Achieving high-efficiency emission depletion nanoscopy by employing cross relaxation in upconversion nanoparticles

Qiuqiang Zhan1, Haichun Liu2, Baoju Wang1, Qiusheng Wu1, Rui Pu1, Chao Zhou1, Bingru Huang1, Xingyun Peng1, Hans Ågren2 & Sailing He1,3,4

Stimulated emission depletion microscopy provides a powerful sub-diffraction imaging modality for life science studies. Conventionally, stimulated emission depletion requires a relatively high light intensity to obtain an adequate depletion efficiency through only light–matter interaction. Here we show efficient emission depletion for a class of lanthanide-doped upconversion nanoparticles with the assistance of interionic cross relaxation, which significantly lowers the laser intensity requirements of optical depletion. We demonstrate two-color super-resolution imaging using upconversion nanoparticles (resolution - 66 nm) with a single pair of excitation/depletion beams. In addition, we show super-resolution imaging of immunostained cytoskeleton structures of fixed cells (resolution - 82 nm) using upconversion nanoparticles. These achievements provide a new perspective for the development of photoswitchable luminescent probes and will broaden the applications of lanthanide-doped nanoparticles for sub-diffraction microscopic imaging.
Luminescent biomarkers, which include fluorescent dyes and proteins, semiconductor quantum dots, carbon dots, aggregation-induced emission molecules, and lanthanide-doped upconversion nanoparticles (UCNPs), offer powerful tools for optical imaging and life science studies since they make it possible to visualize events and structures that are otherwise invisible. While continuously expanding the toolbox of luminophores, researchers are keen on exploring their photophysical properties to improve the quality and scope of imaging modalities. Exploitation of optical switching to control the transition of luminophores between two optically distinguishable states (on/off) is an exciting and popular way to circumvent the diffraction limit and makes it possible to image at a spatial resolution of down to a few nanometers. A major category of these nanoscopy methods, including pulse STED and PALM/STORM, provides sub-diffraction resolution essentially by transiently switching the luminophores between the bright (on) and dark (off) states, causing those within the same diffraction range to emit successively, rather than simultaneously.

Lanthanide-doped UCNPs have been developed as an important group of luminescent biomarkers during the last decade. UCNPs convert low-intensity near-infrared (NIR) excitation light to shorter-wavelength NIR, visible, and ultraviolet (UV) emission. Such unique luminescence properties of UCNPs enable superior bioimaging without many of the constraints associated with conventional optical biomarkers, including photobleaching, photoblinking, tissue autofluorescence, limited imaging depth, and high light toxicity. However, efficient optical modulation of UCNPs has not been well established, which hinders their use in advanced luminescence imaging techniques, such as STED microscopy. In the few studies on optical modulation of UCNPs, highly efficient optical depletion of upconversion luminescence has not been reported in major groups of UCNPs (NaYF₄:Yb³⁺, Er³⁺/Tm³⁺/Ho⁺⁴⁻) that provide bright luminescence upon NIR excitation, except a demonstration of optical inhibition of upconversion luminescence in low-efficiency YAG:Pr³⁺ nanoparticles emitting toxic UV light under a complicated excitation scheme with limited applications in biomedical areas. The research efforts exploring co-irradiation by additional wavelengths have usually resulted in luminescence enhancement rather than silencing due to the presence of complicated photon recycling pathways among the lanthanide dopants under multi-wavelength excitation. In our previous study on another upconversion system (NaYF₄:Yb⁺³, Er⁺³), the first optical depletion of Er⁺³ emission was shown by inducing excited-state absorption (ESA), intrinsically different from the mechanism we propose in the present study, but the efficiency needed improvement for applications.

In this work, the cross relaxation between lanthanide ions provides an auxiliary mechanism for the optical depletion of Er⁺³ emission, allowing efficient optical modulation of UCNPs. The power-dependent depletion efficiency of the 455-nm emission from NaYF₄:18% Yb⁺³, 10% Tm⁺³ UCNPs was shown by inducing excited-state absorption (ESA) intrinsically different from the mechanism we propose in the present study, but the efficiency needed improvement for applications.

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pathway of upconversion luminescence. We establish an efficient optical depletion approach for the blue upconversion luminescence at 455 nm of Tm$^{3+}$ ions in high Tm$^{3+}$-doped NaYF$_4$:Yb$^{3+}$, Tm$^{3+}$ UCNP with enhanced inter-Tm$^{3+}$ interaction. The almost complete emission silencing is achieved by inducing stimulated emission from the emitting state through a second laser beam, while simultaneously suppressing the synergistic emission enhancement effect of the same laser beam by making full use of the concentration-dependent cross relaxation between the doped Tm$^{3+}$ ions. This novel emission depletion approach enables us to implement super-resolution luminescence microscopy. We further extend the super-resolution imaging to a two-color mode using the same pair of excitation/depletion beams. In addition, we implement cytoskeleton protein super-resolution imaging of immunolabeled HeLa cells using antibody-conjugated UCNP and demonstrate the ability of this established UCNP-mediated nanoscopy technique for imaging subcellular structures.

Fig. 2 Optical depletion mechanism of the 455-nm upconversion luminescence in NaYF$_4$:18% Yb$^{3+}$, 10% Tm$^{3+}$ nanoparticles. a Proposed optical emission depletion mechanism of the 455-nm upconversion band of Tm$^{3+}$ of NaYF$_4$:18% Yb$^{3+}$, 10% Tm$^{3+}$ nanoparticles. b Schematics of net stimulated depletion (left) and net synergistic enhancement (right) induced by 810-nm irradiation in high and low Tm$^{3+}$-doped UCNP, respectively. c Decay lifetimes of the 455-nm emission of NaYF$_4$:18% Yb$^{3+}$, 10% Tm$^{3+}$ UCNP at different power densities (Supplementary Table 1) of the depleting 810-nm laser. d The intensity change of different emission bands of NaYF$_4$:18% Yb$^{3+}$, 10% Tm$^{3+}$ UCNP under 975-nm excitation with/without 810-nm irradiation. The spectra were individually normalized in each spectral range, including 360–400, 570–615, 680–715, 980–1140, and 1300–2100 nm, separated by cutoff marks, in order to clearly display the relative intensity changes of different emission bands with and without the 810-nm irradiation. e The Tm$^{3+}$ doping concentration-dependent 1470-nm and 1800-nm downconversion emission spectra of NaYF$_4$:18% Yb$^{3+}$, x% Tm$^{3+}$ UCNPs ($\lambda_{\text{exc}} = 975$ nm). f Dependence of 455-nm depletion efficiency (DE, $\Delta I_{455}/I_{455}$) of NaYF$_4$:18% Yb$^{3+}$, 10% Tm$^{3+}$ on the depleting laser wavelength ($\Delta \lambda = 4$ nm) ranging from 700 to 815 nm. Negative depletion efficiency represents emission enhancement. The intensity of the 975-nm excitation laser and the depletion laser were kept at 700 kW cm$^{-2}$ and 17.7 MW cm$^{-2}$, respectively. g The excitation spectrum for the 455-nm emission of NaYF$_4$:18% Yb$^{3+}$, 10% Tm$^{3+}$ solely excited by the depletion laser with wavelength ranging from 700 to 815 nm; the inset is an enlarged view. Error bars represent ±1 s.d.
Results

Reversible optical depletion of blue emission of Tm-UCNPs. The hexagonal-phase NaYF₄ nanoparticles used in this study were synthesized using previously reported protocols with modifications (Methods)²⁸, ²⁹. Transmission electron microscope micrographs reveal a hexagonal shape of the as-synthesized high Tm³⁺-doped (NaYF₄:18% Yb³⁺, 10% Tm³⁺) nanoparticles with an average diameter of 18.0 (± 1.8) nm (Fig. 1a). When excited by a 975-nm continuous-wave (CW) laser (700 kW cm⁻²), these nanoparticles produce a dominant blue emission band at 455 nm together with several other relatively weak emission bands at 475 nm, 512 nm, 650 nm, and 700 nm (Fig. 1b, Supplementary Fig. 1), similar to the results in previous reports in high Tm³⁺-doped UCNPs under excitation of comparable intensity³⁰. These emission bands can be assigned to the transitions of 1D₂ → 3F₄ (455 nm), 1G₄ → 3H₆ (475 nm), 1D₂ → 3H₅ (512 nm), 1G₄ → 3F₄ (650 nm), and 3F₃ → 3H₄ (700 nm) of Tm³⁺ ions, respectively. Intriguingly, when co-irradiated by an 810-nm CW laser beam (Δλ = 4 nm, 17.7 MW cm⁻²), the most intense 455-nm blue emission is greatly quenched, with a depletion efficiency approaching 90% (Fig. 1b). The emission bands at 475 nm and 650 nm are also depleted, but less significantly (Fig. 1b), while the emission band at around 700 nm (3F₃ → 3H₄) is distinctly enhanced (Fig. 1b).

A concentrated UCNP dispersion was spin-coated onto a glass slide repeatedly to form a uniform film of nanoparticles and then imaged with a multiphoton laser scanning microscope (Supplementary Fig. 2). Figure 1c shows the multiphoton luminescence images of the UCNP film. Here the intensity of the CW 810-nm laser beam, I₈₁₀ was modulated by a chopper, which resulted in dark lines in the scanned image when the luminescence was periodically inhibited. This emphasizes the degree of optical control over the 455-nm upconversion luminescence as well as its reversible nature. The optical modulation of the 455-nm luminescence for dispersed nanoparticles on a slide was measured. As shown in Fig. 1d, compared to 975-nm single excitation, the 455-nm blue luminescence was almost completely depleted by the co-excitation with 975-nm and 810-nm laser light. Interestingly, the depletion efficiency of the 455-nm upconversion luminescence exhibits strong dependence on the Tm³⁺ concentration (Fig. 1e, f). With low Tm³⁺ doping (0.5%) (Supplementary Figs. 1 and 3), strong enhancement was observed for the 455-nm emission with the addition of the second laser at 810 nm (Fig. 1e). Emission depletion for the 455-nm band requires an adequately high Tm³⁺ concentration (Fig. 1f). For different-sized UCNPs with the same composition (NaYF₄:18% Yb³⁺, 10% Tm³⁺), all show efficient depletion of the 455-nm upconversion luminescence (λₑₓc = 975 nm) by the 810-nm laser beam, with slightly different saturation intensities (Supplementary Fig. 4). The degree of luminescence inhibition was measured for various values of I₈₁₀ (Fig. 1g). About 4% residual luminescence can be detected for I₈₁₀ = 17.7 MW cm⁻², confirming the effectiveness of the luminescence inhibition process. In our study, Iₛₘᵢ₉ was estimated to be 849 kW cm⁻² (power about 2.4 mW). It should be noted that optical depletion efficiency data of upconversion luminescence cannot be well fitted with the depletion equation I/Iₐₜ = 1/(1 + I₈₁₀/Iₛₘᵢ₉)²⁷, ³¹, which was derived from a simplified Jablonski diagram of downconversion fluorescence²⁸, ³⁲. The inapplicability of this equation to optical depletion of lanthanide upconversion emission is probably due to the fact that many complicated interionic energy transfer processes are not considered in the derivation. However, we still adopt the parameter—saturation intensity (Iₛₘᵢ₉) to characterize the emission depletion. The saturation power here is significantly lower than those used in CW-STEM microscopy using other biomarkers and those used in the depletion of other lanthanide ions, typically tens to hundreds of milliwatts¹¹. ³⁴-³⁷. Our saturation power is larger than that reported in a similar work about STED microscopy of UCNPs³¹, probably due to the difference in the nanoparticles. The 810-nm laser itself can only generate very weak upconversion emission (Fig. 1b). The background luminescence intensity generated by single 810-nm excitation (17.7 MW cm⁻²) is about 2% of that generated by single 975-nm excitation (700 kW cm⁻²) (Fig. 1b).

Emission depletion mechanism assisted with cross relaxation. Our spectroscopic results suggest an emission depletion mechanism involving stimulated emission supported by interionic cross relaxation that suppresses the synergistic enhancement effect of the second laser beam (810 nm). As illustrated in Fig. 2a, the 1D₂ emitting state, is populated by the 975-nm excitation beam through a five-photon upconversion process³⁸-⁴⁰ and can be depleted via a stimulated emission process induced by the 810-nm beam to populate the 3F₂ state, namely, 1D₂ → 3F₂ (810 nm). It should be noted that the 1D₂ → 3F₂ transition is very easy to be overlooked and was indeed often neglected in the literature. The 3F₂ and 3F₃ states were commonly treated as a single state (denoted as 3F₂,3) while ignoring their conspicuous energy difference since ions at the 3F₂ state quickly decays to the 3F₃ state nonradiatively, and the transition of 1D₂ → 3F₃,₂ was usually recognized to generate a single emission band centered at 740–750 nm.³⁰ Notably the 1D₂ → 3F₃ transition has an even larger branching ratio than the 1D₂ → 3F₂ transition according to previous theoretical studies⁴¹-⁴⁴. The 1D₂ → 3F₃ transition spectrally overlaps with other transitions, including 3H₄ → 3H₆ and 1G₄ → 3H₄, making it barely distinguishable from others and thus causing negligence. The matching of the STED laser light at 810 nm with the emission spectrum (red-shifted relative to the absorption spectrum) of the 1D₂ → 3F₃ transition is supported by many previous reports that indicate the absorption spectrum of 3F₂ → 1D₂ transition is centered around 800 nm and could cover quite a few nanometers.³⁹, 46-49. Subsequently, the ions at the 3F₂ state quickly decay to the lower-lying 3F₃ state and then to the 3H₄ state nonradiatively, which increases the population densities of these states.

The occurrence of the stimulated emission process is well supported by the enhanced emission originating from the 3F₂,₃ states at around 700 nm (3F₂ → 3H₄) and 1064 nm (3F₂ → 3F₄) after the addition of the 810-nm beam (Figs. 1b and 2d), indicating population increase of the 3F₂,₃ states. It is also supported by the observed decreased lifetime of the 455-nm luminescence with increasing the power of 810 nm beam in UCNPs with different Tm³⁺ concentrations (Fig. 2c, Supplementary Fig. 5, and Supplementary Table 1). Given that the emission cross sections of Tm³⁺ transition are typically in the range of 10⁻²¹ to 10⁻²⁰ cm² ⁴², ⁵⁰-⁵², the current depletion laser intensity yields a stimulated emission lifetime from the 1D₂ state, τᵣ, on the order of a few microseconds (Supplementary Fig. 5), which are three orders of magnitude longer than the stimulated emission lifetime in organic fluorescent dyes (ns vs. ps). The less dramatic lifetime change in some samples was probably attributable to the multiple effects of the depletion laser (discussed below), as well as the stepwise upconversion pumping approach in the lifetime measurement, where the measured decay lifetimes are often not solely determined by the lifetime of the emitting state but also reflect the radiative properties of many long-lived intermediate states.⁵⁵. In addition, in UCNPs with low doping (0.5%), where the 475-nm emission (three-photon process) was enhanced with the addition of the depletion laser at 810 nm, the 455-nm emission (five-photon process) was enhanced much less significantly (Fig. 1e). This
result also indicates that stimulated emission occurred for the $^{1}\text{D}_2$ state, because higher-order multiphoton emission from the $^{1}\text{D}_2$ state (455 nm) would be enhanced more significantly than the lower-order multiphoton emission from the $^{3}\text{G}_4$ state (475 nm) if stimulated emission did not occur for the $^{1}\text{D}_2$ state.

Along with the optical depletion through stimulated emission effect of the 810-nm laser beam, a synergistic enhancement effect by the same laser beam is also active due to the presence of a ground-state absorption (GSA) process matching 810 nm, $^{2}\text{H}_6 \rightarrow ^{2}\text{H}_4$, which effectively populates a key intermediate state $^{2}\text{H}_4$ of the upconversion process (Supplementary Fig. 6). The increase of the population density at the $^{3}\text{H}_4$ state, well supported by the observed enhanced emission at around 1470 nm originating from the transition $^{3}\text{H}_4 \rightarrow ^{3}\text{F}_2$ (Fig. 2d, Supplementary Fig. 7), is in favor of upconversion emission originating from higher states above the $^{3}\text{H}_4$ state (including the $^{3}\text{G}_4$ and $^{3}\text{D}_2$ states) under the co-irradiation of the 975-nm excitation beam, although single 810-nm excitation can only generate very weak upconversion luminescence (Fig. 1b, Supplementary Fig. 6). This synergistic enhancement effect competes with the STED effect, compromising the optical depletion efficiency induced by the 810-nm beam (Fig. 2b). A key cross relaxation (CR1) process between Tm$^{3+}$ ions could suppress the aforementioned emission enhancement effect of the 810-nm beam and facilitate the optical depletion, $^{3}\text{H}_4 + ^{3}\text{H}_4 \rightarrow ^{3}\text{F}_4 + ^{3}\text{F}_4$ through transferring electrons at the $^{3}\text{H}_4$ state to the $^{3}\text{F}_4$ state (Fig. 2a, b). The occurrence of this Tm$^{3+}$ concentration-dependent CR1 process is confirmed by the infrared emission measurements on nanoparticles with varied Tm$^{3+}$ concentrations under 975-nm excitation, where the emission originating from the $^{3}\text{F}_4$ to $^{3}\text{H}_4$ transition (1800 nm) is significantly enhanced, while that from the $^{3}\text{H}_4$ state (1470 nm) is significantly quenched as Tm$^{3+}$ concentration increases (Fig. 2e).

Our experimental results have shown that with the increase of the Tm$^{3+}$ doping concentration (from 0.5 to 10%), the ratio $I_{1470}/I_{1380}$ shows a very significant decrease (from about 5:1 to 1:32), indicating an enhancement factor of 160 for I$_{1380}$ with respect to I$_{1470}$ (Fig. 2e, Supplementary Fig. 8). The importance of this cross relaxation process in the blue luminescence inhibition process in our experiments is supported by the large enhancement of the infrared emission at 1800 nm upon the co-irradiation of the 810-nm beam in high Tm$^{3+}$-doped UCNPs (Fig. 2d), and also by the strong dependence of the depletion efficiency on the Tm$^{3+}$ concentration shown in Fig. 1f. These results suggest that the CR1 process can weaken the synergistic enhancement effect of the 810-nm beam and render the STED effect dominant in high Tm$^{3+}$-doped UCNPs. Otherwise, the absence of this CR1 process as in low Tm$^{3+}$-doped UCNPs makes the emission enhancement effect dominant (Fig. 2b, Supplementary Fig. 9).

Furthermore, it should be noted that the CR1 process can simultaneously enhance the STED effect of the 810-nm beam and thus lower the depletion laser intensity requirement. As shown in Fig. 2b, in the case of low Tm$^{3+}$ doping (without the CR1 process), the relaxing electrons from $^{1}\text{D}_2$ to $^{5}\text{F}_2$ via stimulated emission and then to $^{3}\text{H}_4$ would be readily re-pumped back to the $^{1}\text{D}_2$ state via the synergistic excitation of the two lasers. In contrast, in the case of high Tm$^{3+}$ doping the electrons at the $^{1}\text{D}_2$ state are first transferred to the $^{2}\text{H}_4$ state (via $^{5}\text{F}_2$) by the depletion laser, and then the CR1 process consecutively transfers the relaxing electrons from the $^{2}\text{H}_4$ state further to the $^{3}\text{F}_4$ state, where the CR1 process introduces a second energy-dissipating pathway $^{3}\text{F}_4 \rightarrow ^{3}\text{H}_4$. The cooperation of the STED effect of the 810-nm laser beam and the CR1 process makes the population center of gravity shift toward lower energy states and helps suppress the re-population rate of the emitting state under two laser irradiation, thus enhancing the stimulated emission-induced depletion efficacy of the depletion laser. This downward shift of the population center of gravity, yielding a new equilibrium among the population densities of different energy states, can also explain the passive depletion of the $^{3}\text{G}_4$ state (475-nm emission) just below the $^{1}\text{D}_2$ state at high Tm$^{3+}$ doping (Fig. 1b).

The slight decrease of the depletion efficiency induced by the 810-nm beam with the Tm$^{3+}$ doping concentration above 10% (Fig. 1f) could be due to the activation of the otherwise inefficient cross relaxations between Tm$^{3+}$ ions at adequately high Tm$^{3+}$ concentration, for example, $^{3}\text{G}_4 + ^{3}\text{F}_4 \rightarrow ^{1}\text{D}_2 + ^{3}\text{H}_6$ and $^{3}\text{F}_{2,3} + ^{3}\text{H}_4 \rightarrow ^{1}\text{D}_2 + ^{3}\text{H}_6$, as reported in a previous report$^{39}$, which enhances the synergistic excitation effect of the 810-nm laser beam. The critical role of the Tm$^{3+}$ concentration-dependent CR1 process is also supported by the results of our numerical simulations (Supplementary Figs. 10 and 11, Supplementary Table 2). The associated quenching of the emission from higher-lying states (370/383/395/595 nm) above the $^{1}\text{D}_2$ state$^{38}$ precludes depletion pathways through ESA to higher excited states (Fig. 2d, Supplementary Fig. 12).

The proposed optical depletion mechanism of the 455-nm emission for high Tm$^{3+}$-doped (NaYF$_4$:18% Yb$^{3+}$, 10% Tm$^{3+}$) nanoparticles is also well supported by the depletion-laser-wavelength-dependent depletion efficiency data, which were obtained by fixing the wavelength (975 nm) and power of the excitation laser and scanning the wavelength of the second beam from 700 nm to 810 nm while keeping the power constant, as shown in Fig. 2f. A depletion efficiency peak at around 810 nm (corresponding to the $^{1}\text{D}_2 \rightarrow ^{3}\text{F}_2$ transition) was observed. This broad depletion region is in consistence with the $^{1}\text{D}_2 \rightarrow ^{3}\text{F}_2$ emission spectrum$^{49, 54-56}$, indicating that this transition can have a rather broad emission band in various host materials due to spectrum-broadening effects. The existence of this efficient depletion peak is also consistent with the observed excitation spectrum for the 455-nm emission under sole irradiation of the second beam (Fig. 2g), where a second laser beam at around 810 nm gives rise to weaker luminescence background than other wavelengths. Interestingly, an extra depletion region centered at around 750 nm was observed (Fig. 2f), which is associated with the $^{1}\text{D}_2 \rightarrow ^{3}\text{F}_3$ transition$^{31}$. This reveals that the population of the $^{1}\text{D}_2$ state can also be efficiently depleted to the $^{3}\text{F}_3$ state via stimulated emission with laser irradiation of proper wavelength, analogous to our proposed stimulated emission pathway by an 810-nm laser associated with the $^{1}\text{D}_2 \rightarrow ^{3}\text{F}_2$ transition (Supplementary Fig. 13). The difference in the depletion efficiency at these two depletion peaks could be partially due to the different emission cross sections of the $^{1}\text{D}_2 \rightarrow ^{3}\text{F}_2$ and $^{1}\text{D}_2 \rightarrow ^{3}\text{F}_3$ transitions$^{41-44}$. In addition, laser light at around 750 nm could have a larger synergistic enhancement effect (cooperating with the 975-nm excitation light) on the 455-nm emission than the 810-nm light, due to its matching with other energy gaps, for example, the $^{3}\text{H}_4 \rightarrow ^{3}\text{G}_4$ transition$^{49, 54, 55}$, which would also degrade the depletion efficiency at this wavelength (Supplementary Fig. 13). Notable 455-nm emission enhancement was induced by addition of a second beam approaching 700 nm (Fig. 2f), which could be caused by the action of the $^{3}\text{H}_4 \rightarrow ^{3}\text{F}_3$ GSA process that increases the population of the $^{3}\text{F}_3$ state, in favor of multiphoton upconversion of Tm$^{3+}$ ions (Supplementary Fig. 13). In addition, the 455-nm emission enhancement observed under the co-irradiation of a laser beam around 765 nm and the 975-nm excitation laser (Fig. 2f) could be due to the matching of the former with the $^{3}\text{H}_4 \rightarrow ^{3}\text{G}_4$ ESA.
process\textsuperscript{31, 49, 55}, facilitating multiphoton upconversion luminescence by increasing the population of the $^{1}G_{4}$ state (Supplementary Fig. 13).

**Two-color optical nanoscopy with high photostability.** This efficient optical depletion mechanism enabled subsequent STED super-resolution microscopy using NaYF$_4$:18% Yb$^{3+}$, 10% Tm$^{3+}$ (18.0 ± 1.8 nm in diameter) UCNP\s with two NIR laser beams centered at 975 nm and 810 nm. The 975-nm laser beam with a Gaussian distribution intensity profile was the excitation beam, while the 810-nm laser of a donut-shaped intensity distribution provided the depletion beam. In comparison with the single 975-nm excitation, a significant improvement in the spatial resolution was achieved with the addition of the 810-nm depletion laser beam, as demonstrated in Fig. 3a, b. The line profile analysis for a selected area containing two single nanoparticles indicates that the lateral imaging resolution reaches about 66 nm using the excitation wavelength of 975 nm with the 810-nm depletion beam (Fig. 3c). The spatial resolution can likely be further improved by optimizing our lab-made super-resolution imaging system. The power densities for the two CW NIR beams were much lower than those used in typical CW-STED imaging\textsuperscript{11, 34–37, 57}, which is attractive for biological studies and technique commercialization. Another attractive advantage offered by this UCNP-mediated nanoscopy technique is the outstanding photostability of the UCNP\s. In the test of photostability, the region of interest was scanned 3000 times using both the 975-nm excitation and 810-nm depletion beam simultaneously, with a control region scanned using only the 975-nm beam. The results show that the UCNP\s did not show any noticeable photobleaching and photoblinking (Fig. 3d, Supplementary Fig. 14). Such an excellent photostability is highly desired for STED nanoscopic imaging, and the utilization of UCNP\s would enable long-time, continuous super-resolution imaging\textsuperscript{58}.

The proposed emission depletion mechanism can be exploited in multiple types of UCNP\s for multicolor super-resolution imaging using the same pair of laser beams. This can be achieved by incorporating other activators such as Tb$^{3+}$ and Eu$^{3+}$ into the Yb$^{3+}$-Tm$^{3+}$ system to generate luminescent nanoprobes emitting in complementary spectral channels, taking advantage of energy migration-mediated upconversion constructed on the population of higher excited states of Tm$^{3+}$ above the $^{1}D_{2}$ state
Fig. 4 Immunofluorescence labeling of cellular cytoskeleton protein desmin with antibody-conjugated UCNPs and super-resolution imaging. a The multiphoton imaging under 975-nm excitation of some cytoskeleton structures and desmin proteins in HeLa cancer cells incubated with anti-desmin primary antibody and immunostained with UCNPs (11.8 ± 2.2 nm in diameter) bioconjugated with goat anti-rabbit IgG secondary antibody. b The same region with a imaged in the super-resolution mode (975-nm excitation and the 810-nm STED laser beam). Scale bars: 2 μm. c–n Magnified areas selected from a and b (marked by white dotted squares) and line profile analyses; images in c, f, i, and l are taken from the white dotted squares in a. Images in d, g, j, and m are taken from the white dotted squares in b, e, h, k, n. Line profiles analyses of several areas indicated by arrow heads in c and d, f and g, i and j, and l and m, respectively.

As a proof of concept, a two-color super-resolution imaging was carried out using NaYF4:18% Yb3+, 10% Tm3+ nanoparticles (type 1) together with NaGdF4:40% Yb3+, 10% Tm3+@NaGdF4:15% Tb3+ nanoparticles (type 2). Type 2 nanoparticles, upon 975-nm excitation, produce bright green emission at around 547 nm of Tb3+, indirectly pumped via the 1I6 state of Tm3+ (Supplementary Fig. 16). As expected, the green emission of type 2 can be efficiently depleted using the 810-nm beam, here with efficiency of 88% (Supplementary Fig. 16). The mixture of both types of nanoparticles was spin-coated onto a glass coverslip for two-channel imaging. The target was first imaged in the 455-nm channel, which yielded images of both types of nanoparticles (Fig. 3e, h), since both types emitted in this blue channel. Subsequent imaging in the 547-nm channel resulted in images of solely type 2 nanoparticles (Fig. 3f, i), where type 1 nanoparticles were absent because they did not give rise to green emission under excitation of either laser beam or the combination of both. By comparing the images taken in the blue and green channels, type 1 nanoparticles can be completely distinguished (Fig. 3g, j). This super-resolution imaging strategy can be readily extended to even more spectral channels by introducing, for example, Eu3+ and Sm3+-activated nanoparticles, pumped through a high excited state of Tm3+. This strategy eliminates the need for one pair of excitation and depletion beams for each color channel, which currently constitutes a major obstacle for multicolor STED due to the challenge of aligning multiple laser beams. The pixel dwelling time (100 μs) employed in the laser scanning super-resolution imaging is comparable to that used in traditional STED imaging (50–200 μs) and is significantly shorter than that reported in a very recent work regarding nanoscopy using UCNPs (4–6 ms). The image acquisition speed can be increased with STED-parallelization strategies.

Immunofluorescence nanoscopy of cytoskeleton with UCNPs. In the recent decade lanthanide-doped UCNPs have been extensively developed to be an important group of biomarkers for various biological applications. However, there is a long-standing question of whether UCNPs can be used as an alternative fluorophore for subcellular fine structure imaging. In this study, we managed to image the cytoskeleton of HeLa cancer cells with UCNP-mediated nanoscopy. High Tm3+-doped UCNPs (NaGdF4:18% Yb3+, 10% Tm3+) with an average diameter of 11.8 (±2.2) nm were synthesized (Supplementary Fig. 4). After surface modification with poly(acrylic acid) (PAA) (Supplementary Fig. 17), these UCNPs were conjugated with the secondary antibody (goat anti-rabbit IgG antibody) (Supplementary Fig. 18). Fixed HeLa cancer cells were incubated with the secondary antibody and then stained with the secondary antibody IgG-conjugated UCNPs. Such immunostained HeLa cells were imaged in the super-resolution mode under 975-nm and 810-nm co-irradiation, and with a reference under irradiation of only the 975-nm laser. As shown in Fig. 4, subcellular structures were clearly imaged, and a significant improvement in the spatial resolution was
achieved with the assistance of the 810-nm depletion laser beam (Fig. 4a–n). To evaluate the resolution of STED imaging, line profiles of several areas were extracted and fitted with Gaussian functions. The results reveal that the lateral imaging resolution has been improved to about 82 nm, less than λ/10 (λ = 975 nm) (Fig. 4c–e). To the best of our knowledge, this is the first demonstration of successful immuno-labeling of fine subcellular structures as well as the first demonstration of super-resolution cell imaging using UCNPs.

**Discussion**

Optical depletion of upconversion luminescence has long remained a challenge, since the depletion laser wavelength easily matches with multiple energy gaps, complicating the intended role of the depletion laser with both emission enhancement and depletion effects active. In this work, we utilize ion-ion interaction to suppress the enhancement effect and enhance the depletion effect of the depletion laser beam, and achieve large optical depletion of upconversion luminescence in a major group of UCNPs. Specifically, we have successfully achieved 96% depletion of the blue emission at 455 nm (λ_{exc} = 975 nm) in NaYF₄:18% Yb³⁺, 10% Tm³⁺ nanoparticles with an 810-nm beam. The strong cross-relaxation between low-lying states of the emitters (Tm³⁺), H₂ + H₂ → F₂ + F₂, plays a critical role in the optical inhibition of the 455-nm emission. We have successfully implemented STED super-resolution imaging using these UCNPs. By adopting nanoparticles emitting in other color channels, which were indirectly pumped via a high excited state of Tm³⁺ through energy migration upconversion, we have implemented two-color super-resolution imaging using a single excitation/depletion laser beam pair. In addition, we have achieved a successful immuno-labeling of fine subcellular structures and super-resolution cellular imaging using UCNPs. The upconversion emission depletion nanoscopy developed here significantly reduces the requirement of the depletion power density, as also demonstrated in a very recent similar work [31], and removes the need for bulky and expensive pulse lasers and makes STED microscopy more biocompatible, providing alternatives to traditional STED imaging using organic dyes and other luminescent nanoparticles. We believe that our work will provide new insight into developing optical modulation methods for luminophores toward broad applications in super-resolution imaging and beyond and will significantly extend the range of applications of UCNPs in nanoscale photonics and life science.

**Methods**

**Optical setup for spectroscopic and microscopic study.** A lab-made optical system was built to perform spectroscopic measurement and super-resolution imaging, as shown in Supplementary Fig. 2. The 975-nm laser beam was generated by a CW, single-mode diode laser (Laser 1, B&A Technology Co. Ltd., Shanghai). A band-pass filter (F2, LD01-975/10-25, Semrock) was used to purify the laser spectrum. The 810-nm CW laser beam was provided by a Ti:sapphire laser (Laser 2, Mira 900, Coherent) and filtered with a band-pass filter (F1, FF01-810-10-25, Semrock). A pinhole in combination with two lenses was used as a spatial filter to optimize the beam profile. Two pairs of half-wave plate and polarization beam splitter were placed in the optical path to control the laser power. These two NIR laser beams were then spatially overlapped using a 950-nm short-pass dichroic mirror (DM1, ZET950spxrt, Chroma), and then directed into a multiphoton laser scanning microscope. In the super-resolution imaging, a vortex phase plate (VPP-1a, RPC Photonics) was employed to perform a 2π helix phase modulation of the 810-nm beam, generating a donut-shaped beam. Another half-wave plate (F3) combined with a quarter-wave plate attached to the objective lens was used to match the 810-nm laser beam in circular polarization. The coaxial laser beams were reflected by a 775-nm short-pass dichroic mirror (DM2, ZET775sp-2p-UF3, Chroma), and focused on the sample using an oil-immersed objective (OL1, PL- APO ×60/1.35, Olympus). The visible emission (400–755 nm) was collected through the same objective and filtered by a 775-nm short-pass filter (F3, FF01-775SP-25, Semrock) and then captured by a spectrometer (QE65000, Ocean Optics) or a photomultiplier tube (PMT) for spectroscopic or imaging studies. The NIR emission was collected by another objective (OL2, ×20/0.5, Olympus) set on the condenser holder above the sample and filtered by a 980-nm long-pass filter (F6, BLP01-980R-25, Semrock), detected by two NIR spectrometers: NIRQuest, Ocean Optics (spectral range: 1100–2100 nm); and Shamrock 303i-B, iDus Spectroscopy Cameras, Andor (spectral range: 800–1500 nm).

**Luminescence lifetime measurement.** Emission lifetime measurements of UCNPs were performed by modulating the 975-nm excitation laser with a chopper (model SR540, Stanford) and recording the time-resolved luminescence intensity. The emission photons, passing through a 448-nm band-pass filter (F4, ET448/×19, Chroma), were detected by a PMT, and the trigger signal from the chopper was synchronized with a time-correlated single-photon counter (NanoHarp, Picocount). The chopper modulation frequencies were varied with the PMT’s doping concentrations, 480, 530, 796, 890, 1000, 1276, 2000, and 3500 Hz for UCNP’s doped with 0.5% Tm³⁺, 2% Tm³⁺, 5% Tm³⁺, 7% Tm³⁺, 10% Tm³⁺, 15% Tm³⁺, and 20% Tm³⁺, respectively. To minimize the influence of the instrument response function, for example, the falling edge of the mechanically modulated light pulse, the beam was suitably focused and the chopper was placed in the focal plane. In addition, iterative deconvolution was performed with the instrument response function, measured with a rhodamine sample with a lifetime on order of nanoseconds.

**Dual-color super-resolution imaging.** In the dual-color super-resolution microscopic imaging, a 495-nm long-pass dichroic mirror (DM3, FF495-Di03-25x 36, Semrock) was used to separate the luminescence into two detectors. The 448-nm beam was placed in front of PMT to control the PMT’s emission signal from NaYF₄:18% Yb³⁺ +10% Tm³⁺ and NaGdF₄:40% Yb³⁺ +10% Tm³⁺ +NaGdF₄:15% Tb³⁺, and a 550-nm band-pass filter (F5, ET550/×20, Chroma) was placed in front of PMT2 to collect the 547-nm emission signal only from NaGdF₄:40% Yb³⁺ +10% Tm³⁺ +NaGdF₄:15% Tb³⁺. Image subtraction using the data from the two channels allowed for separate imaging of the two types of UCNP’s, enabling dual-color super-resolution imaging.

**Synthesis of Tm³⁺-NaYF₄ UCNPs.** The designed UCNP’s were synthesized following previously reported protocols with some modifications [28, 29]. In a typical synthesis, 5 mL Ln(CH₂CO₂)₃ (Ln = Y/Yb/Tm) stock solution (0.2 M) was added into a 100-mL round bottom flask, followed by the addition of 7.5 mL oleic acid (OA) and 17.5 mL 1-octadecene (ODE). The mixture was heated to 150 °C under stirring for 40 min to form the lanthanide-oleate precursor solution, and then cooled down to room temperature. Then 10 mL of NH₂F-methanol solution (0.4 M) and 2.5 mL of NaOH-methanol solution (1 M) were pipetted into a 15-mL centrifuge tube, and then the mixture solution was quickly injected into the flask. Subsequently, the mixture was heated to 50 °C and kept at that temperature for at least 0.5 h, and then was heated to 110 °C under vacuum to remove methanol. After methanol was evaporated, the mixture was heated to 300 °C and incubated at that temperature for 1.5 h under an argon atmosphere, and then cooled down to room temperature. The UCNP’s were precipitated by the addition of ODE, followed by centrifugation at 7500 rpm for 10 min. The supernatant was removed, and the UCNP’s were washed several times using ethanol and cyclohexane, and were finally re-dispersed into cyclohexane for subsequent use. Besides, the NaYF₄:18% Yb³⁺ +10% Tm³⁺ UCNP’s with an average diameter of 49.5±1.6 nm (Supplementary Fig. 4) were synthesized with minor modification, and the reaction was isothermally kept at 320 °C for 2 h.

**Synthesis of core-shell Tm³⁺-NaGdF₄ UCNPs.** The fabrication of the core NaGdF₄:40% Yb³⁺ +10% Tm³⁺ UCNP’s with an average diameter of 15.7±1.9 nm used the same general synthesis protocol with modifications. In the synthesis, 5 mL Ln(CH₂CO₂)₃ (Ln = Gd/Y/Yb/Tm) stock solution (0.2 M) was added into a 100-mL round bottom flask, followed by the addition of 10 mL OA and 15 mL ODE. The mixture was heated to 150 °C under stirring for 40 min to form the lanthanide-oleate precursor solution, and then cooled down to room temperature. The UCNP’s were precipitated by the addition of OA, followed by centrifugation at 7500 rpm for 5 min. The obtained UCNP’s were washed several times using ethanol and cyclohexane, and were finally re-dispersed into cyclohexane for subsequent use. Besides, the NaYF₄:18% Yb³⁺ +10% Tm³⁺ UCNP’s with an average diameter of 49.5±1.6 nm (Supplementary Fig. 4) were synthesized with minor modification, and the reaction was isothermally kept at 320 °C for 2 h.

The visible emission (400–755 nm) was collected through the same objective and filtered by a 775-nm short-pass filter (F3, FF01-775SP-25, Semrock) and then captured by a spectrometer (QE65000, Ocean Optics) or a photomultiplier tube (PMT) for spectroscopic or imaging studies. The NIR emission was collected by another objective (OL2, ×20/0.5, Olympus) set on the condenser holder above the sample and filtered by a 980-nm long-pass filter (F6, BLP01-980R-25, Semrock), detected by two NIR spectrometers: NIRQuest, Ocean Optics (spectral range: 1100–2100 nm); and Shamrock 303i-B, iDus Spectroscopy Cameras, Andor (spectral range: 800–1500 nm).
mixing 2.5 mL Mn(CH₃CO₂)₂ (Lₓ = 10⁻⁷ / L / Tm³⁺) stock solution with 5 mL of OA and 7.5 mL of ODE in a 100-mL flask. The mixture was heated to 150 °C under stirring for 40 min to form a homogeneous solution, and then cooled down to room temperature. Then, 4 mL of as-prepared core nanoparticle suspension was injected into the reaction flask, followed by the addition of 5 mL NH₄F-methanol stock solution and 1.25 mL NaOH-methanol stock solution. Subsequent steps were the same as in the synthesis of the NaGdF₄:40% Yb³⁺/10% Tm³⁺ UCNPs.

Preparation of hydrophilic Tm³⁺-UCNPs. Two methods were used to prepare hydrophilic Tm-UCNPs. One is the modified method of ligand removal with hydrochloric acid (HCl)64, 65. In a typical experiment, 60 mg dried OA-UCNPs were redispersed into 3 mL 1% HCl solution, and then was sonicated for 1 h. The desorbed OA and naked UCNPs were separated by extraction with ether, followed by centrifugation at 13,000 r.p.m. for 30 min. The supernatant was discarded and the obtained pellets were dispersed into 1 mL deionized water by sonication. Another hydrophilic processing method is ligand exchange to replace the oleic acid on the nanoparticle surface with PAA66, 68. In a typical experiment, 30 mL diethyleneglycol and 0.5 g PAA were added to a 100-mL bottom flask. The mixture was heated to 110 °C with vigorous stirring under an argon atmosphere for 1 h. Then 4 mL OA-UCNPs (ca. 60 mg) was slowly injected into the hot solution and the mixture was kept under vacuum for 30 min to evaporate cyclohexane. Subsequently, the mixture was heated to 240 °C and kept for 3 h under argon protection until the solution became clear. Then the mixture was cooled down to room temperature. After adding 15 mL 1% HCl aqueous solution and stirring for 15 min, a transparent gelatinous system was obtained. The UCNPs were precipitated by centrifugation at 15,000 r.p.m. for 30 min, followed by washing with ethanol and deionized water (v/w = 1:1) for several times. Finally, the products were re-dispersed in 4 mL fresh deionized water to form a clear solution for subsequent use.

Upconversion luminescence immunolabeling of cytoskeleton. Activation of the carboxyl groups on the surface of UCNPs were carried out according to the commonly used methods with modifications67, 68. First, PAA-UCNP aqueous solution (1 mg mL⁻¹) was prepared, and N-(3-dimethylaminopropyl)-N-ethylcarbodiimide hydrochloride (EDC) and N-hydroxysuccinimide (NHS) solids were dissolved in 2-(N-morpholino)ethanesulfonic solution at concentrations of 0.2 and 0.3 mg mL⁻¹, respectively. After adding 20 μL NHS and 20 μL EDC solution into 1 mL PAA-UCNPs aqueous solution, the mixture was stirred for 2 h at room temperature. The UCNPs were obtained via centrifugation at 15,000 r.p.m. for 30 min and then re-dispersed in 1 mL ultrapure water. The pH value of the solution was adjusted to 7.2–7.5 to ensure an efficient reaction between the antibody and the NHS-activated UCNPs. After the addition of 500 μg goat anti-rabbit antibody IgG (dissolved in antibody diluent), the mixture was stored in a refrigerator at 4 °C overnight, followed by centrifugation at 14,000 r.p.m. for 20 min. The obtained precipitate was dispersed into phosphate-buffered saline (PBS) buffer for subsequent use.

A commonly used immunostaining technique was used to perform cytoskeleton labeling using UCNPs67, 68. The HeLa cells were first cultured in 96-well plates with approximately 15,000 cells in each unit and then were rinsed three times using PBS buffer to remove culture media. Subsequently, the cells were fixed and rinsed using 4% paraformaldehyde (PFA) for 15 min at room temperature. Next, the fixed cells were rinsed three times again to remove the residual PFA. The HeLa cells were permeabilized in 0.2% Triton X-100 for 20 min at room temperature and were rinsed again using PBS. Quick Block blocking buffer was then used to block the cells for 20 min at room temperature. The antibody dilutions were prepared using the Quick Block primary antibody dilution buffer. The primary antibody (anti-desmin monoclonal antibody) solution of 5 μg mL⁻¹ was used to incubate the fixed cells at 3 °C overnight. Then, the incubated sample was rinsed three times using PBS buffer. Then, the as-prepared UCNPs biocomjugated with secondary antibodies were dissolved in PBS buffer after centrifugation. A volume of 100 μL UCNPs solution (200 μg mL⁻¹) was added to 96-well plates with the primary antibody-treated fixed cells. The immunostaining reaction of cytoskeleton desmin protein was kept at room temperature for 2 h. The cells were then rinsed gently with pre-warmed (37 °C) PBS buffer twice. Then, the HeLa cells were imaged using the multiphoton laser scanning microscope. The super-resolution imaging was performed with a pixel dwelling time of 100 μs.

Theoretical modeling and numerical simulation. The proposed optical depletion mechanism of the Tm³⁺ emission in NaF₅₋₄Yb³⁺/Tm³⁺ nanocrystals is shown in Supplementary Fig. 10. The upconversion process in Yb³⁺:Tm³⁺ co-doped system can be described by the rate equations of optical direct excitation/depletion and interionic energy transfer. According to the proposed energy transfer upconversion process shown in Supplementary Fig. 10, the rate equations of each energy state are derived as follows:

\[ \frac{d n_i}{d t} = \frac{-n_i}{\tau_i} + \frac{n_{i+1}}{\tau_i} + \frac{n_{i-1}}{\tau_i} + \frac{n_{i0}}{\tau_i} + \frac{n_{i+1}}{\tau_i} + \frac{n_{i-1}}{\tau_i} + \frac{n_{i0}}{\tau_i} \]

Here, \( n_i \) (\( i = 0–8 \)) and \( n_0 \) (\( i = 1–8 \)) represent the population densities and radiative lifetimes of the \( \text{H}_0 \), \( \text{F}_0 \), \( \text{H}_1 \), \( \text{F}_1 \), \( \text{F}_2 \), \( \text{G}_0 \), \( \text{D}_2 \), and \( \text{I}_0 \) states of Tm³⁺ ions, respectively. \( n_0 \) (\( i = 1–8 \)) is the energy transfer rate from the \( \text{F}_0 \) state of Yb³⁺ to the \( \text{H}_0 \), \( \text{F}_0 \), \( \text{H}_1 \), \( \text{F}_1 \), \( \text{F}_2 \), \( \text{G}_0 \), \( \text{D}_2 \), and \( \text{I}_0 \) states of Tm³⁺, respectively. \( n_{i0} \) and \( n_{i0} \) are the cross sections for the absorption processes \( \text{H}_0 \to \text{F}_0 \), \( \text{F}_0 \to \text{F}_1 \), \( \text{F}_1 \to \text{F}_2 \), \( \text{F}_2 \to \text{G}_0 \) of Tm³⁺, respectively. \( n_{i0} \) and \( n_{i0} \) are the cross sections for the stimulated emission processes \( \text{H}_0 \to \text{F}_0 \), \( \text{F}_0 \to \text{F}_1 \), \( \text{F}_1 \to \text{F}_2 \), \( \text{F}_2 \to \text{G}_0 \) of Tm³⁺.
Due to the matching of the 810-nm laser (full width at half maximum of 4 nm) with multiple transitions, including $^{3}H_{6}$ to $^{1}D_{2}$, $^{3}H_{4}$ to $^{1}G_{4}$, and $^{3}F_{4}$ to $^{1}D_{2}$, three light absorption and three stimulated emission processes at 810 nm are all considered in this model, with different cross sections (Supplementary Table 2). Regarding the net effect of the 810-nm laser on the population density of the $^{1}H_{6}$ state, fundamentally determined by the magnitude relation between two terms, namely, $\frac{g_{3}I_{3}}{\tau_{3,4}+\tau_{1,2}}$ and $\frac{g_{2}I_{2}}{\tau_{2,3}+\tau_{1,2}}$, our spectroscopic result of the 1470-nm band ($^{3}H_{4}$ to $^{3}F_{4}$) (Fig. 2d) indicates that the GSA process $^{3}H_{4}$ to $^{1}D_{2}$ is dominant over the stimulated emission process $^{3}H_{4}$ to $^{1}H_{6}$. In addition, the stimulated emission process $^{3}D_{2}$ to $^{3}F_{2}$ is supposed to be dominant over the ESA process $^{3}F_{2}$ to $^{3}D_{2}$, due to a larger population density at the $^{3}D_{2}$ state than at the $^{3}F_{2}$ state and a larger emission cross section than the absorption cross section at this wavelength. For simplicity, only the net effects of the 810-nm laser on the $^{3}H_{4}$ to $^{1}H_{6}$, $^{3}H_{4}$ to $^{1}G_{4}$, and $^{3}F_{4}$ to $^{1}D_{2}$ transitions are illustrated in the mechanism diagram (Fig. 2a). The $T_{1}$ concentration dependence of the depletion efficiency of the 455-nm emission was investigated by varying the parameter values, particularly those for cross-relaxation processes. The values used for the main parameters for both low and high $T_{1}$-doped samples are tabulated in Supplementary Table 2.

Data availability

All the relevant data that support the findings of this work are available from the corresponding authors upon reasonable request.

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Author contributions

Q.Z. conceived and designed this project. Q.Z. directed the experiments with contribution from H.L. B.W., Q.W., and X.P. acquired and processed data. B.W., Q.W., and X.P. were responsible for synthesis, surface functionalization, and characterization of nanoparticles. C.Z. and Q.Z. prepared samples. Q.W., Q.Z., and X.P. were responsible for the theoretical analysis and simulation with contribution from H.Å. Q.Z. and Q.W. built the optical system. B.W., Q.W., Q.Z., and X.P. were responsible for the theoretical analysis and simulation with contribution from H.Å. and S.H. Q.Z. and S.H. supervised the project. H.L., Q.Z., and S.H. wrote the paper with contribution from all authors.

Additional information

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