Tumors have evolved mechanisms to escape anti-tumor immnosurveillance. They limit humoral and cellular immune activities in the stroma and render tumors resistant to immunotherapy. Sensitizing tumor cells to immune attack is an important strategy to revert immunosuppression. However, the underlying mechanisms of immune escape are still poorly understood. Here we discover Indoleamine-2,3-dioxygenase-1 (IDO1) Paneth cells in the stem cell niche of intestinal crypts and tumors, which promoted immune escape of colorectal cancer (CRC). Ido1 expression in Paneth cells was strictly Stat1 dependent. Loss of IDO1 Paneth cells in murine intestinal adenomas with tumor cell-specific Stat1 deletion had profound effects on the intratumoral immune cell composition. Patient samples and TCGA expression data suggested corresponding cells in human colorectal tumors. Thus, our data uncovered an immune escape mechanism of CRC and identify IDO1+ Paneth cells as a target for immunotherapy.
Colon cancer (CRC) is the third most common cancer worldwide and patients with metastases in distant organs have a 5-year survival rate below 13%. Metastatic CRC is currently treated with several combinations of cytotoxic agents. They improved overall survival of patients, treated initially with fluoropyrimidine monotherapy, from 12 to 30 months. However, chemotherapy reached its limits, which fostered clinical trials for immunotherapies. The importance of immunosurveillance in CRC is emphasized by the good prognostic value of CD3+, CD8+, and CD45RO+ T-cell infiltration (immunoscore)\(^{15-17}\). Unfortunately, immunotherapy with checkpoint inhibitors showed clinical benefits only in mismatch-repair-deficient CRC with high neo-antigen load. Durable responses in CRC with different etiologies remained scarce, which is due to immune escape mechanisms\(^8\).

The transcription factor signal transducer and activator of transcription 1 (Stat1) is a key effector in tumor immunosurveillance mediated by natural killer (NK)- and T cells\(^9,10\). Consistently, Stat1 is part of the immunologic constant of rejection gene expression signature, which correlates with good prognosis of CRC\(^7\). In cancer cells, Stat1 inhibits proliferation and promotes apoptosis via induction of cyclin-dependent kinase inhibitors and pro-apoptotic proteins\(^9\). Stat1 also regulates the expression of tissue antigens and proteins of the antigen presentation machinery, which enhance the immunogenicity of tumors\(^31\). Therefore, it is generally considered that Stat1 expression and activation in immune cells and in cancer cells suppresses tumors. Type I and II interferon (IFN) are the major activators of canonical Stat1 signaling, which relies on Tyr701 phosphorylation (p-IFN-Stat1) and mediates tumor suppressive effects of IFN\(^12\). However, the tumor cell-intrinsic role of Stat1 in CRC is not well defined.

Lgr5\(^+\) stem cells at the bottom of intestinal crypts have been identified as possible precursor cells for CRC\(^13\). However, non-stem cells can also acquire tumor-initiating capacity\(^14\) and Lgr5\(^+\) cancer stem cells are not essential for growth of primary tumors\(^15\). These cells are separated by Lysozyme\(^+\) Paneth cells in the small intestine, which provide essential niche factors for stem cell proliferation and self-renewal\(^16\). In the colon, Lysozyme\(^-\) deep crypt base secretory cells support Lgr5\(^+\) stem cells\(^16\) but colonic Paneth cells can appear through epithelial metaplasia\(^17\).

Paneth cells have been identified in intestinal tumors of Apc\(^{Min}\) mice\(^18\) and in sporadic CRC of humans, albeit at varying frequencies ranging from 0.2% to 39%\(^19\). However, intestinal tumors from familial adenomatous polyposis (FAP) patients with inherited Apc mutations harbored more than 90% of Paneth cells\(^20\). The role of Paneth cells is unclear but CRC developed predominantly in colonic mucosal tissue with Paneth cell metaplasia\(^21\) and the presence of Paneth cell-containing adenomas increased the risk for synchronous CRC\(^19\). Therefore, Paneth cells might promote CRC formation.

Here we identified a subset of Paneth cells that displayed Stat1-dependent expression of the immune checkpoint molecule PD1. Loss of these cells in Stat1-deficient intestinal tumors of Apc\(^{Min}\) mice resulted in reduced tumor load and increased infiltration of anti-tumor immune cells.

**Results**

**Epithelial Stat1 promotes formation of intestinal tumors.** We used Apc\(^{Min}\) mice\(^25\) with conditional deletion of Stat1 in intestinal epithelial cells (Stat1\(^{ΔIEC}\) Apc\(^{Min}\))\(^23,24\) to identify Stat1 functions in intestinal tumorigenesis. Deletion of Stat1 was confirmed by PCR (Supplementary Fig. 1a), quantitative PCR (qPCR) of purified intestinal epithelial cells (Supplementary Fig. 1b) and immunohistochemistry (IHC, Supplementary Fig. 1c). Lamina propria cells of Stat1\(^{ΔIEC}\) Apc\(^{Min}\) mice displayed STAT1 expression, which demonstrated specific ablation in intestinal epithelial cells (Supplementary Fig. 1c). Goblet, enterocinocrine, Paneth, and proliferating cells in the intestinal crypts were present at normal numbers in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) mice (Supplementary Fig. 1d-h). These data show that epithelial cell-specific deletion of Stat1 does not affect intestinal cell differentiation and crypt proliferation of Apc\(^{Min}\) mice.

Four-month-old Stat1\(^{ΔIEC}\) Apc\(^{Min}\) and Stat1\(^{ΔIEC}\) Apc\(^{Min}\) mice were used to investigate epithelial cell-intrinsic functions of Stat1 in intestinal tumorigenesis. Tumor formation was reduced in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) male and female mice (Fig. 1a–c). Angiogenesis (Supplementary Fig. 2a), tumor cell proliferation, and apoptosis (Supplementary Fig. 2b) were not affected but numbers of low-grade adenomas were increased (Fig. 1d). These data show that epithelial cell-intrinsic Stat1 promotes the formation and progression of intestinal tumors in Apc\(^{Min}\) mice.

Apc\(^{Min}\) mice develop tumors mainly in the small intestine but also in the colon\(^25\). Similarly, we found tumors in the small intestine and the colon and stained them for STAT1 by IHC. STAT1 was detected in tumor and stroma cells of Stat1\(^{ΔIEC}\) Apc\(^{Min}\) mice. In contrast, STAT1 was not detectable in tumor cells of Stat1\(^{ΔIEC}\) Apc\(^{Min}\) tumors demonstrating efficient conditional deletion (Fig. 1e–g). However, Stat1\(^{ΔIEC}\) Apc\(^{Min}\) tumors displayed a significant upregulation of STAT1 in the tumor stroma (Fig. 1e, g). Numbers of STAT3- and activated pY-STAT3-positive cells were not changed in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) tumors (Supplementary Fig. 2c–e). These data show that tumor cell-intrinsic Stat1 suppresses upregulation of Stat1 in the stroma of Apc\(^{Min}\) tumors.

**Tumor cell-intrinsic Stat1 suppresses immune cell activation.** We have recently shown that enhanced anti-tumor immune cell activity is reflected by increased stromal Stat1 expression in azoxymethane-dextran sodium sulfate (AOM-DSS)-induced colorectal tumors\(^26\). Therefore, we investigated immune cell activation in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) tumors. As human intestinal tumors develop mainly in the colon, we performed RNA sequencing (RNA-seq) experiments with murine colon tumors (Supplementary Data 1 and 2). The analyses showed reduced expression of several IFN-stimulated genes (ISGs) in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) tumors (Supplementary Data 1). In particular, 24 out of 49 orthologs of human genes of the IFN-related gene signature for DNA damage (IRDS)\(^27\) were downregulated in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) tumors (Fig. 1i–j and Supplementary Data 1). We analyzed The Cancer Genome Atlas (TCGA) data to evaluate whether these genes are also regulated by Stat1 in human CRC. A correlation analysis revealed 529 genes that are co-expressed with Stat1 in human CRC (Supplementary Table 1). Among them were 16 IRDS genes. A Venn diagram using (i) human IRDS genes\(^27\), (ii) orthologs of human IRDS genes, significantly downregulated in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) mouse tumors, and (iii) the 529 genes, identified to be co-expressed with Stat1 in the CRC TCGA dataset, revealed a substantial overlap. A signature of 13 genes was present in all three gene sets (Fig. 1h).

Gene Ontology (GO) term-enrichment analyses of deregulated genes in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) tumors and genes that are co-expressed with Stat1 in human CRC showed a substantial overlap of GO terms and revealed mainly pathways implicated in immunological processes (Supplementary Data 3 and 4). These data suggest that Stat1 target genes and Stat1-dependent regulation of immunological processes are similar in human and murine CRC.

Among the most significantly downregulated modulators of immune responses in Stat1\(^{ΔIEC}\) Apc\(^{Min}\) tumors was the enzyme Indoleamine-2,3-dioxygenase-1 (Ido1, Fig. 1f). Supplementary
checkpoint28. Other immune checkpoints were not deregulated in Stat1ΔIEC ApcMin mice and the supernatants of Stat1ΔIEC ApcMin (si: 12 tumors, 4 mice; colon: 12 tumors, 4 mice; Stat1ΔIEC ApcMin si: 12 tumors, 4 mice; colon 12 tumors, 4 mice; Stat1ΔIEC ApcMin si: 24 tumors of 4 mice each). A Venn diagram showing an overlap between human IRDS genes, IDO1 genes downregulated in Stat1ΔIEC ApcMin colon tumors, and genes that positively correlated with Stat1 expression in human CRC TCGA data. Heat map of downregulated IRDS genes in Stat1ΔIEC ApcMin colon tumors. qPCRs for Ifi44, Isg15, and Ido1 mRNA expression in colon tumors of Stat1ΔIEC ApcMin (5 mice) and Stat1ΔIEC ApcMin (6 mice) mice (tumors from each mouse were pooled). ELISA for kynurenine in supernatants of Stat1ΔIEC ApcMin (si: 7 tumors, 5 mice; colon: 7 tumors, 6 mice) and Stat1ΔIEC ApcMin (si: 7 tumors, 7 mice; colon: 7 tumors, 5 mice). Quantification of iNOS+ and Granzyme B+ stroma cells (Stat1ΔIEC ApcMin: 12 tumors, 4 mice; GZMB < 0.5 mm²: 9 tumors, 2 mice; >0.5 mm²: 21 tumors, 4 mice; Stat1ΔIEC ApcMin: iNOS: 14 tumors, 4 mice; GZMB < 0.5 mm²: 23 tumors, 5 mice; >0.5 mm²: 13 tumors, 3 mice). FACS analysis of immune cells of Stat1ΔIEC ApcMin (pooled tumors of 5 mice in 4 experiments) and Stat1ΔIEC ApcMin (pooled tumors of 4 mice in 4 experiments) colon tumors. si: small intestine. Bars represent mean ± SEM.

Fig. 3a, and Supplementary Data 1), which acts as an immune checkpoint28. Other immune checkpoints were not deregulated in Stat1ΔIEC ApcMin tumors but expression of the T-cell activation marker CD28 was increased (Supplementary Fig. 3a). IDO1 converts tryptophan into the immune-suppressive metabolite kynurenine. Consistently, levels of kynurenine were reduced in supernatants of Stat1ΔIEC ApcMin tumors (Fig. 1k). We analyzed expression of inducible nitric oxide synthase (iNOS) and the serine protease Granzyme B, because they are markers for activation of several immune cells such as macrophages, mature
dendritic cells, cytotoxic T cells, or NK cells. IHC characterization of the stroma showed increased numbers of activated INOS⁺ immune cells (Fig. 1). Numbers of Granzyme B⁺ cells were not significantly changed in large tumors but accumulated in small adenomas of Stat1ΔIEC Apc⁰⁻ Min mice (Fig. 1) and Supplementary Fig. 3b).

To address this issue for IHC staining, we performed in-situ hybridization (ISH) experiments and verified expression of Ido1 mRNA in adenoma sheets of Stat1ΔIEC Apc⁰⁻ Min tumor cells (Supplementary Fig. 3d). These data suggest that tumor cell-intrinsic Stat1 suppression stroma immune cell activation in Apc⁰⁻ Min tumors through Ido1.

**Stat1 promotes Ido1 expression in neoplastic Paneth cells.** We performed IHC and IF staining to assess downregulation of Ido1 in Stat1ΔIEC Apc⁰⁻ Min tumors at the cellular level. This analysis revealed specific Ido1⁺ tumor cells in Stat1ΔIEC Apc⁰⁻ Min tumors that were absent in Stat1ΔIEC Apc⁰⁻ Min tumors (Fig. 2a, b, i). It has been shown that commercially available Ido1 antibodies are unspecific and unable to detect Ido1 in western blotting.

To address this issue for IHC staining, we performed in-situ hybridization (ISH) experiments and verified expression of Ido1⁺ tumor cells at the RNA level (Fig. 2a). The Ido1⁺ tumor cells were arranged in an alternating pattern with Ido1⁻ tumor cells in neoplastic adenoma sheets (Fig. 2a). A similar arrangement was described for transformed Lgr5⁺ stem cells and Lysozyme⁺ Paneth cells in adenoma sheets of Apc⁰⁻ Min tumors, which resembles crypt organization of the small intestine. This suggests that Ido1⁺ tumor cells are either related to stem cells or to Paneth cells. Co-expression of Lgr5 and Ido1 mRNA was barely detectable by ISH in Stat1ΔIEC Apc⁰⁻ Min tumor cells (Fig. 2a), indicating that Ido1⁺ cells are Paneth cells. However, the ISH signals for Lgr5 were weak and not clearly attributable to individual cells. Therefore, double immunofluorescence (IF) with Paneth cell markers was performed. These analyses revealed protein expression of Paneth cell markers Lysozyme and MMP7 in Ido1⁺ tumor cells (Fig. 2c, g, h). More than 80% of Ido1⁺ tumor cells expressed Paneth markers (Fig. 2e, k), indicating that Paneth cells are the major source for Ido1 expression in the neoplastic epithelium. Double-positive cells were absent in Stat1ΔIEC Apc⁰⁻ Min tumors (Fig. 2c, g) but the overall numbers of Lysozyme⁺ and MMP7⁺ Paneth cells were not reduced (Fig. 2d, j).

Moreover, about 50% of Paneth cells expressed Ido1 in Stat1ΔIEC Apc⁰⁻ Min tumors (Fig. 2f, l). These data demonstrate that Stat1 is required for the formation of Ido1⁺ Paneth cells in Apc⁰⁻ Min tumors. Ido1⁺ Paneth cells are potential immunosuppressors and their presence might account for immunological changes in Stat1ΔIEC Apc⁰⁻ Min tumors. This assumption is challenged by the low number of Ido1⁺ Paneth cells in Stat1ΔIEC Apc⁰⁻ Min tumors (Fig. 2b, i). However, we identified a much higher percentage of Ido1⁺ Paneth cells in small and early Apc⁰⁻ Min adenomas with almost 30% in the colon (Fig. 2m, n). These data suggest that Ido1⁺ Paneth cells support immune escape during early stages of Apc⁰⁻ Min-induced tumorigenesis.

Ido1 is an ISG in human and murine tumor cells, indicating a role of canonical Stat1 signaling in the formation of Ido1⁺ Paneth cells. IHC staining for canonical Stat1 activation detected <3% pY-STAT1⁺ tumor cells in Stat1⁰⁻/⁰⁻ Apc⁰⁻ Min tumors (Supplementary Fig. 4a, b). The pY-STAT1⁺ tumor cells appeared as cell clusters (Supplementary Fig. 4a), which differed from the alternating arrangement of Ido1⁺ Paneth cells. pY-STAT1 was undetectable in tumor cells of Stat1ΔIEC Apc⁰⁻ Min tumors (Supplementary Fig. 4a, b) but appeared upregulated in the stroma (Supplementary Fig. 4a, c) similar to upregulation of total STAT1 (Fig. 1e, g). We blunted type I IFN signaling by conditional deletion of Ifnar1 in Apc⁰⁻ Min tumor cells. Tumor formation was not affected in the intestine of Ifnar1⁻⁻ Apc⁰⁻ Min mice (Supplementary Fig. 4d-f). Moreover, the number of Ido1⁺ Paneth cells was comparable in Ifnar1⁻⁻ Apc⁰⁻ Min and Ifnar1ΔIEC Apc⁰⁻ Min intestinal tumors (Supplementary Fig. 4g). These data demonstrate that the formation of Ido1⁺ Paneth cells in tumors of Apc⁰⁻ Min mice is independent of type I IFN signaling.

We next investigated whether human CRC contain Ido1⁺ Paneth cells. Biopsies of 149 human T3 and T4 CRC that had not yet metastasized (Supplementary Table 2) were IHC-stained for Stat1 and Ido1 to compare staining patterns with Stat1ΔIEC Apc⁰⁻ Min tumor cells. STAT1⁻⁻ and Ido1⁺ cancer cells were readily detectable in human CRC but unlike mouse tumors, Ido1⁺ cells did not show an alternating pattern with Ido1⁻ cells (Fig. 3a). However, a TCGA-based correlation matrix of ISGs (IRDS genes) and marker genes for cell identities revealed a correlation between the expression of Stat1, Ido1, and Lysozyme. Lysozyme clustered with Ido1- and Stat1-regulated IRDS genes (Fig. 3b). Moreover, we stained 14 early adenomas (5 from FAP patients) for neoplastic Lysozyme⁺ Ido1⁺ Paneth cells. Lysozyme⁺ and Lysozyme⁻ Ido1⁺ Paneth cells were found in ten adenomas (four from FAP patients) and six adenomas (three from FAP patients), respectively. In particular, FAP adenomas showed a perinuclear signal for Ido1 in Paneth cells (Fig. 3c). The relative contribution of Paneth cells to Ido1 expression was assessed by IF staining of the six adenomas harboring Ido1⁺ Paneth cells. About 50% of Ido1⁺ tumor cells were Lysozyme⁺, demonstrating a significant contribution of Paneth cells to Ido1 expression in the neoplastic epithelium (Fig. 3d).

In summary, these data suggest that Ido1⁺ Paneth cells are present in human CRC.

**IDO1⁺ CRC cells promote immune escape.** Subcutaneous implantation of C57BL/6-derived MC38 cells into immunocompetent host mice is an established method for evaluation of pre-clinical immunotherapy approaches. We transplanted green fluorescence protein (GFP)-labeled MC38 CRC cells to test whether deletion of Ido1 in neoplastic cells mimics immunologic consequences of Ido1⁺ Paneth cell ablation in Stat1ΔIEC Apc⁰⁻ Min tumors. Two independent MC38Δid01-GFP subclones (MC38Δid01-GFP-2 and MC38Δid01-GFP-6) with CRISPR/Cas9-mediated deletion of the Ido1 locus were generated. The presence of INDELS in MC38Δid01-GFP cells was verified by sequence analysis. Both clones contained an additional G in exon 6 of Ido1, which is a common CRISPR/Cas9-mediated insertion, and resulted in a truncated Ido1 protein (MC38Δid01-GFP-2) with CRISPR/Cas9-mediated deletion of the Ido1 locus. Sequencing revealed a bigger deletion of the genomic locus upstream of the sgRNA targeting site in MC38Δid01-GFP-2 cells, which might destabilize the Ido1 mRNA. Commercial antibodies are not suitable to detect a specific Ido1 protein by western blotting. However, IFNγ stimulation induced Ido1 mRNA expression in MC38wt-GFP cells but not in MC38Δid01-GFP-2 or MC38Δid01-GFP-6 cells (Fig. 4a and Supplementary Fig. 5c). MC38Δid01-GFP cells displayed a reduced cumulative cell number (Supplementary Fig. 5d), indicating an in-vitro proliferation defect similar to human cells. In vivo, MC38Δid01-GFP cells formed smaller tumors than MC38wt-GFP.
cells in immunocompetent C57BL/6 hosts (Fig. 4b, c), which were strongly infiltrated with CD3+ T cells (MC38ΔIdo1-GFP-2 cells are shown in Fig. 3d, e). In contrast, growth of MC38ΔIdo1-GFP-2 cells was not affected in immunocompromised NOD scid gamma (NSG) hosts, which lack mature T cells, B cells, and NK cells (Fig. 4f, g). We performed transplantation experiments with mixtures of cells to evaluate protective effects acting in trans. MC38ΔIdo1-GFP-6 cells were additionally labeled with dsRed to discriminate them from Ido1-proficient MC38wt-GFP cells. A 1:1 mixture of MC38wt-GFP/MC38ΔIdo1-GFP-6 cells showed comparable growth to MC38wt-GFP cells in immunocompetent C57BL/6 hosts, indicating that MC38wt-GFP cells restored growth of MC38ΔIdo1-G/RFP-6 cells in trans (Fig. 4h). IHC staining detected dsRed-positive cells close to the expected percentage in mixed tumors (Fig. 4i, j). Moreover, the prominent CD3+ T-cell infiltration in MC38ΔIdo1-GFP tumors (Fig. 4d, e) was abolished in mixed tumors (Fig. 4k). IHC staining revealed strong infiltration of Granzyme B+ immune cells in MC38ΔIdo1-G/RFP-6 tumors, which was also abolished in mixed tumors (Fig. 4l). These data demonstrate that Ido1+ MC38 CRC cells are able to promote immune escape of transplanted tumors.

Tumor cell-intrinsic Stat1-Ido1 favors progression of CRC. We investigated correlations of STAT1 and IDO1 protein expression...
in IHC-stained biopsies of 149 human T3 and T4 CRC (Supplementary Table 2). A score of 0–4 was attributed to STAT1 and IDO1 levels in tumor and stroma compartments (Supplementary Fig. 6). STAT1 or IDO1 protein in tumor or stroma cells did not correlate with overall survival and metastasis-free survival of patients (Supplementary Table 2). However, a strong correlation was observed between protein expression of STAT1 and IDO1 in tumor cells (Fig. 5a) and stroma cells (Fig. 5b). Additional analysis of TCGA data was used to increase the sample size. A correlation plot, derived from TCGA data, confirmed strong co-expression (Spearman score of 0.797) at the RNA level (Fig. 5c). IF staining was used for quantification of IDO1+ non-Paneth (LYZ−) and Paneth (LYZ+) cells in the neoplastic epithelium of human adenomas.

in the neoplastic epithelium of CRC. IHC staining of human biopsies demonstrated IFIT1 expression in CRC cancer cells but not in the tumor stroma (Supplementary Fig. 7c). Importantly, IFIT1 expression correlated strongly with STAT1 and IDO1 expression in CRC cancer cells (Supplementary Fig. 7c). Scoring of IDO1 and IFIT1 staining intensities (Supplementary Fig. 7d) confirmed a strong correlation of protein expression (Supplementary Fig. 7e, Pearson coefficient = 0.541). We first evaluated the prognostic value of stromal STAT1 expression in Ifit1low tumors. Ifit1low Stat1low and Ifit1low Stat1high tumors should display low Stat1-Ido1 expression in the neoplastic epithelium and low or high Stat1 expression in stroma cells, respectively. The Ifit1-based stratification significantly improved the prognostic value of STAT1 expression in CRC (p = 0.03, Fig. 5f). Patients with Ifit1low CRC benefited from high stromal STAT1 expression immediately after diagnosis, indicating that low expression of tumor cell-intrinsic Stat1-Ido1 sensitizes tumors to immune attack. We next mimicked conditions of our mouse models and stratified TCGA data into Ifit1high Stat1low and Ifit1high Stat1high CRC. Ifit1high Stat1low CRC (tumor cell-intrinsic Stat1-Ido1↑, stromal Stat1↓, similar to Stat1DEΔIEC ApMin tumors) displayed a higher percentage of late-stage IV tumors than Ifit1low Stat1high CRC (tumor cell-intrinsic Stat1-Ido1↓, stromal Stat1↑, similar to Stat1ALEC ApMin tumors) (Fig. 5g, h), suggesting that tumor cell-intrinsic Stat1-Ido1 promotes CRC progression. Moreover, we employed the DeMiXT algorithm38 to deconvolute tumor cell-intrinsic and stromal expression of Ido1 in TCGA data. We further stratified tumors into CMS1-4 consensus molecular...
subtypes and re-investigated immune cell marker expression. Similarly, immune cell markers were predominantly expressed in the stroma. Moreover, the strongest expression was detected in the stroma of CMS1, a subtype characterized by immune cell infiltration, as exemplified for T-cell markers (Supplementary Fig. 8a, b). Without tumor stroma deconvolution, patients with Idol1 high tumors showed a slight trend towards better prognosis (Fig. 5d, p = 0.55). After deconvolution, survival curves for stroma and tumor compartments were laterally reversed (Supplementary Fig. 8c). Patients with strong Idol1 expression in the neoplastic epithelial cells showed a strong trend (p = 0.074) towards bad prognosis. Taken together, these data suggest that tumor cell-intrinsic Stat1-Idol1 expression favors immune escape and progression of human CRC.

**IDO1** + Paneth cells are present in normal crypts. Similar to tumors, Idol1 mRNA expression was downregulated in small intestinal and colonic IEC preparations of Stat1ΔIEC ApcMin mice (Fig. 6a). Therefore, we wondered whether IDO1 + Paneth cells are present in normal crypts. IHC and ISH analyses revealed IDO1 + vesicle-bearing Paneth cells in a subset of crypts of Stat1lox/lox ApcMin mice (Fig. 6b), which were abolished in Stat1ΔIEC ApcMin crypts (Fig. 6b, c). We detected up to three IDO1 + cells in crypts of the small intestine and up to seven cells in colonic crypts (Fig. 6c). IDO1 + Paneth cells were also identified in Stat1lox/lox mice (Fig. 6c), demonstrating that their formation does not depend on the ApcMin allele. They were more abundant in the distal small intestine, which has a higher bacterial load, than in the proximal small intestine (Fig. 6c). Moreover, their abundance was decreased in mice that were housed in an extra clean special pathogen-free (SPF) facility and treated with antibiotics (Fig. 6c). Treated mice also displayed reduced numbers of Lysozymeox Paneth cells in the proximal intestine (Fig. 6c).
The formation of IDO1+ Paneth cells is induced by the bacterial microbiome. It was previously shown that the TLR9 agonist ISS DNA can induce Ido1 in intestinal epithelial cells, which protects from colitis. Therefore, we isolated intestinal organoids and stimulated them with immunostimulatory (ISS) DNA (ODN 1668). Lysozyme staining showed that Paneth cells were present in organoids but ISS DNA failed to induce Ido1 (Supplementary Fig. 8d, e). However, bacteria also promote IFNγ production by CD4+ T cells in the intestinal lamina propria. IFNγ readily induced Ido1 in all epithelial cells of organoids at the RNA and protein level (Fig. 6f, g and Supplementary Fig. 8e). To further investigate the role IFNγ in vivo, we performed IHC for IDO1 in intestines of Ifngr1−/− mice, which lack functional IFNγ signaling. The number of IDO1+ crypt cells was reduced in different intestinal parts of Ifngr1−/− mice (Fig. 6h, j).

Interestingly, Ifngr1−/− mice displayed reduced numbers of Lysozyme+ Paneth cells in the proximal small intestine similar to mice with antibiotic treatment (Fig. 6d). These data demonstrate a contribution of IFNγ in induction of IDO1+ Paneth cells.

Our data suggest that a bacteria/IFNγ axis is responsible for Ido1 induction in Paneth cells. Therefore, we analyzed single-cell RNA-seq (scRNA-seq) data of Haber et al. to investigate the impact of bacterial infection on Ido1 induction in Paneth cells. Paneth cell clusters were identified in t-distributed stochastic neighbor embedding (t-SNE) maps using marker genes. Only two Ido1+ cells were found in t-SNE maps of healthy mice. Interestingly, Ifngr1−/− mice were both allocated to the Paneth-1 cell cluster, which is located in the distal small intestine and expresses the marker GM2100243. Haber et al. derived also scRNA-seq data from bacteria- and helminth-
Fig. 6 IDO1+ Paneth cells are present in intestinal crypts and reduced in Ifngr1−/− mice. a qPCR for Ido1 mRNA expression in isolated intestinal epithelial cells of Stat1flox/flox ApcMin (si: n = 4; colon: n = 5) and Stat1IEC ApcMin (si: n = 6; colon: n = 5) mice. b IHC staining for IDO1 (upper images) and double ISH for Ido1 and Lgr5 (lower images) in the small intestine of Stat1flox/flox ApcMin and Stat1IEC ApcMin mice. IDO1+ Paneth cells are indicated by arrowheads. Scale bars indicate 20 µm. c-e Quantification of IDO1+ crypts (c), LYZ1+ Paneth cells/crypt (d), and IDO1+ cells/crypt (e) in different intestinal compartments of co-housed Stat1flox/flox ApcMin (n = 4), Stat1IEC ApcMin (n = 5), Stat1IEC ApcMin (n = 4), Stat1IEC ApcMin (n = 4), Stat1IEC ApcMin (n = 4), as well as C57BL/6 mice kept at an extra clean SPF facility with and without ABx treatment (n = 3 each). f-qPCR for Ido1 mRNA expression after IFNγ stimulation in intestinal organoids of Stat1flox/flox (three technical replicates, two mice) and Stat1IEC (three technical replicates, three mice) mice. g IF showing the induction of IDO1 upon IFNγ stimulation in tumor organoids of Stat1IEC ApcMin mice. Scale bars indicate 50 µm. h-j Quantification of IDO1+ crypts (h), LYZ1+ Paneth cells/crypt (i), and IDO1+ cells/crypt (j) in different intestinal compartments of co-housed C57BL/6 (n = 3) and Ifngr1−/− mice (n = 3). n.d.: not detectable; si: small intestine; SPF: special pathogen free. ABx: antibiotics. Bars represent mean ± SEM.

Discussion
We identified an immune escape mechanism of CRC that is based on Stat1-dependent expression of Ido1 in Paneth cells. Paneth cell markers have previously been linked with intestinal tumorigenesis but the significance of the observations remained unclear. The markers Pla2g2a and Mmp7 were identified as modifiers of Min mice and loss of Mmp7, which is essential for Paneth cell

infected mice. Analysis of these data revealed a prominent induction of Ido1 in Paneth cells of bacteria- but not helminth-infected mice (Fig. 7a, b, d, f). Stat1 was induced in both infection models but more prominently by bacteria (Fig. 7a-c, e). These data demonstrate that the formation of Ido1+ Paneth cells is induced by the bacterial microbiome.
Treg
Salmonella enterica
infection. Single-cell RNA-seq data were generated by Haber et al.43 (GEO database: GSE92332).

Moreover, expression of Paneth markers correlated with increased risk for dietary-induced sporadic intestinal cancer in mice47 and a Paneth cell-associated gene expression pattern was increased in human intestinal tumors48. The presence of IDO1+ Paneth cells in intestinal cancers might provide an explanation for these observations. IDO1 increases local kynurenine levels and depletes tryptophan. Effector T cells respond to tryptophan depletion with cell cycle arrest49 and kynurenine promotes Treg cell escape of CRC (for details, see discussion). CTL: cytotoxic T lymphocyte; Treg: regulatory T cell.

Ido1 is expressed by Paneth cells in intestinal cancers might provide an explanation for these observations. IDO1 increases local kynurenine levels and depletes tryptophan. Effector T cells respond to tryptophan depletion with cell cycle arrest49 and kynurenine promotes Treg cell escape of CRC (for details, see discussion). CTL: cytotoxic T lymphocyte; Treg: regulatory T cell.

Ido1 overexpression is commonly observed in human CRC and associated with reduced serum tryptophan levels, whereas kynurenine metabolites are increased53–55. Localization studies have shown that Ido1 is expressed by infiltrating myeloid cells and neoplastic epithelial cells56–58, and both cellular compartments could contribute to kynurenine production. Our results suggest that the neoplastic epithelium is an important source for kynurenine, because loss of IDO1+ Paneth cells in Stat1ΔIEC ApcMin tumors resulted in significantly reduced kynurenine levels that were not compensated by stromal kynurenine production. We speculate that neoplastic cells are major producers of kynurenine in tumors, whereas stromal myeloid cells use different metabolic routes. A contribution of Ido1-expressing neoplastic epithelial cells to immune escape has also been found in pancreatic ductal adenocarcinomas59 and high Ido1 expression in neoplastic epithelial cells at the invasive front is an independent adverse prognostic factor for overall survival and metastasis in CRC58,60,61.

Stat1 is considered as a tumor suppressor in solid cancers9 and we expected tumors of increased size in Stat1ΔIEC ApcMin mice. However, tumors were smaller and contained reduced numbers of Tregs and increased numbers of CD8+ T cells. Similar neoplastic and immunologic aberrations were observed in Ido1−/− ApcMin tumors62. This indicates that loss of IDO1+ Paneth cells and corresponding immunological consequences surpassed tumor-promoting effects of Stat1 deletion in Stat1ΔIEC ApcMin tumors. Interestingly, tumor formation was not affected in Stat1−/− ApcMin mice,63 but this study neglected compensating effects of stromal Stat1 deletion, which interferes with immunosurveillance and alleviates the need for immunosuppression.

Most patients develop sporadic CRC, whereas colitis-associated CRC (CAC) affects only 1–2% of human cases. Recent studies demonstrated that specific deletion of Ido1 in intestinal epithelial cells interfered with AOM-DSS-induced CAC formation in mice30. The oncogenic function of Ido1 in CAC was attributed to tumor cell-intrinsic phosphatidylinositol-3-kinase–Akt-mediated nuclear translocation of β-catenin rather than immunosuppression.30 Our results showed that Stat1 ablation and corresponding Stat1ΔIEC ApcMin tumors resulted in significantly reduced kynurenine levels that were not compensated by stromal kynurenine production. Our results showed that Stat1 ablation and corresponding Stat1ΔIEC ApcMin tumors resulted in significantly reduced kynurenine levels that were not compensated by stromal kynurenine production. We speculate that neoplastic cells are major producers of kynurenine in tumors, whereas stromal myeloid cells use different metabolic routes. A contribution of Ido1-expressing neoplastic epithelial cells to immune escape has also been found in pancreatic ductal adenocarcinomas59 and high Ido1 expression in neoplastic epithelial cells at the invasive front is an independent adverse prognostic factor for overall survival and metastasis in CRC58,60,61.

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sporadic CRC. We found increased AOM-DSS-induced CAC formation in male 
Stat1ΔIEC mice, while Stat1ΔIEC tumors contained reduced numbers of CD8+ T cells, although IDO1+ Paneth cells were absent (unpublished data). Therefore, Stat1-dependent IDO1+ Paneth cells might be particularly important for the development of sporadic tumors but dispensable for CAC.

It is challenging to deduce prognostic information of tumor cell-intrinsic Stat1 and Id1 expression from CRC TCGA data, because both genes are expressed in neoplastic cells and immune cells. Correspondingly, good prognosis of CRC patients with Stat1ΔIEC tumors might be primarily caused by enhanced anti-tumor activity of Stat1ΔIEC immune cells, whereas prognostic information of Stat1 expression in neoplastic cells is masked. Using IHC staining of human samples, we identified IFIT1 as a surrogate marker for STAT1-IDO1 expression in cancer cells of human CRC. IFIT1 was not detectable in stromal cells and regulated the replication of viruses, a function that should not impact on human CRC prognosis. IFIT1 surrogate expression enabled us to discriminate between tumor cell-intrinsic and stromal Stat1-Ido1 function using bulk gene expression TCGA data. These analyses suggested that tumor cell-intrinsic Stat1-Ido1 expression promotes progression of human CRC, correlates positively with Treg numbers and desensitizes tumors to immune attack. Moreover, Stat1-Ido1 expression correlated with Lysozyme expression in human TCGA data and IDO1+ Paneth cells were present in adenomas of FAP patients, indicating that human CRC also contain neoplastic IDO1+ Paneth cells.

Stat1-dependent IDO1+ Paneth cells were also found in normal murine crypts. They did not depend on ApcMin but the presence of the Min mutation affected their spatial distribution in the distal small intestine and colon. Extra clean SPF conditions and treatment of mice with antibiotics substantially reduced the number of IDO1+ crypts in all parts of the intestine. Moreover, IDO1+ Paneth cells in normal crypts were enriched in the distal small intestine, which has a high bacterial load, and we could identify Id1 induction in Paneth cells of bacteria-infected mice using scRNA-seq data. This suggests that IDO1+ Paneth cells are induced by the local microbiome. TLR9 and IFN signaling are candidate pathways that could promote Id1 expression in Paneth cells. The tumor studies with Ifnar1ΔIEC mice and Ifnα/β treated organoids suggested that Id1 is not induced by type I IFN or TLR9 signaling, in contrast, IFNγ readily induced Id1 in epithelial cells of organoids and IDO1+ Paneth cells were reduced in the intestines of Ifngr1−/− mice. However, in contrast to Stat1ΔIEC mice, IDO1+ Paneth cells were not completely abolished in Ifngr1−/− mice, indicating that additional factors are implicated in Stat1-Ido1 induction. Id1 is an IFNγ-inducible gene in human and murine tumor cell lines. The microbiome induces IFNγ production by mucosal T cells in mice and in humans, and depletion of bacteria reduces IFNγ levels. This indicates that interaction of lamina propria cells with the microbiome leads to the production of type II IFN that induces Stat1-dependent Id1 expression in Paneth cells of distinct intestinal crypts.

In summary, we identified Stat1-dependent IDO1+ Paneth cells in intestinal tumors and normal intestinal crypts. They might represent bone fide Paneth cells but need Stat1 for Id11 expression. IDO1+ Paneth cells could act as local immunosuppressors to prevent aberrant immune cell activation in response to bacteria. Hence, they could also provide immune-privileged niches for tumor formation (Fig. 7g). Early adenomas might use these niches to shield anti-tumor immune attack during elimination and equilibrium phases of immunoeediting. Consistent with this idea, neoplastic IDO1+ Paneth cells were particularly abundant in early adenomas of ApcMin mice. Of note, tumor formation was impaired in ApcMin mice kept under germ-free conditions. Targeting IDO1+ Paneth cells might improve efficacy of immunotherapy in microsatellite-stable CRC patients. Besides representing a conceptual advance, our findings will improve precision oncology of CRC.

**Methods**

**Mice.** Mice with floxed alleles of Stat1ΔIEC or Ifnar1ΔIEC were crossed to Villin-cre mice. Villin-cre Stat1ΔIEC and Villin-cre Ifnar1ΔIEC animals were crossed with ApcMin mice (Jackson Laboratory) to generate Villin-cre Stat1ΔIEC ApcMin−/− (Stat1ΔIEC ApcMin−/−) and Villin-cre Ifnar1ΔIEC ApcMin−/− (Ifnar1ΔIEC ApcMin−/−). Mice were kept on a C57BL/6 background and housed under standard conditions at the Dezentrale Biomedizinische Einrichtung of the Medical University Vienna (Stat1ΔIEC ApcMin−/−) and the Zentrale Versuchsanstalten of the Medical University Innsbruck (Ifnar1ΔIEC ApcMin−/−). Experiments were performed with adult (6-8 weeks old) or tumor-bearing (4 months old) male or female mice. To deplete commensal gut microbiota, adult wild-type C57BL/6 mice were given ampicillin (1 g/l), vancomycin (0.5 g/l), neomycin sulfate (1 g/l), and metronidazole (1g/l) in drinking water for 28 days. Water was changed every third day to ensure antibiotic stability. All mouse experiments were performed in accordance with Austrian and European laws (license numbers BMWF-66.009/0191-WFV/3b/2015 and BMWFV-66.009/0189-WFV/3b/2015) and with the general regulations specified by the Good Science Practices guidelines of the Medical Universities Vienna and Innsbruck.

**Human material.** Patient material from the Austrian Breast and Colorectal Cancer Study 91 (ABC91 Study, n=90035943) was used, which is a prospective, multicenter, randomized trial comparing the efficacy of adjuvant chemotherapy in stage II colon cancer. All patients provided written consent and the study was approved by the ethics committees at the participating institutions.

**Isolation of intestinal epithelial cells.** Intestinal epithelial cells were isolated from 10-12-week-old mice as described previously.

**Histology, immunohistochemistry, and immunofluorescence.** Tissues were flushed with phosphate-buffered saline (PBS) and 4% paraformaldehyde fixed and embedded in paraffin as swiss rolls. Swiss rolls were cut into 2 µm sections and IHC/IF-stained with standard procedures using antibodies against β-catenin (Becton Dickinson, 610153, 1:80), BrdU (BrdU In-Situ Detection Kit, Becton Dickinson, 550803), cleaved Caspase 3 (Cell Signalling, 9661, 1:200), Endomucin (eBioscience, 14-5851-82, 1:500), Granzyme B (Abcam, ab6059, 1:200), IDO1 (Biolegend, 122402, 1:80), iNOS (Biolegend, 610431, 1:200), Ki67 (Novacorsa, NKL-K67-P, 1:1000), Lysozyme (Dako, A009902, 1:100), p-STAT1 (Cell Signalling, 9167S, 1:100), p-STAT3 (Cell Signalling, 9145, 1:100), STAT1 (Santa Cruz, sc-592, 1:500), STAT3 (Santa Cruz, sc-7179, 1:80), Synaptophysin (GeneTex, GTX100865, 1:1000), GFP (Roche, 181146001, 1:1000), red fluorescent protein (RFP) (Rockland antibodies and assays, 600-401-379S, 1:500), CD3 (Neomarker RM9107, 1:100), MMP7 (Cell Signalling, 3801, 1:100). IHC staining on human samples was performed using antibodies against IFIT1 (Sigma Aldrich, HPA055380, 1:500), IOD1 (Biolegend, 122402, 1:100), p-STAT1 (Cell Signalling, 9167S, 1:100), and STAT1 (Cell Signalling, 14994, 1:1000).

**CRISPR/Cas9 of MC38 cells and transplantation.** MC3GΔGFP/GFP cells were generated using CRISPR-Cas9 as described previously. ID1 exon 6 was targeted using the following oligos: 5’-CAACCTGGGTTGAGGACTTCCGACA-3’ (forward) and 3’-AAACCTGTCGAGTCACCGAGA-3’ (reverse). Frequency of insertions/deletions in the transfected cell pool was estimated using the TIDE analysis software. The following primers were used for target site amplification: 5’-AAGCTTGGGCTTGGAGTTG-3’ (forward) and 5’-TCTAATCCGCTGAGGCTG-3’ (reverse). Single cells derived from the initially targeted cell pool were expanded independently, sequenced for DNA modifications using the above mentioned primers, and used for transplantation experiments. To label MC3GΔGFP/GFP cells with dsRed, 10 µg of DsRed-plenti plasmid (gift from Venugopal Bhaskara), 8 µg of packaging plasmid, and 2 µg of plasmid expressing envelope vector (SVG-G, Addgene plasmid # 14888), and 61 µl of 2 µM CaCl2 were diluted to 500 µl in ddH2O. The solution was then mixed with 500 µl of 2x HBSS (50 mM HEPES, 10 mM KCl, 12 mM Dextrose, 280 mM NaCl, 1.5 mM NaH2PO4 pH 7.05) and incubated for 10 min at room temperature, before being added to HKE293T cells and cytoplasmic tails. Target cells were incubated with virus-containing supernatant for 5 days. Cells 105 were injected subcutaneously into the flanks of 8-9-week-old male C57BL/6J mice or NSG mice. To account for host effects, cells of different genotypes were implanted in the left and right flanks of the same mouse. Mixed MC3GΔGFP/GFP/MC3GΔGFP/GFP tumors were evaluated for the presence of GFP- and MC3GΔGFP-positive cells via IHC. Individual MC3GΔGFP/GFP and MC3GΔGFP/GFP cells with 100% GFP+ or GFP+GFP−/dsRed− cells were used for normalization of data.

**MC38 cell stimulation.** MC3GΔGFP/GFP and MC3GΔGFP/GFP cells were cultured in Dulbecco’s modified Eagle’s medium containing 10% fetal calf serum (FCS), 1% human material from the Austrian Breast and Colorectal Cancer Study 91 (ABC91 Study, n=90035943) was used, which is a prospective, multicenter, randomized trial comparing the efficacy of adjuvant chemotherapy in stage II colon cancer. All patients provided written consent and the study was approved by the ethics committees at the participating institutions.
Penicillin/Streptomycin (10,000 U/ml) and 1% L-glutamine (200 mM). At 70–80% confluency, cells were stimulated with 100 ng/ml IFNγ (ImmunoTools, 123433S6) for 1 h and 24 h, in triplicates.

**Quantification and grading of ApoMMin tumors.** Swiss rolls were stained with hematoxylin and eosin, and scanned with a Pannoramic MIDI Slide Scanner (3D Histec), and histomorphometrically analyzed with DefiniensTM Developer software (Definiens). Grading was performed by a board certified pathologist (I.K.).

**Flow cytometry.** Intestinal tumors from single mice were pooled, minced and digested in 2 ml PBS containing 0.25% (v/v) FCS and 0.25% (w/v) collagenase IV (Life technologies, 17104-019) for 45° at 37°C under shaking. After spraying through a 70 µm mesh and washing twice with 30 ml PBS, cells were incubated with TrueStain FcX (Biologend, 101320) and Zombie Aqua Fixable Viability Kit (Biologend, 423102). Extracellular staining was performed using antibodies against CD45 (Biologend, 100728), CD45 (Biologend, 103128), CD4 (Biologend, 100408), CD3e (ebioscience, 35-0031-82), and CD25 (ebioscience, 25-0251-81). Cells were fixed (Fixation/Permeabilization Buffer, ebioscience, 00-5123-43), permeabilized (Permeabilization Buffer, ebioscience, 00-8333-56), and intracellular staining of FOXP3 (Biologend, 520011) and Granzyme B (Biologend, 515405) was performed. Data were collected using a FACS Fortessa (BD) and analyzed with FlowJo software.

**Enzyme-linked immunosorbent assay.** Single tumors of the small intestine and colon were homogenized in 60 µl PBS and centrifuged. Supernatants were used for kynurenine ELISA (EMELCA Bioscience, MBS043489) according to the manufacturer’s instructions. The results were normalized for the amount of protein.

**In-situ hybridization.** Duplex ISH was performed on formalin-fixed paraffin-embedded tissue samples using RNAscope assay (ACD, 322436) with probes against Lgr5 (ACD, 312171) and Id1 (ACD, 315971) according to the manufacturer’s instructions.

**RNA sequencing.** Total RNA from tumors was extracted using TRIzol Reagent (Thermo Fisher Scientific, 15596018) and processed for sequencing using the TruSeq RNA Sample Preparation Kit (Illumina, Inc.) according to the manufacturer’s protocol. The mRNA was purified using poly(T)-eligo-attached magnetic beads, fragmented, and applied to first-strand complementary DNA (cDNA) synthesis. Second-strand cDNA synthesis was performed using DNA polymerase I and RNAse H. cDNA was end-repaired, A-tailed, ligated to adapters, and amplified to create the final cDNA library for sequencing (HiSeq2000, Illumina, Inc.). TopHat2 algorithm was used to align raw RNA-seq data to mm10. Aligned bam files were deposited in ArrayExpress database (E-MTAB-5083). Differentially expressed genes were identified using DEseq2 algorithm. An adjusted p < 0.005 and a fold change > 2 or < 0.5 were defined as cut-off for differentially expressed genes. GO enrichment analyses were performed using GOrilla software.

**scRNA-seq analysis.** Pre-processed droplet-based scRNA-seq datasets from Haber et al.44 (GEO: GSE92332) were re-analyzed using the R package Seurat. For comparison of Id1 expression in Paneth cell clusters, the infection model datasets “SH_Salmonella” and “SH_Hpol” were used, as well as the accordingly control sets. Different infection durations (3 days and 10 days) within the “SH_Hpol” dataset were pooled. Furthermore, sequencing data of intestinal cells, specifically sorted with focus on large cells to improve Paneth cell yield, were analyzed. Dimensionality reduction was performed using gene expression data for a subset of variable genes. The variable genes were selected based on dispersion of binned variance to mean expression ratios using FindVariableGenes function of Seurat40 followed by filtering of ribosomal protein and mitochondrial genes. Next, principal component analysis (PCA) was performed and the data were reduced to the top 15 PCA (infection model)/10 PCA (large cells) components (number of components was chosen based on SDs of the principal components—in a plateau region of an elbow plot). Graph-based clustering of the PCA reduced data with the Louvain Method was used after computing a shared nearest-neighbor graph.49 The clusters were visualized on a two-dimensional map produced with t-SNE. The VPL plot function was applied to show expression probability distributions across the clusters and the FeaturePlot function to visualize feature expression within the clusters on a t-SNE plot. These methods were performed for marker genes of our cells of interest to identify Paneth and goblet cell clusters. Violin plot expression levels are depicted on a log transcripts per million (TPM) scale per cluster. Feature plot depicts a color scale for average gene expression. To identify further clusters containing Id1+ cells (TA/Stem, Tuft, enteroctyes), the top 50 specific marker genes for each cluster identified and described in Haber et al.43 were selected using the CaseMatch function and aggregated and matched to gene expression profiles of the clusters identified within this analysis using the MetaFeature function (calculation of relative contribution of each feature to each cell for given set of features).

**Polymerase chain reaction.** Wild-type, floxed and deleted Stat1 alleles were amplified using 5′-TAGGCTCTCCCTTTCTCTTCC-3′, 5′-ACACATTGGCTTCACAATTTCCTC-3′, and 5′-CCCCCTGTCATCTGGAGTGT-3′ primers. The Cre transgene was detected with 5′-CGTCGTCATGCAAGCGTGT-3′ and 5′-CCAAGCGAATATCATGCGTCG-3′ primers. ApoMMin genotyping was performed using 5′-TCTTGTTCTGAGAAGACAGAACT-3′ and 5′-TGTGATCTTCCAAAGCTTGGCTAT-3′ primers, and digestion of amplicons with HindIII.

**Quantitative PCR.** RNA was isolated with TRIzol (Life technologies, 15596-018) and reverse transcribed with QuantiTect Reverse Transcription Kit (Qiagen, 205313). qPCR was performed using Fast SYBR Green Mastermix (Thermo Fisher Scientific, 438656) and Applied Biosystems 7500 Fast Real Time PCR System with primers 5′-TGGTGAAATTGAGACAGGTC-3′ and 5′-TGTTTGCGTACCAAATGATGT-3′ for Stat1, 5′-ATGGGCGTTCCTGCTTCAACA-3′ and 5′-AAGGCTGCCGTCTCAATCA-3′ for Id1, and 5′-TGGTGTGATGGGTTGG-3′ and 5′-TAC TTGGAGGGTTTCCTC-3′ for Gpdh.

**Statistics and reproducibility.** Sample sizes and numbers of replicates are described in detail in the figure legends. Biological replicates were defined as parallel measurements of biologically distinct samples (mice in most cases). Each experiment was repeated at least three times. All values are given as means ± SEM. Normal distribution of data was tested and appropriate tests were performed: Student’s T-test or Mann–Whitney U-test. For more than two groups, one-way analysis of variance and Bonferroni’s post-hoc test or Kruskal–Wallis test, and Dunn’s post-hoc test were used. For analysis of the tumor and TMA gradings, χ2-test was used. Survival analyses using clinical data from CRC TCGA patients were performed using log-rank testing and GraphPad Prism 6 software. Correlation analyses of TCGA data were calculated using cor function of R3.2.1 software and visualization was performed using corplot and ggplot2 packages. No sample size estimation was performed. Samples were excluded as outliers according to Grubbs’ test (α = 0.05). Experiments were performed and analyzed in a blinded, randomized manner. Significant differences between experimental groups were stated as *p < 0.05, **p < 0.01, ***p < 0.001, or ****p < 0.0001.

**Reporting summary.** Further information on research design is available in the Nature Research Reporting Summary linked to this article.

**Data availability** RNA sequencing data were deposited in ArrayExpress database, accession number E-MTAB-5083. Previously generated single-cell RNA sequencing data analyzed here can be found in GEO (GSE92332). Processed (MapSplice aligned, RSEM quantified, quartile normalization standardized; Level 3 RnaSeqV2) RNA sequencing data of the COADREAD dataset were obtained from The Cancer Genome Atlas (TCGA) database. All other data that support the findings of this study are available from the corresponding author upon reasonable request.

Received: 7 August 2019; Accepted: 10 April 2020; Published online: 22 May 2020

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**Funding** This study was supported by ERC grant 803893 (SFBWNR) and was part of the program of the European同步脣脗 Outflow. This work was performed using the SimulaResearch computational infrastructure project supported by the Simula Research Foundation.

**Competing interests** The authors declare no competing interests.

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Acknowledgements
We thank Latifa Bakiri and Erwin F. Wagner for their critical reading of the manuscript. We thank Nina Braun for support with single-cell RNA-seq analysis. This work was supported by the Austrian Science Fund (FWF) Doktoratskolleg-plus grant “Inflammation and Immunity” (R.E., M.S., M.M., and B.S.), the FWF grants P29222-B28 (R.E.), F4709-B20 (G.H.), P26011 (L.K.), P32900 (E.C.), SFB F6101 (M.M. and B.S.), SFB F6106 (M.M. and B.S.), and the European Training Network MSCA-ITN-2015-ETN ALKA-TRAS No 675712 (L.K.).

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Competing interests
The authors declare no competing interest.

Additional information
Supplementary information is available for this paper at https://doi.org/10.1038/s42003-020-0989-y.

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