Diphyllobothriasis is a fishborne cestodiasis caused by infection with adult tapeworms belonging to the genus *Diphyllobothrium* Cobbold, 1858 (1–5); the most frequent etiologic agents are *D. latum* and *D. nihonkaiense*. Humans are infected by ingesting raw or undercooked fish infected with larval plerocercoids. Adult tapeworms can grow to \( \approx 2–10 \) m in length in the human small intestine (1–6). Despite the large size of the tapeworms, clinical symptoms can be absent or mild and include mild abdominal pain, watery diarrhea, and abdominal discomfort (3–7). *D. latum* infection can also cause vitamin B\(_{12}\)-deficiency anemia (5).

Diphyllobothriasis caused by *D. nihonkaiense* has been extensively reported in Japan (3,4), but it has also occurred autochthonously in South Korea (8) and the Far Eastern Federal District of Russia (originally reported as *D. klebanovskii* infection [9]). Sporadic cases have been reported in Europe (6), North America (10), and New Zealand (7) in recent years.

In mainland China, 15 cases of diphyllobothriasis among humans have been reported since the first report in 1927 through 2012; the etiologic species was identified as *D. latum* by morphologic characteristics (11–13; Table) and molecular markers (14,15). No cases of diphyllobothriasis had been reported in large cities such as Beijing and Shanghai during 1954–2007 (11). However, we confirm 4 cases of *D. nihonkaiense* infection in humans in Shanghai, previously identified as *D. latum* infection, during 2008–2011, as well as 1 case in the moderately populous city of Harbin in Heilongjiang Province.

### The Study

We examined 5 recent infections of humans with *Diphyllobothrium* spp. (Table, cases 12, 16–19) that occurred in China. Each case had been originally reported as a *D. latum* infection on the basis of morphologic identification only. Case 12 was reported in Harbin City, Heilongjiang Province, in 2009 (13). The 4 cases reported in Shanghai were diagnosed at the National Institute for Parasitic Diseases, Shanghai, on the basis of morphologic features of passed strobila. Case-patient 16 lived in Japan, but it was suggested that he acquired the tapeworm in Shanghai where he had frequently eaten raw salmon. Case-patient 17 was a 10-year-old girl from Japan. Whether she became infected in Shanghai or Japan was unclear because of lack of information. Case-patients 18 and 19 acquired the infection in Shanghai because they had never been abroad.

Because all patients in Shanghai had eaten raw salmon, we decided to re-examine how the causative *Diphyllobothrium* spp. were identified. *D. latum* infection is associated with consumption of freshwater fish such as perch (*Perca* spp.), not Pacific salmon (*Oncorhynchus keta, O. masou*) and Atlantic salmon (*Salmo salar*) in the Northern Hemisphere (1–5). To expand diagnostic parameters and clarify the point of misidentification, we re-identified *Diphyllobothrium* spp. by examining the tapeworms’ morphologic features and using a molecular marker. In a sample from case-patient 12, only proglottids stained with acetic acid–carbyme were available for testing by both methods (Figure 1, panel A). Proglottids obtained from 4 case-patients in Shanghai were preserved in either 10% formalin (case-patient 16) or 70% ethanol (case-patients 17–19) after collection (Table). Parts of the proglottids were embedded in paraffin, and sagittal sections were prepared for morphologic observation.

For molecular identification of the *Diphyllobothrium* spp., genomic DNA samples were extracted from specimens by using a DNeasy Blood & Tissue Kit (QIAGEN, Hilden, Germany). In specimens from case-patients 17–19, the mitochondrial cytochrome *c* oxidase subunit 1 gene (cox1, 1,566 bp) was amplified by PCR by using *Ex Taq* DNA polymerase (Takara Bio, Shiga, Japan) (7). In formalin-fixed samples of proglottids from case-patients 12

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and 16, DNA degradation caused by the fixative meant that only shorter *cox1* fragments (249 bp, corresponding to sites 880–1128 of *cox1*) could be amplified successfully by PCR by using KOD FX DNA polymerase (Toyobo, Osaka, Japan). DNA sequencing of amplicons was performed with a 3100-Advant Genetic Analyzer or 3730 x1 DNA Analyzer (Life Technologies, Foster City, CA, USA). Phylogenetic analysis was performed by the maximum likelihood method (MEGA 5.05, http://megasoftware.net/mega.php) and Bayesian inference (MrBayes version 3.1.2, http://mrbayes.sourceforge.net/). Clades were assessed by bootstrap resampling (1,000 replicates) and a posterior probability (10^6 generations) for the maximum likelihood and Bayesian inference trees, respectively. *Diphyllobothrium* spp. isolated from case-patients 12 and 16 were identified on the basis of morphologic criteria, tapeworms from case-patients 16–19 were identified as *D. latum* on the basis of their 99%–100% sequence identity to *D. nihonkaiense*. Nonetheless, in this study, on the basis of morphologic criteria, tapeworms from case-patients 16, 17, and 19 were identified as *D. latum* (Figure 1, panels B, C, and E) and the tapeworm found in case-patient 18 was identified as *D. nihonkaiense* (Figure 1, panel D).

Phylogenetic trees based on the complete *cox1* nucleotide sequences showed the same topologies in maximum likelihood and Bayesian inference analyses, implying that the 3 isolates from persons in China (case-patients 17–19; GenBank accession numbers AB684621–AB648623) are *D. nihonkaiense* (Figure 2). The 2 isolates (AB684625 and AB684624) from case-patients 12 and 16, respectively, were excluded from the analysis because they produced smaller PCR products, but they were identified as *D. nihonkaiense* on the basis of their 99%–100% sequence identity to *D. nihonkaiense*.

The 5 *Diphyllobothrium* spp. tapeworms examined in this study were previously identified as *D. latum* on the basis of morphologic characteristics, as were 3 of the 5 when we re-examined their morphologic characteristics. However, the 5 etiologic agents were confirmed as *D. nihonkaiense*.

| Case no. | Residence/place of eating fish, if different | Proglottids expelled | Suspected source of infection | Proglottid specimens fixative | Morphologic identification | Molecular identification | GenBank accession no. for *cox1* gene | Reference |
|----------|-------------------------------------------|----------------------|-----------------------------|-----------------------------|--------------------------|---------------------------|-----------------------------|----------|
| 1–4      | Heilongjiang                              | NA                   | NA                          | NA                          | *D. latum*               | NT                        | NA                          | (11)     |
| 5        | Beijing                                   | NA                   | NA                          | NA                          | *D. latum*               | NT                        | NA                          | (11)     |
| 6        | Shanghai                                  | NA                   | NA                          | NA                          | *D. latum*               | NT                        | NA                          | (11)     |
| 7        | Beijing (returned from United States)     | NA                   | NA                          | NA                          | *D. latum*               | NT                        | NA                          | (11)     |
| 8        | Guangzhou (returned from Argentina)       | NA                   | NA                          | NA                          | *D. latum*               | NT                        | NA                          | (11)     |
| 9        | Heilongjiang                              | NA                   | Raw fish                    | NA                          | *D. latum*               | NT                        | NA                          | (11)     |
| 10       | Jilin                                      | NA                   | Raw fish                    | NA                          | *D. latum*               | NT                        | NA                          | (11)     |
| 11       | Fujian (lived in Yokohama, Japan, until 1996) | 2003 Jan            | *PlecoGLOSSUS altivelis*    | NA                          | *D. latum*               | NT                        | NA                          | (12)     |
| 12       | Heilongjiang                              | 2009 Jan             | Raw fish                    | 10% formalin                | *D. latum*               | *D. nihonkaiense*          | AB684625 | (13) and this study |
| 13       | Heilongjiang                              | NA                   | Salmon                      | NA                          | *D. latum*               | *D. latum*                | NA                          | (14)     |
| 14       | Jilin                                      | NA                   | Salmon                      | NA                          | *D. latum*               | *D. latum*                | NA                          | (14)     |
| 15       | Shanghai, 2008–2011/Japan, China           | 2011 Dec             | Raw sea and freshwater fish | NA                          | *D. latum*               | *D. latum*                | NA                          | (15)     |
| 16       | Shanghai/Japan (returned from Japan in June 2008; ate raw salmon in Shanghai) | 2008 Oct             | Raw salmon                  | 10% formalin                | *D. latum*               | *D. nihonkaiense*          | AB684624 | This study         |
| 17       | Shanghai                                  | 2011 Sep             | Raw salmon                  | 70% ethanol                 | *D. latum*               | *D. nihonkaiense*          | AB684621 | This study         |
| 18       | Shanghai, Ate raw salmon in April 2011     | 2011 Jun             | Raw salmon                  | 70% ethanol                 | *D. latum*               | *D. nihonkaiense*          | AB684622 | This study         |
| 19       | Shanghai, Ate raw salmon in 2011           | 2011 Jul             | Raw salmon                  | 70% ethanol                 | *D. latum*               | *D. nihonkaiense*          | AB684623 | This study         |

*Identification of *Diphyllobothrium* spp. was performed by morphologic identification alone in cases 1–11; organisms in each case were identified as *D. latum*. Cases 12–19 were assessed by morphologic and molecular identification; morphologic identification of all specimens was *D. latum*. Molecular identification varied from morphologic findings in 5 of 8 tested specimens. NA, not available; NT, not tested.
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by molecular analysis. This discrepancy in the identity of these agents may be attributed to the morphologic similarities between the species and the century-long confusion between the parasite D. latum and the parasite that caused human diphyllobothriosis associated with the consumption of Pacific salmon in Japan (1–3). Diphyllobothriosis caused by D. nihonkaiense has also been reported in South Korea (8) and in the Far Eastern Federal District of Russia (9) and is considered to be autochthonous and linked to the consumption of wild Pacific salmon in these regions. Therefore, some cases of diphyllobothriosis reported in mainland China were probably caused by infections with D. nihonkaiense; case-patient 12 (13) in this study is considered to have had such a case. However, a recent report stating that the causative species of 2 diphyllobothriosis cases in northeastern China was D. latum, which suggests that D. latum is also indigenous to mainland China (14).

Conclusions

We confirmed human diphyllobothriosis caused by D. nihonkaiense in mainland China by using a mitochondrial DNA marker. Reassessment of a case in Harbin revealed that some, if not all, of the autochthonous diphyllobothriosis cases were likely initially misdiagnosed as D. latum infection because of morphologic similarities between D. nihonkaiense and D. latum tapeworms. Consequently, molecular analysis is indispensable not only for avoiding diagnostic confusion among Diphyllobothrium spp., but also for facilitating the acquisition of reliable epidemiologic and epizootic information and improving clinical relevance and preventive controls for diphyllobothriosis.

Information on diphyllobothriosis and warnings of the potential risks associated with infection by its local species should be disseminated to food handlers, restaurant owners, physicians, and consumers. Because

Figure 1. Diphyllobothriid samples examined in the present study, China, 2008–2012. A) Proglottids stained with acetic acid–carmine from case-patient 12. B–E) Sagittal sections of proglottids stained with hematoxylin-eosin from case-patients 16–19. cs, cirrus sac; ut, uterus; sv, seminal vesicle; ov, ovary; ga, genital atrium. Scale bar in panel A represents 2 mm; scale bars in panels B–E represent 500 μm.

Figure 2. Phylogenetic tree constructed by using the maximum likelihood algorithm (Kimura’s 2-parameter model) on the basis of the complete cox1 sequences of isolates from Diphyllobothrium species found in persons in China and related Diphyllobothrium species. Numbers at nodes are bootstrap values (1,000 replicates) and posterior probabilities (10⁶ generations) for maximum likelihood and Bayesian inference, respectively. Spirometra erinaceieuropaei was used as an outgroup. Scale bar indicates the number of base substitutions per site.
we cannot determine with certainty whether previous diphyllobothriosis cases in mainland China were caused by *D. latum* or *D. nihonkaiense*, identification of *Diphyllobothrium* spp. should be performed with care. In addition, studies on the distribution and sources of infection of *D. latum* and *D. nihonkaiense* on mainland China should be undertaken.

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