METHOD DEVELOPMENT AND VALIDATION FOR MULTI-COMPONENT ANALYSIS OF LAMIVUDINE & TENOFOVIR DISOPROXIL FUMARATE IN BULK DRUG BY UV-VISIBLE SPECTROPHOTOMETER & RP-HPLC
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ABSTRACT
A novel, simple, precise and accurate method developed for the estimation of Lamivudine and tenofovir disoproxil fumarate (TDF) in bulk drug form has been established. Lamivudine and tenofovir are well known drugs and used in treatment of HIV-1. The method was performed by using C18 column, ODS Hypersil column with UV detection at 262nm by using Acetonitrile and water in ratio 55:45. The retention time was found to be 2.8 and 6.8 min for Lamivudine and tenofovir disoproxil fumarate (TDF). The linearity was found in range of 6-14µg/ml for Lamivudine and 10-50µg/ml for Tenofovir disoproxil fumarate with flow rate 1ml/min. The method was validated for linearity, accuracy, precision and robustness as per ICH guidelines. This method is suitable for simultaneous analysis for both the nucleoside analog reverse- transcriptase inhibitors

INTRODUCTION
Tenofovir disoproxil fumarate (Fig-1) and Lamivudine (Fig-2) are widely used anti-retroviral drugs in the categories of NRTIs i.e. nucleotide analogues reverse transcriptase inhibitors [1-4]. These drugs are used for the prevention and clinical management of acquired immune deficiency syndrome (AIDS) with multiple complications [5-8].

Keywords
Lamivudine, Tenofovir
Disoproxil Fumarate, anti-HIV, RP-HPLC

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Tenofovir is a type of anti-HIV medicine called a nucleoside reverse transcriptase inhibitor (NRTI). It is always used in combination with other antiviral agents to treat patients with HIV. It is used in the form of a prodrug as Tenofovir Disoproxil Fumarate [13-15]. Tenofovir is not a cure for HIV infection but decreases the risk of spreading the disease to others. It is also used to treat the certain type of liver infection called chronic hepatitis B infection [16-18]. The literature review revealed that there are several methods available for single component analysis for lamivudine and Tenofovir (TDF) [19-20].

![Figure 1: Tenofovir (TDF)](image1)

![Figure 2: Lamivudine](image2)

**MATERIAL AND METHOD**

**Instrument**
The lambda max and iso-absorptive point were determined by UV–spectrophotometer using Lab India with UV win software. HPLC (Shimadzu) prominence LC 20 AD, manual sampler, software LC solution and detector (UV-visible), Column C-18, Thermo scientific octadecysilane Hypersil (ODS), ultrasonicater, vacuum filter, analytical balance.

**Selection of wavelength**
The selection of wavelength 10 µg/ml concentration of lamivudine and 10 µg/ml concentration of tenofovir was prepared in 55:45 with ACN: water respectively. The result show iso-absorptive point that was observed at 262 nm (figure 3).

![Figure 3. Overlain spectrum of lamivudine and TDF](image3)

**Selection of chromatographic condition**
The isocratic mode with mobile phase Acetonitrile and water in ratio 55:45 with flow rate 1ml/min. The resulting chromatograms were recorded and the chromatographic responses were measured.

**Analytical Method Validation**
A calibration curve was plotted with the concentration range 6-14 µg/ml for lamivudine and 10-50 µg/ml for tenofovir (TDF). The method was developed and validated as per ICH guidelines. The parameters were studied linearity, accuracy, precision (intraday and interday precision and repeatability) and robustness and the amount recovery, percentage recovery and mean recovery for the same was calculated.

**Preparation of standard stock solution**

**Lamivudine standard stock solution**
Standard lamivudine 100 mg was weighed and transferred to a 100 ml clean and dry volumetric flask and dissolved into the HPLC grade sample solution (ACN: Water in the ratio 55:45) then volume was made up to the mark with solution containing 1000 µg/ml conc. Then 10 ml of solution was pipette out and transferred to 100 ml clean and dry volumetric flask, made up its volume with solvent to get 100 µg/ml conc. solutions.

**TDF standard stock solution**
Standard tenofovir (TDF) 100 mg was weighed and transferred to a 100 ml clean and dry volumetric flask and dissolved into the HPLC grade sample solution (ACN: Water in the ratio 55:45) then volume was made up to the mark with solution containing 1000 µg/ml conc. Then 10 ml of solution was pipette out and transferred to 100 ml clean and dry volumetric flask, made up its volume with solvent to get 100 µg/ml conc. solutions.
Chromatographic conditions
The mobile phase consisting of Acetonitrile: water (55:45) was used and absorbance was measured at 262 with the run time 15 min and the flow rate was set at 1.0 ml/min respectively.

Preparation of mobile phase
Mobile phase was prepared by mixing HPLC grade acetonitrile and water in ratio of 55: 45 respectively, and the chromatographic conditions were made for separation of the drugs at the wavelength of 262nm. Degassing is done before the use of mobile phase.

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Validation of the developed method

Linearity curve for lamivudine
Standard lamivudine stock solution the volume of 0.6, 0.8, 1, 1.2, 1.4ml was pipetted out from 100µg/ml and transferred to different 10ml clean and dry volumetric flasks. The volume was made up to the mark having conc. of 6, 8, 10, 12, 14µg/ml respectively. The injection was prepared 20µg/ml and given with run time of 15 minutes. The linearity peaks was found to be within the limits. The results are shown in figure 7 and table 1.

| S. No | Conc. (µg/mL) | Area (µ volt sec.) |
|-------|---------------|--------------------|
| 1     | 6             | 376003             |
| 2     | 8             | 651314             |
| 3     | 10            | 866679             |
| 4     | 12            | 1108256            |
| 5     | 14            | 1357247            |

Figure 7: Linearity curve of lamivudine at 262nm

Accuracy
To study the accuracy, 3 determinants of conc. range of 8, 10, 12µg/ml for lamivudine and 20, 30, 40 µg/ml for tenofovir (TDF0 were prepared having 80%, 100%, and 120% of spiked level respectively. 3 replicates of above conc. were prepared and different 10 ml clean and dry volumetric flasks. The volume was made up to the mark having conc. of 6, 8, 10, 12, 14µg/ml respectively. The injection was prepared 20µg/ml and given with run time of 15 minutes. The linearity peaks was found to be within the limits. The results are shown in figure 7 and table 1.

Table: 1. Results of linearity curve of Lamivudine at wavelength 262 nm.

| S. No | Conc. (µg/mL) | Area (µ volt sec.) |
|-------|---------------|--------------------|
| 1     | 6             | 376003             |
| 2     | 8             | 651314             |
| 3     | 10            | 866679             |
| 4     | 12            | 1108256            |
| 5     | 14            | 1357247            |

Figure 7: Linearity curve of lamivudine at 262nm
responses were obtained. Percent recovery was calculated for obtained data and calculated according to ICH guidelines (Table 3 and 4).

**Table 2. Result of linearity curve of tenofovir (TDF) at 262nm**

| S. No | Conc. (µg/mL) | Area (µ volt sec.) |
|-------|----------------|--------------------|
| 1     | 10             | 427228             |
| 2     | 20             | 733996             |
| 3     | 30             | 1101386            |
| 4     | 40             | 1481526            |
| 5     | 50             | 1916508            |

**Drug recovery**

\[
\begin{align*}
80\% &= \left( \frac{\text{Mean abs. of 180\% fortified sample} - \text{mean abs. of 80\% unfortified sample}}{\text{Mean abs. of fortified standard solution of 100\% test conc.}} \right) \times 100 \\
100\% &= \left( \frac{\text{Mean abs. of 200\% fortified sample} - \text{Mean abs. of 100\% unfortified sample}}{\text{Mean abs. of fortified standard solution of 100\% test conc.}} \right) \times 100 \\
120\% &= \left( \frac{\text{Mean abs. of 220\% fortified sample} - \text{Mean abs. of 120\% unfortified sample}}{\text{Mean abs. of fortified standard solution of 120\% test conc.}} \right) \times 100
\end{align*}
\]

**Table 3: % Drug Recovery of lamivudine at wavelength 262nm**

| S. No | Unfortified sample | Fortified sample | % Recovery |
|-------|---------------------|------------------|------------|
|       | Conc. (µg/ml) | Area | Mean | Conc. (µg/ml) | Area | Mean |          |
| 1     | 8             | 651315 | 651313 | 651314 | 1517973 | 1517971 | 1517971 | 99.34       |
| 2     | 10            | 866680 | 866678 | 866679 | 1733398 | 1733400 | 1733396 | 100.02      |
| 3     | 12            | 1108257 | 1108255 | 1108254 | 1974929 | 1974927 | 1974927 | 99.05       |

**Table 4: % Drug recovery of tenofovir (TDF) at wavelength 262nm**

| S. No | Unfortified sample | Fortified sample | % Recovery |
|-------|---------------------|------------------|------------|
|       | Conc. (µg/ml) | Area | Mean | Conc. (µg/ml) | Area | Mean |          |
| 1     | 20             | 733996 | 733976 | 733996 | 1835268 | 1835270 | 1835269 | 99.06       |
| 2     | 30             | 1101386 | 1101389 | 1101387 | 2202799 | 2202797 | 2202797 | 100.0       |
| 3     | 40             | 1481526 | 1481429 | 1481428 | 2582813 | 2582811 | 2582812 | 99.76       |
Precision
The precision was done for interday, intraday and repeatability.

Interday & intraday precision
Interday & intraday precision of conc. 8, 10, 12µg/ml was prepared and data was obtained for lamivudine. Interday & intraday precision of conc. 20, 30, 40µg/ml was prepared and data was obtained for tenofovir (TDF). 3 replicates were prepared for 3 days. The results of lamivudine and tenofovir were shown in table 5 to 7.

Table 5: Intraday precision of lamivudine and tenofovir (TDF) at 262nm

| Drug          | Lamivudine | Tenofovir (TDF) |
|---------------|------------|-----------------|
|               | Conc.      | 8 µg/ml         | 10 µg/ml | 12 µg/ml | 8 µg/ml | 30 µg/ml | 40 µg/ml |
| Area (µ volt sec.) |           | 8 µg/ml         | 10 µg/ml | 12 µg/ml |          | 20 µg/ml |          | 40 µg/ml |
|               | 651314     | 866679          | 1108256 |          | 733996  | 1101386 | 1481526 |
|               | 651294     | 866580          | 1108350 |          | 733954  | 1101285 | 1481429 |
|               | 651229     | 866662          | 1108275 |          | 733889  | 1101324 | 1481494 |
| Mean          | 651279     | 866640.3        | 1108294 |          | 733946.3| 1101332 | 1481483 |
| S D           | 44.440     | 52.937          | 49.702  |          | 53.910  | 50.934  | 49.426  |
| % RSD         | 0.0068     | 0.0061          | 0.0044  |          | 0.007   | 0.004   | 0.003   |

Table 6: Interday precision of lamivudine at 262nm

| Day  | Day 1 | Day 2 | Day 3 |
|------|-------|-------|-------|
| Conc. | 8µg/ml | 10µg/ml | 12µg/ml | 8µg/ml | 10µg/ml | 12µg/ml | 8µg/ml | 10µg/ml | 12µg/ml |
| Area (µ volt sec.) | 651314 | 866679 | 1108256 | 651379 | 866679 | 1108256 | 6512134 | 866679 | 1108256 |
| Mean | 651279 | 866640.3 | 1108294 | 651302.7 | 866638.7 | 1108299 | 651276.3 | 866651 | 1108286 |
| S D | 44.440 | 52.937 | 49.702 | 48.585 | 69.859 | 44.015 | 53.346 | 53.777 | 78.805 |
| % RSD | 0.0068 | 0.0061 | 0.0044 | 0.012 | 0.008 | 0.003 | 0.0069 | 0.0062 | 0.0071 |

Table 7: Interday precision of tenofovir (TDF) at 262nm

| Day  | Day 1 | Day 2 | Day 3 |
|------|-------|-------|-------|
| Conc. | 20µg/ml | 30µg/ml | 40µg/ml | 20µg/ml | 30µg/ml | 40µg/ml | 20µg/ml | 30µg/ml | 40µg/ml |
| Area (µ volt sec.) | 733996 | 1101386 | 1481526 | 733987 | 1101287 | 1481437 | 733996 | 1101386 | 1481432 |
| Mean | 733946.3 | 733946.3 | 1481493 | 733958.3 | 1101419 | 1481482 |
| S D | 53.910 | 53.910 | 49.426 | 6.244 | 54.744 | 48.418 | 61.80885 | 49.81298 | 47.35328 |
| % RSD | 0.007 | 0.007 | 0.003 | 0.0008 | 0.0049 | 0.0032 | 0.0084 | 0.0045 | 0.0031 |

Repeatability
For repeatability determination minimum of 6 determinants were prepared of 20µg/ml conc. and the chromatogram responses were obtained. The results of lamivudine and tenofovir (TDF) were shown in table 8.
Robustness
This method was carried out by changing wavelength and flow rate of mobile phase. The results were shown in table 9 for change in mobile phase and table 10 for change in flow rate.

Table: 9 Robustness of lamivudine & tenofovir (TDF) at wavelength 262±2nm.

| Wavelength | Difference | $R_t$ of Lamivudine (min.) | $R_t$ of Tenofovir (min.) |
|------------|------------|---------------------------|--------------------------|
| 260        | -2         | 2.769                     | 6.883                    |
| 262        | 0          | 2.858                     | 6.881                    |
| 264        | +2         | 2.841                     | 6.888                    |

Change in flow rate of mobile phase

Table: 10 Robustness of lamivudine & tenofovir (TDF) at wavelength 262 nm.

| Flow rate (mL/min.) | Difference | $R_t$ of Lamivudine (min.) | $R_t$ of Tenofovir (min.) |
|---------------------|------------|---------------------------|--------------------------|
| 0.9                 | -0.1       | 2.752                     | 6.780                    |
| 1                   | 0          | 2.858                     | 6.880                    |
| 1.1                 | +0.1       | 2.285                     | 6.898                    |

CONCLUSION
The estimation of lamivudine and Tenofovir (TDF) was done by RP-HPLC. The mobile phase was optizied Acetonitrile: water in the ratio of 55:45% v/v. A C18 column contains octa-decylsilane chemically linked to porous silica particles was used as stationary phase. UV detector was used at 262 nm. The solutions were chromatograph at a constant flow rate of 1 ml/min. The linearity range of lamivudine was found 6-14µg/ml and tenofovir (TDF) were found to be 10-50µg/ml. Linear regression coefficient was not more than 0.999.

The results obtained on the validation parameters met ICH and USP requirements. It can be also inferred that the method found to be simple, accurate, precise and linear. The method was found to be having suitable application in routine laboratory analysis with high degree of accuracy and precision.

Table 11: Summary of developed method

| Parameter                  | Lamivudine | Tenofovir |
|----------------------------|------------|-----------|
| Linear range (µg/ml)       | 6-14       | 10-50     |
| Regression coefficient ($R^2$) | 0.999     | 0.999     |
| % Recovery                 | 99.47      | 99.60     |
| Repeatability (n=6)        | % RSD NMT 2 | % RSD NMT 2 |
| Precision                  | Intraday precision | % RSD NMT 2 | Interday precision |

FINANCIAL ASSISTANCE
Nil

CONFLICT OF INTEREST
The authors declare no conflict of interest.

AUTHOR CONTRIBUTION
Ms. Shweta Sharma performed experiment in the laboratory and collected data. Dr. Amar Deep Ankalgi analyzed and helps to perform the studies in laboratory and recorded observation and make necessary correction in the records. He also helps to design the experimental data and read the manuscript and make all the necessary corrections in manuscript. Miss. Pooja Kaushal analyzed the data and help to reading and drafting manuscript and help in research work. Dr. M.S. Ashawat studies all the records and helps to make necessary correction and approved the manuscript.

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