The impact of an invasive ambrosia beetle on the riparian habitats of the Tijuana River Valley, California

John M Boland

The Tijuana River Valley is the first natural habitat in California to be substantially invaded by the Kuroshio Shot Hole Borer (KSHB, Eucallacea sp.), an ambrosia beetle native to Southeast Asia. This paper documents the distribution of the KSHB in the riparian vegetation in the valley and assesses the damage done to the vegetation as of early 2016, approximately six months after the beetle was first observed in the valley. I divided the riparian habitats into 29 survey units so that the vegetation within each unit was relatively homogenous in terms of plant species composition, age and density. From a random point within each unit, I examined approximately 60 individuals of the dominant plant species for evidence of KSHB infestation and evidence of major damage such as limb breakage. In the 22 forested units, I examined the dominant arroyo and black willows (Salix lasiolepis Benth. and S. gooddingii C.R. Ball), and in the seven scrub units, I examined mule fat (Baccharis salicifolia (Ruiz & Pav.) Pers.). Evidence of KSHB infestation was found in 25 of the 29 units. In the forest units, infestation rates ranged from 0 to 100% and were high (>60%) in 16 of the units. In the scrub units, infestation rates ranged from 0 to 33%. Infestation rates were significantly correlated with the wetness of a unit; wetter units had higher infestation rates. Evidence of major physical damage was found in 24 units, and dense stands of willows were reduced to broken trunks in several areas. Overall, I estimated that more than 280,000 (70%) of the willows in the valley were infested, and more than 140,000 had suffered major limb damage. In addition, I recorded evidence of KSHB infestation in the other common plant species in the valley; of the 23 species examined, 14 showed evidence of beetle attack. The four species with the highest rates of infestation were native trees in the Salicaceae family. The three species considered to be the worst invasive plants in the valley, Ricinus communis L., Tamarix ramosissima Ledeb. and Arundo donax L., had low rates of infestation. Several findings from this study have significance for resource managers: (1) the KSHB attack caused extensive mortality of trees soon after being first discovered so, if managers are to control the spread of the beetle, they will need to develop an effective early detection and rapid response program; (2) infestation rates were highest in units that were wet, so resource managers trying to detect the beetle in other areas should thoroughly search trees near water, particularly
nutrient-enriched water; (3) the infestation appears to be a novel form of disturbance, and the affected forests may need special management actions in order to recover; and (4) the infestation has altered the structure of the forest canopy, and this is likely to promote the growth of invasive plant species that were relatively inconspicuous in the forests prior to the beetle attack but will now need more attention.
THE IMPACT OF AN INVASIVE AMBROSIA BEETLE
ON THE RIPARIAN HABITATS OF THE TIJUANA RIVER VALLEY, CALIFORNIA

John M. Boland
Southwest Wetlands Interpretive Association
Imperial Beach, CA 91932
USA

JohnBoland@sbcglobal.net

24 May 2016
The Tijuana River Valley is the first natural habitat in California to be substantially invaded by the Kuroshio Shot Hole Borer (KSHB, Euwallacea sp.), an invasive ambrosia beetle native to Southeast Asia. This paper documents the distribution of the KSHB in the riparian vegetation in the valley and assesses the damage done to the vegetation as of early 2016, approximately six months after the beetle was first observed in the valley.

I divided the riparian habitats into 29 survey units so that the vegetation within each unit was relatively homogenous in terms of plant species composition, age and density. From a random point within each unit, I examined approximately 60 individuals of the dominant plant species for evidence of KSHB infestation and evidence of major damage such as limb breakage. In the 22 forested units, I examined the dominant arroyo and black willows (Salix lasiolepis Benth. and S. gooddingii C.R. Ball), and in the seven scrub units, I examined mule fat (Baccharis salicifolia (Ruiz & Pav.) Pers.). In addition, I recorded evidence of KSHB infestation in other common species as they were encountered.

Evidence of KSHB infestation was found in 25 of the 29 units. In the forest units, infestation rates ranged from 0 to 100% and were high (>60%) in 16 of the units. In the scrub units, infestation rates ranged from 0 to 33%. Infestation rates were significantly correlated with the wetness of a unit; wetter units had higher infestation rates. Evidence of major physical damage was found in 24 units, and dense stands of willows were reduced to broken trunks in several areas. Overall, I estimated that more than 280,000 (70%) of the willows in the valley were infested, and more than 140,000 had suffered major limb damage. Of the 23 common plant species examined, 14 showed evidence of beetle attack. The four species with the highest rates of infestation were native riparian trees in the Salicaceae family. The three species considered to be the worst invasive plants in the valley, Ricinus communis L., Tamarix ramosissima Ledeb. and Arundo donax L., had low rates of infestation.

Several findings from this study have significance for resource managers: (1) the KSHB attack caused extensive mortality of trees soon after being first discovered so, if managers are to control the spread of the beetle, they will need to develop an effective early detection and rapid response program; (2) infestation rates were highest in units that were wet during spring and summer, so resource managers trying to detect the beetle in other areas should thoroughly search trees near water, particularly nutrient-enriched water; (3) the infestation appears to be a novel form of disturbance, and the affected forests may need special management actions in order to recover; and (4) the infestation has altered the structure of the forest canopy, and this is likely to promote the growth of invasive plant species that were relatively inconspicuous in the forests prior to the beetle attack but will now need more attention.
INTRODUCTION

Accidentally-introduced insect pests have caused major economic losses and environmental damages within the U.S. (Pimentel, Zuniga & Morrison 2005). Examples include the elm bark beetle (Scolytus multistriatus Marsham), which has spread Dutch elm disease in North America and killed an estimated 75% of all the elms (Ulmus spp.; Kendrick 2000), and the balsam woolly adelgid (Adelges piceae (Ratz.)), which has severely damaged the spruce-fir forests of southern Appalachia, killing up to 95% of the Fraser fir trees (Abies fraseri (Pursh) Poir) and substantially changing the avian community (Rabenold et al. 1998, Gandhi and Herms 2010).

Two accidentally-introduced ambrosia beetles are threatening to cause similar ecosystem-wide damages to the riparian habitats of southern California. These beetles are the Polyphagous Shot Hole Borer (PSHB; Euwallacea sp. near fornicatus; Coleoptera: Curculionidae: Scolytinae) and the Kuroshio Shot Hole Borer (KSHB, Euwallacea sp.; Eskalen 2016). The two species are morphologically identical and are distinguished by their DNA sequences and by their associated fungi (Eskalen 2016). They are part of a species complex that also includes the Tea Shot Hole Borer (Euwallacea fornicatus (Eichhoff)). The PSHB was first documented in Los Angeles County in 2003, and the KSHB was first observed in San Diego County in 2012 (Eskalen et al. 2013; Eskalen 2016; Umeda, Eskalen & Paine 2016). Both are believed to be native to Southeast Asia and both attack many tree species in southern California, including native species, landscape trees, and the economically important avocado (Persea americana Mill.; Freeman et al. 2013; Eskalen et al. 2013). The ever-increasing list of reproductive host plants used by these species is currently at 41 species for the PSHB and 15 species for the KSHB (Eskalen 2016).

Both ambrosia beetles were initially observed to cause problems in urban and agricultural settings, but their recent appearance in natural settings has raised grave, new concerns. Both damage or kill trees through their boring activities and their spread of fungal pathogens. Females bore into tree trunks, create networks of tunnels in the xylem, inoculate the tunnels with a fungus (e.g., Fusarium sp.), and live in the tunnels eating the fungus and reproducing (Biedermann, Klepzig & Taborsky 2009). Within a few weeks females emerge, fly to new trees, and perpetuate the infestation (Rudinsky 1962). Most trees appear to die from the fungal infection in their tissues (Freeman et al. 2013). Beyond this life cycle, little is known about the effects of any ambrosia beetle in natural habitats because the emphasis of research investigations has been on their presence as pests of commercial agriculture and lumber rather than in natural areas (Wood 1982; Hulcr and Dunn 2011).

In late summer 2015, the KSHB was found in the riparian forests of the Tijuana River Valley, making these forests the first natural habitats in California to be substantially attacked by an invasive ambrosia beetle. These forests are dominated by two willow species, Salix lasiolepis Benth. (arroyo willow) and Salix gooddingii C.R. Ball (Goodding’s black willow), which account for more than 80% of the individuals and create the vertical structure of the forest (Boland 2014a). Both willows were attacked by the KSHB and, within only a few months, tens of thousands of trees were visibly infested. The forests were so obviously negatively impacted by the beetle that the infestation was covered by the local news media (e.g., Graham 2016, Smith 2016).
Here I describe the distribution of the KSHB in the riparian habitats of the Tijuana River Valley during the six-month period after first observation, assess the damage caused by the beetle, and discuss the prospects of the habitats recovering from this unusual damage. The overarching goal of this paper is to alert resource managers to this emerging beetle problem in natural habitats.

**STUDY SITE**

The Tijuana River Valley (32° 33.080'N, 117° 4.971'W) in San Diego County, California, is a coastal floodplain of approximately 1,500 ha at the end of a 448,000 ha watershed (Figure 1). The river is a managed intermittent stream that typically flows strongly in winter and spring and is mostly dry in summer (Boland 2014a). In 2015, however, the main river channels contained water all summer because of unusual rain storms on May 7, May 15, July 18 and September 15. The main river splits into two in the center of the floodplain at Hollister Bridge, and the northern arm carries more of the flows than the southern arm because of extensive sedimentation west of Hollister Street within the southern arm.

Riparian forests in the river bed are numerically and structurally dominated by *S. lasiolepis* and *S. gooddingii*, and the surrounding riparian scrub is numerically and structurally dominated by the perennial shrub, *Baccharis salicifolia* (Ruiz & Pav.) Pers. (mule fat; Boland 2014a). Zonation of these three dominant species across the elevation gradient and the factors that produce their zonation were described in Boland (2014a).

The riparian forest and scrub habitats are preserved within three adjoining parks: the Tijuana River Valley Regional Park, the Border Field State Park, and the Tijuana Slough National Wildlife Refuge. The riparian habitats are relatively undisturbed and support numerous reptile, mammal and bird species, most notably the endangered *Vireo bellii pusillus* Coues (least Bell's vireo) for which most of the riparian habitats are designated critical habitats (U.S. Fish and Wildlife Service 1994).

The beetles causing the damage in the riparian habitats of the Tijuana River Valley during 2015-16 were collected and identified as the KSHB by Dr. Akif Eskalen at University of California Riverside (UCR; Eskalen 2016). These specimens have been stored in the UCR collection.

**MATERIALS & METHODS**

**Infestation and damage rates in the valley**

To estimate the extent and magnitude of infestation and damage in the valley, I surveyed the entire valley in a stratified random manner. I divided the valley’s riparian forest and scrub habitats into 29 survey units so that the vegetation within each was relatively homogenous in terms of plant species composition, age and density. Within each unit, an accessible survey point was chosen at random from among several accessible points. The location of each survey point was recorded using a handheld GPS unit (Garmin eTrex Venture HC) and the units were mapped (ArcGIS 10.2.2. and projected in Universal Transverse Mercator). I did four surveys in each unit focused on the dominant plant species, *S. lasiolepis* and *S. gooddingii* in the forest units, and *B. salicifolia* in the scrub units.
First, I examined *S. lasiolepis*, *S. gooddingii* and *B. salicifolia* for evidence of KSHB infestation, using binoculars when necessary. A plant was counted as infested (or attacked) if it had beetle holes, extrusion of sawdust plugs or frass, or gumming out of sap (Figure 2A-D). Plants with only weeping were noted but included with the non-infested plants in the final count (Hulcr 2012). All plants examined were within 50 m of the survey point. Surveys were conducted between November 2, 2015 and January 22, 2016. The rate of infestation (the percent infested) was calculated for each forest unit based on *S. lasiolepis* and *S. gooddingii* and for each scrub unit based on *B. salicifolia*.

Second, I examined *S. lasiolepis*, *S. gooddingii* and *B. salicifolia* for evidence of major plant damage. A plant was counted as damaged if it had a recently-broken trunk or major limb (Figure 2F). All plants examined were within 50m of the survey point. These surveys were conducted after the first winter storms, between December 14, 2015 and January 22, 2016. The rate of damage (the percent damaged) was calculated for each forest unit based on *S. lasiolepis* and *S. gooddingii* and for each scrub unit based on *B. salicifolia*.

Third, within the forest units I measured the girth of the *S. lasiolepis* and *S. gooddingii* trees at breast height. All plants measured were within 50m of the survey point. An average of 33 (+ 7) trees was measured in each unit, and the median value within each unit was determined.

Fourth, I recorded the species composition and density of all plant species. All perennial trees and shrubs taller than 0.5m were identified and counted within a belt transect (20m x 2m = 40 m²) that started at the survey point. Transects were done between November 2, 2015 and January 22, 2016. To estimate the total number of *S. lasiolepis* and *S. gooddingii* individuals in each forest unit and the total number of *B. salicifolia* individuals in each scrub unit, the density of these species in the transect was multiplied by the area of the unit. The numbers of individuals infested and damaged in each unit were then extrapolated by multiplying the total number of individuals by the infestation and damage rates in each unit.

Finally, I assigned a measure of wetness to each unit based on the distance of the survey point from surface water during summer 2015. The measure was used to test whether infestation rate in a site was correlated with the relative wetness of the site.

**Infestation rates in the common plant species**

I examined all common tree and shrub species for signs of infestation when they were encountered while doing the surveys above. As above, a plant was counted as infested if it had beetle holes, extrusion of sawdust plugs or frass, or gumming out of sap, whereas plants with only weeping were noted but included with the non-infested plants in the final count. In total, 23 species were examined, and the rate of infestation (the percent infested) was calculated for each species.

**RESULTS**

**Infestation and damage rates in the valley**

Evidence of KSHB infestation was widespread throughout the valley, being observed in all but four of the 29 survey units (Table 1). Rates of infestation were high (61 – 100%) in 16 of the 22
forest units (Units 1 – 14, 19, 22). Most of the *S. lasiolepis* and *S. gooddingii* in these units had obvious sawdust and frass coming out of abundant holes indicating active boring of the beetles within. Rates of infestation were low (0 – 10%) in six of the forest units (Units 15 – 18, 20 - 21) and in all of the scrub units (0 – 33%; Units 23 – 29). Overall, an estimated 287,620 willow trees (71% of the total) and 16,641 mule fat shrubs (4%) were infested by the KSHB (Table 1).

The forest units with the highest infestation rates were scattered throughout the valley, and the forest units with the lowest infestation rates were contiguous in the center of the valley (Figure 3). To find a possible cause for this distribution pattern I looked for correlations between the characteristics of the forest units and their infestation rates. There were no significant correlations between the infestation rates of the forest units and the age of the willow stands ($r^2 = 0.355$, $p > 0.05$, $n = 22$), average willow densities ($r^2 = 0.078$, $p > 0.05$, $n = 22$) and median willow girths ($r^2 = 0.1157$, $p > 0.05$, $n = 20$). However, there was a significant negative correlation between infestation rates of the forest units and distance from surface water ($r^2 = 0.639$, $p < 0.01$, $n = 22$), as well as a significant negative correlation between infestation rates of all units and their distance from surface water ($r^2 = 0.577$, $p < 0.01$, $n = 29$; Figure 4). Units that were wet during spring and summer 2015 had the highest infestation rates, whereas units that were dry and far from surface water had the lowest infestation rates. The driest forests were all along the southern arm of the river, which did not receive abundant surface flows during 2015 because of heavy sedimentation.

Major plant damage was also widespread in the valley, with some damage observed in all but five of the 29 survey units (Table 1). The magnitude of the damage was particularly high in the eastern forest units (Units 2 – 13) where more than 50% of the trees had major damage. Before-and-after photos taken of Unit 2 show the kind of damage seen in the more heavily infested and damaged forest units (Figure 5). The native riparian forest in this unit went from a dense stand of tall willows to a jumble of broken limbs in just a few months. Examination of the broken limbs showed evidence of KSHB infestation and extensive tunneling (Figure 2E) and showed that the KSHB had so weakened the trunks that the first strong wind was able to snap them (Figure 2F). The amount of damage within the forest units was significantly positively correlated with the infestation rates in those units ($r^2 = 0.533$, $p < 0.01$, $n = 22$); units with high infestation rates generally had high damage rates. Overall, an estimated 141,011 willow trees (35% of the total) and 4,140 mule fat shrubs (1%) were damaged by the KSHB.

**Infestation rates in the common plant species**

Of the 23 species examined, 14 species showed obvious signs of beetle attack (Table 2). Among the native species, the tree species showed relatively high rates of infestation (>20%) and the shrub species relatively low rates of infestation (<20%). The four species with the highest rates of infestation (>50%) were all native riparian trees belonging to the Salicaceae family, i.e., *S. lasiolepis*, *S. gooddingii*, *S. laevigata* Bebb. and *Populus fremontii* S. Watson. Among the non-native species, the three species considered to be the worst invasive plants in the valley, *Ricinus communis* L., *Tamarix ramosissima* Ledeb., and *Arundo donax* L., had relatively low rates of infestation – 13%, 3% and 0%, respectively. Young, immature plants of *S. gooddingii*, *B. salicifolia* and *R. communis* showed no signs of infestation although all three species were frequently attacked as adults (Table 2).
DISCUSSION

The Tijuana River Valley is the first natural riparian habitat in California to be substantially attacked by an invasive, non-native ambrosia beetle; KSHB attacks in California have previously been observed only in agricultural and urban settings (Eskalen et al. 2013). The goals of this study were to describe the KSHB infestation in the Tijuana River Valley and to alert resource and land managers to the beetle problem in natural habitats. Several findings from this study have significance for resource and land managers.

First, the KSHB attack caused extensive damage to the trees in the riparian forests. Many dense stands of tall willows were reduced to broken trunks, making the magnitude and severity of the beetle attack on a par with a medium-severity fire (Brumby et al. 2001). Furthermore, the damage appeared to occur rapidly – in just a matter of months. The KSHB was first observed in the valley in August 2015 (pers. obs.) and as many as 100,000 trees had their branches or trunks snapped by the first winter wind storms in December 2015. It is likely that the beetle was present in low numbers well before it was first observed and that it had time to build up its population to outbreak levels (Crooks 2005). However the apparent speed of attack means that if resource managers are to control the spread of the beetle in their riparian forests, they will need to develop an early detection program that will discover the beetle when it is at very low population levels (Porter 2007). They will also need a rapid response program in place so that they can respond as soon as a small population of the beetle is discovered. A program currently being discussed in San Diego County involves the monitoring of apparently uninfested riparian habitats and the removal of any infested trees, followed by their chipping or solarization (placing in the sun under a clear tarp for several months; Jones and Paine 2015). A research group at University of California Riverside is exploring many control options, including fungicides, pesticides and biocontrol, primarily for the avocado industry but their findings are likely to assist in the control of the beetles in native habitats as well (Spann 2015, Eskalen 2016).

Second, KSHB infestation rates in the forest units were significantly correlated with the proximity to surface water or the wetness of the unit. This correlation suggests that the beetles were more attracted to or active in the wetter parts of the valley where trees grow in or near water. In these wetter areas, trees would have high internal moisture content, because the moisture content of plant tissues is positively correlated with that of the surrounding soil (Constantz and Murphy 1990), and they would be growing vigorously, because the water and soils are nutrient enriched due to frequent cross-border sewage flows. The idea that an ambrosia beetle would prefer to attack trees growing in wet, nutrient-rich conditions has support in the literature; Rudinsky (1962), in his review of the ecology of the Scolytidae, reported that growth of the fungi and larvae of ambrosia beetles was positively correlated with moisture content of the host plants, and Coyle, Booth & Wallace (2005) found that a clone of the eastern cottonwood, Populus deltoides Bartram, was attacked by ambrosia beetles significantly more frequently when the trees received liquid fertilizer than when they received control treatments. The factors governing KSHB infestation have yet to be fully understood, but the observed correlation with wetness in the Tijuana River Valley suggests that those searching for KSHB infestations in other parts of southern California should focus especially on Salicaceae-dominated forests near nutrient-enriched water.
Third, KSHB impacts appear to be a novel form of disturbance. Riparian habitats in California are adapted to many kinds of disturbances, including floods, fires, avalanches, debris flows, ice scour, windstorms and droughts (Pettit and Naiman 2007). Such disturbances may kill many plants but, importantly, they usually leave large open expanses of sandy sediments and some living trees. The riparian vegetation reestablishes quickly after such events because new seedlings can establish abundantly in the open, sandy areas and the living trees can grow back from stump sprouts (Rood et al. 2007, Wintle and Kirkpatrick 2007, Boland 2014b). The KSHB, however, has disturbed the forest in a unique way; the damaged forests are littered with woody debris that has fallen from the canopy so there are few patches of open sediment for seedlings, and there appear to be few living tree stumps from which sprouts can grow. Opportunities for revegetation via seedlings and resprouts therefore appear to be greatly limited. Whether the riparian vegetation can respond to this unique disturbance caused by the KSHB remains to be seen. Because the KSHB was not observed to attack young willow and mule fat plants, removal of woody debris in sites suitable for plant recruitment might be a good strategy for promoting the recovery of damaged riparian habitats.

Fourth, the KSHB attack appears to be having ecosystem-wide impacts. The KSHB showed a preference for the willows *S. lasiolepis* and *S. gooddingii*, the dominant trees in the Tijuana River Valley (Boland 2014a). Reduction or loss of these foundation species will drastically alter the canopy architecture, understory microclimate, and ecosystem processes such as productivity and water balance within the forest units (Ellison et al. 2005). Three species that are considered to be among the worst invasive species in California (Cal-IPC 2006), *R. communis*, *T. ramosissima* and *A. donax*, are likely to thrive in the damaged forests. All three species were present but relatively inconspicuous in the forest prior to the beetle infestation, and all are likely to grow tall and thrive in the absence of a willow canopy (pers. obs.). A new riparian community composed of invasive shrubs and herbaceous weeds is unlikely to provide the same habitat quality and food-chain support for animal species in the valley, including the endangered *Vireo bellii pusillus*. It is therefore possible that the KSHB is affecting not only its preferred host species but also the structure and function of the entire ecosystem. Managers will need to monitor many aspects of the damaged forests and pay particular attention to the spread of non-native plant species in these new forest habitats.

**ACKNOWLEDGEMENTS**

I thank Deborah Woodward for assistance in the field, helpful discussions, and comments on drafts of this manuscript, Monica Almeida for producing the maps, and Jeff Crooks, Mike Picker, Dezene Huber, Leland Humble, Richard Stouthamer and an anonymous reviewer for their valuable comments on an early draft. I also thank the U.S. Fish and Wildlife Service, U.S. Navy, California State Parks, and the San Diego County Parks and Recreation Department for preserving the Tijuana River Valley riparian habitats and making them available for study.

**REFERENCES**
Biedermann P.H.W., K.D. Klepzig and M. Taborsky. 2009. Fungus cultivation by ambrosia beetles: behavior and laboratory breeding success in three Xyleborine species. *Environmental Entomology* 38: 1096-1105.

Boland, J.M. 2014a. Factors determining the establishment of plant zonation in a southern Californian riparian woodland. *Madroño* 61: 48–63.

Boland, J.M. 2014b. Secondary dispersal of willow seeds: sailing on water into safe sites. *Madrono* 61: 388-398.

Brumby, S.P., N.R. Harvey, J.J. Bloch, J.P. Theiler, S.J. Perkins, A.C. Young, and J.J. Szymanski. 2001. Evolving forest fire burn severity classification algorithms for multispectral imagery. Pages 236-245 in: S.S. Shen and M.R. Descour, editors. *Algorithms for multispectral, hyperspectral, and ultraspectral imagery VII*. Proceedings of a symposium. SPIE Volume 4381. International Society for Optics and Photonics, 16-19 Apr 2001, Orlando, Florida, USA.

California Invasive Plant Council (Cal-IPC). 2006. *California invasive plant inventory*. Cal-IPC Publication 2006-02. California Invasive Plant Council, Berkeley, CA. 39p.

Constantz, J. and F. Murphy. 1990. Monitoring moisture storage in trees using time domain reflectometry. *Journal of Hydrology* 119: 31 - 42.

Coyle, D.R., D.C. Booth and M.S. Wallace. 2005. Ambrosia beetle (Coleoptera: Scolytidae) species, flight, and attack on living eastern cottonwood trees. Journal of Economic Entomology 98: 2049–2057.

Crooks, J.A. 2005. Lag times and exotic species: the ecology and management of biological invasions in slow-motion. *Ecoscience* 12: 316-329.

Ellison A.M., M.S. Bank, B.D. Clinton, E.A. Colburn, K. Elliott, C.R. Ford, D.R. Foster, B.D. Kloeppel, J.D. Knoepf, G.M. Lovett, J. Mohan, D.A. Orwig, N.L. Rodenhouse, W.V. Sobeck, K.A. Stinson, J.K. Stone, C.M. Swan, J. Thompson, B. Von Holle, and J.R. Webster. 2005. Loss of foundation species: Consequences for the structure and dynamics of forested ecosystems. *Frontiers in Ecology and the Environment* 9: 479-486.

Eskalen, A. 2016. Shot Hole Borers/Fusarium Dieback. *Available at: http://eskalenlab.ucr.edu/avocado.html* and *http://eskalenlab.ucr.edu/distribution.html* (accessed 30 March 2016).

Eskalen, A., R. Stouthamer, S.C. Lynch, M. Twizeyimana, A. Gonzalez and T. Thibault. 2013. Host range of *Fusarium* dieback and its ambrosia beetle (Coleoptera: Scolytinae) vector in southern California. *Plant Disease* 97: 938-951.

Freeman S., M. Sharon, M. Maymon, Z. Mendel, A. Protasov, T. Aoki, A. Eskalen and K. O’Donnell. 2013. *Fusarium euwallacea* sp. nov – a symbiotic fungus of *Euwallacea* sp., an invasive ambrosia beetle in Israel and California. *Mycologia* 105: 1595-1606.
Gandhi K.J.K., and D.A. Herms. 2010. Direct and indirect effects of alien insect herbivores on ecological processes and interactions in forests of eastern North America. *Biological Invasions* 12: 389–405.

Graham, M. 2016. Beetles invasion rocks biologists’ world. San Diego Reader January 18, 2016. Available at: [http://www.sandiegoreader.com/news/2016/jan/18/stringers-beetles-invasion-rocks-biologists-world/](http://www.sandiegoreader.com/news/2016/jan/18/stringers-beetles-invasion-rocks-biologists-world/) (accessed 30 March 2016).

Hulcr, J. 2012. What's wrong with ambrosia beetles these days? What we know, what we think we know, what we don't know. Presentation given at the Invasive Ambrosia Beetle Conference in Riverside, CA, August 12-14, 2012. Available at: [http://www.avocadosource.com/journals/iabc_2012/s1_01_hulcr.pdf](http://www.avocadosource.com/journals/iabc_2012/s1_01_hulcr.pdf) and [https://www.youtube.com/watch?v=XMuUMFlwtFg](https://www.youtube.com/watch?v=XMuUMFlwtFg). (accessed 30 March 2016).

Jones, M.E., and T.D. Paine. 2015. Effect of chipping and solarization on emergence and boring activity of a recently introduced ambrosia beetle (*Euwallacea* sp., Coleoptera: Curculionidae: Scolytinae) in Southern California. *Journal of Economic Entomology* 108: 1852-1859.

Kendrick, B. 2000. *The Fifth Kingdom*. Focus Publishing, Newburyport, MA.

Pettit, N.E. and R.J. Naiman. 2007. Fire in the Riparian Zone: Characteristics and Ecological Consequences. *Ecosystems* 10: 673-687.

Pimentel, D., R. Zuniga and D. Morrison. 2005. Update on the environmental and economic costs associated with alien-invasive species in the United States. *Ecological Economics* 52: 273-288.

Porter, R. 2007. *Strategies for Effective State Early Detection/Rapid Response Programs for Plant Pests and Pathogens*. Environmental Law Institute, Washington, DC. 97 pages.

Rabenold, K.N., P.T. Fauth, B.W. Goodner, J.A. Sadowski and P.G. Parker. 1998. Response of avian communities to disturbance by an exotic insect in spruce-fir forests of the Southern Appalachians. *Conservation Biology* 12: 177-189.

Rood, S., L.A. Goater, J.M. Mahoney, C.M. Pearce, and D.G. Smith. 2007. Floods, fire, and ice: disturbance ecology of riparian cottonwoods. *Canadian Journal of Botany* 85: 1019-1032.

Rudinsky, J.A. 1962. Ecology of Scolytidae. *Annual Review Entomology* 7: 327-348.

Smith, J.E. 2016. Beetle leaving wake of destruction. San Diego Union-Tribune January 2, 2016. Available at: [http://www.sandiegouniontribune.com/news/2016/jan/02/tp-beetle-leaving-wake-of-destruction-shot-hole/](http://www.sandiegouniontribune.com/news/2016/jan/02/tp-beetle-leaving-wake-of-destruction-shot-hole/) (accessed 30 March 2016).
Spann, T. 2015. PSHB research update. From the Grove fall: 33-35. Available at: http://www.californiaavocadogrowers.com/sites/default/files/documents/11-PSHB-Research-Update-Fall-2015.pdf (accessed 30 March 2016).

U.S. Fish and Wildlife Service. 1994. Endangered and threatened wildlife and plants: Designation of critical habitat for the least Bell’s vireo. Final Rule. Federal Register 59: 4845-4867.

Umeda, C., A. Eskalen and T.D. Paine. 2016. Polyphagous Shot Hole Borer and Fusarium Dieback in California. Pages 757-767. In: T.D. Paine and F. Lieutier (eds.). Insects and diseases of Mediterranean forest systems. Springer International Publishing, Cham, Switzerland.

Wintle B.C. and J.B. Kirkpatrick. 2007. The response of riparian vegetation to flood-maintained habitat heterogeneity. Austral Ecology 32: 592-599.

Wood, S.L. 1982. The bark and ambrosia beetles of North and Central America (Coleoptera: Scolytidae), a taxonomic monograph. Great Basin Naturalist Memoirs 6: 1-1356.
**FIGURE and TABLE HEADINGS**

**Figure 1.** Map of the Tijuana River Valley. The location of the riparian forest and riparian scrub habitats within the Tijuana River Valley.

**Figure 2.** Photos of the KSHB in the Tijuana River Valley. (A) Two beetles at the entrance to a tunnel. (B) Holes in the bark of a sycamore. (C) Extrusion of sawdust plugs indicating active burrowing within an arroyo willow. (D) Gumming out of beetle holes in mule fat. (E) Beetle tunnels and associated fungus (black staining) in a cross section through a black willow trunk. (F) Beetle-infested trunks of arroyo willows snapped by the wind.

**Figure 3.** The KSHB in the Tijuana River Valley in 2015-16. KSHB infestation levels in the riparian survey units in the Tijuana River Valley. Numbers indicate the 29 survey units and survey points.

**Figure 4.** Infestation rates vs. wetness. KSHB infestation rates of survey units as a function of the distance from surface water. The linear regression line, equation and correlation coefficient are for the forest units only (n = 22).

**Figure 5.** Before and after photos. The forest in Unit 2 during May 2015 and February 2016 showing KSHB-induced damage to the dominant willow trees.

**Table 1.** Infestation and damage rates within the survey units. Description of the survey units (stand age, unit area, median willow girth, plant density and estimated total number of plants) and the rates of KSHB infestation and major plant damage (n = number of individuals examined). The total numbers of plants infested and damaged in each unit are extrapolated. In the forest units, data are for *S. lasiolepis* and *S. gooddingii*; in the scrub units, data are for *B. salicifolia*. ND = no data.

**Table 2.** Infestation rates in the common plant species. KSHB infestation rates in the common riparian plant species in the Tijuana River Valley.
1

Map of the Tijuana River Valley.

The location of the riparian forest and riparian scrub habitats within the Tijuana River Valley.
Figure 2 (on next page)

Photos of the KSHB in the Tijuana River Valley.

(A) Two beetles at the entrance to a tunnel. (B) Holes in the bark of a sycamore. (C) Extrusion of sawdust plugs indicating active burrowing within an arroyo willow. (D) Gumming out of beetle holes in mule fat. (E) Beetle tunnels and associated fungus (black staining) in a cross section through a black willow trunk. (F) Beetle-infested trunks of arroyo willows snapped by the wind.
The KSHB in the Tijuana River Valley in 2015-16.

KSHB infestation levels in the riparian survey units in the Tijuana River Valley. Numbers indicate the 29 survey units and survey points.
**Figure 4** (on next page)

Infestation rates vs. wetness.

KSHB infestation rates of survey units as a function of the distance from surface water. The linear regression line, equation and correlation coefficient are for the forest units only ($n = 22$).
$y = -0.0022x + 0.8628$

$r^2 = 0.639$
Figure 5 (on next page)

Before and after photos.

The forest in Unit 2 during May 2015 and February 2016 showing KSHB-induced damage to the dominant willow trees.
Table 1 (on next page)

Infestation and damage rates within the survey units.

Description of the survey units (stand age, unit area, median willow girth, plant density and estimated total number of plants) and the rates of KSHB infestation and major plant damage (n = number of individuals examined). The total numbers of plants infested and damaged in each unit are extrapolated. In the forest units, data are for *S. lasiolepis* and *S. gooddingii*; in the scrub units, data are for *B. salicifolia*. ND = no data.
| UNIT            | AGE | AREA  | Girth | Density | Infested | Damaged |
|-----------------|-----|-------|-------|---------|----------|---------|
|                 | #   | yr    | ha    | cm      | n        | RATE    | TOTAL  | n    | RATE | TOTAL |
| A. Riparian forests |     |       |       |         |          |         |        |      |      |       |
| 1               | 35  | 14.64 | 130.8 | 2       | 7,319    | 39      | 74%    | 5442 | 50   | 12%   | 878   |
| 2               | 5   | 25.9  | 36    | 16,026  | 143      | 94%    | 15,130 | 67   | 51%  | 8,133 |
| 3               | 15  | 3.02  | 7     | 5,517   | 79       | 100%   | 5,517  | 50   | 98%  | 5,407 |
| 4               | 35  | 5.13  | 1     | 1,282   | 32       | 91%    | 1,162  | 38   | 82%  | 1,046 |
| 5               | 35  | 18.08 | ND    | 3       | 13,561   | 26      | 96%    | 13,040 | 43 | 84%  | 11,354 |
| 6               | 22  | 12.26 | 83.8  | 3       | 9,194    | 37      | 95%    | 8,697 | 44   | 86%  | 7,940 |
| 7               | 10  | 0.80  | 22    | 4,407   | 81       | 100%   | 4,407  | 65   | 94%  | 4,136 |
| 8               | 5   | 2.11  | 72    | 37,953  | 163      | 87%    | 33,063 | 90   | 23%  | 8,856 |
| 9               | 22  | 10.21 | ND    | 4       | 10,211   | 40      | 100%   | 10,211 | 51 | 100% | 10,211 |
| 10              | 22  | 23.04 | 66.0  | 3       | 17,280   | 42      | 98%    | 16,868 | 68 | 78%  | 13,468 |
| 11              | 35  | 4.68  | 81.3  | 8       | 9,365    | 31      | 100%   | 9,365 | 86   | 78%  | 7,296 |
| 12              | 22  | 3.14  | 73.7  | 12      | 9,421    | 42      | 100%   | 9,421 | 54   | 89%  | 8,375 |
| 13              | 22  | 15.01 | 54.6  | 10      | 37,526   | 70      | 97%    | 36,454 | 52 | 58%  | 21,650 |
| 14              | 22  | 17.84 | 34.3  | 19      | 84,717   | 141     | 75%    | 63,688 | 127 | 17%  | 14,008 |
| 15              | 35  | 18.52 | 71.1  | 4       | 16,204   | 50      | 8%     | 1,296 | 93   | 12%  | 1,917 |
| 16              | 35  | 20.75 | 79.5  | 5       | 25,936   | 35      | 6%     | 1,441 | 66   | 14%  | 3,537 |
| 17              | 35  | 21.42 | 83.8  | 3       | 16,069   | 43      | 0%     | 0     | 106  | 12%  | 1,971 |
| 18              | 35  | 7.06  | 67.3  | 4       | 7,062    | 44      | 2%     | 161   | 64   | 28%  | 1,986 |
| 19              | 35  | 6.82  | 79.8  | 5       | 8,524    | 33      | 61%    | 5,166 | 67   | 16%  | 1,399 |
| 20              | 35  | 12.86 | 82.6  | 3       | 9,643    | 40      | 10%    | 964   | 58   | 5%   | 499   |
| 21              | 35  | 9.56  | 77.0  | 3       | 7,172    | 35      | 6%     | 410   | 73   | 7%   | 491   |
| 22              | 10  | 12.83 | 43.9  | 15      | 48,124   | 60      | 95%    | 45,718 | 82 | 13%  | 6,456 |
| Total           |     |       |       |         | 241.6    |         | 402,513 |     |      | 287,620 | 141,011 |
| B. Riparian scrub |     |       |       |         |          |         |        |      |      |       |
| 23              | 35  | 76.6  | ND    | 1       | 19,150   | 40      | 33%    | 6,319 | 30   | 0%   | 0     |
| 24              | 35  | 38.0  | ND    | 2       | 19,016   | 55      | 25%    | 4,754 | 57   | 5%   | 951   |
| 25              | 15  | 39.6  | ND    | 1       | 9,909    | 34      | 24%    | 2,378 | 42   | 0%   | 0     |
| 26              | 25  | 31.7  | ND    | 14      | 110,961  | 91      | 0%     | 0     | 81   | 0%   | 0     |
| 27              | 35  | 106.3 | ND    | 6       | 159,467  | 55      | 2%     | 3,189 | 50   | 2%   | 3,189 |
|   | 28 | 35 | 68.7 | ND | 6 | 102,982 | 50 | 0% | 0 | 56 | 0% | 0 |
|---|----|----|------|----|---|----------|----|----|---|----|----|---|
| 29 | 35 | 57.8 | ND | 3 | 43,378 | 50 | 0% | 0 | 52 | 0% | 0 |
| **Total** | **833.9** | | | | **464,863** | | | **16,641** | | **4,140** |
Table 2 (on next page)

Infestation rates in the common plant species.

KSHB infestation rates in the common riparian plant species in the Tijuana River Valley.
| Species                          | Common name               | Family          | # examined | # infested | % infested |
|---------------------------------|---------------------------|-----------------|------------|------------|------------|
| **NATIVE SPECIES**              |                           |                 |            |            |            |
| *Salix lasiolepis* Benth.       | Arroyo willow             | Salicaceae      | 539        | 442        | 82%        |
| *Salix gooddingii* C.R. Ball    | Black willow              | Salicaceae      | 670        | 499        | 74%        |
| *Salix laevigata* Bebb          | Red willow                | Salicaceae      | 14         | 9          | 64%        |
| *Populus fremontii* S. Watson   | Western cottonwood        | Salicaceae      | 53         | 28         | 53%        |
| *Platanus racemosa* Nutt.       | California sycamore       | Platanaceae     | 28         | 6          | 21%        |
| *Baccharis salicifolia* (Ruiz & Pav.) Pers. | Mule fat       | Asteraceae      | 486        | 76         | 16%        |
| *Baccharis pilularis* DC.       | Coyote brush              | Asteraceae      | 92         | 5          | 5%         |
| *Ambrosia monogyra* *           | Singlewhorl burrobrush    | Asteraceae      | 33         | 1          | 3%         |
| *Salix exigua* Nutt             | Narrow-leaf willow        | Salicaceae      | 88         | 1          | 1%         |
| *Malosma laurina* (Nutt.) Nutt. Ex Abrams | Laurel sumac  | Anacardiaceae   | 7          | 0          | 0%         |
| *Peritoma arborea* (Nutt.) H.H. Iltis | Bladderpod     | Cleomaceae      | 9          | 0          | 0%         |
| *Sambucus nigra* L.             | Blue elderberry           | Adoxaceae       | 31         | 0          | 0%         |
| *Scirpus spp.*                  | Tule                      | Cyperaceae      | 5          | 0          | 0%         |
| *Artemisia dracunculus* L.      | Tarragon                  | Asteraceae      | 19         | 0          | 0%         |
| *Chloracantha spinosa* (Benth.) G.L. Nesom | Spiny aster     | Asteraceae      | 2          | 0          | 0%         |
| *Isocoma vernonioides* (Nutt.) G.L. Nesom | Coastal goldenbush | Asteraceae      | 14         | 0          | 0%         |
| **NON-NATIVE SPECIES**          |                           |                 |            |            |            |
| *Schinus terebinthifolius* Raddi | Brazilian pepper          | Anacardiaceae   | 15         | 7          | 47%        |
| *Nicotiana glauca* Graham       | Tree tobacco              | Solanaceae      | 40         | 8          | 20%        |
| *Ricinus communis* L.           | Castor bean               | Euphorbiaceae   | 123        | 16         | 13%        |
| *Eucalyptus spp.*               | Gum tree                  | Myrtaceae       | 21         | 2          | 10%        |
| *Tamarix ramosissima* Lede.     | Salt cedar                | Tamaricaceae    | 32         | 1          | 3%         |
| *Tropaeolum majus* L.           | Garden nasturtium         | Tropaeolaceae   | 5          | 0          | 0%         |
| *Araujia sericifera* Brot.      | Cruel vine                | Apocynaceae     | 5          | 0          | 0%         |
| *Phytolacca icosandra* L.       | Tropical pokeweed         | Phytolaccaceae  | 5          | 0          | 0%         |
| *Schinus molle* L.              | Peruvian pepper            | Anacardiaceae   | 7          | 0          | 0%         |
| *Myoporum laetum* G. Forst.     | Myoporum                  | Scrophulariaceae | 16     | 0          | 0%         |
| *Acacia cyclops* G. Don         | Cyclops wattle             | Fabaceae        | 2          | 0          | 0%         |
| *Arundo donax* L.               | Giant reed                | Poaceae         | 42         | 0          | 0%         |
| **IMMATURE PLANTS (< 1 year old)** |                           |                 |            |            |            |
| Species                  | Common Name    | Family       | Value | Rating | Percentage |
|-------------------------|----------------|--------------|-------|--------|-------------|
| *Salix gooddingii*      | Black willow  | Salicaceae   | 72    | 0      | 0%          |
| *Baccharis salicifolia* | Mule fat       | Asteraceae   | 30    | 0      | 0%          |
| *Ricinus communis*      | Castor bean    | Euphorbiaceae| 32    | 0      | 0%          |

1 * = (Torr. & A. Gray) Strother & B.G. Baldwin