The transcriptomic response of *Streptococcus pneumoniae* following exposure to cigarette smoke extract

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Exposure to cigarette smoke is a risk factor for respiratory diseases. Although most research has focused on its effects on the host, cigarette smoke can also directly affect respiratory pathogens, in some cases enhancing virulence. *Streptococcus pneumoniae* (the pneumococcus) is a leading cause of community-acquired pneumonia worldwide, however data on the effects of cigarette smoke on the pneumococcus are sparse. Using RNA-seq, we show that pneumococci exposed to cigarette smoke extract in a concentrated acute exposure *in vitro* model initiate a ‘survival’ transcriptional response including the upregulation of detoxification enzymes, efflux pumps and osmoregulator transporters, as well as the downregulation of fatty acid and D-alanyl lipoteichoic acid biosynthesis genes. Except for the downregulation of the pneumolysin gene, there were no changes in the expression of major virulence factors following exposure to cigarette smoke. Compared to unexposed pneumococci, smoke-exposed pneumococci did not exhibit any changes in viability, adherence, hydrophobicity or cell lysis susceptibility. In this study, we demonstrate that pneumococci adapt to acute noxious cigarette smoke exposure by inducing a gene expression signature that allows the bacteria to resist its harmful effects.

Exposure to cigarette smoke increases susceptibility to respiratory infection due to the vast array of chemicals that irritate the airways and cause cell injury, lung inflammation, and reduced lung function1–5. These effects are also observed in young children that are commonly exposed to second-hand cigarette smoke, with infants living in households with smoking family members exhibiting a higher risk of lower respiratory tract infection2,3. Although damage to the host plays a critical role in the increased risk of respiratory infections associated with cigarette smoke exposure, emerging data suggest that other factors may contribute such as the effects of smoke on the pathogen itself. Bacteria can rapidly sense and respond to changes in the environment, including exposure to chemicals such as those in cigarette smoke4,5.

The direct effects of cigarette smoke have been investigated in multiple respiratory pathogens. Cigarette smoke exposure enhances biofilm formation by *Staphylococcus aureus*6, *Streptococcus gordonii*7, *Pseudomonas aeruginosa*8, *Candida albicans*9 and *Porphyromonas gingivalis—S. gordonii* co-cultures10. Enhanced biofilm formation is often caused by smoke-induced changes in the expression of genes that facilitate biofilm formation (e.g. fimbrial and pilus genes, and transcriptional regulators of biofilm formation)9,11. In *S. aureus*, other phenotypes exhibited following cigarette smoke exposure include increased adherence to epithelial cells, reduced susceptibility to macrophage killing, and changes to the charge of the bacterial cell surface that reduce antimicrobial peptide affinity9,11.

*Streptococcus pneumoniae* (the pneumococcus) is a respiratory pathogen that is a common cause of community-acquired pneumonia. The risk of community-acquired pneumonia has been associated with cigarette
smoking in adults, with positive relationships identified with number of cigarettes smoked per day and the use of unfiltered cigarettes. Given these associations, it is not surprising that cigarette smoke is a well-known risk factor for pneumococcal disease. For example, current smoking has been identified as an independent risk factor for septic shock complications during pneumococcal pneumonia. In a colonization model, smoke-exposed mice that were intranasally infected with pneumococci exhibited reduced expression of TNF-α, CXCL-1 and CXCL-2 cytokines, suggesting cigarette smoke impairs the host response in the nasopharynx, facilitating the development of invasive disease. Pneumococcal colonization of the upper respiratory tract is enhanced in mice exposed to cigarette smoke. Pneumococcal burdens in the lungs of mice exposed to cigarette smoke were 4-fold greater at 24 hours post-infection compared to mice exposed to room air, which increased to 35-fold by 48 hours post-infection. Cigarette smoke-exposed mice also exhibited elevated production of cytokines IL-1β, IL-6, IL-10 after pneumococcal challenge as well as impaired complement-mediated phagocytosis by alveolar macrophages.

Few studies have investigated the direct impact of cigarette smoke on the pneumococcus. Pneumococcal cultures exposed to cigarette smoke condensate or black carbon exhibit enhanced biofilm formation. Cigarette smoke also reduces the activity and expression of the major cytolytic toxin, pneumolysin. However, the effect of cigarette smoke on pneumococcal gene expression, as well as on other phenotypes, remains unknown. Transcriptional data provide a broad understanding of how pathogens such as the pneumococcus can exploit hostile environments within a susceptible host. In this study, we aimed to investigate how pneumococcus responds to acute cigarette smoke exposure by assessing gene expression across the pneumococcal genome.

**Results and Discussion**

We hypothesized that the chemicals in cigarette smoke would have profound effects on pneumococcal gene expression. To identify transcriptomic differences mediated by exposure to cigarette smoke, log-phase cultures of pneumococcal strain EF3030 (serotype 19F) were exposed to either THY media or THY treated with cigarette smoke (hereby referred to as CSE, cigarette smoke extract) using a short and concentrated acute exposure *in vitro* model. RNA was extracted from both cultures and RNA-Seq was performed to identify genes that were differentially expressed. Genes with a log2(fold change) of greater than 1 or less than −1 in CSE cultures relative to THY cultures and a false discovery of <0.05 were considered significantly differentially expressed in cigarette smoke.

Following incubation of pneumococcal cultures in CSE, 59 genes were upregulated compared to the THY control (Table 1). These include genes involved in competence, vitamin B6 synthesis and sugar catabolism. Other genes that were upregulated after CSE-exposure included glyoxalase (SPCG_RS00380), which removes methylglyoxals and other aldehydes (both of which can induce oxidative cell damage when accumulated to high levels). Multiple oxidoreductase-encoding genes (SPCG_RS07925, SPCG_RS08280, SPCG_RS09290, SPCG_RS09075, SPCG_RS07475 and SPCG_RS05800) were also upregulated in CSE. Oxidoreductases also play a role in the removal of glyoxals. In addition, multiple metal ion transporters and metal-dependent transcriptional regulators including a zinc efflux (SPCG_RS09490, SPCG_RS09485 and SPCG_RS09480) and copper efflux operon (SPCG_RS03545, SPCG_RS03550, SPCG_RS03555) were upregulated in CSE. Cigarette smoke contains an array of toxic substances including methylglyoxals and glyoxals, as well as metals such as zinc and copper. Interestingly, zinc transporter cccD (SCPG_RS09490) is also upregulated in the presence of cadmium, which is also present in cigarette smoke. The intracellular accumulation of cadmium is also associated with oxidative stress by interfering with manganese uptake. The upregulation of these genes in pneumococcus is likely to detoxify or remove these potentially harmful substances from the cell and combat oxidative stress. Interestingly, viable counts of pneumococci incubated in either THY or CSE did not exhibit any difference in viability (Fig. 1A, Supplementary Fig. S1), suggesting the upregulation of these genes may help the bacteria to survive in the presence of these substances.

The glycine betaine ABC transporter operon encodes proteins that respond to osmotic stress. The gene products in this operon play a role in the accumulation of compatible solutes such as proline and glycine betaine, which are essential for osmoregulation. The genes in this putative operon, including a MarR family transcriptional regulator (SPCG_RS09520), hypothetical protein (SPCG_RS09515), glycine betaine ABC transporter ATP-binding protein, ProV (SPCG_RS09510) and a glycine betaine ABC transporter permease, ProWX (SPCG_RS09505), were all upregulated after CSE exposure (Table 1). These genes were also upregulated during the acid tolerance response of pneumococci to low pH conditions. In our experiments, cigarette smoke treatment of THY media lowered the pH by ~0.8 (mean pH of CSE media 7.09 ± 0.13 compared to 7.91 ± 0.20 for THY), which is lower than the optimal pH for pneumococci. Martin-Galiano et al. proposed a relationship between the pneumococcal acid tolerance response, and other stress responses including oxidative and osmotic stress. As such, the changes in pneumococcal gene expression in response to cigarette smoke may be mediated via direct exposure to chemicals in cigarette smoke and/or changes in physiological pH.

Smoke exposure enhances biofilm formation in bacterial pathogens including *P. aeruginosa*, *S. gordonii* and *S. aureus*. Pneumococcal cultures exposed to cigarette smoke condensate also exhibit augmented biofilm formation. In our study, pcpA, which encodes a choline-binding protein, was upregulated in CSE. PcpA contributes to biofilm formation, supported by the diminished biofilm formation phenotype observed in pcpA mutants. The upregulation of pcpA in CSE we identified in this study is consistent with published data on increased biofilm formation exhibited by pneumococcus following smoke exposure as well as previous reports on the upregulation of this gene during metal stress. Some pneumococci contain a 7 kb insertion encoding a second copy of the *pcpA* gene. This is also the case in EF3030, the strain used in this study. Interestingly, only one pcpA gene (SPCG_RS06505) was upregulated in CSE. PcpA has also been reported to play a role in adherence to nasopharyngeal and lung epithelial cells. Although, CSE-treated pneumococcal cultures exhibit an upregulation of *pcpA*, no increase in adherence to A549 lung epithelial cells was observed (Fig. 1B). This may be because pcpA was the only adhesin that was upregulated in CSE. In the absence of changes in expression of other major adhesins (e.g. PsaA), this may not be sufficient to increase adherence. In addition, cigarette smoke exposure of A549 cells
| Functional group or gene | Description | log2 (fold change) | FDR |
|--------------------------|-------------|--------------------|-----|
| Competence/competence induced cell lysis |            |                    |     |
| SPCG_RS10625 | Competence protein ComGC | 1.81 | 0.048 |
| SPCG_RS04840 | Late competence protein ComEA, DNA receptor | 1.43 | 0.012 |
| SPCG_RS10610 | Competence protein ComGF | 1.37 | 0.044 |
| SPCG_RS11465 | Choline-binding protein D CbpD | 1.29 | 0.033 |
| SPCG_RS06370 | DNA protecting protein DprA | 1.27 | 0.041 |
| SPCG_RS04845 | Late competence protein ComEC, DNA transport | 1.17 | 0.049 |
| Two-component regulatory systems |            |                    |     |
| SPCG_RS10300 | Two-component system transcriptional response regulator 11 | 2.21 | 0.004 |
| SPCG_RS10305 | Two-component system sensor histidine kinase 11 | 2.12 | 0.006 |
| Nucleotide transport/biosynthesis |            |                    |     |
| SPCG_RS01145 | Anaerobic ribonucleoside-triphosphate reductase activating protein NrdG | 1.92 | 0.022 |
| SPCG_RS09500 | Nicotinamide mononucleotide transporter PnuC | 1.89 | 0.006 |
| SPCG_RS01150 | Phosphoribulokinase | 1.73 | 0.038 |
| SPCG_RS09080 | NADPH-dependent 7-cyano-7-deazaguanine reductase QueF | 1.49 | 0.011 |
| Glycine betaine transport |            |                    |     |
| SPCG_RS099515 | Hypothetical protein | 2.75 | 0.008 |
| SPCG_RS09520 | MarR family transcriptional regulator | 2.70 | 0.008 |
| SPCG_RS09510 | Glycine/betaine ABC transporter ATP-binding protein ProY | 2.57 | 0.011 |
| SPCG_RS09505 | Glycine/betaine ABC transporter permease ProWX | 2.41 | 0.007 |
| Sugar catabolism |            |                    |     |
| SPCG_RS09465 | Galactose-1-phosphate uridylyltransferase GalT2 | 2.22 | 0.003 |
| SPCG_RS09700 | Sugar ABC transporter substrate-binding protein MsrE | 1.89 | 0.038 |
| SPCG_RS03585 | 6-phospho-beta-glucoisidase | 1.82 | 0.038 |
| SPCG_RS09705 | Alpha-galactosidase Aga | 1.72 | 0.042 |
| SPCG_RS01445 | Glutamine-fructose-6-phosphate aminotransferase GlmS | 1.54 | 0.005 |
| SPCG_RS09680 | Sacrose phosphorylase GtIa | 1.33 | 0.037 |
| SPCG_RS09470 | Galactokinase GalK | 1.26 | 0.007 |
| Vitamin B6 biosynthesis |            |                    |     |
| SPCG_RS07540 | Pyridoxal 5’-phosphate synthase subunit PdxS | 1.67 | 0.004 |
| SPCG_RS07535 | Pyridoxal 5’-phosphate synthase subunit PdxT | 1.41 | 0.007 |
| Acetolactate synthesis |            |                    |     |
| SPCG_RS02285 | Acetolactate synthase, large subunit IlvB | 1.34 | 0.006 |
| SPCG_RS02290 | Acetolactate synthase, small subunit IlvH | 1.00 | 0.044 |
| Oxidoreductases |            |                    |     |
| SPCG_RS07925 | NAD(P)H-dependent FMN reductase | 3.32 | 0.000 |
| SPCG_RS08280 | NADH-flavin reductase | 2.46 | 0.034 |
| SPCG_RS02990 | NAD(P)H-flavin reductase | 2.35 | 0.004 |
| SPCG_RS09075 | Thioredoxin | 2.02 | 0.004 |
| SPCG_RS07475 | Thioredoxin-diisulfide reductase | 1.97 | 0.004 |
| SPCG_RS05800 | Glutaredoxin | 1.14 | 0.048 |
| Metal transport |            |                    |     |
| SPCG_RS09480 | Zn-dependent alcohol dehydrogenase AdhB | 4.30 | 0.0004 |
| SPCG_RS09485 | MerR family transcriptional regulator | 3.97 | 0.003 |
| SPCG_RS09490 | Cation transporter, CzcD | 3.50 | 0.004 |
| SPCG_RS03555 | Copper-translocating P-type ATPase, CopA | 2.46 | 0.004 |
| SPCG_RS03550 | Cupredoxin domain-containing protein, CopA | 2.42 | 0.004 |
| SPCG_RS03545 | CopY/TcrY family copper transporter transcriptional repressor, CopY | 2.41 | 0.004 |
| SPCG_RS00385 | Metal-sensitive transcriptional repressor, FrmR family | 2.36 | 0.013 |
| SPCG_RS1895 | Peptide ABC transporter ATP-binding protein | 1.31 | 0.006 |
| Adherence |            |                    |     |
| SPCG_RS06505 | Choline-binding protein A, PcpA | 1.34 | 0.046 |
| Other genes |            |                    |     |
| SPCG_RS00380 | Glyoxalase | 4.46 | 0.003 |
| SPCG_RS4530 | Chlorohydrolase, predicted pseudogene | 3.33 | 0.005 |
| SPCG_RS11630 | Membrane protein YhGL, phage infection protein (PIP) family | 2.65 | 0.006 |

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brane functions such as permeability 46. In these bacteria, the release of DNA into the environment leads to the formation of extracellular DNA (eDNA), which can be taken up by neighboring cells and used for genetic recombination.

Among these genes, a substantial proportion is predicted to result in changes to the pneumococcal cell surface, which is an important component of biofilms. Competence genes upregulated in CSE include cbpD, which encodes a protein that is important in biofilm formation and restriction of nutrient accessibility 42. The upregulation of competence genes in response to CSE may explain the enhanced biofilm formation observed previously by Cockeran et al. 20. In addition, the upregulation of competence genes can occur during stress conditions such as antibiotic stress and DNA damage 43–45. Together with the enhanced biofilm formation observed previously by Cockeran et al. 20, these findings on the augmented pneumococcal adherence phenotype described previously are driven more by the effects of cigarette smoke on the host rather than on the pathogen itself.

An important component of biofilms is extracellular DNA. In pneumococci, this is partially mediated by bacterial fratricide, whereby non-competent pneumococci are killed by their competent neighbors. The DNA released from lysed pneumococci can not only be taken up by competent pneumococci, but serves as a mechanism to maintain cell surface hydrophobicity, rather than allowing it to increase. Previous work on S. aureus has shown that cigarette smoke can increase the expression of PAFR (platelet activating factor receptor), which enhances pneumococcal adhesion 34. In our study, A549 cells were not exposed to cigarette smoke. Taken together, these findings on the augmented pneumococcal adherence phenotype described previously are driven more by the effects of cigarette smoke on the host rather than on the pathogen itself.

In the differential gene expression analysis, 63 genes were significantly downregulated in CSE (Table 2). These genes are involved in fatty acid biosynthesis (Table 2). Fatty acids are an important component in phosholipids that make up the lipid bilayer of cell membranes. Bacteria are capable of regulating membrane lipid homeostasis by regulating the expression of genes involved in fatty acid biosynthesis, which affects various membrane functions such as permeability 46. In Streptococcus mutans, deletion of the fatty acid isomerase gene fabM results in significant changes to membrane composition and enhanced ATPase activity, proton permeability and phosphotransferase activity 47. The pneumococcal homolog of fabM (SPCG_RS02150) was among the fatty acid biosynthesis genes that were downregulated. Some ATPases and transporters were also upregulated following CSE exposure (e.g. SPCG_RS10310, SPCG_RS10315 and SPCG_RS07325, Table 1). The downregulation of fatty acid biosynthesis genes in CSE-exposed cells suggested that cigarette smoke may induce changes in the hydrophobicity of the pneumococcal cell surface. However, when pneumococcal hydrophobicity was evaluated, no difference between CSE and THY cultures was observed (Fig. 1C). Cigarette smoke has been shown to increase hydrophobicity in S. aureus 12. It is therefore plausible that cigarette smoke would be acting in a similar manner on pneumococcus. However, unlike S. aureus, pneumococci downregulate fatty acid biosynthesis genes, possibly as a mechanism to maintain cell surface hydrophobicity, rather than allowing it to increase. Previous work on

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Table 1. Pneumococcal genes upregulated following 45 min incubation in cigarette smoke-treated THY (CSE) media relative to cultures incubated in THY media. SPCG number is the locus tag ID for each gene in the CGSP14 genome sequence, which was used to map the EF3030 RNA reads from two independent cultures. Only differentially expressed genes are shown, defined as a log2(fold change) of greater than 1 or less than −1 and a false discovery rate (FDR) of < 0.05. Genes in bold and italics were validated by qRT-PCR.

| Functional group or gene | Description | log2 (fold change) | FDR |
|--------------------------|-------------|--------------------|-----|
| SPCG_RS099070            | DUF469 domain-containing protein | 2.39 | 0.009 |
| SPCG_RS10310             | ABC transporter permease | 2.05 | 0.021 |
| SPCG_RS10320             | Hypothetical protein | 1.94 | 0.022 |
| SPCG_RS10315             | ABC transporter ATP-binding protein | 1.92 | 0.007 |
| SPCG_RS01140             | GNAT family acetyltransferase | 1.75 | 0.044 |
| SPCG_RS09065             | Mari family transcriptional regulator | 1.50 | 0.004 |
| SPCG_RS10895             | Predicted transcriptional regulator, pseudogene | 1.46 | 0.048 |
| SPCG_RS07525             | ABC-type lipoprotein export system, ATPas component | 1.33 | 0.049 |
| SPCG_RS06665             | tRNA-specific 2-thiouridylase MnmA | 1.18 | 0.015 |
| SPCG_RS03595             | Sodium-dependent transporter | 1.13 | 0.008 |
| SPCG_RS10255             | Putative cell surface protein with DUF1542 domain | 1.10 | 0.019 |
| SPCG_RS10750             | tRNA-Ile | 1.08 | 0.008 |
| SPCG_RS10445             | Permease | 1.07 | 0.037 |
| SPCG_RS10695             | MATE family efflux transporter | 1.02 | 0.037 |
the effects of cigarette smoke on S. aureus used a non-encapsulated strain. This is in contrast to our study in which EF3030 produces a serotype 19F capsular polysaccharide. It is plausible that the pneumococcal extracellular capsular polysaccharide may provide a physical barrier that masks the effects of cigarette smoke on cell surface hydrophobicity. Future studies could explore this area further by testing the hydrophobicity of isogenic capsule mutants.

Another cigarette smoke-driven effect on the pneumococcal cell surface with the potential to impact membrane permeability and molecule affinity is the downregulation of the \( \textit{dlt} \) operon (SPCG_RS11320, SPCG_RS11325, SPCG_RS11330, SPCG_RS11335, and SPCG_RS11340). This operon encodes proteins involved in D-alanyl-lipoteichoic acid biosynthesis, which involves the addition of D-alanine onto teichoic acids in the cell wall. This reduces the negative charge of the cell surface, diminishing its affinity to cationic antimicrobial peptides (CAMPs) and promoting the evasion of CAMP-mediated killing. Pneumococcal \( \textit{dlt} \) mutants display increased susceptibility to CAMPs, and upregulation of the \( \textit{dlt} \) operon has been observed in response to the presence of CAMPs such as nisin and LL-37. Interestingly, the lack of D-alanyl lipoteichoic acids (due to repression of the \( \textit{dlt} \) operon) in CSE-exposed pneumococci would increase its negative charge. The changes at the pneumococcal cell surface raise an interesting question around how smoke-exposed pneumococci would behave in the context of the nasopharyngeal environment. The pneumococcal capsule is negatively charged, which facilitates pneumococcal colonization by the capsule mediating electrostatic repulsion of the mucus lining of the upper respiratory tract, which is also negatively charged due high levels of sialic acid. This allows the bacterium to escape mucus-dependent clearance. In our study, the putative CSE-induced increase in negative charge of the cell surface (from the downregulation of the \( \textit{dlt} \) operon) would likely enhance colonization of the nasopharynx by a similar mechanism. Pneumococcal colonization of the upper respiratory tract is enhanced in mice exposed to cigarette smoke. With multiple smoke-induced changes in pneumococcal gene expression that could impact colonization, it is plausible that the effects of smoke on the pneumococcus may contribute to this phenotype.

Two other genes involved in cell wall metabolism were also downregulated in CSE. These included \( \textit{lytB} \) (endo-\( \beta \)-N-acetylglucosaminidase, SPCG_RS04895) and \( \textit{lysM} \) (peptidoglycan-binding protein, SPCG_RS00545). The differential expression of these genes could induce changes in the cell wall architecture, which may influence susceptibility to cell lysis and potentially influence the ability of the bacterium to persist within a host. We therefore assessed whether CSE exposure would impact susceptibility to cell lysis. However, no difference
| Functional group or gene | Description | log2 (fold change) | FDR |
|-------------------------|-------------|--------------------|-----|
| **Fatty acid biosynthesis** | | | |
| SPCG_RS02210 | Acetyl-CoA carboxylase carboxyl transferase subunit alpha AccA | −3.64 | 0.003 |
| SPCG_RS02205 | Acetyl-CoA carboxylase carboxyl transferase subunit beta AccD | −3.59 | 0.003 |
| SPCG_RS02200 | Acetyl-CoA carboxylase biotin carboxylase subunit AccC | −3.34 | 0.004 |
| SPCG_RS02190 | Acetyl-CoA carboxylase biotin carboxyl carrier protein subunit AccB | −3.32 | 0.004 |
| SPCG_RS02195 | 3-hydroxyacyl-ACP dehydratase FabZ | −3.18 | 0.005 |
| SPCG_RS02185 | 3-oxoacyl-ACP reductase FabG | −2.92 | 0.006 |
| SPCG_RS02180 | 3-ketoacyl-ACP reductase FabK | −2.10 | 0.036 |
| SPCG_RS02175 | Acetyl-CoA carboxylase | −2.06 | 0.007 |
| SPCG_RS02170 | 3-hydroxyacyl-ACP dehydratase Enoyl-[acyl-carrier-protein] reductase FabM | −2.82 | 0.026 |
| SPCG_RS02160 | Enoyl-[acyl-carrier-protein] reductase FabK | −2.10 | 0.036 |
| SPCG_RS02165 | Acetyl-CoA carboxylase | −2.06 | 0.007 |
| SPCG_RS02155 | Pneumolysin Ply | −3.39 | 0.004 |
| **Pyrimidine biosynthesis** | | | |
| SPCG_RS03420 | Orotate phosphoribosyltransferase PyrE | −2.54 | 0.003 |
| SPCG_RS03415 | Orotidine 5′-phosphate decarboxylase PyrF | −2.30 | 0.006 |
| SPCG_RS06430 | bifunctional pyrimidine operon transcriptional regulator/uracil phosphoribosyltransferase PyrR | −1.83 | 0.004 |
| SPCG_RS06420 | Carbamoyl-phosphate synthase small chain CarA | −1.82 | 0.004 |
| SPCG_RS06425 | Aspartate carbamoyltransferase PyrB | −1.80 | 0.004 |
| SPCG_RS04890 | Dihydroorotate dehydrogenase PyrD | −1.73 | 0.004 |
| SPCG_RS086415 | Carbamoyl-phosphate synthase large chain CarB | −1.52 | 0.005 |
| SPCG_RS04885 | Dihydroorotate dehydrogenase PyrDII | −1.34 | 0.012 |
| **Virulence** | | | |
| SPCG_RS09950 | Pneumolysin Ply | −3.39 | 0.004 |
| **D-alanyl-lipoteichoic acid biosynthesis** | | | |
| SPCG_RS11330 | D-alanyl-lipoteichoic acid biosynthesis protein DltD | −1.51 | 0.031 |
| SPCG_RS11320 | D-alanyl-lipoteichoic acid biosynthesis protein DltC | −1.47 | 0.025 |
| SPCG_RS11325 | D-alanine-poly(phosphoribitol) ligase subunit 2 DltC | −1.46 | 0.007 |
| SPCG_RS11335 | D-alanine-poly(phosphoribitol) ligase subunit 1 DltA | −1.24 | 0.046 |
| **Carbohydrate transport and metabolism** | | | |
| SPCG_RS07255 | Glucosamine-6-phosphate deaminase NagB | −1.99 | 0.049 |
| SPCG_RS010975 | Glycogen/starch/alpha-glucan family phosphorylase MalP | −1.77 | 0.046 |
| SPCG_RS00515 | Glycosyltransferase | −1.64 | 0.042 |
| SPCG_RS00520 | Polysaccharide biosynthesis protein | −1.48 | 0.037 |
| SPCG_RS04895 | Endo-beta-N-acetylglucosaminidase LysB | −1.40 | 0.004 |
| SPCG_RS08595 | Carbohydrate ABC transporter permease | −1.13 | 0.044 |
| SPCG_RS09625 | Phosphotransferase system trehalose transporter subunit IIABC TreP | −1.07 | 0.014 |
| **Cell wall/membrane biogenesis** | | | |
| SPCG_RS00545 | Peptidoglycan-binding protein LysM | −1.76 | 0.040 |
| SPCG_RS04610 | ABC-type lipoprotein export system, ATPase component | −1.40 | 0.019 |
| SPCG_RS10170 | Membrane protein insertase YidC | −1.16 | 0.021 |
| **Lipid transport and metabolism** | | | |
| SPCG_RS04125 | Glycerol-3-phosphate acyltransferase | −1.27 | 0.006 |
| SPCG_RS08265 | 1-acyl-sn-glycerol-3-phosphate acyltransferase | −1.16 | 0.046 |
| **Proteases** | | | |
| SPCG_RS02225 | CAAX protease self-immunity protein | −2.28 | 0.004 |
| SPCG_RS01770 | ATP-dependent Clp protease ATP-binding protein ClpL | −2.08 | 0.037 |
| SPCG_RS11675 | Serine protease HtrA | −1.23 | 0.044 |
| SPCG_RS11690 | Membrane proteinase | −1.16 | 0.046 |
| **Other genes** | | | |
| SPCG_RS09955 | Hypothetical protein | −2.94 | 0.004 |

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was observed in the ability of CSE and THY cultures to resist detergent-mediated lysis (Fig. 1D), suggesting the integrity of the cell wall has not been modified by CSE exposure.

Following exposure to CSE, the major virulence gene, *ply* (SPCG_RS09950), which encodes the cytolytic toxin pneumolysin, was downregulated (Table 2), a finding that is consistent with what has been described previously20. Although some genes with virulence associated roles in metal transport and adherence were differentially expressed in CSE, most major pneumococcal virulence factors (e.g. *lytA*, *pspA*, *nanA*, capsular polysaccharide biosynthesis genes)21 were not. Taken together with the previous work of Cockeran et al.20, our data suggest that exposure of pneumococci to cigarette smoke promotes survival and biofilm formation.

To confirm whether the genes identified in RNA-Seq are indeed differentially expressed following CSE exposure, a subset of 13 genes (8 upregulated and 5 downregulated) that were differentially expressed in EF3030 CSE cultures by RNA-Seq were validated by quantitative reverse transcription polymerase chain reaction (qRT-PCR). The 13 genes were chosen to represent diverse functional categories including metal transport, competence, fatty acid biosynthesis, virulence, D-alanyl-lipoteichoic acid biosynthesis and transcriptional regulation. Consistent with the RNA-Seq findings, all 13 genes that were differentially expressed in CSE using RNA-Seq were also differentially expressed in EF3030 CSE media relative to cultures incubated in THY media. SPCG number is the locus tag ID for each gene in the CGSP14 genome sequence, which was used to map the EF3030 RNA reads from two independent cultures. Only differentially expressed genes are shown, defined as a log₂(fold change) of greater than 1 or less than −1 and a false discovery rate (FDR) of <0.05. Genes in bold and italics were validated by qRT-PCR.

Table 2. Pneumococcal genes downregulated following 45 min incubation in cigarette smoke-treated THY (CSE) media relative to cultures incubated in THY media. SPCG number is the locus tag ID for each gene in the CGSP14 genome sequence, which was used to map the EF3030 RNA reads from two independent cultures. Only differentially expressed genes are shown, defined as a log₂(fold change) of greater than 1 or less than −1 and a false discovery rate (FDR) of <0.05. Genes in bold and italics were validated by qRT-PCR.

| Functional group or gene | Description | log₂ (fold change) | FDR |
|--------------------------|-------------|-------------------|-----|
| SPCG_RS02220             | Membrane protein | −2.23             | 0.005 |
| SPCG_RS09980             | Hypothetical protein | −2.80             | 0.006 |
| SPCG_RS09965             | DUF4231 domain-containing protein | −2.20             | 0.048 |
| SPCG_RS10500             | Bifunctional acetaldehyde-CoA/alcohol dehydrogenase | −1.91             | 0.024 |
| SPCG_RS08740             | Sodium ABC transporter permease | −1.76             | 0.029 |
| SPCG_RS04615             | FtsX-like ABC transporter permease | −1.67             | 0.007 |
| SPCG_RS03430             | Hypothetical protein | −1.57             | 0.015 |
| SPCG_RS11680             | Chromosome partitioning protein ParB | −1.54             | 0.038 |
| SPCG_RS08185             | Hypothetical protein | −1.42             | 0.006 |
| SPCG_RS39000             | DUF3270 domain-containing protein | −1.28             | 0.048 |
| SPCG_RS03575             | Hypothetical protein | −1.24             | 0.008 |
| SPCG_RS09945             | YeBC/PmPR family DNA-binding transcriptional regulator | −1.24             | 0.044 |
| SPCG_RS04680             | Phage shock protein PspC (stress-responsive transcriptional regulator) | −1.20             | 0.038 |
| SPCG_RS07515             | Putative channel-forming cytolysin, hemolysin III family | −1.12             | 0.019 |
| SPCG_RS08585             | Oxidoreductase | −1.09             | 0.029 |
| SPCG_RS05215             | DUF1002 domain-containing protein | −1.00             | 0.038 |

The role of TCS11 is not clear and the genes regulated by this system have not been described to date. Mutation of the histidine kinase and response regulator genes of TCS11 had no effect in a murine respiratory tract infection model, suggesting it does not play a role in virulence52. It has been postulated that TCS11 may be involved in responding to stress caused by environmental stimuli53, indicated by its upregulation following exposure to the antibiotic vancomycin54. The upregulation of TCS11 following CSE exposure supports this notion.

An interesting question is whether these changes in gene expression also occur within *in vivo* systems. The *in vitro* system is disadvantaged by the fact that it is not a physiologically relevant system and does not accurately reconstruct a pneumococcal carriage episode. Examining pneumococcal gene expression in colonized mice exposed to either cigarette smoke or air will determine whether the transcriptomic response observed in our study is also an *in vivo* occurrence. It will also allow the investigation of whether these changes are tissue-specific (e.g. comparing pneumococcal expression in the nasopharynx vs lung of smoke-exposed mice). To better understand how these changes relate to pneumococcal carriage and smoking behavior in humans, this mouse model can also be used to uncover if smoke-induced changes in pneumococcal gene expression are dependent on dose or exposure duration, which were only briefly investigated in the *in vitro* system of this study (Supplementary Figs S1 and S2). The results from our study suggest that the increased susceptibility to pneumococcal disease in smokers would unlikely be due to smoke-induced changes in the pathogen itself. However, it is still not clear whether this increased susceptibility is due to a synergistic combination of smoke-induced effects on both the pathogen and the host. For example, it is possible that pneumococci modify their gene expression profile for...
enhanced virulence by detecting and responding to the smoke-induced damage to the host (e.g., detection of key inflammatory agents produced by the host in response to cigarette smoke), which would not be detected using *in vitro* systems. A smoke-exposure animal model would be suited to answering this question. In addition, the results from our study demonstrate that pneumococci have the capacity to resist the harmful effects of cigarette smoke. This would allow pneumococci to survive in a smoke-exposed host environment, which together with a more susceptible host may play a significant role in the increased infection risk observed in smokers.

The work described in this study begins to elucidate the genetic basis underlying the behavior of pneumococcus in a cigarette smoke-exposed environment. To our knowledge, this is the first report evaluating the direct effects of cigarette smoke on a respiratory pathogen from a transcriptomic perspective and therefore provides novel insights into the global response of pneumococci to cigarette smoke exposure. These responses include; (1) an upregulation of genes facilitating the detoxification and removal of toxic compounds; (2) activation of genes responsive to physiological stress; (3) upregulation of genes involved in gene exchange and biofilm formation; and (4) repression of genes required for fatty acid biosynthesis and D-alanylation of lipoteichoic acids. These changes ensure survivability and adaptability of pneumococci in a cigarette smoke exposed environment. The phenotypes that were assessed (viability, adherence, hydrophobicity and lysis susceptibility) did not change upon CSE exposure, which may be due to rapid reversion of gene expression profile upon removal of the CSE stimulus at the time the phenotypes were investigated. Alternatively, it is possible that the phenotypes do not change as the transcriptomic changes in gene expression allow pneumococci to resist the harmful effects of CSE. Our data demonstrate that smoke exposure results in a gene expression signature of survivability and adaptability rather than directly impacting pneumococcal virulence. Future studies should focus on testing different lengths of exposure duration as well as concentration of CSE to determine if the same changes in gene expression are observed.

**Materials and Methods**

**Preparation of cigarette smoke extract media.** Cigarette smoke extract (CSE) medium was prepared using a method adapted from a previous study.

In brief, four 3R4F research grade cigarettes (Kentucky Tobacco Research & Development Centre, University of Kentucky) were bubbled through 10 ml of THY medium (3% [w/v] Todd-Hewitt broth, 0.5% [w/v] yeast extract). This produced the highest concentration of CSE without affecting pneumococcal viability (Supplementary Fig. S1) and was thereby considered to provide the strongest transcriptional response. This was performed using a smoke apparatus and syringe to draw the smoke into the medium. Cigarette smoke was drawn in 30 ml tidal volumes with a 20 sec break between draws. The control THY medium was prepared in the same manner except that air was drawn through the apparatus instead of cigarette smoke. Air controls were conducted in a chemical fume hood prior to preparation of CSE to ensure there is no contamination of control media with residual smoke. Media were filter sterilized before use with a 0.22 µm syringe-driven filter.
Exposure of pneumococcal cultures to CSE. Pneumococcal strain EF3030 (serotype 19F, multi-locus sequence type 43) or PMP1287 (serotype 16F, multi-locus sequence type 3117) was inoculated in THY media and incubated at 37 °C, 5% CO₂ to an OD₆₀₀ of 0.1–0.2. Bacteria were then harvested in 3 ml aliquots by centrifugation (1820 × g for 2 min) and the supernatants removed. Pellets were resuspended in 3 ml of THY or CSE and incubated for 45 min. This time point was chosen based on experiments examining expression of the *ply* operon following incubation of pneumococcal cultures in CSE (Supplementary Fig. S2). This operon was chosen as it included the *ply* gene, which has been demonstrated to be differentially expressed in response to cigarette smoke previously. For the enumeration of pneumococci following CSE exposure, cultures were serially diluted in 0.85% saline, plated on horse blood agar plates and incubated overnight at 37 °C, 5% CO₂. Additionally, 0.5 ml aliquots of pneumococcal cultures were mixed with 1 ml RNAprotect Bacteria Reagent (Qiagen), incubated at room temperature for 5 min and stored at −80 °C for RNA extraction.

RNA extraction. Extraction of RNA from pneumococci was performed using the RNeasy Mini Kit (Qiagen) according to the manufacturer's instructions with the incorporation of an additional lysis step. In summary, bacterial preparations that were stored in RNAlater Bacteria Reagent (Qiagen) (as described above) were thawed and incubated at 37 °C for 15 min in 200 µl lysis buffer containing 600 µg/ml lysozyme (Sigma-Aldrich), 15 µg/ml mutanolysin (Sigma-Aldrich), 1U/µg SUPERase In RNase Inhibitor (Ambion) and 400 µg/ml Proteinase K (Qiagen) in TE buffer. The rest of the procedure was conducted as per the RNeasy Mini Kit instructions. An on-column DNase treatment was performed by the addition of 2727 Kunitz U/ml DNase I in buffer RDD, (Qiagen), and incubated at room temperature for 15 min prior to washing and elution in 20 µl RNase-free water. RNA concentration and integrity was examined by Tape Station (Agilent). RNA was stored at −80 °C until use.

RNA-Seq and differential gene expression analysis. RNA, extracted from EF3030 cultures grown in either THY or CSE on two independent occasions, was sequenced. Libraries were prepared using the TrueSeq Stranded Total RNA with Ribo-Zero Bacteria kit (Illumina) to deplete bacterial rRNA. Sequencing was then performed on the Illumina HiSeq. 4000 platform in 2 × 75 bp reads (total read counts for THY samples 68,717,994 and 99,576,470; CSE samples 67,387,854 and 91,871,848). Because the EF3030 genome has not been fully sequenced, reads were mapped to the most closely related complete pneumococcal genome available in the NCBI database at the time of analysis (CGSP14, serotype 14, accession no. NC_010582, which shared four of seven multi-locus sequence type alleles with EF3030) using Bowtie 2.5. Differential expression of the capsular polysaccharide locus was assessed by mapping RNA-Seq reads to the serotype 19F capsule locus. A counts table was constructed using htseq-count to determine the number of reads that mapped to each gene from each condition (CSE versus THY). Differentially expressed genes were then identified from the counts table using the Voom/LIMMA package, with data normalized for read depth and library size. Genes were defined as differentially expressed in CSE if the log2(fold change) in the CSE condition was greater than 1 or less than −1 and a false discovery rate cutoff of <0.05.

Quantitative reverse transcription polymerase chain reaction (qRT-PCR). To validate RNA-Seq data, a selection of differentially expressed genes identified by RNA-Seq were also tested using qRT-PCR. Pneumococcal cultures were once again incubated in THY or CSE and the RNA extracted. This RNA was used for first strand cDNA synthesis using the iScript™ cDNA Synthesis Kit (Bio-Rad). For each sample 2.5 ng RNA was added and incubated in a thermocycler under the following protocol: 5 min at 25 °C, 30 min at 42 °C and then 5 min at 85 °C. The cDNA was then stored at −20 °C until required. cDNA was used for qPCR in duplicate reactions using the GoTaq® qPCR Master Mix (Promega) as per the manufacturer's instructions. Each reaction contained 2 µl of cDNA (or nuclease free water for the no template control) and 0.2 M of forward and reverse primers. Primer sequences are provided in Table S1. Reactions were run on the Mx3005P qPCR system (Agilent Technologies) under the following conditions: 2 min at 95 °C followed by 40 cycles of 15 sec at 95 °C and 1 min at 60 °C. Non-specific amplification was assessed by performing a dissociation curve at the end of the run (1 min at 95 °C, 30 sec at 60 °C and then 30 sec at 95 °C). Analysis of each gene for changes in its expression in CSE relative to THY cultures was performed using the 2⁻ΔΔCt relative quantification method. The gyrA gene was used as a reference gene as described previously.

Pneumococcal adherence assay. The ability of CSE-exposed pneumococci to adhere to A549 lung epithelial cells was evaluated using an adherence assay. A549 cells were seeded overnight in a 24 well tray at 1.5 × 10⁵ cells per well. EF3030 cultures (incubated in either THY or CSE) were harvested by centrifugation and resuspended to ~1.5 × 10⁶ CFU/ml in 0.85% saline. Seeded A549 cells were washed with PBS prior to adding ~1.5 × 10⁶ CFU/ml pneumococci (or saline as a negative control) to duplicate wells. The tray was subject to centrifugation at 114 × g for 3 min before incubating at 37 °C, 5% CO₂ for 1 h. Non-adherent bacteria were removed by washing the wells in PBS three times. Epithelial cells were subsequently lysed by the addition of 0.1% digitonin (Sigma-Aldrich) to each well and incubated at 37 °C, 5% CO₂ for 5 min. Following incubation, cell-associated pneumococci were resuspended in THY and viable counting was conducted. The percentage of remaining pneumococci was calculated by comparing the number of remaining pneumococci to the starting inoculum added to the wells.

Sensitivity to cell lysis. Bacterial cultures incubated in either THY or CSE were prepared as above. Cultures were then washed twice with PBS and resuspended in fresh THY media. To these cultures, 0.005% Triton X-100 was added and were incubated at 37 °C, 5% CO₂. At 0, 15 and 30 min after the addition of Triton X-100, aliquots were taken for viable counting, and results used to calculate percent survival at each time point.
Hydrophobicity assay. To assess cell surface hydrophobicity, the microbial adhesion to hydrocarbons assay was performed as described previously\(^8\). Pneumococcal cultures that had been incubated in either THY and CSE media were washed twice in PBS. At this point, an aliquot was taken for viable counts. Hexadecane was added to the bacterial suspensions and were mixed in a 3:1 ratio by vortexing for 2 min, followed by incubation at room temperature for 30 min. After incubation, an aliquot was taken from the aqueous phase for viable counts. This was used to calculate the proportion of pneumococi remaining in the aqueous phase.

Data Availability
Raw RNA-Seq data are available from the corresponding author upon reasonable request.

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Author Contributions

S.M., E.M.D., S.B. and C.S. conceived the experiments and interpreted results, S.M., A.W., and A.P. conducted the experiments, S.M. and N.E.H. analyzed the results. S.M. and C.S. wrote the manuscript. All authors reviewed and approved the manuscript for submission.

Additional Information

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