Platelet-Derived Growth Factor Is Associated with Progression of Symptomatic Intracranial Atherosclerotic Stenosis

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\textbf{Background and Purpose} We aimed to determine the relationships of 33 biomarkers of inflammation, oxidation, and adipokines with the risk of progression of symptomatic intracranial atherosclerotic stenosis (ICAS).

\textbf{Methods} Fifty-two of 409 patients who participated in the TOSS-2 (Trial of Cilostazol in Symptomatic Intracranial Stenosis-2) showed progression of symptomatic ICAS in magnetic resonance angiography at 7 months after an index stroke. We randomly selected 20 patients with progression as well as 40 age- and sex-matched control patients. We serially collected blood samples at baseline, 1 month, and 7 months after an index stroke. Multiplex analysis of biomarkers was then performed.

\textbf{Results} Demographic features and risk factors such as hypertension, diabetes, and smoking history were comparable between the two groups. Univariate analyses revealed that the levels of platelet-derived growth factor (PDGF)-AA [median (interquartile range)=1.64 (0.76–4.57) vs. 0.77 (0.51–1.71) ng/mL], PDGF-AB/BB [10.31 (2.60–25.90) vs. 2.35 (0.74–6.70) ng/mL], and myeloperoxidase [10.5 (7.5–22.3) vs. 7.8 (5.5–12.2) ng/mL] at 7 months were higher in the progression group. In the multivariate analysis using logistic regression, the PDGF AB/BB level at 7 months was independently associated with the progression of ICAS ($p=0.02$).

\textbf{Conclusions} The PDGF-AB/BB level is associated with the progression of ICAS, and so may play a significant role in the progression of human ICAS.

\textbf{Key Words} platelet-derived growth factor, intracranial stenosis, ischemic stroke, magnetic resonance angiography.

\section*{INTRODUCTION}

Intracranial atherosclerotic stenosis (ICAS) is one of the main causes of ischemic stroke, especially among Asians, Hispanics, and Africans.\textsuperscript{1,3} In the WASID (Warfarin-Aspirin Symptomatic Intracranial Disease) trial, the rates of ischemic stroke in the territory of ICAS (50% to 99% stenosis) at 1 year were as high as 15% and 14% in the aspirin and warfarin arms, respectively.\textsuperscript{4} Since atherosclerotic plaques can change dynamically, ICASs may progress over time. Although the progression of ICAS is an important predictor of recurrent stroke,\textsuperscript{5,6} the mechanisms underlying this progression are still poorly understood.

Chronic inflammation in the arterial wall is a crucial feature associated with the development of atherosclerosis.\textsuperscript{7} Proatherogenic cytokines such as interleukin (IL)-1, IL-2, tumor necrosis factor (TNF)-\alpha, and CD40 ligand (CD40L) were found to be positively associated with the progression of atherosclerosis in the cervical carotid arteries or coronary arteries, whereas antiatherogenic cytokines such as IL-1 receptor antagonist (IL-1ra), IL-6, and IL-10...
had negative associations.8 A few studies have found associations between the progression of ICAS and cytokines including IL-6, C-reactive protein (CRP), and plasminogen activator inhibitor (PAI)-1.8,9

This prospective substudy of TOSS-2 (Trial of Cilostazol in Symptomatic Intracranial Stenosis-2)10 measured 33 biomarkers of inflammation, oxidation, and adipokines, with the aim of identifying their associations with the risk of progression of symptomatic ICAS.

METHODS

Initial TOSS-2 design
In TOSS-2, patients with symptomatic ICAS were randomly allocated to treatment with aspirin plus cilostazol or aspirin plus clopidogrel within 14 days of stroke onset.10 Follow-up magnetic resonance angiography (MRA) was performed 7 months after this allocation. The progression of symptomatic ICAS was indicated by changes in serial MRA findings. As reported previously, the severity of stenosis in both middle cerebral arteries (MCAs) was classified into five grades: normal, mild, moderate, severe, and occlusion.11 Progression was defined as worsening of stenosis by at least one grade in the follow-up MRA relative to the baseline MRA, whereas regression was defined as improvement of stenosis by at least one grade. The change in stenosis in MCAs was evaluated by gathering the raw data from magnetic resonance imaging and MRA data as DICOM files. Two raters blinded to clinical information and the location of symptomatic stenosis independently classified the degree of stenosis on MRA using Petaview, a noncommercial DICOM viewer program of Asan Medical Center. Discrepancies between the two reviewers were referred to the third rater and resolved by consensus among all three reviewers.

Fifty-two (12.7%) of 409 patients who completed follow-up MRA showed progression of symptomatic ICAS in follow-up MRA.

Collection and storage of blood samples
We collected serum from all patients who were included in TOSS-2 to evaluate the vascular risk factors at baseline, 1 month, and 7 months after an index stroke. Blood samples were collected by venipuncture from the participants into EDTA-containing vacutainer tubes after an overnight fast. The freshly drawn blood was centrifuged at 3,000g for 20 min at 4°C. Plasma was then separated before being stored at -70°C in small aliquots. All analyses were performed on frozen plasma samples. We randomly selected 20 patients with progressive ICAS as well as 40 age- and sex-matched controls without progressive ICAS, and analyzed their sera. The study protocol was approved by the Institutional Review Board of Dongguk University Ilsan Hospital. All participants or their legal representatives gave written informed consents.

Cytokine assay
Multianalyte profiling was performed on the Luminex-100 system and the XY Platform (Luminex Corporation, Austin, TX, USA). Calibration microspheres for classification and reporter readings as well as sheath fluid were also purchased from Luminex Corporation. The acquired fluorescence data were analyzed using the MasterPlex™ QT software (version 1.2, MiraiBio, Alameda, CA, USA). Plasma concentrations of fractalkine, IL-1a, IL-1b, IL-1ra, IL-2, IL-6, IL-8, IL-10, monocyte chemoattractant protein (MCP)-1, MCP-3, macrophage derived chemokine (MDC), sCD40L, TNF-α, vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF)-AA, PDGF-AB/BB, serum amyloid A (SAA), matrix metalloproteinase (MMP)-9, E-selectin, intercellular adhesion molecule (ICAM)-1, vascular-cell adhesion molecule (VCAM)-1, and PAI-1 were determined by the Millipore 8-Plex panel (Millipore, Burlington, MA, USA). P-selectin, MMP-2, and MMP-3 were measured using the R&D 8-Plex panel (R&D Systems, Minneapolis, MN, USA). Myeloperoxidase (MPO) and S100 beta were analyzed using the Youngin Frontier 6-Plex panel (Youngin Frontier, Seoul, Korea). All analyses were performed in accordance with the manufacturers’ protocols.

Statistical analysis
All statistical tests were two-sided, and the cutoff for significance was set at 0.05. Continuous variables are expressed as mean±standard-deviation or median (interquartile range) values, as appropriate, and categorical variables are described by numbers and percentages. Statistically significant intergroup differences was detected using the chi-square test for categorical variables and Student’s t-test or the Mann-Whitney U test for continuous variables as appropriate.

Multivariate analyses were performed to examine the independent relationships between biomarkers and the progression of symptomatic ICAS. We included variables for which \( p < 0.10 \) in the univariate analysis and covariates that were potentially associated with the progression of stenosis, including the degree of stenosis, statin use, and baseline low-density lipoprotein (LDL)-cholesterol level. We further divided patients into tertiles and then reran the analyses. All analyses were performed using the SAS statistical software package (version 9.1, SAS Institute, Cary, NC, USA).


**RESULTS**

The baseline characteristics of the subjects were similar in the progression and control groups (Table 1). The age at onset was 62.8±11.8 years in the progression group and 64.7±11.0 years in the control group. Vascular risk factors such as hypertension, diabetes, smoking, family history of stroke, and hyperlipidemia were comparable between the two groups. The severity of the symptomatic stenosis and the baseline neurological severity did not differ between the two groups. There were also no intergroup differences in the baseline blood LDL-cholesterol, high-density lipoprotein (HDL)-cholesterol, glucose, CRP, and hemoglobin A1c (HbA1c) levels. Medication allocation to cilostazol versus clopidogrel occurred equally between the two groups. In addition, the frequency of statin treatment was comparable between the two groups.

None of the analyzed cytokines differed at baseline or after 1 month. However, at 7 months the PDGF-AA, PDGF-AB/BB, and MPO levels were higher in the progression group than in control group (Table 2). Additional candidates for inclusion in the multivariate model, with p values of 0.05–0.10, were higher ICAM-1, PDGF-AB/BB, and P-selectin levels at baseline, and a higher P-selectin level at 7 months (Table 2).

Multivariate analysis using logistic regression identified the PDGF-AB/BB level at 7 months as the single cytokine factor that was independently associated with the progression of ICAS (p=0.02) (Table 3). Considering the ranges of PDGF-AB/BB levels, patients in the highest tertile at 7 months had a substantially higher probability of progression than those in the lowest tertile, with an adjusted odds ratio of 6.93 (95% confidence interval=1.53–31.38, p=0.01). The evolutions of PDGF-AB/BB levels over time in progressors and control subjects are shown in Fig. 1.

**DISCUSSION**

This study found an independent association between the PDGF-AB/BB level at 7 months after an index stroke and the progression of ICAS. Patients in the highest tertile of PDGF-AB/BB levels at 7 months had a sevenfold higher probability of progression of ICAS compared with the lowest tertile. Other atherogenic cytokines (e.g., IL-1, IL-2, IL-6, TNF-α, fractalkine, MCP-1, and CD40L) and antiatherogenic cytokines (e.g., IL-1ra, and IL-10) were not associated with the progression of ICAS. To our knowledge, this is the first study to investigate and demonstrate an association between PDGF-AB/BB and ICAS progression.

PDGF plays a significant role in the formation of blood vessels (angiogenesis) and their growth. PDGF is a potent mitogen for cells of mesenchymal origin, including smooth-muscle cells, and is produced by platelets, smooth-muscle cells, activated macrophages, and endothelial cells. PDGF exists as a disulfide-linked dimer and comprises two chains: A and B. Two receptors for PDGF, called α and β, have been identified. Binding of PDGF to the receptor induces receptor dimerization and activation of the kinase activity. The PDGF-α receptor binds both PDGF-A and -B chains, whereas the PDGF-β receptor binds only the PDGF-B chain. Blockade of the PDGF-β receptor (but not the PDGF-α receptor) prevents the accumulation of vascular smooth-muscle cells in apolipoprotein-E-knockout mice. These results indicate that the PDGF-β receptor plays an important role in the development of fibrous atherosclerotic lesions and that the regulation of signal transduction via the PDGF-β receptor could affect atherosclerosis in mice. Additional support for this mechanism comes from a study finding that a PDGF inhibitor, imatinib, attenuates diabetes-associated atherosclerosis. In addition to direct atherogenic effects, several studies have suggested that PDGF is an important mediator of arterial remodeling, which may indirectly contribute to the progression of stenosis.

In addition to identifying that PDGF-AA/BB is independently associated with ICAS progression, this study is notable for not finding a strong link between other candidate cyto-

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**Table 1. Baseline characteristics of the study population**

| Parameter | Progression (n=20) | Controls (n=40) | p |
|-----------|-------------------|----------------|---|
| Age, years | 62.8±11.8         | 64.7±11.0       | 0.53 |
| Sex, male | 10 (50)           | 21 (52.5)       | 0.86 |
| Hypertension | 12 (60)          | 32 (80)        | 0.10 |
| Diabetes | 8 (40)             | 15 (37.5)       | 0.85 |
| Smoking | 9 (45)            | 18 (45)        | 0.99 |
| Family history of stroke | 5 (25)         | 8 (20)      | 0.66 |
| Hyperlipidemia | 10 (50)      | 20 (50)      | >0.99 |
| NIHSS score | 3 (2–7)         | 3 (2–6)       | 0.29 |
| LDL-cholesterol, mg/dL | 120.9±30.8 | 117.6±48.7 | 0.79 |
| HDL-cholesterol, mg/dL | 42.6±13.4 | 42.6±11.0 | 0.99 |
| Glucose, mg/dL | 151.3±61.8 | 152.4±65.9 | 0.96 |
| CRP, mg/dL | 0.33±0.36 | 0.29±0.49 | 0.78 |
| HbA1c, % | 7.15±2.18 | 6.52±1.61 | 0.21 |
| Degree of stenosis | 0.56* | 0.56* | 0.56* |

Data are mean±standard deviation, number (percentage), or median (IQR) values. *Fisher’s exact test was used.

HDL: high-density lipoprotein, IQR: interquartile range, LDL: low-density lipoprotein, NIHSS: National Institutes of Health Stroke Scale.
Table 2. Cytokine levels at baseline, 1 months, and 7 months

| Cytokine     | Time point | Progression (n=20) | Control (n=40) | p    |
|--------------|------------|--------------------|----------------|------|
| Fractalkine, pg/mL | Baseline  | 1.86 (0–63.66)    | 6.23 (0–41.03) | 0.93 |
|              | 1 month   | 0 (0–22.73)       | 13.89 (0–44.56) | 0.11 |
|              | 7 months  | 2.91 (0–15.32)    | 18.04 (0–59.86) | 0.24 |
| IL-1α, pg/mL | Baseline  | 0 (0–14.89)       | 0 (0–11.96)    | 0.66 |
|              | 1 month   | 0                  | 0 (0–14.99)    | 0.62 |
|              | 7 months  | 0                  | 0 (0–15.83)    | 0.30 |
| IL-1β, pg/mL | Baseline  | 1.36 (0–1.88)     | 0.73 (0.34–2.15) | 0.96 |
|              | 1 month   | 1.13 (0.93–2.10)  | 0.94 (0.29–1.88) | 0.93 |
|              | 7 months  | 1.58 (0.50–2.27)  | 1.0 (0–1.74)   | 0.14 |
| IL-1ra, pg/mL| Baseline  | 7.58 (0–75.63)    | 8.92 (0–37.18) | 0.76 |
|              | 1 month   | 5.81 (0–28.67)    | 6.28 (0–24.29) | 0.96 |
|              | 7 months  | 11.96 (0.17–48.43)| 5.63 (0–28.46) | 0.22 |
| IL-2, pg/mL  | Baseline  | 0.03 (0–3.3)      | 0.91 (0.49–1.91) | 0.38 |
|              | 1 month   | 0 (0–1.53)        | 0.81 (0–4.47)  | 0.13 |
|              | 7 months  | 0.28 (0–2.75)     | 0.75 (0–3.99)  | 0.51 |
| IL-6, pg/mL  | Baseline  | 0 (0–26.13)       | 0.46 (0–11.22) | 0.71 |
|              | 1 month   | 0 (0–0)           | 0 (0–14.07)    | 0.13 |
|              | 7 months  | 0 (0–4.23)        | 0.24 (0–8.37)  | 0.56 |
| IL-8, pg/mL  | Baseline  | 7.65 (4.92–14.18) | 6.25 (4.75–9.39) | 0.33 |
|              | 1 month   | 5.5 (4.71–10.82)  | 6.06 (4.7–10.45) | 0.98 |
|              | 7 months  | 7.11 (4.89–12.53) | 6.13 (5.02–10.29) | 0.57 |
| IL-10, pg/mL | Baseline  | 2.34 (0–5.25)     | 1.71 (0.43–4.91) | 0.83 |
|              | 1 month   | 1.93 (0–5.20)     | 1.37 (0–3.78)  | 0.75 |
|              | 7 months  | 2.04 (0–10.6)     | 1.75 (0.27–4.43) | 0.56 |
| MCP-1, pg/mL | Baseline  | 290.1 (224.8–346.8) | 275.2 (239.23–338.0) | 0.93 |
|              | 1 month   | 288.3 (202.3–364.0) | 240 (182.2–324.8) | 0.22 |
|              | 7 months  | 271.4 (220.1–327.4) | 285.1 (228.8–340.8) | 0.68 |
| MCP-2, pg/mL | Baseline  | 6.62 (6.09–8.08)  | 6.67 (6.21–7.10) | 0.93 |
|              | 1 month   | 6.35 (6.17–7.24)  | 6.55 (6.16–7.04) | 0.41 |
|              | 7 months  | 6.60 (6.26–6.94)  | 6.61 (5.96–7.34) | 0.88 |
| MDC, pg/mL   | Baseline  | 1559.5 (812–2183) | 1335.7 (938–1629) | 0.24 |
|              | 1 month   | 1045 (787–1640)   | 1330.9 (916–1715) | 0.65 |
|              | 7 months  | 1365 (928–1877)   | 1400 (1058–2018) | 0.34 |
| sCD40L, ng/mL| Baseline  | 2.07 (0.84–20.35) | 1.84 (0.54–567.7) | 0.26 |
|              | 1 month   | 1.95 (0.53–5.69)  | 1.79 (0.88–4.36) | 0.86 |
|              | 7 months  | 3.00 (0.69–27.62) | 1.52 (0.72–6.04) | 0.24 |
| TNF-α, pg/mL | Baseline  | 4.38 (2.77–5.46)  | 3.82 (2.76–4.73) | 0.26 |
|              | 1 month   | 4.53 (2.66–5.89)  | 3.91 (3.07–4.99) | 0.68 |
|              | 7 months  | 4.24 (3.08–6.54)  | 4.0 (3.13–5.07) | 0.78 |
| VEGF, pg/mL  | Baseline  | 15.58 (0–105.8)   | 9.47 (0–42.4)  | 0.50 |
|              | 1 month   | 0 (0–28.9)        | 0 (0–26.8)     | 0.96 |
|              | 7 months  | 13.1 (0–66.3)     | 0 (0–28.7)     | 0.27 |
| PDGF-AA, ng/mL| Baseline | 1.76 (0.75–4.16)  | 1.204 (0.66–2.39) | 0.29 |
|              | 1 month   | 1.04 (0.69–1.99)  | 0.88 (0.48–1.61) | 0.28 |
|              | 7 months  | 1.64 (0.76–4.57)  | 0.77 (0.51–1.71) | 0.006* |
| PDGF-AB/BB, ng/mL | Baseline | 7.0 (2.40–2.22)  | 3.40 (1.62–9.30) | 0.09 |
|              | 1 month   | 2.58 (1.51–7.72)  | 2.50 (1.14–5.90) | 0.54 |
|              | 7 months  | 10.31 (2.60–25.90) | 2.35 (0.74–6.70) | 0.004* |
kines and further vessel narrowing. Cytokines such as TNF-α, IL-2, IL-3, IL-6, IL-8, IL-10, and interferon-γ are expressed in human atherosclerotic plaques and have been reported to play roles in the progression of atherosclerosis.7 With respect to the extracranial carotid artery, there are several reports of atherosclerosis being associated with cytokine profiles. Plasma CRP and TNF-α levels are independently associated with the risk of cardiovascular events in patients with atherosclerotic occlusive disease,22 and serum levels of IL-8, IL-6, and MCP-1 have been found to be related to the severity of carotid atherosclerosis.23-26 Furthermore, experimental studies have shown that mitigating these inflammatory cytokines impedes the progression of atherosclerosis.23,24 There is also evidence of associations between cytokines and coronary artery disease.27-29 Compared with extracranial arteries of similar sizes, intracranial arteries have a thinner tunica muscularis, tunica adventitia, and internal elastic lamina.3

Clinical studies have indicated that the risk factors, progression rate, and outcomes may differ between coronary/carotid arteries and intracranial arteries. Thus, the aforemen-

Table 2. Cytokine levels at baseline, 1 months, and 7 months (continued)

| Cytokine  | Time point | Progression (n=20) | Control (n=40) | p       |
|-----------|------------|--------------------|----------------|---------|
| SAA, ng/mL| Baseline   | 2982.0 (1001.2–8596.2) | 3130.5 (1190–7218.5) | 0.89    |
|           | 1 month    | 1875.0 (140.0–6298.7)  | 2242.0 (762.5–5281.7) | 0.42    |
|           | 7 months   | 2664.5 (412.0–4371.3)  | 2052.5 (796.3–7062.5) | 0.70    |
| E-selectin, ng/mL | Baseline | 40.4 (34.6–48.7) | 41.4 (37.1–47.1) | 0.94 |
|           | 1 month    | 41.7 (36.1–49.5) | 41.5 (36.7–47.2) | 0.94 |
|           | 7 months   | 42.3 (39.2–50.9) | 41.3 (38.2–48.26) | 0.42 |
| ICAM-1, ng/mL | Baseline | 92.1 (65.3–114.2) | 103.6 (84.1–143.8) | 0.08   |
|           | 1 month    | 90.0 (68.5–112.7) | 99.3 (64.0–136.2) | 0.83   |
|           | 7 months   | 93.5 (64.5–114.2) | 99.7 (67.3–144.8) | 0.63   |
| VCAM-1, ng/mL | Baseline | 551.4 (481.3–680.8) | 582.2 (478.6–766.8) | 0.49   |
|           | 1 month    | 585.1 (518.3–792.7) | 603.0 (514.0–714.0) | 0.66   |
|           | 7 months   | 627.1 (524.6–682.9) | 599.4 (515.1–673.7) | 0.85   |
| PAI-1, ng/mL | Baseline | 17.0 (12.5–27.2) | 16.9 (11.8–24.9) | 0.89   |
|           | 1 month    | 13.6 (9.8–20.8) | 15.3 (10.3–21.5) | 0.60   |
|           | 7 months   | 19.3 (13.0–32.4) | 18.3 (11.3–27.4) | 0.43   |
| P-selectin, pg/mL | Baseline | 41697.7 (28153.4–73949.5) | 32490.8 (24769.8–38011.8) | 0.06   |
|           | 1 month    | 32620.0 (20263.9–45428.2) | 30852.6 (24503.3–38460.2) | >0.99 |
|           | 7 months   | 49919.0 (28343.0–85196.9) | 33103.0 (24532.9–42906.6) | 0.06   |
| MMP-2, ng/mL | Baseline | 124.4 (100.6–226.7) | 130.8 (110.4–275.1) | 0.57   |
|           | 1 month    | 145.1 (112.9–305.3) | 134.3 (112.7–213.8) | 0.80   |
|           | 7 months   | 133.5 (107.8–269.8) | 169.5 (118.4–352.4) | 0.27   |
| MMP-3, ng/mL | Baseline | 4.8 (3.3–8.3) | 5.9 (4.0–8.4) | 0.41   |
|           | 1 month    | 5.2 (3.5–10.2) | 6.0 (4.0–8.3) | 0.55   |
|           | 7 months   | 5.6 (3.1–8.7) | 6.9 (4.3–9.2) | 0.29   |
| MPO, ng/mL | Baseline | 10.5 (6.7–37.3) | 7.8 (5.4–13.4) | 0.13   |
|           | 1 month    | 11.7 (5.2–17.0) | 9.3 (5.2–12.6) | 0.42   |
|           | 7 months   | 10.5 (7.5–22.3) | 7.8 (5.5–12.2) | 0.014* |
| MMP-9, ng/mL | Baseline | 63.1 (28.4–117.2) | 44.1 (27.2–84.5) | 0.29   |
|           | 1 month    | 41.8 (27.0–60.4) | 43.7 (31.6–56.9) | 0.94   |
|           | 7 months   | 63.3 (36.0–111.9) | 41.5 (25.8–67.4) | 0.09   |
| S100 beta, ng/mL | Baseline | 4.1 (1.1–10.3) | 1.3 (0.8–9.4) | 0.24   |
|           | 1 month    | 2.1 (1.0–6.9) | 1.3 (0.6–3.1) | 0.15   |
|           | 7 months   | 2.0 (0.9–7.9) | 1.2 (0.6–2.2) | 0.15   |

Data are median (IQR) values.
*p<0.05.
CD40L: CD40 ligand, ICAM: intercellular adhesion molecule, IL: interleukin, MCP: monocyte chemoattractant protein, MDC: macrophage derived chemo-
kine, MMP: matrix metalloproteinase, MPP: myeloperoxidase, PAI: plasminogen activator inhibitor, PDGF: platelet-derived growth factor, SAA: se-
rum amyloid A, TNF: tumor necrosis factor, VCAM: vascular-cell adhesion molecule, VEGF: vascular endothelial growth factor.
Table 3. Results from the multivariate analysis of the progression of symptomatic intracranial atherosclerotic stenosis

| Parameter                              | Value    | p*         |
|----------------------------------------|----------|------------|
| Continuous PDGF-AB/BB at 7 months, per 1-ng/mL increase | 1.07 (1.02–1.13) | 0.02      |
| Tertiles of PDGF-AB/BB at 7 months     |          |            |
| First (<1.63 ng/mL)                    | Reference|            |
| Second (1.63–7.16 ng/mL)              | 2.43 (0.51–11.51) | 0.26      |
| Third (>7.16 ng/mL)                   | 6.93 (1.53–31.38) | 0.01      |

Data are adjusted odds ratio (95% confidence interval) values.

*Adjusted for P-selectin and MPO at 7 months, †Odds ratio was calculated in separate models for continuous and categorical PDGF-AB/BB.

MPO: myeloperoxidase, PDGF: platelet derived growth factor.

reported to be associated with atherosclerosis.

In conclusion, the PDGF-AB/BB level is associated with the progression of symptomatic ICAS, suggesting that PDGF-AB/BB plays a role in the progression of human ICAS. Although the causal relationship remains to be elucidated, this result may have clinical implications, and it should be investigated in further studies of PDGF-AB/BB.

Author Contributions

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Conflicts of Interest

The authors have no potential conflicts of interest to disclose.

Acknowledgements

Korea Otsuka Pharmaceutical (KOP) Company. Korea Otsuka International Asia, and Arab Co, Ltd. provided financial support for the TOSS-2 study. However, these organizations played no role in protocol development, data collection, analysis, or manuscript preparation for the current study.

This work was also supported by the Dongguk University Research Fund of 2015.

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