Ancient Mitogenomes Provide New Insights into the Origin and Early Introduction of Chinese Domestic Donkeys

Linying Wang1, Guilian Sheng2,3, Michaela Preick4, Songmei Hu5, Tao Deng6, Uline H. Taron7, Axel Barlow4,7, Jiaming Hu1, Bo Xiao6, Guojiang Sun1, Shiwen Song2, Xindong Hou2,3, Xulong Lai3,8, Michael Hofreiter4* and Junxia Yuan1,3*

1Faculty of Materials Science and Chemistry, China University of Geosciences, Wuhan, China, 2School of Environmental Studies, China University of Geosciences, Wuhan, China, 3State Key Laboratory of Biogeology and Environmental Geology, China University of Geosciences, Wuhan, China, 4Institute for Biochemistry and Biology, University of Potsdam, Potsdam, Germany, 5Shaanxi Provincial Institute of Archaeology, Xi’an, China, 6Key Laboratory of Vertebrate Evolution and Human Origins of Chinese Academy of Sciences, IVPP, Beijing, China, 7School of Science and Technology, Nottingham Trent University, Nottingham, United Kingdom, 8School of Earth Sciences, China University of Geosciences, Wuhan, China

Both molecular data and archaeological evidence strongly support an African origin for the domestic donkey. Recent genetic studies further suggest that there were two distinct maternal lineages involved in its initial domestication. However, the exact introduction time and the dispersal process of domestic donkeys into ancient China are still unresolved. To address these questions, we retrieved three near-complete mitochondrial genomes from donkey specimens excavated from Gaoling County, Shaanxi Province, and Linxia Basin, Gansu Province, China, dated at 2,349-2,301, 469-311, and 2,160-2,004 cal. BP, respectively. Maximum-likelihood and Bayesian phylogenetic analyses reveal that the two older samples fall into the two different main lineages (i.e., clade Ⅰ and clade Ⅱ) of the domestic donkey, suggesting that the two donkey maternal lineages had been introduced into Midwestern China at least at the opening of Silk Road (approximately the first century BC). Bayesian analysis shows that the split of the two donkey maternal lineages is dated at 0.323 Ma (95% CI: 0.583–0.191 Ma) using root-tip dating calibrations based on near-complete mitogenomes, supporting the hypothesis that modern domestic donkeys go back to at least two independent domestication events. Moreover, Bayesian skyline plot analyses indicate an apparent female population increase between 5,000 and 2,500 years ago for clade Ⅰ followed by a stable population size to the present day. In contrast, clade Ⅱ keeps a relatively stable population size over the past 5,000 years. Overall, our study provides new insights into the early domestication history of Chinese domestic donkeys.

Keywords: Chinese domestic donkeys, ancient DNA, mitochondrial genome, maternal lineage, divergence time
INTRODUCTION

The domestication of the donkey (Equus asinus) is a vital event in human history, which played a significant role in the development of human civilization (Liu et al., 2010; Bai et al., 2017; Wang et al., 2020). Donkey deeply transformed ancient societies and land-based transport in Africa and Eurasia, contributed to the growth of the early Egyptian State, and allowed the development of mobile pastoralism and ancient overland trade routes (Denbow et al., 1993; Denham and Iriarte, 2007; Rossel et al., 2008; Kimura et al., 2010). The domestication of the donkey therefore probably indicates a major cultural shift away from sedentary, agrarian life-styles towards more migration and trade in ancient times (Beja-Pereira et al., 2004; Han et al., 2014). However, compared with the other domesticated species of the genus Equus, i.e., the horse, the domestic donkey is greatly underrepresented in the scientific literature (Blench, 2000; Lu et al., 2008; Ma et al., 2020). In the last decades with the promotion of agricultural mechanization and the rapid development of the transportation industry in modern society, the role of domestic donkeys as a means of transportation is decreasing and the number of donkeys has declined greatly (Xie, 1987). Despite these developments, currently donkeys still remain an essential means of transport for people living in mountain areas, deserts, and underdeveloped regions of the world (Blench, 2000; Smith and Pearson, 2005; Kimura et al., 2010; Ma et al., 2020).

Archaeological evidence suggests an African origin for the donkey (Epstein, 1971; Clutton-Brock, 1992; Rossel et al., 2008). The earliest domestic donkey remains, 5,000-year-old ass skeletons, have been excavated from an early pharaonic mortuary complex at Abydos, Middle Egypt, which exhibit a range of osteopathologies consistent with load carrying (Rossel et al., 2008). However, it is often difficult to determine whether the remains from early phases of animal domestication originate from animals that have been domesticated or not (Peters and von der Driessc, 1997; Rossel et al., 2008). Compared with the horse, donkey remains are relatively rare in archaeological sites and are not easily distinguished from the former based on morphological characters alone (Han et al., 2014). Therefore, the available morphological evidence provides limited information about the timing and location of donkey domestication.

Mitochondrial and nuclear DNA have revealed that domestic donkeys originated from African wild asses (Ivankovic et al., 2002; Beja-Pereira et al., 2004; Kimura et al., 2010; Ma et al., 2020; Wang et al., 2020). Mitochondrial DNA studies showed that domestic donkeys harbored two distinct lineages (i.e., clade I and clade II). Clade I (Nubian lineage) contains domestic donkeys and the Nubian wild ass (Equus africanus africanus), while clade II (unknown origin) probably derived from a now extinct African wild ass population, which might have been close to the Somali wild ass (Equus africanus somaliensis) (Kimura et al., 2010, 2013; Ma et al., 2020). Wang et al. (2020) recently analyzed 126 modern domestic donkey nuclear genomes. Their D-statistic analysis showed an African domestication of donkeys, consistent with the results from mitochondrial DNA, and indicated its subsequent spread to Europe and Asia. In addition, the principal component analysis (PCA) suggested that domestic donkeys are divided into three main clusters on the nuclear level, i.e., a Tropical Africa cluster, a North Africa and Eurasia cluster and an Australia cluster. Wang et al. (2020) finally found that domestic donkeys showed reduced levels of Y chromosome variability, which might indicate a discordance of paternal and maternal histories of donkeys, similar to the domestic horse (Lindgren et al., 2004; Lippold et al., 2011).

The history of domestic donkey in China dates back more than 4,000 years (Zheng, 1980; Xie, 1987; Chen et al., 2010). According to literature records, domestic donkeys were bred in present-day Shache County, Xinjiang Uygur Autonomous Region, Northwestern China as early as in the Yin and Shang Dynasties (1,300–1,046 BC) (Yang and Hong, 1989). Regarding the origin of the Chinese domestic donkey, there are two main views: 1) Due to morphological similarities to Asian wild asses, e.g., in fur color, some researchers believed that Chinese domestic donkeys might have originated from Mongolian wild ass (Equus hemionus) (Xie, 1987; Liu et al., 2010). 2) In contrast, genetic studies suggested that Chinese domestic donkeys originate from African wild asses (Sun et al., 2007; Han et al., 2014; Ma et al., 2020). Wang et al. (2020) analyzed mitochondrial DNA and nuclear genomes of Chinese local donkey breeds and revealed that Chinese donkeys are closer to African wild asses than to Asian wild asses (E. hemionus and Equus kiang). So far, most molecular studies on Chinese domestic donkey focus on modern specimens. The only public report on genetic analyses of Chinese ancient donkeys has been presented by Han et al. (2014), but only mitochondrial DNA D-loop and cytochrome b gene fragments were obtained, with the dates of the analyzed samples ranging between 1,200–550 years before present (BP). Han et al. (2014) found that the ancient specimens represent both donkey mitochondrial maternal lineages, i.e., the Nubian lineage (clade I) and the lineage of unknown origin (clade II). Unfortunately, due to a lack of
genetic information from earlier Chinese donkeys, we know little about the initial dispersal process of donkeys into China.

In this study, we retrieved three near-complete mitogenomes from archaeological donkey specimens excavated from Midwestern China, investigated the phylogenetic status of the analyzed individuals and estimated the divergence time of the two donkey lineages. We also carried out a Bayesian skyline plot (BSP) analysis to assess donkey population dynamics. Overall, our study provides new insights into the early domestication history of Chinese donkeys.

RESULTS AND DISCUSSION

Three ancient donkey tooth samples are included in this study. Two specimens (SG1 and SG3) were excavated from Gaoling County, Shaanxi Province, China and one sample (LXH1) was collected from Linxia Basin, Gansu Province, China (Figure 1). ^14C dating of the samples was performed by accelerator mass spectrometry (AMS) at the Archaeological Geochronology Laboratory of Peking University (PKUAMS, China). Calibration was done using IntCal13 (Reimer et al., 2013), yielding ages of 2,349-2,301 (SG1), 469-311 (SG3), and 2,160-2,004 (LXH1) cal. BP, respectively. Detailed information on the samples is listed in Supplementary Table S1. Using hybridization capture technology and an E. asinus mitogenome (GenBank No. X97337) as reference, we successfully retrieved three near-complete mitochondrial genomes from the analyzed samples with a mean depth of 79.5-, 37.2- and 36.8-fold, respectively (Supplementary Table S1). Mitochondrial DNA (mtDNA) fragments show damage patterns characteristic for ancient DNA (Briggs et al., 2007) (Supplementary Figure S1), supporting the obtained sequences as derived from authentic ancient DNA.
Early Dispersal of Domestic Donkey to Ancient China

We reconstructed phylogenetic trees using these newly obtained mitogenomes together with Equus sequences from GenBank. Both maximum-likelihood (ML) and Bayesian methods strongly support that all E. asinus individuals form a separate clade within non-caballine horses (Figures 2, 3). The E. asinus branch is further divided into three clades, i.e., one Somali wild ass clade, which diverges from the E. asinus branch first, and two domestic donkey clades (clade I and clade II), containing modern domesticated donkeys, Nubian wild asses and our ancient individuals (Figures 2, 3). Our results are consistent with previous studies (Kimura et al., 2010; Kimura et al., 2013; Han et al., 2014; Ma et al., 2020; Wang et al., 2020). Interestingly, the three samples investigated in this study fall into different clades of domestic donkey, i.e., specimens SG1 and SG3 cluster within clade I (Nubian lineage), while LXH1 groups into clade II (with no extant wild representatives). Both of these two donkey clades are distant from the Asiatic wild asses (E. kiang and E. hemionus), which reveals that the maternal origin of Chinese domestic donkeys is most likely from African wild asses instead of Asian wild asses, as suggested by previous analyses (Han et al., 2014; Ma et al., 2020; Wang et al., 2020), African wild asses are the most likely ancestor of the domestic donkey. It is commonly believed that donkeys first dispersed from Africa to Northwest China through Central Asia about 4,000 years ago (Xie, 1987; Lu et al., 2008). If correct, this means that domestic donkeys had spread into Northwestern China before the establishment of the Han Dynasty (about the second century BC) (Han et al., 2014). After the Southern and Northern Dynasties (420-589 AD), people from Central China also started raising and breeding donkeys, and its population size gradually increased since then (Yang and Hong, 1989).

Two out of three ancient samples in this study have been dated at similar ages (i.e., 2,349-2,301 cal. BP for SG1 and 2,160-2,004 cal. BP for LXH1), yet they fall into different donkey clades (Figures 2, 3). The results demonstrate that the two donkey maternal lineages had been introduced into China at least at the beginning of Han Dynasty, i.e. around the opening of the Silk Road (about the first century BC). Unfortunately, due to a lack of earlier samples, our knowledge about when and how the two donkey maternal lineages were introduced to China is very limited so far, and further research is needed to explore these questions.

Divergence Time of Different E. asinus Lineages

We carried out a mitogenome relaxed molecular clock analysis to investigate the coalescence times among E. asinus lineages, using
root-and-tip dating calibrations in BEAST (Figure 3). Our analysis reveals that the divergence time between Somali wild ass and domestic donkey is at 0.715 Ma (95% CI: 1.169–0.305 Ma), and the split of the two domestic donkey maternal lineages is dated at 0.323 Ma (95% CI: 0.583–0.191 Ma). The times of the most recent common ancestor (TMRCA) of clade I and clade II are 0.185 Ma and 0.145 Ma, respectively.

Beja-Pereira et al. (2004) also estimated the divergence time of the two donkey maternal clades, and suggested a somewhat more ancient divergence in the time range of 0.910–0.303 Ma. Our point estimate (0.323 Ma) is close to the lower limit of that predicted by Beja-Pereira et al. (2004), while the confidence intervals of the two estimates overlap widely. The difference may at least partially be due to the fact that we use near-complete mitochondrial genomes to calculate the divergence time, while only cyt b gene sequences were included in Beja-Pereira et al. (2004). Another possible reason is that different calibration methods are implemented. Beja-Pereira et al. (2004) chose the previously estimated divergence time between horse and donkey (10–8 Ma) as the calibration node (Xu et al., 1996), whereas we considered the 4.5–4.0 Ma from Orlando et al. (2013) for the TMRCA of all extant Equus representatives and the median radiocarbon date or strata age of specimens as calibration points. Although our estimate is younger, our results together with the previous study (Beja-Pereira et al., 2004) suggest that the split of the two donkey lineages dates much earlier than its first known domestication date.

Our estimation of the TMRCA of the donkey clade I and clade II maternal lineages are 0.185 Ma and 0.145 Ma, respectively. This is much younger than the estimates of Kimura et al. (2010), who analyzed mitochondrial D-loop sequences of historic Nubian wild ass, Somali wild ass and ancient donkey, arriving at ages for clade I of 0.406 Ma, clade II of 0.334 Ma and the Somali wild ass clade of 0.360 Ma, respectively. However, even our younger estimates predate the domestication of donkeys by a large margin, suggesting that in both clades multiple wild lineages were incorporated into the domestic gene pool.

Demographic Dynamics of Domestic Donkeys

The Bayesian skyline plot analyses indicate an apparent population expansion between 5,000 and 2,500 years ago for clade I, following by a relatively stable population to the present day (Figure 4). However, compared to clade I, clade II keeps a relatively stable population size overall, only showing a slight population increase during the past 5,000 years, which is similar to the result obtained by Ma et al. (2020).

Domestication of animals is generally accompanied by population expansion, as seen e.g. in horse (Fages et al., 2019), goat (Al-Araimi et al., 2017) and camel (Chen et al., 2019). The donkey population expansions of clade I and clade II may also relate to their domestication. If the Nubian lineage (clade I) and the lineage of unknown origin (clade II) were domesticated simultaneously, a similar demographic history may be expected from them. Ma et al. (2020) also assessed the population dynamics of the two domestic donkey lineages based on modern donkey mitogenomes. Their analyses suggested that clade II had a constant effective population size during most of its history, while clade I experienced a rapid population expansion starting approximately 8,000 years ago. Our estimates are overall similar to the estimates by Ma et al. (2020). In addition, Wang et al. (2020) found that there were no obvious differences in effective population size of Tropical African donkeys and North African and Eurasian donkeys, proposing that these donkeys were probably derived from the domestication of one common ancestral group. However, they noted that their analyses did not allow determining whether donkeys were domesticated at a single or multiple locations. Thus, currently, the history of donkey domestication remains at least partially unresolved. Therefore, ancient DNA is key to explore this essential question, as shown for other domesticated species such as goat (Daly et al., 2018) or cattle (Verdugo et al., 2019). Our estimates confirm that the two donkey lineages experienced somewhat different past demographic expansion histories. Together with the split time of the two clades, our
results at least suggest that donkeys might have undergone at least two independent domestication events.

METHODS

DNA Extraction
We performed DNA extraction in a dedicated ancient DNA laboratory at the University of Potsdam, following the protocol of Dabney et al. (2013) with several modifications as described in Yuan et al. (2020). The tooth samples were ground into fine powder with mortar and pestle, and for each sample about 50 mg powder was added to 1 ml extraction buffer containing 0.45 M EDTA and 0.25 mg/ml proteinase K. The tooth powder was resuspended by vortexing and incubated overnight at 37°C under constant rotation. Next, we centrifuged the samples for 2 min at 13,300 rpm to pellet the powder, followed by adding the supernatant to 13 ml binding buffer. Then the mixtures were poured into the binding apparatus reservoirs, followed by centrifugation for 4 min at 1,500 rpm. We added 650 μL PE buffer to the silica membrane in the washing step and then carried out centrifugation again at 1,500 rpm for 4 min. DNA was eluted twice by adding 12.5 μL TET buffer each time to the silica membrane, incubating for 10 min at room temperature and centrifugation at 13,300 rpm for 30 s each time. In total, we obtained 25 μL DNA extract. In addition, an extraction blank was included alongside the samples.

Library Construction and Hybridization Capture
Single-stranded DNA libraries were prepared by using 20 μL DNA extract for each sample, following the protocol described by Gansauge and Meyer (2013) with the modifications in Yuan et al. (2020). The amount of Circligase II was reduced to 2 μL (100 U/μL) in the ligation step of the first adapter; accordingly, incubation time was increased to overnight at 60°C. Hybridization capture of the complete mitochondrial genome was carried out following previously published procedures (González-Fortes and Paijmans, 2019). Baits were prepared as in the following protocol. First, total DNA was extracted from a modern horse sample and the mitochondrial genome was amplified using four overlapping long range PCR (LR-PCR) primer pairs (Vilstrup et al., 2013; Yuan et al., 2019). Second, LR-PCR products were sheared, blunt-end repaired and ligated to biotinylated adapters. Subsequently, two rounds of hybridization capture were carried out to improve the enrichment rate as described in Yuan et al. (2019). The enriched libraries were purified using Minelute columns (Qiagen) and DNA was eluted twice by adding 10 μL EB buffer each time. Concentration and fragment size of the DNA were measured on a Qubit 2.0 and a TapeStation 2200 (Agilent). Finally, the enriched libraries were pooled and sequenced on 75 cycle single-end runs on the Illumina NextSeq 500 sequencing platform, following the procedures described in Paijmans et al. (2017). Blanks were also included in single-stranded library preparation and hybridization capture procedures to monitor potential contamination.

Data Analysis
Sequencing reads were processed as follows: 3’ adapter sequences were removed from raw reads by using cutadapt v1.4.2 (Martin, 2011), and reads shorter than 30 bp were discarded. The trimmed reads were mapped to a complete mitochondrial genome of E. asinus (GenBank No. X97337) using the “aln” algorithm in Burrows-Wheeler aligner (BWA) (Li and Durbin, 2010) with default parameters, and converted to bam format using the “samview” algorithm in BWA. Next, reads with a MapQuality score less than 30 and PCR duplicates were removed by using “view” and “rmdup” in samtools v0.1.9 (Li et al., 2009). Finally, a mitochondrial consensus sequence was generated in Geneious (https://www.geneious.com/), called with a minimum coverage of 2 and a base agreement greater than 75%.

Bioinformatics Analysis
To reconstruct the phylogenetic relationships and investigate the phylogenetic status of the analyzed samples among the E. asinus, the three newly obtained near-complete mitochondrial genomes were aligned with 95 Equus mitochondrial genomes from GenBank, including E. asinus, E. kiang, E. hemionus, E. hydruntinus, E. ovodovi, E. zebra, E. greysi, E. burchellii, and E. caballus using MAFFT v7.471 (Katoh et al., 2002) on the CIPRES portal (Miller et al., 2010). The ambiguous section of the D-loop was discarded and the length of the final alignment was 16,621 bp. The substitution model GTR + G for each section was selected and the data set was divided into seven partitions (Supplementary File S1) using PartitionFinder v2.1.1 (Lanfear et al., 2016). We conducted a maximum-likelihood analysis using 1,000 bootstrap replicates in RAxML-HPC v8.2.12 (Stamatakis, 2014) with E. caballus as outgroup.

In addition, in order to estimate the divergence time of the two main donkey maternal lineages, we also performed a Bayesian analysis in BEAST v1.8.2 (Drummond et al., 2012) using the same partitioning as above. The phylogenetic tree was calibrated by root-tip dating, using the median calibrated radiocarbon ages or stratigraphic ages for all sequences (Supplementary Table S2), and assuming a most recent common ancestor (TM RCA) of all equids of 4.0–4.5 million years ago (Ma) (Orlando et al., 2013) as calibration points. We selected constant coalescent to provide the prior distribution for the branch lengths. The GTR + G substitution model was used, running 100,000,000 generations and sampling every 10,000 steps. The first 50,000,000 samples for each chain were discarded as burn-in. The result was analyzed with Tracer v1.7 (Rambaut et al., 2018) to check effective sample size for each model parameter. A Maximum Clade Credibility tree was calculated using TreeAnnotator v1.5.4 (Drummond et al., 2012) and viewed in FigTree v1.4.4 (http://tree.bio.ed.ac.uk/software/figtree). Moreover, the donkey female effective population size changes through time (Supplementary Table S2) were estimated using the Bayesian skyline plot (BSP) analysis in Tracer v1.7 (Rambaut et al., 2018).
DATA AVAILABILITY STATEMENT

The original contributions presented in the study are publicly available in NCBI using accession numbers MZ823384, MZ823385, and MZ823386. Our data can be freely downloaded from NCBI after October 11th 2021.

ETHICS STATEMENT

The animal study was reviewed and approved by Shaanxi Provincial Institute of Archaeology, Xi’an 710054, China; Key Laboratory of Vertebrate Evolution and Human Origins of Chinese Academy of Sciences, IVPP, Beijing 100044, China.

AUTHOR CONTRIBUTIONS

JY, XL, and MH conceived the project. SH and TD collected the samples and performed morphological analyses. GSH and AB guided the experimental and bioinformatics analyses. JY, MP, and UT performed laboratory work. LW, JH, BX, GSU, SS, and XH supported data analyses. LW, GSH, MH, and JY wrote the paper. All authors read and gave comments to the final version of the manuscript.

REFERENCES

Al-Araimi, N. A., Gaafar, O. M., Costa, V., Neira, A. L., Al-Atiyat, R. M., and Beja-Pereira, A. (2017). Genetic Origin of Goat Populations in Oman Revealed by Mitochondrial DNA Analysis. PLoS ONE 12, e0190235. doi:10.1371/journal.pone.0190235

Bai, D., Zhao, Y., Bei, L., Gerelchimeg, B., Zhang, X., and Dugar-Javiin, M. (2017). Progress in the Whole Genome of Equus by Using High-Throughput Sequencing Technologies. Hereditas 39, 974–983. doi:10.16288/j.yczz.17-122

Beja-Pereira, A., England, P. R., Ferrand, N., Jordan, S., Bakhiet, A. O., Abdalla, M. A., et al. (2004). African Origins of the Domestic Donkey. Science 304, 1781. doi:10.1126/science.1096008

Blench, R. M. (2000). A History of Donkeys, Wild Asses and Mules in Africa. London (UK): University College London Press.

Briggs, A. W., Stenzel, U., Johnson, P. L. F., Green, R. E., Kelso, J., Prufer, K., et al. (2007). Patterns of Damage in Genomic DNA Sequences from a Neandertal. Proc. Natl. Acad. Sci. 104, 14616–14621. doi:10.1073/pnas.0704665104

Chen, J., Sun, Y., Manglai, D., Min, L., and Pan, Q. (2010). Maternal Genetic Diversity and Population Structure of Four Chinese Donkey Breeds. Livestock Sci. 131, 272–280. doi:10.1016/j.livsci.2010.04.012

Chen, S., Li, J., Zhang, F., Xiao, B., Hu, J., Cui, Y., et al. (2019). Different Maternal Lineages Revealed by Ancient Mitochondrial Genome of Camelus bactrianus from China. Mitochondrial DNA A 30, 786–793. doi:10.1080/24701394.2019.1659250

Clutton-Brock, J. (1992). Horse Power: A History of the Horse and the Donkey in Human Societies. London (UK): Harvard University Press and the Natural History Museum.

Dabney, K., Knapp, M., Glocke, I., Gansauge, M.-T., Weihmann, A., Nickel, B., et al. (2013). Complete Mitochondrial Genome Sequence of a Middle Pleistocene Cave bear Reconstructed from Ultrashort DNA Fragments. Proc. Natl. Acad. Sci. 110, 15758–15763. doi:10.1073/pnas.1314451110

Daly, K. G., Maisano Delser, P., Mullin, V. E., Scheu, A., Mattiangeli, V., Teasdale, M. D., et al. (2018). Ancient Goat Genomes Reveal Mosaic Domestication in the Fertile Crescent. Science 361, 85–88. doi:10.1126/science.aas9411

Denbow, J., Shaw, T., Sinclair, P., Andah, B., and Okpoko, A. (1993). The Archaeology of Africa: Food, Metals and Towns. J. Field Archaeol. 23, 256. doi:10.2307/530508

FUNDING

This work was supported by the National Natural Science Foundation of China (Nos. 41472014; 42172027) and the “PPP” project jointly funded by China Scholarship Council and DAAD (No. 2016–6041).

ACKNOWLEDGMENTS

We are grateful to Dr. Stefanie Hartmann at the University of Potsdam, Dr. Johanna Pajimans at the University of Leicester, and Dr. Michael V. Westbury at the University of Copenhagen for their help in the process of this research.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene.2021.759831/full#supplementary-material

Denham, T., and Iriarte, J. (2007). Rethinking Agriculture:Archaeological and Ethnoarchaeological Perspectives. Walnut Creek, CA: Left Coast Press.

Drummond, A. J., Suchard, M. A., Xie, D., and Rambaut, A. (2012). Bayesian Phylogenetics with BEAUti and the BEAST 1.7. Mol. Biol. Evol. 29, 1969–1973. doi:10.1093/molbev msq075

Epstein, H. (1971). The Origin of the Domestic Animals of Africa. New York (USA): African Publishing Corporation.

Fages, A., Hanghøj, K., Khan, N., Gauntz, C., Seguin-Orlando, A., Leonard, M., et al. (2019). Tracking Five Millennia of Horse Management with Extensive Ancient Genome Time Series. Cell 177, 1419–1435. doi:10.1016/j.cell.2019.03.049

Gansauge, M., and Meyer, M. (2013). Single-stranded DNA Library Preparation for the Sequencing of Ancient or Damaged DNA. Nat. Protoc. 8, 737–748. doi:10.1038/nprot.2013.038

González-Fortes, G., and Pajimans, J. L. A. (2019). “Whole-Genome Capture of Ancient DNA Using Homemade Baits,” in Ancient DNA: Methods and Protocols (New York (USA): Humana Press), 93–105. doi:10.1007/978-1-4939-9176-1_11

Han, L., Zhu, S., Ning, C., Cai, D., Wang, K., Chen, Q., et al. (2014). Ancient DNA Provides New Insight into the Maternal Lineages and Domestication of Chinese Donkeys. BMC Evol. Biol. 14, 246. doi:10.1186/s12862-014-0246-4

Ivanovics, A., Kavar, T., Caput, P., Mioc, B., Pavic, V., and Dow, P. (2002). Genetic Diversity of Three Donkey Populations in the Croatian Coastal Region. Anim. Genet. 33, 169–177. doi:10.1046/j.1365-2052.2002.00879.x

Katoh, K., Misawa, K., Kuma, K., and Miyata, T. (2002). MAFFT: A Novel Method for Rapid Multiple Sequence Alignment Based on Fast Fourier Transform. Nucleic Acids Res. 30, 3059–3066. doi:10.1093/nar/gkf436

Kimura, B., Marshall, F., Chen, S., Rosenbom, S., Moehlman, P. D., Tuross, N., et al. (2010). Ancient DNA from Nubian and Somali Wild Ass Provides Insights into Donkey Ancestry and Domestication. Proc. R. Soc. B. 278, 50–57. doi:10.1098/rspb.2010.0708

Kimura, B., Marshall, F., Beja-Pereira, A., and Mulligan, C. (2013). Donkey Domestication. Afr. Archaeol. Rev. 30, 83–95. doi:10.1080/14004510.1216–9126–8

Lanfear, R., Frandsen, P. B., Wright, A. M., Senfeld, T., and Calcott, B. (2016). PartitionFinder 2: New Methods for Selecting Partitioned Models of Evolution for Molecular and Morphological Phylogenetic Analyses. Mol. Biol. Evol. 34, ms260–773. doi:10.1093/molbev/msw260

Frontiers in Genetics | www.frontiersin.org 7 October 2021 | Volume 12 | Article 759831
Li, H., and Durbin, R. (2010). Fast and Accurate Long-Read Alignment with Burrows-Wheeler Transform. *Bioinformatics* **26**, 589–595. doi:10.1093/bioinformatics/btp698

Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., et al. (2009). The Sequence Alignment/map Format and Samtools. *Bioinformatics* **25**, 2078–2079. doi:10.1093/bioinformatics/btp532

Lindgren, G., Backström, N., Swinburne, J., Hellborg, L., Einarsson, A., Sandberg, K., et al. (2004). Limited Number of Patrilines in Horse Domestication. *Nat. Genet.* **36**, 335–336. doi:10.1038/ng1326

Lippold, S., Knapp, M., Kuznetsova, T., Leonard, J. A., Benecke, N., Ludwig, A., et al. (2011). Discovery of Lost Diversity of Paternal Horse Lineages Using Ancient DNA. *Nat. Commun.* **2**, 450. doi:10.1038/ncomms1447

Liu, J., Yang, B., Xia, L., Sun, X., Guo, J., and Cheng, S. (2010). Study on Some Fragment Analysis of mtDNA D-loop and Phylogenesis for Nine Domestic Donkey Breeds in China. *Chinese J. Anim. Sci. (Chinese Version)* **46**, 1–6.

Lu, C., Xie, W., Su, R., Ge, Q., Chen, H., Shen, S., et al. (2008). African Origin of Chinese Domestic Donkeys. *Hereditas (Beijing)* **30**, 324–328. doi:10.3724/ sp.j.1005.2008.00324

Ma, X. Y., Ning, T., Adeola, A. C., Li, J., Esmailizadeh, A., Lichoti, J. K., et al. (2020). Potential Dual Expansion of Domesticated Donkeys Revealed by Worldwide Analysis on Mitochondrial Sequences. *Zool. Res.* **41**, 51–60. doi:10.24272/zrs.2095-8137.2020.007

Martin, M. (2011). Cutadapt Removes Adapter Sequences from High-Throughput Sequencing Reads. *EMBnet j.* **17**, 10–12. doi:10.14806/ej.17.1.200

Miller, M. A., Pfeiffer, W., and Schwartz, T. (2010). “Creating the CIPRES Science Gateway for Inference of Large Phylogenetic Trees,” in 2010 Gateway Computing Environments Workshop (GCE), December 14, 2010. doi:10.1109/gce.2010.5676129

Orlando, L., Ginolhac, A., Zhang, G., Froese, D., Albrechtsen, A., Stiller, M., et al. (2013). Recalibrating Equus Evolution Using the Genome Sequence of an Early Middle Pleistocene Horse. *Nature* **499**, 74–78. doi:10.1038/nature12323

Pajmajš, J. I. A., Báleka, S., Henneberger, K., Taron, U. H., Trinks, A., Westbury, M. V., et al. (2017). Sequencing Single-Stranded Libraries on the Illumina NextSeq 500 Platform. *arXiv. arxiv.1711.11004.*

Peters, J., and Driesch, A. V. d. (1997). The Two-Humped Camel (*Caminus bactrianus*): New Light on its Distribution, Management and Medical Treatment in the Past. *J. Zool.* **242**, 651–679. doi:10.1111/j.1469-7998.1997.tb05819.x

Rambaut, A., Drummond, A. J., Xie, D., Baele, G., and Suchard, M. A. (2018). Posterior Summarization in Bayesian Phylogenetics Using Tracer 1.7. *Syst. Biol.* **67**, 901–904. doi:10.1093/sysbio/syy032

Reimer, P. J., Bard, E., Bayliss, A., Beck, J. W., Blackwell, P. G., Ramsey, C. B., et al. (2013). IntCal13 and marine13 Radiocarbon Age Calibration Curves 0–50,000 Years Cal BP. *Radiocarbon* **55**, 1869–1887. doi:10.2458/azu_js_rc.55.16947

Rossel, S., Marshall, F., Peters, J., Pilgram, T., Adams, M. D., and O’Connor, D. (2008). Domestication of the Donkey: Timing, Processes, and Indicators. *Proc. Natl. Acad. Sci.* **105**, 3715–3720. doi:10.1073/pnas.07096292105

Smith, D. G., and Pearson, R. A. (2005). A Review of the Factors Affecting the Survival of Donkeys in Semi-arid Regions of Sub-saharan Africa. *Trop. Anim. Health Prod.* **37**, 1–19. doi:10.1007/s11250-005-9002-5

Stamatakis, A. (2014). RAxML Version 8: a Tool for Phylogenetic Analysis and post-analysis of Large Phylogenies. *Bioinformatics* **30**, 1312–1313. doi:10.1093/bioinformatics/btu033

Sun, W., Yang, B., and Liang, C. (2007). Phylogenetic Relationship and Genetic Diversity of Chinese Four Domestic Donkeys Using mtDNA D-Loop. *China Herbivores (Chinese Version)* **27**, 7–10.

Verdugo, M. P., Mullin, V. E., Scheu, A., Mattiagelli, V., Daly, K. G., Maisano Delser, P., et al. (2019). Ancient Cattle Genomics, Origins, and Rapid Turnover in the Fertile Crescent. *Science* **365**, 173–176. doi:10.1126/science.aav1002

Vilstrup, J. T., Seguin-Orlando, A., Stillier, M., Ginolhac, A., Raghavan, M., Nielsen, S. C. A., et al. (2013). Mitochondrial Phylogenetics of Modern and Ancient Equids. *PLoS ONE* **8**, e59590. doi:10.1371/journal.pone.0059590

Wang, C., Li, H., Guo, Y., Huang, J., Sun, Y., Min, J., et al. (2020). Donkey Genomes Provide New Insights into Domestication and Selection for Coat Color. *Nat. Commun.* **11**, 6014. doi:10.1038/s41467-020-19813-7

Xie, C. (1987). *Horse and Ass Breeds in China*. Shanghai (China): Shanghai Scientific and Technical Publishing House.

Xu, X., Gullberg, A., and Arnason, U. (1996). The Complete Mitochondrial DNA (mtDNA) of the Donkey and mtDNA Comparisons among Four Closely Related Mammalian Species-pairs. *J. Mol. Evol.* **43**, 438–446. doi:10.1007/BF02337515

Yang, Z., and Hong, Z. (1989). Geographic and Population Ecology of Donkey in China. *J. Ecol. (Chinese Version)* **8**, 40–42.

Yuan, J., Hou, X., Barlow, A., Preick, M., Taron, U. H., Alberti, F., et al. (2019). Molecular Identification of Late and Terminal Pleistocene *Equus Ovodovi* from Northeastern China. *PLoS ONE* **14**, e0216883. doi:10.1371/journal.pone.0216883

Yuan, J., Sheng, G., Preick, M., Sun, B., Hou, X., Chen, S., et al. (2020). Mitochondrial Genomes of Late Pleistocene Caballine Horses from China Belong to a Separate Clade. *Quat. Sci. Rev.* **250**, 106691. doi:10.1016/j.quascirev.2020.106691

Zheng, P. (1980). *Donkeys in Livestock Breeds in China and Their Ecological Characteristics*. Beijing (China): Agriculture Press.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Wang, Sheng, Preick, Hu, Deng, Taron, Barlow, Hu, Xiao, Sun, Song, Hou, Lai, Hofreiter and Yuan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.