Transcriptomic model-based IncRNAs and mRNAs serve as independent prognostic indicators in head and neck squamous cell carcinoma

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Abstract. Head and neck squamous cell carcinoma (HNSC) is one of the most common types of cancer worldwide, and mRNAs and long non-coding RNAs (lncRNAs) have been identified as prognostic biomarkers in HNSC. In the present study, using gene expression datasets from multiple platforms, survival-associated genes in HNSC were identified. Subsequently, a combination of 17 genes (14 mRNAs and 3 lncRNA) was optimized using random forest variable hunting and a risk score model for HNSC prognosis was developed using a cohort from The Cancer Genome Atlas. Patients with high-risk scores tend to have earlier disease recurrence and lower survival rates, compared with those with low-risk scores. This observation was further validated in three independent datasets (GSE41613, GSE10300 and E-MTAB-302). Association analysis revealed that the risk score is independent of other clinicopathological observations. On the basis of the results depicted in the nomogram, the lncRNA-mRNA signature-based risk score successfully predicts the survival of HNSC and serves as an indicator of prognosis.

Introduction

Head and neck squamous cell carcinoma (HNSC) is one of the most common types of cancer worldwide (1). According to a recent study, 108,700 new cases were identified and 56,200 mortalities occurred as a result of HNSC in China in 2015 (2). The reasons behind HNSC carcinogenesis include smoking and human papilloma virus (HPV) infection (3). The 5-year survival rate of HNSC is estimated to be ~50% (4); although novel treatment methods have been utilized, the survival rate has not improved significantly over recent decades (5). Therefore, a prognostic model was urgently required.

Non-coding RNAs, particularly long non-coding RNAs (lncRNAs), have been the subjects of considerable attention in recent years, although the abundance of these RNAs is much lower than that of mRNAs (6). lncRNAs serve crucial roles in various cellular processes in HNSC, including carcinogenesis and progression (6-14). Metastasis associated lung adenocarcinoma transcript 1 (MALAT1) was identified as an oncogene, and the high expression of MALAT1 is associated with metastasis and poor survival across different cancer types (15-17). Suppression of HOX transcript antisense RNA expression was reported to induce apoptosis and to inhibit proliferation of HNSC cells (18). In addition to their use as prognostic markers, certain lncRNAs, including growth arrest specific 5, were reported to be participate in treatment response (19). Chemotherapy drugs, including cisplatin and paclitaxel, have been demonstrated to exert effects on lncRNAs (20), to a certain extent.

In line with this, lncRNAs and mRNAs significantly associated with survival were identified using Cox univariate regression based on two independent datasets. To facilitate the utilization and to reduce the size of the panel, random forest variable hunting was implemented and 17 lncRNA-mRNAs were used to develop the model, which estimated the survival with risk scores. The risk score was significantly associated with survival in all the training and test datasets involved. Association analyses revealed that the risk score is a prognostic factor that is independent of other clinical observations.

Materials and methods

Raw data pre-processing. The Cancer Genome Atlas (TCGA) expression data evaluated using RNA-seq was downloaded from the TCGA website (http://cancergenome.nih.gov/), the
upper quantile fragments per kilobase per million (FPKM) method (21) was used to normalize primary HNSC samples. The normal, recurrent and metastatic samples were removed and genes expressed in <80% samples were excluded for further analysis. Half of the minimum FPKM value (except for zero) was used in order to avoid zero values for each gene. Subsequently, the expression data were log2-transformed. The pre-processed data was then z-transformed for further analysis. The raw microarrays data were downloaded from the Gene Expression Omnibus (https://www.ncbi.nlm.nih.gov/geo) and the ArrayExpress (http://www.ebi.ac.uk/arrayexpress/) websites (21-23). Following background correction and normalization, the expression values were calculated. If several probes represented the same gene, the mean value was used as the expression value. Z-scores of each sample in each dataset were also evaluated. The probes were matched to lncRNAs, as described in a previous study (24).

**Prediction of gene selection and Cox multivariate regression model.** Cox univariate regression was performed on the Cancer Genome Atlas Head-Neck Squamous Cell Carcinoma (TCGA-HNSC) (n=407) and GSE41613 (n=97) datasets (24), and the IncRNAs/mRNAs significantly associated with overall survival (OS) (P<0.01) in the two datasets were identified as candidate genes, with 87 genes being identified. To narrow down the panel and optimize the results, random forest variable hunting was performed to identify biomarker combinations for survival prediction with 100 replications and 100 iterations, using z-score-transformed expression values and OS information. The selected 17 IncRNAs and mRNAs were used to develop the Cox multivariate regression model, with the ‘coxph’ function in the R package ‘survival’ (25). In the validation datasets, coefficients were locked and the risk score for each sample was calculated. The risk score was calculated using the following formula; where β indicates the coefficients evaluated with gene expression and x refers to the relative gene expression level.

\[
\text{Risk score} = \sum_{i} \beta_i \cdot x_i
\]

The risk score of each sample was calculated in each dataset, and the median risk score value of each dataset was used to divide the high-risk and low-risk group.

**Statistical analysis.** All statistical analyses involved in the present study were implemented on R (https://www.r-project.org/; v3.0.1) platform and R packages. Normalization of Affymetrix raw data was performed using the R package ‘affy’ (26) using function ‘rma’. The survival analysis and Cox proportional hazards model was performed using the R package ‘survival’ and the random forest for variable hunting performed using the package ‘randomForestSRC’ (27,28). The receiver operating characteristic curves were drawn using R package ‘pROC’ (25), and the nomogram was drawn using R package ‘rms’ (29).

**Results**

**Identification of 87 survival-associated mRNAs and IncRNAs.** Over-fit often occurs in model development. To avoid this, Cox univariate regression was implemented in two independent datasets, TCGA-HNSC (n=407) and GSE41613 (n=97). IncRNAs and mRNAs detected in the two platforms (next-generation sequencing assembly by TCGA and affymetrix HG U133 plus 2) were retained for further analysis. The mRNAs and IncRNAs that were significantly associated with overall survival in the two datasets (Cox univariate regression; P<0.01) were retained for further analysis. In total, 87 genes, including 10 IncRNAs and 77 mRNAs, were identified. To reduce redundancy, the mRNAs and lncRNAs detected in the two platforms (next-generation sequencing assembly by TCGA and affymetrix HG U133 plus 2) were retained for further analysis. The mRNAs and lncRNAs that were significantly associated with overall survival in the two datasets (Cox univariate regression; P<0.01) were retained for further analysis. The risk score was further validated in other independent datasets, GSE41613 (n=97), GSE10300 (n=81) and E-TABM-302 (n=81). Following the locking of coefficients in each dataset, the risk scores were calculated and the high/low-risk groups were divided according to the median risk score in each dataset. The overall survival (OS) and recurrence-free survival (RFS) times (Fig. 2B). The high-risk group samples tend to have lower survival rates, high expression of oncogenes and low expression of tumor suppressor genes (Fig. 2C). In order to test the robustness of the risk score, P-values were calculated by retrieving 80% of samples from the dataset (K-fold), and this process was repeated 10,000 times. In 92.59% of cases, the difference in survival rate between the risk-high and risk-low groups was statistically significant, as depicted in Fig. 2D.

**Risk score performance validation.** The high performance of the risk score may result from over-fit; thus, the robustness of the risk score was further validated in other independent datasets, GSE41613 (n=97), GSE10300 (n=81) and E-TABM-302 (n=81). Following the locking of coefficients in each dataset, the risk scores were calculated and the high/low-risk groups were divided according to the median risk score in each dataset. The overall survival (OS) and recurrence-free survival (RFS) times (Fig. 2B). The high-risk group samples tend to have lower survival rates, high expression of oncogenes and low expression of tumor suppressor genes (Fig. 2C). In order to test the robustness of the risk score, P-values were calculated by retrieving 80% of samples from the dataset (K-fold), and this process was repeated 10,000 times. In 92.59% of cases, the difference in survival rate between the risk-high and risk-low groups was statistically significant, as depicted in Fig. 2D.
survival (RFS) of the high-risk group were significantly lower than those of the low-risk group (Fig. 3A-C, top). The expression patterns of these 17 genes in these datasets was similar to those of the training dataset (Fig. 3A-C). Together, these results indicated that the risk-score staging model was robust across datasets and platforms (RNA-seq and microarray).

To test the performance of random forest variable hunting, which has previously been used for biomarker combination screening (24,26), 20 randomly-selected 17-gene panels were generated, and the risk score model was developed using these genes. However, the majority of combinations were not statistically significantly different in the four datasets (17/20; Table II), indicating that random forest variable hunting is a powerful tool for candidate gene selection.

**Risk score and clinicopathological observations.** Association analyses between risk score and other clinical observations were performed in the largest dataset, TCGA-HNSC. The results revealed that the risk score was able to predict prognosis independently of sex, differentiation grade and primary tumor size (Fig. 4A), indicating that the risk score served as an independent prognostic indicator. To facilitate the utilization of the risk score and to evaluate its importance among clinical observations, a nomogram (Fig. 4B) for the 3-year OS rate was plotted,

![Figure 1. Selection of 17 lncRNAs and mRNAs. (A) The frequency of genes selected during random forest variable hunting and (B) the coefficients of 17 RNAs. (C) The coefficient of each gene was shown. lncRNAs, long non-coding RNAs; TCGA, The Cancer Genome Atlas.](image-url)

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Table II: The coefficient of each gene was shown. lncRNAs, long non-coding RNAs; TCGA, The Cancer Genome Atlas.
Table I. Cox univariate and multivariate regression of genes involved.

| Gene             | Univariate Cox regression | Multivariate Cox regression |
|------------------|---------------------------|-----------------------------|
|                  | HR  | 95% CI   | P-value | HR  | 95% CI   | P-value |
| WDR54            | 2.30| 1.30-4.10| 0.004   | 0.68| 0.32-1.47| 0.330   |
| RPL26L1          | 3.90| 1.40-10.00| 0.007  | 4.95| 1.38-17.73| 0.014  |
| PSMC1            | 5.20| 2.00-14.00| 0.001  | 1.74| 0.60-5.07| 0.311  |
| SRPX             | 1.50| 1.10-2.10| 0.006  | 0.99| 0.68-1.44| 0.945  |
| SMS              | 2.80| 1.40-5.50| 0.003  | 1.49| 0.60-3.66| 0.387  |
| MTFMT            | 4.50| 1.50-13.00| 0.007  | 1.26| 0.32-4.91| 0.736  |
| CNFN             | 0.77| 0.64-0.93| 0.007  | 0.80| 0.62-1.02| 0.070  |
| CLEC10A          | 0.59| 0.44-0.80| 0.001  | 0.79| 0.49-1.25| 0.314  |
| LRRRC8           | 1.30| 1.10-1.60| 0.007  | 1.13| 0.92-1.39| 0.252  |
| RAB24            | 0.42| 0.23-0.76| 0.005  | 1.02| 0.41-2.52| 0.965  |
| PTPN9            | 4.10| 1.40-12.00| 0.008  | 1.52| 0.36-6.38| 0.570  |
| ZNF266           | 0.11| 0.045-0.26| <0.001 | 0.11| 0.04-0.29| <0.001 |
| CIITA            | 0.60| 0.43-0.83| 0.002  | 0.98| 0.59-1.62| 0.924  |
| IL1RAP           | 2.00| 1.20-3.40| 0.009  | 1.23| 0.70-2.18| 0.468  |
| ENSG00000261408  | 0.53| 0.34-0.84| 0.007  | 0.93| 0.51-1.71| 0.826  |
| ENSG00000261777  | 1.90| 1.20-3.10| 0.009  | 1.67| 0.95-2.95| 0.074  |
| ENSG00000265206  | 0.55| 0.42-0.73| <0.001 | 0.79| 0.53-1.20| 0.271  |

HR, hazard ratio; CI, confidence interval; WDR54, WD repeat domain 54; RPL26L1, ribosomal protein L26 like 1; PSMC1, proteasome 26S subunit, ATPase 1; SRPX, sushi repeat containing protein, X-linked; SMS, spermine synthase; MTFMT, mitochondrial methionyl-tRNA formyltransferase; CNFN, cornifelin; CLEC10A, C-type lectin domain containing 10A; LRRRC8, leucine rich repeat containing 38; RAB24, member RAS oncogene family; PTPN9, protein tyrosine phosphatase, non-receptor type 9; ZNF266, zinc finger protein 266; CIITA, class II major histocompatibility complex transactivator; IL1RAP, interleukin 1 receptor accessory protein.

Figure 2. Risk score in The Cancer Genome Atlas (training datasets). (A) Overall survival and (B) recurrence-free survival of the high-risk and low-risk groups. (C) The association between risk score (upper), survival information (middle) and z-score transformed expression value (bottom). (D) The density was assayed.
Figure 3. Risk score validation in three independent datasets. The survival plot (upper), risk score (middle), survival information (bottom) and z-score transformed seventeen gene expression values are presented in the (A) GSE41613 (B) GSE10300 and (C) E-TABM-302 datasets. HR, hazard ratio.

Figure 4. Association between risk score and other clinical information. (A) The association between risk score and clinical information is not significantly different (P>0.05). (B) Nomogram for predicting mortality using the risk score and clinical information.
Table II. Twenty (20) random combination performance in datasets.

| Combination/dataset | E302 | GSE10300 | GSE41613 | TCGA-OS |
|---------------------|------|----------|----------|---------|
| ENSG00000245213, ENSG00000212694, LRR3C8, NEK6, FOXN1, SRPX, CALML5, STC1, DOT1L, SPOCK1, P4HA1, ZNF823, CLEC10A, EXPH5, MAST4, AHSG, CNFN | 0.034280 | 0.029630 | 0.040090 | 0.000003 |
| ZNF266, STC1, B4GALNT1, PIK3C2B, SSR3, RPL26L1, ENSG00000265206, enSG00000261269, GPN1, CALML5, TOR1A, AREG, ZBED6CL, SNX20, PPIFB2, ENSG00000258634, ENSG00000228863 | 0.047550 | 0.029340 | 0.039440 | 0.001180 |
| GABARAPL2, ENSG00000258634, MRPS23, EZH1, TOMM34, VEPH1, SH3B5PL1, IL2RG, AREG, ZNF66, SPOCK1, SRPX, ALG2, FOXN1, ZNF967, OSGIN2, ENSG00000261408 | 0.087940 | 0.258250 | 0.943440 | 0.058420 |
| GABARAPL2, ENSG00000258634, MRPS23, EZH1, TOMM34, VEPH1, SH3B5PL1, IL2RG, AREG, ZNF66, SPOCK1, SRPX, ALG2, FOXN1, ZNF967, OSGIN2, ENSG00000261408 | 0.268740 | 0.807370 | 0.093000 | 0.000001 |
| I1A2, IL2RG, IL1RAP, CLEC10A, SMS, SRPX, PTPN9, RAB24, LRR3C8, ENSG00000261777, MTFMT, CNFN, CITAT, RPL26L1, ENSG00000271870, MRPS23, TOR1A, CNFN, NAT8L, IL2RG, LIF, CTA, LT, P2RY14, PTPN9, SPINK5, UNK, MAST4, ENSG00000212694 | 0.116500 | 0.026790 | 0.000390 | 0.987820 |
| GABARAPL2, ENSG00000258634, MRPS23, EZH1, TOMM34, VEPH1, SH3B5PL1, IL2RG, AREG, ZNF66, SPOCK1, SRPX, ALG2, FOXN1, ZNF967, OSGIN2, ENSG00000261408 | 0.186930 | 0.239400 | 0.239550 | 0.166820 |
| GABARAPL2, ENSG00000258634, MRPS23, EZH1, TOMM34, VEPH1, SH3B5PL1, IL2RG, AREG, ZNF66, SPOCK1, SRPX, ALG2, FOXN1, ZNF967, OSGIN2, ENSG00000261408 | 0.435250 | 0.132970 | 0.000002 | 0.000017 |
| GABARAPL2, ENSG00000258634, MRPS23, EZH1, TOMM34, VEPH1, SH3B5PL1, IL2RG, AREG, ZNF66, SPOCK1, SRPX, ALG2, FOXN1, ZNF967, OSGIN2, ENSG00000261408 | 0.625440 | 0.928520 | 0.928520 | 0.000350 |
| HSD17B7, MAST4, TRIM32, MSANTD3, MSANTD3, NAT8L, CTA, ENSG00000261408, 0.637600 | 0.596900 | 0.600430 | 0.552850 |
| HSD17B7, MAST4, TRIM32, MSANTD3, NAT8L, CTA, ENSG00000261408, 0.643950 | 0.270340 | 0.872530 | 0.322990 |
| HSD17B7, MAST4, TRIM32, MSANTD3, NAT8L, CTA, ENSG00000261408, 0.713340 | 0.078270 | 0.004130 | 0.000030 |
| HSD17B7, MAST4, TRIM32, MSANTD3, NAT8L, CTA, ENSG00000261408, 0.715320 | 0.883560 | 0.130070 | 0.874740 |
| HSD17B7, MAST4, TRIM32, MSANTD3, NAT8L, CTA, ENSG00000261408, 0.723310 | 0.214790 | 0.720550 | 0.196580 |
and it was demonstrated that the risk score was the most important predictor of the 3-year OS rate (range, 0-100). Taken together, the aforementioned results indicated that the 17 lncRNA-mRNA-based risk score is an independent clinical indicator and that it is more efficient in predicting survival than other clinical observations.

**Risk score and radiation therapy.** In the present study, patients were artificially divided into radiation-receiving and non-radiation-receiving groups. These groups were further divided into high-risk and low-risk groups based on their corresponding median risk score values. The high-risk group exhibited a significantly poorer survival rate in the radiation-receiving (Fig. 5A) and non-radiation-receiving groups (Fig. 5B), indicating that the risk score was a prognostic indicator independent to radiation therapy.

**Discussion**

In the present study, using the expression of lncRNAs and mRNAs and random forest variable hunting, a risk-score model was developed and was further validated in another three independent datasets. Furthermore, the risk score is independent of other clinicopathological observations and performs better at survival-prediction than other clinical characteristics.

The clinical outcome of HNSC carcinoma is affected by the heterogeneity of cancer, treatment method and surgery, the latter of which is controllable, while heterogeneity. Therefore, it is important to elucidate the molecular mechanisms underlying the heterogeneity of HNSC carcinoma (27). Clinically, HNSC may result from multiple causes, including smoking, HPV infection and diet, which makes the survival-prediction of HNSC difficult to determine using clinicopathological observations. Another indication of this heterogeneity is that no single gene was associated with survival in all datasets used in the present study. On the other hand, a multigene-based model is more robust in predicting prognosis, according to previous studies (24,28-31). lncRNAs and mRNAs are notable regulators of carcinogenesis and cancer development (32,33).

However, there are limitations to the present study. To begin with, owing to the retrospective nature of the study, important clinical indicators, including the resection margin of

**Table II. Continued.**

| Combination/dataset                                      | E302  | GSE10300 | GSE41613 | TCGA-OS |
|--------------------------------------------------------|-------|----------|----------|---------|
| RPL26L1, PPFIBP2, PLOD2, DOT1L, GABARAPL2, LTB, ZNF823, | 0.876380 | 0.621280 | 0.000017 | 0.000650 |
| ICOS, SPINK5, PCF11, FOXN1, RASSF2, BOD1, NEK6, SRPX,   |       |          |          |         |
| ENSG00000261408, THBS1                                 |       |          |          |         |
| LTB, SPOCK1, BOD1, ALG2, CNFN, SRPX, OSGIN2, PCMT1,    | 0.936720 | 0.264460 | 0.022270 | 0.333580 |
| MTFMT, PSAMC1, GP1, CRIPAK, PIK3C2B, ARHGAP30, AHSG,   |       |          |          |         |
| AQP1, DOT1L                                             |       |          |          |         |
| TRIM32, MTFMT, ENSG00000265206, PTPN9, XPR1, NEK6,    | 0.963480 | 0.342870 | 0.004220 | 0.000067 |
| TOMM34, ZNF266, DENND2D, NAT8L, RAB24, PLOD2, SPOCK1,  |       |          |          |         |
| HMGA2, ENSG00000245213, AQP1, LIF                      |       |          |          |         |

**Figure 5. Risk score and radiation therapy.** Survival rate of high-risk patients who (A) did and (B) did not receive radiation therapy is shorter. HR, hazard ratio.
surgery and the drugs used during the therapeutic period, were not available. Secondly, although the risk score is an independent prognostic factor, pooled data were required to facilitate its utilization.

In summary, the 17 IncRNA-mRNA-based model is robust across different platforms and is a better survival predictor than other factors.

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Availability of data and materials
The datasets generated and/or analyzed during the current study are available from the Gene Expression Omnibus (https://www.ncbi.nlm.nih.gov/geo) and the ArrayExpress (http://www.ebi.ac.uk/arrayexpress/) websites.

Authors' contributions
ZLZ, LJZ, LX, LC, FW and YF conceived the study, participated in its design. ZLZ and YF drafted the manuscript. YPX and SHZ contributed to the acquisition of data, data analysis. All authors read and approved the final manuscript.

Ethics approval and consent to participate
Not applicable.

Patient consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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