Correlation Analysis of Anti-Cardiolipin Antibody/D Dimer/C-Reactive Protein and Coronary Artery Lesions/Multiple-Organ Damage in Children With Kawasaki Disease

Yun-ming Xu, Yan-qiu Chu and Hong Wang*

Pediatric Department of Shengjing Hospital, China Medical University, Shenyang, China

Aim: Kawasaki disease (KD) is a systemic vasculitis with unknown etiology. In addition to cardiovascular system involvement, it can also have other multiple organs involved. This study is aimed at investigating the correlation between anti-cardiolipin antibody (ACA)/D dimer/C reactive protein (CRP) and coronary artery lesions (CAL)/multiple-organ lesions in children with KD.

Methods: Retrospective analysis was performed in 284 KD/IKD patients from May 2015 to April 2016. Among them, 175 were males (61.6%), with average age of 2 years and 5 months old. Patients were divided into ACA+ group and ACA- group, elevated D dimer (DDE) and normal D dimer group (DDN), and coronary artery injury (CAL) group and non-coronary artery injury (NCAL) group.

Results: ACA was most likely tested positive in younger KD children (p < 0.05). ACA+ and hypoproteinemia were correlated with CAL, thrombocytosis, and granulocytopenia (p < 0.05–0.01). Levels of cTnI and CK in the CAL group were significantly higher than those in the NCAL group (p < 0.05). CAL was more frequently detected in younger patients and patients with prolonged fever, later IVIG treatment, and elevated CRP over 100 mg/l, but there was no statistically significant difference (all p > 0.05). In the KD with DDE group, the incidence of granulopenia, thrombocytosis, myocardial damage, cholestasis, hypoproteinemia, and aseptic urethritis was significantly higher than that in the KD with DDN group (p < 0.05–0.01). However, elevated D dimer was not associated with CAL. CRP elevation was highly correlated with D dimer, but not with CAL.

Conclusion: Higher incidence of CAL and myocardial damage occurred in KD patients with positive ACA and hypoproteinemia. In the current study, ACA was only tested for positive and negative, which is a limitation to this study. To further elucidate the association, ACA titers would establish its significance in drawing a conclusion for the
INTRODUCTION

Kawasaki disease (KD) is a systemic vasculitis with unknown etiology. It is one of the common connective tissue diseases in children. Currently, KD has emerged as a major pediatric disorder throughout the developed world (1). In developing countries, KD is currently being diagnosed and reported in both China and India (2–4). Multiple organs/systems can be involved in KD. Coronary artery aneurysm (CAA) is the most significant complication that affects the quality of life in the long term. Even in patients receiving IVIG treatment within 10 days of disease onset, the incidence of CAL is still about 5% (5). Prediction of the occurrence of CAL and MOD by utilizing laboratory examination has become a priority for pediatric cardiologists.

In 2014, four KD children with fever but without identified underlying pathogenesis were hospitalized at the department of rheumatism. Results of the ANA, ANCA, titer test, cytokine test (interferon + interleukin), and anticardiolipin antibody (ACA) test indicated that all the four patients had elevated interleukin and two of them were tested positive for ACA and negative for other tests (6, 7). Meanwhile, two KD patients with KD shock syndrome (KDSS) and macrophage activation syndrome (MAS) were transferred to a pediatric intensive care unit (PICU) where the D dimer test was performed (8). Both had significantly elevated D dimer. These clinical aspects prompted us to review literature on ACA and D dimer. As reported in literature, ACA is positive in infection (9), myocardial infarction (10), infectious endocarditis (11), recurrent abortion (12), KD complications (13), systemic lupus erythematosus (14), and stroke (15). Elevated D dimer is associated with vascular endothelial damage (16), and it has been a supporting tool in early diagnosis of KD (17). Furthermore, it is correlated with CAL (18) and so on. We hypothesized that, since KD in children is systemic vasculitis with a high risk in blood clot, ACA+ and D dimer may be associated with disease progression and be used as indications in KD-related complications. Therefore, we added the ACA/D dimer test to the newly admitted children with KD/IKD.

METHODS

Patients

A total of 284 IKD and KD cases were collected from the pediatric cardiovascular ward of Shengjing Hospital, China Medical University, from May 2015 to April 2016. Among them, there were 175 males (61.62%). The average age of patients was 2 years and 5 months (2 months to 11 years) old. All data were retrospectively analyzed.

Multiple-Organ Injuries

When two or more than two organs are impacted in the same child with KD around the acute and/or subacute stages.

Inclusion Criteria

Patients met diagnostic criteria of KD and IKD (19). The diagnostic criteria for CAL were based on the guidelines of the Japanese Circulator Association (20).

Exclusion Criteria

(1) Incomplete clinical data records; (2) Without ACA and D-dimer tests; and (3) Without IVIG treatment.

Data Collection

All children diagnosed with KD/IKD had routine tests done at admission: blood routine, liver function, CK, CKMB, hs-cTnI, cTnI, NT pro-BNP, and urine routine. The ACA-IgG antibody (ACA antibody tests were done in 2015) was measured before IVIG in 2 ml of blood using enzyme-linked immunosorbent assay (ELISA), following the manufacturer's instruction (Beijing Beier Company), and the test is qualitative (the cutoff is 0.1+ negative control OD value, ACA IgG positive ≥ 0.1+ negative control OD value). D dimer was measured in blood with EDTA anticoagulant using immunoturbidimetry for the routine disseminated intravascular coagulation (DIC) test following instructions (Instrumentation Laboratory Company), and the normal range is <252 µg/l. CRP was measured in blood using the radical immunodiffusion method following the manufacturer's instructions (Abbott's Diagnosis, Inc.), using the bromocresol green (BCG) method. CRP/D dimer and albumin were measured several times within 2 weeks of onset. The highest value of CRP/D dimer and the lowest value of albumin were included in the table for statistical analysis.

Groups

(1) Based on ACA-IgG results, 284 KD/IKD patients were divided into ACA-positive (ACA+) and ACA-negative (ACA−) groups.
(2) Based on D dimer results, 280 KD/IKD (4 children without DIC analysis were excluded) were divided into D dimer-elevated (DDE) and D dimer normal (DDN) groups. (3) According to ECHO results, 284 KD/IKD were divided into coronary artery injury (CAL) and no-coronary artery injury (NO-CAL) groups.

Statistical Analysis

SPSS 22.0 statistical software was used for statistical analysis. The t test was used for data in normal distribution. Statistical differences were measured by t test. Median (M) or quaternary
FIGURE 1 | Age difference for ACA+ group vs. ACA– group.

TABLE 1 | General information.

| Groups | n   | M (%) | Age (Y) | Fever (d) | First IVIG (d) | IVIG -R (%) | CRP ≥ 100 mg/L (%) | CRP ≥ 80 mg/L (%) | CRP ≥ 50 mg/L (%) | CRP ≥ 30 mg/L (%) |
|--------|-----|-------|---------|-----------|---------------|-------------|---------------------|-------------------|-------------------|-------------------|
| ACA+   | 168 | 105(62.5) | 2.1 (1.0, 2.9) | 8.1 | 8.0 | 10 | 38 (22.6) | 52 (31.0) | 95 (56.5) | 124 (73.8) |
| ACA–   | 116 | 70 (60.3) | 2.8 (1.1, 4.0) | 8.0 | 8.0 | 12 | 33 (28.4) | 48 (41.4) | 70 (60.3) | 85 (73.3) |
| P      | 0.714 | 0.002 | 0.763 | 0.581 | 0.173 | 0.265 | 0.071 | 0.524 | 0.920 |
| DDE    | 192 | 113 (58.9) | 1.8 (1.2, 3.0) | 8 (7, 9) | 7 (6, 8) | 14 (7.3) | 57 (29.7) | 82 (42.7) | 135 (70.3) | 165 (85.8) |
| DDN    | 88  | 59 (67.0) | 2.4 (1.2, 4.0) | 8 (6, 9) | 8 (6, 10) | 7 (8.0) | 14 (15.9) | 18 (20.5) | 30 (34.1) | 44 (50.0) |
| P      | 0.191 | 0.222 | 0.274 | 0.002 | 0.845 | 0.014 | 0.000 | 0.000 | 0.000 |
| CAL    | 17  | 11 (64.7) | 2.1 ± 1.3 | 10.2 ± 5.2 | 9.2 ± 4.4 | 3 (17.6) | 7 (41.18) | 7 (41.18) | 10 (58.82) | 14 (82.35) |
| NO-CAL | 267 | 164 (61.4) | 2.4 ± 1.8 | 8.0 ± 2.3 | 7.7 ± 2.0 | 19 (7.1) | 66 (24.72) | 93 (25.34) | 154 (57.68) | 190 (51.77) |
| P      | 0.787 | 0.433 | 0.114 | 0.186 | 0.134 | 0.153 | 0.596 | 0.926 | 0.413 |

DDE, D-dimer elevated; DDN, D-dimer normal; IVIG-R, IVIG resistance; CRP, C reactive protein.

interval (p25–p75) was used for data in non-normal distribution. Enumeration data were shown as rate (%), and the chi-square test was used for comparison. The ROC curve was drawn to analyze the predictive value of relevant laboratory indicators on coronary artery injury in KD. P < 0.05 indicates statistically significant difference.

RESULTS
General Information
Patients in the ACA+ group were significantly younger (2.1 years old) than those in the ACA– group (2.8 years old) (Figure 1, Z = −2.516, p = 0.002).

The average time of the first IVIG treatment in the DDE group was 1 day earlier than that in the DDN group (both groups had fever≥ 5 days) (p < 0.05). CRP was highly associated with D dimer despite the cutoff value of CRP (all p < 0.001), There was no significant correlation between D dimer and ACA (Table 1).

The Correlation Between ACA/D-Dimer and Myocardial/Liver Damage in KD/IKD Children
There were no significant differences in myocardial and liver damages between CAL and NO-CAL groups (p > 0.05).
TABLE 2 | The correlation between ACA/D-dimer and myocardial/liver damage in KD/IKD children.

| Groups | n      | CKMB (>24U/L) | CTNI (>0.04 µg/L) | Hs-CTNT (>0.014 ng/mL) | NT-pro BNP (>300 pg/mL) | TBA (>10 umol/L) | ALT (>40 U/L) | AST (>35 U/L) | ALB (<30 g/L) | AAR |
|--------|--------|---------------|------------------|------------------------|-------------------------|-------------------|---------------|---------------|---------------|----------------|-----|
| ACA+   | 168    | 20            | 0.005            | 0.005                  | 358                     | 5.7               | 20            | 25            | 33.5          | 1.42           |
|        | (17, 27) | (0, 0.011) | (0, 0.007) | (169,1049) | (2.9,9.9) | (11.7,43.5) | (20,37) | (29.9,36.3) | (0.68,1.91) |
| ACA−   | 116    | 20            | 0.009            | 0.005                  | 500                     | 6.3               | 17.5          | 24            | 33            | 1.41           |
|        | (17, 25) | (0, 0.009) | (0,0.007) | (150,1378) | (3.8,12.1) | (11.65) | (17,34.8) | (29.5,37.2) | (0.77,2.01) |
| P      | 0.556  | 0.137         | 0.291            | 0.152                  | 0.698                    | 0.815             | 0.061         | 0.771         | 0.961         |
| DDE    | 192    | (190) 20      | (187) 0.010      | (188) 0.005            | (188) 543.6              | (169) 6.4         | 20            | 25.5          | 32.3          | 1.24           |
|        | (17, 24) | (0.010, 0.012) | (0.003, 0.007) | (228.5,1377) | (3.4,11.3) | (11,66.5) | (19,35.5) | (29.3,35.7) | (0.58, 1.91) |
| DDN    | 88     | (87) 22       | (86) 0.010       | (85) 0.004             | (84) 185.4               | (80) 4.9          | 16            | 23            | 36.2          | 1.61           |
|        | (17.5, 28) | (0.010, 0.010) | (0.003, 0.007) | (66.2, 494.7) | (3.2, 8.4) | (11, 23) | (18, 34) | (33.1, 39.1) | (1.15, 2.08) |
| P      | 0.160  | 0.002         | 0.145            | 0.000                  | 0.002                    | 0.002             | 0.005         | 0.000         | 0.006         |
| CAL    | 17     | 17            | 0.01             | 0.004                  | 358                      | 3                 | 5             | 3             | 32.6          | 1.41           |
|        | (15.5, 21.5) | (0, 0.024) | (0, 0.007) | (130,1323) | (23.1) | (31.3) | (18.8) | (28.0,35.1) | (0.78,1.68) |
| NO-CAL | 267    | 20            | 0                | 0.005                  | 435.5                    | 65                | 72            | 67            | 33.9          | 1.43           |
|        | (17.26) | (0, 0.01) | (0, 0.007) | (153,1116) | (27.8) | (27.3) | (25.4) | (30.2,36.9) | (0.75,2.00) |
| P      | 0.110  | 0.013         | 0.698            | 0.838                  | 1.000                    | 0.775             | 0.768         | 0.364         | 0.933         |

CKMB, creatine kinase isoenzyme MB; CTNI, cardiac troponin I; Hs-CTNT, high sensitive cardiac troponin T; NT-proBNP, N-terminal brain naturetic peptide premise; TBA, total bile acid; ALT, alanine transaminase; AST, aspartate transaminase; DDE, D-dimer elevated; DDN, D-dimer normal; AAR, AST/ALT ratio; the average of AAR is 0.8.

FIGURE 2 | The incidence of hypoproteinemia was significantly higher in the CAL group than in the NO-CAL group (p < 0.05). The X-axis indicates with or without presence of CAL. The Y-axis represents the albumin (g/L).

The Correlation Between ACA/D Dimer and the Incidence of Multiple-Organ Injuries

The incidence of granulocytopenia, thrombocytosis, and CAL in the ACA+ group was significantly higher than in the ACA- group (p < 0.01–0.05).

The incidence of elevated NT pro-BNP and ALT, hypoproteinemia, sterile urethritis, and thrombocytosis was significantly higher in the DDE group than in the DDN group (p < 0.01–0.05) (Table 3).
In the DDE group, there were 113 patients who were ACA+ (59.2%), which was significantly higher than in the DDN group \((p < 0.000)\), but the incidence of DDE was not significantly higher than DDN in the CAL group (Figure 3).

**The Correlation Between CAL and Myocardial/Liver Damages**

CAL group presented with significantly higher incidence of hypoproteinemia and cTnI, \((p < 0.05)\). CAL occurred frequently in patients who were younger, with prolonged fever, later IVIG treatment, CRP elevated over 100 mg/L, and lower AAR. However, there were no significant difference \((p > 0.05)\; \text{Table 4}\).

**The Receiver Operating Characteristics (ROC) of cTnI Used in Predicting Coronary Artery Damage in Children With KD**

The AUC of the area under cTnI curve was 0.657, 95% CI was 0.508–0.805 \((p < 0.05)\). When 0.0175 was used as the cutoff value, the sensitivity and specificity for predicting CAL were 0.412 and 0.876 (Figure 4).

**DISCUSSION**

**The Current Clinical Application of ACA Test**

ACA are antcardiolipin antibodies. They were first detected using radioimmunoassay (22). Under normal conditions, the immune system does not recognize cardiolipin that lies at the inner mitochondrial membrane. However, once cardiolipin is exposed, it will trigger an immune response and produce the autoimmune antibody ACA. Immunophenotypes include IgG, IgM, and IgA. Among them, IgG is a pathogenic autoantibody, and its high titer and long half-life make it an important player in immune response. In a study including 34 Italian KD children, antcardiolipin (aCL) was detected in 30% of KD patients (IgG aCL antibodies were found in 14 patients, IgM aCL in 1, and 1 had both) (23). The target antigen of ACA is negatively charged cardiolipin on the platelet and endothelial cell membranes. ACA can stimulate immune response to the phospholipids of its own cell membranes and damage the cell membranes and thus release a large amount of inflammatory cytokines and chemokines and further trigger vascular inflammation. ACA is commonly found in the following diseases: (1) thrombotic diseases, such as myocardial infarction (24), stroke (25), and habitual abortion. There is marked activation of the endothelium and immune system in KD. Anticardiolipin antibodies (aCL) can cause activation of the endothelium. An increase in IgA anticardiolipin antibodies in KD patients suggests that the degree of increase in these antibodies correlates with the degree of systemic inflammation (26). Low IgG was not a risk factor for CAAs in this study. However, KD patients with relatively high IgG prior to treatment may have an increased risk of resistance to initial IVIG therapy (27). Studies have suggested that (13, 28) ACA, as one of laboratory indications for thrombosis, can cause damage in vascular endothelial cells, interfere with the coagulation system, affect platelet function, and reduce
fibrinolytic activity. Finnazzi et al. (29) performed a blind study on 360 ACA-positive patients and concluded that a high level of ACA was a meaningful indicator for predicting thrombosis. Sueoka et al. (30) raised an alarm in patients who had developed thrombosis in the presence of positive ACA after percutaneous thoracoscopic surgery. There have been discussions on the clinical significance of positive ACA antibodies. In healthy controls, aCL antibodies were found in five patients (22%) (23). In order to differentiate from false-positive interference, many medical institutions have implemented testing ACA antibody titers (31) (our hospital is currently using semi-quantified). (1) ACA is also found in connective tissue diseases, such as systemic lupus erythematosus and KD. (2) ACA can be detected in patients infected with syphilis, AIDS, hepatitis C, tuberculosis, parvovirus, and cytomegalovirus. Thus, there is a limitation in evaluating the association of ACA-IgG with vasculitis in Kawasaki disease. ACA titer may stratify the association between ACA and Kawasaki disease. In our study, the incidence of CAL and the proportion of thrombocytosis in children with elevated ACA were significantly increased, suggesting a possible mechanism that inflammatory mediators and the activation of negatively charged cardiolipin in platelets caused damages in the vascular endothelium, which increased the risk of thrombosis. Our results showed that CAL was more common in ACA+ patients. This is consistent with literature reports (32). In contrast to the correlation between CAL and low AAR reported by Wang et al. (21), our results showed that CAL was not associated with AAR. On the other hand, elevated D dimer was correlated with low AAR and NT-pro-BNP, but not with CAL. Since the COVID-19 outbreak, there have been various complications related to thrombosis, and they are accompanied by DIC and an increase in ACA (33, 34). These results suggest that COVID-19 virus can activate vascular endothelium and cause endothelial damage and intravascular coagulation.

The Significance of Elevated D Dimer

The D dimer test is often used in clinical diagnosis of diffuse intravascular coagulation (DIC), deep vein thrombosis (DVT), pulmonary embolism, myocardial infarction, cerebral infarction, etc. This index can not only be used for the diagnosis of thrombotic diseases but also be used as an index for monitoring the dose of thrombolytic drugs for therapeutic effect. KD itself is an inflammation-mediated vasculitis that activates the vascular endothelium and causes endothelial damage. Symptoms are similar to multisystem inflammatory syndrome in children.
FIGURE 4 | The ROC curve of cTnI levels in predicting CAL in children with KD.

with COVID-19. Elevated D dimer is associated with DIC hypercoagulation and macrophage activation and aggravates tissue disintegration (32). In addition, elevated D dimer has been reported to be correlated with CAL (35) and complications in multiple organs. Our study showed that elevated DD was only correlated with complications in MDO but not with CAL. Kong et al. (36) reported that elevated D dimer was also correlated with IVIG resistance in KD children (18). However, our results showed that D dimer elevation was not associated with IVIG resistance. We also found that KD with MAS complication and necrotizing pneumonia was not associated with CAL, although patients had fever which lasted for more than 2 weeks. CRP increased surprisingly (7). The conclusion drawn from our study disagrees with other reports in literature (37). In this study, there were two KD children with elevated D dimer over 10,000 µg/L (D dimer at 24,007 µg/l in one 19-month-old boy with pneumonia and aseptic meningitis and at 10,773 µg/l in the other 17-month-old girl with hemophagocytic syndrome, hyponatremia, pneumonia, bilateral pleural effusion, liver function damage, hypoproteinemia, pancreatic injury, and aseptic meningitis). Both patients did not have CAL. It is consistent with the report by Ming-Tsan et al. (38). Data at our center showed that there was no relationship between increased D dimer and IVIG resistance either, which is conflicted with reported studies (39). This may be associated with earlier IVIG treatment by 1 day in DDE, or it may be related to the differential expression of immune responsive genes during the KD occurrence and development (40).

D Dimer in Children With Multisystem Inflammatory Syndromes

Last year, the coronavirus disease 2019 (COVID-19) hit the world, and a new manifestation emerged as a multisystem inflammatory syndrome in children (MIS-C) which carries similar clinical symptoms of KD, including toxic shock syndrome and severe sepsis (41). The severity is significantly correlated with D dimer, even KDSS (42). COVID-19 linked with thrombotic microangiopathy triggers multiple vasculitis along with arteriole thrombosis, and medium and large venous and arterial vessels mediate the disseminated intravascular coagulation (DIC) (43). Additionally, 52% of MIS-C has elevated D dimer (44). Although some children with KD in our center last year had very similar symptoms to COVID-19 with MIS-C (45), we did not find any evidence of COVID-19 infection in these patients. However, the ultimate treatments for these patients rely on IVIG and glucocorticoid, which are similar to treatments for rheumatic immune diseases such as KD (46).

CONCLUSION

Our study indicated that ACA+ and hypoproteinemia were correlated with CAL, granulocytopenia, and thrombocytopenia in KD children. In the current study, ACA was only tested for positive and negative, which is a limitation to this study. To further elucidate the association, ACA titers would establish its significance in drawing a conclusion for the significance of
ACA in CAL and myocardial damages. Elevated D dimer was correlated with MOD but not with CAL; CRP was correlated with D dimer, but not with ACA and CAL.

**DATA AVAILABILITY STATEMENT**

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

**REFERENCES**

1. Shulman ST. IVGG therapy in Kawasaki disease: mechanism(s) of action. *Clin Immunol Immunopathol.* (1989) 53(2 Pt 2):S141–6. doi: 10.1016/0090-1229(89)90079-2
2. Chen JJ, Ma XJ, Liu F, Yan WL, Huang MR, Huang M, et al. Epidemiologic features of Kawasaki disease in Shanghai from 2008 through 2012. *Pediatr Infect Dis J.* (2016) 35:7–12. doi: 10.1097/INF.00000000000009914
3. Tang Y, Gao X, Shen J, Sun L, Yan W. Epidemiological and clinical characteristics of Kawasaki disease and factors associated with coronary artery abnormalities in East China: nine years experience. *J Trop Pediatr.* (2016) 62:86–93. doi: 10.1016/j.jtroped.2016.e ng.107
4. Filanía RK, Singh S. Kawasaki Disease. In: Cimaz R., editors. Periodic and Non-Periodic Fevers. *Rheumatic Fever, Anti-phospholipid Syndromes, and the Systemic Lupus Erythematosus (SLE)*. Cham: Springer (2020). 45–63. doi: 10.1007/978-3-030-19055-2_4
5. Soriano M, Martínez E, Negreira S, González M, Romero PC, Fernández E, et al. Risk of coronary artery involvement in Kawasaki disease. *Arch Argent Pediatr.* (2016) 114:107–13. doi: 10.5546/aap.2016.e ng.107
6. Xu YM, Chu YQ, Li XM, Wang C, Ma QM, Yu XM, et al. The complementary relationship between echocardiography and multi-slice spiral CT coronary angiography in the diagnosis of coronary artery thrombosis in children with Kawasaki disease. *Front Pediatr.* (2021) 9:670887. doi: 10.3389/fped.2021.670887
7. Wang H. *Pediatric Kawasaki Disease: Clinical Analysis and Cases.* (2017). People's Medical Publishing House (in Chines). Peking.
8. Wang H. *Pediatric Kawasaki Disease: Clinical Analysis and Cases.* Singapore: Springer (2021).
9. Makino J, Koshy S, Bajaj S, Jeong YG, David C, Perlman DC. Fulminant ecchymosis as the initial manifestation of antiphospholipid syndrome (APS) triggered by respiratory syncytial virus (RSV) infection: a case report and review of the literature. *ID Cases.* (2016) 7:19–22. doi: 10.1016/j.idcr.2016.10.013
10. Andreoli L, Chighizola CB, Banzato A, Pons-Estel GJ, Ramire de Jesús G. Estimated frequency of antiphospholipid antibodies in patients with pregnancy morbidities, stroke, myocardial infarction, and deep vein thrombosis: a critical review of the literature. *Arthritis Care Res.* (2013) 65:1869–73. doi: 10.1002acr.22066
11. Zaratzian C, Gouriet F, Tissot-Dupont H, Casalta J-P, Million M, Alla F. Possible relationship between antiphospholipid antibodies in children with Kawasaki disease. *Ann Rheum Dis.* (2016) 75:1869–73. doi: 10.1136/annrheumdis-2016-209031
12. Zhang N, Leng X, Tian X, Zhao Y, Zeng X. Clinical analysis of 6 patients with drug-induced lupus. *Zhonghua Nai Ke Za Zhi.* (2016) 55:211–5. doi: 10.3760/cma.j.issn.0578-1426.2016.03.011
13. Selton-Suty C, Maigret C-H, Devignes I, Goehringer F, Erpelding M-L, Alla F. Possible relationship between antiphospholipid antibodies and embolic events in infective endocarditis. *Heart.* (2018) 104:509–16. doi: 10.1136/heartjnl-2017-312359
14. Morise T, Takeuchi Y, Takeda R, Karayalcin U, Yachie A, Miyawaki T. Increased plasma endothelin levels in Kawasaki disease: a possible marker for Kawasaki disease. *Angiology.* (1993) 44:719–23. doi: 10.1177/000331979304400908
15. Imamura T, Yoshihara T, Yokoi K, Nakai N, Ishida H, Kasabuchi Y. Impact of increased D-dimer concentrations in Kawasaki disease. *Eur J Pediatr.* (2005) 165:526–7. doi: 10.1007/s00431-005-1699-7
16. Masuzawa Y, Morii M, Hara T, Inaba A, Oba MS, Yokota S. Elevated D-dimer level is a risk factor for coronary artery lesions accompanying intravenous immunoglobulin-unresponsive Kawasaki disease. *Ther Apher Dial.* (2015) 19:171–7. doi: 10.1111/1447-9987.12235
17. Ayusawa M, Sonobe T, Uemura S, Ogawa S, Nakamura Y, Kiyosawa N, et al. Revision of diagnostic guidelines for Kawasaki disease (the 5th revised edition). *Pediatr Int.* (2005) 47:232–4. doi: 10.1111/j.1442-2003.2005.00233.x
18. Japanese Circulation Society Joint Research Group. Guidelines for diagnosis and management of cardiovascular sequelae in Kawasaki disease. *Circ J.* (2010) 74:1989–2020. doi: 10.1253/circj.CJ.10-7-04093
19. Wang IJ, Li J, Ren Y, Shi HY, Rong X, Zhang XT, et al. Association between alamine aminotransferase/aspartate aminotransferase ratio (AST/ALT Ratio) and coronary artery injury in children with Kawasaki disease. *Cardiol Res Pract.* (2020) 2020:8743548. doi: 10.1155/2020/8743548
20. Willis R, Papalard O, Nigel Harris E. Solid phase immunooassay for the detection of antiphospholipid antibodies. *Methods Mol Biol.* (2017) 1646:185–99. doi: 10.1007/978-1-4939-7919-1_16
21. Falcini F, Trapani S, Turchini S, Farsi A, Ermini M, Keser G, et al. Immunological findings in Kawasaki disease: an evaluation in a cohort of Italian children. *Clin Exp Rheumatol.* (1997) 15:685–9.
22. Ertas F, Can O, Acet H, Ozbakkaloglu M. The clinical significance of antiphospholipid antibody levels in patients with acute myocardial infarction: a regional study. *Postepy Kardiol Interwencyjne.* (2013) 9:328–31. doi: 10.5114/pki.2013.38859
23. Gašperši ˇc N, Zaletel M, Kobal J, Žigon P, Cu ˇcnik S, Šemrl SS, et al. Stroke and antiphospholipid syndrome-antiphospholipid antibodies are a risk factor for an ischemic cerebrovascular event. *Clin Rheumatol.* (2019) 38:379–84. doi: 10.1007/s10067-018-4247-3
24. Gupta M, Johann-Liang R, Bussell JB, Gersony WM, Lehman TWM. Elevated IgA and IgM antiphospholipid antibodies in acute Kawasaki disease. *Cardiology.* (2002) 97:180–2. doi: 10.1159/000063118
25. Yanagimoto K, Nomura Y, Masuda K, Hirabayashi M, Morita Y, Yoshisige M, et al. Immunoglobulin G values before treatment are correlated with the responsiveness to initial intravenous immunoglobulin therapy for Kawasaki disease. *Pediatrics.* (2017) 140:1274–7. doi: 10.1542/peds.2016-4734
26. Finazzini G, Brancaccio V, Moia M, Ciavarella N, Mazzuco MG, Schincro PC, et al. Natural history and risk factors for thrombosis in 360 patients with antiphospholipid antibodies. A four-year prospective study from the italian registry. *Ann Med.* (1996) 100:330. doi: 10.1016/0003-3993(96)00060-5
27. Sueoka J, Kataoka M, Shimura N, Inami T, Yanagisawa R, Ishiguro H. Therapeutic efficacy after percutaneous transluminal pulmonary angioplasty in CTEPH with and without clotting disorder according to anti-cardiolipin
antibody. Int J Cardiol. (2015) 201:271–3. doi: 10.1016/j.ijcard.2015.08.104
31. Xiao M, Yan Zhang Y, Zhang SL, Qin XZ, Xia P, Cao W, et al. Antiphospholipid antibodies in critically ill patients with COVID-19. Arthritis Rheumatol. (2020) 72:1998–2004. doi: 10.1002/art.41425
32. Favaloro EJ, Wong RCW. Current clinical and laboratory practice for the investigation of the antiphospholipid syndrome: findings from the 2008 Australasian antiphospholipid antibody survey. Pathology. (2009) 41:666–75. doi: 10.3109/00313020903257731
33. Zhang Y, Xiao M, Zhang S, Xia P, Cao W, Jiang W, et al. Coagulopathy and antiphospholipid antibodies in patients with covid-19. N Engl J Med. (2020) 382:e38. doi: 10.1056/NEJMc2007575
34. Zhou Y, Wang S, Zhao J, Fang P. Correlations of complication with coronary arterial lesion with VEGF, PLT, D-dimer and inflammatory factor in child patients with Kawasaki disease. Eur Rev Med Pharmacol Sci. (2020) 22:5121–6. doi: 10.26355/eurrev_201808_15706
35. Kong WX, Ma FY, Fu SL, Wang W, Xie CH, Zhang YY, et al. Biomarkers of intravenous immunoglobulin resistance and coronary artery lesions in Kawasaki disease. World J Pediatr. (2019) 15:168–75. doi: 10.1007/s12519-019-00234-6
36. Pilania RK, Jindal AK, Johnson N, Prithvi A, Vignesh P, Suri D, et al. Macrophage activation syndrome in children with Kawasaki disease: an experience from a tertiary care hospital in northwest India. Rheumatology. (2020) 60:3413–9. doi: 10.1093/rheumatology/keaa715
37. Ming-Tsan L, Lon-Yen T, Ming-Lin C, Yu-Jun C, Hsiao-Neng C, et al. Absence of hypercoagulability in acute Kawasaki disease. Pediatr Int. (2005) 47:126–31. doi: 10.1111/j.1442-200x.2005.02025.x
38. Maria CM, Giovanni C, Eugenia P, Rolando C. Kawasaki disease in Sicily: clinical description and markers of disease severity. Ital J Pediatr. (2016) 42:92. doi: 10.1186/s13052-016-0306-2
39. Nie H, Wang SZ, Wu QL, Xue DN, Zhou WM. Five immune-gene-signatures participate in the development and pathogenesis of Kawasaki disease. Immun Inflamm Dis. (2020) 9:157–66. doi: 10.1002/iid3.373
40. Elizabeth MD, Emilia HK, Eric JC, Elizabeth MR, Alison M, Jemma R, et al. Multisystem inflammatory syndrome in children in New York State. N Engl J Med. (2020) 383:347–58.
41. Li YD, Zheng Q, Zou LX, Wu JQ, Guo L, Teng LP, et al. Kawasaki disease shock syndrome: clinical characteristics and possible use of IL-6, IL-10 and IFN-γ as biomarkers for early recognition. Pediatr Rheumatol Online J. (2019) 17:1. doi: 10.1186/s12969-018-0303-4
42. Vinayagam S, Sattu K. SARS-CoV-2 and coagulation disorders in different organs. Life Sci. (2020) 260:118431. doi: 10.1016/j.lfs.2020.118431
43. Jun Y, Toshiki K, Hisato T, Naokata S. Clinical characteristics of COVID-19 in children: a systematic review. Pediatr Pulmonol. (2020) 55:2565–75.
44. Mamishi S, Movahedi Z, Mohammad M, Ziaee V, Khodabande H, Abdolsalehi MR, et al. Multisystem inflammatory syndrome associated with SARS-CoV-2 infection in 45 children: a first report from Iran. Epidemiol Infect. (2020) 148:e196. doi: 10.1017/S095026882000196X
45. Shah S, Danda D, Kavachchanda C, Das S, Adarsh MB, Negi VS. Autoimmune and rheumatic musculoskeletal diseases as a consequence of SARS-CoV-2 infection and its treatment. Rheumatol Int. (2020) 40:1539–54. doi: 10.1007/s00296-020-04639-9

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Xu, Chu and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.