Inflammation plays a crucial role in the pathophysiology of acute ischemic stroke. In the ischemic cascade, resident microglia are rapidly activated in the brain parenchyma and subsequently trigger inflammatory mediator release, which facilitates leukocyte-endothelial cell interactions in inflammation. Activated leukocytes invade the endothelial cell junctions and destroy the blood-brain barrier integrity, leading to brain edema. Toll-like receptors (TLRs) stimulation in microglia/macrophages through the activation of intercellular signaling pathways secretes various proinflammatory cytokines and enzymes and then aggravates cerebral ischemic injury. The secreted cytokines activate the proinflammatory transcription factors, which subsequently regulate cytokine expression, leading to the amplification of the inflammatory response and exacerbation of the secondary brain injury.

Traditional Chinese medicines (TCMs), including TCM-derived active compounds, Chinese herbs, and TCM formulations, exert neuroprotective effects against inflammatory responses by downregulating the ischemia-induced microglial activation, microglia/macrophage-mediated cytokine production, proinflammatory enzyme production, intercellular adhesion molecule-1, matrix metalloproteinases, TLR expression, and deleterious transcription factor activation. TCMs also aid in upregulating anti-inflammatory cytokine expression and neuroprotective transcription factor activation in the ischemic lesion in the inflammatory cascade during the acute phase of cerebral ischemia. Thus, TCMs exert potent anti-inflammatory properties in ischemic stroke and warrant further investigation.

1. Introduction

Stroke is the third leading cause of death in developed countries [1] and the major cause of severe long-term disability worldwide [1–3]. Approximately 15 million people experience stroke annually. Of these, one-third die and one-third experience permanent disabilities, thus imposing considerable social and economic burden [4]. Approximately 80%–85% of all stroke events are ischemic caused by cerebral arterial thrombosis or embolism [5, 6]. To date, recombinant tissue plasminogen activator (rtPA) is the only Food and Drug Administration-approved medical therapy for acute ischemic stroke. However, rtPA has severe disadvantages, including the narrow therapeutic time window of 4.5 h and potential risk of hemorrhagic transformation; therefore, the eligibility of rtPA is reduced to only 4%–7% in all the patients with acute ischemic stroke [5]. Thus, potential therapeutic strategies for ischemic stroke are urgently needed.

Increasing evidence has demonstrated that inflammation plays a pivotal role in the pathophysiology of acute ischemic stroke [3, 5, 7]. During acute ischemic stroke, the brain is injured by ischemia- and inflammation-related primary and secondary insults [5]. The primary injury occurs at the beginning of ischemia; it rapidly interrupts the cerebral blood flow to the ischemic core and subsequently causes a significant decrease in oxygen and glucose supply to
cerebral neurons [8, 9]. The secondary injury is attributed to the postischemic inflammatory cascade, which produces various proinflammatory mediators, including cytokines, chemokines, proteases, and cell adhesion molecules, leading to an exacerbated ischemic brain injury [10]. However, the postischemic inflammatory response has a disadvantage and an advantage, exacerbating ischemic brain damage in the early phase and triggering tissue regeneration in the delayed phase, respectively [1, 2].

The lack of effective and widely applicable therapeutic strategies for the treatment of ischemic stroke has triggered increasing interest in traditional medicines, particularly traditional Chinese medicine (TCM) [11, 12]. Several centuries ago, TCM was used in China to treat cerebrovascular disorders, including stroke. Evidence revealed that TCM preparation, Chinese herb medicine, and TCM-derived active compounds exert anti-inflammatory effects by inhibiting inflammatory mediators, leukocyte infiltration, and blood-brain barrier (BBB) disruption in experimental cerebral ischemia [13]. These potent effects of TCMs against cerebral ischemic injury highlight their potential in clinical applications. Therefore, this review summarized the origin and development of the postischemic inflammatory cascade and delineated the anti-inflammatory effects of TCMs (namely, TCM-derived active compounds, Chinese herbs, and TCM formulations) on the basis of the in vivo literature.

2. TCM-Mediated Downregulation of Microglial Activation

2.1. Activation of Microglia in the Initial Phase of Cerebral Ischemia. In the acute phase (min to h) of cerebral ischemia, ischemic injury triggers a rapid activation of resident microglia in the brain parenchyma [3, 14]. During cerebral ischemia, microglial morphology changes from a ramified to an amoeboid shape upon activation [15]. In the initial stage of ischemia, the injured neurons expose damage-associated molecular patterns (DAMPs), which are subsequently recognized by toll-like receptors (TLRs), such as TLR4, and other pattern recognition receptors on the surface of the reactive microglia; this recognition triggers microglia-mediated inflammatory mediators release, contributing to secondary damage after stroke [6, 10, 16]. Reactive microglia/macrophages can be detected as early as 2 h after cerebral ischemia and maintained up to 1 week after the ischemic insult [6]. Reactive microglia are divided into two phenotypes: the classically and alternatively activated phenotypes (M1 and M2, resp.) [17]. The M1 microglia produce proinflammatory mediators, such as cytokines [interleukin- (IL-) 1β, IL-6, IL-18, and tumor necrosis factor- (TNF-) α], chemokines [monocyte chemoattractant protein- (MCP-) 1, and macrophage inflammatory protein- (MIP-) 1α], interferon- (IFN-) γ, matrix metalloproteinase- (MMP-) 9, and reactive oxygen species (ROS) [18], exerting detrimental functions in the early phase. By contrast, the M2 microglia secrete anti-inflammatory mediators, such as IL-4, IL-10, IL-13, transforming growth factor- (TGF-) β, and insulin growth factor-1, exerting neuroprotective effects in the delayed phase [2, 3, 19]. In cerebral ischemia, microglial activation is accompanied by reactive astrogliosis, which also produces an excessive amount of cytokines and causes the exacerbation of ischemic brain injury [18].

2.2. The Effects and Mechanisms of TCMs on Inhibiting Microglial Activation in In Vivo Models of Cerebral Ischemia. Hsieh et al. reported that Paenol, a common compound of Paeonia suffruticosa Andrews (Chinese name, Mu Dan Pi; Moutan cortex), reduces cerebral infarct and neurological deficits at 1.5 h of ischemia and 24 h of reperfusion. Paenol exerts anti-infarct effect mainly by inhibiting microglial activation and IL-1β expression in the ischemic cortex in ischemia/reperfusion- (I/R-) injured rats [20]. Pretreatment with tetramethylpyrazine (TMP), an active compound isolated from Ligusticum wallichii Franch (Chuan Xiong), effectively reduces the cerebral volume by inhibiting myeloperoxidase (an inflammation marker) and ED1 (a microglia/macrophage marker) expression in the ischemic core 72 h after reperfusion. The effect of TMP against microglial activation-mediated neurotoxicity can be further attributed to the suppression of prostaglandin E2 (PGE2) in the ischemic core [21]. A later study reported that TMP provides neuroprotection against ischemic brain injury partially by inhibiting microglial activation and subsequently downregulating MCP-1 expression in the ischemic cortex 72 h after reperfusion [22]. Andrographolide, the major active compound derived from Andrographis paniculata (Chuan Xin Lian) protects against cerebral infarction and ameliorates neurological deficits 24 h after permanent middle cerebral artery occlusion (MCAo). Andrographolide exerts neuroprotective effects partially by inhibiting microglial activation and microglia-mediated IL-1β and TNF-α expression in the ischemic area [23]. Pretreatment with Sophora japonica L. (Hua Hua; intraperitoneal injection) effectively reduced the cerebral infarct area and neurological deficits at 1.5 h of ischemia and 24 h of reperfusion. The effects of S. japonica involve the suppression of microglial activation and microglia-mediated IL-1β expression in the ischemic cortex [24]. Lim et al. demonstrated that an intragastric administration of total isoflavones isolated from Pueraria lobata (Ge Gen; TIPL) significantly reduces the cerebral infarct volume at 2 h of ischemia and 48 h of reperfusion. The anti-inflammatory effect of TIPL is partially attributed to the inhibition of astrocyte and microglial activation in the hippocampal CA1 region 7 d after MCAo [25]. On the basis of these studies, the anti-inflammatory effects of TCMs against cerebral ischemic injury could be attributed to the downregulation of microglial activation and microglia-mediated proinflammatory cytokines production in the ischemic area during the initial phase of MCAo (Figure 1 and Table I).

3. TCM-Mediated Suppression of Leukocyte Infiltration

3.1. The Process of Leukocyte Infiltration during Cerebral Ischemia. The microglial and astrocytic production of proinflammatory mediators rapidly increase the expression of
adhesion molecules on the endothelium [26, 27]. During acute cerebral I/R injury, the peripheral leukocytes first roll, become activated, and consequentially attach to the endothelial cells in the ischemic lesion. Leukocyte-endothelial cell interactions in inflammation are mediated by various adhesion molecules, including selectins, integrins, intercellular adhesion molecule-1 (ICAM-1), and vascular cell adhesion molecule-1 [28]. Selectins, comprising L-selectin on leukocytes and E- and P-selectins on endothelial cells, are a family of lectin-like adhesion glycoproteins that regulate leukocyte rolling and recruitment [28, 29]. Integrins, including leukocyte function-associated antigen-1 (CD11a/CD18) expressed on all leukocytes and macrophage-1 (MAC-1; CD11b/CD18) expressed on neutrophils and monocytes, are transmembrane glycoproteins and mediate leukocyte-endothelium interactions [30]. In acute cerebral ischemia, the upregulation of integrins facilitates a firm adherence of leukocytes to endothelial ICAM-1; the leukocytes subsequently penetrate the endothelial basement membrane into the brain parenchyma. Thus, the tight junctions (TJs) between endothelial cells of the BBB are disrupted and become more permeable, leading to leukocyte infiltration [30]. This evidence revealed that circulating leukocytes adhere to the damaged endothelium as early as 4 h and achieve the peak at approximately 12–48 h after ischemic brain injury. Meanwhile, reactive microglia, platelets, and infiltrating leukocytes further release IL-1β, IL-6, TNF-α, ROS, MCP-1, MIP-1α, IL-8, and MMPs (mainly MMP-9) that exacerbate ischemic injury in MCAo [3, 5, 6].

3.2. The Effects and Mechanisms of TCMs on Inhibiting Leukocyte Infiltration in In Vivo Models of Cerebral Ischemia.

Lu et al. reported that emodin, an active component of the rhizome of *Rheum palmatum* L. (Da Huang), effectively reduces the cerebral infarct size 6 h after MCAo. The neuroprotective effects of emodin can be attributed to ICAM-1 downregulation in the ischemic area in the early phase of cerebral ischemia [31]. Ferulic acid (FA), a major active compound in both *Angelica sinensis* (Oliv.) Diels (Dang Gui)
and *Ligusticum chuanxiong* Hort. (*Chuan Xiong*), effectively reduces the cerebral infarct area and ameliorates the neurological deficit at 1.5 h of reperfusion and 24 h of reperfusion. FA exerts anti-inflammatory effects against cerebral I/R injury, at least partially, by inhibiting ICAM-1, mRNA, and Mac-1 mRNA expression in the ischemic striatum 2 h after reperfusion [32, 33]. Bu-yang Huan-wu decoction (BHD), composed of *Astragalus membranaceus* (Shao Yao), *A. sinensis* (Oliv.) *Diels* (Dang Gui), *Paeonia lactiflora* Pall (Shao Yao), *L. chuanxiong* (Chuan Xiong), *Prunus persica* (L.) Batsch (Tao Ren), *Carthamus tinctorius* L. (Hong Hua), and *Pherezzma aspergilium* (Di Long), effectively ameliorates cerebral infarction and improves neurological deficits 24 h after forebrain ischemia.

The beneficial effects of Borneol involve the reduction of leukocytes (PMNs) in the ischemic area 2 h after ischemia [37]. However, during the acute phase of cerebral ischemia, the infiltrated leukocytes and reactive microglia synthesize and secrete MMPs (mainly MMP-2 and MMP-9) and ROS, thus increasing BBB permeability [72, 73]. Previous studies have reported that MMP-9 activation is initiated as early as 4 h, which reaches the maximum level at 24 h and persists for at least 5 d after cerebral ischemia [74, 75], whereas activated MMP-2 reaches the highest level 5 d after MCAo [75]. Active MMPs disrupt BBB integrity by degrading the

### 4. TCM-Mediated Stabilization of Blood-Brain Barrier Integrity

#### 4.1. Blood-Brain Barrier Disruption during Cerebral Ischemia

Under normal conditions, leukocyte recruitment across the BBB into the brain parenchyma contributes to the maintenance of the central nervous system immune privilege [9]. The BBB comprising endothelial cells, the basement membrane, the astrocyte end-feet, and pericytes provides a highly selective permeability barrier that separates the blood cells from the brain interstitial fluid and maintains brain homeostasis [38]. However, during the acute phase of cerebral ischemia, the infiltrated leukocytes and reactive microglia synthesize and secrete MMPs (mainly MMP-2 and MMP-9) and ROS, thus increasing BBB permeability [72, 73]. Previous studies have reported that MMP-9 activation is initiated as early as 4 h, which reaches the maximum level at 24 h and persists for at least 5 d after cerebral ischemia [74, 75], whereas activated MMP-2 reaches the highest level 5 d after MCAo [75]. Active MMPs disrupt BBB integrity by degrading the
Table 2: TCMs suppress leukocyte infiltration in the inflammatory cascade in ischemic stroke models.

| TCMs                          | Isolated from the Chinese herb (Chinese name) | Anti-inflammatory actions | Models                          | References |
|-------------------------------|---------------------------------------------|---------------------------|---------------------------------|------------|
| Emodin                        | Da Huang                                    | ICAM-1, TNF-α, IL-1β↓     | MCAo 6 h of ischemia            | [31]       |
| Ferulic acid                  | Dang Gui or Chuan Xiong                     | ICAM-1, ICAM-1 mRNA↓, MPO↓, NF-κBp50↓ | MCAo 1.5 h of ischemia followed by 2 or 24 h of reperfusion | [32, 33]   |
| Bu-yang Huan-wu decoction     | Huang Qi, Dang Gui, Shao Yao, Chuan Xiong, Tao Ren, Hong Hua, Di Long | CD11b↓                    | MCAo 0.5 h of ischemia followed by 24 h of reperfusion | [34]       |
| Persimmon leaf flavonoid      | Shi Zhi Ye                                  | ICAM-1↓                   | MCAo 2 h of ischemia followed by 24 h of reperfusion | [35]       |
| Cordyceps sinensis            | Dong Chong Xia Cao                          | ICAM-1, TNF-α, IL-1β, NF-κBp50, iNOS, COX-2↓ | MCAo 2 h of ischemia followed by 22 h of reperfusion | [12]       |
| FuLing-BaiZhu-DangGui          | Fu Ling, Bai Zhu, Dang Gui                  | TNF-α, IL-1β, IL-8, MPO↓, NF-κB↓ | Repetitive BCCAo 10 min of ischemia (repeat 2 times) followed by 24 h of reperfusion | [36]       |
| Borneol                       | Bing Pian                                   | ICAM-1, TNF-α↓            | MCAo 2 h of ischemia followed by 22 h of reperfusion | [37]       |

ICAM-1, intercellular adhesion molecule-1; BCCAo, bilateral common carotid artery occlusion.

extracellular matrix and TJs in endothelial cells and result in vascular and BBB leakage. The BBB disruption facilitates the entry of circulating leukocytes and intravascular fluid into the brain, which cause vasogenic edema and hemorrhagic transformation, leading to the exacerbation of cerebral infarction [6, 9, 76]. TJs, including claudin-5, occludin, and zonula occludens- (ZO-) 1, play a pivotal role in maintaining the structural and functional integrity of the BBB [77]. Previous studies have reported that decreased claudin-5, occludin, and ZO-1 expression is closely related to BBB disruption and ischemic brain edema formation [77, 78]. Thus, MMPs, claudin-5, occludin, and ZO-1 could present the potential targets for pharmacological intervention to stabilize BBB integrity in cerebral ischemic injury and the regulation of their activity may yield therapeutic effects.

4.2. The Effects and Mechanisms of TCMs on Ameliorating Blood-Brain Barrier Disruption in In Vivo Models of Cerebral Ischemia. Posttreatment methylphosphononane- (MO-) A, an active compound isolated from Ophiopogon japonicus (Mai Men Dong), effectively reduces the infarct volume and brain edema and improves neurological deficits 7 d after transient MCAo. The results indicate that MO-A protects against cerebral I/R injury mainly through its property to ameliorate BBB disruption through MMP-9 downregulation, and claudin-3 and claudin-5 upregulation in the ischemic cortex [38]. Tan et al. reported that pretreatment with ligustrazine, an active ingredient of L. wallichii Franchat (Chuan Xiong), effectively preserves BBB integrity by downregulating MMP-9 expression and upregulating claudin-5 and occludin expression in the ischemic area in a rat model of focal cerebral I/R injury [39]. Levo-tetrahydropalmatine (I-THP), a major active ingredient of Rhizoma corydalisis (Yan Hu Suo), protects against cerebral I/R-induced BBB injury at 1.5 h of ischemia and 24 h of reperfusion. The protective effect of I-THP can be partially attributed to MMP-2 and MMP-9 downregulation and claudin-5, occludin, and ZO-1 upregulation in the ischemic area [40].

From these results, we conclude that MMP-9 downregulation and claudin-5, occludin, and ZO-1 upregulation are the potential effects of TCMs on the stabilization of BBB integrity to ameliorate inflammatory responses in the ischemic area during the acute and subacute phases of cerebral I/R injury (Figure 1 and Table 3).

5. TCM-Mediated Regulation of Proinflammatory Mediator Release

5.1. Toll-Like Receptor Stimulation on Microglia/Macrophages during Cerebral Ischemia. In the ischemic core, active microglia are indistinguishable from blood-derived macrophages, and the microglia/macrophages are apparent 3.5–12 h after transient focal cerebral ischemia [79]. Subsequently, active microglia/macrophages are distributed in the entire middle cerebral artery territory at 22–24 h and maintained for up to 1 week after cerebral ischemia [6]. During cerebral ischemic insult, the dying cells release DAMPs, including heat shock proteins (HSPs), β-amyloid, hyaluronan, high mobility group box 1 (HMGB1), heparin sulfate, and ATP, thus stimulating TLRs, which are expressed on microglia/macrophages. Thereafter, the microglia/macrophages transform into M1 and M2 phenotypes upon stimulation and secrete various proteins.
cytokines in response to ischemic injury [73, 80, 81]. TLRs are pivotal components in the innate immune system, and TLRs (mainly TLR2 and TLR4) stimulation in microglia/macrophages and T-lymphocytes also exerts strong regulatory effects on posts ischemic inflammatory responses [2, 73]. During cerebral ischemic injury, TLRs facilitate cytokine and chemokine release and trigger transcription factor activation by activating intercellular signaling pathways. According to recruitment of specific adaptors, TLR signaling can be classified into either myeloid differentiation primary response gene 88-(MyD88-) dependent or independent pathways [81]. The binding of HSPs, such as HSP60 and HSP70, or HMGB1 by activating intercellular signaling pathways. According to recruitment of specific adaptors, TLR signaling can be classified into either myeloid differentiation primary response gene 88-(MyD88-) dependent or independent pathways [81].

5.2. The Effects and Mechanisms of TCMs on Suppressing Toll-Like Receptor Stimulation in In Vivo Models of Cerebral Ischemia. Zhou et al. reported that the anti-infarct effects of puerarin, a major isoflavonoid in Radix puerariae (Ge Gen), can be attributed to the downregulation of TLR4/MyD88/NF-κB/TNF-α signaling in the ischemic region 24 h after transient MCAo [41]. TMP exerts anti-inflammatory effects against neutrophil activation 3 d after permanent MCAo. The beneficial effects of TMP can be partially attributed to HMGB1 and TLR4 downregulation in the ischemic brain at 6 h, peak at 24 h, and decline at 72 h after MCAo [83].

5.3. M1 Microglia/Macrophages Releasing Proinflammatory Mediators during Cerebral Ischemia. The M1 microglia/macrophages produce proinflammatory cytokines, including TNF-α, IL-1β, IL-6, IL-8, IL-12, IL-18, IL-20, and IFN-γ, whereas the M2 microglia/macrophages release anti-inflammatory cytokines, including IL-4, IL-10, IL-13, and TGF-β [1]. TNF-α is a pleiotropic cytokine that possesses both neurotoxic and neuroprotective properties. In the early phase of the inflammatory response, TNF-α binds to TNF receptor 1 (TNFRI) and contributes to the detrimental effects, such as promoting BBB disruption, vasogenic edema, leukocyte infiltration, and endothelial cell apoptosis [6, 84, 85]. Furthermore, TNF-α/TNFRI activates NF-κB signaling, which regulates the expression of cytokines; chemokines; adhesion molecules; and inducible enzymes, namely, iNOS and cyclooxygenase-2 (COX-2), thus exacerbating cerebral ischemic injury [6]. By contrast, in the late phase of posts ischemic inflammation, TNF-α/TNFRI2 facilitates neuroprotection, synaptic plasticity, and tissue repair [6, 86]. TNF-α becomes predominant in the ischemic lesion 24–48 h after MCAo [87]. IL-1, including IL-1α and IL-1β, can bind to IL-1 receptor type 1 (IL-1RI) and majorly contribute to the exacerbation of ischemic brain injury [88]. IL-1β has been clearly implicated in the pathogenesis of cerebral ischemia [6]. Inactive proIL-1β is converted to biologically active IL-1β by an IL-1β-converting enzyme, which belongs to the cysteine protease family. IL-1β is initially upregulated at 1–3 h and peaks at 12–24 h after ischemic injury. The cytotoxic actions of IL-1β include facilitating the activation of microglia, infiltration of leukocytes, and production of other cytokines such as IL-6 [6, 89]. Conversely, IL-1 receptor antagonist, a member of the IL-1 family, binds to IL-1RI and subsequently blocks the detrimental actions of IL-1, exerting neuroprotection in cerebral ischemia process [90]. The role of IL-6 in cerebral ischemia remains controversial. Some studies have reported that IL-6 aggravates cerebral infarction [91, 92], whereas other studies have reported the beneficial effects of IL-6 in preventing damaged neuron from undergoing apoptosis and promoting neuronal survival after cerebral ischemia [93, 94]. Moreover, IL-6 is predominantly expressed in the ischemic area 24–48 h following cerebral I/R injury [87]. IL-8, IL-12, IL-18, and IL-20 play a pivotal role in promoting cerebral ischemic injury [7, 16, 95, 96]. IL-18 is initiated within 24–48 h and peaks at 6 d in the ischemic region after cerebral ischemia [97]. IFN-γ contributes to the exacerbation of cerebral ischemia by increasing ischemia-induced glutamate release [98].

5.4. The Effects and Mechanisms of TCMs on Downregulating Proinflammatory Mediators in In Vivo Models of Cerebral Ischemia. Notoginseng saponins isolated from the root of Panax notoginseng (San Qi) provide beneficial effects

### Table 3: TCMs stabilize blood-brain barrier integrity in the inflammatory cascade in ischemic stroke models.

| TCMs Isolated from the Chinese herb (Chinese name) | Anti-inflammatory actions | Models | References |
|--------------------------------------------------|--------------------------|--------|------------|
| Methyllophiopogonanone A Mai Men Dong | MMP-9↓, claudin-3↑, claudin-5↓ | MCAo 2 h of ischemia followed by 7 d of reperfusion | [38] |
| Ligustrazine Chuan Xiong | MMP-9↓, claudin-5↑, occludin↑ | MCAo 1.5 h of ischemia followed by 22.5 h of reperfusion | [39] |
| Levo-tetrahydropalmatine Yan Hu Suo | MMP-2↓, MMP-9↓, claudin-5↑, occludin↓, ZO-1↑ | MCAo 1.5 h of ischemia followed by 24 h of reperfusion | [40] |

MMP-9, matrix metalloproteinase-9; ZO-1, zonula occludens-1.
against cerebral I/R injury partially through IL-1β mRNA downregulation in the ischemic area after 22 h of reperfusion [43]. Chang et al. reported that pretreatment with puerarin effectively reduces the cerebral infarct size and neurobehavioral deficits 24 h after MCAo. The anti-inflammatory effect of puerarin is, at least partially, because of the inhibition of TNF-α and iNOS expression in the ischemic area [44]. Li et al. explored the effect of osthole, a major active ingredient in *Cnidium monnieri* (L.) Gusson (She Chuang Zi), on acute cerebral I/R injury and reported that pretreatment with osthole markedly reduces the brain infarct volume and ameliorates neurological scores 24 h after MCAo. The neuroprotective effects of osthole are accompanied by the downregulation of proinflammatory mediators, including TNF-α, IL-1β, COX-2, and iNOS, expressed in the ischemic cortex [45]. The caffeic acid ester (Caf) fraction from *Erigeron brevicaudus* (Deng Zhan Hua) significantly reduces the cerebral infarct volume and improves neurobehavioral performance at 1 h of ischemia and 24 h of reperfusion. The inhibition of iNOS, TNF-α, and IL-1β mRNA expression is one of the mechanisms underlying the neuroprotective effects of Caf against cerebral infarction [46]. Pretreatment with arctigenin, an active agent from *Arctium lappa* (Nu Bang Zi), effectively inhibits microglial activation and subsequently downregulates TNF-α and IL-1β expression in the penumbra region 24 h after transient MCAo [47]. Lee et al. reported that schisandrin B isolated from *Fructus schisandrae* (Wu Wei Zi) markedly reduces the cerebral infarct size and neurological deficits 24 h after transient focal cerebral ischemia. The anti-inflammatory effect of schisandrin B involves the inhibition of TNF-α, IL-1β, MMP-2, and MMP-9 expression and suppression of microglial activation in the ischemic area [48]. Posttreatment with asiaticoside, an active compound isolated from *Centella asiatica* (L.) (Ji Xue Cao), attenuates memory deficits by suppressing iNOS, TNF-α, IL-1β, and IL-6 expression in the hippocampus 7 d after transient bilateral common carotid artery occlusion [49]. Chen et al. reported that posttreatment with magnolol, an active ingredient of *Magnolia officinalis* (Hou Pu), ameliorates cerebral infarction partially by dose-dependently inhibiting iNOS, TNF-α, IL-1β, and IL-6 expression in the ischemic area 24 h after transient global ischemia [50]. Posttreatment with danhong, extracted from *Radix salviae miltiorrhizae* (Dan Shen) and *Flos carthami* (Hong Hua), exerts beneficial effects in cerebral I/R injury, at least partially, through dose-dependent IL-1β and TNF-α downregulation in the ischemia area at 1.5 h of ischemia and 14 d of reperfusion [51]. Gastrodin, an active constituent of *Gastrodia elata* Blume (Tian Ma), exerts an initial anti-inflammatory effect by suppressing TNF-α and IL-1β expression in the ischemic hemispheres 6 h after cerebral I/R injury [52].

5.5. The Effects and Mechanisms of TCMs on Regulating Anti-Inflammatory Cytokines in In Vivo Models of Cerebral Ischemia. IL-4, IL-10, IL-13, and TGF-β reduce microglia/macrophages-induced proinflammatory cytokines, such as IL-8 [99]. Moreover, IL-4 promotes long-term recovery after ischemic stroke [100]. IL-10 can inhibit IL-1 and TNF-α expression [55] and prevent the downregulation of the antiapoptotic protein Bcl-2 expressed in ischemic brain lesion [101]. IL-4 mRNA generates as early as 1 h, reaches a peak at 3–24 h, and gradually declines 2 d following ischemic stroke [102]. Pretreatment with danshen, an aqueous extract of the root and rhizome of *Salvia miltiorrhiza* Bunge (Dan Shen), protects against cerebral I/R injury in association with decreased IL-10 and TNF-α mRNA and protein expression in the ischemic area 24 h after transient MCAO [53]. Guizhi fuling capsules, composed of *Cinnamomum cassia* Blume (Gui Zhi), *P. lactiflora* Pall (Shao Yao), *P. suffruticosa* Andrews (Mu Dan Pi), *P. persica* Batsch (Tao Ren), and *Poria* cocos Wolf (Fu Ling), protect against cerebral infarction through TNF-α and IL-1β mRNA and protein downregulation and IL-10 and IL-10 receptor (IL-10R) mRNA and protein upregulation in the ischemia area after 2 h of ischemia and 24 h of reperfusion [54]. Zhang et al. also reported that the Gualou Guizhi decoction composed of *Trichosanthis radix* (Tian Hua Fen), *Ramulus cinnamomi* (Gui Zhi), *P. lactiflora* (Shao Yao), *Glycyrrhiza* (Gan Zao), *Zingiber officinale* Roscoe (Sheng Jiang), and *Fructus jujubae* (Da Zao) exerts neuroprotection against cerebral I/R injury through IL-1, TNF-α, and NF-κB downregulation and IL-10 upregulation in the ischemic area in the subacute phase (7 d) after transient MCAO [55].

5.6. The Effects and Mechanisms of TCMs on Downregulating Proinflammatory Enzymes in In Vivo Models of Cerebral Ischemia. COX-2 and 5-lipoxygenase (5-LO) are rate-limiting enzymes that convert arachidonic acid to prostaglandins and leukotrienes [57]. In the delayed phase of cerebral ischemia, microglia/macrophages produce 5-LO, which converts arachidonic acid to leukotrienes. Leukotrienes are potent inflammatory mediators that trigger chemotaxis of leukocytes and BBB damage and subsequently cause vasogenic edema, thus exacerbating cerebral ischemia [103]. COX-2 and 5-LO expression is markedly enhanced in the ischemic cortex 24 h after cerebral I/R injury [57]. Guo et al. explored the anti-infarct effect of paeoniflorin (PF), the principle component of *P. radix* (Shao Yao), in the subacute phase of cerebral I/R injury and reported that PF protects against cerebral infarction mainly through TNF-α, IL-1β, iNOS, COX-2, and 5-LO downregulation in the ischemic area 14 d after reperfusion [56]. Chen et al. reported that pretreatment with PF effectively ameliorates the cerebral infarct volume and neurological deficits 24 h after reperfusion in a model of pharmacological preconditioning. The neuroprotective effects of PF against cerebral I/R injury are partially related to the inhibition of COX-2, 5-LO, and iNOS expression in the ischemic lesion [57].

According to the aforementioned studies, proinflammatory mediators, such as TLR4, TNF-α, IL-1β, IL-6, IL-18, COX-2, and 5-LO, are predominately expressed in the ischemic area 24 h after MCAo. TCMs effectively ameliorate cerebral I/R injury by downregulating TLR4, TNF-α, IL-1β, IL-6, iNOS, COX-2, and 5-LO expression and upregulating IL-10 expression in the ischemic area during the acute and subacute phases of cerebral ischemia (Figure 1 and Table 4).
Table 4: TCMs regulate the cytokine release in the inflammatory cascade in ischemic stroke models.

| TCMs                   | Isolated from the Chinese herb (Chinese name) | Anti-inflammatory actions | Models                                      | References |
|------------------------|---------------------------------------------|---------------------------|---------------------------------------------|------------|
| Puerarin               | Ge Gen                                      | TLR4↓, MyD88↓, NF-κBp65↓, TNF-α↓ | MCAo 1.5 h of ischemia followed by 24 h of reperfusion | [41]       |
| Tetramethylpyrazine    | Chuan Xiong                                 | TLR4↓, HMGB1↓, Nrf2↑, HO-1↑ | Permanent MCAo 3 d of ischemia              | [42]       |
| Notoginseng            | San Qi                                      | IL-1β↓                    | MCAo                                        | [43]       |
| Puerarin               | Ge Gen                                      | TNF-α↓, iNOS↓             | BCCAo 1 h of ischemia followed by 24 h of reperfusion | [44]       |
| Caffeic acid ester     | She Chuang Zi                              | TNF-α↓, IL-1β↓, COX-2↓    | MCAo 1 h of ischemia followed by 24 h of reperfusion | [45]       |
| Arctigenin             | Nu Bang Zi                                  | TNF-α↓, IL-1β↓, OX-42↓    | MCAo 2 h of ischemia followed by 24 h of reperfusion | [46]       |
| Schisandrin B          | Wu Wei Zi                                   | TNF-α↓, COX-2↓, MMP-2↓, MMP-9↓, OX-42↓ | MCAo 2 h of ischemia followed by 24 h of reperfusion | [47]       |
| Asiaticoside           | Ji Xue Cao                                  | iNOS↓, TNF-α↓, IL-1β↓, IL-6↓ | BCCAo 10 min of ischemia (repeat 2 times) followed by 7 d of reperfusion | [48]       |
| Magnolol               | Hou Pu                                      | iNOS↓, TNF-α↓, IL-1β↓, IL-6↓, NF-κBp65↓ | BCCAo 1.5 h of ischemia followed by 24 h of reperfusion | [49]       |
| Danhong injection      | Dan Shen and Hong Hua                       | TNF-α↓, IL-1β↓            | MCAo 1.5 h of ischemia followed by 14 d of reperfusion | [50]       |
| Gastrodin              | Tian Ma                                     | TNF-α↓, IL-1β↓            | MCAo 1 h of ischemia followed by 6 h of reperfusion | [51]       |
| Danshen                | Dan Shen                                    | IL-10 mRNA↓, TNF-α mRNA↓, IL-10↑, TNF-α↓ | MCAo 1 h of ischemia followed by 24 h of reperfusion | [52]       |
| Guizhi fuling capsules | Gui Zhi, Shao Yao, Mu Dan, Tao Ren, Fu Ling | TNF-α mRNA↓, IL-1β mRNA↓, TNF-α↓, IL-10R mRNA↓, IL-10R↑, IL-10R↑ | MCAo 2 h of ischemia followed by 24 h of reperfusion | [53]       |
| Gualou Guizhi decoction| Tian Hua Fen, Gui Zhi, Shao Yao, Gan Zao, Sheng Jiang, Da Zao | TNF-α↓, IL-1↓, NF-κBp65↓, IL-10↑ | MCAo 2 h of ischemia followed by 7 d of reperfusion | [54]       |
| Paeoniflorin           | Shao Yao                                    | TNF-α↓, IL-1β↓, iNOS↓, COX-2↓, 5-LO↓ | MCAo 1.5 h of ischemia followed by 14 d of reperfusion | [55]       |
| Paeoniflorin           | Shao Yao                                    | COX-2↓, 5-LO↓, iNOS↓      | MCAo 1.5 h of ischemia followed by 24 h of reperfusion | [56]       |

TLR4, toll-like receptor 4; MyD88, myeloid differentiation primary response gene 88; HMGB1, high mobility group box 1; Nrf2, nuclear factor-erythroid 2-related factor 2; HO-1, heme oxygenase-1; iNOS, inducible nitric oxidase synthase; 5-LO, 5-lipoxygenase.
6. TCM-Mediated Regulation of Transcription Factor Activation

6.1. NF-κB Activation during Cerebral Ischemia. NF-κB is a classic transcription factor and plays a crucial role in the regulation of hundreds of genes involved in cell survival and death [104]. Thus, NF-κB can be activated via several intracellular signaling pathways associated with host defense, inflammation, and apoptosis [58]. In the brain, NF-κB regulates the expression of different sets of genes, such as antiapoptotic, proapoptotic, and proinflammatory genes, thereby playing a dual role in neuronal survival and death [63]. The NF-κB family includes five members, namely, p65 (RelA), RelB, c-Rel, p50/p105 (NF-κB1), and p52/p100 (NF-κB2), which form various homo- and heterodimeric complexes [2]. The most common form of NF-κB is the p65/p50 heterodimer [6]. Under an unstimulated condition, the inhibitor of NF-κB proteins (IκBs), including mainly IκBα, IκBβ, and IκBε, retain inactive NF-κB dimmers in the cytosol, whereas in response to various extracellular stimuli, including infection, proinflammatory cytokines, and antigen receptor engagement, the activated IκB kinase complexes phosphorylate IκB proteins, resulting in their ubiquitination and proteasomal degradation and consequently inducing the release of NF-κB for nuclear translocation and the activation of target gene transcription [6, 105]. During cerebral ischemia, the activated NF-κB dimers are subsequently translocated into the nucleus where they selectively bind to specific DNA sequences called κB sites; promoter domains present a large number of proinflammatory genes and subsequently cause TNF-α, IL-1β, IL-6, ICAM-1, PGE2, COX-2, and iNOS translation [6, 105, 106]. NF-κB activators include some proinflammatory cytokines, such as TNF-α and IL-1β, whose genes are regulated by NF-κB itself, inducing a positive feedback loop and resulting in the amplification of the inflammatory response and exacerbation of cerebral ischemic insults [107]. Previous studies have indicated that NF-κB activation occurs as early as 1h, reaches a peak at 6 h, and sustains for at least 72 h in the cerebral ischemic area in rats [62, 107].

6.2. The Effects and Mechanisms of TCMs on Downregulating NF-κB Activation in In Vivo Models of Cerebral Ischemia. Wogonin, a flavonoid derived from Scutellaria baicalensis Georgi (Huang Qin), exerts neuroprotective effects by inhibiting the inflammatory activation of microglia in an in vitro cell culture model. The anti-inflammatory effects of wogonin are partially attributed to the downregulation of NF-κB-mediated iNOS and TNF-α expression in the ischemic hippocampal CA1 area in transient global cerebral ischemia in rats [58]. Tanshinone IIA (Ts IIA) and IIB, the key compounds of S. miltiorrhiza Bunge, effectively reduce the cerebral infarct volume and improve the neurological function 24 h after transient MCAo [59]. Dong et al. further reported that pretreatment with Ts IIA protects against cerebral infarction partially associated with the reduction of ROS-mediated NF-κB activation, leading to the inhibition of iNOS expression in the ischemic area 24 h after permanent MCAo [60]. Silymarin, a bioactive component isolated from Silybum marianum (Shui Fei Ji), provides neuroprotection against cerebral I/R injury by inhibiting oxidative and nitrosative stress in the ischemic area 24 h after cerebral ischemia. The antioxidative and antinitrosative effects of silymarin are partially attributed to the reduction of NF-κB-mediated iNOS, COX-2, ICAM-1, TNF-α, and IL-1β expression in the injured tissues [61]. In addition, Guan et al. reported that ruscogenin, a major effective compound isolated from O. japonicus Ker-Gawl, ameliorates cerebral I/R injury through the downregulation of NF-κB target genes, including ICAM-1, iNOS, COX-2, TNF-α, and IL-1β, in the ischemic area 24 h after reperfusion [62]. Hydroxyasflor yellow A, a major active component of C. tinctorius L. (Hong Hua), reduces cerebral infarction by suppressing cytosolic NF-κBp65 translocation to the nucleus and subsequently downregulates NF-κB-mediated TNF-α, IL-1β, and IL-6 expression in the ischemic area 24 h after permanent MCAo [63]. Chern et al. reported that 2-methoxyxystepandrene (2-Ms), a major active component of Polygonum cuspidatum (Hu Zhang), attenuates the brain infarct size and improves the neurological function, at least partially, by preventing IκBα degradation and a reducing NF-κB-mediated iNOS and COX-2 expression in the peri-infarct cortex 24 h after transient MCAo. The anti-inflammatory effects of 2-Ms can further contribute toward the preserving BBB integrity [64]. Previous studies have indicated that p38 mitogen-activated protein kinase (MAPK), one of the MAPK family members, upregulates NF-κB expression (p38 MAPK/NF-κB signaling) and subsequently causes the transcription of genes encoding proinflammatory cytokines, resulting in the exacerbation of cerebral infarction in the acute phase of transient MCAo [108–110]. In addition, activated p38 MAPK occurs in the ischemic area as early as 2 h and reaches a peak 24–48 h after reperfusion [110]. Piperlongumine from Piper longum (Bi Bo) alkaloids protects against cerebral ischemic injury by inhibiting the activation of p38 MAPK/NF-κB signaling cascade in the ischemic region 24 h after permanent MCAo [65].

6.3. The Effects and Mechanisms of TCMs on Upregulating Peroxisome Proliferator-Activated Receptor Activation in In Vivo Models of Cerebral Ischemia. Peroxisome proliferator-activated receptors (PPARs) include PPARα, PPARγ, and PPARδ/β isoforms, which are members of the nuclear receptor superfamly and represent ligand-activated transcription factors. PPARγ is predominantly expressed in the central nervous system and binds to peroxisome proliferator response elements to regulate its target gene expression [66]. During cerebral ischemia, PPARγ is detected in the peri-infarct area as early as 4 h and is sustained for at least 14 d after ischemia [111]. PPARγ exerts neuroprotective effects against inflammatory mediators to initiate responses by inhibiting the activation of NF-κB signaling in the ischemic area after focal cerebral ischemia [66, 111]. Liu et al. reported that pretreatment with curcumin, a natural polyphenolic component of curcuma longa (Jiang Huang), markedly reduces the cerebral infarct volume by activating PPARγ signaling in the ischemic cortex 24 h after reperfusion. The effects of curcumin on the regulation of PPARγ signaling further contribute to the downregulation of NF-κBp65-mediated TNF-α, IL-1β,
genes encoding antioxidant and anti-inflammatory proteins protect the nervous system from oxidative stress and inflammatory response. The protective transcription factor, Nrf2, is activated by upstream signaling pathways that involve transcription factors such as HO-1. In an in vivo model of cerebral ischemia, Nrf2/HO-1 signaling pathway attenuates inflammatory responses in cerebral ischemia [71]. During transient focal cerebral ischemia, Nrf2 and HO-1 occur in the ischemic cortex as early as 6 h, up to a maximum of 48 h and decline 72 h after cerebral I/R [119]. The activation of c-Jun N-terminal kinase (JNK), one of the MAPK family members, signaling plays a central role in ischemia-induced neuroinflammation. When stimulated, activated JNK translocates into the nucleus and phosphorylates c-Jun, the major component of activating protein (AP-1), which comprises c-Jun and c-Fos proteins, leading to the expression of target genes encoding proinflammatory mediators. JNK/AP-1 signaling amplifies the inflammatory response during cerebral ischemia [71]. JNK/c-Jun/AP-1 signaling factors are predominantly expressed in the ischemic area 2 h after cerebral ischemia [120]. Kao et al. reported that TMP effectively reduces cerebral infarction by inhibiting microglia/macrophages activation in the ischemic cortex 72 h after permanent MCAo. The anti-inflammatory effects of TMP can be further attributed to the upregulation of Nrf2/HO-1 signaling and downregulation of JNK/c-Jun/AP-1 signaling in the ischemic cortex [71].

According to the aforementioned studies, NF-κB and JNK/AP-1 signaling induced in the ischemic brain may amplify inflammatory responses, whereas PPARs and Nrf2/HO-1 signaling are considered to prevent postischemic inflammation and yield potent effects against cerebral ischemic injury. JAK/STAT signaling plays a dual role in the regulation of proinflammatory mediators depending on the experimental models of brain ischemia. TCMs protect against cerebral ischemic injury by inhibiting deleterious transcription factors (NF-κB, JAK3/STAT, and JNK/AP-1), activating neuroprotective transcription factors (PPARs and Nrf2/HO-1) and consequently regulating the expression of transcription factor-mediated proinflammatory genes (TNF-α, IL-β, IL-6, IL-8, iNOS, COX-2, MMP-9, and ICAM-1) in the ischemic area in the early stage (24–72 h) after cerebral ischemia (Figure 2 and Table 5).

7. Conclusions
After the onset of cerebral ischemia, resident microglia are rapidly activated (within a few minutes) and subsequently produce large amounts of cytokines, chemokines, and ROS, thus causing the initial ischemic injury. TCMs can exert neuroprotective effects against the initial ischemic injury by rapidly downregulating ischemia-induced microglial activation and microglia-mediated proinflammatory cytokine production in the ischemic region. The microglial production of proinflammatory mediators subsequently increase adhesion molecule expression, facilitate leukocyte-endothelial cell interactions, and activated leukocytes that penetrate the endothelial cell barrier into the brain parenchyma (as early as 4 h after the ischemic onset). The infiltrating leukocytes further release inflammatory mediators in the...
ischemic lesion, thus exacerbating ischemic injury. TCMs can effectively attenuate leukocyte infiltration by inhibiting ICAM-1 and activated leukocyte-induced cytokine expression in the ischemic region in the early phase of cerebral ischemia. Meanwhile, infiltrated leukocytes and activated microglia secrete MMPs, which cause the disruption of BBB integrity, worsening cerebral infarction. TCMs effectively inhibit MMPs expression and stabilize BBB integrity to ameliorate cerebral infarction. Increased TLRs stimulation in microglia/macrophages (activated microglia and recruited leukocytes) by the activation of intercellular signaling pathways robustly secretes various proinflammatory mediators (cytokines and enzymes) in the ischemic region 6–24 h after ischemia. TCMs can timely rescue the injured neurons by downregulating proinflammatory receptors (TLRs), cytokines, and enzymes and upregulating anti-inflammatory cytokine expression in the ischemic lesion. The proinflammatory transcription factors are subsequently activated by the secreted cytokines, whose genes are regulated by these transcription factors themselves, thus inducing a positive feedback loop, in which the inflammatory response is amplified and secondary brain injury is exacerbated 24–72 h after cerebral ischemia. TCMs protect against inflammatory response-induced secondary brain injury by inhibiting deleterious transcription factors, activating neuroprotective transcription factors, and consequently regulating the expression of transcription factor-mediated proinflammatory genes in the ischemic area. Therefore, TCMs provide promising anti-inflammatory therapeutic strategies in the acute phase of cerebral ischemia. However, further studies are needed to elucidate the precise mechanisms of TCMs against inflammatory responses in the ischemic cascade after stroke.

**Competing Interests**

The authors have declared that no competing interests exist.
Table 5: TCMs regulate transcription factors in the inflammatory cascade in ischemic stroke models.

| TCMs                         | Isolated from the Chinese herb (Chinese name) | Anti-inflammatory actions | Models          | References |
|------------------------------|-----------------------------------------------|---------------------------|-----------------|------------|
| Wogonin                      | Huang Qin                                    | NF-κBp65↓, iNOS↓, TNF-α↓ | 4-VO            | [58]       |
|                              |                                               |                           | 10 min of ischemia followed by 7 d of reperfusion |            |
| Tanshinone IIA               | Dan Shen                                      | NF-κBp65↓, iNOS↓         | MCAo            | [59, 60]   |
|                              |                                               |                           | 2 h of ischemia followed by 24 h of reperfusion |            |
| Silymarin                    | Shui Fei Ji                                   | NF-κBp65↓, iNOS↓, COX-2↓, ICAM-1↓, IL-1β↓, IL-6↓, MPO↓ | MCAo          | [61]       |
|                              |                                               |                           | 1 h of ischemia followed by 24 h of reperfusion |            |
| Ruscogenin                   | Mai Men Dong                                  | NF-κBp65↓, TNF-α↓, IL-1β↓ | MCAo            | [62]       |
|                              |                                               |                           | 1 h of ischemia followed by 24 h of reperfusion |            |
| Hydroxysafflor yellow A      | Hong Hua                                      | NF-κBp65↓, iNOS↓, COX-2↓, IL-1β↓ | MCAo          | [63]       |
|                              |                                               |                           | 40 min of ischemia followed by 24 h of reperfusion |            |
| 2-Methoxystypandrone         | Hu Zhang                                      | NF-κBp65↓, iNOS↓, COX-2↓, IL-1β↓ | MCAo          | [64]       |
|                              |                                               |                           | 24 h of ischemia |            |
| Piperlonguminine             | Bi Bo                                         | NF-κBp65↓, p-p38 MAPK↓   | Permanent MCAo  | [65]       |
|                              |                                               |                           | 24 h of ischemia |            |
| Curcumin                     | Jiang Huang                                   | PPARγ↑, NF-κBp65↓, iNOS↓, COX-2↓, IL-1β↓ | MCAo          | [66]       |
|                              |                                               |                           | 2 h of ischemia followed by 24 h of reperfusion |            |
| Icariin                      | Yin Yang Hu                                   | PPARα↑, PPARγ↑, NF-κBp65↓, IL-1β↓ | MCAo          | [67]       |
|                              |                                               |                           | 2 h of ischemia followed by 24 h of reperfusion |            |
| Kaempferol-3-O-rutinoside    | Hong Hua                                      | STAT3↓, NF-κBp65↓, TNF-α↓, IL-1β↓, iNOS↓, MMP-9↓, ICAM-1↓ | MCAo          | [68]       |
| Kaempferol-3-O-glucoside     |                                               |                           | 2 h of ischemia followed by 24 h of reperfusion |            |
| Astragaloside IV, ginsenoside| Rgl, ginsenoside Rbl, notoginsenoside RI      | JAK1↓, STAT1↓, NF-κBp65↓, p-IκBα↓, TNF-α mRNA↓, IL-1β mRNA↓, ICAM-1 mRNA↓ | BCCAo        | [69]       |
|                              | Huang Qi and San Qi                           |                           | 20 min of ischemia followed by 24 h of reperfusion |            |
| Curcumin                     | Jiang Huang                                   | JAK2↑, STAT3↑, IL-1β↑, IL-8↓ | MCAo          | [70]       |
|                              |                                               |                           | 1.5 h of ischemia followed by 24 h of reperfusion |            |
| Tetramethylpyrazine          | Chuan Xiong                                   | Nrf2↑, HO-1↑, MPO↓, p-c-Jun↑, p-JNK↓, AP-1↓ | Permanent MCAo | [71]       |
|                              |                                               |                           | 72 h of ischemia |            |

4-VO, 4-vessel occlusion; IκBα, inhibitor of NF-κB protein α; PPARγ, peroxisome proliferator-activated receptor γ; STAT3, signal transducer and activator of transcription 3; JAK1, Janus kinase 1; AP-1, activating protein-1.

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References

[1] A. Siniscalchi, L. Gallelli, G. Malferrari et al., “Cerebral stroke injury: the role of cytokines and brain inflammation,” Journal of Basic and Clinical Physiology and Pharmacology, vol. 25, no. 2, pp. 131–137, 2014.

[2] R. Jin, L. Liu, S. Zhang, A. Nanda, and G. Li, “Role of inflammation and its mediators in acute ischemic stroke,” Journal of Cardiovascular Translational Research, vol. 6, no. 5, pp. 834–851, 2013.

[3] R. Jin, G. Yang, and G. Li, “Inflammatory mechanisms in ischemic stroke: role of inflammatory cells,” Journal of Leukocyte Biology, vol. 87, no. 5, pp. 779–789, 2010.

[4] R.-D. Cheng, J.-J. Ren, Y.-Y. Zhang, and X.-M. Ye, “P2X4 receptors expressed on microglial cells in post-ischemic inflammation of brain ischemic injury,” Neurochemistry International, vol. 67, no. 1, pp. 9–13, 2014.

[5] M. K. Tobin, J. A. Bonds, R. D. Minshall, D. A. Pelligrino, F. D. Testai, and O. Lazarov, “Neurogenesis and inflammation after ischemic stroke: what is known and where we go from here,”
its mechanism,” African Journal of Traditional, Complementary, and Alternative Medicines, vol. 11, no. 1, pp. 161–164, 2014.

[38] M. Lin, W. Sun, W. Gong, Z. Zhou, Y. Ding, and Q. Hou, “Methylphenidoponagonone a protects against cerebral ischemia/reperfusion injury and attenuates blood-brain barrier disruption,” PLoS ONE, vol. 10, no. 4, Article ID e0124558, 2015.

[39] F. Tan, W. Fu, N. Cheng, D. I. Meng, and Y. Gu, “Ligustriazine reduces blood-brain barrier permeability in a rat model of focal cerebral ischemia and reperfusion,” Experimental and Therapeutic Medicine, vol. 9, no. 5, pp. 1757–1762, 2015.

[40] X.-W. Mao, C.-S. Pan, P. Huang et al., “Levotetrahydropalmatine attenuates mouse blood-brain barrier injury induced by focal cerebral ischemia and reperfusion: involvement of Src kinase,” Scientific Reports, vol. 5, Article ID 11155, 2015.

[41] F. Zhou, L. Wang, P. P. Liu et al., “Puerarin protects brain tissue against cerebral ischemia/reperfusion injury by inhibiting the inflammatory response,” Neural Regeneration Research, vol. 9, no. 23, pp. 2074–2080, 2014.

[42] C.-Y. Chang, T.-K. Kao, W.-Y. Chen et al., “Tetramethylpyrazine inhibits neutrophil activation following permanent cerebral ischemia in rats,” Biochemical and Biophysical Research Communications, vol. 463, no. 3, pp. 421–427, 2015.

[43] Y. H. Tang, S. P. Zhang, Y. Liang, and C. Q. Deng, “Effects of Panax notoginseng saponins on mRNA expressions of interleukin-1 β, its correlative factors and cysteinyl-aspartate specific protease after cerebral ischemia-reperfusion in rats,” Zhong Xi Yi Jie He Xue Bao, vol. 5, no. 3, pp. 328–332, 2007.

[44] Y. Chang, C.-Y. Hsieh, Z.-A. Peng et al., “Neuroprotective mechanisms of puerarin in middle cerebral artery occlusion-induced brain infarction in rats,” Journal of Biomedical Science, vol. 16, article 9, 2009.

[45] F. Li, Q. Gong, L. Wang, and J. Shi, “Osthole attenuates focal inflammatory reaction following permanent middle cerebral artery occlusion in rats,” Biological and Pharmaceutical Bulletin, vol. 35, no. 10, pp. 1686–1690, 2012.

[46] S.-X. Wang, H. Guo, L.-M. Hu et al., “Caffeic acid ester fraction from erigeron breviscapus inhibits microglial activation and provides neuroprotection,” Chinese Journal of Integrative Medicine, vol. 18, no. 6, pp. 437–444, 2012.

[47] T. Fan, W. L. Jiang, J. Zhu, and Y. Feng Zhang, “Arctigenin protects focal cerebral ischemia-reperfusion rats through inhibiting neuroinflammation,” Biological and Pharmaceutical Bulletin, vol. 35, no. 11, pp. 2004–2009, 2012.

[48] T. H. Lee, C. H. Jung, and D.-H. Lee, “Neuroprotective effects of Schisandrín B against transient focal cerebral ischemia in Sprague-Dawley rats,” Food and Chemical Toxicology, vol. 50, no. 12, pp. 4239–4245, 2012.

[49] S. Chen, Z.-J. Yin, C. Jiang et al., “Asiaticoside attenuates memory impairment induced by transient cerebral ischemia-reperfusion in mice through anti-inflammatory mechanism,” Pharmacology Biochemistry and Behavior, vol. 122, pp. 7–15, 2014.

[50] J.-H. Chen, H.-C. Kuo, K.-F. Lee, and T.-H. Tsai, “Magnolol protects neurons against ischemia injury via the downregulation of p38/MAPK, CHOP and nitrotyrosine,” Toxicology and Applied Pharmacology, vol. 279, no. 3, pp. 294–302, 2014.

[51] Z. Wang, F. Song, J. Li et al., “PET demonstrates functional recovery after treatment by danhong injection in a rat model of cerebral ischemic-reperfusion injury,” Evidence-Based Complementary and Alternative Medicine, vol. 2014, Article ID 430757, 9 pages, 2014.

[52] Z. Peng, S. Wang, G. Chen et al., “Gastrodin alleviates cerebral ischemic damage in mice by improving anti-oxidant and anti-inflammation activities and inhibiting apoptosis pathway,” Neurochemical Research, vol. 40, no. 4, pp. 661–673, 2015.

[53] X.-Y. Liang, H.-N. Li, X.-Y. Yang, W.-Y. Zhou, J.-G. Niu, and B.-D. Chen, “Effect of Danshen aqueous extract on serum hs-CRP, IL-8, IL-10, TNF-α levels, and IL-10 mRNA expression levels, cerebral TGF-β1 positive expression level and its neuroprotective mechanisms in CIR rats,” Molecular Biology Reports, vol. 40, no. 4, pp. 3419–3427, 2013.

[54] T.-J. Li, Y. Qiu, J.-Q. Mao, P.-Y. Yang, Y.-C. Rui, and W.-S. Chen, “Protective effects of Guizhi-Fuling-capsules on rat brain ischemia/reperfusion injury,” Journal of Pharmaco logical Sciences, vol. 105, no. 1, pp. 34–40, 2007.

[55] Z. Zhang, S. Zhang, H. Li et al., “Ameliorative effects of Gualou Guizhi decoction on inflammation in focal cerebral ischemic-reperfusion injury,” Molecular Medicine Reports, vol. 12, no. 1, pp. 988–994, 2015.

[56] R.-B. Guo, G.-F. Wang, A.-P. Zhao, J. Gu, X.-L. Sun, and G. Hu, “Paoniflorin protects against ischemia-induced brain damages in rats via inhibiting MAPKs/NF-κB-mediated inflammatory responses,” PLoS ONE, vol. 7, no. 11, Article ID e49701, 2012.

[57] D.-M. Chen, L. Xiao, X. Cai, R. Zeng, and X.-Z. Zhu, “Involvement of multitargets in paoniflorin-induced preconditioning,” Journal of Pharmacology and Experimental Therapeutics, vol. 319, no. 1, pp. 165–180, 2006.

[58] H. Lee, Y. O. Kim, H. Kim et al., “Flavonoid wogonin from medicinal herb is neuroprotective by inhibiting inflammatory activation of microglia,” The FASEB Journal, vol. 17, no. 13, pp. 1943–1944, 2003.

[59] B. Y. H. Lam, A. C. Y. Lo, X. Sun, H. W. Luo, S. K. Chung, and N. J. Sucher, “Neuroprotective effects of tanshinones in transient focal cerebral ischemia in mice,” Phytomedicine, vol. 10, no. 4, pp. 286–291, 2003.

[60] K. Dong, W. Xu, J. Yang, H. Qiao, and L. Wu, “Neuroprotective effects of Tanshinone IIA on permanent focal cerebral ischemia in mice,” Phytotherapy Research, vol. 23, no. 5, pp. 608–613, 2009.

[61] Y.-C. Hou, K.-T. Liou, C.-M. Chern et al., “Preventive effect of silymarin in cerebral ischemia-reperfusion-induced brain injury in rats possibly through impairing NF-κB and STAT-1 activation,” Phytomedicine, vol. 17, no. 12, pp. 963–973, 2010.

[62] T. Guan, Q. Liu, Y. Qian et al., “Ruscogenin reduces cerebral ischemic injury via NF-κB-mediated inflammatory pathway in the mouse model of experimental stroke,” European Journal of Pharmacology, vol. 714, no. 1–3, pp. 303–311, 2013.

[63] Y. Liu, Z. Lian, H. Zhu et al., “A systematic, integrated study on the neuroprotective effects of hydroxyxafflor yellow a revealed by 1H NMR-based metabolomics and the NF-κB pathway,” Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 147362, 14 pages, 2013.

[64] C.-M. Chern, Y.-H. Wang, K.-T. Liou, Y.-C. Hou, C.-C. Chen, and Y.-C. Shen, “2-Methoxyxystipandrene ameliorates brain function through preserving BBB integrity and promoting neurogenesis in mice with acute ischemic stroke,” Biochemical Pharmacology, vol. 87, no. 3, pp. 502–514, 2014.

[65] T. Yang, S. Sun, T. Wang et al., “Piperlonguminine is neuroprotective in experimental rat stroke,” International Immunopharmacology, vol. 23, no. 2, pp. 447–451, 2014.

[66] Z.-J. Liu, W. Liu, L. Liu, C. Xiao, Y. Wang, and J.-S. Jiao, “Curcumin protects neuron against cerebral ischemia-induced inflammation through improving PPAR-gamma function,” Evidence-Based Complementary and Alternative Medicine
Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 470975, 10 pages, 2013.

[67] D. Xiong, Y. Deng, B. Huang et al., "Icariin attenuates cerebral ischemia-reperfusion injury through inhibition of inflammatory response mediated by NF-κB, PPARα and PPARγ in rats," *International Immunopharmacology*, vol. 30, pp. 157–162, 2016.

[68] L. Yu, C. Chen, L.-F. Wang et al., "Neuroprotective effect of kaempferol glycosides against brain injury and neuroinflammation by inhibiting the activation of NF-κB and STAT3 in transient focal stroke," *PLoS ONE*, vol. 8, no. 2, Article ID e55839, 2013.

[69] X.-P. Huang, H. Ding, J.-D. Lu, Y.-H. Tang, B.-X. Deng, and C.-Q. Deng, "Effects of the combination of the main active components of Astragalus and Panax notoginseng on inflammation and apoptosis of nerve cell after cerebral ischemia-reperfusion," *The American Journal of Chinese Medicine*, vol. 43, no. 7, pp. 1419–1438, 2015.

[70] L. Li, H. Li, and M. Li, "Curcumin protects against cerebral ischemia-reperfusion injury by activating JAK2/STAT3 signaling pathway in rats," *International Journal of Clinical and Experimental Medicine*, vol. 8, no. 9, pp. 14985–14991, 2015.

[71] T.-K. Kao, C.-Y. Chang, Y.-C. Ou et al., "Tetramethylpyrazine reduces cellular inflammatory response following permanent focal cerebral ischemia in rats," *Experimental Neurology*, vol. 247, pp. 188–201, 2013.

[72] H. Könecke and I. Bechmann, "The role of microglia and matrix metalloproteinases in neuroinflammation and gliomas," *Clinical and Developmental Immunology*, vol. 2013, Article ID 914104, 15 pages, 2013.

[73] T. Shichita, R. Sakaguchi, M. Suzuki, and A. Yoshimura, "Post-ischemic inflammation in the brain," *Frontiers in Immunology*, vol. 3, article 132, 2012.

[74] X. Dong, Y.-N. Song, W.-G. Liu, and X.-L. Guo, "MMP-9, a potential target for cerebral ischemic treatement," *Current Neuropharmacology*, vol. 7, no. 4, pp. 269–275, 2009.

[75] A. M. Romanic, R. F. White, A. J. Arleth, E. H. Ohlstein, and F. C. Barone, "Matrix metalloproteinase expression increases after focal ischemia in rats: inhibition of matrix metalloproteinase-9 reduces infarct size," *Stroke*, vol. 29, no. 5, pp. 1020–1030, 1998.

[76] S. E. Lakhani, A. Kirchgeessner, D. Tepper, and A. Leonard, "Matrix metalloproteinases and blood-brain barrier disruption in acute ischemic stroke," *Frontiers in Neurology*, vol. 4, article 32, 2013.

[77] Y.-M. Zhang, H. Xu, H. Sun, S.-H. Chen, and F.-M. Wang, "Electroacupuncture treatment improves neurological function associated with regulation of tight junction proteins in rats with cerebral ischemia reperfusion injury," *Evidence-Based Complementary and Alternative Medicine*, vol. 2014, Article ID 989340, 10 pages, 2014.

[78] H. Jiao, Z. Wang, Y. Liu, P. Wang, and Y. Xue, "Specific role of tight junction proteins claudin-5, occludin, and ZO-1 of the blood-brain barrier in a focal cerebral ischemic insult," *Journal of Molecular Neuroscience*, vol. 44, no. 2, pp. 130–139, 2011.

[79] R. A. Taylor and L. H. Sansing, "Microglial responses after ischemic stroke and intracerebral hemorrhage," *Clinical and Developmental Immunology*, vol. 2013, Article ID 746068, 10 pages, 2013.

[80] R. Barakat and Z. Redzic, "Differential cytokine expression by brain microglia/macrophages in primary culture after oxygen glucose deprivation and their protective effects on astrocytes during anoxia," *Fluids and Barriers of the CNS*, vol. 12, article 6, 2015.

[81] Y.-C. Wang, S. Lin, and Q.-W. Yang, "Toll-like receptors in cerebral ischemic inflammatory injury," *Journal of Neuroinflammation*, vol. 8, article 134, 2011.

[82] Y. Wang, P. Ge, and Y. Zhu, "TLR2 and TLR4 in the brain injury caused by cerebral ischemia and reperfusion," *Mediators of Inflammation*, vol. 2013, Article ID 124614, 8 pages, 2013.

[83] X.-K. Tu, W.-Z. Yang, S.-S. Shi et al., "Spatio-temporal distribution of inflammatory reaction and expression of TLR2/4 signaling pathway in rat brain following permanent focal cerebral ischemia," *Neurochemical Research*, vol. 35, no. 8, pp. 1147–1155, 2010.

[84] A. Maddahi, L. S. Kruse, Q.-W. Chen, and L. Edvinsson, "The role of tumor necrosis factor-α and TNF-α receptors in cerebral arteries following cerebral ischemia in rat," *Journal of Neuroinflammation*, vol. 8, article 107, 2011.

[85] Q. Hua, X.-L. Zhu, P.-T. Li et al., "The inhibitory effects of chloral acid and hyodeoxycholalic acid on the expression of TNFα and IL-1β after cerebral ischemia in rats," *Archives of Pharmacal Research*, vol. 32, no. 1, pp. 65–73, 2009.

[86] W. Chadwick, T. Magnus, B. Martin, A. Keselman, M. P. Mattson, and S. Maudsley, "Targeting TNF-α receptors for neurotherapeutics," *Trends in Neurosciences*, vol. 31, no. 10, pp. 504–511, 2008.

[87] A. Maddahi and L. Edvinsson, "Cerebral ischemia induces microvascular pro-inflammatory cytokine expression via the MEK/ERK pathway," *Journal of Neuroinflammation*, vol. 7, article 14, 2010.

[88] H. Boutin, R. A. LeFeuvre, R. Horai, M. Asano, Y. Iwakura, and N. J. Rothwell, "Role of IL-1α and IL-1β in ischemic brain damage," *The Journal of Neuroscience*, vol. 21, no. 15, pp. 5528–5534, 2001.

[89] G. P. Schielke, G.-Y. Yang, B. D. Shivers, and A. L. Betz, "Reduced ischemic brain injury in interleukin-1β converting enzyme- deficient mice," *Journal of Cerebral Blood Flow and Metabolism*, vol. 18, no. 2, pp. 180–185, 1998.

[90] K. N. Murray, A. R. Parry-Jones, and S. M. Allan, "Interleukin-1 and acute brain injury," *Frontiers in Cellular Neuroscience*, vol. 9, article 18, 2015.

[91] D. Acalovschi, T. Wiest, M. Hartmann et al., "Multiple levels of regulation of the interleukin-6 system in stroke," *Stroke*, vol. 34, no. 8, pp. 1864–1869, 2003.

[92] E. Tarkowski, L. Rosengren, C. Blomstrand et al., "Early interleukin-6 signaling aggravates ischemic cerebral damage in mice: possible involvement of Stat3 activation in the protection of neurons," *Journal of Neurochemistry*, vol. 94, no. 2, pp. 459–468, 2005.

[93] S. A. Lodidick, A. V. Turnbull, and N. J. Rothwell, "Cerebral interleukin-6 is neuroprotective during permanent focal cerebral ischemia in the rat," *Journal of Cerebral Blood Flow & Metabolism*, vol. 18, no. 2, pp. 176–179, 1998.

[94] M. Hedtjärn, A.-L. Leverin, K. Eriksson, K. Blomgren, C. Mallard, and H. Hagberg, "Interleukin-1β involvement in hypoxic-ischemic brain injury," *The Journal of Neuroscience*, vol. 22, no. 14, pp. 5910–5919, 2002.

[95] F. M. Domac and H. Misirli, "The role of neutrophils and interleukin-8 in acute ischemic stroke," *Neurosciences*, vol. 13, no. 2, pp. 136–141, 2008.
[97] S. Jander, M. Schroeter, and G. Stoll, “Interleukin-18 expression after focal ischemia of the rat brain: association with the late-stage inflammatory response,” Journal of Cerebral Blood Flow & Metabolism, vol. 22, no. 1, pp. 62–70, 2002.

[98] K. L. Lambertsen, R. Gregersen, M. Meldgaard et al., “A role for interferon-gamma in focal cerebral ischemia in mice,” Journal of Neopatology and Experimental Neurology, vol. 63, no. 9, pp. 942–955, 2004.

[99] C. Marie, C. Pitton, C. Fitting, and J.-M. Cavaillon, “Regulation of anti-inflammatory cytokines (IL-4, IL-10, IL-13, TGFβ) of interleukin-8 production by LPS- and/or TNFα-activated human polymorphonuclear cells,” Mediators of Inflammation, vol. 5, no. 5, pp. 334–340, 1996.

[100] X. Liu, J. Liu, S. Zhao et al., “Interleukin-4 is essential for microglia/macrophage M2 polarization and long-term recovery after cerebral ischemia,” Stroke, vol. 47, no. 2, pp. 498–504, 2016.

[101] N. Liu, R. Chen, H. Du, J. Wang, Y. Zhang, and J. Wen, “Expression of IL-10 and TNF-α in rats with cerebral infarction after transplantation with mesenchymal stem cells,” Cellular and Molecular Immunology, vol. 6, no. 3, pp. 207–213, 2009.

[102] X. Zhao, H. Wang, G. Sun, J. Zhang, N. J. Edwards, and J. Aronowski, “Neuronal interleukin-4 as a modulator of microglial pathways and ischemic brain damage,” The Journal of Neuroscience, vol. 35, no. 32, pp. 11281–11291, 2015.

[103] L.-S. Chu, S.-H. Fang, Y. Zhou et al., “Minocycline inhibits 5-lipoxygenase activation and brain inflammation after focal cerebral ischemia in rats,” Acta Pharmacologica Sinica, vol. 28, no. 6, pp. 763–772, 2007.

[104] D. A. Ridder and M. Schwaninger, “NF-κB signaling in cerebral ischemia,” Neuroscience, vol. 158, no. 3, pp. 995–1006, 2009.

[105] A. Ockinghaus and S. Ghosh, “The NF-kappaB family of transcription factors and its regulation,” Cold Spring Harbor Perspectives in Biology, vol. 1, no. 4, article a000034, 2009.

[106] F. Wan and M. J. Lenardo, “Specification of DNA binding activity of NF-kappaB proteins,” Cold Spring Harbor Perspectives in Biology, vol. 1, no. 4, Article ID a000067, 2009.

[107] R. Berti, A. J. Williams, J. R. Moffett et al., “Quantitative real-time RT-PCR analysis of inflammatory gene expression associated with ischemia-reperfusion brain injury,” Journal of Cerebral Blood Flow and Metabolism, vol. 22, no. 9, pp. 1068–1079, 2002.

[108] M. Jiang, J. Li, Q. Peng et al., “Neuroprotective effects of bilobalide on cerebral ischemia and reperfusion injury are associated with inhibition of pro-inflammatory mediator production and down-regulation of JNK1/2 and p38 MAPK activation,” Journal of Neuroinflammation, vol. 11, article 167, pp. 1–17, 2014.

[109] L. Wang, Z. Li, X. Zhang et al., “Protective effect of shikonin in experimental ischemic stroke: attenuated TLR4, p-p38MAPK, NF-κB, TNF-α and MMP-9 expression, up-regulated claudin-5 expression, ameliorated BBB permeability,” Neurochemical Research, vol. 39, no. 1, pp. 97–106, 2014.

[110] H. Liu, X. Wei, L. Kong et al., “NOD2 is involved in the inflammatory response after cerebral ischemia-reperfusion injury and triggers NADPH oxidase 2-derived reactive oxygen species,” International Journal of Biological Sciences, vol. 11, no. 5, pp. 525–535, 2015.

[111] J.-H. Yi, S.-W. Park, R. Kapadia, and R. Vemuganti, “Role of transcription factors in mediating post-ischemic cerebral inflammation and brain damage,” Neurochemistry International, vol. 50, no. 7–8, pp. 1014–1027, 2007.

[112] S. Dziennis and N. J. Alkayed, “Role of signal transducer and activator of transcription 3 in neuronal survival and regeneration,” Reviews in the Neurosciences, vol. 19, no. 4-5, pp. 341–361, 2008.

[113] S. Dziennis, T. Jia, O. K. Ronnekleiv, P. D. Hurn, and N. J. Alkayed, “Role of signal transducer and activator of transcription-3 in estradiol-mediated neuroprotection,” The Journal of Neuroscience, vol. 27, no. 27, pp. 7268–7274, 2007.

[114] X. Wang, Q. Liu, A. Ihsan et al., “JAK/STAT pathway plays a critical role in the proinflammatory gene expression and apoptosis of RAW264.7 cells induced by trichothecenes as don and T-2 toxin,” Toxicological Sciences, vol. 127, no. 2, pp. 412–424, 2012.

[115] C. Schindler, D. E. Levy, and T. Decker, “JAK-STAT signaling: from interferons to cytokines,” The Journal of Biological Chemistry, vol. 282, no. 28, pp. 20059–20063, 2007.

[116] C. Lei, J. Deng, B. Wang et al., “Reactive oxygen species scavenger inhibits STAT3 activation after transient focal cerebral ischemia-reperfusion injury in rats,” Anesthesia & Analgesia, vol. 113, no. 1, pp. 153–159, 2011.

[117] H. Zhu, L. Zou, J. Tian, G. Du, and Y. Gao, “SMND-309, a novel derivative of salvianolic acid B, protects rat brains ischemia and reperfusion injury by targeting the JAK2/STAT3 pathway,” European Journal of Pharmacology, vol. 714, no. 1–3, pp. 23–31, 2013.

[118] L. Li, X. Zhang, L. Cui et al., “Ursolic acid promotes the neuroprotection by activating Nrf2 pathway after cerebral ischemia in mice,” Brain Research, vol. 1497, pp. 32–39, 2013.

[119] M. Li, X. Zhang, L. Cui et al., “The neuroprotection of oximatin in cerebral ischemia/reperfusion is related to nuclear factor erythroid 2-related factor 2 (Nrf2)-mediated antioxidant response: role of Nrf2 and hemeoxygenase-1 expression,” The Journal of Biological Chemistry, vol. 282, no. 34, pp. 595–601, 2011.

[120] Y. Dong, H. D. Liu, R. Zhao et al., “Ischemia activates JNK/c-Jun/AP-1 pathway to up-regulate 14-3-3σ in astrocyte,” Journal of Neurochemistry, vol. 109, supplement 1, pp. 182–188, 2009.