A Confirmed Catheter-Related Blood Stream Infection (CRBSI) in an Immunocompetent Patient Due to *Myroides odoratimimus*: Case Report and Literature Review

**Abstract:** The genus *Myroides* are gram-negative bacilli which are completely aerobic, non-motile, non-fermenting and yellow-pigmented with a characteristic fruity odor. *Myroides* species are widely found in the environment, especially in water and soil, and are considered as low-grade opportunistic pathogens for humans. *Myroides* infections are most commonly seen in immunocompromised patients and only rarely occur in immunocompetent patients. We here report the first confirmed catheter-related bloodstream infection (CRBSI) due to *Myroides odoratimimus* in an immunocompetent patient. We also review the literature related to *Myroides* infections.

**Keywords:** *Myroides odoratimimus*, CRBSI, immunocompetent patient

**Introduction**

*Myroides* species originally classified as *Flavobacterium odoratum*, they were renamed *Myroides* in 1996 because of the difference for genotypic and phenotypic data such as nonmotile, good growth at 37°C, halotolerance, and difference in the fatty acid profile. The two best-known species are *Myroides odoratus* and *Myroides odoratimimus*. They are commonly found in the environment, rarely infect humans, but the incidence of *Myroides* infections has increased in recent years. These infections usually occur in immunocompromised patients, including those with diabetes mellitus, cirrhosis, chronic obstructive pulmonary disease, renal failure, or prolonged corticosteroid therapy, with soft tissue infections and urinary tract infections being the most common types. However, infections in immunocompetent hosts have also been described and are occasionally life-threatening. We here report a confirmed CRBSI in an immunocompetent patient due to *Myroides odoratimimus*. Clinicians must be alert to the possibility that this genus can be highly pathogenic even in immunocompetent hosts. Written informed consent was provided by the patient to allow the case details to be published, and our study was approved by the Ethics Committee at Jiangsu Province Hospital.

**Case Report**

A 48-year-old female presented to our emergency department 2 days after cardiopulmonary resuscitation (CPR). Two days before, the patient suffered a sudden loss

**Correspondence:** Fang Ni; Yaning Mei
Department of Laboratory Medicine, Jiangsu Province Hospital, Guangzhou Street No. 300, Nanjing 210029, People’s Republic of China
Tel +86 25-6830-6287
Fax +86 25- 8372-4440
Email 13813972378@163.com; myn303@163.com
of consciousness, with a pale face, salivation at the corners for the mouth, and urinary incontinence, but without nausea, vomiting, or limb twitching. She was immediately admitted to the local hospital resuscitation room. At that time, her blood pressure was as high as 160/103 mmHg while her heart rate dropped to 12 beats per minute, and the electrocardiogram showed third-degree atrioventricular block, serum potassium was only 2.59 mmol/L. There was no significant past medical history other than diarrhea before her loss of consciousness. The patient immediately underwent a series of rescue measures and a temporary pacemaker was implanted to adjust the heart at 60 beats per minute due to her recurrent cardiac and respiratory arrest. Additionally, a chest CT scan revealed exudative lesions in the lungs bilaterally while a head CT scan showed no abnormality. Shortly thereafter, she was transferred to the intensive care unit (ICU) with stable vital signs.

The patient was subsequently admitted to our hospital for further treatment. Upon clinical examination, her temperature was 39.5°C; blood pressure, 92/45 mmHg; heart rate, 60 beats/minute; and respiratory rate, 23 breaths per minute. Physical examination revealed a placed trachea intubation with external mechanically ventilating, an appendectomy incision at the right McBurney’s point, a temporary cardiac pacing catheter at the ipsilateral groin, and a left femoral vein catheterization in place. Laboratory values included a white blood count of 36,610/µL (normal, 4000–10,000) with 88% neutrophils, procalcitonin 4.31 ng/mL (normal, 0–0.05), pro-B type natriuretic peptide 6072 pg/mL (normal, 0–125), hypersensitive troponin T (hs-TropT) 115.5 ng/L (normal, 0–14), ALT 331.5 U/L (normal, 7.0–40.0), and AST 272 U/L (normal, 13.0–35.0). The initial diagnoses of cardiac respiratory arrest, post-CPR syndrome, and third-degree atrioventricular block were put forward and broad empirical therapy was considered to be a possible infection source and therefore removed; the catheter, catheter blood and venous blood cultures were sent for microbiological examination in view of the deteriorating clinical picture.

Three days after hospitalization, the temperature of the patient rose as high as 40.5°C and inflammation indexes were significantly elevated. The left femoral vein catheter was considered to be a possible infection source and therefore removed; the catheter, catheter blood and venous blood cultures were sent for microbiological examination in view of the deteriorating clinical picture.

After 14 h of culture, gram-negative rods were detected from both aerobic blood culture bottles (BACTEC FX, BD Becton, Dickinson and Company) and colistin was applied. Strains were isolated from blood, chocolate, and MacConkey Agar after 24 h of incubation in aerobic conditions, which were round, yellow pigmented, non-fermentative, and had a fruity odor. Initial identification of Myroidesspp. was performed using a VITEK2 (BioMerieux) automated system and subsequently confirmed by matrix-assisted laser desorption ionization-time-of-flight mass spectrometry (MALDI-TOF-MS) (BioMerieux) with 99% confidence. Definitive identification was accomplished via 16SrRNA gene sequencing, revealing a 1474-score homology with Myroides odoratimimus. The same organism was isolated from both the left femoral vein catheter and catheter blood, and the positive time of catheter blood culture was earlier than venous blood. Accordingly, a diagnosis of catheter-related bloodstream infection (CRBSI) was confirmed. In the meantime, the patient developed septic shock.

Antimicrobial susceptibility tests (AST) were carried out by automated microdilution broth test (VITEK2, BioMerieux) and interpreted following CLSI M100 guidelines (2019), using the minimum inhibitory concentration (MIC) breakpoints of other non-Enterobacteriaceae and cefoperazone/sulbactam referred to the breakpoint of cefoperazone. The in vitro susceptibility testing of the isolate proved it resistant to the majority of agents tested (i.e., piperacillin, piperacillin/tazobactam, ceftazidime, ceftriaxone, cefepime, aztreonam, imipenem, meropenem, amikacin, gentamicin, tobramycin, ciprofloxacin, levofloxacin, and trimethoprimsulfamethoxazole), with susceptibility solely to cefoperazone/sulbactam (Table 1). Based on that finding, the patient’s antibiotic treatment was changed to a combination of cefoperazone/sulbactam (1 g IV qd) and levofloxacin (3 g IV q6h).

After 5 days on the regimen, the infection of CRBSI was under control and no causative organism could be detected in blood culture. The patient had cardiac respiratory arrest again during the time of hospitalization, and a temporary pacemaker was reinstalled. Coronary angiography (CAG) showed the patient had a coronary myocardial bridge, for which a permanency cardiac pacemaker should be considered when the patient stabilizes. Afterwards, the patient was transferred to local hospital for rehabilitation.

Discussion
CRBSI have been increased in recent years due to the widespread use of invasive operations. The most common pathogenic bacteria reported to cause CRBSI were coagulase-negative Staphylococcus (especially Staphylococcus epidermidis), followed by Staphylococcus aureus, Candida spp. and Enterococcus.4 CRBSI with Myroidesspp. were rarely described. Douce et al reported an outbreak of central venous CRBSI due to contaminated ampoules of water for injection.5
There were another four CRBSI cases reported in cancer patients. A case involving a baboon after a porcine kidney graft had a CRBSI because of this genus. Among the cases published, our case is the first confirmed CRBSI in immunocompetent patients. Clinical attention should be paid to this emerging pathogen causing CRBSI.

*Myroides* species have been cultured from blood, urine, soft tissue, bronchoalveolar lavage, sputum, bile, pus and pericardial fluid, although rarely isolated from clinical specimens. They have also been found in seafood products, meat-processing, flesh flies, grey mullet’s gut, livestock manure, Sagar catfish, and boar semen. Bacteria existing in these non-human hosts may have some connection with the transmission of the pathogen. *Myroides* infections include cellulitis, urinary tract infection (UTI), bacteremia, necrotizing fasciitis, pneumonia, pericarditis endocarditis, ventriculitis, acalculous cholecystitis, urosepsis, empyema and canaliculitis as well as nosocomial outbreaks. *Myroides odoratus* and *Myroides odoratimimus* are the two main species of the genus to cause corresponding infections. *Myroides odoratus* strains are associated with septic soft tissue infections, whereas *Myroides odoratimimus* is described in cases of urinary infections, but not absolutely. Patients infected with *Myroides* are generally cured or improved, but occasional life-threatening cases have been reported.

Besides the most two common species mentioned above, a series of new species have been identified with the development of molecular techniques and are detailed in Table 2. These uncommon species apparently do well in both aquatic and soil environments, which are mainly isolated from seawater and soil, and have not to date been documented as a source of infections in humans except for the species of *Myroides injenensis* which caused bacteremia and severe cellulitis.

*Myroidesspp.* is not traditionally normal human flora and they are usually considered to be low-grade opportunist pathogens, *Myroidesspp.* infection is strongly associated with immunocompromised patients. There are some infection cases reported in these hosts. Those infected with the organisms often have underlying diseases such as diabetes mellitus, cirrhosis, chronic obstructive pulmonary disease, renal failure, chemotherapy, immunosuppression for transplantation and prolonged corticosteroid therapy, which may damage the body’s immunity and make human susceptible to *Myroides* infections. However, immunocompetent hosts can also be infected although rarely reported. We found only six isolated cases of *Myroidesspp.* infection in immunocompetent hosts (including our case), summarized in Table 3. Increasing cases in immunocompetent hosts have been published in recent 10 years, four cases were caused by *Myroides odoratimimus*, four

### Table 1 In vitro Susceptibility of the *Myroides odoratimimus* Isolate

| Antibiotic             | MIC Value (µg/mL) | Interpretation (S, I, R) |
|------------------------|-------------------|--------------------------|
| Piperacillin           | ≥ 128             | R                        |
| Piperacillin/tazobactam| ≥ 128             | R                        |
| Ceftriaxone            | ≥ 64              | R                        |
| Cefepime               | ≥ 64              | R                        |
| Aztreonam              | ≥ 64              | R                        |
| Imipenem               | ≥ 16              | R                        |
| Meropenem              | ≥ 16              | R                        |
| Amikacin               | ≥ 64              | R                        |
| Gentamicin             | ≥ 16              | R                        |
| Tobramycin             | ≥ 16              | R                        |
| Ciprofloxacin          | ≥ 4               | R                        |
| Levofloxacin           | ≥ 8               | R                        |
| Trimethoprim/sulfamethoxazole | ≥ 320  | R                        |
| Cefoperazone/sulfactam | 4.0              | S                        |

**Abbreviations**: MIC, minimum inhibitory concentration; S, susceptible; I, intermediate; R, resistant.

### Table 2 New Species of the *Myroides*

| Year | New Species                  | Isolated from                  | Country | Reference |
|------|------------------------------|--------------------------------|---------|-----------|
| 2006 | *Myroides pelagicus* sp. nov | Seawater                       | Japan   | [26]      |
| 2008 | *Myroides profundi* sp. nov  | Deep-sea sediment              | China   | [29]      |
| 2011 | *Myroides marinus* sp. nov   | Seawater                       | Korea   | [21]      |
| 2012 | *Myroides phaeus* sp. nov    | Human saliva                    | China   | [22]      |
| 2012 | *Myroides injenensis* M09-0166 T | Prehistoric paintings         | Korea   | [27]      |
| 2013 | *Myroides guanonis* sp. nov  | Forest soil                    | Bulgaria| [24]      |
| 2014 | *Myroides Xuanwuensis* sp. nov | Garden soil                  | China   | [23]      |
| 2015 | *Myroides indicus* sp. nov   | Radiation-polluted soils       | India   | [25]      |
| 2017 | *Myroides* sp. N17-2         |                                | China   | [28]      |
A 69-year-old man who showed no evidence of immunity had a portal of entry from the bloodstream. Licker et al described an outbreak of four UTIs caused by Myroides injenensis because of his occupational exposure as a plumber and contact with sewage as well as his alcoholic cirrhosis. Another case described a Myroides odoratimimus bloodstream infection in a patient with a chronic diabetic foot ulcer who was exposed to a presumably contaminated water source. A 69-year-old man who showed no evidence of immunocompromise frequently swam in a freshwater river and suffered a right lower extremity cellulitis, fever, chills and sepsis because of Myroides odoratus, the portal of entry was probably the small abrasion that became contaminated with water during swimming. Some other reports have been described infections in the same way. Open wounds with exposure to water can be considered as a risk factor to cause rapidly invasive Myroides infections, regardless of the immunity of patients. Maraki et al reported a healthy child developed cellulitis after a pig bite, while Willems et al describe a case of fulminant erysipelas with sepsis following a scratch of the domestic dog in a corticosteroid-treated host, the two cases above indicated that animal attacks on people may be another risk factor to cause the infections of Myroides spp. Licker et al described an outbreak of four UTIs caused by Myroides odoratimimus, all patients were immunocompromised and three of them underwent urinary catheterization with a Foley’s catheter upon admission. Lorenzin et al also reported a Myroides odoratimimus UTI in an immunocompromised patient who had a permanent urinary Foley’s catheter due to urinary retention. There are also some CRBSI cases described. Carrying catheters in immunocompromised patients may be important for Myroidesspp. to cause infections. In our case, the patient who had no history of disease was infected with CRBSI due to a left femoral vein catheterization, this should be observed.

Moreover, some works of literature substantiated Myroidesspp. have the capacity of co-aggregation and self-aggregation to form biofilm using crystal violet binding assay and the bacterial surface is extremely hydrophobic because of a polysaccharide capsule possessed. Biofilms can adhere to both biotic and abiotic surfaces like medical devices and the bacteria can be entrapped within a self-produced extracellular polymeric matrix, which increases pathogenicity in device-related infections and is associated with conventional therapeutic failure, as well as recurrent infections. Some virulence factors (VFs) of the genus Myroides were identified in several works of literature. Hu et al found some similar virulence factors in the genomes of three clinically pathogenic and three environmental Myroides. odoratimimus strains with the help of VFDB protein Set B database, the VFs included capsule/capsular polysaccharide, intracellular survival and invasion factors, molecular chaperone, urease, acinetobactin, Streptococal endolase, heme biosynthesis, acyl carrier protein, and T4SS effectors.
The treatment of Myroides spp. infections is difficult because the genus is often multi-drug-resistant.\textsuperscript{30} Many strains have been recognized as resistant to beta-lactams, monobactams and carbapenems due to the production of chromosome-encoded metallo-beta-lactamase (MUS-1 and TUS-1).\textsuperscript{36} Myroides odoratimimus is intrinsically resistant to colistin.\textsuperscript{36} In addition, a new subclass B1 metallo-beta-lactamase gene \textit{bla}_{MOC} was found which also conferred resistance to most beta-lactams except for aztreonam and cefepime.\textsuperscript{38} There was another report described MUS-2, a novel variant of the chromosome-encoded beta-lactamase MUS-1, reducing the susceptibility to beta-lactams.\textsuperscript{39} Some strains have shown a susceptibility to beta-lactam inhibitor compounds,\textsuperscript{30,33,35} and the isolate in our report is sensitive to cefoperazone/sulbactam which is a kind of beta-lactamase inhibitor. Perhaps beta-lactamase inhibitor compounds can become an option for treating such infections. Myroides spp. has variable susceptibility to aminoglycosides, quinolones, trimethoprim/sulfamethoxazole\textsuperscript{31} and tetracyclines. Our patient’s isolate is resistant to all drugs except for cefoperazone/sulbactam. She ultimately responded well to treatment with cefoperazone/sulbactam combining levofloxacin. Given no clear guidelines for the treatment of this genus and the variability in susceptibility testing of different Myroides species, antimicrobial treatment should be based on drug sensitivity results.

**Conclusion**

In conclusion, with the increasing invasive therapy in clinical, clinicians should be alert to the possibility of Myroides spp. becoming a new highly pathogenic pathogen in immunocompetent hosts. Identification and antimicrobial susceptibility test should be done in time to guide a reasonable clinical medicine application.

**Disclosure**

The authors report no conflicts of interest in this work.

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