Introduction

Open fractures frequently result from high-energy trauma and very often complicated by potentially fatal injuries.\(^1,2\) Open fractures are characterized by soft tissue injury that occurs in communication of the fracture site with the outside environment. This leads to not only contamination of the fracture site with external microorganisms also the introduction of foreign bodies into the wound.\(^1\)

The injury mechanism significantly influences the microbiological profile on the wound site (road traffic accident, agricultural injury, fall from height, gun-shot injury). It is well regarded that many infections in open fractures are very often nosocomial. This is because, on many occasions, causative microorganisms found inward samples cultures are different from initial surveillance culture at admission.\(^3\) The risk of infection can, therefore, also be influenced positively by the operating technique (e.g., debridement, instrumentation, fracture fixation, irrigation) as well...
as the indoor atmosphere at ward. These infections lead to malunion, nonunion, chronic osteomyelitis and sometimes even ultimate amputation.

Tibial shaft fractures account for 2% of all fractures and 44.4% of all open long-bone fractures in adults. Due to the specific anatomical features of the tibia (limited soft coverage) more than 15% of its’ fractures are classified as open and have made most infection-prone bone of the body. The Gustilo classification system is used widely to grade open fractures. The infection rate of open fractures varies with the fracture characteristic. Infection rates progressively increased from 0% to 2% for Gustilo type-I, 2% to 10% for type-II, and 10% to 50% for type-III. Management of open fractures is based on the principles that include assessment of the patient, classification of the injury, antibiotic therapy, debridement and wound management, fracture stabilization, early bone grafting, and supplemental procedures to achieve healing. Surgical debridement is considered the most critical step for open lower limb fracture management.

This infection complication and antibiotic resistance synergistically pose a significant threat to the health care system. It not only increases the risk for complications, but extended hospital stay and economic burden to the hospital resources put an unexpected load to the citizen as well as the Government. Updated knowledge about all these aspects of open tibia fracture is essential for proper management. The present study has evaluated injury characteristics and resistance patterns of microorganisms involved in the infection of open fracture tibia in a single-centre cross-sectional study among the Bangladeshi population.

Materials and Methods
This was a cross-sectional study conducted between January 2020 to July 2020 at the National Institute of Traumatology and Orthopaedic Rehabilitation (NITOR), Sher-E-Bangla Nagar, Dhaka where patients of all ages and both the sexes presented with open fractures of tibia attending emergency, were studied. Sample size of this study was 384. Purposive sampling (non randomised) according to availability of the patients and strictly considering the inclusion and exclusion criteria were done. From Admission till discharge three successive culture sensitivity test were done. Cultures were initial culture/surveillance culture (culture sample from initial wound before prophylactic antibiotic), post debridement culture (last saline irrigation in emergency theater) and finally infection culture (7 to 10 days after admission).

Approval from the Institutional Review Board (IRB) of National Institute of Traumatology and Orthopaedic Rehabilitation (NITOR), Sher-E-Bangla Nagar, Dhaka was taken to conduct this study. Keeping compliance with Helsinki Declaration for medical research involving human subject 1964, the study subjects were informed verbally about the study design, the purpose of the study and right for withdrawing themselves from the project at any time, for any reason. Individual who gave informed written consent to participate voluntarily in the study were included as a study sample.

Data were collected in a structured data collection form filled up by the investigator containing all the variables of interest according to inclusion criteria after taking their permission regarding participation and this research work. At arrival of the patients, time of hospital admission, demographic variables, date and time of injury were noted along with mechanism of injury, Gustilo types and subtypes and fracture injury characteristics. During initial resuscitation at emergency room, surveillance culture sample (from wound before prophylactic antibiotic) was collected and sent to lab. Prophylactic antibiotics (intravenous Flucloxacillin and 3rd generation cephalosporin) was administered. Then patients were sent to emergency theatre for wound debridement, fracture stabilization and soft tissue care. Debridement was done following current practice at NITOR using chlorhexidine (Hexi scrub), normal saline, hydrogen peroxide and povidone-iodine solution. A second post debridement culture (last saline wash from the wound at emergency theatre) was sent. From the theatre after initial fracture stabilisation. Stable patients were sent to post-operative ward followed by to general ward. A third infection culture sample after admission in the ward at 7 to 10 days was sent for culture sensitivity and identification of organism. Results of all culture were noted in same structured data collection form.

The data were tabulated and quantitative parameters such as age of patient were summarized in terms of mean with standard deviation and percentage, χ²-statistic was applied to see significant difference between variables. The significance of the results was determined in 95% confidence interval and a value of $p<0.05$ considered to be statistically significant.
Results
In present study among 384 studied patients, most (209) were from 21-40 years age group, in contrast with the least number of patients (28) from the elderly (>60 years) group (figure 1). Of them, 345 were male and 39 were female. Regarding Injury Characteristics, according to Gustilo classification, Gustilo type III fracture were predominant, 99(55.3%) followed by type II, 56(31.3%) and among Gustilo subtypes Gustilo IIIA were predominant, 48(26.7%) followed by IIIB, 43(24.0%). (table-I). 142 patients (37.0%) had contamination in surveillance culture while the rest 242 had no organism. Noticeably, rate of infection was largely influenced by wound debridement. In post debridement culture, only 101 (26.3%) patients had contamination compared to around 283(73.7%) with no organism in post debridement wound. But from the ward sample it again increased to 171(44.5%) cases of organism positive. (table-II). The highest infection rate was in Gustilo type III, 130(76.0%) followed by type II, 39 (22.8%). While in subtypes, type IIIB is the predominant, 69 (40.4%), followed by type IIIA, 58(33.9%). (table-III). A total of 15 different type of organisms (bacteria) found in three successive cultures. Number of organisms decreased in 2nd culture (194 decreased to 138) after debridement but again increased in 3rd culture (219 in the samples from the ward). (table-IV). Gram-negative organisms were predominant with multidrug resistance. Pseudomonas and Klebsiella species are only sensitive to intravenous imipenem or meropenem only around 50-60% which is the highest sensitivity of all used drugs for these two organisms in this study. Less commonly used antibiotic Co-trimoxazole and Chloramphenicol showed good sensitivity against Staphylococcus Aureus 90-93% and E. coli 65-70%. But they are not effective against Pseudomonas and Klebsiella species (table-V).

Table I: Injury characteristics: As per Gustilo classification (n=384)

| Type of Fracture | Frequency (Main types) | Frequency (subtypes) |
|------------------|------------------------|----------------------|
| Gustilo I        | 55(14.3)               |                      |
| Gustilo II       | 120(31.3)              | Gustilo IIIA 105(27.3) |
| Gustilo III      | 209(54.4)              | Gustilo IIIB 91(23.7) |
|                  |                        | Gustilo IIIC 13(3.4) |
| Total            | 384(100)               | 209(54.4)            |

Percentage in parenthesis.

Table II: Results of three cultures in terms of organism present/absent among the cases

| Organism Present or absent | Surveillance culture (on arrival) | Postdebridement Culture (at emergency Theater) | Third (infection) Culture (from the wards) |
|----------------------------|----------------------------------|-----------------------------------------------|-----------------------------------------|
|                            | Freq | %    | Freq | %    | Freq | %    |
| Present                    | 142  | 37.0 | 101  | 26.3 | 171  | 44.5 |
| Absent                     | 242  | 63.0 | 283  | 73.7 | 119  | 31.0 |
| Total                      | 384  | 100% | 384  | 100  | *290 | 75.5 |

χ² test found significant difference among the three cultures (P < .05).

Table III: Infection: As per Gustilo classification (n=384)

| Type of Fracture | Frequency (Main types) | Frequency (subtypes) |
|------------------|------------------------|----------------------|
| Gustilo I        | 2(1.2)                 |                      |
| Gustilo II       | 39(22.8)               | Gustilo IIIA 58(33.9) |
| Gustilo III      | 130(76.0)              | Gustilo IIIB 69(40.4) |
|                  |                        | Gustilo IIIC 3(1.8)  |
| Infection Absent | 119(31.0)              |                      |
| Wound healed Cases | 94 (24.5%)             |                      |
| Total            | 384(100)               |                      |

Percentage in parenthesis.
### Table IV: Common organisms in three cultures

| Sensitivity | Stap. | Strept. | Ecoli | Pseud. | Klebs. | Citrob. | Prote. | Acine | Serra. | Prov. | Entero. | Flav. | Plesio. | Aero. | Morga. | Total |
|-------------|-------|---------|-------|--------|--------|---------|--------|-------|--------|-------|---------|------|--------|------|-------|-------|
| 1st Culture | Freq  | 19      | 7     | 34     | 38     | 34      | 5      | 7     | 32     | 1     | 2       | 14   | 1      | 0    | 0     | 194   |
| %           | 9.8   | 3.6     | 17.5  | 19.6   | 17.5   | 2.6     | 3.6    | 16.5  | 0.5    | 1.0   | 7.2     | 0.5  | 0      | 0    | 0     | 100   |
| 2nd Culture | Freq  | 6       | 6     | 23     | 35     | 26      | 3      | 8     | 18     | 1     | 1       | 9    | 0      | 1    | 1     | 138   |
| %           | 4.3   | 4.3     | 16.7  | 25.4   | 18.8   | 2.2     | 5.8    | 13.0  | 0.7    | 0.7   | 6.5     | 0.7  | 0.7    | 0    | 0     | 100   |
| 3rd Culture | Freq  | 5       | 4     | 17     | 109    | 52      | 0      | 12    | 12     | 0     | 3       | 4    | 0      | 0    | 0     | 219   |
| %           | 2.3   | 1.8     | 7.8   | 49.8   | 23.7   | 0       | 5.5    | 5.5   | 0      | 3     | 4       | 0    | 0      | 0    | 0     | 100   |

Stap. - Staphylococcus aureus  
Strept. - Streptococcus sp.  
Ecoli. - Escherichia coli  
Pseud. - Pseudomonas sp.  
Klebs. - Klebsiella sp.  
Citrob. - Citrobacter freundii  
Prote. - Proteus sp.  
Acine. - Acinetobacter

### Table V: Resistance pattern of common four organisms

| Antimicrobial agent’s sensitive (S) / Intermediate (I) / Resistance (R) | Staphylococcus aureus (%) | Escherichiacoli (%) | Pseudomonas spp (%) | Klebsiella spp (%) |
|---------------------------------------------------------------------|---------------------------|---------------------|---------------------|-------------------|
|                                                                     | S (I) R                   | S (I) R              | S (I) R             | S (I) R           |
| Ampicillin                                                          | 35.3 0 64.7 3.1 0 96.9 1.7 0 98.3 0 0 100 |
| Amoxicillin                                                         | 35.3 0 64.7 3.1 0 96.9 1.7 0 98.3 2.4 0 97.6 |
| Piperacillin                                                        | 64.7 5.9 29.4 31.3 9.4 59.4 20.5 36.8 42.7 13.3 6.0 80.7 |
| Cephalexin                                                          | 64.7 0 35.3 15.6 0 84.4 11.1 0 88.9 15.7 0 84.3 |
| Ceftriaxone                                                         | 47.1 0 52.9 37.5 3.1 59.4 5.1 2.6 92.3 16.9 0 83.1 |
| Ceftazidime                                                         | 52.9 11.8 35.3 40.6 3.1 56.3 22.2 1.7 76.1 22.9 4.8 72.3 |
| Cefepime                                                            | 17.6 23.5 58.8 40.6 9.4 50.0 32.5 0 67.5 15.7 6.0 78.3 |
| Cefixime                                                            | 17.6 11.8 70.6 28.1 0 71.9 5.1 0.9 94.0 14.5 0 85.5 |
| Imipenem                                                            | 88.2 0 11.8 78.1 9.4 12.5 53.8 0.9 45.3 62.7 25.3 12.0 |
| Meropenem                                                           | 89 0 11 87.5 9.4 3.1 47.9 8.5 43.6 66.3 12.0 21.7 |
| Gentamicin                                                          | 88.8 0 11.2 75 0 25 23.9 0.9 75.2 34.9 0 65.1 |
| Amikacin                                                            | 94.1 0 5.9 71.9 3.1 25 42.7 4.3 53.0 51.8 1.2 47.0 |
| Netilmicin                                                          | 100 0 0 78.1 0 21.9 35.0 9.4 55.6 38.6 8.4 53.0 |
| Doxycycline                                                         | 88.2 5.9 5.9 53.1 3.1 43.8 17.1 9.4 73.5 33.7 6.0 60.2 |
| Ciprofloxacin                                                       | 58.8 5.9 35.3 50 15.6 34.4 38.5 3.4 58.1 26.5 21.7 51.8 |
| Levofoxacin                                                         | 64.7 5.9 29.4 59.4 3.1 37.5 33.3 6.0 60.7 54.2 7.2 38.6 |
| Moxifloxacin                                                        | 58.8 11.8 29.4 46.9 12.5 40.6 27.4 1.7 70.9 30.1 15.7 54.2 |
| Cotrimoxazole                                                       | 88.2 0 11.8 65.6 3.1 31.3 18.8 0 81.2 27.7 1.2 71.1 |
| Chloramphenicol                                                     | 94.1 0 5.9 68.8 0 31.3 12.8 6.8 80.3 55.4 2.4 42.2 |
| Azithromycin                                                        | 52.9 0 47.1 21.9 9.4 68.8 13.7 6.8 79.5 3.6 4.8 91.6 |

Serra. - Serratia sp.  
Prov. - Providencia alcalifaciens  
Entero. - Enterobacter sp.  
Plesio. - Plesiomonas sp.  
Aero. - Aeromonas  
Morga. - Morganella morganii
Antimicrobial resistance open fracture tibia

Islam SS et al

Figure 1: Age of the patients in different groups

Discussion

Open fracture tibia is one of the commonest emergency admissions in orthopaedic surgery. Exposure and contamination of the open fracture with environmental microorganisms are widespread. Tibia as a pedal bone, being mostly subcutaneous it is very prone to accidental injury and easily become bare following the trauma. An updated knowledge about injury characteristics and the resistance pattern of microorganisms involved in infection is the much required area of open fracture tibia management.

In this study, it was observed that the mean age of the patient was 36.2±15.5 years. An epidemiological study for 7 years on tibia fracture and another related study on Bangladeshi population also found similar mean age. In another study conducted in Brazil, the mean age of the patients with open fractures was 36±12.60 years. The results of all these studies are similar to our study. There were 345(89.8%) male and 39(10.2%) female patients in our study, which states of male predominance. Several related studies has reported male predominance.

Considering injury characteristics according to Gustilo and Anderson classification found Gustilo type III fracture were predominant (54.4%) followed by type II (31.3%) and among the Subtypes IIIA were predominant (27.3%) followed by IIIB (23.7%). A quantitative descriptive study conducted at ULBRA university hospital in Canoas, Rio Grande do Sul, Brazil, found similar type injury characteristics, which is in agreement with this study. A review article comprising 32 articles of 3036 patients of tibia also found same type Gustilo III predominance. Still, among the subtypes, IIIB was the highest subtype, while in our study, it is IIIA. But this difference is minimum.

In this study infection rate of the Gustilo type III fracture was the highest (76.0%) followed by Type II 22.8% and among the subtypes, IIIB having the most top (40.4%) then IIIA (33.9%). A study conducted at Hospital de Pronto Socorro de Canoas found a similar type of finding with the highest infection rate in Gustilo type III fracture. A retrospective clinical analysis of 50 patients who underwent tibial open fracture treatment found infection rate highest in Gustilo type III fractures. This finding is similar to our study, but they differ in the infection rate of subtypes, where they found Gustilo type IIIA having the highest rate. Their sample size is around 1/7 of ours (50 versus 384). It may be an important factor of influence.

This study found only 26.3 % contamination in the wound after debridement. According to EFFORT open reviews, surgical debridement is considered as the pivotal and most essential procedures to reduce bacterial load in open lower limb fractures. The infection rate from the ward samples was 44.5 %, where the post-debridement contamination was less (26.3%). Among the patient entering the ward after debridement with no contamination (73.7%) became infected in 34.6% cases, which indicates hospital-acquired infection. An observational study conducted in a tertiary hospital in India over a one-and-a-half-year period has shown that nosocomial infections are a significant issue worldwide, ranging from 1% in a few European and American countries to more than 40% in Asia.

The bacteriological study found common organisms present in our study in three cultures were Escherichia coli, Pseudomonas spp. and Klebsiella spp. A related microbiological study of wound from All India Institute of Medical Sciences, India also reported Gram-negative organisms were predominant, which is almost similar with this study. A prospective multicentered study on open fracture tibia at the various hospitals of Bangladesh done by Ali et al. (2017) found Staphylococcus aureus and Streptococcus pyogenes as dominant infecting organisms. This finding differs from our study findings. It may be due to they had a very small sample size of 12 cases.

In the present study, common gram-negative and gram-positive organisms alarmingly multidrug-resistant. Pseudomonas and Klebsiella species are only sensitive to intravenous imipenem or meropenem only around 50-60%, which is the highest sensitivity of all used drugs for these two organisms in this study.
Amikacin is the next sensitive drug having only 40-50% sensitivity. Older drug Chloramphenicol was more than 55% sensitive for *Klebsiella species*. In the case of *E. coli* sensitivity of these four drugs was around 70-88% and for gram-positive bacteria *Staphylococcus aureus* 90-95%, but in the case of Netilmicin, the sensitivity is 100% for *Staphylococcus aureus*. Nowadays less commonly used antibiotic Co-trimoxazole and Chloramphenicol showed good sensitivity against *Staphylococcus aureus* 90-95% and *E. coli* 65-70%. But these old antibiotics are only 12-18% sensitive for *Pseudomonas species* and 25-55%. For *Klebsiella species*. A study on 126 patients in China in the year 2016 reported that gram-positive bacteria were susceptible to meropenem and imipenem, while sulbactam and ampicilllin were nearly completely resistant.24 In another sample, 50% of the strains displayed an imipenem and meropenem sensitivity close to our sample.22

There is positive correlation of bacterial contamination between times elapsed since injury and debridement. Further multicentred and randomised study required to evaluate the nosocomial infection addressed in this study.

**Conclusion**

The study found Gustilo type III tibia fractures are predominant injury with highest infection rate. Surgical debridement is effective in reducing contamination from the open fracture wound, but the hospital-acquired infection is common in orthopaedic admitted patients where Gram-negative pathogens were dominant, and the antibiograms showed alarming pattern of drug resistance.

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