Serum retinol binding protein 4 is negatively related to beta cell function in Chinese women with non-alcoholic fatty liver disease: a cross-sectional study

Hongmei Yan¹, Xinxia Chang¹, Mingfeng Xia¹, Hua Bian¹, Linshan Zhang¹, Huandong Lin¹, Gang Chen², Mengsu Zeng² and Xin Gao¹*

Abstract

Background: To observe the relationship between serum retinol binding protein 4 (RBP4) and β cell function in Chinese subjects with non-alcoholic fatty liver disease (NAFLD) and without known diabetes.

Methods: 106 patients diagnosed as fatty liver by ultrasonography (M/F: 61/45; aged 47.44 ± 14.16 years) were enrolled in our current cross-sectional study. Subjects with known diabetes, chronic virus hepatitis and excessive alcohol consumption were excluded. Serum RBP4 was detected by ELISA and validated by quantitative Western blotting. β cell function were assessed by HOMA in all subjects and by hyperglycemic clamp in 17 normal glucose tolerance subjects (M = 6, F = 11).

Results: The levels of serum RBP4 in men were higher than that in women (55.96 ± 11.14 μg/ml, p < 0.001). Pearson’s correlation analysis demonstrated that in women, serum RBP4 levels were significantly associated with fasting blood glucose (FBG), HOMA-β, and increment of first phase insulin secretion (1PH), but not associated with age, BMI, waist circumference, WHR, systolic (SBP) and diastolic blood pressure (DBP), TC, TG, HDL-c, LDL-c, 2 h blood glucose, HOMA-IR, ALT, AST, γ-GT, hepatic fat content (HFC), and insulin sensitivity index (ISI). However, in men, serum RBP4 levels were significantly associated with HDL-c, ALT, AST, but not associated with any other parameters as mentioned above. A stepwise multiple linear regression analysis demonstrated that in women, HOMA-IR and RBP4 were significantly associated with HOMA-β, while in men, HOMA-IR and BMI were significantly variables associated with HOMA-β.

Conclusions: Serum RBP4, secreted mainly by liver and adipose tissue, may involve in the pathogenesis of β cell dysfunction in Chinese women patients with NAFLD.

Keywords: Serum retinol binding protein 4 (RBP4), Beta cell function, Non-alcoholic fatty liver disease (NAFLD)

Background

Retinol binding protein 4 (RBP4) is a member of the lipocalin family of proteins that transport retinol from the liver to the peripheral tissues and plasma RBP4 levels positively correlate with retinol levels.

A potential link between RBP4 and type 2 diabetes mellitus was suggested by Yang et al. [1]. The study reported that RBP4 was elevated in insulin-resistant mice with adipose tissue-specific GLUT4 knockout and humans with obesity and type 2 diabetes mellitus. They found that RBP4 expression was increased in insulin resistant adipose tissue specific GLUT4 null mice and reduced in insulin sensitive adipose specific GLUT4 transgenss mice. Transgenic overexpression of human RBP4 or injection of recombinant RBP4 in normal mice causes insulin resistance and glucose intolerance. Conversely, genetic deletion of Rbp4 enhances insulin sensitivity.
Recent studies [2,3] in human found that plasma RBP4 levels were elevated in subjects with IGT or type 2 diabetes mellitus and that RBP4 was related to various clinical parameters known to be associated with insulin resistance. And a cross-sectional study conducted in Chinese found that increased RBP4 levels increased the risk for hyperglycemia, including impaired glucose regulation and newly diagnosed type 2 diabetes mellitus, even after adjustment for a number of confounders [4]. These studies suggested that RBP4 was an adverse impact factor of diabetes mellitus.

Some researchers explored the relationship of RBP4 and insulin resistance. It was reported that serum RBP4 levels associated negatively with insulin sensitivity (determined by a hyperinsulinemic euglycemic clamp) in non-diabetic [5] or impaired glucose metabolism participants [6] and in women with polycystic ovary syndrome [7] or in women with normal glucose tolerance with different obesity [8]. However, the associations between RBP4 levels and insulin resistance were not consistent. Several studies failed to find a significant correlation between RBP4 levels and insulin resistance [9,10]. The suggested explanations for these discrepancies include the differences in renal clearance of RBP4 caused by the different renal functions in the study subjects, the imbalance between RBP4 and retinol, the influence of the collection method, and the antibodies used in measuring RBP4 [11].

Since the relationship between RBP4 and insulin resistance was controversial, and impaired β-cell was another important pathogenesis of diabetes mellitus, the relationship between β cell function and RBP4 need to be further studied. However, few studies showed their relation at present.

Our earlier work found that, the first phase or early phase of insulin secretion were impaired in non-alcoholic fatty liver disease (NAFLD) patients when their hepatic fat content detected by 1H MRS or CT were more than 10%, which indicated that liver cell with excessive fat deposition may release some cytokines to affect the beta cell function [12,13]. Recent studies found that levels of serum RBP4 were increased in NAFLD patients [14], so we speculated that the serum RBP4 may be one of cytokines involved in the cross-talk of adipose tissue and beta cell function in patients with fatty liver.

Thus, the purpose of the present study was to investigate correlations between serum RBP4 levels and β-cell function in Chinese NAFLD subjects without known T2DM.

Results
The clinical characteristics of the subjects are shown in Tables 1 and 2. The women participants were older than men participants. Compared to women, men had higher waist circumference and waist-to-hip ratio (WHR), triglycerides (TG), alanine aminotransferase (ALT), γ-glutamyltranspeptidase (γ-GT), serum RBP4, hepatic fat content, and lower high-density lipoprotein cholesterol (HDL-c). There are no differences between men and women in body mass index (BMI), systolic and diastolic blood pressure, total cholesterol (TC), low-density lipoprotein cholesterol (LDL-c), fasting blood glucose (FBG) and 2 hour blood glucose (2hBG), fasting insulin (FINS) and 2 hour insulin (2hINS), HOMA-IR, HOMA-β, the insulin sensitivity index (ISI), First-phase insulin secretion (1PH), Increment of 1PH, Second-phase insulin secretion (2PH), Increment of 2PH, aspartate aminotransferase (AST), alkaline phosphatase (ALP).

In all the subjects, serum RBP4 levels were significantly associated with sex, waist circumference, WHR, SBP, DBP, TG, HDL-c, γ-GT, but not associated with age, BMI, TC, LDL-c, FBG, 2hBG, FINS, 2hINS, HOMA-IR, HOMA-β, ALT, AST, ALP, HFC. ISI, 1PH, Increment of 1PH, 2PH, Increment of 2PH. In women, serum RBP4 levels were significantly associated with FBG, HOMA-β, Increment of 1PH; but not associated with age, BMI, and the other variables mentioned above. While in men, serum RBP4 levels were significantly associated with HDL-c, ALT, AST, but not associated with age, BMI, and the other variables mentioned above, either (Tables 3 and 4).

A stepwise multiple linear regression analysis was performed using HOMA-β as a dependent variable. RBP4 was selected as an independent variable together with age, BMI, HFC, ALT, AST, γ-GT, waist circumference, WHR, SBP, DBP, TC, TG, HDL-c, LDL-c, HOMA-IR in all subjects. We also performed stepwise multiple linear regression analysis within each gender. In all subjects, HOMA-IR (Standardized Coefficients Beta = 0.626, p < 0.001) together with age (Standardized Coefficients Beta = -0.245, p = 0.013) were determined as significant variables associated with HOMA-β. In women, HOMA-IR (Standardized Coefficients Beta = 0.521, p = 0.001) and RBP4 (Standardized Coefficients Beta = 0.415, p = 0.005) were significant variables associated with HOMA-β. While in men, HOMA-IR (Standardized Coefficients Beta = 0.591, p < 0.001) and BMI (Standardized Coefficients Beta = 0.328, p = 0.013) were significant variables associated with HOMA-β (Table 5).

Discussion
In the current study we found that serum RBP4 levels were inversely associated with β cell function in a cohort of Chinese women NAFLD patients without known diabetes mellitus. To the best of our knowledge, this study is the first showing a quantitative correlation between serum RBP4 levels and beta cell function assessed not only by HOMA-β but also by hyperglycemic clamp in Chinese NAFLD patients. Our study added to the growing evidence that serotonin RBP4, which secreted mainly by liver
and adipose tissue, was a potential adverse impact factor for beta cell function and diabetes mellitus. In addition, the serum RBP4 might be one of cytokines involved in the cross-talk between adipose tissue and beta cell function in patients with fatty liver.

So far, a total of 3 articles reported the relationship between circulating RBP4 and β cell insulin secretion. Similarly, two studies have found that RBP4 negatively related to beta cell function. But the study population and methods to assess insulin secretion were different. Broch et al. reported that [15] circulating RBP4 was negatively associated with acute insulin response (AIRg) (r = -0.48, P = 0.007), calculated as the insulin area during the first 10 min of the frequently sampled intravenous glucose tolerance test, in 107 Spanish men without diabetes mellitus, especially in obese subjects. In multiple regression analyses to predict insulin secretion, RBP4 emerged as an independent factor that contributed independently to AIRg variance (23%) in obese but not in nonobese subjects. There were no related information about women in this study. Another study investigated 867 non-obese Chinese patients with NGT [16] and showed that serum RBP4 correlated with glucose-stimulated insulin secretion, assessed by ΔI30/ΔG30 (increment in plasma insulin concentration plasma glucose concentration 30 min after the oral administration of 75 g glucose) and the total area

| Table 1 Basic characteristics of all the subjects |
|-----------------------------------------------|
| Total (n = 106) | Men (n = 61) | Women (n = 45) | P value |
|-----------------------------------------------|
| Age (y) | 47.39 ± 13.88 | 44.70 ± 15.21 | 50.98 ± 11.04 | 0.016 |
| BMI (kg/m²) | 26.69 ± 3.48 | 26.90 ± 3.45 | 26.39 ± 3.54 | 0.459 |
| Waist (cm) | 91.88 ± 10.04 | 94.26 ± 9.59 | 88.64 ± 9.82 | 0.004 |
| WHR | 0.92 ± 0.07 | 0.95 ± 0.06 | 0.89 ± 0.07 | <0.001 |
| Systolic blood pressure (mmHg) | 129.05 ± 19.57 | 130.67 ± 16.57 | 126.84 ± 23.04 | 0.322 |
| Diastolic blood pressure (mmHg) | 81.45 ± 11.25 | 82.93 ± 10.50 | 79.44 ± 12.02 | 0.115 |
| Total cholesterol (mmol/L) | 4.97 ± 0.83 | 4.93 ± 0.77 | 5.03 ± 0.92 | 0.532 |
| Triglycerides (mmol/L) | 2.15 ± 1.22 | 2.54 ± 1.37 | 1.63 ± 0.74 | <0.001 |
| HDL-c (mmol/L) | 1.16 ± 0.26 | 1.06 ± 0.18 | 1.30 ± 0.27 | <0.001 |
| LDL-c (mmol/L) | 2.89 ± 0.69 | 2.81 ± 0.62 | 2.98 ± 0.76 | 0.217 |
| Fasting blood glucose (mmol/L) | 5.33 ± 0.76 | 5.31 ± 0.80 | 5.35 ± 0.72 | 0.832 |
| 2 h blood glucose (mmol/L) | 7.30 ± 2.52 | 7.08 ± 2.51 | 7.62 ± 2.53 | 0.289 |
| Fasting insulin (mU/L) | 10.85 ± 6.95 | 10.29 ± 7.01 | 11.61 ± 6.89 | 0.354 |
| 2 h insulin (mU/L) | 80.51 ± 61.67 | 77.54 ± 66.68 | 84.74 ± 54.30 | 0.574 |
| HOMA-IR | 2.62 ± 1.76 | 2.46 ± 1.81 | 2.84 ± 1.70 | 0.289 |
| HOMA-β | 138.36 ± 110.55 | 136.84 ± 122.41 | 140.42 ± 93.48 | 0.874 |
| ALT (U/L) | 47.23 ± 43.10 | 56.64 ± 47.86 | 34.82 ± 32.42 | 0.011 |
| AST (U/L) | 33.12 ± 20.13 | 34.51 ± 19.96 | 31.24 ± 20.45 | 0.427 |
| ALP (U/L) | 73.06 ± 20.76 | 74.87 ± 21.86 | 70.68 ± 19.22 | 0.333 |
| γ-GT (U/L) | 49.79 ± 48.67 | 60.62 ± 58.24 | 35.54 ± 26.57 | 0.006 |
| Serum RBP4 (μg/ml) | 51.65 ± 11.85 | 55.96 ± 11.14 | 45.87 ± 10.31 | <0.001 |
| Hepatic fat content (%) | 7.90 ± 6.89 | 9.10 ± 6.85 | 6.26 ± 6.66 | 0.035 |

Data are means ± S.D. BMI: body-mass index, WHR: waist-to-hip ratio, HDL-c: high-density lipoprotein cholesterol, LDL-c: low-density lipoprotein cholesterol, HOMA-IR: homeostatic model assessment of insulin resistance, HOMA-β: homeostatic model assessment of β cell function, ALT: alanine aminotransferase, AST: aspartate aminotransferase, γ-GT: γ-glutamyltransferase, RBP4: retinol binding protein 4.

| Table 2 Basic characteristics of NGT subjects underwent hyperglycemic clamp |
|-----------------------------------------------|
| Total (n = 17) | Men (n = 6) | Women (n = 11) | P value |
|-----------------------------------------------|
| Insulin sensitivity index (ISI) | 15.48 ± 14.41 | 13.55 ± 15.50 | 16.53 ± 14.45 | 0.698 |
| First-phase insulin secretion (1PH) | 288.32 ± 169.41 | 304.60 ± 161.52 | 279.45 ± 180.64 | 0.780 |
| Increment of 1PH | 77.45 ± 41.96 | 85.57 ± 33.54 | 73.01 ± 46.83 | 0.572 |
| Second-phase insulin secretion (2PH) | 120.33 ± 67.90 | 110.21 ± 50.41 | 125.84 ± 77.53 | 0.665 |
| Increment of 2PH | 207.75 ± 145.92 | 216.91 ± 104.04 | 202.76 ± 169.05 | 0.856 |
Table 3 Pearson’s correlation analysis of serum RBP4 in all the subjects

|                      | Total (n = 106) |         | Men (n = 61) |         | Women (n = 45) |         |
|----------------------|----------------|---------|--------------|---------|----------------|---------|
|                      | r    | P     | r    | P     | r    | P     |
| Gender               | −0.423 | 0.000 | 0.061 | 0.574 | 0.114 | 0.432 |
| Age (y)              | 0.048 | 0.653 | 0.034 | 0.814 | −0.018 | 0.915 |
| BMI (kg/m²)          | 0.218 | 0.040 | 0.063 | 0.632 | 0.121 | 0.468 |
| WHR                  | 0.343 | 0.001 | 0.139 | 0.332 | 0.264 | 0.109 |
| Systolic blood pressure (mmHg) | 0.247 | 0.020 | 0.138 | 0.332 | 0.299 | 0.068 |
| Diastolic blood pressure (mmHg) | 0.241 | 0.023 | 0.196 | 0.167 | 0.124 | 0.458 |
| Total cholesterol (mmol/L) | 0.095 | 0.382 | 0.175 | 0.228 | 0.076 | 0.652 |
| Triglycerides (mmol/L)³ | 0.330 | 0.002 | 0.124 | 0.398 | 0.188 | 0.257 |
| HDL-c (mmol/L)³ | −0.392 | <0.001 | −0.281 | 0.059 | −0.183 | 0.279 |
| LDL-c (mmol/L)       | 0.049 | 0.658 | 0.108 | 0.476 | 0.113 | 0.504 |
| Fasting blood glucose (mmol/L)³ | 0.063 | 0.556 | −0.123 | 0.390 | 0.322 | 0.048 |
| 2 h blood glucose (mmol/L)³ | −0.045 | 0.676 | −0.142 | 0.322 | 0.277 | 0.097 |
| Fasting insulin (mU/L)³ | −0.051 | 0.641 | 0.164 | 0.266 | −0.271 | 0.104 |
| 2 h insulin (mU/L)³ | −0.039 | 0.729 | −0.089 | 0.546 | −0.010 | 0.954 |
| HOMA-IR³ | −0.095 | 0.387 | 0.119 | 0.422 | −0.211 | 0.210 |
| HOMA-β³ | −0.101 | 0.360 | 0.106 | 0.472 | −0.326 | 0.049 |
| ALT (U/L)³          | 0.105 | 0.335 | −0.287 | 0.048 | 0.168 | 0.312 |
| AST (U/L)³          | −0.077 | 0.487 | −0.304 | 0.038 | 0.055 | 0.752 |
| ALP (U/L)            | 0.116 | 0.310 | −0.007 | 0.962 | 0.171 | 0.326 |
| γ-GT (U/L)³         | 0.239 | 0.034 | 0.142 | 0.357 | 0.042 | 0.810 |
| Hepatic fat content (%)³ | 0.199 | 0.071 | 0.048 | 0.737 | 0.160 | 0.339 |

Data are means ± S.D. BMI: body-mass index, WHR: waist-to-hip ratio, HDL-c: high-density lipoprotein cholesterol, LDL-c: low-density lipoprotein cholesterol, HOMA-IR: homeostatic model assessment of insulin resistance, HOMA-β: homeostatic model assessment of β cell function, ALT: alanine aminotransferase, AST: aspartate aminotransferase, γ-GT: γ-glutamyltransferase, RBP4: retinol binding protein 4. Log: logarithmic transformation with 10 as its base of skewed variables.

under the curve for insulin over 180 min (AUC-I), in NGT non-visceral obesity subjects, but not in NGT visceral obesity subjects and T2DM patients.

This study and the two studies mentioned above all demonstrated that circulating RBP4 may negatively affect β cell function in different subjects. On the contrary, Stefan et al. found that [17] circulating RBP4 did not correlate with insulin secretion (measured by the IVGTT and OGTT) before or after adjustment for age, sex, and insulin sensitivity in a total of 75 Caucasians without type 2 diabetes mellitus. One of the reasons for this difference may due to different study subjects, another reason may be the excessive adjustment for gender. If the author analyzed the relationship of RBP4 and β cell function in men or women respectively, there might be different results.

It is well known that retinol is pathophysiologically linked to β cell function [18,19]. On the other hand, retinol-binding protein circulates in serum, forming a complex with transthyretin (TTR), a transport protein for thyroxine. An investigation disclosed that TTR constitutes a functional component in pancreatic β cell stimul-
secretion coupling [20]. TTR inhibits the binding of RBP to the receptor [21]. Thus, it is possible that increased serum RBP4 prevents TTR from exerting its β-cell stimulus-secretion effects. In fact, circulating RBP4 is highly bound to TTR in a one-for-one stoichiometric ratio, and there is little or no "free RBP4" in circulation [22]. In a recent study [23] using gel filtration chromatography to analyze the RBP4-TTR complex, it was found that increased serum RBP4 remains bound to TTR in insulin resistant states. Because the affinity of RBP4-TTR binding is very strong [22], the relative stoichiometry and affinity of the two proteins in serum could conceivably influence kinetics of RBP4-antibody binding.

The concentration of serum RBP4 varies by gender, generally lower in women than in men, both among adults and children [2,24,25]. Several previous studies hypothesized that sex hormones may play a role in the gender discrepancy. In support of this hypothesis, RBP4 levels were shown to be different between premenopausal and postmenopausal women [26] and vary according to pubertal status [27]. Although serum RBP4 concentration was higher in men than in women, whereas serum RBP4 didn’t show adverse effect on β cell function of men, which suggested there may be some protective factors, such as testosterone, against RBP4 adverse effect on β cell. McInnes et al. found that androgen receptor (AR) signaling in adipocytes not only protects against high-fat diet induced visceral obesity but also regulates insulin action and glucose homeostasis, independent of adiposity. Androgen deficiency in adipocytes in mice resembles human type 2 diabetes mellitus, with early insulin resistance and evolving insulin deficiency [28]. Palomar et al. found [29] that testosterone, other than progesterone nor estradiol, protects the β cells of men pancreas against STZ-induced apoptosis, which is sex specific and could be mediated through the induction of catalase and Cu/Zn-SOD. It demonstrated that androgens may protect β cell function in men.

The relationship about RBP4 negatively related to beta cell function, restricted to some, but not all of the study population, which suggested that their relationship was complicated and influenced by many other confounding factors and deserved further research.

Limitations
Firstly, the cross-sectional design of the study represents a limitation, implicating that cause-and-effect relationships cannot be discerned. Secondly, we used CT as a tool to determine the hepatic fat content, and its diagnostic accuracy is inferior to pathological diagnosis by liver biopsy. Thirdly, we did not measure blood glucose and insulin at 30 min, so we could not analyze the relationship of serum RBP4 and early phase of insulin secretion. Finally, due to the limitation of conditions, there were only 17 NGT patients undertook clamps in this study. Hence, correlation analysis of serum RBP4 and the increment of 1PH was underpowered because the sample size was too small. So, the data came from hyperglycemic clamp could just be considered as a clue, the real relationship of RBP4 and 1PH or increment of 1PH needs further verification by large-scale studies.

In summary, our results suggest that serum RBP4 may negatively affect β-cell function in Chinese women NAFLD patients without known diabetes mellitus.

Subjects, materials and methods
Subjects
This is a cross-sectional study. A total of 106 subjects of Chinese origin (HanChinese) (61 men and 45 women) diagnosed as fatty liver by ultrasound from the outpatient department of Endocrinology of Zhongshan hospital, Fudan university between Nov. 2005 and Apr. 2006 were recruited for this study.

Subjects with excessive alcohol consumption (≥30 g/d in men and ≥20 g/d in women) were excluded from the study. Subjects were also excluded if they had known diabetes mellitus, hepatitis B or C or other liver diseases,
or if they were receiving hypoglycemic or lipid-regulating (statins, fibrates) drugs, or other drugs that may impact glucose and lipid metabolism. Other exclusion factors were diseases affecting the metabolic state or not suitable to participate in this study, for example, hyperthyroidism, hypothyroidism, mental disease, serious disease with dysfunction of heart, liver, kidney, and cancer, etc.

All subjects underwent comprehensive physical examinations and routine biochemical analyses of blood. A 75-g oral glucose tolerance test (OGTT) with a test of 2 time-points (0 and 120 min post glucose loading) was performed. Among all subjects, 59 had normal glucose regulation (NGR), 13 had isolated-impaired fasting glucose (I-IFG), and 15 had isolated-impaired glucose tolerance (I-IGT), 5 had IFG and IGT, 14 had new diagnosed diabetes mellitus.

The study was approved by the human research ethics committee of Zhongshan hospital, Fudan University and informed consent was obtained from all subjects.

**Anthropometric and biochemical measurements**

Body mass index (BMI) was calculated as body weight in kilograms divided by the square of height in meters. Waist circumference was measured at the midpoint between the inferior costal margin and the superior border of the iliac crest on the mid axillary line.

Blood samples were drawn after an overnight fast and immediately centrifuged. Serum glucose was measured by the glucoseoxidase method. Serum total cholesterol (TC), triglycerides (TG), high-density lipoprotein cholesterol (HDL-c), low-density lipoprotein cholesterol (LDL-c), and liver enzyme levels were determined by enzymatic methods with a Hitachi 7600 analyzer (Hitachi, Ltd. Tokyo, Japan). Serum insulin (mU/L) was measured with an insulin radioimmunoassay kit (kit from Beijing North Biotechnology Research Institute), which had a reactivity of less than 0.2% to human proinsulin. The inter-assay and intra-assay coefficients of variation were 5.8% and 11.0% for insulin.

Insulin resistance (IR) and β cell function were estimated using the homeostasis model of assessment (HOMA). HOMA-IR was calculated as fasting glucose (mmol/L) × fasting insulin (mU/L)/22.5 and HOMA-β was calculated as 20 × fasting insulin (mU/L)/[fasting glucose (mmol/L)-3.5].

Serum RBP4 (μg/ml) was measured in duplicate by a sandwich ELISA developed in-house, using affinity-chromatography purified polyclonal and monoclonal antibodies generated against recombinant human RBP4 protein. The assay system was subsequently cross-validated by Western blot analysis. The intra-assay coefficient of variation was 1.8–7.6% and inter-assay was 3.7–8.85% [30].

**Hyperglycemic clamp**

A total of 17 NGT subjects (men = 6, women = 11) underwent hyperglycemic clamp. None of the subjects was taking any medications known to affect glucose and lipid metabolism. Subjects fasted 12 h before the experiments, in the experiments morning, a dorsal hand vein was cannulated in a retrograde fashion and kept in a constant-temperature device at 50-55°C for sampling arterialized venous blood. An antecubital vein was cannulated for infusion of 20% glucose. After a 30-min baseline period, serum glucose concentrations were increased to 13 mmol/L and subsequently maintained at this level for 150 min using the glucose clamp technique. During the experiments, blood samples for serum insulin determinations were obtained at -30,-15, 0, 2, 4, 6, 8, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, and 150 min [31].

**Calculations**

First-phase insulin secretion (1PH) was considered the sum of the serum insulin concentration from 2–10 min of the hyperglycemic clamp. Increment of 1PH was calculated as 1PH subtracting basal insulin level. Second-phase insulin secretion (2PH) was calculated as the average serum insulin concentration from 20–150 min of the clamp. Increment of 2PH was calculated as 2PH subtracting basal insulin level. The insulin sensitivity index (ISI) was determined as the average glucose metabolized rate during the last 30 min of the hyperglycemic clamp divided by the average serum insulin concentration during the same interval.

**Determination of hepatic fat content with calibrated CT**

CT scans was performed using a Marconi MX8000 machine Philips by the same one experienced doctor to measure hepatic fat content of patients. CT scan was performed at a voltage of 120 kv, electricity of 292 mA, and a slice of 6.5 mm with scan time of 3 seconds. Four slices were acquired in each patient at the following levels: dome of the liver, cephalad and caudal to the hepatic hilum, and the inferior margin of the right lobe. No contrast medium was used. The average of the four Hounsfield values obtained for each slice was measured.

Test tubes containing solutions with CT scan density (CTD) similar to normal liver (“liver-equivalent”) or “fat-equivalent material” in variable proportions were prepared to mimic different degrees of hepatic fat content. At the time of the CT measurement, the test tubes were placed on the abdomen of the patient, the CT numbers of the calibrating solutions were measured at the beginning of the exam, concurrently with those of the liver. A linear regression of the relative fat content of the test tubes solution was then calculated, and the CT number of the liver of the patient derived according to the regression line. The hepatic fat content (HFC) was expressed as a percentage of the entire liver [32].
Statistical analyses
All analyses were performed using Statistical Package for the Social Sciences version 16.0 (SPSS Inc., Chicago, Illinois, USA). Normally distributed data were expressed as mean ± SD. Data that were not normally distributed, as determined using the Kolmogorov-Smirnov test, were logarithmically transformed before analysis. The Student’s unpaired t test was used for comparison between two groups. Pearson’s correlation was used to analyze the relation of serum RBP4 and other parameters. Multiple stepwise regression analysis was used to examine the association of HOMA-β and serum RBP4 and other parameters. In all statistical tests, P values less than 0.05 were considered significant.

Abbreviations
RBP4: Retinol binding protein 4; NAFLD: Non-alcoholic fatty liver disease; FPG: Fasting blood glucose; 1PH: Firstphase insulin secretion; 2PH: Second-phase insulin secretion; WHR: Waist-to-hip ratio; SBP: Systolic blood pressure; DBP: Diastolic blood pressure; HFC: Hepatic fat content; ISI: Insulin sensitivity index; OGTT: Oral glucose tolerance test; NGR: Normal glucose regulation; FBG: Fasting blood glucose; 1PH: First phase insulin secretion; 2PH: Second phase insulin secretion; AST: Aspartate aminotransferase; ALP: Alkaline phosphatase; y-GT: γ-glutamyltransferase; FINS: Fasting insulin; 2hINS: 2 hour insulin; AUC-I: Area under the curve for insulin.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
XG designed the research and revised the manuscript; HY and XC carried out recruiting patients to the study. We thank Prof. Pu Xia for critical reading and editing the manuscript, and HY and XC carried out recruiting patients to the study. We thank Prof. Pu Xia for critical reading and editing the manuscript, and HY and XC carried out recruiting patients to the study.

Acknowledgements
We thank Prof. Pu Xia for critical reading and editing the manuscript, and the colleagues in Zhongshan Hospital, Fudan University for assistance in recruiting patients to the study.

Received: 19 July 2013 Accepted: 25 October 2013

References
1. Yang Q, Graham TE, Mody N, Preitner F, Peroni OD, Zabolotny JM, Kotani K, Quadro L, Kahn BB. Serum retinol binding protein 4 contributes to insulin resistance in obesity and type 2 diabetes. Nature 2005, 436:356–362.
2. Cho YM, Youn BS, Lee H, Lee N, Min SS, Kwak SH, Lee HK, Park KG. Plasma retinol-binding protein-4 concentrations are elevated in human subjects with impaired glucose tolerance and type 2 diabetes. Diabetes Care 2006, 29:2457–2461.
3. Meisinger C, Ruckert IM, Rathmann W, Döring A, Elveback LR, von der Decken H. Plasma retinol-binding protein-4 concentrations are elevated in human subjects with impaired glucose tolerance and type 2 diabetes. Diabetes Care 2006, 29:2457–2461.
4. Xu M, Li XY, Wang JG, Wang XJ, Huang Y, Cheng Q, Huang HE, Li R, Xiang J, Tan JR, et al. Retinol-binding protein 4 is associated with impaired glucose regulation and microalbuminuria in a Chinese population. Diabetologia 2009, 52:1511–1519.
5. Gavi S, Stuart LM, Kelly P, Melendez MW, Mynarcik DC, Gelato MC, McNurlan MA. Retinol-binding protein 4 is associated with insulin resistance and body fat distribution in nonobese subjects without type 2 diabetes. J Clin Endocrinol Metab 2007, 92:1886–1890.
6. Yang Q, Eksurka I, Kiernan UA, Phillips DJ, Bluerer M, Graham TE, Kahn BB. Quantitative measurement of full-length and C-terminal Proteolyzed RBP4 in serum of normal and insulin-resistant humans using a novel mass spectrometry immunomass. Endocrinology 2012, 153:1519–1527.
7. Weiping L, Qingfeng C, Shikun M, Xiong L, Hua Q, Xiaoshu B, Subha Z, Qifu L. Elevated serum RBP4 is associated with insulin resistance in women with polycystic ovary syndrome. Endocrine 2006, 30:283–287.
8. Kowalska I, Straczkowski M, Adamaska A, Nikolajuk A, Karczewska-Kupczewska M, Ozioke M, Gorska M. Serum retinol binding protein 4 is related to insulin resistance and nonoxidative glucose metabolism in lean and obese women with normal glucose tolerance. J Clin Endocrinol Metab 2008, 93:2782–2789.
9. Wallenius V, Elias E, Bergstrom GA, Zetterberg H, Behre CJ. The lipocalins retinol-binding protein-4, lipocalin-2 and lipocalin-type prostaglandin D2-synthase correlate with markers of inflammatory activity, alcohol intake and blood lipids, but not with insulin sensitivity in metabolically healthy SB-year-old Swedish men. Exp Clin Endocrinol Diabetes 2011, 119:75–80.
10. Chavez AG, Coletta DK, Kamath S, Cromack DT, Moroney A, Foll F, Defronzo RA, Tripathy D. J Retinol-binding protein 4 is associated with impaired glucose tolerance but not with whole body or hepatic insulin resistance in Mexican Americans. Am J Physiol Endocrinol Metab 2009, 296:E758–E764.
11. Kotschi P, Fischer-Pospavyzky P, Wabitsch M: RBP4: a controversial adipokine. Eur J Endocrinol 2011, 165:703–711.
12. Hongmei Y, Xin G, Liu M, Qian GJ, Jian GAO. Changes of β-cell function and insulin sensitivity in subjects with non-alcoholic fatty liver disease and normal glucose regulation. Chin J Endocrinol Metab 2007, 1:12–15.
13. Bian H, Lin H, Rao S, Yao X, Zeng M, Zhou J, Weiping J, Gao X. Association of liver fat content with insulin resistance and islet β-cell function in individuals with various statuses of glucose metabolism. Chin J Endocrinol Metab 2010, 26:535–540.
14. Seo JA, Kim NH, Park SY, Kim HY, Ryu OH, Lee KW, Lee J, Kim DL, Choi KM, Baik SH, et al. J Serum retinol-binding protein 4 levels are elevated in non-alcoholic fatty liver disease. Clin Endocrinol (Oxf) 2008, 68:555–560.
15. Broch M, Vendrell J, Ricart W, Richart C, Fernandez-Real JM. Circulating retinol-binding-protein 4, insulin sensitivity, insulin secretion, and insulin disposition index in obese and nonobese subjects. Diabetes Care 2007, 30:1802–1806.
16. Li L, Wang C, Bao Y, Wu H, Lu J, Xiang K, Jia W. Serum retinol-binding protein 4 is associated with insulin secretion in Chinese people with normal glucose tolerance. J Diabetes 2009, 1:125–130.
17. Stefan N, Henning AM, Staiser H, Schlecher E, Fritsche A, Haring HU. Circulating retinol-binding protein-4, insulin sensitivity, insulin secretion, and insulin disposition index in obese and nonobese subjects: response to Broch et al. Diabetes Care 2007, 30:e91. author reply e92.
18. Blaner WS, STRA6, a cell-surface receptor for retinol-binding protein: the plot thickens. Cell Metab 2007, 5:164–166.
19. Chertov BS. Effects of vitamin a deficiency and repletion on Rat insulin secretion in vivo and in vitro from isolated islets. J Clin Invest 1987, 79:163–169.
20. Refai E, DeKKi N, Yang SN, Inreh G, Cabrera O, Yu L, Yang G, Norgren S, Rossner SM, Inverardi L. The interaction of retinol-binding protein with its receptor: a plot thickens. Cell Metab 2007, 5:164–166.
21. Sivaprasadarao AFJ. Plasma retinol binding protein: structure and interactions with retinol, retinoids, and transthyretin. Vitam Horm 2004, 69:271–295.
22. Graham TE, Wason CJ, Bluher M, Kahn BB. Shortcomings in methodology: Circulating retinol binding protein: the plot thickens. Cell Metab 2007, 5:164–166.
23. Lee DC, Lee JW, Im JA: Association of serum retinol binding protein 4 and insulin resistance in apparently healthy adolescents. Metabolism 2007, 56:327–331.
25. Balagopal P, Graham TE, Kahn BB, Altomare A, Funanage V, George D: Reduction of elevated serum retinol binding protein in obese children by lifestyle intervention: association with subclinical inflammation. J Clin Endocrinol Metab 2007, 92:1971–1974.

26. An C, Wang H, Liu X, Li Y, Su Y, Gao X, Jiang W: Serum retinol-binding protein 4 is elevated and positively associated with insulin resistance in postmenopausal women. Endocr J 2009, 56:987–996.

27. Reinehr T, Stoffel-Wagner B, Roth CL: Retinol-binding protein 4 and its relation to insulin resistance in obese children before and after weight loss. J Clin Endocrinol Metab 2008, 93:2287–2293.

28. McInnes KJ, Smith LB, Hunger NI, Saunders PTK, Andrew R, Walker BR: Deletion of the androgen receptor in adipose tissue in male mice elevates retinol binding protein 4 and reveals independent effects on visceral fat mass and on glucose homeostasis. Diabetes 2012, 61:1072–1081.

29. Palomar-Morales MMS, Mendoza-Rodríguez CA, Cerbón MA: The protective effect of testosterone on streptozotocin-induced apoptosis in beta cells is sex specific. Pancreas 2010, 39:193–200.

30. Qi Q, Yu Z, Ye X, Zhao F, Huang P, Hu FB, Franco OH, Wang J, Li H, Liu Y, Lin X: Elevated retinol-binding protein 4 levels are associated with metabolic syndrome in Chinese people. J Clin Endocrinol Metab 2007, 92:4827–4834.

31. Li H, Bao Y, Xu A, Pan X, Lu J, Wu H, Lu H, Xiang K, Jia W: Serum fibroblast growth factor 21 is associated with adverse lipid profiles and gamma-glutamyltransferase but not insulin sensitivity in Chinese subjects. J Clin Endocrinol Metab 2009, 94:2151–2156.

32. Ricci CLR, Gioulis E, Bosco M, Pollesello P, Masutti F, Cocco LS, Paolotti S, de Bernardi B, Tiribelli C, Dalla Palma L: Non-invasive in vivo quantitative assessment of fat content in human liver. J Hepatol 1997, 27:108–113.

Cite this article as: Yan et al.: Serum retinol binding protein 4 is negatively related to beta cell function in Chinese women with non-alcoholic fatty liver disease: a cross-sectional study. Lipids in Health and Disease 2013 12:157.