Research Article

IL-10 and ARG-1 Concentrations in Bone Marrow and Peripheral Blood of Metastatic Neuroblastoma Patients Do Not Associate with Clinical Outcome

Fabio Morandi,1 Michela Croce,2 Giuliana Cangemi,3 Sebastiano Barco,3 Valentina Rigo,2 Barbara Carlini,1 Loredana Amoroso,4 Vito Pistoia,1 Silvano Ferrini,2 and Maria Valeria Corrias1

1 Laboratorio di Oncologia IRCCS Istituto Giannina Gaslini, 16148 Genova, Italy
2 Laboratorio di Bioterapia, IRCCS AOI San Martino-Istituto Nazionale per la Ricerca sul Cancro, 16132 Genova, Italy
3 Laboratorio di Patologica Clinica IRCCS Istituto Giannina Gaslini, 16148 Genova, Italy
4 U.O. Oncologia IRCCS Istituto Giannina Gaslini, 16148 Genova, Italy

Correspondence should be addressed to Maria Valeria Corrias; mariavaleriacorrias@ospedale-gaslini.ge.it

Received 1 August 2014; Accepted 25 September 2014

Academic Editor: Marco Antonio Velasco-Velázquez

Copyright © 2015 Fabio Morandiet al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The expression of the immunosuppressive molecules IL-10 and arginase 1 (ARG-1), and of FOXP3 and CD163, as markers of regulatory T cells (Treg) and macrophages, respectively, was evaluated in bone marrow (BM) and peripheral blood (PB) samples collected at diagnosis from patients with metastatic neuroblastoma (NB). IL-10 and ARG-1 plasma concentrations were measured and the association of each parameter with patients’ outcome was tested. The percentages of immunosuppressive Treg and type-1 regulatory (Tr1) cells were also determined. In both BM and PB samples, IL-10 mRNA expression was higher in metastatic NB patients than in controls. IL-10 plasma concentration was higher in patients with NB regardless of stage. Neither IL-10 expression nor IL-10 plasma concentration significantly associated with patient survival. In PB samples from metastatic NB patients, ARG-1 and CD163 expression was higher than in controls but their expression did not associate with survival. Moreover, ARG-1 plasma concentration was lower than in controls, and no association with patient outcome was found. Finally, in metastatic NB patients, the percentage of circulating Treg was higher than in controls, whereas that of Tr1 cells was lower. In conclusion, although IL-10 concentration and Treg percentage were increased, their contribution to the natural history of metastatic NB appears uncertain.

1. Introduction

It is generally assumed that the efficacy of anticancer immunotherapy is hampered by immune suppressive and tolerogenic mechanisms, including soluble factors, produced by both neoplastic and other tumor-instructed immune suppressive cells [1].

Neuroblastoma (NB) is a pediatric neuroectodermal solid tumor with a heterogeneous clinical behavior [2]. Children presenting with metastatic disease at diagnosis, that is, stage 4 according to the International Neuroblastoma Staging System (INSS) [3] or stage M according to the International Neuroblastoma Risk Group Staging System (INRG-SS) [4], aged more than 18 months, have poor survival rate despite intensive multimodal therapy. In these patients, immunotherapy with anti-disialoganglioside GD2 (GD2) antibody, with or without IL-2 or granulocyte macrophage-colony stimulating factor (GM-CSF), is administered in a minimal residual disease condition after myeloablative therapy, in ongoing high-risk protocols of the Children Oncology Group (COG) [5] and the International Society of Pediatric Oncology-European Neuroblastoma (SIOPEN) [6]. It is of note that, in spite of low/absent Human Leucocyte Antigen (HLA) class I expression by NB cells [7], resting NK cells have limited efficacy against neuroblasts, because the latter cells express an inhibitory ligand [8] and may release immunosuppressive soluble factors [9–13]. Nonetheless, the combination of
anti-GD2 mAb with IL-2 or GM-CSF can activate NK-cell or granulocyte mediated antibody dependent cell-mediated cytotoxicity (ADCC) against NB cells and lead to improved survival [14].

In human NB, information on the presence and activity of specific subsets of immune suppressive cells and soluble factors is limited [15]. A gene expression study performed on primary tumors [16] suggested a role for arginase (ARG)-1 myeloid-derived suppressor cells in the prognosis of metastatic NB patients. Recently, the same authors [17] demonstrated that tumors from high-risk NB patients present greater infiltration with CD163+ M2-type tumor associated macrophages (TAM). In addition, Song et al. showed that NKT cell-mediated killing of TAMs associated with a better outcome [18].

Immune suppressive CD4+CD25+FOXP3+ regulatory T cells (Treg) are increased in several types of tumors, where they may play a relevant tolerogenic role [19]. However, no increase in their number was found in a small cohort of localized and metastatic NB patients [20].

A potential immunosuppressive role of T regulatory type 1 (Tr1) cells has never been investigated in NB patients. Tr1 cells have been firstly identified in mice as CD4+ T cells secreting high amounts of IL-10 [21]. Subsequently, Tr1 cells have also been characterized in humans [22-24] and are now identified as CD4+CD45R0+CD49b+LAG-3+ T cells [25].

The immunosuppressive cytokine IL-10 is produced by both Treg and Tr1 cells and by cells of the innate immunity, such as NK cells and macrophages [26]. Gowda et al. have shown that, in patients with localized NB, plasma IL-10 concentration was higher than in patients with metastatic disease and suggested that high IL-10 levels may reflect the activation of an effective innate immunity [20].

Our working hypothesis is that, in patients with metastatic NB, bone marrow- (BM-) infiltrating and circulating tumor cells could contribute to the instruction of different immune suppressive mechanisms in the periphery. We thus evaluated the mRNA expression of IL-10, FOXP3, as marker of Treg, and ARG-1 and CD163, as markers of myeloid suppressor cells/macrophages in both BM and peripheral blood (PB) samples collected at diagnosis from metastatic NB patients. We also measured IL-10 and ARG-1 plasma levels and these markers of immune suppressive mechanisms were tested for association with event-free and overall survival (EFS and OS, resp.). Finally, we determined the percentage of Treg and Tr1 cells in PB samples from metastatic NB patients and age-matched controls.

2. Materials and Methods

2.1. Patients. Forty-one consecutive patients diagnosed in Italy with stage 4 NB between November 2001 and December 2006, aged >18 months at onset, were included in the molecular analysis if both BM and PB samples collected at diagnosis were available. Disease staging [3] was made at the referring oncology center and centrally reviewed at the Gaslini Institute. The median age at diagnosis was 2.9 years (range: 1.5–9.7) and the median follow-up was 39.2 months (range: 10.4–97.4). Thirty % of the patients presented MYCN amplification. As controls for molecular analysis, 20 BM samples and 20 PB samples from 40 healthy donors were used after informed consent was given.

ELISA was performed on plasma samples collected at diagnosis from a total of 102 NB patients (50 stage 4 and 52 non-stage 4) admitted to the Gaslini Institute between January 1994 and December 2010. As controls, 55 plasma samples collected between January 2010 and December 2012 from children admitted to the Gaslini Institute for accidental injuries were used. The median age at diagnosis of the stage 4 patients was 3.4 years (range: 1.6–9.6) and the median follow-up was 20.3 months (range: 6.8–138.5). Twenty-six % of stage 4 patients and 29.1% of non-stage 4 patients presented MYCN amplification.

IL-10 determination was performed on all plasma samples available, whereas ARG-1 was tested on a fraction of them, since not enough plasma was available to perform both assays; namely, 57 plasma samples from NB patients (49 stage 4 and 8 non-stage 4) and 30 from healthy age-matched controls were analyzed. All samples were stored at −80°C until use.

Follow-up data at January 2014 were retrieved from the Italian Neuroblastoma Registry (INBR) [27]. The studies were approved by the institutions’ ethical committees and all analyses were performed according to the Helsinki declaration.

2.2. RNA Extraction and RT-qPCR Analysis. Total RNA was extracted from PB or BM samples as described in [28]. One hundred ng of total RNA was reverse transcribed and then amplified for each molecular marker in duplicate by qPCR, using the following assays from Life Technology (Life Technologies Europe BV, Monza, Italy): IL-10: Hs00961622_ml, FOXP3: Hs00203958_ml, CD163: Hs00174705_ml, ARG-1: Hs00968979_ml, and primers and probe for β2-microglobulin (β2M) [29]. The level of expression of each marker was normalized to the expression of β2M, according to the delta Ct method [30], and results were reported as 2−ΔΔCt.

To exclude DNA contamination, samples in the absence of reverse transcriptase were analyzed in each qPCR assay, and water was included as negative control.

2.3. ELISA. ARG-1 and IL-10 concentrations were measured using Arginase I Human ELISA kit (Hycult Biotech, Milan, Italy) and human high sensitivity IL-10 ELISA Kit (Diacline Gen-Probe, Besançon Cedex, France), respectively, following manufacturers’ protocols.

2.4. Flow Cytometry. The percentage of Treg and Tr1 cells was evaluated by flow cytometry on whole PB samples. Treg were identified using Human Regulatory T Cell Cocktail (BD Biosciences, Milan, Italy), containing anti-CD4 FITC, anti-CD25 PE-Cy7, and anti-CD127Alexa Fluar 647 antibodies, following manufacturer’s protocol. Tr1 were evaluated using anti-CD4 PE-Cy7 (eBiosciences, San Diego, CA), anti-CD45R0 APC (eBiosciences), anti-CD49b PE (R&D Systems, Space, Milan, Italy), and anti-LAG-3 FITC (R&D Systems) antibodies. Samples were subjected to erythrocytes lysis using...
BD FACS lysis (BD Biosciences) and washed. Cells were run on Gallios cytometer (Beckman Coulter, Cassina De’ Pecchi, MI, Italy) acquiring at least 10^4 events. Data were analyzed using Kaluza software (Beckman Coulter). Treg were identified as CD4^+ CD25^high CD127^low cells, whereas Tr1 were identified as CD4^+ CD45R0^− CD49b^− ARG-1^+ cells [25].

2.5. Statistical Analysis. Gaussian distribution of data was analyzed using Kolmogorov-Smirnov test. t-test or Mann-Whitney test was used to compare values depending on Gaussian distribution, while the Spearman ρ coefficient was used to assess correlation between variables. EFS and OS analyses were performed according to the Kaplan-Meier method and compared by the log-rank test. A P value < 0.05 was considered statistically significant. Analyses were made using the Prism software (GraphPad Software Inc., La Jolla, CA).

3. Results

3.1. Expression of IL-10, FOXP3, ARG-1, and CD163 mRNA in BM and PB Samples and Association with EFS and OS. Expression of IL-10, FOXP3, ARG-1, and CD163 mRNAs was analyzed by RT-qPCR in BM and PB samples collected at diagnosis from 41 children with metastatic NB and from 20 healthy donors. As shown in Figure 1(a), the expression of IL-10 mRNA in BM samples was significantly higher in NB patients (2−ΔΔCt: 1.28 × 10^−3 ± 1.32 × 10^−3) than in controls (2−ΔΔCt: 0.41 × 10^−3 ± 0.26 × 10^−3; P = 0.002), whereas the expression of FOXP3, ARG-1, and CD163 mRNAs was similar between the two groups.

In PB samples, metastatic NB patients showed higher expression of IL-10 (2−ΔΔCt: 2.89 × 10^−3 ± 4.66 × 10^−3 versus 0.41 × 10^−3 ± 0.56 × 10^−3; P < 0.0001), ARG-1 (2−ΔΔCt: 20.21 × 10^−3 ± 24.32 × 10^−3 versus 4.54 × 10^−3 ± 6.54 × 10^−3; P < 0.0001), and CD163 (2−ΔΔCt: 45.2 × 10^−3 ± 62.2 × 10^−3 versus 13.5 × 10^−3 ± 4.7 × 10^−3; P = 0.02) mRNAs than healthy donors (Figure 1(b)). However, no association with EFS or OS was found for any of the four markers analyzed, either in BM (Supplemental Figures 1(a) and 1(b), resp., available online at http://dx.doi.org/10.1155/2015/718975) or in PB samples (Supplemental Figures 2(a) and 2(b), resp.).

3.2. Correlations between IL-10, FOXP3, ARG-1, and CD163 mRNA Expression in BM and PB Samples. We next analyzed potential correlations in the mRNA expression levels of the four markers in both BM and PB samples. In BM samples, IL-10 expression positively correlated with FOXP3 (r = 0.43; P = 0.0046, Figure 2(a)) and CD163 expression (r = 0.45; P = 0.0028, Figure 2(b)). In PB samples, CD163 correlated with ARG-1 (r = 0.51; P = 0.0005, Figure 2(c)), whereas IL-10 showed an inverse correlation with FOXP3 (r = −0.57; P < 0.0001, Figure 2(d)). These data suggested that IL-10 production in BM may be ascribed to both CD163^+ and Treg cells, whereas in PB IL-10 was unlikely produced by FOXP3^+ cells. Conversely, in PB, CD163^+ myeloid cells may be responsible for ARG-1 production.

3.3. NB Patients Display an Increased Plasma Concentration of IL-10 Irrespective of Disease Stage. Given the high expression of IL-10 mRNA in metastatic NB patients, we then measured IL-10 plasma concentration. As shown in Figure 3(a), IL-10 concentration was significantly higher in metastatic NB patients (8.68 ± 4.45 pg/mL) than in age-matched controls (0.41 ± 0.19 pg/mL, P < 0.0001). However, when we compared IL-10 plasma levels in metastatic and localized NB patients, no significant difference was found (Figure 3(b), stage 4: 12.18 ± 7.44 pg/mL; non-stage 4: 6.35 ± 3.5 pg/mL). In addition, as observed for IL-10 mRNA expression, IL-10 plasma levels in metastatic NB patients did not associate with either EFS (Figure 3(c)) or OS (Figure 3(d)).
3.4. NB Patients Display Lower Plasma Concentration of ARG-1 than Healthy Subjects. We next analyzed the plasma concentration of soluble ARG-1 in PB samples. In contrast with the mRNA expression, ARG-1 concentration was significantly lower in NB patients (19.52 ± 4.58 pg/mL) than in age-matched controls (44.89 ± 4.17 pg/mL, \( P < 0.0001 \), Figure 4(a)). However, ARG-1 concentration was significantly higher in stage 4 (22.49 ± 5.41 pg/mL) than in non-stage 4 (8.2 ± 5.1 pg/mL, \( P = 0.01 \)) NB patients (Figure 4(b)), suggesting that ARG-1 release may be related to a more advanced disease stage. However, no significant correlation was found between ARG-1 plasma levels in metastatic NB patients and their EFS (Figure 4(c)) or OS (Figure 4(d)).

3.5. Treg but Not Tr1 Cells Are Expanded in PB from NB Patients. Since we detected increased IL-10 plasma levels in NB patients, we asked whether two regulatory T cell populations able to secrete IL-10, namely, CD4\(^+\)CD25\(^{high}\)CD127\(^{low}\) Treg and CD4\(^+\)CD45R0\(^+\)CD49b\(^+\)LAG-3\(^-\)Tr1 cells, were also increased. As shown in Figure 5, the percentage of Treg was significantly higher in metastatic NB patients (% of cells: 0.91 ± 0.39) than in age-matched controls (% of cells: 0.32 ± 0.06, \( P = 0.02 \)), whereas the percentage of Tr1 cells was lower in NB patients (% of cells: 1.1 ± 0.19) than in controls (2.32 ± 0.4, \( P = 0.01 \)).

4. Discussion

To the best of our knowledge, this is the first report where both BM and PB samples from metastatic NB patients were analyzed to evaluate the prognostic role of immune suppressive soluble factors and cell subsets. In addition, this is the first report on Tr-1 cells in NB.

Our study clearly shows that IL-10 mRNA expression levels were significantly increased in both PB and BM samples and plasma IL-10 concentration was also elevated. However, no significant association with clinical outcome was found. In addition, similar IL-10 plasma levels were found in children with metastatic and localized NB, whose prognosis is completely different [4], confirming that increase in IL-10 concentration is unrelated to stage and prognosis. These findings are different in part from those reported by Gowda.
et al., showing higher IL-10 plasma concentration in the low-risk group of a small cohort of NB patients [20]. This discrepancy may be related to the study of larger cohorts of patients, belonging to stage 4 and non-stage 4, in the present report than in the previous one. Moreover, in our study, only patients >18 months at diagnosis were included, thus excluding the effect of age.

Our study also shows that the significant increase of ARG-1 mRNA levels in PB did not correspond to elevated ARG-1 plasma concentration. This finding may be related to the fact that ARG-1 plasma levels depend on the amount released by MDSC or by granulocytes, following their degranulation. However, neither ARG-1 expression nor ARG-1 plasma concentration significantly associated with survival of metastatic NB patients. Thus, our results do not support a relevant role of IL-10 and ARG-1 in the natural history of metastatic NB.

The finding that IL-10 expression levels in BM significantly correlated with both FOXP3 and CD163 expression suggested that IL-10 can be produced not only by the CD4+/CD25+/CD127low Treg cells, as already demonstrated [31], but also by CD163+ macrophages. These data are in accordance with previous reports showing a significant correlation between CD163+ TAM and IL-10 release [32–34]. Surprisingly, in PB samples from metastatic NB patients, IL-10 expression inversely correlated with FOXP3 expression and no correlation was found with CD163 expression, making it unlikely that Treg or macrophages could be responsible for IL-10 production in PB.

In our study, the increased percentage of Treg found in metastatic NB patients appeared to be compensated by a decrease in the other regulatory T cell subsets able to secrete IL-10 [35], the Tr1 cells. The increased percentage of Treg cells
found in metastatic NB patients is in accordance with the finding reported by Tilak et al. [36] in human NB and with Treg involvement demonstrated in murine NB models [37–40]. However, a direct relationship between the expansion of Treg cells in the PB and the increased levels of IL-10 in sera from NB patients could not be demonstrated. From our findings, also Tr1 cells did not appear to be responsible for the increased IL-10 plasma concentration. The present study is the first reporting on circulating Tr1 cells in NB patients. Thus, in view of the limited number of cases available for Tr1 analysis, further studies on larger cohorts are needed to confirm the reduction of Tr1 cells in metastatic NB. Moreover, this finding is in contrast with studies in head and neck tumors, where increased Tr1 cells were reported [41, 42].

It is of note that, in addition to the cell populations studied here, also NK, dendritic, and B cells can produce IL-10 [26]. Moreover, since IL-10R blockade prolonged survival in a murine NB model [43], we cannot exclude that inhibition of IL-10 levels may improve the clinical outcome of metastatic NB patients. Finally, the inverse correlation between FOXP3 and IL-10 expression in PB could be related to the fact that FOXP3 is not a Treg cell-specific marker but is also associated with T cell activation [44, 45].

Our study also showed that CD163 and ARG-1 mRNA expression were significantly higher in metastatic NB patients and significantly correlated with each other, suggesting that ARG-1 was expressed by myeloid cells. In different human tumors, ARG-1 was found overexpressed [46–48] and

### Figure 4: ARG-1 plasma concentration in 57 NB patients and 30 healthy children (a, b), and association between ARG-1 plasma levels of stage 4 patients with EFS and OS (c, d). In (a), all NB patients and controls are shown. In (b), NB patients are stratified in stage 4 (n = 49) and non-stage 4 (n = 8). Results are expressed as pg/mL. Horizontal bars indicated means. *P* values are indicated where differences are significant. Kaplan-Meyer plot of EFS (c) and OS (d) obtained by stratifying the 49 stage 4 NB patients according to ARG-1 plasma levels above (dotted line) or below (continuous line) median value. Y-axes represent % of event-free or alive patients, respectively. X-axes represent time of survival (months).
5. Conclusions

Our data exclude a prognostic role of IL-10 and ARG-1 in metastatic NB and show that in human NB circulating Tr1 cells are decreased, whereas Treg cells are significantly increased. These data suggest the existence of a complex network of immune regulatory interactions involving IL-10 and Treg cells, whose role in immune escape of NB should be further addressed. Combining agents targeting Treg inhibitory functions with immune enhancing agents may represent an interesting approach for therapeutic interventions.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Authors’ Contribution

Fabio Morandi and Michela Croce equally contributed to the work. Silvano Ferrini and Maria Valeria Corrias equally contributed to the work.

Acknowledgments

This study was supported by Fondazione Italiana per la Lotta al Neuroblastoma (to Maria Valeria Corrias and Michela Croce), by AIRC IG13518 (to Silvano Ferrini), and by Ministero Della Salute (Ricerca Corrente and Cinque per mille to Vito Pistoia and Silvano Ferrini). Michela Croce is a recipient of Fondazione Italiana per la Lotta al Neuroblastoma fellowship. Secretarial assistance by Mrs. Camilla Valentino is deeply acknowledged.

References

[1] M. Kroesen, D. Lindau, P. Hoogerbrugge, and G. J. Adema, “Immunocombination therapy for high-risk neuroblastoma,” Immunotherapy, vol. 4, no. 2, pp. 163–174, 2012.
[2] J. M. Maris, “Recent advances in neuroblastoma,” The New England Journal of Medicine, vol. 362, no. 23, pp. 2154–2211, 2010.
[3] G. M. Brodeur, J. Pritchard, F. Berthold et al., “Revisions of the international criteria for neuroblastoma diagnosis, staging, and response to treatment,” Journal of Clinical Oncology, vol. 11, no. 8, pp. 1466–1477, 1993.
[36] T. Tilak, S. Sherawat, S. Agarwala, R. Gupta, S. Vishnubhatla, and S. Bakhshi, “Circulating T-regulatory cells in neuroblastoma: a pilot prospective study,” Pediatric Hematology-Oncology, vol. 63, no. 5, pp. 501–511, 2014.

[37] V. Rigo, M. V. Corrias, A. M. Orengo et al., “Recombinant IL-21 and anti-CD4 antibodies cooperate in syngeneic neuroblastoma immunotherapy and mediate long-lasting immunity,” Cancer Immunology, Immunotherapy, vol. 63, no. 5, pp. 501–511, 2014.

[38] W. Jing, X. Yan, W. H. D. Hallett, J. A. Gershon, and B. D. Johnson, “Depletion of CD25+ T cells from hematopoietic stem cell grafts increases posttransplantation vaccine-induced immunity to neuroblastoma,” Blood, vol. 117, no. 25, pp. 6952–6962, 2011.

[39] W. Jing, J. A. Gershon, and B. D. Johnson, “Depletion of CD4 T cells enhances immunotherapy for neuroblastoma after syngeneic HSCT but compromises development of antitumor immune memory,” Blood, vol. 113, no. 18, pp. 4449–4457, 2009.

[40] M. Croce, M. V. Corrias, A. M. Orengo et al., “Transient depletion of CD4 T cells augments IL-21-based immunotherapy of disseminated neuroblastoma in syngeneic mice,” International Journal of Cancer, vol. 127, no. 5, pp. 1141–1150, 2010.

[41] C. Bergmann, L. Strauss, R. Zeidler, S. Lang, and T. L. Whiteside, “Expansion of human T regulatory type 1 cells in the microenvironment of cyclooxygenase 2 overexpressing head and neck squamous cell carcinoma,” Cancer Research, vol. 67, no. 18, pp. 8865–8873, 2007.

[42] C. Bergmann, L. Strauss, Y. Wang et al., “T regulatory type 1 cells in squamous cell carcinoma of the head and neck: mechanisms of suppression and expansion in advanced disease,” Clinical Cancer Research, vol. 14, no. 12, pp. 3706–3715, 2008.

[43] C. Brignonle, D. Marimpietri, F. Pastorino et al., “Anti-IL-10R antibody improves the therapeutic efficacy of targeted liposomal oligonucleotides,” Journal of Controlled Release, vol. 138, no. 2, pp. 122–127, 2009.

[44] R. J. deLeeuw, S. E. Kost, J. A. Kakal, and B. H. Nelson, “The prognostic value of FoxP3+ tumor-infiltrating lymphocytes in cancer: a critical review of the literature,” Clinical Cancer Research, vol. 18, no. 11, pp. 3022–3029, 2012.

[45] F. Martin, S. Ladoire, G. Mignot, L. Apetoh, and F. Ghiringhelli, “Human FOXP3 and cancer,” Oncogene, vol. 29, no. 29, pp. 4121–4129, 2010.

[46] F. Mussai, C. de Santo, I. Abu-Dayyeh et al., “Acute myeloid leukemia creates an arginase-dependent immunosuppressive microenvironment,” Blood, vol. 122, no. 5, pp. 749–758, 2013.

[47] P. C. Rodriguez, M. S. Ernstoff, C. Hernandez et al., “Arginase I-producing myeloid-derived suppressor cells in renal cell carcinoma are a subpopulation of activated granulocytes,” Cancer Research, vol. 69, no. 4, pp. 1553–1560, 2009.

[48] R. Rotondo, G. Barisione, L. Mastracci et al., “IL-8 induces exocytosis of arginase I by neutrophil polymorphonuclears in nonsmall cell lung cancer,” International Journal of Cancer, vol. 125, no. 4, pp. 887–893, 2009.

[49] P. C. Rodriguez, D. G. Quiceno, J. Zabaleta et al., “Arginase I production in the tumor microenvironment by mature myeloid cells inhibits T-cell receptor expression and antigen-specific T-cell responses,” Cancer Research, vol. 64, no. 16, pp. 5839–5849, 2004.

[50] V. Bronte and P. Zanolatto, “Regulation of immune responses by L-arginine metabolism,” Nature Reviews Immunology, vol. 5, no. 8, pp. 641–654, 2005.

[51] C. de Santo, P. Serafini, I. Marigo et al., “Nitroaspirin corrects immune dysfunction in tumor-bearing hosts and promotes tumor eradication by cancer vaccination,” Proceedings of the National Academy of Sciences of the United States of America, vol. 102, no. 11, pp. 4185–4190, 2005.