Hormones and Neuropeptide Receptor Heteromers in the Ventral Tegmental Area. Targets for the Treatment of Loss of Control of Food Intake and Substance Use Disorders

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Opinion Statement
Hormones and neuropeptides represent biological correlates of internal homeostatic signals detected and integrated in the hypothalamus, which establishes a robust functional connection with the ventral tegmental area (VTA). The hypothalamus-VTA connection determines the ability of these signals to influence central dopaminergic neurotransmission and, therefore, their ability to increase responsiveness to their reward-associated stimuli and to establish appropriate associative learning. The hypothalamus also provides the main source of the multiple neuropeptides that are released in the VTA. With volume transmission of neuropeptides and hormones, extrasynaptic receptors within the VTA provide a fine-tune mechanism, which depends on the ability of molecularly different G protein-coupled receptors (GPCRs) to form heteromers. GPCR heteromer is defined as a macromolecular complex composed of at least two different receptor units (protomers) with biochemical properties that are demonstrably different from those of its individual components. GPCR heteromers can provide unique allosteric properties to specific ligands, which provides new avenues for drug development. We have identified specific GPCR
heteromers in the VTA that integrate orexin and CRF neurotransmission and opioid and galanin neurotransmission, which play a very significant role in the modulation of dopaminergic neuronal activity and which can constitute targets for the treatment of loss of control of food intake and substance use disorders.

Introduction: hormones and neuropeptides as interoceptive discriminative stimuli

“Drives” and “motivational states” can be operationally approached by specific interoceptive discriminative stimuli, correlating internal signals conveyed by identifiable biological signaling molecules, largely hormones and neuropeptides. For instance, “hunger” and “satiety” can be operationally approached by specific correlative internal orexigenic and anorectic signaling molecules, which respectively facilitate and inhibit food-oriented behavior, eliciting approach to food-related stimuli and consummatory eating behavior or withdrawal from those stimuli. Alterations in the capacity of these biological molecules to signal through their specific receptors lead to pathological implications, such as obesity and anorexia.

Definitions of hormone and neuropeptide

A hormone is defined as a signaling molecule produced by an endocrine cell that is transported by the circulatory system to target distant organs. A neuropeptide is defined as a small proteinaceous substance produced by neurons, released in a regulated fashion and acting on neural substrates, e.g., neurons, glial cells, or non-neuronal target cells, e.g., a gland or muscle [1].

The hypothalamus represents a key center of integration of homeostatic, threat, and reproductive signals. First, it acts like an internal sensory organ that detects internal signals conveyed by hormones and neuropeptides. Second, the hypothalamus “valuates” among the constantly detected internal signals, playing a decision-maker role on the elicitation of the highest priority homeostatic- or reproductive-oriented response.

Several hormones and neuropeptides, such as ghrelin, leptin, insulin, melanocyte-stimulating hormones (α-MSH, β-MSH and γ-MSH), neuropeptide Y, agouti-related protein (AgRP), neurotensin, melanin-concentrating hormone (MCH), orexin, and galanin, provide orexigenic or anorectic signals that are integrated in the hypothalamus to modulate food-oriented behavior. This multiplicity of signals allows an exhaustive control of the internal metabolic environment and more specifically of energy homeostasis [2, 3•, 4•].

Some of the biological signals, particularly ghrelin, leptin, and insulin, are released to the blood circulation and reach the hypothalamic arcuate nucleus (ARC) by way of its proximity to the median eminence [5–7] (Fig. 1). Median eminence is a circumventricular organ with fenestrated capillaries and modified glial cells called tanyocytes that play an important part in the neuroendocrine system by providing the connection of hypothalamic neurons to the pituitary, through the pituitary portal circulation [8, 9]. Nerve terminals from the paraventricular nucleus of the hypothalamus (PV) release corticotropin-releasing factor (CRF) within the median eminence, as part of the initial stress response of the hypothalmo-pituitary-adrenocortical axis. CRF then travels to the anterior and intermediate pituitary lobes and promotes the synthesis and systemic release of proopiomelanocortin (POMC)-derived peptides, which include the melanocortins adrenocorticotropic hormone (ACTH) and α-, β-, and γ-MSH, as well as the endogenous opioid β-endorphin [3•, 10]. The median eminence acts therefore as a two-way gate for hormones of central and peripheral origin.

Ghrelin is an orexigenic hormone mostly produced by the stomach oxyntic cells, which provide plasma levels that fluctuate diurnally with a peak in the day and through at night [11, 12]. Insulin and glucose independently increases and decreases, respectively, the secretion and circulating concentration of ghrelin [13]. Notably, oxyntic cells qualify as food-entrained oscillators, and ghrelin plasma levels increase during anticipated mealtimes and decrease after meals [11].

Leptin is produced by adipocytes in proportion to triglyceride stores, serving as a signal of repletion of long-term energy stores [14]. The central signals provided by peripheral ghrelin and leptin largely contribute to energy homeostasis, which is achieved when the amount of energy consumed (from food intake) equals energy expended (from basal metabolic rate and physical activity). These signals are initially processed in the hypothalamus, within the ARC, by the two phenotypically different POMC- and AgRP-expressing neurons, which project to the PVN and lateral hypothalamus (LH) (Fig. 1). POMC-derived peptides are therefore not only pituitary-derived hormones, but also neuropeptides synthesized within the hypothalamus. Activation of
the POMC-expressing neuron inhibits food intake and food-oriented behaviors, while activation of the AgRP-expressing neuron promotes food intake [4, 15–18].

The orexigenic AgRP-expressing neuron contains high levels of ghrelin receptors. Therefore, ghrelin directly activates AgRP-expressing neurons, which by inhibitory collaterals inhibit the activity of POMC-expressing neurons. On the other hand, both AgRP- and POMC-expressing neurons contain leptin receptors, which stimulate and inhibit the activity of POMC- and AgRP-expressing neurons, respectively [19, 20]. Another level of integration of orexigenic and anorectic signals processed by AgRP- and POMC-expressing neurons takes place at the very melanocortin receptor level. Thus, AgRP is an endogenous orthosteric antagonist of the targets of hypothalamic melanocortins, MC3 and MC4 receptors [3•, 10].

Pathological alterations in the capacity of ghrelin, leptin, and melanocortins to signal through their specific receptors lead to their incapacity to act as appropriate internal homeostatic signals, leading to pathological loss of control of food intake, both in the experimental animal and in human beings. Leptin deficiency and mutations of leptin receptors and MC4 receptors are common monogenic causes of obesity [3•]. Prader-Willi syndrome (PWS), a condition associated with high ghrelin serum levels, is the most common cause of syndromic obesity [21]. PWS patients are incapable of making appropriate food-related decisions and their hyperphagia can be life-threatening. Thus, the voracious feeding habits seen in PWS include intense food foraging, stealing of food and even consumption of inedible foods, which may lead to choking and gastric rupture [22, 23]. The loss of control of food-related behaviors observed in hyperghrelinemic PWS is a strong and persistent food-oriented psychomotor activation equivalent to that induced by food deprivation, but without satiety. It is therefore equivalent to the psychostimulant effects of addictive drugs, which depend largely on activation of the central dopaminergic system [24]. The question is then how ghrelin and other food-related internal signals can influence the central dopaminergic system.

**The hypothalamus-VTA connection**

Ascending dopamine systems originate in the substantia nigra pars compacta (SNC) and the ventral tegmental area (VTA), and their differential functional...
roles depend on their innervation of functionally different striatal compartments as well as their differential source of inputs. The hypothalamus projects mostly to the VTA [25] and in fact constitutes one of its three main sources of inputs, together with the ventral striatopallidal complex and the dorsal raphe [25, 26]. LH is the main contributor of hypothalamic inputs to the VTA, followed by the PV and with a lesser contribution from other nuclei including ARC [25]. Pioneering studies on intracranial electrical stimulation in rodents showed that electrical stimulation of the LH produces voracious feeding behavior [27] as well as reinforcement of lever-pressing behavior to gain additional stimulation, intracranial self-stimulation [28].

It has been classically known that electrical stimulation of the LH facilitates a variety of species-typical, biologically primitive behavior patterns, including eating, drinking and gnawing, in sated animals [28, 30]. Such stimulation does not elicit specific motor responses, but produces psychomotor activation, which implies an increased responsiveness to a variety of environmental stimuli, which implies a facilitation of different behavioral responses, such as feeding, drinking, gnawing of wood, or a predatory attack [29–32]. These differences are not a result of differences in the stimulation region [33] and the dominant response of a given animal changes as a function of the discriminative and rewarding stimuli present [34]. An important amount of data now indicates that most effective intracranial electrical and optogenetic stimulation-induced feeding and reward (intracranial self-stimulation) largely result from the direct activation of VTA dopaminergic cells as well as the LH and the LH-VTA connection, the descending medial forebrain fibers of passage that directly or indirectly activate VTA dopaminergic cells; for review, see [35].

It is then through the LH-VTA connection that hypothalamic activity promotes an increased dopaminergic neurotransmission and, consequently, the dopamine functions: Dopamine release increases responsiveness to rewarding and reward-associated, discriminative stimuli, with orienting and approaching responses to those stimuli [36]; concomitantly, dopamine is directly involved in reinforcement, in the learning (“stamping-in”) of stimulus-reward and reward-response associations that follows the receipt of reward [36]. The reinforcement of stimulus-reward associations establishes new signals that guide and orient to rewards (discriminative stimuli) or which become rewards themselves (conditioned rewarding stimulus). The stamping-in of reward-response associations promotes the learning of the optimal sequential response, the action skill that leads to the reward [36].

The LH-VTA connection includes glutamatergic and GABAergic projecting neurons that potentially activate and disinhibit (by inhibiting GABAergic interneurons) dopaminergic neuronal activity, respectively [37–41] (Fig. 1). Although it was initially believed that the excitatory input was fundamental to the LH-mediation of dopaminergic cell activation in the VTA [37, 38], recent optogenetic studies have demonstrated a predominant role of the GABAergic projecting neurons in feeding-oriented behavior and reward [39–41]. However, the picture is more complex than just the glutamate-GABA dichotomy, with the existence of different subpopulations of cells expressing different neuropeptides [35•, 42]. For instance, neurotensin and galanin are co-expressed in a population of GABAergic neurons that also express leptin receptors [43, 44] and these neurons exert a local inhibitory control of glutamatergic neurons that co-express orexin [43, 44]. The two orexin/hypocretin peptides, orexin-A and orexin-B, are only produced in the brain by neurons of the lateral hypothalamic nucleus (HL).
and to a lesser extent by neurons of the adjacent dorsomedial hypothalamic nucleus [45]. These cells give origin to the ascending orexin arousal system innervating most brain areas, including the VTA [45]. Still another population expresses MCH, which seem to be predominantly GABAergic [46], but these neurons do not connect directly with the VTA [42].

In addition to the LH, VTA receives inputs from other hypothalamic nuclei, such as PV and ARC [25] (Fig. 1). PV provides a main source of CRF to the VTA [47], and ARC provides the source of POMC-derived peptides, melanocortins and β-endorphin [48, 49]. The factors that determine the differential synthesis of POMC-derived peptides are tissue-specific, and the precursor is cleaved in a differential manner. Thus, the processing of POMC in the anterior pituitary is less extensive than in the hypothalamus, where ACTH is all cleaved to produce α-MSH and β-lipotropin is all cleaved to yield β-endorphin [50]. Surprisingly, although the cleavage of POMC to produce α-MSH and β-endorphin depends on the activity of the same enzyme (protein convertase PC2; 50), a recent study showed a cannabinoid CB1 receptor-dependent differential synthesis of both neuropeptides by the hypothalamic POMC-expressing neurons [51]. Under normal conditions, activation of POMC-expressing neurons promotes melanocortin synthesis and release, inhibiting food-oriented behavior (see above). However, activation of CB1 receptors on POMC-expressing neurons promotes β-endorphin synthesis and the opposite behavioral effect, facilitation of feeding [51]. Two additional opioids, the tetra-peptides endomorphin-1 and endomorphin-2, which are the most potent and selective endogenous agonists for the μ-opioid receptor [52], are also synthesized in hypothalamic neurons which project to the VTA and are localized in an area close to the periventricular nucleus and the ARC [53]. Finally, although still a matter of debate [7], there are data suggesting that ghrelin qualifies as a neuropeptide synthesized in the ARC [54], which could provide an additional ligand source for ghrelin receptors localized in the VTA (see below).

Hormone and neuropeptide transmission within the VTA

VTA hormone and neuropeptide GPCRs

NTS1 and NTS2 neotensin receptors [55], Gal1 and Gal2 receptors [56], OX1 and OX2 orexin receptors [57], CRF1 receptors and lower expression of the CRF2 subtype; [58]; melanocortin MC3 receptors and lower expression of the MC4 subtype; [49]; μ- and κ-opioid receptors [59] and ghrelin GHS1a receptor [60].

Neuropeptide neurotransmission plays then a key role in the hypothalamus-VTA connection. The VTA shows a high density of receptors for neuropeptides localized in the soma and/or dendrites of the dopaminergic cells or in the terminals of their excitatory or inhibitory afferents. Neuropeptide receptors are mostly G protein-coupled receptors (GPCRs), belonging to class A or rhodopsin family (receptors for neotensin, galanin, orexins, melanocortins, endogenous opioids, and ghrelin) and class B or secretin family (receptors for CRF). Neuropeptides are up to 50 amino acid-long polypeptide gene products, synthesized as ribosomal pre-hormones that are cleaved and often post-translationally modified [61••].
Neuropeptides are therefore produced in the soma and to lesser extent dendrites and packaged in large dense core vesicles that are transported through axons and dendrites. Cleavage and additional enzymatic modifications such as a common C-terminal α-amidation take place in the large dense core vesicles, yielding the final bioactive peptide [61••]. Neuropeptides are released extrasynaptically and preferentially with high-frequency neuronal firing. Classical neurotransmitters, on the other hand, are usually stored in clear synaptic vesicles and released at the synapse upon low-frequency activity. An additional fundamental difference with classical neurotransmitters is that neuropeptide clearance is slower and does not depend on efficient reuptake and intracellular metabolization mechanisms, but on extracellular breaking down by extracellular peptidases. Neuropeptide neurotransmission is therefore designed to reach larger distances than classical neurotransmitters, to exert a modulatory role by activating extrasynaptic receptors localized in different neuronal or non-neuronal elements within a brain area. In general, neuropeptides demonstrate higher affinity and selectivity for their respective receptors than classical neurotransmitters, which corresponds to their ability to act as extrasynaptic modulatory signals. Synaptic transmission, on the other hand, depends on the release of high concentrations of the classical neurotransmitter [61••, 62].

Neuropeptide transmission within the VTA provides a local hormonal-like effect of signaling molecules that are released in a volume-transmission mode [62] from hypothalamic-VTA nerve terminals and invade the extracellular space that surrounds the soma and dendrites of mesencephalic dopaminergic cells. The same neuropeptide-mediated modulatory signal is therefore broadcasted by the bulk of dopaminergic cells to their widespread terminal fields. However, the VTA also contains a relatively high density of hormone receptors, for ghrelin (also GPCRs) and for leptin and insulin (both belonging to the large tyrosine kinase receptor family) [63]. The same as for leptin and ghrelin, insulin can enter the brain tissue by passive diffusion through the median eminence and activate receptors that are localized in the ARC [64], but the three hormones can cross the blood-brain barrier and reach other hypothalamic areas and the VTA by means of specific saturable transport systems [5, 65–67].

Numerous experiments have addressed the study of the effect of each neuropeptide and hormone in isolation on the activity of their specific receptors on VTA dopaminergic cell function. In this way, for instance, it has been established that acting on receptors localized in the dopaminergic cells, ghrelin, neuropeptin, or melanocortin increase, while leptin or insulin inhibit their activity [44, 49, 60, 68, 69]. Orexin and opioids also increase dopaminergic cell activity, but most probably by acting on receptors localized in glutamatergic and GABAergic nerve terminals, respectively, increasing excitatory and decreasing inhibitory neurotransmission [70–75]. Less clear has been the role of CRF, which seems to activate dopaminergic cells under specific conditions, such as previous psychostimulant exposure [76], and even more mysterious has been the role of galanin in the VTA [77]. This approach, the investigation of the role of a single neuropeptide or hormone, represents an artificial simplification since the presence of multiple modulators diffusely released in the extracellular space of the VTA is probably the norm, which calls for the analysis of neurotransmitter-neurotransmitter and receptor-receptor interactions. With volume transmission of neuropeptides and hormones, extrasynaptic receptors within the
VTA provide an additional molecular fine-tune mechanism, which depends on the ability of GPCRs to form oligomeric complexes, GPCR heteromers.

Hormone and neuropeptide receptor heteromers in the VTA

Since their discovery, receptors have mostly been considered as single functional units. However, in recent years, a fast-growing list of GPCR-forming receptor oligomers has emerged [78–80]. Receptor heteromer is defined as a macromolecular complex composed of at least two (functional) receptor units (protomers) with biochemical properties that are demonstrably different from those of its individual components [78]. A first important concept that arises from the new field of GPCR oligomerization is that the pentameric structure constituted by one GPCR homodimer and one heterotrimeric G protein provides a main functional unit, and oligomeric entities can be viewed as multiples of dimers [79]. This seems to apply particularly to heteromers that include GPCR homodimers with preferential coupling to Gs/olf (Gs for short) proteins and another molecularly different homodimer with preferential coupling to Gi/o (Gi for short) proteins. Such a "GPCR heterotetramer" would sustain a functional pre-coupled macromolecular complex that includes two molecularly different GPCRs, their cognate G proteins, and adenylyl cyclase and would provide the frame for a canonical interaction at the adenylyl cyclase level, the ability of a Gi-coupled GPCR to counteract adenylyl cyclase activation induced by a Gs-coupled GPCR [81]. Recent studies using biophysical techniques and computerized modeling have provided experimental evidence for the existence of several GPCR heterotetramers that fulfill this scheme, such as the dopamine D1–D3 [82] and the adenosine A2A-dopamine D2 receptor heterotetramer [83] (Fig. 2). Using disrupting synthetic peptides with the amino acid sequence of different transmembrane domains of the receptors, we can now determine not only the interfaces involved in hetero- and homomerization in the heterotetramer [82, 83] but also the interfaces involved in the complex formation with adenylyl cyclase. One unexpected and very significant output of these studies is that the canonical Gs-Gi interaction at the adenylyl cyclase level is a specific property of a heterotetramer (in preparation).

GPCR heteromers can convey allosteric modulations between orthosteric ligands (ligands that bind to the same site as the endogenous neurotransmitter), altering their affinity or intrinsic efficacy. A ligand binding to one GPCR unit in the complex can lead to changes in the properties of a ligand binding to a different GPCR unit [79]. For instance, in the well-established A2A-D2 receptor heteromer, an orthosteric A2A receptor agonist decreases the affinity and intrinsic efficacy of dopamine or another orthosteric agonist for the D2 receptor [83, 84]. Interestingly, the negative modulation of the intrinsic activity is functionally selective for the G protein-independent D2 receptor-mediated MAPK activation [84] (Fig. 2). Although this functional selectivity depends on the existence of neuronal calcium-binding proteins that selectively bind to the A2A-D2 receptor heteromer. In their absence, knocking down the expression of these calcium-binding proteins, an A2A receptor agonist is also able to counteract the canonical interaction, in this case, the ability of a D2 receptor agonist to counteract adenylyl cyclase activation induced by an A2A receptor agonist [84]. A common property of receptor heteromers is that, not only agonists but also
antagonists, can act as allosteric modulators within a GPCR heteromer, a phenomenon named cross-antagonism [79]. Commonly, any orthosteric ligand of one of the protomers in the heteromer can lead to changes in the affinity or intrinsic efficacy of an orthosteric agonist of the molecularly different protomer. This is also the case for the A2A-D2 receptor heteromer [83].

Additional considerations about the functional and pharmacological properties, which make GPCR attractive targets for drug development, are ligand-independent allosteric modulations and probe dependence. One of the GPCR protomers can convey a ligand-independent allosteric modulation of ligands binding to the other molecularly different GPCR protomer and this can be ligand specific (probe dependence). Again, the A2A-D2 receptor heteromer provided the proof of concept, since a specific A2A receptor antagonist (SCH-442416) was found to significantly decrease its affinity for the A2A receptor when it heteromerizes with the D2 receptor [85].

We have demonstrated the existence of specific GPCR heteromers in the VTA that integrate orexin and CRF neurotransmission [86] and opioid and galanin neurotransmission [87], which play a very significant role in the modulation of dopaminergic neuronal activity. Furthermore, we are obtaining experimental evidence for an additional functionally significant GPCR heteromer that controls VTA dopaminergic cell function, a ghrelin-dopamine D1/5 receptor heteromer (in preparation). Our methodology includes, first, in vitro techniques in mammalian cells that are transfected with receptors fused to biosensors that can only interact when in close proximity. This allows finding synthetic peptides with amino acid sequences corresponding to the interfaces of the putative receptor heteromers. Specific disrupting peptides are then used as a tool to identify the biochemical properties of the GPCR heteromer (which are specifically disrupted with the peptides), such as an allosteric interaction between specific ligands. Next, the specific disrupting peptides are used with in situ and in vivo approaches to demonstrate the presence of the same GPCR heteromer within the VTA and its functional and pharmacological significance. The approaches include signaling in VTA slices and a modified infusion-microdialysis technique that allows a slow-rate infusion of combinations of neuropeptides and synthetic disruptive peptides and simultaneous measurement of VTA somatodendritic dopamine release [86]. Somatodendritic dopamine release by mesencephalic dopaminergic cells resembles that of the terminal regions, possessing a similar uptake mechanism and a finite releasable

**Fig. 2.** The adenosine A2A-dopamine D2 receptor heteromer. The heteromer has a heterotetrameric structure, constituted by homodimers of the Gs-coupled A2A receptor (A2AR) and the Gi-coupled D2 receptor (D2R). The canonical Gs-Gi antagonistic interaction at the adenylyl cyclase (AC) level and functionally selective allosteric modulation occur in the frame of the A2AR-D2R heteromer (see text). This heteromer constitutes a predominant population of striatal A2AR and D2R.
storage pool [88]. Furthermore, previous studies have conclusively shown that local dopamine release in the VTA is a correlate of dopaminergic cell firing [89]. This in vivo approach is therefore particularly suited to explore the role of local neuropeptide interactions within the VTA on dopaminergic cell activity and dependence on GPCR heteromerization.

Specifically, OX₁ and not OX₂ receptors form heteromers with CRF₁ receptors [86••] (Fig. 3). In signaling experiments in transfected cells and in the VTA, CRF₁-OX₁ heteromer mediated a strong negative crosstalk between orexin-A and CRF and a cross-antagonism, with the ability of OX₁ and CRF₁ receptor antagonists to counteract the effect of CRF₁ and OX₁ receptor agonists, respectively. In the CRF₁-OX₁ heteromer, CRF₁ couples to Gs and OX₁ to Gi, promoting activation and inhibition of adenylyl cyclase, respectively [86••], and therefore probably constituting an additional example of a GPCR heterotrimer [81]. Different to the A₂₅-D₂ receptor heteromer, the negative allosteric modulation of CRF on the intrinsic efficacy of orexin-A did not show functional selectivity. CRF counteracted both signaling events induced by orexin-A, the G protein-dependent ability of orexin to counteract CRF1 receptor-mediated adenylyl cyclase activation, and the G protein-independent ability to activate MAPK activation [86••] (Fig. 3). Orexin-A produced a significant increase in somatodendritic dopamine release in the VTA and CRF was not effective on its own, but significantly counteracted the effect of orexin-A. Cross-antagonism could also be demonstrated, and a CRF receptor antagonist also counteracted the effect of orexin-A [86••]. That these pharmacological interactions were dependent on CRF₁-OX₁ heteromerization was demonstrated by the ability of heteromer-specific peptides to disrupt the negative crosstalk in vivo [86••]. CRF-orexin-A interactions mediated by the CRF₁-OX₁ heteromers could be an extension of the integrative role of internal signals by the hypothalamus. The CRF₁-OX₁ heteromers integrate volume transmission signals driven by neuropeptides from the PV and LH, probably under conditions where the metabolic demands that facilitate food-oriented behavior need to be inhibited in favor of more evolutionary-significant behaviors, such as those linked to threat signals.

An additional pharmacological finding of the CRF₁-OX₁ heteromer was that it complexes with sigma σ₁ receptors and that σ₁ receptor ligands, including cocaine, counteract the allosteric interactions within the heteromer, both in transfected cells and in the VTA. With the infusion-microdialysis technique, it

**Fig. 3.** The corticotropin-releasing factor CRF₁-orexin OX₁ receptor heteromer. The heteromer can have a heterotetrameric structure, such as the A₂₅-D₂ receptor heteromer, constituted by homodimers of a Gs-coupled CRF₁ receptor (CRF₁R) and a Gᵢ-coupled OX₁ receptor (OX₁R). There is no evidence for functional selectivity of the allosteric interactions within the heteromer. CRF counteracts all signaling events induced by orexin-A, including its potential ability to inhibit CRF-induced AC activation (canonical Gₛ-Gᵢ interaction; see text).
could also be shown that under normal conditions, the CRF₁-OX₁ heteromer mediates a tonic inhibitory influence of orexin-A on CRF-induced signaling that can be released by cocaine exposure [86••]. The counteraction of the negative crosstalk between orexin-A and CRF in the VTA by α₁ receptor ligands provides a mechanism by which CRF can only induce glutamate-dependent somatodendritic dopamine release in animals previously exposed to cocaine [76]. Counteraction of the allosteric interactions between the endogenous neuropeptides in the CRF₁-OX₁ receptor heteromer can also explain the previously reported apparent CRF-independent ability of orexin-A to release dopamine in the VTA and to induce cocaine seeking [90]. The localization of CRF₁-OX₁ heteromers in the VTA still remains to be determined, although the previously reported glutamate dependence of the somatodendritic dopamine release in the VTA induced by orexin-A or stress strongly suggests their localization in the glutamatergic terminals of PVN-VTA neurons [76, 90]. CRF₁-OX₁ receptor heteromer can therefore constitute a target for the treatment of cocaine and other psychostimulant use disorders.

Galanin receptors have been said to provide candidates for the treatment of opioid use disorders [91]. This is based, first, on experimental evidence for the existence of antagonistic interactions between the galanin and opioid systems. Second, on genetic studies that find associations of galanin gene polymorphisms with susceptibility to opioid use disorder [92, 93]. The most significant association was observed for the single nucleotide polymorphism rs948854, localized in the promoter region [93]. The experimental results indicate that galanin receptor activation counteracts the psychomotor-activating and reinforcing effects of morphine [91, 94]. The behavioral effects of morphine were enhanced in galanin knock-out mice and were also counteracted by the systemic administration of a non-peptidergic, non-selective galanin receptor agonist [91]. Significantly, galanin knock-out mice showed a selective increase in morphine-induced MAPK activation in the VTA, which was also counteracted by the galanin receptor agonist [91].

It is well established that μ-opioid receptors localized in the striatum and in the ventral midbrain are involved in the reinforcing effects of opioids [94–97]. In the ventral midbrain, endogenous opioids exert a strong inhibitory control of the function of dopaminergic cells in the VTA, which depends on μ-opioid-mediated inhibition of a tonic GABAergic neurotransmission [70], largely mediated by afferents from the striatal patch compartment and from the rostromedial tegmental nucleus or tail of the VTA [72, 73, 75]. We recently demonstrated the existence of heteromers of μ-opioid and, specifically, Gal₁ receptors in the VTA that can underlie the pharmacological interactions of opioids and galanin systems [87••] (Fig. 4). These μ-opioid-Gal₁ receptor heteromers are therefore most probably localized in inhibitory inputs to the VTA and integrate volume transmission signals driven by the hypothalamic inputs that release β-endorphin and endomorphin-1 (ARC and periARC and periventricular areas) and galanin (LH). A hormonal pituitary origin of β-endorphin seems to be unlikely [98].

We first detected μ-opioid-Gal₁ receptor heteromerization in mammalian transfected cells and obtained a specific peptide that disrupted the heteromerization and a negative crosstalk, by which galanin counteracted endomorphin-1-mediated MAPK activation [87••] (Fig. 4). The negative crosstalk constituted therefore a biochemical property of μ-opioid-Gal₁ heteromer, which could also
be identified in situ in VTA slices and in vivo with microdialysis experiments. Thus, galanin completely counteracted somatodendritic dopamine release induced by the local infusion of endomorphin-1. Both in situ and in vivo galanin-opioid interactions were also selectively counteracted by application of the disruptive peptide, demonstrating their dependence on μ-opioid-Gal1 heteromerization \[87\]. These results indicate that dopaminergic cell function in the VTA is modulated by a predominant population of μ-opioid receptors forming heteromers with Gal1 receptors. Therefore, μ-opioid-Gal1 receptor heteromers constitute an obvious target for the treatment of opioid use disorders.

There is also indirect evidence for the existence of ghrelin receptor heteromers and neurotensin receptor heteromers localized in the dopaminergic cells, although they do not fulfill yet the criteria for their identification within the VTA \[78–19, 80\]. Ghrelin receptors are known as growth hormone secretagogue (GHS) receptor or GHS\(_{1a}\) receptors. Cells expressing GHS\(_{1a}\) also express GHS\(_{1b}\) receptors, a truncated variant of GHS\(_{1a}\) receptors lacking the transmembrane domains 6 and 7. Ghrelin does not bind and therefore does not signal through GHS\(_{1b}\) receptors \[99\] and the role of this truncated “receptor” on ghrelin-mediated signaling is just beginning to be understood. Evidence has been provided for the ability of GHS\(_{1a}\) to homodimerize and to heterodimerize with GHS\(_{1b}\) receptors, which allows GHS\(_{1b}\) to produce a dominant negative effect on GHS\(_{1a}\) receptor signaling \[99, 100\]. Using mammalian transfected cells and neuronal cells in culture, we found, first, a significant and complex modulatory role of GHS\(_{1b}\) in the trafficking and signaling of GHS\(_{1a}\) receptors that depends on the relative expression of both proteins \[101\]. An additional finding in striatal and hippocampal neurons in culture was a predominant Gs/olf protein-dependent signaling of ghrelin, which in striatal neurons depended on D\(_1\)-GHS\(_{1a}\)-GHS\(_{1b}\) receptor heteromerization \[101\] (Fig. 5). A D\(_1\) receptor antagonist blocked ghrelin-induced cAMP accumulation in striatal but not hippocampal neurons, indicating the involvement of D\(_1\) receptors in the striatal GHS\(_{1a}\)-Gs/olf coupling. Experiments in transfected cells demonstrated that D\(_1\) receptor co-expression promotes a switch in GHS\(_{1a}\)-G protein coupling, from Gi/o to Gs/olf, but only upon co-expression of GHS\(_{1b}\). In fact, with biophysical techniques (resonance energy transfer experiments), it could be demonstrated that D\(_1\) receptor interacts with GHS\(_{1b}\) but only in the presence of GHS\(_{1b}\) \[101\]. Finally, a negative crosstalk could also be observed upon co-administration of D\(_1\) and GHS\(_{1a}\) receptor agonists \[101\] (Fig. 5). Therefore, GHS\(_{1b}\) not only

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**Fig. 4.** The μ-opioid-galanin Gal\(_1\) receptor heteromer. The heteromer can have a heterotetrameric structure, constituted by homodimers of two Gi-coupled receptors, the μ-opioid (μOR) and the Gal\(_1\) receptor (Gal1R). Within the heteromer, galanin exerts a strong negative allosteric control of μOR-mediated MAPK signaling (its G protein dependence or independence still needs to be determined). This heteromer constitutes a predominant population of μOR localized in the VTA.
The hypothalamic-VTA connection provides a link between the center of integration of internal homeostatic, threat and reproductive signals, and dopaminergic neurotransmission, which determines the degree of psychomotor activation and reinforcing effects elicited by those stimuli. Apart from classical neurotransmitters, the very heterogeneous hypothalamic-VTA connection releases a series of neuropeptides in a volume-transmission mode which, together with several hormones that seem to be able to cross the blood-brain barrier within the VTA by saturable tracers, simultaneously influences the activity of dopaminergic cells. We introduce first the concept of targeting receptors for hormones and neuropeptides localized in the VTA as a therapeutic approach for determining the efficacy of ghrelin-induced GHS$_{1a}$-mediated signaling but also determines the ability of GHS$_{1b}$ to form oligomeric complexes with other receptors promoting profound qualitative changes in ghrelin-induced signaling.

Now, there is evidence for the localization of both GHS$_{1a}$ and D$_1$-like receptors (D$_1$ or D$_5$ receptor subtype) in the VTA dopaminergic cells [102]. It is therefore very likely that these neurons will also express D$_{1/5}$-GHS$_{1a}$-GHS$_{1b}$ receptor heteromers. In fact, using the infusion-microdialysis technique, we have obtained evidence for a strong ghrelin-induced somatodendritic dopamine release that is counteracted by co-perfusion with a D$_1$ receptor antagonist (in preparation). In addition, GHS$_{1a}$ receptors could also oligomerize with melanocortin receptors in the VTA, as the predominant subtype expressed by dopaminergic neurons, MC$_3$ receptors [49], has been reported to form heteromers with GHS$_{1a}$ in the hypothalamic adenylyl cyclase (AC) [103]. Ghrelin receptor heteromers in the VTA could specifically provide new therapeutic targets for obesity associated with high ghrelin serum levels, such as PWS [21]. Finally, NT$_1$-D$_2$ receptor heteromers [104, 105] would also be localized in dopaminergic cells and could mediate the activating effects of neurotensin related to disinhibition of D$_2$ autoreceptor function [44]. NT$_1$-D$_2$ receptor heteromers would therefore provide an additional disinhibitory mechanism of dopaminergic cell activity provided by GABA-neurotensin-expressing HL neurons.

## Conclusions

Fig. 5. The ghrelin GHS$_{1a}$-GHS$_{1b}$-dopamine D$_1$ receptor heteromer. Oligomerization with the truncated ghrelin GHS$_{1b}$ receptor (GHS$_{1b}$R) allows GHS$_{1a}$R to heteromerize with dopamine D$_1$-like receptors (D$_1$R or D$_3$R) and couple to Gs protein. Activation of AC becomes therefore a main signaling pathway of the heteromer. Within the heteromer, dopamine and ghrelin receptor ligands establish negative allosteric interactions. This heteromer has been demonstrated in striatal cells in culture, and indirect evidence indicates that it is also functionally present in the VTA.
the loss of control of food intake and substance use disorders. Second, we introduce the concept that the specific targeting of those receptors can be achieved by targeting specific VTA GPCR heteromers.

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Compliance with Ethical Standards

Conflict of interest

Sergi Ferré declares that he has no conflicts of interest.

Human and animal rights and informed consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

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