Variables affecting outcomes after allogeneic hematopoietic stem cell transplant for cerebral adrenoleukodystrophy

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Key Points

• Cerebral adrenoleukodystrophy manifests as progressive inflammatory demyelination leading to neurological function loss and early death.
• Early allo-HSCT stabilizes cerebral adrenoleukodystrophy progression; TRM remains high, even with improved regimens and supportive care.

Allogeneic hematopoietic stem cell transplantation (allo-HSCT) in early cerebral adrenoleukodystrophy can stabilize neurologic function and improve survival but has associated risks including transplant-related mortality (TRM), graft failure, and graft-versus-host disease (GVHD). An observational study of 59 patients with median age at allo-HSCT of 8 years addressed impact of donor source, donor match, conditioning regimen, and cerebral disease stage on first allo-HSCT outcomes. Efficacy analyses included 53 patients stratified by disease category: advanced disease (AD; n = 16) with Loes score >9 or neurological function score (NFS) >1 and 2 early disease (ED) cohorts (ED1 [Loes ≤4 and NFS ≤1; n = 24] and ED2 [Loes >4-9 and NFS ≤1; n = 13]). Survival free of major functional disabilities and without second allo-HSCT at 4 years was significantly higher in the ED (66%) vs AD (41%) cohort (P = .015) and comparable between ED1 and ED2 cohorts (P = .991). The stabilization of neurologic function posttransplant was greater in the ED vs AD cohort, with a median change from baseline at 24 months after allo-HSCT in NFS and Loes score, respectively, of 0 and 0.5 in ED1 (n = 13), 0.5 and 0 in ED2 (n = 6), and 2.5 and 3.0 (n = 4) in AD cohort. TRM was lower in the ED (7%) compared with the AD (22%) cohort; however, the difference was not significant (P = .094). Transplant-related safety outcomes were also affected by transplant-related characteristics: graft failure incidence was significantly higher with unrelated umbilical cord grafts vs matched related donors (P = .039), and acute GVHD and graft failure incidences varied by conditioning regimen. This study was registered at www://clinicaltrials.gov as #NCT02204904.
Adrenoleukodystrophy (ALD) is a rare X-linked metabolic disorder resulting from ABCD1 gene mutations with an estimated incidence of 1/20,000 to 1/30,000 males.\(^1,2\) Mutations lead to deficiency of the peroxisomal membrane ALD protein important in transporting very long chain fatty acids to the peroxisome for degradation, causing their accumulation in tissues and plasma.\(^3,4\) The most severe, cerebral form (CALD) develops in \textasciitilde 35\% of males \textasciitilde 10 years of age with ALD.\(^5\) If not treated in a timely manner, inflammatory cerebral demyelination in CALD leads to loss of neurological and cognitive function and death, typically in early childhood.\(^6\) Neurologic progression of CALD may manifest as major functional disabilities (MFDs), including loss of communication, movement, and mobility; blindness; tube feeding dependence; and incontinence.\(^6,7\)

Allogeneic hematopoietic stem cell transplantation (allo-HSCT) has become the standard of care with the potential to stabilize neurological function if performed early in the disease process.\(^5,7-9\) The pretransplant radiologic assessment of white matter changes quantitated by Loes score\(^10-12\) correlates with posttransplant outcomes, with improved results in patients without baseline neurologic deficits and Loes scores <10.\(^7,9\)

Transplant-related variables, including donor type, hematopoietic stem cell (HSC) source and conditioning regimen may also influence outcomes. Although allo-HSCT is ideally performed using HSCs from HLA-matched related donors (MRDs), such donors are unavailable in the majority of cases.\(^7,9\) In the absence of MRDs, grafts from unrelated sources, including adult unrelated donors, unrelated umbilical cord blood (UCB),\(^13\) or haploidentical donors\(^14\) have been successfully used for CALD. The conditioning regimen choice can influence transplant-related morbidity, and although busulfan (Bu) remains the favored myeloablative agent in most centers, more recently fludarabine (Flu) has replaced cyclophosphamide (Cy) in attempts to reduce toxicity.\(^15\) However, reduced-toxicity conditioning regimens may be associated with a higher cumulative incidence of graft failure.\(^16\) Moreover, other transplant-related complications, such as infections and acute and chronic graft-versus-host disease (GVHD) can reduce survival and affect quality of life of children during and after allo-HSCT.\(^7,9,15,17\)

To explore how CALD stage and transplant variables affect safety and efficacy outcomes of allo-HSCT in children with CALD treated with current protocols, we performed an international multicenter observational study.

### Methods

#### Study design

This was an international, prospective and retrospective data collection (January 2013-December 2019) of allo-HSCT in males with CALD (NCT02204904). Fifteen centers with extensive experience in allo-HSCT for CALD participated.

The study protocol was reviewed and approved by the relevant institutional review boards/ethics committees. All participants/caregivers provided written informed consent.
Graft failure was not deinvestigator assigned-graft rejection by 24 months posttransplant). HSCT), or secondary (based on loss of neutrophil engraftment or progression).

The incidence of graft failure/rejection (referred subsequently as graft failure) was classified as primary (based on lack of achieving neutrophil engraftment; defined as inability to achieve 3 consecutive absolute neutrophil counts ≥0.5 × 10⁹/μL by 42 days after allo-HSCT), or secondary (based on loss of neutrophil engraftment or investigator assigned-graft rejection by 24 months posttransplant). Graft failure was not defined by a prespecified threshold for the percent donor chimerism. The decision-making approach regarding second allo-HSCT varied across centers, with patient’s CALD status and overall health as important consideration factors.

The incidence and severity of acute GVHD (aGVHD) were reported according to the Acute GVHD Grading Scale (I-IV) and for chronic GVHD (cGVHD) by investigator assessment of limited and extensive cGVHD.

Platelet engraftment was defined as achieving 3 consecutive platelet counts ≥20 × 10⁹ cells/L without platelet transfusions in the preceding 7 days.

Posttransplant serious adverse events were collected throughout the study after first allo-HSCT. Safety assessments included infection frequency and severity, intensive care unit stays and duration, and inpatient hospitalizations and duration.

### Table 1. Patient baseline disease and transplant characteristics

|                     | ED1 n = 26 | ED2 n = 13 | ED1 + ED2 n = 39 | AD n = 16 | All patients n = 59 |
|---------------------|------------|------------|------------------|-----------|--------------------|
| **Median (min-max)**|            |            |                  |           |                    |
| Age at CALD dx, y   | 7 (0-14)   | 7 (5-11)   | 7 (0-14)         | 8 (6-12)  | 7 (0-14)           |
| Age at first allo-HSCT, y | 7 (2-14) | 8 (5-11)   | 8 (2-14)         | 9 (6-10)  | 8 (2-14)           |
| Time from ALD dx to CALD dx, mo | 30.8 (0-125) | 0.2 (0-111.2) | 9.8 (0-125) | 0.03 (0-48.2) | 0.8 (0-125) |
| Time from ALD dx to transplant, mo | 45.0 (2.8-152.2) | 4.4 (0.8-113.2) | 25.3 (0.8-152.2) | 3.5 (1.9-99.5) | 9.4 (0.8-152.2) |
| Time from CALD dx to transplant, mo | 3.6 (1.3-76.3) | 3.0 (0.6-78) | 3.6 (0.6-78) | 3.4 (1.1-51.4) | 3.5 (0.6-78) |
| Length of follow-up, mo | 25.3 (0.9-49.2) | 31.2 (0.9-48.1) | 25.8 (0.9-48.1) | 8.9 (2.1-48.6) | 23.0 (0.9-49.5) |
| **n (%)**           |            |            |                  |           |                    |
| Baseline NFS        |            |            |                  |           |                    |
| 0                   | 26 (100)   | 12 (92.3)  | 38 (97.4)        | 5 (31.3)  | 43 (72.9)          |
| 1                   | 0          | 1 (7.7)    | 1 (2.6)          | 5 (31.3)  | 7 (11.8)           |
| >1-<4               | 0          | 0          | 0                | 1 (6.3)   | 1 (1.7)            |
| >4                  | 0          | 0          | 0                | 1 (6.3)   | 4 (6.8)            |
| Baseline Loes score |            |            |                  |           |                    |
| 0                   | 4 (15.4)   | 0          | 4 (10.3)         | 0         | 6 (10.2)           |
| 0.5-4               | 22 (84.6)  | 0          | 22 (56.4)        | 0         | 22 (37.3)          |
| >4-9                | 0          | 13 (100)   | 13 (33.3)        | 1 (6.3)   | 15 (25.4)          |
| >9                  | 0          | 0          | 0                | 13 (81.3) | 13 (22.0)          |
| Donor source        |            |            |                  |           |                    |
| MRD (BM + PB + UCB) | 7 (26.9)   | 4 (30.8)   | 11 (28.2)        | 1 (6.3)   | 12 (20.3)          |
| UR (BM + PB)        | 9 (46.2)   | 5 (38.5)   | 14 (35.9)        | 4 (25.0)  | 19 (32.2)          |
| UR (UCB)            | 10 (38.5)  | 4 (30.8)   | 14 (35.9)        | 6 (37.5)  | 23 (39.0)          |
| Haploidentical      | 1 (3.8)    | 0          | 1 (2.6)          | 1 (6.3)   | 2 (3.4)            |
| Donor match         |            |            |                  |           |                    |
| Matched             | 20 (76.9)  | 9 (69.2)   | 29 (74.4)        | 6 (37.5)  | 36 (61.0)          |
| Haplo + mismatched  | 6 (23.1)   | 4 (30.8)   | 10 (25.6)        | 10 (62.5) | 23 (39.0)          |
| **HLA match type**  |            |            |                  |           |                    |
| Matched             |            |            |                  |           |                    |
| 10/10               | 8 (30.8)   | 4 (30.8)   | 12 (30.8)        | 3 (18.8)  | 16 (27.1)          |
| 8/8                 | 6 (23.1)   | 4 (30.8)   | 10 (25.6)        | 1 (6.3)   | 11 (18.6)          |
| 6/6                 | 6 (23.1)   | 1 (7.7)    | 7 (17.9)         | 2 (12.5)  | 9 (15.3)           |
| Mismatched          |            |            |                  |           |                    |
| 9/10                | 1 (3.8)    | 1 (7.7)    | 2 (5.1)          | 1 (6.3)   | 3 (5.1)            |
| 5/6                 | 3 (11.5)   | 2 (15.4)   | 5 (12.8)         | 1 (6.3)   | 7 (11.9)           |
| 4/6                 | 2 (7.7)    | 1 (7.7)    | 3 (7.7)          | 3 (18.8)  | 8 (13.6)           |
| Haploidentical      | 0          | 0          | 0                | 5 (31.3)  | 5 (8.5)            |

dx, diagnosis; UR, unrelated donor.
Safety assessments were stratified by donor cell source, type and match, and conditioning regimen (Bu/Cy and Bu/Flu).

Analyses

Patient and transplant characteristics were expressed as number and percentage for categorical variables and median with ranges for continuous variables.

Time of origin in time-to-event analyses was the first allo-HSCT and patients alive without an event were censored at the last follow-up or at the time of study termination. The Kaplan-Meier method was used to estimate survival rates and associated 2-sided 95% confidence intervals (CIs).

Cumulative incidence function (CIF) estimates were calculated from the log-rank test. The subdistribution hazard ratios with 95% CIs were provided from the univariate Cox model and P values comparing subgroups were calculated from the log-rank test.

Figures A and B show Kaplan-Meier analyses of overall survival (A) and MFD-free survival (B) by disease severity cohorts. Time of origin was the first allo-HSCT and patients alive without an event were censored at the last follow-up or at the time of study termination. MFD-free survival included survival without second allo-HSCT or major functional disabilities (MFDs).

Results

Characteristics of patients and transplants

Fifty-nine patients were enrolled at 15 centers, including 5 centers in the United States (n = 36); 2 centers each in the United Kingdom (n = 5), Germany (n = 7), and the Netherlands (n = 5); and 1 center each in Argentina (n = 2), Italy (n = 2), France (n = 1), and Spain (n = 1). Patient and transplant characteristics are shown in Table 1. Twenty-six patients were enrolled before allo-HSCT (prospective), 26 were enrolled posttransplant but before the 24-month visit (retrospective or partially retrospective), and 7 patients that died posttransplant before the study commenced were enrolled retrospectively. Study populations are summarized in supplemental Figure A. The efficacy population included 55 patients (4 were excluded because baseline Loes scores were 0 and either had no follow-up, or all follow-up Loes scores were 0) and efficacy analyses stratified by disease state included 53 patients (2 additional patients were excluded, given missing baseline NFS or central readings of baseline Loes score). Individual patient disease and transplant characteristics are listed in supplemental Table A.

The median (range) duration of follow-up after first allo-HSCT was 23.0 months (0.9-49.5). The median (range) age at CALD diagnosis was 7 years (0-14). The median ALD-to-CALD diagnosis time was <1 month in the ED2 (0-11.2) and AD cohorts (0-48), and 30 months (0-125) in the ED1 cohort. The time from CALD diagnosis to allo-HSCT was a median 3.5 months (range, 0.6-78) and did not differ across cohorts.

Baseline NFS was 0 or 1 in 50/59 patients (85%). Fifty-two patients (88%) had available baseline GdE status and 39/52 (75%) were GdE+. The distribution of centrally read baseline Loes scores is shown in Table 1.

The most common conditioning regimen was Bu/Flu (54%), followed by Bu/Cy (42%), without significant differences between ED and AD cohorts. Two patients had missing regimen information. No systemic exposure data for Bu were available, but Bu was administered with myeloablative intent. Granulocyte-colony stimulating factor was administered per institutional guidelines, and was received during the first allo-HSCT period by 25/59 patients (42%), including 18 who had UCB donors.

In the ED cohort, MRD grafts were used in 11/39 cases (28%) and unrelated donors in 28/39 cases (72%); 14 from BM+PB and 14
Table 2. Effects of disease stage, donor source, donor match, and conditioning regimen on survival and transplant-related outcomes

| Disease stage | OS | MFD-free |
|---------------|----|----------|
| All           | 20.5 | 13.7 |
|               | (11.2-31.7) | (6.3-23.9) |
| Disease stage | Survival analyses from log-rank test; underlining and italics denote significant differences. |

| Disease stage | OS | MFD-free |
|---------------|----|----------|
| ED1           | 86.6 | 67.8 |
|               | (53.9-96.7) | (43.0-83.6) |
| ED2           | 72.9 | 61.7 |
|               | (27.6-92.5) | (25.6-84.3) |
| ED            | 81.9 | 66.1 |
|               | (57.8-93.0) | (48.3-80.0) |
| AD            | 53.0 | 41.3 |
|               | (23.3-75.9) | (17.3-63.9) |
| ED vs AD      | **0.008** | **0.015** |
| P value       | 0.520 | 0.348 |
| HR (95% CI)   | (0.580-0.743) | (0.149-0.847) |
|               | (0.190-2.388) | (0.171-5.113) |
|               | (0.200-1.491) | (0.091-0.734) |
|               | (0.329-2.150) | (0.035-1.317) |
|               | (0.487-3.072) | |

| Donor source | MRD | UR BM + PB | UR UCB | UR BM + PB + UCB vs MRD |
|--------------|-----|------------|---------|-------------------------|
| P value      | **0.060** | **0.015** | **0.068** |
| HR (95% CI)  | 0.733 | 1.796 | 1.434 |
|               | 0.310-1.642 | (0.017-5.113) |
|               | 0.524-1.780 | (0.014-1.000) |
|               | 0.524-1.780 | (0.014-1.000) |

| Donor source | MRD | UR BM + PB vs MRD |
|--------------|-----|--------------------|
| P value      | **0.060** |
| HR (95% CI)  | 0.733 | 1.796 |
|               | 0.310-1.642 |

| Donor source | MRD | UR UCB vs MRD |
|--------------|-----|----------------|
| P value      | **0.060** |
| HR (95% CI)  | 0.733 | 1.796 |
|               | 0.310-1.642 |

| Donor source | MRD | UR UCB vs UR BM + PB |
|--------------|-----|-----------------------|
| P value      | **0.060** |
| HR (95% CI)  | 0.733 | 1.796 |
|               | 0.310-1.642 |

Deaths, MFDs, and second allo-HSC infusions are considered events for MFD-free survival outcome. MFDs included loss of communication, cortical blindness, tube feeding dependence, total incontinence, wheelchair dependence, and complete loss of voluntary movement. Patients who did not experience any event are censored at the time of the last MFD assessment when they are MFD-free. Survival rates estimated using Kaplan-Meier analysis. HR (95% CI) for survival rates calculated by Cox proportional hazards model. P values for survival analyses from log-rank test; underlining and italics denote P values < 0.05 and 0.05-0.1, respectively. Bold numbers indicate a significant difference. HR (95% CI) for cumulative incidence functions (CIF) estimates from a cause-specific hazard model. P values are calculated using competing risks analysis calculated by Gray’s test. NE, not estimable; UR, unrelated.

*Same as at month 48.
1Haploidentical category (donor source) = 5 patients had allo-HSCT using haploidentical donors, but all had <24 months of follow-up after allo-HSCT; therefore, data as of month 24 or month 48 were not available.
from UCB). Conversely, only 1 patient with AD (1/16, 6%) received an MRD graft. Five patients with AD (5/16, 31%) had transplants from haploidentical donors. Matched donors were used for 74% of patients with AD and 38% of patients with ED and AD, respectively.

**OS and TRM and toxicity**

Forty-four of 55 patients (80%) included in OS analyses were alive at last follow-up; 4-year OS (95% CI) Kaplan-Meier estimate was 72.1% (54.4-83.8) (Figure 1; Table 2). OS was significantly higher in the ED (81.9%, 57.8-93.0) vs AD (53%, 23.3-75.9) cohort (P = .008).

In the gene therapy-matched cohort (n = 27; patient and treatment characteristics described in supplemental Table B), the estimated 4-year OS was 77.8% (45.5-92.3) (supplemental Figure B; supplemental Table C) and was similar in those matched with sibling donors (n = 10, 74.1% [28.9-93.0]) or other donors (n = 17, 83.3% [27.3-97.5]), respectively (hazard ratio 1.65 [0.15-18.35]).

There was no difference in OS in the ED or AD cohorts stratified by NAC use. However, the small numbers of patients in AD (12 with NAC and 4 without) limited the assessment.

Eleven patients in the efficacy population (n = 55) died during the first allo-HSCT period because of disease progression (n = 2), cardiac arrest (n = 1), and unknown causes (n = 2). The remaining deaths were characterized as TRM and attributed to aGVHD (n = 3), cGVHD (n = 1), conditioning toxicity (n = 1), and immunosuppression secondary to drugs for posttransplant management (n = 1); cause of death listed as fatal respiratory failure caused by cytomegalovirus viremia and Epstein-Barr viremia. Two TRM-attributed deaths occurred in patients with MRD transplants (from conditioning toxicity [n = 1] and cGVHD [n = 1]). One additional patient from the overall treated population (N = 59) died (TRM from aGVHD).

The CIF (95% CI) of TRM at 24 months was 14.8% (6.3-26.6), without significant differences across transplant characteristics (Table 2). There was a trend for increased TRM in the AD (22%) vs ED (7%) cohort (P = .08). TRM and other transplant-related outcomes in the gene therapy-matched cohort were comparable to the ED cohort (supplemental Table C).

The majority of patients had at least 1 serious adverse event (43/59, 73%); those occurring in ≥2 patients are described in Table 3. One patient experienced veno-occlusive disease attributed to the Bu/Cy conditioning regimen. Data for hospitalizations and intensive care unit stays are shown in Table 4. The initial hospitalization data for allo-HSCT were available for 59 patients, with a median (min-max) duration of 51 (25-240) days.

**Donor engraftment**

Neutrophil engraftment was achieved in 53/59 (89.8%) patients at a median of 17 (12-36) days. Ten patients had either primary (6/59, 10.2%) or secondary 4/53 (7.5%) graft failure, and all had received...
unrelated grafts. All evaluable patients (n = 47) achieved platelet engraftment at a median of 26 (13-67) days. Nine patients with primary or secondary graft failure underwent a second allo-HSCT and 1 patient had a third allo-HSCT. At last follow-up, 6/9 patients were engrafted and alive; among these, 5 were MFD-free.

Among 6 patients with primary graft failure (absolute neutrophil count-based), donor sources were mismatched UCB in 4 (67%) and matched BM in 2 (33%). Four patients, who received grafts from mismatched UCB (n = 1), mismatched UCB (n = 2), or mismatched PB (n = 1), had secondary graft failure. Although donor chimerism threshold was not included in the definition of graft failure, it could have been considered by investigators when assessing secondary graft failure. Donor chimerism (with cell type used for donor chimerism analysis shown in parentheses) for 4 patients with secondary graft failure at time of second allo-HSCT was 0% (BM), 24% (peripheral blood mononuclear cells), 54% (BM), and 61% (BM).

The CIF estimate (95% CI) of graft failure for all patients at 24 months was 17.0% (8.7-27.7) and was significantly lower in patients that received MRD (0/12) vs unrelated UCB transplants (7/23) (CIF: not estimable vs 30.4%, respectively; *P* = .039) (Table 2). Although not significant, a trend (*P* = .09) was noted related to the conditioning regimen, where fewer patients receiving Bu/Cy (2/25; 8%) had graft failure vs those receiving Bu/Flu (8/32; 25%).

**Table 3. Incidence and timing of serious adverse events occurring in 2 or more patients**

| Event Category | D1 to M24 (N = 59) | D1 to M24 (N = 53) | D1 to M24 (N = 29) | D1 to M48 (N = 59) | D1 to M48 (N = 59) |
|---------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| Nervous system disorders | | | | | |
| Neurological decompensation | 1 (1.7) | 1 (1.9) | 0 | 1 (3.4) | 1 (3.4) |
| Aphasia | 0 | 1 (1.9) | 0 | 1 (3.4) | 1 (3.4) |
| Seizure | 0 | 1 (1.9) | 0 | 1 (3.4) | 1 (3.4) |
| Infections and infestations | | | | | |
| Device-related infection | 0 | 4 (7.5) | 0 | 0 | 4 (6.8) |
| BK virus infection | 0 | 3 (5.7) | 0 | 0 | 3 (5.1) |
| Bacteremia | 1 (1.7) | 2 (3.8) | 0 | 0 | 3 (5.1) |
| Staphylococcal infection | 0 | 3 (5.7) | 0 | 0 | 3 (5.1) |
| Clostridium difficile infection | 0 | 2 (3.8) | 0 | 0 | 2 (3.4) |
| Epstein-Barr viremia | 0 | 2 (3.8) | 0 | 0 | 2 (3.4) |
| Human herpesvirus 6 infection | 1 (1.7) | 1 (1.9) | 0 | 0 | 2 (3.4) |
| Lung infection | 1 (1.7) | 1 (1.9) | 0 | 0 | 2 (3.4) |
| Sepsis | 0 | 2 (3.8) | 0 | 0 | 2 (3.4) |
| Septic shock | 0 | 1 (1.9) | 0 | 1 (3.4) | 1 (1.7) |
| General disorders and administration site conditions | | | | | |
| Pyrexia | 0 | 3 (5.7) | 0 | 0 | 3 (5.1) |
| Disease progression | 0 | 2 (3.8) | 0 | 0 | 2 (3.4) |
| Ear and labyrinth disorders | | | | | |
| Hypoacusis | 0 | 2 (3.8) | 0 | 0 | 2 (3.4) |
| Gastrointestinal disorders | | | | | |
| Diarrhea | 0 | 2 (3.8) | 0 | 0 | 2 (3.4) |
| Immune system disorders | | | | | |
| Anaphylactic reaction | 1 (1.7) | 0 | 0 | 1 (3.4) | 1 (1.7) |
| Metabolism and nutrition disorders | | | | | |
| Feeding intolerance | 0 | 0 | 2 (5.9) | 0 | 2 (3.4) |
| Renal and urinary disorders | | | | | |
| Acute kidney injury | 0 | 1 (1.9) | 0 | 1 (2.9) | 0 | 2 (3.4) |
| Respiratory, thoracic, and mediastinal disorders | | | | | |
| Hemorrhage | 0 | 1 (1.9) | 1 (2.9) | 0 | 2 (3.4) |
| Respiratory failure | 0 | 2 (3.8) | 0 | 0 | 2 (3.4) |
| Vascular disorders | | | | | |
| Hypertension | 1 (1.7) | 1 (1.9) | 0 | 0 | 2 (3.4) |

| D | day 1; M, month; NE, neutrophil engraftment. *
|Does not include severe adverse events of hematologic events, death, or engraftment failure.
Among these 7 patients with available follow-up Loes scores and NFS, the NFS change from baseline to the last visit ranged from −2 to 5 (n = 6; 5 with ≤1), whereas the change in Loes score ranged from −1 to 4 (n = 6; 5 with ≤3). Narratives of the NFS components over time for the 5 patients with AD with NFS at last visit ≤2 are provided in Appendix 2.

In the gene therapy-matched cohort (n = 27), the estimated 4-year MFD-free survival was 63.2% (38.2-80.4) because of second transplant and death (none of these patients developed MFDs) (supplemental Figure B; supplemental Tables A and C). There was no significant difference between the recipients of matched sibling grafts (74.1% [28.9-93.0]; n = 10) vs other donor types (58.8% [27.5-80.4]; n = 17).

Neurological outcome

The median change from baseline in Loes score and NFS over time are shown in Figure 2A-B stratified by disease stage and in supplemental Figure C for the gene therapy-matched cohort. For both parameters, there were limited changes from baseline in the ED1 cohort and greater variability in the ED2 and AD cohorts. Individual patient data over time are shown in Figure 2C-D with outcomes of MFD and death indicated.

Table 4. Hospitalizations and ICU stays

| Number of inpatient hospitalizations, n (%) | First hospitalization (up to D/C)† | Post-D/C to M48 (last contact) |
|-------------------------------------------|---------------------------------|--------------------------------|
| N = 59                                    | N = 58                          |
| 0                                         | 28 (48.3)                       | 11 (19.0)                      |
| 1                                         | 57 (96.6)                       | 6 (10.3)                       |
| 2                                         | 2 (3.4)                         | 12 (20.7)                      |
| 3                                         | 0                               | 5 (8.6)                        |
| 4                                         | 0                               | 1 (1.7)                        |
| 5                                         | 0                               | 1 (1.7)                        |

Duration of inpatient hospitalizations (days)

| Number of ICU stays, n (%) | First hospitalization (up to D/C)† | Post-D/C to M48 (last contact) |
|---------------------------|---------------------------------|--------------------------------|
| N = 59                    | N = 58                          |
| 0                         | 58 (98.3)                       | 53 (91.4)                      |
| 1                         | 0                               | 4 (6.9)                        |
| 2                         | 1                               | 1 (1.7)                        |
| 3                         | 1 (1.7)                         | 0                              |

Duration of ICU stays (days)

| Number of ICU stays, n (%) | First hospitalization (up to D/C)† | Post-D/C to M48 (last contact) |
|----------------------------|---------------------------------|--------------------------------|
| N = 59                     | N = 58                          |
| 0                          | 5                               | 17 (30)                        |
| 1                          | 1                               | 5                              |

D/C, discharge after initial hospitalization; ICU, intensive care unit.
†Starts with admission for conditioning and donor cell infusion for first transplant and ends with the first discharge after neutrophil engraftment.
*Analysis includes hospitalization times for patients who did not have neutrophil engraftment after first allo-HSCT and remained in the hospital until neutrophil engraftment was obtained after second or third allo-HSCT procedures.

Survival free of graft failure or MFDs (MFD-free survival)

The 4-year MFD-free survival for 55 patients was 58.1% (95% CI, 42.6-70.8) (Figure 1B; Table 2) and was significantly higher in the ED (66.1%; 95% CI, 46.3-80.0) vs AD group (41.3%; 95% CI 17.3-63.9) (P = .015). Primary events were death and graft failure for the ED group, and death and MFDs for the AD group. The baseline NFS for patients developing MFDs was 0 to 5 (2 with 0, 1 with 1, and 3 with ≥2) and baseline Loes 6 to 18.5 (1 with 6, 4 with ≥10, and 1 missing). (supplemental Table A). All 5 with available GdE data at baseline were GdE−. One of the 6 with MFDs had primary graft failure (as determined by treating physician; also noted to have 0% donor chimerism in blood). The remaining 4 patients had >90% donor chimerism (2 in myeloid cells, 1 in blood, and 1 in peripheral blood mononuclear cells), whereas 1 patient had blood donor chimerism of 97% at 12 months after treatment and 0% shortly thereafter. This patient was alive at last follow-up (48 months posttreatment). Two additional patients with MFDs were alive at study termination. All MFDs occurred between days 12 and 111 and are described in detail in Appendix 1.

Among 7 patients with AD who did not die or develop MFDs, the baseline NFS ranged from 0 to 3 (2 with 0, 3 with 1, and 2 with ≥2), Loes from 11 to 16 (4 with 11, 2 ≥13, and 1 missing) and 4/6 with GdE status at baseline were GdE− (supplemental Table A).

Neurological outcome

The median change from baseline to last visit was observed in 29/30 (97%) patients (Figure 3). Overall, there was no correlation between GdE status or GdE reemergence and clinical status or cerebral progression posttransplant. One patient in the ED1 group who was GdE− at the 1-month assessment had a second allo-HSCT from graft failure. Among 33 patients with at least 6 months of follow-up, GdE− transiently reemerged in 1 patient GdE− at baseline and 3 GdE− at baseline (1 patient each in ED1 and ED2 and 2 in AD). None of these patients had primary or secondary engraftment failure. One patient in the AD cohort with transient reemergence at 12 months and resolution at 24 months had baseline Loes pattern that was described as frontal (Loes score = 6 and NFS = 4) and developed 4 MFDs posttreatment (supplemental Material; patient 23). Both the NFS and Loes score increased to 18 at 12 months. For the remaining 3 patients with transient GdE− reemergence, changes from baseline to final readings for Loes score ranged from −1 to 4.5, and NFS ≤1 for ED1 and ED2 patients and 4 for the patient with AD.

GVHD

The overall CIF estimate (95% CI) of grade II-IV and grade III-IV aGVHD at 24 months was 20.5% (11.2-31.7) and 13.7% (6.3-23.9), respectively. The CIF estimates of limited and extensive cGVHD at 24 months was 13.3% (5.7-24.2) and 7.6% (2.4-17.0), respectively, and cumulative incidence of overall cGVHD was 16.7% (8.1-27.9). Differences in cumulative incidence across donor type and match were not statistically significant (Table 2). For grade II-IV and III-IV, the aGVHD cumulative incidence was significantly higher in patients who received Bu/Cy compared with BuFlu conditioning regimens (P = .006 and 0.02, respectively) (Table 2). There were no apparent differences in donor graft source that might confound this finding.
Among patients receiving Bu/Flu or Bu/Cy conditioning regimens, serotherapy use was similar (25/32 [78%] and 17/25 [68%], respectively). Grade II-IV aGVHD occurred in 13/59 (22%) patients, 8 of whom were among 25 patients who received granulocyte-colony stimulating factor.

**Discussion**

This large, international, primarily prospective study assessed outcomes after the first allo-HSCT in patients with CALD stratified by disease stage and transplant-related variables. The results provide an important supplement to other published reports on the safety and outcomes profile of allo-HSCT treatment of CALD.\(^7,8,13,17,23\)

These data will also provide an important context for the clinical studies of LVV-based HSC gene therapy for CALD, for which interim phase 2/3 study data have been reported\(^6\) and a completed phase 2/3 study manuscript is in preparation.

The study confirms that outcomes following allo-HSCT are dependent on disease stage based on baseline neuroimaging and neurological function. Baseline Loes scores <9 (with parietal-occipital demyelination pattern in most of the patients) have been shown to be associated with better posttransplant survival,\(^17\) although Loes scores ≥4.5 may be associated with worse neuropsychological outcomes after allo-HSCT compared with those with Loes scores <4.5.\(^24\) Patients in the 2 ED cohorts in our study compared with the AD cohort experienced improved OS and MFD-free survival as well as a greater stabilization of neurologic function based on NFS.

By demonstrating that early intervention in CALD leads to more favorable disease-related outcomes, this study emphasizes the importance of newborn screening for ALD, which will prove critically important in establishing an early diagnosis and providing an opportunity for consistent monitoring to identify patients developing cerebral disease.\(^25,26\) Although inclusion of ALD into newborn screening panels has been gaining momentum in the United States in recent years, at this time this life-saving diagnostic method has been

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**Figure 2.** Loes score and Neurological Function Score (NFS) over time. Median change from baseline by disease stage is shown for Loes score (A) and NFS (B). Number of evaluable patients at each time point are shown below the x-axis (data availability for each patient at each visit was influenced by a number of factors, including the survival status, retransplantation status, length of follow-up, whether the visit took place, and whether the assessment was performed). (C-D) Individual patient Loes scores and NFS by disease stage.
adopted by fewer than one-half of the US states and no European countries. Additionally, while increasing efforts to support the earlier diagnosis of ALD, there should also be focus on standardizing approaches for scoring demyelinating lesions to ensure that appropriate patients are treated with allo-HSCT. Among 7 deaths from TRM after first transplant (N = 59), 2 occurred with MRD transplants (n = 12). In our study, aGVHD was the leading cause of TRM (4 cases). Previous work suggested that severe aGVHD is associated with rapid clinical deterioration in patients with CALD. The risk of development of grade II-IV or grade III-IV aGVHD in our study was significantly greater (three- to eightfold higher) with a Bu/Cy vs Bu/Flu conditioning regimen. Conversely, a single-center retrospective study of allo-HSCT in 99 pediatric patients with inherited metabolic disorders, including 43 with CALD, demonstrated that Bu/Cy and Bu/Flu were associated with a similar incidence of grade III-IV aGVHD (9% and 6%, respectively). In the same study, Bu/Flu was used in most transplants with unrelated umbilical cord grafts and led to a higher rate of graft failure compared with Bu/Cy (29% vs 14%, P = .08), which is similar to our study where we noted a trend (~threefold difference in risk) for increased graft failure in patients receiving Bu/Flu vs Bu/Cy. Based on other studies, Bu/Cy is considered more myeloablative, and more toxic, than Bu/Flu. A limitation of our study was the lack of data regarding therapeutic Bu dose monitoring, although the doses administered were assumed to be myeloablative. Because targeting a myeloablative Bu area under the curve is critical to achieve full donor engraftment, it is possible that Bu underexposure might have contributed to these results. We also observed increased graft failure risk with the use of unrelated umbilical cords compared with the use of a graft from an MRD.

MFD-free survival was significantly higher for matched transplants vs haploidentical/mismatched transplants. Patients receiving haploidentical/mismatched transplants were more likely to undergo a second allo-HSCT or to develop MFDs. Overall, haploidentical/mismatched transplants were used in the majority of patients with AD (83%), possibly because of the time required to find matched donors.

Allo-HSCT is offered to patients with AD at limited number of centers. In the absence of treatment, progression in patients with AD is typically rapid and devastating, with onset of multiple MFDs, and death or vegetative state within 2 to 4 years of onset of symptoms. None of the 16 patients with AD treated in our study had MFDs at baseline. For 6 patients with AD who developed MFDs, they occurred <4 months posttreatment. Our study shows that neurologic disease stabilization can occur in a significant proportion of patients with AD: 7/16 (43%) patients were alive and without MFDs at last follow-up, and of these 7, 5 patients had an NFD ≤1 at last assessment. These data suggest that a subset of patients with CALD and advanced disease may benefit from allo-HSCT if a suitable donor is available. In the future, additional treatment options should be explored for these patients. Nevertheless, identifying which patients with AD will develop significant progression and/or achieve an MFD is still quite difficult to ascertain.

Although this study provides important data regarding factors affecting allo-HSCT safety and outcomes, there are several limitations to...
Some analyses were limited by small sample sizes and limited follow-up in some cases, preventing comparisons for several factors. Trends were noted, but additional differences in cell source and/or level of donor matching between cohorts may contribute to observed effects. The study was limited by its small sample size, precluding multivariate analysis. Important endpoints, especially for the patients with ED, are neuropsychological outcomes. However, sufficient information was not available to include these comprehensive assessments.

In summary, patients with early cerebral disease benefit the most from allo-HSCT based on the presented data. Stabilization of neurological disease and improved overall and MFD-free survival are observed. Early disease stages may also be associated with lower TRM. Newborn screening and MRI monitoring of those at risk will provide a crucial opportunity to identify early cerebral disease and intervene expeditiously. Nevertheless, significant risks continue to be observed with allo-HSCT for CALD, which are influenced by donor source, conditioning regimen, and disease status.

**Data sharing**

All authors had access to primary clinical trial data that were analyzed by bluebird bio. Appropriately deidentified patient-level datasets and supporting documents may be shared following attainment of applicable marketing approvals and consistent with criteria established by bluebird bio and/or industry best practices to maintain the privacy of study participants. Requests will be evaluated on a case-by-case basis. For more information, please contact datasharing@bluebirdbio.com.

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Authorship

Contribution: R.C., J.-J.B., A.T., E.M., A.C.D., and P.J.O. contributed to the conception, design, and planning of the analysis; D.J.L. interpreted magnetic resonance imaging data; A.T. contributed to data analysis; and all authors contributed to data acquisition and to critically reviewing or revising the manuscript.

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References

1. Bezman L, Moser AB, Raymond GV, et al. Adrenoleukodystrophy: incidence, new mutation rate, and results of extended family screening. Ann Neurol. 2001;49(4):512-517.
2. Wiesinger C, Eichler FS, Berger J. The genetic landscape of X-linked adrenoleukodystrophy: inheritance, mutations, modifier genes, and diagnosis. Appl Clin Genet. 2015;8:109-121.
3. Moser HW, Mahmood A, Raymond GV. X-linked adrenoleukodystrophy. Nat Clin Pract Neurol. 2007;3(3):140-151.
4. Raymond GV, Moser AB, Fatemi A. X-linked adrenoleukodystrophy. In: Adam MP, Ardinger HH, Pagon RA, et al, eds. GeneReviews®. Seattle, WA: University of Washington, Seattle; 1999:1-17.
5. Mahmood A, Raymond GV, Dubey P, Peters C, Moser HW. Survival analysis of hematopoietic cell transplantation for childhood cerebral X-linked adrenoleukodystrophy: a comparison study. Lancel Neurol. 2007;6(8):687-692.
6. Eichler F, Duncan C, Musolino PL, et al. Hematopoietic stem-cell gene therapy for cerebral adrenoleukodystrophy. N Engl J Med. 2017;377(17):1630-1638.
7. Raymond GV, Aubourg P, Paker A, et al. Survival and functional outcomes in boys with cerebral adrenoleukodystrophy with and without hematopoietic stem cell transplantation. Biol Blood Marrow Transplant. 2019;25(3):538-548.
8. Aubourg P, Blanche S, Jambaqué I, et al. Reversal of early neurologic and neuroradiologic manifestations of X-linked adrenoleukodystrophy by bone marrow transplantation. N Engl J Med. 1990;322(26):1860-1866.
9. Miller WP, Rothman SM, Nascene D, et al. Outcomes after allogeneic hematopoietic cell transplantation for childhood cerebral adrenoleukodystrophy: the largest single-institution cohort report. Blood. 2011;118(7):1971-1978.
10. Loes DJ, Hite S, Moser H, et al. Adrenoleukodystrophy: a scoring method for brain MR observations. AJNR Am J Neuroradiol. 1994;15(9):1761-1766.
11. Loes DJ, Fatemi A, Melhem ER, et al. Analysis of MRI patterns aids prediction of progression in X-linked adrenoleukodystrophy. Neurology. 2003;61(3):369-374.
12. Moser HW, Loes DJ, Melhem ER, et al. X-Linked adrenoleukodystrophy: overview and prognosis as a function of age and brain magnetic resonance imaging abnormality. A study involving 372 patients. Neuropediatrics. 2000;31(5):227-239.
13. Beam D, Poe MD, Provenzale JM, et al. Outcomes of unrelated umbilical cord blood transplantation for X-linked adrenoleukodystrophy. Biol Blood Marrow Transplant. 2007;13(6):665-674.

14. Fernandes JF, Bonfim C, Kerbauy FR, et al. Haploidentical bone marrow transplantation with post transplant cyclophosphamide for patients with X-linked adrenoleukodystrophy: a suitable choice in an urgent situation. Bone Marrow Transplant. 2018;53(4):392-399.

15. Bartelink IH, van Reij EM, Gerhardt CE, et al. Fludarabine and exposure-targeted busulfan compares favorably with busulfan/cyclophosphamide-based regimens in pediatric hematopoietic cell transplantation: maintaining efficacy with less toxicity. Biol Blood Marrow Transplant. 2014;20(3):345-353.

16. Gupta A, Downey M, Shanley R, et al. Reduced-toxicity (BuFlu) conditioning is better tolerated but has a higher second transplantation rate compared to myeloablative conditioning (BuCy) in children with inherited metabolic disorders. Biol Blood Marrow Transplant. 2020;26(3):486-492.

17. Peters C, Charnas LR, Tan Y, et al. Cerebral X-linked adrenoleukodystrophy: the international hematopoietic cell transplantation experience from 1982 to 1999. Blood. 2004;104(3):881-888.

18. Barker JN, Kurtzberg J, Ballen K, et al. Optimal practices in unrelated donor cord blood transplantation for hematologic malignancies. Biol Blood Marrow Transplant. 2017;23(6):882-896.

19. Glucksberg H, Storb R, Fefer A, et al. Clinical manifestations of graft-versus-host disease in human recipients of marrow from HLA-matched sibling donors. Transplantation. 1974;18(4):295-304.

20. Lee SJ. Classification systems for chronic graft-versus-host disease. Blood. 2017;129(1):30-37.

21. Kaplan EL, Meier P. Nonparametric estimation from incomplete observations. J Am Stat Assoc. 1958;53(282):457-481.

22. Kalbfleisch JD, Prentice RL. The Statistical Analysis of Failure Time Data. New York, NY: Wiley; 1980.

23. Kuhl JS, Kupper J, Baqué H, et al. Potential risks to stable long-term outcome of allogeneic hematopoietic stem cell transplantation for children with cerebral X-linked adrenoleukodystrophy. JAMA Netw Open. 2018;1(3):e180769.

24. Pierpont EI, Eisengart JB, Shanley R, et al. Neurocognitive trajectory of boys who received a hematopoietic stem cell transplant at an early stage of childhood cerebral adrenoleukodystrophy. JAMA Neurol. 2017;74(6):710-717.

25. Engelen M, Kemp S, de Visser M, et al. X-linked adrenoleukodystrophy (X-ALD): clinical presentation and guidelines for diagnosis, follow-up and management. Orphanet J Rare Dis. 2012;7(1):51.

26. Mallack EJ, Turk BR, Yan H, et al. MRI surveillance of boys with X-linked adrenoleukodystrophy identified by newborn screening: meta-analysis and consensus guidelines. J Inherit Metab Dis. 2021;44(3):728-739.

27. Lee S, Cliniard K, Young SP, et al. Evaluation of X-linked adrenoleukodystrophy newborn screening in North Carolina. JAMA Netw Open. 2020;3(1):e1920356.

28. Wiens K, Berry SA, Choi H, et al. A report on state-wide implementation of newborn screening for X-linked adrenoleukodystrophy. Am J Med Genet A. 2019;179(7):1205-1213.

29. Lampret BR, Remec ZI, Torkar AD, et al. Expanded newborn screening program in Slovenia using tandem mass spectrometry and confirmatory next generation sequencing genetic testing. Zdr Varst. 2020;59(4):256-263.

30. Barendsen RW, Dijkstra IME, Visser WF, et al. Adrenoleukodystrophy Newborn Screening in the Netherlands (SCAN Study): the X-factor [published correction appears in Front Cell Dev Biol. 2021;9:631655]. Front Cell Dev Biol. 2020;8:499.

31. Chiesa R, Wang J, Blok HJ, et al. Hematopoietic cell transplantation in chronic granulomatous disease: a study of 712 children and adults. Blood. 2020;136(10):1201-1211.

32. Lum SH, Miller WP, Jones S, et al. Changes in the incidence, patterns and outcomes of graft failure following hematopoietic stem cell transplantation for Hurler syndrome. Bone Marrow Transplant. 2017;52(6):846-853.

33. Pierpont EI, Nascone DR, Shanley R, et al. Neurocognitive benchmarks following transplant for emerging cerebral adrenoleukodystrophy. Neurology. 2020;95(5):e591-e600.