Deficiency of the Stress Kinase p38α Results in Embryonic Lethality: Characterization of the Kinase Dependence of Stress Responses of Enzyme-deficient Embryonic Stem Cells

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Abstract

The mitogen-activated protein (MAP) kinase p38 is a key component of stress response pathways and the target of cytokine-suppressing antiinflammatory drugs (CSAIDs). A genetic approach was employed to inactivate the gene encoding one p38 isoform, p38α. Mice null for the p38α allele die during embryonic development. p38α−/− embryonic stem (ES) cells grown in the presence of high neomycin concentrations demonstrated conversion of the wild-type allele to a targeted allele. p38α−/− ES cells lacked p38α protein and failed to activate MAP kinase–activated protein (MAPKAP) kinase 2 in response to chemical stress inducers. In contrast, p38α+/− ES cells and primary embryonic fibroblasts responded to stress stimuli and phosphorylated p38α, and activated MAPKAP kinase 2. After in vitro differentiation, both wild-type and p38α−/− ES cells yielded cells that expressed the interleukin 1 receptor (IL-1R). p38α+/− but not p38α−/− IL-1R-positive cells responded to IL-1 activation to produce IL-6. Comparison of chemical-induced apoptosis processes revealed no significant difference between the p38α+/− and p38α−/− ES cells. Therefore, these studies demonstrate that p38α is a major upstream activator of MAPKAP kinase 2 and a key component of the IL-1 signaling pathway. However, p38α does not serve an indispensable role in apoptosis.

Key words: inflammation • cytokines • mitogen-activated protein kinase • signaling • cytokine-suppressing antiinflammatory drug

Introduction

After binding of the inflammatory cytokines IL-1 and TNF-α to their specific receptors on target cells, complex kinase-mediated signaling cascades are initiated that lead to specific changes in transcriptional and translational activities (1, 2). Many of these same signaling pathways can be engaged by chemical stress stimuli such as sodium arsenite or anisomycin (3, 4), by UV irradiation, (5), and by LPS (6). Several members of the family of mitogen-activated protein (MAP) kinases are components of the stress response, including p38 and c-Jun NH2-terminal protein kinases (7, 8). These MAP kinases regulate activity of key transcription factors including ATF2, Elk, CHOP, MEF2C, and CREB (7, 9–12); this regulation is achieved, in part, by control of downstream kinases such as MAP kinase–activating protein (MAPKAP) kinase 2 and 3 and p38-regulated/activated protein kinase (PRAK; 13–15). Stress-activated MAP kinases, therefore, serve as key coordinators and/ or regulators of a cell's stress responsiveness.

p38 MAP kinase was first identified as an LPS-inducible activity in murine peritoneal macrophages (6). Like other members of the MAP kinase superfamily, p38 phosphorylates its substrates on serine/threonine residues, and itself requires dual phosphorylation on both a threonine and tyrosine residue for activity (16). Several upstream MAP kinase kinases (MKKs) have been reported to phosphorylate p38 in vitro, including MKK3, MKK4, and MKK6 (17, 18). Four human p38 isoforms have been identified (α, β, γ, and δ), each containing the conserved dual phosphorylation sites.
posttranslational processing of IL-1 

ting (39) and in others inhibiting (40), and to impair 

ta studies have demonstrated that CSAIDs prevent IL-

cation that these agents are selective kinase inhibitors; struc-

demonstrated to be insensitive to CSAID inhibition (28), indi-

ding to inhibit this kinase (19). Both the \( \alpha \) and \( \beta \) forms of p38 are reported to be CSAID sensitive (19–

Mechanism of action studies have demonstrated that CSAIDs prevent IL-1

tional events (38). These agents do not inhibit NF-

tive nuclear factor (NF)-\( \kappa \)B-dependent transcriptional events (38). These agents do not inhibit NF-\( \kappa \)B activation per se, but may block activation of accessory factors necessary for NF-\( \kappa \)B-dependent transcription. Finally, CSAIDs are reported to alter apoptosis, in some cases promoting (39) and in others inhibiting (40), and to impair posttranslational processing of IL-1\( \beta \) (41). Whether all reported CSAID effects are attributable to p38 inhibition, and to what extent the various p38 isoforms participate in separate biological processes, are important issues that remain to be clarified. Recent studies demonstrating that CSAIDs can also block cyclooxygenase activity (42) and increase Raf-1 activity (43) highlight the potential for CSAID-mediated cellular effects occurring independently of p38.

To better understand the biological function of p38\( \alpha \), a genetic approach was used to inactivate the gene encoding this kinase by homologous recombination in embryonic stem (ES) cells. Inbred DBA/1 lacj mice that are p38\( \alpha \) null die during early embryonic development. However, growth of p38\( \alpha ^{-/-} \)-ES cells in the presence of increased concentrations of neomycin led to isolation of a p38\( \alpha ^{-/-} \) ES cell line. These enzyme-deficient cells were used to investigate the role of p38\( \alpha \) in stress activation pathways. Results derived from this analysis confirm that p38\( \alpha \) is a major upstream activator of MAPKAP kinase 2 and an important component of the IL-1 signaling pathway. On the other hand, the results provide no indication that p38\( \alpha \) is a necessary component of the chemical-induced apoptotic response mechanism of ES cells. Therefore, these studies help to clarify the role of p38\( \alpha \) in stress response pathways.

**Materials and Methods**

Construction of the p38\( \alpha \) targeting vector. A 759-bp murine p38\( \alpha \) partial genomic PCR fragment was used as a probe to identify genomic clones from a DBA/1 lacj genomic phage library (Stratagene). The p38\( \alpha \) targeting vector (Fig. 1) was constructed by cloning 5.5 kb of 5' homology and 3.5 kb of 3' homology into a pJNS2 (PGK-NEO/PGK-Tk) backbone vector. The targeting vector replaced \( \sim \)8–10 kb of genomic locus, containing the TGY dual phosphorylation site required for kinase activation, with PGK-NEO.

Generation of p38\( \alpha \)-deficient ES cells. DBA/252 ES cells (44, 45) were cultured on primary embryonic fibroblasts (PEFs) in DMEM containing high glucose, 15% heat-inactivated fetal bovine serum, 0.2 mM l-glutamine, 0.1 mM 2 ME, 0.1 mM MEM nonessential amino acids, penicillin-streptomycin, and 400 U/ml recombinant murine leukemia inhibitory factor (SCML). The p38\( \alpha \) targeting vector (25 \( \mu \)g) was electroporated at 1.0 \( \times \)10\( ^{6} \) ES cells/ml (0.4 ml total volume), and the electroporation was achieved with a BTX Elect roc cell M anipulator 600 (200 V, 50 \( \mu \)F, 360 \( \Omega \)). Cells were subsequently subjected to positive/negative selection with 200 \( \mu \)g/ml G418 (geneticin; Life Technologies) and 2 \( \mu \)M gancyclovir (Syntex Laboratories) as described (46). Gene targeting by homologous recombination was determined by

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**Figure 1.** Scheme employed to achieve homologous recombination of the murine p38\( \alpha \) locus. Approximately 8–10 kb of endogenous genomic sequence, including regions necessary for p38\( \alpha \) enzymatic activity, was replaced by the neomycin gene. This creates a deletion from phenylalanine 129 to aspartic acid 283 within the native protein. Hybridization of ES cell DNA with an external probe identified a predicted RFLP of 6.5 kb, created as a result of the introduction of a novel EcoRI restriction site in the targeting vector.
Southern analysis using a 5'-361-bp external PCR probe in conjunction with EcoRI-digested ES cell DNA. As depicted in Fig. 1, the endogenous wild-type allele yielded a band hybridizing at 7.7 kb, whereas the targeted allele revealed a predicted RFLP at 6.5 kb due to the introduction of a novel EcoRI site from the p38α targeting vector. The parental heterozygous p38α ES cell line was subsequently cultured in vitro in SCML supplemented with 2 mg/ml G418, according to published methods, to generate p38α-/- cells (47).

Differentiation of ES Cells. Cultures of wild-type and p38α-/- ES cells were maintained in the absence of SCML for 16 d to promote formation of embryoid bodies. These structures were disassociated by trypsin digestion to generate single cell suspensions, and the cells were seeded into IMDM containing 4.5 × 10^-4 M monothioglycerol, 15% fetal bovine serum, 2 mM glutamine, 1% methylcellulose, 0.1 ng/ml IL-3, 10 ng/ml IL-6, and 3 ng/ml GM-CSF to promote differentiation to a myeloid-like phenotype. Recombinant murine IL-3, IL-6, and GM-CSF were obtained from Life Technologies. The cultures were maintained for 12 d, then were harvested and stained with PE-labeled anti-murine type 1 IL-1R antibody (PharMingen) or with a control PE-labeled IgG (Caltag). The antibody-stained cells were analyzed by FACScal, and IL-1R+ positive cells were collected.

Generation of PEFs. PEFs were isolated from day 12 DBA/2 Iafl embryos by established methods (48). In contrast, p38α-/- embryos were not viable at 12 d postcoitum. Therefore, to derive p38α-/- PEF cell lines, attempts were made to rescue the early embryonic lethality in chimeric embryos p38α-/- ES cells were microinjected into day 2.5 C57BL/6 blastocysts, and embryos were not viable at 12 d postcoitum. Therefore, to derive p38α-/- ES cells were maintained in the absence of SCML for 16 d to promote differentiation of embryoid bodies. These structures were disassociated by trypsin digestion to generate single cell suspensions, and the cells were seeded into IMDM containing 4.5 × 10^-4 M monothioglycerol, 15% fetal bovine serum, 2 mM glutamine, 1% methylcellulose, 0.1 ng/ml IL-3, 10 ng/ml IL-6, and 3 ng/ml GM-CSF to promote differentiation to a myeloid-like phenotype. Recombinant murine IL-3, IL-6, and GM-CSF were obtained from Life Technologies. The cultures were maintained for 12 d, then were harvested and stained with PE-labeled anti-murine type 1 IL-1R antibody (PharMingen) or with a control PE-labeled IgG (Caltag). The antibody-stained cells were analyzed by FACScal, and IL-1R+ positive cells were collected.

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Reverse Transcription PCR Analysis of p38 Kinase Expression. Reverse transcription (RT)-PCR was performed on 1 μg of total ES cell or liver RNA in a 20-μl reaction mixture containing 1× PCR buffer (Life Technologies) supplemented with 500 μM each dNTP, 2.5 mM MgCl₂, 0.01 M DTT, and 200 U SuperScript II reverse transcriptase (Life Technologies). The RT reaction was carried out for 1 h at 42°C, followed by denaturation at 70°C for 15 min. Aliquots (2 μl) of each RT reaction mixture were then amplified in a 50-μl reaction volume containing 1× PCR buffer, 1.4 mM MgCl₂, 200 μM each dNTP, 0.4 μM each primer, and 1 U Taq polymerase (Boehringer Mannheim). PCR cycling conditions were 30 cycles at 94°C for 30 s, 55°C for 30 s, and 72°C for 1 min. In addition to using liver as a positive tissue control for p38 isoform expression, amplification of the housekeeping gene β-actin was carried out as a positive control for the RT reaction. RT-PCR was also performed on all RNA samples in the absence of the reverse transcriptase to eliminate the possibility that contaminating DNA contributed to the PCR amplification; no transcripts corresponding to the p38α species were detected (data not shown).

Primers employed in the RT-PCR analysis consisted of the following: p38α, 5'-ATCTTCTCCAGATGTCACGCC-3' and 5'-ATCCTTCAGGATTCGCAGACG-3'; p38β, 5'-ATCCATCGAGGATTCGCCG-3' and 5'-CCTCCCATGGTCACGCTCC-3'; p38γ, 5'-ATCCATCGAGGATTCGCCG-3' and 5'-CCTCCCATGGTCACGCTCC-3'; p38δ, 5'-ATCCATCGAGGATTCGCCG-3' and 5'-CCTCCCATGGTCACGCTCC-3'; p38ε, 5'-ATCCATCGAGGATTCGCCG-3' and 5'-CCTCCCATGGTCACGCTCC-3'; and 5'-AACAGGCTTGTCAGGCAACT-3' and 5'-CTCCTTCAGGATTCGCAGACG-3'. To ensure that no other murine genes would cross-hybridize to these specific primer nucleotide sequences, they were prescreened against the BLASTN sequence similarity database (available at ncbi.nlm.nih.gov). PCR products were size separated by electrophoresis on a 2% agarose gel, and were visualized by ethidium bromide staining.

IL-6 ELISA. Wild-type and p38α-/- ES cells and PEFs were cultured overnight in 1 ml SCML containing no effector, human TNF-α (10–20 ng/ml), or human IL-1β (10–20 ng/ml); the recombinant cytokines were obtained from Upstate Biotechnology. W here indicated, the CSAD SB-203,580 (Calbiochem) was added to the culture medium. Media were harvested, clarified by centrifugation, and analyzed in an IL-6 ELISA (Endogen). The assay was performed as instructed by the manufacturer.

Western Blot Analysis. Cell monolayers (3.5-cm dishes) were washed with PBS, after which 0.1 ml of 2× concentrated Laemmli sample buffer (49) was added, and cells were dislodged by scraping. Cell lysates were transferred to Eppendorf centrifuge tubes and subjected to several bursts of a microtip sonicator to reduce viscosity. These same samples were subsequently boiled for 3 min, and 20 μl of each extract was subjected to SDS-PAGE using 12% Novex minigels. When completed, gels were boiled for 3 min, and 20 μl of each extract was subjected to SDS-PAGE using 12% Novex minigels. When completed, gels were blotted onto Trans-Blot transfer medium (0.45 μm nitrocellulose; Bio- Rad Laboratories). The resulting blots were blocked by immersion in TBS (10 mM Tris, pH 8, 150 mM NaCl, and 0.1% Tween-20) containing 5% nonfat dry milk for 1 h at room temperature. Blots were rinsed with TBS, then incubated overnight at 4°C in TBS containing 5% BSA and a 1:1,000 dilution of the appropriate primary antibody. For detection of total and phosphorylated p38, a rabbit polyclonal anti-p38 MAP kinase antibody (New England Biolabs) and a rabbit polyclonal antiphospho-p38 MAP kinase antibody (New England Biolabs) were used. After treatment with the primary antibodies, blots were washed with three changes of TBS, then incubated in TBS containing 5% nonfat dry milk with a 1:2,000 dilution of horseradish peroxidase-conjugated goat anti-rabbit IgG. After a 60-min incubation, blots were washed three times with TBS, then placed in Lumiglo™ (New England Biolabs). Immune complexes were visualized by exposure to X-ray film.

MAPKAP Kinase 2 Assay. ES cells were seeded (4 × 10⁶ cells) into 6-well cluster dishes and maintained in SCML medium. The growth medium was removed and replaced with 1 ml of medium containing, where indicated, 50 ng/ml aminosyn or 0.5 mM sodium arsenite, and the cultures were incubated for 15 min at 37°C. Monolayers were then rinsed with PBS and treated with 1 ml PBS containing 1% Triton X-100 to promote cell lysis. After a 10-min incubation on ice, extracts were clarified by centrifugation for 20 min at 50,000 rpm in a TLA-100.3 ultracentrifuge rotor (Beckman Instruments). MAPKAP kinase 2 activity within the resulting supernatants was determined using a peptide-based substrate obtained from Upstate Biotechnology. In brief, 5 μg sheep anti-rabbit MAPKAP kinase 2 antibody was incubated with 1 ml of a 5% suspension of protein G-sepharose in PBS containing 1% Triton X-100 and Complete® protease inhibitor (Boehringer Mannheim). After a 60-min incubation, beads were recovered by centrifugation and washed twice with 50 mM Tris pH 7.5, 1 mM EDTA, 1 mM EGTA, 0.5 mM sodium orthovanadate, 0.1% β-mercaptoethanol, 1% Triton X-100, 5 mM sodium pyrophosphate, 10 mM sodium glycerophosphate, 0.1 mM PM SF, 1 μg/ml leupeptin, 1 μg/ml pepstatin, and 50 mM sodium fluoride (buffer A). Individual bead pellets were suspended in 1 ml of the clarified buffer.
cell extracts, incubated at 4°C for 2 h with gentle rocking, and collected by centrifugation. The beads were washed once with 0.5 ml of buffer A containing 0.5 M sodium chloride, once with 0.5 ml of buffer B, and once with 0.1 ml of 20 mM 4-morpholine propanesulfonic acid, pH 7.2, containing 25 mM β-glycerophosphate, 5 mM EGTA, 1 mM sodium orthovanadate, and 1 mM DTT (kinase buffer). Final bead pellets were suspended in 40 µl of an assay mixture composed of kinase buffer supplemented with 125 µM ATP, 10 mM MgCl₂, 100 µM MAPKAP kinase 2 substate peptide, and 250 µCi/ml [γ-32P]ATP (DuPont). Reaction mixtures were incubated at 30°C for 30 min, after which 25 µl of each was spotted onto individual p81 phosphocellulose squares (Whatman). These filters were washed with four changes of 0.75% acetic acid, 75% ethanol, and air-dried. Filters were then incubated in 1 ml of liquid scintillation fluid for radioactivity determination.

Results

Embryonic lethality associated with p38α deficiency. Mice ES cells in which the p38α gene was disrupted by homologous recombination were generated using the scheme shown in Fig. 1. The targeted ES cells successfully contributed to the germline of resulting chimeric mice. F1 generation +/+—animals were grossly normal, fertile, and displayed no discernible phenotype. Contrary to expected Mendelian segregation, offspring from the F2 generation demonstrated an embryonic lethal phenotype. To determine whether inflammatory cytokine signaling cascades were operative in PEF and ES cells, individual cultures were treated with IL-1 or TNF-α, after which IL-6 released into the medium was determined by ELISA. In the absence of a cytokine effector, PEFs isolated from wild-type embryos produced minimal quantities of IL-6 (Fig. 4 A). However, in response to IL-1 stimulation, these fibroblasts generated large quantities of extracellular IL-6 (Fig. 4 A). Likewise, PEFs responded to TNF-α and generated IL-6, but cytokine levels produced in response to TNF-α were less than those elicited by an equivalent concentration of IL-1 (Fig. 4 A). To assess whether ES cells express all known p38 family members, RT-PCR was performed on total RNA prepared from wild-type (+/+ ) DBA/252 ES cells and from two cell lines that demonstrated either heterozygosity (+/− ) or the complete absence (−/− ) of the p38α allele by Southern blotting. This analysis indicated that wild-type ES cells express mRNAs for p38α, β, γ, and δ (Fig. 3). Likewise, total RNA prepared from liver of an adult DBA wild-type mouse contained transcripts encoding each of the four p38 kinases, although the β species appeared to be in low abundance (Fig. 3). In contrast, total RNA isolated from p38α−/− ES cells showed no evidence of the p38α transcript, but these cells maintained expression of p38β, γ, and δ (Fig. 3). p38α−/− ES cells, on the other hand, yielded RNA transcripts corresponding to each of the four p38 kinases. Within the limits of the RT-PCR approach, no obvious change in the relative expression levels of the p38β, γ, and δ species was observed between the +/+ and −/− ES cell lines.

Comparison of the IL-1 responsiveness of PEF and ES cells. Inflammatory cytokines such as IL-1 can promote IL-6 production by cell types that express the appropriate signaling receptors (35, 36). To determine whether inflammatory cytokine signaling cascades were operative in PEF and ES cells, individual cultures were treated with IL-1 or TNF-α, after which IL-6 released into the medium was determined by ELISA. In the absence of a cytokine effector, PEFs isolated from wild-type embryos produced minimal quantities of IL-6 (Fig. 4 A). However, in response to IL-1 stimulation, these fibroblasts generated large quantities of extracellular IL-6 (Fig. 4 A). Likewise, PEFs responded to TNF-α and generated IL-6, but cytokine levels produced in response to TNF-α were less than those elicited by an equivalent concentration of IL-1 (Fig. 4 A). Attempts to isolate p38α−/− PEFs from day 12 chimeric embryos were unsuccessful; fibroblast-like cells were recovered after G418 selection, but these did not survive during culture. Unstimulated ES cells did not produce IL-6, and addition of IL-1 or TNF-α to their medium did not result in an increase in IL-6 production (Fig. 4 A). This lack of responsiveness suggests that DBA/1Tac ES cells lack signaling receptors for IL-1 and TNF, as has previously been reported for ES cells derived from other mouse strains (50).

IL-1-induced IL-6 production by PEFs appeared to be dependent on p38. Stimulation of these fibroblasts with IL-1 in the presence of the CSAID SB-203,580 inhibited production of IL-6 in a dose-dependent manner (Fig. 4 B); a

Figure 2. Southern analysis of ES cell clones. EcoRI restriction enzyme-digested ES cell genomic DNA was electrophoresed, transferred to nitrocellulose, and hybridized to the 5′ external probe of the p38α locus. (A) A blot showing wild-type (+/+ ) and the parental heterozygous (+/− ) ES cell clones. (B) A blot showing examples of G418-surviving ES cell clones; this blot is representative of 64 surviving clones. The positions of the wild-type (WT) and knockout (KO) alleles are indicated.

Figure 3. Expression of the four murine p38 family members, RT-PCR was performed on total RNA prepared from wild-type (+/+ ) DBA/252 ES cells and from two cell lines that demonstrated either heterozygosity (+/− ) or the complete absence (−/− ) of the p38α allele by Southern blotting. This analysis indicated that wild-type ES cells express mRNAs for p38α, β, γ, and δ (Fig. 3). Likewise, total RNA prepared from liver of an adult DBA wild-type mouse contained transcripts encoding each of the four p38 kinases, although the β species appeared to be in low abundance (Fig. 3). In contrast, total RNA isolated from p38α−/− ES cells showed no evidence of the p38α transcript, but these cells maintained expression of p38β, γ, and δ (Fig. 3). p38α−/− ES cells, on the other hand, yielded RNA transcripts corresponding to each of the four p38 kinases. Within the limits of the RT-PCR approach, no obvious change in the relative expression levels of the p38β, γ, and δ species was observed between the +/+ and −/− ES cell lines.

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half-maximal inhibitory concentration (IC50) of 3 μM was estimated for the inhibitory effect (Fig. 4 B).

ES Cells and PEFs Respond to Stress Stimuli and Activate p38α. In addition to cytokines, chemical stress stimuli can activate p38 (3, 4). Therefore, ES cells and PEFs were treated with two agents previously demonstrated to activate p38, anisomycin and sodium arsenite. To assess p38 activation, cell lysates were prepared and analyzed by Western blotting both for total p38α content and the presence of phosphorylated p38α. The absolute amount of p38α recovered in extracts of ES cells and PEFs did not change in response to the various activation stimuli (Fig. 5). In contrast, quantities of phosphorylated p38α were not equivalent, and were dependent on the activation stimulus (Fig. 5). ES cells that were pretreated with anisomycin (50 ng/ml) or sodium arsenite (0.5 mM) yielded phosphorylated p38α (Fig. 5). On the other hand, stimulation of the ES cells with IL-1 or TNF-α did not promote p38α phosphorylation (Fig. 5); this lack of cytokine responsiveness again suggests that ES cells do not possess the corresponding cytokine receptors. In addition, LPS did not promote phosphorylation of ES cell p38α. Therefore, ES cells possess p38α, and this enzyme can be activated by chemical inducers of stress such as anisomycin and sodium arsenite, but not by inflammatory cytokines (IL-1 or TNF-α) or by LPS.

PEFs also responded to the chemical stress stimuli to generate phosphorylated p38α. Nonstimulated PEFs possessed a low level of phosphorylated p38α, and treatment with either anisomycin or sodium arsenite elevated levels of the phosphorylated enzyme (Fig. 5). Levels of total p38α in the fibroblasts were not affected by the various stimuli (Fig. 5). In addition, PEFs stimulated with IL-1, and to a lesser degree with TNF-α, yielded elevated levels of phosphorylated p38α (Fig. 5); similarly, LPS treatment yielded elevated levels of the phosphorylated enzyme. Therefore, PEFs differ from ES cells in that they generate phosphorylated p38α in response to chemical stress inducers and selective cytokine activators.

p38α-deficient ES cells were subjected to a similar analysis. Extracts derived from anisomycin-treated p38α−/− and p38α−/+ ES cells contained p38α, and this enzyme was phosphorylated in response to the chemical stress inducer (Fig. 6). In contrast, extracts derived from anisomycin-treated p38α−/− ES cells did not contain p38α or the phosphorylated p38α polypeptide species (Fig. 6).
MAPKAP Kinase 2 Activation Is Dependent on p38α. Previous studies established that MAPKAP kinase 2 is a substrate of p38α (4). However, the extent to which other kinases may participate in vivo in the activation of MAPKAP kinase 2 remains unclear. To determine whether stress activation of MAPKAP kinase 2 in ES cells is p38α-dependent, +/+ and −/− ES cells were stimulated with anisomycin or sodium arsenite, and cell extracts were isolated. MAPKAP kinase 2 was subsequently recovered by immunoprecipitation, and kinase activity associated with the immunoprecipitates was assessed. In the absence of a stimulus, both p38α+/+ and p38α−/− ES cells yielded low levels of immunoprecipitable MAPKAP kinase 2 activity (Fig. 7 A). Pretreatment with anisomycin or sodium arsenite stimulated MAPKAP kinase 2 activity in extracts derived from p38α+/+ ES cells. Sodium arsenite-stimulated cells achieved a greater level of activity than did the anisomycin-treated cells (Fig. 7 A). In contrast to this large increase in MAPKAP kinase 2 activity, p38α−/− cells stimulated with anisomycin or sodium arsenite demonstrated greatly attenuated responses (Fig. 7 A). No significant increase in MAPKAP kinase 2 activity was observed when p38α−/− cells were stimulated with anisomycin, and sodium arsenite treatment yielded only a fourfold increase in activity (Fig. 7 A). Sodium arsenite treatment of p38α+/+ ES cells on the other hand, yielded >800-fold increase in MAPKAP kinase 2 activity relative to the nonstimulated level (Fig. 7 A).

Sodium arsenite-induced MAPKAP kinase 2 activation was inhibited by the CSAID SB-203,580. p38α+/+ ES cells treated with sodium arsenite in the presence of 2 μM SB-203,580 yielded 67% less MAPKAP kinase 2 activity than did their counterparts activated in the absence of the CSAID (Fig. 7 B). Likewise, the modest increase in MAPKAP kinase 2 activity observed when p38α−/− ES cells were stimulated with sodium arsenite was inhibited by 63% in the presence of SB-203,580 (Fig. 7 B). Therefore, absence of p38α greatly impaired the ability of stress stimuli to promote MAPKAP kinase 2 activation, but a small CSAID-sensitive component remained in the p38α-deficient cells.

ES C cells differentiated in vitro to IL-1R-positive fibroblast-like cells demonstrate an impaired response to IL-1 in the absence of p38α. ES cells can undergo differentiation-type processes when maintained under appropriate culture conditions (51). In an attempt to derive cells expressing IL-1R, p38α+/+ and p38α−/− ES cells were allowed to form embryoid bodies. These structures then were dissociated, and the resulting individual cells were cultured for an additional 12 d in the presence of a cytokine mixture composed of IL-3, IL-6, and GM-CSF. The resulting cell populations were separated by FACS® on the basis of binding of murine anti-IL-1R antibodies. The percentage of cells staining positive for IL-1R was comparable in both the p38α+/+ and p38α−/− cultures, representing 19 and 16%, respectively (Fig. 8 A). Staining above a nonimmune IgG control was used as a specificity control. Before the in vitro differentiation process, neither ES cell population stained positive for type 1 IL-1R (data not shown).

FACS®-sorted IL-1R-positive cells were cultured in the presence of IL-1. Cytokine stimulation of the IL-1R-positive cells

Figure 6. Demonstration that p38α+/− ES cells lack immunodetectable p38α. Wild-type (ES+/+) and p38α-deficient ES cells were stimulated with sodium arsenite for 15 min and then solubilized by detergent extraction. Duplicate samples of each extract were loaded onto separate gels and processed for Western blotting. Antigen detected after staining these blots with an antibody against total (p38) or one selective for phosphospecific (phospho-p38) forms are indicated. Extracts of IL-1-stimulated PEFs and C6 cells are included as standards. Arrow indicates the migration of p38α

Figure 7. MAPKAP kinase 2 activation in wild-type and p38α−/− ES cells. Cultures of p38α+/+ and p38α−/− ES cells were incubated with the indicated effector for 15 min, after which the cells were solubilized by detergent extraction, and MAPKAP kinase 2 was recovered by immunoprecipitation. The resulting immunoprecipitates were then assayed for kinase activity using a peptide substrate and [γ-32P]ATP. Radioactivity (in cpm) incorporated into the peptide substrate is indicated as a function of treatment; a background (no enzyme control) was subtracted to correct for non–peptide-associated radioactivity sticking to the filter. This experiment was repeated twice with comparable results. Each value is the average of duplicate determinations.

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tive p38α+/+ cells resulted in expression of IL-6 (Fig. 8 B); in the absence of IL-1, little IL-6 was generated (Fig. 8 B). Coincubation with SB-203,580 during the IL-1 activation period led to a dose-dependent decrease in IL-6 production (Fig. 8 B). The IC50 for this response was estimated to be 2 μM, a value consistent with the IC50 observed in PEFs (Fig. 1). In contrast, cultures of IL-1R–positive p38α2/2 cells generated much less IL-6 in response to IL-1 stimulation, and this increase was not reduced by the CSAID (Fig. 8).

Absence of p38α Does Not Alter ES Cell Apoptosis. p38α has been implicated as an important regulator of apoptotic processes (52, 53), and we considered the possibility that a defect in apoptosis contributed to the embryonic lethality observed with the knockout animals. To assess the apoptotic responsiveness of the p38α1/1 and p38α2/2 ES cells, they were initially treated with anti-Fas, a well-characterized inducer of apoptosis (53). However, DBA ES cells failed to bind fluorescently labeled anti-Fas antibody (as detected by FACS®; data not shown), negating use of the Fas-induced apoptotic mechanism. Therefore, chemical stimuli were used to promote apoptosis of p38α1/1 and p38α2/2 ES cells. Staurosporin induces apoptosis in many cell systems, and this process is accompanied by caspase-dependent cleavage of the cytoplasmic protein poly(ADP) ribose polymerase (PARP [54]). p38α1/1 ES cells treated overnight with increasing concentrations of staurosporin demonstrated a dose-dependent increase in levels of the 85-kD PARP cleavage product as assessed by Western analysis (Fig. 9 A). Appearance of the PARP cleavage fragment within extracts of p38α1/1 ES cells demonstrated a similar dependence on staurosporin concentration (Fig. 9 A). No significant difference in the responsiveness of the two cell types was noted in multiple experiments.

Figure 8. IL-1–induced IL-6 production by in vitro–differentiated ES cells. Wild-type and p38α−/− ES cells were subjected to in vitro differentiation. (A) Cells recovered from each culture were stained with a PE-labeled control IgG or PE-labeled anti–IL-1R IgG, and the cell mixtures were analyzed by FACS®. Cells were gated by autofluorescence, an indicator of size (x-axis), and by PE fluorescence intensity (y-axis). The percentage of cells demonstrating a PE fluorescence intensity above the background level is indicated in each panel. (B) IL-1R–positive cells recovered by sorting were plated into culture wells and stimulated with IL-1β (10 ng/ml) in the absence or presence of SB-203,580. After an overnight stimulation, media were harvested and assayed for IL-6 by ELISA. The amount of IL-6 produced is indicated as a function of treatment. These results are representative of four separate experiments. wt, wild-type.

Figure 9. Deletion of p38α does not inhibit ES cell apoptosis in response to chemical stimuli. p38α1/1 and p38α2/2 ES cells were treated with the indicated concentration of staurosporin (A) or adriamycin (B) for 16 h, and then were disaggregated with SDS sample buffer. Samples of the resulting lysates were subjected to Western blot analysis with an anti-PARP antibody. Regions of the blots corresponding to full-length 116-kD PARP and its 85-kD cleavage fragment are shown as a function of the effector concentration.
Adriamycin was recently reported to induce caspase-dependent apoptosis of murine ES cells (55), and sensitivity to this agent was also characterized. Extracts prepared from both p38α+/+ and p38α−/− ES cells yielded the 85-kD PARP cleavage fragment in response to adriamycin treatment (Fig. 9 B), and concentrations of this agent required to generate the cleavage fragment again were similar between the two cell lines (Fig. 9 B).

Apoptosis was also assessed by flow cytometry using the annexin V/propidium iodide staining procedure (56). Adriamycin concentrations between 0.05 and 4.5 μg/ml led to a dose-dependent decrease in cell viability (Table I). Comparison of the sensitivity of p38α+/+ and p38α−/− ES cells revealed a similar concentration dependence in the annexin V/propidium iodide staining profiles (Table I). Moreover, addition of 10 μM SB-203,580 to the culture medium did not significantly alter the percentages of viable cells detected in the adriamycin-treated p38α+/+ and p38α−/− ES cell populations (Table I).

Discussion

p38 is activated by a variety of extracellular stimuli, including inflammatory cytokines such as IL-1 and TNF, growth factors such as fibroblast growth factor (FGF) and CSF, osmolarity changes, UV light, and chemical agents that promote stress response (7). The diverse nature of these activators suggests that p38 serves as a point of convergence for a variety of extracellular effectors. Based on this multiplicity of activators, it is perhaps not surprising that deletion of p38α led to embryonic lethality. A similar fate has been observed for mice engineered to lack other signaling kinases including MKK4 (50, 57); function of these signaling kinases must be essential to normal development. Several individual p38 kinases initiate separate cellular processes (27).

Generation of cells deficient in p38α allowed us to explore this enzyme’s role in several stress response pathways. Chemical agents such as anisomycin and sodium arsenite promote activation of p38 in a variety of cell types (3, 4). Indeed, p38α−/− ES cells and PEFs treated with these agents demonstrated increased phosphorylation of p38α and an increased level of MAPKAP kinase 2 activity. In contrast, p38α−/− ES cells failed to efficiently activate MAPKAP kinase 2 in response to anisomycin or sodium arsenite exposure. Therefore, p38α appears to be a critical component of the MAPKAP kinase 2 activation cascade in response to chemical stress inducers. Interestingly, a small increase in MAPKAP kinase 2 activity persisted after sodium arsenite treatment of p38α−/− ES cells and, as in wild-type ES cells, this increase was blocked by SB-203,580. The magnitude of the CSAID effect against the residual activity was similar to that observed in wild-type cells. Since human p38β, in contrast to the γ and δ isoforms, is CSAID sensitive (20, 21, 23–25), perhaps the mouse equivalent is responsible for the attenuated response.

Mouse PEFs treated with IL-1, and to a lesser degree with TNF−α, responded by phosphorylating p38α and producing and/or secreting IL-6. Fibroblast IL-6 production was inhibited by SB-203,580. ES cells, on the other hand, lacked receptors for IL-1 and TNF−α; IL-1 (or TNF) failed to significantly alter cell viability in these cells (Table I). Since mice, like humans, appear to express multiple p38 kinase family members (59), the developmental arrest suggests that the different enzymes do not perform redundant activities, at least during embryonic development. Deletion of p38α also has been associated with murine embryonic lethality in a 129SvEv × C57BL/6 genetic background, where the arrest was attributed to a defect in placental development (60).

The four human p38 kinases display distinct expression patterns (23, 24). By Northern blot analysis, transcripts for p38α, p38β, and p38δ were found in many human tissues, but the relative abundance of an individual species varied widely from tissue to tissue (23). In contrast, p38γ transcripts were abundantly expressed only in skeletal muscle, with minimal expression in most other tissues (23). Not only is expression regulated, but also within the same cell type two forms of the enzyme may demonstrate differential activation requirements. For example, human macrophages express both p38α and p38δ, but p38α was activated to a greater extent than was p38δ after LPS stimulation (26). This differential activation may be achieved through the use of distinct upstream activators (20, 61, 62). For example, p38α was efficiently activated by MKK3, MKK4, or MKK6 in transfected COS-7 cells, whereas p38β was phosphorylated only by MKK6 (20). Finally, forced activation of overexpressed p38α and p38β was reported to promote cardiomyocyte apoptosis and hypertrophy, respectively, suggesting that the individual p38 kinases initiate separate cellular processes (27).

Table I. ES Cell Viability after Adriamycin-induced A apoptosis

| Adriamycin Concentration (μg/ml) | p38α+/+ | p38α−/− | p38α+/+ | p38α−/− |
|----------------------------------|---------|---------|---------|---------|
|                                  | %       | %       | %       | %       |
| 0.05                             | 72      | 64      | 68      | 66      |
| 0.16                             | 51      | 48      | 43      | 36      |
| 0.5                              | 44      | 37      | 30      | 29      |
| 1.0                              | 28      | 27      | 22      | 21      |
| 1.5                              | 21      | 19      | 17      | 24      |
| 4.5                              | 23      | 20      | 17      | 19      |

| µg/ml | % | % | % | % |
|-------|---|---|---|---|
| 0     | 72| 64| 68| 66|
| 0.05  | 51| 48| 43| 36|
| 0.16  | 44| 37| 30| 29|
| 0.5   | 28| 27| 22| 21|
| 1.0   | 21| 19| 17| 24|
| 1.5   | 23| 20| 17| 19|

ES cells (2 × 106 cells per well in 6-well cluster plates) were incubated for 18 h with the indicated concentration of adriamycin. Where indicated, SB-203,580 was also present at 10 μM. The cells were subsequently harvested and analyzed by flow cytometry after staining with FITC-labeled annexin V and propidium iodide. The percentage of viable cells (those not staining with either annexin V or propidium iodide) was determined based on 10,000 gated events. Data are representative of three separate experiments.
to promote p38α phosphorylation and to stimulate IL-6 production. However, after wild-type ES cells were subjected to an in vitro differentiation process, a percentage of the population expressed the type 1 IL-1R. These receptor-positive cells responded to IL-1 and generated IL-6 via an SB-203,580-inhibitable process.

p38α+/− ES cells also gave rise to a population of IL-1R–positive cells after the in vitro differentiation process. Based on the percentage of cells that expressed this receptor, no difference in the efficiency of the differentiation response to an IL-1R–positive state was observed between p38α+/− and p38α−/− cell types. A large number of cytokine and/or growth factor signaling cascades have been reported to involve p38 (7); for example, SB-203,580 is reported to inhibit GM-CSF-induced cell proliferation (63). Moreover, p38 has been implicated as an important element in cellular differentiation processes. For example, the terminal differentiation of L8 muscle cells is blocked by SB-203,580 (64). Therefore, the similar efficiency at which wild-type and p38α−/− ES cells differentiated to an IL-1R–positive phenotype was somewhat of a surprise. However, the IL-1R–positive p38α−/− cells were greatly impaired in their IL-1 responsiveness as judged by a reduction in IL-6 production relative to wild-type cells.

RT-PCR indicated that wild-type ES cells express all four p38 kinase family members. Knockout of the p38α allele eliminated expression of p38α mRNA without altering expression of the other three kinases. Therefore, the gene-targeting strategy successfully eliminated expression of p38α individually. Despite loss of just one of the four p38 kinases, p38α−/− ES cells, in contrast to their wild-type counterparts, did not respond to anisomycin or sodium arsenite to activate MAPKAP kinase 2, and they did not generate IL-6 in response to IL-1 stimulation after in vitro differentiation. With respect to these activities (and within the context of the ES cell), therefore, p38α does not serve a redundant role to its relatives. The markedly reduced responsiveness of p38α−/− ES cells is surprising based on literature reports indicating that MAPKAP kinase 2 is an in vitro substrate for both p38α and p38β family members (25), and evidence that IL-1 treatment can promote activation of all p38 paralogs (23, 25, 26, 59).

p38 kinases have also been implicated as regulators of apoptosis. For example, activation of p38α is reported to be required for apoptosis of PC-12 cells after nerve growth factor withdrawal (52). Likewise, serum removal from rat-1 fibroblasts led to p38 activation and to apoptosis (40). In contrast, an inhibitor of p38, SB-202,190, is reported to promote apoptosis of Jurkat cells (39); overexpression of p38β, but not p38α, attenuated the SB-202,190 effect, suggesting that the response was isoform dependent. ES cells demonstrated an apoptotic response when treated with the chemical inducers of apoptosis, staurosporin or Adriamycin. Two separate indicators of apoptosis, PARP cleavage and annexin V binding, were observed in ES cells treated with the chemical inducers. Importantly, p38α+/− and p38α−/− ES cells were equally sensitive to the chemical-induced apoptotic response. This suggests that p38α does not play a dominant role in this form of ES cell apoptosis. Moreover, the ES cell apoptotic response was not affected by SB-203,580, suggesting that other CSAID-sensitive proteins also were not involved in the death response.

The ability of CSAIDs to modulate inflammatory cytokine production by monocytes and macrophages highlights p38 as an attractive pharmaceutical target. Human monocytes were recently reported to express both p38α and p38δ, and macrophages derived from in vitro differentiation of blood monocytes continued to express both of these kinases (26). In contrast, p38β and p38γ were not expressed in the monocytes and/or macrophages (26). When macrophages were stimulated with LPS, a large increase in p38α activity was observed, with only a modest activation of p38δ (26). Therefore, from the standpoint of regulating monocyte and/or macrophage inflammatory function, it appears that p38α is the key p38 kinase. The data presented in this report using in vitro-differentiated ES cells clearly support p38α serving as a key mediator of IL-1 signal transduction. IL-1R–positive p38α+/+ cells responded to IL-1 and generated IL-6. In contrast, p38α−/− IL-1R–positive cells generated minimal quantities of IL-6. IL-1, therefore, appears similar to LPS in its utilization of p38α to achieve target cell activation.

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