miRNA-31 over-expression improve synovial cells apoptosis induced by RA

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ABSTRACT

OBJECTIVE: The aim of this study was to evaluate the effects and mechanism of miRNA-31 in synovial cells apoptosis induced by RA.

METHODS: The miRNA-31 gene expressions were extracted from synovial tissues of normal and RA patients by RT-PCR and H & E staining. The synovial cells of RA patients were isolated and randomly divided into Control, Blank and miRNA groups. The cell apoptosis of difference groups were measured by flow cytometry; the TNF-α and IL-1β concentrations of difference groups were measured by Elisa assay; TLR4 and NF-κB proteins expressions were measured by WB assay and the correlation between TLR4 and miRNA-31 were evaluated by double luciferase target experiment.

RESULTS: The miRNA-31 gene expression was significantly suppressed in RA tissues (p<0.001); Compared with control group, the cell apoptosis rate of miRNA group was significantly suppressed (p<0.001); TNF-α and IL-1β concentrations were significantly down-regulation in culture fluid (p<0.001, respectively) and TLR4 and NF-κB proteins expressions were significantly depressed (p<0.001, respectively) in miRNA group. By double luciferase target experiment, the TLR4 was a target gene of miRNA-31.

CONCLUSION: miRNA-31 is a key role in synovial cells apoptosis induced by RA (Fig. 7, Ref. 23). Text in PDF www.elis.sk.

KEY WORDS: miRNA-31, cell apoptosis, RA; TLR4, NF-κB.
Each sample was set up with 3 complex holes for quantitative detection. The reaction takes U6 as the internal reference. The CT value of each hole was recorded, the average value of each hole was taken as the result, and the results were analyzed by 2-ΔΔCT method.

**H & E staining**

The synovial tissues were fixed in the 10% polyoxymethylene for 24 h, Paraffin embedded, made into 5 μm slices, the sections were H & E staining, observation pathological changes under optical microscope.

**RA separation and culture of synovial cells**

The synovial tissues of the knee joint were removed in a sterile environment, washing by PBS, the synovial tissues were cut, added 2 ml DMEM culture fluid and 2 ml I type collagenase (2.5 mg/ml) to culture in incubator (37 °C, 5% CO2) for 4–6 h, filtering, centrifugal, discarding the supernatant. Adding DMEM culture solution to resuspension cell, the cells as 1×10^6 cells/ml were inoculated in the cell culture bottle. The cells were cultured in the incubator (37 °C, 5% CO2).

**Grouping and treatment**

The synovial cells were randomly divided into control group which were treated with normal treatment; Blank group which were transferred with empty vector and miRNA group which were transferred with miRNA-31. The synovial cells were taken and inoculation in 6-hole culture dish, there were 1×10^6 cells/ml in every holes. When the cell density was 80–90%, the cells of Blank and miRNA groups were transferred with Lipofectamine™ 2000 (Sigma, USA) or miRNA-31 (Sigma, USA) following by Supplier Agreement.

**MTT assay**

After transfection for 72 h, the logarithmic growth period synovial cells of difference groups were collected and washed by PBS, added 0.25 % trypsin to digest, centrifugal, made as single cell suspension, regulation cell suspension as 1×10^6 cells/ml, Inoculated in the 96-hole plate, culture overnight, added 20 μl MTT (5mg/ml) and cultured at 37 °C for 4 h, removing the supernatant, added 100 μl DMSO to every holes. After shocking for 10 min, the absorbance value were measured at 570 nm, and measured the cell proliferation.

**Cell apoptosis by flow cytometry**

After transfection for 48 h, the cells of difference groups were collected and washed by PBS for 2 times. Adding 100 μl Binding buffer to resuspension cells, after that, added 5 μl Annexin V and 1 μl PI to culture for 15 min in dark, added 400 μl Binding buffer to measure the cell apoptosis by flow cytometry, using FlowJo 7.6 software to analysis apoptosis.

**TNF-α and IL-1β concentrations by Elisa assay**

After treatment for 48 h, the culture fluid of difference groups were collected and centrifugal to collect supernatant. The TNF-α and IL-1β concentrations were measured by Elisa kits following by supplier agreement.

**The relative proteins expression by WB assay**

The total protein was extracted by total protein extraction kit (Nanjing KeyGen Biotech Co., Ltd., China), the total protein concentration was measured by BCA method (Nanjing KeyGen Biotech Co., Ltd., China), the 20 μg total protein was taken to electrophoretic separation by 10% SDS-PAGE, transferred the protein to PVDF member, adding the first antibodies (GAPDH 1:1000; TLR4 1:1000 and NF-κB 1:500) to culture overnight at 4 °C, washing by PBS, added HRP marked second antibody (1:1000) to culture for 1 h, adding enhanced chemiluminescence (ECL) to color, using Chemiluminescent imaging system (TANON 5200) to analysis the gray value by Imagel software.

**Double luciferase reporter gene experiment**

The TLR4-UTR sequence and its mutants containing the miRNA-31 binding site were inserted into the downstream of the luciferase reporter gene of the pmirGLO vector. The wild type and mutants obtained were named as TLR 4-WT and TLR4-Mul, constructed the plasmid, miRNA-Control and miRNA-31 were transfected into synovial cells; cultured at 48h after transfection, using dual luciferase reporter gene assay kit for detection of luciferase activity in cells.

**Statistical analysis**

The relative data of this study were analysis by SPSS 22.0 software and shown as mean ± SD (standard deviation); The difference was analysis by one way ANOVA with t-test, p < 0.05 was shown the difference had statistical significance.

**Results**

**miRNA-31 gene expression and pathology of difference tissues**

Compared with normal synovial tissue, the miRNA-31 gene expression was significantly suppressed in RA synovial tissue (p < 0.001) (Fig. 1). By H & E staining, In NC rats, the pathology of synovial tissue had no changes. The synovial layer is composed

![Graph showing miRNA-31 gene expression in NC and RA synovial tissues](image)
of the membrane cells and arranged in an orderly manner. The pathology of RA synovial tissues were the thickness of the matrix layer was obviously thickened, the structure of the collagen fibers in the subsynovial layer changed, and the inflammatory cells were mainly infiltrated by lymphocyte (Fig. 2). Depending on these results, we inferred that miRNA-31 knockdown might be the result which lead RA in RA patients. We wanted to verification miRNA-31 effects by following experiment.

The cell proliferation and apoptosis of difference

The cell proliferation rate of miRNA group was significantly up-regulation compared with that of Control group (p < 0.001) (Fig. 3) and there were no significantly difference between Control and Blank groups (p > 0.05) (Fig. 3). Meanwhile, The cell apoptosis rate of miRNA group was significantly suppressed compared with that of Control group (p < 0.001) (Fig. 4); the cell apoptosis rate of Blank group was no significantly difference compared with Control group (p > 0.05) (Fig. 4). These results were shown that miRNA-31 overexpression had effects to improve synovial cell proliferation by suppressing cell apoptosis induced by RA.

The relative inflammatory factors of difference groups

To evaluate the relative inflammatory factors expression in difference groups, we measured the TNF-α and IL-1β concentrations of the different groups.

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**Fig. 2.** The pathology of NC and RA synovial tissues by H&E staining (×200).

**Fig. 3.** The cell proliferation rate of difference groups by MTT assay. *** p < 0.001, compared with Control group.

**Fig. 4.** The cell apoptosis rate of difference groups by flow cytometry. *** p < 0.001, compared with Control group.
of difference groups in culture fluid by Elisa assay. The results were shown that the TNF-α and IL-1β concentrations were significantly reduced with miRNA-31 over-expression in miRNA group (p < 0.001, respectively). The relative data were shown in Figure 5.

**The relative protein expression of difference groups by WB assay**

By WB assay, we measured the relative proteins expression to explain the mechanism of miRNA-31 in RA treatment. The results were shown that TLR4 and NF-κB proteins expressions of miRNA group were significantly depressed compared with those of Control group (p < 0.001, respectively) and there were no significantly differences between Control and Blank groups in TLR4 and NF-κB proteins expression in vitro study (p > 0.05, respectively). The relative data were shown in Figure 6.

**The correlation between miRNA-31 and TLR4**

To analysis the correlation between miRNA-31 and TLR4, we used double luciferase target experiment to verification the cor-

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**Fig. 5. The IL-1β and TNF-α concentrations of difference groups by Elisa assay. *** p < 0.001, compared with Control group.**

**Fig. 6. The relative proteins expressions of difference groups by WB assay. *** p < 0.001, compared with Control group.**

**Fig. 7. Double luciferase target experiment. *** p < 0.001, compared with miRNA-Control.**
relation between miRNA-31 and TLR4. The results were shown that the LUC activities of miRNA-Control and miRNA-31 were no significantly differences in TLR4-Mul, however, the miRNA-31 was significantly suppressed compared with miRNA-Control in TLR4-WT (p < 0.001). Depending on those results, we inferred that TLR4 might be the target gene of miRNA-34. The relative data were shown in Figure 7.

Discussion

Rheumatoid arthritis (RA) is a multisystemic inflammatory autoimmune disease that mainly involves the surrounding joints. Rheumatoid arthritis is a serious threat to human health due to its high disability rate and organ involvement. At present, the effect of clinical treatment is also needed to improve. Therefore, it is necessary to explore the treatment of RA.

miRNA is encoded by the corresponding gene and is transcribed through a series of processing processes to form a mature small RNA molecule. Mature miRNA is retained in functional complexes, and plays an important regulatory role in gene expression by inhibiting the translation of mRNA or affecting the stability of mRNA by binding to target mRNA (13). miRNA is a key part of the regulation of cytokine signaling pathway and rheumatoid arthritis (14–16). Therefore, we study the interaction of rheumatoid arthritis and miRNA and explore the downstream signal pathway, so as to provide a theoretical basis for the rational treatment and intervention of RA (17). In our present study, we firstly evaluated miRNA-31 gene expression and evaluated the pathology in normal and RA synovial tissues, the results were shown that miRNA-31 gene expression was down-regulation with RA pathology change. Depending on those results, we hypothesized that miRNA-31 over-expression may improve synovial cell injury induced by RA. In the vitro cell experiment, the results were showed that miRNA-31 over-expression enhanced synovial cell proliferation by suppressing cell apoptosis.

IL-1β and TNF-α are two important roles of inflammation in RA development (18–20). IL-1β and TNF-α high concentration leaded cell apoptosis (21, 22). TLR4 plays an important role in the body’s immune defense. As a bridge between immune response and inflammatory response, TLR4 ligand can activate NF-κB transcription factor, and the expression of many inflammatory factors needs NF-κB participation (like as IL-1β and TNF-α) (23). In present study, with miRNA-31 supplement, the RA synovial cell apoptosis was suppressed, meanwhile, the results also found that IL-1β and TNF-α concentrations were decreased via suppressing TLR4/NF-κB activation. Depending on those results, we inferred that TLR4 might be a target gene of miRNA-31. By double luciferase target experiment, the result confirmed TLR4 was a target gene of miRNA-31.

Conclusion

miRNA-31 was a potential therapeutic gene to RA. miRNA-31 over-expression had effects to improve synovial cell proliferation by suppress apoptosis and regulate IL-1β and TNF-α concentration via TLR4/NF-κB pathway.

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Received February 12, 2018.
Accepted March 2, 2018.