New Insights on Taxonomy, Phylogeny and Population Genetics of Leishmania (Viannia) Parasites Based on Multilocus Sequence Analysis

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Abstract

The Leishmania genus comprises up to 35 species, some with status still under discussion. The multilocus sequence typing (MLST)—extensively used for bacteria—has been proposed for pathogenic trypanosomatids. For Leishmania, however, a detailed analysis and revision on the taxonomy is still required. We have partially sequenced four housekeeping genes—glucose-6-phosphate dehydrogenase (G6PD), 6-phosphogluconate dehydrogenase (6PGD), mannose phosphate isomerase (MPI) and isocitrate dehydrogenase (ICD)—from 96 Leishmania (Viannia) strains and assessed their discriminatory typing capacity. The fragments had different degrees of diversity, and are thus suitable to be used in combination for intra- and inter-specific inferences. Species-specific single nucleotide polymorphisms were detected, but not for all species; ambiguous sites indicating heterozygosis were observed, as well as the putative homozygous donor. A large number of haplotypes were detected for each marker; for 6PGD a possible ancestral allele for L. (Viannia) was found. Maximum parsimony-based haplotype networks were built. Strains of different species, as identified by multilocus enzyme electrophoresis (MLEE), formed separated clusters in each network, with exceptions. NeighborNet of concatenated species confirmed species-specific clusters, suggesting recombination occurring in L. braziliensis and L. guyanensis. Phylogenetic analysis indicates L. lainsoni and L. naiffi as the most divergent species and does not support L. shawi as a distinct species, placing it in the L. guyanensis cluster. BURST analysis resulted in six clonal complexes (CC), corresponding to distinct species. The L. braziliensis strains evaluated correspond to one widely geographically distributed CC and another restricted to one endemic area. This study demonstrates the value of systematic multilocus sequence analysis (MLSA) for determining intra- and inter-species relationships and presents an approach to validate the species status of some entities. Furthermore, it contributes to the phylogenies of L. (Viannia) and might be helpful for epidemiological and population genetics analysis based on haplotype/diplotype determinations and inferences.

Introduction

Leishmania are the causative agents of leishmaniasis, which can present in different forms, from simple cutaneous to the deadly visceral disease, and are found in most tropical and sub-tropical regions. In spite of their morphological homogeneity, more than 20 species have been described for the Leishmania genus. Phenotypic diversity is observed not only between species, but also even in virulence levels among clones [1] which, upon interaction with the host’s immunological response, contributes to determining the observed clinical pleiotropy and affects the efficiency of therapy applied [2]. Leishmania genetic diversity may also compromise vaccine development, although key antigen genes are highly conserved. Understanding leishmaniasis and the development of measures to counter its spread depend on the ability to identify Leishmania species and characterize genetic variants.

Multilocus enzyme electrophoresis (MLEE) is still considered by many to be the gold standard for Leishmania identification, but several DNA based methods have proven useful to study Leishmania genetic diversity [3,4]. DNA sequencing and PCR-RFLP of hsp70 genes have been shown to be promising for the identification of Leishmania parasites [5,6], although too conserved for intra-specific diversity studies. Highly polymorphic markers, such as microsatellites, perform poorly at taxonomic levels higher than species, whilst most other genotyping methods rely on multiplex genes that are more difficult to analyze.

A standardized, sensitive, practical and reproducible typing method, such as multilocus sequence typing (MLST), must form the basis for a robust classification of Leishmania species, which is achievable in most laboratories even in the advent of fast and cheaper genome sequencing. MLST, as proposed in 1998 for bacterial pathogens [7], provides a portable, reproducible, and quantitative typing system. It has since been applied to diploid organisms [8–10], including
Author Summary

*Leishmania* is a protozoan genus comprising many species, some associated with a human neglected disease called leishmaniasis. This parasite is found worldwide and is transmitted by sand flies, having numerous domestic and sylvatic animals as reservoirs. *Leishmania* is genetically and ecologically diverse and it has been argued that this has an impact on the epidemiology of the disease. Many typing methods have been proposed for the study of this diversity, although a generally agreed methodology is still required. Also, there is still a lack of consensus on the validity of some species. Multilocus sequence typing (MLST) is a method for studying the population structure and diversity of pathogens, but before an MLST scheme can be proposed it is essential to undertake a detailed analysis and selection of the sequences that are to be included in the system. Here, we sequenced four gene fragments of 96 *L. (Vianna)* strains, representing most species from this subgenus. Our results showed a good agreement between the current species assignment and the multilocus sequence analysis. Evidence of genetic recombination was found and the phylogenetic relationships were determined. Overall the results point to the feasibility of an MLST scheme for *Leishmania* and indicate that the four gene fragments analyzed could form part of this typing system. This will certainly be a valuable approach for taxonomy, population genetics, and epidemiological studies of this pathogen.

*Leishmania*, with ten markers described for the *L. donovani* complex [11–13] and four targets proposed for *L. (Vianna)* spp. [14]. MLST databases for *Leishmania* have not yet been implemented and the published number of strains and species typed is relatively low. Alongside MLST, multilocus sequence analysis (MLSA) has been shown to be a good tool for strain characterization and epidemiological surveillance, as well as population structure and evolutionary studies [9,10,15,16]. The implementation of an MLST system demands careful evaluation of markers beforehand to study their diversity and phylogenetic consistency [17,18].

We report here the evaluation of four candidate coding regions, located in different chromosomes, to be included in an MLSA system of the subgenus *L. (Vianna)*. These regions are part of the genomes for the metabolic enzymes glucose-6-phosphate dehydrogenase (G6PDH, EC 1.1.1.49), phosphogluconate dehydrogenase (6PGD, EC 1.1.1.44), mannose phosphate isomerase (MPI, EC 5.3.1.8) and isocitrate dehydrogenase (ICD, EC 1.1.1.42). Three of these genes had been studied previously, but mainly for Peruvian isolates of the *L. braziliensis* complex. Herein a large number of strains and most species of the subgenus *L. (Vianna)* were included. The sequences were used to conduct haplotype analysis, including the construction of haplotype networks. Diploid multilocus analyses were performed and a concatenated tree was defined, as well as clonal complexes (CC). Using an MLSA approach, we report the genetic diversity of the four gene fragments evaluated in the *L. (Vianna)* group and demonstrate their value for taxonomic, phylogenetic and population genetic studies of *Leishmania* parasites.

Methods

Ethics statement

Research in this study was subject to ethical review by the European Commission and approved as part of contract negotiation for Project LeishEpiNetSA (contract 01547); the work conformed to all relevant European regulations. The research was also reviewed and approved by the ethics committee of the London School of Hygiene and Tropical Medicine (approval 5092). The *Leishmania* strains analyzed were principally strains derived from international cryobanks such as Coleção de Leishmaniose do Instituto Oswaldo Cruz (CLIOC) and the cryobank of the London School of Hygiene and Tropical Medicine (LSHTM). In all cases *Leishmania* were isolated from patients as part of normal diagnosis and treatment with no unnecessary invasive procedures and with written and/or verbal consent recorded at the time of clinical examination. Data on isolates were coded and anonymised. The *Leishmania* strains are deposited either at the LSHTM (n = 9) or at CLIOC (n = 87) as open data. CLIOC is a Depository Authority by the Ministry of the Environment [Fiel Depositária pelo Ministério do Meio Ambiente, MMA] (D.O.U. 05.04.2005. All samples were used for research purposes only and the data were analyzed anonymously in the scope of resolution 21 [August 31, 2006 – CGEN/MMA], for which authorization is not required.

Samples and GenBank available sequences

Ninety six strains were obtained from frozen stocks from CLIOC (n = 87), and from the London School of Hygiene and Tropical Medicine (LSHTM) (n = 9) (Table S1). Strains were chosen to be representative of the zymodeme and geographical diversity of species of the subgenus *Leishmania (Vianna)* in South America, and in particular Brazil: *L. (V) braziliensis*, *L. (V) guyanensis*, *L. (V) naiffi*, *L. (V) lainsoni*, *L. (V) shawi*, *L. (V) altenuis* and *L. (V) lindemeri*. Most strains had been previously characterized by MLEE [19]. DNA sequences were retrieved from GenBank of *L. (Vianna)* species: 14 of G6PD; 24 of 6PGD; 20 of MPI and 1 of ICD (Table S2). Only those presenting full coverage with the alignment of sequences obtained in the present study were included in the analysis: for G6PD: 3 *L. braziliensis*, 2 *L. guyanensis*, 1 *L. peruviana*, 1 *L. panamensis*, 1 *L. lainsoni*; for 6PGD: 10 *L. peruviana*, 07 *L. braziliensis*, 1 *L. panamensis*; for MPI: 11 *L. peruviana*, 9 *L. braziliensis*; for ICD: 1 *L. braziliensis*.

DNA extraction and PCR

Promastigotes were grown at 25°C in Schneider’s medium supplemented with 20% (v/v) heat-inactivated fetal bovine serum to a density of 1 × 10^6 cells/mL (late log phase), as estimated by counting in a Neubauer chamber. DNA extraction was performed using the Wizard DNA purification Kit (Promega, Madison, USA) following the manufacturer’s instructions.

The chosen loci are distributed on different chromosomes according to *L. (Vianna) braziliensis* genome sequencing data: 6PGD, on chromosome 34; G6PD, on chromosome 20; MPI, on chromosome 12; ICD, on chromosome 33.

Primers (Table 1) were designed in conserved regions of gene sequences from the published *L. major* and the *L. braziliensis* genomes in Genbank (www.ncbi.nlm.nih.gov/Genbank) to amplify sections of the coding regions of the genes that would be amenable to full sequencing using the PCR primers and that include putative species-specific polymorphisms as well as singleton SNPs. Amplification reactions had, for 50 μl total volume, 0.1 mM of each primer, reaction buffer (100 mM Tris–HCl, pH 8.8; 500 mM KCl, 1% Triton X-100; 15 mM MgCl_2), 0.25 mM deoxyribonucleotide triphosphate (dNTPs), 0.025 U FideliTaq/GoTaq polymerase and approximately 50 ng DNA. Amplification conditions were: 94°C for 2 minutes, followed by 35 cycles at 94°C for 30 seconds, 55°C (for ICD, 6PGD and G6PD) or 58°C (for MPI) for 30 seconds and 68°C for 1 minute, with a final extension at 68°C for 5 min.
DNA sequencing

PCR products were purified with the Wizard SV Clean-up System (Promega). The final DNA concentration was estimated by comparison with a DNA Ladder Marker (Promega) in 2% agarose gel. Sequencing was performed with the same primers used for amplification, using the ABI PRISM BigDye Terminator v3.1 Cycle Sequencing Kit, and products analyzed in an automated DNA sequencer (ABI-3730). Consensus sequences were generated and edited in Phred/Phrap/Consed Version: 0.020425.c [20] from two forward and two reverse strands. Sequences with Phred values below ten over their extent were discarded and only sequence segments with values above twenty were used for contig construction. Ambiguous (heterozygous) sites were coded using the standard IUPAC codes for combinations of two or more bases. Contigs from all samples were manually assembled and aligned in MEGA4 [21]. The homologous sequences available in GeneBank for other *L. braziliensis* isolates were obtained using the Basic Local Alignment Search Tool (BLAST) algorithm hosted by NCBI, National Institute of Health, USA (http://www.ncbi.nlm.nih.gov) and included in the alignments.

Haplotype analyses

Haplotype reconstruction was done through DNAsp5 [22] using the PHASE algorithm, which automatically assigns a haplotype number (H) for each unique haploid sequence. Haplotype data was then used in DNAsp to calculate the synonymous and non-synonymous substitution rates and haplotype diversity (HD). Haplotype diversity (HD) was calculated as: HD = N(1−∑xi^2)/(N−1), where xi is the haplotype frequency of each haplotype in the sample and N is the sample size. The average haplotype diversity for species was compared through ANOVA.

For each sample a sequence type number (ST) was defined with each marker, which in the homozygous strains was identical to the haplotype number (H), and for the heterozygous strains was a combination of the two possible alleles. A diploid sequence type (DST) was defined for the final combination of STs of the four markers. The sequences from the four genes were concatenated using BioEdit v7.0.9 and unique concatenated sequences were identified with the DST previously assigned. To evaluate the phylogenetic information provided by the four markers and to investigate the presence of recombination signatures, a Neighbor-Net network was built in SplitsTree 4.0 [24] based on genetic distances calculated according to the Kimura-2 parameter model of nucleotide substitutions from concatenated data.

| Locus | Gene ID* | Gene length | Amplicon size (bp) | Primer positions | Primers sequence 5’-3’ |
|-------|----------|-------------|--------------------|------------------|-----------------------|
| G6PD  | LbrM.20.0160 | 1686 | 881 | 173 | ATGGAAAGGTGTGATCGAAAT |
| 6PGD  | LbrM.34.3250 | 1440 | 836 | 143 | CTCAAGGAACATAGGACAGGA |
| MPI   | LbrM.32.1750 | 1287 | 681 | 128 | GGCAGAATGTAGGGGAGTT |
| ICD   | LbrM.33.2820 | 1278 | 1022 | 99 | GAATCGGGAGAAGATCACA |

ST data combinations (with the exception of heterozygous strains) were analyzed in e-BURST v.3 (http://eburst.mlst.net/v3/enter_data/single/) to define clonal complexes (CC), which are sets of related strains containing pairs of strains that share at least (L-1) identical alleles at the L loci with at least one other member of the CC.

**Results and Discussion**

We report here the most comprehensive multilocus sequence analysis study to date of the subgenus *Leishmania* (*Viannia*), based on fragments of four different metabolic enzyme coding genes from a wide range of species, zymodemes and geographical origins.

With the aim of developing a working MLST scheme, we selected polymorphic coding regions that could be sequenced with forward and reverse primers. The full coding regions of three of the studied genes (6PGD, G6PD and MPI) had already been sequenced and characterized for some *L. (Viannia)* strains, mainly of Peruvian *L. braziliensis* and *L. peruviana*. However, we also partially analyzed the coding region for ICD and studied a large number of strains (96), to include most known species or complexes of *L. (Viannia)*.

**Genetic diversity of the four housekeeping genes used in the MLSA of Leishmania (Viannia): Polymorphic, ambiguous and multi-allelic sites**

The sequences obtained for each gene varied between 589 (MPI) and 914 bp (ICD) (Table 2) and covered between 38% (6PGD) and 71% (ICD) of the corresponding gene. Sequences were deposited in GenBank (www.ncbi.nlm.nih.gov/Genbank) with accession numbers JN996517–JN996708 and JQ181608–JQ181801 and are also available at the CLIOC website (http://clio.fiocruz.br/).

Gene fragments had 50–78 polymorphic sites (respectively, 7.3–8.7%) for the strains studied here, of which 46 (G6PD) to 76 (ICD) were parsimony informative (PI) sites (Table 2). 6PGD had the...
highest percentage of PI sites (8.4%), with the lowest found in G6PD (6.7%). The studied G6PD region had a much higher number and percentage of singletons, 4 (0.6%), in relation to the respective total template length than the other three gene regions (0.1–0.2%; Table 2).

The four sequenced fragments, representing different loci, had different degrees of diversity between species groups as seen by the haplotype diversity (Figure 1) and are thus suitable to be used in combination for intra- and inter-specific inferences. Although theoretically including more loci improves discriminatory capacity, it has been described that increasing the number of genes above four did not increase discrimination of MLST [18]. Furthermore, the index of diversity [25] for the targets included in the present study implies that the typing results can be interpreted with confidence.

Overall, after including published gene sequences for the same regions as studied here (except for ICD, which had no available sequences) gene fragments provided 51 (MPI) and 64 (6PGD) PI sites (Table 3).

Most polymorphic sites were bi-allelic, although sites with three variants occurred in three genes, G6PD, 6PGD and in ICD (Table S3). Recently, the genome plasticity of some Leishmania species was analyzed [26]. The evaluation of the L. braziliensis genome showed that 30 of 35 chromosomes are clearly trisomic; three are tetrasomic (chromosomes 4, 5 and 29) and one hexasomic (chromosome 31). Moreover, the same study showed that multicopy genes are found preferentially on non-supranumerary chromosomes. Such peculiarities are of great importance in all molecular analyses proposed for the Leishmania genus. In the present work, all markers used are located on different chromosomes, which are described as non-supranumerary [26]. However, the description of a trisomic genome for L. braziliensis indicates that multiallelic possibilities might be quite frequent when polymorphic sites are analyzed in DNA sequences of Leishmania (Viannia) species.

### Table 2. Characteristics of each sequenced region for the 96 strains used in the present study.

| Gene | Length (bp) | PS (bp) | PI (bp) (% of length) | Singletons (bp) | Ambiguous sites (bp) | Number of sites (% of length) | Number of isolates (% total strains) |
|------|-------------|---------|-----------------------|-----------------|----------------------|-----------------------------|-------------------------------------|
| G6PD | 683         | 50      | 46 (6.7%)             | 4 (0.6%)        | 6 (0.9%)             | 8 (8.3%)                    |                                     |
| 6PGD | 716         | 63      | 60 (8.4%)             | 1 (0.1%)        | 5 (0.7%)             | 9 (9.4%)                    |                                     |
| MPI  | 589         | 50      | 49 (8.3%)             | 1 (0.2%)        | 2 (0.3%)             | 2 (2.1%)                    |                                     |
| ICD  | 914         | 78      | 76 (8.3%)             | 2 (0.2%)        | 6 (0.7%)             | 10 (10.4%)                  |                                     |

Percentages are relative to the total number of sites (% of length) or strains (% of total strains). PS – Polymorphic sites; PI - parsimony informative sites.

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Figure 1. Comparison of haplotype diversity values of each locus within species groups. ICD was the only polymorphic marker for L. shawi strains. * G6PD and MPI were the least polymorphic markers (P<0.001) within species for L. lainsoni and L. guyanensis respectively.
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Moreover, gene copy numbers might also change between species and generations of the same strain.

Twenty-six isolates had at least one locus with double peaks in the chromatograms (Table 2 and Table S4). The number of sites showing double peaks was similar in three gene regions, but much smaller for MPI, which also had a much smaller number of strains with double peaks in their sequences compared to the other targets (Table 2). Different strains presenting double peaks at the same site were observed in all markers, except MPI (Table S4).

Although most strains were homozygous, almost a third had at least one locus with double peaks, except L. shawi. MPI was the most homozygous gene, even though it was not the most conserved. All the other markers presented more than one double peak per strain and more than one strain with double peaks. It is possible that this is a sampling effect of the sequenced region in MPI, or that gene conversion in this gene is higher [27].

The presence of ambiguous sites and possible parental alleles among the samples studied strongly suggests real heterozygosis and also the occurrence of some level of recombination. Ambiguous sites were observed in twenty positions over the four gene fragments analyzed (Table S4), and putative homozygous parental were observed in eight of them. Many strains had two or more heterozygous sites. Heterozygosis can be caused by mutation in one allele but it can also be caused by genetic exchange between strains with different alleles. Mutation is more likely for single heterozygous sites but recombination is a more parsimonious explanation for two or more sites [28].

The majority of heterozygous alleles among our sample presented just one ambiguous site. Exceptions were encountered in one L. braziliensis strain showing HP 8/17 in 6PGD and 6/11 in ICD, with two and three ambiguous sites respectively. This observation reinforces the fact that real heterozygosis might be occurring, because these two loci are on different chromosomes and in both sequence alignments at least two ambiguous sites were observed in that sample. Moreover, after determination of the minimum number of recombination events by using DNAsp, recombination between sites was detected for the four markers, with highest frequency for 6PGD and least for G6PD (data not shown).

The assumption that ambiguities in the chromatograms are the result of heterozygosis can only be reliably postulated if the target is a single copy gene. In our case, the four selected genes are single copy genes, as estimated by in silico analysis of the four available Leishmania species reference genomes (data not shown). Heterozygous samples would need confirmation through biological cloning of isolates, to exclude polyclonal populations. The strains we have used were not cloned, except for the reference strains. As far as we know, none of the studies aiming to construct an MLST system for Leishmania have used biologically and/or genetically cloned samples. Following an overview of sample profiles, without cloning, if ambiguous sites are detected, the strains containing them should be selected for deeper analysis and cloning.

Species-specific SNPs

Putative species-specific SNPs, as determined through the data presented here and available in GenBank, were detected for L. guyanensis in 6PGD (A105); for L. naiffi in MPI (15T, 82A, 98A, 270A, 306G, 498C, 543T) and G6PD (320C, 341C, 432A, 467A); for L. lainsoni in MPI (33G, 105G, 294T) and G6PD (116A, 239T, 247G, 326A, 464C, 482C, 572A, 577T) and for L. shawi in MPI (135G). No species-specific SNPs were detected for L. braziliensis. ICD was the only marker that did not present species-specific SNPs.

In a previous study, species-specific SNPs for all species, including L. braziliensis were shown [14]. This incongruence between the two studies might be a consequence of differences in the gene fragments analyzed as well as in the strains studied. SNP markers should thus be used for species identification with great care, even when more strains are studied in future. Either full sequences should be obtained, or a large panel of SNP markers should be used for reliable identification and characterization.

Identification of assorted Leishmania (Viannia) haplotypes

Here we found a large number of unique haplotypes, which are likely to be rare or recent in the population. In contrast, some haplotypes were shared across species, such as haplotype H1 of 6PGD, which was detected in strains from different species: almost all L. braziliensis (n = 40), nine L. peruviana as well as two previously published L. braziliensis sequences, one L. lainsoni (IIOC 2500 from Acre) and one L. naiffi (IIOC 835 from Amazonas) strain. This could be due to recombination or convergent evolution, but it could also represent the most likely ancestral haplotype, as depicted from the haplotype network constructed for this gene (Figure 2). Remarkably, those species present different electrophoretic mobility for the 6PGDH enzyme system. Such incongruence might be due to the fact that the entire coding region was not sequenced here, so sequence sections coding for differences in MLEE might not be present in the current analysis, or it might be due to post-transcriptional and post-translational modifications [28].

Table 3. Characteristics of each sequenced region for all available L. (Viannia) (retrieved from GenBank plus the 96 strains included) and for L. braziliensis complex samples.

| Gene | Length (bp) | PI* | Singletons | Ambiguous sites |
|------|-------------|-----|------------|-----------------|
| G6PD | L. (Viannia) | 683 | 54 (7.9%) | 4 (0.6%) | 6 | 8 |
| L. braziliensis | 11 (1.6%) | 0 | 2 | 5 |
| 6PGD | L. (Viannia) | 716 | 64 (8.9%) | 1 (0.1%) | 5 | 9 |
| L. braziliensis | 27 (3.8%) | 0 | 3 | 5 |
| MPI | L. (Viannia) | 589 | 51 (8.7%) | 1 (0.2%) | 2 | 2 |
| L. braziliensis | 17 (2.9%) | 1 (0.2%) | 3 | 3 |

Percentages are relative to the total number of sites (% of length).

*PI - parsimony informative sites.

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Figure 2. Maximum parsimony-based haplotype networks. A) G6PD; B) 6PGD; C) MPI; D) ICD. The 96 strains selected for the study (Table S1) plus sequences of *Leishmania* (Viannia) strains retrieved from GenBank (Table S2) were included. Haplotype frequency is represented by the size of each node and the numbers of polymorphisms are indicated in the branches by dashes: one = 02 to 05 polymorphism; two = 06 to 10; three = more than 10 polymorphisms. Each species, considering the MLEE characterization, were assigned by different colors as coded by the legend.

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Table 4. Haplotype analysis per marker for the 96 strains included in the present study.

| Gene      | Length (bp) | N° of different alleles | HD* | N° unique alleles |
|-----------|-------------|-------------------------|-----|-------------------|
| G6PD      | 683         | 33                      | 0.78| 21 (63.6%)        |
| 6PGD      | 716         | 34                      | 0.76| 22 (64.7%)        |
| MPI       | 589         | 25                      | 0.77| 16 (64%)          |
| ICD       | 914         | 43                      | 0.85| 31 (72.1%)        |

Percentages are relative to the number of different alleles.

*HD - haplotype diversity.

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Upon haplotype assignment it was found that among the 96 studied strains there were 25 (MPI) to 43 (ICD) different haplotypes, of which 16 (MPI) to 31 (ICD) were represented by only one strain. Nevertheless, ICD had the highest percentage of exclusive haplotypes per strain (72.1%) and G6PD had the lowest (63.6%; Table 4).

Strains identified as *L. braziliensis* represented more than 50% of the strains studied. The greatest number of haplotypes was observed within the *L. braziliensis* group for all markers except G6PD for which the greatest number of haplotypes was observed within the *L. guyanensis* group (Table 3). Around 30% of the haplotypes in each gene were detected in *L. braziliensis* (Table 3). However, almost no singletons and no species-specific SNPs were observed among the *L. braziliensis* strains, but some ambiguous sites were seen (Table S4). *L. utengensis* and *L. lindenberghii* had unique haplotypes for all markers, with the exception of 6PGD for *L. lindenberghii*, as mentioned above.

Regarding G6PD, 12 haplotypes were shared by at least two strains. The most common haplotype, H6, was found in 45 strains, characterized as *L. braziliensis* or *L. peruviana*. Four additional haplotypes were found in previously published sequences, one in *L. panamensis* (H36), two *L. braziliensis* (H34, from Peru, and H35, from Brazil) and one *L. lainsoni* (H37, from Peru). One published *L. guyanensis* sequence had haplotype H4, the most common *L. guyanensis*/ *L. shawi* haplotype in this analysis (Table S2). Except for *L. shawi* and *L. peruviana*, which shared haplotypes with *L. guyanensis* and *L. braziliensis* respectively, the network depicted from this target presented the best clustering accordingly to species (Figure 2A).

Concerning 6PGD, haplotype H15 was only found in one isolate of *L. braziliensis* from Acre, northern Brazil, but it was identical to four previously published *L. braziliensis* strains from Peru. Similarities among strains from these two adjacent areas were reported previously based on MLMT [29], corroborating the suggestion that a geographic cluster and probably a hierarchical population structure of *L. braziliensis* exist in this area. *L. guyanensis* had 10 haplotypes, of which H9 was the most frequent, and *L. shawi* had one distinct haplotype (H3). Only two 6PGD haplotypes found in previously published sequences (one *L. braziliensis* from Peru and one *L. panamensis*) were not detected among the strains studied here, indicating a good coverage of haplotype diversity by our study.

The two most frequent alleles observed, corresponding to 6PGD H1 and H8 (Figure 2B), were also observed combined in one heterozygous *L. braziliensis* strain (IOCL 2833). These haplotypes were also observed in one (H1; IOCL 2494) and two (H8 IOCL 918 and 2538) other heterozygous strains, combined with other haplotypes (Table S4). Some 6PGD haplotypes were unique to heterozygous strains, as H4/H5 and H30/H31 (Table S1).

MPI *L. braziliensis* haplotype H8 was also found in one *L. guyanensis* strain. Haplotype H7 was the most common in *L. braziliensis* (41 strains) and was also found in eight previously published sequences of *L. peruviana* and eight of *L. braziliensis* (Table S2). MPI was previously reported as a good target to discriminate between *L. braziliensis* and *L. peruviana* [14,30]. Our conflicting results could be related to differences in the fragment regions analyzed, or might reflect a bias in the sample analyzed previously. Two published sequences of *L. peruviana* strains presenting an ambiguous site had the most common H7 and a new haplotype 26 after allele reconstruction. One available *L. peruviana* and one *L. braziliensis* sequence from Peru presented additional haplotypes H27 and H28, respectively (Figure 2C; Table S2).

The greatest number of haplotypes was found in ICD, for almost all species, although there were similar levels for G6PD and 6PGD in *L. naiffi*. ICD H6 was by far the most common *L. braziliensis* haplotype, including one previously published sequence (Table S2). ICD H9, the most common *L. guyanensis* haplotype, was also found in one *L. shawi* (IOCL 3200) (Figure 2D; Table S1).

Overall, and including published gene sequences for the same regions as studied here, there are 28 to 43 different described alleles in *L. (Viannia)*, of which 11 to 16 are in the *L. braziliensis* complex.

Haplotype diversity and haplotype network selection of marker combinations for resolution of inter- and intra-species relationships

Overall, haplotype diversity (HD) was higher for ICD (0.85), similar among the other gene regions (0.76–0.78) and as compared in ANOVA not significantly different (P=0.01). However, upon analysis by species, MPI was the least polymorphic marker for *L. guyanensis* (P<0.001) and G6PD was the least polymorphic locus for *L. lainsoni* (P<0.001). Among the three *L. shawi* isolates, ICD was the only polymorphic marker (Figure 1).

The Simpson index of diversity (D), which provides an indication of the discriminatory capacity of each marker, was similar for MPI, G6PD and 6PGD, (0.77, 0.77 and 0.78 respectively), but higher for ICD (D=0.89). Analyses of the discriminatory capacity indicate a high level of strain discrimination, of almost 90%, using these four loci. However, higher values were observed when applied to *L. braziliensis* and *L. guyanensis* (96% and 100%, respectively). Other species were represented by few strains and were not analyzed. This suggests that, even though the number of markers is smaller than that usually used in MLST (7), these four markers are sufficient for studies in *L. (Viannia)*. However, more detailed population genetics studies may require more markers. Regarding the discriminatory power of each marker, the Simpson index showed that ICD had the highest diversity. Indeed, this is a very polymorphic locus in MLEE, which is able to detect intra-species variation [19].

MPI was here found to be a good marker to distinguish between the species, although HP8 was shared among *L. braziliensis* and one *L. guyanensis* strain. This enzyme is used as a marker to differentiate *L. peruviana* from *L. braziliensis*, which was supported by DNA sequencing in a previous study [14] by the detection of a specific SNP, as well as others useful to differentiate closely related *L. (Viannia)* species. We could not confirm this, given that our sequences were shorter and did not include that SNP locus.

Maximum parsimony-based haplotype networks built for each gene (Figure 2) showed that species, as identified by MLEE, clearly formed separated clusters in each gene network, with a few exceptions confined to some strains that for some markers were not grouped in their species haplogroups. Reticulate patterns were observed in some clusters for all loci studied. Even with markers
for which \textit{L. shawi} presented different haplotypes from \textit{L. guyanensis}, these two species always clustered close together.

Haplogroups were, in general, consistent with the species (color-coded nodes), although exceptions occurred for all markers: \textit{G6PD} (H4 and H6; Figure 2A); \textit{6PGD} (H1, Figure 2B); \textit{MPI} (H7, H8 and H19; Figure 2C) and in the ICD network (H9, H22, H23, H24; Figure 2D). A single haplotype MPI (H19) comprising an \textit{L. braziliensis} strain (IOCL 2541; \textit{L. braziliensis} from Pernambuco) was at the base of the \textit{L. lainsoni} cluster, but for all the other markers this strain clustered within the \textit{L. braziliensis} species group. One \textit{L. lainsoni} strain (IOCL 2500 from Acre) was part of the \textit{L. braziliensis} cluster for ICD (H24) and \textit{6PGD} (H1). Conversely, the \textit{L. lainsoni} cluster included two haplotypes exclusively from \textit{L. braziliensis} strains (haplotypes H22 and H23, IOCL 2498 and IOCL 2499 respectively, from Acre), even though both clusters were located in opposite sides of the network.

The most frequent haplotypes (node size) were often the founding haplotype for a given haplogroup, as clearly observed for \textit{L. braziliensis} (\textit{MPI} H7, ICD H6, \textit{G6PD} H6, \textit{6PGD} H1 and \textit{L. gyanenens} (\textit{MPI} H9, ICD H9, \textit{G6PD} H4, \textit{6PGD} H9). Eighteen out of 55 \textit{L. braziliensis} strains were frequently observed in the most common haplotypes, but for \textit{L. gyanenens} the composition of the most common haplotypes was different between the markers. The most frequent haplotypes were also usually those in which inter-specific sharing of sequences was observed (Figure 2).

\textit{L. gyanenens} formed a diverse cluster, whereas \textit{L. shawi} strains presented a profile coherent with a subpopulation of the \textit{L. gyanenens} group for all markers, commonly sharing the most common \textit{L. gyanenens} haplotype or differing from it in at most two polymorphisms (Figure 2C).

Although we used a relevant number of strains, the data analysis by Network software generated median vectors. The presence of median vectors in the networks might indicate that: i) intermediate haplotypes were present in lost populations; ii) haplotypes from populations that not included in this analysis; iii) the ancestors of these strains suffered rapid adaptive evolution with expansion of these extant strains. Therefore, even without sequencing strains representing all genetic diversity, statistical tools may predict the variability.


eighbour Joining (NJ) trees were built for each marker to evaluate the phylogenetic relationship between the haplotypes. Almost no incongruence was observed between the markers. Four monophyletic groups were clearly observed for each marker, representing basically \textit{L. lainsoni} and \textit{L. naffi}, the most divergent groups, and \textit{L. braziliensis} and \textit{L. gyanenens} (in this case, including \textit{L. shawi}), which were very closely related for any marker (data not shown), corroborating the genetic distance MLEE-based tree [19]. \textit{L. lindenberghi} (except in \textit{6PGD}) and \textit{L. utingensis} were each in a separate and independent branch, and grouping closer to \textit{L. naffi} and \textit{L. gyanenens} respectively.

Available sequences of \textit{L. peruviana} for \textit{G6PD}, \textit{6PGD} and \textit{MPI}, and for \textit{L. panamensis} for \textit{G6PD} and \textit{MPI}, were included in the respective network constructions. \textit{L. peruviana} sequences either presented the most frequent haplotype for \textit{L. braziliensis} strains (in \textit{G6PD} H6, \textit{6PGD} H1 MPI H7; Figure 2) or differed from it in one polymorphic site (in the MPI H26 and H27; Figure 2C; Table S2). A previous study using random amplification of polymorphic DNA (RAPD) and MLEE [31] reported that \textit{L. peruviana} and \textit{L. braziliensis} corresponded to two closely related, but distinct monophyletic lines, which was not corroborated by \textit{hop70} gene sequence analysis [32]. \textit{L. panamensis} sequences presented new haplotypes (\textit{G6PD} H36 and \textit{6PGD} H36) within the \textit{L. gyanenens} and \textit{L. shawi} haplogroup. Previously [33] MLEE and RAPD analysis questioned the distinction between \textit{L. panamensis} and \textit{L. gyanenens}, since data did not indicate distinct monophyletic lines.

### Concatenated NeighborNet confirms species-specific clusters and suggests relatively frequently recombination occurring in \textit{L. braziliensis} and \textit{L. gyanenens}

Among the 96 \textit{(Viannia)} strains, 75 final diploid sequence types (DSTs) were assigned. Although we detected a high number of DSTs, many DSTs are unique, while others are more prevalent, widely distributed and presenting temporal stability, which might reflect limited genetic recombination involving these DSTs [17].

The only species with strains sharing the same DST was \textit{L. braziliensis}, DST12, for example, was found in 18 strains of \textit{L. braziliensis} from different Brazilian endemic regions related to the Atlantic rain forest, except one, and present zymodeme diversity although most are classified in zymodeme 27 (Table S1). DST12 is not only the most frequent but also shows temporal stability, as the strains included in this DST had been isolated between 1987 and 2001. Strains typed as DST12 were isolated from patients presenting distinct clinical manifestations. This raises the intriguing proposition that the apparent dominance of DST12 in endemic locations associated with urban areas of the Atlantic rain forest region may be a consequence of higher fitness of this DST to the modified environment.

Three other \textit{L. braziliensis} DSTs (16, 27, and 35) comprised two or three strains. All the other \textit{L. braziliensis} were assigned to unique DSTs (n = 30). DST16 comprised two heterozygous \textit{L. braziliensis} strains from Peru, whilst DST27 comprised two strains (zymodemes 27 and 74) from different localities (Pernambuco and Bahia, respectively), and DST35 three strains from the same zymodeme (IOC/Z27) and geographic origin (Bahia, Northern Brazil).

This demonstrates that the MLSA approach allows both detection of different genotypes and the level of separation between strains through the number of polymorphic sites.

A NeighborNet network was obtained with the concatenated sequences represented by the DSTs (Figure 3). The clusters were in agreement with MLEE for species groups, with the exception of \textit{L. shawi}, which clusters together with all \textit{L. gyanenens} strains. The reticulate aspects of the \textit{L. gyanenens} group suggest recombination events occurring among the strains, including \textit{L. shawi} (Figure 3).

This same aspect was observed for the \textit{L. braziliensis} cluster, but two strains were more divergent (IOC/L2498 and IOC/L2499, DSTs 32 and 57). Both were isolated from Acre state, a region bordering Peru. Recently we have demonstrated that these two strains clustered together with \textit{L. peruviana} and \textit{Peruviana} \textit{L. braziliensis} by microsatellite typing [29]. However, upon removal of ICD sequences from this analysis these two strains grouped very closely to the other \textit{L. braziliensis} strains (data not shown).

It is clear that \textit{L. braziliensis}, \textit{L. gyanenens}, \textit{L. naffi} and \textit{L. lainsoni} all represent distinct species, forming monophyletic groups in the NeighborNet. \textit{L. lindenberghi} and \textit{L. utingensis} were placed close to the monophyletic groups corresponding to \textit{L. naffi} and \textit{L. gyanenens} respectively, corroborating MLEE and MLMT data (unpublished). More isolates from both species should be studied to infer their taxonomic status. However, it is important to mention that these two species shared no alleles with all the other species/strains, except for \textit{L. lindenberghi}, which shared alleles in 6PGD with
L. braziliensis (Figure 2B). The estimate of the average genetic distance between L. lindenbergi or L. utingensis and the other L. (Viannia) species is comparable to those observed between each of the species analyzed. The highest values were observed between L. lainsoni and any other species.

Corroborating the results observed for the haplotype network, L. naiffi and L. lainsoni were the most divergent and the large splits observed suggesting no influence of genetic exchange. DST58, represented by a single L. lainsoni strain was positioned in a split between L. lainsoni and L. naiffi. The extensive reticulation suggests that recombination has occurred relatively frequently in L. braziliensis and L. guyanensis, as proposed by other authors [34]. As expected, L. braziliensis and L. naiffi presented higher within group average genetic distance values than L. guyanensis, even including L. shawi strains in the L. guyanensis group, corroborating previous studies on the genetic diversity of both L. braziliensis and L. naiffi [35,36,37,38]. However, as far as we know, this is the first report of a high level of genetic diversity within L. lainsoni. This was the species giving the highest within group average genetic distance value, although it consists of only two zymodemes. Reports on the occurrence of L. lainsoni and L. naiffi in distinct endemic areas have constantly increased, indicating that both species are adapted to several environments where different sand fly species participate in parasite transmission. Notably, L. lainsoni was the most divergent species, followed by L. naiffi, which might contribute to the widespread dispersion of these two species [39–41].

**BURST analysis contributes to the understanding of relatedness among DSTs**

Considering that a large number of unique DSTs were detected, the BURST algorithm was employed to evaluate the relationships between 52 homozygous DSTs, which corresponded to 70 strains. Here we applied eBURST to analyze the allele profiles as an efficient method to recognize natural discontinuities that would merit taxonomic status as species or subspecies. The BURST algorithm first identifies mutually exclusive groups of related genotypes in the population (typically an MLST database), and attempts to identify the founding genotype (sequence type, ST) of each group. The algorithm then predicts the descent from the predicted founding genotype to the other genotypes in the group, displaying the output as a radial diagram, centered on the predicted founding genotype. The procedure was developed for use with the data produced by MLST (STs and their allelic profiles).

The clonal complex model has proved to be valuable in analyzing many MLST datasets, making the results more amenable for epidemiological analysis. A clonal complex (CC) comprises genetically related, but not identical, organisms. In this study six CCs were observed (Figure 4); each one represented by strains classified as the same Leishmania species. Strains typed as L. braziliensis were grouped in two clonal complexes. The CCs observed were: CC1, L. lainsoni (comprising 2 of 6 strains; 2/6); CC2, L. naiffi (3/5); CC3, L. shawi (3/3); CC4, L. braziliensis (30/37); CC5, L. braziliensis (2/37); CC6, L. guyanensis (13/17). CC6 included 13 L. guyanensis DSTs from 17 analyzed, indicating DST37 as the putative founder genotype. CC4 included 12 out of 19 L. braziliensis DSTs analyzed and the predicted founder genotype was DST12, the most common DST observed for this species.

Most of the isolates were not grouped in any clonal complex, showing a high prevalence of singletons (n = 15; 4 L. lainsoni, 2 L. naiffi, 5 L. braziliensis, 4 L. guyanensis), differing at two or more loci from all other DSTs (Table S1), typical of populations with a high rate of recombination relative to mutation and for which eBURST does not reliably indicate ancestry [42]. Although eBURST does not contribute to phylogenetic analysis in Leishmania, this is a helpful clustering tool to analyze MLST results from this organism, which minimizes the differences observed between each Leishmania strain. For example, although all L. guyanensis strains were assigned to a distinct DST, almost all of them clustered in the same clonal group (CC5), corroborating the already demonstrated homogeneity of the L. guyanensis population in Brazil [19] and contrasting with the results with microsatellites showing diversity within the L. guyanensis complex [34]. We cannot infer the dispersion capacity of this group, but temporal stability is certainly a characteristic of this clonal complex.

Another scenario was observed for L. braziliensis. Out of 55 L. braziliensis, we obtained groups of 18, 3, 2 and 2 strains that were assigned to the same DSTs. However, two clonal complexes were observed for this species, one represented by 57.5% of the strains analyzed (CC4), the other one by only two strains (CC5) and 27.5% representing singletons. This result would fit a hypothesis of L. braziliensis having better fitness than L. guyanensis, leading to an improved dispersion. However, more L. guyanensis isolates from other regions not located in Brazil and its ‘sister’ species, L. panamensis, would benefit this discussion.

Although the main purpose of this study was not to investigate the epidemiological findings from MLSA of L. (Viannia), it is important to mention some aspects. The capacity of dispersion and adaptation of L. braziliensis clonal complex CC4 is evident as well as its temporal stability. Although formed by only two L. braziliensis strains, CC5 represents an interesting clonal complex. The two strains were isolated from the same locality, a municipality in Pernambuco state, in northeastern Brazil, where a unique zymodeme is found. Furthermore, it is interesting that a high level of genetic diversity was already demonstrated for the L. braziliensis population in Pernambuco, depicted by MLEE analysis. It seems that in this region two transmission cycles co-exist; involving or not involving the sylvatic environment [35,43]. This could influence or favor the maintenance of two clonal complexes in the area.

The putative founder for the clonal cluster CC4 (L. braziliensis) and CC6 (L. guyanensis) were determined. However, it is not possible to infer if they represent the ancestor of each group. In fact, for L. braziliensis CC4 the putative founder is the most prevalent DST, but for L. guyanensis this is not true as for this species each strain was assigned as one DST.

To evaluate the robustness of each CC and the relationship between them, a NJ tree was built using the concatenated sequences of DSTs included by eBURST in clonal complexes, excluding the singletons (Figure 5). Clusters with high bootstrap supports were observed, corresponding to each CC. DST24 and DST30, from CC5, were the only two DSTs inside the L. braziliensis cluster forming a group supported by the bootstrap value. Inside the L. guyanensis group a high supported cluster was
observed composed of all CC3 DSTs, corresponding to L. shawi. Low bootstrap values were observed among almost all DSTs from CC4 and CC6, corroborating the fact that they belong to the same complex. Evidence was again found for the divergence of L. lainsoni and L. naiffi clonal complexes.

Concluding remarks
The results obtained here strongly supported the established taxonomy of L. (Viannia), considering the species that have been found in circulation in Brazil. Specifically, our data support monophyly of all but one Brazilian L. (Viannia) species analyzed here and highlight the close relationships between L. braziliensis and L. guyanensis and the recombination events occurring in both species. Some aspects merit special mention, however. The taxonomic validity of L. shawi has been questioned and indeed the markers studied here suggested that L. shawi isolates were closely related to or were part of the L. guyanensis group. The same was observed in the NeighborNet network with concatenated sequences. However, the eBURST analysis indicated that these isolates form a distinct clonal complex from L. guyanensis, although closely related as observed in the NeighborNet network. Recently, hsp70 gene sequence analysis has indicated that L. shawi is a subgroup of L. guyanensis [5]. In addition, L. shawi and L. guyanensis had the lowest average genetic distance of the L. (Viannia) studied here. Our results, thus, indicate that L. guyanensis has different clonal populations, such as those observed for L. braziliensis, with L. shawi being one of them.

It is noteworthy that the MLSA clades derived here are in good agreement with MLEE clusters reported previously. Taken together, our data point to the combined four gene scheme used here as a reasonable approach that provides extensive differentiation and offers evolutionarily accurate clustering. The analysis performed herein should be extended to species other than those studied here and should be used as a starting point to develop an MLST scheme for Leishmania spp. genetic typing. While providing complete genome sequences is not possible as a routine approach, MLST generates evidence for similarities and differences between Leishmania species and/or strains, offering a number of advantages over most typing methods and providing results helpful for taxonomic, population genetics, evolutionary and, in consequence, epidemiological studies [4].

Such data are likely to revolutionize the systematics of Leishmania, consolidate our view of what constitutes a Leishmania ‘species’, provide evidence concerning the epidemiology of these
Multilocus Analysis of Leishmania Species

L. guyanensis

L. shawi

L. braziliensis

L. naiffi

L. lainsoni
pathogens and might even be applicable directly to clinical samples.

Definition of Leishmania species and knowledge of the genetic structure of all Leishmania species will provide a useful framework for exploring the evolutionary dynamics and phylogenetic distribution of relevant strain properties. Recognizing the urgent need for a standardized globally acceptable species definition and typing method for Leishmania, we are now sequencing other genes and including more species and strains in our analysis aiming to propose that species within the Leishmania genus could be defined as a group of strains that share a determined level of similarity in the concatenated nucleotide sequences of the genes selected. To achieve this, establishment of a consensus MLST gene set that provides optimum differentiation for Leishmania species and/or strains is required.

Supporting Information

Table S1 Taxonomic and collection data (clinical, geographical, biochemical and molecular Information) for L. (Viannia) strains used for MLSA.

Table S2 Sequences retrieved from GenBank and those included in the analysis performed in the present study.

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