High fat diet enhances cardiac abnormalities in SHR rats: Protective role of heme oxygenase-adiponectin axis

Jian Cao1†, Komal Sodhi2†, Nitin Puri2, Sumit R Monu2, Rita Rezzani3 and Nader G Abraham2*

Abstract

Background: High dietary fat intake is a major risk factor for development of cardiovascular and metabolic dysfunction including obesity, cardiomyopathy and hypertension.

Methods: The present study was designed to examine effect of high fat (HF) diet on cardio-vascular structure and function in spontaneously hypertensive rats (SHR), fed HF diet for 15 weeks, a phenotype designed to mimic metabolic syndrome.

Results: Development of metabolic syndrome like phenotype was confirmed using parameters, including body weight, total cholesterol and blood pressure levels. High fat diet impaired vascular relaxation by acetylcholine and exacerbated cardiac dysfunction in SHRs as evidenced by lower left ventricular function, and higher coronary resistance (CR) as compared to controls (p < 0.05). The histological examination revealed significant myocardial and peri-vascular fibrosis in hearts from SHRs on HF diet. This cardiac dysfunction was associated with increased levels of inflammatory cytokines, COX-2, NOX-2, TxB2 expression and increase in superoxide (O2-) levels in SHR fed a HF diet (p < 0.05). HO-1 induction via cobalt-protoporphyrin (CoPP, 3 mg/kg), in HF fed rats, not only improved cardiac performance parameters, but also prevented myocardial and perivascular fibrosis. These effects of CoPP were accompanied by enhanced levels of cardiac adiponectin levels, pAMPK, peNOS and iNOS expression; otherwise significantly attenuated (p < 0.05) in HF fed SHRs. Prevention of such beneficial effects of CoPP by the concurrent administration of the HO inhibitor stannic mesoporphyrin (SnMP) corroborates the role of HO system in mediating such effects.

Conclusion: In conclusion, this novel study demonstrates that up-regulation of HO-1 improves cardiac and vascular dysfunction by blunting oxidative stress, COX-2 levels and increasing adiponectin levels in hypertensive rats on HF diet.

Keywords: heme oxygenase, adiponectin, high fat diet, COX-2, oxidative stress

Background

Obesity and hypertension are two major risk factors that lead to increased incidence of cardiac diseases including coronary artery disease, heart failure and cardiomyopathy [1-3]. Blood pressure, which strongly correlates with body mass index, is one of the most important determinants of cardiovascular function [4]. In addition, obesity also leads to abnormal cardiac function through mechanisms that are independent of hypertension [5,6]. Metabolic syndrome is a clinico-pathological condition which entails superimposition of these abnormalities and is characterized by systemic inflammation and oxidative stress [3,7] A combination of these risk factors leads to disruption of metabolic homeostasis and may further contribute towards progressive cardiovascular dysfunction.

The heme-HO system, comprising of HO-1 (inducible) and HO-2 (constitutive) isoforms, is one of the key defense mechanisms against oxidative stress [8]. This

© 2011 Cao et al; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.
The beneficial role of HO enzyme system in animal models of obesity and hypertension are clearly defined but paucity of evidence exists regarding similar effects in co-morbid conditions such as hypertension and obesity. In light of this evidence, the aim of this novel study was to explore the potential effect of HO-1 induction in spontaneously hypertensive rats (SHR) fed a high fat diet, a phenotype designed to mimic metabolic syndrome. We tested our hypothesis by using a well-described high fat regimen [28] that does not cause atherosclerotic lesion formation in mice [29], to address the effects of a known HO-1 inducer, cobalt protoporphyrin (CoPP). To verify that the effects of CoPP were due to an increase in HO-activity, we also treated a group of SHR concurrently with stannous mesoporphyrin (SnMP) to inhibit HO activity. Our results show that obesity exacerbates myocardial and vascular damage in SHRs, and HO-1 induction improves heart function in parallel with increased adiponectin levels and reduced expression of myocardial pro-inflammatory enzymes such as COX-2 and iNOS. Thus, HO-1 appears to play a critical role in the cellular defense against obesity-induced cardiovascular dysfunction in a hypertensive animal model fed a high fat diet. These findings may have important clinical implications in the management of patients with metabolic syndrome.

Methods

Animal treatment
All animal studies were approved by the New York Medical College Animal Care and Use Committee in accordance with the National Institutes of Health Guidelines for Care and Use of Laboratory Animals. Fifty-eight seven-week-old male SHRs were purchased from Charles River Laboratories and were divided into four groups: A) SHR control, B) SHR-fat, C) SHR-fat and CoPP treatment, D) SHR-fat and CoPP and SnMP treatment. SHR rats were fed ad libitum either with a normal diet (group A) containing 11% fat, 62% carbohydrate, and 27.0% protein total, 12.6 KJ/g or a high fat diet (groups B, C, D) containing 58% fat from lard, 25.6% carbohydrate, and 16.4% protein yielding 23.4 KJ/g (Bio-SERV, Frenchtown, NJ) for 15 weeks [28,30]. The diet used is distinct from the so-called “Western” or “atherosclerotic” diet which contains, in addition to high fat, cholesterol and bile acids. While the high fat diet used in the present study results in obesity, it does not cause atherosclerotic lesion formation in mice [29]. After 4 weeks of high fat diet, cobalt protoporphyrin (CoPP), an inducer of HO-1, was administered intraperitoneally once a week (3 mg/kg) for 11 weeks to SHR rats maintained on a high fat diet. Some of the SHR treated with CoPP were concurrently treated with tin mesoporphyrin IX dichloride (SnMP), to inhibit HO activity, which was administered intraperitoneally three times a week (20 mg/kg) [11] to ascertain that any effects of CoPP treatment were related to increased HO activity. The untreated SHR rats maintained on the high fat diet were administered the vehicle for CoPP and SnMP once a week and 3 times a week respectively (0.1 mM sodium citrate buffer pH 7.8) for 11 weeks.

Rats were weighed every 7 days and systolic blood pressure was determined weekly by the tail-cuff method. After a 6-hour fast, rats were anesthetized with sodium pentobarbital (65 mg/kg, i.p.) and blood was obtained from a tail vein for glucose measurement using a glucometer (Lifescan Inc., Miligitas, CA). Blood samples were then collected and stored as previously described [15].

Isolated Heart Preparation
Three days after the last CoPP (or vehicle) injection, rats were anesthetized with pentobarbital, i.p., and hepариized via the left femoral vein (250 units/kg). The heart was rapidly excised, placed in cold perfusion medium and weighed. The isolated hearts were attached to the Langendorff apparatus and retrogradely perfused (at 37°C) using constant perfusion pressure of 80 cm H2O, then perfusion pressure was decreased to 20 mmHg for 30 min, and then pressure was increased back to 80
mmHg for the remaining 30 min (reperfusion) [29]. The perfusion medium consisted of oxygenated Krebs-Henseleit buffer [31,32]. For measurement of ventricular systolic and end diastolic pressure (EDP), latex balloons were inserted into the left ventricle of the heart through the mitral valve and connected to a Harvard pressure transducer. In each experiment EDP was set at 10 mmHg and kept stable during the first 30 minutes of perfusion. Coronary perfusion pressure (CPP) was monitored by a second pressure transducer connected to the aortic cannula. Left ventricular developed pressure (LVDevP), EDP, dP/dTmax and dP/dTmin were all derived or calculated from the continuous monitoring of the LV pressure signal. In all experiments, coronary flow was continuously monitored by collecting the cardiac effluent. Coronary resistance (CR) was defined as input pressure divided by coronary flow per gram of myocardial tissue (mmHg x minxg/mL). At the end of each experiment, hearts were collected, half were used for histology examination and half of them were rapidly frozen in liquid nitrogen and stored at -80°C.

Assessment of Myocyte Cross-Sectional Area, Myocardial Fibrosis and Collagen in Myocardial Tissue
Hearts were fixed in 10% buffered formalin, and embedded in paraffin wax and sectioned to 5 μm. For measurement of the cross-sectional area, 100 cells (per animal) from the left ventricular wall were randomly chosen and analyzed in hematoxylin staining. The myocyte cross-sectional area and myocardial fibrosis were quantitatively analyzed with Image Pro-Plus 4.5.1 software in digitalized microscopic images. Myocardial fibrosis in the tissue sections was quantitatively analyzed by morphometry in 2 ways: (1) on the perivascular fibrosis, and (2) on myocardial tissue (total fibrosis index). The collagen in myocardial tissue was visualized by Sirius Red staining under polarization microscopy and then quantified.

Assessment of Vascular Reactivity
The aorta was removed, cleaned of fat and loose connective tissue, placed in cold Krebs-bicarbonate solution, and sectioned into 3-mm-long rings. Vasoactivity responses of phenylephrine-constricted arteries to cumulative increments in acetylcholine (10⁻⁹ to 10⁻⁴ mol/L) were examined in the presence of indomethacin (10 μmol/L) as described [33].

Western Blot Analysis of Cardiac Tissue for protein expression
At the time of sacrifice, hearts were harvested, and stored at -140°C. Frozen hearts were pulverized under liquid nitrogen and placed in a homogenization buffer prior to immunoblotting with antibodies against HO-1, and HO-2 (Stressgen Biotechnologies Corp., Victoria, BC), COX-2, TX synthase, NOX-2, AKT, AMPK, pAMPK(Thr172), pAKT and adiponectin (Cell Signaling Technology, Inc., Beverly, MA) and eNOS, peNOS(serine 1177), and iNOS (Santa Cruz Biotechnology, Santa Cruz, CA). Immunoblotting was performed in cardiac tissue as previously described [15,33].

Measurement of HO activity
HO activity in heart tissue was assayed as described previously [15] using a technique in which bilirubin, the end product of heme degradation, was extracted with chloroform, and its concentration was determined spectrophotometrically (dual UV-visible beam spectrophotometer Lambda 25; PerkinElmer Life and Analytical Sciences, Waltham, MA) using the difference in absorbance at a wavelength from 460 to 530 nm, with an extinction coefficient of 40 mM⁻¹ cm⁻¹.

Measurements of O₂⁻ production and total cholesterol levels
Total cholesterol was measured in serum using a cholesterol Quantification Kit (Biovision, Mountainview, CA) according to the manufacturer’s instructions. For the detection of O₂⁻, homogenized hearts were placed in plastic scintillation vials containing 5 μmol/l lucigenin in a final volume of 1 ml of air-equilibrated Krebs solution as described previously [15].

Plasma Adiponectin and inflammatory cytokines Measurements
The high molecular weight (HMW) HMW form of adiponectin, IL-6, TNF-α and TXB2 levels were determined using an ELISA assay (Pierce Biotechnology, Inc., Woburn, MA) as described previously [15].

Statistical Analysis
The data are presented as mean ± standard error (SEM) where n = 6/group for the results. For comparison between treatment groups, the Null hypothesis was tested by a single factor analysis of variance (ANOVA) for multiple groups or unpaired t-test for two groups. Statistical significance (p < 0.05) between the experimental groups was determined by the Fisher method of analysis for multiple comparisons.

Results
Effect of a high-fat diet on body weight and metabolic response
Figure 1A shows the percent change in body weight over its baseline values in the 4 groups. In untreated SHR rats body weight increased 54% ± 5.5 on a normal diet over a period of 15 weeks, whereas in rats fed a high fat diet body weight increased 79% ± 3.7 (p < 0.05).
The total body weight observed after 15 weeks of study was 367 ± 10.7 gms in SHR controls and 419 ± 6.3 gms in SHR rats fed a high fat diet (data not shown). We also examined the effect of long-term CoPP treatment on body weight gain in response to a high fat diet. Weekly treatment with CoPP was started 4 weeks after the initiation of the high fat diet and was well tolerated by the SHR (n = 14/group); activity and grooming were maintained during CoPP treatment. Rats fed a high fat diet and concurrently exposed to CoPP, showed reduction in body weight as compared to SHR rats on high fat diet, 68% ± 2.4 (p < 0.05). A significant increase in body weight was seen when animals fed a high fat diet were exposed to CoPP + SnMP. The weight gain was 75% ± 4.9 and was not significantly different from animals fed a high fat diet. The total body weight observed after 15 weeks of study in rats fed a high fat diet and concurrently exposed to CoPP was 386 ± 9.7 gms and was increased to 416 ± 8.1 gms in SHR rats fed a high fat diet and treated with CoPP and SnMP (data not shown).

Systolic blood pressure was increased over the 15-week period in SHR rats (Figure 1B; n = 6/group). The systolic blood pressure was 175 ± 11 mmHg in the SHR control and was significantly increased in the rats fed a high fat diet, 211 ± 9 mmHg (p < 0.05). The elevation in systolic pressure was attenuated by CoPP treatment in SHR fed a high fat diet whereas SnMP treatment nulled the antihypertensive effect of CoPP in SHR fed a high-fat diet (Figure 1B). The mean blood glucose level in the SHR rats maintained on a normal diet was 128 ± 4 mg/dl, and was increased to 173 ± 14 mg/dl by a high fat diet (p < 0.05; n = 6/group) (data not shown). This increase in blood glucose levels was significantly attenuated by CoPP treatment in SHR rats fed a high fat diet (137 ± 4.5 mg/dl) and this effect was reversed by treatment with SnMP (180 ± 7.8 mg/dl) (data not shown).

Plasma cholesterol levels remained elevated in SHRs fed a high-fat diet as compared to their controls. Plasma cholesterol levels were 0.55 ± 0.11 in SHRs fed a normal diet for 15 weeks, and levels were increased to 1.25 ± 0.15 mg/dL by 15 weeks on the high-fat diet (P < 0.05).
CoPP treatment prevented the increase in cholesterol levels in SHR while concomitant treatment with SnMP blocked the effect of CoPP.

Effect of high fat diet on cardiac parameters

The collagen III was higher in hearts of SHRs fed a high fat diet ($p < 0.05$) when compared to untreated animals (Figure 2A). The perivascular fibrosis index was higher in SHRs fed a high fat diet than those animals fed a normal diet ($p < 0.05$) (Figure 2B). CoPP administration prevented the occurrence of these increases in animals fed a high fat diet on perivascular fibrosis while concurrent administration of SnMP did not significantly reversed the effect of CoPP (Figure 2B). The myocyte cross-sectional area was increased by a high fat diet in SHRs. CoPP treatment prevented the increase in myocyte cross-sectional area while concurrent administration of SnMP did not significantly reversed the effect of CoPP (Figure 2C).

Effect of high fat diet on CR and cardiac function during ischemia/reperfusion

Our results show that during low perfusion pressure (i.e. ischemia), CR increased over baseline values in all groups, but CR in SHR mice was significantly higher than in controls ($p < 0.05$) (Figure 3A). This phenomenon, defined as ‘paradoxical vasoconstriction’, has been described previously by our group in both control and diabetic animals [34]. CoPP modulated coronary tone during the ischemic period significantly reducing vasoconstriction. After 30 min of reperfusion, CR was still significantly increased over baseline values in high fat hearts ($p < 0.05$), while CR in High fat CoPP group returned to baseline values (Figure 3A). The CoPP-
“normalization” of coronary tone at reperfusion in HF hearts was mirrored by better overall cardiac function during both low pressure ischemia and reperfusion times. Indeed, LVDevP (Figure 3B), dP/dtmax (Figure 3C) and dP/dtmin (Figure 3D) were all significantly improved compared to the untreated group (p < 0.05).

Effect of high fat diet on Vascular Reactivity and superoxide levels
Aortic endothelial dilatory responses to acetylcholine (at concentration of $10^{-5}$ and $10^{-4}$ mmol/L respectively) were significantly impaired in SHRs after 15 weeks of a high-fat diet compared with those fed a normal diet ($P < 0.05$) (Figure 4A). Endothelial function was improved in SHRs as a result of the CoPP treatment ($P < 0.05$), but exacerbated by SnMP (Figure 4A) indicating that it is specifically the endothelial dilatory response that is impaired by a high fat diet in this animal model. Cardiac oxidative stress was increased as cortical superoxide generation was greater in SHR fed a high fat diet compared with rats fed a normal diet (n = 6/group), ($P < 0.05$). CoPP treatment prevented the increase in cardiac O$_2^-$ generation in SHR maintained on a high fat diet ($P < 0.01$), an effect abolished by concurrent administration of SnMP.

Effect of high fat diet on plasma adiponectin, inflammatory cytokines and TxB2 Levels
Plasma IL-6 and TNF-α (Figure 5A and 5B) levels were greater in SHR fed a high fat diet compared to rats fed a normal diet (n = 6/group), ($P < 0.05$). Increasing HO-1 by CoPP administration significantly decreased plasma cytokines and this effect was prevented by concurrent SnMP treatment ($P < 0.01$, Figure 5A and 5B). Similar
pattern was observed in plasma TxB2 levels as shown in Figure 5C (n = 6/group), (p < 0.05). Plasma adiponectin levels were lower in rats fed a high fat diet when compared to control animals fed a normal diet (p < 0.05; n = 6/group) (Figure 5D). This effect was reversed when rats were treated with CoPP (p < 0.05). Indeed, in SHR rats maintained on a high-fat diet and treated with CoPP, plasma adiponectin levels were higher than those in the respective control groups (p < 0.05). Concurrent administration of SnMP with CoPP in the SHR fed a high fat diet prevented the increase in adiponectin, so that the levels of this protein were not different from those in the untreated SHR.

Effect of high fat diet on Cardiac COX-2, TxA2 and NOX-2 Levels
Hearts isolated from SHRs fed a high fat diet showed a significant increase in markers of oxidative stress compared to animals fed a normal diet (p < 0.05, respectively) (Figures 6A, B and 6C). Treatment with CoPP resulted in a decrease in COX-2, TxA2 and NOX-2 expression in SHRs fed a high fat diet (p < 0.01 respectively), an effect abolished by concurrent administration of SnMP.

Effect of high fat diet on cardiac HO-1
First, we confirmed that CoPP treatment for 11 weeks resulted in up-regulation of HO-1. HO-1 protein in the hearts of SHR fed a high fat diet was significantly less than that of the respective control group (Figure 7A where n = 6/group) when the latter was fed a normal diet (p < 0.05). Treatment with CoPP resulted in a significant increase in HO-1 levels in SHR fed a high-fat diet. Although SnMP treatment showed a significant increase in HO-1 expression (Figure 7A), it is a potent inhibitor of HO activity as shown previously [11,35] and thus prevents heme degradation and inhibits formation of CO and biliverdin. HO-2 levels were unaffected either by high fat diet or by CoPP treatment (Figure 7A). Consistent with protein expression, HO activity was significantly decreased in obese SHR hearts compared to the control group (Figure 7B). CoPP treatment significantly increased HO activity in SHR fed a high fat diet, 1.45 ± 0.20 nmol bilirubin/mg/hr compared to 0.39±0.09 nmol bilirubin/mg/hr in untreated SHR fed a high fat diet (p < 0.001). The concurrent administration of SnMP resulted in significant decrease of HO activity as shown in Figure 7B.

Effect of high fat diet on Cardiac adiponectin, pAMPK and pAKT Expression
Cardiac adiponectin levels, normalized against β-actin, exhibited a similar pattern to plasma adiponectin levels. Thus, feeding SHR a high fat diet for 15 weeks resulted in a decrease in adiponectin compared to untreated SHR (Figure 8; n = 6/group). Induction of HO-1 with CoPP increased cardiac adiponectin levels in hypertensive rats (p < 0.01) and the increase in SHR was prevented and reversed to a decrease when the rats were, also, treated with SnMP to inhibit HO activity (Figure 8A).
8). A high fat diet resulted in significant decreases in pAMPK and pAKT expression in hearts from SHR (p < 0.05; n = 6/group) (Figure 8). CoPP administration caused a significant increase in the expression of pAKT and pAMPK in the rats fed a high fat diet (p < 0.05) compared to untreated rats fed a high fat diet. The changes in expression of pAMPK and pAKT paralleled those seen with HO-1 protein expression. In SHR main-
tained on a high fat diet and treated with CoPP, the concurrent administration of SnMP prevented the increase in pAKT and pAMPK; indeed, the expression of both pAKT and pAMPK was reduced to levels lower than those seen in SHR on the high fat diet alone (p < 0.01).

**Effect of high fat diet on Cardiac eNOS, peNOS and iNOS Levels**

Compared to animals fed a normal diet, SHR animals fed a high fat diet exhibited lower levels of eNOS and peNOS protein (p < 0.05) (Figure 8) CoPP administration produced an enhanced expression of eNOS and peNOS protein (p < 0.05 compared to untreated ani-
imals) in SHRs fed a high fat diet (Figure 8). In contrast, SnMP administration resulted in eNOS and peNOS protein in SHRs fed a high fat diet (Figure 8). Hearts iso-
lated from SHRs fed a high fat diet showed a significant increase in iNOS expression compared to animals fed a normal diet (p < 0.05, respectively) (Figures 8). Treatment with CoPP resulted in a decrease in iNOS in SHRs fed a high fat diet (p < 0.0, Figure 8). In contrast, SnMP did not prevent the increase of iNOS expression in SHRs fed a high fat diet (Figures 8).

**Discussion**

The results of the present study demonstrate that SHR fed a high fat diet develop patho-physiological abnormalities similar to that observed in metabolic syndrome. This phenotype is characterized by increased levels of body weight, blood cholesterol and blood pressure along with an accelerated decline in cardiac function when compared to SHR maintained on a normal diet. We, also, demonstrated that cardiac HO-1 induction,
accompanied by increased plasma and tissue adiponectin levels, resulted in the improvement of cardiovascular function as manifested by a decrease in blood pressure, coronary resistance (CR), myocardial fibrosis; and increase in left ventricular function and vascular relaxation, as compared to control. The upregulation of HO-1 was associated with a concomitant decrease in the levels of O$_2^-$, COX-2 and iNOS, markers of oxidative stress. Furthermore, there was a decrease in cardiac remodeling, and an increase in the expression of cardiac pAKT, pAMPK and peNOS via induction of HO-1-adiponectin axis. To the best of our knowledge, this is the first report showing a protective effect of HO-adiponectin axis in a co-morbid condition where a pre-existing cardio-vascular pathology is further aggravated by addition of a HF diet.

High fat intake increased body weight, serum cholesterol and blood pressure in SHR and these changes in metabolic indices were associated with cardiovascular dysfunction in these animals. Previous studies have shown that HO-1 induction decreases obesity, reduces levels of visceral and subcutaneous fat and normalizes the metabolic profile in obese rats and mice [15,17,36,37]. Also HO-1 overexpression is known to improve cardiovascular dysfunction in hypertensive rats [7,11]. In contrast, in the current study we induced a metabolic syndrome-like phenotype in hypertensive animals. SHR demonstrate chronic hypertension, oxidative stress and cardiac damage [38]. All of these parameters were worsened by the addition of high fat diet, strengthening our hypothesis that obesity and the associated metabolic abnormalities accelerate pathological pre-existing cardiovascular changes. Reversal of these pathophysiological abnormalities by HO-1-adiponectin induction corroborates the protective effects of the heme-oxygenase system in such a setting.

Metabolic syndrome-mediated increases in oxidative stress contribute to cardiovascular dysfunction via

![Figure 6 Western blot and densitometry analysis of A) COX-2; B) TXB2; C) NOX-2 in hearts obtained from SHR. Rats were fed a high fat diet and treated with CoPP and CoPP + SnMP. Data are shown as the COX-2/actin ratio, TXB2/actin ratio and NOX-2/actin ratio, respectively. n = 6, *p < 0.05 vs. control, #p < 0.05 vs. HF, †p < 0.05 vs. HF+ CoPP.](image-url)
endothelial cell sloughing and beta cell apoptosis [39]. Sustained increases in O$_2^-$ levels and cytokines, including TNF-$\alpha$ and its receptor, lead to monocyte phenotype transition, myocytic apoptosis, and activation of matrix metalloproteinase. This, in turn, modifies the interstitial matrix, augmenting further ventricular remodeling [40,41]. COX-2 is considered a pro-inflammatory enzyme as free radicals and prostaglandins (PGs) are produced during its catalytic cycle [8]. It has been shown in our previous reports that upregulation of HO-1 decreases vasoconstrictors, such as cyclooxygenase (COX-2), PGs and thromboxane synthases (TxA$_2$) levels [8,42] by regulating the cellular heme levels and ROS. The heme-HO system is a stress response system (reviewed in [8] that undergoes activation under conditions of increased oxidative stress such as those presented here. Induction of HO-1 resulted in decreased cardiac levels of superoxide and NOX-2 expression which may be due to a decrease in the levels of NADPH oxidase [43], a heme-dependent protein, and/or an increase in the levels of superoxide dismutase EC-SOD [44]. Also in the present study, increased cardiac iNOS expression and impaired vascular relaxation in rats fed a high-fat diet was reversed by HO-1 induction which may involve the interplay of one of the various mechanisms including, CO generation, HO-1-induced increase in eNOS expression and increased NO bioavailability due to an increase in cellular antioxidants [37,45-47].

In the present study, a decrease in coronary vascular reactivity manifested by coronary resistance, myocardial fibrosis and cardiac function was found in SHRs fed a high fat diet. The increase in expression of HO-1/adiponectin reverses these deleterious effects with a resultant improvement in energy metabolism and an amelioration of the damaged endothelial and cardiac function seen in SHRs fed a high fat diet. We studied coronary microvascular reactivity and hemodynamics in the isolated, empty, beating heart of SHRs fed a high fat diet. This was prevented in CoPP-treated animals by SnMP suggesting the seminal role of increased HO activity in instigating the changes attributable to increased HO-I expression. This finding highlights the role of the HO system in the preservation of microvascular and cardiac function.

Apart from effects on heme degradation products, HO1 up-regulation was associated with increased cardiac and plasma levels of adiponectin. This causality between HO activity and adiponectin release was strengthened by the inhibitory effects of SnMP on both HO activity and adiponectin levels. It has been recently shown that the beneficial effects of heme-HO system in established cardiovascular-metabolic disorders is mediated, at least in part, via its effect on adiponectin-dependent pathways [15,48,49]. Results presented in the current study support and advance our hypothesis that, in addition to its antioxidant properties, the heme-oxygenase system enhances the adiponectin axis which, in...
turn, modulates multiple physiological processes and may contribute towards HO-mediated attenuation of cardiac dysfunction [17,18,50].

The HO-1-mediated increase in adiponectin was associated with an increase in cardiac pAMPK-pAKT signaling and cross-talk between AMPK and AKT levels appear to correlate with HO-1 and adiponectin levels [16,18,25,51]. This is of particular importance in the setting of myocardial ischemia of SHR rats fed a high fat diet due to the very-high-energy demands and low-energy reserves of the heart. Amplifying signaling through AMPK by HO-1 induction during early reperfusion is beneficial to the injured myocardium due to the ability of AMPK to promote ATP generation [52,53] and to attenuate cardiomyocyte apoptosis [54]. An increase in AMPK-AKT signaling is considered an important metabolic response that is necessary for the attenuation of ROS-mediated cardiac and endothelial dysfunction [55] and both pAMPK and pAKT use eNOS as a substrate and enhance the levels of peNOS [8,56,57]. The results of this study support this link as induction of HO-1-adiponectin axis, also, increased peNOS expression in the heart of SHR. The seminal role of increased HO-1 expression and HO activity in cardiac protection is further strengthened by the results obtained when SnMP was concurrently administered with CoPP; the inhibition of HO activity prevented the beneficial effects of HO-1 induction in obese SHR with regard to blood pressure, adiponectin, pAKT and pAMPK. In summary, these observations support the beneficial role of pharmacogenetic interventions targeted towards HO-1-adiponectin axis in patients with metabolic syndrome. Such patients often exhibit chronic energy imbalance along with a wide array of cardiovascular abnormalities amenable to aggravation by confounding factors such as diet induced obesity. Restoration of metabolic homeostasis by activation of HO-1-adiponectin axis could not only improve the energy profile but also attenuate associated cardiovascular patho-physiological alterations observed in the patients with metabolic syndrome.
Conclusion
In conclusion, the results of the present study demonstrate that upregulation of HO-1 in association with increased levels of adiponectin prevents vascular and cardiac dysfunction in SHR mice fed a high fat diet, a phenotype designed to mimic metabolic syndrome. The pharmacological enhancement of HO-1 expression, resulting in a phenotype resistant to injurious stimuli, permits the heart to initiate a crucial and immediate defense against the events associated with the metabolic syndrome, thereby preventing the continued deterioration in cardiac function associated with this disease.

Acknowledgements
All authors had full access to the data and take responsibility for its integrity. All authors have read and agree with the manuscript as written. We also thank Jennifer Brown for her outstanding editorial assistance in the preparation of the manuscript.

This work was supported by NIH grants DK068134, HL55601 and HL34300 (NGA).

Authors details
1. Department of Geriatric Cardiology, Chinese PLA General Hospital, Beijing 100853, China. 2. Department of Physiology and Pharmacology, College of Medicine, University of Toledo, Toledo, Ohio, 43614, USA. 3. Department of Biomedical Science, Division of Anatomy, University of Brescia, Brescia Italy.

Authors' contributions
*JC and KS contributed equally to this work
JC drafted the manuscript. KS performed all the experiments except vascular function. RRM carried out the morphological studies in heart. NGA conceived the study, and participated in its design and coordination. All authors read and approved the final manuscript.

Competing interests
The authors declare that they have no competing interests.

Received: 11 October 2011 Accepted: 23 December 2011 Published: 23 December 2011

References
1. Zaleski KC, Franklin BA, Miller WM, Peterson ED, McCullough PA. Impact of obesity on cardiovascular disease. Med Clin North Am 2011, 95:919-937.
2. Hall JE: The kidney, hypertension, and obesity. Hypertension 2003, 41:625-633.
3. Knight SF, Quigley JE, Yuan J, Roy SS, Elmarakby A, IMG JD: Endothelial dysfunction and the development of renal injury in spontaneously hypertensive rats fed a high-fat diet. Hypertension 2008, 51:352-359.
4. Garrison RJ, Kannel WB, Stokes J, Castelli WP: Incidence and precursors of hypertension in young adults: the Framingham Offspring Study. Prev Med 1987, 16:235-251.
5. Kenchahsl A, Evans JC, Levy D, Wilson PW, Benjamin EJ, Larson MG, Kannel WB, Vasan RS: Obesity and the risk of heart failure. N Engl J Med 2002, 347:305-313.
6. Mottolino S, Filon KB, Genest J, Joseph L, Pilote L, Poirier P, Rinfret S, Schiffrin EL, Eisenberg MJ: Heme oxygenase-1: a key sensor of cellular energy status. J Am Coll Cardiol 2010, 56:1131-1132.
7. Berg AH, Scherer PE: Adipose tissue, inflammation, and cardiovascular disease. Circ Res 2005, 96:939-949.
8. Abraham NG, Kappas A: Pharmacological and clinical aspects of heme oxygenase. Pharmacol Rev 2008, 60:79-127.
9. Wu L, Wang R: Carbon monoxide: endogenous production, physiological functions, and pharmacological applications. Pharmacol Rev 2005, 57:585-630.
10. Sacchi N, Escalante B, Abraham NG, McGiff JC, Lavery BD: Treatment with tin prevents the development of hypertension in spontaneously hypertensive rats. Science 1989, 243:388-390.
11. Botros FT, Schwartzman ML, Steir CT Jr, Goodman AI, Abraham NG: Increase in heme oxygenase-1 levels ameliorates renovascular hypertension. Kidney Int 2005, 68:2745-2755.
12. Sabaawy HE, Zhang F, Nguyen Y, Elhassene A, Nasjletti A, Schwartzman M, Denney P, Kappas A, Abraham NG: Human heme oxygenase-1 gene transfer lowers blood pressure and promotes growth in spontaneously hypertensive rats. Hypertension 2001, 38:210-215.
13. Buja LM: Myocardial ischemia and reperfusion injury. Cardiovasc Pathol 2005, 14:70-75.
14. Cao J, Inoue K, Li X, Drummond G, Abraham NG: Physiological significance of heme oxygenase in hypertension. Int J Biochem Cell Biol 2009, 41:1025-1033.
15. Li M, Kim DH, Tsenovoy PL, Peterson SJ, Rezzani R, Rodella LF, Aronow WS, Kappas A, Abraham NG: Treatment of obese diabetic mice with a heme oxygenase inducer reduces visceral and subcutaneous adiposity, increases adiponectin levels, and improves insulin sensitivity and glucose tolerance. Diabetologia 2008, 51:1526-1535.
16. Li M, Peterson S, Husney D, Inaba M, Guo K, Terada E, Monta T, Patil K, Kappas A, Ikehara S, Abraham NG: Interdiction of the diabetic state in NOD mice by sustained induction of heme oxygenase: possible role of carbon monoxide and bilirubin. Antioxid Redox Signal 2007, 9:855-863.
17. Kim DH, Burgess AP, Li M, Tsenovoy PL, Addabbo F, McClung JA, Puri N, Abraham NG: Heme oxygenase-mediated increases in adiponectin decrease fat content and inflammatory cytokines, tumor necrosis factor-alpha and interleukin-6 in Zucker rats and reduce adipogenesis in human mesenchymal stem cells. J Pharmacol Exp Ther 2008, 325:833-840.
18. Nicolai A, Li M, Kim DH, Peterson SJ, Vanella L, Postiano V, Gastaldelli A, Rezzani R, Rodella LF, Drummond G, Kusmic C, L’Abbate A, Kappas A, Abraham NG: Heme Oxygenase-1 Induction Remodels Adipose Tissue and Improves Insulin Sensitivity in Obesity-Induced Diabetic Rats. Hypertension 2009, 53:508-515.
19. Iwashita Y, Otsubo S, Ishizuka T, Uchida K, Nitta K: Influence of serum high-molecular-weight and total adiponectin on arteriosclerosis in IgA nephropathy patients. Nephron Clin Pract 2008, 108:e226-e232.
20. Huang KC, Chen CL, Chuang LM, Ho SR, Tai TY, Wang YS: Plasma adiponectin levels and blood pressures in nonobese adolescent females. J Clin Endocrinol Metab 2003, 88:4130-4134.
21. Iwashima Y, Katsuya T, Ishikawa K, Ouchi N, Ohishi M, Sugimoto K, Yu F, Motome M, Yamamoto K, Matsuo A, Ohashi K, Khara S, Funahashi T, Rakugi H, Matsuzawa Y, Ogihara T: Hypoadiponectinemia is an independent risk factor for hypertension. Hypertension 2004, 43:1318-1323.
22. Abraham NG, Kruger A, Peterson S: High serum levels of adiponectin in HO-1-preconditioning in mice and rats with Type 2 diabetes improve vascular function. American Heart Association 2007.
23. Cao J, Drummond G, Inoue K, SoDhi K, Li XY, Omura S: Upregulation of Heme Oxygenase-1 Combined with Increased Adiponectin Lowers Blood Pressure in Diabetic Spontaneously Hypertensive Rats through a Reduction in Endothelial Cell Dysfunction, Apoptosis and Oxidative Stress. J Int Med 2008, 9:2388-2406.
24. Peterson SJ, Drummond G, Kim DH, Li M, Kruger AL, Ikehara S, Abraham NG: L-4F treatment reduces adiposity, increases adiponectin levels and improves insulin sensitivity in obese mice. J Lipid Res 2008, 49:1658-1669.
25. Peterson SJ, Kim DH, Li M, Positano V, Vanella L, Rodella LF, Piccolomini F, Puri N, Gastaldelli A, Kusmic C, L’Abbate A, Abraham NG: The L-4F mimic peptide prevents insulin resistance through increased levels of HO-1, pAMPK, and pAKT in obese mice. J Lipid Res 2009, 50:1293-1304.
26. Hardie DG: Minireview: the AMP-activated protein kinase cascade: the key sensor of cellular energy status. Endocrinology 2003, 144:5179-5183.
27. Hopkins TA, Ouchi N, Shibata R, Walsh K: Adiponectin actions in the cardiovascular system. Cardiovasc Res 2007, 74:11-18.
28. Schreyer SA, Wilson DL, LeBoeuf RC. C57BL/6 mice fed high fat diets as models for diabetes-accelerated atherosclerosis. Atherosclerosis 1998, 136:17-24.

29. Molnar J, Yu S, Mhavva N, Paug C, Ceresahev N, Danays HM. Diabetes induces endothelial dysfunction but does not increase neointimal formation in high-fat diet fed C57BL/6 mice. Circ Res 2005, 96:1178-1184.

30. Survit R, Kahn CM, Cochrane C, McCubbin JA, Feinglos MN. Diet-induced type II diabetes in C57BL/6 mice. Diabetes 1988, 37:1163-1167.

31. L'Abbate A, Neglia D, Vecoli C, Novelli M, Ottaviano V, Baldi S, Barsacchi R, Paolucci A, Masiero P, Drummond GS, Mc Clung JA, Abraham NG. Beneficial effect of heme oxygenase-1 expression on myocardial ischemia-reperfusion involves an increase in adiponectin in mildly diabetic rats. Am J Physiol Heart Circ Physiol 2007, 293:H532-H5341.

32. Paolucci N, Biondi R, Bettini M, Lee CI, Berlowitz CO, Rossi R, Xia Y, Ambroso G, L'Abbate A, Kass DA, Zweier JL. Oxygen radical-mediated reduction in basal and agonist-evoked NO release in isolated rat heart. J Mol Cell Cardiol 2001, 33:671-679.

33. Sodhi K, Houe K, Gottlinger K, Canestramo M, Vanella L, Kim DH, Manthali R, Koduru SR, Falck JR, Schwartzman ML, Abraham NG. Epoxyeicosatrienoic acid agonist rescues the metabolic syndrome phenotype of HO-2-null mice. J Pharmacol Exp Ther 2009, 331:906-916.

34. Lara-Castro C, Luo N, Wallace P, Klein RL, Garvey WT. Adiponectin multimeric complexes and the metabolic syndrome trait. Diabetes 2006, 55:249-259.

35. Sardana MK, Kappas A. Dual control mechanism for heme oxygenase; (IV)-protoporphyrin plays a potentially inhibitory enzyme activity while markedly increasing content of enzyme protein in liver. Proc Natl Acad Sci USA 1987, 84:2464-2468.

36. Cobdian AD, Davies MJ, Previtt RL, Lauterio TJ. Development of hypertension in a rat model of diet-induced obesity. Hypertension 2000, 35:1009-1015.

37. Cobdian AD, Davies MJ, Schriner SD, Lauterio TJ, Previtt RL. Oxidative stress in a rat model of obesity-induced hypertension. Hypertension 2001, 37:554-560.

38. Bing CH, Brooks WW, Robinson KG, Slavsky MT, Hayes JA, Litwin SE, Shen S, Conrad CH. The spontaneously hypertensive rat as a model of the transition from compensated left ventricular hypertrophy to failure. J Mol Cell Cardiol 1995, 27:383-396.

39. Kruger AL, Peterson S, Turkseven S, Karninski PM, Zhang FF, Su Q, Wolin MS, Abraham NG. DF-4 induces heme oxygenase-1 and extra cellular superoxide dismutase, decreases endothelial cell sloughing, and improves vascular reactivity in rat model of diabetes. J Pharmaco Exp Ther 2008, 326:1347-1357.

40. Dimeoller S, Fleming I, Frischthaler B, Hermann C, Busse R, Zeiher AM. Activation of nitric oxide synthase in endothelial cells by Akt-dependent phosphorylation. Nature 1999, 399:601-605.

41. Chun ZP, Mitchelhill KI, Michell B, Stapleton D, Rodriguez-Crespo I, Witters LA, Power DA, Ornt De Montellano PR, Kemp BE. AMP-activated protein kinase is expressed in endothelial cells and decreases NO synthase. Cell Biol Int 2005, 25:9554-9575.

42. Sambucetti G, Morbelli S, Vanella L, Kuzmic C, Marin C, Masillo M, Augeri C, Corselli M, Ghersi C, Chiavarina B, Rodella LF, L'Abbate A, Drummond G, Abraham NG, Frassoni F. Diabetes Impairs the Vascular Recruitment of Normal Stem Cells by Oxidant Damage; Reversed by Increases in pAMPK, Heme Oxygenase-1 and Adiponectin. Stem Cells 2009, 27:399-407.

43. Cotterl DF, Kurfth EL, Winder WW. AICA riboside increases AMP-activated protein kinase, fatty acid oxidation, and glucose uptake in rat muscle. Am J Physiol 1997, 273:E1107-E1112.

44. Kudo N, Gillespie JG, Kung L, Witters LA, Schulz R, Clanchan AS, Lopashuk GD. Characterization of SAMP-activated protein kinase activity in the heart and its role in inhibiting acetyl-CoA carboxylase during reperfusion following ischemia. Biochim Biophys Acta 1996, 1301:67-75.

45. Terai K, Hiramoto Y, Masaki M, Sugiyama S, Kuroda T, Hori M, Kawase I, Hirota H. AMP-activated protein kinase protects cardiomyocytes against hypoxic injury through attenuation of endoplasmic reticulum stress. Mol Cell Biol 2005, 25:9554-9575.

46. Di Marco A, Incheva M, Ciampi A, Mafi B, Merighi E, Zaninotto G, Kaur K. Adiponectin Protects Mesenchymal Stem Cells from Oxidative Stress in an In Vitro Model of Aging. Mol Med 2010, 16:183-191.

47. Di Marco A, Incheva M, Ciampi A, Mafi B, Merighi E, Zaninotto G, Kaur K. Adiponectin Protects Mesenchymal Stem Cells from Oxidative Stress in an In Vitro Model of Aging. Mol Med 2010, 16:183-191.

48. Mathew AV, Okada S, Sharma K. Obesity related kidney disease. Curr Diabetes Rev 2011, 7:41-49.

49. Ik Jr, Sharma K. Mechanisms linking obesity, chronic kidney disease, and fatty liver disease: the roles of fetuin-A, adiponectin, and AMPK. J Am Soc Nephrol 2010, 21:406-412.

50. Tan J, Xiong X, Liu W, Guo S, Li Q, Zhang R, Lao G. Increased plasma adiponectin closely associates with vascular endothelial dysfunction in type 2 diabetic patients with diabetic nephropathy. Diabetes Res Clin Pract 2010, 88:177-183.

51. Cao et al. High fat diet enhances cardiac abnormalities in SHR rats: Protective role of heme oxygenase-adiponectin axis. Diabetology & Metabolic Syndrome 2011 3:7.