Letter to the Editor
Clinical Microbiology

The First Case of Clostridium saudiense Bacteremia in a Patient with Hepatocellular Carcinoma

Eungjun Yoon a, M.D. 1, Tae Yeul Kim a, M.D. 1, Won Young Heo a, M.D. 1, Onkyun Kang a, M.T. 1, Hui-Jin Yu a, M.D. 1, Joon Hyeok Lee a, M.D., Ph.D. 2, Jae-Hoon Ko a, M.D., Ph.D. 3, Nam Yong Lee a, M.D., Ph.D. 1, and Hee Jae Huh a, M.D., Ph.D. 1

1Department of Laboratory Medicine and Genetics, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea; 2Department of Medicine, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea; 3Division of Infectious Diseases, Department of Medicine, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea

Dear Editor,

Clostridium saudiense (formerly known as C. saudii) is a spore-forming, anaerobic, gram-positive bacillus, first identified by Angelakis, et al. [1] from the stool of an obese 24-year-old Saudi individual and named C. saudii. As C. saudii was not a validly published name, the bacterium was later renamed C. saudiense [2]. Extensive culturomics studies of stool samples from healthy individuals have indicated that C. saudiense is a commensal bacterium colonizing the human intestinal tract [3-5]. To date, no apparent infections caused by C. saudiense have been reported. We report the first case of C. saudiense bacteremia. The Institutional Review Board (IRB) of Samsung Medical Center, Seoul, Korea (IRB number SMC 2021-08-077), approved this study and waived the need for informed consent as this study poses minimum risk to the patient.

A 49-year-old male patient with hepatocellular carcinoma visited the Samsung Medical Center emergency room complaining of persistent fever and abdominal discomfort. He had undergone a transcatheter arterial chemoembolization (TACE) procedure one month prior. His temperature was 38.4°C, blood pressure 98/65 mmHg, pulse rate 99 beats/min, and respiratory rate 20 breaths/min. Laboratory parameters were as follows: leukocyte count, 14.76x10^9/L (neutrophils, 78.8%) and C-reactive protein (CRP) level, 150.5 mg/L (reference interval, 0.0–5.0 mg/L). Abdominopelvic computed tomography revealed ischemic biliopathy and intrahepatic retention of bile, forming a biloma (Fig. 1).

Two sets of blood samples from peripheral veins were collected in BACT/ALERT FA/FN PLUS bottles (bioMérieux, Marcy-l’Étoile, France) and incubated in the BACT/ALERT VIRTUO system (bioMérieux). The patient was subjected to empirical antibiotic therapy with ceftriaxone 2,000 mg intravenously (IV) every 24 hours and metronidazole IV 500 mg every 8 hours on day 1. The next day, the antibiotic was switched to piperacillin/tazobactam IV 3,375 mg every 6 hours owing to persistent fever. On day 3 of incubation, positive growth was observed in one anaerobic bottle of the two sets of blood cultures. Gram staining revealed gram-variable, rod-shaped bacteria of various lengths and occasional endospores (Fig. 2). The positive blood culture was plated onto blood agar, MacConkey agar, and Brucella agar for subculture. Growth was observed only on Brucella agar under anaerobic conditions. Bacterial identification using matrix-assisted laser desorption ionization time-of-flight mass spectrometry (VITEK MS; bioMérieux) was unsuccessful. The VITEK 2 system
using the ANC card (bioMérieux) identified the isolate as *Clostridium clostridiolforme* with a probability of 88%, failing precise identification.

For accurate identification, we performed 16S ribosomal RNA gene sequence analysis as described previously [6]. The sequence obtained was compared with those in the GenBank database using the basic local alignment search tool (https://blast.ncbi.nlm.nih.gov/Blast.cgi). The sequence matched that of *C. saudiense* JCC (GenBank accession number NR_144696.1) perfectly (100.0%, 1,411/1,411 bp). The second-best match was *Clostridium celatum*, with 98.9% sequence identity (1,397/1,412 bp). Based on these findings, the isolate was identified as *C. saudiense* [7].

The patient was treated with piperacillin/tazobactam IV 3,375 mg every 6 hours for two weeks followed by an additional four weeks of oral metronidazole 250 mg every 8 hours and oral ceffixime 300 mg every 24 hours. *C. saudiense* was re-cultured from bile samples obtained via percutaneous drainage and two sets of additional blood cultures on day 3. Follow-up blood cultures on days 5 and 7 were negative, and the patient’s CRP level started to decline. After scheduled antibiotic therapy, the patient showed no symptoms or signs of infection and the CRP level normalized.

*Clostridium* species are widespread in nature, and some are present in the human intestinal tract, female genital tract, and oral mucosa as commensals [8]. *Clostridium* species are the second most common anaerobes isolated from blood [9], especially in patients with predisposing factors, such as gastrointestinal anomalies or immunocompromise [8, 10]. However, *C. saudiense* infections, particularly bacteremia, have not been reported. In our case, commensal *C. saudiense* translocated from the gastrointestinal system to biliary system led to a loculated biliary and eventually developed into a bloodstream infection. Like in other cases of clostridial bacteremia, the patient had a predisposing factor, namely a recent intervention history (TACE). The patient responded well to empirical treatment for *Clostridium* species.

In conclusion, we report the first case of *C. saudiense* bacteremia. Our findings suggest that *C. saudiense* be considered an opportunistic pathogen with the potential to cause biliary infection and bacteremia in patients with damage to the biliary system.

**ACKNOWLEDGEMENTS**

None.

**AUTHOR CONTRIBUTIONS**

Huh HJ designed the study. Heo WY, Kang OK, Kim TY, and Yu HJ analyzed the data. Yoon EJ drafted the manuscript. Lee JH, Ko JH, Huh HJ, and Lee NY reviewed the manuscript. Submission of the final manuscript was approved by all authors.
CONFLICTS OF INTEREST

None declared.

FUNDING

None declared.

ORCID

Eungjun Yoon https://orcid.org/0000-0002-6210-7290
Tae Yeul Kim https://orcid.org/0000-0002-4605-5305
Won Young Heo https://orcid.org/0000-0003-2695-9634
Onkyun Kang https://orcid.org/0000-0002-1031-1991
Hui-Jin Yu https://orcid.org/0000-0002-7064-1968
Joon Hyeok Lee https://orcid.org/0000-0003-3547-7434
Jae-Hoon Ko https://orcid.org/0000-0002-9490-6609
Nam Yong Lee https://orcid.org/0000-0003-3688-0145
Hee Jae Huh https://orcid.org/0000-0001-8999-7561

REFERENCES

1. Angelakis E, Bibi F, Ramasamy D, Azhar EI, Jiman-Fatani AA, Aboushoushah SM, et al. Non-contiguous finished genome sequence and description of Clostridium saudii sp. nov. Stand Genomic Sci 2014;9:8.
2. Oren A and Garrity GM. List of new names and new combinations previously effectively, but not validly, published. Int J Syst Evol Microbiol 2016;66:2463-6.
3. Diakite A, Dubourg G, Dione N, Afouda P, Bellali S, Ngom II, et al. Extensive culturomics of 8 healthy samples enhances metagenomics efficiency. PLoS One 2019;14:e0223543.
4. Lagier JC, Hugon P, Khelafia S, Fournier PE, La Scola B, Raoult D. The rebirth of culture in microbiology through the example of culturomics to study human gut microbiota. Clin Microbiol Rev 2015;28:237-64.
5. Dubourg G, Baron S, Cadoret F, Couderc C, Fournier PE, Lagier JC, et al. From culturomics to clinical microbiology and forward. Emerg Infect Dis 2018;24:1683-90.
6. Park JH, Kim TS, Park H. A Hungatella effluvii isolate in blood culture of a patient with hematochezia. Anaerobe 2020;63:102211.
7. CLSI. Interpretive criteria for identification of bacteria and fungi by targeted DNA sequencing. 2nd ed. CLSI MM18. Wayne, PA: Clinical Laboratory Standards Institute, 2018.
8. Jorgensen J, Pfaller M, et al. eds. Manual of clinical microbiology. 11th ed. Washington DC: American Society of Microbiology, 2015:940-66.
9. Simmon KE, Mirrett S, Reiler LB, Petti CA. Genotypic diversity of anaerobic isolates from bloodstream infections. J Clin Microbiol 2008;46:1596-601.
10. Brook I. The role of anaerobic bacteria in bacteremia. Anaerobe 2010;16:183-9.