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A new paraprobiotic-based treatment for control of *Haemonchus contortus* in sheep

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**ABSTRACT**

*Haemonchus contortus* is a critical parasite of goats and sheep. Infection by this blood-feeding gastrointestinal nematode (GIN) parasite has significant health consequences, especially in lambs and kids. The parasite has developed resistance to virtually all known classes of small molecule anthelmintics used to treat it, giving rise in some areas to multidrug resistant parasites that are very difficult to control. Thus, new anthelmintics are urgently needed. *Bacillus thuringiensis* (Bt) crystal protein 5B (Cry5B), a naturally occurring protein made by a bacterium widely and safely used around the world as a bioinsecticide, represents a new non-small molecule modality for treating GINs. Cry5B has demonstrated anthelmintic activities against parasites of monogastric animals, including some related to those that infect humans, but has not yet been studied in a ruminant. Here we show that *H. contortus* adults are susceptible to Cry5B protein in vitro. Cry5B produced in its natural form as a spore-crystal lysate against *H. contortus* infections in goats had no significant efficacy. However, a new Active Pharmaceutical Ingredient (API) paraprobiotic form of Cry5B called IBaCC (Inactivated Bacterium with Cytosolic Crystals), in which Cry5B crystals are encapsulated in dead Bt cell wall ghosts, showed excellent efficacy in vitro against larval stages of *H. contortus* and relative protein stability in bovine rumen fluid. When given to sheep experimentally infected with *H. contortus* as three 60 mg/kg doses, Cry5B IBaCC resulted in significant reductions in fecal egg counts (90%) and parasite burdens (72%), with a very high impact on female parasites (96% reduction). These data indicate that Cry5B IBaCC is a potent new treatment tool for small ruminants in the battle against *H. contortus*.

**1. Introduction**

There are many species of gastrointestinal nematodes (GINs) that parasitize small ruminants, such as sheep and goats. The main GIN group of concern for producers and veterinarians is the trichostrongyles, common nematode parasites that cause major production loss and disease. They possess a direct life cycle, where the adults in the host produce eggs that are shed in the feces. The eggs develop to third stage larvae, or L3, in the feces and then circulate into the ambient environment. The infective L3 stage (L3i) is ingested by the host and develops into an egg-laying adult in the gastrointestinal tract of the animal (Scott and Sutherland, 2009). One species stands out as particularly...
pathogenic due to the fact it feeds on blood. This parasite, *Haemonchus contortus*, is an abomasal species that can cause severe anemia, hypoproteinemia, weight loss, lethargy, rough hair coats, poor wool/milk/-meat production, and death in heavily infected individuals (Zajac, 2006; Gilleard, 2013). Lambs and kids are at the greatest risk of developing disease due to their immature immune response and high rate of infection from environmental contamination due to peri-parturient egg production by infected ewes and does (Getachew et al., 2007).

Until recently, this parasite was controlled almost exclusively with anthelmintics. Unfortunately, the wide and frequent use of these drugs has created selection pressure that favors individual worms resistant to these anthelmintics (Kaplan, 2004a). The offspring of the adult parasites that survive treatment make up greater and greater portions of the worm population as more treatments are administered, leading to a resistant population and treatment failures (Jackson and Coop, 2008). Resistance in small ruminant GINs has been reported to all widely available anthelmintics, including multidrug resistance (Kaplan, 2004b; Wostenholme et al., 2004; Howell et al., 2006; Kaplan and Vidyashankar, 2012). For each major class of anthelmintic, resistance was reported within a decade of the drug becoming commercially available (Kaplan, 2004b); very recently resistance was seen within a few short years of introduction of monepantel, which belongs to one of the newest classes of anthelmintics for *H. contortus* control (Mederos et al., 2014; Van den Brom et al., 2015). Thus, anthelmintics involving new mechanisms of action are needed.

We have shown that the nematode-intoxicating crystal (Cry) protein Cry5B made by the soil bacterium *Bacillus thuringiensis* (Bt) has broad in vivo anthelmintic properties against GINs in monogastric animals (mice, hamsters, dogs, and pigs (Cappello et al., 2006; Hu et al., 2010a; Urban et al., 2013; Hu et al., 2018b)). Furthermore, Bt Cry proteins related to Cry5B are used safely, globally, and extensively as the number one Cry protein in transgenic crops (Karthickumar and Balasubramanian, 2017; Xiao and Hamer, 2013). Lambs and kids are at the greatest risk of developing infection. At 50 days post inoculation (dpi), the goats were euthanized and adult worms were screened from the abomasal contents then pan-ned using a glass bowl and fine forceps to remove the worms. Tubes containing about 20–30 adult worms were placed in 50 ml of RPMI-1640 media with antibiotics after washing by sedimentation at least 10 times in sterile media and shipped overnight to the Aroian laboratory. In vitro assays were carried out as described for hookworms using the same medium, same temperature, and same CO2 concentration with the addition of 25 mM HEPES pH 7.2 (Cappello et al., 2006; Hu et al., 2012, 2013a) in a 24-well plate with volume of 500 μL, four adult *H. contortus* per well, and two wells per dose. Cry5B was purified from spore crystal lysates as described (Griffitts et al., 2001). Adult parasites were scored once per day as live/dead as described (dead = no motility even after repeated touching); (Hu et al., 2013a; Urban et al., 2013).

2. Materials and methods

2.1. Animal approvals

All protocols in the study were approved by the Virginia Tech Institutional Biosafety Committee (IBC, Protocol #17–006) and the Institutional Animal Care and Use Committee (IACUC, Protocol #18–135, #18–141) and USDA Beltsville IACUC #12–025 and IBC #271.

2.2. *H. contortus* in vitro assays

Parasite eggs were isolated from infected sheep stool as described (Mes et al., 2007). Egg-to-larval assays (larval development assays) were carried out as described for *C. elegans* and cyathostome eggs (Hu et al., 2010b, 2018a). Briefly, parasite eggs were placed in S Medium in a 96-well format supplemented with *Escherichia coli* OP50 and incubated at 25 °C for seven days. The total number that developed to the L3i stage were counted. Feces from sheep experimentally mono-infected with *H. contortus* were shipped overnight from Virginia Tech to the Aroian Lab for these studies. Cry5B IBaCC was prepared as described (Li et al., 2020).

Adult *H. contortus* (Isolate Hc/2004A originally provided by Dr. Ray Kaplan under an MTA between the University of Georgia and USDA/ARS) was isolated from French Alpine goats. This strain is resistant to 3 major classes of anthelmintics 1 benzimidazole (fenbendazole, albendazole, others), 2 imidazothiazole/tetrahydropyrimidine (levamisole, moranil, others) and 3 avermectin (ivermectin, doramectin, others), and slightly resistant to moxidectin. Three goats each at approximately three months of age were orally inoculated with a single dose of 5000 *H. contortus* L3i and maintained for an additional 50 days (primary infection). At 50 days post inoculation (dpi), the goats were euthanized and adult worms were screened from the abomasal contents then pan-ned using a glass bowl and fine forceps to remove the worms. Tubes containing about 20–30 adult worms were placed in 50 ml of RPMI-1640 media with antibiotics after washing by sedimentation at least 10 times in sterile media and shipped overnight to the Aroian laboratory. In vitro assays were carried out as described for hookworms using the same medium, same temperature, and same CO2 concentration with the addition of 25 mM HEPES pH 7.2 (Cappello et al., 2006; Hu et al., 2012, 2013a) in a 24-well plate with volume of 500 μL, four adult *H. contortus* per well, and two wells per dose. Cry5B was purified from spore crystal lysates as described (Griffitts et al., 2001). Adult parasites were scored once per day as live/dead as described (dead = no motility even after repeated touching); (Hu et al., 2013a; Urban et al., 2013).

2.3. In vivo goat study

Male Sire (Boer) x Dam (Kiko x Savanna x Boer) goats 19–28 weeks of age were purchased locally before relocating to a pasture at the Beltsville Agricultural Research Center in early spring; the pasture had been free of livestock for the previous three years. Goats on pasture were supplemented ad libitum with orchard grass hay. The goats had been treated with moxidectin by the producer 26 days before shipment and received a CDT vaccine at 26 and 5 days prior to arrival in Beltsville. Initial egg counts from feces were evaluated as described (Gasbarre et al., 2015). Upon arrival in Beltsville, the goats showed strongylo eggs at <50 eggs per gram of feces or egg in 7 of 24 goats (the remainder had no strongylo eggs), Moniezia proglottids in 7 of 24, Strongyloides eggs in 10 of 24 goats, *Trichuris* eggs in 1 of 24 goats and coccidia oocysts in all samples. The average initial body weight was 27.3 kg (22–34 kg range). All goats were given a subcutaneous injection of 200 mg of ceftiofur hydrochloride upon arrival in Beltsville.

*Haemonchus contortus* L3i were obtained from sheep infected at Virginia Tech (see below for details) and sent overnight as a suspension to Beltsville. The goats were each subsequently given an oral inoculation of 5000 *H. contortus* L3i per animal two weeks after arrival and placement on pasture. Weights and fecal samples were collected after three weeks to evaluate the acquisition of parasite infection and changes in weight. The average weight at that time was 26.8 kg (19–33 kg range) with an average strongylo egg of 5843 (8–21,249 range). Full fecal egg counts of all relevant time points are given in Fig. S1. Two weeks later, goats were relocated to an open-aired concrete-surfaced enclosure and randomly assigned to three groups of eight designated as 1) untreated,
2) Cry5B-SCL-treated twice (40 mg/kg body weight once per day one day apart) and 3) moxidectin-treated (single dose 0.2 mg/kg body weight), with groups 2 and 3 inoculated by oral gavage. Cry5B SCL was prepared as described (Marronqiu et al., 2006; Hu et al., 2010a). The goats were fed orchard grass hay ad libitum and a supplement of pelleted grain concentrate while on the enclosure.

For oral dosing, animals were manually restrained, and the anthelmintic suspensions were given with a 50 ml syringe and ball-tipped inoculating needle. Animals were observed daily for any adverse reactions; none were seen throughout the course of the study. The necropsy of the goats was conducted over three days between eight and 10 days after moxidectin treatment and after the initial treatment with a two-dose regimen of Cry5B-SCL based on the logistics of availability of facilities and personnel. After euthanasia, the contents of the abomasum and small intestine, a rectal fecal sample, and tissue from the large intestine were collected. Fecal egg counts (Gasbarre et al., 2015) were determined from feces taken from the rectum at necropsy. The abomasum and small intestine contents were evaluated for parasitic burden using a modified agar gel method originally used to detect luminal parasites of swine (Slotved et al., 1996; Urban et al., 2013). Very few parasites were found in the contents from the small intestine. Aliquots of samples of abomasum contents collected from the gel after a 3-h incubation in normal saline at 37°C were examined for worms using a dissecting microscope and gridded Petri dish. All worms present from the abomasum contents were identified as adult *H. contortus* based on morphologic characteristics.

2.4. In vitro rumen fluid stability study

Cry5B crystals were isolated from Cry5B IBaCC (Li et al., 2020) and incubated in duplicate in cow rumen fluid for 12, 24, or 48 h as described (Goesser, 2008; Goesser et al., 2009; Goesser and Combs, 2009). Incubations were carried out courtesy of Zinpro Corporation. To determine any percent degradation of Cry5B, the amount of full length Cry5B was quantitated by SDS PAGE relative to BSA standards and compared to the total amount used in the in vitro assay.

2.5. In vivo sheep study

Dorset, Suffolk, and Dorset/Suffolk cross lambs aged 7–8 months were used for this study. The six female and six castrated male lambs had been previously pastured but were maintained in housing for the duration of the study to prevent additional natural *H. contortus* exposure. All animals were orally administered albendazole (7.5 mg/kg), ivermectin (0.2 mg/kg), and levamisole (8.0 mg/kg) sequentially to remove the abomasum and small intestine, a rectal fecal sample, and tissue from the large intestine were collected. Fecal egg counts (Gasbarre et al., 2015) were determined from feces taken from the rectum at necropsy. The abomasum and small intestine contents were evaluated for parasitic burden using a modified agar gel method originally used to detect luminal parasites of swine (Slotved et al., 1996; Urban et al., 2013). Very few parasites were found in the contents from the small intestine. Aliquots of samples of abomasum contents collected from the gel after a 3-h incubation in normal saline at 37°C were examined for worms using a dissecting microscope and gridded Petri dish. All worms present from the abomasum contents were identified as adult *H. contortus* based on morphologic characteristics.

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2.6. Statistics

All graphs and analyses were generated using GraphPad Prism v. 8. The comparisons between multiple groups in the goat study were carried out using one-tailed Dunnett’s multiple comparison test, comparing to control. The comparison of FEC in the sheep study (Fig. 5B) was carried out using two-way analysis of variance (mixed-effects analysis Time x Treatment, as there was one data point missing for one sheep). All comparisons between two groups were carried out using one-tailed Mann-Whitney test with the assumption that treatment would reduce the infection, except for starting FEC for the sheep study (two-tailed Mann-Whitney to test for equality at the beginning).

3. Results

3.1. Cry5B is active against *H. contortus* adults in vitro

We first determined if Cry5B on its own was active against *H. contortus*. Adult parasites from a triple anthelmintic-resistant strain of *H. contortus* maintained at the USDA were isolated from French-Alpine goats and set up for testing an in vitro dose-response using Cry5B purified from spore-crystal lysates (SCLs) (Griffitts et al., 2001, 2005; Cappello et al., 2006). We found that purified Cry5B intoxicates triple anthelmintic-resistant adult *H. contortus* (Fig. 1).

3.2. Cry5B spore crystal lysate was not effective against *H. contortus* in goats

We next tested whether Cry5B spore crystal lysates could be used against an experimental *H. contortus* infection in goats. SCLs are formed when Bt sporulates, producing crystal(s) in the cytosol of the mother cell. The cell subsequently lyses and releases the crystal(s) and the spore. SCLs are the form of Bt Cry proteins used in topical applications worldwide as insecticides (Roh et al., 2007; Koch et al., 2015). Goats experimentally infected with *H. contortus* larvae were treated with either nothing (placebo, negative control), Cry5B SCL (40 mg/kg) given twice one day apart *per os*, or a single dose of moxidectin (0.2 mg/kg; positive control). Moxidectin treatment resulted in a statistically significant drop in the number of worms in the entire 20% aliquot of abomasal contents from each animal was counted using a dissecting microscope and gridded Petri dish. All worms present were identified as adult *Haemonchus contortus* based on morphologic characteristics.
in parasite burdens and fecal egg counts (FEC), while Cry5B SCL treatment did not, although both parasite burdens and FEC for Cry5B treated goats were slightly lower than control (see Fig. 2).

3.3. The paraprobiotic Cry5B IBaCC is effective against H. contortus larvae and crystals are stable in rumen fluid

We recently developed a new Active Pharmaceutical Ingredient (API) containing Cry5B called IBaCC for inactivated bacterium with cytosolic crystal (Fig. 3A (Li et al., 2020)). In this form, Cry5B is expressed in a sporulation defective Bt strain under control of a non-sporulating promoter, giving rise to production of a Cry5B crystal inside a vegetative Bt cell. This vegetative Bt is then inactivated (killed) with food-grade plant essential oil (monoterpenoid), giving rise to a dead bacterial cell wall ghost, harboring within its peptidoglycan shell a Cry5B crystal that is fully bioactive. Such a dead probiotic is known as a paraprobiotic (Taverniti and Guglielmetti, 2011). An appropriate negative control for IBaCC is IBa (inactivated bacterium). IBa is identical to IBaCC in all ways except that the plasmid harboring the Cry5B gene has no insert (empty vector). The parent Bt strain, plasmid backbone, and all processing steps are otherwise identical for IBa and IBaCC.

Although we were no longer able to access H. contortus adults for Cry5B efficacy testing, we were able to obtain fecal samples from H. contortus-infected sheep to evaluate larval development in vitro in the presence/absence of Cry5B IBaCC. From these fecal samples, we isolated parasite eggs and carried out egg to larval development assays in a 96 well format culture system over the course of 7 days as described (Hu et al., 2018a). Three independent experiments were carried out, looking to see how many H. contortus eggs developed to the infective L3 stage (L3i) in 7 days at 28 °C. As shown (Fig. 3B), H. contortus larval development is exquisitely sensitive to Cry5B IBaCC, showing near complete (98%) inhibition at 56 ng/mL. These results were confirmed with an independent batch of IBaCC, showing complete (100%) inhibition of L3i development at 37.5 ng/mL (Fig. S2).

These results demonstrated that IBaCC was potent against H. contortus larvae. Since the goat study failed to show an effect of Cry5B on H. contortus, we were concerned that Cry5B crystals might not be stable during passage through the rumen. To determine if Cry5B from IBaCC might be stable in a ruminant, Cry5B crystals isolated from Cry5B IBaCC were incubated in cow rumen fluid in vitro for 12, 24, and 48 h. No appreciable degradation was seen at 12 and 24 h (Fig. 4). At 48 h, approximately 40% of the Cry5B was degraded, indicating that Cry5B crystals in IBaCC would be stable up to 24 h in the rumen after ingestion and passage to the abomasum.

3.4. Cry5B IBaCC is effective against H. contortus infestations in sheep

Based on Cry5B IBaCC efficacy against H. contortus larvae and on stability studies with Cry5B IBaCC crystals in rumen fluid, we tested the efficacy of Cry5B IBaCC against H. contortus infections in sheep (Fig. 5A). Twelve sheep were experimentally inoculated with H. contortus L3i. Fecal samples were taken and pre-treatment FEC were determined on the day of treatment with IBaCC. Six sheep were given
water (placebo control) and six sheep were given 60 mg/kg Cry5B in IBaCC daily for three consecutive days. Fecal egg counts were determined daily (on days when treatment occurred, fecal samples were taken before treatment). The sheep were euthanized seven days after the first treatment to determine the worm burden in the abomasum.

Sharp reductions in FEC were seen within two days after treatment that was statistically significant (Fig. 5B). Although starting FEC were similar in both control and treated groups, by the end of the study they were statistically much lower in the treated group (Fig. 5C). Between 88% and 96% reductions in FEC were seen relative to untreated controls starting three days after the first treatment.

By the end of the study, parasite burdens in Cry5B IBaCC treated sheep were significantly reduced 72% relative to infected but untreated control sheep (Fig. 6A). The larger reduction in FEC relative to reduced adult worm burden was likely due to a 96% reduction in female worms versus a 60% reduction in male worms (both statistically significant; Fig. 6B and C).

4. Discussion

New therapies against *H. contortus* in small ruminants are desperately needed due to the highly pathogenic nature of the parasite and its propensity for developing anthelmintic drug resistance to small molecules. Here, we have shown *in vitro* that Cry5B protein intoxicates adult *H. contortus* parasites and is very potent against *H. contortus* larval development (complete inhibition ~50 ng/mL). We have presented data showing for the first time that a Bt Cry protein, Cry5B, when produced as part of the paraprobiotic IBaCC (crystal encapsulated within the shell of an inactivated bacterium), significantly reduced *H. contortus* infection in sheep. Three 60 mg/kg doses of Cry5B in IBaCC cleared >70% of the adult parasites and 96% of the female parasites, which led to a comparable reduction in parasite FEC. Although three doses were given, further testing may show that 1–2 doses can achieve a biologically significant effect that reduces infection transmission. Further dosing studies to determine the minimal effective dose and development of formulations to promote delivery to the abomasum and posterior GI tract are warranted.

![Figure 5. Experimental design and FECs of curative sheep study with IBaCC (n = 6 sheep/group). (A) Experimental design of sheep study. (B) Fecal egg counts (FECs) over time (eggs per gram of feces) relative to the day of first treatment for control (water) and treated (IBaCC) groups (six sheep per group). FECs were always determined before treatment on any given day. The difference between fecal egg counts between control and treated groups based on two-way analysis of variance (P = 0.0009) was significant. (C) Comparison of starting and ending FECs for both groups.](image1)

![Figure 6. *H. contortus* abomasal parasite burdens in curative sheep study with IBaCC. (A) Total parasite burdens in control vs IBaCC treated sheep. (B) Male parasite burdens in control vs IBaCC treated sheep. (C). Female parasite burdens in control vs IBaCC treated sheep. Data from one sheep in the control group was not collected (see Materials and Methods).](image2)
Our data show that Cry5B crystals from IBaCC will survive the rumen and that *H. contortus* adults can ingest Cry5B since they are intoxicated in vitro. How Cry5B is ingested by *H. contortus* in the abomasum is an area for investigation but there is precedent for a blood feeder accessing Cry5B in vivo with hookworms (Kalkofen, 1970; Cappello et al., 2006; Hu et al., 2012, 2013b, 2018a, 2018b; Wu et al., 2015; Li et al., 2020).

Although Cry5B IBaCC was effective in *H. contortus*-infected sheep, we found that Cry5B delivered as part of Bt spore crystal lysates (SCLs) had no significant impact on *H. contortus* infections in goats. The reasons for these disparate results are yet unclear but could include: 1) differences between goats and sheep; 2) differences in dosing (2 × 40 mg/kg versus 3 × 60 mg/kg); and/or 3) differences between spore crystal lysates and IBaCC and the ability of IBaCC and/or Cry5B crystals in IBaCC to better survive rumen conditions before entering the abomasum to intoxicate *H. contortus*.

The mechanism of action of Cry5B has been studied extensively in *C. elegans* and to some extent in hookworms and *Ascaris*, where the mechanism of action was shown to be conserved with *C. elegans* (Cappello et al., 2006; Hu et al., 2012; Urban et al., 2013). Cry5B, which is a three-domain Cry protein, needs to be ingested by the target nematode to act, following which it binds to invertebrate-specific glycosphin-golipids on nematode intestinal cells and forms pores, resulting in intoxication, dysfunction, and death as has been demonstrated in insects targeted by three-domain Cry proteins (Griffitts et al., 2001, 2003, 2005; Huffman et al., 2004; Griffitts and Aroian, 2005; Barrows et al., 2007; Bischof et al., 2008; Kao et al., 2011; Los et al., 2011; Hui et al., 2012). The mechanism of action against *H. contortus* is likely the same and can be subjected to more study in the future (e.g., investigation of Cry5B binding to *H. contortus* intestinal glycosphin-golipids).

Other studies have examined the impact of Bt and Cry proteins on *H. contortus*. One group showed that a Cry5B-containing strain of Bt was toxic to *H. contortus* adults and larvae (Kotze et al., 2005). Although this study did not confirm that the anthelmintic activity was due to Cry5B since other Cry proteins were also produced by this strain, the level of toxicity against larvae and adults in vitro was remarkably similar to that shown here (Figs. 1 and 3), suggesting that Cry5B was likely the active component in those studies. In other studies, either the toxicity seen (e.g., against larvae) was 2–3 orders of magnitude less effective than shown here, or the doses used were not indicated, or the efficacy could not be attributed to Cry protein (e.g., activity was seen against non-feeding egg stages or activity seen when soluble fractions were injected by intra-peritoneal or intramuscular routes) (Lopez-Arellano et al., 2002; Lopez et al., 2006; Linares et al., 2008; Vázquez-Pineda et al., 2010; Sinott et al., 2012; de Lara et al., 2016; Beena et al., 2019).

One interesting finding here was the higher sensitivity of female versus male *H. contortus* in vivo. Since Cry proteins need to be ingested to intoxicate, this may mean that the females are feeding more than males or their feeding behavior is more conducive towards ingesting Cry proteins. The significant difference in size between female and male worms are consistent with the former. Alternatively, the metabolic requirements of egg-producing females may make them more sensitive to intoxication.

Development of anthelmintic resistance in *H. contortus* is an important consideration. In this regard, Cry5B may have some advantages. Forward genetic screens for resistance showed that it is 3X more difficult to target nematode toxicity (e.g., feeding) versus male

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