ROLE OF SUNKIST PEELS EXTRACT AGAINST THE LEVEL OF ESTRADIOL AND MATURITY OF FOLLICLE IN THE FEMALE WISTAR RAT

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ABSTRACT

Objective: The purpose of this study is to investigate the effect of Sunkist peel extract against the estradiol level and ovarian follicle in the female Wistar rats.

Methods: This study was used 25 rats that were divided into five groups, namely, control (Na-CMC), standard (ethinylestradiol), and Sunkist peels extract-I, II, and III (300 mg/kg BW, 450 mg/kg BW, and 600 mg/kg BW). After 14 days, all rats were sacrificed by chloroform inhalation, and the ovarium was excised to histopathology preparation; meanwhile, the blood was collected for the serum. The histology and serum were determined the percentage of each type of follicle and the estradiol level, respectively.

Results: The result of this study showed that increasing of the extract significantly increased not the level of estradiol level (p<0.05) but also the percentage of secondary and de Graaf follicle (p<0.05). Nevertheless, the percentage of primordial and primer follicle decreased, followed by increasing the extract dosage (p<0.05).

Conclusion: It can be concluded that the Sunkist peels Sunkist peel increase both the level of estradiol serum and maturity of the follicle that increases the opportunity of female rats to be fertilized.

Keywords: Sunkist peels, Ethinylestradiol, Estradiol, Follicle.

INTRODUCTION

Infertility is a common case which can be found today. This can be seen from the latest data, which was reported whether the prevalence of infertility was 9 to 18% in the general population. In 2008, 61,430 neonates born from assisted reproductive technology [1].

About 15% of married couples suffered from infertility that was around 48.5 million couples over the world. Meanwhile, about 17% of the couple living in the industrialized countries reported that they required treatment related to infertility, which can be caused by ovulation failure, tubal damage or endometriosis, or a low sperm count. When puberty, primary oocytes in primary follicles in the ovary undergo meiosis to form polar bodies and an ovum that will be released for conception during ovulation. Typically, the ovarium content 300,000 reserve follicles. However, around 400 follicles become mature and can be ovulated; meanwhile, remain 99.7% will degrade [2].

Ovulation is affected by various hormones such as follicle-stimulating hormone (FSH), luteinizing hormone (LH), estrogen, and progesterone. Before ovulation, estrogen takes a dominant role, while after ovulation is the progesterone. Normally, estrogen level before menopause or premenopause is 30-400 pg/ml and reduces when postmenopausal age into 0-30 pg/ml [2,3].

In Indonesia, “Badan Pusat Statistik” reported that around 39.8 million women were in the childbearing age. However, 10–15% of them were infertile [4]. Various factors can cause infertility; however, more than 70% come from female problems. Infertility in female can be caused by ovarium, uterine tube, uterine, hypothyamic, hypophysis, or systemic disease. Around 27% of the cases come from ovarium cause due to ovulation failure, either congenital or imbalanced hormones [5-8].

Various herbs enrich Indonesia. Some herbs may content phytoestrogens. Phytoestrogen is a natural compound from herbs, which has a similar structure with 17-β-estradiol (E2). It was a common sexual hormone among the female. The phytoestrogens are commonly found as isoflavone, pentyflavonoid, coumestans, and ligans [8]. One of the natural products that have these compounds is Sunkist (Citrus sinensis L. Osbeck) peel. Sunkist ethanol extract contains various phytochemicals; one is a flavonoid that the derivate may act as the phytoestrogen. Phytoestrogen has a similar structure to the 17-β-estradiol (E2). It affects the ovulation cycle. Due to these reasons, this study was aimed to investigate the effect of Sunkist peel extract against the estradiol level and ovarian follicle in the female Wistar rats as the animal trial [9,10].

METHODS

This study was an experimental study with post-test only group control design in June 2020 at the Pharmacology Laboratory at the Faculty of Medicine, Universitas Prima Indonesia. Health Research Ethics Committee from Universitas Prima Indonesia has approved the protocol of this study with registration no. 012/KEPK/UNPRI/V/2020.

The materials used in this study were 96% ethanol, Sunkist peels, sodium carboxymethylcellulose (Na-CMC), ethinylestradiol (Lynoral®), distilled water, standard rat pellets, chloroform, alcohol, formalin, xylol, paraffin, and hematoxylin-eosin (HE) staining powder.

The Sunkist was obtained from one of the traditional markets in Medan City, then it was identified at the Medanese Herbarium at FMIPA, University of North Sumatra. Amount of 500 grams Sunkist peel was dried by aerating for 7 days then pulverized to become simplicia powder. The simplicia powder was extracted by maceration method. First, simplicia powder was macerated by 70% ethanol as a solvent in a ratio of 1:10 for 3 days. The mixture was stirred regularly every day. After 3 days, it was filtered by filter paper, and then, the residue...
was re-macerated with 600 ml of 70% ethanol for 3 days. The filtered maceration and maceration were evaporated by rotary evaporator at a temperature of 70°C and then followed by concentrated using the oven at 40°C until they become thick extracts [11,12].

After that, the ethanol extract of Sunkist peels was suspended into the sodium carboxymethylcellulose to form an oral suspension. The oral suspension used 0.5% sodium carboxymethyl cellulose as vehicle. It was made by mixing 0.5 grams sodium carboxymethyl cellulose powder into 30 ml hot distilled water in the mortar for 15 min until it formed a clear phrase. Then, it was grounded until homogenous and dissolved in a volumetric flask by the remaining distilled water. Moreover, this suspension was used to vehiculum for extract and standard drugs. Amount of 600 mg, 900 mg, and 1200 mg ethanol extract of Sunkist peel was dissolved into 10 ml [11].

The treatment was performed into 25 rats that were divided into five groups. The rats as the animal trial were a female rat, 150–200 g of body weight. Each group was received the following treatments [13].

a. Control: The rats received 1 ml of 0.5% sodium carboxymethyl cellulose suspension
b. Standard [ethinylestradiol 50 µg/kgBW]: The rats received 1 ml of ethinylestradiol (Lynoral®) oral suspension
c. Sunkist peels extract-I (300 mg/kg BW): The rats received 1 ml of an oral suspension of Sunkist orange peel extract at a dose of 300 mg/kg BW
d. Sunkist Peels Extract-II (450 mg/kg BW): The rats received 1 ml of an oral suspension of Sunkist orange peel extract at a dose of 600 mg/kg BW
e. Sunkist Peels Extract-III (600 mg/kg BW): The rats received 1 ml of an oral suspension of Sunkist orange peel extract at a dose of 1200 mg/kg BW.

All rats in each group can access food and drink freely and received some treatments for 14 days. After that, all rats were sacrificed by inhalation of chloroform in the closed room. After the rat was anastized, the rat’s abdominal and chest wall were incised vertically, and the blood was collected from the cardiac punctured by 5 ml spuit and 25G needle, and then, it was saved in a red tube. Furthermore, both ovarian tissues were obtained and rinsed in normal saline. After that, the tissues were soaked in buffer neutral formalin 10% and were stained by HE staining.

The estradiol level was determined from obtained serum by ELISA Methods. After that, the estradiol level was analyzed by One Way ANOVA and followed by post-hoc Test Tukey HSD. Table 1 is shown the result of the analysis.

Based on Table 1 above, all intervention groups showed significant differences in estradiol level one another. It was shown by the p-value and superscript that was obtained by one-way ANOVA and post-hoc test Tukey HSD.

**Estimation of follicle development**

Other than the estradiol level, this study also investigated the number of follicles in rats’ ovarian tissue. The number of follicles was expressed as the percentage and shown by the following Table 2.

Based on Table 2 above, it can be seen that the number of primordial, primer, secondary, and de Graaf follicle was significant difference due to p-value lower than 0.05. The further analysis showed that the increasing Sunkist peels extract dosage reduced percentage of primordial and primer follicle but increased percentage of secondary and de Graaf follicle. The remain follicle, namely, corpus luteum and atresia, was not shown a significant change in the percentage. It can be seen in p-value that higher than 0.05. Fig. 1 is shown histology of the ovarian structure.

Based on the Fig. 1 above, ovarium was a structure that consists of cortex and medulla part and lined by germinal epithelium. All types of follicles can be found in the cortex of ovarium such as the primordial, primer, and atresia follicle shown by the figure above. Furthermore, the clear structure of each follicle can be seen in the following Fig. 2.

Based on the Fig. 2 above, it can be seen the structure of primordial, primer, and secondary follicle at ×400 magnified. Primer follicle was characterized by an oocyte that was lined by a single layer of squamous follicular cells.

The primordial follicle will develop into primer follicle. The single layer of follicular cells transformed into cuboid form and form primer follicle. Over time, the primer follicle develops into the secondary follicle. Follicular layer develops to form a more stratified structure known as granulosa zone, and the follicle also develops an antrum in a pole of the follicle. The antrum will become wider at the appropriate time, and the corona radiata and cumulus oophorus around the oocyte become more obvious. Hence, the secondary follicle develops into de Graaf follicle. Fig. 3 is shown the microscopic view of de Graaf follicle in the two different magnified.

Fig. 3 above is shown that the de Graaf follicle had wider antrum structure and obvious cumulus oophorus and corona radiata as de Graaf follicle’s specific properties.

**DISCUSSION**

The result of this study clearly answered the purpose of this study to investigate the effect of Sunkist peel extract against the estradiol level and ovarian follicle. Ethanol extract of Sunkist obviously affects the estradiol

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**Table 1: Analysis of the estradiol serum level in all intervention groups**

| Groups                  | Level of estradiol (pg/ml) | p-value |
|-------------------------|----------------------------|---------|
| Control                 | 34.00±1.58<sup>a</sup>     | <0.05   |
| Standard                | 53.60±1.14<sup>b</sup>     |         |
| Sunkist peels extract-I | 43.80±1.30<sup>c</sup>     |         |
| Sunkist peels extract-II| 46.20±1.30<sup>d</sup>     |         |
| Sunkist peels extract-III| 49.00±1.58<sup>d</sup>   |         |

Data were expressed as Mean±SD and P value was obtained by one-way ANOVA. Different superscript in the same column significantly differentiates according to post-hoc test Tukey HSD.
level and significantly affects the follicle’s percentage in the ovarium. The increasing of extract dosage increases estradiol serum level, but it was not as high as standard drugs. Moreover, this study was also shown that the increasing of extract dosage increased percentage of secondary and de Graaf follicle but reduced percentage of primordial and primer follicle.

The development of primordial follicle affects by the FSH. Moreover, this hormone will affect the primordial follicle’s maturity into the primer and secondary hormone as long as the follicular layer was developed more stratified that will increase the level of estrogen hormone. It due to the secretion of hormone occurred in the follicular cell. The precursor

Table 2: Analysis of the percentage of follicle in all intervention groups

| Groups                    | Percentage of follicle (%) | Primordial | Primer | Secondary | De Graaf | Corpus Luteum | Atresia |
|---------------------------|----------------------------|------------|--------|-----------|-----------|---------------|---------|
| Control                   |                            | 65.98±0.55 | 27.82±1.19 | 5.35±1.55 | 0.75 (0.00–1.5) | 0.00 (0.00–0.50) | 0.00 (0.00–0.00) |
| Standard                  |                            | 33.50±0.80 | 35.28±4.96 | 13.16±5.25 | 5.30 (5.00–8.33) | 0.00 (0.00–0.50) | 0.00 (0.00–0.25) |
| Sunkist peels extract-I   |                            | 51.06±0.75 | 37.75±1.09 | 9.79±1.68 | 1.00 (0.00–1.50) | 0.00 (0.00–1.25) | 0.00 (0.00–0.25) |
| Sunkist peels extract-II  |                            | 43.95±1.11 | 34.39±0.95 | 14.83±2.70 | 3.50 (2.50–4.00) | 1.00 (0.00–1.50) | 0.00 (0.00–0.25) |
| Sunkist peels extract-III |                            | 45.34±0.79 | 34.18±1.80 | 17.55±1.03 | 4.50 (3.40–5.00) | 1.00 (0.00–2.00) | 0.50 (0.00–0.50) |

*p-value*<0.05* **Data were expressed as mean±SD and P value was obtained from the one-way ANOVA and followed by post-hoc test Tukey HSD, **Data were expressed as median (min-max) and P value was obtained from Kruskal–Wallis test and followed by Mann–Whitney U-test; Different superscript in the same column significantly differentiates according to either post-hoc test Tukey HSD or Mann–Whitney U-test
of estrogen was formed in the theca cell after that, it diffused into the follicular cell and converted into estradiol by 17β-HSD or estrone by aromatase. FSH regulated the expression of these enzymes [2,14,15].

The increasing maturity of follicle affects the level of estrogen. The high level of estrogen has two main effects on the maturity of the follicle. First, it gives positive feedback against the hypothalamus-k-hypophysis to amplify the secretion of estrogen hormone while the granulosa cell is secreting inhibin to reduce the secretion of FSH. Another effect is the induction of ovulation; however, it also required the LH surge to induce the ovulation [2,14,16].

These information support the result of this study. The increasing of estradiol level causes the increasing number of secondary and de Graaf follicle. However, the increasing estradiol level was not occupied by LH surge; hence, the ovulation did not occur. On the other hand, the percentage of primordial and primer follicle was reduced; it was caused by the increasing number of these follicles developed into the more mature follicle.

None of the studies investigates the effect of Sunkist peels against female fertility by in-vivo methods. Nevertheless, an in-vitro study has been performed to determine the phytoestrogen level of Sunkist peel. Phytoestrogen has a similar structure to endogenous estradiol and binds with alpha and beta receptors of estrogen. Moreover, this complex will influence the gene expression of the cell. Kuhnle et al. reported that orange peels contented 36 µg/100 g wet weight of phytoestrogen based on the liquid chromatography–mass spectrometry and 13C-Labeled standard [17]. Another study performed by Shalaby and Shanab on the both C. sinensis leaf and Citrus aurantium peel reported that flavonones, flavones, and flavonoids could act as powerful estrogen-like activity [13,18,19].

Further study was required to investigate the role of other hormones such as FSH, LH, and progesterone simultaneously against the follicle development and it should be performed as a multidiscipline study to support the result of this study.

CONCLUSION

It can be concluded that the Sunkist peel can increase the level of estradiol serum and increase the develop of de Graaf follicle. It can be concluded that the Sunkist peel can increase the level of estradiol serum and increase the develop of de Graaf follicle that can increase the opportunity of female rats to be fertilized.

ACKNOWLEDGMENTS

The author(s) disclosed receipts of the financial support for the research and/or publication of this article.

AUTHORS’ CONTRIBUTION

Maya Sari Mutia – Writing manuscript, evaluation of microscopic view, concept and designing the study, and final approval. Suhartina – Data collection, data analysis, interpretation of data, and final approval.

CONFLICT OF INTEREST

The authors declare that there was no conflict of interest.

AUTHORS FUNDING

This study was fully funded by the Ministry of Research, Technology, and Higher Education by Funding Contact No. 282/LLI/PG/2020.

REFERENCES

1. Hanson B, Johnstone E, Dorais J, Silver B, Peterson CM, Hotaling J. Female infertility, infertility-associated diagnoses, and comorbidities: A review. J Assist Reprod Genet 2017;34:167-77.
2. Sherwood L. Fisiologi Manusia dari Sel ke Sistem. Jakarta: EGC; 2013. p. 811-70.
3. Etuk EU, Okolo RU, Okpokwasili MO, Ume ES. The role of fennel infusion on estrous cycle and follicles development of white rats. J Ilmu Kefarmasian Indonesia 2016;24:197-203.
4. Sharabi OJ, Afolayan AJ. Biochemical and hormonal effects of Citrus sinensis peel extract against isoniazid and rifampicin-induced liver injury in wistar rats. Maj Obat Tradis 2013;1:164-70.
5. Hestiantoro A, Wiweko B, Pratama G, Yusuf D. Konsensus Penanganan infertil di klinik fertilitas dan bayi tabung tiara cita rumah sakit putri Surabaya. J Biom Kependud 2016;5:61-9.
6. Igbokwe VU, Ugwah-Oguejiofor CJ, Bello SO, Okolo RU, Etuk EU, Ugwah MO. Konsensus Penanganan infertil di klinik fertilitas dan bayi tabung tiara cita rumah sakit putri Surabaya. J Biom Kependud 2016;5:61-9.
7. Desmawati D, Sulastri D. Phytoestrogens and their health effect. Open Access Muced J Med Sci 2017;7:495-9.
8. Rietjens IM, Liau J, Weckx K. The potential health effects of dietary phytoestrogens. Br J Pharmaco 2017;174:1263-80.
9. Bakti Q. Phytoestrogens and their health effect. Open Access Muced J Med Sci 2017;7:495-9.