Does Phenoloxidase Contributed to the Resistance? Selection with Butane-Fipronil Enhanced its Activities from Diamondback Moths

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Abstract: Using microtitration method, the relationship between Phenoloxidase activity and the resistance of the diamondback moth Plutella xylostella (Linnaeus) to the novel insecticide butane-fipronil was determined in vitro. After selection of the tenth-generation by butane-fipronil, the resistance of the fourth instar larvae was increased 83.80-fold as compared to the susceptible strain. Phenoloxidase activity of the resistant strain (POx) was 1.29-fold higher than the susceptible one (POs). However, the Kₘ and optimum pH values were similar in resistant and susceptible strains, which were 1.11 mM and 6.5, respectively. Both POx and POs have maximum stability at pH values less than 7.0, although POx was less stable at lower pH values than POs. In addition, the thermal stabilities of the two phenoloxidases were very similar. It is suggested that PO may play an important role in the increasing resistance of pests to pesticides.

Keywords Butane-fipronil, phenoloxidase, Plutella xylostella, resistance.

INTRODUCTION

The diamondback moth, Plutella xylostella (Linnaeus), is one of the most widely distributed insects in the world and lives in more than 80 countries [1]. The indiscriminate use of insecticides coupled with the ability of P. xylostella L. to produce multiple generations per annum and the high migratory potential of the species has led to its resistance to many insecticides [2, 3]. It has been shown that the resistance occurring included alterations to insecticide target sites and powerful metabolic resistance catalyzed by some enzyme such as mixed function oxidase, esterase or glutathione-s-transferases [4]. For example, the chlorfluazuro-resistant P. xylostella has higher glutathione S-transferase-3 (GST-3) gene expression and GST activity, suggesting that GST-3 is involved in chlorfluazuron detoxification in diamondback moths [5]. While no differences in detoxication enzyme activities of permethrin-selected diamondback moths compared to non-selected populations have been found, the observed pyrethroid resistance is most likely due to decreased target site sensitivity [6].

Fipronil is the first member of the phenylpyrazole group of insecticides, and is highly toxic to both piercing–sucking and chewing insects. In addition, it has shown excellent activity against a broad spectrum of insect orders [7], with no obvious cross-resistance to other insecticides with different mechanisms of action [8]. Its resistance mechanism was demonstrated as involving low cuticle permeability, monooxygenase metabolism and increased activity of detoxification enzymes in Musca domestica, Plutella xylostella, and Chilo suppressalis [9-12]. Butene-fipronil is a novel compound obtained via the structural modification of phenylpyrazole. It has similar activity as fipronil in controlling pests on rice and vegetable crops, and has low toxicity to fish, which makes it an excellent pesticide [13].

Phenoloxidase (PO) (EC 1.14.18.1), also known as tyrosinase, is one of the key enzymes of the insect molting process, which is involved in hardening and stabilizing the newly secreted exoskeleton. Phenoloxidase catalyzes the oxidation of various phenolic compounds to the corresponding quinones, and then quinones combine with proteins to form the hard dark-colored layer of the cuticle [14]. Phenoloxidases have been proposed to participate in cuticle tanning. Arakane et al. reported that phenoloxidases extracted from Tribolium castaneum are required for larval, pupal, and adult cuticle sclerotization and pigmentation [15]. Phenoloxidase is also a key enzyme of insect immune response, and can be found in cuticular matrix [16], hemocytes [17], and hemolymph [18].

It is known that insecticide resistance is due to behavioral changes, physiological modifications, and metabolic detoxification [19]. Insect cuticle penetration is involved in physiological modifications, and some insecticides must pass through the insect cuticle in order to reach the site of action before lethal effects can occur. Some reports indicated that decreased penetration confers 2-3 fold resistance by itself [20, 21]. There have been no reports regarding phenoloxidase (PO) activity in insecticide resistance, although PO is involved in hardening and stabilizing the newly secreted exoskeleton. In fact, more attention must be paid to this enzyme when studying insecticide resistance, because PO is
one of the key enzymes of the insect molting process. In addition, PO influences the cuticle structure to allow insecticide penetration.

In the present paper, PO activity and the properties of the resistant and the susceptible diamondback moth *P. xylostella* L. are investigated, the relationship between PO and pest resistance are also discussed. The results of this study may lead to a better understanding of the mechanism of pest resistance.

MATERIALS AND METHODOLOGY

Chemicals and Instruments

L-Dihydroxyphenylalanine (L-DOPA) was purchased from Sigma (USA), and butane-fipronil (97%) was supplied by Dalian Raiser Pesticide Co. LTD (China). All other reagents were local products of analytical grade. The 96-well microtiter plate reader used was a Thermo Mustiskan Mk3 (Thermo electron corporation).

Insect

The susceptible diamomback moth strain was reared on Chinese cabbage *Brassica parachineisi* (Bailey) in a greenhouse at 25±1 °C with a 14:10 h light:dark cycle, and had never been exposed to insecticides [22]. Half of the susceptible strain population was selected for resistance to butane-fipronil for 10 generations to make the resistant strain, using dipping methods described by Ismail F. [23]. Briefly, butane-fipronil was dissolved in acetone and five serially diluted concentrations were prepared with distilled water containing 0.05% triton X-100. Cabbage leaves (4 cm × 4 cm) were cut and dipped in the serially diluted solution for 10 s. Three-day-old third instar larvae of parental generations were allowed to feed on the butane-fipronil treated leaf for 48 h, followed by a fresh leaf until it pupae.

Resistant Selection Bioassay

Technical grade butane-fipronil was dissolved in acetone and five serially experimental concentrations were prepared after protest to larvae of diamonback moth. Then fourth-instar larvae (2-3 mg/larva) were treated with 0.3 μL of the dilution on the mesothorax notum by a hand microapplicator to determine the median lethal concentration (LC50). Each test was repeated six times and acetone treatment was used as a control [24].

Preparation and Purification of PO

Fourth instar larvae (about 4 g) of the two strains were homogenized using a blender in ice-cold extraction buffer [20 mL, 0.02 M phosphate buffer solution (PBS), pH 6.5]. The homogenates were centrifuged at 6 770 g for 30 min. The supernatant was collected and brought to 40% saturation with solid ammonium sulfate. The precipitate was collected by centrifugation at 6 770 g for 30 min, redissolved in a minimum volume of the same buffer, and dialyzed against 20 mM PBS (pH 6.5) for 40 h. The partially purified enzyme was used in the present investigation.

Protein Estimation

Total protein concentration was determined by Coomassie blue G-250 dye-binding using bovine serum albumin as the standard [25].

PO Assay

Phenoloxidase activity was assayed by determining the initial linear increase in absorbance at 490 nm using a Thermo Mustiskan Mk3 microtiter plate reader at room temperature, using L-dihydroxyphenylalanine (L-DOPA) as the substrate. The reading was taken instantly after enzyme addition.

Assay of Optimal pH and Stability of PO Under Different pH Conditions

The optimum pH and the pH stability were determined according to Xue et al. [26]. Briefly, to determine the optimum pH, 140 μL of different buffers (pH range: 4.4-9.0), and 40 μL of 10 mM L-DOPA were added to a microtiter plate well. Next, 20 μL of enzyme (103.55mg/mL, as same below) was added to each reaction mixture, and the plate was read immediately to record the absorbance at 490 nm at room temperature.

To determine the pH stability of PO, 20 μL of enzyme was added to 180 μL of reaction mixture (140 μL of buffer, 40 μL of 10 mM L-DOPA) under different pH conditions to assay the PO activity (as full activity). Another 20 μL of enzyme was added to 140 μL of different buffers, and incubated for 30 min at 4 °C before 40 μL of 10 mM L-DOPA was added and the remaining PO activity was assayed immediately at room temperature. The specific value of the remaining activity to full activity was used to evaluate PO stability.

Assay of PO Thermal Stability

To test the thermal stability of PO, 20 μL of enzyme was incubated at different temperatures (25°C, 30°C, 35°C, 40°C, 50°C, 60°C, 80°C) for 5 min and 15 min, and then added to 180 μL of reaction mixture (containing 40 μL of 10 mM L-DOPA, and 140 μL of PBS, pH 6.5) to assay the PO activity immediately. The thermal stability was evaluated by the ratio of remaining PO activity.

RESULTS

Resistance Selection

The responses of the fourth instar larvae of *P. xylostella* to butene-fipronil in different generations of the susceptible and resistant strains after selection are presented in Fig. (1). The resistance of the fourth instar larvae of *Plutella xylostella* (L.) diamondback moths to butene-fipronil increased 83.80-fold in comparison with the susceptible strain until the election for the tenth generation. The resistance ratio of the parental generation increased 78.02-fold after nine generations of selection. In the experiment the eighth generation was not selected by butene-fipronil. Results showed that at the first three generations and the last three generations, the resistance increased slowly.

Determination of Kinetic Parameters of PO

Under the conditions employed in the present study, the oxidization of L-DOPA by *P. xylostella* PO followed Michaelis-Menten kinetics. The kinetic parameters for the enzyme were determined using a Lineweaver-Burk plot, with the results shown in Fig. (2). The $K_m$ and $V_{max}$ values for PO from the butene-fipronil resistant strain (POr) were 1.108 mM and 153.84 μmol/L·min·mg⁻¹ protein, respectively,
while the $K_m$ and $V_{max}$ values for the PO from the susceptible strain (POs) were 1.114 mM and 67.114 µmol/L·min$^{-1}$·mg$^{-1}$ protein, respectively.

The maximum stability of PO$_r$ and PO$_s$ was at pH 7.0 (the specific values were 0.94 and 0.88, respectively), with no obvious differences between them. As much of the PO became insoluble, the PO activity (from susceptible larvae) at pH 4.4 and pH 5.3, and the PO activity from resistant larvae at pH 4.4 were not determined.

**Effect of pH on Enzyme Activity and pH Stability**

The optimum pH for PO was determined to be about 6.5 (Fig. 3). The enzyme activity decreased rapidly when the pH was higher or lower than 6.5. The optimum pH of PO$_r$ was the same as PO$_s$. Interestingly, when PO$_s$ was added to the pH 4.4 and pH 5.3 buffers, some floccules appeared; but only in pH 4.4 buffer did PO$_s$ turn into floccules.

After the enzyme was incubated in different pH buffers, the remaining activity was assayed, and the specific value of remaining activity to full activity was calculated (Fig. 4).
Thermal Stability of PO

After the enzyme was incubated at different temperatures (25°C, 30°C, 35°C, 40°C, 50°C, 60°C, 80°C) for 5 min and 15 min, the remaining PO activity was assayed. The results showed that both PO_{r} and PO_{s} are relatively stable in the range from 25 °C to 40 °C. However, when the temperature was going greater than 40 °C the enzyme activity decreased rapidly (Fig. 5). When the temperature reached 80 °C the enzyme was completely denatured and no enzyme activity was detected (data not shown).

![Fig. 5. Stability of phenoloxidase at different temperatures. The ratio of remaining PO activity after a 15 min incubation to PO activity after a 5 min incubation was calculated.](image)

DISCUSSION AND CONCLUSION

Using microtitration, the PO activity of *P. xylostella* selected for resistance to butane-fipronil was investigated. The resistant strain was found to have significantly higher PO activity than the susceptible strain, but the difference in PO activity between the two strains could not be explained by variations in total protein content (PO_{r}:PO_{s} was 1.5:1 for a 20 μL enzyme sample). This result is similar to data obtained with Sydney rock oysters *Saccostrea glomerata* [27] in which resistance was selected for QX disease for three generations, and its PO activity was found to be higher than in the non-selected populations. In addition, a novel form of PO protein in selected oysters was detected by native PAGE. In the present paper, PO_{r} and PO_{s} had the same *K_m* values, indicating that they have the same catalysis ability but whether they are the same proteins is still unknown.

Armitage and Siva-Jothy [28] reported that *Tenebrio molitor* beetle adults with darker cuticular color have higher PO activity. Also, the black larvae of *Bombyx mori* silkworms have higher PO activities than white larvae [29]. In the present study, the insect *P. xylostella* was selected for ten generations, and the cuticles of larvae were darker than in the non-selected larvae. We deduced that the darkened cuticle has more PO content which can catalyze more phenolic compounds to quinines, and the melanin content is also increased during this time. This is beneficial for defense against the penetration of butane-fipronil or infection of microorganisms.

On the other hand, insects have an unusual innate immune system and PO is the key enzyme responsible for melanization [30]. For instance, microorganism-infected locusts can form many black nodules to eliminate invaders [31]. Undoubtedly, PO plays a major role in the innate response of insects towards infection. Some chemicals, such as the pesticide monocrotophos, can cause the PO activity to increase in the hemolymph of *Penaeus chinensis* [32]; chlorfluazuron can also increase the PO activity in the cuticle of *Ostrinia furnacalis* [33]. In our previously work [34], the physiological effect of quercetin, an inhibitor of PO, was determined in vivo on *Tenebrio molitor*, results suggested that quercetin at low concentration can cause immune response to the tested insect. The response of PO to chemicals suggests that PO may participate in the detoxification of insects by immune response. With more PO content, insects will easily survive from all kinds of invaders, including pesticides. The fact that butane-fipronil selected *P. xylostella* has higher PO activity supports this view.

So, the contribution of PO to the resistance of *P. xylostella* to butane-fipronil can be attributed to two aspects: lower cuticle penetration and higher immune ability (Fig. 6).

![Fig. 6. Two pathways of phenoloxidase contributed to *P. xylostella* resistance to butane-fipronil.](image)

Although the enzyme activity of PO_{r} was higher than PO_{s} they have similar *K_m* and optimum pH values or thermal stabilities. The only notable difference was that PO_{r} was stable under pH 5.3 conditions, while PO_{s} were unstable under the same conditions. Maybe this phenomenon is due to different protein content in the reaction mixtures.

It was be concluded that the resistance-selected diamondback moths *P. xylostella* have higher PO activity than the susceptible ones. Consequently, PO may play an impor-
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tant role in the increasing resistance of pests to pesticides, the detailed mechanism of which needs further study.

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