Yiqi-Huoxue Granule (YQHX) Downregulates Prothrombotic Factors by Modulating KLF2 and NF-κB in HUVECs following LPS Stimulation

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The Yiqi-Huoxue granule (YQHX) is a traditional Chinese medication widely used in the therapy of the traditional Chinese medicine diagnosis “Qi deficiency” or “blood stasis” in China. Both these symptoms are related to inflammation, but the mechanisms of YQHX against inflammation are largely unknown. Thus, our present study investigated the effects of YQHX on regulating inflammatory responses induced by lipopolysaccharides (LPS) in HUVECs. Our data found that YQHX remarkably inhibits the production of prothrombotic factors, plasminogen activator inhibitor-1 (PAI-1) and tissue factor (TF), while it upregulated the protein expression of Kruppel-like factor 2 (KLF2). The increase in PAI-1 and TF was significantly attenuated through a transgenic knockdown in KLF2 with a Lenti-shKLF2 vector. YQHX also decreases the phosphorylation of nuclear factor-κB (NF-κB) p65 and IκB following LPS stimulation, and it effectively suppresses PAI-1 and TF via a NF-κB-dependent mechanism. Taken together, our results suggest that YQHX provides a notable antithrombotic activity via regulating the KLF2 expression and NF-κB signaling pathway in HUVECs. The KLF2 and NF-κB may be potential therapeutic targets for interventions of inflammation associated with atherosclerosis.

1. Introduction

Acute ischemic heart disease is a leading cause of death and disability worldwide [1]. It is usually associated with luminal thrombosis resulting from vulnerable atherosclerotic erosion or plaque rupture initiated by endothelial dysfunction [2]. Chronic inflammation is involved in the development of luminal thrombosis [3], due to its role in exacerbating endothelial injury and provoking atherosclerotic plaque rupture [2, 4]. However, current antiatherosclerotic agents and antiplatelet therapy only prevent the physical formation of thrombosis [5, 6], but do not alleviate inflammation or endothelial dysfunction. Thus, developing new strategies aimed at protecting against inflammation and endothelial dysfunction may have important clinical implications in antithrombotic therapies.

The Yiqi-Huoxue granule (YQHX) is a traditional Chinese medication widely used in the therapy of the traditional Chinese medicine diagnosis “Qi deficiency” or “blood stasis” in China. It is composed of Ginseng Radix et Rhizoma (Panax ginseng C. A. Mey), Astragali Radix (Astragalus membranaceus (Fisch.) Bge.), Paeoniae Rubra Radix (Paeonia veitchii Lynch), and Carthami Flos (Carthamus tinctorius L.). The Paeoniae Rubra Radix, Carthami Flos, and Ginseng Radix et Rhizoma extracts have been recognized to produce antithrombotic effects [7–9]. The components of Ginseng
Radix et Rhizoma and Astragali Radix in YQHX are effective to inhibit inflammatory responses [10, 11]. Our previous studies have demonstrated that YQHX could inhibit the expression of prothrombotic factors, plasminogen activator inhibitor-1 (PAI-1) and tissue factor (TF), induced by thrombin in human umbilical vein endothelial cells (HUVECs) [12]. It also reduces platelet aggregation associated with myocardial infarction in rats [13]. However, so far, it is largely unknown if the antithrombotic effect of YQHX is associated with its anti-inflammatory activity.

Kruppel-like factor 2 (KLF2) is a transcriptional regulator highly expressed in endothelial cells. Overexpression of KLF2 prolongs thrombotic time and mediates in vivo rapamycin-induced arterial thrombosis in mice [14, 15]. Conversely, KLF2 deficiency inhibits antithrombotic genes [16]. KLF2 also mediates acute and chronic inflammations [17, 18]. The anti-inflammatory effects of KLF2 mechanistically are linked to the suppression of nuclear factor-kappa B (NF-κB) signaling [17, 19] that regulates a variety of genes related to inflammatory responses [20, 21]. Thus, KLF2 may be a key thrombotic regulator due to its effects on regulating both endothelial function and inflammation.

Lipopolysaccharide (LPS) is a component of Gram-negative bacteria. It alters the fibrinolytic system leading to a procoagulant state or thrombosis [22, 23]. LPS stimulates both proinflammatory mediators [24, 25] and prothrombotic factors [26, 27]. In the present study, we used a LPS-incubated endothelial cell model to mimic inflammatory conditions in human atherosclerotic lesions. We investigated the effects of YQHX on regulating the NF-κB signaling pathway and KLF2 expression in response to inflammatory stimulation. We also examined if the production of prothrombotic factors PAI-1 and TF could function as a downstream readout to test the potential therapeutic effects of YQHX against cardiovascular diseases.

2. Materials and Methods

2.1. Reagents. Endothelial cell culture media (ECM), endothelial cell growth supplies (ECGS), and fetal bovine serum (FBS) were purchased from ScienCell Research Laboratories (ScienCell, CA, USA) or Gibco (Gibco, CA, USA). All of the chemicals, including LPS, simvastatin (ST), pyrrolidine dithiocarbamate (PDTC), and 5-diphenyl-2-H-tetrazolium bromide (MTT) dye, were purchased from Sigma-Aldrich (St. Louis, MO, USA). YQHX, composed of Ginseng Radix et Rhizoma, Astragali Radix, Paeoniae Rubra Radix, and Carthami Flos, was produced by Sichuan Neo-Green Pharmaceutical Technology Development Co., Ltd. (Sichuan, China). It was freshly prepared in a phosphate-buffered solution before use.

2.2. HUVECs. The HUVECs were purchased from ScienCell Research Laboratories (ScienCell, CA, USA), and they were cultured in an incubator at 37°C with 5% CO₂ in ECM medium supplemented with 5% FBS and 0.03 mg/ml ECGS. The HUVECs (around 70-80% confluence) were incubated with YQHX (from 0.25 to 1.25 mg/ml) and ST (3 μM) for 3 hours. This was followed by LPS stimulation (25 μg/ml) for 12 hours.

2.3. Cell Viability. The HUVECs were seeded in 96-well plates with a density of 0.7 × 10⁴ cells/well and cultured overnight. The cells were then incubated with YQHX at a concentration of 0-10.0 mg/ml prior to treatment with 10 μl of MTT. The absorbance at 570 nm was detected using a microplate reader (Thermo Fisher Scientific, Waltham, USA).

2.4. Transfection of HUVECs. 293T cells in the logarithmic growth phase were cotransfected with recombinant pBOB plasmid, PAX-2, and VSV-G for 48 h. The supernatant was centrifuged at 4000g for 10 min at 4°C to remove cell debris and further concentrated to obtain lentivirus. To overexpress KLF2, the HUVECs were infected with lentivirus containing a KLF2-overexpressing sequence (Lenti-KLF2). As shown in Table 1, the KLF2 short hairpin RNA (Lenti-shKLF2, Cyagen Biosciences Inc., Guangzhou, China) was used to knockdown KLF2 as described previously. The cells were also transfected with a scrambled Lenti-GFP as the negative control. The efficiency of transfection was detected with fluorescence microscopy.

2.5. Western Blot Analysis. The cellular lysates extracted from HUVECs were used to quantify protein expression by Western blot. Briefly, the protein concentration was determined by a BCA protein assay kit, and equal amounts of proteins were loaded into 10% SDS-PAGE gels. Following transfer and blocking with 5% skim milk for 1 h at room temperature, the membranes were then incubated with primary antibodies against KLF2 (1 : 2000 dilution), PAI-1 (1 : 500 dilution), TF (1 : 500 dilution), p65 (1 : 500 dilution), p-p65 (1 : 500 dilution), GAPDH (1 : 2000 dilution), or β-actin (1 : 2000 dilution) overnight at 4°C. Following incubation with a secondary antibody and ECL, the proteins were visualized using a Bio-Rad Gel Doc XR ++ Imaging System (Bio-Rad, CA, USA). The intensity of the bands was assessed using ImageJ software (available at http://rsbweb.nih.gov/ij/).

2.6. Statistical Analysis. The data were analyzed using either parametric test or nonparametric Mann-Whitney U test depending on the pattern of data distribution, and the results are presented as the mean ± SD. A p value less than 0.05 was considered statistically significant.

3. Results

3.1. LPS Upregulates PAI-1 and TF Expression in a Time-Dependent Manner. LPS regulates PAI-1 and TF in both cell and animal models [26, 27]. The present study investigated
the effects of LPS on regulating the expressions of prothrombotic factors, PAI-1 and TF, in HUVECs. We observed that following LPS stimulation (25 μg/ml), the protein levels of PAI-1 and TF significantly increased in a time-dependent manner in HUVECs (Figure 1). The parallel enhancement of PAI and TF suggests that LPS treatment significantly induces prothrombotic reactions in HUVECs by upregulating both PAI-1 and TF.

3.2. YQHX Inhibits LPS-Induced Expressions of PAI-1 and TF in HUVECs. A low concentration of YQHX (no more than 1.25 mg/ml) incubated with HUVECs did not affect cell survival (Figure 2(a)). Thus, we cotreated the cells with YQHX and LPS in order to identify the protective effects of YQHX on regulating LPS-induced prothrombotic reactions. Our previous study has demonstrated that YQHX could inhibit thrombin-induced PAI-1 and TF expressions in HUVECs [12]. Our present findings further indicate that YQHX at a low concentration can inhibit LPS-induced PAI-1 and TF (Figure 2(b)). The inhibitory effect of YQHX on both prothrombotic factors PAI-1 and TF is similar to the effect of simvastatin (ST), which is an HMG CoA reductase inhibitor [28] (Figure 2(b)).

3.3. YQHX Inhibits LPS-Induced PAI-1 and TF Expression through KLF2. KLF2 regulates both acute and chronic inflammation and owns antithrombotic properties [17, 18]. LPS treatment significantly reduced the expression of KLF2 in HUVECs (Figure 3(a)). In the present study, we then overexpressed KLF2 in HUVECs to identify its effect on the regulation of the prothrombotic factors PAI-1 and TF. Indeed, Lenti-KLF2 transduction in HUVECs upregulates the KLF2 expression in the protein level (Figure 3(b)). It significantly suppresses the expression of PAI-1 and TF with or without LPS stimulation (Figure 3(c)). In contrast, shRNA targeting KLF2 downregulates the expression of KLF2 in HUVECs (Figure 4(a)), which results in increased PAI-1 and TF expression in the presence or absence of LPS (Figure 4(b)). At the concentration range of 0.25 mg/ml to 1.25 mg/ml, YQHX was able to overcome the attenuation of KLF2 expression caused by LPS (Figure 3(a)), which is associated with the attenuation of PAI-1 and TF expression. The knockdown of KLF2 with Lenti-shKLF2 failed to inhibit PAI-1 and TF, suggesting that YQHX might modulate the expression of prothrombotic factors through the upregulation of KLF2.

3.4. YQHX Inhibits the Phosphorylation of NF-κB p65 and IκB. The activation of the NF-κB signaling pathway is initiated by IκB phosphorylation and degradation. To explore if YQHX regulates the prothrombotic factors by the NF-κB signaling pathway, the phosphorylation levels of NF-κB p65 and IκB were quantified by Western blot. Our data suggest that LPS treatment for 3 hours in HUVECs increased the phosphorylation of NF-κB p65 and IκB, and this was

| Table 1: The sequence of specific KLF2 shRNAs used in the present study. All of the shRNAs correspond to Homo sapiens. |
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| Name | Sense (5’-3’) | Antisense (5’-3’) |
| KLF2 (shRNA1) | GCTGCACATGAAACGGCACAT | ATGTGCCGTTTCATGTGCAGC |
| KLF2 (shRNA2) | TTGTGATGCCTTGTGAGAAAT | ATTTCTCACAAGGCAATCAAA |
| KLF2 (shRNA3) | CCAACTGTAAGCTGATATTTA | TAAAATTACGACACATACAGTTGG |
| NC (scramble shRNA) | CCTAAGGTTAAGTCGCCCTCG | CGAGGGCGACTTAACCITAGG |

Figure 1: LPS upregulates the protein level of PAI-1 and TF in HUVECs. The cells were treated with 25 μg/ml LPS from 0 to 12 h. The levels of (a) PAI-1 and (b) TF were determined using Western blot. Data are expressed as means ± SD (n = 4). *p < 0.05 vs. 0 h.

Figure 2: YQHX inhibits LPS-induced expression of PAI-1 and TF in HUVECs. The cells were treated with 25 μg/ml LPS and/or 0.25 mg/ml YQHX for 0 to 12 h. The levels of (a) PAI-1 and (b) TF were determined using Western blot. Data are expressed as means ± SD (n = 4). *p < 0.05 vs. 0 h.
significantly inhibited by YQHX at a concentration range of 0.25 to 1.25 mg/ml (Figure 5). The inhibition of phosphorylation of NF-κB p65 and IκB was more robust following a higher concentration (1.25 mg/ml) of YQHX treatment compared to a lower concentration (0.25 mg/ml) (Figure 5). The high concentration of YQHX displayed the same inhibitory effect on the NF-κB signaling pathway as simvastatin, which is a well-known NF-κB-specific inhibitor. Together, our data suggest that YQHX inhibits LPS-induced inflammation by repressing the activation of the NF-κB signaling pathway in a dose-dependent manner.

3.5. YQHX Inhibits PAI-1 and TF through an NF-κB-Dependent Mechanism. NF-κB regulates KLF2 expression, and KLF2 inhibits PAI-1 and TF. Therefore, we investigated if YQHX inhibits the LPS-induced expression of PAI-1 and TF through the NF-κB signaling pathway. Inhibition of the NF-κB pathway by PDTC, which is a specific inhibitor of NF-κB, successfully reversed the reduction in KLF2 expression caused by LPS (Figure 6). This suggests that the NF-κB pathway is important in mediating KLF2 expression. Interestingly, the upregulated levels of KLF2 following treatments with PDTC and YQHX were not comparable. The effects of YQHX on KLF2 seem more robust (Figure 6). Furthermore, cotreatment with PDTC and YQHX impeded the ability of YQHX to increase KLF2 expression. This suggests that the upregulation of KLF2 by YQHX is partially mediated by the NF-κB pathway. We also observed that treatment with PDTC significantly reduced the expression of PAI-1 and TF, and PDTC partially attenuates the effects of YQHX on PAI-1 and TF (Figure 6). Together, our results indicate that YQHX may play a key role in modulating the expression of PAI-1 and TF through an NF-κB-dependent mechanism.

3.6. The Schematic Mechanism of YQHX’s Antithrombotic Effects following LPS Stimulation. LPS binds with the cell membrane receptor, TLR4, and activates the NF-κB pathway through a MyD88-dependent pathway. NF-κB then binds with p65 to form a complex, which is further translocated into the nucleus to inhibit KLF2 gene transcription. The KLF2 negatively regulates PAI-1 and TF genes. P65 indirectly promotes the expression of PAI-1 and TF. YQHX downregulates LPS-induced prothrombotic factors through an NF-κB/KLF2 pathway (Figure 7).
4. Discussion

Chinese herbs have been used to prevent cardiovascular diseases for thousands of years in China. YQHX is exclusively prescribed as a traditional Chinese medication for blood stasis of cardiovascular disease in the clinical practice in China. The present study is the first to demonstrate that YQHX attenuates the expression of prothrombotic factors, PAI-1 and TF, following LPS stimulation. YQHX mediates the NF-κB/KLF2 pathway leading to the reduction of the PAI-1 and TF expression in HUVECs. Our study may provide an important insight into the utilization of traditional Chinese medicine in the prevention of thrombosis formation driven by inflammation.

Increasing evidence suggests that inflammation plays a critical role in plaque stability, which eventually leads to rupture and thrombosis [29, 30]. Increased expression of TF and PAI-1 in the endothelium and circulating inflammatory cells might accelerate thrombosis enlargement in patients with acute coronary syndrome (ACS) [31]. Statins have been recognized to have anti-inflammatory effects independent of their hypolipidemic actions [32]. However, patients with ACS still remain vulnerable even following statin treatment. Ginsenoside Rb1, which is extracted from Ginseng Radix et Rhizoma, was found to potentially inhibit inflammatory responses by skewing macrophages toward the M2 phenotype [33]. The Astragali Radix polysaccharide significantly reduces LPS-induced gene expression of tumor necrosis factor alpha (TNF-α) and interleukin-8 [34]. The extracts of Carthami Flos also possess remarkable anti-inflammatory activity [35]. These findings demonstrate that active extracts from Chinese herbs may regulate inflammation. In fact, a traditional Chinese medicine, Xiangqi Tang (XQT), contains the above active components, and it has an anti-inflammatory function in LPS-treated rat cardiac microvascular endothelial cells. It inhibits the secretion of prothrombotic genes, such as TF and PAI-1, and inflammatory factors, such as TNF-α and intercellular cell adhesion molecule-1 (ICAM-1) [36]. The previous study also found that Tongqiaohuoxue decoction (THD), which includes

Figure 3: YQHX inhibits LPS-induced PAI and TF expression through KLF2. (a) HUVECs were incubated with YQHX (0.25 and 1.25 mg/ml) or ST (3 μM) for 3 h followed with LPS (25 μg/ml) stimulation for 12 h. (b) HUVECs were infected with Lenti-GFP or Lenti-KLF2 for 24 h. (c) HUVECs were preinfected with Lenti-GFP or Lenti-KLF2, followed by LPS (25 μg/ml) stimulation for 12 h. Cell lysates were then prepared, and Western blot analysis was performed to determine the expressions of KLF2, PAI-1, and TF. Data are expressed as means ± SD (n = 4). *p < 0.05 vs. control and *p < 0.05 vs. group with 25 μg/ml LPS.
Figure 4: YQHX inhibits the PAI-1 and TF expression in a KLF2-dependent manner. (a) HUVECs were infected with Lenti-GFP or Lenti-shKLF2 for 24 h. (b) HUVECs were infected with Lenti-GFP or Lenti-shKLF2. Then, they were incubated with YQHX (1.25 mg/ml) for 3 h prior to 25 μg/ml LPS stimulation for 12 h. Cell lysates were prepared and subjected to Western blot measurements to determine the expression of KLF2, PAI-1, and TF. Data are expressed as means ± SD (n = 4). # p < 0.05 vs. control, * p < 0.05 vs. group with 25 μg/ml LPS or LPS plus Lenti-GFP, and ▲ p < 0.05 vs. LPS plus YQHX or LPS plus YQHX and Lenti-GFP.

Figure 5: YQHX regulates the phosphorylation of NF-κB p65 and IκB. HUVECs were pretreated with YQHX (0.25 and 1.25 mg/ml) or ST (3 μM) for 3 h before being exposed to LPS (25 μg/ml) for 3 h. Cell lysates were prepared and subjected to Western blot analysis to determine the phosphorylation levels of (a) NF-κB p65 and (b) IκB proteins. Data are expressed as means ± SD (n = 4). # p < 0.05 vs. control and * p < 0.05 vs. group with 25 μg/ml LPS.
Ginseng Radix et Rhizoma, Astragali Radix, Paeoniae Rubra Radix, and Carthami Flos, also exerts anti-inflammatory and antithrombotic effects by regulating PAI-1 and fibrinolysis [37].

KLF2 functions as a “molecular switch” to regulate vascular homeostasis by maintaining the integrity of the endothelial barrier under physiological and pathological conditions [16]. KLF2 overexpression represses PAI-1 and TF gene expression with or without cytokine TNF-α stimulation [16]. Statins and shear stress also provide antithrombotic actions by stimulating the KLF2 expression in a dose-dependent manner [38–40]. Thus, the upregulation of KLF2 may be a potential therapeutic strategy to limit thrombosis. The present study is the first to demonstrate that YQHX has anti-inflammatory and antithrombotic effects through a KLF2-dependent mechanism. Upregulation of the KLF2 expression by YQHX markedly decreased the production of PAI-1 and TF induced by LPS stimulation, and transgenic knockdown of KLF2 reversed the antithrombotic effects of YQHX.

Typically, NF-κB heterodimers are sequestered with the repressive protein IκB and are inactive in the cytoplasm. Various inflammatory stimuli activate the NF-κB pathway by inducing the phosphorylation and degradation of IκB [41, 42]. The activation of NF-κB facilitates the expressions of proinflammatory genes during chronic inflammation and atherosclerotic development [43]. Thus, the suppression of NF-κB is an important strategy for overcoming inflammation [44–46]. Previous studies have indicated that the Chinese traditional medicine, XQT, and its active components promote anti-inflammatory effects by inhibiting mitogen-activated protein kinase (MAPK) and NF-κB signaling pathways [36]. Our results are the first to suggest that YQHX suppresses LPS-induced phosphorylation of both NF-κB p65 and IκB. More importantly, YQHX partially inhibits the production of PAI-1 and TF through an NF-κB-dependent mechanism.

KLF2 inhibits the NF-κB pathway by affecting the recruitment of p300/CBP [19]. In contrast, the NF-κB p65 protein incorporates with histone deacetylase 4 to suppress...
the levels of KLF2. It primarily acts to inhibit the binding of myocyte enhancer factor 2 (MEF2) with the KLF2 promoter [47]. The present study demonstrated that an inhibitor of NF-κB, PDTC, partially inhibits the upregulation of KLF2 induced by YQHX, suggesting that NF-κB is partially involved in YQHX-regulated KLF2 expression.

5. Conclusion

In summary, we demonstrated that YQHX provides anti-inflammatory and antithrombotic effects. It markedly decreases expressions of PAI-1 and TF following stimulation, which are associated with an upregulation of KLF2 in HUVECs. YQHX also inhibits the phosphorylation of NF-κB p65 and IκB proteins, which partially regulate the KLF2 expression. The present findings revealed a mechanism for the inhibitory effects of YQHX on thrombosis formation, suggesting that YQHX may be a promising strategy to prevent inflammation-related diseases.

5.1. Limitations of Our Present Study. Our present findings about YQHX are only based on in vitro cell culture models. Our future studies will focus on exploring the effects of YQHX against inflammation or endothelial dysfunction in in vivo animal models.

Data Availability

The datasets used or analyzed during the current study are available from the corresponding author on reasonable request.

Conflicts of Interest

No conflict of interest was declared.

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References

[1] E. J. Benjamin, M. J. Blaha, S. E. Chiuve et al., "Heart disease and stroke statistics-2017 update: a report from the American Heart Association," Circulation, vol. 135, no. 10, pp. e146–e603, 2017.
[2] L. Badimon and G. Vilahur, "Thrombosis formation on atherosclerotic lesions and plaque rupture," Journal of Internal Medicine, vol. 276, no. 6, pp. 618–632, 2014.
[3] C. Cochain and A. Zernecke, "Macrophages in vascular inflammation and atherosclerosis," Pflügers Archiv - European Journal of Physiology, vol. 469, no. 3-4, pp. 485–499, 2017.
[4] P. Shah, S. Bajaj, H. Virk, M. Bikkina, and F. Shamoon, "Rapid progression of coronary atherosclerosis: a review," Thrombosis, vol. 2015, no. 634983, 6 pages, 2015.
[5] M. Zhang, J. He, C. Jiang et al., "Plaque-hyaluronidase-responsive high-density-lipoprotein-mimetic nanoparticles for multistage intimal-macrophage-targeted drug delivery and
enhanced anti-atherosclerotic therapy,” International Journal of Nanomedicine, vol. 12, pp. 533–558, 2017.

[6] N. Papageorgiou, E. Zacharia, A. Ioannou et al., “Novel anti-platelets in stable coronary artery disease,” Current Pharmaceutical Design, vol. 22, no. 29, pp. 4537–4567, 2016.

[7] Y. H. Li and N. S. Wang, “Antithrombotic effects of Danggui, Honghua and potential drug interaction with clopidogrel,” Journal of Ethnopharmacology, vol. 128, no. 3, pp. 623–628, 2010.

[8] P. Xie, L. Cui, Y. Shan, and W. Y. Kang, “Antithrombotic effect and mechanism of Radix Paoniae Rubra,” BioMed Research International, vol. 2017, Article ID 9475074, 9 pages, 2017.

[9] M. Endale, W. M. Lee, S. M. Kamruzzaman et al., “Ginsenoside-Rp1 inhibits platelet activation and thrombus formation via impaired glycoprotein VI signalling pathway, tyrosine phosphorylation and MAPK activation,” British Journal of Pharmacology, vol. 167, no. 1, pp. 109–127, 2012.

[10] W. J. Zhang and B. Frei, “Astragaloside IV inhibits NF-kB activation and inflammatory gene expression in LPS-treated mice,” Mediators of Inflammation, vol. 2015, Article ID 274314, 11 pages, 2015.

[11] J. H. Kim, “Cardiovascular diseases and Panax ginseng: a review on molecular mechanisms and medical applications,” Journal of Ginseng Research, vol. 36, no. 1, pp. 16–26, 2012.

[12] H. Wu, X. Wang, S. Gao, Z. Lei, Z. Wang, and L. Han, “Effect of Yiqi Huoxue decoction on expression of pro-thrombotic factor PAI-1 and TF in endothelial cells induced by thrombin,” Acta Chinese Medicine, vol. 32, no. 9, pp. 1683–1685, 2017.

[13] L. Han, H. Wu, Z. Wang, and J. Li, “Effects of Yiqi Huoxue decoction on platelet aggregation and apptt/ptt/tt in rats after myocardial infarction with left ventricular remodeling,” Jiangsu Zhong Yi Yao, vol. 26, no. 1, pp. 55–56, 2005.

[14] X. M. Nie, L. X. Su, R. X. Xu, Y. L. Guo, Y. J. Zhou, and J. J. Li, “Kruppel-like factor 2 might mediate the rapamycin-induced arterial thrombosis in vivo: implications for stent thrombosis in patients,” Chinese Medical Journal, vol. 126, no. 14, pp. 2636–2640, 2013.

[15] L. Nayak, H. Shi, G. B. Atkins, Z. Lin, A. H. Schmaier, and M. K. Jain, “The thromboprotective effect of bortezomib is dependent on the transcription factor Kruppel-like factor 2 (KLF2),” Blood, vol. 123, no. 24, pp. 3828–3831, 2014.

[16] Z. Lin, A. Kumar, S. SenBanerjee et al., “Kruppel-like factor 2 (KLF2) regulates endothelial thrombotic function,” Circulation Research, vol. 96, no. 5, pp. e48–e57, 2005.

[17] L. Nayak, L. Goduni, Y. Takami et al., “Kruppel-like factor 2 is a transcriptional regulator of chronic and acute inflammation,” The American Journal of Pathology, vol. 182, no. 5, pp. 1696–1704, 2013.

[18] R. Pathak, L. Shao, S. M. Chafekar et al., “IKKβ regulates endothelial thrombomodulin in a KLF2-dependent manner,” Journal of thrombosis and haemostasis, vol. 12, no. 9, pp. 1533–1544, 2014.

[19] S. SenBanerjee, Z. Lin, G. B. Atkins et al., “KLF2 is a novel transcriptional regulator of endothelial proinflammatory activation,” The Journal of Experimental Medicine, vol. 199, no. 10, pp. 1305–1315, 2004.

[20] R. M. P. Gutierrez and C. Hoyo-Vadillo, “Anti-inflammatory potential of Petiveria alliacea on activated raw264.7 murine macrophages,” Pharmacognosy magazine, vol. 13, no. 50, pp. 174–178, 2017.

[21] J. Gdula-Argasinska, P. Pasko, K. Sulikowska-Ziaja, K. Kala, and B. Muszyńska, “Anti-inflammatory activities of garlic sprouts, a source of α-linolenic acid and 5-hydroxy-1-tryptophan, in raw 264.7 cells,” Acta Biochimica Polonica, vol. 64, no. 3, pp. 551–559, 2017.

[22] A. F. Suffredini, P. C. Harpel, and J. E. Parrillo, “Promotion and subsequent inhibition of plasminogen activation after administration of intravenous endotoxin to normal subjects,” The New England Journal of Medicine, vol. 320, no. 18, pp. 1165–1172, 1989.

[23] P. H. Quax, C. M. van den Hoogen, J. H. Verheijen et al., “Endotoxin induction of plasminogen activator and plasminogen activator inhibitor type 1 mRNA in rat tissues in vivo,” The Journal of Biological Chemistry, vol. 265, no. 26, pp. 15560–15563, 1990.

[24] Y. H. Choi and H. J. Kang, “Fructus sophorae attenuates secretion of proinflammatory mediators and cytokines through the modulation of NF-kB and MAPK signaling pathways in LPS-stimulated RAW 264.7 macrophages,” General Physiology and Biophysics, vol. 35, no. 3, pp. 323–331, 2016.

[25] L. A. Cox, L. T. van Eijk, B. P. C. Ramakers et al., “Inflammation-induced increases in plasma endocan levels are associated with endothelial dysfunction in humans in vivo,” Shock, vol. 43, no. 4, pp. 322–326, 2015.

[26] M. Y. Gao, L. Chen, L. Yang, X. Yu, J. P. Kou, and B. Y. Yu, “Berberine inhibits LPS-induced TF procoagulant activity and expression through NF-kB/p65, Akt and MAPK pathway in THP-1 cells,” Pharmacological Reports, vol. 66, no. 3, pp. 480–484, 2014.

[27] N. Ohkura, K. Oishi, F. Kihara-Negishi, G. Atsumi, and T. Tatefuji, “Effects of a diet containing Brazilian propolis on lipopolysaccharide-induced increases in plasma plasminogen activator inhibitor-1 levels in mice,” Journal of Intercultural Ethnopharmacology, vol. 5, no. 4, pp. 439–443, 2016.

[28] Y. Kunieda, K. Nakagawa, H. Nishimura et al., “HMG CoA reductase inhibitor suppresses the expression of tissue factor and plasminogen activator inhibitor-1 induced by angiotensin II in cultured rat aortic endothelial cells,” Thrombosis Research, vol. 110, no. 4, pp. 227–234, 2003.

[29] I. Tabas, “Macrophage death and defective inflammation resolution in atherosclerosis,” Nature Reviews Immunology, vol. 10, no. 1, pp. 36–46, 2010.

[30] K. Ley, Y. I. Miller, and C. C. Hedrick, “Monocyte and macrophage dynamics during atherogenesis,” Arteriosclerosis, Thrombosis, and Vascular Biology, vol. 31, no. 7, pp. 1506–1516, 2011.

[31] T. Sakai, S. Inoue, M. Takei et al., “Activated inflammatory cells participate in thrombus size through tissue factor and plasminogen activator inhibitor-1 in acute coronary syndrome: immunohistochemical analysis,” Thrombosis Research, vol. 127, no. 5, pp. 443–449, 2011.

[32] G. Vogiatzi, E. Oikonomou, G. Siasos et al., “Statins and inflammation in cardiovascular disease,” Current Pharmaceutical Design, vol. 23, 2017.

[33] X. Zhang, M. H. Liu, L. Qiao et al., “Ginsenoside Rb1 enhances atherosclerotic plaque stability by skewing macrophages to the M2 phenotype,” Journal of Cellular and Molecular Medicine, vol. 127, no. 5, pp. 443–449, 2011.

[34] Y. Yuan, M. Sun, and K. S. Li, “Astragalus mongholicus polysaccharide inhibits lipopolysaccharide-induced production of TNF-alpha and interleukin-8,” World Journal of Gastroenterology, vol. 15, no. 29, pp. 3676–3680, 2009.
[35] Y. Wang, P. Chen, C. Tang, Y. Wang, Y. Li, and H. Zhang, "Antinociceptive and anti-inflammatory activities of extract and two isolated flavonoids of Carthamus tinctorius L," Journal of Ethnopharmacology, vol. 151, no. 2, pp. 944–950, 2014.

[36] C. L. He, P. F. Yi, Q. J. Fan et al., "Xiang-Qi-Tang and its active components exhibit anti-inflammatory and anticoagulant properties by inhibiting MAPK and NF-xB signaling pathways in LPS-treated rat cardiac microvascular endothelial cells," Immunopharmacology and Immunotoxicology, vol. 35, no. 2, pp. 215–224, 2013.

[37] S. H. Kim, H. S. Park, M. J. Hong et al., "Tongqiaohuoxue decoction ameliorates obesity-induced inflammation and the prothrombotic state by regulating adiponectin and plasminogen activator inhibitor-1," Journal of Ethnopharmacology, vol. 192, no. 11, pp. 201–209, 2016.

[38] A. Undas, K. E. Brummel-Ziedins, and K. G. Mann, "Anticoagulant effects of statins and their clinical implications," Thrombosis and Haemostasis, vol. 111, no. 3, pp. 392–400, 2014.

[39] P. Libby, I. Tabas, G. Fredman, and E. A. Fisher, "Inflammation and its resolution as determinants of acute coronary syndromes," Circulation Research, vol. 114, no. 12, pp. 1867–1879, 2014.

[40] R. Sathanoori, F. Rosi, B. J. Gu et al., "Shear stress modulates endothelial KLF2 through activation of P2X4," Purinergic Signalling, vol. 11, no. 1, pp. 139–153, 2015.

[41] N. D. Perkins, "Integrating cell-signalling pathways with NF-kappaB and IKK function," Nature Reviews Molecular Cell Biology, vol. 8, no. 1, pp. 49–62, 2007.

[42] M. Karin and Y. Ben-Neriah, "Phosphorylation meets ubiquitination: the control of NF-kappaB activity," Annual Review of Immunology, vol. 18, no. 1, pp. 621–663, 2000.

[43] A. Oeckinghaus and S. Ghosh, "The NF-kappaB family of transcription factors and its regulation," Cold Spring Harbor Perspectives in Biology, vol. 1, no. 4, article a000034, 2009.

[44] X. Palomer, D. Álvarez-Guardia, M. M. Davidson, T. O. Chan, A. M. Feldman, and M. Vázquez-Carrera, "The interplay between NF-kappaB and E2F1 coordinately regulates inflammation and metabolism in human cardiac cells," PLoS One, vol. 6, no. 5, article e19724, 2011.

[45] A. Kauppinen, T. Suuronen, J. Ojala, K. Kaarniranta, and A. Salminen, "Antagonistic crosstalk between NF-kB and SIRT1 in the regulation of inflammation and metabolic disorders," Cellular Signalling, vol. 25, no. 10, pp. 1939–1948, 2013.

[46] N. Fakhrudin, B. Waltenberger, M. Cabaravdic et al., "Identification of plumericin as a potent new inhibitor of the NF-kB pathway with anti-inflammatory activity in vitro and in vivo," British Journal of Pharmacology, vol. 171, no. 7, pp. 1676–1686, 2014.

[47] A. Kumar, Z. Lin, S. SenBanerjee, and M. K. Jain, "Tumor necrosis factor alpha-mediated reduction of KLF2 is due to inhibition of MEF2 by NF-xB and histone deacetylases," Molecular and Cellular Biology, vol. 25, no. 14, pp. 5893–5903, 2005.
