Transformation of Morphine by Resting Cells and Cell-Free Systems of Arthrobacter Sp.

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Received for publication 30 January 1975

Morphine can be transformed into 14-hydroxymorphine and a related unidentified material by resting cells of an Arthrobacter species. Cell-free extracts containing the transforming enzyme(s) have been obtained. O₂, Fe²⁺, and reduced nicotinamide adenine dinucleotide stimulate the transformation.

Transformation of steroids by microorganisms has been a widely used procedure to produce commercial preparations of physiologically active steroids (4). Other complex natural products have also been examined as substrates in microbial transformation, for example, ergot (2), indoles (5), yohimbines (6), or opium alkaloids (8). Cell-free systems have been described which transform nicotine (7) and steroids (14, 15), and the hydroxylation of the main constituents of marijuana by rat liver extracts has been well studied (3). We have previously reported the hydroxylation of morphine and codeine by cultures of an Arthrobacter species (P. Liras et al., In L. A. Unterkofler (ed.), Developments in Industrial Microbiology, vol. 16, in press). Such transformation of morphine might result in new analgesics, possibly nonaddictive derivatives, and transforming enzymes are promising tools as reagents to detect trace amounts of morphine. The present work describes the favorable conditions for hydroxylation of morphine by cultures and cell-free extracts of this strain, and a partial purification of the involved enzyme has been achieved.

MATERIALS AND METHODS

Growth conditions. Cells were grown in 1-liter flasks containing 300 ml of 0.5% K₂HPO₄, 0.5% yeast extract, and 0.5% tryptone. Three milliliters of a 24-h culture were used as inoculum. The cells were grown at room temperature on a 2-in. (5.08-cm) throw rotary shaker (150 rpm) for 20 h, centrifuged at 5,000 × g for 10 min, washed twice with 10 mM phosphate buffer, pH 7.5, and suspended in the same buffer at a concentration of 10 mg of dry weight/ml. Cell suspensions were stored frozen at −20 °C without appreciable loss of activity for at least 3 months.

Preparation of cell-free extracts. The cells were suspended at 25 mg of dry weight/ml of tris(hydroxymethyl)aminomethane (Tris)-hydrochloride buffer (20 mM, pH 8.4) and were sonically treated for 2 min, in four intervals of 30 s each, in a sonifier cell disruptor, model W 185D (Heat Systems Ultrasonic, Inc., Plainview, Long Island, N.Y.). The cells were kept at 4 °C. The extracts were centrifuged twice at 5,000 × g for 10 min, and the pellets were discarded. The yellow supernatant was centrifuged at 18,000 × g for 40 min in a Sorvall RC-2 centrifuge, producing a gelatinous pellet (P₁). The remaining supernatant was centrifuged at 102,000 × g for 60 min in a Spinco Model L centrifuge, producing a pellet, P₂, and the final supernatant. Both pellets were suspended separately in the original volume of 20 mM Tris buffer, pH 8.4, containing 0.03% Triton X-100 and sonically treated for 10 s to produce a finely divided, almost transparent suspension. The processes were carried out at 4 °C, and the cell-free preparation was assayed immediately.

Assay of hydroxylating activity. Whole cells were suspended at 4 mg of dry weight/ml in 10 mM phosphate buffer, pH 7.5, containing 100 μg of morphine/ml, 20 mmol of FeSO₄, and 0.05 μCi [N-methyl-14C]morphine (500 μg of morphine/ml), in a total volume of 4 ml. The cell suspension was incubated with shaking at 30 °C.

In cell-free extracts, the reaction was started by adding the desired amount of enzymes to the 25-ml flasks containing 60 mmol of Tris-hydrochloride buffer, pH 8.4, 0.05 μCi [N-methyl-14C]morphine, 500 μg of morphine/ml, 15 mmol of FeSO₄, and 4.8 mmol of reduced nicotinamide adenine dinucleotide (NADH) in a final volume of 3 ml. The flasks were incubated at 30 °C with shaking up to 18 h, and the reaction was stopped by addition of 10% NH₄OH (vol/vol). To avoid contamination, the system was occasionally sterilized through membrane filters (Millipore Corp.) before incubation, although no difference in the results was found.

Extraction and determination of transformed products. After incubation the cultures or extracts were adjusted to pH 10 with ammonium hydroxide and extracted three times with 2 volumes of chloroform:ethanol (2:1 vol/vol) for 40 min each. The mixed extracts were dried in an air stream, and the residue was suspended in methanol and spotted on silica gel G plates (20 by 20 cm), 250 microns (Analtech, Inc.). The
plates were developed using the solvent system ethyl acetate:methanol:ammonium hydroxide (14:8 M) (86:10:4) and were visualized with \( \text{I}_4 \) vapors. Standard samples of 14-hydroxymorphine were always included on the plates. The plates were scraped off, the silica gel was placed into radioactivity vials, and the \( \text{I}_4 \) was allowed to evaporate before the scintillation cocktail was added. Toluene containing 4 g of 98% 2,5-diphenyloxazole/2% Bis MSB per liter (Omni-scint, ICN, Inc.) was used as scintillation cocktail. Radioactivity was measured in a Beckman LS-250 model liquid scintillation counter.

**Protein.** Protein was determined spectrophotometrically, measuring optical density at 260 and 280 nm (10).

**Chemicals.** [\( N \)-methyl-\( ^{14} \text{C} \)]morphine hydrochloride (143 Ci/mg) was purchased from Amersham/Searle (Arlington Heights, Ill.). Morphine and codeine were obtained from Merck and Co. (Rahway, N.J.). 14-Hydroxymorphine was a kind gift from Everett L. May of the National Institutes of Health (Bethesda, Md.).

### RESULTS

**Transformation products: factors affecting the kinetics of transformation.** Two main transformation products are formed when cells or cell-free extracts of *Arthrobacter* sp. 86 are incubated with morphine. Thin-layer and gas chromatography parameters of those substances have already been described and one of the products has been identified as 14-hydroxymorphine (P. Liras et al., *In L. A. Unzkofer* (ed.), in press). The structure of the second substance formed, product “C” has not yet been established. Both 14-hydroxymorphine and transformation product C are the main components formed from morphine by cells, and are formed by whole cells in about the same proportion under the different conditions described below. Minor proportions of other unknown products were detected, but they are not considered in this study. A nonspecific degradation of morphine also occurs, as observed by the quantitative difference between the total initial morphine, the morphine transformed, and the morphine remaining in the medium. This material appears as radioactivity in the aqueous phase after extraction, but thin-layer chromatography and gas chromatography showed that this radioactivity is not present as a single component.

The maximum yield of both transformation products is reached after 3 days of incubation of morphine with a cell suspension of 4 mg of dry weight/ml (Fig. 1). The time required for maximum production of both transformation products was inversely proportional to the concentration of cells in the preparation. The transforming process was greatly facilitated by increasing the dissolved oxygen in the cell preparation (Fig. 2). An increase in aeration from about 150 to 500 ml of \( O_2 \)h produced a substantial increase in 14-hydroxymorphine production, although no net increase in the cell mass occurred. Higher aeration—up to 6,000 ml of \( O_2 \)h—further increased the production but not proportionally to the increase in aeration. Cells under anaerobiosis did not transform or degrade morphine. Although partial cell death takes place under anaerobiosis, this is not the only explanation for the lack of transformation, since cell-free extracts of *Arthrobacter* sp. do not degrade morphine under anaerobiosis (Table 1).

The transformation does not appear to require the induction of specific enzymes. Cells which had been preincubated for 12 h with or without morphine, when transferred to fresh medium plus morphine, produced approximately the same yield of products at the same transformation rate. Young cells transformed morphine at a faster rate than older cells.

The effect of some reagents on the hydroxylation activity of the cells was studied.
Ethylenediaminetetraacetic acid at concentrations above 1 mM partially or completely inhibited the formation of 14-hydroxymorphine, 5 mM being completely inhibitory (Table 2). This suggested that metal cations may be involved in the hydroxylation. Consequently, the effect of different cations was tested. At 2 mM concentration, Zn** (data not shown) and Cu** almost completely inhibited the transformation of 14-hydroxymorphine and product C; Mo**, Mn** and Mg** had no effect (data not shown), and Fe** clearly increased the yield of products. Different iron concentrations produced an increase in the formation of 14-hydroxymorphine with a maximum effect at 5 mM, higher concentrations being inhibitory (Table 2). The formation of product C is not affected or is slightly inhibited by the presence of iron. Fe** produced a similar effect as Fe** although possibly both cations reach an equilibrium in the incubation media. In the presence of iron, the decrease of morphine in the medium was smaller than in its absence (Table 2), therefore producing a much higher specific transformation of the drug. This fact was also found in cell-free preparations. Glutathione at a concentration of 5 mM does not affect the formation of 14-hydroxymorphine, but increases the yield of product C at least three times.

**Hydroxylation of morphine by cell-free extracts of Arthrobacter sp. 86.** Cell-free extracts of Arthrobacter sp. hydroxylated morphine at about the same rate than whole cells.

Table 1 shows the effect of pH, iron, oxygen, and reduced cofactors on the hydroxylation of morphine. Both NADH and reduced nicotinamide adenine dinucleotide phosphate (NADPH) produced an increase in the hydrox-
fraction EDTA and FeSO₄(5) 16,500 Glutathione(5) pH free Additions Fe₂⁺ ions The considerable difference in the soluble fraction after centrifugation at 100,000 × g (Table 3). Addition of one or both pellets did not increase the activity. All the enzymatic activity precipitated between 35 and 60% saturation of ammonium sulfate. The activity for formation of 14-hydroxymorphine and product C appeared together in the same fraction (Table 4). However, the relative proportion of both substances differs widely from one enzyme preparation to another. Conditions

### Table 2. Effect of EDTA⁺, glutathione, Cu⁺⁺, and Fe⁺⁺ ions on the transformation of morphine by resting cells of Arthrobacter sp. 86

| Additions (mM) | Morphine in the medium at 72 h (counts/min per ml) | 14-Hydroxy-morphine (counts/min per ml) | Product C (counts/min per ml) |
|---------------|---------------------------------------------|----------------------------------------|-------------------------------|
| None          | 16,500                                      | 5,600                                  | 5,450                         |
| Glutathione (5) | 15,228                                      | 5,220                                  | 13,400                        |
| EDTA (4)      | 29,330                                      | 424                                    | 632                           |
| CuSO₄ (2)     | 4,608                                       | 560                                    | 320                           |
| FeSO₄ (1)     | 19,820                                      | 8,200                                  | 4,400                         |
| FeSO₄ (2)     | 20,860                                      | 9,130                                  | 4,450                         |
| FeSO₄ (5)     | 21,200                                      | 10,454                                 | 4,360                         |
| FeSO₄ (10)    | 27,113                                      | 6,680                                  | 4,120                         |

* EDTA, Ethylenediaminetetraacetic acid.
* Cells (4 mg/ml) in 10 mM phosphate buffer, pH 7.5, with 100 μg of morphine per ml (0.02 μCi/ml; 40,250 counts/min per ml), and the indicated additions were incubated for 72 h at 30°C.

ylation of morphine, although NADH was more effective. The specific decrease in absorption of NADH at 340 nm could not be detected in the presence of morphine, probably because several other enzymes in the cell-free preparation also oxidized the cofactor. The oxidized cofactors NAD⁺ and NADP⁺ did not affect the transforming activity in the cell-free extracts. Iron stimulated the hydroxylating activity of cell-free extracts as well as of whole cells.

The pH of the medium affected differentially the yield of 14-hydroxymorphine and product C by whole cells, resulting in a twofold increase of 14-hydroxymorphine when the pH was raised from 7 to 9 (data not shown; T. Atherholt, personal communication). Transformation by cell-free systems was even more affected by the pH than the transformation by the whole cells, in the previously indicated range of pH (Table 1, experiments no. 3 and 4).

A considerable difference in the amount of 14-hydroxymorphine and product C formed appeared in different cell-free preparations (Tables 3 and 4), although both were produced in the same proportion by treatment of morphine by whole cells (Fig. 1, Table 2).

The hydroxylating activity was completely recovered in the soluble fraction after centrifugation at 100,000 × g (Table 3). Addition of one or both pellets did not increase the activity. All the enzymatic activity precipitated between 35 and 60% saturation of ammonium sulfate. The activity for formation of 14-hydroxymorphine and product C appeared together in the same fraction (Table 4). However, the relative proportion of both substances differs widely from one enzyme preparation to another. Conditions
which positively affect the formation of 14-hydroxymorphine may also affect positively the formation of product C (as NADH and NADPH, or pH) or have a negative effect (as Fe²⁺ ions). These results, together with the fact that product C formation is highly variable, indicate two separate enzymatic activities.

**DISCUSSION**

Transformation of morphine by resting cells, cell-free extracts and ammonium sulfate fractionated extracts of *Arthrobacter* sp. 86 was studied. 14-Hydroxymorphine was identified as one of the transformation products (P. Liras et al., *In L. A. Unterkofler (ed.), in press*). The formation of 14-hydroxymorphine is a process requiring oxygen (Fig. 2; Table 1, experiment no. 6), which suggests that the oxygen of the hydroxyl group in 14-hydroxymorphine comes from the air, as has been shown in the formation of 14-hydrocodeineone from thebaine (1). The enzymatic hydroxylating activity is partially inhibited by ethylenediamine tetraacetic acid, Zn²⁺, and Cu²⁺ and stimulated by Fe²⁺, NADH, and NADPH (Table 1). Hydroxylation partially takes place in the absence of external NADH, presumably due to endogenous coenzyme present in the cell-free extracts.

Other biological systems involved in drug transformation, such as the mono-oxygenase system present in rat liver microsomes (3), also requires NADPH, NADH, and atmospheric oxygen. In microorganisms, mixed function oxygenases constitute a coupled complex of soluble enzymes (9, 12). In *Pseudomonas* sp., this system is formed by at least three protein components (9, 11, 12) and requires Fe²⁺, NADH, and molecular oxygen. The similarity in requirements for maximal activity in the morphine-transforming system suggests that enzymes similar to those mentioned above may be responsible for the transformation of the drug, although in our case they appear to be non-morphine-inducible enzymes.

It seems that the formation of product C is not catalyzed by the same enzyme or enzyme complex as the 14-hydroxylation of morphine, since the response of each activity to iron is different from the other. This is in agreement with the report that substance C appears as a product of transformation of 14-hydroxymorphine itself (P. Liras et al., *In L. A. Unterkofler (ed.), in press*). However, adequate amounts of pure, radioactive 14-hydroxymorphine are not yet available to make a more complete study of this transformation.

**ACKNOWLEDGMENTS**

This study was supported by U. S. Public Health Service grant PHS-DA 378-01 from the National Institutes of Health.

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