INTRODUCTION

The genetic pathogenesis remains unclear in most congenital hypothyroidism (CH) cases. Familial cases are associated with genetic defects to a certain extent, approximately 20% CH cases being caused by inherited defects in the pathway of thyroid hormone synthesis.\(^1,2\)

A critical step in the synthesis of thyroid hormone is the generation of hydrogen peroxide (H\(_2\)O\(_2\)). H\(_2\)O\(_2\) is produced by dual oxidase (DUOX)/dual oxidase maturation factor (DUOXA) at the apical membrane of follicular thyroid cells, and is used as a substrate by thyroid peroxidase (TPO) in the organification of iodide and incorporation of iodine into thyroglobulin. DUOXA is essential for DUOX maturation and activation, thereby determining normal DUOX function.\(^3,4\)

To date, mutations in the DUOX2/DUOXA2 system have been identified to be associated with CH, and the system is one of the important candidate genes for CH.\(^5-9\) In addition, the most prevailing causes of goitrous congenital hypothyroidism (GCH) are TPO defects. Until now, more than 90 inactivating mutations in the TPO gene have been reported.\(^2,10-12\)

Currently, the genetic defects in CH are not fully understood. In this study, the DUOX2, DUOXA2, and TPO genes were considered for screening genetic defects in a girl and her younger sister with GCH.

CASE REPORT

The girl and her sister with GCH were recruited for molecular analysis of mutations. CH was diagnosed on the basis of serum levels of thyrotropin, free thyroxine and free triiodothyronine at the neonatal screening stage. Thyroid ultrasound demonstrated that both the girl and her sister had goiter. Their parents were not consanguineous and had normal thyroid function (Table 1). The girls were regularly followed up. Until now,
they had no obvious complications. This study was approved by the Ethics Committee of the hospital. Written informed consent was obtained.

Venous blood samples were obtained from the girls. Genomic DNA was extracted by using the phenol-chloroform extraction method from peripheral blood leukocytes. Primers were designed to target the flanking intron regions of the exons. All exons of the DUOX2 (MIM# 606759, GenBank NM_014080.4), DUOXA2 (MIM# 612772, GenBank NM_207581.3), and TPO (MIM# 606765, GenBank NM_000547.5) genes were amplified by PCR. PCR products were purified and Sanger sequenced directly for variance analysis. If an mutation was identified, the target fragment was also amplified in the girl’s parents and in 105 control individuals.

As shown in Fig. 1, the genetic analysis demonstrated that the two girls carried the same compound heterozygous mutations: G to T transversion at 2654 position \([c.2654G>T (p.\text{Arg885Leu}, p.R885L)]\) in the exon 20 of the DUOX2 gene and C to G transversion at nucleotide position 738 \([c.738C>G (p.Y246X)]\) in the exon 5 of DUOXA2 gene.

The missense mutation \(p.R885L\) is presented in 1000 Genomes Project database, but not found in Pubmed and con-

| Table 1. Thyroid Blood Tests and Clinical Data of the Family in September 2015 |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Variables       | Normal range    | III 1*          | III 2*          | II 1           | II 2           | I 1            | I 2            |
| Age (yr)        | /               | 2.5            | 0.5            | 28            | 30            | 49            | 60            |
| Height (cm)     | /               | 87             | 60             | 160           | 169           | 160           | 160           |
| Weight (kg)     | /               | 11             | 6.4            | 55            | 65            | 60            | 60            |
| Thyrotropin (mIU/mL) | 0.34–5.44   | >100.00        | >100.00        | 2.13          | 2.73          | 1.66          | 1.24          |
| Free triiodothyronine (pmol/L) | 2.92–5.93       | 2.00           | 4.64           | 3.93          | 5.19          | 4.10          | 5.32          |
| Free thyroxine (pmol/L) | 7.91–20.59       | 1.30           | 5.82           | 16.88         | 12.75         | 16.00         | 16.70         |
| Thyroid size    | /               | Goiter         | Goiter         | Normal        | Normal        | Normal        | Normal        |

*Thyroid function tests at the neonatal screening stage.

c.738C>G (p.Y246X) in the exon 5 of the DUOXA2 gene

| Mutant type | Wild type |
|-------------|-----------|
| 244 Thr     | 244 Thr   |
| 245 Gln     | 245 Gln   |
| 246 Tyr/Stop| 246 Gly   |
| 247 Ala     | 247 C     |
| 248 C       | 248 C     |

c.2654G>T (p.R885L) in the exon 20 of the DUOX2 gene

| Mutant type | Wild type |
|-------------|-----------|
| 881 Phe     | 881 Phe   |
| 882 Thr     | 882 Thr   |
| 883 Met     | 883 Met   |
| 885 Arg/Leu | 885 Arg   |

Fig. 1. Genotypes indicating the pedigrees with congenital hypothyroidism due to the coexistence of heterozygous mutations, c.2654G>T (p.R885L) in the DUOX2 gene and c.738C>G (p.Y246X) in the DUOXA2.
controls. Bioinformatics analysis with Mutation Taster, PolyPhen-2 and PROVEAN revealed that the p.R885L mutation is capable of causing disease (Mutation Taster score of 120, prob ≥0.99), is most likely damaging (PolyPhen-2 score of 0.993), or is likely to have deleterious effects (PROVEAN score of -6.040).

Heterozygous mutations were detected in the family members. Pedigree analysis showed that the pattern was consistent with autosomal recessive inheritance. No mutations in the TPO gene was detected in our pedigree. None of the controls showed the same pathogenic variants.

**DISCUSSION**

In the present study, mutations in the DUOX2, DUOXA2, and TPO genes were screened in the two patients with GCH. The compound heterozygous mutations, p.R885L in the DUOX2 gene and p.Y246X in the DUOXA2, were detected. Pedigree analysis revealed an autosomal recessive inheritance pattern.

Mutations in the DUOX2/DUOXA2 heterodimer are the common genetic defects of CH. Previous studies reported four mutations in the DUOX2 (p.I26M, p.Y138X, p.C189R, and p.Y246X) associated with CH. The patients with homozygous nonsense mutation (p.Y246X) exhibited mild permanent CH and goiter, which is similar to our patients. The p.Y246X mutation of DUOX2 leads to a complete loss of function in the homozygous patient, thus ensuring to a secondary deficiency of DUOX2 activity. Together with our results, the findings indicate that the p.Y246X mutation frequently occurs in Chinese population, and the frequency of affected homozygous for p.Y246X is estimated to be one of 34000 newborns in Chinese population.

To date, over 40 mutations in the DUOX2 gene have been described to be correlated with CH. Maruo, et al. firstly reported the p.R885Q (c.2654G>A) mutation in the DUOX2 gene. However, the patients in this study was found to have G to T transversion (c.2654G>T) at the same position, resulting in the replacement of an acidic residue at amino acid position 885 (p.Arg885Leu, p.R885L). In silico analysis revealed that this alteration (p.R885L) affected the structure/function of the DUOX2 protein and were capable of causing disease. To our best knowledge, p.R885L is the first ever reported in patients with CH. In addition, the two girls in our study presented with a normally located but enlarged thyroid gland. Their parents, each with a single heterozygous mutation, exhibited normal thyroid function. Our patients in the present study developed no physical or cognitive developmental defects; Timely and effective treatment contributed to the satisfactory results.

In conclusion, the present study identified two missense mutations in two patients with CH: c.2654G>T (p.R885L) in the DUOX2 gene and c.738C>G (p.Y246X) in the DUOXA2. The findings indicate the importance of molecular diagnosis and classification of CH.

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