Etavopivat, a Pyruvate Kinase Activator in Red Blood Cells, for the Treatment of Sickle Cell Disease

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ABSTRACT
Etavopivat is an investigational, oral, small molecule activator of erythrocyte pyruvate kinase (PKR) in development for the treatment of sickle cell disease (SCD) and other hemoglobinopathies. PKR activation is proposed to ameliorate the sickling of SCD red blood cells (RBCs) through multiple mechanisms, including reduction of 2,3-diphosphoglycerate (2,3-DPG), which consequently increases hemoglobin (Hb)-oxygen affinity; increased binding of oxygen reduces sickle hemoglobin polymerization and sickling. In addition, PKR activation increases adenosine triphosphate (ATP) produced via glycolytic flux, which helps preserve membrane integrity and RBC deformability. We evaluated the pharmacodynamic response to etavopivat in nonhuman primates (NHPs) and in healthy human subjects and evaluated the effects in RBCs from patients with SCD after ex vivo treatment with etavopivat. A single dose of etavopivat decreased 2,3-DPG in NHPs and healthy subjects. Hb-oxygen affinity was significantly increased in healthy subjects after 24 hours. After daily dosing of etavopivat over 5 consecutive days in NHPs, ATP was increased by 38% from baseline. Etavopivat increased Hb-oxygen affinity and reduced sickling in RBCs collected from patients with SCD with either homozygous hemoglobin S or hemoglobin S and C disease. Collectively, these results demonstrate the ability of etavopivat to decrease 2,3-DPG and increase ATP, resulting in increased Hb-oxygen affinity and improved sickle RBC function. Etavopivat is currently being evaluated in clinical trials for the treatment of SCD.

SIGNIFICANCE STATEMENT
Etavopivat, a small molecule activator of the glycolytic enzyme erythrocyte pyruvate kinase, decreased 2,3-diphosphoglycerate in red blood cells (RBCs) from nonhuman primates and healthy subjects and significantly increased hemoglobin (Hb) oxygen affinity in healthy subjects. Using ex vivo RBCs from donors with sickle cell disease (SCD) (homozygous hemoglobin S or hemoglobin S and C genotype), etavopivat increased Hb-oxygen affinity and reduced sickling under deoxygenation. Etavopivat shows promise as a treatment for SCD that could potentially reduce vaso-occlusion and improve anemia.

Introduction
Sickle cell disease (SCD) is an autosomal recessive blood disorder caused by a single point mutation in the beta-globin gene, resulting in the expression of sickle hemoglobin (Hbs). HbS tends to polymerize when deoxygenated in the low oxygen tension of capillaries or even in arterioles. HbS polymerization alters red blood cell (RBC) morphology and negatively affects its function (Sundd et al., 2019). The two most common SCD genotypes are due to homozygosity for the beta-S mutation (p.Glu6Val) (HbSS) or compound heterozygosity for beta-S and beta-C (p.Glu6Lys) (HbSC) (da Guarda et al., 2020). The formation of cytoplasmic HbS polymers causes the RBCs to adopt a rigid, sickle-like shape that is the defining characteristic of SCD (Kato et al., 2018). Sickled RBCs display abnormal

ABBREVIATIONS: AUC0–24h, area under the concentration time curve from time 0 to 24 hours after dosing; EI, elongation index; HbS, sickle hemoglobin; HbSC, hemoglobin S and C; HbSSc, homozygous hemoglobin S; HU, hydroxyurea; LC-MS/MS, liquid chromatography-tandem mass spectrometry; NHP, nonhuman primate; PD, pharmacodynamics; PK, pharmacokinetics; PKR, erythrocyte pyruvate kinase; pO2, partial pressure of dissolved oxygen; P50, the pO2 at which hemoglobin is 50% saturated with oxygen; PoS, point of sickling; RBC, red blood cell; SCD, sickle cell disease; VOS, vaso-occlusive crisis.
rhological properties, aggregate, and can lead to microcapillary blockage causing tissue hypoxia, which is experienced as a painful vaso-occlusive crisis (VOC). Repeated HbS polymerization and depolymerization with changing oxygen pressure, as well as oxidative damage, alters the RBC membrane, ultimately leading to a significantly shortened RBC lifespan (i.e., hemolysis) (Sundd et al., 2019).

Despite increased reticulocyte production, this decreased RBC lifespan results in anemia. The physiologic metabolic response of RBCs to anemia is to increase 2,3-diphosphoglycerate (2,3-DPG), thereby promoting the release and offloading of oxygen from hemoglobin (Hb) to tissues (Charache et al., 1970). This shift to lower oxygen affinity promotes HbS deoxygenation and consequently polymerization, leading to RBC sickling. Resulting changes in the RBC membrane, such as expression of phosphatidyl serine on the cell surface, lead to altered interaction with other blood cells and the endothelium. Together with the release of proinflammatory RBC contents upon hemolysis, these changes elicit an inflammatory response, including the activation of white blood cells and increased adhesion of sickle RBCs and white blood cells to the endothelium (Telen, 2007), resulting in the sudden, severe, and painful VOC episodes. VOC is the leading diagnosis associated with emergency department visits for patients with SCD (Yusuf et al., 2010); upwards of 40% of such emergency department visits can result in hospital admission (Lanzkron et al., 2010). VOCs also lead to secondary complications, such as acute chest syndrome, that cause end-organ damage and premature death in patients with SCD (Novelli and Gladwin, 2016).

Erythrocyte pyruvate kinase (PKR) is the RBC-expressed isofrom of the key glycolytic enzyme pyruvate kinase. PKR catalyzes the last and rate-limiting step of glycolysis from phosphoenol-pyruvate to pyruvate while generating adenosine phosphoenol-pyruvate to pyruvate while generating adenosine triphosphate (ATP) from adenosine diphosphate. In patients with pyruvate kinase deficiency, decreased PKR activity results in anemia (Zanella et al., 2005), whereas activation of PKR improves RBC survival and Hb levels (Grace et al., 2019). We hypothesized that PKR activation above normal levels would alter several steps of the glycolytic pathway utilizing glucose to generate pyruvate. Increased PKR activity would be expected to decrease production of 2,3-DPG and increase production of ATP within RBCs (Koralkova et al., 2014; Bianchi et al., 2019). PKR is therefore a logical target to activate for treatment of SCD, as a decrease in 2,3-DPG levels would increase the affinity of Hb for oxygen, lowering the potential for HbS polymerization and RBC sickling.

Furthermore, an increase in ATP synthesis can enhance overall RBC function and health. Reduced ATP within RBCs adversely affects ion homeostasis, contributing to cell dehydration (Ortiz et al., 1990; Gallagher, 2017), membrane damage (de Jong et al., 2001; Soupen and Kuypers, 2006; Weiss et al., 2012), and impaired RBC deformability (Clark et al., 1980), all of which contribute to the pathophysiologic features of SCD. By attenuating these adverse effects, an increase in ATP synthesis via PKR activation within sickle RBCs would potentially have the dual beneficial effect of addressing the chronic anemia associated with SCD and reducing the incidence of VOC.

Here, we describe the cellular effects of etavopivat (FT-4202), a potent, selective, orally bioavailable PKR activator (Ericsson et al., 2020). Using oral dosing studies in nonhuman primates (NHPs) and healthy subjects, we demonstrate that etavopivat decreases 2,3-DPG and increases ATP in RBCs. We also present the results of a series of pharmacodynamic (PD) assays after in vivo treatment of healthy RBCs and ex vivo treatment of sickle RBCs with etavopivat, supporting the relevance of these biomarkers when evaluating response to this investigational agent. This work demonstrates the translational value of measurements adopted in preclinical and early clinical development, which are indicative of RBC health. We present target engagement data (single 700-mg dose) from the first-in-human clinical trial that provides a platform of evidence supporting further clinical development of etavopivat for the treatment of SCD.

Materials and Methods

In Vivo Studies

Procedures in Cynomolgus Monkeys. The pharmacokinetics (PK) and PD of etavopivat were assessed in male cynomolgus monkeys. The decision to use male cynomolgus monkeys was based on availability only. There were no PK/PD gender differences observed during preclinical multiple-dose safety studies. Animals (AlphaGenesis Inc., Yemassee, SC) were of Chinese origin, in good health (nonnaive), and housed, cared for, and acclimated to study procedures in compliance with the testing facility Institutional Animal Care and Use Committee Guidelines and Standard Operating Procedure (Biomere, MA). NHP experiments were approved by the Institutional Animal Care and Use Committee in Biomere, MA, and conducted according to the National Research Council’s Guide for the Care and Use of Laboratory Animals (National Research Council, 2011).

A spray-dried dispersion of etavopivat was delivered in a vehicle solution containing 0.5% hydroxypropyl methylcellulose and was administered to monkeys by oral gavage. In a single-dose experiment, monkeys ($n = 4$) received etavopivat at a dose of 50 mg/kg. In a multiple-dose experiment, animals received a dose of 0.5% hydroxypropyl methylcellulose (control); $n = 3$ or a dose of etavopivat at 3, 8, or 22 mg/kg ($n = 4$ per group) once daily on 5 consecutive days. The dose volume for all groups was 5 ml/kg. For the single-dose experiment, whole blood samples (~0.5 ml) for determination of plasma concentrations of etavopivat and whole blood 2,3-DPG and ATP were collected predose, 30 minutes postdose, and 1, 2, 4, 6, 8, 12, and 24 hours postdose. For the multiple-dose experiment, blood samples were collected 1, 6, 12, and 24 hours postdose on day 1 (~0.5 ml) and predose, 30 minutes postdose, and 1, 2, 4, 6, 8, 12, and 24 hours postdose on day 5 (~0.8 ml).

Single-Dose Plasma PK and PD of Etavopivat in Healthy Subjects. Key aspects of the first-in-human study of etavopivat are summarized below. The study was performed in accordance with the Declaration of Helsinki and Good Clinical Practice guidelines and was approved by the relevant institutional review board. Subjects who took part in the study were healthy volunteers aged 18 to 60 years inclusive and provided written informed consent.

The single-ascending dose portion of the study was a phase 1, single-center, randomized, placebo-controlled, double-blind dose escalation study to assess the safety, tolerability, PK, and PD after a single oral dose of etavopivat in healthy subjects (ClinicalTrials.gov identifier: NCT03815695). Placebo and etavopivat were administered orally to subjects after overnight fasting. Successively escalated dose levels of etavopivat at 200, 400, 700, and 1000 mg were evaluated. For each sequential dose cohort, eight subjects were randomly assigned 2:6 to receive a single oral dose of placebo or etavopivat, respectively. Blood samples were collected pre- and post-etavopivat dosing at serial time points for PK/PD assessment. Etavopivat plasma concentrations were measured using a validated liquid chromatography-tandem mass spectrometry (LC-MS/MS) method. The lower and upper limits of quantitation were 1 and 1000 ng/ml, respectively. 2,3-DPG and ATP in whole blood were measured using a separate LC-MS/MS method as described below. Hb-oxygen affinity and values for the partial pressure of dissolved oxygen ($pO_2$) at which Hb is 50% saturated with oxygen ($P_{50}$)
were measured in whole blood by generating oxygen equilibrium curves as described below. Noncompartmental PK analysis of etavopivat was conducted using Phoenix WinNonlin (Certara, Princeton, NJ).

In Vitro Studies

Activation of PKR in Human Red Blood Cells. The activation of wild-type PKR by etavopivat in mature human RBCs ex vivo was evaluated in purified RBCs purchased from Research Blood Components (Watertown, MA). Varying concentrations of etavopivat stock solutions (2.5 μl) were added to 22.5 μl of washed human RBCs (0.75 million) in the range of 2 nM to 30 μM in Roswell Park Memorial Institute Medium (Corning Inc., Corning, NY) and incubated for 3 hours at 37°C. Cells were spun at 1500 rpm for 5 minutes, and the supernatant was removed. Cells were washed twice using 200 μl of PBS and a 5-minute 1500 rpm spin cycle. After the second wash, the RBC pellet was lysed for 30 minutes in 200 μl of assay buffer. After a 5-minute spin at 1500 rpm, lysisate was assayed using a Biovision (Milpitas, CA) Pyruvate Kinase Assay (Catalog # K709-100).

Whole Blood from Patients with SCD. Whole blood samples from female and male patients homozygous for HbS (HbSS) or compound heterozygous for HbS and HbC (HbSC) were collected in vacutainers with K2-EDTA as an anticoagulant, at Cincinnati Children's Hospital, with informed consent and in accordance with an Independent Review Board-approved protocol. Whole blood samples (250 μl) from normal control donors (HbAA) and patients with SCD were washed twice with 1 ml Hanks' Balanced Salt Solution (Thermo Fisher Scientific, Wal- tham, MA) containing 10 mM HEPES, 10 mM MgCl2, and 5.5 mM glucose at pH 7.4 (Buffer 1) and centrifuged at 400 × g for 5 minutes. Supernatant was removed by aspiration. The RBC pellet was resuspended in 1 ml of Buffer 1 and centrifuged for 5 minutes at 800 × g to pack the cells. Supernatant was aspirated and packed RBCs were resuspended to 20% (v/v) (20% hematocrit) with Buffer 1 and incubated for 2 hours at 37°C. Cells were spun at 1500 rpm, 5-minute spin at 1500 rpm, and packed cells were resuspended in 1 ml of Buffer 1 and centrifuged for 5 minutes at 800 × g to pack the cells. Supernatant was aspirated and packed RBCs were resuspended to 20% (v/v) (20% hematocrit) with Buffer 1 and incubated for 4 hours at 37°C with 20 μl etavopivat in dimethyl sulfoxide (DMSO) or equivalent volume of vehicle control (DMSO only) on a VWR Incubating Orbital Shaker 3500 (VWR International, Radnor, PA) at 25rpm. The concentration of 20 μM etavopivat was selected based on preliminary work showing that this concentration consistently decreased P50 in RBCs incubated in vitro with etavopivat for 3 hours at 37°C. Hb-oxygen dissociation curves and oxygen scans were performed as described below to determine P50 and point of sickling (PoS), respectively.

Simultaneous Measurement of 2,3-DPG and ATP in Whole Blood. Whole blood samples of monkeys or humans using LC-MS/MS. The analysis was performed using an API-5500 triple-quadrupole mass spectrometer (AB Sciex, Redwood City, CA) coupled with an Aglient 1200 series high-performance liquid chromatography system (Aglient 1260 Infinity Bin Pump and G3179B Degasser, Agilent, Santa Clara, CA) and a Thermo Scientific CTC-PAL autosampler (Thermo Fisher Scientific). Analyst 1.6.2 (Sciex) and Aria MX software (Thermo Fisher Scientific) were used for instrument control, data acquisition, and data analysis. Calibration standards (25.0–1500 μg/ml), quality controls, and study samples were thawed on wet ice and vortex-mixed for approximately 2 minutes before being pipetted. Because ATP and 2,3-DPG are endogenous molecules present in whole blood at high levels, the low, middle, and high quality control calibration standards were prepared in deionized water (surrogate matrix). Whole blood samples (15 μl) were spiked with stable isotope-labeled internal standard (13C10, 15N5-ATP, and D3-2,3-DPG), processed by protein precipitation extraction, and analyzed using ZIC-pHILIC separation with Turbo Ion Spray MS/MS detection. Negative (M-H)+ ions for ATP and 2,3-DPG and their respective internal standards (13C10, 15N5-ATP, and D3-2,3-DPG) were monitored in multiple reaction monitoring mode. Analyte to internal standard peak area ratios for the standards were used to create a quadratic calibration curve using 1/x2 weighted least-squares regression analysis. For both analytes, the overall accuracy of the method was within ± 10.5% (relative error), and the intra- and interassay precision percent CV (%CV) were less than 7%.

Hb-Oxygen Dissociation Curve and P50 Assessment. The Hb-oxygen equilibrium curves were collected using a TCS HEMOX Analyzer (Guarnone et al., 1995) (TCS Scientific Corp., New Hope, PA) providing the level of oxygenated Hb (expressed as percentage) as a function of PO2 (mm Hg). Whole blood samples from healthy subjects were collected in K2-EDTA tubes 24 hours after etavopivat treatment. For ex vivo treatment, RBCs from patients with SCD were prepared as described above. The whole blood samples or the ex vivo etavopivat-treated RBCs were diluted 50-fold into prewarmed (37°C) N-tris(hydroxymethyl)methyl-2-aminoethanesulfonic acid (TES) sodium salt buffer (Sigma-Aldrich) containing 20-μl bovine serum albumin-20 additive A and 10-μl antifoaming agent-25 per 5 ml of buffer.

Samples were oxygenated using compressed air. Measurement using the HEMOX Analyzer (TCS Scientific) started at an oxygen pressure of 145 mm Hg after replacement of air with nitrogen and was stopped automatically at an oxygen pressure of 1.9 mm Hg. Results were analyzed using the HEMOX Analyzer Software; nonlinear regression analysis was performed to obtain the P50 value.

Oxygen scan and PoS Calculations. Oxygen gradient ektometry (oxygen scan) using a Laser Optical Rotational Red Cell Analyzer (Lorrrca) (RR Mechatronics, Zwaag, The Netherlands) (Rab et al., 2019; Sadaf et al., 2021) was performed to evaluate changes in the deformability of RBCs with or without previous etavopivat treatment as a function of oxygen pressure under steady shear stress. 200 × 106 RBCs from patients with SCD treated ex vivo with DMSO or etavopivat, as described above, were suspended in 5 ml of Oxy Iso (mosmolality 282–286 mOsm/kg, pH 7.35–7.45) at room temperature. The RBC suspension was injected into a rotating wall cylinder maintained at 37°C with a fixed shear stress of 50 Pascal. Deoxygenation and reoxygenation of the RBC suspension were achieved by gradual entry of nitrogen gas and ambient air, respectively. One oxygen scan consisted of approximately 80 measurements of the elongation index (EI) during one round of deoxygenation (1300 seconds) and was followed by reoxygenation (280 seconds). Oxygen saturation in the RBC suspension was calculated every 20 seconds based on quenching of the signal using a lumino- phore oxygen sensor. Ambient air PO2 in the cup of the device started at approximately 150 mm Hg and was gradually lowered to below 20 mm Hg of oxygen. The PoS in oxygen scan is calculated as the Po2 (mm Hg) at which the EI falls below 5% of the maximum EI (EI_max) during deoxygenation and indicates the oxygen pressure at which the polymerization of HbS starts to show effects on RBC deformability.

Statistics

Statistical analysis was performed using Prism, Version 8.4 (GraphPad Software, San Diego, CA). Paired t test or Wilcoxon tests were used as appropriate. A P value of less than 0.05 was considered statistically significant for all tests. All preclinical experiments should be considered exploratory. The sample size for the phase 1 study was considered adequate to evaluate the safety, tolerability, and PK/PD of etavopivat; no formal power calculations were performed.

Results

Etavopivat Activates PKR in Healthy Human RBCs In Vitro. The activation of wild-type PKR by etavopivat in mature human RBCs ex vivo was evaluated in purified RBCs. Wild-type PKR was activated in a concentration-dependent manner. The calculated mean maximum activation was 89% and the half-maximal effective concentration (EC50) was 113 nM.

Etavopivat Modulates 2,3-DPG and ATP in Cynomolgus Monkeys. After the administration of a single 50-mg/kg dose of etavopivat in NHPs, the etavopivat plasma concentration reached its maximal value (Cmax) between 1 and 2 hours postdose, and 2,3-DPG declined to 47% of the baseline concentration between 12 and 24 hours postdose (P = 0.04)
(Fig. 1), which approaches the theoretical maximum calculated by 2,3-DPG PK/PD modeling (Supplemental Text 1). After five consecutive daily doses of etavopivat at lower dose levels (3, 8, and 22 mg/kg), the maximum mean decrease in the concentration of 2,3-DPG occurred 12 hours postdose on day 1 (Fig. 2A), 12 hours post-etavopivat Cmax (Fig. 2B). The mean concentrations of 2,3-DPG observed on day 5 were comparable with those observed on day 1 (Fig. 2A), with mean decreases from baseline of 19% (P = 0.09), 25% (P = 0.02), and 36% (P = 0.002), respectively, 12 hours after the last dose on day 5.

Due to the temporal delay between etavopivat plasma concentration and 2,3-DPG modulation, direct plots between etavopivat PK and 2,3-DPG response are not meaningful. An indirect PK/PD model, previously applied to describe 2,3-DPG PK/PD relationships (Kha et al., 2015) and described in Supplemental Text 1, was used to quantitate the exposure-response relationship. This analysis estimates a calculated 2,3-DPG EC50 of ~18 ng/ml (34 nM) and a 50% 2,3-DPG reduction from baseline as the maximum pharmacodynamic effect in NHPs dosed with etavopivat. Primary PK/PD parameters in NHPs are summarized in Supplemental Table 1.

After a single 50-mg/kg dose, ATP was not significantly elevated from baseline up to 24 hours postdose. To understand the temporal behavior of ATP elevation upon etavopivat administration, ATP was measured in whole blood from NHPs dosed with 3, 8, or 22 mg/kg once daily for 5 consecutive days. Concentrations of ATP were not significantly altered at any dose on day 1 (Fig. 2C); however, by day 5, ATP concentrations were elevated at the 8- and 22-mg/kg doses, with day average increases of 18% (P = 0.0004) and 38% (P = 0.004), respectively, relative to average levels on day 1. As the dynamics of ATP formation and clearance were not adequately captured, this precluded a quantitative assessment of in vivo EC50 for ATP. Etavopivat Cmax and area under the concentration time curve from time 0 to 24 hours after dosing (AUC0–24) after single and repeated escalating doses in NHPs are presented in Supplemental Table 2. Day 5 percentage changes of 2,3-DPG and ATP, along with exposure (day 5 AUC0–24) are summarized in Fig. 3.

![Fig. 1. Relationship between the PK of etavopivat and the PD of 2,3-DPG in nonhuman primates. PK profiles of NHPs (n = 4) administered a single 50-mg/kg dose of etavopivat demonstrated rapid absorption and mean Cmax of 1823 ng/ml etavopivat and an AUC0–24 of 8877 ng·h/ml (AUC0–inf = 9007 ng·h/ml). At this dose level, 2,3-DPG decreased by an average of 47% between 12 and 24 hours postdose vs. baseline (paired t test; P = 0.04). Data are expressed as mean ± S.D. AUC0–inf, area under the concentration time curve from time 0 to infinity.](https://jpet.aspetjournals.org/)

![Fig. 2. Dose-response time course modulation of 2,3-DPG and ATP after 5 consecutive days of once-daily dosing of etavopivat in nonhuman primates (n = 4 per treatment group). For all graphs, data are expressed as mean ± S.D. and analyzed using paired t tests (2,3-DPG, baseline vs. 12 hours post–last dose on day 5; ATP, day 1 vs. day 5). (A) Blood 2,3-DPG profile on day 1 and day 5 of consecutive once-daily dosing of 3, 8, or 22 mg/kg. (B) Plasma PK profile of etavopivat on day 1 and day 5 of consecutive once-daily dosing of 3, 8, or 22 mg/kg. (C) Blood ATP profile on day 1 and day 5 of consecutive once-daily dosing of 3, 8, or 22 mg/kg.](https://jpet.aspetjournals.org/)
Ex Vivo Etavopivat Treatment of RBCs from Patients with SCD Improved Hb-Oxygen Affinity and Reduced PoS. The ability of etavopivat to increase Hb-oxygen affinity in healthy subjects treated with a single dose (Fig. 4A) is indicative of the potential to mitigate the rate and extent of RBC sickling in patients with SCD. We tested this possibility ex vivo, using whole blood from donors with either HbSS or HbSC. The \( P_{50} \) in the patients’ specimens before the in vitro incubation with vehicle control or etavopivat was in the range of 28–34 mm Hg. Despite optimization of the in vitro incubation media, especially of the buffer composition to maintain stable \( CO_2 \) and pH as far as possible, the \( P_{50} \) values of the sickle red cells incubated under control conditions were relatively lower than baseline, as shown in Fig. 4, B and C. Nevertheless, etavopivat-incubated RBCs under the same conditions as vehicle control signified a leftward shift of the Hb-oxygen dissociation curve in RBCs from donors with either HbSS or HbSC (Fig. 4, D and E), suggesting an improved oxygen-carrying capacity of Hb under conditions of decreased oxygen pressure.

We tested the deformability of RBCs across a \( pO_2 \) range, from 150 to <20 mm Hg, using oxygen gradient ektacytometry. PoS was calculated as the \( pO_2 \) at which RBC maximal deformability (\( E_{max} \)) decreased by 5%. The mean (S.D.) decrease in PoS after incubation with etavopivat for 4 hours versus vehicle was 3.53 ± 4.29 mm Hg and 3.91 ± 2.40 mm Hg in HbSS and HbSC donors, respectively. Ex vivo incubation of RBCs from patients with HbSS with etavopivat decreased the PoS (mean ± S.D.) to 28.3 ± 8.09 mm Hg compared with 31.9 ± 7.93 mm Hg in the paired samples treated with vehicle control (\( P = 0.0244 \)) (Fig. 5, A and C). PoS was also decreased in etavopivat-treated (19.3 ± 5.11 mm Hg) versus vehicle-treated (23.2 ± 5.61 mm Hg) RBCs from donors with HbSC (\( P = 0.0313 \)) (Fig. 5, B and D).

Human PKR of Etavopivat. After a single oral dose of 700 mg in healthy subjects (one female, five male), etavopivat was rapidly absorbed and median time to maximum concentration was approximately 0.5 hours. After reaching peak concentrations, mean concentrations of etavopivat declined in a multieponential manner (Fig. 6). The estimated elimination half-life (\( t_{1/2} \)) was approximately 13 hours. Area under the concentration time curve from time 0 extrapolated to infinity (geometric mean ([%CV]) was 6995 (29.4) ng·h/ml. Moderate intersubject variability in plasma etavopivat concentrations was observed.

Etavopivat Decreases 2,3-DPG and Increases Hb-Oxygen Affinity in Healthy Subjects. The PD profile after a single 700-mg dose of etavopivat in healthy subjects was comparable with a single 50-mg/kg dose in NHPs. The maximum inhibition of 2,3-DPG was observed 24 hours postdose, lagging behind the etavopivat \( C_{max} \), which was observed approximately 0.5 hours postdose. In healthy subjects, concentrations of 2,3-DPG were collected for 7 days after a single 700-mg dose; maximal reductions of 2,3-DPG (mean 49%; \( P = 0.0313 \)) were durable, lasted 48 hours postdose, and returned to baseline by day 7 (Fig. 6). Changes in whole blood concentrations of ATP after a single dose were not significant. After a single 700-mg dose in healthy subjects, etavopivat was also shown to increase Hb-oxygen affinity (or decrease \( P_{50} \)) by 4.85 mm Hg at 24 hours, when the greatest decrease in 2,3-DPG was observed (Supplemental Table 3). 2,3-DPG and \( P_{50} \) were correlated positively, providing a PKR activation-mediated PD response to etavopivat that increased Hb-oxygen affinity (Fig. 7).

Discussion

Advancements in disease-modifying therapies are urgently needed for patients with SCD. Through methodical medicinal chemistry efforts, we have identified etavopivat, a potent, selective, orally bioavailable, and novel PKR activator. Here, we present data from preclinical NHP and early-stage clinical human studies to show that etavopivat decreased 2,3-DPG and increased ATP in NHPs after daily dosing. In RBCs from donors with SCD, etavopivat increased Hb-oxygen affinity of RBCs and delayed sickling upon deoxygenation. Decreasing the level of deoxygenation that RBCs can tolerate, before sickleing, may reduce the incidence of VOC in patients with SCD.

Targeting PKR activation to reduce the concentration of 2,3-DPG is based on the rate-limiting role that PKR plays in regulating glycolysis in RBCs. A decrease in 2,3-DPG with PKR activation has been demonstrated in preclinical studies, in healthy subjects, and in patients with PKR deficiency after
treatment (Chubukov et al., 2016; Kung et al., 2017; Yang et al., 2019). Decreases in 2,3-DPG in both NHPs and humans were delayed relative to peak plasma concentrations of etavopivat. More specifically, in NHPs, maximum decreases in whole blood 2,3-DPG were observed between 12 and 24 hours postdose, sometime after the mean C_{max} was achieved, typically 1 hour postdose. A comparable observation was made after a single 700-mg dose of etavopivat was durable for 48 hours before slowly returning to baseline by day 7. After a single dose, maximum decreases in 2,3-DPG in NHPs dosed with 50 mg/kg and healthy human subjects treated with 700 mg were 47% and 49%, respectively, consistent with comparable unbound exposure at those dose levels (2342 and 2099 ng·h/ml for NHPs and humans, respectively). Mitigating 2,3-DPG has been proposed as a means to reduce HbS polymerization in patients with SCD (Eaton and Bunn, 2017). Basal concentrations of 2,3-DPG are higher in patients with SCD compared with healthy controls (Charache et al., 1970; Castro, 1980;
Ould Amar et al., 1996), and depletion of 2,3-DPG has been shown to reduce the tendency for RBC sickling in patients with SCD (Poillon et al., 1995). 2,3-DPG strongly influences the binding of oxygen to Hb; by binding within the central cavity of the Hb tetramer, 2,3-DPG causes allosteric changes and reduces the molecule’s affinity to oxygen (MacDonald, 1977). Increased Hb-oxygen affinity (decreased P50) with a mean delta P50 of 4.85 mm Hg was observed in healthy subjects after the administration of a single 700-mg dose of etavopivat. P50 and 2,3-DPG in blood were strongly correlated before and 24 hours after a single dose of etavopivat. The 2,3-DPG-mediated effect provides a PD-based rationale for the increased Hb-oxygen affinity after treatment with etavopivat. Furthermore, ex vivo experiments using RBCs from donors with HbSS or HbSC showed decreases in P50, indicating increased Hb-oxygen affinity in the presence of etavopivat. Since higher concentrations of 2,3-DPG are associated with increases in deoxyhemoglobin, therapeutic

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**Fig. 5.** Etavopivat reduced the dissolved oxygen pressure at which HbSS or HbSC RBCs initiate sickling. Isolated RBCs from donors with either HbSS or HbSC genotype were treated ex vivo with either 20 μM etavopivat or DMSO (vehicle) for 4 hours at 37°C. Curves for RBC deformability as a function of dissolved oxygen pressure (oxygenscan) were constructed using Lorrrca. The PoS at which the deformation of RBCs reached 5% from baseline was calculated and plotted. Treatment with etavopivat significantly decreased the PoS in RBCs from both HbSS (n = 11) (A) and HbSC (n = 6) (B) donors. Representative oxygenscans for RBCs from an HbSS donor (C) and an HbSC donor (D) are shown. Data were analyzed using the nonparametric Wilcoxon matched-pairs signed rank test. *P < 0.05.
approaches that increase Hb-oxygen affinity and consequently reduce deoxygenation of HbS are expected to decrease HbS polymerization, reduce sickling, and attenuate the clinical consequences of SCD.

In addition to the reduction in 2,3-DPG and associated improvement in Hb-oxygen affinity, the increased glycolytic flux induced by PKR activation is expected to increase RBC function and health by increasing ATP and reducing oxidative stress. Although no meaningful elevation in ATP was observed after a single dose in monkeys or healthy subjects, ATP concentrations were elevated by as much as 38% ($P = 0.004$) relative to day 1 in NHPs after 5 consecutive days of once-daily doses of etavopivat. In a recent study in Berkeley sickle cell mice, ATP levels were increased ($P < 0.01$) after 2 weeks of treatment with etavopivat, corresponding with etavopivat plasma levels of $7702 \pm 796$ ng/ml (Shrestha et al., 2021). ATP plays an essential role in maintenance of RBC membrane-cytoskeletal integrity (Betz et al., 2009), membrane repair via activation of flipase to reinternalize phosphatidylserine to the inner leaflet of the RBC membrane (Soupene and Kuypers, 2006; Weiss et al., 2012), cell hydration (Gallagher, 2017), maintenance of ionic ($\text{Mg}^{2+}$ and $\text{Ca}^{2+}$) gradients (Ortiz et al., 1990; Raftos et al., 1999; Rivera et al., 2005), RBC deformability (Weed et al., 1969; McMahon, 2019), and protection from oxidative damage (Banerjee and Kuypers, 2004). Therefore, increasing ATP concentrations is likely to have broad beneficial effects for patients with SCD. Moreover, in the current study, ex vivo treatment with etavopivat significantly improved the PoS in the RBCs from donors with the HbSS genotype, likely due to a reduction in 2,3-DPG. These dual effects of decreased 2,3-DPG and increased ATP suggest that etavopivat has the potential to reduce RBC sickling, hemolysis, anemia, and consequently VOC.

Pharmacologic interventions in clinical practice or development for the treatment of SCD include approaches to increase fetal Hb or Hb oxygenation to decrease Hb polymerization and to decrease vascular adhesion. The current standard of care for most patients with SCD is hydroxyurea (HU), which reduces sickling by increasing fetal Hb in RBCs and reduces painful episodes of VOC by ~50%; however, the effectiveness of HU can be confounded by poor patient compliance and discontinuation rates can be high (Brandow and Panepinto, 2010; Agrawal et al., 2014; Shah et al., 2019). In addition, although HU is broadly efficacious, response to treatment can be variable and a proportion of patients (10%–20%) are nonresponsive (Steinberg et al., 1997; Ma et al., 2007; Steinberg, 2008). Furthermore, HU is a myelosuppressive agent that may cause neutropenia and thrombocytopenia and thus requires routine blood monitoring for side effects (Agrawal et al., 2014). Although HU has been used to treat SCD for over 30 years, there remains a need for improved disease management, more pharmacological options, and the possibility of combination therapies to target different aspects of SCD pathology. Crizanlizumab was recently approved to reduce the frequency of VOC in adults and pediatric patients aged 16 years and older with SCD. However, no improvement in hemolysis, anemia, or markers of inflammation have been observed (Ataga et al., 2017; Blair, 2020a). These data suggest that although targeting endothelial adhesion may...
decrease the incidence and severity of VOC, these agents are unlikely to address the anemia or chronic tissue damage associated with SCD. Drugs that increase Hb-oxygen affinity, such as the recently approved agent voxelotor, directly interfere with HbS polymerization by maintaining a higher proportion of Hb in an oxygenated state (Vichinsky et al., 2019; Blair, 2020b). Although the clinical safety profile for voxelotor to date has been positive, there is a recognized risk that excessive oxygen affinity could cause hypoxia by preventing release of oxygen into tissues, for which routine neurovascular monitoring has been recommended (Hebbel and Hedlund, 2018). Activation of PKR follows a physiologic pathway that increases Hb-oxygen affinity by decreasing 2,3-DPG and is likely to counteract the pathophysiology of SCD while enabling oxygen release in hypoxic tissues. As a result of the combination of decreased 2,3-DPG and increased ATP, it is proposed that targeting PKR activation may have a broad and significant impact on SCD, both in HbSS and HbSC, by improving the membrane integrity of RBCs.

In conclusion, by virtue of its ability to promote activity in the glycolytic pathway, etavopivat is hypothesized to have unique, beneficial, disease-modifying effects on SCD. As the enzyme that catalyzes the last step of glycolysis, PKR underpins reactions that directly impact the metabolic health and primary functions of RBCs. Etavopivat has two key effects: first, it decreases 2,3-DPG, which can reduce HbS polymerization and potentially attenuate clinical sequelae of vaso-occlusion in SCD; and second, it increases ATP, which provides metabolic resources to support membrane integrity and protect against the loss of RBC deformability and potentially to mitigate hemolysis in SCD. The PD response to etavopivat in relation to PKR was demonstrated preclinically and confirmed in the first-in-human trial, providing valuable evidence of target modulation, which supports the rationale for subsequent clinical trials. Furthermore, the preliminary safety profile of etavopivat supports its advancement to additional clinical trials to determine the risk/benefit profile of PKR activation in SCD. Clinical trials of etavopivat in patients with SCD are currently in progress.

Authorship Contributions

Participated in research design: Schroeder, Fulzele, Forsyth, Ribadenira, Guichard, Wilker, Marshall, Drake, Konstantinidis, Seu, Kalifa.

Conducted experiments: Ribadenira, Fessler, Konstantinidis, Seu. Contributed new reagents or analytic tools: Konstantinidis, Seu.

Performed data analysis: Schroeder, Fulzele, Forsyth, Ribadenira, Guichard, Fessler, Konstantinidis, Seu, Kalifa.

Wrote or contributed to the writing of the manuscript: Schroeder, Fulzele, Forsyth, Ribadenira, Guichard, Wilker, Marshall, Drake, Fessler, Konstantinidis, Seu, Kalifa.

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References

Agrawal RK, Patel RK, Shah V, Nainiwal L, and Trivedi B (2014) Hydroxyures in sickle cell disease: drug review. Indian J Hematol Blood Transfus 30:91–96.

Ataga KI, Kutlar A, Kanter J, Liles D, Cancado R, Friedriech J, Guthrie TH, Knight-Madden J, Alvarez OA, Gordeuk VR, et al. (2017) Crizanlizumab for the prevention of pain crises in sickle cell disease. N Engl J Med 376:429–439.

Bhanejee T and Kuyper FA (2004) Reactive oxygen species and phosphatidylserine externalization in murine sickle red cells. Br J Haematol 124:391–402.

Betz T, Lenz M, Joanny JE, and Sykes C (2009) ATP-dependent mechanisms of red blood cells. Proc Natl Acad Sci USA 106:15320–15325.

Bianchi P, Ferro E, Glader B, Kanno H, Agarwal A, Barcellini W, Eber S, Hoyer JD, Kuster EM, Maai TM, et al. (2020) The European Reference Network in Rare Hematological Diseases (2019) Addressing the diagnostic gaps in pyruvate kinase deficiency: consensus recommendations on the diagnosis of pyruvate kinase deficiency. Am J Hematol 94:149–161.

Blair HA (2020a) Crizanlizumab: first approval. Drugs 80:79–84.

Blair HA (2020b) Voxelotor: first approval. Drugs 80:209–215.

Brandon AM and Panepinto JA (2010) Hydroxures in sickle cell disease: the battle for new prescription rates, poor patient compliance and fears of toxicities. Expert Rev Hematol 3:255–260.

Castronuovo E (1973) Vascular occlusion in sickle cell disease: a physiologic pathway that increases Hb-oxygen affinity. J Clin Invest 52:1039–1046.

Eaton WA and Bunn HF (2017) Treating sickle cell disease by targeting HbS polymerization. Blood 129:2719–2726.

Eriksen A, Green N, Gustafson G, Lancia Jr DR, Marshall G, Mitchell L, Richard D, Wang S (2020) inventors, FORMA Therapeutics, Inc., assignee. Activating pyruvate kinase R. U.S. patent 10,675,274. 2020-Jun-9.

Gallagher PG (2017) Disorders of erythrocyte hydration. Blood 130:2699–2708.

Grace RF, Mark Layton D, and Barcellini W (2019) How we manage patients with pyruvate kinase deficiency. Br J Haematol 184:721–731.

Guarneri C, Centenara E, and Barosi G (1995) Performance characteristics of Hemox/Analyzer for assessment of the hemoglobin dissociation curve. Haematologica 80:430–438.

Hebbel RP and Hedlund BE (2018) Sickle hemoglobin oxygen-affinity-shifting strategies have unequal cerebrovascular risks. Am J Hematol 93:321–325.

Kato GJ, Fiel FB, Reid CD, Gaston MH, Ohene-Frempong K, Krishnamurti L, Smith WR, Panepinto JA, Weatherall DJ, Costa FP, et al. (2018) Sickle cell disease. Nat Rev Dis Primers 4:18010.

Kla H, Cohen M, Chen Y, Kim H, Silver B, Agresta S, Merica E, Kung C, Kosinski P, Silverman L, et al. (2015) Population pharmacokinetics and pharmacodynamics of AG-348 in healthy human volunteers guide dose selection for the treatment of pyruvate kinase deficiency. Br J Clin Pharmacol 80:274–280.

Kla H, Cohen M, Chen Y, Kim H, Silver B, Agresta S, Merica E, Kung C, Kosinski P, Silverman L, et al. (2015) Population pharmacokinetics and pharmacodynamics of AG-348 in healthy human volunteers guide dose selection for the treatment of pyruvate kinase deficiency. Br J Pharmacol 172:1249–1263.

Koralpova P, van Solinge WW, and van Wijk DR (2014) Rare hereditary red blood cell enzyme deficiencies associated with hemolytic anemia – pathophysiology, clinical aspects, and laboratory diagnosis. Int J Lab Hematol 36:388–397.

Kung C, Hixon J, Kosinski PA, Chianetta G, Histen G, Chen Y, Hill C, Gross S, Si Y, Johnson K, et al. (2017) AG-348 enhances pyruvate kinase activity in red blood cells from patients with pyruvate kinase deficiency. Blood 130:1347–1356.

Lanzkron S, Carroll CP, and Haywood Jr C (2010) The burden of emergency department use for sickle-cell disease: an analysis of the national emergency department sample database. Am J Hematol 85:787–796.

Ma Q, Wyszyński DF, Farrell JJ, Kutlar A, Farrer LA, Baldwin CT, and Steinberg MH (2007) Fetal hemoglobin in sickle cell anemia: genetic determinants of response to hydroxures. Pharmacogenomics 8:386–394.

MacDonald R (1977) Red cell 2,3-diphosphoglycerate and oxygen affinity. Eur J Biochem 86:703–709.

McMahon TJ (2019) Red blood cell deformability, vasoactive mediators, and adhesion. Front Physiol 10:1417.

National Research Council (2011) Guide for the Care and Use of Laboratory Animals, 8th ed, National Academies Press, Washington, D.C.

Novelli EM and Gladwin MT (2016) Crises in sickle cell disease. Chest 149:1082–1093.

Oritz OE, Lew VL, and Bozkchin RM (1990) Deoxygenation permeabilizes sickle cell anemia red cells to magnesium and reverses its gradient in the dense cells. J Clin Invest 87:211–226.

Ould Amar AK, Kerob-Lauchet B, Robert P, Leconte C, Maier H, Bera O, Plumelle Y, Hyronimus JC, and Caspere JC (1996) Assessment of qualitative functional parame ters of stored red blood cells from donors with sickle cell trait (AS) or with heterozygote (AC) status. Br J Haematol 92:11–16.

Poillon WN, Kim BC, Labotka RJ, Hicks CU, and Kark JA (1995) Rapid and reproducible method for 2,3-diphosphoglycerate depletion. Blood 85:3289–3296.

Rah MAE, van Oorschot BA, Bo J, Merkx TH, van Wesel ACW, Abdulmalik O, Saffi T, Verdija IA, Houwig ME, Cnossen MH, et al. (2019) Rapid and reproducible characterization of sickling during automated deoxygenation in sickle cell disease patients. Am J Hematol 94:575–584.

Rah JR, Lew VL, and Packer DW (1999) Refinement and evaluation of a model of Mg2+ buffering in human red cells. Eur J Biochem 263:635–645.

Rivera A, Ferreira A, Bertioli D, Romero-JR, and Brugna X (2005) Abnormal regulation of Mg2+ transport via Na/Mg exchanger in sickle erythrocytes. Blood 106:382–386.

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