Review Article

TRAIL, OPG, and TWEAK in kidney disease: biomarkers or therapeutic targets?

Stella Bernardi1, Rebecca Voltan2, Erika Rimondi2, Elisabetta Melloni2, Daniela Milani2, Carlo Cervellati3, Donato Gemmati4,5, Claudio Celeghini2, Paola Secchiero2, Giorgio Zauli2 and Veronica Tisato2

1Department of Medical Sciences, University of Trieste, Trieste, Italy; 2Department of Morphology, Surgery and Experimental Medicine and LTTA Centre, University of Ferrara, Italy; 3Department of Biomedical and Specialist Surgical Sciences, University of Ferrara, Italy; 4Centre of Hemostasis & Thrombosis, Department of Biomedical and Specialty Surgical Sciences, University of Ferrara, Italy; 5Department of Biomedical and Specialty Surgical Sciences, University Center for Studies on Gender Medicine, University of Ferrara, Italy

Correspondence: Veronica Tisato (veronica.tisato@unife.it)

Ligands and receptors of the tumor necrosis factor (TNF) superfamily regulate immune responses and homeostatic functions with potential diagnostic and therapeutic implications. Kidney disease represents a global public health problem, whose prevalence is rising worldwide, due to the aging of the population and the increasing prevalence of diabetes, hypertension, obesity, and immune disorders. In addition, chronic kidney disease is an independent risk factor for the development of cardiovascular disease, which further increases kidney-related morbidity and mortality. Recently, it has been shown that some TNF superfamily members are actively implicated in renal pathophysiology. These members include TNF-related apoptosis-inducing ligand (TRAIL), its decoy receptor osteoprotegerin (OPG), and TNF-like weaker inducer of apoptosis (TWEAK). All of them have shown the ability to activate crucial pathways involved in kidney disease development and progression (e.g. canonical and non-canonical pathways of the transcription factor nuclear factor-kappa B), as well as the ability to regulate cell proliferation, differentiation, apoptosis, necrosis, inflammation, angiogenesis, and fibrosis with double-edged effects depending on the type and stage of kidney injury. Here we will review the actions of TRAIL, OPG, and TWEAK on diabetic and non-diabetic kidney disease, in order to provide insights into their full clinical potential as biomarkers and/or therapeutic options against kidney disease.

Introduction

The tumor necrosis factor (TNF) superfamily consists of ligands and receptors sharing a key regulatory role in the immune system development and functions, as confirmed by evolutionary studies suggesting that the expansion of this family took place together with the fine-tuning of adaptive immunity [1]. In addition, it has been shown that TNF-superfamily signaling modulates hematopoiesis and morphogenesis as well as different disease states, including cancer and diabetes [1–3].

TNF-superfamily ligands are type II homo-heterotrimeric transmembrane proteins, with a molecular structure characterized by the presence of a TNF Homology Domain, which binds to cystein-rich regions of specific TNF-superfamily receptors [3]. In addition, proteolytic processes, such as those mediated by ADAM, matrylisin, subtilisin-like furin family, and perhaps cysteine proteases, can release soluble TNF superfamily ligands by cleavage of their membrane-bound forms [3]. Likewise, also the receptors of this family, which are mostly type I–III transmembrane proteins, can be present as soluble molecules, due to either a missing transmembrane domain (such as in the case of osteoprotegerin), or proteolytic cleavage, or alternative splicing events [3]. All the main structural characteristics of TNF-superfamily signaling have been recently reviewed by Vanamee and Faustman [4]. In short, the basic signaling unit consists of a trimeric ligand and three receptors, each binding at the interface of two ligand monomers [4]. In addition, TNF-superfamily receptor resting and activation states depend on clustering processes at the cell surface.
Overall, due to the complexity of TNF-superfamily signaling, ligands and receptors exert pleiotropic peripheral effects, such that some TNF-superfamily members are studied in different disease states [5]. These members include TNF-related apoptosis-inducing ligand (TRAIL), osteoprotegerin (OPG), and TNF-like weaker inducer of apoptosis (TWEAK).

The first of them, TRAIL, whose peripheral actions epitomize the complexity of the entire superfamily [4], has attracted much attention as a potential anti-cancer therapeutic target. TRAIL has the ability to induce apoptosis in tumor cells while preserving the normal ones, thanks to the activation of survival/proliferative and autophagy pathways [6–10]. This ability to selectively induce different/opposite cellular responses seems to be due to the high number of receptors that can differentially mediate TRAIL signaling, as well as to the balance with other circulating enzymes and cytokines, such as OPG. During the last two decades, several groups have addressed the role of TRAIL not only in cancer but also in other non-neoplastic diseases. Along with other Authors, we have found an inverse correlation between circulating TRAIL and the presence or the adverse prognosis of different pathological conditions including cardiovascular disease (CVD), type 1 and type 2 diabetes, and neurological disorders [11–21]. In addition, our results have provided insights into the therapeutic potential of recombinant TRAIL administration [22–26]. The second TNF-superfamily member, OPG, is a molecule that was initially identified as a bone resorption inhibitor, as it prevented the binding of receptor activator of nuclear factor kappa-β (RANK) to its ligand RANKL [27–29]. Further studies showed that OPG was a decoy receptor (DcR) also for TRAIL [30], and that it could have additional direct effects on the vasculature and the immune system, independent of its binding to RANKL or TRAIL. Today, also OPG has emerged as a TNF-superfamily member with important effects on acute and chronic cardio-metabolic diseases [31], which might partly depend on the OPG/TRAIL ratio [32]. Third, TWEAK is another TNF-superfamily member with clinical potential. TWEAK was discovered at the same time of TRAIL and OPG [33] and it has recently gained much attention because of its ability to activate canonical and non-canonical pathways of the transcription factor nuclear factor-kappa B (NF-kB) leading to cell proliferation, differentiation, apoptosis, necrosis, inflammation, angiogenesis, and fibrosis [34].

Due to their belonging to the same family, in this review we will focus on the actions of TRAIL, OPG, and TWEAK on the kidney, in order to provide insights into their full clinical potential as biomarkers and/or therapeutic options against kidney disease. Kidney disease represents a global public health problem, whose prevalence is rising worldwide, due to the aging of the population and the increasing prevalence of diabetes, hypertension, obesity, and immune disorders [35]. It is estimated that some 10–15% of the population is affected by chronic kidney disease (CKD), whose end stage requires to be treated by dialysis or kidney transplantation. In addition, CKD is an independent risk factor for the development of CVD, which further increases kidney-related morbidity and mortality [35].

**Overview on TRAIL, OPG, and TWEAK biology**

**TRAIL biology**

Since its discovery dating back to the nineties [36,37], TRAIL (also known as Apo-2 Ligand, CD253, or TNF-SF10) has emerged as a highly pleiotropic cytokine. Encoded by a gene located on chromosome 3 at position 3q26, human TRAIL is a 281 amino acid type II transmembrane protein (291 in the murine form) that can be cleaved and self-organize into soluble homotrimers, which are the ligand biologically active forms (Figure 1). Wiley and colleagues have reported that many tissues, as well as immune system cells, constitutively express significant levels of TRAIL, suggesting that TRAIL – unlike other TNF superfamily members – must not be cytotoxic to most tissues in physiological conditions [36]. The key biological feature of TRAIL is indeed its ability to selectively induce apoptosis in cancer/transformed cells while sparing the normal ones [38]. The mechanisms underlying this sort of ‘TRAIL resistance’ in normal (healthy) cells are still unclear, but scientific evidence suggests that this phenomenon might depend on the local conditions, such as inflammatory and/or pro-oxidative states [7,39,40]. For instance, it has been shown that oxidative stress promoted the redistribution of TRAIL receptors in membrane proteins (rafts), which could facilitate TRAIL signal transduction [7,41,42].

In cancer/transformed cells, TRAIL generally triggers the apoptotic cascade by binding to its two specific type I transmembrane death receptors (DRs) (TRAIL-R1/DR4 and TRAIL-R2/DR5), which harbor an intracellular death domain (DD) that induces caspase activation leading to cell death [43]. The details of the single intracellular steps leading to apoptosis, as well as all the anti-cancer therapeutic strategies targeting DRs, have been reviewed elsewhere [44,45]. In addition to the DRs, TRAIL can bind two transmembrane decoy receptors (DcRs), DcR1 (also known as TRAIL-R3) and DcR2 (also known as TRAIL-R4), which are unable to transmit the apoptotic signal since they do not contain any functional DD (i.e. DcR1 lacks the DD and DcR2 has only a truncated DD) (Figure 2A) [30,46]. Moreover, TRAIL can bind to soluble OPG (also known as TNF receptor superfamily member 11B, or TNFRSF11B),
Figure 1. Graphical structure of TRAIL/OPG/TWEAK genes and proteins
The picture shows a schematic representation of TRAIL/OPG/TWEAK genes (GENATLAS) where exons are indicated by red squares. For TWEAK, a transcript variant representing a shorter transcript encoding the functional protein is also shown. Each gene is shown with the respective schematic protein reporting the main relevant structural domains. Note: figures not drawn to scale. Abbreviation: TM, transmembrane.

which lacks both transmembrane and cytoplasmic residues, such that it acts as a soluble DcR for TRAIL (Figure 2A) [30]. Overall, by binding to its receptors, TRAIL can activate not only apoptosis but also additional cell death pathways, such as necroptosis [47,48] and autophagy [49,50], as well as survival/proliferative and proinflammatory pathways, including those relying on NF-κB, ERK1/ERK2, and Akt signaling (Figure 2A) [51–54]. Although some mediators of TRAIL signaling have been identified, such as caspases and RIPK (receptor-interacting protein kinases), which act as apoptotic and necroptotic regulators [55], it remains to be fully elucidated how TRAIL could activate alternative pathways leading to opposite peripheral effects. So far, TRAIL multifaceted biology seems to be related not only to its complex receptor system, which can be influenced by local stimuli, but also to other conditions, such as the balance between metalloproteinase-2 and tissue inhibitor of metalloprotease-2, which seems to mediate the clearance of circulating TRAIL [56]. In addition, TRAIL biology is unavoidably linked to that of its DcR OPG.

In healthy subjects, the blood levels of TRAIL are in the picogram (per milliliter) range [57] with reported physiological fluctuations due to age and gender [20], and significant changes in several pathological conditions. For example, along with other authors, we have found that TRAIL levels were significantly decreased in patients affected by CVD, such as acute myocardial infarction and coronary syndromes [58,59], as well as in patients with diabetes [17,19]. In addition, clinical studies have shown that there was an inverse correlation between circulating TRAIL and C-reactive protein [59], and between TRAIL and adverse cardio-metabolic outcomes [17,60,61], supporting a prognostic role for TRAIL. Several works carried out in animal models of cardio-metabolic diseases have attempted to understand if TRAIL had a causative role in cardio-metabolic disease development and progression. In this setting, we reported that TRAIL administration had protective effect against atherosclerosis, type 1 and type 2 diabetes, and non-alcoholic fatty liver disease, via the regulation of innate and adaptive immune system [22,25,26,62,63]. Nevertheless, other studies focusing on other tissues/conditions, such as Alzheimer disease and ischemic stroke, showed that TRAIL expression and signaling increased in disease states of the central nervous system (CNS) and promoted their progression [14,59]. It can be speculated that these conflicting effects might be due to the fact that TRAIL is not normally expressed in the CNS [36], while it is in the majority of the other organs and tissues [36].
Figure 2. Schematic representation of TRAIL/OPG/TWEAK signaling pathways

The figure shows a schematic representation of the key pathways mediated by TRAIL/OPG/TWEAK. In (A) TRAIL-mediated apoptosis (intrinsic and extrinsic pathways) and cell survival signaling after interaction between TRAIL and DR4/5 are shown. In the figure, DcR1 and DcR2 are membrane-bounded TRAIL DcRs while OPG acts as soluble DcR for TRAIL. In (B) OPG takes place in the OPG/RANK/RANKL axis as DcR for RANKL. In (C) TWEAK-mediated pathways by interaction with its membrane bounded Fn14 receptor are shown. Soluble CD163 acts as TWEAK DcR. The monocyte/macrophage scavenger membrane receptor CD163 able to bind and internalize TWEAK is also shown. FADD is Fas-associated DD. TRAF is TNF receptor-associated factor. The figure legend shows a schematic representation of all ligands and receptors.

OPG biology

OPG was firstly described as a glycoprotein involved in bone resorption regulation [27–29], and then it emerged as a TNF-superfamily member with multiple implications. The OPG gene, which is located on chromosome 8 at position 8q24 [27], encodes a circulating 401 amino acid glycoprotein with an N-terminal fragment harboring four tandem cysteine-rich motifs and a C-terminal fragment that seems to function as a linker of OPG monomers (Figure 1) [27]. The protein can indeed be detected as a free monomer of 60 kDa or as a homodimer of 120 kDa, which is reported as the most biologically active form [27]. OPG is generally released by several cell types, such as vascular smooth muscle cells, endothelial cells, osteoblasts and immune cells [27,64] and it can have either direct or indirect tissue effects that are due to its binding to RANKL (receptor activator of nuclear factor kappa-β ligand) and/or TRAIL. By binding to RANKL [65], and inhibiting the interaction between RANKL and its membrane receptor RANK, OPG prevents bone remodeling (Figure 2B) [66,67], which represents the rationale behind its current use in patients with osteoporosis [66]. On the other hand, by binding to TRAIL, OPG inevitably affects/modulates TRAIL effects. Finally, by virtue of its heparin-binding domain, OPG can also interact directly with heparin sulfate proteoglycans, which are expressed at the cell surface, whereby it mediates additional ligand-independent effects that are still largely unexplored [68,69]. Not surprisingly, OPG biology lies at the intersection of different pathways, such as those regulating the vessels and the bone [31].

As reported for TRAIL, the blood levels of OPG are in the picogram (per milliliter) range with reported physiological fluctuations due to age and gender as well the blood-group, with higher circulating levels associated with non-zero blood groups [70]. OPG circulating levels change in several diseases. For example, along with other authors, we have found that OPG levels were significantly increased in patients affected by atherosclerosis [71], heart failure [72], as well as metabolic disturbances and diabetes [32,73]. In addition, OPG levels were significantly associated with markers of vascular damage [32]. Consistent with these results, several studies have shown that OPG is an independent risk factor for cardiovascular (CV) morbidity and mortality [71,74,75], such that OPG is considered a biomarker of...
CVD onset and progression [76]. Nevertheless, it remains to be clarified whether OPG has any causative/pathogenic role in vascular damage, as the studies carried out so far have provided conflicting results [16,31,32].

**TWEAK biology**

TWEAK (also known as TNFSF12 or Apo3L) was firstly described in 1997, when Chicheportiche and colleagues carried out a macrophage cDNA library screening and found that one of the cDNAs was a TNF family member, which, like other members of the TNF-supersfamily, was able to induce apoptosis even though in a weaker and non-DD-dependent fashion [33]. Encoded by a gene located on chromosome 17 at position q26, TWEAK is a 249 amino acid type II transmembrane protein that can be cleaved by a furin endoprotease, with the generation of a 18 kDa soluble protein (156 amino acids), which is the most common form of TWEAK ligand [33] (Figure 1). Interestingly, both soluble and membrane TWEAK seem to activate different intracellular signaling pathways, leading to different biological responses [34]. So far, two receptors of TWEAK have been identified: the transmembrane fibroblast growth factor-inducible molecule 14 (Fn14) [77], which is the only receptor mediating TWEAK actions/effects, and the macrophage-derived scavenger receptor CD163. CD163 is expressed by monocytes and macrophages as a type I transmembrane protein and as a soluble protein (sCD163), which can both act as substitute receptors in the cells lacking Fn14 [78]. Nevertheless, the existence of other TWEAK receptors has been speculated [79]. Like TRAIL and OPG, soluble TWEAK is expressed by several cells of the immune system, such as monocytes, natural killer (NK) cells, and dendritic cells [80], where it seems to promote the switch from innate to adaptive immunity responses [80]. Additional non-hematopoietic sources of TWEAK are represented by endothelial and smooth muscle cells [81]. Generally, the activation of the TWEAK/Fn14 pathway is strictly dependent on Fn14 up-regulation. In normal (healthy) conditions, both TWEAK and Fn14 appear to be expressed at low levels; by contrast, in various disease conditions, including autoimmune and inflammatory/neuro-inflammatory diseases [82–87] as well as several types of cancer [88–93], both Fn14 and TWEAK expressions are up-regulated.

At the molecular level, TWEAK/Fn14 signaling initiates with TWEAK binding to the extracellular domain of its receptor, then the signal is transduced by TNF receptor-associated factors (TRAFs), which mediate the activation of both canonical and non-canonical NF-κB pathways (NF-κB1 and NF-κB2, respectively) [94–97] (Figure 2C). Apart from NF-κB, TWEAK can activate other pathways, such as mitogen-activated protein kinase (MAPK) with ERK, JNK and p38 signaling, as well as AKT/PI3K, Wnt/GSK3B, and NIK/MAP3K14 pathways (Figure 2C). As expected, TWEAK-mediated biological effects span from cell survival/proliferation, migration/invasion, cell apoptosis, and angiogenic activity, to the overexpression of inflammatory cytokines and inflammation as well as modulation of cell differentiation pathways. In addition, TWEAK seems to interact with Dicer (a ribonuclease involved processing microRNA precursors into microRNAs), reducing its pre-microRNA processing activity and thus affecting microRNA-regulated gene expression [98]. In general, the multiple (and sometimes opposite) effects of TWEAK seem to be due to different types of activation with a ‘transient’ activation involved into reparative/regenerative processes and an ‘over/sustained activation’ responsible for progressive tissue damage and cancer [99].

**TRAIL, OPG, and TWEAK in the normal kidney**

Several TNF superfamily members, such as TNF, Fas ligand, as well as TRAIL, OPG, and TWEAK are expressed in the human kidney [27,33,100–102]. Their sources include not only intrinsic kidney cells, but also the vessels [16,103], as well as infiltrating macrophages or lymphocytes. In general, given that knockout mice lacking the gene for TRAIL, OPG, or TWEAK do not exhibit signs of kidney pathology, it has been argued that these ligands are not strictly required for kidney development and physiology [80,104,105].

When looking at intrinsic kidney cells, studies on the tissue distribution of TRAIL and its receptors have shown that – in healthy conditions – TRAIL is expressed by the renal tubules, but not by the glomeruli [100]. In particular, Spierings and colleagues found that TRAIL, TRAIL-R1, and TRAIL-R2 were expressed in the tubuli contorti, while TRAIL-R2 was expressed in the Henle’s loop cells [106]. In vitro experiments on tubular cells have shown that TRAIL has different effects ranging from apoptotic [100] to proliferative actions [107], depending on local conditions. Also OPG has been found expressed in the kidney, including kidney samples [27,101], cultured tubular cells [100], and urinary exosome-like vesicles [108]. In particular, Benito-Martín and colleagues found that cultured human proximal tubular epithelial cells constitutively secreted exosome-like vesicles containing OPG [108], hypothesizing that tubular cell exosomes might have a regulatory role in matrix deposition, inflammation, and apoptosis. By contrast, exosome-like vesicles did not contain TRAIL or TWEAK [108].

As compared with TRAIL and OPG, experimental/animal studies have shown that TWEAK and its receptor Fn14 are constitutively expressed not only by tubular but also by glomerular cells [109,110], such as mesangial cells and...
podocytes [110,111]. In normal conditions, TWEAK and Fn14 expression is rather low but it becomes significant in the presence of tissue damage/inflammatory processes [111,112]. This is consistent with experimental and human studies, showing that Fn14 was not expressed in the normal kidney, but its expression together with that of TWEAK greatly increased in diverse forms of acute kidney injury and CKD, such as proteinuric and non-proteinuric, immune- and non-immune-mediated kidney disease [113]. In addition, experimental studies have shown that, as many other TNF-superfamily members, such as TRAIL, TWEAK can induce different cellular responses, including proliferation [114], inflammation [110], as well as apoptosis [115], depending on the milieu conditions.

### Acute kidney injury and chronic kidney disease

As a premise to the studies investigating the actions of TRAIL, OPG, and TWEAK on kidney disease, it has to be said that the loss of kidney function can be acute (<90 days) or chronic (>90 days) [116], and it can have several causes that contribute and/or cause local activation of common damaging pathways such as inflammation, fibrosis, and cell death (with apoptosis, necroptosis, and necrosis) [117,118]. Acute kidney injury (also referred to as acute kidney disease when it lasts more than 7 and less than 90 days) has an estimated incidence rate of 20–30% [119]. Classically, the causes of AKI/AKD can be classified based on the mechanisms leading to kidney damage. These include pre-renal (e.g. reduced kidney perfusion), intra-renal (direct renal parenchymal injury), and post-renal causes (e.g. obstruction of urinary tract). The etiology can significantly vary in different regions of the world [116]. In high-income countries AKI/AKD is generally seen in adults with multiple comorbidities – especially diabetes and CVD – who are exposed to nephrotoxins, such as radiocontrast media, antibiotics, or to infections, and/or dehydration. In low- and middle-income countries AKI/AKD affects younger patients and it is most often due to episodes of infection, diarrhea, and/or obstetric complications [116].

It has to be noted that an episode of AKI/AKD is not only associated with short-term but also with long-term adverse outcomes, such as incomplete or non-recovery of kidney function with development of CKD and a reduction in survival. Apart from AKI/AKD, the most common contributing factors to CKD include genetic abnormalities and ageing, while the most common underlying diseases associated with CKD are diabetes [120] and hypertension [117]. However, CKD can be also due to infectious diseases, autoimmune diseases (such as lupus), and medications. Overall, it is estimated that CKD affects 10-15% of the population. As CKD progresses, it may require renal replacement therapies, such as peritoneal dialysis, hemodialysis, or kidney transplantation with severe consequences on patient survival. It has in fact been reported that life expectancy is one-third of that of the general population for patients on dialysis, while for patients receiving a kidney transplant is 45–85% of that of the general population [117]. Given that one of the most common cause of AKI/AKD/CKD is diabetes and that diabetes accounts for 33% of patients initiating renal replacement therapies worldwide [120], we will divide the studies on the actions of TRAIL, OPG, and TWEAK into those focusing on non-diabetic and those focusing on diabetic kidney disease.

### TRAIL, OPG, and TWEAK in non-diabetic kidney disease

#### TRAIL in non-diabetic kidney disease

Most of the works on TRAIL have explored its anti-cancer activity for the treatment of renal carcinoma. As a result, the full significance of TRAIL on non-neoplastic kidney disease has not been fully elucidated yet. Overall, experimental and clinical studies have demonstrated that TRAIL is up-regulated in different kidney diseases, ranging from diabetic nephropathy [100] to non-diabetic conditions, which include lupus nephritis [107], minimal-change nephrotic syndrome (MCNS) [121], rejected kidney transplant, as well as acute kidney injury [122], or renal ischemia reperfusion injury [123]. Nevertheless, in these studies, TRAIL up-regulation was not necessarily followed by an increase in circulating TRAIL. For example, although TRAIL was up-regulated on mononuclear cells in patients with MCNS, circulating TRAIL levels did not change between the groups. In addition, in patients with autosomal dominant polycystic kidney disease, circulating TRAIL was actually lower than in healthy volunteers [124]. This has been ascribed to the fact that TRAIL is widely expressed and its circulating levels might come from different tissue sources outside the kidney. Other explanations include a tissue uptake of TRAIL by its specific receptors (including the DcR OPG). Consistent with this hypothesis, TRAIL receptors were found up-regulated in kidney biopsies of various glomerular diseases [125]. Interestingly, in a recent paper evaluating the association between a vast number of proteins and kidney function, TRAIL-R2 turned out to be the protein most strongly associated with kidney function decline [126].

Interestingly, Liabeuf and colleagues have recently reported that circulating TRAIL levels are inversely associated with the mortality risk in CKD patients [127], suggesting a protective role for TRAIL. Consistent with this, in vitro experiments on primary renal proximal tubular epithelial cells showed that TRAIL did not stimulate apoptosis, but rather proliferation [107]. By contrast, blockade of TRAIL ameliorated tissue damage in two animal models of kidney..
Table 1 Studies including the TNF-superfamily triad members TRAIL/OPG/TWEAK as biomarkers and/or therapeutic targets in renal pathological conditions (reported on www.clinicaltrials.gov)

| Disease pathological conditions                  | Study type | TNF-superfamily member | Intervention(s)                                                                                                           | Phase | Status | Study ID     |
|--------------------------------------------------|------------|------------------------|--------------------------------------------------------------------------------------------------------------------------|-------|--------|--------------|
| CKD                                              | Int.       | OPG levels (biomarker) | Plasma OPG level; FGF-23 level; vascular calcification score                                                            | na    | R      | NCT02808572 |
| CKD                                              | Int.       | OPG levels (biomarker) | Plasma OPG level; FGF-23 level; vascular calcification score                                                            | na    | R      | NCT02813642 |
| Renal failure, haemodialysis                     | Int.       | OPG levels (biomarker) | Low molecular weight heparin; unfractioned heparin                                                                      | na    | U      | NCT00669721 |
| Vitamin D Def., vascular calcification, ESRF, peritoneal dialysis | Int.       | OPG levels (biomarker) | Cholecalciferol; placebo                                                                                               | IV    | A      | NCT02598635 |
| Chronic renal failure                            | Obs.       | OPG levels (biomarker) | Identification of CV risk factors linked to renal failure progression                                                    | na    | Unk    | NCT00608998 |
| Renal transplant                                 | Int.       | OPG levels (biomarker) | Zortress, everolimus                                                                                                     | na    | With.  | NCT01612299 |
| Renal insufficiency, CKD, hyperphosphatemia bone diseases | Int.       | OPG levels (biomarker) | Lanthanum carbonate; calcium carbonate                                                                                   | IV    | E      | NCT02237534 |
| Kidney failure, chronic haemodialysis            | Int.       | OPG levels (biomarker) | Apabetalone; placebos                                                                                                     | I-II  | A*     | NCT03160430 |
| Clear-cell, metastatic renal cell carcinoma, bone metastases | Int.       | OPG levels (biomarker) | XOFIGO radium-223 dichloride                                                                                             | I-II  | A      | NCT02880943 |
| Chronic kidney failure; inflammation vascular calcification | Obs.     | OPG levels (biomarker) | Vascular calcification's risk factors in haemodialysis patients                                                          | na    | C      | NCT00694824 |
| Fabry disease                                    | Int.       | OPG levels (biomarker) | RVX000222                                                                                                               | I-II  | A      | NCT03228940 |
| Osteoporosis and CKD                             | Int.       | OPG levels (biomarker) | Denosumab; placebo                                                                                                       | IV    | A*     | NCT02792413 |
| Renal cell carcinoma                             | Obs.       | OPG levels (biomarker) | Assessment of bone biomarkers for TKI response in RCC with bone metastases, HRQol. and comparison of imaging techniques | na    | T      | NCT02747173 |
| Renal transplantation, immunosuppression         | Int.       | OPG levels (biomarker) | Everolimus                                                                                                               | IV    | Unk    | NCT01239472 |
| Renal cancer                                     | Int.       | TRAIL (therapeutic)    | T-cells transduced with T-cell receptor recognizing TRAIL bound to the DR4                                              | I/II  | T      | NCT00923390 |
| Diabetic patients with proteinuria               | Int.       | TWEAK levels (biomarker) | Calcium channel blocker amloidine                                                                                       | IV    | C      | NCT01738945 |
| Diabetes, hypertension proteinuria               | Int.       | TWEAK levels (biomarker) | Renin angiotensin system blockade, calcium channel blocker amloidine; valsartan                                         | IV    | C      | NCT00921570 |
| Lupus nephritis                                  | Int.       | TWEAK (therapeutic)    | BIB023 anti TWEAK monoclonal antibody; mycophenolate mofetil; oral corticosteroids                                       | II    | T      | NCT01930890 |
| Lupus nephritis                                  | Int.       | TWEAK (therapeutic)    | BIB023 anti TWEAK monoclonal antibody; placebo; mycophenolate mofetil; oral corticosteroids                             | II    | T      | NCT01499355 |
| Advanced solid tumours                           | Int.       | Anti Fn14 (therapeutic) | PDL192 anti-TWEAKR monoclonal antibody                                                                                   | I     | C      | NCT00738784 |

A: active; C: completed; T: terminated; Int: interventional; Obs: observational; With: withdrawn; R: recruiting; Unk: unknown; *: not recruiting yet; na.: not available (accessed November 2018).

disease, namely renal ischemia reperfusion injury [123], and burn-related acute kidney injury [122]. These contradictory findings between human and animal studies might be partly due to a different regulation and function of TRAIL between humans and mice, as well as in different settings (i.e. AKI or CKD) [102]. In any case, to date, it has not been clarified if TRAIL up-regulation at a tissue level is harmful or protective, and/or if TRAIL could be used as a biomarker or as a therapeutic target in patients with CKD.

**OPG in non-diabetic kidney disease**

On the other hand, there is growing interest in OPG as a biomarker in patients with CKD (Table 1). Several clinical
studies have shown that OPG increases in CKD [128,129], including patients undergoing renal replacement therapy [130]. The only ‘exception to the rule’ seems to be nephrotic syndrome, where Mohamed and colleagues found that OPG levels were significantly lower as compared with control patients, which might have been due to either protein loss or steroid effect that are typical of this condition [131]. Nevertheless, along with other Authors, we have recently reported that there is a significant inverse association between OPG and renal function [132]. Consistent with these data, high levels of OPG have been associated with long-term risk of renal decline, renal disease hospitalizations, and/or deaths in elderly women [133]. Similar results, which will be summarized in the following paragraphs, have been obtained in patients with diabetic kidney disease. Given that in one of our previous works we found that OPG administration was associated with proinflammatory and pro-oxidative renal changes, it can be speculated that OPG might be not only a risk marker but also a risk factor of CKD [132].

Apart from circulating OPG, also urinary OPG has raised interest as a potential biomarker of renal disease. Several studies have reported a correlation between urinary OPG and both disease status and prognosis of patients with lupus nephritis [134,135]. In particular, Gupta and Kiani showed an association between urinary OPG and renal disease activity. In addition, urinary OPG was elevated in patients with poor response to therapy, disease relapse or CKD development [134]. Overall, all these studies suggest that OPG could serve as a biomarker of CKD development and progression [136,137]. For this reason, several ongoing clinical studies have included OPG measurement to monitor CKD progression and its complications (Table 1).

Besides CKD development and progression, it has been argued that measuring OPG could help differentiate between low-turnover and high-turnover osteodystrophy in hemodialysis patients. Nevertheless, the studies focusing on this topic have shown contradictory results. In a study on 39 chronic hemodialysis patients who underwent bone biopsies, OPG levels were lower in the patients with low-turnover bone disease than in those with predominant hyperparathyroidism or mixed osteodystrophy, for intact PTH levels lower than 300 pg/ml [138]. By contrast, Haas and colleagues found that OPG levels were lower in patients with high-turnover bone disease as compared with those with low or normal-turnover bone disease [139].

**TWEAK in non-diabetic kidney disease**

As compared with TRAIL and OPG, there is much more information on the actions of TWEAK and Fn14 on the kidney. Table 2 summarizes the effects that TWEAK has on different kidney cells in vitro [140], where it promotes fibrosis, inflammation, proliferation, as well as cell death [81,85,94,109,110,114,115,141–159]. In line with these effects, in vivo studies suggest that TWEAK/Fn14 blockade might protect against renal injury/disease (Table 2). In models of AKI, such as folic acid nephropathy, which is characterized by tubular cell death, proliferation, and inflammation, TWEAK deficiency or Ab anti-TWEAK decreased kidney apoptosis and inflammation [114,145,146] and improved kidney function [114,160]. Similar effects were found in a model of AKI induced by ischemia-reperfusion, where Fn14 blockade reduced renal apoptosis, fibrosis, and inflammation [161]. Consistent with these observations, TWEAK administration exacerbated proinflammatory changes in the kidney of high-fat diet-fed ApoE-knockout mice, while anti-TWEAK neutralizing antibodies significantly reduced renal, as well as vascular inflammation [162]. Also in a model of chronic graft-versus-host lupus nephritis, Fn14 deficiency and/or anti-TWEAK neutralizing antibodies were able to significantly decrease proinflammatory cytokine expression, macrophage infiltration, as well as proteinuria [163]. In addition, TWEAK deficiency significantly reduced apoptosis, inflammation, and fibrosis after ureter ligation [143], as well as the cell proliferation in the remnant kidney after unilateral nephrectomy [114], suggesting an involvement of TWEAK/Fn14 axis in the compensatory tubular cell proliferation that follows uninephrectomy [114] (Table 2).

Overall, these studies have provided sound evidence for the development and use of anti-TWEAK therapies against renal injury. Consistent with this, a humanized anti-TWEAK monoclonal IgG antibody BI023 is currently under investigation. The first study on the effects of BI023 showed that a single dose had a favorable safety and tolerability profile in patients with rheumatoid arthritis [164], where it reduced serum biomarkers of inflammation. However, in the ATLAS study (NCT01499355), the same treatment did not demonstrate sufficient efficacy when added to mycophenolate and corticosteroids, in patients with lupus nephritis, such that the best modality to use anti-TWEAK therapies is still under investigation [164,165].

Apart from the potential of anti-TWEAK therapies, accumulating evidence suggests that TWEAK could serve also as a CKD biomarker [166] (Table 1). Rather surprisingly, circulating TWEAK seems to be lower in patients undergoing hemodialysis than healthy controls [167,168], and to increase in patients who have undergone renal transplant [168]. Nevertheless, within the hemodialysis range, higher TWEAK levels have been found associated with a shorter time to death, and hemodialysis patients with high TWEAK have been found at increased CV and all-cause mortality...
Table 2 Renal actions of TWEAK/Fn14: key findings from *in vitro* and *in vivo* models

| **In vitro key effect** | **Cell type**       | **Mechanisms**                                                                                     | **Ref.** |
|------------------------|---------------------|---------------------------------------------------------------------------------------------------|----------|
| Fibrosis               | Mesangial cells     | TGFbeta1 and fibronectin increase through PKG-I down-regulation                                    | [141]    |
|                        | Tubular cells       | Endothelial–mesenchymal transition via NF-κB and ERK activation: F-actin redistribution, loss of epithelial and tight junction proteins, vimentin expression | [142]    |
| Renal fibroblasts      |                     | Decrease in collagen I and fibronectin protein levels                                              | [143]    |
| Inflammation           | Tubular cells       | MCP-1, RANTES increase via NF-κB and JAK2 kinase activation                                        | [144]    |
|                        |                     | MCP-1, RANTES, and IL-6 increase                                                                   | [145]    |
|                        |                     | CCL21 increase via non-canonical NF-κB activation                                                   | [96]     |
|                        |                     | MXCL16 increase via NF-κB                                                                          | [146]    |
|                        |                     | CD74 and DDT increase                                                                               | [147]    |
|                        |                     | IL-6 and other chemokines via EGFR activation, ERK activation                                       | [148]    |
|                        |                     | CXCL10 increase via MAPK14 and non-canonical NF-κB pathway                                          | [96]     |
|                        |                     | Modulation of NF-κB components: Bcl-3 overexpression – which decreases anti-inflammatory anti-apoptotic effects | [151]    |
|                        |                     | Induction of multiple inflammatory cytokines/chemokines and adhesion molecules                     | [110]    |
|                        |                     | MCP-1 increase via NF-κB                                                                          | [152]    |
|                        |                     | CCL19, RANTES increase via NF-κB                                                                    | [153]    |
|                        |                     | CCL21 increase via non-canonical NF-κB pathway                                                     | [96]     |
|                        |                     | Induction of multiple inflammatory cytokines/chemokines and adhesion molecules                     | [110]    |
|                        |                     | MCP-1, RANTES, CXCL10, and CXCL1 increase                                                          | [109]    |
| Proliferation          | Tubular cells       | Cell number increase, cyclin D1 expression via MAPK (ERK/p38), PI3K/Akt, NF-κB                    | [114]    |
|                        | Mesangial cells     | Promotion of cell proliferation and cell cycle activity                                            | [85]     |
|                        |                     | Increase in mitosis number, cyclin D1 expression via Ras/ERK pathway                               | [143]    |
| Cell death             | Tubular cells       | (Late) Necroptosis via RIPK1, RIPK3, MLKL                                                          | [154]    |
|                        |                     | Apoptosis and increased inflammatory gene expression                                               | [155]    |
|                        |                     | In an inflammatory milieu, induction of apoptosis, activation of caspase-8,-9,-3, Bid cleavage and mitochondrial injury | [115]    |
| Others                 | Tubular cells       | Klotho down-regulation via NF-κB                                                                    | [156]    |
|                        |                     | MAGE02 up-regulation – modulation of electrolyte transport                                         | [157]    |
|                        |                     | PGC-1alpha and mitochondrial function down-regulation                                               | [158]    |
|                        | Endothelial cells   | Endothelin-1 increase and ECE1 up-regulation via AP-1 and NF-κB                                      | [159]    |
| Vascular smooth muscle cells |                     | Cell growth and migration, enhanced FGF-2 and VEGF-A mitogenic activity                          | [81]     |
|                        |                     | Enhanced inorganic phosphate-induced calcification via both canonical and non-canonical NF-κB pathways | [94]     |

| **In vivo eperimental model** | **Intervention** | **Effect**                                                                                     | **Ref.** |
|-------------------------------|------------------|------------------------------------------------------------------------------------------------|----------|
| Healthy state                 | TWEAK administration | Increase in inflammation (chemokines and IL-6) via NF-κB activation                               | [145]    |
|                               |                   | Increase in inflammation (CCL21) via non-canonical NF-κB                                         | [96]     |
|                               |                   | Increase in inflammation (MXCL16 and CD6 infiltration) via NF-κB                                   | [146]    |
|                               |                   | Klotho reduction                                                                                 | [156]    |
|                               |                   | PGC-1alpha reduction                                                                             | [158]    |
| Folic acid nephropathy        | Anti-TWEAK AB     | Reduction in inflammation (chemokines and IL-6)                                                 | [145]    |
|                               | TWEAK-KO mice     | Reduction in apoptosis and proliferation and renal function improvement                           | [114]    |
|                               | Anti-TWEAK AB TWEAK-KO mice | Reduction in inflammation (CCL21)                                                               | [96]     |
|                               | Anti-TWEAK AB     | Reduction in inflammation (MXCL16 and CD6 infiltration)                                          | [146]    |
|                               | Anti-TWEAK AB TWEAK-KO mice | Reversal in klotho down-regulation                                                              | [156]    |
| I/R injury                    | Anti-TWEAK AB     | Reversal PGC-1alpha                                                                              | [158]    |
|                               | Fn14 blockade     | Reduction in fibrosis and increased survival                                                     | [161]    |
Table 2 Renal actions of TWEAK/Fn14: key findings from in vitro and in vivo models (Continued)

| In vivo experimental model | Intervention | Effect | Ref. |
|---------------------------|--------------|--------|------|
| HFD-fed ApoE-KO mice     | TWEAK admin.  | Increase in inflammation (RANTES, MCP-1, macrophage infiltration) via NF-κB | [162] |
|                           | Anti-TWEAK AB | Reduction in inflammation (RANTES, MCP-1, macrophage infiltration) | |
| cGVH-induced lupus        | Fn14 blockade | Reduction in IgG deposition, IL-6, MCP-1, RANTES, IP-10, macrophage infiltration | [163] |
|                           | Anti-TWEAK AB | Reduction in proteinuria | |
| Unilateral nephrectomy    | TWEAK-KO mice | Reduction in tubular cell proliferation in the remnant kidney | [114] |
| Ureter ligation           | TWEAK-KO mice | Reduction in apoptosis, inflammation, and fibrosis | [143] |
|                           | Adeno-TWEAK   | Increase in apoptosis, inflammation, and fibrosis | [143] |

Abbreviations: AB: antibody; AP-1: activator protein 1; Bcl3: B-cell lymphoma 3-encoded protein; CCL19: chemokine (C–C motif) ligand 19; CCL21: chemokine (C–C motif) ligand 21; CD74: cluster of differentiation 74; CXCL1: chemokine (C–X–C motif) ligand 1; CXCL10: C–X–C motif chemokine 10 (also known as IP-10); CXCL16: chemokine (C–X–C motif) ligand 16; DDT: D-dopachrome tautomerase cytokine (also known as MIF-2); ECE-1: endothelin-converting enzyme-1; EGFR: epidermal growth factor receptor; ERK: extracellular signal-regulated kinase; FGF-2: fibroblast growth factor 2; IL: interleukin; I/R: ischemia/reperfusion; JAK2: Janus Kinase 2; KO: knockout; MAGED2: melanoma antigen-encoding gene D2; MAPK: mitogen-activated protein kinase; MAP3K14: mitogen-activated protein kinase kinase kinase 14 (also known as NIK); MCP-1: monocyte chemoattractant protein 1; MLKL: mixed lineage domain-like protein; NF-κB: nuclear factor kappa-light-chain-enhancer of activated B cells; NF-κBz: NF-κB inhibitor Zeta; PGC-1 alpha: peroxisome proliferator-activated receptor-γ coactivator-1 alpha; PI3K: phosphoinositide 3-kinase; PKG-I: protein kinase G-I; RANTES: regulated on activation, normal T cell expressed and secreted (also known as CCL5); RIPK: receptor interacting protein kinase; TGFβ1: transforming growth factor beta 1; VEGF-A: vascular endothelial growth factor A.

risk [167]. Based on these results, Carrerro and colleagues have put forward that TWEAK might be an additive, but not a primary marker, of the high mortality rate seen in hemodialysis patients [167]. With respect to lupus nephritis, while circulating TWEAK does not discriminate between patients with and without lupus nephritis [169,170], urinary (uTWEAK) has been found selectively higher in patients with lupus nephritis as compared with other autoimmune disease patients and healthy subjects [169], such that uTWEAK measurement has been proposed as a non-invasive tool to monitor this disease [171,172].

TRAIL, OPG, and TWEAK in diabetes and diabetic kidney disease

OPG and diabetic nephropathy

Diabetes mellitus is now globally the single leading cause of end-stage renal disease (ESRD) and renal replacement therapy worldwide [173]. So far, OPG is probably one of the TNF superfamily members that have been studied the most in the setting of diabetic nephropathy. First of all, experimental studies have shown that the onset of diabetes is associated with an increase in circulating OPG [174], and that diabetes significantly increases OPG expression in peripheral tissues, such as the vessels [16,175]. In addition, microarray studies have confirmed that OPG is up-regulated in the kidneys of diabetic patients [100]. Moreover, it has been shown that OPG levels, which are elevated in patients with type 1 diabetes mellitus (T1DM) [176], correlate with glycemic status [177]. Similar results have been found in patients with type 2 diabetes mellitus (T2DM), who displayed significantly higher levels of OPG as compared with healthy controls, and the patients whose glucose levels were poorly controlled had the highest levels of OPG. All these studies are summarized in a recent review by Perez de Ciriza and colleagues [178].

With respect to diabetic nephropathy, OPG levels have been found significantly higher in patients with T1DM and diabetic nephropathy as compared with those with normal kidney function [177,179]. In these patients, OPG levels have been associated not only with the presence but also with the severity of diabetic kidney disease [180]. Likewise, OPG levels have been found significantly higher in patients with T2DM and kidney damage as compared with those with normal kidney function, where OPG was found to be independently associated with the severity of diabetic nephropathy [181]. Interestingly, several experimental studies suggest that OPG is not only a risk marker but also a risk factor of disease, meaning that it has a causal role on disease development and progression. This is supported by the observations that OPG administration was associated not only with pancreatic [182] and adipose tissue damage [73], and metabolic abnormalities, but also with kidney damage [132].
TRAIL and diabetic nephropathy

In keeping with the view that TRAIL and OPG have opposite effects [32], the literature suggests that TRAIL might have a protective role against diabetes. The first evidence of a link between TRAIL and diabetes was provided by the observation that TRAIL-knockout mice developed heightened autoimmune responses and diseases, such as autoimmune diabetes [183]. Further experimental studies strengthened this relationship, as TRAIL blockade exacerbated T1DM [184] and TRAIL delivery attenuated disease severity [26,185]. In line with these observations, we have demonstrated that TRAIL treatment significantly ameliorated glucose abnormalities in high-fat diet-fed mice [22,25].

Other Authors have reported that TRAIL deficiency worsened glucose control in T2DM animal models [186,187]. In addition, looking at the kidney, the same authors found that TRAIL deficiency contributed to diabetic nephropathy in a mouse model of T2DM, as TRAIL-knockout mice displayed increased urinary protein loss and greater glomerular damage as compared with wild-type mice [187].

Clinical studies have shown that TRAIL levels are significantly lower in patients with T1DM [17] and T2DM [188,189] at onset as compared with healthy subjects, and that this reduction becomes less pronounced a few months after the start of the therapy [17,188]. A decrease in circulating TRAIL has been found also in patients with diabetic nephropathy as compared with healthy controls [190–192]. By contrast, studies on the tissue expression of TRAIL have shown that in T1DM patients there is an increase in TRAIL positive T-cells [193,194]. As for the kidney, TRAIL expression was found increased in renal biopsies of diabetic patients, where it became detectable also in the glomeruli. Based on in vitro studies showing that TRAIL had apoptotic effects on tubular cells exposed to high glucose and pro-inflammatory conditions, Lorz and colleagues have hypothesized that TRAIL could contribute to diabetic nephropathy [100]. Nevertheless, given the limited number of papers and their partly contradictory results, at this stage, it is difficult to draw any conclusion on the exact role of TRAIL in the development and progression of diabetic nephropathy. Further studies are needed to clarify TRAIL significance and potential applications in diabetic kidney disease.

TWEAK and diabetic nephropathy

Despite being largely studied in acute kidney injury, TWEAK actions/significance in the setting of diabetic nephropathy remain to be clarified. Nevertheless, cross-sectional studies have shown a link between the soluble circulating TWEAK and diabetes. In particular, T1DM patients exhibited lower levels of TWEAK as compared with healthy subjects [195]. Likewise, also T2DM patients had lower levels of TWEAK and, in this setting, kidney disease had an additive and independent effect on it. In other words, as compared with healthy subjects, whose TWEAK levels were 669 ± 201 ug/l, in patients with diabetes TWEAK levels were 516 ± 187 ug/l, and in those with diabetes and chronic hemodialysis TWEAK levels were 317 ± 132 ug/l [196]. In addition, a large prospective nested case–control study has shown that TWEAK concentrations were significantly lower in patients who develop diabetes, highlighting that decreased circulating TWEAK could be used to predict T2DM [197].

Given that TWEAK-transgenic mice exhibited greater body weight gain and fat mass as well as insulin resistance and glucose intolerance, and that TWEAK inhibited insulin-stimulated glucose uptake in vitro, it has been argued that TWEAK might promote metabolic abnormalities and diabetes development [198]. Therefore, TWEAK reduction in patients with diabetes is counterintuitive and the mechanisms underlying such findings are poorly understood. The same could be said for nephropathy/diabetic nephropathy, as TWEAK administration worsened renal damage in apoE-knockout mice [162], and yet diabetic patients undergoing hemodialysis exhibit low levels of TWEAK [196]. To explain this apparent paradox the hypotheses that have been postulated include a possible uptake of TWEAK by Fn14, which increases in disease states/inflammatory conditions [199], or the increase in CD163, which is a DcR for TWEAK [78], and it increases in inflammatory conditions, as well as in CKD, leading to a reduction in the TWEAK/CD163 ratio [200].

Role of TRAIL, OPG, and TWEAK in cardiovascular disease of CKD patients

CKD and CVD are often associated with co-existing pathologies that can begin, intensify and sustain each other [201,202]. CKD patients are at risk of developing CVD proportionally to the severity of kidney disease, decrease in glomerular filtration rate and increase in proteinuria [203]. As a consequence, CKD patients have an early onset and a higher risk of CVD, including stroke, coronary artery disease, peripheral vascular disease, and congestive heart failure, as compared with the general population [202]. Epidemiological data underline that patients with CKD die at an accelerated rate, which is 10-30 times higher than that of subjects without CKD [204,205]. Of note, in ESRD patients, CVD is the leading cause of morbidity and mortality with the ability of reducing the 5-year survival rate of...
patients on dialysis to only 40–50% [206,207]. Given that the traditional risk factors for CVD (including diabetes, dyslipidemia, hypertension, smoking, age and gender, as well as medical and family history) do not fully explain the accelerated development and progression of CVD in patients with CKD, it is current scientific opinion that other non-traditional risk factors must be involved. These non-traditional risk factors include hemodialysis, disorders of phospho-calcium metabolism, such as vitamin D deficiency, hyperparathyroidism, hyperphosphatemia [208,209], as well as the modification of the plasma levels of soluble proteins involved in vascular homeostasis, such as fibroblast growth factor-23 (FGF-23) and its co-receptor klotho, which are examples of recognized systems that contribute to CVD in course of CKD [210].

Also OPG, TRAIL, and TWEAK seem to be involved in CVD development and progression in patients with CKD. OPG has been recently identified as a potential risk marker of CVD in CKD patients [127]. Endothelial cells are a source of OPG when exposed to an inflammatory milieu, suggesting a role for OPG – and possibly the entire OPG/RANK/RANKL axis – in vascular injury and atherosclerosis. Increased OPG levels are associated with an increased CVD risk, although the increase in OPG production can be the attempt to protect the vessel walls from vascular damage/calcification [31]. Increased OPG levels have been reported in pre-dialysis patients and those undergoing dialysis, as well as in transplant recipients, where it may predict vascular calcification progression and patient survival. In this regard, Montanez-Barragan and colleagues have argued that, even in the context of uremia, the increase in OPG might be a protective response against kidney and vascular injury [137]. Consistent with this concept, recombinant OPG has been evaluated in phase I clinical trials, although additional studies are needed to confirm its potential [137]. Overall, so far, OPG levels appear a promising independent biomarker of CVD in patients with acute/chronic cardio-metabolic disease [31].

By contrast to OPG, circulating TRAIL levels are significantly decreased in patients with myocardial infarction and/or acute coronary syndrome [32,58] and they are inversely associated with an increased risk of CVD and cardiovascular mortality [58,60]. Similarly, an inverse association between TRAIL levels and mortality risk was observed in patients with advanced heart failure and CKD [32]. In particular, Kuzniecki and colleagues reported that CKD patient exhibited high OPG levels and OPG/TRAIL ratio and that OPG levels were associated with a higher risk of CVD and mortality among patients with renal failure [211]. The same Authors observed that OPG levels and OPG/TRAIL ratio predicted long-term mortality (all-cause and CVD mortality) and that they correlated with aortic pulse wave velocity and with N-terminal pro b-type natriuretic peptide, that are two independent predictors of CVD morbidity and mortality, supporting the role of OPG and OPG/TRAIL ratio as biomarkers of CV dysfunction and predictor of mortality in late stage of CKD. In the context of the NEFRONA study [212], our group has recently investigated the role of circulating TRAIL in predicting the progression of subclinical atheromatosis in CKD patients [213]. We reported that the baseline levels of soluble TRAIL were independently and inversely associated with novel plaque formation in 3–5-stage patients, establishing the proof of concept for the use of recombinant TRAIL to prevent/attenuate atheromatous progression in CKD patients not undergoing dialysis [213].

Recent studies have highlighted also the importance of TWEAK in the evaluation of the CV risk in kidney disease. It is known that TWEAK contributes to endothelial cell dysfunction. As a matter of fact, both TWEAK and Fn14 have been detected in human atherosclerotic plaques, suggesting that they can both play a role in the atherogenic process, probably also via reactive oxygen species (ROS) and mtROS modulation [214]. In the setting of kidney disease, it has been demonstrated that circulating TWEAK decreases across CKD stages and that this decrease is associated with an increased risk of CVD. For these reasons, TWEAK has been indicated as predictor of mortality in dialysis and non-dialysis CKD patients, as well as a biomarker of CVD outcomes [215]. Moreover, it has been observed that atherosclerotic burden and atheromatosis progression in CKD patients free of CVD are accompanied by low TWEAK, suggesting that TWEAK could serve as biomarker to predict CVD risk even before clinical manifestations [216,217].

Mineral and bone disorders are common in CKD patients and contribute to the burden of CV and bone complications in CKD. Vascular calcification of the arterial wall and cardiac valves is a hallmark of atherosclerosis. Bozic and colleagues analyzed the serum levels OPG, Osteopontin (OPN), and TWEAK as predictors of CVD outcomes in CKD patients (with or without CVD risk factors) without any history of CVD [218]. In detail, they observed that higher levels of OPG (or OPN) alongside lower levels of TWEAK were associated with significantly greater risk of non-fatal and fatal CV events [218]. Although other inflammatory markers are associated with plaque progression and instability in the general population [219,220], the combination of OPG, OPN, and TWEAK showed good discriminative power suggesting that marker combinations integrating different biological mechanisms might better stratify CVD risk in CKD [218].

In this scenario, CKD can be considered as an atypical disease, whose CVD outcomes depend on both traditional and new risk factors [210]. Although the underlying mechanisms linking CKD to accelerated CVD remain to be fully elucidated, the level fluctuations of OPG, TRAIL, and TWEAK influence the predictability of CVD outcomes in CKD.
Figure 3. Snapshot of key suggestions on the involvement of the TRAIL/OPG/TWEAK triad in kidney disease

The figure summarizes evidence and suggestions about the role of the TNF-superfamily triad in kidney disease in the light of potential clinical translations as disease biomarkers and/or therapeutic strategies.

Conclusion perspectives

Several lines of evidence suggest a specific involvement of TRAIL, OPG, and TWEAK in the pathogenesis of kidney disease, such that they are regarded as new potential biomarkers of kidney disease and its complications, as well as potential therapeutic targets (Figure 3). In particular, ongoing clinical studies on renal diseases have already included OPG and TWEAK as biomarkers to monitor disease-related progression and complications. Besides circulating OPG levels, also urinary OPG has been proposed as a biomarker of disease status and prognosis in this setting. Likewise, due to a large body of evidence that has highlighted the harmful role of TWEAK in kidney injury, the TWEAK–Fn14 system is emerging not only as a potential biomarker of renal disease but also as a promising therapeutic target, and strategies blocking TWEAK/Fn14, which appear to be safe and well tolerated, are under clinical investigation. By contrast, the significance of TRAIL in the setting of kidney disease remains to be fully elucidated.

Further studies are needed to clarify if OPG has a causative role in kidney disease, to understand the exact role of TRAIL in renal disease development/progression, and to optimize TWEAK-Fn14-targeting therapeutics. Overall, due to their belonging to the same family, a better comprehension of their changes and specific effects, alone and in combination, might facilitate the identification of new diagnostic/therapeutic strategies, in line with the concept that...
it is the integration of biological, molecular, and genetic data that will allow the identification and validation of new clinically-relevant biomarkers for kidney disease [221].

Competing Interests
The authors declare that there are no competing interests associated with the manuscript.

Abbreviations
AKI, acute kidney injury; CKD, chronic kidney disease; CNS, central nervous system; CV, cardiovascular; CVD, cardiovascular disease; DCRI, decoy receptor; DD, death domain; DR, death receptors; ERK, extracellular signal regulated kinase; ESRD, end-stage renal disease; FGF-23, fibroblast growth factor-23; Fn14, fibroblast growth factor-inducible molecule 14; MAPK, mitogen-activated protein kinase; MCNS, minimal-change nephrotic syndrome; NF-kB, nuclear factor kB; OPG, osteoprotegerin; OPN, osteopontin; PI3K, phosphatidylinositol-4,5-bisphosphate 3-kinase; RANK, receptor activator of nuclear factor kappa-β; RANKL, RANK ligand; ROS, reactive oxygen species; T1DM, type 1 diabetes mellitus; T2DM, type 2 diabetes mellitus; TNF, tumor necrosis factor; TRAF, TNF receptor-associated factors; TRAIL, TNF-related apoptosis inducing ligand; TWEAK, TNF-like weaker inducer of apoptosis; uTWEAK, urinary TWEAK.

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