Genomewide Association Study Identifies Novel Genetic Loci That Modify Antiplatelet Effects and Pharmacokinetics of Clopidogrel

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Genetic variants in the pharmacokinetic (PK) mechanism are the main underlying factors affecting the antiplatelet response to clopidogrel. Using a genomewide association study (GWAS) to identify new genetic loci that modify antiplatelet effects in Chinese patients with coronary heart disease, we identified novel variants in two transporter genes (SLC14A2 rs12456693, ATP-binding cassette [ABC]A1 rs2487032) and in N6AMT1 (rs2254638) associated with P2Y12 reaction unit (PRU) and plasma active metabolite (H4) concentration. These new variants dramatically improved the predictability of PRU variability to 37.7%. The associations between these loci and PK parameters of clopidogrel and H4 were observed in additional patients, and its function on the activation of clopidogrel was validated in liver S9 fractions (P < 0.05). Rs2254638 was further identified to exert a marginal risk effect for major adverse cardiac events in an independent cohort. In conclusion, new genetic variants were systematically identified as risk factors for the reduced efficacy of clopidogrel treatment.

Study Highlights

WHAT IS THE CURRENT KNOWLEDGE ON THE TOPIC?
Clopidogrel is a first-line antiplatelet therapy for CHD, but it is associated with substantial variability in PK and pharmacodynamic response. To date, gene variants explain only a small proportion of the variability.

WHAT QUESTION DID THIS STUDY ADDRESS?
The study aimed to identify new genetic loci modifying antiplatelet response to clopidogrel in Chinese patients with CHD by a systematic analysis combining antiplatelet effects and PK.

WHAT THIS STUDY ADDS TO OUR KNOWLEDGE
We identified novel variants in two transporter genes (rs12456693 in SLC14A2 and rs2487032 in ABCA1) and in N6AMT1 (rs2254638) associated with clopidogrel-treated PRU and plasma H4 concentration. The new variants, together with CYP2C19*2 and clinical factors, dramatically improved the predictability of PRU variability to 37.7% compared with the published value of ~20%. Meanwhile, N6AMT1 rs2254638 was identified to exert a marginal risk effect for MACE in an independent cohort.

HOW THIS MIGHT CHANGE CLINICAL PHARMACOLOGY OR TRANSLATIONAL SCIENCE
Our study findings enhanced the understanding of the absorption and metabolic mechanisms that influence antiplatelet response to clopidogrel treatment.

Clopidogrel is a first-line antiplatelet therapy for coronary heart disease (CHD) and an extensively prescribed drug worldwide.1,2 However, clopidogrel therapy is associated with highly variable pharmacodynamic response, and individuals with high residual on-treatment platelet reactivity have a great risk of death, myocardial infarction, and stent thrombosis.3–8 Reduced efficacy of clopidogrel may be caused not only by nongenetic factors, such as age and use of interacting drugs, but also by genetic polymorphisms, particularly those in transporters and enzymes participating in clopidogrel absorption and metabolic transformation. Several studies have been conducted using a candidate gene approach to identify genetic variants associated with antiplatelet and pharmacokinetic (PK) response to clopidogrel.9–12 However, such candidate gene studies have shown inconsistent results, except for those of CYP2C19 variants. A recent genomewide association study (GWAS) identified an association with CYP2C19*2 and diminished platelet response to clopidogrel treatment and poor cardiovascular outcomes,12 and this result was confirmed by other studies.11,13,14

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Gene variants that have been investigated to date explain only 12% of the variability in the clopidogrel response and 20.6% when combined with clinical factors. The substantial variability in the response to clopidogrel remains unexplained. Published GWAS may have missed single-nucleotide polymorphisms (SNPs) with small or moderately sized genetic effects on the P2Y12 reaction unit (PRU) variations under clopidogrel treatment, because of the stringent cutoff imposed by Bonferroni correction. Furthermore, a vast racial difference exists in the distribution of genetic variants. Studies focusing on white subjects may have also overlooked significant associations with Chinese subjects.

Considering that genetic variants in PK mechanisms are the main underlying factors that modify the antiplatelet effects of clopidogrel, joint analysis of genetic variants modifying pharmacodynamic and PK response to clopidogrel should be effective for identifying the biochemical mechanism. Therefore, additional GWAS using combination analysis of antiplatelet effects and PKs are necessary to identify new potential genetic variants that affect the antiplatelet response to clopidogrel, particularly in non-European populations.

In this study, we conducted a genome-wide investigation on the effects of genetic variants on the clopidogrel-treated antiplatelet response and H4 (an active metabolite of clopidogrel) exposure. For SNPs associated with platelet function in response to clopidogrel and H4 exposure, we then evaluated their relationship with PK parameters of clopidogrel and H4 in additional patients with CHD, as well as investigated the function of these SNPs on clopidogrel activation in human liver tissues. Finally, the SNPs suggestively associated with antiplatelet effects and H4 exposures were further determined by their link to major adverse cardiac events (MACEs) in patients with CHD after percutaneous coronary intervention.

**RESULTS**

**Patient characteristics and their effects on the antiplatelet response to clopidogrel and H4 exposure**

A schematic of the design of the GWAS is illustrated in Figure 1. In stage I, 3 of 120 patients were excluded for failure of the VerifyNow test, whereas 2 patients were excluded for deviation from the population, as shown by principal component analysis (PCA; Supplementary Figure S1 online). Thus, 115 patients passed quality control and were included in the GWAS analysis. Baseline clinical characteristics and their effects on the H4 concentration and PRU of 115 patients are shown in Supplementary Table S1 online. In stage IIa, baseline clinical characteristics and their effects on PKs of clopidogrel and H4 in 31 patients are shown in Supplementary Table S2 online.

Patient characteristics and their effects on antiplatelet and PK response to clopidogrel in stages I and IIa can be found in the Supplementary Data online.

**Genomewide association study of the antiplatelet response to clopidogrel and H4 exposure**

To identify SNPs associated with antiplatelet response to clopidogrel treatment, we performed a GWAS of PRU in 4 h in 115 patients with CHD. The antiplatelet effects of clopidogrel at 4 h widely varied (Supplementary Figure S2a online). Although no genomewide significant association signals were detected in single marker-based GWAS of PRU or PRU >208 with a Bonferroni-adjusted \( P \) value \( (P < 7.11 \times 10^{-8}) \), a total of 125 loci in 25 genes showed suggestive evidence of association with PRU or PRU >208 \( (P < 1 \times 10^{-4}) \) excluding CYP2C19*3 (rs4986893, \( P_{PRU >208} = 0.5003, P_{PRU >208} = 0.3513 \)).

The plasma concentration of clopidogrel, H4, and H3 at 2 h widely varied (Supplementary Figure S2 online). Among 125

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**Figure 1** Schematic of the design of the genomewide association study (GWAS). CHD, coronary heart disease; PK, pharmacokinetic; PRU, P2Y12 reaction unit; SNP, single-nucleotide polymorphism.
SNPs associated with PRU or PRU > 208 \((P < 1 \times 10^{-4})\), 27 SNPs were associated with the H4 concentration at 2 h at the significance level of \(P < 0.01\) (Supplementary Table S3 online), among which 23 SNPs were within the HELLS-CYP2C18-CYP2C19 cluster (Figure 2a). These SNPs were in strong linkage disequilibrium (LD) with one another with pairwise \(r^2 = 0.49–1.0\), including CYP2C19*2 (rs4244285, \(P = 1.66 \times 10^{-5}\)).

Univariate linear regression analysis revealed that CYP2C19*2 was associated with increased PRU, which explained 11% PRU variation. Others included rs12456693 in SLC14A2 \(\left( P = 8.52 \times 10^{-3} \right)\), rs2254638 in N6AMT1 \(\left( P = 6.29 \times 10^{-3} \right)\), rs471806 on chromosome 9 \(\left( P = 2.40 \times 10^{-3} \right)\), and rs7292279 on chromosome 22 \(\left( P = 9.08 \times 10^{-3} \right)\).

**Association of single-nucleotide polymorphisms in absorption, distribution, metabolism, and excretion genes with P2Y12 reaction unit and H4 concentration**

A total of 399 loci in 95 candidate absorption, distribution, metabolism, and excretion (ADME) genes were significantly associated with PRU or PRU > 208 \((P < 0.01)\). Among these loci, 77 SNPs were associated with the H4 concentration \((P < 0.05)\) (Supplementary Table S4 online). Besides SNPs in the CYP2C19 cluster and SLC14A2, four loci (ATP-binding cassette \([ABC]\)A1 rs2487032, ABCA4 rs4147820, MGST1 rs1236968, and UGT2B11 rs13123057) were significantly associated with the H4 concentration \((P < 0.01)\).

**Selected single-nucleotide polymorphism genotyping for replication studies in stages II and III**

SNPs were selected based on the following criteria for further validation: (1) \(P < 1.0 \times 10^{-4}\) for PRU or PRU > 208 for all SNPs and \(P < 0.05\) for the H4 concentration; (2) \(P < 0.01\) for PRU or PRU > 208 for SNPs in ADME genes and \(P < 0.05\) for the H4 concentration in stage I; (3) only the SNP with the lowest \(P\) value was chosen to represent multiple SNPs, which were in high LD \((r^2 \geq 0.8)\) for each gene. Eighteen SNPs satisfied the criteria for replication (Supplementary Table S5 online, Table 1).

**Contribution of the novel loci to variability of antiplatelet effects and pharmacokinetic of clopidogrel**

To identify the independent contribution of each factor to the interindividual variability in antiplatelet and PK response to clopidogrel, multivariate linear regression analysis was used. The results showed that CYP2C19*2, rs2254638 in N6AMT1, rs2487032 in ABCA1, use of calcium channel blockers, and sex were independent predictors for variability of the antiplatelet response to clopidogrel. These predictors explained 37.7% of the variance in PRU, among which 28.2% was explained by genotypes. CYP2C19*2, rs12456693 in SLC14A2, rs2254638 in N6AMT1, and age were independent predictors for variability of the H4 concentration. These predictors explained up to 28.3% of variability, of which 23.4% was explained by genotypes (Table 2).
| SNP         | CHR | Gene symbol | PRU >208 | PRU | H4 C<sub>2h</sub> | Clop T<sub>max</sub> | Clop C<sub>max</sub> | Clop AUC0-4h | H4 T<sub>max</sub> | H4 C<sub>max</sub> | H4 AUC0-4h | P value (n = 115) in stage I | P value (n = 31) in stage Ia | P value (n = 32) in stage Iib |
|-------------|-----|-------------|---------|-----|----------------|---------------------|---------------------|----------------|----------------|----------------|----------------|--------------------------------|--------------------------------|-------------------------------|
| rs74460025  | 5   | GABRB2      | 2.61 × 10^-5 | 5.08 × 10^-5 | 5.19 × 10^-3 | 0.679 | 0.973 | 0.672 | 0.760 | 0.740 | 0.668 | 0.9402 | 0.00861 | 0.0334 |
| rs17145154  | 7   | DNAH11      | 4.72 × 10^-5 | 6.36 × 10^-4 | 2.40 × 10^-2 | 0.672 | 0.557 | 0.799 | 0.0513 | 0.950 | 0.794 | 0.4269 | 0.0135 |
| rs4741806   | 9   | Intergenic  | 4.04 × 10^-4 | 6.28 × 10^-5 | 2.40 × 10^-3 | 0.972 | 0.483 | 0.544 | 0.495 | 0.918 | 0.541 | 0.9502 |
| rs1048196   | 10  | HELLS       | 1.45 × 10^-5 | 4.05 × 10^-5 | 4.20 × 10^-4 | 0.978 | 0.402 | 0.587 | 0.760 | 0.740 | 0.668 | 0.9402 |
| rs1926711   | 10  | CYP2C18     | 8.24 × 10^-6 | 3.51 × 10^-5 | 6.61 × 10^-3 | 0.667 | 0.153 | 0.291 | 0.0513 | 0.950 | 0.794 | 0.4269 |
| rs4244285   | 10  | CYP2C19     | 1.66 × 10^-5 | 6.01 × 10^-5 | 2.20 × 10^-4 | 0.955 | 0.125 | 0.205 | 0.773 | 0.517 | 0.0843 | 0.0153 |
| rs2852213   | 11  | GRK4        | 8.77 × 10^-5 | 5.12 × 10^-5 | 1.62 × 10^-2 | 0.593 | 0.147 | 0.0794 | 0.121 | 0.679 | 0.495 | 0.9842 |
| rs774392    | 12  | GRIP1       | 2.49 × 10^-5 | 3.18 × 10^-4 | 6.72 × 10^-3 | 0.432 | 0.444 | 0.369 | 0.382 | 0.611 | 0.387 | 0.961 |
| rs12913988  | 15  | ATP10A      | 1.57 × 10^-3 | 9.69 × 10^-6 | 1.47 × 10^-2 | 0.307 | 0.831 | 0.691 | 0.895 | 0.538 | 0.617 | 0.9948 |
| rs12456693  | 18  | SLC14A2     | 6.83 × 10^-5 | 1.07 × 10^-2 | 8.52 × 10^-3 | 0.00485 | 0.262 | 0.172 | 0.0444 | 0.0245 | 0.129 | 0.4131 |
| rs1571678   | 21  | N6AMT1      | 1.21 × 10^-3 | 2.65 × 10^-5 | 6.85 × 10^-3 | 0.0249 | 0.969 | 0.958 | 0.0551 | 0.286 | 0.358 | 0.0643 |
| rs2254638   | 21  | N6AMT1      | 1.87 × 10^-3 | 5.37 × 10^-5 | 6.29 × 10^-3 | 0.0249 | 0.969 | 0.958 | 0.0551 | 0.286 | 0.358 | 0.0386 |
| rs17209532  | 22  | Intergenic  | 2.46 × 10^-3 | 1.92 × 10^-5 | 3.08 × 10^-2 | 0.218 | 0.266 | 0.167 | 0.0670 | 0.520 | 0.306 | 0.7581 |
| rs729279    | 22  | Intergenic  | 7.39 × 10^-4 | 4.34 × 10^-5 | 9.08 × 10^-3 | 0.730 | 0.689 | 0.863 | 0.385 | 0.944 | 0.860 | 0.1483 |
| rs13123057  | 4   | UGT2B11     | 8.69 × 10^-3 | 8.94 × 10^-3 | 8.69 × 10^-4 | 0.334 | 0.984 | 0.847 | 0.332 | 0.396 | 0.850 | 0.7324 |
| rs12369968  | 12  | MGST1       | 3.43 × 10^-4 | 3.57 × 10^-3 | 9.18 × 10^-4 | 0.459 | 0.440 | 0.448 | 0.131 | 0.9094 | 0.135 | 0.9919 |
| rs2487032   | 9   | ABCA1       | 3.46 × 10^-3 | 3.75 × 10^-2 | 5.34 × 10^-3 | 0.243 | 0.0211 | 0.00590 | 0.0189 | 0.382 | 0.329 | 0.8812 |
| rs4147820   | 1   | ABCA4       | 1.74 × 10^-3 | 8.42 × 10^-6 | 7.00 × 10^-3 | 0.495 | 0.448 | 0.604 | 0.759 | 0.673 | 0.679 | 0.662 |

AUC, area under the curve; Clop, clopidogrel; C<sub>max</sub>, peak plasma concentration; PRU, P2Y<sub>12</sub> reaction unit; SNP, single-nucleotide polymorphism; T<sub>max</sub>, time of maximum plasma concentration.

SNPs were associated with PRU or PRU >208 (P < 1 × 10^-3) and H4<sub>2h</sub> (P < 0.01), or SNPs in absorption, distribution, metabolism, and excretion gene were associated with PRU or PRU >208 (P < 0.05) and H4<sub>2h</sub> (P < 0.01) in stage I.

SNPs in bold indicate associations with pharmacokinetic parameters of clopidogrel or its active metabolite H4 in stage Ila.
Effects of single-nucleotide polymorphisms on the pharmacokinetic of clopidogrel and H4 in patients with coronary heart disease
Supplementary Figure S3 online presents the plasma concentration-time profiles of clopidogrel and its metabolites obtained in 31 patients after the administration of clopidogrel in stage Ia. The absorption of clopidogrel from the gastrointestinal tract widely varied. The PK parameters calculated for clopidogrel and its metabolites are presented in Supplementary Table S6 online.

To investigate the absorption and metabolic mechanisms that influenced antplatelet response to clopidogrel treatment, SNPs associated with antplatelet effects and PKs of clopidogrel were further studied in 31 patients with CHD along with PK parameters of clopidogrel and H4 (Supplementary Table S7 online, Table 1). Among the 18 significant SNPs identified in stage I, rs1048196 in HELLs, rs1926711 in CYP2C18, and CYP2C19*2 in the HELLs-CYP2C18-CYP2C19 cluster were associated with decreased H4 concentration (9.21 ± 3.68 ng/mL/mg protein for TT; 7.69 ± 6.48 ng/mL/mg protein for TC; and 3.36 ± 3.75 ng/mL/mg protein for CC; P = 0.0386). Other SNPs were not associated with the H4 concentration in human liver S9 fractions (P > 0.05).

Effects of the novel loci on the risk of major adverse cardiac event in patients with coronary heart disease
To validate whether the significant SNPs identified potentially influence the clinical outcome, these SNPs were genotyped for 299 patients with CHD after percutaneous coronary intervention by the Sequenom’s MassARRAY system. Rs1571678 was not included, because it was in strong LD with rs2254638 in N6AMT1 at r² = 0.93. The call rate of the SNPs ranged from 96.33–99.80% (Supplementary Table S9 online).

Logistic regression analysis showed that diabetes mellitus (odds ratio [OR] = 1.666; 95% confidence interval [CI] = 0.977–2.84; P = 0.0608), heart failure (OR = 5.101; 95% CI = 2.617–9.94; P < 0.0001), arrhythmia (OR = 3.695; 95% CI = 1.793–7.615; P = 0.0004), and use of calcium channel blockers (OR = 2.251; 95% CI = 1.338–3.788; P = 0.0022) exerted a significant effect on the risk of MACE (Table 3). Rs2254638 in N6AMT1 was marginally associated with the occurrence of MACE (P = 0.0653; false discovery rate [FDR] = 0.4726). Patients with the C allele had a higher risk of MACE than those without (OR = 1.428; 95% CI = 0.978–2.086). Rs12456693 in SLC14A2 was marginally associated with the occurrence of MACE (P = 0.0953; FDR = 0.4726). No significant association was observed between CYP2C19*2 and MACE (P = 0.9463; FDR = 0.9463). Rs12913988 in ATP10A was found to be significantly associated with the occurrence of MACE (P = 0.0011; FDR = 0.0187). Patients with the T allele had a higher risk of MACE in 1.5 years (OR = 1.876; 95% CI = 1.285–2.740). Other SNPs were not associated with the occurrence of MACE (P > 0.1; FDR >0.5; Table 4).
DISCUSSION

This work is the first systematic GWAS combining analysis of antiplatelet effects and PKs to investigate functional genetic variants that modify clopidogrel efficacy in Chinese patients with CHD. We identified new genetic variants that affect the antiplatelet response to clopidogrel by changing PK behavior in patients with coronary heart disease. The new variants in *N6AMT1* and *ABCA1*, together with *CYP2C19* and clinical factors, dramatically

Figure 3  Association of significant single-nucleotide polymorphisms (a) rs4244285 in CYP2C19, also known as CYP2C19*2; (b) rs12456693 in SLC14A2; (c) rs2254638 in N6AMT1; and (d) rs2487032 in ABCA1 with pharmacokinetic (PK) parameters of clopidogrel and H4 in 31 patients with coronary heart disease. The PK parameters include area under the concentration-time curve (AUC), peak concentration (Cmax), and peak concentration (Tmax).
improved the predictability of PRU variability to 37.7% compared with the published value of ∼20%. Rs12456693 in SLC14A2 was also an independent predictor for the variability of the H4 concentration. N6AMT1 rs2254638 and SLC14A2 rs12456693 were further identified to exert a marginal risk effect on the occurrence of MACE in an independent patient with CHD cohort.

We confirmed the association between variants in the HELLS-CYP2C18-CYP2C19 cluster and antiplatelet effects and PK of clopidogrel.\textsuperscript{12,22} CYP2C19*2 independently explained 16% of the variation in the logarithm of the H4 concentration at 2 h and 11% of PRU variation after LD of clopidogrel, similar to a published value of ∼12% of the variation in the clopidogrel response.\textsuperscript{12} Whereas CYP2C19*3 did not have a significant contribution to antiplatelet effects, PK, and activation of clopidogrel. The reason may be due to the limited effect of CYP2C19*3 on the activation of clopidogrel, small sample number, and low

Table 3 Association of baseline characteristics with the occurrence of major adverse cardiac event in 299 patients with coronary heart disease

| Characteristics     | Case Mean ± SD | Control Mean ± SD | P value | OR per variant allele (95% CI) |
|---------------------|----------------|-------------------|---------|-----------------------------|
| Demographic data    |                |                   |         |                             |
| Age, y              | 65.71 ± 9.95  | 64.08 ± 11.43     | 0.2388  | 1.014 (0.991–1.037)         |
| Male                | 73 (81.11)    | 166 (79.43)       | 0.7386  | 1.112 (0.595–2.079)         |
| Medical history     |                |                   |         |                             |
| Diabetes mellitus   | 32 (35.56)    | 52 (24.88)        | 0.0608* | 1.666 (0.977–2.84)*         |
| Hypertension        | 60 (66.67)    | 121 (57.89)       | 0.1556  | 1.455 (0.867–2.44)          |
| Heart failure       | 28 (31.11)    | 17 (8.13)         | <0.0001* | 5.101 (2.617–9.94)*         |
| ACS                 | 71 (78.89)    | 155 (74.16)       | 0.3837  | 1.302 (0.719–2.357)         |
| Arrhythmia          | 20 (22.22)    | 15 (7.18)         | 0.0004* | 3.695 (1.793–7.615)*        |
| Biochemical measures|                |                   |         |                             |
| ALT                 | 38.99 ± 63.15 | 29.48 ± 29.15     | 0.1311  | 1.005 (0.999–1.011)         |
| AST                 | 38.23 ± 41.41 | 39.75 ± 72.11     | 0.8548  | 1 (0.996–1.004)             |
| LDLC                | 2.57 ± 1.02   | 2.84 ± 2.12       | 0.2416  | 0.862 (0.673–1.105)         |
| HDLC                | 0.97 ± 0.25   | 0.99 ± 0.25       | 0.5219  | 0.717 (0.259–1.986)         |
| TRIG                | 1.65 ± 1.24   | 1.69 ± 1.26       | 0.8092  | 0.975 (0.793–1.199)         |
| APOA                | 0.98 ± 0.25   | 1.03 ± 0.26       | 0.1739  | 0.45 (0.142–1.423)          |
| CHOL                | 4.24 ± 1.14   | 4.53 ± 1.25       | 0.0665  | 0.81 (0.647–1.014)          |
| CK                  | 191.44 ± 295.10 | 181.62 ± 345.37  | 0.8293  | 1.0 (0.999–1.001)           |
| CKMB                | 9.22 ± 11.75  | 8.44 ± 9.83       | 0.5874  | 1.007 (0.982–1.032)         |
| CREA                | 102.32 ± 83.21 | 94.41 ± 43.20     | 0.3425  | 1.002 (0.998–1.006)         |
| GLUC                | 6.97 ± 2.82   | 6.45 ± 2.41       | 0.3109  | 1.002 (0.998–1.006)         |
| LPa                 | 274.97 ± 323.41 | 306.37 ± 340.55  | 0.4993  | 1 (0.999–1.001)             |
| Medication          |                |                   |         |                             |
| Statins             | 89 (98.89)    | 208 (99.52)       | 0.55    | 0.428 (0.026–6.918)         |
| β-blockers          | 84 (93.33)    | 195 (93.3)        | 0.9919  | 1.005 (0.373–2.705)         |
| ACE inhibitors      | 72 (80.00)    | 151 (72.25)       | 0.1599  | 1.536 (0.844–2.795)         |
| CCBs                | 39 (43.33)    | 53 (25.36)        | 0.0022* | 2.251 (1.338–3.788)*        |
| Proton pump inhibitors | 56 (62.22) | 116 (55.50)    | 0.2815  | 1.32 (0.796–2.19)           |

ACE, angiotensin-converting enzyme; ACS, acute coronary syndrome; ALT, alanine aminotransferase; APOA, apolipoprotein A; AST, aspartate aminotransferase; CCB, calcium channel blockers; CHOL, total cholesterol; CI, confidence interval; CK, cytokeratin; CKMB, creatine kinase-MB; CREA, creatinine; GLUC, glucose; HDLC, high-density lipoprotein cholesterol; LDLC, low-density lipoprotein cholesterol; LPa, lipoprotein; OR, odds ratio; TRIG, triglyceride.

*Indicates the associations between the corresponding baseline characteristics and the occurrences of major adverse cardiac event are significant.
Table 4  Association of genetic polymorphisms with the occurrence of major adverse cardiac event in 299 patients with coronary heart disease

| SNP      | Gene symbol | Genotype | With MACE | Without MACE | OR per variant allele (95% CI) | P value | FDR |
|----------|-------------|----------|-----------|--------------|--------------------------------|---------|-----|
| rs74460025 | GABRB2      | TT       | 64 (71.91) | 160 (77.29) | 1.487 (0.9–2.457)              | 0.1216  | 0.4726 |
|          | TC          | 21 (23.60) | 46 (22.22) |
|          | CC          | 4 (4.49)  | 1 (0.48)   |
| rs17145154 | DNAH11      | TT       | 51 (59.3)  | 104 (51.23) | 1.272 (0.859–1.884)           | 0.2299  | 0.6292 |
|          | TG          | 28 (32.56) | 78 (38.42) |
|          | GG          | 7 (8.14)  | 21 (10.34) |
| rs4741806 | Intergenic  | AA       | 72 (80.9)  | 158 (76.33) | 0.718 (0.402–1.282)          | 0.2623  | 0.6292 |
|          | AC          | 17 (19.1)  | 45 (21.74) |
|          | CC          | 0 (0)     | 4 (1.93)   |
| rs1048196 | HELLS       | CC       | 41 (46.07) | 92 (45.1)   | 0.942 (0.638–1.393)          | 0.7661  | 0.9422 |
|          | CT          | 41 (46.07) | 93 (45.59) |
|          | TT          | 7 (7.78)  | 19 (9.31)  |
| rs1926711 | CYP2C18     | GG       | 33 (37.5)  | 74 (36.63)  | 1.022 (0.693–1.505)         | 0.9143  | 0.9463 |
|          | GA          | 45 (51.14) | 105 (51.98) |
|          | AA          | 10 (11.36) | 23 (11.39) |
| rs4244285 | CYP2C19     | GG       | 42 (46.67) | 98 (47.34)  | 0.987 (0.666–1.461)         | 0.9463  | 0.9463 |
|          | GA          | 42 (46.67) | 90 (43.48) |
|          | AA          | 6 (6.67)  | 19 (9.18)  |
| rs2852213 | GRIK4       | CC       | 13 (14.61) | 62 (30.10)  | 1.302 (0.918–1.847)         | 0.139   | 0.4726 |
|          | CT          | 54 (60.67) | 89 (43.20) |
|          | TT          | 22 (24.72) | 55 (26.70) |
| rs774392  | GRIP1       | CC       | 3 (3.33)   | 3 (1.44)    | 0.904 (0.548–1.49)          | 0.6913  | 0.9422 |
|          | CA          | 20 (22.22) | 49 (23.56) |
|          | AA          | 67 (74.44) | 156 (75.00) |
| rs12913988 | ATP10A      | CC       | 7 (7.78)   | 43 (20.77)  | 1.876 (1.285–2.740)         | 0.0011  | 0.0187 |
|          | CT          | 40 (44.44) | 99 (47.83) |
|          | TT          | 43 (47.78) | 65 (31.4)  |
| rs12456693 | SLC14A2     | CC       | 66 (74.16) | 141 (67.46) | 0.654 (0.398–1.077)         | 0.0953  | 0.4726 |
|          | CT          | 23 (25.84) | 58 (27.75) |
|          | TT          | 0 (0)     | 10 (4.78)  |
| rs22546388 | N6AMT1      | TT       | 13 (14.77) | 42 (20.59)  | 1.428 (0.978–2.086)         | 0.0653  | 0.4726 |
|          | TC          | 45 (51.14) | 113 (55.39) |
|          | CC          | 30 (34.09) | 49 (24.02) |
| rs17209532 | Intergenic  | GG       | 33 (36.67) | 92 (45.10)  | 0.822 (0.57–1.187)          | 0.2961  | 0.6292 |
|          | GA          | 46 (51.11) | 88 (43.14) |
|          | AA          | 11 (12.22) | 24 (11.76) |
| rs7292279 | Intergenic  | GG       | 31 (34.44) | 82 (39.42)  | 0.974 (0.68–1.396)          | 0.8876  | 0.9463 |
|          | GA          | 48 (53.33) | 94 (44.98) |
variant allele frequency.\textsuperscript{23} Although many studies have shown a positive association between \textit{CYP2C19} functional variants and cardiovascular outcomes, we did not find a significant association between \textit{CYP2C19*2} or \textit{*3} polymorphism and MACE. This result was in line with the findings of two previous meta-analyses\textsuperscript{24,25} and the finding from a recent study.\textsuperscript{26} One of the novel findings in the study is that rs2254638 in \textit{N6AMT1} may be a potential genetic biomarker to predict the individual difference of the intermediate efficacy response and clinical endpoint to clopidogrel treatment through metabolic and PK mechanisms. Our study showed that rs2254638 in \textit{N6AMT1} could lower liver metabolic function, reduce the plasma H4 concentration, decrease the antiplatelet effects of clopidogrel with a higher treated PRU, and confer a risk effect on the occurrence of MACE. \textit{N6AMT1} rs2254638 independently explained 4.5% of the variation in the logarithm of the H4 concentration 2 h and 14.8% of PRU variation. The mechanistic basis for the function of \textit{N6AMT1} in clopidogrel activation remains unclear. The \textit{N6AMT1} gene was identified in humans and encodes an N(6)-adenine-specific DNA methyltransferase.\textsuperscript{27} This enzyme possesses specific activity in converting the arsenic metabolite, monomethylarsonous acid, to the less toxic dimethylarsinic acid.\textsuperscript{28} Harari \textit{et al}.\textsuperscript{29} reported that \textit{N6AMT1} polymorphisms are associated with arsenic methylation in Andean women. Whether \textit{N6AMT1} is involved in the activation of clopidogrel directly or through other mechanisms, such as methylation of other metabolic enzymes, should be elucidated in further studies.

Compared with the metabolism of clopidogrel, the current knowledge about its absorption is limited. Only one transporter, a P-glycoprotein encoded by \textit{ABCB1}, has been reported to be involved in the intestinal absorption of clopidogrel.\textsuperscript{30} Almost all previous publications concerning clopidogrel absorption variability focused on the effects of genetic polymorphism of \textit{ABCB1} on antiplatelet and PK response to clopidogrel and clinical outcomes. However, published data on the association of \textit{ABCB1} genetic variants and antiplatelet effects and PKs of clopidogrel are inconsistent.\textsuperscript{30–33} We did not find an SNP consistently associated with PRU and H4 concentration. \textit{ABCB1} genetic variants may not be a key factor contributing to the absorption variability of clopidogrel. Genetic variants in other transporters involving clopidogrel disposition may contribute to the large coefficient of variation in \textit{Tmax} and \textit{Cmax} values. The PK of clopidogrel and H4 may be affected not only by the transporter in the absorption phase but also by the transporter in the excretion phase.

We identified novel variants in two transporter genes, \textit{SLC14A2} and \textit{ABCA1}, associated with antiplatelet or PK response to clopidogrel that may enhance our understanding of the absorption and excretion of clopidogrel. \textit{ABCA1} was independently associated with PRU but not with H4 at 2 h after a loading dose of clopidogrel, whereas \textit{SLC14A2} was independently associated with H4 in the elimination phase. These findings suggested that the mechanisms for \textit{ABCA1} and \textit{SLC14A2} involved in the PK of clopidogrel and H4 differed.

\begin{table}[h]
\centering
\caption{Continued}  
\begin{tabular}{lcccccc}
\hline
SNP & Gene symbol & Genotype & With MACE & Without MACE & OR per variant allele (95\% CI) & \textit{P} value & FDR \\
\hline
 & & & No. (%) & No. (%) & & \\
rs13123057 & UGT2B11 & AA & 11 (12.22) & 33 (15.79) & & \\
 & & GT & 28 (31.11) & 72 (34.78) & & \\
 & & AG & 7 (7.78) & 16 (7.73) & & \\
 & & TT & 9 (10.00) & 12 (5.80) & & \\
 & & AT & 5 (5.56) & 7 (3.38) & & \\
 & & AA & 0 (0) & 2 (0.97) & & \\
rs12369968 & MGST1 & AA & 32 (33.71) & 73 (34.93) & 1.16 (0.814–1.655) & 0.4114 & 0.7052 \\
 & & AG & 40 (44.94) & 104 (49.76) & & \\
 & & GG & 19 (21.35) & 32 (15.31) & & \\
rs2487032 & ABCA1 & GG & 36 (40.91) & 83 (40.1) & 0.954 (0.688–1.322) & 0.7759 & 0.9422 \\
 & & GA & 31 (35.23) & 82 (39.61) & & \\
 & & AA & 21 (23.86) & 42 (20.29) & & \\
rs4147820 & ABCA4 & GG & 44 (48.89) & 120 (57.42) & 1.177 (0.796–1.739) & 0.4148 & 0.7052 \\
 & & GA & 41 (45.56) & 73 (34.93) & & \\
 & & AA & 5 (5.56) & 16 (7.66) & & \\
\hline
\end{tabular}
\end{table}

\textsuperscript{CI}, confidence interval; FDR, false discovery rate; MACE, major adverse cardiac event; OR, odds ratio; SNP, single-nucleotide polymorphism. FDR control was used to correct for multiple comparisons. *Indicates the associations of genetic polymorphisms with the occurrence of MACE are significant.
SLC14A2 is a member of the solute carrier family and encodes six urea transporters (UTs), UT-A1 to UT-A6 isoforms, in mammals. UT transporters are found mainly in the kidneys and heart,35 cochlea,35 placenta,36 colon, brain, and liver.37 UT’s facilitate the reabsorption and recycling of the urinary urea, thereby increasing medullary urea concentration.38 Variants in SLC14A2 were associated with blood urea nitrogen levels in a Japanese population, but no clear phenotypic effect was demonstrated.39 We hypothesized that SLC14A2 was involved in the excretion of clopidogrel, but this hypothesis should be tested in further studies. The newly identified genetic variants in SLC14A2 associated with antiplatelet effects and PKs of clopidogrel were found to have a marginal effect on clinical outcomes for patients with CHD after percutaneous coronary intervention.

ABCA1 encodes a member of the ABC transporter family and is located on chromosome 9q22.31. ABCA1 is expressed in many tissues and cells, including enterocytes and hepatocytes,40 and plays a major role in cholesterol efflux. In enterocytes, ABCA1 is localized on the basolateral plasma membrane,41 and induction of ABCA1 enhances basolateral (systemic) absorption of dietary cholesterol that is apically assimilated by enterocytes.42 McNeish et al.43 reported that ABCA1 disruption in mice leads to the accumulation of intestinally derived cholesterol in plasma very low-density lipoprotein. Our results showed that a variant of rs2487032, located in an ABCA1 promoter, increased the concentration of plasma clopidogrel’s active metabolite H4. This effect increased inhibition of platelet function. The mechanism may involve an increase in the speed of absorption of clopidogrel and H4. However, the SNP was not associated with clinical outcomes.

We also identified a new genetic variant (rs12913988) in ATP10A that functions as a significant risk factor for MACE occurrence, and it enhances the antiplatelet effects of clopidogrel with a low treated PRU and increases the plasma H4 concentration in stage I. The underlying reason leading to the result is unclear and requires further study. ATP10A encodes a protein belonging to the family of P-type cation transport ATPases and to the subfamily of aminophospholipid-transporting ATPases. The appearance of phosphatidylserine on the surface of animal cells triggers phagocytosis and blood coagulation.44 ATPases transport a variety of different compounds, including ions and phospholipids, across a membrane using ATP hydrolysis for energy.

The novel SNPs detected in our study escaped detection in a previous European GWAS.12 One of the reasons could be frequency differences among populations for SNPs. For example, rs2254638 in N6AMT1 and rs12913988 in ATP10A are more frequent in East Asians (~50–60%) than Europeans (~10–15%). Another reason could be the difference of chips used. For example, the previous GWAS in European populations adopted Affymetrix GeneChip Human Mapping 500K or 1M (version 6.0) arrays12 did not capture rs12913988 in ATP10A.

Our study had two limitations that merit mention. (1) The sample size was small; therefore, findings may exhibit false positivity because of multiple comparisons, a problem inherent in GWAS. To address this issue, we jointly analyzed antiplatelet effects and PKs of clopidogrel with genomewide SNPs and adopted a P value at the significance level of CYP2C19*2 for prioritizing functional SNPs. Furthermore, we used not only other patient populations to replicate and confirm findings but in liver S9 fractions to validate the impact of SNPs in vitro. The validated SNPs with biological plausibility of ADME have a high likelihood of being true-positive findings. We further investigated the association of genetic polymorphisms with the clinical outcome in another large population. (2) The H4 concentrations at 2 h after clopidogrel were used as a substitute PK parameter in GWAS of stage I and may have compromised the power to some extent. The PK parameters, including AUIC and clearance, were originally designed to be estimated by a nonlinear mixed-effects modeling method. However, given the large individual differences in the absorption phase of clopidogrel, T_max reached 4 h, which was the last timepoint of blood drawn, in some patients. Nonlinear mixed-effects modeling of population PK may be biased. In addition, the H4 concentration at 2 h was highly correlated with AUIC_{0–4h} (r = 0.8141; P < 0.0001) in 31 patients (Supplementary Figure S4 online).

In conclusion, we systematically identified new genetic variants as risk factors for reduced efficacy of clopidogrel and highlighted related genes that may be involved in antiplatelet effects and PK of clopidogrel. Our results enhance our understanding of the absorption and metabolic mechanisms that influence antiplatelet response to clopidogrel treatment.

METHODS
A detailed Methods section is provided in the Supplementary Data online.

Study population and study design
A three-stage GWAS in Han Chinese patients with CHD was performed. In stage I, the GWAS of on-treatment platelet reactivity was performed to identify SNPs associated with antiplatelet effects of clopidogrel in 115 patients with CHD. The concentration of H4 was analyzed to identify a PK explanation for SNPs associated with antiplatelet effects in the same cohort. In stage IIa, SNPs associated with antiplatelet effects and PKs of clopidogrel were further studied in 31 patients with CHD along with PK parameters of clopidogrel and H4. In stage IIb, the function of these SNPs on clopidogrel activation was further investigated in 32 human liver tissues. In stage III, clinical outcomes (MACE) were used to study the association between these SNPs and response to clopidogrel in 299 patients with CHD. This study was approved by the Medical Ethical Review Committee of Guangdong General Hospital. All subjects provided written informed consent.

Sample processing
In stage I, patients received a 300 mg loading dose of clopidogrel and a 100 mg dose of aspirin. Blood samples were collected for determining PRU and the plasma concentration of clopidogrel, active metabolite isomer H4, and inactive metabolite isomer H3 at 2 and 4 h after the loading dose. In stage IIa, blood samples for the PK assessment of clopidogrel and its metabolites were collected at 0, 0.25, 0.5, 1, 2, and 4 h after a 300 mg loading dose of clopidogrel.

Antiplatelet effect analysis
The antiplatelet effectiveness of clopidogrel was measured using the VerifyNow, assay as described by the manufacturer (Accumetrics, San Diego, CA). The test of platelet function using PRU was assessed separately as a continuous measure and a categorical measure, which was defined as...
high or low platelet reactivity on clopidogrel using a PRU cutoff value higher than 208 based on previous studies.1,45

Bioanalytical analysis
A high-performance liquid chromatography tandem mass spectrometry assay was developed and validated for the simultaneous determination of clopidogrel, metabolite stereoisomers (H3 and H4), and internal standard carbamazepine in human plasma and mixtures of liver S9 fraction. The representative tandem mass spectrometry spectrum and liquid chromatography mass spectrometry profiles are shown in Supplementary Figure S5 online. The PK parameters of clopidogrel, H3, and H4 were estimated using the noncompartmental analysis function in Phoenix WinNonlin version 6.3 software (Pharsight, Cary, NC).

Genotyping and quality control in the genomewide association study
A total of 900,015 SNPs in a GWAS scan of 117 subjects were genotyped with the Illumina HumanOmniZhongHua-8 BeadChip according to the Infinium HD protocol from Illumina. Prior to association analysis, a systematic quality control procedure was applied to the raw genotyping data to filter unqualified SNPs and samples. We detected population outliers using a method based on PCA. After quality filtering and cleaning, 115 subjects with 703,143 SNPs were included in further analyses.

Candidate single-nucleotide polymorphism association in absorption, distribution, metabolism, and excretion genes
We focused particularly on the genes involved in ADME to identify genetic variants with PK mechanisms associated with response to clopidogrel using combination analysis of antiplatelet effects and PK. We closely examined 16,202 SNPs in 295 candidate ADME genes from PharmaADME (http://www.pharmaadme.org/) and their surrounding regions (±2 kb) for association with PRU or PRU > 208 and the H4 concentration in 115 patients with CHD.

Pharmacokinetic replication study
To confirm the association between significant SNPs with PK parameters of clopidogrel and H4, as well as further assess possible mechanisms for the association between significant SNPs and PRU, a replication study in 31 patients with CHD was performed in stage IIa. The plasma concentration was determined at 0, 0.25, 0.5, 1, 2, and 4 h.

Functional replication study on clopidogrel activation
In stage IIb, the function of these SNPs on clopidogrel activation was further investigated in 32 human liver tissues to identify a plausible mechanism for newly identified genetic variants. The formation of H4 in the liver S9 fraction was analyzed using a validated liquid chromatography tandem mass spectrometry assay. Genotyping of 18 SNPs was performed using the iPLEX MassARRAY platform (Sequenom).

Clinical validation study
In stage III, we used clinical outcomes as a measure to study the association between these SNPs and clopidogrel efficacy. A total of 299 Han Chinese patients with CHD who received clopidogrel therapy were sequentially recruited and followed up to 1.5 years, including 91 patients with MACE and 208 patients without MACE as control. Genotyping was conducted by Sequenom’s MassARRAY system (Sequenom). The effect of genetic variants on the clinical outcome of MACE in 1.5 years was assessed.

Statistics
Details on statistical analysis can be found in the Supplementary Data online. Additional Supporting Information may be found in the online version of this article.
transmission aggregometry from the responsiveness to clopidogrel and stent thrombosis 2-acute coronary syndrome (RECLUSE 2-ACS) study. J. Thromb. Thrombolysis 40, 76–82 (2015).

9. Zhu, H.-J., Wang, X., Gawronski, B.E., Brinda, B.J., Angiolillo, D.J. & Markowitz, J.S. Carboxylesterase 1 as a determinant of clopidogrel metabolism and activation. J. Pharmacol. Exp. Ther. 344, 665–672 (2013).

10. Viviani Anselmi, C. et al. Routine assessment of on-clopidogrel platelet reactivity and gene polymorphisms in predicting clinical outcome following drug-eluting stent implantation in patients with stable coronary artery disease. JACC Cardiovasc. Interv. 6, 1166–1175 (2013).

11. Mega, J.L. et al. Reduced-function CYP2C19 genotype and risk of adverse clinical outcomes among patients treated with clopidogrel predominantly for PCI: a meta-analysis. JAMA 304, 1821–1830 (2010).

12. Shuldiner, A.R. et al. Association of cytochrome P450 2C19 genotype with the antiplatelet effect and clinical efficacy of clopidogrel therapy. JAMA 302, 849–857 (2009).

13. Perry, C.G. & Shuldiner, A.R. Pharmacogenomics of anti-platelet therapy in patients with coronary heart disease in terms of CYP2C19 polymorphism [in Russian]. Donakanyan, S.A. The possibility of selecting optimal antiplatelet metabolites in patients with cardiovascular diseases. J. Clin. Pharmacol. Ther. 874–880 (2014).

14. Bokeriya, O.L., Kudzoeva, Z.F., Shvarts, V.A., Koasari, A.K. & Bouman, H.J. et al. Polymorphism in arsenic-induced toxicity. Environ. Health Perspect. 119, 771–777 (2011).

15. Ahn, K., Luo, J., Berg, A., Keefe, D. & Wu, R. Functional mapping of smoking, co-medications (including proton pump inhibitors), and pre-treatment in on-treatment platelet reactivity explained by CYP2C19*2 genotype is modest in clopidogrel pretreated patients undergoing coronary stenting. Heart 97, 1239–1244 (2011).

16. Frelinger, A.L. 3rd et al. Clopidogrel pharmacokineticians and pharmacodynamics vary widely despite exclusion or control of polymorphisms (CYP2C19, ABCB1, PON1), noncompliance, diet, smoking, co-medications (including proton pump inhibitors), and pre-existent variability in platelet function. J. Am. Coll. Cardiol. 61, 872–879 (2013).

17. Chan, N.C. et al. Role of phenotypic and genetic testing in managing clopidogrel therapy. Blood 124, 689–699 (2014).

18. Ahn, K., Luo, J., Berg, A., Keefe, D. & Wu, R. Functional mapping of drug response with pharmacodynamic-pharmacokinetic principles. Trends Pharmacol. Sci. 31, 306–311 (2010).

19. Wu, R. et al. A conceptual framework for pharmacodynamic genome-wide association studies in pharmacogenomics. Drug Discov. Today 16, 884–890 (2011).

20. Goswami, S. et al. Genetic variants in transcription factors are associated with the pharmacokinetics and pharmacodynamics of metformin. Clin. Pharmacol. Ther. 96, 370–379 (2014).

21. Karazhnievivz-Lada, M., Danielak, D., Rubi, B., Burchardt, P., Oszkinis, G. & Gloska, F. The influence of genetic polymorphism of Cyp2c19 isoenzyme on the pharmacokinetics of clopidogrel and its metabolites in patients with cardiovascular diseases. J. Clin. Pharmacol. 54, 874–880 (2014).

22. Brandt, J.T. et al. Common polymorphisms of CYP2C19 and CYP2C9 affect the pharmacokinetic and pharmacodynamic response to clopidogrel but not prasugrel. J. Thromb. Haemost. 5, 2429–2436 (2007).

23. Man, M. et al. Genetic variation in metabolizing enzyme and transporter genes: comprehensive assessment in 3 major East Asian populations with comparison to Caucasians and Africans. J. Clin. Pharmacol. 50, 929–940 (2010).

24. Holmes, M.V., Perel, P., Shah, T., Hingorani, A.D. & Casas, J.P. CYP2C19 genotype, clopidogrel metabolism, platelet function, and cardiovascular events: a systematic review and meta-analysis. JAMA 306, 2704–2714 (2011).

25. Baurer, T., Bouman, H.J., van Werkum, J.W., Ford, N.F., ten Berg, J.M. & Taubert, D. Impact of CYP2C19 variant genotypes on clinical efficacy of antiplatelet treatment with clopidogrel: systematic review and meta-analysis. BMJ 343, d4588 (2011).

26. Doni, J.A. et al. Impact of CYP2C19 metabolizer status on patients with ACS treated with prasugrel versus clopidogrel. J. Am. Coll. Cardiol. 67, 936–947 (2016).

27. Ratel, D. et al. Undetectable levels of N6-methyl adenine in mouse DNA: cloning and analysis of PRED28, a gene coding for a putative mammalian DNA adenine methyltransferase. FEBS Lett. 580, 3179–3184 (2006).

28. Ren, X. et al. Involvement of N6 adenine-specific DNA methyltransferase 1 (N6AMT1) in arsenic biomethylation and its role in arsenic-induced toxicity. Environ. Health Perspect. 119, 771–777 (2011).

29. Harari, F., Engström, K., Concha, G., Colque, G., Vahter, M. & Broberg, K. N-6-adenine-specific DNA methyltransferase 1 (N6AMT1) polymorphisms and arsenic methylation in Andean women. Environ. Health Perspect. 121, 797–803 (2013).

30. Taubert, D. et al. Impact of P-glycoprotein on clopidogrel absorption. Clin. Pharmacol. Ther. 80, 486–501 (2006).

31. Karazhnievivz-Lada, M. et al. Impact of common ABCB1 polymorphism on pharmacokinetics and pharmacodynamics of clopidogrel and its metabolites. J. Clin. Pharm. Ther. 40, 226–231 (2015).

32. Nassar, S., Anmo, O., Abu-Rmaileh, H., Alshaer, I., Korachi, M. & Ayesh, S. ABCB1 C3435T and CYP2C19*2 polymorphisms in a Palestinian and Turkish population: a pharmacogenetic perspective to clopidogrel. Meta Gene 2, 314–319 (2014).

33. Su, J.F., Hu, X.H. & Li, C.Y. Risk factors for clopidogrel resistance in patients with ischemic cerebral infarction and the correlation with gene rs1045642 polymorphism. Exp. Ther. Med. 9, 267–271 (2015).

34. Duchesne, R. et al. UT-A urea transporter protein in heart: increased abundance during uremia, hypertension, and heart failure. Circ. Res. 89, 139–145 (2001).

35. Kwan, Y.S. et al. Immunohistochemical localization of urea transporters A and B in the rat cochlea. Hear. Res. 183, 84–96 (2003).

36. Damiano, A.E., Zotta, E. & Ibarra, C. Functional and molecular expression of AQP9 channel and UT-A transporter in normal and preeclamptic human placentas. Placenta 27, 1073–1081 (2006).

37. Fenton, R.A. et al. Characterization of mouse urea transporters UT-A1 and UT-A2. Am. J. Physiol. Renal Physiol. 283, F817–F825 (2002).

38. Stewart, G. The emerging physiological roles of the SLC14A family of transporters A and B in the rat cochlea. Hear. Res. 272, 273–283 (2002).

39. Okamura, T. et al. Dominant expression of ATP-binding cassette transporter-1 on basolateral surface of Caco-2 cells stimulated by LXR/RXR ligands. Biochem. Biophys. Res. Commun. 296, 625–630 (2002).

40. Myrthy, S., Born, E., Mathur, S.N. & Field, F.J. LXR/RXR activation and UT-A2. Trends Pharmacol. Sci. 31, 139–145 (2010).

41. Hammerschmidt, K., Burchardt, P., Oszkinis, G. & Gloska, F. The influence of genetic polymorphism of Cyp2c19 isoenzyme on the pharmacokinetics of clopidogrel and its metabolites in patients with cardiovascular diseases. J. Clin. Pharmacol. 34, 874–880 (2014).

42. Brandt, J.T. et al. Common polymorphisms of CYP2C19 and CYP2C9 affect the pharmacokinetic and pharmacodynamic response to clopidogrel but not prasugrel. J. Thromb. Haemost. 5, 2429–2436 (2007).

43. Mao, M. et al. Genetic variation in metabolizing enzyme and transporter genes: comprehensive assessment in 3 major East Asian populations with comparison to Caucasians and Africans. J. Clin. Pharmacol. 50, 929–940 (2010).

44. Holmes, M.V., Perel, P., Shah, T., Hingorani, A.D. & Casas, J.P. CYP2C19 genotype, clopidogrel metabolism, platelet function, and cardiovascular events: a systematic review and meta-analysis. JAMA 306, 2704–2714 (2011).

45. Baurer, T., Bouman, H.J., van Werkum, J.W., Ford, N.F., ten Berg, J.M. & Taubert, D. Impact of CYP2C19 variant genotypes on clinical