Large-scale assessment of olfactory preferences and learning in *Drosophila melanogaster*: behavioral and genetic components

Elisabetta Versace\(^1,2\) and Julia Reisenberger\(^1\)

\(^1\) Institut für Populationsgenetik, Vetmeduni, Vienna, Austria
\(^2\) Center for Mind/Brain Sciences, University of Trento, Rovereto, Italy

**ABSTRACT**

In the Evolve and Resequence method (E&R), experimental evolution and genomics are combined to investigate evolutionary dynamics and the genotype-phenotype link. As other genomic approaches, this method requires many replicates with large population sizes, which imposes severe restrictions on the analysis of behavioral phenotypes. Aiming to use E&R for investigating the evolution of behavior in *Drosophila*, we have developed a simple and effective method to assess spontaneous olfactory preferences and learning in large samples of fruit flies using a T-maze. We tested this procedure on (a) a large wild-caught population and (b) 11 isofemale lines of *Drosophila melanogaster*. Compared to previous methods, this procedure reduces the environmental noise and allows for the analysis of large population samples. Consistent with previous results, we show that flies have a preference for orange vs. apple odor. With our procedure wild-derived flies exhibit olfactory learning in the absence of previous laboratory selection. Furthermore, we find genetic differences in the olfactory learning with relatively high heritability. We propose this large-scale method as an effective tool for E&R and genome-wide association studies on olfactory preferences and learning.

**INTRODUCTION**

Ongoing evolutionary dynamics and genotype-phenotype mapping can be studied during experimental evolution through subsequent phenotyping and genomic sampling (*Travisano & Lenski, 1996; Wiser, Ribeck & Lenski, 2013*). This method is known as Evolve and Resequence (E&R) (*Turner et al., 2011*) and can be applied to entire populations by sequencing at the same times hundreds of individuals (Pool-seq, see *Futschik & Schlötterer, 2010*) of the same population. Thanks to the advancements of high-throughput sequencing techniques, the E&R method has been used to track the changes in genomic composition not only across thousand of generations in bacteria (*Wiser, Ribeck & Lenski, 2013*) but also in eukaryotes with a fast life cycle, such as yeast (*Barrick & Lenski, 2013*) and fruit flies (*Schlötterer et al., 2015*). This approach provides a very promising opportunity...
to investigate the evolution of complex traits and their genetic architecture with a limited budget (Schlötterer et al., 2014), thus paving the way to the analysis of the evolutionary dynamics of traits that cannot be inferred through fossil records, including complex behavioral phenotypes (Versace, 2015).

The first implementation of E&R on a complex behavior focused on phenotypic and genomic change in response to artificial selection for shorter/longer inter-pulse interval in male courtship song in Drosophila melanogaster (Turner & Miller, 2012). In this study, thousand of loci have been identified that responded to artificial selection and differed between populations selected for different behaviors. Similar outcomes, with thousand of alleles that significantly change in frequency between generations and treatments, have been found also for morphological or physiological traits (Orozco-terWengel et al., 2012; Tobler et al., 2014), showing that the same methodological issues apply to behavioral and other traits. Despite the success in identifying some causative genes (e.g., Zhou et al., 2011; Martins et al., 2014), theoretical (Schlötterer et al., 2015; Kofler & Schlötterer, 2014) and empirical evidence (Tobler et al., 2014; Franssen et al., 2015) has clarified that many of the significantly changed variants are in fact false positives derived by short or long-distance linkage disequilibrium.

Another limit that E&R shares with other genome-wide approaches, is low statistical power in identifying unknown causative variants (Rockman, 2012; e.g., Kofler & Schlötterer, 2014). Although haplotype-blocks can be used to study the dynamics of selected genomic regions during experimental evolution (Franssen et al., 2015), an effective E&R study should primarily minimize the false positives rate and maximize statistical power. As shown in recent theoretical and simulation work (Baldwin-Brown, Long & Thornton, 2014; Kessner & Novembre, 2015; Schlötterer et al., 2015; Kofler & Schlötterer, 2014), to reach this aim several issues have to be taken into account in the design of the experiment: (a) use a large starting population (possibly hundreds or thousands of individuals); (b) use a large population size; (c) use at least 5–10 replicate populations; (d) run the experiment for dozens of generations; (e) reduce linkage disequilibrium. In the light of this, during E&R researchers should phenotype and propagate thousands of individuals in multiple replicate populations for many generations (Kofler & Schlötterer, 2014; Versace, 2015). In E&R studies, a practical limitation is imposed by the stages of propagation and phenotyping: when flies have to be individually phenotyped and manipulated, the time and working load required can force a reduction of the census size. For this reason, investigating behavioral traits, that often require a large effort in phenotyping, poses a methodological challenge.

In this work, we focus on the development of a fast and reliable method for phenotyping and propagating medium-size and large-size populations of fruit flies assessed for olfactory behavior. This approach can be effectively used in E&R of olfactory behavior, in particular olfactory preferences and olfactory learning after conditioning with an aversive stimulus, as well as for any large-scale assay. Fruit flies show complex behaviors, can be easily maintained at a large census size, have a fast generation cycle and low linkage disequilibrium (Mackay et al., 2012), and thus are a convenient model to investigate the evolutionary dynamics and genotype-phenotype map of behavioral traits.
Olfactory behavior (olfactory preferences and olfactory learning) is a good candidate for E&R investigation of both spontaneous and learned responses, because of its remarkable conservation between *Drosophila* and vertebrates (Davis, 2005; Wilson, 2013) and the presence of standing variation for both olfactory preferences and olfactory learning (Mery et al., 2007; Ruebenbauer et al., 2008; Rollmann et al., 2010; Van den Berg et al., 2011). Genetic variability in the experimental population is crucial to apply E&R to fruit flies, because within the time scale of feasible experiments (50 generations of selection take about two years to be completed) new mutations have little impact on evolutionary change.

Different preferences for specific odors and odor concentrations have been documented in *Drosophila* using T-mazes and olfactometers (Kutsukake et al., 2000; Wang et al., 2003; e.g., Suh et al., 2004; Revadi et al., 2015). Phenotypic variability in olfactory behavior is associated with polymorphisms that influence reactions to different compounds (Wang et al., 2007; Wang et al., 2010) but to date E&R has not been applied to odor preferences (see Rhodes & Kawecki, 2004 for a similar idea). Olfactory learning has been extensively studied at the behavioral, genetic and neurobiological level (McGuire, Deshazer & Davis, 2005; Davis, 2005; Fiala, 2007; Tabone & de Belle, 2014). Wild and mutant flies have been tested in associative conditioning tasks, typically the association between an olfactory conditioned stimulus and an electric or mechanical aversive stimulus. In a first paradigm developed to measure olfactory learning, Quinn and colleagues (1974) tested groups of about 40 flies. From a starting tube flies could approach a light source at the end of a second tube painted with odor A (or B). When flies entered the second tube an electric shock was delivered and could be associated with odor A (or B). After being returned to the starting tube, flies could enter a third tube containing odor B (or A), that was not associated with any electric shock. At test flies could choose to enter either a tube with odor A or one with odor B. A performance index compared the fraction of flies that avoided the unpaired odor and those which avoided the shock-paired odor. Not all flies explored the tubes, and their performance was affected also by phototaxis, thus this small-scale assay produced low learning scores. Tully & Quinn (1985) modified the paradigm to test groups of about 100 flies in an apparatus in which odor A matched with pulses of electric shock was followed by odor B in the absence of electric shock. The odors were delivered by vacuum so that all flies were exposed to the odor-shock contingencies. After training, the flies were tested in a T-maze where could choose to approach either odor A or B. The learning scores for this procedure ranged between 0.7 and 0.9 (McGuire, Deshazer & Davis, 2005) but the need of dedicated machines and hands-on operations on the flies limit the application of this method to large-scale long-term experiments. Other methods used to score olfactory behaviors require individual handling/scoring of the flies (Swarup et al., 2013) and/or air flow and automated systems (Steck et al., 2012; Brown et al., 2013) that can hardly be implemented to the large scales required for long-term evolutionary experiments.

To date, the main method used in experimental evolution for enhanced learning (Mery & Kawecki, 2002; Dunlap & Stephens, 2014) is the oviposition paradigm. This method is a medium/large-scale procedure based on the habit of flies to use the same medium for foraging and egg laying. The procedure starts exposing hundreds of free ranging flies to
olfactory (or visual) stimuli associated to palatable or aversive media displaced in petri dishes located in a box (e.g., orange juice smell associated with palatable food, apple juice smell associated with aversive flavor). After exposure to olfactory (or visual) and associated gustatory stimuli, flies are tested for their olfactory (or visual) preferences for stimuli previously associated or not associated with the aversive flavor. Compared to flies that do not remember the association, flies that remember the contingencies presented during the exposure phase are expected to lay more eggs in the substrate whose smell (or color) was never associated with aversive flavor. The proportion of eggs laid in the substrate associated with the palatable flavor is used as a proxy for learning.

To select for enhanced learning across generations, *Mery & Kawecki (2002)* rinsed, moved to a neutral medium and propagated only the eggs laid in the medium previously associated with palatable food (alternatively *Dunlap & Stephens (2009)* displaced eggs individually with a needle). As effect of this regime, in about 15 generations the proportion of flies which made the “correct” choice significantly increased. In spite of this, several aspects make the oviposition procedure less than ideal for E&R studies: (a) this method is prone to experimental noise, as shown by the lack of learning effects before selection; (b) only females are exposed to selection, because learning is measured using laid eggs, thus reducing the selective pressure to half of the propagated individuals and preventing to investigate behavioral domains different from oviposition and sex effects; (c) the oviposition paradigm imposes selection for fertility, egg laying during the few hours of the test and resistance to egg washing; (d) this paradigm does not control for the experience provided during the conditioning phases (many flies might not experience all stimuli before making a choice in the test phase); (e) extensive work to rinse/displace the eggs and propagate the flies is required, in turn reducing the number of experimental replicates that can be propagated.

Aiming to use E&R for investigating the evolution of behavior in fruit flies, we have developed a simple and effective method based on a T-maze to assess olfactory preferences and learning in large samples (hundreds) of fruit flies of both sexes as well as in smaller samples (dozens of individuals). This method can hence be used for any behavioral or genomic approach which requires medium or large samples. We find evidence for olfactory preferences and learning in a large population of *D. melanogaster* originally caught in South Africa and in a population of inbred lines originally caught in Portugal. Our procedure reduces the impact of undesired selective pressures and the effort in propagation and phenotyping. Furthermore, this method is sensitive enough to detect olfactory preferences and learning, and the heritability of these traits. We discuss the relevance of this T-maze based procedure for E&R and genome-wide association studies as well as behavioral/chemical ecology studies.

**MATERIALS AND METHODS**

**Subjects**

All experiments were run on isofemale lines of the same species, *D. melanogaster*. Flies were maintained on standard cornmeal-soy flour-syrup-yeast medium, except during the
experimental assays. Before the beginning of the experiments we kept all lines for at least two generations at 22 °C in a constant 14:10 h light:dark cycle.

The population-wide experiments were ran on 670 lines derived from a natural population of *D. melanogaster* collected in Paarl (South Africa) in March 2012. In each trial we used a group of 250 adult flies (males and females 2 days old or older), randomly picked from the 670 isofemale lines of the South African population.

The individual-line experiments were ran on 11 inbred lines derived from a *D. melanogaster* population of 113 lines originally collected during the summer in Povoa de Varzim (northern Portugal) in July 2008 and maintained as isofemale lines. From these isofemale lines, 11 inbred lines were generated through full-sibling mating of 3 lines of the base stock population (B101, B192 and B211), 8 lines of a population maintained for 53 generations in a hot regime at 25 °C (R1, R2, R3, R5) or for 33 generations in a cold regime at 18 °C (R6, R7, R9 and R10). The hot and cold temperature regimes are described in detail in Orozco-terWengel et al. (2012) and Tobler et al. (2014). Inbred lines were generated through full-sib mating for 17 or more generations (B101: 17 generations; B192: 18 generations; B211: 19 generations; R1 and R3: 27 generations; R2: 29 generations; R5: 21 generations, R6 and R9: 20 generations; R7 and R10: 22 generations). For each line a virgin female and a randomly collected male were allowed to mate and from their offspring another virgin female and a random male were used to create the next generation. After inbreeding, these lines were kept as isofemale lines until the experimental assays. In each trial, we used a group of 40 flies (males and females 2 days old or older) of the same line.

**Apparatus and stimuli**

The T-maze (31 × 17.5 cm) used for the experimental assays (Fig. 1A) consisted of a starting chamber and a central chamber (12 × 8 × 1.5 cm) connected on each side to a food chamber. The starting chamber (9.5 × 2.5 cm) contained the flies at the beginning of each experimental phase. Food chambers (9.5 × 2.5 cm) were filled with 4 ml of experimental food. In each experimental phase, flies begun the exploration of the apparatus from the starting chamber. The central chamber was connected to the food chambers with a funnel that prevents flies to re-enter the central chamber once they have approached the food. A similar trapping technique has been previously used for fruit flies.

Experimental media were prepared with juice fruit (either orange or apple juice from 100% concentrate) and agar (14 g/l). Aversive flavor was obtained adding 8 g/l of quinine to the experimental medium. The use of complex mixtures of odorants, as those found in fruit juice, is justified both by the previous literature in the field (Mery & Kawecki, 2002) and by chemical ecology studies which showed that in *D. melanogaster* most olfactory receptors are responsive to complex mixtures of fruit odors and many fruit odors strongly activate multiple receptors (Hallem & Carlson, 2006). Hence, complex fruit odorants provide a wide evolutionary basis of experimental evolution studies.
**T-maze olfactory paradigm**

A Exposure 1: odor 1 and flavor 1 on both sides. In the learning assay flavor 1 is made aversive adding quinine.

B Exposure 2: odor 2 and flavor 2 on both sides.

C Test: odor 1 and 2 are simultaneously presented on different sides of the apparatus.

**Figure 1** T-maze apparatus and experimental paradigm. (A) During Exposure 1, flies are presented with one odor (apple or orange) and the aversive flavor (apple or orange juice supplemented with quinine in the learning assay, without quinine in the olfactory preference assay). (B) In Exposure 2, flies are presented the second odor (orange or apple) and the second flavor (orange or apple juice without quinine). (C) At Test, both stimuli (orange and apple odor) are presented (without quinine) on different sides of the apparatus.
Procedure

Olfactory preferences

The olfactory preferences assay is based on overnight starvation (15–16 h) followed by 2 h exposure to the first odor (Exposure 1), 2 h exposure to the second odor (Exposure 2), 4 h of starvation, and 2 h of Test.

We assessed preferences for apple and orange odor in the absence of punishment by using the same procedure described for the learning assays (see below), with the only difference that no food supplemented with quinine (aversive stimulus) was provided during the exposure phases. By comparing the results obtained with this procedure and those obtained with the aversive stimulus we could evaluate the role of the aversive stimulus in determining olfactory choice.

Olfactory learning assays

The learning assay is based on overnight starvation (15–16 h) followed by 2 h exposure to the first odor-flavor contingency (Exposure 1), 2 h exposure to the second odor-flavor contingency (Exposure 2), 4 h of starvation and 2 h of Test.

We used CO₂ anesthesia to collect flies and starve them 15–16 h before the beginning of the conditioning procedure. After starvation flies were moved to the starting chamber for Exposure 1.

In a pilot study we observed that in a time course of two hours flies tend to explore only one arm of the T-maze. Hence we decided to expose flies serially to the olfactory and taste stimuli, presenting the first contingency in Exposure 1 and the second contingency in the subsequent Exposure 2. During Exposure 1, for two hours flies were exposed to the odor associated with the aversive flavor and could approach the aversive flavor (e.g., orange odor and orange juice supplemented with quinine) located in the food chambers. To make sure to score the first choice for all the flies and reduce the exposure to food, after one hour of exposure we substituted the food chambers with new vials filled with the same type of experimental food and moved the trapped flies in an empty vial. We repeated the same procedure at the end of Exposure 1. Flies who entered the food chambers during Exposure 1 were moved to the starting chamber for Exposure 2. During Exposure 2, for two hours flies were exposed to the odor associated with the palatable flavor and the palatable flavor (e.g., apple odor and apple juice). After one hour of exposure, we substituted the food chambers with new vials filled with the same type of experimental food and moved the trapped flies in an empty vial. We repeated the same procedure at the end of Exposure 2. Flies who entered the food chambers during Exposure 2 were starved for four hours prior to the Test.

In half trials we conditioned flies on apple odor associated with aversive flavor and orange odor associated with palatable food (A-/O), and also conditioned flies on orange odor associated with aversive flavor and apple odor associated with palatable food (O-/A). It has been shown that in D. melanogaster appetitive long-term memory occurs after single-cycle training also in the absence of fasting (Krashes & Waddell, 2008), while aversive long-term memory by single-cycle training requires previous fasting.
Hirano et al., 2013). For this reason, the exposure to the aversive stimulus was always conducted immediately after fasting (Exposure 1).

Flies began the Test from the starting chamber. Differently from the exposure phases, during the Test the odor associated with the aversive flavor and the odor associated with the palatable flavor were presented simultaneously, each on a different food chamber (Fig. 1C). Across different trials, we alternated the right/left side in which the two odors were presented. No food was supplemented with quinine during this phase. We counted flies that chose to enter either the orange odor side or the apple odor side.

**Data analysis**

In the test for olfactory preferences between the orange and apple odor we compared the proportion of flies that chose the orange odor vs. the random choices level using a $t$-test single sample against the random choice proportion of 0.5. Beforehand we controlled for deviations from the normal distribution of the data using the Shapiro–Wilk normality test. To check for any effect of the order of presentation (O/A: Orange followed by Apple odor, A/O Apple followed by Orange odor) we calculated the order score $o$ as the difference in the proportion of flies that in each trial chose orange odor after being exposed to O/A vs. A/O:

$$o = (\text{proportion orange choices O/A flies}) - (\text{proportion orange choices A/O flies})$$

An order score significantly different from zero was expected if the order of presentation of the two odors/flavors had an effect in determining subsequent choices.

Similarly, in the test for conditioned preferences between the orange and apple odor, we compared the proportion of flies that chose the orange odor vs. the chance level using a $t$-test single sample against the random choices proportion of 0.5. Beforehand, we controlled for deviations from the normal distribution of the data using the Shapiro–Wilk normality test.

To obtain a measure of learning (learning score, $l$) we calculated the difference in the proportion of flies that in each trial chose orange odor after being conditioned on apple flavor vs. orange flavor:

$$l = (\text{proportion orange choices O-/A flies}) - (\text{proportion orange choices A-/O flies})$$

A learning score significantly different from zero was expected if the conditioning had an effect.

To investigate the heritable component of learning, we repeatedly tested the same inbred lines ($n = 10$) and derived the intraclass correlation $t$, which is an estimate of the genetic heritability ($h^2$) of learning for the tested population, using the variance between ($V_b$) and within ($V_w$) inbred lines (Hoffmann & Parsons, 1988):

$$t = \frac{V_b - V_w/n}{V_b + (n-1)(V_w/n)} = \frac{nV_b - V_w}{nV_b + (n-1)V_w}$$

As discussed by David et al. (2005), the intraclass correlation can be used as a proxy for heritability (see also Hoffmann & Parsons, 1988; Betti, Soto & Hasson, 2014).
RESULTS

Population experiments: olfactory preferences and learning assays

To evaluate the sensitivity of our method in detecting olfactory preferences and learning in large groups of naturally derived fruit flies, we investigated the differences in olfactory preferences and learning in a large South African *D. melanogaster* population (see Data S1 and S2). In each trial, we tested 250 flies of both sexes.

In the olfactory preference experiment, across 28 test trials the overall population showed a marginally significant preference for the orange odor (*t*27 = 1.953, *p* = 0.06). This preference is consistent with the preference for citrus previously documented in *D. melanogaster* (*Mery & Kawecki, 2002* but see *Betti, Soto & Hasson, 2014*; e.g., *Dweck et al., 2013*), that is likely a behavioral strategy against the attack of parasitic wasps (*Dweck et al., 2013*).

Before testing flies, we exposed them to both odors/flavors: in half trials flies were exposed first to orange then to apple (O/A), in half trials first to apple then to orange (A/O). We have derived the order effect score *o* to investigate the effect of the order in which the orange/apple stimuli had been presented. We observed a significant order effect score (*t*13 = 3.09, *p* = 0.009; Fig. 2B), indicating that A/O flies (flies first exposed to Apple, then to Orange) had a significantly higher preference for orange odor than O/A flies (flies first exposed to Orange, then to Apple). A post-hoc *t*-test on A/O and O/A flies vs. the chance
Figure 3  Orange choices after conditioning and learning score. (A) Proportion of orange choices of flies conditioned with Orange aversive/Apple palatable (O-/A, yellow) and Apple aversive/Orange palatable (A-/O, orange). (B) Learning score: difference in the proportion of orange odor choices between flies conditioned A-/O (Apple aversive, Orange palatable) and O-/A (Orange aversive, Apple palatable).

level (0.5) revealed that only A/O flies had a significant preference for the orange odor: for A/O flies $t_{13} = 3.18$, $p = 0.007$; for O/A flies $t_{13} = −0.242$, $p = 0.81$.

In the conditioning experiment, the overall population showed a preference for the orange odor (mean = 0.59, $t_{57} = 3.95$, $p < 0.001$). Flies that previously experienced Apple as aversive/Orange as palatable (A-/O) were more likely to choose orange than flies exposed to the opposite contingency (O-/A) ($t_{56} = 2.24$, $p = 0.029$; Fig. 3A). The population showed a significant learning score ($t_{28} = 2.88$, $p = 0.007$; Fig. 3B).

We also checked for differences in the proportion of orange choices after the conditioning procedure and after the olfactory preference exposures. Overall, after conditioning flies had a stronger preference for orange odor than in the absence of conditioning ($t_{81} = 2.50$, $p = 0.014$), thus indicating a significant effect of the conditioning procedure. These results suggest that exposure to aversive stimuli can influence the preferences of flies towards specific odors/flavors. Given that conditioning with the aversive stimulus was done more than 4 h prior to the test, we showed that our procedure is sensitive to memory capabilities that last at least 4 h.

When comparing samples that had the same order of presentation of apple and orange odor in the olfactory preference and conditioning and procedure, we observed a significant difference for the A/O but not for the O/A presentation (A-/O: $t_{40} = 2.49$, $p = 0.017$; O-/A: $t_{59} = 1.24$, $p = 0.22$). These results indicate that the conditioning procedure is more effective in the A-/O exposure than in the O-/A exposure. As suggested by an anonymous reviewer, this could result from the unconditioned preference for the orange stimuli, that
produces an asymmetry in the strength of conditioning, with the O- stimulus less effective in conditioning than the A- stimulus.

**Inbred lines: olfactory preferences**

To study olfactory preferences in 11 inbred lines of *D. melanogaster* derived from a population collected in Portugal, we used the same procedure adopted for the large population using 40 flies from the same isofemale line in each trial (see Data S3). Line R6 consistently did not enter the food chambers in both experiments, so we excluded this line and run the analyses in the ten remaining lines.

In the olfactory preference assay, the overall distribution of the orange odor choices was significantly different from the normal distribution (Shapiro–Wilk normality test: $W = 0.98$, $p = 0.03$) and we analyzed the data using non-parametric tests (Wilcoxon signed-rank test and Kruskal–Wallis test). Overall, the group of ten responsive lines showed an olfactory preference for the orange odor (mean $= 0.56$; $V = 7862$, $p < 0.001$; Fig. 4A). We did not observe significant differences across lines (Kruskal–Wallis Chi squared $= 14.14$, $p = 0.12$; see Fig. 4B) and in the effect of the order of presentation (A/O vs. O/A) (Kruskal–Wallis Chi squared $= 1.48$, $p = 0.22$).

We calculated the order effect score—(proportion of orange odor choices after A/O exposure — proportion of orange odor choices after O/A exposure)—for the overall sample of ten lines tested and found no significant effect ($V = 1300.5$, $p = 0.23$).

**Inbred lines: learning assays**

To study learning in the 11 inbred lines of *D. melanogaster* derived from a population collected in Portugal, we used the same procedure adopted for the large population using
Figure 5 Conditioned orange odor choices for a population of ten inbred lines and learning score. (A) Proportion of orange odor choices for flies conditioned with Orange aversive/Apple palatable (O-/A) and Apple aversive/Orange palatable (A-/O) in the overall sample of ten inbred lines. (B) Learning score: difference in the proportion of orange odor choices between flies conditioned on A-/O (Apple aversive, Orange palatable) and O-/A (Orange aversive, Apple palatable).

40 flies from the same isofemale line in each trial (see Data S4). Line R6 consistently did not enter the food chambers in both experiments, so we excluded this line and run the analyses in the ten remaining lines.

In the learning assay, the overall distribution of the orange odor choices was significantly different from the normal distribution (Shapiro–Wilk normality test: $W = 0.98$, $p = 0.005$) and we analyzed the data using non parametric tests (Wilcoxon signed-rank test and Kruskal–Wallis test). Overall, after the conditioning procedure the ten responsive lines showed a preference for orange odor (mean = 0.59, Wilcoxon signed-rank test, $V = 8630$, $p = 9.159x^{10^{-6}}$), Fig. 5A. No significant difference in the overall choices was observed between the olfactory preference and the learning assay ($W = 13655.5$, $p = 0.30$). Differently from the olfactory preference assay though, significant differences in the proportion of orange choices were apparent between the two conditioning treatments (A-/O vs. O-/A: Kruskal–Wallis Chi square: 34.93, $p = 3.424x^{10^{-9}}$) (Fig. 5A), and we documented a significant learning effect (Fig. 5B). We also detected significant differences in the proportion of orange choices between lines (Kruskal–Wallis Chi square: 23.45, $p = 0.005$).

We ran post-hoc tests to evaluate the performances of each line in the A-/O and O-/A conditioning procedure. For each line we measured the proportion of orange choices after conditioning with A-/O and O-/A (Fig. 6). After using the Bonferroni–Holmes correction for multiple comparisons, we found that in the A-/O procedure three lines (R10, R7, R9)
had a preference significant at 5% level for orange and four lines (R1, R2, R3, R5) had a preference significant at 10% level, whereas in the O-/A procedure only R5 had a preference significant at 10% level. These results indicate that most of the tested inbred lines are able to discriminate between apple and orange odor.

We calculated the learning score—(proportion of orange odor choices after A-/O exposure − proportion of orange odor choices after O-/A exposure)—for the overall population and found a significant effect of learning (mean = 0.22, \( V = 2752.5, p < 0.001 \)). Since there was a significant effect of Line in the learning score (Kruskal–Wallis Chi square = 22.15, \( p = 0.008 \)) we ran also post-hoc tests on each line (Fig. 6). All lines exhibited a higher proportion of orange choices after being conditioned with Orange as palatable stimulus (and all lines increased the proportion of orange odor choice after conditioning compared to the olfactory preference assay, see Fig. 4B). Using the Bonferroni–Holmes correction for multiple comparisons, we found that three lines (R3, R7, R9) showed a learning score significant at 5% level and three lines (R1, R2 and R5) that were significant at 10% level. These results suggest that most of the tested lines are able to learn through our conditioning procedure.

**Inbred lines: heritability of olfactory behavior**

We derived an estimate of the genetic heritability of olfactory preferences and olfactory learning using the variance between \( (V_b) \) and the variance within \( (V_w) \) lines and calculating the intraclass correlation \( t \) as a proxy for heritability in inbred lines (Hoffmann & Parsons, 1988; David et al., 2005).

---

**Figure 6** Proportion of orange odor choices for each tested line conditioned with Orange aversive/Apple palatable (O-/A exposure) and Apple aversive/Orange palatable (A-/O exposure).
In the olfactory preferences, the variability between lines \((V_b = 0.09)\) was larger than the variability within lines \((V_w = 0.06)\) and the intraclass correlation is \(t = 0.6\). The same pattern holds true for olfactory learning: the variability between lines \((V_b = 0.017)\) is much higher than the variability within lines \((V_w = 0.004)\), thus leading to \(t = 0.80\). The high intraclass correlations show a moderate to high heritability of olfactory preferences and learning and suggest that our method is suitable to investigate these traits.

**DISCUSSION**

Historically, the evolutionary dynamics of behavioral traits have been particularly hard to catch. This is not only due to lack of fossil record as a tool to help reconstructing evolutionary change but also to limits in investigating organisms with a complex behavior for hundreds or thousands of generations, as can be done with yeast (e.g., [Goddard, Godfray & Burt, 2005]) and bacteria ([Wiser, Ribeck & Lenski, 2013]. *Drosophila* is a model system which shows either complex behavior and a life cycle fast enough for being studied in experimental evolution. For instance, in few generations of targeted selection it has been possible to obtain a significant increase in learning in a wild-derived population of *Drosophila melanogaster* ([Mery & Kawecki, 2002]), or to change its responsiveness to specific (odor-flavor or color-flavor) associations ([Dunlap & Stephens, 2014]. These behavioral findings have not been accompanied by a correspondent genomic investigation, partly due to the costs and difficulties associated until recently to this enterprise. The recent development of high-throughput sequencing technologies, together with advancements in statistical and bioinformatics tools, has changed this scenario. In particular, using the Evolve and Resequence method ([Törner et al., 2011]) entire populations can be propagated and investigated for genomic changes at subsequent time points by sequencing collectively hundreds of individuals (a method known as Pool-seq, see [Futschik & Schlötterer (2010)]. This approach paves the way to the analysis of complex behavioral phenotypes such as olfactory preferences and learning in *Drosophila* ([Versace, 2015]).

Empirical ([Turner & Miller, 2012; Orozco-terWengel et al., 2012; Tobler et al., 2014; Franssen et al., 2015] and theoretical studies ([Schlötterer et al., 2015; Kofler & Schlötterer, 2014]) have shown the current limits of E&R in terms of false positive and false negative errors. Different strategies have been suggested to reduce the error rate and increase the efficiency of this method in investigating evolutionary dynamics and the genotype-phenotype link ([Baldwin-Brown, Long & Thornton, 2014; Schlötterer et al., 2015; Kofler & Schlötterer, 2014]), including propagate and phenotype large samples of several replicate populations for multiple generations. The oviposition method ([Mery & Kawecki, 2002]) is the experimental paradigm currently used for experimental evolution of learning in fruit flies ([Mery & Kawecki, 2002; Dunlap & Stephens, 2009; Dunlap & Stephens, 2014]). This procedure though is not optimal for E&R due to drawbacks in the effort required for propagation, reduced sample size and selective pressures for traits different from those of interest. With the aim of overcoming these limitations, we have established a method based on subsequent exposure and test in a T-maze used to assess olfactory preferences and learning in large (hundreds or even thousands of individuals) and medium/small
(dozens) samples of fruit flies. With our experimental schedule (4 h of exposure following starvation, 4 h of starvation, 2 h of test) it is possible to conduct the assay in a single day, during the light cycle of not sleep-deprived fruit flies. This simple procedure can impose selection on both sexes and does not entail selection for fast egg laying and egg washing to propagate flies.

We have used the T-maze procedure to investigate olfactory preferences and learning in a large population of *D. melanogaster* originally caught in South Africa and in 11 inbred lines of another population of *D. melanogaster* originally caught in Portugal. Overall both populations show a preference for orange vs. apple odor. The preference for citrus media had been previously documented in *D. melanogaster* (*Mery & Kawecki, 2002; Dweck et al., 2013*; but see *Betti, Soto & Hasson, 2014*). We find a significant effect of learning in both populations. The presence of olfactory learning in the absence of selection for this trait shows the sensitivity of our method. The fact that we show learning and olfactory preferences in fruit flies of both tested populations, in the absence of selective pressures, suggests that this method can be successfully applied to different genetic pools.

Small population size and inbreeding negatively affect the resolution of genomic scans (*Schlötterer et al., 2015; Kofler & Schlötterer, 2014; Franssen et al., 2015*), thus limiting the power of E&R and genome-wide association studies. This limitation could be overcome using the T-maze procedure on large samples, since no individual handling of eggs is necessary in this method, and it is not necessary the separate male and female flies.

Moreover, repeatedly testing inbred lines we have detected genetic differences in olfactory behavior between lines. Differences between wild-derived inbred lines have been previously documented (*Nepoux, Haag & Kawecki, 2010; Nepoux et al., 2015*). We have also calculated the intraclass correlation *t* (*Hoffmann & Parsons, 1988; David et al., 2005*) as an estimate of heritability, showing a medium/high heritability for the investigated traits.

We have showed that our method is suitable to be used with large samples with the aim of investigating the evolution of spontaneous preferences and learning performances in large groups of fruit flies with limited effort. The ease of implementation of our procedure will enable researchers to investigate the perceptual/learning abilities of their population of interest, including variability between strains, the phenotypic distribution of large populations, and the hereditability of these traits, before starting long-term projects. On this basis, we suggest to use T-mazes in large-scale experiments as a tool for E&R and genome-wide association studies on olfactory preferences and learning and for other traits. Our experimental paradigm can be easily adapted for the needs of chemical ecology and pest management to assess olfactory behavior with different odors and odor concentrations in a comparative perspective; to test visual behaviors by changing the color/texture of the central and food chambers; and for investigating navigation performance by increasing the number of food chambers, as well the role of social information transmitted (*Kohn et al., 2013*; see for instance *Battesti et al., 2015*) by one or both sexes. Varying the delay between conditioning and test it is possible to investigate the duration of memory. We suggest the use of large-scale T-mazes to widen the
investigation of behavioral traits and cognitive abilities in invertebrates at the behavioral and genomic level.

ACKNOWLEDGEMENTS
We are grateful to Christian Schlötterer for his support through all stages of the project. Special thanks to the members of the Institut für Populationsgenetik at Vetmeduni and the Vienna Graduate School of Population Genetics for helpful discussions throughout the project. We thank Lino Ometto for fruitful discussions on the genetic analyses.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding
The project has been funded through a grant of the Austrian Science Fund (FWF, P22725) awarded to C. Schlötterer. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures
The following grant information was disclosed by the authors: Austrian Science Fund: FWF, P22725.

Competing Interests
The authors declare there are no competing interests.

Author Contributions
• Elisabetta Versace conceived and designed the experiments, performed the experiments, analyzed the data, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
• Julia Reisenberger performed the experiments, wrote the paper, reviewed drafts of the paper.

Supplemental Information
Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.1214#supplemental-information.

REFERENCES
Baldwin-Brown JG, Long AD, Thornton KR. 2014. The power to detect quantitative trait loci using resequenced, experimentally evolved populations of diploid, sexual organisms. *Molecular Biology and Evolution* 31:1040–1055 DOI 10.1093/molbev/msu048.
Barrick JE, Lenski RE. 2013. Genome dynamics during experimental evolution. *Nature Reviews. Genetics* 14:827–839 DOI 10.1038/nrg3564.
Battesti M, Pasquaretta C, Moreno C, Teseo S, Joly D, Klensch E, Petit O, Sueur C, Mery F, Biologie D. 2015. Ecology of information: social transmission dynamics within groups of non-social insects. *Proceedings of the Royal Society B: Biological Sciences* 282:20142480 DOI 10.1098/rspb.2014.2480.
Betti MIL, Soto EM, Hasson E. 2014. Oviposition site preference for natural breeding sites in *Drosophila melanogaster* (Diptera: Drosophilidae) populations from Argentina. *Annals of the Entomological Society of America* **107**:944–953 DOI 10.1603/AN14050.

Brown EB, Layne JE, Zhu C, Jegga AG, Rollmann SM. 2013. Genome-wide association mapping of natural variation in odor-guided behavior in *Drosophila*. *Genes, Brain, and Behavior* (12):503–515 DOI 10.1111/gbb.12048.

David JR, Gibert P, Legout H, Pétavy G, Capy P, Moreteau B. 2005. Isofemale lines in *Drosophila*: an empirical approach to quantitative trait analysis in natural populations. *Heredity* **94**:3–12 DOI 10.1038/sj.hdy.6800562.

Davis RL. 2005. Olfactory memory formation in *Drosophila*: from molecular to systems neuroscience. *Annual Review of Neuroscience* **28**:275–302 DOI 10.1146/annurev.neuro.28.061604.135651.

Dunlap AS, Stephens DW. 2009. Components of change in the evolution of learning and unlearned preference. *Proceedings of the Royal Society B: Biological Sciences* **276**:3201–3208 DOI 10.1098/rspb.2009.0602.

Dunlap AS, Stephens DW. 2014. Experimental evolution of prepared learning. *Proceedings of the National Academy of Sciences of the United States of America* **111**:11750–11755 DOI 10.1073/pnas.1404176111.

Dweck HKM, Ebrahim SAM, Kromann S, Bown D, Hillbur Y, Sachse S, Hansson BS, Stensmyr MC. 2013. Olfactory preference for egg laying on citrus substrates in *Drosophila*. *Current Biology* **23**:2472–2480 DOI 10.1016/j.cub.2013.10.047.

Fiala A. 2007. Olfaction and olfactory learning in *Drosophila*: recent progress. *Current Opinion in Neurobiology* **17**:720–726 DOI 10.1016/j.conb.2007.11.009.

Franssen SU, Nolte V, Tobler R, Schlötterer C. 2015. Patterns of linkage disequilibrium and long range hitchhiking in evolving experimental *Drosophila melanogaster* populations. *Molecular Biology and Evolution* **32**:495–509 DOI 10.1093/molbev/msu320.

Futschik A, Schlötterer C. 2010. The next generation of molecular markers from massively parallel sequencing of pooled DNA samples. *Genetics* **186**:207–218 DOI 10.1534/genetics.110.114397.

Goddard MR, Godfray HCJ, Burt A. 2005. Sex increases the efficacy of natural selection in experimental yeast. *Nature* **434**:636–640 DOI 10.1038/nature03405.

Hirano Y, Masuda T, Naganos S, Matsuno M, Ueno K, Miyashita T, Horiuchi J, Saitoe M. 2013. Fasting launches CRTC to facilitate long-term memory formation in *Drosophila*. *Science* **339**:443–446 DOI 10.1126/science.1227170.

Hoffmann AA, Parsons PA. 1988. The analysis of quantitative variation in natural populations with isofemale strains. *Genetics, Selection, Evolution* **20**:87–98 DOI 10.1186/1297-9686-20-1-87.

Kessner D, Novembre J. 2015. Power analysis of artificial selection experiments using efficient whole genome simulation of quantitative traits. *Genetics* **199**:991–1005 DOI 10.1534/genetics.115.175075.

Kofler R, Schlötterer C. 2014. A guide for the design of evolve and resequencing studies. *Molecular Biology and Evolution* **31**:474–483 DOI 10.1093/molbev/mst221.

Kohn NR, Reaume CJ, Moreno C, Burns JG, Sokolowski MB, Mery F. 2013. Social environment influences performance in a cognitive task in natural variants of the foraging gene. *PLoS ONE* **8**:e81272 DOI 10.1371/journal.pone.0081272.
Krashes MJ, Waddell S. 2008. Rapid consolidation to a radish and protein synthesis-dependent long-term memory after single-session appetitive olfactory conditioning in Drosophila. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience* **28**:3103–3113 DOI 10.1523/JNEUROSCI.5333-07.2008.

Kutsukake M, Komatsu A, Yamamoto D, Ishiwa-Chigusa S. 2000. A tyramine receptor gene mutation causes a defective olfactory behavior in *Drosophila melanogaster*. *Gene* **245**:31–42 DOI 10.1016/S0378-1119(99)00569-7.

Mackay TFC, Richards S, Stone EA, Barbadilla A, Ayroles JF, Zhu D, Casillas S, Han Y, Magwire MM, Criddle JM, Richardson MF, Anholt RRH, Barron M, Bess C, Blankenburg KP, Carbone MA, Castellano D, Chaboub L, Duncan L, Harris Z, Javaid M, Jayaseelan JG, Jhangiani SN, Jordan KW, Lara F, Lawrence F, Lee SL, Librado P, Linheiro RS, Lyman RF, Mackey AJ, Munidasa M, Muzny DM, Nazareth L, Newsham I, Perales L, Pu L-L, Qu C, Rámia M, Reid JG, Rollmann SM, Rozas J, Saada N, Turlapati L, Worley KC, Wu Y-Q, Yamamoto A, Zhu Y, Bergman CM, Thornton KR, Mittelman D, Gibbs RA. 2012. The *Drosophila melanogaster* genetic reference panel. *Nature* **482**:173–178 DOI 10.1038/nature10811.

Martins NE, Faria VG, Nolte V, Schlötterer C, Teixeira L, Sucena E, Magalhaes S. 2014. Host adaptation to viruses relies on few genes with different cross-resistance properties. *Proceedings of the National Academy of Sciences of the United States of America* **111**(16):5938–5943 DOI 10.1073/pnas.1400378111.

McGuire SE, Deshazer M, Davis RL. 2005. Thirty years of olfactory learning and memory research in *Drosophila melanogaster*. *Progress in Neurobiology* **76**:328–347 DOI 10.1016/j.pneurobio.2005.09.003.

Mery F, Belay AT, So AK-C, Sokolowski MB, Kawecki TJ. 2007. Natural polymorphism affecting learning and memory in *Drosophila*. *Proceedings of the National Academy of Sciences of the United States of America* **104**:13051–13055 DOI 10.1073/pnas.0702923104.

Mery F, Kawecki TJ. 2002. Experimental evolution of learning ability in fruit flies. *Proceedings of the National Academy of Sciences of the United States of America* **99**:14274–14279 DOI 10.1073/pnas.222371199.

Nepoux V, Babin A, Haag C, Kawecki TJ, Le Rouzic A. 2015. Quantitative genetics of learning ability and resistance to stress in *Drosophila melanogaster*. *Ecology and Evolution* **5**(3):543–556 DOI 10.1002/ece3.1379.

Orozco-terWengel P, Kapun M, Nolte V, Kofler R, Flatt T, Schlötterer C. 2012. Adaptation of *Drosophila* to a novel laboratory environment reveals temporally heterogeneous trajectories of selected alleles. *Molecular Ecology* **21**:4931–4941 DOI 10.1111/j.1365-294X.2012.05673.x.

Quinn WG, Harris WA, Benzer S. 1974. Conditioned behavior in *Drosophila melanogaster*. *Proceedings of the National Academy of Sciences of the United States of America* **71**:708–712 DOI 10.1073/pnas.71.3.708.

Revadi S, Eccher F, Mazzoni V, Ani S Al, Carlin S, Vrhovsek U, Anfora G. 2015. Olfactory responses of *Drosophila suzukii* to host plant volatiles. *Physiological Entomology* **40**(1):54–64 DOI 10.1111/phen.12088.

Rhodes JS, Kawecki TJ. 2004. Behavior and neurobiology. In: Garland T, Rose MR, eds. *Experimental evolution*. Berkeley: University of California Press, 263–300.
Rockman MV. 2012. The QTN program and the alleles that matter for evolution: all that’s gold does not glitter. *Evolution; International Journal of Organic Evolution* **66**:1–17 DOI 10.1111/j.1558-5646.2011.01486.x.

Rollmann SM, Wang P, Date P, West SA, Mackay TFC, Anholt RRH. 2010. Odorant receptor polymorphisms and natural variation in olfactory behavior in *Drosophila melanogaster*. *Genetics* **186**:687–697 DOI 10.1534/genetics.110.119446.

Ruebenbauer A, Schlyter F, Hansson BS, Löfstedt C, Larsson MC. 2008. Genetic variability and robustness of host odor preference in *Drosophila melanogaster*. *Current Biology* **18**:1438–1443 DOI 10.1016/j.cub.2008.08.062.

Schlötterer C, Kohler R, Versace E, Tobler R, Franssen SU. 2015. Combining experimental evolution with next-generation sequencing: a powerful tool to study adaptation from standing genetic variation. *Heredity* **114**:431–440 DOI 10.1038/hdy.2014.86.

Schlötterer C, Tobler R, Kohler R, Nolte V. 2014. Sequencing pools of individuals—mining genome-wide polymorphism data without big funding. *Nature Reviews Genetics* **15**:749–763 DOI 10.1038/nrg3803.

Steck K, Veit D, Grandy R, Badia SBI, Badia SBI, Mathews Z, Verschure P, Hansson BS, Knaden M. 2012. A high-throughput behavioral paradigm for *Drosophila* olfaction—the flywalk. *Scientific Reports* **2**:361 DOI 10.1038/srep00361.

Suh GSB, Wong AM, Hergarden AC, Wang JW, Simon AF, Benzer S, Axel R, Anderson DJ. 2004. A single population of olfactory sensory neurons mediates an innate avoidance behaviour in *Drosophila*. *Nature* **431**:854–859 DOI 10.1038/nature02980.

Swarup S, Huang W, Mackay TFC, Anholt RRH. 2013. Analysis of natural variation reveals neurogenetic networks for *Drosophila* olfactory behavior. *Proceedings of the National Academy of Sciences of the United States of America* **110**:1017–1022 DOI 10.1073/pnas.1220168110.

Tabone C, De Belle S. 2014. Olfactory learning and memory assays. In: Dubnau J, ed. *Behavioral genetics of the fly* (Drosophila melanogaster). Cambridge: Cambridge University Press, 231–249.

Tobler R, Franssen SU, Kohler R, Orozco-terWengel P, Nolte V, Hermisson J, Schlötterer C. 2014. Massive habitat-specific genomic response in *D. melanogaster* populations during experimental evolution in hot and cold environments. *Molecular Biology and Evolution* **31**:364–375 DOI 10.1093/molbev/mst205.

Travisano M, Lenski RE. 1996. Long-term experimental evolution in *Escherichia coli*. IV. Targets of selection and the specificity of adaptation. *Genetics* **143**:15–26.

Tully T, Quinn WG. 1985. Classical conditioning and retention in normal and mutant *Drosophila melanogaster*. *Journal of Comparative Physiology A* **157**:263–277 DOI 10.1007/BF01350033.

Turner TL, Miller PM. 2012. Investigating natural variation in *Drosophila* courtship song by the evolve and resequence approach. *Genetics* **191**:633–642 DOI 10.1534/genetics.112.139337.

Turner TL, Stewart AD, Fields AT, Rice WR, Tarone AM. 2011. Population-based resequencing of experimentally evolved populations reveals the genetic basis of body size variation in *Drosophila melanogaster*. *PLoS Genetics* **7**:e1001336 DOI 10.1371/journal.pgen.1001336.

Van den Berg M, Duivenvoorde L, Wang G, Tribuhi S, Bukovinszky T, Vet LEM, Dicke M, Smid HM. 2011. Natural variation in learning and memory dynamics studied by artificial selection on learning rate in parasitic wasps. *Animal Behaviour* **81**:325–333 DOI 10.1016/j.anbehav.2010.11.002.

Versace E. 2015. Experimental evolution, behavior and genetics: associative learning as a case study. *Current Zoology* **61**:226–241.
Wang Y, Chiang A-S, Xia S, Kitamoto T, Tully T, Zhong Y. 2003. Blockade of neurotransmission in Drosophila mushroom bodies impairs odor attraction, but not repulsion. *Current Biology* 13:1900–1904 DOI 10.1016/j.cub.2003.10.003.

Wang P, Lyman RF, Mackay TFC, Anholt RRH. 2010. Natural variation in odorant recognition among odorant-binding proteins in Drosophila melanogaster. *Genetics* 184:759–767 DOI 10.1534/genetics.109.113340.

Wang P, Lyman RF, Shabalina SA, Mackay TFC, Anholt RRH. 2007. Association of polymorphisms in odorant-binding protein genes with variation in olfactory response to benzaldehyde in Drosophila. *Genetics* 177:1655–1665 DOI 10.1534/genetics.107.079731.

Wilson RI. 2013. Early olfactory processing in Drosophila: mechanisms and principles. *Annual Review of Neuroscience* 8:217–241 DOI 10.1146/annurev-neuro-062111-150533.

Wiser MJ, Ribeck N, Lenski RE. 2013. Long-term dynamics of adaptation in asexual populations. *Science* 342:1364–1367 DOI 10.1126/science.1243357.

Zhou D, Udpa N, Gersten M, Visk DW, Bashir A, Xue J, Frazer KA, Posakony JW, Subramaniam S, Bafna V, Haddad GG. 2011. Experimental selection of hypoxia-tolerant Drosophila melanogaster. *Proceedings of the National Academy of Sciences of the United States of America* 108:2349–2354 DOI 10.1073/pnas.1010643108.