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In patients with moderate-to-severe COVID-19 pneumonia, an aberrant post-viral alveolitis with excessive inflammatory responses and immunothrombosis underpins use of immunomodulatory therapy (eg, corticosteroids and interleukin-6 receptor antagonism). By contrast, immunosuppression in individuals with mild COVID-19 who do not require oxygen therapy or in those with critical disease undergoing prolonged ventilation is of no proven benefit. Furthermore, a window of opportunity is thought to exist for timely immunosuppression in patients with moderate-to-severe COVID-19 pneumonia shortly after clinical presentation. In this Viewpoint, we explore the shortcomings of a universal immunosuppression approach in patients with moderate-to-severe COVID-19 due to disease heterogeneity related to ongoing SARS-CoV-2 replication, which can manifest as RNAaemia in some patients treated with immunotherapy. By contrast, immunomodulatory therapy has overall benefits in patients with rapid SARS-CoV-2 clearance, via blunting of multifaceted, excessive innate immune responses in the lungs, potentially uncontrolled T-cell responses, possible autoimmune responses, and immunothrombosis. We highlight this therapeutic dichotomy to better understand the immunopathology of moderate-to-severe COVID-19, particularly the role of RNAaemia, and to refine therapy choices.

**Introduction**

There is ongoing interest in understanding the link between SARS-CoV-2 infection and inappropriate or excessive immune responses, which might contribute substantially to mortality from COVID-19.1 These responses were first reported in early 2020, when cases of severe systemic inflammation with coagulopathy, which were superficially reminiscent of cytokine storm syndromes, were described.2,4 A common example of a cytokine storm syndrome is macrophage activation syndrome, which encompasses a consumptive bleeding diathesis termed disseminated intravascular coagulation.3,5 The observation of high levels of systemic inflammation with elevated concentrations of C-reactive protein, ferritin, and serum cytokines; cytopenias; and suspected cardiac immune toxicity, in conjunction with coagulopathy, in patients with severe COVID-19 also suggested a cytokine storm with macrophage activation syndrome.6,8 Detection of SARS-CoV-2 RNA in oropharyngeal swab tests for only 8–10 days following SARS-CoV-2 infection, as well as hyperinflammation with increased risk of mortality in the subsequent weeks, also supported the concept of a hyperactivation of the immune system in the post-viral replication phase, which could be modified therapeutically.9,10 This apparent post-infectious inflammation is of considerable interest to rheumatologists, who are familiar with treating conditions such as post-infection inflammation and macrophage activation syndrome.

Data from open-label studies of patients with severe COVID-19 treated with tocilizumab, the first licenced interleukin (IL)-6 receptor blocker, and the efficacy of tocilizumab in treating cytokine storm syndrome associated with chimeric antigen receptor (CAR) T-cell therapy, invigorated a global trial agenda of clinical trials to test immunomodulatory therapy in patients with COVID-19 pneumonia.11,12 As data from randomised controlled trials on immunomodulatory therapy in patients with COVID-19 have emerged, including corticosteroid13 and IL-6 receptor blockade strategies,12,14 the benefits have proven to be modest compared with the benefit of these therapies in patients with genuine cytokine storm syndromes, in whom they can be curative.2 The most encouraging randomised controlled trial in patients with severe COVID-19 pneumonia showed an incremental benefit, with a 4% reduction in mortality when tocilizumab was added to corticosteroids—far lower than that seen with similar strategies for CAR T-cell therapy-associated cytokine storm syndromes.15 The benefits of corticosteroid or IL-6 receptor blockade in patients with COVID-19 have not been replicated in every study,15,16 indicating an incomplete understanding of disease mechanisms and disease heterogeneity. Although tocilizumab has been widely heralded as efficacious with COVID-19 on the basis of large, open-label platform trials, small randomised controlled trials have not shown a survival benefit of the drug.17 Beyond corticosteroids, IL-6 receptor blockade, and antagonism of the Janus kinase (JAK) pathway,17,18 there is insufficient evidence for other immunomodulatory therapies (including IL-1 antagonism) due to the relative paucity of data from large, phase 3 clinical trials; therefore, we do not discuss immunomodulatory therapies further.

In this Viewpoint, we contend that much of the heterogeneity in benefit of immunotherapy is linked to ongoing SARS-CoV-2 replication in a subgroup of patients (figure 1). We focus on patients with moderate-to-severe COVID-19, rather than on those with mild disease who do not require oxygen therapy or those with critical disease undergoing prolonged ventilation, in whom immunotherapy can be less effective (figure 1). Whereas immunotherapy seems to show overall benefits at the population level, we argue that a window of opportunity for immunotherapy might not exist universally in patients with moderate-to-severe COVID-19.
Ongoing active SARS-CoV-2 infection
At the beginning of the COVID-19 pandemic, studies from China indicated that the duration of viral shedding as detected by RT-PCR (not direct viral cultivation) was 7–11 days from illness onset (figure 2). However, the view emerged that a post-viral alveolitis and hyper-inflammation state could be effectively targeted with immunomodulatory strategies. ARDS=acute respiratory distress syndrome.

Figure 1: Viral clearance and immunosuppression recommendations in patients with COVID-19
Patients with mild COVID-19 who do not need oxygen therapy might be immunocompetent and in an early phase of a self-limiting disease with minimal viral replication. Immunosuppression might increase viral replication and is of no benefit in these patients. In patients with moderate or severe COVID-19 who are not mechanically ventilated, some immunosuppression strategies might be beneficial, although optimal strategies await definition. Most of these patients clear the virus by day 8–11; however, some patients with ongoing viral replication might not benefit from immunosuppression. For critically ill patients with COVID-19 who are mechanically ventilated, post-mortem studies have reported extensive fibrosis, lung destruction, and associated pulmonary infarction; therefore, immunosuppression therapy might not be beneficial. ARDS=acute respiratory distress syndrome.

Viral RNA in respiratory tract secretions or in the blood, as detected by PCR, does not necessarily equate to active viral replication, and therapeutic suppression of innate immune responses to viral nucleic acids could be beneficial, provided viral replication is no longer occurring. Although this concept was not well understood at the outset of the COVID-19 pandemic, several studies have since directly investigated active viral replication of SARS-CoV-2 in individuals who are immunocompetent or immunosuppressed, using virological culture techniques that are considered to be the gold standard (table). In one study, only 29% of RT-PCR-positive SARS-CoV-2 cases had cultivatable virus, although this did not last beyond 8 days, and most patients included in the study had mild COVID-19. The positivity rate in oropharyngeal cultures from otherwise healthy people with mild COVID-19 was 74% at 1 week and 20% at 2 weeks. In another study, viral cultivation was more difficult in patients with moderate-to-severe COVID-19 older than 41 years, perhaps indicating excessive immune responses following viral clearance in some cases. In that study, the risk of infection from respiratory secretions declined considerably to 6% in patients with mild-to-moderate disease after 10 days. In another study of 129 patients hospitalised with severe PCR-positive COVID-19, infectious viral particle shedding from the upper airways was detected by viral cultures in only because unrestrained viral replication is still taking place in some. We frame our arguments through the established concept that SARS-CoV-2-related alveolitis triggers an intrapulmonary macrophage activation syndrome-like state with a distinctive immunothrombosis of the lung (termed pulmonary intravascular coagulopathy). This pathology has been supported by single-cell analysis of bronchoalveolar fluid from patients with severe COVID-19, and intrapulmonary macrophage activation has been confirmed. We propose that immunotherapy would improve survival only in clinical settings in which excessive innate and adaptive immune responses (including autoimmune responses) occur in the context of rapid control of SARS-CoV-2 replication. We argue that distinguishing ongoing infection in the alveolar territory, which manifests as RNAemia, from excessive inflammatory responses and immunothrombosis occurring after clearance of infection, could be useful to improve survival.

Figure 2: Optimal trajectory of COVID-19 pneumonia
Typically, SARS-CoV-2 alveolar infection proceeds with a viral phase, an inflammatory phase, and an associated immunothrombotic phase. In severe disease, ARDS can develop. In patients who are otherwise immunocompetent, obesity and other cardiovascular risk factors can lead to cardiopulmonary system decompensation. ARDS=acute respiratory distress syndrome.
23 (18%) patients and correlated with RT-PCR viral loads of more than 10 million copies per mL. The median duration of infectious virus shedding was 8 days after symptom onset; the probability of cultivating infectious viral was below 5% after a duration of symptoms of 15 days. Increases in the number of anti-SARS-CoV-2 antibodies correlated with an inability to detect infectious virus.

At the population level, patients with moderate-to-severe COVID-19 are generally immunocompetent, unlike patients with cancer who have been treated with chemotherapy and are considered to be immunodeficient (figure 3). In patients with cancer receiving chemotherapy who are concomitantly infected with SARS-CoV-2, replication-competent virus was retrievable from the upper airways for 3–8 weeks after symptom onset. Several other studies have provided evidence for ongoing SARS-CoV-2 replication in patients with severe COVID-19 pneumonia or in those receiving therapeutic immunosuppression. However, ongoing viral replication has generally been overlooked in therapy decisions for critically ill patients, in whom prompt immunosuppression is likely to worsen outcomes in the face of potentially rampant viral replication (figure 3).

The viral reservoir in these patients is poorly understood; however, its persistence over weeks might be linked to permissive bronchial mucosal or upper airway environment, and an inability to kill virus-infected cells. In addition to patients with COVID-19 receiving chemotherapy for cancer, studies using RT-PCR showed higher amounts of SARS-CoV-2 RNA in patients with acute respiratory distress syndrome (ARDS) treated with corticosteroids compared with those treated with standard of care. High viral loads are associated with immunosuppression, as well as extended stay in intensive care, and prolonged intubation, but the relationship between viral load and mortality remains controversial.

Cytokine elevation in moderate-to-severe COVID-19 pneumonia and RNAemia

The magnitude of elevation in serum IL-6 concentrations in patients with moderate-to-severe COVID-19 is in the range of 100 pg/mL or less, compared with approximately 10,000 pg/mL typically observed in cytokine storm syndromes in patients receiving CAR T-cell therapy. Low IL-6 concentrations have been used to stratify patients with COVID-19 with a good prognosis who do not require anti-cytokine therapy. However, high serum cytokine concentrations might be linked to ongoing SARS-CoV-2 viral replication. The concept that active viral replication might be detrimental in patients with severe COVID-19 is supported by the early termination of several studies on IL-6 receptor blockade, in which increased mortality was reported in the group receiving tocilizumab. Additionally, data from a meta-analysis of trials suggested that tocilizumab was less effective in patients requiring ventilation than in those not requiring ventilation (figure 3). One study reported that only seven (4%) of 168 patients with severe SARS-CoV-2 infection had elevations in serum cytokines indicative of cytokine storm, and that these elevations were of a lower magnitude than in patients with severe influenza infection, challenging the concept of a systemic cytokine storm and highlighting pulmonary-centred pathology in patients with severe COVID-19.

A key observation in patients with severe COVID-19 is that RNAemia or detectable SARS-CoV-2 RNA in the blood, but not proven cultivable virus, is linked to serum concentrations of IL-6 up to 10 times higher than in patients without RNAemia. In a study of 192 patients with severe COVID-19, 71 (37%) had RNAemia (defined as a positive result by real-time PCR for E, RdRp, or N genes in plasma samples at any timepoint), which was associated with increased risk of invasive mechanical ventilation support, admission into intensive care, multi-organ dysfunction, and death. Baseline disease severity, baseline corticosteroid use, and viral titre were also associated with risk of death in the same study. Furthermore, heavily infected secretory cells of the human airway epithelium expressed IL-6 abundantly. Modest RNAemia (as compared with elevated RNA concentrations measured by PCR of respiratory secretions), has been associated with high IL-6 concentrations and mortality in critically ill patients with COVID-19 pneumonia (table 1). Patients with COVID-19 and RNAemia had higher viral loads in respiratory secretion samples than did those without RNAemia, and high plasma RNAemia has been associated with severe COVID-19 requiring admission to intensive care. It is incompletely understood how RT-PCR positivity for SARS-CoV-2 in upper respiratory tract secretions aligns with cultivable virus in the alveolar territory, and this is a major consideration for therapy. Worryingly, ongoing SARS-CoV-2 replication in the alveolar compartment is expected to drive potentially lethal, diffuse alveolar damage.

| PCR at 1 week | PCR at 2 weeks | Viral culture |
|---------------|---------------|---------------|
| Oropharynx    | +++           | +             | Detectable in <80% of cultures at week 1, but in only 6% after 10 days; might persist for weeks in immunosuppressed patients |
| Airways (endotracheal tube aspirate) | +++ | ++ | + |
| Blood         | Negative in mild or moderate COVID-19; variably positive in severe COVID-19 (+ or ++) | Negative in mild or moderate COVID-19; variably positive in severe COVID-19 (+ or ++) | No culture attainable in any group; infection of endothelial cells in vitro not usually attainable |

+, ++, and +++ refer to the strength of the positivity of PCR. The magnitude of elevation by RT-PCR in the blood is 3–4 logs lower than in the oropharynx or airways. Viral detection by PCR at low-cycle thresholds is associated with SARS-CoV-2 replication, especially in the oropharynx and airways during the first week of infection. By contrast, little evidence exists for replication in the circulation.
are considered to be important factors in severe pneumonia. Furthermore, post-viral cytokine storm scenarios robustness of adaptive immunity and an increased reliance on innate immune mechanisms via IL-6, IL-8, and IL-1, alveolar damage, have been suspected but not proven. Additionally, the effects of ageing, with a reduced immunothrombosis. Excessive cytotoxic CD8 T-cell responses, leading to alveolar pneumocyte killing and diffuse concentrations of proinflammatory cytokines, which drive macrophage and neutrophil activation, and suggested that helper T-cell cytokine responses in the pulmonary compartment might contribute to high local immunosuppression in patients with controlled SARS-CoV-2 viral replication. Given that most mortality occurs corticosteroid therapy in patients not requiring oxygen. (B) Potential mechanisms underlying benefit of as type-1 interferon pathway defects, immunosuppression regimens in autoimmune diseases, organ transplantation settings, cancer-related immunosuppression or chemotherapy, or the administration of corticosteroid therapy in patients not requiring oxygen: (A) Factors leading to prolonged viral replication include pre-existent, unrecongnised immunodeficiency states, such as type-1 interferon pathway defects, immunosuppression regimens in autoimmune diseases, organ transplantation settings, cancer-related immunosuppression or chemotherapy, or the administration of corticosteroid therapy in patients not requiring oxygen: (B) Potential mechanisms underlying benefit of interferon responses.63 Other mechanisms, such as antiviral effects (eg, JAK inhibition) also been reported.60,61 Genome-wide association studies in patients with severe COVID-19 have also indicated dysregulation of the interferon pathway in critically ill patients, including single-nucleotide polymorphisms in OAS1, OAS2, and OAS3 genes, and adjacent to TYK2 and IFNAR2,62 although the precise functional correlates of these polymorphisms need further evaluation. In general, studies showing absence of measurable interferon in critical cases of COVID-19 attest to the multifaceted mechanisms in which SARS-CoV-2 can disable antiviral interferon responses.61

Acquired immunodeficiency states, such as those secondary to prolonged corticosteroid treatment, B-cell depleting therapy, or immunosuppressive drugs (eg, calcineurin inhibitors, mycophenolate, azathioprine), have marked effects on T-cell function in patients with rheumatological conditions, and these states have been associated with an increased risk of COVID-19-related mortality.64,65 Likewise, pan-cytokine inhibition with corticosteroids in patients with mild COVID-19 was associated with increased mortality.66 Collectively, these factors are likely to contribute to ongoing viral replication, which might be a major challenge in selecting patients for immunosuppression and means that consideration of early immunomodulatory therapy in patients with moderate-to-severe COVID-19 pneumonia needs careful re-evaluation (figure 3). This challenge is already well recognised with respect to hepatitis B virus reactivation in patients receiving rituximab-containing regimens during lymphoma therapy.67 Although haematologists pay close attention to active viral infection when treating patients with primary haemophagocytic lymphohistiocytosis, scant consideration has been given to virus-induced
development of ARDS (figure 3). Direct measurement of alveolar infection is not possible; however, RNAemia in patients critically ill with COVID-19 could be a surrogate for severe alveolitis with damage to the alveolar–vascular barrier.4 This theory is supported by post-mortem reports describing SARS-CoV-2 in the damaged alveolar compartment by use of electron microscopy, RNAscope assays, and SARS-CoV-2 protein immunohistochemistry, which point to active alveolar viral replication in patients with late-stage COVID-19.19 However, the presence of viral protein or viral nucleic acid in post-mortem examinations does not necessarily equate to actual viral replication at the time of sampling.35 Nevertheless, bronchoalveolar lavage from patients with COVID-19 in intensive care showed viral particles inside mononuclear cells through electron transmission microscopy, confirmed by immunostaining of antiviral capsid and spike antibodies.36 Collectively, these findings support the idea that unrestrained viral replication is a credible factor that could account for the heterogeneity of outcomes in clinical trials of patients with severe COVID-19 pneumonia.

**Immunodeficiency states and SARS-CoV-2 persistence**

Autoinflammatory syndromes and autoimmune diseases might be intimately intertwined with primary immunodeficiency states.55 For example, genetic defects in the perforin pathway machinery in primary haemophagocytic lymphohistiocytosis are linked to simultaneous immunodeficiency in CD8 T cells and natural killer cells, immune dysregulation, and a hyperinflammatory state; such scenarios are uncommon in COVID-19, although some studies have shown heterozygous mutations in the perforin pathway in patients with severe COVID-19.56,57 Loss-of-function mutations in genes involved in interferon signalling pathways (including toll-like receptors) have also been reported.61,62 Genome-wide association studies in patients with severe COVID-19 have also indicated dysregulation of the interferon pathway in critically ill patients, including single-nucleotide polymorphisms in OAS1, OAS2, and OAS3 genes, and adjacent to TYK2 and IFNAR2,62 although the precise functional correlates of these polymorphisms need further evaluation. In general, studies showing absence of measurable interferon in critical cases of COVID-19 attest to the multifaceted mechanisms in which SARS-CoV-2 can disable antiviral interferon responses.61

**Figure 3:** Factors and mechanisms causing prolonged viral replication in the alveolar territory

(A) Factors leading to prolonged viral replication include pre-existent, unrecongnised immunodeficiency states, such as type-1 interferon pathway defects, immunosuppression regimens in autoimmune diseases, organ transplantation settings, cancer-related immunosuppression or chemotherapy, or the administration of corticosteroid therapy in patients not requiring oxygen: (B) Potential mechanisms underlying benefit of interferon responses.63 Other mechanisms, such as antiviral effects (eg, JAK inhibition) also been reported.60,61 Genome-wide association studies in patients with severe COVID-19 have also indicated dysregulation of the interferon pathway in critically ill patients, including single-nucleotide polymorphisms in OAS1, OAS2, and OAS3 genes, and adjacent to TYK2 and IFNAR2,62 although the precise functional correlates of these polymorphisms need further evaluation. In general, studies showing absence of measurable interferon in critical cases of COVID-19 attest to the multifaceted mechanisms in which SARS-CoV-2 can disable antiviral interferon responses.61

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immunodeficiency or underlying, unrecognised immunodeficiency in the context of moderate-to-severe COVID-19, in which a one-size-fits-all approach has been used (figure 3). By contrast, the history of cryoglobulinaemia related to hepatitis C virus has shown that immunosuppressive treatments are not deleterious, even in the presence of increased viral replication.⁶⁹ In patients with moderate-to-severe COVID-19 pneumonia, the evidence to date suggests that immunosuppression in the context of active replication of SARS-CoV-2, which quickly damages the alveolar territory and causes rapidly developing diffuse alveolar damage, might be detrimental.

**ARDS development during immunosuppression in COVID-19**

Patients hospitalised with COVID-19 might require high-dependency respiratory support, given that many of these patients meet the ARDS Berlin Definition of diffuse pulmonary infiltrates and severe disease (PaO₂ <100 mm Hg or FiO₂ <100%) not explained by fluid overload.⁴⁴ In patients with ARDS without COVID-19, the role of corticosteroid treatment remains controversial. Pre-emptive or late use of corticosteroids in these patients is not beneficial and could be harmful,⁶⁹ with late use probably reflecting the ineffectiveness of immunosuppression in the setting of fibroproliferative or fibrotic ARDS.⁷⁰ A meta-analysis of trials in patients with severe COVID-19 found that combination therapy with corticosteroids plus tocilizumab was not efficacious in patients on invasive mechanical ventilation,⁴³ many of whom probably had ARDS.

Although our understanding of the lung pathology in ARDS is based on post-mortem samples rather than on ante-mortem studies, the data suggest that ARDS is linked to diffuse alveolar damage in most cases.⁷¹,⁷² Another major pathological finding is that the diffuse immunothrombosis seen in severe COVID-19 pneumonia is completely distinctive to that seen in disseminated intravascular coagulation.⁷³ Regardless of nuanced pathological differences, most patients who die from severe COVID-19 respiratory failure ultimately show loss of lung compliance and the pathology of ARDS, suggesting that regardless of whether the genesis is viral pneumonia or vascular thrombosis associated with evolving ARDS, the final pathway is similar. Unremitting viral alveolitis secondary to immune deficiency or immunosuppression might also fuel the associated development of ARDS (figure 3).

**Treating immune responses when SARS-CoV-2 viral replication is contained**

Despite the tentative link between corticosteroid use and viral replication, as measured by RNAaemia and elevated concentrations of IL-6, some types of immunotherapy seem to improve survival from severe COVID-19 at the population level. Having addressed ongoing active viral replication, it is worth discussing the scenarios in which viral replication is controlled or restricted, yet an exaggerated immune response occurs and manifests with immunothrombosis, which helps to contain dissemination of viral nucleic acids (figure 3). Pulmonary immunothrombosis is evident in over 90% of severe COVID-19 cases,⁷⁴ occurs independent of disease duration, and is more prevalent in COVID-19 than in influenza.⁷⁵ This pulmonary immunopathology has been well described and termed as pulmonary intravascular coagulopathy, as opposed to overt disseminated intravascular coagulation.⁷⁶,⁷⁷ Multiple molecular mechanisms have been shown to connect inflammation and coagulation under the integrated umbrella of immunothrombosis,⁷⁸ which include dynamic bidirectional crosstalk between coagulation and inflammation in vivo. Conversely, activated coagulation proteases are able to cleave cell-surface receptors and can thereby trigger proinflammatory signalling pathways in various cell types, including macrophages, endothelial cells, and platelets. Besides endotheliopathy, which appears to occur independent of direct viral infection, several processes might contribute to pulmonary intravascular coagulopathy, including platelet activation, formation of neutrophil extracellular traps (NETosis), complement activation, and down-regulation of fibrinolysis. Consistent with this multifactorial process, distinctive clot morphologies have been characterised in patients with severe COVID-19, including the presence of abundant necrotic neutrophils, free DNA, and platelet-rich areas with elevated megakaryocyte numbers.⁷⁹ Megakaryocytes typically occur in the bone marrow and lungs under physiological conditions; however, the number of CD61-positive megakaryocytes was reported to be significantly higher in the lungs of patients with COVID-19 pneumonia than in patients with ARDS without COVID-19, suggesting active platelet production, aggregation, and consumption.⁷⁹ Independently, peripheral blood neutrophilia and systemic neutrophil activation have also been consistently linked to poor prognosis in cases of severe COVID-19.⁸⁰,⁸¹ This dysregulated, local immunothrombosis seems to occur independent of disease duration and has specific deleterious effects on the monolayer lining of endothelial cells within the pulmonary microvasculature.⁷⁵,⁷⁷ A key consideration is that infection might be compartmentalised to the alveolar compartment, with endothelial and associated vascular thrombosis occurring independent of infection and compartmentalisation between sites.⁴ Electron microscope studies claiming direct viral infection of the pulmonary endothelium have been strongly challenged, with some suggesting that the morphology of cellular organelles was misinterpreted as SARS-CoV-2.⁸² The general inability to culture SARS-CoV-2 in endothelial cells raises the possibility that the endothelial orchestration of vascular damage might occur solely via severe inflammation and multifaceted, injurious immune mechanisms.⁸³–⁸⁵ The effect of ACE2 receptor expression...
on the severity of COVID-19 pneumonia remains unclear, but it is probably not linked to the endothelium given the resistance of the endothelium to SARS-CoV-2 infection. RNAemia does not seem to occur in mild COVID-19 due to sparing of the alveolar territory or possibly because of viral containment by immunothrombosis. The presence of SARS-CoV-2 RNA in the plasma of severely ill, but not mildly ill, patients supports the idea that the alveolar–vascular barrier breaks down, resulting in viral RNA dissemination and systemic immunothrombosis (figure 4).

We have argued that this complex and prognostically relevant immunothrombosis without viral replication, especially in the vascular compartment, could partially explain the benefits of immunomodulatory therapy when used in the post-viral replication phase in patients with severe COVID-19 pneumonia. Unlike with immunomodulatory therapy, a therapeutic dose of anticoagulation with heparin did not improve outcomes in patients with severe or critical COVID-19, and their risk of major bleeding was increased (3-7% in treated patients vs 1-8% in controls), which supports the pre-eminence of immunothrombosis. However, in patients with mild COVID-19, full-dose anticoagulation conferred a small survival advantage. This scenario is similar to Behçet’s disease, whereby thromboinflammation requires immunotherapy rather than anticoagulation.

**Excessive adaptive and innate immune responses in COVID-19**

In patients with COVID-19 in whom active virus replication is controlled, various mechanisms have been suggested to explain how immunotherapy controls immunothrombosis. With respect to humoral immunity directed by B cells, production of a multiplicity of anti-cytokine, anti-endothelial, and other autoantibodies has been reported. Extensive pulmonary infarction and tissue necrosis in the presence of abundant viral RNA and other adjuvants is likely to result in temporary failure of immunological tolerance against many self-proteins, with emergence of multiple autoantibodies. Such a transient autoimmune process is well recognised following tissue infarction in other settings, such as myocardial infarction or stroke. Likewise, extensive pulmonary infarction, viral infection, and tissue necrosis might be key factors in the secondary production of anti-interferon and other autoantibodies that characterise moderate-to-severe COVID-19. If this is the case, immunosuppression of the secondary autoantibody responses might be of little value in moderate-to-severe COVID-19. However, the presence of anti-endothelial cell autoantibodies in patients with severe COVID-19 was shown to trigger NETosis and facilitate venous thrombosis in murine models. In another COVID-19 mouse model, such autoantibodies were shown to be immunosuppressive and exacerbate disease (figure 3). It is therefore possible that suppression of some of the plethora of autoantibodies reported in severe COVID-19 might be a factor in the beneficial effects of immunotherapy.

In cases of COVID-19 in which antiviral T-cell responses become detrimental, it is probable that immunosuppression blunts this response (figure 3). An attractive theory—based on the timing of viral clearance from day 10 onwards in many patients with COVID-19, which broadly correlates with the timing of emerging T-cell responses—is that the potential success of corticosteroid therapy is
linked to the taming of overzealous CD4+ and CD8+ T-cell responses. Therefore, the excessive production of T-helper (Th)-1 cytokines in the lungs and CD8+ T-cell cytotoxicity could be driving immunopathology.59 One study, in which single-cell RNA sequences were analysed from the airways of patients with COVID-19, reported cytotoxic T cells with perforin and granzyme production in the vicinity of stressed pulmonary epithelial cells, suggesting a link between epithelial cell infectivity and adjacent lymphocyte toxicity.58 An obvious question is whether or not suppression of T-cell killing of infected pneumocytes might be beneficial in this scenario (figure 4). This notion is relevant because, experimentally, a low level of infection of type 2 pneumocytes with influenza A virus has not been shown to be detrimental to the functional activity of the cells (including cell division).58 Consequently, excessive cytotoxic T cell-mediated elimination of type 2 pneumocytes, which are stem cells for the alveolus, might be detrimental and could contribute to diffuse alveolar damage with ARDS development.

Older patients (aged >80 years) are at increased risk of poor outcomes from COVID-19 pneumonia and, in general, do not have robust T-cell and B-cell responses. This patient group also shows immunosenescence and an increased magnitude of inflammation driven by the innate immune system (also known as inflammaging), which probably underscores pathogen–host interactions that drive tissue damage and immunothrombosis (figure 3). Functional assays of samples from patients with severe COVID-19 showed reduced production of T-cell-derived cytokines (eg, IFNγ, IL-17, and IL-22) and prominent T-cell exhaustion in critically ill patients, whereas innate immune responses were intact or increased.100

**Future directions and conclusions**

In this Viewpoint, we have focused on patients with moderate-to-severe COVID-19 pneumonia who are thought to represent the best target group for immunotherapy, given that patients with mild COVID-19 or those who are critically ill and mechanically ventilated with extensive tissue destruction might not respond to these agents. Timing treatment with respect to the clinical phase of the disease, and not delaying such treatment for too long, are considered to be key potential factors in the selection of patients who are most likely to respond to immunotherapy. However, a roadmap for therapy stratification needs to be thoroughly evaluated, especially because some patients with rapidly progressing disease can have unrestrained viral replication. This patient group, who have poor prognosis related to ongoing viral replication, seems to be hidden among the larger subgroup of patients with COVID-19 who are responsive to immunotherapy, a factor that might be important in blunting immunotherapy responses, rendering them modest, incremental, or even futile.

We have reviewed the evidence suggesting that RNAemia in patients with comparatively high elevations in cytokine concentrations probably reflects ongoing viral replication, and it is unlikely that these patients will respond to immunosuppression. Strategies to differentiate this patient group and to establish the PCR cycle threshold for detection of viral RNA or identify viral antigenaemia could resolve the question of whether immunosuppression should be completely avoided in these patients and anti-SARS-CoV-2 antibody cocktails and antiviral therapies would be more appropriate. Given that our understanding of the immunopathogenesis of COVID-19 has improved greatly over the past 18 months, we advocate for a careful, simultaneous evaluation of blood inflammatory markers and the magnitude and persistence of RNAemia to formally identify which patients might optimally respond to therapy (figure 4). High initial RNAemia or persistent RNAemia might be indicators to exclusively pursue standard-of-care with antiviral therapy strategies, including anti-spike antibody cocktails and RNA transcriptase antagonism. The cellular and molecular mechanisms underscoring the beneficial effects of immunotherapy are likely to be multifaceted, including innate and adaptive immune mechanisms that remain incompletely understood. However, it is probable that the beneficial effects of immunotherapy are limited to the post-viral alveolitis phase of COVID-19 and that identifying the patient group with ongoing viral replication in the alveolar territory should help to optimise use of immunosuppressive therapy. It is possible that immunosuppression in patients with moderate-to-severe COVID-19 and marked inflammatory responses might facilitate viral replication and medium-term and long-term lung damage; long-term survival studies in such patients are needed.

**Contributors**

DM developed the initial concepts for this paper. All authors contributed to the first draft writing, the literature review, critical revision, and editing. All authors have participated sufficiently in this work, take public responsibility for the content, and have made substantial contributions to this research. All authors approved the final version and had final responsibility for the decision to submit for publication.

**Declaration of interests**

All authors declare no competing interests.

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References

1. Bredin P. Immune determinants of COVID-19 disease presentation and severity. Nat Med 2021; 27: 28–33.
2. McGonagle D, Bridgewood C, Ramanan AV, Meaney JFM, Wadat A. COVID-19 vasculitis and novel vasculitis mimics. Lancet Rheumatol 2021; 3: e224–33.
3. McGonagle D, O’Donnell JS, Sharif K, Emery P, Bridgewood C. Immune mechanisms of pulmonary intravascular coagulopathy in COVID-19 pneumonia. Lancet Rheumatol 2020; 2: e437–44.
4. McGonagle D, Plein S, O’Donnell JS, Sharif K, Bridgewood C. Increased cardiovascular mortality in African Americans with COVID-19. Lancet Respir Med 2020; 8: 649–51.
5. McGonagle D, Ramanan AV, Bridgewood C. Immune cartography of macrophage activation syndrome in the COVID-19 era. Nat Rev Rheumatol 2021; 17: 145–57.
6. Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. Lancet 2020; 395: 1054–62.
7. Liu D, Cui P, Zeng S, et al. Risk factors for developing into critical COVID-19 patients in Wuhan, China: a multicenter, retrospective, cohort study. Clinical Medicine 2020; 25: 100471.
8. Sinha P, Matthyse MA, Calfee CS. Is a “Cytokine Storm” relevant to COVID-19? JAMA Intern Med 2020; 180: 1152–54.
9. Shereen MA, Khan S, Kazmi A, Bashir N, Siddique R. COVID-19 infection: origin, transmission, and characteristics of human coronaviruses. J Adv Res 2020; 24: 91–98.
10. Li TZ, Cao ZH, Chen Y, et al. Duration of SARS-CoV-2 RNA shedding and factors associated with prolonged viral shedding in patients with COVID-19. J Med Virol 2021; 93: 506–12.
11. Guaraldi G, Meschiari M, Cozzi-Lepri A, et al. Tocilizumab in patients with severe COVID-19: a retrospective cohort study. Lancet Rheumatol 2020; 2: e74–84.
12. Salama C, Han J, Yau L, et al. Tocilizumab in patients hospitalized with COVID-19 pneumonia. N Engl J Med 2021; 384: 20–30.
13. Horby P, Lim WS, Emberson JR, et al. Dexamethasone in hospitalized patients with COVID-19. N Engl J Med 2021; 384: 693–704.
14. Gordon AC, Mouncey PR, Al-Beidh F, et al. Interleukin-6 receptor antagonists in critically ill patients with COVID-19. N Engl J Med 2021.
15. Yang JW, Yang L, Luo RG, Xu JF, Corticosteroid administration for viral pneumonia: COVID-19 and beyond. Clin Microbiol Infect 2020; 26: 1711–71.
16. Li Y, Li, J, et al. Adverse outcomes associated with corticosteroid use in COVID-19: a retrospective multicenter cohort study. Front Med (Lausanne) 2021; 8: 604263.
17. Alam W, Bizri AR. Efficacy of tocilizumab in COVID-19: a review of the current evidence. Sci Prog 2021; 104: 36850421100372.
18. Guimarães PO, Quirk D, Furtado RH, et al. Tofacitinib in patients hospitalized with COVID-19 pneumonia. N Engl J Med 2021; 384: 855–65.
19. Liao M, Liu Y, Yuan J, et al. Single-cell landscape of bronchialveolar immune cells in patients with COVID-19. Nat Med 2020; 26: 842–44.
20. Carasana I, Sonzogni A, Nasr A, et al. Pulmonary post-mortem findings in a series of COVID-19 cases from northern Italy: a two-centre descriptive study. Lancet Infect Dis 2020; 20: 1135–40.
21. Zou L, Ruan F, Huang M, et al. SARS-CoV-2 viral load in upper respiratory specimens of infected patients. N Engl J Med 2020; 382: 1177–79.
22. Siddiqi HK, Mehra MR. COVID-19 illness in native and immunosuppressed states: a clinical-therapeutic staging proposal. J Heart Lung Transplant 2020; 39: 405–07.
23. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ. COVID-19: consider cytokine storm syndromes and immunosuppression. Lancet 2020; 395: 1033–34.
24. Liu Y, Yan LM, Wan L, et al. Viral dynamics in mild and severe cases of COVID-19. Lancet Infect Dis 2020; 20: 565–71.
25. Jacobs JL, Mellors JW. Detection of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) RNA in blood of patients with coronavirus disease 2019 (COVID-19): what does it mean? Clin Infect Dis 2020; published online Sept 8. https://doi.org/10.1093/cid/ciaa1116.
26. Zheng S, Fan J, Yu F, et al. Viral load dynamics and disease severity in patients infected with SARS-CoV-2 in Zhejiang province, China. January-March 2020: retrospective cohort study. BMJ 2020; 369: m4143.
27. Munker D, Osterman A, Stubbe H, et al. Dynamics of SARS-CoV-2 shedding in the respiratory tract depends on the severity of disease in COVID-19 patients. Eur Respir J 2021; 58: 2002734.
28. Bueti N,icky P-H, Le Hingrat Q, et al. SARS-CoV-2 detection in the lower respiratory tract of invasively ventilated ARDS patients. Crit Care 2020; 24: 610.
29. Bullard J, Dust K, Funk D, et al. Predicting infectious SARS-CoV-2 from diagnostic samples. Clin Infect Dis 2020; 71: e2661–66.
30. Singanayagam A, Patel M, Charlett A, et al. Duration of infectiousness and correlation with RT-PCR cycle threshold values in cases of COVID-19, England, January to May 2020. Euro Surveill 2020; 25: 2001483.
31. van Kampen JJA, van de Vijver DAMC, Friaaij PLA, et al. Duration and key determinants of infectious virus shedding in hospitalized patients with coronavirus disease-2019 (COVID-19). Nat Commun 2021; 12: 267.
32. Camps J, Gaya A, Marcos MA, et al. Persistent replication of SARS-CoV-2 in a severely immunocompromised patient treated with several courses of remdesivir. Int J Infect Dis 2021; 104: 379–81.
33. Avanzato VA, Matson MJ, Seifert SN, et al. Case study: prolonged infectious SARS-CoV-2 shedding from an asymptomatic immunocompromised individual with cancer. Cell 2020; 183: 1901–12.e9.
34. Hendsky MK, Bain WG, Jacobs J, et al. Intractable COVID-19 and prolonged SARS-CoV-2 replication in a CAR-T-cell therapy recipient: a case study. Clin Infect Dis 2021; 73: e815–21.
35. Nakajima Y, Ogai A, Furukawa K, et al. Prolonged viral shedding of SARS-CoV-2 in an immunocompromised patient. J Infect Chemother 2021; 27: 387–89.
36. Tarhini H, Reicoing A, Bridier-Nahmias A, et al. Long term SARS-CoV-2 infectiousness among three immunocompromised patients: from prolonged viral shedding to SARS-CoV-2 superinfection. J Infect Dis 2021; 223: 1522–27.
37. Matthyse MA, Wick KD. Corticosteroids, COVID-19 pneumonia, and acute respiratory distress syndrome. J Clin Invest 2020; 130: 6218–21.
38. Maltzou HC, Raptopoulos V, Vorou R, et al. Association between upper respiratory tract viral load, comorbidities, disease severity and outcome of patients with SARS-CoV-2 infection. J Infect Dis 2021; 223: 1132–38.
39. Quarantico L, Sonaglia A, McGonagle D, et al. Profiling COVID-19 pneumonia progressing into the cytokine storm syndrome: results from a single Italian Centre study on tocilizumab versus standard of care. J Clin Virol 2020; 129: 104444.
40. Uterno-Rico A, Ruiz-Hornillos J, Gonzalez-Cuadrado C, et al. IL-6-based mortality prediction model for COVID-19: validation and update in multicenter and second wave cohorts. J Allergy Clin Immunol 2021; 147: 1652–61.
41. Quarantico L, Fabris M, Sonaglia A, et al. Interleukin 6, soluble interleukin 2 receptor alpha (CD25), monocyte colony-stimulating factor, and hepatocyte growth factor linked with systemic hyperinflammation, innate immunity hyperactivation, and organ damage in COVID-19 pneumonia. Cytokine 2021; 140: 155438.
42. Vega VC, Prats JAGG, Farias DLC, et al. Effect of tocilizumab on clinical outcomes at 15 days in patients with severe or critical coronavirus disease 2019: randomised controlled trial. BMJ 2021; 372: n84.
43. WHO. Rapid Evidence Appraisal for COVID-19 Therapies (REACT) Working Group. Association between administration of IL-6 antagonists and mortality among patients hospitalized for COVID-19: a meta-analysis. JAMA 2021; 326: 499–518.
44. Mudd PA, Crawford JC, Turner JS, et al. Distinct inflammatory profiles describe US COVID-19 from influenza with limited contributions from cytokine storm. Sci Adv 2020; 6: eabe3024.
45. Li H, Gu X, Li H, et al. Risk factors of viral maemia and its association with clinical prognosis among patients with severe COVID-19. Chest 2021; 159: 1582–86.
Fiege JK, Thiede JM, Nanda HA, et al. Single cell resolution of SARS-CoV-2 tropism, antiviral responses, and susceptibility to therapies in primary human airway epithelium. PLoS Pathog 2021; 17:e1009292.

Fajny-Pelzer J, Regan J, Coxen K, et al. SARS-CoV-2 viral load is associated with increased disease severity and mortality. Nat Commun 2020; 11:5493.

Martín-Vicente M, Almansa R, Martínez I, et al. Absent or insufficient anti-SARS-CoV-2 S antibodies at ICU admission are associated to higher viral loads in plasma, antigenemia and mortality in COVID-19 patients. medRxiv 2021; published online March 8. https://doi.org/10.1101/2021.03.08.21235121 (preprint).

Colagrossi L, Antonello M, Renica S, et al. SARS-CoV-2 RNA in plasma samples of COVID-19 affected individuals: a cross-sectional proof-of-concept study. BMC Infect Dis 2021; 21:184.

Tang K, Wu L, Luo Y, Gong B. Quantitative assessment of SARS-CoV-2 RNAemia and outcome in patients with coronavirus disease 2019. J Med Virol 2021; 93:1165–75.

Bermejo-Martín JF, Gonzáles-Rivera M, Almansa R, et al. Viral RNA load in plasma is associated with critical illness and a dysregulated host response in COVID-19. Crit Care 2020; 24:691.

Deshmukh V, Motwani R, Kumar A, Kumari C, Raza K. Histopathological observations in COVID-19: a systemic review. J Clin Pathol 2021; 74:76–83.

Schurink B, Ross E, Radonc T, et al. Viral presence and immunopathology in patients with lethal COVID-19: a prospective autopsy cohort study. Lancet Microbe 2020; 1:e290–99.

Pandolfi I, Fossali T, Frangipane V, et al. Broncho-alveolar inflammation in COVID-19 patients: a correlation with clinical outcome. BMC Pulm Med 2020; 20:301.

Hoyos-Bachigilou R, Chou J. Autoimmunity and immunodeficiency. Curr Opin Rheumatol 2020; 32:168–74.

Opoka-Winiarska V, Grywalska E, Rolinski J. Could hemophagocytic lymphohistiocytosis be the core issue of severe COVID-19 cases? Eur Med J 2020; 11:214.

Cron RQ, Behrens EM, Shkayoor B, Ramanan AV, Chatham WW. Does viral hemorrhagic fever represent reactive hemophagocytic syndrome? J Rheumatol 2015; 42:1078–80.

Calvera-Marante O, Rodriguez de Frias E, Plegeuzuelo DE, et al. Performin gene variant A91V in young patients with severe COVID-19. Haematologica 2020; 105:2844–46.

Luo H, Liu D, Liu W, et al. Germline variants in UNC13D and CABP1 are enriched in COVID-19 patients experiencing severe cytokine storm. Eur J Hum Genet 2021; 29:1112–15.

van der Made CI, Simons A, Schuurs-Hoeijmakers J, et al. Presence of genetic variants among young men with severe COVID-19. JAMA 2020; 324:663–73.

Zhang Q, Bastard P, Liu Z, et al. Inborn errors of type I IFN immunity in patients with life-threatening COVID-19. Science 2020; 370:eabd4570.

Pairo-Castineira E, Clohisy S, Klarc L, et al. Genetic mechanisms of critical illness in COVID-19. Nature 2021; 591:92–98.

Manito E, Bukreyeva N, Maruyama J, Paesler S, Huang C. Antiviral activities of type I interferons to SARS-CoV-2 infection. Antiviral Res 2020; 179:104811.

Akiyama S, Hamdeh S, Micic D, Sakuraba A. Prevalence and clinical recommendations for the management of Behçet’s syndrome. Autoimmun Rev 2020; 19:102537.

Berastegui-Cabrera J, Salto-Alejandre S, Valerio M, et al. SARS-CoV-2 RNAemia is associated to higher viral loads in plasma, antigenemia and mortality in COVID-19 patients. medRxiv 2021; published online March 8. https://doi.org/10.1101/2021.03.08.21235121 (preprint).

Huang H, Li X, Zhu J, et al. Entecavir vs lamivudine for prevention of hepatitis B virus reactivation among patients with untreated chronic hepatitis B. Cochrane Database Syst Rev 2019; 7:CD004472.

Meduri GU, Headley AS, Golden E, et al. Effect of prolonged methylprednisolone therapy in unresolving acute respiratory distress syndrome: a randomized controlled trial. JAMA 1998; 280:159–65.

Masaud T, Duarte-Neto AN, da Silva LFF, et al. Tracking the time course of pathological patterns of lung injury in severe COVID-19. Respir Res 2021; 22:32.

Polak SB, Van Gool IC, Cohen D, von der Thiesen JH, van Paasen J. A systematic review of pathological findings in COVID-19: a pathophysiologic timeline and possible mechanisms of disease progression. Med Pathol 2020; 33:2128–38.

McGonagle D, O’Donnell JS, Sharif K, Emery P, Bridgewood C. Pulmonary intravascular coagulopathy in COVID-19 pneumonia–Authors’ reply. Lancet Rheumatol 2020; 2:e60–61.

Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. J Thromb Haemost 2020; 18:844–47.

Ackermann M, Verleden SE, Kuehnel M, et al. Pulmonary vascular endotheliopathy, thrombosis, and angiogenesis in COVID-19. N Engl J Med 2020; 383:120–28.

McGonagle D, Sharif K, O’Regan A, Bridgewood C. Bridgewood CJAr. The role of cytokines including interleukin-6 in COVID-19 induced pneumonia and macropathogia activation syndrome-like disease. Autoimmun Rev 2020; 19:102537.

Foley JL, Conway EM. Conway EMJC. Cross talk pathways between coagulation and inflammation. Circ Res 2016; 118:1392–408.

Rakpiewicz AV, Mai X, Carsons SE, et al. Megakaryocytes and platelet-fibrin thrombi characterize multi-organ thrombosis at autopsy in COVID-19: a case series. EClinicalMedicine 24:100434.

Dwiputra Hermangruto K, Novembrino UT, Dorami H, et al. Thromboembolic involvement and its possible pathogenesis in COVID-19 mortality: lesson from post-mortem reports. Eur Rev Med Pharmacol Sci 2021; 25:1670–79.

Zuo Y, Yalavarthi S, Shi H, et al. Neutrophil extracellular traps in COVID-19. JCI Insight 2020; 5:138999.

Busch MH, Timmernans SAMEG, Nagy M, et al. Neutrophils and contact activation of coagulation as potential drivers of COVID-19. Circulation 2020; 142:1787–90.

Goldsmith CS, Miller SE, Martines RB, Bullock HA, Zaki SR. Electron microscopy of SARS-CoV-2: a challenging task. Lancet 2020; 395:e99.

Ahmet-Shala B, Peaceock TP, Baillon L, et al. Resistance of endothelial cells to SARS-CoV-2 infection in vitro. bioRxiv 2020; published online Nov 9. https://doi.org/10.1101/2020.11.08.372581 (preprint).

McCracken IR, Sagin G, He L, et al. Lack of evidence of angiotensin-converting enzyme 2 expression and replicative infection by SARS-CoV-2 in human endothelial cells. Circulation 2021; 143:865–68.

Stahl K, Bräsen JH, Hoepner MM, David S. Absence of SARS-CoV-2 RNA in COVID-19-associated intestinal endothelialitis. Intensive Care Med 2021; 47:359–60.

McCracken IR, Sagin G, He L, et al. Lack of evidence of angiotensin-converting enzyme 2 expression and replicative infection by SARS-CoV-2 in human endothelial cells. Circulation 2021; 143:865–68.

Wölfel R, Corman VM, Guggemos W, et al. Virological assessment of hospitalized patients with COVID-2019. Nature 2020; 581:465–69.

Berzategui-Cabreja J, Salto-Alejandre S, Valerio M, et al. SARS-CoV-2 RNAemia is associated with severe chronic underlying diseases but not with nasopharyngeal viral load. J Infect 2021; 82:e38–41.

Sholzberg M, Tang GH, Raahil H, et al. Heparin for moderately ill patients with COVID-19. medRxiv 2021; published online July 12. https://doi.org/10.1101/2021.07.08.21255311 (preprint).

Hatem M, Christensen R, Bang D, et al. 2018 update of the EULAR recommendations for the management of Behçet’s syndrome. Ann Rheum Dis 2018; 77:808–18.

Zuo Y, Estes SK, Ali RA, et al. Prothrombotic autoantibodies in serum from patients hospitalized with COVID-19. Sci Transl Med 2020; 12:eabd876.
92 Bastard P, Rosen LB, Zhang Q, et al. Autoantibodies against type I IFNs in patients with life-threatening COVID-19. Science 2020; 370: eabd4585.
93 Javidi E, Magnus T. Autoimmunity after ischemic stroke and brain injury. Front Immunol 2019; 10: 686.
94 Liao Y-H, Cheng X. Autoimmunity in myocardial infarction. Int J Cardiol 2006; 112: 21–26.
95 Zuo Y, Estes SK, Ali RA, et al. Prothrombotic autoantibodies in serum from patients hospitalized with COVID-19. Sci Transl Med 2020; 12: eabd1876.
96 Wang EY, Mao T, Klein J, et al. Diverse functional autoantibodies in patients with COVID-19. Nature 2021; 595: 283–88.
97 Altmann DM, Boyton RJ. SARS-CoV-2 T cell immunity: specificity, function, durability, and role in protection. Sci Immunol 2020; 5: eabd6160.
98 Chua RL, Lukassen S, Trump S, et al. COVID-19 severity correlates with airway epithelium-immune cell interactions identified by single-cell analysis. Nat Biotechnol 2020; 38: 970–79.
99 Weinheimer VK, Becher A, Tönjes M, et al. Influenza A viruses target type II pneumocytes in the human lung. J Infect Dis 2012; 206: 965–74.
100 Janssen NAF, Grondman I, de Nooijer AH, et al. Dysregulated innate and adaptive immune responses discriminate disease severity in COVID-19. J Infect Dis 2021; 223: 1322–33.

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