulin-producing destiny, but lose the molecular characteristics of mature beta-cells (Fig 9).

We further noticed that the numbers of YFP-labeled cells in beta-cell-specific Pax6-deficient islets were always significantly inferior to those found in control islets. We therefore checked cell survival and cell proliferation in control and mutant islets. While apoptosis of insulin producing cells was not affected (S3 Fig), their proliferation was indeed defective. At 7 weeks of age (4 weeks post-tamoxifen induction), YFP$^+$ Ki67$^+$ cells could not be detected in the beta-cell-specific Pax6-deficient islets. However, such double positive cells were present in the control islets (Fig 10). It appears that Pax6 depletion may provoke decreased cell proliferation.

Pax6-deficiency may affect insulin-processing machinery

Pax6 was shown to directly down regulate Pcsk1n expression and thereby to affect pro-insulin processing [29]. To analyse whether Pax6 inactivation has an impact on insulin processing and release, we examined the expression of c-peptide, PC1/3, PC2, proSAAS, and 7B2. In Pax6-deficient insulin-producing cells the expression of c-peptide was lost before that of insulin...
suggesting a defect in insulin processing. However, the expression of pro-hormone convertases 1/3 and 2 that are required for such process was normal (Fig 11 and S1 Fig). On the other hand, expression of proSAAS and 7B2, the regulatory peptides for PC1/3 and PC2, respectively, was increased (Fig 12). Elevated expression of proSAAS is known to inhibit the activity of PC1/
3 supporting the speculation that insulin processing may indeed be disturbed. In contrast, the significance of increased 7B2 expression is still not clear.

In addition, we checked the expression of Glut2 and GLP1R, both of which play an important role in glucose stimulated insulin secretion (GSIS). Glut2 expression was lost in both the YFP labeled and unlabeled beta-cells (Fig 13), sustaining the notion that Glut2 expression is affected by hyperglycemia [30]. ChIP analysis clearly indicates that Pax6 does not appear to interact with Glut2 promoter (S4 Fig). This suggests that Pax6 may indirectly control the expression of Glut2. In contrast, GLP1R expression was lost in Pax6 KO islets only (Fig 14). This corroborates previous in vitro studies where Pax6 was shown to directly act on the expression of GLP1R [17].

Regeneration of beta-cells was shown to occur in several diabetic mouse models [21, 31, 32]. As the beta-cell-specific Pax6 KO mice developed a sustained hyperglycemia, we were also interested whether some beta-cell regeneration may have occurred. For this purpose, we injected the mice (expressing YFP reporter) with tamoxifen at 3 weeks and 1.5 months of age and analyzed the pancreata at 10 weeks and 4.5 months post-tamoxifen induction, respectively. During this time period, the non-fasting blood glucose level was also monitored over regular intervals. In both cases, blood glucose levels started to rise until they reached a plateau. Afterwards, the blood glucose level slightly decreased in the mice that were injected at 3 weeks of age but not in those injected at 1.5 months of age (Fig 15a and 15b). Furthermore, a significant increase in the number of insulin+ YFP+ cells was observed at both ages indicating newly generated insulin+ cells following the KO induction. Again, this increase was more pronounced in the mice that were injected at 3 weeks of age as compared to the ones injected at 1.5 months of age (Fig 15c and 15d). This result is also in accordance with the levels of blood glucose. In conclusion, we noticed a modest and partial recovery from hyperglycemia in the mice injected at 3

**Fig 6. Co-expression of ghrelin with insulin, iAPP, and PC1/3 in the islets of beta-cell-specific Pax6 KO mice.** Double immunofluorescence staining of pancreatic cryosections from 2.5 month old mice at 2 weeks after tamoxifen induction. Ghrelin+ cells are not detected in the control islets (a-c). In the beta-cell-specific Pax6 KO islets, ghrelin expression is up regulated (d-f) and some of the ghrelin+ cells co-express insulin (d). Additionally, these ghrelin+ cells co-express iAPP (e) and PC1/3 (f).

[doi:10.1371/journal.pone.0144597.g006]
weeks but not in the mice injected at 1.5 months. This is probably due to the more pronounced proliferation of beta-cells at younger age. However, in young as well as in old mice the observed increase in beta-cell number is not sufficient to allow recovery to normoglycemia.

**Pax6**-deficient alpha-cells lose MafB and glucagon expression, while sustaining Arx expression

To generate alpha-cell-specific Pax6 KO mice we used both non-inducible and tetracycline inducible Glucagon-Cre mouse lines [11, 22]. Lineage tracing was again performed in combination with Pax6 ablation to unambiguously identify and follow the destiny of Pax6-deficient glucagon producing alpha-cells. Efficiency of knockout was nearly 68% in case of Glucagon-Cre and nearly 75% when using Tetracycline inducible Glucagon-Cre (Fig 16).

Our immunohistochemical analyses demonstrate that in alpha-cell-specific Pax6 knockout pancreata, the majority of the YFP-labeled alpha-cells lost glucagon expression and become positive for ghrelin (Fig 17). Of interest was the observation that Pax6-deficient reporter-labeled cells were MafB+/Arx+ (Fig 18 and S5–S7 Figs). This indicates that similar to what has
been found in beta-cell-specific Pax6 knockout pancreata, the loss of Pax6 gene activity in glucagon-producing cells provoked the erasure of alpha-cell mature differentiation characteristics, and induced the activation of increased levels of ghrelin expression. However, alpha-cells lacking Pax6 continue to express Arx and may represent alpha-cells that have lost their molecular maturation characteristics.

As Pax6 ablation affects insulin processing in beta-cells, we were interested to see whether this also occurs for glucagon in alpha-cells too. In YFP labeled Pax6-deficient alpha-cells, the expression of PC2 was preserved but that of 7B2 was highly up regulated (Fig 19).

Alpha-cell hyperplasia has been shown to occur in animals where glucagon signaling was impaired, providing strong evidence for glucagon neogenesis [9, 11, 33–35]. Given the observation that our Pax6 glucagon-specific knockout was accompanied with the loss of glucagon expression, we were interested to examine whether the induced alpha-cell shortage may provoke alpha-cell neogenesis. Using the non-inducible Glucagon-Cre, in islets of Glu-Cre;

Fig 8. Ghrelin cells originate from beta-cells in the beta-cell-specific Pax6 KO islets. Double immunofluorescence staining of pancreatic cryosections from 4 month old mice at 6 weeks after tamoxifen induction (a-f) and 6 month old mice at 4.5 months after tamoxifen induction (g-i). Ghrelin expression is not detected in the control islets (a-c). In beta-cell-specific Pax6 KO islets, ghrelin expression is up regulated and that of insulin down regulated in the YFP labeled Pax6-deficient cells (d-f). Most of the ghrelin+ cells have lost the expression of insulin but some are still insulin+ (arrows d-f). Some YFP+ cells in the KO islets are negative for both insulin and ghrelin (arrowhead d-f). At 4.5 months after tamoxifen induction many of the YFP+ cells are negative for both insulin and ghrelin (arrowheads g-i).
R26-YFP;Pax6fl/fl mice at 1 or 3 month of age, the number of total glucagon+ cells was not significantly different from that observed in islets of control animals. However, a clear reduction of glucagon-labeled cells was detected at 8 months of age (Fig 20a). A similar phenotype was noticed when using tetracycline-inducible Glucagon-Cre, in 4 months old Glu-rtTA;TetO-Cre; R26-YFP;Pax60/0 mice, following 6 weeks of Doxycycline treatment (Fig 20a). This may indicate that in younger animals where glucagon cells have lost Pax6, some alpha-cell regeneration

Fig 9. Expression of Rfx6 and Pdx1 in the islets of beta-cell-specific Pax6 KO mice. Immunofluorescence staining of pancreatic cryosections from 6 month old mice at 4.5 month after tamoxifen induction. Expression of Rfx6 (e, f) and Pdx1 (g, h) is maintained in the YFP+ Pax6-deficient cells of the beta-cell-specific Pax6 KO islets.

doi:10.1371/journal.pone.0144597.g009
**Fig 10. Pax6-deficient beta-cells do not proliferate.** Immunofluorescence staining of pancreatic cryosections from 7 week old mice at 4 weeks after tamoxifen induction. Ki67+ YFP+ cells are detected in the control islets (arrows a, b) but not in the beta-cell-specific Pax6 KO islets (c, d). On the other hand, Ki67+ YFP+ cells are detected in both the control and KO islets (arrowhead a-d). Quantification of Ki67+ YFP+ cells (e) in the islets of 7 week old mice at 4 weeks after tamoxifen induction (n = 3). Error bars represent SEM; *p<0.05.

doi:10.1371/journal.pone.0144597.g010

**Fig 11. Expression of C-peptide, PC1/3, and PC2 in the islets of beta-cell-specific Pax6 KO mice.** Double immunofluorescence staining of pancreatic cryosections from 2 month old mice at 4 weeks after tamoxifen induction. In beta-cell-specific Pax6 KO islets, C-peptide expression is lost from majority of the YFP+ Pax6-deficient cells (d). However, PC1/3 (e) and PC2 (f) expression is maintained in the YFP+ Pax6-deficient cells.

doi:10.1371/journal.pone.0144597.g011
was taking place. To clarify this further, we quantified the number of YFP\(^{-}\)/glucagon\(^{+}\) and YFP\(^{+}\)/glucagon\(^{+}\) cells in control and KO islets. We found that the number of YFP\(^{-}\)/glucagon\(^{+}\) cells was significantly increased in the KO islets at 1 month and 3 months of age but not at 8 months of age in mice from the non-inducible Glucagon-Cre line (Fig 20c). Similarly, this is also true for the Pax6 KO islets generated using the Tetracycline-inducible Glucagon-Cre mice (Fig 20b). The significant increase in the number of such reporter negative/glucagon\(^{+}\) cells may
correlate with glucagon neogenesis in the KO animals. At the same time, the number of YFP+ glucagon+ cells was significantly diminished in the KO islets, as such cells become glucagon negative upon Pax6 depletion, and further initiate the expression of ghrelin.

**Pax6 ablation in somatostatin and PP cells of the adult pancreas**

Finally, we were also interested to examine the consequences of Pax6 ablation in somatostatin- and PP-producing cells. As no mouse line specifically expressing the Cre recombinase in delta- or PP-cells was available, we decided to use ubiquitous Cre line [25] to induce the inactivation of Pax6 in the whole body of adult mice. Successful knockout was monitored by double immunostaining using Pax6 and glucagon/somatostatin antibodies (Fig 21). We checked the expression of ghrelin in combination with glucagon, insulin, somatostatin, and PP at 1 week, 2 week, and 3 week following tamoxifen induction. Our analyses indicate that glucagon cells appeared to be most sensitive to the loss of Pax6 gene activity. In fact, already at 1-week post induction, most of the glucagon positive cells start to express the ghrelin hormone. Two and three week later glucagon/ghrelin co-positive cells became nearly undetectable suggesting that by that time, all of the Pax6-deficient alpha-cells had lost the expression of glucagon. In contrast, in beta-cells of these Pax6 knockout mice, we noticed insulin/ghrelin co-positive cells at all the time points analysed with a slight increase in number with time. Interestingly, ghrelin positive cells expressing either somatostatin, or PP could not be detected at any stage analyzed (Fig 22). Hence, delta- and PP-cell content was not affected in these mutant islets. Furthermore, consistent with the alpha- and beta- cell specific Pax6 ablation, the ghrelin cells appearing in these ubiquitous KO islets were also distinguishable based on the fact that they were either Pdx1+.
(coming from beta cells) or Pdx1⁻ (coming from alpha cells) (Fig 23). It therefore appears that Pax6 inactivation has a variable effect in different subtypes of endocrine cells of the islet. Moreover, the effect of Pax6 loss appears to be more pronounced in alpha-cells correlating with the phenotype described in global Pax6 knockout [4].
In the present report we studied the consequences of \textit{Pax6} conditional inactivation in distinct endocrine cells in the adult mouse pancreas. While somatostatin- and PP-cells remained unaffected by the specific loss of \textit{Pax6} gene activity, insulin-producing beta-cells and glucagon-labeled alpha-cells lost their maturing molecular characteristics and were shunted towards ghrelin marked cells. Based on the findings presented above it is appropriate to conclude that \textit{Pax6} is required for the maturation of alpha- and beta-cells, and \textit{Pax6} inactivation does not alter cell-subtype specification.

\textbf{\textit{Pax6} in adult beta-cells}

The use of \textit{RIP-CreERT;Pax6\textsuperscript{0/\textmu}} mice allowed us to inactivate \textit{Pax6} in beta-cells upon tamoxifen induction. Our findings sustain the idea that \textit{Pax6} controls the maturation of insulin producing cells. Hence, in the absence of \textit{Pax6} we could demonstrate that following tamoxifen induction two crucial beta-cell-specific markers, Glut2, and MafA were found depleted. We further noticed a gradual decrease of the expression of an additional insulin-producing cell factor, Nkx6.1. Nkx6.1 expression has been shown to be reduced in hyperglycemic \textit{db/db} mice and T2DM human islets [36] as well as in beta-cells upon in vitro \textit{Pax6} knockdown [17].
Therefore, it seems that the Nkx6.1 expression may initially be affected by the absence of Pax6 and subsequently also by the developing hyperglycemia in these mice. In contrast, Pax6 depletion does not appear to affect Pdx1 gene activity that remained unchanged even when followed over longer time period. Moreover, lineage-tracing experiments using RIP-CreERT; Rosa26-YFP; Pax6fl/fl mice also demonstrated that Pax6-deficient beta-cells undergo a progressive loss of insulin labeling which is accompanied with an activation of ghrelin expression. This is corroborated by the detection of a transient insulin+/ghrelin+ cell population. Collectively, our data provide strong evidence for the crucial role of Pax6 in beta-cell maturation. However, in the absence of Pax6 the expression of Pdx1, and Rfx6, as well as PC1/3 and IAPP persists even few months following tamoxifen induction, clearly indicating that such cells, although immature, preserve their beta-cell destiny. However, in earlier studies in cell culture (Pax6 knockdown) [17], and in mice PC1/3 and Pdx1 were found reduced [7]. On the other hand, using our Pax6 conditional knockout mice crossed with Pax6 Cre, did not reveal any change in
Fig 17. Ghrelin+ cells originate from alpha-cells in the alpha-cell-specific Pax6 KO islets. Double immunofluorescence staining of pancreatic cryosections from 1 month old mice. Ghrelin expression is not detected in the control islets (a-c). In alpha-cell-specific Pax6 KO islets ghrelin expression is up regulated in YFP labeled cells (d-f) and rarely ghrelin expression co-localizes with glucagon expression (arrows d-f).

doi:10.1371/journal.pone.0144597.g017

Fig 18. Expression of alpha-cell-related transcription factors in ghrelin+ cells of alpha-cell-specific Pax6 KO islets. Double immunofluorescence staining of pancreatic cryosections from 1 month old mice. Ghrelin expression is not detected in the control islets (a-c). In alpha-cell-specific Pax6 KO islets ghrelin expression is up regulated in YFP labeled cells and ghrelin+ cells are negative for Pax6 (d) and MafB (e) expression but positive for Arx (f) expression.

doi:10.1371/journal.pone.0144597.g018
PC1/3 expression and still displayed some Pdx1+ and Cre-labeled cells [5]. Finally, Rfx6 was recently shown to play an important role in beta-cell maturation [37, 38]. Thus, the unaltered expression of Rfx6 in Pax6-deficient beta-cells suggests that it acts upstream of Pax6.

In addition, it appears that Pax6 might act on cell proliferation, as documented by our lineage tracing analyses where the number of YFP-labeled beta-cells in mutant islets of older

Fig 19. Expression of 7B2 and PC2 in alpha-cell-specific Pax6 KO islets. Immunofluorescence staining of pancreatic cryosections from 1 month old mice. 7B2 expression is very low in the control islets (a, b). In alpha-cell-specific Pax6 KO islets, 7B2 expression is highly up regulated in YFP labeled Pax6-deficient cells (e, f). Compared to control islets PC2 expression is unchanged in the alpha-cell-specific Pax6 KO islets (c, d and g, h).

doi:10.1371/journal.pone.0144597.g019
animals were significantly decreased as compared to controls, indicating that Pax6-deficient beta-cells may have lost the ability to proliferate over time. Pax6 binds and activates the promoter of GLP-1R [17]. Considering the role of GLP-1 in stimulating the glucose-dependent insulin secretion as well as beta-cell proliferation [39–41], a loss of GLP-1 receptor may contribute to defective insulin secretion and reduced proliferation of Pax6-deficient beta-cells. We therefore conclude that Pax6 may control beta-cell proliferation capacity and their maturation characteristics. Interestingly, Pax6 has been already shown to act on cell proliferation to control the pool of cell progenitors in the spinal cord [42]. On the other hand, the numbers of non-reporter labeled beta-cells (that are generated after tamoxifen induced Pax6 depletion) were increased in the mutant islets, and this augmentation was more pronounced in animals injected at younger age. In accordance with that a partial and modest recovery in hyperglycemia was
also apparent in young but not in older animals. This is probably due to the increased proliferation of beta-cells at younger age.

Finally, Pax6 may act on insulin processing, as documented by the augmented expression levels of proSAAS in mutant insulin producing cells. In Pax6-deficient beta-cells, the expression of proSAAS (the regulatory peptide for PC1/3) and 7B2 (the regulatory peptide for PC2) were highly augmented. Increased levels of proSAAS lead to defective proinsulin processing by inhibiting the activity of PC1/3 [29]. On the other hand, 7B2 may help to rescue the remaining processing by promoting the activity of PC2 [43, 44]. The finding of increased 7B2 in our study is in contrast to a previous study where 7B2 expression was down regulated after Pax6 knock-down [14]. However, we noticed an increased 7B2 expression not only during the loss of Pax6 in beta-, but also following Pax6 depletion in alpha-cells, as well as in all endocrine cells (using ubiquitous Cre recombinase) (S8 Fig). Interestingly, both proSAAS and 7B2 may also possess prohormone convertase unrelated activities as anti-aggregation chaperones [45, 46]. Therefore, their increased expression might also be related to the deterioration of the Pax6-lacking cell.

**Pax6 in alpha-cells**

The ablation of Pax6 function in glucagon-producing alpha-cells revealed a similar phenotype to what we found when inactivating Pax6 in insulin producing beta-cells. Accordingly, the absence of Pax6 in alpha-cells provokes a gradual loss of glucagon expression that is accompanied with a depletion of MafB labeling. Similar to MafA in insulin producing beta-cells, MafB is a marker for alpha-cell maturation [47, 48]. Interestingly, the expression of the alpha-cell determination factor Arx persists in glucagon producing cells that lost Pax6 gene activity. Thus, Pax6 does not act on Arx and therefore does not appear to promote alpha-cell specification, as suggested previously [16]. Hence, these findings suggest that, in contrast to PP- and somatostatin-expressing delta-cells, alpha-cells and beta-cells of the endocrine pancreas require Pax6 gene
activity for their maturation. This is corroborated by an earlier report using Pax6 global knock-out mice where the number of ghrelin positive cells was found increased [7]. However, in this study the fate of Pax6-deficient cells was not determined, and the described increase in ε-cells was not directly associated with the conversion of Pax6/− cells to ghrelin labeled cells. In fact, mutant cells eventually lose insulin/glucagon expression and appear only ghrelin positive, when analysed at later time points [7]. In contrast to the previous study we have used lineage tracing...
to unequivocally show the conversion of Pax6-/− cells into ghrelin marked cells. Moreover, during the analysis of the endocrine hormones following the inactivation of Pax6 using the ubiquitous Cre line, we could detect insulin+/ghrelin+, as well as glucagon+/ghrelin+ cells in mutant islets. Collectively, our analyses may also sustain the conclusion that ghrelin represents a marker for immature endocrine cells in the pancreas.

Supporting Information

S1 Fig. Expression of insulin, ghrelin, and beta-cell related factors in the pancreata of beta-cell-specific Pax6 KO mice. Quantitative RT-PCR of insulin, MafA, Nkx6.1, Glut2, Pdx1, Nkx2.2, PC1/3, PC2, and ghrelin mRNA in the pancreata of 4.5 month old mice at 3 months after tamoxifen induction (n = 2). Error bars represent SEM; *p<0.05.

(TIF)

S2 Fig. Alterations in the population of glucagon+, somatostatin+, and PP+ cells in the islets of beta-cell-specific Pax6 KO mice. Quantification of glucagon+, somatostatin+, and PP+ cells from 2 month old mice at 4 weeks after tamoxifen induction (n = 3). Error bars represent SEM; *p<0.05.

(TIF)

S3 Fig. Apoptosis is not induced in the Pax6-deficient cells of the beta-cell-specific Pax6 KO islets. TUNEL staining of pancreatic cryosections from 7 week old mice at 4 weeks after tamoxifen induction. TUNEL+ cells are not detected in the beta-cell-specific Pax6 KO islets (c, d).

(TIF)

S4 Fig. Pax6 does not bind to Glut2 and ghrelin promoters. Cross-linked chromatin from Min6 cells was precipitated by anti-Pax6 antibody and analyzed by PCR for the corresponding promoter regions. Pax6 interacts with MafA promoter region 3 but not with Glut2 and ghrelin promoters.

(TIF)

S5 Fig. Ghrelin expression is upregulated in Pax6-deficient cells of the alpha-cell-specific Pax6 KO islets. Double immunofluorescence staining of pancreatic cryosections from 1 month old mice. Ghrelin expression is not detected in the YFP+ Pax6+ cells of the control islets.

Fig 23. Ghrelin expressing cells, in the islets of adult-ubiquitous Pax6 KO mice, can be differentially identified. Double immunofluorescence staining of pancreatic cryosections from mice that were injected with tamoxifen at 2 months of age and analysed at 3 weeks post tamoxifen induction. Ghrelin+ cells are not detected in the control islets (a). In the ubiquitous Pax6 KO islets, ghrelin expression is up regulated and ghrelin+ cells may or may not express Pdx1 depending on the fact that they originally come from insulin+ or glucagon+ cells, respectively (b).

doi:10.1371/journal.pone.0144597.g023
In alpha-cell-specific Pax6 KO islets ghrelin expression is upregulated in YFP labeled Pax6-deficient cells (d-f).

**S6 Fig. Expression of alpha-cell related transcription factors in ghrelin+ cells of alpha-cell-specific Pax6 KO islets.** Double immunofluorescence staining of pancreatic cryosections from 1 month old mice. Ghrelin expression is not detected in the control islets (a-c). In alpha-cell-specific Pax6 KO islets ghrelin expression is upregulated in YFP labeled cells and ghrelin+ cells are negative for MafB expression (d-f). Rarely some YFP+ ghrelin+ cells are also detected in the KO islets and they are also negative for MafB (d-f).

**S7 Fig. Expression of alpha-cell related transcription factors in ghrelin+ cells of alpha-cell-specific Pax6 KO islets.** Double immunofluorescence staining of pancreatic cryosections from 1 month old mice. Ghrelin expression is not detected in the control islets (a-c). In alpha-cell-specific Pax6 KO islets ghrelin expression is upregulated in YFP labeled cells and ghrelin+ cells are positive for Arx expression (d-f). Rarely some YFP+ ghrelin+ cells are also detected in the KO islets and they are also positive for Arx (d-f).

**S8 Fig. Gradual increase in the population of ghrelin and 7B2 expressing cells in the islets of adult-ubiquitous Pax6 KO mice.** Double immunofluorescence staining of pancreatic cryosections from mice that were injected with tamoxifen at 2 months of age and analysed at 1 week and 3 weeks post tamoxifen induction. Ghrelin+ cells are not detected and the expression of 7B2 is very low in the control islets (a-c). At 1 week after tamoxifen induction few cells start to express 7B2 and ghrelin at a higher level in the KO islets (d-f). At 3 weeks after tamoxifen induction a large number of cells express high levels of 7B2 and ghrelin in the KO islets (g-i). 7B2 expression in the KO islets may or may not co-localize with ghrelin expression (d-i).

TI = tamoxifen induction.

**S1 Text. Supplementary materials and methods.**

**Acknowledgments**

We would like to thank Dr. J.F. Habener, Dr. C. Wright, Dr. C. Tomasetto, Dr. I. Lindberg, and Developmental Studies Hybridoma Bank for kindly providing antibodies. We are indebted to Dr. P. Herrera, Dr. D. Melton, Dr. A. McMahon, Dr. J. Whitsett, and Dr. F. Constantini for providing mouse lines. We would like to thank Prof. H. Jäckle for his constant support.

**Author Contributions**

Conceived and designed the experiments: AM ZA PC. Performed the experiments: ZA MR. Analyzed the data: ZA MR PC AM. Contributed reagents/materials/analysis tools: ZA MR PC AM. Wrote the paper: ZA PC AM.

**References**

1. Collombat P, Hecksher-Sorensen J, Serup P, Mansouri A. Specifying pancreatic endocrine cell fates. Mech Dev. 2006; 123(7):501–12. PMID: 16822656
2. Pan FC, Wright C. Pancreas organogenesis: from bud to plexus to gland. Dev Dyn. 2011; 240(3):530–65. doi: 10.1002/dvdy.22584 PMID: 21337462
3. Sosa-Pineda B, Chowdhury K, Torres M, Oliver G, Gruss P. The Pax4 gene is essential for differentiation of insulin-producing beta cells in the mammalian pancreas. Nature. 1997; 386(6623):399–402. PMID: 9121556

4. St-Onge L, Sosa-Pineda B, Chowdhury K, Mansouri A, Gruss P. Pax6 is required for differentiation of glucagon-producing alpha-cells in mouse pancreas. Nature. 1997; 387(6631):406–9. PMID: 9163426

5. Ashery-Padan R, Zhou X, Marquardt T, Herrera P, Toube L, Berry A, et al. Conditional inactivation of Pax6 in the pancreas causes early onset of diabetes. Dev Biol. 2004; 269(2):479–88. PMID: 1510714

6. Wang J, Elghazi L, Parker SE, Kizilokac H, Asano M, Sussel L, et al. The concerted activities of Pax4 and Nkx2.2 are essential to initiate pancreatic beta-cell differentiation. Dev Biol. 2004; 266(1):178–89. PMID: 14729487

7. Hart AW, Mella S, Mendrychowski J, van Heyningen V, Kleinjan DA. The developmental regulator Pax6 is essential for maintenance of islet cell function in the adult mouse pancreas. PLoS One. 2013; 8(1):e54173. doi: 10.1371/journal.pone.0054173 PMID: 23326594

8. Sander M, Neubuser A, Kalamaras J, Ee HC, Martin GR, German MS. Genetic analysis reveals that Pax6 is required for normal transcription of pancreatic hormone genes and islet development. Genes Dev. 1997; 11(13):1662–73. PMID: 9224716

9. Collombat P, Xu X, Ravassard P, Sosa-Pineda B, Dussaud S, Billestrup N, et al. The ectopic expression of Pax4 in the mouse pancreas converts progenitor cells into alpha and subsequently beta cells. Cell. 2009; 138(3):449–62. doi: 10.1016/j.cell.2009.05.035 PMID: 19665969

10. Collombat P, Mansouri A. Turning on the beta-cell identity in the pancreas. Cell Cycle. 2009; 8(21):3450–1. PMID: 19838059

11. Al-Hasani K, Pfeifer A, Courtney M, Ben-Othman N, Gjernes E, Vieira A, et al. Adult duct-lining cells can reprogram into beta-like cells able to counter repeated cycles of toxin-induced diabetes. Dev Cell. 2013; 26(1):86–100. doi: 10.1016/j.devcel.2013.05.018 PMID: 23810513

12. Pfeifer A, Courtney M, Ben-Ohman N, Al-Hasani K, Gjernes E, Vieira A, et al. Induction of multiple cycles of pancreatic beta-cell replacement. Cell Cycle. 2013; 12(20):3243–4. doi: 10.4161/cc.26357 PMID: 24036539

13. Nishimura W, Rowan S, Salameh T, Maas RL, Bonner-Weir S, Sell SM, et al. Preferential reduction of beta cells derived from Pax6-MafB pathway in MafB deficient mice. Dev Biol. 2008; 314(2):443–56. doi: 10.1016/j.ydbio.2007.12.009 PMID: 18199433

14. Katz LS, Gosmain Y, Marthinet E, Philippe J. Pax6 regulates the proglucagon processing enzyme PC2 and its chaperone 7B2. Molecular and cellular biology. 2009; 29(8):2322–34. doi: 10.1128/MCB.01543-08 PMID: 19223471

15. Gosmain Y, Marthinet E, Cheyssac C, Guerardel A, Mamin A, Katz LS, et al. Pax6 controls the expression of critical genes involved in pancreatic (alpha) cell differentiation and function. The Journal of biological chemistry. 2010; 285(43):33881–93. doi: 10.1074/jbc.M110.147215 PMID: 20592023

16. Heller RS, Stoffers DA, Liu A, Schedl A, Crenshaw EB 3rd, Madsen OD, et al. The role of Brn4/Pou3f4 and Pax6 in forming the pancreatic glucagon cell identity. Dev Biol. 2004; 268(1):123–34. PMID: 15031110

17. Gosmain Y, Katz LS, Masson MH, Cheyssac C, Poisson C, Philippe J. Pax6 is crucial for beta-cell function, insulin biosynthesis, and glucose-induced insulin secretion. Mol Endocrinol. 2012; 26(4):696–709. doi: 10.1210/me.2011-1256 PMID: 22403172

18. Zhang X, Rowan S, Yue Y, Heaney S, Pan Y, Brendolan A, et al. Pax6 is regulated by Meis and Pbx homeoproteins during pancreatic development. Developmental biology. 2006; 300(2):748–57. PMID: 17049510

19. Delporte FM, Pasque V, Devos N, Manifold I, Voz ML, Motte P, et al. Expression of zebrafish pax6b in pancreas is regulated by two enhancers containing highly conserved cis-elements bound by PDX1, PBX and PREP factors. BMC developmental biology. 2008; 8:53. doi: 10.1186/1471-213X-8-53 PMID: 18485195

20. Ashery-Padan R, Marquardt T, Zhou X, Gruss P. Pax6 activity in the lens primordium is required for lens formation and for correct placement of a single retina in the eye. Genes Dev. 2000; 14(21):2701–11. PMID: 11069887

21. Dor Y, Brown J, Martinez OI, Melton DA. Adult pancreatic beta-cells are formed by self-duplication rather than stem-cell differentiation. Nature. 2004; 429(6987):41–6. PMID: 15129273

22. Herrera PL. Adult insulin- and glucagon-producing cells differentiate from two independent cell lineages. Development. 2000; 127(11):2317–22. PMID: 10804174

23. Perl AK, Wert SE, Nagy A, Lobe CG, Whitsett JA. Early restriction of peripheral and proximal cell lineages during formation of the lung. Proc Natl Acad Sci U S A. 2002; 99(16):10482–7. PMID: 12145322
Zhu X, Lindberg I. 7B2 facilitates the maturation of proPC2 in neuroendocrine cells and is required for the expression of enzymatic activity. J Cell Biol. 1995; 129(6):1641–50. PMID: 7790360

27. Fortenberry Y, Hwang JR, Apletalina EV, Lindberg I. Functional characterization of ProSAAS: similarities and differences with 7B2. J Biol Chem. 2002; 277(7):5175–86. PMID: 11719503

28. Spijker HS, Song H, Ellenbroek JH, Roefs MM, Engelse MA, Bos E, et al. Loss of beta-Cell Identity Occurs in Type 2 Diabetes and Is Associated With Islet Amyloid Deposits. Diabetes. 2015; 64(8):2926–38. doi: 10.2337/db14-1752 PMID: 25918235

29. Liu T, Zhao Y, Tang N, Feng R, Yang X, Lu N, et al. Pax6 directly down-regulates Pcsk1n expression thereby regulating PC1/3 dependent proinsulin processing. PLoS One. 2012; 7(10):e46934. doi: 10.1371/journal.pone.0046934 PMID: 23056534

30. Thorens B, Wu YJ, Leahy JL, Weir GC. The loss of GLUT2 expression by glucose-unresponsive beta cells of db/db mice is reversible and is induced by the diabetic environment. J Clin Invest. 1992; 90(1):77–85. PMID: 1634622

31. Xu X, D’Hoker J, Stange G, Bonne S, De Leu N, Xiao X, et al. Beta cells can be generated from endogenous progenitors in injured adult mouse pancreas. Cell. 2008; 132(2):197–207. doi: 10.1016/j.cell.2007.12.015 PMID: 18243096

32. Bonner-Weir S, Baxter LA, Schuppin GT, Smith FE. A second pathway for regeneration of adult exocrine and endocrine pancreas. A possible recapitulation of embryonic development. Diabetes. 1993; 42(12):1715–20. PMID: 8243817

33. Furuta M, Yano H, Zhou A, Rouille Y, Holst JJ, Carroll R, et al. Defective prohormone processing and altered pancreatic islet morphology in mice lacking active SPC2. Proc Natl Acad Sci U S A. 1997; 94(13):6646–51. PMID: 9192619

34. Gelling RW, Du XQ, Dichmann DS, Romer J, Huang H, Cui L, et al. Lower blood glucose, hyperglycogonemia, and pancreatic alpha cell hyperplasia in glucagon receptor knockout mice. Proc Natl Acad Sci U S A. 2003; 100(3):1438–43. PMID: 12552113

35. Courtney M, Gjernes E, Druelle N, Vieira A, Ben-Othman N, et al. The Inactivation of Arx in the Adult Endocrine Pancreas. Diabetes. 2015; 64(8):3305–16. doi: 10.1172/JCI65390 PMID: 23863625

36. Chandra V, Albargi-Curiel O, Hastoy B, Piccand J, Randriamampita C, Vaillant E, et al. RFX6 regulates insulin secretion by modulating Ca2+ homeostasis in human beta cells. Cell reports. 2014; 9(6):2206–18. doi: 10.1016/j.celrep.2014.11.010 PMID: 25497100

37. Piccand J, Strasser P, Hodson DJ, Meunier A, Ye T, Keime C, et al. Rfx6 maintains the functional identity of adult pancreatic beta cells. Cell reports. 2014; 9(6):2219–32. doi: 10.1016/j.celrep.2014.11.033 PMID: 25497096

38. MacDonald PE, El-Kholy W, Riedel MJ, Salapatek AM, Light PE, Wheeler MB. The multiple actions of GLP-1 on the process of glucose-stimulated insulin secretion. Diabetes. 2002; 51 Suppl 3:S434–42. PMID: 12475787

39. Buteau J, Foisy S, Joly E, Prentki M. Glucagon-like peptide 1 induces pancreatic beta-cell proliferation via transactivation of the epidermal growth factor receptor. Diabetes. 2003; 52(1):124–32. PMID: 12502502

40. Wang Q, Li L, Xu E, Wong V, Rhodes C, Brubaker PL. Glucagon-like peptide-1 regulates proliferation and apoptosis via activation of protein kinase B in pancreatic INS-1 beta cells. Diabetologia. 2004; 47 (3):478–87. PMID: 14762654

41. Bel-Vialar S, Medevielle F, Pitulello F. The on/off of Pax6 controls the tempo of neuronal differentiation in the developing spinal cord. Dev Biol. 2007; 305(2):659–73. PMID: 17399698

42. Farber CR, Chitwood J, Lee SN, Verdugo RA, Islas-Trejo A, Rincon G, et al. Overexpression of Scg5 increases enzymatic activity of PCSK2 and is inversely correlated with body weight in congenic mice. BMC Genet. 2008; 9:34. doi: 10.1186/1471-2156-9-34 PMID: 18439298

43. Helwig M, Lee SN, Hwang JR, Ozawa A, Medrano JF, Lindberg I. Dynamic modulation of prohormone convertase 2 (PC2)-mediated precursor processing by 7B2 protein: preferential effect on glucagon synthesis. J Biol Chem. 2011; 286(49):42504–13. doi: 10.1074/jbc.M111.281980 PMID: 22013069
45. Helwig M, Hoshino A, Berridge C, Lee SN, Lorenzen N, Otzen DE, et al. The neuroendocrine protein 7B2 suppresses the aggregation of neurodegenerative disease-related proteins. J Biol Chem. 2013; 288(2):1114–24. doi:10.1074/jbc.M112.417071 PMID: 23172224

46. Hoshino A, Helwig M, Rezaei S, Berridge C, Eriksen JL, Lindberg I. A novel function for proSAAS as an amyloid anti-aggregant in Alzheimer's disease. J Neurochem. 2014; 128(3):419–30. doi:10.1111/jnc.12454 PMID: 24102330

47. Artner I, Hang Y, Mazur M, Yamamoto T, Guo M, Lindner J, et al. MafA and MafB regulate genes critical to beta-cells in a unique temporal manner. Diabetes. 2010; 59(10):2530–9. doi:10.2337/db10-0190 PMID: 20627934

48. Artner I, Le Lay J, Hang Y, Elghazi L, Schisler JC, Henderson E, et al. MafB: an activator of the glucagon gene expressed in developing islet alpha- and beta-cells. Diabetes. 2006; 55(2):297–304. PMID: 16443760