LETTER TO THE EDITORS

Guillain-Barré syndrome following COVID-19: new infection, old complication?

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Received: 9 April 2020 / Revised: 13 April 2020 / Accepted: 17 April 2020 / Published online: 24 April 2020 © Springer-Verlag GmbH Germany, part of Springer Nature 2020

Dear Sirs,

The coronavirus disease-2019 (COVID-19) pandemic originated in Wuhan (China) on December 2019. So far, more than 1,500,000 cases have been confirmed worldwide (> 150,000 cases in Italy) [1]. Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) infection is a systemic disorder typically presenting with fever, fatigue and prevalent respiratory disturbances (dry cough, dyspnea, chest pain, interstitial pneumonia) [2], although neurological manifestations are increasingly reported [3]. Guillain-Barré syndrome (GBS) is an acute/subacute immune-mediated polyradiculoneuropathy characterized by varying degrees of limbs or cranial-nerves weakness, loss of deep tendon reflexes, sensory and dysautonomic symptoms due to peripheral nerves and roots demyelination and/or axonal damage [4]. According to a recent review about two-third of all GBS are preceded by upper respiratory infection or enteritis [4]. Here we describe a case of GBS following a clinically resolved paucisymptomatic COVID-19.

A 70-year-old-woman was referred to our emergency department (ED) on Mar-28 complaining of asthenia, hands and feet paresthesia and gait difficulties progressing within 1 day. On Mar-4 she had developed fever (body temperature—BT = 38.5 °C) and dry cough. A day later she had been tested positive for SARS-CoV-2-RNA on RT-PCR with a nasopharyngeal swab. Symptoms of COVID-19 had resolved in a few days. The epidemiological survey had revealed a previous hospital visit to an inpatient in an area with high incidence for COVID-19 (Piacenza, Italy) on Feb-28, 29.

At the ED admission BT was 36.5 °C, oxygen saturation was 98% on room air. Arterial blood gas analysis showed pO2 = 76 mmHg with normal p/f ratio (= 363). Hematological investigations revealed slightly increased white blood cells (10.41 × 10^9/L, normal = 4–10 × 10^9/L) with 8.15 × 10^9/L neutrophils (normal = 2–8 × 10^9/L) and lymphocytes in the normal range. D-dimer, creatine phosphokinase, blood glucose, hepatic and renal function were normal, as well as c-reactive protein, erythrocyte sedimentation rate, folate and vitamin B12 blood levels. A chest high-resolution computed tomography revealed some small “ground glass” areas in both lungs. A repeated nasopharyngeal swab for SARS-CoV-2-RNA was negative. Mycoplasma Pneumoniae and Cytomegalovirus (CMV) serology (IgM and IgG), Legionella Pneumophila and Streptococcus Pneumoniae urinary tests were unrevealing. The neurological examination disclosed moderate (Medical Research Council grade 4/5) symmetric distal upper and lower limbs weakness, loss of deep tendon reflexes, preserved light touch and pinprick sensation. On Mar-31 a lumbar puncture was performed. The cerebrospinal fluid (CSF) analysis revealed slight aluminocytological dissociation (CSF proteins = 48 mg/dL, normal = 0–40 mg/dL, white blood cells = 1 × 10^6/L, normal = 0–8 × 10^6/L). Microbiologic testing on CSF was negative (including herpes simplex virus, varicella zoster virus, Epstein-Bar virus, CMV, HIV-1, Borrelia Burgdorferi IgM and IgG). Neurophysiologic findings were consistent with a diagnosis of GBS (Table 1), according to current criteria [5]. A trial with 400 mg/die intravenous immunoglobulin (IV Ig) for 5 days was started. On Apr-1 the patient was intubated and mechanical ventilation was applied, because of respiratory failure due to the worsening of muscle weakness.

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Figure 1 displays timeline of main clinical events and diagnostic investigations.

To date, only a prior case of GBS concomitant with SARS-CoV-2 infection and a parainfectious profile has been reported [6]. In our patient, the temporal evolution of neurological manifestations resembles that of a postinfectious etiology, although a single repeated negative nasopharyngeal swab was available. Therefore, we may speculate an association between the acute polyradiculopathy and SARS-CoV-2 infection. This hypothesis is supported by the notion of 24-days home isolation before the onset of neurological symptoms and by the comprehensive exclusion of most

| Table 1 Results of neurophysiologic study |
|------------------------------------------|
| Motor nerve conductions                  |
| Distal latency (ms)                      |
| Amplitude (mV)                           |
| Conduction velocity (m/s)                |
| F waves                                  |

**Median nerve**
- Wrist-abductor pollicis brevis: R = NE; L = 4.5 (n.v. < 4.2)
- Antecubital fossa-wrist: R = NE; L = 12.3
- Amplitude: R = 1.3; L = 4.5 (n.v. > 6)
- Conduction velocity: R = 38.0; L = 30.8 (n.v. > 50)

**Ulnar nerve**
- Wrist-abductor digitii minimi: R = 4.6; L = 1.5 (n.v. < 3.2)
- Below elbow-wrist: R = 9.7; L = 9.8
- Amplitude: R = 2.8; L = 2.3 (n.v. > 7)
- Conduction velocity: R = 43.1; L = 30.1 (n.v. > 52)

**Tibial nerve**
- Medial malleolus-abductor hallucis brevis: R = 8.7; L = 8.5 (n.v. < 5)
- Amplitude: R = 1.3; L = 1.1 (n.v. > 5)
- Conduction velocity: R = A; L = A

**Orthodromic sensory nerve conduction studies**
- Left median nerve
  - Digit 3-wrist: NE (n.v. > 4)
- Ulnar nerve
  - Digit 5-wrist: R = 0.4; L = NE (n.v. > 3.5)
  - Amplitude: R = 37.5; L = NE (n.v. > 45)

**Soleus H reflex** was absent bilaterally. Evocable distal compound muscle action potentials (CMAPs) showed reduced amplitude because of temporal dispersion due to demyelination. Needle electromyography disclosed reorganization of motor units with polyphasic and long-duration potentials, without signs of acute denervation.

**R** right, L left, n.v. normal values adjusted for age (65–75 years), NE not evocable, A absent

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**Fig. 1** Timeline of clinical events and diagnostic investigations. COVID-19 coronavirus disease-2019, GBS Guillain-Barré syndrome, LP lumbar puncture, IVIg intravenous immunoglobulin, MV mechanical ventilation

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common infectious agents related to GBS (negative IgM/IgG for CMV and Mycoplasma Pneumoniae, negative history for enteritis). However, we could not rule out with certainty a COVID-19 parainfectious neurological syndrome, given the reported suboptimal sensitivity of the RT-PCR for swab due to laboratory error or insufficient viral material in the specimen [7]. Moreover, other less common infectious agents (eg. West Nile and Toscana viruses), which were not tested but are endemic in Northern Italy, might be responsible for the present clinical picture [8]. A shortcoming of our report is the lack of antiganglioside antibody testing to identify specific targets of the autoimmune GBS process [9]. Furthermore, we did not perform a complete paraneoplastic/autoimmune screening in the acute phase including testing for serum onconeural and vasculitis-related antibodies (e.g. antineutrophil cytoplasmic antibodies—ANCA). Hence, we could not exclude the possibility of an autoimmune or paraneoplastic polyradiculoneuropathy mimicking GBS [10]. Nevertheless, the postinfectious onset, the acute clinical course and the typical neurophysiologic findings (polyradiculoneuropathy with predominant demyelination of both motor and sensory fibers, sural sparing pattern), together with the negative history for autoimmune, neoplastic or neurologic antecedences, make these alternative diagnoses less suitable. Another major limitation relies on the unavailability of SARS-CoV-2 serology and CSF validated test (e.g. PCR on CSF) in our center.

Taking together all these findings, the causal association between GBS and COVID-19 remains speculative, but more probable, given that GBS and Bickerstaff’s encephalitis have been already described as postinfectious complications of other coronavirus, sharing similarities with SARS-CoV-2 (Middle East respiratory syndrome, MERS-CoV) [11]. If our hypothesis will be confirmed in larger case series, neurologists and other clinicians should be aware of the important early recognition and treatment of the potential neuromuscular and autonomic worsening leading to cardio-respiratory failure in patients with GBS and mild or controlled pulmonary COVID-19. Notwithstanding the causative relationship remains unproved, we believe that our case description provide further evidence to the heterogeneous and multi-systemic complications associated with SARS-CoV-2. Future research and data acquisition are needed to clarify the possible pathophysiological correlation, as well as to characterize the clinical/electrophysiological pattern of new cases of GBS observed in the context of COVID-19 pandemic.

Availability of data and material

Deidentified data and material inherent to the case report and not included in the manuscript are available on request to the corresponding author by any qualified investigator.

Author contributions PM: acquisition of data, analysis and interpretation of data, drafted the manuscript for intellectual content. MV, AGM, PMG, ARS, QP: acquisition of data, revised the manuscript for intellectual content. CC: acquisition of data, table creation, revised the manuscript for intellectual content, FM: study concept and design, acquisition of data, figure creation, analysis and interpretation of data, study supervision, drafted and revised the manuscript for intellectual content.

Funding Not targeted funding reported.

Compliance with ethical standards

Conflicts of interest The Authors declare no conflicts of interest relevant to the manuscript.

Ethical approval This article does not contain any studies involving human participants performed by any of the Authors.

Informed consent Written informed consent was collected from the patient for the inclusion of deidentified clinical data in a scientific publication, in accordance with the Declaration of Helsinki.

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