Genetic interactions and functional analyses of the fission yeast gsk3 and amk2 single and double mutants defective in TORC1-dependent processes

Charalampos Rallis†,*, StJohn Townsend* & Jürg Bähler

The Target of Rapamycin (TOR) signalling network plays important roles in aging and disease. The AMP-activated protein kinase (AMPK) and the Gsk3 kinase inhibit TOR during stress. We performed genetic interaction screens using synthetic genetic arrays (SGA) with gsk3 and amk2 as query mutants, the latter encoding the regulatory subunit of AMPK. We identified 69 negative and 82 positive common genetic interactors, with functions related to cellular growth and stress. The 120 gsk3-specific negative interactors included genes functioning in translation and ribosomes. The 215 amk2-specific negative interactors included genes functioning in chromatin silencing and DNA damage repair. Both amk2- and gsk3-specific interactors were enriched in phenotype categories related to abnormal cell size and shape. We also performed SGA screen with the amk2 gsk3 double mutant as a query. Mutants sensitive to 5-fluorouracil, an anticancer drug are under-represented within the 305 positive interactors specific for the amk2 gsk3 query. The triple-mutant SGA screen showed higher number of negative interactions than the double mutant SGA screens and uncovered additional genetic network information. These results reveal common and specialized roles of AMPK and Gsk3 in mediating TOR-dependent processes, indicating that AMPK and Gsk3 act in parallel to inhibit TOR function in fission yeast.

The nutrient-responsive Target of Rapamycin (TOR) signalling pathway regulates fundamental cellular processes such as growth in response to environmental or physiological factors. In the fission yeast, Schizosaccharomyces pombe, two Tor kinases (as opposed to a single kinase in mammals) function within two protein complexes, TORC1 and TORC2. Tor2 is part of TOR Complex 1 (TORC1) and positively regulates cellular growth, while Tor1 predominates in the antagonistically acting TOR Complex 2 (TORC2). We have previously demonstrated that the fission yeast TOR pathway can be pharmacologically inhibited by the combined action of rapamycin and caffeine; TORC1 inhibition mimics the effects of nitrogen starvation and results in increased chronological lifespan (CLS).

The duration and intensity of TORC1 activation depends on the abundance and quality of different nutrients. Genetic studies in fission yeast have demonstrated that the TSC1-TSC2 complex is a negative regulator upstream of TORC1. Effectors downstream of TORC1 have been extensively characterised, including the AGC kinases Psk1, Sck1 and Sck2 that phosphorylate a variety of regulatory proteins to promote processes such as protein synthesis. For example, a major regulator of protein translation, the translation initiation factor eIF2α, can be phosphorylated by the Gcn2 kinase to inhibit translation; in nitrogen-rich conditions, Gcn2 is inactivated in a TORC1-dependent manner, thus providing a mechanism for TORC1 to promote translation in the presence of nutrients.

The AMP-activated protein kinase (AMPK) can detect decrease in cellular energy via AMP levels; in fission yeast, AMPK consists of Ssp2, Amk2 and Cbs2. AMP is an allosteric activator of AMPK: binding to the βγ...
regulatory subunit (Amk2 and Cbs2) renders the α subunit (Ssp2) a better target for the stress-responsive kinase Ssp1, and Ssp2 phosphorylation activates AMPK, thus promoting catabolic and inhibiting anabolic processes12. AMPK is functionally linked to TORC1-dependent processes: the budding yeast Ssp2 ortholog, Snf1, mediates the phosphorylation and activation of Gen2 in response to nutritional stress, leading to phosphorylation of eIF2α and a global reduction in translation9.

Gsk3 is a conserved kinase that inhibits the canonical Wnt pathway13 and has clinical significance14. However, its role in unicellular eukaryotes remains poorly characterized. We have shown that gsk3 mutant cells are resistant to TORC1 inhibition by caffeine and rapamycin and feature a shortened CLS when glucose is depleted15. Human GSK3 inhibits AMPK by interacting with the AMPK β subunit while phosphorylating and inactivating the α subunit. Disruption of GSK3 function within the AMPK complex promotes high AMPK activity and cellular catabolic processes even under anabolic conditions, indicating that GSK3 acts as a critical sensor for anabolic signalling to regulate AMPK16. To investigate the cellular roles and relationships of AMPK and Gsk3 in fission yeast, we have applied classical and genome-wide genetic approaches. We show that AMPK and Gsk3 are functionally implicated in the same TOR-dependent processes, involving both common and specific genetic interactors.

Materials and Methods

Fission yeast strains and media. Cell cultures were grown in YES liquid medium at 32 °C, with shaking at 130 rpm. Deletion mutants were generated as previously described17. For wild-type the deletion by combined treatment with caffeine and rapamycin (Fig. 1a)15. To gain insight into functional relationship of TORC1 inhibition negatively correlated with cell growth, with combined deletion of tco89Δ and drug treatment representing the most severe inhibition (Fig. 1b). Deletion of gsk3Δ was able to rescue the growth defect associated with a mild TORC1 inhibition (tco89Δ cells) as well as the phenotype of severe TORC1 inhibition (Fig. 1b, combination of tco89Δ and drug treatment). Furthermore, deletion of gsk3Δ largely rescued the reduced cell size at division triggered by both pharmacological and genetic TORC1 inhibition (Fig. 1c). Cells deleted for both gsk3 and drug treatment. Furthermore, deletion of gsk3Δ largely rescued the reduced cell size at division triggered by both pharmacological and genetic TORC1 inhibition (Fig. 1c). Cells deleted for both gsk3Δ and drug treatment. Furthermore, deletion of gsk3Δ largely rescued the reduced cell size at division triggered by both pharmacological and genetic TORC1 inhibition (Fig. 1c).
and tco89 also showed a short CLS, similar to the gsk3Δ single mutant (Fig. 1d). Together, these results indicate that fission yeast Gsk3 counteracts TORC1 function. This could be achieved either upstream of TORC1 by reducing TORC1 activity itself and/or downstream of TORC1 by inhibiting cell growth. These two scenarios are
not mutually exclusive, because Gsk3 is an upstream regulator affecting the TSC2 inhibitor of TORC125, which is conserved in fission yeast26 and has also been shown to function downstream of TORC127.

Gsk3 and AMPK act in parallel to inhibit TORC1-dependent processes. Previous studies have shown that AMPK negatively regulates TORC1-related processes28. We therefore tested the response of cells lacking amk2 (AMPKα) or amk2 (AMPKβ) for caffeine and rapamycin resistance. Like gsk3Δ cells, deletion of amk2 or amk2 resulted in cells whose growth was less inhibited in the presence of caffeine and rapamycin compared to wild-type cells (Fig. 2a). Furthermore, combined deletion of gsk3 and either amk2 or amk2 resulted in an even lower inhibition of cell growth by caffeine and rapamycin than shown by any of the three single mutants. This additive resistance to caffeine and rapamycin of the double mutants suggests that Gsk3 and AMPK operate in parallel to negatively regulate TORC1-dependent processes.

Ssp1 is a direct upstream activator of AMPK and ssp1 deletion results in loss of AMPK activity28,29. We therefore expected that the phenotype of ssp1Δ cells will mimic that of amk2Δ and ssp2Δ cells. Indeed, ssp1Δ cells showed lowered inhibition of growth by caffeine and rapamycin (Fig. 2b). However, the ssp1Δ gsk3Δ double mutant cells showed higher inhibition of cell growth by caffeine and rapamycin, in stark contrast to the amk2Δ gsk3Δ and ssp2Δ gsk3Δ double mutants. This result suggests that Ssp1 is required for the resistance of gsk3Δ cells to caffeine and rapamycin. To further explore the relationship between Ssp1 and Gsk3, we measured cell sizes at division of ssp1Δ and gsk3Δ double mutants (Fig. 2c). Thus, deletion of gsk3Δ exacerbates the elongated phenotype of ssp1Δ cells. The PInt tool30 predicts a physical interaction between Gsk3 and Ssp1, implying that Gsk3 can operate upstream of AMPK. However, using co-immunoprecipitation assays, we found no evidence for any direct interaction between Gsk3 and Ssp1 (data not shown). Together, these results reveal a functional relationship between Gsk3 and Ssp1 that is distinct from the relationship between Gsk3 and AMPK components.

To further investigate relationships between Gsk3 and AMPK, we performed CLS assays. As reported before15, gsk3Δ cells showed a reduced CLS in rich medium (Fig. 1d), whilst the CLS of amk2Δ cells was similar to that

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**Figure 2. Deletions of gsk3 and amk2 additively repress phenotypes associated with TORC1 inhibition.**

(a) Graph showing the times to achieve 25% increase in cell mass for wild-type (wt) and mutant cells as indicated under caffeine and rapamycin treatment. (b) Times to achieve 25% increase in cell mass as in panel A, with strains as indicated. (c) Cell size upon division for strains shown in panel B. (d) CLS assays for different strains as indicated. Error bars indicate standard deviations from three biological repeats.
of wild-type cells (Fig. 2d). However, the CLS of amk2Δ gsk3Δ double mutant cells was reduced compared to wild-type cells: initially the lifespan of the double mutant was intermediate of the single mutants, whilst in later time points was severely reduced. Together, these findings suggest that Gsk3 can act independently of AMPK in different contexts. In mammalian cells, GSK3 and AMPK signalling are integrated at TSC2, which then inhibits mTORC125 and the relationship between AMPK, the TSC1/2 complex and TORC1 is conserved in fission yeast28. It is not known whether Gsk3 and Tsc2 functionally interact in fission yeast. AMPK is linked to transcriptional responses, such as the nuclear accumulation of the transcription factor Ste11, which triggers sexual differentiation in response to nitrogen starvation29. Taken together, these results suggest that Gsk3 and AMPK largely act in parallel to mediate cellular responses related to TORC1 signalling.

Genetic interactions of gsk3 and amk2. Genetic (or epistatic) interactions reveal functional relationships between genes and pathways. Negative genetic interactions suggest compensatory, redundant pathways or protein complexes, while positive genetic interactions can reflect functions within the same pathway31, but most genetic interactions reveal broad phenotypic connections between functional modules or biological processes32. We conducted synthetic genetic array (SGA) analyses15,20 to identify genome-wide genetic interactions of gsk3 and amk2.

The query strains, which all showed equal fitness when grown on YES (data not shown), were crossed against all non-essential mutants in a deletion library19. We analysed genes showing positive or negative interactions with gsk3Δ for enrichments with AnGeLi 24, using the non-essential genes present in the deletion library as a reference19. The data for all genetic interactions are presented in Supplemental Table 1. As expected, a bias of negative interactions with gsk3 was evident around the genomic locus of gsk3, reflecting genetic linkage (blue line in Fig. 3a). For the same reason, a bias towards positive interactions was evident around the ade6 locus, because ade6 SGAs were used for control and normalisation purposes (red line in Fig. 3a).

Negative interactions with gsk3 were enriched for Gene Ontology (GO) categories related to protein translation, histone modification, endosome to vacuole transport, protein metabolic process and protein modification.
by small protein conjugation or removal, i.e., ubiquitination or ubiquitin-proteasome system (UPS). Further investigations will be required to define how and under which conditions Gsk3 may be functionally associated with the UPS. As Gsk3 suppresses TORC1-related phenotypes (Fig. 1), a plausible mechanism is through regulation of protein synthesis, which is a major determinant of CLS4,15,33. Positive interactions were enriched for genes related to abnormal cell shape, decreased population growth and stress (Supplemental Table 2).

Among the positive interactors of gsk3 was ste20 which encodes the Rictor homologue, a core component of TORC22. To validate this genetic interaction, we performed CLS assays. While ste20 mutant cells were even more short-lived than gsk3 mutant cells, the double mutant cells, albeit short-lived compared to wild-type, showed a somewhat longer CLS than either single mutant (Fig. 3b). Thus, gsk3 and ste20 show a positive genetic interaction in the phenotypic context of CLS. TORC2 is required for various stresses including oxidative stress 34. Both gsk3 and ste20 single mutants showed increased resistance to hydrogen peroxide (Fig. 3c), exhibiting a shorter lag-phase extension between treated and untreated conditions compared to wild-type 4,15. Surprisingly, the double mutant was sensitive to the same oxidative-stress treatment (Fig. 3c). As stress and growth programs are antagonistic35, this result of decreased stress resistance of the double mutant is consistent with the improved growth recorded in the SGA screens. The ste20Δ cells were larger upon division compared to wild-type, and this phenotype was further enhanced in the gsk3Δ ste20Δ double mutant (Fig. 3d). In contrast, the phenotype of the elongated sin1Δ mutant cells, which are also defective in TORC2, was diminished in the gsk3Δ sin1Δ double mutant. These results reveal diverse, complex and antagonistic genetic interactions of gsk3 with genes encoding TORC2 components. We conclude that the relationship between Gsk3 and TORC2 is multifaceted, and it is possible that TORC2 activities are determined by several distinct thresholds of activation.

As expected, the SGA screens with amk2 showed a negative interaction bias around the amk2 locus on chromosome 2 (blue line, Fig. 4a) and a positive interaction bias around the ade6 locus (red line, Fig. 4a). The negative interactions with amk2 were enriched for phenotypes36 such as sensitivity to chemical during vegetative growth.

**Figure 4. Relationships of genetic interactions of gsk3 and amk2.** (a) Physical mapping along the three chromosomes of mutants that genetically interact with gsk3. Blue and red lines indicate the amk2 and ade6 loci, respectively. (b) Distribution of gsk3 interactions with respect to amk2 interactions. Red lines indicate cut-offs used for positive and negative interactions. (c) Genetic interactions were classified as positive (interaction value i ≥ 0.15) negative (i ≤ 0.15) or neutral (−0.15 < i < 0.15) for both gsk3 and amk2. The numbers of genes for each category are indicated. (d) Venn diagram showing overlaps of negative genetic interactions for gsk3 and amk2. (e) Venn diagram showing overlaps of positive genetic interactions for gsk3 and amk2.
abnormal cellular physical quality (FYPO: 0004638), and chromatin silencing at centromere (FYPO: 0002346, 0003412) (Supplemental Table 2). Genes showing positive genetic interactions were enriched for phenotypes related to stress and abnormal cell shape. Although there was no significant enrichment in GO terms, many interacting genes were related to amino acid metabolism, lipid homeostasis, and DNA repair. Notably, loss of AMPK reduces the activity of enzymes responsible for UVB-induced DNA repair in mammalian cells.

Figure 5. Triple mutant SGA using gsk3 amk2 as query. (a) Physical mapping along the three chromosomes of mutants that genetically interact with gsk3 amk2 double mutant. Blue lines on Chromosomes I and III represent the genetic loci of gsk3 and amk2, respectively, while the red line indicates the ade6 locus. (b) Distributions of genetic interaction values for the three SGAs conducted in this study. Vertical dotted lines represent the cut-offs used. (c) Plot showing distribution of the gsk3 interaction values compared to the interaction values of the gsk3 amk2 double mutant. Red lines represent the cut-offs used. (d) Plot showing distribution the amk2 interaction values compared to the interaction values of the gsk3 amk2 double mutant. Red lines represent the cut-offs used. (e) Venn diagram showing overlaps of negative genetic interactions for the gsk3 and amk2 single mutants and the gsk3 amk2 double mutant. (f) Venn diagram showing overlaps of positive genetic interactions for the gsk3 and amk2 single mutants and the gsk3 amk2 double mutant.
The genetic interactions of gsk3 and amk2 were moderately correlated with each other (Fig. 4b). Genes that either positively or negatively interacted with both gsk3 and amk2 were more abundant than genes that interacted only with one of the two genes (Fig. 4c). There were 69 common negative and 82 common positive genetic interactions (Fig. 4d). These common gene lists showed no functional enrichments in AnGeLi44. We conclude that Amk2 and Gsk3 play roles in the same phenotypes (such as stress and abnormal cell shape) through both common and specific interactors.

**Triple mutant SGA: providing additional insights into Gsk3 and Amk2 functions.** Given that gsk3 and amk2 mutants suppress TOR-related phenotypes additively (Fig. 2), we also conducted an SGA screen to interrogate triple mutant cells, with gsk3Δ amk2Δ double mutant cells as query38,39. As in the case of the single mutant query SGA screens, linkage effects for positive and negative interactions were present in the double mutant query (Fig. 5a, blue and red lines for negative and positive interactions, respectively). The distribution of interaction values of the double mutant query showed a larger dynamic range than the values of the single mutant queries. This result reflects that a larger number of interactions were identified with the gsk3Δ amk2Δ query (Fig. 5b). The interactions of the gsk3Δ amk2Δ double mutant were positively correlated with those of both single mutant queries: gsk3Δ (R = 0.38, Fig. 5c) and amk2Δ (R = 0.46, Fig. 5d). A bias towards negative interactions was evident (see data points’ displacement in Figure c and d, and tail of double mutant line in Fig. 5b towards left part of graph). While yeast genomes only contain a few hundred essential genes, there are hundreds of thousands of double mutants that are lethal. For example, in budding yeast there are 1100 essential genes and 550,000 gene pairs that are essential together46. By extension, it is not surprising that triple mutants result in an even larger number of cases of synthetic lethality or sickness.

We identified 425 positive and 777 negative genetic interactions with gsk3 amk2 double mutants (applying cut-offs of +0.15 and −0.15, respectively). While positive interactions showed no functional enrichment (Supplemental Table 2), negative interactions were enriched in the GO terms ‘chromatin and gene silencing’, ‘protein translation’ and ‘gene products for localisation to the mitochondria’. Gsk3 and AMPK might together regulate factors linked to chromatin function and to respiration and detoxification of metabolic waste products. Interestingly, negative interactions were also enriched for genes induced during the oxidative stress response45. Compared to the single mutant queries, 305 positive and 496 negative genetic interactions were specific for the double mutant query. While the negative interactions were enriched for highly expressed genes, mutants with sensitivity to 5′-fluorouracil, a uracil analogue commonly utilised as a chemotherapeutic agent in the treatment of a range of cancers44 are significantly under-represented. Notably, AMPK is emerging as an interesting metabolic tumour suppressor and a promising target for cancer prevention and therapy43. This result highlights the principle that targeting multiple proteins simultaneously, e.g. by combined drugs, can greatly increase the effectiveness to combat cancer cells44, and triple action treatments open up even more possibilities to kill cancer cells.

In mammalian cells, Gsk3 is primarily known for its role in canonical Wnt signalling via β-catenin45. Previous work has also shown that Gsk3 operates in parallel to AMPK for mTORC1 inhibition via TSC218. Functional relationships of AMKP, Gsk3 and TOR are likely conserved from fission yeast to mammals. Our work provides rich data sets for both common and specific interactors that may relate to TORC1 function and associated cell physiology and disease.

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Author Contributions
C.R. designed the experiments, C.R. and S.T. did the experiments, C.R., S.T. and J.B. wrote the paper.

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This article contains a typographical error in the Materials and Methods section under the subheading ‘Growth assays’.

‘Caffeine and rapamycin were used at concentrations of 100 mM and 100 ng mL$^{-1}$, respectively while hydrogen peroxide at a concentration of 0.5 mM’

should read:

‘Caffeine and rapamycin were used at concentrations of 10 mM and 100 ng mL$^{-1}$, respectively while hydrogen peroxide at a concentration of 0.5 mM’.

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