Contrasting contributions of TNF from distinct cellular sources in arthritis

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ABSTRACT

Objectives Neutralisation of tumour necrosis factor (TNF) is widely used as a therapy for rheumatoid arthritis (RA). However, this therapy is only effective in less than a half of patients and is associated with several side effects. We hypothesised that TNF may possess non-redundant protective and immunomodulatory functions in vivo that cannot be blocked without a cost. The present work aimed to identify cellular sources of protective and pathogenic TNF, and its molecular forms during autoimmune arthritis.

Methods Mice lacking TNF expression by distinct cell types, such as myeloid cells and T or B lymphocytes, were subjected to collagen-induced arthritis (CIA) and collagen antibody-induced arthritis. Mice lacking soluble TNF production were also employed. The severity and incidence of the disease, as well as humoral and cellular responses were assessed.

Results Myeloid cell-derived TNF contributes to both induction and pathogenesis of autoimmune arthritis. Conversely, T cell-derived TNF is protective during the induction phase of arthritis via limiting of interleukin-12 production by dendritic cells and by subsequent control of autoreactive memory T cell development, but is dispensable during the effector phase of arthritis. B cell-derived TNF mediates severity of CIA via control of pathogenic autoantibody production.

Conclusions Distinct TNF-producing cell types may modulate disease development through different mechanisms, suggesting that in arthritis TNF ablation from restricted cellular sources, such as myeloid cells, while preserving protective TNF functions from other cell types may be superior to pan-anti-TNF therapy.

INTRODUCTION

In humans, tumour necrosis factor (TNF) is over-expressed in several autoimmune diseases such as rheumatoid arthritis (RA), ankylosing spondylitis and others. Furthermore, TNF overexpression in mice leads to polyarthritis. Although other factors are also involved in arthritis pathogenesis, TNF inhibition as monotherapy is beneficial for a fraction of patients with RA and is widely used as treatment. The success of such therapy points to the contribution of non-redundant and deleterious functions of TNF in the development of RA. On the other hand, TNF has been implicated in multiple aspects of immune regulation. Specifically, TNF may control the development of CD4+ T helper (Th1) and Th17 T cells in vivo and contributes to germinal centre (GC)-mediated antibody induction. Moreover, TNF induces expression of chemokines and adhesion molecules that mediate homing of proinflammatory cells.

Key messages

What is already known about this subject?

- Anti-tumour necrosis factor (TNF) therapy is widely used as a treatment for rheumatoid arthritis (RA) and is associated with several side effects that can be explained by beneficial functions of TNF that should not be neutralised.

What does this study add?

- The study links deleterious and protective TNF functions in arthritis to distinct cellular sources and molecular forms.
- TNF produced by myeloid cells controls arthritis induction through regulation of synovial fibroblast activation, while B cell-derived TNF regulates severity via induction of autoantibodies.
- TNF produced by T cells limits arthritis development via control of T cell responses during induction phase of arthritis but is dispensable for the pathogenesis.

How might this impact on clinical practice or future developments?

- Multifaceted contributions of TNF from distinct cellular sources at different stages of autoimmune arthritis suggest the advantage of cell type-specific TNF blockade for patients with RA and lay the ground for exploring myeloid cell-restricted TNF inhibition in arthritis.
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produced by myeloid cells as a critical factor for arthritis induction and pathogenesis. Second, we showed that T cell-derived TNF, likely in its membrane-bound form, provides protective functions through regulation of autoreactive T cell development during arthritis induction, but is dispensable during effector phase of arthritis. Finally, B cell-derived TNF controls development of autoantibodies during collagen-induced arthritis (CIA) and regulates severity of arthritis. Thus, our findings dissect the multifaceted contributions of TNF at different stages of autoimmune arthritis and underline intrinsic limitations of a non-selective pan-blockade of TNF.

RESULTS

Soluble TNF is critical for CIA development

TNF is a trimeric proinflammatory cytokine initially expressed in a membrane-bound form and then cleaved off the membrane by metalloproteases, such as ADAM17.17 18 Membrane-bound TNF is biologically active on cell-to-cell contacts and can also mediate reverse signalling.19–21 In order to address the contribution of soluble versus membrane-bound TNF in experimental arthritis, we first studied CIA in mice expressing only transmembrane form of TNF (tmTNF KI)22 or in mice lacking TNF expression by all cells (TNF KO).23 Around 50% of wild-type (WT) animals consistently developed CIA,24 while TNF- deficient and tmTNF KI mice failed to show any disease symptoms (figure 1A and online supplementary table S1), indicating that soluble TNF is critical for disease induction.

Figure 1 Soluble TNF is required for arthritis induction. (A) Incidence (left) and clinical scores (right) of CIA in tmTNF KI, WT and TNF KO mice. (B) Anti-collagen IgG titres in tmTNF KI, WT and TNF KO mice at day 14 after second immunisation. (C, D) Frequency of splenic CII-specific Th1 (IFNγ+) and Th17 (IL-17A+) CD4 T cell responses in tmTNF KI, WT and TNF KO mice at day 14 after second immunisation. All data are representative of two or more independent experiments with n≥10 for the analysis of arthritis incidence and severity and n≥3 for analysis of cellular and humoral responses. *p<0.05, **p<0.01, ***p<0.001, as calculated by Kruskal-Wallis non-parametric test with Dunn’s multiple comparisons test. CIA, collagen-induced arthritis; tmTNF, transmembrane form of TNF; TNF, tumour necrosis factor; WT, wild-type.

Taking into account that both autoreactive cytokine-producing T cells and autoantibodies may contribute to arthritis development and disease severity, we next evaluated these parameters in mice lacking TNF entirely or only its soluble form. Anticollagen II IgG titres were severely diminished in both TNF KO and tmTNF KI mice (figure 1B), indicating a crucial role of soluble TNF for autoantibody production. TNF has been reported to control both development and persistence (pathogenic memory) of autoreactive T cell responses.5 25 Given the importance of autoreactive memory T cells in disease perpetuation, we next quantified the frequency of collagen-specific CD4 T cells during active disease. Complete ablation of TNF resulted in augmented autoreactive CII-specific Th1 T cell responses characterised by increased IFNγ when compared to WT or tmTNF KI mice (figure 1C and D). Notably, these cytokine-producing CD4 T cells were found to be antigen-specific (online supplementary figure S1). Altogether these data suggested that soluble TNF is critical for disease induction and autoantibody production, whereas membrane-bound TNF limits autoreactive T cell development.

Opposite contributions of TNF produced by myeloid cells and T cells in CIA

TNF produced by various cell types may possess distinct non-redundant functions in various disease models.5 26 27 Thus, we next compared the contribution of TNF expressed by myeloid cells and T cells in arthritis induction. Mice lacking TNF in myeloid cells (M-TNF KO) exhibited significantly decreased disease incidence (figure 2A, C and D and online supplementary table S1), indicating that TNF from myeloid cells is pathogenic. In contrast, mice with TNF ablation in T lymphocytes (T-TNF KO) were highly susceptible to arthritis development (figure 2B and D and online supplementary table S1), suggesting that T-TNF may actually play a protective immunomodulatory role in the same disease model. Furthermore, analysis of infiltration of myeloid cells (acute inflammation) and lymphocytes (chronic inflammation) in affected joints revealed...
that T-TNF ablation was associated with increased CIA severity (figure 2D). Notably, deletion of myeloid cell-derived TNF did not affect accumulation of lymphoid cells, but led to decreased cartilage destruction (figure 2D), suggesting that M-TNF contributed to joint destruction during CIA. Thus, myeloid cell-derived and T cell-derived TNF exhibited opposing effects both in arthritis induction and in the control of disease severity.

**Soluble TNF produced by myeloid cells is required for arthritis induction**

Given that TNF produced by myeloid cells, on one hand, and the soluble form of TNF, on the other hand, are essential contributors for RA development, we next tested whether soluble form of TNF from myeloid cells is critical for arthritis induction. To do so, we generated mice expressing only transmembrane TNF by myeloid cells (tm-M-TNF KI mice) by crossing tmTNF KI mice to M-TNF KO animals. The resulting mice (tm-M-TNF KI) expressed only membrane-bound TNF on myeloid cells (online supplementary figure S2A,B), whereas both molecular forms of TNF were intact in all other cell types. Ablation of soluble TNF in myeloid cells resulted in decreased serum TNF levels during arthritis (figure 3A), whereas T cells did not contribute significantly to the systemic levels of TNF...
during CIA (figure 3A). Strikingly, ablation of soluble TNF production by myeloid cells resulted in reduced incidence of arthritis (figure 3B and online supplementary table S1), while autoantibody responses and autoreactive T cell responses were not affected (figure 3C and D). Thus, soluble TNF from myeloid cells controlled the arthritis development without affecting autoreactive T cell and humoral responses.

Recent reports have highlighted the importance of local joint macrophages in the pathogenesis of arthritis. Therefore, we next analysed the degree of TNF deletion in variable myeloid subsets. M-TNF KO mice exhibited efficient TNF deletion in neutrophils, while TNF has been only partially deleted in inflammatory monocytes (online supplementary figure S2C, D, E). Interestingly, analysis of TNF production by cells infiltrating inflamed paws revealed that in the joints of M-TNF KO mice TNF is produced only by T cells, indicating that neither infiltrating, nor resident myeloid cells are the source of TNF in paws of arthritic M-TNF KO animals. Furthermore, joint swelling is associated with influx of neutrophils in M-TNF KO mice, while both neutrophils and inflammatory monocytes were detected in arthritic paws of WT and T-TNF KO animals (online supplementary figure S3). Thus, it is plausible that TNF from both resident and circulating myeloid cells might contribute to arthritis induction.

Enhanced production of TNF may result in activation of synovial fibroblasts characterised by upregulation of podoplanin (PDPPN) and Thy1, as well as expression of metalloproteases and interleukin-6, leading to tissue destruction. Thus, we next addressed whether TNF produced by myeloid cells may regulate synovial fibroblast activation. Notably, TNF KO mice showed no signs of synovial fibroblast activation as revealed by Thy1/PDPPN staining (figure 3E,F and data not shown), while TNF ablation in myeloid cells resulted in reduced presence of activated fibroblasts during CIA. Consistently, expression levels of MMP9, TNF and IL-6 were significantly decreased in M-TNF KO joints when compared with WT counterparts (figure 3G and online supplementary figure S2). Altogether, these data suggested that M-TNF, likely in soluble form, may control arthritis induction via activation of synovial fibroblasts.

Concomitant ablation of TNF in myeloid cells and T cells results in decreased CIA incidence

Next, we tested that M-TNF may be both necessary and sufficient for disease induction, while T cell-derived TNF exhibits its protective role during arthritis. For this, we induced arthritis in mice lacking TNF production by both cell types (M-T-TNF KO) and found that disease development was significantly abrogated in M-T-TNF KO mice as compared with WT counterparts (figure 3H and online supplementary table S1). Furthermore, M-T-TNF KO animals exhibited elevated levels of IFNγ-producing T cells in spleen (figure S3I), thus phenocopying CIA development in TNF KO. These findings further corroborated on the importance of myeloid cell-derived TNF for CIA induction, even in the absence of protective T-TNF.

Membrane-bound TNF produced by T cells controls disease severity by limiting the expansion of autoreactive T cells

Since arthritis development is driven by arthritogenic Th1 and Th17 CD4 T cells, we hypothesised that TNF from various cellular sources may have distinct effects on arthritogenic T cell populations. To get further insights into how TNF produced by T cells may control arthritis development, we analysed the arthritogenic, cytokine-producing CD4+ T cells in various TNF-deficient mice during active disease. Importantly, T-TNF KO mice contained more autoreactive IFNγ-producing and IL-17A-producing CD4+ T cells in their spleens when compared with WT counterparts (figure 4A and B). Of note, myeloid cell-derived TNF did not influence frequencies of CII-specific Th1 or Th17 CD4 T cells (figure 4A and B). TNF–TNFR2 axis is known to modulate T reg frequencies and function via control of FoxP3 expression and drive pathologic state of sinovial fibroblasts (SFs). However, ablation of TNF only in T cells or in myeloid cells had no impact neither on T reg numbers in lymphoid organs, nor on FoxP3 expression levels (online supplementary figure S4A, B), suggesting that T-TNF is dispensable for T regs modulation in this model. Finally, blockade of IFNγ, but not of TNFR2 or IL-17A, significantly reduced arthritis incidence in those mice during CIA (online supplementary figure S4C and data not shown). Altogether, this suggests that TNF derived from T cells controls CIA development by limiting autoreactive Th1 response.

Figure 4 T-TNF regulates memory T cell responses during CIA. (A) Analysis of TNF production in inflamed paws in TNF KO, T-TNF KO, M-TNF KO and WT mice at day 14 after second immunisation. (B, C) Frequency of anticollegen type II Th1 (IFNγ+) and Th17 (IL-17A+) CD4 T cell responses in T-TNF KO, M-TNF KO and WT mice spleen at day 14 after second immunisation. Frequency of total IL-12p40-producing cells (D), IL-12p40-producing inflammatory monocytes (CD11b+Ly6C+Gr1low) and (CD11c+MHCIi+) (E) in spleens of WT, T-TNF KO and M-TNF KO mice at day 6 after immunisation. Splenocytes were stimulated with heat-killed M.TB H37Ra (100 mcg/ml) for 6 hours in the presence of brefeldin A and were analysed for the IL-12p40 expression using flow cytometry. All data are representative of two or more independent experiments with n≥3. *p<0.05, **p<0.01, ***p<0.001, as calculated by Kruskal-Wallis non-parametric test with Dunn’s multiple comparisons test. CIA, collagen-induced arthritis; M-TNF, mice lacking TNF in myeloid cells; T-TNF, TNF ablation in T lymphocytes; TNF, tumour necrosis factor; WT, wild-type.
Since TNF is known to control proinflammatory cytokine production by myeloid cells, we next quantified the number of IL-12p40-producing cells at day 6 after immunisation. Ablation of T cell-derived TNF, but not of TNF from myeloid cells, augmented frequencies of IL-12p40-producing cells in the spleen (figure 4C), suggesting that in WT mice T-TNF may dampen IL-12p40 production in vivo during immune responses. Further analysis revealed increased numbers of IL-12p40-producing monocytes and dendritic cells on T-TNF deletion (figure 4D), while the frequency of inflammatory monocytes was not changed in these mice (online supplementary figure S4D, E). Altogether, these data suggest that T-TNF regulates Th1 and Th17 responses via control of IL-12p40 production by inflammatory monocytes and dendritic cells.

**Myeloid cell-derived TNF drives pathogenesis in collagen-antibody induced arthritis**

Cell type-specific TNF deletion strategy during CIA affected gene expression both during induction of autoimmune responses and during effector phase of autoimmune disease. In order to further delineate the contribution of distinct cellular sources to arthritis pathogenesis, we utilised the model of collagen-antibody induced arthritis (CAIA) that is driven by preformed autoantibodies, is independent on the adaptive immune system and is known to be critically dependent on soluble TNF.37 TNF KO mice were largely protected from the disease development (figure 5A). Strikingly, myeloid cell-derived TNF was found to be crucial for CAIA development (figure 5A–C), while T-TNF was dispensable for disease manifestation (figure 5A–C). Furthermore, both TNF KO and M-TNF KO mice exhibited reduced activation of synovial fibroblasts during CAIA (figure 5D), when compared with WT mice. Of note, gp38 Thy1SFs that are important for the tissue destruction seemed to be absent in mice lacking TNF in myeloid cells on CAIA (figure 5D). Finally, M-TNF KO mice had significantly lower IL-6 levels in their blood during CAIA (figure 5E). Altogether, myeloid cell-derived TNF drives arthritis induction during CAIA via activation of synovial fibroblasts and IL-6 production.

**B cell-derived TNF controls autoantibody production and disease severity during CIA**

Since TNF controls autoantibody development during CIA (figure 1), we next wondered which cellular source is crucial for anti-CII antibody production during CIA. Several studies have described the critical contribution of B cell-derived TNF for the follicular dendritic cell development and GC reactions.8 9 38 Thus, we next analysed anti-CII IgG titres (figure 6A) in various knockout strains during CIA and observed the reduction of anti-collagen type II-specific IgG, IgG1 and IgG2a titres in B-TNF KO mice during CIA (figure 6A). Interestingly, B-TNF KO developed disease with similar incidence when compared to WT littermate controls (figure 6B), but with significantly lower clinical score (figure 6C), suggesting that B-TNF controls severity of CIA. Furthermore, B cell-derived TNF controlled frequency of GC B cells in spleen during CIA, but did not influence numbers of either IFNγ-producing or IL-17A-producing CD4 T cells (figure 6D).

**DISCUSSION**

There are multiple biological therapies currently approved for RA treatment with predominance of anti-TNF biologics.1 2 However, anti-TNF therapy is associated with multiple, although relatively rare, side effects: susceptibility to intracellular infections, particularly reactivation of Mycobacterium tuberculosis, secondary autoimmune manifestations and an increased risk of lymphoma development, all due to several known non-redundant beneficial functions of TNF in host defence, immune and body homeostasis.13 40 Here we uncover the significance of non-redundant contributions of TNF from distinct cellular sources in the pathogenesis of experimental arthritis that implicate cell type-restricted TNF blockade as a promising strategy for the treatment of RA. Our data suggest that TNF produced by T cells controls activation of T cells in the secondary lymphoid organs and that T-TNF blockade using conventional anti-TNF therapy may abolish this protective effect, resulting in activation of autoreactive T cells capable of initiating secondary autoimmunity in patients treated with anti-TNF therapeutics.

Myeloid cells, a component of innate immune system, are long implicated in arthritis pathogenesis.41 Selective depletion of myeloid cell subsets, such as monocytes, macrophages and neutrophils, further revealed their critical impact on disease induction and severity.42 43 Using highly selective genetic TNF ablation in mice, we have demonstrated the functional importance of TNF produced by myeloid cells in disease pathogenesis. Mechanistically, M-TNF acts in its soluble form to activate

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**Figure 5** Myeloid-derived TNF controls severity of CAIA. (A) Clinical score of CAIA in M-TNF KO, T-TNF KO, TNF KO and WT mice. (B) Safranin O, fast green and haematoxylin staining of knee joints from T-TNF KO, M-TNF KO, TNF KO and WT mice at day 10 after induction of CAIA. (C) Histopathological scores of knee joint sections from T-TNF KO, M-TNF KO, TNF KO and WT mice at day 10 after induction of CAIA. (D) Immunofluorescent staining (THY1, gp38) of knee joints from WT, T-TNF KO, M-TNF KO and TNF KO at day 10 after induction of CAIA. (E) Serum levels of IL-6 in WT, T-TNF KO, M-TNF KO and TNF KO at day 10 after induction of CAIA. All data are representative of two or more independent experiments with n≥6 for the analysis of arthritis clinical scores and serum IL-6 levels and n≥3 for histopathological evaluation. **p<0.05, ***p<0.01, ****p<0.001, as calculated by Kruskal-Wallis non-parametric test with Dunn’s multiple comparisons test. CAIA, collagen antibody-induced arthritis; M-TNF, mice lacking TNF in myeloid cells; T-TNF, TNF ablation in T lymphocytes; CIA, collagen-induced arthritis; WT, wild-type.
Figure 6  B cell-derived TNF controls severity of CIA. (A) Anticollagen type II antibody titres in WT, T-TNF KO, M-TNF KO and TNF KO mice at day 21 after immunisation. (B) Incidence of CIA in B-TNF KO, M-TNF KO and WT mice. (C) Clinical scores of CIA in B-TNF KO and WT mice. (D) Frequency of B cells (CD19+ cells) and GC B cells (CD19+GL7+PNA+) in WT, B-TNF KO and TNF KO mice. (E) Frequency of anticontigen type II Th1 (IFNγ+) and Th17 (IL-17A+) CD4 T cell responses in WT, B-TNF KO and TNF KO mice at day 14 after second immunisation. All data are representative of two or more independent experiments with n=10 for the analysis of arthritis incidence and severity and n=3 for analysis of cellular and humoral responses. *p<0.05, **p<0.01, ***p<0.001, as calculated by Kruskal-Wallis non-parametric test with Dunn’s multiple comparisons test with Dunn’s multiple comparisons test. GC, CIA, collagen-induced arthritis; GC, germinal centre; M-TNF, mice lacking TNF in myeloid cells; T-TNF, TNF ablation in T lymphocytes; TNF, TNF, tumour necrosis factor; WT, wild-type.

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synovial fibroblasts. On the other hand, myeloid cells are the main contributors to systemic serum TNF in arthritic mice, suggesting that increased TNF levels may initiate arthritis in the presence of arthritogenic adaptive immunity. Indeed, M, T-TNF KO mice that lack TNF from myeloid cells and T cells displayed strongly reduced arthritis induction despite enhanced autoreactive cytokine-producing CD4 T cells and autoantibody development. TNFR signalling in synovial fibroblasts is crucial for synovial fibroblasts activation and may serve as the initial signal for arthritis development. Altogether, it is plausible that soluble TNF produced by myeloid cells initiates arthritis via activation of fibroblasts in the joint in the presence of autoantibodies. Recent report described the development of bispecific TNF inhibitor that selectively inhibits TNF production by F4/80-expressing myeloid cells and is intended for arthritis treatment through selective ablation of TNF on macrophages.

TNF exerts its biological actions via two distinct receptors, TNFR1 and TNFR2 with distinct functions; while signalling via TNFR1 is driving arthritis pathogenesis, TNFR2 signalling exhibits protective functions. Membrane-bound TNF was reported to preferentially bind to TNFR2, and it is possible that the tmTNF–TNFR2 signalling axis is crucial for antigen presenting cells tuning by T cells in the secondary lymphoid organs. Indeed, blockade of both TNFR1 and TNFR2, but not TNFR1, not only ameliorated the disease, but also enhanced T cell responses. Finally, TNFR2 expressed on Tregs is instrumental for their suppressive action. Altogether, this data suggest that novel anti-TNF therapies should be designed not to interfere with tmTNF–TNFR2 axis to avoid enhanced development of autoimmune T cells. Finally, an earlier study revealed that T cell-derived TNF is critical for the control of chronic experimental M. tuberculosis infection, whereas myeloid cell-derived TNF is dispensable for the survival on M. tuberculosis challenge.

Based on these findings we propose that the next generation anti-TNF therapy should preserve TNF produced by T cells, and that myeloid cell TNF constitutes reasonable selective therapeutic target for the treatment of arthritis.

**MATERIALS AND METHODS**

Detailed explanation of each procedure was described in the online supplementary file 1.

Mice with ablation of TNF in various cell types used in this study were described elsewhere. All animal procedures were carried out in accordance with German and Russian regulations for animal protection. CIA was performed by immunisation of mice with chicken collagen II in complete Freund’s adjuvant. CAIA was induced by injection of monoclonal anti-CII antibodies (Chondrex). Histological analysis of knee joints was performed during CIA and CAIA. Analysis of cytokine production by T cells during CIA was performed by flow cytometry after restimulation with collagen II. Cytokine production and autoantibody production were assessed by ELISA. Gene expression was measured by real-time PCR. All results were statistically evaluated using by Kruskal-Wallis non-parametric test with Dunn’s multiple comparisons test unless otherwise stated. P values (p<0.05) were considered as statistically significant.

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**Contributors** AK and SN designed research. AK, MD, DS, KK, EG and LM performed experiments. AK, MD and SN wrote the manuscript.

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**Competing interests** None declared.

**Patient and public involvement** Patients and/or the public were not involved in the design, or conduct, or reporting or dissemination plans of this research.

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