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Scientific Appraisal and Therapeutic Properties of Plants Utilized for Veterinary Care in Poonch District of Jammu and Kashmir, India

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Simple Summary: In the rural areas of the Himalaya, ethnobotanical knowledge is crucial for preserving plant diversity and treating a variety of diseases. Ethno-veterinary medicines can provide leads for drug development, but probably a more practical and lucrative exercise would be to develop a preferred preparation by pharmacological research, and the ensuing medication can be returned to the society with extra impact. Further, this valuable knowledge base has now become obsolete due to industrialization, urbanization and, above all, lack of recognition by the younger generations. Therefore, there is a dire need to review, document and authenticate the valuable traditional knowledge of ethno-medicinal plants for human welfare. By comprehending the traditional knowledge system, the current study conducted in Jammu and Kashmir, India, could serve as a pilot to document the sustainable exploitation of regularly utilized bioresources and would provide crucial leads for the manufacturing of pharmaceuticals and medicines.

Abstract: The importance of traditional and indigenous knowledge is acknowledged on a worldwide scale for its coexistence principles and sustainable use techniques. In view of this, the present study is an attempt to document the ethno-veterinary plants used by the tribal communities of Western Himalaya. This study also provides the scientific validation of herbal medicines used in ethno-veterinary practices through a reverse pharmacological approach. A total of 59 informants were selected through a non-probability sampling method. Detailed information on the medicinal plants used in ethno-veterinary practices along with their habits and habitats, part/s used, remedy preparation methods, additives/ingredients used during preparation and administration, dosages administered, and route of administration was collected. Data was analyzed for the Relative Frequency of Citations (RFC), Use Values (UV), Informant Consensus Factor (ICF), and Jaccard Index (JI). Further, a reverse pharmacological approach was used for scientific validations of the documented herbal knowledge of plant species. During the study, 56 plant species belonging to 54 genera and 39 families were documented. Asteraceae was the dominant family followed by Lamiaceae, Amaranthaceae and Fabaceae. Life forms were dominated by herbaceous species and leaves were the most common plant parts used. The highest Relative Frequency of Citations (RFC) and Use Values (UV) were recorded for Brassica napus L. (Brassicaceae). The Pearson correlation coefficient between RFC and UV shows a strong positive correlation between the proportion of uses of a plant species within a sample of informants and the number of times that a particular use of a plant species was mentioned by the informant.
Studies of the biological activity of ethno-veterinary plants can provide clues of promising leads for the isolation and identification of useful compounds that may be developed into pharmaceuticals for human welfare.

**Keywords:** animal care; informant consensus; Jaccard index; reverse pharmacology; phytochemicals

1. Introduction

India has one of the primeval and sophisticated healthcare systems with a history of more than 5000 years [1]. This priceless information came together over the course of many centuries based on different traditional healthcare systems such as Rigveda, Atharveda, and several post Vedic treatises such as Charakasamhita, Dhanwanthari, and Nighantu [2]. Materia Medica provides a huge knowledge base of traditional healthcare systems of India [3]. Approximately, 25,000 plant-based formulations are used as traditional healthcare systems by different rural communities of India [4], and out of these, only 5–10% have been validated scientifically [3]. In various communities across India, the use of plant-based formulations has long been a crucial component in the treatment of various illnesses and has led manufacturers to produce a large number of medication candidates that are currently widely used in commercial markets [5]. However, various ethnomedicinal plants used in Indian healthcare systems have been investigated scientifically for nearly three decades [6]. Since then, the Government of India has made several attempts to investigate the possibility of evaluating these systems for their therapeutic potential as they were originally practiced, as well as to generate data to include them in national healthcare programs. This is due to the growing interest worldwide in adopting and studying traditional systems and in utilizing their potential from various healthcare perspectives [7]. According to the WHO report (2018), 66% of India’s population resides in rural areas with a large dependency on agriculture. In the Himalayan region, the rearing of animals to supplement the family income and sustain crop production constitutes an important component of the rural economy of the region [8–10]. In the mixed crop–livestock farming systems, livestock and food production systems are closely integrated in the Himalaya [11]. The Himalayan region’s rural economy is based mostly on animal husbandry, and the growth of this industry could raise the standard of living in rural areas. Farmers look after their livestock by using ethno-veterinary practices [12]. To keep domestic animals healthy and productive, ethno-veterinary remedies are widely and very successfully utilized for basic healthcare. People in remote rural areas continue to rely heavily on herbal and common domestic remedies to treat veterinary ailments. The system comprises conventional beliefs, knowledge, expertise, approaches, practices and traditions of a particular society [13,14]. Through a process of experience spanning hundreds of years, the traditional communities have identified the folk knowledge of ethno-veterinary medicine and its relevance. One of the most crucial prevailing systems in the region where modern veterinary healthcare facilities are scarce or in extremely bad condition is the traditional method of treatment [15]. Traditional herbalists (Pashu Vaidyas) pass down their native knowledge of the veterinary healthcare system orally from one generation to the next [16,17]. Thus, documentation of ethno-veterinary medicinal knowledge can generate leads for drug development, but on the basis of modern scientific approaches, there is inadequate data available on the evaluation and validation of this folk wisdom. One of the modern scientific approaches to validate the traditional knowledge of ethnic groups is reverse pharmacology. It is a multidisciplinary procedure for connecting traditional knowledge bases to promising research strategies, tools, technologies and innovations [18]. A notable component of this methodology is the blending of information gained from traditional or folk medicine with the advanced technology to guarantee better and more secure leads [19]. The attempt to study firm findings would assist not only in the recognition of the candidate drugs but also in the understanding of their underlying molecular mechanisms [20]. Reverse
pharmacology has an immense scope to evaluate the efficacy and quality of traditionally used medicines clinically and, furthermore, to identify new drugs from the natural products used in traditional medicinal systems since times immemorial [18].

In the Himalayan region, people have strong a belief and faith in traditional herbal medicinal practices for healthcare, and therefore, the plants have immense cultural and medicinal significance within the region [21]. Nomadic tribes and pastoral communities residing within the Northwest and Trans-Himalaya are reputed to have mastered their conventional practices and knowledge regarding the ethno-medicinal and ethno-veterinary utilization of plants [22,23]. Jammu and Kashmir (J&K) is home to several tribal communities that are scattered throughout the region and form an inherent part of its culture and tradition [24]. The Gujjars and Bakerwals, seminomadic and nomadic tribes respectively, are the third-biggest ethnic groups in J&K, constituting more than 11.9% of the total population, and have maintained their culture and heritage throughout the ages [25]. These tribals depend on cattle rearing for their livelihood, and since they feed their livestock in the upper reaches of mountain terrain, they do not have easy access to modern veterinary medicine or doctors. They depend solely on folk medicine and herbal remedies for treating animal diseases. Oral transmission has been the only medium through which the traditional knowledge regarding the use of plants for healing purposes has been passed from generation to generation [26]. Lack of recognition by the younger generation has led to the decline of this valuable knowledge-base, and now it has become obsolete [27]. Furthermore, the indigenous knowledge of the use of lesser-known plants is also rapidly declining. Many researchers have documented the ethno-medicinal plants of J&K [28–36]; however, very limited attempts have been made to document the traditional herbal knowledge of veterinary systems. There is a dire need to review and document the valuable traditional knowledge regarding the use of ethno-veterinary plants, especially in remote and far-flung areas. In view of this, the present study aimed to provide a contemporary compilation of medicinal plants used for the treatment of animal diseases by the tribal communities of the district Poonch, Jammu and Kashmir, through quantitative ethnomedical analysis. Further, the study attempted to validate the remedies through reverse pharmacological correlations.

2. Materials and Methods

2.1. Study Area

Poonch, renowned as ‘Mini Kashmir’ is one of the far-flung and backward districts of J&K situated on the Line of Control (LOC) located at 33.77° N and 74.1° E (Figure 1), and is delimited by the Actual Line of Control (ALC) from three sides. The Pir Panjal range of mountains separates Poonch valley from the Kashmir valley by the Pir Panjal Range. It is bordered by Baramulla, Budgam, Shopian and Kulgam Districts of Kashmir in the north east, Rajouri in the south and Azad Kashmir (Pakistan) in the west. Topographically, Poonch district is hilly and mountainous, exclusive of a few low-lying valleys. The vegetation of the study area is largely influenced by monsoon rainfall and varies from the humid zone to the temperate zone. Although people from many different linguistic groups live in the area, Gujjars and Bakerwals predominate. Both tribes have a similar ethnic make-up, speak the same language (Gojri), and rely on the same ecology to meet their daily requirements. The only distinction is that Gujjars raise buffalo and cows, whilst Bakerwals raise sheep and goats. The economic condition of people residing in the study area is not satisfactory and depends on herbal medicines for human and animal health care. Both groups have access to herbal healers to treat their common health issues, and oral transmission is the sole way this traditional knowledge is passed along.
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Figure 1. Map showing the location of the study area.

2.2. Data Collection

A field assessment, which involved plant collection, photography, and data recording, was carried out between March 2019 and December 2020 following Heinrich et al. [37]. A total of 59 informants were selected through non-probability sampling, using convenience sampling methods based on easy access, availability, and relevance of informants. Out of the total 59 informants, 38 were males and 21 were females. The age group of the informants varies from 20 to 70 years, with a different level of education, from illiterate to above higher secondary level. The information of the respondents has been collected through questionnaires and direct interviews. Prior to interviewing and having discussions, formal written ethical consent for the study was obtained from the local tribal committee (vide number SPHC-B/108; Dated 19 February 2019) and oral consent from all participating informants was also given. Open semi-structured questionnaires developed by following Edwards et al. [38] with some modifications were used for collecting ethnobotanical data, as this approach permits several respondents to be cross-examined in a comparatively short period by posing the same questions within a flexible structure. Interviews were based on a checklist of questions set earlier in the English language and concurrently translated into Gojri, the local language, following Negi et al. [39]. Interviews focused on the demographic features of participants, including gender, age, matrimonial status, educational background, and the extent of time that an informant spends in the study area in a year. The foremost
A segment of the interviews focused on the local names of medicinal plants used, their life forms and habitats, part/s used, modes of remedial preparations, additives and ingredients used in the preparations, and dosages required. Besides the collection of data in written form, photographs and audio recordings were also taken for thorough analysis of the data and later verification following Ribeiro et al. [40].

Local guides/personnel were hired for the collection of plant specimens from the field. Fresh plant samples were collected from the field and were pressed, dried, sprayed with 1% mercuric chloride (preservative) and mounted on herbarium sheets following routine herbarium practices [41]. Collected plant samples were identified following regional floras [42–44], and the botanical nomenclature of the collected species was authenticated using the Plants of the World Online database (http://www.plantsoftheworldonline.org accessed on 21 March 2022).

2.3. Quantitative Data Analysis

The Microsoft Office Excel spreadsheet (2010, Microsoft Corporation, Redmond, WA, USA) was used for data scrutinization, computation, and drawing graphs. Some of the graphs were prepared using ‘ggplots2’ package in R statistical software (v 4.0.3; R Development Core Team 2021). The following quantitative and similarity metrics were used to analyze the data:

(A). Relative Frequency of Citation (RFC): RFC was calculated by using the following formula

\[
RFC = \frac{FC}{N}
\]

where FC is the number of informants reporting the use of a particular species and N is the total number of informants.

(B). Use Value (UV): Use value is an index in ethnobotany that has been commonly used to enumerate the relative significance of useful plants [45,46]. It is the relative importance of a particular plant species used by the indigenous inhabitants and was calculated following Phillips et al. [47]

\[
UV = \frac{\sum U_i}{N}
\]

where ‘U_i’ is the number of use-reports mentioned by each participant for the particular species and ‘N’ is the total number of informants. The use value ranges from 0 to 1. UV does not help in distinguishing whether a plant is used for single or multiple purposes [48].

(C). Informant Consensus Factor (ICF): To assess consistency of familiarity with the medicinal plants, the Informant Consensus Factor (ICF) was used by following [49]. Prior to analysis, all the ailments were categorized following Heinrich et al. [49] and Bhatia et al. [28]. The ICF was calculated by using the formula

\[
ICF = \frac{n_{ur} - n_t}{n_{ur} - 1}
\]

where ‘n_{ur}’ is the number of use reports and ‘n_t’ is the number of taxa utilized for a particular use category by all informants. ICF values range from 0.00 to 1.00. When only one or a few taxa are accounted to be used by a high proportion of informants for curing a particular illness, ICF values are higher, whereas low ICF values indicate that informants disagree about which plant to use and prefer different plants for curing a particular disease [49]. In the present study, eight disease categories were identified (Figure 2) and the ICF was calculated for each disease category.
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Figure 2. Disease categories for calculation of ICF.

(D). Jaccard Index (JI): In order to compare the present study with similar previously published literature and to access comparison of knowledge among different communities, the Jaccard Index was calculated using the following formula [50–52]

\[ JI = \frac{c \times 100}{(a + b) - c} \]

where “a” is the number of species of area A (our study area); “b” is the number of species of the neighboring area B; “c” is the number of species common to both A and B.

2.4. Statistical Analysis

The Pearson correlation between the RFC and UV was determined using SPSS version 26.0 (IBM, Armonk, NY, USA); \( r^2 \) was also computed in order to find out the cross-species inconsistency in RFC clarified by the discrepancy in UV following Amjad et al. [50].

2.5. Reverse Pharmacological Correlations

Eclectic screening of the relevant literature was carried out to review the pharmacology and phytochemistry of each documented plant species to validate these traditional ethnoveterinary medicines. Data were retrieved from Google Scholar, Science Direct, PubMed, Scopus, SciFinder and Web of Science by searching for the following keywords: phytochemicals, phytochemistry, biologically active compounds, biological activities, pharmacology, and medicinal uses in combination with each plant species.

3. Results

3.1. Demography of the Informants

Out of the total 59 informants, 64.40% were males and 35.59% were females. Based on age, the informants were divided into three age groups: 20–35 years (25.42%), 36–50 years (38.98%), and above 50 years (35.59%). Out of the total informants, 37 were married and 22 were unmarried. Concerning education, 32.20% were illiterate, 22.03% had attended school up to primary level, 16.95% up to middle level, 13.56% up to secondary level, 8.47% up to intermediate level, and 6.78% above higher secondary level (Table 1). The informants belong to two tribal groups: Gujjars (35 informants) and the Bakerwals (24 informants). The duration of time spent by the informants in the study area varies and it was found that Gujjars spend more time in the forest ecosystem for utilizing natural resources and either did not migrate or migrated within the district; Bakerwals are nomadic and
migrate to other areas such as Kashmir valley during summer, and thus spend less time in the study area.

Table 1. Demographic data of informants.

| S. No. | Variable             | Category | No. of Informants | % Age |
|--------|----------------------|----------|-------------------|-------|
| 1      | Gender               | Male     | 38                | 64.40 |
|        |                      | Female   | 21                | 35.59 |
| 2      | Marital status       | Married  | 37                | 62.7  |
|        |                      | Unmarried| 22                | 37.3  |
| 2      | Age (in years)       | 20–35    | 15                | 25.42 |
|        |                      | 36–50    | 23                | 38.98 |
|        |                      | Above 50 | 21                | 35.59 |
| 3      | Educational Qualification | Illiterate | 19       | 32.20 |
|        |                      | Primary  | 13                | 22.03 |
|        |                      | Middle   | 10                | 16.95 |
|        |                      | Secondary| 8                 | 13.56 |
|        |                      | Hr. Sec. | 5                 | 8.47  |
|        |                      | Above    | 4                 | 6.78  |

3.2. Medicinal Plant Diversity

During the present study, a total of 56 plant species belonging to 54 genera and 39 families were reported. The details of botanical names, vernacular names, families, life forms, ethno-veterinary uses, parts used, methods of use, FC, RFC, and UV are given in Table 2. Asteraceae with five species was dominant, followed by the Lamiaceae, Fabaceae, and Amaranthaceae with three species each. The remaining families contribute less than two species in the listed ethno-veterinary plants, out of which 28 families are represented by single species. The reason behind the dominance of Asteraceae, Lamiaceae, and Fabaceae in the traditional healthcare system may be due to the abundance of plant species of these families in the study area [53]. Residents of the study region have therefore been using these plants for many generations as a result of their easy accessibility, and as a result, they are familiar with the plant families. These plant families have been reported to be dominant in other ethno-botanical studies as well [54–56]. The reason for the overall dominance of the plant species belonging to these families may probably be due to the presence of secondary metabolites in them. Many active essential oils have been isolated from members of the family Lamiaceae [57]. Furthermore, plants of the family Asteraceae are known for their pharmacological importance [58–60] and this family is extensively strewn and is regarded as the largest angiosperm family in the world [61].

Life forms are dominated by herbs, followed by trees, shrubs and climbers (Figure 3). In other similar studies [62–64], the life forms are dominated by herbaceous plants. The dominance of herbs in conventional and their use indigenous medicinal systems may be due to their ease availability, high therapeutic potential, and presence of biologically active phytochemicals [65–67]. Further, the presence of soft tissues in herbaceous plants makes their extractions and preparations uncomplicated and easy. Most of the documented plant species grow in the wild, and few (Allium cepa, Allium sativum, Foeniculum vulgare, Brassica rapa, Lagenaria siceraria, Trigonella foenum-graecum, Mentha sylvestris, Oryza sativa, and Prunus persica) are cultivated. The forests, farmlands, roadsides, fallow lands and riversides are the habitats where the wild medicinal plants are found.
Table 2. Ethno-veterinary plants used by Gujjar and Bakerwal tribes in Poonch, J&K.

| Taxa/Local Name                  | Family/Voucher Number | Life Form | Ethno-Veterinary Uses | Part(s) Used | Preparation | Method of Use               | FC   | RFC | UV  |
|---------------------------------|-----------------------|-----------|-----------------------|--------------|-------------|-----------------------------|------|-----|-----|
| *Amaranthus viridis* L. {Ganar} | Amaranthaceae/BGSBU-01| H         | Tonic                 | AP           | Raw         | Aerial parts of the plant are cut into small pieces and mixed with wheat husk. This mixture is fed preferably twice a day for two weeks. | 23   | 0.38| 0.35|
| *Chenopodium album* L. {Bathua} | Amaranthaceae/BGSBU-02| H         | Wound healing         | L            | Paste       | Paste prepared from the leaves boiled in mustard oil is applied externally. | 32   | 0.53| 0.58|
| *Achyranthes aspera* L. {Phutkanda} | Amaranthaceae/BGSBU-03| H         | Fever                 | R            | Paste       | Root paste is given orally. | 39   | 0.65| 0.68|
| *Allium cepa* L. {Gandha}       | Amaryllidaceae/BGSBU-04| H         | Loss of appetite      | B            | Paste       | Part of bulbs mixed with salt is fed to the cattle. | 37   | 0.61| 0.58|
| *Angelica glauca* Edgew. {Chora} | Apiaceae/BGSBU-06     | H         | Cold                  | R            | Decoction   | Decoction of root is given to cattle thrice a day. | 31   | 0.51| 0.54|
| *Foeniculum vulgare* Mill. {Sounf} | Apiaceae/BGSBU-07    | H         | Indigestion           | AP           | Decoction   | Decoction prepared by boiling fruits in water is given to cattle. | 43   | 0.71| 0.78|
| *Calotropis procera* (Aiton) W.T.Aiton {Aak} | Apocynaceae/BGSBU-08 | S         | Removal of retained placenta | Ltx          | Raw         | Tail of buffaloes is dipped for 4–5 min into latex. | 38   | 0.63| 0.61|
| *Hedera nepalensis* K.Koch {Harbem-bel} | Araliaceae/BGSBU-09  | C         | Leech removing        | L            | Extract     | Leaf extract is put in the nostrils. | 41   | 0.68| 0.65|
| Taxa/Local Name | Family/Voucher Number | Life Form | Ethno-Veterinary Uses | Part(s) Used | Preparation | Method of Use | FC  | RFC | UV  |
|----------------|------------------------|-----------|------------------------|--------------|-------------|--------------|-----|-----|-----|
| *Taraxacum officinale* F.H.Wigg. {Handh} | Asteraceae/ BGSBU-10 | H | Enhance milk production | WP | Raw | Whole plant is fed to cattle with other feeds. | 45  | 0.75 | 0.78 |
| | | | Stretch of bones and ligaments | AP | Decoction | Decoction prepared by boiling aerial parts into water in 1:1 ratio is given for about 15 days. |     |      |      |
| *Erigeron canadensis* L. [Kuttey Haddi] | Asteraceae/ BGSBU-11 | H | Indigestion | AP | Paste | Aerial parts of the plants are crushed and the paste is fed to the cattle. | 29  | 0.49 | 0.47 |
| *Saussurea costus* (Falc.) Lipsch. [Kuth] | Asteraceae/ BGSBU-12 | H | Tonic | R | Powder | Root powder mixed with crushed onion bulbs, gur (raw sugar) and water is fed to the cattle. | 21  | 0.36 | 0.38 |
| *Sonchus arvensis* Linn. [Sonchal] | Asteraceae/ BGSBU-13 | H | Enhance milk production | WP | Raw | For increasing milk production, fresh plants are fed to cattle. | 39  | 0.66 | 0.71 |
| *Achillea millefolium* L. [Chou] | Asteraceae/ BGSBU-14 | H | Deworming | WP | Raw | Whole plants are fed to animals. | 42  | 0.71 | 0.67 |
| *Brassica rapa* L. [Sariyoon] | Brassicaceae/ BGSBU-17 | H | | Sd | Cakes | Seed cakes locally called *Khahl* is fed to lactating cows and buffaloes to enhance milk production. | 48  | 0.81 | 0.87 |
| | | | Vigor maintenance of bulls | Sd | Cakes | Mixture of seed cakes and rice husk is fed to bulls. |     |      |      |
| | | | Skin Infection | Sd | Paste | Seeds are ground and mixed with mustard oil. This paste is applied externally on infected parts for a week. |     |      |      |
| *Buxus wallichiana* Baill. [Chikhri] | Buxaceae/ BGSBU-18 | T | Skin infection | L | Decoction | Decoction of fresh or dried leaves is given orally. | 24  | 0.41 | 0.45 |
| Taxa/Local Name | Family/ Voucher Number | Life Form | Ethno-Veterinary Uses | Part(s) Used | Preparation | Method of Use | FC  | RFC  | UV  |
|-----------------|------------------------|-----------|-----------------------|--------------|-------------|---------------|-----|------|-----|
| *Cannabis sativa* L. [Bhang] | Cannabaceae/ BGSBU-19 | H         | Removal of lice and ticks | L            | Paste       | Paste of crushed leaves is applied externally. | 31  | 0.53 | 0.51 |
| *Valeriana jatamansi* Jones ex Roxb. [Balo] | Caprifoliaceae/ BGSBU-20 | H         | Fatigue                | Rh           | Powder     | Dried rhizomes are ground to fine powder which is dissolved in about 200–300 mL of normal water and is given to the cattle in the morning for about a week. | 29  | 0.49 | 0.53 |
| *Lagenaria siceraria* (Molina) Standl. [Doberi] | Cucurbitaceae/ BGSBU-21 | C         | Yoke Galls             | F            | Paste      | Fruits are burned. Ash is mixed with luke warm mustard oil and the paste is applied on yoke galls of bulls. | 22  | 0.37 | 0.31 |
| *Mallotus philippensis* (Lam.) Mull. Arg. [Kamila] | Euphorbiaceae/ BGSBU-22 | T         | Deworming              | F            | Powder     | Powdered dry fruits are mixed with flour and given to animals for 2–3 days. | 29  | 0.49 | 0.47 |
| *Ricinus communis* L. [Arand] | Euphorbiaceae/ BGSBU-23 | S         | Dysentery              | Sd           | Paste      | Seeds are crushed in small quantity, mixed with fodder and given to cattle. | 30  | 0.51 | 0.53 |
| *Trifolium pratense* L. [Shatul] | Fabaceae/ BGSBU-24 | H         | Enhance milk production | WP           | Raw        | Whole plant is fed to cattle | 45  | 0.76 | 0.79 |
| *Trifolium repens* L. [Srieh] | Fabaceae/ BGSBU-25 | H         | Enhance milk production | WP           | Raw        | Whole plant is fed to cattle | 39  | 0.66 | 0.63 |
| *Trigonella foenum-graecum* L. [Methi] | Fabaceae/ BGSBU-26 | H         | Diarrhea               | L            | Sd         | Leaves and seeds are fed to animal for 3–4 days. | 34  | 0.58 | 0.62 |
| *Juglans regia* Linn. [Khor] | Juglandaceae/ BGSBU-27 | T         | Enhancing Milk Production | F            | Cakes      | The oil cakes obtained by grinding of fruit kernels are fed to cows to enhance their milk production. | 26  | 0.44 | 0.40 |
| *Vitex negundo* L. [Banno] | Lamiaceae/ BGSBU-28 | S         | Fever                  | L            | Paste      | Young leaves are crushed and given orally. | 24  | 0.41 | 0.38 |
| *Ajuga pareiflora* Benth. [Jan-i-adam] | Lamiaceae/ BGSBU-29 | H         | Weakness               | L            | Infusion   | Water extract of fresh leaves is given to cattle | 30  | 0.51 | 0.53 |
|                         |                        |           | Indigestion            | L            | Decoction  | Decoction is given to animals orally | 30  | 0.51 | 0.53 |
|                         |                        |           | Fever                  | L            | Decoction  | Decoction is given to animals orally | 30  | 0.51 | 0.53 |
### Table 2. Cont.

| Taxa/Local Name | Family/Voucher Number | Life Form | Ethno-Veterinary Uses | Part(s) Used | Preparation | Method of Use | FC  | RFC | UV |
|-----------------|-----------------------|-----------|-----------------------|--------------|-------------|--------------|-----|-----|----|
| Mentha sylvestris L. [Pootno] | Lamiaceae/BGSBU-30 | H | Deworming | L | Raw | Leaves are fed to live stock | 27  | 0.46  | 0.41 |
| Punica granatum L. [Daruno] | Lythraceae/BGSBU-31 | T | Jaundice | F | Decoction | Decoction of fruit exocarp is given orally. | 37  | 0.63  | 0.71 |
| Malva neglecta Wall. [Sonchal] | Malvaceae/BGSBU-32 | H | Constipation | L | Paste | Soaked leaves are crushed, mixed with cow-butter and fed to newly born calves. | 28  | 0.47  | 0.53 |
| Trillium govanianum Wall. ex D.Don [Traepatri] | Melanthiaceae/BGSBU-33 | H | Deworming | Rh | Paste | Crushed rhizome is given to the cattle. | 16  | 0.27  | 0.21 |
| Melia azedarach L. [Dreck] | Meliaceae/BGSBU-34 | T | Foot and mouth disease | L | Paste | Fresh leaves are crushed with sugar and water. The paste so formed is given orally to the cattle for 2–3 days. | 31  | 0.53  | 0.48 |
| Cissampelos pareira L. | Menispermaceae/BGSBU-35 | C | Eye problems | L | Infusion | Infusion of leaves is put in eyes | 21  | 0.36  | 0.38 |
| Ficus palmata Forssk. [Kemer] | Moraceae/BGSBU-36 | T | Wounds | BK | Raw | Bark is applied on wound for quick healing. | 36  | 0.61  | 0.55 |
| Boerhavia diffusa L. [Itt-sitt] | Nyctaginaceae/BGSBU-37 | H | Removal of retained placenta | WP | Raw | Whole plants are fed twice a day. | 28  | 0.47  | 0.45 |
| Olea europaea Subsp. cuspidata (Wall. ex. G. Don) Cif. [Kaou] | Oleaceae/BGSBU-38 | T | Deworming | L | Raw | Fresh leaves are given for deworming. | 39  | 0.66  | 0.58 |
| | | | | Bk | Raw | Fresh stem bark is tied over broken bones. | | | |
### Table 2. Cont.

| Taxa/Local Name | Family/ Voucher Number | Life Form | Ethno- Veterinary Uses | Part(s) Used | Preparation | Method of Use | FC | RFC | UV |
|-----------------|------------------------|-----------|------------------------|--------------|-------------|---------------|----|-----|----|
| *Cedrus deodara* (Roxb. ex D. Don.) Don. [Dyar] | Pinaceae/ BGSBU-39 | T | Vomiting | W | Oil | Small pieces of wood are heated in a vessel which causes an oil to ooze out from them. This oil is given to livestock in vomiting. | 27 | 0.46 | 0.51 |
| | | | Hair fall in Goats | W | Oil | Oil is also applied externally to cure hair fall in goats. | | | |
| | | | Removal of ticks and lice | W | Oil | Oil is also applied externally. | | | |
| *Plantago lanceolata* Linn. [Chamch-e-pater] | Plantaginaceae/ BGSBU-40 | H | Yoke galls | WP | Paste | Paste is applied externally. | 29 | 0.49 | 0.54 |
| *Picrorhiza kurroa* Royle. ex Benth. [Koudh] | Plantaginaceae/ BGSBU-41 | H | Pneumonia | Rh | Powder | Dried rhizome powder mixed with wheat flour, gur and water is fed to cattle. | 16 | 0.27 | 0.29 |
| | | | Tapeworm | Rh | Paste | Paste is given orally. | | | |
| | | | Detachment and expulsion of placenta | Sd | Raw | Grains are fed to cows after delivery to facilitate the detachment and expulsion of placenta. | 35 | 0.59 | 0.56 |
| | | | Constipation | Sd | Paste | Paste of rice flour is made which is given to sheep. | | | |
| *Oryza sativa* Linn. [Chaval] | Poaceae/ BGSBU-42 | H | Detachment and expulsion of placenta | Sd | Raw | Grains are fed to cows after delivery to facilitate the detachment and expulsion of placenta. | 35 | 0.59 | 0.56 |
| | | | Constipation | Sd | Paste | Paste of rice flour is made which is given to sheep. | | | |
| | | | Gaseous bloats | R | Paste | Fresh roots are crushed, salt is added and small balls are made which are given to cattle. | 33 | 0.56 | 0.51 |
| | | | Cough | R | Paste | Paste is fed to cattle. | | | |
| | | | Sprained body parts | R | Paste | Paste is mixed with salt and is given orally. | | | |
| *Persicaria hydropiper* L.) Delarbre [Pipla] | Polygonaceae/ BGSBU-44 | H | Tongue infection | L | Raw | Chopped leaves are applied on the tongue. | 25 | 0.42 | 0.44 |
| *Aconitum heterophyl- lum* Wallich ex Royle [Patris] | Ranunculaceae/ BGSBU-45 | H | Flatulence | R | Powder | Root powder is given with water. | 17 | 0.29 | 0.34 |
Table 2. Cont.

| Taxa/Local Name                  | Family/Voucher Number | Life Form | Ethno-Veterinary Uses | Part(s) Used | Preparation | Method of Use                                | FC  | RFC | UV  |
|---------------------------------|-----------------------|-----------|-----------------------|--------------|-------------|---------------------------------------------|-----|-----|-----|
| **Prunus persica** (Linn.) Batsch. [Aarou] | Rosaceae/BGSBU-46     | T         | Wound healing         | L            | Paste       | Paste is applied externally.               | 25  | 0.42| 0.47|
| **Galium aparine** L. [Beeso]    | Rubiaceae/BGSBU-47    | H         | Wound healing         | WP           | Paste       | Paste is applied externally.               | 21  | 0.36| 0.29|
| **Salix alba** L. [Beeso]        | Salicaceae/BGSBU-48   | T         | Deworming             | Bk L         | Decoction   | Bark decoction and leaves are given to animals. | 32  | 0.54| 0.56|
| **Aesculus indica** (Wall. ex Camb.) Hook. [Bankhori] | Sapindaceae/BGSBU-49  | T         | General weakness      | F            | Paste       | Crushed fruits mixed with onion and salt are fed to cattle. | 31  | 0.53| 0.47|
| **Dodonaea viscosa** Jacqin [Sanatha] | Sapindaceae/BGSBU-50  | S         | Deworming             | L Extract    | Leaf extract is given to cattle.           | 20  | 0.34| 0.39|
| **Bergenia ciliata** (Haw.) Sternb. [Bat mevo] | Saxifragaceae/BGSBU-51 | H         | Diarrhoea             | R Powder     |  | Dried roots are powdered which is given to cattle with luke-warm water. | 30  | 0.51| 0.46|
| **Verbascum thapsus** L. [Gidharh tamako] | Scrophulariaceae/BGSBU-52 | H         | Flatulence            | AP Decoction | Decoction is prepared by boiling aerial parts in water for about 2 h, is added to the paddy husk and given to the cattle to cure flatulence. | 26  | 0.44| 0.3 |
| **Datura stramonium L. [Daturo]** | Solanaceae/BGSBU-53   | H         | Leech removing        | Sd Raw       | Dried seeds are heated on fire to release smoke which is used to expel leeches from the nasal cavity. | 21  | 0.36| 0.31|
| **Urtica dioica** L. [Kiyarie]   | Urticaceae/BGSBU-54   | H         | Fractured bones       | R Paste      | Root paste is applied on the fractured bones for early healing. | 23  | 0.39| 0.47|
| **Sambucus wightiana** Wall.     | Viburnaceae/BGSBU-55  | H         | Foot and mouth disease | R Paste      | Paste is applied externally.               | 27  | 0.46| 0.39|
| **Viburnum grandiflorum** Wall ex. Wt & Arn. [Kuch] | Viburnaceae/BGSBU-56  | S         | Wound healing         | R Powder     | Powdered roots mixed with mustard oil is applied externally. | 37  | 0.63| 0.64|

**Abbreviations used:** H = Herb; S = Shrub; T = Tree; C = Climber; R = Root; F = Fruit; L = Leaf; B = Bulb; Sd = Seed; W = Wood; AP = Aerial parts; WP = Whole plant; Rh = Rhizome; Ltx = Latex; Bk = Bark.
3.3. Utilization Pattern

Depending on hereditary knowledge and the accessibility of those plants and plant parts to the local population, different plant parts, such as leaves, roots, stems, flowers, fruits, and even resins, gums, and galls of some plants, are used in diverse ways in traditional health care systems. In the present study, leaves were the most common plant parts used (35.71%), followed by roots (16.07%), whole plants (12.5%), fruits (10.71%), aerial parts (8.92%), seeds (7.14%), rhizome and bark (5.35% each), bulbs (3.57%), grains, and wood and latex (1.78%) (Figure 4). Leaves are often used in herbal preparations and have been documented as the most-used plant parts in many studies [68–72]. The reason for the preference of leaves in herbal medicines may be due to their easy collection [73] and the presence of bioactive metabolites such as alkaloids, flavonoids, terpenoids, saponins, phenolic compounds in leaves [74]. Further, the collection of leaves causes less damage to the plant [40]. Similarly, a high concentration of biologically active compounds in roots make them favored parts for curative uses, besides leaves [50].

Figure 3. Growth form of the documented plant species.

Figure 4. Utilization pattern of the documented plant species.
3.4. Methods of Herbal Drug Preparations and Administration

Herbal preparations were made through different modes i.e., infusion, poultice, powder, decoction, extract, juice, paste, tea, scorched, gel, cooked and steamed. In the present study, the main method of preparation was paste (36.6%), followed by raw (28.3%), powder (13.3%), decoction (11.1%), infusion, extract and cakes (3.33% each) and oil (1.6%) (Figure 5). Pastes are one of the common modes of preparation in ethno-medicinal practices due to their ease in preparations and handling. Furthermore, in pastes the originality and purity of plant material is maintained as no heat or any other treatment is used and thus does not alter the phytochemical composition of the plant material, leading to the acceleration of biological activities. Similar findings have been reported from some other studies also [75,76]. Preparations are applied both externally as well as internally. Dosages are determined on the basis of age, physical appearance health conditions, and the sternness of disease/ailment. Most of the plants are used directly but in some cases plant materials are mixed with additive materials and the amalgam is used as medicine. Common additives used include the husk of wheat and rice, salt, oil, flour and gur (raw sweet).

![Figure 5. Mode of preparations of the documented plant species.](image)

3.5. Quantitative Analysis

Relative Frequency of Citation: In the present study, the RFC value ranges from 0.27–0.81 (Table 2). The highest RFC was recorded for *Brassica rapa*, and the lowest RFC was recorded for *Picrorhiza kurroa* and *Trillium govanianum*. Some other species with a high RFC include *Taraxacum officinale* and *Trifolium pratense* (0.76 each), *Foeniculum vulgare* (0.73) and *Achillea millefolium* (0.71). The plant species with high RFC grow abun-
dantly in the study area, so the local people are well aware of their therapeutic properties. Thus, among the indigenous people, their unique abilities to treat various illnesses and conditions have gained popularity. Such plant species should be evaluated for their phytochemical and pharmacological properties, as these plant species could lead to identifying bioactive compounds for novel drug discoveries [77].

Use Value: In the present study, UV ranges from 0.87 for *Brassica rapa* to 0.21 for *Trillium govanianum* (Table 2). Other species with UV high include *Trifolium pratense* (0.79) and *Taraxacum officinale* (0.78). The reason for the high UV of a species may be due to its
easy availability, extensive distribution and high therapeutic properties for curing various diseases [78,79]. Plants having more use reports have higher use values, while those plants having fewer use reports have lower use values [50]. In the present study, the plant species having higher use values are common plants growing abundantly in the study area. Such plants with a higher UV suggest the presence of bioactive compounds and thus require thorough phytochemical evaluations [80].

**Informant Consensus Factor:** ICF is used to gauge how well the community agrees on the usage of several plant species for a given disease category. The ICF for the eight disease categories identified during the present study ranges from 0.93 to 0.97 (Table 3). The ICF for all disease categories was recorded as being high, revealing that the informants have the same opinion on which plants to use in the treatment of common diseases. The highest ICF was recorded for dermatological/wounds and jaundice/Foot and Mouth disease (0.97 each) followed by pregnancy and post-pregnancy (0.96), the musculoskeletal system, the gastrointestinal system and insecticide/antidote categories (0.95 each), pneumonia/cough/cold/fever (0.94) and hair loss, and ENT/ophthalmic system (0.93). The high ICF values signify a plausibly high consistency of informants on the uses of medicinal plant species [81]. For treating a single disease category, a high ICF value is frequently linked to a small number of specific plants with high use reports [82], whereas low values are linked to many plant species with comparable or high use reports, implying a lower level of agreement among the informants on the use of these plant species for treating a specific disease category [50].

| Disease Category               | Ailments Included                                                                 | Nur | Nt  | ICF |
|-------------------------------|----------------------------------------------------------------------------------|-----|-----|-----|
| Musculoskeletal System        | Body weakness, Stretch of bones and ligaments, fatigue, Fracture, sprains        | 196 | 10  | 0.95|
| Pneumonia/Cough/Cold/Fever    | Pneumonia, Cough, Cold, Fever                                                    | 88  | 6   | 0.94|
| Dermatological/wounds         | Skin diseases, wounds, yolk galls                                               | 268 | 9   | 0.97|
| Alopecia/Ophthalmic           | Hair loss, eye disease                                                           | 47  | 3   | 0.93|
| Gastrointestinal System       | Indigestion, vomiting, diarrhea, dysentery, flatulence, tongue infection, gaseous bloats, constipation, loss of appetite | 315 | 16  | 0.95|
| Jaundice/Foot and Mouth disease| Jaundice, Foot and Mouth disease                                                | 126 | 4   | 0.97|
| Pregnancy and post pregnancy  | Stimulate oestrus cycle, Removal of retained placenta, increased milk production, Vigor maintenance | 322 | 12  | 0.96|
| Insecticide/Antidote          | Poisonous bite, deworming, removing of lice and ticks, removing of leaches       | 345 | 15  | 0.95|

**Jaccard Index:** The resemblances and disparities in ethno-medicinal studies seem to mark the significance of traditional medicinal knowledge in different regions, where historical, phytochemical and ecological factors interact in their selection [83,84]. Further, indigenous communities have differences in their origins and cultures leading to differences in ethnobotanical knowledge within these communities [85]. The results of regional, national, and international studies were compared with the data from the current study, and the observed percentage of similarity spans from 1.3 to 23.1 with an average value of 8.95 (Table 4). The maximum level of similarity was found with the studies conducted by Ch et al. [86], Sharma et al. [87], Khuroo et al. [88], and Khan et al. [89], with JI values of 23.1, 14.6, 11.9 and 10.5, respectively. It is interesting to note that all these studies were carried out in areas which are in close vicinity to our study area and thus have similar ecological factors and ethnic values. The lowest index of similarity was found in the study conducted by Harsha et al. [90] in Uttara Kannada district of Karnataka, and the reason for this may be the difference in topography, climate, vegetation cover and difference in socio-cultural values. According to JI, various tribal societies have their own traditional knowledge
systems that employ identical plant bioresources in various ways. The great amount of information among groups can be revealed by recording and comparing ethnic/traditional knowledge, which can lead to new sources of drug discovery [83].

Table 4. Jaccard index comparing the present study with some of the earlier reports.

| Area                                      | Study Year | No. of Recorded Plant Species | Plants with Similar Use | Plants with Dissimilar Use | Total Species Common in Both Areas | Species Enlisted Only in Aligned Area | Species Enlisted Only in Study Area | % of Plants with Similar Use | % of Plants with Dissimilar Use | Jaccard Index | Citation       |
|--------------------------------------------|------------|-------------------------------|-------------------------|---------------------------|-----------------------------------|---------------------------------------|-----------------------------------|-------------------------------|-------------------------------|----------------|----------------|
| Kashmir Himalaya                          | 2007       | 24                            | 5                       | 2                         | 7                                 | 17                                    | 49                                | 20.8                          | 8.3                           | 11.9           | [88]           |
| Kathua district of J&K                     | 2012       | 72                            | 5                       | 8                         | 13                                | 59                                    | 43                                | 6.9                           | 11.1                          | 14.6           | [87]           |
| Samahni valley, district Bhimber (Azad Kashmir) Pakistan | 2006 | 54                            | 9                       | 6                         | 15                                | 39                                    | 41                                | 16.7                          | 11.1                          | 23.1           | [86]           |
| Poonch valley, Azad Kashmir                | 2012       | 19                            | 3                       | 3                         | 6                                 | 13                                    | 50                                | 15.8                          | 15.8                          | 10.5           | [89]           |
| Tosham block of district Bhiwani (Haayana) | 2014       | 52                            | 3                       | 2                         | 5                                 | 47                                    | 51                                | 5.8                           | 3.8                           | 5.4            | [91]           |
| Jhansi district, UP                        | 2010       | 47                            | 0                       | 7                         | 7                                 | 40                                    | 49                                | 0                             | 14.9                          | 8.5            | [92]           |
| Tehri district of Garwal Himalaya          | 2013       | 35                            | 0                       | 5                         | 5                                 | 30                                    | 51                                | 0                             | 14.3                          | 6.6            | [93]           |
| Uttara Kannada district of Karnataka       | 2005       | 24                            | 0                       | 1                         | 1                                 | 23                                    | 55                                | 0                             | 4.2                           | 1.3            | [90]           |
| Visakhapatnam and Vizianagarm districts, AP | 2015 | 61                            | 0                       | 3                         | 3                                 | 58                                    | 53                                | 0                             | 4.9                           | 2.8            | [94]           |
| Kudavasal taluk of Thiruvarur district, TN | 2016       | 54                            | 0                       | 6                         | 6                                 | 48                                    | 50                                | 0                             | 11.1                          | 6.5            | [12]           |

3.6. Statistical Analysis

The Pearson correlation coefficient between the use value and relative frequency of citations is 0.94, revealing a considerable and strong positive correlation between the proportion of uses of a plant species within a sample of respondents and the frequency with which a particular use of a species was mentioned by the informant (Table 5). This implies a direct relationship between the number of informants and use reports of a particular species. In the present study, R² is 0.88, implying that 88% of the variation in RFC can be elucidated in terms of the UV [95,96], implying the empirical robustness among the two indices [66]. It also implies that Figure 6 illustrates the positive correlation between the values of RFC and UV.
Table 5. Correlation between RFC and UV.

|       | RFC       | UV       |
|-------|-----------|----------|
| RFC   | Pearson   | 0.940 ** |
| Sig.  | Correlation| 0.000    |
| N     | 56        | 56       |
| UV    | Pearson   | 1        |
| Sig.  | Correlation| 0.000    |
| N     | 56        | 56       |

** Correlation is significant at the 0.01 level (2-tailed).

Figure 6. Relationship between RFC and UV.

3.7. Reverse Pharmacological Correlation of Ethno-Vetenairy Plants

Plant species documented during the present study have been found to show many biological activities due to the presence of various biologically active phytochemicals (Table 6). Some of the biological activities associated with these plants are antiviral, antitumorogenic, spermicidal, hepatoprotective, nephroprotective, antidiabetic, anti-inflammatory, immunomodulatory, antimicrobial, antiparasitic, anti-allergic, antioxidant, hypolipidemic, antifungal, anthelmintic, antimicrobial, antidiabetic, and antipyretic. Prominent biologically active chemicals reported from the documented plants include quercetin, betaine, allin, allicin, linalool, borneol, stigmasterol, lupeol, β-sitosterol, campesterol, rutin, saussureine, eugenol, limonene, α-thujone, artemetin, α-pinene, β-pinene, salicylic acid, berberine, berbamine, cannabigerol, cannabidiol, betulin, kaempferol, penogenin, diosgenin, apigenin 7-glucoside, aconitine, heterophylline A, heterophylline B, and bergenin.
Table 6. Reverse pharmacological correlations of documented ethno-veterinary plants from Poonch district of J&K.

| Taxa            | Phytochemicals                                                                 | Pharmacological Activities                                                                 | Reference(s) |
|-----------------|--------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------|--------------|
| *Amaranthus viridis* | rutin, quercetin, spinosterol, amasterol                                        | antifungal, hepatoprotective, anthelmintic, antioxidant, antimicrobial, antidiabetic, antipyretic, anti-inflammatory | [97–100]    |
| *Chenopodium album* | 3-O-glycosides of caempferol, quercetin, andisoramnetin, kaempferol-3-O-(4-β-D-xylopyranosyl)-α-L-rhamnopyranoside-7-O-α-L-rhamno-pyranoside, 3-O-(4-β-D-apiofuranosyl)-α-L-rhamnopyranoside-7-O-α-L-rhamnopyranoside, 5-O-α-L-rhamno-pyranoside, 3,7-di-O-α-L-rhamnopyranoside, 3-O-glucopyranoside, quercetin, 3,7-di-O-β-D-glucopyranoside, 3-O-glucosylglucuronide, 3-O-α-L-rhamnopyranosyl-(1→6)-β-D-glucopyranoside, 3,7-O-β-D-glucopyranoside, kaempferol-3-O-arabinogluucose, quercetin, quercetin-3-O-xyllosylglucoside, quercetin-3-orhamnoglucoside | antiviral, antifungal, anti-inflammatory, antiallergic, antiseptic, anti-nociceptic, sperm immobileizing, immunomodulating, antiparasitic, antispasmodic, antibacterial, anti-helminthic, hypotensive, spasmolytic, hepatoprotective | [101–103] |
| *Achyranthes aspera* | betaine, achyranthine, hentriacontane, ecldyosterone, achyranthes saponins A, B, C, D, α-Lrhamnopyranosyl-(1→4)-(β-D-glucopyranosyluronic acid)-(1→3)-oleanolic acid, trigmasta-5, 2-dien-3-β-ol, trans-13-docasenoic acid, n-hexacosanyl-n-decaniate, hexacos-17-enoic acid | antiviral, anticarcinogenic, spermicidal, hepatoprotective, nephroprotective, anti-diabetic, anti-inflammatory, immunomodulatory, antiparasitic, anti-allergic, anti-oxidant, hypolipidemic | [104–106] |
| *Allium cepa*    | quercetin 3,7′,4′-O-β-triglucopyranoside, quercetin 3,4′-O-β-diglucopyranoside, taxifolin 4′-O-β-glucopyranoside | analgesic, anti-diabetic, antioxidant, antidepressant, aphrodisiac, antihyperlipidemic    | [107–109] |
| *Allium sativum* | alliin, allicin, ajoenes, vinyldithiins, quercetin                             | antitumor, cortical, anti-atherosclerotic, anti-bacterial, anti-inflammatory, antihypertensive, antifungal, antiprotozoal, antioxidant, anti-inflammatory, and anticancer | [110–112] |
| *Angelica glauca* | aleric acid, angelic acid, angelisine, phellandrene, coumarins, bergapten, linalool, borneol, anthotoxin, umbelliferene                       | cardioactive, carminative, digestive, sudorific, expectorant and stomachic, antispasmodic, anti-bacterial, antifungal | [113–117] |
| *Foeniculum vulgare* | *trans*-anethole, fenchone, methylchavicol, eriodictol-7-rutinoside, quercetin-3-rutinoside, rosmarinic acid, quercetin-3-glucuronide, isouqueritin, quercetin-3-arabinoside, kaempferol-3-glucuronide, kaempferol-3-arabinoside, isorhamnetin glucoside, 3-O-cafeoylquinic acid, 4-O-cafeoylquinic acid, 5-O-cafeoylquinic acid, 1,3-O-di-cafeoylquinic acid, 1,4-O-di-cafeoylquinic acid, 1,5-O-di-cafeoylquinic acid | antimicrobial, antiviral, anti-inflammatory, antimutagenic, antinoceptive, antipyretic, antispasmodic, antithrombotic, apoptotic, cardiovascular, chemomodulatory, antitumor, hepatoprotective, hypoglycemic, hypolipidemic, and memory enhancing property | [118] |
| Taxa                  | Phytochemicals                                                                                           | Pharmacological Activities                                                                 | Reference(s) |
|----------------------|-----------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------|---------------|
| *Calotropis procera* | voruscharin, uscharidin, uzarigenin, calotroposide, calactin, calotoxin, uscharin, ascleposide, calotropagin, coroglaucigenin, calotropin, proceroside, proceragenin, syriogenin, rutin, cyanidin-3-rhamnoglucoside, cycloart-23-en-3β, 25-diol, cyclosadol, multiflorenol, procestrol, quercetin-3-rutinoside, β-sitosterol, β-sitost-4en-3-one, stigmasteral, cyanidin-3-rhamnoglucoside, ascorbic acid, calactin, calotoxin, calatropagenin, calotropin, polysaccharide containing D-arabinose, D-glucose, D-glucosamine L-rhamnose, calotropagin, 3-proteinase, calotropin, α-calotropeol, 3-epimoretenol, gigantin, giganteol, isogiganteol, α-lactuceryl acetate, α-lactuceryl isovalerate, lupeol, procestrin, procerain, procerain-B, calactinic acid, choline, O-pyrocatechuic acid, β-sitosterol, taraxasterol, calotroprocerol A, calotroprocerol acetate A, calotroprocerone A, calotroproceryl acetate B | analgesic, antinoceptive, anticonvulsant, antimalarial, anthelminthic, antioxidant, antidiabetic, anticancer, antimicrobial, anti-inflammatory, immunomodulatory, antipyretic, antiarrheal, antidematogenic, antiplasmodial | [119–122] |
| *Hedera nepalensis*  | lupeol, hederacoside C, α-hederin                                                                       | anticancer, neuroprotective, antidiabetic, antioxidant                                       | [123,124] |
| *Taraxacum officinale* | stigmasterol, campesterol, syringing, dihydroxyringin, dihydroconiferin, luteolin 7 glucoside, luteolin 7 diglucosides, cichorin, ascinulin, dicafeoyltartaric acid, rutin, luperoside, quercetin | antibacterial, antioxidant, anticancer, diuretic, hepatoprotective, antiviral, anti-inflammatory | [125–129] |
| *Erigeron canadensis*| onyzolide, conyzapyranone A, conyzapyranone B, 4 Z,8 Z-maticraria-γ-lactone, 4 E,8 Z-maticraria-γ-lactone, 9,12,13-trihydroxy-10(E)-octadecenoic acid, epifriedelanol, friedeline, taraxerol, simiareol, spinasterol, stigmasteral, β-sitosterol, quercetin-7-O-beta-D-galactopyranoside, quercetin, luteolin, apigenin, 5,7,4′-trihydroxy-3′-methoxy flavone, quercetin-3-alpha-rhamnopyranoside, quercetin-3-O-beta-D-glucopyranoside, apigenin-7-O-beta-D-gluco pyranoside, luteolin-7-O-beta-D-glucuronide methyl ester, 4′-hydroxy baicalein-7-O-beta-D-glucopyranoside, baicalein, rutin | antimicrobial, antioxidant, anticoagulant, anti-inflammatory, anticancer, antifungal | [130–132] |
| *Saussurea costus*   | costunolide, dehydrocostuslactone, costic, palmitic, linoleic acids, cyclocoastunolide, alantolactone, isoalantolactone, isodehydrocostus lactone, iso-zaluzanin-C, guainolides, 12-methoxydehydrocostuslactone, 4-methoxydehydrocostus lactone, saussurealdehyde, isodehydrocostus-lactone-15-aldehyde, cyanaropicrin, reynosin, santamarine, Saussureal, pregnenolone, sitosterol, daucosterol, syringe, chlorogenic acid, saussurine | anti-inflammatory, anticancer, hepatoprotective, immunomodulatory, anti-ulcer, antimicrobial, hypoglycemic, antiparasitic | [133–138] |
| Taxa                        | Phytochemicals                                                                 | Pharmacological Activities                                                                 | Reference(s) |
|-----------------------------|--------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------|--------------|
| **Sonchus arvensis**        | alkaloids, flavonoids, phenols, saponins and tannins                         | anti-fatigue activity, antioxidant, hepatoprotective, kidney-protective, antidiabetic, antibacterial | [139–141]    |
| **Achillea millefolium**    | borneol, camphene, azulene, carophyllene, 1,8-cineole, p-cymene, eugenol, farnesene, limonene, myrcene, α-pinene, β-pinene, salicylic acid, achillin, achillin, terpinolene, α-thujone, artemetin, casticin, isorhamnetin, luteolin, rutin | antibacterial, antifungal, antiparasitic, hemostyptic, anti-inflammatory, antispasmodic, antioxidant, anticancer, hepatoprotective, | [142–144]    |
| **Berberis lycium**         | berberine, berbamine, chinabine, karakoramine, palmatine, balauchistanamine, giglitine, jhelumine, punjabine, sindamine, chinabine acetate acid, maleic acid, ascorbic acid, berberine, berberine hydrochloride, berberine hydroiodide, oxyberberine, umbellatine | antidiabetic, hepatoprotective, anti-hyperlipidemic, antimutagenic, antioxidant, anticancer, anti-hyperlipidemic, anti-inflammatory, antispasmodic, anti-hyperlipidemic, anti-inflammatory, antispasmodic, | [145,146]    |
| **Alnus nitida**            | Diarylheptanoids                                                              | anticancer, anti-inflammatory, anti-influenza, hepatoprotective, antitumor and antioxidant | [147–149]    |
| **Brassica rapa**           | lutein, β-carotene, glucobrassicin, 4-methoxyglucobrassicin, 1-methoxyglucobrassicin, isothiocyanates, nitriles, thiocyanates, epithionitrites, oxazolidine, aciton, citric, ketoglutaric, malic, shikimic, fumaric, oxalic, ascorbic, succinic and glutamic acids | anticancer, antioxidant, anti-inflammatory, chemopreventive | [150,151]    |
| **Buxus wallichiana**       | buxenolen E, buxatine H, Buxiramin D, buxatinebuxadrine F, buxidine F, (+)-16a, 31-diacetylbuxadine, semperviraminol, buxamine F | bitter tonic, diaphoretic, anti-rheumatic, vermifuge, anti-helminthic, analgesic, purgative diuretic, antiepileptic, antitumoric, hemorrhoids | [152–154]    |
| **Cannabis sativa**         | cannabigerol, cannabichromene, cannabidiol, tetrahydrocannabinol, 9-tetrahydrocannabivarin, annabicyclol, annabinol, D-limonene, beta-myrcene, alpha-pinene, caryophyllene oxide, D-linalool, beta-caryophyllene | anticonvulsant, antibiotic, antifungal, anti-inflammatory, analgesic, anxiolytic, antipsychotic, antioxidant, antispasmodic, anti-emic, sedative | [155]         |
| **Valeriana jatamansi**     | baldrinal, homobaldrinal, decyl baldrinal, valtoxal, isovalepotriate, acetoxvalexopatrate, isovalemxyhydroxy-dihydrovratate, rupesin, linarin, linarin-isovalerianate, linarin-2-O-methylbutyrate, hispidulin, hesperetin-7-O-β-rutinoside, hesperidin, kaempferol 3-O-β-D-glucopyranoside, quercetin 3-O-β-D-glucopyranoside, kaempferol, quercetin, 7-O-β-D-glucopyranoside, apigenin 7-O-β-D-glucopyranoside, larin cesinol, pinsepil, syringaresinol, pisoresinol, berchemol, podophyllotoxin, hydroxyvalencenic acid, acetoxvalexeric acid, valerenic acid | neuroprotective, sedative, cytoxic, cardio-protective, anxiolytic, antidepressant, anti-inflammatory, antispasmodic, hepatoprotective, antioxidant | [156]         |
| **Lagenaria siceraria**     | β-carotene, 22-deoxocurcubitacin-d, 22-deoxoisocurcubitacin d, avenasterol, codisterol, elesterol, isofucasterol, stigmasterol, sitosterol, compesterol, spinasterol, 7-O-glucosyl-6-C-glucoside apigenin, 6-C-glucoside apigenin, 6-C-glucoside luteolin, 7,4′-O-diglucosyl-6-C-glucoside apigenin | analgesic, anti-inflammatory, anti-hyperlipidemic, diuretic, hepatoprotective, anthelmintic, antibacterial, immunomodulatory, antistress, hepatoprotective, antioxidant | [157]         |
| Taxa                      | Phytochemicals                                                                 | Pharmacological Activities                        | Reference(s) |
|--------------------------|-------------------------------------------------------------------------------|---------------------------------------------------|---------------|
| Mallotus philippensis    | rotterin, mallotoxin, iso rotterin, crotoxigenin, betulin, friedelin, kamaladiol-3-acetate, lipoe, tannic acid, 3-hydroxy-D-A-friedoolean-3-en-2-one, 2β-hydroxy-D-A-friedooleanan-3-one, 3α-hydroxy-D-A-friedooleanan-2-one, betulin-3- acetate lupeol acetate, berginin acetylateuruite acid, sitosterol, crotoxigenin, coroghcignin, homorottlerin | antioxidant, antimicrobial, anticaner, anti-viral, anti-inflammatory, hepatoprotective, anti-helminthic, analgesic | [158,159]    |
| Ricinus communis         | kaempferol-3-O-β-D-xylopyranoside, Kaempferol-3-O-β-rutinoside, Kaempferol-3-O-β-D-glucopy-ranoside, Quercetin, Ricin, Rutin, Ellagic acid, Epicatechin, Fucosterol, Stigmasterol, α-pinene, 1,8-pinene, 1,8-cineole, β-caryophyllene, Rinicine, demetilricinicine, Tririncinolein, Lupeol | abortifacient, analgesic, antiasthmatic, anti-fertility, contraceptive, antidiabetic | [160]         |
| Trifolium pratense       | biochanin A, benzaldehyde, (Z)-β-caryophyllene, 3-methyl-1-butanol, 3-octanone, Z)-β-caryophyllene, β-famesene, 6,10,14-trimethyl-pentadecanone, 3-methylbutyl butanoate, 3-methyl-1-butanol,3-methylbutanoic acid, ethyl-2-methylpentanoate | estrogenic, antiproliferative, antioxidant | [161,162]    |
| Trifolium repens         | 4′,5,6,7,8-pentahydroxy-3-methoxyflavone and 5,6,7,8-tetrahydroxy-3-methoxyflavone, 3,7-dihydroxy-4′-methoxyflavone, 5,6,7,8-tetrahydroxy-4′-methoxyflavone, 3,5,6,7,8-pentehydroxy-4′-methoxyflavone, 6-hydroxy-kaempferol, 4′,5,6,7,8-pentahydroxyflavone, 3,4′-dimethoxykaempferol, quercetin and kaempferol, chalcone, chalconol glucosides, repensin A and repensin B, gallocatechin, epigallocate-chin, gallocatechin-(4α-8)-epigallocatechin | antioxidant, antifungal, anticaner, antiaging, hepatoprotective, anti-inflammatory, antidiabetic | [163]         |
| Trigonella foenum-graecum| disogenin, gitogenin, neogitogenin, homorientin saponaretin, neogigogenin, trigogenin, trigonelline and choline | immunomodulatory, antioxidant, chemo preventive, anticancer, antidiabetic, gastro protective, anti-inflammatory, anti hypertytic, anthelmintic, antigenotoxic, anti-plasmodial, hypocholesterolemic, antiseptic, aphrodisiac, astringent, bitter, demulcent, emollient, expectorant | [164,165]    |
| Juglans regia            | quercetin, and caffeic acid, paracomaric acid, juglone, ascorbic acid, queratin arabinoside, queretin xyloside and queretin rhamnoside, Glueltins, globulins, albumin and prolamins, glansrins A, B and C, casuarin, stenophyllarin | antioxidiant, antidiabetic, antimicrobial, and hepatoprotective, antifungal, anti-hypertensive, renal protective, anti-inflammatory, antinoceptive, anticancer | [166,167]    |
### Table 6. Cont.

| Taxa                        | Phytochemicals                                                                 | Pharmacological Activities                                         | Reference(s) |
|-----------------------------|------------------------------------------------------------------------------|--------------------------------------------------------------------|--------------|
| *Vitex negundo*             | friedelin, casticin, artemetin, terpinen-4-ol, α-terpineol,                   | anxiolytic, analgesic, anti-inflammatory, neuroprotective, antibacterial, antipyretic, anticancer, antioxidant    | [168,169]    |
|                             | sabenine, globulol, spathulenol, β-farnesene, farnesol, α-pinene, β-pinene,  |                                                                     |              |
|                             | linalool, terpinyl acetate, caryophyllene epoxide, caryophyllenol, vitexicarpin, |                                                                     |              |
|                             | viridiflorol, 5-odesmethylmobbiletin, gardenin A, gardenin corymbosin,         |                                                                     |              |
|                             | terpinen-4-ol, α-copaene, β-caryophyllene, β-elemene, camphene, α-thujene,     |                                                                     |              |
|                             | α-pinene, sebinene, α-elemen, δ-elemene, β-elemene, β-eudesmol, camphene,     |                                                                     |              |
|                             | careen, 1,8-cineol, α-phellendrene, β-phellendrene, α-guaiene, neral, gerarial, |                                                                     |              |
|                             | bornyl acetate, nerolidol, β-bisabolol, cedrol, agnuside, lagundinin,         |                                                                     |              |
|                             | viridiflorol, betulinic acid, ursolic acid, dimethoxyflavonone, vitexoside,    |                                                                     |              |
|                             | agnuside, R-dalbergiphenol, negundin A, negundin B, vitexin cafeate, epifriedelinol |                                                                     |              |
| *Ajuga parviflora*          | ajugin A, ajugin B                                                             | antioxidant, antibacterial                                         | [170–172]    |
| *Mentha sylvestris*         | menthol, menthone, isomenthone, limonene, neomenthol, methyl acetate,        | carminative, stimulant, stomachic, aromatic, antiseptic, antispasmodic, sudorific, emmenagogue, anesthetic, anodyne | [173–175]    |
|                             | beta-caryophyllene, piperitone, alpha- and beta-pipene, acacetin, chrysoeriol,|                                                                     |              |
|                             | diosmin, eriocitrin (eriodictyo-7-β-rutinoside), hesperidin, hesperidoside,   |                                                                     |              |
|                             | isorhoifolin, larin, luteolin, menthoxide, methyl rosmarinate, rutin,         |                                                                     |              |
|                             | tilianine, narirutin, nodiflorein, caffeic acid, lithospermic acid, rosmarinic|                                                                     |              |
|                             | acid, protocatechuic acid, protocatechuic aldehyde, phytosterols, β-sitosterol,|                                                                     |              |
|                             | daucosterol, alo-e-emodin, chrysophansol, emodin                              |                                                                     |              |
| *Punica granatum*           | punicalin, punicalagin, gallic acid, ellagic acid, cyanidin, delphinidin,     | antioxidant, antibacterial, anticancer, anti-inflammatory,         | [176–178]    |
|                             | uetolin, quercetin, kaempferol, naringenin, estrone, estriol, testosterone,   | antiagulant, antimutant, cardioprotective, antifungal, antidiabetic  |              |
|                             | betasistosterol, coumestrole, gammacopherol, punicic acid, campesterol,      |                                                                     |              |
|                             | stigmasterol                                                                  |                                                                     |              |
| *Malva neglecta*            | hydrotrystosyl, coumaroylhexoside, kaempferol-3-(p-coumaroyldiglucoside)-7-glucoside, quercetin-3-O-rutinoside, epicatechin-3-O-(4-O-methyl)-gallate, oleic acid, taurine, ethylene dimercaptan, isoeugenol, patchoulean, methyl 12-methyltetradecanoate, isopropyl myristate | antimicrobial, antioxidant, anti-inflammatory, anti-ulcerogenetic, hepatoprotective, neuroprotective  | [179–182]    |
| *Trillium govanianum*       | pennogenin, diosgenin, borassoside E, govanoside A                             | analgesic, anti-inflammatory, antifungal, antioxidant, anticancer, antispasmodic, diuretic | [183]        |
Table 6. Cont.

| Taxa                  | Phytochemicals                                                                 | Pharmacological Activities                          | Reference(s)         |
|-----------------------|--------------------------------------------------------------------------------|------------------------------------------------------|-----------------------|
| Melia azedarach       | surinol, melanin b, sendanoloactone, 3-α-hydroxy-4,4,14alpha-trimethyl-5-apreg-8-en-20-one, ochinin acetate, 4-methoxy-1-vinyl-beta-carboline, 4,8-dimethoxy-1-vinyl-beta-carboline, kuline, kulactone, kulolactone, kulinone, nimbine, azaridine, paraistine, isochuanliansu, 6-H-β-hydroxy-4-stigmasten-3-one, 6β-hydroxy-4-campesten-3-one, quercetin, quercetin-3-0-β-rutinoside, kaempferol3-0-β-rutinoside, rutin, kaempferol-3-L-ramno-Dglucoside, azaridine, bakayanin, bakalactone, margosine, azadrione, azadiradione, epoxyazadiradione, ochinin, ochchinol, ochchinolal, ochchinolides A and B, nimbolinin B, 1-desacytlnimboline B, nimbildins A and B, triterpene B, melaicos A1,A2, B1, B2, sendanin, sendenal, 1-cinnamoylmelianolone, meliantriol, melinanone, melianol, lupeol, β-sitosterol, catechin, vanillin, cinnamic acid, salannal, meliacarpinin E | analgesic, immunomodulatory, antifeedant, antifungal, antibacterial, antiviral, cytotoxic, anthelmintic, antilithic, antifertility | [184–186] |
| Cissampelos pareira   | hayatine, hayatinine, hayatidine, quercitol, sterol, espeperine, berberine, cissampeline, pelosine, cycleamine, menisamin iodine, cissamin chloride, paretrin, cissamine chloride, cissampareine, dehydrodicentrine, dicentrine and insularine | antipyretic, anti-inflammatory, antiarhritic, antilucler, anti diabetic, anticancer, antifertility, antimicrobial, antioxidant, antivenom, antimalarial, and immunomodulatory | [187,188] |
| Ficus palmata         | germanicol acetate, vanillic acid, germanicol acetate methyl ether, rutin | hepatoprotective, nephroprotective, anti ulcer and anticoagulant | [189] |
| Boerhavia diffusa    | eupatitin, eupatitin-3-0-β-Dgalactopyranos, eupatillin-7-O-β-Dgalactopyranoside, eupatilin 7-O-α-Lramnopyranosyl (1→2) α-Lramnopyranosyl (1→6)- β-Dgalactopyranoside, quercetin-3-O- β-Dglucopyranoside- 7-O-β-Dglucopyranosyl, -3-O-α-Lramnopyranosyl (1→6)-β-Dgalactosyloside, kaempferol, quercitin, borhavone, punarnavoside, alkalamide, ferulic acid, gentisic acid, caffeoyltartaric acid, boerhaaviac acid, boeravinone, coccineon, isomethone, limonene, menthol, phellandrene, safranal, α-pinene, geranylacetone, eugenol, stigmasterol, campesterol, β-sitosterol | antidiabetic, immunosuppressive, nephroprotective, antilithic, antioxidant, hepatoprotective, antiviral, anticancer, cardiovascular | [190–193] |
| Cedrus deodara        | wikstromal, matairesinol, dibenzybutyro lactol, cedrin, taxifolin, cedearadin, dihydroxymeticin, cedrinoside, deodardione, diosphenol, limonene,carboxylic acid, deodarin, sitosterol, deodarone, atlantone, α-himacholone, β-himacholone, α-pinene, β-pinene, myrcene, himachalene, cis-atlantone, α-atlantone | anti-inflammatory, analgesic, anti spasmodic, immunomodulatory, anti-hyperglycemic, anticancer, antibacterial | [194] |
| Plantago lanceolata   | pyrocatechol, picatechen, vanillic, verbascoside, taxifolin, luteolin 7-glucoside, hesperidin, hyperoside, apigenin 7-glucoside, pinoresinol, eriodictyol, quercetin, luteolin, kaempferol, apigenin, baicalein, plantaginin, aucubin, indicain, plantagonin | antiluercative, anti diabetic, anti diarrhoeal, anti inflammatory, anticancer, antinociceptive, antioxidant, antifatigue, antibacterial, antiviral | [195,196] |
Table 6. Cont.

| Taxa                | Phytochemicals                                                                 | Pharmacological Activities                                                                 | Reference(s) |
|---------------------|-------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------|--------------|
| Picrorhiza kurroa   | veronicoside, pikuroside, cucurbitacins, 4-hydroxy-3-methoxy acetophenone, apocyanin, drosin | hepatoprotective, anticholestatic, antioxidant, anti-inflammatory, anticancer, antiviral, hepatoprotective, anticonvulsant, nephroprotective | [197]        |
| Rumex dentatus      | rumejaposides E, cassialoin, emodin                                           | astringent, antioxidant, antibacterial                                                    | [198]        |
| Persicaria hydropiper | catechin, epicatechin, hyperin, isoquercitrin, isorhamnatin, kaempferol, quercetin, quercitrin, rhamnatin, rutin, 3-β-angelyloxy-7-epifuturonolide, 7-ketoisodrimenin, changweikangic acid A, dendocarbin L, fuegine, futuronolide, polygonumate, winterin, conferifolin, isodrimeninol | antioxidant, antibacterial, antifungal, anthelmintic, cytotoxic, anti-inflammatory, antifertility, neuroprotective | [199]        |
| Aconitum heterophyllum | aconitine, heterophylline A, heterophylline B, heteratisine, heterophyllidine, mesaconitine, 3 acetylaconitine, atidine, isoaconitine, hetidine, hetsinone, benzoybetalisnine, acetic acid, N-Diethyl-N-formyllaconitine, O-methylaconitine, methyl-N-succinoylanthranilate, hapyaconitine, benzoylalacine, benzoylmesaconine, benzoymphaconine, 6-dehydroacetylsesapaconitine, 13-hydroxylappaconitine | antidiarrheal, expectorant, diuretic, hepatoprotective, antipyretic, analgesic, antioxidant, alexipharmic, anodyne, anti-irritant, immunostimulant, febrifuge, anthelmintic, anti-cancerous, anti-emetic, anti-inflammatory, anti-flatulent, antiperiodic, anti-phlogistic, anti-diabetic, antifungal, antimicrobial, antiviral and carminative | [200–203] |
| Prunus persica      | lutein, β-cryptoxanthin, β-carotene, champagne, hexahydroxydiphenic acid, gallic acid, protocatechuic acid, quercetin, kaempferol, isorhamnatin, myricetin, cyanidin-3-O-glucoside, cyanidin-3-O-rutinoside, catechin, epicatechin | antioxidant, antimicrobial, anti-diabetic, anti-inflammatory, cardioprotective, neuroprotective, anticancer | [204]        |
| Galium aparine      | gallic acid, caffic acid, isoquercitrin, rutin, quercetin, luteolin, asperulosidic acid, deacetylasperulosidic acid, β-sitosterol, daucosterol, vanillin, γ-sitosterol | antioxidant, antimicrobial, anticancer, hepatoprotective, antifeedant | [205–207]   |
| Salix alba          | anthocyanins, p-hydroxybenzoic, gallic acid, Gentisic acid, sisymbriifolin      | antibacterial, antifungal, antioxidant                                                    | [208]        |
| Aesculus indica     | quercetin, mandelic acid                                                      | Immunomodulatory                                                                           | [209,210]    |
| Dodonaea viscosa    | aliarin, pinocembrin, viscosol, sakuranetin, isokaempferide, dodoviscins A-J, isorhamnatin, quercetin, jegosapogenol, β-pinene, myrcene, limonene, p-cymene, citronellal, linalool, linalyl acetate, β-terpineol, geraniol, α-spinasterol, 4-hydroxy-3,5-diprenylbenzaldehyde, β-sitosterol, stearic acid, syringic acid, fraxetin, cleomiscosin A, cleomiscosin C, and β-sitosterol β-D-glucoside | Antidiabetic, antimicrobial, antioxidant, cytotoxic, antifertility, anti-inflammatory, analgesic, anti-ulcer, anti-spasmodic | [211]        |
| Bergenia ciliata    | bergenin, tannic acid, gallic acid, catechin, β-sitosterol, arbutin, afzelechin, quercetin 3-o-β-D xylopyranoside, quercetin 3-o-α-L-arabinofuranoside, Linalool, | anti-diabetic, anti-cancer, anti-ulcer, antibacterial, antimalarial, antifungal, antiviral | [212]        |
| Verbascum thapsus   | verbascoside, aucubin, harpagoside, ajugol, isocatapal, methylcatapal, 6-O-α-L-rhamnopyranosylcatapal, saccatoside, harpagide, ningpoegenin, 10-deoxyeucosmiol, jiojoglutulideforsythoside B, alyssenoside, arenarioside, saikogenin A, thapsuine A, β-spinasterol, veratic acid | antioxidant, antimicrobial, antiviral, anti-cancer, antihyperlipidemic | [213]        |
Table 6. Cont.

| Taxa                     | Phytochemicals                                                                 | Pharmacological Activities                  | Reference(s) |
|--------------------------|-------------------------------------------------------------------------------|---------------------------------------------|--------------|
| *Datura stramonium*      | atropine, scopoline, 3-(hydroxyacetoxyl) tropan, 3-hydroxy-6-(2-methylbutyryloxy) tropan, aponorscopolamine, 7-hydroxyhyoscyamin, aponorscopolamine, aposcopolamine, hygrine, hyoscyamine, littorine, meteloidine, scopine scopolamine, tropinone, tropine | antimicrobial, analgesic, anti-asthmatic, anticancer, antioxidant | [214] |
| *Urtica dioica*          | acetylcholine, histamine, and 5-hydroxytryptamine, formic acid, histamine, serotonin, carvacrol, carvone, isolectins | anti-inflammatory, analgesic, antimicrobial, hepatoprotective, anti-colitis | [215] |
| *Sambucus wightiana*     |                                                                                | Antimicrobial                               | [216] |
| *Viburnum grandiflorum*  | luteolin                                                                      | Antifungal                                  | [217, 218] |

These phytochemicals are associated with several biological activities and thus play a vital role in healthcare systems. For example, allicin extracted from *Allium sativum* is a defensive phytochemical with a broad range of biological activities. It inhibits the proliferation of both bacteria and fungi, including antibiotic-resistant strains and furthermore, allicin induces cell-death and inhibits cell proliferation in mammalian cancer cells [219]. Similarly, berberine derived from *Berberis lycium* shows anti-inflammatory, antioxidant, anti-depressant and anti-hypertensive activities [220]. The biological activities of some other phytochemicals isolated from the documented plant species are given in Table 7.

Table 7. Biological activities and mechanism of action of some of the phytochemicals.

| Phytochemical | PubChem CID | Chemical Formula | Biological Activities | Mechanism of Action | Reference(s) |
|--------------|-------------|------------------|-----------------------|---------------------|--------------|
| Rutin        | 5280805     | C_{27}H_{36}O_{16} | Modifies the cognitive and various behavioral symptoms of neurodegenerative diseases | Effects processing, aggregation and action of amyloid beta (Aβ); shift of the oxidant–antioxidant balance allied with neuronal cell loss | [221] |
| Antihyperglycemic | Decrease in carbohydrates’ absorption from the small intestine, inhibition of tissue gluconeogenesis, an increase of tissue glucose uptake, stimulation of insulin secretion from beta cells, and protecting Langerhans’ islet against degeneration | [222] |
| Quercetin    | 5280343     | C_{15}H_{10}O_{7} | Anti-inflammatory     | Inhibits LPS-induced mRNA levels of cytokines in colloid cells, such as tumor necrosis factor (TNF)-a and IL-1a | [223] |
| Anticancer   | Suppression of many kinases involved in the growth of cancer cells, proliferation and metastasis | [224] |
| Lupeol       | 259846      | C_{30}H_{50}O    | Anti-inflammatory     | Suppresses and alters the phagocytic activity of macrophages and T lymphocytes, and suppresses CD4+ T cell mediated cytokine generation | [225] |
Table 7. Cont.

| Phytochemical   | PubChem CID | Chemical Formula | Biological Activities | Mechanism of Action                                                                 | Reference(s) |
|-----------------|-------------|------------------|-----------------------|-------------------------------------------------------------------------------------|--------------|
| Apigenin        | 5280443     | C\(_{15}\)H\(_{10}\)O\(_5\) | Anxiolytic            | Inhibits the binding of flunitrazepam to brain membranes without influencing the binding of muscimol to GABA\(_A\) receptors. | [226]        |
|                 |             |                  | Anti-inflammatory      | Downregulates the expression of IL-1\(\beta\) and TNF-\(\alpha\) in LPS-stimulated mouse macrophages and human monocytes | [227]        |
| Calotropin      | 16142       | C\(_{20}\)H\(_{46}\)O\(_9\) | Cytotoxic              | Upregulated the expression of p27 leading to cell arrest by downregulating the G2/M regulatory proteins, cyclins A and B, and by upregulating the cdk inhibitor, p27 | [228]        |
| α-hederin       | 319412227   | C\(_{41}\)H\(_{66}\)O\(_{12}\) | Anticancer             | Induces depolarization of mitochondrial membrane potential which released Apaf-1 and cytochrome c from the intermembrane space into the cytosol, where they promoted caspase-3 and caspase-9 activation | [229]        |
| Luteolin        | 5280445     | C\(_{15}\)H\(_{10}\)O\(_6\) | Antibacterial          | Disrupts the integrity of the bacterial cell membrane and cell wall, resulting in the leakage of cell contents and damage to the barrier function of the cell wall and membrane | [230]        |
| Rupesin         | 134715087   | C\(_{15}\)H\(_{22}\)O\(_5\) | Antitumor              | Inhibits the proliferation of Glioma Stem Cells (GSCs)                               | [231]        |
| Kaempferol      | 5280863     | C\(_{15}\)H\(_{10}\)O\(_6\) | Anticancer             | Induces apoptosis, cell cycle arrest at the G2/M phase, downregulation of Epithelial–Mesenchymal Transition (EMT)-related markers, and phosphoinositide 3-kinase/protein kinase B signaling pathways | [232]        |
| Rottlerin       | 5281847     | C\(_{30}\)H\(_{28}\)O\(_8\) | Antitumor              | Sensitizes MCF-7 breast cancer cells to TRIAL-mediated apoptosis by PKC\(\delta\)-dependent inhibition of the transcription factor nuclear factor \(\kappa\)B (NF\(\kappa\)B), | [233]        |
| Heterophylline  | 251575      | C\(_{22}\)H\(_{26}\)N\(_2\)O\(_4\) | Alzheimer’s disease    | Inhibits muscle-contracting enzymes acetylcholinesterase and butyrylcholinesterase | [234]        |

On earth, there are approximately 250,000 plant species [234], and only 5–15% of higher plants have been thoroughly examined for the presence of bioactive compounds [235]. Plants have an immeasurable potential to produce a vast diversity of unusual chemical structures known as secondary metabolites that modulate the relationships of organisms with the environment [235]. Additionally, plants possess a precise mechanism to fight infection through the production of phytoalexins, possessing anti-infective activities. Such phytochemicals hold much potential for medicinal applications, and thus, it is coherent and reasonable to investigate the potential of such compounds for both human and animal health care. Studies of the biological activity of ethno-veterinary plants can provide clues about promising leads for the isolation and identification of useful compounds that may
be developed into pharmaceuticals. The present study reported the use of *Chenopodium album* for wound healing by the local inhabitants, and Said et al. [236] have revealed that *C. album* can promote wound healing and tissue regeneration through the modulation of growth factors and their receptors. Similarly, *Foeniculum vulgare* was reported to cure gastrointestinal disorders and Birdane et al. [237] has revealed a protective effect of *F. vulgare* against ethanol-induced gastric mucosal lesions in rats. Thus, following up on ethno-medicinal leads is one of the best approaches to selecting plants for bioactivity screening. There is frequently an overlap between medicinal plants used to alleviate animals and humans. It would make sense that similar treatments are used to treat comparable ailments in humans and their livestock [238]. Ethno-veterinary medicines can function as leads for drug development, but probably a more practical and lucrative exercise would be to develop a preferred preparation through pharmacological research and development, and the ensuing medication can be returned to society with extra impact. Further, local farmers can cultivate such plants, which improves the economic condition of the farmers, and such commercialization can aid in biodiversity conservation. The growing interest in traditional medicine and increasing admiration of ethno-veterinary medicinal plants has been restricted in terms of more advancement by the inaccessibility of information on the effectiveness and assurance of these practices.

4. Conclusions

In the isolated rural areas of the Himalayan region, ethnobotanical knowledge is crucial for preserving plant diversity and treating a variety of diseases. One of the most crucial prevailing systems in the region where modern veterinary healthcare facilities are scarce or in extremely bad condition is the traditional method of treatment. The folk knowledge of ethno-veterinary medicine has its significance in the treatment of livestock diseases in remote and rural areas of the J&K. Farmers alleviate the health-related problems of their livestock using ethno-veterinary medicine, as it is affordable and a more reliable surrogate to synthetic drugs. The seminomadic and nomadic tribes of the Indian Himalayan region in general and the Gujjar and Bakerwal tribes of J&K, in particular, still possess a rich heritage of traditional healthcare systems. In our literature survey, it was found that plant species documented during the present study possess biological activities owing to the presence of various biologically active phytochemicals. Thus, more studies of the biological activities of ethno-veterinary plants can provide clues of promising leads for the isolation and identification of useful compounds that may be developed into pharmaceuticals. Further, there is recurrently an overlap between medicinal plants used to alleviate animals and humans. Thus, ethno-veterinary medicines can function as leads for drug development, but probably a more practical and lucrative exercise would be to develop a preferred preparation by pharmacological research, and the ensuing medication can be returned to the society with extra impact. However, this valuable knowledge-base has become obsolete due to industrialization, urbanization and, above all, lack of recognition by the younger generations. Therefore, there is a dire need to review, document and authenticate the valuable traditional knowledge of ethno-medicinal plants for the human welfare. This study could be a pilot to document the sustainable utilization of frequently used bioresources by understanding the traditional knowledge systems, and will provide important leads for the formulation of novel drugs and medicine.

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