Molecular Mechanism Underlying the Synergetic Effect of Jasmonate on Abscisic Acid Signaling during Seed Germination in Arabidopsis

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Running title: JAZ-ABI5/ABI3 module integrates JA and ABA

One-sentence summary: JAZ repressors of the jasmonic acid (JA) pathway directly regulate abscisic acid (ABA)-responsive transcription factors to integrate JA and ABA signals during germination and post-germinative growth.

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ABSTRACT

Abscisic acid (ABA) is known to suppress seed germination and post-germinative growth of Arabidopsis thaliana, and jasmonate (JA) enhances ABA function. However, the molecular mechanism underlying the crosstalk between the ABA and JA signaling pathways remains largely elusive. Here, we show that exogenous coronatine (COR), a JA analog structurally similar to the active conjugate jasmonate-isoleucine (JA–Ile), significantly enhances the delayed seed germination response to ABA. Disruption of the JA receptor, CORONATINE INSENSITIVE1 (COI1), or accumulation of the JA signaling repressor, JASMONATE ZIM-DOMAIN, reduced ABA signaling, while jaz mutants enhanced ABA responses. Mechanistic investigations revealed that several JAZ repressors of JA signaling physically interact with ABSCISIC ACID INSENSITIVE3 (ABI3), a critical transcription factor that positively modulates ABA signaling, and that JAZ proteins repress the transcription of ABI3 and ABI5. Further genetic analyses showed that JA activates ABA signaling and requires functional ABI3 and ABI5. Overexpression of ABI3 and ABI5 simultaneously suppressed the ABA-insensitive phenotypes of the coil-2 mutant and JAZ-accumulating (JAZ-ΔJas) plants. Together, our results reveal a previously uncharacterized signaling module in which JAZ repressors of the JA pathway regulate the ABA-responsive ABI3 and ABI5 transcription factors to integrate JA and ABA signals during seed germination and post-germinative growth.

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INTRODUCTION

The processes of seed germination and subsequent seedling establishment are precisely regulated by external and internal cues, including phytohormones. Abscisic acid (ABA) is an important phytohormone that regulates numerous physiological processes in plants, including seed germination, stomatal aperture, and seedling growth, as well as plant responses to various abiotic and biotic stresses (Mauch-Mani and Mauch, 2005; Finkelstein et al., 2008; Fujita et al., 2011; Hauser et al., 2011; Nakashima and Yamaguchi-Shinozaki, 2013). For example, ABA promotes seed dormancy and suppresses seed germination. In the presence of ABA, the ABA receptors PYRABACTIN RESISTANCE (PYR)/REGULATORY COMPONENT OF ABSCISIC ACID RECEPTOR (RCAR) recognize the ABA molecule (Ma et al., 2009; Miyazono et al., 2009; Nishimura et al., 2009; Santiago et al., 2009). Upon binding to ABA, these receptors form a stable complex with type 2C protein phosphatases (PP2Cs), leading to the release of SNF1-related kinases 2 (SnRK2s) from PP2C-SnRK2 complexes (Cutler et al., 2010). The activated SnRK2s subsequently phosphorylate downstream transcription factors such as ABSCISIC ACID-INSENSITIVE5 (ABI5), ABI4, and ABI3 to mediate ABA responses (Kobayashi et al., 2005; Furihata et al., 2006; Fujii et al., 2007; Fujii and Zhu, 2009; Nakashima et al., 2009; Zhu et al., 2020).

The transcription factors ABI3 and ABI5 are highly induced by ABA and positively modulate ABA responses to suppress seed germination (Finkelstein, 1994; Lopez-Molina et al., 2001; Brocard et al., 2002; Finkelstein et al., 2005). Mechanistic investigations have revealed that ABI3 and/or ABI5 physically interact with several critical regulators to integrate multiple signaling pathways during seed germination. For instance, Park et al. (2011) demonstrated that ABI3 forms a protein complex with PHYTOCHROME INTERACTING FACTOR3-LIKE5 (PIL5) that activates the expression of SOMNUS (SOM) in imbibed seeds. Therefore, the SOM promoter integrates ABA and light signals to mediate seed germination. Previously, we showed that ABI5 physically interacts with the BRASSINOSTEROID INSENSITIVE2 (BIN2) brassinosteroid (BR)-related protein kinases, and that the BIN2-ABI5 cascade mediates the antagonism between BRs and ABA during seed germination (Hu and Yu, 2014). Lim et al. (2013) showed that ABI3, ABI5, and DELLA proteins interact to integrate ABA and gibberellin (GA) signals to modulate the expression of a subset of high temperature-inducible genes, leading to the inhibition of seed germination.

The phytohormone jasmonate (JA) functions as a critical signaling molecule with roles in plant growth, development, and stress responses (Pauwels et al., 2010; Hu et al., 2013; Qi et al., 2015; Du et al., 2017). It is perceived by the F-box protein CORONATINE INSENSITIVE1 (COI1), which assembles in the SCFCOI1 complex.
protein complex that targets JASMONATE ZIM-DOMAIN (JAZ) proteins for degradation (Xu et al., 2002; Chini et al., 2007; Thines et al., 2007; Yan et al., 2009). The degradation of JAZ repressors alleviates the inhibition of downstream transcription factors, thus activating the transcriptional reprogramming of JA signaling (Chini et al., 2007; Thines et al., 2007; Pauwels et al., 2010). JAZ proteins have been shown to interact with multiple transcription factors to regulate diverse aspects of JA-regulated physiological processes, such as root growth, male fertility, anthocyanin accumulation, trichome development, senescence, and stress responses (Cheng et al., 2009; Niu et al., 2011; Fernández-Calvo et al., 2011; Kazan and Manners, 2013; Schweizer et al., 2013). In addition, JA signaling is involved in seed germination and interacts with ABA signaling. For example, methyl jasmonate (MeJA) has an inhibitory effect on the seed germination of several crops and Arabidopsis thaliana (Wilen et al., 1991; Krock et al., 2002; Preston et al., 2002; Norastehnia et al., 2007; Barrero et al., 2009; Dave et al., 2011; Dave et al., 2016). Several studies have provided evidence for crosstalk between ABA and JA signaling. For example, the genes encoding the ABA receptors PYRABACTIN RESISTANCE (PYR)/PYR1-LIKE (PYL)/REGULATORY COMPONENT OF ABSCISIC ACID RECEPTOR (PYL) 4 and PYL5 were found to be up-regulated by JA, and pyl4 and pyl5 knockout mutants were hypersensitive to JA (Lackman et al., 2011). Conversely, MYC2-overexpressing plants exhibited hypersensitivity to ABA (Abe et al., 2003; Lorenzo et al., 2004). The increased inhibition of germination in a line overexpressing a mutant MYC2 was associated with ABA hypersensitivity (Goossens et al., 2015). The results of those studies indicated that the ABA receptors PYL4/5 and the transcriptional factor MYC2 play vital roles in the crosstalk between JA and ABA. Further research showed that the E3 RING ligase KEG interacts with JAZ12 and regulates its stability (Pauwels et al., 2015). Another study provided evidence for the link between ABA and JA signaling through the direct interaction of the ABA receptor PYL6 with MYC2 (Aleman et al., 2016); the pyl6 mutant was more sensitive to the combination of JA and ABA than to ABA alone. Dave et al. (2011) found that 12-oxo-phytodienoic acid (OPDA, the precursor of JA) inhibited wild-type germination and acted with ABA to regulate seed germination in Arabidopsis. The abundance of ABI5 protein was increased by the inhibitory effect of OPDA and the combination of OPDA and ABA on seed germination. Ju et al. (2019) reported that JAZ proteins interact with ABI5 to modulate seed germination.

Although JA signaling has been implicated in seed germination in several species, the detailed molecular mechanism of how JA regulates those crucial physiological processes remains largely unknown. In this study, we found that exogenous application of 1 μM COR or 10 μM MeJA alone did not inhibit seed germination.
However, JA significantly delayed germination in the presence of ABA, showing that JA stimulates ABA signaling to delay seed germination. We discovered that the COI1/JAZ-mediated JA signaling pathway is positively involved in ABA-delayed seed germination. Mechanistic analyses revealed that several JAZ proteins physically interact with the ABA-responsive ABI3 transcription factor and repress ABI3/ABI5-mediated transcriptional activation of downstream targets. Further genetic analyses showed that JA activates ABA signaling to delay seed germination and requires functional ABI3 and ABI5. Overexpression of ABI3 and ABI5 simultaneously suppressed the ABA-insensitive phenotypes of coil mutants and JAZ-ΔJas (JAZ1, JAZ5, and JAZ8 lacking the Jas domain) plants. Our results provide a mechanistic understanding of how the integration of the JA and ABA signaling pathways is mediated by the JAZ-ABI3/ABI5 module during seed germination.

RESULTS

Exogenous JA Enhances ABA Signaling to Delay Seed Germination

To investigate the molecular mechanisms underlying JA regulation of seed germination, we first confirmed the regulatory effect of JA by evaluating the germination of wild-type (Col-0) Arabidopsis seeds on medium supplemented with MeJA and/or ABA. To avoid the effects of sucrose and/or nitrate on seed germination, we performed germination assays using water agar medium. Consistent with a previous study (Dave et al., 2011), the Col-0 seeds displayed similar percentages of germination and expanded cotyledons on medium with or without 10 μM MeJA (Supplemental Figure 1), confirming that 10 μM MeJA alone has no inhibitory effect on germination. We then analyzed the performance of Col-0 seeds on medium containing 10 μM MeJA with or without 0.5 μM ABA. Compared with seeds treated with MeJA or ABA alone, those treated with both MeJA and ABA showed much lower percentages of germination and expanded cotyledons (Supplemental Figure 1), suggesting that MeJA enhances ABA signaling during seed germination (Staswick et al., 1992; Ellis and Turner, 2002). To verify these observations, we used coronatine (COR), a JA analog that is structurally similar to the active conjugate jasmonate-isoleucine (JA–Ile). Whereas 1 μM COR did not inhibit germination and cotyledon expansion, Col-0 plants exhibited lower percentages of germination and of expanded cotyledons on both COR medium and on ABA medium (Figure 1A to 1C).

To corroborate the regulatory effect of COR on ABA signaling during seed germination, we used reverse transcription quantitative PCR (RT-qPCR) to determine the transcript levels of several well-characterized ABA-responsive genes, including ALCOHOL DEHYDROGENASE1 (ADH1), LATE EMBRYOGENESIS
ABUNDANT6 (EM6) and EMI (Finkelstein and Lynch, 2000; Carles et al., 2002; Lopez-Molina et al., 2002), in Col-0 germinating seeds treated with ABA and COR simultaneously and individually. The transcript levels of these genes were higher in seeds treated with both ABA and COR than in seeds treated with ABA or COR alone (Figure 1D). Taken together, these results demonstrate that exogenous JA enhances ABA signaling to delay seed germination.

COI1/JAZ-Mediated JA Pathway Is Positively Associated with ABA Signaling during Seed Germination

Having ascertained that JA interacts with ABA synergistically to delay seed germination, we then queried whether critical components of the JA signaling pathway participate in these ABA-regulated processes. The JA receptor F-box protein COI1 is a positive modulator of JA signaling (Xie et al., 1998; Yan et al., 2009). To investigate whether COI1 is involved in ABA responses during seed germination, we analyzed the loss-of-function mutant coil-2. Compared with Col-0 seeds, coil-2 seeds exhibited much higher percentages of germination and cotyledon greening on medium containing ABA (Figure 2A to 2D; Supplemental Figure 2A; Supplemental Figure 3). As shown in Supplemental Figure 3, sucrose delayed seed germination, and nitrate enhanced this process. Then, in comparison with seeds exposed to both ABA and nitrate medium, the germination of wild type was similar to coil-2 and coil-16 mutant, indicating that nitrate had an effect on ABA signaling during seed germination. Similarly, there were no differences in the percentages of germination and expanded cotyledon between the coil mutants with Col-0 on the presence of both ABA and sucrose, suggesting that sucrose participated in ABA signaling to delay seed germination (Supplemental Figure 3). These results suggested that the phenotype of coil mutants is conditioned by the absence of nitrate or sucrose. Therefore, we used water agar medium to avoid the effects of sucrose and/or nitrate. In parallel experiments, compared with Col-0, in our hands and experimental conditions, coil-16 (another loss-of-function mutant of COI1) showed reduced sensitivity to ABA during seed germination and post-germinative growth (Figure 2A to 2D; Supplemental Figure 2A; Supplemental Figure 3). To further confirm that disruption of COI1 is responsible for the ABA-insensitive phenotypes of coil-2 and coil-16, we introduced the full-length COI1 cDNA under the control of the Cauliflower mosaic virus (CaMV) 35S promoter into coil-2 and coil-16 mutant plants (coil-2 COI1 and coil-16 COI1). Expression of COI1 in coil-2 and coil-16 fully complemented the MeJA-insensitive root elongation phenotype (Supplemental Figure 4). Moreover, coil-2 COI1 and coil-16 COI1 plants had similar responses to those of Col-0 to ABA
during seed germination (Supplemental Figure 5). To avoid unspecific effects due to misexpression of \textit{COII} under the control of the 35S promoter, we also introduced \textit{COII} into the \textit{coil-2} and \textit{coil-16} backgrounds under the control of its native promoter (\textit{coil-2}:/\textit{COII}pro:\textit{COII} and \textit{coil-16}:/\textit{COII}pro:\textit{COII}). The \textit{coil-2}:/\textit{COII}pro:\textit{COII} and \textit{coil-16}:/\textit{COII}pro:\textit{COII} seeds consistently exhibited percentages of germination and cotyledon greening similar to those of Col-0 under ABA treatment (Supplemental Figure 5). These results suggest that endogenous JA perception or signaling positively mediates ABA responses during seed germination and early seedling growth.

To further confirm the involvement of JA signaling components in ABA responses, we investigated whether genetic manipulation of JAZ proteins, which are the substrates of the SCF\textsuperscript{COII} E3 ubiquitin ligase and repressors of JA signaling (Chini \textit{et al.}, 2007; Thines \textit{et al.}, 2007), affects ABA-repressed seed germination. Phenotypic analysis showed that the \textit{jazQ} mutant (with T-DNA insertion mutations in \textit{JAZ1}/3/4/9/10 genes; Campos \textit{et al.}, 2016) was more sensitive than Col-0 to ABA during seed germination and seedling establishment (Figure 2A to 2D). Consistently, compared with seeds of wild-type, seeds of the \textit{jaz} decuple mutant (\textit{jazD}; defective in \textit{JAZ1}-7,-9, -10 and -13; Guo \textit{et al.}, 2018) were more sensitive to ABA (Supplemental Figure 2B). To further confirm this possibility, we expressed the full-length cDNA of \textit{JAZ1} under the control of the CaMV 35S promoter (\textit{JAZ1-OE}). However, there were no differences in the percentages of germination and cotyledon greening between the homozygous \textit{JAZ1-OE} plants and Col-0 plants with or without ABA. Similar results were obtained for \textit{JAZ5-OE} and \textit{JAZ8-OE} seeds (Supplemental Figure 6A and 6B).

Next, we generated transgenic plants overexpressing \textit{JAZ1}, \textit{JAZ5}, or \textit{JAZ8} with a deleted \textit{Jas} domain (\textit{JAZ1-ΔJas}, \textit{JAZ5-ΔJas} or \textit{JAZ8-ΔJas}). The \textit{JAZ1-ΔJas} plants have been shown to accumulate large amounts of the JAZ1 repressor and are insensitive to JA treatment (Chini \textit{et al.}, 2007; Thines \textit{et al.}, 2007). As \textit{JAZ1-ΔJas} homozygous plants are completely sterile, seeds from heterozygous plants were used in the germination assays. When incubated on medium containing 0.5 µM ABA, \textit{JAZ1-ΔJas} heterozygous (F1 progeny from a cross with wild type) lines displayed much higher percentages of germination and cotyledon greening in comparison with Col-0 (Figure 2A to 2D). Compared with wild-type seeds, those of \textit{JAZ5-ΔJas} or \textit{JAZ8-ΔJas} were also less insensitive to ABA (Figure 2A to 2D). Among the \textit{JAZ1-ΔJas} heterozygous seeds germinated on medium containing 0.75 µM ABA for 3.5 d, most of the seedlings with green cotyledons were \textit{JAZ1-ΔJas} transgenic plants (\textit{n} = 80, 91.7%). The transcript levels of several ABA-responsive genes were consistently lower in \textit{coil-2} than in Col-0 in response to ABA (Figure 2E), but
were higher in the jazQ mutant than in Col-0 (Figure 2E).

Taken together, these findings provide further evidence that the endogenous JA pathway is positively involved in ABA signaling during seed germination.

**JAZ Repressors Physically Interact with ABI3**

Recent studies have revealed that JAZ repressors physically interact with numerous transcription factors to modulate different aspects of JA signaling processes (Qi et al., 2011; Song et al., 2011; Zhu et al., 2011; Hu et al., 2013; Jiang et al., 2014). To test the interactions between JAZ1 and transcription factors, the full-length coding sequence (CDS) of JAZ1 was fused to pGBK-T7 to construct a bait vector, which was transformed into the yeast strain Y2HGOLD (Clontech, Palo Alto, CA, USA). Yeast screening was performed as described previously (Hu et al., 2013), and the cDNA library was acquired from Clontech (Cat. no. 630487). After screening, more than 10 independent colonies were isolated. Among these positive clones, clones encoding ABI5, TOE1, TOE2, MYC2, and NINJA were frequently sequenced. ABI3 was sequenced seven times through this screening procedure.

To further confirm the interaction between JAZs and ABI3, we generated the full-length ABI3 in the bait vector pGBKT7 (BD-ABI3), and each JAZ (Supplemental Data Set 1) gene was introduced into the Gal4 activation domain of the pGAD-T7 vector to act as prey (AD-JAZ). The bait and prey vectors were co-transformed into yeast. Previous studies have reported the interactional relationships of ABI4-MED18, JAZ4-WRKY57, JAZ8-WRKY57, IAA29-WRKY57 (Lai et al., 2014; Jiang et al., 2014; Supplemental Figure 7). Physical interactions were detected between ABI3 and JAZ1, JAZ5, and JAZ8 (splicing variant of JAZ1, JAZ4, JAZ5, and JAZ8 was AT1G19180.1, AT1G48500.1, AT1G17380.1, and AT1G30135.1, respectively; Figure 3A and 3B; Supplemental Figure 7; Supplemental Data Set 1). ABI4 is a well-known transcription factor that positively modulates ABA responses during seed germination. However, no interaction was detected between ABI4 and JAZ (Figure 3B), suggesting that JAZ proteins involved in the ABA response interact with ABI3 but not ABI4.

These interactions between JAZ proteins and ABI3 were confirmed in plant cells using bimolecular florescence complementation (BiFC) and co-immunoprecipitation (Co-IP) assays. For the BiFC assays, each JAZ was fused to the N-terminal yellow fluorescent protein (YFP) fragment to generate JAZ-nYFP, while ABI3 and ABI4 were each fused to the C-terminal fragment of YFP (cYFP) to produce ABI3-cYFP and ABI4-cYFP, respectively. When ABI3-cYFP was transiently co-expressed with JAZ1-nYFP, JAZ5-nYFP, or
JAZ8-nYFP in leaf cells of *Nicotiana benthamiana*, reconstituted YFP fluorescence was observed in the nucleus, as revealed by staining with 4',6-diamidino-2-phenylindole (DAPI) (Figure 3C). As the negative control, ABI3-cYFP (or ABI4-cYFP) was co-expressed with JAZ4-nYFP and accumulated (or JAZ1-nYFP, JAZ5-nYFP and JAZ8-nYFP) in *N. benthamiana* leaf cells and no fluorescence was detected (Figure 3C; Supplemental Figure 8). For the Co-IP assays, the 3MYC-ABI3 construct was co-expressed with HA-JAZ1, HA-JAZ5, or HA-GFP (green fluorescent protein) in *N. benthamiana* leaves. Cell extracts were immunopurified with an anti-MYC antibody, and the resultant immunoprecipitants were separated by SDS-PAGE and probed with an anti-HA antibody. The HA-JAZ1 and HA-JAZ5 fusion proteins were detected in cell extracts when they were co-expressed with 3MYC-ABI3 in *N. benthamiana* leaves (Figure 3D and 3E; Supplemental Figure 9). The control HA-GFP fusion protein was not detected in cell extracts when it was co-expressed with 3MYC-ABI3 (Figure 3D and 3E; Supplemental Figure 9). These results show that JAZ proteins physically interact with ABI3 in planta.

Having ascertained that JAZ proteins physically interact with ABI3, we wondered whether JAZs are involved in ABI3-mediated ABA signaling during seed germination. To test this possibility, we further analyzed the properties of JAZs in more detail. To examine JAZ expression in different tissues, we analyzed its transcript levels by RT-qPCR. Interestingly, relatively high transcript levels of *JAZ1*, *JAZ5*, and *JAZ8* were detected in dry seeds (Supplemental Figure 6C to 6E), indicating that JAZ proteins play roles in seed germination.

**The N-Terminal Fragment of ABI3 Interacts with the Middle Region of JAZ that Contains ZIM Domain**

To examine which regions of ABI3 are required to interact with JAZ proteins, we performed directed yeast two-hybrid analyses. ABI3 was divided into the N-terminal region (BD-ABI3-N) and the C-terminal region containing the B2 and B3 domains (BD-ABI3-C; Figure 4A). Deletion of the C-terminal residues of ABI3 (BD-ABI3-C) did not affect its interactions with JAZ1 and JAZ5 (Supplemental Data Set 1); however, deletion of the N-terminal fragment of ABI3 completely abolished its interactions with these proteins (Figure 4A). Therefore, the N-terminal region of ABI3 is required for its interactions with JAZ1 and JAZ5.

We used the same approach to identify the domains of JAZ proteins that are required for their interactions with ABI3. JAZ1 and JAZ8 (Supplemental Data Set 1) were divided into different truncated versions and then independently fused with the Gal4-activation domain to generate prey vectors (Figure 4B and 4C). The
potential interactions between these derivatives and ABI3 were then assayed using the yeast two-hybrid system. Deletion of the C-terminal 133 residues of JAZ1 (AD-JAZ1 1–120) eliminated its interaction with ABI3, while deleting its N-terminal fragment (AD-JAZ1 121–253) had no effect (Figure 4B). Further mapping revealed that the middle region of JAZ1 containing the ZIM domain was required for the JAZ1–ABI3 interaction. Likewise, the middle region of JAZ8 containing the ZIM domain was essential for the JAZ8–ABI3 interaction (Figure 4C).

**JA Activation of ABA Signaling Requires Functional ABI3 and ABI5**

The ABI3 and ABI5 transcription factors are critical regulators of ABA signaling during seed germination and early seedling growth. Phenotypic analyses have indicated that the loss-of-function mutants abi3 and abi5 are insensitive to ABA (Koornneef et al., 1984; Giraudat et al., 1992; Finkelstein, 1994; Finkelstein and Lynch, 2000). Given that several JAZ repressors of the JA signaling pathway physically associate with ABI3 and ABI5 (Figure 3; Ju et al., 2019), we hypothesized that JA stimulates ABA responses during seed germination and that this requires ABI3 and/or ABI5. To test this possibility, we determined the percentages of germination and expanded cotyledons of abi3 (abi3-1 mutant backcrossed six times with Col-0 wild type) and abi5 (abi5-1 mutant backcrossed six times with Col-0 wild type) seeds on media containing ABA and/or COR. Consistent with previous studies (Koornneef et al., 1984; Giraudat et al., 1992; Finkelstein, 1994), abi3 and abi5 seeds germinated and grew faster than Col-0 seeds on ABA-containing medium, and the germination percentages of abi3-8 and abi5-7 were higher than that of Col-0 (Figure 5). Furthermore, compared with Ler and Ws, the abi3-1 and abi5-1 single mutants exhibited much higher percentages of germination and expanded cotyledons on media containing ABA, COR, or both ABA and COR (Figure 5). Because ABI5 physically interacts with ABI3 (Nakamura et al., 2001), we generated an abi5 abi3 double mutant (backcrossed abi5 crossed with backcrossed abi3) and analyzed its response to ABA and COR. Compared with abi3, abi5, and Col-0, a higher percentages of the double mutant had expanded cotyledons in the presence of both ABA and COR (Figure 5). These observations indicate that JA activates ABA signaling to delay seed germination and that this requires functional ABI3 and ABI5.

The ABI3 and ABI5 transcription factors are recognized as key regulators of seed dormancy in *Arabidopsis* (Bentsink and Koornneef, 2008). Because JAZ proteins physically interact with ABI3, we questioned whether COI1 and JAZ are also involved in modulating seed dormancy. To test this possibility, we examined the fresh mature siliques of coi1-2 and *JAZ1-ΔJas* plants after 5 d on water-saturated filter
paper at 22°C without stratification. The seed dormancy levels of coi1-2 and JAZ1-ΔJas were similar to that of wild type (Supplemental Figure 10). However, seed dormancy was significantly reduced in the abi3 mutant, consistent with previous studies (Ooms et al., 1993; Finkelstein et al., 1994; Liu et al., 2013). These results suggest that the COI1/JAZ-mediated JA pathway does not regulate seed dormancy in Arabidopsis.

JAZ Proteins Repress the Transcriptional Activation Roles of ABI3 and ABI5

Because JAZ proteins directly interact with ABI3 (Figure 3) and ABI5 (Ju et al., 2019), we investigated whether JAZ proteins interfere with the transcriptional activation roles of ABI3 and/or ABI5. To this end, we used dual luciferase (LUC) reporter assays to examine the effects of JAZ1, JAZ5, or JAZ8 on the transcriptional functions of ABI3 or ABI5 in Arabidopsis mesophyll protoplasts (Yoo et al., 2007). Because EM1, EM6, and ABI5 are downstream targets of ABI5 and ABI3 (Lopez-Molina and Chua, 2000; Nakamura et al., 2001; Carles et al., 2002; Lopez-Molina et al., 2002), their promoters were fused to the LUC gene to generate reporters (Figure 6A). The effector constructs contained ABI5, ABI3, JAZ, or GFP driven by the CaMV 35S promoter (Figure 6A). ABI3 expression activated LUC expression driven by the EM6 or ABI5 promoter with ABA treatment, compared with the expression of the GFP control (Figure 6B and 6C). Moreover, compared with expression of ABI3 alone, co-expression of JAZ1 or JAZ5 with ABI3 repressed the LUC expression level (Figure 6B and 6C). Similarly, ABI5 expression dramatically activated LUC expression driven by the EM6 or EM1 promoter in the presence of ABA in comparison with expression of the GFP control (Figure 6D and 6E). However, compared with expression of ABI5 alone, co-expression of ABI5 with JAZ8 suppressed LUC expression (Figure 6D and 6E). These results demonstrate that JAZ proteins repress the transcriptional activation roles of ABI3 and ABI5, thereby preventing activation of their downstream targets.

Overexpression of ABI3 and ABI5 Simultaneously Suppresses ABA-Insensitive Phenotypes of coi1-2 and JAZ8-ΔJas

Having ascertained that JAZ proteins suppress the transcriptional roles of ABI3 and ABI5 (Figure 6), we queried whether overexpression of ABI3 and/or ABI5 could overcome JAZ repression and rescue the ABA-insensitive phenotypes of the coi1 mutants in which JAZ proteins are highly accumulated (Chini et al., 2007; Thines et al., 2007). To test this hypothesis, we generated transgenic plants overexpressing ABI3 (ABI3-OE). The RT-qPCR analyses showed that the ABI3-OE lines constitutively expressed ABI3 at high
levels (Supplemental Figure 11A). Then, we overexpressed ABI3-OE-5 in the coi1-2 and coi1-16 backgrounds to produce coi1-2 ABI3-OE and coi1-16 ABI3-OE plants. Compared with Col-0, the coi1-2 ABI3-OE and coi1-16 ABI3-OE plants displayed higher percentages of germination and expanded cotyledons (Figure 7; Supplemental Figure 12A and 12B). However, coi1-2 ABI3-OE and coi1-16 ABI3-OE seeds were more sensitive to ABA and COR during germination than were coi1-2 and coi1-16 seeds. Because ABI5 and ABI3 positively modulate ABA signaling during seed germination, we simultaneously overexpressed ABI3 and ABI5 by overexpressing ABI5 (the validated overexpression line ABI5-OE-8; Supplemental Figure 11B) in the coi1-2 ABI3-OE and coi1-16 ABI3-OE backgrounds (coi1-2 ABI3-OE ABI5-OE and coi1-16 ABI3-OE ABI5-OE). Compared with coi1-2 and coi1-16, the coi1-2 ABI3-OE ABI5-OE and coi1-16 ABI3-OE ABI5-OE lines exhibited significantly lower percentages of germination and expanded cotyledons when grown on medium supplemented with both ABA and COR (Figure 7; Supplemental Figure 12A and 12B). Thus, simultaneous overexpression of ABI3 and ABI5 suppressed the ABA-insensitive phenotypes of the coi1-2 and coi1-16 mutants.

To further understand the regulatory relationship between JAZ repressors and ABI3 or ABI5, we determined whether overexpression of ABI3 and/or ABI5 could rescue the ABA-insensitive phenotypes of JAZ-ΔJas transgenic plants in which JAZ proteins are also highly accumulated (Chini et al., 2007; Thines et al., 2007; Figure 2). To test this hypothesis, we generated JAZ8-ΔJas plants and then crossed them with ABI3-OE-5 to create JAZ8-ΔJas ABI3-OE plants. The percentages of germination and expanded cotyledons were lower in JAZ8-ΔJas ABI3-OE lines than in JAZ8-ΔJas lines in response to both ABA and COR (Figure 8). Moreover, overexpression of ABI5 (ABI5-OE-8) in the JAZ8-ΔJas ABI3-OE background rendered the transgenic lines much more sensitive to ABA and COR during seed germination (Figure 8). Similar data were also obtained when ABI3 and ABI5 were expressed in the JAZ5-ΔJas background (Supplemental Figure 12C and 12D). Thus, simultaneous overexpression of ABI3 and ABI5 suppressed the ABA-insensitive phenotypes of JAZ-ΔJas plants. These findings support the idea that ABI3/ABI5 and JAZ proteins modulate ABA and JA signaling during seed germination.

Because ABI3 interacts with JAZ, the jazQ and jazD mutants were sensitive to ABA during seed germination, and overexpression of ABI3 and ABI5 suppressed the ABA-insensitivity of coi1-2 and coi1-16. We speculated that signaling mediated by the receptor COI is required to induce ABA responses. Therefore, we performed RNA-seq experiments to profile the transcriptomes of coi1-2, coi1-16, and Col-0 with or without ABA treatments. Total RNAs were isolated from seeds treated with or without ABA. We sequenced
each sample on the Illumina HiSeq X Ten platform. Sequenced reads were trimmed to remove adaptor sequences, and low-complexity or low-quality sequences were removed using Trimmomatic (0.36) with the following parameters: LEADING:3 TRAILING:3 SLIDINGWINDOW: 4:15 MINLEN:50. Clean reads were mapped to the genome of TAIR10.1_NCBI_year_2018 using Hisat2 (2.2.1.0) with default parameters. The read counts of each gene were obtained using htseq-count (0.9.1).

Under normal conditions (no ABA treatment), compared with wild type, coi1-2 and coi1-16 had 84 up-regulated genes and 160 down-regulated genes (Figure 9A). We classified the genes co-regulated by COI1 under normal conditions and found that they encode proteins with various biological functions, including seed oil body biogenesis, lipid storage, and response to ABA (Figure 9B). Under ABA treatment, 106 genes were up-regulated and 210 genes were down-regulated in both coi1-2 and coi1-16, compared with wild type (Figure 9C).

To identify the genes responsible for the ABA-insensitive phenotypes of the coi1-2 and coi1-16 mutants, we examined the overlap between co-regulated genes in the two mutants and those regulated by ABA signaling in the wild type. Among all the co-regulated genes in coi1-2 and coi1-16, 16 were responsive to ABA (Figure 9D, Supplemental Data Set 2). We then compared the expression levels of these co-regulated and ABA-responsive genes under normal and ABA-treatment conditions. A heat map analysis indicated that the expression levels of most co-regulated genes were lower in coi1-2 and coi1-16 than in the wild type under both normal and ABA-treatment conditions (Figure 9E).

We used reverse transcription-quantitative PCR to confirm the transcript levels of selected genes from the set of co-regulated and ABA-responsive genes, including CRU1, CRU3, LEA and RAB18. The analyses revealed that those genes’ transcript levels were lower in coi1-2 and coi1-16 than in the wild type (Figure 9F).

Ju et al. (2019) found that ABA treatments could promote jasmonate biosynthesis. Consistent with this, qRT-PCR confirmed that several JA biosynthesis genes (ACX1, AOC3, KAT5, and OPR3) were up-regulated by ABA treatment in wild type (Supplemental Figure 13E to 13H). The transcription of ABA biosynthesis genes (ABA1, ABA2, ABA3, and NCED3) also was up-regulated by exogenous ABA (Supplemental Figure 13A to 13D). However, under ABA treatment, the ABA induction of these ABA biosynthetic genes was reduced in coi1-2 and coi1-16 compared with wild type (Supplemental Figure 13I to 13L). Similarly, the transcript levels of these JA biosynthetic genes were decreased in the coi1-2 and coi1-16 mutants (Supplemental Figure 13M to 13P). These results indicate that ABA treatment promotes JA and ABA
biosynthesis, while ABA-induced JA and ABA biosynthesis is reduced in the coi1-2 and coi1-16 mutants, and increased in the jazQ mutant (Supplemental Figure 13).

Discussion

The phytohormone JA is ubiquitous in the plant kingdom and regulates multiple physiological aspects of plant development, growth, and stress responses. Several studies have highlighted the involvement of JA in seed germination, and have suggested that it may interact with ABA to mediate these processes (Wilen et al., 1991; Krock et al., 2002; Preston et al., 2002; Norastehnia et al., 2007; Barrero et al., 2009; Dave et al., 2011). However, the detailed molecular mechanisms underlying JA regulation of seed germination and its crosstalk with ABA remain elusive. In this study, we further investigated the regulatory role of JA in modulating ABA responses during seed germination and subsequent post-germinative growth. Consistent with previous studies, our results demonstrate that exogenous JA (COR) has a positive role in activating ABA responses to delay seed germination. Col-0 seeds displayed low percentages of germination and of expanded cotyledons in the presence of both ABA and JA (Figure 1; Supplemental Figure 1), while lines with blocked endogenous JA perception or signaling were less sensitive to ABA during seed germination (Figure 2; Supplemental Figure 2). Based on these findings, we conclude that the JA signal activates ABA responses to delay seed germination in Arabidopsis.

Intriguingly, we found that mutation of COI1 leads to decreased ABA signaling during seed germination. This finding is in disagreement with the results of two earlier studies (Ellis and Turner, 2002; Fernández-Arbaizar et al., 2012). In those studies, the authors analyzed one of the same coi1 alleles that we used, coi1-16, and found that it was hypersensitive to ABA in comparison with Col-0. To verify the role of COI1 in mediating the ABA response, we also investigated another coi1 allele, coi1-2, in the presence of ABA. Like our coi1-16 mutant, coi1-2 seeds also exhibited much higher percentages of germination and greening cotyledons than Col-0 seeds on medium containing ABA (Figure 2; Supplemental Figure 2A). Consistent with this phenotype, the induced expression levels of several well-characterized ABA-responsive genes were lower in coi1-2 than in wild type (Figure 2E). More importantly, to further corroborate that mutation of COI1 is responsible for the ABA-insensitive phenotypes of coi1-2 and coi1-16, we expressed the full-length COI1 gene driven by its native promoter or the CaMV 35S promoter in the coi1-2 and coi1-16 mutant backgrounds. As expected, expression of full-length COI1 in the coi1-2 or coi1-16 background resulted in the mutant plants responding similarly to Col-0 under ABA treatment (Supplemental
Figure 5). Moreover, on medium containing ABA, the germination and greening percentages were much higher in heterozygous JAZ-ΔJas plants, which accumulate higher levels of JAZ proteins, than in Col-0 (Figure 2 and 8), similar to the effects seen for the coi1 mutants. In contrast, the germination rates were lower in the jazQ and jazD mutants than in wild type under ABA treatment. Therefore, our results show that jasmonate synergistically regulates ABA signaling during seed germination.

The coi1-16 phenotype and germination rates of seeds may be dependent on our experimental conditions because many external factors, including light, humidity, temperature, and availability of nitrogenous compounds are known to affect ABA responses during seed germination. For example, the light intensity under long-light conditions is one factor that differs among various laboratories. The germination media also differ among different studies. Some studies conducted germination assays on water agar medium to avoid the effects of nitrate and sucrose (this study; Dave et al., 2011), whereas different media were used in earlier studies (Ellis and Turner, 2002; Fernández-Arbaizar et al., 2012). Differences in these factors may account for the inconsistency of the germination rates and the coi1-16 phenotype in response to ABA among different studies. In our hands and experimental conditions, the ABA sensitivity of coi1 mutants was changed under conditions where exogenous nitrate or sucrose was present (Supplemental Figure 3). The data suggested that coi1 mutants may have different ABA sensitivity according to the imbibition conditions. Furthermore, our germination data were not identical on half-strength MS and water agar medium, but seeds displayed a similar percentage of expanded cotyledons in both media.

Recently, several studies have provided evidence of crosstalk between JA and ABA signaling. The results of those studies supported that JA interacts with ABA signaling to regulate physiological process (Lackman et al., 2011; Pauwels et al., 2015). Consistent with the results of Dave et al. (2011), our results show that COR acts along with ABA to regulate seed germination in Arabidopsis. Despite recent advances in research on the link between JA and ABA signaling, the exact mechanisms underlying the crosstalk between JA signaling and other developmental processes and signaling pathways remain to be elucidated. Characterization of the physical interactions between JAZ proteins and the critical components of other signaling pathways may shed light on the molecular basis of the regulation of the JA signal transduction network.

In this study, we found that JAZ proteins physically associate with an important transcription factor in ABA signaling, ABI3, and further demonstrated that those JAZ proteins repress the transcriptional activation functions of ABI3 and ABI5 (Figure 3 and 6). In our analyses, these JAZ repressors specifically interacted
with ABI3 and ABI5, but did not form complexes with ABI4, an APETALA2 (AP2) family transcription factor that positively modulates ABA signaling in yeast (Figure 3; Ju et al., 2019). Further analyses revealed that the ZIM domain of JAZ is required for interactions between JAZ and ABI3 (Figure 4). Deletion of the C-terminal 133 residues of JAZ that contain the ZIM domain eliminated the interaction between JAZ with ABI3, while deletion of the N-terminal region (121–253 residues) did not affect this interaction. Interestingly, this finding differs from that of several previous studies, which showed that the Jas domain is essential for most of the interactions between JAZ repressors and downstream transcription factors (Pauwels and Goossens, 2011). Recently, Zhai et al. (2015) found that the deletion of a 53-amino acid residue N-terminal region (including the ZIM domain) of JAZ1 eliminated its interaction with TOE1, indicating that this region is required for the JAZ–TOE1 interaction. Other studies also reported that the region containing the ZIM domain is vital for interactions with other proteins (Cheng et al., 2011; Song et al., 2011; Jiang et al., 2014). Importantly, the ZIM domain of most JAZ proteins can recruit TPL or TPR proteins indirectly through the ethylene-response factor amphiphilic repression (EAR) motif-containing NOVEL INTERACTOR OF JAZ (NINJA) adaptor protein to repress JA responses (Shyu et al., 2012; Thatcher et al., 2016; Howe et al., 2018). In addition, sequence variations in the hypervariable region of the degron affect JAZ stability and JA-regulated physiological responses. JAZ8-mediated repression depends on an EAR motif at the JAZ8 N terminus, which binds the corepressor TOPLESS and represses its transcriptional activation function (Shyu et al., 2012). Therefore, different domains of JAZ proteins interact with different proteins. In future studies, the identification of JAZ-associated transcription factors and further mapping of specific JAZ domains or residues required for these interactions may enhance our understanding of JAZ-regulated targets.

The transcription factors ABI3 and ABI5, which are mainly expressed in seeds and are strongly induced by ABA, play critical roles in modulating ABA responses to suppress seed germination and early seedling growth (Finkelstein, 1994; Finkelstein and Lynch, 2000; Lopez-Molina and Chua, 2000; Lopez-Molina et al., 2001, 2002; Suzuki et al., 2001; Brocard et al., 2002; Finkelstein et al., 2005; Bedi et al., 2016). Previous studies have shown that the loss-of-function mutants abi3 and abi5 are insensitive to ABA treatment (Koornneef et al., 1984; Giraudat et al., 1992; Finkelstein, 1994; Finkelstein and Lynch, 2000). Interestingly, our phenotypic analyses showed that abi3, abi5, and an abi5 abi3 double mutant also exhibited higher percentages of expanded cotyledons than Col-0 on medium containing ABA and COR (Figure 5). These findings demonstrate that JA stimulates ABA signaling to delay seed germination and post-germinative
growth, and that this requires functional ABI3 and ABI5. We found that overexpression of ABI3 and ABI5 simultaneously suppresses the ABA-insensitive phenotypes of coi1-2 and JAZ8-ΔJas mutants (Figure 7 and 8). This finding further supports the hypothesis that ABI3/ABI5 and JAZ proteins modulate ABA and JA signaling during seed germination. Together, the results of our biochemical and genetic analyses reveal a previously unknown signaling module in which JAZ repressors in the JA pathway directly modulate ABA-responsive ABI3 and ABI5 transcription factors to integrate JA and ABA signals during seed germination and post-germinative growth.

In a recent study (Pan et al., 2018), two VQ (containing the conserved FxxxVQxxTG motif) proteins, VQ18 and VQ26, were shown to physically associate with ABI5 and interfere with its transcriptional role. As with the JAZ proteins in the present study, VQ18 and VQ26 do not mediate seed dormancy but, rather, negatively modulate ABA signaling during seed germination and early seedling establishment. However, Liu et al. (2013) found that two transcription factors of auxin signaling, AUXIN RESPONSE FACTOR 10 (ARF10) and ARF16, positively regulate ABI3 expression and promote both seed dormancy and ABA signaling during seed germination. Furthermore, Vaistij et al. (2013) revealed that MOTHER-OF-FT-ANDTFL1 (MFT), a member of the phosphatidyl ethanolamine-binding protein (PEBP) family, regulates ABI5 expression to stimulate seed dormancy but represses ABA responses during seed germination. Together, the results of these studies show that the components that regulate or interact with ABI3 and/or ABI5 may have distinct roles in regulating seed dormancy or ABA signaling. Further research is required to dissect the crucial regulators of ABI3/ABI5 and elucidate the exact mechanisms underlying ABI3/ABI5-mediated seed dormancy and regulation of the ABA signaling network.

JA is a hormone induced by pathogen attack, wounding, and some abiotic stresses (Kazan, 2015; Howe et al., 2018), and ABA also is induced by several abiotic stresses. It is easy to imagine that when seeds are exposed to soil, environmental factors induce the accumulation of JA and ABA. In plants, ABA and JA are two important hormones that regulate diverse aspects of plant developmental and stress responses. Some observations have revealed clues about the crosstalk between the ABA and JA signaling pathways. For instance, ABA induces the expression of PLIP2 and PLIP3, which may participate in JA biosynthesis (Wang et al., 2018). Another study showed that ABA does indeed promote JA biosynthesis (Ju et al., 2019). Consistent with these studies, we found that ABA treatments enhanced JA and ABA biosynthesis. Then, the accumulated JA may cooperate with ABA to delay seed germination for avoiding a bad environment. Our results reveal some of the mechanisms underlying the crosstalk between ABA and JA: JAZ proteins
physically interact with the transcription factors ABI3/ABI5 and repress their activities to delay seed germination. As shown in Figure 10, we propose the following working model: under unfavorable growth conditions, JAZ proteins are degraded, then ABI3 and ABI5 are released from JAZ-mediated repression to activate ABA signaling; simultaneously, the transcription factors ABI3 and ABI5 are activated by ABA, thereby delaying the process of seed germination.

METHODS

Plant Materials and Growth Conditions

Most of the *A. thaliana* lines used in this study were in the Columbia (Col-0) background. Additional *Arabidopsis* accessions used were Wassilewskija (Ws), and Landsberg erecta (Ler). The *abi3-1* (Jiang et al., 2009), *abi3-8* (Lin et al., 2020), *abi5-1* (Finkelstein, 1994; Finkelstein and Lynch, 2000), and *abi5-7* (Zhou et al., 2015) mutants have been described previously. Seeds of *abi3* and *abi5* were obtained from Dapeng Zhang (Tsinghua University, Beijing, IN, China), seeds overexpressing *ABI5* (Chen et al., 2012; Hu and Yu, 2014) were obtained from Chuanyou Li (Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, Beijing, IN, China), and *coi1-2* was kindly provided by Zhixiang Chen (Purdue University, West Lafayette, IN, USA) and Daoxin Xie (Tsinghua University, Beijing, IN, China). The *coi1-16* mutant was obtained from the ABRC.

The seeds were surface-sterilized in 20% (v/v) bleach for 15 min before being sown on water agar medium supplemented with 0.7% (w/v) agar, and were then maintained at 4°C for 3 d. After 7 d, the seedlings were transferred to soil. Then, plants were grown in chambers under long-day conditions, consisting of 14-h light (120 μE m⁻²s⁻¹ at 22°C, white fluorescent bulbs, full wavelength of light) and 10-h darkness (0 μE m⁻²s⁻¹ at 19°C), or short-day conditions, consisting of 8-h light (120 μE m⁻²s⁻¹ at 22°C) and 16-h darkness (0 μE m⁻²s⁻¹ at 19°C). *N. benthamiana* plants were grown in a controlled environment cabinet under long-day conditions (120 μE m⁻²s⁻¹) at 22°C for BiFC and Co-IP assays.

Chemicals and Enzymes

The plant hormones ABA, MeJA, and COR were purchased from Sigma (St Louis, MO, USA), and the 26S proteasome inhibitor MG132 was purchased from Sigma-Aldrich (St Louis, MO, USA). The Taq DNA polymerase was purchased from TaKaRa Biotechnology Co. Ltd (Dalian, China), and other common chemicals were purchased from Shanghai Sangon Biotechnology Co. Ltd. (Shanghai, China).
Generation of Transgenic Arabidopsis Lines

To generate JAZ- and ABI3-OE transgenic lines, the full-length cDNA of ABI3 was cloned into the binary vector pOCA30 (Pan et al., 2018) in the sense orientation behind the CaMV 35S promoter (Hu et al., 2013). The JAZ1, JAZ5, or JAZ8 cDNAs had the Jas domain deleted (Thines et al., 2007), and each was independently cloned into the binary vector pOCA30 in the sense orientation behind the CaMV 35S promoter. The heterozygous JAZ-ΔJas plants were the F1 progeny of JAZ-ΔJas crossed with wild type, and the seeds of this cross were used in a germination assay. The primers and restriction enzyme sites used to amplify sequences and generate vectors are listed in Supplemental Table 1.

The coi1-2 ABI3-OE (or coi1-16 ABI3-OE) and JAZ8-ΔJas ABI3-OE (JAZ5-ΔJas ABI3-OE) plants were generated by crossing an appropriate transgenic plant with ABI3-OE-5. The results of those crosses were respectively crossed with ABI5-OE-8 plants to produce coi1-2 ABI3-OE ABI5-OE (or coi1-16 ABI3-OE ABI5-OE) and JAZ8-ΔJas ABI3-OE ABI5-OE (JAZ5-ΔJas ABI3-OE ABI5-OE) plants.

Because the abi5-1 mutant was in the Ws background, we backcrossed it with the Col-0 wild type six times before use. Similarly, the abi3-1 mutant (in the Ler background) was backcrossed with the Col-0 wild type six times before use. The abi5 abi3 double mutant was generated by crossing the backcrossed abi5 mutant with the backcrossed abi3 mutant.

Seed Germination and Dormancy Assays

Plants of each genotype were grown side by side under the same conditions. Seeds of each genotype were harvested from independent plants and stored at the same time, and pooled before germination assays. We scored germination as radicle emergence from the seed coat and endosperm, and we also recorded cotyledon opening and cotyledon expansion (Dave et al., 2011; Piskurewicz and Lopez-Molina, 2016). For the stratification treatment, seeds were stratified at 4°C in the dark for 3 d.

To examine the phenotypes of germinated materials, every batch of seeds was grown on water agar medium supplemented with or without ABA and/or COR for the indicated periods in long-day conditions (14-h light and 10-h dark). Every batch of dry mature seeds for each genotype was pooled from at least 15 independent plants (each figure represents data from seeds that matured and were harvested at the same time). Average values were calculated from three biological replicates and compared using statistical tests to determine the significance of differences. For every biological replicate, we tested the seeds from the same
batch at least three times as technical replicates.

For seed dormancy analysis, for each genotype, we examined fresh mature siliques on plants grown under the same conditions and maturing at the same time after 5 d of growth on water-saturated filter paper at 22°C without stratification (Liu et al., 2013).

**Expression Analyses**

Seeds were placed on media containing hormones as indicated, and total RNA was extracted from seeds after the indicated periods. Total RNA was extracted using TRIzol reagent (Invitrogen) for real-time RT-PCR analysis. Superscript II was used according to the manufacturer’s instructions (Invitrogen, Carlsbad, CA, USA), and 1 μg DNase-treated RNA was reverse transcribed in a 20 μl reaction volume. Each qRT-PCR was conducted using 1 μl cDNA and the SYBR Premix Ex Taq kit (Takara) on a Roche LightCycler 480 real-time PCR instrument. At least three independent biological samples for each replicate were analyzed. *AT1G13320* was used as an internal control. The independent biological data were subjected to ANOVA (Supplemental Data Set 3). The 2^{-ΔΔCt} method was used for relative quantification of gene transcript levels (Livak and Schmittgen, 2001). The RT-qPCR primers used in these analyses are listed in Supplemental Table 2.

**RNA Sequencing and Data Analysis**

The Col-0, *coi1*-2, and *coi1*-16 seeds were grown on media with or without 1 μM ABA for 24 h, and then the materials were collected. Three biological replicates were prepared for each sample, and RNA was extracted from a mixed sample of seeds. RNA was extracted using TRIzol reagent (Invitrogen) using the ethanol precipitation protocol, and then sequenced (OEBIOTECH, Shanghai, China). Clean reads were mapped to the *Arabidopsis* genome (TAIR10; www.arabidopsis.org) after screening and trimming. Cufflinks software was used to determine expression values (Tarazona et al., 2011). Genes with estimated absolute fold changes ≥1 were identified as reliable differentially expressed genes (DEGs). DEGs were analyzed using DESeq (Anders and Huber, 2012). Multiple testing was corrected via false discovery rate estimation and q-values below 0.05 were considered to indicate differential expression. Subsequently, gene ontology (GO) enrichment analysis was performed using TopGO (Alexa and Rahnenfuhrer, 2010). The RT-qPCR primers used in the analysis shown in Figure 9F are listed in Supplemental Table 2.
Y2H Screening and Confirmation

The full-length CDS of ABI3 was fused to pGBK7 to construct the bait vector, which was transformed into the yeast strain Y2HGOLD (Clontech). Yeast screening was performed as described previously (Hu et al., 2013). Each of the 12 JAZs (JAZ1, JAZ2, JAZ3, JAZ4, JAZ5, JAZ6, JAZ7, JAZ8, JAZ9, JAZ10, JAZ11, and JAZ12) was introduced into the prey vector pGADT7. These plasmids were co-transfected into yeast strain AH109. The transfected yeast cells were plated on SD/-Leu/-Trp medium and SD/-Ade/-His/-Leu/-Trp medium and cultured at 28°C for 4 days. The primers and restriction enzyme sites used to amplify sequences and generate vectors are listed in Supplemental Table 3.

BiFC Assays

The full-length CDSs of JAZ1, JAZ4, JAZ5, and JAZ8 were fused in-frame to the YFP C-terminus of the C-YFP to form JAZ1-cYFP, JAZ4-cYFP, JAZ5-cYFP and JAZ8-cYFP, respectively. The ABI3 and ABI4 full-length CDSs were cloned in-frame to the YFP N-terminus, respectively. Each of the cloning plasmids was introduced into Agrobacterium tumefaciens strain EHA105, and then infiltrated into N. benthamiana leaves as described previously (Wang et al., 2016). At 48 h after infiltration, infected tissues were analyzed under a confocal laser-scanning microscope (Leica, Wetzlar, Germany). The primers and restriction enzyme sites are listed in Supplemental Table 3.

Co-IP Assays

The Co-IP assays were performed as described elsewhere (Pan et al., 2018). Briefly, the full-length CDSs of JAZ1 and JAZ5 were separately cloned into a tagging vector with HA-tag in the sense orientation downstream of the CaMV 35S promoter. ABI3 was fused to a 3×MYC-tag to form the ABI3-3MYC fusion protein (Pan et al., 2018). The fusion proteins were transiently coexpressed in N. benthamiana leaves. All of the infected leaves were treated with 50 µM MG132 and 5 µM ABA after being infiltrated for 40 h. The leaves were collected after 48 h and homogenized in extraction buffer [50 mM Tris-HCl, pH 7.5, 50 mM NaCl, 1 mM EDTA, 0.1% (w/v) Trition-X-100, 0.2% Nonidet P-40, 0.6 mM PMSF, and 25 µM MG132 with 1×Roche protease inhibitor cocktail]. Then, MYC-fused ABI3 was immunoprecipitated using an anti-MYC mouse antibody (1:5000; Sigma-Aldrich; catalog no. A7470; Pan et al., 2018), and the co-immunoprecipitated proteins were detected using an anti-HA rabbit antibody (1:10000; Sigma-Aldrich; catalog no. H9658). The primers and restriction enzyme sites are listed in Supplemental Table 1. The
antibodies used for this study included anti-mouse MYC (1:5000; Sigma-Aldrich; Pan et al., 2018) and anti-rabbit HA (1:10000; Sigma-Aldrich).

**Transient Transactivation Assay**

To detect the transactivation activity of ABI3 and ABI5 proteins, the promoter regions of *ABI5* (~2,326 bp), *EM6* (~1,273 bp), and *EM1* (~2,000 bp) were amplified by PCR and independently cloned into the pGreenII 0800-LUC vector as reporter plasmids (Hellens et al., 2005). JAZ1, JAZ5, or JAZ8 were amplified and independently cloned into the pGreenII 62-SK vector as effector plasmids (Hellens et al., 2005). Combinations of plasmids were introduced into mesophilic protoplasts from *Arabidopsis* according to the protocol of Sheen (2001). Transfected cells were cultured for 10 to 16 h with or without 5 µM ABA, and then relative LUC activity was determined using the Dual-Luciferase Reporter Assay Protocol machine (Promega, Madison, WI, USA), which measures the activities of firefly LUC and the internal control Renilla luciferase (REN). The primers and restriction enzyme sites are listed in Supplemental Table 4.

**Statistical analysis**

Analysis of variance (ANOVA) was performed using SPSS software. A value of P < 0.05 was considered to be statistically significant. The results of statistical analyses are shown in Supplement Data Set 3.

**Accession Numbers**

Arabidopsis Genome Initiative numbers for the genes discussed in this article are as follows: *ABA1*, AT5G67030; *ABA2*, AT1G52340; *ABA3*, AT1G16540; *ABI3*, AT3G24650; *ABI4*, AT2G40220; *ABI5*, AT2G36270; *ACX1*, AT4G16760; *ADHI*, AT1G77120; *AOC3*, AT3G25780; *AT1G13320*; *COII*, AT2G39940; *CRU1*, AT5G44120; *CRU3*, AT4G28520; *EM1*, AT3G51810; *EM6*, AT2G40170; *KAT5*, AT5G48880; *LEA*, AT2G21490; *IAA29*, AT4G32280; *JAZ1*, AT1G19180; *JAZ4*, AT1G48500; *JAZ5*, AT1G17380; *JAZ8*, AT1G30135; *MED18*, AT2G22370; *MYC2*, AT1G32640; *NCED3*, AT3G14440; *OPR3*, AT2G06050; *RAB18*, AT1G43890; *WRKY57*, AT1G69310.

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Author contributions
J.P., G.A.H., and D.Y. designed the experiments; J.P., H.W., Q.G., and Y.G. performed the experiments; J.P., G.A.H., and D.Y. analyzed the data; J.P., and Y.H. wrote the manuscript. All authors read and approved the final article.

Supplemental Data
Supplemental Figure 1. JA Enhances ABA Signaling during Seed Germination. Supports Figure 1.
Supplemental Figure 2. The ABA-sensitivity of coi1-2, coi1-16, and jazD Seeds. Supports Figure 2.
Supplemental Figure 3. The Phenotype of wild type (Col-0), coi1-2, coi1-16, and abi3-8 under Different Conditions. Supports Figure 2.
Supplemental Figure 4. The MeJA-sensitivity of coi1-2 COI1, coi1-2/COI1pro:COI1, coi1-16 COI1, and coi1-16/COI1pro:COI1 Seedlings. Supports Figure 2.
Supplemental Figure 5. The ABA-sensitivity of coi1-2 COI1, coi1-2/COI1pro:COI1, coi1-16 COI1, and coi1-16/COI1pro:COI1 Seeds. Supports Figure 2.
Supplemental Figure 6. The Phenotype of Plants Overexpressing the Full-length JAZ and Expression Patterns of JAZ. Supports Figure 2 and 3.
Supplemental Figure 7. Y2H Assays for the Interaction of JAZ and ABI3, ABI4, WRKY57. Supports Figure 3.
Supplemental Figure 8. Expression of ABI3, ABI4, JAZ1, JAZ4, JAZ5, or JAZ8 Protein in N. benthamiana Leaves. Supports Figure 3.
Supplemental Figure 9. Expression of ABI3, JAZ1, JAZ5, or GFP Protein in N. benthamiana Leaves. Supports Figure 3.
Supplemental Figure 10. The Phenotype of Seed Dormancy in Col-0, ABI3-OE, abi3, coi1-2 and JAZ1-Ajas. Supports Figure 5.

Supplemental Figure 11. RT-qPCR Analysis of ABI3 or ABI5 Expression in Overexpression Lines. Supports Figure 7 and 8.

Supplemental Figure 12. Overexpressing ABI3 and ABI5 Simultaneously Rescued the ABA-insensitive Phenotypes of coi1-16 and JAZ5-Ajas. Supports Figure 7 and 8.

Supplemental Figure 13. RT-qPCR Analysis of Several JA and ABA Biogenesis Genes in Col-0, coi1-2 and coi1-16. Supports Figure 9 and 10.

Supplemental Table 1. Primers used for transgenic plants construction and CoIP assay.

Supplemental Table 2. Primers used for qRT-PCR.

Supplemental Table 3. Primers used for yeast-two hybrid assays and BiFC assay.

Supplemental Table 4. Primers used for transient expression assay.

Supplemental Data Set 1. Sequence of JAZ’Splicing Variant. Supports Figure 3 and 4.

Supplemental Data Set 2. Data of Transcriptomic Analysis for COI1 regulated genes. Supports Figure 9.

Supplemental Data Set 3. Statistical Analysis of ANOVA Results for the Data Shown in Figures.

References

Abe, H., Urao, T., Ito, T., Seki, M., Shinozaki, K., and Yamaguchi-Shinozaki. (2003) Arabidopsis AtMYC2 (bHLH) and AtMYB2 (MYB) function as transcriptional activators in abscisic acid signaling. Plant Cell 15: 63–78.

Aleman, F., Yazaki, J., Lee, M., Takahashi, Y., Kim, AY., Li, Z., Kinoshita, T., Ecker, J.R., and Schroeder, J.J. (2016) An ABA-increased interaction of PYL6 ABA receptor with MYC2 Transcription Factor: A putative link of ABA and JA signaling. Sci Rep. 6: 28941.

Alexa, A., and Rahnebfuhrer, J. (2010) TopGO: enrichment analysis for gene ontology. R package version 2.8.

Anders, s., and Huber, W. (2012) Differential expression of RNA-Seq data at the gene level-the DESeq package [J]. Heidelberg, Germany: European Molecular Biology Laboratory.

Barrero, J.M., Talbot, M.J., White, R.G., Jacobsen, J.V., and Gubler, F. (2009) Anatomical and transcriptomic studies of the coleorhiza reveal the importance of this tissue in regulating dormancy in barley. Plant Physiol. 150: 1006–1021.

Bedi, S., Senqupta, S., Ray, A. Naq., and Chaudhuri, R. (2016) ABI3 mediates dehydration stress
recovery response in *Arabidopsis thaliana* by regulating expression of downstream genes. Plant Sci. 250: 125–140.

Bentsink, L., and Koornneef, M. (2008) Seed dormancy and germination. *Arabidopsis* Book 6: e0119.

Brocard, I.M., Lynch, T.J., and Finkelstein, R.R. (2002) Regulation and role of the *Arabidopsis* abscisic acid-insensitive 5 gene in abscisic acid, sugar, and stress response. Plant Physiol. 129: 1533–1543.

Campos, M.L., et al. (2016) Rewiring of jasmonate and phytochrome B signaling uncouples plant growth-defense tradeoffs. Nat Commun. 7: 12570.

Bentsink, L., Koornneef, M. (2008) Seed dormancy and germination. *Arabidopsis* Book 6: e0119.
Fernández-Arbaizar, A., Reqalado, J.J., and Lorenzo, O. (2012) Isolation and characterization of novel mutant loci suppressing the ABA hypersensitivity of the Arabidopsis coronatine insensitive 1-16 (coi1-16) mutant during germination and seedling growth. Plant Cell Physiol. 53: 53–63.

Fernández-Calvo, P., et al. (2011) The Arabidopsis bHLH transcription factors MYC3 and MYC4 are targets of JAZ repressors and act additively with MYC2 in the activation of jasmonate responses. Plant Cell 23: 701–715.

Finkelstein, R.R. (1994) Mutations at two new Arabidopsis ABA response loci are similar to the abi3 mutations. Plant J. 5: 756–771.

Finkelstein, R., Gampala, S.S., Lynch, T.J., Thomas, T.L., and Rock, C.D. (2005) Redundant and distinct functions of the ABA response loci ABA-INSENSITIVE (ABI)5 and ABRE-BINDING FACTOR (ABF) 3. Plant Mol Biol. 59: 253–267.

Finkelstein, R.R., and Lynch, T.J. (2000) The Arabidopsis abscisic acid response gene ABI5 encodes a basic leucine zipper transcription factor. Plant Cell 12: 599–609.

Finkelstein, R., Reeves, W., Ariizumi, T., and Steber, C. (2008) Molecular aspects of seed dormancy. Annu. Rev. Plant Biol. 59: 387–415.

Fujii, H., Verslues, P.E., and Zhu, J.K. (2007). Identification of two protein kinases required for abscisic acid regulation of seed germination, root growth, and gene expression in Arabidopsis. Plant Cell 19: 485–494.

Fujii, H., and Zhu, J.K. (2009). Arabidopsis mutant deficient in 3 abscisic acid-activated protein kinases reveals critical roles in growth, reproduction, and stress. Proc. Natl. Acad. Sci. USA 106: 8380–8385.

Fujita, Y., Fujita, M., Shinozaki, K., and Yamaguchi-Shinozaki, K. (2011) ABA-mediated transcriptional regulation in response to osmotic stress in plants. J. Plant Res. 124: 509–525.

Furihata, T., Maruyama, K., Fujita, Y., Umezawa, T., Yoshida, R., Shinozaki, K., and Yamaguchi-Shinozaki, K. (2006). Abscisic acid-dependent multisite phosphorylation regulates the activity of a transcription activator AREB1. Proc. Natl. Acad. Sci. USA 103: 1988–1993.

Giraudat, J., Hauge, B.M., Valon, C., Smalle, J., Parcy, F., and Goodman, H.M. (1992) Isolation of the Arabidopsis ABI3 gene by positional cloning. Plant Cell 4: 1251–1261.

Goossens, J., Swinnen, G., Vanden Bossche, R., Pauwels, L., and Goossens, A. (2015) Change of a conserved amino acid in the MYC2 and MYC3 transcription factors leads to release of JAZ repression and increased activity. New Phytol. 206: 1229–1237.

Guo, Q., Yoshida, Y., Major, I.T., Wang, K., Sugimoto, K., Kapali, G., Havko, N.E., Benning, C., Howe, G.A. (2018) JAZ repressors of metabolic defense promote growth and reproductive fitness in Arabidopsis. Proc Natl Acad Sci U S A. 115: E10768–E10777.
Hauser, F., Waadt, R., and Schroeder, J.I. (2011) Evolution of abscisic acid synthesis and signaling mechanisms. Curr. Biol 21: R346–355.

Hellens, R.P., Allan, A.C., Friel EN, Bolitho, K., Grafton, K., Templeton, M.D., Karunairetnam, S., Gleave, A.P., and Laing, W.A. (2005) Transient expression vectors for functional genomics, quantification of promoter activity and RNA silencing in plants. Plant Methods 1: 13.

Howe, G.A., Major, I.T., and Koo, A.J. (2018) Modularity in jasmonate signaling for multistress resilience. Annu Rev Plant Biol. 69: 387–415.

Hu, Y., Jiang, L., Wang, F., and Yu, D. (2013) Jasmonate regulates the INDUCER OF CBF EXPRESSION-C-REPEAT BINDING FACTOR/DRE BINDING FACTOR1 cascade and freezing tolerance in Arabidopsis. Plant Cell 25: 2907–2924.

Hu, Y., and Yu, D. (2014) BRASSINOSTEROID INSENSITIVE2 interacts with ABSCISIC ACID INSENSITIVE5 to mediate the antagonism of brassinosteroids to abscisic acid during seed germination in Arabidopsis. Plant Cell 26: 4394–4408.

Jiang, Y., Lang, G., Yang, S., and Yu, D. (2014) Arabidopsis WRKY57 functions as a node of convergence for jasmonic acid- and auxin-mediated signaling in jasmonic acid-induced leaf senescence. Plant Cell 26: 230–245.

Jiang, W., and Yu, D. (2009) Arabidopsis WRKY2 transcription factor mediates seed germination and postgermination arrest of development by abscidic acid. BMC Plant Biology 9: 96.

Ju, L., Jing, Y., Shi, P., Liu, J., Chen, J., Yan, J., Chu, J., Chen, K., and Sun, J. (2019) JAZ proteins modulate seed germination through interaction with ABI5 in bread wheat and Arabidopsis. New Phytol. 223: 246–260.

Kazan, K. (2015) Diverse roles of jasmonates and ethylene in abiotic stress tolerance. Trends Plant Sci. 20: 219–229.

Kazan, K., and Manners, J.M. (2013) MYC2: the master in action. Mol. Plant 6: 686–703.

Kobayashi, Y., Murata, M., Minami, H., Yamamoto, S., Kagaya, Y., Hobo, T., Yamamoto, A., and Hattori, T. (2005). Abscisic acid activated SNRK2 protein kinases function in the gene-regulation pathway of ABA signal transduction by phosphorylating ABA response element-binding factors. Plant J. 44: 939–949.

Koornneef, M., Reuling, G., and Karssen, C.M. (1984). The isolation and characterization of abscisic acid-insensitive mutants of Arabidopsis thaliana. Physiol. Plant 61: 377–383.

Krock, B., Schmidt, S., Hertweck, C., and Baldwin, I.T. (2002) Vegetation derived abscisic acid and four terpenes enforce dormancy in seeds of the post-fire annual, Nicotiana attenuata. Seed Sci. Res. 12: 239–252.
Lackman, P., et al. (2011) Jasmonate signaling involves the abscisic acid receptor PYL4 to regulate metabolic reprogramming in *Arabidopsis* and tobacco. Proc Natl Acad Sci U S A. 108: 5891–5896.

Lai, Z., Schluttenhofer, C.M., Bhide, K., Shreve, J., Thimmapuram, J., Lee, S.Y., Yun, D.J., and Mengiste, T. (2014) MED18 interaction with distinct transcription factors regulates multiple plant functions. Nat Commun. 5: 3064.

Lin, J., Yu, L., and Xiang, C. (2020) *ABABIDOPSIS NITRATE REGULAED 1* acts as a negative modulator of seed germination by activating ABI3 expression. New phytol. 225: 835–847.

Lim, S., et al. (2013) ABA-INSENSITIVE3, ABA-INSENSITIVE5, and DELLAs interact to activate the expression of SOMNUS and other high-temperature-inducible genes in imbibed seeds in *Arabidopsis*. Plant Cell 25: 4863–4878.

Liu, X., Zhang, H., Zhao, Y., Feng, Z., Li, Q., Yang, H.Q., Luan, S., Li, J., and He, Z.H. (2013) Auxin controls seed dormancy through stimulation of abscisic acid signaling by inducing ARF-mediated ABI3 activation in *Arabidopsis*. Proc. Natl. Acad. Sci. USA 110: 15485–15490.

Livak, K.J., and Schmittgen, T.D. (2001) Analysis of relative gene expression data using real-time quantitative PCR and the 2−ΔΔCT method. Methods 25: 402–408.

Lopez-Molina, L., and Chua, N.H. (2000) A null mutation in a bZIP factor confers ABA-insensitivity in *Arabidopsis thaliana*. Plant Cell Physiol. 41: 541–547.

Lopez-Molina, L., Mongrand, S., and Chua, N.H. (2001) A postgermination developmental arrest checkpoint is mediated by abscisic acid and requires the ABI5 transcription factor in *Arabidopsis*. Proc. Natl. Acad. Sci. USA 98: 4782–4787.

Lopez-Molina, L., Mongrand, S., McLachlin, D.T., Chait, B.T., and Chua, N.H. (2002) ABI5 acts downstream of ABI3 to execute an ABA-dependent growth arrest during germination. Plant J. 32: 317–328.

Lorenzo, O., Chico, J.M., Sánchez-Serrano, J.J., and Solano, R. (2004) JASMONATE-INSENSITIVE1 encodes a MYC transcription factor essential to discriminate between different jasmonate-regulated defense responses in *Arabidopsis*. Plant Cell 16: 1938–1950.

Ma, Y., Szostkiewicz, I., Korte, A., Moes, D., Yang, Y., Christmann, A., and Grill, E. (2009) Regulators of PP2C phosphatase activity function as abscisic acid sensors. Science 324:1064–1068.

Mauch-Mani, B., and Mauch, F. (2005) The role of abscisic acid in plant-pathogen interactions. Curr. Opin. Plant Biol. 8: 409–414.

Miyazono, K., et al. (2009). Structural basis of abscisic acid signaling. Nature 462: 609–614.

Nakamura, S., Lynch, T.J., and Finkelstein, R.R. (2001) Physical interactions between ABA response loci
of *Arabidopsis*. Plant J. **26**: 627–635.

Nakashima, K., *et al.* (2009). Three *Arabidopsis* SnRK2 protein kinases, SRK2D/SnRK2.2, SRK2E/SnRK2.6/OST1 and SRK2I/SnRK2.3, involved in ABA signaling are essential for the control of seed development and dormancy. Plant Cell Physiol. **50**: 1345–1363.

Nakashima, K., and Yamaguchi-Shinozaki, K. (2013) ABA signaling in stress-response and seed development. Plant Cell Rep. **32**: 959–970.

Nishimura, N., Hitomi, K., Arvai, A.S., Rambo, R.P., Hiromi, C., Cutler, S.R., Schroeder, J.I., and Getzoff, E.D. (2009) Stuctural mechanism of abscisic acid binding and signaling by dimeric PYR1. Science **326**: 1373–1379.

Niu, Y.J., Figueroa, P., and Browse, J. (2011) Characterization of JAZ- interacting bHLH transcription factors that regulate jasmonate responses in *Arabidopsis*. J. Exp. Bot. **62**: 2143–2154.

Norastehnia, A., Sajedi, R.H., and Nojavan-Asghari, M. (2007) Inhibitory effects of methyl jasmonate on seed germination in maize (*Zea mays*): effect on a-amylase activity and ethylene production. Gen, Appl. Plant Physiol. **33**: 13–23.

Ooms, J., Leon-Kloosterziel, K.M., Bartels, D., Koornneef, M., and Karssen, C.M. (1993) Acquisition of desiccation tolerance and longevity in seeds of *Arabidopsis thaliana* (A comparative study using abscisic acid-insensitive abi3 mutants). Plant Physiol. **102**: 1185–1191.

Pan, J., Wang, H., Hu, Y., and Yu, D. (2018) *Arabidopsis* VQ18 and VQ26 proteins interact with ABI5 transcription factor to negatively modulate ABA response during seed germination. Plant J. **95**: 529–544.

Park, J., Lee, N., Kim, W., Lim, S., and Choi, G. (2011) ABI3 and PIL5 collaboratively activate the expression of SOMNU5 by directly binding to its promoter in imbibed *Arabidopsis* seeds. Plant Cell **23**: 1404–1215.

Pauwels, L., *et al.* (2010) NINJA connects the co-repressor TOPLESS to jasmonate signaling. Nature **464**: 788–791.

Pauwels, L., *et al.* (2015) The RING E3 Ligase KEEP ON GOING Modulates JASMONATE ZIM-DOMAIN12 Stability. Plant Physiol. **169**: 1405–1417.

Pauwels, L., and Goossens, A. (2011) The JAZ proteins: a crucial interface in the jasmonate signaling cascade. Plant Cell **23**: 3089–3100.

Piskurewicz, U., and Lopez-Molina, L. (2016) Basic Techniques to Assess Seed Germination Responses to Abiotic Stress in *Arabidopsis thaliana*. Methods Mol Biol. **1398**: 183-196.

Preston, C.A., Betts, H., and Baldwin, I.T. (2002) Methyl jasmonate as an allelopathic agent: sagebrush inhibits germination of a neighboring tobacco *Nicotiana Attenuata*. J. Chem. Ecol. **28**: 2343–2369.
Qi, T., et al. (2011) The Jasmonate-ZIM-domain proteins interact with the WD-Repeat/bHLH/MYB complexes to regulate Jasmonate-mediated anthocyanin accumulation and trichome initiation in *Arabidopsis thaliana*. Plant Cell 23: 1795–1784.

Qi, T., Huang, H., Song, S., and Xie, D. (2015) Control of jasmonate-regulated stamen development and seed production by a bHLH-MYB complex in *Arabidopsis*. Plant Cell 27: 1620–1633.

Santiago, J., Dupeux, F., Round, A., Antoni, R., Park, S.Y., Jamin, M., Cutler, S.R., Rodriguez, P.L., and Márquez, J.A. (2009). The abscisic acid receptor PYR1 in complex with abscisic acid. Nature 462: 665–668.

Schweizer, F., et al. (2013) *Arabidopsis* basic helix-loop-helix transcription factors MYC2, MYC3, and MYC4 regulate glucosinolate biosynthesis, insect performance, and feeding behavior. Plant Cell 25: 3117–3132.

Sheen, J. (2001) Signal transduction in maize and *Arabidopsis* mesophyll protoplasts. Plant Physiol. 127: 1466–1475.

Shyu, C., Figueroa, P., DePew, C.L., Cooke, T.F., Sheard, L.B., Moreno, J.E., Katsir, L., Zheng, N., Browse, J., and Howe, G.A. (2012) JAZ8 lacks a canonical degron and has an EAR motif that mediates transcriptional repression of jasmonate responses in *Arabidopsis*. Plant Cell 24: 536–550.

Staswick, P.E., Su, W., and Howell, S.H. (1992) Methyl jasmonate inhibition of root growth and induction of a leaf protein are decreased in an *Arabidopsis thaliana* mutant. Proc. Natl. Acad. Sci. USA 89: 6837–6840.

Song, S., et al. (2011) The Jasmonate-ZIM-domain proteins interact with the R2R3-MYB transcription factors MYB21 and MYB24 to affect Jasmonate-regulated stamen development in *Arabidopsis*. Plant Cell 23: 1000–1013.

Suzuki, M., Kao, C.Y., Cocciolone, S., and McCarty, D.R. (2001) Maize VP1 complements *Arabidopsis abi3* and confers a novel ABA/auxin interaction in roots. Plant J. 28: 409–418.

Tarazona, S., García-Alcalde, F., Dopazo, J., Ferrer, A., and Conesa, A. (2011) Differential expression in RNA-seq: a matter of depth. Genome Res. 21: 2213–2223.

Thatcher, L.F., et al. (2016) Characterization of a JAZ7 activation-tagged *Arabidopsis* mutant with increased susceptibility to the fungal pathogen *Fusarium oxysporum*. J. Exp. Bot. 67: 2367–2386.

Thines, B., et al. (2007) JAZ repressor proteins are targets of the SCF(COI11) complex during jasmonate signaling. Nature 448: 661–665.

Vaistij, F.E., et al. (2013) Differential control of seed primary dormancy in *Arabidopsis* ecotypes by the transcription factor SPATULA. Proc. Natl. Acad. Sci. USA 110: 10866–10871.

Wang, H., Pan, J., Li, Y., Lou, D., Hu, Y., and Yu, D. (2016) The DELLA-CONSTANS cascade
integpercentages gibberellin and photoperiod signaling to regulate flowering in *Arabidopsis*. Plant Physiol. 172: 479–488.

Wang, K., Guo, Q., Froehlich, J. E., Hersh, H. L., Zienkiewicz, A., Howe, G. A., and Benning, C. (2018) Two abscisic acid-responsive plastid lopase genes involved in jasmonic acid biosynthesis in *Arabidopsis thaliana*. Plant Cell 30: 1006-1022.

Wilen, R.W., van Rooijen, G.J., Pearce, D.W., Pharis, R.P., Holbrook, L.A., and Moloney, M.M. (1991) Effects of jasmonic acid on embryo specific processes in *Brassica* and *Linum* oilseeds. Plant Physiol. 95: 399–405.

Xie, D.X., Feys, B.F., James, S., Nieto-Rostro, M., and Turner, J.G. (1998) *COII*: an *Arabidopsis* gene required for jasmonate-regulated defense and fertility. Science 280: 1091–1094.

Xu, L., Liu, F., Lechner, E., Genschik, P., Crosby, W.L., Ma, H., Peng, W., Huang, D., and Xie, D. (2002) The SCF(COII) ubiquitin-ligase complexes are required for jasmonate response in *Arabidopsis*. Plant Cell 14: 1919–1935.

Yan, J., et al. (2009) The *Arabidopsis* CORONATINE INSENSITIVE1 protein is a jasmonate receptor. Plant Cell 21: 2220–2236.

Yoo, S.D., Cho, Y.H., and Sheen, J. (2007) *Arabidopsis* mesophyll protoplasts: a versatile cell system for transient gene expression analysis. Nat. Protoc. 2: 1565–1572.

Zhai, Q., Zhang, X., Wu, F., Feng, H., Deng, L., Xu, L., Zhang, M., Wang, Q., and Li, C. (2015) Transcriptional mechanism of jasmonate receptor COII-mediated delay of flowering time in *Arabidopsis*. Plant Cell 27: 2814–2828.

Zhou, X., et al. (2015) *SOS2*-LIKE PROTEIN KINASE5, an SNF1-RELATED PROTEIN KINASE3-Type protein kinase, is important for abscisic acid responses in *Arabidopsis* through phosphorylation of ABSCISIC ACID-INSENSITIVE5. Plant Physiol. 168: 659-676.

Zhu, Y., et al. (2020) The *Arabidopsis* nodulin homeobox factor AtNDX interacts with AtRING1A/B and negatively regulates abscisic acid signaling. Plant Cell 32: 703-721.

Zhu, Z., et al. (2011) Derepression of ethylene-stabilized transcription factor (EIN3/EIL1) mediates jasmonate and ethylene signaling synergy in *Arabidopsis*. Proc. Natl. Acad. Sci. USA 108: 12539–12544.
Figure 1. JA Enhances ABA Signaling during Seed Germination. (A) Germination of wild-type seeds was scored 1-7 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. (B) The percentage of wild-type seedlings with expanded cotyledons was scored 2-7 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. (C) Seedlings of wild type (Col-0) observed 4.5 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. Scale bar, 30 mm. The average and significance were calculated over three biological replicates by analyzing seeds of different batches. Each batch of seeds for Col-0 was pooled from 50 independent plants. For every biological replicate, we examined seeds from the same batch at least three times as technical replicates. The value of each biological replicate was the average calculated over three technical replicates by analyzing more than 200 seeds. (D) Expression of several ABA-responsive genes in response to ABA and COR treatment. Seeds of wild type were germinated on water agar medium containing 1 µM COR, 0.5 µM ABA, and both 0.5 µM ABA and 1 µM COR, respectively, then those seeds grown on these medium 4 h in long-day conditions. The average and significance were calculated over three biological replicates. Total RNA was extracted from at least three batches of seeds as biological replicates. Each batch of seeds for the wild type was pooled from 50 independent plants. For each biological replicate, more than 250 seeds of the same batch were used for RNA extraction. AT1G13320 gene was used as control. Data from three biological replicates were analyzed by ANOVA. Values with different letters are significantly different from each other (P < 0.05). Error bars show SD from three biological replicates.
Figure 2. The COI1/JAZ-Mediated JA Pathway Is Positively Associated with ABA Signaling during Seed Germination. (A) to (B) Germination of wild type (Col-0), jazQ, coi1-2, coi1-16, JAZ1-ΔJas, JAZ5-ΔJas, and JAZ8-ΔJas seeds was scored 1-5 d after stratification on water agar medium supplemented without and with 0.5 µM ABA. (C) The percentage of Col-0, jazQ, coi1-2, coi1-16, JAZ1-ΔJas, JAZ5-ΔJas, and JAZ8-ΔJas seedlings with expanded cotyledons was scored 5 d after stratification on water agar medium supplemented with 0.5 µM ABA. (D) Seedlings of Col-0, jazQ, coi1-2, coi1-16, JAZ1-ΔJas, JAZ5-ΔJas, and JAZ8-ΔJas observed 4 d after stratification on water agar medium supplemented with 0.5 µM and 1 µM ABA. Scale bar, 30 mm. The average and significance were calculated over three biological replicates by analyzing seeds of different batches. Each batch of seeds for each genotype was pooled from at least 15 independent plants. For every biological replicate, we examined the seeds from the same batch at least three times as technical replicates. The value of each biological replicate was the average calculated over three technical replicates by analyzing more than 150 seeds. Data from three biological replicates were analyzed by ANOVA. Values with different letters are significantly different from each other (p < 0.05). (E) Expression of several ABA-responsive genes in jazQ and coi1-2. Dry seeds of Col-0, jazQ, and coi1-2 were collected. The average and significance were calculated over three biological replicates. Total RNA was extracted from at least three batches of seeds as biological replicates. Each batch of seeds for Col-0, jazQ, and coi1-2 was pooled from at least 15 independent plants, respectively. For each biological replicate, more than 250 seeds of the same batch were used for RNA extraction. AT1G13320 gene was used as control. Data from three biological replicates were analyzed by ANOVA. Values with different letters are significantly different from each other (p < 0.05). Error bars show SD from three biological replicates.
Figure 3. Physical Interactions between JAZ and ABI3. (A) and (B) Y2H assay to detect interactions of ABI3 and ABI4 with JAZ proteins. Interaction was indicated by the ability of cells to grow on dropout medium lacking Leu, Trp (A). Interaction was indicated by the ability of cells to grow on dropout medium lacking Leu, Trp, His, and Ade and containing 20 mM 3-aminotriazole for 4 d. pGBK7 vector was used as the negative control (B). (C) BiFC analyses. JAZ1, JAZ4, JAZ5, and JAZ8 were fused to the N-terminal fragment of YFP (nYFP); ABI3 and ABI4 were fused to the C-terminal fragment of YFP (cYFP). Fluorescence was observed in the nuclear compartment of transformed cells, which resulted from complementation of the C-terminal part of YFP fused with ABI3 (ABI3-cYFP) or ABI4-cYFP with the N-terminal part of YFP fused with JAZ (JAZ-nYFP). The localization of the nuclei was detected by 4',6-diamidino-2-phenylindole (DAPI) staining. Scale bar, 15 µm. (D) and (E) Co-IP analyses. Protein extracts from N.benthamiana leaves coexpressing MYC-tagged ABI3 with HA-fused JAZ1, JAZ5, or GFP were immunoprecipitated (IP) with HA antibody-bound agarose beads, and the immunoprecipitated proteins were analyzed by immunoblotting using anti-MYC or anti-HA antibody. The experiments were repeated three times with similar results.
Figure 4. Y2H Assays for Identifying ABI3 and JAZ Regions Required for Their Interaction. (A) The N-terminal fragment of ABI3 is involved in its interaction with JAZ proteins. Left: Diagram of full-length and truncated ABI3 constructs with specific deletions. Right: Interaction was indicated by the ability of cells to grow on dropout medium lacking Leu, Trp, His, and Ade and containing 20 mM 3-aminotriazole for 4 d. The vectors pGBKT7 (BD) and pGADT7 (AD) were used as negative controls. (B) and (C) The ZIM domain of JAZ1 or JAZ8 is responsible for their interactions with ABI3. Left: Diagram of full-length and truncated JAZ constructs with specific deletions. Right: Interaction was indicated by the ability of cells to grow on dropout medium lacking Leu, Trp, His, and Ade and containing 20 mM 3-aminotriazole for 4 d. The vectors BD and AD were used as negative controls.
Figure 5. JA Promotes ABA Signaling during Seed Germination Requiring ABI3 and ABI5. (A) to (D) Germination of Col-0, abi3, abi5, abi5 abi3, abi3-8, abi5-7, Ler, abi3-1, Ws, and abi5-1 seeds was scored 1-5 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. (E) The percentage of Col-0, abi3, abi5, abi5 abi3, abi3-8, abi5-7, Ler, abi3-1, Ws, and abi5-1 seedlings with expanded cotyledons was scored 5 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. (F) Seedlings of Col-0, abi3, abi5, abi5 abi3, abi3-8, abi5-7, Ler, abi3-1, Ws, and abi5-1 observed 4 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. Scale bar, 30 mm. The average and significance were
calculated over three biological replicates by analyzing seeds of different batches. Each batch of seeds for each genotype was pooled from at least 15 independent plants. For every biological replicate, we examined the seeds from the same batch at least three times as technical replicates. The value of each biological replicate was the average calculated over three technical replicates by analyzing more than 150 seeds. Data from three biological replicates were analyzed by ANOVA. Values with different letters are significantly different from each other (p < 0.05). Error bars show SD from three biological replicates.
Figure 6. JAZ Proteins Repress the Transcriptional Activities of ABI3 and ABI5. (A) Schematic of the reporter and effectors used in the transient transactivation assays. (B) and (C) JAZ1 and JAZ5 repress ABI3 to activate EM6 and ABI5 in response to 5 µM ABA. Error bars show SD from three biological replicates; each biological replication was from different wild type leaves. Data were analyzed by ANOVA. Values with different letters are significantly different from each other (p < 0.05). (D) and (E) JAZ8 represses ABI5 to activate EM6 and EM1 in response to 5 µM ABA. Error bars show SD from three biological replicates; each biological replication was from different wild type leaves. Data were analyzed by ANOVA. Values with different letters are significantly different from each other (p < 0.05).
Figure 7. Overexpression of ABI3 and ABI5 Simultaneously Suppresses the ABA-Insensitive Phenotypes of coi1-2 Mutant. (A) to (D) Germination of Col-0, ABI3-OE, ABI5-OE, coi1-2, coi1-2 ABI3-OE, and coi1-2 ABI3-OE ABI5-OE seeds was scored 1-5 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. (E) The percentage of Col-0, ABI3-OE, ABI5-OE, coi1-2, coi1-2 ABI3-OE, and coi1-2 ABI3-OE ABI5-OE seedlings with expanded cotyledons was scored 5 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. (F) Seedlings of Col-0, ABI3-OE, ABI5-OE, coi1-2, coi1-2 ABI3-OE, and coi1-2 ABI3-OE ABI5-OE observed
4 d after stratification on water agar medium supplemented with 1 μM COR, 0.5 μM ABA, or both 0.5 μM ABA and 1 μM COR, respectively. Scale bar, 30 mm. The average and significance were calculated over three biological replicates by analyzing seeds of different batches. Each batch of seeds for each genotype was pooled from at least 18 independent plants. For every biological replicate, we examined the seeds from the same batch at least three times as technical replicates. The value of each biological replicate was the average calculated over three technical replicates by analyzing more than 150 seeds. Data from three biological replicates were analyzed by ANOVA. Values with different letters are significantly different from each other (p < 0.05). Error bars show SD from three biological replicates.
Figure 8. Overexpression of ABI3 and ABI5 Simultaneously Suppresses the ABA-insensitive Phenotypes of JAZ8-ΔJas. (A) to (D) Germination of Col-0, ABI3-OE, ABI5-OE, JAZ8-ΔJas, JAZ8-ΔJas ABI3-OE, and JAZ8-ΔJas ABI3-ΔJas ABI5-OE seeds was scored 1-5 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. (E) The percentage of Col-0, ABI3-OE, ABI5-OE, JAZ8-ΔJas, JAZ8-ΔJas ABI3-OE, and JAZ8-ΔJas ABI3-OE ABI5-OE seedlings with expanded cotyledons was scored 5 d after stratification on water agar medium supplemented with 1 µM COR, 0.5 µM ABA, or both 0.5 µM ABA and 1 µM COR, respectively. (F) Seedlings of Col-0, ABI3-OE, ABI5-OE, JAZ8-ΔJas, JAZ8-ΔJas ABI3-OE, and JAZ8-ΔJas ABI3-OE
ABI5-OE observed 4 d after stratification on water agar medium supplemented with 1 μM COR, 0.5 μM ABA, or both 0.5 μM ABA and 1 μM COR, respectively. Scale bar, 30 mm. The average and significance were calculated over three biological replicates by analyzing seeds of different batches. Each batch of seeds for each genotype was pooled from at least 18 independent plants. For every biological replicate, we examined the seeds from the same batch at least three times as technical replicates. The value of each biological replicate was the average calculated over three technical replicates by analyzing more than 150 seeds. Data from three biological replicates were analyzed by ANOVA. Values with different letters are significantly different from each other (p < 0.05). Error bars show SD from three biological replicates.
Figure 9. Transcriptomic Analysis of Regulatory Gene Expression Profiles by COI1 in Response to ABA. (A) Venn diagram shows the number of overlapped up- and down-regulated DEGs in coi1-2 and coi1-16 mutant compared with the wild type without ABA treatments. (B) GO analysis of coi1-2 and coi1-16 co-regulated genes. (C) Venn diagram shows the overlapped number of up- and down-regulated DEGs in coi1-2 and coi1-16 mutant compared with the wild type under ABA treatment. (D) Venn diagram shows the overlapped number of co-up and down-regulated genes in coi1-2 and coi1-16 mutant, which are ABA responsive genes in the wild type. (E) Heat map shows that ABA responsive genes co-regulated by coi1-2 and coi1-16 mutant. (F) RT-qPCR analysis of expression level of several selected ABA responsive genes co-regulated by ABA and COI1. Seeds of wild type, coi1-2, and coi1-16 were germinated on medium containing 0.5 µM ABA treated with 4 h. The average and significance were calculated over three biological replicates. Total RNA was extracted from at least three batches of seeds as biological replicates. Each batch of seeds for each genotype was pooled from at least 20 independent plants. For each biological replicate, more than 250 seeds of the same batch were used for RNA extraction. AT1G13320 gene was used as control. Three biological replicates data were analyzed by ANOVA. Values with different letters are significant different from each other (P < 0.05).
Figure 10. Proposed Working Model Describing the Interaction of Jasmonate and Abscisic acid Signaling in Mediating Seed Germination. Under normal growth conditions, the JAZ proteins interact directly with ABI3/ABI5 and repress the transcriptional activities of ABI3/ABI5, leading to repression of the ABA pathway. Under unfavorable growth conditions, JA biosynthesis is increased, which leads to the degradation of JAZ proteins, and then ABI3/ABI5 are released from JAZ-mediated inhibition to activate ABA signaling; simultaneously, the accumulation of ABA activates the transcription factors ABI3 and ABI5, thereby delaying the process of seed germination.
Molecular Mechanism Underlying the Synergetic Effect of Jasmonate on Abscisic Acid Signaling during Seed Germination in Arabidopsis

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