Genomics and clinical correlates of renal cell carcinoma

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Abstract
Purpose Clear cell, papillary cell, and chromophobe renal cell carcinomas (RCCs) have now been well characterised thanks to large collaborative projects such as The Cancer Genome Atlas (TCGA). Not only has knowledge of the genomic landscape helped inform the development of new drugs, it also promises to fine tune prognostication.

Methods A literature review was performed summarising the current knowledge on the genetic basis of RCC.

Results The Von Hippel–Lindau (VHL) tumour suppressor gene undergoes bi-allelic knockout in the vast majority of clear cell RCCs. The next most prevalent aberrations include a cohort of chromatin-modifying genes with diverse roles including PBRM1, SETD2, BAP1, and KMD5C. The most common non-clear cell renal cancers have also undergone genomic profiling and are characterised by distinct genomic landscapes. Many recurrent mutations have prognostic value and show promise in aiding decisions regarding treatment stratification. Intra-tumour heterogeneity appears to hamper the clinical applicability of sparsely sampled tumours. Ways to abrogate heterogeneity will be required to optimise the genomic classification of tumours.

Conclusion The somatic mutational landscape of the more common renal cancers is well known. Correlation with outcome needs to be more comprehensively furnished, particularly for small renal masses, rarer non-clear cell renal cancers, and for all tumours undergoing targeted therapy.

Keywords Renal cancer · Genomics · Mutations · Evolution · Prognosis · Therapy

Introduction
In this review, we consider what is currently known of the genetic landscape of the commonest subtypes of renal cell cancer (RCC). A glossary has been provided to aid the understanding of specialist terminology (Table 1). Clear cell, papillary, and chromophobe cancers have now been well characterised thanks to the development of sequencing technologies (Table 2) and large collaborative projects such as The Cancer Genome Atlas (TCGA). Not only has knowledge of the genomic landscape helped inform the development of new drugs, this understanding also promises to improve risk stratification of tumours and to determine their sensitivity to systemic therapies. We shall consider each subtype in turn describing genes and pathways of oncogenesis and how these relate to prognosis and treatment response. We finish by discussing limitations of these metrics before widespread clinical applicability may be adopted.

Methods
A non-systematic literature search was conducted using Medline, updated to May 2018. The reference lists of selected manuscripts were checked manually for eligible articles. The most relevant articles summarising existing knowledge on RCC genomics, including tumour cell evolution and progression, were selected for this review. Recurrent aberrations have been defined as those with false discovery rates of < 0.1 and reported in multiple studies in the literature. For prognostic markers, those events that
Table 1  Glossary of terms used in this manuscript

| Term                        | Definition                                                                                                                                 |
|-----------------------------|------------------------------------------------------------------------------------------------------------------------------------------|
| 5′UTR                       | The 5′ untranslated region is located downstream of where transcription begins but upstream of the first protein-coding region                  |
| A:T-to-T:A transversions    | The substitution of a purine for a pyrimidine or vice versa in DNA. This changes the base from adenine (A) to thymine (T) (or vice versa)       |
| Allele                      | Different versions of the same gene are called alleles. Humans have two alleles at each genetic locus, with one allele inherited from each parent |
| Arm-level loss              | Loss of genetic material from the end of a chromosome (telomere) to the centre (centromere)                                               |
| Autosome                    | Any chromosome that is not a sex chromosome (i.e., in chromosomes 1–22 in humans)                                                        |
| Bi-allelic knockout         | Inactivation of both copies of a gene                                                                                                    |
| Chromatin                   | Chromatin is a set of molecules found in cells whose primary function is to package DNA into a more compact structure                       |
| Clonal expansion            | In cancer evolution, clonal expansion is the production of daughter cells with the same genetic makeup as the original cell               |
| Convergent evolution        | A process where independent clones evolve with similar traits, likely as a result of pressures to survive and grow within the tumour microenvironment |
| Copy number aberration      | Gain or loss of part of a chromosome                                                                                                     |
| Driver gene                 | A driver gene is one whose mutations increase the oncogenic potential of a tumour                                                        |
| Epigenetic                  | Epigenetic refers to non-genetic influences on gene expression                                                                         |
| Epithelial to mesenchymal transition | Epithelial–mesenchymal transition describes the process by which epithelial cells lose their cell polarity and cell–cell adhesion, and gain migratory and invasive properties |
| Focal deletions             | Deletion of genomic material within a chromosome spanning in general less than 5 million base pairs                                         |
| Gene fusion                 | The result of a re-arrangement between different parts of the genome that aligns two genes                                              |
| Germline mutation           | A germline mutation is one present in the germ cells, i.e., can be passed onto offspring                                                |
| GWAS                        | A genome-wide association study (GWAS), is an observational study of a genome-wide set of genetic variants in different individuals to see if any variant is associated with a trait |
| Haploinsufficiency          | Presence of only one functional copy of a gene (see mono-allelic inactivation)                                                            |
| Loci                        | Loci refers to a position within the genome                                                                                               |
| Loss of heterozygosity      | The loss of one allele of a genetic locus                                                                                                 |
| Methyltransferase           | Methyltransferases are a large group of enzymes that all methylate their substrates. In genetics, this affects gene transcription            |
| Mono-allelic inactivation   | Inactivation of one copy of a gene. We are born with two copies of all genes (aside from those on the sex chromosomes)                     |
| Mutational burden           | The total number of mutations present within a cell or tumour                                                                            |
| Mutational signature        | For the context of this paper, mutational signatures relate to the effect of mutational processes such as age or smoking on the specific types of point mutation seen in the tumour. For instance, signature 1 is found in all cancer types and is associated with age at diagnosis. Signature 2 is attributed to the AID/APOBEC family of cytidine deaminases. See https://cancer.sanger.ac.uk/cosmic/signatures for details |
| Non-synonymous mutation     | A mutation in the protein-coding part of the genome that results in a change in the resulting amino acid sequence                           |
| Point mutations             | Alteration of a single base in the genome to an alternative base                                                                          |
| Proto-oncogene              | A normal gene which, when altered by mutation, becomes an oncogene—one that can contribute to cancer                                         |
| Somatic mutation            | Mutations that are acquired during the lifetime of an individual, i.e., are not inherited                                                  |
| Splice-site variants        | A genetic alteration in the DNA sequence that occurs at the boundary of a protein-coding and non-coding region. This change can, therefore, alter the protein-coding sequence |
| Stochastically              | A random probability distribution that cannot be precisely predicted                                                                      |
| Structural variants         | Variation in the structure of a chromosome. This encompasses many changes including the abnormal joining of different chromosomal regions and copy number aberrations |
| Telomeres                   | Repetitive genetic sequence at the ends of a chromosome that protect against degradation or fusion with other chromosomes                |
| Trisomy                     | A trisomy is where there are three copies of a particular chromosome, instead of the normal two (in humans)                                  |
| Ubiquitination              | The addition of ubiquitin to a substrate protein which affects their subsequent use, interaction, localisation, or breakdown               |
are significant after multiple hypothesis testing have been included (adjusted \( p \) value < 0.1). The commonly accepted significance threshold (\( p < 5 \times 10^{-8} \)) has been used for genome-wide association studies (GWAS).

**Results**

**Clear cell renal cell carcinoma**

**Epidemiology and genetics**

The mutational landscape of clear cell RCC (ccRCC) has been defined most recently through several large-scale whole genome-sequencing studies [1–4]. These studies reveal that recurrent somatic mutations occur in only a handful of genes, with an overall mutational burden of roughly 1–2 per Mb. In addition, there are only a small number of recurrent copy number aberrations and rare gene fusions. Some insights into clinical risk factors and their genomic correlates have been made. These include patient age, with mutational burden correlating strongly with age via the predominance of the clock-like mutational signature in these genomes [5]. The higher incidence of ccRCC in male patients may partially be accounted by mono-allelic inactivation of the chromatin remodelling gene, \( KDM5C \) on the X chromosome [6]. No tobacco-specific mutational process has been detected despite the strong clinical risk factor [5]. The high frequency of A:T-to-T:A transversions consistent with mutational damage as a result of aristolochic acid exposure was detected via sequencing of patients from Eastern Europe [1] and has directly influenced primary prevention strategies.

Genetic risk factors are known to play a role in sporadic RCC development [7, 8]. Patients who have at least one first degree relative with RCC are at an increased risk of developing the disease (OR 1.4, 95% CI 0.71–2.76) [9]. The first renal large-scale GWAS in Europe revealed susceptibility loci at 2p21 and 11q13.3 [10]. The two correlated variants on 2p21 map to \( EPAS1 \), a transcription factor previously implicated in RCC, whereas the variant on 11q13.3 contains no characterised genes. An additional susceptibility locus on 12p11.23 was later discovered containing two variants in the \( ITPR2 \) gene, though direct functional evidence between \( ITPR2 \) and oncogenesis is lacking [11]. Subsequently, a locus on intron 2 of the \( ZEB2 \) gene was discovered which may play a role in decreasing regulation of epithelial to mesenchymal transition [12].

Most recently, a variant on 8q24.21 was discovered via interrogation of GWAS from an Icelandic population [13], prior to discovery of an additional seven new loci in the largest such GWAS study to date [14]. More comprehensive details of these loci are shown in Table 3.

**Somatic mutations**

**VHL** The Von Hippel–Lindau (\( VHL \)) tumour suppressor gene, located at 3p25, undergoes bi-allelic knockout in the majority of ccRCCs [15]. Haploinsufficiency of \( VHL \) occurs via arm-level loss of chromosome 3p in over 90%

### Table 1 (continued)

| Term                        | Definition                                                                 |
|-----------------------------|---------------------------------------------------------------------------|
| Warburg-like metabolic shift | This is a phenomenon, whereby cells can produce additional energy through increased oxygen-dependent glycolysis followed by lactic acid fermentation |

### Table 2 Overview of the development of sequencing technologies that have enabled the understanding of the genetic component of cancer development

| Methods                          | Explanation                                                                                                                                 |
|----------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------|
| Cytogenetics                     | These are methods used to study the structure and function of chromosomes. In cancer, they commonly refer to methods such as karyotyping, fluorescent in situ hybridisation (FISH), and comparative genomic hybridization (CGH), and give an overview of which areas of the chromosome may have been lost or gained during oncogenesis |
| Polymerase chain reaction (PCR)  | The amplification of a few copies of a short region of DNA, generating thousands to millions of copies of this sequence. This allows the detection of mutations within the amplified sequence |
| Next generation sequencing       | Next generation sequencing (NGS), massively parallel or deep sequencing are related terms that describe a DNA sequencing technology that can sequence an entire human genome within a single day. This is a catch-all term used to describe a number of different sequencing technologies such as Illumina (Solexa), Roche 454, Ion torrent or SOLiD sequencing |

DNA sequencing technologies allow us to “read” DNA. By comparing the sequence of DNA in cancer compared to normal cells, we can identify changes that might be driving the growth of cancer. Comparisons between patients with cancer and those without may also reveal the presence of inherited mutations.
of tumours. Astonishingly, this event appears to occur in a handful of cells in childhood or late adolescence, often many decades prior to diagnosis [16] (Fig. 1). The second copy of \textit{VHL} is lost, usually much later in life, by either non-synonymous mutation or epigenetic down-regulation [1–4, 16]. \textit{VHL} inactivation prevents the ubiquitination of hypoxia-inducible factor (HIF) for degradation. Upregulation of HIF is advantageous to tumour cell survival due to increased expression levels of angiogenic factors, lower rates of apoptosis, and higher rates of cellular proliferation (Fig. 2). The ubiquitous nature of upregulated HIF pathways and, therefore, neo-angiogenesis has provided the rationale for treatment with vascular endothelial growth factor (VEGF) inhibitors. Perhaps, unsurprisingly, given its ubiquitous role, no consistent relationship between \textit{VHL} status and clinical outcome has been found [17, 18].

**Chromatin-modifying genes** A cohort of chromatin-modifying genes with diverse roles including \textit{PBRM1}, \textit{SETD2}, \textit{BAP1}, and \textit{KMD5C} constitutes the next most prevalent somatic mutations. The first three of these genes are also co-located with \textit{VHL} on chromosome 3p, meaning that after 3p loss, any further non-synonymous mutation will result in complete inactivation of these haploinsufficient genes. \textit{PBRM1}, a methyltransferase is the second most commonly mutated gene in ccRCC, found in 30–50% of tumours [1–2]. \textit{PBRM1}’s inactivation could lead to loss of DNA methylation via reduction of H3K36me3 [2]. \textit{SETD2} is mutated in 10–30% of ccRCCs [1, 19–21]. \textit{SETD2}’s intracellular roles are numerous, including the regulation of transcription elongation, RNA processing, and double-stranded DNA break repair [22] that may then activate the p53-mediated checkpoint in the absence of specific \textit{p53} mutations [23]. \textit{BAP1}, a histone deubiquitinase, is mutated in up to 5–15% of ccRCCs [1–3, 24, 25]. \textit{BAP1}’s other roles include control of cellular proliferation and regulation of DNA damage repair [24].

Due to their prevalence, \textit{PBRM1}, \textit{SETD2}, and \textit{BAP1} have all been investigated as prognostic markers and for possible treatment stratification. A retrospective, validated analysis found that tumours with \textit{BAP1} mutations conferred a worse prognosis, higher grade, and worse overall survival when compared to those with \textit{PBRM1} mutations or when compared to those without \textit{BAP1} mutations [2, 24–26]. The presence of \textit{BAP1} and \textit{PBRM1} mutants appeared anti-correlated, though when co-existing, their presence conferred the worst overall survival. Similarly, the presence of \textit{SETD2} confers worse overall survival by a hazard ratio of 1.7 [26].

Genomic profiling of tumours from patients with ccRCC is beginning to illustrate how the presence of mutations in chromatin-modifying genes may aid systemic treatment stratification. For instance, in the RECORD-3 trial [27], different sequences of everolimus (an mTOR inhibitor) and sunitinib (a VEGF inhibitor) appeared to affect progression free survival in metastatic patients according to \textit{PBRM1} and \textit{BAP1} status. Immuno-oncological agents are now showing increasing promise in metastatic ccRCC settings, where \textit{PBRM1} mutations appear to confer clinical benefit after treatment with these agents [28].

**TERT** Somatic mutations have been detected within the core promoter [27, 29, 30] and 5’UTR [16] of telomerase reverse transcriptase (\textit{TERT}) in 6–14% of ccRCCs. Their functional corollary appears to include the lengthening of telomeres [16]. Furthermore, the presence of \textit{TERT} promoter mutations has been shown to decrease cancer-specific survival [29] and increased disease stage [30].

| Name          | Chromosome band | Possible gene | Putative mechanism                        | References |
|---------------|-----------------|---------------|-------------------------------------------|------------|
| rs11894252    | 2p21            | \textit{EPAS1} | Transcription factor encoding hypoxia-inducible-factor-2 alpha | [10]       |
| rs7579899     | 2p21            | \textit{EPAS1} | Transcription factor encoding hypoxia-inducible-factor-2 alpha | [10]       |
| rs7105934     | 11q13.3         | –             | –                                         | [10]       |
| rs718314      | 12p11.23        | \textit{ITPR} | Possibly through obesity related effects  | [11]       |
| rs1049380     | 12p11.23        | \textit{ITPR} | Possibly through obesity related effects  | [11]       |
| rs12105918    | 2q22.3          | \textit{ZEB2} | Regulation of the epithelial to mesenchymal transition | [12]       |
| rs35252396    | 8q24.21         | –             | Within potential regulatory region associated with other solid cancers | [13]       |
| rs4381241     | 1p32.3          | \textit{FAF1} | Facilitates increased expression of genes driving cell proliferation | [14]       |
| rs67311347    | 3p22.1          | \textit{CTNNB1} | Facilitates increased expression of genes driving cell proliferation | [14]       |
| rs10936602    | 3q26.2          |               |                                           | [14]       |
| rs2241261     | 8p21.3          | \textit{GFRA2} | Activation of the RET tyrosine kinase receptor | [14]       |
| rs11813268    | 10q24           | \textit{OBFC1} | Regulator of telomere length               | [14]       |
| rs74911261    | 11q22.3         | \textit{KDELRC2} | Functional disruption in endoplasmic reticulum | [14]       |
| rs4903064     | 14q24.2         | \textit{DPF3} | Chromatin remodelling                      | [14]       |

Summary of the inherited genetic locations believed to be associated with the development of RCC

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PTEN- and mTOR-signalling pathway The PTEN gene undergoes both recurrent point mutations (2–12% of samples) and focal deletions (approximately 7% of samples) in ccRCC [2, 3, 27]. The gene encodes a protein and phospholipid phosphatase that controls the balance between cell proliferation, apoptosis, and migration via the PI(3)K/AKT/mTOR pathway. Specific mutations may act in a dominant negative manner, implying that bi-allelic knockout is not always required to suppress function [31]. An interrogation of the TCGA data set revealed that bi-allelic loss of PTEN was uncommon but conferred worse overall survival. Mono-allelic loss was also associated with higher stage and histological grade [32]. Tumours with mutant PTEN status showed a non-significant increase in rates of progression when compared to non-PTEN mutant tumours after either VEGFR or mTOR inhibition in metastatic patients in the RECORD-3 trial [27].

The PI3K-AKT-mTOR signalling axis is directly augmented via MTOR mutations, observed in 4–9% of ccRCC neoplasms [1, 3]. Numerous other genes are involved in the mTOR pathway including PTEN, whose importance is discussed above. Other members of the pathway, such as TSC1/TSC2/PIK3CA, are infrequently mutated. Although FGFR4 undergoes copy gain as part of an arm-level gain of the long-arm of chromosome 5 in approximately 50% of ccRCC, a direct causal link between this event and PI3K-AKT has not yet been conclusively found. The PI3K-mTOR pathway is an important growth factor-signalling cascade that alters cellular metabolism. It is, therefore, an attractive target for systemic therapies via compounds collectively named rapalogs that bind to FKBP12 to inhibit the PI3K–AKT–mTOR pathway via abrogated mTORC1 kinase activity [33]. There is some evidence that tumours with mutations in MTOR/TSC1/TSC2 have a better response...
to rapalogs, although statistical significance has not been reached [27, 34, 35].

**TP53** TP53 appears relatively infrequently mutated in ccRCC (2–9% of tumours) [2, 3]. However, aberrations in genes involved in the P53 pathway are relatively common implying that the p53 pathway and cell-cycle checkpoint inhibition play significant roles in ccRCC [3, 23]. TP53 appears to be more frequently mutated in metastases [36] and on survival analysis, confers a worse cancer-specific survival than any other single mutation [37].

**Structural variants** Structural variants promote oncogenesis by altering the number of copies of genes in the genome or re-arranging the order of the genome such that either new genes are formed or the expression of a gene is altered. Aside from translocation renal cell carcinoma, gene fusions are uncommon in renal cell cancer. Large-scale copy number variations are common, however, including the almost ubiquitous heterozygous loss of the short arm of chromosome 3. The next most common aberrations are: gain of chromosome 5 (~ 60% of samples) and loss of chromosome 14 (~ 40% of samples), loss of chromosomes 6q, 8p, and 9p, and gain of chromosome 7 (~ 30% of samples) [1–4, 16, 37, 38]. Multiple authors have investigated copy number aberrations as potential biomarkers, mainly using array comparative genomic hybridisation and cytogenetic studies. Although many of these aberrations have been shown to predict prognosis [38–43], only few were repeatedly validated on multivariable analyses. Due to most copy number alterations covering large segments of the genome, it is difficult to uncover the mechanism by which the change confers oncogenic advantage. Some interesting and notable genes within these regions include CDKN2A on chromosome 9p, which has been shown to modulate VEGF expression via its interaction with HIF-1alpha, encoded by HIF1A on 14q [44].

**Immunotherapy and mutational burden**

Targeted immunotherapy, for instance, in the form of programmed death 1 (PD-1) checkpoint inhibition or cytotoxic T-lymphocyte associated antigen 4 (CTLA-4) inhibition is being increasingly used in both first- and second-line therapies [45]. Prediction of favourable response has not been correlated with PD-1 ligand biomarker expression [46]. In bladder cancer amongst others [47], mutational burden as a surrogate for neoantigen levels has been associated with enhanced response to targeted immunotherapy. Although the total mutational burden is relatively low in renal cancer, it may be a useful adjunct for response prediction to novel immunotherapeutic agents. Recently, a small study correlated total mutational burden with response to targeted immunotherapy in RCC [48]. In this study, estimated tumour mutational burden was similar in those with progressive disease and clinical benefit (10 vs. 11, \( p = 0.8 \)), as was the duration of therapy in patients with high- and low-tumour mutational burden (71 vs. 70 days, \( p = 0.39 \)). However, this study was fairly small (n = 31) and included both patients with ccRCC and non-clear cell RCC who received several different targeted immunotherapies.

| VHL mutation                  | Inducing angiogenesis | Sustaining cellular proliferation | Deregulating cellular energetics |
|-------------------------------|-----------------------|----------------------------------|---------------------------------|
| 40-70% of tumours             |                       |                                  |                                 |

| Chromatin modifying gene mutation | Disrupting chromatic regulation |
|----------------------------------|---------------------------------|
| 30-50% PBRM1                     |                                 |
| 10-30% SETD2                     |                                 |
| 5-15% BAP1                       |                                 |
| 5-15% KDM5C                      |                                 |

| TERT mutation                   | Enabling replicative immortality |
|---------------------------------|---------------------------------|
| 6-14% of tumours                |                                 |

| Chromosomal aberrations         | Activating invasion and metastasis |
|---------------------------------|-----------------------------------|
| chr14 loss ~40%                 |                                   |
| chr9 loss ~30%                  |                                   |

**Fig. 2** Mutation frequencies of the most commonly mutated genes in ccRCC and their effect as described by the Hallmarks of Cancer [64]
Heterogeneity

Intratumoural heterogeneity in ccRCC is well understood through bulk tissue DNA sequencing with branching evolution occurring more commonly and earlier than other cancer types [4, 49]. It dominates the evolutionary landscape at the genomic, transcriptomic, and proteomic levels [50]. Phylogenetic analyses led to the trunk-branch model of RCC development (Fig. 1). Somatic mutations that are found in all sampled tumour regions present in the trunk of the phylogenetic tree, including VHL mutations and 3p loss [51, 52]. This finding supports the Knudson two-hit hypothesis [53], where two ‘hits’ (i.e., bi-allelic inactivation of VHL) are required for clonal expansion to yield a clone large enough to stochastically acquire independent branches. Less prevalent mutations appear more commonly on branches with the same gene sometimes mutated on different branches in a fashion consistent with convergent evolution [4, 52]. One direct corollary of these findings is that the estimation of driver gene prevalence based on single regional sequencing significantly underestimates the true tumour-based estimation. Increased driver prevalence was seen, particularly in PBRM1, BAP1, TP53, PTEN, PIK3CA, and TSC2. Incomplete molecular profiling from single biopsies may hinder accurate prognosis and response to therapy. Extensive multi-regional sequencing appears to demarcate tumour behaviour according to evolutionary subtypes [4].

Clearly, the development of methods to infer tumour behaviour without resorting to exhaustive spatial tissue sampling is vital. Current estimates of the sampling density required to adequately represent tissue biology lie between 3 and 8 [4, 51], making these methods impractical in clinical practice. Alternatively, tumour behaviour and the Darwinian phylogeny may be predicted from other methods of molecular profiling such as transcriptomic analysis or functional imaging.

Papillary renal cell carcinoma

Papillary RCC (pRCC) represents approximately 20% of all kidney cancers and accumulates mutations at a similar rate to ccRCC [54], again with a predominance of a clock-like process. Signature 2, associated with APOBEC family of cytidine deaminases is the next most common signature. Classified as either type I or type II in roughly equal proportions, pRCC occurs either sporadically or as an inherited form [7]. In general, type I cancers are often multifocal and confer a better prognosis than the more aggressive and typically unifocal type II cancers [55]. Although classified as separate entities, types I and II pRCCs share many molecular features, including chromatin modifications seen in ccRCC [56]. Some of the shared genomic features, such as gene fusions involving TFE3 or TFEB, are present in approximately 10% of samples and show no particular disposition to type I or type II cancers [56]. The functional implication of these events remains unclear [57]. Due to the molecular overlap between both types and the fact that prognostic significance of types I vs. II was not confirmed on multivariable analyses, the clinical utility of papillary type has been questioned [55].

Type I

Hereditary papillary renal cancer (HPRC) predisposes to type I pRCC via autosomal dominant inheritance of a mutation in the MET proto-oncogene [7]. Increased MET mRNA expression is commonly observed in the sporadic form of the disease [56]. This increased expression is potentially directly driven by whole chromosomal copy number aberrations, in particular trisomy 7 which is present in the majority of type I pRCC tumours. In addition, approximately fifteen per cent of sporadic cases harbour activating mutations in the tyrosine kinase domain or contain splice-site variants [56, 58]. The MET pathway interacts with other key oncological pathways such as RAS and PIK3 causes increased angiogenesis and increased cell dissociation and is, therefore, the subject of interest for targeted inhibition. A multicentre phase II study investigated foretinib, an inhibitor of MET, VEGFR2, RON, and AXL tyrosine kinase in sporadic and HPRC-associated papillary RCC [59]. Although overall response rates were moderate, half of patients with germine MET mutations had a partial response. Unfortunately, no other pathological or molecular review was undertaken, but improved treatment stratification by type I or MET mutational status shows promise.

Additional genes recurrently mutated in type I pRCC include KDM6A, SMARCB1 and NFE2L2 [56]. Despite widening the net to discover other candidate driver mutations through known-cancer associated genes, one-third of tumours had no clearly discernible driver, other than trisomy of broad copy number alterations, most commonly chromosome 7.

Type II

The inherited form of the more aggressive, type II pRCC tumours is caused by germline mutation of the gene encoding fumarate hydratase (FH) [7]. Sporadic FH mutations are rarely found; however, mutations of genes in the downstream NRF2–antioxidant response element (ARE) pathway such as NFE2L2 are recurrently detected. CDKN2A alterations are present in 25% of type II pRCC tumours when loss of heterogeneity, promoter hypermethylation, and somatic mutations are considered together [56]. Increased expression of cell-cycle related genes was seen, most likely via retinoblastoma protein. The presence
of CDKN2A alterations was also adversely associated survival in univariate analysis of the whole cohort and when limited to the more aggressive type II phenotype [56].

A small subset of type II pRCC; CpG island methylator phenotype (CIMP) had universal hypermethylation of the CDKN2A promoter and also a high prevalence of germline or somatic mutations in FH [56]. These tumours expressed increased levels of hypoxia-related genes and evidence of a Warburg-like metabolic shift. These effects underpin the rationale to trial agents such as bevacizumab (VEGF inhibition) and erlotinib (TKI), in the phase II setting (NCT01130519).

The chromatin-modifying genes SETD2, BAP1, and PBRM1 are recurrently mutated in the absence of consistent loss of heterozygosity or promoter hypermethylation [56].

Chromophobe renal cell carcinoma

Chromophobe RCC (chRCC) accounts for 5% of renal carcinomas, but this figure is higher amongst young women. It is relatively indolent, although sarcomatoid differentiation renders it highly aggressive [55]. These tumours derived from the distal nephron accumulate mutations at a low rate (~0.4 per Mb) [54]. The most characteristic feature is extensive whole chromosomal loss of heterozygosity involving chromosomes 1, 2, 6, 10, 13, 17, and 21 [60]. The TP53 and PTEN genes were recurrently mutated almost exclusively in classic (i.e., non-eosinophilic) variants [60, 61]. Recurrent aberrations in the TERT gene were also detected with some harbouring the canonical 228T mutations, but mainly via structural variants that correlated strongly with increased TERT expression [60]. The eosinophilic subtype, describing an eosinophilic cytoplasm with densely packed mitochondria, harboured cases that were devoid of copy number aberrations and some that were enriched for the mitochondrial MT-ND5 gene mutations [60, 61]. The causal mechanism between this mutation and the histopathological phenotype has not yet been ascertained.

There are little data on genomic correlates with patient outcomes. Sun et al. analysed 66 patients from the TCGA database. TP53 mutations were found in 33% of tumours, while loss of HNF1B was seen in 88%. Prevalence of both TP53 mutations and loss of HNF1B increased with tumour stage and were linked with poor survival [62]. Casuscelli et al. [63] studied genomic outcome correlates of 79 chRCCs of all stages. TP53 mutations (58%), PTEN mutations (24%), and imbalanced chromosome duplication (duplication of ≥ 3 chromosomes, 25%) were enriched in patients with metastatic disease. While each feature was associated with inferior survival, the combination of all three changes yielded the worst prognosis.

Oncocytoma

Oncocytomas are benign tumours that share many features with eosinophilic chRCCs, including derivation from the distal tubule and recurrent mutations in mitochondria-encoded proteins [61]. Type 2 oncocytomas also contains recurrent whole chromosome losses that resemble those seen in eosinophilic chRCC [61]. The absence of PT53 mutations and activation of the p53 pathway in oncocytomas highlights p53 as a barrier to oncocytoma progression.

Conclusions

Scientific literature provides a detailed view of the genomic landscapes for each of the more common renal cancers. The genomic archaeology of clear cell tumours is particularly well characterised through exhaustive multi-regional sequencing. Through this knowledge, there is the potential to better stratify the risk of progression and survival for kidney cancer. Emerging evidence is showing that the presence or the absence of certain mutations may relate to therapeutic sensitivity or resistance. There are a number of gaps in our knowledge; these particularly relate to the behaviour of small renal masses, rarer subtypes of renal cell cancer, and the response of tumours to newer targeted agents.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no relevant conflict of interest.

Research involving human participants and/or animals The following manuscript is a review of existing data. Therefore, this article does not contain any studies with human participants or animals performed by any of the authors.

Informed consent For this type of study (review), formal consent is not required.

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