Research Article

The Late Osteoblast/Preosteocyte Cell Line MLO-A5 Displays Mesenchymal Lineage Plasticity In Vitro and In Vivo

Dongqing Yang 1, Stan Gronthos 2,3, Sandra Isenmann 2,3, Howard A. Morris 4 and Gerald J. Atkins 1

1 Biomedical Orthopaedic Research Group, Centre for Orthopaedic and Trauma Research, Adelaide Medical School, Faculty of Health and Medical Sciences, University of Adelaide, Adelaide, SA 5005, Australia
2 Mesenchymal Stem Cell Laboratory, Adelaide Medical School, Faculty of Health and Medical Sciences, Adelaide University, Adelaide, SA 5005, Australia
3 South Australian Health and Medical Research Institute, Adelaide, SA 5000, Australia
4 Musculoskeletal Biology Research, School of Pharmacy and Medical Sciences, University of South Australia, Adelaide, SA 5000, Australia

Correspondence should be addressed to Gerald J. Atkins; gerald.atkins@adelaide.edu.au

Received 4 June 2018; Revised 28 October 2018; Accepted 11 November 2018; Published 17 January 2019

Academic Editor: Hector Mayani

Copyright © 2019 Dongqing Yang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The process of osteoblast switching to alternative mesenchymal phenotypes is incompletely understood. In this study, we tested the ability of the osteoblast/preosteocyte osteogenic cell line, MLO-A5, to also differentiate into either adipocytes or chondrocytes. MLO-A5 cells expressed a subset of skeletal stem cell markers, including Sca-1, CD44, CD73, CD146, and CD166. Confluent cultures of cells underwent differentiation within 3 days upon the addition of osteogenic medium. The same cultures were capable of undergoing adipogenic and chondrogenic differentiation under lineage-appropriate culture conditions, evidenced by lineage-specific gene expression analysis by real-time reverse-transcription-PCR, and by Oil Red O and alcian blue (pH 2.5) staining, respectively. Subcutaneous implantation of MLO-A5 cells in a gel foam into NOD SCID mice resulted in a woven bone-like structure containing embedded osteocytes and regions of cartilage-like tissue, which stained positive with both alcian blue (pH 2.5) and safranin O. Together, our findings show that MLO-A5 cells, despite being a strongly osteogenic cell line, exhibit characteristics of skeletal stem cells and display mesenchymal lineage plasticity in vitro and in vivo. These unique characteristics suggest that this cell line is a useful model with which to study aging and disease-related changes to the mesenchymal lineage composition of bone.

1. Introduction

Osteoblasts derive from mesenchymal skeletal stem cells (SSCs) and actively lay down the organic phase of bone matrix and regulate its mineralisation. The differentiating osteoblast is commonly thought to have three possible fates, including apoptosis or differentiation into either bone lining cells or mature osteocytes. Osteoblasts, rather than being terminally differentiated, have been shown to retain mesenchymal lineage plasticity [1] and have been shown capable of differentiating into adipocytes and chondrocytes [2–6]. Recently, in mice, it was found that even apparently mature osteocytes, expressing green fluorescent protein (GFP) under control of the dentin matrix protein-1 (Dmp1) promoter, can escape from the bone matrix ex vivo, and in doing so revert to a more immature osteoblastic phenotype [7]. The phenomenon of lineage plasticity among osteoblast lineage cells is of increasing interest with the emerging knowledge surrounding the important regulatory role of the osteocyte in bone homeostasis. Furthermore, the observation that the tightly regulated process of bone formation (osteogenesis) decreases with age concomitant with an increase in bone marrow fat accumulation warrants the characterisation of cell line models permissive for the study of both cell fates.

The cell line MLO-A5 [8] was established from osteoblasts derived from the long bones of transgenic mice...
expressing SV40 large T antigen under control of a 2.6 kb osteocalcin promoter [9]. Since its isolation in 2001, multiple studies have demonstrated that this is a useful cell model, with which to study the differentiation of late-stage osteoblasts [10], the process of biomineralisation [11, 12], and the transition of osteoblasts to osteocytes [13]. MLO-A5 was initially described as an osteoblastic model that mineralises its extracellular matrix in culture [8], with a pattern of mineralisation indistinguishable from that found in lamellar bone [11]. MLO-A5 has been characterised as a late-osteoblast/preosteocyte model because of the high level of the mineralisation phase marker, tissue nonspecific alkaline phosphatase (Tnap) and the preosteocyte marker E11/gp38, but very low levels of the more mature osteocyte genes Sost and Fgf23 [14].

In order to test whether this osteogenic cell line also exhibited lineage plasticity, we first examined the immunophenotype of these cells and found they expressed markers in common with SSCs. We found that under the appropriate culture conditions, MLO-A5 cells poised to form a bone-like phenotype of these cells and found they expressed markers

2.1. Cell Culture and Osteogenic Differentiation. MLO-A5 cells were maintained in α-MEM media containing 10% FCS, 100 U/ml penicillin, 100 mg/ml streptomycin, 2 mM L-glutamine, and 10 mM HEPES (growth medium), as described previously [15]. For osteogenic differentiation, growth medium was supplemented with β-glycerol phosphate (10 mM; Sigma, Australia), dexamethasone (10 nM), and ascorbic acid (50 μg/ml; Sigma Chemical Company, St. Louis, MO, USA) [15]. For differentiation experiments, 3 × 10^4 cells/well were seeded into 24-well plates in growth medium and allowed to proliferate for 3 days to achieve 100% confluence to a “predifferentiation” stage, and this time point was recorded as differentiation day 0. From day 0, fresh osteogenic differentiation media were supplied to cultures every three days until day 12. Total RNA was extracted from duplicate wells on days 3, 6, and 12, as described below. To quantify mineral deposition at these time points, triplicate wells were stained using the alizarin red technique, as described previously [16].

2.2. Cell Surface Marker Staining and Flow Cytometry Analysis. Mouse primary bone marrow stromal cells (BMSCs) were isolated as described previously [17]. MLO-A5 cells and BMSCs were subjected to immunofluorescence and flow cytometry for reactivity with the monoclonal antibodies STRO-1 [18], anti-Sca-1, CD44, CD73, CD105, CD90, CD106, CD146, and CD166.

2.3. Adipogenic Differentiation. For adipogenic induction, confluent cultures of MLO-A5 cells grown in growth medium, as above, were switched (day 0) to media also containing media 60 μM indomethacin, 100 nM dexamethasone, and 50 μg/ml ascorbate-2-phosphate [19]. Fresh adipogenic differentiation medium was supplied to cultures every three days till day 21. In vitro fat droplet formation was visualised by Oil Red O (Sigma-Aldrich, Australia) staining on days 7, 14, and 21 of culture.

2.4. Chondrogenic Differentiation. For chondrogenic assays, MLO-A5 cells were harvested from 100% confluent 75 cm² tissue culture flasks generated in growth medium (day 0) as above. Aliquots of 1 × 10^6 cells were pelleted by centrifugation at 600 g for 5 min. Cell pellets were supplied with chondrogenic medium consisting of DMEM with 0.125% w/v bovine serum albumin, tissue culture additives (100 unit/ml penicillin and 100 mg/ml streptomycin, 2 mM L-glutamine, and 10 mM HEPES), 10 μM dexamethasone, 50 μg/ml ascorbate-2-phosphate, and 10 ng/ml TGF-β1 (R&D Systems, Minneapolis, MN, USA). Fresh chondrogenic medium was supplied to all cultures every two days till day 21. Total RNA was collected on days 3, 6, 12, and 21. On day 12, a cell pellet culture was also fixed in 4% w/v paraformaldehyde overnight for paraffin embedding and further histological analyses including staining with safranin O and alcin blue (pH2.5) [19].

2.5. Glycosaminoglycan (GAG) Assay. Chondrogenic differentiation was assessed in high-density (0.5-1 × 10^6 cells per well) cultures in 96-well plates with 10 ng/ml TGF-β1 for 48 hours as described previously [19]. Glycosaminoglycan synthesis was measured in quadruplicate wells by incorporation using a TopCount NXT Microplate Scintillation & Luminescence counter (Perkin Elmer Life and Analytical Sciences, Downers Grove, IL). Values for GAG synthesis were normalised to DNA content per well.

2.6. Xenograft Model. Approximately 5.0 × 10^6 MLO-A5 cells from confluent cultures, as above, were added to a gel-foam carrier (Zimmer Inc., Warsaw, IN) and then transplanted subcutaneously into the dorsal surface of 10-week-old immunocompromised NOD/SCID mice for 10 weeks as previously described [19]. MLO-A5-implanted carriers were fixed in 10% w/v buffered formalin and decalcified. Safranin O staining and alcin blue staining (with pH 2.5) were performed to characterise osteogenesis and chondrogenesis. An anti-SV40 large T antigen antibody (Santa Cruz Biotechnology, Texas, USA) followed by a peroxidase detection system (Envision Detection Systems, Peroxidase/DAB, Dako, Glostrup, Denmark) was used to confirm the cell origin within the tumour tissue. These procedures were performed in accordance to specifications of an approved animal protocol (University of Adelaide Animal Ethics Committee number M-2012-207 and SA Pathology Ethics Committee number 141/12).

2.7. Gene Expression Analysis. Total RNA was extracted from the cell cultures using the TRIZOL method (Invitrogen, Australia), and cDNA samples were synthesised
To measure the expression of genes of interest in mRNA level by real-time PCR, as described previously [15]. The primer sequences for measuring each gene are listed in Table 1. Gene expression was normalised for the housekeeping gene β-actin.

| Gene     | Sequence (5′-3′)                          | GenBank accession no. | Amplicon (bp) |
|----------|------------------------------------------|-----------------------|---------------|
| β-Actin  | Forward: cacaccgcgcaccagg<br>Reverse: ggaatacagcccgggga | NM_007393.5           | 115           |
| Runx2    | Forward: cagaagacgatcgcgagag<br>Reverse: ggtctggtgcgtgtgcct | NM_001146038.2        | 122           |
| Osx      | Forward: cgcggcccccgccccgg<br>Reverse: ggcgtgtggtgcctgcttcc | NM_130458.3           | 137           |
| Col1a1   | Forward: cccgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_007742.3           | 155           |
| Tnap     | Forward: gctgcggcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_007431.2           | 101           |
| Dmp1     | Forward: cgcgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_016779.2           | 121           |
| Ocn      | Forward: gctgcggcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_010288.3           | 79            |
| E11      | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_010329.2           | 158           |
| Phex     | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_011077.2           | 156           |
| Cebp     | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_001287514.1        | 89            |
| Pparg    | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_001308352.1        | 160           |
| Adipoq   | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_009605.5           | 98            |
| Slca2a4  | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_012751.1           | 370           |
| Fdhp4    | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_024406.2           | 129           |
| Acan     | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_007424.2           | 251           |
| Col2a1   | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_00113515.2         | 136           |
| Col10a1  | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_009925.4           | 346           |
| Sox9     | Forward: cgcgcgcgcgcgcgc<br>Reverse: ggtctggtgcggtgccca | NM_011448.4           | 120           |

(Superscript III) to measure the expression of genes of interest in mRNA level by real-time PCR, as described previously [15]. The primer sequences for measuring each gene are listed in Table 1. Gene expression was normalised for the housekeeping gene β-actin.

### 3. Results

#### 3.1. MLO-A5 Cells Express a MSC-Like Immunophenotype.

Flow cytometric analyses demonstrated that while the SSC surface markers STRO-1, CD90, CD105, and CD106 were barely detectable in MLO-A5 cells, the SSC markers Sca-1, CD44, CD73, CD146, and CD166 were expressed at high levels (Figure 1(a)). In comparison, mouse BMSCs expressed the expected immunophenotype. This suggested that MLO-A5 cells may possess at least some SSC-like qualities with respect to mesenchymal lineage commitment.

The mRNA levels of osteogenic specific transcriptional factors, runt-related transcription factor 2 (Runx2) and osterix (Osx) (Figure 1(b)), were on average 20-fold higher in osteogenic cultures compared to the cultures treated with adipogenic or chondrogenic conditions. Notably, both Runx2 and Osx mRNA levels rapidly increased from the day 0 value when cells were cultured in osteogenic media, peaking after 3 days, while the levels of these two genes remained unchanged with culturing under adipogenic or chondrogenic conditions.

#### 3.2. Osteogenic Differentiation.

Confluent cultures switched to osteogenic inductive conditions and underwent extensive mineralisation after just 3 days (Figure 2(a)), suggesting that by virtue of reaching confluence at day 0, the cells had already reached a pre-differentiated osteogenic state. The degree of mineral deposition increased progressively.
with time evidenced by alizarin red staining (Figure 2(a)). The mRNA levels of osteogenic differentiation-related genes in MLO-A5 cultures, including collagen type I alpha 1 (Col1a1), Tnap, osteocalcin (Ocn), and X-linked phosphate-regulating endopeptidase (Phex) (Figure 2(b)), increased significantly by day 3 compared to day 0 levels. The mRNA expression of dentin matrix protein 1 (Dmp1) and E11 did not change significantly over this period.

3.3. Adipogenic Differentiation. Under adipogenic inductive conditions, predifferentiated cultures of MLO-A5 developed oil droplet formation over a period of 7 days, as evidenced by Oil Red O staining (Figure 3(a)). The mRNA levels of adipogenic differentiation-related genes in MLO-A5 cultures including CCAAT/enhancer-binding protein, alpha (Cebpa), peroxisome proliferator-activated receptor gamma (Pparg), adiponectin (Adipoq), solute carrier family...
2 (Slc2a4), and fatty acid-binding protein 4 (Fabp4) all significantly increased compared to day 0 levels (Figure 3(b)).

3.4. Chondrogenic Differentiation. MLO-A5 cultures switched to chondrogenic inductive conditions stained negative by the safranin O method but positive under alcian blue staining at pH 2.5, suggesting the formation of unsulphated but not sulphated glycosaminoglycans [20–22] (Figure 4(a)). In vitro production of glycosaminoglycan synthesis under chondrogenic conditions was significantly increased by 30% \((p < 0.01)\) with the addition of TGF-β1 (Figure 4(b)). The mRNA level of the chondrogenic transcriptional factor, sex-determining region Y-box 9 (Sox9), increased compared to day 3 level (Figure 4(c)). The mRNA levels of Col1a1 and collagen type II alpha 1 (Col2a1) showed no significant changes over the 21-day culture period. Notably, the levels of Col1a1 were on average approximately 1500-fold higher than Col2a1 mRNA levels (Figure 4(c)). The mRNA expression of the chondrocyte matrix-related genes, aggrecan (Acan) and collagen type X alpha 1 (Col10a1), was undetectable, based on a 45-cycle real-time PCR reaction used in this study (data not shown).

3.5. Xenograft Implantation. After 10 weeks postimplantation of MLO-A5 grafts into NOD SCID mice, an ectopic bone/cartilage organ was observed. Histological analyses (H&E staining) revealed a woven bone-like tissue inside the graft containing embedded osteocyte-like cells (Figure 5(a)). Both alcian blue staining at pH 2.5 (Figure 5(b)) and safranin O staining (Figure 5(c)) confirmed that there was at least some genuine cartilage formation within the tumour graft. Specific immunostaining for the SV40 small and large T antigens confirmed that a large proportion of the cells was of MLO-A5 origin (Figures 5(d) and 5(e)).
4. Discussion

The osteoblast cell line MLO-A5 is a useful model, with which to study the process of mineralisation during osteoblast-osteocyte transition under osteogenic culture conditions, and does so in manner entirely consistent with this process in vivo [11]. In this study, we explored the mesenchymal lineage plasticity of the late osteoblast, using MLO-A5 cells already cultured to confluence and poised to rapidly become mineralised osteoblasts/preosteocytes [8]. Consistent with previous characterisation [8], “predifferentiated” MLO-A5 cells displayed strong capacity for osteogenic differentiation in vitro, evidenced by the rapid and extensive deposition after just 3 days of calcium phosphate mineral in monolayer cultures, and the expression of a panel of osteogenic differentiation-related markers including Runx2, Osx, Tnap, Ocn, E11, Phex, and Dmp1. Besides the osteoblastic phenotype, MLO-A5 cells display certain immunophenotypic features of SSC, evidenced by the cell surface expression of Sca-1, CD44, CD73, CD146, and CD166 [17, 23]. However, the minimal criteria for a true multipotent stromal cell include the expression of CD73, as well as CD90 and CD105 [24]. The absence of CD90 and CD105, together with the absence of the markers, STRO-1 and CD106, indicates that this cell line represents a late rather than an early SSC model. Despite this, MLO-A5 retains the capacity to differentiate into adipocyte and chondrocyte lineages, as well as undergoing osteogenesis, which to our knowledge is unique among available cell lines.

By switching these predifferentiated osteogenic cultures to adipogenic media, the formation of oil droplets and the mRNA levels of adipogenic-related genes including Cebpa, Pparg, Adipoq, Slc2a4, and Fabp4 were all induced, consistent with MLO-A5 cells retaining adipogenic lineage plasticity [17]. In chondrogenic cultures, the cartilage matrix products, GAGs, are visualised by alcian blue staining at pH 2.5, with both sulphated and unsulphated forms being detected [21, 22]. In contrast, safranin O staining only detects sulphated GAGs [20]. In this study, chondrogenic pellet cultures of MLO-A5 cells after 21 days did not stain positive by safranin O but were positive by alcian blue. In articular cartilage in vivo, unsulphated GAGs are connected
by the protein aggregan, encoded by the Acan/ACAN gene, which results in the formation of sulphated GAGs, and which are indicative of mature cartilage matrix [25]. Consistent with the staining pattern observed, the expression of Acan mRNA in the MLO-A5 chondrogenic cultures was undetectable after 45 cycles of real-time RT-PCR. Hence, in this study, only unsulphated and possibly nonfunctional GAGs are synthesised by MLO-A5 cells in vitro, presumably because of the lack of aggregan expression. Although the mRNA expression level of the chondrogenic transcription factor, Sox9, increased significantly over the 21-day period, the level of the downstream gene Col2a1 encoding type II collagen, the major collagenous protein in the developing articular cartilage tissue [26], remained unchanged. Our findings demonstrate that MLO-A5 cells are able to undergo a limited degree of adipogenic and chondrogenic differentiation when cultured under the appropriate culture conditions in vitro. However, the mechanism of MLO-A5 cell plasticity either via direct or indirect conversion to other lineages requires further assessment. For

Figure 4: (a) In vitro chondrogenesis by MLO-A5 in pellet cultures, as described in Materials and Methods, visualised by safranin O staining and alcian blue staining at pH 2.5. (b) GAG synthesis in MLO-A5 cells cultured under chondrogenic conditions in the presence or absence of recombinant TGF-β1. Data shown are means ± SEM of readings from quadruplicate wells; significance is indicated by **p < 0.01. (c) Chondrogenic gene expression determined by real-time RT-PCR analysis of Sox9 and Col2a1 as well as the Col1a1/Col2a1 ratio. Data are shown as mean fold – change ± SEM of biological triplicates assayed in duplicate from the day 3 value for each gene. Significant changes are indicated by **p < 0.01.
example, the extent to which cell division is required for lineage switching to occur, or whether a subpopulation of the cells differentiate directly into either adipocytes or chondrocytes, will be important to determine. Future studies will also investigate the transcriptional and epigenetic mechanisms that mediate dedifferentiation and lineage determination. In the xenograft model, where MLO-A5 cells in a gel-foam carrier were implanted subcutaneously into immunodeficient mice, MLO-A5-derived woven bone containing embedded osteocyte-like cells together with cartilage deposits was observed, evidenced by both safranin O and alcian blue staining within the ectopic organ formed, suggesting that additional host factors promote complete chondrogenic differentiation in this model.

5. Conclusions

To summarise, we have demonstrated that the MLO-A5 cell line displays mesenchymal plasticity in vitro and in vivo and is capable of undergoing either adipogenic or chondrogenic differentiation, despite this being a late-osteoblast/preosteocyte, strongly osteogenic cell line. However, the capacity of MLO-A5 cells to undergo adipogenesis and chondrogenesis is incomplete. Whether the lack of the stem cell markers CD90, CD105, and CD106 explains the inability of MLO-A5 to fully differentiate towards adipocytes and chondrocytes is yet to be determined. Furthermore, due to its original method of isolation from transgenic mice, with insertion of the transgene into the genome, and the possibility that the cell line carries unidentified chromosomal abnormalities that may impact on its ability to undergo multilineage differentiation, findings using this cell line should be treated with appropriate caution. Nevertheless, our findings with this cell line suggest that even up to a late stage of osteogenic differentiation, osteoblasts are still capable of switching to an alternative mesenchymal lineage. Recently, methotrexate-based chemotherapy induced bone loss and fat formation was shown to be attributed to the disruption of balance between adipogenic, osteogenic, and osteoclastogenic differentiation within the bone marrow [27]. Therefore, in addition to this cell line being a useful model, with which to study mesenchymal plasticity in the context of a cell line that can form mineralised bone, MLO-A5 is also potentially useful for studying the molecular and cellular basis of the age and disease-associated phenomenon of bone to fat formation.

Abbreviations

| Abbreviation | Description |
|--------------|-------------|
| Dmp1         | Dentin matrix protein-1 |
| Tnap         | Tissue nonspecific alkaline phosphatase |
| E11/gp38     | Podoplanin |
| Sost         | Sclerostin |
| Fgf23        | Fibroblast growth factor 23 |
| Runx2        | Run-related transcription factor 2 |
| Osx          | Osterix |
| Colla1       | Collagen type I alpha 1 |
| Ocn          | Osteocalcin |
| Phex         | X-linked phosphate-regulating endopeptidase |
| Cebpa        | CCAAT/enhancer-binding protein, alpha |
Pparg: Peroxisome proliferator-activated receptor gamma
Adipoq: Adiponectin
Scl2a4: Solute carrier family 2
Fabp4: Fatty acid-binding protein 4
Sox9: Sex-determining region Y-box 9
Col2a1: Collagen type II alpha 1
Col10a1: Collagen type X alpha 1
Acan: Aggrecan
BMSC: Bone marrow stromal cell
SCC: Skeletal stem cell
GAG: Glycosaminoglycan.

Data Availability
The data used to support the findings of this study are included within the article.

Conflicts of Interest
The authors declare that they have no conflicts of interest.

Acknowledgments
The authors thank Prof. Lynda Bonewald (Indiana University, IN, USA) for the provision of MLO-A5 cells. This study was supported by funding by the Project Grant Scheme of the National Health and Medical Research Council of Australia (NHMRC) (ID 1106029). SG and GJA were supported by NHMRC Research Fellowships.

References
[1] F. Liu, L. Malaval, and J. E. Aubin, “The mature osteoblast phenotype is characterized by extensive plasticity,” Experimental Cell Research, vol. 232, no. 1, pp. 97–105, 1997.
[2] A. D. Berendsen and B. R. Olsen, “Osteoblast–adipocyte lineage plasticity in tissue development, maintenance and pathology,” Cellular and Molecular Life Sciences, vol. 71, no. 3, pp. 493–497, 2014.
[3] M. H. Lafage-Proust, T. Thomas, A. Guignandon, L. Malaval, A. Rattner, and L. Vico, “Plasticity of osteoprogenitor cells,” Joint Bone Spine, vol. 74, no. 6, pp. 536–539, 2007.
[4] U. Noth, A. M. Osyczka, R. Tuli, N. J. Hickok, K. G. Danielson, and R. S. Tuan, “Multilineage mesenchymal differentiation potential of human trabecular bone-derived cells,” Journal of Orthopaedic Research, vol. 20, no. 5, pp. 1060–1069, 2002.
[5] Y. Yoshiko, K. Oizumi, T. Hasegawa et al., “A subset of osteoblasts expressing high endogenous levels of PPARγ switches fate to adipocytes in the rat calvaria cell culture model,” PLoS One, vol. 5, no. 7, article e11782, 2010.
[6] P. G. Robey, “Mesenchymal stem cells: fact or fiction, and implications in their therapeutic use,” F1000Res, vol. 6, 2017.
[7] E. Torreggiani, B. G. Matthews, S. Pejda et al., “Preosteocytoid osteocytes have the potential to dedifferentiate to a source of osteoblasts,” PLoS One, vol. 8, no. 9, article e75204, 2013.
[8] Y. Kato, A. Boskey, L. Spervak, M. Dallas, M. Hori, and L. F. Bonewald, “Establishment of an osteoid preosteocyte-like cell MLO-A5 that spontaneously mineralizes in culture,” Journal of Bone and Mineral Research, vol. 16, no. 9, pp. 1622–1633, 2001.
[9] Y. Kato, J. J. Windle, B. A. Koop, G. R. Mundy, and L. F. Bonewald, “Establishment of an osteocyte-like cell line, MLO-Y4,” Journal of Bone and Mineral Research, vol. 12, no. 12, pp. 2014–2023, 1997.
[10] J. D. Eick, C. Barragan-Adjemian, J. Rosser et al., “Silorane resin supports proliferation, differentiation, and mineralization of MLO-A5 bone cells in vitro and bone formation in vivo,” Journal of Biomedical Materials Research Part B: Applied Biomaterials, vol. 100B, no. 3, pp. 850–861, 2012.
[11] C. Barragan-Adjemian, D. Nicolella, V. Dusevich, M. R. Dallas, J. D. Eick, and L. F. Bonewald, “Mechanism by which MLO-A5 late osteoblasts/early osteocytes mineralize in culture: similarities with mineralization of lamellar bone,” Calcified Tissue International, vol. 79, no. 5, pp. 340–353, 2006.
[12] S. L. Dallas, P. A. Veno, J. L. Rosser et al., “Time-lapse imaging techniques for comparison of mineralization dynamics in primary murine osteoblasts and the late osteoblast/early osteocyte-like cell line MLO-A5,” Cells Tissues Organs, vol. 189, no. 1-4, pp. 6–11, 2009.
[13] J. Rosser and L. F. Bonewald, “Studying osteocyte function using the cell lines MLO-Y4 and MLO-A5,” in Bone Research Protocols, M. Helfrich and S. Ralston, Eds., vol. 816 of Methods in Molecular Biology, pp. 67–81, Humana Press, Totowa, NJ, USA, 2012.
[14] S. M. Woo, J. Rosser, V. Dusevich, I. Kalajzic, and L. F. Bonewald, “Cell line IDG-SW3 replicates osteoblast-to-late-osteocyte differentiation in vitro and accelerates bone formation in vivo,” Journal of Bone and Mineral Research, vol. 26, no. 11, pp. 2634–2646, 2011.
[15] D. Yang, A. G. Turner, A. R. Wijenayaka, P. H. Anderson, H. A. Morris, and G. J. Atkins, “Osteocyte-like cell line MLO-A5 that spontaneously mineralizes in culture: comparison with cetylpyridinium chloride extraction,” Analytical Biochemistry, vol. 329, no. 1, pp. 77–84, 2004.
[16] C. A. Gregory, W. Grady Gunn, A. Peister, and D. J. Prockop, “An alizarin red-based assay of mineralization by adherent cells in culture: comparison with cetylpyridinium chloride extraction,” Analytical Biochemistry, vol. 329, no. 1, pp. 77–84, 2004.
[17] Y. Kato, O. You, T. Yamabayashi et al., “Characterization of bone marrow derived mesenchymal stem cells in suspension,” Stem Cells Translational Medicine, vol. 3, 2012.
[18] P. J. Simmons and B. Torok-Storb, “Identification of stromal cell precursors in human bone marrow by a novel monoclonal antibody, STRO-1,” Blood, vol. 78, no. 1, pp. 55–62, 1991.
[19] S. Gronthos, A. C. Zannettino, S. J. Hay et al., “ Molecular and cellular characterisation of highly purified stromal stem cells derived from human bone marrow,” Journal of Cell Science, vol. 116, no. 9, pp. 1827–1835, 2003.
[20] L. Rosenberg, “Chemical basis for the histological use of safranin O in the study of articular cartilage,” The Journal of Bone & Joint Surgery, vol. 53, no. 1, pp. 69–82, 1971.
[21] S. Yamabayashi, “Periodic acid — Schiff — alcian blue: a method for the differential staining of glycoproteins,” The Histochemical Journal, vol. 19, no. 10-11, pp. 565–571, 1987.
[22] R. Jones and L. Reid, “The effect of pH on alcian blue staining of epithelial acid glycoproteins. II. Human bronchial submucosal gland,” The Histochemical Journal, vol. 5, no. 1, pp. 19–27, 1973.

[23] H. Yang, L. N. Gao, Y. An et al., “Comparison of mesenchymal stem cells derived from gingival tissue and periodontal ligament in different incubation conditions,” Biomaterials, vol. 34, no. 29, pp. 7033–7047, 2013.

[24] M. Dominici, K. le Blanc, I. Mueller et al., “Minimal criteria for defining multipotent mesenchymal stromal cells. The international society for cellular therapy position statement,” Cytotherapy, vol. 8, no. 4, pp. 315–317, 2006.

[25] C. Kiani, L. Chen, Y. J. Wu, A. J. Yee, and B. B. Yang, “Structure and function of aggrecan,” Cell Research, vol. 12, no. 1, pp. 19–32, 2002.

[26] D. Eyre, “Articular cartilage and changes in arthritis: collagen of articular cartilage,” Arthritis Research & Therapy, vol. 4, pp. 30–35, 2002.

[27] K. R. Georgiou, T. J. King, M. A. Scherer, H. Zhou, B. K. Foster, and C. J. Xian, “Attenuated Wnt/β-catenin signalling mediates methotrexate chemotherapy-induced bone loss and marrow adiposity in rats,” Bone, vol. 50, no. 6, pp. 1223–1233, 2012.