RESEARCH NOTE

Cell signaling promoting protein carbonylation does not cause sulfhydryl oxidation: Implications to the mechanism of redox signaling [version 1; peer review: 1 approved, 2 approved with reservations]

Yuichiro J. Suzuki¹, Faisal Almansour¹, Camilla Cucinotta¹, Vladyslava Rybka¹, Lucia Marcocci²

¹Department of Pharmacology and Physiology, Georgetown University Medical Center, Washington, DC, 20057, USA
²Department of Biochemical Sciences “A. Rossi Fanelli”, Sapienza University of Rome, Rome, Italy

Abstract

Reactive oxygen species (ROS) have been recognized as second messengers, however, targeting mechanisms for ROS in cell signaling have not been defined. While ROS oxidizing protein cysteine thiols has been the most popular proposed mechanism, our laboratory proposed that ligand/receptor-mediated cell signaling involves protein carbonylation. Peroxiredoxin-6 (Prx6) is one protein that is carbonylated at 10 min after the platelet-derived growth factor (PDGF) stimulation of human pulmonary artery smooth muscle cells. In the present study, the SulfoBiotics Protein Redox State Monitoring Kit Plus (Dojindo Molecular Technologies) was used to test if cysteine residues of Prx6 are oxidized in response to the PDGF stimulation. Human Prx6 has a molecular weight of 25 kDa and contains two cysteine residues. The Dojindo system adds the 15 kDa Protein-SHifter if these cysteine residues are reduced in the cells. Results showed that, in untreated cells, the Prx6 molecule predominantly exhibited the 55 kDa band, indicating that both cysteine residues are reduced in the cells. Treatment of cells with 1 mM H₂O₂ caused the disappearance of the 55 kDa band and the appearance of a 40 kDa band, suggesting that the high concentration of H₂O₂ oxidized one of the two cysteine residues in the Prx6 molecule. By contrast, PDGF stimulation had no effects on the thiol status of the Prx6 molecule. We concluded that protein carbonylation is a more sensitive target of ROS during ligand/receptor-mediated cell signaling than sulfhydryl oxidation.

Keywords
cell signaling, protein oxidation, reactive oxygen species, redox signaling
Corresponding author: Yuichiro J. Suzuki (ys82@georgetown.edu)

Competing interests: No competing interests were disclosed.

Grant information: This work was supported by the National Institutes of Health, National Heart, Lung, and Blood Institute and National Institute of Aging (Grants R01 HL72844 and R03 AG047824) to YJS. The content is solely the responsibility of the authors and does not represent the official views of the National Institutes of Health.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Copyright: © 2017 Suzuki YJ et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. Data associated with the article are available under the terms of the Creative Commons Zero "No rights reserved" data waiver (CC0 1.0 Public domain dedication).

How to cite this article: Suzuki YJ, Almansour F, Cucinotta C et al. Cell signaling promoting protein carbonylation does not cause sulfhydryl oxidation: Implications to the mechanism of redox signaling [version 1; peer review: 1 approved, 2 approved with reservations] F1000Research 2017, 6:455 https://doi.org/10.12688/f1000research.11296.1

First published: 10 Apr 2017, 6:455 https://doi.org/10.12688/f1000research.11296.1
Introduction

Reactive oxygen species (ROS) have been shown to play important roles in cell signaling (Finkel, 2011; Suzuki et al., 1997). In particular, the roles of ROS in cell growth signaling have been well documented (Rao & Berk, 1992; Sundaresan et al., 1995). For the mechanism of ROS signaling, the receptor activation producing ROS via NAD(P)H oxidase is a widely accepted concept (Griendling et al., 1994). However, molecular targeting mechanisms for ROS in cell signaling have been unclear. ROS targeting protein cysteine thiols has been the most popular proposed mechanism (‘D’Autreaux & Toledano, 2007; Forman et al., 2010; Moran et al., 2001; Rhee et al., 2000; Sen, 2000; Truong & Carroll, 2012; Veal et al., 2007), yet the occurrence of thiol oxidation requires levels of ROS that are much higher than what is expected to occur during cell signaling (Burgoyne et al., 2007).

Our laboratory has proposed that ligand/receptor-mediated cell signaling involves protein carbonylation (Wong et al., 2008; Wong et al., 2010), which occurs on four susceptible amino acid residues: proline, arginine, lysine, and threonine (Amici et al., 1989; Berlett & Stadtman, 1997). Notably, in cultured cells, hydrogen peroxide \((\text{H}_2\text{O}_2)\) as low as 0.5 µM was found to promote protein carbonylation (Wong et al., 2008).

More recently, we identified proteins that are carbonylated in response to the platelet-derived growth factor (PDGF) stimulation. Among them, peroxiredoxin-6 (Prx6) was found to be carbonylated in response to a 10-min treatment of human pulmonary artery smooth muscle cells with PDGF (Wong et al., 2013). Peroxiredoxins have been shown to regulate cell signaling (Woo et al., 2010). The present study tested whether this signaling mechanism also promotes sulfhydryl oxidation within the Prx6 molecule.

Methods

HPASMCs (ScienCell Research Laboratories, Carlsbad, CA, USA) were serum-starved overnight and treated with recombinant human PDGF-BB or \(\text{H}_2\text{O}_2\) for 10, 15 or 30 min. Protein thiol states were monitored using SulfoBiotics Protein Redox State Monitoring Kit Plus (Dojindo Molecular Technologies, Rockville, MD, USA) in accordance with the manufacturer’s instructions. Briefly, cells were washed, proteins precipitated with trichloroacetic acid andlysate samples were prepared in accordance with the manufacturer’s instructions. Four proteins were subjected to Western blotting, the human Prx6 molecule with a molecular weight of 25 kDa has two cysteine residues. Our results indicated that untreated human pulmonary artery smooth muscle cells predominantly contain the 55 kDa species, consistent with the Prx6 molecule, which has two Protein-SHifters incorporated, indicating that both cysteine residues occur in reduced form in the cells (Figure 1A, lane 1). Treatment of cells with PDGF (10 ng/ml) for 10 min, which promoted protein carbonylation of Prx6 (Wong et al., 2013), did not alter the thiol state of Prx6 (Figure 1A, lane 1 and lane 2). The PDGF treatment

![Figure 1](image)

**Figure 1.** The thiol state of the Prx6 molecule is not altered by PDGF stimulation. Human pulmonary artery smooth muscle cells were treated with PDGF (10 ng/ml) for 10 or 30 min as described in Wong et al. (2013), or with \(\text{H}_2\text{O}_2\) (1 mM) for 15 min. Cellular proteins were precipitated with trichloroacetic acid and lysate samples were prepared in accordance with the manufacturer’s instructions for SulfoBiotics Protein Redox State Monitoring Kit Plus (Dojindo). The Protein-SHifter Plus that covalently binds to reduced protein thiols was added and the samples were subjected to electrophoresis through a 12% polyacrylamide gel. Each Protein-SHifter Plus causes ~15 kDa shift of the protein bands. After electrophoresis, the gel was exposed to UV irradiation to excise the Protein-SHifter Plus moiety, and then subjected to electrophoresis to a nitrocellulose membrane and Western blotting with the Prx6 antibody. (A) Representative Western blotting image of six experiments. (B) Diagram of the native 25 kDa Prx6 molecule, the 40 kDa Prx6 molecule with one Protein-SHifter attached, and the 55 kDa Prx6 molecules with two Protein-Shifters attached. (C) The bar graph represents means (+ SEM) of the intensity of the 55 kDa band (N = 5). The symbol (*) denotes that the value is significantly different from all other values.

**Results**

The technology developed for SulfoBiotics Protein Redox State Monitoring Kit Plus, by Dojindo Molecular Technologies adds a 15 kDa Protein-SHifter on free sulfhydryl groups, allowing the visualization of the thiol status of a given protein by coupling with immunoblotting. The human Prx6 molecule with a molecular weight of 25 kDa has two cysteine residues. Our results indicated that untreated human pulmonary artery smooth muscle cells predominantly contain the 55 kDa species, consistent with the Prx6 molecule, which has two Protein-SHifters incorporated, indicating that both cysteine residues occur in the reduced form in the cells (Figure 1A, lane 1). Treatment of cells with PDGF (10 ng/ml) for 10 min, which promoted protein carbonylation of Prx6 (Wong et al., 2013), did not alter the thiol state of Prx6 (Figure 1A, lane 1 and lane 2). The PDGF treatment

![Image](image)
for 30 min did not alter the thiol state of Prx6 either (Figure 1A, lane 1 and lane 3). By contrast, treatment of H₂O₂ at a high concentration (1 mM) eliminated the 55 kDa band and generated a 40 kDa band that is consistent with one sulphydryl group being oxidized (Figure 1A, lane 4). These results were reproduced at least five times. Dataset 1 (Suzuki et al., 2017) contains the uncropped version of Figure 1A and the uncropped repeats. The bar graph shows the data from five separate experiments with five separate cell treatments. Control experiments were performed to ensure that PDGF stimulated protein phosphorylation as well as carbonylation.

Protein carbonylation is promoted by metal-catalyzed generation of hydroxyl radicals, which are known to promote oxidation indiscriminately. However, the caged and site-directed production of hydroxyl radicals via metals could confer specificity (Stadtman & Berlett, 1991; Wong et al., 2010).

Data availability
Dataset 1. The uncropped version of Figure 1A and the uncropped repeats.
DOI, 10.5256/f1000research.11296.d157362 (Suzuki et al., 2017)

Author contributions
YJS conceived the study and designed the experiments. CC, FA, LM, VR, and YJS carried out the research. YJS prepared the first draft of the manuscript. All authors were involved in the revision of the draft manuscript and have agreed to the final content.

Competing interests
No competing interests were disclosed.

Grant information
This work was supported by the National Institutes of Health, National Heart, Lung, and Blood Institute and National Institute of Aging (Grants R01 HL72844 and R03 AG047824) to YJS. The content is solely the responsibility of the authors and does not represent the official views of the National Institutes of Health.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

References

Amici A, Levine RL, Tsai L, et al.: Conversion of amino acid residues in proteins and amino acid homopolymers to carbonyl derivatives by metal-catalyzed oxidation reactions. J Biol Chem. 1989; 264(6): 3341–3346. PubMed Abstract
Berlett BS, Stadtman ER: Protein oxidation in aging, disease, and oxidative stress. J Biol Chem. 1997; 272(33): 20313–20316. PubMed Abstract | Publisher Full Text
Burgoyne JR, Madhani M, Cuello F, et al.: Cysteine redox sensor in PKGIIa enables oxidant-induced activation. Science. 2007; 317(5843): 1393–1397. PubMed Abstract | Publisher Full Text
D’Autréaux B, Toledano MB: ROS as signalling molecules: mechanisms that generate specificity in ROS homeostasis. Nature Rev Mol Cell Biol. 2007; 8(10): 813–824. PubMed Abstract | Publisher Full Text
Finkel T: Signal transduction by reactive oxygen species. J Cell Biol. 2011; 194(1): 7–15. PubMed Abstract | Publisher Full Text | Free Full Text
Forman HJ, Maiorino M, Ursini F: Signaling functions of reactive oxygen species. Biochemistry 2010; 49(5): 835–842. PubMed Abstract | Publisher Full Text | Free Full Text
Griendling KK, Minieri CA, Ollerenshaw JD, et al.: Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. Circ Res. 1994; 74(6): 1141–1148. PubMed Abstract | Publisher Full Text
Moran LK, Gutteridge JM, Quinlan GJ: Thiols in cellular redox signalling and control. Curr Med Chem. 2001; 8(7): 763–772. PubMed Abstract | Publisher Full Text
Rao GN, Berk BC: Active oxygen species stimulate vascular smooth muscle cell growth and prote- oncogene expression. Circ Res. 1992; 70(3): 593–599. PubMed Abstract | Publisher Full Text
Rhee SG, Bae YS, Lee SR, et al.: Hydrogen peroxide: a key messenger that modulates protein phosphorylation through cysteine oxidation. Sci STKE. 2000; 2000(53): pe1. PubMed Abstract | Publisher Full Text
Sen CK: Cellular thiols and redox-regulated signal transduction. Curr Top Cell Regul. 2000; 36: 1–30. PubMed Abstract | Publisher Full Text
Stadtman ER, Berlett BS: Fenton chemistry. Amino acid oxidation. J Biol Chem. 1991; 266(26): 17201–17211. PubMed Abstract
Sundaresan M, Yu ZX, Ferrans VJ, et al.: Requirement for generation of H₂O₂ for platelet-derived growth factor signal transduction. Science. 1995; 270(5234): 296–299. PubMed Abstract | Publisher Full Text
Suzuki Y, Almansour F, Cucinotta C, et al.: Dataset 1 in: Cell signaling promoting protein carbonylation does not cause sulphydryl oxidation: Implications to the mechanism of redox signaling. F1000Research. 2017. Data Source
Suzuki YJ, Forman HJ, Sevanian A: Oxidants as stimulators of signal transduction. Free Radic Biol Med. 1997; 22(1–2): 269–285. PubMed Abstract | Publisher Full Text
Truong TH, Carroll KS: Redox regulation of epidermal growth factor receptor signaling through cysteine oxidation. *Biochemistry.* 2012; 51(50): 9954–9965. PubMed Abstract | Publisher Full Text | Free Full Text
Veal EA, Day AM, Morgan BA: Hydrogen peroxide sensing and signaling. *Mol Cell.* 2007; 26(1): 1–14. PubMed Abstract | Publisher Full Text
Wong CM, Cheema AK, Zhang L, et al.: Protein carbonylation as a novel mechanism in redox signaling. *Circ Res.* 2008; 102(3): 310–318. PubMed Abstract | Publisher Full Text
Wong CM, Marcocci L, Das D, et al.: Mechanism of protein decarbonylation. *Free Radic Biol Med.* 2013; 66: 1126–1133. PubMed Abstract | Publisher Full Text | Free Full Text
Wong CM, Marcocci L, Liu L, et al.: Cell signaling by protein carbonylation and decarbonylation. *Antioxid Redox Signal.* 2010; 12(3): 393–404. PubMed Abstract | Publisher Full Text | Free Full Text
Woo HA, Yim SH, Shin DH, et al.: Inactivation of peroxiredoxin I by phosphorylation allows localized H$_2$O$_2$ accumulation for cell signaling. *Cell.* 2010; 140(4): 517–528. PubMed Abstract | Publisher Full Text
The authors describe the effects of PDGF and H2O2 treatment on the oxidation state of Prdx6 using a thiol probe, that when attached to free thiols increases the molecular weight of the protein by 15 kDa for each probe attached to the protein. The authors demonstrate that H2O2 treatment causes a change in the redox status of Prdx6 as compared to PDGF treatment. There are a number of issues that need to be resolved and validated by the authors before they can make some of the statements made within the manuscript.

It is essential that the authors fully describe the sample preparation before analysis as this could greatly affect the results and interpretations made. In Figure 1 the authors describe that Prdx6 when the “Protein-SHifter” is added the protein has a mol weight of 55 kDa in controls and the PDGF treatments and one free thiol with the H2O2 treatment, but in Fig1B they show the native state of Prdx6 forming an intra-disulphide, was a reducing agent used in the sample preparation to reduce this disulphide? Does the catalytic Cys47 of this 1-Cys peroxiredoxin form an intra-disulphide with Cys91? It would also be helpful if a non “Protein-SHifter” treated sample was included in the blot to demonstrate the native band at 25 kDa. From Fig1A it would appear that there is a much more intense band for Prdx6 in the H2O2 treated samples, is there a loading control that can be included for this blot?

Carbonylation usually refers to the introduction of an aldehyde or ketone group on an amino acid, I am not sure if this is what the authors are referring to in the title and throughout the manuscript. It is well known that Cys47 of Prdx6 forms a sulphinic (-SO2H) and/or sulphonic (-SO3H) acid. Indeed Prdx6 has been described as having quite a number of various modifications (Jeong, J et al, Proteomics, 2012) so the authors need to confirm the carbonylation or other modifications by mass spectrometry. It is clear that one of the Cys residues is not amenable to “Protein-SHifter” after H2O2 treatment, it would be helpful if they could identify which cysteine residue is susceptible to oxidation.

Is the work clearly and accurately presented and does it cite the current literature?
Partly

**Is the study design appropriate and is the work technically sound?**
Partly

**Are sufficient details of methods and analysis provided to allow replication by others?**
No

If applicable, is the statistical analysis and its interpretation appropriate?
Yes

**Are all the source data underlying the results available to ensure full reproducibility?**
Partly

**Are the conclusions drawn adequately supported by the results?**
No

**Competing Interests:** No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

---

**Author Response ( ) 07 Jun 2017**

**Yuichiro Suzuki,** Georgetown University Medical Center, Washington, USA

Referee: It is essential that the authors fully describe the sample preparation before analysis as this could greatly affect the results and interpretations made.

Authors’ response: As stated in the Methods section “Protein thiol states were monitored using SulfoBiotics Protein Redox State Monitoring Kit Plus (Dojindo Molecular Technologies, Rockville, MD, USA) in accordance with the manufacturer's instructions.” The instructions for SulfoBiotics Protein Redox State Monitoring Kit Plus including the sample preparations can be viewed at [http://www.dojindo.com/store/p/942-SulfoBiotics-Protein-Redox-State-Monitoring-Kit-Plus.html](http://www.dojindo.com/store/p/942-SulfoBiotics-Protein-Redox-State-Monitoring-Kit-Plus.html).

Referee: In Figure 1 the authors describe that Prdx6 when the “Protein-SHifter” is added the protein has a mol weight of 55 kDa in controls and the PDGF treatments and one free thiol with the H2O2 treatment, but in Fig1B they show the native state of Prdx6 forming an intra-disulphide, was a reducing agent used in the sample preparation to reduce this disulphide? Does the catalytic Cys47 of this 1-Cys peroxiredoxin form an intra-disulphide with Cys91?

Authors’ response: The referee is correct that the scheme in Fig. 1B is confusing. In this figure, we did not imply that the 25kD species actually has a disulphide bond but the cartoon merely depicts that both sulphydryl groups are oxidized with “0 SH”.
Referee: It would also be helpful if a non “Protein-SHifter” treated sample was included in the blot to demonstrate the native band at 25 kDa.

Authors’ response: We have done these control experiments many times. Without “Protein-SHifter”, Prx6 gives a band at 25 kDa.

Referee: From Fig1A it would appear that there is a much more intense band for Prdx6 in the H2O2 treated samples, is there a loading control that can be included for this blot?

Authors’ response: Other than that BCA protein assay can be used to monitor total protein levels in the cell lysates prepared using Dojindo SulfoBiotics Protein Redox State Monitoring Kit Plus, neither Dojindo Molecular Technology, Inc nor our laboratory have yet developed loading control to be used in this system. Thus, we rely on performing multiple experiments to make appropriate conclusions.

Referee: Carbonylation usually refers to the introduction of an aldehyde or ketone group on an amino acid, I am not sure if this is what the authors are referring to in the title and throughout the manuscript.

Authors’ response: The referee is correct that we refer protein carbonylation as a process that forms reactive ketones or aldehydes that can be reacted by 2,4-dinitrophenylhydrazine (DNPH) to form hydrazones.

Referee: It is well known that Cys47 of Prdx6 forms a sulphinic (-SO2H) and/or sulphonic (-SO3H) acid. Indeed Prdx6 has been described as having quite a number of various modifications (Jeong, J et al, Proteomics, 2012) so the authors need to confirm the carbonylation or other modifications by mass spectrometry. It is clear that one of the Cys residues is not amenable to “Protein-SHifter” after H2O2 treatment, it would be helpful if they could identify which cysteine residue is susceptible to oxidation.

Authors’ response: This particular Research Note was intended to communicate with the scientific community that, under the condition where protein carbonylation is elicited as previously described by our laboratory (Wong et al., 2013), thiol oxidation was not detected by using a novel method of Dojindo SulfoBiotics Protein Redox State Monitoring Kit Plus. The referee raises exciting and important questions. Based on some of the results obtained while performing experiments for the present study, our laboratory is further investigating the redox biology of peroxiredoxin 6 and we wish to publish a full paper in the near future.

**Competing Interests:** None
Tanea T. Reed  
Department of Chemistry, Eastern Kentucky University, Richmond, KY, USA

The authors studied the response of the antioxidant protein, peroxiredoxin-6 to treatment with PDGF and hydrogen peroxide. By using a commercially available kit, the authors discovered oxidation in one of the cysteine residues at high concentrations of H2O2. My only issue with this work is for Figure 1A. The authors state that they tested three time points of hydrogen peroxide, but only one is shown in the figure. By showing all three time points could further verify the finding of this report as the 40kD would be most potentially pronounced at 30 min.

Is the work clearly and accurately presented and does it cite the current literature?  
Yes

Is the study design appropriate and is the work technically sound?  
Yes

Are sufficient details of methods and analysis provided to allow replication by others?  
Yes

If applicable, is the statistical analysis and its interpretation appropriate?  
Yes

Are all the source data underlying the results available to ensure full reproducibility?  
Partly

Are the conclusions drawn adequately supported by the results?  
Yes

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.
three time points could further verify the finding of this report as the 40kD would be most potentially pronounced at 30 min.

Authors' Response: The reviewer is correct that the statement in the Methods section "HPASMCs were serum-starved overnight and treated with recombinant human PDGF-BB or H2O2 for 10, 15 or 30 min." is confusing. More precisely, it should have stated "HPASMCs were serum-starved overnight and treated with recombinant human PDGF-BB for 10 or 30 min and H2O2 for 15 min." Experimental design was based on our previous report (Wong et al., 2013), showing that PDGF causes carbonylation of Prx6 at 10 min and decarbonylation at 30 min. H2O2 was merely used as a positive control in accordance with the instruction for the Dojindo SulfoBiotics Protein Redox State Monitoring Kit Plus. We have performed time course experiments with H2O2 and found that the appearance of the 40 kD band occurs as early as 5 min and the level is sustained up to at least 30 min.

**Competing Interests:** none

---

**Reviewer Report 25 April 2017**

https://doi.org/10.5256/f1000research.12189.r22184

© 2017 Hussain S. This is an open access peer review report distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**Sabah N.A. Hussain**

Meakins-Christie Laboratories, Translational Research in Respiratory Diseases Program, Department of Critical Care Medicine, Research Institute of the MUHC, McGill University Health Centre (MUHC), Montreal, QC, Canada

The authors provided indirect evidence that peroxiredoxin-6 does not undergo sulphydryl oxidation when human pulmonary artery smooth muscle cells are exposed to PDGF but this protein undergo sulphydryl oxidation when these cells were exposed to H2O2. It was concluded that protein carbonylation is more sensitive target of ROS during ligand/receptor-mediated cell signaling than sulphydrul oxidation.

Major comments: I believe that the conclusion of this study is too general and the authors should restrict themselves to the main findings of this study and do not extend their observation beyond one type of cells exposed to one growth factor (PDGF).

In addition, the authors used an indirect method to assess sulphydryl oxidation rather than a direct measurement. Moreover, the authors did not provide evidence in the current study that PDGF actually produced carbonylation of Prx6. This data is required to document the differential oxidation response of this protein to these two interventions (H2O2 vs. PDGF exposure).
Finally, the authors need to provide data as to the time course of Prx6 oxidation in response to H2O2 exposure. They have only shown one time point.

Is the work clearly and accurately presented and does it cite the current literature?  
Yes

Is the study design appropriate and is the work technically sound?  
Yes

Are sufficient details of methods and analysis provided to allow replication by others?  
Yes

If applicable, is the statistical analysis and its interpretation appropriate?  
Yes

Are all the source data underlying the results available to ensure full reproducibility?  
Yes

Are the conclusions drawn adequately supported by the results?  
No

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Angiogenesis, ROS signaling, NO biology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

---

**Author Response ( ) 29 Jul 2017**

**Yuichiro Suzuki**, Georgetown University Medical Center, Washington, USA

The authors provided indirect evidence that peroxiredoxin-6 does not undergo sulfhydryl oxidation when human pulmonary artery smooth muscle cells are exposed to PDGF but this protein undergo sulfhydryl oxidation when these cells were exposed to H2O2. It was concluded that protein carbonylation is more sensitive target of ROS during ligand/receptor-mediated cell signaling than sulfhydryl oxidation.

[RESPONSE: Please note that, in this study, a high concentration of H2O2 was merely used as a positive control to ensure that our experimental system works in accordance with the instruction for the Dojindo SulfoBiotics Protein Redox State Monitoring Kit Plus.]

Major comments: I believe that the conclusion of this study is too general and the authors should restrict themselves to the main findings of this study and do not extend their observation beyond one type of cells exposed to one growth factor (PDGF).
[RESPONSE: The reviewer is correct that we herein report findings concerning PDGF-signaling in human pulmonary artery smooth muscle cells, as indicated in Abstract, Introduction, Methods, Results, and Discussion sections.]

In addition, the authors used an indirect method to assess sulphydryl oxidation rather than a direct measurement. Moreover, the authors did not provide evidence in the current study that PDGF actually produced carbonylation of Prx6. This data is required to document the differential oxidation response of this protein to these two interventions (H2O2 vs. PDGF exposure).

[RESPONSE: Using mass spectrometry, we have recently identified the formation of glutamic semialdehyde on the Prx6 protein molecule in response to the PDGF treatment of cultured human pulmonary artery smooth muscle cells, confirming the induction of protein carbonylation. This work is currently ongoing, and we wish to publish these results soon. Also, please note that, in this Research Note, a high concentration of H2O2 was merely used as a positive control to ensure that our experimental system works in accordance with the instruction for the Dojindo SulfoBiotics Protein Redox State Monitoring Kit Plus.]

Finally, the authors need to provide data as to the time course of Prx6 oxidation in response to H2O2 exposure. They have only shown one time point.

[RESPONSE: Please note that, in this reported study, H2O2 (1 mM, 15 min) was merely used as a positive control to ensure that our experimental system works in accordance with the instruction for the Dojindo SulfoBiotics Protein Redox State Monitoring Kit Plus. For the subsequent study, our laboratory has performed time course and dose response experiments with H2O2. We found that the appearance of the 40 kD band occurs as early as 5 min, and the level is sustained up to at least 30 min. We wish to publish these results along with other new findings conceding the redox regulation of Prx6 soon.]

**Competing Interests:** No competing interests were disclosed.
The benefits of publishing with F1000Research:

• Your article is published within days, with no editorial bias
• You can publish traditional articles, null/negative results, case reports, data notes and more
• The peer review process is transparent and collaborative
• Your article is indexed in PubMed after passing peer review
• Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com