Review

The UPS: a promising target for breast cancer treatment
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Abstract

During the past decade, progress in endocrine therapy and the use of trastuzumab has significantly contributed to the decline in breast cancer mortality for hormone receptor-positive and ERBB2 (HER2)-positive cases, respectively. As a result of these advances, a breast cancer cluster with poor prognosis that is negative for the estrogen receptor (ESR1), the progesterone receptor (PRGR) and ERBB2 (triple negative) has come to the forefront of medical therapeutic attention. DNA microarray analyses have revealed that this cluster is phenotypically most like the basal-like breast cancer that is caused by deficiencies in the BRCA1 pathways. To gain further improvements in breast cancer survival, new types of drugs might be required, and small molecules targeting the ubiquitin proteasome system have moved into the spotlight. The success of bortezomib in the treatment of multiple myeloma has sent encouraging signals that proteasome inhibitors could be used to treat other types of cancers. In addition, ubiquitin E3s involved in ESR1, ERBB2 or BRCA1 pathways could be ideal targets for therapeutic intervention. This review summarizes the ubiquitin proteasome pathways related to these proteins and discusses the possibility of new drugs for the treatment of breast cancers.

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Protein pathway involvement in disease

The UPS in breast cancer
The ubiquitin proteasome system (UPS) consists of several crucial enzymes: a ubiquitin-activating enzyme (E1), a ubiquitin-conjugating enzyme (E2), a ubiquitin ligase (E3) and the 26S proteasome [1,2]. The E3 catalyzes the formation of polyubiquitin chains (and sometimes monoubiquitylation), utilizing ubiquitin monomers that have been activated by the E1 and E2 enzymes, and transfers them onto a specific substrate(s). Depending upon the type of ubiquitin chain, the ubiquitin modifications signal a variety of processes, including 26S proteasome-dependent degradation [3-5]. By contrast, ubiquitin modifications are negatively controlled by deubiquitylation enzymes (DUBs) [6].

Remarkably, there are ~1000 E3s, categorized into sub-families based on the structure of their catalytic site, including 300–500 Cullin-ROC/Rbx complexes, ~450 RING-type proteins, ~40 HECT-type proteins and ~20 U-box-type proteins. When comparing this with the ~500 mammalian protein kinases, it is easy to appreciate that the UPS contributes to most, if not all, cellular events. Therefore, it is realistic to anticipate major drug discoveries from this field, just like there have been in the kinase
 arena. Indeed, startling breakthroughs have been achieved recently with proteasome inhibitors.

**Estrogen receptors and the UPS**

The α subunit of the estrogen receptor (ESR1) is degraded by the UPS, and compounds inhibiting its degradation could accelerate breast cancer growth [7]. However, the mechanism underlying its proteolysis might not be straightforward as there are at least two pathways for degradation: ligand-independent and ligand-dependent. For ligand-independent degradation, unliganded ESR1 associates with a protein complex containing Hsc/Hsp70 (a protein chaperone) and STUB1 (CHIP), an E3 ligase containing a U-box. STUB1 preferentially recognizes misfolded ESR1 and targets this protein for ubiquitin-mediated proteolysis. This pathway is important for the quality control of ESR1 [8,9]. Inhibition of this pathway could increase the active ESR1 pool. Alternatively, a dominant-negative effect could be induced by accumulation of misfolded ESR1.

Ligand-dependent degradation of ESR1 is mediated by different E3 ligases and is required for estrogen-induced transactivation. In HeLa cells expressing ESR1, treatment with MG132, a proteasome inhibitor, resulted in ESR1 stabilization but impaired ESR1-mediated transcription [10]. Cyclical recruitment of E3 ligases to ESR1 and binding of ESR1 to the proteasome is necessary for transcriptional activation of estrogen-responsive promoters [11,12]. During this process, the proteasome plays a central role in the clearance of ESR1-regulated transcription complexes, and inhibition of proteasomal activity prevents cycling of ESR1 onto promoters. Putative proteins involved in this process include: i) E3 ligases UBE3A (E6AP) [13,14], MDM2 [15] and TRI25 (EFP) [16]; ii) the 20S catalytic proteasome subunit beta type-9 (PSB9; also known as LMP2) [12] and the 26S proteasome regulatory subunit 8 (PR8; also known as Rpt6/TRIP1/SUG1), which is a subunit of the 19S regulatory cap of the proteasome [11]; and iii) NCOA3 (SRC), which interacts directly with PSB9 [12] and NCOA3 (AIB1), which interacts with UBE3A [17]. UBE3A and TRI25 are preferentially recruited to estrogen-liganded ESR1, whereas MDM2 preferentially, but not exclusively, associates with unliganded ESR1 [11,16]. NCOA3 is the p160 ESR1 transcriptional coactivator and is amplified or overexpressed in breast cancer. An Ncoa3 transgenic mouse shows a high incidence of tumors in multiple organs, including breast [18,19].

**ERBB2, EGFR and UPS**

ERBB2 (HER2) and EGFR (ERBB1, HER1) are growth factor receptors (GFRs), members of the transmembrane receptor tyrosine kinase family, and are overexpressed in 25–30% and 7–20% of breast cancers, respectively [20-22]. Similar to ESR1 ligand-dependent and ligand-independent degradation, ubiquitylation is involved in the downregulation of GFRs. Ligand-dependent dimerization of EGFR increases tyrosine kinase activity and autophosphorylation of cytoplasmic tyrosine residues [23], and this enables interaction with the RING-domain-containing E3 ubiquitin-protein ligase CBL (c-CBL) in addition to downstream effectors SHC1 (SHC) and GRB2. CBL is phosphorylated by EGFR, which increases its ubiquitin ligase activity towards EGFR [24]. EGFR is then translocated to the endosomal compartment by SH3K1 (CIN85) and endophilins [25] and subsequently sorted for either recycling to the plasma membrane or destruction [26,27]. By contrast, phosphorylated ERBB2 only weakly associates with CBL and is resistant to CBL-induced downregulation [28]. In addition, ERBB2-EGFR heterodimerization blocks CBL association with EGFR and inhibits its lysosomal degradation, resulting in enhanced signaling [28-31]. Interestingly, antibody-induced ERBB2 tyrosine phosphorylation at Tyr1112 mediates interaction of the receptor with CBL, leading to receptor ubiquitylation and degradation [32]. By contrast, ligand-independent degradation of ERBB2 is mediated by STUB1 in collaboration with heatshock proteins Hsp70 and Hsp90 [33-36].

**BRCA1 and basal-like cancer**

Gene-expression profiling identified basal-like breast cancer as an exceptional cluster with poor prognosis and a unique chemosensitivity. These cancers express basal/myoepithelial cell markers such as cytokeratins 5/6, 14 and 17 or vimentin [37-39] but do not express ESR1, progesterone receptor (PRGR) or ERBB2 (triple negative) [37]. Approximately 15% of sporadic breast cancers are characterized by this phenotype [37], but this particular cluster additionally possesses the dominant characteristics of aggressive breast cancers that are insensitive to both hormone therapy and trastuzumab (Herceptin, Genentech) [38], a humanized monoclonal antibody directed against ERBB2. Therefore, treatments that specifically target this subset of breast cancers could dramatically improve the overall prognosis for breast cancers. Interestingly, 80–90% of hereditary breast cancers with mutations in the gene encoding BRCA1 display a basal-like phenotype, suggesting that a deficiency in the BRCA1 pathway might cause this specific phenotype [39-43]. Indeed, BRCA1 dysfunction in sporadic basal-like cancers has been reported [44-46], and conditional deletion of exons encoding the C-terminus of BRCA1 in the mammary gland of mice results in basal-like cancer [47]. Thus, investigating the BRCA1 pathway could be an important approach for the treatment of basal-like cancer.

BRCA1 acts as a hub protein that coordinates a diverse range of cellular pathways to maintain genomic stability [48]. Figure 1 shows some representative functions of BRCA1. Many proteins whose mutation is implicated in
familial breast cancer, such as serine-protein kinase ATM [49,50] and serine/threonine-protein kinase CHK2 [51-53], are involved in this functional network. Mutation of the central \textit{BRCA1} gene results in ~80% penetrance of breast cancer, and \textit{BRCA1} gene methylation or the accumulated dysfunctions of other proteins whose single mutation causes low penetrance are thought to result in sporadic breast cancer [48,54]. \textit{BRCA1} is a component of several different super-complexes, and, importantly, \textit{BRCA1} partners with \textit{BARD1} to form a RING heterodimer ubiquitin ligase [55,56] in most of these complexes [57]. This suggests that the \textit{BRCA1}-\textit{BARD1} complex directs the ubiquitylation of distinct substrates within each complex. \textit{BRCA1}-\textit{BARD1} catalyzes the formation of unconventional polyubiquitin chains that include Lys6-linked chains [58-60] or catalyzes the monoubiquitylation of some substrates [61-64]. The putative substrates of \textit{BRCA1} in each complex are shown in Figure 1. Histones [61,62], γ-tubulin [63] and ESR1 [64] are monoubiquitylated, whereas NPM1 [65], RPBI [66,67], RBPP8 (CtIP) [68], RPAB3 (RPB8) [69], PRGR [70] and T2EA/T2EB [71], as well as for \textit{BRCA1} auto ubiquitylation [59], it has been proposed that the ubiquitylation is not a signal for degradation. Although the biochemical mechanism regarding how the ubiquitin modifications affect intermolecular functions remains to be clarified, some biological consequences of the modifications have been reported. RBPP8 ubiquitylation depends on the phosphorylation-mediated interaction between RBPP8 and \textit{BRCA1} BRCT domains (a phosphoserine/threonine binding motif), and ubiquitylated RBPP8 associates with chromatin after DNA damage to participate in G2/M checkpoint control [68]. RPAB3 is polyubiquitylated by \textit{BRCA1} after UV irradiation, and HeLa cells expressing a ubiquitin-resistant form of RPAB3 exhibit UV hypersensitivity [69]. \textit{BRCA1}-mediated ubiquitylation of the T2EA subunit of T2EA/T2EB blocks the initiation of mRNA synthesis by preventing the association between the pre-initiation complex and both TFIIIE and the general transcription factor IIH (TFIHI) [71].

Disease models, knockouts and assays
Basal-like cancer most closely resembles features of hereditary breast cancer associated with \textit{BRCA1} mutation. It also displays frequent mutations in the \textit{TP53} gene (encoding cellular tumor antigen P53). Based on this correlation, conditional mouse mutants with somatic deletion of \textit{Brca1} and \textit{Tp53} in mammary epithelium have been generated. Female mice of this strain show a high incidence of mammary tumors. Furthermore, the phenotype of these tumors mimics many aspects of human basal-like breast cancer [73]. More specifically, conditional mouse mutants with a deletion of the C-terminus of \textit{Brca1} and \textit{Tp53} blocks the initiation of mRNA synthesis by preventing the association between the pre-initiation complex and both TFIIIE and the general transcription factor IIH (TFIHI) [71].

It has recently been established that chimeric proteins containing the tyrosine kinase and RING domain of CBL and substituted Src-homology 2 domains from GRB2, from the P85 regulatory subunits of phosphoinositide 3-kinase (PI3K) or from SRC were capable of mediating ubiquitin-dependent proteolysis of ERBB2 (HER2) [74]. Although no physiological E3 ligases for the estrogen receptor (ESR1) and ERBB2 ubiquitylation have yet been identified, this model could prove useful for understanding ERBB2-positive cancer. Despite the fact that there are no mouse models of ERBB2 and ESR1 function in breast
cancer, there is good potential for novel models to be generated for dissecting the roles of these two proteins.

**Disease targets and ligands**

**The UPS and proteasome inhibitors**

The first proteasome inhibitor that has come into clinical practice is bortezomib (Velcade, PS-341, Millennium) [75]. Approximately a third of relapsed, refractory multiple myeloma patients show a significant response to bortezomib [76]. The US FDA approved bortezomib for use as a therapy for multiple myeloma in 2003. Bortezomib inhibits proteasome function in a slowly reversible manner by means of an interaction between boronic acid at the C-terminus of bortezomib and an active threonine in the chymotryptic catalytic site of the 20S proteasome [77]. The mechanisms underlying the therapeutic effect of bortezomib in multiple myeloma have been investigated intensively. Inhibition of the transcription factor NFκB by blocking the degradation of its inhibitory partner IκB is one such putative model. However, recent studies suggest that multiple factors might contribute to the efficacy of the drug [78]. An interesting recent study showed that inhibition of the 26S proteasome by MG132 causes a depletion of available nuclear ubiquitin because of the accumulation of nondegraded polyubiquitylated proteins in the cytosol. The depletion of free nuclear ubiquitin resulted in a loss of monoubiquitylated histones, and consequently this might have impaired many nuclear regulatory systems, including the DNA-damage responses [79,80].

Clinical trials of bortezomib in many hematologic (e.g. a phase II trial on cutaneous T-cell lymphoma by Jonsson Comprehensive Cancer Center, NCI00182637) and non-hematologic malignancies (e.g. phase II trials on advanced bronchiolo-aveolar carcinoma [BAC] or adenocarcinoma by the California Cancer Consortium, NCT00118144, and on pleural mesothelioma by the Irish Clinical Oncology Research Group, NCT00513877) are ongoing. Thus far, bortezomib has failed to show a significant clinical effect on breast cancer. Although bortezomib was well tolerated, no responses were observed in 12 patients with aggressive metastatic breast cancers, with extremely poor prognoses, when used as a single agent [81]. However, the effect of combination therapy and the therapeutic effect for selected patients, such as those with tumors expressing a particular hormone receptor, ERBB2 (HER2) status or those in earlier stages of breast cancer, remain to be determined.

There are increasing numbers of small molecules that target the UPS. The UPS Patent Portfolio table and UPS Drugs & Biologicals table on the Targeted Proteins database show the patents and drugs in development or on the market (and the associated organizations and companies) designed to inhibit the UPS that might be useful in breast cancer therapy. An orally active proteasome inhibitor salinosporamide A (NPI0052, Nereus), a natural product derived from a marine actinomycete, resembles lactacystin and irreversibly targets the proteasome [82,83,72]. Epoxomicin and eponemycin are epoxyketone-containing natural products that exhibit antitumor activity [84,85]. Epoxomicin, currently the most specific proteasome inhibitor, reacts irreversibly with the chymotrypsin-like site, whereas the less-potent epoxyketone eponemycin reacts with both the caspase-like and chymotrypsin-like sites of the proteasome [86-88]. PR171 (carfilzomib, Proteolix), a synthetic derivative of epoxomicin, has been shown recently to have antiproliferative and proapoptotic effects on primary human acute myeloid leukemia cells [89].

**Estrogen receptors and the UPS**

Many anti-breast cancer drugs are involved in degradation of the estrogen receptor (ESR1) [90]. The pure antagonist fulvestrant (ICI 182780, AstraZeneca) and RU58668 promote degradation of ESR1, whereas 4-hydroxy-tamoxifen does not [91]. Fulvestrant removes ESR1 from cyclic recruitment to its target promoters by direct targeting of ESR1 to the proteasome without associated transcription [11]. NCOA3 (AIB1) also induces agonist-induced, but not antagonist-induced, degradation of ESR1. Therefore, the suppression of NCOA3 that results in inhibition of estrogen-induced degradation of ESR1, but not degradation induced by fulvestrant or GW5638 [17], could have additional clinical effects for ESR1-positive breast cancer. Other compounds that induce degradation of ESR1 include 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD), a ligand of the aryl hydrocarbon receptor (AHR) [92], and PPARγ agonists ciglitazone and 15-deoxy-delta 12,14-prostaglandin J2 [93].

Although these existing drugs have not been designed to modify the UPS, it is possible that they function primarily through UPS modulation. In this regard, revealing the precise mechanisms of degradation of ESR1 with respect to the UPS might further improve the effectiveness of such compounds for the treatment of breast cancer. Compounds that affect the interactions of E3 ligases, coactivators and proteasome subunits in ESR1-regulated complexes on promoters could work as anti-breast cancer agents.

**ERBB2, EGFR and the UPS**

As mentioned previously, ligand-independent degradation of ERBB2 is mediated by STUB1 (CHIP) in collaboration with Hsp70 and Hsp90 [33-36]. Therefore, acceleration of this pathway might have additive antitumor activity when introduced with trastuzumab. The potent anticancer agent geldanamycin, a benzoquinone
ansamycin that binds to Hsp90, is one such candidate [33,34]. In addition, the STUB1-dependent degradation pathway of ERBB2 can be stimulated by tyrosine kinase inhibitors such as CI1033 (Pfizer) [35]. CI1033 and geldanamycin additively inhibit tumor cell growth.

Thus, downregulation of ERBB2 by means of acceleration of the UPS is of crucial importance to breast cancer treatment. Interestingly, however, the proteasome inhibitor bortezomib has an additive or synergistic effect with trastuzumab in the induction of apoptosis in ERBB2-positive breast cancer cell lines [94]. It is likely that inhibition of other UPS pathways contributes to this effect. Alternatively, it could be caused by a depletion of available nuclear ubiquitin due to the accumulation of nondegraded polyubiquitinated proteins, as mentioned previously [79,80].

Targets downstream of ERBB2 could also be affected by the UPS. BIRC5 (also known as survivin), an anti-apoptotic protein, is expressed only in tumors. Expression of BIRC5 is associated with a poor prognosis in a variety of malignancies, including breast cancer [95,96]. Recent studies revealed that BIRC5 is regulated by ERBB2 and ERBB3 but not by EGFR [97]. Interrupting the ERBB2-ERBB3 heterodimer using lapatinib (GW572016, Glaxo-SmithKline), a potent small-molecule inhibitor of EGFR and ERBB2 tyrosine kinases, leads to proteasomal degradation of BIRC5 and induces apoptosis both in breast cancer cell lines overexpressing ERBB2 and in primary tumors [97]. Understanding the mechanisms that protect ERBB2-overexpressing cancer cells from apoptosis might result in new targets for therapeutic intervention.

New frontiers in drug discovery

The therapeutic effect of proteasome inhibitors on breast cancer remains to be determined but is greatly anticipated. Additionally, interest is focused on the discovery of other potent anticancer drugs that affect substrate ubiquitylation by E3 ligases as well as their deubiquitylation catalyzed by DUB enzymes. Because inhibition of the proteasome, whose activity is crucial for all cell types, imparts such a specific clinical effect, inhibiting the catalytic site of E3 ligases that regulate a broad range of cellular processes, for example the RBX1 (ROC1) RING finger subunit of Cullin-based E3 complexes [98,99], could be equally valuable. Alternatively, E3 ligases found to be oncogenes or tumor suppressor genes that regulate more restricted pathways could be promising targets for small-molecule inhibitors and activators, respectively, with fewer side effects [100]. The following sections describe such specific pathways that link directly to the breast cancer clinic.

Estrogen receptors and the UPS

Progress in hormone therapy, including aromatase inhibitors and the gonadotrophin-releasing hormone agonist goserelin, has been a major contribution to the recent improvements in breast cancer prognosis. As there already exist several effective hormone therapy agents, it might be conjectured that discovering additional drugs for the same pathway is not necessary. However, approximately half of hormone receptor-positive breast cancers do not respond sufficiently to the current hormone therapies [101], and therefore alternative drugs free from cross-tolerance are needed. Compounds targeting the UPS in hormone receptor signaling could be one such alternative.

ERBB2, EGFR and UPS

Trastuzumab prolongs the survival of patients with metastatic ERBB2 (HER2)-positive breast cancer and leads to dramatic improvements in prognosis when used in adjuvant therapy [102]. Indeed, trastuzumab has switched ERBB2-positive breast cancer from being a subset with the worst prognosis to one that is curable with existing therapy [102]. However, even in a subset of patients with highly ERBB2-positive tumors, the response rate to trastuzumab is still limited, and alternative compounds that target the ERBB2 pathway could have additive or synergistic roles. The UPS is definitely one such candidate. In addition to ERBB2, EGFR is overexpressed in basal-like breast cancer and is a possible target for new anti-breast cancer reagents.

BRCA1 and basal-like cancer

Although the significance of the E3 activity of BRCA1 in the BRCA1 functional network (Figure 1) is only partially understood, it is obvious that its activity is crucial in the prevention of a certain subset of breast cancers [56,103-105]. In terms of the potential for targeting this activity for breast cancer therapy, tumor suppressors such as BRCA1 are not ideal targets for therapy because small molecule activators can be more difficult to produce than small molecule inhibitors. This is absolutely true for the treatment of breast cancer caused by BRCA1 gene mutations. However, if the breast cancer results from other factors besides BRCA1 gene mutations, it might be conjectured that discovering additional drugs for the same protein or the UPS in hormone receptor signaling could be one such alternative.

List of abbreviations used

DUB: deubiquitylation enzyme; GFR: growth factor receptor; PI3K: phosphoinositide 3-kinase; UPS: ubiquitin proteasome system.

Competing interests

T. Ohta is the author of two patents:
1) Ohta T. Carcinostatic method using BRCA1-BARD1 pathway. WO2005073379 (08/11/2005).

2) Ohta T and Wu W. A method for the ubiquitination of common subunits of RNA polymerases. WO2007046538 (04/26/2007).

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Additional TPdb reviews on the ubiquitin-proteasome system are also available in BMC Biochemistry – see Volume 8 Supplement 1 http://www.biomedcentral.com/1471-2091/8?issue=S1#issue51.

References
1. Hershko A, Ciechanover A: The ubiquitin system. Annu Rev Biochem 1998, 67:425-79.
2. Pickart CM, Edkins MJ: Ubiquitin: structures, functions, mechanisms. Biochim Biophys Acta 2004, 1695(1-3):55-72.
3. Pickart CM: Back to the future with ubiquitin. Cell 2004, 116:181-90.
4. Mukhopadhyay D, Riezman H: The co-chaperone protein CHIP regulates protein triage in the endoplasmic reticulum. Trends Cell Biol 2003, 13(5):212-20.
5. Ohta T, Fukuda M: Ubiquitin and breast cancer. Oncogene 2004, 23(11):1789-98.
6. Nijman SM, Luna-Vargas MP, Velds A, Brummelkamp TR, Dirac AM, Nawaz Z, Lonard DM, Dennis AP, Smith CL, O'Malley BW: Fan M, Park A, Nephew KP: Torres-Arzayus MI, Yuan J, Dellagatta JL, Lane H, Kung AL, Brown M: Targeting the AIB1 oncogene through mammalian target of rapamycin inhibition in the mammary gland. Cancer Res 2006, 66(23):11381-8.
7. Harari D, Yarden Y: Molecular mechanisms underlying ErbB2/HER2 action in breast cancer. Oncogene 2000, 19(53):6102-6114.
8. Bhargava R, Gerald WL, Li AR, Pan Q, Lal P, Ladanyi M, Chen B: EGFR gene amplification in breast cancer: correlation with epidermal growth factor receptor mRNA and protein expression and HER-2 status and absence of EGFR-activated MAPK signalling. J Pathol 2004, 203(4):586-93.
9. Park K, Han S, Shin E, Kim HJ, Kim JY: EGFR gene and protein expression in breast cancers. Eur J Surg Oncol 2007, 33(8):956-60.
10. Olayioye MA, Neve RM, Lane HA, Hynes NE: The ErbB signaling network: receptor heterodimerization in development and cancer. EMBO J 2000, 19(13):3159-67.
11. Moseyson S, Shitkman K, Katz M, Zwang Y, Vereg B, Szollosi J, Yarden Y: Endocytosis of receptor tyrosine kinases is driven by monoubiquitination, not polyubiquitination. J Biol Chem 2007, 282(24):13233-43.
12. Dikic I, Giordano S: Negative receptor signalling. Curr Opin Cell Biol 2005, 17(2):128-35.
13. Bucci C, Thomsen P, Nicciziani P, McCarthy J, van Deurs B: Rab7: a key to lysosome biogenesis. Mol Biol Cell 2001, 12(2):167-80.
14. de Rensi S, Sonnichsen B, Zerial M: Divalent Rab effectors regulate the sub-compartmental organization and sorting of early endosomes. Nat Cell Biol 2002, 4(2):124-33.
15. Muthuswamy SK, Gilman M, Brugge JS: Controlled dimerization of ErbB receptors provides evidence for differential signaling by homo- and heterodimers. Mol Biol Cell 1999, 10(9):6845-57.
16. Graus-Porta D, Beerli RR, Daly JM, Hynes NE: Erb-B-2, the preferred heterodimerization partner of all ErbB receptors, is a mediator of lateral signaling. EMBO J 1997, 16(7):1647-55.
17. Saji S, Okumura N, Eguchi H, Nakashima S, Suzuki A, Toi M, Nozawa X, Shang Y: ErbB signaling complexes for cancer therapy. Mod Pathol 2005, 18(8):1027-33.
18. Torres-Arzayus MI, Yuan J, DellaGatta JL, Lane H, Kung AL, Brown M: Tumor-inhibitory antibodies to HER-2/HER2-2 may act by recruiting c-Cbl and enhancing ubiquitination of HER-2. Cancer Res 2000, 60(13):3384-8.
19. Zhou P, Ferrandes N, Dodge IL, Reddi AL, Rao N, Safran H, DiPietrillo TA, Wazer DE, Band V, Band H: ErbB2 degradation mediated by the c-chaaperone protein CHIP. J Biol Chem 2003, 278(16):13829-37.
20. Graus-Porta D, Beerli RR, Daly JM, Hynes NE: Erb-B-2, the preferred heterodimerization partner of all ErbB receptors, is a mediator of lateral signaling. EMBO J 1997, 16(7):1647-55.
21. Lonard DM and Nawaz Z: The co-chaperone CHIP regulates protein triage in the endoplasmic reticulum. Trends Cell Biol 2003, 13(5):212-20.
22. Douroumis D, Karamizoglou M, Mavridis K, Isola J: erbB signaling and activation of the PI3K/AKT pathway in transgenic mice define AIB1 as an oncogene. Cancer Cell 2004, 6:263-74.
23. Torres-Arzayus MI, Yuan J, DellaGatta JL, Lane H, Kung AL, Brown M: Targeting the AIB1 oncogene through mammalian target of rapamycin inhibition in the mammary gland. Cancer Res 2006, 66(23):11381-8.
24. Shao W, Keeton EK, McDonnell DP, Brown M: Coactivator AIB1 links estrogen receptor transcriptional activity and stability. Mol Endocrinol 2004, 18(12):1599-604.
25. Torres-Arzayus MI, Font de Morra J, Yuan J, et al.: High tumor incidence and activation of the PI3K/AKT pathway in transgenic mice define AIB1 as an oncogene. Cancer Cell 2004, 6:263-74.
26. Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, et al.: Molecular portraits of human breast tumours. Nature 2000, 406(6797):747-752.
27. Tannir M, Kapanen Al, Junttila T, Raheem O, Grenman S, Elo J, Elenius K, Isola J: Characterization of a novel cell line established
from a patient with Herceptin-resistant breast cancer. Mol Cancer Ther 2004, 3(12):1585-92.

39. Srinivasan, Tatham, R. Parker, J. Hasteie, T. Marron JS, Nobel A, Deng S, Johnsen H, Pesich R, Geisler S, et al.: Repeated observation of breast subtype expression in independent gene expression data sets. Proc Natl Acad Sci USA 2003, 100(14):8418-8423.

40. Fouls KE, Stanford JS, Chappuis PO, Begin LR, Goffin JR, Wong N, Trudel M, Alaei L: Germline BRCA1 mutations and a basal epithelial phenotype in breast cancer. J Natl Cancer Inst 2003, 95(19):1482-1485.

41. Lakhani SR, Reis-Filho JS, Hattersley T, Sung HY, Vujic I, et al.: Basal-like breast cancer is associated with BRCA1 and BRCA2 gene mutations and a basal epithelial phenotype. J Natl Cancer Inst 2005, 11(14):5175-5180.

42. Horndro E, Osorio A, Palacios J, Benitez J: Pathology and gene expression of hereditary breast tumors associated with BRCA1 and CHEK2 gene mutations. Oncogene 2006, 25(43):5837-45.

43. Turner N, Tutt A, Ashworth A: Hallmarks of BRCA1 deficiency in sporadic cancers. Nat Rev Cancer 2004, 4:814-819.

44. Staff S, Isola J, Tanner M: Haplo-insufficiency of BRCA1 in sporadic breast cancer. Cancer Res 2003, 63(16):479-483.

45. Turner NC, Reis-Filho JS, Russell AM, Springall R, Ryder K, Steele D, Savage K, Gillett CE, Schmitt FC, Ashworth A, et al.: BRCA1 dysfunction in sporadic basal-like breast cancer. Oncogene 2007, 26(14):2126-2132.

46. Richardson SE, Al, Wang ZC, De Nicolo A, Lu X, Brown M, Mirou J, Liao X, Iglehart JD, Livingston DM, Ganesan S: X chromosome abnormalities in basal-like human breast cancer. Cancer Cell 2006, 9(2):121-32.

47. McCarthy A, Savage K, Gabriel A, Naceur C, Reis-Filho JS, Ashworth A: A mouse model of basal-like breast canceroma with metastatic plastic elements. J Pathol 2007, 211(4):389-98.

48. Zheng L, Li S, Boyer TG, Lee WH: Lessons learned from BRCA1 and BRCA2. Oncogene 2000, 19(53):6159-75.

49. Swift M, Morrell D, Crompton E, Chamberlin AR, Skl尼克 MH, Bishop DT: The incidence and gene frequency of ataxia-telangiectasia in the United States. Am J Hum Genet 1986, 39(5):573-83.

50. Ahmed M, Rahman N: ATM and breast cancer susceptibility. Oncogene 2006, 25(33):5905-11.

51. Meijers-Heijboer H, Ouweland A, van den, Klijn J, Wasielewski M, de Man M, Adams J, Schenkein DP, Anderson KC: A mechanism for transcriptional repression dependent on K6 of ubiquitin. J Biol Chem 2003, 278(37):34743-6.

52. Nishikawa H, Ooka S, Kato S, Arima K, Okamoto N, Klevit RE, Fukuda M, Ohta T: Mass spectrometric and mutational analyses reveal Lys-6-linked polyubiquitin chains catalyzed by BRCA1-BARD1 ubiquitin ligase. J Biol Chem 2004, 279(9):3916-24.

53. Chen A, Kleiman FE, Manley JL: Autoubiquitination of the BRCA1-BARD1 RING ubiquitin ligase. J Biol Chem 2002, 277(24):22085-92.

54. Mallory DL, Vandenberg CJ, Hiom K: Activation of the E3 ligase function of the BRCA1-BARD1 complex by polyubiquitin chains. EMBO J 2002, 21(4):6755-62.

55. Starita LM, Machida Y, Sarkanen S, Elias JE, Griffin K, Schlegel BP, Gygi SP, Parvin JD: BRCA1-dependent ubiquitination of gammab-tubulin regulates centrosome number. Mol Cell Biol 2004, 24(8):6847-66.

56. Eakin CM, Maccoss MJ, Finney GL, Klevit RE: Estrogen receptor alpha is a putative substrate for the BRCA1 ubiquitin ligase. Proc Natl Acad Sci USA 2007, 104(14):5794-9.

57. Sato K, Hayami R, Wu W, Nishikawa T, Nishikawa H, Okuda Y, Ogata H, Fukuda M, Ohta T: Nuclear phosphoribosyl transferase is a candidate substrate for the BRCA1-BARD1 ubiquitin ligase. J Biol Chem 2004, 279(30):30919-22.

58. Kleiman FE, Wu-Baer F, Forsca D, Kaneko S, Baer R, Manley JL: BRCA1/BARD1 inhibition of mRNA 3' processing involves targeted degradation of RNA polymerase II. Genes Dev 2005, 19(10):1227-37.

59. Starita LM, Horwitz AA, Keogh MC, Ishioica C, Parvin JD, Chibna B: BRCA1/BARD1 ubiquitinate phosphorylated RNA polymerase II. J Biol Chem 2005, 280(26):26289-96.

60. Yu X, Fu S, Lai M, Baer R, Chen J: BRCA1 ubiquititates its phosphorylation-dependent binding partner CtDP. Genes Dev 2006, 20(13):1721-6.

61. Wu W, Nishikawa H, Hayami R, Sato K, Honda A, Aratani S, Nakajima T, Fukuda M, Ohta T: BRCA1 ubiquititates RB8 in response to DNA damage. Cancer Res 2007, 67(3):951-8.

62. Poole AJ, Li Y, Kim Y, Lin SC, Lee WH, Lee EY: Prevention of BRCA1-mediated mammary tumorigenesis in mice by a progestosterone antagonist. Science 2006, 314(5804):1467-70.

63. Horwitz AA, Affar E, B, Hiene GF, Shy, I, Parvin JD: A mechanism for transcriptional repression dependent on the BRCA1 E3 ubiquitin ligase. Proc Natl Acad Sci USA 2007, 104(16):6614-9.

64. Joaozeiro CA, Anderson KC, Hunter T: Proteasome inhibitor drugs on the rise. Cancer Res 2006, 66(16):7840-2.

65. Li X, Holtrege H, Gulden H van der, Tumur-Mulder M, Zevenhoven J, Velds A, Velds AH, Velds LM, Vliet MH, Wessels LF, Peterle J, Berns A, Jonkers J: Somatic loss of BRCA1 and p53 in mice induces mammary tumors with features of human BRCA1-mutated basal-like breast cancer. Proc Natl Acad Sci USA 2007, 104(23):9290-5.

66. Li X, Shen L, Zhang J, Ju S, Shen L, Liu X, Han H, Han W, Yao L: Degradation of HER2 by Cbl-based chimeric ubiquitin ligases. Cancer Res 2007, 67(18):9761-24.

67. Hideshima T, Richardson P, Chauhan D, Palombella VJ, Elliott Pj, Adams J, Anderson KC: The proteasome inhibitor PS-341 inhibits growth, induces apoptosis, and overcomes drug resistance in human multiple myeloma cells. Cancer Res 2001, 61(7):3071-6.

68. Richardson PR, Barlogie B, Berenson J, Singhal S, Jagannath S, Irwin D, Rajkumar SV, Srkalovic G, Alsina M, Alexanian R, Siegel D, Orlowski RZ, Kuter D, Limentani SA, Lee S, Hideshima T, Esseltine DL, Kauff- man M, Adams J, Schenken DP, Anderson KC: A phase 2 study of bortezomib in relapsed, refractory myeloma. N Engl J Med 2003, 348(26):2609-17.

69. Adams J: The proteasome: structure, function, and role in the cell. Cancer Treat Rev 2003, 29(Suppl 1):1-90.

70. Hideshima T, Chauhan D, Richardson P, Mitsiades C, Mitsiades N, Hayashi T, Munsch N, Dang L, Castro A, Palombella V, Adams J,
98. Ohta T, Michel JJ, Schottelius AJ, Xiong Y: A novel active proteasome inhibitor induces apoptosis in multiple myeloma cells with mechanisms distinct from Bortezomib. *Cancer Cell* 2005, 8(5):407-19.

99. Hanada M, Sugawara K, Kaneta K, Oki T: Epoxomicin, a new antibiotic of microbial origin. *J Antibiot (Tokyo)* 1990, 43(1):1-8.

100. Sin N, Kim KB, Elofsson M, Meng L, Kwok BH, Crews CM: Total synthesis of the potent proteasome inhibitor epoxomicin: a useful tool for understanding proteasome biology. *Bioger Med Chem Lett* 1999, 9(15):2283-8.

101. Perez EA, Hortobagyi GN: Ongoing and planned adjuvant trials with trastuzumab. *Semin Oncol* 2000, 27(6 Suppl 11):26-32.

102. Ruffner H, Joazeiro CA, Hemmati D, Hunter T, Verma IM: Cancer-predisposing mutations within the RING domain of BRCA1: loss of ubiquitin protein ligase activity and protection from radiation hypersensitivity. *Proc Natl Acad Sci USA* 2001, 98(9):5134-5139.

103. Brzovic PS, Keeffe JR, Nishikawa H, Miyamoto K, Fox D 3rd, Fukuda M, Ohta T, Klevit R: Binding and recognition in the assembly of an active BRCA1/BARD1 ubiquitin-ligase complex. *Proc Natl Acad Sci USA* 2003, 100(10):5646-5651.

104. Morris JR, Pangon L, Boullet C, Katagiri T, Keep NH, Solomon E: Genetic analysis of BRCA1 ubiquitin ligase activity and its relationship to breast cancer susceptibility. *Hum Mol Genet* 2006, 15(4):599-606.

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Anderson KC: NF-kappa B as a therapeutic target in multiple myeloma. *J Biol Chem* 2002, 277(19):16639-47.

97. Tanaka K, Iwamoto S, Gon G, Nohara T, Iwamoto M, Tanigawa N: Expression of survivin and its relationship to loss of apoptosis in breast carcinomas. *Clin Cancer Res* 2000, 6(1):127-134.

98. XIA W, Bai J, Strum J, Liu L, Carrick K, Graham KM, Treece AL, Hardwick MA, Dush M, Liao Q, et al.: Regulation of survivin by ErbB2 signaling: therapeutic implications for ErbB2-overexpressing breast cancers. *Cancer Res* 2006, 66(3):1640-1647.

99. Ohsa T, Michel JJ, Schottelius AJ, Xiong Y: ROCL, a homolog of APOC1, represents a family of cullin partners with an associated ubiquitin ligase activity. *Mol Cell 1999, 3(4):535-41.

100. Kamura T, Kepp DM, Conrad MN, Skowryda D, Moreland RJ, Illiopoulos O, Lane WS, Kaelin WG Jr, Elledge SJ, Conaway RC, Harper JW, Conaway JW: Rbx1, a component of the VHL tumor suppressor complex and SCF ubiquitin ligase. *Science* 1999, 284(5414):657-61.

101. Chen C, Seth AK, Aplin AE: Genetic and expression aberrations of E3 ubiquitin ligases in human breast cancer. *Mol Cancer Res* 2004, 6(10):695-707.

102. Early Breast Cancer Trialists' Collaborative Group (EBCTCG): Effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: an overview of the randomised trials. *Lancet* 2005, 365(9472):1687-717.

103. Perez EA, Hortobagyi GN: Ongoing and planned adjuvant trials with trastuzumab. *Semin Oncol* 2000, 27(6 Supplement 11):26-32.

104. Ruffner H, Joazeiro CA, Hemmati D, Hunter T, Verma IM: Cancer-predisposing mutations within the RING domain of BRCA1: loss of ubiquitin protein ligase activity and protection from radiation hypersensitivity. *Proc Natl Acad Sci USA* 2001, 98(9):5134-5139.

105. Brzovic PS, Keeffe JR, Nishikawa H, Miyamoto K, Fox D 3rd, Fukuda M, Ohta T, Klevit R: Binding and recognition in the assembly of an active BRCA1/BARD1 ubiquitin-ligase complex. *Proc Natl Acad Sci USA* 2003, 100(10):5646-5651.

106. Morris JR, Pangon L, Boullet C, Katagiri T, Keep NH, Solomon E: Genetic analysis of BRCA1 ubiquitin ligase activity and its relationship to breast cancer susceptibility. *Hum Mol Genet* 2006, 15(4):599-606.

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