Exercise activates the PI3K-AKT signal pathway by decreasing the expression of 5α-reductase type 1 in PCOS rats

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Hyperandrogenism and hyperinsulinemia are main clinical endocrine features of PCOS. Exercise can adjust the androgen level, as well as increase the sensitivity of insulin by activating PI3K-Akt insulin signaling pathways. So 5αR1 may be the target of androgen and insulin for exercise-induced regulation. To investigate the role of 5αR1 in the PI3K-Akt signaling pathway in skeletal muscle of PCOS rats activated by exercise, fifty-four female rats were randomly divided into the PCOS group (n = 42) and the control group (n = 12). After injection of testosterone propionate for 28 days, the remaining 36 rats in the PCOS group were randomly assigned to six groups: the sedentary group (PS, n = 6), sedentary and 5αR1 (5α-reductase inhibitor) group (PS + R1, n = 6), sedentary and 5αR2I (5α-reductase type 2 selective inhibitor) group (PS + R2I, n = 6), exercise group (PE, n = 6), exercise and 5αR1 group (PE + R1, n = 6), and exercise and 5αR2I group (PE + R2I, n = 6). The rats undergoing exercise were trained to swim for 14 days. Finasteride (5α-reductase type 2 selective inhibitor) and dutasteride (5α-reductase inhibitor) were administered once daily and were dosed based on weight. At the end, the expression of 5αR1 proteins, the phosphorylation level of PI3K and AKT, were determined by Western blot. The PCOS non-exercise group and the PE + R2I group displayed significantly lower phosphorylation of Akt, PI3K p85 and GLUT4 expression, while in the PE + R2I group, the level of Akt phosphorylation and PI3K p85 expression was significantly higher than that of the PCOS non-exercise group and the PE + R1 group. In summary, our study demonstrated that exercise can activate the PI3K/AKT signal pathway of PCOS rats by decreasing the expression of 5αR1.

Polycystic ovary syndrome (PCOS) is a clinically heterogeneous syndrome characterized by anovulation, clinical or biological hyperandrogenism and abnormalities of the ovary1. It is one of the most common endocrine diseases in women of reproductive age2. The main clinical endocrine features of this syndrome are hyperandrogenism and hyperinsulinemia1,3. Currently, a variety of medications (such as ovulation drugs, anti-androgen drugs, and insulin sensitizer) have been used for the treatment of PCOS; however, the majority of these drugs are directed only to one specific feature of this syndrome and there remains some difficulty in decreasing the level of androgen while improving insulin sensitivity. Exercise can simultaneously improve the two major endocrine characteristics of PCOS.

Exercise therapy can increase insulin sensitivity and decrease the level of androgen, and the effects of exercise therapy in PCOS patients are becoming increasingly important4–6. Obese PCOS patients can restore normal menstrual cycle and pregnancy after diet and exercise intervention, and if exercise therapy is performed prior to receiving assisted reproductive technology, the pregnancy rate can be significantly increased7,8. Previously, we have successfully generated an obesity rat model of PCOS. In these experiments, exercise and dietary intervention were used in the obese PCOS rats to demonstrate that exercise training improved insulin sensitivity and decreased androgen levels9. The signal transduction pathway of insulin and glucose metabolism has been extensively studied. In recent years, many scholars have paid close attention to the relationship between the

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Boda confirmed that the presence of polycystic ovary syndrome in women was due to an abnormal expression of group (PCOS, n = 42) and the Control group (control, n = 12). The PCOS rat model was generated by injecting testosterone propionate for 28 days and being fed a high-fat diet according to our previous studies. The day after the 28th injection, a total of 12 rats from the PCOS group and the Control group were sacrificed by cervical dislocation. When rats were sacrificed, blood samples were obtained from all the 12 rats and ovaries were dissected from connective tissue to verify the PCOS phenotype. Then six rats were included in each group.

The remaining 36 rats in the PCOS group were randomly assigned to six groups (Fig. 1): the sedentary group (PS, n = 6), sedentary and 5αR1 group (PS + RI, n = 6), sedentary and 5αR2 group (PS + R2, n = 6), exercise group (PE, n = 6), exercise and 5αR1 group (PE + RI, n = 6), and exercise and 5αR2 group (PE + R2, n = 6). All the remaining 42 rats were fed with an ordinary diet for two weeks. The exercised rats in the PE, PE + RI and PE + R2 groups were exercised for 5 min and 30 s on a treadmill with a 15° inclination. The PE group was exercised for 5 months, while the PE + RI and PE + R2 groups were exercised for 3 months and 1 month, respectively.

Androgen is a protein and an anabolic hormone that plays a role in promoting the use of energy reserves in muscle cells and participates in the signal transduction pathway of glucose metabolism in skeletal muscle cells. In skeletal muscle cultures from normal SD rats, increasing exogenous androgen concentration could modulate the activation of PKC and Akt. The activation of Akt could be inhibited by androgen related inhibitors. Thus, PKC/Akt/GLUT-4 is likely to be a common pathway for insulin and androgen signaling pathways in glucose metabolism. In 2014, the latest international research results showed that androgen did not interact with insulin in regulating the level of glucose metabolism of skeletal muscle in patients with PCOS. This was shown through direct glucose uptake experiments. This raises a contradiction and allows us to speculate that there is a genetic or protein expression deficit in PCOS patients and that there is lack of some androgen triggering mechanism of glucose metabolism.

An important metabolic enzyme in the body that regulates androgens is known as the 5α-reductase enzyme (5αR). Two types of isoenzymes named 5α-reductase type 1 (5αR1) and 5α-reductase type 2 (5αR2) are encoded by the 5α-reductase gene type 1 (SRD5A1) and the 5α-reductase gene type 2 (SRD5A2), respectively. In 2009, Boda confirmed that the presence of polycystic ovary syndrome in women was due to an abnormal expression of 5αR. A 2014 population study confirmed that 5αR blockers increased insulin resistance and were more likely to be after 5αR1 was inhibited. Population studies confirmed that 5αR1 has certain effects on insulin resistance. Exercise can adjust the androgen level, as well as increase the sensitivity of insulin. Thus, 5αR1 may be the target of androgen and insulin for exercise-induced regulation (Fig. 1).

In the present study, we activated PI3K-Akt insulin signaling pathways in the skeletal muscle of PCOS rats by exercise. To investigate the role of 5αR1 in the PI3K-Akt signaling pathway in skeletal muscle of PCOS rats activated by exercise, we compared dutasteride, a dual 5αR1 and 5αR2 inhibitor, with finasteride, a 5αR2 selective inhibitor.

Materials and Methods

Ethics statement. Fifty-four Wistar female rats (50–70 g, 21 days old) were purchased from the Beijing Vital River Laboratory Animal Technology Co. (Beijing, China). The animals were housed in temperature-controlled rooms (22 °C), with light from 08:00 to 20:00 h and dark from 20:00 to 08:00 h and had free access to water. The experimental protocols were approved by the Institutional Animal Care and Use Committees of Nanjing Medical University (permit number: NJMU/IA-CUC_20120210_01) and strictly followed. The experimental protocols were in strict accordance with the Guidelines for the Care and Use of Laboratory Animals (National Research Council of People's Republic of China, 2010).

Study procedure. All rats were randomly divided into two experimental groups (Fig. 2): the PCOS model group (PCOS, n = 42) and the Control group (control, n = 12). The PCOS rat model was generated by injecting testosterone propionate for 28 days and being fed a high-fat diet according to our previous studies. The day after the 28th injection, a total of 12 rats from the PCOS group and the Control group were sacrificed by cervical dislocation. When rats were sacrificed, blood samples were obtained from all the 12 rats and ovaries were dissected from connective tissue to verify the PCOS phenotype. Then six rats were included in each group.

The remaining 36 rats in the PCOS group were randomly assigned to six groups (Fig. 1): the sedentary group (PS, n = 6), sedentary and 5αR1 group (PS + RI, n = 6), sedentary and 5αR2 group (PS + R2, n = 6), exercise group (PE, n = 6), exercise and 5αR1 group (PE + RI, n = 6), and exercise and 5αR2 group (PE + R2, n = 6). All the remaining 42 rats were fed with an ordinary diet for two weeks. The exercised rats in the PE, PE + RI and
Figure 2. The experimental design. O: ordinary diet; F: high fat diet; T: testosterone propionate; Control: control group; PCOS: PCOS model group; S: no exercise; PS + R1: sedentary with 5 alpha R inhibitor in PCOS rats; PS + R2I: sedentary with 5 alpha R2 inhibitor in PCOS rats; E: exercise; PE + R1: exercise with 5 alpha R inhibitor in PCOS rats; PE + R2I: exercise with 5 alpha R2 inhibitor in PCOS rats.

PE + R2I groups swam simultaneously without a load for 120 min/d, 6 days/week for two weeks in a barrel filled with water maintained at 33–35 °C at a depth of 40–50 cm. Every rat was given enough area to swim freely. The cages of non-exercised rats in the PS, PS + RI and PS + R2I groups were placed nearby the barrels to eliminate the influence of exercise noise. Finasteride (5 mg/kg/day) and dutasteride (1 mg/kg/day) were administered by gavage in the PE + RI, PE + R2I, PS + RI and PS + R2I groups, once daily. All of the rats were sacrificed between 08:00 and 11:00 AM under fasting conditions 24 h after the end of the last training session with 3% pentobarbital abdominal anesthesia followed by cervical dislocation. Blood and skeletal muscle (mainly composed of gastrocnemius muscle) samples were collected from all animals.

Measurements of body weight and body length. The body weight and length of rats were measured weekly beginning at 21 days of age. The body length was defined as the distance from the nose to the anus of the rats. The Lee index reflects the body fat as a parameter

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LI = \frac{\text{body weight (g)}}{1000/\text{body length (cm)}}
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Oral glucose tolerance test (OGTT). Prior to sacrificing, all rats were fasted overnight and infused intragastrically with 2 g glucose per kilogram of body weight. Blood samples were collected at 0 min, 30 min, 60 min and 120 min after oral glucose was administered to evaluate fasting blood glucose (FBG), 30 min postprandial blood glucose (PBG30′), 1 h postprandial blood glucose (PBG60′) and 2 h postprandial blood glucose (PBG120′), respectively.

Serum analysis. Blood samples obtained from all rats fasting for 12 h were centrifuged at 2500 g for 10 min and stored at −80 °C. Fasting serum levels of insulin (FINS) and testosterone (T) were determined using commercial ELISA kits (Jiancheng Bioengineering Institute, Nanjing, P. R. China). Fasting blood glucose (FBG), 30 min postprandial blood glucose (PBG30′), 1 h postprandial blood glucose (PBG60′) and 2 h postprandial blood glucose (PBG120′) were analyzed by GOD-PAP. AUCglucose (Area under the curve of blood glucose) = 1/4 × FBG + 1/2 × PBG30′ + 3/4 × PBG60′ + PBG120′. The insulin resistance index is defined as the ratio of HOMA-IR (Homeostasis model assessment).

Western blotting analysis. One hundred milligrams of skeletal muscle from the gastrocnemius muscle of the different groups were stored at −80 °C and were homogenized in 10–20 volumes with a buffer containing 1% sodium dodecylsulfate (SDS), 100 mmol/L Tris HCl (pH 6.8), 1 mmol/L phenylmethyl sulfonyl fluoride (PMSF), and 0.1 mmol/L β-mer-captoethanol. The homogenate was centrifuged at 12000 × g for 10 min at 4 °C and supernatant A was collected. Then, supernatant A for the protein GLUT4 was centrifuged at 4 °C, 40000 × g for 1 h to precipitate the cell membrane debris. The supernatant was centrifuged again at 4 °C, 40000 × g for 10 min, with the proper amount of membrane protein extraction reagent to obtain the supernatant after high-speed mixing using a vortex mixer after 5 seconds. Then, the sample was placed in an ice bath precipitation for 5–10 minutes, which was repeated 3 times to obtain the full extraction of protein. At 4 °C, 40000 × g/15 min, supernatant B for PM GLUT4 was collected for the membrane protein solution. The protein concentration was determined by the BCA protein assay. For each sample, proteins were separated by electrophoresis via 10% sodium dodecylsulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The gel was transferred to a nitrocellulose (NC) membrane (Bio-Rad Instruments, CA, USA) in transfer buffer containing 25 mmol/L Tris, 192 mmol/L glycine, and 20% methanol. The NC membranes were blocked using PBS containing 2% bovine serum albumin (BSA) and 0.05% Tween-20 at 4 °C for 1 h at room temperature and incubated with primary antibody overnight. The following antibodies were used for the supernatant A: anti-Akt antibody, anti-Phospho-Akt(Ser473) antibody, anti-Phospho-Akt (Thr308) antibody, anti-PI3 Kinase p85 antibody (Cell Signaling Technology, Danvers, MA), anti-SRD5A1 antibody, and anti-GLUT4 antibody (Abcam, Cambridge, UK). The next day, membranes were incubated with goat anti-rabbit-HRP secondary antibody (Abcam, Cambridge, UK) at room temperature for 1 h. Photos were acquired using a gel imaging analysis system (Tanon 6600; Tanon Shanghai). Quantification analysis of blots was performed with Image-Pro plus 6.0 software. The same procedure was also for the supernatant B to detect the expression of PM GLUT4. The relative amount of target proteins were represented by the protein/β-actin gray-scale ratio and phosphorylation was expressed as the ratio of phosphorylated to total protein.
Immunohistochemical staining for GLUT4 protein expression. The formalin-fixed paraffin-embedded tissue sections were deparaffinized in xylene, followed by rehydration in decreasing ethanol concentrations at room temperature. Retrieval with high-temperature heating in citrate buffer (10 mM citrate, pH 6.0) was performed and the slides were incubated with inhibitor buffer (3% H2O2) and washed with PBS (0.01 mol/L). The slides were treated with goat anti-rat GLUT4 primary antibody (Abcam, Cambridge, UK) overnight at 4 °C. After another washing with PBS, the slides were incubated with rabbit anti-goat secondary antibody polymer-based En Vision-HRP-enzyme conjugate (Abcam, Cambridge, UK) at room temperature. The slides were observed under an optical microscope and 10 non-overlapping high-magnification fields were selected after being stained with 3,3′-diaminobenzidine (DAB). The average optical density values of every field were measured using Image-Pro plus 6.0 software.

Statistical analyses. Data were expressed as the means ± SD. Statistical evaluations were performed with SPSS software (version 16.0, SPSS, Chicago, IL, USA). All of the data were analyzed by T test or one-way ANOVA, and P < 0.05 was considered significant. When the ANOVA revealed significant differences among the three experimental groups, post hoc analysis was performed using least significant difference (LSD), and statistical difference was set at P < 0.05.

Results

Changes in metabolic and endocrine variables in a PCOS model induced by a high-fat diet and testosterone. The metabolic and endocrine variables were measured on the 28th day of the experiment in PCOS and Control groups. The Lee index in the PCOS group was significantly increased in comparison with the Control group. Serum levels of FINS and T were significantly increased in the PCOS group than in the Control group. The levels of FBG and PBG1 were significantly higher in the PCOS group than in the Control group. The levels of AUCglucose were significantly increased in the PCOS group compared to the Control group. The value of HOMA in the PCOS group was also significantly higher compared with the Control group (Table 1).

5αR1 expression in skeletal muscle of PCOS rats. Compared with the Control group, the expression of 5αR1 was significantly higher in skeletal muscle of the PCOS group (Fig. 3).

Changes in metabolic and endocrine variables in response to exercise and 5αR inhibition. The metabolic and endocrine variables were measured at the end of the experiment in the CO, PS, PS + RI, PS + R2I, PE, PE + RI and PE + R2I groups. Compared with the PS group, the levels of FINS were significantly decreased in the PE and PE + R2I groups. The levels of PBG120′ were significantly increased in the PS group compared with the CO group and they were significantly decreased in the PE + R2I group compared to the PS and PS + RI groups.

Table 1. Changes of metabolic and endocrine variables in PCOS model.

| Variable               | Control       | PCOS          | P (PCOS vs. Control) |
|------------------------|---------------|---------------|----------------------|
| Lee Index              | 295.44 ± 5.48 | 304.11 ± 5.25 | 0.039                |
| T (nmol/L)             | 31.58 ± 1.19  | 38.96 ± 5.18  | 0.025                |
| DHT (nmol/L)           | 29.72 ± 2.93  | 30.69 ± 2.87  | 0.597                |
| FINS (mU/L)            | 10.28 ± 2.02  | 11.35 ± 1.21  | 0.048                |
| FBG (mmol/L)           | 2.95 ± 0.33   | 3.88 ± 0.74   | 0.021                |
| PBG30′ (mmol/L)        | 7.86 ± 1.50   | 8.97 ± 1.87   | 0.315                |
| PBG60′ (mmol/L)        | 7.74 ± 0.90   | 10.92 ± 1.55  | 0.003                |
| PBG120′ (mmol/L)       | 4.58 ± 0.93   | 5.48 ± 1.26   | 0.208                |
| AUC glucose (h:mmol/L) | 13.24 ± 1.56  | 15.88 ± 1.40  | 0.016                |
| HOMA-IR                | 1.35 ± 0.32   | 1.77 ± 0.31   | 0.045                |

Figure 3. Ratios of 5αR1 to β-actin. *Compared with the Control group, P < 0.05.
levels of FBG, PBG30′ and PBG60′ in all of these groups (Table 2).

Table 2. Changes of metabolic and endocrine variables after exercise. Values are means ± SD. aCompared with CO group, p < 0.05; bcompared with PS group, p < 0.05; ccompared with PS + RI group, p < 0.05; dcompared with PS + R2I group, p < 0.05; ecompared with PE + RI group, p < 0.05.

|                     | No-exercise | PS         | PS + RI    | PS + R2I    | PE         | PE + RI    | PE + R2I   |
|---------------------|-------------|------------|------------|------------|------------|------------|------------|
| TINS (mU/L)         | 10.30 ± 0.69| 11.45 ± 0.71 | 11.01 ± 0.68 | 10.93 ± 0.71 | 10.13 ± 0.64 | 11.42 ± 0.87 | 9.67 ± 1.23 |
| FBG (mmol/L)        | 3.12 ± 0.34 | 3.06 ± 0.38 | 3.35 ± 0.38 | 3.60 ± 0.60 | 2.93 ± 0.41 | 3.33 ± 0.31 | 3.05 ± 0.27 |
| PBG30′ (mmol/L)     | 7.2 ± 1.04  | 9.22 ± 2.44 | 8.13 ± 0.61 | 8.02 ± 1.40 | 8.47 ± 0.92 | 8.06 ± 1.13 | 9.65 ± 2.96 |
| PBG60′ (mmol/L)     | 6.32 ± 0.34 | 7.57 ± 0.77 | 8.03 ± 1.44 | 7.70 ± 1.02 | 6.73 ± 0.88 | 7.92 ± 0.52 | 7.90 ± 2.13 |
| PBG120′ (mmol/L)    | 4.20 ± 0.42 | 5.12 ± 0.74 | 4.80 ± 0.43 | 4.37 ± 0.97 | 4.33 ± 0.44 | 4.46 ± 0.48 | 3.68 ± 0.72 |
| AUCglucose (h:mmol/L) | 13.35 ± 0.97 | 16.04 ± 1.37 | 15.73 ± 0.81 | 15.05 ± 1.30 | 14.35 ± 0.41 | 15.27 ± 0.81 | 15.14 ± 3.34 |
| HOMA-IR             | 1.43 ± 0.14 | 1.56 ± 0.10 | 1.64 ± 0.21 | 1.74 ± 0.22 | 1.32 ± 0.16 | 1.70 ± 0.25 | 1.31 ± 0.22 |

Figure 4. Ratios of 5αR1 to β-actin; response to exercise and 5αR1. aCompared with the CO group, p < 0.05; bcompared with the PS group, p < 0.05; ccompared with the PS + RI group, p < 0.05; dcompared with the PS + R2I group, p < 0.05; ecompared with the PE + RI group, p < 0.05.

groups. The levels of AUG glucose and HOMA-IR were significantly elevated in the PS, PS + RI, PS + R2I and PE + RI groups compared to the CO group, while the levels of HOMA-IR were significantly reduced in the PE + R2I compared to the PS, PS + RI, PS + R2I and PE + RI groups. No significant differences were found in the levels of FBG, PBG30′ and PBG60′ in all of these groups (Table 2).

5αR1expression in skeletal muscle in response to exercise and 5αR inhibition. After 14 days of intervention, the expression of 5αR1 in the PS, PS + RI, PS + R2I and the PE + RI groups were significantly increased compared with the CO group. Compared with the PS group, the expression of 5αR1 was significantly lower in the PS + RI, PE, PE + RI and PE + R2I groups (P < 0.05). No significant difference was observed between the CO group, the PE group and the PE + R2I group (Fig. 4).

PI3Kp85 and Akt phosphorylation expression in skeletal muscle in response to exercise and 5αR inhibition. Compared with the CO group and the PE + R2I group, the expression of PI3Kp85 was significantly reduced in the PS, PS + RI, PS + R2I, PE and PE + RI groups. Compared with the PE group, the expression of PI3Kp85 was significantly lower in the PS, PS + RI and PS + R2I groups (P < 0.05). There was no significant difference in the expression of PI3Kp85 between the CO group and the PE + R2I groups, as well as between the PS, PS + RI, PS + R2I and PE + R2I groups (Fig. 5).

Compared with the CO group and the PE + R2I group, the relative Akt(Ser 473) phosphorylation ratios were significantly lower in the PS, PS + RI, PS + R2I, PE and PE + RI groups (P < 0.05). Compared with the PE group, the relative Akt(Ser 473) phosphorylation ratios were significantly lower in the PS, PS + RI, PS + R2I and PE + RI groups (P < 0.05). There was no significant difference in the expression of PI3Kp85 between the CO, PE and PE + R2I groups, as well as between the PS, PS + RI, PS + R2I and PE + R2I groups (Fig. 5).

Compared with the CO and the PE + R2I group, the relative Akt(Thr308) phosphorylation ratios were significantly lower in the PS, PS + RI, PS + R2I and PE + RI groups (P < 0.05). Compared with the PE group, the relative Akt(Thr308) phosphorylation ratios were significantly reduced in the PS group (P < 0.05). No significant difference was found in the expression of PI3Kp85 between the CO and PE + R2I groups, as well as between the PS, PS + RI and PE + R2I groups (Fig. 5).
Effects of exercise and 5αR inhibition on GLUT4 expression. Compared with the CO, PE and PE + R2I groups, the expression of PM GLUT4 was significantly lower in the PS, PS + RI, PS + R2I and PE + RI groups (P < 0.05). Compared with the CO, the expression of total GLUT4 was significantly reduced in the PS, PS + RI and PS + R2I groups (P < 0.05). There was no significant difference in the expression of PM GLUT4 and total GLUT4 between the CO, PE and PE + R2I groups, as well as between the PS, PS + RI, PS + R2I and PE + R2I groups. The ratio of PM GLUT4 to total GLUT4 also tend to be significantly lower in the PS, PS + RI, PS + R2I and PE + RI groups (P < 0.05) (Fig. 6).

GLUT4 immunocytochemistry. Signals were visualized by DAB. Brown staining represents GLUT4 and blue staining represents nuclei. The value of the average optical density of GLUT4 in the CO, PS, PS + RI, PS + R2I, PE, PE + RI and PE + R2I groups were 0.20 ± 0.02, 0.10 ± 0.02, 0.11 ± 0.05, 0.09 ± 0.02, 0.19 ± 0.03, 0.19 ± 0.02 and 0.09 ± 0.02, respectively. The value of the average optical density of GLUT4 in the skeletal muscle of rats in the PS, PS + RI, PS + R2I and PE + RI groups were much less than that of the CO, PE and PE + R2I groups (P < 0.05) (Fig. 7).

Discussion
Several studies have suggested that insulin resistance plays a key role in the pathogenesis of PCOS22,23. By measuring the activity of 5αR in urine, an increase of the 5αR activity was observed with an increase of insulin resistance index in related studies24–26. A small sample population-based research study showed that the elevated activity of 5αR was associated with obesity27 and type2 diabetes28 and could be regulated by the composition of dietary nutrients. Compared with weight-matched normal women, PCOS showed more severe insulin resistance with increased 5αR activity4. In our experiment, the PCOS rat model also showed severe insulin resistance with raised activity of 5αR1 in skeletal muscle. Tomlinson et al. found that the level of 5αR increased in obese populations, with no gender differences. Therefore, it is suggested that the increase of reductase activity may be a compensatory mechanism after the decrease of insulin sensitivity29.

5αRI, which reduce circulating and prostatic 5α-dihydrotestosterone (DHT) levels, are currently widely used in androgen treatment for prostate hyperplasia and other diseases30–32. Finasteride(5αR1) inhibits 5αR selectively, whereas dutasteride(5αR1) inhibits both 5αR1 and 5αR2. Previous studies have suggested that there is almost no significant difference between the two inhibitors in terms of clinical efficacy and related parameters33,34. However, increasing studies show that the two inhibitors are very different in their biochemical effects15,35. Related studies have shown that people who received dutasteride displayed increased body fat and decreased insulin sensitivity. These features were not found in people who received finasteride35. In fact, finasteride, in contrast, can even improve insulin function. Therefore, 5αR1 may play an important role in regulating metabolism. In 2014, the study of skeletal muscle cultures of PCOS patients showed no interaction between insulin and androgen13. In our present study, PCOS rats showed no improvement in glucose and lipid metabolism after 14 days of intragastroic administration of finasteride and dutasteride. Typical characteristics of insulin resistance were observed in these PCOS rats. It was further confirmed that PCOS rats may have abnormalities in some genes or proteins, thus blocking the interaction between androgen and insulin36.
Exercise itself reduced 5α-reductase in PCOS rats, and the expression of 5αR1 in skeletal muscle was significantly lower in the PCOS exercise group than in the PCOS stationary group. Exercise can reduce body weight and improve insulin sensitivity. Studies have shown that weight loss may be accompanied by a decrease in reductase activity, and the mechanism may be related to the decrease of the glucocorticoid production rate and the role of the hypothalamus pituitary adrenal axis. Similarly, PCOS patients who received an oral insulin sensitizer (thiazolidinediones) showed a decrease in reductase activity. It is considered that the reduction of the 5αR activity was associated with the level of insulin-like growth factor 1 (IGF-1), adiponectin, and growth hormone in the circulation. Animal experiments in 2011 suggest that aerobic exercise training induced to increase circulating and muscle DHT level, and the increased muscle DHT level leads to an increase in glucose metabolism in skeletal muscle of obese and diabetic rats. The results of these studies seem to be in contradiction with the results of reducing the activity of reductase in our experiment. But different from obese and diabetic patients, PCOS patients have the endocrine environment of hyperandrogenism and this study does not pay much attention to androgen pathway. So it remains to be further studied.

It has been recognized and accepted that exercise can improve insulin resistance in diabetes. The main mechanism is to increase the utilization efficiency of insulin by activating the non-insulin dependent AMPK signaling pathway and the insulin-dependent PI3K/AKT signaling pathway. Previous studies have shown that PCOS patients have abnormal glucose metabolism as represented by their levels of AKT. Additionally, animal experiments demonstrated that the changes in glucose metabolism could be induced by exogenous androgen in normal SD rats. Therefore, we chose the PI3K/AKT signaling pathway to investigate. Exercise can improve insulin resistance of PCOS with the main target organ of exercise being skeletal muscle. Therefore, it may be considered that the executive organ of exercise for glucose metabolism regulated by insulin and androgen is skeletal muscle. Insulin is now thought to activate PKC and Akt, as well as AS160, which is a downstream signaling molecule of Akt. By activating PI3K, the sugar transporter GLUT-4 is translocated to the cell membrane and the sugar transport capacity of the muscle cells are all increased, thereby allowing this pathway to be utilized by contracting muscles. Our study has proved that there is an abnormal activation of the PI3K/AKT signaling pathway in the skeletal muscle of the PCOS rat model. After exercise intervention, the PCOS exercise group showed a significantly higher expression of PI3Kp85 protein, as well as the phosphorylation levels of Akt(Ser473) and Akt(Thr308) in comparison to the PCOS stationary group. This suggests that exercise activates the PI3K/AKT signaling pathway in skeletal muscles. In the PCOS non-exercise group, only the reductase inhibitor was given with no increase in the phosphorylation levels of PI3K and AKT being achieved regardless of whether 5αRI or 5αR2I was used. This suggests that the reductase inhibitor itself, although similar to exercise, may decrease reductase activity; however, it cannot activate the PI3K/AKT signaling pathway. In the PCOS exercise group, when type 1

**Figure 6.** Effect of exercise and 5αRI on GLUT4 expression in skeletal muscle. (A) Western blots of PM GLUT4 and total GLUT4. (B) Ratios of PM GLUT4 to β-actin. (C) Ratios of total GLUT4 to β-actin. (D) Ratios of PM GLUT4 to total GLUT4. *a* compared with the CO group, p < 0.05; *b* compared with the PS group, p < 0.05; *c* compared with the PS + RI group, p < 0.05; *d* compared with the PS + R2I group, p < 0.05; *f* compared with the PE + RI group, p < 0.05.
reductase is retained, exercise can activate the signaling pathway by repressing 5αR1 action. These results suggest that 5αR1 may play a key role in the activation of the PI3K/AKT signaling pathway in PCOS rats. Of course, exercises play a comprehensive role and do not exclude the activation of other substances, such as IGF and insulin receptor. Our research suggests that reductase may be another important mediator in addition to known mediators that can regulate PI3K/AKT pathway. And the relationship between these mediators needs further research.

This study confirmed that the expression of 5αR1 protein was increased in skeletal muscle of PCOS rats, and exercise can decrease 5αR1 to activate the PI3K/Akt pathway in skeletal muscle of PCOS rats. Because 5αR1 acts as a key enzyme in the androgen metabolism pathway, this study further confirms that 5αR1 may be at the intersection of two pathways for exercise, androgen therapy and insulin. Thus, the treatment of 5αR1 may be the key to activate the two pathways of androgen and insulin. And, 5αR1, as the target of exercise therapy, maybe can provide a new idea for the formulation of exercise prescription and the evaluation of exercise efficacy in the future.

There were still some deficiencies in this experiment: (1) In this experiment, we only studied the glucose metabolism pathway and did not assess androgen metabolism in skeletal muscle; (2) The sample size of this study is too small and the study is limited to animal research; (3) In this study, we mainly observed the changes of 5αR1. The relationship between 5αR2 and 5αR1 should be further studied.

Conclusion
In this study, the PCOS model rats were treated with the combination of 5αR inhibitors and exercise. The results demonstrate that exercise activates glucose metabolism signaling pathways in skeletal muscle by reducing the expression of 5αR1 protein in PCOS rats.
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Author Contributions
Chuyan Wu designed the study, performed experiments, analyzed the data and wrote the manuscript, Feng Jiang performed experiments, analyzed the data and critically reviewed the manuscript, Ke Wei performed experiments and critically reviewed the manuscript, Zhongli Jiang interpreted results of experiments and critically reviewed and edited the manuscript.

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