Does therapeutic hypothermia during extracorporeal cardiopulmonary resuscitation preserve cardiac function?

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Abstract

Background: Extracorporeal cardiopulmonary resuscitation (E-CPR) is increasingly used as a rescue method in the management of cardiac arrest and provides the opportunity to rapidly induce therapeutic hypothermia. The survival after a cardiac arrest is related to post-arrest cardiac function, and the application of therapeutic hypothermia post-arrest is hypothesized to improve cardiac outcome. The present animal study compares normothermic and hypothermic E-CPR considering resuscitation success, post-arrest left ventricular function and magnitude of myocardial injury.

Methods: After a 15-min untreated ventricular fibrillation, the pigs (n = 20) were randomized to either normothermic (38 °C) or hypothermic (32–33 °C) E-CPR. Defibrillation terminated ventricular fibrillation after 5 min of E-CPR, and extracorporeal support continued for 2 h, followed by warming, weaning and a stabilization period. Magnetic resonance imaging and left ventricle pressure measurements were used to assess left ventricular function pre-arrest and 5 h post-arrest. Myocardial injury was estimated by serum concentrations of cardiac TroponinT and Aspartate transaminase (ASAT).

Results: E-CPR resuscitated all animals and the hypothermic strategy induced therapeutic hypothermia within minutes without impairment of the resuscitation success rate. All animals suffered a severe global systolic left ventricular dysfunction post-arrest with 50–70% reductions in stroke volume, ejection fraction, wall thickening, strain and mitral annular plane systolic excursion. Serum concentrations of cardiac TroponinT and ASAT increased considerably post-arrest. No significant differences were found between the two groups.

Conclusions: Two-hour therapeutic hypothermia during E-CPR offers an equal resuscitation success rate, but does not preserve the post-arrest cardiac function nor reduce the magnitude of myocardial injury, compared to normothermic E-CPR.

Trial registration FOTS 4611/13 registered 25 October 2012

Keywords: Cardiac function, Cardiopulmonary resuscitation, Extracorporeal circulation, Extracorporeal membrane oxygenation, Therapeutic hypothermia

Background

Survival after cardiac arrest is greatly influenced by early post-arrest cardiac function [1, 2]. Hence, cardiopulmonary resuscitation (CPR) strategies that preserve post-arrest cardiac function may improve outcome. Extracorporeal CPR (E-CPR) by veno-arterial extracorporeal membrane oxygenation (ECMO) is increasingly used when standard CPR fails. Promising results have been reported by using E-CPR as a rescue method within brief timeframes for selected cases [3–6]. Therapeutic hypothermia (HT, 32–34 °C) is widely used for patients resuscitated from cardiac arrest as it is believed to exhibit cardiovascular [7, 8] and neurological benefits [9–11].
To achieve cardiac benefit from HT the importance of early and rapid cooling has been emphasized in experimental studies [12–14]. E-CPR provides the opportunity to rapidly induce HT, but whether hypothermic E-CPR preserves post-arrest cardiac function and hence improves outcome, is not known.

A severe cardiac dysfunction following normothermic E-CPR is recently demonstrated in pigs [15]. The present study aimed to investigate if HT during E-CPR improves cardiac outcome early post-arrest. We hypothesized that hypothermic E-CPR offers an equal resuscitation success rate, but with a better preserved post-arrest left ventricular (LV) function and less myocardial injury compared to normothermic E-CPR.

Methods
Design
A prospective controlled block-randomized animal study was completed to compare a normothermic (38-ECPR, n = 10) and a hypothermic (32-ECPR, n = 10) E-CPR group.

Animal welfare
The experimental protocol was approved by the Norwegian National Animal Research Authority and the animal experiments were performed in accordance with the European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes (European Council, ETS No. 170). Detailed information according to the ARRIVE guidelines is presented in Additional file 1: Table S1 [16]. With respect to animal welfare’s 3R-principle, eight 38-ECPR animals were included in a separate methodological study demonstrating E-CPR associated post-arrest LV dysfunction by cardiac magnetic resonance imaging (MRI) per se [15].

Animal preparation
The animal preparation included premedication in the pig enclosure and total intravenous anaesthesia in the operating theatre, mechanical ventilation and a succeeding surgical tracheotomy and placements of intravascular catheters and ECMO-cannulas as recently described [15]. Ringer’s acetate solution was infused at 10 ml/kg/h.

Experimental protocol
Baseline assessments
After the preparation and a following 30-min stabilization period, baseline cardiac MRI (Philips Achieva 3 Tesla, Philips Medical Systems, DA Best, Netherland) and haemodynamic measurements of LV function (MPR-500, Millar Instruments, Houston, TX, USA) were obtained. Arterial and mixed venous blood samples were analyzed (ABL 800 Flex, Bergman Diagnostika, Kjeller, Norway) and serum concentrations of cardiac Troponin T (cTnT) and Aspartate aminotransferase (ASAT) were measured.

ECMO and cardiac arrest
After baseline assessments, the pig was connected to a Ringer’s acetate-primed femoro-jugular veno-arterial ECMO circuit (Biopump + BPX-80/Affinity NT/Biomedicus 550, Medtronic Inc, Minneapolis, MN, USA) featuring an oxygen/air mixer (Sechrist Model 20090, Sechrist Industries, Anaheim, CA, USA) to adjust sweep gas oxygen content and sweep gas flow rate. A connected heat-exchanger (Stöckert Heater-Cooler System 3T, Sorin Group, Milano, Italy) enabled animal blood temperature control.

After an intravenous injection of 2 mg/kg heparin, an electrical stimulator connected to a right ventricular pacing lead (Qstim 5Fr, VascoMed GmbH, Binzen, Germany) induced ventricular fibrillation (VF), confirmed by ECG shape and aortic fibrillation (VF), confirmed by ECG shape and aortic fibrillation (VF). After an 15 min of untreated VF the animals received either normothermic [pulmonary artery blood temperature 38.0 °C (normothermia in the pig)] or hypothermic (32.0–33.0 °C) E-CPR at maximum ECMO blood flow rate with a 100% oxygen sweep gas set at the same flow rate as the ECMO blood flow rate.

HT in the 32-ECPR group was achieved using 20.0 °C priming solution with later adjustments at the heat-exchanger. The heat-exchanger thermostat at 38.0 °C ensured normothermia in the 38-ECPR group.

After 5 min of E-CPR 360 Joule monofasic defibrillations (CodeMaster XL + Hewlett Packard, Lexington, KY, USA) were provided until regain of spontaneous cardiac beating (ROSB) with extracorporeal support continuing at unchanged blood flow rate and temperature target for 120 min. In the 32-ECPR group a 30-min warming period followed, whereas a corresponding 30-min continued run at 38.0 °C was provided in the 38-ECPR group.

The mean aortic blood pressure (MAP) target (≥50 mmHg) and pulse-pressure target (≥15 mmHg) after ROSB were met using repeated 10–25 µg adrenaline (epinephrine) intravenously followed by dobutamine infusion if needed [17].

Weaning from ECMO
After the 120 + 30-min extracorporeal support, with all animals being normothermic, a step-wise separation from ECMO (weaning) was completed during a 60-min period, and the animals were allowed to stabilize after weaning for another 60 min before the post-arrest assessments.
Post-arrest assessments
At 285 min post-arrest LV function was re-assessed by LV pressure measurements and MRI. Finally, a second arterial and mixed venous blood sample were analyzed and blood samples for cTnT and ASAT measurements were collected, before the pig was euthanized.

Measurements
MRI and haemodynamic measurements
Magnetic resonance imaging was used to assess LV volumes and function (end-systolic volume (ESV), end-diastolic volume (EDV), stroke-volume (SV = EDV – ESV), ejection fraction (EF = SV/EDV), cardiac output [CO = SV heart rate (HR)] mid LV radial wall thickening, mitral annular plane systolic excursion (MAPSE), and peak global systolic LV circumferential strain) together with LV pressure measurements (maximum systolic LV pressure (LVP\textsubscript{max}), maximum positive and negative first time derivate of LV pressure (dP/dt\textsubscript{max} and dP/dt\textsubscript{min}), end-diastolic LV pressure (EDP), end-systolic LV pressure (ESP), isovolumetric relaxation constant (tau), and arterial elastance (E_a = ESP/SV)) [15, 18]. Continuous measurements of blood temperature (Figs. 1, 2), HR and MAP (Fig. 3) outside the MRI were recorded throughout the experiments.

cTnT and ASAT
The serum concentration measurements of cTnT and ASAT pre-arrest and 6 h post-arrest were performed by an electro-chemiluminescence immunoassay (Troponin T hs, Roche Diagnostics, Rotkreuz, Switzerland) and by an UV-test with pyridoxal phosphate activation (ASAT, Roche Diagnostics) using an automated clinical chemistry analyzer (Modular analytical platform, module E170 and P800, Roche Diagnostics) to estimate myocardial injury.

Myocardial tissue staining
Eleven (38-ECPR n = 5, 32-ECPR n = 6) hearts excised immediate post-mortem were sliced and stained in 1% triphenyl tetrazolium chloride (TTC, Sigma Chemical Co., St. Louis, MO, USA) in phosphate buffer and examined for myocardial infarction [19, 20].

Statistical analysis
Statistical analyses were made using Graphpad prism 6.04 (GraphPad Software, La Jolla, CA, USA). Data are reported as mean ± standard deviation if not otherwise stated. The statistical significance level α was set to 0.05 and power 1 − β to 0.80.

The study sample size was estimated by a prospective power analysis. The least detectable difference considered as clinically significant between cardiac function variables was 15% of baseline values. Cardiac function variability in range of 5–15% of baseline values in pilot experiments made a calculated sample size of 20 necessary to achieve the desired power.
A paired two-tailed Student’s t test (t) was used to compare baseline and post-arrest measurements within each treatment group, and an unpaired two sample t-test (T) was used to compare post-arrest measurements between the two different treatment groups. Alternatively, a two-tailed Mann–Whitney test (MW) of group differences with exact p-value was used for data not normally distributed.

**Results**

Animal weight (49.2 ± 2.8 kg), preparation time (122 ± 22 min) and cardiac MRI scan time (63 ± 15 min) were similar in the two groups. Neither activated clotting-time prior to cardiac arrest (330 ± 76 s) nor ECMO circuit priming volume (544 ± 22 ml) differed between the two groups, and the experiment durations were also similar, averaging 762 ± 63 min (32-ECPR 777 ± 58 min vs. 38-ECPR 745 ± 67 min; T, p = 0.27).

One animal in the 38-ECPR group was euthanized after ROSB, and was thus excluded from further analyses, because MAP could not be sustained as dictated by the protocol, due to ECMO venous cannula malfunctioning. At initiation of VF the blood temperature was 38.0 ± 0.2 °C and was maintained at this level in the 38-ECPR group (Fig. 1). In the 32-ECPR group the temperature quickly dropped after initiation of hypothermic E-CPR and was 33.3 ± 1.0 °C at the time of defibrillation (Fig. 2). It further dropped to the targeted 32–33 °C within the 1st min after defibrillation (5.25 ± 4.8 min from start E-CPR) and was kept stable at this level until warming.

**Defibrillation**

All animals were successfully defibrillated by a median of 1 shock (range 1–6) with no significant differences between the groups (MW, p = 0.99). After ROSB three animals in the 38-ECPR group spontaneously had a second VF, and immediately received a median of 1 (range 1–3) additional defibrillations. No additional defibrillations were needed in the 32-ECPR group (MW, p = 0.21).

MAP at the time of defibrillation was 54 ± 9 mmHg in the 38-ECPR group and 60 ± 5 mmHg in the 32-ECPR group (T, p = 0.082) (Fig. 3) with an ECMO blood flow rate of 4.6 ± 0.1 l/min and 4.4 ± 0.2 l/min in the two groups, respectively (T, p = 0.055).
Inotropes

The adrenaline dosage [median (range)] was 100 µg (0–340 µg) in the 38-ECPR group vs. 183 µg (50–1700 µg) in the 32-ECPR group (MW, p = 0.074). The dobutamine requirements differed between the groups [38-ECPR dobutamine [median (range)] = 15.8 µg (0–31.2 mg) vs. 32-ECPR dobutamine = 36.2 mg (10–93.6 mg)] (MW, p = 0.0076), but the dosages after ECMO weaning to the end of the experiments were not significantly different; four of nine animals in the 38-ECPR group and five of ten in the 32-ECPR group received dobutamine dosages [median (range)] of 2 mg (0.5–2.2) and 2.6 mg (0.01–5 mg), respectively, after weaning (MW, p = 0.40).

Haemodynamic measurements and blood gas analyses

HR increased similarly post-arrest in both groups (Table 1), limiting the reductions in CO. The increased Ea post-arrest did not differ between the groups, and the dP/dt_max, dP/dt_min, EDP, EDP/EDV relationship, and tau did not change significantly from baseline values in either group.

Mixed venous oxygen saturation dropped considerably and group-similarly post-arrest (Table 1). A small decrease in haemoglobin content, base excess and pH was also measured post-arrest, with an accompanying increase in arterial lactate.

Cardiac MRI

In both groups the LV function was severely affected post-arrest (Fig. 4). EF decreased from 61 ± 4% to 33 ± 8% (t, p < 0.001) and SV decreased from 61 ± 10 to 30 ± 6 ml (t, p < 0.001) in the 38-ECPR group. In the 32-ECPR group EF decreased from 65 ± 7 to 34 ± 7% (t, p < 0.001) (T, p = 0.94) and SV decreased from 61 ± 13 to 29 ± 8 ml (t, p < 0.001) (T, p = 0.60). In the 38-ECPR group the lowered EF was only related to the increase in ESV from 38 ± 6 to 61 ± 14 ml (t, p = 0.001) as the EDV was maintained at 92 ± 14 ml from a baseline EDV value of 99 ± 13 ml (p = 0.44). There was a similar increase in ESV in the 32-ECPR group from 34 ± 10 to 56 ± 11 ml (t, p < 0.001). In this group, however, EDV was moderately reduced from a baseline EDV value of 95 ± 18 ml to post-arrest EDV 85 ± 14 ml (t, p = 0.0044). Despite severe tachycardia CO was reduced in most animals post-arrest (six of nine in the 38-ECPR group; nine of ten in the 32-ECPR group), but CO did not significantly differ between the two groups post-arrest; 38-ECPR CO = 4.5 ± 1.0 l/min vs. 32-ECPR CO = 4.1 ± 0.9 l/min (T, p = 0.38).

Consistent with the reduced SV the strain decreased group-alike; from −17 ± 4 to −5 ± 3% (t, p < 0.001) in the 38-ECPR group and from −18 ± 3 to −7 ± 4% (t, p < 0.001) in the 32-ECPR group (T, p = 0.22). MAPSE in the 38-ECPR group was reduced from 12 ± 1 to 6 ± 1 mm (t, p < 0.001) and likewise in the 32-ECPR group from 12 ± 2 to 6 ± 1 mm (t, p < 0.001) (T, p = 0.98). Correspondingly, the wall thickening in the 38-ECPR group was severely affected by a reduction from 54 ± 11 to 18 ± 7% (t, p < 0.001). A similar reduction from 57 ± 14 to 13 ± 24% (t, p < 0.001) was found in the 32-ECPR group (T, p = 0.87).

cTnT and ASAT

The serum concentrations of cTnT and ASAT remained minimal after the surgical preparation (38-ECPR cTnT (median (range)) = 13 (10–32) ng/L vs. 32-ECPR cTnT = 16 (7–28) ng/L; 38-ECPR ASAT = 27 (18–33) U/L vs. 32-ECPR ASAT = 28 (18–31) U/L, but increased considerably post-arrest without significant differences between the groups; 38-ECPR cTnT = 2295 (782–5203) ng/L vs. 32-ECPR cTnT = 2452 (925–5475) ng/L (MW, p = 0.44) and 38-ECPR ASAT = 123 (61–1630) U/L vs. 32-ECPR ASAT = 266 (112–969) U/L (MW, p = 0.13).

TTC staining post mortem

The assessment of myocardial infarction by TTC staining post mortem did not demonstrate regional infarctions.

Discussion

In the present controlled animal study comparing hypothermic and normothermic E-CPR considering resuscitation success, post-arrest LV function, and myocardial injury, surprisingly, and contrary to our hypothesis, no beneficial effects of HT during E-CPR could be demonstrated.

A better preserved LV function would be desirable in resuscitated patients as post-arrest cardiac function is related to patient survival. In the present study, HT during E-CPR was hypothesized as being beneficial because HT has demonstrated cardioprotective effects in various animal studies with regional myocardial ischaemia (i.e. coronary occlusion) [21, 22].

The best strategy of E-CPR to preserve early post-arrest cardiac function is not known and no guidelines exist to assist clinicians deciding on an E-CPR strategy for patients in refractory cardiac arrest. To date HT is not recommended as a cardioprotective intervention in patients with acute myocardial infarction without associated cardiac arrest [22, 23]. In resuscitated patients, however, HT is an established treatment due to neurological benefits, irrespective of any cardioprotection [9–11]. Whole body cooling targeting 32–36 °C is the latest recommendation (preferably a constant temperature in this range) and a HT induction time-frame of 4–6 h post-arrest is usually accepted [24]. A delay of several hours
from resuscitation to target temperature may exclude a cardioprotective effect of hypothermia per se. HT may nevertheless be favorable to the cardiovascular function as it may reduce cardiac work load as a consequence of reduced whole body metabolism during HT [25].

Correct timing of HT has been emphasized in recent years as HT is claimed to be cardioprotective only if induced either shortly before or at the time point of myocardial reperfusion [12, 13, 26, 27]. Maeng and co-workers found that HT induced at the time of coronary reperfusion did not reduce myocardial infarct size in pigs [28]. Despite the efficient HT induction, the present study did not demonstrate cardioprotection by hypothermic E-CPR (i.e. reperfusion) of the fibrillating heart. Cooling the myocardium prior to myocardial reperfusion may thus be a crucial procedure, but effort is needed to achieve hypothermia this early in the clinical coronary occlusion scenario [29, 30]. Correspondingly, a rationale for cardioprotective intra-arrest HT (i.e. HT induced before ROSB) exists in cardiac arrested patients, and is supported by animal studies [13, 31, 32], but the suggested post-arrest cardiac function benefits are not specifically investigated in humans. If interventions to reperfuse the myocardium are postponed until HT is established, the harm of prolonged ischaemia may cancel any benefits of HT. This issue could be further investigated experimentally using topical cooling of the arrested heart prior to reperfusion.

Two-hour HT duration was investigated as brief cooling has been sufficient to achieve cardioprotective effects in previous animal studies [12] and 3-h cooling has made myocardial damage worse [33]. Whether a longer HT duration would be beneficial in our study is not known, but cannot be excluded. On the other hand, extracorporeal circulatory support by ECMO is not without complications and side effects, and clinical practice is to wean as soon as the heart is capable to independently handle the circulation.

Compared to our study, rewarming of patients after cardiac arrest is slow (0.3–0.5 °C/h), tailored for neuroprotection [24]. In cardiac surgery, however, a quick rewarming is well tolerated by the heart even after

### Table 1 Haemodynamic measurements and blood gas analyses

| Variable                  | Normothermic E-CPR group | Hypothermic E-CPR group | Post-arrest, difference between groups |
|---------------------------|--------------------------|-------------------------|---------------------------------------|
|                           | Baseline | Post-arrest | Baseline | Post-arrest | MD (95% CI) | p value |
| a. Haemodynamic measurements |          |            |          |            |              |          |
| HR (beats/min)            | 88 ± 23 | 152 ± 31*  | 101 ± 26 | 148 ± 33*  | −4 (−35.27) | p = 0.79 |
| MAP (mmHg)                | 842 ± 12.0 | 73.6 ± 16.1 | 93.1 ± 14.4 | 65.6 ± 14.1* | −81 (−22.6.65) | p = 0.26 |
| CVP (mmHg)                | 5.7 ± 2.8 | 6.1 ± 3.1  | 6.9 ± 1.6 | 8.7 ± 2.9  | 2.6 (−0.3.5) | p = 0.078 |
| LVP, max (mmHg)           | 99.6 ± 7.0 | 91.0 ± 13.5 | 107.7 ± 13.2 | 86.4 ± 14.8* | −46 (−18.9.2) | p = 0.49 |
| EDP (mmHg)                | 14.0 ± 4.7 | 14.9 ± 2.4  | 12.9 ± 3.7 | 11.7 ± 3.7 | −3.2 (−6.3) | p = 0.041 |
| dP/dt max (mmHg/s)        | 1427 ± 207 | 1963 ± 557 | 1786 ± 414 | 2398 ± 1466 | 435 (−663.1553) | p = 0.41 |
| dP/dt min (mmHg/s)        | −212 ± 270 | −1683 ± 404 | −2225 ± 315 | −1792 ± 832 | −109 (−755.537) | p = 0.73 |
| tau (ms)                  | 324 ± 3.0 | 33.3 ± 6.8 | 323.3 ± 3.2 | 285.8 ± 8.3 | 0.68 (5.7.71) | p = 0.19 |
| EDP/EDV (mmHg/ml)         | 0.14 ± 0.04 | 0.16 ± 0.03 | 0.14 ± 0.04 | 0.14 ± 0.04 | −0.02 (−0.06.01) | p = 0.15 |
| Ea (mmHg/ml)              | 1.08 ± 0.25 | 1.65 ± 0.44* | 1.16 ± 0.35 | 1.68 ± 0.72* | 0.03 (−0.56.62) | p = 0.91 |
| b. Blood gas analyses     |          |            |          |            |              |          |
| Hb (g/dl)                 | 8.3 ± 0.9 | 8.1 ± 0.7*  | 8.7 ± 0.8 | 7.8 ± 1.1*  | −0.3 (−1.3.06) | p = 0.45 |
| PaO2 (kPa)                | 22.5 ± 1.9 | 20.6 ± 3.7 | 22.6 ± 1.4 | 21.5 ± 2.4 | 0.90 (−2.1) | p = 0.93 |
| PaCO2 (kPa)               | 5.4 ± 0.7 | 5.4 ± 0.3  | 4.9 ± 0.4 | 4.9 ± 0.4  | −0.45 (−0.82) | p = 0.019 |
| pH                       | 7.49 ± 0.04 | 7.49 ± 0.03 | 7.51 ± 0.05 | 7.48 ± 0.05 | −0.01 (−0.03) | p = 0.63 |
| BaseExcess (mmol/l)       | 6.77 ± 1.15 | 4.97 ± 1.82* | 6.23 ± 2.38 | 5.36 ± 2.12* | −1.41 (−3.39) | p = 0.15 |
| Lactate (mmol/l)          | 0.85 ± 0.19 | 1.28 ± 0.54* | 1.34 ± 0.85 | 2.8 ± 1.57* | 1.52 (0.36.268) | p = 0.013 |
| Ca2+ (mmol/l)             | 1.30 ± 0.06 | 1.27 ± 0.06 | 1.25 ± 0.06 | 1.27 ± 0.06 | 0.003 (−0.06.06) | p = 0.91 |
| SvO2 (%)                  | 65.7 ± 14.8 | 42.4 ± 8.7* | 68.4 ± 7.4 | 42.3 ± 13.4* | −0.2 (−11.2) | p = 0.98 |

Values are expressed as mean ± standard deviation. Comparison post-arrest to baseline within group by paired student’s t-test. Post-arrest comparison of groups by unpaired two-sample student’s t-test. MD mean difference, CI 95% confidence interval

HR heart rate, MAP mean aortic blood pressure, CVP central venous pressure, LVP, max systolic left ventricular pressure maximum, EDP end-diastolic pressure, dP/dt max maximum left ventricular pressure first time derivate, dP/dt min minimum left ventricular pressure first time derivate, tau isotropic relaxation constant, Ea arterial elastance, SvO2 mixed venous oxygen saturation

*p ≤ 0.05
long-lasting cardioplegic arrest, and we have no indications that a longer and slower rewarming period would have influenced our results.

**Inotropes**

Inotropic support is regularly used during VA–ECMO to sustain aortic ejections with a sufficient pulse-pressure to avoid LV distention and failure. The increased dobutamine requirements observed during HT may be related to altered pharmacologic properties of dobutamine with a reduced effect [34] as supported by the similar requirements in the two groups after rewarming. HT also affects LV function, causing slower LV contraction and relaxation velocities [35], and may thus increase the need for inotropic stimulation to reach preset targets. The optimal MAP target during HT is not known, and could possibly differ from MAP target at normothermia, but for comparison, they were set at the same level.

**LV function**

In the present study, the 50–70% reductions in SV, EF, wall thickening, strain and MAPSE were consistent, demonstrating a severe global systolic LV dysfunction post-arrest with uniform impairments in all directions of systolic LV motion, and with no differences between the two treatment groups.

The diastolic LV function assessed by dP/dt<sub>min</sub>, tau and EDP/EDV relationship was preserved in both groups post-arrest, and neither EDV (preload) nor Ea (afterload) differed between the two groups.

**Myocardial injury evaluation**

The post-arrest LV dysfunction indicated a severe and global myocardial injury as was confirmed by the considerable increase in serum concentrations of cTnT and ASAT in both groups. A global injury without distinguished regional areas was also confirmed by the TTC assessment, as no regional infarctions could be demonstrated, excluding coronary thrombus or embolic complications.

The success rate of resuscitation by hypothermic E-CPR was not inferior to the normothermic strategy, and neither LV function nor myocardial injury was exacerbated. E-CPR initiated by a room tempered ECMO may be convenient, as a normothermic ECMO (or a circuitry heating device) will not always be available in clinical emergency settings that may include emergency rooms, ambulance transfers and even pre-hospital use.

**Limitations**

The clinical scenario of E-CPR differs from a controlled animal experiment as the period of no-flow is usually

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| Ejection fraction | Stroke volume | Cardiac output |
|-------------------|---------------|---------------|
|                   |               |               |

| Strain | Wall thickening | MAPSE |
|--------|----------------|-------|
|        |                |       |

![Fig. 4 Cardiac MRI measurements. The systolic left ventricle function variables did not differ between the normothermic and the hypothermic E-CPR group at baseline or post-arrest. Strain, peak global systolic left ventricular circumferential strain; Wall thickening, radial mid left ventricular wall thickening; MAPSE, mitral annular plane systolic excursion. Line at mean ± standard deviation (short line)](image-url)
short (preferably <5 min) and the patient is cannulated at ongoing CPR in a low-flow period of varying duration. In the present study, a healthy pig heart suffered an electrically induced VF and the total ischemic insult was prepared to be substantial and consistent to assure a significant post-arrest cardiac dysfunction and injury that could be compared between the two different treatment groups. The duration of no-flow thus exceeded usual clinical limits, and the low-flow period was bypassed.

Conclusions
E-CPR is an effective resuscitation technique for prolonged cardiac arrest that may rapidly induce HT. In the present animal study, 2-h HT during E-CPR offered an equal resuscitation success rate, but did not preserve the post-arrest cardiac function nor reduce the magnitude of myocardial injury, compared to normothermic E-CPR.

Additional file

Additional file 1: Table S1. Detailed information on the experimental animals according to ARRIVE* guidelines.

Abbreviations
ASAT: aspartate transaminase; CO: cardiac output; CPR: cardiopulmonary resuscitation; CVP: central venous pressure; dP/dt max and dP/dt min: maximum and minimum LVP first time derivatives; Ea: arterial elastance; ECMO: extracorporeal membrane oxygenation; E-CPR: extracorporeal cardiopulmonary resuscitation; EDP: end-diastolic pressure; EDV: end-diastolic volume; ESP: end-systolic pressure; HR: heart rate; LVP: left ventricular pressure; LVP max: systolic left ventricular pressure maximum; MAP: mean aortic pressure; MRI: magnetic resonance imaging; ROSB: regain of spontaneous cardiac beating; SV: stroke volume; Svo2: mixed venous oxygen saturation; tau: isovolumetric relaxation constant; cTnT: cardiac Troponin T.

Authors’ contributions
HAB: study design, data collection, data analyses, and manuscript preparation; PSH: study design, data collection, data analyses, and revision of manuscript; HS: study design, revision of manuscript; EF: study design and revision of manuscript; JFB: study design, data collection, data analyses, and manuscript revision. All authors read and approved the final manuscript.

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Competing interests
All authors are employed by Oslo University Hospital. The authors declare that they have no competing interests.

Availability of data and materials
Data supporting the findings are presented in the article.

Ethics approval
The experimental protocol was approved by the Norwegian National Animal Research Authority and the animal experiments were performed in accordance with the European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes (European Council, ETS No. 170).

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References
1. Laver S, Farrow C, Turner D, Nolan J. Mode of death after admission to an intensive care unit following cardiac arrest. Intensiv Care Med. 2004;30(1):2126–8.
2. Laurent I, Monchi M, Chiche JD, Joly LM, Spaulding C, Bourgeois B, Caricou A, Rozenberg A, Carli P, Weber S, et al. Reversible myocardial dysfunction in survivors of out-of-hospital cardiac arrest. J Am Coll Cardiol. 2002;40(12):2110–6.
3. Chen YS, Lin JW, Yu HY, Ko WJ, Jeng JS, Chang WT, Chen JW, Huang SC, Chi NH, Wang CH, et al. Cardiopulmonary resuscitation with assisted extracorporeal life-support versus conventional cardiopulmonary resuscitation in adults with in-hospital cardiac arrest: an observational study and propensity analysis. Lancet. 2008;372(9638):554–61.
4. Haneya A, Philipp A, Diez C, Schopka C, Bein T, Zimmermann M, Lubnow M, Luchner A, Agha A, Hilker M, et al. A 5-year experience with cardiopulmonary resuscitation using extracorporeal life support in non-postcardiomyopathy patients with cardiac arrest. Resuscitation. 2012;83(11):1331–7.
5. Shin TG, Choi JH, Jo JJ, Sim MS, Song HG, Jeong WT, Chen JW, Huang SC, Cha NH, Wang CH, et al. Extracorporeal cardiopulmonary resuscitation in patients with inhospital cardiac arrest: a comparison with conventional cardiopulmonary resuscitation. Crit Care Med. 2011;39(1):1–7.
6. Lasa JJ, Rogers RS, Localio R, Shults J, Raymond T, Gaies M, Thiagarajan R, Laussen PC, Kilbaugh T, Berg RA et al. Extracorporeal-cardiopulmonary resuscitation (E-CPR) During pediatric in-hospital cardiopulmonary arrest is associated with improved survival to discharge: A report from the American heart association’s get with the guidelines(R)—resuscitation registry (GW1TG-R). Circulation. 2015.
7. Tissier R, Chenoune M, Ghaebi B, Cohen MV, Downey JM, Berdeaux A. The small chill: mild hypothermia for cardioprotection? Cardiovasc Res. 2010;88(3):406–14.
8. Zobel C, Adler C, Kranz A, Seck C, Pfister R, Hellmich M, Kochanek M, Reuter H. Mild therapeutic hypothermia in cardiogenic shock syndrome. Critical Care Med. 2012;40(6):1715–23.
9. Bernard SA, Gray JW, Buist MD, Jones BM, Silvester W, Gutteridge G, Smith K. Treatment of comatose survivors of out-of-hospital cardiac arrest with induced hypothermia. N Engl J Med. 2002;346(9):557–63.
10. Hypothermia after Cardiac Arrest Study Group. Mild therapeutic hypothermia to improve the neurologic outcome after cardiac arrest. N Engl J Med. 2002;346(24):495–506.
11. Nagao K. Therapeutic hypothermia following resuscitation. Curr Opin Crit Care. 2012;18(3):239–45.
12. Nozari A, Safar P, Siezoski SW, Wu X, Kostelnik S, Radosvky A, Fisherman S, Kochanek PM. Critical time window for intra-arrest cooling with cold saline flush in a dog model of cardiopulmonary resuscitation. Circulation. 2006;113(23):2690–6.
13. Zhao D, Abella BS, Beiser DG, Alvarado JP, Wang H, Hamann KJ, Hoek TL, Becker LB. Intra-arrest cooling with delayed reperfusion yields higher survival than earlier normothermic resuscitation in a mouse model of cardiac arrest. Resuscitation. 2008;77(2):242–9.
14. Abella BS, Zhao D, Alvarado J, Hamann K, Vanden Hoek TL, Becker LB. Intra-arrest cooling improves outcomes in a murine cardiac arrest model. Circulation. 2004;109(22):2786–91.
15. Bergan HA, Halvorsen PS, Skulstad H, Edvardsen T, Fosse E, Bugge JF. Successful ECMO-cardiopulmonary resuscitation with the associated post-arrest cardiac dysfunction as demonstrated by MRI. Intensiv Care Med Exp. 2015;3(1):61.

16. Kilkenney C, Browne WJ, Cuthill IC, Emerson M, Altman DG. Improving bioscience research reporting: the ARRIVE guidelines for reporting animal research. PLoS Biol. 2010;8(6):e1000412.

17. Vasquez A, Kern KB, Hilliwg RW, Heidenreich J, Berg RA, Ewy GA. Optimal dosing of dopamine for treating post-resuscitation left ventricular dysfunction. Resuscitation. 2004;61(2):199–207.

18. Schütz-Menger J, Bluemke DA, Bremerich J, Flamm SD, Fogel MA, Friedrich MG, Kim RJ, von Knobelsdorff-Brenkenhoff F, Kramer CM, Pennell DJ, et al. Standardized image interpretation and post processing in cardiovascular magnetic resonance: Society for cardiovascular magnetic resonance (SCMR) board of trustees task force on standardized post process. J Cardiovasc Magn Reson. 2013;15:35.

19. Klein HH, Puschmann S, Schaper J, Schaper W. The mechanism of the tetrazolium reaction in identifying experimental myocardial infarction. Virchows Arch. 1981;393(3):287–97.

20. Fishbein MC, Meeraum S, Rit J, Lando U, Kanamatake K, Mercier JC, Corday E, Ganz W. Early phase acute myocardial infarct size quantification: validation of the triphenyl tetrazolium chloride tissue enzyme staining technique. Am Heart J. 1981;101(5):593–600.

21. Chien GL, Wolff RA, Davis RF, van Winkle DM. “Normothermic range” temperature affects myocardial infarct size. Cardiovasc Res. 1994;28(7):1014–7.

22. Delhaye C, Mahmoudi M, Waksman R. Hypothermia therapy: neurological and cardiac benefits. J Am Coll Cardiol. 2012;59(3):197–210.

23. Steg PG, James SK, Atar D, Badano LP, Blomstrom-Lundqvist C, Borger MA, Chen SL, Dickstein K, Fernandez-Aviles F, et al. ESC guidelines for the management of acute myocardial infarction in patients presenting with ST-segment elevation. Euro Heart J. 2012;33(20):2569–619.

24. Nolan JP, Soar J, Cariou A, Cronberg T, Moulaert VRM, Deakin CD, Böttiger BW, Friberg H, Sunde K, Sandroni C. European resuscitation council and European society of intensive care medicine guidelines for post-resuscitation care 2015: section 5 of the European resuscitation council guidelines for resuscitation 2015. Resuscitation. 2015;95:202–22.

25. Schmidt-Schweda S, Ohler A, Post H, Pieske B. Moderate hypothermia for severe cardiogenic shock (COOL Shock Study I & II). Resuscitation. 2013;84(3):319–25.

26. Tissier R, Ghaleh B, Cohen MV, Downey JM, Berdeaux A. Myocardial protection with mild hypothermia. Cardiovasc Res. 2012;94(2):217–25.

27. Gottberg M, van der Pals J, Gottberg M, Olivecrona GK, Kanski M, Koul S, Otto A, Engblom H, Ugander M, Arshedh E, et al. Optimal timing of hypothermia in relation to myocardial reperfusion. Basic Res Cardiol. 2011;106(5):697–708.

28. Maeng M, Mortensen UM, Kristensen J, Kristiansen SB, Andersen HR. Hypothermia during reperfusion does not reduce myocardial infarct size in pigs. Basic Res Cardiol. 2006;101(1):61–8.

29. Erlinge D, Gottberg M, Lang I, Holzer M, Noc M, Clemmensen P, Jensen U, Metzler B, James S, Botker HE, et al. Rapid endovascular catheter core cooling combined with cold saline as an adjunct to percutaneous coronary intervention for the treatment of acute myocardial infarction. The CHILL-MI trial: a randomized controlled study of the use of central venous catheter core cooling combined with cold saline as an adjunct to percutaneous coronary intervention for the treatment of acute myocardial infarction. J Am Coll Cardiol. 2014;63(18):1857–65.

30. Dae MW, Gao DW, Seissler DI, Chair K, Stillson CA. Effect of endovascular cooling on myocardial temperature, infarct size, and cardiac output in human-sized pigs. Am J Physiol Heart Physiol. 2002;282(5):H1584–91.

31. Tsai MS, Barbut D, Wang H, Guan J, Sun S, Inderbitzen B, Well MH, Tang W. Intra-arrest rapid head cooling improves postresuscitation myocardial function in comparison with delayed postresuscitation surface cooling. Crit Care Med. 2008;36(1 Suppl):S434–9.

32. Yannopoulos D, Ziviman M, Castro V, Kolandaivelu A, Ranjan R, Wilson RF, Halperin HR. Intra-cardiopulmonary resuscitation hypothermia with and without volume loading in an ischemic model of cardiac arrest. Circulation. 2009;120(14):1426–35.

33. Leonov V, Sterz F, Safar P, Radovsky A. Moderate hypothermia after cardiac arrest of 17 minutes in dogs. Effect on cerebral and cardiac outcome. Stroke. 1990;21(11):1600–6.

34. Erlinge D, Sjögren P, Scholte CS, Grottke O, Hein M, Ackermann D, Rossaint R, Schalte G. Influence of temperature on the positive inotropic effect of levosimendan, dobutamine and milrinone. Euro J Anaesthesiol. 2009;26(11):946–53.

35. Kerans V, Espinoza A, Skulstad H, Halvorsen PS, Edvardsen T, Bugge JF. Systolic left ventricular function is preserved during therapeutic hypothermia, also during increases in heart rate with impaired diastolic filling. Intensiv Care Med Exp. 2015;3(1):41.