Early anteroposterior regionalisation of human neural crest is shaped by a pro-mesodermal factor

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Abstract The neural crest (NC) is an important multipotent embryonic cell population and its impaired specification leads to various developmental defects, often in an anteroposterior (A-P) axial level-specific manner. The mechanisms underlying the correct A-P regionalisation of human NC cells remain elusive. Recent studies have indicated that trunk NC cells, the presumed precursors of childhood tumour neuroblastoma, are derived from neuromesodermal-potent progenitors of the postcranial body. Here we employ human embryonic stem cell differentiation to define how neuromesodermal progenitor (NMP)-derived NC cells acquire a posterior axial identity. We show that TBXT, a pro-mesodermal transcription factor, mediates early posterior NC/spinal cord regionalisation together with WNT signalling effectors. This occurs by TBXT-driven chromatin remodelling via its binding in key enhancers within HOX gene clusters and other posterior regulator-associated loci. This initial posteriorisation event is succeeded by a second phase of trunk HOX gene control that marks the differentiation of NMPs toward their TBXT-negative NC/spinal cord derivatives and relies predominantly on FGF signalling. Our work reveals a previously unknown role of TBXT in influencing posterior NC fate and points to the existence of temporally discrete, cell type-dependent modes of posterior axial identity control.

Editor’s evaluation This paper presents an interesting model of a bi-phasic regulation for Hox genes in which Wnt drives HOX regulation in neuromesodermal precursors, but it does not control HOX levels in neural crest or...
spinal cord cells in human cells. The paper makes an important contribution to the literature and is of general interest.

Introduction

The neural crest (NC) is a multipotent cell population, which arises in the dorsal neural plate/non-neural ectoderm border region during vertebrate embryogenesis and generates a variety of cell types following epithelial-to-mesenchymal transition and migration through diverse routes. NC cells emerge at all levels of the anteroposterior (A-P) axis and their A-P position determines the identity of their derivatives: cranial NC produces mesoectodermal cell types (e.g., dermis, cartilage, and bone), melanocytes, neurons, and glial cells colonising the head; vagal NC cells, arising between somites 1 and 7, contribute (together with their sacral counterparts emerging at axial levels posterior to somite 28) predominantly to the enteric nervous system and include a subpopulation (termed cardiac NC) that generates various heart structures; trunk NC generates dorsal root/sympathetic neurons, adrenal chromaffin cells, and melanocytes (reviewed in Le Douarin et al., 2004; Rocha et al., 2020; Rothstein et al., 2018). Defects in the specification, differentiation, or migration of NC cells (known collectively as neurocristopathies) lead to a wide spectrum of serious developmental disorders, often in an axial level-specific manner (Pilon, 2021; Vega-Lopez et al., 2018). The use of human pluripotent stem cell (hPSC) differentiation offers an attractive platform for the study of human NC biology and neurocristopathies as well as an indefinite in vitro source of clinically relevant NC-associated cell populations for regenerative medicine applications. However, the cellular and molecular mechanisms directing the acquisition of distinct A-P axial identities by human NC cells remain largely undefined. In turn, this obviates the design of optimised hPSC differentiation protocols aiming to produce NC derivatives as well as the dissection of the links between human neurocristopathy emergence and the axial identity of the NC cells affected.

In vivo, the A-P patterning of the vertebrate embryonic body and its nascent cellular components relies on the coordinated action of Hox gene family members (arranged as paralogous groups [PG] in four distinct chromosomal clusters: A, B, C, and D). In mammals, transcriptional activation of Hox genes is initiated during gastrulation, within the posterior part of the embryo around the primitive streak and proceeds in a sequential manner reflecting their 3'-to-5' genomic order, a phenomenon described as temporal collinearity or the Hox clock (Deschamps and Duboule, 2017; Dollé et al., 1989; Izpisúa-Belmonte et al., 1991). The Hox clock continues to operate after gastrulation, until the end of somitogenesis, in the posterior growth zone comprising the caudal lateral epiblast-late primitive streak, and later the tail bud (Deschamps and Duboule, 2017; Wymeersch et al., 2019; Wymeersch et al., 2021). The colinear activation of Hox genes in the posterior growth region is thought to be tightly coupled to the assignment of the terminal A-P coordinates of the cell lineages that make up the postcranial axis, including the NC (Deschamps and Duboule, 2017; Wymeersch et al., 2019; Wymeersch et al., 2021). Eventually, the terminal expression domains of Hox genes along the A-P axis correspond to their ordering within their chromosomal clusters so that the earliest activated, 3' Hox PG members are expressed more anteriorly compared to their later-activated 5' counterparts. In the case of NC, anterior cranial NC is Hox-negative, posterior cranial NC cells express Hox PG(1-3) genes, while vagal NC cells are marked by the expression of Hox PG(3-5) members and positivity for Hox PG(5-9) genes denotes a trunk NC character (Rocha et al., 2020).

The post-gastrulation posterior growth zone is marked by high levels of WNT/FGF signalling activity and harbours a pool of multipotent axial progenitors driving embryonic axis elongation in a head-tail direction. These include neuromesodermal progenitors (NMPs) that generate a large fraction of the spinal cord as well as presomitic/paraxial mesoderm, the building blocks of the musculoskeleton (reviewed in Henrique et al., 2015; Wymeersch et al., 2021). NMPs are marked by the co-expression of neural and mesodermal genes, such as those encoding the transcription factors Sox2 and Brachyury (T) (Guillot et al., 2021; Henrique et al., 2015; Martin and Kimelman, 2012; Olivera-Martínez et al., 2012; Tsakiridis et al., 2014), as well as a number of other posteriorly expressed genes such as Nkx1-2, Cdx2, Tbx6, and Hox family members (Amin et al., 2016; Dias et al., 2020; Gouti et al., 2017; Guillot et al., 2021; Javali et al., 2017; Koch et al., 2017; Rodrigo Albors et al., 2018; Wymeersch et al., 2019; Young et al., 2009).
Fate mapping, lineage tracing, and single-cell transcriptomics studies in vertebrate embryos have revealed that trunk NC cells, which give rise to neuroblastoma (the most common extracranial solid tumour in infants) following oncogenic transformation, are derived from NMPs (Javali et al., 2017; Lukoseviciute et al., 2021; Shaker et al., 2021; Tzouanacou et al., 2009; Wymeersch et al., 2016). Moreover, recent work has demonstrated that the in vitro generation of trunk NC cells and their derivatives from hPSCs relies on the induction of an NMP-like intermediate following stimulation of WNT/FGF signalling pathways (Abu-Bonsrah et al., 2018; Faustino Martins et al., 2020; Frith et al., 2018; Frith and Tsakiridis, 2019; Hackland et al., 2019; Kirino et al., 2018). These and other studies have pointed to a model where a generic posterior axial identity is installed early within the NMP precursors of trunk NC cells under the influence of WNT/FGF activities prior to their differentiation and commitment to an NC fate (Frith et al., 2018; Metzis et al., 2018). However, it is unclear whether induction of a developmentally plastic, Brachyury-positive state is an obligatory step in the posteriorisation of prospective NC cells, or whether trunk NC cells can still acquire a posterior axial identity without passing through a mesoderm-competent progenitor stage.

Here we dissect the links between human NMP induction and trunk NC specification using hPSC differentiation as a model. We show that disruption of NMP ontogeny via knockdown of the NMP/ mesoderm regulator TBXT (the human homologue of Brachyury) abolishes the acquisition of a posterior axial identity by NC and early spinal cord cells, as well as the adoption of an NC fate. This is linked to a failure of hPSC-derived NMPs to activate properly and subsequently maintain, during their differentiation, the expression of HOX genes and key signalling pathway components. We demonstrate that TBXT regulates these events by directly orchestrating an open chromatin landscape in regions associated with HOX clusters and loci encoding pivotal posterior/NC identity determinants. Our results also indicate that, as TBXT levels gradually decrease to extinction during the differentiation of NMPs toward early spinal cord/NC cells, the control of trunk HOX gene expression/posteriorisation becomes WNT-independent and relies predominantly on FGF signalling activity. Collectively, our data reveal a previously unappreciated role for TBXT in prospectively shaping the A-P patterning and specification of NC cells. They also indicate the existence of two distinct phases of posterior axial identity control: (i) an early NMP-based phase that involves the TBXT/WNT-driven establishment/fixed of a HOX-positive, posterior character in uncommitted progenitors, and, (ii) a later one, which is defined by the FGF-based maintenance of a posterior axial identity within the early spinal cord/NC derivatives of NMPs.

Results
NC-fated axial progenitors are marked by T expression

We and others have recently shown that hPSC-derived neuromesodermal-potent axial progenitors exhibiting expression of TBXT, a well-established regulator of NMP/axial progenitor maintenance (Amin et al., 2016; Gouti et al., 2014; Guibentif et al., 2021; Koch et al., 2017; Martin and Kimelman, 2010; Rashbass et al., 1991), are the optimal source of trunk NC cells in vitro (Cooper et al., 2022; Frith et al., 2018; Frith and Tsakiridis, 2019; Gomez et al., 2019; Hackland et al., 2019). In the mouse embryo, homotopic grafting-based fate mapping has indicated that the equivalent NM-potent axial progenitors giving rise to trunk NC are located in the lateral-most caudal epiblast of somitogenesis-stage embryos (marked as ‘LE’ in Figure 1; Wymeersch et al., 2016). We confirmed by wholemount immunofluorescence analysis that, similar to their human in vitro counterparts, the LE domain in mouse is also marked by expression of T (Brachyury) immediately before the time of early posterior NC emergence (embryonic day [E] 8.75–E9.0. Theiler stage [TS] 13–14) (Figure 1A). Low-level expression of T/Fp2a, potentially marking nascent NC cells (Mitchell et al., 1991; Rothstein and Simoes-Costa, 2020), was first detected within the posterior LE domain from E9.0 (Figure 1B-g-h vs B-c-d). ‘Committed’ NC cells, located anteriorly at the same time points, were found to also express Sox9 alongside T/Fp2a (Figure 1B-a-b and B-e-f). These data demonstrate that NC-fated axial progenitors in the mouse embryo exhibit T expression in line with the previous lineage tracing studies showing that T-positive embryonic cells contribute to the posterior neural tube and trunk NC (Anderson et al., 2013; Mugele et al., 2018; Shaker et al., 2021).
TBXT controls posterior axial identity acquisition by NMP-derived NC cells

Our embryo analysis results suggest that the induction of trunk NC may rely on the prior generation of mesoderm-competent, T+axial progenitors emerging in the posterior growth zone during embryonic axis elongation. To test this hypothesis, we examined the effects of attenuating TBXT during human embryonic stem cell (hESC) differentiation using an hESC line engineered to exhibit short hairpin RNA (shRNA)-mediated, tetracycline (Tet)-inducible knockdown of TBXT expression (Bertero et al., 2016). We first generated NMPs from these hESCs following their treatment with the WNT agonist CHIR99021 (CHIR) and FGF2 for 3 days (Frith et al., 2018), in the presence or absence of Tet (Figure 2A). We confirmed that Tet addition induced a considerable reduction in TBXT expression both at the transcript and protein level compared to untreated controls (Figure 2B–D). No effect on TBXT induction was observed in NMPs generated from an isogenic control Tet-inducible shRNA hESC line targeting the B2M gene (Bertero et al., 2016) following Tet treatment (Figure 2—figure supplement 1A).
Figure 2. Effect of TBXT reduction on human embryonic stem cell (hES)-derived neuromesodermal progenitors (NMPs). (A) Differentiation/tetracycline (Tet) treatment scheme. (B) Immunofluorescence analysis of the expression of TBXT in shRNA hESC-derived NMPs in the presence and absence of Tet. Scale bar = 100 μm. (C) Mean fluorescence intensity of TBXT protein in Tet-treated and control NMP cultures. (D) Normalised expression values of TBXT transcripts in control, Tet-treated NMPs and undifferentiated hESC samples following RNA sequencing (RNA-seq) analysis. (E) Gene ontology (GO) term analysis of differentially expressed genes in hESC-derived NMPs following TBXT knockdown. (F) Representative significantly downregulated and upregulated transcripts following TBXT depletion. (G) Quantitative PCR (qPCR) expression analysis of indicated HOX genes in control vs Tet-treated NMPs. Error bars represent SD (n=3). *p<0.05, **p<0.005, ***p<0.0005 (paired t-test). n.s. not significant.

Figure 2 continued on next page
To define the impact of TBXT depletion during the transition of hPSCs toward an NMP state, we carried out transcriptome analysis of Tet-treated and control hESCs cultured in NMP-inducing conditions (i.e. presence of CHIR and FGF2) for 3 days, using RNA sequencing (RNA-seq). We found that 346 and 293 genes were significantly up- and down-regulated respectively in TBXT knockdown cells compared to untreated controls (padj<0.05, Wald test; log2FC>|0.5|; Supplementary file 1). Gene ontology (GO) biological processes enrichment analysis revealed that most differentially expressed genes were established regulators of A-P regionalisation/posterior development (Figure 2E, Supplementary file 2). Significantly downregulated genes included presumptive mesoderm-associated transcription factors (TBX6, MSGN1, and FOXC1/2) as well as WNT (e.g. RSPO3, WISP1, WNT5A/B, WNT8A, and LEF1), FGF (FGF3/8/17 and DUSP7), and NOTCH (HES7 and DLL1/3) signalling pathway-linked transcripts (Figure 2F, Supplementary file 1), most of which have been reported to be present in NMPs and their immediate mesodermal derivatives in vivo (Guillot et al., 2021; Koch et al., 2017; Wymeersch et al., 2019). Moreover, we found that TBXT knockdown triggered a significant reduction in the expression of various HOX genes belonging to anterior, central, and posterior PG(1-9) (Figure 2G, Supplementary file 1) while no effect was observed in Tet-treated B2M shRNA hESC-derived NMPs (Figure 2—figure supplement 1B) ruling out the possibility that the observed decrease in HOX transcript levels may be due to nonspecific effects of Tet and/or interference of shRNAs with the microRNA processing machinery. On the contrary, the most highly upregulated genes included anterior visceral endoderm (AVE)/endoderm, anterior neurroectoderm, and pluripotency/early post-implantation epiblast-associated genes such as SOX17, LEFTY1/2, OTX2, CER1, HHEX, NANOG, and GDF3 (Figure 2F, Supplementary file 1). Taken together, these results indicate that TBXT knockdown severely impairs the induction of NMPs and their immediate presomitic mesoderm progenitor derivatives from hPSCs. They also suggest that TBXT directs the establishment of a posteriorising signalling environment associated with early HOX gene activation, upon pluripotency exit, since in its absence hPSCs adopt an identity that resembles the anterior epiblast despite the combined presence of caudalising WNT (CHIR) and FGF (FGF2) signalling agonists.

We next assessed the effect of TBXT disruption/failure to induce NMPs/presomitic mesoderm on trunk NC specification. To this end, we attempted to generate trunk NC cells from inducible TBXT knockdown shRNA hESCs via an NMP intermediate. We treated cells initially with CHIR-FGF2 for 3 days to induce NMPs, followed by their replating under NC-promoting conditions (i.e. low CHIR and moderate BMP levels) for a further 5 days, as previously described (Frith et al., 2018; Frith and Tsakiridis, 2019) either in the continuous presence or absence of Tet (Figure 3A). We found that the expression of most HOX PG(1-9) genes examined was dramatically reduced in the resulting Tet-treated cultures compared to their untreated counterparts (Figure 3B and C) indicating that the failure of TBXT-depleted NMPs to properly induce HOX gene expression persists in their NC derivatives. The expression levels of CDX2, an NMP/spinal cord/early trunk NC regulator (Sanchez-Ferras et al., 2016; Sanchez-Ferras et al., 2012) were also found to be severely decreased (Figure 3D). A statistically significant reduction in the levels of some NC-associated transcripts such as SOX10 and PAX3 was observed in the presence of Tet (p<0.05 and p<0.01 respectively; paired t-test; Figure 3D) though no change in SOX10 protein levels was detected (Figure 3E). We also observed a trend towards a slight upregulation in the expression of the anterior NC markers OTX2 and ETS1 (Simoes-Costa and Bronner, 2016) as well as the neural progenitor marker SOX1 (Wood and Episkopou, 1999) following TBXT knockdown (Figure 3D). However, we found no significant (paired t-test) expansion of the minor fraction of SOX1 protein-positive cells present in the trunk NC cultures, indicating that TBXT knockdown does not induce a cell identity switch from NC to CNS neurroectoderm (Figure 3—figure supplement 1). Based on these data we conclude that the acquisition of a posterior axial identity by trunk NC cells is shaped by the action of TBXT, an NMP/pro-mesodermal regulator.
TBXT-driven programming of a posterior NC axial identity is an early event

To further dissect the temporal role of TBXT in controlling HOX gene expression dynamics, we assessed the effect of manipulating its levels via Tet treatment during distinct time windows of NMP induction and differentiation (Figure 4A). We opted to differentiate day 3 TBXT shRNA hESC-derived NMPs in the presence of basal, serum-free media devoid of any signalling pathway agonists/antagonists for a further 4 days to minimise any potential influence of the extrinsic signalling environment on the expression status of HOX genes. Under these conditions, human NMPs give rise mainly to Tlow/-SOX2+ posterior neural progenitors (Gouti et al., 2014) as well as more differentiated, mutually exclusive NC and CNS spinal cord subpopulations marked by SOX10 and SOX1 protein expression respectively (Figure 4C and D). We found that both early (days 0–3; NMP induction) and late (days 3–7; NMP...
Figure 4. TBXT-driven programming of a posterior neural crest (NC) axial identity is an early event. (A) Differentiation/tetracycline (Tet) treatment scheme. The temporal Tet treatment regimens employed were: no Tet control (−−); Tet treatment between days 3 and 7 (−+); Tet treatment between days 0 and 7 (++); Tet treatment between days 0 and 3 (+−). (B) qPCR expression analysis of representative lineage identity/posterior markers in early spinal cord/NC cells generated following culture in ‘neutral’ basal media in the presence of Tet during the time windows shown in A. Error bars represent SD (n=3). Only statistically significant changes are indicated. *p<0.05, **p<0.005 (paired t-test). (C) Immunofluorescence analysis of the expression of HOXC9, SOX1, and SOX10 in early spinal cord/NC cells generated following culture in ‘neutral’ basal media in the presence of Tet during the time windows shown in A. Scale bar = 100 µm. Quantification of the cell populations expressing these proteins in relation to the different Tet treatment regimens is shown in (D). Error bars represent SD (n=3). H: HOXC9; S1: SOX1; S10: SOX10. (E) qPCR expression analysis of indicated HOX
genes in early spinal cord/NC cells generated following culture in ‘neutral’ basal media in the presence of Tet during the time windows shown in A. Error bars represent SD (n=3). Only statistically significant changes are indicated. *p<0.05, **p<0.005 (paired t-test).

differentiation) time window Tet treatments resulted in a marked reduction in TBXT transcript levels but had a minor impact on the levels of early and definitive neural identity markers (such as SOX2 and SOX1, respectively) on day 7 of differentiation (Figure 4B). However, we observed a statistically significant decrease in the number of cells co-expressing SOX1 and HOXC9 proteins (denoting a CNS posterior spinal cord identity) in the case of cultures treated with Tet between days 0 and 3 compared to their untreated counterparts (from ~10 to ~1%; p<0.05, paired t-test) (Figure 4C; compare dark blue bars in the middle graph in Figure 4D). A more pronounced effect of continuous (days 0–7) TBXT knockdown on both SOX10 transcript and protein levels was also detected: the mean percentage of SOX10+ cells decreased from ~23 to ~3.6% (p<0.05, paired t-test) while the mean percentage of SOX10+HOXC9+ cells decreased from ~20 to ~1% (p<0.05, paired t-test; Figure 4B and C; compare purple bars in the top graph in Figure 4D). The sharp decline in HOXC9 protein positivity linked to early Tet treatment (Figure 4C and D) was accompanied by a global downregulation in the expression of all HOX genes examined by quantitative PCR (qPCR) as well as CDX2 (Figure 4B and E). On the contrary, late days 3–7 Tet treatment had a minimal effect on HOX/CDX2 transcription (Figure 4B and E). Together, these data further confirm that TBXT controls the acquisition of a posterior axial identity in prospective spinal cord/NC cells and the induction of an NC fate in the absence of any extrinsic signals. Moreover, our results indicate that TBXT exerts its effect on these processes at an early stage that temporally coincides with the induction of NMPs from pluripotent cells.

**Early encoding of a posterior axial identity in NMP-derived NC cells is primarily WNT-dependent**

We next sought to decipher the interplay between extrinsic signals and TBXT in orchestrating the establishment of an NC posteriorising environment. Our RNA-seq data revealed the concomitant downregulation of WNT targets and upregulation of WNT antagonists (e.g. DKK4) in day 3 FGF-CHIR-treated cultures in the presence of Tet (Supplementary file 1, Figure 2F) suggesting that the inability of TBXT-depleted NC cells to activate HOX genes may be linked to an early reduction in WNT activity. This is further supported by our observation that TBXT knockdown-triggered reduction of HOX gene expression in our NC cultures was also accompanied by a significant reduction in the expression of WNT signalling target genes such as TCF1, LEF1, and CDX2 (Figure 3D). We, therefore, examined the temporal effects of WNT signalling perturbation on posterior axial identity acquisition during the transition from hPSCs to NMPs and subsequently NC. We first assessed the effects of WNT signalling inhibition during the 3-day induction of NMPs from hPSCs, by removing CHIR and treating the cultures with the tankyrase inhibitor XAV939 (XAV; Huang et al., 2009) in order to eliminate any endogenous paracrine signalling, in the presence of FGF2 (the other major signalling agonist included in the media; Figure 5A). Given the reported role of FGF signalling in controlling the HOX clock in NMPs and their derivatives (Delfino-Machín et al., 2005; Hackland et al., 2019; Liu et al., 2001; Mouilleau et al., 2021; Nordström et al., 2006), we also tested the effect of attenuating this pathway in a similar manner, by omitting FGF2 and culturing cells only in the presence of CHIR and the FGF pathway-MEK1/2 inhibitor PD0325901 (PD03) between days 0 and 3 (Figure 5A). Signalling inhibition was verified by confirming the downregulation/extension of WNT (AXIN2, TCF1, and LEF1) and FGF signalling target genes (SPRY4) in inhibitor-treated D3 cultures relative to untreated controls (Figure 5B). Our results confirmed that maximal induction of NMP markers (TBXT, CDX2, and NKX1-2) and all HOX genes examined can be achieved only in the combined presence of WNT and FGF agonists (Figure 5C and D, white bars). However, WNT signalling stimulation alone, in the absence of any FGF activity, rescued the induction of most HOX genes and TBXT/CDX2 with variable efficiency (Figure 5D, red bars) indicating that WNT is the main instructive pathway controlling global HOX transcription and the expression of major axis elongation regulators during the differentiation of hPSCs toward NMPs. The HOXC genes were an exception, as their activation was found to be equally dependent on WNT and FGF signalling (Figure 5D) suggesting the additional existence of HOX cluster-specific modes of transcriptional control, in line with similar findings in the embryo (Ahn et al., 2014; van den Akker et al., 2001).
Figure 5. Early programming of a posterior axial identity in neuromesodermal progenitor (NMP)-derived neural crest (NC) cells is primarily WNT-dependent. (A) Scheme of treatments during the differentiation of human pluripotent stem cells (hPSCs) toward NMPs. (B) qPCR expression analysis of representative WNT-FGF targets in NMP cultures treated with the indicated combinations of WNT-FGF agonists/antagonists. Error bars represent SD (n=3–4). In all cases, changes (agonist-antagonist treatments vs corresponding WNT-FGF controls) were significantly different (ratio paired t-test) unless otherwise stated (n.s. not significant). (C–D) qPCR expression analysis of key NMP markers (C) and HOX genes (D) in NMP cultures treated with the indicated combinations of WNT-FGF agonists/antagonists. Error bars represent SD (n=3–4). In all cases, changes (agonist-antagonist treatments vs corresponding WNT-FGF controls) were significantly different (ratio paired t-test). (E) Scheme of treatments during the differentiation of hPSC-derived NMPs toward trunk NC cells. (F–G) qPCR expression analysis of representative lineage-specific, axial identity (F) and HOX genes (G) in NMP-derived trunk NC cultures treated with the indicated combinations of WNT-BMP agonists/antagonists. Error bars represent SD (n=3). Statistically significant changes are indicated. *p<0.05, **p<0.005.

The online version of this article includes the following figure supplement(s) for figure 5:

Figure supplement 1. Influence of WNT and BMP signalling on trunk NC specification.

Figure supplement 2. WNT signalling dynamics during posterior NC emergence.
We next interrogated the signalling pathway dependence of HOX gene expression during the transition of NMPs toward trunk NC cells, which involves a 5-day treatment with a lower amount of the WNT agonist CHIR and an intermediate level of BMP activity; the latter is achieved through the addition of a saturating amount of recombinant BMP4 together with the BMP type I receptor inhibitor DMH-1 to antagonistically modulate the effects of BMP4 (Figure 5E; Frith et al., 2018; Frith and Tsakiridis, 2019; Hackland et al., 2017). We examined the effect of perturbing the two main signalling pathways driving the specification of NC from NMPs (WNT and BMP) using agonist/antagonist combinations between days 3 and 8, in a manner similar to the days 0–3 treatments (Figure 5E). Treatment of differentiating NMPs with XAV to inhibit WNT signalling in combination with BMP stimulation resulted in a statistically significant reduction in the expression of many of the HOX gene family members examined albeit to a lesser degree compared to days 0–3 (Figure 5G, dark blue vs purple bars). No significant decrease in the overall number of cells expressing HOXC9 protein was observed (Figure 5—figure supplement 1). WNT signalling attenuation in the presence of BMP activity also led to a moderate increase in the expression of anterior (OTX2 and ETS1) and a decrease in posterior (CDX2) NC markers while the levels of neural/NC-specific transcripts appeared unaffected (Figure 5F, dark blue vs purple bars). On the contrary, WNT stimulation in combination with the BMP inhibitor LDN193189 (LDN) resulted in significant upregulation in the expression of most HOX genes relative to the WNT-BMP controls (Figure 5G, light blue vs purple bars) with overall HOXC9 protein levels being unaffected (Figure 5—figure supplement 1). Moreover, this treatment resulted in the complete extinction of the expression of the definitive NC marker SOX10 and a concomitant increase in the CNS neural progenitor marker SOX1 both at the transcript and protein levels (Figure 5F, light blue vs purple bars; the percentage of SOX1 + cells increased from 0.7 to 45%, p<0.05 paired t-test; Figure 5—figure supplement 1, compare dark blue bars in the graph in Figure 5—figure supplement 1B) pointing to a role for BMP signalling in steering NMPs toward an NC fate at the expense of a spinal cord neuroectoderm identity, in agreement with previous observations (Leung et al., 2016). Taken together, our results demonstrate that HOX gene activation in hPSC-derived NMPs and its tight coupling to the early programming of a posterior axial identity in NMP-derived NC cells are primarily driven by a WNT-TBXT regulatory loop. However, the early dependence of HOX gene expression on WNT signalling diminishes with time as NMPs gradually differentiate toward NC.

We went on to examine whether the reliance of early trunk NC progenitor patterning on WNT signalling occurs in vivo. We examined mouse R26-WntVis reporter embryos, in which graded nuclear EGFP expression relates to the degree of WNT signalling strength (Takemoto et al., 2016), at the time of posterior NC formation (E8.75–E9.0, TS13–TS14). In agreement with previous work (Ferrer-Vaquer et al., 2010), the highest levels of WNT activity were confined within the T+ posterior growth region (Figure 5—figure supplement 2A). Wholemount immunostaining followed by 3D modelling of the high anti-GFP (a-GFP)-positive fraction (a-GFP<sup>100%</sup>) demonstrated that NC-fated regions within the T+ lateral-most caudal epiblast domain are marked by higher WNT signalling levels compared to more rostral epiblast positions (dashed lines in Figure 5—figure supplement 2B). These findings further confirm that early specification of NM-potent T+ trunk NC progenitors correlates with high WNT activity. However, this association becomes less prevalent as cells gradually commit to a definitive NC fate in line with our in vitro observations showing the progressively decreasing WNT-dependence of TBXT-driven posterior axial identity programming in NC cells.

**Posterior axial identity acquisition by WNT-FGF-induced pre-neural spinal cord progenitors is TBXT-independent and FGF-dependent**

Our data so far indicate that TBXT affects the induction of a posterior axial identity in NC and CNS spinal cord derivatives of NMPs in extrinsic signal-free culture conditions (Figure 4C and D). We have recently reported the generation of early pre-neural spinal cord progenitors following a culture of hPSC-derived NMPs under the influence of combined WNT and high FGF signalling activity; these progenitors can then be directed to efficiently generate posterior motor neurons of a thoracic axial identity (Wind et al., 2021). We thus examined whether TBXT knockdown disrupts HOX gene expression dynamics in the presence of these signals (i.e. combined WNT-high FGF). To this end, we derived early pre-neural spinal cord progenitors from TBXT shRNA hESCs: cells were treated with CHIR-FGF2 for 3 days to produce NMPs, followed by their replating and further culture in the presence of CHIR and high FGF levels for 4 days, combined with continuous (i.e. throughout the entire
7-day differentiation) Tet treatment to mediate TBXT knockdown (Figure 6A). The levels of most HOX transcripts examined were largely unaffected in day 7 pre-neural spinal cord progenitors generated from TBXT-depleted cells and similar to untreated controls (Figure 6B). No dramatic changes in the expression of CDX2 and SOX2 (markers of an early pre-neural spinal cord character at this stage) were observed (Figure 6C). In line with our previous observations and published in vivo data (Gofflot et al., 1997; Nordström et al., 2006; Wind et al., 2021), we also detected low levels of TBXT transcripts in control cultures, and surprisingly, these were comparable to their Tet-treated counterparts (Figure 6C). This finding suggests that selection of cells evading shRNA knockdown and maintaining low levels of TBXT may occur upon culture in pre-neural spinal cord cell-promoting culture conditions. Later addition of Tet on day 3, at the start of NMP differentiation toward pre-neural spinal cord cells, appeared to restore efficient TBXT knockdown (Figure 6D and F) but, again, had no major impact on either HOX PG(1-9) gene expression or the levels of the early spinal cord/pre-neural transcripts CDX2 and NKX1-2 in the resulting day 7 cultures (Figure 6E and F). These results indicate that HOX
expression dynamics/posterior axial identity in NMP-derived, pre-neural spinal cord progenitors, generated under the influence of WNT/high FGF signalling activity, are largely unaffected by TBXT depletion. They also suggest that the combined presence of extrinsic WNT/FGF signals compensates for the reduced levels of TBXT, rescuing the effects of its knockdown on HOX gene expression in ‘neutral’ culture conditions (Figure 4).

We next investigated in more detail the role of WNT and FGF in the posteriorisation of pre-neural spinal cord progenitors since both signalling pathways have been previously implicated in the control of the HOX gene expression in the neural tube, through cooperation with the key axis elongation factor CDX2 (Bel-Vialar et al., 2002; Lippmann et al., 2015; Liu et al., 2001; Mazzoni et al., 2013; Metzis et al., 2018; Mouilleau et al., 2021; Nordström et al., 2006; Olivera-Martinez et al., 2014; Takemoto et al., 2006; van de Ven et al., 2011). We carried out signalling agonist/inhibitor combination experiments as described earlier (Figure 5) treating hPSC-derived NMPs differentiating toward pre-neural spinal cord progenitors (following their replating in the presence of CHIR-high FGF2) either with the WNT inhibitor XAV in the presence of FGF2 or the FGF/MEK inhibitor PD03 in combination with CHIR between days 3 and 7 of differentiation (Figure 7A). We found that WNT signalling inhibition in the presence of FGF activity during this time window does not appear to have a major effect on HOX or CDX2 transcript levels (blue vs grey bars in Figure 7B and C). In contrast, blocking FGF signalling in combination with CHIR treatment resulted in a dramatic reduction in the expression of most HOX genes examined, particularly those belonging to PG(4-9), as well as CDX2 (red bars vs grey bars, Figure 7B and C). The NMP/pre-neural spinal cord marker NKX1-2 was equally affected by the two treatments while the levels of later spinal cord markers (PAX6 and SOX1) were modestly elevated (Figure 7C). Immunofluorescence analysis further revealed that FGF signalling attenuation in the presence of CHIR results in the elimination of SOX2'HOX9+' (their mean percentage decreased from ~45 to ~9%; n=3 p<0.05 paired t-test; Figure 7D, compare purple bars in the graph in Figure 7B) and CDX2'HOX9+' pre-neural spinal cord populations (Figure 7—figure supplement 1). Based on these data, we conclude that maintenance of a posterior axial identity in NMP-derived pre-neural early spinal cord cells is driven primarily by FGF signalling, which gradually becomes functionally equivalent to TBXT in controlling trunk HOX gene/CDX2 expression as NMP differentiation progresses.

### TBXT controls posterior axial identity acquisition by influencing chromatin accessibility

We next examined whether TBXT controls HOX gene activation and potentiates the action of posteriorising extrinsic signals in CHIR-FGF-treated hPSCs via direct genomic binding or, indirectly, by affecting the expression of other key axis elongation regulators such as CDX2. To this end, we carried out chromatin immunoprecipitation followed by high-throughput sequencing (ChIP-seq) on hESC-derived NMPs and undifferentiated hESCs (control) to map TBXT targets genome-wide. We identified 24,704 TBXT-binding regions in NMPs (Supplementary file 3, Figure 8—figure supplement 1A), a large fraction of which were located within introns (~50%) and distal intergenic (~38%) regions (Figure 8—figure supplement 1B), reflecting similar findings on mouse T genomic binding (Beisaw et al., 2018; Tosic et al., 2019). A small number (n=2088) of non-overlapping TBXT binding sites was also detected in undifferentiated hESCs, in line with the reported low-level expression of this transcription factor in mesoderm-biased pluripotent cells (Stavish et al., 2020; Tsakiridis et al., 2014). The GO biological processes enrichment analysis of target genes associated with TXBT binding sites around the transcriptional start site (~2000 to +500 bp; 3084 peaks, 2841 genes) revealed an over-representation of established developmental regulators of A-P regionalisation (Benjamini–Hochberg padj <0.05; Figure 8—figure supplement 1C, Supplementary file 4). Transcription factor motif enrichment analysis revealed that the TBXT binding regions in hPSC-derived NMPs were enriched, as expected, for the Brachyury consensus binding motif as well as other T-box binding motifs such as EOMES and TBX6 (Figure 8A, Supplementary file 5), in agreement with previous findings in the mouse demonstrating that a fraction of the genomic targets of these transcription factors are also occupied by TBXT (Koch et al., 2017; Tosic et al., 2019). Other over-represented motifs included WNT signalling effectors (LEF1/TCF3) and Homeobox genes such as CDX factors (CDX2/4) and various HOX family members (Figure 8A, Supplementary file 5) reflecting possible cooperative binding between these factors and TBXT in human NMPs. Comparison of TBXT ChIP-seq targets with our list
Figure 7. Posterior axial identity acquisition by NMP-derived pre-neural spinal cord progenitors is FGF-dependent. (A) Scheme of treatments during the differentiation of hPSC-derived NMPs toward early spinal cord progenitors. (B–C) qPCR expression analysis of indicated HOX genes (B) and representative NMP/early spinal cord/neural markers (C) in NMP-derived early spinal cord progenitor cultures generated as depicted in A. Error bars represent SD (n=3). Statistically significant changes are indicated. *p<0.05, **p<0.005. (D) Immunofluorescence analysis of SOX2 and HOXC9 protein expression in NMP-derived early spinal cord progenitor cultures treated with the indicated combinations of WNT-FGF agonists/antagonists. Quantification of the cell populations expressing these proteins in relation to the different treatment regimens is shown in (E). Error bars represent SD (n=3). H: HOXC9; S2: SOX2.

The online version of this article includes the following figure supplement(s) for figure 7:

Figure supplement 1. Posterior axial identity acquisition by NMP-derived pre-neural spinal cord progenitors is FGF-dependent.
Figure 8. TBXT controls HOX gene expression and WNT signalling in NMPs by influencing chromatin accessibility. (A) Representative transcription factor-binding motifs enriched in TBXT binding sites. (B) Graph showing the percentages of differentially expressed genes (padj<0.05, log2FC>|1|) following TBXT knockdown during the transition of hESCs toward NMPs that are bound directly by TBXT. (C) Table showing all differentially expressed genes following TBXT knockdown that exhibit TBXT binding within their promoter region. (D) Volcano plot of differentially accessible ATAC-seq peaks between TBXT-depleted and control NMPs. (E) Graph showing the number of significantly upregulated and downregulated direct TBXT targets in relation to changes in chromatin accessibility associated with TBXT knockdown. (F) Correspondence between TBXT binding (ChIP) and chromatin accessibility changes (ATAC-seq) in the presence and absence of tetracycline (Tet) in indicated HOX clusters, WNT/presomitic mesoderm-linked loci.

Figure 8 continued on next page
of differentially expressed genes following Tet-induced TBXT knockdown in day 3 FGF-CHIR treated cultures (Figure 2) revealed that the majority (~85%) of the downregulated (padj <0.05, log2FC>|1|) genes, including HOX PG(1-9) members, pro-mesodermal factors and WNT-FGF signalling components, were directly bound by TBXT, while most (~60%) of their upregulated counterparts were not (Figure 8B, C and F, Figure 8—figure supplement 1D, Supplementary file 6). This indicates that TBXT acts primarily as a transcriptional activator of key downstream NMP/mesoderm regulators and HOX genes as previously reported in the mouse (Amin et al., 2016; Beisaw et al., 2018; Guibentif et al., 2021; Koch et al., 2017; Lolas et al., 2014).

To further dissect the impact of TBXT binding, we interrogated the chromatin accessibility landscape in TBXT-depleted and control hESC-derived NMPs using ATAC-seq (assay for transposase accessible chromatin with high-throughput sequencing; Buenrostro et al., 2013). We detected 4421 and 4027 unique regions corresponding to loss or gain of chromatin accessibility (a readout of DNA regulatory element/enhancer activity) following Tet treatment, respectively (distance from TSS ranging from –1–1 kb, log2FC cutoff = 2 and p-value<0.05; Figure 8D, Supplementary file 7). Genes associated with loss of chromatin accessibility were predominantly linked to A-P patterning, mesoderm specification, and WNT signalling (Supplementary file 7). Moreover, the Tet-induced downregulation of most transcripts found to be TBXT targets (Supplementary file 3) was also accompanied by a significant loss of ‘open’ chromatin regions within the TBXT binding sites (Figure 8E and F, Supplementary file 7). These included sites that were dispersed within all HOX clusters as well as key presomitic mesoderm regulators (e.g. TBX6 and MSGN1) and WNT signalling components (e.g. WNT3A/8 A and LEF1; Figure 8F, Figure 8—figure supplement 1D). On the contrary, there was no obvious correlation between upregulation of expression following TBXT knockdown and chromatin accessibility change (Figure 8E, Supplementary file 7). Gain of accessible regions in TBXT-depleted hPSC-derived NMPs appeared to occur predominantly within/around loci linked to neural development (e.g. SOX3 and DLG2; Tao et al., 2003; Wood and Episkopou, 1999), most of which were not bound directly by TBXT (Figure 8G, Supplementary file 7). Moreover, examination of transcription factor binding motifs (annotated using HOMER; Heinz et al., 2010; Benjamini–Hochberg padj<0.05) showed that ATAC-seq sites marked by loss of chromatin accessibility following TBXT knockdown were uniquely enriched in CDX, T-box factor and HOX binding-associated DNA sequence elements whereas the regions marked by a gain of chromatin accessibility were mainly characterised by motifs indicating binding of SOX, OCT/POU, and Forkhead family transcription factors (Figure 8—figure supplement 1E, Supplementary file 8).

Further ATAC-seq analysis of trunk NC cells generated from hESC-derived NMPs in the continuous presence (from days 0–8) and absence of Tet revealed that, like in NMPs, various key loci associated with posterior patterning such as HOX clusters and CDX2 were marked by a significant loss of ‘open’ chromatin regions in TBXT knockdown cultures (Figure 8—figure supplement 2A,B; Supplementary file 9). We also detected a major disruption of the chromatin accessibility landscape within/around WNT/BMP/FGF signalling-linked genes in Tet-treated trunk NC cells (Figure 8—figure supplement 2C,D; Supplementary file 9). Some affected components of these pathways, such as WNT1 and BMP7, have been shown to play a pivotal role in trunk NC specification and delamination (Burstyn-Cohen et al., 2004; Nguyen et al., 2000). Many of the disrupted regions, particularly HOX-associated enhancers, were bound by TBXT and exhibited reduced chromatin accessibility already at the NMP stage (see boxed areas in Figure 8—figure supplement 2A). However, we also observed a loss of trunk NC-specific accessible regions, which were not direct TBXT genomic targets in NMPs (marked by red asterisks in Figure 8—figure supplement 2). No major changes in chromatin accessibility were detected in Tet-treated day 7 early spinal cord cultures compared to untreated controls (Figure 8—figure supplement 2) reflecting the minimal impact of TBXT knockdown under these conditions.
Collectively, these data suggest that TBXT actively reconfigures the chromatin landscape in HOX clusters and other axis elongation-associated loci including WNT signalling components, by direct binding to key enhancer elements to promote the acquisition of a posterior axial character/mesoderm identity during the transition of pluripotent cells toward an NMP state. Some of these TBXT-controlled, NMP-specific ‘posteriorisation’ enhancers appear to remain active at later stages, in TBXT-negative trunk NC derivatives of NMPs. Moreover, our data also demonstrate the existence of trunk NC-specific enhancers that are influenced indirectly by TBXT, possibly through its binding and regulation of critical upstream transcription factors (e.g. CDX2) or signalling specifiers of NC fate during the early TBXT-positive stages of the transition of NMPs towards trunk NC.

Discussion

A number of recent studies have demonstrated that the emergence of the trunk NC is tightly coupled to the induction of NM-potent progenitors localised in the post-gastrulation/somitogenesis stage posterior growth region of vertebrate embryos. Here we provide evidence indicating that the encoding of a posterior axial identity in human NC/early spinal cord cells, embodied by the sequential activation of Hox genes and the induction of posterior markers, relies on their transition through this developmentally plastic entity. This ‘primary regionalisation’ process (Metzis et al., 2018) is mediated by the action of the pro-mesodermal transcription factor TBXT, which, together with WNT signalling effectors and, possibly, CDX2, directs global HOX gene cluster transcription and the expression of posterior/presomitic mesoderm regulators (Figure 9). The early WNT dependence of HOX expression dynamics diminishes during the transition of NMPs towards their NC/spinal cord derivatives; this second phase of HOX gene expression maintenance is in turn controlled predominantly by FGF signalling (Figure 9). Our data also revealed another novel role of TBXT, namely its influence on the adoption of an NC but not CNS spinal cord lineage identity.

Our proposed model synthesises previous findings showing that WNT-driven posterior axial identity acquisition in neural derivatives of NMPs takes place prior to neural induction (Lippmann et al., 2015; Metzis et al., 2018; Neijts et al., 2017; Neijts et al., 2016; Takemoto et al., 2006) with data illustrating that temporally discrete modes of trunk axial patterning in CNS spinal cord/NC rely on both FGF and WNT activities (Bel-Vialar et al., 2002; Delfino-Machín et al., 2005; Dunty et al., 2014; Hackland et al., 2019; Liu et al., 2001; Mazzoni et al., 2013; Mouilleau et al., 2021; Muhr et al., 1999; Nordström et al., 2006; Sanchez-Ferras et al., 2016; Sanchez-Ferras et al., 2014; Sanchez-Ferras et al., 2012; van Rooijen et al., 2012; Zhao et al., 2014). Our work also reflects results showing the existence of distinct phases of Hox gene expression program implementation in vivo: (i) an early ‘plastic’ phase linked to A-P patterning of multipotent axial progenitors/NMPs within their posterior niche and (ii) a later phase, which marks the instalment and fixing of lineage-specific Hox gene expression patterns and the sharpening of their final boundaries in the neural and mesodermal derivatives of axial progenitors as and after they exit the posterior growth zone (Ahn et al.,
We demonstrate that TBXT function is crucial for proper activation of HOX genes during NMP induction from hPSCs in vitro by channelling extrinsic WNT activity toward the establishment of a ‘posteriorising’/pro-mesodermal niche (in line with previous data; Martin and Kimelman, 2010) that is essential for the subsequent transduction of a posterior character to NMP-derived NC/spinal cord cells. This prospective TBXT-WNT-driven transmission of positional information via an early axial progenitor intermediate and the potential inductive interaction between nascent presomitic mesoderm and NC progenitors reinforces previous observations on the effect of non-organiser mesoderm and vertical signalling as determinants of HOX gene expression and axial identity in early neural cells (Bardine et al., 2014; Forlani et al., 2003; Grapin-Botton et al., 1997). However, further work is required to disentangle the cell vs non-cell autonomous effects of TBXT on the A-P patterning of NC cells.

Our results are in line with previous studies reporting that TBXT attenuation in hPSC-derived mesoderm progenitors and spinal cord organoids, as well as in Xenopus morphant and mouse Tc/2J/2J mutant embryos abolishes Hox gene expression (Faial et al., 2015; Libby et al., 2021; Lolas et al., 2014; Wacker et al., 2004). Moreover, they further expand the repertoire of actions that orchestrate embryonic axis elongation/axial progenitor ontogeny and are exerted via the regulatory axis centred on WNT signalling, TBXT, and the HOX clock (Denans et al., 2015; Mariani et al., 2021; Ye et al., 2021; Ye and Kimelman, 2020). Interestingly, global Hox gene expression appears minimally affected in some Brachury mutants (such as those with the T^{mu} genotype; Koch et al., 2017; Tosic et al., 2019), as well as in T^{mu}:: wild type chimeras (Guibentif et al., 2021) suggesting that the nature of the T gene mutation and non-cell autonomous rescue effects from the surrounding wild-type environment, similar to those described previously in the case of grafted Cdx mutant axial progenitors (Bialecka et al., 2010), are critical actors in influencing the effect of Brachury on its target genes. Moreover, the developmental arrest exhibited by T mutant embryos around the time of trunk NC emergence and the associated lack of posterior axial tissue precludes a detailed assessment of Hox gene expression in T null NC cells.

TBXT has been shown to regulate its downstream mesoderm/axis elongation-associated targets in mouse embryos by promoting chromatin remodelling and directing permissive chromatin modifications in key regulatory elements (Amin et al., 2016; Beisaw et al., 2018; Koch et al., 2017; Tosic et al., 2019). Our data confirm that this is also the case during the transition of hPSCs toward NMPs and reveal that TBXT additionally contributes to global control of HOX cluster transcription in a similar way and in line with previous reports showing direct TBXT binding in HOX loci during hPSC differentiation (Faial et al., 2015). We propose that concomitant activation of the HOX gene clusters and induction of WNT signalling components, via TBXT-driven chromatin landscape reconfiguration, comprise a critical early step in primary A-P regionalisation and the transition of pluripotent cells toward a caudal epiblast/axial progenitor state. This is supported by our demonstration that TBXT knockdown results in the acquisition of an anterior epiblast/AVE identity associated with WNT antagonism/head formation-promoting activity as well as increased chromatin accessibility in regulatory elements controlling pro-neural differentiation genes. Previous data on the crucial role of Brachury in countering the default neurectodermal differentiation of mouse pluripotent cells by altering chromatin accessibility in key enhancers also point to such a model (Tosic et al., 2019). Moreover, early A-P regionalisation driven by the TBXT-WNT-HOX axis is likely to involve the cooperative action of CDX factors and the participation of other key transcriptional factors such as EOMES and SOX2 given their previously reported functions in the mouse (Amin et al., 2016; Blassberg et al., 2022; Metzis et al., 2018; Neijts et al., 2017; Neijts et al., 2016; Tosic et al., 2019). Thus, an additional thorough dissection of the individual roles of these factors via loss/gain of function approaches is required to elucidate their crosstalk as they fine-tune the balance between pluripotency exit, cell fate decision-making, and adoption of a posterior character in human cells.

We also provide new evidence that TBXT influences the acquisition of a SOX10-positive NC identity early on during in vitro differentiation, a time window that coincides with the specification of NM-potent progenitors from hPSCs. This is likely to occur through two complementary routes: (i) via early direct TBXT binding and activation of DNA regulatory elements responsible for the induction of key signalling and transcriptional determinants of NC fate such as WNT1, BMP7, and CDX2...
mented with CHIR (3 μM, Tocris), FGF2 (20 ng/ml, R&D Systems), and Rho- associated coil kinase (ROCK) inhibitor Y-27632 2HCl (10 μM, Adooq Biosciences) with the latter being withdrawn from the differentiation medium after the first day of NMP induction. For TBXT inducible knockdown, NMP (ROCK) inhibitor Y-27632 2HCl (10 μM, Adooq Biosciences) with the latter being withdrawn from the differentiation medium after the first day of NMP induction. For TBXT inducible knockdown, NMP (ROCK) inhibitor Y-27632 2HCl (10 μM, Adooq Biosciences) with the latter being withdrawn from the differentiation medium after the first day of NMP induction. For TBXT inducible knockdown, NMP (ROCK) inhibitor Y-27632 2HCl (10 μM, Adooq Biosciences) with the latter being withdrawn from the differentiation medium after the first day of NMP induction. 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In summary, we provide mechanistic insight into the cellular and molecular basis of posterior axial identity acquisition during hPSC differentiation. Our data demonstrate a novel role for TBXT in controlling HOX gene expression and early posteriorisation supporting the idea that A-P patterning of at least some axial progenitor derivatives, such as the trunk NC, occurs prior to their specification, within their multipotent precursors. We speculate that the close links between TBXT-driven posterior axial identity programming in the NC and NMP ontogeny may explain some cases of spina bifida observed in individuals carrying mutations within the TBXT locus (Agopian et al., 2013; Carter et al., 2011; Fellous et al., 1982; Morrison et al., 1996; Shields et al., 2000), especially in light of the potential involvement of impaired NC specification and HOX gene dysregulation in neural tube defects (Anderson et al., 2016; Degenhardt et al., 2010; Poncet et al., 2020; Rochtus et al., 2015; Yu et al., 2019).

Materials and methods

Cell culture and differentiation

The use of hPSCs has been approved by the Human Embryonic Stem Cell UK Steering Committee (SCSC15-23). The TBXT and B2M shRNA sOPTiKD hESC lines (H9 background; Bertero et al., 2016) were employed for all TBXT loss-of-function and sequencing experiments whereas signalling agonist/antagonist treatments were performed in H9 hESCs and SFCi55-ZsGr human-induced PSCs (Lopez-Yrigoyen et al., 2018; Thomson et al., 1998). All cell lines were verified by confirming the expression of pluripotency-specific genes such as OCT4, NANOG, and TRA-1–60 using a combination of qPCR, immunofluorescence, and FACS together with human-specific primers/antibodies. Cultures were routinely monitored for bacterial and fungal contamination as well as the presence of mycoplasma. None of the cell lines is in the list of commonly misidentified cell lines maintained by the International Cell Line Authentication Committee. All cell lines were cultured routinely in feeder-free conditions in either Essential 8 (Thermo Fisher or made in-house) or mTeSR1 (Stem Cell Technologies) medium on laminin 521 (Biolamina) or Geltrex LDEV-Free reduced growth factor basement membrane matrix (Thermo Fisher). Cells were passaged twice a week after reaching approximately 80% confluency using PBS/EDTA as a passaging reagent. The TBTX inducible knockdown in the T8BTX shRNA sOPTiKD hESC line was achieved using Tet hydrochloride (Merck Life Science) at 1 μg/ml as described previously (Bertero et al., 2016). The hESCs were cultured in the presence/absence of Tet for 2 days prior to the initiation of differentiation and the Tet treatment was continued throughout the differentiation for the periods indicated in the results section/schemes.

For NMP differentiation, hPSCs (70–80% confluent) were dissociated using Accutase solution (Merck Life Science) and plated at a density of approximately 50,000 cells/cm² on vitronectin (Thermo Fisher)-coated culture plates in N2B27 basal medium containing 50:50 Dulbecco’s Modified Eagle’s Medium (DMEM) F12 (Merck Life Science) / Neurobasal medium (Gibco) and 1 x N2 supplement (Gibco), 1 x B27 (Gibco), 1 x GlutaMAX (Gibco), 1 x Minimum Essential Medium Non-Essential Amino Acids (MEM NEAA) (Gibco), 2-Mercaptoethanol (50 μM, Gibco). The N2B27 medium was supplemented with CHIR (3 μM, Tocris), FGF2 (20 ng/ml, R&D Systems), and Rho-associated coil kinase (ROCK) inhibitor Y-27632 2HCl (10 μM, Adooq Biosciences) with the latter being withdrawn from the differentiation medium after the first day of NMP induction. For TBXT inducible knockdown, NMP medium was supplemented with 1 μg/ml Tet hydrochloride and replenished every other day. For
culture under ‘neutral’ conditions, control and Tet-treated NMPs were dissociated into single cells using Accutase and replated at a density of 50,000 cells/cm² on Geltrex-coated culture plates in N2B27 basal medium (made as described above for NMP differentiation) in the presence/absence of 1 μg/ml Tet hydrochloride. No small molecules were included apart from ROCK inhibitor Y-27632 2HCl (10 μM), which was added for the first 2 days to facilitate plating and increased cell survival. The basal media was replenished every other day and the cells were fixed/harvested on day 7 of differentiation for downstream analysis. For early spinal cord progenitor differentiation, day 3 hPSC-derived NMPs were dissociated into single-cell suspension using Accutase and replated at a density of 37,500 cells/cm² on Geltrex-coated culture plates in N2B27 containing FGF2 (100 ng/ml), CHIR (3 μM), and ROCK inhibitor Y-27632 2HCl (10 μM; for the first day only) in the presence or absence of Tet hydrochloride (1 μg/ml). The medium was replaced every other day until day 7 of differentiation. For trunk NC differentiation, day 3 hPSC-derived NMPs were dissociated using Accutase and replated at a density of 50,000 cells/cm² on Geltrex-coated culture plates directly into NC inducing medium consisting of DMEM/F12, 1 × N2 supplement, 1 × GlutaMAX, 1 × MEM NEAA, the TGF-beta/Activin/Nodal inhibitor SB-431542 (2 μM, Tocris), CHIR (1 μM), BMP4 (15 ng/ml, Thermo Fisher), the BMP type-I receptor inhibitor DMH-1 (1 μM, Tocris), and ROCK inhibitor Y-27632 2HCl (10 μM). The medium was refreshed every two days without ROCK inhibitor and was supplemented with 1 μg/ml Tet hydrochloride throughout the differentiation for tet-mediated inducible TBXT knockdown. Trunk NC cells were analysed on day 8 of differentiation. The following signalling pathway inhibitors were employed: the WNT antagonist tankyrase inhibitor XAV (1 μM, Tocris), the MEK1/2 inhibitor PD03 (1 μM, Merck), and LDN (100 nM, Tocris).

Immunofluorescence and imaging

Cells were fixed in 4% Paraformaldehyde (PFA) for 10 min at room temperature, rinsed twice with PBS and permeabilised with 0.5% Triton X-100 in PBS containing 10% foetal calf serum (FCS) and 0.1% bovine serum albumin (BSA) for 15 min. Blocking was then performed in a blocking buffer consisting of 10% FCS and 0.1% BSA in PBS at room temperature for 1–2 hr or overnight at 4°C. Primary antibodies were diluted in the blocking buffer and cells were incubated with primary antibodies overnight at 4°C. Following three washes, cells were incubated with secondary antibodies conjugated to Alexa fluorophores (Invitrogen) diluted in blocking buffer for 1–2 hr at room temperature and in the dark. Cell nuclei were counterstained with Hoechst 33342 (Thermo Fisher, 1:1000) and fluorescent images were acquired using the InCell Analyser 2200 system (GE Healthcare). Images then were processed in Fiji (Schindelin et al., 2012) or Photoshop (Adobe) using identical brightness/contrast settings to allow comparison between different treatments. At all times, the positive/negative threshold was set using a sample incubated with secondary antibody only. The following primary antibodies were employed: anti-TBXT (Abcam, ab209665; 1:1000); anti-CDX2 (Abcam, ab76541; 1:200); anti-HOXC9 (Abcam, ab50839; 1:50); anti-SOX10 (Cell Signalling Technology, 89356; 1: 500); anti-SOX1 (R&D Systems, AF3369; 1:100); and anti-SOX2 (Abcam, ab92494; 1:400).

For quantification of TBXT protein expression in NMP-like cells, nine random fields per experiment were scored (two biological replicates) and processed in the image analysis software CellProfiler (Carpenter et al., 2006). Nuclei were first identified using Hoechst staining and subsequently mapped onto the other fluorescent channels for single-cell fluorescence intensity quantification. Cells stained with secondary antibody only were used as a negative control to set the negative/positive threshold. A histogram was created using GraphPad Prism (GraphPad Software). For quantification of SOX10 and HOXC9 protein expression in WNT/BMP-induced trunk NC, five random fields per experiment were scored (three biological replicates) and processed as described above. For quantification of SOX1 protein levels in control vs Tet-treated NMP-derived trunk NC cells, 28 random fields per experiment were scored (three biological replicates) and processed in CellProfiler. For quantification of HOX9/SOX10, HOXC9/SOX1, and SOX10/SOX1 in ‘neutral’ basal media and in trunk NC cultures following WNT-BMP agonists/antagonist combinations, 25 and 30 fields per experiment were scored respectively (three biological replicates) and processed in CellProfiler using a custom pipeline. Cells were classified as single/double positive based on a pair of measurements analysis for the two channels of interest. For quantification of SOX2 and HOXC9 protein expression in WNT-FGF-induced early spinal cord progenitors, 5–10 random fields per experiment were scored (three biological replicates) and processed as described above. Graphs
were generated using GraphPad Prism (GraphPad Software), which was also employed for statistical analysis.

**Flow cytometry**

For flow cytometric analysis of intracellular markers, a single cell suspension was prepared following Accutase treatment and cells were fixed in 4% PFA for 10 min at room temperature. Cells were permeabilised with 0.5% Triton X-100 in PBS containing 10% FCS and 0.1% BSA for 10 min and subsequently washed with a blocking buffer consisting of 10% FCS and 0.1% BSA in PBS. Primary antibodies diluted at the appropriate concentration in blocking buffer were added to cells followed by overnight incubation at 4°C on a shaking platform and washed off the following day. Secondary antibody diluted in blocking buffer was added for 1 hr in the dark and subsequently washed with blocking buffer. Cells were resuspended in 0.5 ml PBS for flow cytometric analysis. A separate negative control stained with secondary only and an unstained sample was used to set the gates. Flow cytometric analysis was performed using a BD LSR II flow cytometer and the data were processed using the software package FlowJo.

**Immunofluorescence analysis of wholemount embryos and 3D reconstruction**

E8.75 and E9.0 (TS13-14) R26-WntVis embryos were dissected in the M2 medium (Nowotschin et al., 2010). Embryos with yolk sac and amnion removed were fixed for 25–30 min at 4°C in 4% PFA in PBS, followed by permeabilisation in 0.5% Triton X-100 in PBS for 15 min, then incubated in 0.5 M glycine in PBS in 0.1% Triton X-100 (PBST) for 20 min, after which they were washed three times in PBST and blocked overnight at 4°C in 10% serum (Merck) in PBS/0.3% Triton X-100. Both primary and Alexa Fluor-conjugated secondary antibodies (Thermo Fisher Scientific, and Alexa Fluor 647 donkey-anti-chicken (#703-606-155) from Jackson Immunoresearch; all at 2 µg/ml final concentration) were diluted in blocking buffer and samples were incubated for 48 hr on a rocking platform at 4°C. A minimum of four 25 min washes were performed with PBST on a rocking platform at room temperature after the primary and secondary antibody incubation. Antibodies used (supplier, final concentration): anti-GFP (Abcam; ab13970; 10 µg/ml); anti-Brachyury (R&D; AF2085; 1 µg/ml); anti-Lamin B1 (Abcam; ab16048; 1:800–1:1000); anti-Sox9 (Merck, AB5535; 2 µg/ml); anti-Tfap2a (DSHB, B3, 3.1 µg/ml). Embryos were counterstained with Hoechst33342 (Thermo Fischer Scientific; 5 µg/ml). Confocal microscopy was performed after dehydration through an increasing methanol/PBS series (25, 50, and 75%, 10 min each) and two 5 min washes in 100% methanol and clearing in 1:1 v/v methanol/benzyl alcohol:benzyl benzoate(BA:BB; 2:1; Sigma), and two washes in 100% BA:BB. Embryos were imaged in BA:BB under an LSM800 confocal system with Airyscan and GaAsP detectors (Zeiss). Two to three replicate embryos were imaged per staining and stage. Wholemount immunostaining data were processed using Zeiss or Imaris software (Oxford Instruments) using channel alignment, background subtraction and deconvolution tools. In Imaris, 3D volumes were created from single channels. These volumes represent positively stained areas. As many cells in the tail bud exhibit some level of EGFP expression, only the fraction of a-GFP-stained cells with a higher intensity (a-GFP^high) were displayed in the analyses for clarity reasons.

**Quantitative real time PCR**

Total RNA was extracted using the total RNA purification kit (Norgen Biotek) following the manufacturer’s instructions. The cDNA preparation was completed using the High-Capacity cDNA Reverse Transcription kit (Thermo Fisher). Quantitative real-time PCR was carried out using the QuantStudio 12 K Flex (Applied Biosystems) thermocycler in combination with the Roche UPL system and the TaqMan Fast Universal PCR Master Mix (Applied Biosystems).

Primer sequences are shown in Supplementary file 10. Graphs were generated using GraphPad Prism (GraphPad Software), which was also employed for statistical analysis.

**Mouse husbandry**

All mice were maintained on a 12 hr-light/12 hr-dark cycle and housed at 18–23°C with 40–60% humidity. Homozygote R26-WntVis mice were obtained from the Laboratory for Animal Resources and Genetic Engineering at the RIKEN Center for Biosystems Dynamics Research, Kobe, Japan (accession...
no. CDB0303K; RBRC10162). Homozygote R26-WntVis males were crossed with ICR females (JAX stock #009122, The Jackson Laboratory); all experiments were performed on heterozygote embryos. For timed matings, noon on the day of finding a vaginal plug was designated E0.5. All animal experiments were approved by the Institutional Animal Experiments Committee of RIKEN Kobe Branch (A2016-03-10). Mice were handled in accordance with the ethics guidelines of the institute.

**RNA-seq**

Sample preparation

Total RNA was harvested from day 3 NMPs obtained from TBXT shRNA sOPTiKD hESC in the presence and absence of Tet (three biological replicates) using the total RNA purification plus kit (Norgen BioTek) according to the manufacturer’s instructions. Sample quality control, library preparation, and sequencing were carried out by Novogene (http://en.novogene.com). Library construction was carried out using the NEB Next Ultra RNA Library Prep Kit and sequencing was performed using the Illumina NoveSeq platform (PE150). Raw reads were processed through FastQC v0.11.2 and Trim Galore. Reads were aligned using STAR v2.4.2a (Dobin et al., 2013) to the human reference genome assembly GRCh38 (Ensembl Build 79) in the two-pass mode. The RSEM v1.3.0 (Li and Dewey, 2011) was used to extract expected gene counts, where genes expressed in <3 samples or with total counts ≤5 among all samples were excluded. We identified genes showing significant differential expression with DESeq2 (Love et al., 2014), with log2FoldChange>|0.5| and Benjamini–Hochberg-adjusted p<0.05. Data were deposited to GEO (Accession number: GSE184620).

**ChIP-seq**

Chromatin immunoprecipitation followed by next-generation sequencing (ChIP-seq) was performed on approximately 10 million cells/sample (three pooled replicates per sample), fixed with 1% formaldehyde solution (11% formaldehyde, 0.1 M NaCl, 1 mM EDTA [pH 8.0], 50 mM HEPES [pH 7.9]) for 15 min at room temperature on a shaking apparatus. Fixation was quenched with 125 mM of glycine (1/20 volume of 2.5 M stock) for 5 min and then adherent cells were scraped thoroughly from the culture surface. Cells were washed, centrifuged at 800 × g for 10 min at 4°C and pellets were resuspended in 10 ml chilled PBS-Igepal (1 × PBS [pH 7.4], 0.5% Igepal CA-630). This pellet wash was repeated and cells were resuspended in 10 ml chilled PBS-Igepal and 1 mM phenylmethylsulfonyl fluoride (PMSF). Samples were centrifuged at 800 × g for 10 min at 4°C for a third time, after which the supernatant was removed and pellets were snap-frozen on dry ice and stored at ~80°C. Samples were sent to Active Motif (Carlsbad, CA) for ChIP-seq. Active Motif (https://www.activemotif.com) prepared the chromatin, performed ChIP reactions, generated libraries, sequenced them, and performed basic data analysis. Chromatin was isolated by adding lysis buffer, followed by disruption with a Dounce homogenizer. Lysates were sonicated and the DNA sheared to an average length of 300–500 bp with Active Motif’s EpiShear probe sonicator (cat# 53051). Genomic DNA (Input) was prepared by treating aliquots of chromatin with RNase, proteinase K, and heat for de-crosslinking, followed by SPRI beads clean up (Beckman Coulter) and quantitation by Clariostar (BMG Labtech). Extrapolation to the original chromatin volume allowed the determination of the total chromatin yield. An aliquot of chromatin (50 µg) was precleared with protein G agarose beads (Invitrogen) and genomic DNA regions of interest were isolated using 4 µg of antibody against Brachyury (R&D Systems, cat# AF2085, lot# KQP0719121). Complexes were washed, eluted from the beads with sodium dodecyl sulfate (SDS) buffer, and subjected to RNase and proteinase K treatment. Crosslinks were reversed by incubation overnight at 65°C, and ChIP DNA was purified by phenol-chloroform extraction and ethanol precipitation. qPCR reactions were carried out in triplicate on specific genomic regions using SYBR Green Supermix (Bio-Rad). The resulting signals were normalized for primer efficiency by carrying out QPCR for each primer pair using Input DNA. Illumina sequencing libraries were prepared from the ChIP and Input DNAs by the standard consecutive enzymatic steps of end-polishing, dA-addition, and adaptor ligation. Steps were performed on an automated system (Apollo 342, Wafergen Biosystems/Takara). After a final PCR amplification step, the resulting DNA libraries were quantified and sequenced on Illumina’s NextSeq 500 (75 nt reads, single end). Reads were aligned to the human genome (hg38) using the BWA algorithm (default settings; Li and Durbin, 2009). Duplicate reads were removed and only uniquely mapped reads (mapping quality >25) were used for further analysis. Alignments were extended in silico at their 3’-ends to a length of 200 bp, which is the average genomic fragment length.
in the size-selected library, and assigned to 32-nt bins along the genome. The resulting histograms (genomic ‘signal maps’) were stored in bigWig files. Peak locations were determined using the MACS algorithm (v2.1.0) (Zhang et al., 2008) with a cutoff of p-value=1e–7. Peaks that were on the ENCODE blacklist of known false ChIP-Seq peaks were removed. Signal maps and peak locations were used as input data to Active Motifs proprietary analysis program, which creates Excel detailed information on sample comparison, peak metrics, peak locations, and gene annotations. Motif analysis was carried out using the Homer software (Heinz et al., 2010). Regions of 200 bp surrounding the summit of the top 2500 peaks (based on MACS2 p-values) were analysed. Data were deposited to GEO (Accession number: GSE184622).

**ATAC-seq**

Day 3 NMPs and day 8 trunk NC cells (50,000 cells) obtained from TBXT shRNA sOPTiKD hESC in the presence and absence of Tet (three biological replicates) were harvested and samples were prepared following the Illumina Tagment DNA Enzyme and Buffer Small kit (Illumina), 1% Digitonin (Promega), and EDTA-free protease inhibitor cocktail (Roche). Following DNA purification with the MinElute kit eluting in 12 μl, 1 μl of eluted DNA was used in a qPCR reaction to estimate the optimal number of amplification cycles. The remaining 11 μl of each library were amplified for the number of cycles corresponding to the Cq value (i.e. the cycle number at which fluorescence has increased above background levels) from the qPCR. Library amplification was followed by AMPure beads (Beckman Coulter) size selection to exclude fragments smaller than 150 bp and larger than 1200 bp. Library amplification was performed using custom Nextera primers. DNA concentration was measured with a Qubit fluorometer (Life Technologies) and library profile was checked with Bioanalyzer High Sensitivity assay (Agilent Technologies). Libraries were sequenced by the Biomedical Sequencing Facility at CeMM (Research Center for Molecular Medicine of the Austrian Academy of Sciences, Vienna) using the Illumina HiSeq 3000/4000 platform and the 50 bp single-end configuration. Base calling was performed by Illumina Real Time Analysis (v2.7.7) software and the base calls were converted to short reads using the IlluminaBasecallsToSam tool from the Picard toolkit (v2.19.2) (‘Picard Toolkit’ 2019. Broad Institute, GitHub Repository. http://broadinstitute.github.io/picard/; Broad Institute). Sequencing adapters were removed, and the low-quality reads were filtered using the fastp software (v 0.20.1) (Chen et al., 2018). Alignment of the short reads on GRCh38 was performed using Bowtie2 (v2.4.1) (Langmead and Salzberg, 2012) with the ‘-very-sensitive’ parameter. The PCR duplicates were marked using samblaster (v0.1.24) (Faust and Hall, 2014), and the reads mapped to the ENCODE black-listed (Amemiya et al., 2019) regions were discarded prior to peak calling. To detect the open chromatin regions, we performed peak calling using the MACS2 (v2.2.7.1) (Zhang et al., 2008) software with the ‘--nomodel’, ‘--keep-dup auto’, and ‘--extsize 147’ options. Peaks in the format of bed files were analysed for differential analysis to compare signals corresponding to the + vs –Tet samples using the GUAVA software. Differential peaks with a distance from TSS ranging from –1–1 kb, log2FC cutoff = 1 and p-value<0.05 were extracted. Finally, HOMER findMotifs (v4.11) (Heinz et al., 2010) was used for motif enrichment analysis over the detected open chromatin regions. Data were deposited to GEO (Accession number: GSE184227).

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**Author contributions**

Antigoni Gogolou, Celine Souilhol, Formal analysis, Validation, Investigation, Visualization, Methodology, Writing – review and editing; Ilaria Granata, Data curation, Formal analysis, Validation, Investigation, Visualization, Writing – review and editing; Filip J Wymeersch, Formal analysis, Funding acquisition, Investigation, Visualization, Writing – review and editing; Ichcha Manipur, Data curation, Formal analysis, Investigation, Visualization, Writing – review and editing; Matthew Wind, Formal analysis, Investigation, Writing – review and editing; Thomas JR Frith, Funding acquisition, Investigation, Writing – review and editing; Maria Guarini, Investigation; Alessandro Bertero, Resources, Writing – review and editing; Christoph Bock, Data curation, Methodology, Writing – review and editing; Florian Halbritter, Methodology, Writing – review and editing; Minoru Takasato, Mario R Guarracino, Funding acquisition, Writing – review and editing; Anestis Tsakiridis, Conceptualization, Supervision, Funding acquisition, Investigation, Visualization, Methodology, Writing – original draft, Project administration, Writing – review and editing

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**Ethics**

All animal experiments were approved by the Institutional Animal Experiments Committee of RIKEN Kobe Branch (A2016-03-10). Mice were handled in accordance with the ethics guidelines of the institute.

**Decision letter and Author response**

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**Additional files**

**Supplementary files**

- Supplementary file 1. Significantly up- and downregulated transcripts in Tet-treated, TBXT-depleted hESC-derived NMPs.
- Supplementary file 2. List of GO terms and corresponding gene lists enriched in Tet-treated, TBXT-depleted hESC-derived NMPs.
- Supplementary file 3. List of all genomic regions (Intervals) with peak p-value below the applied threshold bound by TBXT in hESC-derived NMPs and undifferentiated hESCs.
• Supplementary file 4. List of GO terms and corresponding gene lists associated with TBXT binding sites in hPSC-derived NMPs.

• Supplementary file 5. List of known HOMER database motifs enriched in TBXT binding sites in hESC-derived NMPs.

• Supplementary file 6. List of TBXT target genes which are differentially expressed following Tet treatment (padj<0.05 log2FC>0.5) and Gene Ontology Biological Processes enrichment analysis. Genes are listed in relation to the genomic position (in relation to TSS) of TBXT binding within their proximity. Blue highlight denotes downregulation while red represents upregulation in expression.

• Supplementary file 7. List of ATAC-seq peaks associated with gain or loss of chromatin accessibility following TBXT depletion in hESC-derived NMPs. Gene Ontology Biological Processes enrichment analysis, list of HOX genes as well as other genes (padj<0.05 log2FC>1) affected by TBXT depletion and are associated with changes in chromatin accessibility are also included.

• Supplementary file 8. List of transcription factor DNA binding motifs enriched in ATAC-seq sites associated with chromatin accessibility gain, chromatin accessibility loss or both.

• Supplementary file 9. ATAC-seq analysis of trunk NC cells generated from hPSCs via NMPs in the presence of Tet. Includes: (i) Summary: Summary of detected peak numbers; (ii) all_peaks: List of all the peaks with P-value <0.05 (gained open, gained close, no change) with the relative annotations; (iii) Accessibility Loss padj <0.05: Significant Gained close peaks (pvalue adjusted (padj) <0.05); (iv) HOX genes: Peaks in HOX gene regions, yellow colour highlights the significant peaks (padj <0.05); (v) Enrichment-Accessibility Loss: Enrichment of Significant Gained close peaks (pvalue adjusted <0.05); (vi) Accessibility gain padj <0.05: Significant Gained close peaks (pvalue adjusted (padj) <0.05).

• Supplementary file 10. List of primers used.

• Transparent reporting form

Data availability
Sequencing data have been deposited in GEO under accession codes GSE184622, GSE184620 and GSE184227.

The following datasets were generated:

| Author(s) | Year | Dataset title | Dataset URL | Database and Identifier |
|-----------|------|---------------|-------------|------------------------|
| Tsakiridis A | 2021 | Early anteroposterior regionalization of human neural crest is shaped by a pro-mesodermal factor | https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE184227 | NCBI Gene Expression Omnibus, GSE184227 |
| Granata I, Tsakiridis A | 2021 | Early anteroposterior regionalization of human neural crest is shaped by a pro-mesodermal factor | https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE184622 | NCBI Gene Expression Omnibus, GSE184622 |
| Manipur I, Granata I, Tsakiridis A | 2021 | RNA sequencing of control and TBXT-depleted human NMP-like axial progenitors | https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE184620 | NCBI Gene Expression Omnibus, GSE184620 |

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