A novel formulation of chitosan nanoparticles functionalized with titanium dioxide nanoparticles

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ABSTRACT

Herein, chitosan nanoparticles (CS-NPs) were prepared and functionalized chemically with titanium dioxide nanoparticles (TiO$_2$-NPs) to allow on-demand degradation of CS-NPs, using ultraviolet (UV) irradiation as a trigger. This is expected to allow drug release depending on patients’ needs or physiological circumstances. Eleven formulations were arranged and their particle size, charge, and polydispersity were determined. The effect of CS-NPs size and the amount of TiO$_2$-NPs, on the system collapse, was studied accordingly. Moreover, the collapse of these systems was examined using a fluorescence microscope after loading CS-NPs with Rhodamine. The formulations showed high monodispersity and had sizes ranged between 170 and 440 nm and charges ranged between +5 and +34 mV. Scanning electron microscope, Fourier-transform infrared spectroscopy, and X-ray diffraction proved the chemical deposition of TiO$_2$-NPs on CS-NPs. The dye test showed that there are two factors that oppose each other and affected the deposition of TiO$_2$-NPs on CS-NPs, the size of CS-NPs, and the amount of TiO$_2$-NPs used. In addition, the dye test showed that the deposition of TiO$_2$-NPs is a saturated process that relies on the amount of TiO$_2$-NPs used initially. Finally, the intensity of Rhodamine released from these systems after illumination with UV light was related to the amount of TiO$_2$-NPs deposited on CS-NPs. In conclusion, functionalization of CS-NPs with TiO$_2$-NPs can be controlled and used to rupture CS-NPs on demand by illumination with UV light.

Key words: Chitosan nanoparticles, Rhodamine, titanium dioxide nanoparticles, ultraviolet irradiation

INTRODUCTION

Polymeric nanoparticles (PNPs) are widely used in many fields today, including the pharmaceutical industry and drug delivery. PNPs have numerous advantages to be used. First, they use lower drug concentrations, and therefore, less frequent doses are needed. Second, the side effects related to drugs loaded in PNPs are avoided or reduced. Third, they enhance the diffusion through biological membranes and the penetration of drugs into the cells. Fourth, they enhance drug’s stability and prolong drugs’ activity in vivo and in vitro. Fifth, several methods and polymers are available to prepare biocompatible nanoparticles. Finally, they have a huge potential in the targeted distribution of drugs and biological molecules. This targeting allows the selective transport of any

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therapeutic agent to its site of action independently on the method of administration.[6]

Recently, numerous polymers were used to formulate nanoparticles. Interestingly, chitosan (CS) is one of the most used polymers to formulate nanoparticles. CS is a natural, linear polysaccharide, biocompatible, biodegradable, nontoxic, and bioadhesive polymer. CS nanoparticles (CS-NPs) have been used as a colloidal drug carrier to target and control the drug delivery to specific sites in the body.[7,8] It is used for gene and vaccine delivery and in cancer therapy.[9] CS-NPs are prepared by different methods, where the method used in the preparation of NPs, parameters, and properties of the starting materials affect the prepared NP physicochemical properties and the drug release profile.[10]

One of the important drawbacks of PNPs, including CS-NPs, is the drug release at a predetermined rate regardless of the patient’s needs or the disease physiological circumstances. A system that triggers drug delivery may allow controlling the therapeutic effect depending on time. This on-demand drug release from nanoparticles is very important in chemotherapy, where it maximizes tumor killing and minimizes metastatic spread.[7]

In general, the drug release from PNPs depends on the diffusion through the polymers or on the degradation of the polymeric chain. To allow on-demand drug release from PNPs, these PNPs may be functionalized with materials that respond to specific triggers, such as pH, redox, proteins, temperature, light, or magnetic field. Ultraviolet (UV) light is an important trigger that is used in preparing photo-controlled release systems.[8,9] In such formulations, PNPs are noncovalently or covalently assembled with a material that is photosensitive such as TiO$_2$-NPs. Adding TiO$_2$-NPs to CS-NPs may allow on-demand drug release by photolytic degradation. In previous research, CS/PVA blend was applied as a nanoreactor for Ag and Au nanoparticles and showed promising anticancer applications. Other researchers find that triggering hydrophilic polymers such as CS could control drug release in response to UV irradiation.[10]

TiO$_2$-NPs are inorganic chemicals that are widely used in cosmetics, pollution treatment, food preservation, pharmaceutical, and painting fields. They are also used in biomaterials due to their high stability, antimicrobial, and anticorrosive properties. TiO$_2$-NPs have unique photocatalytic properties that guided many research to be used as disinfectants, biological sensors, antibiotics, and tumor-cell killing agents.[11]

In this study, the chemical functionalization of CS-NPs with TiO$_2$-NPs was studied. CS-NPs were fabricated using TPP as a crosslinker. Subsequently, the nanoparticles were functionalized chemically with TiO$_2$-NPs. The mean particle size (PS), polydispersity index (PDI), charge (ZP), and physical morphology of CS-NPs before and after functionalization were characterized. The deposition of TiO$_2$-NPs on CS-NPs was explored and measured using Fourier-transform infrared spectroscopy (FTIR), X-ray diffraction (XRD), scanning electron microscope (SEM), and dye test. Finally, functionalized CS-NPs were loaded with Rhodamine and the release of Rhodamine after triggering with UV light was studied.

**MATERIALS AND METHODS**

**Materials**

Sodium tripolyphosphate (TPP) was purchased from AZchem (China). CS (50–190kDa, 75%DDA), titanium (IV) oxide (TiO$_2$-NPs) (99.5%,<100 nm), and Rhodamine B (95%) were purchased from Sigma-Aldrich (St. Louis, MO). All chemicals used were of analytical grade.

**Preparation and functionalization of chitosan nanoparticles**

CS-NPs were prepared according to the ionic gelation method, and the experimental conditions were chosen according to our previous findings to control the sizes of the NPs.[10] Two solutions were prepared: CS (0.5 mg/mL) in 1% acetic acid solution and TPP solution in water. TPP was added to the CS solution. Part of these NPs was purified by placing the dispersion into a dialysis bag with a cutoff of 12–14 KD. Consequently, CS-NPs were freeze-dried and stored in a tightly closed container in the fridge.

The other portion was functionalized by adding TiO$_2$-NPs. TiO$_2$-NPs were dispersed in 1% acetic acid solution and added to the CS-NPs dispersion at 25s by a syringe pump. The flow rate was 2.5 ml/min and the speed of stirring of 700 rpm. The final mixture was purified and dried. To study the effect of the size of the NPs on the functionalization, CS-NPs with sizes around 200, 250, and 400 nm were prepared. The different parameters applied are described in Table 1.

**Characterization of chitosan nanoparticles before and after functionalization**

The PS, PDI, and ZP of CS-NPs before and after functionalization with TiO$_2$-NPs were determined using a Zetasizer Nano ZS90 (Malvern, UK). SEM (Thermo scientific, Germany) was used to study the morphologies of all formulations after being coated with carbon film.

Shimadzu IR Spectrophotometer (Shimadzu, Kyoto, Japan) and Ultima IV X-Ray Diffractometer (Rigaku, Japan) were used to study the formulations.

**Dye test**

Functionalized CS-NPs were mixed with 50 mg/L Direct Blue 78 (DB78) (50 mg/L) at pH=2 under magnetic stirring
at 200 rpm for 60 min. The changes in the absorbance at $\lambda_{\text{max}}=600$ nm of solution samples were determined at predetermined intervals. The percentage absorption reduction was calculated as follows:

\[
\%\text{absorption reduction} = \frac{\text{Absorption before adding the NPs} - \text{Absorption after adding the NPs}}{\text{Absorption before adding the NPs}} \times 100
\]

**Rhodamine thin film fluorescence**

The formulations were loaded with Rhodamine to study the rupture of the systems upon exposure to UV light. Rhodamine was dissolved in the CS solution in the preparation step and the particles were characterized. A drop of a solution of ethanol/water (50:50) was added to each formulation on a slide and covered with a coverslip and analyzed by fluorescence microscope (Motic AE31E, USA). The formulations were illuminated by UV light at $\lambda=370$ nm. The fluorescence of Rhodamine was detected after 30 min of illumination at $\lambda=520$ nm. Fluorescent intensities of each sample were normalized and evaluated using ImageJ software.

**RESULTS AND DISCUSSION**

**Characterization of chitosan-TiO$_2$ nanoparticles**

The conditions applied in this work to prepare CS-NPs were chosen depending on our previous work to get three different sizes of CS-NPs. The targeted sizes were around 200, 250, and 400 nm. The ratio of CS to TPP and the rate of adding TPP to CS were controlled to realize the sizes. The PS of all formulations is summarized in Table 2.

F$_0$, F$_1$, and F$_8$ were functionalized using different amounts of TiO$_2$-NPs. The effect of the amount of TiO$_2$-NPs used on the NP properties is shown in Table 2. Unfunctionalized CS-NPs have smaller sizes in comparison to functionalized ones, which may prove the accumulation of TiO$_2$-NPs on the surface of CS-NPs. Further, when the amount of TiO$_2$-NPs used in the coating increased from 3:1 to 1:1, a huge increase in the sizes can be noticed, while increasing the amount of TiO$_2$-NPs further from 1:1 to 1:3 has a smaller effect on the particles’ size. This behavior is observed in the three groups of sizes prepared. This may indicate saturation of the sites of interaction between CS-NPs and TiO$_2$-NPs at certain concentrations. The PS increased after loading CS-NPs with Rhodamine in F$_1$, F$_4$, and F$_8$ and the EE was around 35%, as shown in Table 3. All systems prepared carried positive charges, but it is obvious that functionalization decreases the charges.

SEM confirmed that CS-NPs prepared in this study are spherical. The coated CS-NPs showed small particles on their surfaces, which may be related to the precipitated TiO$_2$-NPs. Figure 1 shows a representative SEM image of CS-NPs before and after functionalization (F$_0$ and F$_1$).

**Table 1: Weight ratios and parameters used in preparing chitosan nanoparticles**

| Flow rate (ml/min) | CS: TPP | CS: TiO$_2$-NPs |
|-------------------|---------|----------------|
| 0.25              | 1:2     | F$_0$          |
| 0.25              | 1:2     | F$_1$          |
| 0.25              | 1:2     | F$_2$          |
| 0.25              | 1:2     | F$_3$          |
| 0.25              | 2:1     | F$_4$          |
| 0.25              | 2:1     | F$_5$          |
| 0.25              | 2:1     | F$_6$          |
| 0.25              | 2:1     | F$_7$          |
| 0.25              | 1:7.5   | F$_8$          |
| 0.25              | 1:7.5   | F$_9$          |
| 0.25              | 1:7.5   | F$_{10}$       |
| 0.25              | 1:7.5   | F$_{11}$       |

PS: Particle size, PDI: Polydispersity index

**Table 2: Particle size (nm), polydispersity index, and charge (mV) for the formulations prepared (mean±standard deviation, n=3)**

| PS       | PDI    | ZP     |
|----------|--------|--------|
| F$_0$    | 170.67±1.35 | 0.451±0.021 | 19.54±0.56 |
| F$_1$    | 220.25±3.41 | 0.513±0.085 | 9.62±0.44  |
| F$_2$    | 395.23±4.00 | 0.373±0.111 | 5.97±0.86  |
| F$_3$    | 376.43±8.41 | 0.411±0.033 | 6.61±1.01  |
| F$_4$    | 234.37±2.13 | 0.521±0.005 | 26.33±1.04 |
| F$_5$    | 290.37±3.00 | 0.379±0.028 | 15.50±1.21 |
| F$_6$    | 439.33±5.39 | 0.408±0.060 | 18.33±0.52 |
| F$_7$    | 442.80±5.99 | 0.714±0.303 | 16.72±0.33 |
| F$_8$    | 391.17±2.47 | 0.618±0.0615| 34.04±0.88 |
| F$_9$    | 344.93±3.25 | 0.547±0.004 | 8.71±0.34  |
| F$_{10}$ | 408.43±10.21| 0.392±0.010 | 11.63±0.11 |
| F$_{11}$ | 420.56±3.70 | 0.481±0.002 | 7.46±0.18  |

PS: Particle size, PDI: Polydispersity index

**Table 3: Particle size (nm), polydispersity index, charge (mV), and EE % of the chitosan nanoparticles loaded with Rhodamine before functionalization (mean±standard deviation, n=3)**

| EE (%) | PS       | PDI    | ZP     |
|--------|----------|--------|--------|
| 37.81  | F$_0$    | 175.12±2.50 | 0.321±0.101 | 18.70±0.55 |
| 35.29  | F$_4$    | 241.41±1.98 | 0.494±0.113 | 24.47±0.75 |
| 33.76  | F$_{11}$ | 412.05±4.16 | 0.700±0.0835| 31.07±1.07 |

PS: Particle size, PDI: Polydispersity index, EE: Entrapment Efficiency

FTIR of CS, TiO$_2$-NPs, CS-NPs, and functionalized CS-NPs is demonstrated in Figure 2. The spectrum of CS was compared to that of CS-NPs to confirm the cross-linking in CS-NPs. In CS spectrum, a characteristic band related to NH$_2$ and OH groups stretching was observed at 3447 cm$^{-1}$. This band gets shallower in CS-NPs. Further, a new double peak at 2360 cm$^{-1}$ related to NH$_2$ in CS-NPs spectrum appeared.[13]

The spectrums of TiO$_2$-NPs, CS-NPs, and functionalized CS-NPs showed the following differences: for TiO$_2$-NPs,
the characteristic band at 3500 cm$^{-1}$ that corresponds to OH stretching gets shallower in the functionalized CS-NPs. The second peak in TiO$_2$-NPs around 1630 cm$^{-1}$ that corresponds to bending modes of water Ti-OH, disappeared in the functionalized NPs. In addition, the peak at 2368 cm$^{-1}$ in CS-NPs disappeared in the spectrum of functionalized NPs. These differences in the spectrums of TiO$_2$-NPs, CS-NPs, and the functionalized NPs give a clear proof of the interaction between TiO$_2$-NPs and CS-NPs.$^{[14]}$

XRD was carried out for CS, CS-NPs, TiO$_2$-NPs, and functionalized CS-NPs to explore any interactions. Figure 3 shows that CS exhibited an amorphous structure with one large broad peak. CS-NPs showed a spectrum without any peaks that is somewhat different from the spectrum of CS, which reveals the formation of CS-NPs. The spectrum of TiO$_2$-NPs showed a crystalline behavior as indicated by the sharp peaks. When the spectrums of TiO$_2$-NPs, CS-NPs, and the functionalized CS-NPs are compared, it can be noticed that the functionalized CS-NPs spectrum is similar to TiO$_2$-NPs. This however indicates that TiO$_2$-NPs precipitated on the surface of CS-NPs.$^{[15]}$

**Dye test**

Since the dye adsorption is a mass transfer process, then the higher the reduction of the absorption indicates the higher amount of TiO$_2$-NPs available to interact with the dye. This indirectly indicates the higher deposition of TiO$_2$-NPs on CS-NPs. The percentage reduction in the dye absorbance was measured and referred to as dye removal percentage and the results are shown in Figure 4.$^{[12]}$

Adsorption is affected by many factors including temperature, pH, size, surface area, and the amounts of the adsorbent and adsorbate. Herein, the effect of the particles size and the amount of TiO$_2$-NPs used was studied.

![Figure 2: Fourier-transform infrared spectroscopy of chitosan, chitosan nanoparticles, titanium dioxide nanoparticles, and functionalized chitosan nanoparticles](image)

![Figure 3: X-ray diffraction of chitosan, chitosan nanoparticles, titanium dioxide-nanoparticles, and functionalized chitosan nanoparticles](image)

![Figure 4: The %dye removal by functionalized chitosan nanoparticles versus time measured at 600 nm (mean ± % relative standard deviation, n = 3)](image)
First, it is obvious that as the size of the NPs increases the dye removal decrease (adsorption decrease). This behavior can be observed when F5, F2 and F11 are compared, F2, F6 and F10 are compared, and when F1, F4, and F6 are compared.

Formulations prepared using the highest amount of TiO2-NPs showed the highest dye removal (1:3). For the ratios 1:1 and 3:1, the dye removal from the ratio 3:1 was higher than that of 1:1. For example, F6 showed a high %dye removal in comparison to F2. Furthermore, F5 showed a high %dye removal in comparison to F6. In addition, F9 showed a high %dye removal in comparison to F10. This may be due to the lower sizes recorded for F5, F6, and F9 that were prepared using lower amounts of TiO2-NPs. These results indicate that the effect of the two factors, the size of CS-NPs and the amount of TiO2-NPs, are controverting each other. Increasing the particles sizes decreases the surface area, which is expected to decrease the adsorption, while increasing the amount of TiO2-NPs used is expected to increase the sites available for dye adsorption.

Rhodamine thin film fluorescence

Figure 5a shows the Rhodamine release from F5 after illustration with UV light for 30 min. Figure 5b shows the intensity of Rhodamine released from the 11 formulations prepared. From these optical micrographs, it is clear that the lower intensities of Rhodamine were recorded for F5, F6, and F9 that were prepared using lower amounts of TiO2-NPs. These results indicate that the effect of the two factors, the size of CS-NPs and the amount of TiO2-NPs, are controverting each other. Increasing the particles sizes decreases the surface area, which is expected to decrease the adsorption, while increasing the amount of TiO2-NPs used is expected to increase the sites available for dye adsorption.

CONCLUSION

In this study, CS-NPs were prepared and functionalized with TiO2-NPs to allow on-demand degradation of CS-NPs using UV irradiation as a trigger. Monodispersed nanoparticulate systems were prepared successfully with sizes and charges related to the size of CS-NPs and the amount of TiO2-NPs used in the preparation. Further, the sizes of the NPs and the amount of TiO2-NPs used have a great effect on NPs collapse upon irradiated with UV light. Moreover, the deposition of TiO2-NPs increases as the size of CS-NPs decreases and as the amount of TiO2-NPs used in the formulation increases. Finally, the fluorescence intensity test showed a significant difference in the release of Rhodamine as the functionalized and unfunctionalized CS-NPs were irradiated with UV light. Therefore, we can conclude that the chemical deposition of TiO2-NPs on PNP’s could allow on-demand remotely controlled drug release that depends on therapy and patient circumstances.

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Conflicts of interest

There are no conflicts of interest.

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