Review

Fucoidan and Cancer: A Multifunctional Molecule with Anti-Tumor Potential

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Abstract: There is a wide variety of cancer types yet, all share some common cellular and molecular behaviors. Most of the chemotherapeutic agents used in cancer treatment are designed to target common deregulated mechanisms within cancer cells. Many healthy tissues are also affected by the cytotoxic effects of these chemical agents. Fucoidan, a natural component of brown seaweed, has anti-cancer activity against various cancer types by targeting key apoptotic molecules. It also has beneficial effects as it can protect against toxicity associated with chemotherapeutic agents and radiation. Thus the synergistic effect of fucoidan with current anti-cancer agents is of considerable interest. This review discusses the mechanisms by which fucoidan retards tumor development, eradicates tumor cells and synergizes with anti-cancer chemotherapeutic agents. Challenges to the development of fucoidan as an anti-cancer agent will also be discussed.

Keywords: fucoidan; cancer; apoptosis; synergy
1. Introduction to Cancer

Cancers are multifactorial diseases of various etiologies. They arise largely as a result of acquired genetic changes that alter cell function leading neoplastic cells to gain survival or growth advantages [1]. For cancer cells to survive, the generation of new blood vessels (angiogenesis) is required. Cancer leads to death mostly through tumor cell spread to distal organs (metastasis). Various pathways are disrupted in tumor development, which result from unbalanced programmed cell death, disordered signaling pathways, angiogenesis and poor immune response against cancer. Most of the chemotherapeutic agents used in cancer treatment target these major deregulated pathways. Unfortunately, as many of these therapies cause severe side effects, the toxicities limit the dose and thus the efficacy of treatment. Therefore, there is strong interest in developing better-tolerated anti-cancer agents.

2. A Role for Natural Products for Cancer Treatment

Chemotherapy has been a cornerstone of the standard cancer treatment regimens since the 1960s. A variety of chemicals ranging from traditional agents such as methotrexate and folic acid analogues to novel chemicals such as anthracyclines have been used in cancer treatment [2]. Despite promising tumor growth-inhibitory effects in pre-clinical tests, many fail in clinical trials when adverse unexpected side effects are revealed. Traditionally anti-cancer chemotherapy targets rapidly dividing and proliferating cells. Therefore, normal cells which have high-proliferating potential are also affected.

Novel therapeutic agents are designed to target specific molecules (targeted therapy). However, these targeted therapies are not always completely free of side effects either. For instance, vemurafenib, a B-Raf enzyme inhibitor, is specific for oncogenic mutant V600E B-Raf positive melanoma cells. This drug was the first targeted molecular therapy, which was approved for use in advanced stages of melanoma. Although vemurafenib has shown significant beneficial anti-cancer effects, several studies have reported the rapid emergence of acquired resistance and adverse dermatological effects. It also stimulates B-Raf expression in V600E B-Raf negative patients promoting melanoma growth [3,4]. Monoclonal antibodies are another example of targeted therapy and are designed to specifically target the cancer antigens located on tumor cells. Monoclonal antibodies are generally safer than chemotherapy and the side effects caused by them include mild allergic reactions such as urticaria. But they can also cause severe reactions such as infusion reactions and serum sickness. As an example, rituximab (anti-CD20), which is widely used in treating B-cell lymphoma, generally causes only mild toxicities, however, reports have described occasional cases with severe complications such as anaphylactic reactions and myocardial infarction as well as high risk of tumor lysis syndrome in patients who have a high burden of tumor cells in their circulation [5].

Concerns over toxicity, tumor cell resistance and development of secondary cancers from chemotherapeutic chemicals have generated interest in exploiting natural products for cancer treatment. Flavopiridol is a flavonoid derived from the indigenous Indian plant *Dysosyllum binectariferum*, which inhibits cell cycle progression. It is the first cyclin-dependent kinase (CDK) inhibitor to be approved for use in clinical trials [6]. Natural products are also being tested as adjuvants for use in synergy with chemotherapeutic agents. For example those with immunomodulatory effects can reduce immune
suppression and the associated increased risk of infection. In George et al. [7] study, Indukantha Ghritha (IG), a polyherbal preparation consisting of 17 plant components, was used as an adjuvant to cyclophosphamide cancer chemotherapy and shown to stimulate the hematopoietic system and induce leukopoiesis in tumor-bearing mice. When administrated in combination with cyclophosphamide, it reversed myelosuppression induced by cyclophosphamide suggesting its potential to minimize or reverse chemotherapy-induced leukopenia.

Polysaccharides include a large family of diverse biopolymers. They are constituted by monosaccharide residues linked together by O-glycosidic bonds that are found in natural and semi-synthetic structures [8]. Due to structural diversity, polysaccharides display the highest biological properties among macromolecules. Many natural polysaccharides obtained from natural sources such as plants and algae have anti-cancer properties. The multifunctional structure of natural polysaccharides also allows them to be used in conjugation with anti-cancer agents that lack physiochemical and biopharmaceutical properties [8,9].

3. Fucoidan

Fucoidan is a natural sulfated polysaccharide that exists mainly in the cell wall matrix of various species of brown seaweed such as mozuku, kombu, limumouci, bladderwrack and wakame [10]. Various forms of fucoidan have also been recognized in some marine invertebrates such as sea urchins [11] and sea cucumbers [12]. The brown seaweeds containing fucoidan are widely consumed as part of the normal diet in East Asia, particularly Japan, China and Korea.

3.1. Fucoidan’s Anti-Cancer Potential

The anti-cancer property of fucoidan has been demonstrated in vivo and in vitro in different types of cancers. Nevertheless, it has been rarely investigated for its anti-cancer properties in clinical trials. Fucoidan mediates its activity through various mechanisms such as induction of cell cycle arrest, apoptosis and immune system activation. Additional activities of fucoidan have been reported that may be linked to the observed anti-cancer properties and these include induction of inflammation through immune system, oxidative stress and stem cell mobilization. These activities have been reviewed by Kwak [13].

3.1.1. Fucoidan and Cell Cycle

Fucoidan treatment results in sub G0/G1 cell accumulation (suggestive of dead cells/apoptotic cells) in a variety of cell types [14,15]. It can also induce cell cycle arrest in other phases; Riou et al. [16] and Mourea et al. [17] reported arrest in G1 phase in a chemo-resistant non-small-cell bronchopulmonary carcinoma line by fucoidan from Ascophyllum nodosum and Bifurcaria bifurcate, respectively.

In an investigation of the mechanism of the action, fucoidan demonstrated significant down regulation of cyclin D1, cyclin D2 and CDK4 in cancer cells [18–20]. The crude fucoidan from Fucus vesiculosus increased the level of p21/WAF1/CIP1 in PC3 cells and down-regulated E2F; a transcription factor that controls progression of cells from G1 to S phase [18].
Table 1. Effects of fucoidan on cell cycle and apoptosis molecules.

| Ref   | Cell Type                  | Fucoidan Source | Dose (µg/mL) | Effects on Cell Cycle | Effects on Apoptosis Pathways | Extrinsic | Intrinsic | Common |
|-------|----------------------------|-----------------|--------------|-----------------------|-------------------------------|-----------|-----------|---------|
| [15]  | Human lymphoma HS-sultan cells | *F. vesiculosus* | 100          | ↑ sub G0/G1           | ↓ MMP                         |           |           |         |
|       |                            |                 |              | No G0/G1 or G2/M arrest |                               |           |           |         |
| [20]  | HTLV-1 infected T-cell HUT-102- cells | *C. okamurans* | 3000         | G1 arrest             | Apoptosis was reversed by caspase 8 inhibitor | Caspase 9 activation |           |         |
|       |                            |                 |              | ↓ cyclin D2, c-myc    |                               | No changes in Bcl-2 and Bcl-XL |           |         |
|       |                            |                 |              | No changes in p21,p53 |                               | ↓ survivin, cIAP-2            |           |         |
| [21]  | Human hepatocellular carcinoma cells | *Okinawa mozuku* | 22.5         | ↑ G2/M phase in HAK-1A, KYN-2, KYN-3 cell lines | -                             | No clear caspase 9 activation in HAK-1B cell line |           |         |
| [22]  | Human breast cancer MCF7 cells | Not mentioned   | 1000         | ↑ sub-G1 fraction     | Caspase 8 activation          | Caspase 9 activation |           |         |
|       |                            |                 |              | Caspase inhibitors blocked apoptosis completely | ↓ Bid, cytosolic Bax          | No changes in Bcl-2 or Bax |          | Caspase 3 activation |
| [23]  | Human acute leukemia NB4 and HL-60 cells | *F. vesiculosus* | 150          | ↑ sub-G1 fraction     | Caspase 8 activation          | Caspase 9 activation |           |         |
|       |                            |                 |              | Caspase inhibitors blocked apoptosis completely | ↓ Mcl-1, ↑ cytochrome C       | No changes in Bcl-2 or Bax |          | PARP cleavage |
| [24]  | Human colon cancer HT-29 and HCT116 cells | *F. vesiculosus* | -            | ↑ sub-G1 fraction     | Caspase 8 activation          | Caspase 9 activation |           |         |
|       |                            |                 |              | Caspase inhibitors blocked apoptosis completely | ↑ Fas, DR5, TRAIL            | ↑ cytochrome C, Smac/Diablo, Bak, t-Bid |          | PARP cleavage |
|       |                            |                 |              | No significant effects on FasL and DR4 | No changes in Bcl-2, Bel-xL, Bax, Bad, Bin, Bik | No changes in Bcl-2, Bel-xL, Bax, Bad, Bin, Bik | Caspase 3 and 7 activation |
| [25]  | Human lung cancer AS49 cells | *U. pinnatifida* | 50, 100, 200 | ↑ Sub-G1 fraction     | Caspase-9 activation          | Caspase-9 activation |           | ↓ procaspase-3 |
|       |                            |                 |              | ↓ Bel-2, ↑ Bax        |                               | ↓ Bel-2, ↑ Bax |          |         |
| [14]  | Human breast cancer MCF-7 cells | *Cladosiphon novae-caledoniae* | 82, 410, 820 | ↑ Sub-G1             | No changes in caspase-8      | Mitochondrial dysfunction | AIF and cytochrome C release | No activation of PARP and caspase-7 |
|       |                            |                 |              | No significant changes in cell cycle distribution | No cleavage of caspase-9 and Bid. | ↑ Bel-2, Bel-xL, ↑ Bax, Bad | All caspase inhibitors failed to attenuate FE-induced apoptosis |
| [26]  | Hela cells | *Sargassum filipendula* | 1500 | - | No effect on caspase 9 activation | ↑ cytool AIF | No effect on caspase 3 (Caspase independent) |
| [19]  | Human breast cancer MCF-7 cells | *F. vesiculosus* | 400, 800, 1000 | G1 phase arrest | Caspase-8 activation | ↓ Bel-2 | Caspase-dependent pathway |
|       |                            |                 |              | ↑ Sub G0/G1 ↓ cyclin D1 and CDK-4 gene expression | ↑ Bax | No effect on caspase 3 |
|       |                            |                 |              | Caspase-8 activation | Release of cytochrome C and APAF-1 | Caspase-8 activation |          |         |
|       |                            |                 |              | ↑ Bax | No effect on caspase 3 |
|       |                            |                 |              | Caspase-dependent pathway | | Caspase-dependent pathway |          |         |
| Ref  | Cell Type                      | Fucoidan Source | Dose (µg/mL) | Effects on Cell Cycle                                      | Effects on Apoptosis Pathways Extrinsic Intrinsic Common |
|------|--------------------------------|-----------------|--------------|-----------------------------------------------------------|----------------------------------------------------------|
| [18] | Human prostate cancer          | U. pinnatifida   | 100          | • G0/G1 phase arrest                                      | • ↓ Bcl-2                                                |
|      | PC-3 cells                     |                 |              | • ↓ E2F-1                                                | • ↑ Bax,                                                  |
|      |                                |                 |              | • ↑ p21Cip1/Waf                                           | • Caspase 9 activation                                   |
|      |                                |                 |              |                                                           | • Caspase-3 activation                                   |
| [27] | Human Hepatocellular Carcinoma | U. pinnatifida   | 1000         | • Non-significant accumulation in S-phase                | • Caspase-9 activation                                   |
|      | SMMC-7721 cells                |                 |              | • Caspase-8 activation                                    | • MMP dissipation, Cytochrome C release                  |
|      |                                |                 |              |                                                           | • ↓ Bcl-2, ↑ Bax                                         |
|      |                                |                 |              |                                                           | • ↓ XIAP, livin mRNA expression                          |
| [28] | Human bladder                  |                 | 100          | • ↑ G1-phase, p21WAF1                                     | • Caspase-3 activation                                   |
|      | carcinoma 5637 and T-24 cells  | F. vesiculosus   |              | • ↓ Cyclin E, D1, DK2, CDK4                              | • Caspase-3 activation                                   |
|      |                                |                 |              | • No change in p27KIP,p53                                  | • Caspase-3 activation                                   |
|      |                                |                 |              | • ↑ p21WAF1 and CDK4 binding                              | • Caspase-3 activation                                   |
In a recent study, fucoidan down-regulated cyclin E, CDK2, CDK4 resulting in G0/G1 arrest in human bladder cancer 5637 cells. Furthermore, immunoprecipitation assays revealed a significant increase in the binding of p21/WAF1/CIP1 to CDK2 and CDK4 in cells treated with fucoidan, suggesting that the induced G0/G1 arrest is due to suppression of CDK activity following direct binding of this CDK inhibitor to CDKs 2 and 4 [28]. Table 1 summarizes findings of studies examining the effects of fucoidan on cell cycle.

3.1.2. Fucoidan and the Apoptosis Pathway

Apoptosis characterized by cytoplasmic shrinkage and chromatin condensation facilitates the removal of cells without inducing inflammation [29]. Apoptosis occurs through either the extrinsic (cytoplasmic) pathway whereby death receptors trigger the apoptosis, or the intrinsic (mitochondrial) pathway in which changes in mitochondrial membrane potential (MMP) lead to cytochrome C release and death signal activation. Both pathways activate executive caspases that cleave regulatory and structural molecules [30]. Several studies examining a variety of cancers such as hematopoietic, lung, breast and colon cancers have shown that fucoidan-mediated cell death occurs through triggering apoptosis (Table 1) [14,22,24]. A very low dose of fucoidan from F. vesiculosus (20 µg/mL) activated common caspases 3 and 7 in human colon cancer cells [24], whereas it induced the same activity in T-cell leukemia at a much higher concentration (3 mg/mL) [20]. Caspase 8 and 9, two of the best characterized molecules of the extrinsic and intrinsic pathways respectively are activated by fucoidan [24]. Yamasaki-Miyamoto et al. showed that pre-treatment with caspase 8 inhibitor completely blocked fucoidan mediated apoptosis in MCF-7 breast cancer cell line [22]. In contrast, in Zhang et al. [14] study, the mediated apoptosis by fucoidan from Cladosiphon okamuranus in MCF-7 human breast cancer cell line was shown to be caspase independent. As cytochrome C and apoptosis inducing factor (AIF) increased in the cytosol, it was concluded that fucoidan performed its activity through mechanisms altering mitochondrial function.

Fucoidan also affects other components of extrinsic and intrinsic pathways. Analyzing the extrinsic pathway, 20 µg/mL crude fucoidan from F. vesiculosus increased the levels of the death receptors Fas, DR5 and TRAIL but not FasL and DR4 in human colon cancer cell lines [24]. Bcl-2 family members include anti-apoptotic, pro-apoptotic and regulatory proteins, which are mainly involved in the apoptosis intrinsic pathway. Contradictory results have been described in the expression of these regulatory molecules in response to fucoidan (Table 1). Treatment of MDA-MB231 breast cancer cells with 820 µg/mL of low molecular weight (LMW) fucoidan resulted in a significant decrease in anti-apoptotic proteins Bcl-2, Bcl-xl and Mcl-1 [31]. In contrast, no changes in expression of Bcl-2, Bcl-xl, Bad, Bim and Bik were observed in colon cancer cells when they were treated with 20 µg/mL fucoidan from Fucus vesiculosus [24]. Taken together, the results suggest that fucoidan may interact with several components of the apoptosis pathway.

3.1.3. Fucoidan and Angiogenesis

Fucoidan inhibits the formation of new vessels by which tumor cells receive their oxygen and required nutrients. Fucoidan has been found to inhibit the binding of VEGF, a key angiogenesis promoting molecule, to its cell membrane receptor [32]. Xue et al. examined the anti-angiogenic properties of
fucoidan in 4T1 mouse breast cancer cells both in vitro and in vivo and observed a significant
dose-dependent decrease in VEGF expression in cells treated with fucoidan. Further, in a mouse breast
cancer model using 4T1 cells, intraperitoneal injections of 10 mg/kg body weight fucoidan from
F. vesiculosus for 20 days markedly reduced the number of microvessels. Using immunohistochemistry,
fucoidan was shown to reduce VEGF expression compared to the control group [33]. In contrast, Zhu
et al. reported that fucoidan did not suppress angiogenesis and VEGF expression in human
hepatocarcinoma cell lines treated with 10 to 200 µg/mL of a commercial fucoidan purified from
Sargassum spp. Similarly no changes in VEGF expression were observed in xenograft tumors developed
in nude mice following 20 to 200 mg/kg/body weight fucoidan injected intraperitoneally once a day over
25 days [34]. It is postulated that different effects are observed with fucoidans of various MWs and
molecular structures and this is reviewed by Kwak [13].

3.1.4. Fucoidan and Metastasis

In 1987, Coombe et al. demonstrated that fucoidan significantly decreased tumor cells metastasis to
the lungs in animals that were intravenously injected with rat mammary adenocarcinoma 13762 MAT
cells [35]. It was first reported that fucoidan inhibits cell invasion through competing with tumor cell
binding with laminin in the basement membrane [36]. Subsequent studies then revealed that fucoidan
binds to fibronectin with high affinity and prevent attachment of tumor cells. In agreement with this
study, fucoidan reduced the spread of human breast adenocarcinoma cells plated on a surface containing
fibronectin [37].

Selectin inhibition by fucoidan interferes with tumor cell–platelet interaction. In Cumashi et al.
study [38], highly metastatic MDA-MB-231 breast cancer cells were plated in platelet-coated plates in
the presence or absence of 100 µg/mL fucoidan. The number of cells attached to the platelets
decreased by 80% in the presence of fucoidan. Interaction of tumor cells with platelets is one of the
key factors in facilitating the early steps of tumor cell migration. During tumor cell migration, most
circulating tumor cells do not survive attack from immune cells or the shear forces of the blood stream.
However, they can attach to platelets to induce platelet aggregation allowing the tumor cell cluster to
survive in the micro-vascular system. It was concluded that fucoidan inhibited P-selectin residing on
the platelet surface and led to reduced number of attached tumor cells. Fucoidan can also inhibit other
adhesion molecules such as integrins residing on the tumor cell surface and can modify distribution of
their subunits.

Tumor invasion requires the secretion of proteolytic enzymes by tumor cells to break down
the extracellular matrix (ECM) proteins (e.g., collagen, fibronectin and laminin), with the matrix
metalloproteinases (MMPs) MMP-2 and MMP-9 playing a major role. Fucoidan attenuates both
expression and activity of these enzymes [39].

3.1.5. Fucoidan and Signaling Pathways

The extracellular signal-regulated kinase (ERK) pathway (or Ras/Raf/MAPK pathway) is often
hyperphosphorylated and upregulated in a variety of human cancers. The potential for developing
anticancer agents that cause ERK’s dephosphorylation and pathway blockade have been explored.
Various studies have shown that fucoidan inhibits tumor cell proliferation by decreasing ERKs activity
through reduction of its phosphorylation [15,40] while several studies have proposed that fucoidan causes ERK activation rather than inactivation [41,42]. To explain these contradictions, it should be noted that the ERK signaling pathway is highly complex. It induces a range of different responses including cell proliferation, differentiation, migration and apoptosis depending on cell type, the type of stimulus and duration of activation [43]. Therefore, some of the contradictory results of the aforementioned studies can be explained by different fucoidan extracts with different molecular structures being used on different tumor cell types. Another complication is that different studies have examined ERK phosphorylation over different time periods ranging from 10 min to 48 h. Jin et al. reported increased ERK1/2 phosphorylation in HL-60 leukemic cell line 10–15 min after fucoidan treatment. The phosphorylation returned to the basal level after 1 h [23]. In Lee et al. study, crude fucoidan progressively diminished phosphorylation of ERK1/2 from 1 h to 9 h after treatment [39].

JNK and p38 are other MAPK superfamily members whose activity is altered by fucoidan. Fucoidan induced cell death in breast cancer cells through phosphorylation and activation of JNK and p38 after 30 min. The fucoidan-induced apoptosis significantly annulled in the presence of JNK inhibitor, indicating critical role of JNK in fucoidan-mediated apoptosis [14].

![Figure 1. Overview of main signal transduction pathways involved in cell proliferation and apoptosis.](image)

Similarly, the PI3K/AKT, GSK and Wnt pathways have been shown to be triggered by fucoidan. PI3K/AKT pathway generally inhibits apoptosis. AKT over-activation is also associated with drug resistance and tumor cell survival. As a result, deactivating this pathway could be another potential target for anti-cancer drug development. Most of the studies have reported inactivation of AKT by
fucoidan. PI3k, an upstream molecule of AKT, is also inhibited by fucoidan [39]. Upregulation of the Wnt signaling pathway is believed to have a critical role in prostate cancer development, survival and progression. Fucoidan from F. vesiculosus activated GSK-3β in PC3 human prostate cancer cells resulting in hypo-phosphorylation and inactivation of β-catenin, a critical component of the Wnt pathway (Figure 1) [18]. Figure 1 represents an overview of the mentioned signaling pathways.

3.1.6. Fucoidan and the Immune System

The effects of fucoidan on molecules of the immune system have been studied both in vitro and in vivo and effects on both cellular and humoral elements have been described. Fucoidan increases both activity and number of natural killer (NK) cells in vivo [44,45]. Increase in the number of cytotoxic T-cells (CTLs) has also been reported. A high-molecular-weight (HMW) fucoidan from Cladosiphon okamuranus (200–300 kDa) induced a large increase in the proportion of murine cytotoxic T cells [46]. Investigation of the role of fucoidan on dendritic cell (DC)-mediated T-cell cytotoxicity has revealed that the stimulation of CTLs was more effective in fucoidan-treated DCs as CTLs co-cultured with fucoidan-treated DCs exerted a high level of specific lysis of breast cancer cells [47].

In a recent study, the role of fucoidan in DCs function and its adjuvant effect have been examined in vivo. Fucoidan was systemically administrated to mice by intraperitoneal injection. Examination of the spleen DCs revealed up-regulation of maturation markers as well as production of IL-6, IL-12 and TNF-α. Fucoidan was then used as an adjuvant in vivo with ovalbumin antigen and induced Th1 mediated immune response and CTL activation [48].

3.1.7. Fucoidan and Malignant Transformation in Vitro and in Vivo

Few studies have reported the potential of fucoidan to inhibit neoplastic transformation. Teas et al. fed rats with dietary seaweed (Laminira) for 55 days and administrated the carcinogen 7,12-dimethylbenz(a) anthracene intragastrically. Following 26 weeks monitoring, experimental rats showed a significant delay in the median time for tumor appearance (19 vs. 11 weeks in the control group) [49].

Transforming growth factor β1 (TGFβ1) is believed to promote tumor development and metastasis through epithelial to mesenchymal transition (EMT), a process that enables epithelial cells migrate to distant areas during late stages of breast cancer development [50]. To trigger tumor progression, TGFβ1 recruits TGF receptors (TGFR) residing on the cell surface. The investigations of effects of fucoidan on TGFβ1-promoted carcinogenesis in MDA-MB-231 breast cancer cells have indicated that fucoidan decreased the expression of TGFRs and affected the downstream signaling molecules, which are involved in TGFβ1-mediated EMT [41].

Epidermal growth factor (EGF) is another carcinogenesis promoter, which induces tumor transformation through overexpression and activation of EGF receptor (EGFR). EGFR has a key role in cell proliferation and differentiation and many carcinomas arise from its mutations [51]. Lee et al. examined the role of fucoidan on the activation of EGFR and EGF-mediated neoplastic transformation [52]. They utilized murine JB6 Cl41 epidermal cells and induced cell transformation by EGF in the presence of fucoidan from L. guryanovae. Fucoidan markedly reduced the EGFR activation through hypo-phosphorylation. It also inhibited EGF-tumorigenic activity through inhibition of AP-1, a transcription factor responsible for cell proliferation regulation.
3.2. Fucoidan Metabolism

Fucoidanase, the enzyme responsible for fucoidan hydrolysis, has only been found in brown seaweed and marine microorganisms such as some marine bacteria and fungi [53] and not in humans. It is possible that the acidic conditions in the stomach could degrade fucoidan, but it has been reported that the low gastric pH does have restricted effects on fucoidan [54]. Small amounts of dietary fucoidan can be endocytosed and cross the intestinal wall directly without breaking down [54]. In Tokita et al. study, 10 volunteers were given oral fucoidan and the concentrations of fucoidan in the serum and urine were analyzed. Fucoidan was detectable 3 h after administration and increased to 100 ng/mL in serum and 1000 ng/mL in urine. However the rate of absorption in the small intestine was highly variable among the participants. The MW of fucoidan in serum was similar to administered fucoidan indicating that fucoidan was not hydrolyzed by digestive enzymes [55]. However, the MW of the fucoidan detected in urine was significantly smaller than the ingested fucoidan suggesting that fucoidan is degraded in the excretory system and possibly the kidney and not by intestinal enzymes or normal flora.

To evaluate the fucoidan uptake process by cells, the internalization of LMW fucoidan into rabbit smooth muscle cells (SMCs) was analyzed. Fucoidan was shown to be internalized by endocytosis at 6 h. The number of vesicles containing fucoidan increased in the peri-nuclear region at 24 h, but nuclear internalization was not observed at any time during the study [56]. However, examining the transport of a native fucoidan from Cladosiphon okamuranus with MW of 80 kDa revealed a poor permeation of fucoidan across the human colon adenocarcinoma Caco-2 cell monolayer [57].

Regarding the specific ligands by which fucoidan binds to the cells surface, several molecules have been implicated including class A macrophage scavenger receptors for fucoidan attachment to macrophages [58] as well as adhesion molecules such as L-selectin and P-selectin [59] and integrins [60]. However, some reports have shown fucoidan mediates apoptosis through selectin-independent mechanisms [15].

3.3. Fucoidan as a Synergistic Anti-Cancer Agent

The ability of fucoidan to synergize with standard anti-cancer agents and/or reduce toxicity has recently been investigated. Ikeguchi et al. examined the synergistic effect of a HMW fucoidan with colorectal cancer chemotherapy agents; oxaliplatin plus 5-fluorouracil/leucovorin (FOLFOX) or irinotecan plus 5-fluorouracil/leucovorin (FOLFIRI). The test patients received 150 mL/day for 6 months of liquid that contained 4.05 g fucoidan. From the commencement of chemotherapy, toxicities and chemotherapy efficiency were compared. Fucoidan showed no side effects such as allergic dermatitis. Diarrhea, neurotoxicity and myelosuppression were not suppressed by fucoidan, whereas general fatigue was significantly decreased from 60% to 10%. The patients were followed for approximately 15 months and the survival rate of the patients who received fucoidan was longer than that of the control participants; however the difference was not significant, probably due to the small numbers [61].

Fucoidan affects the migration and invasion of multiple myeloma (MM) cells treated with chemotherapy drug cytarabine. The human myeloma cell lines RPMI8226 and U266 were treated with crude fucoidan from F. vesiculosus for 72 h and then cytarabine for 6 h. Fucoidan reduced cell migration
through a Boyden chamber and down-regulated expression of CXCR4 and MMP-9 [62]. Fucoidan from *Saccharina cichorioides* has been reported to synergize with the anti-tumor activity of low dose resveratrol (a natural polyphenol extracted from foods and beverages) on invasive and highly motile HCT 116 colon cancer cell line [63]. In the colony formation assay, fucoidan plus resveratrol reduced the colony number by 60% compared to 34% and 27% in resveratrol alone or fucoidan alone, respectively.

Zhang *et al.* studied the combinatory effect of fucoidan and three commonly used anti-cancer agents; cis-platin (CDDP), tamoxifen (TAM) and paclitaxel (Taxol) on signal transduction pathways. Fucoidan from *Cladosiphon navae-caledoniae* plus anti-cancer agents reduced the ERK phosphorylation in MDA-MB-231 breast cancer cells compared to untreated control or fucoidan alone [64]. Dietary fucoidan synergistically reduced cell growth in the OE33 cell line when it was combined with lapatinib, a targeted therapy that acts as a tyrosine kinase inhibitor in advanced HER2-positive breast cancer cells [65].

In a xenograft transplantation study, the effect of fucoidan alone or in combination with cyclophosphamide was examined on tumor growth. Nine days after the injection of Lewis lung carcinoma cells into mice, fucoidan from *Fucus evanescens* was administered to animals alone or combined with cyclophosphamide. The fucoidan group showed marked antitumor (33% tumor growth inhibition) and anti-metastatic (29% reduction of the number of metastases) activities. However, fucoidan did not exhibit a synergistic effect with cyclophosphamide on tumor growth, but significantly decreased the lung cancer cells metastasis [66].

### 3.4. Why Fucoidan Usage is Complicated?

Despite the promising results about the anti-cancer effect of fucoidan, there are still challenges impeding utilization of fucoidan in the clinic. Variable and contradictory results being influenced by endogenous and exogenous factors in fucoidan usage are of the main concerns. In this section we will summarize important conditions, which have been undertaken in different experiments and have led to such variable results in reported studies.

#### 3.4.1. Structure and Molecular Weight Variation

Fucoidan is composed of α-(1-2) or α-(1-3)-linked L-fucose with a fucose content of 34-44%. It also contains various amounts of other monosaccharide such as galactose, mannose, xylose and uronic acid all of which make up less than 10% of the total fucoidan structure [67,68]. The sulfate groups in fucoidan structure are mainly at position 4 but they can also occupy position C2 and occasionally C3 [53]. The fucoidan structure and monosaccharide composition vary depending on different factors such as the source of fucoidan, the time and location of harvesting and the extraction method, which can affect the fucoidan’s bioactivities. Most anti-cancer studies of fucoidan have used a commercially available crude fucoidan extracted from *Fucus vesiculosus* (Sigma Co. St. Louis, MO, USA). Some groups have extracted and purified fucoidan in their own laboratories. Okinawa mozuku, *C. Okamuranus* tokida, *Sargassum* sp. and *Undaria pinnatifida* are the most common fucoidans examined in cancer studies.

Cumashi *et al.* studied different biological aspects of fucoidan from nine different species of brown seaweed in rats [38]. Analysis of P-selectin-mediated neutrophil adhesion to platelets revealed that
extracted fucoidans from only some sources like *F. evanescens* and *A. nodosum* could serve as more efficient P-selectin inhibitors. Furthermore, in contrast to other sources, fucoidan from *C. okamuranus* did not exert anti-coagulant activity, which was suggested to be due to high content of 2-O-a-D-glucuronyl substituent in the polysaccharide chain of fucoidan from *C. okamuranus*.

Sulfation is another key factor in fucoidan bioactivity. More sulfation is linked with greater bioactivity and thus researchers have produced over-sulfated fucoidans to enhance its biological properties [36]. It has been suggested that over-sulfation causes higher negative charge in the molecule which can facilitate formation of fucoidan-protein complexes involved in cell proliferation [69].

Molecular weight is another crucial factor in fucoidan activity. Cho *et al.* produced three fucoidan fractions with molecular weights of <5, 5–30 and >30 kDa and reported that the F5-30K showed the most tumor growth inhibitory effect despite the sulfate amount in F<5K being greater than in the two other fractions [70].

The extraction method can also affect fucoidan’s bio-properties. Fucoidan from *Undaria pinnatifida* was hydrolyzed using different hydrolysis conditions and their anti-cancer activity was compared *in vitro*. The native fucoidan showed 37% anti-cancer activity; hydrolyzed fucoidan generated under mild conditions (in boiling water with HCl for 5 min) exhibited 75.9% anti-tumor activity; whereas hydrolyzed fucoidan generated under harsh conditions (microwave for more than 90 s) slightly enhanced the anti-cancer effect [71].

### 3.4.2. Fucoidan Dose and Route of Administration

As fucoidan is a large highly branched molecule, the dosage for *in vitro* studies mostly resides in the range of µg/mL and not ng/mL. However, there is a large variation in the doses. Vischchuk *et al.* treated HCT-116 colon cancer cells with 100–800 µg/mL fucoidan from the brown alga *Saccharina cichorioides* Miyabe and observed that fucoidan exerted a low cytotoxicity and there was less than 15% reduction in cell number with the high dose of 800 µg/ml after 24 h [63]. In contrast, Kim *et al.* demonstrated that 20 µg/mL fucoidan from *F. vesiculosus* caused 37% growth inhibition in the same cell line after 72 h [24]. Though the difference between incubation times (24 h vs. 72 h) should be considered, the dose difference (800 µg/mL vs. 20 µg/mL) was substantial. The source of fucoidan appears to be the main factor leading to variation in results. Though most researchers have utilized dosages of less than 1 mg/mL, there are reports of use of up to 3 mg/mL fucoidan.

Regarding the *in vivo* studies, both dose and the route of administration can affect outcome. To select the most effective dose, mice were treated with various doses of fucoidan (10–400 mg/kg body weight) followed by total-body irradiation. The mice injected with 100 mg/kg body weight fucoidan showed the best survival rate at 30 days post-irradiation [72]. Other studies have used various doses ranging from 5 mg/kg to 100 mg/kg and occasionally doses up to 500 mg/kg/body weight of different fucoidan extracts. The amount and number of doses of fucoidan administration has also been shown to be important for *in vivo* studies. Alekseyenko *et al.* studied mice with lung carcinoma that were treated with fucoidan from *Fucus evanescence*. They found that a single injection of 25 mg/kg/body weight of fucoidan did not inhibit tumor cell proliferation, while three-time injections of 10 mg/kg/body weight significantly reduced tumor growth and metastasis [66]. Most *in vivo* studies of anti-tumor activity have selected intraperitoneal (IP) injections, but subcutaneous (SC) or intravenous (IV) routes of
administration have also been used. Oral fucoidan is another route for in vivo delivery either for its anti-tumor properties following tumor induction or as a neoplastic transformation inhibitor administered prior to cancer induction. Taken together, these studies indicate that different delivery routes can affect the fucoidan metabolism in vivo and lead to variable outcomes.

3.5. Fucoidan Toxicity

Whilst fucoidan consumed in food in the form of 4% of the total dry weight of brown seaweeds is generally regarded as safe, the fucoidan used for research is a highly purified extract. For in vitro studies, researchers have utilized normal cells such as normal fibroblasts alongside tumor cell lines and reported that fucoidan did not induce apoptosis within normal cells at the doses which were toxic for cancer cell lines. A very high dose of 3 mg/mL fucoidan suppressed the viability of peripheral blood mononuclear cells from healthy donors by 25% compared to 60%–90% in five different leukemic T-cells [20]. In vivo, oral administration of up to 1 g/mL/body weight Undaria pinnatifida fucoidan was non-toxic in mice but higher doses (2 g/mL/body weight) induced changes in thyroid weight and altered levels of triglyceride and alanine transaminase activity [73]. In another study, daily administration of 300 mg/kg/body weight fucoidan from Laminaria japonica in Wister rats over 6 months did not induce any adverse side effects, but higher doses (900–2500 mg/mL) resulted in coagulopathy and markedly elevated clotting time [74].

Toxicity has also been examined in the context of fucoidan use as adjuvant. Oh et al. examined the combinatory effect of fucoidan with the standard anti-Her2 inhibitor lapatinib in different breast cancer cell lines in vitro [65] and found that fucoidan decreased the efficiency of lapatinib and exerted antagonistic effects on cell proliferation in a few cell lines. Examining the effect of combination of fucoidan from Fucus evanescence with cyclophosphamide, 7 out of 10 mice that were injected with 25 mg/kg/body weight fucoidan plus cyclophosphamide died whereas of the mice that were treated with fucoidan alone, 3 out of 10 died [66].

Fucoidan has been examined in several clinical trials mainly for its anti-coagulant and anti-viral properties. Administration of capsules containing 560 mg fucoidan from Undaria pinnatifida for up to 24 months did not induce any side effect when the participants took 4 capsules a day [75]. In Mori et al. [76] and Irhimeh et al. [77] studies, daily consumption of 5 capsules contained 166 mg fucoidan from C okamuranus Tokida for over one year and 3 g HMW fucoidan from Undaria pinnatifida for up to 12 days, respectively, were revealed to be safe. However, Irhimeh et al. demonstrated that orally administered fucoidan affected coagulation tests and prolonged the aPTT, thrombin time and AT-III. Other studies have also shown the potential of bleeding complication development due to fucoidan’s anti-thrombotic property [78]. Diarrhea is another reported side effect, which was seen in 4 out of 17 participants within 1 month of daily administration of 6 g fucoidan [79].

When a blend of three different extracts (from Fucus vesiculosus (85% w/w), Macrocystis pyrifera (10% w/w), and Laminaria japonica (5% w/w)) in capsules containing up to 187.5 mg were daily given to volunteers, a statistically significant change in the potassium level was seen after 28 days. Although, the change was minor and within the clinical reference range [80].
4. Conclusions

The goal of cancer treatment is eradication of tumor cells ideally with minimal damage to healthy tissues. Because of the side-effects of many current treatments, the use of natural substances of low toxicity is of interest. A number of in vitro and in vivo studies have indicated that fucoidan contains strong anti-cancer bioactivity. Since fucoidan also possesses immunomodulatory effects, it is postulated that it may have protective effects against development of side effects when it is co-administered with chemotherapeutic agents and radiation.

In this report, we reviewed the underlying cellular mechanisms by which fucoidan induces cell death within tumor cells and increases the survival rate of tumor-bearing animal models by suppression of metastasis and angiogenesis. However despite numerous promising pre-clinical reports, there are few reported clinical studies so far [61]. In this review we also discussed the challenges impeding utilization of fucoidan in the clinic which include the complex heterogeneous structure of fucoidan, highly variable doses, different administration routes and possible negative interactions with chemotherapy. Due to the wide variation of fucoidan structure and to make future experiments reproducible, it is recommended that the critical bioactivity factors such as fucoidan content, sulfate content, monosaccharide constituents and molecular weight be reported. Attention to these factors will be likely to lead to more consistent reports and ultimately produce the required evidence to underpin clinical studies in near future.

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Authors Contribution

FA conducted the literature research and drafted the manuscript. RML carried out the supervision and edited the manuscript. GMW carried out the supervision and edited the manuscript. AFH carried out the supervision and edited the manuscript. JLD carried out the supervision and edited the manuscript.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Cooper, G.M. The development and causes of cancer. In The Cell: A Molecular Approach, 2nd ed.; Sinauer Associates: Sunderland, MA, USA, 2000.
2. Joo, W.D.; Visintin, I.; Mor, G. Targeted cancer therapy—Are the days of systemic chemotherapy numbered? Maturitas 2013, 76, 308–314.
3. Cohen, P.R.; Bedikian, A.Y.; Kim, K.B. Appearance of new vemurafenib-associated melanocytic nevi on normal-appearing skin: Case series and a review of changing or new pigmented lesions in patients with metastatic malignant melanoma after initiating treatment with vemurafenib. *J. Clin. Aesthet. Dermatol.* 2013, 6, 27–37.

4. Huang, V.; Hepper, D.; Anadkat, M.; Cornelius, L. Cutaneous toxic effects associated with vemurafenib and inhibition of the braf pathway. *Arch. Dermatol.* 2012, 148, 628–633.

5. Dotan, E.; Aggarwal, C.; Smith, M.R. Impact of rituximab (rituxan) on the treatment of b-cell non-hodgkin’s lymphoma. *P T* 2010, 35, 148–157.

6. Senderowicz, A.M. Flavopiridol: The first cyclin-dependent kinase inhibitor in human clinical trials. *Investig. New Drugs* 1999, 17, 313–320.

7. George, S.K.; Rajesh, R.; Kumar, S.S.; Sulekha, B.; Balaram, P. A polyherbal ayurvedic drug—Indukantha ghritha as an adjuvant to cancer chemotherapy via immunomodulation. *Immunobiology* 2008, 213, 641–649.

8. Caliceti, P.; Salmaso, S.; Bersani, S. Polysaccharide-based anticancer prodrugs. In *Macromolecular Anticancer Therapeutics*; Reddy, L.H., Couvreur, P., Eds.; Springer: New York, NY, USA, 2010; pp. 163–166.

9. Aravind, S.R.; Joseph, M.M.; Varghese, S.; Balaram, P.; Sreelekha, T.T. Antitumor and immunopotentiating activity of polysaccharide pst001 isolated from the seed kernel of tamarindus indica: An in vivo study in mice. *Sci. World J.* 2012, 2012, 361382.

10. Kalimuthu, S.; Kim, S. Fucoidan, a sulfated polysaccharides from brown algae as therapeutic target for cancer. In *Handbook of Anticancer Drugs from Marine Origin*; Kim, S., Ed.; Springer International Publishing: Cham, Switzerland, 2015; p. 147.

11. Mulloy, B.; Ribeiro, A.; Alves, A.; Vieira, R.; Mourao, P. Sulfated fucans from echinoderms have a regular tetrasaccharide repeating unit defined by specific patterns of sulfation at the o-2 and o-4 positions. *J. Biol. Chem.* 1994, 22113–22123.

12. Ribeiro, A.; Vieira, R.; Mourao, P.; Mulloy, B. A sulfated a-l-fucan from sea cucumber. *Carbohydr. Res.* 1994, 255, 225–240.

13. Kwak, J.Y. Fucoidan as a marine anticancer agent in preclinical development. *Mar. Drugs* 2014, 12, 851–870.

14. Zhang, Z.; Teruya, K.; Eto, H.; Shirahata, S. Fucoidan extract induces apoptosis in mcf-7 cells via a mechanism involving the ros-dependent jnk activation and mitochondria-mediated pathways. *PLoS ONE* 2011, 6, e27441.

15. Aisa, Y.; Miyakawa, Y.; Nakazato, T.; Shibata, H.; Saito, K.; Ikeda, Y.; Kizaki, M. Fucoidan induces apoptosis of human hs-sultan cells accompanied by activation of caspase-3 and down-regulation of erk pathways. *Am. J. Hematol.* 2005, 78, 7–14.

16. Riou, D.; Colliec-Jouault, S.; Pinczon du Sel, D.; Bosch, S.; Siavoshian, S.; Le Bert, V.; Tomasoni, C.; Sinquin, C.; Durand, P.; Roussakis, C. Antitumor and antiproliferative effects of a fucan extracted from ascophyllum nodosum against a non-small-cell bronchopulmonary carcinoma line. *Anticancer Res.* 1996, 16, 1213–1218.
17. Moreau, D.; Thomas-Guyon, H.; Jacquot, C.; Jugé, M.; Culioli, G.; Ortalo-Magné, A.; Piovetti, L.; Roussakis, C. An extract from the brown alga bifurcaria bifurcata induces irreversible arrest of cell proliferation in a non-small-cell bronchopulmonary carcinoma line. *J. Appl. Phycol.* 2006, 18, 87–93.

18. Boo, H.J.; Hong, J.Y.; Kim, S.C.; Kang, J.I.; Kim, M.K.; Kim, E.J.; Hyun, J.W.; Koh, Y.S.; Yoo, E.S.; Kwon, J.M.; *et al.* The anticancer effect of fucoidan in pc-3 prostate cancer cells. *Mar. Drugs* 2013, 11, 2982–2999.

19. Banafa, A.M.; Roshan, S.; Liu, Y.Y.; Chen, H.J.; Chen, M.J.; Yang, G.X.; He, G.Y. Fucoidan induces g1 phase arrest and apoptosis through caspases-dependent pathway and ros induction in human breast cancer mcf-7 cells. *J. Huazhong Univ. Sci. Technol. Med. Sci.* 2013, 33, 717–724.

20. Haneji, K.; Matsuda, T.; Tomita, M.; Kawakami, H.; Ohshiro, K.; Uchihara, J.; Masuda, M.; Takasu, N.; Tanaka, Y.; Ohta, T.; *et al.* Fucoidan extracted from cladosiphon okamuranus tokida induces apoptosis of human t-cell leukemia virus type 1-infected t-cell lines and primary adult t-cell leukemia cells. *Nutr. Cancer* 2005, 52, 189–201.

21. Fukahori, S.; Yano, H.; Akiba, J.; Ogasawara, S.; Momosaki, S.; Sanada, S.; Kuratomi, K.; Ishizaki, Y.; Moriya, F.; Yagi, M.; *et al.* Fucoidan, a major component of brown seaweed, prohibits the growth of human cancer cell lines in vitro. *Mol. Med. Rep.* 2008, 1, 537–542.

22. Yamasaki-Miyamoto, Y.; Yamasaki, M.; Tachibana, H.; Yamada, K. Fucoidan induces apoptosis through activation of caspase-8 on human breast cancer mcf-7 cells. *J. Agric. Food Chem.* 2009, 57, 8677–8682.

23. Jin, J.O.; Song, M.G.; Kim, Y.N.; Park, J.I.; Kwak, J.Y. The mechanism of fucoidan-induced apoptosis in leukemic cells: Involvement of erk1/2, jnk, glutathione, and nitric oxide. *Mol. Carcinog.* 2010, 49, 771–782.

24. Kim, E.J.; Park, S.Y.; Lee, J.Y.; Park, J.H. Fucoidan present in brown algae induces apoptosis of human colon cancer cells. *BMC Gastroenterol.* 2010, 10, 96, doi:10.1186/1471-230X-10-96.

25. Boo, H.J.; Hyun, J.H.; Kim, S.C.; Kang, J.I.; Kim, M.K.; Kim, S.Y.; Cho, H.; Yoo, E.S.; Kang, H.K. Fucoidan from undaria pinnatifida induces apoptosis in a549 human lung carcinoma cells. *Phytother. Res.* 2011, 25, 1082–1086.

26. Costa, L.S.; Telles, C.B.; Oliveira, R.M.; Nobre, L.T.; Dantas-Santos, N.; Camara, R.B.; Costa, M.S.; Almeida-Lima, J.; Melo-Silveira, R.F.; Albuquerque, I.R.; *et al.* Heterofucan from sargassum filipendula induces apoptosis in hela cells. *Mar. Drugs* 2011, 9, 603–614.

27. Yang, L.; Wang, P.; Wang, H.; Li, Q.; Teng, H.; Liu, Z.; Yang, W.; Hou, L.; Zou, X. Fucoidan derived from undaria pinnatifida induces apoptosis in human hepatocellular carcinoma smmc-7721 cells via the ros-mediated mitochondrial pathway. *Mar. Drugs* 2013, 11, 1961–1976.

28. Cho, T.M.; Kim, W.J.; Moon, S.K. Akt signaling is involved in fucoidan-induced inhibition of growth and migration of human bladder cancer cells. *Food Chem. Toxicol.* 2014, 64, 344–352.

29. Elmore, S. Apoptosis: A review of programmed cell death. *Toxicol. Pathol.* 2007, 35, 495–516.

30. Ghobrial, I.M.; Witzig, T.E.; Adjei, A.A. Targeting apoptosis pathways in cancer therapy. *CA Cancer J. Clin.* 2005, 55, 178–194.

31. Zhang, Z.; Teruya, K.; Eto, H.; Shirahata, S. Induction of apoptosis by low-molecular-weight fucoidan through calcium- and caspase-dependent mitochondrial pathways in mda-mb-231 breast cancer cells. *Biosci. Biotechnol. Biochem.* 2013, 77, 235–242.
32. Koyanagi, S.; Tanigawa, N.; Nakagawa, H.; Soeda, S.; Shimeno, H. Oversulfation of fucoidan enhances its anti-angiogenic and antitumor activities. *Biochem. Pharmacol.* **2003**, *65*, 173–179.

33. Xue, M.; Ge, Y.; Zhang, J.; Wang, Q.; Hou, L.; Liu, Y.; Sun, L.; Li, Q. Anticancer properties and mechanisms of fucoidan on mouse breast cancer *in vitro* and *in vivo*. *PLoS ONE* **2012**, *7*, e43483.

34. Zhu, C.; Cao, R.; Zhang, S.X.; Man, Y.N.; Wu, X.Z. Fucoidan inhibits the growth of hepatocellular carcinoma independent of angiogenesis. *Evid Based Complement. Alternat. Med.* **2013**, *2013*, 692549.

35. Coombe, D.R.; Parish, C.R.; Ramshaw, I.A.; Snowden, J.M. Analysis of the inhibition of tumour metastasis by sulphated polysaccharides. *Int. J. Cancer* **1987**, *39*, 82–88.

36. Soeda, S.; Ishida, S.; Shimeno, H.; Nagamatsu, A. Inhibitory effect of oversulfated fucoidan on invasion through reconstituted basement membrane by murine lewis lung carcinoma. *Jpn. J. Cancer Res.* **1994**, *85*, 1144–1150.

37. Liu, J.M.; Bignon, J.; Haroun-Bouhedja, F.; Bittoun, P.; Vassy, J.; Fernandjian, S.; Wdzieczak-Bakala, J.; Boisson-Vidal, C. Inhibitory effect of fucoidan on the adhesion of adenocarcinoma cells to fibronectin. *Anticancer Res.* **2005**, *25*, 2129–2133.

38. Cumashi, A.; Ushakova, N.A.; Preobrazhenskaya, M.E.; D’Incecco, A.; Piccoli, A.; Totani, L.; Tinari, N.; Morozovich, G.E.; Berman, A.E.; Bilan, M.I.; *et al.* A comparative study of the anti-inflammatory, anticoagulant, antiangiogenic, and antiadhesive activities of nine different fucoidans from brown seaweeds. *Glycobiology* **2007**, *17*, 541–552.

39. Lee, H.; Kim, J.S.; Kim, E. Fucoidan from seaweed fucus vesiculosus inhibits migration and invasion of human lung cancer cell via pi3k-akt-mtor pathways. *PLoS ONE* **2012**, *7*, e50624.

40. Patel, M.K.; Mulloy, B.; Gallagher, K.L.; O’Brien, L.; Hughes, A.D. The antimitogenic action of the sulphated polysaccharide fucoidan differs from heparin in human vascular smooth muscle cells. *Thromb. Haemost.* **2002**, *87*, 149–154.

41. Hsu, H.Y.; Lin, T.Y.; Hwang, P.A.; Tseng, L.M.; Chen, R.H.; Tsao, S.M.; Hsu, J. Fucoidan induces changes in the epithelial to mesenchymal transition and decreases metastasis by enhancing ubiquitin-dependent tgfbeta receptor degradation in breast cancer. *Carcinogenesis* **2013**, *34*, 874–884.

42. Hyun, J.H.; Kim, S.C.; Kang, J.I.; Kim, M.K.; Boo, H.J.; Kwon, J.M.; Koh, Y.S.; Hyun, J.W.; Park, D.B.; Yoo, E.S.; *et al.* Apoptosis inducing activity of fucoidan in hct-15 colon carcinoma cells. *Biol. Pharm. Bull.* **2009**, *32*, 1760–1764.

43. Zhuang, S.; Schnellmann, R.G. A death-promoting role for extracellular signal-regulated kinase. *J. Pharmacol. Exp. Ther.* **2006**, *319*, 991–997.

44. Ale, M.T.; Maruyama, H.; Tamauchi, H.; Mikkelsen, J.D.; Meyer, A.S. Fucoidan from sargassum sp. And fucus vesiculosus reduces cell viability of lung carcinoma and melanoma cells *in vitro* and activates natural killer cells in mice *in vivo*. *Int. J. Biol. Macromol.* **2011**, *49*, 331–336.

45. Azuma, K.; Ishihara, T.; Nakamoto, H.; Amaha, T.; Osaki, T.; Tsuka, T.; Imagawa, T.; Minami, S.; Takashima, O.; Ifuku, S.; *et al.* Effects of oral administration of fucoidan extracted from cladosiphon okamuranus on tumor growth and survival time in a tumor-bearing mouse model. *Mar. Drugs* **2012**, *10*, 2337–2348.
46. Shimizu, J.; Wada-Funada, U.; Mano, H.; Matahira, Y.; Kawaguchi, M.; Wada, M. Proportion of murine cytotoxic t cells is increased by high molecular-weight fucoidan extracted from okinawa mozuku (cladosiphon okamuranus). J. Health Sci. 2005, 51, 394–397.

47. Hu, Y.; Cheng, S.C.; Chan, K.T.; Ke, Y.; Xue, B.; Sin, F.W.; Zeng, C.; Xie, Y. Fucoidin enhances dendritic cell-mediated t-cell cytotoxicity against ny-eso-1 expressing human cancer cells. Biochem. Biophys. Res. Commun. 2010, 392, 329–334.

48. Jin, J.O.; Zhang, W.; Du, J.Y.; Wong, K.W.; Oda, T.; Yu, Q. Fucoidan can function as an adjuvant in vivo to enhance dendritic cell maturation and function and promote antigen-specific t cell immune responses. PLoS ONE 2014, 9, e99396.

49. Teas, J.; Harbison, M.L.; Gelman, R.S. Dietary seaweed (laminaria) and mammary carcinogenesis in rats. Cancer Res. 1984, 44, 2758–2761.

50. Wakefield, L.M.; Roberts, A.B. Tgf-beta signaling: Positive and negative effects on tumorigenesis. Curr. Opin. Genet. Dev. 2002, 12, 22–29.

51. Humphrey, P.A.; Wong, A.J.; Vogelstein, B.; Zalutsky, M.R.; Fuller, G.N.; Archer, G.E.; Friedman, H.S.; Kwatra, M.M.; Bigner, S.H.; Bigner, D.D. Anti-synthetic peptide antibody reacting at the fusion junction of deletion-mutant epidermal growth factor receptors in human glioblastoma. Proc. Natl. Acad. Sci. USA 1990, 87, 4207–4211.

52. Lee, N.Y.; Ermakova, S.P.; Zvyagintseva, T.N.; Kang, K.W.; Dong, Z.; Choi, H.S. Inhibitory effects of fucoidan on activation of epidermal growth factor receptor and cell transformation in jb6 cl41 cells. Food Chem. Toxicol. 2008, 46, 1793–1800.

53. Silchenko, A.S.; Kusaykin, M.I.; Kurilenko, V.V.; Zakharenko, A.M.; Isakov, V.V.; Zaporozhets, T.S.; Gazha, A.K.; Zvyagintseva, T.N. Hydrolysis of fucoidan by fucoidanase isolated from the marine bacterium, formosa algae. Mar. Drugs 2013, 11, 2413–2430.

54. Irhimeh, M.R.; Fitton, J.H.; Lowenthal, R.M.; Kongtawelert, P. A quantitative method to detect fucoidan in human plasma using a novel antibody. Methods Find Exp. Clin. Pharmacol. 2005, 27, 705–710.

55. Tokita, Y.; Nakajima, K.; Mochida, H.; Iha, M.; Nagamine, T. Development of a fucoidan-specific antibody and measurement of fucoidan in serum and urine by sandwich elisa. Biosci. Biotechnol. Biochem. 2010, 74, 350–357.

56. Deux, J.F.; Meddahi-Pelle, A.; le Blanche, A.F.; Feldman, L.J.; Colliec-Jouault, S.; Bree, F.; Boudghene, F.; Michel, J.B.; Letourneur, D. Low molecular weight fucoidan prevents neointimal hyperplasia in rabbit iliac artery in-stent restenosis model. Arterioscler. Thromb. Vasc. Biol. 2002, 22, 1604–1609.

57. Kimura, R.; Rokkaku, T.; Takeda, S.; Senba, M.; Mori, N. Cytotoxic effects of fucoidan nanoparticles against osteosarcoma. Mar. Drugs 2013, 11, 4267–4278.

58. Thelen, T.; Hao, Y.; Medeiros, A.I.; Curtis, J.L.; Serezani, C.H.; Kobzik, L.; Harris, L.H.; Aronoff, D.M. The class a scavenger receptor, macrophage receptor with collagenous structure, is the major phagocytic receptor for clostridium sordellii expressed by human decidual macrophages. J. Immunol. (Baltimore, Md. 1950) 2010, 185, 4328–4335.

59. Ding, Z.; Issekutz, T.B.; Downey, G.P.; Waddell, T.K. L-selectin stimulation enhances functional expression of surface cxcr4 in lymphocytes: Implications for cellular activation during adhesion and migration. Blood 2003, 101, 4245–4252.
60. Yamasaki, Y.; Yamasaki, M.; Tachibana, H.; Yamada, K. Important role of beta1-integrin in fucoidan-induced apoptosis via caspase-8 activation. *Biosci. Biotechnol. Biochem.* 2012, 76, 1163–1168.

61. Ikeguchi, M.; Yamamoto, M.; Arai, Y.; Maeta, Y.; Ashida, K.; Katano, K.; Miki, Y.; Kimura, T. Fucoidan reduces the toxicities of chemotherapy for patients with unresectable advanced or recurrent colorectal cancer. *Oncol. Lett.* 2011, 2, 319–322.

62. Lv, Y.; Song, Q.; Shao, Q.; Gao, W.; Mao, H.; Lou, H.; Qu, X.; Li, X. Comparison of the effects of marchantin c and fucoidan on sflt-1 and angiogenesis in glioma microenvironment. *J. Pharm. Pharmacol.* 2012, 64, 604–609.

63. Vishchuk, O.S.; Ermakova, S.P.; Zvyagintseva, T.N. The effect of sulfated (1-->3)-alpha-l-fucan from the brown alga saccharina cichorioides miyabe on resveratrol-induced apoptosis in colon carcinoma cells. *Mar. Drugs* 2013, 11, 194–212.

64. Zhang, Z.; Teruya, K.; Yoshida, T.; Eto, H.; Shirahata, S. Fucoidan extract enhances the anti-cancer activity of chemotherapeutic agents in mda-mb-231 and mcf-7 breast cancer cells. *Mar. Drugs* 2013, 11, 81–98.

65. Oh, B.; Kim, J. Anticancer effect of fucoidan in combination with tyrosine kinase inhibitor lapatinib. *Evid. Based Complement. Alternat. Med.* 2014, 2014, 865375.

66. Alekseyenko, T.V.; Zhanayeva, S.Y.; Venediktova, A.A.; Zvyagintseva, T.N.; Kuznetsova, T.A.; Besednova, N.N.; Korolenko, T.A. Antitumor and antimetastatic activity of fucoidan, a sulfated polysaccharide isolated from the okhotsk sea fucus evanescens brown alga. *Bull. Exp. Biol. Med.* 2007, 143, 730–732.

67. Mabeva, S.; Kloareg, B.; Joseleau, J. Fractionation and analysis of fucans from brown algae. *Phytochemistry* 1990, 29, 2441–2445.

68. Black, W. The seasonal variation in the combined l-fucose content of the common british laminariaceae and fucaceae. *J. Sci. Food Agric.* 1954, 5, 445–448.

69. Haroun-Bouhedja, F.; Ellouali, M.; Sinquin, C.; Boisson-Vidal, C. Relationship between sulfate groups and biological activities of fucans. *Thromb. Res.* 2000, 100, 453–459.

70. Cho, M.L.; Lee, B.Y.; You, S.G. Relationship between oversulfation and conformation of low and high molecular weight fucoidans and evaluation of their in vitro anticancer activity. *Molecules* 2010, 16, 291–297.

71. Yang, C.; Chung, D.; Shin, I.S.; Lee, H.; Kim, J.; Lee, Y.; You, S. Effects of molecular weight and hydrolysis conditions on anticancer activity of fucoidans from sporophyll of undaria pinnatifida. *Int. J. Biol. Macromol.* 2008, 43, 433–437.

72. Lee, J.; Kim, J.; Moon, C.; Kim, S.H.; Hyun, J.W.; Park, J.W.; Shin, T. Radioprotective effects of fucoidan in mice treated with total body irradiation. *Phytother. Res.* 2008, 22, 1677–1681.

73. Chung, H.J.; Jeun, J.; Hounge, S.J.; Jun, H.J.; Kweon, D.K.; Lee, S.J. Toxicological evaluation of fucoidan from undaria pinnatifidain vitro and in vivo. *Phytother. Res.* 2010, 24, 1078–1083.

74. Li, N.; Zhang, Q.; Song, J. Toxicological evaluation of fucoidan extracted from laminaria japonica in wistar rats. *Food Chem. Toxicol.* 2005, 43, 421–426.

75. Cooper, R.; Dragar, C.; Elliot, K.; Fitton, J.H.; Godwin, J.; Thompson, K. Gfs, a preparation of tasmanian undaria pinnatifida is associated with healing and inhibition of reactivation of herpes. *BMC Complement. Altern. Med.* 2002, 2, 11; doi:10.1186/1472-6882-2-11.
76. Mori, N.; Nakasone, K.; Tomimori, K.; Ishikawa, C. Beneficial effects of fucoidan in patients with chronic hepatitis c virus infection. World J. Gastroenterol. 2012, 18, 2225–2230.
77. Irhimeh, M.R.; Fitton, J.H.; Lowenthal, R.M. Fucoidan ingestion increases the expression of cxcr4 on human cd34+ cells. Exp. Hematol. 2007, 35, 989–994.
78. Millet, J.; Jouault, S.C.; Mauray, S.; Theveniaux, J.; Sternberg, C.; Boisson Vidal, C.; Fischer, A.M. Antithrombotic and anticoagulant activities of a low molecular weight fucoidan by the subcutaneous route. Thromb. Haemost. 1999, 81, 391–395.
79. Araya, N.; Takahashi, K.; Sato, T.; Nakamura, T.; Sawa, C.; Hasegawa, D.; Ando, H.; Aratani, S.; Yagishita, N.; Fujii, R.; et al. Fucoidan therapy decreases the proviral load in patients with human t-lymphotropic virus type-1-associated neurological disease. Antivir. Ther. 2011, 16, 89–98.
80. Myers, S.P.; O’Connor, J.; Fitton, J.H.; Brooks, L.; Rolfe, M.; Connellan, P.; Wohlmuth, H.; Cheras, P.A.; Morris, C. A combined phase I and II open label study on the effects of a seaweed extract nutrient complex on osteoarthritis. Biologics 2010, 4, 33–44.

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