Impaired Plasma Membrane Targeting of Grb2–Murine Son of Sevenless (mSOS) Complex and Differential Activation of the Fyn–T Cell Receptor (TCR)-ζ-Cbl Pathway Mediate T Cell Hyporesponsiveness in Autoimmune Nonobese Diabetic Mice

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Summary

Nonobese diabetic (NOD) mouse thymocytes are hyporesponsive to T cell antigen receptor (TCR)-mediated stimulation of proliferation, and this T cell hyporesponsiveness may be causal to the onset of autoimmune diabetes in NOD mice. We previously showed that TCR-induced NOD T cell hyporesponsiveness is associated with a block in Ras activation and defective signaling along the PKC/Ras/MAPK pathway. Here, we report that several sequential changes in TCR-proximal signaling events may mediate this block in Ras as activation. We demonstrate that NOD T cell hyporesponsiveness is associated with the (a) enhanced TCR-β--associated Fyn kinase activity and the differential activation of the Fyn–TCR-ζ-Cbl pathway, which may account for the impaired recruitment of ZAP70 to membrane-bound TCR-ζ; (b) relative inability of the murine son of sevenless (mSOS) Ras GDP releasing factor activity to translocate from the cytoplasm to the plasma membrane; and (c) exclusion of mSOS and PLC-γ1 from the TCR-ζ-associated Grb2/pp36–38/ZAP70 signaling complex. Our data suggest that altered tyrosine phosphorylation and targeting of the Grb2-containing complex may block the downstream activation of Ras and Ras-mediated amplification of TCR/CD3-mediated signals in hyporesponsive NOD T cells. These findings implicate mSOS as an important mediator of downregulation of Ras signaling in hyporesponsive NOD T cells.

1Abbreviations used in this paper: GRF, GDP releasing factor; GST, glutathione S-transferase; IDDM, insulin-dependent diabetes mellitus; MAPK, mitogen-activated protein kinase; mSOS, murine son of sevenless; NOD, nonobese diabetic; PLC, phospholipase C; PTK, protein tyrosine kinase; p-Tyr, phosphotyrosine; SH, Src homology domain.

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ness may be causal to the onset of IDDM, and stimulated us to investigate the biochemical mechanisms of induction of NOD T cell hyporesponsiveness. We found that R as activation is deficient in quiescent and TCR-stimulated NOD thymocytes, and that this deficiency does not result from either a decreased amount or activity of the R as GAP activating protein p120 R as GAP, a negative regulator of R as activation (8). Our findings demonstrated that T cell hyporesponsiveness is linked to a block in R as activation in vivo. Reduced R as activity was also shown to correlate with the reduced activity and tyrosine phosphorylation of the mitogen-activated protein kinase (MAPK) in NOD thymocytes (8). Since MAPK activity is required for progression to S phase of the cell cycle, reduced tyrosine phosphorylation of MAPK may abrogate its activity in TCR-stimulated NOD T cells and elicit their proliferative hyporesponsiveness (8). Deficient R as activation and altered MAPK and Jnk protein kinase activities were also recently observed in anergic murine CD4+ T cells (9, 10).

After TCR engagement by MHC-bound peptides, R as activation is positively regulated by a protein tyrosine kinase (PTK)-dependent signaling pathway that elicits T cell differentiation and proliferation (11, 12). The phosphotyrosine (p-Tyr)-dependent recruitment of GDP releasing factors (GRFs) to the membrane and the assembly of R as-GRF complexes are essential for R as activation. GRFs activate R as by promoting the conversion of GDP-bound R as to the active GTP-bound state (13, 14). M urine of sevenless (mSOS) is a GRF that positively activates R as in T cells (15, 16). The adaptor protein Grb2 recruits and translocates mSOS to the plasma membrane (16, 17), which leads to R as-mediated signaling even in the absence of TCR stimulation (18). R as activation may also be indirectly controlled by phospholipase C (PLC)-γ1 (19, 20), after PLC-γ1 is activated and translocated to the plasma membrane after its tyrosine phosphorylation (20).

Translocation of mSOS is dependent on the Lck and Fyn PTK-mediated tyrosine phosphorylation of TCR-associated CD3 subunits, which form docking sites for the binding of secondary signaling proteins containing Src homology 2 (SH2) domains (e.g., Grb2) and recruit the ZAP70 cytoplasmic PTK to the TCR-CD3 complex (21–25). In TCR-activated cells, Grb2 also forms a complex with a 36–38-kD tyrosine phosphoprotein, pp36–38 (26–28), another docking site for SH2 domain-containing proteins. The PTK-dependent pathway of TCR activation, which induces the association of mSOS, ZAP70, PLC-γ1, and other phosphoproteins with Grb2 and ζ-chain of the TCR complex (28), is essential for R as activation.

In this study, we further investigated the mechanism of inhibition of R as activation in TCR-stimulated hyporesponsive NOD thymocytes. We show that in NOD thymocytes the differential activation of the Fyn-TCR-ζ-Cbl pathway may account for the impaired recruitment of ZAP70 to membrane-bound TCR-ζ. We also demonstrate a significant reduction in mSOS GRF activity in TCR-stimulated NOD thymocytes, which is mediated by the inability of mSOS to be translocated in association with Grb2 to the plasma membrane. Our findings implicate mSOS as an important mediator of downregulation of TCR-mediated R as signaling in hyporesponsive NOD thymocytes.

Materials and Methods

Mice. Female NOD/Del as well as control C57BL/6j and BALB/cj (insulitis-free and diabetes-resistant) mice were either bred in our specific pathogen-free animal facility at The John P. Robarts Research Institute colony or were purchased from The Jackson Laboratory (Bar Harbor, ME), and were used at 6–8 wk of age. C57BL/6j and BALB/cj were chosen as control strains, since we previously showed that C57BL/6j and BALB/cj thymocytes proliferate normally in response to TCR stimulation of proliferation in vitro (3, 5). Antibodies and Glutathione S-transferase (GST) Fusion Proteins. The mAbs used were the following: biotin-conjugated hamster H57-597 anti-TCR-β and rat L3T4 anti-CD4 (PharMingen, San Diego, CA); rat IgG1 anti-H-Ras, mouse anti-Lck, mouse anti-TCR-ζ (6B10.2), and mouse PY20 (IgG2b) anti-p-Tyr (Santa Cruz Biotechnology, Santa Cruz, CA); mouse anti-Grb2, mouse anti-mSOS1, and mouse anti-mZAP70 (Transduction Laboratory, Lexington, KY). Mouse anti-Fyn and anti-TCR-ζ mAbs and a polyclonal anti-ZAP70 antiserum were gifts from Dr. J. Bolen (Bristol-Myers Squibb, Princeton, NJ). The following polyclonal rabbit Abs were supplied by Santa Cruz Biotechnology: anti-mSOS1/2, anti-Grb2, anti-Cbl, and anti-PLC-γ1C. The polyclonal rabbit anti-mouse Lck and anti-mouse Fyn Abs were provided by Dr. A. Veillette (McGill University, Montreal, Quebec, Canada). R rabbit polyclonal anti-TCR-ζ serum 387 was obtained from Dr. L. Samelson (National Institutes of Health, Bethesda, MD). GST murine Grb2-SH2 domain fusion protein (GST–Grb2-SH2) was provided by Drs. D. Moto and G. Koretsky (University of Iowa, Iowa City, IA).

Cell Activation and Lysis. NOD, C57BL/6j, and BALB/cj thymocytes or peripheral splenic T cells purified on murine T cell enrichment columns (R&D Systems, Minneapolis, MN) (purity >95%) were maintained on ice in DM EM supplemented with 20 mM HEPES (all GIBCO BRL, Burlington, Ontario, Canada) until stimulation. The total number of thymocytes and percent distribution of CD4+CD8+ double-positive and CD4+ and CD8+ single-positive thymocytes are very similar in NOD and C57BL/6j control mice (3). This ensures that the differences observed do not reflect an unappreciated change in thymocyte composition in these mouse strains. Where not otherwise indicated, quiescent thymocytes or T cells (4 × 10^6/ml) were stimulated (3 min at 37°C) with 1 μg/10^6 cells of the biotin-conjugated H57-597 hamster anti-mouse TCR-β mAb either alone or together with the biotin-conjugated rat anti-mouse CD4 L3T4 mAb. Crosslinking of mAbs was accomplished using streptavidin or protein G (Sigma Chemical Co., St. Louis, MO) for various times at a 4:1 wt/wt ratio. Cells were lysed in ice-cold 50 mM Tris, pH 8.0, 150 mM NaCl, 5 mM EGTA lysis buffer containing 1% Brij 97 or Triton X-100, 0.02% NP-40, 5% glycerol, and supplemented with a mixture of protease and phosphatase inhibitors (100 μM β-naphthylamidemthylketone, 1 mM PM SF, 10 μg/ml aprotinin, 10 μg/ml leupeptin, 5 μg/ml pepstatin, 2 mM NaF, 10 mM Na3PO4, and 10 mM NaF) (all obtained from Sigma Chemical Co.). Alternatively, RIPA buffer (20 mM Tris pH 8.0, 0.15 M NaCl, 1 mM EDTA, 1 mM EGTA, 0.1% SDS, 1% NP-40, 0.5% Na deoxycholate, plus above mixture of protease and phosphatase inhibitors) was used to lyse cells and immunoprecipitate...
Fyn and Cbl in Fig. 5. All subsequent steps were performed at 4°C. Lysates were clarified of detergent-insoluble material by centrifugation (10 min, 14,000 rpm), precleared with protein A–Sepharose CL-4B (Pharmacia Biotech, Inc., Baie d’Urfé, Quebec, Canada), and 50–100 μl aliquots were quantified for their amounts of protein by the Bradford assay using BSA as standard.

Subcellular Fractionation. Cells (10⁶) were resuspended and lysed by brief sonication in ice-cold 10 mM Tris, pH 7.4, 10 mM KCl, 1.5 mM MgCl₂, 2 mM EGTA hypotonic buffer containing the above-described mixture of protease and phosphatase inhibitors (buffer A). Lysates were adjusted to 150 mM NaCl, centrifuged to remove nuclei and debris, and particulate membrane-containing (P100) and soluble cytoplasm-containing (S100) fractions (buffer A). Membrane fractions were washed with ice-cold buffer A and solubilized by sonication in buffer A supplemented with 150 mM NaCl and 1% Triton X-100. After recentrifugation (10 min, 100,000 g), the detergent-insoluble cytoskeleton-containing fraction was extracted with RIPA buffer and recentrifuged.

Immunoprecipitations and Affinity Precipitations of Cellular Proteins. Precleared postnuclear fractions obtained from 2–4 × 10⁹ cells were normalized for protein concentration levels and immunoprecipitated (3 h at 4°C) with the specific polyclonal Abs or control isotype-matched preimmune Ig precoupled to 25 μl of protein A–Sepharose CL-4B, protein A/G agarose (Santa Cruz Biotechnology) or streptavidin immobilized on 4% beaded agarose (Sigma Chemical Co.). This was followed by four washes of the precipitates with ice-cold lysis buffer. For affinity precipitations, GST fusion proteins (10 μg) or control GST (10 μg) noncovalently coupled to glutathione–agarose beads were reacted (2 h at 4°C) with cell lysates and washed extensively in lysis buffer.

Cell Electrophoresis and Immunoblotting. Precipitated proteins were solubilized in 2–× Laemmli sample buffer containing 2-ME, 20 mM EDTA, and 2 mM NaVO₄, resolved by SDS-PAGE (8–16% gradient gel; Novex, San Diego, CA) under reducing conditions, transferred to Immobilon (Millipore Corp., Bedford, MA) or nitrocellulose (Schleicher & Schuell, Keene, NH) membranes and immunoblotted with the indicated Abs. Blots were developed by enhanced chemiluminescence (Amersham Life Science, Inc., Arlington Heights, IL). Signal intensities were quantified using a Molecular Imager System and Molecular Analyzing software (Bio-Rad, Hercules, CA). Immunoblots revealed that quiescent NOD and control thymocytes contain equivalent amounts of the TCR-ζ, CD3e, ZAP70, Syk, Fyn, Lck, Grb2, Cbl, mSOS1, PLC-γ1, and phosphatidylinositol 3-kinase (p85 subunit) proteins.

In Vitro Kinase Assay. Precipitated proteins (2 × 10⁷ T cell equivalents/sample) were assayed for activated in vitro kinase activity after washing the beads in kinase buffer (25 mM Hapes, pH 7.4, 5 mM MgCl₂) containing 0.1% NP-40 and incubation (15 min at 20°C) with either [γ-³²P]ATP (10 μCi; New England Nuclear, Boston, MA) or 500 μCi cold ATP in 25 μl kinase buffer containing 0.1% NP-40. Reactions were stopped by addition of an ice-cold buffer containing 50 mM Tris, pH 8.0, 150 mM NaCl, 5 mM MgCl₂, 20 mM EDTA, 0.2% NP-40, and 10 mM NaF. Proteins were resolved by SDS-PAGE as described above, transferred to Immobilon membranes, and treated (1 h at 55°C) with 1 M KOH to remove alkali-labile phosphate groups from threonine- and serine-phosphorylated proteins (29). Membranes were immunoblotted serially, and overlay of autoradiograms confirmed the nature of the phosphoproteins.

³²P-Labeling and Peptide Mapping of Fyn. Cells were incubated for 2 h in phosphate-free DMEM supplemented with 2% dialyzed FCS and 20 μM Hepes, pH 7.3, containing 0.5 μCi/ml of carrier-free ortho³²P]phosphate (New England Nuclear), washed with phosphate-free DMEM, and lysed. Immunoprecipitated ³²P-labeled Fyn was resolved by SDS-PAGE, transferred onto a nitrocellulose membrane and cleaved by cyanogen bromide (50–100 mg/ml; Sigma Chemical Co.) for 1 h in 70% (vol/vol) formic acid (30). ³²P-labeled Fyn fragments were resolved by SDS-PAGE (24% gel) in a Tricine cathode buffer, and autoradiograph signal intensities were quantified.

Results

Ras GTPase Activity Is Deficient in TCR-stimulated NOD T thymocytes due to the Inability of mSOS to Translocate to the Plasma Membrane in Association with Grb2. Upon TCR cross-linking, SOs generally redistributes to a plasma membrane–localized Ras and TCR–CD3 complex in association with Grb2 (25, 26). To investigate whether this type of redistribution occurs in TCR-stimulated NOD thymocytes, we analyzed the plasma membrane associated mSOS-regulated RasGRF activity before and after TCR cross-linking. In NOD thymocytes, the amount of TCR-induced mSOS-dependent Ras as GDP releasing activity in the membrane-containing fraction was lower than that observed in control C57BL/6J thymocytes, and this low level of mSOS GTPase activity was not increased significantly after TCR-β cross-linking (Fig. 1 A). Anti-mSOS mAb immunoblotting performed after normalization for protein concentration levels, and the approximately equivalent amounts of constitutively membrane-associated Ras were observed in each sample revealed that relatively small amounts of mSOS associated with the plasma membrane in NOD and control thymocytes in the absence of TCR-β ligation (Fig. 1 B). Whereas TCR-β and TCR-β–CD4 cross-linking induced the recruitment of mSOS to the plasma membrane in control C57BL/6J and BALB/c thymocytes, this recruitment was markedly decreased in activated NOD thymocytes. Since Ras as deficiency in TCR-stimulated hyporesponsive NOD thymocytes does not result from altered
amounts or activity of p120 Ras-GAP (8), deficient mSOS-associated Ras GRF activity and impaired recruitment of mSOS to the plasma membrane may account for the decrease in Ras activation observed here.

To investigate whether deficient Ras activation is caused by the inability of Grb2 to translocate to the plasma membrane and/or its incapacity to recruit mSOS and PLC-γ1C into TCR signaling complexes, we determined the relative amounts of Grb2 in cytoplasmic and membrane fractions of NOD and control thymocytes. Immunoblotting of Grb2 immunoprecipitates from membrane fractions with anti-Grb2, anti-sOS, and anti-PLC-γ1C Abs demonstrated that at 1.5 min after TCR-β and TCR-β-CD4 cross-linking, the amounts of Grb2 and Grb2-associated mSOS and PLC-γ1C translocated to the plasma membrane were lower in NOD thymocytes than control thymocytes (Fig. 1C). Moreover, TCR-β and TCR-β-CD4 cross-linking induced marked increases in the membrane/cytoplasmic ratio for Grb2 and Grb2-associated mSOS and PLC-γ1C only in control strain thymocytes. The membrane/cytoplasmic ratios observed reflect the amounts of Grb2 and mSOS and PLC-γ1C translocated from the cytoplasm to the plasma membrane.
the plasma membrane in the close vicinity of the TCR signaling complex and its downstream effectors, including Ras. Thus, the TCR-induced deficiency in Ras GRF activity in NOD thymocytes appears to be mediated by the inability of Grb2 to translocate to the plasma membrane. The observed differences in the amounts of the Grb2-mediated recruitment of mSOS and PLC-γ1C to the plasma membrane in NOD and control thymocytes persisted at longer times (5 and 20 min after TCR-β-CD4 cross-linking) of TCR stimulation, and are not due to differences in the kinetics of Grb2 recruitment (data not shown). In addition, we did not observe any plasma membrane translocation of the Shc phosphoprotein in TCR-stimulated NOD and C57BL/6J thymocytes (our unpublished observations), suggesting that Shc may not influence the membrane targeting of SOs in thymocytes.

Altered Tyrosine Phosphorylation and Targeting of the Grb2/pp36–38/ZAP70 Complex to the Plasma Membrane and Cytoskeleton Impair TCR-Induced Signaling in NOD T-lymphocytes. We investigated whether the TCR-stimulated tyrosine phosphorylation and interactions of various docking phosphoproteins, which associate with Grb2 and recruit mSOS and PLC-γ1C into TCR-Ras signaling complexes, are deficient and contribute to a block in Ras activation in TCR-stimulated NOD thymocytes. In particular, we examined whether TCR ligation (a) induces an increase in TCR-ζ-ZAP70 association and ZAP70 tyrosine phosphorylation in the membrane fraction of NOD thymocytes and (b) enables Grb2 and ZAP70 to translocate to the TCR and promote the membrane targeting of the Grb2-SO complex. Immunoprecipitation of TCR-ζ immunoprecipitates from the membrane fraction with anti-ZAP70 and anti-Grb2 Abs revealed that relatively small amounts of ZAP70 and Grb2 associate with TCR-ζ in NOD and control thymocytes, even in the absence of TCR-β ligation (Fig. 2 A). TCR-β and TCR-β-CD4 cross-linking induced the recruitment of ZAP70 and Grb2 to the TCR-ζ-containing complex in control C57BL/6J and BALB/cj thymocytes; however, this recruitment was markedly decreased in activated NOD thymocytes. Similarly, TCR-CD4-induced tyrosine phosphorylation of ZAP70 was reduced considerably in NOD thymocytes compared with control thymocytes. Both before and after TCR-β and TCR-β-CD4 cross-linking, immunoblotting of TCR-ζ after anti-TCR-ζ immunoprecipitation showed that equal amounts of TCR-ζ were present in the membrane fractions of C57BL/6J, BALB/cj, and NOD thymocytes.

pp36–38 may mediate the inducible membrane targeting and recruitment of Grb2-SO and Grb2-PLC-γ1 by binding to the Grb2-SH2 domain (26–28). To determine if deficient recruitment of Grb2 to the plasma membrane correlates with diminished tyrosine phosphorylation of pp36–38 and decreased binding of Grb2 to pp36–38, phospho-pp36–38 was affinity precipitated with a GST-Grb2-SH2 fusion protein and immunoblotted with anti–p-Tyr. The extent of tyrosine phosphorylation of pp36–38 bound to GST-Grb2-SH2 was significantly reduced in TCR-β- and TCR-β-CD4-stimulated NOD thymocytes compared with control thymocytes (Fig. 2 B). Control affinity precipitations with GST alone did not yield any appreciable amount of bound pp36–38. Immunoprecipitation of mSOS1/2 and blotting with anti–p-Tyr revealed that while phospho-pp36–38 was induced to associate with mSOS1/2 in control C57BL/6J and BALB/cj thymocytes upon TCR-β

![Figure 2](image-url)

**Figure 2.** (A) TCR-induced association of TCR-ζ with ZAP70 and Grb2 in the plasma membrane and tyrosine phosphorylation of ZAP70 is markedly decreased in NOD thymocytes. NOD and control thymocytes were either unstimulated (N one) or stimulated for 15 min with anti-TCR-β or anti-TCR-β plus anti-CD4. TCR-ζ was immunoprecipitated from thymocyte membrane fractions (10⁶ cell equivalents/sample), and immunoprecipitates were immunoblotted with either anti-ZAP70, anti-Grb2, anti-p-Tyr, or anti-TCR-ζ Abs. The latter anti-TCR-ζ immunoblots showed that equal amounts of TCR-ζ were precipitated from these thymocyte membrane fractions before and after TCR-β and TCR-β-CD4 cross-linking. (B) Affinity precipitation of phospho-pp36–38 from the membrane fractions of TCR-β- and TCR-β-CD4-stimulated NOD and control thymocytes with the GST-Grb2-SH2 fusion protein (lanes 1–9) and GST alone (lanes 10) immobilized on glutathione-agarose beads. Precipitated proteins were immunoblotted with anti–p-Tyr. (C) Tyrosine phosphorylation of pp36–38 bound to mSOS1/2 in NOD thymocytes. NOD and control thymocytes were either unstimulated (N one) or stimulated as in A. Equal amounts of mSOS1/2 were immunoprecipitated from total cell lysates (4 × 10⁷ cell equivalents/sample), and immunoprecipitates were immunoblotted with either anti–p-Tyr or anti-mSOS1 mAbs. The results shown are representative of one of two separate reproducible experiments.
and TCR-β-CD4 cross-linking, about fourfold less phospho-pp36-38 was bound to mSOS 1/2 in stimulated NOD thymocytes (Fig. 2B). As Grb2-associated phospho-pp36-38 partitions exclusively in the particulate (membrane) fraction of cells (26, 27), these results suggest that a decrease in tyrosine phosphorylation of pp36-38 is responsible for the impaired recruitment of the Grb2-SOS complex to the plasma membrane in TCR-stimulated NOD thymocytes.

TCR ligation can induce the association of tyrosine-phosphorylated TCR-ζ chains with a cytoskeletal-enriched subcellular fraction of T cells, and this cytoskeletal association generally correlates with IL-2 production (32). Since TCR-stimulated NOD splenic T cells are deficient in IL-2 production (7), we tested whether TCR-β-CD4 cross-linking induces a decrease in the association of TCR-ζ, Grb2, and ZAP70 with the cytoskeleton in NOD T cells. Immunoblotting of proteins in a cytoskeleton-enriched fraction, isolated by differential centrifugation as described (25), with anti-TCR-ζ, anti-Grb2, and anti-ZAP70 Abs confirmed that these proteins are inducibly associated with the cytoskeleton after TCR-β-CD4 cross-linking in control BALB/cζ and C57BL/6ζ splenic T cells (Fig. 3). However, the cytoskeletal associations of TCR-ζ, Grb2, and ZAP70 are reduced considerably in NOD splenic T cells after coligation of TCR-β and CD4. These results suggest that the TCR-ζ-induced deficiencies in association of TCR-ζ, Grb2, and ZAP70 with the cytoskeleton may mediate the proliferative hyporesponsiveness and reduced IL-2 secretion of NOD T cells.

TCR Ligation Induces Enhanced TCR-ζ-associated Fyn Kinase Activity in NOD T Cells. In anergic CD4+ peripheral T cells, Fyn activity is elevated (33, 34) and Fyn, but neither Lck nor ZAP70, associates with TCR-ζ (29). To test whether similar events occur in hyporesponsive NOD thymocytes, the levels and kinetics of TCR-ζ-associated Fyn activity and the relative capacities of Fyn to bind to TCR-ζ induced upon TCR ligation were determined. A rapid increase in tyrosine phosphorylation of membrane-localized Fyn, CD3, and TCR-ζ was noted (Fig. 4A). The most striking difference between NOD and control C57BL/6ζ thymocytes was the elevated autophosphorylation of membrane-bound Fyn both in quiescent (no cross-linking) and TCR-β-stimulated NOD thymocytes. This was accompanied by the transient tyrosine hyperphosphorylation of CD3 and TCR-ζ. At each time of analysis (0–20 min), slightly increased amounts of Fyn coprecipitated with TCR-β before and after cross-linking, indicating that Fyn is constitutively associated with TCR in the membrane. Anti-Lck and anti-ZAP70 mAb immunoblotting confirmed that increased Fyn activity was not due to the presence of Lck or ZAP70 in TCR-β immunoprecipitates (data not shown). A significant increase in basal (no cross-linking) and TCR-β cross-linking–induced tyrosine phosphorylation of total cellular Fyn was also observed in NOD thymocytes (Fig. 4B), and a similar result was obtained using an in vitro kinase assay (Fig. 4C).

To analyze the basis of increased Fyn activity, we investigated whether Fyn is present in a constitutively active form in quiescent NOD thymocytes relative to control thymocytes. The status of phosphorylation of Fyn residue Tyr528 was determined, as this residue is the site of negative regulation by the Csk PTK and positive regulation by the CD45 protein tyrosine phosphatase. Hyperphosphorylation of Tyr528 inactivated Fyn, whereas a reduction in or absence of phosphorylation of Tyr528 stimulates Fyn kinase activity. Cyanogen bromide cleavage of 32P-labeled Fyn and subsequent phosphopeptide mapping studies were performed to distinguish between 32P-labeled peptides resulting from phosphorylation of Fyn at NH2-terminal sites of phosphorylation and phosphorylation at the Tyr528 autoregulatory site. In NOD thymocytes, the COOH-terminal Tyr528-containing Fyn phosphopeptide is phosphorylated only very weakly relative to the NH2-terminal peptide containing sites of serine, threonine, and tyrosine phosphorylation, yielding a NH2/COOH peptide 32P-labeling ratio of 1.7 (Fig. 4D). In contrast, the levels of phosphorylation of the COOH-terminal Tyr528-containing Fyn phosphopeptide and NH2-terminal peptide were essentially equivalent in C57BL/6ζ thymocytes, as a NH2/COOH peptide 32P-labeling ratio of 0.9 was observed. The relative amounts of 32P-label in the NH2-terminal Fyn phosphopeptides were about equal in NOD and C57BL/6ζ thymocytes.

The above-described studies suggest that TCR-associated Fyn is directly responsible for the observed TCR-induced increases in its autophosphorylation, based on our inability to detect Lck and ZAP70 in Fyn immunoprecipitates (our unpublished data). This raised the possibility that this heightened autophosphorylation and binding of Fyn to TCR-ζ may account for the impaired recruitment of ZAP70 to membrane-bound TCR-ζ in TCR-stimulated NOD thymocytes as noted earlier in Fig. 2A. To test this possibility further, we compared the relative amounts of TCR-ζ that associate with ZAP70 independently of Fyn as a result of TCR-β-CD4 cross-linking in NOD and control thymocytes. A marked increase in Fyn-associated TCR-ζ was found in activated NOD thymocytes (Fig. 4C). When supernatants of these Fyn immunoprecipitates were preclaved for detectable Fyn, then immunoprecipitated with anti-ZAP70 and immunoblotted with anti-TCR-ζ, less ZAP70-associated TCR-ζ was detected in stimulated NOD thymocytes. These data support the idea that the in-
creased binding of TCR-ζ to TCR-associated Fyn diminishes the capacity of TCR-ζ to interact with ZAP70 in the plasma membrane of TCR-stimulated NOD thymocytes.

TCR Stimulation Induces the Differential Activation of the Fyn-Cbl Signaling Pathway in NOD Thymocytes. In T cells, Fyn preferentially interacts with Cbl, and TCR cross-linking-induced tyrosine phosphorylation of Cbl is mediated by Fyn activity (35, 36). The ability of Cbl to inhibit Syk PTK activity indicates that Cbl may function as a negative regulator of intracellular signaling (37). Therefore, we analyzed whether differential activation of the Fyn signaling pathway affects the function of Cbl in NOD thymocytes. TCR-β cross-linking rapidly induced the tyrosine hyperphosphorylation of total cellular Cbl in NOD thymocytes but not control C57BL/6 J and B6.BR/J thymocytes (Fig. 5 A). In contrast, upon TCR-β-C4D cross-linking, this difference between NOD and control thymocytes was less pronounced, which indicates that the high levels of tyrosine phosphorylation of Cbl induced in control thymocytes might mask the difference observed upon TCR cross-linking.

The ability of Fyn to phosphorylate Cbl as a substrate was examined after immunoprecipitation of Cbl from precleared postnuclear fractions of unstimulated thymocyte lysates. The specificity of the in vitro cold kinase assay was maintained by using the high stringency RIPA buffer for cell lysis and immunoprecipitation of Fyn and Cbl. Fyn and Cbl RIPA immunoprecipitates did not contain associated kinase activity, based on the detection of Fyn and cbl but neither Lck, ZAP70, Syk, nor any additional phosphoproteins in these precipitates. Anti-p-Tyr immunoblotting revealed that TCR-β cross-linking in NOD thymocytes rapidly (within 1.5 min) enhanced Fyn autophosphorylation and Fyn-mediated tyrosine phosphorylation of Cbl, which was significantly higher in NOD than control C57BL/6 J thymocytes (Fig. 5 B). Thus, the Fyn-Cbl signaling pathway seems to be highly activated in TCR-stimulated NOD thymocytes.

Figure 4. (A) Time course of activation of TCR-β-associated kinase activity in TCR-β-stimulated NOD and B6 thymocytes. NOD and control B6 thymocytes (2 × 10⁶ cells/lane) were incubated for 3 min in the presence of biotinylated anti-TCR-β. Cell-bound mAbs were either not cross-linked (0 min) or were cross-linked for the indicated times in the presence of protein G. Cells were washed to remove unbound mAbs, and TCR-β-immune complexes were immunoprecipitated from precleared postnuclear fractions of thymocyte lysates using streptavidin immobilized on 4% beaded agarose and then assayed for their associated in vitro kinase activity. Membranes were then immunoblotted serially with different mAbs, and overlay of autoradiograms and immunoblots demonstrated equal loading in each lane (data not shown) and confirmed the nature of the detected phosphoproteins (anti-Fyn immunoblotting, data not shown; anti-TCR-ζ; immunoblotting, middle). Supernatants precleared of Fyn were immunoprecipitated with anti-ZAP70, and the amounts of residual (Fyn-independent) TCR-ζ in these precipitates were analyzed by immunoblotting (bottom). (D) The level of Tyr528 phosphorylation of Fyn is decreased in quiescent NOD thymocytes. [32P]Phosphate labeling and peptide mapping of Fyn immobilized to membrane. [32P]Phosphate-labeled Fyn fragments were resolved by SDS-24% PAGE in a Tricine cathode buffer, and the signal intensities from autoradiographs were quantified. Results are expressed as the ratio of the signal intensity of the potential NH2-terminal sites of serine, threonine, and tyrosine phosphorylation (NH2) relative to the Tyr528-containing regulatory COOH-terminal site (COOH). The results shown are representative of one of two separate reproducible experiments.
Our major findings are that NOD T cell hyporesponsiveness in NOD thymocytes by conducting biochemical analyses of the pathway of downregulation of TCR-proximal signaling and inhibition of Ras activity. Our major findings are that NOD T cell hyporesponsiveness is mediated by (a) enhanced TCR-β-associated Fyn activity and the differential activation of the Fyn-TCR-ζ-Cbl pathway; (b) deficient translocation of mSOS and PLC-γ1 in association with Grb2 from the cytoplasm to the plasma membrane; and (c) a reduction in the capacity of the mSOS GRF to activate Ras and deliver downstream signals essential for T cell proliferation.

This impaired recruitment of Grb2-mSOS and Grb2-PLC-γ1C complexes to a membrane-localized TCR complex correlates with the diminished abilities of Grb2 and ZAP70 to interact with TCR-associated pp36-38 and TCR-ζ. Accordingly, changes in the tyrosine phosphorylation of the TCR complex may interfere with the targeting of these Grb2-containing complexes to membrane-localized Ras, and seem to be key steps in the pathway that lead to the consequent block in Ras activation in hyporesponsive T cells. It is possible that the uncoupling of the Grb2-mSOS complex from pp36-38 and TCR-ζ may prevent the derepression of the COOH-terminal autoinhibitory domain of mSOS, which normally occurs upon the formation of a trimeric complex between Grb2, mSOS, and docking phosphoproteins (38).

In contrast with our observations presented here, the block in Ras activation in an anergic murine CD4+ Th1 clone does not result from either a defect in association between Shc, Grb2, and mSOS or from a defect in Shc phosphorylation (9). The reasons for the discrepancies between the latter data and ours are presently unclear. However, it is intriguing to note that, in B cells, Ras as activation may be inhibited under negative signaling conditions, and that this block in Ras as activation is associated with diminished Shc-Grb2 interaction and attenuation of the Grb2-mSOS signal (39). Thus, different signaling mechanisms appear to mediate T cell hyporesponsiveness, dependent on the origin of the T cells and their state of differentiation and activation. Nonetheless, a common feature of the different mechanisms of T cell hyporesponsiveness reported is that a block in Ras activation accompanies the anergic state.

Fyn activity may transduce the signals that mediate T cell hyporesponsiveness by a mechanism in which TCR-dependent signals originate from the association of Fyn with TCR-ζ in the TCR-CD3 complex. In contrast with TCR ligation in the presence of appropriate costimulation, which elicits the association of TCR-ζ with ZAP70, ligation of TCR by an alloantigen alone stimulates the tyrosine phosphorylation of TCR-ζ and its association with Fyn (29). The latter Fyn-TCR-ζ interaction mediates T cell hyporesponsiveness. Our analyses of Fyn activity in response to TCR-β cross-linking demonstrated a relative increase in both the tyrosine phosphorylation of Fyn and binding of TCR-associated phospho-Fyn to TCR-ζ in NOD versus control thymocytes. Phosphopeptide mapping studies revealed that the high basal activity of Fyn in quiescent NOD thymocytes is attributable to the dephosphorylation of Fyn at its Tyr528 autoregulatory site. Thus, increased Fyn activity may play a major role in the induction of NOD thymocyte hyporesponsiveness.

This suggested role of increased Fyn activity in T cell hyporesponsiveness is consistent with the following observations. First, in mice homozygous for the lpr and gld mutations, TCR stimulation of hyporesponsive peripheral T lymphocytes elicits increased Fyn activity and the constitutive tyrosine hyperphosphorylation of TCR-ζ as well as other components of the TCR-CD3 complex (40, 41). Second, in Th1 cells, an increase in Fyn activity correlates...

**Figure 5.** (A) Cbl tyrosine phosphorylation in NOD and control thymocytes after TCR-β or TCR-β-CD4 stimulation. Cbl was immunoprecipitated from precleared postnuclear RIPA fractions of unstimulated thymocyte lysates, and was used as a substrate for Fyn. Kinase assay samples were resolved on SDS-PAGE under reducing conditions, transferred to nitrocellulose, and immunoblotted with an anti-p-Tyr mAb.

**Table 1.**

| Condition | B6 | B/c | NOD | B6 | B/c | NOD | B6 | B/c | NOD |
|-----------|----|----|-----|----|----|-----|----|----|-----|
| Crosslinking | None | TCR | TCR/CD4 | IP: anti-Cbl | 1.0 | 0.7 | 3.2 | 3.6 | 2.3 | 13.3 | 5.9 | 5.5 | 7.8 |

Discussion

Previously, we found that TCR-induced T cell hyporesponsiveness may be causal to the onset of autoimmune diabetes in NOD mice (3, 6, 7), and that this hyporesponsiveness is associated with a block in Ras activation and altered tyrosine phosphorylation of downstream effectors of Ras as activation in NOD T cells (8). In this report, we extended our investigation of the mechanism(s) of TCR-induced hyporesponsiveness in NOD thymocytes by conducting biochemical analyses of the pathway of downregulation of TCR-proximal signaling and inhibition of Ras as activation. Our major findings are that NOD T cell hyporesponsiveness is mediated by (a) enhanced TCR-β-associated Fyn activity and the differential activation of the Fyn-TCR-ζ-Cbl pathway; (b) deficient translocation of mSOS and PLC-γ1 in association with Grb2 from the cytoplasm to the plasma membrane; and (c) a reduction in the capacity of the mSOS GRF to activate Ras and deliver downstream signals essential for T cell proliferation.

This impaired recruitment of Grb2-mSOS and Grb2-PLC-γ1C complexes to a membrane-localized TCR complex correlates with the diminished abilities of Grb2 and ZAP70 to interact with TCR-associated pp36-38 and TCR-ζ. Accordingly, changes in the tyrosine phosphorylation of the TCR complex may interfere with the targeting of these Grb2-containing complexes to membrane-localized Ras, and seem to be key steps in the pathway that lead to the consequent block in Ras as activation in hyporesponsive T cells. It is possible that the uncoupling of the Grb2-mSOS complex from pp36-38 and TCR-ζ may prevent the derepression of the COOH-terminal autoinhibitory domain of mSOS, which normally occurs upon the formation of a trimeric complex between Grb2, mSOS, and docking phosphoproteins (38).

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with the induction of anergy (42, 43), suggesting that these changes in Fyn activity play a crucial role in maintaining hyporesponsiveness to antigen. Third, we recently found that increased Fyn activity is not mediated by Lck in activated NOD thymocytes (Zhang, J., K. Salojin, and T.L. Delovitch, manuscript submitted for publication). Therefore, it is conceivable that the increased activity of Fyn in hyporesponsive T cells is due to a deficiency in its tyrosine dephosphorylation. Analyses of the activity of several protein tyrosine phosphatases in NOD thymocytes are required to explore this possibility.

Interestingly, TCR-ζ-CD4 stimulation induces an increased binding of TCR-ζ-associated phospho-Fyn to TCR-ζ in NOD thymocytes. This may give rise to the impaired recruitment of ZAP70 to membrane-bound TCR-ζ and the deficient phosphorylation of TCR-ζ-associated ZAP70 observed in NOD thymocytes. Thus, the increased association of Fyn with TCR-ζ may represent an important early TCR-induced event of an anergic response in T cells.

Our observation that the Fyn–TCR-ζ-Cbl signaling pathway is differentially activated in TCR-stimulated NOD T cells may also explain how increased Fyn activity contributes to the induction of hyporesponsiveness in NOD T cells. Fyn preferentially interacts with Cbl in T cells (35, 36), because tyrosine phosphorylated Cbl is found only in Fyn-containing and not in Lck-containing immune complexes (36; our unpublished observations). The amount of Fyn-associated Cbl and extent of Fyn-mediated tyrosine phosphorylation of Cbl are generally enhanced upon TCR cross-linking (35). In this regard, we found that TCR cross-linking significantly augments the tyrosine phosphorylation of Cbl by Fyn in NOD thymocytes relative to control thymocytes. These findings may have important implications since overexpression of Cbl blocks Syk activity, Syk–Fc receptor interactions and intracellular signaling in rat basophils (37), and the Cbl homologue sl-1 functions as a negative regulator of the Let-23 receptor PTK pathway of activation that involves Grb2 and Ras homologues in Caenorhabditis elegans (44). Thus, elevated and/or altered Fyn–TCR-ζ-Cbl interactions may disrupt downstream ZAP70–Grb2–Ras activation and signaling in T cells. Further studies are required to elucidate the mechanism and physiological role of differential activation of the Fyn–TCR-ζ-Cbl pathway in hyporesponsive NOD thymocytes.

We thank Drs. J. Bolen, A. Veillette, L. Samelson, D. Otto, and G. Koretsky for their kind gifts of reagents; S. Rowland and C. Richardson for maintaining our mouse colony; all members of our laboratory for their valuable advice and encouragement; and Dr. J. Madrenas for his critical evaluation of the manuscript. We also thank M. A. Leaist for her expert assistance with the preparation of this manuscript.

This work was supported by grants from the Juvenile Diabetes Foundation International, a Medical Research Council of Canada Juvenile Diabetes Foundation International Diabetes Interdisciplinary Research Program, and the Helen M. Armstrong grant from the Canadian Diabetes Association. K. Salojin, J. Zhang, and B. Gill were recipients of Juvenile Diabetes Foundation International postdoctoral fellowships, and G. Arreaza was the recipient of a Canadian Diabetes Association postdoctoral fellowship.

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Received for publication 11 April 1997 and in revised form 10 July 1997.

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