A Rare Case of Negative Serum Calcitonin in Metastatic Medullary Thyroid Carcinoma: Diagnosis, Treatment, and Follow-Up Strategy

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Patient: Male, 39-year-old
Final Diagnosis: Medullary thyroid carcinoma
Symptoms: Painful left latero-cervical mass of 2.5 cm
Medication: —
Clinical Procedure: —
Specialty: Endocrinology and Metabolic • Surgery

Objective: Rare disease
Background: Medullary thyroid carcinoma is a rare tumor and represents less than 5% of all thyroid cancers. Tumor size and the presence of possible metastases are strictly related to serum calcitonin level, which also suggests prognosis. A serum calcitonin level above the normal range is highly suspicious for medullary cancer. This tumor has no characteristic patterns on ultrasound and fine-needle aspiration, so serum calcitonin level is the only significant element for diagnosis. The literature contains 47 cases of serum calcitonin-negative medullary thyroid cancer with heterogeneous behavior, ranging from non-aggressive to aggressive form.

Case Report: We report a rare case of a young man with negative serum calcitonin but with lateral cervical node metastases from medullary thyroid carcinoma. At the beginning, diagnosis was not achieved due to normal range of serum calcitonin and a negative thyroid ultrasonography for large nodules. Nevertheless, in another medical center he underwent a neck biopsy of the lateral cervical mass. The histologic findings showed metastases of medullary thyroid cancer. Therefore, a total thyroidectomy plus cervical lymphadenectomy was performed. During the follow-up, due to uselessfulness of serum markers, and as CEA levels were negative pre- and post-operatively, an 18F-DOPA PET scan was performed. In 3 years of follow-up, there is still no evidence of disease relapse.

Conclusions: Our case shows how a normal serum level of calcitonin cannot exclude medullary thyroid cancer. Early diagnosis is crucial because it influences appropriate surgical therapy and patient outcome. The current lack of an optimal strategy for the diagnosis and postoperative follow-up for patients with negative serum calcitonin tumor presents a diagnostic challenge for physicians.

Keywords: Calcitonin • Procalcitonin • RET Protein, Human • Thyroid Cancer, Medullary

Full-text PDF: https://www.amjcaserep.com/abstract/index/idArt/935207
Background

Medullary thyroid carcinoma (MTC) represents 3-5% of all differentiated thyroid tumors [1] and 14% of all deadly thyroid cancers [2,3]. Diagnosis is based on serum tumor markers (calcitonin, CEA, chromogranin A, Ca 19-9 and pro-calcitonin), on ultrasound imaging of the neck, and on cytology.

Regarding medullary thyroid carcinoma markers, serum calcitonin (s-CT) has the highest sensitivity (100%) [1], allowing the correct diagnosis. Other MTC markers helpful in evaluating prognoses and follow-up are carcino-embryonic antigen (CEA), chromogranin A (CgA), Ca 19-9, and pro-calcitonin (PCT) [4].

MTC has no specific patterns at ultrasonography [3,4]; many samples are indeterminate with variable appearances on fine-needle aspiration cytology [5,6]. Thus, ultrason and cytology support MTC diagnosis in patients with positive s-CT, but they are not sufficient by themselves for diagnosis. Another supportive diagnostic tool is immunohistochemical (IHC) analysis of FNA specimens, which might reduce indeterminate diagnoses, but it is not routinely available in most institutions.

s-CT level is directly proportional to tumor volume and potential metastases. Nevertheless, MTC patients with normal s-CT level, called CT-negative MTC, are diagnosed at an advanced stage [7,8]. In s-CT-negative MTC patients, ultrasonography and FNA may not help in establishing a diagnosis. The biological mechanism of negative serum is still unknown; CT-negative status may depend on the undifferentiation of the tumor cells, which is related to poorer prognosis [3]. There are many hypotheses: one considers that negative serum can be related to alteration of process of storage/secretion, while others suggest that it can be linked to protein-misfolding [5,9,10]. The literature contains 47 cases of CT-negative MTC. We report a case of a patient with an advanced MTC, s-CT-negative with RET mutation (positive in exon 14 with a heterozygous mutation c.2410 G>A, p.Val804Met). In this paper, we discuss the hypotheses of this biological presentation and the difficulties related to establishing the diagnosis of medullary thyroid carcinoma in s-CT-negative patients, which is essential to determine the best therapeutic strategy.

Case Report

A 39-year-old man with a slightly painful 2.5-cm left laterocervical mass at level III presented in April 2017, at which time the maxillofacial surgeon service report showed normal thyroid function, physiological range of antibodies against thyroid peroxidase (Ab TPO), and antibodies against thyroglobulin (Ab Tg); CT: 4 pg/L, CEA: 3.70 ng/mL (range 0-5 ng/mL). Thyroid ultrasonography demonstrated a hypoechoic nodule measuring 0.7×0.38 cm in the left middle lobe. Ultrasonography, computed tomography (CT), and MRI of the neck and fine-needle aspiration (FNA) of the laterocervical mass finding were compatible with diagnosis of schwannoma, and all findings were incompatible with a diagnosis of metastatic MTC. The patient also had bilateral congenital neurosensory deafness.

Lateral cervical mass biopsy was performed as indicated. Histologic findings showed metastases of MTC; the immunohistochemistry (IHC) showed CT+ (calcitonin +), CK7+ (cytokeratin 7+), CgA+, synaptophysin (SPH) +, CEA+, Vimentin+, thyroglobulin-, and this IHC profile was compatible with MTC metastases.

Therefore, the patient was referred to the Endocrinology Unit of the Paolo Giaccone Hospital in Palermo, where he was screened for multiple neuroendocrine malignancies using neuroendocrine marker dosage (NSE: measured by a Roche Cobas e 602 automatic electrochemical luminescence immune-analyzer and its matched reagents [Roche Diagnostics GmbH]). The NSE was slightly above the normal range, but not significantly. Plasma metanephrine and fractionated 24-h urinary metanephrines were performed to exclude pheochromocytoma. Parathyroid hormone was normal (PTH: 44.6 pg/ml). The family history was negative for adrenal and parathyroid tumors. Finally, a genetic study for RET mutation was performed and was positive for heterozygous mutation in exon 14 (c.2410 G>A, p.Val804Met), confirmed on 2 independent samples.

The patient has been referred to us at Vittorio Emanuele Hospital of Catania. Preoperative thyroid ultrasonography revealed a single hypoechoic nodule, 0.4 cm in diameter in the right lobe and a single nodule in the left lobe, measuring 0.7×0.38 cm in diameter. After collecting all data and excluding MEN syndrome (multiple endocrine neoplasia), a total thyroidectomy with bilateral central neck and lateral left compartments dissection and lymphadenectomy were recommended. On the morning of surgery, blood samples were drawn 3 times, ensuring that the patient was drug-free so as not to alter the CT serum level. The samples were carried to the laboratory within 30 min, and they were centrifuged at 3000 r/min for 10 min, then serum was separated and analyzed. For analysis of serum calcitonin, our laboratory used a Siemens IMMULITE® 2000 automatic chemiluminescence immunoassay analyzer (reference range <8 pg=L) using a standard assay kit for in-vitro diagnostic (Siemens Healthcare Diagnostics Products Limited). Carbohydrate-associated antigen 19-9 (CA19-9), and CEA were analyzed using a Siemens Centaur® XP automatic chemiluminescence immunoassay analyzer (Siemens Healthcare Diagnostics, Inc). It was also dosed Pro-CT and was in normal range.

Surgical specimen histological findings (Figures 1, 2) showed bilateral medullary thyroid cancer with an amyloid stroma; the
immunohistochemical analysis was positive for CT and CEA. Both tumors measured 3 mm and all remnants of thyroid tissue revealed C cell hyperplasia and venous vascular invasion but no extra-thyroidal extension. Two lymph nodes of the central compartment were positive for metastases, but no other metastases were found at the left lateral cervical level (only 1 was excised during the previous biopsy), so the final staging was pT1a(m) N1b, stage IVa.

A month and a half after surgery, blood tests showed Tg<1 ng/ml, CT 1.8 pg/mL, and CEA 1.14 ng/mL. Three months after surgery, blood tests showed chromogranin A 96.2 ng/ml (0-100) and NSE 9.4 mcg/L (0-18.3). The patient’s mother and son tested positive for RET mutation with a heterozygous mutation c.2410 G>A, p.Val804Met. During the follow-up, serum CT level was normal (1.7 pg/mL), as were the other markers. Ultrasound of the neck and of the liver were negative for metastases, as was the RX-chest. The patient underwent 18F-DOPA PET, as he was CT-negative, to be sure he was metastases-free. Thus, to date (February 2022) there is no evidence of disease relapse.

Discussion

The medullary thyroid tumor markers used for diagnosis and follow-up are Ct, Pro-CT, chromogranin A, CEA, and Ca19-9. Normally, a high level of serum CT is strongly suggestive for MTC. Serum concentration of CT is related to the mass of the tumor, to parafollicular C cell hyperplasia, and to presence of loco-regional cervical lymph nodes or distant-organ metastases [3]. Calcitonin is the most accurate marker for the diagnoses of MTC, but it is subjected to degradation if the test is not performed properly. Calcitonin is a 32-amino acid monomeric peptide resulting from cleavage of the post-translational processing of Pro-CT; it has a half-life of 15-40 min in physiologic condition and 3-30 h if in high concentrations [11]. The serum range reflects the pulsating secretion of the CT, which can be influenced by food intake and by temperature. At low temperature, CT decays by 23% after 12 h, by 35% after 24 h, and by 65% after 7 days, causing false-negative test results [11,12]. Pro-CT has been reported as a promising alternative marker for diagnosis and follow-up in uncertain cases of MTC without a high CT concentration, whereas chromogranin A is elevated in more than 50% of MTC cases, mainly at advanced disease stage, so it cannot be considered a useful marker for MTC diagnosis [1]. CgA is stored in neuroendocrine cells into aggregates; its serum level is directly proportional to tumor mass size and to cellular secretory activity [1]. Intracellular concentration CgA and its low serum level validate the hypothesis of impaired secretion function. CEA is not a specific marker for diagnosis of MTC, with high levels associated with other malignancies, but it can be useful in MTC patient follow-up [1]. Patients with aggressive MTC may have a disproportionately high CEA/CT ratio and a rapid CEA doubling time [13], so it might be irrelevant for the diagnosis, but it can be significant in follow-up. Carbohydrate antigen 19.9 (Ca 19.9) is a serum marker for gastrointestinal tumor, present in approximately 6% of MTC patients [5,14,15]. Further, the association between MTC and Ca 19.9 serum level has been demonstrated [4]. Based on these last observations, Ca19.9 level identifies a subgroup of patients with aggressive disease and with high mortality risk. All these tumor markers can be useful for diagnosis and follow-up of MTC, but against this background, dedifferentiation of the tumoral cells determines a reduction of CT secretion, known as CT-negative MTC. Several cases of

Figure 1. Intense immunohistochemical staining of tumor cells for calcitonin. IHC of a histologic section of Ct-negative medullary thyroid carcinoma stained with anti-calcitonin antibody.

Figure 2. Transection of MTC. Histologic section of medullary thyroid carcinoma characterized by dense bands of fibrous tissue (black arrow) and trabecular or sheetlike growth pattern (red arrow). Cells appear round, polygonal, or spindle-shaped. The cytoplasm is eosinophilic and finely granular (blue circle).
non-secretory CT MTC, also called CT-negative MTC, are reported in the literature; since normal serum tumor markers are useless, their diagnosis is given only by surgical specimen IHC analysis. CT-negative MTC is very rare, with an estimated prevalence of 0.83%, as reported by Franck-Raue et al [5]. To date, the biological mechanism of normal or reduced CT levels in MTC patients has not yet been clarified. It has been assumed that low CT serum level could be linked to impaired production of CT, as shown by CT immunostaining. This hypothesis is related to dedifferentiation of tumor cells, as tumors with only 25% cells IHC-positive for CT are related to a poor prognosis [12]. Frank-Roue et al reported that diagnoses of these patients are heterogeneous, varying from slow-progression disease to aggressive biological behavior [5]. Another hypothesis supports the hook effect: false-negative serum CT with high-level tumor marker at immunoassay analysis identified by exceeding antibody binding capacity [15]. The diagnosis in CT-negative MTC, which would allow selection of an optimal surgery strategy, is difficult to obtain. Relating to our patient, negative CT serum and imaging diagnosis of schwannoma of the lateral mass led to a wrong diagnosis. The histologic and immunohistochemical analysis findings after lateral cervical mass biopsy showed metastases by medullary thyroid carcinoma. Subsequently, at our center, we dosed all tumor markers related to MCT, and to prevent errors related to the hook effect, we performed 1:10 and 1:100 dilutions of the patient’s serum; findings were within normal ranges. In accordance with thyroid ultrasound imaging (showing a left lobe nodule measuring 0.7×0.38 cm) and immune-histological findings (mts of MTC), the patient underwent total thyroidectomy and left lateral cervical lymph nodes dissection. At histological examination, metastatic lymph nodes of about 1 cm were found in the central compartment, so it was staged as pT1a(m) N1a. However, the final TNM staging (AJCC staging criteria, 8th edition), adding the lymph node metastases of MTC biopsied at the beginning of all patient’s journey, was pT1a(m) N1b.

Table 1. Cases of CT-negative MTC reported in the literature, with their time of diagnosis and surgical specimen characteristics.

| Author                  | Case(s) | Preoperative diagnosis | Postoperative diagnosis | Central compartment MTS | Laterocervical compartment MTS |
|-------------------------|---------|------------------------|-------------------------|------------------------|--------------------------------|
| Sobol et al (1989) [30] | 1       | 0                      | 1                       | 0                      | 0                              |
| Schmid et al (1998) [31]| 4       | 0                      | 4                       | 1                      | (relapse of disease after 9 months) |
| Redding et al (2000) [32]| 1      | 0                      | 1                       | 0                      | 0                              |
| Bockhorn et al (2004) [33]| 1      | 1                      | 0                       | 0                      | 0                              |
| Sand et al (2006) [34] | 1       | 0                      | 1                       | 0                      | 1                              |
| Dora et al (2008) [35] | 1       | 0                      | 1                       | 1                      | 0                              |
| Wang et al (2008) [36] | 1       | 1                      | 0                       | 0                      | 0                              |
| Giovanella et al (2008) [37]| 1        | 1                      | 0                       | 0                      | 0                              |
| Alapat et al (2011) [38]| 1       | 1                      | 0                       | 0                      | 0                              |
| Chernyavsky et al (2011) [39]| 1        | 1                      | 0                       | 0                      | 0                              |
| Nakazawa et al (2015) [40]| 1         | 1                      | 0                       | 0                      | 0                              |
| Frank-Raue et al (2013) [5]| 9       | 7                      | 0                       | 0                      | 2                              |
| Ismi et al (2014) [41]| 1       | 0                      | 1                       | 0                      | 0                              |
| Brutsaert et al (2015) [42]| 1       | 1                      | 0                       | 0                      | 0                              |
| Kim et al (2015) [24] | 1       | 1                      | 0                       | 0                      | 0                              |
| Kasajima et al (2016) [43]| 1        | 1                      | 0                       | 0                      | 0                              |
| Parmer et al (2017) [44]| 1       | 0                      | 1                       | 0                      | 0                              |
| Zhou et al (2017) [19]| 19      | 0                      | 19                      | 3                      | 0                              |
| Licata L. et al: Metastatic medullary thyroid carcinoma serum-negative calcitonin [47]| 47 | 16 | 29 | 5 | 3 |
Table 2. CT-negative patients’ outcome in relation to preoperative and postoperative tumor marker concentrations.

| Author          | Patient | TNM       | Preoperative CT (pg/ml) | Preoperative CEA (ng/ml) | Postoperative CT (pg/ml) | Postoperative CEA (ng/ml) | Outcome                                                                 |
|-----------------|---------|-----------|-------------------------|--------------------------|--------------------------|---------------------------|--------------------------------------------------------------------------|
| Sobol et al (1989) [30] | 1       | /         | NA                      | NA                       | NA                       | NA                        | 6 months: subcutaneous masses scalp/chest wall; 11 months: regrowth scar of scalp resection; 14 months: markers (chromogranin A); 15 months: right supraclavicular lymphadenopathy. Liver and bone metastases. The patient died |
| Redding et al (2000) [32] | 1       | /         | Normal range            | ≤0.5                     | Undetectable after stimulation. Immunohistochemical stains: 75% | Undetectable after stimulation. Immunohistochemical stains: 6% | 30 months post-op TCs negative for residual or recurrent MTC |
| Sand et al (2006) [34] | 1       | pT3 L1 V1 R1 N1b | 5.3                     | Positive staining        | ≤2.47                     | /                         | Central lung emboli in the right pulmonary artery. Lung and intracerebral metastases. The patient died 6 weeks after diagnosis |
| Frank-Raue et al (2013) [5] | 7       | T1 N0 M0 | 2.9                     | 1.3                      | NA                       | NA                        | Alive, no tumor detectable                                                                 |
|                 |         |           | ≤2                      | 2.1                      |                          |                           | Alive with pulmonary metastases, reoperation because of local tumor infiltration |
|                 |         |           | ≤0.8                    | 2.8                      |                          |                           | Alive, recurrence after 10 years indicated by CEA increase; after reoperation, disease free |
|                 |         |           | 2.6                     | 3.1                      |                          |                           | Alive, reoperation because of local tumor infiltration |
|                 |         |           | NA                      | NA                       |                          |                           | Alive with lymph node metastases in the neck, bone and brain; after first operation, CT and CEA were normal despite multiple lymph node metastases; during follow-up, CT became slightly elevated |
|                 |         |           | T1 N1 M0                | 11                       | NA                       | NA                        | Dead due to disease, locally invasive tumor, and pulmonary metastases; 3 reoperations because of local tumor infiltration, during follow-up CT became slightly elevated |
|                 |         |           | T3 N1 M1                | 1.5                      | 1.7                      |                            | Dead because of pulmonary failure (metastatic MTC) |
| Brutsaert et al (2015) [42] | 1       | T2N0Mx | 2                       | 3.1                      | Positive immuno-staining | NA                        | US: no lymph nodes or nodules; TC: unremarkable; MRI: liver hemangioma |
| Zhou et al (2017) [19] | 19      | /         | NA                      | NA                       | Increase in value        | NA                        | After 6 years: 2 cases of local recurrence; 1 case of lymph nodes metastases; 1 case of lymph nodes metastasis and local recurrence |
### Table 3. Histologic characteristics and RET mutation in CT-negative and CT-positive patients.

|                | Histological features     | IHC                              |
|----------------|---------------------------|----------------------------------|
|                | Amaloid deposition        | Capsular invasion                | Lympho-vascular invasion | C cell hyperplasia | Mitosis | Necrosis | Pancytokeratin | CK7 | CK8 | CK18 | Chromogranin A | Synaptophysin |
| Sobol et al (1989) [30] | NA                        | NA                               | NA                      | NA                | NA      | NA       | NA            | NA  | NA  | NA   | NA              | +              |
| Schmid et al (1998) [31]    | –                         | +                                | –                       | NA                | NA      | NA       | NA            | +   | +   | –/+  | –                | +              |
| Redding et al (2000) [32]   | –                         | NA                               | –                       | NA                | NA      | NA       | NA            | NA  | NA  | –    | –                | +              |
| Bockhorn et al (2004) [33]  | NA                        | –                                | –                       | NA                | NA      | NA       | NA            | NA  | NA  | NA   | +                | NA             |
| Sand et al (2006) [34]       | NA                        | NA                               | NA                      | NA                | NA      | NA       | NA            | NA  | NA  | NA   | NA              | NA             |
| Dora et al (2008) [35]       | +                         | +                                | +                       | NA                | +       | NA       | NA            | NA  | NA  | NA   | NA              | +              |
| Wang et al (2008) [36]       | –                         | +                                | (vascular)              | NA                | NA      | NA       | (AE1/AE3)     | +   | NA  | NA   | –                | +              |
| GiovANELLA et al (2008) [37] | NA                        | NA                               | +                       | NA                | NA      | NA       | NA            | NA  | NA  | NA   | +                | NA             |
| Alapat et al (2011) [38]     | –                         | –                                | –                       | NA                | NA      | NA       | NA            | NA  | NA  | NA   | +                | NA             |
| Chernovskii et al (2011) [39]| –                         | –                                | –                       | –                 | –       | +        | NA            | +   | +   | +    | +                | +              |
| Nakazawa et al (2015) [40]   | –                         | –                                | +                       | –                 | <1/10 HFPs | NA         | (AE1/AE3)     | NA  | NA  | NA   | +                | +              |
| Frank-Raue et al (2013) [5]  | NA                        | 1. –                             | 2. mts and tumor infiltration | 3. recurrence     | 4. tumor infiltration | 5. lymph node mts | 6. pulmonary mts, tumor infiltration | 7. mts | NA  | NA  | NA               | NA             |
| Ismi et al (2014) [41]       | NA                        | –                                | –                       | –                 | NA      | NA       | NA            | NA  | NA  | NA   | +                | +              |
| Brutsaert et al (2015) [43]  | NA                        | +                                | –                       | NA                | +       | +        | (AE1/AE3)     | NA  | NA  | NA   | +                | NA             |
| Kim et al (2015) [24]        | NA                        | NA                               | –                       | NA                | NA      | NA       | NA            | NA  | NA  | NA   | +                | +              |
| Kasajima et al (2016) [43]   | –                         | –                                | –                       | –                 | <1/10 HFPs | –          | (AE1/AE3)     | –   | –   | +    | +                | +              |
| Parmar et al (2017) [44]      | –                         | –                                | NA                      | NA                | –       | –        | –             | NA  | NA  | NA   | +                | +              |
| Zhou et al (2017) [19]        | 11pz + 8pz –             | 8pz + 11pz –                    | Lymph node 3pz + 16pz – Vascular thrombus 1pz + 18pz –| NA  | NA  | NA      | NA            | NA  | NA  | 18pz + 1pz – | 19pz + 0pz – |
Table 3 continued. Histologic characteristics and RET mutation in CT-negative and CT-positive patients.

| IHC | ISH | Gene mutation |
|-----|-----|---------------|
| Calcitonin | CGRP | TTF-1 | TTF-2 | PAX8 | Thyroglobulin | Thyroperoxidase | CEA | KI-67 | Calcitonin | Thyroglobulin | RET gene mutation | H-RAS, K-RAS, B-RAF gene mutation |
| Sobol et al (1989) [31] | – | – | NA | NA | NA | – | NA | + | NA | NA | NA | NA | NA |
| Schmid et al (1998) [32] | – | + | NA | NA | NA | – | NA | – | NA | NA | NA | NA | NA |
| Redding et al (2000) [33] | + | NA | NA | NA | NA | – | NA | + | NA | – | – | – | NA |
| Bockhorn et al (2004) [34] | + | NA | NA | NA | NA | NA | + | NA | NA | NA | NA | NA | NA |
| Sand et al (2006) [35] | + | NA | NA | NA | NA | NA | NA | + | NA | – | NA | NA | NA |
| Dora et al (2008) [36] | + | NA | NA | NA | NA | NA | – | NA | NA | – | – | – | NA |
| Wang et al (2008) [37] | + (minimally) | NA | + | NA | NA | – | NA | + | NA | + | NA | NA | NA |
| Giovanella et al (2008) [38] | + | NA | NA | NA | NA | NA | NA | + | NA | – | NA | NA | NA |
| Alapat et al (2011) [39] | + | NA | + | NA | NA | – | NA | + | NA | NA | NA | NA | – |
| Chernyavsky et al (2011) [40] | – | NA | NA | NA | NA | + | NA | + | NA | NA | NA | – | – |
| Nakazawa et al (2015) [41] | – | + | + | – | (nuclear) | + | (cytoplasmic) | + | – | – | – | < 2% | – | – | – | NA |
| Frank-Raue et al (2013) [5] | 1. + | 2. + | 3. + | 4. ++ | 5. +++ | 6. + | 7. ++ | NA | NA | – | NA | – | NA | 1. <2 | 2. <2 | 3. <5 | 4. <10 | 5. >50 | 6. >50 | 7. >50 | NA | NA | NA | Codon 918,%: 1. 0 | 2. 0 | 3. 39 | 4. 0 | 5. 91 | 6. 60 | 7. 22 |
| Ismi et al (2014) [42] | – | NA | – | NA | NA | – | NA | – | 70% | – | NA | NA | NA |
| Brutsaert et al (2015) [43] | + | NA | + | NA | NA | NA | NA | NA | NA | – | NA | NA | NA | + | (MEN) | NA |
| Kim et al (2015) [25] | – | NA | + | NA | NA | + | NA | – | NA | NA | NA | NA | NA |
| Kasajima et al (2016) [44] | – | + | + | – | (nuclear) | + | (cytoplasmic) | + | – | – | – | 0,3% | – | – | – | – |
| Parmer et al (2017) [45] | – | NA | + | NA | NA | – | NA | + | 1% | NA | NA | NA | NA |
| Zhou et al (2017) [20] | 8 pz + 11 pz | NA | NA | NA | NA | 5 pz + 14 pz | NA | 4 pz + 15 pz | NA | NA | NA | NA | NA | – | NA | – | NA |
Conventional follow-up with neck and liver ultrasound were negative for persistence/relapse of disease; RX-chest was negative for pulmonary metastases. However, as it was a non-secretory MTC, so we focused on finding a different follow-up approach compared to the conventional one.

In the literature, the use of positron emission tomography (PET) with radiolabeled amino acid dihydroxyphenylalanine (DOPA), usually performed for gastro-pancreatic neuroendocrine tumors [17,18], has been proposed for CT-negative MTC patient follow-up [16]. 18F-DOPA PET for MTC has a sensitivity of 72% and a specificity of >90%, but limitations of this test are its high cost and its unavailability in many medical centers; consequently, it is not a regularly performed procedure [16]. In our case, 18F-DOPA PET [26,27] was performed, with a negative result for distant metastases.

Only 47 cases of CT-negative MTC are described in the literature (Table 1): 16 had a preoperative diagnosis and 29 had a postoperative incidental diagnosis; only 3 of them had lateral lymph node metastases. A retrospective analysis of 19 cases by Zhou et al [19] evaluated the immunohistochemical examination of the surgical specimens of 158 patients with a diagnosis of medullary thyroid carcinoma. Frank-Roue [5] reported 7 cases with preoperative diagnoses of MTC CT-negative, finding for the first time the prevalence of non-secretory MTC patients with sporadic diseases (prevalence of 0.83%). RET mutation analysis was not performed in all patients reported in the literature, and this is a critical point for collection and analysis of MTC CT-negative patient data.

It is also possible to classify CT-negative MTCs into single-negative, double-negative, and triple-negative. In the single-negative group, only serum CT is negative. In the double-negative group, MTC markers are negative at the time of diagnosis and in case of relapse [28]. Triple-negative patients have a typical medullary thyroid carcinoma histopathological pattern with high negative serum calcitonin, no calcitonin expression on immunohistochemistry, and absence of mRNA in situ hybridization [29]. The patient that we report in this paper belongs to the double-negative group.

As shown in Table 2, CT-negative MTC has a large spectrum of biological behavior, from indolent to life-threatening tumors [20,21]. Moreover, CT-negative and CT-positive patients show no difference in histologic characteristics and RET mutation (Table 3).

The reliability of FNA in CT-negative MTC patients is compromised by intracellular calcitonin concentration. Different MTC intracellular markers could be investigated to be helpful in FNA examination, but this can be difficult due to economic reasons and limited cellular sample availability.

Conclusions

In conclusion, we present a rare clinical case of double-negative, stage IVA MTC. CT-negative MTC is a very rare disease, showing a non-clear biological mechanism crucial for negative serum calcitonin; therefore, CT-negative MTCs, due to their biological characteristics, are usually diagnosed at an advanced stage. To date, due to the rarity of this pathology diagnosis, there are no protocols for CT-negative MTC management that can help physicians in these patients’ clinical journey. Further studies are needed to determine the optimal diagnostic, therapeutic, and follow-up pathways for these patients.

Declaration of Figures’ Authenticity

All figures submitted have been created by the authors who confirm that the images are original with no duplication and have not been previously published in whole or in part.

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