Review
How the Tumor Micromilieu Modulates the Recruitment and Activation of Colorectal Cancer-Infiltrating Lymphocytes

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Abstract: The successful treatment of advanced colorectal cancer disease still represents an insufficiently solved clinical challenge, which is further complicated by the fact that the majority of malignant colon tumors show only relatively low immunogenicity and therefore have only limited responsiveness to immunotherapeutic approaches, such as, for instance, the use of checkpoint inhibitors. As it has been well established over the past two decades that the local tumor microenvironment and, in particular, the quantity, quality, and activation status of intratumoral immune cells critically influence the clinical prognosis of patients diagnosed with colorectal cancer and their individual benefits from immunotherapy, the enhancement of the intratumoral accumulation of cytolytic effector T lymphocytes and other cellular mediators of the antitumor immune response has emerged as a targeted objective. For the future identification and clinical validation of novel therapeutic target structures, it will thus be essential to further decipher the molecular mechanisms and cellular interactions in the intestinal tumor microenvironment, which are crucially involved in immune cell recruitment and activation. In this context, our review article aims at providing an overview of the key chemokines and cytokines whose presence in the tumor micromilieu relevantly modulates the numeric composition and antitumor capacity of tumor-infiltrating lymphocytes.

Keywords: colorectal cancer; tumor-infiltrating lymphocytes; chemokines; cytokines

1. Introduction
Colorectal cancer (CRC) represents the third most common malignant tumor disease worldwide and was found to be the underlying diagnosis in almost 10% of all cancer-related deaths in 2020 [1]. In the subgroup of CRC patients who have already developed distant metastases (stage IV disease), the average 5-year relative survival rate is only 12% [2]. Therefore, significant scientific efforts are being made to gain a more detailed understanding of the CRC-specific pathogenesis and to translate this into clinically applicable strategies for improved early diagnosis and optimized therapy [3].

It is well accepted that the tumor microenvironment and, in particular, the local presence of immune and stromal cells critically determine CRC prognosis [4]. Already in 2005, in a pioneering study published by the team of Jérôme Galon, it was demonstrated that the accumulation of early memory and effector memory CD8+ T cells in the tumors of CRC patients correlated significantly with an improved clinical outcome (absence of early metastatic invasion, a less advanced pathological stage, and increased survival) [5]. Meanwhile, this recognized relevance of the tumors’ immune profile for CRC prognosis has been successfully translated into clinically applied diagnostic concepts. For instance, the Immunoscore, a standardized immune assay based on the immunohistochemical staining of tumor-infiltrating CD3+ and CD8+ T cells in combination with automated artificial intelligence-assisted signal detection and interpretation, has found its way into
international consensus guidelines for the clinical management of CRC disease and has been proven to refine prognosis and significantly improve CRC risk assessment [6,7].

However, although the development of CRC usually results in the induction of profound antitumor immune responses indicated, for instance, by the presence of tumor antigen-specific adaptive immune cells in the peripheral blood of affected patients [8], the local tumor microenvironment often succeeds in generating local tumor-beneficial conditions, which are able to dampen the accumulation and cytotoxic activity of infiltrating effector T cells and to promote the immune escape properties of tumor cells [9]. As a prime example, DNA mismatch repair-proficient or microsatellite-stable CRC colon cancer, which represent around 85% of all diagnosed CRC diseases, are characterized by low overall immunogenicity and the decreased infiltration of CD8\(^+\) T lymphocytes [10–12]. The phenomenon of tumor immune escape includes, for instance, the enhanced infiltration of myeloid-derived suppressor cells and suppression-competent FoxP\(^{\text{high}}\) regulatory T cells (T\(_{\text{reg}}\) cells), which results in suppressed effector T cell function and was described to be associated with CRC progression, lymphatic invasion, and metastasis [13,14]. As we will consider T\(_{\text{reg}}\) cells in the following as an antitumor immune response-suppressing and thus colon tumor growth-promoting T cell subpopulation, it is important to mention, at least briefly, the functional heterogeneity of FoxP3\(^{\text{high}}\) CD45RA\(^{-}\) T cells in the CRC microenvironment [15]. While CRC-infiltrating FoxP3\(^{\text{high}}\) CD45RA\(^{-}\) T cells are characterized by a good capacity to regulate and suppress effector T cell function, FoxP3\(^{\text{low}}\) CD45RA\(^{-}\) T cells turned out to be non-suppressive but produced pro-inflammatory cytokines [15]. The intratumoral co-existence of suppression-competent FoxP3\(^{\text{high}}\) T\(_{\text{reg}}\) cells and FoxP3\(^{\text{low}}\) non-T\(_{\text{reg}}\) cells might most likely be responsible for the partly controversial discussion about the prognostic role of CRC-infiltrating FoxP3\(^{+}\) T cells [16,17].

Understanding that the fate of tumor disease is significantly influenced by the efficiency of the antitumor immune response and the overcoming of immune-evasive local conditions has also shaped the development of modern therapeutic strategies for the treatment of CRC patients. While classic chemotherapeutic treatment regimens predominantly target the proliferation and survival of tumor cells directly, more recent immunotherapeutic approaches instead aim at supporting anticancer immunity by modulating the tumor microenvironment. Thus, the principle of immunotherapy is primarily to increase the number of tumor-infiltrating and tumor antigen-specific cytotoxic T lymphocytes (e.g., by adaptive cell transfer [18,19] or by optimized recruitment of systemic effector cells) and to increase their cytolytic activity (e.g., by checkpoint inhibitors [11] or by inhibition of immunosuppressive components). However, a large proportion of CRC patients show only a rather disappointing response to monotherapy with clinically established PD-1 checkpoint inhibitors [11]. Thus, it currently remains an important challenge to better understand CRC-specific aspects in the interaction between tumor cells and their local microenvironment (including accumulating immune cells) in order to develop innovative immunotherapeutic strategies to optimize the efficacy of the immune system in controlling intestinal tumor growth.

2. Prognostic-Relevant Chemokine Signaling Impacting the Immune System-Mediated Control of CRC

The demonstrated relevance of tumor-infiltrating lymphocytes to the fate of CRC disease has turned our attention also to those mediators in the close tumor environment that, through their chemotactic capacity, underlie the local accumulation of T cells and are able to influence the ratio between tumor-suppressing T lymphocytes and tumor-promoting T\(_{\text{reg}}\) cells. Several chemokines (e.g., CCL1, CCL3, CCL4, CCL7, CCL20, CCL25, CCL26, CX3CL1, CXCL1, CXCL8, CXCL9, CXCL10, and CXCL16) and partly also their respective G protein-coupled receptors (e.g., CCR1, CCR5, CCR6, CCR8, CXCR2, CXCR6, and CX3CR1) were found to be upregulated or selectively expressed in human colon cancer tissue compared to corresponding non-tumor control tissue [20–25]. Moreover, during the last two decades, significant progress has been achieved in defining specific chemokines
or chemokine receptors whose intratumoral expression profile might allow a prediction of tumor prognosis in individual CRC patients. For instance, high expression levels of CXCR3, CXCR4, and CCL20 could be identified as a predictor of poor prognosis [26–29], while increased levels of CXCL16, CCL4, CX3CL1, CXCR6, CX3CR1, and CCR5 in CRC lesions turned out to indicate prolonged disease-free survival and were mostly associated with the profound accumulation of effector T cells in the tumor [4,21,30–33] (Figure 1).

Figure 1. Chemokine signaling of described relevance for the immune system-mediated control of CRC. Abbreviations: TAM = tumor-associated macrophages; CTL = cytotoxic T lymphocytes; ↑ = increased; ↓ = decreased.

2.1. CXCR3

CXCL9, CXCL10, and CXCL11 are ligands of the chemokine receptor CXCR3, which is preferentially expressed by CD4+ and CD8+ T cells, but could also be detected on the surfaces of cancer cells [34]. In tumors of individual CRC patients, a strong degree of co-expression has been described for all three of these CXCR3 ligands, whose gene transcription in tumor cells and tumor-associated stromal cells (e.g., endothelial cells and fibroblasts) can obviously be triggered by the presence of interferons and the pro-inflammatory cytokine TNFα [31]. The enhanced intratumoral expression of CXCL11, as the presumably most potent CXCR3 agonist [34], turned out to correlate with the increased presence of CD4+ T helper (Th) cells and CD8+ cytotoxic T cells in human CRC lesions. Regarding the impact of CXCL11 on the different Th cell subtypes, its presence in CRC did not correlate with the accumulation of FoxP3+ Treg cells and additionally performed transcriptional analyses implicated its particular relevance for T-bet-expressing Th1 cells, strongly pointing to a supportive role of CXCR3 engagement during the adaptive immune system-mediated tumor control [31]. This is also in accordance with another study describing the particular relevance of CXCL9 and CXCL10 for the intratumoral enrichment
of memory CD8\(^+\) T cells and the finding that CXCL10 secretion in colon cancer tissue was found to correlate with the frequency of granzyme B-expressing CD8\(^+\) tumor-infiltrating lymphocytes [30,35]. However, in contrast to its ligands, CXCL9, CXCL10, and CXCL11, the protein expression of the CXCR3 chemokine receptor protein in colon cancer tissue could be associated with increased tumor size, the occurrence of lymph node or distant metastasis, and poor survival [28]. Most likely, the implicated tumor-promoting effects of CXCR3 can be explained by the tumor cell autonomous expression of this chemokine receptor and its described positive impact on cancer cell motility, resulting in an increased metastasis-forming capacity [36,37]. In this context, CXCL4, as another CXCR3 ligand, might be of significant relevance. Indeed, increased expression of CXCL4 in tumor tissue indicated a decreased survival time in CRC patients [38], and experimental in vivo CRC models described the capacity of this chemokine to promote the proliferation and activation of T\(_{\text{reg}}\) cells, while dampening CD8\(^+\) T cell-mediated antitumor immune responses [39]. Thus, it becomes crucial to consider and further elucidate the, at least partially, ambiguous and target cell-specific role of CXCR3 signaling [36,40] when discussing this chemokine receptor and its ligands as potential targets for CRC immunotherapy.

2.2. CXCR6

Unlike this complex role of CXCR3 signaling in the context of CRC disease, which apparently can directly influence tumor cell behavior in addition to immune cell recruitment, chemokine receptors with a more restricted expression pattern limited to specific types of immune cells may represent more favorable therapeutic targets, in the sense that their effect on the fate of tumor development can be more precisely predicted. For instance, the chemokine receptor CXCR6 is expressed almost selectively by CD4\(^+\) T cells, CD8\(^+\) T cells, NKT cells, NK cells, plasma cells [41], while no or only very weak CXCR6 protein expression could be detected in colon cancer cells and, of particular relevance, its increased expression and the enhanced presence of its unique ligand CXCL16 in colon tumor tissue turned out to be associated with marked T cell infiltration and improved tumor prognosis [25,32,42]. Although human colon cancer tissue characterized by high expression levels of CXCL16 showed increased infiltration by CD4\(^+\) and CD8\(^+\) lymphocytes, data acquired in an experimental in vivo model of CRC metastasis interestingly indicated the particular relevance of the CXCL16/CXCR6-driven NKT cell recruitment for the control of hepatic CRC metastasis, while the beneficial effect of CXCL16 in this context seemed to be mostly independent of the presence or absence of CD8\(^+\) T cells [42]. However, CXCR6 expression can obviously be induced on the surface of colon tumor-infiltrating CD8\(^+\) T cells by yet-to-be-defined mediators of the local microenvironment, and CXCR6\(^+\) CD8\(^+\) T cells in primary human CRC lesions could recently be characterized by the marked expression of cytotoxic effector markers, indicating their potent antitumor effector function [25]. Indeed, the absence of CXCR6 in the transplantable MC38 colon tumor model drastically reduced the response to immune checkpoint therapy and decreased the number and cytotoxic effector activity of tumor-infiltrating CD8\(^+\) T cells [25]. Thus, the ex vivo selection or enrichment of CXCR3\(^+\) CD8\(^+\) T cells appears as a promising strategy for optimizing the efficacy of therapeutic concepts, which are based on the adoptive transfer of tumor-targeting T cells in the clinical context of CRC [25].

2.3. CX3CR1

Regarding CX3CL1 and its receptor CX3CR1, recent data interestingly indicated that antigen-specific tumor-infiltrating CD8\(^+\) T cells expressing high levels of CX3CR1 show a lower proliferative capacity upon antigenic rechallenge than their CX3CR1-negative counterparts and might be less relevant for the efficient immunological control of tumor growth [43]. However, considering the mentioned positive association between CX3CL1 and tumor prognosis [30,44], as well as its inverse correlation with lymph node metastasis [23] and the fact that the absence or decreased expression of the CX3CR receptor itself was found to be associated with the development of lymph node metastasis and
poor CRC prognosis [4], it can be concluded that this chemokine/chemokine receptor interaction relevantly influences the fate of CRC disease by means other than triggering the antitumor activity of recruited immune cells [23]. Indeed, however, taking advantage of the fact that CX3CL1 seems to be present in more than 80% of colorectal tumors but showed a significantly lower expression level in the normal colon mucosa [22,23], ectopic expression of its receptor CX3CR on ex-vivo-expanded T cells designated for adoptive T cell immunotherapy has been discussed and successfully validated in vivo in a humanized CRC mouse model as a strategy to guide the transferred T cells from the peripheral circulation into the tumor tissue [22].

2.4. CCR5

As another chemokine typically expressed in the intestinal mucosa and usually upregulated in colorectal cancer tissue, CCL4 (previously known as macrophage inflammatory protein (MIP)-1β) is mainly produced by hematopoietic cells and, besides mediating the chemoattraction of macrophages and NK cells, also promotes the intestinal recruitment of T cells via its receptor CCR5 [45–47]. In addition to tumor-infiltrating immune cells, colon tumor cells might represent a CRC-intrinsic and β-catenin-regulated source of CCL4 [48], as suggested in analogy to a similar phenomenon observed in the context of cutaneous melanoma disease [49]. Although experimental in vivo administration of recombinant CCL4 failed to promote the cytolytic activation of mucosal CD8+ T cells and preferentially triggered Th2 cytokine responses [50], the intratumoral release of CCL4 can be expected to promote the recruitment of CD103+ dendritic cells and to subsequently enhance T cell priming against tumor-associated antigens [49]. Accordingly, increased protein expression levels of the CCL4-regulating protein β-catenin and low mRNA levels of CCR5 in CRC tissue turned out to be associated with decreased numbers of tumor-infiltrating CD8+ T cells [33,48] and, moreover, low CCL4 protein expression levels in colon tumor tissue were associated with a significantly decreased cancer-specific survival rate in CRC patients [21]. Despite this obviously beneficial role of CCL4 for the immunological control of CRC disease, it cannot be neglected that colon tumor-infiltrating regulatory T cells (Treg cells) could be characterized by significantly increased expression of the CCL4 receptor CCR5 compared to tumor-infiltrating conventional T cells but also in comparison to Treg cells located in the mucosa of the distal colon [47]. As the targeted migration of CRC patient-derived Treg cells towards CCL4 could be demonstrated in vitro [47], it thus appears likely that CCL4 can also support the Treg cell-mediated immune evasion of colorectal tumors. Indeed, in the CT26 mouse colon cancer model, which is based on the subcutaneous injection of colon tumor cells, CCR5-deficient mice could be characterized by significantly decreased numbers of tumor-infiltrating Treg cells and reduced tumor growth [51]. In the human system, the intratumoral protein expression of CCR5 markedly varies between individual CRC patients, being absent or only weakly expressed in almost 50% or 30% of patients, respectively. In accordance with the previously described beneficial role of the CCR5 ligand CCL4 in the immunological control of colonic tumorigenesis, CRC tumors with intermediate to strong CCR5 expression seemed to have a better prognosis, indicated by Union International Contre Cancer (UICC) staging and less frequent lymph node involvement [33]. However, CCL4 is not the only ligand of CCR5. In particular, CCL3 and CCL5 should also be considered as potential modulators of antitumor immune responses in the context of CRC, with CCL3 being significantly upregulated on the mRNA and protein level in colon tumors compared to tumor-free tissue of the distal colon [47] and CCL5 showing enhanced secretion in colon tumors whose gene expression profile indicated increased type-1 T cell activity [35]. Although CCL5 does not seem to mediate direct chemotactic effects on CD8+ T cells, its absence in orthotopic CRC tumor models interestingly resulted in the marked accumulation of intratumoral effector CD8+ T cells and in significantly reduced tumor growth [52]. These in-vivo-acquired data implicated a tumor-promoting role of CCL5, which can, at least partly, be explained by its capacity to positively influence the secretion of the calcium-binding protein S100a9 by tumor-associated macrophages (TAMs) [52], and
thereby to suppress T cell activation and proliferation [53]. Moreover, CCL5 could be identified as a potent promoter of intratumoral T\textsubscript{reg} accumulation and activation, which also significantly enhances their ability to kill CD8\textsuperscript{+} T cells and to dampen the T cell-driven antitumor immune response [51]. The concept that the colon tumor-promoting effect of CCL5 crucially depends on the involvement of the adaptive immune system was further supported by the observation that the inoculation of CCL5-deficient colon tumor cells into immunocompetent recipient mice, but not into B and T cell-lacking Rag1-deficient animals, resulted in impaired tumor growth [51]. Similar to CCL5, CCL3 has also been linked to increased colon tumor development, although, at least to our knowledge, no relevant impact of this CCR5-binding chemokine on the T cell-mediated control of colon cancer has been described so far. Obviously, it is more the potency of CCL3 to trigger the colonic accumulation of cancer-associated fibroblast and the subsequent production of fibroblast-derived growth factors that underlies its pro-tumorigenic character [54]. Regarding the ability of CCL3 to promote the recruitment of fibroblasts, it should be mentioned that cancer-associated fibroblasts represent an important source for the chemokine CXCL12, which has also been linked to an immunosuppressive and thus tumor-promoting microenvironment in the context of CRC and whose receptor CXCR4 showed increased expression levels in the tumor tissue of CRC patients with poor clinical prognosis [29,55,56]. Overall, the described involvement of the CCR5 ligands CCL5 and CCL3 in CRC-driven immune escape mechanisms and their tumor-promoting properties seemed to outweigh the capacity of CCL4 to trigger the recruitment of peripheral effector T cells in CRC metastases via CCR5 engagement [57]. Accordingly, the therapeutic use of the CCR5 inhibitor maraviroc in patients with advanced-stage metastatic colorectal cancer resulted in the marked induction of tumor cell death and decreased tumor cell proliferation within metastatic lesions, while the number of infiltrating CD8\textsuperscript{+} T cells detected in the invasive margin of liver metastasis remained mostly stable [57].

2.5. CCR6

Also of particular interest in the context of metastatic CRC disease, elevated levels of the chemokine receptor CCR6 in primary CRC lesions were found to be associated with the presence of liver metastases [58]. In accordance with these data, another study demonstrated that increased expression of the CCR6 ligand CCL20 in colon tumor tissue was associated with worse overall survival of CRC patients under neoadjuvant chemotherapy [27]. CCL20 is produced by colon cancer cells and tumor-associated macrophages and its release into the tumor microenvironment can even be intensified by the standard chemotherapeutic agent 5-fluorouracil [27,59]. As the suggested key mechanism underlying the enhancing influence of CCL20/CCR6 on CRC progression and chemoresistance, CCL20 is able to trigger the intratumoral accumulation of CCR6\textsuperscript{+} T\textsubscript{reg} cells and, thereby, to counteract the antitumor immune response [27,59]. In some analogy to the enhanced CCL20-driven T\textsubscript{reg} recruitment during neoadjuvant chemotherapy, surgical interventions might also bear the risk of promoting local T\textsubscript{reg} accumulation in CRC and thereby tumor progression, with CCL18 discussed as the responsible chemokine [60].

3. Cytokines Influencing the Efficacy of the Antitumor Immune Response

While the primary role of chemokines is to induce and coordinate the recruitment of immune cells into the tumor microenvironment, cytokines serve predominantly as modulators of the activation, differentiation, and function of tumor-infiltrating immune cells [13,61] (Figure 2). Although a plethora of different cell types within the intestinal tumor tissue—such as, for instance, innate immune cells, cancer-associated fibroblasts, and dysplastic epithelial cells—relevantly contribute to and/or depend on the local cytokine network, tumor-infiltrating lymphocytes seem to function as the main actors in this context [3,13]. Locally accumulated CD4\textsuperscript{+} and CD8\textsuperscript{+} T cells do not only represent an important source for the release of pro- and anti-inflammatory cytokines into the tumor microenvironment, but are also key targets of intratumoral cytokine signaling. Dependent on their differentiation
into distinct T helper cell subsets, T lymphocytes can either function as potent promoters of the antitumor immune response (e.g., Th1 cells and cytotoxic T lymphocytes (CTLs)) or support the further progression of tumor development (e.g., Th17 cells, Th22 cells, and Treg cells) [13].

Figure 2. Cytokines of described relevance for the immune system-mediated control of CRC. Abbreviations: PD-L1 = programmed death ligand 1; TAM = tumor-associated macrophages; CTL = cytotoxic T lymphocytes; MDSC = myeloid-derived suppressor cell; ECM = extracellular matrix; ↑ = increased; ↓ = decreased.

3.1. IFNγ

It is well established that the pro-inflammatory cytokine IFNγ mediates potent antitumor effects. In addition to its direct anti-proliferative impact on colon tumor cells [62] and its capacity to inhibit tumor-associated angiogenesis [63], IFNγ particularly supports the immune cell-driven tumor defense, including the MHC-I- and MHC-II-dependent presentation of tumor antigens and the activation, expansion, and survival of tumor-infiltrating effector T cells [13,64]. Accordingly, the presence of Th1 cells, NK cells, and CD8+ T cells,
as the main cellular producers of the pro-inflammatory cytokine IFNγ, in colon tumors correlates with improved clinical prognosis [5,65,66], while downregulation of the IFNR1 chain of type I interferon on the protein level in the cancer cell compartment or the stromal cell compartment of CRC tumors was found to be associated with poor disease prognosis [67]. Interestingly, the microenvironmentally induced degradation of the IFNR1 protein in tumor-associated stromal cells and particularly in CD8+ T cells could recently be identified as a common process during CRC development, which is required for efficient tumor growth [67]. Analyses in the murine MC38 colon cancer model clearly indicated that the inhibition of the local IFN signaling via degradation of IFNAR1 resulted in the significantly decreased survival of tumor-infiltrating CD8+ T cells as the key cytolytic mediators of the immunological tumor control, and in the subsequent downregulation of cytolytic effector genes such as granzyme B and IFNγ [67]. Moreover, experimental data from the same study strongly suggested that the targeted stabilization of IFNRI expression in colon tumors might improve the efficacy of anti-PD-1 therapy [67].

Despite the traditionally assumed and well-validated antitumor capacity of IFNγ, there is also growing evidence for a more ambiguous role of this cytokine in the fine-tuning of CD8+ T cell-driven CRC defense. In particular, the ability of IFNγ to promote the expression of PD ligand 1 (PD-L1) in cancer cells is considered evidence that it also has immunosuppressive properties. Indeed, the intratumoral expression of PD-L1 in a cohort of 181 CRC patients was found to be positively associated with the mRNA levels of IFNG and its downstream signaling molecules JAK2 and STAT1, and, on a functional level, IFNγ was able to induce enhanced PD-L1 expression in human colorectal cancer cells, thus strongly suggesting its supportive impact on the tumor cell-driven initiation of immune escape mechanisms [68]. In this context, it is noteworthy that a recent study pointed to the histone methyltransferase WHSC1 as an essential mediator of the IFNγ-promoted MHC-I expression in colon cancer cells, whose capacity to epigenetically modulate gene transcription did not seem to be involved in the IFNγ-induced expression of PD-L1. The identification of strategies to promote the enzymatic activity of WHSC1 in colon tumors thus appeared as a promising strategy to trigger selectively the antitumor properties of IFNγ in colon tumors without enhancing its potential involvement in immunosuppressive processes [69].

3.2. IL-17A and IL-17F

Almost two thirds of primary sporadic CRCs were estimated to show increased expression of IL-17A. In general, the enhanced presence of this cytokine was found to be associated with worse disease prognosis and more severe dysplasia [70–72]. In line with this, its deficiency in the azoxymethane (AOM)/dextran sodium sulfate (DSS) model of colitis-associated cancer significantly reduced the intestinal tumor burden. In contrast, augmented experimental colon tumor induction has been described in IL-17F-deficient mice, suggesting an at least partially opposing role of these two IL-17 family members in the pathogenesis of colon cancer [72]. In addition to distinguishing between the different members of the IL-17 family, it is further essential to differentially consider their influence on the various pathogenetic aspects involved in the complex process of CRC development—for example, their effect on tumor cells, angiogenesis, or immunological tumor defense. Taking into account the thematic focus of our article, which is the modulation of antitumor immune responses, it should only be briefly mentioned here that for IL-17A, the ability to increase tumor cell growth and tumor-associated neoangiogenesis has been described in the context of CRC, whereas IL-17F seems to influence both processes in the opposite direction [72]. In addition to the prognostically nonbeneficial role of IL-17A in the local microenvironment of CRC, CRC patients could be characterized by an increased frequency of IL-17+ CD4+ T cells in the peripheral blood [73], and increased serum levels of IL-17A in patients with advanced-stage colon tumors have been linked to the reduced tumor infiltration of CD8+ T cells [74]. Of mechanistical relevance, advanced-stage CRC patients could also be characterized by the significantly decreased expression of the chemokine receptor CXCR3 on the surfaces of
circulating CD8+ T cells, while the CXCR3 ligand CXCL10 turned out to be upregulated in the tumor tissue. Together with the described capacity of IL-17A to inhibit the expression of CXCR3 in CD8+ T cells via STAT3 signaling, the acquired results thus implicated that Th17 cell-released IL-17A is able to dampen the immunological control of CRC disease via the IL-17A/STAT3/CXCR3 axis [74]. Together with γδT cells, Th17 cells appeared as the predominant source of IL-17 in the micromilieu of colon tumors, where their development turned out to be particularly dependent on the transcription factor Batf [14,75,76]. As already indicated by the description of the partly very different effects of IL-17A and IL-17F on CRC pathogenesis, it is obviously far too simplistic to automatically conclude from the pro-tumorigenic effect of IL-17A that an increased presence of Th17 cells in tumor tissue might be an indicator of poor disease prognosis. Rather, CRC-infiltrating Th17 cells showed some Smad7-induced plasticity towards T-bet/RORγt double-positive T cells characterized by low IL-17A and increased IFNγ expression [77] and could in general be described as multifunctional modulators of the local antitumor immune response, which are able to secrete a broad spectrum of different cytokines and chemokines in addition to IL-17 and, thereby, to support also important mechanisms of tumor defense [75]. In accordance with the described double-edged role of Th17 cells in the context of CRC, the number of tumor-infiltrating IL-17+ cells did not allow the disease outcome to be predicted [75]. However, the increased intratumoral frequency of the specific fraction of IL-17+ γδT cells indeed seemed to be associated with clinical parameters such as enhanced tumor size, vascular invasion, and lymph node metastasis, thus indicating CRC invasiveness and progression [14]. As with Th17 cells, tumor-infiltrating γδT cells do not exclusively produce IL-17 but are also considered as an important source of cytokines such as IL-8, GM-CSF, and TNFα, enabling them, in particular, to recruit polymorphonuclear myeloid-derived suppressor cells [78] into the tumor tissue and, thus, to contribute significantly to the establishment of an immunosuppressive and tumor-promoting milieu [14].

3.3. IL-10

As one of the cytokines integrally involved in the formation of an immunosuppressive tumor micromilieu, IL-10 was found to be markedly upregulated on the mRNA level in CRC-infiltrating Treg cells compared to colorectal polyps and control colon tissue derived from healthy volunteers, and increased serum levels of IL-10 could be linked to therapy failure, decreased survival, and post-surgical disease recurrence in CRC patients [79–82]. Besides Treg cells, i.e., Th2 cells, monocytes, intestinal macrophages, and colon tumor cells are relevant sources of IL-10 in the context of CRC [80]. In accordance with its prognostically negative role, experimental silencing of IL-10 secretion in the local environment of MC38 tumors was able to inhibit tumor growth and to improve the efficacy of combined chemoimmunotherapy by reducing the local accumulation of myeloid-derived suppressor cells and Treg cells and by triggering Th1-mediated antitumor immunity [83,84]. However, in the APCmin−/+ model of sporadic CRC, IL-10 seemed to be responsible for the capacity of adoptively transferred Treg cells to inhibit intestinal tumor development and growth. It can thus be hypothesized that IL-10 mediates a growth/survival-supporting effect directly on intestinal tumor cells and/or damps the activity of effector CD4+ T cells in tumor-promoting chronic inflammatory processes [85,86], whereby both effects would oppose the experimentally demonstrated suppressive influence of IL-10 on the antitumor immune response and its negative prognostic relevance in human CRC disease. Thus, further studies are needed to better decipher the distinct and partly contrary effects of IL-10 on the different aspects of intestinal tumor disease and possibly to be able to therapeutically target them separately in the future.

3.4. IL-6

Increased concentrations of IL-6 in the peripheral blood of CRC patients turned out to be associated with an advanced tumor stage, the development of metastasis, tumor recurrence, and reduced survival [87–90]. Accordingly, the significantly diminished growth
of experimentally induced orthotopic or inflammation-associated colon tumors could be observed in IL-6-deficient mice compared to tumor development in IL-6-proficient control mice. Moreover, tumor growth in wild-type mice could successfully be inhibited by anti-IL6R treatment [91–93]. IL-6 can be secreted by a large panel of different immune and non-immune cells [94], whereby lamina propria CD4+ T cells and, in particular, PU.1-driven Th9 cells, dendritic cells, and tumor cells could be identified as the main producers of this cytokine in colon tumors [93,95]. Mechanistically, the binding of IL-6 to the IL-6 receptor (IL6R) and the subsequent association with the glycoprotein gp130 is known to initiate STAT3 signaling in immune cells and epithelial cells [93]. The suggested unbeneﬁcial inﬂuence of IL-6 on the fate of CRC disease was mainly attributed to its inhibitory effects on T cell-mediated aspects of antitumor immunity, as well as to its proliferative and anti-apoptotic effects on colon tumor cells, which are mainly mediated via the soluble IL-6 receptor (sIL6R) and IL-6 trans-signaling [93,96]. Regarding its dampening inﬂuence on the antitumor immune response, it has been demonstrated that the absence of IL-6 in host mice in an orthotopic CT26 colon tumor model resulted in the increased expression of MHC class I molecules on the surfaces of tumor cells and in the intratumoral accumulation of mature dendritic cells, CD4+ helper T cells, and CD8+ cytolytic effector T cells [91]. Moreover, for the same colon tumor model, another study described the additional IL-6-triggered regulation of the number of tumor-inﬁltrating Treg cells and myeloid-derived suppressor cells, whereby overexpression of the cytokine in tumor cells resulted in the increased detectability of these two immunosuppressive cell types [87]. Moreover, in the context of colitis-associated colon cancer, IL-6 is known to shift macrophage polarization towards tumor-promoting M2 macrophages [97]. However, for completeness, it should also be mentioned that despite its predominantly pro-tumorigenic function, IL-6 has been described as an important activator of the macrophage-mediated phagocytosis of colon tumor cells, driving the required cytoskeleton rearrangement [98]. Moreover, in general, there is not only a unilateral inﬂuence of IL-6 on the composition and activation of the CRC immune microenvironment, but IL-6-mediated signaling is conversely also controlled by components of the tumor micromilieu. For instance, TGFβ could be identiﬁed as a potent inhibitor of IL-6-mediated tumor-supportive effects [93], whereas Th9 cell-released IL-9 is apparently able to increase I-L6 and IL-9 production by intratumoral Th9 cells in an auto-amplifying loop [95].

The fact that, in parallel to the upregulation of MHC class I molecules in the IL-6-deﬁcient host milieu, a concomitant increase in PD-L1 expression was observed on the surface of IL-6-proficient CT26 tumor cells [91], implied that the therapeutic blockade of IL-6 signaling can potentially improve the responsiveness of CRC patients to checkpoint inhibitor therapies. Notably, the quality of the IL-6-mediated regulation of PD-L1 expression in colon tumor cells obviously depends on the source of IL-6 production, as the signiﬁcant induction of this immunoregulatory molecule on the tumor cell surface could be detected in a similar manner either after the overexpression of IL-6 in the tumor cell compartment [87], after antibody-mediated systemic IL-6 blockade [92], or in the absence of IL-6 in the surrounding non-tumor cells [91]. However, in vivo ﬁndings provided experimental evidence for the synergistic and beneﬁcial therapeutic effects of combined IL-6/ PD-L1 blockade in the CT26 colon cancer model [87,91,92].

3.5. IL-12 Family of Cytokines

The IL-12 family of cytokines consists of IL-12, IL-23, IL-27, and IL-35, with IL-12, IL-23, and IL-27 being mainly produced by macrophages and dendritic cells, while IL-35 is predominantly secreted by regulatory T cells but can also be expressed by tumor cells [99–101]. All four family members have in common that they represent heterodimers consisting of an α chain and an αβ chain and that their particular receptors initiate the JAK-STAT signaling pathways, while they markedly differ with regard to their immunological function and show a unique heterodimeric combination of their subunits. IL-12 consists of a p35 and a p40 chain, IL-23 comprises the p19 and the p40 protein, IL-27 was defined as
a heterodimer of p28 and EBI3, and, in the most recently identified family member, IL-35 EBI3 forms a complex with p35 [101,102].

IL-12 is well known as a classic mediator of antitumor immunity, mainly attributed to its capacity to promote Th1 cell and CD8\(^+\) T cell differentiation and NK cell activation and, subsequently, to enhance IFN\(\gamma\) secretion, which is required for the efficient presentation of tumor antigens, T cell recruitment to the tumor site, and the intratumoral survival and expansion of cytolytic effector T cells [103–108]. Accordingly, the experimentally induced local increase in IL-12 levels in the tumor microenvironment showed clear therapeutic efficacy in different in vivo models of CRC disease and colon cancer metastasis [101,105,106,109,110]. Interestingly, a successful therapeutic response to local IL-12 therapy in a model of hepatic colon cancer metastasis was associated with increased expression of the IL-15 receptor alpha (IL15Ra) in tumor-specific CD8\(^+\) T cells, indicating the significant relevance of IL-12 for IL-15-supported long-lasting memory CD8\(^+\) T cell responses [111]. In accordance with this link between IL-12 and IL-15, there also exists experimental evidence that some of the IL-12-mediated immunological antitumor effects can even be observed in the absence of IFN\(\gamma\), but then require the presence of IL-15 [112]. Moreover, another study reported that mice with a good therapeutic response to combined cyclophosphamide plus IL-12 induction therapy in the orthotopic CT26 colon cancer model were characterized by significantly decreased numbers and immunosuppressive activity of systemic and tumor-infiltrating T\(_{\text{reg}}\) cells [113]. Despite promising preclinical data, clinical studies performed so far on the applicability of IL-12-based strategies for the treatment of CRC are relevantly limited by the high level of toxicity associated with the systemic delivery of the cytokine [114] and fail to report a significant therapeutic advantage [101,115]. As a promising approach to potentially overcome the limitations caused by the poor systemic tolerability of IL-12, a recent preclinical study described a marked beneficial effect of intratumoral IL-12p70 mRNA therapy on tumor growth and tumor regression in the MC38 colon tumor model. Indeed, the intratumoral delivery of lipid nanoparticle-encapsulated IL-12p70 mRNA resulted in the increased secretion of IL-12p70 protein by the colon tumor cells and, subsequently, resulted in a Th1 shift in the tumor microenvironment, triggering IFN\(\gamma\)-driven and CD8\(^+\) T cell-dependent antitumor immunity [116].

In a quite similar way as described for IL-12, overexpression of IL-27 in colon cancer cells was able to trigger local IFN\(\gamma\) secretion, NK cell-mediated antitumor effects, and T cell-mediated tumor-specific cytotoxicity [117]. In particular, in CD4\(^+\) T cells and NK cells, but also in CD8\(^+\) T cells, a marked mRNA expression of the alpha subunit of the IL-27 receptor alpha could be detected [101]. Although experimental overexpression of IL-27 in the micromilieu of colon cancer resulted in decreased tumor growth even in the absence of IL-12p40, the functional repertoire of IL-27 also includes effects that could only be observed as synergistic effects together with IL-12, such as, for instance, the enhancement of IFN\(\gamma\) production in CD8\(^+\) T cells [118]. Other aspects of the tumor-controlling capacity of IL-27, such as its ability to induce the proliferation of naïve CD8\(^+\) T cells or the differentiation of CTLs, seem to be independent of the presence of the transcription factor T-bet, which crucially controls the fate of Th1 cells and CTLs [118,119]. Most likely, IL-27 is able to compensate for the lack of T-bet during the process of CTL differentiation by its demonstrated capacity to induce the augmented expression of Eomes [118], representing an alternative T-box transcription factor of described prognostic relevance in the clinical context of CRC [120].

In contrast to the beneficial effects of IL-12 and IL-27 on the immune system-mediated control of CRC development, IL-23 was found to promote the immune evasion of colon tumor cells [101,121]. Indeed, IL-23p19-deficient animals turned out to be protected against the experimental induction of colon cancer [76,101,122]. In human CRC patients, IL-23p19 mRNA and protein expression levels were found to be significantly increased in the tumor tissue compared to adjacent tumor-free colon tissue [76,123], and individuals with elevated IL-23p19 mRNA expression showed lower disease-free survival rates [124]. Mechanistically, the early CRC-induced loss of local intestinal barrier integrity and the subsequent
entry of microbial products into the tumor microenvironment relevantly trigger the activation of IL-23-producing intratumoral myeloid cells [125]. In general, antigen-presenting cell-derived or neutrophil-derived IL-23 [126] is known to be crucially involved in the differentiation, maintenance, survival, and IL-23-responsiveness of Th17 cells [76,127,128] and thereby contributes to increased levels of IL-17A in the tumor microenvironment [76], favoring an immunosuppressive and tumor-promoting milieu. Accordingly, the presence of intratumoral IL-17A+ IL-23R+ IL-6+ CD4+ T cells was described to be associated with the development of colitis-associated cancer [76].

Compared with IL-12, IL-23, and IL-27, relatively little is known so far about the exact functional influence of the most recently discovered IL-12 family member, IL-35, on CRC pathogenesis. In good accordance with the generally suggested immunosuppressive and tumor-promoting function of IL-35 [129,130], the overall increased presence of the two IL-35 subunits p35 and EBI3 in colon tumor tissue reached the most pronounced expression levels in advanced and poorly differentiated colon tumors [99]. Notably, serum levels of IL-35 in patients diagnosed with CRC disease positively correlated with the number of circulating FoxP3 T<sub>reg</sub> cells in the peripheral blood [99], which may indicate, beyond the fact that T<sub>reg</sub> cells are considered the main source of IL-35, that IL-35 is involved in the CRC-associated induction of peripheral T<sub>reg</sub> cells and thus in the initiation of tumor immune escape mechanisms.

3.6. TGFβ

In the colon tumor microenvironment, the anti-inflammatory cytokine TGFβ turned out to be mainly produced by cancer-associated fibroblasts but also by tumor-infiltrating T<sub>reg</sub> cells and tumor cells [93,131]. Its increased expression in primary colon tumor specimens correlated with metastatic disease and poor prognosis. It should nevertheless be mentioned, at this point, that the direct interplay between TGFβ and colon cancer cells can obviously mediate both tumor-promoting and tumor growth-arresting effects depending on the stage and type of disease, emphasizing the complexity of its involvement in the pathogenesis of CRC [132,133]. However, focusing on its impact on the immune system-mediated control of colon cancer and CRC metastasis, TGFβ has been definitively identified as a driver of tumor immune escape mechanisms, and its targeted inhibition in different preclinical in vivo models successfully dampened the development and metastasis of colon cancer and increased the efficacy of adoptive NK cell therapy [131,134]. Besides its immunosuppressive function, which includes the local induction of T<sub>reg</sub> cell differentiation from naïve CD4<sup>+</sup> T cells and the generation of monocytic myeloid-derived suppressor cells from CD14<sup>+</sup> monocytes [135–137], TGFβ serves as a key regulator of extracellular matrix (ECM) homeostasis. Via its ECM-modulating capacity, TGFβ contributes to the phenomenon of tumor T cell exclusion, which means that the accessibility of the tumor tissue for invading immune cells can be blocked by the formation of a collagen-rich ECM barrier [138]. Indeed, genes induced by TGFβ in normal colonic fibroblasts were found to be upregulated in colon tumors of CRC patients with poor prognosis [139]. Moreover, on a functional level, it could be demonstrated, in the in vivo MC38 colon cancer model, that the therapeutic blockade of TGFβ resulted in the increased tumor infiltration of CD8<sup>+</sup> T cells and NK cells [138]. Regarding the effect of TGFβ on colon tumor-infiltrating CD4<sup>+</sup> T cells, the increased presence of TGFβ in sporadic colon tumors was obviously accompanied by the augmented presence of IL-17A+ IL-22+ CD4<sup>+</sup> T cells. On a functional level, in vivo studies in colitis-associated cancer were able to confirm that TGFβ relevantly contributed to the intratumoral accumulation of IL-17A+ IL-22+ CD4<sup>+</sup> T cells and thereby obviously triggered IL-22-dependent tumor-promoting effects [140,141]. Finally, results from the same preclinical models interestingly suggested that the tumor micromilieu is able to further increase the TGFβ responsiveness of colon tumor-infiltrating CD4<sup>+</sup> T cells by suppressing their expression of the TGFβ signaling-regulator Smurf2, with the consecutive induction of TGFβ receptor type II levels [142]. In summary, TGFβ can thus be seen as a relevant antagonist of the immunological control of CRC disease, both by limiting the presence...
of cytolytic effector immune cells in the tumor and by promoting the T cell-mediated secretion of pro-tumorigenic cytokines. However, it should also be considered that the T cell-specific blockade of TGFβ-signaling in the AOM/DSS model of colitis-associated cancer resulted in enhanced tumor formation [93], which was obviously driven by the increased intratumoral presence of T cell-released IL-6 and its capacity to trigger the growth of dysplastic epithelial cells.

4. Conclusions and Targets beyond Chemokines and Cytokines

The highlighted multifaceted influence of chemokines and cytokines on the tumor microenvironment and, in particular, on the recruitment, intratumoral accumulation, and activation of lymphocytes suggests their strong potential as therapeutic targets to specifically enhance the immunological control of tumor disease. In this context, however, it is important to carefully take into account that most of these secreted immunomodulators rarely act selectively on a single immune cell type, depending on the expression profile of their respective receptors, and often also have relevant direct effects on tumor cells or mediate systemic effects outside the tumor environment. Preclinically validated strategies to limit the systemic side effects of cytokine/chemokine-based treatment strategies in the therapy of solid tumors, for instance, include the intratumoral induction of non-secretable membrane-bound but still functionally active cytokines or the ex vivo exposure of effector immune cells to selected cytokines prior to adoptive cell therapy.

Moreover, cytokines and chemokines are of course not the only players and potential therapeutic targets within the colon tumor microenvironment that are able to modify the efficiency of immune cell recruitment into the intestinal tumor tissue and the locally induced antitumor immune response. In particular, tumor-associated neoangiogenesis, which is promoted by local hypoxia, the deprivation of nutrients, and the secretion of different growth factors, represents an important and complex process that significantly impacts the successful infiltration of immune cells from the circulating blood into the tumor tissue [143]. For instance, binding of Vascular Endothelial Growth Factor A (VEGF-A) to its receptor, VEGFR-2, represents a very relevant trigger for promoting angiogenesis and neovascularization in the context of colorectal cancer. However, this is again a highly complex scenario, as the newly formed vascular network induced under the influence of the intratumoral disbalance between pro- and anti-angiogenic factors often shows structural and functional abnormalities, resulting in insufficient tissue perfusion and the hampered delivery of chemokine-attracted effector T cells [144,145]. Moreover, VEGF-A has also been demonstrated to directly promote the proliferation of VEGFR-2-expressing regulatory T cells in colon tumor-bearing mice. Accordingly, the treatment of CRC patients with the clinically approved VEGF-A-targeting antibody Bevacizumab resulted in a significantly reduced frequency of regulatory T cells in the peripheral blood, and the in vivo administration of a fusion protein consisting of an anti-VEGFR2 antibody and IFNα successfully enhanced the accumulation of CD8+ T cells in a murine colorectal cancer xenograft model [145–148].

Besides the sensing of a chemokine-mediated recruitment signal and the accessibility of the tumor tissue ensured by existing blood vessels, the ability of circulating immune cells to migrate from the blood into the tumor tissue also crucially depends on their surface expression profile of tissue-specific homing receptors. As a surface molecule of key relevance for T cell gut homing, the integrin β7 as a heterodimer either with integrin α4 or integrin αE promotes the adhesion of T cells at the endothelial wall and their subsequent retention in the intestinal tissue, respectively [149]. Indeed, the functional and prognostic relevance of integrin β7-mediated T cell homing could also be confirmed for the pathogenesis of CRC disease, although different studies reported partly conflicting data on the quality of its involvement. The increased intratumoral presence of integrin β7-positive cells was found to be associated with enhanced immune cell-mediated cytolytic activity and the improved survival of CRC patients [150], while another study described that decreased expression levels of MAdCAM-1, the ligand of α4β7, in human colon cancer tissue pointed to an improved colon tumor prognosis [151]. In the orthotopic MC38 colon tumor model, mice
carrying a genetic deficiency for integrin β7 showed the reduced intratumoral infiltration of activated T cells, B cells, NK cells, and DCs and developed larger tumors, overall implicating that integrin β7 is required for efficient antitumor immunity. However, besides these quite recent data, there also exist experimental data published by another group already in 2017 that functionally linked the absence of integrin β7 to the slower growth of sporadic colon tumors [151]. Irrespective of the fact that the exact clinical consequence and the underlying mechanisms of β7 integrin-mediated endothelial adhesion and the intratumoral accumulation of immune cells, as well as the possible involvement of other integrins and integrin receptors, certainly need to be investigated in more detail in future studies, anti-adhesion and anti-integrin therapies not only represent a valuable treatment approach in the therapy of chronic inflammatory diseases [152], but might also emerge as a promising and exciting therapeutic option for triggering the immunological tumor control in CRC disease.

In conclusion, we hope that, in this review article, we have succeeded in demonstrating the diversity and functional complexity but also the great therapeutic potential of the various factors involved in immune cell recruitment and local immune cell activation in the colon tumor microenvironment, which are highly critical for modulating the antitumor immune response, for determining the responsiveness to immunotherapy, and, finally, for improving CRC prognosis.

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References
1. Hossain, S.; Karuniawati, H.; Jairoun, A.A.; Urbi, Z.; Ooi, D.J.; John, A.; Lim, Y.C.; Kibria, K.M.K.; Mohiuddin, A.M.; Ming, L.C.; et al. Colorectal Cancer: A Review of Carcinogenesis, Global Epidemiology, Current Challenges, Risk Factors, Preventive and Treatment Strategies. Cancers 2022, 14, 1732. [CrossRef]
2. Miller, K.D.; Nogueira, L.; Mariotto, A.B.; Rowland, J.H.; Yabroff, K.R.; Alfano, C.M.; Jemal, A.; Kramer, J.L.; Siegel, R.L. Cancer treatment and survivorship statistics, 2019. CA Cancer J. Clin. 2019, 69, 363–385. [CrossRef]
3. Schmitt, M.; Greten, F.R. The inflammatory pathogenesis of colorectal cancer. Nat. Rev. Immunol. 2021, 21, 653–667. [CrossRef]
4. Yue, Y.; Zhang, Q.; Sun, Z. CX3CR1 Acts as a Protective Biomarker in the Tumor Microenvironment of Colorectal Cancer. Front. Immunol. 2022, 12, 758040. [CrossRef]
5. Pagès, F.; Berger, A.; Camus, M.; Sanchez-Cabo, F.; Costes, A.; Molidor, R.; Mlecnik, B.; Kirilovsky, A.; Nilsson, M.; Damotte, D.; et al. Effector memory T cells, early metastasis, and survival in colorectal cancer. N. Engl. J. Med. 2005, 353, 2654–2666. [CrossRef]
6. Boquet, I.; Kassambara, A.; Lui, A.; Tanner, A.; Latil, M.; Lovera, Y.; Arnoux, F.; Hermitte, F.; Galon, J.; Catteau, A. Comparison of Immune Response Assessment in Colon Cancer by Immunoscore (Automated Digital Pathology) and Pathologist Visual Scoring. Cancers 2022, 14, 1170. [CrossRef]
7. Argilés, G.; Tabernero, J.; Labianca, R.; Hochhauser, D.; Salazar, R.; Iveson, T.; Laurent-Puig, P.; Quirke, P.; Yoshino, T.; Taieb, J.; et al. Localised colon cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann. Oncol. 2020, 31, 1291–1305. [CrossRef]
8. Liu, F.; Dong, X.; Pang, X.; Xing, Q.; Wang, H.; Zhang, H.; Li, Y.; Yin, Y.; Fant, M.; Ye, Y.; et al. The Specific Immune Response to Tumor Antigen CP1 and Its Correlation With Improved Survival in Colon Cancer Patients. Gastroenterology 2008, 134, 998–1006. [CrossRef]
9. Pancione, M.; Giordano, G.; Remo, A.; Febbraro, A.; Sabatino, L.; Manfrin, E.; Ceccherini, M.; Colantuoni, V. Immune Escape Mechanisms in Colorectal Cancer Pathogenesis and Liver Metastasis. J. Immunol. Res. 2014, 2014, 1–11. [CrossRef]
10. Fidelis, M.; Yonekura, S.; Picard, M.; Cogdill, A.; Hollebecque, A.; Roberti, M.P.; Zitvogel, L. Resolving the Paradox of Colon Cancer Through the Integration of Genetics, Immunology, and the Microbiota. Front. Immunol. 2020, 11, 600886. [CrossRef]
11. Breakstone, R. Colon cancer and immunotherapy—can we go beyond microsatellite instability? Transl. Gastroenterol. Hepatol. 2021, 6, 12. [CrossRef]
12. Michael-Robinson, J.M.; Biemer-Hüttmann, A.-E.; Purdie, D.M.; Walsh, M.D.; Simms, L.A.; Biden, K.G.; Young, J.P.; Leggett, B.A.; Jass, J.R.; Radford-Smith, G.L. Tumour infiltrating lymphocytes and apoptosis are independent features in colorectal cancer stratified according to microsatellite instability status. *C Gut* 2001, 48, 360–366. [CrossRef]

13. DiToro, D.; Basu, R. Emerging Complexity in CD4+ T-Lineage Programming and Its Implications in Colorectal Cancer. *Front. Immunol.* 2021, 12, 3362. [CrossRef]

14. Wu, P.; Wu, D.; Ni, C.; Ye, J.; Chen, W.; Hu, G.; Wang, Z.; Wang, C.; Zhang, Z.; Xia, W.; et al. γδT17 cells promote the accumulation and expansion of myeloid-derived suppressor cells in human colorectal cancer. *Immunity*. 2014, 40, 785–800. [CrossRef]

15. Saito, T.; Nishikawa, H.; Wada, H.; Nagano, Y.; Sugiyama, D.; Atarashi, K.; Maeda, Y.; Hamaguchi, M.; Ohkura, N.; Sato, E.; et al. Two FOXP3+CD4+ T cell subpopulations distinctly control the progression of colorectal cancers. *Nat. Med.* 2016, 22, 679–684. [CrossRef]

16. Hanke, T.; Melling, N.; Simon, R.; Sauter, G.; Bokemeyer, C.; Lebok, P.; Terracciano, L.M.; Izbicki, J.R.; Marx, A.H. High intratumoral FOXP3+ T regulatory cell (Tregs) density is an independent good prognosticator in nodal negative colorectal cancer. *Int. J. Clin. Exp. Pathol.* 2015, 8, 8227–8235.

17. Liu, Z.; Huang, Q.; Liu, G.; Deng, L.; Chu, D.; Tao, K.; Wang, W. Presence of FOXP3(+)+Treg cells is correlated with colorectal cancer progression. *Int. J. Clin. Exp. Med.* 2014, 7, 1781–1785.

18. Carluccio, S.; Delbue, S.; Signorini, L.; Setola, E.; Bagliani, A.; Della Valle, A.; Galli, A.; Ferrante, P.; Bregn, M. Generation of tumor-specific cytotoxic T-lymphocytes from the peripheral blood of colorectal cancer patients for adoptive T-cell transfer. *J. Cell. Physiol.* 2015, 230, 1457–1465. [CrossRef]

19. Aparicio, C.; Belver, M.; Enriquez, L.; Espeso, F.; Nuñez, L.; Sánchez, A.; de la Fuente, M.; González-Vallinas, M. Cell Therapy for Colorectal Cancer: The Promise of Chimeric Antigen Receptor (CAR)-T Cells. *Int. J. Mol. Sci.* 2021, 22, 11781. [CrossRef]

20. Erreni, M.; Bianchi, P.; Laghi, L.; Mirolo, M.; Fabbri, M.; Locati, M.; Mantovani, A.; Allavena, P. Chapter 5 Expression of Chemokines and Chemokine Receptors in Human Colon Cancer. *Methods Enzymol.* 2009, 460, 105–121. [CrossRef]

21. Shamoun, L.; Landeholm, K.; Ramilo, A.B.; E Andersson, R.; Dimberg, J.; Wågsäter, D. Association of gene and protein expression and genetic polymorphism of CC chemokine ligand 4 in colorectal cancer. *World J. Gastroenterol.* 2021, 27, 5076–5087. [CrossRef]

22. Siddiqui, I.; Erreni, M.; van Brakel, M.; Debets, R.; Allavena, P. Enhanced recruitment of genetically modified CX3CR1-positive human T cells into Fractalkine/CX3CL1 expressing tumors: Importance of the chemokine gradient. *J. Immunother. Cancer*. 2016, 4, 21. [CrossRef]

23. Erreni, M.; Siddiqui, I.; Marelli, G.; Grizzi, F.; Bianchi, P.; Morone, D.; Marchesi, F.; Celesti, G.; Pesce, S.; Doni, A.; et al. The Fractalkine-Receptor Axis Improves Human Colorectal Cancer Prognosis by Limiting Tumor Metastatic Dissemination. *J. Immunol. 2015*, 196, 902–914. [CrossRef]

24. Hojo, S.; Koizumi, K.; Tsuneyama, K.; Arita, Y.; Cui, Z.; Shinohara, K.; Minami, T.; Hashimoto, I.; Nakayama, T.; Sakurai, H.; et al. High-Level Expression of Chemokine CXCL16 by Tumor Cells Correlates with a Good Prognosis and Increased Tumor-Infiltrating Lymphocytes in Colorectal Cancer. *Res. 2007*, 67, 4725–4731. [CrossRef]

25. Wang, B.; Wang, Y.; Sun, X.; Deng, G.; Huang, W.; Wu, X.; Gu, Y.; Tian, Z.; Fan, Z.; Xu, Q.; et al. CXCR6 is required for antitumor efficacy of intratumoral CD8+ T cell. *J. Immunother. Cancer*. 2021, 9, e031010. [CrossRef]

26. Nagasawa, S.; Tsuchida, K.; Shiozawa, M.; Hiroshima, Y.; Kimura, Y.; Hashimoto, I.; Watanabe, H.; Kano, K.; Numata, M.; Aoyama, T.; et al. Clinical Significance of Fractalkine/CX3C14 and CXCR4 mRNA Expression in Patients With Colorectal Cancer. *Anticancer Res.* 2021, 41, 4489–4495. [CrossRef]

27. Wang, D.; Yang, L.; Yu, W.; Wu, Q.; Lian, J.; Li, F.; Liu, S.; Li, A.; He, Z.; Liu, J.; et al. Colorectal cancer cell-derived CCL20 recruits regulatory T cells to promote chemoresistance via FOXO1/CEBPB/NF-κB signaling. *J. Immunother. Cancer*. 2019, 7, 215. [CrossRef]

28. Wu, Z.; Han, X.; Yan, J.; Fan, Y.; Gong, J.; Di, J.; Cheng, Z.; Jin, Z.; Wang, Z.; Zheng, Q.; et al. The prognostic significance of chemokine receptor CXCR3 expression in colorectal carcinoma. *Biomed. Pharmacother.* 2012, 66, 373–377. [CrossRef]

29. Kim, J.; Takeuchi, H.; Lam, S.T.; Turner, R.R.; Wang, H.-J.; Kuo, C.; Foshag, L.; Bilchik, A.J.; Hoon, D.S. Chemokine Receptor CXCR4 Expression in Colorectal Cancer Patients Increases the Risk for Recurrence and for Poor Survival. *J. Clin. Oncol.* 2005, 23, 2744–2753. [CrossRef]

30. Mlcnek, B.; Tosolini, M.; Charoentong, P.; Kirilovsky, A.; Bindea, G.; Berger, A.; Camus, M.; Gillard, M.; Bruneval, P.; Fridman, W.H.; et al. Biomolecular Network Reconstruction Identifies T-Cell Homing Factors Associated With Survival in Colorectal Cancer. *Gastroenterology*. 2010, 138, 1429–1440. [CrossRef]

31. Kistner, L.; Doll, D.; Holtorf, A.; Nitsche, U.; Janssen, K.-P. Interferon-inducible CX3-CHEMOKINES are crucial immune modulators and survival predictors in colorectal cancer. *OncoTarget*. 2017, 8, 89998–90012. [CrossRef] [PubMed]

32. Guo, J.-N.; Li, M.-Q.; Deng, S.-H.; Chen, C.; Ni, Y.; Cui, B.-B.; Liu, Y.-L. Prognostic Immune-Related Analysis Based on Differentially Expressed Genes in Left- and Right-Sided Colon Adenocarcinoma. *Front. Oncol*. 2021, 11, 640196. [CrossRef] [PubMed]

33. Zimmermann, T.; Moehler, M.; Gockel, I.; Sgourakis, G.G.; Biesterfeld, S.; Müller, M.; Berger, M.R.; Lang, H.; Galle, P.R.; Schimsanski, C.C. Low expression of chemokine receptor CCR5 in human colorectal cancer correlates with lymphatic dissemination and reduced CD8+ T-cell infiltration. *Int. J. Col. Dis.* 2010, 25, 417–424. [CrossRef] [PubMed]

34. Karin, N. CXCR3 Ligands and Autoimmunity, Chemotractionf of Effector T Cells, and Beyond. *Front. Immunol.* 2020, 11, 976. [CrossRef]
59. Liu, J.; Zhang, N.; Li, Q.; Zhang, W.; Ke, F.; Leng, Q.; Wang, H.; Chen, J.; Wang, H. Tumor-Associated Macrophages Recruit CCR6+ Regulatory T Cells and Promote the Development of Colorectal Cancer via Enhancing CCL20 Production in Mice. *PloS ONE* **2011**, *6*, e19495. [CrossRef]

60. Sun, Z.; Du, C.; Xu, P.; Miao, C. Surgical trauma-induced CCL18 promotes recruitment of regulatory T cells and colon cancer progression. *J. Cell. Physiol.* **2019**, *234*, 4608–4616. [CrossRef]

61. Märlö, F.; Huynh, D.; Endres, S.; Kobold, S. Utilizing chemokines in cancer immunotherapy. *Trends Cancer* **2022**, *8*, 670–682. [CrossRef]

62. Wang, L.; Wang, Y.; Song, Z.; Chu, J.; Qu, X. Deficiency of Interferon-Gamma or Its Receptor Promotes Colorectal Cancer Progression. *Cancer Cell* **2017**, *31*, 194–207. [CrossRef]

63. Qin, Z.; Schwartzkopff, J.; Pradera, F.; Kammertoens, T.; Seliger, B.; Pircher, H.; Blankenstein, T. A critical requirement of interferon-gamma-mediated angiostasis for tumor rejection by CD8+ T cells. *Cancer Res.* **2003**, *63*, 4095–4100.

64. Burke, J.D.; Young, H.A. IFN-γ: A cytokine at the right time, is in the right place. *Semin. Immunol.* **2019**, *43*, 101280. [CrossRef] [PubMed]

65. Ling, A.; Lundberg, I.V.; Eklöf, V.; Wikberg, M.L.; Öberg, Å.; Edin, S.; Palmqvist, R. The infiltration, and prognostic importance, of Th1 lymphocytes vary in molecular subgroups of colorectal cancer. *J. Pathol. Clin. Res.* **2016**, *2*, 21–31. [CrossRef]

66. Coppola, A.; Arriga, R.; Lauro, D.; Del Principe, M.L.; Bucchiano, F.; Maurillo, L.; Palomba, P.; Venditti, A.; Sconocchia, G. NK Cell Inflammation in the Clinical Outcome of Colorectal Carcinoma. *Front. Med.* **2015**, *2*, 33. [CrossRef]

67. Katlinski, K.V.; Gui, J.; Katlinskaya, Y.V.; Ortiz, A.; Chakraborty, R.; Bhattacharya, S.; Carbone, C.J.; Betting, D.P.; Girondo, M.A.; Peck, A.R.; et al. Inactivation of Interferon Receptor Promotes the Establishment of Immune Privileged Tumor Microenvironment. *Cancer Cell* **2017**, *31*, 132, e153167. [CrossRef]

68. Zhao, T.; Li, Y.; Zhang, J.; Zhang, B. PD-L1 expression increased by IFN-γ via JAK2-STAT1 signaling and predicts a poor survival in colorectal cancer. *Oral Oncol. Lett.* **2020**, *20*, 1127–1134. [CrossRef]

69. Ren, J.; Li, N.; Pei, S.; Lian, Y.; Li, L.; Peng, Y.; Liu, Q.; Guo, J.; Wang, X.; Han, Y.; et al. Histone methyltransferase WHSC1 loss dampens MHC-I antigen presentation pathway to impair IFN-γ-stimulated antitumor immunity. *J. Clin. Investig.* **2022**, *132*, e153167. [CrossRef]

70. Hurtado, C.G.; Fan, F.; Houseau, F.; Sears, C.L. Roles for Interleukin 17 and Adaptive Immunity in Pathogenesis of Colorectal Cancer. *Gastroenterology* **2018**, *155*, 1706–1715.

71. Cui, G.; Yuan, A.; Goll, R.; Florholmen, J. IL-17A in the tumor microenvironment of the human colorectal adenoma–carcinoma sequence. *Scand. J. Gastroenterol.* **2012**, *47*, 1304–1312. [CrossRef] [PubMed]

72. De Simone, V.; Pallone, F.; Monteleone, G.; Stolfi, C. Role of TH17 cytokines in the control of colorectal cancer. *Oncoimmunology* **2013**, *2*, e26617. [CrossRef] [PubMed]

73. Mao, H.; Pan, F.; Guo, H.; Bu, F.; Xin, T.; Chen, S.; Guo, Y. Feedback mechanisms between M2 macrophages and Th17 cells in colorectal cancer patients. *Tumor Biol.* **2016**, *37*, 12223–12230. [CrossRef] [PubMed]

74. Wang, D.; Yu, W.; Lian, J.; Wu, Q.; Liu, S.; Yang, L.; Li, F.; Huang, L.; Chen, X.; Zhang, Z.; et al. TH17 cells inhibit CD8+ T cell migration by systematically downregulating CXCR3 expression via IL-17A/STAT3 in advanced-stage colorectal cancer patients. *J. Hematol. Oncol.* **2020**, *13*, 1–15. [CrossRef]

75. Amicarella, F.; Muraro, M.G.; Hirt, C.; Cremonesi, E.; Padovan, E.; Mele, V.; Governa, V.; Han, J.; Huber, X.; Droezer, R.A.; et al. Dual role of tumor-infiltrating T helper 17 cells in human colorectal cancer. *Gut* **2017**, *66*, 692–704. [CrossRef]

76. Punkenburg, E.; Vogler, T.; Büttrm, M.; Amann, K.; Waldner, M.; Atrey, R.; Abendroth, B.; Mudter, J.; Merkel, S.; Gallmeier, E.; et al. Batf-dependent Th17 cells critically regulate IL-23 driven colitis-associated colon cancer. *Gut* **2015**, *65*, 1139–1150. [CrossRef]

77. Rizzo, A.; De Mare, V.; Rocci, C.; Stolfi, C.; Colantoni, A.; Neurath, M.F.; Macdonald, T.T.; Pallone, F.; Monteleone, G.; Fantini, M.C. Smad2 induces plasticity in tumor-infiltrating Th17 cells and enables TNF-alpha-mediated killing of colorectal cancer cells. *Carcinogenesis* **2014**, *35*, 1536–1546. [CrossRef]

78. Groth, C.; Weber, R.; Lasser, S.; Ozbay, E.G.; Kurzay, A.; Petrova, V.; Allevogt, P.; Utikal, J.; Umansky, V. Tumor promoting activity of polymorphonuclear myeloid-derived suppressor cells and their neutralization. *Int. J. Cancer* **2021**, *149*, 1628–1638. [CrossRef]

79. Johdi, N.A.; Ait-Tahar, K.; Sagap, I.; Jamal, R. Molecular Signatures of Human Regulatory T Cells in Colorectal Cancer and Polyps. *Front. Immunol.* **2017**, *8*, 620. [CrossRef]

80. Evans, C.; Dalgleish, A.G.; Kumar, D. Review article: Immune suppression and colorectal cancer. *Aliment. Pharmacol. Ther.* **2006**, *24*, 1163–1177. [CrossRef]

81. Giacomelli, L.; Gianni, W.; Belfiore, C.; Gandini, O.; Repetto, L.; Filippini, A.; Frati, L.; Agliano, A.M.; Gazzaniga, P. Persistence of epidermal growth factor receptor and interleukin 10 in blood of colorectal cancer patients after surgery identifies patients with high risk to relapse. *Clin. Cancer Res.* **2003**, *9*, 2678–2682.

82. Galizia, G.; Orditura, M.; Romano, C.; Lieto, E.; Castellano, P.; Peloso, L.; Imperatore, V.; Catalano, G.; Pignatelli, C.; De Vita, F. Prognostic Significance of Circulating IL-10 and IL-6 Serum Levels in Colon Cancer Patients Undergoing Surgery. *Clin. Immunol.* **2002**, *102*, 169–178. [CrossRef] [PubMed]

83. Rossowska, J.; Anger, N.; Szczygiel, A.; Mierzejewska, J.; Pajtasz-Piasecka, E. Reprogramming the murine colon cancer microenvironment using lentivectors encoding shRNA against IL-10 as a component of a potent DC-based chemoimmunotherapy. *J. Exp. Clin. Cancer Res.* **2018**, *37*, 1–14. [CrossRef] [PubMed]
84. Li, J.; Huang, L.; Zhao, H.; Yan, Y.; Lu, J. The Role of Interleukins in Colorectal Cancer. *Int. J. Biol. Sci.* 2020, 16, 2323–2339. [CrossRef]

85. Erdman, S.E.; Sohn, J.J.; Rao, V.P.; Nambiar, P.R.; Ge, Z.; Fox, J.G.; Schauer, D.B. CD4+CD25+ Regulatory Lymphocytes Induce Regression of Intestinal Tumors in ApcMin/+ Mice. *Cancer Res.* 2005, 65, 3998–4004. [CrossRef]

86. Rizzo, A.; Pallone, F.; Monteleone, G.; Fantini, M.C. Intestinal inflammation and colorectal cancer: A double-edged sword? *World J. Gastroenterology.* 2011, 17, 3092–3100.

87. Li, W.; Wu, Z.; Meng, W.; Zhang, C.; Cheng, M.; Chen, Y.; Zou, Y.; Li, K.; Lin, S.; Xiong, W.; et al. Blockade of IL-6 inhibits tumor expression in tumors of mice receiving systemic therapy with IL-12. *J. Immunol.* 2011, 186, 15901–15909. [CrossRef] [PubMed]

88. Koper-Lenkiewicz, O.; Dymicka-Piekarska, V.; Milewksa, A.; Zitchczuk, J.; Kamińska, J. The Relationship between Inflammation Markers (CRP, IL-6, sCD40L) and Colorectal Cancer Stage, Grade, Size and Location. *Diagnostics* 2021, 11, 1382. [CrossRef]

89. Lehtomäki, K.; Mustonen, H.; Kellokumpu-Lehtinen, P.-L.; Joensuu, H.; Hermunen, K.; Soeveri, L.-M.; Boisen, M.; Dehlendorff, C.; Johansen, J.; Haglund, C.; et al. Lead Time and Prognostic Role of Serum CEA, CA19-9, IL-6, CRP, and YKL-40 after Adjuvant Chemotherapy in Colorectal Cancer. *Cancers* 2021, 13, 3892. [CrossRef]

90. Lu, C.-C.; Kuo, H.-C.; Wang, F.-S.; Jou, M.-H.; Lee, K.-C.; Chuang, J.-H. Upregulation of TLRs and IL-6 as a Marker in Human Colorectal Cancer. *Int. J. Mol. Sci.* 2014, 15, 15901–15909. [CrossRef] [PubMed]

91. Ohno, Y.; Toyoshima, Y.; Yurino, H.; Monna, N.; Xiang, H.; Sumida, K.; Kaneumi, S.; Terada, S.; Hashimoto, S.; Ikek, K.; et al. Lack of interleukin-6 in the tumor microenvironment augments type-1 immunity and increases the efficacy of cancer immunotherapy. *Cancer Sci.* 2017, 108, 1999–1966. [CrossRef]

92. Li, W.; Wu, Z.; Meng, W.; Zhang, C.; Cheng, M.; Chen, Y.; Zou, Y.; Li, K.; Lin, S.; Xiong, W.; et al. Blockade of IL-6 inhibits tumor immune evasion and improves anti-CD8+ T cell responses in colorectal cancer patients. *Cytokine* 2022, 158, 155976. [CrossRef] [PubMed]

93. Becker, C.; Fantini, M.C.; Schramm, C.; Lehr, H.A.; Wirtz, S.; Becker, A.; Burg, J.; Strand, S.; Kiesslich, R.; Huber, S.; et al. TGF-beta suppresses tumor progression in colon cancer by inhibition of IL-6 trans-signaling. *Immunity* 2004, 42, 139. [CrossRef] [PubMed]

94. Hunter, C.A.; Jones, S.A. IL-6 as a keystone cytokine in health and disease. *Nat. Immunol.* 2017, 18, 1271. [CrossRef]

95. Gerlach, K.; Popp, V.; Wirtz, S.; Al-Saifi, R.; Acera, M.G.; Atreya, R.; Dregelies, T.; Vieth, M.; Fichtner-Feigl, S.; McKenzie, A.N.J.; et al. Modified nanoparticle mediated IL-12 immunogene therapy for colon cancer. *Nanomed. Nanotechnol. Biol. Med.* 2012, 8, 294–309. [CrossRef] [PubMed]

96. Neurath, M.F.; Finotto, S. IL-6 signaling in autoimmune, chronic inflammation and inflammation-associated cancer. *Cytokine Growth Factor Rev.* 2011, 22, 83–89. [CrossRef]

97. Wunderlich, C.M.; Ackermann, P.J.; Ostermann, A.L.; Adams-Quack, P.; Vogt, M.C.; Tran, M.L.; Nikolajev, A.; Waismann, A.; Garbers, C.; Theurich, S.; et al. Obesity exacerbates colitis-associated cancer via IL-6-regulated macrophage polarisation and CCL-20/CCR-6-mediated lymphocyte recruitment. *Nat. Commun.* 2018, 9, 1646. [CrossRef] [PubMed]

98. Zhong, C.; Wang, L.; Hu, S.; Huang, C.; Xia, Z.; Liao, J.; Yi, W.; Chen, J. Poly(I:C) enhances the efficacy of phagocytosis checkpoint blockade immunotherapy by inducing IL-6 production. *J. Leukoc. Biol.* 2021, 110, 1197–1208. [CrossRef] [PubMed]

99. Zeng, J.-C.; Zhang, Z.; Li, T.-Y.; Liang, Y.-F.; Wang, H.-M.; Bao, J.-J.; Zhang, J.-A.; Wang, W.-Y.; Kong, B.; et al. Assessing the role of IL-35 in 35 colorectal cancer progression and prognosis. *Int. J. Clin. Exp. Pathol.* 2013, 6, 1806–1816.

100. Wang, Z.; Liu, J.-Q.; Liu, Z.; Shen, R.; Zhang, G.; Xu, J.; Basu, S.; Feng, Y.; Bai, X.-F. Tumor-Derived IL-35 Promotes Tumor Growth by Enhancing Myeloid Cell Accumulation and Angiogenesis. *J. Immunol.* 2013, 190, 2415–2423. [CrossRef]

101. Engel, M.A.; Neurath, M.F. Anticancer properties of the IL-12 family–focus on colorectal cancer. *Curr. Med. Chem.* 2010, 17, 3303–3308. [CrossRef] [PubMed]

102. Vignali, D.A.; Kuchroo, V.K. IL-12 family cytokines: Immunological playmakers. [PubMed]

103. Fujiwara, H.; Clark, S.C.; Hamaoka, T. Cellular and Molecular Mechanisms Underlying IL-12-Induced Tumor Regression. *Ann. New York Acad. Sci.* 1996, 795, 294–309. [CrossRef] [PubMed]

104. Chi, X.; Yang, P.; Zhang, E.; Gu, J.; Xu, H.; Li, M.; Gao, X.; Li, X.; Zhang, Y.; Xu, H.; et al. Significantly increased anti-tumor activity of carcinoembryonic antigen-specific chimeric antigen receptor T cells in combination with recombinant human IL-12. *Cancer Med.* 2019, 8, 4753–4765. [CrossRef]

105. Uemura, A.; Takehara, T.; Miyagi, T.; Suzuki, T.; Tsurumi, R.; Okhawa, K.; Kanto, T.; Hiramatsu, N.; Hayashi, N. Natural killer cell is a major producer of interferon gamma that is critical for the IL-12-induced anti-tumor effect in mice. *Cancer Immunol. Immunother.* 2011, 60, 155976. [CrossRef] [PubMed]

106. Rodolfo, M.; Zilocchi, C.; Melani, C.; Cappetti, B.; Arioli, I.; Parmiani, G.; Colombo, M.P. Immunotherapy of experimental metastases by vaccination with interleukin gene-transduced adenocarcinoma cells sharing tumor-associated antigens. Comparison between IL-12 and IL-2 gene-transduced tumor cell vaccines. *J. Immunol.* 1996, 157, 5536–5542.

107. Tannenbaum, C.S.; Wicker, N.; Armstrong, D.; Tubbs, R.; Finke, J.; Bukowski, R.M.; A Hamilton, T. Cytokine and chemokine expression in tumors of mice receiving systemic therapy with IL-12. *J. Immunol.* 1996, 156, 693–699.

108. Nastala, C.L.; Edington, H.D.; McKinney, T.G.; Tahara, H.; A Nalesnik, M.; Brunda, M.J.; Gately, M.K.; Wolf, S.F.; Schreiber, R.D.; Storkus, W.J. Recombinant IL-12 administration induces tumor regression in association with IFN-gamma production. *J. Immunol.* 1994, 153, 1697–1706.

109. Liu, X.; Gao, X.; Zheng, S.; Wang, B.; Li, Y.; Zhao, C.; Muftuoglu, Y.; Chen, S.; Li, Y.; Yao, H.; et al. Modified nanoparticle mediated IL-12 immunogene therapy for colon cancer. *Nat. Nanotechnol.* 2017, 13, 993–2004. [CrossRef]
10. Rodriguez-Madoz, J.R.; Prieto, J.; Smerdou, C. Semliki forest virus engineered to express higher IL-12 levels induce efficient elimination of murine colon adenocarcinoma. Mol. Ther. 2005, 12, 153–163. [CrossRef]

11. Quetglas, J.I.; Rodriguez-Madoz, J.R.; Beznarartea, J.; Ruiz-Guillen, M.; Casales, E.; Medina-Echeverrz, J.; Prieto, J.; Berraondo, P.; Hervas-Stubbis, S.; Smerdou, C. Eradication of liver-implanted tumors by Semliki Forest virus expressing IL-12 requires efficient long-term immune responses. J. Immunol. 2013, 190, 2994–3004. [CrossRef] [PubMed]

12. Gri, G.; Chiodoni, C.; Gallo, E.; Stoppani, A.; Liew, F.Y.; Colombo, M.P. Antitumor effect of interleukin (IL)-12 in the absence of endogenous IFN-gamma: A role for intrinsic tumor immunogenicity and IL-15. Cancer Res. 2002, 62, 4390–4397.

13. Malvicini, M.; Ingoliotti, M.; Piccioni, F.; Garcia, M.; Bayo, J.; Atorrasagasti, C.; Alaniz, L.; Aquino, J.B.; Espinoza, J.A.; Gidekel, M.; et al. Reversal of gastrointestinal carcinoma-induced immunosuppression and induction of antitumoral immunity by a combination of cyclophosphamide and gene transfer of IL-12. Mol. Oncol. 2011, 5, 242–255. [CrossRef] [PubMed]

14. Xu, Y.X.; Gao, X.; Janakiraman, N.; Chapman, R.A.; Gautam, S.C. IL-12 Gene Therapy of Leukemia with Hematopoietic Progenitor Cells without the Toxicity of Systemic IL-12 Treatment. Clin. Immunol. 2001, 98, 180–189. [CrossRef]

15. Lenz, R.; Rosenblum, M.; Verschraegen, C.; Kudelka, A.P.; Kavanagh, J.J.; Hicks, M.E.; Lang, E.A.; Nash, M.A.; Levy, L.B.; Garcia, M.E.; et al. Phase I study of intraperitoneal recombinant human interleukin 12 in patients with Mullerian carcinoma, gastrointestinal polyps, and mesothelioma. Clin. Cancer Res. 2002, 8, 3686–3695.

16. Hewitt, S.L.; Bailey, D.; Zielinski, J.; Apte, M.; Musenge, F.; Karp, R.; Burke, S.; Garcon, F.; Mishra, A.; Gurumurthy, S.; et al. Intratumoral IL-12 mRNA Therapy Promotes TH1 Transformation of the Tumor Microenvironment. Clin. Cancer Res. 2020, 26, 6284–6298. [CrossRef] [PubMed]

17. Chiyo, M.; Shimozato, O.; Yu, L.; Kawamura, K.; Iizasa, T.; Fujisawa, T.; Tagawa, M. Expression of IL-27 in murine carcinoma cells produces antitumor effects and induces protective immunity in inoculated host animals. Int. J. Cancer 2005, 115, 437–442. [CrossRef] [PubMed]

18. Morishima, N.; Ovaki, T.; Asakawa, M.; Kamiya, S.; Mizuguchi, J.; Yoshimoto, T. Augmentation of Effector CD8+ T Cell Generation with Enhanced Granulocyte B Expression by IL-27. J. Immunol. 2005, 175, 1686–1693. [CrossRef] [PubMed]

19. Powell, N.; Canavan, J.B.; Macdonald, T.T.; Lord, G. Transcriptional regulation of the mucosal immune system mediated by T-bet. Mucosal Immunol. 2010, 3, 567–577. [CrossRef] [PubMed]

20. Attirea, I.; Schimanski, C.C.; Becker, C.; Wirtz, S.; Dornhoff, H.; Schnüer, E.; Berger, M.R.; Galle, P.R.; Herr, W.; Neurath, M.F. The T-box transcription factor eomesodermin controls CD8 T cell activity and lymph node metastasis in human colorectal cancer. Gut 2007, 56, 1572–1578. [CrossRef]

21. Neurath, M.F. IL-23 in inflammatory bowel diseases and colon cancer. Cytokine Growth Factor Rev. 2018, 45, 1–8. [CrossRef]

22. Richter, C.; Herrero San Juan, M.; Weigmann, B.; Bergis, D.; Dauber, K.; Muders, M.H.; Baretton, G.B.; Bonig, H.; Brenner, S.; et al. Defective IL-23/IL-17 Axis Protects p47phox-/- Mice from Colon Cancer. Front. Immunol. 2021, 12, 76. [CrossRef] [PubMed]

23. Lan, F.; Zhang, L.; Wu, J.; Zhang, J.; Zhang, S.; Li, K.; Qi, Y.; Lin, P. IL-23/IL-23R: Potential mediator of intestinal tumor progression from adenomatous polyps to colorectal carcinoma. Int. J. Color. Dis. 2011, 26, 1511–1518. [CrossRef] [PubMed]

24. Panneerselvam, J.; Madka, V.; Rai, R.; Morris, K.T.; Houchen, C.W.; Chandrakesan, P.; Rao, C.V. Inflammatory Mediators and Gut Microbial Toxins Drive Colon Tumorigenesis by IL-23 Dependent Mechanism. Cancers 2021, 13, 5159. [CrossRef] [PubMed]

25. Grivennikov, S.I.; Wang, K.; McIver, D.; Stewart, C.A.; Schnabl, B.; Jauch, D.; Taniguchi, K.; Yu, G.Y.; Österreicher, C.H.; Hung, K.E.; et al. Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. Nature 2012, 491, 254–258. [CrossRef] [PubMed]

26. Kvedaraite, E.; Lourda, M.; Ideström, M.; Renstrom, M.; Chen, P.; Olsson-Åkefeldt, S.; Forkel, M.; Galle, P.; Brenner, S.; et al. Tissue-infiltrating neutrophils represent the main source of IL-23 in the colon of patients with IBD. Gut 2015, 64, 1632–1641. [CrossRef] [PubMed]

27. Zhou, L.; Ivanov, I.I.; Spolski, R.; Min, R.; Shenderov, K.; Egawa, T.; Levy, D.E.; Leonard, W.J.; Litman, D.R. IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nat. Immunol. 2007, 8, 967–974. [CrossRef] [PubMed]

28. McGechy, M.J.; Chen, Y.; Tato, C.M.; Laurence, A.; Joyce-Shaikh, B.; Blumenschein, W.M.; McClanahan, T.K.; O’Shea, J.J.; Cua, D.J. The interleukin 23 receptor is essential for the terminal differentiation of interleukin 17–producing effector T helper cells in vivo. Nat. Immunol. 2009, 10, 314–324. [CrossRef]

29. Sawant, D.V.; Yano, H.; Chikina, M.; Zhang, Q.; Liao, M.; Liu, C.; Callahan, D.J.; Sun, Z.; Sun, T.; Tabib, T.; et al. Adaptive plasticity of IL-10+ and IL-35+ Treg cells cooperatively promotes tumor T cell exhaustion. Nat. Immunol. 2019, 20, 724–735. [CrossRef] [PubMed]

30. Liu, K.; Huang, A.; Nie, J.; Tan, J.; Xing, S.; Qu, Y.; Jiang, K. IL-35 Regulates the Function of Immune Cells in Tumor Microenvironment. Front. Immunol. 2021, 12, 68332. [CrossRef] [PubMed]

31. Tauriello, D.V.F.; Palomo-Ponce, S.; Stork, D.; Berenguer-Llergo, A.; Badia-Ramentol, J.; Iglesias, M.; Sevillano, M.; Ibiza, S.; Cañellas, A.; Hernando-Momblona, X.; et al. Targeting IFN-Gamma to Enhance Tumor Immune Response. Nature 2018, 554, 538–543. [CrossRef] [PubMed]

32. Stolfi, C.; Troncone, E.; Barfus, I.; Monteleone, G. Role of TGF-Beta and Smad7 in Gut Inflammation, Fibrosis and Cancer. Biomolecules 2020, 11, 17. [CrossRef] [PubMed]

33. Owyang, S.Y.; Zhang, M.; Walkup, G.A.; Chen, G.E.; Grasberger, H.; El-Zaatari, M.; Kao, J.Y. The effect of CT26 tumor-derived TGF-beta on the balance of tumor growth and immunity. Innate Immun. 2019, 17, 47–54. [CrossRef] [PubMed]
134. Otegbeye, F.; Ojo, E.; Moreton, S.; Mackowski, N.; Lee, D.A.; de Lima, M.; Wald, D.N. Inhibiting TGF-beta signaling preserves the function of highly activated, in vitro expanded natural killer cells in AML and colon cancer models. *PLoS ONE* **2013**, *8*, e58886. [CrossRef]

135. Fantini, M.C.; Becker, C.; Tubbe, I.; Nikolaev, A.; Lehr, H.A.; Galle, P.; Neurath, M.F. Transforming growth factor beta induced FoxP3+ regulatory T cells suppress Th1 mediated experimental colitis. *Gut* **2006**, *55*, 671–680. [CrossRef]

136. Gneo, L.; Rizkalla, N.; Hejmadi, R.; Mussai, F.; de Santo, C.; Middleton, G. TGF-beta orchestrates the phenotype and function of monocytic myeloid-derived suppressor cells in colorectal cancer. *Cancer Immunol. Immunother. CII* **2022**, *71*, 1583–1596. [CrossRef]

137. Becker, C.; Fantini, M.; Neurath, M. TGF-beta as a T cell regulator in colitis and colon cancer. *Cytokine Growth Factor Rev.* **2006**, *17*, 97–106. [CrossRef]

138. Horn, L.A.; Chariou, P.L.; Gameiro, S.R.; Qin, H.; Iida, M.; Flies, D.; Langermann, S.; et al. Remodeling the tumor microenvironment via blockade of LAIR-1 and TGF-beta signaling enables PD-L1-mediated tumor eradication. *J. Clin. Investig.* **2022**, *132*, e135148. [CrossRef]

139. Calon, A.; Lonardo, E.; Berenguer-Llengó, A.; Espinet, E.; Hernando-Momblama, X.; Iglesias, M.; Sevillano, M.; Palomo-Ponce, S.; Tauriello, D.V.F.; Ballester, M.; et al. Stromal gene expression defines poor-prognosis subtypes in colorectal cancer. *Nat. Genet.* **2015**, *47*, 320–329. [CrossRef]

140. Perez, L.G.; Kamei, K.; McGee, H.M.; Pelzcar, P.; Agalioti, T.; Giannou, A.; Konczalla, L.; Brockmann, L.; Wahib, R.; Xu, H.; et al. TGF-beta signaling in Th17 cells promotes IL-22 production and colitis-associated colon cancer. *Nat. Commun.* **2020**, *11*, 2608. [CrossRef]

141. Zhuang, Y.; Peng, L.-S.; Zhao, Y.-L.; Shi, Y.; Mao, X.-H.; Guo, X.; Chen, W.; Liu, X.-F.; Zhang, J.-Y.; Liu, T.; et al. Increased intratumoral IL-22-producing CD4+ T cells and Th27 cells correlate with gastric cancer progression and predict poor patient survival. *Cancer Immunol. Immunother.* **2012**, *61*, 1965–1975. [CrossRef] [PubMed]

142. Dornhoff, H.; Becker, C.; Wirtz, S.; Strand, D.; Tenzer, S.; Rosfa, S.; Neufert, C.; Mudter, J.; Markl, J.; Siebler, J.; et al. A variant of Smurfl2 protects mice against colitis-associated colon cancer by inducing transforming growth factor beta signaling. *Gastroenterology* **2012**, *142*, 1183–1194.e4. [CrossRef] [PubMed]

143. Malla, R.R.; Marni, R. Angiogenesis: Promising therapeutic target of metastatic colon cancer. *Crit. Rev. Oncog.* **2020**, *25*, 161–173. [CrossRef]

144. Kampoli, K.; Foukas, P.G.; Tzavatzikos, A.; Arkadopoulos, N.; Koumarianou, A. Interrogating the interplay of angiogenesis and immunity in metastatic colorectal cancer. *World J. Methodol.* **2022**, *12*, 43–53. [CrossRef] [PubMed]

145. Voron, T.; Marcheata, E.; Perrot, S.; Colussi, O.; Tartoure, E.; Taieb, J.; Terme, M. Control of the immune response by pro-angiogenic factors. *Front Oncol.* **2014**, *4*, 70. [CrossRef]

146. Terme, M.; Perrot, S.; Marcheata, E.; Sandoval, F.; Benhamouda, N.; Colussi, O.; Dubreuil, O.; Carpentier, A.H.; Tartoure, E.; Taieb, J. VEGFA-VEGFR Pathway Blockade Inhibits Tumor-Induced Regulatory T-cell Proliferation in Colorectal Cancer. *Cancer Res.* **2013**, *73*, 539–549. [CrossRef]

147. Li, Z.; Zhu, Y.; Li, C.; Trinh, R.; Ren, X.; Sun, F.; Wang, Y.; Shang, P.; Wang, T.; Wang, M.; et al. Anti-VEGFR2-interferon-α regulates the tumor microenvironment and exhibits potent antitumor efficacy against colorectal cancer. *OncoImmunology* **2017**, *6*, e129038. [CrossRef]

148. Wang, P.; Gao, R.; Zhu, Y.; Zhang, X.; Wang, Y.; Guo, M.; Peng, H.; Wang, M.; Zhang, J. VEGFR2-targeted antibody fused with IFN mu调节 the tumor microenvironment and exhibits potent anti-tumor and anti-metastasis activity. *Acta Pharm. Sin. B* **2020**, *11*, 420–433. [CrossRef]

149. Zundler, S.; Becker, C.; Weidinger, C.; Siegmund, B. Anti-Adhesion Therapies in Inflammatory Bowel Disease—Molecular and Clinical Aspects. *Front. Immunol.* **2017**, *8*, 891. [CrossRef]

150. Zhang, Y.; Xie, R.; Zhang, H.; Zheng, Y.; Lin, C.; Yang, L.; Huang, M.; Li, M.; Song, F.; Lu, L.; et al. Integrin beta7 Inhibits Colorectal Cancer Pathogenesis via Maintaining Antitumor Immunity. *Cancer Immunol. Res.* **2021**, *9*, 967–980. [CrossRef]

151. Das, S.; Donas, C.; Akeus, P.; Quiding-Jarbrink, M.; Mora, J.R.; Villablanca, E.J. beta7 integrins contribute to intestinal tumor growth in mice. *PLoS ONE* **2018**, *13*, e0204181. [CrossRef] [PubMed]

152. Wiendl, M.; Becker, E.; Müller, T.M.; Voskens, C.J.; Neurath, M.F.; Zundler, S. Targeting Immune Cell Trafficking—Insights From Research Models and Implications for Future IBD Therapy. *Front. Immunol.* **2021**, *12*, 1546. [CrossRef] [PubMed]