Downregulation of epithelial sodium channel (ENaC) activity in human airway epithelia after low temperature incubation

Sangya Yadav,1 Ciaran A Shaughnessy,1 Pamela L Zeitlin,1,2 Preston E Bratcher 3,1

ABSTRACT
Introduction The incubation of airway epithelia cells at low temperatures is a common in vitro experimental approach used in the field of cystic fibrosis (CF) research to thermo-stabilise F508del-CFTR and increase its functional expression. Given that the airway epithelium includes numerous ion transporters other than CFTR, we hypothesised that there was an impact of low temperature incubation on CFTR-independent ionoregulatory mechanisms in airway epithelia derived from individuals with and without CF.

Methods After differentiation at the air–liquid interface, nasal epithelia were incubated at either 37°C or 29°C (low temperature) for 48 hours prior to analysis in an Ussing chamber.

Results While F508del-CFTR activity was increased after low temperature incubation, activity of CFTR in non-CF epithelia was unchanged. Importantly, cultures incubated at 29°C demonstrated decreased transepithelial potential difference (TEPD) and short-circuit currents (Isc) at baseline. The predominant factor contributing to the reduced baseline TEPD and Isc in 29°C cultures was the reduced activity of the epithelial sodium channel (ENaC), evidenced by a reduced responsiveness to amiloride. This effect was observed in cells derived from both non-CF and CF donors.

Discussion Significant transcriptional downregulation of ENaC subunits β and γ were observed, which may partially explain the decreased ENaC activity. We speculate that low temperature incubation may be a useful experimental paradigm to reduce ENaC activity in vitro epithelial cultures.

INTRODUCTION
The conducting and respiratory airway epithelia of the nose, mouth, trachea, bronchi and alveoli process inhaled air for gas exchange while protecting from potentially harmful inhaled particles and pathogens. Key to both the respiratory and protective functions of airway epithelia is maintaining hydromineral balance of the periciliary fluid compartment of the airway surface liquid (ASL), a thin layer of fluid that serves to trap and flush out debris and pathogens.1 ASL hydration is maintained by the osmotic movement of water driven by ion transport across the epithelium. Three of the most prominent ion transporters on the apical side of the airway epithelium are: (i) epithelial Na+ channel (ENaC), which transports Na+ from the ASL into the cell; (ii) cystic fibrosis transmembrane conductance regulator (CFTR), a Cl− channel that transports Cl− out of the cell to the ASL and (iii) Ca2+-activated Cl− channels (CaCCs), which also transport Cl− out to the ASL.2

Cystic fibrosis (CF) is an autosomal recessive disease caused by mutations in the CFTR gene resulting in a deterioration of the ASL and an overaccumulation of mucus in the airways leading to respiratory dysfunction and chronic lung disease.3 The most common CFTR mutation, F508del,4 results
in misfolding and retention of the mutant CFTR protein in the endoplasmic reticulum, leading to the premature degradation of F508del-CFTR by proteasomes and an inadequate amount of F508del-CFTR at the cell surface. The misfolding of F508del-CFTR is temperature-sensitive and can be partially rescued by incubation at low temperature. Thus, a widely used experimental methodology to correct F508del-CFTR folding in vitro is to thermo-stabilise F508del-CFTR using low temperature incubation (at ~27°C–29°C rather than 37°C) of cells harbouring F508del-CFTR. Low temperature correction of F508del-CFTR is also an important experimental tool in translational research on CF as it has served as a positive control treatment in studies screening for and investigating the efficacy of novel pharmacological correctors of F508del-CFTR.

In CF research, ENaC has also been widely investigated for its abundance on the airway epithelium and its prominent contribution to ion and fluid homeostasis on the airway surface. The balance of ENaC-mediated ion absorption and CFTR-mediated ion secretion largely determines proper ASL homeostasis. CFTR and ENaC interact on the apical membrane of the airway epithelium, both through direct and indirect means. In non-CF epithelia, membrane-expressed CFTR directly interacts with and downregulates the function of ENaC. Likewise, ENaC activity is increased in CF epithelia, which have low membrane expression of CFTR. This relationship between ENaC and CFTR is so crucial that it has been suggested that ENaC should be a target in treating CF.

For example, it has been posited that pharmacologically reduced ENaC activity on the apical surface could lead to increased periciliary fluid and enhanced lung function in CF airways. This paradigm is perhaps most conspicuous in murine models of CF, of which models overexpressing ENaC in the airway more closely exhibit pulmonary symptoms of CF than Cfr-/- models.

Given the use of low temperature incubation experiments to investigate F508del-CFTR and airway epithelial ion transport, the current study sought to evaluate the impact of low temperature on the activities of other epithelial ion transporters of the airway epithelium. In this study, we exposed patient-derived non-CF and CF (F508del homozygous) nasal epithelial cells to low temperature. We performed electrophysiological analyses to examine the effect of low temperature incubation on the activities of amiloride-sensitive ENaC, forskolin-activated CFTR and ATP-responsive CaCCs. This report provides further characterisation of the impact of low temperature on the electrophysiological properties and ion channel functions of cultured airway epithelia.

**MATERIALS AND METHODS**

**Primary human nasal epithelial cell expansion and maintenance**

Primary nasal cells were obtained from three to four individuals with and without CF by nasal brushing and cultured as previously described. Cells were expanded on irradiated National Institutes of Health (NIH)-registered 3T3 feeder layers with F-media (73% complete Dulbecco's Modified Eagle's Medium (DMEM) (89% DMEM, 9% fetal bovine serum, 1% L-glutamine, 1% penicillin/streptomycin), 0.1% hydrocortisone/EGF mix, 0.00086% cholera toxin, 1.6% adenine, 0.1% insulin, 25% Ham's F-12) containing 10 µM Y27632 Rho kinase inhibitor. For differentiation at air-liquid interface (ALI), expanded cells were seeded on type I bovine collagen-coated (3 mg/mL) cell culture permeable polyester transwell inserts (Corning) at a density of 250 000 cells/cm² in PneuMaCult-Ex Plus (STEMCELL Technologies). Apical media was removed and basal media was replaced by PneuMaCult-ALI media (STEMCELL Technologies) after 2 days of expansion. The basal media was changed every 2–3 days with fresh PneuMaCult-ALI media. Cells were well-differentiated and analysed after 21–28 days at ALI. For low temperature incubation, cells were either held at 37°C (control) or transferred to 29°C for 48 hours prior to analysis in the Ussing chamber (four replicates per group for each individual donor).

**Electrophysiological measurement by Ussing chamber**

Electrophysiological measurements were performed in an Ussing chamber (Physiological Instruments) under open-circuit conditions with continuous measurement of transepithelial potential difference (TEPD). Monolayers were intermittently voltage-clamped to 0 mV for measurement of short-circuit current (Isc). During periods of short-circuiting, epithelia were pulsed (200 ms of ±5 mV) and measurements of transepithelial electrical resistance (TEER) were obtained. Cells were placed between apical and basolateral compartments filled with Ringer's solution (120 mM NaCl, 10 mM D-glucose, 3.3 mM KH₂PO₄, 0.83 mM K₂HPO₄, 1.2 mM MgCl₂, 1.2 mM CaCl₂, saturated with 95% O₂/5% CO₂, pH 7.4 at 37°C). After mounting in the Ussing chamber, solutions were continuously gassed with 5% CO₂ and 95% O₂. Acute treatments of the monolayers in the Ussing chamber consisted of apical 100 µM amiloride (Alfa Aesar), apical/basal 20 µM forskolin (Fsk; Tocris)/100 µM 3-isobutyl-1-methylxanthine (IBMX; Sigma), apical 1 µM VX-770 (Selleck Chemicals), apical 10 µM CFTR(inh)-172 (CFTR Chemical Compound Distribution Program) and apical 100 µM ATP (Sigma). For non-CF cultures, CFTR was potentiated with VX-770 then activated by Fsk/IBMX. For CF cultures, CFTR was activated by Fsk/IBMX then potentiated with VX-770. After Ussing chamber analysis, monolayers were immediately frozen and stored at −80°C.

**Gene expression analysis**

Total RNA was isolated from frozen cells using the TRIzol method following manufacturer’s protocol (Molecular Research Center) and quantified and assessed for purity using a NanoDrop spectrophotometer (ND-1000; NanoDrop Technologies). First-strand complementary DNA (cDNA) was synthesised using a reverse transcription kit.
following the manufacturer’s protocol (M-MLV Reverse Transcriptase; Invitrogen). Real-time quantitative PCR (RT-qPCR) was performed using a StepOnePlus system (Applied Biosystems) and carried out in 10 µl reactions containing 10 ng cDNA, 150 nM forward and reverse primers and 1X SYBRSelect master mix, following the manufacturer’s protocol (ThermoFisher). The thermal profile of the reactions was 2 min at 50°C, 2 min at 95°C (holding and activation), 40 cycles of 15 s at 95°C, 30 min at 60°C, 30 s at 72°C (cycling), then finally a ramp from 60°C to 95°C (melt curve analysis) was used to confirm a single product in each reaction. Relative mRNA expression of SCNN1A, SCNN1B and SCNN1G (the genes corresponding to α, β and γ subunits of ENaC, respectively) was calculated using the comparative method (2-ΔΔCT) using α-tubulin (ATURAJA) as a reference gene. Nucleotide sequences of primer pairs used for analysis were as follows: SCNN1A forward, 5’-AGGGGACAAGCGTGAGGAA-3’; SCNN1A reverse, 5’-GGTGGAACCTGAATCAAGG-3’; SCNN1B forward, 5’-CCGGCTACACGTTCAAGG-3’; SCNN1B reverse, 5’-CCCTACACGATTGCCTT-3’; SCNN1G forward, 5’-GCACCGGAGAAGATCAA-3’; SCNN1G reverse, 5’-TACCACCGCATTAGCTCTTA-3’.

Western blot analysis
Frozen monolayers were washed with ice-cold phosphate-buffered saline and lysed with 1X Laemmli buffer. Equal amounts of protein (20 µg) were loaded for SDS-PAGE, then transferred on to nitrocellulose membrane (GE-Amersham Biosciences). Blots were subsequently blocked with 5% Blotto (non-fat dry milk; Sigma) in tris-buffered saline with 0.1% Tween-20 (TBS-T) for 1 hour at room temperature (RT), followed by overnight incubation with primary antibodies: ENaC (1:1000 dilution; Invitrogen, Cat. No. 920A); actin (1:3000 dilution; Cell Signaling Technology, Cat. No. 12262). After primary incubation, blots were washed with TBS-T then incubated in HRP-conjugated secondary antibodies (1:3000 dilution; Cell Signaling Technology, Cat. No. 7074) for 1 hour at RT, then washed again before chemiluminescent detection (ECL western blotting substrate; Pierce, Cat. No. 32106). ENaC α-subunit band densities were normalised to β-actin and quantified using ImageJ software (NIH).

Calculations and statistical analysis
To control for donor-to-donor variation, data were normalised to the donor-corrected 37°C control. Statistical analyses were carried out on normalised replicates from each donor (n=16–34 replicates from three to four donors) by either one-way or two-way analysis of variance with Tukey’s multiple comparisons test, unpaired t-test or linear regression as described in figure legends. All data are presented as mean±SD. Figure assembly and all statistical analyses were completed in Prism V.6.0 (GraphPad Software).

RESULTS
Non-CF and CF primary nasal epithelial cell cultures formed visibly ciliated and electrically tight monolayers, ranging 93–358Ω/cm² at baseline. All monolayers exhibited an immediate response and subsequent stabilisation after exposure to the test compounds amiloride, VX-770, Fsk/IBMX, CFTR(inh)-172 and ATP (figure 1A, B). Non-CF and CF epithelia had significantly different baseline TEPD (non-CF: −6.3±2.3 mV; CF: −10.5±2.5 mV), amiloride-responsive ΔTEPD (non-CF: 5.9±2.8 mV; CF: 13.6±2.5 mV) and VX-770/Fsk/IBMX-mediated ΔTEPD (non-CF: −3.8±1.8 mV; CF: −0.92±0.29 mV) (figure 1C–E).

We first sought to compare the effects of low temperature (29°C) incubation on baseline values (prior to administration of test compounds) for TEPD, Isc and TEER between non-CF and CF nasal epithelial cell cultures. We observed a significant reduction in baseline TEPD and Isc after 29°C incubation compared with the control temperature (37°C), and this effect was consistent in both non-CF and CF epithelial cell cultures (figure 2A, B). Low temperature-mediated decreases in baseline TEPD and Isc were greater in CF cells (TEPD...
was reduced 82%±14%; Isc was reduced in 80%±17% compared with non-CF cells (TEPD was reduced 60%±13%; Isc was reduced in 67%±13%). Low temperature incubation had only modest influences on TEER, slightly increasing TEER in non-CF cells and having no effect in CF cells (figure 2C).

The acute application of amiloride, a potent inhibitor of ENaC, had large and temperature-dependent effects on TEPD and Isc across both non-CF and CF epithelia. The magnitude of amiloride-responsive ΔTEPD and ΔIsc were significantly reduced at 29°C compared with 37°C control epithelia (figure 3A,B). In non-CF epithelia at 29°C, amiloride-responsive ΔTEPD and ΔIsc were decreased to 47%±22% and 59%±21%, respectively, compared with 37°C controls. In CF epithelia at 29°C, amiloride-responsive ΔTEPD and ΔIsc were decreased to 62%±9% and 59%±13%, respectively. In non-CF epithelia, CFTR activation by VX-770 and Fsk/IBMX was not different between 29°C and 37°C treatments (figure 3C,D). However, in CF epithelia, CFTR activation by Fsk/IBMX and VX-770 resulted in a significant increase in ΔTEPD at 29°C compared with the 37°C control (figure 3C). Low temperature incubation had no detectable effect on CFTR-activated ΔIsc in either non-CF or CF epithelia (figure 3D). It is important to note that the effect of temperature correction on F508del-CFTR is very moderate when assayed under symmetrical Cl− conditions; more dramatic increases in F508del-CFTR activity after low-temperature incubation are observed in the presence of a Cl− gradient. Although the acute addition of ATP elicited a strong response in Cl− conductance for both non-CF and CF epithelia (figure 1), no significant effect of low temperature incubation on the ATP response was detected in either non-CF or CF epithelia (figure 3E).
In both CF and non-CF subunit protein expression was observed for glycosylated ENaC (αsylated ENaC).

29°C non-CF treatment with normal (37°C) and low (29°C) temperature resulted in a modest (~25%) but significant decrease in levels of ENaC activity may be attributed to the downregulation of ENaC α-subunit—the 97 kDa band corresponding to the glycosylated, thus active, form of ENaC α-subunit.

Low temperature incubation is a widely used experimental tool in vitro to enhance folding and trafficking of F508del-CFTR. Fewer studies have sought to identify the impact of experimental temperature manipulation on other epithelial ion transport processes. In

DISCUSSION

The balance between ENaC and CFTR on the airway epithelium is critical in maintaining airway surface ion and fluid homeostasis. In the present study, we demonstrated that low temperature treatment of airway epithelial cells in vitro, which is an experimental approach often used to modulate F508del-CFTR, also reduced ENaC activity in both non-CF and CF airway epithelia. We further demonstrated that ENaC activity is highly correlated to baseline TEPD, implying that ENaC is critical in producing the lumen-negative TEPD present in non-CF and CF airway epithelia. Protein-level and transcript-level data indicate that temperature-sensitive changes in ENaC activity may be attributed to the downregulation of ENaC expression.

Low temperature incubation is a widely used experimental tool in vitro to enhance folding and trafficking of F508del-CFTR.

(figure 4B): 37°C non-CF: m=1.053, r²=0.6530, p=0.0002; 29°C non-CF: m=0.6273, r²=0.5941, p=0.0005. After the addition of amiloride in non-CF epithelial cells were analysed. Differences in TEPD and Isc (but not TEER) remained non-significant (t-test; p<0.01).
CF bronchial epithelial (CFBE) cells, low temperature incubation increased CFTR surface expression and channel activity, and resulted in lower activity of another Cl− channel, the outwardly rectifying Cl− channel. In the present investigation, low temperature incubation resulted in an expected increase in F508del-CFTR activity after incubation at 29°C compared with 37°C, a decrease in ENaC activity and no effect on the ATP response, which is traditionally viewed as an indicator of CaCC activity. Thus, the impact of low temperature, which could affect many protein regulatory processes, such as protein synthesis, proteolytic activation or protein turnover, appears to be protein-specific rather than general. Importantly, the impact of low temperature incubation on ENaC activity was substantial and will be the primary topic of the discussion that follows.

In the 37°C controls for our study, ENaC activity was greater in CF than non-CF airway epithelia (figure 1D). The upregulation of ENaC activity in CF cultures is firmly established. In both epithelial and non-epithelial cells, CFTR membrane expression and activity is negatively associated with ENaC expression or ENaC-mediated Na+ transport. The reciprocal balance of the activities of ENaC and CFTR on the airway epithelium may aid in maintaining proper osmoregulation of periciliary fluid in non-CF epithelia, but likely exacerbates the dysregulation of ASL homeostasis caused by mutations in CFTR. When CFTR membrane expression on airway epithelium decreases due to improper folding and membrane-trafficking (such as in the case of F508del-CFTR), ENaC functional expression is enhanced, leading to decreased ASL volume, thus worsening mucociliary clearance.

The mechanism by which CFTR and ENaC interact is not well understood. Investigations into the CFTR interacting proteome (ie, the CFTR ‘interactome’) using co-immunoprecipitation (co-IP) of endogenously expressed CFTR did not find any interaction between ENaC and wild-type-CFTR or F508del-CFTR at 37°C or F508del-CFTR at 30°C. Conversely, studies using systems exogenously overexpressing CFTR and ENaC have shown a direct interaction between CFTR and ENaC α, β and γ subunits in co-IP and fluorescence energy transfer (FRET) experiments. Interestingly, low-temperature (27°C) incubation is required for F508del-CFTR to exhibit direct interaction (co-IP) and physical proximity (FRET) with ENaC, indicating that the interaction between CFTR and ENaC likely takes place only once the proteins have trafficked to the plasma membrane. Neither incubation at 37°C nor 27°C resulted in any such interaction between ENaC and the Cl− channel CLC-1, although ENaC is known to interact with another CLC family member, CLC-2. From these previous studies, it appears that the functional relationship between CFTR and ENaC is likely occurring through direct and indirect mechanisms, which both require future investigation. It appears that CFTR can physically interact with ENaC on the plasma membrane to reduce ENaC function in cell lines exogenously expressing CFTR and ENaC, and that the defective membrane-trafficking of F508del-CFTR prevents direct interaction with ENaC. However, the lack of detection of ENaC in the interactome of wild-type-CFTR or F508del-CFTR in cell lines endogenously expressing CFTR indicates that indirect functional interaction between CFTR and ENaC, facilitated through intermediary or accessory proteins, is likely a predominant mode of interaction between CFTR and ENaC.

In the present study, low temperature incubation of F508del-CFTR monolayers resulted in increased CFTR activity and decreased ENaC activity (figure 3A and C). This finding agrees with the well-known effects of low temperature in modulating F508del-CFTR membrane presence. A novel aspect of the present study is that we assessed the influence of low temperature in non-CF in addition to CF primary-derived epithelial cells. In CFBE41o cells, which exogenously express CFTR, low temperature (29°C) incubation reduced wild-type-CFTR activity. In contrast, in the present study we found no change in CFTR activity in non-CF epithelia at 29°C (figure 3C). The difference between the exogenous expression of CFTR in CFBE41o cells and the endogenous expression of CFTR in our primary-derived nasal epithelial cells may account for this difference in the CFTR-regulatory response at low temperature.

An important finding in the present study was that in both non-CF and CF epithelial cells, low temperature incubation reduced apparent ENaC activity (figure 3A). Furthermore, the reduction in ENaC activity substantially altered baseline transepithelial electrical properties such as membrane potential (figure 3A). The strong relationship between ENaC activity and baseline TEPD demonstrates the important influence of ENaC function on hydromineral balance on the airway epithelium. ENaC activity appeared to have a greater influence on baseline TEPD in CF than non-CF epithelia, which is likely due to lower CFTR activity in CF epithelia. Furthermore, in CF epithelia, the relationship between ENaC activity and baseline TEPD was similar regardless of temperature incubation (figure 3A). From these data, it appears that low temperature treatment affects ENaC function through a mechanism that is not necessarily dependent on CFTR, and that the reduced ENaC function at low temperature has a direct impact on baseline transepithelial membrane potential.

The cellular/molecular mechanisms responsible for the downregulation of ENaC activity at low temperature are not clear. Dysregulation of ENaC channel gating, membrane localization and surface residency time are all known pathologies for several human diseases. Along with reduced CFTR function, hyperactivity of ENaC-mediated Na+ absorption exacerbates mucous dehydration and worsens symptoms of CF. Hyperactivation of ENaC as a contributing factor of CF lung disease specifically is thought to be the result of increased ENaC open probability (P0) and/or increased ENaC surface expression. It has been shown in Xenopus oocytes that CFTR...
activation decreases the ENaC $P_o$ without affecting its surface expression.\textsuperscript{45} Increased ENaC $P_o$ can occur by proteolytic cleavage of an extracellular loop by various proteases.\textsuperscript{46} Increased ENaC surface expression can be regulated by cAMP/PKA inhibition of NEDD4-mediated ubiquitination and proteasomal degradation of ENaC.\textsuperscript{46}

The extent to which altered $P_o$ and/or surface expression of the reduced ENaC activity that we observed at 29°C is only partially answered in the present study. Direct comparisons of the results in the present study to the literature are difficult. Although many studies have used low temperature incubation to experimentally enhance F508del-CFTR, we could not identify any studies describing the effect of low temperature incubation on the ENaC activity. It was shown in CFBE41o cells that low temperature (29°C) incubation increased TEER independently of CFTR membrane presence or activity,\textsuperscript{47} although the impact of low temperature incubation on ENaC expression or activity was not disclosed. In the present study, we similarly demonstrate an increase in TEER after low temperature incubation (figure 1C) and attribute this change primarily to a change in ENaC activity. Experimentally applied acute low temperature treatment has been shown to increase ENaC $P_o$.\textsuperscript{47} It is possible that prolonged exposure to low temperature (such as in the present study) results in sustained increases in ENaC $P_o$, thus eliciting a regulatory response by the cells to decrease ENaC activity.

Our RT-qPCR and western blot analyses demonstrated that transcript and/or protein expression of all three ENaC subunits (α, β and γ) were downregulated. As ENaC activity requires all three subunits to be present, the downregulation of any one of the ENaC subunits could explain the observed decreases in ENaC activity. Our results indicate differential regulation of ENaC subunits; transcription of SCNN1B and SCNN1G was strongly downregulated at low temperature whereas SCNN1A was not (figure 5A–C). Despite no observed reduction at the transcript level, protein expression of the ENaC α subunit was still reduced (figure 5E). Given this finding, we surmise that protein expression of β and γ subunits, which were strongly downregulated at the transcript level, were likely reduced. Thus, our results indicate that it is possible that downregulation of ENaC transcript and protein synthesis at low temperature can fully explain the downregulation in ENaC activity. It is important to note that both microarray\textsuperscript{12} and proteomic\textsuperscript{37} analyses have revealed that many cellular functions are affected by low temperature incubation. Therefore, there could be other cellular mechanisms responding to low temperature incubation that may be affecting ENaC activity. Given that, in the present study, differences in the TEED and Isc of non-CF epithelia were observed between 37°C and 29°C treatments after the addition of amiloride (figure 4C–E), we think it is likely that low temperature incubation affects other processes related to epithelial ion transport in addition to the downregulatory effect low temperature incubation has on ENaC activity.

In conclusion, we have reported that ENaC activity in primary-derived airway epithelia is substantially reduced after low temperature incubation. This effect of low temperature appears to be specifically acting on some ion transport mechanisms, namely F508del-CFTR and ENaC, but not others, such as CaCCs. Importantly, the reduction in ENaC activity and expression after low temperature incubation was observed in both non-CF and CF cultures indicating that this effect may occur through a CFTR-independent mechanism. Future studies seeking to use low temperature incubation to experimentally correct mutant CFTR should consider the impact low temperature incubation may have on other ion transporters as well as on epithelial function more broadly, including impacts on TEED, ASL hydration and mucociliary transport.

Lastly, for investigators interested in studying ENaC function, we speculate that low temperature incubation may be a useful experimental paradigm to induce specific downregulation of ENaC activity in order to study the ENaC dysfunction in various disease and model systems.

Acknowledgements The authors would like to thank Dr Robert Bridges, the Rosalind Franklin University of Medicine and the Cystic Fibrosis Foundation for the contribution of CFTRinh-172 through the CFTR Chemical Compound Distribution Programme.

Contributors SY, CAS and PEB conceived of and designed the research and performed experiments; SY, CAS and PEB analysed data and prepared the figures; SY and CAS produced the original manuscript draft; SY, CAS, PLZ and PEB revised and approved the final version of manuscript; PLZ and PEB facilitated the research.

Funding This research was supported by the Cystic Fibrosis Foundation (ZeitLI2010 to PLZ and Bratcher1610 to PEB), the Eugene F. and Easton M. Crawford Charitable Lead Unitrust (to CAS and PEB), the Gilead Sciences Research Scholars Programme in Cystic Fibrosis (to PEB) and the Department of Paediatrics at National Jewish Health.

Competing interests None declared.

Patient and public involvement Patients and/or the public were not involved in the design, or conduct, or reporting, or dissemination plans of this research.

Patient consent for publication Not required.

Ethics approval This study was approved by the Institutional Review Board of National Jewish Health: HS-2832.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement All data relevant to the study are included in the article.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non-Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.

ORCID iD Preston E Bratcher http://orcid.org/0000-0003-2240-8632

REFERENCES

1. Boucher R. Regulation of airway surface liquid volume by human airway epithelia. Pflugers Arch - Eur J Physiol 2003;445:495–8.
2. Bartoszewski R, Matalon S, Collawn JF. Ion channels of the lung and their role in disease pathogenesis. Am J Physiol Lung Cell Mol Physiol 2017;313:L859–72.
3. Rowe SM, Miller S, Sorscher EJ. Cystic fibrosis. N Engl J Med 2005;352:1992-2001.
4. Kerem B, Rommens J, Buchanan J, et al. Identification of the cystic fibrosis gene: genetic analysis. Science 1989;245:1073–80.
Kashlan OB, Kinlough CL, Myerburn 2016;55:323–36.

Progenitor clone formation is enhanced by Y-27632–Dependent Nat Commun the human airway epithelium. 2019;10:1763.

Chioccioli M, Feriani L, Kotar J, and coordination in response to CFTR- modulation in airway epithelia cultured from nasal brushings. 2020;8:e14603.

Ouabain mimics low temperature rescue of F508del- Am J Physiol Cell Physiol 2008;366:1025–9.

Rescue of the mutant CFTR by VX-809 and suggest multiple agents EMBO Rep 2001;2:1047–51.

Malik B, Elkhaib J, et al. CFTR-mediated inhibition of epithelial Na+ conductance in human colon is defective in cystic fibrosis. Am J Physiol Gastrointest Liver Physiol 1999;277:O709–16.

Yan W, Samahaa FF, et al. Cystic fibrosis transmembrane conductance regulator differentially regulates human and mouse epithelial sodium channels in Xenopus oocytes. J Biol Chem 2004;279:23183–92.

Cell 1990;63:827–34.

Bratcher PE, Yuan C, Liao J, Robert R, et al. The cystic fibrosis transmembrane conductance regulator impedes proteolytic stimulation of the epithelial Na+ channel. Journal of Biological Chemistry 2010;285:32227–32.

Ismailov II, Awaysda MS, Jovov B, et al. Regulation of epithelial sodium channels by the cystic fibrosis transmembrane conductance regulator. Journal of Biological Chemistry 1996;271:4725–32.

König J, Schreiber R, Voeiker T, et al. The cystic fibrosis transmembrane conductance regulator (CFTR) inhibits EnAc through an increase in the intracellular CI− concentration. EMBO Rep 2001;2:1047–51.

Mall M, Bleich M, Kuehr J, et al. CFTR-mediated inhibition of epithelial Na+ conductance in normal but not in cystic fibrosis airways. J Clin Invest 1998;102:15–21.

SNSO E, Tervo M, Calci M, et al. Rescue of the mutant CFTR chloride channel by pharmacological correctors and low temperature analyzed by gene expression profiling. Am J Physiol Cell Physiol 2011;301:C872–85.

Yadav S, et al., Tzotzos ST, Bedak M, et al. Rescue of the mutant CFTR by VX-809 and suggest multiple agents EMBO Rep 2001;2:1047–51.

Gomes-Alves P, Neves S, Coelho AV, et al. Low temperature restoring effect on F508del-CFTR misprocessing: a proteomic approach. J Proteomics 2009;73:218–30.

Gentzsch M, Dang h, Dang Y. The cystic fibrosis transmembrane conductance regulator impedes proteolytic stimulation of the epithelial Na+ channel. Journal of Biological Chemistry 2010;285:32227–32.

Yadav S, et al., BMJ Open Resp Res 2021;8:000861. doi:10.1136/bmjresp-2020-000861