The effect of zinc supplementation on pro-inflammatory cytokines (TNF-α, IL-1 AND IL-6) in mice with Escherichia coli LPS-induced diarrhea

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ABSTRACT

Background and Objectives: Inflammation in the intestine causes diarrhea due to an increased release of pro-inflammatory cytokines such as TNF-α, IL-1, and IL-6. These are triggered by the exposure of E. coli-LPS to epithelial cells of the intestinal mucosa as well as low concentration of zinc in plasma such as in infants or children who are experiencing diarrhea. This paper aims to determine the effects of zinc supplementation on pro-inflammatory cytokines (TNF-α, IL-1 and IL-6) in mice with E. coli-LPS-induced diarrhea.

Materials and Methods: This study used a controlled trial experimental design in the laboratory. A sample size of 20 mice were randomly divided into 4 groups: 1) Control group was given standard foods, 2) Trial group was given E. coli-LPS 2.5 mg/kg/oral once on day1, 3) Prevention group was given E. coli-LPS + 30 mg/kg/oral of zinc once daily for 12 days, 4) Therapeutic group was given E. coli-LPS, and were then given 30 mg/kg/oral of zinc once daily for 12 days if diarrhea occurred. Blood samples of mice were taken through the orbital sinus on the 0, 5th, 10th hour, and on the 4th, 8th and 12th days.

Results: Positive effects of zinc supplementation on levels of pro-inflammatory cytokines were observed, in which the higher levels of zinc were present, the lower levels of pro-inflammatory cytokines, especially TNF-α were observed. However, there was an increase of IL-1 and IL-6 levels on the 8th day in the prevention and therapeutic groups.

Conclusion: Oral zinc supplementation had a significant positive effect on the levels of pro-inflammatory cytokines. Where there were higher levels of zinc, lower levels of pro-inflammatory cytokines TNF-α were present.

Keywords: Zinc; Tumor necrosis factor-α; Interleukin 1; Interleukin 6; Diarrhea

INTRODUCTION

Zinc deficiency is still common, especially in developing countries. This can be related to lack of intake, increased needs, and the loss of zinc due to diseases, especially infection. There is an associ-
EFFECT OF ZINC PRO-INFLAMMATORY CYTOKINES

atation between infection and zinc deficiency which influence each other. Zinc requirements of the body will increase during infection, formation of immune functions, and new cells. Meanwhile, zinc deficiency can cause suppression of immune function, making it easier for infection to occur (1, 2).

Zinc plays a role in maintaining the integrity of the intestinal mucosa through its function in cell regeneration and membrane stability. Furthermore, Zinc has a direct impact on intestinal villi, disaccharidase brush border activity, intestinal water, and electrolyte transport. Zinc also plays a role in T cell function and enhances immunity that can reduce the severity of diarrhea (3).

Escherichia coli, despite being the main occupant of healthy colon flora, can also cause various diseases such as diarrhea by releasing endotoxins, which trigger the release of pro-inflammatory mediators. Lipopolysaccharide (LPS) is the main component of the cell wall of Gram-negative bacteria which is also called endotoxin and is known to be the trigger of several types of inflammatory or infectious reactions in macrophage cells and other cells that have CD14 receptors. In fact, the endotoxin (LPS) of bacteria that binds to the Toll Like Receptor (TLR) in Dendritic Cells (DC) can stimulate monocyte and macrophage cells to secrete nitric oxide (NO) and inflammatory substances (cytokines) such as Tumor Necrosis Factor-alpha (TNF-α) and interleukin-1-beta (IL-1-β), IL-6, and IL-8 (4). As a result, excessive pro-inflammatory cytokines can cause symptoms of decreased blood pressure, fever, and diarrhea (5).

In experimental studies, zinc deficiency has proven to have a direct effect on the digestive tract, by forming villous atrophy, decreasing disaccharidase enzyme activity in brush borders, and impairing intestinal transport. However, the exact mechanism that explains the pathophysiology of diarrhea due to zinc deficiency has not been agreed upon yet. Despite the uncertain mechanism, there has been proof that incidence of persistent diarrhea could be reduced by zinc supplementation and administration of oral rehydration solutions (ORS). Zinc has substantially reduced the duration and severity of diarrhea in children with acute and persistent diarrhea (6). Although the mechanism of how zinc supplementation can reduce diarrhea is still unknown, zinc therapy in diarrhea patients has shown better absorption of water and electrolytes by the intestine, faster regeneration of intestinal epithelial cells, increased levels of enzymes from the brush border, and increased immune response that can eliminates pathogens in the intestine (7). Thus, the purpose of this study was to determine the effect of zinc supplementation on pro-inflammatory cytokine (TNF-α, IL-1 and IL-6) in mice with E. coli-LPS -induced diarrhea.

MATERIALS AND METHODS

This study deployed a Controlled Trial Design (8) and was conducted at The Laboratory of Nutrition and Food Centre (PSPG) at Gadjah Mada University Yogyakarta. The sample size was 20 white mice Sprague Dawley which were randomly chosen and randomly divided into 4 groups, with each group consisting of 5 mice. The 1st group (G1) or the Control Group, was given standard nutrition. The 2nd group, the Trial Group (G2), was given 2,5 mg/kg/oral of E. coli-LPS for three days, from day 1 to day 3. The 3rd group, the Prevention Group (G3) was given 2,5 mg/kg/oral of E. coli-LPS on day 1 and also 30 mg/kg/oral of zinc from day 1 until Day 12, and the 4th group, the Therapeutic Group (G4), was given 2,5 mg/kg/oral of E. coli-LPS on day 1. When diarrhea occurred, the mice in G4 were given 30 mg/kg/oral of zinc from the 1st until the 12th day. A 2ml sample of the mice blood was taken from sinus orbital with ELISA test technique (9) at the baseline, on the 5th, and 10th hour, and on the 4th, 8th, and 12th day to analyze zinc and pro-inflammatory cytokines (TNF-α, IL-1, IL-6) level in its serum.

RESULTS

The characteristics of subjects in this study including weight and Hemoglobin level (Hb) are presented in Table 1.

The level of zinc based on treatment time. The results of zinc level can be seen in Fig 1. In the control group (G1), we observed that there was no difference in zinc level based on treatment time in 5 mice, with the level ranging from 1, 14 to 1, 2 µg/dl. However, zinc level in the trial group (G2) seemed to be lower compared to group G1, G3 and G4 which could be seen in Fig. 1. Starting from the 4th, 8th and 12th day of treatment, the 3rd, 4th and 5th mice in group G2 experienced diarrhea, respectively.
The difference could also be seen in the prevention group (G3), where zinc level increased at the 5th and 10th hour after being given 0.94 µg/dl and 0.77 µg/dl LPS, respectively. Meanwhile on the 4th, 8th and 12th day, zinc level was higher compared to group G2, as it can be seen in Fig. 1. This event was observed in the 3rd, 4th and 5th mice in group G3, despite that fact that all mice in this group were given zinc every day.

The same finding was observed in the therapeutic group (G4), in which zinc levels were higher in the 3rd, 4th and 5th mice on the 4th, 8th and 12th day. Actually, zinc level was observed higher in the 3rd mouse of group G3, 3 days after receiving zinc. The same happened for the 4th mouse in G4, in which zinc level was observed to be higher on the 6th day after receiving zinc. Meanwhile, zinc level was higher after 10 days of receiving zinc on the 5th mouse in G4, as it is shown in Fig. 1.

Zinc level in trial group (G2) was lower than control group (G1) on all the observation points (5 & 10 hours, 4, 8 and 12 days after being given LPS) as it was shown in Fig. 1. Moreover, zinc levels were also lower in the Trial group (G2) compared to the Prevention (G3) and the Therapeutic group (G4) especially on the 4th, 8th, and 12th day. This might be because the mice in the Prevention and Therapeutic groups were given zinc every day, while the mice in trial group (G2) were not.

**Level of TNF-α based on treatment duration.**

The results of TNF-α levels examination were shown in Fig. 2. In the trial group (G2), a higher level of TNF-α was observed, starting from the 5th & 10th hour, and the 4th, 8th, and 12th day compared to the other three groups (G1, G3, and G4). The TNF-α levels in G3 and G4 were noticeably lower than in G2 on the 4th day. The TNF-α levels in G3 and G4 became higher on 8th, and 12th day, but were still not as high as in G2. This finding was interesting because zinc was administered daily for mice in G3 and G4, but not for mice in G2.

The TNF-α level in G2 was higher at the 5th hour after being given LPS compared to G1, G3, and G4. The TNF-α level was higher at the 10th hour after being...
ing given LPS in G3, and G4 compared to the level on the 5th hour in respective groups, as shown in Fig. 2.

**The level of IL-1 based on treatment duration.**
In this study the level of IL-1 in the control group (G1) was found to be higher on the 8th day of mice (G1) 4 compared to mice (G1) 1, (G1) 2, (G1) 3, and (G1) 5, however, diarrhea did not occur in all mice in this group. In the trial group (G2), IL-1 levels were higher at the 5th and 10th hour after being given LPS before it went lower on the 4th day in mice (G2) 3. However, subsequently higher IL-1 levels were observed on 8th and 12th day in mice (G2) 4 and (G2) 5, as shown in Fig. 3. In this group, three out of five mice (mice (G2) 3, (G2) 4, and (G2) 5) started to have diarrhea from the 3rd day after given LPS.

From Fig. 3, it could also be seen that IL-1 levels in the therapy group (G4) were the highest among all the groups (16.33 ng/ml) at the 5th hour after given LPS, but then declined to its lowest level at the 10th hour. Interestingly, the IL-1 level then increased again on the 4th and 8th day (its peak) to just decline again on the 12th day (12.30 ng/ml only).

**The level of IL-6 based on treatment duration.**
The IL-6 level in this study was observed and is shown in Fig. 4. In the control group (G1) of the 5 experimental mice, a very high IL-6 level (5.70 pg/ml) was observed on the 12th day in mouse (G1) 5, although the mouse did not experience diarrhea. However in the trial group (G2), IL-6 level was higher at the 5th and 10th hour after given LPS compared to the control group (G1), which was 4.86 pg/ml and 4.10 pg/ml respectively. However, the IL-6 level in this group (G2) decreased on the 4th and 8th day in mouse (G2) 3 and (G2) 4, further higher than 4.22 pg/ml on the 12th day of mice (G2) 5, shown in Fig. 4.

As shown in Fig. 4, the IL-6 level in the prevention group (G3) showed an increase at the 5th hour after being given LPS. The level of IL-6 were almost the same as IL-6 levels in the control group (G1) which was 1.74 pg/ml, higher than twice from the previous value at the 10th hour after treatment from mice (G3) 2. Meanwhile, in the therapeutic group (G4), the level of IL-6 increased 5 hours after treatment (2.67 pg/ml), but then declined again down to 1.60 pg/ml on the 4th day from mice (G4) 3.
DISCUSSION

Pro-inflammatory cytokine TNF-α, IL-1 and IL-6. Existing literatures mention that the expression of pro-inflammatory cytokines increase 90 minutes after infection, reach their peak in 3 hours and return to normal 6 hours after infection. Meanwhile, the secretion of pro-inflammatory cytokines reach its maximum level in 4-6 hours after infection and return to their normal level 12 hours later (10). In this study, TNF-α level in trial group (G2) was the highest at every measurement point. Despite a small decrease on the 4th day, its level was still the highest among all groups as shown in Fig. 2. Furthermore, the IL-1 level in trial group (G2) was also observed to be higher compared to the control group (G1) and the prevention group (G3). Although the IL-1 level in Group 2 was higher than in the therapeutic group (G4) at the 10th hour, 4th day, and 12th day, however, the level of IL-1 in G4 was significantly higher than in G2 on the 5th hour, and 8th day as seen in Fig. 3. Although zinc can influence the production and signaling of numerous inflammatory cytokines in various cell types, zinc supplementation is not always found beneficial. Adding zinc at very high concentrations (>100 µM) has been suggested to increase cytokine production in some cell types, as has been observed in human peripheral blood mononuclear cells (PB-MCs) harvested from healthy adults and in human promonocytic HL-CZ cells (11).

In this study, the levels of TNF-α, IL-1, and IL-6 were seen higher in the trial group (G2) compared to the control group (G1) as seen in Figs. 1-3. The levels of IL-1 and IL-6 were particularly high on the 8th day in all experimental groups. Zinc deficiency, as occurred in G2, causes decreased cellular immunity which will induce the inflammatory process, characterized by an increase in the expression of pro-inflammatory cytokines in response to the exposure reaction of an antigen, as shown in this study. Free intracellular zinc acts as a suppressor of cytokine production by stimulating the immune cells. In contrast, zinc positively regulates the expression of cytokines. Zinc supplementation enhances cytokine production in immune cells. These conflicting effects may be due to the concentration-dependent effect of zinc on different signalling molecules involved in cytokine expression (12).

Tumor Necrosis Factor (TNF) is the main cytokine in the acute inflammatory response to Gram-negative bacteria and other microbes. Severe infection can trigger production of large amounts of TNF which can cause systemic reactions (4). According to Prasad, 2008, there was an increase in the release of pro-inflammatory cytokines such as TNF-α and IL-1β by monocytes and macrophages which were activated by the inflammatory process and were associated with decreased zinc status in patients. This is in accordance with the finding in this study, which showed an increase in TNF-α levels in the trial group (G2) compared to the control group (G1) seen in 2 (13). In the condition of diarrhea, zinc deficiency occurs and might reduce the production and activity of SOD enzymes, subsequently increasing the activity of free radicals that can also cause inflammatory reactions in the intestinal mucosa, which would trigger an increase of TNF-α by competent immune cells. This condition is not ideal, as a high concentration of TNF-α will damage the tight junction in enterocyte cells of the intestinal mucosa (14).

Increased levels of IL-1 and IL-6 in the diarrhea group compared to the control group in this study were due to the presence of microbes that release endotoxins and/or exotoxins, both of which promote the release of pro-inflammatory mediators. LPS is a component of the cell wall of Gram-negative bacte-
Zinc also increases the formation of ADP ribosylation in intestinal mucosal epithelial cells. Furthermore, superoxide anions inhibiting the apoptosis process acts as the main antioxidant enzyme that controls radicals by increasing the formation of SOD, which means it can also control the immune response to free radicals, a process made possible by its ability to inhibit the formation of free radicals. The occurrence of intestinal mucosal inflammation is proven by the presence of faecal TNF-α and based on the role of TNF-α. It is suggested that TNF-α is responsible for damage to the intestinal mucosa because this cytokine is pleotropic, meaning it can stimulate inflammation and signal death/cell apoptosis (17).

Zinc deficiency has been associated with increase in inflammatory cytokines and other markers of inflammation. However, during the time this study was carried out, the precise mechanisms that explain how zinc deficiency contributes to increased inflammation remains a topic that can be clarified through further study (18).

Zinc is also able to stimulate the production of tumor necrosis factor-alpha (TNF-α) by monocyte cells, so that the ability of phagocytosis increases. TNF-α is a mediator for non-specific immune responses and belongs to the cytokine group. Helge and Rink reported that in vitro mononuclear cell incubation in the zinc medium can increase the production of interleukin 1, interleukin 6, tumor necrosis factor (TNF), IL-2R, and interferon (19).

Monocytes in deficient mice fail to kill intracellular parasites. The research conducted by Bires reported that phagocytic activity increased after the administration of zinc, showing an increase in the number of monocytes by 14% and granulocytes by 86% (20). Furthermore, Linder also suggested that insufficiency or excess zinc minerals can cause damage to the immune system components (21).

Cytokines are directly involved in regulating permeability and ion response in the epithelium and TNF-α has a direct effect on almost all enterocyte cell functions including transportation, permeability, regeneration, and proliferation. With the administration of zinc, inflammation of the intestinal mucosa can be reduced as proven by the decrease in serum TNF-α levels by inhibiting the production of TNF-α and IL-6. TNF-α plays an important role in the mechanism of diarrhea in zinc deficiency, while serum TNF-α level can be used as a sign of inflammation that triggers the production of cytokines (22, 23).

In conclusion, zinc supplementation has a positive effect on reducing pro-inflammatory cytokines level. The higher level of zinc is, the lower level of pro-inflammatory cytokines will be, particularly for
TNF-α. However, it is important to notice the increase of IL-1 and IL-6 levels on the 8th day in the prevention and therapeutic groups in mice with E. coli-LPS-induced diarrhea.

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