Objectives: The present study was undertaken to screen the analgesic and the antidepressant activity of *Gaultheria fragrantissima* Wall. an ethnomedicinal plant of Meghalaya, India. 

Methods: In this study, the analgesic effect was induced by hot plate method and tail immersion test. The analgesic effect of two doses of the methanolic extract of *Gaultheria fragrantissima* Wall. was tested and diclofenac was used as a standard drug. The antidepressant activity was brought about by force swimming test and tail suspension test. The antidepressant effect of two doses of the methanolic extract of *G. fragrantissima* Linn. was tested and imipramine was used as a standard drug. The statistical analysis was carried out by one-way analysis of variance (ANOVA) followed by Tukey-Kramer multiple comparison tests using GraphPad InStat 3.0 software. 

Results: The analgesic study shows that the plant extract at two different doses (200 mg/kg and 400 mg/kg) possesses significant analgesic activity (p<0.001), whereby the two different doses of the plant extract showed a significant increase in the latency time were obtained at 120 min in hot plate method and increase in the latency time were obtained at 90 min in the tail immersion method, respectively. The antidepressant study shows that the methanolic extract at 200 and 400 mg/kg produced significant reduction (p<0.001) in the immobility period when compared with that of control group animals in both the tests. 

Conclusion: The result of the study shows that the methanolic bark extract of *G. fragrantissima* Wall. possesses analgesic activity and antidepressant activity. 

Keywords: *Gaultheria fragrantissima*, Analgesic, Antidepressant, Diclofenac, Latency.

INTRODUCTION

The most ancient form of health care in the world is usually used in the prevention, and treatment of physical and mental illnesses is traditional medicine. Many societies developed through history various useful healing methods to fight against a variety of health and life-threatening ailments [1]. The foundations of traditional systems of medicine through ages of years have been formed indeﬁnitely from the plants. The plants thus remain to offer mankind with new medicinal options [2]. Different forms of traditional medicine if applied successfully will assist in the development of new drugs entities, producing abundant beneﬁcial advantages such as signiﬁcantly lower cost of drugs [3]. There is a humongous prospect for future ﬁndings from plants and various natural products which offers a huge boost in procuring useful data about novel chemical structures and new features of the types of activities related to the development of drugs [4].

The international association for the study of pain deﬁnes pain as a bad sensory and emotional experience related to tissue injury [5]. Pain is a protective mechanism. It occurs when tissues are damaged and there is a natural reaction from the individual to remove pain stimulus. The pain receptors are free nerve endings [6]. The therapeutic actions of nonsteroidal anti-inﬂammatory drugs (NSAIDs) are manifested by their ability to block certain prostaglandins (PGs) synthesis through the cyclooxygenase enzymes (COX-1 and COX-2) inhibition [7]. NSAIDs are used as pain reducers, however, they cause adverse effects such as acidity, gastric lesions, and the use of these drugs has not been successful in all the cases. Hence, a wide range of research is underway to find alternative options for NSAIDs [8].

One of the most well-acclaimed mental disorders is depression which manifests itself through symptoms mentally and the psychological state of mind of a person. A person suffering from depression is indiﬁdent, the energy level is down, thoughts and activity are slowed down, and is accompanied with a mixed feeling of sadness and dejection [9]. Antidepressants are the drugs that act in helping in reducing the symptoms of depressive disorders by transforming certain imbalances of chemicals of neurotransmitters in the regions of our brain. The antidepressants are used in the medication several cases which not only include depression but some anxiety disorder, nervousness, obsessive-compulsive disorder, mania-depressive disorders, major depressive disorder, diabetic peripheral neuropathic pain, and post-traumatic stress disorder [10]. One of the ﬁrst drugs used in the treatment of depression is benzodiazepines but they have unwanted effects such as the relaxation of muscle, a sedating effect, addiction, and drug interaction [11].

The genus *Gaultheria* (Ericaceae) is widely distributed around the Paciﬁc Ocean, westwards to western slopes of the Himalayas, and the southern areas of India. Modern research has demonstrated that these plants exhibit anti-inﬂammatory, antioxidative, antibacterial, and analgesic activities. The occurrence of *Gaultheria fragrantissima* in Nigiriri, Travancorens, and Toungo hills in Burma, Ceylon, and Nepal to Bhutan at an altitude of 6,000–8,000 ft. Assam and also in Khali hills, Meghalaya and the Western Ghats are being reported [12].

There are 19 ﬂavonoids, 10 lignans, and even simple phenylpropanoids, steroids, and terpenoids in the genus *Gaultheria*. From the aerial parts of *G. yunnanensis* it displays signiﬁcant analgesic and anti-inﬂammatory activities, which has impelled several studies on the...
phytochemistry and biology of the genus Gaultheria [13]. In the present study, G. fragrantissima Wall. has been selected as it is locally used as a medicinal plant. However, scientific data regarding the use of G. fragrantissima Wall. for analgesic and antidepressant activity are not available for plant leaves. Hence, the present study has been chosen to evaluate the in vivo analgesic and antidepressant activity.

METHODS

Collection of plant and authentication
The leaves of G. fragrantissima Wall. were collected in February 2019 from Mawawwa, Mylliem Village, East Khasi Hills, Meghalaya. The plant was taxonomically identified and authenticated by Botanical Survey of India (BSI), Shillong. The identification number given was No. BSI/ERC/Tech/2019-20/26.

Chemical and reagents
The standard drug diclofenac was obtained from Merck. All the other chemicals were analytical standard.

Preparation of the extract
The collected plants were dried under shade and the dried leaves were pulverized to get a powder leaves and were extracted by Soxhlet apparatus with petroleum ether followed by chloroform then 95% methanol. The extraction was carried out exhaustively and the solvents were recovered. The concentrated extracts were kept in the refrigerator at 4°C for further use.

Phytochemical screening
Phytochemical screening of the methanol extract of G. fragrantissima Wall. was carried out by employing standard procedures and tests to detect the presence or absence of chemical constituents such as glycosides, saponins, alkaloids, triterpenoids, carbohydrates, protein and amino acids, steroids, tannins, phenolics, volatile oil, flavonoids, and pectins [14].

Animal care and handling
Both male and female Wistar rats (150–200 g) used in the study were obtained from the inbred colony and are acclimatized at least for 7 days at the Animal House Facility of Department of Pharmacy, Regional Institute of Paramedical and Nursing Sciences, Mizoram, India. Animals were kept under standard laboratory condition, that is, temperature (25±2°C), humidity (55–60%), and 12 h light and dark cycles were maintained. Animals were allowed to free access to food and water. All animal experiments were carried out according to NIH, USA, and Indian National Science Academy, New Delhi, India, guidelines. Before the experiment, the Institutional Animal Ethics Committees approval was taken with the reference No. IAEC/RIPANS/56.

Acute toxicity studies
Acute oral toxicity was performed as per OECD guideline 425. Two groups of animals were used for the study. Group I served as control and received distilled water. Group II received a single oral dose of G. fragrantissima (2000 mg/kg). The animals were observed for gross behavioral, neurological, autonomic, and toxic effects at short intervals of time for 24 h and then daily for 14 days for any sign of delayed toxicity [15].

In vivo analgesic methods
Hot plate method
This test consists of introducing a rat into an open-ended cylindrical space with a floor consisting of a metallic plate which is heated at a constant temperature. This produces two behavioral components, which are measured in terms of their reaction times, namely, paw licking and jumping. These responses are considered to be supraspinally integrated. Rats of either sex were divided into four groups, each consisting of five animals.

All animals fasted overnight. Two groups were given different doses of the plant extract (determined based on acute toxicity, p.o.), while one group was given a vehicle (control, p.o.) and diclofenac sodium (100 mg/kg) was used, respectively, as standard [16,17]. Then, 1 h after the orally administration, rats were placed onto a hot plate (55±2°C), and the reaction time for licking of paw or jumping for the control and treated mice was recorded (in seconds). Before the treatment, the reaction time of each animal was recorded. The latency to lick the paw or jump from the hot plate was noted as the reaction time. The reaction times were noted at 30, 60, 90, 120, and 180 min. The cutoff time was considered as 15 s [19].

Tail immersion test
A tail immersion test was used to assess the analgesic activity of G. fragrantissima. In this method, five rats per group were used. Tail immersion method involved immersing the extreme 3 cm of the rat’s tail in a glass beaker containing water at a temperature of 55.0±0.5°C. After immersing within a few minutes, the rat reacted by withdrawing the tail. The reaction time was noted on a stopwatch. The test groups were given a methanolic extract of G. fragrantissima (400 mg/kg and 200 mg/kg, p.o.) and diclofenac sodium (100 mg/kg, p.o) was used, respectively, as standard and 1 ml/kg saline solution for the control group (p.o.). The reaction time of the test groups was taken at 30, 60, 90, 120, and 180 min after a latency period of 30 min following the administration of the tests substances. The cutoff time, that is, time of no response was put at 12 s. The reaction time was measured and calculated [19].

In vivo antidepressant
Force swimming test
The method used was similar to that described by Porsolt et al., animals were randomly divided into four groups. Thirty minutes after treatment with the methanol extract of G. fragrantissima and imipramine (a reference) antidepressant drug, control is normal saline. Rats were individually placed in an open cylindrical container (35 cm height × 24 cm diameter) containing 13.5 cm of water at 22–25°C for 6 min. The duration of immobility was scored during the last 4 min of the 6 min test period. Rats were recorded as immobile when floating motionless or making only those movements necessary to keep the head above water. A decrease in the duration of immobility during the forced swimming test was taken as a measure of antidepressant activity [20].

Tail suspension test (TST)
The total duration of immobility induced by tail suspension was measured according to the method of Steru et al. Animals were randomly divided into four groups. Thirty minutes after treatment with the methanol extract of G. fragrantissima and imipramine (a reference) antidepressant drug, control is normal saline. Rats were suspended 50 cm above the floor by adhesive tape placed approximately 1 cm from the tip of the tail. Immobility time was recorded during a 6 min test. A decrease in the duration of immobility during the forced swimming test was taken as a measure of antidepressant activity [21].

Statistical analysis
All values were shown as Mean±SEM (n=4). Statistical analysis was performed using one-way analysis of variance (ANOVA) followed by Tukey-Kramer multiple test. p<0.05 was considered statistically significant.*p<0.05 and **p<0.01 when compared with control. All analyses were made with the statistical software GraphPad InStat.

RESULTS

Result of phytochemical screening
Table 1 shows the results of the phytochemical screening of the methanolic extract of G. fragrantissima.

The tests identify the presence of phenols and tannins, flavonoid, steroid and triterpenoid, saponin, carbohydrates, volatile oil, and pectin in methanol extract of G. fragrantissima.

Acute toxicity test
Since extract of the plant was already used as traditional medicine and there was no sign of toxicity from its consumption, an acute toxicity test was performed using one-way analysis of variance (ANOVA) followed by Tukey-Kramer multiple test. p<0.05 was considered statistically significant.*p<0.05 and **p<0.01 when compared with control.
was performed according to the limit test of OECD 425 guidelines. It was found that at the dose of 2000 mg/kg body weight did not produce any mortality to the subjected test animal (female albino rats). The animals were observed for 14 days after dosing and no clinical and behavioral changes were observed during this period.

In vivo analgesic activity
Hot plate method
The results of the analgesic activity of the methanolic extract of G. fragrantissima extract using the hot plate method are presented in Fig. 1.

There was no significant difference in the thermal stimulus in mice treated with the vehicle (control) throughout the whole time of the experiment. Diclofenac administration significantly increased the response time of the animal to reach 7.83±0.390 (at 120 min) and is considered as extremely significant when compared to the control group (p<0.001). All doses of G. fragrantissima extract showed a significant increase in the latency time of mice when compared to the control group. As shown in Fig. 1, the maximum activities were obtained after 2 h with 7.114±0.469 and 7.538±0.070 s for 200 and 400 mg/kg (G. fragrantissima) and are considered as significant when compared to the control group (p<0.01). Fig. 1 demonstrates the relative activity of G. fragrantissima with respect to the standard drug (diclofenac) in the hot plate method. The analgesic activity in the standard and extracts treated in animals is noted from 30 min onward till 180 min in which response peaked at 120 min for diclofenac sodium and at 120 min for both the extracts at low and high dose. The result shows us that when in comparison, the high dose is almost equivalently potent to the standard diclofenac sodium, though standard is more potent than the low dose.

Table 1: Results of preliminary phytochemical tests

| S. No. | Phytochemical tests | Methanol extract |
|--------|---------------------|-----------------|
| 1.     | Alkaloids           | -               |
| 2.     | Glycosides          | +               |
| 3.     | Phenols and tannins | +               |
| 4.     | Flavonoids          | +               |
| 5.     | Steroids and triterpenoids | + |
| 6.     | Saponins            | +               |
| 7.     | Carbohydrates       | +               |
| 8.     | Protein and amino acid | +  |
| 9.     | Fats and fixed oils | -               |
| 10.    | Volatile oil        | +               |
| 11.    | Pectin              | +               |

(+) indicates the presence of the tested group, (−) indicates the absence of the tested group

Fig. 1: Analgesic activity of GFE by hot plate method in rats. Values are expressed in Mean±SEM, n=5. ***p<0.001 considered extremely significant when compared to the control group. *p<0.05 compared with control. **p<0.01 compared with control. +++p<0.001 compared with control. Data were analyzed using one-way ANOVA followed by Tukey-Kramer multiple test.

Tail immersion test
The results of the analgesic activity of the methanolic extract of G. fragrantissima extract using the tail immersion method are presented in Fig. 2.

There was no significant difference in the thermal stimulus in mice treated with the vehicle (control) throughout the whole time of the experiment. Diclofenac administration significantly increased the response time of the animal to reach 4.902±0.2595 s (at 90 min) and is considered as extremely significant when compared to control group (p<0.001). All doses of G. fragrantissima extract showed a significant increase in the latency time of rats when compared to the control group. As shown in Fig. 2, the maximum activities were obtained after 90 min with 4.01±.222 and 2.704±0.2515 s for 400 and 200 mg/kg (G. fragrantissima) and are considered as significant when compared to the control group (p<0.01). Fig. 2 demonstrates the relative activity of G. fragrantissima extract with respect to the standard drug (diclofenac sodium) in the tail immersion method. The analgesic activity in the standard and extracts treated in animals is noted from 30 min onward till 120 min in which response peaked at 90 min for diclofenac sodium and at 90 min for both the extracts at low and high dose. The result shows us that when in comparison, the high dose is almost equivalently potent to the standard diclofenac sodium, though the standard is more potent than the low dose.

Findings from Figs. 1 and 2 demonstrate that the methanol extract at the dose of 400 mg/kg prolonged the reaction time in the hot plate method and the tail immersion method. Taken together sensitivity of both methods as well as the mechanism involved may explain the analgesic effects observed in this study.

In vivo antidepressant activity
The antidepressant effect of the methanolic extract of G. fragrantissima (200 and 400 mg/kg) and imipramine was studied by observing the changes in the duration of immobility in the two models: Forced swim test (FST) and TST. These tests are used to monitor depressive-like behavior and are based on the assumption that immobility reflects a measure of behavioral despair. The methanol extract of G. fragrantissima (200 and 400 mg/kg) produced a significant reduction (p<0.001) in the immobility period when compared with that of control group animals. The immobility time in sec of the high dose of G. fragrantissima is 185±3.661 in FST. The immobility time in sec of the high dose of G. fragrantissima is 100.8±7.116 as compared to the control which is 202±4.247 in the TST as seen in the results tabulated in Figs. 3 and 4.

DISCUSSION

After performing the acute toxicity studies, it was found that at the dose of 2000 mg/kg body weight did not produce any mortality to the
subjected test animal (female albino rats). The animals were observed for 14 days after dosing, and no clinical and behavioral changes were observed during this period. Hence, the LD₅₀ of *G. fragrantissima* extract was more than 2000 mg/kg body weight and was safe for further use.

Results from the analgesic activity studies reveal that the methanol extract of the leaf of *G. fragrantissima* acts as an analgesic agent. Analgesics are drugs that act on the central or peripheral nervous system to relieve pain selectively without altering consciousness. Centrally acting analgesics act by increasing the threshold for pain and altering the physiological response to pain. However, peripherally acting drugs act by inhibiting the generation of pain impulses at the chemoreceptor level [22].

It also revealed that the plant possesses significant analgesic activities as the presence of flavonoids, tannins were confirmed. Flavonoids were reported to have a role in an analgesic activity primarily by targeting PGs [23,24]. There are also reports on the role of tannins in anti-nociceptive activity [25]. The methanolic extracts of the plants in NSAIDs inhibit peripheral pain [26].

The analgesic effect of the plants in both models suggests that they have been acting through a peripheral mechanism. The analgesic activity may be due to its ability to inactivate the pain receptors at the receptor site level. It may also inhibit endogenous pain substances, which are involved in the peripheral analgesia.

The methanol extract of *G. fragrantissima* (200 and 400 mg/kg) produced a significant reduction (p<0.001) in the immobility period when compared with that of control group animals that received only the vehicle in both the FST and TST.

Forced swimming and TSTs which represent the behavioral despair model claimed to reproduce a condition similar to human depression [27]. The tests are based on the observation that animals, following initial escape oriented movements, develop an immobile posture when placed in an inescapable chamber. The immobility is thought to reflect either a failure of persistence in escape-directed behavior (i.e., behavioral despair) or the development of passive behavior that disengages the animal from active forms of coping with stressful stimuli [28].

It is well known that clinically effective antidepressants (such as imipramine) typically increase the swimming efforts of the animal seeking a solution to the problem and, therefore, decrease the duration of immobility in the forced swimming test and this was observed in this study. Phytochemical analysis showed that the presence of flavonoids and phenolic compounds has been reported to have multiple biological effects such as central nervous system disorders [29].

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