Post-COVID-19 arthritis: a case report and literature review

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Abstract
Severe acute respiratory syndrome coronavirus 2 (SARS-CoV2) is the novel pathogen responsible for the coronavirus disease 19 (COVID-19) outbreak. Researchers and clinicians are exploring the pathogenetic mechanisms of the viral-induced damage and growing interest is focusing on the short-term and long-term immune-mediated consequences triggered by the infection. We will focus on post-SARS-CoV2 infection arthritis which may arise as a new pathological condition associated with COVID-19. In this article, we describe a case of acute oligoarthritis occurring 13 days after a SARS-CoV2 severe pneumonia in a middle-aged Caucasian man and we go over a brief review of the current available literature. We hypothesize that molecular mimicry might be the basic immunological mechanism responsible for the onset of COVID-19-related arthritis based on the current knowledge of SARS-CoV2 and on the known pathogenetic mechanism of viral-induced arthritis.

Keywords  Arthritis · COVID-19 · Molecular mimicry · Reactive arthritis · SARS-CoV2

Introduction
In the midst of COVID-19 outbreak, researchers from all over the world are studying the pathogenetic mechanisms of SARS-CoV2 respiratory infection but growing interest is also focusing on the immune-mediated consequences that could be secondarily triggered by the virus. The most severe cases are characterized by a marked pro-coagulant state [1, 2] and by an inflammatory cytokine storm similar to that found in macrophage activation syndrome [3–5]. A dysregulated hyperimmune response definitely contributes to the severity of damage and may elicit autoimmune processes in predisposed individuals. Viral infections are supposed to be involved in the pathogenesis of many rheumatological conditions and several cases of autoimmune-induced diseases after SARS-CoV2 infection are reported in the literature [6–9].

Case report description
We describe a case of acute arthritis after a SARS-CoV2 infection in a 60-year-old Caucasian man without relevant comorbidities (Table 1). In April 2020, he was hospitalized for hyperpyrexia, headache, asthenia, and a worsening dyspnea. At the emergency room (ER), the thoracic ultrasound and chest X-ray revealed an interstitial pneumonia; a nasopharyngeal swab was positive for SARS-CoV2 and his blood test revealed a marked inflammatory state characterized by CRP (C-reactive protein) 240 mg/L, fibrinogen 9.83 g/L, interleukin-6 162 ng/L, ferritin 944 μg/L, and D-dimer 993 μg/L. He was admitted to the Internal Medicine department and treated with azithromycin, ceftriaxone, hydroxychloroquine (HCQ) (400 mg/die), anticoagulation for thromboembolism prophylaxis, and low-flow oxygen. For progressive respiratory failure, he was referred to the intensive care unit where he underwent nasotracheal intubation and received broad-spectrum antibiotics (meropenem, linezolid), antimycotic prophylaxis, continuous diuretic infusion, noradrenalin for hemodynamic support, and therapeutic dose of anticoagulants for elevation in D-dimer values. Due to a progressive improvement of respiratory gas exchange and chest X-ray, he was extubated after 10 days. He was discharged in good general conditions and low-grade inflammation on blood tests after overall 19 days of hospitalization. The weekly surveillance nasopharyngeal swabs for SARS-CoV2 persisted negative. Nevertheless, 13 days after
discharge, he complained tenderness of the right ankle, knee, and hip in association with low-grade fever. He presented to the ER with oligoarthritis of the right lower limb and high CRP level (237 mg/L) on blood tests. Physical and ultrasound examination confirmed slight right ankle inflammation and clear right knee arthritis. Arthrocentesis led to the evacuation of 20 cc of a cloudy, yellow, and highly inflammatory synovial fluid (SF) (Fig. 1) whose analysis revealed 20,000/mm$^3$ white blood cells of which 90% polymorphonucleates and 10% monocytes; no crystals were detected. Synovial RT-PCR (real-time polymerase chain reaction) for SARS-CoV2, as well as SF culture for bacterial agents, was negative (Table 2). He denied any infectious symptom, recent history of physical trauma, dyspnea, previous episode of arthritis, dactylitis, conjunctivitis or uveitis nor inflammatory diarrhea, and personal or familial history of psoriasis. To proceed with further investigations, the patient was hospitalized again. A new nasopharyngeal swab and a negative research of SARS-CoV2 nucleic acid on sputum excluded a recurrence of the systemic infection while serology showed indisputable seroconversion (a-SARS-CoV2 IgG 37.120 KUA/L and a-SARS-CoV2 IgM 9.163 KUA/L). Urine and blood cultures were negative and procalcitonin within the normal range; urethral swab and stool culture did not show evidence of bacterial infection. The in-depth examination for systemic rheumatic causes of arthritis were negative including antinuclear antibodies (ANA), extractable antinuclear antibodies, rheumatoid factor (RF), anti-citrullinated peptide, and HLA-B27 typing (Table 3). The knees, ankles, and hip X-ray did not show erosions or intra-articular calcifications (Fig. 2). Even though the patient’s SF was markedly inflammatory, which is an infrequent finding in infectious-related arthritis, the temporal relation with SARS-CoV2 infection made the hypothesis of post-viral acute arthritis the most probable. A nonsteroidal anti-inflammatory (NSAID) therapy with ibuprofen 600-mg; bid was started with clinical benefit and decrease of CRP. The patient was discharged after 9 days and continued the NSAIDs for other 3 weeks. Up to 6 months after therapy discontinuation, he presented no signs of arthritis recurrence.

### Table 1 Laboratory findings during clinical course in our patient

|                      | Day of discharge for COVID-19 hospitalization | Day of hospitalization for oligoarthritis arthritis | Day of discharge |
|----------------------|---------------------------------------------|-----------------------------------------------------|------------------|
| WBCs (4400–11,000/mmc)| 6490                                        | 9020                                                | 5510             |
| Neutrophils (1.800–7.800/mmc) | 2750                              | 5750                                                | 2990             |
| Lymphocytes (1.100–4800/mmc) | 2080                                | 990                                                 | 1760             |
| Monocytes (200–960/mmc)       | 910                                       | 630                                                 | 560              |
| Eosinophils (0–500/mmc)         | 690                                       | 220                                                 | 190              |
| Basophils (0–200 /mmc)            | 6                                         | 10                                                  | 4                |
| Hb (14–17.5 g/dL)            | 11.8                                      | 11.8                                                | 11.7             |
| PLTs (150,000–410,000/mmc)    | 380,000                                   | 330,000                                             | 388,000          |
| ESR (2–37 mm/h)              | -                                         | 111                                                 | 72               |
| CRP (0–6 mg/L)               | 12                                        | 237                                                 | 14               |
| D-dimer (0–300 μg/L)         | 1490                                      | 839                                                 | -                |
| Fibrinogen (1.5–4.5 g/L)     | 7.66                                      | 10.61                                               | -                |
| LAD (135–225 U/L)            | 260                                       | 213                                                 | 167              |
| Ferritin (20–250 μg/L)       | 700                                       | 446                                                 | 248              |
| Procalcitonin (0–0.5 μg/L)   | < 0.04                                    | < 0.04                                               | -                |
| SARS-CoV2 swab               | Negative                                   | Negative                                             | Negative         |

WBC white blood cell, Hb hemoglobin, PLT platelet, ESR erythrocyte sedimentation rate, CRP C-reactive protein, LAD lactate dehydrogenase, SARS-CoV2 severe acute respiratory syndrome coronavirus 2

Fig. 1 Patient’s synovial fluid
Discussion and review of the literature

The pathogenesis of viral associated arthritis is only partially understood but one of the mechanisms supposed to mediate the activation of the inflammatory process is molecular mimicry [16], well known to be responsible for eliciting autoimmune responses in predisposed individuals [17, 18]. Examples of molecular mimicry concerning SARS-CoV2 are reported [18] and this mechanism is hypothetically involved in the pathogenesis of both the acute systemic infection and the post-infective viral-related immunological consequences [19, 20]. Previous and actual studies demonstrate that coronaviruses share molecular epitopes with human proteins (e.g., spike glycoprotein S) that play a key role to host cell invasion and escape immune response attacks, giving to the infectious agent an immune-evasive capacity [21, 22]. SARS-CoV2 shares three sequences of six amino acids with as many brainstem human proteins and the cross-reaction between human and viral epitopes may lead to brainstem damage and respiratory failure [23]. Other suggestive examples of molecular mimicry-driven diseases after COVID-19 come from recent publications reporting cases of Guillain-Barre and Miller-Fisher syndrome [24, 25]. Mimicking epitopes may also be present in synovial membrane and cause, with similar mechanism, an acute local inflammation.

To study in depth the relationship between SARS-CoV2 infection and post-COVID-19 arthritis, we performed a comprehensive search on PubMed of all the reported cases of acute arthritis in patients with SARS-CoV2 infection from January 2020 to October 2020 combining the following keywords: acute arthritis, reactive arthritis, viral arthritis, COVID-19, coronavirus, and SARS-CoV2. We considered only English-written case reports of adult patients. Thirteen articles met our searching inclusion criteria; four were excluded because they are not pertinent with the purpose of our review, one because it was a correspondence letter to an already considered article with no case report included in the text, and two because of describing cases of drug-induced gouty arthritis or patients with previous history of gout and therefore not strictly categorizable in among the group of post-COVID-19 arthritis. The six articles which satisfied inclusion and exclusion criteria refer to 6 case reports [10–15] which are included in Table 3.

The low prevalence of this clinical condition we found in patients with COVID-19 could be due to the use of HCQ and corticosteroids for the treatment of the viral infection which may prevent or weaken the inflammatory joint manifestations. Despite HCQ has not demonstrated to be effective in the treatment of COVID-19 [26], it has proven efficacy in the management of systemic rheumatological diseases, especially with inflammatory joint involvement. HCQ is the anchor drug in systemic lupus erythematosus, acting as immunomodulator, and prevents or mitigates lupus clinical manifestation in autoantibody positive asymptomatic subjects [27]; it is also part of the treatment for the milder forms of rheumatoid arthritis and Sjogren syndrome with frequent episodes of joint pain [28].

Analyzing more specifically the six cases of suspected COVID-19-related arthritis, it is apparent how the SF analysis was not performed in three cases [13–15]; therefore a microcrystalline etiology cannot be certainly ruled out. In the remaining three cases [10–12], monosodium urate and calcium pyrophosphate crystals were not detected at polarized light microscope examination, thus configuring an essential step forward in the exclusion diagnostic process. No other chemical-physical characteristics, including the number of white blood cells and their differential count, have been reported. As shown in Table 3, the lag time between SARS-CoV2 infection and onset of arthritis is variable but joint symptoms generally present days after the acute viral infection and usually during the healing period. Clinical and epidemiological data show a prominent involvement of lower limb joints with mono- or oligoarticular symptoms and a predilection for male sex. The clinical presentation that emerges from these case reports may deviate from the classic picture of a viral-related arthritis where joint involvement usually occurs during the viremia period and presents with a polyarticular pattern sometimes resembling rheumatoid arthritis [29].

### Table 2 Synovial fluid analysis

| Aspect       | Yellow turbid                        |
|--------------|--------------------------------------|
| Differential count of WBC | WBC 20,000/mmnc, PMN 90%, M 10% |
| Polarized light microscopy | No crystals                          |
| SARS-CoV2 RT-PCR | Negative                             |
| Culture      | Negative                             |

WBC white blood cell, PMN polymorphonucleate, M monocyte, RT-PCR real-time polymerase chain reaction

Fig. 2 Knees X-ray

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| Age and sex       | Yokagawa et al. [10] | Liew et al. [11] | Ono et al. [12] | Saricaoglu et al. [13] | Danssaert et al. [14] | Parisi et al. [15] | Present case |
|------------------|----------------------|------------------|------------------|------------------------|----------------------|-------------------|--------------|
| Time to arthritis| 57-year-old man      | 47-year-old man  | Male in his 50s  | 73-year-old man        | 37-year-old female   | 58-year-old female| 60-year-old man|
| COVID19 diagnosis| 15 days after        | At diagnosis of  | 21 days after    | 15 days after          | 12 days after        | 25 days after     | 32 days after  |
|                  | COVID19 diagnosis    | COVID19          | COVID19 diagnosis| COVID19 diagnosis      | COVID19 diagnosis    | COVID19 diagnosis| COVID19 diagnosis|
| Affected joints  | Right knee           | Right knee       | Left and right   | Left I MTP, PIP, DIP   | Tendonitis of the II, III and IV extensor of the right hand| Ankle             | Right knee and ankle|
|                  |                      |                  | ankle            | and DIP                |                      |                   |              |
| Therapy          | No therapy (self-recovery) | Oral NSAID and intra-articular corticosteroid | Oral NSAID and intra-articular corticosteroid | Oral NSAID | Topical NSAID, oral opioid, gabapentin | Oral NSAID | Oral NSAID |
| SF polarised microscopic examination | No crystals | No crystals | No crystals | - | - | - | No crystals |
| SF culture       | -                     | (negative Gram stain) | Negative | - | - | - | Negative |
| SF PCR for SARS-CoV2 | Negative | Negative | - | - | - | - | Negative |
| M. pneumoniae and C. pneumoniae serology | - | - | Negative | - | - | - | Negative |
| Gonococcus       | Negative             | Negative         | Negative         | - | - | - | - |
| C. trachomatis   | PCR/serology         | PCR/serology     | PCR/serology     | PCR/serology           | PCR/serology         | PCR/serology     | PCR/serology |
| M. urealyticum   | -                     | -                | -                | - | - | - | - |
| Urethral swab    | -                     | -                | -                | - | - | - | Negative |
| Uric acid        | -                     | -                | Within the normal range | Within the normal range | Within the normal range | - | Within the normal range |
| RF               | -                     | -                | Negative         | Negative | Negative | Negative | Negative |
| ACPA             | -                     | -                | Negative         | Negative | - | Negative | Negative |
| ANA              | -                     | -                | Negative         | - | Negative | Negative | Negative |
| Anti-ENA         | -                     | -                | -                | - | Negative | Negative | Negative |
| HLA-B27          | -                     | -                | -                | - | Negative | Negative | Negative |
| HBV and HCV      | -                     | -                | Negative         | - | - | - | Negative |
| antibodies       | -                     | -                | -                | - | - | - | Negative |
| CMV-DNA and EBV-DNA | -                  | -                | -                | - | - | - | < 1000 copies/mL |
| Urine culture    | -                     | -                | -                | - | - | - | Negative |
| Blood culture    | -                     | -                | -                | - | - | - | Negative |
| Stool culture    | -                     | -                | -                | - | - | - | Negative |

*COVID19, Coronavirus disease 19; MTP, metatarsophalangeal; PIP, proximal interphalangeal; DIP, distal interphalangeal; NSAID, non-steroidal anti-inflammatory drug; SF, synovial fluid; PCR, polymerase chain reaction; SARS-CoV2, severe acute respiratory syndrome coronavirus 2; RF, rheumatoid factor; ACPA, anti-citrullinated peptide antibodies; ANA, anti-nuclear antibodies; ENA, Extractable nuclear antibodies; HLA, human leukocyte antigen; HBV, hepatitis B virus; HCV, hepatitis C virus*
Viral-related arthritis remains, in the most cases, a diagnosis of exclusion and this underlines the importance to exhaustively perform all the tests to rule out other possible diagnosis. Unfortunately, the diagnostic workup carried out in the reported cases is often partial and incomplete. In particular, a broad microbiological investigation comprehensive of blood, urine and stool cultures, urethral swab, and serological tests for bacteria responsible for reactive arthritis is in some cases lacking. By contrast, in our case report, the chemical-physical characteristics and microbiological cultures of SF excluded septic and microcrystalline arthritis; broad-spectrum microbiological and serological tests for the common agent of bacterial reactive arthritis did not show any evidence of active or recent infections; the autoantibody immune profile resulted negative too. Finally, RT-PCR for the detection of SARS-CoV2 nucleic acids did not show the presence of the virus in the SF and this validates the hypothesis of an immune-mediated process.

Interestingly, all these suspected post-COVID-19 arthritis cases share a complete and prompt response to NSAID and/or glucocorticoids that, together with the onset timing and joint localization of arthritis, play in favor of a strict relation with SARS-CoV2 infection.

If we suppose a molecular mimicry-based pathogenesis, where the antibody response to the virus is crucial to induce joint inflammation, the rapid lowering of post-infection immunity along weeks could in turn have contributed to the fading of arthritic manifestations. Finally, this pathogenetic hypothesis, based on immune system hyperactivation, could also explain why arthritis has been reported only in patients with a severe infection; in milder forms of COVID-19, joint involvement may have a subclinical course and therefore less frequently come to medical attention.

Conclusions

Before COVID-19 outbreak, no cases of coronavirus-related arthritis have been reported in literature but SARS-CoV2 represents a new devastating entity still under study on worldwide scale. Many steps forward in the comprehension of the viral pathogenicity have been made since the start of the pandemic, but much of the infection-related consequences remain to be discovered. A growing number of cases of COVID-19-related arthritis are being reported in literature, configuring this condition worthy of further study. Complete clinical and laboratory data, SF analysis, and a strict follow-up of the patient are of paramount importance to perform a careful differential diagnosis and to better define the characteristic of inflammatory joint involvement related to SARS-CoV2 infection.

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Data availability All data relevant to the clinical case are included in the article.

Compliance with ethical standards

Disclosures None.

Ethical approval Not applicable; ethical approval not required.

Patient consent for publication Consent is obtained directly from patient, signed on July 15, 2020.

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