Structural basis for the function and regulation of the receptor protein tyrosine phosphatase CD45

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CD45 is the prototypic member of transmembrane receptor–like protein tyrosine phosphatases (RPTPs) and has essential roles in immune functions. The cytoplasmic region of CD45, like many other RPTPs, contains two homologous protein tyrosine phosphatase domains, active domain 1 (D1) and catalytically impaired domain 2 (D2). Here, we report crystal structure of the cytoplasmic D1D2 segment of human CD45 in native and phosphotyrosyl peptide–bound forms. The tertiary structures of D1 and D2 are very similar, but doubly phosphorylated CD3/H9256 immunoreceptor tyrosine–based activation motif peptide binds only the D1 active site. The D2 "active site" deviates from the other active sites significantly to the extent that excludes any possibility of catalytic activity. The relative orientation of D1 and D2 is very similar to that observed in leukocyte common antigen–related protein with both active sites in an open conformation and is restrained through an extensive network of hydrophobic interactions, hydrogen bonds, and salt bridges. This crystal structure is incompatible with the wedge model previously suggested for CD45 regulation.

Like many other RPTPs, CD45 consists of an extracellular receptor–like region, a short transmembrane segment, and a cytoplasmic region comprising tandem PTP domains (see Fig. 1 A). The length of the extracellular segment varies among the CD45 isoforms generated by alternative splicing (15). Mutational analyses have shown that the membrane proximal PTP domain 1 (D1), but not the membrane distal domain 2 (D2), is catalytically active (16). The entire cytoplasmic region of CD45 is conserved among vertebrates, from shark to mammalian (17). The strong conservation of the CD45 D2 primary structure implies a functional role, but this role is not yet clearly defined. Nonetheless, there are observations that suggest a regulatory role of the catalytically inert D2 domain (18). D2 perhaps influences the activity of D1 by direct intermolecular and intramolecular interaction, as shown by in vitro binding studies (19, 20). Phosphorylation of D2 by casein kinase 2 enhances the phosphatase activity of D1, consistent with a regulatory role of the D2 domain (21). It was also reported that the CD45 D2 domain could, by itself, bind one of the important substrates, Lck, facilitating its dephosphorylation by D1.
domain (22). In the case of another RPTP, leukocyte common antigen–related (LAR) protein, we observed a high degree of similarity between the crystal structures of the D1 and D2 domains (23). Perhaps consistent with the structural similarity of D2 and D1, only two amino acid substitutions were required to convert the otherwise inert D2 domains of LAR and PTPα into active enzymes (23–25). These findings led us to propose that D2 might function as an auxiliary catalytic site that would be activated under specific cellular conditions or in the presence of a proper substrate (23).

A generalized model, known as the dimeric inhibition model or the wedge hypothesis, for the regulation of RPTPα activity has been proposed based on the crystal structure of the RPTPα D1 domain (26). The RPTPα D1 domain formed a homodimeric structure in the crystal, in which the active site of one monomer was blocked by an NH2-terminal helix-turn-helix wedge motif of the other monomer, suggesting that dimer formation on the membrane negatively regulates the PTP activity. Experimental data that seem to corroborate this model have been reported. For example, ligand-induced dimerization of the epidermal growth factor receptor–CD45 chimeric molecule and enforced covalent dimerization of full-length PTPα resulted in down-regulation of their PTP activities (27, 28). Impairment of dimer formation, by introducing mutations into the wedge region, led to apparent PTP activation both in vivo and in vitro (28–30). Transiently expressed PTPα was shown to exist predominantly as homodimers (31). In the same study, however, mutations in the wedge region reduced but did not completely abolish PTPα dimerization, suggesting that other factors are also contributing to the dimer formation.

Here, we report crystal structures of the CD45 cytoplasmic region containing both the D1 and D2 domains. We also determined crystal structures of the CD45 cytoplasmic region bound to phosphopeptide substrates; one bound to a nonspecific short phosphopeptide, and the other bound to the membrane-proximal immunoreceptor tyrosine-based activation motifs (ITAMs) of the CD3ξ chain. We will describe the key features of CD45 and its substrate interactions and discuss the implications of these structures as they impact on our understanding of the mechanism of D1 and the potential function of D2. In these structures, CD45 exists as a monomer with the D1 active site cleft, as well as the D2 site, clearly unobstructed by the rest of the protein chain. The observed intramolecular orientation of the D1 and D2 domains precludes the formation of a dimer of the type predicted by the dimeric inhibition model. Such a D1 dimer interaction is impossible because of significant steric hindrance generated from the overlap of the attached D2 domains.

Results

Overview of the CD45 D1D2 structure determination

In human leukocytes, various CD45 isoforms of different lengths are expressed by combinatorial alternative splicing (15). All isoforms, however, share the exactly identical cytoplasmic region of 707 amino acids, which is composed of a...
juxtamembrane spacer segment, the PTP D1 domain, a short interdomain linker, the PTP D2 domain, and a COOH-terminal tail (Fig. 1 A). In this report, we will refer to the amino acid positions as they appear in the mature protein of the longest isoform (15). According to this convention, the cytoplasmic domain is from 575 to 1281, and the catalytic site cysteine residues are at 828 (Cys828) in D1 and at 1144 (Cys1144) in D2, respectively. An alignment of the D1 and D2 sequences, as well as several key features in the PTP domain, are shown in Fig. 1 B. Previously, we and others have shown that the CD45 D2 PTP domain has no catalytic activity (16, 32), whereas others reported different findings (33). Later in this report, we will present the structural basis of why the CD45 D2 domain cannot have a catalytic activity. For convenience sake, however, we will use terms such as “D2 active site,” “D2 catalytic site,” etc., to refer to the region in D2 that corresponds in PTP domain structure to the D1 active site, without implying an actual catalytic activity.

To obtain the materials for crystallographic studies, the CD45 cytoplasmic region containing both the D1 and D2 domain was expressed in Escherichia coli as six-His–tagged proteins and purified to apparent homogeneity. In our initial attempts where we used the entire cytoplasmic region of CD45, we could not obtain sufficiently good crystals. This was likely due to the presence of an extended COOH-terminal segment that seems flexible. To circumvent this problem, we generated a series of CD45 cytoplasmic fragments, differing both at the NH2- and COOH-terminal positions, to obtain the optimal domain boundaries for crystallization. Thus, we obtained two different crystals, CD45 D1D2 (597–1213) with Cys828 to Ser (C828S) mutation, and wild-type CD45 D1D2 (603–1203), which gave optimal diffraction data. In addition, a number of datasets were analyzed from crystals grown in the presence of different ions (sulfate, phosphate, and tungstate ions), monomeric phosphotyrosine, and two different phosphotyrosyl-peptides.

The best quality data have been collected from monoclinic crystals of selenomethionine (Sel-Met)–substituted CD45-D1D2 (597–1213) (C828S) protein with phosphotyrosine-containing peptides (derived from polyoma middle T antigen and CD3ζ ITAM motif; see below for more details). The crystal structures for both complexes were solved to 2.9 Å resolution by the molecular replacement method. Data collection and the final refinement statistics for these complexes are summarized in Table I. Both datasets are of similar quality with respect to resolution and agreement of R factor for symmetry-related reflections (Rsym). The corresponding protein structures demonstrate correct geometric parameters as well as the desired lack of model bias. The current CD45 model includes 589 residues (601–1205), excluding the disordered acidic loop segment of D2.

In Fig. 2, the structure of the CD45 cytoplasmic region is viewed from three directions. For comparison, the structure of LAR D1D2, the only other RPTP for which the tandem PTP domain structure has been solved (23), is reproduced in the inset as viewed from the same direction as the CD45 structure shown at the top left. It is clear that the observed orientation of the D1 and D2 domains in CD45 is very similar to that adopted by LAR. It also can be seen that the overall structures of the D1 and D2 domains are essentially identical to each other. Both D1 and D2 domains consist of a highly twisted nine-stranded mixed β sheet flanked by sheets flanked by α helices that are tilted to the β-sheets and are in registry, thus forming a sandwich-like structure. The α helices are perpendicular to the β-sheets, and each β-sheet is composed of at least two parallel strands. The PTP domain of CD45 is flanked by a short linker that forms a helical turn, which is connected to the D2 domain by a short flexible segment. The D1 domain is connected to the D2 domain by a long link that is flanked by alternating β-strands and α-helices.

Table I. Crystallographic data and refinement statistics

| Crystallographic data | Polyoma middle T antigen | CD3ζ ITAM-1 |
|-----------------------|--------------------------|-------------|
| Space group           | P2(1)                    | P2(1)       |
| Cell parameters       | a = 86.9 Å, b = 57.9 Å,  | a = 86.0 Å, b = 59.7 Å, |
|                       | c = 159.3 Å, β = 98.78   | c = 160.5 Å, β = 99.58 |
| Z                     | 4                        | 4           |
| X-ray source          | X12, BNL                 | X12, BNL    |
| Resolution            | 30–2.9 Å                 | 30–2.9 Å    |
| Observations (n)      | 99,669                   | 99,514      |
| Unique reflections (complete) | 32,025 (93.0%) | 29,466 (81.5%) |
| Rsym                  | 0.079                    | 0.071       |
| Refinement statistics of current model |                       |             |
| Resolution            | 30–2.9 Å                 | 30–2.9 Å    |
| Protein atoms (n)     | 9,457                    | 9,416       |
| Substrate atoms (n)   | 74                       | 122         |
| Rwork(work/free)      | 0.263/0.306              | 0.258/0.315 |
| Rmsd bond lengths     | 0.010 Å                  | 0.010 Å     |
| Rmsd bond angles      | 1.59°                    | 1.57°       |

*Rsym = Σh|Fobs| - |Fcalc||Σh|Fobs|, where |Fobs| is the mean of the observations l_h of reflection h.

*Rwork = Σ|Fobs| - |Fcalc|/Σ|Fcalc|.

*Rfree = R factor for a selected subset (5%) of the reflections that was not included in prior refinement calculations.
by four α helices on one side and two α helices on the other. In general, this domain structure is similar to those of previously reported PTPs (26, 34, 35). Secondary structure elements in D1 and D2, as determined in our structure, are shown in Fig. 1 B.

Interaction of phosphopeptides at the D1 active site
To understand the mechanisms of substrate recognition and dephosphorylation by CD45, purified CD45 D1D2 (597–1213) C828S was crystallized in the presence of either phosphotyrosine (i.e., monomeric amino acid) or one of two different phosphopeptides. C828S mutant was used to prevent dephosphorylation of the substrates.

The first phosphopeptide (QQQNQLpYNELNLGR-REEpYDVLDKRRG, where pY is phosphotyrosine) is the membrane proximal ITAM of the CD3ε chain (ζITAM-1) with both the tyrosine residues phosphorylated. CD3ε is an important CD45 substrate and contains three ITAMs in its cytoplasmic domain. An ITAM is defined as a 20–25–amino acid sequence containing two YxxL/I segments separated by six to eight amino acid residues (36). Each of the three ITAMs in CD3ε is tyrosine phosphorylated when TCR is activated. The doubly phosphorylated ITAMs serve as docking sites for the tandem SH2-containing ZAP-70, which initiates a cascade of signal transduction events (37, 38). It has been demonstrated that CD3ε and CD45 physically associate in vivo and that CD45 efficiently dephosphorylates CD3ε in vitro (39, 40). Mutational analyses of the tyrosine residues in the membrane-proximal ITAM of CD3ε have shown that the two YxxL segments in the ITAM are functionally distinct (41), indicating the critical importance of regulating ITAM phosphorylation states.

The second phosphopeptide (NPTpYS) is derived from the polyoma middle T antigen. Although this peptide is probably not a physiologically meaningful substrate of CD45, it has the advantage of being a well-known short phospho-substrate.

When CD45 was crystallized in the presence of monomeric phosphotyrosine, there was a well-defined density corresponding to this moiety in the D1 active site pocket, but no comparable density in the D2 domain that would suggest the presence of phosphotyrosine (not depicted). In the crystal structure of the CD45 D1D2 complexed with ζITAM-1, although there are two phosphotyrosyl residues in ζITAM-1, assignable extra densities were visible only at the D1 active site. No extra densities are discernible at the D2 active site, or in fact anywhere else associated with the protein. Electron densities for six amino acids surrounding the second phosphotyrosyl residue (REEpYDV) are visible at the D1 active site (Fig. 3, A and B). This result appears consistent with the
previous in vitro analyses showing that the CD45 phosphatase prefers the second phosphotyrosine in ζITAM-1 over the first phosphotyrosine (42). Similarly, in the structure of CD45 bound with the polyoma phosphopeptide (NPT-pYS), electron density corresponding to four residues (PT-pYS) was visible only at the D1 active site (Fig. 4, A and B).

For both the ζITAM-1 peptide and the polyoma peptide, the general features of the phosphotyrosine binding to the CD45 D1 active site are similar. The active site pocket of CD45 D1 is composed of a base formed by the residues of the PTP signature motif, surrounded by three additional segments: the KNRY motif, the WPD motif, and the Q-rich motif (Fig. 1 B). Extensive hydrogen bonding occurs between the phosphotyrosyl residue and the active site pocket. Within the PTP signature motif, the active site Cys828 (Ser in our structure) is positioned for nucleophilic reaction with the substrate phosphorus. Hydrogen bonds are observed between the phosphophoryl moiety and the main chain amide groups of the PTP signature motif, as well as the basic side chain of Arg834. The tyrosine residue of the phosphopeptide is stabilized by aromatic ring stacking with Tyr658 in the KNRY motif. The WPD motif, which contains catalytically important Asp796, adopts an open conformation when crystallized with sulfate ions. This loop, however, shows a closed conformation in all of our substrate-bound structures. The imidazole ring of His797 makes hydrophobic contact with the phenyl ring of the substrate phosphotyrosyl residue as well as a hydrogen bond with the backbone carbonyl group of the phosphotyrosyl residue. The acidic side chain of Asp660 in the KNRY loop is making a hydrogen bond with the amide group of the pY residue. This positioning of the two residues of opposite charges (His797 and Asp660) seems to determine the backbone configuration of the substrate phosphotyrosyl residue and residues COOH-terminal to it. The conserved Gln872 and Gln876 residues in the Q-rich motif are positioned surrounding the active site to generate the required charge distribution and pocket shape that accommodate a phosphate leaving group during catalysis. Overall, most interactions between the substrate phosphotyrosyl residue and the D1 active site are similar to those seen in the substrate-bound complex of PTP1B and Yersinia PTP (43, 44), with the exception of the one between the

Figure 3. Structure of the CD45 D1D2 bound to the CD3 ζITAM-1 phosphopeptide. (A) A stereo view of the D1 active site with the CD3 ζITAM-1 phosphopeptide. Relevant hydrogen bonds between the side chains of the CD45 protein and the ζITAM-1 peptide are shown by green lines. (B) The phosphopeptide model is shown with a 2|Fobs|−|Fcalc| composite omit map at the 0.9 σ contour level. (C) Molecular surface representation of the D1 active site bound to the CD3 ζITAM-1 peptide. The surfaces are colored according to the electrostatic potential: blue for positive and red for negative. The peptide model is shown using a ball and stick representation.
carbonyl of the phosphotyrosyl residue and the basic side chain of His797. The residue corresponding to His797 is a phenylalanine in PTP1B. In the CD45 structure, His797 may add extra interactions and strengthen the binding of the substrate phosphotyrosyl residue.

Fig. 3A shows detailed interactions between six amino acids (REEpYDV) of the ITAM-1 substrate and the CD45 protein. On the NH₂-terminal side of the phosphotyrosyl residue, the Glu (pY-1) side chain is pointing in the direction opposite that of the phosphotyrosine. The Glu (pY-1) side chain is exposed to solvent without any interaction with the protein; thus, the presence of a hydrophilic residue at this position might be favored for binding. The carboxyl group of the Glu (pY-2) makes a hydrogen bond contact with the guanidium group of Arg734. The surface charge distribution of the CD45 protein at this region shows an extended positively charged surface contributed by residues Arg657, Arg704, Arg734, and Lys736 (Fig. 3, A and C). The negatively charged Glu (pY-2) fits nicely into this positively charged pocket and seems to be important for binding to the CD45 PTP. The positive charges of the protein at this region may repel the basic side chain of Arg (pY-3), thus allowing no direct interactions with the protein.

On the COOH-terminal side of ITAM-1, the Asp (pY+1) side chain points toward the CD45 protein and makes a hydrogen bond with the side chain of Gln872. The main chain backbone of the Val (pY+2) runs almost perpendicular to the rest of the peptides. The overall extent of these contacts between the ITAM-1 substrate and the D1 domain is consistent with the previous results from PTP1B–substrate complexes (43), indicating that residues between pY-3 and pY+2 of the substrate determine phosphatase specificities.

In the structure of CD45 bound with the polyoma phosphopeptide (NPTpYS), electron densities corresponding to four residues (PpYS) are visible at the D1 active site (Fig. 4). In addition to the interactions between the phosphotyrosine moiety and the protein, the NH₂-terminal peptide residues make contact with the KNRY loop. Thr (pY-1) is in van der Waals contact with the side chain of Asp660, and the ring of Pro (pY-2) is making a van der Waals contact with the Tyr658 ring. The terminal oxygen of Ser (pY+1) is hydrogen bonded with the side chain of residue Gln872 in the Q-rich motif.

Based on these structures, we propose that the optimal interaction of CD45 active site and phosphotyrosyl substrate occurs when a hydrophilic residue occupies the pY-1 position and negatively charged amino acids are located around the pY-2 position. The Lck and Fyn protein tyrosine kinases, two of the best known CD45 substrates, fit into these criteria. These protein kinases have two tyrosine phosphorylation sites: one in their activation loops (DENEpYTAR in Lck and EDNEpYTAR in Fyn) and another near their COOH termini (TEGQpYQPQ in Lck and TEPQpYQPG in Fyn). All of these peptides contain hydrophilic residues at the pY-1 position, as in the ITAM-1 sequence. In the activation loop peptides, the acidic amino acid at pY-3 (and perhaps at pY-4) is likely to interact with the positively charged pocket of CD45. In the COOH-terminal peptides, we can more confidently speculate that the negatively charged Glu (pY-3) interacts with the positively charged pocket of CD45, because the residues at the Y-2 position, namely Gly and Pro, are optimal at generating conformations to accommodate such interactions. The NH₂-terminal phosphorylation site of ITAM-1 (NQLpYNE) lacks these criteria. The presence of the hydrophobic Leu residue at the pY-1 position, and lack of any negatively charged amino acids at the pY-2 and pY-3 positions, explain the reduced affinity of the NH₂-terminal phosphorylation site for the CD45 PTP.

The CD45 D2 domain is a totally inactive phosphatase

The strong structural conservation of the CD45 D2 domain implies that it serves a cellular function, but it is still of ma-
Our CD45 structure suggests that the function of the CD45 D2 domain is very different from its LAR counterpart. In the LAR D1D2 structure we determined previously, the inactive D2 domain maintains an active site topology similar to that of D1. With only two substitutions of the key residues in the surrounding loops, LAR D2 could be converted to a very active enzyme (23). In contrast, the PTP signature motif in CD45 D2 deviates from the consensus sequence to the extent that it can no longer accommodate a phosphoryl group (VHCSAGVGRGTG in D1 vs. IHCRDGSSQQTG in D2). As a result, the CD45 D2 domain has a significantly reduced number of available ligands for oxygen binding, a significantly altered shape of the active site pocket, as well as a reduced accessibility of the Cys nucleophile. Fig. 5, A and B, show the details of the D1 and D2 active sites, respectively. The structure of the loops surrounding the D2 active site is also quite different from that of the D1 active site. This can be easily visualized by the surface representation of the active sites with their respective charge distributions (Fig. 5, C and D). The WPD loop in D2 shows a partially open conformation, and the invariant Asp, which acts as general acid/base in active PTPs, is substituted with Val1102 in D2. This substitution will certainly impair catalytic activity. The KNRY loop also shows a quite different conformation in D2. The conserved Tyr involved in the stabilizing $\pi$-$\pi$ interactions with the phenyl ring of the ligand phosphoryl residue is replaced by Asn949 in D2, further contributing to the reduced affinity for phosphoryl substrates. In conclusion, the CD45 D2 structure does not have even the remotest possibility of retaining a catalytic activity. The KNRY loop also shows a quite different conformation in D2. The conserved Tyr involved in the stabilizing $\pi$-$\pi$ interactions with the phenyl ring of the ligand phosphoryl residue is replaced by Asn949 in D2, further contributing to the reduced affinity for phosphoryl substrates. In conclusion, the CD45 D2 structure does not have even the remotest possibility of retaining a catalytic activity.
lytic activity. This represents a clear mechanistic distinction between CD45 and other RPTPs, even as compared with its close relative, LAR.

**Possible role of the CD45 D2 domain deduced from its structure**

Inspection of the D2 structure, as detailed in this section, suggests that it has a role in substrate recruitment. The CD45 D2 domain has two unique features that are not found in any other PTPs, including LAR. Those are the two extra loops, referred to, respectively, as the acidic and basic loops, whose positions are shown in Fig. 2 by color. The 20-amino acid acidic loop (SKESEHDSDESSDDSSEE [969–988]), located between the β1 and β2 strands, is mostly disordered and flexible. Although not all of the residues in the acidic loops could be seen in the crystal structure, the location and approximate model can be visualized based on the rest of the structure and the electron density observed in this area. The 11-amino acid basic loop (KNSSEGNKHHK [1126–1136]) is located between the α3 helix and the β12 strand. The entire basic loop could be refined in one of the two molecules in the P2(1) space group, but the temperature factor for these residues is quite high (average main chain B factor is ~20 Å² higher than the rest of the molecule), reflecting the flexible nature of the basic loop.

It has been reported that deletion of the acidic loop, or substitution of four Ser residues to Ala in the acidic loop, diminished CD3-mediated activation of T lymphocytes (45). Furthermore, phosphorylation of Ser residues in the acidic loop by casein kinase 2 is important for CD45 functions (21). Our structure lends a mechanistic explanation to such observations. Both the acidic and basic loops in D2 are located on the same side of the surface of the molecule with respect to the D1 active site. Because both loops have significant charge biases and are located very close to the active site in D1, they could easily serve as initial substrate recruiting sites. When phosphorylated, these loops may undergo a conformational change or induce a change in the catalytic site to facilitate docking of incoming substrates.

**Extensive interaction between the D1 and D2 domains**

In this work, we obtained crystals of CD45-D1D2, without peptide, in two different space groups: P2(1) and P6(3)22. Although the crystallographic contacts between the neighboring molecules are totally different in the two crystal forms, the intramolecular spatial relationship between the D1 and D2 domains is essentially identical in both crystal forms. Several cocrystals of CD45-D1D2 with phosphopeptides also had very similar D1–D2 domain orientation. If the atoms of the D1 domains from two different crystal forms, P2(1) and P6(3)22, are superimposed, the atoms in the D2 domains differ by an average (root-mean-square) distance of 3.2 Å between the equivalent Cα atoms. They differ by 2.9 Å between the two noncrystallographically related molecules in the P2(1) space group. The average distances are even smaller between the residues located at the interface of the D1 and D2 domains (~1.5 Å). In all cases, the D1 and D2 domains are connected by a four-residue linker, GETE (891–894), and packed tightly against each other. This linking segment is conserved between CD45 and LAR (GETE and GHTE, respectively) and is bound to both D1 and D2 domains in a similar fashion in the two different proteins. The interdomain orientation between D1 and D2 is further stabilized by an extensive network of interactions, consisting of hydrogen bonds, van der Waals interactions, and salt bridges (Fig. 6). Hydrophobic interactions were observed at two regions of the interdomain surface. Phe890 at the end of D1 α6 is making van der Waals contact with the Phe1173 in D2 α5, and Pro765 (in the loop between B9 and B10 of D1) is in contact with Tyr1001 and Trp1002 (in the loop be-
between β2 and β3 of D2). Hydrophobic interactions are further stabilized by salt bridges, observed between the basic side chains (His804, Lys808, Arg811, and Arg812) in D1 α3 and acidic side chains (Glu1167 and Asp1171) in the D2 domain. Among these residues, Arg811 and Arg812 are highly conserved among most RPTP D1 domains, and Glu1167 and Asp1171 are conserved in most RPTP D2 domains. Overall, there are extensively complementary surfaces between the D1 and D2 domains, including the residues of the α3 and α6 helices of D1 and the α4 and α5 helices of D2.

These results suggest that the intramolecular D1–D2 orientation reported here is a likely state of the CD45 molecule in the cell. Furthermore, this relative orientation of the D1 and D2 domains of CD45 is very similar to that of LAR, which we reported previously (23), suggesting that this is typical of most, if not all, two-domain RPTPs. The energy required to separate D1 from D2 is ~16 kcal/mol, which is probably sufficient to keep the two domains bound together even if they are not tethered by the linker. This conclusion is consistent with the earlier observations, based on fluorescence resonance energy transfer and coprecipitation analyses, which showed strong intramolecular and intramolecular interactions between D1 and D2 domains of various RPTP proteins (46, 47). Nonetheless, it should be noted that the extracellular domain and transmembrane segment, which are not included in our structure, may have a significant influence on the conformation of D1–D2 orientation in cells.

Because of these extensive D1–D2 interactions, even subtle structural changes in D2 might affect the D1 conformation. Indeed, a mutation in D2 is known (Q1180G in mouse CD45, which is equivalent to Q1192G in our human structure) that nearly completely abolishes the D1 catalytic activity (48). Because Gln1192 is not making any direct contact with D1 residues, its effect must be propagated indirectly through the D1–D2 interface. Exactly how this is effected will require the structural solution of the CD45 Q1192G mutant protein. Thus, stabilization of the D1 domain seems an important role of D2. We can hypothesize that the primordial two-domain RPTPs had two enzymatically active PTP domains, and over the history of RPTP evolution, D1 and D2 domains established extensive association to the extent that one requires another for maintenance of its proper structure. Thus, although the CD45 D2 domain has lost its catalytic activity, it may still be needed to support the D1 structure and activity.

**DISCUSSION**

An outstanding question concerning RPTPs is how extracellular stimuli, such as ligand binding, may affect the catalytic activity of the cytoplasmic PTP domains. A major model concerning this question was proposed based on the crystal structure of the RPTPα D1 domain that crystallized as a dimer (26). In the RPTPα dimer structure, the active site in each monomer was blocked by the NH2-terminal α1-turn-α2 segment (called “wedge”) of an opposing monomer, thus preventing substrate access to the active site.

It was further suggested that ligand-induced dimerization activates the RPTP phosphatase activity. The wedge hypothesis is aesthetically attractive because, if proven, it would depict RPTPs as a perfect mirror image of receptor tyrosine kinases, where ligand-induced dimerization typically activates the kinases. This hypothesis was apparently supported by in vivo and in vitro analyses of RPTPα and epidermal growth factor receptor–CD45 chimeric molecules (28–30).

Nonetheless, our present CD45 structure shows that the cytoplasmic region of CD45 does not dimerize even when highly concentrated, and the dimeric interaction as observed for RPTPα D1 would be impossible given the D1–D2 intramolecular domain orientation. The CD45 D1–D2 interaction seems too tight to allow the deformation needed for D1–D1 interaction, given the extensive interface and the essentially identical D1–D2 orientation observed among different CD45 crystal forms and LAR. Furthermore, the CD45 wedge region (610–634; refer to Fig. 2) is not involved in any intermolecular interaction. We previously reached a similar conclusion regarding lack of dimerization of the D1D2 domains based on the LAR structure (23). At that time, we could not know for sure whether what we observed for LAR represented a general phenomenon or merely an isolated case of that particular RPTP. However, now that we have solved the CD45 structure itself, we can convincingly conclude that the steric hindrance from the D2 domain prevents D1–D1 dimer formation in CD45 as proposed previously (26).

Our data, therefore, indicate that the regulation of CD45 cannot be simply explained by the dimeric inhibition model mediated by the NH2-terminal wedge. Nonetheless, the wedge region (helix-turn-helix motif) is conserved, suggesting that it has a regulatory function (29, 30). If so, however, it is likely that the wedge region influences the CD45 function by a mechanism other than the dimer formation. Indeed, it was recently shown that CD45 could dimerize without the cytoplasmic domain (49). Furthermore, a CD45 wedge mutation that had significant effect on the CD45 function did not affect the dimerization. Obviously, more studies, including the structures of additional RPTPs as well as the extracellular portion of CD45, will be needed to further such understanding.

**MATERIALS AND METHODS**

**Nomenclature.** An individual amino acid is indicated by the standard three-letter or one-letter code, followed by its amino acid position. Thus, both Cys828 and C828 represent the cysteine at position 828. Missense mutations are designated by the one-letter code. C828S, for example, indicates the mutation of Cys828 to serine. Peptide sequences are indicated by the one-letter amino acid code. pY is a phospho-tyrosine residue. pY-1 indicates the amino acid position immediately on the COOH-terminal side of pY, and pY+2 represents the second amino acid from pY on the COOH-terminal side.

**Preparation of recombinant proteins.** Proteins were expressed using the T7 expression system essentially as described previously (23, 50). The catalytic site mutant (C828S) of CD45-D1D2 (597–1213) was tagged at the NH2-terminus with cleavable six-His, whereas the wild-type CD45-D1D2...
lographic symmetry restraints were used throughout refinement. Composite (CNS) software packages, followed by manual rebuilding (53). Noncrystallographic factor refinement options of the crystallography & NMR system of refinement were performed using minimization and anisotropic temperature factor refinement revealed four unambiguous positions of phosphatase domains. Determination of the peptides would be expected in an asymmetric unit. For the structure determination, the CD45-D1D2 (597–1213) C828S model and Sel-Met data. The map showed strong corresponding densities at most of the Met positions, validating the model.

Crystallization. Two different crystal forms of CD45 D1D2 protein were obtained. Monoclinic crystals of CD45-D1D2 (597–1213) C828S were grown by vapor diffusion in hanging drops in 100 mM sodium acetate, pH 5.0, 10–13% (wt/vol) polyethylene glycol (PEG) 4000, 10 mM diithiothreitol (DTT), and 25% glycerol, with various additives such as ammonium sulfate, phosphotyrosine, or phosphopeptide. For cocrystallization experiments with phosphopeptides, either NTPtPrS derived from polymyx middle T antigen or QQQQNQpYPYENLNGREEpYDVLKRG, the membrane proximal ITAM of the CD3γ chain (ITAM-1), was added to the purified CD45-D1D2 (597–1213) C828S protein so that the final concentration of the peptides would be ~10 mM.

Hexagonal crystals of CD45-D1D2 (603–1203) with noncleavable COOH-terminal six-His tag were grown by vapor diffusion in hanging drops in 100 mM MOPS/NaOH, pH 6.5, 13–15% (wt/vol) PEG 8000, 0.5 M ammonium sulfate, 10 mM DTT, and 15% glycerol.

X-ray data collection. Crystals were flash-frozen with the crystallization solution. Diffraction data were collected at various synchrotron sources including the F1 station of the Cornell High Energy Synchrotron Source, the SBC-CAT of Advanced Photon Source in Argonne National Laboratory, and the X12 station of Brookhaven National Laboratory (BNL). Images were processed with DENOZO, and data were scaled and processed with SCALEPACK (51).

Structure determination. Initially, the structure of CD45-D1D2 (597–1213) C828S bound with sulfate ion was determined. In this crystal form, two molecules were expected in an asymmetric unit. The structure was determined by molecular replacement using an unaltered model of the D2 domain of RPTP LAR as a search model with the program AmoRe (52). An initial search revealed four unambiguous positions of phosphatase domains. After rigid body refinement, the identity of the PTP domains was determined based on the locations of the N and COOH termini, and amino acid sequence was changed to reflect the CD45 molecule. Several iterative cycles of refinement were performed using minimization and anisotropic temperature factor refinement options of the crystallography & NMR system (CNS) software packages, followed by manual rebuilding (53). Noncrystallographic symmetry restraints were used throughout refinement. Composite omit map in CNS was used for most of the refinement steps.

A refined model of the CD45-D1D2 (597–1213) C828S–sulfate complex was then used to determine other crystal structures. A Sel-Met dataset was collected with the intention of confirming the refined structure. Because of the anisomorphism, the Sel-Met data could not be used for phase determination. Instead, an Fobs–Fcalc map was generated using the CD45-D1D2 (597–1213) C828S model and Sel-Met data. The map showed strong (>3.5σ) corresponding densities at most of the Met positions, validating the structure. For structure determination of Sel-Met CD45-D1D2 (597–1213) C828S–phosphopeptide complexes, a refined model of Sel-Met CD45-D1D2 (597–1213) C828S was used to generate an Fobs–Fcalc map. For both phosphopeptides, ITAM-1 and polyoma middle T antigen, initial maps revealed clear peptide density at each D1 active site. Refinement procedures include minimization and individual temperature factor refinement using the CNS software package.

In the hexagonal crystal of CD45-D1D2 (603–1203), one molecule was expected in an asymmetric unit. For the structure determination, the refined model of the CD45-D1D2 (597–1213) C828S was used for molecular replacement. The CD45 D1 and D2 domains were used either together or separately for the search. In both cases, the highest solutions were the same, and the relative orientation of the D1 and D2 domains was basically identical as in monomeric crystals.

Calculation of the free energy required to separate the D1 and D2 domains. Environment-free energies of the CD45 D1D2 proteins have been calculated using the program ENVIRON (54). The atomic model of the protein was manually modified so that there were no interdomain interactions between the D1 and D2 domains, with the linker region still attached. The environment-free energies of the several modified models were calculated, and the energy required for separation of the D1 and D2 domains was calculated by subtracting the free energy of the native protein from that of the modified proteins.

Coordinates of the CD45/CD3 ITAM-1 and the CD45/polyoma middle T antigen complex have been deposited into the Protein Data Bank with the ID clones IYGR and IYGU, respectively.

Graphics. Molecular models were generated using the programs ALSCRIPT (55), MOLSERIP (56), RASTER3D (57), BOBSCRIPT (56), and GRASP (59).

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