Detection of SARS-coronavirus-2 in the central nervous system of patients with severe acute respiratory syndrome and seizures

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Abstract
This study was designed to evaluate whether severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) can directly target the central nervous system (CNS). We present four patients suffering from the loss of consciousness and seizure during the clinical course of COVID-19 infection. In addition to positive nasopharyngeal swab tests, SARS-CoV-2 has been detected in their cerebrospinal fluid. This report indicates the neuroinvasive potential of SARS-CoV-2, suggesting the ability of this virus to spread from the respiratory tract to the CNS.

Keywords Neuroinvasion · Convulsion · SARS-CoV-2 · Brain · CNS

Introduction
Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) can affect various organs (Paramasivam et al. 2020; Sepehrinezhad et al. 2020; Tian and Ye 2020). Coronavirus disease 2019 (COVID-19) has been reported to be associated with a wide range of neurological symptoms (Ellul et al. 2020). However, whether the neurological manifestations reflect a direct effect of SARS-CoV-2 on the central nervous system (CNS) or indirect effects of parainfluenza virus that may gain access to the CNS by infecting endothelial cells via transcytosis to neural tissue (Paniz-Mondolfi et al. 2020). Furthermore, the post-mortem findings in patients who died of COVID-19 have revealed the presence of SARS-CoV-2 in the cortical neurons associated with minimal immune cell infiltrates in brain tissues (Song et al. 2020). Despite several reports of neurological manifestations and neuroinvasiveness of SARS-CoV-2 during infection (Guan et al. 2020; Li et al. 2020; Mao et al. 2020), additional evidence of neuroinvasiveness of SARS-CoV-2 is warranted. To evaluate the potential in previous outbreaks (Hung et al. 2003; Li et al. 2020). The brain autopsy findings in a patient with SARS-CoV-2 have suggested that this virus may gain access to the CNS by infecting endothelial cells via transcytosis to neural tissue (Paniz-Mondolfi et al. 2020). Furthermore, the post-mortem findings in patients who died of COVID-19 have revealed the presence of SARS-CoV-2 in the cortical neurons associated with minimal immune cell infiltrates in brain tissues (Song et al. 2020). Despite several reports of neurological manifestations and neuroinvasiveness of SARS-CoV-2 during infection (Guan et al. 2020; Li et al. 2020; Mao et al. 2020), additional evidence of neuroinvasiveness of SARS-CoV-2 is warranted. To evaluate the potential

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of SARS-CoV-2 to spread from the respiratory tract to the CNS, the present study was designed to assess SARS-CoV-2 in the cerebrospinal fluid (CSF) of patients with COVID-19 infection, who suffers from severe neurological manifestations. Here, we report the detection of SARS-CoV-2 in the CSF of four patients with loss of consciousness and seizure during the course of COVID-19 infection.

**Case presentation**

**Patient 1**

A 56-year-old female presented to the emergency department with a loss of consciousness. Seven days before, she developed myalgia, stomachache, and mild headache, which were superimposed with behavioral disturbances, hallucinations, and altered mental status later (Table 1). There was a history of upward gaze and left deviation of the face. Except for migraine headaches, her previous medical history was unremarkable. On the day of admission, she was unconscious with an eye-opening response only to painful stimulations. She was unable to move the left extremities with the impairment of the left plantar reflex. The pupils were midsized and reactive to light.

Due to the progressive loss of consciousness, increased level of PCO₂, and pulmonary secretions, the patient underwent mechanical ventilation after 24 h. Biochemical and hematological parameters were normal, except for a decreased lymphocyte count (Table 1). A lung high-resolution computed tomography (HRCT) has shown peripheral ground-glass consolidations in the apical zone of the right lung. Besides, a mild pleural effusion of both lungs accompanied by subsegmental collapse consolidations in the basal segments was noted (Fig. 1A).

MRI revealed bilateral asymmetrical areas of hyper signal intensity of the temporal lobes, inferior area of the frontal lobes, insula, and right parietal lobe (Fig. 1B: a). There was no absorption in post-contrast imaging.

The specimen of the nasopharynx and the CSF sample were both positive for SARS-CoV-2 (Fig. 1B: b-b'). In parallel, the expression of HSV was detected in the CSF. The value of protein and cellular counts were abnormal in the CSF analysis (Table 1).

**Patient 2**

The second patient was a 24-year-old man, who developed progressive dizziness and incoherent responses after 3 days of generalized weakness and myalgia. A few hours later, he had an attack of a generalized tonic-clonic seizure lasting for a few minutes (Table 1). No previous history of seizures or other medical conditions was reported.

The lung HRCT revealed multifocal and multi-lobar patchy ground-glass consolidations with peripheral distribution (Fig. 1A). The brain MRI showed small-size hyperintense foci in the subcortical regions of the parietal and temporal lobes in T2 and FLAIR images (Fig. 1B: c).

In this case, the specimen of the nasopharynx for COVID-19 was negative (Fig. 1B: d'). The PCR test for HSV in the CSF was also negative. However, the RNA of SARS-CoV-2 was detected in CSF (Fig. 1B: d). Analysis of the CSF showed abnormal protein levels and cellular components (Table 1).

**Patient 3**

A 65-year-old woman, after a few days of malaise and flu-like symptoms, was admitted with loss of consciousness followed by a seizure attack (lower limb shaking and urinary incontinence; Table 1). Lung HRCT revealed a few patchy areas of peripheral ground-glass consolidations and hilar lymphadenopathy (Fig. 1A). A CT scan of the brain was normal (Fig. 1B: e).

The nasopharyngeal swab and CSF samples were positive for COVID-19 (Fig. 1B: f-f'). The RT-PCR test for HSV in the CSF was negative. Elevated levels of protein and white blood cell count (with mononuclear predominance) in the CSF were reported (Table 1).

**Patient 4**

A 71-year-old man was hospitalized with loss of consciousness and seizure (Table 1). One week before, he was diagnosed with COVID-19 infection after 3 days of malaise, cough, and fever and received azithromycin (250 mg/day). No other symptoms were reported. On the day of admission, he was lethargic and afebrile. Lung HRCT revealed peripheral multifocal and multi-lobar patchy ground-glass consolidations in both lungs (Fig. 1A). A CT scan of the brain showed only senile atrophies and small vessel disease (Fig. 1B: g).

The nasopharyngeal swab and CSF samples were positive for COVID-19 (Fig. 1B: h-h'). The RT-PCR test for HSV in the CSF was negative. Increased levels of protein and white blood cell count were observed in the CSF (Table 1).

**Discussion**

This report described four COVID-19-positive patients with seizures and loss of consciousness, who were positive for SARS-CoV-2 in their CSF. Our findings suggest the ability of SARS-CoV-2 to spread from the respiratory tract to the CNS. These patients did not have any significant risk factors.
Table 1  Clinical characteristics, serum biochemical parameters, hematological laboratory values, CSF cellular components, herpes PCR results, and culture of CSF in different patients

| Variable                                           | Patient 1 | Patient 2 | Patient 3 | Patient 4 | Normal value |
|----------------------------------------------------|-----------|-----------|-----------|-----------|--------------|
| **Demographic information and clinical symptoms**  |           |           |           |           |              |
| Age (years)                                        | 56        | 24        | 65        | 71        | -            |
| Gender                                             | Female    | Male      | Female    | Male      | -            |
| Neurologic complaint                               | Altered mental status-seizure | Altered mental status-seizure | Loss of consciousness and seizure | Loss of consciousness and seizure | - |
| Primary presentation                               | Myalgia-mild headaches | Myalgia | Malaise-flu-like syndrome | Malaise, cough and fever | - |
| Time from onset of the disease to neurologic       | 7 days    | 3 days    | 3 days    | 10 days   | -            |
| presentation                                       |           |           |           |           |              |
| Fever on admission                                 | Yes       | No        | Yes       | No        | -            |
| Outcome                                            | ICU admission | Good | Good | Good | - |
| Nasopharynx for COVID-19 PCR                       | Positive  | Negative  | Positive  | Positive  | -            |
| CSF for COVID-19 PCR                               | Positive  | Positive  | Positive  | Positive  | -            |
| **Serum biochemical analysis**                     |           |           |           |           |              |
| Blood sugar (mg/dl)                                | 101       | 107       | 119       | 193       | < 200        |
| Urea (mg/dl)                                       | 25        | 52        | 52        | 45        | 18–55        |
| Creatinine (mg/dl)                                 | 0.7       | 1         | 1.3       | 1.1       | 0.7–1.4      |
| Sodium (mg/dl)                                     | 132       | 142       | 140       | 137       | 135–145      |
| Potassium (mg/dl)                                  | 3.3       | 4.4       | 4.1       | 5.2       | 3.5–5.3      |
| Calcium (mg/dl)                                    | 9.0       | 9.2       | -         | 9.8       | 8.6–10.3     |
| Magnesium (mg/dl)                                  | 2.1       | 2.5       | -         | 2.1       | 1.2–2.6      |
| Aspartate transaminase (U/L)                       | -         | 31        | 44        | -         | 5–40         |
| Alanine transaminase (U/L)                         | -         | 43        | 24        | -         | 5–40         |
| **Hematological laboratory values**                |           |           |           |           |              |
| White blood cell (×10^3 mcl)                       | 11.9      | 9.5       | 9.5       | 10.3      | 4–10         |
| Hemoglobin (g/dl)                                  | 13.3      | 13.6      | 13.7      | 13.7      | 13–17        |
| Hematocrit (%)                                     | 40.5      | 41.3      | 43.1      | 42.2      | 40–50        |
| Red blood cell (×10^6 mcl)                         | 4.77      | 4.02      | 5.09      | 5.04      | 4.5–5.5      |
| Polymorphonuclear leukocytes (%)                   | 88.4      | 67        | 70.8      | 80        | 30–70        |
| Lymphocyte (%)                                     | 8         | 26.8      | 16.3      | 15        | 20–50        |
| ESR mm                                             | 34        | 27        | 44        | 34        | < 15         |
| Platelet (×10^3 mcl)                               | 213       | 263       | 230       | 282       | 150–450      |
| C-reactive protein (mg/dl)                         | 61.3      | 19.7      | 19.1      | 20.1      | < 6          |
| **CSF parameters analysis**                        |           |           |           |           |              |
| Sugar (mg/dl)                                      | 63        | 64        | 66        | 133       | > 2/3 concurrent BS |
| Protein (mg/l)                                     | 130       | 85        | 124       | 64        | 15–45        |
| White blood cell (/mm^3)                           | 70        | 10        | 90        | 0         | 0–1          |
| Polymorphonuclear leukocytes (%)                   | 5         | 10        | 10        | 0         | 0            |
| Mononuclear (%)                                    | 95        | 90        | 90        | 0         | -            |
| LDH (U/l)                                          | 197       | 42        | 57        | 82        | < 100        |
| Red blood cell (/mm^3)                             | 70        | 20        | 20        | 0         | 0–100        |
| Herpes-PCR                                         | Positive  | Negative  | Negative  | Negative  | (Negative)   |
| Bacterial culture                                  | Negative  | Negative  | Negative  | Negative  | (Negative)   |

*BS* blood sugar, *ESR* erythrocyte sedimentation rate, *g/dl* gram/deciliter, *mcl* microliter, *mg/dl* milligram/deciliter, *mm3* millimeter cubed, *U/l* Unites/liter
for neurological diseases and none of them had a history of neurological disorders.

During the COVID-19 pandemic, several studies have reported neurological manifestations in patients with COVID-19 (Farhadian et al. 2020; Mao et al. 2020; Moriguchi et al. 2020). Recently, several research groups have focused on the explanation of neurological manifestations of COVID-19 via its neuroinvasive potential (Farhadian et al. 2020; Li et al. 2020; Moriguchi et al. 2020; Sepehrinezhad et al. 2020). An increasing amount of evidence suggests the neuroinvading potential of SARS-CoV-2, which may lead to clinical symptoms and brain damage (Najjar et al. 2020; Wood 2020; Zangbar et al., 2021). The hypothesis of the SARS-CoV-2 invasion into the CNS has been supported by studies on the coronaviruses that caused previous outbreaks (Sepehrinezhad et al. 2020). The neuroinvasion of SARS-CoV-2 has been detected in human brain autopsy specimens (Paniz-Mondolfi et al. 2020; Song et al. 2020). The present study confirmed
the neuroinvasive propensity of SARS-CoV-2 in patients who suffer from severe neurological symptoms.

Taken together, we demonstrated the ability of SARS-CoV-2 to attack the CNS, which was associated with the loss of consciousness and seizure. Although this study demonstrated the neuro-invading properties of SARS-CoV-2, further research will be required to determine the mechanisms underlying SARS-CoV-2-mediated neurological manifestation and brain injury.

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Authors' contributions F.R. contributed to the interpretation of the neurological results, worked on the manuscript, and performed CSF collection. S.AJ and S.H. performed RT-PCR tests and contributed substantially to the conception of the analysis and interpretation of PCR results. F.R, A.R, F.M, F.S.E.F, and M.E.R. processed the clinical data, contributed to the case presentation. A.S. conceived the study, designed the tables, and contributed to the writing of the manuscript. A.G. critically revised the manuscript and provided the final outlines of the version to publish. S.S.N. conceived the study, wrote the manuscript, and supervised the work. All authors provided critical feedback and helped shape the research and manuscript.

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Data availability The data collection method and RT-PCR protocol are available in the supplementary materials. Additional data and materials can be available from the corresponding authors upon request.

Compliance with ethical standards

Ethics approval and consent to participate The ethics committee of Mashhad University of Medical Sciences approved the study. Written informed consent has been obtained from the patients in accordance with the Declaration of Helsinki.

Conflict of interest The authors declare that they have no conflict of interest.

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