Minireview

Classifying the Linkage between Adipose Tissue Inflammation and Tumor Growth through Cancer-Associated Adipocytes

Yae Chan Song¹,³, Seung Eon Lee¹,³, Young Jin², Hyun Woo Park¹,*, Kyung-Hee Chun²,*, and Han-Woong Lee¹,*

¹Department of Biochemistry, College of Life Science and Biotechnology and Yonsei Laboratory Animal Research Center, Yonsei University, Seoul 03722, Korea, ²Department of Biochemistry and Molecular Biology, Yonsei University College of Medicine, Seoul 03722, Korea, ³These authors contributed equally to this work.

*Correspondence: hwl@yonsei.ac.kr (HWL); khchun@yuhs.ac (KHC); hwp003@yonsei.ac.kr (HWP)

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INTRODUCTION

To date, heterogeneous features of tumor cells have emphasized the challenges to comprehensively treat cancer. Moreover, a lethal characteristic of cancer to develop multidrug resistance has contributed to the continuous modification of current treatment strategies in cancers (Tolios et al., 2020). Thus, therapeutic targeting has shifted from tumor cell-intrinsic pathways to the bidirectional communication between the tumor and its microenvironmental compartments (Hanahan and Coussens, 2012; Hanna et al., 2009; Roma-Rodrigues et al., 2019). Investigating these interactions has manifested extensive hallmarks for changes in tumor behavior; however, identifying substantial crosstalk between various cell types within TME remains complex (Hui and Chen, 2015). The prevalence of obesity worldwide has tripled since...
## Table 1. Bidirectional communication between CAAs and cancer cells in adipocyte abundant tissues

| Tissue | Species | CAA status | Secretion | Cancer | Experiment | Reference |
|--------|---------|------------|-----------|--------|------------|-----------|
| Pancreas | Human | Dedifferentiation | FFA, matrix remodeling factors, EM induction, Angiogenic factors | Enhanced migration, Delipidation | Co-culture, RNA analysis, trans-well, Transwell analysis | (Carbone et al., 2018) |
| Pancreas | Human | Overexpression of Wnt | WNT paracrine factors, EMT inducing factors | EMT, desmoplasia | Co-culture, Wound healing assay, Invasion assay, CM treatment, gene expression profile, RNA analysis | (Bouche et al., 2013) |
| Pancreas | Mouse | Dedifferentiation | Lipolysis, FHDA | Enhanced migration, Desmoplasia | CM treatment, Adipocyte co-culture, FACS | (Okumura et al., 2017) |
| Breast | Mouse | Dedifferentiation | IL-6, IL-8, MCP1, TIMP-1, Adiponectin | Enhanced homing, Migration, Invasion | Co-culture, Protein array, H&E staining, Xenograft assay, Lipidomic analysis | (Nieman et al., 2011) |
| Breast | Human | Overexpression of Wnt | WNT paracrine factors, EMT inducing factors | EMT, desmoplasia | Co-culture, CM treatment, Gene expression profile, RNA analysis | (Carbone et al., 2018) |
| Breast | Mouse | Dedifferentiation | Lipolysis, FHDA | Enhanced migration, Desmoplasia | CM treatment, Adipocyte co-culture, FACS | (Okumura et al., 2017) |
| Breast | Human | Overexpression of Wnt | WNT paracrine factors, EMT inducing factors | EMT, desmoplasia | Co-culture, CM treatment, Gene expression profile, RNA analysis | (Carbone et al., 2018) |
| Ovary | Mouse | Dedifferentiation | Lipolysis, FHDA | Enhanced migration, Desmoplasia | CM treatment, Adipocyte co-culture, FACS | (Okumura et al., 2017) |
| Ovary | Human | Overexpression of Wnt | WNT paracrine factors, EMT inducing factors | EMT, desmoplasia | Co-culture, CM treatment, Gene expression profile, RNA analysis | (Carbone et al., 2018) |
| Ovary | Mouse | Dedifferentiation | Lipolysis, FHDA | Enhanced migration, Desmoplasia | CM treatment, Adipocyte co-culture, FACS | (Okumura et al., 2017) |
| Ovary | Human | Overexpression of Wnt | WNT paracrine factors, EMT inducing factors | EMT, desmoplasia | Co-culture, CM treatment, Gene expression profile, RNA analysis | (Carbone et al., 2018) |
| Prostate | Mouse | Dedifferentiation | CCL7, CXCR4, CXCR5, CXCR6, Leptin | Enhanced migration, Invasion | Co-culture, Orthotopic transplantation, FACS | (Laurent et al., 2016) |
| Colon | Mouse | Dedifferentiation | CCL7, CXCR4, CXCR5, CXCR6, Leptin | Enhanced migration, Invasion | Co-culture, Orthotopic transplantation, FACS | (Laurent et al., 2016) |

CAAs, cancer-associated adipocytes; FFA, free fatty acid; EMT, epithelial-mesenchymal transition; CM, conditioned media; HGF, hepatocyte growth factor; FACS, fluorescence-activated cell sorting; MMP, matrix metalloproteinase; IL, interleukin; PAI1, plasminogen activator inhibitor 1; CAF, cancer-associated fibroblast; FSP-1, fibroblast-specific protein-1; IHC, immunohistochemistry; IGFBP2, insulin growth factor-binding protein 2; ANGPTL4, angiopoietin-like 4; MIF, macrophage migration inhibitory factor; PDGF, platelet-derived growth factor; TGF-β, transforming growth factor-beta; MAPK, mitogen-activated protein kinase; PI3K, phosphoinositide 3-kinase; Akt, protein kinase B; STAT3, signaling transducer and activator of transcription 3; AMPK, 5′-AMP-activated protein kinase; CC17, chemoattractant protein 7; CXC, chemokine (C-X-C motif) receptor 4; CCR, C-C chemokine receptor; OXCR, OX-C motif chemokine ligand; CR, C-C chemokine receptor; OCR, oxygen consumption rate.
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1975, increasing the incidence of chronic inflammation, insulin resistance, type 2 diabetes mellitus (T2DM), and cardiovascular diseases (CVD). Adipose tissue (AT) inflammation and metabolic dysfunction are proposed to influence tumorigenesis and cancer progression (Lengyel et al., 2018). This interaction between AT and cancer has been well documented, with severe effects observed among adipocyte-rich cancers such as breast, colon, prostate, ovarian, and pancreatic cancers (Table 1). Thus, the profound influence of systemic inflammation on the whole body emphasizes the importance of an individual’s metabolic health during cancer treatment.

A small number of approaches have attempted to list the commonalities between growing tumors and inflammatory AT. However, numerous factors such as cytokines, extracellular matrix (ECM), immune cells, hypoxic factors, and fibroblasts affect tumor growth and AT expansion in similar manners. During proliferation, similar arrays of cytokines, such as transforming growth factor-β (TGF-β), tumor necrosis factor-α (TNF-α), monocyte chemoattractant protein-1 (MCP-1), and other pro-inflammatory cytokines, are secreted from both tumor cells and adipocytes to their surroundings, reprogramming the ECM to favor growth (Fig. 1) (Berraondo et al., 2019). The reconstruction of the TME and AT is a comparable phenomenon since both conditions pursue adipogenic and tumorigenic niche via vascularization during expansion (Hinrichs and Rosenberg, 2014; Romagnani et al., 2001). Therefore, the classification of environmental changes in the AT provides profound insights into unraveling the mechanism underlying the dynamic tumor behavior, especially when the tumor is adjacent to a large population of adipocytes. Furthermore, this comparison may suggest effective therapeutic targets for TME, which could alleviate inflammatory stress from both tumorigenic regions and AT.

OVERVIEW OF ADIPOSE TISSUE INFLAMMATION

AT is mainly composed of adipocytes that play a pivotal role as energy storage. Additionally, AT senses the nutrient status to regulate energy mobilization depending on its nutritional accessibility (Zhang et al., 2003). As a crucial endocrine organ, AT relies on adipocytes that secrete hormones through coordinating with other adjacent and distant tissues (Reilly...
and Saltiel, 2017). However, under conditions of overnutrition, AT expansion occurs due to hypertrophic adipocytes. During nutritional excess, the enlarged adipocytes cause AT dysfunction, augmenting systemic inflammation that is highly implicated in insulin resistance and T2DM (Fischer-Posovszky et al., 2011). Numerous physiological outcomes are manifested during AT inflammation: for example, increased lipolysis resulting in hydrolysis of triglyceride (TG) to three free fatty acids (FFAs) and glycerol. In turn, the metabolic health of an individual is severely impaired, and numerous adipokines are altered.

Fibrosis, a phenomenon of dysfunctional AT, is characterized by excessive deposition of ECM. Here, myofibroblasts produce ECM components, namely collagen, fibronectin, and glycoproteins. TGF-β, platelet-derived growth factor-α (PDGF-α), and HIF1-α represent the activators of profibrotic myofibroblast differentiation (Buechler et al., 2015; Marcellin et al., 2019). Excessive deposition of ECM enhances its rigidity, resulting in angiogenic properties of AT. For instance, desmoplasia occurs with a process of disrupted tissue homeostasis during inflammation regulation mediated by myofibroblasts, which deposit stiff matrix components, including fibrillar collagens and fibronectins (Quail and Dannenberg, 2019). Moreover, TGF-β and PDGF-α control ECM dynamics by regulating metalloproteinases (MMPs) and tissue inhibitor of matrix metalloproteinase-1 (TIMP-1) to mediate ECM reconstruction (Sun et al., 2013). Adjustment of ECM stiffness, rigidity, and composition is an essential procedure to accommodate hypertrophic adipocytes (Seo et al., 2015).

AT components, such as fibroblasts, preadipocytes, tissue-resident macrophages, and vascular constituents termed adipose tissue macrophages (ATMs), contribute to propagating pro-inflammatory responses under hypotherpic conditions (Huh et al., 2014; Ikeoka et al., 2010). ATMs represent the most abundant class of leukocytes, originating from blood monocytes (Russo and Lumeng, 2018). These macrophages are arranged around dead adipocytes, forming crown-like structures (CLS), which are hallmarks of the pro-inflammatory process in AT. Though the macrophage polarization status M1 (pro-inflammatory) and M2 (anti-inflammatory) could be an over-simplification, macrophages in inflammatory AT generally exhibit more M1 polarized phenotypes (Huh et al., 2014; Nguyen et al., 2007; Patsouris et al., 2008; Quail and Dannenberg, 2019).

The secretory pro-inflammatory response in AT involves TNF-α, IL-6, MCF-1, inducible nitric oxide synthase (iNOS), TGF-β, and HIF1-α (Greenberg and Obin, 2006; Hotamisligil et al., 1993). Indeed, macrophages infiltrating into AT and cancer are both responsible for local TNF-α and IL-6 secretion (Weisberg et al., 2003). This secretion mediates direct communication between adipocytes and nearby compartments, followed by induction of angiogenic factors. For instance, HIF1-α, expressed from adipocytes during AT inflammation, directly regulates the transcription of vascular endothelial growth factor (VEGF) (He et al., 2011). Reduced adiponectin secretion during obesity promotes TNF-α-induced monocyte adhesion and vascularization (Palanisamy et al., 2019). Conversely, leptin induces angiogenesis, vascular fenestration, and vascular remodeling (Buechler et al., 2015).

AN OVERVIEW OF THE TUMOR MICROENVIRONMENT

TME comprises a variety of cellular (fibroblasts, endothelial cells, pericytes, and adipocytes) and non-cellular (growth factors, cytokines, RNA, DNA, metabolites, and ECM) components exchanging a substantial network of signals. The vast networks of communication within the TME compartments include proliferative signals, resistance to apoptosis, uncontrolled multiplication, vascularization, and immune evasion (Petrova et al., 2018). Often followed by hypoxic conditions, the ECM of malignant cancers undergo dynamic alterations in its components, which are critical determinants of tumor behavior and differentiation (Walker et al., 2018).

The ECM comprises a complex mixture of macromolecules with major components providing structural support. Fibroblasts maintain the ECM homeostasis via collagen synthesis, which is implicated in the structural deposition that undergoes constant reconstruction during tumor progression (Eble and Niland, 2019; Fang et al., 2014). Fibroblasts are stromal tissue that forms a baseline for membrane compartments, regulate differentiation of epithelial cells, modulate immune responses, and mediate tissue homeostasis (Kalluri and Zeisberg, 2006; Quail and Joyce, 2013). Fibroblasts adjacent to malignant tumor cells are activated by growth factors, cell-cell communication, and reactive oxygen species (ROS), acquiring abnormal features to transform into cancer-associated fibroblasts (CAFs). Particularly, during tumor progression, TGF-β, fibroblast growth factor (FGF), and PDGF stimulate the transition from normal fibroblasts to CAFs. CAFs then produce MMPs, modulating Notch and p53 signaling pathways, releasing angiogenic factors, reprogramming ECM plasticity and architecture (Bauer et al., 2010; Quail and Joyce, 2013; Xing et al., 2010).

Fibillar collagen is one of the dominant components of ECM. Growing evidence has indicated that collagen undergoes qualitative and quantitative reorganization to provide a supportive structure for tumor progression (Sun et al., 2013; Zhou et al., 2017). Collagens directly interact with cancer cells through the discoidin domain receptor (DDR) to mediate the behavioral changes during cancer. Furthermore, cancer cells also reshape collagen to reinforce the cell-collagen loop, which gradually fosters cancer progression. Additionally, the direct binding of integrin and collagen leads to the activation of the Akt/PI3K, MAPK, Rho, and MEK/ERK signaling pathways, thereby inducing cell proliferation and evasion of apoptosis (Xu et al., 2019a).

The proliferating tumor inevitably develops a hypoxic environment that drives signals transduction via the upregulation of HIFs within the microenvironment. In the tumor, the most compelling consequence of excessive oxygen and nutrient demand is the secretion of angiogenic and lymphangiogenic factors (Ji, 2014). The correlation between the HIF pathway and pro-angiogenic genes to promote vascular permeability, endothelial cell proliferation, migration, adhesion, and tube formation is well elicited (Ros et al., 2005; Bryant et al., 2010). Additionally, hypoxia is involved in the regulation of multiple transcription factors, including Notch and Wnt signaling, to induce multiple gene expressions associated with EMT, cell cycle progression, and survival (Jung et al., 2015).
Thus, hypoxia exerts a powerful impact on the surrounding hyperplastic tumor cells.

Based on epidemiological studies, the link between obesity and cancer progression has been proposed. In addition to the role of VAT to store energy, it also acts as an endocrine organ, secreting an extensive variety of adipokines communicating with both proximal and distant organs (Paz et al., 2011). Though overnutrition is accompanied by multiple systemic effects, the co-culture system suggests the direct effects of adipokines and FFA on tumor behavior. Interestingly, it has been reported that dysfunctional AT determines tumor malignancy, entailing massive contributions of adipocytes toward cancer progression and metastasis (Johnson et al., 2012).

INVESTIGATING THE INTERRELATIONSHIP BETWEEN ADIPOSE TISSUE INFLAMMATION AND TUMOR MICROENVIRONMENT

Cancer-associated adipocytes
The association between obesity and cancer prognosis has been well established, and epidemiological studies support a strong correlation between the prevalence of obesity and cancer. However, the direct mechanism remains elusive due to the complex network of TME compartments (Quail and Dannenberg, 2019). Thus, to inspect the impact of adipocytes on proliferating tumors, several studies have conducted a co-culture system to observe the direct interplay between cancer cells and adipocytes. Though yet to be further established, adipokines and cytokines exchanged between two compartments dramatically alter the morphology of both cell types. Common phenotypes exhibited by the co-culture system included adipocytes when grown along with cancer cells originating from adipocyte-rich environments, undergoing dedifferentiation, and delipidation. Along with the down-regulation of mature adipocyte markers, these observations imply that the cancer-associated adipocytes (CAAs) supply the tumorigenic niche to nearby cancer cells followed by adipocyte death (Table 1).

Notably, CAAs from different tumor origins release important secretory factors. The chemokine ligand C-X-C motif chemokine ligand (CXCL) 12 and the receptors, C-X-C chemokine receptor (CXCR) 4 and 7, are overexpressed in tumors and implicated in cell proliferation, migration, and tumor metastasis (Zhou et al., 2019). The binding of CXCL12 to CXCR4 is now documented to activate divergent signals on multiple pathways, such as extracellular signal-regulated kinase 1/2 (ERK1/2), p38, stress-activated protein kinase (SAPK)/c-Jun NH(2)-terminal kinase (JNK), Akt, mammalian target of rapamycin (mTOR), and Bruton tyrosine kinase. CXCL12/CXCR4 flux regulates intracellular calcium flux, chemotaxis, transcription, and cell survival (Scala, 2015). These cytokines are not only secreted by malignant tumors but also secreted from adipocytes when co-cultured with certain cancers (Laurent et al., 2016).

Interleukin 6 (IL-6), another major cytokine in the TME, is frequently deregulated in cancers. Interestingly, IL-6 secretion has been implicated in obese patients and in CAAs co-cultured with breast, ovary, and prostate cancer cells, indicating the correlation between IL-6 and AT inflammation (Table 1). IL-6 overexpression can promote tumorigenesis and evasion of immune surveillance via Janus kinases (JAK) and signal transducer and activator of transcription 3 (STAT3) activation (Fisher et al., 2014; Kang et al., 2019). This mediates anti-apoptotic signals such as B-cell lymphoma 2 (Bcl-2), survivin, and myeloid cell leukemia-1 (Mcl-1), and enhances proliferation by overexpression of c-Myc, Cyclin D1, and MMP (Cho et al., 2019).

Extracellular matrix
ECM remodeling is essential for both AT inflammation and tumor growth. Similar to inflammatory AT, the TME experiences modification of ECM stiffness via CAF-mediated desmoplasia. Myofibrolasts in TME also deposit stiff matrix components, such as fibrillar collagens and fibronectins. As previously mentioned, transformation into CAFs is mediated via TGF-β, FGF, and PDGF expression from the cancerous region, stimulating angiogenesis, cell proliferation, invasion, and motility (Karagiannis et al., 2012). These outcomes are relatively common in TME with enriched ECM: for example, breast cancer becomes abundant with myofibroblasts during obesity to promote ECM stiffness (Seo et al., 2015). The transition of normal fibroblast to CAFs initiates cancer fibrosis by increasing the stiffness of the ECM in TME and AT. ECM isolated from inflammatory stromal cells promotes the invasive behavior of breast cancer cells by stimulating mechanosignal transduction, however, this could be reversed by weight loss (Seo et al., 2015). Nintedanib, a tyrosine kinase inhibitor targeting platelet-derived growth factor receptor (PDGFR) and vascular endothelial growth factor receptor (VEGFR), is a well-known chemotherapeutic drug and has been approved to treat idiopathic pulmonary fibrosis (IPF) (Rivera-Ortega et al., 2018). Similarly, drugs capable of simultaneously alleviating both inflammatory AT and TME are listed in Table 2.

Another major regulator of the ECM structure is collagen. Collagens are known to be the dominant components of ECM in both TME and AT. Specifically, collagen VI deposition plays a pivotal role in AT to induce epithelial hyperplasia (Quail and Dannenberg, 2019). During obesity, collagen VI is highly synthesized, which reflects a broad shift toward AT fibrosis (Khan et al., 2009). During inflammatory states, AT experiences robust changes in ECM, that coincide with the delivery of the pro-tumorigenic niche as they disrupt tissue homeostasis. Fibrillar collagens that reside in TME directly interact with tumor cells to modify the environment, resembling the function of collagens under inflammatory AT.

Angiogenesis
Hypoxic conditions within malignant tumor and inflammatory AT secrete HIF1-α mediated angiogenic factors into their adjacent environment. The most prominent factor involved in this condition is VEGF. Since oxygen and nutrient demands from both proliferating tumor cells and hypertrophic adipocytes are immense, they need to be highly vascularized. However, the vessel function is often dysfunctional or insufficient in both cases. In the case of inflammatory AT, CLS forming near the adipocytes are inaccessible to blood vessels. This phenomenon is exaggerated in obese patients due to vascu-

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Table 2. Anti-inflammatory drugs can be applied to suppress adipocyte-rich cancer progression

| Drug name       | Types of cancer                                                                 | Inflammatory disease | Mechanism                          | Reference/trial identifier                                                                 |
|-----------------|---------------------------------------------------------------------------------|----------------------|------------------------------------|---------------------------------------------------------------------------------------------|
| Aspirin         | Gastrointestinal cancer                                                          |                      | COX inhibitor                       | (Huang et al., 2015; Panka et al., 2017; Tougeron et al., 2014)                               |
|                 | Colorectal cancer                                                               |                      |                                    | (Bombardier et al., 1995; Broadfield et al., 2019; Hawley et al., 2012; Liang et al., 2015; Wu et al., 2019; Yuan et al., 2001) |
| Salsalate       | Prostate cancer treated with radiotherapy                                        |                      |                                    | (Huang et al., 2015; Panka et al., 2017; Tougeron et al., 2014)                               |
|                 | Colorectal cancer treated with Curcumin                                          |                      |                                    | (Bombardier et al., 1995; Broadfield et al., 2019; Hawley et al., 2012; Liang et al., 2015; Wu et al., 2019; Yuan et al., 2001) |
| Etanercept      | Breast cancer                                                                    |                      |                                    | (Aksu et al., 2011; Burge, 2003; Cane-staro et al., 2016; Madhusudan et al., 2004; Pfeifer et al., 2017; Stanley et al., 2011; Yang et al., 2017; Zhao et al., 2018) |
|                 | Bone cancer                                                                      |                      |                                    | (Aksu et al., 2011; Burge, 2003; Cane-staro et al., 2016; Madhusudan et al., 2004; Pfeifer et al., 2017; Stanley et al., 2011; Yang et al., 2017; Zhao et al., 2018) |
|                 | Non-Hodgkin’s lymphoma                                                           |                      |                                    | (Aksu et al., 2011; Burge, 2003; Cane-staro et al., 2016; Madhusudan et al., 2004; Pfeifer et al., 2017; Stanley et al., 2011; Yang et al., 2017; Zhao et al., 2018) |
|                 | Skin cancer                                                                      |                      |                                    | (Aksu et al., 2011; Burge, 2003; Cane-staro et al., 2016; Madhusudan et al., 2004; Pfeifer et al., 2017; Stanley et al., 2011; Yang et al., 2017; Zhao et al., 2018) |
| Rosiglitazone   | Ovarian cancer treated with Olaparib                                             | T2DM                 | Agonist for PPARγ nuclear receptor | (Dang et al., 2018; Lau et al., 2019; Quintanilla Rodriguez and Correa, 2020; Wang et al., 2020; Wei et al., 2019; Xu et al., 2019b) |
|                 | Breast cancer                                                                    |                      |                                    | (Dang et al., 2018; Lau et al., 2019; Quintanilla Rodriguez and Correa, 2020; Wang et al., 2020; Wei et al., 2019; Xu et al., 2019b) |
|                 | Colorectal cancer treated with 5-fluorouracil                                    |                      |                                    | (Dang et al., 2018; Lau et al., 2019; Quintanilla Rodriguez and Correa, 2020; Wang et al., 2020; Wei et al., 2019; Xu et al., 2019b) |
| Metformin       | Colorectal cancer                                                                | T2DM                 | Not specified                       | (Coyle et al., 2016; Rangarajan et al., 2018; Rena et al., 2017)                             |
|                 | Prostate cancer                                                                  | IPF                  |                                    | (Coyle et al., 2016; Rangarajan et al., 2018; Rena et al., 2017)                             |
|                 | Breast cancer                                                                    | Obesity              |                                    | (Coyle et al., 2016; Rangarajan et al., 2018; Rena et al., 2017)                             |
|                 | Urothelial cancer                                                                |                      |                                    | (Coyle et al., 2016; Rangarajan et al., 2018; Rena et al., 2017)                             |
|                 | Kidney cancer                                                                    |                      |                                    | (Coyle et al., 2016; Rangarajan et al., 2018; Rena et al., 2017)                             |
| Imatinib        | Gastrointestinal stromal tumors CML                                              |                      | PDGFR antagonist                    | (Hantel and Larson, 2018; Hasselbalch et al., 2003; Joensuu et al., 2017; Madke and Khopkar, 2011) |
| Nintedanib      | Pancreas cancer                                                                  | SSc                  | VEGFR/PDGFR antagonist              | (Distler et al., 2019; Reguera-Nunez et al., 2019; Rivera-Ortega et al., 2018; Rossi et al., 2017; Varone et al., 2018) |
|                 | Breast cancer                                                                    | IPF                  |                                    | (Distler et al., 2019; Reguera-Nunez et al., 2019; Rivera-Ortega et al., 2018; Rossi et al., 2017; Varone et al., 2018) |
|                 | Colorectal cancer                                                                |                      |                                    | (Distler et al., 2019; Reguera-Nunez et al., 2019; Rivera-Ortega et al., 2018; Rossi et al., 2017; Varone et al., 2018) |
| 17-AAG (tanespimycin) | Breast cancer                                                              | SSc                  | HIF-1α inhibitor                   | (Sontake et al., 2017; Talamantez-Lyburn et al., 2019)                                     |

ARDS, acute respiratory distress syndrome; COX, cyclooxygenase; NF-κB, nuclear factor-kappa B; AMPK, AMP-activated protein kinase; T2DM, type 2 diabetes mellitus; IPF, idiopathic pulmonary fibrosis; TNF, tumor necrosis factor; BDL, bile duct ligation; PPARγ, peroxisome proliferator-activated receptor γ; CML, chronic myelomonocytic leukemia; PDGFR, platelet-derived growth factor receptor; SSc, systemic sclerosis; VEGFR, vascular endothelial growth factor receptor.
lar control, which is unable to meet the necessary demand (Quail and Dannenberg, 2019). Obese patients also demonstrate higher IL-6 and FGF-2 expression levels, which can potentially result in resistance to anti-VEGF therapy. Reportedly, a mouse model treated with IL-6 blockade abrogated resistance to this therapy. Additionally, the downregulation of the FGF-2 expression by metformin or FGF receptor inhibition sensitized obese mice to anti-VEGF therapy (Incio et al., 2018). Thus, both the inflammatory AT and TME communicate with nearby elements to facilitate vascularization and meet demands.

**Macrophages**

Complicated networks of TME involve notable players of unique classes and subclasses of the tumor immune microenvironment (TIME), including tumor-associated macrophages (TAMs), myeloid-derived suppressor cells (MDSCs), natural killer cells (NKS), dendritic cells (DCs), and T cells (Binnews et al., 2010). TAMs are responsible to maintain the host defense and tissue homeostasis: thus, they are well characterized in both inflammatory TME and AT (Erreni et al., 2011; Thomas and Apovian, 2017). Correspondingly, TAMs are considered major components of tumor-infiltrating immune cells, providing a favorable TME for cancer to develop uncontrolled growth, neovascularization, and resistance to apoptosis (Channee et al., 2014). TAMs are characterized by the polarization status, which alternates between M1 and M2 polarized macrophages. Frequently expressing M2-like phenotypes, TAMs promote tumor progression and contribute to chemoresistance. Thus, M2 macrophage density often correlates with poor prognosis in different types of cancer patients. While M1 macrophages counteract cancer progression and metastasis via pro-inflammatory and antimicrobial phenotypes, M2 macrophages facilitate anti-inflammatory responses, tissue repair and remodeling, and immune tolerance (Channee et al., 2014). M2 can be further classified into M2a, b, c, and d according to their phenotypes, indicating the diverse functions of M2 macrophages (Yao et al., 2019). M2d macrophages are the most prevalent TAMs to drive angiogenesis, immunosuppression, and tumor progression (Channee et al., 2014). Although the macrophage polarization status is contradictory between inflammatory AT and TME, both conditions are associated with high-grade macrophage infiltration (Pang et al., 2008). In obese mice, CXCL12 enhances macrophage recruitment and insulin resistance, which could be reversed via CXCL12 blockade (Kim et al., 2014). In parallel, CXCL12/CXCR4 7 flux in the TME shapes monocyte differentiation to a distinct type of macrophages with pro-angiogenic and immunosuppressive phenotypes (Sanchez-Martin et al., 2011). MCP-1, the master regulator of monocyte/macrophage recruitment, has been implicated in multiple adipocyte-rich cancers, such as breast, prostate, ovarian, and non-small lung cancers (Table 1). Therefore, macrophages, demonstrating alternating behavior of M1 and M2 polarization, actively communicate with adjacent compartments to support the proliferative niche.

**CONCLUSION**

The worldwide prevalence of obesity has been dramatically increasing for the past decades. Both clinically and experimentally, pathological consequences of AT inflammation have been substantially demonstrated. In the presence of cancer, obese patients develop chemoresistance with high frequency, suggesting that distinct therapeutic approaches are required depending on an individual's metabolic health. Body mass index (BMI) and waist circumference are the main assessments in assigning obesity; however, this fails to fully examine the metabolic health. As AT homeostasis is critical for TME, a qualitative analysis, including a systematic histological, biochemical, and gene expression analysis, of AT function is necessary when treating cancer.

Notably, there exist commonalities between growing tumors and dysfunctional AT, indicating that both conditions undergo environmental reconstruction to favor growth (Fig. 1). Interestingly, during simultaneous growth, tumors grow more rapidly and invasively (Table 1). Both tumor cells and adipocytes secrete cytokines, promote immune cell infiltrations, and reconstruct their microenvironment, supporting the transition of tumor behavior to be more malignant. Adipocytes and tumor cells experience hypoxic conditions during proliferation, that activates HIF1α-mediated VEGF, PDGF, and FGF expression to further induce angiogenic factors. These features of cancer cells and adipocytes reconstruct ECM to favor the supply of both inflammatory and tumorigenic niche.

An intensive inflammation is a common feature among obese cancer patients. Especially those with adipocyte-rich cancers experience fatal growth of tumor fueled by chronic inflammation within the TME (Quail and Dannenberg, 2019). Hence, to counteract the TME which favors rapid progression, more attempts to reduce secretion of the inflammatory cytokines and chemokines should be made by administration of non-steroidal anti-inflammatory drugs (NSAIDs). Aspirin, for example, plays a critical role in suppressing inflammatory environment through inhibition of COX2, making it possible to be applied in cancer treatment. In addition to aspirin, we provide a potential therapeutic approach to target both inflammatory AT and TME with a single drug delivery (Table 2) (Cuzick et al., 2009). For instance, the inhibitory effect of metformin on AMPK/mTOR pathway is now implemented with numerous chemotherapeutic drugs (Sarai et al., 2019). Taking this into account, our rearrangement provides a broader application of anti-inflammatory drugs to inhibit the secretion of pro-inflammatory agents. Here, we not only suggest both direct and combined applications of anti-inflammatory drugs against tumor progression, but also emphasize the necessity to identify the patients’ metabolic health.

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