C-type natriuretic peptide modulates quorum sensing molecule and toxin production in *Pseudomonas aeruginosa*

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*Pseudomonas aeruginosa* coordinates its virulence expression and establishment in the host in response to modification of its environment. During the infectious process, bacteria are exposed to and can detect eukaryotic products including hormones. It has been shown that *P. aeruginosa* is sensitive to natriuretic peptides, a family of eukaryotic hormones, through a cyclic nucleotide-dependent sensor system that modulates its cytotoxicity. We observed that pre-treatment of *P. aeruginosa* PAO1 with C-type natriuretic peptide (CNP) increases the capacity of the bacteria to kill *Caenorhabditis elegans* through diffusive toxin production. In contrast, brain natriuretic peptide (BNP) did not affect the capacity of the bacteria to kill *C. elegans*. The bacterial production of hydrogen cyanide (HCN) was enhanced by both BNP and CNP whereas the production of phenazine pyocyanin was strongly inhibited by CNP. The amount of 2-heptyl-4-quinolone (HHQ), a precursor to 2-heptyl-3-hydroxy-4-quinolone (*Pseudomonas* quinolone signal; PQS), decreased after CNP treatment. The quantity of 2-nonyl-4-quinolone (HNQ), another quinolone which is synthesized from HHQ, was also reduced after CNP treatment. Conversely, both BNP and CNP significantly enhanced bacterial production of acylhomoserine lactone (AHL) [e.g. 3-oxo-dodecanoyl-homoserine lactone (3OC12-HSL) and butanoylhomoserine lactone (C4-HSL)]. These results correlate with an induction of *lasI* transcription 1 h after bacterial exposure to BNP or CNP. Concurrently, pre-treatment of *P. aeruginosa* PAO1 with either BNP or CNP enhanced PAO1 exotoxin A production, via a higher *toxA* mRNA level. At the same time, CNP led to elevated amounts of *algC* mRNA, indicating that *algC* is involved in *C. elegans* killing. Finally, we observed that in PAO1, Vfr protein is essential to the pro-virulent effect of CNP whereas the regulator PtxR supports only a part of the CNP pro-virulent activity. Taken together, these data reinforce the hypothesis that during infection natriuretic peptides, particularly CNP, could enhance the virulence of PAO1. This activity is relayed by Vfr and PtxR activation, and a general diagram of the virulence activation cascade involving AHL, HCN and exotoxin A is proposed.

**INTRODUCTION**

*Pseudomonas aeruginosa* is a ubiquitous Gram-negative opportunistic pathogen that can infect different hosts including mammals (Rahme et al., 1995), insects (Jander et al., 2000), nematodes (Tan & Ausubel, 2000) and plants (Rahme et al., 2000). In humans, *P. aeruginosa* causes serious infections in immunocompromised hosts, is one of the major micro-organisms responsible for nosocomial diseases (Govan & Deretic, 1996) and is the predominant

Abbreviations: BNP, brain natriuretic peptide; CNP, C-type natriuretic peptide; ETA, exotoxin A; HCN, hydrogen cyanide; H-HQ, 2-heptyl-4-quinolone; HNQ, 2-nonyl-4-quinolone; LPS, lipopolysaccharide; PQS, *Pseudomonas* quinolone signal; qRT-PCR, quantitative reverse transcription-PCR; QS, quorum sensing.

A supplementary table of primer sequences is available with the online version of this paper.
cause of morbidity and mortality in cystic fibrosis patients (Adams et al., 1998; Govan & Deretic, 1996).

During infection, the stress status of the host promotes the release and circulation of eukaryotic signal molecules facilitating their contact with bacteria. Some reports suggest that P. aeruginosa PAO1 may actively sense eukaryotic signal molecules (Lesouhaitier et al., 2009), such as gamma-interferon (Wu et al., 2005) or dynorphin (Zaborina et al., 2007) which trigger bacterial virulence factor production. Natriuretic peptides represent a family of eukaryotic hormones which comprises at least three major members: atrial natriuretic peptide (ANP), brain natriuretic peptide (BNP) and C-type natriuretic peptide (CNP). These peptides are principally produced by atrial (ANP and BNP) and endothelial (CNP) cells (de Bold et al., 1996; Suga et al., 1992). It has been observed that BNP concentration is increased in human blood during septic shock (Post et al., 2008; Rudiger et al., 2006) and that plasma N-terminal pro-brain natriuretic peptide (NT-proBNP) increases in a model of systemic exposure to Escherichia coli endotoxin (or lipopolysaccharide, LPS) in healthy volunteers (Vila et al., 2008). Moreover, LPS potently stimulates CNP secretion by endothelial cells (Suga et al., 1993). Thus, it appears that bacteria are exposed to natriuretic peptides in the plasma during the host infection process. Natriuretic peptides possess a structure similar to cationic antimicrobial peptides (Bulet et al., 2001) and have been first proposed as antimicrobial peptides active against a large spectrum of microorganisms (Krause et al., 2001). However, we have recently demonstrated that, at a micromolar range, neither human BNP nor CNP act on P. aeruginosa survival but are able to enhance the global cytotoxicity of the bacterium (Veron et al., 2007). The sensitivity of P. aeruginosa to natriuretic peptides appears to be relayed by the activation of a cyclase leading to an increase of intracellular cyclic AMP concentration and to a stimulation of the Vfr global regulator (Veron et al., 2007). This implication of the Vfr protein suggests that numerous virulence factors or bacterial physiological parameters may be regulated by CNP in P. aeruginosa.

To make its infectious process successful, P. aeruginosa uses a large arsenal of both secreted and cell-associated virulence factors (Kipnis et al., 2006; Lyczak et al., 2000). In addition, it has been proposed that P. aeruginosa outer membrane vesicles, which contain numerous secreted factors and cell-membrane-associated virulence factors, may represent another weapon used by this bacterial pathogen (Bomberger et al., 2009; Kuehn & Kesty, 2005; Mashburn & Whiteley, 2005). In Pseudomonas, the regulation of the expression of a wide variety of survival and virulence mechanisms is controlled in a cell-density-dependent manner through a process called quorum sensing (QS) (Juhas et al., 2005; Rumbaugh et al., 2000). Global regulation through QS systems controls population behaviours and plays fundamental roles in bacterial pathogenicity (Fuqua et al., 2001; Smith & Iglewski, 2003; Winzer & Williams, 2001). P. aeruginosa QS is principally represented by the Las and Rhl systems. LasI and RhlI synthesize their cognate signal molecules 3-oxo-dodecanoyl-homoserine lactone (3O-C12-HSL) and butanoylhomoserine lactone (C4-HSL), respectively. These two diffusible molecules induce gene expression by binding to their respective receptor proteins, LasR and RhlR, which are also transcriptional activators (Pearson et al., 1995). A third signal has been found to participate in the P. aeruginosa QS network, the quinolone 2-heptyl-3-hydroxyl-4-quinolone, termed Pseudomonas quinolone signal (PQS) (Pesci et al., 1999). PQS fits the functional criteria of a QS signal, such as cell-density-dependent accumulation (Lépine et al., 2003) and recognition by bacterial cells in which it triggers a specific transcriptional response (Camilli & Bassler, 2006). PQS is synthesized by the mono-oxygenase PqsH which converts the 2-heptyl-4-quinolone (HHQ) into PQS (Gallagher et al., 2002). Since it is released from and taken up by bacteria, HHQ is also considered to act as an intercellular message molecule (Déziel et al., 2004). PQS signalling plays an important role in P. aeruginosa pathogenesis because it regulates diverse target genes including those coding for elastase, rhamnolipid, the PA-IL lectin and pyocyanin, as well as influencing biofilm development (Diggle et al., 2006).

Among P. aeruginosa secreted virulence factors, exotoxin A (ETA) appears to be essential for the success of bacterial infection (Fogle et al., 2002; Hamood et al., 1996; Storey et al., 1998). The blue phenazine pigment pyocyanin is also an important exotoxin secreted by P. aeruginosa (Diggle et al., 2003). This molecule is directly toxic to both prokaryotic (Hassan & Fridovich, 1980) and eukaryotic (Denning et al., 1998) cells and also metazoans such as Caenorhabditis elegans (Mahajan-Miklos et al., 1999). Moreover, pyocyanin induces apoptosis in human neutrophils, favouring the persistence of P. aeruginosa in host tissues and promoting bacterial host defence evasion (Usher et al., 2002). Hydrogen cyanide (HCN) is a diffusible gas produced by P. aeruginosa and also represents a formidable toxic factor against eukaryotic organisms including C. elegans (Gallagher & Manoil, 2001) and humans (Anderson et al., 2010).

In the present study, we examined the impact of natriuretic peptides on the global virulence of PAO1 using the infectious model C. elegans. The effect of natriuretic peptides on P. aeruginosa production of both major virulence factors and communication signal molecules was determined. Finally, we identified key steps of the molecular mechanism involved in CNP action on P. aeruginosa.

**METHODS**

**Bacterial cultures and tested molecules.** P. aeruginosa PAO1 was obtained from an international collection (Veron et al., 2007). P. aeruginosa MPAO1 and mutants obtained by transposon insertion
CNP enhances *Pseudomonas aeruginosa* virulence

**C. elegans** synchronization and virulence assays. The *C. elegans* wild-type Bristol strain N2 was obtained from the Caenorhabditis Genetics Center (Minneapolis, USA). *C. elegans* was maintained under standard culturing conditions at 22 °C on nematode growth medium (NGM; all per litre, 3 g NaCl, 2.5 g peptone, 17 g agar, 5 mg cholesterol, 1 ml 1 M CaCl₂, 1 ml 1 M MgSO₄, 25 ml 1 M KH₂PO₄) agar plates with *E. coli* OP50 as a food source (Sulston & Hodgkin, 1988). Synchronous cultures of worms were generated after exposure of the adult worm population to a sodium hypochlorite/sodium hydroxide solution as described previously (Stiernagle, 1999). The resulting eggs were incubated at 22 °C on an *E. coli* OP50 lawn until the worms reached the L4 (48 h) life stage (confirmed by light microscopy).

Pathogen lawns used for *C. elegans* survival assays were prepared by spreading 50 μl *P. aeruginosa* strains (control and treated) on 35 mm peptoneglucose/sorbitol (PGS; 1 % Bacto-peptone, 1 % NaCl, 1 % glucose, 0.15 M sorbitol, 1.7 % Bacto-agar) conditioned Petri dishes for fast-killing evaluation or on 35 mm NGM conditioned Petri dishes supplemented with 0.05 mg 5-fluoro-2′-deoxyuridine ml⁻¹ for slow killing determination. This nucleotide analogue blocks the DNA synthesis, thus preventing offspring from the experimental animals. The plates were incubated overnight at 37 °C and then placed at room temperature for 4 h. Between 15 and 20 L4 synchronized worms were harvested with M9 solution (per litre, 3 g KH₂PO₄, 6 g NaHPO₄, 5 g NaCl, 1 ml 1 M MgSO₄), placed on the 35 mm assay Petri dishes and incubated at 22 °C. Worm survival was scored at 1 h, 24 h and each subsequent day, using an Axiovert S100 optical microscope (Zeiss) equipped with a Nikon digital camera DXM 1200F (Nikon Instruments). The results were expressed as the percentage of living worms and are the average of three independent assays.

**Pyocyanin assay.** *P. aeruginosa* PAO1 was grown at 37 °C, in shaking conditions at 180 r.p.m. in ONB medium with either physiological water (control), BNP or CNP (1 μM). Following 24 h incubation, bacterial cells were centrifuged at 8000 g for 10 min, and 2 ml of supernatant was extracted using 2 ml chloroform and re-extracted with 1 ml 0.5 M HCl. The pyocyanin concentration was determined by measurement of OD₅₃₀.

**HSL and PQS extraction and quantification.** C4-HSL and 3OC12-HSL were extracted and quantified by liquid chromatography-mass spectrometry (LC-MS-MS) according to Morin et al. (2003). PQS, HHQ and 2-nonyl-4-quinolone (HNQ) were extracted as the HSLs (Morin et al., 2003) and characterized as described by Détzel et al. (2004) and Bazire et al. (2005). After the bacterial cells were removed from the growth medium by centrifugation (2500 g, 10 min), the supernatants were extracted twice with one vol. HPLC-grade dichloromethane. The dichloromethane extracts were dried over anhydrous magnesium sulfate, filtered and evaporated. The residue was dissolved in HPLC-grade acetonitrile and analysed using LC-MS-MS. The proportion of each PQS-related molecule was quantified from the corresponding m/z [M + H]⁺ chromatograms by integration of peak areas.

**Elastase assay.** The elastolytic activity of *P. aeruginosa* culture supernatants was determined by using the elastin Congo red (ECR) assay (Pearson et al., 1997). *P. aeruginosa* PAO1 was grown at 37 °C, in shaking conditions at 180 r.p.m. in ONB medium and exposed to either physiological water (control), BNP or CNP (1 μM). After 3 h incubation, bacterial cells were centrifuged at 2500 g for 10 min and the supernatants were filtered. Triplicate 50 μl samples of culture filtrates were added to tubes containing 20 mg ECR (Sigma-Aldrich) and 1 ml buffer (0.1 M Tris, pH 7.2, 1 mM CaCl₂). The samples were incubated for 18 h at 37 °C with rotation; they were then placed on ice and 0.1 ml 0.12 M EDTA was added. Insoluble ECR was removed by centrifugation, and the OD₄₅₃ of the supernatant was measured.

**Azocasein assay.** Proteinase secretion was quantified by the azocasein assay as outlined by Kessler et al. (1982) with modifications. Azocasein (Sigma-Aldrich) was dissolved to 5 mg ml⁻¹ in assay buffer containing 0.1 M Tris/HCl (pH 8). The *P. aeruginosa* PAO1 culture was incubated for 3 h with either physiological water (control), BNP and the results were expressed as the percentage of living worms and are the average of three independent assays.

Table 1. Strains used in this study

| Strain         | Characteristic | Reference or source                      |
|----------------|----------------|------------------------------------------|
| *P. aeruginosa*|                |                                          |
| PAO1           | Wild-type      | Veron et al. (2007)                      |
| MPAO1          | Wild-type      | Holloway et al. (1979)                   |
| PW6882         | MPAO1 mutant   | University of Washington Genome Center   |
|                | rhlK-B10::I(SlacZ)hah |                                    |
| PW3597         | MPAO1 mutant   | University of Washington Genome Center   |
|                | lasR-B10::I(SlacZ)hah |                                      |
| PW3601         | MPAO1 mutant   | University of Washington Genome Center   |
|                | lasl-F07::I(SlacZ)hah |                                   |
| PW6880         | MPAO1 mutant   | University of Washington Genome Center   |
|                | rhlL-D03::I(SphoA)hah |                                  |
| PW2181         | MPAO1 mutant   | University of Washington Genome Center   |
|                | vfr-G11::I(SlacZ)hah |                                  |
| PW2283         | MPAO1 mutant   | University of Washington Genome Center   |
|                | toxR/regA-E04::I(SlacZ)hah |                               |
| PW4833         | MPAO1 mutant   | University of Washington Genome Center   |
|                | ptxR-G01::I(SlacZ)hah |                               |
| PW9967         | MPAO1 mutant   | University of Washington Genome Center   |
|                | algC-D07::I(SphoA)hah |                               |
| PW2810         | MPAO1 mutant   | University of Washington Genome Center   |
|                | phnB-H01::I(SlacZ)hah |                               |
| *E. coli*      |                |                                          |
| OP50           |                | Stiernagle (1999)                        |

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or CNP, and media were removed and centrifuged to pellet the cells. The azocasein solution (250 μl) was mixed with 250 μl bacterial supernatants and incubated in a 37 °C water bath for 30 min. The reactions were stopped by adding 500 μl 10% TCA, and the reaction mixtures were allowed to stand at −20 °C for 10 min. Samples were then centrifuged for 5 min at 8000 g, and 500 μl of each supernatant was added to 500 μl 0.5 M NaOH. A$_{450}$, corresponding to released azocasein, was determined with a spectrophotometer.

**HCN assay.** HCN concentration in bacterial culture medium was determined by polarographic analysis. Voltametric measurements were made using a Metrohm (Herisau, Swiss) 757 VA Compratrace. Analysis was carried out in a three-electrode configuration using an Ag/AgCl 3 mol KCl 1$^{-1}$ reference, a platinum wire as a counter electrode and a multi-mode mercury electrode as a working electrode. All experiments were performed at a room temperature. Bacterial cell cultures were centrifuged for 10 min at 8000 g and filtered at 0.22 μm. The filtered supernatants were diluted in a 0.2 mol borate 1$^{-1}$ electrolyte (pH 10.2). The solution was purged for 3 min with nitrogen to remove dissolved oxygen and then for 20 s more between each cyanide addition. A potential scan was carried out in a negative direction from −0.1 to −0.5 V with a sweep rate of 10 mV s$^{-1}$. The pulse amplitude was 0.05 V with a pulse duration of 0.04 s. The peak height of cyanide was measured at −200 mV in a differential pulse mode and the cyanide concentration was determined by adding four successive amounts of potassium cyanide standard (10 mg l$^{-1}$).

**ETA quantification.** The assay was done using a protocol adapted from Coligan et al. (2001). Briefly, throughout the assay, the 96-well microplates (MaxiSorp F; Nunc) were washed with PBST (0.02%, v/v, Tween 20 in PBS). Each well was coated with 100 μl diluted goat anti-ETA antibody (0.25 mg ml$^{-1}$ in 100 mM Na$_2$HCO$_3$) (Sigma-Aldrich) overnight at 4 °C. The plates were washed and treated with 1 mg BSA ml$^{-1}$ in PBST for 1 h at 37 °C. The plates were then washed twice and incubated with supernatant fractions (100 μl per well) for 1 h at 37 °C. Serial dilutions (512–2 pg ml$^{-1}$) of purified ETA (Sigma-Aldrich) in PBST were used as standards. The plates were washed six times and incubated for 1 h at 37 °C with rabbit anti-ETA (100 μl per well) (Sigma-Aldrich) diluted in PBST. The plates were then washed six times and incubated for 1 h at 37 °C with goat anti-rabbit IgG conjugated to horseradish peroxidase (Sigma-Aldrich). The plates were washed again six times, and bound ETA (Sigma-Aldrich) in PBST were used as standards. The plates were then washed six times and incubated for 1 h at 37 °C with rabbit anti-ETA (100 μl per well) (Sigma-Aldrich) diluted in PBST. The plates were washed six times, incubated for 1 h at 37 °C with goat anti-rabbit IgG conjugated to horseradish peroxidase (Sigma-Aldrich). The plates were washed again six times, and bound ETA was revealed by incubation at 37 °C for 30 min with 100 μl TMB (ImmunoPure TMB substrate, Sigma-Aldrich). The reaction was stopped by adding 100 μl 2 M H$_2$SO$_4$ per well. A$_{450}$ was read using an ELISA plate reader (Bio-Rad 680 XR). The values were standardized by dividing the amount of ETA (pg ml$^{-1}$) from each supernatant fraction by the OD$_{340}$ of the culture from which that fraction was obtained.

**mRNA assay by quantitative reverse transcription-PCR (qRT-PCR).** RNA quantification was performed as described previously (Bazire et al., 2010) from bacteria grown for 3 h in liquid ONB medium. The primers used are given in Supplementary Table S1 (available with the online version of this paper). 16S rRNA was used as an endogenous control (Corbella & Puyet, 2003). PCRs were performed in triplicate and the standard deviations were lower than 0.15 C$_T$. The relative quantifications were obtained as described previously (Bazire et al., 2009), using the comparative C$_T$ (2$^{-\Delta\Delta C_T}$) method (Livak & Schmittgen, 2001).

**Statistical analysis.** For the killing assay, nematode survival was calculated by using the Kaplan–Meier method, and survival differences were tested for significance by using the log–rank test (GraphPad Prism version 4.0, GraphPad Software). For other results, each value reported for the assays is the mean measurement for a minimum of three independent preparations. The non-parametric Mann–Whitney test was used to compare the means within the same set of experiments.

**RESULTS**

**Effect of natriuretic peptides on PAO1 virulence towards C. elegans**

Depending on the experimental conditions, *P. aeruginosa* kills *C. elegans* in different ways. In a low-salt medium such as NGM, *P. aeruginosa* kills the nematodes over a period of a few days (‘slow killing’) by an infection-like process in which the bacteria accumulate in the worm intestines (Tan et al., 1999).

When feeding worms with untreated PAO1 (control), it took 6 days to kill 50% of *C. elegans* (Fig. 1a). Using PAO1 treated with CNP (1 μM) as food, the worms died more rapidly and only 30% of worms were alive at day 5 (Fig. 1a). Using the statistical log–rank test we observed that CNP-treated bacteria were more virulent to *C. elegans* than the control bacteria (PAO1 control versus CNP-exposed PAO1; P=0.0002). In contrast, bacteria exposed to BNP (10$^{-6}$ M) appeared less virulent towards *C. elegans* with a median survival at day 7 (Fig. 1a).

*P. aeruginosa* is also able to kill *C. elegans* within a few hours (‘fast killing’) by producing a number of diffusible toxins, including phenazines (Aballay et al., 2003; Mahajan-Miklos et al., 1999; Tan et al., 1999) or HCN (Gallagher & Manoil, 2001). This phenomenon is observed when bacteria are spread in a lawn onto a high-osmolarity medium. When *C. elegans* was dropped on PAO1 control bacteria grown on a high-osmolarity medium, we observed that 60.8±5.2% of worms were still alive after 48 h (Fig. 1b). The percentage of living worms on CNP-treated bacteria fell to 43.4±3.6% (Fig. 1b). When *C. elegans* was dropped on BNP-exposed bacteria we observed a slight, but not significant, decrease in the number of worms surviving after 48 h (49.6±5.9%) (Fig. 1b). Dead worms were stiff and colourless, whereas living worms were dark and curved (Fig. 1c).

**Effect of natriuretic peptides on HCN production by PAO1**

In order to identify virulence factors produced by PAO1 after exposure to BNP and CNP that are involved in *C. elegans* killing, we investigated the effect of natriuretic peptides on HCN, which is the primary diffusible toxic factor that PAO1 produces to kill *C. elegans* (Gallagher & Manoil, 2001).

The amount of HCN was measured after exposure of bacteria (3 h) to BNP, CNP (both 10 μM) or physiological water (control) during their exponential phase of growth. The results were standardized by dividing the amount of HCN (μg l$^{-1}$) measured in each fraction by the quantity of bacterial proteins from which the fraction was obtained.
The treatment of *P. aeruginosa* PAO1 with BNP or CNP significantly increased HCN production, which reached 144.0 ± 8.7% and 149.7 ± 22.3% of the control value, respectively (Fig. 2a). In order to determine the origin of the increase in HCN production, we examined the *hcnB* mRNA levels by qRT-PCR 1 h after exposing bacteria to BNP or CNP. The *hcnB* mRNA level rose (4.18-fold) following CNP exposure compared with untreated PAO1, whereas BNP did not significantly affect the level of *hcnB* mRNA (Fig. 2b).

**Effect of natriuretic peptides on production of pyocyanin, PQS and PQS-related molecules by PAO1**

Pyocyanin is a virulence factor involved in the lethal activity of *P. aeruginosa* on *C. elegans*. The impact of pyocyanin, however, depends on the *P. aeruginosa* strain. For instance, in strain PA14, pyocyanin production triggers...
C. elegans death (Mahajan-Miklos et al., 1999) whereas in PAO1 there is apparently no correlation between pyocyanin and worm killing (Gallagher & Manoil, 2001). We nevertheless investigated whether pyocyanin could be involved in C. elegans killing by PAO1 in our culture conditions and in the pro-virulent effect of CNP.

We first observed that the \( \text{phnB} \) mutant of PAO1 grown in ONB medium killed C. elegans with the same kinetics as the wild-type strain (data not shown). Pyocyanin production was measured in culture supernatant (24 h, 37 °C) after exposure of bacteria to BNP or CNP (10\(^{-6}\) M). We observed a decrease in pyocyanin production of 65.1 ± 4.3 % for CNP-treated bacteria (Fig. 3a). In contrast, when bacteria were exposed to BNP, no significant variation in pyocyanin was observed (Fig. 3a). The \( \text{phzC1} \) mRNA level was halved after CNP exposure compared with untreated PAO1, whereas BNP did not affect the amount of \( \text{phzC1} \) mRNA (Fig. 3b).

Pre-treatment of \( P. \text{aeruginosa} \) PAO1 with CNP induced a slight decrease in the capacity of the bacteria to produce PQS. Three hours after CNP exposure, PQS production was lowered by 12.3 ± 11.7 % compared with the control value (Fig. 3c). HHQ production by bacteria exposed to CNP was significantly lowered, by 26.7 ± 7.1 %, compared with the control value (Fig. 3d). Pre-treatment of \( P. \text{aeruginosa} \) PAO1 with CNP also induced a significant decrease in the bacterial capacity to produce HNQ. Three hours after exposure to CNP, HNQ production was reduced in the same range as was HHQ (a 29.2 ± 7.0 % decrease compared with the control value) (Fig. 3e).

**Effect of natriuretic peptides on HSL production by PAO1**

The \( P. \text{aeruginosa} \) PAO1 QS systems LasI/LasR and RhlI/RhlR regulate virulence factor production (Smith & Iglewski, 2003) and are involved in the bacterial killing of...
C. elegans (Darby et al., 1999). The amount of C4-HSL and 3OC12-HSL produced by P. aeruginosa was measured after exposure of bacteria to BNP, CNP or physiological water for 3 h during their exponential phase of growth. Pre-treatment of P. aeruginosa with either BNP or CNP led to an increase in C4-HSL production compared with untreated bacteria of 156.2 ± 23.7% and 160.6 ± 10.6%, respectively (Fig. 4a). We observed that the increase in C4-HSL production was maintained 5 h after exposure to BNP and CNP, and the levels of C4-HSL returned to the control values 9 h after the onset of the experiment (data not shown). We examined the rhlI mRNA levels by qRT-PCR. RNA was extracted from bacteria harvested after 1 h of exposure to BNP, CNP or physiological water. We observed that the rhlI mRNA levels were not modified by exposure of PAO1 to BNP or CNP compared with untreated bacteria (Fig. 4b).

Pre-treatment of P. aeruginosa with BNP and CNP for 3 h also induced an increase in the capacity of the bacteria to produce 3OC12-HSL. Bacteria exposed to BNP and CNP showed an increase in 3OC12-HSL production which reached 156.0 ± 33.7% and 150.1 ± 25.6% of the control value (Fig. 4c). This increase in 3OC12-HSL production was maintained up to 5 h after exposure of PAO1 to BNP and CNP, and returned to the basal level within 9 h (data not shown). We examined the lasI mRNA levels by qRT-PCR. While the lasI mRNA level slightly increased after BNP exposure (×1.37), PAO1 exposure to CNP induced a significantly higher rise in lasI mRNA levels (×2.51) compared with untreated PAO1 (Fig. 4d).

Using mutants, we evaluated the involvement of LasI/LasR and RhlI/RhlR systems on the lethal activity of PAO1 on C. elegans after growth in ONB medium. We first observed that lasI and rhlI mutants possessed a capacity to kill C. elegans in the same range as the wild-type (Fig. 5a). In contrast, lasR and rhlR mutants appeared less virulent than the wild-type strain (Fig. 5a).

In order to evaluate a direct toxic effect of HSLs on C. elegans, we exposed worms to E. coli OP50 cultivated on NGM plates supplemented with C4-HSL or 3OC12-HSL (0.5 mg l⁻¹). We observed that neither C4-HSL nor 3OC12-HSL had a significant effect on C. elegans survival kinetics (Fig. 5b).

**Effect of natriuretic peptides on ETA production by PAO1**

P. aeruginosa secretes numerous extracellular products, including ETA, elastase and protease, that are important for virulence and that are regulated by QS molecules (de Kievit & Iglewski, 2000). The treatment of P. aeruginosa PAO1 with BNP or CNP for 3 h did not modify elastase and protease (azocasein hydrolytic) activities of the bacteria (data not shown).

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**Fig. 4.** Effect of BNP and CNP on AHL production by P. aeruginosa PAO1. (a) and (c) Relative amounts of C4-HSL (a) and 3OC12-HSL (c) in P. aeruginosa PAO1 supernatants 3 h after exposure to physiological water (1) or natriuretic peptides [BNP (2) and CNP (3)]. Data are the mean ± SEM of 11 values obtained from five independent experiments for C4-HSL and of eight values obtained from four independent experiments for 3OC12-HSL. The mean C4-HSL and 3OC12-HSL levels in control conditions were 188.4 ± 25.8 μM and 19.2 ± 2.9 μM, respectively. Significant difference is indicated by asterisks: *P<0.05; **P<0.01; ***P<0.001. (b) and (d) Expression levels of rhlI and lasI in PAO1-treated bacteria relative to those in the PAO1 control. RNA was extracted 1 h after exposure of PAO1 to physiological water or natriuretic peptides (BNP and CNP) and assayed by qRT-PCR. See the legend to Fig. 2 for further information.
The amount of ETA produced by *P. aeruginosa* PAO1 was determined by ELISA (Coligan et al., 2001). The results were standardized by dividing the amount of ETA (pg ml⁻¹) measured in each fraction by the OD₅₉₀ of the culture medium from which the fraction was obtained. The treatment of *P. aeruginosa* PAO1 with BNP or CNP for 3 h significantly increased ETA production, which reached 127.1 ± 8.4 % and 129.7 ± 8.1 % of the control values, respectively (Fig. 6a).

In order to determine the origin of the increase in ETA production, we examined the toxA mRNA levels by qRT-PCR. The toxA mRNA level was increased by exposure to either BNP or CNP compared with untreated PAO1 (× 2.47 and × 2.83, respectively) (Fig. 6b). ToxR (RegA) is a regulator of ETA production in PAO1. We observed that a toxR (regA) mutant lost all lethal activity in the *C. elegans* model and that both BNP and CNP are unable to increase toxR mutant virulence (Fig. 6c).

### Implication of Vfr in PAO1 response to natriuretic peptides

The protein Vfr is a global regulator of PAO1 virulence and is activated by cAMP. Since it has been shown that CNP induces a twofold increase of cAMP into PAO1 (Veron et al., 2007), we investigated the role of Vfr in the CNP pro-virulence effect in the *C. elegans* model.

We observed that the vfr mutant dramatically lost its ability to kill *C. elegans* in both slow-killing (Fig. 7a) and fast-killing (Fig. 7b) tests. When the vfr mutant was exposed to BNP or CNP, we observed no modification in the virulence activity of this strain (Figs 7a and 8a). In addition, we noted that the vfr mutant produced a small amount of ETA, the levels of which remained unchanged even after exposure to BNP or CNP (Fig. 7c).

### Role of PtxR in the effect of natriuretic peptides on PAO1

PtxR regulates ETA production in PAO1 (Hamood et al., 1996) and is under the control of Vfr (Ferrell et al., 2008). We decided to evaluate the role of the PtxR regulator on the effect of CNP on PAO1.

We observed that the ptxR mutant partially lost its virulence towards *C. elegans* in the slow-killing model (Fig. 8a). In contrast, the virulence of this mutant was retained in a fast-killing test (data not shown). When the ptxR mutant was exposed to BNP or CNP, we observed no modification of the virulence pattern of this strain (Fig. 8a). In addition, we remarked that, as in the vfr mutant, the level of ETA produced by the ptxR mutant was low and not affected by BNP and CNP (Fig. 8b). We examined ptxR mRNA levels by qRT-PCR. The level of ptxR mRNA increased slightly after exposure to BNP (× 1.76), whereas exposure to CNP had a more pronounced effect (the ptxR mRNA level was 3.42 times greater than that of untreated PAO1) (Fig. 8c).

### Role of algC in the effect of natriuretic peptides on PAO1

It has been shown that cAMP, after binding to Vfr, modulates the structure of LPS in PAO1 (Veron et al., 2007) and that algC is required for the synthesis of a complete LPS core, which is involved in the lethal effect of bacteria on *C. elegans* (Gallagher & Manoil, 2001). We therefore investigated the role of algC in the pro-virulence effect of CNP on PAO1. We observed that the algC mutant grown in ONB medium dramatically lost its ability to kill *C. elegans* in the slow-killing conditions (Fig. 9a). When
the algC mutant was exposed to BNP or CNP we observed no modification in the virulence activity of this strain (Fig. 9a). In addition, the level of algC mRNA measured by qRT-PCR was not significantly modified after BNP exposure (×1.50) whereas CNP exposure doubled the algC mRNA level compared with untreated PAO1 (Fig. 9b).

DISCUSSION

Originally, we described that natriuretic peptides, in particular CNP, enhance P. aeruginosa PAO1 cytotoxicity through the activation of Vfr, a global regulator of bacterial virulence (Veron et al., 2007). Since the pro-cytotoxic effect of natriuretic peptides was only observed in vitro, we decided to investigate further the mechanism of action of these molecules by evaluating their impact on bacterial virulence using the more complex pluricellular model C. elegans. In the present study, we provide evidence that CNP enhances the global virulence of PAO1 to C. elegans, by increasing the secretion of HCN and ETA through stimulation of the regulatory proteins Vfr and PtxR and activation of QS.

Our experiments indicate that CNP is involved in the regulation of the virulence of P. aeruginosa to C. elegans, both through the infection-like process (slow killing) and the production of diffusible toxins (fast killing). The lethal effect of Pseudomonas on C. elegans is multifactorial but HCN appears to be the primary diffusible toxic factor killing C. elegans (Gallagher & Manoil, 2001). The lethal effect of Pseudomonas on C. elegans could also be mediated by the blue phenazine pigment pyocyanin, which is known to be essential in the P. aeruginosa PA14 fast-killing test (Mahajan-Miklos et al., 1999). Pyocyanin is the major phenazine produced by P. aeruginosa and it acts as a virulence factor (Lau et al., 2004) that is directly toxic to isolated eukaryotic cells (Denning et al., 1998), prokaryotic cells (Hassan & Fridovich, 1980) and complex organisms such as C. elegans (Mahajan-Miklos et al., 1999). We observed that pyocyanin produced by PAO1 in ONB medium (this study) is not involved in C. elegans killing, confirming that PAO1 does not act on C. elegans through this diffusible factor (Gallagher et al., 2002). However, we noted that although pyocyanin is not involved in the PAO1 killing activity, CNP modulates pyocyanin production. This result is consistent with our hypothesis that CNP acts on Pseudomonas through different mechanisms and toxic molecules.

Bacteria must integrate extra- and intracellular information to respond appropriately to environmental changes (Miller et al., 1989). The pathways by which pathogenic bacteria are
activated by host signals appear to be, like receptor–ligand interactions, specific to bacteria and to the host signal involved (for a review see Lesouhaitier et al., 2009). Concerning *P. aeruginosa*, the sensitivity to eukaryotic signal molecules comes with the regulation of bacterial QS molecule production (Wu et al., 2005; Zaborina et al., 2007). Since QS-dependent exoproducts such as pyocyanin production are under the influence of PQS (Déziel et al., 2004; Gallagher et al., 2002), we evaluated the impact of CNP on the production of PQS and related molecules. By analysing the PQS production profile as a function of growth, it has been suggested that PQS is not maximally produced until the late stationary phase and that very little PQS is present when the bacterial cells enter the stationary phase (McKnight et al., 2000). However, although PQS levels are indeed maximal in late stationary phase, the molecule is already detectable in the exponential phase of growth, and it was demonstrated that PQS, apart from being important in the late stationary phase, may also have a function much earlier in the growth phase (Diggle et al., 2003; Guina et al., 2003). Then, we measured the impact of CNP on the capacity of *P. aeruginosa* PAO1 to produce PQS and its precursor HHQ at the end of the exponential growth phase. We observed that CNP slightly reduces PQS production, apparently as a consequence of a significant decrease in its precursor HHQ synthesis. This hypothesis is supported by the observation that a reduction in HNQ, which is derived from HHQ, as is PQS. Nevertheless, these results indicate that the effect of CNP on PAO1 virulence is not mainly mediated by the PQS system.

The production of PQS in *P. aeruginosa* is finely regulated and interconnected with cell-to-cell signals AHLs (Wade et al., 2005). Among AHLs, the two chemically distinct molecules (C4-HSL, 3OC12-HSL), which act in concert with PQS, control the expression of many genes that are eventually involved in the global virulence of the bacterium (Diggle et al., 2007; Xiao et al., 2006). We observed that the pro-virulent effects of natriuretic peptides on *P. aeruginosa* PAO1 could be supported by modifications of C4-HSL and 3OC12-HSL production. Both BNP and CNP significantly enhance the bacterial production of C4-HSL and 3OC12-HSL. These effects were observed during the end of the exponential phase and 3 h after bacterial exposure to BNP or CNP, and appeared transient, since the levels of C4-HSL and 3OC12-HSL returned to the control values 9 h later.
These results are consistent with previous studies showing that the cytotoxic effect of PAO1 is enhanced by BNP and CNP (Veron et al., 2007) and we suggest that the increase in AHL supports a part of the pro-virulent action of natriuretic peptides on P. aeruginosa PAO1. It has been proposed that 3OC12-HSL, in addition to its informative activity, can act by itself as a virulence factor able to induce inflammation in vivo (Smith et al., 2002), to accelerate apoptosis in macrophages and neutrophils (Tateda et al., 2003), and to interact directly with immune cell membranes (Davis et al., 2010). However, we verified that both 3OC12-HSL and C4-HSL are unable to kill C. elegans directly, indicating that HSL are not or are only slightly toxic on a complex organism such as C. elegans. Analysis of the time-course synthesis of HSL in PAO1 showed that 3OC12-HSL is first synthesized under lasI activation and that C4-HSL is subsequently produced under rhlI activation (Pesci et al., 1997). In our study, we observed that 1 h after PAO1 exposure to CNP, the amount of lasI mRNA was enhanced by 2.5-fold whereas the amount of rhlI mRNA remained unchanged. The fact that both C4-HSL and 3OC12-HSL are overproduced 3 h after PAO1 exposure to CNP is in agreement with the 3OC12-HSL-dependent mechanism of C4-HSL synthesis and suggests that CNP activates bacterial internal regulators that act through the same cascade of events. Both LasI/R and RhlI/R QS systems of P. aeruginosa are involved in C. elegans killing (Steindler et al., 2009). We observed that only lasR and rhlR mutants lose their virulence, whereas the virulence of lasI and rhlI mutants is preserved, which shows a clear dissociation between the functions of lasR and rhlR receptor and their classical ligands, as previously suggested (Darby et al., 1999; Steindler et al., 2009). Moreover, these results reinforce the recent hypothesis proposing that LasR can switch into a functional conformation in the absence of an AHL ligand (Sappington et al., 2011).

Among P. aeruginosa secreted virulence factors, ETA is the most toxic, and this toxin appears indispensable to the infectious success of the bacterium (Fogle et al., 2002; Hamood et al., 1996; Storey et al., 1998). The production of ETA by P. aeruginosa depends on environmental conditions such as iron concentration (Hamood et al., 2004; Liu, 1973), but it was also reported to be regulated by a neurotransmitter such as norepinephrine (Li et al., 2009). We investigated a possible action of natriuretic peptides on ETA production by PAO1. We observed that both BNP and CNP increase the ETA production by PAO1. We observed that both BNP and CNP increase the ETA production by PAO1. This effect appears to be correlated with an increase in the transcription of the toxA gene, suggesting that ETA synthesis is genomically regulated by natriuretic peptides. It has been shown that, in P. aeruginosa PA14, ETA is involved in C. elegans fast and/or slow killing (Tan et al., 1999). Using a toxR mutant, we confirmed that ETA from P. aeruginosa PAO1 is at least partially involved in the effect of CNP on virulence in this strain, indicating that, in PA14 as in PAO1, ETA supports the killing of C. elegans. In contrast, elastase and protease appear to have no role in the
pro-virulence effect of CNP on PAO1 in the C. elegans model.

Whereas only CNP actually enhances PAO1 virulence, we noticed that BNP is also able to induce ETA and HSL production by the bacteria. Three subtypes of natriuretic peptide receptors (NPRs) are expressed on eukaryotic cell membranes. Of these receptors, two (NPR-B and NPR-C) are preferentially activated by CNP rather than BNP (Potter et al., 2006). Our results suggest that the PAO1 natriuretic peptide sensor could be related to one of these two NPR subtypes. Nevertheless, these NPRs can also be activated, with a lower efficiency, by BNP, and this difference could explain how, in our study, the effect of BNP on PAO1 toxin production is limited compared with CNP. Furthermore, we cannot rule out the presence of different subtypes of Pseudomonas natriuretic peptide sensors, as observed in eukaryotic cells (Anand-Srivastava et al., 1996; Potter & Hunter, 2001). Indeed, we observed previously that the sensitivity to natriuretic peptides diverges between Pseudomonas species, for instance between P. fluorescens and P. aeruginosa (Veron et al., 2008). Consistent with this notion of high stereospecific recognition of eukaryotic messengers in bacteria, it has been shown that Helicobacter pylori can recognize only one of the subtypes of the somatostatin receptor agonists (Yamashita et al., 1998).

In order to go further into the mechanism of action of natriuretic peptides in P. aeruginosa we decided to investigate the transcriptional regulator(s) potentially involved in the effect of CNP. In P. aeruginosa, numerous proteins such as Vfr, GacA, MvaT, QscR, PtxR, RsaL, VqsM and VqsR regulate (positively or negatively) the expression of QS genes (Albus et al., 1997; Chugani et al., 2001; Diggle et al., 2002; Dong et al., 2005; Juhas et al., 2004; Reimann et al., 1997; Schuster & Greenberg, 2006). ETA synthesis itself is controlled both by a cascade of positive regulators, including PvdS and RegA (ToxR) protein (Hamood et al., 2004; Ochsner et al., 1996) or the PtxR regulator (Hamood et al., 1996), and also by direct binding of the PAO1 global virulence regulator Vfr on the toxA promoter region (Davinic et al., 2009). We have demonstrated previously that the Vfr protein is involved in the effect of natriuretic peptides on P. aeruginosa PAO1 virulence (Veron et al., 2007). We observed here that Vfr is not only essential for expression of the lethal effect of PAO1 on C. elegans in both fast and slow killing conditions but also involved in the pro-virulent effect of BNP and CNP. Since the PtxR regulator, which regulates ETA production in PAO1 (Hamood et al., 1996), is under the control of Vfr (Ferrell et al., 2008), and ETA synthesis is promoted by CNP (and BNP) we naturally examined the role of PtxR in CNP action. We observed that CNP induced a 3.5-fold increase of ptxR mRNA synthesis 1 h after its contact with PAO1, suggesting that PtxR is involved in the effect of CNP on PAO1. Moreover, the observation that PtxR negatively regulates the expression of pyocyanin while it positively regulates the production of 3OC12-HSL through lasI (Carty et al., 2006) strongly reinforces our hypothesis that PtxR is a member of the transduction cascade triggered by CNP in PAO1 (Fig. 10).

However, PtxR appears to regulate only a part of the effect of CNP, since a ptxR mutation does not lead to a total suppression of bacterial virulence as observed with the vfr mutant, and whereas PtxR is necessary for the expression of the slow-killing activity of PAO1, this protein is not required to kill C. elegans in the fast-killing test (data not shown). We know that Vfr is involved in LPS structural rearrangements induced by CNP (Veron et al., 2007) and an algC mutant is found to be avirulent, an effect which has been specifically ascribed to LPS biosynthesis modifications (Goldberg et al.,...
It is interesting to note that although CNP interferes with iron uptake and in return reduces infection and aids in the clearance of this pathogen. Our study suggests that chemicals able to block or reverse the effect of natriuretic peptides on bacteria may serve as a basis for the development of compounds inhibiting the global virulence of *P. aeruginosa*. These data, added to those of Zaborina et al. (2007) on dynorphin or Hegde et al. (2009) on norepinephrine, show that QS is involved in the bacterial response to eukaryotic communication molecules and suggest that inter-kingdom signal exchanges are certainly essential in the pathogenic activity of bacteria.

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**Fig. 10.** Schematic model representing the mechanism of action of natriuretic peptides on *P. aeruginosa* PAO1. Activation of the membrane natriuretic peptides sensor (NPS) by CNP induces a rise in intra-bacterial cAMP concentration through adenylate cyclase (AC) activation (Veron et al., 2007). Then, the global regulator Vfr, activated by cAMP, regulates directly or indirectly (after PtxR activation) the expression of several virulence factors such as ETA, HCN and pyocyanin through the modulation of QS signalling molecules. Finally, CNP binding on *P. aeruginosa* PAO1 increases global bacteria virulence.
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