The Scent of the Waggle Dance

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Introduction

More than fifty years ago, Karl von Frisch demonstrated through a series of elegant experiments that the waggle dance of honey bees uses symbolic communication to convey information about a subject that is both spatially and temporally removed from the receiver of the signal [1]. The waggle dance is therefore a unique animal signal that exhibits several of the important properties of true language, which are generally attributed only to “advanced” organisms such as marine mammals, nonhuman primates, and humans. Honey bees are themselves quite advanced, however; a honey bee society consists of a large group of individuals of overlapping generations, living permanently together with cooperative care of young and a division of labor. To coordinate the complex interactions among the members of such a society, a sophisticated system of communication is necessary.

The role of the waggle dance in this sophisticated system of communication is primarily to direct the colony’s foraging effort toward nectar- and pollen-producing flowers. Successful foragers perform the dance within the nest to recruit other bees to a profitable food source. By closely following a dancer, potential recruits acquire information about the location and richness of the advertised food source [1,2]. However, despite our considerable knowledge of the information contained in the dance, we still do not understand how dancers attract and convey information to recruits in the darkness of the hive [2]. Airborne sounds [3,4], substrate vibrations [5,6], and tactile cues [7] seem to play some role in attracting recruits to waggle dancers or conveying dance information, but each of these modalities appears to be neither necessary nor sufficient for recruitment [2].

Another modality that may be involved in waggle dance communication is olfaction. Evidence to date that olfaction plays a role in honey bee waggle-dance communication is limited to odors acquired from the environment at or en route to the floral food source [2,8,9]. (These odors are thought to serve as cues that allow foraging recruits to pinpoint the food source advertised by the waggle dancer.) However, the production of olfactory signals by the waggle dancers themselves could attract recruits and convey dance information. Pheromones and other semiochemicals are used frequently by honey bees and other social insects to coordinate the activities of colony members [10,11] and hence may be expected to help organize the vital task of foraging. Furthermore, evidence suggests that closely related Hymenopterans, such as bumblebees, use pheromones within the nest to organize foraging activity [12]. Whereas bumblebees do not communicate via waggle dances, olfactory-based foraging communication may be an ancestral trait and thus present in honey bees. A preliminary study suggested indeed that the scent of active honey bee foragers could encourage other bees to forage [13].

The goal of this study was to investigate whether waggle dancers produce and release into the air chemical compounds that distinguish them from other foragers. We addressed this first goal by using solid phase microextraction (SPME) and gas chromatography coupled with mass spectrometry (GC/MS). If these distinguishing compounds are semiochemicals or pheromones, then adding these compounds into the hive may affect the behavior of foraging bees. To test whether the compounds are behaviorally active, we measured foraging activity before and after we injected the volatilized compounds into the hive. Our results show that honey bee waggle dancers produce four characteristic volatile compounds that increase foraging activity.

Results

We discovered four conspicuous compounds in the airspace surrounding dancing bees that were not present in the airspace surrounding nondancing bees (Figure 1). We obtained similar results for the airspace surrounding waggle
dancers on an artificial swarm as compared with the air over an area of the swarm with no waggle dancers. From extracts made of three intact waggle dancers immersed for 1 min into 250 µl of hexane, we identified the four substances that generated peaks 1, 2, 3, and 4 as marked in Figure 1 to be Z-(9)-tricosene, tricosane, Z-(9)-pentacosene, and pentacosane, respectively.

All four compounds were present in significantly higher amounts on the abdomens of waggle dancers than in either (a) nondancing foragers that returned from the same unscented nectar source, or (b) nonforaging bees [Figure 2, one-way analysis of variance (ANOVA) for each compound, degrees of freedom (df) = 2, 49, \( p < 0.001 \) for all compounds; Tukey HSD for unequal \( n \), \( p < 0.0005 \) for all compounds, experiment-wise \( \alpha = 0.05 \)]. Intriguingly, although only marginally significant, waggle dancers that danced more vigorously (i.e., appeared to perform waggle runs at higher rates and with more exaggerated movement of the abdomen as assessed subjectively by one person, \( n = 5 \) bees) tended to emit more of all four compounds than less vigorous dancers did \((n = 13)\) (Figure 3, Mann-Whitney U test, \( p = 0.05, 0.07, 0.13, \) and 0.08 for peaks 1, 2, 3, and 4, respectively). Nondancing foragers and nonforaging bees did not emit different amounts of these compounds (Tukey HSD for unequal \( n \), \( p = 0.076, 0.074, \) and 0.400, for peaks 2, 3, and 4, respectively) with the exception of Z-(9)-tricosene \((p < 0.001)\).

To test whether the waggle-dance-specific substances affect behavior, we injected onto the dance floor a gaseous mixture of the three commercially available compounds dissolved in hexane [hereafter called TTP (Z-(9)-tricosene, tricosane, pentacosane) solution]. TTP trials lasted 32 min, during which we recorded the number of bees exiting the hive each minute, and during which we made two injections: Injection 1, which consisted of 50 µl of pure hexane, was made during minute 1, and Injection 2, which consisted of 50 µl of TTP solution, was made during minute 16. To control for solvent and treatment, we performed “Hexane trials”, in which Injection 2, like Injection 1, consisted of 50 µl of pure hexane.

Experiments were done with two colonies, C1 and C2, using one colony at a time. We performed a total of 20 trials (10 TTP trials and 10 Hexane trials) with C1, and a total of 28 trials (15 TTP trials and 13 Hexane trials) with C2. Only one trial was performed per day, and all trials were performed at the same time of day. Because the compounds originate from waggle dancers under natural conditions, we ascertained that at least one dancer was present during each trial.

Injection of TTP increased the number of bees exiting the hive (Figure 4). The normalized mean number of bees exiting the hive during minutes 25–32 differed significantly between TTP trials (i.e., Injection 2 = TTP solution) and Hexane trials (i.e., Injection 2 = hexane) (two-sample T-test; Colony 1: \( T = 2.66, p = 0.004, df = 18 \); Colony 2: \( T = 2.25, p = 0.033, df = 26 \)). This difference between TTP trials and Hexane trials was not observed following Injection 1, which consisted of hexane for both types of trial (Colony 1: \( T = 1.63, p = 0.121, df = 18 \); Colony 2: \( T = -0.47, p = 0.643, df = 26 \)).

We also observed within TTP trials an increase in the number of bees exiting the hive following Injection 2, as compared with the number exiting following Injection 1, but this was significant for only one colony (two-sample T-test; Colony 1: \( T = -1.63, p = 0.121, df = 18 \); Colony 2: \( T = -2.66, p = 0.013, df = 28 \)). This increase could possibly be the result of circadian foraging patterns, but this would not explain the lack of such an effect during Hexane trials (two-sample T-test; Colony 1: \( T = 0.83, p = 0.415, df = 18 \); Colony 2: \( T = -0.68, p = 0.511, df = 18 \)).
which were conducted at the same time of day as TTP trials.

**Discussion**

We have identified in this study four compounds that are unique to waggle-dancing bees and that are behaviorally active. This waggle-dance scent originates from the waggle dancers themselves; it is not acquired from the environment while foraging, nor is it a byproduct of a bee’s age or task, because it is emitted in only minute quantities by nondancing foragers returning from the same food source.

The prominent compounds of the waggle-dance scent are the cuticular hydrocarbons Z-(9)-tricosene, tricosane, Z-(9)-pentacosene, and pentacosane. Hydrocarbons such as these are produced subcutaneously and are not stored within a gland [14]. The waggle-dance scent is therefore best classified as a semiochemical or nonglandular pheromone. Waggle dancers could raise the levels of the compounds through increased synthesis and/or increased release onto the epicuticle. The compounds could then be released passively into the air, perhaps assisted by the relatively high body temperature of the waggle dancers.

The chemical nature and source of the compounds of the waggle-dance scent differ from those of the bumblebee foraging recruitment pheromone [15]. The main compounds of the bumblebee recruitment pheromone were identified as eucalyptiol, ocimene, and farnesol, which are terpene derivatives. These compounds are produced in the bees’ tergal glands. Their different chemistry and source suggest different evolutionary origins for the bumblebee foraging recruitment pheromone and the waggle-dance scent of honey bees. Hence, it is unlikely that the waggle-dance scent of honey bees has evolved from the bumblebee foraging recruitment pheromone.

The compounds of the waggle-dance scent have been identified in earlier studies on honey bees. Z-(9)-tricosene, tricosane, Z-(9)-pentacosene, and pentacosane have been previously identified in hexane washes of the cuticles of foraging-age worker bees [16] and have been shown to be perceptible to honey bee workers [17]. More recent work has shown that, among many others, the four compounds are present in the air surrounding foragers at a feeder station [18]. However, with the exception of demonstration of a possible kin-recognition function of Z-(9)-tricosene [19], past studies did not link specific compounds with specific behaviors of honey bees. In insects other than honey bees, however, the individual compounds of the waggle-dance scent have been linked to specific behaviors. For example, tricosane, pentacosane, and Z-(9)-pentacosene are compounds of a pheromone that foragers of the social wasp *Vespula vulgaris* use to lay and follow terrestrial trails [20]. Compounds of the waggle-dance scent are also well-known sex attractants in other insects, such as flies [14].

The waggle-dance scent may have a similar marking or attracting function, which the context and results of our experiments link to recruitment behavior. Honey bees produce the scent when they perform waggle dances, both

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**Figure 2.** Relative Abundance of Alkanes and Alkenes

Relative abundance of Z-(9)-tricosene, tricosane, Z-(9)-pentacosene, and pentacosane sampled from three types of bees. The height of each bar indicates the mean peak area for each compound from GC/MS chromatograms. The relative abundance of compounds is significantly higher for waggle-dancing foragers (black bars) than for either nondancing foragers (hatched bars) or nonforaging bees (white bars, for statistical tests, see text). The relative abundance of compounds for nondancing foragers does not differ from that for nonforaging bees, with the exception of Z-(9)-tricosene. Error bars represent the standard error of the average of six daily mean values, each daily mean comprising a sample of two–three bees of each type. Peak areas represent relative abundance only and thus do not enable comparison among different compounds.

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**Figure 3.** Comparison of Vigorous and Less-Vigorous Dancers

The relative abundance for Z-(9)-tricosene, tricosane, Z-(9)-pentacosene, and pentacosane tended to be higher for vigorous (n = 5, black bars) than less-vigorous (n = 13, white bars) dancers. Vigor of dancing was assessed subjectively by one experimenter and included appearance of abdomen movement and the rate of waggle runs. More vigorous dancers appeared to move their abdomen farther to the side and perform more waggle runs than less-vigorous dancers. Error bars represent the standard error. Statistical tests are given in the text.

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in the hive and on swarms, and in both contexts, workers are recruited to the advertised site. In the more common context of foraging, a general measure of recruitment is the number of foragers that leave the hive. Our results show that injection of the waggle-dance scent onto the dance floor increased the number of bees that left the hive. These bees can be assumed to be foragers, because only foragers leave the hive without noticeably hesitating, performing orientation flights, or gathering at the hive entrance.

Our experiments show that the waggle-dance scent increases the number of foragers leaving the hive, but the exact mechanism underlying the effect is still unclear. Given the function of the waggle-dance scent in other hymenoptera, we propose that the waggle-dance scent, which in our experiments was fanned onto the dance floor, attracts potential recruits to the dance floor, thereby increasing the probability of encounters between potential recruits and dancers, and finally the number of recruits. Under natural conditions, the scent would originate from the dancers themselves, thus the odor plume would mark not only the dance floor, but the individual dancers. This should enable recruits to locate the dancers themselves, which could enhance recruitment efficiency. Recruits could even seek out more vigorous dancers, who typically advertise especially profitable food sources [21], and who seem to emit higher concentrations of the scent. Hence, the waggle-dance scent may mark and attract recruits to successful foragers, and thus provide information to a recruit, even if she lost contact with the dancer herself. This hypothesis is supported by the observation that a mechanical model of a waggle dancer recruits bees to a food source only after the model has touched a waggle dancer (H. E. Esch, personal observation).

Whereas the waggle-dance signal is likely a signal intended for new recruits, two other groups of bees, namely foragers already devoted to a food source and in-hive receiver bees, could glean cues from the waggle-dance scent. Foragers that are already devoted to a food source do not readily follow new dances if their source becomes unavailable, but rather wait for it to replenish [8]. Because the waggle-dance scent does not seem to identify specific food sources, it can provide only limited information to these foragers. However, high concentrations of the scent could alert them to generally good foraging conditions. This could be useful at the beginning of daily foraging or if foraging can be resumed after a spell of bad weather. This mechanism may have been responsible for the effect observed in an earlier preliminary study, in which the number of foragers visiting an empty feeder increased following exposure to the air from a foraging colony [13]. In our experiments, however, the increase in foragers was not likely caused by already-devoted foragers for three reasons. First, experiments were done well after the time that colonies started daily foraging. Second, external conditions such as the weather were remarkably stable, which made strong fluctuations in the numbers of already-devoted foragers unlikely. Third, we did not record a conspicuous drop in the number of bees that left the hive after the initial increase (Figure 4); if the increase would have been due mostly to already-devoted foragers, we would have expected a quick drop to original levels once these foragers found that there was no change in food-source profitability.

**Figure 4. Injection of Scent and Behavioral Response**
Comparison of foraging activity in Colony 2 following injection of the TTP solution versus pure hexane. The normalized mean number of bees exiting the hive following Injection 2 (during minutes 25–32) was greater during TTP trials (black bars) than during Hexane trials (white bars). Results were similar for Colony 1. Statistical tests are given in the text.
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However, it is possible that such a drop could be hidden by the more substantial numbers of newly recruited foragers. Receiver bees are the second group of bees that could glean cues from the waggle-dance scent. Receiver bees unload nectar from newly returned foragers, which relieves foragers of the time-consuming search for empty storage cells. A high concentration of the waggle-dance scent would indicate a high demand for receiver bees, and could help attract receiver bees to the dance floor. The tremble dance, which is performed by successful foragers that perceive a shortage in receiver bees [22], may help to additionally spread the scent to potential receiver bees.

Materials and Methods

Bees. Bees were kept indoors in four-frame observation hives at the Carl Hayden Bee Research Center in Tucson, Arizona, United States. To test whether waggle dancers produce specific chemical compounds, we performed experiments with two colonies, using first one, then the other colony. Foragers were trained to an artificial feeder [1] ~ 100 m from the hive that offered nonscented sugar water. Foragers visiting the feeder were marked on the thorax with powdered paint and thus could be recognized in the hive.

Sampling of chemical profiles. We used a SPME fiber (65 μm polydimethylsiloxane/divinylbenzene; Supelco; http://www.sigmaaldrich.com/Brands/Supelco_Home.html) to sample chemicals. After sampling, the SPME fiber was injected into a GC, CP-3800 (Varian; http://www.varian.com) coupled to a MS (Saturn 2200 Ion Trap,Varian).

We compared the chemical profile of the air over dancing bees to that over nondancing bees, and we also compared the chemical profile of waggle dancers’ abdomen scent to that of both nondancing foragers and nonforaging bees. For air samples, we exposed the fiber for 5 consecutive min approximately 2 cm above the surface of the comb. For abdomen profiles, we briefly touched the fiber to the tip of the abdomen of an individual bee [23–26]. Waggle dancers were sampled during a waggle run, shortly after they began dancing. Sampled bees were observed from the moment they entered the hive until the sample was taken. It is possible that we classified as nondancing bees that danced before observation began (e.g., in the entrance tunnel to the hive) or that danced after SPME sampling, but this would bias our results in only a conservative direction.

Compound identification. After SPME sampling, the fiber was desorbed for 3 min in the GCMS, and compounds separated on a Varian VF-5MS 30 m x 0.25 mm inner diameter (ID) column with an injector temperature of 250 °C, and a column temperature of 40 °C for 5 min, which was then ramped at 50 °C/min to 150 °C, followed by a ramp at 15 °C/min to 260 °C with a 4.5-min hold; flow rate was 1 ml/min. The MS was operated in electron ionization mode at 150 eV. Tentative identification of the peaks was made by comparing MS fragment patterns with spectra from the National Institute of Standards and Technology (NIST) 98 and Wiley library databases. Chemical ionization with acetoniitride was used to determine molecular weights and to assign double-bond position through derivatization of the double bond and formation of characteristic addition compounds [27,28]. The identities of peaks 1, 2, and 4 were further confirmed by the production of identical retention times and fragment patterns in both electron and chemical ionization modes when compared with chemical standards.

Behavioral tests. TTP trials consisted of 32 min during which we made two injections: Injection 1, 50 μl of pure hexane, during minute 1; Injection 2, 50 μl of TTP solution, during minute 16. Hexane trials were similar to TTP trials except that Injection 2, like Injection 1, consisted of 50 μl of pure hexane.

The TTP solution contained 2-(9)-tricosene, tricosane, and pentacosane, each diluted 1:100 in hexane and mixed at a ratio of 1:23, respectively (this ratio produced chromatograms with peak heights that approximately matched those from samples of waggle dancers), and then further diluted 1:10 in hexane. To volatilize the TTP solution, we injected it into a heated glass tube with 0.5-cm ID. Immediately after injection, we inserted into the tube a fan to blow the vaporized solution through the tube and into a funnel (10-cm diameter) positioned 1 cm above the comb surface of a colony’s dance floor. The identical method was used to volatilize and deliver hexane only during Hexane trials (Injection 1: hexane, Injection 2: hexane). To avoid contamination between TTP and Hexane trials, we used separate equipment for TTP solution and hexane. The temperature of the gaseous mixture arriving on the dance floor was maintained between 35 °C and 40 °C. At least one waggle dancer was present on the dance floor during each trial.

We conducted each trial on a different day between 14 July and 8 October 2004. To avoid seasonal effects, we randomized the type of trial (TTP or Hexane) performed each day. To reduce the effect of time of day, trials for a colony started during the same hour every day. Trials for the first colony started between 1200 and 1300, and for the second colony between 1030 and 1130. To further account for day and time effects, the data for each trial were normalized by dividing each 1-min count by the average number of bees exiting the hive per minute during the 10 min immediately preceding the trial.

Statistics. To compare the presence of the four compounds on the abdomens of waggle dancers with either nondancing foragers that returned from the same unscented food source or nonforaging bees, we used a one-way ANOVA for each compound using Box-Cox transformed data, and a Tukey HSD for unequal n, with data for both colonies pooled.

To compare the effect of injection of TTP solution with injection of hexane, we used two-sample T-tests on normally distributed sample groups with equal variances. To account for day and time effects, the data for each trial were normalized by dividing each 1-min count by the average number of bees exiting the hive per minute during the 10 min immediately preceding the trial.

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