Prevalence of enterotoxins and other virulence genes of *Staphylococcus aureus* caused subclinical mastitis in dairy cows

Rania M. Ewida¹ and Amira A. T. Al-Hosary²

1. Department of Food Hygiene (Milk Hygiene), Faculty of Veterinary Medicine, New Valley University, New Valley 72511, Egypt; 2. Department of Animal Medicine (Infectious Diseases), Faculty of Veterinary Medicine, Assiut University, Assiut 71526, Egypt.

Corresponding author: Rania M. Ewida, e-mail: r_ewida@aun.edu.eg

Co-author: AATA: amirah hosary@yahoo.com

Received: 10-11-2019, Accepted: 05-05-2020, Published online: 26-06-2020

**doi:** www.doi.org/10.14202/vetworld.2020.1193-1198

**How to cite this article:** Ewida RM, Al-Hosary AAT (2020) Prevalence of enterotoxins and other virulence genes of *Staphylococcus aureus* caused subclinical mastitis in dairy cows, *Veterinary World*, 13(6): 1193-1198.

---

**Abstract**

**Background and Aim:** Milk production is one of the main props for the national economy. One of the crucial problems in this industry is subclinical mastitis, which harms this industry that considered the backbone of the economy. It is an infectious and zoonotic disease; the infection can spread between dairy animals through milkers’ hands, and milking machines, while the human infection occurs due to the consumption of apparently hygienic milk. *Staphylococcus aureus* is one of the main causative agents of clinical and subclinical mastitis. It is also considered one of the bacteria incriminated in food intoxication of humans due to its virulence factors as enterotoxins and toxic shock syndrome. The current study was designed to assess the prevalence of *S. aureus* and its enterotoxins, as well as, its other virulence factors in milk collected from cows that suffer from subclinical mastitis.

**Materials and Methods:** Sixty cows were collected from different dairy farms located in Assiut Governorate, Egypt. These cows were subjected to the clinical examination of the udder and its lymph nodes before sampling. Milk samples were collected from clinically healthy udders. All the milk samples were examined by California mastitis test (CMT), polymerase chain reaction (PCR), and enzyme-linked immunosorbent assay (ELISA) for confirmation subclinical mastitis, presence of *S. aureus* and its enterotoxins genes and other virulence factors in the examined milk samples.

**Results:** The cows included in the current study had healthy udders. The sixty collected milk samples were tested by CMT. 48/60 (80.0%) were positive samples; from the 48 positive samples, 46 (95.83%) samples were confirmed positive by *S. aureus* 16s rRNA PCR assay. Multiplex PCRs confirmed the presence of staphylococcus enterotoxin gene C (*sec*) in one sample, staphylococcus enterotoxin gene D (*sed*) in 23 samples, while ELISA assay confirmed the presence of the same enterotoxin in only two samples. On the other hand, other groups of genes responsible for some other virulence factors of *S. aureus* like the extracellular thermostable nuclease (*nuc*) gene were found in 33 samples, while toxic shock syndrome (*tsst*) gene and methicillin restraint *S. aureus* (*mecA*) gene were not detected in this study.

**Conclusion:** Subclinical mastitis is one of the hidden factors that adversely affect the health of both animals and humans. The milk is usually appeared good and may be consumed by humans especially children; however, it causes severe public health problems. In addition, the infected animals with this form of mastitis can spread the infection to other dairy animals and may be turned to a clinical case of contagious mastitis that may be ended by animal culling or death. *S. aureus* is one of the main causes of subclinical mastitis in cattle. In addition to extracellular thermostable nuclease (*nuc*) gene, staphylococcus enterotoxin gene C (*sec*) and staphylococcus enterotoxin gene D (*sed*) are the most common virulence genes confirmed in subclinical mastitis milk. These results highlighted the need to apply more hygienic measures in the dairy farms to avoid spreading the infection between animals to ensure the production of safe and healthy food to humans.

**Keywords:** enterotoxins, *mecA*, *nuc*, polymerase chain reaction, *Staphylococcus aureus*, subclinical mastitis.

**Introduction**

Mastitis is one of the most common infectious diseases affecting dairy animals. It is considered the most economical disease of the dairy industry worldwide, including Egypt [1,2]. Mastitis, especially subclinical mastitis, is responsible for severe economic losses in the form of reduction of milk production, low-quality milk, costs of treatment, and veterinary service, besides, the increase in the culling rate [3]. Mastitis is one of the multifactorial diseases that usually resulting due to synergistic action between some risk factors that enhance the action of the mastitis pathogens. Bacteria are considered the major cause of bovine mastitis. It is responsible for both environmental mastitis caused by *Escherichia coli* and contagious mastitis caused by *Staphylococcus* spp. and *Streptococcus* spp. [4].

*Staphylococcus aureus* is the main cause of mastitis as well as it is the second identified bacteria causing food poisoning in humans through its ability...
to produce several enterotoxins in milk and milk products. These enterotoxins include SEA to SEE and SEG to SEQ. The previous studies confirmed that 25% of food poisoning outbreaks usually are caused by classical enterotoxins (SEA to SEE) [5]. According to the official European Union data from 2011, 346 foodborne outbreaks were attributed to Staphylococcus spp. This represented 6.4% of all reported outbreaks. Furthermore, S. aureus can produce other virulence factors such as exfoliative toxin A and B and toxic shock syndrome (TSST-1) [6,7]. Although, pasteurization can destroy S. aureus but cannot affect their enterotoxins which are transmitted to humans and cause a public health hazard. Intramammary and systemic administrations are common methods for the treatment of mastitis around the world. The long history of using antibiotics in the dairy farms to treat infectious diseases, especially mastitis and this un-responsible application lead to developing the resistance of bacteria to the antibiotics. Methicillin-resistant S. aureus (MRSA) has been characterized by the presence of meca A gene, which can resist the β-lactam antibiotics [8,9]. MRSA was isolated from milk samples in Egypt in two different localities, including Assiut and El-Mansoura governorates [10-12].

The present study was planned to estimate the prevalence of S. aureus and its enterotoxins’ genes in bulk milk samples from cows infected with subclinical mastitis using California mastitis test (CMT), molecular, and serological assays.

**Materials and Methods**

**Ethical approval**

Sampling and lab work were followed the ethical guidelines and principles by both Assiut University and veterinary authorities in Assiut Governorate for scientific research involving animals.

**Study area, study period and samples collection**

Milk samples were collected from different cows’ farms located in Assiut Governorate, Egypt. Assiut is the biggest governorate in Upper Egypt and it is the capital of Upper Egypt [13]. The collection of samples and samples analysis were done from April to September 2019.

A total of 60 bulk milk samples were collected from 60 cows. The samples were collected in clean, dry, and sterile 50 ml screw-capped Falcon tubes, under aseptic conditions after cleaning and disinfection of the udder. Each sample was tested individually using CMT then divided into two parts, each one stored at −20°C until DNA extraction and enzyme-linked immunosorbent assay (ELISA) analysis.

**Clinical examination**

All the cows’ udders were clinically examined before sampling according to the previously described methods for the examination of the udder and its regional lymph nodes [14].

**CMT**

Milk samples were subjected to CMT as a screening test for subclinical mastitis. An equal amount of milk and CMT solution was gently mixed for 10 s in CMT plastic plate, and then the result was recorded by a single test reader. Positive milk samples in CMT were examined for confirmation of S. aureus infection and its enterotoxins' genes by molecular assays [15].

**Polymerase chain reaction (PCR)**

**DNA extraction**

DNA was extracted from positive CMT bulk milk samples using Qiagen DNA Blood Mini kit (Cat. No. 51104, Hilden, Germany) according to manufacturer instruction, and then the extracted DNA was stored at −20°C.

**DNA amplification**

The S. aureus and its enterotoxin genes were tested using different primers for 16S rRNA, sea-see, tssst, nuc, and meca A (Table 1) [16-21]. PCR reactions were performed in final volume 25 μl including 12.5 μl of 2× PCR master mix (Green Master, Promega, USA), 150 ng of DNA template, 1 μl of each primer, and 5.5 μl of Nuclease-free water. Multiplex PCR was performed for the detection of sea-sec genes (reaction A) and sed-see genes (reaction B) in final volume 25 μl including 12 μl of Multiplex PCR master mix (Qiagen, German), 150 ng of DNA template, and 1 μl of each primer and Nuclease-free water was added until the volume was completed to 25 μl. The amplification was performed in Gradient Thermal Cycler (Veriti Applied Biosystem, USA). The thermal profile for all genes was 94°C for 5 min, followed by 35-40 cycles of denaturation at 94°C for 50 s., and the annealing temperatures were set according to each primer (s) (Table 1) followed by extension at 72°C for 1 min and the final extension at 72°C for 10 min then PCR products were preserved at 4°C.

**Gel electrophoresis**

PCRs products were electrophoresed in 1% agarose gel (GX 040.90, Gen Agarose, L.E., Standard DNA/RNA agarose, Molecular Biology Grade, InnoTrain Diagnostic, D–61476, Kronberg/Taunus) containing 10% diluted ethidium bromide 1 μl/ml electrophoresis buffer at 100 volts for 60 min. Using 100 bp DNA–ladder in Scie–PLAS, HU 10, 5636, UK. Then, the gel was visualized by the high-performance ultraviolet (UV) transilluminator (UV, INC, UK) and DOC–It ® LS, Image acquisition–software (UVP, INC, UK).

**ELISA**

The milk samples were prepared and tested as described in the manual kit of RIDASCREEN® SET (Art. No.: R4101). It is a sandwich ELISA used for the detection and the identification of staphylo-cocal enterotoxins A, B, C, D, and E in milk. The test was performed according to the manufacturer’s instructions.
Statistical analysis

The prevalence of *S. aureus* as a cause of the subclinical mastitis, its virulence enterotoxins, and their statistical significance was calculated using Chi-square test (SPSS, Chicago, USA). Any difference was considered significant at 5% threshold values [22].

Results

The udders of the sixty cows were clinically examined, and all were apparently healthy, and the related cranial and caudal supra-mammary lymph nodes were normal in size and constituent. Forty-eight (80%) milk samples were positive using CMT out of the sixty collected cow's bulk milk samples. 16S rRNA PCR confirmed the presence of *S. aureus* in 46/48 (95.83%) samples, which were CMT positive (Table-2). The PCR assays confirmed the presence of sec gene (C enterotoxin) only in one sample (2.17%), the sed gene (D enterotoxin) was found in 23 (50%) samples. In addition, the occurrence of virulence genes (*nuc*) was confirmed in 33 (71.73%) of the examined milk samples (Figure-1). The statistical analysis revealed highly significant differences between the tested enterotoxin genes and virulence genes with p=0.00001. ELISA assay confirmed the occurrence of enterotoxin D in only two samples (4.34%) (Table-3).

Discussion

Subclinical mastitis is a trigger factor for contagious mastitis in dairy farms and considered one of the most important diseases that affect dairy animals. Clinical mastitis is characterized by signs of inflammation on the udder associated with physical, chemical, cellular, and bacteriological changes in milk in addition to systemic signals as fever, anorexia, lameness, and toxemia. Contrariwise, the subclinical mastitis produces no clinical signs, but it produces cellular changes in the mastitis milk. All the examined cows showed no clinical signs of mastitis with apparently healthy udders. This finding came in agreement with the results of some previous studies which confirmed the subclinical mastitis using CMT in cows with apparently healthy udder [23].

| Table-1: Primer sets used to amplify fragments of 16S rRNA, enterotoxins, and virulence factors genes in *S. aureus*. |
| --- |
| **Primer Name** | **Sequence (5′-3′)** | **Amplified product size (bp)** | **Annealing Temperature** | **No. of cycles** | **Reference** |
| 16S rRNA | GTA GGT GGC AAG CGT TAT CC CGC ACA TCA GGC TCA G | 228 | 50°C/50 s | 40 cycles | [16] |
| sea | GCA GGG AAC AGC TTT AGG C GTT CTG TAG AAG TAT GAA ACA CG | 521 | 48°C/1 min | 44°C/1 min | 15 cycles | 20 cycles | [17] |
| seb | ACA TGT AAT TTT GAT ATT CGC ACT G TGC AGG CAT CAT GTC ATA CCA | 667 | (reaction A) | 20 cycles | [16] |
| sec | CCA ATA ATA GGA GAA AAT AAA AG ATT GGT ATT TTT TTT CGT TC | 284 | | | |
| sed | AGG TTT TTT CAC AGG TCA TCC CTT TTT TTT CCG TCA ATC | 278 | 57°C/2 min | 35 cycles | [18] |
| see | sea | 209 | | [19] |
| tsst | GCT TGC GAC AAC TGC TAC AG TGG ATC CGT CAT TCA TGG TTA T | 559 | 50°C/50 s | 40 cycles | [16] |
| nuc | GCG ATT GAT GAT GAC GTT AGC TAA ACT GGG TTA ATC | 270 | of 50°C/1 min | 40 cycles | [20] |
| meCA | AAA ACT CAT GGT AAA GGT TGG C ATC TGT ACT GGG TTA ATC | 533 | of 50°C/1 min | 40 cycles | [21] |

*S. aureus*: *Staphylococcus aureus*

| Table-2: Prevalence of subclinical mastitis in cow’s milk samples (n=60) caused by *S. aureus*. |
| --- |
| **Results** | **CMT** | **16S rRNA gene of *Staphylococcus aureus*** |
|  | **Positive/ Total** | **%** | **Positive/ Total** | **%** |
| + ve | 48/60 | 80 | 46/48 | 95.83 |
| - ve | 12/60 | 20 | 2/48 | 4.17 |

*S. aureus*: *Staphylococcus aureus*, CMT: California mastitis test

![Figure-1: Amplicons of the genes 16S rRNA, sec, sed, and nuc gene. Lane (M) DNA ladder 100 bp; lanes (1-3) positive samples of 16S rRNA gene at 228 bp; lane (4) positive sample of sec gene at 284 bp, lanes (5-7) positive samples of sed gene at 278 bp and lanes (8-10) positive samples nuc gene at 270 bp.](image-url)
the examined milk samples, 80% were positive for the subclinical mastitis through the results of CMT. This finding confirmed that subclinical mastitis is more hazardous than the clinical mastitis because it is a hidden form of the infection and the udder and milk are normal without any physical changes, only the somatic cell count (SCC) increases more than normal count which is 100,000: 200,000 cell/ml. This SCC includes both the bacterial cell responsible for mastitis like *S. aureus* and the immune cells responsible for defense. These animals act as a potential source for infection of other healthy susceptible animals. This finding came in agreement with the previous studies; however, the result in this study is slightly higher than the results found in Ismailia Province, Egypt; Musanze district, Rwanda; India and Ethiopia, where the subclinical mastitis was confirmed in 71.6%, 60%, 48.14%, and 36.67% of the examined milk samples, respectively [24-27]. *S. aureus* is one of the relevant and important major mastitis pathogens and responsible for food intoxication transmitted to humans through the consumption of contaminated milk. The current study confirmed that 95.83% of the CMT examined bovine bulk milk samples were positive for *S. aureus* subclinical mastitis. This incidence was in agreement with some previous studies in Ethiopia and Egypt, where the authors confirmed that *S. aureus* was a predominant pathogen confirmed in subclinical mastitis cases [27-29]. Moreover, the obtained result was higher than the previous finding confirmed in the previous study in Benha Province, Dakahlia Province, and West Bank-Palestine. In these studies, the authors confirmed the presence of *S. aureus* in 90.4%, 81.3%, and 68.4% of the examined milk samples, respectively [30-32]. This high incidence of *S. aureus* may be attributed to several factors: (1) Its ability to be hidden inside the udder especially with poor hygienic measures before and after milking; (2) the neglected periodical cleaning; (3) the disinfection of milking pens, milking machine, and/or milker’s hands which act as a potential vehicle that can carry the infection from animal to another; and (4) the neglected periodical milk examination using direct and indirect methods that can detect any increase in the SCC that acts as an indicator for subclinical mastitis. On the other hand, the less effective or insufficient veterinary services and irresponsible or insufficient use of antimicrobial drugs in cases of clinical mastitis may lead to the occurrence of antimicrobial-resistant strains of *S. aureus*. This leads to the formation of botryomycosis, which is one of the most serious complications and residues of *S. aureus* infection in the udder. In these cases, the bacteria become protected inside the udder tissue and resist the treatment. Furthermore, these animals will become a potential source of the infections even with no clinical signs observed and this makes *S. aureus* one of the major contagious subclinical mastitis pathogens all over the world [33]. In addition, the genes of the *S. aureus* enterotoxins were detected using two different techniques include molecular and serological assays (PCR and ELISA). The enterotoxin D was found in 50% and 4.34% of the CMT and *S. aureus* positive bulk milk samples using PCR and ELISA, respectively, while the enterotoxin C gene was found only in one sample (2.17%) by PCR and did not detect by ELISA. The result of enterotoxin D was lower than the finding obtained in Iran, where the authors confirmed the presence of enterotoxin D in 12.5% of the examined samples using ELISA. The enterotoxin C finding came in agreement with a previous study in Egypt, in which the author confirmed the enterotoxin C in only one sample and failed to identify other enterotoxins in the same sample [34,35]. Our finding came in agreement with the previous result [36] and was higher than the result reported by Abd El-Tawab et al. [37] where the authors reported *sed* gene in 36.36% of raw milk samples. In this study, the ELISA test could not identify other classical enterotoxins in subclinical mastitis milk positive for *S. aureus* and this result agreed with Inganas et al. [38], Fey et al. [39] who confirmed that sandwich ELISA is the best available method for SE detection. However, the presence of protein A produced by *S. aureus* may be interfered with its sensitivity because of Protein A, which binds with Fe portion of the antibodies (IgG), causing false-negative results. The occurrence of both SEC and SED enterotoxins in *S. aureus* isolated from milk samples confirmed that these strains responsible for subclinical mastitis are from the animal origin where these enterotoxins are dominant if compared with other classical enterotoxins [40]. Furthermore, SED is produced in foods under a wider range of pH, redox potential (E<sub>r</sub>), and water activity (a<sub>w</sub>) than other SEs, which explain why SED is principal toxins involved in staphylococcal food poisoning [41,42].

Other virulence factors present in the *S. aureus*, including extracellular thermostable nuclease (*nuc*),

### Table-3: Frequency distribution of some genes of enterotoxins and virulence factors of *S. aureus* occurred in bovine subclinical mastitic milk.

| Genes of enterotoxins and virulence factors | PCR assay | ELISA assay |
|-------------------------------------------|-----------|-------------|
| Positive/Total | % | p-value | Positive/Total | % |
| C | 1/46 | 2.17 | 0.00001 | 0/46 | 0 |
| D | 23/46 | 50 | 0.000001 | 2/46 | 4.34 |
| nuc | 33/46 | 71.73 | 0.00001 | 0/46 | 0 |

The result is significant at p<0.05. *S. aureus*: *Staphylococcus aureus*, PCR: Polymerase chain reaction, ELISA: Enzyme-linked immunosorbent assay.
toxic shock syndrome gene (tsst), and methicillin-restraint Staphylococcus (mecA) were investigated using PCR. Only the extracellular thermostable nuclease (nuc) gene was detected and confirmed in 71.73% of the examined milk samples confirmed positive for S. aureus. These findings were in fair agreement with the previous study [43] carried out in Egypt and confirmed the presence of nuc gene in all S. aureus strains, while, the (tsst) gene was confirmed in only one sample and they detected only the mecA gene in 53% of the examined samples [12]. Moreover, some phenotypically S. aureus strains were shown negative results for nuc gene, probably due to non-optimal experimental conditions for PCR method. The differences in the nucleotide sequence among the nuc genes were caused by some mutation or the absence of nuc gene in some S. aureus strains [44]. The results of this study confirmed no S. aureus MRSA strains. It concluded that S. aureus responsible for subclinical mastitis in this study mainly comes from animal origin rather than human ones according to Kitai et al. [45] who suggested that the source of food contamination by MRSA is mainly not of animal origin but from human as the food handlers and not from animal infection.

Conclusion

S. aureus is one of the most important mastitis pathogens; it is a potential pathogen in cases of subclinical mastitis and considered one of the common public health hazards. The molecular assay is recommended for confirmation of the presence of S. aureus infection and its enterotoxin, especially enterotoxin D and extracellular thermostable nuclease (nuc) genes, which are the most common enterotoxin and virulence factors detected in this study. Human consumption of subclinical mastitis milk with the above-mentioned bacteria and its enterotoxins may lead to milk-borne intoxication, especially in children and old aged people. Therefore, CMT test is recommended to use as a screening test for the periodical examination of the dairy animals for the early detection of subclinical mastitis to help in rapid and effective treatment and prevention of this disease and avoid its spreading between dairy animals. In addition, the molecular and ELISA techniques are important to detect the different virulence factors carried in S. aureus. Moreover, the application of pre- and post-milking disinfection that will ensure the quality of milk and reflected on human health is required.

Authors’ Contributions

RME and AATA designed the experiments, conducted the lab works, then drafted and edited the manuscript. Both authors read and approved the final manuscript.

Acknowledgments

We greatly indebted to the Molecular Biology Research Unit, Assiut University (Certified ISO/IEC: 17025-2005) where we carried out our work, we would like to express our hearty appreciation, sincere thanks for the staff of this unit for all the offered facilities, great help, and encouragement. This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

Competing Interests

The authors declare that they have no competing interests.

Publisher’s Note

Veterinary World remains neutral with regard to jurisdictional claims in published institutional affiliation.

References

1. Miller, R.H., Paape, M.J., Fulton, L.A. and Schutz, M.M. (1993) The relationship of somatic cell count to milk yields for Holstein heifers after first calving. J. Dairy Sci., 76(3): 728-733.
2. Seleim, R.S., Rashed, Y.M. and Fahmy, B.G.A. (2002) Mastitis pathogens attachment-related virulence features, whey protein markers and antibiotic efficacy in cows. Vet. Med. J. Giza, 50(3): 405-409.
3. Barker, A.R., Schrick, F.N., Lewis, M.J., Dowlen, H.H. and Oliver, S.P. (1998) Influence of clinical mastitis during early lactation on reproductive performance of Jersey cows. J. Dairy Sci., 81(5): 1285-1290.
4. Sayed, M. and Rady, A.A. (2008) Acute clinically mastitic animals in villages of Assiut governance: Diagnosis and treatment. Vet. World, 1(9): 261-264.
5. Letertre, C., Perelle, S., Dilasser, F. and Fach, P. (2003) Identification of a new putative enterotoxin SEU encoded by the egc cluster of Staphylococcus aureus. J. Appl. Microbiol., 95(1): 38-43.
6. European Food Safety Authority (2014) The European Union Summary Report on Trends and Source of Zoonoses, Zoonotic Agents and Food-borne Outbreaks in 2012. EFSA J., 12(2): 3547.
7. Fagundes, H. and Oliveira, C.A.F. (2004) Staphylococcus aureus intramammary infections and its implications in public health. Cienc. Rural, 34(4): 1315-1320.
8. Vishnupriya, S., Anthony, P.X., Mukhopadhyay, H.K., Pillai, R.M., Thanislass, J. and Vivek, S.V.M. (2014) Methicillin-resistant staphylococci associated with bovine mastitis and their zoonotic importance. Vet. World, 7(6): 422-427.
9. Jahan, M., Rahman, M., Parvej, S., Chowdhury, Z.H., Haque, E., Talukder, A.K. and Ahmed, S. (2015) Isolation and characterization of Staphylococcus aureus from raw cow milk in Bangladesh. J Adv. Vet. Anim. Res., 2(1): 49-55.
10. Ewida, R.M. (2009) Some Studies on Staphylococcus aureus in Milk and Some Milk Products Sold in Assiut City with Special Reference to Antibiotics Resistant Staphylococcus aureus. Ph. D Thesis. Assiut University, Faculty of Veterinary Medicine, Egypt.
11. ElShall, S.M. (2019) Enterotoxins and Enterotoxigenic Staphylococci in Milk and Some Milk Products. Ph. D. Thesis. Assiut University, Faculty Of Veterinary Medicine, Egypt.
12. Al-Ashmawy, M.A., Salam, K.I., Abd-Elghany, S.M. Elhadidy, M.M. and Tamura, T. (2016) Prevalence, molecular characterization, and antimicrobial susceptibility of methicillin-resistant Staphylococcus aureus. Foodborne Pathog. Dis., 13(3): 1-7.
13. Baines, J., Malek, J. and Speake, G. (20000) Cultural Atlas of Ancient Egypt. Checkmark Books, New York.
14. Radostitiz, O., Gay, C., Blood, D.C. and Hinchcliff, K.W. (2006) Veterinary Medicine: A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs and Goats. 10th ed. Saunders Ltd., England.
15. National Mastitis Council. (1999) Laboratory and Field Handbook on Bovine Mastitis. 2nd ed. National Mastitis Council, Madison Wisconsin, USA.
16. Monday, S.R. and Boohah, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
17. Lavseth, A., Loncarevic, S. and Berdal, K.G. (2004) Modified multiplex PCR method for detection of pyrogenic exotoxin genes in staphylococcal isolates. J. Clin. Microbiol., 42(8): 3869-3872.
18. Bayles, K.W. and Iandolo, J.J. (1989) Genetic and molecular analyses of the gene encoding staphylococcal enterotoxin D. J. Bacteriol., 171(9): 4799-4806.
19. Couch, J.L., Soltis, M.T. and Betley, M.J. (1988) Cloning and nucleotide sequence of the type E staphylococcal enterotoxin gene. J. Bacteriol., 171(7): 2976-2980.
20. Kuźma, K., Malinowski, E.H. and Kłossowska, A. (2003) Genetic and molecular studies on toxigenic Staphylococcus aureus species isolated from raw chicken meat in Japan. Bull. Vet. Inst. Pulawy, 47(1): 183-190.
21. Murakami, K., Minamide, W., Wada, K., Nakamura, E., Teraoka, H. and Watanabe, S. (1991) Identification of methicillin-resistant strains of staphylococci by polymerase chain reaction. J Clin. Microbiol., 29(10): 2240-2244.
22. Thrusfield, M. (2007) Veterinary Epidemiology. 3rd ed. Blackwell Science Publishing, New Jersey.
23. Schalm, O.W. and Noorlander, D.O. (1957) Experiments and observations leading to development of the California mastitis test. JAVMA, 130(5): 199-207.
24. Elhaig, M.M. and Selim, A. (2004) Molecular and genetic analyses of the gene encoding staphylococcal enterotoxin D. J. Bacteriol., 176(1): 391-394.
25. Reddy, B.S.S., Kumari, K.N., Reddy, Y.R., Reddy, M.V.B., Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
26. Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
27. Thrusfield, M. (2007) Veterinary Epidemiology. 3rd ed. National Mastitis Council, Madison Wisconsin, USA.
28. Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
29. Adwan, G. and Isayed, H. (2018) Prevalence and characterization of Staphylococcus aureus isolated from bulk tank milk dairy cow farms in West Bank-Palestine. Microbiol. Res. J. Int., 23(5): 1-13.
30. Jones, G.M., Bailey, T.L. and Roberson, J.R. (1998) Staphylococcus aureus mastitis: Cause, detection and control. Virginia Cooperative Extension, Virginia. p404-429.
31. Hunt, K., Butler, F. and Jordan, K. (2014) Factors affecting staphylococcal enterotoxin C bovine production in milk. Int. Dairy J., 39(1): 41-46.
32. Rajkovic, A. (2016) Staphylococcus: Food poisoning. In: Caballero, B., Finglas, P.M. and Toldra, F., editors. Encyclopedia of Food and Health. 1st ed. Academic Press, Oxford. p133-139.
33. Ahmed, A.A.H. (1980) Studies on the Enterotoxigenicity of S. aureus isolated from Milk and Milk Products. Ph. D. Thesis, Assiut University, Faculty Of Veterinary Medicine, Egypt.
34. Hunt, K., Butler, F. and Jordan, K. (2014) Factors affecting staphylococcal enterotoxin C bovine production in milk. Int. Dairy J., 39(1): 41-46.
35. Sahebnasagh, R., Saderi, H. and Owlia, P. (2014) The prevalence of resistance to methicillin in Staphylococcus aureus strains isolated from patients by PCR method for detection of mecA and nuc genes. Iran. J. Public Health, 43(1): 64-92.
36. Sadek, O.A. (2008) Human Health Risks Associated with Consumption of Milk from Subclinically Mastitic Animals in Assiut Governorate. Ph. D. Thesis. Assiut University, Faculty of Veterinary Medicine, Egypt.
37. El-Gendy, A.M. (2015) Bacteriological and Molecular Studies on Staphylococcus Species Isolated from Raw Milk. M.V.Sc. Thesis. Benha University, Egypt.
38. Roy, K., Reddy, B.S.S., Kumari, K.N., Reddy, Y.R., Reddy, M.V.B., Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
39. Radostitiz, O., Gay, C., Blood, D.C. and Hinchcliff, K.W. (2006) Veterinary Medicine: A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs and Goats. 10th ed. Saunders Ltd., England.
40. Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
41. Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
42. Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
43. Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.
44. Monday, S.R. and Bohach, G.A. (1999) Use of multiplex PCR to detect classical and newly described pyrogenic toxin genes in staphylococcal isolates. J. Clin. Microbiol., 37(10): 3411-3414.