Introduction

Diabetes mellitus is a complex chronic metabolic disorder that is a major source of ill health worldwide. It is characterized by hyperglycemia and disturbances of carbohydrate, protein and fat metabolisms secondary to an absolute or relative lack of hormone insulin [1]. The elevated blood glucose level is considered the principal cause of complications in diabetes mellitus [2]. Diabetic complications usually arise as a result of non-enzymatic protein glycation, which leads to the formation of heterogeneous, toxic and antigenic advanced glycation end products (AGEs) [3]. The accumulation of AGEs in vivo has been considered to play a major role in the pathogenic process of diabetes and its complications, including neuropathy, nephropathy, retinopathy, cataract and in other health disorder such as Alzheimer’s disease and aging [4, 5,6]. Thus, the investigation of compounds with an AGEs inhibitor activity, would certainly offer a potential therapeutic approach for the prevention of diabetes or other pathogenic complications. As currently, there is growing interest in herbal remedies due to the side effects associated with the oral hypoglycemic agents for the treatment of diabetes mellitus; it was decided to study the effect of extracts of some indigenous plants known for their antidiabetic activity namely A. lebbeck, B. aristata and M. pruriens on non-enzymatic glycosylation of hemoglobin. Albizia lebbeck Benth. (Family: Leguminosae) is a deciduous tree with compound leaves, flat oblong fruits, round cream colored seeds, grows wild. Barks are used in toothache and diseases of the gum. Decoction of the leaves and barks are protective against bronchial asthma and other allergic disorders. Barks and seeds are astringent and are given in piles and diarrhea. Ethanolic extract of pods possesses antiprotozoal, hypoglycemic and anticancer properties [7].

Berberis aristata DC. (Family: Berberidaceae) is used in Ayurveda medicines from very long time. This plant has been considered as a valuable medicine for the treatment of remittent fevers, oxidative stress and used as a cooling laxative to children and as a tonic remedy for liver and heart.

It exhibits febrifugal, hypotensive, immuno-stimulating, anti-inflammatory, antidiabetic, antimicrobial, antiprotozoal, anticholinergic and antiarrhythmic activities [8]. Mucuna pruriens Linn. (Family: Fabaceae) is one of the popular drug in Ayurvedic system of medicine [9]. Various preparations from the seeds of this plant are used for the management of several free radical mediated diseases such as ageing, rheumatoid arthritis,
Mangesh B, Somnath B, Dheeraj R, Ganesh W, Sachin T. Studies on In-Vitro Antiglycation Potential of Some Indigenous Antidiabetic Plants. Glob J Pharmaceu Sci. 2017; 3(5): 555624. DOI: 10.19080/GJPPS.2017.03.555624

Materials and Methods

Plant material

The bark of A. lebbeck and seeds of M. pruriens were collected from local areas of Karad, whereas the roots of B. aristata were purchased from local market. The plant material was further identified and authenticated by the Department of Botany, Science College, Karad. The bark of A. lebbeck, roots of B. aristata and the seeds of M. pruriens were cleaned, dried in a hot air oven (50 °C), powdered, passed through 60 mesh sieve (BSS) and stored in an airtight container at 4 °C till further use.

Chemicals

Hemoglobin was purchased from Sigma Aldrich, USA. All the chemicals used in the study were of extra pure analytical grade.

Preparation of plant extracts

Aqueous extracts were prepared by extracting the powders of bark of A. lebbeck, roots of B. aristata and the seeds of M. pruriens with hot water (70 °C) in a mechanical shaker (24h), filtered and freeze dried.

Determining the best condition for hemoglobin glycosylation

To find the best glucose concentration, hemoglobin 5g/100ml in 0.01 M phosphate buffer, pH 7.4 was incubated with different concentrations of glucose [12]. The extent of glycosylation was measured by colorimetric method. Thereafter, to find the most useful time for glycosylation, hemoglobin 5g/100ml with the best concentration of glucose was incubated at different times and the amount of glycosylation was measured.

Assay

1ml of hemoglobin solution 5g/100ml and 1ml of the solution containing glucose 2g/100ml and Gentamycin 20mg/100ml in 0.01M phosphate buffer, pH 7.4 were incubated in the dark at room temperature. Then, the glycosylation degree of hemoglobin in the presence of different concentrations of plant extracts and their absences were measured by the colorimetric method.

Results and Discussion

To find the inhibitory effects of selected plant extracts on glycosylation of hemoglobin, initially hemoglobin 5g/100ml was incubated in the presence of different concentrations of glucose and then the degree of glycosylation was measured by the colorimetric method. It was observed that up to the concentration of 2g/100ml stock solution of glucose, the amount of glycosylation increased linearly (Figure 1). So as to find the suitable time for incubation in this study, hemoglobin 5g/100ml was incubated in the presence of glucose 2g/100ml at different times and the degree of glycosylation was measured by the colorimetric method.

The results showed that glycosylation increased up to the time of 72h linearly and therefore, 72h was chosen as the best time for this study (Figure 1). Different concentrations of the plant extracts were used in the study viz. 250µg/ml, 500µg/ml, 750µg/ml and 1000µg/ml. The inhibitory effects on glycosylation of hemoglobin at these concentrations were estimated as 19.08%, 29.22%, 37.86% and 48.55%, for A. lebbeck extracts. For B. aristata it was observed as 29.66%, 37.05%, 49.0%, 56.17%, whereas for M. pruriens extract the inhibitory effects on hemoglobin glycosylation were observed as 45.83%, 58.73%, 70.14%, and 75.60% at a concentration of 250 µg/ml, 500 µg/ml, 750µg/ml and 1000 µg/ml respectively.

Figure 1: 1ml of hemoglobin solution 5g/100ml and 1ml of the solution containing different concentrations of glucose in 0.01M phosphate buffer, pH 7.4 were incubated at room temperature at different time intervals.
Increased concentration of glucose in the blood as observed in diabetes mellitus leads to its binding to hemoglobin and other plasma proteins which may result in the formation of the reactive oxygen species. Plant extracts may play an important role in the inhibition of the glycosylation end products. Our studies revealed that glycosylation of hemoglobin increases markedly on its incubation with the increasing concentration of the glucose (2mg, 4mg, 8mg, 10mg and 20mg) over a period of 72hrs (Figure 1). However, the plant extracts promisingly inhibited the glycosylation of hemoglobin as shown in (Figure 2).

The results of the studies clearly demonstrated that non-enzymatic nature of hemoglobin glycosylation could be effectively inhibited by the extracts of M. pruriens, B. aristata and A. lebbeck at a desirable concentration. Amongst the plant extracts studied, M. pruriens exhibited higher inhibition of glycosylation indicating that the extract of M. pruriens decreases the formation of the glucose- hemoglobin complex and thus amount of free hemoglobin increases. The vivo effect should be investigated so that it can be utilized to prevent or treat complication associated with diabetes.

Conflict of Interest Statement

We declare that we have no conflict of interest.

Acknowledgement

The authors are grateful to the Principal, Govt. College of Pharmacy, Karad for providing all the research facilities.

References

1. Bhutkar MA, Bhise SB (2011) Spices and condiments in the management of Diabetes mellitus. Res J Pharm Technol 4(1):1-6.

2. Peppa M, Uribarri J, Vlassara H (2003) Glucose, advanced glycation end products, and diabetes complications: what is new and what works. Clin Diabetes 21(4): 186-187.

3. Zhicai Zhang, Pengjie Cui (2007) Non-enzymatic glycosylation reaction contributes to a rise of blood glucose in alloxan-induced diabetic rats. Int J Diabetes and Metabolism 15: 52-59.

4. Ahmed N (2005) Advanced glycation end products role in pathology of diabetic complications. Diabetes Res Clin Pract 67(1): 3-21.

5. Vitek MP, Bhattacharya K, Glendening JM, Stopes E, Vlassara H, et. al (1994) Advanced glycation end products contribute to amyloidosis in Alzheimer disease. Proc Natl Acad Sci U S A 91(11): 4766-4770.

6. Brownlee M (1995) Advanced protein glycosylation in diabetes and ageing. Annu Rev Med 46: 223-234.

7. Rahul C, Lincy J, Methew G, Pradhan P (2010) Pharmacognostic standardization and phytochemical screening of Albizia lebbeck. J Chem Pharm Res 2(1):432-443.

8. Rimbau V, Cerdan C, Vila R, Iglesia J (1999) Anti-inflammatory activity of some extracts from plants used in traditional medicines of North-African Countries. Phytother Res 13(2): 128-132.

9. Sharma BK, Shamim A, Singh R (2012) A review on M. pruriens: its phytoconstituents and therapeutic uses. Novel Sci Int J Pharm Sci 1(6): 308-312.

10. Vaidya RA, Allorkar SD, Seth AR, Pandey SK (1978) Activity of bromoergocryptine, M. pruriens and L-Dopa in the control of hyperprolactenaemia. Neurology 26: 179-186.

11. Bhutkar MA, Bhise SB (2013) In vitro hypoglycemic effects of Albizia lebbeck and Mucuna pruriens. Asian Pac J Trop Biomed 3(11): 866-870.

12. Fluckiger R, Winterhalter KH (1976) In vitro synthesis of hemoglobin. North-Holland Publishing Company, Amsterdam (71): p.354-356.
How to cite this article: Mangesh B, Somnath B, Dheeraj R, Ganesh W, Sachin T. Studies on In-Vitro Antiglycation Potential of Some Indigenous Antidiabetic Plants. Glob J Pharmaceut Sci. 2017; 3(5): 555624. DOI: 10.19080/GJPPS.2017.03.555624