Les circuits des ventilateurs, l’humidification et la pneumonie associée au ventilateur

RÉSUMÉ : Les aspects techniques de la prise en charge des patients ventilés mécaniquement comprennent ceux qui ont rapport au circuit du ventilateur, à l’humidification et à la pneumonie associée au ventilateur. Les aspects principaux concernant le circuit du ventilateur comprennent les fuites et le volume de compression. Le volume de compression du circuit affecte le volume courant fourni aussi bien que les mesures d’auto-pression expiratoire positive (PEEP) et de la \( P_{CO_2} \) expirée. La résistance générée à travers le circuit du ventilateur contribue à une désynchronisation patient-ventilateur dans les modes assistés de la ventilation mécanique. Une humidification adéquate du gaz inspiré est nécessaire pour prévenir une perte de chaleur et d’humidité. Les méthodes couramment employées pour humidifier l’air inspiré lors de la ventilation mécanique comprennent l’utilisation d’humidificateurs chauffants et de nez artificiels passifs. Les nez artificiels sont moins efficaces que les humidificateurs chauffants et sont mieux adaptés à un usage à court terme. Avec les humidificateurs chauffants, le circuit peut être chauffé pour éviter la formation de condensation. Cependant, quand on utilise des circuits chauffés, on doit faire attention à ne pas faire baisser humidité relative qui entraînera un assèchement des sécrétions dans le tube endotrachéal. Bien que les pneumonies soit une complication de la ventilation mécanique, celles-ci ressortent souvent d’une aspiration des sécrétions pharyngées et sont rarement liées au circuit du ventilateur. Il n’est pas nécessaire de changer le circuit du ventilateur plus d’une fois par semaine pour des raisons de contrôle des infections; l’incidence des pneumonies associées au ventilateur pourrait même être plus élevée lorsque les changements du circuit sont plus fréquents.

**Key Words:** Humidification, Ventilator-associated pneumonia, Ventilator circuits

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Care of mechanically ventilated patients requires attention to both physiological and technical issues. The outcome from mechanical ventilation is often affected by the interface between the patient and the ventilator. To deliver an adequate tidal volume (VT), this interface must be unobstructed, leak-free, and have minimal compliance and compressible volume.

THE VENTILATOR CIRCUIT

The ventilator circuit delivers gas to the patient and conditions the inspired gases by filtering and humidification. Principal issues related to ventilator circuits include leaks (Table 1) and compression volume. Compression volume within the circuit is determined by the volume of the circuit, the compliance (elasticity) of the tubing material and the ventilation pressure. Circuit compression volume does not reach the patient and becomes clinically important with high pressures and low VTs. The volume that leaves the expiration valve of the ventilator includes the expired volume from the patient and the volume of gas compressed in the ventilator circuit. Unless volume is measured directly at the patient’s airway, the expired volume displayed by the ventilator overestimates the patient’s actual VT by the amount of the compressible volume. Some current microprocessor ventilators correct measured volume for circuit compression volume.

Compressible volume can be calculated by multiplying the compression factor by the airway pressure. The compression factor is about 2 to 4 mL/cm H2O for circuits with a humidifier. The delivered VT is the volume leaving the expiration valve minus the compression volume:

\[ VT = V_{exp} - \text{(factor)}(PIP - \text{PEEP}) \]

where \( V_{exp} \) is the volume leaving the expiration valve, VT is tidal volume corrected for compression, PIP is peak inspiratory pressure and PEEP is positive end-expiratory pressure.

Consideration of compression volume is important for several reasons. Most important, it decreases the delivered VT. Failure to consider compression volume results in overestimation of lung compliance. Auto-PEEP measurements are also affected by circuit compression volume, as follows:

\[ \text{auto-PEEP} = [(\text{Crs} + \text{Cpc})/\text{Crs}] \times \text{(estimated auto-PEEP)} \]

where Crs is the compliance of the respiratory system, Cpc is the compliance of the circuit (ie, the compression factor) and estimated auto-PEEP is the value that is measured. Compression volume also affects the measurement of mixed expired carbon dioxide tension (\( P_{\text{E}}CO_2 \)). Because the inspired gas is compressed, it contributes a volume in excess of VT, which dilutes the mixed expired \( PCO_2 \); this may be corrected by applying the following equation:

\[ P_{\text{E}}CO_2 = (P_{\text{exp}}CO_2)(V_{exp}/V_t) \]

where \( P_{\text{E}}CO_2 \) is the true mixed expired \( PCO_2 \) and \( P_{\text{exp}}CO_2 \) is measured mixed expired \( PCO_2 \) (including gas compressed in the ventilator circuit). This correction is important for dead space calculations. Compression volume does not affect measurements of rates of oxygen consumption and carbon dioxide elimination.

During assisted modes of ventilation, the resistance through the ventilator circuit may contribute to patient-ventilator dysynchrony. The inspiratory work of breathing due to circuit resistance is a function of the inspiratory flow. When this is coupled to the resistance through an endotracheal tube, the imposed work may be clinically important at high flows. The effects of inspiratory circuit resistance becomes important during assisted and spontaneous modes of breathing and can be decreased by triggering at the proximal airway. The resistance through the expiratory limb of the circuit is primarily that due to the expiration valve. Mushroom and scissor valves have significant expiratory resistance. Current generation ventilators use an expiration valve with a large electrically controlled diaphragm that produces a more consistent circuit pressure regardless of flow. Excessive expiratory circuit resistance can prolong expiration and produce auto-PEEP.

HUMIDIFICATION

Inspired gases are conditioned in the airway so that they are fully saturated with water at body temperature by the time they reach the alveoli (37°C, 100% relative humidity, 44 mg/L absolute humidity, 47 mmHg water vapour pres-
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Figure 2  Temperature, relative humidity (RH) and absolute humidity levels at three sites in the respiratory tract. The output of any therapeutic gas delivery system should match the normal conditions at that point of entry into the respiratory system.

TABLE 2
Brief comparison of different types of humidifiers used with critically ill patients

| Humidifier          | Description                                                                                                                                 |
|---------------------|--------------------------------------------------------------------------------------------------------------------------------------------|
| Saline instillation | Simple and low cost; risk of contamination; aids removal of thick tenacious secretions, but should not be used routinely                     |
| Heated humidifier   | Efficient but expensive; condensation in circuit unless circuit heated; maintains body temperature; contributes to compression volume of circuit |
| Nebulizer           | Simple and efficient; condensation in circuit; contributes to compression volume; may contribute to water load to patient; aids removal of secretions; airway cooling (unless heated); airway contamination |
| Artificial nose     | Convenient and requires no external power source; marginal efficiency of some devices; adds dead space and resistance; serves as filter; may clog with secretions |

Circuits, humidification and VAP

decreases the insensible water loss that normally occurs during breathing. Failure to consider this could result in a positive water balance (250 mL/day). With humidification systems, significant heat gain is unlikely and tracheal injury due to high temperature output of a humidifier is an infrequent occurrence. Because the specific heat of gas is low, it is difficult to transfer significant amounts of heat to cause tracheal burns without an aerosol. In hypothermic patients, super-warming of inspired gases has little effect in the facilitation of core rewarming. Breathing gases warmed and humidified to normal body conditions, however, complements other rewarming techniques because it prevents further heat loss from the respiratory tract.

The output of any therapeutic gas delivery system should match the normal conditions at that point of entry into the respiratory system (Figure 2). If the temperature and humidity are less than this, a humidity deficit is produced; if the temperature and humidity are greater than this, fluid overload and patient discomfort may occur. Inspired gases that bypass the upper respiratory tract (e.g. endotracheal tubes and tracheostomy tubes) should be heated to 32°C to 34°C at 95% to 100% relative humidity.

Techniques to humidify the lower respiratory tract are summarized in Table 2. Heated humidifiers are capable of providing a relative humidity of nearly 100% at temperatures near body temperature. Specific devices include the pass-over, cascade, wick and vapour phase humidifiers. The water level in the reservoir of a humidifier can be maintained by manually adding water, adding water from a bag attached to the humidifier or by a continuous-feed system that keeps the water level constant. Closed-feed systems avoid interruption of ventilation to fill the humidifier. Continuous-feed systems avoid fluctuations in the temperature of delivered gas and maintain a low compression volume. Many heated humidifier systems are servo-controlled with a thermistor at the proximal airway to maintain the desired gas delivery temperature. Cooling of the gas between the humidifier and the patient
results in condensation (rain-out), which should be collected in a water trap and removed aseptically.

The circuit that carries gas from the humidifier to the patient can be heated. This prevents a temperature drop in the circuit, and a more precise gas temperature can be delivered. If the temperature of the circuit is greater than the temperature of the gas leaving the humidifier, then the relative humidity of the gas will drop and the circuit will remain dry.

Artificial noses passively humidify the inspired gases by collecting the patient’s expired heat and moisture and returning them during the following inspiration (Figure 5). These devices are attractive alternatives to heated humidifiers because of their passive operation and their relatively low cost. Important considerations in the performance of artificial noses are their humidity output, dead space, flow resistance and cost. Some artificial noses provide greater than 30 mg/L of water to the airway at minute ventilation less than 10 L/min. However, other devices perform poorly and should not be used – all artificial noses are not created equally. There is an increase in resistance to flow through these devices as they become saturated with water, increasing the work of breathing during spontaneous breathing. Although the most efficient devices provide greater than 30 mg/L of water, the output of artificial noses is less than that with a heated humidifier. When the artificial nose is used during prolonged mechanical ventilation, the patient must be frequently assessed for signs of inadequate humidification (eg, thick secretions, bronchial casts, mucus plugging). If signs of inadequate humidification are present, heated humidification...
Contraindications to the use of artificial noses

| Contraindications                                                                 | Summary |
|----------------------------------------------------------------------------------|---------|
| Copious secretions                                                                | Causes  |
| Secretions increase resistance to flow in the artificial nose. If a patient has  |
| copious amounts of secretions, the lack of therapeutic humidification may       |
| result in thickening of secretions when artificial noses are used.              |
| Very small or very large tidal volumes                                           | Causes  |
| With small tidal volumes, the dead space of the artificial nose may compromise  |
| ventilation and lead to CO2 retention. With large tidal volumes, the ability   |
| of the artificial nose to humidify inspired gases is compromised.               |         |
| Low synchronized intermittent mandatory ventilation rates and high spontaneous   |
| minute ventilation (>10 L/min)                                                  |         |
| Artificial noses should be used cautiously in patients with mandatory rates ≤4/min. The resistance through artificial noses increases with time and makes spontaneous breathing difficult |
| Low ventilatory reserve with spontaneous breathing                               | Causes  |
| The pressure drop required for flow through these devices may result in         |
| decreased breathing ability for patients who have low ventilatory reserves.     |
| Expired tidal volume <70% of inspired tidal volume                               | Causes  |
| To function properly, both inspired gases and expired gases must travel through |
| the artificial nose.                                                             |         |
| Hypothermia                                                                       | Causes  |
| Artificial noses are contraindicated with a body temperatures <32°C             |         |
| Aerosol medication treatments                                                     | Causes  |
| Artificial noses should be removed from the circuit during aerosol treatments    |
| when the nebulizer or inhaler is placed in the circuit.                         |         |

Summary of published papers related to ventilator circuit change frequency

| Author, year (reference) | Results                                                                 |
|--------------------------|-------------------------------------------------------------------------|
| Lareau et al, 1978 (23)  | No difference in VAP with 8, 16 and 24 h ventilator circuit changes      |
| Craven et al, 1982 (12)  | No difference in ventilator circuit contamination with circuit changes  |
|                          | at 24 h and 48 h intervals; contaminants in the circuits originated     |
|                          | in the respiratory tracts of patients; water in cascade humidifiers     |
|                          | remained sterile in spite of heavy circuit contamination near patient   |
| Craven et al, 1986 (13)  | The odds of VAP were over two times greater when circuits were changed   |
|                          | at 24 h rather than 48 h intervals                                     |
| Dreyfuss et al, 1991 (15) | VAP rates were similar when circuits were changed every 48 h compared  |
|                          | with no circuit changes; colonization of the circuit usually followed  |
|                          | colonization of the pharynx and trachea; colonization of water in cascade |
|                          | humidifier was rare                                                    |
| Dreyfuss et al, 1995 (14) | VAP rates were similar for use of heated humidifier use and artificial |
|                          | noses; no circuit changes in either group; results suggested that       |
|                          | circuit colonization plays a small role in occurrence of VAP           |
| Hess et al, 1995 (20)    | In a large study of more than 3000 patients and nearly 20,000          |
|                          | ventilator days, there was no significant difference in VAP            |
|                          | with circuits changed at 48 h intervals and 7 day intervals           |
| Kollef et al, 1995 (22)  | VAP rates were similar for 7 day circuit changes and no routine circuit |
|                          | changes                                                               |

VAP Ventilator-associated pneumonia should be initiated. The artificial nose should be replaced at 24 h intervals. There are clinical conditions that contraindicate the use of an artificial nose (Table 3).

VENTILATOR CIRCUITS AND NOSOCOMIAL PNEUMONIA

Intubated mechanically ventilated patients are at risk for nosocomial pneumonia. These pneumonias are associated with high morbidity and mortality, increased length of hospitalization and increased cost of care. The ventilator circuit has been historically associated with the risk of ventilator-associated pneumonia (VAP). The condensate in mechanical ventilator circuits is often contaminated, raising the question as to whether this might pose a risk for VAP. It is now appreciated, however, that organisms contaminating the circuit usually originate from the patient. The patient contaminates the circuit, and VAP may not be ventilator-circuit related. VAPs are usually the result of aspiration of pharyngeal secretions and do not arise from the ventilator circuit. Ventilator circuits do not need to be changed more frequently than weekly for infection control purposes. Recently published papers have shown that the VAP rate does not increase when ventilator circuits are changed at weekly or less frequent intervals (Table 4), and the incidence of VAP may increase with more frequent circuit changes.

There are a number of issues related to ventilator circuits and the risk of VAP. Unlike wick humidifiers, cascade humidifiers produce microaerosols that can carry bacteria. However, humidifiers operate at a high temperature that is bactericidal for nosocomial pathogens. Heated circuits minimize condensation in ventilator circuits. Because circuit condensate almost always arises from the patient, the role of heated circuits to avoid VAP is unclear. For unheated circuits, water traps should be used and evacuated aseptically at regular intervals to avoid bolus lavage of the patient with circuit condensate. Artificial noses and heated circuits maintain a dry circuit, but have not been shown to affect the incidence of VAP. Use of medication nebulizers can deliver contaminated aerosols to the lower respiratory tract and metered dose inhalers may be superior to nebulizers in this
respec. Closed suction systems may have useful infection control implications because they prevent spraying of ventilator circuit condensate and tracheal secretions into the intensive care environment during suctioning.

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