Pilot study of $^{89}$Zr-bevacizumab positron emission tomography in patients with advanced non-small cell lung cancer

Idris Bahce1*, Marc C Huisman2, Eline E Verwer2, Rogier Ooijevaar1, Firdaouss Boutkourt1, Danielle J Vugts2, Guus AMS van Dongen2,3, Ronald Boellaard2 and Egbert F Smit1

Abstract

**Background:** The aim of this pilot study was to evaluate whether the uptake of $^{89}$Zr-bevacizumab in non-small cell lung cancer (NSCLC) tumors could be visualized and quantified. The correlation between tumor $^{89}$Zr-bevacizumab uptake and tumor response to antitumor therapy with a bevacizumab-based regimen was explored.

**Methods:** Seven NSCLC patients underwent static PET scans at days 4 and 7 after injection of 36.4 ± 0.9 MBq (mean ± SD) $^{89}$Zr-bevacizumab, prior to commencing carboplatin-paclitaxel-bevacizumab chemotherapy (CPB). Overall survival (OS) and progression-free survival (PFS) to CPB followed by bevacizumab maintenance therapy was correlated to tumor tracer uptake, quantified using peak standardized uptake values (SUV$_{\text{peak}}$).

**Results:** Zr-bevacizumab uptake (SUV$_{\text{peak}}$) was approximately four times higher in tumor tissues (primary tumor and metastases) than in non-tumor tissues (healthy muscle, lung, and fat) on days 4 and 7. A positive trend but no significant correlation could be found between SUV$_{\text{peak}}$ and OS or PFS.

**Conclusions:** This pilot study shows that $^{89}$Zr-bevacizumab PET imaging in NSCLC is feasible. Further investigation to validate this technique as a predictive biomarker for selecting patients for bevacizumab treatment is warranted.

**Keywords:** NSCLC; VEGF; Immuno-PET; $^{89}$Zr-bevacizumab

**Background**

Vascular endothelial growth factor (VEGF) is an important mediator in non-tumoral and tumoral angiogenesis. VEGF-A, one of the five members of the VEGF family and generally referred to as VEGF, is overexpressed in many tumors [1,2]. In response to hypoxia, cells of mesenchymal, stromal, and epithelial origin (e.g., myocytes, blood platelets, and stromal cells in tumors) show paracrine secretion of VEGF-A. This increases VEGF-A concentrations locally in the tumor, causing a strong pro-angiogenic stimulus. The binding of VEGF-A to VEGF receptor subtypes 1 and 2 (VEGFR1 and VEGFR2), both transmembrane monomers expressed on vascular endothelial cells, causes dimerization of the VEGFR1/2 monomers, leading to the activation of intracellular pro-angiogenic signaling pathways [3-6]. Also, non-angiogenic effects of increased VEGF-A have been described [3,7,8].

The addition of bevacizumab, a monoclonal antibody directed against VEGF-A, to cytotoxic chemotherapy improves tumor response rates and provides a survival advantage over chemotherapy alone in several malignancies [9-12]. A phase III study showed a survival benefit for patients with non-small cell lung cancer (NSCLC) treated with carboplatin-paclitaxel-bevacizumab compared with chemotherapy alone [13]. However, on an individual basis, it is unclear who would benefit from bevacizumab therapy, as predictive markers are lacking, despite a large body of clinical research available to date.

Imaging tumor uptake of radiolabeled bevacizumab in patients using positron emission tomography (PET) may provide such a predictive marker, as PET imaging enables *in vivo* monitoring of physiological processes. To date, several clinical studies have been performed using antibodies with $^{89}$Zr labeling. In these studies, positive
image contrast is observed, and its relation to therapy response discussed [14-17]. The aim of this pilot study was to evaluate whether $^{89}$Zr-bevacizumab enables visualization of NSCLC tumors and to assess the ranges of tumor-to-background ratios and standardized uptake values. Additionally, the correlation between $^{89}$Zr-bevacizumab uptake and tumor response following treatment with a bevacizumab-based regimen was assessed.

**Methods**

**Study design**

**Patients**

In this prospective pilot study, patients with stage IV adenocarcinoma of the lung who were scheduled for combined carboplatin-paclitaxel-bevacizumab treatment were enrolled. Inclusion criteria were histologic diagnosis of non-squamous NSCLC, age of 18 years or older, performance status of 0 to 2 (WHO), a life expectancy of at least 12 weeks, and presence of at least one NSCLC lesion within the chest of at least 1.5-cm diameter as measured by computed tomography (CT). Exclusion criteria included claustrophobia, pregnancy, lactation, and use of concurrent or previous treatment with anticancer drugs within 30 days prior to scanning. The patients gave informed consent, and this study was approved by the Medical Ethics Review Committee of the VU University Medical Center.

**Imaging**

Prior to the start of therapy, the patients underwent PET/CT scans using $[^{18}F]$fluorodeoxyglucose (FDG) and $^{89}$Zr-bevacizumab.

**Therapy**

As scheduled prior to inclusion in this study, the patients received treatment with carboplatin (area under the concentration-time curve of 6.0 mg/mL/min)-paclitaxel (200 mg/m² of body surface area)-bevacizumab (15 mg/kg) and continuation maintenance bevacizumab upon non-progression after 4 cycles. Chemotherapy was given every 21 days until disease progression or unacceptable toxicity [13].

**Preparation of $^{89}$Zr-bevacizumab**

Bevacizumab was labeled with zirconium-89 using N-succinyl-desferrioxamine (N-suc-Df) as described previously [18,19]. All procedures were performed under aseptic conditions in a shielded laminar flow cabinet. In short, desferrioxamine (Desferal (Df), Novartis, Basel, Switzerland) was converted to N-succinyl-Df. Next, the hydroxamate groups were blocked with iron, and the succinic acid group converted to its tetrafluorophenol ester. The resulting Fe-N-suc-Df-TFP ester was reacted with bevacizumab under basic conditions (pH 9.5 to 9.7) for 30 min at room temperature. Subsequently, iron was removed using an excess of ethylenediaminetetraacetic acid (EDTA) for 30 min at 35°C, and the resulting N-suc-Df-bevacizumab was purified over a c and radiolabeled with $^{89}$Zr for 60 min in HEPES buffer at room temperature. Finally, $^{89}$Zr-N-suc-Df-bevacizumab was purified over a PD-10 column using 5 mg/mL gentisic acid in 0.9% NaCl (pH 4.9 to 5.4). The mean labeling efficiency was 71.1% ± 5.3%. The product was formulated and filter sterilized to 5 mg bevacizumab and 37 MBq Zr-89 per patient injection. These procedures resulted in a sterile final product with endotoxin levels <2.5 EU/mL. The radiochemical purity was >97% according to iTLC and HPLC, and the immunoreactivity as determined by an ELISA assay was >75%, which is optimal for this assay.

**PET scanning**

All patients underwent a routine FDG PET/CT scan within 4 weeks prior to starting therapy. One week prior to therapy, the patients were injected intravenously with 36.4 ± 0.9 MBq $^{89}$Zr-bevacizumab, and 10-min static PET/CT scans of the thoracic NSCLC lesion were performed on days 4 and 7 post-injection. These parameters were based on previous $^{89}$Zr-trastuzumab analysis, showing appropriate doses and timing for visualization and quantification of uptake in HER2-positive tumors to be 37 MBq and days 4 to 5 after injection, respectively [20]. The scans were performed on a Gemini TF-64 PET/CT scanner (Philips Medical Systems, Cleveland, USA). PET data were normalized, and all appropriate corrections were applied for dead time, decay, randoms, and scatter. Reconstruction of PET data was performed using the BLOB-OS-TF reconstruction algorithm with CT-based attenuation correction, resulting in a final voxel size of 4 × 4 × 4 mm³, matrix of 144 × 144 × 45, and a spatial resolution of 5 to 7 mm full-width at half maximum [21]. Additional smoothing was performed, as this was shown to optimize quantitative accuracy and harmonized image quality [22].

**$^{89}$Zr-bevacizumab uptake analysis**

FDG PET/CT scans were used to identify the exact location of the tumors. The PET data were analyzed using in-house-developed analysis tools within the IDL environment (Interactive Data Language Virtual Machine 6.2, RSI Inc., Boulder, CO, USA). Volumes of interest (VOI) were drawn manually on the CT images around the contours of the primary tumor (PT) and, if present, lymph node metastases (LNM) and non-lymph node metastases (NLNM). Additionally, VOI were drawn within the descending aorta (AD) and non-tumor tissue muscle (M), healthy lung (HL), and fatty tissue (FT). Standardized uptake value (SUV) parameters were calculated by normalizing VOI activity concentrations to injected.
dose and patient weight: SUV\textsubscript{peak} (spheric VOI of 1.2-cm diameter positioned around the voxel with the highest uptake) and SUV\textsubscript{mean} (mean activity in VOI/cc) from the AD VOI to calculate the tissue-to-blood ratio (TBR). SUV\textsubscript{peak} and TBR for the abovementioned VOI were then compared for the scans at days 4 and 7.

**Statistical analysis**

Statistical analysis was performed using SPSS software (SPSS for Windows 15.0, SPSS, Inc.). Correlations were explored using Spearman’s correlation coefficient (\( r_s \)). A two-tailed probability value of \( P < 0.05 \) was considered significant. Values in the text and tables are presented as mean ± standard deviation (SD) unless stated otherwise.

**Results**

Seven patients were included in this study. The patient characteristics are summarized in Table 1. Nine mediastinal LNM were identified using FDG PET/CT scans within the thoracic field of view. Six patients showed NLNM. All tumor lesions showed visible \(^{89}\text{Zr}\)-bevacizumab uptake. To illustrate the findings as reported below, typical examples of fused PET/CT images using FDG and \(^{89}\text{Zr}\)-bevacizumab in three NSCLC patients are provided in Figure 1.

In all tumorous tissues (PT, LNM, and NLNM), \(^{89}\text{Zr}\)-bevacizumab uptake (SUV\textsubscript{peak}) was approximately four times higher than those in non-tumorous tissues (M, HL, and FT) on days 4 and 7, as shown in Table 2. For tumorous tissues, the SUV\textsubscript{peak} in NLNM and LNM was approximately 54% and 26% higher as compared to that in PT on day 4, and 49% and 19% higher on day 7, respectively (see Figure 2).

Remarkably, on day 4, activity concentration in blood (AD) was approximately the same as that in the PT, while the healthy tissue activity was clearly lower than that in PT (see Table 2 and Figures 1 and 2). However, between days 4 and 7, tumor TBR increased from 1.23 ± 0.4 to 2.2 ± 1.2.

SUV\textsubscript{peak} correlated strongly between days 4 and 7, as indicated by an \( r_s \) of 1 \( (P < 0.001) \). Figure 3 shows the correlation between days 4 and 7 for SUV\textsubscript{peak}.

As shown in Figure 4, correlations between tumor SUV\textsubscript{peak} and outcome parameters showed a positive trend, but did not reach statistical significance. SUV\textsubscript{peak} showed weak correlation with both PFS and OS on days 4 and 7. \( r_s \) was 0.64 with a \( P = 0.139 \) for both PFS and OS on days 4 and 7, respectively.

**Discussion**

**Visualization of tumor lesions**

Prior to this study, it was unclear whether NSCLC tumors could be imaged by \(^{89}\text{Zr}\)-bevacizumab. The results of this pilot study show that all tumor lesions (PT, LNM, and NLNM) had a higher \(^{89}\text{Zr}\)-bevacizumab uptake as compared to non-tumor background tissues, allowing for visualization and analysis of tumor \(^{89}\text{Zr}\)-bevacizumab uptake.

This increased uptake may be caused by accumulation of VEGF in the tumor, due to high paracrine expression and subsequent binding to extracellular matrix glycoproteins such as heparan sulfate proteoglycans (HSPG) and neuropilins (NRP). These glycoproteins act as non-signaling co-receptors that facilitate binding of VEGF to VEGFR molecules [6]. Another mechanism that may contribute is the internalization of \(^{89}\text{Zr}\)-bevacizumab into cells within the tumor. After internalization, the \(^{89}\text{Zr}\) label may become trapped in the lysosomes and show up on the PET scan [23].

Our findings are in accordance with the limited number of publications of previous preclinical and clinical data showing that high-VEGF-A-expressing tumors are associated with high tumor-to-background \(^{89}\text{Zr}\)-bevacizumab uptake. Nagengast et al. [19,24] showed

**Table 1 Patient characteristics**

| Number | Sex | Age (years) | Primary tumor | Evaluable metastasis | Histology | Bevacizumab maintenance therapy | Best response | Site of progression | PFS (weeks) | OS (weeks) |
|--------|-----|-------------|---------------|----------------------|-----------|-------------------------------|---------------|---------------------|-------------|------------|
| 1      | F   | 56          | RLL           | -                    | AC        | Yes                           | PR            | LLL                 | 56          | 79         |
| 2      | M   | 51          | RUL           | Peritoneum           | AC        | Yes                           | PR            | PC                  | 13          | 17         |
| 3      | M   | 59          | RLL           | Bone                 | AC        | Yes                           | PR            | RUL\textsuperscript{a} | 27          | 48         |
| 4      | M   | 61          | RLL           | Bone                 | LCC       | Yes                           | CR            | RA                  | 35          | 63         |
| 5      | F   | 54          | RLL           | Bone                 | AC        | Yes                           | PR            | Brain               | 54          | 74         |
| 6      | F   | 69          | RH            | Pericardium          | AC        | Yes                           | PR            | MC                  | 34          | 60         |
| 7      | M   | 51          | LH            | Bone                 | AC        | Yes                           | No            | PD                  | 3           | 3          |

Evaluable metastases are non-primary lesions, located within the PET field of view. All patients received bevacizumab maintenance therapy, except for one patient, who died before bevacizumab maintenance therapy could be administered. The best tumor response to chemotherapy is shown. Progressive disease was seen in all patients. \( \text{\textsuperscript{a}} \)Two patients developed clinical disease progression before radiological progression. One patient developed progression due to brain metastasis. Sites of radiological progression are shown. PFS and OS were calculated in weeks, between start of chemotherapy and date of progression and death, respectively. AC, adenocarcinoma; CR, complete response; F, female; LCC, large cell carcinoma; LH, left hilum; LLL, left lower lobe; M, male; MC, meningitis carcinomatosa; OS, overall survival; PC, peritonitis carcinomatosa; PD, progressive disease; PFS, progression-free survival; PR, partial response; RA right adrenal gland; RH, right hilum; RLL, right lower lobe; RUL, right upper lobe; SDC, salivary duct carcinoma.
that high-VEGF-producing SKOV-3 ovarian tumor xenografts had a higher uptake of $^{89}$Zr-bevacizumab than of $^{89}$Zr-labeled IgG, which served as a control. In a follow-up study, the same researchers found that $^{89}$Zr-bevacizumab uptake decreased in (NVP-AUY922 sensitive) A2780 ovarian tumor xenografts after 2 weeks of antitumor therapy (using NVP-AUY922), while in the therapy-resistant CP70 xenografts, $^{89}$Zr-bevacizumab uptake did not change. van der Bilt et al. [25] showed a decrease in $^{89}$Zr-bevacizumab uptake in A2780 ovarian tumor xenografts after 2 weeks of antitumor therapy (using NVP-AUY922), while in the therapy-resistant CP70 xenografts, $^{89}$Zr-bevacizumab uptake did not change.

Table 2

| Tumorous tissue | Blood (AD) | Non-tumorous tissue |
|-----------------|------------|---------------------|
| Tumor tissue    | PT LNM NLNM | Muscle Fat Lung     |
| Day 4           |            |                     |
| $\text{SUV}_{\text{peak}}$ | 4.3 (2.1 to 6.9) | 4.0 (2.1 to 12.1) | 5.0 (2.9 to 14.5) | 3.9 (1.7 to 4.9) | 1.0 (0.4 to 1.1) | 0.6 (0.3 to 0.7) | 0.8 (0.5 to 1.4) |
| TBR             | 1.2 (0.7 to 1.8) | 1.1 (0.8 to 7.2) | 1.2 (0.7 to 8.6) |                     | 0.3 (0.1 to 0.3) | 0.2 (0.1 to 0.2) | 0.3 (0.2 to 0.3) |
| Day 7           |            |                     |
| $\text{SUV}_{\text{peak}}$ | 3.3 (0.7 to 4.9) | 2.7 (1.0 to 9.0) | 3.9 (1.1 to 9.7) | 1.4 (0.9 to 3.5) | 0.5 (0.2 to 1.3) | 0.4 (0.1 to 0.6) | 0.4 (0.2 to 1.1) |
| TBR             | 0.8 (1.0 to 4.0) | 1.6 (1.1 to 15.3) | 1.9 (0.9 to 16.5) |                     | 0.5 (0.3 to 0.6) | 0.3 (0.1 to 0.4) | 0.4 (0.2 to 0.6) |

AD, aorta descendens; LNM, lymph node metastases; NLNM, non-lymph node metastases; PT, primary tumor; TBR, tumor-to-blood ratio. Median (range) is shown.
xenografts after 2 weeks of everolimus therapy, while \( ^{89}\text{Zr}\)-bevacizumab uptake remained unchanged in other organs, matching \textit{ex vivo} measures of VEGF-A levels [19,24,25]. Recently, a first clinical study using \( ^{89}\text{Zr}\)-bevacizumab was reported. In breast cancer patients, tumors with elevated VEGF-A levels showed higher \( ^{89}\text{Zr}\)-bevacizumab uptake than the background of healthy breast tissue [26].

We found that tumor-to-background ratios were high at days 4 and 7; however, tumor-to-blood ratios were higher on day 7, due to a relative decrease of blood activity concentrations as compared to tumor activity concentrations. However, image quality at day 7 was hampered due to physical decay of the tracer. The optimal time for \( ^{89}\text{Zr}\)-bevacizumab seems to be between 4 and 7 days post-injection.

Lymph node metastases and especially NLNM showed higher uptake than pulmonary lesions. This may be caused by the fact that pulmonary lesions suffered more from breathing movement-induced partial volume effects than LNM and NLNM, which were relatively fixed to bone and peritoneum. Although all tumoral lesions were visible in this study, the quantification of uptake in small-volume lesions needs to be interpreted with caution, as these tumors suffer even more from this partial-volume-induced underestimation of tracer uptake.

**Clinical outcome**

Although our results did not show a significant correlation between tracer uptake and PFS, a positive trend was observed. As tumor \( ^{89}\text{Zr}\)-bevacizumab uptake may represent the level of tumor VEGF, this technique might offer a predictive biomarker for bevacizumab treatment efficacy. At present, there is an absence of clinically useful predictive biomarkers. For example, several blood biomarkers, such as VEGF-A and NRP-1 using newly developed sensitive assays, have been proposed to predict bevacizumab treatment efficacy, but their value is still the subject of debate [6,27].

**Limitations**

As this was a pilot study, only a limited number of patients were included. However, the results obtained were consistent for all patients, indicating that larger clinical trials are warranted.

Another limitation was the absence of arterial blood sampling for blood and plasma activity and metabolite analysis, which could have provided a more accurate quantification of tracer uptake. However, because patients already underwent several PET/CT scans, additional blood sampling for research purposes was considered too high a burden. Furthermore, previously published data show a good correlation between image-derived activity from \( ^{89}\text{Zr}\)-labeled antibodies and blood activity [15,17]. Additionally, \( ^{89}\text{Zr}\)-bevacizumab was found to be highly stable in plasma, as only a 6% decrease in protein-bound radioactivity was seen after 168 h (stored in serum at
Although NSCLC was initially diagnosed in all patients, no extra tumor biopsy was taken prior to scanning for VEGF-A staining, again because this was considered too burdensome.

Future perspectives
The results of this study show tumor specific uptake of \(^{89}\text{Zr}\)-bevacizumab. Future studies should consider not to include small lesions that could suffer from partial volume effects. Furthermore, the optimal timing should be investigated, as this can be expected to be between day 4 and day 7 post-injection. To better understand the physiological processes causing tracer accumulation, concurrent pathology data (e.g., angioproliferative markers) should be assessed by taking biopsies prior to scanning. Quantification of uptake may be improved by using blood sampling for plasma radioactivity assessment.

The effect of perfusion on \(^{89}\text{Zr}\)-bevacizumab should be analyzed. In this study, an assessment of tumor blood perfusion, e.g., with \(^{15}\text{O} \text{H}_{2}\text{O}\) PET scan, was not performed. Using \(^{15}\text{O} \text{H}_{2}\text{O}\) PET scans, van der Veldt et al. [28] showed that the intratumoral distribution of docetaxel followed the tumor perfusion patterns. Therefore, tumor perfusion data may have provided additional understanding of the distribution of tumoral \(^{89}\text{Zr}\)-bevacizumab uptake.

\(^{89}\text{Zr}\)-bevacizumab PET may be used as an imaging agent, as tracer uptake showed a trend towards a positive correlation with PFS and OS. It should be noted that patients were treated with carboplatin-paclitaxel-bevacizumab (CPB) followed by bevacizumab maintenance therapy. PFS was the result of both CPB therapy and bevacizumab. In future studies, \(^{89}\text{Zr}\)-bevacizumab scans should ideally be performed at two time points, i.e., prior to CPB therapy and also prior to bevacizumab maintenance therapy. This is because the post-CPB-altered tumor VEGF status is unknown; however, this new post-CPB VEGF status may be a better predictor for sensitivity to subsequent bevacizumab maintenance therapy.

Conclusions
This pilot study demonstrates that \(^{89}\text{Zr}\)-bevacizumab PET imaging in tumors is feasible. Larger studies are needed to validate and substantiate these findings. \(^{89}\text{Zr}\)-bevacizumab PET merits further investigation, aiming to evaluate its use as a predictive imaging biomarker.

Abbreviations
AD: aorta descendens; CPB: carboplatin-paclitaxel-bevacizumab; CT: computed tomography; FDG: fluorodeoxyglucose; FT: fatty tissue; HL: healthy lung; LNM: lymph node metastasis; M: muscle; MBq: megabecquerels; NLNM: non-lymph node metastasis; NRP: neuropilin; NSCLC: non-small cell lung cancer; OS: overall survival; PET: positron emission tomography; PFS: progression-free survival; PT: primary tumor; SUV: standardized uptake value; TBR: tumor-to-blood ratio; VEGF: vascular endothelial growth factor; VOI: volume of interest; WHO: World Health Organization; Zr: zirconium.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
IB was involved in the design of the study, data collection, and initial writing of the manuscript. MH, EV, RO, and FB carried out the multiple processing steps of the acquired raw PET data and drafted the manuscript. DV and GvD carried out tracer synthesis. RB and ES participated in conceiving, designing, and coordinating the study. All authors read and approved the final manuscript.

Acknowledgements
This study was performed within the framework of the Center for Translational Molecular Medicine (CTMM, Netherlands) AIRFORCE project (grant 03O-103). Funding was used for tracer synthesis, PET scanning, and personnel costs. CTMM had no role in the study design; in the collection, analysis, and interpretation of data; in the writing of the manuscript; and in the decision to submit the manuscript for publication.
Identity with respect to tumor angiogenesis.

1. Okines AFC, Reynolds AR, Cunningham D. Targeting angiogenesis in esophageal adenocarcinoma. Oncologist 2011, 16(44-58).
2. Burger RA. Overview of anti-angiogenic agents in development for ovarian cancer. Gynecol Oncol 2011, 121:230-238.
3. Maharaj ASR, D’Amore PA. Roles for VEGF in the adult. Microvasc Res 2007, 74:100-113.
4. Kerbel RS. Tumor angiogenesis. N Engl J Med 2008, 358:2039-2049.
5. Maharaj ASR, Saint-Geniez M, Maldonado AE, D’Amore PA: Vascular endothelial growth factor localization in the adult. Am J Pathol 2006, 168:659-668.
6. Wu FTH, Stefanini MO, Mac Gabhann F, Kontos CD, Annex BH, Popel AS: A systems biology perspective on SVEF1: its biological function, pathogenic role and therapeutic use. J Cell Mol Med 2010, 14:528-552.
7. Lee S, Chen TT, Barber CL, Jordan MC, Murdock J, Desai S, Ferrara N, Nagy A, Ross RP, Ingle-Anispe CL: Autoimmune VEGF signaling is required for vascular homeostasis. Cell 2007, 130:701-709.
8. D’Amore PA: Vascular endothelial cell growth factor-A: not just for endothelial cells anymore. Am J Pathol 2007, 171:141-18.
9. Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N: Humanization of an anti-vascular endothelial growth factor monoclonal antibody for the therapy of solid tumors and other disorders. Cancer Res 1997, 57:4593-4599.
10. Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N: Neutralizing anti-vascular endothelial growth factor antibody completely inhibits angiogenesis and growth of human prostate carcinoma micro tumors in vivo. Prostate 1998, 35:3-10.
11. Bartström P, Bourdon MA, Hillan KJ, Sriramarao P, Ferrara N: Neutralizing anti-vascular endothelial growth factor antibody completely inhibits angiogenesis and growth of human prostate carcinoma micro tumors in vivo. Prostate 1998, 35:3-10.
12. Reck M, von Pawel J, Zatloukal P, Ramlau R, Gorbounova V, Hirsh V, Leighl N, Mezger J, Archer V, Moore N, Manegold C: Predictive biomarkers for bevacizumab: are we there yet? Clin Cancer Res 2013, 19:8284-8287.
13. van der Veldt AA, Lubbrenik M, Bahce I, Walraven M, de Boer MP, Greuter HJNM, Hendrikse NH, Eriksson J, Windhorst AD, Postmus PE, Verheul HM, Serné EH: Rapid decrease in delivery of chemotherapy to tumors after anti-VEGF therapy: implications for scheduling of anti-angiogenic drugs. Cancer Cell 2012, 21:89-21.
14. Perk LR, Visser OJ, Stigter-van Walsum M, Vosjan MWD, Visser GWM, Zijlstra JM, Huijgens PC, van Dongen GA, de Vries EG, Schröder CP: Zr: from 89Zr:bevacizumab PET of early antiangiogenic tumor response to treatment with HS990 inhibitor NVP-AUY922. J Nucl Med 2010, 51:761-767.
15. van der Veldt AA, van Scheltinga AGT, Timmer-Bosscha H, Schröder CP, Pot L, Kostikin JGW, van der Zee AGI, Luther DE, de Zee EG, Reynolds AKL: Measurement of tumor VEGF-A levels with 89Zr-bevacizumab PET as an early biomarker for the antiangiogenic effect of everolimus treatment in an ovarian cancer xenograft model. Clin Cancer Res 2012, 18:630-6314.
16. Gaylema SMB, Brouwers AH, De Boer MP, Greuter HJNM, Pleijhuis RG, Timmer-Bosscha H, Pot L, van der Veeren BJ, van der Meulen SB, de Jong JR, Bart J, de Vries EJ, de Zee EG, Schröder CP: 89Zr-bevacizumab PET imaging in primary breast cancer. J Nucl Med 2013, 54:1014-1018.
17. Maru D, Venook AP, Ellis LM: Predictive biomarkers for bevacizumab: are we there yet? Clin Cancer Res 2013, 19:8284-8287.
18. van der Veldt AA, Lubbrenik M, Bahce I, Walraven M, de Boer MP, Greuter HJNM, Hendrikse NH, Eriksson J, Windhorst AD, Postmus PE, Verheul HM, Serné EH: Rapid decrease in delivery of chemotherapy to tumors after anti-VEGF therapy: implications for scheduling of anti-angiogenic drugs. Cancer Cell 2012, 21:89-21.