Bone Phenotyping Approaches in Human, Mice and Zebrafish – Expert Overview of the EU Cost Action GEMSTONE (“GEnomics of MusculoSkeletal traits Translational NEtwork”)

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A synoptic overview of scientific methods applied in bone and associated research fields across species has yet to be published. Experts from the EU Cost Action GEMSTONE (“GEneromics of MusculoSkeletal Traits translational Network”) Working Group 2 present an overview of the routine techniques as well as clinical and research approaches employed to characterize bone phenotypes in humans and selected animal models (mice and zebrafish) of health and disease. The goal is consolidation of knowledge and a map for future research. This expert paper provides a comprehensive overview of state-of-the-art technologies to investigate bone properties in humans and animals – including their strengths and weaknesses. New research methodologies are outlined and future strategies are discussed to combine phenotypic with rapidly developing –omics data in order to advance musculoskeletal research and move towards “personalised medicine”.

Keywords: bone and skeletal diseases, phenotyping, imaging, animal models, GEMSTONE, COST

INTRODUCTION

Bone metabolism and its regulation involves complex interactions and crosstalk across multiple tissues, physiological systems and pathways from fat and muscle to the immune system and gut-bone axis (1–3). With this knowledge and based on the recent advances in our understanding of genetics and genomics, this narrative overview of technological evidence...
intends a practical information for young researchers and/or scientists outside the respective bone areas to enable crosstalk between the disciplines.

The ultimate goal is to translate between clinical and preclinical research and aim for mutual interaction and development of future diagnostic and therapeutic approaches, drug development and risk assessment. This task is being undertaken by experts from the EU Cost Action GEMSTONE (“Genomics of MusculoSkeletal traits Translational Network”). One remit is to facilitate interaction between researchers in animal and human bone science and establish common phenotypic terminology across different spheres of expertise, thus enabling translational comparability of phenotypic signatures. The first step in this process, undertaken by GEMSTONE working group 2, Phenotyping, is to curate a comprehensive catalogue of bone phenotyping methods used within GEMSTONE in human and give a compressed overview on comparable methodology used in mice and zebrafish studies.

In this publication, we summarise the current state of the art, identify gaps in knowledge and suggest future directions/needs to be addressed. We provide insights in how the presented animal models can be used to model bone disease and complement human studies in order to advance bone phenotyping. Integrating the aims of this working group and the larger GEMSTONE action, we briefly outline how –omics technologies can contribute to the phenotypic dissection of skeletal traits. Finally, we offer our perspective on triangulation of the diagnostic evidence and lay out strengths and limitations of the respective techniques.

Discrepancies in the translation of clinical and preclinical research results are an important issue that complicates the understanding and progress in the care for patients with bone disease but also in associated disciplines in the bone field. Bone diseases are complex and multifactorial and require more than the just traditional methods to aim for new horizons with future diagnostic and therapeutic approaches.

Phenotyping and endophenotyping can be mechanistically oriented towards drug development, targeted treatment or prognostically oriented towards treatment stratification and treatment decisions. With this expert view on phenotyping methods across species, we aim for building bridges between animal and human bone science to establish common phenotypic terminology including growth-specific aspects, enabling translational comparability of phenotypic signatures for all researchers involved.

There are many open questions and unmet needs in the field of bone diseases in humans, such as the achievement of an optimal peak bone mass, robust evaluations of bone strength in clinical practice, including cross-validation between measurement methods and more holistic approaches in diagnosis as well as personalized, tailored treatment (4). An increased understanding of perspectives in animal models might help to solve a number of these open questions, as they are important issues for millions of people, e.g., diagnosis and treatment in children, adolescents or young adults, questions of the ideal use of current imaging techniques including new technologies to measure bone quality and strength, the interaction of epigenetic factors and the microbiome with bone quantities and qualities and future treatment options. Many new aspects might be answered by specific animal models, which are described in more detail.

Mice models are popular in studies of skeletal physiology due to their relatively low cost, high rates of reproduction, and ease of handling and care (5). They also provide the opportunity to collect phenotype data not available from humans, and to study the effect of single, specific interventions that are not possible in patients such as changes to diet, age, or genetics (6).

A number of features of bone biology are shared between the mouse and human skeleton. Like humans, mice experience age-related bone loss (5). They also undergo similar patterns of bone turnover and bone healing to humans (7, 8). However, there are differences that should be considered such as the lack of Haversian organisation and non-closure of growth plates at skeletal maturity (6).

Mice have a high homology to the human genome, making them suitable models for many human genetic disorders (6, 7). Manipulation of the mouse genome has allowed for the creation of models for numerous human musculoskeletal diseases. Transgenic and gene-targeted mice have allowed for studies of global overexpression or deletion of genes of interest for decades, but more recent technologies are making more specific genetic manipulation possible. The Cre-lox system applies for cell-specific and temporal deletion of target genes. In this system, LoxP sites are inserted on either side of the target gene or sequence, and when bred to a mouse expressing the Cre recombinase the relevant segment of DNA is excised in the desired cell type or developmental stage (9). CRISPR/Cas9 is the most recently developed technology and uses adapted bacterial proteins which cleave double stranded DNA at specific sites, offering a quick and accurate option for gene editing (10).

An additional model for bone research are small teleost fish such as zebrafish (Danio rerio) and medaka (Oryzias latipes). This model has been increasingly used to interrogate the biology of human skeletal conditions. Here, we will focus on zebrafish as an emerging and alternative model system used for the study of molecular mechanisms and gene function associated with human skeletal diseases. Zebrafish show conserved physiology compared to mammals and display advantages as animal model, such as the generation of a high number of embryos per cross (over 150), their rapid and transparent embryonic development that combined with the availability of a number of bone specific transgenic lines, allow in vivo cell trackability (11, 12). Moreover, genetic manipulation in zebrafish is relatively simple and highly efficient. Evaluation of the first bones in larval stages and adult whole skeleton can be performed in high-resolution with reasonably high throughput (13, 14). Zebrafish have been used for genetic and drug screening, and they pose an attractive model system to accelerate functional validation of human-omics findings.

Despite being evolutionarily more distant from humans than mice, zebrafish share key bone similarities, showing the same bone cell types (osteoblasts, osteocytes osteoclasts and...
chondrocytes) and types of ossification (intramembranous and endochondral) as those found in mammals, with the advantage that the first bones and cartilage are available for studies from the 3rd day of development (11). During ageing, zebrafish show bone macro and microstructure reminiscent of osteoporosis (15) and osteoarthritis (16). Furthermore, non-invasive bone fracture experiments in zebrafish allow investigation of bone healing and fracture repair (17, 18). At the molecular scale, zebrafish bone is reminiscent of mammalian bone up to the level of aligned mineralized collagen fibrils (19). Zebrafish also show some differences that should be considered. Unlike in humans, zebrafish bones do only show few bones with trabeculae, whereas long bones are absent. The bone marrow in zebrafish is fatty and does not harbour a site for haematopoesis, but blood vessels invade the bone marrow similar to mammals (20). Zebrafish have growth plates, but the main source of longitudinal growth relies on cartilage proliferation and not from accumulation of hypertrophic chondrocytes, as only a small portion of chondrocytes become hypertrophic (21).

For further information and details, see also the GEMSTONE WG3 publication on “Gene & Therapeutic Discoveries in Bone Mass Disorders”.

Insights into mouse and zebrafish biology and pathophysiology will allow for a better understanding of human investigations and open clinical questions. There are substantial differences between the views of experts in human disease on various aspects of bone. Therefore, a translational approach for new research reducing the discrepancy between the animal and human models is highly warranted. Even in case, techniques cannot be directly compared, they may be tailored to specific research questions in the future.

Many links liaise this publication to those of Working Group 3 and 4 of the GEMSTONE COST Action with important details to many topics mentioned in this manuscript. This comprehensive overview allows us to better classify and detect bone diseases, predict disease progression using radiographic and clinical scores, clustering (identification of different groups/phenotypes of patients with bone diseases), pinpoint the most important characteristics that could affect disease progression and identify patients who will be rapid progressors for the development of late sequelae, e.g. multiple fractures. This paper aims to link the knowledge and understanding of different aspects of bone disease from various expert viewpoints, contributing to a solid basis for further and more effective cooperation between various specialties to enable a personalized care in this field in the future.

1 MUSCULOSKELETAL PHENOTYPING OF BONE CONDITIONS

Musculoskeletal phenotyping is a broad and multi-faceted process that provides essential information for establishing a diagnosis of bone conditions, with or without bone fragility and muscle weakness. For all common or rare forms of musculoskeletal disorders, a comprehensive evaluation of clinical and functional aspects is required since fragility depends on much more than bone mineral density (BMD) alone (Figure 1).

1.1 Medical History and Physical Examination

A fragility fracture in children or adults is often the first sign of an underlying primary or secondary disease. A detailed medical history and thorough clinical examination can provide valuable insights into the overall state of musculoskeletal health. The content of the medical history depends on a patient’s age. In a child, family history of bone fragility, joint laxity or hearing loss gives essential clues towards the presence of genetic disorder, such as osteogenesis imperfecta. For humans of all ages, a history of back pain can relate to the presence of low-impact vertebral fractures, which may in turn increase the risk for future fractures. In addition, chronic or acute underlying conditions such as rheumatoid arthritis, diabetes, malabsorption, hypogonadism or premature menopause and stroke and neural damage may cause cytokine-, glucocorticoid- or immobility-induced metabolic disease that in turn can affect skeletal and muscular strength. The physical examination includes anthropometry, inspection of limbs and spine for deformities, assessing sclerae and teeth, palpation of spine and extremities along with observing the patient’s posture, limb length, muscle tone and strength. The spine is assessed for tenderness, and deformities (such as scoliosis, hyperkyphosis, or hyperlordosis). Decreased mobility and low lean mass predict low bone mass in humans according to the mechanostat theory (22). Sarcopenia, pain, presence of gait, balance and vision disturbances therefore provide important information on the risk of falling and future fractures. These parameters may be summarized in the concept of Patient Reported Outcome Measures from Questionnaires (PROMs) or Clinician Reported Outcome Measures (CROMs), respectively (see Figure 1) and may include a large number of additional terms, including psychological and social approaches. For more detailed phenotyping in genetic musculoskeletal diseases, see also, “Careful patient phenotyping is key to disease discovery” in the publication by GEMSTONE WG3.

Limitations: Taking a thorough medical history and assessing a deep clinical phenotype is time consuming and requires profound expertise of an experienced examiner. Studies may not even employ sufficient clinical phenotyping or time into this important investigation. An additional limiting factor may be a lack of knowledge and patients’ recall bias as well as the non-availability of x-rays or other clinical imaging for the clinician to confirm a patient’s fracture history and assess the radiological bone phenotype.

Strengths: Medical history and careful physical examination provide essential hints for the further diagnostic workup and avoid unnecessary or repetitive testing.

In mice models, detailed records and breeding charts should be kept for all mouse colonies, and these can and should be used as a proxy for medical history. These detailed colony records allow tracing of recurring skeletal problems or fractures. Physical examination is equally important in mice as in humans, and should include inspection of their condition, behaviour and environment. In regard to the skeleton and muscle tonus, this inspection should include examination of the incisors, gait
abnormalities or lameness, and manually manipulating of the limbs (23).

In zebra fish, the skeleton can be regionalized into functional groups including the craniofacial skeleton and the vertebral column (together with the tail fin, being parts of the axial skeleton), and fins (pectoral, dorsal, anal fins). Since different genetic mutations often affect several skeletal compartments, it is common to perform skeletal phenotyping as a whole (13). Gross skeletal deformities such as scoliosis, hyperkyphosis, hyperlordosis, emaciation, specific swimming patterns, are readily visible (16). Severe abnormal spinal curvature can be detected as the fish swim in the tank. It can be argued that “family history” is as relevant for the genetically-modified fish as for the mice of inbred lines; although the exact parents are usually not known for every specific fish, parental pairs usually come from well-documented strains/established mutants. Although there may be a lack of one-to-one relationship with the fish musculoskeletal phenotype during aging, the latter is measurable (24).

1.2 Questionnaires and Patient-Reported Outcome Measures

Patient-reported outcome measures (PROMs), which are collected using questionnaires are essential to comprehend the full extent of how musculoskeletal diseases influence the quality of life. In adults, questionnaires are often used to systematically collect information on self-reported socio-demography, medical conditions, family and fracture history, medication use, lifestyle such as dietary intake, smoking habits, alcohol consumption, physical activity and quality of life. Such information is important given that environmental factors, in combination with genetic susceptibility contribute to general frailty and risk for fracture.

In premenopausal women, there might be special attention to pregnancy and lactation based on hormonal changes and challenged calcium metabolism, due to the nutritional demands of the foetus and neonate (25–27). Female specific questions may address age at menarche, cycle abnormalities and conditions such as gestational diabetes (28) and preeclampsia, also for the child (29). In men, hypogonadism and other endocrine disturbances, but also exogenous toxins might be asked for. In children and adolescents, where heritable forms of osteoporosis are mostly diagnosed, questionnaires are not commonly used and emphasis is put on family history and physical examination.

A wide variety of patient-reported outcomes (pain, mobility, anxiety/depression, fatigue, peer relationship, physical function, sexual function) are available and can be collected as part of the European Registry for Rare Bone and Mineral conditions (https://eurr-bone.com). This EU initiative will provide extended phenotype information and increase knowledge about rare bone disorders.

Limitations: The quality of the patient interview is critical for the successful diagnostic support. Many PROMs questionnaires
are validated and tested in a population-based setting and they are preferable over non-validated questionnaires. However, regular, systematic collection and assessment of PROMs in a clinical setting takes time and resources, which may not be available to doctors and patients alike. Self-reported information may not be well formulated to provide sufficient levels of detail, therefore, recall bias and other uncertainties have to be taken into consideration.

Strengths: PROMs questionnaires are widely available and at relatively low costs, and they are easy to administer, allow for repeated assessments and may use different formats (in person, postal, telephone, or electronic).

1.3 Fracture History

Family and personal fracture history are strong risk factors for fragility fractures in humans of all ages and can give hints to frailty in the elderly and genetic disorders and abnormalities in children. This reflects the genetic component of risk for fracture, particularly for hip fragility fracture (30).

Predicting the ‘first fracture’ is still challenging, since the majority who fracture do not have osteoporosis (31). A first fracture of any type doubles the risk of a new fracture (32). The timeframe for a new fracture is partially dependent on age and type; for a first fracture in young adulthood, the next may be 20 years ahead, but for an octogenarian, 2-3 years. Stress fractures – including both fatigue fractures (from abnormal, or repetitive loading on normal bone) and insufficiency fractures by normal loading on abnormal bone (33) - are important events in a patient’s history and should be an additional indication for a thorough clinical exploration for potential secondary causes (34). Fracture type and location are of particular relevance. Lower limb and vertebral fractures are typical for young children with osteogenesis imperfecta, vertebral fractures associated with back pain in acute lymphoblastic leukaemia and distal femur fractures in immobilized persons.

Limitations: Recall of elderly patients fracture history may be poor. Silent vertebral fractures can also come with little or no symptoms or they may be non-specific and therefore prone to be misinterpreted or overlooked (35).

Strengths: Information is easily ascertained in a healthcare setting and the well-established link with family history and previous fracture should be sufficient to merit bone characterization and potential pharmacological and/or non-pharmacological musculoskeletal management via a Fracture Liaison Service (FLS) (36).

In mouse models, most studies describe changes in material properties and histology at a certain timepoint (37). Therefore, fracture history for an individual mouse will not be evaluated. However, in the context of mouse strains, probability and time until a fragility fracture occurs might provide important information.

In the zebrafish skeleton, ribs and fins should be given attention when analysing fractures. Zebrafish models for osteogenesis imperfecta e.g. show recurrent fractures in the ribs and fins (38-40). Rib fractures can be evaluated through life using radiographs, as well as analysing the fins under a transmitted light microscope. Fracture recurrence can be annotated longitudinally.

1.4 Functional Tests

The functional assessment of an increased risk of fall via the muscle-bone unit involves evaluation of a) muscle force using tools such as dynamometer, leg press and chest press; and b) physical performance using tools such as 30sec or 6min walk test (gait speed test), chair rise test, short physical performance battery (SPPB) and timed-up-and-go test (TUG) (reviewed in detail elsewhere) (41).

In children, the chair rise test, the 30sec or 6min walk test or the BOT™-2 (Bruininks-Oseretsky-Test of Motor Proficiency, second edition) test are commonly used.

Limitations: The results of the functional tests are largely influenced by the presence of chronic diseases and the patient’s cooperation as well as trained health care personnel.

Strengths: The dynamometer and the gait speed tests can be of greatest utility given the fact they can be used in research settings, in specialist clinical settings and in primary care settings at very low expenditures. These techniques provide valuable information on muscle mass and function, important determinants of falls and fragility fracture risk.

In mice, gait analysis can be used to detect abnormalities in speed, stride length, and limb-force profile (42). This technique has been used to measure altered stride length, velocity and limb angle after fracture fixation in mice (43). For muscle mass and strength assessment, multiple methods such as grip strength test (44), wire hang test (45), treadmill test (46), vertical pole test (47) and swimming endurance (48). Additionally, invasive methods include in vitro and in situ muscle force measurement (49).

Adult swimming behaviour analysis in the fish provides information on how the skeletal system is functioning as a whole (bone and muscles), with potential measurements of angle achieved during the swim, velocity achieved after tail propulsion, as well the time that it takes for exhaustion and induction of fractures (50).

1.5 Fracture Risk Prediction Tools

For a potential prediction of future fragility fractures, information gathered from the above described tools can be used with risk calculators that combine several risk factors, with or without BMD testing, e.g. the Fracture Risk Assessment Tool (FRAX®) algorithm1, the Garvan Fracture Risk Calculator2 and QFracture®. These tools provide a valuable risk stratification for the screening and management of osteoporotic patients (51). As an example, the FRAX®-based community screening in the elderly is increasingly used to provide individualized 10-year probability estimates of hip and major osteoporotic fractures (52). However, to date, there is no consensus on the discriminative ability of these tools to predict fragility fracture risk, except FRAX® with BMD, Garvan with BMD and QFracture® (53). Furthermore, the holistic approach of data collection together with physical and clinical measurements could help the construction of frailty index scores (54, 55) to

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1https://www.sheffield.ac.uk/FRAX/.
2https://www.garvan.org.au/bone-fracture-risk.
3https://qfracture.org/.
identify subjects at higher risk of fragility fractures (56), and mortality (57).

In children, such prediction programs have not been developed since the underlying conditions vary in nature; osteoporosis can be transient (e.g., acute leukaemia) or permanent (genetic). For example, vertebral fractures may spontaneously reshape in a leukemic child if the remaining growth potential suffices but this would be highly unlikely in a child with osteogenesis imperfecta (58).

Limitations: Some tools might be less representative for a number of important factors, such as probably an individual bone turnover. A lack of medical history data or the number of prior fractures might result in over- or underestimating a person’s personal risk.

Strengths: Community screening is more easily feasible and patients may be more adherent to bone-active treatment options in view of numer risk estimation.

In a quadrupedal mouse model, studying bipedal fracture risk and the link between muscle mass/strength and falls is difficult. However genetically modified models, as well as induced fracture models, allow for the study of changes in motion and function of the muscle bone unit which may provide insight into human cases.

Zebrafish fractures, their numbers and recurrence can be easily evaluated in vivo and longitudinally. As in mammals, fractures that happen early in life would indicate higher risks of fracture recurrence in zebrafish. Nevertheless, there are no estimates available, yet, for fracture risk predictions in zebrafish.

### 2. Bone Density and Imaging - 2D

Many different imaging modalities have been used to quantify bone density, strength, fracture risk and remodelling (Table 1). Some of these methods are specific for the human, but many can be used as well (in modified form) for animals (Figure 2).

Essentially, imaging methods can be 2D (slices or projections) or 3D. In this section we focus on the 2D imaging methods while the next section deals with 3D methods.

Different options are available for 2D bone assessment based on imaging in humans, which include plain radiography, bone densitometry by dual energy x-ray absorptiometry (DXA), bone scintigraphy, as well as vertebral fracture assessment (VFA) and trabecular bone score (TBS) based on lumbar spine DXA. The different imaging modalities have specificities in their local availabilities, as well as varying advantages and disadvantages depending on the technology, like radiation exposure, spatial resolution and the information that can be obtained.

#### 2.1 Plain Radiography

Conventional and digital x-rays are widely available and are frequently used as the first-line overview for imaging almost all pathological changes in the bone e.g. to assess bone structure and morphology in case of a suspected vertebral fracture. The main feature of osteoporosis in radiographs is increased radiolucency of the trabecular bone and cortical thinning, though this is mostly subjective and with low specificity, found at advanced stages of osteoporosis when bone mass is substantially reduced or bone mass accrual was insufficient as in osteogenesis imperfecta (OI) (59), and shows other mechanical or inflammatory changes of the vertebrae.

The Genant classification of vertebral fractures has been implemented using a semi quantitative technique (60) in five subtypes (OFI-5) (61) based on lateral vertebral imaging with a relatively low interobserver variation [see Vertebral Fracture Assessment (VFA)].

Limitations: The biggest disadvantage of plain radiography for assessing changes in the bone structure is the 2D nature, resulting in superposition of three-dimensional structures consisting of soft tissue and hard tissue onto a 2D plane. Thus, the interpretation can be difficult due to the superposition of shadows (62, 63). Another drawback is the limited resolution (order 200 microns) and the inability to discriminate between low bone mass and mineralization defects. As for all the techniques involving x-rays, there should be careful consideration between examination outcome and radiation dosage.

Strengths: Nevertheless, plain radiographs are widely available and some additional software techniques for bone density estimation from radiographs are under development. Generally, radiography is the first assessment due to the wide availability of the equipment, and the low cost (64). Radiographs may also provide an initial differential diagnosis covering also scoliosis assessment and other diseases of the spine presenting with back pain.

In mouse, 2D radiography is a highly sensitive method to study bone properties. The x-ray microradiography imaging is a useful tool for phenotyping. With this technique an X-ray tube with a small spot size (around 10 microns) is used that enables magnified projections of bone details. It can be used to assess changes in bone size and cortical thickness, and if used with appropriate standards, it can also provide a quantitative measure of mineral content (65). It also has the ability to detect cortical thinning and bone loss as seen in humans suffering osteoporosis (66). Lateral x-ray imaging has been applied in high-throughput format to identify bones with altered length and mineral content (66). It has the benefit of being fast and non-destructive, but drawbacks include that it only provides a two-dimensional image and may be affected by poor or inconsistent positioning of the animal or bone.

In zebrafish, radiographs are useful for rapid evaluation of skeletal deformities and bone density. As an example for the power of the technique, Fisher et al. have identified the zebrafish mutant Chihuahua (chi) (mutation in the ε1 chain of collagen type I) through a zebrafish forward genetic screening in which the authors leveraged from radiographies to screen a high number of adult zebrafish for skeletal abnormalities (67). Radiographs allow longitudinal studies of the zebrafish spine. Imaging takes a few seconds, allowing anesthetized zebrafish to be imaged without water, and followed by full recovery. However, due to the small size of the zebrafish bones, many aspects of bone morphology, microarchitecture, and mineralization, are limited in radiographic analysis, while μCT captures all these metrics.
| Imaging technique | Human | Mouse/rat models | Zebrafish models |
|-------------------|-------|-----------------|-----------------|
| **Strengths**     |       |                 |                 |
| Plain radiographs | Widely available | 2D analyses availability | Longitudinal skeletal assessment |
|                   | Additional density estimation in development | Potential superposition | Detailed aspects of bone morphology and density are not captured due to the imaging resolution, overlay with soft tissues, and small bones in zebrafish |
|                   | Low cost |                |                 |
| Moderate radiation dose | Artifacts from bone fractures, osteophytes, vascular calcifications and other superpositions | 2D information only | Most suitable method for BMD measurement in small animals |
| DXA               | Low radiation |                | General anaesthesia needed |
|                   | Fast and highly reproducible measurements | Artefacts from bone fractures, osteophytes, vascular calcifications and other superpositions | Poor or inconsistent positioning of the animal or bone. |
|                   | Widely available and full automatization | 2D information only | Poor edge detection and accuracy for very small animals (<50 gr) |
|                   | WHO/ISCD definition for osteoporosis/osteopenia Individual longitudinal monitoring possible | No differentiation of trabecular vs cortical compartments | Accurate positioning of the animals and placement of the region of interest can be challenging |
| TBS               | Non-invasive | No direct relation to fracture risk published | Measurements affected by size and weight of the animal |
|                   | Tool for trabecular bone structure Discrimination in secondary osteoporosis e.g. in diabetes mellitus | Improvement of risk prediction via FRAX Potential artefacts |
|                   | Information on vertebral fractures Low radiation exposure | Lateral positioning of patient sometimes difficult |
| VFA               | Transportable | No WHO definitions of osteoporosis/osteopenia Many different devices – no standardization Individual monitoring difficult | N.A. |
|                   | Quick | No direct translation to bone structure | N.A. |
|                   | Non-invasive |               | N.A. |
|                   | Radiation free |               | N.A. |
|                   | Inexpensive |               | N.A. |
|                   | It can be used apart from specialised centres |               | N.A. |
| Bone scintigraphy | Widely available | Potential false positive results inferior to SPECT in 3D questions | Mainly use of SPECT (see Single-Photon Emission Computed Tomography (SPECT)) N.A. |
|                   |               | Mainly use of SPECT (see Single-Photon Emission Computed Tomography (SPECT)) | N.A. |

DXA, dual energy x-ray absorptiometry; FRAX, fracture risk assessment tool; QUS, quantitative ultrasound; SPECT, Single-photon emission computed tomography; TBS, trabecular bone score; VFA, vertebral fracture assessment.

N.A., not applicable.
2.2 Dual Energy X-Ray Absorptiometry (DXA)

DXA provides a two-dimensional (2D) representation of bone, but also information about body composition including lean and fat mass. Measurement sites include the lumbar spine, hip, radius and whole body. Though anterior-posterior scans are generally obtained, lateral spine scanning is also performed to assess vertebral morphology and fractures (see Vertebral Fracture Assessment (VFA)). The DXA image comprises a series of pixels containing information about mineral content. Total mineral content within a region of interest is defined, from which bone mineral density [BMD, in g/cm², also often noted as areal BMD (aBMD)] is obtained after dividing bone mass by bone area. aBMD measured by DXA predicts fracture risk in adults (68), for which this method is widely used for clinical and research purposes. In clinical settings, aBMD is compared to a young reference cohort of the same ethnic background and sex, generating a T-score. The International Society of Clinical densitometry (ISCD) defines osteoporosis in adults as a T score ≤2.5, representing 2.5 standard deviations below the young reference mean value (69). Another score reported from DXA measurements is the Z-score. This score quantifies the number of SDs above/below the mean value of an age and sex matched population. This score is not used for the diagnosis of osteoporosis but provides information about an individual’s fracture risk compared to peers (70). Volumetric bone mineral density (mass per volume, (vBMD)) also noted as bone mineral apparent density (BMAD) is strongly correlated with bone strength in experimental studies (71). Although DXA just provides an ‘areal’ density, this remains the most common technique of assessing bone strength clinically. BMD thresholds also contribute, but less than in adults, to the diagnosis of osteoporosis in children (58, 72). In children, aBMD data require adjustment for body size to avoid misinterpretation from size artefacts, by using lumbar spine BMAD and total body less head (73). The ISCD definitions of osteoporosis in children are mainly based on the presence of fractures (74). Hip structural analysis (HSA) (75, 76), has been developed to derive other parameters related to bone strength, for example by calculating femoral neck width (77). Finite element analysis has also been applied to hip DXA images, which may provide additional information on fracture risk (78). In addition, current DXA devices enable specific morphological features to be assessed such as vertebral...
fractures and osteophytes. Due to the strong relationships with fat and particularly lean mass, DXA scans provide regional body composition measures which are particularly useful in evaluating android and gynoid fat distribution (79). In fact, DXA is considered the gold standard method for body composition assessment in clinical practice due to its advantages of high accuracy and precision, low cost, low radiation dose and short scan time. It has a variety of clinical applications, such as diagnosis and follow-up of lipodystrophy and sarcopenia, as well as being widely used in research studies of body composition. Further technologies based on DXA are analyses of fractures by Vertebral Fracture Assessment (VFA), and the trabecular bone score (TBS) described in section Vertebral Fracture Assessment (VFA) and Trabecular Bone Score (TBS).

Limitations: BMD can be artificially elevated by collapsed vertebrae or mineral deposits at sites that do not contribute to bone strength, such as osteophytes and aortic calcification. In addition, due to its 2D nature, DXA is unable to capture the complex 3D morphological characteristics of skeletal elements. For instance, the trabecular vs cortical compartments cannot be differentiated and DXA gives no information on the bone microarchitecture (80). Moreover, DXA measurements are not corrected for skeletal size, hence DXA underestimates BMD in humans and animals with short stature, and overestimates BMD in those with tall stature. Such artefacts are not generally corrected for in adult medicine and in many animal studies. In children or growing animals alike, interpretation of DXA results requires adjustment not just for age and sex but also for body or bone size, and skeletal maturity (bone age or pubertal status).

Strengths: DXA scans use very low radiation doses and a fast-scanning mode, making this method suitable for research as well as clinical use. Derivation of commonly used measures such as BMD is fully automated and highly reproducible, enabling small changes to be detected in longitudinal studies.

In mice, as in humans, DXA is the most commonly used method for measuring BMD (81). DXA has been demonstrated to be accurate and precise in measuring total bone and bone mineral content in mice (82). It has been used to characterise the bone loss in multiple models of post-menopausal osteoporosis in mice (83). Benefits in animal characterisation include the ability for live imaging, its low cost, relatively fast speed, and low-radiation emission. Limitations of the application of DXA in animal models are the low resolution of the technique, and the need for correct (sometimes repeated) positioning. For zebrafish, techniques of bone density measurements are reflected in section MicroCT (μCT).

2.3 Vertebral Fracture Assessment (VFA)
VFA uses lateral DXA imaging of the thoracic and lumbar spine for the presence of vertebral fractures (VF). Images can be obtained at the same time as areal (aBMD) measurement. The radiation exposure is lower than in plain radiographs of the spine (84). According to the ISCD, Genant’s semi-quantitative fracture assessment is the method of reference for the diagnosis of VF on VFA or other lateral spine imaging (74, 85–87).

Limitations: In some devices, the analysis requires a lateral positioning of the patient, which is sometimes not feasible. This limitation can be solved by the use of a “C-arm”, allowing supine lateral spine imaging. The upper thoracic vertebrae (Th4 to Th7) might be poorly visible.

Strengths: The low radiation exposure of VFA is an advantage in all, but especially in paediatric patients (according to the latest ISCD paediatric recommendations (74, 88), as well as the combination of both aBMD and VFA in one session.

In mouse, VFA is not used in the mouse skeletal phenotyping.

In zebrafish, vertebral fractures can be assessed longitudinally through radiographs and post-mortem through μCT and whole mount staining (Alizarin Red S staining). Although vertebral fractures in zebrafish are not commonly observed, compressive forces applied ex vivo, anteroposterior at the vertebral column and visualized using μCT, have demonstrated points of stress in the vertebrae where it is subjected to fracture in zebrafish (89). Recently, a non-invasive method to induce fractures in zebrafish has been established. By using physical pressure applied to the fin rays of the anaesthetized fish, one can easily cause fin fractures (90). This allows the assessment and the study of fractures from the initial moment that they happened.

2.4 Trabecular Bone Score (TBS)
Trabecular bone score (TBS) is a texture-based index that provides an indirect assessment of trabecular bone microarchitecture. It is calculated based on the pixel gray-level variations in lumbar spine DXA images (78, 91). While this index is increasingly used in adult human patients, there are no animal studies to date, except ex vivo comparisons with porcine vertebrae (92). TBS provides additional information on fracture risk and is mainly used in secondary osteoporosis, e.g. in diabetes (93) and ankylosing spondylitis (94). However, the proportion of risk prediction in a more general osteoporosis approach warrants further studies, and also depends on the software used (TBS iNsight®, Version 4.0) with a 54% (OR 1.54; 95% CI, 1.18 to 2.00) increase of having a major osteoporotic fracture (MOF) for each standard deviation decline in TBSv4.0 values (95). Chronological age and TBS are related; significant age-related changes seem to occur with a turning point to higher TBS values at age 8 in girls and age 10 in boys (96). The use of the TBS has not yet been sufficiently explored or recommended for clinical use in children, (see the current ISCD position at iscd.org/learn/official-positions, last access Dec 2020). Assessment of TBS at other bone sites than the lumbar spine might be an interesting development (97), also in view of comparisons with animal measurements and high-resolution (peripheral) quantitative computed tomography HR(p)QCT or bone biopsies including state-of-the-art histomorphometry.

Limitations: TBS is used as an add-on tool to DXA-scans; patterns for specific osteoporosis risk prediction are warranted. An independent contribution of TBS to fracture prediction seems to be small (98), and potential artefacts can be due to collapsed vertebrae.

Strengths: The respective software is a widely used non-invasive tool for indirect assessment of trabecular bone structure based on already existing compatible DXA scans. TBS might allow for a discrimination of patients at risk, e.g. in secondary osteoporosis, where DXA alone does not.
The inclusion of TBS into the Fracture Risk Assessment Tool (FRAX®) may improve the fracture prediction beyond FRAX® without TBS.

In mice and zebrafish, TBS or similar scores are not currently used.

2.5 Quantitative Ultrasound (QUS)

QUS provides a measure of bone quality and quantity (99). Broadband ultrasound attenuation (BUA, dB/MHz) reflects anisotropic characteristics of trabecular bone, and speed of sound (SOS, m/s) refers to the division of sound waves by the length of the bone and transmission time. Some devices combine BUA and SOS values to provide a quantitative ultrasound index (QUI) or stiffness index (STI). Though providing a distinct measure to DXA-evaluated BMD, QUS and DXA have similar predictive value for hip fracture risk in elderly populations (100). QUS is used to assess easily accessible bones like the calcaneus, which is the most widely used measurement site, and patella, tibia, metatarsal bone at weight-bearing sites, as well as phalanges and radius (101). Clinically, QUS is used as a screening tool for osteoporosis. QUS has also provided major insights for genetic discovery, through its incorporation in UK Biobank, based on estimated bone mineral density (eBMD, g/cm²) derived as a linear combination of SOS and BUA (102). The device is not recommended for routine use in children and adolescents.

Fracture sonography is a special field of application of medical ultrasound diagnostics (sonography) for the detection of bone fractures. In addition, there are other applications of bone sonography, such as osteoporosis diagnostics and for the representation of callus. In patients younger than 12, proximal humerus or clavicle fractures can be visualized by ultrasound due to the changes at the bone surface (103).

Limitations: As WHO definitions of osteoporosis and osteopenia require DXA measurements (104), confirmation of QUS findings by DXA measurement is needed. A lack of standardization hampers result comparison, in view of the many different QUS devices available and of the influence of environmental conditions (temperature etc.). Furthermore, in contrast to DXA, the precision of QUS is insufficient for monitoring individual patients (101). QUS cannot assess bone structure.

Strengths: Advantages of QUS include being transportable, quick, non-invasive, radiation free, inexpensive, and useful for large population screening studies even apart from health care centres.

In mice and zebrafish QUS is not used.

2.6 Bone Scintigraphy

Bone scintigraphy detects an increase in osteoblastic activity or vascularization, which may be associated with osteoporotic fracture or localized bone lesions. Radionuclides such as technetium-99m [99m Tc], often linked to a bone-avid tracer molecule such as bisphosphonates, e.g. 99mTc-methylene diphosphonate (MDP) emit gamma-radiation in proportion to their attachment to a target structure. This technology may supplement radiographs with additional information about recent/old fractures or may identify radiographically occult injuries and differential diagnoses such as metastatic disease (63).

Limitations: Bone scans are a sensitive technique, but may produce false positive results and cannot determine the extension of a fracture, whereas SPECT is superior in the detection of vertebral fractures (see section CT-Based Techniques).

Strengths: Bone scintigraphy is widely available in specialized nuclear medicine departments and may be used to address clinical questions not only for oncological diagnosis. It can discriminate recent from healed spinal fractures and demonstrate evidence for radiographically difficult to assess fractures, e.g. atypical femoral fractures (105). Furthermore, positive tracer uptake is reported in areas that subsequently develop osteonecrosis of the jaw (ONJ) (106).

In mice, radioactive methods are widely used, but mainly in context with SPECT (see section CT-Based Techniques).

In zebrafish, scintigraphy is nor used mainly due to the water-based environment.

3 BONE DENSITY AND IMAGING - 3D

Development of three-dimensional (3D) methods for bone imaging allowed for new approaches in bone phenotyping, such as computed tomography (CT), single-photon emission computed tomography (SPECT) and magnetic resonance imaging (MRI) (Table 2). Implementation in clinical use and research protocols depend on local availability and technical knowledge. For scientific purposes, international cooperation of researchers might be an additional benefit by bringing bone scientists together.

3.1 CT-Based Techniques

3.1.1 Computed Tomography (CT)

Computed tomography is a sectional imaging method that allows a representation of soft tissues, bones and vessels. Thanks to the spiral technology (except QCT), clinical CTs produce small isotropic voxels, which enables a high spatial resolution in any spatial direction. The multiplanar slicing also allows sagittal and coronal representations of high quality and 3D visualization provides structural and morphological information. The x-ray-based imaging technique is widely used for characterization of degenerative changes, vascular and soft tissue calcifications (84).

The voxel size varies according to the method, e.g., 250 - 1000 µm for clinical whole-body CTs, 50 - 80 µm for HR-pQCT [see High Resolution Peripheral Quantitative Computed Tomography (HR-pQCT)] and 10 - 30 µm for microCT (see MicroCT, used for instance for microscopic bone structure analysis on bone biopsies) (107).

Quantitative CT enables the measurement of volumetric BMD (vBMD) at the spine and any bone, and allows separate evaluation of cortical and trabecular bone. Further details are available in Quantitative CT.

Limitations: CT scans involve high radiation exposure. A direct comparison with DXA is not possible.

Strengths: CT scans are widely available and can be used for the characterization of morphological changes and differential diagnoses.

In mice and zebrafish, clinical CT scanners are not used due to the low resolution (See μCT section MicroCT).
3.1.2 Single-Photon Emission Computed Tomography (SPECT)

Non-quantitative bone scintigraphy using 99mTc-MDP may be combined with CTs for Single-photon emission computed tomography (SPECT)/CTs. Standardized uptake volume (SUV) is also used for bone metabolism measurements. The range of SUV in normal lumbar spine is roughly coherent with 18F-fluoride in positron emission tomography (PET). In addition, it correlates positively with Hounsfield units (HU) of the lumbar spine and negatively with age (108). As a fusion method SPECT/CT has been shown to be superior to SPECT alone in the identification of vertebral lesions especially in distinguishing acute fractures in a multiple fracture setting (109) and is consistent with MRI in patients with osteoporotic vertebral compression fractures (109).

Limitations: SPECT should be avoided in children unless oncologic or inflammatory conditions are suspected (110).

| Imaging technique | Human | Mouse/rat models | Zebradfish models |
|-------------------|-------|------------------|------------------|
| **CT** | Widely available | High radiation exposure | See µCT below | Low resolution |
| | Morphological use | No direct comparison to 2D methods (e.g. DXA) | See µCT below | N.A. |
| | Concomitant differential diagnosis | | N.A. | N.A. |
| **SPECT/CT** | Correlation of skeletal standardized uptake values (SUVs) and BMD possible | Not useful in children due to radiation and inflammation concerns radiation exposure | | |
| | | Good spatial resolution | Radiation | |
| | | Useful for bone growth and repair | N.A. | |
| | | Non-invasive and longitudinal tracking of changes | N.A. | |
| **QCT** | Volumetric bone density information | Considerable costs | See µCT below | See µCT below |
| | Can be used for continuum FE models | Higher radiation dosage | See µCT below | See µCT below |
| **pQCT** | Evaluation of cortical and trabecular bone density, structure and strength | | See µCT below | See µCT below |
| Relatively low radiation dose | | Long operational time | See µCT below | See µCT below |
| | | Thresholding and difficulties with standardisation at distal sites | See µCT below | See µCT below |
| | | Size artefacts by partial volume effect | See µCT below | See µCT below |
| | | Needs adjustment for bone length | See µCT below | See µCT below |
| **HR-pQCT** | Only existing non-invasive imaging method obtaining bone microarchitecture | Only for distal extremities | See µCT below | See µCT below |
| | | Fast and safe | | |
| | | Low radiation dose 3 μSv/scan | | |
| | | Good reproducibility No side effects | | |
| **MicroCT (µCT)** | In bone specimens, fast and non-destructive assessment | Ex vivo use in humans only | Most suitable method for skeletal measurement | High radiation allows only ex vivo bone assessment |
| | | Excellent reproducibility and accuracy | | |
| | | Lack of specificity for soft tissues | Stabilization required | |
| | | | Radiation exposure | |
| **MRI** | No radiation exposure | No direct comparison to 2D methods (DXA) | Longitudinal assessment | Bones and muscles can be visualized |
| | | Widely available | Long scanning time | Aquatic flow cell system is needed for in vivo scanning |
| | | | Low resolution due to small sample | Low resolution |
| | | | | Difficult in use |

CT, computed tomography; µCT, microCT; FE, finite element analysis; HR-pQCT, high resolution peripheral computed tomography; MRI, magnetic resonance imaging; pQCT, peripheral quantitative CT; QCT, quantitative CT; SPECT, Single-photon emission computed tomography. N.A., not applicable.
Strengths: The correlation of skeletal standardized uptake values (SUVs) and BMD suggests its use for clinical and research purposes (108).

In mice, SPECT scanners designed for pre-clinical models, can have a spatial resolution of <0.5mm due to pinhole and multi-pinhole collimators (111). This is useful when used in combination with CT scanning (SPECT/CT) which allows co-registration of the area of activity, and the skeleton, hence areas of new bone formation or high bone turnover (112). Multi-pinhole SPECT has successfully been used to track bone growth and repair in a mouse model for 12 weeks – specifically to track the temporal and spatial positioning of hydroxyapatite deposition in a bone defect mouse model (113). Benefits of this technique in mice are the non-invasive character allowing longitudinal tracking of changes in individual animals, which may reduce cost, animal numbers and inter-animal variability. Fast scan times (minutes) require less time under anaesthetic. However, exposure to radiation is required and this may be significant if repeated scans are taken.

In zebrafish, as for other radioactivity-based measurements, no special scintigraphy technology is available due to the water-based environment.

3.1.3 Quantitative CT
Quantitative CT (QCT) enables 3D imaging of bone in vivo while providing quantitative information about the spatial bone density distribution at a resolution of around 0.5 mm (114, 115). The possibility to calculate vBMD provides a true density measure of the whole bone cross-section, in contrast to areal BMD obtained from DXA. QCT images can also be used as the basis for Finite Element (FE) models (116, 117). With such models, the bone geometry is represented by a large number of sub-volumes (the 'elements'), typically one or a few mm³ in size. As the QCT resolution is not enough to resolve the trabecular or cortical microstructure, these models represent bone as a 'continuum' in which the bone microstructure is homogenized and represented by its density only (116, 118). Using such models, it is possible to calculate the bone stiffness, the stresses in the bone and the bone strength for a specified set of forces ('boundary conditions') (116, 119, 120). Such loading conditions can represent physiological loading (e.g. vertebral forces in the spine, or hip joint forces) to calculate physiological stress values in the bone tissue, or can represent loading conditions that typically lead to fracture (e.g. a fall) to calculate bone strength. QCT can be used in clinical trials aiming at quantifying the effects of drugs or other treatments on bone strength or in research studies correlating e.g. nutrition, lifestyle or genetic factors with bone strength. In addition, images can be analysed as an "add-on" screening tool in cases where QCT images are made for other reasons, e.g. during virtual colonoscopy or cardiovascular research. In which vertebrae are in the field of view (116, 121, 122). In both human and animal, FE modelling of bone in young versus older ages may differ. In particular, the growth plate can lead to artefacts, as these may appear as gap regions. In addition, the tissue mineralization in young versus old bone can differ, which may require using different empirical relationships to translate density to material properties.

Limitations: QCT images involve considerable costs, radiation dose and operational time. QCT based FE is not suitable as a screening tool (120). It is not possible to account for bone microstructure (other than its mere density), and thus empirical relationships between bone density and material properties are needed. Routine use of QCT in children is not established.

Strengths: A particular strength of QCT-based continuum FE models is that the technique is based on well-validated mechanical principles. This is in contrast to stochastic models is that the technique is based on well-validated mechanical principles. This is in contrast to stochastic models which are less suitable for small animals because of the smaller size of their bones (123). Thereby, the assumption that the bone microstructure can be homogenized to a continuum becomes less accurate. The resolution is not enough to resolve thin cortices. For small animals, the use of high-resolution micro-finite element analysis (micro-FE) therefore is more appropriate.

See section 3.1.6 MicroCT (µCT).

3.1.4 Peripheral Quantitative CT (pQCT)
Peripheral quantitative CT (pQCT) is used to image the radius and tibia. The spatial distribution of fat, muscle and bone within the cross section is obtained after applying density thresholds for each of these tissues. At diaphyseal sites, cortical bone indices are obtained including cortical vBMD, periosteal circumference and cortical thickness, as well as muscle and fat cross-sectional area (124). In addition, estimates of cortical bone strength can be generated, such as cross-sectional moments of inertia. At the distal radius (i.e. metaphysis), trabecular vBMD is obtained at a pre-defined central region of the medullary space.

Limitations: A disadvantage is the poor standardisation at distal sites (positioning of reference line relative to the growth plate), which in children and growing animals limits reproducibility. In addition, in humans and animals alike, the partial volume effects (the situation where a voxel volume is only partially filled by bone tissue) lead to size artefacts, i.e. cortical vBMD is artificially reduced in individuals with reduced cortical thickness as a larger part of the voxel is not within the bone tissue. Adjustment for bone length may be required for subjects with tall or small stature, to correct for bone size.

Strengths: The ability of pQCT to evaluate cortical and trabecular vBMD, structure and strength variables separately represents an important advantage compared to DXA scans, and has provided the basis for separate genetic studies of cortical vBMD (125), trabecular vBMD (126) and cortical thickness (127). This may be even more important for the assessment of bone conditions where the relations between cortical and trabecular bone are shifted (128). Furthermore, pQCT scans are associated with a relatively low radiation dose, making this method suitable for clinical studies.

In mouse models, pQCT is useful to accurately measure both trabecular and cortical vBMD, as well as predicting bone strength (129). pQCT has been shown to be accurate and precise in mouse models, confirmed by both µCT and histology (130). It can be
used in vivo on live animals. There is however the potential for errors in vBMD measurements based on specimen thickness and positioning (129, 131).

See section 3.1.6 MicroCT (µCT).

### 3.1.5 High Resolution Peripheral Quantitative Computed Tomography (HR-pQCT)

Like the classical pQCT, HR-pQCT assesses bone microarchitecture in the cortical and trabecular compartments, but with higher resolutions of 82 µm for the first-generation devices and 61 µm for the second-generation devices (132, 133). Depending on technical developments, the clinically standardized volume of interest (VOI) is to be set to 9.5 mm in length for the first generation and 10.2 mm for the second-generation of devices. In adults, the beginning of the VOI is situated at a fixed distance proximally from a reference line through the joint at the distal cortex of radius and tibia (134). Thus, in taller individuals, the VOI is relatively more distal and has greater cross-sectional and trabecular areas as well as thinner cortices. For HR-pQCT imaging, the participant’s extremity is placed in a cast, which reduces motion. The cast is then inserted into the device and is fixed in position, while the x-ray source rotates around the extremity. The scanning time is around 2.5 min for these standard measurements. Cortical and trabecular vBMD, cortical and medullary cross-sectional area, cortical thickness, cortical porosity, trabecular spacing (Tb.Sp), trabecular number (Tb.N = 1/Tb.Sp) and Tb.Sp standard deviation are typically reported and satisfactorily accurate (135). For the first generation HR-pQCT only Tb.N was directly measured, while other parameters were derived from Tb.N and BV/TV using standard methods adapted from histomorphometry (134). For the second generation all parameters were measured directly. HR-pQCT is used for longitudinal assessment of changes in bone microarchitecture, e.g., of age-related bone loss (136). However, bone loss at the endocortical bone surface results in trabecularisation of the inner cortex and errors in estimation of cortical and trabecular bone loss (137). More recent software permits to transfer the initial endocortical contour on the follow-up scans and assess bone loss in the cortical and trabecular compartment. HR-pQCT scans may be used for micro-FE analysis to estimate bone strength (138, 139). In addition, vascular and tissue calcifications are targets of HR-pQCT measurements and currently under development.

Limitations: For now, HR-pQCT is available only for distal extremities, although with the second generation scanning of areas up to the knee and elbow has become possible as well (140, 141). As HR-pQCT is sensitive to movements, some scans have to be excluded due to poor quality. The occurrences of movement artefacts are higher for radius than for tibia scans, probably because the sitting position is less comfortable. Furthermore, the necessity to stay in a resting position without any movement of the scanned limb might be challenging especially in the elderly patient with tremor and people with pain in joints may have trouble being positioned. Any x-ray based method is not conclusive in areas with metallic or other implants (133).

In the structural analysis, identification of the endocortical limit between cortical and trabecular compartments by software is challenging (142). Thus, estimates of cortical thickness and area and that of trabecular area may be inaccurate. Manual analysis is time-consuming and has only moderate-to-good reproducibility. However, the endocortical limit on the HR-pQCT scan is not always evident even for experts and the manual analysis does not improve its identification. Several algorithms assess cortical porosity, but they are based on unverified assumptions (143–145).

Strengths: HR-pQCT is the only existing non-invasive imaging method obtaining bone microarchitecture in clinical studies. It is fast and safe (low radiation dose 3 µSv/scan), has good reproducibility (<1% for vBMD, <4% for structural variables) and gives no side effects (146).

HR-pQCT permits to assess the structural basis of the effects of the risk factors of osteoporosis (e.g., sex steroid deficit), predict fragility fracture and the effect of anti-osteoporotic treatments on bone (147–149). The biomechanical parameters assessed by micro-FE can improve fracture prediction (148). Scanning protocols for children are being developed.

For mouse and zebrafish context see section 3.1.6 MicroCT (µCT).

### 3.1.6 MicroCT (µCT)

Micro-computed tomography (MicroCT or µCT) is a high-resolution imaging modality that offers quantitative analysis of trabecular and cortical bone morphology in animals and human specimens. First introduced in the late 1980s (150), µCT now has become the gold standard for the evaluation of bone microarchitecture throughout species.

The method, such as the other CT-methods, is based on the use of x-rays to create cross-sections of an object. For µCT voxel sizes lower than 10 µm can be obtained (151). The degree of x-ray beam absorption is recorded, so that the 3D structure of the object can be visualized and numerous bone structural parameters can be quantified with a high degree of accuracy, such as cortical and trabecular vBMD, cortical thickness, and if used at high enough resolution/voxel size, cortical porosity.

As an ex vivo imaging modality in humans, µCT enables 3D characterization of small bone specimens acquired from bone biopsies, or of larger cadaveric specimens such as vertebrae (152, 153). Studies have shown that µCT can reproducibly quantify 3D microarchitecture of the trabecular and cortical bone in iliac crest biopsies, demonstrating significant changes in 3D trabecular structural parameters in postmenopausal samples, including a decrease in BV/TV, an increase in trabecular separation and a shift from platelike to rodlike structure (154). µCT quantification of bone structure from iliac crest biopsies is an important end point in longitudinal drug efficacy studies (155). Assessment of 3D trabecular and cortical structural characteristics may improve our ability to understand the pathophysiology of osteoporosis, to test the efficacy of pharmaceutical intervention, and to predict bone biomechanical properties.

Limitations: Due to the high radiation exposure, the use in humans is restricted to ex vivo measurements and thereby limits the clinical application of µCT. High-resolution scans produce
large amounts of data that require support for data acquisition, processing and management. Even though a considerable limitation of the µCT technology is the lack of specificity for soft tissues, it can be combined with contrast agents for the visualization and quantification of soft tissues like vascular structures and bone marrow adiposity within the bone specimens (156, 157).

Strengths: Compared with histology, µCT has many advantages as larger volumes are analysed, 3D-measurements can be performed faster with higher resolution, excellent reproducibility and accuracy. The assessment of bone morphology is non-destructive and does not require fixing agents, enabling subsequent analyses of specimens for histology, mechanical testing and biochemical analysis.

In mice, µCT is a widely used method for analysing bones of small animal models in vivo and ex vivo, due to its high resolution, with the ability to achieve resolutions as small as 1-µm (158) and identifying body composition. Guidelines for µCT assessment of rodent bone specimens have been recommended including sample preparation, image acquisition, processing and analysis (159).

Ex vivo µCT can be used to measure cortical and trabecular vBMD, cortical thickness, and if used at high enough resolution/voxel size, cortical porosity (160). It has been used at high-throughput format to identify bones with altered BV/TV, trabecular thickness (Tb.Th) and trabecular number (Tb.N) (66) and can be used for longitudinal assessment of the same animal over time due to the non-destructive character. It can be performed on living animals, although long scan times do require large doses of anaesthetics and radiation (above 400 mGy/scan) can affect osteoblasts and subsequent evaluation of bone formation (161, 162).

In adult zebrafish, µCT is a well-established and widely used tool for the detection of skeletal abnormalities (13, 40, 163–165). Due to the small size of the bones in zebrafish, the visualisation of the skeleton through standard µCT has been mostly limited to skeletally mature animals. The use of contrast agents, such as AgN03, has been shown useful for the visualisation of earlier ages of the zebrafish skeleton, as well as for soft tissue [28]. 3D tissue mineral density (TMD) examines the amount of mineral per unit volume of bone tissue and is used to measure cortical TMD in zebrafish. TMD values of 450-600 mg HA/cm³ have been reported in the vertebrae of adult zebrafish (13) which is noticeably less than the TMD values of 800-1000 mg HA/cm³ in the cortical bone of adult mice (166) or human cancellous bone (167). These differences in TMD have been attributed to differences from human bone in material properties and mineralization dynamics (12) and as a possible reflection of adaptation to mechanical loading and bi- or quadrupedalism in terrestrial mammals (168). In parallel with TMD, values of the vertebrae length, area, volume, thickness and other measurements of shape are often used for phenotypic characterisation of the zebrafish vertebral column (13, 164, 169). The vertebral column, as a major skeletal structure of the zebrafish adult skeleton, is most commonly studied by µCT. However, it is also used for the analysis of other parts, such as the zebrafish craniofacial skeleton (11, 164, 170, 171). Semi- to full automation of bone segmentation from µCT imaging data would allow rapid and robust analysis. In this line, a supervised segmentation algorithm (Fish-µCT) enables segmentation of each vertebrae and profiling of phenotypic measures (13, 39).

3.2 Magnetic Resonance Imaging of Bone (MRI)

Magnetic resonance imaging is an intersectional imaging method. It technically uses a combination of a strong magnetic field (1.5-9T) and stimulation of protons by radiofrequency pulses. MRI provides high contrast resolution and better soft tissue display than computed tomography. Due to different imaging techniques, like fat suppression, it provides a high sensitivity for findings like periosteal edema and bone marrow changes as well as intracortical signal abnormalities (172). Frequent findings in acute and subacute vertebral fractures are vertebral edemas with a low signal on T1-weighted images (WI, using basic pulse sequences in MRI) and a high signal on T2-WI, and high signals on STIR (Short tau inversion recovery), while old fractures show the opposite (109). MRI has been analysed for “M-scores”, deviated from signal to noise ratios (SNR) in the vertebrae L1-L4 as compared to T-scores using DXA. The SNR in L1-L4 is negatively related to BMD, but the cut-off value for M-scores is still under debate (173). Some of the novel MRI imaging techniques are able to quantify bone composition and may generate precise phenotypes of bone changes related to age (174). Future developments should define calibration phantoms for routine imaging. Artificial intelligence (AI) algorithms may be used for existing images to identify patients at risk for bone fractures.

Limitations: MRI requires expensive equipment and training. A direct comparison to 2D DXA is not possible, and limited resolution is often not sufficient for morphological analysis. Especially high field MR scanners (over 7T) are not widely available and costly, therefore only accessible in well-equipped institutions. In general, this equipment is exclusively used for research purposes. MR examinations are time-consuming due to the longer scanning time and therefore more susceptible to motion artefacts, which can affect the accuracy of evaluations. Furthermore, MRI is very susceptible to artefacts caused by metallic implants, for example postoperatively in the case of spondylodesis, which in turn reduces image quality and makes partial evaluations impossible.

Strengths: MR technologies are widely available, at least in developed countries. There is no radiation exposure, therefore repeated and large areal scans are possible. Well-defined morphological tools may help to characterize significant changes in clinical work-up. A powerful strength of magnetic resonance is the excellent soft tissue imaging. It is superior for imaging muscle pathologies and, through special techniques such as the Dixon technique (175), for quantifying adipose tissue and muscle mass in a reasonable time frame, which might also add important information in connection with osseous pathologies. Furthermore, cartilage damage and degenerative as well as inflammatory changes in articular cartilage and intervertebral disc tissue can be identified and
quantified, which is not technically possible to the same extent using computer tomography.

MRI is a useful tool in mouse phenotyping as it allows concurrent imaging of soft tissue (cartilage, bone marrow, muscle, fat) and bone with good spatial resolution. MRI has been used to image bone injuries in mice with good distinction between the bone, soft tissue and injury sites, with a good signal-to-noise ratio (176). MRI is particularly useful for monitoring endochondral fracture healing, which involves a cartilaginous tissue callus (177). MRI has the benefit of providing 3D images, and allowing longitudinal assessment of single animals. Disadvantages include a long scanning time (up to hours), the potential for artefacts at the bone-soft tissue interfaces, and low resolution due to the size of the sample (176, 178).

MRI has not been widely used in zebra fish. However, recent studies demonstrated the use of the imaging technology for longitudinal and non-invasive studies. 3D scans covering the thoracic region of the same adult zebra fish at an isotropic voxel resolution of 31 µm allowed longitudinal studies of the zebra fish heart. Bone and muscles were observed with MRI (179). To overcome the limitations of the aquatic system, a flow cell system has been developed for MRI imaging, allowing to monitor the zebra fish during the scan and to fully recover the animal (180). However, the methodology needs to be further improved to establish it as a routine bone assessment in zebra fish.

4 BONE BIOPSY AND LOCAL MEASUREMENTS

Investigations at the tissue level have a long tradition for histology and several microscopic technologies which are important in clinical practice for the differential diagnosis of disease entities. However, new approaches will help to expand our understanding of bone properties using microindentation (see Microindentation) or compositional bone matrix analyses via quantitative backscattered electron microscopy imaging (qBEI) and vibrational spectroscopy (see Compositional Bone Matrix Analysis Using Quantitative Backscattered Electron Microscopy Imaging (qBEI) and Vibrational Spectroscopy) – these new approaches and their use in human and animal bone research is of increasing importance.

4.1 Histology

Histology of bone biopsies provide qualitative information about bone cells, matrix (e.g. the orientation of collagen fibres), mineralization and bone marrow. Evaluation of bone biopsy should comprise its histological (visual, qualitative) and histomorphometric (quantitative) assessment (181). The biopsy should be examined for the presence of mast cells and cancer cells infiltrating the bone marrow or the bone. It should be noted if the bone has the normal lamellar texture or if woven bone is present (182).

Limitations: Histology only provides information on the 2D structure of tissues and cells which can lead to an over- or underestimation of morphological features. However, the stacking of layers can be applied to regain 3D-information. It is a destructive method, and only the remaining parts of an embedded sample can be analysed with other techniques than the histological assessment.

Strengths: Histology is one of the most established and versatile methods to identify different types of tissues, and osseous cell components at high resolution. Various staining protocols are readily available for the detection of bone matrix alterations due to diseases or treatment.

In mouse and zebrafish studies, histology is widely used. As with human studies, it can provide information on cell type and number, bone matrix and mineralisation and help to characterize specific disease models.

4.2 Histomorphometry (Static and Dynamic)

In addition to specific histology, a histomorphometric evaluation of bone modelling and remodelling can provide quantitative information about mineralization disorders, metabolic bone diseases, and secondary bone diseases including cancer. “Static” bone histomorphometry (HM) consists in counting cells and measuring bone tissue components. For “dynamic” purposes, oral tetracyclines are administered separated by 10-12 days. Tetracycline is incorporated into new bone at the “mineralization front”, and its fluorescence allows for the assessment of bone turnover (183–185).

Histomorphometry from patients requires bone biopsies obtained standardly from the iliac crest under local or (in children) general anaesthesia.

Bone samples are processed without prior decalcification according to published protocols. The stains should allow the differentiation between mineralized bone tissue and osteoid, and the identification of bone and marrow cells by using several methods, with Goldner’s trichrome and toluidine blue being most widely used. Solochrome cyanine R allows the observation of bone texture under polarized light. Unstained sections are prepared for the observation of the tetracycline labels by fluorescence microscopy. May–Grünewald–Giemsa or toluidine blue are used for the analysis of bone marrow and especially for the identification of mast cells and TRAP-staining is common to assess osteoclast parameters.

Quantitative analysis is performed on complete and unbroken samples. Measurements are performed by using automatic or semi-automatic image analysers. Parameters can be measured separately on periosteal, cortical, endocortical and cancellous bone. The bone histomorphometric parameters with abbreviations have been standardized by the American Society for Bone and Mineral Research (ASBMR) Histomorphometric Nomenclature Committee (186, 187).

For some specific diagnoses histomorphometric examination is required. For example, osteomalacia shows an accumulation of osteoid i.e. non-mineralised bone. While the experienced examiner can give the diagnosis of osteomalacia without quantification, the degree of the delay of mineralization requires HM. Hyperparathyroidism (HPTH) is associated with high bone turnover and an increased amount of immature bone showing a diverged picture from the usual lamellar structure.
referred to as woven bone, as well as marrow fibrosis. An important indication for bone biopsies is chronic kidney disease (CKD) with potential high or low turnover conditions. In mild CKD, changes may be similar to HPTH with woven bone and peri-trabecular marrow fibrosis, referred to as osteitis fibrosa. Osteomalacia and adynamic bone disease are showing with low turnover features in bone histomorphometry. Both conditions require careful therapy adaptation. Bone fragility disorders such as osteogenesis imperfecta are associated with typical static and dynamic HM. Reference values of healthy children and adolescents (188), adult osteoporosis (183) as well as patients with OI type 1 (189) are used for interpretation of HM results.

Limitations: This invasive method depends on established procedures and trained personnel. Localised bone diseases like Paget’s disease of bone and fibrous dysplasia are usually not seen in iliac biopsies. Analysis is performed on an iliac bone sample, an unloading site not prone to fracture in contrast to vertebra, forearm or femoral neck. Despite differences in microarchitecture and turnover between iliac crest and the other skeletal sites, significant correlations were found (190).

Strengths: Bone HM remains the only method allowing the study of bone at the tissue and cell levels to enable measurements at intermediary levels of organization of bone i.e., the osteon. It also remains the only established method to diagnose osteomalacia.

In the study of mouse bone, both static and dynamic histomorphometry are widely used mostly on sections of the distal femoral metaphysis and, for cancellous bone, in the appendicular skeleton. Several staining methods are used to measure osteoblast parameters, such as Toluidine blue or Von Kossa and McNeal stain (191). Osteoclast parameters additionally are measured using the TRAP staining (192). In dynamic histomorphometry, bone formation and apposition rates are calculated using a timed fluorescent agent which is incorporated into newly formed bone, much like described for human studies. Fluorochromes such as calcine, tetracycline and alizarine red (193), can even be combined for double labelling that has demonstrated both increased and reduced bone formation and mineral apposition rates in cortical and trabecular bone (160, 192).

As in human and mice, static and dynamic histomorphometry are used in zebrafish. Vertebral endplates are active sites of bone formation, providing a suitable region for typical static bone histomorphometry (50). Number of osteoblasts per bone perimeter (N.Ob/B.Pm), osteoid thickness (O.Th), osteoid surface per bone surface (OS/BS) and osteocyte density (N.Ot/B.Ar) can be assessed (40). As zebrafish are transparent during skeletogenesis and through juvenile stages, bone staining are often readily observed in whole-mount, alleviating the time and resources required for tissue sectioning. Alizarin Red and Calcein staining are used to label mineralizing tissues (194), which can be monitored in vivo in bones that are optically accessible, such as early developing vertebrae, growing vertebrae, scales and adult fin rays. Pulse labelling with Alizarin and Calcein can demarcate bone formation between labelling periods (50, 195–197), similar to dynamic histomorphometric approaches in mammals. Mineralised bone can also be assessed by Von-Kossa staining and activity of alkaline phosphatase (ALP) (198). Cartilage is frequently visualized using Alcian Blue (199–201). Moreover, the use of transgenic lines also allows in vivo assessment of specific cell types, including osteoblasts (202, 203), osteocytes (203) and osteoclasts (204, 205). Osteoclast activity can be observed in whole-mount or histological sections using TRAP staining (206).

4.3 Microindentation

Micro- and nano-indentation have been in many studies to quantify the modulus (stiffness) and hardness (resistance to yielding) of bone tissue (207, 208). A limitation of these techniques is that they can be applied only to extracted bone samples or biopsies. Reference Point Indentation (RPI) estimates the resistance of the cortical bone to fracture (209, 210). It is based on the hypothesis that the microindentation of the bone surface induces the separation of mineralized collagen microfibers and the initiation of micro-cracks (211). Whereas this measure is related to the resistance of bone tissue to fracture, it is incompletely understood which mechanical properties of bone are captured by RPI. For this reason, measurements typically quantify a parameter called Bone Material Strength index (BMSi) units representing the ratio between the penetration of the probe into the bone and its penetration in a methyl methacrylate reference phantom (209, 210). In RPI, a probe is applied to the outer surface of the cortical bone of the tibia under local anaesthesia to produce a microindentation (of a size similar to a resorption lacuna), and thus, to measure the distance the probe can penetrate the bone. The higher this distance, the less the bone is able to resist the formation and propagation of micro-cracks, and thus the weaker it is (210).

Two distinct RPI techniques exist (208). The first to be developed was the cyclic reference point microindentation (CMI) using the BioDent™ device (211). CMI was used in the first human clinical studies and is currently the most used technique in animal studies (209). The second technique called impact microindentation (IMI) is conducted with the Osteoprobe R device (212). IMI was developed for in vivo use in clinical studies exclusively from 2013 on (210), and in larger animals (209). As the two techniques differ in mechanical challenges, and do not exactly measure the same mechanical properties, the preclinical results from CMI cannot be extrapolated to clinical results from IMI (209).

Limitations: The use of IMI in the clinical practice is still hampered by methodological and technical limitations and the lack of reference values validated according to ethnicity, sex and geographical regions. The development of standardized procedure (213) and future prospective multicentre studies will clarify the benefit of the methods for the assessment of the pathophysiology and the response to treatment interventions (208–210).

Strengths: This technique holds great promise as it provides clinicians a minimally invasive, simple and safe tool for assessing the material properties of bones in vivo (209). Existing data support IMI as a valuable technique for the assessment of bone
fragility in research studies and possibly for its follow-up (209, 210). Importantly, IMI has been proposed as an additional tool to assess and comprehend bone quality, instead of replacing the existing techniques (208–210).

In mice, micro- and nanoindentation is not a commonly used technique, but has been used to assess bone homeostasis and bone repair following micro-damage (214, 215). Among the techniques, RPI is used most frequently. Benefits are the capacity for longitudinal, in vivo assessment of the mechanical properties of bone. However, during ex vivo technique validation, it has been shown that RPI testing data is poorly correlated with fracture data from traditional biomechanical testing, and has relatively large variability (216, 217).

In zebrafish, nano-indentation is used instead of micro-indentation due to the small size of the zebrafish bones (vertebral length ~ 500 μm, and width ~ 50 μm). It allows the determination of local mechanical properties in sagittal or transverse planes of individual zebrafish bone (40, 218–220). Specifically, the modulus of elasticity, hardness, and modulus-to-hardness ratio (E/H; used as a surrogate measure bone fracture toughness (221) can be extracted and correlated to compositional parameters. In this context, an increase in mineralization under physiological conditions, e.g. with aging, results in an increasing elasticity of zebrafish vertebrae, homologous to human and mammalian bone in general. However, in case of a more disoriented bone matrix, e.g. due to collagen pathologies, an altered organization of the mineral has been correlated with a decrease in mechanical performance. Given the high resolution of nano-indentation experiments, i.e. penetration depth of several 100 nm with a Berkovic tip, heterogeneities in mechanical performance can be assessed, e.g. vertebral end plate region vs. vertebral centrum.

4.4 Compositional Bone Matrix Analysis Using Quantitative Backscattered Electron Microscopy Imaging (qBEI) and Vibrational Spectroscopy

Blocks from bone biopsies can be analysed using quantitative backscattered electron microscopy imaging (qBEI). For qBEI, specimens are commonly embedded, polished coplanar and coated with carbon to provide stable electron conductivity. Assessed is bone mineralization density distribution (BMDD), reflecting the calcium content of cortical and trabecular bone matrix (222–224). With qBEI, the phenotype of several conditions can be further delineated, for example the typically elevated bone tissue density in osteogenesis imperfecta (223). Using a backscattered electron (BSE) detector, variations of intensity of the BSE-signal are measured. Backscattered electrons interact mainly with the sample surface, whereby the intensity is dependent on the local mean atomic number of the sample. Calcium, being the heaviest element in bone, is used to quantify the degree of mineralization based on a linear correlation between the calcium content and the grey value of the BSE image. With the help of reference materials, the brightness and contrast of the image are calibrated and the mean calcium weight percent distribution can be determined based on the grey value histograms of the BSE image, which allow to assess the average calcium content in the mineralized bone tissue area (Ca_mean), the heterogeneity of mineralization (Ca_width), as well as areas of high and low mineralization (Ca_high and Ca_low, respectively).

Whereas qBEI provides compositional information mainly on the inorganic component of bone, vibrational spectroscopy can be used for the simultaneous analysis of mineral- and protein-related parameters in bone. The identification of molecular components based on their energy-specific vibrations is used in both Fourier-Transform Infrared spectroscopy and Raman spectroscopy (225). In the context of bone quality assessment, vibrational spectroscopy is a specialized tool to evaluate the “structural fingerprint” for the identification of molecular bonding involved. During vibrational spectroscopy, the sample is irradiated with a specific wavelength, which leads to changes in the vibrational modes of specific molecules, allowing to detect mineral-related components in a spectrum (peaks of phosphate, carbonate) and protein-related components (peaks of amide I and II, phenylalanine, hydroxyproline and proline). Typical Raman and FTIR parameters of bone quality include the mineral-to-matrix-ratio (e.g. phosphate-to-amide I, indicative of the degree of mineralization), the carbonate-to-phosphate ratio (indicative of carbonate substitution in the crystal lattice), and crystallinity (fill-width-at-half-maximum of the phosphate peak, related to crystal size), e.g. with different reactions of bound-water compartments for collagen and mineral-bound water. Certain aspects of collagen are also assessable (226). For a more in-depth view also on strengths and limitations, please see e.g (227).

Limitations: Electron microscopy is a complex method, depending on specific clinical/research questions and requires established highly specialized procedures and trained personnel. Moreover, both qBEI and vibrational spectroscopy are generally limited to 2D information.

Strengths: True bone density distribution at the tissue level is measured. Additional information on molecular components and mineralisation enables for new approaches in the interpretation of bone metabolism and structure.

For studying mouse bone, backscattered electron scanning electron microscopy is particularly useful. Resin embedded samples have been used to determine local tissue level mineralisation of bone and the high resolution allows for the investigation of lacunar properties, such as their size and appearance of their surface. Osteoblasts and osteocytes bound to the surface of the specimen can be assessed for their various phenotypic stages. Macerated, non-embedded samples can be used to identify changes in microarchitecture and surface values (192, 227).

Similar to performing qBEI and vibrational spectroscopy in humans, zebrafish bones can be investigated in terms of calcium content and heterogeneity of mineralization as well as Raman spectral parameters. For instance, an increased calcium content has been observed after exercise of zebrafish, as well as in zebrafish OI models (40). Raman and Fourier-transform infrared (FTIR) spectroscopy (19) imaging in zebrafish have shown that zebrafish bone contains carbonated hydroxyapatite as well as other mineral phases, similar to mammalian bone. Moreover, in a zebrafish model of OI, lower matrix maturity is
confirmed through reduced collagen maturity and altered carbonate-to-phosphate ratio using FTIR (40).

4.5 Immunohistochemistry of Bone (IHC)

Immunohistochemistry (IHC) allows to determine the cellular localization of proteins and their expression within tissues (228). This technique requires two phases: 1) specimen fixation and tissue processing and 2) interpretation and quantification of the obtained expression (229). Tissue properties can be analysed in depth, making it possible to study not only bone but also the surrounding tissues like cartilage, muscle and tendons. There are different approaches in immunohistochemistry analysis and reporting and for some IHC markers like bone morphogenetic proteins (BMP), osteocalcin (OCN), osteopontin (OPN), and few others scoring systems are available (230), which may include IHC markers of the surrounding tissue. Sequential antibody immunostaining for quantification is used to detect antigens of interest. This is a complementary method to in situ hybridization histochemistry (ISHH) which detects cellular nucleic acids based on the formation of double-stranded hybrids between a nucleic acid fragment (the probe) and a DNA or RNA sequence present within bone cells (231).

Limitations: Decalcified bone samples are widely used for IHC. However, during the decalcification process, the integrity of the trabecular network is lost, which can cause changes in the overall appearance of the bone morphology. Careful specimen preparation and analysis by experienced researchers should be applied. As an alternative, methyl methacrylate embedding retains the mineral fraction of the bone tissue. However, the hard embedding makes sectioning more difficult and epitope retrieval complex (228). The process of optimization of the method for each target can be relatively time-consuming, costly and labour intense.

Strengths: For humans, a great variety of antibodies are available and established for IHC. On its own and in combination with different antibodies and other staining techniques, this enables a multitude of possibilities to visualize complex interactions.

In mouse studies, IHC can provide useful information on the temporo-spatial expression of key factors important for musculoskeletal development and function. However, careful and experienced specimen handling of the small mouse samples is needed due to the tendency for samples to detach from slides. IHC has been used to locate many important antigens in bone such as SOX9, OSX and sclerostin (232, 233). As in humans, decalcified bone specimens are most commonly used.

Immunohistochemistry is applied in histological sections (234) and whole-mount zebrafish samples, often performed in larval stages and dissected adult tissues (171, 235–237). While a plethora of antibodies are available for human and mice proteins, only few are available for zebrafish. Antibody tests and protocol optimization need to be performed in zebrafish as for mice and human samples.

5 BIOCHEMISTRY FOR BONE PHENOTYPING

In patients, a number of general laboratory analyses are necessary to assess a patient’s general health and potential causes of secondary bone disease (238, 239). These include a full blood count, erythrocyte sedimentation rate (ESR) or C-reactive protein (CRP), markers of liver and kidney function and markers of calcium/phosphate metabolism. Further, optional tests include serum proteins (including electrophoresis; to exclude multiple myeloma), markers of thyroid function (to exclude thyrotoxicosis), sex hormones (to exclude hypogonadism) and measurement of free cortisol in 24-hour urine for screening for Cushing’s syndrome. Some additional tests might be useful to exclude other pathologies, e.g. celiac disease via transglutaminase antibodies or systemic mastocytosis via serum tryptase and/or urine methyl histamine.

Limitations: Optional laboratory parameters require a diagnostic plan for the individual patient to be useful and may be more expensive.

In mice, most studies use a defined mouse model for distinct research questions. Therefore, it is not necessary to perform biochemistry for the diagnosis of a disease in this context. However, biochemistry can be performed on blood and plasma samples. Other than in humans, the amount of blood obtained from the living animal can be a limiting factor for such tests. They are therefore mostly performed with terminal blood collection in mice.

In zebrafish, although neither pregnancy nor lactation exist, the sex-hormonal changes and sexual-development milestones (analogous to “puberty” or “post-reproductive” age) are well characterized. Sex in zebrafish (as well as amphibians and reptiles) is not determined by a particular chromosome, but by the interaction between gene and environment. Sex hormones are well studied, hypogonadism could be easily obtained and well as orchid- and ovariectomy. Catecholamines, mineralocorticoids and microelements are measured similarly to mammals, as well as thyroid stimulating hormone and its receptor (13). Blood can be collected from the adult zebrafish through the aorta and decapitation. Recently, it has been shown that repeated blood collection can be performed from the same adult zebrafish longitudinally for the measurements of triglycerides and glucose (240). Due to the small size of zebrafish, a limitation of the method is the total blood sample volume that can be collected at time, ≤0.4% of body weight per week for repeated measurements.

5.1 Controllers of Bone Mass and Mineralisation

Many factors are involved in the regulation of bone mineralization, among them calcium, phosphates, calcitriol, fibroblast growth factor 23 (FGF23) and parathyroid hormone (PTH). A major clinical problem worldwide is vitamin D deficiency, which has attracted considerable interest over the last two decades (241). Active Vitamin D is the main supplier of bone minerals to bone tissue. In patients with sufficient vitamin D levels, additional supplementation has no effect on bone (242). Hence, recent studies in osteoporosis have seriously questioned the routine use of vitamin D supplementation for the purpose of preventing osteoporotic fractures (243). Bone mineralization disorders in general (osteomalacia/rickets) can easily be excluded, diagnosed
and treated - there is global consensus for what constitutes sufficient vitamin D levels and calcium intake for the prevention of osteomalacia and rickets (244).

The calcium sensing receptor senses decreased dietary calcium supply and increases PTH secretion. Hence, PTH levels are inversely correlated with 25-hydroxycholecalciferol [25(OH)D] levels and dietary calcium supply. However, while PTH is generally known as an indicator of vitamin D status, there is no consensus regarding the accuracy of measuring PTH to determine vitamin D depletion (245). Dietary calcium supply is the likely reason for this (245). Through an unknown phosphate sensing mechanism, FGF23 controls renal phosphate reabsorption. Bone hypo-mineralisation (osteomalacia, rickets) only develops when serum phosphate is low (246) and is accompanied by elevated (bone) alkaline phosphatase and PTH concentrations.

Laboratory analysis of calcitropic hormones, mainly 25-hydroxycholecalciferol (25(OH)D) as a surrogate for the individual pool of vitamin D and several metabolites including the active 1,25-dihydroxycholecalciferol (1,25(OH2)D) have gradually undergone worldwide standardization with European and U.S. quality and accuracy methods including reference samples used as gold standards for both mass spectrometry and enzyme-linked assays (247). PTH measurements have undergone considerable development over the years, discriminating between the entire molecule and distinct fragments. Currently used assays identify intact PTH and assays for subforms are still available for specific questions.

Limitations: There is an urgent need to define non-invasive diagnostic criteria for osteomalacia (248). There is some discussion about the analytical approaches and the conversion of units used e.g. in 25(OH)D measurements. There is no single biochemical marker that represents normal bone mineralization.

Strengths: Bone mineralization disorders can be excluded by simple blood tests.

In mice, assays are available for testing serum levels of vitamin D and PTH in mice. Although these tests have been used in the study of dietary intervention and bone health, they are not regularly used in the routine monitoring of bone health in mice (249). On the other hand, a particular benefit of mouse models for studying skeletal disease is that they have less natural variation than humans. Identical diets, environments and genetics mean that mice should have minimal variation in vitamin D or PTH levels.

In zebrafish, measurements have not yet been performed in vivo in longitudinal studies. Upon fish decapitation, blood and serum readings could be potentially performed for levels of PTH, calcium phosphate (dependent on food intake), FGF23 levels and vitamin D. It would be interesting to test if such assays could be performed using small volumes of blood/serum that can be collected from zebrafish allowing longitudinal studies.

5.2 Bone Turnover Markers (BTMs)

Systemic markers of bone turnover (BTMs) reflect bone remodelling in adults (250), but also a combination of bone remodelling, modelling and 3-dimensional bone growth in children (251) (Figure 3).

Bone formation markers such as osteocalcin (OC), and N-terminal and C-terminal propeptides of type I procollagen (PINP, PICP) are proteins secreted by osteoblasts and represent the activity of bone formation. PINP is also expressed in other tissues (e.g., skin) and during fibrotic processes. Therefore, its concentration may be elevated in skin diseases and in case of active fibrosis (e.g., liver, lungs, heart) (252). Bone alkaline phosphatase (bone ALP) is an ectoenzyme present on the outer surface of osteoblasts. Their serum levels are correlated positively with histomorphometric measures of bone formation (e.g., osteoid surface, appositional rate, mineralization) (253).

The markers of bone resorption activity comprise C-terminal and N-terminal telopeptides of type I collagen (CTX, NTX), deoxypyridinoline (DPD) and hydroxyproline (HPro). They are products of bone collagen degradation. Blood and urinary levels of bone resorption markers reflect the activity of bone resorption. They are correlated positively with histomorphometric measures of bone resorption and decrease rapidly after administration of an anti-resorptive agent (254). Among bone resorption markers, CTX is the most specific for bone. Tartrate-resistant acid phosphatase 5b (TRAP5b) is an enzyme expressed by osteoclasts. It is an indicator of the presence (number) of osteoclasts, but not necessarily of their resorptive activity. Therefore, in some situations (osteopetrosis, treatment with cathepsin K inhibitors), discrepancy between the TRAP5b concentration and the levels of collagen degradation products may be observed (255, 256).

Serum PINP and CTX are defined as the reference markers of bone formation and bone resorption (257).

Limitations: Their specific technical and analytical limitations are categorized in two main groups (258):

Analytical variability: Despite the reduced analytical coefficient of variation (CV) of the techniques used to measure these markers, lack of a uniform standardization technique has resulted in difficulties in comparing values obtained by dissimilar methods in different laboratories. The intra-individual biological variability of BTMs is still of concern, especially when therapeutic approaches should be chosen based on a single measurement.

Pre-analytical variability: The BTMs’ preanalytical variability due to uncontrollable and controllable factors should be considered during their clinical interpretation. Uncontrollable factors are for instance age, sex, renal function, growth rate, pubertal or menopausal status, comorbidities or recent fracture. The use of appropriate reference ranges, in particular in children, and suitable adjustments can help overcome this variability to some extent. Controllable factors are for instance food intake/fasting status, circadian and menstrual cycle or exercise. The effects of these factors can be minimized by standardizing the timing and conditions of sample collection.

Measurements of BTMs are not helpful for diagnosis of osteoporosis, bone fragility or skeletal dysplasia. For instance, only 20% of osteoporotic women had serum CTX-I concentration exceeding the upper limit of the reference values in premenopausal women (259). However, elevated BTM levels (especially urinary bone resorption markers and bone ALP) may point to more rapid bone loss and higher risk of hip fracture in adults, mainly in postmenopausal women (260, 261). On an individual level, BTMs may be helpful for monitoring
anti-osteoporotic treatments in adults (262) and children (263). Circadian rhythms and fasting are important factors in the clinical validation - blood samples for PINP can be collected at any time of the day due to only a slight circadian variation comparable to the measurement error of the assay (264). By contrast, blood CTX decreases rapidly after breakfast (265) and blood for CTX assay must be collected in the fasting status in the morning before 10 am.

In children, specific age- and development-associated reference ranges are used for most of the markers but the diagnostic value is limited (266).

**Strengths:** BTMs are non-invasive to bone. They closely monitor systemic bone turnover and are increasingly established and accessible. Standardization of the analytes is ongoing, but assay results may differ between providers.

BTMs are easily measured in mice using ELISA or commercially available assays. In an ovariectomized mouse (as a model of osteoporosis) OC and CTX provided conclusive outcomes on bone turnover when compared to μCT imaging (267, 268). Although serum BTMs hold the benefit of allowing multiple tests to be taken from one animal over a time course, it should be considered when planning an experiment whether more useful data can be gained by qPCR or western blot of bone tissue post mortem.

Bone formation (osteocalcin, alkaline phosphatase) and resorption markers (TRAP) are measured ex vivo through immunoassays in whole-mount, dissected tissues and on histological sections. For in vivo studies Alizarin Red and Calcein can be used to measure bone formation as well as the use of transgenic lines for osteocalcin, and TRAP line labelling osteoclasts (205, 269). In vivo longitudinal studies for such markers through blood and serum collection have not yet been performed in zebrafish.

### 6 NEW PHENOTYPING AND FUTURE ASPECTS

For the purpose of personalized medicine and to avoid a “one size fits all” approach, a differentiated pattern of patients’ characteristics might be a goal for future investigations based on age, sex and ethnicity and other background information, which should be taken into consideration. Duration of exposure to any harmful agent or environmental condition is important during the life course and of great interest from the epigenetic perspective. In the young, damaging environmental exposures have not yet accumulated. One example is obesity, where
duration of exposure in relation to bone health is something comparatively little is known, but which could be explored in animal models.

There are numerous important new angles, e.g. the view on bone health early in life, shown by the recent insights in the paediatric field. Children with severe illness or skeletal disorders often have short stature, scoliosis, joint contractures and bone deformities. Hence, despite size adjustment, DXA data can be falsely low or unobtainable. Therefore, clinical assessment of bone health and research studies focus on defining bone phenotypes from bone biopsy, x-rays and vertebral fractures and less on DXA. This is improving our understanding of bone physiology of rare diseases. For example, skeletal effects associated with transgender pharmacotherapy have not yet been widely studied. Issues include start-time for treatment, particularly with regard to puberty, and the short- and long-term effects from cross-sex therapies on mineral metabolism.

Looking to the future, a more holistic approach to musculoskeletal health is needed given the multifactorial, polygenic nature of osteoporosis, to facilitate healthy aging/frailty prevention.

6.1 Future Developments in 2D and 3D Imaging
To date, bone mass measured by DXA is the most widely used bone phenotypic measure for genetic population studies of osteoporosis, in large part due to the ubiquitous presence of DXA scanners, which are widely used clinically and in large scale population studies. More recent genetic studies have examined other related phenotypes including hip geometry and shape (270). Looking to the future, DXA scans are likely to contribute to genetic studies of other age-related musculoskeletal conditions. For example, a range of other phenotypes, more closely related to osteoarthritis, are currently being generated in hip and knee DXA scans from approximately 100,000 individuals from UK Biobank\(^4\). However, a broad number of new technical approaches are under development.

For mammalian and fish studies, an optical coherence tomography (OCT) and synchrotron radiation microcomputed tomography (SR-\(\mu\)CT) may be developed for future use. OCT provides non-invasive high-resolution three-dimensional (3D) images of biological tissue and quantification of chromophores in tissues (271). Fish and human bones contain hydroxyapatite crystals so it can be compared and quantification of BMD is possible to obtain by comparison to sample with known hydroxyapatite levels. Better resolution (100 nm) can be achieved with synchrotron equipped \(\mu\)CT technologies (SR-\(\mu\)CT) with better assessing of bone micro-architecture (11). Among recently developed MRI techniques is ultrashort echo time (UTE) MRI. Clinical MRI cannot detect water bound to organic matrix, or the free water in the pores of the Haversian system of cortical bone due to the very short apparent transverse relaxation times (\(T2^*\)). Therefore, a new class of sequences, ultrashort-TE (UTE) sequences have been developed recently, with TEs of less than 100 \(\mu\)s. This is much shorter than TEs of conventional sequences. These sequences can be used to detect water signals from within cortical bone (272, 273).

PET scanning can be used for some analyses in bone, and has been used in animal models to assess changes in e.g. bone vascularity or stress fractures (274). However, the reduction of radiation and dose exposures are critical in medical and research imaging, since high doses of radiation are associated with DNA damage. Multiple researchers are actively engaged in the development of clinical total-body PET hardware, promising improvements in dose reductions, reduced scan times, and quantitative kinetic modelling capabilities (275).

Artificial intelligence as new methods for automatic image segmentation, and prediction of fracture risk shows promising clinical value (276). Advances in artificial intelligence (deep learning) also perform well in classifying skeletal radiographs (277). Drug and genetic screening as well as longitudinal studies in zebrafish would benefit from AI, towards implementing novel platforms for gene functional validation through rapid skeletal phenotypic assessment.

6.2 Contribution of -omics Technologies to the Phenotypic Dissection of Musculoskeletal Traits
Historically, genome-wide association studies (GWAS) and next generation sequencing have revolutionized genetic diagnostic services and our understanding of common and rare bone diseases. For example, there are now 20+ genes identified that cause osteogenesis imperfecta through whole genome, exome and RNA sequencing and thousands of genetic variants arising from meta-analyses of the Genetic Factors of Osteoporosis (GEFOS) consortium, and the UK Biobank.

Nevertheless, several steps are needed before GWAS discoveries can be translated to biologic processes underlying the genotype-phenotype relationship. First, variants identified by GWAS need to be linked to the gene(s) in the region. Second, such target genes [identified through GWAS or whole exome sequencing (WES)] need to be placed in the context of pathways affected by the genetic variation. Third, functional, mechanistic studies need to establish how the given alteration of the biologic pathway(s) results in a phenotype.

It is here, where the confluence of a roadmap of gene functional evaluations and a detailed assessment of the laboratory and musculoskeletal phenotype can provide mechanistic insight into the relevant biological processes underlying disease. Multi-omics approaches are used to identify laboratory, phenotypic, disease-specific signatures.

Here we provide a succinct overview of multi-omics layers and their importance for bone phenotyping:

Genomics and Epigenomics: Gene coding regions (underlying most Mendelian disorders) make up less than 3% of the human genome while, approximately 90% of the single-nucleotide polymorphisms (SNPs) that are associated with human disease lie within intergenic or intronic regions. As such, genetic variation (polymorphisms, insertion/deletions and mutations) in intergenic regions, such as enhancers, can strongly affect gene expression, demonstrating a tight regulatory network between the coding and noncoding parts of the genome.

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\(^4\)https://www.ukbiobank.ac.uk/.
Epigenetic changes at DNA level include methylation, where methyl groups are added to the DNA molecule and change DNA activity, e.g. the repression of gene transcription without change of the DNA sequence. Important examples for these effects include genomic imprinting, X-chromosome inactivation, or repression of transposable elements. Effects on bone have been described in ageing (278) and oncology (279), but also increasingly during metabolic challenges, such as chronic kidney disease (280).

Another important epigenetic modification among others is considered histone tail modifications, where covalent post-translational modification (PTM) of nuclear histone proteins occur via methylation, phosphorylation, acetylation, ubiquitination, crotonisation, or sumoylation processes. These changes are currently addressed in a number of bone fields, such as bone and cancer as well as inflammation and rheumatic diseases (281). As an example, for bone/vasculature calcification interactions, a crosstalk between osteogenic transcription factors and histone deacetylases has been described. The inhibition/activation of histone deacetylases might help to develop potential therapeutic interventions in future (282).

For more specific information, see also the publication of GEMSTONE WG3 “Gene & Therapeutic Discoveries in Bone Mass Disorders”.

Transcriptomics: RNA Sequencing, gene expression, eQTLs, Non-coding RNAs (IncRNA and miRNA)

Non-coding RNAs (ncRNAs) are recent candidates to become future diagnostic bone biomarkers (283). ncRNA transcripts vary in length from around 22 nucleotides for microRNAs (miRNAs) to more than 200 nucleotides for long non-coding RNAs (IncRNAs). They are found in tissues, but importantly also in body fluids, where they are easier accessible for detection (283, 284).

Different types of ncRNAs are involved in several processes like DNA replication (285, 286), translation (287), RNA splicing (288) and transcriptional regulation (289). Especially miRNAs and IncRNAs are in focus as biomarkers for many conditions, including osteoporosis with some commercially available assays (290–292). They are stable in the bloodstream and protected from RNase digestion. In the exosomal fraction of body fluids, both miRNAs and IncRNAs are enriched, indicating potentially active secretion of these RNA species from their cells of origin (293, 294).

miRNAs are best studied for their involvement in the control of bone formation and homeostasis through their regulatory functions in osteoblast and osteoclast development (295), not only in metabolic bone diseases, but also in case of cancer and bone metastases. Currently, ncRNA assays are not routinely used and require in most cases a dedicated lab with established PCR procedures. However, ncRNAs may help in the multi-omics phenotype characterization of rare bone diseases and defining disease state in common bone diseases.

Proteomics and metabolomics: The protein contents, i.e. the proteomes, of tissues and cells logically occupy a central position within the biologic processes underlying genotype-phenotype relationships. Proteomes can be studied qualitatively and/or quantitatively at a large scale by proteomics (296). Nowadays, mostly based on the use of liquid chromatography and mass spectroscopy methodologies, proteomic approaches can be employed for protein identification and quantification in samples as diverse as tissues, blood and cells, to provide a comprehensive and quantitative information on their proteomes (296).

Therefore, a closer inspection of the bone proteome by proteomics is surely a fundamental tool to put in place towards the phenotypic dissection of musculoskeletal traits. For instance, proteomic approaches can help detect changes in the signal transduction of bone cells, in the regulatory mechanisms that govern bone cell differentiation, among other cellular and tissue processes enrolled in bone metabolism in both physiologic and pathological contexts (297). Accordingly up to now, several studies have made important contributions to our knowledge of the bone proteome, as well as, of the proteomes of individual bone cells (reviewed in (297–299). A special emphasis has been given to identify proteomic changes in osteoporosis (300).

Modern metabolomic analysis (analytical chemistry and bioinformatics) is capable of detecting hundreds of metabolites in human serum and hence identify novel biomarkers and biochemical signatures of disease. These topics are further described in the publication of GEMSTONE WG4.

Microbiomics: Animal studies using germ free mice, antibiotics, probiotics (i.e., microorganisms which confer a health benefit on the host) or prebiotics (i.e., nutrients capable to modify the gut microbiota) have shown that the complex community of microbes colonizing the gastrointestinal tract may regulate bone mass (301, 302). The impact of major alterations of the gut microbiota has been evaluated using either germ-free mice raised in sterile isolators and completely devoid of microbiota, or rodents depleted of gut microbiota by antibiotic use. These rodent models may be inoculated with specific microbes or communities of microbes to examine the effects they trigger on the skeleton in their host. All these studies using germ free mice or antibiotic treated mice exemplify extreme situations and it might be more physiological to look at the bone effects of treatments resulting in minor but specific changes of an already present gut microbiota. Probiotics and prebiotics given to rodents with an already present gut microbiota have shown that specific changes in the gut microbiota may protect against ovariectomy-induced and inflammation-induced bone loss (303). To characterize the skeletal phenotypes in these rodent bone loss models, standard analyses including DXA of areal BMD, MicroCT of cortical and trabecular bone parameters in the axial and appendicular skeleton, static and dynamic bone histomorphometry, bone strength estimates using three-point bending tests of long bones, in vitro studies of primary cultures of osteoblasts and osteoclasts precursors as well as analyses of circulating BTMs can be used (304).

The first promising findings of two randomized clinical probiotic treatment trials recently revealed that certain probiotic
treatments had some bone sparing effects on DXA measurements of areal BMD at the lumbar spine (305) or by CT measurements of volumetric BMD of the distal tibia in humans (306).

The results from the first human cross-sectional association studies between the gut microbiota composition and bone related parameters have yielded conflicting results, most likely as some of the studies were underpowered and not adjusted for major confounders affecting the gut microbiota composition. Large-scale population-based studies assessing the association between the gut microbiome composition, as assessed by cost efficient 16S rRNA sequencing, and DXA-derived phenotypes adjusting for relevant confounders including lifestyle factors, diet and medications are underway. Furthermore, meta-genome wide association studies using state of the art sequence methodology in combination with other -omics platforms should be performed to characterize functional gut microbiota signatures associated with human bone health in detail. In addition, these analyses should not be restricted to DXA-derived areal BMD but include bone architecture and dimensions, specific cortical and trabecular bone parameters and incident fracture risk.

CONCLUSION AND THE TRIANGULATION OF [DIAGNOSTIC] EVIDENCE: THE PATH TO PERSONALIZED MEDICINE

Understanding complex systems such as the skeleton requires the integration of multiple layers of evidence arising from a combination of analytical methods, in humans and animals, as outlined in this publication. Integration across many disciplines is required to solve outstanding questions and create a “deep phenotype” which accurately captures disease signatures. Key to this is translatability – whether across methodologies or species – and synchronization of efforts. The ‘triangulation of evidence’ (307) can overcome logistical and ethical constraints related to experimental design and speed the rate at which the reality of personalized medicine is attained. Looking to the future, there are several areas where the utility of animal models is obvious. For example, skeletal effects associated with transgender pharmacotherapy have not yet been widely studied. Issues include start–time for treatment, particularly with regard to puberty, and the short- and long-term effects from cross-sex therapies on mineral metabolism. Further, on a broader scale ‘duration of exposure’ to a given risk factor during the life course is of interest for skeletal health from an epigenetic perspective. In the young, damaging exposures have not yet accumulated, while with age, these contribute to accelerated biological aging. In conclusion, the idea of triangulation of evidence emerges as a solid way to weigh the robustness of each layer of evidence, but most importantly to gain insight from the integrated systems perspective. Similarly, the triangulation approach helps to overcome severe logistical and ethical constraints on experimental design potentially arising at each level. Once etiologic validity has been satisfactorily established across the different dimensions, the next challenge is to amalgamate these into a “deep phenotype”. This assembled deep phenotype will pave the road to find drug targets and clinical applications, ultimately charting the course toward personalized medicine approaches, for each of us.

AUTHOR CONTRIBUTIONS

IF and BO-P initiated and organised the manuscript. IF, FM, WH, and BO-P generated the figures for the manuscript. All authors contributed to the article and approved the submitted version.

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REFERENCES

1. Sobacchi C, Menale C, Villa A. The RANKL-RANK Axis: A Bone to Thymus Round Trip. Front Immunol (2019) 10. doi: 10.3389/fimmu.2019.00629
2. Zaiss MM, Jones RM, Schett G, Pacifici R. The Gut-Bone Axis: How Bacterial Metabolites Bridge the Distance. J Clin Invest (2019) 129:3018–28. doi: 10.1172/JCI128521
3. Behera J, Ison J, Tyagi SC, Tyagi N. The Role of Gut Microbiota in Bone Homeostasis. Bone (2020) 135. doi: 10.1016/j.bone.2020.115317
4. Lems WF, Raterman HG. Critical Issues and Current Challenges in Osteoporosis and Fracture Prevention. An Overview of Unmet Needs. Ther Adv Musculoskelet Dis (2017) 9(12):299–316. doi: 10.1177/ 1759720X17732562
5. Kan L. Animal Models of Bone Diseases–A. In: Animal Models for the Study of Human Disease. Elsevier Inc (2013). p. 353–90.
6. Maynard RD, Ackert-Bicknell CL. Mouse Models and Online Resources for Functional Analysis of Osteoporosis Genome-Wide Association Studies. (2013) J. Endocrinol. 217(2):97–108. doi:10.1530/JOE-12-0600

7. Haffner-Luizter M, Kovtun A, Rapp AE, Ignatius A. Mouse Models in Bone Fracture Healing Research. Curr Mol Biol Rep (2016) 2(2):101–111. doi:10.1007/s00464-016-0037-3

8. Ilkka RL. The Relevance of Mouse Models for Investigating Age-Related Bone Loss in Humans. Biol Sci J 1(1) J Gerontol A Biol Sci Med Sci (2013) 68(10):1209–17. doi:10.1093/gerona/glt046

9. Song AJ, Palmeter RD. Detecting and Avoiding Problems When Using the Cre–lox System. Trends Genet (2018) 34:333–40. doi:10.1016/j.tig.2017.12.008

10. Hsu PD, Lander ES, Zhang F. Development and Applications of CRISPR-Cas9 for Genome Engineering. Cell (2014) 157:1262–78. doi:10.1016/j.cell.2014.05.010

11. Bergen DJM, Kagwe F, Hammond CL. Zebras as an Emerging Model for Osteoporosis: A Primary Testing Platform for Screening New Osteo-active Compounds. Front Endocrinol (2019) 10:66. doi:10.3389/fendo.2019.00006

12. Kwon RY, Watson CJ, Karasik D. Using Zebras to Study Skeletal Genomics. Bone (2019) 126:37–50. doi:10.1016/j.bone.2019.02.009

13. Hur M, Gistelincn CA, Huber P, Lee J, Thompson MH, Monstad-Rios AT, et al. MicroCT-Based Phenomics in the Zebrafish Skeleton Reveals Virtues of Deep Phenotyping in a Distributed Organ System. Elife (2017) 6:e26014. doi:10.7554/eLife.26014

14. Pardo-Martin C, Allalou A, Medina J, Eimon PM, Walhly C, Yanik MF. High-Throughput Hyperdimensional Vertebrate Phenotyping. Nat Commun (2013) 4(1):1–9. doi:10.1038/ncomms2475

15. Momma Y, Shimada N, Nakayama H, Zang L, Nishimura N, Tanaka T. Aging-Associated Microstructural Deterioration of Vertebra in Zebrafish. Bone Rep (2019) 11:100215. doi:10.1016/j.bonre.2019.100215

16. Hayes AJ, Reynolds S, Nowell MA, Meakin LB, Habicher J, Ledin J, et al. Spinal Deformity in Aged Zebrafish Is Accompanied by Degenerenerative Changes to Their Vertebras That Resemble Osteoarthritis. Heymann D, Editor. PLoS One (2013) 8(9):e75787. doi:10.1371/journal.pone.0075787

17. Gilad S, Meiri E, Yoge Y, Benjamin S, Lebanaoy D, Yerushalmi N, et al. Serum MicroRNAs Are Promising Novel Biomarkers. Williams S, Editor. PLoS One (2008) 3(9):e3148. doi:10.1371/journal.pone.0003148

18. Tomecka MJ, Ethiraj LP, Sanchez LM, Roehl HH, Carney TJ. Clinical Pathologies of Bone Fracture Modelled in Zebrafish. Dis Model Mech (2019) 12(9):dmm037630. doi:10.1242/dmm.037630

19. Mahamad J, Sharir A, Addadi L, Weiner S. Amorphous Calcium Phosphate is a Major Component of the Forming Fin Bones of Zebras. Blackwell Science, 2001. doi:10.1038/srep42323

20. Giovannone D, Paul S, Schindler S, Arata C, Farmer DT, Patel P, et al. Risk Factors for Hip Fracture in White Women. N Engl J Med (1995) 332(12):767–73. doi:10.1056/NEJM19950323332102

21. Siris ES, Chen YT, Abbott TA, Barrett-Connor E, Miller PD, Wehren LE, et al. Bone Mineral Density Thresholds for Pharmacological Intervention to Prevent Fractures. Arch Intern Med (2004) 164(10):1108–12. doi:10.1001/archinte.164.10.1108

22. Kanis JA, Johnell O, De laet C, Johansson H, Oden A, Delmas P, et al. A Meta-Analysis of Previous Fracture and Subsequent Fracture Risk. Bone (2004) 35(2):375–82. doi:10.1016/j.bone.2004.03.024

23. Mattuck GR, Mahanty SR, Skalski MR, Patel DB, White EA, Gottsegan CJ. Stress Fractures: Pathophysiology, Clinical Presentation, Imaging Features, and Treatment Options. Emergency Radiol (2016) 23:365–75. doi:10.1007/s10140-016-1390-5

24. Saumier J, Chapurlat R. Stress Fracture in Athletes. Joint Bone Spine (2018) 85:307–10. doi:10.1016/j.jbspin.2017.04.013

25. Jager PL, Jonkman S, Koalaas W, Stiekema A, Wolfenbuttel BHR, Slat JHA. Combined Vertebral Fracture Assessment and Bone Mineral Density Measurement: A New Standard in the Diagnosis of Osteoporosis in Academic Populations. Osteoporos Int (2011) 22(4):1059–68. doi:10.1007/s00198-010-1293-3

26. Ganda K, Puch M, Chen JS, Speerin R, Blesel J, Center JR, et al. Models of Care for the Secondary Prevention of Osteoporotic Fractures: A Systematic Review and Meta-Analysis. Osteoporosis Int (2013) 24:393–406. doi:10.1007/s00198-012-2090-y

27. Nilsson KH, Hemming P, Shahawy M, Nethander M, Andersen TL, Ejersted C, et al. RSP03 Is Important for Trabecular Bone and Bone Fracture Risk in Mice and Humans. Nat Commun (2021) 12(1):4923. doi:10.1038/s41467-021-25124-2

28. Kagve E, Roy P, Asselin G, Hu G, Simonet J, Stanley A, et al. Osterix/Sp Limitis Cranial Bone Initiation Sites and is Required for Formation of Sutures. Dev Biol (2016) 413(2):160–72. doi:10.1016/j.ydbio.2016.03.011

29. Gistelincn C, Kwon RY, Malafit S, Symoens S, Harris MP, Henke K, et al. Zebras: Zebrafish Type I Collagen Mutants Faithfully Recapitulate Human Type I Collagenopathies. Proc Natl Acad Sci USA (2018) 115(34):E8037–46. doi:10.1073/pnas.1722001115

30. Fiedler IAK, Schmidt FN, Wolfel EM, Plumeyer C, Milovanovic P, Gioia R, et al. Severely Impaired Bone Material Quality in Chihuahua Zebra Reveals Classical Similar Nam Human Osteogenesis Imperfecta. J Bone Miner Res (2018) 33(8):1489–99. doi:10.1002/jbmr.3445

31. Bergquist R, Weiler M, Schwenk W, Ulseth S, Helbostad JL, Vereijken C, et al. Performance-Based Clinical Tests of Balance and Muscle Strength Used in Young Seniors: A Systematic Literature Review. BMC Geriatr (2019) 19(1):9. doi:10.1186/s12877-018-1011-0

32. Clarke KA, Still J. Gait Analysis in the Mouse. Physiol Behav (1999) 66(5):723–9. doi:10.1016/S0031-9384(98)00343-6

33. Histing T, Kristen A, Roth C, Holstein JH, Garcia P, Mathys S, et al. In Vivo Gait Analysis in a Mouse Femur Fracture Model. J Biomech (2010) 43(16):3240–3. doi:10.1016/j.jbiomech.2010.07.019

34. Takeshita H, Yamamoto K, Nozato S, Inagaki T, Tsuchimochi H, Shirai M, et al. Modified Forelimb Grip Strength Test Detects Aging-Associated Physiological Decline in Skeletal Muscle Function in Male Mice. Sci Rep (2017) 7(1):1–9. doi:10.1038/s41598-017-01233-x

35. Connolly AM, Keeling RM, Mehta S, Pestronk A, Sanes JR. Three Mouse Models of Muscular Dystrophy: The Natural History of Strength and Fatigue in Dystrophin-, Dystrophin/Utrophin-, and Lamnin G2-Deficient Mice. Neuromuscul Disord (2001) 11(8):703–12. doi:10.1016/S0960-8966(01)00232-2
101. Knapp KM. Quantitative Ultrasound and Bone Health. J Soc Interface (2019) 16(160):20190430. doi: 10.1098/rsif.2019.0430

102. Kemp JP, Morris JA, Binkley N, Bilezikian JP, Kendrick DL, et al. Special Report on the 2007 Adult and Pediatric Position Development Conferences of the International Society for Clinical Densitometry. Osteoporos Int (2008) 19(10):1369–78. doi: 10.1007/s00198-008-0689-9

103. Cosman F, de Beur SJ, LeBoff MS, Leissner EM, Tanner B, Randall S, et al. Clinician’s Guide to Prevention and Treatment of Osteoporosis. Osteoporos Int (2014) 25(10):2399–81. doi: 10.1007/s00198-014-2794-2

104. Hans D, Dargent-Molina P, Schott AM, Sebert JL, Cormier C, Kotzki PO, et al. The Bone Phenotyping Approaches

105. Lentle BC, Hammond I, Firth GB, Sutton RAL. Imaging of Osteoporotic Fractures on XR, CT, and MR. Eur J Radiol Rep (2017) 1:1–9. doi: 10.1016/j.ejrr.2017.04-04X

106. O’Ryan FS, Khoury S, Liao W, Han MM, Hui RL, Baer D, et al. Intravenous Bisphosphonate-Related Osteonecrosis of the Jaw: Bone Scintigraphy as an Early Indicator. J Oral Maxillofac Surg (2009) 67(7):1363–72. doi: 10.1016/j.oms.2009.03.005

107. Grünebohm A, Kling L, Christiansen S, Mill L, Maier A, Engelke K, et al. Next-Generation Imaging of the Skeletal System and its Blood Supply. Nat Rev Rheumatol (2019) 15:533–49. doi: 10.1038/s41584-019-0274-y

108. Huang K, Feng Y, Liu D, Liang W, Li Q. Quantification Evaluation of 99mTc-MDP Concentration in the Lumbar Spine With SPECT/CT: Compare With Bone Mineral Density. Ann Nucl Med (2020) 34(2):136–43. doi: 10.1007/s12149-019-01425-x

109. Li YR, Zheng X, Wang R, Wu H, Han S, Deng YZ, et al. SPECT-CT Versus MRI in Localizing Active Lesions in Patients With Osteoporotic Vertebral Compression Fractures. Nucl Med Commun (2018) 39(7):610–7. doi: 10.1097/MNM.0000000000000857

110. Butterfield NC, Logan JG, Waung J, Williams GR, Bassett JHD. Quantitative X-Ray Imaging of Mouse Bone by Faxitron. In: Methods in Molecular Biology. New York, NY: Humana Press (2019). p. 559–69.

111. Khalil MM, Tremoleda JL, Bayomy TB, Gsell W. Molecular SPECT Imaging: An Overview. Int J Mol Imaging (2011) 2011:1–15. doi: 10.1155/2011/7796025

112. Tremoleda JL, Khalil M, Gompels ML, Wylezinska-Arridge M, Vincent T, Gsell W. Imaging Technologies for Preclinical Models of Bone and Joint Disorders. ClinNMMRI Res (2011) 1(1):1–14. doi: 10.1186/2191-219X-1-11

113. Liemann PS, Metzger S, Kivelio AS, Blance A, Papageorgiou P, Astalo A, et al. Longitudinal In Vivo Evaluation of Bone Regeneration by Combined Measurement of Multi-Pinhole SPECT and Micro-CT for Tissue Engineering. Sci Rep (2015) 5:10238. doi: 10.1038/srep10238

114. Adams JE. Quantitative Computed Tomography. Eur J Radiol (2009) 71(3):415–24. doi: 10.1016/j.ejrad.2009.04.074

115. Lang TF. Quantitative Computed Tomography. Radiologic Clinics North America (2010) 48:589–600. doi: 10.1016/j.rcl.2010.03.001

116. Engelke K, von Rietbergen B, Zysset P. FEA to Measure Bone Strength: A Review. Clin Rev Bone Miner Metab (2016) 14:26–37. doi: 10.1016/j.crbmm.2015.09.009

117. Johannesdottir F, Allaire B, Bouxsein ML. Fracture Prediction by Computed Tomography and Finite Element Analysis: Current and Future Perspectives. Eur J Radiol Rep (2018) 16(4):411–22. doi: 10.1016/j.ejrr.2018.04.050-z

118. Lengfelder M, Schmitt J, Alter P, Kaminsky J, Leppke R. Comparison of Geometry-Based and CT Voxels-Based Finite Element Modelling and Experimental Validation. Med Eng Phys (1998) 20(7):515–22. doi: 10.1016/S1350-4533(98)00034-4

119. Zysset PK, D’AlAra E, Varga P, Pahr DH. Finite Element Analysis for Prediction of Bone Strength. Bonekey Rep (2013) 2:386. doi: 10.1038/bonekey.2013.120

120. Viecchiotti M. Predicting Bone Strength From CT Data: Clinical Applications. Morphologie (2019) 103(343):180–6. doi: 10.1016/j.morpho.2019.09.007

121. Keavney TM. Biomechanical Computed Tomography–Noninvasive Bone Strength Analysis Using Clinical Computed Tomography Scans. Ann N Y Acad Sci (2010) 1192(1):57–65. doi: 10.1111/j.1749-6632.2009.05348.x

122. Hoffmann U, Massaro JM, D’Agostino RB Sr, Katheresan S, Fox CS, O’Donnell CJ. Cardiovascular Event Prediction and Risk Reclassification by Coronary, Aortic, and Valvular Calcification in the Framingham Heart Study. J Am Heart Assoc (2016) 5(2):e003144. doi: 10.1161/JAHA.115.003144

123. Ryan TM, Shaw CN. Trabecular Bone Microstructure Scales Allometrically in the Primate Humerus and Femur. Proc R Soc B Biol Sci (2013) 280(1758):20130172. doi: 10.1098/rspb.2013.0172

124. Stagi S, Cavalli L, Cavalli T, De Martino M, Brandi ML. Peripheral Quantitative Computed Tomography (pQCT) for the Assessment of Bone Strength in Most of Bone Affecting Conditions in Developmental Age: A Review. Ital J Pediatr (2016) 42:1–20. doi: 10.1186/s13052-016-0297-9

125. Paternoster L, Lorentzon M, Vandenput L, Karlsson MK, Ljunggren Ö, Brandt J, et al. The Bone Phenotyping Approaches
and potential pleiotropic effects on bone. Gibson G, Editor. *PloS Genet* (2010) 6(11):e1001217. doi: 10.1371/journal.pgen.1001217

132. Paternoster L, Lorentzon M, Lehtimäki T, Eriksson J, Kähönen M, Raitakari O, et al. Genetic determinants of trabecular and cortical volumetric bone mineral densities and bone microstructure. Richards JB, Editor. *PloS Genet* (2013) 9(2):e1003247. doi: 10.1371/journal.pgen.1003247

137. Zheng H-F, Tobias JH, Duncan E, Evans DM, Eriksson J, Paternoster L, et al. WNT16 influences bone mineral density, cortical bone thickness, bone strength, and osteoporotic fracture risk. Gibson G, Editor. *PloS Genet* (2012) 8(7):e1002745. doi: 10.1371/journal.pgen.1002745

139. Gasser JA, Willnecker J. Bone measurements by peripheral quantitative computed tomography (pQCT) in the mouse skeleton compared with histology and microcomputed tomography (μCT). *J Bone Miner Res* (2003) 18(8):1486–96. doi: 10.1359/jbmr.2003.18.8.1486

140. Kroker A, Zhu Y, Manske SL, Barber R, Mohtadi N, Boyd SK. Quantitative nucleotide sequencing of human trabecular bone. *Bone* (2015) 79:213–21. doi: 10.1016/j.bone.2015.06.006

141. Thwaitt DE, Boyd SK, Burghardt AJ, Paccou J, Ghasem-Zadeh A, Chapurlat R, et al. Guidelines for the assessment of bone density and microarchitecture in *In Vivo* using high-resolution peripheral quantitative computed tomography. *Osteoporos Int* (2020) 31(9):1607–27. doi: 10.1007/s00198-020-05438-5

142. Boutroy S, Bouxsein ML, Munoz F, Delmas PD. In *Vivo* assessment of trabecular bone microarchitecture by high-resolution peripheral quantitative computed tomography. *J Clin Endocrinol Metab* (2005) 90(12):6508–15. doi: 10.1210/jc.2005-1258

143. Zebaze R, Ghasem-Zadeh A, Mbala A, Seeman E. A new method of segmentation of compact-appearing, transitional and trabecular cortices at the distal radius and tibia by high-resolution peripheral quantitative computed tomography. *Bone* (2010) 47(3):519–28. doi: 10.1016/j.bone.2010.05.034

144. Burghardt AJ, Buie HR, Laib A, Majumdar S, Boyd SK. Reproducibility of direct quantitative measures of cortical bone microarchitecture of the distal radius and tibia by HR-pQCT. *Bone* (2010) 47(3):519–28. doi: 10.1016/j.bone.2010.05.034

145. Zebarz R, Ghasem-Zadeh A, Mbala A, Seeman E. A new method of segmentation of compact-appearing, transitional and trabecular cortices at the distal radius and tibia by high-resolution peripheral quantitative computed tomography. *Bone* (2010) 47(3):519–28. doi: 10.1016/j.bone.2010.05.034

146. Nishiyama KK, Macdonald HM, Buie HR, Hanley DA, Boyd SK. Postmenopausal women with osteopenia have higher cortical porosity and thinner cortices at the distal radius and tibia than women with normal aBMD: An *In Vivo* HR-pQCT study. *J Bone Miner Res* (2009) 24(4):901-910. doi: 10.1359/jbmr.091020

147. Burghardt AJ, Pialat JB, Kakazia GJ, Boutroy S, Engkelke K, Patsch JM, et al. Multicenter precision of cortical and trabecular bone quality measures assessed by high-resolution peripheral quantitative computed tomography. *J Bone Miner Res* (2013) 28(3):524–36. doi: 10.1002/jbmr.1795

148. Piot A, Chapurlat RD, Claustre B, Sulpiz P. Relationship between sex steroids and deterioration of bone microarchitecture in older men: The prospective STRAMBO study. *J Bone Miner Res* (2019) 34(9):1562–73. doi: 10.1002/jbmr.3746

149. Samelson EJ, Broe KE, Xu H, Yang L, Boyd S, Biver E, et al. Cortical and trabecular bone microarchitecture as an independent predictor of incident fracture risk in older women and men in the Bone Microarchitecture International Consortium (BoMIC). A prospective study. *Lancet Diabetes Endocrinol* (2019) 7(1):34–43. doi: 10.1016/S2223-8587(18)30308-5

150. Tsai JN, Ulhein AV, Burnett-Bowie SM, Neer RM, Derrico NP, Lee H, et al. Effects of two years of teriparatide, denosumab, or both on bone microarchitecture and strength (DATA-HRPQCT study). *J Clin Endocrinol Metab* (2016) 101(5):2023–30. doi: 10.1210/jc.2016-1160

151. Feldkamp LA, Goldstein SA, Parfitt MA, Leson J, Kleecker M. The direct examination of three-dimensional bone architecture in *In Vitro* by computed tomography. *J Bone Miner Res* (1989) 4(1):3–11. doi: 10.1002/jbmr.5650040103

152. Campbell GM, Sophocleous A. Quantitative analysis of bone and soft tissue by micro-computed tomography: Applications to *Ex Vivo* and *In Vivo* studies. *Bonekey Rep* (2014) 3:564. doi: 10.1308/bonekey.2014.559

153. Ozan F, Pekedis M, Koyuncu S, Altay T, Yıldız H, Kayah C. Micro-computed tomography and mechanical evaluation of trabecular bone structure in osteopenic and osteoporotic fractures. *J Orthop Surg* (2017) 25 (1):2309499017692717. doi: 10.1177/2309499017692717

154. Perilli E, Parkinson IH, Reynolds JK. Micro-CT examination of human bone: From biopsies towards the entire organ. *Ann Ist Super Sanita* (2012) 48(1):73–82. doi: 10.4415/ANN_12_01_13

155. Jiang Y, Zhao J, Liao EY, Dai RC, Wu XP, Genant HK. Application of micro-CT: Assessment of 3-D bone microarchitecture in preclinical and clinical studies. *J Bone Miner Res* (2005) 23(2):122–31. doi: 10.1359/jbmr.05026336

156. Burghardt AJ, Link TM, Majumdar S. High-resolution computed tomography for clinical imaging of bone microarchitecture. *Clin Orthopaedics Related Res* (2011) 469(6):2179–93. doi: 10.1097/BIO.0b013e31822e7599

157. De Bouronville S, van Grunen S, Kerckhofs G. Contrast-enhanced microCT for virtual 3D anatomical pathology of biological tissues: A literature review. *Contrast Media Mol Imaging* (2019) 1914:533. doi: 10.1002/cmmi.201914

158. Dall’Ara E, Boudiffa M, Taylor C, Schug D, Fiegle E, Kennerley AJ, et al. Direct quantitative measures of cortical bone microarchitecture of the human upper arm: An *In Vivo* HR-pQCT study. *J Bone Miner Res* (2018) 33(6):763–71. doi: 10.1002/jbmr.2817

159. Hamilton EH, Ghasem-Zadeh A, Gianatti E, Lim-Joon D, Bolton D, Zebaze R, et al. Structural decay of bone microarchitecture in men with prostate cancer treated with androgen deprivation therapy. *J Clin Endocrinol Metab* (2010) 95(12):E547–62. doi: 10.1210/jc.2010-0902

160. van Rietbergen B, Ito K. A survey of micro-Finite element analysis for clinical assessment of bone strength: The first decade. *J Biomech* (2015) 48(8):382–41. doi: 10.1016/j.jbiomech.2014.12.024

161. Whittier DE, Manske SL, Kiel DP, Bouxsein M, Boyd SK. Harmonizing finite element model for non-invasive strength estimation by high-resolution peripheral quantitative computed tomography. *J Biomech* (2018) 86:63–71. doi: 10.1016/j.jbiomech.2018.08.030

162. Crocker A, Zhu Y, Manske SL, Barber R, Mohtadi N, Boyd SK. Quantitative *In Vivo* assessment of bone microarchitecture in the human knee using HR-pQCT. *Bone* (2017) 97:43–8. doi: 10.1016/j.bone.2016.12.015

163. Saka K, Chiba K, Kajiyama S, Okazaki N, Yonekura A, Tomita M, et al. Bone mineral density and microstructure of the elbow in baseball pitchers: An analysis by second-generation HR-pQCT. *J Clin Densitom* (2020) 23(2):322–8. doi: 10.1016/j.jocd.2019.03.001

164. Nishiyama KK, Macdonald HM, Buie HR, Hanley DA, Boyd SK. Postmenopausal women with osteopenia have higher cortical porosity and thinner cortices at the distal radius and tibia than women with normal aBMD: An *In Vivo* HR-pQCT Study. *J Bone Miner Res* (2009) 24(4):901-910. doi: 10.1359/jbmr.091020
Wang XM, Frank Eames B, Blanco-Sánchez B, Peng G, He X, Swartz ME, et al. Zebrafish Sp7:EGFP: A Transgenic for Studying Otic Vesicle Formation, Skeletogenesis, and Bone Regeneration. Genesis (2010) 48 (8):505–11. doi: 10.1002/dvg.20639.

Singh SP, Holdway JE, Poss KD. Regeneration of Amputated Zebrafish Fin Rays From De Novo Osteoblasts. Dev Cell (2012) 22(4):879–86. doi: 10.1016/j.devcel.2012.03.006.

Sharif F, De Bakker MA, Richardson MK. Osteoclast-Like Cells in Early Embryonic Zebrafish. Morpho (2001) 250 (3):197–207. doi: 10.1002/jmor.1065.

Nyman JS, Granke M, Singleton RC, Pharr GM. Tissue-Level Mechanical Properties of Bone Contributing to Fracture Risk. Curr Osteoporosis Rep (2016) 1(14):138–50. doi: 10.1007/s11914-016-0314-3.

Allen MR, McNerny EM, Organ JM, Wallace JM. True Gold or Pyrite: A Bioengineered Reference Point Indentation for Assessing Bone Mechanical Properties In Vivo. J Bone Miner Res (2015) 30:1539–50. doi: 10.1002/jbmr.2603.

Herrera S, Diez-Perez A. Clinical Experience With Microindentation In Vivo in Humans. Bone (2017) 95:175–82. doi: 10.1016/j.bone.2016.11.003.

Schoeb M, Handy NAT, Malgo F, Winter EM, Appelman-Dijkstra NM. Bone Mineralization and Fracture Resistance in Multiple Myeloma. J Bone Miner Res (2017) 32(23):4557–63. doi: 10.1002/jbmr.20201.6.

Zhang Y, Cui FZ, Wang XM, Peng QL, Zhu XD. Mechanical Properties of Skeletal Bone in Gene-Mutated Stöpselfli28d and Wild-Type Zebrafish (Danio Rerio) Measured by Atomic Force Microscopy-Based Nanoindentation. Bone (2002) 30(4):541–6. doi: 10.1016/S7567-3282(02)00676-2.

Chang Z, Chen PY, Chuang YJ, Akhtar R. Zebrafish as a Model to Study Bone Maturation: Nanostructure and Mechanical Characterization of
Fractures in Men (MrOS) Study. Osteoporos Int (2016) 27(1):331–8. doi: 10.1007/s00198-015-3536-y
240. Zang L, Shimizu Y, Nishimura Y, Tanaka T, Nishimura N, Novel A. Reliable Method for Repeated Blood Collection From Aquarium Fish. Zebrafish (2013) 10(3):425–32. doi: 10.1089/zeb.2012.0862
241. Holick MF. The Vitamin D Deficiency Pandemic: Approaches for Diagnosis, Treatment and Prevention. Rev Endocrine Metab Disord (2017) 18:153–65. doi: 10.1007/s11154-017-9424-1
242. Reid IR. Vitamin D Effect on Bone Mineral Density and Fractures. Endocrinol Metab Clinics North America (2017) 46:935–45. doi: 10.1016/j.ecl.2017.07.005
243. Buschhoff-Ferrari HA. Influence of Vitamin D on Fracture Reduction Among Older Adults: A Discussion of Recent Meta-Analysis Findings. Osteology (2019) 28(2):136–9. doi: 10.1055/a-0861-2813
244. Munns CF, Shaw N, Kiely M, Specker BL, Thacher TD, Ozono K, et al. Global Consensus Recommendations on Prevention and Management of Nutritional Rickets. J Clin Endocrinol Metab (2016) 101(2):394–415. doi: 10.1210/jc.2015-2175
245. Cherniack EP, Troen BR. Calcitropic Hormones. In: Holick MF. The Vitamin D De. 2016:1–31. doi: 10.1007/s00198-015-3145-7
246. Tiosano D, Hochberg Z. Hypophosphatemia: The Common Denominator of Many Disorders. Histol Histopathol (2017) 32. doi: 10.1002/his.21359
247. Cavalier E, Souberbielle JC. Vitamin D and Its Metabolites: From Now and Beyond. EJIFCC (2018) 29(2):105–10.
248. Uday S, Högler W. Spot the Silent Sufferers: A Call for Clinical Diagnostic Standards in Osteoporosis. J Bone Miner Res (2019) 34(8):1436–45. doi: 10.1002/jbmr.3715
249. Szulc P, Delmas PD, Deniaux B, Malaval L, Chapuy MC, Edouard C, Meunier PJ. Delmas PD, Demiaux B, Malaval L, Chapuy MC, Edouard C, Meunier PJ. Biochemical Markers of Bone Turnover in Men. Calcified Tissue Int (2003) 73(4):207–15. doi: 10.1007/s00223-001-0320-1
250. Szulc P, Naylor KE, Jacques RM, Paggiossi M, Gossiel F, Peel NFA, McCloskey EV, et al. Response of Bone Turnover Markers to Three Oral Bisphosphonate Therapies in Postmenopausal Osteoporosis: The TRIO Study. Osteoporos Int (2016) 27(1):21–31. doi: 10.1007/s00198-015-3145-7
251. Shaw N, Högler W. Biochemical Markers of Bone Turnover in Men. Calcified Tissue Int (2003) 73(4):207–15. doi: 10.1007/s00223-001-0320-1
252. Reid IR. Vitamin D Effect on Bone Mineral Density and Fractures. Endocrinol Metab Clinics North America (2017) 46:935–45. doi: 10.1016/j.ecl.2017.07.005
253. Delmas PD, Demiaux B, Malaval L, Chapuy MC, Edouard C, Meunier PJ. Delmas PD, Demiaux B, Malaval L, Chapuy MC, Edouard C, Meunier PJ. Biochemical Markers of Bone Turnover in Men. Calcified Tissue Int (2003) 73(4):207–15. doi: 10.1007/s00223-001-0320-1
254. du Toit JG, Broux O, Clabaut A. New Insights Into the Epigenetics of Osteoporosis. Genomics (2019) 111:793–8. doi: 10.1007/j.14783.2019.00501
279. Astleford K, Campbell E, Norton A, Mansky KC. Epigenetic Regulators Involved in Osteoclast Differentiation. *Int J Mol Sci* (2020) 21:1–15. doi: 10.3390/ijms21093246

280. Martinez-Moreno JM, Fontecha-Barrusso M, Martin-Sanchez D, Guerrero-Mauveic I, Goma-Garcés E, Fernandez-Fernandez B, et al. Epigenomic Modifiers as Potential Therapeutic Targets in Diabetic Kidney Disease. *Int J Mol Sci* (2020) 21:1–26. doi: 10.3390/ijms21141143

281. Kwon DH, Ryu J, Kim YK, Kook H. Roles of Histone Acetylation Modifiers and Other Epigenetic Regulators in Vascular Calcification. *Int J Mol Sci* (2020) 21(9):3246. doi: 10.3390/ijms21093246

282. Lin W, Li Y, Chen F, Yin S, Liu Z, Cao W. Klotho Preservation via Histone Deacetylase Inhibition Attenuates Chronic Kidney Disease-Associated Bone Injury in Mice. *Sci Rep* (2017) 7(1):46195. doi: 10.1038/srep46195

283. Silva AM, Moura SR, Teixeira JH, Barbosa MA, Santos SG, Almeida MI. Long Noncoding RNA: A Missing Link in Osteoporosis. *Bone Res* (2019) 7:10. doi: 10.1038/s41414-019-0048-9

284. Swinivasan S, Duval MX, Kaimal V, Cuff C, Clarke SH. Assessment of Pircher A, Gebetsberger J, Polacek N. Ribosome-Associated ncRNAs: An Emerging Class of Translation Regulators. *RNA Biol* (2014) 11:1335–9. doi: 10.1080/15476286.2014.966549

285. Will CL, Lührmann R. Spliceosome Structure and Function. *Cold Spring Harb Perspect Biol* (2011) 3(7):1–2. doi: 10.1101/cshperspect.a003707

286. Dykes IM, Emanuelli C. Transcriptional and Post-Transcriptional Gene Regulation by Long Non-Coding RNA: *Genomics Proteomics Bioinf* (2017) 15:177–86. doi: 10.1016/gpb.2016.12.005

287. Foessl I, Kobelt P, Obermayer-Pietsch B. miRNAs as Novel Biomarkers for Bone Related Diseases. *J Lab Precis Med* (2019) 4:2–2. doi: 10.21037/jrpm.2018.12.06

288. Yavropoulou M, Anastasilakis A, Makras P, Grammatiki M, Kotas K, Yovos J. Circulating microRNAs in Postmenopausal Women With Osteoporosis and Vertebral Fractures. *Bone Abstr* (2016) 5:245. doi: 10.1530/boneab.5.P245

289. Hassan MQ, Tye CE, Stein GS, Lian JB. Non-Coding RNAs: Epigenetic Regulators of Bone Development and Homeostasis. *Bone* (2015) 81:746–56. doi: 10.1016/j.bone.2015.05.026

290. Haidl M, Heilmieier U, Wehner S, Grillari J. Circulating microRNAs as Novel Biomarkers for Bone Diseases - Complex Signatures for Multifactorial Diseases? *Mol Cell Endocrinol* (2016) 432:83–95. doi: 10.1016/mce.2015.10.015

291. Lee YR, Kim G, Tak WY, Jang SY, Kweon YO, Park KG, et al. Circulating Exosomal Noncoding RNAs as Prognostic Biomarkers in Human Hepatocellular Carcinoma. *Int J Cancer* (2019) 144(6):144–52. doi: 10.1002/ijc.31931

292. Lian JB, Stein GS, van Wijnen AJ, Stein JL, Hassan MQ, Gaur T, et al. MicroRNA Control of Bone Formation and Homeostasis. *Nat Rev Endocrinol* (2012) 8(4):212–27. doi: 10.1038/nrendo.2011.234

293. Masuda T, Mori A, Ito S, Ohtsuki S. Quantitative and Targeted Proteomics-Based Identification and Validation of Drug Efficacy Biomarkers. *Drug Metab (2021) 36:100361. doi: 10.1016/j.dmpk.2020.09.006

294. Lee JH, Cho JY. Proteomics Approaches for the Studies of Bone Metabolism. *BMB Rep* (2014) 47(3):141–8. doi: 10.5483/BMBRep.2014.47.3.270

295. Sijbren KN, Jacobs JM, Orwell ES. Proteomic Studies of Bone and Skeletal Health Outcomes. *Bone* (2019) 126:18–26. doi: 10.1016/j.bone.2019.03.032

296. Kocic E, Donos N. Proteinomic and Transcriptomic Approaches for Studying Bone Regeneration in Health and Systemically Compromised Conditions. *Proteomics – Clin Appl* (2020) 14(3):1900084. doi: 10.1002/prca.201900084

297. Yang TL, Shen H, Liu A, Dong SS, Zhang L, Deng FY, et al. A Road Map for Understanding Molecular and Genetic Determinants of Osteoporosis. *Nat Rev Endocrinol* (2020) 16:91–103. doi: 10.1038/s41574-019-0282-7

298. Ohlsson C, Sjögren K. Effects of the Gut Microbiota on Bone Mass. *Trends Endocrinol Metab* (2015) 26:69–74. doi: 10.1016/j.tem.2014.11.004

299. Medina-Gomez C. Bone and the Gut Microbiome: A New Dimension. *J Lab Precis Med* (2018) 3:96–6. doi: 10.21037/jrpm.2018.11.03

300. Ohlsson C, Sjögren K. Osteomicrobiology: A New Cross-Disciplinary Research Field. *Calcified Tissue Int* (2018) 102:426–32. doi: 10.1007/s00227-017-0336-6

301. Guss JD, Horsfield MW, Fontenele FF, Sandoval TN, Luna M, Apoorva F, et al. Alterations to the Gut Microbiome Impair Bone Strength and Tissue Material Properties. *J Bone Miner Res* (2017) 32(6):1343–53. doi: 10.1002/jbmr.3114

302. Jansson PA, Curieq D, Lazou Ahrén I, Hansson F, Martinsson Niskanen T, Sjögren K, et al. Probiotic Treatment Using a Mix of Three Lactobacillus Strains for Lumbar Spine Bone Loss in Postmenopausal Women: A Randomised, Double-Blind, Placebo-Controlled, Multicentre Trial. *Lancet Rheumatol* (2019) 1(3):e154–62. doi: 10.1016/S2665-9913(19)30068-2

303. Nilsson AG, Sundh D, Bäckhed F, Lorenzont M. *Lactobacillus Reuteri* Reduces Bone Loss in Older Women With Low Bone Mineral Density: A Randomized, Placebo-Controlled, Double-Blind, Clinical Trial. *J Intern Med* (2018) 284(3):307–17. doi: 10.1111/jim.12805

304. Heale R, Forbes D. Understanding Triangulation in Research. *Evidence-Based Nursing* (2013) 16:98. doi: 10.1136/eb-2013-101494