Case Report

Hematochezia: An Uncommon Presentation of Colonic Tuberculosis

Fares Ayoub,1 Vikas Khullar,2 Harry Powers,1 Angela Pham,2 Shehla Islam,3 and Amitabh Suman2

1Department of Medicine, University of Florida, Gainesville, FL 32608, USA
2Department of Medicine, Division of Gastroenterology, University of Florida, Gainesville, FL 32608, USA
3Department of Medicine, Division of Infectious Disease, University of Florida, Gainesville, FL 32608, USA

Correspondence should be addressed to Fares Ayoub; fares.ayoub@medicine.ufl.edu

Received 16 February 2017; Accepted 28 March 2017; Published 3 April 2017

1. Introduction

After a resurgence in the incidence of tuberculosis (TB) infections in the United States between 1985 and 1992 due to the human immunodeficiency virus (HIV) epidemic, the incidence of TB had annually declined. However, as of 2015, TB incidence has leveled in the US and TB elimination (defined as <1 TB case per 1 million persons annually) remains elusive [1]. Gastrointestinal tuberculosis is a manifestation of extrapulmonary tuberculosis. In 2014, extrapulmonary TB constituted 20.57% of TB cases reported to the CDC and continues to be a missed diagnosis for providers not considering TB in their differential diagnoses [2].

Tuberculosis affecting the gastrointestinal tract was recognized as early as the fourth century BC in texts by Hippocrates [3]. While TB of the gastrointestinal tract is not as common as pulmonary TB, it is an important cause for TB related morbidity and mortality. The pathophysiology of this form of tuberculosis involves spread of mycobacteria to the gastrointestinal tract by number of means: hematogenous spread, swallowing of sputum contaminated with live M. tuberculosis bacilli, ingestion of contaminated food, or direct spread from adjacent organs [4]. The terminal ileum and the ileocecal valve are the most commonly affected parts, followed by the ascending colon which is usually affected through continuous involvement extending from the cecum. This predilection for the terminal ileum has been attributed to the presence of large amounts of lymphoid tissue and the longer contact duration of gastrointestinal contents with the lumen [5].

Classically, gastrointestinal TB may present with fever, weight loss, anorexia, abdominal pain, nausea, vomiting, or diarrhea. Hematochezia is a less common presentation [5]. Physical exam findings are nonspecific but may include abdominal tenderness, ascites, and hepatomegaly [4, 5]. Diagnosis is often delayed, as this form of tuberculosis is commonly misdiagnosed as Crohn’s disease or carcinoma.
Urinalysis was remarkable for pyuria with negative human immunodeficiency virus (HIV). A metabolic panel revealed hyponatremia of 110 mmol/L, a sodium level of 123 mmol/L, and an elevated anion gap metabolic acidosis with a pH of 7.3, a serum potassium of 6.2 mmol/L, acute renal failure with a creatinine of 8.0 mg/dL, and anemia with a hemoglobin of 12 g/dL and an MCV of 81.0 fL. Laboratory testing revealed a myriad of abnormalities. A complete blood count was significant for normocytic anemia with a hemoglobin of 12 g/dL and an MCV of 81.0 fL, as well as thrombocytosis with a platelet count of 552 × 10^9 /microliter. A metabolic panel revealed hyponatremia with a sodium level of 123 mmol/L, hyperkalemia with a potassium of 6.2 mmol/L, acute renal failure with a creatinine of 8.0 mg/dL, and an elevated anion gap metabolic acidosis with a pH of 7.3, a serum CO2 of 11 mmol/L, and an anion gap of 28 mmol/L. Testing for human immunodeficiency virus (HIV) was negative. Urinalysis was remarkable for pyuria with >180 WBC/high powered field and a negative urine nitrite. This was thought to indicate sterile pyuria, as conventional urine culture techniques were initially negative. A chest X-ray revealed extensive ill-defined bilateral parenchymal opacities in the perihilar, mid, and lower lung zones (Figure 1). The patient was admitted to the hospital for further management of acute renal failure and respiratory abnormalities. Blood and urine cultures were obtained. The patient was hydrated with intravenous normal saline and intravenous ceftriaxone therapy was initiated for a tentative diagnosis of community acquired pneumonia. Over the course of the following three days, the patient's hemoglobin decreased from 12 g/dL to 8 g/dL. A fecal immunohistochemical test for blood in stool was positive and the gastroenterology consult service recommended both an upper endoscopy and lower endoscopy.

Esophagogastroduodenoscopy was unremarkable. Colonoscopy was marred by inadequate bowel preparation but was significant for diverticulosis in the sigmoid and descending and transverse colon with purulent discharge associated with one diverticular opening. It also demonstrated an ulcer in the sigmoid colon which was biopsied (Figure 2). The presence of other ulcers was unclear due to the poor bowel preparation. These findings raised suspicion for acute diverticulitis and the patient's antimicrobial coverage was broadened to include ciprofloxacin and metronidazole. The following day the patient passed a large amount of blood per rectum, developed hypotension with a blood pressure of 79/49 mmHg, and was transferred to the medical intensive care unit. A repeat urgent colonoscopy revealed multiple ulcers at the anus and transverse and ascending colon as well as the cecum; the terminal ileum appeared normal (Figure 3).

A CT scan of the chest obtained for acute hypoxic respiratory failure showed extensive patchy consolidation and reticulonodular opacities throughout the lungs with a 1.3 cm cavitory lesion in the right apex as well as mediastinal and
hilar lymphadenopathy. These findings were suspicious for atypical infection (Figure 4). The patient underwent bronchoscopy with bronchoalveolar lavage and a lavage sample was AFB stain positive. Pathology of the sigmoid ulcer biopsy revealed focal active colitis with cryptitis, crypt abscesses, and mild stromal lymphoplasmacytic inflammation. AFB stain of the sigmoid ulcer biopsy was positive with findings consistent with *mycobacterium* (Figure 5). AFB stain of a urine sample obtained to explain sterile pyuria was also positive. The patient was placed under respiratory isolation and was started on appropriate antitubercular therapy.

Over the course of our patient's hospitalization, his clinical condition continued to decline. Multiorgan failure requiring vasopressor support later ensued and the patient ultimately passed away 29 days into admission.

3. Discussion

We present a case of diffuse systemic tuberculosis (TB) infection involving the colon, urinary tract, and lungs complicated by multiorgan failure and death. Unique to our patient was the lack of typical risk factors for TB infection. The patient had no history of travel to TB endemic areas and denied incarceration, homelessness, or exposure to infected individuals. It remains unclear where the patient may have contracted the infection; however, earlier exposure to infected individuals or family members that the patient does not recall is possible. Also unique in our patient's case is the presentation of colonic tuberculosis with hematochezia causing hemodynamic instability. Rectal bleeding is a rare presentation of colonic TB; only a handful of case reports have documented this manifestation [11].

Our patient's initial colonoscopy findings of a rectal ulcer, as well as diverticulosis showing signs of active infection, were highly suspicious for acute diverticulitis (Figure 2). However, initial colonoscopy was marred by poor bowel preparation and did not demonstrate the extent of colonic involvement with ulceration and there was low suspicion of active tuberculosis as a potential cause for the patient's drop in hemoglobin. Given our patient's advanced age as well as the presence of multiple diverticulae, angiodysplastic changes in colonic vessels were also part of our differential for lower gastrointestinal bleeding. However, massive hematochezia the following day prompted repeat colonoscopy which showed multiple new ulcers involving all segments of the colon, raising concern for an infectious process (Figure 3). No biopsies were obtained during the second colonoscopy due to active bleeding. Acid-fast stain of the sigmoid ulcer biopsy obtained during the first colonoscopy showed acid-fast organisms consistent with mycobacteria (Figure 5).
Tuberculosis remains a “great mimicker” and must continue to be part of the differential diagnosis in cases of colitis especially in individuals who have additional findings suspicious for infection on chest imaging. While staining for acid-fast bacilli is not routine practice in pathology laboratories, this should be considered in select patients who present with concerns for infection, especially both in the lungs and in the gastrointestinal tract. As described above however, only 15–20% of intestinal TB is associated with active pulmonary TB, thus providers must maintain a high index of suspicion to allow early diagnosis and management.

Our case also serves as a reminder for the importance of postexposure testing and prophylaxis in healthcare workers. Nosocomial transmission of TB in the healthcare setting is well described; cases of multidrug resistant TB transmitted to providers and patients in hospitals have also been reported [12, 13]. In hospitals that receive more than 200 TB admissions a year, incidence of hospital worker infection has been reported to be as high as 10% a year [14].

Hospital-based infection control programs are essential in the control of such transmission. Airborne infection isolation rooms (previously termed negative pressure isolation rooms) should be utilized for patients with suspected or confirmed tuberculosis. N95 masks that filter particles ≥1 micrometer in diameter with at least 95 percent efficiency should be utilized by healthcare staff and offered to visitors of patients with suspected or confirmed tuberculosis. Nonessential invasive diagnostic procedures ought to be postponed unless absolutely necessary [15].

Providers exposed to patients with active pulmonary tuberculosis should be assessed for symptoms suggestive of active TB infection (hemoptysis, weight loss, and fever)
and, if present, should undergo chest X-ray with sputum acid-fast stain and culture. Providers without symptoms of active infection are to undergo tuberculin skin testing or an interferon-gamma release assay at baseline and again 8 to 12 weeks postexposure [12].

In our case, all providers who may have been exposed to the patient underwent interferon-γ release assay testing (QuantiFERON, Cellestis Ltd., Australia) at baseline and 8 weeks postexposure. The patient’s family members were also contacted and offered screening and appropriate treatment for TB.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

References

[1] J. L. Salinas, G. Mindra, M. B. Haddad, R. Pratt, S. F. Price, and A. J. Langer, “Leveling of tuberculosis incidence—United States, 2013–2015,” Morbidity and Mortality Weekly Report, vol. 65, no. 11, pp. 273–278, 2016.

[2] Department of Health and Human Services (US DHHS), Centers for Disease Control and Prevention (CDC), and Division of TB Elimination, “Online Tuberculosis Information System (OTIS), National Tuberculosis Surveillance System, United States, 1993–2014,” CDC WONDER Online Database, January 2016. Data for all years updated by June 5, 2015, http://wonder.cdc.gov/tb-v2014.html.

[3] T. A. Sheer and W. J. Coyle, “Gastrointestinal tuberculosis,” Current Gastroenterology Reports, vol. 5, no. 4, pp. 273–278, 2003.

[4] K. D. Horvath and R. L. Whelan, “Intestinal tuberculosis: return of an old disease,” The American Journal of Gastroenterology, vol. 93, no. 5, pp. 692–696, 1998.

[5] S. Mukewar, S. Mukewar, R. Ravi, A. Prasad, and K. S. Dua, “Colon tuberculosis: endoscopic features and prospective endoscopic follow-up after anti-tuberculosis treatment,” Clinical and Translational Gastroenterology, vol. 3, article e24, 2012.

[6] Y. Lee, S. Yang, J. Byeon et al., “Analysis of colonoscopic findings in the differential diagnosis between intestinal tuberculosis and crohn’s disease,” Endoscopy, vol. 38, no. 6, pp. 592–597, 2006.

[7] B. Nagi, R. Kochhar, D. K. Bhasin, and K. Singh, “Colorectal tuberculosis,” European Radiology, vol. 13, no. 8, pp. 1907–1912, 2003.

[8] W.-K. Lee, F. Van Tonder, C. J. Tartaglia et al., “CT appearances of abdominal tuberculosis,” Clinical Radiology, vol. 67, no. 6, pp. 596–604, 2012.

[9] K. M. Kim, A. Lee, K. Y. Choi, K. Y. Lee, and J. J. Kwak, “Intestinal tuberculosis: clinicopathologic analysis and diagnosis by endoscopic biopsy,” The American Journal of Gastroenterology, vol. 93, no. 4, pp. 606–609, 1998.

[10] V. K. Kapoor, “Abdominal tuberculosis,” Postgraduate Medical Journal, vol. 74, no. 874, pp. 459–467, 1998.

[11] M. Kela, V. B. Agrawal, R. Sharma, R. Agarwal, and A. Agarwal, “Ileal tuberculosis presenting as a case of massive rectal bleeding,” Clinical and Experimental Gastroenterology, vol. 2, article 129, 2009.

[12] M. S. Bader and D. S. McKinsey, “Postexposure prophylaxis for common infectious diseases,” American Family Physician, vol. 88, no. 1, pp. 25–32, 2013.

[13] Centers for Disease Control (CDC), “Nosocomial transmission of multidrug-resistant tuberculosis to health-care workers and HIV-infected patients in an urban hospital—Florida,” Morbidity and Mortality Weekly Report, vol. 39, no. 40, pp. 718–722, 1990.

[14] D. Menzies, A. Fanning, L. Yuan, and M. Fitzgerald, “Tuberculosis among health care workers,” New England Journal of Medicine, vol. 332, no. 2, pp. 92–98, 1995.

[15] Centers for Disease Control and Prevention, “Guidelines for preventing the transmission of Mycobacterium tuberculosis in health-care settings, 2005,” MMWR 54.RR-17, 2–107, 2005.