Elevated blood lactate in resting conditions correlate with post-exertional malaise severity in patients with Myalgic encephalomyelitis/Chronic fatigue syndrome

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Elevated blood lactate after moderate exercise was reported in some patients with myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS). We hypothesised that blood lactate could be also elevated in resting conditions. We aimed investigating the frequency of elevated lactate at rest in ME/CFS patients, and comparing characteristics of ME/CFS patients with and without elevated lactate. Patients fulfilling international consensus criteria for ME/CFS who attended the internal medicine department of University hospital Angers-France between October 2011 and December 2017 were included retrospectively. All patients were systematically hospitalised for an aetiological workup and overall assessment. We reviewed their medical records for data related to the assessment: clinical characteristics, comorbidities, fatigue features, post-exertional malaise (PEM) severity, and results of 8 lactate measurements at rest. Patients having ≥1 lactate measurement ≥2 mmol/L defined elevated lactate group. The study included 123 patients. Elevated (n = 55; 44.7%) and normal (n = 68; 55.3%) lactate groups were comparable except for PEM, which was more severe in the elevated lactate group after adjusting for age at disease onset, sex, and comorbidities (OR 2.47, 95% CI: 1.10–5.55). ME/CFS patients with elevated blood lactate at rest may be at higher risk for more severe PEM. This finding may be of interest in ME/CFS management.

Myalgic encephalomyelitis also known as chronic fatigue syndrome (ME/CFS) is a long-term and debilitating multisystem condition of unknown aetiology affecting several millions of individuals worldwide1,2. It represents a significant public health issue due to the high levels of health care resource use by ME/CFS patients, and the resulting loss of productivity3. ME/CFS is characterised by persistent or relapsing unexplained fatigue of at least 6 months duration that is not alleviated by rest and leads to a substantial reduction of previous levels of different activities of patients. Beside fatigue, ME/CFS patients report many symptoms, especially unrefreshing sleep, cognitive difficulties, orthostatic intolerance, and pain. The frequency and the severity of symptoms differ greatly between patients and over the course of the disease. Nevertheless, the cardinal feature of ME/CFS is the post-exertional malaise (PEM), which is defined as the exacerbation of some or all of a patient’s symptoms due to physical or cognitive stressors that were normally tolerated before disease onset4. PEM helps distinguish ME/CFS from other causes of chronic fatigue such as depression and fibromyalgia (FM), and recent diagnostic criteria of ME/CFS require its presence4,5.

At present the underlying mechanism of ME/CFS is not yet well defined and its diagnosis, in the absence of a specific diagnostic marker, is based on recognised criteria4,5, and established after eliminating other diseases that could contribute to the patient’s complaints.

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Giving to the fact that ME/CFS is a heterogeneous illness, many studies tried to subtype ME/SFC patients according to various clinical variables such as number of symptoms\textsuperscript{6}, type of fatigue\textsuperscript{7}, mode of illness onset\textsuperscript{8}, age at onset\textsuperscript{9}, and illness course. Biological variables were also used to group ME/CFS patients including natural killer cell activity\textsuperscript{10}, cytokines\textsuperscript{11}, and gene expression in the peripheral blood\textsuperscript{12}.

Mitochondrial dysfunction is known to be linked to fatigue\textsuperscript{13}, and almost all studies that investigated mitochondrial function in ME/CFS patients observed dysfunction, for instance low levels of carnitine\textsuperscript{14}, and CoQ10\textsuperscript{15}, and elevated ventricular lactates\textsuperscript{16,17}. During exercise, low levels of ATP\textsuperscript{18} and elevated blood lactate concentration\textsuperscript{19} were found in some ME/CFS patients. High levels of oxidative stress markers were observed not only after exercise\textsuperscript{20} but also at rest\textsuperscript{21} and correlated with CFS symptoms\textsuperscript{22}.

We hypothesised that a proportion of ME/CFS patients could display abnormal blood lactate concentrations at resting conditions as a result of mitochondrial dysfunction. We thus aimed to examine whether this group of patients represent a distinct phenotype.

Results

Among the 168 patients who were fulfilling inclusion criteria, we excluded 45 patients in whom 24 did not undergo lactate assessment, and 21 patients in whom results of lactate assay were incomplete, mainly due to blood sample haemolysis. Hence a total of 123 patients constituted the study population (Fig. 1).

Table 1 presents the characteristics of the study population. The PEM assessment showed that the median PEM score on the CDC SI questionnaire was equal to 12 [7.5–12]. A more severe PEM (scores \( \geq 12 \)) were observed in 62 (50.4\%) patients. Elevated blood lactate levels at rest \( \geq 2 \text{ mmol/L} \) on one or more measurements were observed in 55 (44.7\%) patients, who constituted the elevated lactate group. Data concerning all lactate measurements can be found as Supplementary Table S1. Figure 2 shows that the overall lactate levels in the elevated lactate group were significantly higher (\( p < 0.0001 \)) than those of the normal lactate group. Lactate values significantly differed between the 2 groups for each time point.

As summarised in Table 2, the comparison between the group of patients with elevated lactate levels and the group of those with normal lactate showed that the only significant difference between both groups was the...
number of patients who display more severe PEM, which was significantly higher in the elevated lactate group (34/55; 61.8%) than in the normal lactate group (28/68; 41.2%), (p = 0.02). This difference persisted after adjusting for age at disease onset, sex, comorbidities, and laboratory findings with an OR for more severe PEM in the elevated lactate group of 2.47 [95% CI: 1.10–5.55] (p = 0.03) (Table 3).

| Epidemiological characteristics                  |          |
|-------------------------------------------------|----------|
| Patients, n (%)                  | 123      |
| Female/Male                      | 89/34    |
| Duration of illness (months)     | 72 [36–120] |
| Age at disease onset (years)     | 34 [26–41] |
| 13–19 n (%)                     | 11 (8.9) |
| 20–29 n (%)                     | 34 (27.6) |
| 30–39 n (%)                     | 42 (34.1) |
| 40–49 n (%)                     | 23 (18.7) |
| ≥50 n (%)                       | 13 (10.6) |
| Family history of fatigue        | 13 (10.6) |
| Identified disease trigger       | 87 (70.7) |
| Infectious triggers              | 58 (47.2) |

| Fatigue features                  |          |
|-----------------------------------|----------|
| Fatigue severity scale (n = 96)   | 5.5 [5–6.2] |
| Fatigue scale (n = 103)           | 24 [20–27.5] |
| Modified fatigue impact scale (n = 104) | 29 [25–32] |
| Physical subscale                 |          |
| Cognitive subscale                |          |
| Psychosocial subscale             |          |

| Clinical manifestations, n (%)    |          |
|-----------------------------------|----------|
| More severe post-exertional malaise* | 62 (50.4) |
| Difficulty processing information | 118 (95.9) |
| Short-term memory loss            | 101 (82.1) |
| Headaches                         | 89 (72.4) |
| Myalgia                           | 107 (87) |
| Arthralgia                        | 72 (58.5) |
| Disturbed sleep patterns          | 109 (88.6) |
| Unrefreshed sleep                 | 118 (95.9) |
| Neurosensory and perceptual disturbances | 114 (92.7) |
| Motor disturbances                | 112 (91.1) |
| Flu-like symptoms                 | 92 (74.8) |
| Recurrent infections              | 47 (38.2) |
| Gastrointestinal impairments      | 105 (85.4) |
| Urinary impairments               | 38 (30.9) |
| Orthostatic intolerance           | 41 (31.3) |
| Palpitation                       | 79 (64.2) |

| Main comorbidities, n (%)         |          |
|-----------------------------------|----------|
| Reactive depression               | 36 (29.3) |
| Fibromyalgia                      | 19 (15.5) |
| Irritable bowel syndrome          | 50 (40.7) |

| Main laboratory abnormalities, n (%) |          |
|-------------------------------------|----------|
| Elevated blood lactate at rest†    | 55 (44.7%) |
| Serum zinc deficit‡                | 59 (31.7) |
| Serum 25-hydroxyvitamin D deficit§ | 61 (49.6) |
| Low plasma 8 am and/or 8 pm cortisol levels¶ | 23 (18.7) |

Table 1. Demographic characteristics, fatigue features, clinical manifestations, comorbid conditions, and laboratory abnormalities of the study population. Notes: * Score ≥12 on Centres for Disease Control and Prevention Symptom Inventory auto-questionnaire. † One or more blood lactate measurements at rest. ‡ Normal range = 0.70–1.25 mg/L, measured by atomic emission spectroscopy/high-frequency induction plasma. § Normal range = 75–250 nmol/L, measured by chemiluminescence technology (CLIA). ¶ Normal values = 5–49 µg/L at 8 am, and 30–100 µg/L at 8 pm, measured by Immunoenzymatic chemoluminescence (12000 ABBOT) method. Cortisol deficit was retained if one or both measurements were reduced. Categorical data were expressed as absolute number and percentage. Continuous data were expressed as median and interquartiles.
Figure 2. Comparison between the overall blood lactate levels in the elevated lactate group and the normal lactate group. Notes: The solid line represents the curve of blood lactate of the elevated lactate group, and the dotted line represents the curve of blood lactate of the normal lactate group. Blood lactate concentrations were measured for each patient at 8 time points on a one-day period: T1 = before breakfast after overnight fasting for 16 hours; T3, T5, and T7 = 30 minutes before lunch, 4 pm snack, and dinner at 4-hour intervals; T2, T4, T6, T8 = 1 hour after the 4 meals. The comparison of both curves was realized by a 2-way ANOVA, and the numerical p-value on the graph is tied to the interaction factor. Lactate values are presented as means with 95% confidence interval. Post-hoc analyses for each lactate time point were realised with Bonferroni corrected t-tests and p-values of these tests were summarised as following: ***p < 0.01, ****p < 0.001 and *****p < 0.0001.

Discussion

ME/CFS is characterised by persistent or recurrent fatigue and the occurrence of PEM, which is a key feature of this debilitating illness. Due to the heterogeneous character of ME/CFS and the absence of biological marker, many studies tried to subtype ME/CFS patients on the basis of clinical or biological variables. Mitochondrial dysfunction has been shown in some of ME/CFS patients. In the current study we identified a group of ME/CFS patients with elevated blood lactate in resting conditions and displaying more severe PEM.

The demographic characteristics, clinical manifestations, and comorbid conditions of our population were comparable to that reported in prior studies. For instance, the age peak in the age range 30–39 (34% of patients) and the median age of disease onset of 34 [26–41] years, the higher prevalence of women and the median duration of illness, and the family history of chronic fatigue syndrome (CFS) in one or more first-degree relatives (11% of patients) were in accord with that previously reported. Nevertheless, the percentage of patients who identified at least one ME/CFS precipitating factor (70.7%) and that of those who reported an infectious trigger (47.2%) was somewhat lower than that of prior studies.

One or more comorbidities were found in 76 (61.8%) patients, which corresponds to previous reports that found comorbidities prevalence between 50 and 65%, but higher prevalence was reported in one study. Laboratory anomalies observed in our population including reduced 8am and/or 8 pm plasma cortisol levels (18.7%), low serum zinc (32%), and low serum 25-hydroxy vitamin D (49.6%) were comparable to that reported in literature.

Blood lactate assay results showed elevated lactate levels in resting conditions in 55 (44.7%) patients. Lane et al. found elevated lactate concentrations in a proportion of CFS patients (31/96; 32%) after short periods of exercise below the predicted anaerobic threshold. They also reported that patients displaying abnormal lactate responses to exercise had relative deficiency of mitochondria type I fibres on muscle biopsies, which may explain elevated lactate responses to exercise, and low intracellular pH during recovery phase when examined by magnetic resonance spectroscopy of muscle. Other teams showed elevated ventricular lactates in CFS patients pointing to metabolic dysfunction in these patients. In the current study, almost half of patients had elevated blood lactate concentrations in resting conditions. To the best of our knowledge, our study is the first to show these results. Lactate values were significantly different between the 2 groups and this difference was observed for each time point. Nevertheless, this difference appeared to be more prominent at T2. A hypothesis that could explain, at least in part, the predominant elevation of lactate levels at T2 is that eating after 16-hour fasting would abolish the beneficial effect of fasting on mitochondrial functioning and redox signalling, leading to a sort of rebound effect responsible for mitochondrial dysfunction with increased blood lactate concentration after breakfast. The positive effect of fasting was reported in FM patients, which could be interesting to evaluate its impact in ME/CFS patients.

When we classified our study population into two groups with or without elevated lactate to compare their phenotypic characteristics, we found that they only differ in the number of patients experiencing more severe PEM, which was significantly higher in patients with elevated lactate. This association persisted after adjusting for age at disease onset, sex, and comorbidities (OR 2.47, 95% CI: 1.10–5.55; p = 0.03). PEM severity was assessed in all patients by the standardised CDC SI self-reported questionnaire, which includes an item designed to measure the presence, frequency, and intensity of PEM. The CDC SI is one of two tools proposed by the IOM for PEM assessment. We used the median PEM score on the CDC SI questionnaire to set the cut-off score ≥12 defining more severe PEM in our study.
Elevated blood lactate in resting conditions in ME/CFS patients was not reported before. In our study, patients did not display a state of extreme fatigue at the time of lactate assay, and strict resting conditions were followed carefully in all patients all over the period of different samples’ collection during hospitalisation. As well, all patients received standardised meals. Thus, the rise of lactate levels in a proportion of patients could not be related to exertion or due to variation in caloric intake.

It is interesting to note that although the whole study population was suffering from high levels of fatigue and fatigue-related impairment, results of fatigue scales showed that fatigue severity was comparable between both groups; FS median score (elevated lactate group 24 [21–28] vs. normal lactate group 23 [17–27]; p = 0.14) and FSS median score (elevated lactate group 5.6 [4.8–6.2] vs. normal lactate group 5.4 [5.1–6]; p = 0.90). In this way, elevated lactate levels were correlated with PEM severity, but not with fatigue severity. One can expect that patients who display severe PEM are suffering from high level of general fatigue, however some authors report

| Epidemiological characteristics | Elevated lactate | Normal lactate | p |
|--------------------------------|------------------|----------------|---|
| Patients, n (%)                | 55 (44.7)        | 68 (55.3)      |   |
| Female/Male                    | 35/20            | 54/14          | 0.052 |
| Duration of illness (months)   | 60 [63–96]       | 78 [36–144]    | 0.11 |
| Age at disease onset (years)   | 36 [25.5–42]     | 32 [26–39]     | 0.26 |

Fatigue features

| Fatigue severity scale | Elevated lactate (n = 43) | Normal lactate (n = 53) | p |
|------------------------|----------------------------|-------------------------|---|
|                        | 5.6 [4.8–6.2]              | 5.4 [5.1–6]             | 0.90 |

| Fatigue scale | Elevated lactate (n = 46) | Normal lactate (n = 57) | p |
|---------------|----------------------------|-------------------------|---|
|               | 24 [21–28]                 | 23 [17–27]              | 0.14 |

| Modified fatigue impact scale | Elevated lactate (n = 47) | Normal lactate (n = 57) | p |
|-------------------------------|---------------------------|-------------------------|---|
| Physical subscale             | 28 [24–32]                | 30 [27–32]              | 0.09 |
| Cognitive subscale            | 28 [20–33]                | 26 [20–32]              | 0.59 |
| Psychosocial subscale         | 6 [4–7]                   | 6 [4–6]                 | 0.43 |

Patients’ comorbidities, n (%)

| Reactive depression | Elevated lactate 15 (27.3) | Normal lactate 21 (30.9) | 0.66 |
| Fibromyalgia        | Elevated lactate 8 (14.6)   | Normal lactate 11 (16.2)  | 0.80 |
| Irritable bowel syndrome | Elevated lactate 18 (32.7) | Normal lactate 32 (47.1)  | 0.11 |

Clinical manifestations n (%)

| More severe post-exertional malaise* | Elevated lactate 34 (61.8) | Normal lactate 28 (41.2) | 0.02 |
| Difficulty processing information  | Elevated lactate 54 (98.2)  | Normal lactate 64 (94.1)  | 0.26 |
| Short-term memory loss             | Elevated lactate 46 (83.6)  | Normal lactate 55 (80.9)  | 0.69 |
| Headaches                          | Elevated lactate 42 (76.4)  | Normal lactate 47 (69.1)  | 0.37 |
| Myalgia                            | Elevated lactate 50 (90.9)  | Normal lactate 57 (83.8)  | 0.25 |
| Arthralgia                         | Elevated lactate 36 (65.5)  | Normal lactate 36 (53)    | 0.16 |
| Disturbed sleep patterns           | Elevated lactate 51 (92.7)  | Normal lactate 58 (85.3)  | 0.20 |
| Unrefreshed sleep                  | Elevated lactate 51 (92.7)  | Normal lactate 67 (98.5)  | 0.11 |
| Neurosensory and perceptual disturbances | Elevated lactate 48 (87.3) | Normal lactate 66 (97.1)  | 0.16 |
| Motor disturbances                 | Elevated lactate 50 (90.9)  | Normal lactate 61 (89.7)  | 0.56 |
| Flu-like symptoms                  | Elevated lactate 42 (76.4)  | Normal lactate 50 (73.5)  | 0.72 |
| Recurrent infections               | Elevated lactate 25 (45.5)  | Normal lactate 22 (32.4)  | 0.14 |
| Gastrointestinal impairments       | Elevated lactate 43 (78.2)  | Normal lactate 62 (91.2)  | 0.63 |
| Urinary impairments                | Elevated lactate 15 (27.3)  | Normal lactate 23 (33.8)  | 0.43 |
| Orthostatic intolerance            | Elevated lactate 15 (27.3)  | Normal lactate 26 (38.2)  | 0.20 |
| Palpitation                        | Elevated lactate 35 (61.8)  | Normal lactate 44 (64.7)  | 0.90 |

Laboratory abnormalities, n (%)

| Serum zinc deficit† | Elevated lactate 20 (36.4) | Normal lactate 19 (28) | 0.31 |
| Serum 25-hydroxyvitamin D deficit‡ | Elevated lactate 31 (56.4) | Normal lactate 30 (44.1) | 0.18 |
| Low plasma 8 am and/or 8 pm cortisol levels§ | Elevated lactate 10 (18.2) | Normal lactate 13 (19.1) | 0.90 |

Table 2. Comparison of patients according to blood lactate levels at rest. Notes: *Score ≥12 on Centres for Disease Control and Prevention Symptom Inventory auto-questionnaire. †Normal range = 0.70–1.25 mg/L, measured by atomic emission spectroscopy/high-frequency induction plasma. ‡Normal range = 75–250 nmol/L, measured by chemiluminescence technology (CLIA). §Normal values = 5–49 µg/L at 8 am, and 30–100 µg/L at 8 pm, measured by Immunoenzymatic chemoluminescncze (12000 ABBOT) method. Cortisol deficit was retained if one or both measurements were reduced. Categorical data were expressed as absolute number and percentage. Continuous data were expressed as median and interquartiles.

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that some of patients who experienced severe PEM did not report high fatigue levels because they were reducing their activities to stay within their energy envelope and so their fatigue levels may be low but they can experience exhaustion if they exceed their functional capacities or exposed to a PEM trigger. Another possible explanation is that there is currently no gold standard measurement of fatigue, and a ceiling effect for FSS and FS was observed in multiple sclerosis patients, but not in those with ME/CFS. Nevertheless, we have used this scale alongside two other scales, FS and FSS, which explore fatigue severity as recommended by the CDC-NINDS project.

Finally, the MFIS scale that we used to measure the impact of the fatigue on function has been mainly validated in patients with an identifiable medical condition that could account for chronic fatigue, and those with primary psychiatric disorders or substance dependence were excluded. Patients with diseases or drugs susceptible to be associated with elevated lactate levels were also excluded. According to ICC 2011, comorbidities such as FM, irritable bowel syndrome, Hashimoto’s thyroiditis, and reactive depression did not constitute an exclusionary condition.

Epidemiological characteristics, fatigue features, and comorbidities were extracted from patients’ medical records as well as reported patients’ symptoms including the frequency and intensity of PEM.

The following biological data were also gathered for each patient: lactate assessment, immunological assay, infectious disease screen for HIV, hepatitis B and C, Lyme disease, enterovirus, Mycoplasma pneumoniae, Chlamydia pneumoniae, serum zinc, 25- hydroxyvitamin D, morning (8 am) and evening (8 pm) plasma cortisol, testosterone, iron studies, plasma vitamin B12 and serum folate, full blood count and differential, erythrocyte

| p-value | OR [95% CI] |
|---------|-------------|
| Sex (male) | 0.42 | 1.46 [0.58–3.66] |
| Age at onset disease ≥34 years | 0.93 | 1.04 [0.46–2.31] |
| PEM score ≥12 | 0.03 | 2.47 [1.10–5.55] |
| Reactive depression | 0.81 | 0.86 [0.37–2.17] |
| Fibromyalgia | 0.52 | 0.570[0.23–2.12] |
| Irritable bowel syndrome | 0.31 | 0.66 [0.29–1.48] |
| Serum zinc deficit | 0.44 | 1.41 [0.59–3.35] |
| Serum 25-hydroxyvitamin D deficit | 0.38 | 1.45 [0.63–3.33] |
| Low plasma cortisol levels | 0.77 | 1.16 [0.43–3.17] |

Table 3. Multivariate analysis of severe post-exertional malaise in the group of elevated lactate. Notes: OR [95% CI]: Odds Ratio with 95% Confidence interval; PEM: post-exertional malaise; Multivariate analysis was performed with logistic regression. The variable to explain was the lactate group (elevated or normal). The variables included in the model were those present in the table as well treated sleep apnoea syndrome (p = 0.10) and treated Hashimoto’s thyroiditis (p = 0.41).
sedimentation rate, electrolytes, calcium, phosphate, fasting glucose, C-reactive protein, liver function, serum protein electrophoresis, renal function, thyroid function, and creatine kinase.

**Fatigue assessment tools.** Fatigue level was assessed by means of standardised auto-questionnaires; the fatigue scale (FS)\(^{14}\) and the fatigue severity scale (FSS)\(^{15}\). The impact of fatigue on patient’s activities was assessed by the modified fatigue impact scale (MFIS)\(^{16}\).

**PEM assessment.** The PEM item from the standardised auto-questionnaire of Centres for Disease Control and Prevention Symptom Inventory (CDC SI)\(^{18}\) was used to measure PEM symptoms over the past month. Perceived frequency of PEM was rated on a 4-point scale (1 = a little of the time, 2 = some of the time, 3 = most of the time, 4 = all of the time), and its intensity was measured on a 3-point scale (1 = mild, 2 = moderate, 3 = severe). The intensity score was converted to equidistant score (0 = symptom not reported, 1 = mild, 2.5 = moderate, 4 = severe). The frequency and intensity scores were then multiplied to create the PEM severity score ranging from 0–16. In the absence of validated threshold that defines PEM severity, we used the median PEM score on the CDC SI questionnaire to set the cut-off score ≥12 defining more severe PEM.

**Lactate assay.** The following procedure was applied for all patients included in the study: each patient was admitted at 3 pm (day 1). A peripheral venous catheter was inserted as soon possible after admission and patient was asked to follow resting and fasting conditions (drinking water was allowed till midnight) until next day (day 2). At day 2, eight blood samples were taken from each patient without tourniquet in resting conditions for at least 30 minutes and without preceding exercise\(^{14}\). The first blood sample was obtained before breakfast at 7.30 am that corresponds to time point 1 (T1) after overnight fasting for 16 hours. The other 3 pre-prandial samples were obtained 30 minutes before lunch (T3), 4 pm snack (T5), and dinner (T7) at 11.30 am, 3.30 pm, and 7.30 pm, respectively. Four post-prandial samples were obtained 1 hour after meals (T2, T4, T6, T8). The different meals were standardised for all patients during hospital stay. Caloric values for every day breakfast and 4 pm snack were exactly the same; 358 Kcal and 135 Kcal, respectively. The mean caloric values of lunch and dinner over the 3-day hospitalisation were 885 Kcal and 446 Kcal, respectively for all patients. For lactate measurements, 2 ml of venous blood was collected in lithium heparin tubes and immediately transported to the Biochemistry Laboratory on ice. The plasma is then separated by centrifugation at 20° for 10 minutes X 2255 g. Quantitative assay of plasma lactate was measured with ADVIA® Chemistry XPT system (Siemens Healthcare Diagnostics Ltd., UK) by an enzymatic reaction using lactate oxidase to oxidize lactate into pyruvate and hydrogen peroxide. Catalysed by peroxidase, the hydrogen peroxide formed reacts with a chromogen yielding a purple coloured substance. The resulting modification in absorbance at 545/694 nm measured by spectrophotometry is proportional to the lactate concentration. References values for blood lactate were as follows: T1 (0.55–1.15 mmol/L), T2 (0.90–1.60 mmol/L), T3 (0.70–1.40 mmol/L), T4 (0.90–1.40 mmol/L), T5 (0.56–1.06 mmol/L), T6 (0.70–1.50 mmol/L), T7 (0.55–1.18 mmol/L), and T8 (0.73–1.45 mmol/L). A cut-off ≥2 mmol/L at any lactate time point defined elevated lactate levels. The lower limit of quantitation was 0.03 mmol/L, and the error measurement of the assay was 5.22%. Results of lactate cycle were unexploitable if more than 2 out of 8 time points measurements were missing, mainly due to haemolysis.

**Patients’ grouping.** According to blood lactate concentrations, we stratified patients into two groups; the first one included patients in whom ≥1 lactate measurements were ≥2 mmol/L, and the other one comprised patients with normal lactate concentrations.

**Statistical analysis.** Categorical data were expressed as absolute number and percentage. Continuous data were expressed as median and interquartile (IQ). Data were compared using the Student t test for continuous variables and the χ² test or Fisher’s test for categorical data. Lactate analyses were compared using 2 way-ANOVA and lactate values were presented as means with 95% confidence interval in Fig. 2. Post-hoc analyses for each lactate time point were realised after 2-way ANOVA with Bonferroni corrected t-tests. The multivariate analysis for testing the association of comorbidities and the group of elevated lactates were carried out using binomial logistic regression. The continuous variable Age was transformed into a categorical variable with a cut-off equal to the median of the whole population (< or ≥34 years). Odds-ratio (OR) were calculated with their 95% confidence interval (CI). A p value of <0.05 was considered significant. The analyses were performed using the Graphpad Prism v6.01 software (GraphPad Software, Inc., La Jolla, CA 92037 USA).

**Data availability**
Data that support the findings of this study are available from the corresponding author upon reasonable request.

**References**
1. Prins, J. B., Meer, J. Wvander & Bleijenberg, G. Chronic fatigue syndrome. *The Lancet* **367**, 346–355 (2006).
2. Beyond Myalgic Encephalomyelitis/Chronic Fatigue Syndrome: Redefining an Illness. *Mil. Med.*** **180**, 721–723 (2015).
3. Collin, S. M. & Crawley, E. Specialist treatment of chronic fatigue syndrome/ME: a cohort study among adult patients in England. *BMC Health Serv. Res.*** **17** (2017).
4. Carruthers, B. M. *et al.* Myalgic encephalomyelitis: International Consensus Criteria: Review: ME: Intl. Consensus Criteria. *J. Intern. Med.*** **270**, 327–338 (2011).
41. Chalder, T.
40. Myalgic Encephalomyelitis/Chronic Fatigue Syndrome Standards - NINDS Common Data Elements. https://www.commondataelements.ninds.nih.gov/MECFS.aspx#tab-Data_Standards.
37. Jason, L.
29. Van Den Eede, F., Moorkens, G., Van Houdenhove, B., Cosyns, P. & Claes, S. J. Hypothalamic-Pituitary-Adrenal Axis Function in Chronic Fatigue Syndrome: preliminary evidence for a subgroup. J. Psychosom. Res. 60, 559–566 (2006).
36. Moneghetti, K. J. et al. Value of Circulating Cytokine Profiling During Submaximal Exercise Testing in Myalgic Encephalomyelitis/Chronic Fatigue Syndrome. Sci. Rep. 8, (2018).
35. Michalsen, A.
34. Craig, C. Mitoprotective dietary approaches for Myalgic Encephalomyelitis/Chronic Fatigue Syndrome: Caloric restriction, fasting, and ketogenic diets. Med. Hypotheses 85, 690–693 (2015).
33. Michalsen, A. et al. In-Patient Treatment of Fibromyalgia: A Controlled Nonrandomized Comparison of Conventional Medicine versus Integrative Medicine including Fasting Therapy. Evid. Based Complement. Alternat. Med. 2013, 1–7 (2013).
32. Wagner, D. et al. Psychometric properties of the CDC Symptom Inventory for assessment of Chronic Fatigue Syndrome. BMC Med. 8, 204–209 (1998).
31. Lane, R. J. et al. Muscle fibre characteristics and lactate responses to exercise in chronic fatigue syndrome. J. Neurol. Neurosurg. Psychiatry 64, 362–367 (1998).
30. Lane, R. J. M., Barrett, M. C., Taylor, D. J., Kemp, G. J. & Lodi, R. Heterogeneity in chronic fatigue syndrome: evidence from magnetic resonance spectroscopy of muscle. Neuromusc. Disord. 8, 204–209 (1998).
29. Craig, C. Mitoprotective dietary approaches for Myalgic Encephalomyelitis/Chronic Fatigue Syndrome: Caloric restriction, fasting, and ketogenic diets. Med. Hypotheses 85, 690–693 (2015).
28. Castro-Marrero, J. et al. Comorbidity in Chronic Fatigue Syndrome/Myalgic Encephalomyelitis: A Nationwide Population-Based Cohort Study. Psychosomatics 58, 533–543 (2017).
27. Aaron, L. A., Burke, M. M. & Buchwald, D. Overlapping Conditions Among Patients With Chronic Fatigue Syndrome, Fibromyalgia, and Temporomandibular Disorder. Arch. Intern. Med. 160, 221 (2000).
26. Castro-Marreiro, J. et al. Comorbidity in Chronic Fatigue Syndrome/Myalgic Encephalomyelitis: A Nationwide Population-Based Cohort Study. Psychosomatics 58, 533–543 (2017).
25. Shungu, D. C. Precipitating factors for the chronic fatigue syndrome. J. Psychosom. Res. 147–153 (1993).
24. Chu, L., Valencia, I. J., Garvert, D. W. & Montoya, J. G. Onset Patterns and Course of Myalgic Encephalomyelitis/Chronic Fatigue Syndrome. Adv. Exp. Med. Biol. 132–138 (1995).
23. Bakken, I. J.
22. Richards, R. S., Roberts, T. K., McGregor, N. R., Dunstan, R. H. & Butt, H. L. Blood parameters indicative of oxidative stress are associated with symptom expression in chronic fatigue syndrome. J. Neurol. Neurosurg. Psychiatry 59(1), 112–120 (2007).
21. Fulle, S. et al. Specific oxidative alterations in vastus lateralis muscle of patients with the diagnosis of chronic fatigue syndrome. Free Radic. Biol. Med. 29, 1252–1259 (2000).
20. Kennedy, G. et al. Oxidative stress levels are raised in chronic fatigue syndrome and are associated with clinical symptoms. Free Radic. Biol. Med. 39, 584–589 (2005).
19. Lane, R. J. M., Burgess, A. P., Flint, J., Riccio, M. & Archard, L. C. Exercise responses and psychiatric disorder in chronic fatigue syndrome. Arch. Psychosomatics 79, 59–65 (1997).
18. Lengert, N. & Drossel, B. In silico analysis of exercise intolerance in myalgic encephalomyelitis/chronic fatigue syndrome. Biophys. Chem. 202, 21–31 (2015).
17. Jason, L. A.
16. Shungu, D. C. Precipitating factors for the chronic fatigue syndrome. J. Psychosom. Res. 55, 112–120 (2007).
15. Maes, M.
14. Parikh, S.
13. Filler, K.
12. Kerr, J. R. et al. Gene Expression Subtypes in Patients with Chronic Fatigue Syndrome/Myalgic Encephalomyelitis. J. Infect. Dis. 197, 1171–1184 (2008).
11. Moneghetti, K. J. et al. Value of Circulating Cytokine Profiling During Submaximal Exercise Testing in Myalgic Encephalomyelitis/Chronic Fatigue Syndrome. Sci. Rep. 8, (2018).
10. Stegel, S. D. et al. Impaired natural immunity, cognitive dysfunction, and physical symptoms in patients with chronic fatigue syndrome: preliminary evidence for a subgroup. J. Psychosom. Res. 60, 559–566 (2006).
9. Kidd, E.
8. Jason, L.
7. Jason, L. A. et al. Development of a fatigue scale. J. Psychosom. Res. 43, 147–153 (1993).
6. Krupp, L. B., LaRocca, N. G., Muir-Nash, J. & Steinberg, A. D. The Fatigue Severity Scale: Application to Patients With Multiple Sclerosis and Systemic Lupus Erythematosus. Arch. Neurol. 46, 1121–1123 (1989).
5. Fukuda, K. et al. The chronic fatigue syndrome: a comprehensive approach to its definition and study. International Chronic Fatigue Syndrome Study Group. Ann. Intern. Med. 121, 953–959 (1994).
4. Collin, S. M., Heron, J., Nikolaus, S., Knoop, H. & Crawley, E. Chronic fatigue syndrome (CFS/ME) symptom-based phenotypes and 1-year treatment outcomes in two clinical cohorts of adult patients in the UK and The Netherlands. J. Psychosom. Res. 104, 29–34 (2018).
3. Jason, L. A. et al. Classification of Myalgic Encephalomyelitis/Chronic Fatigue Syndrome by Types of Fatigue. Behav. Med. 36, 24–31 (2010).
2. DeLuca, J., Johnson, S. K., Ellis, S. P. & Natileon, B. H. Sudden vs gradual onset of chronic fatigue syndrome differentiates individuals on cognitive and psychiatric measures. J. Psychiatr. Res. 31, 83–90 (1997).
1. Kidd, E. et al. The Relationship between Age and Illness Duration in Chronic Fatigue Syndrome. Diagnostics 6, 16 (2016).

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Author contributions
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Competing interests
The authors declare no competing interests.

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