Numerical Chromosomal Abnormalities in Patients with Acute Lymphoblastic and Myeloid Leukemia in Iran

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Abstract The majority of cases of ALL demonstrate an abnormal karyotype, either in chromosome number or structural changes. Abnormal chromosome number in childhood acute lymphoblastic leukemia defines distinct biological subgroups with a different response to treatment. The tubes are cultured with three different protocols to save time if one protocol failed. Cultures are then harvested, and cells are fixed and chromosome spreads are prepared. Of 25 patient studied, one patient had psudodiploid karyotype, three patients had tetra-ploid karyotype, four patients had low hypo-diploid karyotype, four patients had high hyper-diploid karyotype, five patients had low hyper-diploid karyotype and eight patients had normal karyotype. Some other factors like, Age, Sex, Consanguinity, Hemoglobin, WBC count, and Type of the leukemia cell also have been evaluated. We found excess number of patients having hypodiploid karyotype but still response to treatment protocols were satisfactory. By comparison between 4 different cultures methods, we find direct method to be more efficient for ploidy analysis.

Keywords ALL, AML, Karyotype, Ploidy, Leukemia

1. Introduction

Acute lymphoblastic leukemia (ALL) is characterized by clonal proliferation, accumulation, and tissue infiltration of neoplastic cells. They are mainly regarded as childhood diseases, with an early incidence peak at two to five years of age, where they represent about 80% of the childhood leukemia in the United States, and occur with an incidence of 30 cases per one million population per year[1].

A basic principle of the WHO classification is that the diagnosis of myeloid and lymphoid malignancies should be based on the integration of the information derived from genetic, immunophenotypic, biological, and clinical features in order to better define specific disease entities. In fact, genetic findings may predict the prognosis and biologic properties of the leukemia more consistently than does morphology[2].

Chromosomal abnormalities in childhood ALL had important significance related to prognosis, diagnosis and management. Chromosomal classification in ALL is based on the number of chromosomes in abnormal cells (ploidy pattern) as well as the structural changes.

Abnormal chromosome numbers in childhood acute lymphoblastic leukemia, defines distinct biological subgroups with a different response to treatment[3,4]. The largest subgroup within the non-T cells ALL (approximately 25%) is the hyper-diploid group, which is associated with favorable characteristics and outcome[3,4,5]. In contrast the hypo-diploid and near-haploid ALL is much rarer (1%) and has shown a very poor outcome with the few intensive protocols reported[6,7]. Near haploid has been reported in around 41 children[5,8,9] and in six additional children within a clinical study[9].

The majority of cases of ALL demonstrate an abnormal karyotype either in chromosome number or as structural changes such as trans-locations, inversions, deletions, etc. These changes were detected in only half of ALL patients in the first banding studies[10]. The scantiness of information gained from chromosomal findings in ALL has been in large part, due to technical difficulties. The emerging theory for the role of constitutional trisomy 21 in leukaemia predisposition is that genes on this chromosome contribute to the expansion of haematopoetic compartments during early development that result in an increased pool of potential tumour precursor cells[11].

Chromosome studies in ALL exhibit poor morphology. Chromosomes tend to spread poorly, and appear blurred and fuzzy with indistinct margins, making banding studies challenging or even impossible[12,13] Improvements in spreading and banding techniques have resulted in higher rates of detection, and most studies now report chromosomal changes in 60% to 85% of ALL cases [14,15].

We have conducted this study to evaluate numerical
abnormalities in these patients for better management of patients admitted to this hospital by using different cell culture and chemotherapy treatment protocols.

2. Material and Methods

This work was conducted on 25 cases (five cases with AML and 19 cases with ALL) referred to our laboratory for karyotype and ploidy determination of chromosomes after getting a written consent from their parents. Only new diagnosed cases (from December 2007 to July 2008) were included in this survey and old cases which have been already started chemotherapy treatment were excluded.

0.5-1 ml of bone marrow aspirate were collected in centrifuge tube containing five ml RPMI complete with 1% preservative-free heparin. In few cases where the bone marrow samples were unavailable or failed to grow in culture, blood samples were used. The optimum cell density for a bone marrow culture is 10⁶ cell/ml. All samples were optimized for cell density, using hemocytometer (slide counter chamber).

In most cases, the preferred culture method was overnight colcemid (ONC), followed by flurodeoxy uridine (FudR) and 24 hour protocols. In AML samples the preferred technique was FudR, followed by 24 hours cultures and then ONC. For each patient we set up at least three culture tubes including one direct cultures method[12] and two out of the three fore-mentioned methods. Quality of spreads in these methods were compared. Good and analysable spreads could obtain with direct method for ploidy determination in most cases, but for structural chromosomal analysis, flurodeoxy uridine (FudR) method proved to be satisfactory.

3. Results

3.1. Number of Patient, Age, Sex, Ethnic Groups, Consanguinity, Hemoglobin Rate and WBC Count Factors

Bone marrow (19 patient) or blood (six patients) samples from 25 patients, suffering from leukemia referred from Shafa hospital have been analyzed. Among 25 patients participated, 19 ALL (including two cases with T-ALL and seven cases with B-ALL), five AML and one unidentified case was present.

Out of 25 patients, 16 males (64%) and nine females (36%) are recorded. The age of patients ranges from 1.5 to 12 with median age of 5.82 years.

Of 25 parents participated, eight couples (32%) have not reported the type of their marriage, (consanguinity) and out of remaining 17 (68%) couple, 11 couples (44%) had consanguineous marriage (10 firsts and one second) and six couples (24%) had non-consanguineous marriage. Table 1

| Patient No. | Ploidy groups | Leukemia subtype | HB/g/dl | WBC/μl | Treatment result | Type of cell involved | Primary diagnosis | Sex | Consanguinity | Ethnic | Age |
|-------------|---------------|------------------|---------|--------|------------------|----------------------|------------------|-----|--------------|--------|-----|
| 1           | High hyper diploid | AML-M5           | 5.5     | 1900   | Remission       | Myeloid              | Aplastic anemia   | Male | First cousin | Arab   | 1.5 |
| 2           | Low hyper diploid | B-ALL-L1         | 7.7     | 7500   | Remission       | B cell               | anemia            | Female| Non consan. | lur    | 6   |
| 3           | Low Hypo diploid | AML-M4           | 7.9     | 6300   | No Remission    | Myeloid              | AML               | Female| First cousin | lur    | 5.5 |
| 4           | Near tetraploid | 46, Ring Ch.     | 7.7     | 3400   | Remission       | Pansitopy            | Male              | Unkno wn      | lur | 6.5 |
| 5           | Low Hyper diploid | BALL-L1          | 10.7    | 10400  | Remission       | B cell               | Pansitopy         | Male | First cousin | Arab   | 3.5 |
| 6           | Normal | T-ALL-L2         | 6.9     | 20000  | Remission       | T cell               | Burkit lymphoma    | Male | Unkno wn | lur | 11 |
| 7           | Normal | B-ALL-L2         | 13.5    | 4200   | Remission       | B cell               | Pansitopy         | Female| First cousin | fars   | 3   |
| 8           | Normal | B-ALL-L1         | 5       | 5700   | Remission       | B cell               | ALL               | Female| Non consa. | Arab   | 3   |
| 9           | Low Hyper diploid (Down) | B-ALL-L2 | 2500   | 5700   | Remission       | B cell               | ALL               | Male | Non consa. | lur | 5   |
| 10          | High hyper diploid | AML-M1           | 6       | 90000  | Remission       | Myeloid              | AML               | Female| Unkno wn | Arab   | 11 |
| 11          | Pseudo diploid | B-ALL-L2         | 5.1     | 7400   | Remission       | B cell               | Anemia            | Male | First cousin | Arab   | 9   |
Except six cases (Nos. 5, 7, 12, 13, 18, 24), the hemoglobin rate of the remaining patients ranges from 2.6 to 7.9, and is considered as anemic.

Of 25 patients, 17 (68%) had WBC of less than 10000 c/μl, five patients had 10000-50000 c/μl and three patients had extremely high WBC count, Table 1.

Out of four Major ethnic groups living in this province, including Persians (Farsis), Iranian Arabs (Arab) Bakhtiari Lurs (lurs), and Behbahanies, 14 Arab (56%), 9 Lurs (36%) and two Farsis (8%) are recorded.

3.2. Response to Treatment

Of these 25 leukemic cases, 23 (92%), achieved complete remission but with different resistant. Two patients (8%) did not respond to treatment and survival, one of them with hypo-diploid karyotype had Philadelphia chromosome, and other one had complex karyotype of hypo-diploid and near-haploid. Both of them were among AML patients with M3 and M4 subgroup, Table 3.

Regarding hematological parameters, cases with lower Hb (< 8 gm/dl) and high WBC (>50000/mm3) showed a better remission rate. Cases with age < 5 years, male sex and couples with positive consanguinity also showed a better remission rate, although not significant.

3.3. Clonal Abnormalities

Of 25 patients studied, 17 (68%) cases with clonal ploidy abnormalities and eight cases (32%) of normal karyotype were recorded. Among 17 cases with more than one abnormal karyotype, 10 cases (40%) were hyper-diploid, [six low (47-50 chromosomes) and four highs (> 50 chromosomes) hyper-diploid], three cases (12%) were hypo-diploid (30-44 chromosomes), Three cases (12%) were near tetra-ploid, (> 80 chromosomes) Figures 1, 2 and 3. and one patient was pseudo-diploid, Table 2.

Although some of these patients are hypo-diploid (three cases), or pseudo-diploid (one case), most of them (92%) had good remission with treatment protocols (U.K.C.C.S.G) for ALL, and (B.F.M) AML patients. Only two patients (8%) had no responses to treatment, both of which were among AML leukemic cases.

Of 17 patients with abnormal karyotype, eleven B-cells ALL and five AML with M1 to M5 subgroups were recorded. Classification of one of the patient could not be determined. Eight patients had complex karyotype with more than two clonal chromosomal abnormalities. Details are available on request.
Table 2. Ploidy groups and number of patients in each group

| Ploidy group     | Chromosome number | Number of Patients present in each group | Patients number | Percentage |
|------------------|-------------------|-----------------------------------------|-----------------|------------|
| Normal           | 46                | 8                                       | 6-7-8-13-14-20-22-24 | 32%        |
| Near haploid     | 23-29             | 8                                       | 2-5-9-16-19-21   | 24%        |
| Low hypodiploid  | 36-44             | 11                                      | 3-15-18         | 12%        |
| Pseudo diploid   | 46                | 6                                       | 1-10-12-25      | 16%        |
| Low hyper diploid| 47-50             | 3                                       | 2-5-6-7-8-9-14-15-16-17-20-21-24-25 | 24%        |
| High hyper diploid| 50               | 6                                       | 3-15-18         | 12%        |
| Near triploid    | 66-80             | 3                                       | 4-17-23         | 12%        |
| Near tetraploid  | 80                | 25                                      | 25              | 100%       |
| Total            |                   | 25                                      |                 |            |

Table 3. Type of Samples in each leukemia subgroup

| Sample and leukemia type | Total No. of patients |
|-------------------------|-----------------------|
|                         | AML                  | ALL                  |
| Peripheral blood(PN)    |                      |                      |
| AML                     | 23                   | 1-3-10-18            |
| ALL                     | 4 & 12               | 2-5-6-7-8-9-14-15-16-17-20-21-24-25 | 25 |
| Bone marrow(PN)         |                      |                      |
| AML                     | 2                    | 4                    |
| ALL                     | 4                    | 14                   |
| Total                   | 25                   | 25                   |

PN=Patient’s number

4. Discussion

Most of the leukemia patients have cytogenetical or molecular abnormality and it is shown that there are some relations between chromosomal abnormalities and morphological and immunological characteristics of the cancer cells in these patients. More important are karyotype changes which show important prognosis without any relations to other variables like age, sex and primary leukocyte count, and so permits to differentiate between high risk and low risk patients at the time of diagnosis for proper treatment.

Cytogenetic analysis of 25 cases in our study shows 8 patients (32%) to have normal diploid karyotype, similar to many reports. Pseudo-diploid on the other hand were found in only one case (4%) which is much lower than the earlier reports which suggested that pseudo-diploid comprised the largest cytogenetic group (41.5%) in childhood ALL[16]. This could be due to misclassification of pseudo-diploid karyotype by failure of detection of some submicroscopic genetic abnormalities at the conventional cytogenetic analysis.

Low hypo-diploid karyotype (30-44 chromosomes) were present in 12% of the our cases, which are higher than those reported by others who considered hypo-diploidy to be a relatively uncommon (>9%) finding in ALL[17,18]. Many studies reported low percent of patients in hypo-diploid group, but some studies[19-20], including our study (12%) found much higher percent of the patient in this group with good remission. In studies in which only cytogenetic investigations are carried out, some hypo-diploid cells may not diagnosed correctly due to presence of many hyper-diploid cells and only after relapse of the disease and repeat investigation of these patients with FISH, or other advanced molecular methods, the true karyotype can be detected.
Hyper-diploid (low + high hyper-diploid) represented a majority of ALL cases in our study and many other authors have also reported same higher frequency[21,22]. The remission rates in our study cases were almost same among hypo-diploid, pseudo-diploid and hyper-diploid groups, but the normal diploid cases had better remission rate.

Diploid group was reported to have the best remission by some authors[19] although others considered it to be of an intermediate remission rate[23,24].

Response to treatment rates varied among chromosome ploidy groups. The highest response rates were seen in patients with normal karyotypes and a modal number >50 chromosomes. In our study, after normal karyotype cases, we also found the best remission rate in this group. Hyper-diploidy was found by some authors to have a good prognosis[25,26] unless associated with structural aberrations making it less favorable[19,27,28].

In our study, we had two resistant cases, one mixed numerical aberration (near haploid + low hypo-diploid) and the other one with low Hypo diploid, both of which were among AML subtype.

Near-haploid (< 30 chromosomes) ALL, were not present in our case, except the one with mixed numerical aberration (near haploid + low hyper-diploid) between two resistant patients. Near-haploid is a rare and unique subgroup associated with a very poor outcome. It may be underestimated being masked by a coexisting hyper-diploid line.

In our study although any chromosome could be involved in tetrasomy, most of chromosomes especially 5,9,10, 12,18, 20,22 and X were duplicated more frequently.

Of four Majors Ethnic groups in Khoezestan province including Persians, Iranian Arabs, Bakhtiari Lurs, and Behbahantis, 14 Arab (56%), 9 Lurs (36%) and two Farsi (8%) are recorded. Prevalence of Arabs were significant in these four major ethnic groups, although it may be unreliable due to small sample size, Table 1. We could not find any data showing prevalence of leukemia in these Ethnic groups in Iran.

The most common FAB type among our cases (48%) were L2 subtype with better remission compared to other immunophenotypes[29]. Some studies reported a poor prognosis associated with L2 and other studies reported that significance of L2 remained controversial[30].

Although full analyzing of karyotype spreads for determination of structural, numerical and submicroscopic genetic abnormalities at the conventional cytogenetic analysis and its easier detection by the molecular and FISH technique is mandatory, but determination of only numerical chromosomal changes (ploidy pattern) for typing of patients into different ploidy groups can also be useful as a primary step. Therefore we decided to conduct this study because of increasing number of ALL patients and need for immediate report for ploidy determination.

We compare three standard methods including, overnight colcemid (ONC), fluoroxy uridine (FudR) and 24 hour protocols, with a modified direct culture method (12), and found the modified direct culture method to be easy and satisfactory for ploidy pattern determination in most (68%) cases.

5. Conclusions

We found excess number of patients having hypo-diploid karyotype but still response to treatment protocols were satisfactory. Variation in number of hypo-diploidy in different studies could be due to difference in ethnic group. Out of four different cultures methods used, we found the direct method to be satisfactory for ploidy analysis.

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REFERENCES

[1] Young JL Jr, Ries LG, Silverberg E, Horm JW, Miller RW. Cancer incidence, survival and mortality for children younger than age 15 years. Cancer 1986;58:589
[2] Vardiman JW, Harris NL, Brunning RD. The World Health Organization (WHO) classification of the myeloid neoplasms. Blood 2002;100: 2292–302
[3] Pui C-H, Evans WE. Acute lymphoblastic leukemia. N Engl J Med 1998;339:605–15
[4] Raimondi SC, Current status of cytogenetic research in childhood acute lymphoblastic leukemia. Blood 1993;81:2237–51
[5] Ito C, Kamagai M-A, Manabe A, Coustan-Smith E, Hyperdiploid acute lymphoblastic leukemia with 51 to 65 chromosomes: a distinct biological entity with a marked propensity to undergo apoptosis. Blood 1999;93:315–20
[6] Prieto, F; Badia, L; Mayans, J; Gomis, F; Marty, M L. Hipodiploidia de 26 cromosomas en leuemia linfoblástica aguda. Sangre 1978;23: 484–8
[7] Heerema NA, Nachman JB, Sather HN, Sensel MG, Lee MK, Hutchinson R, et al. Hypodiploidy with less than 45 chromosomes confers adverse risk in childhood acute lymphoblastic leukemia: a report from the Children’s Cancer Group. Blood 1999;94:4036–46
[8] Ritterbach J, Hiddemann W, Beck JD, Schrappe M, Janka-Schaub G, Ludwig WD, et al. Detection of
hyperdiploid karyotypes (50 chromosomes) in childhood acute lymphoblastic leukemia (ALL) using fluorescence in situ hybridization (FISH). Leukemia 1998;12:427–33

[9] Chessel JM, Swansbury GJ, Reeves B, Bailey CC, Richards SM. Cytogenetics and prognosis in childhood lymphoblastic leukemia: results of MRC UKALL X. Br J Haematol 1997;99:93–100

[10] Mitelman F: Cytogenetic methods and terminology, in Heim S, Mitelman F (eds): Cancer Cytogenetics New York, NY, Wiley-Liss, 1995, p 20

[11] Lo KC, Chalker J, Strehl S, Neat M, Smith O, Dastugue N, et al. Array comparative genome hybridization analysis of acute lymphoblastic leukaemia and acute megakaryoblastic leukaemia in patients with Down syndrome. Blackwell Publishing Ltd, British Journal of Haematology, 2008;142:934–945

[12] Williams DL, Harris A, Williams KJ, Brosius MJ, Lemonds W. A direct bone marrow chromosome technique for acute lymphoblastic leukemia. Cancer Genet Cytogenet, 1984;13:239

[13] Mitelman F: The Third International Workshop on Chromosomes in Leukemia. Cancer Genet Cytogenet, 1981;4:96

[14] Secker-Walker LM, Prentice HG, Durrant J, Richards S, Hall E, Harrison G. Cytogenetics adds independent prognostic information in adults with acute lymphoblastic leukaemia on MRC trial UKALL XA. Br J Haematol, 1997;96:601

[15] Groupe Français de Cytogénétique Hématologique: Cytogenetic abnormalities in adult acute lymphoblastic leukemia: Correlations with hematologic findings and outcome. A collaborative study of the Groupe Français de Cytogénétique Hématologique. Blood, 1996;87:3135

[16] Pui CH, Grist WM, Look AT. Biology and clinical significance of cytogenetic abnormalities in childhood acute lymphoblastic leukemia. Blood 1990;76:1449–1463.

[17] Kobayashi H, Maseki N, Homma C. Clinical significance of chromosome abnormalities in childhood acute lymphoblastic leukemia in Japan. Leukemia 1994;8(11):1930–1944.

[18] Bloomfield CD, Lindquist LL, Arthur D, McKenna R W, LeBien T W, Peterson B A, et al. Clinical significance of chromosomal abnormalities in acute lymphoblastic leukemia. Cancer Res 1981;41:4838.

[19] Hafez M, Mansour AK, Mahmoud LA, et al. Clinical significance of chromosome abnormalities in children with acute lymphoblastic leukemia. Egypt J Haematol 1999;24.

[20] Settin A, Alhaggar M, Al dosoky T, Al baz R, Abdelrazik N, Fouda M, et al. Prognostic cytogenetic markers in childhood acute lymphoblastic leukemia: Cases from Mansoura Egypt. Hematology, April 2007;12(2):103–111

[21] Fletcher JA, Kimball VM, Lynch E, Donnelly M, Pavelka K, Gelber RD, et al. Prognosis implications of cytogenetic studies in an intensively treated group of children with acute lymphoblastic leukemia. Blood 1989;74:2130.

[22] Heerema NA, Nachman JB, Sather HN, Sensel MG, Lee MK, Hutchinson R, et al. Hypodiploidy with less than 45 chromosomes confers adverse risk in childhood acute lymphoblastic leukemia: A report from the children’s cancer group. Blood 1999;94(12):4036–4045.

[23] Bloomfield CD, Secker-Walker LM, Goldman AI, Van Der Bergh E, de la Chapelle A, Ruutu T. Six year follow up of the clinical significance of karyotype in acute lymphoblastic leukemia. Cancer Genet Cytogenet 1989;40(2):171–185.

[24] Secker-Walker LM, Goldman JM. Cytogenetics and leukemogenesis. In: Hoffbrand AV, Lewis SM, editors. Postgraduate hematology. 3rd ed. London: Heinemann Professional Publishing Ltd. 1989. p 359.

[25] Ito C, Kumagi M, Manabe A, Coustan-Smith E, Raimondi SC, Behm FG, et al. Hyperdiploid acute lymphoblastic leukemia with 51 to 65 chromosomes: A distinct biological entity with a marked propensity to undergo apoptosis. Blood 1999;93(1):315–20.

[26] Mehdirpour P, Mirfakhraie R, Jahani M, Mehdirpour AR. Karyotypic evolution: Cytogenetics follow-up study in childhood acute lymphoblastic leukemia. Asian Pac J Cancer Prev 2003;4(4):358–68.

[27] Nordgren A, Nordenskjöld M, Söderhäll S, Porwit-MacDonald A, Blenno E. A case of acute lymphoblastic leukemia, near-triploidy, and poor outcome. Cancer Genet Cytogenet 1997;99(2):93–6.

[28] Zemanova Z, Michalova K, Sindelarova L, Smisek P, Brezinova J, Ransdorfova S, et al. Prognostic value of structural chromosomal rearrangements and small cell clones with high hyperdiploidy in children with acute lymphoblastic leukemia. Leuk Res 2005;29(3):273–81.

[29] Lilleyman JS, Hann IM, Stevens RF, Eden OB, Richards SM. French American British (FAB) morphological classification of childhood lymphoblastic leukemia and its clinical importance. J Clin Pathol 1986;39(9):998–1002.

[30] van Eys J, Pullen J, Head D, Boyett J, Crist W, Falletta J, et al. The French American British (FAB) classification of leukemia. The paediatric oncology group experience with lymphoeytic leukemia. Cancer 1986;57:1046–51.