Review Article

Understanding of ROS-Inducing Strategy in Anticancer Therapy

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Redox homeostasis is essential for the maintenance of diverse cellular processes. Cancer cells have higher levels of reactive oxygen species (ROS) than normal cells as a result of hypermetabolism, but the redox balance is maintained in cancer cells due to their marked antioxidant capacity. Recently, anticancer therapies that induce oxidative stress by increasing ROS and/or inhibiting antioxidant processes have received significant attention. The acceleration of accumulative ROS disrupts redox homeostasis and causes severe damage in cancer cells. In this review, we describe ROS-inducing cancer therapy and the anticancer mechanism employed by prooxidative agents. To understand the comprehensive biological response to certain prooxidative anticancer drugs such as 2-methoxyestradiol, buthionine sulfoximine, cisplatin, doxorubicin, imexon, and motexafin gadolinium, we propose and visualize the drug-gene, drug-cell process, and drug-disease interactions involved in oxidative stress induction and antioxidant process inhibition as well as specific side effects of these drugs using pathway analysis with a big data-based text-mining approach. Our review will be helpful to improve the therapeutic effects of anticancer drugs by providing information about biological changes that occur in response to prooxidants. For future directions, there is still a need for pharmacogenomic studies on prooxidative agents as well as the molecular mechanisms underlying the effects of the prooxidants and/or antioxidant-inhibitor agents for effective anticancer therapy through selective killing of cancer cells.

1. Introduction

Reactive oxygen species (ROS) are generally defined as chemically reactive molecules containing oxygen, produced as a result of cellular metabolism [1]. A moderate level of ROS plays an essential role in the cellular signaling that regulates cell proliferation and cell survival [2]. However, an increase in ROS levels can damage cellular components such as lipids, proteins, and DNA, causing an imbalance between cellular reduction-oxidation (redox) conditions and resulting in the disruption of homeostasis [3]. Chronically increased ROS cause severe cellular damage and lead to carcinogenesis by modulating cell signaling in biological processes including cell proliferation and survival, angiogenesis, and metastasis [4, 5].

Anticancer therapies based on oxidative damage through the acceleration of accumulative ROS or the defective antioxidant system in cancer cells have been developed [2, 6]. Due to uncontrolled metabolic processes during hyperproliferation, cancer cells have a higher basal ROS level than normal cells [7]. Adaptation to excessive ROS conditions in cancer cells has been reported, suggesting they have a higher level of antioxidative capacity and ROS than normal cells [2]. ROS-inducing approaches rely on the fact that increasing the ROS level over the cytotoxic threshold can selectively kill cancer cells. The elevated ROS level breaks the redox homeostasis and consequently causes cancer cell death. If exogenous ROS-generating agents are triggered, the redox-imbalanced cancer cells become more vulnerable than normal cells, thereby leading to cell death [8] (Figure 1). Accordingly,
prooxidative agents have been investigated as anticancer drugs that interrupt redox adaptation and eventually induce cytotoxicity in ROS-dependent cancer cells [9].

In this review, we summarize the mechanisms underlying the effects of anticancer drugs utilized in oxidative stress-inducing chemotherapy for direct or indirect ROS generation. To grasp the biological alterations mediated by prooxidative drugs, the drug-focused pathways were analyzed and visualized using big data-based network analysis software. We also suggest crucial therapeutic strategies for anticancer drugs and provide information regarding potential side effects and drug resistance based on the results of the pathway analysis.

2. Basic Concepts of ROS: Generation and Elimination

Oxygen is an essential molecule for maintaining metabolism and life in organisms. However, the metabolism of oxygen produces highly reactive molecules called ROS, a major source of oxidative stress. There are many types of ROS, including superoxide (O$_2^-$), hydroxyl radicals (OH), hydrogen peroxide (H$_2$O$_2$), and singlet oxygen (¹O$_2$) [10]. The cellular redox state refers to the balance between the oxidized and reduced states in cells. In living organisms, redox equilibrium is important for cellular homeostasis [11]. As previously demonstrated, the impairment of redox homeostasis mediated by an excess of oxidized biological molecules is associated with cellular toxic effects [12]. Accordingly, proper regulation of the redox status through ROS generation and elimination is crucial.

Most endogenous ROS are mainly generated in the mitochondrial electron transport chain (ETC) and NADPH oxidase complex (NOX) [13, 14]. During oxidative phosphorylation, the leakage of electrons by ETC complexes I and III occurs in the inner mitochondrial membrane, leading to the reduction of oxygen into superoxide. Subsequently, superoxide dismutase (SOD) converts superoxide into hydrogen peroxide in the intermembrane space or the matrix of mitochondria [8, 14]. Hydrogen peroxide can be converted into hydroxyl radicals in the presence of Fe$^{2+}$ [15]. Likewise, NOX, a transmembrane enzyme complex consisting of seven subunits, catalyzes the oxidation of NADPH by transferring electrons to molecular oxygen, leading to the production of superoxide [16].

To avoid endogenous ROS overproduction, cells have diverse defense systems to eliminate ROS using antioxidant molecules and enzymes such as glutathione (GSH),...
peroxiredoxin (Prx), thioredoxin (Trx), SOD, and catalase [17]. GSH protects cellular components against oxidative damage through interactions with a cofactor of GSH peroxidase (GPx) and/or participation in other antioxidant components [18, 19]. In the presence of NADPH, GSH reductase catalyzes the reduction of GSH. Two reduced GSH molecules are oxidized into GSH disulfide (GSSG) via a reaction with GPx, which catalyzes the reduction of hydrogen peroxide to water and oxygen molecules through the redox cycle [20, 21]. GSH deficiency has been shown to reduce tissue ascorbate levels and increase oxidative stress, ultimately resulting in diverse disorders such as mitochondrial disease, hepatic injuries, and HIV [19, 22, 23]. Several anticancer drugs and xenobiotics have been developed for GSH-targeted chemotherapies or detoxifying agent-based chemoprevention [24]. Both Prx and Trx, which contain cysteine residues with redox-reactive thiol groups, can scavenge hydrogen peroxide via thiol/disulfide exchange [25]. Hydrogen peroxide is reduced by Prx, which is simultaneously oxidized to form a disulfide bond, and Prx is subsequently reduced by transferring the disulfide bond to Trx [26]. In the presence of NADPH, Trx is reduced by a reaction with Trx reductase [27, 28]. SOD catalyzes the breakdown of superoxide to molecular oxygen and hydrogen peroxide using metal ion cofactors including copper, zinc, and manganese [29, 30]. Catalases reduce hydrogen peroxide to water and oxygen with a manganese ion cofactor [31].

Although cellular antioxidant systems have a vital role in balancing endogenous ROS levels and the redox status for cell protection against oxidative stress [32, 33], exogenously prooxidants-induced ROS levels and an ineffective cellular defense system result in significant imbalance between prooxidants and antioxidants [34], possibly enabling cellular damage and cell death.

3. Application of ROS Induction for Anticancer Strategies

A lot of anticancer therapies have employed antioxidant supplements as a strategy to prevent or treat cancer cells. tert-Butylhydroquinone (tBHQ) mediates the dissociation of Nrf2 via oxidative modification of the Keap1 cysteine residues by ROS generated during the metabolic process [35]. Nrf2 activation promotes the regulation of downstream cytoprotective genes, which play important roles in cancer prevention [36]. Selenocompounds exhibit anticancer effects through potentiating the antioxidative defense system from ROS-induced cellular damage [37, 38] and through redox modification of redox-active, cysteine-rich regions of protein kinase C (PKC), a receptor for tumor promoters [39, 40].

However, controversial issues remain regarding the chemotherapeutic activities of antioxidants. Indeed, it has been widely reported that Nrf2 activation contributes to chemoresistance in cancer cells [41–44]. Additionally, a high concentration of tBHQ has been reported to increase carcinogenic risk [45, 46]. The efficacy and safety of selenium are also actively discussed due to its toxicity and side effects [47, 48]. Thus, chemotherapies involving antioxidants may not be sufficient to kill cancer cells and further studies are needed to determine whether they have unexpected adverse effects.

ROS has double-edged sword characteristics in terms of its low-dose cell signaling and high-dose cytotoxicity [49]. A mild level of ROS regulates cell development and homeostasis, whereas a high level inflicts severe cellular damage [50, 51]. Cancer cells are more sensitive to the presence of prooxidants and the inhibition of antioxidants due to their excessive ROS levels [52–54]. The ROS-inducing approach for killing cancer cells relies on oxidative stress-dependent cytotoxic effects through apoptosis, necroptosis, and autophagic cell death [55].

In the early stages, cancer cells exhibit uncontrolled cell growth and proliferation via the modulation of transcription factors and are vulnerable to DNA damage [56, 57] through therapeutic strategies focused on inducing genetic damage using radiation or oxidative stress [58–60] (Figure 2). In the late stages, metastatic cancers undergo metabolic changes such as increased endogenous antioxidant levels to buffer oxidative stress conditions [61]. Indeed, the GSH/GSSG ratio tends to be lower in circulating melanoma or metastatic cancers, suggesting that late-stage cancers have better antioxidant processes than early-stage cancers [62, 63]. Although NADPH-independent catalase activity has been reported to decrease with cancer progression [64], the remarkable antioxidant capacity is one of the reasons for chemoresistance in advanced cancer cells [65, 66]. ROS-inducing and/or antioxidant-suppressing approaches can be applied appropriately for the treatment of malignant cancer cells. Oxidative stress-modulated therapeutics for attacking cancer cells are being actively researched in the anticancer field [67, 68]. The cell-killing potential of ROS has been harnessed for anticancer therapies with two major approaches: direct ROS generation and antioxidant process inhibition [6].

3.1. Direct ROS Generation. Electrons derived from metabolism and respiratory processes are representative ROS sources in cells [69]. Impairing the respiratory cycles with the alteration of radical intermediates produces superoxide by which motexafin gadolinium and anthracyclines function [69–71]. Motexafin gadolinium, an avid electron acceptor, enhances the therapeutic index of radiotherapy, since it can inhibit the repair activities of cancer cells after irradiation [72, 73]. It is effective in patients with brain tumors, brain metastases, and pediatric gliomas [72]. Indeed, anthracycline-based anticancer drugs such as doxorubicin can induce the chelation of intracellular iron, leading to the accumulation of hydroxyl radicals and ultimately to cell death [74]. These drugs are effective for malignant lymphomas, acute leukemia, and diverse solid tumors [75]. Cisplatin, a well-known anticancer agent with cross-linking activity, directly damages mitochondrial DNA (mtDNA), which leads to ETC impairment [76]. It can also interfere with DNA replication and consequently induce oxidative stress to target cancer cells [77]. The drug is effective for diverse cancer types, especially ovarian cancer [78, 79]. 2-Methoxyestradiol is known to inhibit ETC complex I [80], inducing mitochondrial production of hydrogen peroxide [81]. Subsequently, it rapidly activates c-Jun N-terminal
kinase (JNK), resulting in cytochrome c release and caspase-9 activation to initiate apoptosis [82, 83]. The drug can promote the therapeutic capability of other anticancer agents [84–86]. In vitro and in vivo studies have demonstrated that 2-methoxyestradiol-mediated chemotherapy can inhibit malignant cell proliferation as its own activity or in combination with synergistic drugs [87–90]. The ROS-accelerating anticancer agents described above are listed in Table 1.

Although anticancer drugs with direct ROS-accumulating activity have been shown to be effective for treating different types of cancer, the effects on normal cells are still controversial as they damage not only cancer cells but also normal cells. For instance, the radiosensitizer motexafin gadolinium interrupts the DNA repair process and causes injuries to surrounding normal cells [91]. Additionally, anthracyclines induce cardiotoxicity since their metabolites (e.g., oxygen-centered free radicals) can cause heart failure or cardiomyopathy, with a higher risk for younger patients [92–94]. Cisplatin-induced ototoxicity has been reported, attributed to its direct binding to DNA and consequent activation of the inflammatory cascade [95]. Additionally, liver function

| Name                  | Mechanism of action                                      | Reference |
|-----------------------|----------------------------------------------------------|-----------|
| **Direct ROS generation** |                                                          |           |
| Motexafin gadolinium  | Accepts electrons to form superoxide                      | [69]      |
| Doxorubicin           | Induces chelation of iron to generate hydroxyl radical    | [74]      |
| Cisplatin             | Damages mtDNA and ETC                                     | [76]      |
| 2-Methoxyestradiol    | Inhibits ETC complex I                                    | [80]      |
| **Antioxidant process inhibition** |                                                            |           |
| Buthionine sulfoximine| Binds to enzyme related to GSH synthesis                   | [101]     |
| Imexon                | Binds to thiol to GSH activity disruption                 | [102, 103]|
abnormalities, fatigue, and diarrhea have been reported in patients treated with 2-methoxyestradiol [85, 96, 97].

3.2. Antioxidant Process Inhibition. Although direct ROS induction is one of the effective strategies for treating malignant cancer cells [98], its combination with the disruption of antioxidative processes leads to the best results for overcoming the resistance characteristics of cancer cells. Depletion of GSH activity is regarded as an indirect method of generating oxidative stress. Cells can synthesize GSH via an ATP-dependent process catalyzed by glutamate-cysteine ligase (GCL) and GSH synthetase [99, 100]. For instance, buthionine sulfoximine, a typical GSH synthesis inhibitor, can bind to the GCL site that normally binds to the acceptor amino acid [101]. Imexon, a small-molecule chemotherapeutic agent, is widely used to treat advanced cancers of the breast, lung, and prostate. It can disrupt GSH activity by binding to the thiol functional group of reduced GSH [102, 103] and subsequently deplete the GSH pool for antioxidative activity. Due to a decrease in the GSH level by imexon treatment, loss of the mitochondrial membrane potential and the accumulation of oxidative stress occur in cancer cells.

Although anticancer therapy needs to disrupt, both directly and indirectly, the redox adaptation status of cancer cells, the inhibition of antioxidative enzyme has deleterious side effects on normal cells in tissues and organs. For instance, buthionine sulfoximine is known to be associated with cardiac hypertrophy and heart failure by inducing soluble epoxide hydrolase [104]. Imexon has potential side effects in normal cells due to its cytotoxicity [105–107]. For the future direction of oxidative stress-accelerating anticancer therapy, further study is needed to identify ways to not only reduce the side effects but also increase cancer cell-specific killing efficiency. For instance, cotreatment with antioxidant supplements that attenuate cisplatin-mediated nephrotoxicity through Nrf2 signaling has been investigated [108]. Moreover, plant-derived phytochemicals such as flavonoids and carotenoids that act as both antioxidants and prooxidants to improve the therapeutic effects and to reduce the cytotoxic effect have been reported [109–111].

4. Pathway Analysis to Understand the Process of Prooxidative Cancer Therapy

Identifying biological changes in cancer cells caused by anticancer drugs is meaningful to improve their therapeutic effect. Although several mechanism studies have been actively conducted to determine the mode of action of

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Figure 3: Proposed biological pathways related to prooxidative anticancer drugs. Comprehensive illustration of the drug-gene, drug-cell process, and drug-disease relationships for certain anticancer drugs with prooxidative activity (buthionine sulfoximine, cisplatin, doxorubicin, imexon, 2-methoxyestradiol, and motexafin gadolinium). Green and red lines denote the positive and negative effects of each drug, respectively. The legend for the diagrams is located at the bottom part of the figure. Target proteins (red), drug molecules (green), cell processes (yellow), and diseases (purple) are symbolized and organized in a complex biological network.
Table 2: List of proteins, cell processes, and diseases targeted by anticancer drugs.

| Drugs              | Target type | Relation | Relation effect | Target                                                                 |
|--------------------|-------------|----------|-----------------|----------------------------------------------------------------------|
| 2-Methoxyestradiol | Expression  | Positive | BAX, TP53       |                                                                         |
|                    | Expression  | Negative | HIF1A, IL6, PCNA, TNF, VEGFA |                                                                  |
|                    | Regulation  | Positive | CASP9, MAPK8    |                                                                         |
|                    | Regulation  | Negative | BCL2, HIF1A, SOD2 |                                                                       |
|                    | Cell process | Positive | Apoptosis, autophagy, cell cycle arrest, cell death, DNA damage, mitochondrial damage, oxidative stress, ROS generation |                                                                 |
|                    | Cell process | Negative | Angiogenesis, cell cycle, cell growth, cell invasion, cell proliferation, cell survival, mitochondrial respiration, tumor growth |                                                                 |
|                    | Disease     | Regulation | Negative | Atherosclerosis, breast cancer, hepatocellular carcinoma, melanoma, pancreatic cancer |
| Buthionine         | Expression  | Positive | BCL2, HMOX1, JUN, NFE2L2, SOD2, TNF |                                                                 |
| sulfoximine        | Expression  | Negative | GPX1, IL6, NOS2 |                                                                         |
|                    | Regulation  | Positive | BCL2, CASP3, MAPK14 |                                                                 |
|                    | Regulation  | Negative | GCLC |                                                                         |
|                    | Cell process | Positive | Apoptosis, autophagy, cell death, cytoxicity, DNA damage, lipid peroxidation, oxidative stress, ROS generation |                                                                 |
|                    | Cell process | Negative | Cell growth, cell proliferation, tumor growth |                                                                      |
|                    | Disease     | Regulation | Positive | Cataract, liver injury, necrosis, neurotoxicity, toxicity |                                                                    |
|                    | Disease     | Regulation | Negative | Hepatocellular carcinoma, lung cancer |                                                                      |
| Cisplatin          | Expression  | Positive | ABCC1, BAX, BBC3, BECN1, CASP3, CASP7, CASP8, CYCS, DDIT3, FAS, FASLG, GPT, H2AFX, HMOX1, IL1B, IL6, JUN, NFE2L2, NOS2, TNF, TP53 |                                                                 |
|                    | Expression  | Negative | BCL2, SOD2, XIAP |                                                                         |
|                    | Regulation  | Positive | CASP3, CASP7, CYCS, G6PD, MAPK14, MAPK3, MAPK8, TP53 | SOD1 |                                                                         |
|                    | Regulation  | Negative | |                                                                         |
|                    | Cell process | Positive | Apoptosis, autophagy, cell cycle arrest, cell death, cytoxicity, DNA damage, lipid peroxidation, mitochondrial damage, oxidative stress, ROS generation |                                                                 |
|                    | Cell process | Negative | Angiogenesis, cancer cell growth, cell growth, cell invasion, cell proliferation, cell survival, tumor growth |                                                                      |
|                    | Disease     | Regulation | Positive | Acute kidney injury, kidney disease, liver injury, necrosis, neurotoxicity, renal dysfunction, toxicity |                                                                    |
|                    | Disease     | Regulation | Negative | Breast cancer, colorectal cancer, gastric cancer, hepatocellular carcinoma, lung cancer, lymphoma, melanoma, metastasis, ovarian cancer, pancreatic cancer |                                                                    |
| Doxorubicin        | Expression  | Positive | ABCC1, BAX, BBC3, BECN1, CASP3, CASP7, CASP8, CASP9, CAT, CYCS, DDIT3, FAS, FASLG, GPT, H2AFX, HMOX1, IL1B, IL6, MAPK3, MAPK8, NFE2L2, NOS2, SOD1, TNF, TP53 |                                                                 |
|                    | Expression  | Negative | BCL2, PCNA, VEGFA, XIAP |                                                                         |
|                    | Regulation  | Positive | ANXA5, CASP3, CASP7, CASP8, FAS, GPT, IL6, MAPK14, MAPK3, MAPK8, NOS2, TP53 | HIF1A |                                                                         |
|                    | Regulation  | Negative | |                                                                         |
|                    | Cell process | Positive | Apoptosis, autophagy, cell cycle arrest, cell death, cytoxicity, DNA damage, lipid peroxidation, mitochondrial damage, oxidative stress, ROS generation |                                                                 |
|                    | Cell process | Negative | Angiogenesis, cancer cell growth, cell growth, cell proliferation, cell survival, DNA repair, mitochondrial respiration, tumor growth |                                                                      |
|                    | Disease     | Regulation | Positive | Acute kidney injury, kidney disease, liver injury, necrosis, neurotoxicity, renal dysfunction, toxicity |                                                                    |
|                    | Disease     | Regulation | Negative | Breast cancer, colorectal cancer, gastric cancer, hepatocellular carcinoma, lung cancer, lymphoma, melanoma, metastasis, ovarian cancer, pancreatic cancer |                                                                    |
Table 2: Continued.

| Drugs                      | Target type | Relation | Relation effect | Target                                      |
|----------------------------|-------------|----------|-----------------|---------------------------------------------|
| Doxorubicin, Imexon, Motexa | Protein     | Expression | Negative     | HIF1A                                       |
|                            | Protein     | Regulation | Positive      | CASP3, CASP9                                |
|                            | Cell process | Regulation | Positive      | Apoptosis, cell cycle arrest, oxidative stress |
|                            | Disease     | Regulation | Negative      | Cancer cell growth, cell cycle, cell growth, tumor growth |
|                            | Protein     | Regulation | Negative      | Lymphoma, melanoma, multiple myeloma, splenomegaly |
| Motexa gadolinium          | Cell process | Regulation | Positive      | HMOX1, TXN                                  |
|                            | Disease     | Regulation | Negative      | Atherosclerosis, cerebral neoplasm, glioblastoma, lung cancer, metastasis |

Anticancer drugs for cancer treatment, the efficacy and toxicity of anti- and prooxidants remain controversial. In this regard, pathway analysis has the advantage of comprehensively elucidating the molecular network involved in the response to certain drugs. However, very few studies have been performed to explore biological modulation during treatment with prooxidant anticancer agents. In this review, we explore and visualize key information on drug-gene, drug-cell process, and drug-disease relationships for six anticancer drugs abovementioned with prooxidative activity (2-methoxyestradiol, buthionine sulfoximine, cisplatin, doxorubicin, imexon, and motexa gadolinium) using a text mining-based biological network analysis tool, Pathway Studio ver. 12.2 (Elsevier, USA). This database provides information describing the relationships between the entities including the drugs, genes, cell processes, and diseases through a curated resource based on text mining from biology articles.

Each drug molecule was first inputted to build a network, and then the genes, cell processes, and diseases associated with the drugs were analyzed based on data provided in five or more references (Figure 3). Cisplatin and doxorubicin had the largest networks, implying that these two drugs have been extensively studied compared to the others, while imexon and motexa gadolinium had the fewest connections. Figure 3 comprehensively illustrates the biological pathways including the target genes, key cellular processes, and target types of cancer that can be positively or negatively affected by these anticancer drugs. There were two types of relationships in the identified networks: Expression and Regulation. In Expression relationship, the drug alters the protein abundance by affecting the levels of transcript or protein stability. In Regulation relationships, the drug directly or indirectly changes the activity of the genes, cell processes, and diseases. In addition, we evaluated the possible side effects related to the prooxidant anticancer drugs such as neurotoxicity and cardiovascular diseases. Table 2 summarizes the detailed information obtained from pathway analysis regarding the relationship of each drug with the targeted genes, cell processes, and diseases. We also explored the association of drug resistance with each drug through network analysis.

Based on the high number of references in the pathway analysis, we found that 2-methoxyestradiol is not only a potent inhibitor of HIF1A and VEGFA, which play important roles in angiogenesis [112], it also activates MAPK8, which triggers apoptosis [113]. Consistent with these results, 2-methoxyestradiol has been shown to be closely associated with cellular processes such as apoptosis, cell proliferation, and angiogenesis. Breast cancer, melanoma, and pancreatic cancer were predicted to be major targets for this drug, and atherosclerosis can also be attenuated due to its antiangiogenic effects. Moreover, 2-methoxyestradiol-mediated autophagy promoting cancer cell survival could lead to drug resistance [114].

Buthionine sulfoximine was shown to effectively inhibit GCLC, blocking GSH synthesis [115]. The expression of GPX1 was also found to decrease while that of NFE2L2, HMOX1, and SOD2 increased in direct response to GSH depletion [116]. Oxidative stress, apoptosis, and cell death were identified as the main cell processes induced by buthionine sulfoximine-mediated GSH inhibition. Hepatocellular carcinoma and lung cancer were predicted to be the main target diseases, and cataract can be evoked by increased lipid peroxidation in the lens [117]. The increased NFE2L2 can upregulate ABCC1, which is a cell membrane transporter protein [118]. Accordingly, increased drug efflux through the transporter leads to drug resistance [119]. Buthionine sulfoximine-mediated autophagy can also negatively affect drug sensitivity.

Cisplatin was shown to significantly induce expression of the well-known tumor suppressor TP53 as well as proapoptotic genes such as TNF, BAX, CASP3, and FAS, while decreasing antiapoptotic BCL2 and XIAP expression. Consistently, cell processes including apoptosis, ROS generation, DNA damage, and mitochondrial damage were found to be significantly induced by cisplatin treatment. Diseases effectively targeted by cisplatin were predicted to be ovarian, lung, gastric, and breast cancer. However, cisplatin-induced proinflammatory cytokines IL1B, IL6, and TNF are at risk of causing side effects such as acute kidney injury and renal dysfunction. Cisplatin also plays important roles in drug resistance by inducing autophagy and activating NFE2L2 and ABCC1, which elevate drug efflux.

Doxorubicin was shown to have similar effects to cisplatin on targeted genes and cell processes. It also significantly increases TP53, BAX, TNF, CASP3, and FAS expression and decreases BCL2 and XIAP expression, promoting apoptosis. Oxidative stress, DNA damage, and lipid peroxidation
were suggested to be doxorubicin-mediated cell processes. Doxorubicin is mainly used to treat breast, ovarian, and lung cancer as well as lymphoma, but there is a risk of heart failure and neurotoxicity. Drug resistance in doxorubicin was predicted to be attributable to increased autophagy and the upregulation of NFE2L2 and ABCC1.

Imexon was found to positively regulate the activity of CASP3 and CASP9 which have critical roles in apoptosis. Oxidative stress and cell cycle arrest can be stimulated by imexon, which was predicted to have therapeutic effects on multiple myeloma and splenomegaly.

Motexafin gadolinium was shown to inhibit the activity of TXN and HMOX1, leading to apoptosis. It was suggested to exhibit anticancer effects by promoting ROS generation and oxidative stress and by disrupting the DNA repair process. Motexafin gadolinium was expected to target diseases including lung cancer and cerebral neoplasm.

5. Conclusions

Redox homeostasis plays an essential role in maintaining diverse cellular processes [120]. The disruption of redox homeostasis is being actively investigated in the field of chemotherapy since cancer cells can be effectively killed by accelerating their oxidative stress state. In this review, we presented an overview of ROS-inducing anticancer therapy and the anticancer strategy using prooxidative agents in terms of direct and indirect ROS accumulation. For a comprehensive understanding of biological network of prooxidant drugs and molecular targets, our pathway analysis highlighted the crucial effects of each anticancer drug on genes, cell processes, and diseases related to ROS generation and antioxidant inhibition. Our explanation of changes in biological processes relevant to specific drugs and potential side effects would be meaningful for better understanding of the toxicological aspects as well as for predicting the efficacy of chemotherapies using prooxidative anticancer drugs with undetectable side effects. Although several previous studies have investigated the modes of action for prooxidant drugs, pharmacogenomic studies evaluating the drug treatments are still required to elucidate the exact anticancer mechanisms and potential molecular targets. Our review will help researchers better understand the current gene-targeting anticancer strategies involving prooxidative drugs in order to overcome their controversial side effects.

Conflicts of Interest

The authors declare no conflict of interest.

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References

[1] K. Brieger, S. Schiavone, J. Miller, and K. H. Krause, “Reactive oxygen species: from health to disease,” Swiss Medical Weekly, vol. 142, article w13659, 2012.
[2] D. Trachootham, J. Alexandre, and P. Huang, “Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach?,” Nature Reviews Drug Discovery, vol. 8, no. 7, pp. 579–591, 2009.
[3] C. E. Cross, B. Halliwell, E. T. Borish et al., “Oxygen radicals and human disease,” Annals of Internal Medicine, vol. 107, no. 4, pp. 526–545, 1987.
[4] A. Glasauer and N. S. Chandel, “Targeting antioxidants for cancer therapy,” Biochemical Pharmacology, vol. 92, no. 1, pp. 90–101, 2014.
[5] S. C. Gupta, J. H. Kim, S. Prasad, and B. B. Aggarwal, “Regulation of survival, proliferation, invasion, angiogenesis, and metastasis of tumor cells through modulation of inflammatory pathways by nutraceuticals,” Cancer Metastasis Reviews, vol. 29, no. 3, pp. 405–434, 2010.
[6] J. Wang and J. Yi, “Cancer cell killing via ROS: to increase or decrease, that is the question,” Cancer Biology & Therapy, vol. 7, no. 12, pp. 1875–1884, 2008.
[7] P. Storz, “KRas, ROS and the initiation of pancreatic cancer,” Small GTPases, vol. 8, no. 1, pp. 38–42, 2017.
[8] M. H. Raza, S. Siraj, A. Arshad et al., “ROS-modulated therapeutic approaches in cancer treatment,” Journal of Cancer Research and Clinical Oncology, vol. 143, no. 9, pp. 1789–1809, 2017.
[9] C. Martin-Cordero, A. J. Leon-Gonzalez, J. M. Calderon-Montano, E. Burgos-Moron, and M. Lopez-Lazaro, “Pro-oxidant natural products as anticancer agents,” Current Drug Targets, vol. 13, no. 8, pp. 1006–1028, 2012.
[10] K. J. Davies, “Oxidative stress: the paradox of aerobic life,” Biochemical Society Symposium, vol. 61, pp. 1–31, 1995.
[11] H. Sies, “Oxidative stress: a concept in redox biology and medicine,” Redox Biology, vol. 4, pp. 180–183, 2015.
[12] M. Valko, H. Morris, and M. T. Cronin, “ Metals, toxicity and oxidative stress,” Current Medicinal Chemistry, vol. 12, no. 10, pp. 1161–1208, 2005.
[13] Y. Liu, G. Fiskum, and D. Schubert, “Generation of reactive oxygen species by the mitochondrial electron transport chain,” Journal of Neurochemistry, vol. 80, no. 5, pp. 780–787, 2002.
[14] J. L. Meitzler, S. Antony, Y. Wu et al., “NADPH oxidases: a perspective on reactive oxygen species production in tumor biology,” Antioxidants & Redox Signaling, vol. 20, no. 17, pp. 2873–2889, 2014.
[15] S. Chen, X. F. Meng, and C. Zhang, “Role of NADPH oxidase-mediated reactive oxygen species in podocyte injury,” BioMed Research International, vol. 2013, Article ID 839761, 7 pages, 2013.
[16] G. Minotti and S. D. Aust, “The requirement for iron (III) in the initiation of lipid peroxidation by iron (II) and hydrogen peroxide,” The Journal of Biological Chemistry, vol. 262, no. 3, pp. 1098–1104, 1987.
[17] C. Nicco and F. Batteux, “ROS modulator molecules with therapeutic potential in cancers treatments,” Molecules, vol. 23, no. 1, p. 84, 2017.
[18] R. Bigley, M. Riddle, D. Layman, and L. Stankova, “Human cell dehydroascorbate reductase. Kinetic and functional
properties,” Biochimica et Biophysica Acta (BBA) - Enzymology, vol. 659, no. 1, pp. 15–22, 1981.

[19] J. Martensson and A. Meister, “Glutathione deficiency decreases tissue ascorbate levels in newborn rats: ascorbate spares glutathione and protects,” Proceedings of the National Academy of Sciences, vol. 88, no. 11, pp. 4656–4660, 1991.

[20] F. J. Giblin, “Glutathione: a vital lens antioxidant,” Journal of Ocular Pharmacology and Therapeutics, vol. 16, no. 2, pp. 121–135, 2000.

[21] D. P. Jones, “Redox potential of GSH/GSSG couple: assay and biological significance,” Methods in Enzymology, vol. 348, pp. 93–112, 2002.

[22] G. M. Enns, T. Moore, A. le et al., “Degree of glutathione deficiency and redox imbalance depend on subtype of mitochondrial disease and clinical status,” PLoS One, vol. 9, no. 6, article e100001, 2014.

[23] A. Pompella, A. Visvikis, A. Paolicchi, V. D. Tata, and A. F. Y. Chen, H. Dong, D. C. Thompson, H. G. Shertzer, D. W. D. P. Jones, “The changing faces of glutathione, a cellular protagonist,” Biochemical Pharmacology, vol. 66, no. 8, pp. 1499–1503, 2003.

[24] Y. Chen, H. Dong, D. C. Thompson, H. G. Shertzer, D. W. Nebert, and V. Vasiliiou, “Glutathione defense mechanism in liver injury: insights from animal models,” Food and Chemical Toxicology, vol. 60, pp. 38–44, 2013.

[25] R. Xiao, J. Lundström-Ljung, A. Holmgren, and H. F. Gilbert, “Catalysis of thiol/disulfide exchange. Glutaredoxin 1 and protein-disulfide isomerase use different mechanisms to enhance oxidase and reductase activities,” The Journal of Biological Chemistry, vol. 280, no. 22, pp. 21099–21106, 2005.

[26] R. Benfeitas, M. Uhlen, J. Nielsen, and A. Mardinoglu, “New challenges to study heterogeneity in cancer redox metabolism,” Frontiers in Cell and Development Biology, vol. 5, p. 65, 2017.

[27] K. Aoyama and T. Nakaki, “Glutathione in cellular redox homeostasis: association with the excitatory amino acid carrier (EAAC1),” Molecules, vol. 20, no. 5, pp. 8742–8758, 2015.

[28] H. Miki and Y. Funato, “Regulation of intracellular signalling through cysteineoxidation by reactive oxygen species,” Journal of Biochemistry, vol. 151, no. 3, pp. 255–261, 2012.

[29] J. V. Bannister, W. H. Bannister, and G. Rotilio, “Aspects of the structure, function, and applications of superoxide Dismutase,” Critical Reviews in Biochemistry, vol. 22, no. 2, pp. 111–180, 1987.

[30] I. N. Zelko, T. J. Mariani, and R. J. Fola, “Superoxide dismutase multigene family: a comparison of the Cu/Zn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) genes structures, evolution, and expression,” Free Radical Biology & Medicine, vol. 33, no. 3, pp. 337–349, 2002.

[31] P. Cheilikani, I. Fita, and P. C. Loewen, “Diversity of structures and properties among catalases,” Cellular and Molecular Life Sciences, vol. 61, no. 2, pp. 192–208, 2004.

[32] C. Nathan and A. Cunningham-Bussel, “Beyond oxidative stress: an immunologist’s guide to reactive oxygen species,” Nature Reviews Immunology, vol. 13, no. 5, pp. 349–361, 2013.

[33] X. Zhao and K. Drlica, “Reactive oxygen species and the bacterial response to lethal stress,” Current Opinion in Microbiology, vol. 21, pp. 1–6, 2014.

[34] H. Sies, “Oxidative stress: oxidants and antioxidants,” Experimental Physiology, vol. 82, no. 2, pp. 291–295, 1997.

[35] Y. Abiko, T. Miura, B. H. Phuc, Y. Shinkai, and Y. Kumagai, “Participation of covalent modification of Keap1 in the actvation of Nrf2 by tert-butylbenzoquinone, an electrophilic metabolite of butylated hydroxyanisole,” Toxicology and Applied Pharmacology, vol. 255, no. 1, pp. 32–39, 2011.

[36] A. Giudice and M. Montella, “Activation of the Nrf2-ARE signaling pathway: a promising strategy in cancer prevention,” BioEssays, vol. 28, no. 2, pp. 169–181, 2006.

[37] M. Kieliszek and S. Blazejak, “Current knowledge on the importance of selenium in food for living organisms: a review,” Molecules, vol. 21, no. 5, p. 609, 2016.

[38] H. W. Tan, H.-Y. Mo, A. Lau, and Y.-M. Xu, “Selenium species: current status and potentials in cancer prevention and therapy,” International Journal of Molecular Sciences, vol. 20, no. 1, p. 75, 2018.

[39] R. Gopalakrishna, Z. H. Chen, and U. Gundimeda, “Seleno-compounds induce a redox modulation of protein kinase C in the cell, compartmentally independent from cytosolic glutathione: its role in inhibition of tumor promotion,” Archives of Biochemistry and Biophysics, vol. 348, no. 1, pp. 37–48, 1997.

[40] R. Gopalakrishna and S. Jaken, “Protein kinase C signaling and oxidative stress,” Free Radical Biology & Medicine, vol. 28, no. 9, pp. 1349–1361, 2000.

[41] X. Bai, Y. Chen, X. Hou, M. Huang, and J. Jin, “Emerging role of Nrf2 in chemoresistance by regulating drug-metabolizing enzymes and efflux transporters,” Drug Metabolism Reviews, vol. 48, no. 4, pp. 541–567, 2016.

[42] J. H. No, Y. B. Kim, and Y. S. Song, “Targeting nrf2 signaling to combat chemoresistance,” Journal of Cancer Prevention, vol. 19, no. 2, pp. 111–117, 2014.

[43] P. Telkoparan-Akililar, S. Suzen, and L. Saso, “Pharmacological applications of Nrf2 inhibitors as potential antineoplastic drugs,” International Journal of Molecular Sciences, vol. 20, no. 8, p. 2025, 2019.

[44] X. J. Wang, Z. Sun, N. F. Villeneuve et al., “Nrf2 enhances resistance of cancer cells to chemotherapeutic drugs, the dark side of Nrf2,” Carcinogenesis, vol. 29, no. 6, pp. 1235–1243, 2008.

[45] N. Gharavi, S. Haggarty, and A. O. El-Kadi, “Chemoprotective and carcinogenic effects of tert-butylhydroquinone and its metabolites,” Current Drug Metabolism, vol. 8, no. 1, pp. 1–7, 2007.

[46] Y. Li, A. Seacat, P. Kuppusamy, J. L. Zweier, J. D. Yager, and M. A. Trush, “Copper redox-dependent activation of 2-tert-butyl(1,4)hydroquinone: formation of reactive oxygen species and induction of oxidative DNA damage in isolated DNA and cultured rat hepatocytes,” Mutation Research, vol. 518, no. 2, pp. 123–133, 2002.

[47] J. Brozmanova, D. Mániková, V. Vlčková, and M. Chovanec, “Selenium: a double-edged sword for defense and offence in cancer,” Archives of Toxicology, vol. 84, no. 12, pp. 919–938, 2010.

[48] R. Muecke, L. Schomburg, J. Buentzel, K. Kisters, O. Micke, and German Working Group Trace Elements and Electro-lytes in Oncology, “Selenium or no selenium—that is the question in tumor patients: a new controversy,” Integrative Cancer Therapies, vol. 9, no. 2, pp. 136–141, 2010.

[49] K. R. Martin and J. C. Barrett, “Reactive oxygen species as double-edged swords in cellular processes: low-dose cell signaling versus high-dose toxicity,” Human & Experimental Toxicology, vol. 21, no. 2, pp. 71–75, 2002.

[50] M. Schieber and N. S. Chandel, “ROS function in redox sig-naling and oxidative stress,” Current Biology, vol. 24, no. 10, pp. R453–R462, 2014.
[51] E. L. Yarosz and C. H. Chang, "The role of reactive oxygen species in regulating T cell-mediated immunity and disease," Immune Network, vol. 18, no. 1, article e14, 2018.

[52] N. Aykin-Burns, I. M. Ahmad, Y. Zhu, L. W. Oberley, and D. R. Spitz, "Increased levels of superoxide and H2O2 mediate the differential susceptibility of cancer cells versus normal cells to glucose deprivation," The Biochemical Journal, vol. 418, no. 1, pp. 29–37, 2009.

[53] A. Singh, V. Misra, R. K. Thimmulappa et al., "Dysfunctional KEAP1-NRF2 interaction in non-small-cell lung cancer," PLoS Medicine, vol. 3, no. 10, article e420, 2006.

[54] J. Wang, B. Luo, X. Li et al., "Inhibition of cancer growth in vitro and in vivo by a novel ROS-modulating agent with ability to eliminate stem-like cancer cells," Cell Death & Disease, vol. 8, no. 6, article e2887, 2017.

[55] C. A. Neumann and Q. Fang, "Are peroxiredoxins tumor suppressors?," Current Opinion in Pharmacology, vol. 7, no. 4, pp. 375–380, 2007.

[56] P. Birner, M. Schindl, A. Obermair, C. Plank, G. Breitenecker, and G. Oberhuber, "Overexpression of hypoxia-inducible factor 1alpha is a marker for an unfavorable prognosis in early-stage invasive cervical cancer," Cancer Research, vol. 60, no. 17, pp. 4693–4696, 2000.

[57] J. C. Soria, S. J. Jang, F. R. Khuri et al., "Overexpression of cyclin B1 in early-stage non-small cell lung cancer and its clinical implication," Cancer Research, vol. 60, no. 15, pp. 4000–4004, 2000.

[58] K. J. Davies, "The broad spectrum of responses to oxidants in proliferating cells: a new paradigm for oxidative stress," IUBMB Life, vol. 48, no. 1, pp. 41–47, 1999.

[59] L. R. Prosnitz, I. S. Goldenberg, R. A. Packard et al., "Radiation therapy as initial treatment for early stage cancer of the breast without mastectomy," Cancer, vol. 39, 2 Suppl, pp. 917–923, 1977.

[60] R. Timmerman, R. Paulus, J. Galvin et al., "Stereotactic body radiation therapy for inoperable early stage lung cancer," JAMA, vol. 303, no. 11, pp. 1070–1076, 2010.

[61] M. Peiris-Pages, U. E. Martinez-Outschoorn, F. Sotgia, and M. P. Lisanti, "Metastasis and oxidative stress: are antioxidants a metabolic driver of progression?," Cell Metabolism, vol. 22, no. 6, pp. 956–958, 2015.

[62] E. Piskounova, M. Agathocleous, M. M. Murphy et al., "Oxidative stress inhibits distant metastasis by human melanoma cells," Nature, vol. 527, no. 7577, pp. 186–191, 2015.

[63] K. Le Gal, M. X. Ibrahim, C. Wiel et al., "Antioxidants can increase melanoma metastasis in mice," Science Translational Medicine, vol. 7, no. 308, article 308re8, 2015.

[64] R. Benfeitas, G. Bidkhori, B. Mukhopadhyay et al., "Characterization of heterogeneous redox responses in hepatocellular carcinoma patients using network analysis," EbioMedicine, vol. 40, pp. 471–487, 2019.

[65] Q. Kong and K. O. Lillehei, "Antioxidant inhibitors for cancer therapy," Medical Hypotheses, vol. 51, no. 5, pp. 405–409, 1998.

[66] T. Ozben, "Oxidative stress and apoptosis: impact on cancer therapy," Journal of Pharmaceutical Sciences, vol. 96, no. 9, pp. 2181–2196, 2007.

[67] C. Gorrini, I. S. Harris, and T. W. Mak, "Modulation of oxidative stress as an anticancer strategy," Nature Reviews Drug Discovery, vol. 12, no. 12, pp. 931–947, 2013.

[68] Z. Zou, H. Chang, H. Li, and S. Wang, "Induction of reactive oxygen species: an emerging approach for cancer therapy," Apoptosis, vol. 22, no. 11, pp. 1321–1335, 2017.

[69] D. Magda and R. A. Miller, "Motexafin gadolinium: a novel redox active drug for cancer therapy," Seminars in Cancer Biology, vol. 16, no. 6, pp. 466–476, 2006.

[70] O. Tacar, P. Siamonsn, and C. R. Dass, "Doxorubicin: an update on anticancer molecular action, toxicity and novel drug delivery systems," The Journal of Pharmacy and Pharmacology, vol. 65, no. 2, pp. 157–170, 2013.

[71] G. T. Wondrak, "NQO1-activated phenothiazinium redox cyclers for the targeted bioreductive induction of cancer cell apoptosis," Free Radical Biology & Medicine, vol. 43, no. 2, pp. 178–190, 2007.

[72] D. Khuntia and M. Mehta, "Motexafin gadolinium: a clinical review of a novel radioenhancer for brain tumors," Expert Review of Anticancer Therapy, vol. 4, no. 6, pp. 981–989, 2004.

[73] M. P. Mehta, P. Rodrigus, C. H. Terhaard et al., "Survival and neurologic outcomes in a randomized trial of motexafin gadolinium and whole-brain radiation therapy in brain metastases," Journal of Clinical Oncology, vol. 21, no. 13, pp. 2529–2536, 2003.

[74] S. Kottaram, C. R. Chitambar, S. V. Kalivendi, J. Joseph, and B. Kalyanaraman, "Transferrin receptor-dependent iron uptake is responsible for doxorubicin-mediated apoptosis in endothelial cells: role of oxidant-induced iron signaling in apoptosis," The Journal of Biological Chemistry, vol. 277, no. 19, pp. 17179–17187, 2002.

[75] Y. J. Kang, Y. Chen, and P. N. Epstein, "Suppression of doxorubicin cardiotoxicity by overexpression of catalase in the heart of transgenic mice," The Journal of Biological Chemistry, vol. 271, no. 21, pp. 12610–12616, 1996.

[76] R. Marullo, E. Werner, N. Degtyareva et al., "Cisplatin induces a mitochondrial-ROS response that contributes to cytotoxicity depending on mitochondrial redox status and bioenergetic functions," PLoS One, vol. 8, no. 11, article e81162, 2013.

[77] G. A. Omura and G. Gynecologic Oncology, "Progress in gynecologic cancer research: the Gynecologic Oncology Group experience," Seminars in Oncology, vol. 35, no. 5, pp. 507–521, 2008.

[78] S. Dasari and P. B. Tchounwou, "Cisplatin in cancer therapy: molecular mechanisms of action," European Journal of Pharmacology, vol. 740, pp. 364–378, 2014.

[79] C. W. Helm and J. C. States, "Enhancing the efficacy of cisplatin in ovarian cancer treatment - could arsenic have a role," Journal of ovarian research, vol. 2, no. 1, p. 2, 2009.

[80] T. Hagen, G. D’Amico, M. Quintero et al., "Inhibition of mitochondrial respiration by the anticancer agent 2-methoxyestradiol," Biochemical and Biophysical Research Communications, vol. 322, no. 3, pp. 923–929, 2004.

[81] W. A. Moolberry, "Mechanism of action of 2-methoxyestradiol: new developments," Drug Resistance Updates, vol. 6, no. 6, pp. 355–361, 2003.

[82] M. Djavaheri-Mergny, J. Wietzerbin, and F. Besancon, "2-Methoxyestradiol induces apoptosis in Ewing sarcoma cells through mitochondrial hydrogen peroxide production," Oncogene, vol. 22, no. 17, pp. 2558–2567, 2003.

[83] R. Kachadourian, S. I. Liochev, D. E. Cabelli, M. N. Patel, I. Fridovich, and B. J. Day, "2-Methoxyestradiol does not inhibit superoxide dismutase," Archives of Biochemistry and Biophysics, vol. 392, no. 2, pp. 349–353, 2001.
A. O. Mueck and H. Seeger, "D. Francis, G. M. Richards, A. Forouzannia, M. P. Mehta, and M. Kataoka, G. Schumacher, R. J. Cristiano, E. N. Atkinson, N. J. Lakhani, M. A. Sarkar, J Venitz, and W. D. Figg, E. J. Bowles, R. Wellman, H. S. Feigelson et al., P. Huang, L. Feng, E. A. Oldham, M. J. Keating, and I. Ben Mosbah, Y. Mouchel, J. Pajaud et al., S. Waissbluth and S. J. Daniel, D. Khuntia, for brain tumors, in vivo, transgene delivery inhibits growth of metastatic lung cancer tumor suppressor transgene product coupled with systemic killing of cancer cells, pp. 2041–2049, 1998.

M. Cushman, H. M. He, J. A. Katzenellenbogen, C. M. Lin, and E. Hamel, "Synthesis, antitubulin and antimitotic activity, and cytoxicity of analogs of 2-methoxyestradiol, an endogenous mammalian metabolite of estradiol that inhibits tubulin polymerization by binding to the colchicine binding site," Journal of Medicinal Chemistry, vol. 38, no. 12, pp. 2041–2049, 1995.

M. Kataoka, G. Schumacher, R. J. Cristiano, E. N. Atkinson, J. A. Roth, and T. Mukhopadhyay, "An agent that increases tumor suppressor transgene product coupled with systemic transgene delivery inhibits growth of metastatic lung cancer in vivo," Cancer Research, vol. 58, no. 21, pp. 4761–4765, 1998.

T. H. Lippert, H. Adlercreutz, M. R. Berger, H. Seeger, W. Elger, and A. O. Mueck, "Effect of 2-methoxyestradiol on the growth of methyl-nitroso-urea (MNU)-induced rat mammary carcinoma," The Journal of Steroid Biochemistry and Molecular Biology, vol. 84, no. 1, pp. 51–56, 2003.

A. O. Mueck and H. Seeger, "2-Methoxyestradiol—biology and mechanism of action," Steroids, vol. 75, no. 10, pp. 625–631, 2010.

D. Francis, G. M. Richards, A. Forouzannia, M. P. Mehta, and D. Khuntia, "Motexafin gadolinium: a novel radiosensitizer for brain tumors," Expert Opinion on Pharmacotherapy, vol. 10, no. 13, pp. 2171–2180, 2009.

J. G. Blanco, C. L. Sun, W. Landier et al., "Anthracycline-related cardiomyopathy after childhood cancer: role of polymorphisms in carbonyl reductase genes—a report from the Children’s Oncology Group," Journal of Clinical Oncology, vol. 30, no. 13, pp. 1415–1421, 2012.

E. J. Bowles, R. Wellman, H. S. Feigelson et al., "Risk of heart failure in breast cancer patients after anthracycline and trastuzumab treatment: a retrospective cohort study," Journal of the National Cancer Institute, vol. 104, no. 17, pp. 1293–1305, 2012.

D. Cardinale, A. Colombo, G. Bacchiani et al., "Early detection of anthracycline cardiotoxicity and improvement with heart failure therapy," Circulation, vol. 131, no. 22, pp. 1981–1988, 2015.

S. Waissbluth and S. J. Daniel, "Cisplatin-induced ototoxicity: transporters playing a role in cisplatin toxicity," Hearing Research, vol. 299, pp. 37–45, 2013.

I. Ben Mosbah, Y. Mouchel, J. Pajaud et al., "Pretreatment with mangafodipir improves liver graft tolerance to ischemia/reperfusion injury in rat," PLoS One, vol. 7, no. 11, article e50235, 2012.

S. Kono, G. R. Merriam, D. D. Brandon, D. L. Loriaux, and M. B. Lipsett, "Radioimmunoassay and metabolism of the catechol estrogen 2-hydroxyestradiol," The Journal of Clinical Endocrinology and Metabolism, vol. 54, no. 1, pp. 150–154, 1982.

S. Kumari, A. K. Badana, M. M. G, S. G, and R. R. Malla, "Reactive oxygen species: a key constituent in cancer survival," Biomarker Insights, vol. 13, article 1177271918755391, 2018.

Y. Chen, Y. Yang, M. L. Miller et al., "Hepatocyte-specific Gclc deletion leads to rapid onset of steatosis with mitochondrial injury and liver failure," Hepatology, vol. 45, no. 5, pp. 1118–1128, 2007.

C. C. White, H. Viernes, C. M. Krejsa, D. Botta, and T. J. Kavanagh, "Fluorescence-based microtiter plate assay for glutamate-cysteine ligase activity," Analytical Biochemistry, vol. 318, no. 2, pp. 175–180, 2003.

O. W. Griffith and A. Meister, "Potent and specific inhibition of glutathione synthesis by buthionine sulfoximine (S-n-butyl homocysteine sulfoximine)," The Journal of Biological Chemistry, vol. 254, no. 16, pp. 7558–7560, 1979.

S. Moulder, N. Dhillion, C. Ng et al., "A phase I trial of imexon, a pro-oxidant, in combination with docetaxel for the treatment of patients with advanced breast, non-small cell lung and prostate cancer," Investigational New Drugs, vol. 28, no. 5, pp. 634–640, 2010.

E. V. Sheveleva, T. H. Landowski, B. K. Samulitis, G. Bartholomeusz, G. Powis, and R. T. Dorr, "Imexon induces an oxidative endoplasmic reticulum stress response in pancreatic cancer cells," Molecular Cancer Research, vol. 10, no. 3, pp. 392–400, 2012.

G. Abdelhamid and A. O. El-Kadi, "Buthionine sulfoximine, an inhibitor of glutathione biosynthesis, induces expression of soluble epoxide hydrolase and markers of cellular hyper trophy in a rat cardiomyoblast cell line: roles of the NF-kB and MAPK signaling pathways," Free Radical Biology & Medicine, vol. 82, pp. 1–12, 2015.

K. Dvorakova, C. M. Payne, T. H. Landowski, M. E. Tome, D. S. Halperin, and R. T. Dorr, "Imexon activates an intrinsic apoptosis pathway in RPMI8226 myeloma cells," Anti-Cancer Drugs, vol. 13, no. 10, pp. 1031–1042, 2002.

K. Dvorakova, C. M. Payne, M. E. Tome, M. M. Biehl, T. McClure, and R. T. Dorr, "Induction of oxidative stress and apoptosis in myeloma cells by the aziridine- containing agent imexon," Biochemical Pharmacology, vol. 60, no. 6, pp. 749–758, 2000.

K. Dvorakova, C. N. Waltmire, C. M. Payne, M. E. Tome, M. M. Biehl, and R. T. Dorr, "Induction of mitochondrial changes in myeloma cells by imexon," Blood, vol. 97, no. 11, pp. 3544–3551, 2001.

U. Kilic, E. Kilic, Z. Tuzcu et al., "Melatonin suppresses cisplatin-induced nephrotoxicity via activation of Nrf-2/HO-1 pathway," Anticancer Research, vol. 33, no. 5, pp. 2429–2435, 2013.

J. Fiedor and K. Burda, "Potential role of carotenoids as antioxidants in human health and disease," Nutrients, vol. 6, no. 2, pp. 466–488, 2014.

K. Sak, "Cytotoxicity of dietary flavonoids on different human cancer types," Pharmacognosy Reviews, vol. 8, no. 16, pp. 122–146, 2014.

Z. Tavan and H. A. Kayali, "Flavonoids showed anticancer effects on the ovarian cancer cells: involvement of reactive oxygen species, apoptosis, cell cycle and invasion," Biomedicine & Pharmacotherapy, vol. 116, article 109004, 2019.
and augments paclitaxel efficacy in head and neck squamous cell carcinoma,” *Clinical Cancer Research*, vol. 10, no. 24, pp. 8665–8673, 2004.

[113] T. L. Yue, X. Wang, C. S. Louden et al., “2-Methoxyestradiol, an endogenous estrogen metabolite, induces apoptosis in endothelial cells and inhibits angiogenesis: possible role for stress-activated protein kinase signaling pathway and Fas expression,” *Molecular Pharmacology*, vol. 51, no. 6, pp. 951–962, 1997.

[114] Y. J. Li, Y. H. Lei, N. Yao et al., “Autophagy and multidrug resistance in cancer,” *Chinese Journal of Cancer*, vol. 36, no. 1, p. 52, 2017.

[115] J. J. Haddad, “L-Buthionine-(S,R)-sulfoximine, an irreversible inhibitor of gamma-glutamylcysteine synthetase, augments LPS-mediated pro-inflammatory cytokine biosynthesis: evidence for the implication of an IkappaB-alpha/NF-kappaB insensitive pathway,” *European Cytokine Network*, vol. 12, no. 4, pp. 614–624, 2001.

[116] J. F. Ewing and M. D. Maines, “Glutathione depletion induces heme oxygenase-1 (HSP32) mRNA and protein in rat brain,” *Journal of Neurochemistry*, vol. 60, no. 4, pp. 1512–1519, 1993.

[117] M. Khorsand, M. Akmali, S. Sharzad, and M. Beheshtitabar, “Melatonin reduces cataract formation and aldose reductase activity in lenses of streptozotocin-induced diabetic rat,” *Iranian Journal of Medical Sciences*, vol. 41, no. 4, pp. 305–313, 2016.

[118] Y. Liu, Q. Li, L. Zhou et al., “Cancer drug resistance: redox resetting renders a way,” *Oncotarget*, vol. 7, no. 27, pp. 42740–42761, 2016.

[119] C. Holohan, S. van Schaeybroeck, D. B. Longley, and P. G. Johnston, “Cancer drug resistance: an evolving paradigm,” *Nature Reviews Cancer*, vol. 13, no. 10, pp. 714–726, 2013.

[120] A. Ayer, C. W. Gourlay, and I. W. Dawes, “Cellular redox homeostasis, reactive oxygen species and replicative ageing in Saccharomyces cerevisiae,” *FEBS Yeast Research*, vol. 14, no. 1, pp. 60–72, 2014.