Identification of Bacterial Hydrocarbonoclastic in Waste Tanks, Petapahan, Riau, Indonesia, using 16srRNA

Irda Sayuti¹, Yusni Ikhwan Siregar¹, Bintal Amin¹, Anthoni Agustien² and Akmal Djamaan³

¹Department of Environmental Science, University of Riau, Riau, Indonesia.
²Department of Biology, Faculty Mathematic and Natural Science, Andalas University, Padang-West Sumatra, Indonesia.
³Laboratory of Biotechnology Biotan Sumatra/Faculty of Pharmacy, Andalas University, Padang-West Sumatra, Indonesia.

http://dx.doi.org/10.22207/JPAM.12.2.25

(Received: 04 March 2018; accepted: 20 April 2018)

This research was conducted to identify the bacterial hydrocarbonoclastic in waste tanks, Petapahan, Riau, Indonesia. Bacteria hydrocarbonoclastic are bacteria that have traffic in degrade petroleum. Analysis of 16S rRNA used the primers pair 63 Forward and Reverse 1387 produced 1300 bp. Results of the analysis of the 16S rRNA sequence in bacterial isolates obtained that IMB-09 09 was similar to Pseudomonas tolsii, IMB-10 was similar to Bacillus cereus, IMB 11 had similarities with Bacillus toyonensis strain BCT-7112, IMB 12 was Lysinibacillus fusiform strain NBRC 15717, IMB 15 was Pseudomonas stutzeri strain ATCC 17588 entirely did have the ability to degrade petroleum.

Keywords: 16S rRNA, hydrocarbonoclastic, bacteria, waste tank, petroleum

As a source of energy, oil and gas have many benefits, it is quite efficient and economical as well as it exists is quite abundant, but when it is spilled or lost to the environment, the oil will be impurities that can become harmful pollutants. Petroleum includes hazardous materials and toxic waste. Petroleum pollution can come from drops and spills of petroleum during the activities of drilling, production, refining, and transportation of oil, resulting in the disruption in the balance of ecosystems, land, water or sea¹. One of the contaminants that are difficult to render is the hydrocarbon compounds derived from petroleum or petroleum sludge. These compounds can be toxic if accumulated in the cell.

Biodegradation of hydrocarbons such as petroleum compounds usually require the cooperation of more than one species of bacteria, it is because petroleum is formed from many different hydrocarbon compounds and bacteria can only use hydrocarbon in a specified range. The difference in the ability of the bacteria in the use of hydrocarbon compounds can be used to maximize the biodegradation process, therefore, the ability of a characterization of bacteria can degrade the hydrocarbon compounds became very important to do²,³.

The process of degraded petroleum start from inoculated the bacterias and take interaction with oil. Processing degradation was occure in aerobic conditions. Degradation petroleum in aerobic more quick than in aerobic conditions, because bacteria produce more energy compare anaerobe reactions⁴. Interaction between microbe and hydrocarbon by adhetion or emulsification for eliminate anionic heteropolysacarade at wall cell or capsule. All of precess ware activated by oxigenase enzyme.

Petroleum degradation be occure at peripheral to transform hydrocarbon become
intermediat compounds such as asetile CoA. CoA was going to come to threecarbocsilate cycle and produce Co$_2^+$ H$_2$O and energy for growth$^5$. Many factors take impact in petroleum degradation such as petroleum chemistry compounds, microbials community, themperature, oxigen, pH and nutrients.

**MATERIAL AND METHODS**

**Genomic DNA Isolation Using Kit invitrogen**

All bacteria samples had the ability to degrade petroleum waste, will be tested and analysis of the 16S rRNA(Figure 1). this method had to mixture all samples in the best liquid media. Nutrient Broth (NB) during the 16 hours at 28 ° C with 120 rpm. Genomic extraction using DNA purification KIT Wizard (Invitrogen) follows the KIT instruction: the first stage of Digestion: 1 ml bacterial suspension 1.5 ml microtube put into the centrifuge bacterial suspension with a speed of 5000 rpm for 5 minutes. Add lysozyme digestion buffer as much as 180 µl vortex, incubation temperature 37°C for 30 minutes. Next add 20 µl proteinase K, vortex and then added 200 µl of Lysis buffer and then bindings genomic vortex and incubated for 30 minutes at a temperature of 55°C, absolute ethanol added as many as 200 µl and vortex.

The second stage of binding: put spin Kulum into the tube, pour the liquid from the binding process into spin Kulum and centrifuge with a speed of 10,000 rpm for 2 minutes, remove the collection tube, replace with a new one. The third stage of washing: Add 500 µl wash buffer I, centrifuge with a speed of 10,000 rpm for 2 minutes, remove the collection tube, and replaced with a new one. Add 500 µl wash buffer 2, centrifuges with speed 12,000 rpm for 3 minutes, then taken spin Kulum, enter into a sterile 1.5 ml micro tube. The fourth stage Elusi: add 200 µl genomic elusion buffer, leave for 1 minute centrifuge with speed 12,000 rpm for 3 minutes, remove spin Kulum, next electrophoresis agarose gel with 1.5% and see in UV light.

**DNA amplification**

Amplification using 16s rRNA Primer from Promega, stages that are conducted in accordance with the instructions of the manufacturer of the kit Protocol. DNA had extracted (Eppendorf, Westbury, NY) for 15 µl of 100 minutes then added DNA rehydration (Promega). Amplification of the 16S gene rRNA had done in 50µl containing 10 pmol 63f primer (5'-AGAGTTTGATC(A/C) TGGCTCAG-3') and 1387r (5'-GG (C/T) TACCTTGTTACGACTT-3'). Around of 25 µl green tag Promega, DNA template 3 µl (10 ng/ µl), the addition of a nuclease-free water 20 µl of PCR Amplification cycle was 35. beginning with the denaturation temperature was 94°C for 3, then proceed with stage 94°C denaturation for 1 minute. 55°C temperature Annealing lasted for 30 seconds, the expansions for 30 seconds at a temperature of 72°C and end satthe end of the expansions at72°C for 5 seconds. PCR cycle last sasmuchas 35cycles, 3 hours. 7 µl of PCR results are examined by gel electrophoresis. Visualization of results of PCR was performed on agarose electrophoresis 75 grin 50 ml of 0,5TBE.

**RESULTS AND DISCUSSION**

**Genomic DNA Isolation Using KIT INVITROGEN**

The genomic DNA isolation using DNA isolation kit from INVITROGEN beginning from the taking of the pellet suspension of bacteria in culture liquid (Nutrient broth) using the centrifuge. Isolation of DNA results can be seen from Figure 1 below.

Each sample of DNA isolation result showed the results good enough, this is because the isolation technique that is used is the isolation techniques already adopted from the isolation kit used, namely INVITROGEN. Samples of IMB and IMB-10 shows the results of isolation are not good, look at the picture that the very thin ribbon, possibly this is due to a number of pellets obtained from bacterial suspensions IMB and IMB-10 bit. The results of this DNA isolation continued to amplify with a PCR.

**Amplified genomic DNA with primer 16s rRNA**

Amplification Genomic DNA with the primer 16s rRNA Amplification using PCR machine Thermo. The PCR cycle run is 35 cycles. The results of PCR with primer 63F and 1387R can be seen in Figure 2 below.

The results of PCR using primer 16s rRNA couples (63F and 1387R) indicates the length of the product PCR obtained i.e. 1,300 BP. Ribbon-Ribbon amplicons result from PCR primer pairs
match showed 63F and 1387R against bacterial genome samples are used. The intensity of the PCR product obtained varies in each sample, samples of IMB-9 and IMB-10, this is due to the concentration of the template used appropriate low, due to the thin insulation results (Figure 10).

Samples IMB-09, IMB-10, IMB-11, IMB-12, IMB and IMB-15 show the results of the nice amplicon and assertive. It also had the same correlations with genomic DNA isolation results obtained as a template on the activities of the PCR.

This bacteria had capability to degraded compound in human body including aromatic hydrocarbon. Inhibition mushroom fungi pathogens.

Phylogenetic analysis shows that the IMB-10 sample was very similar to the 16s sequencer RNA with Bacillus cereus with value NR 074540.1. Analysis was done using Phylogenetic analysis shows rRNA with Bacillus cereus with value NR 074540.1. Analysis using bootstrap 1,000, this figure states that the sequence stability rate will be estimated use this analysis. Bacillus cereus does have ability to degrade petroleum. The influence of oil contamination the earth in the water will change the response of the membrane bacterial cells thus affecting absorption and the need for bacterial oxygen, so influential to the degradation process. Bacillus cereus a lot found in wastewater tanks at refinerie soil.

*Bacillus cereus* had reported could degradation the phenolic compound in petroleum until 95% and more. *Bacillus cereus* strain JMG-01 in enhanced anthracena degradation along the utilization of other hydrocarbons. Another reported, Bacillus cereus can remove contaminated petroleum in soil or about 30 days.

IMB 11 had similarities with Bacillus toyonensis strain BCT-7112. Bacillus toyonensis had been reported that it is able to survive in areas that are contaminated by petroleum. Petroleum use as carbon sources. Analysis of the sequence of the bases of a sample of IMB-11 with all isolates of bacteria from the Blast using a program bio edit and phylogenetic tree analysis using the programe MEGA6. The discovery two isolates that can be degrade petroleum and the two isolates utilized the source petroleum as the only source carbon, as...
for the two bacterial isolates. Bacteria degrade petroleum and obtained type of bacteria 94% similarity for Bacillus thuringiensis and Bacillus bombysepticus, and 95% similarity for Bacillus toyonensis BCT-711213,14.

16s rRNA sequence of samples of IMB-12 had similarities with Lysinibacillus fusiform strain NBRC15717 obtained from the results of the blast. This bacteria was reported to be used to protect the hulls of the activities of bio fouling17. From the results of research that had been done that the data obtained, a sample of IMB-12 has the ability to degrade petroleum of 62.61%. Data that had been retrieved it states that Lysinibacillus fusiform also had the ability to degrade the oil, which can be applied in the future to handle cases of environmental pollution due to oil spills.

Obtained that samples of IMB-15 had similarities with Pseudomonas stutzeri strain ATCC 17588. Samples of the IMB-15 has the ability to degrade petroleum valued at 76.63%. Pseudomonas stutzeri had been reported to have the ability to degrade petroleum (Celik et al., 2008). These bacteria were also reported to have the same capabilities, it has been reported by (Sazinsky et al., 2004). Researched by Lalucat et al., (2006) this bacteria opportunity pathogen of human and Park et al., (2013) according this bacteria have been reported in patients undergoing continuous ambulatory peritoneal dialysis (CAPD).

Over all according to the isolation and identification, bacteria degradation petroleum

Fig. 3. Phylogenetic tree sample IMB-09 using Mega-6 program.

Fig. 4. Phylogenetic tree sample IMB-10 using Mega-6 program.
in waste tank show that bacteria isolate can obtain nutrients from hydrocarbon in waste tank via degradation process. This process comes from hydrocarbon was degraded by peripheral pathway by oxygenase enzyme to become a simple compounds like acetate CoA. Five bacteria found in waste tank can survive because their can degrade hydrocarbon becoming food for life. Microbial community had big impact for degrad petroleum.

**Fig. 5.** Phylogenetic tree sample IMB-11 using Mega-6 program

**Fig. 6.** Phylogenetic tree sample IMB-12 using Mega-6 program

**Fig. 7.** Phylogenetic tree sample IMB-15 using Mega-6 program
CONCLUSIONS

Fragment of PCR results obtained 16SrRNAs were 1300bp. Blast shown samples of the IMB-09 was similar to Pseudomonas tolaasii, IMB-10 was similar to Bacillus cereus, IMB-11 was similar to Bacillus toyonensis, IMB-12 was similar to the sample and the fusiform Lysinibacillus fusiformis and IMB-15 Pseudomonas stutzeri. all bacterias have the ability to degrade petroleum.

REFERENCES

1. Chaan, B. Biodegradasi Kimia Organik 2013. http://chaan-biodegradasi-senyawa-organik.html.
2. Hajar, D. 2012. Islation, Identification and Isolasi, Identifikasi and Degradation Capability Analysis Hydrocarbon Bacteria Soil Samples B, Cilegon, Banten. [Essay]. Depok. University Indonesia.
3. Milic, J. S., Beskoski, V. P., Ilic, M. V., Ali, S.A. M., Gojgic-Cvijovic, G. D., and Vrvic, M.M. Bioremediation of soil heavily contaminated with crude oil and its product: composition of the microbial consortium. Journal of the Serbian Chemical Society. 2009; 74(4): 455–460.
4. Malatova, K. 2005. Isolation and Characterization of Hydrocarbon Degrading Bacteria from Environmental Habitats In Western New York State Department of Chemistry Rochester Institute of Technology. New York: xi+97hlm.
5. Fritsche, W and Hofrichter, M. 2000. Aerobic Degradation by Microorganisms In: H. Rehm and G. Reed (eds). In Biotechnology: Environmental Habitats In Western New York State Department of Chemistry Rochester Institute of Technology, New York: xi+97hlm.
6. Hesham Ael L, Mawad AM, Mostafa YM, Shoreit A. 2014. Biodegradation ability and catabolic genes of petroleum degrading Sphingomonas koreensis strain ASU-06 isolated from Egyptian oily soil. Biomed Res Int: 127674. doi:10.1155/2014/127674
7. Cantore, P. L., Giorgio, A., Lacobellis, N. 2015. Bioactivity of volatile organic compounds produced by Pseudomonas tolaasii. Journal Frontiers In Microbiology: 6. doi: 10.3389/fmicb.01082.
8. Benerjee, A., Ghosh, A. K. 2016. Biodegradation of Real Petroleum Waste water by Immobilized Hypee Phenol- Tolerant Strain of Bacillus cereus in A Fluidized Bed Bioreactor. Journal Soil Sediment Contamination, 2017; 26(5).
9. Agarry S.E., Aremu M.O., Aworanti O.A. 2013. Biodegradation of 2, 6-dichlorophenol wastewater in soil column reactor in the presence of pineapple peels-derived activated carbon, palm kernel oil and inorganic fertilizer. J Environ. Protect. 4: 537–547.
10. JDas,M., Bhattacharya, A., Bamu,S., Kotoky, Enhanced Biodegradation of Anthracene by Bacillus cereus Strain IMG-01 Isolated from Hydrocarbon Contaminated Soils. Journal Soil Sediment Contamination, 2017; 26(5).
11. Maddela, N.R., Burgos,R., Kadiyala,V., Camon, A. R. Removal of Petroleum Hydrocarbons from Crude Oil In Solid and Slurry Phase By Mixed Soil Microorganisms Isolated From Ecuadorian Oil Fields. Journal International Biodeterioration and Biodegradation. 2016; 108: 85–90.
12. Djamaan, A., Mhd Riza Marjoni, and Friardi Ismed. The Influence of Pretreatment Time, Type and the Concentration of Yeast on Ethanol Production from Rice Straw. Research Journal of Pharmaceutical, Biological and Chemical Sciences. 2015, 6(3): 583-591.
13. Gayathri E, Bharathim B, Natarajan S. 2014. Isolation, identification and characterization of diesel degrading bacteria. International Journal of Current Research; 6(6): 7295-300.
14. Janssens, T.K.S., Boer, T.E. De., Anamennone, V., Zaalman, N. M., Kraai, M. V., Roelofs, D. 2016. Draft Genome Sequence Of Bacillus toyonensis VU-DES13, Isolated From Folsomia Candida (Collembola:Entomobrydae). Journal American Society for Microbiology.
15. Jiménez N., Morris B. E. L., Cai M., Grnüdger Donato ,and Stephen J Lippard, Crystal Structure and Chemical Sciences, Research Journal of Pharmaceutical, Biological and Chemical Sciences, 2017; 26(3): 583-591.
16. Kumar, Arun, and Pradhan Nilotpala. 2013, Inhibition of Pathogenic Bacterial Biofilm by Biosurfactant Produced by Lysinibacillus Fusiformis S9."doi:10.1007/s00449-013-0976-5.
17. Rivai, H, Asia A, Rina W, Alen Y. Handayani D, Aldi Y, Marlina, and Djamaan A., Isolation of Endophytic Bacteria from Bark, Leaf, and Pericarp of Mangosteen (Garcinia mangostana L.) and Testing of The Antimicrobial Activity, Research Journal of Pharmaceutical, Biological and Chemical Sciences, 2016, 7(1): 1910-1920
18. Sazinsky, Matthew H, Joel Bard, Alberto Di Donato, and Stephen J Lippard, Crystal Structure of the Toluene / O -Xylene Monooxygenase Hydroxylase from Pseudomonas Stutzeri OX1.
19. Lalucat, J., Bennasar, A., Bosch, R., Valdes, E. G., Palleroni, N. Biology of Pseudomonas stutzeri. *Journal Microbiology and Molecular Biology Reviews*. 2006; 7(2): 510-547.

20. Subhraveti, P., Ong, Q., Keseler, I., Kothari, A., Caspi, R., Karp, P. D. Summary of Bacillus toyonensis, Strain BCT-7112, Version 21.5. BioCYC Database Collection 2016.

21. Park, S. W., Lee, S. W., Shin, C. H., Kim, G. E., Kim, M. J. Successful Antibiotic Treatment of Pseudomonas stutzeri Induced Peritoritis without Peritoneal Dialysis Catheter Removal In Countinous Ambulatory Peritoneal Dialysis. *Journal Kidney Research and Clinical Practice*. 2013; 32(2): 81-83.

22. Zam, S. I. Bioremediation of Petroleum Refining Waste Pertamina UP II Pakning River by Using Indigenous Bacteria. Thesis. Bandung Institute of Technology. Bandung, 2006