Evaluation of Inotropic Activity of Fluorobenzene Derivative Using an Isolated Rat Heart Model

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Abstract

There are studies which indicate that some fluorobenzene derivatives have inotropic activity; nevertheless, the cellular site and mechanism of action at cardiovascular level is very confusing. To clarify these phenomena in this study, a new fluorobenzene derivative was synthesized to evaluate its biological activity on perfusion pressure and left ventricular pressure. The Langendorff technique was used to measure changes on perfusion pressure and coronary resistance in an isolated rat heart model in absence or presence of the fluorobenzene derivative [0.001 nM]. Additionally, to characterize the molecular mechanism involved in the inotropic activity induced by the fluorobenzene derivative was evaluated by measuring left ventricular pressure in absence or presence of following compounds; ouabain, digoxin, levosimendan, cyclopiazonic acid and thapsigargin. The results showed that the fluorobenzene derivative significantly increased the perfusion pressure and coronary resistance in comparison with the control conditions. Additionally, other data indicate that fluorobenzene derivative increase perfusion pressure in a form similar to ouabain and digoxin; however, this effect was different compared with levosimendan. Other results showed that biological activity induced by the fluorobenzene on left ventricular pressure was significantly inhibited by both cyclopiazonic acid [50 mM] and thapsigargin [300 mM]. These data suggest that positive inotropic activity induced by the fluorobenzene on perfusion pressure and left ventricular pressure was via changes of biological activity both Na,K-ATPase and Ca²⁺-ATPase. This phenomenon is a particularly interesting because the positive inotropic activity induced by this fluorobenzene derivative involves a molecular mechanism different in comparison with other positive inotropic drugs.

Keywords
Fluorobenzene, Derivative, Inotropic Activity, ATPase

1. Introduction

Congestive heart failure is one of the main health problems worldwide [1-3]. Since several years ago several drugs with positive inotropic activity have been used for the treatment of congestive heart failure [4, 5]. For example, the use of digoxin (ATP-ase inhibitor) in patients with heart failure; however, its narrow therapeutic window and even slight exposure changes have been associated with adverse reactions; this has resulted in close monitoring of digoxin serum levels in patients with heart failure [6-8]. This problem has resulted in the use of other therapeutic alternatives such as dobutamine (adrenergic agonist); however, its poor oral bioavailability results in this drug being administered intravenously [9]. This has led to studies comparing the effect of dobutamine with levosimendan (positive inotropic, calcium sensitizer); the results showed an improvement in hemodynamic function with levosimendan compared to the group treated with dobutamine [10]. However, it has been observed that high doses of levosimendan in patients with episodes of myocardial infarction may induce changes in the inotropic effect by inhibiting the activity of some phosphodiesterases [11]. In this latter aspect, it is important to mention that there are other types of drugs with positive inotropic activity, which act by inhibiting the isoenzyme III of phosphodiesterase’s [12-14]. For example, the milrinone at a dose of 0.75 μg / kg per minute increased cardiac output and reduce systemic vascular resistance in patients with...
heart failure [15]; however, it can increase the incidence of ischemic heart disease and ventricular arrhythmias in a dose-dependent manner [16]. In addition, other drugs for heart failure have been used such as angiotensin converting enzyme inhibitors and spironolactone [17-20]; however, there data which indicate that the abrupt increase in the prescription of spironolactone can induce hypercalcemia [21]. All these data have led to the search for new therapeutic alternatives for the treatment of heart failure; for example, a study showed that a spiro cycloalkane one induce positive inotropic activity via phosphodiesterase inhibition in an animal model [22]. Other report showed that the compound OPC-18790 is a positive inotropic and vasodilating agent which increases intracellular cyclic AMP and stimulates intracellular Ca^{2+} currents in an isolated dog heart model [23]. In addition, a study indicated that the compound YS-49 (1,2,3,4-Tetrahydro-1-(1-naphthalenylmethyl)-6,7-isoquinolinediol hydrobromide monohydrate) induce positive inotropic activity in rat vascular smooth muscle via nitric oxide synthase inhibition [24]. Other data indicate that showed that a fluorobenzene derivative (piperazino-fluorobenzene) exert both intropic and chronotropic activity in isolated guinea-pig atria preparations via phosphodiesterase’s inhibition [25]. All these data show that several compounds can induce inotropic effects in the cardiovascular system; however, there are few reports on the inotropic activity exerted by the fluorobenzene derivatives. To provide more information about this phenomenon, this study was designed to investigate the effects of a fluorobenzene derivative on perfusion pressure and coronary resistance in isolated rat hearts using the Langendorff technique [26]. In addition, to evaluate the molecular mechanism involved in the inotropic activity induced by the fluorobenzene derivative on left ventricular pressure several drugs were used as pharmacological tools.

2. Methods

Synthesis Chemical

General Methods

All the compounds used in this work were purchased from Sigma-Aldrich Co. Ltd. The melting points for the different compounds were determined on an Electrothermal (900 models). Flash chromatography was performed on silica gel 60 (0.040-0.063 mm, Merck). Infrared spectra (IR) were recorded using KBr pellets on a Perkin Elmer Lambda 40 spectrometer. 1H and 13C NMR spectra were recorded on a Varian VXR-300/5 FT NMR spectrometer at 300 and 75.4 MHz in CDCl3 using TMS as internal standard. EIMS spectra were obtained with a Finnigan Trace GC Polaris Q. Spectra were recorded on a Varian VXR-300/5 FT NMR spectrometer. All drugs were dissolved in methanol and different dilutions were obtained using Krebs-Henseleit solution (≤ 0.01%, v/v).

Experimental Design

All experimental procedures and protocols used in this investigation were reviewed and approved by the Animal care and use Committee of University Autonomous of Campeche (No. PI-420/12) and were in accordance with the guide for the care and use of laboratory animals [27]. Male Wistar rats; weighing 200-250 g were obtained from University Autonomous of Campeche.

Reagents

A solution of 2,4-dinitrofluorobenzene (100 mg, 0.54 mmol) eugenol (212 µl, 1.21 mmol), potassium carbonate (4:2:1) purified by crystallization from methanol: hexane: water (4:2:1).

Biological Methods

All experimental procedures and protocols used in this investigation were reviewed and approved by the Animal care and use Committee of University Autonomous of Campeche (No. PI-420/12) and were in accordance with the guide for the care and use of laboratory animals [27]. Male Wistar rats; weighing 200-250 g were obtained from University Autonomous of Campeche.

Reagents

All drugs were dissolved in methanol and different dilutions were obtained using Krebs-Henseleit solution (≤ 0.01%, v/v).

Experimental Design

Briefly, the male rat (200 - 250 g) was anesthetized by injecting them with pentobarbital at a dose rate of 50 mg/Kg body weight. Then the chest was opened, and a loose ligature passed through the ascending aorta. The heart was then rapidly removed and immersed in ice cold physiologic saline solution. The heart was trimmed of non-cardiac tissue and retrograde perfused via a non-circulating perfusion system at a constant flow rate. The perfusion medium was the Krebs-Henseleit solution (pH = 7.4, 37°C) composed of (mmol); 117.8 NaCl; 6 KCl; 1.75 CaCl2; 1.2 NaH2PO4; 1.2 MgSO4; 24.2 NaHCO3; 5 glucose and 5 sodium pyruvate. The solution was actively bubbled with a mixture of O2/CO2 (95:5/5 %). The coronary flow was adjusted with a variable speed peristaltic pump. An initial perfusion rate of 15 ml/min for 5 min was followed by a 15 min equilibration period at a perfusion rate of 10 ml/min. All experimental measurements were done after this equilibration period.
**Perfusion Pressure**

Evaluation of measurements of perfusion pressure changes induced by drugs administration in this study were assessed using a pressure transducer connected to the chamber where the hearts were mounted and the results entered into a computerized data capture system (Biopac).

**Inotropic Activity**

Contractile function was assessed by measuring left ventricular developed pressure (LV/dP), using a saline-filled latex balloon (0.01 mm, diameter) inserted into the left ventricle via the left atrium. The latex balloon was bound to cannula which was linked to pressure transducer that was connected with the MP100 data acquisition system.

**Biological Evaluation**

First stage

Ischemia/Reperfusion model

After of 15-minute equilibration time, the hearts were subjected to ischemia for 30 minutes by turning off the perfusion system [28]. After this period, the system was restarted and the hearts were reperfused by 30 minutes with Krebs-Henseleit solution. The hearts were randomly divided into 4 major treatment groups with n = 9:

**Group I.** Hearts were subjected to ischemia/reperfusion but received vehicle only (Krebs-Henseleit solution).** Group II.** Hearts were subjected to ischemia/reperfusion and treated with 3,5-dinitrobenzoic acid [0.001 nM] before ischemia period (for 10 minutes) and during the entire period of reperfusion.** Group III.** Hearts were subjected to ischemia/reperfusion and treated with the compound 2 [0.001 nM] before ischemia period (for 10 minutes) and during the entire period of reperfusion.** Group IV.** Hearts were subjected to ischemia/reperfusion and treated with the compound 3 [0.001 nM] before ischemia period (for 10 minutes) and during the entire period of reperfusion.

At the end of each experiment, the perfusion pump was stopped, and 0.5 ml of fluorescein solution (0.10%) was injected slowly through a sidearm port connected to the aortic cannula. The dye was passed through the heart for 10 sec to ensure its uniform tissue distribution. The presence of fluorescein was used to demarcate the tissue that was not subjected to regional ischemia, as opposed to the risk region. The heart was removed from the perfusion apparatus and cut into two transverse sections at right angles to the vertical axis. The right ventricle, apex, and atrial tissue were discarded. The areas of the normal left ventricle non risk region, area at risk, and infarct region were determined using the technique reported by [28]. Total area at risk was expressed as the percentage of the left ventricle.

Second Stage

**Effects induced by 3,5-dinitrobenzoic acid or compounds 2 or 3 on perfusion pressure:** Changes in perfusion pressure as a consequence of increases in time (3 to 18 min) in absence (control) and presence of 3,5-dinitrobenzoic acid or compounds 2 or 3 at a concentration of 0.001 nM were determined. The effects were obtained in isolated hearts perfused at a constant-flow rate of 10 ml/min.

**Evaluation of effects exerted by 3,5-dinitrobenzoic acid or compounds 2 or 3 on coronary resistance:** The coronary resistance in absence (control) and presence of 3,5-dinitrobenzoic acid or compounds 2 or 3 at a concentration of 0.001 nM was evaluated. The effects were obtained in isolated hearts perfused at a constant flow rate of 10 ml/min. Since a constant flow was used changes in coronary pressure reflected the changes in coronary resistance.

Third Stage

**Effects induced by noradrenaline or dobutamine or compound 3 on perfusion pressure via adrenergic.** Changes in perfusion pressure as a consequence of increases in time (3 to 18 min) in absence (control) and presence of the compound 3 [0.001 nM] or noradrenaline [1 nM] or dobutamine [1 nM] were determined. The effects were obtained in isolated hearts perfused at a constant-flow rate of 10 ml/min.

**Effects induced by digoxine or ouabaine or the compound 3 on perfusion pressure via K,Na-ATPase.** Biological activity exerted by digoxine [0.01 nM] or ouabaine [0.001 nM] or the compound 3 [0.001 nM] on perfusion pressure as a consequence of increases in time (3 to 18 min). The effects were obtained in isolated hearts perfused at a constant-flow rate of 10 ml/min.

**Effect exerted by the compound 3 on left ventricular pressure in presence or absence of cyclopiazonic acid or thapsigargin via Ca$^{2+}$-ATPase.** The compound 3 [0.001 to 100 nM] was administered (intracoronary boluses, 50 μl) and the corresponding effect on the left ventricular pressure was determined. The dose response curve (control) was repeated in the presence of cyclopiazonic acid [50 nM] or thapsigargin [300 nM] (duration of the pre-incubation with cyclopiazonic acid was for a period of 10 min).

**Evaluation theoretical of activity exerted by compound 3 on both Na,K-ATPase and Ca$^{2+}$-ATPase** Docking calculations were carried out using Docking Server [29]. The MMFF94 force field [30] was used for energy minimization of ligand molecule using the Docking Server. Gasteiger partial charges were added to the ligand atoms. Non-polar hydrogen atoms were merged, and rotatable bonds were defined. Docking calculations were carried out on the Na,K-ATPase (1KJU) [31] and Ca$^{2+}$-ATPase.
(3WGU) [32] protein models. Essential hydrogen atoms, Kollman united atom type charges, and solvation parameters were added with the aid of AutoDock tools. Affinity (grid) maps of 20 × 20 × 20-Å grid points and 0.375-Å spacing were generated using the Autogrid program. AutoDock parameter set and distance dependent dielectric functions were used in the calculation of the Vander Waals and the electrostatic terms, respectively. Docking simulations were performed using the Lamarckian genetic algorithm (LGA) and the Solis and Wets local search method [33]. Initial position, orientation, and torsions of the ligand molecules were set randomly. Each docking experiment was derived from two different runs that were set to terminate after a maximum of 250,000 energy evaluations. The population size was set to 150. During the search, a translational step of 0.2Å and quaternion and torsion steps of 5 were applied.

### Statistical Analysis

The obtained values are expressed as average ± SE, using each heart (n = 9) as its own control. The data obtained were put under Analysis of Variance (ANOVA) with the Bonferroni correction factor [34] using the SPSS 12.0 program. The differences were considered significant when p was equal or smaller than 0.05.

### 3. Results

#### Chemical Synthesis

The yield of compound 2 (fluoro-methoxybenzene derivative, Figure 1) was 44% with a melting point of 62-62 °C. In addition, the chemical shifts of the spectroscopic analyses of 1H NMR and 13C NMR for the compound 2 is shown in the table 1. Finally, the results of mass spectroscopy (EI-MS) (70 electron volts) shown; m/z 420.17. Additionally, the elementary analysis data for the compound 2 (C26H25FO4) were calculated (C, 74.27; H, 5.99; F, 4.52; O, 15.22) and found (C, 74.22; H, 5.90).

Other data showed a yield of 67% for the compound 3 (Fluoro-cyclobutenyl-2-ol derivative, Figure 1) with a melting point of 80-82 °C. 1H NMR and 13C NMR spectra for the compound 3 are shown in the table 1. Finally, the results of mass spectroscopy (MS) (70 electron volts) shown; m/z 616.32. Additionally, the elementary analysis data for the compound 3 (C38H45FO6) were calculated (C, 74.00; H, 7.35; F, 3.08; O, 15.56) and found (C, 74.00; H, 7.26).

![Figure 1](image-url) Preparation of the fluoro-cyclobutenyl-2-ol derivative (3). Reaction of 1-fluoro-2,4-dinitro-benzene (1) with eugenol (i) to form the fluoro-di-methoxybenzene derivative (2). Then, 2 was reacted with 1-hexyn-3-ol (ii) to synthesis of 3.

| Compound | 1H NMR (300 MHz, CDCl3) | 13C NMR (75.4 Hz, CDCl3) |
|----------|------------------------|-------------------------|
| Compound 2<br>δH: 3.30 (m, 4H), 3.80 (m, 4H), 5. 02 (d,d, 2H), 5.08 (d,d, 2H), 5.98 (d,d, 2H), 6.50-7.18 (m, 9H) ppm. | δC: 39.94, 55.94, 113.11, 113.97, 114.03, 114.09, 114.11, 115.19, 115.31, 115.96, 120.12, 121.1, 130.21, 133.85, 137.47, 145.21, 147.35, 148.25, 148.31, 149.7, 151.7 ppm. |
| Compound 3<br>δH: 0.96 (s, 6H), 1.16-2.28 (m, 8H), 2.50 (broad, 2H), 2.54-2.62 (m, 4H), 2.68-2.72 (m, 2H), 2.94 (m, 2H), 3.30 (m, 2H), 3.84 (s, 6H), 5.34 (d,d, 2H), 6.16-7.18 (m, 9H) ppm. | δC: 9.95, 32.28, 33.91, 36.9, 37.59, 47.33, 55.94, 71.30, 112.19, 113.18, 113.34, 114.11, 115.19, 115.31, 120.46, 121.44, 129.7, 133.32, 133.85, 145.21, 146.1, 147.08, 147.62, 147.68, 149.7, 151.7 ppm. |
Biological Activity

First Stage

In the Figure 2 is showed the biological activity exerted by the compound 3 on ischemia/reperfusion injury; the results showed that compound 3 reduce area of infarction to different dose [0.001 to 100 nM].

Other results (Figure 3) indicate that compound 3 significantly \( p = 0.05 \) reduces infarct size expressed as a percentage of the area at risk compared with 2,4-dinitrofluorobenzene, the compound 2 and vehicle treated hearts (control).

![Figure 2](image1.png)

**Figure 2.** Biological activity exerted by compound 3 at a dose of 0.001 (1e-3) to 100 nM on the functional recovery of rat hearts subjected to ischemia and reperfusion. The results indicate that the compound 3 reduce the area infarct to different dose compared with control conditions.

![Figure 3](image2.png)

**Figure 3.** Biological activity exerted by compound 3 or 2,4-dinitrofluorobenzene (2,4-DNFB) or compound 2 at dose of 0.001 nM on the functional recovery of rat hearts subjected to ischemia and reperfusion. The results indicate that the compound 3 significantly reduce \( p = 0.05 \) the area infarct to different dose compared with 2,4-DNFB, compound 2 and control conditions.

Second Stage

The activity induced by 2,4-dinitrofluorobenzene, the compounds 2 and 3 on perfusion pressure and coronary resistance in the isolated rat hearts was evaluated (Figure 4). The showed that compound 3 \( p = 0.05 \) significantly increase the perfusion pressure in comparison with 2,4-dinitrofluorobenzene, compound 2 [0.001 nM] and control conditions.
Figure 4. Effect induced by the compounds 2 or 3 or 2,4-dinitrofluorobenzene on perfusion pressure. The results showed that the compound 3 significantly increase perfusion pressure ($p = 0.05$) in a manner dependent of time in comparison with 2,4-dinitrofluorobenzene, the control conditions and compound 2. Each bar represents the mean ± S.E. of 9 experiments.

In addition, the coronary resistance (Figure 5), calculated as the ratio of perfusion pressure at coronary flow assayed (10 ml/min) was significantly higher ($p = 0.05$) in presence of the compound 3 [0.001 nM] in comparison with 2,4-dinitrofluorobenzene or the compounds 2 and control conditions.

Figure 5. Biological activity exerted by the compound 3 or 2,4-dinitrofluorobenzene or compound 2 at dose of 0.001 nM on coronary resistance. The results showed that coronary resistance was higher ($p = 0.05$) in the presence of the compound 3 compared with 2,4-dinitrofluorobenzene acid, the control conditions and compound 2. Each bar represents the mean ± S.E. of 9 experiments.
Third Stage

The Figure 6 show that perfusion pressure was significantly higher ($p = 0.05$) in presence of the compound 3 compared with norepinephrine and dobutamine in a manner dependent of time.

![Figure 6](image)

**Figure 6.** Biological activity exerted by norepinephrine, dobutamine and the compound 3 on perfusion pressure in a manner dependent of time. The results showed that perfusion pressure was significantly higher ($p = 0.05$) in presence of the compound 3 compared with norepinephrine and dobutamine. Each bar represents the mean ± S.E. of 9 experiments.

In addition, the Figure 7 showed that biological activity of the compound 3 on perfusion pressure was similar compared with digoxin and ouabain; however, significantly different ($p = 0.05$) to levosimendan and this effect was inhibited ($p = 0.05$) by cyclopiazonic acid and thapsigargin (Figure 8).

![Figure 7](image)

**Figure 7.** Effect induced by digoxin, ouabain, levosimendan and the compound 3 on perfusion pressure in a manner dependent of time. The results showed that compound 3 increasing the perfusion pressure through of time in a similar form that digoxin and ouabain. In addition this effect was significantly different ($p = 0.05$) that levosimendan. Each bar represents the mean ± S.E. of 9 experiments. LVP = left ventricular pressure
Intracoronary boluses (50 μl) of the compound 3 [0.001 to 100 nM] were administered and the corresponding effect on the LVP was determined in the absence and presence of cyclopiazonic acid or thapsigargin. The results showed that compound 3 increase the LVP in a dependent dose manner and this effect was significantly inhibited ($p = 0.05$) in the presence of cyclopiazonic acid and thapsigargin. Each bar represents the mean ± S.E. of 9 experiments. LVP = left ventricular pressure.

Other data showed (Figures 9 and 10) the interaction of the compound 3 with amino acids residues of both K,Na-ATP-ase and Ca$^{2+}$-ATP-ase

![Scheme showing the contact site of amino acid residues involved in the interaction of K,Na-ATP-ase with 3. Visualized with GL mol, viewer after docking analysis with one-click docking.](image-url)
4. Discussion

Chemical Synthesis

In this study we report a straightforward route for the preparation of a fluorobenzene-2,3-methoxy-di-phenoxy-cyclobuten-diol derivative (compound 3) using some strategies. The first stage was achieved by the preparation of a fluoro-methoxy-benzene derivative (compound 2) by the reaction of 2,4-dinotrofluorobenzene with eugenol in basic medium. Then 2 was reacted with 1-hexyn-3-ol using CopperII as catalyst to form 3. The structure of the compounds 2 and 3 was confirmed using NMR spectroscopy (Table 1).

Biological Evaluation

There are several reports which indicate that some fluoro-benzene derivatives exert effects on cardiovascular system [25]; however, the cellular site and molecular mechanism involved in its inotropic activity are very confusing; perhaps this phenomenon is due to differences in the chemical structure of fluoro-benzene derivatives or to the different pharmacological approaches used.

Analyzing this hypothesis, in this study the biologic activity of the compound 3 on reperfusion/injury was evaluated using an isolated heart model. The results (Figure 2) showed that the compound 3 reduced infarct size (expressed as a percentage of the area at risk) to different dose. For evaluate whether the activity exerted by compound 3 could depend on the functional groups involved in its chemical structure; therefore, in this study, also the effect of 2,4-dinitrofluorobenzene and the compound 2 on reperfusion/injury was evaluated. The results showed that only the compound 3 significantly reduce the infarction area compared with 2,4-dinitrofluorobenzene and the compound 2, these data indicate that functional groups of the compound 3 are responsible of their biological activity; these phenomenon could be the result of activation of some structure biological (p.e. ionic channels or specific receptors) involved in the endothelium of coronary artery; or possibly by the influence exerted by the compound 3 on blood pressure which results as a reduction in the infarct size, and decrease the myocardial injury after ischemia-reperfusion. Analyzing this hypothesis and other reports [35, 36] which indicate that some compounds reduce the infarct area by induce changes on perfusion pressure; in this study, the biological activity of compounds 2,4-dinitrofluorobenzene, 2 and 3 on perfusion pressure was evaluated in an isolated rat heart model. The results show that only the compound 3 significantly increases the perfusion pressure over time compared with compounds 2,4-dinitrofluorobenzene, or 2 and the control conditions. This phenomenon indicate that compound 3 exerts effects on perfusion pressure which could translated as changes in the vascular tone and coronary resistance of heart. To evaluate this hypothesis, the activity exerted by the compounds 2,4-dinitrofluorobenzene, 2 and 3 on coronary resistance was evaluated. The results indicate that only the compound 3 exert biological activity on coronary resistance in comparison with the compounds 2,4-dinitro-fluoro-benzene or 2 and the control conditions.

Analyzing these data and other reports which indicate that phenylephrine and dobutamine exert changes on perfusion pressure [37, 38]; in this study, the biological activity of phenylephrine, dobutamine and compound 3 was evaluated to analyze the possibly molecular mechanism involved in the effect exerted by the compound 3 on perfusion pressure. The results showed that compound significantly increased the perfusion pressure in comparison with phenylephrine and dobutamine; this phenomenon, suggest that molecular mechanism involved in the effect exerted on perfusion pressure was not via adrenergic.

Analyzing this data and other studies which indicate that digoxin and ouabain (Na/K-ATPase inhibitors) exert biological activity on perfusion pressure [39]; in this study, the effect exerted by compound 3 on perfusion pressure was compared with the activity of these cardiac glucosides.
The results showed that biological activity of compound 3 on perfusion pressure was similar to both digoxin and ouabain; these data indicated that compound 3 could exert their inotropic activity via Na/K-ATPase inhibition; it is noteworthy, that some reports indicate that Na,K-ATPase is a regulator of intracellular calcium in the heart [40]. Here it is important to mention that also the Ca\(^{2+}\)-ATPase can modulates cardiac contraction and this phenomenon can be inhibited by both cyclopiazonic acid and thapsigargin. Analyzing these data, in this study the biological activity of compound 3 on left ventricular pressure was evaluated in absence or presence of both cyclopiazonic acid and thapsigargin. The results indicate that inotropic activity exerted by the compound 3 was inhibited in presence of cyclopiazonic acid and thapsigargin. All these data indicate that the compound 3 exert a positive inotropic on perfusion pressure and left ventricular pressure via changes of biological activity both Na,K-ATPase and Ca+2-ATPase, this phenomenon could be similar to effect induced by the palmitoylcarnitine in an isolated dog heart model [41].

Docking Evaluation

To evaluate the possibility of compound 3 interacting with both ATPases (Na,K-ATPase and Ca\(^{2+}\)-ATPase), in this study we used a model of molecular coupling (serverdoking) [29, 30]. In the Figure 9 shown theoretical results on hydrogen-interaction between compound 3 and Na,K-ATPase (3WGU) which involves several amino acid residues such as Glu 244, Thr 371, Leu 449, Phe 475, Asn 476, Na,K-ATPase (3WGU) which involves several amino acid residues which is translated as increase in the positive inotropic activity compared with ouabain and digoxine.

5. Conclusions

The fluorobenzene-2,3-methoxy-di-phenoxy-cyclobuten-diol derivative (compound 3) is a particularly interesting drug, because its positive inotropic activity exerted on perfusion pressure involves a molecular mechanism different in comparison with other drugs; this phenomenon may constitute a novel therapy for heart failure.

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