Review Article

Advances in Magnetic Nanoparticle-Driven Delivery of Gene Therapies towards Prostate Cancer

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Conventional treatment possibilities for one of the most common diseases among men, i.e., prostate cancer has several side effects. Gene-based therapy such as siRNA, CRISPR/Cas9, pDNA, and miRNA have emerged as an alternative, combating posttherapy side effects and drug resistance. Magnetic nanoparticles have been appropriately modified and functionalized to efficiently deliver the gene therapy-based active compounds to prostate tumor cells. The main purpose of this review article is to highlight the strategies currently being utilized for the treatment of prostate cancer using magnetic nanoparticles for delivery of genetic material using both the passive and active (ligand-based) targeting. It further discusses the challenges in efficient delivery of therapeutics to tumor sites and their remedial approaches. Finally, it provides a glimpse of future advances for tumor-specific modifications of magnetic nanoparticles to combat prostate cancer.

1. Introduction

Other than lung cancer, prostate cancer (PCa) is the most common cancer among men, especially in the developed world [1, 2]. It is the second most prevalent cause of mortality among males in the USA [2]. Since the disease is caused by accumulation of mutations in normal prostate cells and conferring them malignant phenotype, treatment strategies depend on the stage of PCa in the body. The conventional strategies for the treatment are surgery, chemotherapy, radiation therapy, and hormone therapy [3]. If the cancer is confined, chemotherapy can be utilized for treatment [4, 5]. However, the lack of specificity, systemic toxicity, and development of drug resistance in tumor cells limits the uses of chemotherapy [6]. Radical prostatectomy (RP) and radiation therapy (RT) are another choices for treatment [7] especially, for the most common localized prostate cancer [8]. But these types of treatment can be relatively unsafe, unreliable, cause sexual and urinary dysfunctions, and may induce drug resistance [9].

However, the first-line therapy for improving the survival and reducing the morbidity in primary metastatic prostate cancer is the androgen-deprivation therapy (ADT) also known as called hormonal therapy [10, 11]. It is the primary method of treatment for metastatic prostate and the relapsed cancer [3]. Its aim is to control the cancer by lowering the levels of blood testosterone on which the tumor cells are dependent for survival and are achieved by using antiandrogens, luteinizing hormone-releasing hormone (LHRH) antagonists and LHRH agonists [12, 13]. However, these treatment measures are not specific for PCa and cause side effects which can be serious with chances of recurrence of
more aggressive androgen-independent form called castrate-resistant prostate cancer (CRPC) [14, 15]. Treatment for CRPC after ADT is very difficult because of the slow decline in treatment efficacy [2].

Therefore, eradication of prostate cancer has especially been challenging due to its highly heterogeneous and specific nature. To improve the efficiency of conventional drugs, nanocarriers and nanoparticles have been developed for delivery purposes [16]. Recently, a large number of antiprostate cancer drug delivery approaches have been proposed [17]. For instance, chemotherapeutic prostate cancer drugs such as paclitaxel and docetaxel can be delivered in high doses to specifically target prostate tumor, avoiding many side effects on normal cells [11].

However, unlike the conventional therapeutic delivery methods, gene delivery using nucleic acids has many advantages such as being usable in nondruggable targets [15, 18]. Moreover, nucleic acids are highly specific and act rapidly with higher potency [19]. Knockdown or activation of different types of oncogenes and tumor suppressor genes has shown potential results in prostate cancer treatment. Magnetic nanoparticles can also function as MRI contrast agent for imaging the solid tumors such as prostate tumor [20]. With so many different roles from being delivery vehicles to MRI contrast agent, this review specifically highlights the potential applications of magnetic nanoparticles in prostate cancer gene therapy. It also discusses the techniques used for targeted delivery and sheds light on various obstacles for gene delivery along with their possible mitigation approaches.

2. Magnetic Nanoparticles for Gene Therapy

The great potential benefit of gene therapy lies in the disease treatment of genetic disorders, viral infections, and cancer. Recently, different types of magnetic nanoparticles have been developed for cancer diagnosis and treatment [21]. Their dimension makes them an ideal candidate for surface functionalization. These modifications form the basis of highly diverse pharmaceutical and biomedical applications, such as diagnosis, drug delivery, gene delivery, and enzyme delivery [22]. Magnetic nanoparticles have received considerable attention, in order to develop an efficient delivery system to carry in vivo drugs and other active compounds. Since magnetic nanoparticles can be catered to below 100 nm size, their special characteristics like higher surface-volume ratio, large surface energy, and high reactivity permits them to have potential applications in biomedical fields [23]. Despite all these benefits, very few magnetic nanoparticles have been used in clinical practice partly because of limited research in this field. However, some magnetic nanoparticles are currently in early clinical trials for medical imaging purpose (Table 1).

Both the viral-based and nonviral-based delivery vehicles have been employed for gene therapy treatment of prostate cancer. The most common viral vectors currently being used include adenovirus, adeno-associated virus, pox virus, retrovirus, herpes simplex virus, and lentivirus, whereas the nonviral-based vectors mostly used are polymer-based nanoparticles or nanocarriers [24–26]. These vectors can be easily modified to develop different types of therapeutic systems, and their efficiency and safety are also being evaluated in clinical trials.

3. Delivery Systems

Iron oxide nanoparticles have been better suited as MRI agents as compared to gadolinium-based MRI agents based on their lower toxicity, lower detection limit, and stronger proton relaxation features [27]. There are two types by which magnetic nanoparticles can be targeted to the tumor site, namely, active and passive targeting. In passive targeting, the magnetic nanoparticles are accumulated to the required tumor site by enhanced permeability and retention (EPR) effect caused by leaky blood vasculature and dysfunctional lymph system [22]. The accumulation highly depends on the size of magnetic nanoparticles, pore size of leaky blood vessels, and the half-life of nanoparticles in blood circulation. For active targeting, magnetic nanoparticles are modified with some specific targeting molecules such as ligands, antibodies, and peptides depending on the type of tumor and its target tissue expressing a specific surface receptor.

There are various methods of delivery; here, we discuss about targeted delivery using external magnet and specific ligands for prostate cancer.

3.1. Targeted Delivery Using External Magnet. Magnetic nanoparticles can be externally controlled to guide the nanoparticles to specific solid tumor/tissues and used for imaging the tumor. This feature is more restricted to diagnosis and imaging of tumor in specific tissue (Figure 1). For this purpose, a specific concentration of Fe in MNPs is injected into the body, and then, after guiding the nanoparticles to a specific tumor site using external magnet, the tumor is imaged for which the contrast is provided by the iron oxide nanoparticles. For example, ferumoxtran-10, a dextran-coated MNP which has a mean size of about 30 nm, is in phase III clinical trials for diagnosis of prostate cancer [28]. Its sensitivity and specificity are 90.5% and 97.8%, respectively, for detecting the prostate cancer lymph node disease. It functions by passively accumulating in the metastatic lymph nodes of prostate cancer.

Similarly, another application of magnetic nanoparticles includes the guided photothermal therapy of tumor itself. External magnets can be used to guide the magnetic nanoparticles to their specific location, and then, the drug release can be controlled photothermally using a near infrared or infrared light generating heat in the nanoparticles and releasing the drug bound to them [29]. These externally triggered drug release could accumulate in the tumor site due to the EPR effect. One study revealed the optimum size of Fe3O4 nanoparticles to be 220 nm resulting in higher temperature and better therapeutic effect [21]. In another study, magnetic carbon nanoparticles were used for photothermal delivery of dyes and plasmids [30]. The irradiated nanoparticles with NIR laser beam causes rise in temperature allowing for internalization inside the cells. An improved gene
delivery system was developed using the photothermal delivery composed of liposomal carbon dots nanohybrids (PEG-Fe/CDs) and showed about 16°C temperature rise after 5 min of irradiation of the tumor tissue [31].

3.2. Nonmagnetic (Ligand-Based) Targeting. More specific than the passive targeting, active targeting involves the modification of nanoparticles with some specific ligand molecules such as folic acid, sugars, RGD peptides, antibodies, and hyaluronic acid which then bind to their specific receptor molecule in the tumor cell (Table 2) [32, 33]. Highly overexpressed receptor targeting makes this method more specific towards one specific type of cancer. For example, HSP-70-linked magnetic nanoparticles are capable of attaching to CD40 receptors which is expressed on glioma cells [34].

For diagnosis of prostate cancer, one of the most common targeting molecules is prostate-specific membrane antigen (PSMA) which can be targeted via antibody. PSMA is an ideal target for prostate cancer, specifically produced and expressed in prostate cancer [44]. In one study, magnetic nanoparticles were conjugated to PSMA-targeting antibody, J591, for targeting prostate cancer [43]. The J591-bound magnetic NPs enhanced the cellular iron uptake. MRI of tumors was enhanced in vivo when using PSMA-targeting NPs as compared to nontargeted NPs. Therefore, they provided a proof that PSMA-targeting can be a potential mode for enhancing the MRI of prostate cancer.

4. Delivery of Different Gene Therapy by MNPs

Magnetic nanoparticles after functionalization with nucleic acid carrier can be used for delivering different types of nucleic acids (Figure 2). Here, we discuss the recent strategies on how these nanoparticles can be utilized in various applications and strategies.
4.1. siRNA-Based Gene Therapy of Prostate Cancer. In recent years, different strategies have been developed for treatment of prostate cancer using siRNA-based gene therapy [45, 46]. siRNA against different genes have been utilized for gene silencing purpose to inhibit cellular function [47]. For instance, Wang and coworkers, modified magnetic nanoparticles with PEI to develop a multifunctional nanocarrier for both MRI as well as in vitro gene therapy of cancer cells [48]. They constructed a surviving siRNA-PEI-MNP delivery system which efficiently inhibited cellular proliferation and induced apoptosis and autophagy of glioblastoma multiforme (U251) cells. The major function of cationic polymer such as PEI used in this study is to effectively absorb siRNA via electrostatic interaction and thereby protecting from enzymatic degradation in vitro. In another study, glycol chitosan-based nanoparticles also acted as an efficient delivery vehicle for encapsulating siRNA along with a chemotherapeutic drug to accomplish maximum efficiency for overcoming the drug resistance [49]. In this study, Bcl-2 siRNA-loaded and DOX-loaded glycol chitosan NPs both

| Table 2: Different types of magnetic nanoparticles and their application in treatment and diagnosis of cancer. |
|---------------------------------------------------------------|
| Type of MNP          | Drug/gene         | Target gene                  | Route   | Mouse model                                      | Application                                      | Reference |
|----------------------|-------------------|------------------------------|---------|-------------------------------------------------|-------------------------------------------------|----------|
| PEG-Ga<sup>68</sup>-Lys-urea-Glu | DM1               | PSMA                         | i.v.    | SCID mice with PC3-PIP and PC3-flu tumors       | PET/CT scan and therapy                         | [35]     |
| MNP-PEI-PEG          | —                 | Folate receptor              | i.v.    | Balb/c nude mice                                | MRI                                             | [36]     |
| MNP-PCL              | Gemcitabine       | —                            | i.v.    | Xenograft mice                                  | Pancreatic cancer treatment                     | [37]     |
| MNP-PEI-PEI          | —                 | —                            | i.v.    | Xenograft mice                                  | MRI                                             | [38]     |
| MNP-PEI-PEG          | PSCA (prostate stem cell antigen) | —        | —       | —                                               | Prostate cancer targeting in vitro               | [39]     |
| MNP-citrate          | —                 | —                            | i.v.    | Hepatic fibrosis Wistar rats                     | MRI of liver                                    | [40]     |
| MNP-soybean oil      | —                 | —                            | Nasal   | CD-1 female mice                                | MRI of nasal septum                             | [41]     |
| MNP-Au-HA            | —                 | CD44                         | i.v., i.t. | Balb/c nude mice                                      | MRI, CT and photothermal therapy               | [42]     |
| MNP-CD-J591          | Docetaxel         | PSMA (prostate specific membrane antigen) | i.v.    | C4-2 and PC3 tumor mice                          | Prostate cancer treatment                       | [43]     |

MNP: magnetic nanoparticles; i.v.: intravenous; i.t.: intratumoral; PSCA: prostate stem cell antigen; PSMA: prostate-specific membrane antigen.

**Figure 2:** Different types of gene therapy systems that are being investigated for delivery using magnetic nanoparticles. A simplified overview of the various mechanisms with which these systems act upon internalization inside the cells. All the systems act in the cytosol except for CRISPR/Cas9 system which localizes in nucleus.
showed quite similar physiochemical features such as size, surface properties, and pH sensitive behavior. Both nanodelivery vehicles showed similar in vivo biodistribution and pharmacokinetics and enhanced treatment in a dose-dependent manner.

Instead of delivering the siRNA directly to cells, attaching a specific receptor can specifically target tumor cells for silencing of target genes. For example, folate receptor (FR) is a target specifically overexpressed in tumor cells. In a study conducted by Biswal and coworkers, dihydrofolate reductase (DHFR) siRNA expressing-plasmid was complexed with folate-polyethylene glycol-polyethylenimine (PEI-PEG-Fol) and delivered into FR positive human epidermal carcinoma (KB) cells as well as FR negative A549 cells [50]. It was shown that siRNA was delivered inside the KB cells where it inhibited the DHFR gene expression, whereas A549 cells were unable to uptake the siRNA-nanocomplex. They concluded that DHFR siRNA complexed with PEG-PEI-Fol can be specifically taken up by FR positive cells which makes them highly specific delivery vehicles. This strategy is designed for particular cancer types; however, it can be extended to various tissue treatment deliveries.

There are still some bottlenecks facing nonviral delivery of siRNA which hinder the successful application in vivo [51]. In one study, PEI-based siRNA delivery system was developed to protect the siRNA from nucleolytic enzyme degradation while delivering to specific tissue [52, 53]. They modified PEI by noncovalently attaching phospholipid liposomes which decreased the cytotoxicity and at the same time increased the delivery efficiency of siRNA.

Functionalization of low molecular weight PEI with tyrosine can also improve the biocompatibility, stability, and silencing efficacy of modified nanoparticles [54]. If ligand-mediated cellular uptake is required, PEI can be conjugated with PEG to shield surface charges of PEI, prior to coupling a ligand such as anti-HER1 antibody for targeted delivery [52, 55]. The analyses are also extended beyond tumor cell lines, involving tumor xenograft tissues which reflect more convincingly the in vivo experiment. This is evident by the siRNA-mediated silencing of target genes, i.e., surviving and HER2 in intact tissue and stroma revealing nanoparticle penetration and cellular efficacy. Liposomes containing iron oxide nanoparticles can also work as a heating element for temperature-triggered drug release [56]. In one study, 1,2-dipalmitoyl-sn-glycero-3-phosphocholine (DPPC) and cholesterol-containing thermosensitive magnetic carrier was developed for codelivery of drug doxorubicin as well as shRNA (to target special AT-rich binding protein (SATB1) into gastric tumor cells [57]. SATB1, also known as genome organizer, is involved in regulation of malignant biological behavior of many cancers, and its knockdown can lead to reduced cell proliferation, metastasis, and induction of apoptosis. DPPC is thermosensitive which causes liposomes to change from gel to liquid phase at higher temperatures (41°C), thereby increasing permeability for drug release. This study showed that there was higher tumor inhibition when DOX and SATB1 were codelivered with magnetic targeting to the tumor site than other single formulations.

4.2. CRISPR/Cas9-Based Gene Therapy of Prostate Cancer. Clustered regularly interspaced short palindromic repeats (CRISPR) was identified for the first time in bacteria which encode CRISPR RNAs to guide CRISPR-associated (Cas) nucleases by invading genetic material and act as adaptive immune response system [58–60]. Later, type II CRISPR/Cas9 system of S. pyogens which is similar to eukaryotic CRISPR system, was also discovered and mostly chosen for different applications in gene expression regulation due to its highly efficient DNA-targeting system [60]. To locate the site-specific DNA sequence, gRNA was used by Cas9 to form base pairs and then introduced as a double strand break [61]. The first 20 nucleotides of gRNA can be altered to direct Cas9 to specific site providing unique flexibility and accessibility for DNA editing [60]. The most common method, i.e., the direct delivery of Cas9 protein along with sgRNA, is the easiest method of genome editing since there is no requirement for transcription or translation [62]. Because of larger size of Cas9 protein (160 kD) along with its positive charge and the negative charge of sgRNA, there is major problem associated with the cellular delivery of the CRISPR/Cas9 complex. To alleviate Cas9 protein, an alternative use of Cas9 mRNA with sgRNA was proposed which could be easily obtained by in vitro transcription and by-passing nuclear transcription. Moreover, Cas9 mRNA rapidly initiates genome editing.

Many researches have been using CRISPR/Cas9 system for inhibition of some targeted gene to develop new therapeutics for different diseases [63, 64]. But the major problem is the efficient and safe delivery of CRISPR/Cas9 specifically to targeted cells [62]. To solve this limitation, in one study, a flexible aptamer-liposome CRISPR/Cas9 chimera was developed to effectively combine the drug delivery and targeting [58]. This new chimera contained an RNA aptamer which specifically targets PC3 cells. The aptamers were then linked to cationic liposomes by postinjection method and were utilized to transport a specific survival gene, polo-like kinase 1-based CRISPR-Cas9 in tumor cells. Their results showed a significant gene silencing activity in vitro with highly specific binding of aptamer-liposome complexes. This method describes a feasible way to deliver cell-type-specific CRISPR/Cas9 which will form the basis of therapeutic application in the long term of CRISPR/Cas9-based treatment.

4.3. pDNA-Based Gene Therapy of Prostate Cancer. Several researches have focused on delivery of plasmid DNA (pDNA) into the target tumor cells including prostate cancer [65]. In one interesting study, magnetic nanoparticles have been modified by low molecular weight linear PEI (Mw 423 and 1800) to develop an efficient and novel DNA delivery vehicle as well as transfection agent [66]. This study found that a very low N/P ratio of 1 was efficient to increase the binding capacity of DNA when compared with PEI only. Moreover, these nanoparticles could be easily controlled and driven by an external magnetic force to specific target site. Because of low N/P ratio, very low amounts of PEI were required for DNA delivery and transfection purpose, thereby reducing the cytotoxicity of cells relative to higher PEI concentrations. This new method allowed smaller amounts of
PEI to be effectively distributed across the surface of magnetic nanoparticles.

Another study revealed that APTES-modified silica nanoparticles can easily deliver a relatively higher ratio of nanoparticle to DNA cargo [67]. These nanoparticles have silane end which binds both, hydroxyl groups to iron oxide surface and the negatively charged DNA particles to amino groups of the molecule [68].

Similarly, APTES-modified magnetic nanoparticles have been studied for various purposes such as medical imaging and DNA delivery as well [69, 70]. In one such study, APTES-MNPs were modified with cationic polymers to develop a combo gene delivery vehicle [71]. The addition of cationic polymer increased the DNA binding capacity, improved the gene transfection ability, and also protected the DNA from degradation. With greater biocompatibility, lower cytotoxicity, and higher DNA loading capacity, the prepared delivery system can be largely utilized for wide applications especially in gene therapy.

4.4. miRNA-Based Gene Therapy. miRNAs are small, non-coding, single-stranded RNA molecules, endogenously expressed in cells which regulate the gene expression [72–74]. miRNAs regulate the gene expression by first binding to the 3′-UTR region and inhibiting the mRNA translation at posttranscriptional level [75]. miRNA recognizes the target via sequence complementarity of 2-8 nucleotides at 5′-end of miRNA and the 3′ UTR region of specific mRNA [76, 77].

Recently, many studies have been carried out to understand the role of miRNA in gene therapy of prostate cancer. In one report, the polyarginine peptide (R11) was labeled to PEI nanoparticles for the miR-145 delivery to prostate cancer in vivo [72]. The advantage of this combination was the formation of an efficient transfection agent which could be taken up by prostate cancer. Moreover, the prepared R11-PEI nanocarrier displayed very low cell toxicity. Following R11-PEI/miR-145 nanocomplex delivery, a significant inhibition of tumor growth leading to prolonged survival period was reported.

4.5. Aptamer-Based Gene Therapy. Alternative to nonprotein-based antibodies, nucleic acid aptamers can also be potentially used as targeting molecules for targeted delivery of chemotherapeutic agents. Because of their nucleic acid composition, nucleic acid-based aptamers possess some unique properties such as higher specificity, small size, easy in vitro selection, lower immunogenicity, and higher sensitivity [78–80]. As such, Hicke and Stephens have introduced the concept of “escort aptamers” as a new horizon for aptamer function [81]. Usually, aptamers are designed to target PSMA receptor in prostate cancer cells and attached to nanoparticles for specific delivery [58].

In one study, A10 RNA aptamer, a ligand for PSMA receptor in prostate cancer cells, was attached to magnetic nanoparticles for targeted therapy as well as imaging [82]. In the study, A10 aptamer was used to intercalate the doxorubicin drug into the GC sequence of aptamer. The same researchers developed a multifunctional, SPION-TCL-A10 aptamer nanocarriers which could be used for MRI imaging of PCa detection and targeted drug delivery [83]. The A10 aptamer was composed of a 57bp nucleotide, stabilized 2-fluoropyrimidine, modified RNA molecule, or targeting PSMA receptor in PCa cells and was used as DOX carrier. Moreover, SPION-TCL was coated with carboxy-PEG-derived polymer which was used for MR contrast agent. The PEG coating prevented the nanoparticles from binding to proteins where its carboxyl group was used to conjugate targeting molecules such as A10 aptamer.

MRI has been utilized for diagnosis of cancer and drug delivery to specific tissue/cells in many researchs [84]. Some hypoxia-induced cancer stem cells have been implicated in tumor metastasis, which have been targeted by hypoxia inducible factor 1a-based aptamer after conjugating with the Mn(II)-modified magnetic nanoparticles (Fe3O4/Mn) [85]. These nanoparticles had a good biocompatibility with low toxicity and therefore could be used for enhanced T1- and T2-weighted MR images. The major advantage for these nanoparticles was their specific targeting to the HIF-1α which is expressed in cancer stem cells under hypoxic conditions.

5. Challenges in Gene Therapy Using Nanoparticles

There are several pathological barriers that need to be overcome before the nanoparticle can specifically deliver their active compound [86–88]. The tumor blood vessels are random and discontinuous with poor alignment of endothelial cells and fenestrations ranging from 100 to 500 nm in size [89, 90]. Therefore, if the nanoparticles are sufficiently smaller, i.e., below 500 nm, they can easily pass through the fenestrations and get accumulated in the tumor tissues [91–93]. Along with tumor endothelial vessel, there are several other barriers which need to be overcome by delivery systems (Table 3).

With much effort in the last few years, the challenges related to siRNA stability, delivery, immune response, and off-target effects are being slowly mitigated, and next generation of delivery systems has been developed and proposed [97]. In the last decade, owing to their avoidance of viral-based vectors and features of highly unstable surface properties, shape, and size, nonviral polymer-based nanodelivery systems have gained a lot of attention especially in gene/drug delivery [18, 98]. Recently, different nucleic acid delivery systems prepared from polymers, inorganic compounds, carbohydrates, and lipids have also been developed [99].

About four different issues have been identified that need to be solved before the gene therapy can be successful for cancer treatment [100]. They are

(i) Identify the major target genes which are critical for pathology and progression of cancer
(ii) Identify the best gene that could be therapeutically used to halt cancer progression
(iii) Optimize gene expression so that the target gene is suppressed

(iv) Delivery of nanoparticles (drug carrier) to target site at an effective level

6. Conclusions

Despite significant progress in various nonviral-based delivery systems being at an early stage of clinical trials, significant study still remains to be overcome before the RNAi therapy can be widely used for prostate cancer gene therapy. The safety outlines of different delivery systems need to be further considered with major emphasis on decreasing the toxicity of system and elimination of nonimportant immune stimulation. The safety of enzymatically degraded products of chemically modified nanoparticles as well as nucleic acids after delivery is also important. For targeted delivery systems using ligand molecules, their immune response should also be addressed. Moreover, a detailed investigation into the major mechanisms related to avoidance of ill effects of nanoparticle vehicles is imperative.

As our horizon of prostate cancer research broadens, integration of novel treatment ideas becomes important for clinicians, biologists, and biochemical engineers. Developing gene delivery vehicles for tumor therapy, based on the unique tumor microenvironment of prostate cancer, is crucial to the delivery of clinically relevant amounts of active therapeutics to the tumor site while by-passing various biological hindrances. Nanotechnology will have an important role in realizing the goal for early diagnosis and personalized medicine for prostate cancer patients at different stages of disease, avoiding unnecessary toxicities associated with the current treatment regimens.

Table 3: Different types of pathological barriers and their prevention methods.

| Pathological barrier     | Effector cells/chemicals                     | Effect on nanoparticles                  | Prevention method                              | Nanoparticle modification required | Reference |
|--------------------------|----------------------------------------------|------------------------------------------|------------------------------------------------|-----------------------------------|-----------|
| Tissue vasculature       | Different angiogenesis structure, pH value, and metabolic rate of tumor tissues | Different accessibility and knockdown levels | NA                                             | NA                                | [91, 94]  |
| Immune system            | TLR-3,7,8                                     | Trigger innate immune response            | Longer siRNA molecule binding to nanoparticles | Dicer-substrate siRNA (25-30 nt long) | [95]      |
| Reticulo endothelial system | Kupfur cells, macrophages                    | Sequestration from circulation             | Coating biocompatible polymer                  | PEGylation, RGD                   | [96]      |
| Plasma and tissue nucleases | Nucleases and lipases                         | Degradation of either nanoparticle polymer and/or RNAi drug (siRNA) | Increase circulation time                      | PEGylation                         | [53]      |
| Endosomes and lysosomes  | Endocytic vesicles                            | Digestion of nanoparticles and siRNA     | Surface conjugation of ligand molecules, decrease the NP size | RGD, PEGylation                   | [92]      |

Conflicts of Interest

The authors declare that they have no conflict of interest in this work.

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