Sophorolipid-Based Oligomers as Polyol Components for Polyurethane Systems

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Abstract: Due to reasons of sustainability and conservation of resources, polyurethane (PU)-based systems with preferably neutral carbon footprints are in increased focus of research and development. The proper design and development of bio-based polyols are of particular interest since such polyols may have special property profiles that allow the novel products to enter new applications. Sophorolipids (SL) represent a bio-based toolbox for polyol building blocks to yield diverse chemical products. For a reasonable evaluation of the potential for PU chemistry, however, further investigations in terms of synthesis, derivatization, reproducibility, and reactivity towards isocyanates are required. It was demonstrated that SL can act as crosslinker or as plasticizer in PU systems depending on employed stoichiometry. (ω-1)-hydroxyl fatty acids can be derived from SL and converted successively to polyester polyols and PU. Additionally, (ω-1)-hydroxyl fatty acid azides can be prepared indirectly from SL and converted to A/B type PU by Curtius rearrangement.

Keywords: polyurethane; polyol; bio-based; sophorolipid-based polyols; hydroxyl fatty acid based polyols; platform chemicals

1. Introduction

The main components of well-known and very versatile polyurethanes (PU) [1,2] are polyisocyanates and polyols [3,4]. The morphology and thus the properties of PU are mainly determined by the intrinsic structure of the polyol component(s) since their content exceeds typically 60 wt.% of the PU [5–7]. The polyols most commonly utilized are polyether, polyester, and polycarbonate polyols [5,8]. All types of polyols are usually produced by means of petrochemical resources. Considering the need for a sustainable economic activity [9], the development of polyols based on renewable resources and the research on their applicability for PU systems is of increased significance [10–18]. Among bio-based polyol platforms, sophorolipids (SL) are well documented and show high potential to provide a versatile toolbox for polyol building blocks [19–26]. Specific advantages of SL are their non-pathogenic production organisms, possessing high productivity and an efficient rate of substrate conversion [21]. SL consist of sophorose, a hydrophilic di-glucose, coupled to a hydrophobic hydroxyl fatty acid (HFA) by a glycosidic bond (Figure 1(1B)). The resulting SL is amphiphilic and thus readily utilized as bio-based surfactant in e.g., surface technology [24,27,28]. The most effective organism to produce SL is bombicola, which is capable of precipitating up to 400 g L⁻¹ SL into the fermentation medium [20]. Moreover, structural variation is feasible by feeding different lipid derivatives with C16 and C18 fatty acids, which incorporate in particularly good yields [29,30]. The most common SL derivatives are lactonic SL (LSL) and acidic SL (ASL), as depicted in Figure 1 [24,31].
Dihydroxyl alkyl glycosides are structurally similar to SL and were already converted successfully to PU by reaction with diisocyanates [14,32]. In these procedures, the double bond of the fatty acid moiety is transformed into a vicinal diol structure, which is subsequently converted to PU by reaction with the isocyanate (NCO) groups of isophorone diisocyanate (IPDI). Consequently, SL should perform similarly to give PU. Moreover, LSL has six hydroxyl (OH) groups (two acetylated primary OH groups and four secondary OH groups) and ASL seven OH groups (two acetylated primary OH groups and five secondary OH groups) per molecule. This means that a successive conversion with (di)isocyanates is generally possible because the secondary OH groups are readily available for the reaction with NCO groups, whereas the primary OH groups become available after, e.g., saponification [33]. The complete deacetylated ASL should also allow a certain selectivity since primary OH groups react with NCO groups about three times faster than secondary OH groups [34–38]. Utilizing appropriate reaction conditions, primary OH groups can purposely be consumed first in order to produce linear and thus thermoplastic PU [38–43]. In contrast, complete conversion of all OH groups leads to significant crosslinking and therefore to a thermosetting material. Considering the Flory–Stockmayer relation [44], the degree of crosslinking and thus the nature of the thermosetting material can be tailored to a certain extent, e.g., by appropriate choice of the NCO/OH ratio (index) [44–46]. In summary, SL provide a high number of potential reaction sites with different reactivity toward NCO groups and should therefore be suited as crosslinking agent and potentially as a linear building block [32]. Additionally, SL are suited as internal emulsifier in PU dispersion technology because of their amphiphilic nature [47–49].

HFA form renewable resources like castor oil, which can be converted to polyester polyols or directly to PU [17,50–52]. Typically for such HFA is that the OH group is localized in the center of the molecule. The resultant polyester or PU systems consequently contain dangling chains that reduce crystallinity and glass transition temperature [52,53]. In contrast to that, \( \omega \) or \((\omega-1)\)-HFA yield systems without side chains and with lower ester- or urethane group concentrations. Therefore, dangling chains are not expected and lipophilicity is increased suggesting enhanced hydrolysis resistance and improved interactions to hydrophobic surfaces [54]. \( \omega \) or \((\omega-1)\)-HFA with varying chain lengths can be obtained by fragmentation of SL [26] and successively converted chemically [55] or enzymatically [56–60] to yield polyesters via A/B self-condensation. Note that successive conversion with diisocyanates to high molecular weight PU requires two-fold OH termination, which can be accomplished, e.g., by initiation with or addition of suitable diols [55,61]. Additionally, the acid moiety of HFA can be converted to azide...
in order to accomplish Curtius rearrangement [62] yielding A/B type polyurethanes [51,63–68].

The present work shows that LSL can act as crosslinker or as plasticizer in PU systems depending on employed stoichiometry. (ω-1) HFA-based polyester polyols and (ω-1) HFA-based PU systems are feasible; however, low amounts of glucose impurities seem to limit the versatility of the reaction since there is a substantial extent of crosslinking and branching. Additionally, we demonstrate the fundamental access to A/B type PU systems by applying Curtius rearrangement on (ω-1) HFA azides obtained from LSL, confirming the Curtius approach to A/B type PU systems of other groups [51,64–69].

2. Materials and Methods

2.1. Materials

1,4-Dioxane (pure), dimethyl sulfoxide (DMSO, ≥99%), oxalyl chloride (for synthesis), and tetrabutylammonium hydroxide (in 2-propanol/methanol, TitriPUR) were from Merck Millipore. 4-tert-Butylcatechol (99%), N-methylpyrrolidin-2-one (NMP, for synthesis), and sodium azide (NaN₃, >99%) were from Acros Organics. 12-Hydroxystearic acid (95%), sodium sulfate (Na₂SO₄, anhydr., 99%), and tert-butyl methyl ether (MTBE, ≥99%,) were from Alfa Aesar. 1-Hexylisocyanate (HIC, 97%), dibutylamine (99.5%), phenylisocyanate (PIC, for synthesis), and tin(II) chloride (SnCl₂, 98%) were from Sigma Aldrich. 1,6-Hexanediol (for synthesis), 1,6-hexamethyleneisocyanate (HDI, >98%), 2-butanone (MEK, >99%), 2-propanol (≥99%), acetonitrile (≥99.9%), N,N-dimethylformamide (DMF, ≥99.8%), hydrochloric acid (HCl, 1 M, TitriPUR), potassium hydroxide (KOH, 0.1 M in methanol, TitriPUR), and sodium chloride (NaCl, ≥99%) were from VWR Chemicals. Deuterated chloroform (CDCl₃ with 0.03% TMS, 99.8 atom%D) and deuterated THF (THF-d₈ with 0.03% TMS, 99.8 atom%D) were from Deutero. Acetone (≥99%), cyclohexane (≥99.5%), chloroform (CHCl₃, >99%), dichloromethane (CH₂Cl₂, for analysis), diethyl ether (≥99.5%), ethanol (abs.), ethyl acetate (for analysis), methanol (HPLC grade), petroleum ether (for analysis), tetrahydrofuran (THF, HPLC grade), and toluene (≥99.9%) were from Fisher Chemical. Sodium hydroxide (NaOH, pure) was from Bernd Kraft. n-Hexane (≥95%) was from Carl Roth. Dibutyltindilaurate (DBTDL, technical grade) and poly(1,4-butylene)adipate (PBA, technical grade, M₀ = 2250 g mol⁻¹) were kindly provided by Covestro Deutschland AG. SL (1A) (LSL) (Figure 1) was synthesized and purified according to the methods published elsewhere and provided by Zerhusen and Schörken [19,25,27]. 1,6-Hexanediol and PBA were dried for at least 2 h under reduced pressure, and kept at elevated temperature (50 mbar, 80 °C) before use. All other chemicals were used as received or purified applying established and well-known procedures [70].

2.2. Measurements and Equipment

Size exclusion chromatography (SEC) was carried out on a PSS Polymer SECcurity system based on Agilent 1260 hardware modules equipped with SECcurity isocratic pump, vacuum degasser, refractive index and UV-Vis detector (254 nm), column oven, and a standard auto sampler. A styrene-divinylbenzene copolymer column (SDV linear XL (100–3,000,000 Da)) with 5 µm particle size and 1000 Å porosity was calibrated with polystyrene ReadyCal Kit (PSS Polymer) standards. Measurements were carried out in THF at 30 °C with a flow rate of 1.0 mL min⁻¹. Integration of the signals was performed via software package “WinGPC Unity” from PSS Polymer.

¹H- and ¹³C-NMR spectra were recorded using a Bruker Ascend 400 spectrometer (400 MHz) at room temperature using CDCl₃ or THF-d₈ as solvent (sample conc. = 0.05 mg µL⁻¹). Tetramethylsilane (TMS) was used in all experiments as internal standard.

ATR-IR spectra were recorded using a Bruker Platinum-ATR equipped with a MIR-RT-DLaTGS detector and a KBr radiation plate. OPUS software of Bruker Optic GmbH was used for data handling. Spectra were recorded at room temperature, applying 24
scans, and a resolution of 4 cm⁻¹. Background spectra were recorded in ambient atmosphere prior to each measurement. For reaction monitoring, inline-IR-spectroscopy was used applying Thermo Fisher Nicolet IS 50-spectrometer equipped with a probe coupler and the ZnSe-ATR-tube FlexiSpec® from art photonics GmbH, Berlin. HgCdTe detector was cooled with liquid N₂. The software “ Macros Basic” enabled 20 automated scans per minute in a resolution of 2 cm⁻¹. Background spectra were recorded in dry N₂ prior each measurement. “ OMNIC” software was used to calculate peak areas.

Differential scanning calorimetry (DSC) was conducted applying Q2000 DSC from TA instruments consisting of an auto sampler and a RCS90 cooling unit calibrated by an indium standard. Samples were placed in Tzero aluminum crucibles. SL containing samples were heated to 150 °C and cooled to −60 °C twice using a rate of 10 K min⁻¹. The results of the second cycle were used for evaluation.

X-ray diffraction (XRD) was conducted with a Bruker D2 Phaser 2nd Gen equipped with a Cu tube (λ = 1.54184 Å), fixed slit 0.4 mm, 4° grid, and a Lynx 1D Modus detector as single measurements in order to obtain data for crystallinity (angle of incidence: 5–80°; rotation: 0–15 rpm; increment: 0.03°; time per step: 0.02 s; scan type: Coupled Two Theta Theta; scan modus: Continuous PSD fast). Samples were applied as films or ground powder. For evaluation, Bruker’s Diffrac EVA software was used.

High-performance liquid chromatography (HPLC) was conducted using Shimadzu Nexera XR equipped with a BM-20 A communications bus module, two LC-20 AD XR, a SIL-D0ACXR auto sampler, a column oven, and a VWR-ELSD 80 detector. Conditions: N₂; 3.5 bar; 40 °C; column: 4.6 mm × 250 mm, 5 µM C18 RP La Chrom II (Hitachi, Tokio, Japan); flow rate: 1.0 m min⁻¹; 30 °C; solvent: acetonitrile/water (50 vol.%:50 vol.% for 25 min. to 99 vol.% acetonitrile linearly within 60 min.); samples: 2.5 mg mL⁻¹ in THF.

Acid number, hydroxyl number and the content of NCO groups were determined according to DIN EN ISO 660 2009, DIN 53240-2 (ASTM E1899-08), and DIN EN ISO 11909 2007, respectively. Automated titration was conducted with a TitroLine 7000 titration unit from SI Analytics.

Shore A hardness was measured five times for each sample according to DIN EN ISO 868 applying SAUTER HBA 100-0 and SAUTER TI-A0 with a contact pressure of 5 kg and calibrated with Durometer Test Block Kit AHBA-01 (SAUTER). Tensile tests were conducted according to DIN EN ISO 527-1/2 (ASTM D 638) on a Shimadzu Autograph AG-X plus tensile testing machine (50 mm min⁻¹). TrapeziumX (Shimadzu, Kyoto, Japan) software was used for evaluation. The samples were polymer films produced by casting the liquid polymer (2000 µM) on a Teflon plate with a squeegee.

The preparation of SL solutions was as follows: 100 mg (0.145 mmol) of (1A) (LSL) or 90.5 mg (0.145 mmol) (1B) (ASL) was transferred at room temperature to 2 mL of solvent and the solubility evaluated optically.

2.3. Synthesis of 17-Hydroxyoctadec-9-Ene Acid (ω-1) HFA

(ω-1) HFA (2) (Figure 2) was produced by a modification of published procedures in absence of carcinogenic dioxane, but at the expense of reaction time [26,71]. A typical protocol was as follows. LSL (1A), derived from starmerella bombicola (97% purity) [19], was dissolved in 5 M NaOH solution. To this solution, a further 5 M NaOH was added dropwise until a constant pH was reached. After pH adjustment to 3.5 with diluted HCl, the intermediate (1B) was crystallized at 7 °C and purified by lyophilization to give a white powder. CasH₁₈O₂: M = 622.75 g mol⁻¹; SEC: M₅₅ = 530 g mol⁻¹; Mₛₛ = 660 g mol⁻¹; PDI = 1.3; yield: 89.2%; FT-IR (ATR) ν/cm⁻¹: 3324 (m), 2921 (m); 2851 (m); 1726 (m); 1559 (w); 1155 (m); 1070 (s); 1019 (s); 894 (m); 643 (m); 511 (m); HPLC: 10 min.

A total of 5.0 g (0.008 mol) (1B) was placed in 100 mL round-necked flask and dissolved under stirring in 50 mL 1 M HCl. The reaction proceeds at 80 °C, turning the clear liquid to a turbid solution comprising particles and further to a biphasic system with yellowish, oily droplets. The reaction progress was monitored by HPLC. After completion of hydrolysis, the pH was adjusted to 3.5 by addition of diluted NaOH. The product was
purified by solvent extraction (distilled H2O/CHCl3) and removal of excessive solvent in vacuo. The product (2) accumulated as yellowish oil in 83.5% yield. Since dimers and oligomers were identified, (2) was stirred in 5 M NaOH at 80 °C for a further 16 h. Repeated solvent extraction and removal applying the same conditions yielded a brownish oil.

![Figure 2](image-url) 17-Hydroxyoctadec-9-ene acid ((ω-1) HFA) (2).

C₉₆H₁₆₄O₇; M = 298.47 g mol⁻¹; OH number = 240; acid number = 86.1; yield: 54.3%; FT-IR (ATR) ν/cm⁻¹: 3393 (w), 3004 (w), 2924 (s), 2853 (s), 1709 (s), 1460 (m), 1375 (m), 1243 (m), 1191 (m), 1081 (m), 754 (s); HPLC: 40 min. (monomer), 89 min. (dimer); ¹H NMR (400 MHz, CDCl₃, δ): 5.40–5.28 (m, 2H; H10, H11), 3.81 (m, 1H; H2), 2.34 (t, J = 7.5 Hz, 2H; H18), 2.07–1.96 (m, 4H; H9, H12), 1.69–1.52 (m, 4H; H4, H7, H17), 1.43 (d, J = 3.7 Hz, 4H; H8, H13), 1.37–1.22 (m, 12H; H5–7, H14–16), 1.19 (dd, 3H; H3); ¹³C NMR (101 MHz, CDCl₃, δ): 178.66 (C19), 129.85 (C10, C11), 68.33 (C2), 36.75 (C3), 34.78 (C18), 29.69–28.99 (C6–C8, C13–C16), 27.19 (C9, C12), 25.75 (C5), 24.73 (C17), 23.43 (C3).

2.4. Synthesis of (ω-1) HFA Based Polyester Diol

OH-terminated (ω-1) HFA-based polyester diol (3) (Figure 3) was prepared according to a published procedure [61]. A total of 9.0 g (0.03 mol) (ω-1) HFA (2) and 0.7 g (0.006 mol) 1,6-hexanediol was introduced in a 100 mL flask equipped with a Vigreux column, thermometer, and distilling link. The reaction mixture was heated to 200 °C within 1 h. Reaction water was removed by distillation. In contrast to the published procedure, no catalyst (SnCl₂) was used. Instead, 7.0 mg (0.042 mmol) 4-tert-butylcatechol was added in order to prevent undesired reactions at the double bond. After 24 h, the reaction was stopped, leaving a highly viscous and sticky product, which was partially soluble in THF.

![Figure 3](image-url) (ω-1) HFA based polyester diol (3).

SEC (soluble fraction): Mₘ = 2370 g mol⁻¹ (Mₘ,tho = 2250 g mol⁻¹), Mₜ = 99,700 g mol⁻¹, PDI = 42.1; acid number = 2.8; yield: not determined; FT-IR (ATR) ν/cm⁻¹: 3350 (m), 2930 (s), 2858 (s), 1726 (s), 1460 (m), 1374 (m), 1055 (s), 727 (w); ¹H NMR (400 MHz, THF-d₈, δ): 5.33 (t, J = 4.8 Hz, 4H; H9, H10, H31, H32), 4.89–4.78 (m, 1H; H1), 4.15–4.05 (m, 2H; H42), 4.01 (t, J = 6.6 Hz, 3H; H39, H47), 2.31–2.16 (m, 4H; H17, H23), 2.03 (m, 8H; H8, H11, H30, H33), 1.72–1.45 (m, 12H; H3, H16, H25, H38, H43, H46), 1.45–1.20 (m, 49H; H4–H7, H12–15, H21, H26–29, H34–H37, H44, H45), 1.16 (d, J = 6.2 Hz, 1H; H41); ¹³C NMR (101 MHz, THF-d₈, δ): 172.30 (C18, C22), 129.54 (C9, C10, C31, C32), 69.85 (C1), 66.87 (C39), 63.47 (C47), 35.92 (C38), 34.04 (C3), 33.66 (C17, C23), 29.69 (C36), 29.22–28.62 (C5–C8, C12–C14, C27–C29, C34–C36), 27.03 (C8, C11, C30, C33), 25.61–24.74 (C16, C25, C37, C45), 23.95 (C41), 19.35 (C21).

2.5. 12-Hydroxystearic Acid Based Polyesterdiol
OH-terminated 12-hydroxystearic acid based polyesterdiol (4) (Figure 4) was prepared according to a published procedure [61]. A total of 5.0 g (16.7 mmol) 12-Hydroxystearic acid and 0.41 g (3.5 mmol) 1,6-hexanediol was introduced in a 20 mL flask equipped with a Vigreux column, thermometer, and distilling link. The reaction mixture was heated to 200 °C within 1 h. Reaction water was removed by distillation. After 24 h, SnCl2 was added in order to complete the conversion. After another 24 h, vacuum was applied to remove water residues leaving a highly viscous liquid as the product.

![Figure 4: 12-Hydroxystearic acid based polyesterdiol (4).](image)

SEC: $M_n = 2210$ g mol$^{-1}$ ($M_{n,thio} = 2250$ g mol$^{-1}$), $M_w = 5800$ g mol$^{-1}$, PDI = 2.6; acid number = 2.1; $T_b = -20$ °C; yield: quantitative; FT-IR (ATR) μ/cm$^{-1}$: 3350 (w), 2922 (s), 2852 (s), 1731 (s), 1464 (m), 1376 (w), 1174 (m), 1100 (w), 723 (w); $^1$H NMR (400 MHz, THF-ds, δ): 4.85 (m, $J = 6.2$ Hz, 1H; H12), 4.01 (t, $J = 6.6$ Hz, 2H; H21), 3.41 (s, 1H; H41), 3.25 (s, 2H; H62), 2.24 (m, $J = 7.4$, 3.1 Hz, 4H; H2, H31), 1.59 (m, $J = 8.3$, 5.2, 4.4 Hz, 8H; H3, H22, H25, H32), 1.56–1.47 (m, 16H; H4, H11, H13, H23, H24, H33, H40, H42), 1.40–1.19 (m, 44H; H4–H10, H14–H17, H33–H39, H43–H46), 0.89 (t, $J = 6.5$ Hz, 6H; H18, H47); $^{13}$C NMR (101 MHz, THF-ds, δ): 172.05 (C1, C29), 73.03 (C12), 70.34 (C41), 65.98 (C21), 63.45 (C26), 37.91 (C40, C42), 34.21 (C31), 34.03 (C11, C13), 31.79 (C23, C45), 29.90–28.63 (C4–C9, C15, C33–C37, C44), 25.80 (C3, C24, C32), 24.90 (C17, C46), 13.47 (C18, C47).

2.6. Synthesis of 17-Hydroxyoctadec-9-Enyl Azide ((ω-1) HFA Azide)

The synthesis of (5) (Figure 5) follows a modification and combination of several published procedures in order to improve reaction time and yield [72,73]. In a 100 mL flask, 0.75 g (2.50 mmol) (ω-1) HFA (2) was dissolved in 15 mL dry CH2Cl2. To this solution, 0.25 mL (3.0 mmol) oxalyl chloride and 2.30 mL (0.03 mmol) DMF was added, and the reaction mixture was stirred at room temperature for 1 h. Then the solution was cooled to 0 °C and 0.65 mg (10 mmol) of an aqueous NaN3 solution was added dropwise. The resulting mixture was stirred at 0 °C for 3 h. The organic phase was extracted by 3 × 30 mL CHCl3. The combined organic layers were washed with 40 mL brine, dried over anhydrous Na2SO4, and concentrated under reduced pressure. The product accumulated as a yellow oil.

![Figure 5: 17-Hydroxyoctadec-9-Enyl azide ((ω-1) HFA azide) (5).](image)

Cas$_3$H$_5$N$_3$O$_2$: $M = 323.47$ g mol$^{-1}$; yield: 49%; FT-IR (ATR) μ/cm$^{-1}$: 3335 (w), 2926 (s), 2854 (m), 2158 (w), 1704 (s), 1658 (s), 1368 (m), 1288 (w), 1168 (m), 1064 (m), 976 (w), 832 (w), 721 (w); $^1$H NMR (400 MHz, CDCl3, δ): 5.34 (t, $J = 11.71$, 7.3 Hz, 2H; H9, H10), 4.83–4.71 (m, 1H; H-OH), 3.72 (m, 1H; H17), 2.31 (t, $J = 7.53$ Hz, 2H; H2), 2.01 (m, 4H; H8, H11),
1.68–1.57 (m, 4H; H3, H16), 1.49–1.37 (m, 6H; H7, H15, H17), 1.18 (m, 10H; H4–H6, H13, H14), 1.18 (d, 3H; H18); $^{13}$C NMR (75 MHz, CDCl$_3$, δ): 178.4 (C1), 130.0 (C9, C10), 68.2 (C17), 39.3 (C16), 29.8 (C2), 29.2 (C4), 29.1 (C7, C12), 29.0 (C5, C6, C13), 28.9 (C5, C11), 27.2 (C14), 25.6 (C15), 25.2 (C3), 23.7 (C18).

2.7. Preparation of Urethanes

2.7.1. LSL and PBA-Based Urethanes

In a typical procedure, LSL ((1A), M = 694.44 g mol$^{-1}$, $f_{(OH)} = 4$) or PBA ($f_{(OH)} = 2$) was placed in a three-necked flask, dissolved in 50–80 mL acetone, and heated to the desired temperature shown in Table 1. The corresponding amounts of HIC, PIC, or HDI and DBTDL were added to the solution and the reaction was monitored by inline IR and NCO titration. Residual amounts of diisocyanate were quenched with excessive MeOH prior to further investigations. Figure 6 shows characteristic IR spectra obtained applying this procedure exemplarily for the sample LSL-HDI1.1.

Table 1. Conditions for the reaction of LSL or PBA with 1-isocyanatohexane (HIC), phenylisocyanate (PIC) or 1,6-hexamethylenediisocyanate (HDI) in acetone.

| Sample       | Polyol | Isocyanate | $n_{NCO}$/mol | $n_{NCO}$/nOH | DBTDL/ppm$^1$ | T/°C |
|--------------|--------|------------|---------------|---------------|--------------|------|
| LSL–HIC–S1   | LSL    | HIC        | 0.0317        | 1.1           | 125          | 30   |
| LSL–HIC–S2   | LSL    | HIC        | 0.0317        | 1.1           | 125          | 50   |
| LSL–HIC–S3   | LSL    | HIC        | 0.0317        | 1.1           | 500          | 50   |
| LSL–HIC–S4   | LSL    | HIC        | 0.0432        | 1.5           | 500          | 50   |
| LSL–PIC–S1   | LSL    | PIC        | 0.0317        | 1.1           | 125          | 30   |
| LSL–PIC–S2   | LSL    | PIC        | 0.0317        | 1.1           | 125          | 50   |
| LSL–PIC–S3   | LSL    | PIC        | 0.0317        | 1.1           | 500          | 50   |
| LSL–PIC–S4   | LSL    | PIC        | 0.0432        | 1.5           | 500          | 50   |
| LSL–PIC–S5   | LSL    | PIC        | 0.1440        | 5.0           | 500          | 50   |
| LSL–HDI0.5   | LSL    | HDI        | 0.0072        | 0.5           | 500          | 50   |
| LSL–HDI1.1   | LSL    | HDI        | 0.0158        | 1.1           | 500          | 50   |
| LSL–HDI10    | LSL    | HDI        | 0.1440        | 10.0          | 500          | 50   |
| PBA + PIC    | PBA    | PIC        | 0.0220        | 1.1           | 500          | 50   |
| PBA + HIC    | PBA    | HIC        | 0.0220        | 1.1           | 500          | 50   |
| PBA + HDI    | PBA    | HDI        | 0.0048        | 1.1           | 500          | 50   |

$^1$ relative to nOH.
2.7.2. Synthesis of PU Based on (ω-1) HFA-Based Polyesterdiol and 12-Hydroxystearic Acid Based Polyesterdiol

(ω-1) HFA-based polyesterdiol (3) was dissolved in THF for 7 d. After this period, solubility was complete and \( M_n \) was measured to be 1561 g mol\(^{-1}\). A total of 25 mL THF solution containing 3.5 g (2.3 mmol) (3) was heated to 60 \( ^\circ\)C and 500 ppm DBTDL and 0.58 g (3.5 mmol) HDI was added. According to NCO titration, the reaction was completed after 16 h, cooled to room temperature and the solvent removed under reduced pressure. Residual HDI was quenched by 0.1 M dibutylamine solution in acetone leaving a dark brown rubberlike product.

SEC: \( M_n = 3530 \) g mol\(^{-1}\) \((M_n,\text{theo} = 3406 \) g mol\(^{-1}\)), \( M_w = 772,330 \) g mol\(^{-1}\), PDI = 218.7; \( T_g = -43 \) \( ^\circ\)C; FT-IR (ATR) \( \text{\nu/cm}^{-1} \): 3333 (w), 3002 (w), 2925 (s), 2854 (s), 1732 (s), 1621 (m), 1533 (s), 1459 (m), 1373 (m), 1234 (s), 1172 (s), 726 (m), 590 (w).

12-Hydroxystearic acid based polyesterdiol (4) was dissolved in THF for 7 d. After this period, \( M_n \) was measured to be 1862 g mol\(^{-1}\). A total of 25 mL THF solution containing 3.5 g (2.3 mmol) (4) was heated to 60 \( ^\circ\)C and 500 ppm DBTDL and 0.47 g (2.8 mmol) HDI was added. According to NCO titration, the reaction was completed after 16 h, cooled to room temperature, and the solvent removed under reduced pressure. Residual HDI was quenched by 0.1 M dibutylamine solution in acetone, leaving a dark brownish oil as product.

SEC: \( M_n = 4170 \) g mol\(^{-1}\) \((M_n,\text{theo} = 5209 \) g mol\(^{-1}\)), \( M_w = 11,530 \) g mol\(^{-1}\), PDI = 2.8; \( T_g = -43 \) \( ^\circ\)C; FT-IR (ATR) \( \text{\nu/cm}^{-1} \): 3333 (w), 3002 (w), 2925 (s), 2854 (s), 1732 (s), 1621 (m), 1533 (s), 1459 (m), 1373 (m), 1234 (s), 1172 (s), 726 (m), 590 (w).

2.7.3. Preparation of A/B Type PU

In a 100 mL round-necked flask, 0.80 g (2.5 mmol) 17-Hydroxyoctadec-9-enoyl azide (5) was dissolved in 1 mL THF under N\(_2\) atmosphere. Curtius rearrangement and successive polymerization was started applying different temperatures and allowed to proceed for different periods of time (for details see Section 3). The resulting A/B type polymer (6) is depicted in Figure 7.
Figure 7. A/B Type PU (6) derived by Curtius rearrangement of azide (5).

2.7.4. Ternary Urethane Systems

For kinetic investigations, 16.42 g (7.0 mmol) PBA and 5 g (7.0 mmol) LSL were dissolved in 80 mL acetone. At 50 °C, 6.1 g (0.048 mol) HIC or 5.7 g (0.048 mol) PIC was added. The reaction was immediately started by addition of 500 ppm DBTDL and monitored by NCO titration.

For variation of the polyol composition, corresponding amounts of LSL and PBA (Table 2) were placed in an appropriate flask, dissolved in acetone, and heated to 50 °C. After addition of HDI and 500 ppm DBTDL, the reaction was started and monitored by NCO titration. After completion of the reaction, the product was cast on a Teflon plate applying a squeegee (2000 μm) and dried at room temperature.

Table 2. Conditions to produce ternary PU systems based on LSL and PBA. (solvent = acetone, T = 50 °C, \text{DBTDL} = 500 ppm, \text{x}_{\text{LSL}} = \text{molar fraction of LSL}, \text{T}_{\text{M}} = \text{melting point}).

| Sample           | \text{x}_{\text{LSL}} | \text{n}_{\text{LSL}}/\text{mol} | \text{n}_{\text{PBA}}/\text{mol} | \text{n}_{\text{NCO}}/\text{mol} | \text{T}_{\text{M}}\text{°C} |
|------------------|------------------------|-----------------------------------|----------------------------------|-------------------------------|-----------------------------|
| PBA–HDI–0%LSL    | 0.0                    | -                                 | 0.0222                           | 0.0110                       | 47.51                       |
| PBA–HDI–10%LSL   | 0.1                    | 0.0022                            | 0.0200                           | 0.0122                       | 41.39                       |
| PBA–HDI–20%LSL   | 0.2                    | 0.0050                            | 0.0200                           | 0.0150                       | 40.65                       |
| PBA–HDI–50%LSL   | 0.5                    | 0.0178                            | 0.0178                           | 0.0263                       | 42.57                       |
| PBA–HDI–70%LSL   | 0.7                    | 0.0100                            | 0.0235                           | 0.0285                       | 46.22 ¹                      |
| PBA–HDI–100%LSL  | 1.0                    | 0.0073                            | -                                | 0.0073                       | 46.22 ¹                      |

¹\text{T}_{\text{g}}

3. Results and Discussion

3.1. Sophorolipids as Polyol Component for PU Systems

3.1.1. Solubility Studies

Lactonic SL (LSL, Figure 1 (1A)), and acidic SL (ASL, Figure 1 (1B)) can be produced in good yields and with a purity of at least 97% [19,25,27]. The melting points of LSL and ASL ranges between 55 °C and 65 °C. Above this temperature range, the SL remain highly viscous. The reaction with isocyanates in bulk is thus quite challenging, which is why the utilization of an appropriate solvent becomes highly favorable. The solubility of LSL and ASL in selected solvents is listed in Table 3.

LSL is soluble in several solvents, particularly in acetone, MEK, and THF, all being solvents commonly used in PU synthesis. In contrast to that, ASL shows poor solubility in these solvents. In fact, solubility is sufficient in distilled water and methanol, both being solvents with relatively high polarity. This is assumed to be the result of the higher intrinsic polarity of ASL due to the additional OH- and COOH- functionalities. Note however that acceptable solubility is also measured using less polar DMF and DMSO, suggesting a more complex solubility behavior that is determined not alone by polarity.

Water and methanol are well-known to be inappropriate solvents to study the conversion of polyols with isocyanates since they react themselves with isocyanates
forming urea and urethane groups [1,3]. DMF and DMSO are improper solvents as well because DMF reacts with isocyanates forming amidines and biurets [74,75] and DMSO yields sulfide esters by reaction with the carboxyl group of ASL [76]. Hence, the reaction of SL with isocyanates in solution is studied applying LSL.

**Table 3.** Visually evaluated solubility of lactonic diacetylated sophorolipid (1A) (LSL) and acidic deacetylated sophorolipid (1B) (ASL) in selected solvents sorted by increasing solvent polarity (normalized molar transition energy ($E_T^{\perp}$) values [77,78]). (n$_{LSL}$ = n$_{ASL}$ = 0.145 mmol; solvent volume = 2 mL; + = soluble, − = not soluble).

| Solvent                          | $E_T^{\perp}$/kJ mol$^{-1}$ [77,78] | Solubility LSL | Solubility ASL |
|---------------------------------|-------------------------------------|----------------|----------------|
| Petroleum ether                 | −                                   | −              | −              |
| Cyclohexane                     | 0.025                               | −              | −              |
| n-Hexane                        | 0.038                               | −              | −              |
| Toluene                         | 0.414                               | +              | −              |
| Diethyl ether                   | 0.490                               | −              | −              |
| tert-Butyl methyl ether (MTBE)  | 0.519                               | −              | −              |
| 1,4-Dioxane                     | 0.687                               | +              | −              |
| Tetrahydrofuran (THF)           | 0.867                               | +              | −              |
| Ethyl acetate                   | 0.955                               | −              | −              |
| Chloroform                      | 1.084                               | +              | −              |
| Dichloromethane                 | 1.294                               | +              | −              |
| 2-Butanone (MEK)                | 1.369                               | +              | −              |
| Acetone                         | 1.486                               | +              | −              |
| N-Methylpyrrolidin-2-one (NMP)  | 1.486                               | +              | −              |
| N,N-Dimethylformamide (DMF)     | 1.616                               | +              | +              |
| Dimethyl sulfoxide (DMSO)       | 1.859                               | +              | +              |
| Acetonitrile                    | 1.926                               | +              | −              |
| 2-Propanol                      | 2.286                               | −              | −              |
| Ethanol                         | 2.738                               | −              | −              |
| Methanol                        | 3.190                               | +              | +              |
| Water                           | 4.187                               | +              | +              |

1 not listed.

3.1.2. Reaction of LSL with Monoisocyanates

LSL comprises four secondary OH groups (Figure 1 (1A)). The extent of the conversion of the OH groups can be investigated by reaction with monofunctional
isocyanates HIC and PIC. Assuming complete conversion, the reactions of LSL with HIC or PIC should yield products with $M_n,\text{theo,LSL-HIC} = 1197 \text{ g mol}^{-1}$ and $M_n,\text{theo,LSL-PIC} = 1165 \text{ g mol}^{-1}$, respectively. As shown in Figure 8, the experimental molecular weights are determined to be in the range of about 1000 g mol$^{-1}$ for all systems applying slight excess of isocyanate. This means that on average, 2.5 OH groups of the LSL reacted with the isocyanate. In contrast to this, a five-fold excess of PIC must yield a product with $M_n,\text{theo,LSL-PIC} = 1155 \text{ g mol}^{-1}$, thus indicating complete conversion. In addition to the urethane products, by-product formation is observed (Figure 8). Residual isocyanates could not be detected in a significant amount. For the PIC systems with low isocyanate excess, the molecular weight of the by-product is about 140 g mol$^{-1}$ and can be assigned to phenyl urea ($M_n = 136.15 \text{ g mol}^{-1}$) being an impurity of PIC. It is well-known that monoisocyanates dimerize to form uretdiones [79,80]. According to $^{13}$C-NMR analysis and conversion-time-investigations (results not shown) [81], the signals at ca. 250 g mol$^{-1}$ correspond to the uretdione dimers of HIC ($M_n,\text{theo,HIC-uretdion} = 254.37 \text{ g mol}^{-1}$) and PIC ($M_n,\text{theo,PIC-uretdion} = 238.24 \text{ g mol}^{-1}$), respectively. The dimerization explains thus excellently the incomplete conversion of OH groups of LSL applying minor excess of isocyanate. With large excess, however, uretdione formation becomes prominent, but remains insufficient to prevent completion of the reaction.

![Figure 8](image.jpg)

**Figure 8.** Size exclusion chromatograms of products from the reaction of LSL with (a) 1-isocyanatohexane (HIC); (b) phenylisocyanate (PIC) applying different reaction conditions (Table 1).

3.1.3. Reaction of LSL with Diisocyanates

**Stoichiometric Influence**

To produce high molecular weight polyurethanes, LSL has to be converted with isocyanates with a functionality $f$ of at least two. In contrast to the reaction with monofunctional isocyanates, the reaction with HDI does not lead to measurable uretdione formation. However, because $f_{\text{LSL}}$ is larger than two, crosslinking is substantial and thus the point of gelation must be considered when different stoichiometric NCO/OH ratios (indices) are applied. The point of gelation can be calculated applying the Flory–Stockmayer relation (1) [44–46].

$$
(p_{\text{OH}} \times p_{\text{NCO}})_{\text{gel}} = (f_{\text{OH}} - 1)^{-1}(f_{\text{NCO}} - 1)^{-1}
$$

with $p_{\text{OH}}$ and $p_{\text{NCO}}$ = conversion of OH and NCO groups, respectively, and $f_{\text{OH}}$ and $f_{\text{NCO}}$ = functionality of the OH and the NCO component. For the LSL/HDI system, the point of gelation is calculated to be 1/3. Thus, gelation is to be expected within an index range of 0.33 to 3.03. A series of experiments applying different indices allows detailed inspection.
of the reaction of LSL with HDI (Figure 9). The reaction of LSL with 10-fold excess of HDI leads to a product mixture that is still soluble in THF. SEC shows the nature of the components of this mixture (Figure 9, dotted line). Peak c ($M_n = 2190 \, \text{g mol}^{-1}$) relates to the oligomeric/polymeric adducts formed by the reaction of LSL with HDI. In addition to this, low molecular weight fractions can be detected (a and b Peaks in Figure 9), which correspond to residual HDI and to mono- and diurethanes from quenching residual HDI with excessive MeOH. Reducing the index to 1.1 leads to a product mixture that is not completely soluble any more, indicating significant crosslinking as predicted by the Flory–Stockmayer (Equation (1)). SEC analysis of the soluble fraction proves crosslinking/branching to a substantial extent (Figure 9, dashed line). Inverting the NCO/OH ratio to an index of 0.5 leads to a product with incomplete crosslinking (Figure 9, solid line). This appears to be contradicting the prediction for the lower limit of 0.33 using the Flory–Stockmayer relation. Presumably, unreacted OH groups are sterically shielded by vicinal urethane groups formed first. This reduces the effective amount of OH groups for the reaction with HDI. Consequently, the effective lower limit of the Flory–Stockmayer gelation range is somewhat increased and seems to be larger than 0.5.

![Figure 9](image-url)

**Figure 9.** Size exclusion chromatograms of products from the reaction of LSL with HDI applying different NCO/OH ratios (index). Reaction conditions: solvent = acetone; $T = 50 \, ^\circ\text{C}$; $c_{\text{MeOH}} = 500 \, \text{ppm}$. Peak a corresponds to residual HDI, b to mono- and diurethanes from quenching residual HDI with excessive MeOH, and c to oligomers/polymers from the reaction of LSL with 10-fold excess of NCO groups.

**Kinetic Investigations**

The reaction between OH and NCO groups is generally accepted to follow second order kinetics [36,82] and can be studied by investigation of the corresponding second order plots and determination of $k_{\text{app}}$ according to Equation (2) [38].

$$c_{\text{NCO}}^{-1} = k_{\text{app}} \, t + c_{\text{NCO}}^{-1}$$  \hspace{1cm} (2)

with $c_{\text{NCO}} =$ concentration of NCO groups at time $t$ and $c_{\text{NCO}} =$ concentration of NCO groups at the beginning of the reaction. Figure 10 shows second order kinetic plots of reactions of PIC, HIC, and HDI with LSL and PBA, respectively. The relative reactivity of LSL and PBA towards the isocyanates is similar and follows the order PIC > HIC > HDI. All determined $k_{\text{app}}$ are in the typical range for rate constants of conversions of isocyanates with primary or secondary alcohols [83]. In fact, PBA reacts with the corresponding isocyanates about 2.5 to 3.5 times faster than LSL (compare $k_{\text{app}}$ in Figure 10a with the corresponding $k_{\text{app}}$ in Figure 10b). This is because PBA is terminated by primary OH groups, whereas LSL contains secondary OH groups only. The difference in the reactivity
is also observable in the second order plots of ternary reaction mixtures comprising isocyanate and an equimolar mixture of PBA and LSL (Figure 11). From this, two different $k_{\text{app}}$ for each reaction can be calculated being $k_{\text{app1}} = 16.2 \times 10^{-4}$ L mol$^{-1}$ s$^{-1}$ and $k_{\text{app2}} = 7.99 \times 10^{-4}$ L mol$^{-1}$ s$^{-1}$ for the PBA/LSL/PIC mixture and $k_{\text{app1}} = 4.64 \times 10^{-4}$ L mol$^{-1}$ s$^{-1}$ and $k_{\text{app2}} = 2.61 \times 10^{-4}$ L mol$^{-1}$ s$^{-1}$ for the PBA/LSL/HIC reaction. The differences between $k_{\text{app1}}$ and $k_{\text{app2}}$ is about a factor of two in each ternary system, suggesting that LSL and PBA do not affect each other in terms of reactivity towards isocyanates.

**Figure 10.** Second order kinetic plots of reactions of PIC, HIC, and HDI with (a) LSL; (b) commercially available polyester PBA. Reaction conditions: solvent = acetone; $T = 50$ °C; $c_{\text{DBTDL}} = 500$ ppm; index = 1.1.

**Figure 11.** Second order kinetic plots of reactions of PIC and HIC with an equimolar mixture of LSL and PBA. Reaction conditions: solvent = acetone; $T = 50$ °C; $c_{\text{DBTDL}} = 500$ ppm; index = 1.1. Rate constants $k_{\text{app}}$ shown in the inlet are in $\times 10^{-4}$ L mol$^{-1}$ s$^{-1}$.

**Behavior of Ternary Systems**

Ternary PU systems containing LSL, PBA, and HDI are well-suited model formulations to study the influence of LSL on the product performance. For this, the content of LSL is systematically varied in the polyol mixture. To prevent significant crosslinking and enable reasonably high molecular weights, an index of 0.5 is applied. SEC analysis reveals a monomodal molecular weight distribution for the binary system PBA/HDI (0 mol.% LSL) with an average molecular weight of $M_n = 4820$ g mol$^{-1}$, which agrees well to $M_{n,\text{theo}}$ of 4619 g mol$^{-1}$ according to Flory (Figure 12) [84]. In contrast to that, increasing the amount of LSL in the polyol mixture leads to a broadening of the molecular weight distribution and an increase of modality. The SECs of the samples containing 70
mol % and 50 mol % LSL in particular reveal signals at molecular weights that correspond to pure LSL. This strongly suggests incomplete incorporation of LSL in the PU system. This is most likely because PBA is consumed first due to the reduced reactivity of the secondary OH groups of LSL. Consequently, residual LSL remains in the reaction mixture. An increased amount of LSL in the product leads also to a reduction of properties such as shore A hardness and crystallinity, suggesting a plasticizing character of LSL (Figure 13).

![Figure 12](image-url)  
**Figure 12.** Size exclusion chromatograms of products from the reaction of HDI with LSL/PBA polyol mixtures comprising different molar LSL fractions. Reaction conditions: solvent = acetone; T = 50 °C; cDBTDL = 500 ppm; index = 0.5. Peak a corresponds to residual LSL, peak b to LSL-HDI-LSL trimer.

![Figure 13](image-url)  
**Figure 13.** Crystallinity obtained by XRD and shore A hardness of products from the reaction of HDI with LSL/PBA polyol mixtures comprising different molar LSL fractions (xLSL). Reaction conditions: solvent = acetone; T = 50 °C; cDBTDL = 500 ppm; index = 0.5.

3.2. (ω-1) HFA from LSL as Polyol Component for PU Systems

3.2.1. (ω-1) HFA-Based Polyester Polyols and PU Systems Thereof

The preparation of (ω-1) HFA (2) proceeds according to the sequence shown in Figure 14. Successive alkaline and acidic hydrolysis lead to a mixture of monomeric HFA with its dimers and some oligomers up to an overall yield of 74.5%. Further purification to produce pure monomeric HFA is possible by repeated saponification and solvent
OH-terminated A/B-type polyesters can be produced by modifying a procedure published elsewhere [61]. In this synthesis, the mixture of monomeric and dimeric/oligomeric (ω-1) HFA is polymerized in presence of 1,6-hexanediol. The product is highly viscous, sticky, and not completely soluble in THF (Mn of soluble fraction ≈ 2200 g mol⁻¹). For comparison, a polyester consisting of 12-hydroxy stearic acid and 1,6-hexanediol is produced applying equal conditions. This product is oily and highly viscous as well; however, remains fully soluble in THF (Mn ≈ 2600 g mol⁻¹). These outcomes support other investigations [81], which indicate that notable crosslinking has occurred during the synthesis of (ω-1) HFA-based polyester diols and is most likely the result of inseparable glucose impurities remaining in the (ω-1) HFA.

The reaction of HDI with the soluble fraction of (ω-1) HFA-based polyester diols yields a dark brownish oily product that is not completely soluble in THF. IR and SEC analysis reveals the product to be PU with Mn = 3500 g mol⁻¹ and a significant amount of species with Mn > 10⁶ g mol⁻¹ (results not shown). This is also attributed to residual glucose moieties leading to notable crosslinking and branching during PU synthesis.

3.2.2. Direct Conversion of (ω-1) HFA to PU by Curtius Rearrangement

A/B type PU (6) can be obtained by Curtius rearrangement [51,62–68], thus, fatty acid derivatives such as (ω-1) HFA represent potential A/B type monomers for the synthesis of PU [69]. By modification of model reactions [72,73], a reaction procedure comprising (ω-1) HFA (2) → (ω-1) HFA azide (5) → (ω-1) HFA-based A/B PU (6) could be developed (Figure 15). This sequence leads to macromolecular species with Mn of max. 30,000 g mol⁻¹ after extended reaction time (Figure 16a). With a reaction temperature of 60 °C, the product obtained after 16 h shows a notably high PDI that indicates significant side reactions such as crosslinking, branching, or the formation of macrocycles. Reduction of the reaction temperature to 50 °C leads to species with Mn of about 15,000 g mol⁻¹ and PDIs in the range between 1.5 and 2.0. An increase of the temperature to 80 °C leads to a significant
reduction of obtained molecular weight most likely because of favored formation of macrocycles [51,69].

\[
\text{HO} \quad \text{2} \quad \text{OH}
\]

1. oxalyl chloride, DMF, \( \text{CH}_2\text{Cl}_2 \), 0 °C, 1h
2. \( \text{NaN}_3 \), acetone, 0 °C, 0.5h

\[
\text{N}_3 \quad \text{5} \quad \text{OH}
\]

\[ \Delta, \text{DBTDL, THF} \]

\[
\text{O} \quad \text{H}
\]

\[ \text{N} \quad \text{6} \quad \text{O} \]

**Figure 15.** Reaction scheme for the reaction of (ω-1) HFA (2) producing (ω-1) HFA azide (5) and successive A/B type polymerization via Curtius rearrangement.

**Figure 16.** (a) Evolution of molecular weight and PDI of A/B type polymerization of (ω-1) HFA azide (5) via Curtius rearrangement determined by SEC; (b) reaction monitoring of the polymerization by FT-IR spectroscopy for \( T = 60 \) °C.

FT-IR reaction monitoring of the polymerization step clearly shows the typical strong asymmetric stretching frequency of triatomic N\(_3\) groups at 2150 cm\(^{-1}\) for the starting material (Figure 16b) [85]. The reaction signals at 1795, 1720, and 1540 cm\(^{-1}\) become distinct after 16 h, indicating the formation of the urethane functionality [86–89]. In addition to these signals, the asymmetric stretch vibration of the NCO group [85,86,90,91] at 2264 cm\(^{-1}\) becomes evident after 30 minutes, suggesting effective decomposition of the azide. Note that this signal is still observable after 16 h reaction time, indicating incomplete conversion and most likely the formation of NCO-terminated species.
4. Conclusions

In contrast to acetic and deacetylated sophorolipid (ASL), the lactonic and acetylated form of sophorolipid (LSL) is soluble in solvents commonly used in PU synthesis. The conversion of LSL with mono- or diisocyanates is feasible and the products obtained show the expected behavior for PU systems based on polyols with functionalities higher than two, i.e., crosslinking in the reaction with diisocyanates applying indices below the point of gelation. The reactivity of LSL is in the typical range for conversions of isocyanates with secondary alcohols and about two to three times lower compared to polyols containing exclusively primary OH end groups such as commercially available PBA. This difference leads to incomplete incorporation of LSL in, e.g., ternary HDI/PBA/LSL systems applying an index of 0.5. Consequently, LSL acts here like a plasticizer reducing for instance crystallinity and shore A hardness. (ω-1) HFA-based polyester polyols are in general producible; however, low amounts of residual glucose impurities seem to lead to considerable crosslinking during the synthesis. Subsequent reaction of soluble fractions of this polyester with HDI leads to different PU products with Mn of about 3500 g mol⁻¹ and >10⁶ g mol⁻¹, respectively, indicating substantial crosslinking and branching. (ω-1) HFA-based A/B type PU systems are also feasible by applying a reaction procedure comprising (ω-1) HFA (2) → (ω-1) HFA azide (5) → (ω-1) HFA-based A/B PU (6). Such systems show molecular weights up to ca. 30,000 g mol⁻¹. The products obtained show a significant amount of NCO groups remaining in the system, suggesting incomplete conversion and formation of NCO-terminated PU species.

**Author Contributions:** Writing — original draft preparation, M.S., S.G.A., M.C.L.; writing — review and editing, M.S., S.G.A., A.M.S., M.C.L.; supervision, A.M.S., M.C.L.; project administration, M.C.L.; funding acquisition, M.C.L. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the Fachagentur Nachwachsende Rohstoffe e.V. (FNR) as promoter of Germanys Federal Ministry of Food and Agriculture (BMEL). Project number: 22013514.

**Acknowledgments:** The authors would like to acknowledge Christian Zerhusen and Ulrich Schörken for the synthesis of sophorolipids and their fruitful discussions.

**Conflicts of Interest:** The authors declare no conflicts of interest.

**References**

1. Bayer, O. Das Di-Isocyanat-Polyadditionsverfahren (Polyurethane). Angew. Chem. 1947, 59, 257–272, doi:10.1002/ange.19470590901.
2. Bayer, O.; Rinke, H.; Siefken, W. Verfahren zur Herstellung von Polyurethenen bzw. Polyharnstoffen, German Patent 728981, 13 November 1937.
3. Oertel, G.; Becker, G.W.; Braun, D. (Eds.) Polyurethane, 3rd ed.; Hanser: München, Germany, 1993; ISBN 3446162631.
4. Breton, G.; Emanuel, R.; Lomax, R.; Pennington, K.; Ryan, T.; Tebbe, H.; Timm, M.; Ware, P.; Winkler, K.; Yuan, T.; et al. Polyurethanes. Ullmann’s Encyclopedia of Industrial Chemistry; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2000; pp. 1–76, ISBN 9783527306732.
5. Meier-Westhues, U.; Danielmeier, K.; Kruppa, P.; Squiller, E. Polyurethanes: Coatings, Adhesives and Sealants; 2nd ed.; Vincentz Network: Hannover, Germany, 2019; ISBN 3866307829.
6. Müller, B.; Rath, W. Formulierung von Kleb- und Dichtstoffen: Das kompetente Lehrbuch für Studium und Praxis; 2nd ed.; Vincentz Network: Hannover, Germany, 2009; ISBN 9783866308626.
7. Yilgör, I.; Yilgör, E.; Wilkes, G.L. Critical parameters in designing segmented polyurethanes and their effect on morphology and properties: A comprehensive review. Polymer 2015, 58, A1–A36, doi:10.1016/j.polymer.2014.12.014.
8. Ionescu, M. Chemistry and Technology of Polys or Polyols for Polyurethanes; Rapra Technology Ltd.: Shawbury, UK, 2005; ISBN 9781847350350.
9. Bundesministerium für Ernährung und Landwirtschaft. Neue Produkte: Aus Natur gemacht—Nachwachsende Rohstoffe im Alltag; Bundesministerium für Ernährung und Landwirtschaft: Berlin, Germany, 2018.
10. Athawale, V.D.; Nimbalkar, R.V. Polyurethane Dispersions Based on Sardine Fish Oil, Soybean Oil, and Their Interesterification Products. J. Dispers. Sci. Technol. 2011, 32, 1014–1022, doi:10.1080/01932691.2010.497459.
11. Azam Ali, M.; Ooi, T.L.; Salmiaa, A.; Ishiaku, U.S.; Ishak, Z. New polyester acrylate resins from palm oil for wood coating application. J. Appl. Polym. Sci. 2001, 79, 2156–2163, doi:10.1002/1097-4628(20010321)79:12<2156::AID-APP1023>3.0.CO;2-K.

12. Brister, E.H.; Johnston, T.; King, C.L.; Thames, S.F. New Monomers from Vegetable Oils. In Specialty Monomers and Polymers; Havelka, K.O., Mc Cormick, C.L., Eds.; ACS Symposium Series; American Chemical Society: Washington, DC, USA, 2000; Volume 755, pp. 159–169.

13. Bunker, S.P.; Wool, R.P. Synthesis and characterization of monomers and polymers for adhesives from methyl oleate. J. Polym. Sci. A Polym. Chem. 2002, 40, 451–458, doi:10.1002/pola.10130.

14. Cramail, H.; Boyer, A.; Epoune Lingome, C.; Alfos, C.; Gadenne, B.; Cloutet, E.; Queneau, Y.; Moebes, S. Preparation of Polyurethanes and Polysters from Glycolipid Type Compounds. U.S. Patent 20104235814, 2 October 2012.

15. Homann, J.G.; Yu, X.-H.; Connor, T.J.; Cooper, S.L. Castor oil based UV-curable polyurethane-acrylate interpenetrating networks. J. Appl. Polym. Sci. 1991, 43, 2249–2257, doi:10.1002/app.1991.070431214.

16. Kalita, H.; Kalita, D.; Alam, S.; Chernykh, A.; Tarnavchyk, I.; Bahr, J.; Samanta, S.; Jayasooriyama, A.; Fernando, S.; Selvakumar, S.; et al. Novel Biobased Polymers for Coating Applications. In Biobased and Environmental Benign Coatings; Tiwari, A., Galanis, A., Soucek, M.D., Eds.; Scrivener Publishing: Hoboken, NJ, USA; John Wiley & Sons: Salem, MA, USA, 2016; pp. 1–16, ISBN 9781119185055.

17. Noreen, A.; Zia, K.M.; Zubcr, M.; Tabasum, S.; Zahoor, A.F. Bio-based polyurethane: An efficient and environment friendly coating systems: A review. Prog. Org. Coat. 2016, 91, 25–32, doi:10.1016/j.porgcoat.2015.11.018.

18. Tiwari, A.; Galanis, A.; Soucek, M.D. (Eds.) Biobased and Environmental Benign Coatings; Scrivener Publishing: Hoboken, NJ, USA; John Wiley & Sons: Salem, MA, USA, 2016; ISBN 9781119185055.

19. Bollmann, T.; Zerhusen, C.; Glüsen, B.; Schörken, U. Structures and Properties of Sophorolipids in Dependence of Microbial Strain. Lipid Substrate and Post-Modification. Tenside Surfactants Deterg. 2015, 56, 367–377, doi:10.3139/113.110640.

20. Daniel, H.-J.; Reuss, M.; Syldatk, C. Production of sophorolipids in high concentration from deproteinized whey and rapeseed oil in a two stage fed batch process using Candida bombicola ATCC 22214 and Cryptococcus curvatus ATCC 20509. Biotechnol. Lett. 1998, 20, 1153–1156, doi:10.1023/A:1005332605003.

21. Develte, D.W.G.; Laursen, L.M. Properties and industrial applications of sophorolipids. Eur. J. Lipid Sci. Technol. 2010, 112, 628–638, doi:10.1002/ejlt.200901535.

22. Dolman, B.M.; Kaisermann, C.; Martin, P.J.; Winterburn, J.B. Integrated sophorolipid production and gravity separation. Process Biochem. 2017, 54, 162–171, doi:10.1016/j.procbio.2016.12.021.

23. Konishi, M.; Fukuoka, T.; Morita, T.; Iimura, T.; Kitamoto, D. Production of new types of sophorolipids by Candida batistae. J. Oleo Sci. 2008, 57, 359–369, doi:10.5650/jos.57.359.

24. Price, N.P.J.; Ray, K.J.; Vermillion, K.E.; Dunlap, C.A.; Kurtzman, C.P. Structural characterization of novel sophorolipid biosurfactants from a newly identified species of Candida yeast. Carbohydr. Res. 2012, 346, 33–41, doi:10.1016/j.carres.2011.07.016.

25. Zerhusen, C.; Brotsmann, J.; Barbe, S.; Schörken, U. Process optimization concepts for cost-efficient production of sophorolipids. Chem. Eng. Tech. 2018, 40, 1273, doi:10.1002/ceat.20185510.

26. Rau, U.; Hammer, S.; Heckmann, R.; Wray, V.; Lang, S. Sophorolipids: A source for novel compounds. Ind. Crop. Prod. 2001, 13, 85–92, doi:10.1016/S0926-6690(00)00055-8.

27. Zerhusen, C.; Bollmann, T.; Gödderz, A.; Fleischer, P.; Glüsen, B.; Schörken, U. Microbial Synthesis of Nonionic Long-Chain Sophorolipid Emulsifiers Obtained from Fatty Alcohol and Mixed Lipid Feeding. Eur. J. Lipid Sci. Technol. 2020, 122, 1900110, doi:10.1002/ejlt.201900110.

28. Kurtzman, C.P.; Price, N.P.J.; Ray, K.J.; Kuo, T.-M. Production of sophorolipid biosurfactants by multiple species of the Starmerella (Candida) bombicola yeast clade. FEMS Microbiol. Lett. 2010, 311, 140–146, doi:10.1111/j.1574-6968.2010.02082.x.

29. Delbeke, E.I.P.; Everaert, J.; Uitterhagen, E.; Verweire, S.; Verlee, A.; Talou, T.; Soetaert, W.; van Bogaert, I.N.A.; Stevens, C.V. Petrosoinic acid purification and its use for the fermentation of new sophorolipids. AMB Express 2016, 6, 28, doi:10.1186/s13568-016-0199-7.

30. Ashby, R.D.; Solaiman, D.K.Y.; Foglia, T.A. Property control of sophorolipids: Influence of fatty acid substrate and blending. Biotechnol. Lett. 2008, 30, 1093–1100, doi:10.1007/s10529-008-9653-1.

31. Delbeke, E.I.P.; Movsiyan, M.; van Geem, K.M.; Stevens, C.V. Chemical and enzymatic modification of sophorolipids. Green Chem. 2016, 18, 76–104, doi:10.1039/C5GC02187A.

32. Boyer, A.; Lingome, C.E.; Condassamy, O.; Schappacher, M.; Moeb-Sanchez, S.; Queneau, Y.; Gadenne, B.; Alfos, C.; Cramail, H. Glycolipids as a source of polysols for the design of original linear and cross-linked polyurethanes. Polym. Chem. 2013, 4, 296–306, doi:10.1039/C2PY02588B.

33. Becker, H.G.O.; Beckert, R. Organikum: Organisch-Chemisches Grundpraktikum, 24th ed.; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2015; ISBN 352733968X.

34. Wissman, H.G.; Rand, L.; Frisch, K.C. Kinetics of polyetheropolys-diisocyanate reactions. J. Appl. Polym. Sci. 1964, 8, 2971–2978, doi:10.1002/app.1964.070808639.

35. Rand, L.; Thir, B.; Reegen, S.L.; Frisch, K.C. Kinetics of alcohol-isocyanate reactions with metal catalysts. J. Appl. Polym. Sci. 1965, 9, 1787–1795, doi:10.1002/app.1965.070090514.

36. Entelis, S.G.; Nesterov, O.V. Kinetics and Mechanism of the Reactions of Isocyanates with Compounds Containing “Active” Hydrogen. Russ. Chem. Rev. 1966, 35, 917–930, doi:10.1070/RC1966v035n12ABEH001555.
37. Thiele, L. Kinetik der unkatalysierten Urethanbildung. *Monatsh. Chem. Mon. Chem. Mon.* 1992, 123, 865–874, doi:10.1007/BF00811541.

38. Wenning, C.; Schmidt, A.M.; Leimenstoll, M.C. Evolution of Reaction-Induced Phase Segregations in Bio-Soft Segment Urethane Oligomers. *Macromol. Chem. Phys.* 2020, 221, 1900458, doi:10.1002/macp.201900458.

39. Sugihara, J.M. Relative Reactivities of Hydroxyl Groups of Carbohydrates. In *Advances in Carbohydrate Chemistry*; Hudson, C.S., Wolfrom, M.L., Eds.; Academic Press: New York, NY, USA, 1953; pp. 1–44, ISBN 9780120072088.

40. Kurita, K.; Hikakawa, N.; Iwakura, Y. Synthetic polymers containing sugar residues. 1. Direct addition polymerization of D-glucosamine and diisocyanates. *Macromol. Chem.* 1977, 178, 2939–2941, doi:10.1002/macp.1977.021781016.

41. Kurita, K.; Hikakawa, N.; Iwakura, Y. Synthetic polymers containing sugar residues, 2*. Synthesis and properties of poly(urea-urethane) derived from D-glucosamine and diisocyanates. *Makromol. Chem.* 1979, 180, 2331–2340, doi:10.1002/macp.1979.021801007.

42. Kurita, K.; Hikakawa, N.; Iwakura, Y. Synthetic polymers containing sugar residues, 3. Direct addition polymerization of D-cellobiose and diisocyanates to form novel polyurethanes. *Makromol. Chem.* 1979, 180, 855–858, doi:10.1002/macp.1979.021800335.

43. Kurita, K.; Hikakawa, N.; Morinaga, H.; Iwakura, Y. Synthetic polymers containing sugar residues, 6. Novel polyurethanes by direct addition polymerization of α,α-trehalose with diisocyanates. *Makromol. Chem.* 1979, 180, 2769–2773, doi:10.1002/macp.1979.021801120.

44. Stockmayer, W.H. Molecular distribution in condensation polymers. *J. Polym. Sci.* 1952, 9, 69–71, doi:10.1002/pol.1952.120090106.

45. Fogiel, A.W. Effective Functionality and Intramolecular Reactions of Polyisocyanates and Polyls. *Macromolecules* 1969, 2, 581–587, doi:10.1021/ma00021a005.

46. Yang, W.T.; Deng, J.Y.; Jin, G.T. A new gelation theory. *Polym. Bull.* 1994, 33, 733–740, doi:10.1007/BF00296089.

47. Dieterich, V.D.; Reiff, H. Polyurethan-dispersiogenous durch schmelz-dispergiervorverfahren. *Angew. Makromol. Chem.* 1972, 26, 85–106, doi:10.1002/apmc.1972.050260107.

48. Dieterich, D. Neuere währige PUR-Systeme. *Angew. Makromol. Chem.* 1981, 98, 133–165, doi:10.1002/apmc.1981.059801007.

49. Dieterich, D. Aqueous emulsions, dispersions and solutions of polyurethanes; synthesis and properties. *Prog. Org. Coat.* 1981, 9, 281–340, doi:10.1016/0033-0655(81)80002-7.

50. Muthu, H.; Meier, M.A.R. Castor oil as a renewable resource for the chemical industry. *Eur. J. Lipid Sci. Technol.* 2010, 112, 10–30, doi:10.1002/ejlt.200901038.

51. Palaskar, D.V.; Boyer, A.; Cloutet, E.; Alfos, C.; Cramail, H. Synthesis of biobased polyurethane from oleic and ricinoleic acids as the renewable resources via the AB-type self-condensation approach. *Biomacromolecules* 2010, 11, 1202–1211, doi:10.1021/bm100233v.

52. Petrović, Z.S.; Cvetković, I.; Hong, D.; Wan, X.; Zhang, W.; Abraham, T.; Malsam, J. Polyester polyols and polyurethanes from ricinoleic acid. *J. Appl. Polym. Sci.* 2008, 108, 1184–1190, doi:10.1002/app.27783.

53. Radi, B.; Wellard, R.M.; George, G.A. Effect of dangling chains on the structure and physical properties of a tightly crosslinked poly(ethylene glycol) network. *Soft Matter* 2013, 9, 3262, doi:10.1039/C3SM27819K.

54. Herlinger, H.; Müller, P.; Müller-Dolezal, H.; Stoltz, R.; Sill, H. (Eds.) *Houben-Weyl Methods of Organic Chemistry: Macromolecular Compounds II*; Georg Thieme Verlag: Stuttgart, 1994; ISBN 978313180540.

55. Gubrels, E.; Heitz, T.; Yamamoto, M.; Chilekar, V.; Zarbaresh, S.; Gepraegs, M.; Köpnick, H.; Schmidt, M.; Brügg, W.; Rüter, J.; et al. Polyesters. *Ullmann’s Encyclopedia of Industrial Chemistry*; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2000; pp. 1–30, ISBN 978352706732.

56. Kobayashi, S. Lipase-catalyzed polyester synthesis—A green polymer chemistry. *Proc. Jpn. Acad. Ser. B* 2010, 86, 338–365, doi:10.2183/pjab.86.338.

57. Kobayashi, S.; Uyama, H.; Kimura, S. Enzymatic Polymerization. *Chem. Rev.* 2001, 101, 3793–3818, doi:10.1021/ct990121l.

58. Gross, R.A.; Kumar, A.; Kalra, B. Polymer Synthesis by In Vitro Enzyme Catalysis. *Chem. Rev.* 2001, 101, 2097–2124, doi:10.1021/cr0002590.

59. Douka, A.; Vouyiouka, S.; Papaspyridi, L.M.; Papaspyrides, C.D. A review on enzymatic polymerization to produce polycondensation polymers: The case of aliphatic polyesters, polyamides and polyesteramides. *Prog. Polym. Sci.* 2018, 79, 1–25, doi:10.1016/j.progpolymsci.2017.10.001.

60. Ortiz, C.; Ferreira, M.L.; Barbosa, O.; dos Santos, J.C.S.; Rodrigues, R.C.; Berenguer-Murcia, Á.; Briand, L.E.; Fernandez-Lafuente, R. Novozym 435: The “perfect” lipase immobilized biocatalyst? *Catal. Sci. Technol.* 2019, 9, 2380–2420, doi:10.1039/C9CY00415G.

61. Netzger, H.; Bauer, E.; Kleszewski, B.; Schlossmacher, J.; Mayer-Ahrens, S.; Schmidt, M. Method for Producing Flexible Polyurethane Foams. World Patent 2012/069386, 18 November 2011.

62. Smith, P.A.S. The Curtius Reaction. *Org. React.* 1946, 3, 337–449, doi: 10.1002/0471264180.or003.09.

63. Stoermer, R. Umlagerung aromatischer Säureazide in Arylsulfonate. *Ber. Dtsch. Chem. Ges.* 1909, 42, 3133–3134, doi:10.1002/berh.19090420332.

64. Kinstle, J.F.; Sepulveda, L.E. A novel “apparent sublimation” of a polymer. *J. Polym. Sci. B Polym. Lett. Ed.* 1977, 15, 467–469, doi:10.1002/pol.1977.130150804.
65. Ghatge, N.D.; Jadhav, J.Y. Synthesis, characterization, and properties of novel poly(ether urethanes). J. Polym. Sci. Polym. Chem. Ed. 1983, 21, 1941–1950, doi:10.1002/pol.170210706.

66. Kumar, A.; Ramakrishnan, S. A novel one-pot synthesis of hyperbranched polyurethanes. J. Chem. Soc. Chem. Commun. 1993, 1453, doi:10.1039/c3993001453.

67. Kumar, A.; Ramakrishnan, S. Hyperbranched polyurethanes with varying spacer segments between the branching points. J. Polym. Sci. A Polym. Chem. 1996, 34, 839–848, doi:10.1002/(SICI)1099-0518(19960415)34:5<839::AID-POLA13>3.0.CO;2-J.

68. Ranganathan, T.; Ramesh, C.; Kumar, A. Synthesis of new thermotropic liquid crystalline polyurethanes containing biphenyl mesogens using a novel AB-type self-polycondensation. Chem. Commun. 2004, 154–155, doi:10.1039/b312826a.

69. More, A.S.; Gadenne, B.; Alfos, C.; Cramail, H. AB type polyaddition route to thermoplastic polyurethanes from fatty acid derivatives. Polym. Chem. 2012, 3, 1594, doi:10.1039/c2py20123b.

70. Armarego, W.L.F.; Chai, C.L.L. Purification of Laboratory Chemicals, 6th ed.; Elsevier/BH: Oxford, UK, 2009; ISBN 0080878245.

71. Gorin, P.A.J.; Spencer, J.F.T.; Tullloch, A.P. Hydroxy Fatty Acid Glycosides of Sophorose from Torulopsis Magniloliae. Can. J. Chem. 1961, 39, 846–855, doi:10.1139/v61-104.

72. Tale, R.H.; Patil, K.M. 3,4,5-Trifluorobenzeneboronic acid: A mild and versatile catalyst for the one-pot synthesis of acyl azides from carboxylic acids. Tetrahedron Lett. 2002, 43, 9715–9716, doi:10.1016/S0040-4039(02)02256-6.

73. Qiu, Y.; Zhang, Y.; Li, Y.; Ren, J. Discovery of Uracil Derivatives as Potent Inhibitors of Fatty Acid Amide Hydrolase. Molecules 2016, 21, 2299, doi:10.3390/molecules2102229.

74. Joel, D.; Müller, P.; Ahl, R. Isocyanate reactions in and with N,N-dimethylformamide. Angew. Makromol. Chem. 1992, 197, 131–139, doi:10.1002/apmc.199205190111.

75. Weiner, M. Notes: Reaction of Phenyl Isocyanate with N,N-Dimethylformamide. J. Org. Chem. 1960, 25, 2245–2246, doi:10.1021/jo01082a019.

76. Sorenson, W.R. Reaction of an Isocyanate and a Carboxylic Acid in Dimethyl Sulfoxide. J. Org. Chem. 1959, 24, 978–980, doi:10.1021/jo01089a024.

77. Reichardt, C. Solvatochromic Dyes as Solvent Polarity Indicators. Chem. Rev. 1994, 94, 2319–2358, doi:10.1021/cr00032a005.

78. Reichardt, C. (Ed.) Solvents and Solvent Effects in Organic Chemistry, 3rd ed.; Wiley-VCH: Weinheim, Germany, 2004; ISBN 3527306188.

79. Meyer-Stork, L.S.; Höcker, H.; Berndt, H. Syntheses and reactions of urethanes of cellobiose and cellulose-containing uretdione groups. J. Appl. Polym. Sci. 1992, 44, 1043–1049, doi:10.1002/app.1992070440613.

80. Frisch, K.C.; Rumao, L.P. Catalysis in Isocyanate Reactions. J. Macromol. Sci. Polym. Rev. 1970, 5, 103–149, doi:10.1080/15583727008085365.

81. Sonnabend, M. Sphorolipide und Deren Derivate als Biobasierte Bausteine für Polurethansysteme; Universitäts- und Stadtbibliothek Köln: Köln, Germany, 2019.

82. Delebecq, E.; Pascault, J.-P.; Boutevin, B.; Ganachaud, F. On the versatility of urethane/urea bonds: Reversibility, blocked isocyanate, and non-isocyanate polyurethane. Chem. Rev. 2013, 113, 80–118, doi:10.1021/cr300195n.

83. Dyer, E.; Taylor, H.A.; Mason, S.J.; Samson, J. The Rates of Reaction of Isocyanates with Alcohols. I. Phenyl Isocyanate with 1- and 2-Butanols. J. Am. Chem. Soc. 1949, 71, 4106–4109, doi:10.1021/ja01180a046.

84. Flory, P.J. Molecular Size Distribution in Linear Condensation Polymers 1. J. Am. Chem. Soc. 1936, 58, 1877–1885, doi:10.1021/ja01301a016.

85. Eyster, E.H.; Gillette, R.H. The Vibration Spectra of Hydrazoic Acid, Methyl Azido, and Methyl Isocyanate the Thermodynamic Functions of Hydrazoic Acid. J. Chem. Phys. 1940, 8, 369–377, doi:10.1063/1.1750669.

86. Merten, V.R.; Lauerer, D.; Braun, G.; Dahm, M. Über den aufbau von polyurethan-kunststoffen. II. Die struktur von polyurethan-saumstoffen in abhangigkeit von der temperatur, der zeit und der rezeptur; IR-spektroskopische nachweisgrenzen für urethan- und harstoffgruppen in polymeren. Makromol. Chem. 1967, 101, 337–366, doi:10.1002/macp.1967.02101119.

87. Nakayama, K.; Imo, T.; Matsubara, I. Infrared Spectra and Structure of Polyurethane Elastomers from Polytetrahydrofuran, Diphenylmethane-4, 4’-diisocyanate, and Ethylenediame. J. Macromol. Sci. Chem. 1969, 3, 1005–1020, doi:10.1080/10601326908051929.

88. Zharkov, V.; Strikovskiy, A.; Verteleetskaya, T. Amide I absorption band: Description of the urethane group association scheme in polyether urethane elastomers. Polymers 1993, 34, 938–941, doi:10.1021/13322861(93)90211-R.

89. Strikovskiy, A.G.; Zharkov, V.V. Infra-red spectroscopy study of equilibrium association of urethane groups in poly(ether urethane)s. Polymers 1993, 34, 3397–3401, doi:10.1021/13322861(93)90467-O.

90. Hoyer, H. Die Infrarotabsorption der Isocyanatgruppe. Ber. Dtsch. Chem. Ges. 1956, 89, 2677–2681, doi:10.1002/cber.19560891132.

91. Hirschmann, R.P.; Knieley, R.N.; Fassel, V.A. The infrared spectra of alkyd isocyanates. Spectrochim. Acta 1965, 21, 2125–2133, doi:10.1016/0371-1591(65)80228-4.