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Wnt-1 Modulates Cell-Cell Adhesion in Mammalian Cells by Stabilizing β-Catenin Binding to the Cell Adhesion Protein Cadherin

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Abstract. Wnt-1 homologs have been identified in invertebrates and vertebrates and play important roles in cellular differentiation and organization. In Drosophila, the products of the segment polarity genes wingless (the Wnt-1 homolog) and armadillo participate in a signal transduction pathway important for cellular boundary formation in embryonic development, but functional interactions between the proteins are unknown. We have examined Wnt-1 function in mammalian cells in which armadillo (β-catenin and plakoglobin) is known to bind to and regulate cadherin cell adhesion proteins. We show that Wnt-1 expression results in the accumulation of β-catenin and plakoglobin. In addition, binding of β-catenin to the cell adhesion protein, cadherin, is stabilized, resulting in a concomitant increase in the strength of calcium-dependent cell-cell adhesion. Thus, a consequence of the functional interaction between Wnt-1 and armadillo family members is the strengthening of cell-cell adhesion, which may lead to the specification of cellular boundaries.

Wnt-1 is a striking example of a protein involved in both normal development and oncogenic transformation (Nusse and Varmus, 1992). In mammalian cells, Wnt-1 is thought to function as a growth factor that is secreted and then becomes associated with the cell surface (Bradley and Brown, 1990; Papkoff and Schryver, 1990); in Drosophila, the Wnt-1 homolog, wingless, is thought to function as a locally restricted morphogen (van den Heuvel et al., 1989). Wingless plays an essential, but poorly defined role in a signal transduction pathway that determines segmental patterning in early embryogenesis (Morata and Lawrence, 1977). In vertebrates, regulated Wnt-1 expression is required for murine neural development (McMahon and Bradley, 1990; Thomas and Capecchi, 1990), and its overexpression in Xenopus results in anterior duplication of the embryonic axis (McMahon and Moon, 1989). However, Wnt-1 was originally identified in vertebrates as a proto-oncogene (Nusse and Varmus, 1982; Nusse et al., 1984). Unregulated expression of Wnt-1 was found to participate in the induction of mammary hyperplasias in mice (Tsukamoto et al., 1988), and morphological transformation of some cultured cell lines (Brown et al., 1986; Rijswijik et al., 1987). These apparently disparate effects of Wnt-1 expression have been difficult to reconcile in part because the function(s) of the protein is unknown.

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cell adhesion is regulated by the interactions of $\alpha$-catenin and armadillo family members with cadherin.

Given that members of the armadillo protein family play key roles in both the wingless signaling pathway in segmental patterning and in cadherin mediated cell adhesion, a fundamental question arises: does Wnt-1 function through $\beta$-catenin and plakoglobin to modulate cell-cell adhesion? In Drosophila, wingless expression leads to the accumulation of armadillo protein, but it is not known whether cell-cell interactions are altered. Here, we have examined the effects of Wnt-1 expression on armadillo-cadherin complex formation and the consequences on cell-cell adhesion in mammalian cells. We show that Wnt-1 expression increases the levels of armadillo family members in two different mammalian cell types and the consequence is increased strength of cell-cell adhesion. The mechanism through which Wnt-1 regulates cell-cell adhesion appears to be stabilization of the cadherin/ $\beta$-catenin complex at the cell surface. These effects are specific to armadillo family members as the expression and binding of $\alpha$-catenin to cadherin is unaffected by Wnt-1 expression. We suggest that one function of Wnt-1 is to regulate cell adhesion and, thereby, influence both cell boundary formation and cell proliferation.

Materials and Methods

Cell Lines and Antisera

Control and Wnt-1-expressing AIT20 (Papkoff and Schryver, 1990) and C57MG cell lines (Blasband et al., 1992) and the MDCK cell line (Nelson and Veshnock, 1986) have been described previously.

A pan-cadherin antibody that recognizes E-cadherin has previously been described (a gift from Dr. James Mann, Stanford University, Stanford, CA) (Marrs et al., 1993). An antibody directed against the cytoplasmic domain of murine N-cadherin was a gift from Dr. Dietmar Vestweber, Max-Planck-Institut für Immunobiologie, Freiburg, Germany. Antibodies directed against $\beta$-catenin, plakoglobin, and $\alpha$-catenin were raised in rabbits against peptides with the sequences, PGDNSQLAWFFTDLC, CIDTYS-DGLRPYPTADH, and KKHVNPVALSEFKAC, respectively, conjugated through the cysteine to KLH using the Imject kit (Pierce Chemical Co., Rockford, IL).

Immunoprecipitation and Immunoblots

For the immunoprecipitation and immunoblots (Figs. 1, A and B and 2 A), replicate cultures of cells were extracted with radioimmunoprecipitation assay (RIPA) buffer (Papkoff and Schryver, 1990) for 15 min at 4°C, scraped from the tissue culture dish, and centrifuged at 12,000 g for 15 min at 4°C. The total protein concentration of the clarified extracts was determined using BCA reagents (Pierce Chemical Co.). An equivalent amount of two-fold concentrated Lactam sample buffer was added to aliquots of each extract, which were normalized to total protein. For the E-cadherin immunoprecipitate (Fig. 1 A, E-cad IP), an MDCK cell extract was incubated for 2 h on ice with a pan-cadherin antibody that recognizes E-cadherin. The antibody was prepared for immunoprecipitation by preincubation with a 10% suspension of protein A-Sepharose 4B (Pharmacia Fine Chemicals, Gaithersburg, MD) for 1 h on ice followed by washing the complex once with PBS. The immunoprecipitate was washed as previously described (Wollner et al., 1992). The samples were boiled for 5 min in Laemmli sample buffer, separated in a SDS 7.5% polyacrylamide gel (Laemmli, 1970), and electrophoretically transferred (Towbin et al., 1979) to Immobilon-P filters (Millipore Corp., Bedford, MA). The molecular weight standards are $\beta$-galactosidase (M, 116,000), phosphorylase b (M, 97,000), and bovine serum albumin (M, 68,000). The filters were incubated in gelatin wash buffer (GWB) (Wollner et al., 1992) containing 5% nonfat milk for 2 h. Antibody incubation was followed by extensive washing in GWB, incubation with 125I-protein A (0.1 $\mu$Ci/ml) (Dupont New England Nuclear, Cambridge, MA) for 1 h, extensive washing in GWB and exposure to x-ray (XAR-5) film at ~80°C. For Figs. 1 A and B and 5, $\beta$-catenin, plakoglobin, and $\alpha$-catenin were specifically detected by incubating the filters with polyclonal antipeptide sera. Reactivity of each antibody with protein was completely blocked by incubating the filters with polyclonal antipeptide sera. Reactivity of each antibody with protein was completely blocked by preincubation with excess cognate peptide. For Fig. 2 A, N-cadherin was specifically detected by incubating the filter with an antibody generated against the cytoplasmic domain of murine N-cadherin. For the immunoprecipitation assay.

1. Abbreviations used in this paper: ECL, enhanced chemiluminescence; GWB, gelatin wash buffer; HGF, hepatocyte growth factor; RIPA, radioimmunoprecipitation assay.

![Figure 1](image-url)
Figure 2. (A) Wnt-1 expression results in an increase in the level of N-cadherin protein. Relative levels of N-cadherin and α-catenin in control (−Wnt-1) and Wnt-1-expressing (+Wnt-1) AtT20 cells were determined by immunoblotting equivalent amounts of total cell protein with antibodies directed against N-cadherin and α-catenin. (B) Complex formation between N-cadherin, β-catenin, and plakoglobin. Cell extracts from control (−Wnt-1) and Wnt-1-expressing (+Wnt-1) AtT20 cells were prepared with immunoprecipitated (IP Ab) with either N-cadherin (N-cad), β-catenin (β-cat), or plakoglobin (PG) antibodies. The immunoprecipitates were separated by SDS-PAGE, and immunoblotted (Blot Ab) with antibodies directed against β-catenin (A), plakoglobin (B), or N-cadherin (C).

Sequential immunoprecipitation/immunoblot (Fig. 2 B), replicate cultures of control (AtT20−) and Wnt-1-expressing (AtT20+) cells were extracted with RIPA buffer and protein equivalent aliquots of the extracts were immunoprecipitated with specific antibodies (IP Ab) as described in Fig. 1. The immunoprecipitates were separated in a SDS 7.5% polyacrylamide gel followed by immunoblotting with antibodies specific for β-catenin, plakoglobin and N-cadherin as described in Fig. 1. The immunoblot was exposed to x-ray (XAR-5) film at −80°C. The films were analyzed using a scanning densitometer (Molecular Devices Corp., Menlo Park, CA). For the steady state biotinylation, replicate cultures of AtT20− and AtT20+ cells were biotinylated on the cell surface with S-NHS-biotin as previously described (Wolffner et al., 1992). Cells were extracted with RIPA buffer and protein equivalent aliquots of each extract were immunoprecipitated, as described above, with the indicated antibodies. The immunoprecipitates were separated in a SDS 7.5% polyacrylamide gel followed by immunoblotting with antibodies specific for β-catenin or α-catenin as described in Fig. 1. To detect the biotinylated N-cadherin, the blot was blocked overnight in 5% ovalbumin, washed in Tris-buffered saline (10 mM Tris, pH 8.0, 150 mM NaCl) containing 0.1% Tween-20 and incubated with a complex of Avidin DH: biotinylated horseradish peroxidase H (Vectorstain ABC kit; Vector Laboratories, Inc., Burlingame, CA) as directed by the manufacturer. The blot was processed with enhanced chemiluminescence (ECL) detection system (Amersham Corp., Arlington Heights, IL).

**Metabolic Labeling and Immunoprecipitation**

For metabolic labeling (Fig. 1 D), replicate cultures of control (AtT20−) and Wnt-1-expressing (AtT20+) cells were incubated in the absence of methionine for 20 min, pulse-labeled in the presence of 150 μCi/ml [35S]methionine (Du Pont NEN) for either 5, 10, or 15 min and extracts prepared as described above. Protein equivalent aliquots of each extract were immunoprecipitated with antibodies and washed as described above. The immunoprecipitates were boiled for 5 min in Laemmli sample buffer and separated in a SDS 7.5% polyacrylamide gel (Laemmli, 1970). The gel was subsequently immersed in Amplify as directed by the manufacturer (Amersham Corp.), dried under vacuum, and exposed to x-ray (XAR-5) film at −80°C. The films were analyzed using a Molecular Devices scanning densitometer. The molecular weight standards are β-galactosidase (M, 116,000), phosphorylase b (M, 97,000) and bovine serum albumin (M, 68,000).

For the pulse-chase analysis (Fig. 3) replicate cultures of control (AtT20−) and Wnt-1-expressing (AtT20+) cells were incubated in the absence of methionine for 20 min, pulse-labeled with 150 μCi/ml [35S]methionine for 30 min, washed once, and incubated in medium containing excess unlabelled methionine for 1, 2, 4, 6, and 10 h. At the indicated times, cells were extracted with RIPA buffer and protein equivalent aliquots of each extract were immunoprecipitated with specific antibodies as described above. The immunoprecipitates were boiled for 5 min in Laemmli sample buffer and separated in a SDS 7.5% polyacrylamide gel (Laemmli, 1970) which was subsequently treated as described above. For analysis of cell surface N-cadherin, replicate cultures of control (AtT20−) and Wnt-1-expressing (AtT20+) cells were incubated in the absence of methionine for 20 min, pulse-labeled with [35S]methionine for 45 min, washed once, and incubated in medium containing excess unlabelled methionine for 1, 2, 4, 6, and 8 h. At the indicated times, a single culture of AtT20− and AtT20+ cells was biotinylated on the cell surface with NHS-S-S-biotin as previously described (Wolffner et al., 1992). Cells were extracted with RIPA buffer and protein equivalent aliquots of each extract were immunoprecipitated with specific antibodies as described above. The immunoprecipitates were washed and the immunocomplex dissociated in 0.1 M glycine, pH 2.5, as previously described (Wolffner et al., 1992). Biotinylated proteins were precipitated from neutralized supernatant by the addition of avidin-agarose (Pierce Chemical Co.) for 1 h. The immunoprecipitates were washed as above, the pellets were boiled for 5 min in Laemmli sample buffer and analyzed in a
**Results**

**Specific Antibodies Distinguish Armadillo Protein Family Members in Mammalian Cells**

To distinguish closely related members of the *armadillo* family of proteins, specific antipeptide antibodies were generated against unique COOH-terminal amino acid sequences of β-catenin and plakoglobin. Immunoblots of extracts from C57MG and AtT20 cell lines demonstrated that proteins with distinct electrophoretic mobilities were recognized by these antibodies (Fig. 1 A). The apparent molecular weights of these proteins were similar to those of β-catenin (∼94 kD) and plakoglobin (∼83 kD), which were recognized by the antipeptide antibodies in both immunoblots of MDCK cell extracts (Fig. 1 A) and in complexes coimmunoprecipitated with E-cadherin antibodies from MDCK cells (Fig. 1 A: E-cad IP); this E-cadherin/catenin complex has been previously well-characterized (Ozawa et al., 1989; McCrea and Gumbiner, 1991). Antibody specificity was demonstrated by blocking reactivity with excess cognate peptide (Fig. 1 A; block).

**Wnt-1 Regulates the Accumulation of β-catenin and Plakoglobin Proteins in Mammalian Cells**

The effects of Wnt-1 expression on β-catenin and plakoglobin protein levels were examined in C57MG and AtT20 cells transfected with Wnt-1 cDNA. The C57MG murine, mammary epithelial cell line is routinely used to analyze the protein levels were examined in C57MG and AtT20 cells. The effects of Wnt-1 expression on β-catenin and plakoglobin, which were recognized by the antibodies (Fig. 2 A). Note that there is little or no difference in the amount of α-catenin between the Wnt-1 expressing and control AtT20 cells. We were unable to identify the cadherin expressed in C57MG cells with antibodies due either to differences in antigenicity, or to low levels of cadherin expression. However, functional studies of C57MG+ cells, described below, revealed the presence in these cells of calcium-dependent cell-cell adhesion that has characteristics expected of a cadherin-mediated mechanism.
β-catenin and Plakoglobin Form Mutually Exclusive Complexes with Cadherin in Mammalian Cells

Complexes between N-cadherin, β-catenin, and plakoglobin were examined by high stringency immunoprecipitation of AtT20− and AtT20+ cell extracts with either N-cadherin (N-cad)-, β-catenin (β-cat)-, or plakoglobin (PG)-specific antibodies, followed by SDS-PAGE analysis and immunoblotting with one of the other antibodies (Fig. 2 B). Complexes comprising N-cadherin and both β-catenin (Fig. 2 B, A and B) and plakoglobin (Fig. 2 B, B and C) were detected. It appears that N-cadherin, β-catenin, and plakoglobin interact with high affinity because the complexes are stable in buffers containing 1 M NaCl and 0.1% SDS. Note that in both AtT20− and AtT20+ cells, plakoglobin was not detected in the β-catenin immunoprecipitate (Fig. 2 B, B), and vice versa (Fig. 2 B, A), demonstrating that β-catenin and plakoglobin form mutually exclusive complexes with N-cadherin. We do not know the functional significance of these different complexes, but it raises the possibility that cadherin function is regulated by binding either β-catenin or plakoglobin (see below).

Examination of complexes formed between N-cadherin and β-catenin or plakoglobin confirmed the increased steady state levels of N-cadherin (Fig. 2 B, C) and β-catenin (Fig. 2 B, A) in AtT20+ cells. Note that fourfold more β-catenin was coimmunoprecipitated in a complex with N-cadherin from AtT20+ cells than from AtT20− cells, (Fig. 2 B, A and C; see also Fig. 3, A and B). This indicates that a consequence of Wnt-1-induced accumulation of β-catenin and N-cadherin in the AtT20+ cells is a corresponding increase in the absolute amount of N-cadherin/β-catenin complexes in the cell. However, similar amounts of plakoglobin were coimmunoprecipitated in a complex with N-cadherin from AtT20+ and from AtT20− cells (Fig. 2 B, B). This difference in loading capacity of cadherins for β-catenin and plakoglobin may reflect differences in binding affinities. In MDCK cells, for example, only a small fraction of total plakoglobin is detected in a cadherin-associated complex (Kapprell et al., 1987) (unpublished results), whereas most of the β-catenin is bound to a cadherin complex (unpublished results).

Wnt-1 Expression Results in the Stabilization of Complex Formation Between β-catenin and N-cadherin

The effect of Wnt-1 expression on the formation and stability of complexes of N-cadherin and β-catenin or plakoglobin was examined by pulse-chase analyses. Replicate cell cultures were pulse-labeled with [35S]methionine for 30 min, and then chased in the absence of label for various times. Cells were harvested and aliquots of the extracts immunoprecipitated with antibodies directed against N-cadherin, β-catenin, plakoglobin, or α-catenin. Fig. 3 A shows that the level of synthesis and rate of turnover of newly synthesized N-cadherin was not affected by Wnt-1 expression.

In contrast, there were significant Wnt-1-dependent differences in the stability of the pool of newly synthesized β-catenin complexed with newly synthesized N-cadherin; this complex was immunoprecipitated with antibodies specific for either N-cadherin or β-catenin (Fig. 3, A and B). At the end of the pulse label (time 0), immunoprecipitation with N-cadherin antibodies revealed that the ratio of N-cadherin to β-catenin in the complexes was similar in both AtT20− (1:1.3) and AtT20+ (1:1.1) cells (Fig. 3 A). However, within 1 h of the chase period in AtT20− cells, 60% of the newly synthesized β-catenin was lost from the N-cadherin/catenin complex; the remaining 40% decayed more slowly (Fig. 3 A). This biphasic turnover of β-catenin in N-cadherin complexes was confirmed by direct immunoprecipitation of β-catenin (Fig. 3 B). In contrast to these results, a distinctly different profile of β-catenin turnover from the N-cadherin complex was found in AtT20+ cells. The ratio of N-cadherin/β-catenin in the complexes was approximately constant over the time course of the experiment, and both N-cadherin and β-catenin were degraded with similar kinetics (Fig. 3 A). Therefore, in AtT20+ cells, N-cadherin and β-catenin assemble and turn over coordinately as a protein complex similar to the degradation profile of these proteins in other cells (McCrea and Gumbiner, 1991; Shore and Nelson, 1991; Ozawa and Kemler, 1992). Note that in AtT20+ cells the ratio of N-cadherin/β-catenin in the complexes was different depending on the immunoprecipitating antibody (Fig. 3, A and B). More β-catenin was immunoprecipitated with the β-catenin antibody (Fig. 3 B), than was coimmunoprecipitated with the N-cadherin antibody (Fig. 3 A), although both antibodies were used in excess of protein. These results reveal a N-cadherin-independent pool of β-catenin.

These data demonstrate that Wnt-1 expression in AtT20+ cells results in the stabilization of β-catenin in the N-cadherin complex. It is possible that Wnt-1 expression results in a posttranslational modification of β-catenin, increasing its avidity for N-cadherin and resulting in a more stable association. Alternatively, β-catenin may exchange between an N-cadherin assembled and unassembled pool of β-catenin; in AtT20− cells, the free pool of β-catenin is very small, whereas in AtT20+ cells it is relatively large (Fig. 3 B).

Newly synthesized N-cadherin did not appear to coimmunoprecipitate with newly synthesized plakoglobin (Fig. 3, A and C). That this complex forms at steady state, however, was established in the sequential immunoprecipitation/immunoblots shown in Fig. 2 B, B and C. The absence of a detectable complex after a pulse label may reflect noncoordinated assembly or the weak association of N-cadherin and plakoglobin, supporting the idea that differences in the N-cadherin complexes between β-catenin and plakoglobin may be functionally significant (see above). Nonetheless, the results confirmed the Wnt-1-dependent increase in plakoglobin accumulation. In both the AtT20− and AtT20+ cells, plakoglobin decayed exponentially.

Changes were not detected in the synthesis and turnover of α-catenin as a consequence of Wnt-1 expression (Fig. 3 D). Although the association of α-catenin with cadherin was disrupted under the conditions used to extract the complexes shown here (Fig. 3 D) (McCrea and Gumbiner, 1991), other extraction conditions (Wollner et al., 1992; Piepenhagen and Nelson, 1993) revealed complexes of these proteins in both AtT20− and AtT20+ cells (data not shown). That there were no changes in α-catenin synthesis and turnover, or in the synthesis of total cellular protein (data not shown), indicates that the Wnt-1 effects on β-catenin synthesis and stabilization in cadherin complexes do not reflect a Wnt-1-dependent, global alteration of protein synthesis and stability.
Wnt-1 Expression Results in Increased Stability of Cell Surface N-cadherin

We sought to investigate the effects of increased loading and stability of β-catenin/cadherin complexes in Wnt-1-expressing cells on cell surface expression and function of N-cadherin. The time course of cell surface expression of N-cadherin was determined by pulse-labeling proteins with [35S]methionine, followed by a chase period in the absence of labeled methionine, and detection of newly synthesized protein at the plasma membrane by cell surface biotinylation (Fig. 4). The results showed a short term increase in stability of the population of N-cadherin expressed on the surface of AtT20+ cells (Fig. 4), which could be sufficient to account for the increased steady state levels of N-cadherin (Fig. 2 A). The increased stability of the cell surface pool of cadherin is not reflected in an increased stabilization of the total population of newly synthesized cadherin (Fig. 3 A). This may reflect that only a fraction of total cadherin is stabilized. Nevertheless, our results show that the increased stability of the cell surface population and the increased steady state accumulation of N-cadherin correlates with the accumulation of β-catenin and the stabilized assembly of β-catenin in the N-cadherin complex.

Under the experimental procedures used to isolate the biotinylated pool of N-cadherin, β-catenin was dissociated from the N-cadherin complex (Fig. 4). Therefore, we performed this experiment in a different way to confirm that the additional N-cadherin molecules on the cell surface of Wnt-1-expressing cells were complexed with β-catenin. To detect these complexes, proteins on the cell surface were biotinylated, cells were extracted as before, and protein complexes were immunoprecipitated with either β-catenin or α-catenin antibodies. After SDS-PAGE, proteins were blotted with either β-catenin or α-catenin antibodies, or with HRP-avidin (Fig. 5). A dramatic increase in β-catenin levels was detected in AtT20+ cells compared to control cells as expected (see Fig. 1 B). In addition, a corresponding increase in cell surface biotinylated cadherin was immunoprecipitated in a complex with β-catenin from the Wnt-1-expressing cells. Note that there is little difference in the amount of α-catenin between the Wnt-1-expressing and control cells. These results confirm that there is increased β-catenin and cadherin in Wnt-1-expressing cells and, significantly, that the increased cadherin at the cell surface is complexed with β-catenin.

Wnt-1 Expression Correlates with Increased Stability of the Cell Surface N-cadherin

We sought to determine directly whether increased N-cadherin stability and N-cadherin/β-catenin complex formation affected cell-cell adhesion. For these studies, we used a previously characterized assay for cell adhesion in which cell aggregates, formed in suspension culture, were sheared by vigorous trituration (Schmidt et al., 1992). This assay was chosen because it provides a measure of the strength of cell-cell adhesion. Since both Wnt-1+ and Wnt-1− AtT20 cell lines express cell surface cadherins, we expected that both cell lines would adhere; however, we considered that the increased stability of the catenin/cadherin complex in Wnt-1+ cells might result in an increase in the strength of the interaction between cells.

Both AtT20− and AtT20+ cells formed equally large cell aggregates (>200 cells/aggregate) within 2 h (Fig. 6 A) as expected since both cell lines express cell surface N-cadherin. However, trituration of the aggregates resulted in the dissociation of ~90% of control aggregates (−Wnt-1) into groups of <15 cells (Fig. 6, A and B). In contrast, ~45% of Wnt-1-expressing cells (AtT20+) survived this treatment with larger aggregates intact (>15 cells/aggregate) (Fig. 6, A and B). Analysis of the AtT20+ cell aggregates at higher magnification revealed a greater degree of compaction compared to the AtT20− cells. These results demonstrate an increased adhesion of Wnt-1-expressing AtT20 cells. We also performed the trituration assay on Wnt-1-expressing and control cells suspended in calcium-free growth media (Fig. 6 C). Aggregates formed in calcium-free growth media, indicating the presence of calcium-independent cell-cell adhesion mech-

![Figure 4](image-url) **Figure 4.** Wnt-1 expression results in increased stability of N-cadherin at the surface of AtT20 cells. The kinetics of synthesis and turnover of the cell surface population of N-cadherin were determined by cell surface biotinylation of newly synthesized protein in control (−Wnt-1) and Wnt-1-expressing (+Wnt-1) AtT20 cells. Representative cultures of cells were pulse-labeled (0 h chase) with [35S]methionine and chased in the absence of label for the indicated times (1, 2, 4, 8 h chase). At each time point, cell surface proteins were biotinylated and cell extracts prepared. Protein equivalent aliquots were immunoprecipitated with a pan-cadherin antibody and, following dissociation of the immune complex, the biotinylated population of N-cadherin was isolated by precipitation with avidin-agarose. As a control, the immunoprecipitation was specifically blocked by preincubating the antiserum with the cognate fusion protein (B/0).

![Figure 5](image-url) **Figure 5.** Wnt-1 expression results in increased steady state levels of N-cadherin/β-catenin complexes at the surface of AtT20 cells. The steady state protein levels of cell surface N-cadherin complexed with β-catenin were determined by biotinylation of cell surface proteins in control (−Wnt-1) and Wnt-1-expressing (+Wnt-1) AtT20 cells. Cell extracts were prepared and protein equivalent aliquots were immunoprecipitated with an antibody directed against β-catenin or α-catenin. The immunoprecipitates were separated by SDS-PAGE and biotinylated N-cadherin, immunoprecipitated in association with β-catenin, was detected by blotting with HRP-avidin. The levels of β-catenin and α-catenin were determined by immunoblotting with antibodies directed against β-catenin and α-catenin.

![Table](image-url) **Table 1.**
anisms. However, trituration of the Wnt-1-expressing and control cell aggregates formed in calcium-free media resulted in their complete dissociation. This suggests that calcium is required for strong cell-cell adhesion in both cell lines, and for the resistance to trituration exhibited by the Wnt-1-expressing cell line.

**Wnt-1 Expression Increases Calcium-dependent Cell–Cell Adhesion in C57MG Cells**

Previous studies in C57MG mammary epithelial cells have shown that ectopic expression of Wnt-1 causes morphological transformation (Brown et al., 1986; Blasband et al., 1992; Mason et al., 1992), although the cells did not show a change in growth rate, significant anchorage independence, nor increased tumorigenicity in syngeneic or athymic mice (Brown et al., 1986). Here, we examined the effects of ectopic Wnt-1 expression on the degree of cell-cell adhesion in the same clone of cells used in those previous studies (Fig. 7 A and B). In the presence of extracellular calcium, the Wnt-1-expressing and control cells formed large cell aggregates. Trituration of the Wnt-1-expressing cells demonstrated that these cells were resistant to disruption as compared to control cells (Fig. 7 A and B). We quantified the representative micrographs shown in Fig. 7 A by counting the number of cells in an aggregate (Fig 7 B). After trituration, 40% of the Wnt-1-expressing cells remained in aggregates larger than 11 cells compared to 8% of the control cells. Aggregates formed by the control C57MG cells were easily disrupted (Fig. 7 A and B); after trituration 91% of the control cell aggregates were dissociated to aggregates of <10 cells. When cells were suspended in calcium-free growth medium, both C57MG cells expressing Wnt-1 and control cells exhibited little or no propensity to form aggregates (Fig. 7 A, w/o Calcium). This suggests that in these cell lines, a calcium-dependent cell adhesion mechanism is the primary mechanism responsible for cell-cell adhesion and the strength of this adhesion is enhanced by Wnt-1 expression.

**Discussion**

**Wnt-1 and Formation of the Catenin/Cadherin Complex**

The results of this study indicate a causative role of Wnt-1 expression in regulating cell-cell adhesion in mammalian cells. Strikingly, regulation of cell adhesion correlates with Wnt-1-dependent accumulation of β-catenin and plakoglobin and, specifically, increased loading of β-catenin onto cadherin complexes. Accumulation of armadillo protein family members may be controlled at a posttranscriptional level in these mammalian cells. Preliminary analysis of β-catenin and plakoglobin mRNA levels indicate no change in Wnt-1-expressing compared to control AtT20 cells. Significantly, previous studies in Drosophila also demonstrated that wingless expression regulates the accumulation of armadillo protein by a post-transcriptional mechanism during embryogenesis (Riggelman et al., 1990). Those studies in Drosophila, however, were not able to shed light on the consequences of posttranslational accumulation of armadillo protein family members that are known to interact with cadherin cell adhesion proteins in vertebrates. In the present study, we show that the accumulation of β-catenin in mammalian cells correlates directly with a stabilization of the β-catenin/cadherin complex at the cell surface and increased strength of calcium-dependent cell-cell adhesion.

We observed increases in the steady state levels of β-catenin and plakoglobin, and increased stability of β-catenin with cadherin, but we did not detect changes in the steady state level of α-catenin or binding of α-catenin to cadherin as a consequence of Wnt-1 expression. This result indicates that α-catenin is not part of the same pathway involving Wnt-1 regulation of β-catenin levels and, hence, cadherin function in mammalian cells. In this context, it is interesting to note that the distribution of Drosophila α-catenin is rather homogeneous compared to the discrete segmental patterns of wingless and armadillo proteins (Oda et al., 1993). This indicates that the regulatory mechanisms involved in determining the distributions of α-catenin and armadillo protein in Drosophila are also different.

In mammalian cells, the interaction between catenins and cadherins is known to be critical for the formation and strength of calcium-dependent cell–cell adhesion (Takeichi, 1990; Kemler, 1992). For example, cells that express cadherin and β-catenin, but lack α-catenin, do not form cell aggregates until α-catenin is expressed by cDNA transfection (Hirano et al., 1992). In addition, posttranslational modulation of the catenin/cadherin complex by phosphorylation of β-catenin and cadherin by v-src tyrosine kinase coincides with a decrease in cell adhesion (Matsuyoshi et al., 1992; Behrens et al., 1993). Here, we have shown that strengthening of cell–cell adhesion occurs after stabilized loading of β-catenin onto cadherin/α-catenin complexes as a consequence of Wnt-1 expression. Thus, our results provide the first evidence that binding of β-catenin to cadherin promotes cell–cell adhesion, and thus β-catenin, specifically, plays a direct role in modulating cadherin function.

Our analysis of the assembly of the cadherin/catenin complex in AtT20 cells has uncovered an unusual degree of dynamics and plasticity in the association of β-catenin and cadherin. In the absence of Wnt-1 expression, β-catenin initially bound to, but then dissociated from cadherin; in the presence of Wnt-1, the β-catenin/cadherin complex was stabilized. These dynamics are different from the prevailing view of cadherin/catenin complex assembly in MDCK cells, fibroblasts and A6 cells in which α- and β-catenin form a stable complex with cadherin (Ozawa and Kemler, 1992; Knudsen and Wheelock, 1993; Peifer et al., 1993; Piepenhagen and Nelson, 1993). However, the recent availability of catenin-specific antibodies and the development of cross-linking methods to analyze the Triton X-100 insoluble complex has revealed a remarkable plasticity in the pathway of complex formation in MDCK cells, and uncovered temporal and spatial regulation of assembly, disassembly, and exchange of α/β-catenin and cadherin complexes (Hinck, L., I. S. Nätke, J. Papkoff, and W. J. Nelson, manuscript submitted for publication; Nätke, I. S., L. Hinck, J. Swedlow, J. Papkoff, and W. J. Nelson, manuscript submitted for publication). These new insights into cadherin/catenin complex assembly indicate that there are multiple steps in the assembly and exchange of proteins in the complex each of which are potential targets for regulation. Our present studies in AtT20 cells, therefore, show that Wnt-1 expression regulates
one aspect of this dynamic assembly pathway at the level of stabilizing $\beta$-catenin loading on cadherin.

**Wnt-1 and Regulation of Cell–Cell Adhesion**

Since both control and Wnt-1-expressing cell lines have cadherin on the cell surface, we reasoned that both cell types would adhere, but that the changes in the cadherin/catenin complex of Wnt-1-expressing cells might modulate the strength of adhesion. A stringent test of the strength of cell–cell adhesion is vigorous trituration of cells following their aggregation in suspension culture (Schmidt et al., 1992). Initial aggregation of AtT20 cells expressing Wnt-1 and control cells was not dependent on the presence of extracellular calcium, indicating that these cells expressed a cell adhesion mechanism in addition to cadherin. It is important to note, however, that in the absence of extracellular calcium, trituration of either AtT20− or AtT20+ cell aggregates resulted in rapid disruption of the aggregates and the generation of a population of single cells. In contrast, when cells were allowed to

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**Figure 6.** Wnt-1 expression increases the adhesiveness of AtT20 cells. (A) Micrographs showing the morphology of control (−Wnt-1) and Wnt-1-expressing (+Wnt-1) AtT20 cells before and after trituration following aggregation in suspension culture. (B) Histogram representing the data collected by counting the control (−Wnt-1) and Wnt-1-expressing (+Wnt-1) AtT20 cell aggregates that remain following trituration. Three separate experiments, representing 18 micrographs, were quantified by counting the number of aggregates containing less than 15 cells (% < 15 cells/aggregate: −Wnt-1 = 89%, SD = 7; +Wnt-1 = 54%, SD = 9) and more than 15 cells (% > 15 cells/aggregate: −Wnt-1 = 11%, SD = 7; +Wnt-1 = 46%, SD = 9). (C) Micrograph showing the morphology of control (−Wnt-1) and Wnt-1-expressing (+Wnt-1) AtT20 cells before and after trituration following aggregation in calcium free suspension culture (w/o Calcium).
aggregate in the presence of a normal concentration of calcium in the growth medium (1.8 mM), the cell aggregates formed from Wnt-1-expressing cells were dramatically more resistant to disruption by trituration than control cells. Together, these results indicate that calcium-independent adhesion in these cells does not contribute significantly to the increased strength of adhesion. These results show a direct correlation between increases in steady state levels of N-cadherin, stabilized binding of β-catenin to the cadherin complex, and strength of calcium-dependent cell-cell adhesion in Wnt-1-expressing AtT20 cells.

Unlike AtT20 cells, initial cell-cell adhesion of C57MG cells was dependent on the presence of extracellular calcium. Both the Wnt-1-expressing and control cells formed large cell aggregates in the presence of extracellular calcium. Similar to the results with the AtT20 cells, the Wnt-1-expressing C57MG cells survived trituration with larger aggregates intact. This indicates that the strength of cell-cell adhesion in this cell line also requires a calcium dependent cell-cell adhesion mechanism.

It is interesting to note that while we found Wnt-1 effects in C57MG and AtT20 cells, we have not detected significant Wnt-1-dependent modulation of cadherin/β-catenin complexes or increased cell adhesion in MDCK cells (unpublished results). In MDCK cells, which adhere rapidly and strongly in the presence of calcium (Gumbiner and Simons, 1986), the Wnt-1 signal may already be complemented by another factor, perhaps another Wnt family member. Based on these results, we hypothesize that Wnt-1 acts as a signal for cell adhesion in cells expressing a receptive cell adhesion mechanism that can be modulated by an increase in β-catenin and plakoglobin protein levels.

Wnt-1 and Control of Cell Proliferation and Formation of Cell Boundaries

The data presented in this paper demonstrate, in two independent cell types, that one consequence of Wnt-1 expression is increased strength of cell-cell adhesion. This new phenotype of Wnt-1 may be construed to be at odds with previous
Figure 7. Wnt-1 expression increases the adhesiveness of C57MG cells. (A) Micrographs showing the morphology of control (-Wnt-1) and Wnt-1-expressing (+Wnt-1) C57MG cells after trituration following aggregation in suspension culture containing 1.8 mM calcium (Calcium), and after aggregation in calcium-free media (w/o Calcium). (B) Histogram representing the data collected by counting the control (-Wnt-1) and Wnt-1-expressing (+Wnt-1) C57MG cell aggregates that remain following trituration. Representative micrographs were selected (shown in A) from three separate experiments and the number of single cells and aggregates containing 2–10, 11–20, 21–30, 31–50, and 50+ cells were counted.
ideas about the function of Wnt-1 as a regulator of cell proliferation and differentiation. However, reanalysis of previous studies indicates that the effects of Wnt-1 on cell proliferation and cell–cell adhesion are not mutually exclusive. We will briefly discuss those studies in the context of the new Wnt-1 phenotype.

The C57MG mammary epithelial cell line has been used extensively as a model system to analyze Wnt-1 effects in vitro. Initial studies showed that Wnt-1 expression in these cells caused partial transformation (Brown et al., 1986). Unlike the control cell lines, Wnt-1-expressing C57MG cells continue to divide beyond confluence. They exhibit a phenotypic alteration characterized by elongated cell morphology and multilayered growth. Of note is the observation that there is no change in the growth rate of Wnt-1-expressing cells in subconfluent culture compared to control cells. Also, by established criteria, these cells are not fully transformed; they do not grow in soft agar or form tumors in syngeneic or athymic mice. In an independent study, we confirmed, using the same C57MG cell line, that Wnt-1 expression leads to a similar phenotype (Blasband et al., 1992). Significantly, however, we found in the present study that this same cell line also exhibits an increase in calcium-dependent cell–cell adhesion. Together, these data demonstrate that the effects of Wnt-1 expression in these cells on cell proliferation and cell–cell adhesion are not mutually exclusive events.

In invertebrates, the Wnt-1 homolog, wingless plays a critical role in establishing segmental boundaries. It has been suggested that wingless acts to modify the character of the segmental boundary in order to prevent diffusion of morphogens (Sampedro et al., 1993). The sealing of boundaries between groups of cells could be accomplished by a Wnt-1-induced increase in cell–cell adhesion as suggested by the results of our experiments. It has also been shown in other studies that wingless acts as a morphogen in the regulation of cell proliferation and differentiation of Malpighian tubules (Skaer and Martinez-Arias, 1992). It is interesting to note that a null mutant of wingless is dwarfish, but rescued mutants that ubiquitously express wingless in the embryo are nearly returned to wildtype size (Sampedro et al., 1993). Thus, one interpretation of these data is that wingless expression affects cell proliferation. Together, these data could be interpreted as evidence that in Drosophila Wnt-1 has an effect on both cell proliferation and boundary formation/cell–cell adhesion.

In vertebrates, Wnt-1 was originally identified as a target for MMTV insertional activation that leads to mammary hyperplasias (Nusse and Varmus, 1982; Nusse et al., 1984). In Xenopus development, ectopic expression of Wnt-1 causes duplication of the embryonic axis (McMahon and Moon, 1989), although it is not known whether this effect is at the level of cell proliferation, cell adhesion, or cell differentiation. In contrast, however, expression of Wnt-1 in early Xenopus embryos results in increased gap junctional communication (Olson et al., 1991), which in mammalian cell has been shown to require calcium-dependent cell–cell adhesion (Musil et al., 1990; Jongen et al., 1991).

The duality of effects of Wnt-1 on both cell proliferation and cell–cell adhesion observed in our studies fit the paradigm of other growth factors that affect both morphogenetic movements and cell proliferation. One example is hepatocyte growth factor (HGF). This factor was originally described as "scatter factor" because it caused MDCK cells to migrate away from each other and divide (Stoker et al., 1987). However, HGF was shown in the same cell line to cause branching morphogenesis of epithelial tubules formed in vitro (Montesano et al., 1990a,b). Activin, a mesoderm inducer in Xenopus embryos, causes cell proliferation and elongation of Xenopus animal caps in vitro (Smith et al., 1990), which mimics a normal morphogenetic movement that occurs during gastrulation. At present, it is not clear how Wnt-1, or other growth factors (see above), function in the regulation of both cell adhesion and cell proliferation. One possibility is that Wnt-1 may regulate cell proliferation independent of its effects on β-catenin. Alternatively, Wnt-1 may regulate the interaction of β-catenin with other proteins that play a role in the regulation of cell proliferation. In this context it is interesting to note that β-catenin has recently been found to interact with the protein encoded by the tumor suppressor gene, APC (Rubinfeld et al., 1993; Su et al., 1993). The identification of other proteins that interact with β-catenin may provide new insights into targets of Wnt-1 regulation.

In general, the formation of different tissues requires coordinate cell proliferation and morphogenetic movements of groups of cells (e.g., radial intercalation and convergent-extension during gastrulation) (Gerhart and Keller, 1986). During these events, the integrity of groups of cells is maintained by cell–cell adhesion, but at the same time, the cells proliferate and slide past or dissociate themselves from other groups of cells (Trinkaus, 1984; Gumbiner, 1992). Based upon the results of this study, we speculate that Wnt family members could play a general role in modulating cell–cell interactions during these morphogenetic movements. Wnt-1 signaling could be mediated by its own specific cell surface receptor, as is the case of HGF (Naldini et al., 1991) and activin (Mathews and Vale, 1991; Attisano et al., 1992). That both C57MG and AT20 cells respond to Wnt-1 expression indicates that these cells carry an active receptor and signaling pathway. In these cells, the binding of Wnt-1 to, and activation of, its specific receptor may indirectly regulate the stability and accumulation of β-catenin, plakoglobin and cadherin leading to the increased strength of cell–cell adhesion. However, it remains a possibility that these effects are through direct binding of Wnt-1 to cadherin or a protein closely associated with cadherin. In either case, such a posttranscriptional mechanism for modulating cadherin/cadherin complex formation and cadherin function in cell adhesion means that the cell would be able to respond rapidly to coordinate transient cell–cell contacts in the formation of cellular boundaries during development.

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