Association of *NAT2* polymorphisms with risk of colorectal adenomas: Evidence from 3,197 cases and 4,681 controls

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Received June 4, 2012; Accepted August 29, 2012

DOI: 10.3892/etm.2012.695

**Abstract.** Previous studies have implicated *NAT2* polymorphisms as risk factors for various types of cancer. Colorectal adenomas are recognized as a pre-neoplastic lesion. A growing body of research documenting the association of *NAT2* polymorphisms with the risk of colorectal adenomas has yielded conflicting results. The aim of the present study was to derive a more precise estimation of this association. Meta-analyses assessing the association of *NAT2* variants with colorectal adenomas were conducted and subgroup analyses on smoking status and the source of the controls were also performed. Eligible studies were identified for the period before March 2012. A total of seven case-control studies, including 3,197 cases and 4,681 controls, were selected following extensive searching and screening. In the overall data, no associations between *NAT2* polymorphisms and colorectal adenomas were observed (odds ratio (OR), 1.04; 95% confidence interval (CI), 0.90-1.21). However, in the subgroup analysis concerning smoking status, slow acetylator variants were revealed to be correlated with increased colorectal adenoma risk in individuals who have smoked (OR, 1.31; 95% CI, 1.04-1.64). In conclusion, the data of the present study suggested that *NAT2* polymorphisms may be a risk factor for colorectal adenomas in individuals who have a history of smoking.

**Introduction**

Colorectal cancer is one of the most common malignancies worldwide, particularly in Western populations, and is thought to be correlated with colorectal adenoma, a type of pre-neoplastic lesion. The mechanisms of colon adenoma genesis are still unclear.

Evidence indicates that certain epidemiological factors, including cigarette smoking, alcohol use and meat consumption, may contribute to the risk of colorectal adenomas (1,2). Moreover, exposure to certain toxins, such as the heterocyclic aromatic amines formed during the cooking of meat at high temperatures, may increase colorectal adenoma risk (3). However, although numerous individuals are exposed to environmental risk factors, colorectal adenomas develop in only a small proportion of these individuals. Additionally, previous studies suggested a marked association between colon adenoma risk and a family history of adenomas (4,5), indicating that host genetic factors may play a critical role in the genesis of colorectal adenomas.

Colonic tissue is vulnerable to the effects of external toxins by direct exposure. Xenobiotics can be bio-activated into their ultimate carcinogen forms by phase I enzymes and subsequently detoxified by phase II enzymes, such as *CYP1A1* and *GSTM1*, respectively (6). Genetic variation in the genes encoding these enzymes may affect carcinogen activation/detoxification and modulate DNA repair, possibly by altering the genes’ expression and function. This may result in oncogenesis.

Acetylation is an important biotransformation route for these chemicals. In humans, the N-acetyltransferase 2 (*NAT2*) gene encodes a phase II enzyme that plays an essential role in the metabolism of aromatic heterocyclic amines and hydrazines via N- and O-acetylation (7). Alterations to the *NAT2* acetylator status caused by variations in the *NAT2* gene have been reported to reduce enzymatic activity, resulting in inefficient detoxification and thus leading to increased cancer susceptibility (8). Several *NAT2* genetic variants have been identified in humans, of which *NAT2*\(^*4\) is regarded as the most common allele linked to rapid acetylation. *NAT2*\(^*12A*, *NAT2*\(^*12C*, *NAT2*\(^*13\) and *NAT2*\(^*18\) have also been classified as rapid alleles. The remaining alleles are considered to be slow alleles (9-11).

Published studies have been conducted on the association of *NAT2* genetic variants with colorectal adenoma risk and have yielded inconclusive results. Whether *NAT2* polymorphisms are a risk factor for colorectal adenoma remains uncertain. Therefore, in the present study, evidence-based quantitative meta-analyses of the published studies were performed to derive a more precise estimation of this association.

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**Key words:** *NAT2*, susceptibility, colorectal adenoma, meta-analysis, polymorphism
Materials and methods

Literature search strategy. A search of the Medline, EMBASE, OVID, Sciencedirect, Google scholar and CNKI databases was performed covering all studies published before March 2012 with no language limitations. Combinations of the following keywords were used: NAT2, N-acetyltransferase 2, colon, colorectal, neoplasm, polyp, adenoma and polymorphism. The bibliographies of all the retrieved studies were searched for further relevant publications. Review articles and the bibliographies of other identified relevant studies were searched manually to identify additional eligible studies.

Inclusion criteria. The following criteria were used for the literature selection: i) studies should be concerned with the association of NAT2 polymorphisms with colorectal adenoma risk; ii) studies should be observational (case-control or cohort); iii) studies must present the sample size, odds ratios (ORs) and the corresponding 95% confidence intervals (CIs) and genetic distributions or information that may aid the interpretation of the results. After extensive searching, all studies were reviewed in accordance with the criteria defined above for further analysis.

Data extraction. Data were extracted and entered into a database by two reviewers independently. In the case of conflicting evaluations, agreement was reached following discussion. If a consensus could not be reached, a third reviewer was consulted to resolve the dispute and a final decision was then made according to the majority decision. Carriers of at least one of the high-activity alleles were identified as rapid acetylators and individuals carrying two low-activity alleles were classified as slow acetylators, as stated in the primary literature.

Statistical analysis. The OR of the NAT2 polymorphisms and colorectal adenoma risk was estimated for each study. To detect possible sample size biases, the OR and corresponding 95% CI of each study were plotted against the number of participants. A Chi-square-based Q statistic test was performed to assess heterogeneity. If the result of the heterogeneity test was P>0.1, ORs were pooled according to the fixed-effects model (Mantel-Haenszel). Otherwise, the random-effects model (DerSimonian and Laird) was used. The significance of the pooled ORs was determined using a Z-test.

Publication bias was assessed by visual inspection of funnel plots (12) in which the standard error of the log(OR) of each study was plotted against the corresponding log(OR). An asymmetric plot indicates a possible publication bias. The symmetry of the funnel plot was evaluated using Egger's linear regression test (13). Statistical analyses were performed using the STATA 11.0 software (StataCorp, College Station, TX, USA).

Results

Literature search and meta-analysis databases. As shown in Fig. 1, a total of 59 publications were searched and screened for retrieval, of which 43 irrelevant studies were excluded. Thus, 16 studies were primarily identified, of which one review was excluded (14). Subsequently, two articles which were not case-control studies (15,16) and five studies lacking sufficient information (17-21) were also excluded. Eight publications were identified (22-29). Two studies conducted by Tiemersma et al (22,28) concerned the same research subjects. Thus, the study concerning cigarette smoking was selected (28). Finally, seven case-control studies were selected (23-29).

The included studies were all written in English, of which one involved a Caucasian population (28) and the remaining six involved multi-ethnic populations. We were able to extract information about smoking status from two of the studies (25,28).

We established a database of the extracted information from each study. The relevant information is shown in Table I. The first author and the number and characteristics of cases and controls for each study are presented, as well as the other necessary information. The genetic distributions of the control groups were in Hardy-Weinberg equilibrium. The distributions of the NAT2 acetylator variants (classified as rapid or slow) are presented in Table II.

Test of heterogeneity. As shown in Table III, the heterogeneity of the overall data was significant since the P-value of the Q tests was <0.1 and thus the random-effects model was used. However, when subgroup analysis with regard to the source of the controls was conducted, no heterogeneity in the hospital-based subgroup was found.

Quantitative data synthesis. Table III lists the main results of the meta-analysis. The overall data from the seven studies containing 3,197 cases and 4,681 controls revealed no significant associations between NAT2 polymorphisms and colorectal adenoma risk (OR, 1.04; 95% CI, 0.90-1.21; P=0.043 for heterogeneity), suggesting that NAT2 polymorphisms may have little association with colorectal adenoma risk (Fig. 2).

To further assess the possible impact of smoking and the source of the controls on the results, relevant data were extracted to conduct subgroup analyses. In the subgroup analysis of the source of the controls, no significant associations were observed in either the hospital-based group.
In the smoking status subgroups, the data showed that slow acetylator variants may be associated with increased colonic adenoma risk in smokers (OR, 1.31; 95% CI, 1.04-1.64; P=0.795 for heterogeneity). Nevertheless, no marked associations were observed in the never smoked subgroup (OR, 0.92; 95% CI, 0.53-1.58; P=0.051 for heterogeneity; Fig. 4).

An attempt was made to extract relevant data concerning meat and alcohol consumption to use in the subgroup analysis, but insufficient data were available.

### Table I. Characteristics of the studies included in the meta-analysis.

| First author (ref.) | Publication year | Cases (male/female) | Controls (male/female) | Type of controls | Age range (mean), years Cases Controls Ethnicity Country |
|---------------------|------------------|---------------------|------------------------|------------------|---------------------------------------------|
| Lang (24)           | 1994             | 41 (28/13)          | 205 (129/76)           | 205 volunteers (population-based) | 36-84 (60.6) 20-80 (46.9) Mixed USA |
| Probst-Hensch (26)  | 1996             | 441 (280/161)       | 484 (326/158)          | 484 controls (age- and gender-matched; population-based) | 50-74 (61.7) 50-74 (61.6) Mixed USA |
| Ishibe (23)         | 2002             | 146 (111/35)        | 228 (144/84)           | 228 controls (age- and gender-matched; hospital-based) | 18-74 (58) 18-74 (59) Mixed USA |
| Tiemersma (28)      | 2004             | 431 (236/195)       | 433 (160/273)          | 433 controls (age- and gender-matched; hospital-based) | 18-75 (58.8) 18-75 (50.4) Caucasian Netherlands |
| Moslehi (25)        | 2006             | 772 (535/237)       | 777 (536/241)          | 777 controls (gender- and ethnicity-matched; hospital-based) | 55-74 (NA) 55-74 (NA) Mixed USA |
| Shin (27)           | 2008             | 557 (418/139)       | 1493 (1250/243)        | 1493 controls (hospital-based) | 40-75 (59.6) 40-75 (57.2) Mixed USA |
| Wang (29)           | 2011             | 914 (550/364)       | 1185 (745/440)         | 1185 controls (age-, gender- and ethnicity-matched; hospital-based) | NA (61) NA (62) Mixed USA |

NA, not available.

### Table II. Distribution of NAT2 acetylator variants among colorectal adenoma cases and controls included in the meta-analysis.

| First author | Year | Genotyping method | Cases Rapid | Slow Controls Rapid | Slow |
|--------------|------|-------------------|-------------|----------------------|------|
| Lang         | 1994 | Use of caffeine   | 25          | 16                   | 92   | 113  |
| Probst-Hensch| 1996 | AS-PCR            | 213         | 228                  | 226  | 258  |
| Ishibe       | 2002 | PCR-RFLP          | 64          | 79                   | 98   | 110  |
| Tiemersma    | 2004 | PCR-RFLP          | 168         | 259                  | 179  | 253  |
| Moslehi      | 2006 | Taqman            | 272         | 413                  | 317  | 376  |
| Shin         | 2008 | Taqman            | 243         | 311                  | 609  | 880  |
| Wang         | 2011 | PCR-RFLP          | 449         | 457                  | 631  | 539  |

NAT2, N-acetyltransferase 2; AS-PCR, allele-specific polymerase chain reaction; PCR-RFLP, polymerase chain reaction-restriction fragment length polymorphism.
Sensitivity analysis. In order to compare the differences and evaluate the sensitivity of the meta-analyses, the results of the fixed-effects model for the overall data were also reported. The combined OR and 95% CI were 1.07 and 0.97-1.17, respectively, similar to the results of the random-effects model, suggesting that the meta-analyses were stable. Additionally, one-way sensitivity analysis (30) was also used to evaluate the stability of the meta-analysis. The statistical significance of any of the results was not altered by the omission of any single study, suggesting that the data in this meta-analysis were relatively stable and credible.

Bias diagnostics. A funnel plot was created to assess possible publication biases (Fig. 5A). Egger's linear regression test was then used to assess the symmetry of the plot (Fig. 5B). The data suggested that the funnel plot was symmetrical (t=-1.25, P>0.05), indicating that the results of the meta-analyses were relatively stable and that publication bias had little effect on the results.

Discussion

In the present study, the overall data of the meta-analyses showed that the NAT2 polymorphism may not have a significant association with colorectal adenoma risk. Nevertheless, in subgroup analysis, slow acetylator variants may modify the

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Table III. Main results of the pooled data in the meta-analysis.

| Characteristic        | Number of cases/controls | OR   | 95% CI    | P-value for Q-test |
|-----------------------|--------------------------|------|-----------|--------------------|
| Overall               | 3197/4681                | 1.04 | 0.90-1.21 | 0.043              |
| Smoking status        |                          |      |           |                    |
| Never smoked          | 387/481                  | 0.92 | 0.53-1.58 | 0.051              |
| Have smoked           | 665/561                  | 1.31 | 1.04-1.64 | 0.795              |
| Source of control     |                          |      |           |                    |
| Population-based      | 1167/1382                | 0.96 | 0.66-1.39 | 0.019              |
| Hospital-based        | 2030/3299                | 1.05 | 0.90-1.23 | 0.172              |

OR, odds ratio; 95% CI, 95% confidence interval.
activation of aromatic and heterocyclic amine carcinogens by two pathways. This metabolic reaction may result in detoxification by N-acetylation or bioactivation by O-acetylation which is often proceeded by CYP450 hydroxylation (41). Polymorphisms of \( \text{NAT2} \) may lead to differences in the rate of arylamine metabolism and consequently cancer risk (42). Individuals with a history of smoking and \( \text{NAT2} \) genetic variants who are exposed to cigarette toxins, such as nickel sulphate and benzo(b)fluoranthene, may have a reduced detoxification ability, thus resulting in an increased risk of colon adenoma as well as cancer. However, only two of the studies provided sufficient data concerning smoking status for subgroup analysis. Biases may be noted due to the limited number of selected studies and sample sizes. Therefore, these results should be interpreted with care.

Subgroup analyses of the source of the controls also failed to reveal significant associations in studies using hospital- or population-based controls. Since hospital-based controls may not always truly represent the general population, particularly when the controls have any disease-related conditions relevant to the studied genotypes, inherent selection biases may exist. Further well-designed studies, including population-based controls with strict matching criteria, are required to reduce such possible biases.

Inter-study heterogeneity was observed in this study. Thus, the random-effects model was used. Subgroup analyses were then conducted. As expected, the heterogeneities of the subgroups were reduced or removed. The data suggested that the source of the controls and smoking status may partly contribute to the heterogeneities. Additionally, other factors, including age, gender, ethnicity and the prevalence of lifestyle factors, may also generate heterogeneities.

Publication bias is an important factor that should be considered in a meta-analysis. In the present study, a funnel plot and Egger’s test were used to assess publication biases. The results indicated that publication biases exerted little effect on the overall results, suggesting that the meta-analysis was robust and credible.

Certain limitations may be included in this study. Firstly, in this meta-analysis, the majority of published studies and papers written in English were searched. However, it is possible that some related published or unpublished studies that may have met the inclusion criteria were missed. Thus, it is inevitable that publication biases may exist in the results. Secondly, all the studies concerned mixed ethnicities, with the exception of one study by Tiemersma (28) involving a Caucasian population. Therefore, subgroup analysis concerning ethnicity was not conducted and we were unable to assess the possible effects of ethnic variation on the results. Further studies conducted on separated ethnicities in various areas are required. Thirdly, subgroup analyses of age, gender, alcohol use and meat consumption were not performed since insufficient information was available in the primary literature. Gene-gene and gene-environment interactions should also be considered in the further investigations.

In conclusion, although no significant associations of \( \text{NAT2} \) genetic variations with the overall risk of colorectal adenoma were revealed, statistically significant findings were noted in individuals with a history of smoking, suggesting that \( \text{NAT2} \) genetic variations may modify the colorectal adenoma risk of colorectal adenoma susceptibility of individuals who have a history of smoking.

Possible correlations of \( \text{NAT2} \) polymorphisms with cancer risk were evaluated by the meta-analyses. Previously, \( \text{NAT2} \) polymorphisms have been indicated to be associated with an increased susceptibility to prostate cancer and laryngeal cancer in Asian populations (31,32). However, meta-analyses with regard to lung and gastric cancer did not reveal any marked associations (33,34). Similarly, meta-analyses published in 2002, 2006 and 2012 failed to demonstrate a significant association between \( \text{NAT2} \) polymorphisms and colorectal cancer risk (35-37). In the present study, the overall data also failed to reveal a significant association of the \( \text{NAT2} \) variants with colorectal adenoma, in line with the meta-analyses concerning colorectal cancer.

A previous meta-analysis suggested that smoking affected the formation and aggressiveness of colon adenomas (38). Another meta-analysis suggested that smoking may interact with certain gene variants, such as the \( \text{NQO1} \) genetic variant (39), in the development of colorectal adenomas. Thus, we attempted to extract relevant data from the primary literature to conduct a subgroup analysis. The data showed that slow acetylator variants may be associated with an increased adenoma risk in individuals who have a history of smoking. The data indicate possible interactions between smoking with slow acetylator variants in the pathological mechanisms of colorectal adenoma. The underlying mechanisms by which \( \text{NAT2} \) polymorphisms affect colon adenomas in individuals with a history of smoking are not fully understood. The \( \text{NAT2} \) gene is located on chromosome 8p22, encodes a 290-amino acid protein (40) and catalyzes the detoxification and/or
smokers. Further investigations are required to confirm this conclusion.

References
1. Toyomura K, Yamaguchi K, Kawanomoto H et al: Relation of cigarette smoking and alcohol use to colorectal adenomas by subsite: the self-defense forces health study. Cancer Sci 95: 72-76, 2004.
2. Robertson DJ, Sandler RS, Haile R, et al: Fat, fiber, meat and the risk of colorectal adenomas. Am J Gastroenterol 93: 2789-2795, 1998.
3. Sharma S, Iwasaki M, Kunieda C, et al: Development of a quantitative food frequency questionnaire for assessing food, nutrient, and heterocyclic aromatic amines intake in Japanese Brazilians for a colorectal adenoma case-control study. Int J Food Sci Nutr 60 (Suppl 7): 128-139, 2009.
4. Mansoor-ul-Haq and Faisal N: Familial adenomatous polyposis. J Coll Physicians Surg Pak 21: 46-48, 2011.
5. Wilschut JA, Hubbard JD, Ramsey SD, Boer R, Looman CW and van Baaljogoem M: Increased risk of adenomas in individuals with a family history of colorectal cancer: results of a meta-analysis. Cancer Causes Control 21: 2287-2293, 2010.
6. Zhuo W, Wang Y, Zhuo H, et al: CYP1A1 and GSTM1 polymorphisms and oral cancer risk: association studies via evidence-based meta-analyses. Cancer Invest 27: 86-95, 2009.
7. Hein DW: N-acetyltransferase 2 genetic polymorphism: effects of carcinogen and haplotype on urinary bladder cancer risk. Oncogene 25: 1649-1658, 2006.
8. Zhou W, Liu G, Thurston SW, et al: Genetic polymorphisms in N-acetyltransferase-2 and microsomal epoxide hydrolase, cumulative cigarette smoking, and lung cancer. Cancer Epidemiol Biomarkers Prev 11: 15-21, 2002.
9. Bolt HM, Selinski S, Danappel D, Blaszkewicz M and Golka K: Re-investigation of the concordance of human NAT2 phenotypes and genotypes. Arch Toxicol 79: 196-200, 2005.
10. Hein DW and Doll MA: Accuracy of various human NAT2 SNP genotyping panels to infer rapid, intermediate and slow acetylator phenotypes. Pharmacogenomics 13: 31-42, 2012.
11. Taja-Chayeb L, González-Fierro A, Miguez-Muñoz C, et al: Acetyltransferase status and N-acetyltransferase 2 gene polymorphisms: phenotype-genotype correlation with the sulfamethazine test. Pharmacogenet Genomics 21: 894-901, 2011.
12. Munafò MR, Clark TG and Flint J: Assessing publication bias in genetic association studies: evidence from a recent meta-analysis. Psychiatry Res 129: 39-44, 2004.
13. Egger M, Davey Smith G, Schneider M and Minder C: Bias in meta-analysis detected by a simple, graphical test. BMJ 315: 629-634, 1997.
14. Raimondi S, Botteri E, Iodice S, Lowenfels AB and Maisonneuve P: Gene-smoking interaction on colorectal adenoma and cancer risk: review and meta-analysis. Mutat Res 670: 6-14, 2009.
15. Potter JD, Bigler J, Fosdick L, et al: Colorectal adenomatous and hyperplastic polyps: smoking and N-acetyltransferase 2 polymorphisms. Cancer Epidemiol Biomarkers Prev 8: 69-75, 1999.
16. Moonen H, Engels L, Kleinjans J and Kok T: The CYP1A2-164A>C polymorphism (CYP1A2*1F) is associated with the risk for colorectal adenomas in humans. Cancer Lett 229: 25-31, 2005.
17. Bunschoten A, Tiemersma E, Scholjs L and Kampman E: Simultaneous determination of polymorphism in N-acetyltransferase 1 and 2 genes by reverse line blot hybridization. Anal Biochem 285: 156-162, 2000.
18. Crabtree MD, Fletcher C, Churchman M, et al: Analysis of candidate modifier loci for the severity of colonic familial adenomatous polyposis, with evidence for the importance of the N-acetyltransferases. Gut 53: 271-276, 2004.
19. Lin HJ, Probst-Hensch NM, Hughes NC, et al: Variants of N-acetyltransferase NAT1 and a case-control study of colorectal adenomas. Pharmacogenetics 8: 269-281, 1998.
20. Probst-Hensch NM, Haile RW, Ingles SA, et al: Acetylation polymorphism and prevalence of colorectal adenomas. Cancer Res 55: 2017-2020, 1995.
21. Voskuil DW, Kampman E, Grubben MJ, et al: Meat consumption and preparation, and genetic susceptibility in relation to colorectal adenomas. Cancer Lett 114: 309-311, 1997.
22. Tiersmerswa EW, Voskuil DW, Bunschoten A, et al: Risk of colorectal adenomas in relation to meat consumption, meat preparation, and genetic susceptibility in a Dutch population. Cancer Causes Control 15: 225-236, 2004.
23. Ishibe N, Sinha R, Hein DW, et al: Genetic polymorphisms in heterocyclic amine metabolism and risk of colorectal adenomas. Pharmacogenet Genomics 12: 145-150, 2002.
24. Lang NP, Butler MA, Massengill J, et al: Rapid metabolic phenotypes for acetyltransferase and cytochrome P4501A2 and putative exposure to food-borne heterocyclic amines increase the risk for colorectal cancer or polyps. Cancer Epidemiol Biomarkers Prev 3: 675-682, 1994.
25. Moslehi R, Chatterjee N, Church TR, et al: Smoking, N-acetyltransferase genes and the risk of advanced colorectal adenoma. Pharmacogenomics 7: 819-829, 2006.
26. Probst-Hensch NM, Haile RW, Li DS, et al: Lack of association between the polyadenylation polymorphism in the NAT1 (acetyltransferase 1) gene and colorectal adenomas. Carcinogenesis 17: 2125-2129, 1996.
27. Shin A, Shrubsbole MJ, Rice JM, et al: Meat intake, heterocyclic amine exposure, and metabolizing enzyme polymorphisms in relation to colorectal polyp risk. Cancer Epidemiol Biomarkers Prev 17: 320-329, 2008.
28. Tiersmerswa EW, Bunschoten A, Kok FJ, Glatt H, de Boer SY and Kampman E: Effect of SULT1A1 and NAT2 genetic polymorphism on the association between cigarette smoking and colorectal adenomas. Int J Cancer 108: 97-103, 2004.
29. Wang H, Yamamoto JF, Caberto C, et al: Genetic variation in the bioactivation pathway for polycyclic hydrocarbons and heterocyclic amines in relation to risk of colorectal neoplasia. Carcinogenesis 32: 203-209, 2011.
30. Tobias A: Assessing the influence of a single study in the meta-analysis estimate. Statat Techm Bull 47: 15-17, 1999.
31. Gong C, Hu X, Gao Y, Cao Y, Gao F and Mo Z: A meta-analysis of the NAT1 and NAT2 polymorphisms and prostate cancer: a huge review. Med Oncol 28: 365-376, 2011.
32. Ying XJ, Dong P, Shen B, Wang J, Wang S and Wang G: Possible association of NAT2 polymorphism with laryngeal cancer risk: an evidence-based meta-analysis. J Cancer Res Clin Oncol 137: 1661-1667, 2011.
33. Cui D, Wang Z, Zhao E, Ma J and Lu W: NAT2 polymorphism and lung cancer risk: a meta-analysis. Lung Cancer 73: 153-157, 2011.
34. Zhong X, Hui C, Xiao-Ling W, Yan L and Na L: NAT2 polymorphism and gastric cancer susceptibility: a meta-analysis. Arch Med Res 41: 275-280, 2010.
35. Ye Z and Parry JM: Meta-analysis of 20 case-control studies on the N-acetyltransferase 2 acetylation status and colorectal cancer risk. Med Sci Monit 8: CR558-CR565, 2002.
36. Bolrajk and Reamom-Buttner SM: N-acetyltransferase 2 (NAT2) gene polymorphisms in colon and lung cancer patients. BMC Med Genet 7: 58, 2006.
37. Zhang LQ, Zhou JN, Wang J, et al: Association between N-acetyltransferase 2 acetylator status and colorectal cancer susceptibility: based on evidence from 40 studies. PLoS One 7: e32425, 2012.
38. Botteri E, Iodice S, Raimondi S, Maisonneuve P and Lowenfels AB: Cigarette smoking and adenomatous polyps: a meta-analysis. Gastroenterology 134: 388-395, 2008.
39. Zhou JY, Shi R, Yu HL, Zheng WL and Ma WL: Association of NQO1 Pro187Ser polymorphism with the risks for colorectal cancer: a meta-analysis. Gastroenterology 134: 388-395, 2008.
40. Taja-Chayeb L, González-Muñoz C, et al: Acetyltransferase status and N-acetyltransferase 2 gene polymorphisms: phenotype-genotype correlation with the sulfamethazine test. Pharmacogenet Genomics 21: 894-901, 2011.
41. Tiersmerswa EW and Voskuil DW: Effect of SULT1A1 and NAT2 genetic polymorphisms in colon and lung cancer patients. Int J Cancer 60 (Suppl 7): 128-139, 2009.