Epigenetic control of IL-23 expression in keratinocytes is important for chronic skin inflammation

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The chronic skin inflammation psoriasis is crucially dependent on the IL-23/IL-17 cytokine axis. Although IL-23 is expressed by psoriatic keratinocytes and immune cells, only the immune cell-derived IL-23 is believed to be disease relevant. Here we use a genetic mouse model to show that keratinocyte-produced IL-23 is sufficient to cause a chronic skin inflammation with an IL-17 profile. Furthermore, we reveal a cell-autonomous nuclear function for the actin polymerizing molecule N-WASP, which controls IL-23 expression in keratinocytes by regulating the degradation of the histone methyltransferases G9a and GLP, and H3K9 dimethylation of the IL-23 promoter. This mechanism mediates the induction of IL-23 by TNF, a known inducer of IL-23 in psoriasis. Finally, in keratinocytes of psoriatic lesions a decrease in H3K9 dimethylation correlates with increased IL-23 expression, suggesting relevance for disease. Taken together, our data describe a molecular pathway where epigenetic regulation of keratinocytes can contribute to chronic skin inflammation.
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berrant cytokine expression is believed to be crucial for the development of inflammatory skin diseases such as psoriasis. Although the cause of psoriasis is unknown, development and maintenance of the disease occurs through the crosstalk between immune cells and keratinocytes. In particular, the interleukin (IL)-23/IL-17 axis and the tumor necrosis factor (TNF) pathway are of central importance in psoriasis as demonstrated by successful therapeutic intervention against these cytokines. IL-23 is moreover increased in the skin of patients suffering from atopic dermatitis (AD) or alopecia areata and in the serum of patients with autoimmune diseases as systemic lupus erythematosus (SLE) or Crohn’s disease.

IL-23 is required for the development and expansion of IL-17-producing immune cells, but the mechanism underlying the increased expression of IL-23 is less clear. Expression of IL-23 is regulated by nuclear factor (NF)-κB signaling, which is activated by many cytokines including TNF. TNF and IL-23 are produced by activated innate immune cells, but also by psoriatic keratinocytes, which may contribute to the development of the disease. We now provide evidence that an epigenetic mechanism involving TNF and the neural Wiskott-Aldrich syndrome protein (N-WASP) controls IL-23 expression in keratinocytes.

N-WASP is a ubiquitously expressed member of the WASP/Scar family, which promotes actin polymerization via the Arp2/3 complex. In vitro data indicate a role for N-WASP in the formation of filopodia, the assembly of intercellular junctions, and clathrin-mediated endocytosis, which may contribute to the development of the disease. In addition to these cytoplasmic functions, N-WASP might also have a nuclear role, since it was reported to shuttle between the nucleus and the cytoplasm and to bind to a nuclear complex containing actin and RNA polymerase II.

Genetic ablation of the N-WASP gene leads to early embryonic lethality in mice; however, filopodia formation is intact. Keratinocyte-restricted deletion of the N-WASP gene revealed an important function for N-WASP in the control of the hair cycle, corresponding to increased transforming growth factor-β (TGFβ) signaling and reduction of hair follicle stem cells.

Closer investigation of the interaction between N-WASP function in keratinocytes and the immune system revealed now that loss of N-WASP in keratinocytes provokes IL-23 expression in keratinocytes and a chronic skin inflammation with an IL-17 profile. Furthermore, we observed increased amounts of autoimmune antibodies in the serum. In vitro studies revealed that both TNF and N-WASP are regulating IL-23 expression by controlling H3K9 methylation via modulating protein degradation of the H3K9 methyltransferases G9a and GLP. These data suggest that keratinocytes might contribute to initiation or progression of inflammatory skin diseases such as psoriasis by producing IL-23 and reveal an unexpected nuclear function of N-WASP in epigenetic repression of IL-23, which is regulated by TNF.

Results

N-WASP ko in keratinocytes causes chronic skin inflammation. Mice with a keratinocyte-restricted deletion of the N-WASP gene (N-WASP fl/fl K5 cre; called ko) show increased TGFβ signaling in the epidermis, suggesting the possibility of a local inflammatory reaction. To investigate in more detail a potential inflammatory response in N-WASP ko mice, we performed full necropsies of 8–26-week-old control and N-WASP ko mice and analyzed tissue sections by hematoxylin–eosin (H&E) staining. Skin sections from back, tail, eye lid ear, nose, and facial skin of ko mice of all ages indicated slight to moderate hyperplasia of the epidermis and increased cellularity of the dermis consisting of mast cells, granulocytes, and other mostly non-lymphoid cells (Fig. 1a). Staining of back skin for mast cells confirmed increased number of mast cells in N-WASP ko mice (Fig. 1g). This phenotype was particularly prominent in mechanically stressed tissues such as tail, lip, and ear, suggesting that N-WASP-null skin is oversensitive to mechanical stress. Macroscopically, the tails of older N-WASP mutant mice appeared often scaly (Fig. 1b) and mechanically stressed tissues of older mutant mice showed occasionally focal ulcerations.

Testing proliferation of interfollicular keratinocytes in N-WASP ko mice in an earlier study with only few mice, we had observed a high variation in proliferation and could not detect a significant change. Repeating the proliferation measurement by testing for Ki67+ cells using more mice and more samples, we now show that proliferation in N-WASP ko epidermis is indeed normal in 6- and 9-day-old mice, but increased at 3 and 7 weeks compared to littermate controls (Fig. 1d). These data indicate that N-WASP ko mice develop a skin hyperplasia after birth.

K6, which is often upregulated in hyperproliferation, was not detectable in interfollicular keratinocytes of 6-day-old N-WASP ko mice, confirming the absence of hyperplasia in very young mice. However, in 9-day-old N-WASP-ko mice, rare, focal upregulation of K6 expression was observed in the interfollicular epidermis, which then became more widespread in older mice (Fig. 1c). In our earlier analysis, no interfollicular K6 was detectable in 9-day-old N-WASP ko mice. The reason for this difference is not clear, but might be influenced by differences in the genetic background of the outbred mice used for the analysis and by a different animal house environment.

Next, we analyzed whether the increased cellularity of the dermis in ko mice is due to an increased number of immune cells by staining for the pan-leukocyte marker CD45 (Ly-5.2). While we could see no difference in the frequency of Ly-5.2+ cells in skin of 6-day-old control and ko mice, N-WASP ko skin showed increased numbers of CD45+ cells at 9 days and later indicating the presence of an leukocyte infiltrate (Fig. 1e). Fluorescence-activated cell sorting (FACS) analysis confirmed an increased number of immune cells infiltrating epidermis of N-WASP ko mice at 2 and 4 weeks (Fig. 1f).

To test whether the activation of the immune system also includes the adaptive immune response and the production of antibodies, we determined the deposition of immunoglobulin G (IgG) in the skin of control and ko mice at different ages by immunofluorescent staining and western blot. The 7-week-old N-WASP ko mice showed strongly elevated levels of IgG by western blot of skin lysates and immunofluorescent staining of back skin sections (Fig. 2a, b). In older mice, IgG deposits were also found in the kidney (Fig. 2b). No increased IgG was detectable in the skin of 3-week-old ko mice (Fig. 2a).

Next, we investigated whether inflammatory changes could also be observed in other tissues than skin. Skin-draining inguinal lymph nodes were increased in size in 8-week-old N-WASP ko mice and showed significantly increased cellularity (Fig. 2c). Furthermore, about 20% of the ko mice older than 4 months developed a mostly unilateral kidney hydronephrosis, corresponding to an inflammation of the ureter, which might lead to obstructions preventing the transport of urine from the kidney to the bladder (Fig. 2d, e). Since K5 is also expressed in ureter epithelium, K5 cre-driven deletion of the N-WASP gene might also take place in ureter epithelium and mediate inflammation.

Finally, we screened serum samples (1:100) for antibodies against mouse keratinocytes. Of the 8 ko mice tested, all showed clear presence of self-reactive antibodies against nuclear (4 of 8) or cytoplasmic (4 of 8) antigens. No or clearly weaker autoreactivity was detected at this dilution in the sera of control mice (Fig. 2f). Further analysis of sera with antibodies against
Nuclear antigens revealed increased concentration of antibodies against double-stranded DNA (dsDNA) in serum samples of N-WASP ko mice (Fig. 2g).

These data indicate that loss of N-WASP in keratinocytes results in chronic skin inflammation together with hyperplasia of the epidermis, increased expression of K6, and increased proliferation. Additionally, increased amounts of self-reactive antibodies are observed in serum of aged N-WASP ko mice.

**Increased cytokine expression in N-WASP-null back skin.** Chemokines, cytokines, and growth factors are soluble mediators involved in the activation of immune cells. We therefore analyzed...
by real-time quantitative reverse transcription-polymerase chain reaction (qRT-PCR) the expression of 17 proinflammatory mediators suggested to contribute to inflammatory skin diseases such as psoriasis, atopic dermatitis, or irritant contact dermatitis in the back skin before immune cell infiltration (6 days), at the onset of immune cell infiltration (9 days), and when IgG is detectable in the skin (7 weeks). The 6-day-old ko mice showed increased expression of IL-23A, CXCL10, and CCL20 (Fig. 3a) compared to controls. At 9 days, IL-12A, IFNy, IL-4, IL-13, IL-17A, IL-17F, IL-22, CCL2, and IL-1β are also upregulated (Supplementary Figure 2a), while in 7-week-old ko mice all cytokines tested including IL-1α, IL-6, IFNα, IFNβ, and TNFα are increased
Fig. 3  Increased cytokine expression in N-WASP-null skin.  

a qRT-PCR analysis of indicated genes from back skin RNA of con and ko at indicated ages (n > 3/3, two-tailed unpaired t-test). All normalized to GAPDH.  

b Representative toluidine blue staining for barrier defect in con and ko mice at indicated ages (P: E15.5 embryos used as positive controls; 2 days (2d) n: 6/5, 6 days (6d) n: 4/5, 3 independent experiments).  

c Representative immunofluorescence staining and quantification of IL-23p19 of con and ko back skin at indicated ages (n: 3/3, two-tailed paired t-test). Scale bar: 100 µm.  

d Representative sorting strategy of CD45+, α6H, and α6L cells from con and ko epidermal cells and qRT-PCR analysis of IL-23A and IL-17A expression in sorted cells (n: 3/3, mean ± SD, two-tailed unpaired t-test, 3 independent sortings).  

e Representative immunoblots and quantification of indicated proteins from epidermal lysates of 7–8-week old con and ko mice (n: 4/4, mean ± SD, two-tailed unpaired t-test).  

f Representative immunoblots and quantification of indicated proteins of co-precipitations of IL-23p19 with IL-12p40 from epidermal lysates of 7-8-week-old con and ko mice (n: 4/4, mean ± SD, two-tailed unpaired t-test; *p ≤ 0.05; **p ≤ 0.01; ***p ≤ 0.001)
in the back skin compared to controls (Fig. 3a). IL-23 is a dimer consisting of the proteins IL-23p19 (gene: IL-23A) and IL-12p40 (gene: IL-12B). Both of these subunits are increased in N-WASP ko skin on the protein level (Fig. 3e), indicating increased expression of IL-23 in the absence of N-WASP. Co-purification of IL-23p19 with IL-12p40 demonstrated increased amounts for IL-23 heterodimers in N-WASP ko epidermis (Fig. 3f).

To investigate whether the increased cytokine expression is related to impaired barrier function and increased Toll-like receptor signaling, we performed barrier assays and tested for phosphorylation of NF-κB (pNF-κB (S536)) and pIRAK4 (T345, S346). Dye penetration assay did not indicate impaired barrier function in 2- or 6-day-old mice (Fig. 3b). Furthermore, no increased phosphorylation of NF-κB or IRAK4 could be detected in skin lysates of 6-d-old mice (Supplementary Figure 2c). These data suggest that the increased cytokine expression is not caused by a primary barrier defect. In 7-week-old N-WASP ko mice, expression of involucrin, filaggrin (50 ± 9.8-fold increase as determined by qRT-PCR, n: 2/2), cystatin A, Tgm1, and a number of small proline-rich genes and late cornified envelope genes was increased, while expression of loricrin and cornesodemin was unchanged compared to controls, suggesting maybe subtle changes of the barrier at the stage of chronic skin inflammation (Supplementary Table 1).

Immunofluorescent staining for IL-23p19 in back skin sections of 6-day and 4-week-old control and ko mice confirmed expression of IL-23p19 in the epidermis of 6-day-old ko mice in both basal and suprabasal keratinocytes with further enhanced expression in the epidermis of 4-week-old ko mice (Fig. 3c). These data indicate that increased expression of IL-23p19 is a very early event in skin inflammation in N-WASP ko mice, which precedes the expression of most other cytokines and immune cell infiltration.

To analyze the cellular source of IL-23, epidermis of 4-week-old control and ko mice was trypsinized and the cell suspension stained and sorted by fluorescent staining for IL-23p19 in back skin sections (Fig. 3a). Since IL-17A-producing cells are of critical importance in skin inflammation. However, Smad2 phosphorylation was not detected for IL-17A, IL-22, S100A8, CCL2, CCL20, CXCL10, and IFNγ in dko compared to ko, while IL-22 and IL-12B were strongly reduced. IL37, which was described to be a key factor in activating plasmacytoid dendritic cells in psoriasis21, IL-19, IL-20, and IL-24 were even increased in dko mice.

Repeating the gene expression analysis with FACS purified keratinocytes, we obtained similar results as for total skin for S100A8, S100A9, CCL2, CCL20, CXCL10, and IFNγ were not altered in dko compared to ko, while IL-12A and IL-12B were strongly reduced. IL37 was decreased in both ko and dko keratinocytes compared to controls, the changes were not significant. Low expression of these mediators indicated that they are mainly produced by immune cells. In keratinocytes, no expression was detected for IL-17A, IL-22, and IFNγ.

These data demonstrate that the skin inflammation in N-WASP ko mice is dependent on keratinocyte-produced IL-23.

**Different immune cells contribute to IL-17 in N-WASP ko mice.** Since IL-17A-producing cells are of critical importance in psoriasis and probably also in other chronic inflammatory diseases such as AD and SLE20, we wanted to confirm the production of IL-17A on the protein level and identify the cells producing this cytokine. For this reason we tested epidermal cell suspensions of 4-week-old mice by FACS for IL-17A-producing leukocytes and investigated skin-draining inguinal lymph nodes of 2- and 4-week-old control and ko mice for intracellular IL-17A and the surface markers CD45, CD3, CD4, and γδTCR. In the epidermis of 4-week-old ko mice, only a small percentage of the leukocytes produced IL-17A, about 4 times more than in controls (Fig. 4a). In skin-draining inguinal lymph nodes of 2- and 4-week-old ko mice, frequency and absolute numbers of CD4 cells and γδ T cells expressing IL-17A was strongly increased (Fig. 4b, c). No significant increase was observed in mesenteric lymph nodes of 4-week-old mice (Fig. 4g), suggesting that the IL-17A production is a consequence of the skin inflammation and not due to a systemic defect in T-cell development. These data show that different T-cell subsets contribute to the increased IL-17 levels in N-WASP ko mice.

**Skin inflammation is dependent on keratinocyte-derived IL-23.** To test whether the skin inflammation observed in N-WASP ko mice is dependent on keratinocyte-derived IL-23, we generated mice with a conditional knockout of IL-23A using CRISPR/Cas9 genome editing (Fig. 5a), and crossed them with N-WASP ko mice. The resulting mice with a keratinocyte-restricted loss of both N-WASP and IL-23A (dko) showed a dramatically reduced expression of IL-23A messenger RNA (mRNA), indicating efficient depletion of the IL-23A gene in keratinocytes (Fig. 5d). Importantly, infiltration of leukocytes, keratinocyte proliferation (Ki67+ cells), and K6 expression were strongly reduced compared to N-WASP ko mice (Fig. 5b, c). Interestingly, dko mice still showed epidermal thickening and loss of hairs. The reduced inflammation in dko mice correlated with a decreased expression of IL-17A and S100A9 in skin (Fig. 5d). Average values of IL-17F, IL-22, S100A8, CCL2, CCL20, CXCL10, and IFNγ were not altered in dko compared to ko, while IL-12A and IL-12B were strongly reduced. IL37, which was described to be a key factor in activating plasmacytoid dendritic cells in psoriasis21, IL-19, IL-20, and IL-24 were even increased in dko mice.

Primary N-WASP-null keratinocytes express IL-23A in vitro. To understand the molecular mechanism underlying the increased IL-23A expression in N-WASP-null keratinocytes in vivo, we tested whether IL-23A expression is also increased in cultured keratinocytes lacking N-WASP. We therefore isolated primary keratinocytes from 3-day-old control and ko mice,
**Fig. 4** Increased IL-17A-producing cells in N-WASP ko mice. 

a. Representative and quantification of intracellular FACS analysis of IL-17A+ cells in epidermis (n: 3/3, mean ± SD, two-tailed unpaired t-test).  

b. Representative and quantification of intracellular FACS analysis of IL-17A+ cells in iLN (n: 3/3, mean ± SD, two-tailed unpaired t-test).  

c. Representative and quantification of CD4, CD8 distribution in thymus, spleen, and bone marrow from 9-day-old con and ko mice (n: 3/3, two-tailed unpaired t-test).  

d. Representative H&E staining of thymus from 9-day-old con and ko mice (n: 3/3). Scale bar: 50 µm.  

g. Representative and quantification of intracellular FACS analysis of IL-17A+ cells in mLN (n: 6/6, mean ± SD, two-tailed unpaired t-test; *p ≤ 0.05; **p ≤ 0.01; ****p ≤ 0.0001)
cultured them for 5–7 days in vitro and then analyzed by qRT-PCR the expression of various cytokines. We observed increased expression of IL-23A, CXCL10, and CCL2, while CCL20, TNF, IL-1α, IL-1β, IL-6, S100A8, and CXCL5 were not increased (Fig. 6b). Culturing primary keratinocytes under differentiating high-calcium conditions did not change IL-23A expression in control or N-WASP ko cells (Fig. 6c).

These data suggest that N-WASP also controls expression of IL-23A in cultured neonatal keratinocytes. Analysis of gene expression by microarray analysis from primary keratinocytes isolated from 3-week-old mice did not reveal any additional cytokine genes that showed a more than 1.5-fold alteration of the average value, supporting that increased IL-23A expression by
keratinocytes is crucial for triggering the complex inflammation observed in adult N-WASP ko mice.

The establishment of an in vitro system allowed us therefore to study the molecular mechanism underlying the increased IL-23A expression N-WASP-null keratinocytes in a less complex model system.

**N-WASP regulates IL-23 expression by H3K9 dimethylation.** N-WASP is known to promote actin polymerization, which is also crucial for the N-WASP-dependent regulation of RNA polymerase II-mediated transcription and retinoic acid-induced expression of HoxB. For this reason we tested whether treatment of keratinocytes with the actin polymerization

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**Figure Legends:**

**A** Purified keratinocytes in vivo

**B** In vitro

**C** IL-23A

**D** IL-23A

**E** 3w Phalloidin DAPI

**F** Phalloidin

**G** IL-23A
inhibitor latrunculin or the Arp2/3 inhibitor CK548 increases IL-23A expression in these cells. However, both inhibitors strongly reduced IL-23A expression (Fig. 6d). In addition, we determined the amount of F-actin in primary control and N-WASP-null keratinocytes by fluorescence microscopy and FACS analysis after staining with phallolidin–fluorescein isothiocyanate (FITC). Absence of N-WASP did not reduce the total amount of F-actin in keratinocytes (Fig. 6e, f). Taken together, these data suggest that loss of N-WASP is not promoting IL-23A expression by decreased F-actin.

The N-WASP-related molecule WASP, which is only expressed in hematopoietic cells, is interacting with epigenetic enzymes leading to increased activity of RBBP5-dependent H3K4 methyltransferases and the JMJ2D2A demethylase and elevated gene expression24. To test whether N-WASP regulates IL-23A expression by altering epigenetic histone marks, we treated primary keratinocytes with different epigenetic inhibitors: BIX01294 (BIX), inhibiting the H3K9 dimethyltransferases G9a and GLP and thus decreasing the repressive H3K9me2, HC (tranylcypromine–hydrochloride), which blocks the lysine demethylase LSD1 and increases particularly H3K4 demethylation, IOX-1, which is an inhibitor of several JmjC-domain containing histone demethylases, and trichostatin A, a histone deacetylase inhibitor, which increases histone acetylation.

The H3K9 methyltransferase inhibitor BIX induced IL-23A expression in control keratinocytes, while it had only a relatively small effect on N-WASP-deficient cells (Fig. 6g; right panel shows fold change relative to DMSO). One explanation for this low effect of BIX on N-WASP-null cells could be that H3K9 methylation is already decreased in untreated N-WASP-deficient keratinocytes. Indeed, western blot analysis revealed that N-WASP ko keratinocytes display 50% reduced H3K9me2 levels than control cells (Fig. 7a) Treatment with BIX reduced H3K9me2 levels in controls by about 2-fold, while it had no significant influence on ko cells.

A crucial role for H3K9me2 in the regulation of IL-23A expression in keratinocytes was supported by the result that IOX-1 treatment, which increases H3K9me2 (Fig. 7b), strongly reduced IL-23A expression in both control and N-WASP ko keratinocytes (Fig. 6g). Interestingly, IOX-1-treated N-WASP ko cells showed H3K9me2 and IL-23A levels similar to untreated control cells.

Also, in vivo, H3K9me2 was reduced in N-WASP ko keratinocytes as shown by western blot analysis of epidermal lysates of 6-day-old control and mutant mice and by immunofluorescent staining (Fig. 7c, d).

To interfere with H3K9 methylation by a different method and to test the role of this modification for the regulation of IL-23A expression in vivo, we analyzed IL-23 p19 expression in FACS sorted, interfollicular epidermal keratinocyte stem cells of mice with a keratinocyte-restricted deletion of the G9a gene. G9a ko keratinocytes showed increased expression of IL-23A to a similar extent in N-WASP ko cells (Fig. 7e).

To assess whether reduced H3K9 dimethylation is directly affecting the IL-23A promoter or only indirectly controlling IL-23A expression by regulating expression of transcription factors, a chromatin Immunoprecipitation (ChIP) analysis for different regions of the IL-23A promoter was performed. Indeed, we could identify an 0.3 kb region about 0.5 kb upstream of the transcription start site of IL-23A which showed significantly reduced H3K9me2 in the absence of N-WASP (Fig. 7f, “B”). This promoter region contains binding sites for IRF3, Smad3, and ATF-2, which were found previously to be critical for IL-23A expression25,26. Other IL-23A promoter regions tested showed no detectable alteration in H3K9 dimethylation (Fig. 4j; “A”, “C”).

Deleting the N-WASP gene in vitro by Cre expression in Il1b primary keratinocytes decreased protein amounts of G9a and H3K9me2 and increased mRNA expression of IL-23A, confirming that this mechanism is cell autonomous (Fig. 7g, h).

These findings indicate that N-WASP, H3K9me2, and the H3K9 methyltransferases G9a and GLP are crucially regulating IL-23A expression in keratinocytes in vitro and in vivo.

**N-WASP regulates protein stability of G9a and GLP.** Since loss of N-WASP had a similar effect on IL-23A expression as inhibition of the H3K9 dimethylation transfers G9a and GLP, N-WASP might regulate expression, degradation, or functional activity of G9a and GLP. Gene expression analysis of N-WASP-null epidermis and N-WASP-null keratinocytes by microarray, however, did not indicate decreased mRNA level of G9a and GLP. qRT-PCR of epidermal and primary keratinocyte RNA for G9a and GLP confirmed that the mRNA levels are not significantly altered in vivo and in vitro (Fig. 8a, b). This was confirmed by analysis of the protein degradation kinetic in the presence of cycloheximide, which revealed an accelerated degradation of G9a and GLP in the absence of N-WASP (Fig. 8e).

**N-WASP is associated with chromatin-bound G9a and GLP.** Based on these results we tried to rescue the H3K9 methylation defect by treating N-WASP-null keratinocytes with the proteasome inhibitor MG-132. In addition, we assessed whether loss of N-WASP affects chromatin binding of G9a and GLP, hypothesizing that methyltransferases not attached to chromatin have a reduced half-life. To this end, keratinocytes were fractionated into a soluble cytoplasmic (C; marker: glyceraldehyde 3-phosphate dehydrogenase (GAPDH)), a soluble nuclear (SN; marker: proliferating cell nuclear antigen (PCNA)), and a pellet fraction containing chromatin (P; marker: H3). N-WASP protein was found in the cytoplasmic (65% ± 4%; n: 3) and in the chromatin

Fig. 6 IL-23A expression is increased in N-WASP-null keratinocytes. a qRT-PCR analysis of indicated genes in FACS purified primary keratinocytes from 6-week-old mice (n: 4/4/4, mean ± SD, two-tailed unpaired t-test, 2 independent sorting). b qRT-PCR analysis of indicated cytokine mRNA expression in primary keratinocytes. All normalized to GAPDH (n: 3/3, mean ± SD, two-tailed unpaired t-test). c Primary keratinocytes were differentiated with 2 mM CaCl2 (High Ca2+) for 48 h and then analyzed by qRT-PCR for IL-23A expression, compared to untreated cells (Low Ca2+): n: 3/3, mean ± SD, two-tailed unpaired t-test. d qRT-PCR analysis of IL-23A mRNA expression in primary keratinocytes in response to 24 h treatment with indicated inhibitors. All normalized to GAPDH (n: 3/3, mean ± SD, two-tailed unpaired t-test). e Representative immunofluorescence staining of phallolidin in con and ko primary keratinocyte (n: 3/3, mean ± SD, two-tailed unpaired t-test). f qRT-PCR analysis of IL-23A mRNA expression in primary keratinocytes treated with indicated inhibitors for 24 h. All normalized to GAPDH. The right panel shows the same data, but normalized to the respective DMSO-treated samples, thus showing the relative effect of the treatment as fold change (n: 3/3, mean ± SD, two-tailed unpaired t-test; p ≤ 0.05; **p ≤ 0.01; ***p ≤ 0.001; ****p ≤ 0.0001).
fraction (33% ± 2%), while the soluble nuclear fraction contained only low amounts (2% ± 0.5%) (Fig. 8f).

Treatment of control and ko cells with MG-132 increased the apparent molecular weight of G9a and GLP, which might reflect ubiquitination of the proteins (Fig. 8f). In addition, protein amounts of G9a and GLP in N-WASP ko keratinocytes treated with MG-132 increased, corresponding to normal levels of H3K9me2. Moreover, MG-132 also increased the protein level of N-WASP in the soluble nuclear and the chromatin fraction, but not in the cytosol, indicating that nuclear N-WASP is more prone to proteasomal degradation (Fig. 8f). G9a and GLP were nearly exclusively found in the chromatin pellet fraction and loss
Fig. 8 N-WASP co-precipitates with and regulates the stability of the H3K9 methyltransferases G9a and GLP. Expression of G9a and GLP mRNA (a, b) and protein (c, d) in epidermis of 6-day-old mice (a, c) and in primary keratinocytes (b, d). (n (a, b, c): 3/3; n (d): 5/5). e Immunoblot analysis of protein stability of G9a and GLP in primary keratinocytes in the presence of cycloheximide (G9a n: 4/4, GLP n: 6/6, mean ± SD, two-tailed unpaired t-test). f Cell fractionation of keratinocytes after 90 min of treatment with the proteasome inhibitor MG-132 followed by immunoblot for indicated proteins, the indicated protein levels were quantified on the right (n: 3/3, mean ± SD, two-tailed unpaired t-test). g Cell fractionation of primary keratinocytes with treatment of the pellet fraction with micrococcal nuclease followed by immunoblot for indicated proteins (MNase) (n: 3). h Co-precipitation of N-WASP with G9a and GLP in primary keratinocyte nuclear lysates (n: 3). C: cytosolic fraction, SN: soluble nuclear fraction, P: pellet (*p ≤ 0.05; **p ≤ 0.01; ***p ≤ 0.001)
of N-WASP did not increase G9a and GLP levels detected in the soluble nuclear fraction, indicating that N-WASP is not required for the chromatin association of the methyltransferases. To confirm that N-WASP is indeed bound to chromatin and not to insoluble cytoskeleton, we repeated the fractionation, but digested chromatin with micrococcal nuclease (Fig. 8g). This treatment increased the amount of N-WASP, G9a, GLP, H3, and PCNA in the soluble nuclear fraction, indicating that N-WASP is strongly associated with chromatin, similar to G9a and GLP. We then tested whether N-WASP is forming a complex with G9a and GLP. Indeed, N-WASP co-immunoprecipitated with both H3K9 dimethyltransferases (Fig. 8h). Reciprocally, G9a immunoprecipitated together with GLP and N-WASP. Control precipitation with IgG did not result in detectable amounts of G9a, GLP, or N-WASP. These data suggest the possibility that direct or indirect association of N-WASP with chromatin-bound G9a-GLP heterodimers is preventing the degradation of the methyltransferases.

**TNF regulates IL-23 expression via H3K9 methylation.** In psoriasis, TNF is an important activator of IL-23 expression. We therefore speculated that TNF is able to induce IL-23A expression in keratinocytes and that this is mediated by N-WASP-dependent changes in H3K9 dimethylation (H3K9me2). To test this hypothesis, we stimulated both control and N-WASP-null keratinocytes with TNF and checked IL-23A expression after 4 h by qRT-PCR. While TNF treatment of control cells increased IL-23A expression to a level comparable to N-WASP-deficient keratinocytes, it only slightly increased expression in N-WASP-null cells (Fig. 9a). Normalizing the values for TNF-treated cells by the respective values for untreated cells showed that N-WASP-null keratinocytes are hardly responding to TNF with respect to IL-23A expression, suggesting that TNF and N-WASP regulate IL-23A expression by the same pathway.

To examine whether this effect is caused by an increased TNF signaling in N-WASP-null cells, we tested surface levels of TNF-R1 and activation of several TNF effectors in N-WASP-null and control keratinocytes. However, FACS analysis did not reveal a significant change of TNF-R1 at the cell membrane (Fig. 9b). Furthermore, TNF induced phosphorylation of the TNF effector molecules p38, c-Jun N-terminal kinase (JNK), and NF-κB was unchanged in N-WASP ko cells compared to control keratinocytes (Fig. 9c).

We then checked how treatment with TNF is affecting protein levels of G9a and GLP and H3K9 dimethylation in control and N-WASP-null keratinocytes. In control keratinocytes, TNF resulted in a dose-dependent decrease of G9a, GLP, and H3K9me2 levels already after 10 min (Fig. 9c). H3K9me2 slowly increased thereafter, while G9a and GLP remained also after 4 h at a low level. N-WASP-null keratinocytes displayed low amounts of G9a, GLP, and H3K9 dimethylation, which did not decrease further after treatment with TNF, further supporting that TNF and N-WASP work along the same pathway (Fig. 9c).

**TNF induces phosphorylation of N-WASP.** If N-WASP is acting downstream of TNF, how is TNF signaling modifying nuclear N-WASP function? TNF activates different serine/threonine kinases as for example PKCα or IKKα. N-WASP, on the other hand, was shown to be phosphorylated by casein kinase II at serine 480/481, which decreases N-WASP-dependent actin polymerization. Therefore, we speculated that TNF-induced serine kinase activation might lead to phosphorylation of N-WASP at serine 480/481 affecting the nuclear function of N-WASP. To investigate this, we fractionated keratinocytes 10 min after treatment with TNF. The chromatin fraction was extracted with a 300 mM NaCl buffer to elute weakly chromatin-associated proteins. Treatment with TNF resulted in a shift of nuclear N-WASP from the strongly chromatin-bound fraction (Fig. 9d; P300) to the weakly chromatin-bound fraction (S300). Interestingly, within the time frame of the experiment, no significant increase of N-WASP was observed in the cytoplasmic fraction (C). Clearly, TNF stimulation reduced chromatin association of N-WASP.

Moreover, TNF increased phosphorylation of N-WASP at serine 480/481 (Fig. 6d). Phosphorylated N-WASP was found in the cytoplasmic (C) and the weakly associated chromatin fraction (S300), but not in the strongly associated nuclear fraction (P300). These data suggest that phosphorylation of N-WASP at serine 480/481 is incompatible with strong chromatin binding. Since our previous experiments demonstrated an increased proteolytic degradation of G9a-GLP heterodimers in the absence of N-WASP, the decreased chromatin attachment of phosphorylated N-WASP might be associated with a dissociation of N-WASP from the G9a-GLP heterodimers, resulting in increased degradation of the H3K9 methyltransferases.

Indeed, in a ChIP experiment, TNF treatment for 10 min resulted in a decreased H3K9me2 of the IL-23A promoter in the same region (B) as loss of N-WASP decreased H3K9me2 (Fig. 9e). IOX-1 treatment of N-WASP ko keratinocytes rescued H3K9me2 and IL-23A expression (Figs. 6g and 7b), but it did not restore the ability of TNF to increase IL-23A expression (Fig. 9f). This inability corresponds to the unchanged levels of G9a, GLP, and H3K9me2 in TNF-treated N-WASP ko keratinocytes, suggesting that the N-WASP/G9a/GLP/H3K9me2 pathway is crucial for the stimulation of IL-23A expression by TNF. Collectively, these data indicate that regulation of H3K9me2 by N-WASP is crucial for TNF-induced IL-23A expression in keratinocytes in mice.

**H3K9me2 is regulating IL-23 expression in human keratinocytes.** Since our previous data indicated an important role for H3K9 methylation for the regulation of IL-23A expression in mouse keratinocytes, we wanted to test whether this mechanism is also relevant in human keratinocytes and in psoriasis, an IL-23-dependent chronic skin inflammation.

Treatment of primary human keratinocytes for 30 min with TNF significantly decreased G9a and H3K9me2 levels, while it increased NF-κB phosphorylation (Supplementary Figure 3e). Treatment with the G9a/GLP inhibitor BIX for 24 h decreased H3K9me2 levels and increased IL-23A expression in a concentration-dependent manner (Supplementary Figure 3a). On the other hand, H3K9me2 levels were increased and IL-23A mRNA levels decreased by incubation for 24 h with the histone-demethylase inhibitor IOX-1 (Supplementary Figure 3c). Importantly, TNF-induced IL-23A expression was completely blocked by pretreatment for 24 h with IOX-1, while BIX pretreatment increased TNF-induced IL-23A expression (Supplementary Figure 3b, d).

These data demonstrate that in human keratinocytes H3K9me2 levels are also crucially important for regulating basal and TNF-induced IL-23A expression.

Furthermore, we stained three sections of lesional and non-lesional skin from three psoriasis patients for H3K9me2. Compared to skin from healthy donors, skin from psoriasis patients showed increased proliferation (Ki67; Fig. 10a). While non-lesional epidermis showed a relatively uniform nuclear staining of H3K9me2, H3K9me2 was decreased by about 50% in psoriatic lesions, particularly in the suprabasal region (Fig. 10a, c). Interestingly, skin of healthy controls stained in parallel presented two-fold higher amounts of H3K9me2 than non-lesional skin of psoriasis patients. Co-staining for IL-23p19 revealed an inverse correlation of H3K9me2 and IL-23 as
expression of IL-23p19 was increased in H3K9me2 low suprabasal regions of psoriatic lesions (Fig. 10b, c; Supplementary Figure 2b).

These correlative findings support a role of H3K9me2 in the regulation of IL-23 expression in keratinocytes in psoriasis.
Discussion

The TNF/IL-23/IL-17 cytokine cascade is suggested to be important for different inflammatory diseases. In psoriasis, inhibitors against these cytokines are already used in the therapy of this chronic skin inflammation. Here, we show in a mouse model that keratinocytes can produce IL-23 at levels sufficient to cause differentiation of IL-17A-producing T cells and skin inflammation. We furthermore describe a TNF/N-WASP/G9a, GLP/H3K9me2 pathway that crucially regulates IL-23 expression in human and mouse keratinocytes (Supplementary Figure 3F). Since DNA damage response leads to decreased G9a levels, and polyinosinic-polycytidylic acid-induced Toll-like receptor signaling decreases H3K9 methylation and increases IL-23 expression in dendritic cells, other signaling pathways might also converge on this epigenetic control of IL-23 production in keratinocytes.

As IL-23 is conceived to be also important in other inflammatory diseases such as AD or SLE, epigenetic regulation of IL-23 expression in non-immune cells could also play a role in these illnesses. Indeed, antibodies against dsDNA observed in the serum of N-WASP ko mice are well known in SLE, but not in psoriasis.

In conclusion, we report here a nuclear mechanism for TNF/N-WASP-dependent epigenetic regulation of IL-23 that could explain how environmental stress affects IL-23 expression, thus contributing to chronic inflammatory diseases dependent on the IL-23/IL-17 axis.

For Supplementary Discussion see Supplementary Information.

Methods

Mice. Mice with keratinocyte-restricted deletion of the N-WASP gene (N-WASP/fl/fl K5 cre) were 129sv/C56Bl6 outbred. The control mice used in all experiments are either N-WASP/fl/fl or IL-17/K5 cre (heterozygous ko), as we did not observe a difference between these two genotypes. In all experiments, age- and gender-matched litter mates were used as controls.

Mice with keratinocyte-restricted deletion of the IL23A gene were produced by CRISPR-Cas9-mediated insertion of loxP sites upstream of exon 2 and downstream of exon 4. Two single-guide RNAs (sgRNAs) were designed to target intron 1 and 4, respectively. Insertion of loxP sites was facilitated by 60 bp homologous arms identical to the genomic sequences flanking the cut sites (for sgRNA primers and homologous arms see Supplementary Table 2). The resulting mutant mice were than mated with N-WASP/fl/fl K5 mice to generate mice with keratinocytes-restricted deletion of both N-WASP and IL-23a genes (dko).

Mice with keratinocyte-restricted deletion of the G9a gene were generated by crossing G9a/fl/fl mice with K14-Cre mice (purchased from The Jackson Laboratory, USA), resulting in G9a conditional KO mice.

All mice were kept in an Aaalaac (Association for Assessment and Accreditation of Laboratory Animal Care)-accredited animal house under specific pathogen-free conditions. Licenses for breeding and experimentation were obtained from the Danish Administration for Animal Experiments.

Barrier assay. To perform barrier assay, mouse E15.5 embryos, 2- and 6-day pups were killed and rinsed in phosphate-buffered saline (PBS), and then immersed in 25, 50, 75, 100, 75, 50, and 25% ice-cold methanol for 2 min each. Subsequently, embryos and pups were immersed in 0.1% toluidine blue for 10 min on ice, afterwards washed in PBS and photographed.

Cell isolation, culture and in vitro knockout. Primary keratinocytes were isolated from skin adult mice by digestion with 0.8% trypsin in PBS for 30 min at 37 °C. Keratinocytes were cultured in minimum essential medium Eagle with 5 mg l−1 insulin (Sigma #H5500), 10 µg l−1 EGF (Sigma #E9644), 10 mg l−1 transferrin (Sigma #T8158), 10 µM phosphoethanolamine (Sigma #P0503), 10 µM ethanolamine (Sigma #E0135), 0.36 µg l−1 hydrocortisone (Calbiochem #386698), 1x GlutaMAX (Invitrogen #35050), 1x penicillin/streptomycin (Invitrogen #15140), and 40 ml of chelated fetal calf serum. Normal human primary keratinocytes were cultured in Lonza KGM-2 BulletKit (CC-3107, Lonza, Swiss).

To introduce in vitro recombination, HEK293T cells (ATCC CRL-3126) were transfected with pRRl-GFP or pRRl-Cre-GFP plasmid together with psVSGv. 48 h later, HEK293T were cultured in keratinocytes medium for 48 h to collect virus. Primary keratinocytes with fl/fl genotype were transduced with either green fluorescent protein (GFP) or Cre-GFP transducing virus for 48 h and analyzed after 5–7 days in culture.

Inhibitor experiments. For inhibitor experiments primary keratinocytes isolated from con and ko mice were treated for 24 h with 2 µM BIX01294 (3364, Tocris UK), 2 µM HC (Translycromyine hydrochloride; 3852, Tocris UK), 200 µM iox (4464, Tocris UK), 0.1 µM TSA (1406, Tocris UK), 50 µM CK548 (C7499, Sigma, USA), 0.5 µM Latrunculin A (L5163, Sigma, USA), 10 µg MG-132 (1748, Tocris, UK), or DMSO (Sigma, USA). For TNFa stimulation, keratinocytes were treated with 10 ng ml−1 mouse TNFa (GF027, Millipore, USA) or as indicated in the legend for indicated times. For TNFa and iox-1 experiment, keratinocytes were treated with 200 µM iox for 24 h and then followed by 4 h of TNFa stimulation. To prevent protein synthesis, primary keratinocytes were treated with 20 µg ml−1 cycloheximide (46401, Sigma, USA) for indicated times. Normal human primary keratinocytes were treated as indicated in the legend.

Subcellular fractionation. Fractionation of primary keratinocytes was done as previously described. Briefly, keratinocytes were collected by scraping, resuspended in buffer A (10 mM HEPES, pH 7.9, 10 mM KCl, 1.5 mM MgCl2, 0.34 M sucrose, 10% glycerol, 1 mM dithiothreitol (DTT), 0.1% Triton X-100, protease and phosphatase inhibitors) for 10 min on ice, and centrifuged for 5 min at 1300 × g. The supernatant was cleared by centrifuging and collected as cytosol fraction (C). The pellet was the resuspended in buffer B (3 mM EDTA, 0.2 mM EGTA, 1 mM DTT, protease and phosphatase inhibitors) on ice for 30 min, and centrifuged for 5 min at 1800 × g. The supernatant was collect as soluble nuclear fraction (S), and the pellet was directly lysed as chromatin-enriched fraction (P). Alternatively, the pellet fraction was further extracted with buffer C (20 mM HEPES-KOH, 1.5 mM MgCl2, 300 mM NaCl, 1 mM DTT, 0.2 mM EDTA, 20% (V/V) glycerol, pH 7.9) instead of buffer B for 30 min, and then centrifuged for 5 min at 1800 × g. The supernatant was collected as loose chromatin binding fraction (S300) and pellet as strong chromatin binding fraction (P300).

For micrococcal nuclease digestion, the pellet fraction was resuspended in buffer A with addition of 1 mM CaCl2 and 2 µM Mn2+ (MNuclease (N5386, Sigma, USA) for 2 min at 37 °C. The sample was then centrifuged for 5 min at 1800 × g. The supernatant and pellet were collected and analyzed.

Flow cytometry. Preparative and analytical FACS was carried out according to standard protocols. For preparative FACS sorting, single-cell suspensions were prepared from epidermis, blocked by rat anti-CD16/32 (14-0161, eBiosciences, USA, 1:100 dilution) for 10 min on ice, and then stained with CD49f-FITC (555735, BD Pharmingen, USA, 1:100 dilution) for 10 min on ice, and then stained with CD49f-FITC (555735, BD Pharmingen, USA, 1:100 dilution). For inhibitor experiments primary keratinocytes isolated from skin adult mice by digestion with 0.8% trypsin in PBS for 30 min at 37 °C. Keratinocytes were cultured in minimum essential medium Eagle with 5 mg l−1 insulin (Sigma #H5500), 10 µg l−1 EGF (Sigma #E9644), 10 mg l−1 transferrin (Sigma #T8158), 10 µM phosphoethanolamine (Sigma #P0503), 10 µM ethanolamine (Sigma #E0135), 0.36 µg l−1 hydrocortisone (Calbiochem #386698), 1x GlutaMAX (Invitrogen #35050), 1x penicillin/streptomycin (Invitrogen #15140), and 40 ml of chelated fetal calf serum. Normal human primary keratinocytes were cultured in Lonza KGM-2 BulletKit (CC-3107, Lonza, Swiss).
cell strainer (40 μm). To promote IL-17 expression and prevent secretion, cells were first stimulated for 6 h in RPMI containing cell stimulation cocktail plus protein transport inhibitors (00-4975-93, eBioscience, USA) at 37 °C. The cell suspension was then stained for surface markers for 20 min on ice in the dark, fixed with 4% paraformaldehyde for 10 min at room temperature, permeabilized with 0.1% Triton X-100 on ice for 2 min, blocked with blocking buffer on ice for 10 min and stained for intracellular markers. Data were collected by FACS Calibur (BD Biosciences, USA) using the CellQuestPro software and analyzed by Flowjo 8.8.2. The following antibodies were used: CD45.2-FITC (11-0454, 1:100 dilution), CD4-FITC (11-0041, 1:100 dilution), TCRγ–FITC (11-5711, 1:100 dilution), IL-17a-APC (17-7177-81, all eBioscience, USA, 1:100 dilution), TNFRI-PE (113003, 1:100 dilution), and CD3ε-CY5.5 (100328, 1:100 dilution, all BioLegend, USA).

ChIP. Chromatin Immunoprecipitation was done essentially as described in ref. [35]. Primary keratinocytes isolated from con and ko mice skin were grown to an approximate final count of 5 × 10⁶–1 × 10⁷ cells for analysis. Cells were chemically crosslinked with 1% formaldehyde solution for 10 min at room temperature with gentle agitation and then quenched with 0.125 M glycine. Cells were rinsed three times with 1× PBS. Cells were resuspended, lysed, and sonicated to solubilize and shear crosslinked DNA. Sonication buffer (50 mM HEPES pH 7.5, 140 mM NaCl, 1 mM EDTA, 1% Triton X-100, 0.1% Na-deoxycholate, 0.1% SDS, and 2 mM PMSE, 1 μg ml⁻¹ leupeptin, 1 μg ml⁻¹ aprotinin). Bioruptor (Diagenode) was used to ensure consistent sonication between samples. Sonication was performed for 4–6 cycles (30 s-ON, 30 s-OFF). The resulting chromatin extract was precleared and appropriate amount was taken to incubate at 4 °C with 2–10 μg of H3K9me2 (4658, all from Cell Signaling, USA) antibody overnight. Next morning, pre-blocked beads were added and rotated for 3 h at 4 °C. Afterwards, beads were washed with sonication buffer twice, washing buffer (2 mM Tris pH 8.0, 0.02 mM EDTA, 50 mM LiCl, 0.1% NP-40 0.1% Na-deoxycholate) once, TE buffer once, followed by reverse crosslink for 5 h at 65 °C. DNA was then purified using the QIAGEN PCR purification kit according to the manufacturer’s protocol. H3K9me2 relative occupancies were determined by calculating the immunoprecipitation efficiency (ratios of the amount of immunoprecipitated DNA to that of the input sample). Antibodies against H3K9me2 (4658, Cell Signaling, USA) were used for immunoprecipitation. Primers used for qRT-PCR of ChIP are listed in Supplementary Table 2.

Gene expression analysis. RNA from skin and keratinocytes culture was prepared using the GeneElute Mammalian Total RNA miniprep kit (RNT350, Sigma, USA) according to the instructions of the manufacturer. Skin samples for RNA were first

**Fig. 10** Decreased H3K9me2 in psoriatic lesions correlating with increased IL-23p19 expression. a, b H3K9me2 staining (a) and quantification (b) in healthy skin, non-lesioned and lesioned psoriatic skin (n: 3/3, mean ± SD, one-way ANOVA, with Tukey’s multiple comparisons). c H3K9me2 and IL-23p19 staining in non-lesioned and lesioned psoriatic skin (n: 3/3). Scale bars: 100 μm (a, c). (*p ≤ 0.05; **p ≤ 0.01)
kept in RNA later (R0901, Sigma) until isolation. RNA was reverse transcribed to cDNA and calculated based on duplicates and normalized to the housekeeping gene GAPDH.

Microarray Center. Total RNA samples were isolated from total back skin of 3 pairs of mice as well as primary keratinocytes cultured on glass slides were analyzed by using the Affymetrix, and scanning of the arrays was carried out at the Copenhagen University Hospital Microarray Center. Data are presented as means with error bars representing standard deviation (SD). Statistical significance was determined either by the two-tailed Student’s t-test, or by one-way analysis of variance (ANOVA), with Tukey’s multiple comparisons if not stated otherwise. Significant differences are indicated by asterisks $(p \leq 0.05; * p \leq 0.01; ** p \leq 0.001; *** p \leq 0.0001)$. All other data are available from the corresponding author upon request.

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Author contributions

H.L. designed and conducted experiments and wrote the manuscript. Q.Y., A.G.M., X. W., J.H., P.L., H.N., M.S.M., L.V., and B.R. conducted experiments. A.W., C.V., S.F.T., A. A., and S.A.B. provided crucial reagents. E.P. and K.H. provided important advice. C.B. designed experiments and wrote the manuscript.

Additional information

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