Urine metabolomics and microbiome analyses reveal the mechanism of anti-tuberculosis drug-induced liver injury, as assessed for causality using the updated RUCAM: A prospective study

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Background: Anti-tuberculosis drug-induced liver injury (ATB-DILI) is one of the most common adverse reactions that brings great difficulties to the treatment of tuberculosis. Thus, early identification of individuals at risk for ATB-DILI is urgent. We conducted a prospective cohort study to analyze the urinary metabolic and microbial profiles of patients with ATB-DILI before drug administration. And machine learning method was used to perform prediction model for ATB-DILI based on metabolomics, microbiome and clinical data.

Methods: A total of 74 new TB patients treated with standard first-line anti-TB treatment regimens were enrolled from West China Hospital of Sichuan University. Only patients with an updated RUCAM score of 6 or more were accepted in this study. Nontargeted metabolomics and microbiome analyses were performed on urine samples prior to anti-tuberculosis drug ingestion to screen the differential metabolites and microbes between the ATB-DILI group and the non-ATB-DILI group. Integrating electronic medical records, metabolomics, and microbiome data, four machine learning methods was used, including random forest algorithm, artificial neural network, support vector machine with the linear kernel and radial basis function kernel.

Results: Of all included patients, 69 patients completed follow-up, with 16 (23.19%) patients developing ATB-DILI after antituberculosis treatment. Finally, 14 ATB-DILI patients and 30 age- and sex-matched non-ATB-DILI patients were subjected to urinary metabolomic and microbiome analysis. A total of 28 major differential metabolites were screened out, involving bile secretion, nicotinate and nicotinamide metabolism, tryptophan metabolism, ABC...
transporters, etc. Negativicoccus and Actinotignum were upregulated in the ATB-DILI group. Multivariate analysis also showed significant metabolic and microbial differences between the non-ATB-DILI and severe ATB-DILI groups. Finally, the four models showed high accuracy in predicting ATB-DILI, with the area under the curve of more than 0.85 for the training set and 1 for the validation set.

**Conclusion:** This study characterized the metabolic and microbial profile of ATB-DILI risk individuals before drug ingestion for the first time. Metabolomic and microbiome characteristics in patient urine before anti-tuberculosis drug ingestion may predict the risk of liver injury after ingesting anti-tuberculosis drugs. Machine learning algorithms provides a new way to predict the occurrence of ATB-DILI among tuberculosis patients.

**KEYWORDS**
metabolomic, microbiome, anti-tuberculosis drug-induced liver injury (ATB-DILI), machine learning, cohort, updated RUCAM

**Introduction**

Tuberculosis (TB) is caused by *Mycobacterium tuberculosis* infection, which is an infectious disease with the highest mortality before the novel coronavirus pneumonia pandemic (1). According to the report of the World Health Organization, there were 9.9 million new cases of tuberculosis worldwide, and approximately 1.3 million patients died in 2020 (1). After treatment with a first-line anti-tuberculosis regimen containing isoniazid and rifampicin, 86% of patients were treated successfully (1–3). However, it is often accompanied by various adverse drug reactions, such as gastrointestinal reaction, drug-induced liver injury (DILI), hyperuricemia, leucopenia, allergy, peripheral neuritis and so on (3–7). Anti-tuberculosis drug-induced liver injury (ATB-DILI) is one of the most common adverse reactions in the treatment of tuberculosis (4–6) and may lead to treatment interruption, prolonged treatment time, decreased treatment success rate and increased hospitalization rate (8–10). Early identification and evaluation of ATB-DILI will provide new ideas for the precise treatment of tuberculosis patients.

Currently, the identification of biomarkers by metabolomics has been widely used in pathophysiological mechanisms in many scientific fields, such as plant biology (11), toxicology (12) and disease diagnosis and prognosis (13–16). Ultrahigh performance liquid chromatography tandem mass spectrometry (UPLC-MS) is one of the most effective means of metabolomics research (17). Through metabolomics research, Xie et al. found that 31 metabolites were related to DILI and were closely related to the severity of DILI (18). Prospective studies show that there are significant differences in serum metabolites between the DILI group and the non-DILI group prior to polygonum multiflorum ingestion, and the unique metabolic characteristics may be used to predict the risk of DILI after taking polygonum multiflorum (19). This suggests that metabolomics can be used to evaluate and predict DILI. There are a few clinical and animal experiments using metabolomics to explore the toxic mechanism and biomarkers of ATB-DILI (20–25). Nontargeted metabolomics found that 28 metabolites can be used as important distinguishing factors between ATB-DILI and non-ATB-DILI patients, and ATB-DILI affects the tricarboxylic acid cycle, arginine and proline metabolism, purine metabolism and pentose phosphate pathway (24). In our previous study, 11 urine differential metabolites were identified between ATB-DILI patients and non-ATB-DILI patients by gas chromatography-mass spectrometry (GC-MS) (26). These studies indicate that metabolomics is helpful for a new understanding of the pathophysiological process of ATB-DILI and for screening new markers of ATB-DILI. Moreover, no study has evaluated the metabolic characteristics of ATB-DILI patients and non-ATB-DILI patients before taking anti-tuberculosis drugs.

There are thousands of microbial species in the human microbial ecosystem that play a key role in maintaining host immunity, metabolism, drug metabolism, vitamin production and carbohydrate metabolism (27–29). Research interest has been focused on the interaction between the microbiota and the host, and how the composition of the human microbiota may have a potential impact on the development of certain diseases, such as metabolic syndrome, obesity (30), diabetes (31) and liver injury (26, 32, 33). Previous studies have shown that the
quantities of the urine microbiota differ significantly between patients with ATB-DILI and without ATB-DILI (26).

At present, the data on metabolomic or microbiota changes related to ATB-DILI are limited, especially premedication data. In addition, a model for the prediction of ATB-DILI is lacking. In this study, we hypothesized that the metabolome and microbiome are related to ATB-DILI. Therefore, we performed urine metabolomic and microbiota analyses of ATB-DILI prior to medication. Meanwhile, four machine learning methods was used to establish a clinical prediction model of ATB-DILI based on metabolomics, microbiome and clinical data.

Methods

Study population and sample collection

This prospective cohort study included patients with tuberculosis who visited the tuberculosis clinic of West China Hospital of Sichuan University from March 2021 to December 2021. The study was approved by the Ethics Committee of West China Hospital of Sichuan University. All research subjects were required to sign a written informed consent form by themselves or their representatives before being included in the study. Demographic datasets of patients with laboratory test data were obtained through electronic medical records and questionnaires.

Inclusion criteria are as follows: 1) age ≥16 years and <80 years old; 2) newly diagnosed TB patients, including etiologically confirmed, pathologically confirmed and clinically diagnosed cases; 3) standard first-line anti-TB treatment regimens (including 2-month HRZE intensive treatment and at least 4 months of HRE consolidation therapy), and can be followed up regularly; 4) Han nationality in Southwest China; 5) voluntarily participate in this study and sign the informed consent form. Therefore, urine is more advantageous compared to other sample types and was used as a study sample in this study.

Clean midstream urine from patients before medication was collected, divided into three 1 ml aliquots, and immediately stored at −80°C. We discarded samples that were at room temperature for >2 hours.

Metabolite extraction was primarily performed according to previously reported methods (40, 41). In short, 100 µL samples were extracted by directly adding 300 µL of precooled methanol and acetonitrile (2:1, v/v), and internal standards mix (contains L-Leucine-d3, L-Phenylalanine (13C9, 99%), L-Tryptophan-d5, Progesterone-2,3,4-13C3) were added for quality control of sample preparation. After vortexing for 1 min and incubating at −20°C for 2 h, the samples were centrifuged for 20 min at 4000 rpm, and the supernatant was then transferred for vacuum freeze drying. The metabolites were resuspended in 150 µL of 50% methanol and centrifuged for 30 min at 4000 rpm, and the supernatants were transferred to autosampler vials for LC-MS analysis. A quality control (QC) sample was prepared by pooling the same volume of each sample to evaluate the reproducibility of the whole LC-MS analysis.

Sample preparation

Urine is the common sample type used to perform metabolomics studies (38, 39). Compared to other samples, urine has easy sampling, low protein levels and less complexity (38). Also the urine metabolites are products of normal and abnormal cellular biological processes and can reflect a wide range of phenotypes including genetic modifications (38). Therefore, urine is more advantageous compared to other sample types and was used as a study sample in this study.

Metabolite detection and comments

This experiment used a Waters 2D UPLC (Waters, USA) tandem Q Exactive HF high resolution mass spectrometer
The full scan range was 70 eV. The stepped normalized collision energy was set to 20, 40 and 60 eV.

Chromatographic separation was performed on a Waters ACQUITY UPLC BEH C18 column (1.7 µm, 2.1 mm × 100 mm, Waters, USA), and the column temperature was maintained at 45°C. The mobile phase consisted of 0.1% formic acid (A) and acetonitrile (B) in the positive mode, and in the negative mode, the mobile phase consisted of 10 mM ammonium formate (A) and acetonitrile (B). The gradient conditions were as follows: 0-1 min, 2% B; 1-9 min, 2%-98% B; 9-12 min, 98% B; 12-12.1 min, 98% B to 2% B; and 12.1-15 min, 2% B. The flow rate was 0.35 mL/min and the injection volume was 5 µL.

The mass spectrometric settings for positive/negative ionization modes (ESI+/−) were as follows: spray voltage, 3.8/−3.2 kV; sheath gas flow rate, 40 arbitrary units (arb); aux gas flow rate, 10 arb; aux gas heater temperature, 350°C; capillary temperature, 320°C. The full scan range was 70–1050 m/z with a resolution of 70000, and the automatic gain control (AGC) target for MS acquisitions was set to 3e6 with a maximum ion injection time of 100 ms. The top 3 precursors were selected for subsequent MSMS fragmentation with a maximum ion injection time of 50 ms and resolution of 30,000, and the AGC was 1e5. The stepped normalized collision energy was set to 20, 40 and 60 eV.

**LC–MS/MS analysis**

The original data (raw file) collected by LC-MS/MS were imported into Compound Discoverer 3.1 (Thermo Fisher Scientific, USA) for data processing, including peak extraction, retention time correction, background peak labeling, and metabolite identification. We calculate the coefficient of variation of the relative peak area in all QC samples, and delete the compounds with coefficient of variation greater than 30%. The identification of metabolites was a combined result of the BGI Metabolome Database (BMDB), mzCloud and ChemSpider (Human Metabolome Database (HMDB), Kyoto Encyclopedia of Genes and Genomes (KEGG), LipidMaps) databases. Main parameters of metabolite identification: Precursor Mass Tolerance <5 ppm, Fragment Mass Tolerance <10 ppm, RT Tolerance <0.2 min. The identification level of metabolites was divided into five confidence levels, and the credibility of Level 1 to Level 5 decreased in order. The original data exported by LipidSearch were imported into metaX for data preprocessing and subsequent analysis (42). Multivariate statistical analysis (principal component analysis (PCA) and partial least squares-discriminant analysis (PLS-DA)), and univariate analysis (fold-change, FC and Student’s t test) were combined to screen for differential metabolites between groups. Differential metabolite screening conditions: 1) variable projected importance (VIP) ≥ 1, 2) fold-change ≥ 1.2 or ≤ 0.83, 3) p-value <0.05. Metabolic pathway enrichment analysis of differential metabolites was performed based on the KEGG database.

**Urine DNA extraction and 16S sequencing**

Microbial genomic DNA extraction was performed as described previously (43). Urine microbial DNA was extracted using a QiaGen Mini Kit (Qiagen, Hilden, Germany) following the manufacturer’s instructions. Primers targeting the hypervariable V3+V4 region of the 16S gene were used to amplify the extracted DNA samples (the forward primer was 5′- ACTCCTACGGGAGGCAGCA -3′, and the reverse primer was 5′- GGACTACHVGGGTWTCTAAT -3′). All samples were sequenced via Illumina HiSeq 2500.

**Sequencing data analysis**

Cutadapt v2.6 software was used to process the raw data to obtain fragments of the target region. FLASH (Fast Length Adjustment of Short reads, v1.2.11) was used for sequence splicing, and UCHIME (v4.2.40) software was used to remove chimeras. Sequences were clustered with a 97% similarity level by using USEARCH (v7.0.1090_i86linux32) to cluster the spliced tags into OTUs. The OTU representative sequences were aligned with the database for species annotation by RDP classifier (1.9.1) software, and the confidence threshold was set to 0.6. The VennDiagram package of R (v3.1.1) software was used to display the number of common and unique OTUs for each group. Principal coordinate analysis was performed using QIIME (v1.8.0) software to present similarities or differences in data. Line Discriminant Analysis (LDA) Effect Size (LEFSE) was used to calculate the differences in species abundance between the two groups and then to research the biomarkers related to ATB-DILI.

**Statistical analysis**

Differences between two groups were compared by using Student’s t test for normal continuous variables and χ²-test for categorical variables. Differences with a p value <0.05 (two-
sided) were considered statistically significant. Statistical analyses were performed using SPSS V.21.0 for Windows (SPSS, Chicago, Illinois, USA). Moreover, correlations between the microbiota and metabolites and between the metabolites and clinical parameters were analyzed. Integrating electronic medical records, metabolomic and microbiome data, and machine learning methods was used to establish a clinical prediction model of ATLI. We used four machine learning algorithms: random forest, artificial neural network, support vector machine (SVM) with the linear kernel (SVM-linear), and SVM with radial basis function kernel (SVM-rbf) (44). The stratified sampling method was used to divide the training set (80%) and the validation set (20%), and the R4.1.2 software (R Foundation for Statistical Computing, Vienna, Austria) was used for data screening and model building. The importance of each feature in the occurrence of ATB-DILI was scored, and area under receiver operating characteristic (ROC) curves were employed to assess the accuracy of the models.

### Results

#### Baseline characteristics

A total of 74 patients diagnosed with TB were recruited for this study from March 2021 to December 2021 at West China Hospital of Sichuan University (Sichuan Province, China). Finally, 5 patients were lost to follow-up. Of the remaining 69 patients, 16 (23.19%) developed ATB-DILI after antituberculosis treatment (Supplementary Figure 1). The general clinical characteristics of the two groups and the results of liver function tests when DILI occurred are shown in Table 1. Compared with the non-ATB-DILI group, the levels of albumin (43.2(40.1-44.4) g/L vs. 44.9(42.2-46.8) g/L, P: 0.033) and hemoglobin (125.5(118.5-135.8) g/L vs. 137.0(128.5-145.5) g/L, P: 0.019) were significantly lower in the ATB-DILI group. No significant differences were observed in other baseline characteristics between the two groups of patients (P>0.05).

| Characteristic | ATB-DILI (n=16) | Non-ATB-DILI (n=53) | P |
|----------------|----------------|---------------------|---|
| Age, years, median(IQR) | 39.0(24.0-53.8) | 33.0(27.0-52.0) | 0.717 |
| Females, n(%) | 11(68.75) | 30(56.60) | 0.386 |
| Weight, kg, median(IQR) | 51.0(50.0-55.0) | 55.0(49.0-60.0) | 0.289 |
| BMI, kg/m2, median(IQR) | 19.9(18.9-20.9) | 20.0(18.6-21.9) | 0.771 |
| Smoking, n(%) | 1(6.25) | 7(13.21) | 0.527 |
| Drinking, n(%) | 0(0.00) | 7(13.21) | 0.210 |
| Extrapulmonary tuberculosis, n(%) | 5(31.25) | 14(26.42) | 0.704 |
| TBil umol/L, median(IQR) | 8.1(6.6-12.5) | 9.2(7.0-12.1) | 0.495 |
| ALT IU/L | 14.5(12.3-16.8) | 14.0(10.0-20.5) | 0.499 |
| AST IU/L | 21.5(15.3-26.0) | 19.0(16.0-23.0) | 0.339 |
| Alkaline phosphatase, IU/L | 74.5(57.8-89.0) | 75.0(66.5-107.0) | 0.518 |
| Glutamyltranspeptidase, IU/L | 18.0(11.8-34.3) | 19.5(11.3-32.3) | 0.750 |
| Albumin, g/L | 43.2(40.1-44.4) | 44.9(42.2-46.8) | 0.033 |
| Creatinine, μmol/L | 66.5(59.0-78.3) | 67.0(58.5-76.0) | 0.915 |
| Uric acid, mmol/L | 265.5(227.8-373.0) | 314.0(265.5-370.0) | 0.191 |
| Hemoglobin, g/L | 125.5(118.5-135.8) | 137.0(128.5-145.5) | 0.019 |
| White blood cell ×10^12/L | 6.8(5.4-8.5) | 6.0(4.9-7.5) | 0.060 |
| Platelet×10^9/μL | 242.5(182.0-417.3) | 249.5(202.0-282.5) | 0.957 |
| ESR mm/h | 20.0(10.8-83.3) | 14.0(9.3-29.3) | 0.098 |
| C-reactive protein, mg/L | 3.8(2.8-27.1) | 3.4(2.1-8.6) | 0.359 |
| Triglyceride, mmol/L | 1.1(0.7-1.5) | 1.1(0.8-1.5) | 0.889 |

**Continuous variables:**
- ALT: alanine aminotransferase; AST: aspartate aminotransferase; TBil: total bilirubin; ESR: erythrocyte sedimentation rate; BMI: body mass index; IQR: Interquartile distance.
The median time to DILI occurred on day 29 after taking anti-TB drugs (Table 1).

After excluding patients with urinary system diseases, 14 ATB-DILI patients and 30 age- and sex-matched non-ATB-DILI patients were included for urinary metabolomics and microbiome analysis (Supplementary Figure 1). It is important to note that of these 14 patients, 8 met the definition of liver adaptation (45). The other 6 patients with ALT ≥ 5 times the upper limit of normal were stopped using antituberculosis drugs according to Chinese guidelines, so it was hard to distinguish which were liver adaptation (36).

As shown in Table 2, there were no significant differences in sex, age, body weight, BMI, body mass index (BMI), smoking, drinking or tuberculosis site between the two groups of patients who underwent urine nontargeted metabolome and microbiome analysis (P>0.05). All participants had normal liver function before anti-tuberculosis drug ingestion. It was suggested that the general conditions of the two groups were consistent and comparable.

**Metabonomic analysis of urine**

PCA and OPLS-DA were performed for both positive ion mode (ESI+) and negative ion mode (ESI−). As shown in the figure (Figures 1A, B), the QC samples (blue circles) were significantly aggregated, indicating that the instrument was stable and that the reproducibility of the acquired data was good. The ATB-DILI (n=14, red circles) and tolerance groups (n=30, green circles) were not well separated in PCA. As shown (Figures 1C, D), the PLS-DA model clearly separated the ATB-DILI and non-ATB-DILI groups in both ionization modes. Differential metabolites between the two groups were screened according to multivariate and univariate statistical significance criteria (VIP≥1, FC≥1.2 or ≤0.83, and P<0.05). In general, there were 1256 urine differential metabolites screened in the positive ion mode and 334 in the negative ion mode (Figure 2). Finally, 28 differential metabolites with secondary classification names and reliable identification results (Level 1-3) were selected (Table 3), including choline, cherry base, N-acetyl, pseudohadine, N8-acetyl spermamine, glycolic acid, etc. As shown in Table 4, a total of 7 significant enrichment pathways for differential metabolites were found in both positive and negative ion modes. The differential metabolites were mainly involved in the metabolism of bile secretion, nicotinate and nicotinamide metabolism, tryptophan metabolism, ABC transporters, neuroactive ligand–receptor interaction, arginine and proline metabolism, and porphyrin and chlorophyll metabolism (P<0.05, Count≥2) (Table 4).

**Correlation analysis of metabolic and clinical data**

Correlation analysis was conducted between urine differential metabolites and clinical data, including baseline ALT, AST, TBIL, Alkaline phosphatase, hemoglobin, uric acid and albumin. We found that many different metabolites were significantly correlated with clinical data (Supplementary Table 1). The urine differential metabolite 11 dehydrothromboxane B2 was positively correlated with the baseline total bilirubin concentration, while the urine differential metabolite N8-acetyl spermidine was negatively correlated with the hemoglobin content, and uric acid was also negatively correlated with the baseline serum uric acid level (Supplementary Table 2).

**Microbiome analysis of urine**

As shown in the Figure (Figure 3A), 1079 OTUs were shared between the ATB-DILI group and the non-ATB-DILI group, 607 OTUs were unique to the non-ATB-DILI group, and the other 189 OTUs were unique to the ATB-DILI group. The Shannon curve (Figure 3B) shows that the amount of sequencing data in this study was large enough to reflect the vast majority of microbial information in the sample. The top 10 key species between the two groups are shown in Figure 3C. Weighted UniFrac principal coordinate analysis (PCoA) was applied to detect the changes in microbial community structures

| Characteristic                      | Non-ATB-DILI group (n=30) | ATB-DILI group (n=14) | P     |
|-----------------------------------|--------------------------|----------------------|-------|
| Age, years, median(IQR)           | 33.0 (27.0-52.0)         | 43.5 (22.8-55.5)     | 0.772 |
| Females, n(%)                     | 16(53.3)                 | 9(64.3)              | 0.495 |
| Weight, kg, median(IQR)           | 52.8 (48.0-58.8)         | 50.0 (49.9-55.0)     | 0.495 |
| BMI, kg/m2, median(IQR)           | 19.9 (18.6-22.0)         | 19.5 (18.7-20.1)     | 0.473 |
| Smoking, n(%)                     | 2(6.7)                   | 1(7.1)               | 0.976 |
| Drinking, n(%)                    | 2(6.7)                   | 0(0.0)               | 0.314 |
| Extrapulmonary tuberculosis, n(%) | 23(76.7)                 | 11(78.6)             | 0.888 |

IQR, Interquartile distance.
The results indicate that the ATB-DILI group and the control group were significantly separated along the PC2 axis, which explained 19.79% of the total variation. LEFSE analysis was used to determine the key attribute differences between the two groups. The differential microbiota (LDA score>2) screened between the two groups were Negativicoccus and Actinotignum, which were all upregulated in the ATB-DILI group (Figure 3D).

**Correlation of the urine microbiota and metabolism**

We further investigated the correlation of urinary differential metabolites with altered urinary microbiota. Significant correlations were found between some differential metabolites and microbial groups by calculating rank correlation coefficients (Figure 4 and Supplementary Table 2). Carbendazim was positively correlated with synergistia but negatively correlated with mollicutes (p<0.05) (Supplementary Table 2). D-(-)-lyxose was positively correlated with four microbial groups, including synergistia, ktedonobacteria, fibrobacteria and fusobacteria (p<0.05) (Supplementary Table 2). Altogether, these results showed that distinctive metabolites were closely related to urinary microbiome variation, and distinctive metabolites and microbiomes were closely related to the occurrence of ATB-DILI.

**Subgroup analysis**

According to RUCAM criteria, metabolome and microbiome analysis were performed between the ALT≥5 ULN group and normal patients. Significant metabolic differences were observed between the two groups, there were 1122 different metabolites were screened in positive ion mode and 386 in negative ion mode (Supplementary Figure 3). Finally, 26 different metabolites were selected, including choline, 11-dehydrothromboxane b2, and N8-acetylspersmidine. (Supplementary Table 3). Consistent with our results in Section 3.2, 8 common different metabolites were found in the
subgroup analysis to be related to liver injury after medication, especially when ALT > 5 ULN occurred (Table 3 and Supplementary Table 3). The eight differential metabolites were choline, N8-acetylspermidine, carbendazim, N-acetylputrescine, 1-methylnicotinamide, creatine, porphobilinogen, and nonanoic acid (Table 3 and Supplementary Table 3). And these different metabolites had the same label direction in the two groups of patients with liver injury (Table 3 and Supplementary Table 3).

There were 795 OTUs shared between the DILI patients with ALT ≥ 5 ULN and normal patients, 68 OTUs were unique to the DILI group, and the other 891 OTUs were unique to the control group (Supplementary Figure 4-A). The top 10 key species between the two groups are shown in Supplementary Figure 4-B. Finally, 3 differential microbiotas (LDA score > 2) were found between the two groups (Supplementary Figure 4-B). The Actinotignum was down regulated in DILI group, while the Bradyrhizobiaceae, and Bradyrhizobium were upregulated in the non-DILI group (Supplementary Figures 4-C). Combined with the results in Section 3.4, we have sufficient evidence to show that Actinotignum was closely related to the occurrence of liver injury after medication, regardless of the DILI standards.

Comparison of the models for the prediction of ATB-DILI

Random forest analysis was performed on the screened differential metabolites (Table 3), differential microbiota (Figure 2), and relevant clinical data of 44 patients. For clinical characteristics, we included albumin and hemoglobin, which were significantly different between the two groups, as well as other factors that may be associated with the occurrence of ATB-DILI (including age, sex, BMI, baseline ALT, AST, and TBil). A total of 38 variables are included. When ntree=500 and mtry=6, the model reaches the optimum. The score of the 38 variables was shown in Figure 5A. The larger the absolute value is, the greater the importance of the indicator. After sorting the variables from high to low according to the absolute value, the cross-validation curve was obtained by performing tenfold cross-validation repeated 5 times (Supplementary Figure 5). The top 10 variables were selected for model building with the lowest error (Supplementary Figure 5). The area under the ROC curve of the four models were shown in Table 5. At training set, the random forest model performed significantly better than the remaining three models (area under the curve 0.98 vs. 0.87 (ANN), 0.89 (SVM-linear) and 0.89 (SVM-rbf) (Table 5). Overall, random forest model, artificial neural network model and two support vector machine models (both SVM-linear and SVM-rbf) all have excellent prediction value for the validation set (Figure 5B and Table 5). The consistent results between the training set and the validation set indicate that those models have high accuracy for predicting the occurrence of ATB-DILI.

Discussion

Evidence that the human urine microbiome and metabolome contribute to the development of ATB-DILI is
accumulating. Thus, characterization of the urinary microbiota and metabolites in ATB-DILI is highly warranted, especially before medication. Herein, we first reported the characterization of urine metabolomics and the microbiome in patients with ATB-DILI before medication and identified key metabolites and bacteria that may be involved in the development of ATB-DILI. Meanwhile, we first proposed and successfully built four ATB-DILI clinical prediction models using our metabolomics, microbiome and clinical data.

In this study, the levels of ALT, AST, TBil or ALP were within the normal range in all enrolled patients before ingestion of anti-tuberculosis drugs. Approximately 23.2% of the patients had markedly elevated ALT and AST after ingesting anti-tuberculosis drugs. According to China’s 2019 guidelines for anti-tuberculosis treatment, a total of 22 patients were diagnosed with ATB-DILI. Clinical details of the patients are shown in Table 1. We subsequently investigated the urinary metabolomics and microbiota in ATB-DILI patients before medication and compared the results with healthy controls.

TABLE 3  Identified differential metabolites between two groups.

| Name               | MW   | RT  | VIP  | FC   | P       | Label |
|--------------------|------|-----|------|------|---------|-------|
| Choline            | 103.1| 0.7 | 3.3  | 1.79 | 0.003   | Up    |
| Trigonelline       | 137.0| 0.7 | 2.1  | 3.19 | 0.016   | up    |
| N-acetylputrescine | 130.1| 0.7 | 1.8  | 1.23 | 0.009   | up    |
| Pseudoephedrine    | 165.1| 3.3 | 2.5  | 0.01 | 0.009   | down  |
| N8-acetylspermidine| 187.2| 0.7 | 2.4  | 1.43 | 0.004   | up    |
| Glycocholate       | 465.3| 8.0 | 2.2  | 4.06 | 0.013   | up    |
| Uric acid          | 168.0| 1.0 | 1.9  | 1.66 | 0.021   | up    |
| Eagonine           | 185.1| 4.9 | 1.8  | 1.96 | 0.028   | up    |
| 1-methylnicotinamide| 136.1| 0.8 | 1.8  | 2.09 | 0.012   | up    |
| 6-methylnicotinamide| 143.1| 3.5 | 1.6  | 0.63 | 0.035   | down  |
| Sebacic acid       | 202.1| 6.4 | 1.6  | 1.97 | 0.049   | up    |
| Picolinic acid     | 123.0| 3.5 | 1.5  | 0.64 | 0.012   | down  |
| 3-hydroxyanthranilic acid | 153.0| 2.8 | 1.1  | 1.26 | 0.049   | up    |
| Mannitol           | 182.1| 0.7 | 1.9  | 1.57 | 0.022   | up    |
| Carbendazim        | 191.1| 0.7 | 1.6  | 0.15 | 0.023   | down  |
| Liposomamide       | 205.1| 4.0 | 1.4  | 1.78 | 0.033   | up    |
| Ophthalmic acid    | 289.1| 2.4 | 1.4  | 0.47 | 0.041   | down  |
| Valerenophene      | 162.1| 5.5 | 1.2  | 0.71 | 0.030   | down  |
| D-(-)-lyxose       | 150.1| 0.7 | 1.0  | 0.77 | 0.030   | down  |
| Cretine            | 131.1| 0.7 | 1.6  | 0.32 | 0.030   | down  |
| L-glutamic acid    | 147.1| 0.7 | 1.1  | 0.71 | 0.029   | down  |
| Methylmalonic acid | 118.0| 0.7 | 1.9  | 0.46 | 0.014   | down  |
| Porphobilinogen    | 226.1| 0.7 | 1.5  | 0.49 | 0.001   | down  |
| Epiinephrine       | 183.1| 3.9 | 2.2  | 2.44 | 0.008   | up    |
| Heptanoic acid     | 130.1| 5.4 | 1.4  | 0.50 | 0.047   | down  |
| 11-dehydrothromboxane b2 | 368.2| 6.8 | 1.1  | 1.40 | 0.040   | up    |
| Nonanoic acid      | 158.1| 6.9 | 1.5  | 0.51 | 0.004   | down  |
| Taurolithocholic acid 3-sulfate | 563.3| 7.8 | 2.2  | 1.53 | 0.017   | up    |

VIP, variable important for the projection; FC, fold-change; MW, molecular weight; RT, retention time.

TABLE 4  Differential metabolite pathway analysis.

| Pathway                          | Ion modes | Count | Count All | P    |
|----------------------------------|-----------|-------|-----------|------|
| Bile secretion                   | positive  | 4     | 97        | <0.001|
| Nicotinate and nicotinamide      | positive  | 2     | 55        | 0.001|
| Tryptophan metabolism            | positive  | 2     | 81        | 0.002|
| ABC transporters                 | positive  | 2     | 124       | 0.005|
| Neuroactive ligand-receptor       | negative  | 2     | 52        | <0.001|
| Arginine and proline metabolism  | negative  | 2     | 78        | <0.001|
| Porphyrin and chlorophyll        | negative  | 2     | 142       | 0.003|
the diagnosis and treatment of ATB-DILI (36), for DILI caused by anti-tuberculosis drugs, when ALT ≥ 3ULN or TBil ≥ 2ULN, the relevant anti-tuberculosis drugs need to be discontinued, and when ALT ≥ 5ULN or TBil ≥ 3ULN, it is necessary to stop all anti-tuberculosis drugs. Indicates that DILI needs to be taken seriously in TB patients. Therefore, we first analyzed the characteristics of the metabolomics and microbiome of DILI patients with ALT ≥ 3 ULN. As a large number of domestic and foreign studies both recommend the use of RUCAM to assess DIL (46–48), we did a subgroup analysis for those DILI was defined as serum ALT level ≥5ULN. What was exciting was that no matter which DILI standard, we have found the same differential metabolites and microorganisms.

Metabolomics and the microbiome were used to analyze the urine of the ATB-DILI susceptible group and normal liver function control group, and the two groups could be distinguished significantly on the PLS-DA scatter plot. Consistent with those of a previous study (23, 24, 26), our results also indicated that ATB-DILI susceptible individuals may have specific metabolomic and microbiological patterns. We identified 28 major differential metabolites between the two groups in urine, including choline, trigonelline, N-acetylputrescine, uric acid, etc. The biological properties of each metabolite were searched from the human metabolome database (https://hmdb.ca/), and summarized in Supplementary Table 4. The differential metabolites selected in this study were consistent with the biospecimen locations in the database. This process involves bile secretion, nicotinate and nicotinamide metabolism, tryptophan, ABC transporters, neuroactive ligand-receptor interaction, arginine and proline metabolism, porphyrin and chlorophyll metabolism. Two major differential microbial, Negativicoccus and Actinotignum, were identified between the two groups.

As an essential nutrient, choline in the urine of patients with overactive bladder was 34.8% lower in urine metabolomic analysis than patients without overactive bladder (P = 0.014) (49). The urinary excretion of choline metabolites in term breast-fed infants was significantly higher than that in term formula-fed infants (P < 0.05) (50). This study found that urinary choline levels may be a noninvasive biomarker for predicting ATB-DILI. For the first time, upregulated trigonelline in urine before medication was found to be associated with ATB-DILI in TB patients. This may be related to the possible inhibition of key enzymes in lipid metabolism and absorption by trigonelline (51). Previous studies have found that glycocholic acid levels were significantly increased in DILI (52, 53), and the increased levels were positively correlated with the severity of DILI (52). Combined with the results of this study, glycocholic acid may have a role as a biomarker for DILI. As a proinflammatory and proapoptotic molecule, uric acid
plays an intermediary role in the process of liver and kidney injury (54, 55), and animal experiments have shown that the elevation of uric acid may lead to alcohol-induced steatosis, endoplasmic reticulum stress, and cell apoptosis, death and liver damage (56). Cao et al. found that uric acid levels in urine can be used to differentiate ATB-DILI from non-ATB-DILI patients (24). In this study, we found that uric acid in urine generation before medication could be used as a biomarker to predict the occurrence of ATB-DILI after medication, indicating that uric acid in urine metabolism may have great potential in predicting and identifying ATLI.

Additionally, differential metabolite enrichment analysis showed that metabolic pathways, including bile secretion, niacin and nicotinamide metabolism, ABC transporters, and etc., were involved in the occurrence of ATB-DILI after medication, indicating that uric acid in urine metabolism may have great potential in predicting and identifying ATLI.

Each disease has its own unique microbial alterations (29, 62). Microorganisms in the gut originate from the digestive system, while urine microorganisms reflect the entire body including the intestinal tract, oral system, respiratory system, etc (63). Studies have shown that the urinary microbiota is associated with diseases outside the urinary system (64, 65). Previous studies have indicated that microbiota alterations are associated with drug-induced liver injury (26, 32, 33). Our previous study found that six microbiota including o_Bacteroidales, f_Prevotellaceae, etc., were associated with ATB-DILI (26). Compared with control group, this prospective study found that the Negativicoccus and
Actinotignum were upregulated in the ATB-DILI group before medication. Negativicoccus was found to be significantly increased in the oral cavity of hamsters using smokeless tobacco products (66). Among patients with nonmuscle-invasive bladder cancer, BCG-vaccinated patients had significantly more negativicoccus in their urine than nonvaccinated BCG patients (67). Negativicoccus was also found to be one of the core flora in all ground glass nodules and normal tissue samples (68). However, studies have proven that Negativicoccus and Actinotignum are associated with ATB-DILI.

Furthermore, our results also suggested that there may be specific metabolomic and microbiological patterns in individuals.

![Figure 5](image)

**Figure 5**
Machine learning models. (A) Score the importance of variables. The larger the value, the more important the variable is. (B) Receiver operating characteristic curve for the models developed with the top 10 important variables as inputs. ALT, alanine aminotransferase; AST, aspartate aminotransferase; TBIL, total bilirubin; ALB, albumin; BMI, body mass index; HB, hemoglobin.

| Model          | AUC Training set | Test set |
|---------------|------------------|----------|
| RF            | 0.98             | 1        |
| ANN           | 0.87             | 1        |
| SVM_linear    | 0.89             | 1        |
| SVM_rbf       | 0.89             | 1        |

AUC, area under the curve; RF, random forest; ANN, artificial neural network; SVM, support vector machine; SVM_rbf, support vector machine with radial basis function kernel.
susceptible to severe ATB-DILI when compared with the mild ATB-DILI group. The discovery of these biomarkers may help with the early identification of TB patients at risk of developing severe DILI, thus providing new ideas for the individualized treatment of TB. However, due to the limited sample size, the results of this study cannot be directly generalized to other populations.

This study is the first to establish the early prediction models of ATB-DILI by combining clinical data and metabolomics data using a machine learning method. The random forest algorithm was used to analyze multiple variables, the importance of each variable was scored, and the optimal variable (top 10) combination was obtained by adjusting the parameters to form the ATB-DILI prediction models. The results of the training set and the validation set were consistent (all ROC ≥ 0.85) (Table 5). Based on clinical and genomic data, researchers from Taipei Medical University compared the accuracy of multiple machine learning methods in predicting ATB-DILI, among which the artificial neural network showed the best prediction performance (69). In their study, the area under the ROC curve of the training set in the random forest algorithm was 0.724 and 0.718 for the validation set (69). Combined with our study, machine learning techniques show great potential in predicting ATB-DILI and may provide new opportunities for the diagnosis and treatment of ATB-DILI.

This study has some limitations. First, the number of participants was limited. However, this was a prospective study, which enhanced reliability of the results. Further validation in more centers with more patients needs to be verified in the future. Second, even though this study adds to the understanding of metabolome and microbiological patterns on the progress of ATB-DILI, this study only analyzed predose characteristics and lacked data at multiple time points after drug use. There is much work yet to be performed to understand these changes entirely. Finally, the current study obtained good predictive value in both the training set and the validation set, but limited by the limited sample size and geographical limitations, further verification is required in studies with more regions and larger samples in the future.

Conclusion

In conclusion, our findings extend our knowledge of the relationship between urinary metabolites and microbiota and host ATB-DILI susceptibility, indicating that certain metabolomic and microbiome changes from the host can be used to identify and predict an individual’s susceptibility to ATB-DILI. In the future, prospective cohorts with a larger number of subjects are needed to investigate the potential clinical utility of metabolic markers and key microbiota in identifying susceptible individuals. More time points are needed to investigate the potential clinical utility of metabolic markers and key microbiota in identifying susceptible individuals.

Data availability statement

The original contributions presented in the study are publicly available. This data can be found here: https://www.ncbi.nlm.nih.gov/, PRJNA870240.

Ethics statement

The studies involving human participants were reviewed and approved by West China Hospital of Sichuan University [Approval No.: 761 (2019)]. The patients/participants provided their written informed consent to participate in this study.

Author contributions

All authors contributed substantially to the study design, data interpretation, and the writing of the manuscript. J-QH contributed to the study design. M-GW and S-QW contributed to data collection and analysis and completed the full text. M-MZ contributed to data collection. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2022.1002126/full#supplementary-material

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SUPPLEMENTARY FIGURE 3
Principal co-ordinates analysis.

SUPPLEMENTARY FIGURE 4
Subgroup analysis of 165 sequencing data of urine samples. (A) Venn diagram. The left is the severe DILI group, the right is the non-DILI group. (B) Difference comparison of the top 10 key species. (C) LEfSe analysis. Species with LDA greater than the set value of 2 are presented. The length of the bar indicates the magnitude of LDA influence.

SUPPLEMENTARY FIGURE 5
Cross validation curve. The abscissa is the number of variables, and the ordinate is the cross-validation error rate.
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