Heart bailout by cell therapy: introducing an acceptable test for comparing cell accountability

Paolo Madeddu*

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Abstract

Cell therapy for cardiovascular disease is still in its initial phase of development and hence stringent studies are now required for comparison between available approaches using validated experimental models. The best cell for regenerative purposes should have the ability to stimulate vascular repair and cardiomyogenesis in a time-programmable fashion, cooperating with reparative processes afforded by resident cells. However, these requirements are often unreachable with individual cell types currently used in clinical trials as documented by an interesting article from Barclay and colleagues in the current issue of Stem Cell Research and Therapy.

Cell therapy is an emerging field that promises to help cardiovascular patients to regain normal function. Contemporary approaches rely on subsidising the ischaemic heart and limb muscles with vascular growth factors or stem cells with the hope that encouraging the formation of new blood vessels may eventually result in myocyte salvage/regeneration. However, these requirements are often unreachable with individual cell types currently used in clinical trials as documented by an interesting article from Barclay and colleagues in the current issue of Stem Cell Research and Therapy.

Simultaneously compare mononuclear cells from normal peripheral blood and haematopoietic progenitor cell-rich cell sources (umbilical cord blood, mobilised peripheral blood, bone marrow), CD34+-enriched or -depleted subsets of the above, and outgrowth cell populations from these cells. The results are highlighting yet somehow surprising: CD34+ cells from mobilised peripheral blood or umbilical cord blood are the only cells able to promote new vessel growth; however, they do not incorporate into vessels. Conversely, endothelial outgrowth cells incorporate into vessels, without promoting vessel growth. Data from this study confirm that we are far from using an optimal cell product for promotion of therapeutic vascularisation since the most available cells from peripheral blood would simply provide indirect stimulation of the spontaneous angiogenesis process, whereas the more rare population of true endothelial progenitors is angiogenetically inactive. It could be interesting to investigate if a combination of the two populations could be therapeutically utilitarian.

Several caveats need to be considered when interpreting results from this interesting study. First, cells were originated from human healthy donors and injected into an immunodeficient mouse model. Species-related differences could affect the integration of human cells in functional vascular networks. Moreover, it is not clear if human cells from different sources express surface antigens that can differentially affect the cohesion to mouse vasculature. Second, the plug assay does not necessarily reflect the typical environment of an ischemic tissue, with particular reference to the activated state of endothelial cells and the expression of adhesion molecules required for engraftment of donor cells [2]. Therefore, studies in animal models may not be sufficient to draw conclusions on the mechanisms of integration of autologous cells in a clinical setting. Humanized mice for studying human leukocyte integrins in vivo might be useful to address those pressing questions [3]. Obviously, clinical trials comparing different cell populations will give a definitive answer.

Since the initial description by Asahara and colleagues [4], the characteristics of cultured endothelial progenitors...
continue to remain uncertain. The same author recently reported that floating cells from primary bone marrow outgrowth are able to form thick/stable tubes, with hypoxia or shear stress inducing further enhancement of these endothelial-like features [5]. Owing to the difference in the procedural isolation of haematopoietic cells from mouse and human bone marrow, it is difficult to conclude if this refined protocol can help us to select optimal angiogenic cells from total human haematopoietic cells. Enrichment using functional assays rather than antigenically based sorting could be useful for this purpose [6].

The status of the donor source is also very important in determining the fate of injected cells. In fact, disease state can impinge upon the integrity of the stem cell niche, thus undermining the final therapeutic effect [7]. In this context, a comparison of different cell types from the same individual could be crucial, although difficult to realise, for designing new cell therapy strategies.

Obviously, when deciding the best cell therapy, other critical issues need to be considered apart from promotion of angiogenesis. Recent studies have highlighted the possibility that circulating calcifying cells might be deeply intertwined in the development of osteoporosis and vascular calcification [8]. Enhanced imaging systems could help to rule out the possibility of calcifications in hearts receiving different types of human cells. Furthermore, the time window for cell harvesting is important as the same author recently concluded if this refined protocol can help us to select optimal angiogenic cells from total human haematopoietic cells.

In conclusion, current approaches for heart bailout with donations of unspecialised cells are inadequate. To this end, rigorous extension of the seminal work of Barclay and colleagues could accelerate the clinical refinement of cell therapy for the benefit of patients.

**Competing interests**
The author declares that he has no competing interests.

**Published:** 14 August 2012

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doi:10.1186/scrt123

Cite this article as: Madeddu P: Heart bailout by cell therapy: introducing an acceptable test for comparing cell accountability. Stem Cell Research & Therapy 2012, 3:32.