Metformin in Combination with Malvidin Prevents Progression of Non-Alcoholic Fatty Liver Disease via Improving Lipid and Glucose Metabolisms, and Inhibiting Inflammation in Type 2 Diabetes Rats

Wenlan Zou 1,*
Chen Zhang 1,*
Xuefang Gu 2
Xiaohong Li 3
Huiming Zhu 4

1Department of Endocrinology, Fifth People’s Hospital of Suzhou, Suzhou, Jiangsu, 215100, People’s Republic of China; 2Department of Endocrinology, Xiangcheng District Second People’s Hospital of Suzhou, Suzhou, Jiangsu, 215100, People’s Republic of China; 3Department of Liver Disease, Fifth People’s Hospital of Suzhou, Suzhou, Jiangsu, 215100, People’s Republic of China; 4Department of Gastroenterology, Fifth People’s Hospital of Suzhou, Suzhou, Jiangsu, 215100, People’s Republic of China

*These authors contributed equally to this work

Methods: Sprague-Dawley rats were divided into five groups: normal control group (NC), diabetic control group (DC), DC+MET group, DC+MAL group, and DC+MET+MAL group and treated for eight weeks. Blood and liver tissue samples were collected for metabolic parameters, histological, and RT-qPCR analysis.

Results: Our findings indicated that hyperglycemia, insulin resistance, hyperlipidemia, and non-alcoholic fatty liver disease (NAFLD) in diabetic rats were alleviated after oral treatment with MET and MAL, particularly their combination therapy. Besides, the expression of SREBP-1c, ACC, FAS, IL-6, IL-8, and NF-κB mRNA was down-regulated by MET+MAL, and the expression of PPARα, CPT1, and LPL was up-regulated by MET+MAL.

Conclusion: The evidence of this research indicated that the combination therapy may represent an efficient strategy against NAFLD in T2DM rats via improving lipid and glucose metabolisms, and inhibiting inflammation.

Keywords: non-alcoholic fatty liver disease, diabetes mellitus, combination, lipogenesis, malvidin, inflammation

Introduction
Type 2 diabetes mellitus (T2DM) is a chronic metabolic disorder that is characterized by persistent hyperglycemia due to the impaired islet cell function and is considered a serious global health problem. 1,2 T2DM causes not only high blood glucose, but also non-alcoholic fatty liver disease (NAFLD), diabetic nephropathy, and dyslipidemia. 3 NAFLD is defined by excessive deposition of fat in hepatocytes, which is closely linked to several metabolic diseases, including insulin resistance, T2DM, and dyslipidemia. 4,5 Additionally, a previous report indicated that the prevalence of NAFLD in conjunction with diabetes and obesity. T2DM may be a major risk factor for the progression from steatosis to steatohepatitis, even cirrhosis. 6 Furthermore, NAFLD is rapidly becoming the primary cause of end-
stage liver disease and hepatocellular carcinoma. Thus, in long-term treatment, alleviating the complications of T2DM is just as important as lowering blood glucose levels. A growing number of the report indicated that persistent hyperglycemia affects the inflammatory mechanisms of the liver, which in turn promotes liver damage in aspects of liver fibrosis and lipid accumulation. Additionally, previous reports have demonstrated that anti-inflammatory compounds could alleviate liver damage, control blood glucose, and improve insulin resistance. Thus, therapeutic suppression of hepatic inflammation may be an effective treatment for T2DM and its metabolic syndrome.

Metformin (MET) is the most prescribed oral anti-diabetic drug and derived from plant Galega officinalis. It has been reported that MET could decrease hepatic lipid levels in T2DM patients, MET also inhibited the inflammatory response in non-alcoholic steatohepatitis via STAT3-mediated autophagy induction. Despite the health benefits of MET, previous studies have reported several side effects of MET, including diarrhea, lactic acidosis, and gastrointestinal disorders. Therefore, researchers have proposed the combination therapy to eliminate the side effects of oral anti-diabetic drugs and also to enhance its effectiveness, such as MET combined with other herbal agents.

Recent researches on natural products, especially anthocyanins, have focused on preventing NAFLD and T2DM. For example, the anthocyanins extract from black soybean seed coat was shown to hypolipidemic and hypoglycemic effects in T2DM, it has been reported that an anthocyanin-rich berry could improve insulin sensitivity in obese and overweight populations, bilberry anthocyanins were shown to alleviate western diet-induced NAFLD via improving gut microbiome dysbiosis and hyperlipidemia. Malvidin (MAL) is an anthocyanins compound belonging to a subgroup of flavonoids, is highly present in berries, flowers, and red grape skin. It has been extensively investigated due to its anti-inflammatory, anti-tumor, and cardioprotective effects. Additionally, MAL could protect endothelial cells via suppressing peroxynitrite-induced NF-κB activation; MAL was shown to inhibit hepatocellular carcinoma via regulating metastasis, proliferation, and apoptosis. However, there were no reports on the effects of MET combined with MAL on NAFLD. Therefore, the present study aimed to explore the therapeutic effects of MET or MAL alone or in combination on hyperglycemia, insulin resistance, hyperlipidemia, and NAFLD in diabetic rats, and we hypothesized that MET in combination with MAL could alleviate NAFLD via improving lipid and glucose metabolisms, and suppressing inflammation.

Materials and Methods
Animals and Treatments
Male Sprague-Dawley rats (6–8 weeks old, 180–220 g) were procured from the Model Animal Research Center of Nanjing University (Nanjing, China). All rats were housed in an experimental animal center of Suzhou Fifth People’s Hospital at 50 ± 5% humidity and 20 ± 2°C under a 12/12 h light/dark cycle with a freely available diet and water. The protocol involving animals was approved by the Ethics of Animals Experiments Committee of Suzhou Fifth People’s Hospital (approval number: 2019011B) and performed following the National Standard (GB/T 35892–2018) for Laboratory Animals Care and Use.

After seven days of acclimation, except for the normal control group, other group rats were administrated with a high-fat diet (HFD) for 12 weeks. HFD consisting of 0.5% sodium cholate, 3% cholesterol, 10% lard, 20% sucrose, and 66.5% standard chow diet. After six weeks of HFD, DC, DC+MET, DC+MAL, and DC+MET+MAL groups were fasted overnight and intraperitoneally injected with a single dose of STZ (35 mg/kg body weight) in freshly citrate buffer, while NC group rats were intraperitoneally injected with an equivalent volume of citrate buffer. After one week of STZ injection, the fasting blood glucose levels were assayed, and glucose levels >11.1 mmol/L were considered diabetic. The animals were randomly divided into five groups (n=10 per group). Normal control group (NC): rats received standard chow diet neither drug treatment nor diabetes induction; diabetic control group (DC): rats received HFD/STZ and no drug administration; MET protection group (DC+MET): rats received HFD/STZ and administrated with MET (100 mg/kg body weight); MAL protection group (DC+MAL): rats received HFD/STZ and administrated with MAL (100 mg/kg body weight); MET +MAL protection group (DC+MET+MAL): rats received HFD/STZ and administrated with the combination of MET (100 mg/kg body weight) and MAL (100 mg/kg body weight). From week 0 to week 12, the MET+MAL or MET or MAL or vehicle (DMSO) was treated to rats by oral gavage daily. The dosage of MET or MAL used was based on previous reports. The body weight of rats was recorded throughout the experiment. The detailed experimental scheme is shown in Figure 1A.
experiment, all rats were anesthetized with pentobarbital (30 mg/kg body weight) after fasting overnight. Then, the blood sample was collected from inferior vena cava and centrifuged at 4°C for 15 min at 3500 × g, and the supernatant was collected and stored at −20°C for further analysis. The liver tissues were quickly collected and washed with physiological saline, and weighted. The liver tissue samples were fixed in 10% paraformaldehyde solution for histopathological assessment or stored at −20°C for further analysis.

Oral Glucose Tolerance Test (OGTT) and Insulin Tolerance Tests (IPITT)
One day before the termination of the experiment, the rats fasted overnight. The rats were received glucose (2 g/kg body weight) by oral gavage for OGTT. The rats were injected intraperitoneally with insulin (2 units/kg body weight) for IPITT. Blood samples were obtained from the tail vein over the time course of 0 to 120 min and the blood glucose levels were measured using commercial kits.

Serum Biochemical Analysis
The levels of interleukin 6 (IL-6), IL-8, blood glucose, insulin, leptin, triglyceride (TG), total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol (HDL-C), aspartate aminotransferase (AST), and alanine aminotransferase (ALT) in serum were measured using commercial kits (Jiancheng Bioengineering Institute, China) based on the manufacturer’s protocol. The item number of the corresponding kits was as follows: PEPCK (A131-1-1), G6Pase (H580-1), GS (H388-1), IL-6 (H007), IL-8 (H008), and NF-κB (H202).

Histopathology
The liver tissues were washed with physiological saline and then fixed with polyformaldehyde (4%) for 24 h. Then, the tissue samples were embedded in paraffin and cut into slices with a thickness of 4 µm. These sections were deparaffinized in xylene and rehydrated with a series of concentrations of ethanol. After washing with physiological saline, these sections were incubated in Oil Red O reagent (Jiancheng Bioengineering Institute, China) based on the manufacturer’s protocol. Additionally, the liver sections were examined by hematoxylin and eosin staining (H&E). The stained sections were observed under a light microscope (Olympus, Tokyo, Japan).

Quantitative Real-Time PCR
Total RNA of liver tissues was extracted using the Trizol reagent (Invitrogen, Carlsbad, CA, USA). 2 µg of extracted RNA was reverse transcribed into cDNA by the Prime Script RT reagent Kit (Takara, Shiga, Japan). Then, quantitative real-time PCR was carried out using the SYBR Green Master Mix (Thermo Fisher, USA) on an Applied Biosystems StepOnePlus System (Thermo Fisher, USA). The primer sequences used for measuring gene expression were shown in Supplementary file (Table 1S). The qPCR was carried out in duplicate, and the gene expression was reported as $2^{-\Delta\Delta C_t}$. The GAPDH is used as the internal control.

Statistical Analysis
The data were presented as mean ± SD and analyzed using the GraphPad Prism 6.0 software (San Diego, CA, USA). Statistical differences among multiple groups were evaluated by one-way analysis of variance (ANOVA), followed by Dunnett’s Multiple Comparison Test. It was considered statistically significant when $P < 0.05$. 

Drug Design, Development and Therapy downloaded from https://www.dovepress.com/ by 207.241.232.188 on 18-Jun-2021
For personal use only.
Results

General Effects of Combined Drug Therapy on HFD/STZ-T2DM Model Rats

The diabetic rats exhibited typical diabetic symptoms, including emaciation, polyphagia, and polydipsia. We observed these diabetic symptoms in the DC group. The body weight of rats in HFD groups increased continuously during the first 6 weeks of the experiment. However, the body weight of rats in the HFD/STZ group declined obviously after injection of STZ (Figure 1B). The final body weight of HFD/STZ rats was lower than that of NC rats (Figure 1C), while the food consumption and water consumption of HFD/STZ rats were higher than that of NC rats (Figure 1F and G). The combination of MET and MAL alleviates these diabetic symptoms in the DC group ($P < 0.01$). Besides, the liver weight and liver index of HFD/STZ rats were higher than that of NC rats (Figure 1D and E). The combination of MET and MAL decreased the liver weight, and liver index of rats compared to the DC group ($P < 0.01$).

Combined Drug Therapy Alleviated Insulin Resistance

As shown in Figure 2, treatment with MET or MAL alone obviously decreased glucose, and insulin intolerance (AUC) compared with the DC group ($P < 0.001$). Interestingly, the reduction in the DC+MET+MAL was higher than that of the MET or MAL alone. Besides, insulin resistance was evoked in the DC group as displayed by hyperglycemia, hyperinsulinemia, and increased HOMA-IR as compared to the NC group (Figure 3A–C). Treatment with MET or MAL alone decreased fasting blood glucose level, serum insulin level, and HOMA-IR compared with the DC group. Interestingly, the decline in the DC+MET+MAL was higher than that of the MET or MAL alone. These findings indicated that the combined drug therapy alleviated glucose and insulin intolerance in T2DM model rats.

Combined Drug Therapy Ameliorated HFD/STZ-Induced Adverse Changes in Serum Biochemical Markers

As shown in Figure 3D–G, HFD/STZ resulted in an obvious increase in the serum levels of leptin, TC, TG, and LDL-C compared to those in the NC group. These parameters were reduced in T2DM rats administered with MET or MAL alone. Importantly, the reduction in the DC+MET+MAL was higher than that of the MET or MAL alone. HFD/STZ also resulted in a significant decrease in the serum levels of HDL-C compared to that in the NC group (Figure 3H). The HDL-C levels were increased in T2DM rats administered with MET or MAL alone. Importantly, the combination of MET and MAL caused an increase in the serum levels of HDL-C as compared to MET or MAL alone treated groups.

Combination of MET and MAL Alleviated NAFLD in HFD/STZ-Induced T2DM Rats

The liver function indexes, including serum ALT and AST, were observed to be increased in the DC group (Figure 4A

![Figure 1](https://example.com/figure1.png)

**Figure 1** Effects of MET, MAL, and MET+MAL on T2DM related characteristics in HFD/STZ-T2DM model rats. (A) The detailed experimental scheme; (B) Body weight changes during the experiment; (C) Final body weight; (D) Liver weight; (E) Liver index; (F) Food consumption; (G) Water consumption.
These liver function indexes were declined in T2DM rats administered with MET or MAL alone. Importantly, the combination of MET and MAL led to a decrease in the serum levels of ALT and AST as compared to MET or MAL alone treated groups. Besides, H&E staining showed regular cell morphology, uniform cytoplasm, and clear liver lobules structure in the NC group under a light microscope. In the DC group, hepatocytes were swollen and disordered arrangement; lipid vacuoles were widely distributed in the hepatocytes; there was obvious infiltration of inflammatory cells. MET, MAL, and combination of MET and MAL all alleviated these liver pathological changes in T2DM rats, and the effect on the combined group was greater.

Hepatic levels of TC and TG were increased by HFD/STZ and declined by treatment with MET or MAL alone. Interestingly, the decline in the DC+MET+MAL was higher than that of the MET or MAL alone and D). The Oil red O staining was performed to further evaluate the effects of combined drug therapy on hepatic lipid accumulation (Figure 4E). MET, MAL, and combination of MET and MAL all decreased hepatic neutral lipids in T2DM rats, and the effect on the combined group was greater.

The Combination of MET and MAL Inhibited the Expression of Hepatic Genes Related to Lipogenesis

As shown in Figure 5A–C, the expression of genes involved in lipogenesis (SREBP-1c, ACC, and FAS) were measured by RT-qPCR to further investigate the underlying mechanisms of combined drug therapy regulated hepatic fat accumulation on HFD/STZ-induced T2DM rats. The expression of SREBP-1c, ACC, and FAS was higher in the hepatic tissues of the DC group compared with the NC group. Treatment with MET or
Figure 3 Effects of MET, MAL, and MET+MAL on serum biochemical parameters in HFD/STZ-T2DM model rats. Insulin resistance relative marker, including fasting blood glucose (A), insulin (B), HOMA-IR (C), and leptin (D). Lipid metabolism parameters, including serum TC (E), serum TG (F), serum LDL-C (G), and serum HDL-C (H). The data were presented as mean ± SD, (n = 8 for all groups). ***P < 0.001; **P < 0.01; *P < 0.05.

Figure 4 Effects of MET, MAL, and MET+MAL on NAFLD in HFD/STZ-T2DM model rats. (A) The activity of ALT in serum of rats; (B) The activity of AST in serum of rats; (C) Hepatic levels of TC; (D) Hepatic levels of TG; (E) Lipid accumulation in the hepatic tissue of rats was evaluated by Oil red O staining, the scale bar is 200 µm; (F). Representative photos of H&E staining from liver tissue, the scale bar is 100 µm. The data were presented as mean ± SD, (n = 8 for all groups). ***P < 0.001; **P < 0.01.
MAL alone significantly down-regulated these hepatic genes involved in lipogenesis. Interestingly, the downregulation in the DC+MET+MAL was higher than that of the MET or MAL alone.

**Combination of MET and MAL**

**Up-Regulated Expression of Hepatic Genes Involved in Fatty Acid Oxidation and Lipid Metabolism**

As shown in Figure 5D–F, the gene expression involved in fatty acid oxidation (PPARα and CPT1) and lipid metabolism (LPL) were down-regulated by HFD/STZ and up-regulated by treatment with MET or MAL alone. Interestingly, the upregulation in the DC+MET+MAL was higher than that of the MET or MAL alone.

**The Combination of MET and MAL Relieves Hepatic Inflammation Induced by HFD/STZ**

The serum and hepatic levels of IL-6, IL-8, and NF-κB were increased in the DC group compared to the NC group (Figures 6 and 7). Treatment with MET or MAL alone decreased IL-6, IL-8, and NF-κB levels induced by HFD/STZ. Interestingly, the reduction in the DC+MET+MAL was higher than that of the MET or MAL alone. Besides, the gene expression of IL-6, IL-8, and NF-κB in hepatic tissues was measured by RT-qPCR also to further demonstrate that hepatic inflammation caused by HFD/STZ was inhibited by the combination of MET and MAL (Figure 8). These results indicated the potency of the combination of MET and MAL in inhibited hepatic inflammation in HFD/STZ-induced T2DM rats.

**Effect of Combined Drug Therapy on the Activities of Enzymes Related to Glucose Metabolism**

The activities of glycogen synthesis- and gluconeogenesis-related enzymes were measured to explore the effects of combined drug therapy on hepatic glucose metabolism. As shown in Figure 9, the activities of G6Pase and PEPCK in the DC group were significantly higher than those in the NC group. Treatment with MET or MAL alone decreased G6Pase, and PEPCK activities induced by HFD/STZ. Interestingly, the decrease in the DC+MET+MAL was higher than that of the MET or MAL alone. Besides, the
**Figure 6** Effect of combined drug therapy on serum inflammatory cytokines in HFD/STZ-T2DM model rats. The serum levels of IL-6 (A) and IL-8 (B) of rats were measured by ELISA. The data were presented as mean ± SD, (n = 8 for all groups). ***P < 0.001; **P < 0.01; *P < 0.05.

**Figure 7** Effect of combined drug therapy on hepatic inflammation in HFD/STZ-T2DM model rats. The levels of IL-6 (A), IL-8 (B), and NF-κB (C) in hepatic tissue of rats were measured by ELISA. The data were presented as mean ± SD, (n = 8 for all groups). ***P < 0.001; **P < 0.01; *P < 0.05.

**Figure 8** Effect of combined drug therapy on hepatic gene expression involved in inflammation. The relative expression of hepatic genes involved in hepatic inflammation, including IL-6 (A), IL-8 (B), and NF-κB (C). The data were presented as mean ± SD, (n = 6 for all groups). ***P < 0.001; **P < 0.01; *P < 0.05.
activity of GS in the DC group was significantly lower than those in the NC group. Treatment with MET or MAL alone increased GS activity induced by HFD/STZ. Interestingly, the increase in the DC+MET+MAL was higher than that of the MET or MAL alone.

Discussion

Increased incidence of metabolic syndromes, and T2DM due to lack of exercise and excessive intake of HFD, as a lipid metabolic disease, NAFLD is considered the most common chronic liver disease worldwide, with no approved pharmacotherapy up to date. Thus, it is necessary to explore novel therapy for NAFLD. The current report aimed to investigate the beneficial effects of the combination of MET and MAL in a T2DM rat model.

A T2DM rat model with NAFLD was established by HFD/STZ in the present study. Consistent with previous researches, the DC rats exhibited significant increases in fasting blood glucose, serum TC, and TG; coupled with glucose and insulin intolerance. Besides, hepatic lipid accumulation was found in the DC group, which is characteristic of NAFLD. Interestingly, the combination of MET and MAL not only reduced fasting blood glucose, serum insulin, serum lipid profiles but also alleviated insulin sensitivity. Additionally, we also observed decreased hepatic lipid accumulation in the MET+MAL group. These findings indicated that combined drug therapy could alleviate hyperglycemia, hyperlipidemia, and NAFLD in HFD/STZ-induced T2DM rats.

The previous reports have indicated that the inhibition of lipogenesis and promotion of fatty acid oxidation are important approaches for the suppression of hepatic fat accumulation. It has been reported that hepatic de novo lipogenesis is involved in the progression of NAFLD. SREBP-1c is a transcription factor that regulates the genes involved in hepatic lipogenesis and TG synthesis, including ACC, and FAS. In the current report, the mRNA levels of SREBP-1c, ACC, and FAS were up-regulated by HFD/STZ, resulting in elevations in hepatic TG synthesis. The combination of MET and MAL down-regulated these genes expression, leading to the declines in lipid levels. Besides, PPARα could regulate several key genes involved in fatty acid oxidation and PPARα activation could improve hepatic steatosis and inflammation in NAFLD. CPT-1 is an activator of PPARα expression, which accelerates mitochondrial fatty acid oxidation and uptake of lipid. In the present study, the combination of MET and MAL upregulated PPARα, CPT-1, and LPL expression, causing an increase in fatty acid β-oxidation. These findings indicated that combined drug therapy promoted TG and fatty acid metabolism, resulting in declines in serum and liver lipid levels.

A “two-hit” process has been demonstrated that is involved in the pathogenesis of NAFLD. The hepatic fat accumulation evokes a series of cytotoxic events that result in a hepatic inflammatory response. Besides, accumulating evidence showed that inflammation is closely accompanied by lipid metabolism, is involved in the etiopathogenesis of NAFLD. Pro-inflammatory cytokines, such as IL-6, IL-8, and TNF-α, have been indicated to implicate in the pathogenesis of NAFLD. NF-xB is an important regulator of liver damage and hepatic inflammatory recruitment in steatohepatitis. In the present study, the mRNA levels of IL-6, IL-8, and NF-xB were up-regulated by HFD/STZ, resulting in the elevations in pro-inflammatory cytokines secretion. The combination of MET and MAL has a better effect on alleviating hepatic inflammation in HFD/STZ-induced T2DM rats.

Figure 9 Effect of combined drug therapy on the activities of enzymes related to glucose metabolism. The activities of G6Pase (A), PEPCK (B), and GS (C) involved in glucose metabolism. The data were presented as mean ± SD, (n = 8 for all groups). ***P < 0.001; **P < 0.01; *P < 0.05.
The liver is an essential place for the regulation of normal glucose metabolism. Insulin could suppress hepatic glycogenolysis and gluconeogenesis, and improve glucose metabolism through regulating the activity of glucose metabolism-related enzymes such as G6Pase, PEPCK, and GS under physiological conditions. However, under T2DM conditions, these normal glucose metabolism processes were disordered due to insulin resistance. Our findings showed that a combination of MET and MAL could inhibit the activities of G6Pase and PEPCK, thus inhibiting hepatic gluconeogenesis. Besides, the combination of MET and MAL also could increase the activity of GS, thus promoting glycogenesis. These results indicated that combined drug therapy improved hepatic glucose metabolism via regulating the activities of glucose metabolism-related key enzymes.

Overall, for the first time, we demonstrated that a combination of MET and MAL could alleviate NAFLD via regulating lipid and glucose metabolisms, and inhibiting hepatic inflammation using HFD/STZ-induced T2DM model. The combined drug therapy presents to be more effective in inhibiting lipogenesis, promoting fatty acid oxidation, suppressing inflammatory cytokines secretion, and improving glucose metabolism than MET or MAL alone. Figure 10 indicates the role of MET+MAL in the alleviation of NAFLD. Therefore, this therapy could become a potential alternative therapy in the prevention and treatment of NAFLD.

Disclosure
The authors declare no conflicts of interest in this work.

References
1. GBD 2016 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 328 diseases and injuries for 195 countries, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. Lancet (London, England). 2017;390(10100):1211–1259.
2. Mäkimattila S, Virkamäki A, Groop PH, et al. Chronic hyperglycemia impairs endothelial function and insulin sensitivity via different mechanisms in insulin-dependent diabetes mellitus. Circulation. 1996;94(6):1276–1282. doi:10.1161/01.CIR.94.6.1276
3. Bello NA, Pfeffer MA, Skali H, et al. Retinopathy and clinical outcomes in patients with type 2 diabetes mellitus, chronic kidney disease, and anemia. BMJ Open Diabetes Res Care. 2014;2(1):e000011. doi:10.1136/bmjdr-2013-000011
4. Cheung O, Sanyal AJ. Recent advances in nonalcoholic fatty liver disease. Curr Opin Gastroenterol. 2009;25(3):230–237. doi:10.1097/MOG.0b013e3283294a18
5. Abenavoli L, Milic N, Di Renzo L, Preveden T, Medić-Stojanoska M, De Lorenzo A. Metabolic aspects of adult patients with nonalcoholic fatty liver disease. World J Gastroenterol. 2016;22(31):7006–7016. doi:10.3748/wjg.v22.i31.7006
6. Mantovani A, Zoppini G, Targarher G, Golia G, Bonora E. Non-alcoholic fatty liver disease is independently associated with left ventricular hypertrophy in hypertensive Type 2 diabetic individuals. J Endocrinol Invest. 2012;35(2):215–218. doi:10.1007/BF03345421

7. Liu D, Wong CC, Fu L, et al. Squalene epoxidase drives NAFLD-induced hepatocellular carcinoma and is a pharmaceutical target. Sci Transl Med. 2018;10(437). doi:10.1126/scitranslmed.aap9840

8. Kelley DE, McKolans TM, Hegazi RA, Kuller LH, Kalhan SC. Fatty liver in type 2 diabetes mellitus: relation to regional adiposity, fatty acids, and insulin resistance. Am J Physiol Endocrinol Metab. 2003;285(5):E596–616. doi:10.1152/ajpendo.00117.2003

9. Dharmalingam M, Yamasandhi PG. Nonalcoholic fatty liver disease and type 2 diabetes mellitus. Indian J Endocrinol Metab. 2018;22(3):421–428. doi:10.4103/ijem.IJEM_585_17

10. Zhang C, Lu X, Tan Y, et al. Diabetes-induced hepatic pathogenic damage, inflammation, oxidative stress, and insulin resistance was exacerbated in zinc deficient mouse model. PLoS One. 2012;7(12):e49257. doi:10.1371/journal.pone.0049257

11. Ford RJ, Fullerton MD, Pinkosky SL, et al. Metformin and salicylate inhibition in vitro and in vivo. Cell Metabolism. 2015;68(6):2197–2211. doi:10.1016/j.cmet.2014.09.018

12. Pektaş MB, Sadi G, Koca HB, et al. Resveratrol ameliorates the components of hepatic inflammation and apoptosis in a rat model of streptozotocin-induced diabetes. Drug Dev Res. 2016;77(1):12–19. doi:10.1002/ddr.21287

13. Jiang C, Wang Y, Jin Q, et al. Cyclocarya paliurus triterpenoids improve mitochondrial functions and inhibit NAFLD progression by regulating SREBP-1c/FAS signaling pathways in LO2 cells. J Agric Food Chem. 2019;67(2):625–636. doi:10.1021/acs.jafc.8b06209

14. Sumita Y, Yoneda M. Current and future pharmacological therapies of NAFLD/NASH: J Gastroenterol. 2018;53(3):362–376. doi:10.1007/s00535-017-1415-1

15. Li Y, Li Q, Li Q, et al. TET1 promotes fatty acid oxidation and inhibits NAFLD progression by hydroxymethylation of PPARα promoter. J Nutr Biochem. 2021;87:108505. doi:10.1016/j.jnutbio.2020.108505

16. Solverson PM, Henderson TR, Debello H, Ferruzzi MG, Baer DJ, Novotny JA. An anthocyanin-rich mixed-berry intervention may improve insulin sensitivity in a randomized trial of overweight and obese adults. Nutrients. 2019;11(12):2876. doi:10.3390/nu1122876

17. Liang A, Wu S, Sakao K, et al. bilberry anthocyanins ameliorate NAFLD by improving dyslipidemia and gut microbiome dysbiosis. Diabetes. 2020;12(11):3252. doi:10.1016/j.diabres.20213252

18. Bognar E, Sarzegi Z, Szabo A, et al. Antioxidant and anti-inflammatory effects in RAW264.7 macrophages of malvidin, a major red wine polyphenol. PLoS One. 2013;8(6):e65535. doi:10.1371/journal.pone.0065535

19. Wei H, Li H, Fan S, et al. Cardioprotective effects of malvidin against isoproterenol-induced myocardial infarction in rats: A Mechanistic Study. Med Sci Monit Int Med J Exp Clin Res. 2017;23:2007–2016. doi:10.12659/MSM.902196

20. Srinivasan K, Viswanad B, Asrat L, Kaul CL, Rama Rao P. Combination of high-fat diet-fed and low-dose streptozotocin-treated rat: a model for type 2 diabetes and pharmacological screening. Pharmacol Res. 2005;52(4):313–320. doi:10.1016/j.yphr.2005.05.004

21. Wang Y, Viscarra J, Kim SJ, Sul HS. Transcriptional regulation of hepatic lipogenesis. Nature Rev Mol Cell Biol. 2015;16(11):678–689. doi:10.1038/nrm4074

22. Fang K, Wu F, Chen G, et al. Diosgenin ameliorates palmitic acid-induced lipid accumulation via AMPK/ACC/CPT-1A and its impact on lipid metabolism, inflammation and fibrosis in non-alcoholic fatty liver disease. J Hepatol. 2015;62(3):720–733. doi:10.1016/j.jhep.2014.10.015
39. Kersten S, Seydoux J, Peters JM, Gonzalez FJ, Desvergne B, Wahli W. Peroxisome proliferator-activated receptor alpha mediates the adaptive response to fasting. *J Clin Invest*. 1999;103(11):1489–1498. doi:10.1172/JCI6223

40. Day CP, James OF. Steatohepatitis: a tale of two “hits”? *Gastroenterology*. 1998;114(4):842–845. doi:10.1016/S0016-5085(98)70599-2

41. Manne V, Handa P, Kowdley KV. Pathophysiology of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis. *Clin Liver Dis*. 2018;22(1):23–37. doi:10.1016/j.clld.2017.08.007

42. Cobbina E, Akhlaghi F. Non-alcoholic fatty liver disease (NAFLD) - pathogenesis, classification, and effect on drug metabolizing enzymes and transporters. *Drug Metab Rev*. 2017;49(2):197–211. doi:10.1080/03602532.2017.1293683

43. Tilg H, Moschen AR. Evolution of inflammation in nonalcoholic fatty liver disease: the multiple parallel hits hypothesis. *Hepatology (Baltimore, Md)*. 2010;52(5):1836–1846. doi:10.1002/hep.24001

44. Boden G, She P, Mozzoli M, et al. Free fatty acids produce insulin resistance and activate the proinflammatory nuclear factor-kappaB pathway in rat liver. *Diabetes*. 2005;54(12):3458–3465. doi:10.2337/diabetes.54.12.3458

45. Han P, Cui Q, Lu W, et al. Hepatocyte growth factor plays a dual role in tendon-derived stem cell proliferation, migration, and differentiation. *J Cell Physiol*. 2019;234(10):17382–17391. doi:10.1002/jcp.28360

46. Klover PJ, Mooney RA. Hepatocytes: critical for glucose homeostasis. *Int J Biochem Cell Biol*. 2004;36(5):753–758. doi:10.1016/j.biocel.2003.10.002