Review

Adverse Biological Effect of TiO$_2$ and Hydroxyapatite Nanoparticles Used in Bone Repair and Replacement

Jiangxue Wang 1, Liting Wang 2 and Yubo Fan 1,2,*

1 Key Laboratory for Biomechanics and Mechanobiology of Ministry of Education, School of Biological Science and Medical Engineering, Beihang University, Beijing 100191, China; wangjiangxue@buaa.edu.cn
2 National Research Center for Rehabilitation Technical Aids, Beijing 100176, China; wlt6301@sohu.com
* Correspondence: yubofan@buaa.edu.cn; Tel.: +86-10-8233-9428

Academic Editor: Michael Routledge
Received: 8 April 2016; Accepted: 19 May 2016; Published: 24 May 2016

Abstract: The adverse biological effect of nanoparticles is an unavoidable scientific problem because of their small size and high surface activity. In this review, we focus on nano-hydroxyapatite and TiO$_2$ nanoparticles (NPs) to clarify the potential systemic toxicological effect and cytotoxic response of wear nanoparticles because they are attractive materials for bone implants and are widely investigated to promote the repair and reconstruction of bone. The wear nanoparticles would be prone to binding with proteins to form protein-particle complexes, to interacting with visible components in the blood including erythrocytes, leukocytes, and platelets, and to being phagocytosed by macrophages or fibroblasts to deposit in the local tissue, leading to the formation of fibrous local pseudocapsules. These particles would also be translocated to and disseminated into the main organs such as the lung, liver and spleen via blood circulation. The inflammatory response, oxidative stress, and signaling pathway are elaborated to analyze the potential toxicological mechanism. Inhibition of the oxidative stress response and signaling transduction may be a new therapeutic strategy for wear debris–mediated osteolysis. Developing biomimetic materials with better biocompatibility is our goal for orthopedic implants.

Keywords: wear debris particles; hydroxyapatite; TiO$_2$ nanoparticles; nanotoxicology; cytotoxicity; blood protein; inflammatory response; oxidative stress

1. Introduction

With the advent and rapid development of nanoscience and nanotechnology, every synthetic event occurs at the nanoscale (including fibers, capsules, or particles with at least one dimension from 1 to 100 nm). Especially the engineered nanoparticles (NPs) are massively produced and widely used in the fields of electronics, environment, agriculture, pharmacy, and medicine, etc. It is well known that the nanometer regime is the fundamental unit of length over which cells and molecules interact with biological environments. The molecular basic blocks of proteins, nucleic acids, and lipids are materials that possess unique properties at the nanoscale. For example, the width of a DNA strand is approximately 2 nm. The extracellular matrix, providing structural and biochemical support to surrounding cells, has a hierarchical structure with spatial and temporal levels from nanometer to centimeter scale. Now, inspired by the innate nanostructure of biological tissue and biomolecules, many researchers have attempted to fabricate some biomedical nanomaterials with nanoscale surface features to improve biological application in orthopedics [1–4].

Bone is viewed as a nanofibrous composite with a hierarchical structure composed of organic compounds (mainly collagen) reinforced with inorganic hydroxyapatite (HA). HA crystals are approximately 2 nm thick by 25–50 nm wide, embedded in the holes within the collagen molecule structures and increasing the rigidity of bone. The organization of bone spans three or more orders
of magnitude from large ~200 µm osteons with subunits of ~200 nm collagen fibrils to the 20 nm crystallized HA platelets. The specific structure of bone provides mechanical support, metabolic function and protects bone marrow with nutrients in the body. The fracture of bone often occurs because of high force impact and stress, and is also a result of certain medical conditions such as osteoporosis, bone cancer and osteogenesis imperfecta. The broken bone is a lot more than painful and inconvenient, and is sometimes a costly and permanent health problem. According to the National Institutes of Health, approximately 1.5 million hip fractures occur worldwide each year, and this number might increase to 2.6 million by 2025 and 4.5 million by 2050 [5]. The commercial implants, from ceramics to metals to polymers, have some clinical limitations including fatigue, fracture, poor osseointegration, extrusion, and infection. Due to the natural nanostructure of bone, nanotechnology is used to tailor orthopedic implants aimed at helping bone formation and increased integration into the host tissue. To fabricate biomimetic functional bone, many nanomaterials are designed and manufactured, such as titanium dioxide (TiO$_2$), HA, ceramics, and nanofibers of polymers. In this review, TiO$_2$ and HA are selected as the representative nanomaterials used in orthopedics because they are generally studied as potential biomedical materials, as shown in the following.

2. Benefits of TiO$_2$ and HA Nanoparticles in Bone Repair

HA with the chemical formula of Ca$_{10}$(PO$_4$)$_6$(OH)$_2$, being the main inorganic constituent of natural bone, has been widely used for biomedical applications owing to good biocompatibility and osteoconductivity [6]. Recently, nano-hydroxyapatite (n-HA) with its small size, high surface area and roughness is more often used than microscale HA for bone substitutes, tissue engineering scaffolds, coatings, and so on [7]. It is playing a more and more important role in bone repair and remodeling [8]. Many studies report that n-HA is used to form a three-dimensional biomimetic composite with chitosan, collagen [9], polymer, and other bioactive molecules [10]. The composite materials of n-HA with natural or synthetic polymer mimic the natural bone’s inorganic and organic phase composition [11]. The n-HA composite scaffolds with appropriate porous structure, biodegradability and mechanical properties can induce osteoblast adhesion, proliferation, and differentiation, increasing their osteoinductivity and osseointegrative capacity [9,12]. The osteoblastic MG63 cells prefer to attach on the gelatin/HA nanocomposites with small-sized apatite crystals, to proliferate, and to secrete alkaline phosphatase (ALP) and osteocalcin (OCN) [13]. Recently, the porous n-HA/collagen scaffold is used to load adriamycin-encapsulated poly(lactic-co-glycolic acid) (PLGA) NPs, aiming to develop an osteoconductive material with the ability of inhibiting osteosarcoma [14].

TiO$_2$ NPs, which have poor solubility and are chemically inert, are stable in the organism and have good biocompatibility and tissue compatibility. TiO$_2$ is considered as a promising bone scaffolding material because it promotes osteoblast adhesion and induces bone formation [15–18]. Tiainen et al. fabricated TiO$_2$ scaffolds by themselves and analyzed the bone ingrowth into the scaffold structure after implanting the scaffolds into surgically modified extraction sockets in miniature pigs [18]. Results revealed that the new bone formed in the scaffold pore space by 73.6% ± 11.1%, and the volume of the bone mineral density of the new bone was comparable to that of the native cortical bone. The bone tissue was in direct contact with 50.0 ± 21.5% of the TiO$_2$ scaffolds. Webster et al. firstly reported that osteoblast cells showed a greater synthesis function of proliferation and osteogenic differentiation on nanophase TiO$_2$, ceramics, and HA than conventional ceramics with grain sizes greater than 100 nm [19,20]. TiO$_2$ NPs are also used to reinforce the porosity and mechanical properties of polymer scaffolds, aiming to increase bone-forming ability [21,22]. In recent decades, the anodized TiO$_2$ nanotubes gain much attention in orthopedic implants due to the nanostructured surface stimulating the growth and mineralization of osteoblasts in vitro and inducing bone regeneration and osseointegration in vivo [23,24]. The titanium implants with TiO$_2$ nanotubes could significantly increase the bone-implant contact and the bone bonding strength [25]. The energy-dispersive X-ray spectroscopy (EDX) mapping exhibited that the strong signals of calcium and phosphorus covered 41.7% of the TiO$_2$ nanotube implant surface, but not on the Ti grit-blasted implant surface, indicating the
strong osseointegration of TiO$_2$ nanotubes [25]. At three weeks after implantation, the titanium implant with 70 nm TiO$_2$ nanotubes increased the bone-implant contact by 55.46% ± 9.71%, and the fluorescent labels (xylene orange and calcein labeled) in the surrounding bone of implants with TiO$_2$ nanotubes were more obvious and continuous than those of machined implants [16]. The newly formed bone was observed within the defect site [18] and the high gene expression of ALP, osterix (OSX) and collagen-I were detected in the bone around implants with TiO$_2$ nanotubes [16]. The experimental results have clarified that the novel nanoscale HA and TiO$_2$ show superior characteristics for bone-related cell adhesion and proliferation due to their distinctive nanoscale features and novel physical properties. On the surface of the titanium disk coated with nanoscale anatase and rutile, human mesenchymal stem cells and osteoblasts are successfully grown and expanded. The bone-specific genes including OSX, ALP, OCN, and bone sialoprotein (BSP) are increased on all nanoscale features in vitro [22,26].

In addition, it is reported that n-HA and TiO$_2$ nanotubes coated on a titanium surface could prevent bacterial colonization and bacterial biofilm formation on the implant surface [27,28].

However, nanoscale materials usually exhibit unique physical and chemical properties, including magnetic, catalytic, electrical, and mechanical features. Compared to bulk materials, nanomaterials behave very differently. They more easily enter the body and couple with biological entities such as protein, cells, and tissues. There is great concern about the health consequences of nanomaterials due to their extremely small size, high specific surface area, quantum and surface boundary effects, and increased surface activity. The nanostructured surfaces can stimulate cells through initial surface protein adsorption, and activate cell adhesion and proliferation. Evidently, engineered NPs can act as a double-edged sword, either as a platform for osseointegration or as a toxic agent, depending on the context. This has led to increased interest in obtaining an understanding of the biological response to NPs. The potential toxicity of engineered NPs on different cells and tissue has been explored for decades and continues to be investigated. Therefore, the goal of this paper is to analyze the biological data about n-HA and TiO$_2$ NPs used as materials of bone implants, to clarify their possible negative effects on the organism and the correlated mechanism.

3. Interaction of NPs with Biological Tissue

3.1. Interaction with Protein

NPs, when entered into the body, will interact initially with proteins present in biological fluids, such as blood. Protein adsorption onto NPs in biological medium has been taken into consideration as an important factor for the assessment of biological responses to NPs [29–31]. Binding to plasma components and forming a nanoparticle-protein corona potentially determine the fate of NPs in the systemic circulation and influence their biological activity [32,33]. In human plasma, a typical protein corona formed on NPs consists of proteins such as serum albumin, immunoglobulins, fibrinogen, and apolipoproteins, etc. Nygren et al. [34] explored the surface-adsorbed plasma protein and cells on the annealed and acid-oxidized TiO$_2$ surface using immunofluorescence technology with short durations of exposure to capillary blood. They detected that serine proteases were the dominating protein adsorbed onto the annealed TiO$_2$ and there was a high amount of fibrinogen on the acid-oxidized surface. Platelets could adhere and spread on the annealed surface within 5 s of blood-material contact. When the platelets adhere to the surface they start the signal cascade that eventually leads to cross-linking of fibrin and subsequently to blood clot formation. Using fresh non-anticoagulated human blood, Ekstrand-Hammarström et al. [35] detected that TiO$_2$ NPs at 50 ng/mL induced strong activation of platelets in the contact system, which could lead to a thrombotic reaction and the generation of bradykinin. Ellingsen [36] examined the reaction of human serum proteins with TiO$_2$ and HA surface-adsorbed calcium ions. They identified that the surface of TiO$_2$ and HA with calcium binding could selectively take up the albumin, prealbumin and IgG from human serum. The binding of TiO$_2$ NPs with human serum albumin involves the van der Waals force and hydrogen bonding formation, and electrostatic interactions might be included [37]. Sousa et al. [38] observed that the
adsorbed human serum albumin easily exchanged with other albumin molecules or albumin itself. The fibrinogen, γ-globulins, and apolipoprotein could also be adsorbed onto TiO\textsubscript{2} NPs with the highest affinity than albumin [39,40]. The quantity of adsorbed protein on the large particles is higher than that adsorbed on the small-sized particles [39]. TiO\textsubscript{2} nanorods evidently adsorb with immunoglobulins IgM and IgG, but TiO\textsubscript{2} nanotubes mainly bind with fibrinogen. Albumin and apolipoprotein can be adsorbed both on the TiO\textsubscript{2} nanorods and on the nanotubes [40]. The surface topography and toughness of TiO\textsubscript{2} nanorods are the only factors that influence the amount of adsorbed fibrinogen [41]. Shen et al. [42] studied the adsorption and desorption behavior of fibronectin on the HA surface by molecular dynamics (MD) and steered MD simulations. The research showed that the electrostatic energy played a dominant role in the interaction between the model protein and the HA surface. The bovine serum albumin (BSA) is physically adsorbed through electrostatic attraction between the COOH group of BSA and the calcium ion exposed on the surface of HA [43], which apparently decreases the activation energy and the adsorption heat [44]. The adsorption of BSA, however, onto titanium is tight at low concentrations because the hydration and electrostatic effects are important in the adsorption process [45]. In general, the surface charge and the hydrodynamic diameter of NPs affected by the particle–protein complexes are likely to be what the cell sees in the cell culture medium [46,47]. NP binding, internalization, transport, and even immune response at cellular-level events are highly influenced by the protein corona [48]. In protein-free medium, L929 mouse fibroblasts can uptake more anatase TiO\textsubscript{2} particles than those in the medium containing human serum albumin. While in the presence of fibrinogen, the particle uptake is lowered by fibroblast cells [39]. Fleischer and Payne [49] reported that in the medium containing BSA, the cellular binding of positively charged BSA–NP complexes was increased, but the cellular binding of negatively charged BSA–NP complexes was inhibited.

3.2. Interaction with Blood Cells

It is generally reported that NPs can enter the vascular system intentionally. As a type of active particles, the interaction of NPs with blood cannot be neglected. The previous investigators demonstrated that NPs may have an effect on blood coagulation and hemolysis [50–52]. Chen et al. [53] evaluated the blood compatibility of n-HA and TiO\textsubscript{2} both in vivo (rats) and in vitro (rabbit erythrocytes). The results showed that HA sol obviously prolonged the time of bleeding, clotting and prothrombin in rats after intravenous injection while TiO\textsubscript{2} sol had no effect. In vitro, both NPs’ sol did not cause hemolysis but did induce the aggregation of rabbit red blood cells.

The blood cell is dominated by erythrocytes (99%), with much fewer leukocytes and platelets (1%). The interaction of NPs with erythrocytes has been investigated both in vivo and in vitro. In a hamster model, Nemmar et al. [54] reported the ultrafine carboxylate-polystyrene particles (60 nm) inhibited thrombus formation, while amine-polystyrene significantly enhanced thrombosis and platelet aggregation at 50 µg/mL. The erythrocytes treated with nano-TiO\textsubscript{2} (20 nm) in vitro underwent abnormal sedimentation, hemagglutination and dose-dependent hemolysis, totally differing from those treated with micro-TiO\textsubscript{2} (200 nm). The ghost cells were observed after exposure to nano-TiO\textsubscript{2}. Han et al. [55] investigated the influence of size and surface charge of HA particles on a suspension of red blood cells (RBCs). n-HA NPs induced an obvious aggregation of RBCs, which was different from the HA microparticles. The negatively charged HA NP’s inhibited the aggregation of RBCs by heparin modification. Therefore, they concluded that the crucial factor influencing the hemocompatibility of n-HA NPs was the surface charge rather than the particle size [55]. However, the ultra-small TiO\textsubscript{2} NPs at 1–3 nm showed no cytotoxicity, no oxidative ability and no genotoxicity on human dermal microvascular endothelial cells (HMEC-1) [56]. In early developing zebrafish, TiO\textsubscript{2} NPs caused mortality and malformations in the form of pericardial edema when injected. The anti-angiogenic effects were also detected both in HMEC-1 cells and in zebrafish embryos, accompanied by a decreased nitric oxide concentration [56]. For the peripheral blood monocytes and lymphocytes, titanium particles induced the production of interleukin (IL)-1β in monocytes, but did not activate lymphocytes because
DNA synthesis and IL-2 secretion were unchanged [57]. The toxic effects of TiO$_2$ NPs on human umbilical vein endothelial cells (HUVECs) were reported by some scientists. Results showed that TiO$_2$ NPs (about 25 nm) at 10, 20, and 40 $\mu$g/mL induced higher lactic dehydrogenase (LDH) and superoxide dismutase (SOD) activities in the culture medium (0 $\mu$g/mL). When HUVECs were exposed to nano-TiO$_2$, IL-6 was significantly released in the medium at 20 and 40 $\mu$g/mL and tumor necrosis factor $\alpha$ (TNF-$\alpha$) was highly released at 5 and 20 $\mu$g/mL [58]. The individual or aggregated TiO$_2$ NPs internalized into HUVECs strongly inhibited cell proliferation and induced apoptosis by 20% and necrotic death by 60%, even at 5 $\mu$g/cm$^2$. TiO$_2$ NPs provoked the activation of HUVECs through an increase in U937 monocyctic adhesion and in the expression of adhesion molecules (E- and P-selectins, ICAM-1, VCAM-1 and PECAM-1), associated with increased reactive oxygen species (ROS) production and nuclear factor (NF)-$\kappa$B pathway activation [59]. A similar outcome on the cytocompatibility assessment of HA NPs was obtained in THP-1 monocyte–HUVEC endothelial cell co-culture models [60]. HA NPs were taken up by both monocytes and HUVECs, causing the indirect activation and proinflammatory effect of HUVECs through p38/JNK mitogen-activated protein kinases (MAPK) and NF-$\kappa$B signaling activation. The n-HA resulted in increased total oxidative stress levels, decreased antioxidant capacity, and increased genotoxic effects (increase of sister chromatid exchange, micronuclei, chromosome aberration rates and 8-oxo-2-deoxyguanosine levels) in primary human blood cells in a dose-dependent manner [61]. The potential interaction and adverse effects of NPs on blood cells and cells in the vascular system suggest that NPs could increase the risk of vascular disease and potentially damage the main tissues in the organism through blood circulation.

3.3. Local Toxicity

Generally, there are two kinds of tissue responses to particles, the macrophage-mediated response and the lymphocyte-dominated response. In the periprosthetic tissue, Lohmann et al. [62] found macrophages containing metal particles and perivascular lymphocytic infiltration and fibrin exudation by histological examination in failed metal-on-metal total hip arthroplasties. By intra-articular injection of TiO$_2$ NPs to simulate the release of wear NPs into the joint cavity, Wang et al. [63] revealed that the smaller aggregated TiO$_2$ NPs penetrated the synovial capillaries and were transported to the heart or lung tissues of rats with blood circulation, and the larger aggregated TiO$_2$ particles were deposited in the knee joint, resulting in the injury of the local tissues. For the synovium, the intra-articular TiO$_2$ NPs induced fibroblast-like synoviocyte proliferation, lymphocyte and plasma cell infiltration and synovium hypertrophy [63]. The aseptic lesion and enlargement of soft tissue called a pseudotumor is observed by clinicians in the knee/hip joint with prostheses, composed of fibrous connective tissue infiltrated by immunocompetent cells, fibrin deposition and necrosis. Based on the histological changes in periprosthetic tissues, the cause of pseudotumors is thought to be either cytotoxicity of wear particles or a delayed hypersensitivity reaction (DHR or type IV) [64,65]. DHR is not an antibody-mediated response but a type of cell-mediated response with a large number of T cell responses to an external or internal agent.

In the joint, the articular cartilage is avascular and alymphatic and is innervated. The chondrocyte is the only living element of the cartilage. The effect of wear debris on the articular cartilage is investigated through the intra-articular injection of TiO$_2$ nanoparticles. The authors [66] detected the decreased thickness of articular cartilage in rats with TiO$_2$ NPs at post-exposure days 1, 7, 14, and 30 in a time-dependent manner using the contrast-enhanced high resolution micro-computed tomography. The decreased cartilage volume was detected too. The histopathological changes showed that the chondrocytes had edema with shrunken nuclei in the radial and calcified zone of the cartilage. The cartilage ultrastructure observed by transmission electron microscopy showed the degenerated chondrocytes with condensed chromatin, a dilated endoplasmic reticulum, and rich mitochondria [66].

The wear NPs released from metal-on-metal and metal-on-polyethylene implants circulate in the organism both locally and systemically, penetrate the cell membrane, couple with cellular proteins, and mediate the inflammatory response and expression of cytokines [67]. The macrophages, fibroblasts,
and lymphocytes can phagocytose wear debris particles produced from the prosthesis. Titanium and cobalt-chromium particles alter the immune responses when injected into the peritoneal cavity of female mice. At 8 and 12 weeks of injection, titanium and cobalt-chromium particles inhibited cytokine release by lymphocytes (IL-2, IL-4, IFN-γ), proliferation of T and B cells, and immunoglobulin production by B cells. However, these particles are not cytotoxic to murine lymphocytes [68].

The activated macrophage exhibits a spectrum of polarization states, the classically activated M1 phenotype by pro-inflammatory signals (e.g., TNF-α) and the alternatively activated M2 state by anti-inflammatory signals (e.g., IL-4). The proinflammatory signals stimulate the expression of nuclear factor kappa-B ligand (RANKL) on the surface of osteoblasts, and increase the RANKL/OPG (osteoprotegerin) ratio and ROS production by NADPH-oxidase. RANKL interacts with RANK to activate osteoclasts and regulate bone resorption [69,70]. It is also reported that resident macrophages can differentiate into multinucleated functional osteoclasts with exposure to wear particle-mediated cytokine products in an in vitro experiment [71].

The inflammatory responses occur and the cytokines are produced including TNF-α, IL-1α, IL-1β, and IL-6. These cytokines stimulate osteoblasts to release soluble cytokines, such as granulocyte-macrophage colony-stimulating factor (GM-CSF), RANKL, IL-6 and prostaglandin E2 (PGE2), or they indirectly stimulate osteoclasts to activate osteolysis [72–74]. As a result, the osteogenic activity of osteoblasts is inhibited and predominated by bone resorption at the bone-implant interface. The human synovial cells exhibit a 1.72-fold increase in matrix metalloproteinase (MMP)-2 activity when exposed to a Ti disc and a 3.95-fold increase when exposed to Ti particles [75,76]. Although the cytokine IL-1 only induces a 6.76-fold increase in MMP-2 activity, the combination of Ti particles and inflammatory cytokines induces a 10.54-fold increase of MMP-2 activity. Cytokines released from macrophages and MMPs secreted by fibroblasts can affect the attachment and synthetic activities of osteoblasts and cause the reduction of the bone matrix. The ROS is over-produced in the rat synovial cell line 364 (RSC-364) especially when exposed to 30 and 300 µg/mL anatase TiO₂ NPs [77,78]. The significantly decreased activities of superoxide dismutase (SOD), intracellular glutathione (GSH) and catalase (CAT) enzymes and the increased lipid peroxidation product (malondiadehyde, MDA) are induced, which means there is oxidative stress and oxidative damage in RSC-364 cells. Choi et al. [79] determined the effect of various sizes of Ti particles on the bone-remodeling process both in vivo and in vitro. By injecting Ti particles with different sizes into the spaces surrounding the Ti alloy pins, they demonstrated that the osteoblast function significantly attenuated with the increased expression of the receptor activator of RANKL, which was a dominant signal for osteoclast recruitment and depended on the size of the Ti particle. Therefore, the chronic proinflammatory response induced by wear particles has a negatively effect on osteoblast function, and decreases bone formation and osseointegration. This maybe a major factor in osteolysis and the subsequent aseptic loosening of arthroplasty implants [80].

3.4. Dissemination and Systemic Toxicity

The translocation and accumulation of NPs would occur in the body regardless of the entry routes. There is evidence that NPs can reach and accumulate in the second target organs across body membranes, such as the lung, heart, liver, spleen, and brain. NPs used for bone repair and reconstruction, without exception, would diffuse and redistribute with blood circulation and be entrapped by the reticuloendothelial system (RES). Urban et al. [81] firstly reported that metallic wear particles (less than 1 µm) were detected in the paraaortic lymph nodes and further disseminated to the liver and spleen in patients with a failed hip arthroplasty. The submicrometer metal wear particles generated at nonbearing surfaces were identified within macrophages in the liver and spleen of patients with a revised arthroplasty and with primary hip arthroplasty [82]. Using I-125 radiolabeling on n-HA at 80 nm, Sun et al. [83] quantitatively analyzed that intravenously administrated n-HA was distributed everywhere in the body with blood circulation, but mainly accumulated in the lung, liver and spleen for over one month.
It is well known that the liver, spleen and kidney are the important metabolic and excretory organs. The special biochemical parameters in serum partially reflect the function of these organs, such as alanine aminotransferase (ALT) and aspartate aminotransferase (AST) for liver function, blood urea nitrogen (BUN) for kidney function and alkaline phosphatase (ALP) for liver function or bone formation–related diseases, and so on. Liu et al. [84] found that the n-HAs with a rod shape resulted in acute increases in ALT, AST, BUN and ALP in rabbits after injection by vein. BUN and ALP reached a peak 24 h later, and then decreased rapidly to normal levels. The intraperitoneally injected rod-shaped n-HA (about 20–30 nm in length and 5 nm in width) can be uptaken and disseminate to liver and kidney tissues with the blood circulation; it does not induce the AST, ALT and BUN changes in the serum of rats, but produces apoptosis in the liver cells and renal tubular epithelial cells [85]. However, Wang et al. [86] detected that the nano-hydroxyapatite/chitosan (n-HA/CS) composite induced the significant elevation of BUN, CR and total bilirubin (T-Bil) in the serum of SD rats as well as the apoptosis in the liver and kidney tissues with no inflammation and necrosis at eight weeks of exposure by intraperitoneally injection. Sun et al. [83] reported that n-HA particles could inhibit the growth of cells and induce cell toxicity in different tissues after intravenous administration, which might be closely related with the residence time of nanoparticles in tissues and the metabolism characteristics of organs.

The translocation of TiO$_2$ NPs released from implants is investigated in rats by intra-articular injection [63]. The results demonstrate that the intra-articularly injected TiO$_2$ NPs can potentially affect major organs such as the heart, lung and liver, which indicated the circulation of intra-articular NPs from the joint cavity to the system [63]. The slightly pathological change of the heart, lung, and liver is induced by 0.2 mg/kg TiO$_2$ and the severe pathological impairment of major organs resulted in the rats after exposure to 2 and 20 mg/kg TiO$_2$ suspensions. In the lung tissue, the particle-laden macrophages and the inflammatory cells, such as neutrophils, lymphocytes and eosinophils, are observed in the pulmonary alveoli with thickened alveolar walls. The fatty degeneration of hepatocytes and inflammatory cell infiltration of the portal area prove the injury of the liver. The difficult clearance of TiO$_2$ NPs in the lung and liver results in the increased coefficients of organs and the significant upregulation of the serum AST/ALT level, especially in the 20 mg/kg group.

Overall, regardless of whether the NPs were injected or were produced from the prosthesis, once they entered the body, they would circulate everywhere along with the blood circulation and deposit to interact with cells or tissues, leading to the injury of some organs.

3.5. Immune Response and Oxidative Stress

As stated above, wear debris particles can activate an inflammatory response locally and systemically. This is mediated by monocytes/macrophages, lymphocytes, synoviocytes, osteoblasts and fibroblasts to secret the products of the proinflammatory cytokines, such as PGE2, IL-6, IL-1$\beta$, IL-18, TNF-\(\alpha\) [73,87,88], and reactive oxygen intermediates, such as ROS and reactive nitrogen species (RNS). The nanomaterials induce inflammation mainly through oxidative stress, especially for inorganic NPs and metal oxide NPs [89,90]. Both TiO$_2$ and HA NPs cause an elevated ROS level and expression of inflammatory transcripts in human oral epithelial cells [91]. Oxidative stress is defined as an imbalance between oxidants and antioxidants and is commonly known to play a role in the formation of a fibrous pseudocapsule around hip implants and to exert negative biological effects [92,93]. Kinov et al. [94] detected the high glutathione disulfide (GSSG) and MDA level and low GSH/GSSG ratio in total knee arthroplasties with high wear and osteolysis, suggesting that ROS may be involved in the formation of fibrotic pseudocapsular tissues. In arthrofibrotic tissues, Freeman et al. [95] analyzed that the elevated amounts of myeloperoxidase, an enzyme that generates ROS/RNA, downregulated the expression of SOD, glutathione S-transferase (GST) and antioxidants and mediated the oxidative DNA and matrix protein modifications (DNA hydroxylation and protein nitrosylation). The over-production of ROS is implicated in the degeneration and necrosis of tissues in fibrous pseudocapsule tissues, and even in diseases. Ding [96] revealed that nano-HA with different sizes could induce pseudo-tubercles in the
lung. Additionally, the nanoscale particles with a small size resulted in a vacuolar degeneration of the nephric tubule epithelium after intravenous injection for seven days. In the retrieved hip tissues from total hip arthroplasty patients, the degree of osteolysis shows a correlation with high mobility group protein-B1 (HMGB1), cyclooxygenase-2 (COX2), and 4-hydroxynonenal (4-HNE). This demonstrates that the wear particle–induced oxidative stress mediates bone resorption and osteolysis [97,98].

Many reports state that the structural features of nanoparticles including physical and chemical properties, such as size, shape, crystal phases and surface coating, can exert cytotoxicity through oxidative stress [87,99,100]. Wang and Fan [99] elaborated in detail on the correlation between lung toxicity and pulmonary cell impairment related to TiO$_2$ NPs and its unusual physicochemical characteristics, including the size, shape, surface coating, and crystal phases. The ROS mechanism is also elucidated in the toxicity of TiO$_2$ NPs. Nano-HA with rod-, spherical-, or needle-shaped crystals can inhibit the growth of primary rat osteoblasts in a dose-dependent manner and induce apoptosis via p53 and the cytochrome C-dependent mitochondrial pathways [101]. Zhao et al. [102] demonstrated that nano-HA with smaller specific surface areas induced lower apoptosis, but HA with higher surface area increased the cell-particle interaction and elevated the ROS generation.

The overproduction of ROS is thought of as the best-developed paradigm for the toxicity of NPs [92,93]. With its increase, ROS tends to initiate the significant damage of cell structures and trigger an inflammatory response via the oxidative stress–responsive MAPK, redox-responsive NF-$\kappa$B and activator protein-1 (AP-1) signaling pathways [103]. Further, wear particle–induced ROS elevation also activates the endoplasmic reticulum (ER) stress markers and promotes apoptosis in osteolysis [104,105]. Yang et al. [106] found the phagocytosed wear particles induced the high expression of apoptosis-related markers iNOS, ONOO$^-$, cleaved caspase-3/4/8/9, cytochrome C, glucose regulated protein 78 (GRP78), and growth arrest and DNA damage-inducible gene 153 (GADD153) in macrophages in the periprosthetic interface membrane, proving that the ER stress pathways were the apoptotic pathways of macrophages in the interface membrane of aseptic loosening with the exception of the death receptor pathway and the mitochondrial caspase-dependent pathway. ROS also can improve nanoparticle-induced genotoxicity, which is characterized by chromosomal aberrations, DNA strand breaks, oxidative DNA damage, and mutations [107,108]. In human blood cells, the increases of sister chromatid exchange, micronuclei, chromosome aberration rates and 8-oxo-2-deoxyguanosine levels were caused by HA NPs in a dose–dependent manner [61]. The micronucleus test only showed the type-III foci formation in mouse fibroblasts after exposure to rutile TiO$_2$ NPs [109], while there was no increase in DNA damage in human fibroblasts at sub-cytotoxic concentrations [110].

### 4. Conclusions

Depending on good biocompatibility and osteoconductivity, nano-HA and TiO$_2$ NPs are generously exploited to increase the repair and reconstruction of bone. However, because of their small size and high surface activity, the adverse biological effects of nanoparticles are an unavoidable scientific problem, which gains much attention from clinicians and scientists. The wear nanoparticles would be prone to adsorb blood proteins to form protein–particle complexes, and further to be phagocytosed by macrophages or fibroblasts and to deposit in the local tissue, leading to delayed hypersensitivity reactions. Meanwhile, the dissemination of nanoparticles into the main organs such as the lung, liver and spleen occurs in the blood circulation system. The inflammatory response, oxidative stress and signaling pathways are elaborated to analyze the toxicological mechanism. Inhibition of redox-related signaling transduction may be a new therapeutic strategy for wear debris–mediated osteolysis. Developing biomimetic materials with better biocompatibility is our goal for orthopedic implants.

**Acknowledgments:** This work was financially supported by the National Key Technology R & D Program of China (973 Program, 2011CB70901), the National Natural Science Foundation of China (NSFC) Research Grant (31271008, 11120101001, 61272902, 11421202), the 111 Project (B13003), the International Joint Research Center of Aerospace Biotechnology and the Medical Engineering, Ministry of Science and Technology of China, Specialized
Research Fund for the Doctoral Program of Higher Education, and National High Technology Research and Development Program of China (863 program, 2011AA02A102).

**Author Contributions:** Yubo Fan and Jiangxue Wang proposed the conception and designed the paper; Jiangxue Wang and Liting Wang collected and analyzed the data; Jiangxue Wang and Liting Wang wrote the paper; Yubo Fan conceived the principal idea and revised the paper.

**Conflicts of Interest:** The authors declare no conflict of interest.

**References**

1. Christenson, E.M.; Anseth, K.S.; van den Beucken, L.; Chan, C.K.; Erkan, B.; Jansen, J.A.; Laurencin, C.T.; Li, W.J.; Murugan, R.; Nair, L.S.; *et al.* Nanobiomaterial applications in orthopedics. *J. Orthop. Res.* 2007, 25, 11–22. [CrossRef] [PubMed]

2. Engel, E.; Michiardi, A.; Navarro, M.; Lacroix, D.; Planell, J.A. Nanotechnology in regenerative medicine: the materials side. *Trends Biotechnol.* 2008, 26, 39–47. [CrossRef] [PubMed]

3. Sato, M.; Webster, T.J. Nanobiotechnology: Implications for the future of nanotechnology in orthopedic applications. *Expert Rev. Med. Devices* 2004, 1, 105–114. [CrossRef] [PubMed]

4. McMahon, R.E.; Wang, L.; Skoracki, R.; Mathur, A.B. Development of nanomaterials for bone repair and regeneration. *J. Biomed. Mater. Res.* 2013, 101B, 387–397. [CrossRef] [PubMed]

5. Smith, R. Bone Health and Osteoporosis: A Report of the Surgeon General. In *US Department of Health and Human Services*; Rockville, M., Ed.; Public Health Service: Washington, DC, USA, 2004; pp. 68–70.

6. Dorozhkin, S.V. Calcium orthophosphates in nature, biology and medicine. *Materials* 2009, 2, 399–498. [CrossRef]

7. Gong, T.; Xie, J.; Liao, J.; Zhang, T.; Lin, S.; Lin, Y. Nanomaterials and bone regeneration. *Bone Res.* 2015, 3, 15029. [CrossRef] [PubMed]

8. Huebsch, N.; Mooney, D.J. Inspiration and application in the evolution of biomaterials. *Nature* 2009, 462, 426–432. [CrossRef] [PubMed]

9. Wang, T.; Yang, X.; Qi, X.; Jiang, C. Osteoinduction and proliferation of bone-marrow stromal cells in three-dimensional poly(epsilon-caprolactone)/hydroxyapatite/collagen scaffolds. *J. Transl. Med.* 2015, 13, 152. [CrossRef] [PubMed]

10. Kane, R.; Ma, P.X. Mimicking the nanostructure of bone matrix to regenerate bone. *Mater. Today* 2013, 16, 418–423. [CrossRef] [PubMed]

11. Gupta, D.; Venugopal, J.; Mitra, S.; Giri Dev, V.R.; Ramakrishna, S. Nanostructured biocomposite substrates by electrospinning and electrospraying for the mineralization of osteoblasts. *Biomaterials* 2009, 30, 2085–2094. [CrossRef] [PubMed]

12. Guo, B.; Lei, B.; Li, P.; Ma, P.X. Functionalized scaffolds to enhance tissue regeneration. *Regen. Biomater.* 2015, 2, 47–57. [CrossRef] [PubMed]

13. Kim, H.W.; Kim, H.E.; Salih, V. Stimulation of osteoblast responses to biomimetic nanocomposites of gelatin-hydroxyapatite for tissue engineering scaffolds. *Biomaterials* 2005, 26, 5221–5230. [CrossRef] [PubMed]

14. Rong, Z.J.; Yang, L.; Cai, B.T.; Zhu, L.X.; Cao, Y.L.; Wu, G.F.; Zhang, Z.J. Porous nano-hydroxyapatite/collagen scaffold containing drug-loaded ADM-PLGA microspheres for bone cancer treatment. *J. Mater. Sci. Mater. Med.* 2016, 27, 1–12. [CrossRef] [PubMed]

15. Tsukimura, N.; Kojima, N.; Kubo, K.; Att, W.; Takeuchi, K.; Kameyama, Y.; Maeda, H.; Ogawa, T. The effect of superficial chemistry of titanium on osteoblastic function. *J. Biomed. Mater. Res. A* 2008, 84, 108–116. [CrossRef] [PubMed]

16. Wang, N.; Li, H.; Li, W.; Li, J.; Wang, J.; Zhang, Z.; Liu, Y. Effects of TiO$_2$ nanotubes with different diameters on gene expression and osseointegration of implants in minipigs. *Biomaterials* 2011, 32, 6900–6911. [CrossRef] [PubMed]

17. Brammer, K.S.; Oh, S.; Cobb, C.J.; Bjursten, L.M.; van der Heyde, H.; Jin, S. Improved bone-forming functionality on diameter-controlled TiO$_2$ nanotube surface. *Acta Biomater.* 2009, 5, 3215–3223. [CrossRef] [PubMed]

18. Tainen, H.; Wohlfahrt, J.C.; Verket, A.; Lyngstadaas, S.P.; Haugen, H.J. Bone formation in TiO$_2$ bone scaffolds in extraction sockets of minipigs. *Acta Biomater.* 2012, 8, 2384–2391. [CrossRef] [PubMed]
19. Webster, T.J.; Ergun, C.; Doremus, R.H.; Siegel, R.W.; Bizios, R. Enhanced functions of osteoblasts on nanophase ceramics. *Biomaterials* 2000, 21, 1803–1810. [CrossRef]
20. Webster, T.J.; Siegel, R.W.; Bizios, R. Osteoblast adhesion on nanophase ceramics. *Biomaterials* 1999, 20, 1221–1227. [CrossRef]
21. Hashimoto, M.; Takadama, H.; Mizuno, M.; Kokubo, T. Mechanical properties and apatite forming ability of TiO$_2$ nanoparticles/high density polyethylene composite: Effect of filler content. *J. Mater. Sci. Mater. Med.* 2007, 18, 661–668. [CrossRef] [PubMed]
22. Webster, T.J.; Smith, T.A. Increased osteoblast function on PLGA composites containing nanophase titania. *J. Biomed. Mater. Res. A* 2005, 74, 677–686. [CrossRef] [PubMed]
23. Brammer, K.S.; Frandsen, C.J.; Jin, S. TiO$_2$ nanotubes for bone regeneration. *Trends Biotechnol.* 2012, 30, 315–322. [CrossRef] [PubMed]
24. Wang, J.; Li, H.; Sun, Y.; Bai, B.; Zhang, Y.; Fan, Y. Anodization of highly ordered TiO$_2$ nanotube arrays using orthogonal design and its wettability. *Int. J. Electrochem. Sci.* 2011, 6, 710–723.
25. Bjursten, L.M.; Rasmusson, L.; Oh, S.; Smith, G.C.; Brammer, K.S.; Jin, S. Titanium dioxide nanotubes enhance bone bonding in vivo. *J. Biomed. Mater. Res. A* 2010, 92, 1218–1224. [PubMed]
26. Mendonça, G.; Mendonça, D.B.S.; Simões, L.G.P.; Araújo, A.L.; Leite, E.R.; Duarte, W.R.; Aragão, F.J.L.; Cooper, L.F. The effects of implant surface nanoscale features on osteoblast-specific gene expression. *Biomaterials* 2009, 30, 4053–4062. [CrossRef] [PubMed]
27. Bhardwaj, G.; Webster, T.J. Functionalized nanophase hydroxyapatite (HA) for orthopedic applications. In *Proceedings of the 40th Annual Northeast Bioengineering Conference (NEBEC)*, Boston, MA, USA, 25–27 April 2014; pp. 1–2.
28. Kazemzadeh-Narbat, M.; Lai, B.F.L.; Ding, C.; Kizhakkedathu, J.N.; Hancock, R.E.W.; Wang, R. Multilayered coating on titanium for controlled release of antimicrobial peptides for the prevention of implant-associated infections. *Biomaterials* 2013, 34, 5969–5977. [CrossRef] [PubMed]
29. Cedervall, T.; Lynch, I.; Lindman, S.; Berggård, T.; Thulin, E.; Nilsson, H.; Dawson, K.A.; Linse, S. Understanding the nanoparticle–protein corona using methods to quantify exchange rates and affinities of proteins for nanoparticles. *Proc. Natl. Acad. Sci. USA* 2007, 104, 2050–2055. [CrossRef] [PubMed]
30. Lundqvist, M.; Sethson, I.; Jonsson, B.-H. Protein adsorption onto silica nanoparticles: Conformational changes depend on the particles’ curvature and the protein stability. *Langmuir* 2004, 20, 10639–10647. [CrossRef] [PubMed]
31. Lynch, I.; Dawson, K.A. Protein-nanoparticle interactions. *Nanotoday* 2008, 3, 40–47. [CrossRef]
32. Pino, P.D.; Pelaz, B.; Zhang, Q.; Maffre, P.; Nienhaus, G.U.; Parak, W.J. Protein corona formation around nanoparticles—from the past to the future. *Mater. Horiz.* 2014, 1, 301–313. [CrossRef]
33. Saptarshi, S.R.; Duschl, A.; Lopata, A.L. Interaction of nanoparticles with proteins: relation to bio-reactivity of the nanoparticle. *J. Nanobiotechnol.* 2013, 11, 26. [CrossRef] [PubMed]
34. Nygren, H.; Tengvall, P.; Lundström, I. The initial reactions of TiO$_2$ with blood. *J. Biomed. Mater. Res.* 1997, 34, 487–492. [CrossRef]
35. Ekstrand-Hammarström, B.; Hong, J.; Davoodpour, P.; Sandholm, K.; Ekdahl, K.N.; Bucht, A.; Nilsson, B. TiO$_2$ nanoparticles tested in a novel screening whole human blood model of toxicity trigger adverse activation of the kallikrein system at low concentrations. *Biomaterials* 2015, 51, 58–68. [CrossRef] [PubMed]
36. Ellingsen, J.E. A study on the mechanism of protein adsorption to TiO$_2$. *Biomaterials* 1991, 12, 593–596. [CrossRef]
37. Cooper, L.F. The effects of implant surface nanoscale features on osteoblast-specific gene expression. *Int. J. Mol. Sci.* 2016, 17, 798.
41. Chen, H.P.; Chen, H.L.; Chen, D.H.; Chen, M. Synergistic effect of carbon microstructure and topography of TiO$_2$ nanorod arrays on hemocompatibility of carbon/TiO$_2$ nanorod arrays composites. J. Mater. Sci. 2014, 49, 5299–5308. [CrossRef]

42. Shen, J.-W.; Wu, T.; Wang, Q.; Pan, H.-H. Molecular simulation of protein adsorption and desorption on hydroxyapatite surfaces. Biomaterials 2008, 29, 513–532. [CrossRef] [PubMed]

43. Wassell, D.T.H.; Hall, R.C.; Embery, G. Adsorption of bovine serum albumin onto hydroxyapatite. Biomaterials 1995, 16, 697–702. [CrossRef]

44. Yin, G.; Liu, Z.; Zhan, J.; Ding, F.; Yuan, N. Impacts of the surface charge property on protein adsorption on hydroxyapatite. Chem. Eng. J. 2002, 87, 181–186. [CrossRef]

45. Wassell, D.T.H.; Embery, G. Adsorption of bovine serum albumin onto titanium powder. Biomaterials 1996, 17, 859–864. [CrossRef]

46. Walczyk, D.; Bombelli, F.B.; Monopoli, M.P.; Lynch, I.; Dawson, K.A. What the Cell “Sees” in Bionanoscience. J. Am. Chem. Soc. 2010, 132, 5761–5768. [CrossRef] [PubMed]

47. Lynch, I.; Salvati, A.; Dawson, K.A. Protein-nanoparticle interactions: What does the cell see? Nat. Nanotechnol. 2009, 4, 546–547. [CrossRef] [PubMed]

48. Yan, Y.; Gause, K.T.; Kamphuis, M.M.J.; Ang, C.-S.; O’Brien-Simpson, N.M.; Lenzo, J.C.; Reynolds, E.C.; Nice, E.C.; Caruso, F. Differential roles of the protein corona in the cellular uptake of nanoporous polymer particles by monocyte and macrophage cell lines. ACS Nano 2013, 7, 10960–10970. [CrossRef] [PubMed]

49. Fleischer, C.C.; Payne, C.K. Nanoparticle–cell interactions: Molecular structure of the protein corona and cellular outcomes. Acc. Chem. Res. 2014, 47, 2651–2659. [CrossRef] [PubMed]

50. Mayer, A.; Vadon, M.; Rinner, B.; Novak, A.; Wintersteiger, R.; Frohlich, E. The role of nanoparticle size in hemocompatibility. Toxicology 2009, 258, 139–147. [CrossRef] [PubMed]

51. Aisaka, Y.; Kawaguchi, R.; Watanabe, S.; Ikeda, M.; Igisu, H. Hemolysis caused by titanium dioxide particles. Inhal. Toxicol. 2008, 20, 891–893. [CrossRef] [PubMed]

52. Ghosh, M.; Chakraborty, A.; Mukherjee, A. Cytotoxic, genotoxic and the hemolytic effect of titanium dioxide (TiO$_2$) nanoparticles on human erythrocyte and lymphocyte cells in vitro. J. Appl. Toxicol. 2013, 33, 1097–1110. [CrossRef] [PubMed]

53. Chen, X.; Feng, L.; Peng, R.; Cao, X. Studies on nano-particle sols of hydroxyapatite and titanium dioxide for haemo-compatibility. Wei Sheng Yan Jiu J. Hyg. 2002, 31, 197–199.

54. Nemmar, A.; Hoylaerts, M.F.; Hoet, P.H.M.; Dinsdale, D.; Smith, T.; Xu, H.; Vermullen, J.; Nemery, B. Ultradefine particles affect experimental thrombosis in an in vivo hamster model. Am. J. Respir. Crit. Care Med. 2002, 166, 998–1004. [CrossRef] [PubMed]

55. Han, Y.C.; Wang, X.Y.; Dai, H.L.; Li, S.P. Nanosize and surface charge effects of hydroxyapatite nanoparticles on red blood cell suspensions. ACS Appl. Mater. Interfaces 2012, 4, 4616–4622. [CrossRef] [PubMed]

56. Bayat, N.; Lopes, V.R.; Schoelermann, J.; Jensen, L.D.; Cristobal, S. Vascular toxicity of ultra-small TiO$_2$ nanoparticles and single walled carbon nanotubes in vitro and in vivo. Biomaterials 2015, 63, 1–13. [CrossRef] [PubMed]

57. Kohilas, K.; Lyons, M.; Loffthouse, R.; Frondoza, C.G.; Jinnah, R.; Hungerford, D.S. Effect of prosthetic titanium wear debris on mitogen-induced monocyte and lymphoid activation. J. Biomed. Mater. Res. 1999, 47, 95–103. [CrossRef]

58. Yan, Q.Q.; Yang, L.; Zhao, J.; Li, J.; Wang, Z.L. Comparative experiment on nanoparticle-induced toxicity in human vascular endothelial cells. Zhonghua Lao Dong Wei Sheng Zhi Ye Bing Chin. J. Ind. Hyg. Occup. Dis. 2012, 30, 820–824.

59. Montiel-Dávalos, A.; Ventura-Gallegos, J.L.; Alfaro-Moreno, E.; Soria-Castro, E.; García-Latorre, E.; Cabañas-Moreno, J.G.; Ramos-Godinez, M.D.P.; López-Marure, R. TiO$_2$ nanoparticles induce dysfunction and activation of human endothelial cells. Chem. Res. Toxicol. 2012, 25, 920–930. [CrossRef] [PubMed]

60. Liu, X.; Sun, J. Potential proinflammatory effects of hydroxyapatite nanoparticles on endothelial cells in a monocyte–endothelial cell coculture model. Int. J. Nanomed. 2014, 9, 1261–1273.

61. Turketz, H.; Yousef, M.J.; Sonmez, E.; Togar, B.; Bakan, F.; Sozio, P.; Stefano, A.D. Evaluation of cytotoxic, oxidative stress and genotoxic responses of hydroxyapatite nanoparticles on human blood cells. J. Appl. Toxicol. 2014, 34, 373–379. [CrossRef] [PubMed]
62. Lohmann, C.H.; Meyer, H.; Nuechtern, J.V.; Singh, G.; Junk-Jantsch, S.; Schmotzer, H.; Morlock, M.M.; Pfüiger, G. Periprosthetic metal response to retrieved nanoscale metal oxide particles in total hip arthroplasties. *J. Bone Jt. Surg.* 2013, 95, 1561–1568. [CrossRef] [PubMed]

63. Wang, J.; Fan, Y.; Gao, Y.; Hu, Q.; Wang, T. TiO₂ nanoparticles translocation and potential toxicological effect in rats after intraarticular injection. *Biomaterials* 2009, 30, 4590–4600. [CrossRef] [PubMed]

64. Hallab, N.; Merritt, K.; Jacobs, J.J. Metal sensitivity in patients with orthopaedic implants. *J. Bone Jt. Surg. Am.* 2001, 83, 428–436.

65. Mahendra, G.; Pandit, H.; Klixsey, K.; Murray, D.; Gill, H.S.; Athanasou, N. Necrotic and inflammatory changes in metal-on-metal resurfacing hip arthroplasties: Relation to implant failure and pseudotumor formation. *Acta Orthop.* 2009, 80, 653–659. [CrossRef] [PubMed]

66. Wang, J.; Fan, Y.; Gao, Y.; Hu, Q.; Wang, T. TiO₂ nanoparticles up-regulate the activity of matrix metalloproteinase-2 in human synovial cells. *Calcif. Tissue Int.* 1996, 59, 392–396. [CrossRef] [PubMed]

67. Polyzois, I.; Nikolopoulos, D.; Michos, I.; Patsouris, E.; Theocharis, S. Local and systemic toxicity of nanoscale debris particles in total hip arthroplasty. *J. Appl. Toxicol.* 2012, 32, 255–269. [CrossRef] [PubMed]

68. Wang, J.Y.; Wicklund, B.H.; Gustilo, R.B.; Tsukayama, D.T. Prosthetic metal impairs murine immune response and cytokine release in vivo and in vitro. *J. Orthop. Res.* 1997, 15, 688–699. [CrossRef] [PubMed]

69. Nich, C.; Takakubo, Y.; Pajarinen, J.; Ainola, M.; Salem, A.; Sillat, T.; Rao, A.J.; Raska, M.; Tamaki, Y.; Takagi, M.; et al. Macrophages-Key cells in the response to wear debris from joint replacements. *J. Biomed. Mater. Res. A* 2013, 101, 3033–3045. [CrossRef] [PubMed]

70. Mantovani, A.; Sica, A.; Sozzani, S.; Allavena, P.; Vecchi, A.; Locati, M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol.* 2004, 25, 677–686. [CrossRef] [PubMed]

71. Sabokbar, A.; Fujikawa, Y.; Neale, S.; Murray, D.W.; Athanasou, N.A. Human arthroplasty derived macrophages differentiate into osteoclastic bone resorbing cells. *Ann. Rheum. Dis.* 1997, 56, 414–420. [CrossRef] [PubMed]

72. Horowitz, M.S.; Gonzales, B.J. Inflammatory response to implant particulates in a macrophage/osteoblast coculture model. *Calcif. Tissue Int.* 1996, 59, 392–396. [CrossRef] [PubMed]

73. Maloney, W.J.; James, R.E.; Smith, R.L. Human macrophage response to retrieved titanium alloy particles in vitro. *Clin. Orthop. Relat. Res.* 1996, 322, 268–278. [CrossRef] [PubMed]

74. Song, W.; Wang, J.; Liu, M.; Li, P.; Zhou, G.; Li, Z.; Fan, Y. Titanium dioxide nanoparticles induced proinflammation of primary cultured cardiac myocytes of rat. *J. Nanomater.* 2013, 2013, 349140. [CrossRef]

75. Fu, C.; Xie, J.; Hu, N.; Liang, X.; Chen, R.; Wang, C.; Chen, C.; Xu, C.; Huang, W.; Paul Sung, K.L. Titanium particles up-regulate the activity of matrix metalloproteinase-2 in human synovial cells. *Int. Orthop.* 2014, 38, 1091–1098. [CrossRef] [PubMed]

76. Fu, C.; Xie, J.; Chen, R.; Wang, C.; Xu, C.; Chen, C.; Wang, Z.; Lin, L.; Huang, W.; Liang, X.; Paul Sung, K.L. Effect of titanium particles and TNF-α on the gene expression and activity of MMP-1,2,3 in human knee joint synovial cells. *J. Biomed. Eng.* 2013, 30, 1022–1026.

77. Wang, J.; Ma, J.; Dong, L.; Hou, Y.; Jia, X.; Niu, X.; Fan, Y. Effect of anatase TiO₂ nanoparticles on the growth of RSC-364 rat synovial cell. *J. Nanosci. Nanotechnol.* 2013, 13, 3874–3879. [CrossRef] [PubMed]

78. Wang, J.; Hou, Y.; Dong, L.; Niu, X.; Fan, Y. Influence of TiO₂ nanoparticles on glutathione in rat synovial cell line RSC-364. In *World Congress on Medical Physics and Biomedical Engineering*; Long, M., Ed.; Springer-Verlag Berlin Heidelberg: Berlin, Germany, 2012; pp. 75–78.

79. Choi, M.G.; Koh, H.S.; Kluess, D.; O’Connor, D.; Mathur, A.; Truskey, G.A.; Rubin, J.; Zhou, D.X.F.; Sung, K.L.P. Effects of titanium particle size on osteoblast functions in vitro and in vivo. Proc. Natl. Acad. Sci. USA 2005, 102, 4578–4583. [CrossRef] [PubMed]

80. Jacobs, J.J.; Hallab, N.J. Loosening and osteolysis associated with metal-on-metal bearings: A local effect of metal hypersensitivity? *J. Bone Jt. Surg.* 2006, 88, 1171–1172. [CrossRef] [PubMed]

81. Urban, R.M.; Tomlinson, M.J.; Hall, D.J.; Jacobs, J.J. Migration of corrosion products from modular hip prostheses. Particle microanalysis and histopathological findings. *J. Bone Jt. Surg. Am.* 1994, 76, 1345–1359.

82. Urban, R.M.; Tomlinson, M.J.; Hall, D.J.; Jacobs, J.J. Accumulation in liver and spleen of metal particles generated at nonbearing surfaces in hip arthroplasty. *J. Arthroplast.* 2004, 19, 94–101. [CrossRef]

83. Sun, J.; Xie, G. Tissue distribution of intravenously administrated hydroxyapatite nanoparticles labeled with I-125. *J. Nanosci. Nanotechnol.* 2011, 11, 10996–11000. [CrossRef] [PubMed]
84. Liu, L.; Xiao, Y.; Xiao, Z.; Wang, Z.; Li, C.; Gong, X. Toxicity of hydroxyapatite nanoparticles on rabbits. *Wei Sheng Yan Jiu 2005*, 34, 474–476. [PubMed]

85. Wang, L.; Zhou, G.; Liu, H.; Niu, X.; Han, J.; Zheng, L.; Fan, Y. Nano-hydroxyapatite particles induce apoptosis on MC3T3-E1 cells and tissue cells in SD rats. *Nanoscale 2012*, 4, 2894–2899. [CrossRef] [PubMed]

86. Wang, L.; Zhou, G.; Fan, Y. Effects of nano-hydroxyapatite/chitosan (N-HA/CS) on MC3T3-E1 cell and metabolic organ in SD rats. *Chin. J. Biomed. Eng.* 2013, 32, 595–600.

87. Albrecht, C.; Scherbart, A.M.; van Berlo, D.; Braunbarth, C.M.; Schins, R.P.; Scheel, J. Evaluation of cytotoxic effects and oxidative stress with hydroxyapatite dispersions of different physicochemical properties in rat NR8383 cells and primary macrophages. *Toxicol. In Vitro 2009*, 23, 520–530. [CrossRef] [PubMed]

88. Scheel, J.; Weimans, S.; Thiemann, A.; Heisler, E.; Hermann, M. Exposure of the murine RAW 264.7 macrophage cell line to hydroxyapatite dispersions of various composition and morphology: Assessment of cytotoxicity, activation and stress response. *Toxicol. In Vitro 2009*, 23, 531–538. [CrossRef] [PubMed]

89. He, X.; Young, S.H.; Schwegler-Berry, D.; Chisholm, W.P.; Fernback, J.E.; Ma, Q. Multivalved carbon nanotubes induce a fibrogenic response by stimulating reactive oxygen species production, activating NF-κB signaling, and promoting fibroblast-to-myofibroblast transformation. *Chem. Res. Toxicol. 2011*, 24, 2237–2248. [CrossRef] [PubMed]

90. Lin, W.; Xu, Y.; Huang, C.-C.; Ma, Y.; Shannon, K.B.; Chen, D.-R.; Huang, Y.-W. Toxicity of nano- and micro-sized ZnO particles in human lung epithelial cells. *J. Nanopart. Res. 2009*, 11, 25–39. [CrossRef]

91. Tay, C.Y.; Fang, W.; Setyawati, M.I.; Chia, S.L.; Tan, K.S.; Hong, C.H.L.; Leong, D.T. Nano-hydroxyapatite and nano-titanium dioxide exhibit different subcellular distribution and apoptotic profile in human oral epithelium. *ACS Appl. Mater. Interfaces 2014*, 6, 6248–6256. [CrossRef] [PubMed]

92. Tee, J.K.; Ong, C.N.; Bay, B.H.; Ho, H.K.; Leong, D.T. Oxidative stress by inorganic nanoparticles. *WIREs Nanomed. Nanobiotechnol. 2015*, 99, A179. [CrossRef] [PubMed]

93. Nel, A.; Xia, T.; Madler, L.; Li, N. Toxic potential of materials at the nanolevel. *Science 2006*, 311, 622–627. [CrossRef] [PubMed]

94. Kinov, P.; Leithner, A.; Radl, R.; Bodo, K.; Khoschhorou, G.-A.; Schauenstein, K.; Windhager, R. Role of free radicals in aseptic loosening of hip arthroplasty. *J. Orthop. Res. 2006*, 24, 55–62. [CrossRef] [PubMed]

95. Freeman, T.A.; Parvizi, J.; della Valle, C.J.; Steinbeck, M.J. Reactive oxygen and nitrogen species induce protein and DNA modifications driving arthrofibrosis following total knee arthroplasty. *Fibrogenes. Tissue Repair 2009*, 2, 5. [CrossRef] [PubMed]

96. Ding, T.; Xue, Y.; Lu, H.; Huang, Z.; Sun, J. Effect of particle size of hydroxyapatite nanoparticles on its biocompatibility. *IEEE Trans. Nanobiosci. 2012*, 11, 336–340. [CrossRef] [PubMed]

97. Steinbeck, M.J.; Jablonskowski, L.J.; Parvizi, J.; Freeman, T.A. The role of oxidative stress in aseptic loosening of total hip arthroplasties. *J. Arthroplast. 2014*, 29, 843–849. [CrossRef] [PubMed]

98. Jablonskowski, L.J. The Role of Inflammation and Oxidative Stress in Total Hip Replacement Revisions: Development of a Diagnostic Panel for Osteolysis; Drexel University: Philadelphia, PA, USA, 2011.

99. Wang, J.; Fan, Y. Lung injury induced by TiO$_2$ nanoparticles depends on their structural features: Size, shape, crystal phases, and surface coating. *Int. J. Mol. Sci. 2014*, 15, 22258–22278. [CrossRef] [PubMed]

100. Shi, Z.; Huang, X.; Cai, Y.; Tang, R.; Yang, D. Size effect of hydroxyapatite nanoparticles on proliferation and apoptosis of osteoblast-like cells. *Acta Biomater. 2009*, 5, 338–345. [CrossRef] [PubMed]

101. Xu, Z.; Liu, C.; Wei, J.; Sun, J. Effects of four types of hydroxyapatite nanoparticles with different nanocrystal morphologies and sizes on apoptosis in rat osteoblasts. *J. Appl. Toxicol. 2012*, 32, 429–435. [CrossRef] [PubMed]

102. Zhao, X.; Heng, B.C.; Xiong, S.; Guo, J.; Tan, T.T.; Boey, F.Y.; Ng, K.W.; Loo, J.S. In vitro assessment of cellular responses to rod-shaped hydroxyapatite nanoparticles of varying lengths and surface areas. *Nanotoxicology 2011*, 5, 182–194. [CrossRef] [PubMed]

103. Manke, A.; Wang, L.; Rojanasakul, Y. Mechanisms of nanoparticle-induced oxidative stress and toxicity. *BioMed Res. Int. 2013*, 2013, 942916. [CrossRef] [PubMed]

104. Liu, G.; Wang, R.; Dong, L.; Zhang, J.; Zhao, J. Endoplasmic reticulum stress-induced apoptosis of osteoblasts within the osteolytic craniums. *Zhongguo Zuzhi Gongcheng Yanjiu 2014*, 18, 5257–5265.

105. Wang, Z.; Huang, Z.; Gan, J.; Liu, N.; Zhou, G.; Shi, T.; Wang, Z.; Wang, R.; Bao, N.; Guo, T.; et al. The fibroblast expression of RANKL in CoCrMo-particle-induced osteolysis is mediated by ER stress and XBP1s. *Acta Biomater. 2015*, 24, 352–360. [CrossRef] [PubMed]
106. Yang, F.; Wu, W.; Cao, L.; Huang, Y.; Zhu, Z.; Tang, T.; Dai, K. Pathways of macrophage apoptosis within the interface membrane in aseptic loosening of prostheses. *Biomaterials* **2011**, *32*, 9159–9167. [CrossRef] [PubMed]

107. Xie, H.; Mason, M.M.; Wise, J.P., Sr. Genotoxicity of metal nanoparticles. *Rev. Environ. Health* **2011**, *26*, 251–268. [CrossRef] [PubMed]

108. Song, M.F.; Li, Y.S.; Kasai, H.; Kawai, K. Metal nanoparticle-induced micronuclei and oxidative DNA damage in mice. *J. Clin. Biochem. Nutr.* **2012**, *50*, 211–216. [CrossRef] [PubMed]

109. Uboldi, C.; Urban, P.; Gilliland, D.; Bajak, E.; Valsami-Jones, E.; Ponti, J.; Rossi, F. Role of the crystalline form of titanium dioxide nanoparticles: Rutile, and not anatase, induces toxic effects in Balb/3T3 mouse fibroblasts. *Toxicol. In Vitro* **2016**, *31*, 137–145. [CrossRef] [PubMed]

110. Franchi, L.P.; Manshian, B.B.; de Souza, T.A.J.; Soenen, S.J.; Matsubara, E.Y.; Rosolen, J.M.; Takahashi, C.S. Cyto- and genotoxic effects of metallic nanoparticles in untransformed human fibroblast. *Toxicol. In Vitro* **2015**, *29*, 1319–1331. [CrossRef] [PubMed]

© 2016 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC-BY) license (http://creativecommons.org/licenses/by/4.0/).