A review of plant leaf fungal diseases and its environment speciation

Archana Jain, Surendra Sarsaiya, Qin Wu, Yuanfu Lu, and Jingshan Shi

Key Laboratory of Basic Pharmacology and Joint International Research Laboratory of Ethnomedicine of Ministry of Education, Zunyi Medical University, Zunyi, Guizhou, China; Bioresource Institute for Healthy Utilization, Zunyi Medical University, Zunyi, Guizhou, China

ABSTRACT
There is increasing difficulty in identifying new plant leaf diseases as a result of environmental change. There is a need to identify the factors influencing the emergence and the increasing incidences of these diseases. Here, we present emerging fungal plant leaf diseases and describe their environmental speciation. We considered the factors controlling for local adaptation associated with environmental speciation. We determined that the advent of emergent fungal leaf diseases is closely connected to environmental speciation. Fungal pathogens targeting the leaves may adversely affect the entire plant body. To mitigate the injury caused by these pathogens, it is necessary to be able to detect and identify them early in the infection process. In this way, their distribution, virulence, incidence, and severity could be attenuated.

ARTICLE HISTORY
Received 26 June 2019
Revised 20 July 2019
Accepted 24 July 2019

KEYWORDS
Emergent diseases; environmental speciation; fungal pathogens; host plants; plants leaf disease

Introduction

Over 19,000 fungi are known to cause diseases in crop plants worldwide. They may remain dormant but alive on both living and dead plant tissues until conditions are conducive to their proliferation. Certain fungi may develop inside host plant tissues. Fungal spores are readily dispersed by wind, water, soil, insects, and other invertebrates. In this way, they may infest an entire crop [1]. In contrast, other fungi are, in fact, beneficial to the host plant and may support its growth and development. For example, mycorrhizae form a mutualistic relationship with host plant root systems.

On the other hand, pathogenic fungi cause plant diseases such as anthracnose, leaf spot, rust, wilt, blight, coils, scab, gall, canker, damping-off, root rot, mildew, and dieback. Systemic foliar pathogens are major causes for yield and commercial crop losses and diminished crop quality [2]. The rapid identification of fungal disease by timely recognition of their symptoms is an effective management practice and may help control and prevent their spread and progress. Leaves with fungal lesions are photographed and compared with descriptions in databases uploaded on the internet. Fungal leaf diseases may be controlled if detected and...
correctly identified in a timely manner. Part of the crop disease management process includes assessing the negative impacts of the pathogens on crop yield [3].

Human populations rely heavily upon consistent and stable agricultural production. Foliar fungal diseases may pose serious threats to food security. Various disease recognition techniques are required for the rapid detection of leaf diseases in the early phases of plant growth and fungal infection [4]. The current approach for leaf disease recognition is based on the identification and detection of fungal pathogens by comparison with databases from national or regional biological information systems. However, this approach is not fully reliable. Over the past two decades, leaf diseases have been investigated using visible array images. Nevertheless, the methodology to date is imperfect and tends to rely upon idealized disease models and scenarios [5]. Although the focus here is the accurate identification of foliar fungal diseases, environmental speciation may render this task difficult and challenging. Details of research methodology including the networks and software packages used were omitted as they are beyond the scope of this article. This review article surveys emerging fungal leaf diseases, addresses the effects of environmental speciation, and discusses the complications associated with foliar fungal disease recognition.
Significance and characteristics of emerging foliar fungal diseases

In environmental speciation, barriers impede the flux of genetic material between populations, but environmental factors induce mutations that result in new species. In optional speciation, the growths of pathogens are dependent on the environment basis at the earlier stage. The human actions are

Figure 2. Disease control of plant leaf spot.

Figure 3. An overview of plant leaf diseases, its microbial and environmental interaction.
Table 1. Common leaf diseases caused via pathogenic fungi with its symptoms.

| Year   | Region                  | Plant          | Causal agent                          | Disease          | Symptoms                                                                 | References |
|--------|-------------------------|----------------|---------------------------------------|------------------|--------------------------------------------------------------------------|------------|
| 2004   | Rajasthan and Sorghum, India | Sorghum        | Perenosclerospora sorghi, Perenosclerospora heteropogoni | Downy mildew    | Pale yellow to whitish discolorations on the leaf blade. Tassels may be deformed, and ears may be aborted. | [51]       |
| 2005   | Western Turkey           | Pear           | Erwinia amylovora                     | Fire blight      | First leaves turn yellow-brown and after that all are dying.             | [52]       |
| 2007   | New York                 | Carrot cultivars | Alternaria dauci and Cercospora carotae | Leaf blight     | Brownish spotting on leaf.                                               | [15]       |
| 2008   | Belle Glade, USA         | Celery         | Cercospora apiïres.                   | Early leaf blight | Light-brown spots somewhat circular or slightly bony. Spots might be oily look with or without nearby yellow coronas. | [53]       |
| 2009   | Japan                    | Corunus Florida | Anamorphic fungi                      | Leaf blight      | Dead and brown blotches on leaves.                                       | [54]       |
| 2010   | Egypt                    | Potato         | Phytophthora infestans                | Late leaf blight | Pale green spots.                                                         | [55]       |
| 2011   | France                   | Pear           | Erwinia amylovora                     | Fire blight      | First leaves turn yellow-brown and after that all are dying.             | [56]       |
| 2011   | Switzerland              | Pea and apple  | Erwinia amylovora                     | Fire blight      | Red-brown to black streaking may be apparent in wood just under the bark. | [57]       |
| 2011   | Parana Southern Brazil    | Soybean        | Peronospora meliorica                 | Downy mildew    | Upper (light green spots) and lower (greyish/beige downy tufts) on leaf surface. | [10]       |
| 2012   | Germany                  | Apple          | Erwinia amylovora                     | Fire blight      | Red brown or black, may be outward in wood.                               | [21]       |
| 2012   | Spain                    | Grapevine      | Plasmodoa viticola                   | Downy mildew    | Lesions on leaves are angular, yellowish, sometimes oily, and located between the veins. | [58]       |
| 2013   | Temanggung, Malaysia     | Sengon         | Uromycladium tepperianum             | Gall rust       | It starts with local swelling (tumefaction).                             | [59]       |
| 2013   | North eastern India      | Capsicum assimamicum | Cercosporafresen         | Leaf spot       | Round or oblong spots occur; first water soaked and later turn tan with a dark marginal ring. | [60]       |
| 2013   | France                   | Apple          | Erwinia amylovora                     | Fire blight      | The scorched appearance of plant leaves.                                  | [61]       |
| 2014   | Australasia and Southeast Asia | Grapevine    | P. montana                            | Leaf rust       | Angular brown spots on the top side of the leaf.                        | [62]       |
| 2015   | USA                      | Tomato         | Phytophthora infestans                | Late leaf blight | Pale green spots.                                                         | [63]       |
| 2015   | Sanandaj, Iran           | Chickpea       | Fusarium oxysporum                   | Fusarium wilt    | Grayish-green chlorosis, typically affecting lower leaves first and extending up the plant. | [64]       |
| 2015   | Kamataka, Tamil Nadu, Andhra Pradesh, Kerala and Maharashtra, India | Tomato | Alternaria solani                    | Early leaf blight | On leaves, circular lesions are produced. Inside these lesions turn dark, concentric rings could be seen. | [25]       |
| 2015   | India                    | Pearl millet   | Sclerospora graminicola              | Downy mildew    | Pale color, broad streaks spreading from bottom to top of leaves. Advancement of disease, the leaf streaks turn brown and leaves become shredded longitudinally. | [65]       |
| 2016   | Egypt                    | Wheat          | Puccinia triticina Eriks              | Leaf rust       | Rusty colored pustules erupting through on surface.                      | [66]       |
| 2016   | Yunnan China             | Pea            | Erysiphe pisi D.C.                   | Powdery mildew  | Brownish spots on leaves.                                                | [67]       |
| 2016   | Shandong, China          | Apple          | Glomerella cingulata                 | Leaf spot       | Pale yellow, olive green and dark velvety spots.                         | [68]       |

(Continued)
| Year     | Region                  | Plant     | Causal agent                | Disease                        | Symptoms                                                                 | References |
|----------|-------------------------|-----------|-----------------------------|-------------------------------|--------------------------------------------------------------------------|------------|
| 2016     | Sweden                  | Potato    | *Oomycete Phytophthora infestans* | Late blight                  | Pale green spots.                                                        | [24]       |
| 2016     | Hungary                 | Potato    | *Phytophthora infestans*      | Late blight                  | Pale green spots.                                                        | [69]       |
| 2016     | Delhi, India            | Maize     | *Bipolaris maydis (Nisik)*    | Leaf blight                  | In maize include tan, elongated lesions between veins with light brown to brown borders. | [70]       |
| 2016     | Syngenta, Basel, Switzerland | Sweet Potato | *Altemaria solani*         | Early blight                 | Leaves start to exhibit small irregular dark brown spots on the lowest part. | [71]       |
| 2017     | Georgia, USA            | Soybean   | *Cercospora sojina*          | Leaf spot                    | Dark brown centers with red or dark reddish margins.                     | [72]       |
| 2017     | Mumbai, India           | Tomato    | *Fusarium oxysporum*         | Leaf blight                  | Yellowing and wilting of lower leafs.                                    | [73]       |
| 2017     | Islamabad, Pakistan     | Southern corn | *Cochliobolus heterostrophus* | Leaf blight                  | Southern corn include tan, elongated lesions between veins with light brown to brown borders. | [74]       |
| 2018     | Tamilnadu               | Groundnut | *Puccinia Arachidis Speg*    | Leaf rust                    | Orange pustules on lower leaflet and rupture as masses of reddish brown urediniospores. | [13]       |
| 2018     | Greece                  | Pea       | *Erysiphe pisi DC*           | Powdery mildew              | Brownish spots on leafs.                                                 | [75]       |
| 2018     | Peninsular Malaysia     | Mango     | *Fusarium*                   | Leaf spot                    | Gray to grayish brown spots with dark irregular margins.                 | [76]       |
| 2018     | Chandigarh, India       | Eggplant  | *Fusarium oxysporum*         | Fusarium wilt                | One side of leaf will wilt and stop expanding while the other side continues to develop. | [22]       |
| 2018     | Northeastern Thailand   | Rice      | *Xanthomonas oryzae*         | Leaf blight                  | Pale-green to gray-green than turn yellowish.                            | [29]       |
| 2018     | Hawaii, Honolulu        | Basil     | *Peronospora belbahri*       | Downy mildew                 | Yellowish appearance, similar to a nutritional problem.                  | [27]       |
| 2019     | Japan                   | Rose      | *Phragmidium satoanum*       | Leaf rust                    | Yellow black brown spots on both sides.                                  | [23]       |
| 2019     | Beijing china           | Cucumber  | *Oidium neolycopersici*      | Powdery mildew              | Appears dusty white yellow and gray coating on leaf surface.             | [26]       |
| 2019     | Canada                  | Strawberry| *Sphaerotheca macularis*     | Powdery mildew              | Grayish-white appearance on underside of leafs.                          | [76]       |
| 2019     | Chandigarh              | Grapevine | *Erysiphe necator*           | Powdery mildew              | Gray white powdery development on tissue.                                | [77]       |
| 2019     | Guangzhou, China, Netherland | Peanut  | *Puccinia Arachidis Speg*    | Leaf spot                    | Circular dark brown spots surrounded by a yellow halo.                   | [78]       |
| 2019     | Nederland               | Potato    | *Phytophthora infestans*     | Leaf blight                  | Pale green spots.                                                        | [63]       |
| 2019     | Shandong chins          | Cucumber  | *Trichoderma*                | Fusarium wilt               | Yellowing and wilting of lower leafs.                                    | [20]       |
| 2019     | USA                     | Cotton    | *Fusarium oxysporum*         | Fusarium wilt               | Seem in the spring.                                                      | [79]       |
| 2019     | Australia               | Eucalyptus| *Teratocephria destructans*  | Early blight                 | On leaves yellow brown patches and after some time they turn into dark color. | [80]       |
| 2019     | Flakkebjerg, Denmark    | Potato    | *Altemaria solani*           | Early blight                 | When leaves start turning to the small uneven dark brownish spots on top of the lower section. | [53]       |
| Disease          | Host plant                      | Treatment                                                                 | Prevention                                                                 |
|------------------|---------------------------------|---------------------------------------------------------------------------|---------------------------------------------------------------------------|
| Powdery Mildew   | Pea, cucumber, strawberry, and  | Treated with sulfur; apply chemicals on vegetables once the disease       | Pathogen endures in dry and cool conditions; avoid such conditions to      |
|                  | grapevine                       | signs appear; Eradicate the diseased crops to mitigate the pathogen       | mitigate the pathogen.                                                    |
| Fusarium wilt    | Cucumber, cotton, eggplant,     | Best approach to control disease by using disease-free seedlings or       | Harmful microbes are prevented by maintaining the garden temperature      |
|                  | chickpea, tomato                | plant-certified seed; Kill the pathogens by applying fungicides, for      | lower; disinfect and clean farming tools before using them.               |
| Fire blight      | Pear & apple                    | avoid heavy or low application of fertilizer, both encourage the growth,  | Never apply high nitrogen fertilizer                                      |
|                  |                                 | avoid planting nearby to wild crops. Once fire blight is appeared, prune   |                                                                           |
|                  |                                 | off diseased branches one-foot underneath the diseased segments and      |                                                                           |
|                  |                                 | prevent by burning them to reduce further infection.                     |                                                                           |
| Leaf spot        | Peanut, mango apple, soybean    | Not use overhead water; Decrease humidity among plants by providing      | Remove and dispose of spotted leaves on plants that have fallen.          |
|                  | and capsicum                    | adequate space among plants and by pruning lesser branches; Use mulch     | Apply sulfur sprays with copper base fungicides in a week after first     |
|                  |                                 | below trees.                                                             | disease sign to stop its spread from sprouts. Safely manage utmost      |
| Leaf rust        | Sengon, grapevine, wheat, ground & rose | The best way to control white rust is to practice crop rotation. Concerning treatment, try appropriate fungicide to destroy the microbes. | Prevent Albugo through disposing crop separate before planting the crop. |
| Downy mildew     | Basil, soybean, pearl millet,   | Control downy mildew through uprooting the diseased crop along with      | Prevent the pathogen by avoiding wet and cold conditions; prevent        |
|                  | grapevine & maize               | burn; treat it through forestry-al fungicide.                            | excess overhead irrigation and watering.                                  |
| Leaf blight      | Maize, rice, Cornus florida,    | In summer, eliminate limbs, cutting well external infected area and        | Avoid excess of feed along with severe pruning.                          |
|                  | corn, carrot                    | dispose.                                                                 |                                                                           |
| Early blight     | Eucalyptus, potato, cerealy and  | Spread copper base fungicides early sign before 2 weeks of disease;      | Stake or prune plants to expand air flow and decrease fungal              |
|                  | tomato                         | cover equally the tops and undersides leafs.                             | complications; Disinfect pruning shears after each cut; Keep the clean   |
| Late blight      | Potato and Tomato               | Remove diseased parts prior to planting and space plants far sufficient   | soil under plants; Trickle irrigation and downpour hoses are useful to     |
|                  |                                | apart to permit for sufficiently of air passage; Watering in early       | support keep the foliage. Burn infected plant portions.                   |
|                  |                                | morning time, to give plants period to dry ready during the day evade     | Keep eyes on the plants for understand the disease complications at early  |
|                  |                                | overhead irrigation.                                                     | stage; Protection by spraying plants with fungicides which help to        |

[62,63,67,81–83]
contributed toward the variations of bio network. There is growing evidence that these fluctuations play major roles in determining the establishment of microbial pathogens in plants [6]. In this regard, many researchers have concentrated on the increasing incidence of fungal leaf diseases on the host owing to its significance with consequences of the profusion of recognized cases. The significances of different fungal illnesses on the host are important for agricultural yield under the natural environment. However, this theoretical structure does not account for adaptation or differences among pathogens despite evidence that the contamination of a new plant could increase the incidence of developing diseases [7]. Information about fungal leaf diseases is required for the study of well-known views of environment speciation. It can enhance the host plant’s physiology and the growth due to its environmental speciation interaction with pathogens. There are several indications that plant pathogens may undergo rapid environmental speciation via changes in the host itself. Elucidation of the connections among emergent diseases and environmental speciation will help in the development of more realistic models and simulations that integrate the characteristics of the fungal pathogen and new experimental methods to study them. These incidences are responsible for the growth of fungal leaf illnesses [8].

**Characteristics of environmental speciation**

Pathogen virulence may be determined by a single diallelic loci in the pathogen and the host. Host resistance may be identified when the host transports a resistant allele and the pathogen carries an avirulent allele. The allelic mixtures facilitate contagion as the host does not transport the unaffected allele, or while the pathogen carries the ‘deadly’ allele that escapes host recognition. Pathogen alleles are called as virulent strain. It can denoted as virulence for the qualitative capacity of the host genotype(Figure 1) [9].

Plants could change speciation through concentration on a fresh mass. Obligate biotope is a pathogenic living organism that gains resources from the host. Though, lots of mysterious varieties have been discovered inside morphological varieties through utilizing the hereditary concordance of phylogenetic strain for reorganization measures. These strain criterions are used for the phylogenetic concordance of many nonlinked genes. The phylogenetic strain reorganization decisive factor has proven considerably useful for fungi as it is frequently more discerning than other criteria and is also more convenient to use. We regard the whole fungi capable of reproduction. Moreover, the cross-strain communication of pathogen can be stored in the organization of new tissue [10].

**Characteristics of fungal pathogens of plant leaves promoting environmental speciation**

Some history characteristics of fungal pathogens are favorable to environment speciation as they reduce the constraint for harmful speciation. We detail these features below and speculate their significances for the probability of environment changes. The number of spores is a particularly important determiner determination of alteration input for individuals. Pathogenic fungi can generate thousands of microspores in lesions and lead to considerably increased propagation in similar hosts that could give way to split the diseases [11]. Furthermore, high spore numbers permit the rapid generation of gene differences via mutation. Original variety of resistance gene can confer the full confrontation inside for only some months [12].

Such quick development could arise uniformly for growing pathogens and could be identical for lower inherited multiplicity. If the progeny is lower, the selection of plant could be vital intended for the progeny through the superlative probable inherited with ecological circumstantial. This ‘magical trait’ state is a single trait and is mainly favorable for environmental speciation. In theoretical models, the restriction on gene flow is the reduced viability of emigrants. With a very robust choice, it can totally avoid the impartial genetic transformation. Several studies have provided an experimental evidence for the generalization of a mechanism in fungal host pathogens [13,14].
Emerging plant leaf fungal diseases

Pathogenic fungi are responsible for ≤30% of all crop diseases. Epidemics caused by a fungal pathogen and the changes they cause to ecosystems must be reported. Well-documented outbreaks of Cryphonectria parasitica on chestnut trees have resulted in nearly 100% defoliation. Another more recent blight was caused by Phytophthora cinnamomica which is responsible for root rot or dieback in numerous crops. Devastating fungal diseases may affect multiple host plant species and indirectly involve beetles, birds, and other animals. Phytophthora infestans was responsible for potato blight in Ireland. Fungal blast has infested wheat in Brazil since the 1980s and has advanced toward other South American nations. The fungal pathogen Puccinia graminis has case wheat stem rust [15, 16]. Following are descriptions of various important fungal plant pathogens (Table 1).

Fungal leaf spot

Plants cause an effect on the main production with economic losses of farming crop. It can be improved by identifying the leaf sicknesses inside the early-on leaf with disease identity. It can be capable of making easy the recognition of the diseases by a suitable strategy. Here, the current processes for an earlier finding of leaf diseases inside the host are based on the numerous significant structures examined from its leaf descriptions. Leaf spots become visible about 1 week after they first appear. They are white to grayish-white on the edges and enclosed by reddish-brown, brownish, or yellowish margins [17]. They are first observed on the adaxial leaf surfaces and then become apparent on the abaxial leaf surface as well. Ultimately, they perforate the lamina. These holes may vary in diameter and outline shape or may be identical to the initial spots. In general, it is the newer (younger) leaves that first present with fungal lesions. There may be a few small spots that enlarge over time. Others may coalesce or merge into blotches [18].

The main control measure for fungal leaf spot diseases is to keep the foliage as dry as possible. Plants in greenhouses should not be placed directly under overhead watering. Irrigation should only be applied to the roots in the potting media. The spores of the causative fungi are dispersed by water droplets, personnel working with wet infected plants, and mites and insects. The spores may germinate in water or inside the natural leaf openings (hydathodes, lenticels, stomata, etc.) [19].

Leaf blight

In leaf blight diseases, the tips and margins turn yellow (chlorotic). Necrosis ensues, spreads, and forms dark-brown patches variable in size. As tissue death progresses, eventually the entire lamina dies. Older leaves are generally more susceptible to this disease [20].

Fire blight

Erwinia amylovora is a bacterial pathogen causing fire blight on pear, apple, cotoneaster, and other Rosaceae. Numerous genes associated with the pathogenesis of E. amylovora and fire blight have already been characterized. The bacteria penetrate the host plant via its natural openings such as nectarthodes as well as wounds on the damp aerial organs. Unlike certain plant fungal pathogens, E. amylovora causes a localized necrosis and does not radiate to or colonize adjacent plant tissues [21].

Fusarium wilt

Fusarium wilt is a serious vascular wilt disease in crop plants. It is caused by Fusarium oxysporum which may be morphologically indistinguishable from non-pathogenic strains. The spores spread through soil, plant debris, and seeds and are difficult to eliminate from infested fields and plants. Fusarium wilt attacks potato, tomato, and other Solaneceae. The leaves and possibly the stems of infected plants lose turgidity, turn light green to greenish-yellow to brown, and finally collapse and die. Biological controls have generally been effective in the management of this disease. Plant resistance proteins also confer protection by directly or indirectly reacting with fungal pathogen virulence proteins [22].
**Leaf rust**

There are >4,000 known species of leaf rust. They infest beans, tomatoes, roses, and other crops. The lesions first appear as white, slightly raised spots on the lower epidermis of the lower (older) leaves of mature plants. Over time, the lesions are covered with red-orange spore masses. Later, pustules form and turn yellow-green and eventually black. Severe infestations cause foliar chlorosis, deformation, and eventual abscission [23].

**Late blight**

Late blight is a systemic infection caused by the fungus *Phytopthora infestans*. It often occurs during plant growth and development and may appear after flowering. It starts with the oldest leaves and manifests as green and gray spots on the leaf surfaces. As the disease progresses, the spots darken and white mycelial masses form on the lower leaf surfaces. Finally, the entire plant is infected. This pathogen does not overwinter in the soil or garden litter. It is introduced via contaminated tubers, transplants, and seeds [24].

**Early blight**

This disease commonly occurs on potato and tomato. The causative agent is *Alternaria solani*. Lesions first appear on lower epidermis of older leaves. They appear as small brown spots consisting of concentric rings arranged in a bullseye pattern. As the disease progresses, the lesions spread and cause the leaves to turn yellow, wither, and die. Finally, the infection may also radiate to other parts of the plant. The pathogen overwinters in contaminated plant tissue and is transmitted by rain, irrigation, insects, and gardening tools. It is also transmitted by infested potato tubers and tomato seeds. Despite its name, early blight may occur at any time during the plant growth period. It often attacks malnourished or distressed plants [25].

**Powdery mildew**

This disease is caused by a wide variety of fungal pathogens with limited host ranges. It first appears early in the plant growth period. Powdery mildew starts on budding leaves and forms blisters causing them to curl upwards and expose their lower epidermis. The upper surfaces of the infected leaves eventually become covered with a fine white to grayish powdery mycelial mass. Flower buds infested with powdery mildew may be inhibited from opening. Eventually, the leaves may become necrotic and absctise. The pathogen preferentially infests younger leaves with high moisture content. Mature leaves are less commonly infected [26].

**Downy mildew**

Downy mildew (*Plasmopara viticola*) affects numerous plant species. It manifests as yellow-to-white patches on the upper surfaces of mature leaves. The fungus forms downy mycelial masses that appear after rain and heavy dew and then rapidly regress after sun exposure resumes. Infected leaves turn brown and papery and may absctise despite adequate internal moisture. Downy mildew is cold tolerant and the pathogen resides in the plants and the soil. The spores are transmitted by insects, wind, rain, and gardening tools [27].

**Appearance of plant leaf fungal diseases in host alteration speciation**

Emergent diseases could arise from a rapid increase in virulence owing to geographic variation in an earlier overlooked pathogen, followed by infection of a fresh host, which is the most common cause of the development of new fungal diseases. Infection of new plants could be an outcome from an association, plant array enlargement, or host alteration. A lot of attention has been dedicated toward the extrinsic features that facilitate interaction between a pathogen and a fresh host, for example, changes in surrounding climatic conditions owing to global trade [28]. Even though, the pathogens could be premodified from contamination with communication on an original kind of tissue. It is mainly dominant illness appearance in the form of pathogen [19].

The existing development concepts are expressed as the revision on the new plant leaf. The movement of inherited genes could be terminated partially or completely. These are building the development of propagate the separation of growing diseases . Differentiations are found when observed for the
variation inside C. parasitic. Another historic case of disease appearance is occurred due to the environmental speciation via host shifts. These factors are generally confined the prospect of speciation [29]. The current concept of environment speciation might be accustomed to better understand the to better understand the aspects or opinion inside natural science as well as development encouraging the revision of pathogens toward a novel plant leaf. The horizontal genes can move through inter-specific hybridization. It has been revealed that the pathogens can give the appearance of fungal disease on the new plant leaf, but as revision is generally the outcomes of assortment with obtainable gene’s alternatives or new changes inside fungal inhabitants [21,30].

Inferences of concern regarding growing fungal diseases and environment speciation

Know that the increase in illnesses can be caused through the fungal host pathogens frequently. The abundant characters of fungal host pathogens can cause under favorable condition of environment speciation. This will have better insights into modern tools used for investigation of illness with the multiplicity of fungi. Moreover, this has implications for our understanding of how disease development affects variation, in addition to manipulative well-organized sustainable management plans. First, if changes in the host alone might be enough for speciation, then interfertility, one of the major’s criterion, generally applies for the delimiting varieties. Interfertility might be retained along gene flow between the host pathogen’s types. On the other hand, the plant leaf variation might be a well-organized barrier to facilitate speciation via plant alterations. It is indicated that two dissimilar pathogenic variabilities can be measured in the same manner as in other studies [31]. This could allow for improvement of regulatory processes that determine the specificities of every class, for example, definite fungicide resistance. Furthermore, host adaptation only could be allowed for identical quick speciation through a host shift, so the rapid development of disease indicators would be desirable to identify the strains.

Additionally, it is significant toward linkage of emergent diseases with environment speciation in the path of measure if novel diseases owe toward spread, host variety growth, or host alteration speciation. These changed circumstances are affected the control measures to be taken. Furthermore, the changing aspects and disease development will be dissimilar if the pathogens are modified toward an only against numerous plants [32,33]. At last, if host variation is adequately intended for speciation on an original plant by host changes, then the illness appearance could be reasonably spread more steadily. Furthermore, samples for the development of fungal pathogen characteristics can acquire in the account for the specificity of the pathogen’s growth. Furthermore, the disease management strategy is considered for which the pathogens are caused the main grave symptoms like plants fungal pathogen’s interact inside their hosts and cause disease appearance on new tissues [34].

Recent approaches and research trends for disease management

Single and static management strategies

The control of plant leaf pathogens is difficult partly because of their progressive changing behavior and development associated with high genetic variation and rapid growth periods, which allow them to overcome the recently highest effectual disease (Table 2 and Figure 2). It is described that the proportions of augmented pesticide are used to indicate the reduced competence with economic comeback of used pesticides toward plant disease. Generally, pesticides could be used in a regime that regulates the timing, frequency, and amount of application, independent of the specific resistance status, and ecological environments. The safe and immobile approach of pesticide utilization does not only decrease the effectiveness of the management and increase the expenses, but also has adverse environmental consequences, such as toxicity toward livestock [11,35].

Gray box of plant disease plague mechanisms

Rapid and prompt crop disease prediction, recognition, and analysis are essential. Molecular methods are currently being used to characterize and analyze plant pathogenesis. Despite the availability of these techniques, they are not universally applied here as the agricultural industry may still rely upon agronomists with traditional knowledge
using conventional research methods. Accurate and effective crop disease assessment requires an in-depth understanding of the pathogenesis, pathogen–environment intercommunication, and influential biotic and abiotic factors involved. In view of these complexities, with a few notable exceptions, accurate, timely crop disease forecasting remains imperfect [34].

Financial examination of host leaf disease management

Optimistic externalities can show the profits by the disease management within the farms for reduce the possible pathogens. Presently, these optimistic externalities are not listed in the commercial investigation of host disease management [36]. Growers must account for expenses incurred from pesticide application but not for those connected to residue elimination or site remediation. As growers are only compensated for their crop disease management costs, they select strategies that generate the highest immediate returns possibly while discounting the potential negative ecological impacts of the pesticide deployments. Certain highly effective disease management strategies are in current use without adequate regard for their long-term adverse environmental effects [37].

Prospects of plant leaf disease management

Sustainable leaf disease management is needed for the multidimensional concern, includes social, financial side with environmental science. The host leaf disease management approach could be useful to explore not only single traits but also to shield the natural atmosphere. Future investigations in biological plant leaf disease management must focus on:

1. global and evolutionary outlines of leaf disease under changing environmental conditions and production demands;
2. ecological impacts on crop yield and pathogen resistance;
3. social and financial evaluation of leaf disease outbreaks and their management; and
4. technological development for the combined management of major crop diseases and environmental protection [38].
5. assurance of food security, safety, quality, and variety as human populations grow and living standards improve;
6. declining crop production as a result of competitive use and overuse of farmlands and possible soil nutrient depletion;

Future plant disease management should aim to improve food safety for the growing population while simultaneously preserving ecosystem integrity by reducing dependence on the most commonly used resources. The leaf organizations are the chief component for the normal adjustment of confrontation, removal and remediation strategy. It is directed through characters of host–pathogen relations by evolutionary bionetwork values to make ecological (biotic and abiotic) circumstances positive for host development [39,40].

Fungal diseases intimidate financially prominent plants leading toward important financial losses universal. Fungal pathogens can damage the food production for worldwide food demand. The significance of emergent fungal plant leaf pathogens must be assessed with progressive, rapid, and accurate disease recognition tools. Unfortunately, many currently available technologies and strategies are inadequate in these respects. They may be cost- and labor-intensive, slow, inefficient, and complex. Molecular and immunological tools generate highly informative data but require special equipment and personnel trained and qualified to use them. Novel methods and approaches must be able to deliver accurate results rapidly and identify plant leaf diseases early in their progression. Biosensors help improve the disease recognition capabilities of diagnostic tools. Several of them have been described in the literature [41,42]. Their efficacy has been tested in artificial field trials under controlled conditions, but they still require evaluation on imminent disease outbreaks under natural and real crop field conditions.

Biosensors may be designed and programmed for high throughput using nanomaterials, nanofabrication technology, and DNA sequencing [37]. The ongoing emergence of microbial plant pathogens
could result in major financial and yield losses in agriculture. Bacterial leaf diseases are treated with copper-based compounds and antibiotics. However, pathogens may acquire resistance to them and they may have negative ecological impacts. Moreover, the improvement of antibiotics has been helpful for the ability of a well-organized crop shield. This condition is driving the investigators toward travel around novel and non-conventional methods. Antimicrobial peptides have attracted attention as potential biological controls for plant leaf diseases. These compounds have amphipathic structures that interact with the anionic phospholipids of fungal membrane and cause their disruption.

**Challenges and upcoming outlooks**

Accurate diagnostic methods that can be used *in situ* to detect plant disease must be developed, tested, and perfected in the near future. New fungal leaf disease identification technologies designed for field use may incorporate DNA sequencing. However, sampling and the use of this technology in the form of transportable biosensors may be logistically challenging. Moreover, onsite DNA sequencing expertise may be difficult to acquire as plant leaves have large and complex genomes. For data mining and analysis, high-throughput computational biology and informatics will be essential. At this time, there are no initiatives being undertaken to improve transportable DNA sequencing biosensors that effectively identify fungal plant pathogens. Sequencing with microfluidic devices will facilitate the recognition of numerous fungal plant pathogens in the future. The application of biosensors in fungal diagnostics will enable the development of miniaturized, cost-effective, accurate, simple, and rapid apparatus that can readily be used on-site and in the field even by untrained personnel.

Extensive investigation is necessary for additional manipulation of biological approaches and viable applications. However, recognition of molecular markers connected through disease-resistant key genes and quantitative trait loci display the possible application of marker-aided choice in plant disease resistance. PCR and flow cytometry may eventually be implemented in the genetic recognition of existing pathogens and the identification of emergent ones. Genetic investigations could detect sources of pathogen and host resistance in diseases such as powdery mildew. Biological control in the form of seed dressing has been evaluated against root rot fungal pathogens such as *Trichoderma*. RNA sequencing and double-stranded RNA may be deployed to recognize exclusively fungal genetic biomarkers.

Management of foliar fungal diseases involves both physical and cultivation approaches including the manipulation of plant resistance and the use of synthetic fungicides, bioproducts such as natural plant extracts, and defense activators or biostimulants. The application of thermography for disease evaluation is limited and flawed as it is highly sensitive to fluctuations in ambient environmental conditions. Another option is the examination of individual targeted molecules. However, it is not generally accepted as it discloses only a fraction of the entire chemical profile. Analysis of untargeted volatile organic compounds provides a broader view of the host biochemical organization. This technique may be applied even when highly dissimilar volatile organic compounds must be immediately and simultaneously detected and quantified. Remote-sensing methods may also prove useful in fungal crop disease monitoring.

**Conclusions**

Integration of biosensing mechanisms and DNA sequencing technologies will enhance the accuracy of fungal crop disease detection. Here, we reviewed several major plant foliar fungal pathogens and the factors that induce environmental speciation and host shifts in them. Fungal leaf diseases are responsible for major yield losses in commercially important crops worldwide. In addition to evaluating the agricultural significance of fungal leaf diseases, it is necessary to develop tools that enable the rapid recognition of these disease tools. One possible approach is the identification of environmental speciation in these pathogens.

**Highlights**

- This review explored the factors that controlled the emergence of leaf fungal diseases;
• Recent approaches and research trends are helpful to leaf disease management;
• Economic analysis played an important role in planning of plant disease management.

Acknowledgements

The authors are grateful for the financial support from National Natural Science Foundation of China (NSFC-81460632), Distinguished High-Level Talents Research Grant from a Guizhou Science and Technology Corporation Platform Talents Fund (Grant No.: [2017]5733-001 & CK-1130-002), and Zunyi Medical University, China, for their support and cooperation for this investigation.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by the National Natural Science Foundation of China [NSFC-81460632]; Guizhou Science and Technology Corporation Platform Talents Fund ([2017] 5733-001 & CK-1130-002).

ORCID

Surendra Sarsaiya http://orcid.org/0000-0002-0850-8592
Jingshan Shi http://orcid.org/0000-0002-2537-1751

References

[1] Lazarovits G, Turnbull A, Johnston-Monje D. Plant health management: biological control of plant pathogens. In: Van Alfen NK, editor. Encyclopedia of agriculture and food systems. New York, NY: Academic Press; 2014. p. 388–399.

[2] Iqbal Z, Khan MA, Sharif M, et al. An automated detection and classification of citrus plant diseases using image processing techniques: A review. Comput Electron Agric. 2018;153:12–32.

[3] Baijee B, Ito S, Sunpapao A. *Trichoderma asperellum* T1 mediated antifungal activity and induced defense response against leaf spot fungi in lettuce (*Lactuca sativa* L.). Physiol Mol Plant P. 2019;106:96–101.

[4] Singh V, Misra AK. Detection of plant leaf diseases using image segmentation and soft computing techniques. IPA. 2017;4(1):41–49.

[5] Barbedo JGA. A review on the main challenges in automatic plant disease identification based on visible range images. Biosyst Eng. 2016;144:52–60.

[6] Prado S, Li Y, Nay B. Chapter 8 - Diversity and ecological significance of fungal endophyte natural products. In: -ur-Rahman A, editor. Studies in natural products chemistry. Vol. 36. UK: Elsevier; 2012. p. 249–296.

[7] Tör M, Woods-Tör A. Fungal and oomycete diseases. In: Thomas B, Murray BG, Murphy DJ, editors. Encyclopedia of applied plant sciences. second ed. New York, NY: Academic Press; 2017. p. 77–82.

[8] Ferentinos KP. Deep learning models for plant disease detection and diagnosis. Comput Electron Agric. 2018;145:311–318.

[9] Rahman SFSA, Singh E, Pieterse CMJ, et al. Emerging microbial bio control strategies for plant pathogens. Plant Sci. 2018;267:102–111.

[10] Silva OC, Santos HAA, Pria MD, et al. Potassium phosphate for control of downy mildew of soybean. Crop Prot. 2011;30(6):598–604.

[11] Giraud T, Gladieux P, Gavrilets S. Linking the emergence of fungal plant diseases with ecological speciation. Trends Ecol Evol. 2010;25(7):387–395.

[12] Myo EM, Liu B, Ma J, et al. Evaluation of *Bacillus velezensis* NKG-2 for bio-control activities against fungal diseases and potential plant growth promotion. Biol Control. 2019;134:23–31.

[13] Sathiyabam M, Balasubramanian R. Protection of groundnut plants from rust disease by application of glucan isolated from a bio control agent *Acremonium obclavatum*. Int J Biol Macromol. 2018;116:316–319.

[14] Straat L, Graaff LH. Pathway transfer in fungi. Bioengineered. 2014;5(5):335–339.

[15] Gugino BK, Carroll JE, Widmer TL, et al. Field evaluation of carrot cultivars for susceptibility to fungal leaf blight diseases in New York. Crop Prot. 2007;26(5):709–714.

[16] Neto YAAH, Garzon NGR, Pedezzi R, et al. Specificity of peptidases secreted by filamentous fungi. Bioengineered. 2018;9(1):30–37.

[17] Sarsaiya S, Shi J, Chen J. A comprehensive review on fungal endophytes and its dynamics on Orchidaceae plants: current research, challenges, and future possibilities. Bioengineered. 2019;10(1):316–334.

[18] Matthalli C, Gedefaye E, Endalamaw F, et al. Plant leaf diseases detection and auto-medicine. IoT. 2018;1:2:67–73.

[19] Camó C, Bonaterra A, Badosa E, et al. Antimicrobial peptide KSL-W and analogues: promising agents to control plant diseases. Peptide. 2019;112:85–95.

[20] Li M, Ma GS, Lian H, et al. The effects of *Trichoderma* on preventing cucumber fusarium wilt and regulating cucumber physiology. J Integr Agric. 2019;18(3):607–617.

[21] Flachowsky H, Halborth H, Treutter D, et al. Silencing of flavanone-3-hydroxylase in apple (*Malus × domestica Borkh.*) leads to accumulation of flavanones, but not to reduced fire blight susceptibility. Plant Physiol Biochem. 2012;51:18–25.
[22] Nalini S, Parthasarathi R. Optimization of rhamnolipid biosurfactant production from Serratia rubidaea SNAU02 under solid-state fermentation and its biocontrol efficacy against Fusarium wilt of eggplant. Ann Agric Sci. 2018;16(2):108–115.

[23] Ono Y, Wabayuno D. Phragmidium satoanum, a new rust pathogen of Rosa hirtula in Japan. Mycosci. 2019;60(4):1–29.

[24] Liljeroth E, Lankinen A, Wiik L, et al. Potassium phosphate combined with reduced doses of fungicides provides efficient protection against potato late blight in large-scale field trials. Crop Prot. 2016;86:42–55.

[25] Babu AN, Jogaiah S, Ito SI, et al. Improvement of growth, fruit weight and early blight disease protection of tomato plants by rhizosphere bacteria is correlated with their beneficial traits and induced biosynthesis of antioxidant peroxidase and polyphenol oxidase. Plant Sci. 2015;231:62–73.

[26] Bai X, Fu Z, Stankovski S, et al. A three-dimensional threshold algorithm based on histogram reconstruction and dimensionality reduction for registering cucumber powdery mildew. Comput Electron Agric. 2019;158:211–218.

[27] Shao D, Tian M. A PCR approach to quantify the growth of basil downy mildew pathogen Peronospora belbahrii during infection. Curr Plant Biol. 2018;15:2–7.

[28] Bate AM, Jones G, Kleczkowski A, et al. Modelling the impact and control of an infectious disease in a plant nursery with infected plant material inputs. Ecol Model. 2016;334:27–43.

[29] Wongkhamchan A, Chankaew S, Monkham T, et al. Broad resistance of RD6 introgression lines with xa5 gene from IR62266 rice variety to bacterial leaf blight disease for rice production in Northeastern Thailand. Anres. 2018;52(3):241–245.

[30] Liu L, Yang H, Shin H, et al. How to achieve high-level expression of microbial enzymes. Bioengineered. 2013;4(4):212–223.

[31] Ghobanpour M, Omidvari M, Dahaji PA, et al. Mechanisms underlying the protective effects of beneficial fungi against plant diseases. Biol Control. 2018;117:147–157.

[32] Yang P, Elsas JDV. Mechanisms and ecological implications of the movement of bacteria in soil. Appl Soil Ecol. 2018;129:112–120.

[33] Kemppainen MJ, Pardo AG. Gene knockdown by ihpRNA-triggering in the ectomycorrhizal basidiomycete fungus Laccariabicolor. Bioengineered Bugs. 2010;1(5):354–358.

[34] Steenkamp ET, Wingfield MJ, McTaggart AR, et al. Fungal species and their boundaries matter – definitions, mechanisms and practical implications. Fungal Biol Rev. 2018;32(2):104–116.

[35] Tenorio-Salgado S, Tinoco R, Vazquez-Duhalt R, et al. Identification of volatile compounds produced by the bacterium Burkholderiatropicaplant disease models with cultural control strategies and economic thresholds. Math Comput Simul. 2010;80(5):894–921.

[36] Tang S, Xiao Y, Cheke RA. Dynamical analysis of plant disease models with cultural control strategies and economic thresholds. Math Comput Simul. 2010;80(5):894–921.

[37] Ray M, Ray A, Dash S, et al. Fungal disease detection in plants: traditional assays, novel diagnostic techniques and biosensors. Biosens Bioelectron. 2017;87:708–723.

[38] Bamisile BS, Dash CK, Akutse KS, et al. Prospects of endophytic fungal entomopathogens as bio control and plant growth promoting agents: an insight on how artificial inoculation methods affect endophytic colonization of host plants. Microbiol Res. 2018;217:34–50.

[39] Wade EW, Ma C, White J. Nanoparticles for plant disease management. Curr Opin Environ Sci Health. 2018;6:66–70.

[40] Shi TQ, Peng H, Zeng SY, et al. Microbial production of plant hormones: opportunities and challenges. Bioengineered. 2017;8(2):124–128.

[41] Ayodele VO, Olowe OM, Afolabi CG, et al. Identification, assessment of diseases and agronomic parameters of Curcuma amada Roxb (Mango ginger). Curr Plant Biol. 2018;15:51–57.

[42] Kemppainen MJ, Pardo AG. Transformation of the mycorrhizal fungus Laccariabicolor using Agrobacterium tumefaciens. Bioengineered Bugs. 2011;2(1):38–44.

[43] Miara MD, Bendif H, Rebbas K, et al. Medicinal plants and their traditional uses in the highland region of Bordj Bou Arreridj (Northeast Algeria). J Herbs Med. 2019;1–56. DOI:10.1016/j.hermed.2019.100262.

[44] Silva NID, Brooks S, Lumyong S, et al. Use of endophytes as bio control agents. Fungal Biol Rev. 2019;33(2):133–148.

[45] He DC, Zhan JS, Xie LH. Problems, challenges and future perspectives. Trends Plant Sci. 2018;33(2):133–148.

[46] Singh AK, Ganapathysubramanian B, Sarkar S, et al. Deep learning for plant stress phenotyping: trends and future perspectives. Trends Plant Sci. 2018;23(10):883–898.

[47] Pandey AK, Buralakoti RR, Kenyon L, et al. Perspectives and challenges for sustainable management of fungal diseases of mungbean [Vigna radiata (L.) R. Wilczek var. radiata]: a review. Front Environ Sci. 2018;6:53.

[48] Sarasaiya S, Jia Q, Fan X, et al. First report of leaf black circular spots on Dendrobium nobile caused by Trichoderma longibrachiatum in Guizhou Province, China. Plant Dis. 2019. DOI:10.1094/pdis-03-19-0672-pdn.

[49] McLaughlin AG, Wytnick N, Walker PL, et al. Identification and application of exogenous dsRNA confers plant protection against Sclerotinia sclerotiorum and Botrytis cinerea. Sci Rep. 2018;8:7320.

[50] Martinelli F, Scalenge R, Davino S, et al. Advanced methods of plant disease detection. A review. Agron Sustain Dev. 2015;35(1):25.
[51] Nair SK, Prasanna BM, Rathore RS, et al. Genetic analysis of resistance to sorghum downy mildew and Rajasthan downy mildew in maize (Zea mays L.). Field Crops Res. 2004;89(2–3):379–387.
[52] Gunen Y, Misirli A, Gulcan R. Leaf phenolic content of pear cultivars resistant or susceptible to fire blight. Sci Hort. 2005;105(2):213–221.
[53] Abuley IK, Nielsen BJ. Integrating cultivar resistance into the TOMCAST model to control early blight of potato, caused by Alternaria solani. Crop Prot. 2019;117:69–76.
[54] Masuya H, Kusunoki M, Kosaka H, et al. Haradamyces foliicola anam. gen. et sp. nov., a cause of zonate leaf blight disease in Cornus florida in Japan. Mycol Res. 2009;113(2):173–181.
[55] Abou-Taleb EM, Aboshosha SM, Sherif EM, et al. Genetic diversity among late blight resistant and susceptible potato genotypes. Saudi J Biol Sci. 2010;17:133–138.
[56] Djennane S, Cesbron C, Sourice S, et al. Iron homeostasis and fire blight susceptibility in transgenic pear plants overexpressing a pea ferritin gene. Plant Sci. 2011;180(5):694–701.
[57] Kiewnick AB, Altenbach D, Oberhansli T, et al. A rapid lateral-flow immunoassay for phytosanitary detection of Erwinia amylovora and on-site fire blight diagnosis. J Microbiol Methods. 2011;87(1):1–9.
[58] Gindro K, Villaverde VA, Voinesco F, et al. Susceptibility to downy mildew in grape clusters: new microscopical and biochemical insights. Plant Physiol Biochem. 2012;52:140–146.
[59] Lestari P, Rahayu S, Widiyatno. Dynamics of gall rust disease on Sengon (Falcataaria Moluccana) in various agroforestry patterns. Procedia Environ Sci. 2013;17:167–171.
[60] Meghvansi MK, Khán MH, Gupta R, et al. Identification of a new species of Cercospora causing leaf spot disease in Capsicum annuum in northeastern India. Res Microbiol. 2013;164(9):894–902.
[61] Gaucher M, Beronville TDD, Guyot S, et al. Same amino, different weapons: enzymatic extracts from two apple genotypes with contrasted susceptibilities to fire blight (Erwinia amylovora) differentially convert phloridzin and phloretin in vitro. Plant Physiol Biochem. 2013;72:178–189.
[62] Pota S, Chatasiri S, Unartngam J, et al. Taxonomic identity of a Phakopsora fungus causing the grapevine leaf rust disease in Southeast Asia and Australasia. Mycol Sci. 2015;56(2):198–204.
[63] IM S, Joseph L, Fry WE. Development and implementation of the BlightPro decision support system for potato and tomato late blight management. Comput Electron Agric. 2015;115:57–65.
[64] Mohammadi P, Bahramnejad B, Badakhshan H, et al. DNA methylation changes in fusarium wilt resistant and sensitive chickpea genotypes (Cicer arietinum L.). Physiol Mol Plant P. 2015;91:72–80.
[65] Anup CP, Melvin P, Shilpa N, et al. Proteomic analysis of elicitation of downy mildew disease resistance in pearl millet by seed priming with β-aminobutyric acid and Pseudomonas fluorescens. J Proteomics. 2015;120:58–74.
[66] Shabana YM, Abdalla ME, Shahin AA, et al. Efficiency of plant extracts in controlling wheat leaf rust disease caused by Puccinia triticina. Egypt J Basic Appl Sci. 2017;4(1):67–73.
[67] He SSY, Dai C, Duan C, et al. Two major er1 alleles confer powdery mildew resistance in three pea cultivars bred in Yunnan Province, China. Crop J. 2016;4(5):353–359.
[68] Liu Y, Li B, Wang C, et al. Genetics and molecular marker identification of a resistance to glomerella leaf spot in apple. Hortic Plant J. 2016;2(3):121–125.
[69] Hajarifar R, Kolics B, Cernák I, et al. Expression of biotic stress response genes to Phytophthora infestans inoculation in White Lady, a potato cultivar with race-specific resistance to late blight. Physiol Mol Plant P. 2016;93:22–28.
[70] Kumar R, Mina U, Gogoi R, et al. Effect of elevated temperature and carbon dioxide levels on maydis leaf blight disease tolerance attributes in maize. Agric Ecosyst Environ. 2016;231:98–104.
[71] Nassar AMK, Ibrahim AA, Adss IAA. 2,4-Dichlorophenoxy acetic acid, abscisic acid, and hydrogen peroxide induced resistance-related components against potato early blight (Alternaria solani, Soraier). Ann Agric Sci. 2016;61(1):15–23.
[72] Zeng F, Wang C, Zhang G, et al. Draft genome sequence of Cercospora sojina isolate S9, a fungus causing frogeye leaf spot (FLS) disease of soybean. Genom Data. 2017;12:79–80.
[73] Singh VK, Singh HB, Upadhyay RS. Role of fusaric acid in the development of Fusarium wilt symptoms in tomato: physiological, biochemical and proteomic perspectives. Plant Physiol Biochem. 2017;118:320–332.
[74] Mubeen S, Rafique M, Munis MFH, et al. Study of southern corn leaf blight (SCLB) on maize genotypes and its effect on yield. J Saudi Soc Agric Sci. 2017;16(3):210–217.
[75] Ganopoulos I, Mylona P, Mellidou I, et al. Microsatellite genotyping and molecular screening of pea (Pisum sativum L.) germplasm with high-resolution melting analysis for resistance to powdery mildew. Plant Genet. 2018;15:1–5.
[76] Mahmud MS, Zaman QU, Esau TJ, et al. Development of an artificial cloud lighting condition system using machine vision for strawberry powdery mildew disease detection. Comput Electron Agric. 2019;158:219–225.
[77] Goyal N, Bhatia G, Sharma S, et al. Genome-wide characterization revealed role of NBS-LRR genes during powdery mildew infection in Vitis vinifera. Genomics. 2019;1–11. DOI:10.1016/j.jgeno.2019.02.011
[78] Chen T, Zhang J, Chen Y, et al. Detection of peanut leaf spots disease using canopy hyperspectral reflectance. Comput Agric Electron Agric. 2019;156:677–683.
[79] Cox KL, Babilonia K, Wheeler T, et al. Return of old foes-recurrence of bacterial blight and Fusarium wilt of cotton. Curr Opin Plant Biol. 2019;50:95–103.

[80] Andjic V, Carnegie AJ, Pegg GS, et al. 23 years of research on Teratosphaeria leaf blight of Eucalyptus. For Ecol Manage. 2019;443:19–27.

[81] Pacilly FCA, Bueren ETLV, Groot JCJ, et al. Moving perceptions on potato late blight control: workshops with model-based scenarios. Crop Prot. 2019;119:76–87.

[82] Omar NH, Mohd M, Nor NMIM, et al. Characterization and pathogenicity of Fusarium species associated with leaf spot of mango (Mangifera indica L.). Microb Pathog. 2018;114:362–368.

[83] Raid RN, Pernezny K, Havranek N, et al. Weather-based forecasting systems reduce fungicide use for early blight of celery. Crop Prot. 2008;27 (3–5):396–402.