Cdk1 and SUMO Regulate Swe1 Stability

Kobi J. Simpson-Lavy, Michael Brandeis*

The Department of Genetics, The Silberman Institute of Life Sciences, The Hebrew University of Jerusalem, Jerusalem, Israel

Abstract

The Swe1/Wee1 kinase phosphorylates and inhibits Cdk1-Clb2 and is a major mitotic switch. Swe1 levels are controlled by ubiquitin mediated degradation, which is regulated by interactions with various mitotic kinases. We have recently reported that Swe1 levels are capable of sensing the progress of the cell cycle by measuring the levels of Cdk1-Clb2, Cdc5 and Hsl1. We report here a novel mechanism that regulates the levels of Swe1. We show that S. cerevisiae Swe1 is modified by Smt3/ SUMO on residue K594 in a Cdk1 dependent manner. A degradation of the swe1K594R mutant that cannot be modified by Smt3 is considerably delayed in comparison to wild type Swe1. swe1K594R cells express elevated levels of Swe1 protein and demonstrate higher levels of Swe1 activity as manifested by Cdk1-Y19 phosphorylation. Interestingly this mutant is not targeted, like wild type Swe1, to the bud neck where Swe1 degradation takes place. We show that Swe1 is SUMOylated by the Siz1 SUMO ligase, and consequently siz1Δ cells express elevated levels of Swe1 protein and activity. Finally we show that swe1K594R cells are sensitive to osmotic stress, which is in line with their compromised regulation of Swe1 degradation.

Introduction

In S. cerevisiae, Swe1 (S. pombe wee1 homologue) inhibits mitotic Cdk1-Clb2 (Cdc28-Clb2) activity by phosphorylating Y19 of Cdk1, equivalent to Y15 of Cdk1/cdc2 in S. pombe and higher eukaryotes [1]. This modification is reversed by dephosphorylation by Mih1 (S. pombe Cdc25) [2]. Swe1 does not inhibit Cdk1 when associated with its cyclins Clb5 or Clb6, moderately inhibits Cdk1-Clb3/4 and strongly inhibits Cdk1-Clb2 [3]. When Swe1 is first synthesized in late G1 it is predominantly nuclear, but after bud emergence it is additionally localized to the bud-side of the mother-bud neck in an Hsl1 kinase, Hsl7 and septin dependent manner [4]. Hsl1 and Hsl7 are also required for Cdc5 (polo kinase) bud-neck localization [5]. Prior to its destruction in late G2, Swe1 is hyperphosphorylated by Cdc4, Cdk1-Clb2 and Cdc5, all of which are present at the bud-neck [5,6,7,8,9]. Recently we have found that although Cdk1-Clb3/4 activity is essential for Swe1 destruction, the presence of Clb2 or its interaction with Swe1 is dispensable for Swe1 degradation [10].

Small Ubiquitin-related MOdifier (SUMO, Smt3, 17% identical to ubiquitin) is conjugated to its targets by a system analogous to ubiquitin. Smt3 is activated in an ATP-dependent reaction by thioester bond formation with the E1 activator Aos1/Uba2 [11], transferred to the E2 ligase Ubc9 [12] and passed to a substrate lysine, usually in the sequence ΦKxKxΦ/E, where Φ is a hydrophobic amino acid, and x is any amino acid. There are four SUMO-E3 ligases in S. cerevisiae, Siz1, Siz2 [13], Mms21 [14], and the meiotically expressed Cst9 [15]. Siz1 is responsible for the majority of vegetative growth sumoylation, with Siz2 conducting most of the remainder [13]. Despite Smt3, Aos1, Uba2 and Ubc9 being essential genes [11,12], a siz1Δsiz2Δ strain is viable [13], albeit with a clonal lethality, manifested by a nibbled phenotype which is caused by the 2μ plasmid [16]. In contrast, mms21Δ cells are not viable, though mutations in the RING finger domain that abolish its SUMO-ligase activity such as mms21Δ, mms21Δ11 or mms21Δ134 are not lethal [14,17,18], suggesting that Mms21 executes another, non-SUMO, essential function. Whereas other SUMO-E3 ligases are nuclear, Siz1 is additionally localized to the bud-neck [13,14,19,20]. Many proteins have been reported to be SUMOylated with effects being substrate dependent but including ubiquitin mediated proteolysis and re-localization. Different types of proteins are known to be SUMO substrates, many of them are involved in DNA replication stress response.

Methods

Yeast growth, synchronization and manipulation

Yeast were transformed by the frozen lithium acetate method [21] and are listed in Table S1. Plasmids used are listed in Table S2. Strains containing Cdk1<sup>as1</sup>, cdc5<sup>1753F</sup> and swe1<sup>18Δ</sup> were kind gifts from D. Kellogg [6]. Strains in W303<sup>Δ</sup> lacking hce3 or slx8 were kind gifts from X. Zhao [22]. Strains in the JD52 background lacking SUMO E3 ligases were kind gifts from E. Johnson [13,17]. Mutagenesis of plasmids to introduce K394R and K328R mutations into Swe1 was performed using the Stratagene Quikchange kit and verified by sequencing. Swe1 was tagged with 6myc using pRS306-S6M or pRS306-S6M-K394R cut with ClaI, or by using pRS405-S6M cut with SnaBI. Taggings and knockouts were confirmed by PCR. Standard Yeast-Pepitone and synthetic media (pH 5.8) supplemented with the appropriate carbon source (2%) were used throughout. Cells were grown at 30°C. S-phase arrest and release was achieved by releasing cells from G1 arrest (growth to saturation) for 1 hour, adding 0.2 M hydroxyurea (Sigma) for 2 h followed by three washes with DDW and release into media containing 5 μg/ml nocodazole (Sigma). For growth rates of cells under stress...
conditions, \( \text{OD}_{600} \) was measured before and after 9 hours incubation in YPD with 0.75 M NaCl, 7.5 mM caffeine or water. For osmolarity experiments, cells were synchronized by two doses of alpha factor (5 \( \mu \)g/ml) and released for 1 hour prior to addition of 0.5 M NaCl. For pulse chases, Swe1-3myc was expressed for one hour following release from G1 arrest and glucose added to halt transcription, or was expressed for 1 hour in \( cdkl^{+} \) cells arrested in G2 with 0.5 \( \mu \)M 1NM-PP1 after which cells were released back into the cell-cycle. Cycloheximide was not added, to allow the cell cycle to continue.

### Immunological procedures and antibodies

Cells were harvested and killed using 20% TCA, broken with glass beads and extracted into 2x sample buffer. 10% acrylamide gels were used for SDS-PAGE. Antibodies used were mouse-anti-myc 1/1000 (a kind gift from M. Goldberg), rabbit-anti-Cdc5 1/200 (Santa-Cruz), rabbit anti-Clb2 1/500 (a kind gift from A. Amon), goat-anti-smt3 1/200 (Santa-Cruz), phospho-tyrosine 15 cdk1 1/1000 (Cell Signaling Technologies) which recognizes phosphorylated tyrosine 19 of S.c.Cdk1 (pY19), rabbit anti-phospho p38 1/1000 (Cell Signaling Technologies) which

---

**Figure 1. Swe1 is SUMoylated on K594 in a Cdk1-clb dependant manner.**

A. \( \alpha \)-myc-Immunoprecipitation of Swe1-6myc cells shows Swe1 to be SUMOylated. Cells were grown for one hour before addition of 0.5 \( \mu \)M 1NM-PP1 for 30 minutes. SUMOylation is abolished when Cdk1 is inhibited.

B. Whole cell extracts probed with anti-Smt3 show a SUMO band at 110 kDa (the size at which Swe1-6myc runs at) which is absent in swe1\( ^{-} \), and Swe1\( ^{K594R} \) cells.

C. The 110 kDa Smt3 band is abolished by 0.2 M hydroxyurea. Quantification is relative to the maximum level.

D. Morphologies of cells expressing wild-type and mutated Swe1 overnight from a GAL promoter. Wild type Swe1 and Swe1\( ^{K594R} \) induced bud elongation (active) whereas Swe1\( ^{K328R} \) does not (inactive). doi:10.1371/journal.pone.0015089.g001
recognizes dually phosphorylated Hog1 (ppHog1), rabbit-anti-β-actin 1/500 (Epitomics), rabbit-anti-Aco1 1/20000 (a kind gift from O. Pines) and mouse-anti-Tubulin 1/1000 (B512, Sigma). Secondary antibodies were from Jackson Laboratories.

For immunoprecipitation, cells were released from G1 arrest for 1 hour and then 0.5 mM INM-PP1 (Toronto Research Chemicals) added for an additional 30 minutes. Cells were lysed in the presence of 10 mM NEM, yeast protease inhibitors (100×, Sigma) and yeast phosphatase inhibitor cocktails 1 and 2 (100×, Sigma). Extracts were incubated with mouse anti-c-myc (a kind gift from T. Ravid) overnight and with anti-Protein A beads for 2 hours before extraction into 2× sample buffer.

For Swe1-GFP experiments, Swe1-GFP was expressed for 90 minutes following release from G1 arrest and glucose added to halt transcription. Cells were immobilized using ConA on glass bottom tissue culture dishes (Mattek) and filmed using a Roper CCD Camera mounted on an Olympus IX70 microscope with a 60× oil objective.

**Results and Discussion**

Swe1 is SUMOylated on K594 in a Cdk1 dependent manner

Ubiquitin mediated degradation of Swe1 is one of the major modes of its cell cycle specific activity. The regulation of this degradation has been found to be exceptionally complex and dependent on a multitude of cellular inputs. Cdk1-Clb and Cdc5 activities are both required for destruction of Swe1, however interaction between Swe1 and Clb2 is dispensable [6,10]. As Cdk1 activity is not required for Cdc5 to be active [5] we considered other potential mechanisms as to how Cdk1 regulates elimination of Swe1. The SUMO E3 ligase Siz1 is a Cdk1 substrate [23] whose stability is negatively regulated by the replication-fork poison hydroxyurea and by inhibition of Cdk1 [24]. We explored the possibility that SUMO is involved in Swe1 regulation.

Immunoprecipitation of a 6myc tagged Swe1 shows Swe1 to be SUMOylated (Figure 1A left lane). Tagged Swe1 was also precipitated from extracts of cdk1Δ cells, which expresses cdk1 that is sensitive to the ATP analogue 1NM-PP1 [25]. In the presence of the inhibitor the precipitated Swe1 was not SUMOylated (Figure 1A middle lane), suggesting that sumoylation of Swe1 depends on Cdk1 activity.

Analysis of the Swe1 sequence with the SUMO site prediction program SUMOplot (Abgent) found a high probability (0.73) SUMOylation site at K594, which is within the Swe1 kinase-domain. This lysine is conserved from yeasts to man, with the SUMOylation site present in

| End | Start | Species |
|-----|-------|---------|
| 614 | ALRFIHDSCH1VHLDKLPANVMTFEGHNLKLGDFGMATHLPLEDKSFENE | 565 | Senus stricto | S. cerevisiae |
| 624 | ALRFIHESCH1VHLDKLPANVMTFEGHNLKLGDFGAHMLPLEDKSFENE | 575 | Senus stricto | S. bayanus |
| 610 | ALRFIHESCH1VHLDKLPANVMTFEGHNLKLGDFGMATHLPLEDKSFENE | 561 | Senus stricto | S. kudriavzevi |
| 618 | ALRFIHESCH1VHLDKLPANVMTFEGHNLKLGDFGMATHLPLEDKSFENE | 569 | Senus stricto | S. mikatae |
| 613 | ALRFIHESCH1VHLDKLPANVMTFEGHNLKLGDFGMATHLPLEDKSFENE | 565 | Senus stricto | S. paradoxus |
| 643 | ALRFIHDSCH1VHLDKLPANVMTFEGHNLKLGDFGMATHLPLEDKSFENE | 564 | Senus stricto | S. castellii |
| 594 | ALRFIHDCSH1VHLDKLPANVMTFEGHNLKLGDFGMATHLPLEDKSFENE | 594 | Petite negative | S. kluyveri |

The SUMO site is highlighted, identical residues in yellow and similar in green. However, mutation of G596A is predicted to abolish the SUMOylation site (Abgent SUMOplot).

doi:10.1371/journal.pone.0015089.t001

**Table 1. Alignment of Swe1-K594 using fungal alignment on the SGD.**
increased, persistent Swe1 activity to be a consequence of higher intrinsic Swe1 levels in the swe1K594R mutant cells. Indeed, swe1K594R is present at higher levels than wild type Swe1 during log-phase growth (Figure 1B) or when cells are arrested in S-phase (Figure 2A). To determine whether swe1K594R levels are linked to Swe1 activity, swe1K594R levels were examined in cells expressing cdk1Y19F, which lacks the Swe1 phosphorylation site on Cdk1. Figure 3A shows that in these mutants, in which Swe1 activity could not phosphorylate Cdk1, Swe1 levels remained higher in Swe1K594R cells but without a delay in the initiation of its destruction.

To eliminate the possibility that Swe1 is affecting Cdk1 by binding to it we tested the effect of the K594R mutation on the accumulation of swe118A, in which all 18 Cdk1 phosphorylation sites were mutated to alanine resulting in a loss of Swe1 kinase activity and lack of interaction with Cdk1-Clb2 [6]. Figure 3B shows that also in this case, swe1K594R was present at higher levels in S-phase arrested cells and again the initiation of its destruction was not delayed. Therefore, the increased stability of swe1K594R is independent of its kinase activity and its interaction with Cdk1-Clb2.

Siz1 is the SUMO E3 ligase of Swe1

Of the three SUMO E3-ligases expressed during a mitotic cell cycle, the bulk of SUMOylation is carried out by Siz1 [13]. Levels of Swe1 in cells with knocked-out or inactive SUMO E3-ligases were examined under unstressed conditions or upon arrest in S-phase. Figure 4A shows that Swe1 levels and activity were elevated in siz1D cells. In contrast, Swe1 levels were not elevated above WT levels in siz2D or mms21sp cells, even though mms21 deficient cells are hypersensitive to hydroxyurea [27] whereas siz1A cells are not sensitive [13,28] (and E Johnson, personal communication). Further knockout of SIZ2 did not significantly elevate Swe1 levels in siz1A cells. As expected deletion of the meiotic SUMO E3 ligase Cst9 also did not increase Swe1 abundance (Figure S3A). Figure 4B shows that indeed the 110 kDa SUMO band corresponding to Swe1 was absent in siz1D cells. Pulse chase of Swe1-3myc in wild-type and siz1D cells shows Swe1 to be degraded slower in siz1D cells (Figure 4C). Similarly to swe1K594R, Swe1 levels were elevated in siz1A cells when growing in mid-log phase or when arrested in S-phase. When released from hydroxyurea arrest, siz1A cells showed a considerable delay in Cdc5 accumulation and Swe1 destruction (Figure 4D and Figure S2). Although the effects of the Siz1 deletion could be through lack of SUMOylation of other proteins, the congruence of phenotypes with swe1K594R suggests Siz1 does indeed SUMOylate Swe1 on K594. Furthermore, Siz1 is the only SUMO E3-ligase that co-localizes with Swe1 at the bud-neck, suggesting that the bud-neck may serve as an organizing platform for components of the Swe1 regulatory pathway additionally to Hsl1/7, Cdc5 and Clb2. SUMOylation of Swe1 by Siz1 is thus a novel mechanism by which Cdk1-Clb regulates Swe1 stability.

A subset of SUMOylated proteins are ubiquinylated by Hex3/Slx8 or Ris1 Figures S3A and S3B) [29,30]. Cdc5 accumulation following release from S-phase arrest was delayed in hex3D or slx8D cells, probably as a consequence of the sensitivity of these strains to hydroxyurea [30] but without affecting Swe1 abundance. This observation indicates that SUMOylation is unlikely to be required for Swe1 degradation per se.

Swe1K594R cells are osmosensitive

Swe1 plays a central role in stress response and adaptation, consequently perturbations in Swe1 regulation often lead to stress
**Figure 3. Swe1^K594R abundance do not depend on its activity or interaction with Cdk1.** A. Swe1^K594R is intrinsically more abundant, even in a cdk1^-/^ background. However, the delay in Swe1 destruction is abolished. Cells were arrested for 2 hours with 0.2 M hydroxyurea and released into fresh media containing 5 μg/ml nocodazole. B. Swe1^K594R is intrinsically more abundant, even when all 18 Cdk1 phosphorylation sites present in Swe1 are mutated. Cells were arrested for 2 hours with 0.2 M hydroxyurea and released into fresh media containing 5 μg/ml nocodazole.

doi:10.1371/journal.pone.0015089.g003

**Figure 4. Siz1 is the SUMO ligase of Swe1.** A. Siz1Δ cells exhibit elevated Swe1 levels. JD52^u cells were left unsynchronized in mid-log phase (un), or arrested in S-phase with 0.2 M hydroxyurea (HU). B. The 110 kDa SUMO band (Figure 1B) is also absent in siz1Δ cells. C. Accumulation of Cdc5 and destruction of Swe1 is delayed in siz1Δ cells. Swe1-3myc was expressed from a GAL promoter in JD52^u cells for 1 hour before being shut-off by addition of glucose. D. Swe1 activity is more persistent with a delay in Cdc5 accumulation and destruction of Swe1 in siz1Δ cells. W303a cells were arrested for 2 hours with 0.2 M hydroxyurea and released into fresh media containing 5 μg/ml nocodazole.

doi:10.1371/journal.pone.0015089.g004
sensitivity. We stressed wild type and swe1\textsuperscript{K594R} cells with caffeine and with elevated levels of NaCl and determined their growth rate. Caffeine activates the cell-wall integrity (CWI) pathway and leads to dual-phosphorylation of the Slt2 MAPK [31]. Hyperosmolarity activates the Hog1 MAPK [32] pathway. Figure 5A shows that, in comparison to wild type cells, swe1\textsuperscript{K594R} cells are sensitive to osmotic stress but not to caffeine. Cells were synchronized with a-factor, released into S-phase and the phosphorylation of Cdk1-Y19 and dual-phosphorylation of Hog1 monitored upon addition of 0.5 M NaCl. Phosphorylation of Cdk1-Y19 peaked at 40 minutes following stress, but was stronger and more persistent in the swe1\textsuperscript{K594R} cells (Figure 5B and C). Hog1 was equally dually-phosphorylated after 5 minutes in both wild-type and swe1\textsuperscript{K594R} cells, but dual-phosphorylation of Hog1 attenuated more rapidly in swe1\textsuperscript{K594R} cells (Figure 5B and C). Together, this increased G2 delay and reduced Hog1 signalling duration could account for the osmo-sensitivity of swe1\textsuperscript{K594R} cells.

Why would increasing Swe1 abundance through the swe1\textsuperscript{K594R} mutation affect sensitivity to hyperosmotic stress but not to CWI stress? Whereas the only effector of hyperosmotic stress signalling upon Cdk1-Cib2 activity is through regulation of Swe1 destruction [33] CWI stress regulates Cdk1-Cib2 activity through multiple mechanisms [31] and thus the stress response is probably more buffered against increases in Swe1 abundance.

**Figure 5. Swe1\textsuperscript{K594R} cells are hypersensitive to high osmolarity but not to caffeine.**

A. Growth rates of cells under stress conditions, OD\textsubscript{600} was measured before and after 9 hours incubation in YPD with 0.75 M NaCl, 7.5 mM caffeine or water. For osmolarity experiments, cells were synchronized by two doses of alpha factor (5 \textmu g/ml) and released for 1 hour prior to addition of 0.5 M NaCl. Growth error bars are two standard deviations from three independent experiments. B and C. Swe1\textsuperscript{K594R} activity is elevated and prolonged upon exposure to 0.5 M NaCl. Cells were synchronized in late G1 with alpha-factor (5 \textmu g/ml) and released for 1 hour prior to addition of 0.5 M NaCl. The difference between the blots is the different time points.
doi:10.1371/journal.pone.0015089.g005
Conclusions
Although Cdk1-Clb activity is essential for Swe1 destruction, activation of Cdc5 [5], Cdk1-Clb2 activity or interaction with Swe1 are dispensable for Swe1 degradation [6,10]. We have found that Swe1 is SUMOylated on K594 and that this SUMOylation is prevented by inhibition of Cdk1-Clb activity or by knock out of SIZ1. Mutation of Swe1 to swe1K594R or knock out of SIZ1 results in a greater abundance of Swe1 and consequently Swe1 destruction is delayed. Siz1 abundance is negatively regulated by hydroxyurea and inhibition of Cdk1 [24]. This provides a mechanism through which Cdk1-Clb activity indirectly decreases Swe1 abundance, and may serve to limit Swe1 levels during an unperturbed cell-cycle or following recovery from stress.

Supporting Information
Figure S1 Quantification of figure 2A. (TIF)
Figure S2 Quantification of figure 4D. (TIF)
Figure S3 Cst9, ris1, hex3 and slx8 do not seem to be involved in swe1 SUMOylation. A. Deletion of csl9 or ris1 does not increase Swe1 abundance. Cells were released from G1 arrest (saturation) for 90 minutes, or synchronized in S-phase with 0.2M hydroxyurea for 90 minutes following release from saturation and then released into fresh media containing 3μg/ml nocodazole. B. Deletion of hex3 (upper) or of slx8 (lower) does not increase Swe1 abundance. W303′ a cells were arrested for 2 hours with 0.2M hydroxyurea and released into fresh media containing 5μg/ml nocodazole. (TIF)

Table S1 (DOC)
Table S2 (DOC)

Acknowledgments
Many heartfelt thanks to O. Aparicio, E. Johnson, D. Kellogg, J. Thorner, K. Lee, X. Zhao, A. Amon and Y. Wang for strains and plasmids. We also thank Y. Lavy, O. Feine, D. Zenwirth, S. Creat and J. Sajman for many stimulating discussions.

Author Contributions
Conceived and designed the experiments: KJSL. Performed the experiments: KJSL. Analyzed the data: KJSL. MB. Wrote the paper: KJSL. MB.

References
1. Booher RN, Deshaies RJ, Kirschner MW (1993) Properties of Saccharomyces cerevisiae weel and its differential regulation of p34cdc28 in response to G1 and G2 cyclins. Embo J. 12: 3417–3426.
2. Russell P, Moreno S, Reed SI (1989) Conservation of mitotic controls in fission and budding yeasts. Cell 57: 295–303.
3. Hu F, Aparicio OM (2005) Swe1 regulation and transcriptional control restrict the activity of mitotic cyclins toward replication proteins in Saccharomyces cerevisiae. Proc Natl Acad Sci U S A 102: 8910–8915.
4. Longtine MS, Thesefiel CL, McMillan JN, Weaver E, Pringle JR, et al. (2000) Septin-dependent assembly of a cell-cycle- regulatory module in Saccharomyces cerevisiae. Mol Cell Biol 20: 4048–4061.
5. Asano S, Park JE, Sakaiashi K, Yu LR, Song S, et al. (2005) Concerted regulation of Swe1/Wce1 regulation by multiple kinases in budding yeast. Embo J 24: 2194–2204.
6. Harvey SL, Charlet A, Haas W, Gygi SP, Kellogg DR (2005) Cdk1-dependent regulation of the mitotic inhibitior Swe1. Cell 122: 407–420.
7. Hood-DeGrenier JK, Boulton CN, Lvo Y (2007) Cytosolic Csl9 is required for timely inactivation of the mitotic inhibtor Swe1 and normal bud morphogenesis in Saccharomyces cerevisiae. Curr Genet 51: 1–18.
8. McMillan JN, Thesefiel CL, Harrison JC, Bardes ES, Lew DJ (2002) Determinants of Swe1p degradation in Saccharomyces cerevisiae. Mol Biol Cell 15: 3560–3575.
9. Sakaiashi K, Asano S, Yu LR, Shiwelz MJ, Park CJ, et al. (2004) Coupling morphogenesis to mitotic entry. Proc Natl Acad Sci U S A 101: 4124–4129.
10. Simpson-Lavy KJaBM (2010) Clb2 and the APC/Cdh1 regulate Swe1 stability. Cell Cycle in press.
11. Johnson ES, Schwierhorst I, Dohnen RJ, Blobel G (1997) The ubiquitin-like protein Smtp3p is activated for conjugation to other proteins by a Ubc1p/Uba2p heterodimer. Embo J 16: 5309–5319.
12. Johnson ES, Blobel G (1997) Ubc1p is the conjugating enzyme for the ubiquitin-like protein Smtp3p. J Biol Chem 272: 26799–26802.
13. Johnson ES, Gupta AA (2001) An E3-like factor that promotes SUMO conjugation to the yeast septins. Cell 106: 735–744.
14. Zhao X, Blobel G (2005) A SUMO ligase is part of a nuclear multiprotein complex that affects DNA repair and chromosomal organization. Proc Natl Acad Sci U S A 102: 4777–4782.
15. Cheng CH, Lo YH, Liang SS, Ti SC, Lin FM, et al. (2006) SUMO modifications control assembly of synaptonomal complex and polycomplex in meiosis of Saccharomyces cerevisiae: Genes Dev 20: 2067–2081.
16. Chen XL, Reindel A, Johnson ES (2005) Misregulation of 2 microm circle copy number in a SUMO pathway mutant. Mol Cell Biol 25: 4311–4320.
17. Reindel A, Belichenko I, Byblyl GR, Chen XL, Gandhi N, et al. (2006) Multiple domains in the Siz SUMO ligases contribute to substrate selectivity. J Cell Sci 119: 4749–4757.
18. Takahashi Y, Dulev S, Liu X, Miller NJ, Zhao X, et al. (2008) Cooperation of sumoylated chromosomal proteins in rDNA maintenance. PLoS Genet 4: e1000215.
19. Agarwal S, Roeder GS (2000) Zip3p provides a link between recombination enzymes and synaptonomal complex proteins. Cell 102: 245–253.
20. Takahashi T, Shimoo H, Ito K (2001) Identification of genes required for growth under ethanol stress using transposon mutagenesis in Saccharomyces cerevisiae. Mol Genet Genomics 265: 1112–1119.
21. Knop M, Siegers K, Pereira G, Zachariae W, Winsor B, et al. (1999) Epitope tagging of yeast genes using a PCR-based strategy: more tags and improved practical routines. Yeast 15: 963–972.
22. Burgess RC, Rahman S, Lisy M, Rothstein R, Zhao X (2007) The Siz3-Shh1 complex affects sumoylation of DNA repair proteins and negatively regulates recombination. Mol Cell Biol 27: 6153–6162.
23. Holt LL, Tisch BB, Villen J, Johnson AD, Gygi SP, et al. (2006) Global analysis of Cdk1 substrate phosphorylation sites provides insights into evolution. Science 325: 1682–1686.
24. Takahashi Y, Kikuchi Y (2005) Yeast PIAS-type Ubl/Siz1 is composed of a SUMO ligase and regulatory domains. J Biol Chem 280: 35822–35829.
25. Bishop AG, Ubersax JA, Petch DT, Mathews DP, Gray NS, et al. (2000) A chemical switch for inhibitor-sensitive alleles of any protein kinase. Nature 407: 395–401.
26. Liu H, Wang Y (2006) The function and regulation of budding yeast Swe1 in response to interrupted DNA synthesis. Mol Biol Cell 17: 2746–2756.
27. Brzezni D, Soller J, Liberi G, Zhao X, Maeda D, et al. (2006) Ubc9- and mms21-mediated sumoylation counteracts recombiniogic events at damaged replication forks. Cell 127: 509–522.
28. Kats ES, Enermark JM, Martinez S, Kolodner RD (2009) The Saccharomyces cerevisiae Rds1 postreplication repair and Siz1/Srs2 homologous recombination-inhibiting pathways process DNA damage that arises in asf1 mutants. Mol Cell Biol 29: 5226–5237.
29. Uzunova K, Gottsche K, Miteva M, Weisshaar SR, Glanemann C, et al. (2007) The ubiquitin-dependent proteolytic control of SUMO conjugates. J Biol Chem 282: 34167–34175.
30. Xie Y, Kreischer O, Kroetz MB, McConchie HJ, Sung P, et al. (2007) The yeast Hex3p-S88p heterodimer is a ubiquitin ligase stimulated by substrate sumoylation. J Biol Chem 282: 34176–34184.
31. Levin DE (2005) Cell wall integrity signaling in Saccharomyces cerevisiae. Microbiol Mol Biol Rev 69: 292–301.
32. O’Rourke SM, Herskowitz I, O’Shea EK (2002) Yeast go the whole HOG for the hyperosmotic response. Trends Genet 18: 405–412.
33. Chrost J, Esote X, Astron MA, Yalkov G, Gari E, et al. (2006) Phosphorylation of H11 by HOG1 leads to a G2 arrest essential for cell survival at high osmolality. Embo J 25: 2338–2346.