Effectiveness of Transmitted Drug Resistance Testing Before Initiation of Antiretroviral Therapy in HIV-Positive Individuals

Sara Lodi, Huldrych F. Günthard, John Gill, Andrew N. Phillips, David Dunn, Quang Vu, Peter Reiss, Federico Garcia, Roger Logan, Sophie Jose, Heiner C. Bucher, Alexandra U. Scherrer, Peter Reiss, Ard van Sighem, T. Sonia Boender, Kholoud Porter, Richard Gilson, Dimitrios Paraskevis, Metallidis Simeon, Georgia Vourli, Santiago Moreno, Inmaculada Jarrin, Caroline Sabin, and Miguel A. Hernán on behalf of the HIV-CAUSAL Collaboration

Background: For people living with HIV, major guidelines in high-income countries recommend testing for transmitted drug resistance (TDR) to guide the choice of first-line antiretroviral therapy (ART). However, individuals who fail a first-line regimen can now be switched to one of several effective regimens. Therefore, the virological and clinical benefit of TDR testing needs to be evaluated.

Methods: We included individuals from the HIV-CAUSAL Collaboration who enrolled <6 months of HIV diagnosis between 2006 and 2015, were ART-naive, and had measured CD4 count and HIV-RNA. Follow-up started at the date when all inclusion criteria were first met (baseline). We compared 2 strategies: (1) TDR testing within 3 months of baseline versus (2) no TDR testing. We used inverse probability weighting to estimate the 5-year proportion and hazard ratios (HRs) of virological suppression.
Transmitted drug resistance (TDR) occurs when an individual is infected with a strain of HIV that is resistant to a single antiretroviral, an entire drug class, or multiple classes. The prevalence of TDR in ART-naive individuals in high-income countries ranges between 7% and 14%. Clinical practice guidelines recommend TDR testing—a genotypic test—in all individuals newly diagnosed with HIV before initiating antiretroviral therapy (ART). The expectation is that TDR test results will guide the choice of ART agents and reduce the risk of using a less effective regimen. Use of a drug to which the virus is resistant could be detrimental in the long term because it prolongs the time to achieve virological suppression and increases the risk of developing resistance to active treatments.

Although TDR testing was shown to be beneficial in the early 2000s' rapid changes in the landscape of HIV treatment require a reevaluation of the value of TDR testing. Treatments containing agents with a high barrier to resistance, such as integrase inhibitors and boosted darunavir, as well as several other potent second- and third-line ART options are now widely available in high-income settings. These innovations imply that although most individuals will achieve adequate virological suppression from their initial regimen, those who do not will likely do so after switching to an alternative treatment.

Here, we used observational data to estimate the effectiveness of TDR testing, when used according to current guidelines, on virological and clinical outcomes up to 5 years after HIV diagnosis among individuals diagnosed with HIV in Europe and Canada between 2006 and 2015.

METHODS

Study Data

The HIV-CAUSAL Collaboration is a consortium of prospective HIV cohorts from Europe and the Americas. All cohorts recorded routinely collected data in clinical practice within settings with universal access to care. Data collected include patient characteristics (age, sex, geographical origin, and transmission category), use of ART (type of regimes and dates of start and stop), CD4 cell counts, and plasma HIV-RNA, AIDS-defining conditions, and death (cause(s) and date). Each cohort submits data in a standardized format (http://www.hicdep.org/) to the coordinating center. The analyses presented here are based on data pooled from 7 cohorts within the HIV-CAUSAL Collaboration that contributed data on genotypic drug resistance testing conducted as part of routine clinical care: AMACS (Greece), ATHENA (Netherlands), CoRIS (Spain), Swiss HIV Cohort Study (Switzerland), the South Alberta Cohort Study (Canada), UK CHIC/UK HIV Drug Resistance Database (United Kingdom), and UK Register of HIV Seroconverters (United Kingdom). The date of a TDR testing was the date the blood sample for the testing was drawn, rather than the date of sequencing.

We defined a TDR test as any genotypic drug resistance test conducted while the individual was ART-naive. Predicted resistance was derived for all individuals using the Genotypic Resistance Interpretation Algorithm, version 7.0 (HIVdb Program, Stanford University, http://hivdb.stanford.edu). TDR was defined as predicted intermediate- or high-level resistance to any of the following antiretroviral drugs in use during the study period: atazanavir, darunavir, fosamprenavir, indinavir, lopinavir, nelfinavir, saquinavir, tipranavir (protease inhibitors [PIs]); lamivudine, emtricitabine, abacavir, didanosine, tenofovir, stavudine, zidovudine [nucleoside reverse transcriptase inhibitors (NRTIs)]; nevirapine, efavirenz, etravirine, rilpivirine [nonnucleoside reverse transcriptase inhibitors (NNRTIs)]. Resistance to integrase inhibitors, fusion inhibitors, and CCR antagonists were not routinely measured in the participant cohorts. Resistance data on integrase inhibitors were collected in only one cohort and were rare even in this cohort.

The date of ART initiation was the date that a patient initiated HIV treatment including at least 2 NRTIs plus one or more of the following: an integrase inhibitor, PIs, an NNRTI, an entry inhibitor, or a fusion inhibitor. Regimens consisting of abacavir or tenofovir with 2 or more additional nucleoside reverse transcriptase inhibitors were also considered ART regimens, but they were rare in our population (<2%).

Eligibility Criteria

The analyses were restricted to individuals who met the following eligibility criteria between January 1, 2006, and December 31, 2015: aged 18 years and older, CD4 cell count and HIV-RNA measurements within 3 months of each other while ART-naive, and less than 6 months since HIV diagnosis. We restricted the analysis to individuals enrolled in clinics within cohorts where at least 30% of patients had received a genotypic drug resistance test while naïve.

TDR Testing Strategies

We compared the following 2 strategies: (1) TDR testing under current recommendations, defined as genotypic
drug resistance testing within 3 months while ART-naive, and (2) no TDR testing, defined as no genotypic drug resistance before receiving ART. The 3-month time window for testing (the “grace period”) corresponds to the period within which 90% of TDR testing took place in our data. Note that our analysis emulates a pragmatic trial to estimate the effectiveness of TDR testing in which decisions concerning ART initiation are left to the discretion of the treating physicians. For descriptive purposes, we compared the proportion of individuals who initiated ART under the 2 strategies.

**Outcomes**

We considered the following 2 outcomes: (1) virological suppression, defined as the second of 2 consecutive HIV-RNA <50 copies/mL after initiating ART, and (2) a combined endpoint of an AIDS-defining condition or death. Separately for each outcome, we computed the 5-year cumulative incidence under each strategy, the difference in cumulative incidence, and the hazard ratio (HR).

**Follow-up**

For each individual, follow-up started at baseline, defined as the earliest date that all eligibility criteria were met, and ended at the earliest of developing the outcome, 12 months after the most recent HIV-RNA or CD4 count measurement (censoring due to infrequent laboratory measurements), cohort-specific administrative end of follow-up (ranging from December 2013 to November 2015), date of pregnancy when known, or initiation of an ART combination not defined as ART.

**Statistical Analyses**

For each outcome, we estimated the HR of TDR testing versus no testing using a pooled logistic regression model with an indicator for TDR testing strategy, month of follow-up (restricted cubic splines with 4 knots), and the following covariates at baseline: cohort, sex, HIV acquisition group (heterosexual contact, homosexual contact, or bisexual contact, injecting drug use, or other/unknown), geographical origin (Western countries, sub-Saharan Africa, other, unknown), age (<35, 35–50, >50 years), HIV-RNA (<4, 4–5, >5 log10 copies/mL), CD4 cell count strata (<100–199, 200–349, 350–499, ≥500 cells/mm³), AIDS, and calendar year (2006–2009, 2010–2015) at HIV diagnosis. In the model for the virological outcome, we considered death as a censoring event. We also fit pooled logistic regression models for each outcome like the one above and added an interaction term between month and the indicator of TDR testing strategy. The model’s predicted probabilities were then used to estimate the 5-year cumulative incidence under each strategy and the difference in these values.

TDR testing strategies are defined over a 3-month grace period. For individuals who were not tested for TDR in the baseline month, TDR testing is a postbaseline variable. Therefore, to avoid immortal time bias, we replaced each individual without a TDR test in the baseline month by 2 identical clones. The first clone was assigned to the TDR testing strategy and was censored on deviation from that strategy, that is, at 3 months if the individual had not received a TDR test by that time, or earlier at the time of ART initiation, if the individual initiated ART before receiving a TDR test during the grace period. The second clone was assigned to the no TDR testing strategy and was censored on deviation from that strategy, that is, when the individual received a TDR test. Examples of how cloning and artificial censoring occurred under each TDR testing strategy are presented in Appendix Figure 1, Supplemental Digital Content, http://links.lww.com/QAI/B368.

To adjust for the potential selection bias induced by censoring, we weighted each clone at each time by the inverse of the probability of remaining uncensored. To estimate the weights for censoring due to deviation from the assigned TDR testing strategy, we fit 2 separate pooled logistic regression models for ART initiation before TDR testing and for TDR testing while ART-naive in the original data set. Each model included the previously listed covariates plus the most recent CD4 cell count (restricted cubic splines with knots at 200, 350, 500 and 1000 cells/mm³), HIV-RNA category (<1000, 1000–10,000, >10,000 copies/mL), diagnosis of AIDS (when the outcome was virological suppression), and months since the last CD4 and last HIV-RNA measurements. Individuals who had TDR testing at baseline month were assigned a weight equal to 1. To estimate weights for censoring due to infrequent laboratory measurements, we used a pooled logistic regression model with the same covariates as in the models listed above on the original data set. We stabilized and multiplied the 2 sets of weights. The mean of the weights was 1.02 (min 0.20, max 7.8). Estimates of the effect of TDR testing remained basically unchanged in analyses with weights truncated at the 99th percentile.

We used a nonparametric bootstrap procedure based on 500 samples to obtain percentile-based 95% confidence intervals (CIs). All analyses were conducted with SAS version 9.4.

**Sensitivity Analyses**

We conducted several sensitivity analyses to examine the robustness of our findings. First, because the proportion of individuals tested for TDR varied by center and cohort, we restricted the analyses to (1) the 3 cohorts with highest overall occurrence of TDR testing (UK CHIC/UK Drug Resistance Database, Swiss HIV Cohort Study and Southern Alberta Cohort) and (2) centers in each cohort where at least 50% of patients were tested for TDR. Second, because some TDR may revert relatively quickly after infection, we restricted the analyses on the subset of individuals with early HIV disease defined as having a CD4 count >500 cells/mm³ at baseline. Third, to account for changes over time of antiretroviral combinations, we repeated the analysis separately by calendar period (2006–2009 versus 2009–2015).
TABLE 1. Baseline Characteristics of 25,672 Eligible Individuals, HIV-CAUSAL Collaboration 2006–2015

| Characteristic                  | Individuals (%) | Median (IQR) | Individuals With TDR Testing* (%) | Individuals With TDR Detected† (%) |
|--------------------------------|-----------------|--------------|----------------------------------|-----------------------------------|
| CD4 count, cells/mm³           |                 |              |                                  |                                   |
| <100                           | 3094 (12)       | 34 (14–62)   | 1937 (64)                        | 104 (5)                           |
| 100–200                        | 2542 (10)       | 33 (14–60)   | 1686 (66)                        | 82 (5)                            |
| 200–349                        | 5421 (21)       | 31 (13–59)   | 3803 (70)                        | 206 (5)                           |
| 350–499                        | 5961 (23)       | 27 (13–53)   | 4106 (69)                        | 255 (6)                           |
| ≥500                           | 8654 (34)       | 25 (11–50)   | 5657 (65)                        | 352 (6)                           |
| HIV-RNA, copies/mL             |                 |              |                                  |                                   |
| <1,000                         | 6231 (24)       | 28 (12–55)   | 3880 (62)                        | 238 (6)                           |
| 1,000–10,000                   | 10,984 (43)     | 28 (13–55)   | 7560 (69)                        | 455 (6)                           |
| 10,000                         | 8457 (33)       | 29 (13–55)   | 5749 (68)                        | 306 (5)                           |
| Sex                            |                 |              |                                  |                                   |
| Male                           | 21,101 (82)     | 29 (13–56)   | 14,358 (68)                      | 847 (6)                           |
| Female                         | 4571 (18)       | 27 (11–53)   | 2831 (62)                        | 152 (5)                           |
| HIV acquisition group          |                 |              |                                  |                                   |
| Heterosexual contact           | 7713 (30)       | 29 (13–57)   | 4863 (63)                        | 261 (5)                           |
| Homosexual contact             | 15,429 (60)     | 29 (14–56)   | 10,765 (70)                      | 652 (6)                           |
| Injecting drug use             | 719 (3)         | 18 (11–38)   | 431 (60)                         | 16 (4)                            |
| Other/unknown                  | 1811 (7)        | 23 (11–50)   | 1130 (68)                        | 70 (6)                            |
| Geographical origin            |                 |              |                                  |                                   |
| Western countries              | 10,430 (41)     | 33 (14–62)   | 6340 (61)                        | 405 (6)                           |
| Sub-Saharan Africa             | 1133 (4)        | 30 (12–62)   | 666 (59)                         | 33 (5)                            |
| Rest of the world              | 2470 (10)       | 28 (11–53)   | 1362 (55)                        | 79 (6)                            |
| Unknown country                | 11,639 (45)     | 25 (12–49)   | 8821 (76)                        | 482 (5)                           |
| Calendar year                  |                 |              |                                  |                                   |
| 2006–2009                      | 12,249 (48)     | 56 (24–75)   | 8334 (68)                        | 484 (6)                           |
| 2010–2015                      | 13,423 (52)     | 19 (11–32)   | 8855 (66)                        | 515 (6)                           |
| Age, yr                        |                 |              |                                  |                                   |
| <35                            | 11,921 (46)     | 25 (12–50)   | 8087 (68)                        | 451 (6)                           |
| 35–49                          | 10,651 (41)     | 32 (14–59)   | 7131 (67)                        | 423 (6)                           |
| >50                            | 3100 (12)       | 31 (14–58)   | 1971 (64)                        | 125 (6)                           |
| Late HIV presentation          |                 |              |                                  |                                   |
| No                             | 14,257 (56)     | 26 (12–51)   | 9513 (67)                        | 595 (6)                           |
| Yes                            | 11,415 (44)     | 32 (14–60)   | 7676 (67)                        | 404 (5)                           |
| All                            | 25,672 (100)    | 28 (13–55)   | 17,189 (100)                     | 999 (100)                         |

*TDR test = genotypic HIV resistance test within 3 months of baseline in an ART-naive patient.
†Proportions with TDR among individuals who were tested.
MSM, men who have sex with men; MSW, men who have sex with women.

2010–2015). Fourth, because some TDR tests might have been conducted retrospectively on previously collected blood samples, we reran the analyses excluding individuals who initiated an ART combination including a treatment to which they were resistant. Fifth, we restricted the analyses to homosexual or bisexual males, the group with the highest prevalence of TDR testing and TDR detection in our data. Sixth, ART initiation might be prioritized over TDR testing in individuals with more advanced HIV disease such as late presenters. We, therefore, rerun the analyses changing the definition of TDR testing by allowing genotypic tests occurring within 2 weeks after ART initiation. Because individuals with late diagnosis are at risk of early virological failure and possibly faster clinical progression, we restricted the analyses to individuals with late HIV diagnosis defined as having a CD4 <200 cells/mm³ or AIDS at baseline and to individuals with advanced HIV disease, defined as having a CD4 <200 cells/mm³ or AIDS at baseline. Finally, in the analyses where virological suppression was an outcome, death was treated as a competing risk rather than a censoring event.

RESULTS

Of 25,672 eligible individuals, 82% were men, 44% were late HIV presenters, and 52% started follow-up in 2010 or later. Their median (interquartile range (IQR)) CD4 cell count, HIV-RNA, and age at baseline were 390 (221–570) cells/mm³, 4.6 (4.0–5.2) log₁₀ copies/mL, and 36 (29–44) years, respectively (Table 1). A total of 16,354 (64%) individuals initiated ART at a median (IQR) CD4 cell count of 276 (160–383) cells/mm³. Of these, 9953 (61%) started with an NNRTI regime, 5384 (33%) with a boosted PI regime, 778 (5%) with an INSTI regime, 228 (1%) with an unboosted PI regime, and 11 (<1%) with a 3 NRTI regime.

TDR testing while ART-naive occurred in 67% of individuals within 3 months of baseline (91% of them during the baseline month). Median (IQR) follow-up was 31 (14–58) months for individuals with TDR testing and 44 (13–94) months for individuals without TDR testing. Of 17,189 individuals with TDR testing while ART-naive occurred in 67% of individuals within 3 months of baseline (91% of them during the baseline month). Median (IQR) follow-up was 31 (14–58) months for individuals with TDR testing and 44 (13–94) months for individuals without TDR testing. Of 17,189 individuals with TDR testing while ART-naive occurred in 67% of individuals within 3 months of baseline (91% of them during the baseline month). Median (IQR) follow-up was 31 (14–58) months for individuals with TDR testing and 44 (13–94) months for individuals without TDR testing. Of 17,189 individuals with TDR testing while ART-naive occurred in 67% of individuals within 3 months of baseline (91% of them during the baseline month). Median (IQR) follow-up was 31 (14–58) months for individuals with TDR testing and 44 (13–94) months for individuals without TDR testing. Of 17,189 individuals with TDR testing while ART-naive occurred in 67% of individuals within 3 months of baseline (91% of them during the baseline month). Median (IQR) follow-up was 31 (14–58) months for individuals with TDR testing and 44 (13–94) months for individuals without TDR testing. Of 17,189 individuals with TDR testing while ART-naive occurred in 67% of individuals within 3 months of baseline (91% of them during the baseline month). Median (IQR) follow-up was 31 (14–58) months for individuals with TDR testing and 44 (13–94) months for individuals without TDR testing.
Of those with a detected TDR, 5% initiated ART with a combination containing a drug to which they were resistant. The proportion of individuals with a detected TDR varied by cohort but was similar across subgroups defined by late HIV presentation status, sex, age, calendar year, CD4 count, and HIV-RNA at baseline (last column, Table 1). Detected TDR was less common among injecting drug users than in other HIV acquisition groups.

Figure 1 and Appendix Figure 2, Supplemental Digital Content, http://links.lww.com/QAI/B368, show the cumulative incidence curves under the 2 TDR testing strategies assuming no informative loss to follow-up. As shown in Table 2, the estimated 5-year risk (95% CI) of AIDS or death was 6.0% (5.6 to 6.4) under TDR testing and 5.7% (5.1 to 6.4) under no TDR testing; difference 0.3% (−0.3 to 0.9); HR 1.03 (0.95 to 1.12).

The estimated 5-year proportion (95% CI) who achieved virological suppression was 77.4% (76.6 to 78.2) under TDR testing and 73.7% (72.0 to 75.4) under no TDR testing; difference 3.7% (2.1 to 5.3); HR 1.06 (1.03 to 1.09). The estimated proportion (95% CI) of individuals who had ever initiated ART by 5 years of baseline was 87% (85 to 88) under TDR testing and 87% (83 to 90) under no TDR testing.

The results of the sensitivity analyses are presented in Table 3. The results did not materially change when the analyses were restricted to individuals from the 3 cohorts with the highest rate of TDR testing, to those who were enrolled in centers with more than 50% TDR testing, to individuals with late HIV diagnosis, or to homosexual or bisexual males; when excluding individuals who initiated ART with a combination containing a drug to which they were resistant; or when death was considered as a competing risk. The estimated HRs of virological suppression for TDR testing versus no TDR testing were similar for individuals with baseline in the period 2005–2010 and 2011–2015.

**DISCUSSION**

In individuals diagnosed with HIV in high-income countries, we estimated that current recommendations for TDR testing, compared with no TDR testing, increased the proportion achieving virological suppression by 5 years from diagnosis by 3.7% but with no effect on AIDS or death in this time frame.

One explanation for the lack of any short-term clinical benefit is the low prevalence of TDR (approximately 6% in our population), which imposes a hard limit on the maximum gain that can be achieved by TDR testing. As TDR testing may be beneficial in populations with higher TDR prevalence, care should be taken with extrapolating our results to resource-limited settings because of the differences in the TDR prevalence, HIV subtypes, CD4 count distribution at HIV diagnosis, and treatment patterns. Also, there may be other benefits of TDR testing that our study could not quantify, including some decrease in HIV transmission because of a modest increase in virological suppression, and the generation of genotypic information to describe the dynamics of local HIV transmission (through phylogenetic tree studies) and the changing patterns of drug resistance.

Previous studies have compared the virological response after ART initiation for individuals with and without a previously detected TDR. However, this approach might lead to selection bias if TDR testing affects the likelihood of ART initiation or loss to follow-up. Unlike these studies, we estimated the effect of TDR testing rather than of having a detected TDR. Importantly, we quantified the risk of the virological and clinical outcomes since baseline, a proxy of entry into HIV care and thus of the recommended time for testing, rather than since ART initiation. In fact, our approach parallels the design and emulation of a pragmatic randomized trial of TDR testing in which individuals are randomly assigned to either TDR testing or no testing while not intervening on other aspects of their care. Another strength of our study is the large sample size of over 30,000 individuals.
25,000 individuals and the setting in HIV clinics in Europe and Canada that are considered representative of routine clinical practice where TDR testing is part of routine clinical routine.53

TDR testing is controversial in individuals who present for HIV care with advanced immunosuppression and are therefore in need of immediate initiation of effective treatment.56,57 Although TDR testing might be beneficial to identify the optimal treatment combination in a population with high mortality, it might also cause delays in initiating treatment and most clinicians would not want to wait for the result of the resistance test before initiating ART.58 Also, because of the availability of several effective second-line options, patients who do not respond have the opportunity to be switched to an effective drug and virological suppression may end up being delayed only by a few months. A large proportion of patients in our population (44%) presented late for HIV diagnosis. We found that the HRs of both the clinical and virological outcomes in the subset of individuals with late HIV diagnosis or advanced HIV disease at baseline were similar to the one in the overall population. This supports current TDR testing recommendations that do not vary by HIV presentation status.

Current guidelines worldwide recommend ART initiation with integrase inhibitors.39–42 Only a small proportion of patients (5%) in our study initiated treatment with integrase inhibitors; so, we could not appropriately assess TDR for this drug class, which seems to be rare at this point.41,47 In high-income countries, it is common practice not to delay ART initiation until the results of the drug resistance test become available, which usually takes 2 weeks.10 Instead, because resistance can emerge quickly in low-resistance barrier containing drugs,11 when an NNRTI-based regimen is considered as first-line treatment, the results of the resistance test should be available before starting treatment.

Our study has several limitations. First, the validity of our estimates relies on the untestable assumption that all confounders were adequately adjusted for. We expect this assumption to approximately hold because we adjusted for the most important known predictors for TDR including risk group, sex, and calendar year. Second, some TDR tests could have been performed retrospectively on stored blood samples, possibly in response to virological failure in individuals believed to have good adherence, which might have introduced bias. However, this is unlikely to have affected our estimates because only 5% of patients with a TDR test started with a combination containing an ART they were resistant to. Third, 5 years might not be a long enough period to examine differences in clinical outcomes. Finally, although TDR testing was recommended in all countries of the study between 2006 and 2015,44–47 the large proportion of individuals who were not tested (33%) might reflect incomplete data. The robustness of our results to sensitivity analyses restricting to centers with more than 50% patients tested for TDR and to cohorts with high TDR testing uptake is reassuring.

In conclusion, we found a low prevalence of TDR in high-income countries. Although TDR testing seems to improve virological suppression after ART initiation, we found no evidence that it reduced the incidence of AIDS or death for up to 5 years after baseline. These results call for more evidence to establish which populations would benefit from selective TDR screening.

**REFERENCES**

1. Frentz D, Boucher C, Van De Vijver D. Temporal changes in the epidemiology of transmission of drug-resistant HIV-1 across the world. *AIDS Rev.* 2012;14:17–27.
2. Frentz D, van de Vijver D, Abecasis A, et al. Patterns of transmitted HIV drug resistance in Europe vary by risk group. *PLoS One.* 2014;9:e94495.
3. Monge S, Guillot V, Alvarez M, et al. Clinically relevant transmitted drug resistance to first line antiretroviral drugs and implications for recommendations. *PLoS One.* 2014;9:e90710.
4. Resistance UKCGoHDI, Dolling D, Sabin C, Delpech V, et al. Time trends in drug resistant HIV-1 infections in the United Kingdom up to 2009: multicentre observational study. *BMJ.* 2012;345:e5253.
5. Yang WL, Kouyos R, Scherrer AU, et al. Assessing the paradox between transmitted and acquired HIV type 1 drug resistance mutations in the Swiss HIV cohort study from 1998 to 2012. *J Infect Dis.* 2015;212:28–38.
6. Rhee SY, Clutter D, Fessel WJ, et al. Trends in the molecular epidemiology and genetic mechanisms of transmitted HIV-1 drug resistance in a large U.S. clinic population. *Clin Infect Dis.* 2019;68:213–221.
7. Hofstra LM, Sauvageot N, Albert J, et al. Transmission of HIV drug resistance and the predicted effect on current first-line regimens in Europe. *Clin Infect Dis.* 2016;62:655–663.
8. Hirsch MS, Gunthard HF, Schapiro JM, et al. Antiretroviral drug resistance testing in adult HIV-1 infection: 2008 recommendations of an International AIDS Society–USA panel. Clin Infect Dis. 2008;47:266–285.

9. Vandamme AM, Camacho RJ, Cecherini-Silberstein F, et al. European recommendations for the clinical use of HIV drug resistance testing: 2011 update. AIDS Rev. 2011;13:77–108.

10. Gunthard HF, Saag MS, Benson CA, et al. Antiretroviral drugs for treatment and prevention of HIV infection in adults: 2016 recommendations of the International Antiviral Society–USA panel. JAMA. 2016;316:191–210.

11. Gunthard HF, Calvez V, Paredes R, et al. Human immunodeficiency virus drug resistance: 2018 recommendations of the International Antiviral Society–USA panel. Clin Infect Dis. 2018;68:177–187.

12. Kuritzkes DR, Lalama CM, Ribaudo HJ, et al. Preeexistence to nonnucleoside reverse-transcriptase inhibitors predicts virologic failure of an efavirenz-based regimen in treatment-naïve HIV-1-infected subjects. J Infect Dis. 2008;197:867–870.

13. Wittkop L, Gunthard HF, de Wolf F, et al. Effect of transmitted drug resistance on virological and immunological response to initial combination antiretroviral therapy for HIV (EuroCoord-CHAIN joint project): a European multicohort study. Lancet Infect Dis. 2011;11:363–371.

14. Little SJ, Holte S, Routy JP, et al. Antiretroviral-drug resistance among patients recently infected with HIV. N Engl J Med. 2002;347:385–394.

15. Aves T, Tambe J, Siemieniuk RA, et al. Antiretroviral resistance testing in HIV-positive people. Cochrane Database Syst Rev. 2018;11:CD006495.

16. Sax PE, Islam R, Walensky RP, et al. Should resistance testing be performed for treatment-naïve HIV-infected patients? A cost-effectiveness analysis. Clin Infect Dis. 2005;41:1316–1323.

17. Scherrer AU, Yang WL, Koyous RD, et al. Successful prevention of transmission of integrase resistance in the Swiss HIV cohort study. J Infect Dis. 2016;214:399–402.

18. Ancelle-Park R. Expanded European AIDS case definition. Lancet. 1993;341:441.

19. Hernán MA, Robins JM. Using big data to emulate a target trial when a randomized trial is not available. Am J Epidemiol. 2016;183:758–764.

20. Caniglia EC, Sabin C, Robins JM, et al. When to monitor CD4 cell count and HIV RNA to reduce mortality and AIDS-defining illness in virologically suppressed HIV-positive persons on antiretroviral therapy in high-income countries: a prospective observational study. J Acquir Immune Defic Syndr. 2016;72:214–221.

21. Cain LE, Robins JM, Lanoy E, et al. When to start treatment? A systematic approach to the comparison of dynamic regimes using observational data. Int J Biostat. 2010;6:18.

22. Caniglia EC, Cain LE, Sabin CA, et al. Comparison of dynamic monitoring strategies based on CD4 cell counts in virally suppressed, HIV-positive individuals on combination antiretroviral therapy in high-income countries: a prospective, observational study. Lancet HIV. 2017;4:e251–e259.

23. Cole SR, Hernan MA. Constructing inverse probability weights for marginal structural models. Am J Epidemiol. 2008;168:656–664.

24. Hernan MA, Robins JM. Estimating causal effects from epidemiological data. J Epidemiol Community Health. 2006;60:578–586.

25. Yang WL, Koyous RD, Boni J, et al. Persistence of transmitted HIV-1 drug resistance mutations associated with fitness costs and viral genetic backgrounds. PLoS Pathog. 2015;11:e1004722.

26. Antinori A, Coenen T, Costaglia D, et al. Late presentation of HIV infection: a consensus definition. HIV Med. 2011;12:61–64.

27. Cohen MS, Chen YQ, McCaulley M, et al. Antiretroviral therapy for the prevention of HIV-1 transmission. N Engl J Med. 2016;375:830–839.

28. Consensus Statement. Risk of Sexual Transmission of HIV From a Person Living With HIV Who Has Undetectable Viral Load. 2018. Available at: https://www.preventionaccess.org/consensus.