8. Killick-Kendrick R, Killick-Kendrick M, Tang Y. Anthroponotic cutaneous leishmaniasis in Kabul, Afghanistan: the low susceptibility of Phlebotomus papatasi to Leishmania tropica. Trans R Soc Trop Med Hyg. 1994;88:252–3. http://dx.doi.org/10.1016/0035-9203(94)90320-4

9. Khan NH, Bari AU, Hashim R, Khan I, Munee A, Shah A, et al. Cutaneous leishmaniasis in Khyber Pakhtunkhwa province of Pakistan: clinical diversity and species-level diagnosis. Am J Trop Med Hyg. 2016;95:1106–14. http://dx.doi.org/10.4269/ajtmh.16-0343

10. van Thiel P-P, Leenstra T, de Vries HJ, van der Sluis A, van Gool T, Knull AC, et al. Cutaneous leishmaniasis (Leishmania major infection) in Dutch troops deployed in northern Afghanistan: epidemiology, clinical aspects, and treatment. Am J Trop Med Hyg. 2010;83:1295–300. http://dx.doi.org/10.4269/ajtmh.2010.10-0143

Address for correspondence: Shahzad Munir, Yunnan Agricultural University, Kunming 650201, China; email: shazid_10@yahoo.com

Ocular Vaccinia Infection in Dairy Worker, Brazil

Mauricio Teixeira Lima, Grazielle Pereira Oliveira, Felipe Lopes Assis, Danilo Bretas de Oliveira, Sidiner Mesquita Vaz, Giliane de Souza Trindade, Jônatas Santos Abrahão, Erna Geessien Kroon

Author affiliation: Universidade Federal de Minas Gerais, Belo Horizonte, Brazil (M.T. Lima, G.P. Oliveira, F.L. Assis, D.B. de Oliveira, G. de Souza Trindade, J.S. Abrahão, E.G. Kroon); Casa de Caridade de Carangola, Carangola, Brazil (S.M. Vaz); Universidade Federal dos Vales do Jequitinhonha e Mucuri, Diamantina, Diamantina, Brazil (D.B. de Oliveira)

DOI: https://doi.org/10.3201/eid2401.170430

We studied a clinical case of vaccinia virus that caused an ocular manifestation in a dairy worker in Brazil. Biologic and molecular analyses identified a co-infection with 2 isolates from different Brazilian vaccinia virus phylogenetic groups.

Detachment of co-infections for various viral pathogens recently has increased (1,2). However, the presence of ≥2 viral etiologic agents often is not considered (1). The range of pathogens that can present co-infections and the association of these infections with the occurrence and severity of disease remain unclear (1,2). Vaccinia virus (VACV), the prototype virus of the genus Orthopoxvirus, has been associated with exanthematic outbreaks in Asia and South America that affect mainly dairy cattle and rural workers (3,4). In Brazil, several Brazilian VACV (VACV-BR) have been isolated and characterized biologically and phylogenetically. These studies demonstrated that circulating viruses belonged to at least 2 genetic clusters (4–8). Previous studies have demonstrated the co-circulation of distinct VACV isolates during the same outbreak and VACV co-infecting horses and cattle (6–8).

We obtained 2 distinct VACV isolates from the same clinical sample from 1 eye of a rural worker. Our data show that the eye was co-infected with 2 VACV and demonstrates the detection and isolation of VACV from a natural case of ocular vaccinia infection.

In September 2015, an unvaccinated 45-year-old man who worked on a farm in Carangola County, Minas Gerais State, Brazil (20°44’06”S, 42°01’52”W), showed development of typical manifestations of vaccinia infection, including fever and painful vesiculopustular lesions (online Technical Appendix Figure, panel A, https://wwwnc.cdc.gov/EID/article/24/1/17-0430-Techapp1.pdf). He had lesions on the left hand, right arm, and nose and an atypical manifestation in the left eye with aches in the ocular globe and periorbital region. The clinical condition progressed to major visual acuity losses in the affected eye. He reported recent contact with sick cows on the farm during milking.

Dried swab specimens from his lesions were soaked in 200 µL phosphate-buffered saline containing amphotericin B (4 µg/mL), penicillin (200 U/mL), and streptomycin (100 µg/mL); homogenized; and centrifuged at 3,000 g for 5 min (4). The supernatants were used for molecular diagnosis using orthopoxvirus-specific PCR that targeted the C11R gene, which encodes viral growth factor, and the A56R gene, which encodes viral hemagglutinin protein (4). All samples were positive for both orthopoxvirus targets.

Vero cells were cultured in 25-cm² culture flasks and infected with the specimen supernatants to isolate the virus at 37°C until a cytopathic effect was detected (4). VACV was isolated from the hand, nose, and eye samples. These isolates were tested for their plaque phenotypes in BSC-40 cells incubated at 37°C for 48 h (4), which demonstrated the presence of at least 2 types of viral populations comprising small and large plaques in an estimated ratio 2:1. Two viral plaques (1 forming large and 1 forming small plaques) were obtained from the eye sample after 3 additional rounds of plaque purification in BSC-40 cells (online Technical Appendix Figure, panel B) (4). The viral plaques were propagated and titrated by plaque assay in Vero cells, and their DNA was extracted (4).

We obtained the complete genomes using the Illumina MiSeq instrument (Illumina, San Diego, CA, USA) with the paired-end application. The sequence reads were assembled de novo using ABYSS software (http://www.bcgsc.ca/platform/bioinfo/software/abyss), and the resulting contigs were ordered by the python-based CONTIGuator.py software (http://contiguator.sourceforge.net). The GenBank accession numbers are MG012795 (small) and MG012796 (large).
Analysis of the complete genome revealed 92% similarity between the 2 isolates, and some genes confirm a remarkable variability (online Technical Appendix Figure, panel C). We constructed a phylogenetic tree (online Technical Appendix Figure, panel D) using the A56R gene sequence by the maximum-likelihood method and 1,000 bootstrap replicates in MEGA 6.02 (http://www.megasoftware.net). The analysis demonstrated a co-infection with viruses from both VACV-BR groups, such that the large-plaque clone clustered with group 2 VACV-BR isolates and the small-plaque clone clustered with group 1 VACV-BR isolates. We named these isolates Carangola eye virus 1 (small) and Carangola eye virus 2 (large).

Our study demonstrated the genetic and phenotypic variability between 2 viruses isolated from the same sample in a natural human co-infection with VACV. The viruses belong to 2 distinct VACV-BR groups, reinforcing and expanding previous work with other hosts (6–8). These results raise new questions about how co-infections with these viruses might change the aspects of an infection and its signs and symptoms, such as development of ocular vaccinia. Although cases of ocular vaccinia have been reported after vaccination and accidental laboratory infection (9,10), we proved the association and isolate VACV samples from a natural ocular vaccinia infection. The effort to understand singular aspects of VACV-BR co-infections should be increased, and further molecular and biologic characterizations of these samples should be conducted to identify and better understand the natural dynamics and signs and symptoms caused by VACV-BR.

About the Author
Mr. Lima is a PhD candidate at the Laboratório de Vírus, Departamento de Microbiologia, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais. His primary research interest is the poxviruses.

References
1. Stefanska I, Romanowska M, Donevski S, Gawryluk D, Brydak LB. Co-infections with influenza and other respiratory viruses. Adv Exp Med Biol. 2013;756:291–301. http://dx.doi.org/10.1007/978-94-007-4549-0_36
2. Griffiths EC, Pedersen AB, Fenton A, Petchev OL. The nature and consequences of coinfection in humans. J Infect. 2011;63:200–6. http://dx.doi.org/10.1016/j.jinf.2011.06.005
3. Damon IK. Poxviruses. In: Knipe DM, Howley PM, Griffin DE, Lamb RA, Martin MA, Roizman B, et al., editors. Fields virology. Vol II. 5th ed. Philadelphia: Lippincott Williams and Wilkins; 2007. p. 2947–75.
4. Kroon E, Santos Abrahão J, de Souza Trindade G, Pereira Oliveira G, Moreira Franco Luiz AP, Barbosa Costa G, et al. Natural vaccinia virus infection: diagnosis, isolation, and characterization.Curr Protoc Microbiol. 2016;42:14A.5.1–14A.5.43. http://dx.doi.org/10.1002/cpmt.13
5. Damaso CR, Exposito JI, Condit RC, Moussatché N. An emergent poxvirus from humans and cattle in Rio de Janeiro State: Cantagalo virus may derive from Brazilian smallpox vaccine. Virology. 2000;277:439–49. http://dx.doi.org/10.1006/viro.2000.0603
6. Trindade GS, Lobato ZI, Drumond BP, Leite JA, Trigueiro RC, Guedes MI, et al. Short report: isolation of two vaccinia virus strains from a single bovine vaccinia outbreak in rural area from Brazil: implications on the emergence of zoonotic orthopoxviruses. Am J Trop Med Hyg. 2006;75:486–90.
7. Campos RK, Brum MC, Nogueira CE, Drumond BP, Alves PA, Siqueira-Lima L, et al. Assessing the variability of Brazilian vaccinia virus isolates from a horse exanthematic lesion: coinfection with distinct viruses. Arch Virol. 2011;156:275–83. http://dx.doi.org/10.1007/s00705-010-0857-z
8. Oliveira G, Assis F, Almeida G, Albarnaz J, Lima M, Andrade AC, et al. From lesions to viral clones: biological and molecular diversity amongst autochthonous Brazilian vaccinia virus. Viruses. 2015;7:1218–37. http://dx.doi.org/10.3390/v7031218
9. Lewis FMT, Chernak E, Goldman E, Li Y, Karem K, Damon IK, et al. Ocular vaccinia infection in laboratory worker, Philadelphia, 2004. Emerg Infect Dis. 2006;12:134–7. http://dx.doi.org/10.3201/eid1201.051126
10. Hu G, Wang MJ, Miller MJ, Holland GN, Bruckner DA, Civen R, et al. Ocular vaccinia following exposure to a smallpox vaccinee. Am J Ophthalmol. 2004;137:554–6. http://dx.doi.org/10.1016/j.ajo.2003.09.013

Address for correspondence: Maurício Teixeira Lima and Erna Geessien Kroon, Instituto de Ciencias Biologicas, Universidade Federal de Minas Gerais, Dept Microbiologia, Lab de Virus, bloco F4, sala 258, Av. Antonio Carlos 6627, 31270-901 Belo Horizonte, Minas Gerais, Brazil; emails: maurili15@hotmail.com and ermakroon@gmail.com

Estimation of Undiagnosed Naegleria fowleri Primary Amebic Meningoencephalitis, United States

Almea Matanock, Jason M. Mehal, Lindy Liu, Diana M. Blau, Jennifer R. Cope

Author affiliation: Centers for Disease Control and Prevention, Atlanta, Georgia, USA

DOI: https://doi.org/10.3201/eid2401.170545

Primary amebic meningoencephalitis is an acute, rare, typically fatal disease. We used epidemiologic risk factors and multiple cause-of-death mortality data to estimate the number of deaths that fit the typical pattern for primary amebic meningoencephalitis; we estimated an annual average of 16 deaths (8 male, 8 female) in the United States.

1Preliminary results of this study were presented at the Infectious Diseases Society of America Conference; October 8–12, 2014, Philadelphia, Pennsylvania, USA.
Ocular Vaccinia Infection in Dairy Worker

Technical Appendix

Figure. Ocular vaccinia infection in a 45-year-old male dairy farm worker, Brazil, 2015. A) Lesion on the hand. B) Optical microscopy images of plaque phenotypes showing the lysis
plates of Carangola eye viruses 1 and 2 in BSC-40 cell monolayers stained with crystal violet. Original magnification 400×. C) Genome draft of Carangola eye viruses 1 and 2 shown some genes with nucleotide deletions and substitutions that differentiate the isolates. The percentage of identity is shown. D) Phylogenetic analysis using the maximum-likelihood method based on orthopoxvirus hemagglutinin genes. The tree was constructed with 1,000 bootstrap replicates using the Tamura-Nei model in MEGA 6.06 (http://www.megasoftware.net/). Numbers along branches are bootstrap values. Green circles indicate the vaccinia virus strains isolated in this study. Red and blue brackets indicate isolates belonging to groups 1 and 2, respectively, of the Brazilian vaccinia virus taxonomy. Cutoff value is 50%.