Original Article
Isolation of Bioactive Compounds That Relate to the Anti-Platelet Activity of Cymbopogon ambiguus

I. Darren Grice,1 Kelly L. Rogers,2,3 and Lyn R. Griffiths3

1 Institute for Glycomics, Gold Coast campus, Griffith University, Queensland, 4222, Australia
2 Plate-forme d’imagerie dynamique, Institut Pasteur, Paris Cedex 15, France
3 Genomics Research Centre, Gold Coast campus, Griffith University, Queensland, 4222, Australia

Correspondence should be addressed to I. Darren Grice, d.grice@griffith.edu.au

Received 27 May 2009; Accepted 19 November 2009

Copyright © 2011 I. Darren Grice et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Infusions and decoctions of Cymbopogon ambiguus have been used traditionally in Australia for the treatment of headache, chest infections and muscle cramps. The aim of the present study was to screen and identify bioactive compounds from C. ambiguus that could explain this plant’s anti-headache activity. A dichloromethane extract of C. ambiguus was identified as having activity in adenosine-diphosphate-induced human platelet aggregation and serotonin-release inhibition bioassays. Subsequent fractionation of this extract led to the isolation of four phenylpropenoids, eugenol, elemicin, eugenol methylether and trans-isoelemicin. While both eugenol and elemicin exhibited dose-dependent inhibition of ADP-induced human platelet serotonin release, only eugenol displayed potent inhibitory activity with an IC50 value of 46.6 μM, in comparison to aspirin, with an IC50 value of 46.1 μM. These findings provide evidence to support the therapeutic efficacy of C. ambiguus in the non-conventional treatment of headache and inflammatory conditions.

1. Introduction

The native Australian lemongrass species, Cymbopogon ambiguus A. Camus. (Poaceae) is a strongly aromatic perennial grass found on rocky hillsides throughout the Northern Territory of Australia [1]. The leaves have been used traditionally to treat chest infections, sores, muscle cramps as well as headache and associated complaints (infusions and decoctions) [1–3]. Interestingly, a study on Australian medicinal plants identified very weak anti-viral (Ross River Virus) activity in a leaf extract of C. ambiguus [4], while another study found no anti-bacterial activity in extracts of Australian C. ambiguus against four Gram-positive and four Gram-negative bacterial species [5]. Little is known about the chemical constituents present in C. ambiguus apart from a GC-MS study by Barr et al. [3], which identified camphene, borneol, limonene, α-pinene, α-terpineol, camphor, isoborneol, 4-terpineol, myrcene, β-ocimene as being present in the essential oil. Nothing has been reported to date in relation to substantiating its use as a non-conventional traditional remedy for headache, apart from our original report showing that dichloromethane (DCM) (more potent extract) and methanolic (MeOH) extracts displayed potent inhibition of human platelet aggregation and serotonin (5-HT, 5-hydroxytryptamine) release [6]. To further investigate the basis for use of C. ambiguus as a remedy for headache, we report here the isolation and identification of principal bioactive constituents causing inhibition of human platelet aggregation and serotonin (5-HT) release.

Cymbopogon ambiguus is a headache remedy used in Australian traditional medicine for generalized headache disorders. Headaches associated with nausea, such as migraine are complex disorders, with abnormalities in platelet function reported. Altered 5-HT transport, decreased platelet 5-HT content, altered platelet cytosolic free-calcium concentrations and varying sensitivity to platelet agonists such as adenosine diphosphate (ADP) and collagen have been reported in association with headache/migraine episodes [7–14]. The serotonergic system is thus thought to play an important role in the pathophysiology of these disorders [15–20]. We and others have previously reported on the use of platelets as a model to assess the therapeutic serotonergic potential of tested chemicals in relation to potential headache/migraine treatment [6, 21, 22].
**Cymbopogon ambigus** has been used as a traditional Australian aboriginal headache treatment and our studies have shown that it displays potent anti-platelet activity. To identify the bioactive constituent(s) responsible for this activity, lemongrass leaves were sequentially extracted with DCM then MeOH. Fractionation of the DCM extract led to the identification of four phenylpropenoids as the principal constituents. We describe herein the isolation and purification of these four phenylpropenoids from *C. ambigus* along with pharmacological evaluation of two of these compounds. This is the first report identifying specific constituents that underpin the use of *C. ambigus* as a non-conventional traditional Australian anti-headache medicine.

### 2. Methods

#### 2.1. Plant Collection. **Cymbopogon ambigus** (whole plant) was collected in 1998 in the vicinity of Alice Springs, Northern Territory, Australia. Identity of the material was confirmed by the Alice Springs Herbarium, with voucher specimens deposited at the Herbarium and also in the Genomics Research Centre, Gold Coast campus, Griffith University (sample ID: DN3081).

#### 2.2. Extraction and Isolation. Dried whole plants (283 g) of *C. ambigus* were powdered and exhaustively extracted (vigorous stirring in 2.5 L of solvent for 3 h at 22°C, then procedure repeated) with DCM followed by MeOH at room temperature. The DCM extract was then concentrated and dried under reduced pressure (~50 mmHg, 30°C) to give a dark green aromatic solid residue (7.53 g) and the residue loaded onto a normal phase chromatography column (silica gel, 150 g, Merck, 60 μM, 4.3 × 11.7 cm). The column was then eluted by passing 600 ml volume through the column from 10% hexane in chloroform, to 100% hexane in 10% increments. Fractions eluted with 50% and 60% hexane were both found to exhibit significant inhibition of ADP-induced platelet [14C]-5-HT release. Analytical HPLC [Luna C18, 5 μM analytical, 4.6 mm × 250 mm, photo-diode-array (PDA) detection at 270 nm, 1.0 ml min⁻¹, method: 25%–35% acetonitrile/H₂O gradient over 10 min, followed by an increase to 45% acetonitrile over a further 50 min] of these fractions indicated five common constituents, which were separated by semi-preparative HPLC [Luna C18, 5 μM, 10 mm × 250 mm, PDA, 4.5 ml min⁻¹, method as above] to give the known compounds: eugenol (4-allyl-2-methoxyphenol) (1) (75.4 mg, 0.02% w/w); elemicin (5-allyl-1,2,3-trimethoxybenzene) (2) (520 mg, 0.18% w/w); eugenol methylether (4-allyl-1,2-dimethoxybenzene) (3) (322 mg, 0.11% w/w); and trans iso-elemicin (4) (1,2,3-trimethoxy-5-(1-propenyl) benzene) (162 mg, 0.06% w/w) (Figure 3).

#### 2.3. Chemicals and Solvents. ADP, acetyl salicylic acid (ASA or aspirin), dimethyl sulfoxide (DMSO) and deuterated chloroform (CDCl₃) were all obtained from Sigma chemical company (St Louis, USA). [14C]-5-hydroxytryptamine ([14C]-5-HT) (Amersham, specific activity 57 mCi mmol⁻¹, 50 μCi ml⁻¹). All solvents used were of HPLC grade, including acetonitrile (CH₃CN) (omnisolv. EM Science, Merck), chloroform (CHCl₃) (chromasolv. Riedel-de Haen), DCM (chromasolv. Riedel-de Haen), ethyl acetate (Bankers Scientific Co. Pty. Ltd.), hexane (Malinckrodt chromAR HPLC) and methanol (Omnisolv, EM Science, Merck).

#### 2.4. Characterization of Isolated Compounds. Structural characterization of the four phenylpropenoids was achieved by mass spectrometry (MS) (Bruker Daltonics BioAPEX 47e), extensive NMR experiments (Bruker AC-300, Karlsruhe, Germany) (1H, 1H-1HC O S Y , 1H-13C HMBC, JMOD) and comparison of 1H NMR (300 MHz, CDCl₃) data (see below) with previously reported assignments. Eugenol (1) [23]: 1H NMR δ 3.33 (d, 2H, 3J = 6.7 Hz, CH₂); 3.88 (s, 3H, OCH₃); 5.06 (dd, 1H, 2J = 2 Hz, 3Jcis = 10 Hz, HCHCH₂); 5.07 (dd, 1H, 2J = 2 Hz, 3Jtrans = 17 Hz, HCHCH₂); 5.50 (OH); 5.93 (ddd, 1H, 3Jcis = 10 Hz, 3Jtrans = 17 Hz, 3Jmeta = 6.7 Hz, HCHCH₂); 6.67–7.62 (m, 2H, CCHCH(OH), CCHC(OCH₃)); 6.86 (dd, 1H, Jortho = 8.5 Hz, Jmeta = 4.2 Hz, CCHCH(OH)). MS calculated for C₁₀H₁₆O₂ [M + H]⁺: 165.08. Found: 165.08. Elemicin (2) [24]: 1H NMR δ 3.33 (d, 2H, 3J = 6.6 Hz, CH₂); 3.82 (s, 3H, OCH₃); 3.84 (s, 6H, (H₃CO)CC(OCH₃)CC(OCH₃)); 5.08 (dd, 1H, 2J = 1.8 Hz, 3Jcis = 10 Hz, HCHCH₂); 5.11 (dd, 1H, 2J = 1.8 Hz, 3Jtrans = 17 Hz, HCHCH₂); 5.95 (ddd, 1H, 3Jcis = 10 Hz, 3Jtrans = 17 Hz, 3Jmeta = 6.6 Hz, HCHCH₂); 6.60 (2H, 2 × aromatic H). MS calculated for C₁₀H₁₆O₃Na [M + Na⁺]: 231.11. Found: 231.11. Eugenol methylether (3) [23]: 1H NMR δ 3.31 (d, 2H, 3J = 6.6 Hz, HC = CH₂); 3.84 (s, 3H, OCH₃); 3.85 (s, 3H, OCH₃); 5.04 (dd, 1H, 2J = 1.5 Hz, 3Jcis = 10.1 Hz, HCHCH₂); 5.05 (dd, 1H, 2J = 1.5 Hz, 3Jtrans = 16.8 Hz, HCHCH₂); 5.95 (ddd, 1H, 3Jcis = 10.1 Hz, 3Jtrans = 16.8 Hz, 3Jmeta = 6.6 Hz, HCHCH₂); 6.75 (m, 3H, Jortho = 8.4 Hz, Jmeta = 1.4 Hz, 3 × aromatic H). MS calculated for C₁₀H₁₆O₃[M + H⁺]: 179.09. Found: 179.09. trans-Isoelemicin (4) [25]: 1H NMR δ 1.86 (dd, 3H, 3J = 6.4 Hz, 4J = 1.5 HCHC₂); 3.81 (s, 3H, OCH₃); 3.83 (s, 6H, 2 × OCH₃); 6.13 (dd, 1H, Jtrans = 15.6 Hz, 4J = 6.4 Hz, HCHCH₂); 6.31 (dd, 1H, Jtrans = 1.5 Hz, Jtrans = 15.6 Hz, HCHCH₂); 6.54 (s, 2H, 2 × aromatic H). MS calculated for C₁₂H₁₇O₃[M + H⁺]: 209.11. Found: 209.11.

#### 2.5. Platelet Aggregation and [14C]-5HT Release Bioassays. The bioassays utilised are based on a modified published method, described previously by our group [26] and by Groenewegen and Heptinstall [27]. Fresh blood (45 ml) was collected from healthy human volunteers (20–35 year old, not taking aspirin or other drugs likely to interfere with platelet function for at least 2 weeks prior to blood sampling). Platelet aggregation and 5-HT release were measured simultaneously. [14C]-5-HT (6 μl) (specific activity 50 μCi ml⁻¹) was added to 9 ml of citrated blood to label intracellular storage granules. Platelet-poor plasma (PPP) and platelet-rich plasma (PRP) fractions were prepared, with the PRP being adjusted to 300 × 10⁸ platelets/l by dilution with PPP. A sample consisting of 450 μL of PPP and 100 μL of either, PBS with 1% ethanol or DMSO (control), or...
Evidence-Based Complementary and Alternative Medicine 3

Cymbopogon ambiguus
(283 g dry weight)

DCM extract (7.53 g)

MeOH extract (16.84 g)

Silica flash column
hexane —→ CHCl₃
10% gradient elution

Acetonitrile soluble portion
of combined 50% + 60%
hexane fraction (1.16 g)

50% aq. MeOH
fraction (5.73 g)

C₁₈ flash column
H₂O —→ MeOH

100% H₂O fraction
active (601 mg)

Eugenol (1)
(75 mg, 0.03%),
33.8 min

Elemicin (2)
(520 mg, 0.18%),
49.1 min

Eugenol methyl ether (3)
(322 mg, 0.11%),
50.7 min

trans-Iso-elemicin (4)
(162 mg, 0.06%),
53.3 min

Figure 1: Bioassay-guided fractionation of C. ambiguus (whole plant) to isolate compounds (1–4).

Figure 2: HPLC chromatogram of the combined 50:50 and 60:40 hexane/CHCl₃ fractions: eugenol (33.8 min); elemicin (49.1 min); eugenol methylether (50.7 min); trans-i-so-elemicin (53.3 min).

the test sample, was initially measured in order to set 100% aggregation (a decrease in optical density reflects an increase in platelet aggregation). The final concentration of ethanol or DMSO did not exceed 0.2% (v/v). Labeled PRP (450 μl) was then stirred (1000 r.p.m. at 37°C) for 3 min in the presence of 100 μl of the C. ambiguus extract (or isolated compound) or PBS containing either 1% ethanol or DMSO (control). ADP (50 μl) was then added to samples, and platelet aggregation was monitored for 6 minutes. Platelet aggregation was measured in a four-channel platelet aggregometer (Monitor IV plus, Helena Laboratories, Beaumont, USA). Samples of PRP were incubated without the addition of ADP to measure the amount of [¹⁴C] 5-HT that was not taken up by platelets, and which may have been spontaneously released during the procedure (blank). ASA (50 μl, 14 mm) was then added to irreversibly inhibit further 5-HT release, and samples were placed on ice for ~5 min. Samples were then centrifuged (6000 r.p.m.) for 8 min, and duplicate 50 μl aliquots were taken from the supernatant and counted (Wallac 1450, MicroBeta counter) for determination of % [¹⁴C] 5-HT release. Percentage 5-HT release was calculated as follows:

\[
\text{Percentage 5-HT release} = \frac{\text{Amount of } [¹⁴C] 5-HT - \text{blank}}{\text{Total counts} - \text{blank}} \times 100\%.
\]

IC₅₀ values were generated using the software of Graphpad prism (3.0).

2.6. Bioethics Clearance. Blood samples for the platelet studies were collected by a qualified phlebotomist and the study had full ethical clearance from the Griffith University Ethics Committee for experimentation on human subjects. Informed written consent was obtained from all participants.

2.7. Statistical Analysis. Platelet aggregation and 5-HT release response, was expressed as a percentage of the control value. IC₅₀ values (concentration producing 50% inhibition of the maximum response) were calculated by non-linear regression analysis of the dose-response curves using the software of Graphpad Prism (3.0). Results are presented as the means ± SEM of n experiments.
3. Results

3.1. Extraction. Both DCM and MeOH extracts of *C. ambiguus* produced a dark green residue that was strongly aromatic (lemon scented). The total weight of the DCM extract after drying was 7.53 g, and for the MeOH extract, 16.84 g.

3.2. Isolation and Structural Characterization. Silica column chromatography (10% gradient increments from 10:90 hexane:CHCl₃ through to 100:0 hexane:CHCl₃ elution) of the DCM fraction eluted an active 50:50–60:40 hexane fraction. The dried residue (weighing ~1.16 g) was resuspended in CH₃CN and fractionation of the soluble components utilising HPLC, afforded five compounds (Figures 1 and 2). Four of these compounds were purified further (purity was ascertained by HPLC analysis) and then subjected to MS and NMR analysis. The fifth compound (eluting at 54.11 min) (Figure 2) was not successfully purified and was therefore not structurally characterised in these studies.

The assignment of the structures for eugenol, elemicin, eugenol methylether and *trans*-iso-elemicin (Figure 3), which are known compounds, was established on the basis of the ¹H NMR and MS data. Chemical shifts were identical to those previously reported (see Methods section—Characterization of Isolated Compounds). The chemical structure and identity of each compound is shown in Figure 3. The isolation of these compounds was guided by the activity of the active hexane fraction (50% and 60%) in the ADP-induced platelet 5-HT release bioassay. Both eugenol and elemicin were investigated pharmacologically in further studies.

3.3. Platelet Aggregation and [¹⁴C]-5-HT Release Bioassay. For the platelet aggregation and 5-HT release studies, a submaximal concentration of ADP was used to stimulate the platelet response. Elemicin’s affect on inhibiting ADP-induced platelet aggregation was less potent than eugenol in our results and therefore we focussed our interests on eugenol. Effects of increasing eugenol concentrations (0, 46, 61 and 152 μM) on the ADP-induced platelet aggregation clearly showed that concentrations above 61 μM did not inhibit platelet aggregation further. Interestingly, eugenol and elemicin exhibited dose-dependent inhibition of ADP (2 μM)-induced platelet 5-HT release, although eugenol at a much lower concentration than elemicin (see Table 1 and Figure 4 (eugenol data only)). Concentration ranges of eugenol, elemicin and aspirin chosen to examine their inhibitory activity on human platelet 5-HT release were 2.8 × 10⁻⁶ to 2.8 × 10⁻⁴ M, 1.7 × 10⁻⁴ to 3.5 × 10⁻³ and 2.5 × 10⁻⁶ to 2.5 × 10⁻⁴ M, respectively. Despite structural similarities, there were remarkable differences in the level of activity between eugenol and elemicin [1]. Eugenol was up to 50 times more potent than elemicin, with IC₅₀ values calculated to be 46.6 μM and 1729.8 μM, respectively. ASA is an irreversible inhibitor of platelet cyclooxygenase (COX) activity, and still remains the standard to which other anti-platelet drugs are often compared [28]. The dose-dependent effect of eugenol compared favorably to ASA (Figure 4), which exhibited an IC₅₀ value of 46.1 μM in these studies.

4. Discussion

Bioassay-directed fractionation of the DCM extract of *C. ambiguus* afforded the isolation and structural characterization of four compounds, eugenol, elemicin, eugenol methylether and *trans*-iso-elemicin. Like 1,8-cineole (the major active constituent of *Melaleuca, Eucalyptus* and

---

**Figure 3:** Structure of the four phenylpropenoides [eugenol (4-allyl-2-methoxyphenol) (1); elemicin (5-allyl-1,2,3-trimethoxybenzene) (2); eugenol methylether (4-allyl-1,2-dimethoxybenzene) (3) and *trans*-iso-elemicin (1,2,3-trimethoxy-5-(1-propenyl) benzene) (4)] isolated from the Australian native lemongrass species *C. ambiguus*.

**Figure 4:** Dose-dependent inhibition of ADP (2 μM)-induced human platelet [¹⁴C]-5-HT release. Results represent the mean of duplicate analyses performed on PRP obtained from four different individuals.
Table 1: IC₅₀ values for components of C. ambiguus and aspirin against ADP-induced platelet 5-HT release.

| Compound                      | IC₅₀ (µM) |
|-------------------------------|-----------|
| Eugenol                       | 46.6 ± 3.7|
| Elemicin                      | 1729.8 ± 147.5 |
| Acetylsalicylic acid (ASA, Aspirin) | 46.1 ± 3.1 |

Results represent IC₅₀ ± SEM (n = 4) as determined by the software of Graphpad prism (3.0).

![Diagram](image)

**Figure 5:** Platelet aggregation trace showing an example of the effect of low to high doses (0, 46, 61 and 152 µM) of eugenol on ADP-induced platelet aggregation. The change in light transmission is represented as percentage of platelet aggregation using a four-channel platelet aggregometer. Results are duplicates obtained from PRP from two different individuals.

Prostanthera sp.), eugenol and elemicin are volatile monoterpenoids (i.e., C₁₀ compounds) with anti-inflammatory effects [29]. Indeed, other similar compounds have previously been isolated from Cymbopogon sp. including, limonene and α-terpineol, which are generally common components of citrus oils [30, 31]. Eugenol and elemicin have also been isolated from several other plants including Myristica fragrans (nutmeg) and Syzygium aromaticum (clove oil), where they are believed to be some of the major constituents responsible for the biological activities of these plants [29, 32, 33]. Elemicin has been identified previously in another Australian native, Cymbopogon procerus [3]; however, this is the first report of eugenol in the Cymbopogon sp. and indeed the first evidence for the presence of these compounds in the Australian native lemongrass species, C. ambiguus.

In these studies, eugenol was found to be up to 50-fold more active than elemicin at inhibiting 5-HT release from human platelets. From dose-dependant responses IC₅₀ values of 46.6 µM and 1729.8 µM were determined, respectively, in our 5-HT release inhibition assay (Figure 4 and Table 1). In previous studies using rabbit platelets, eugenol was found to be up to 1000 times more potent than elemicin and was found to compare favorably with indomethacin [34]. An additional study found eugenol to be 29 times more potent than aspirin at inhibiting arachidonic acid-induced human platelet aggregation [35]. Based on reports in the literature and the structural characteristics of eugenol methylether and trans-iso-elemicin, which were also isolated in the active fraction, these two compounds were not investigated further in our pharmacological studies [34]. Although, interestingly it has been demonstrated that eugenol methylether has antinociceptive effects on formalin-induced hyperalgesia [36]. However, our interests and discussion here focus on eugenol due to its principal activity in our bioassays.

Studies have demonstrated that eugenol has anti-oxidant, anti-inflammatory, anti-platelet (using rabbit platelets), anti-nociceptive and anti-ulcerogenic effects [37–42]. Furthermore, this drug has also been shown to potentiate GABAₐ transmission [43, 44]. Eugenol is reported to cause inhibition of thromboxane A₂ and B₂ formation, without any influence on the lipoxygenase pathway and appears to be an inhibitor of COX activity similarly to aspirin. In addition, more recent data indicates that this drug inhibits the rise in intracellular Ca²⁺ caused by collagen, adrenalin, ADP and AA [38]. Hence, eugenol appears to have pharmaceutical features that might be considered ideal characteristics of a migraine treatment. In support of these earlier studies [29], our results provide evidence that eugenol does not have significant effects on the primary phase of ADP induced platelet aggregation. Concentrations of eugenol above 61 µM did not inhibit platelet aggregation further (Figure 5) and this result strongly suggests that eugenol selectively targets the second phase of platelet aggregation. This second phase of aggregation involves the “release reaction,” which is dependent on cytosolic increases in Ca²⁺ and the activation of PLC (phospolipase C) and the COX pathway [45], which mediate platelet dense granule secretion. Therefore, 5-HT release could be completely inhibited even when aggregation is not. In line with this, high concentrations of eugenol that inhibited 5-HT release only inhibited the second phase of platelet aggregation (Figures 4 and 5). The modulation of 5-HT release was the principal pathway we were interested in investigating.

Aspirin is a potent anti-inflammatory analgesic, known to cause inhibition of the COX pathway, namely the COX-1 and COX-2 isoforms of this enzyme. In general, aspirin is used to assist in prevention of heart attacks, strokes, arthritis, diabetes and migraines and may also slow the mental decline of old age or provide neuroprotection in a mouse model of Parkinson’s disease [46, 47]. Previous studies have also shown that a single concentration of eugenol, inhibits both COX-1 and COX-2 enzymes [48]. Our results show that eugenol exhibits pharmacological effects that resemble those of aspirin and this data supports previous evidence that eugenol is a COX inhibitor and acts as a potent anti-platelet drug. Interestingly, eugenol is reported to exhibit analgesic properties [38] and has also been used to treat gastrointestinal upsets and chronic diarrhoea and is approved by the Food and Drug Administration of the USA. It is noteworthy that other traditional medicines...
known to modulate 5-HT activity have also been reported as treatments for gastrointestinal tract [49] and allergic disorders [50]. Eugenol is also likely to have potential in the treatment of other diseases related to platelet aggregation, such as, thrombosis, transient ischemia, inflammation, tumor growth and promotion of atherosclerosis [51, 52]. However, there is only limited data relating to its potential use as an anti-headache/migraine or anti-platelet drug [29]. Few side effects have been reported in relation to the use of eugenol. One report on its use as a traditional dental material does however document local irritative and cytotoxic effects along with hypersensitivity reactions when used in contact with soft oral tissues [53].

5. Conclusions

In conclusion, four known compounds (eugenol, elemicin, eugenol methyl ether and trans-isoelemicin) were isolated from the DCM extract of C. ambiguus. Eugenol and elemicin were identified as contributing the principal activity of this extract and demonstrated dose-dependent inhibition of ADP-induced human platelet [14C]5-HT release, with IC_{50} values of 46.6 μM and 1729.8 μM, respectively. Eugenol was also identified as having potent affects on the second-phase of platelet activation. The results of these studies provide evidence identifying specific constituents (principally eugenol) that have anti-platelet activity (Figure 6) that establish the basis of the therapeutic activity and traditional use of C. ambiguus as a non-conventional remedy for headache conditions.

Funding

This research was supported by Griffith University and a Griffith University Postgraduate Scholarship for Kelly Rogers.

Acknowledgments

We are very grateful to Mr Des Nelson, for collection and identification of Cymbopogon ambiguus samples. The authors would also like to thank the Northern Territory Herbarium in Alice Springs for confirmation of identification and incorporation of voucher specimens.
References

[1] P. K. Latz, *Bushfires and Bushtucker. Aboriginal Plant Use in Central Australia*, IAD Press, Alice Springs, NT, Australia, 1995.

[2] E. V. Lassak and T. McCarthy, *Australian Medicinal Plants*, Reed, Kew, Victoria, Australia, 1983.

[3] A. Barr, J. Chapman, N. Smith, and M. Beveridge, *Traditional Bush Medicines. An Aboriginal Pharmacopoeia. Aboriginal Communities of the Northern Territory of Australia*, Greenhouse Publications, Richmond, Victoria, 1988.

[4] S. J. Semple, G. D. Reynolds, M. C. O’Leary, and R. L. P. Flower, “Screening of Australian medicinal plants for antiviral activity,” *Journal of Ethnopharmacology*, vol. 60, no. 2, pp. 163–172, 1998.

[5] E. A. Palombo and S. J. Semple, “Antibacterial activity of traditional Australian medicinal plants,” *Journal of Ethnopharmacology*, vol. 77, no. 2-3, pp. 151–157, 2001.

[6] K. L. Rogers, I. D. Grice, and L. R. Griffiths, “Modulation of *in vitro* platelet 5-HT release by species of *Erythrina* and *Cymbopogon*,” *Life Sciences*, vol. 69, no. 15, pp. 1817–1829, 2001.

[7] G. D’Andrea, K. M. A. Welch, J. M. Riddle, S. Grunfeld, and R. Joseph, “Platelet serotonin metabolism and ultrastructure in migraine,” *Archives of Neurology*, vol. 46, no. 11, pp. 1187–1189, 1989.

[8] G. D’Andrea, A. R. Cananzi, F. Perini et al., “Decreased collagen-induced platelet aggregation and increased platelet arginine levels in migraine: a possible link with the NO pathway,” *Cephalalgia*, vol. 14, no. 5, pp. 352–356, 1994.

[9] S. Evers, F. Quibeldey, K.-H. Grottemeyer, B. Suhr, and I.-W. Husstedt, “Dynamic changes of cognitive habituation and serotonin metabolism during the migraine interval,” *Cephalalgia*, vol. 19, no. 5, pp. 485–491, 1999.

[10] R. Joseph, K. M. A. Welch, S. Grunfeld, S. B. Oster, and G. D’Andrea, “Cytosolic ionized calcium homeostasis in platelets: an abnormal sensitivity to PAF-activation in migraine,” *Headache*, vol. 28, no. 6, pp. 396–402, 1988.

[11] O. Lingjaerde and P. Monstad, “The uptake, storage, and efflux of serotonin in platelets from migraine patients,” *Cephalalgia*, vol. 6, no. 3, pp. 135–139, 1986.

[12] T. G. Pukhal’skaya, “Effects of steroid hormones and antimigraine drugs on serotonin transport in platelets of patients suffering from migraine and in those of healthy subjects,” *Bulletin of Experimental Biology and Medicine*, vol. 115, pp. 679–682, 1993.

[13] M. G. Tozzi-Ciancarelli, G. De Matteis, C. Di Massimo, C. Marini, I. Ciancarelli, and A. Carolei, “Oxidative stress and platelet responsiveness in migraine,” *Cephalalgia*, vol. 17, no. 5, pp. 580–584, 1997.

[14] G. D’Andrea, F. Granella, M. Leone, F. Perini, A. Farruggio, and G. Bussone, “Abnormal platelet trace amine profiles in migraine with and without aura,” *Cephalalgia*, vol. 26, no. 8, pp. 968–972, 2006.

[15] M. D. Ferrari, “Migraine,” *Lancet*, vol. 351, no. 9108, pp. 1043–1051, 1998.

[16] L. Fioroni, G. D’Andrea, M. Alecci, A. Cananzi, and F. Facchinetti, “Platelet serotonin pathway in menstrual migraine,” *Cephalalgia*, vol. 16, no. 6, pp. 427–430, 1996.

[17] R. Hering, V. Glover, K. Pattichis, T. Catarcì, and T. J. Steinier, “SHT in migraine patients with medication-induced headache,” *Cephalalgia*, vol. 13, no. 6, pp. 410–412, 1993.

[18] A. Panconesi, “Serotonin and migraine: a reconsideration of the central theory,” *Journal of Headache and Pain*, vol. 9, no. 5, pp. 267–276, 2008.

[19] E. Hamel, “Serotonin and migraine: biology and clinical implications,” *Cephalalgia*, vol. 27, no. 11, pp. 1295–1300, 2007.

[20] T. J. Schwedt, “Serotonin and migraine: the latest developments,” *Cephalalgia*, vol. 27, no. 11, pp. 1301–1307, 2007.

[21] I. A. Jagoop and D. P. Mikhailidis, “An investigation of the serotonicergic effects of fenfluramine, dexfenfluramine and dexnorfenfluramine using platelets as neuronal models,” *Platelets*, vol. 11, no. 3, pp. 161–165, 2000.

[22] R. Joseph, K. M. Welch, and G. D’Andrea, “Serotonergic hypofunction in migraine: a synthesis of evidence based on platelet dense body dysfunction,” *Cephalalgia*, vol. 9, no. 4, pp. 293–299, 1989.

[23] SDBS, *Integrated Spectral Database System for Organic Compounds*, National Institute of Advanced Industrial Science and Technology, Tsukuba, Ibaraki, Japan, 2001, http://www. aist.go.jp/RIODB/SDBS/menu-e.html.

[24] A. M. Giesbrecht, N. C. Franca, O. R. Gottlieb, and A. I. Da Rocha, “The neolignans of Licaria canella,” *Phytochemistry*, vol. 13, no. 10, pp. 2285–2293, 1974.

[25] R. G. Enquies and M. A. Chavez, “Propenylbenzenes from *Guatteria guameri*,” *Phytochemistry*, vol. 19, no. 9, pp. 2024–2025, 1980.

[26] K. L. Rogers, I. D. Grice, and L. R. Griffiths, “Inhibition of platelet aggregation and 5-HT release by extracts of Australian plants used traditionally as headache treatments,” *European Journal of Pharmaceutical Sciences*, vol. 9, no. 4, pp. 355–363, 2000.

[27] W. A. Groenewegen and S. Heptinstall, “A comparison of the effects of an extract of feverfew and parthenolide, a component of feverfew, on human platelet activity in-vitro,” *Journal of Pharmacy and Pharmacology*, vol. 42, no. 8, pp. 553–557, 1990.

[28] J. S. Bennett, “Novel platelet inhibitors,” *Annual Review of Medicine*, vol. 52, pp. 161–184, 2001.

[29] J. Janssens, G. M. Laekeman, L. A. C. Pieters, J. Totte, A. G. Herman, and A. J. Vlietinck, “Nutmeg oil: identification and quantitation of its most active constituents as inhibitors of platelet aggregation,” *Journal of Ethnopharmacology*, vol. 29, no. 2, pp. 179–188, 1990.

[30] A. C. De Oliveira, L. F. Ribeiro-Pinto, and J. R. Paumgartten, “In-vitro inhibition of CYP2B1 mono oxygenase by beta-mucrine and other monoterpoid compounds,” *Toxicology Letters*, vol. 92, pp. 39–46, 1997.

[31] N. Dudaí, Z. G. Weinberg, O. Larkov, G. Ashbell, and E. Putievsky, “Changes in essential oil during enzyme-assisted ensiling of lemongrass (*Cymbopogon citratus* Stapf.) and lemon eucalyptus (*Eucalyptus citriodora* Hook.),” *Journal of Agricultural and Food Chemistry*, vol. 49, pp. 2262–2266, 2001.

[32] K. C. Srivastava, “Antiplaetelet principles from a food spice clove (*Syzygium aromaticum* L.),” *Prostanoids Leukotrienes and Essential Fatty Acids*, vol. 48, no. 5, pp. 363–372, 1993.

[33] K. C. Srivastava and N. Malhotra, “Acetyl eugenol, a component of oil of cloves (*Syzygium aromaticum* L.) inhibits aggregation and alters arachidonic acid metabolism in human blood platelets,” *Prostanoids Leukotrienes and Essential Fatty Acids*, vol. 42, no. 1, pp. 73–81, 1991.

[34] A. Rasheed, G. M. Laekeman, A. J. Vlietinck, J. Janssens, G. Hatfield, J. Totte et al., “Pharmacological influence of nutmeg and nutmeg constituents on rabbit platelet function,” *Planta Medica*, vol. 50, pp. 222–226, 1984.
[35] R. H. Raghavendra and K. A. Naidu, “Spice active principles as the inhibitors of human platelet aggregation and thromboxane biosynthesis,” Prostaglandins Leukotrienes and Essential Fatty Acids, vol. 81, no. 1, pp. 73–78, 2009.

[36] S. Yano, Y. Suzuki, M. Yuzurihara et al., “Antinociceptive effect of methyleugenol on formalin-induced hyperalgesia in mice,” European Journal of Pharmacology, vol. 553, no. 1–3, pp. 99–103, 2006.

[37] J. N. Sharma, K. C. Srivastava, and E. K. Gan, “Suppressive effects of eugenol and ginger oil on arthritic rats,” Pharmacology, vol. 49, no. 5, pp. 314–318, 1994.

[38] Y.-C. Chen, J.-J. Chen, Y.-L. Chang et al., “A new aristolactam alkaloid and anti-platelet aggregation constituents from Piper taiwanense,” Planta Medica, vol. 70, no. 2, pp. 174–177, 2004.

[39] T. Ohkubo and M. Shibata, “The selective capsaicin antagonist capsazepine abolishes the antinociceptive action of eugenol and guaiacol,” Journal of Dental Research, vol. 76, no. 4, pp. 848–851, 1997.

[40] R. Capasso, L. Pinto, M. L. Vuotto, and G. Di Carlo, “Preventive effect of eugenol on PAF and ethanol-induced gastric mucosal damage,” Fitoterapia, vol. 71, no. 1, pp. S131–S137, 2000.

[41] K.-G. Lee, A. Mitchell, and T. Shibamoto, “Antioxidative activities of aroma extracts isolated from natural plants,” BioFactors, vol. 13, no. 1–4, pp. 173–178, 2000.

[42] J. Szabadics and L. Erdélyi, “Pre- and postsynaptic effects of eugenol and related compounds on Helix pomatia L. neurons,” Acta Physiologica Hungarica, vol. 51, no. 2–4, pp. 265–273, 2000.

[43] G. Bovill, “Mechanisms of actions of opioids and non-steroidal anti-inflammatory drugs,” European Journal of Anaesthesiology, vol. 128, no. 5, supplement, pp. 9–15, 1997.

[44] P. Teismann and B. Ferger, “Inhibition of the cyclooxygenase isoenzymes COX-1 and COX-2 provide neuroprotection in the MPTP-mouse model of Parkinson’s disease,” Synapse, vol. 39, no. 2, pp. 167–174, 2001.

[45] B. Everts, P. Wahrborg, and T. Hedner, “COX-2-specific inhibitors—the emergence of a new class of analgesic and anti-inflammatory drugs,” Clinical Rheumatology, vol. 19, no. 5, pp. 331–343, 2000.

[46] K. Tominaga, T. Kido, M. Ochi, C. Sadakane, A. Mase, H. Okazaki et al., “The traditional Japanese medicine Rikkunshito promotes gastric emptying via the antagonistic action of the 5-HT3 receptor pathway in rats,” Evidence-Based Complementary and Alternative Medicine, vol. 6, pp. 1–8, 2009.

[47] N. Sarrami, M. N. Pemberton, M. H. Thornhill, and E. D. Theaker, “Adverse reactions associated with the use of eugenol in dentistry,” British Dental Journal, vol. 193, no. 5, pp. 257–259, 2002.
Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:
Grice, ID; Rogers, KL; Griffiths, LR

Title:
Isolation of Bioactive Compounds That Relate to the Anti-Platelet Activity of Cymbopogon ambiguus

Date:
2011-01-01

Citation:
Grice, I. D., Rogers, K. L. & Griffiths, L. R. (2011). Isolation of Bioactive Compounds That Relate to the Anti-Platelet Activity of Cymbopogon ambiguus. EVIDENCE-BASED COMPLEMENTARY AND ALTERNATIVE MEDICINE, 2011, pp.1-8. https://doi.org/10.1093/ecam/nep213.

Persistent Link:
http://hdl.handle.net/11343/255519

License:
CC BY