Multiple evolutionary routes of the single polar capsule in *Thelohanellus* species (Myxozoa; Myxobolidae)

Xiuping Zhang\(^a,1\), Yang Lii\(^a,1\), Christopher M. Whippsc, Qingxiang Guoa, Zemao Gua,b,\(^*\)

\(^a\) Department of Aquatic Animal Medicine, College of Fisheries, Huazhong Agricultural University, Wuhan, 430070, China
\(^b\) Hubei Engineering Technology Research Center for Aquatic Animal Diseases Control and Prevention, Wuhan, 430070, China
\(^c\) SUNY-ESF, State University of New York College of Environmental Science and Forestry, Environmental and Forest Biology, 246 Illick Hall, 1 Forestry Drive, Syracuse, NY, 13210, USA

**A B S T R A C T**

*Thelohanellus* Kudo, 1933 is a species rich genus of Myxosporea, sharing many morphological similarities with species of *Myxobolus* but the former possesses a single polar capsule, and the latter has two. Based on molecular phylogenetic analyses, this single distinguishing feature is not monophyletic, and members of *Thelohanellus* are intermixed with *Myxobolus* species, calling into question the validity of genus *Thelohanellus*. The occurrence of two polar capsules in a small proportion of *Thelohanellus* spores as observed in this study suggests that these species have the capacity to express this *Myxobolus*-like trait, clouding the distinction of these two genera further. Herein, using the most comprehensive data set to date, we explored the phylogenetic relationships of *Thelohanellus* to other myxobolids, to investigate the evolutionary history of the genus *Thelohanellus* and the origins of single polar capsule in this group. The phylogenetic analyses and statistical tests of topology revealed *Thelohanellus* as a strongly supported polyphyletic lineage, clustering in five distinct branches within *Myxobolus* clade. Ancestral state reconstruction for polar capsule number showed that *Thelohanellus* species have evolved from myxosporean species with two polar capsules at least four times, which could be classified in three possible evolutionary pathways. The polyphylly of *Thelohanellus* and the multiple evolutionary origins of single polar capsule of *Thelohanellus* demonstrate that the distinction of this genus from *Myxobolus* is largely for convenience, and does not reflect their evolutionary history.

**1. Introduction**

Myxosporeans are a group of diverse and widely distributed metazoan endoparasites with extremely reduced body size and structure (Okamura et al., 2015). Under the recent advance of multi-gene analyses, myxosporeans were found to diverge from free-living cnidarian ancestors in ancient times (Jiménez-Guri et al., 2007; Feng et al., 2014; Okamura et al., 2015). Under the recent advance of multi-gene analyses, myxosporeans were found to diverge from free-living cnidarian ancestors in ancient times (Jiménez-Guri et al., 2007; Feng et al., 2014; Okamura et al., 2015). Under the recent advance of multi-gene analyses, myxosporeans were found to diverge from free-living cnidarian ancestors in ancient times (Jiménez-Guri et al., 2007; Feng et al., 2014; Okamura et al., 2015).

By construction, myxosporean species (Myxozoa; Myxobolidae) are often referred to as having a single polar capsule. This traditional assumption is based on the morphological diversity of myxosporeans, especially those that formed the new myxosporean genera (Myxidium, Chloromyxum, Kudoa, and Ceratomyxa, to name a few (Fiala, 2006; Lom and Dyková, 2006; Fiala et al., 2015a)). In fact, most myxosporean genera where molecular sequences are available were resolved as non-monophyletic. Such great discrepancies between traditional classification and DNA sequence-based phylogenies have resulted in a number of taxonomic revisions (Whipp et al., 2004; Gleeson and Adlard, 2012; Bartošová et al., 2013) and have motivated taxonomists to consider both morphological and genetic data when classifying myxosporeans.

To date, approximately 2600 myxosporean species have been described and classified in 2 orders, 16 families and 62 genera based mainly on the structure and shape of myxosporites (Lom and Dyková, 2006; Fiala et al., 2015a; Okamura et al., 2018). However, this classical spore-morphology-based taxonomic system is often in conflict with DNA sequence-based phylogenies (Fiala, 2006; Lom and Dyková, 2006; Bartošová et al., 2013). Specifically, there are a number of poly- or paraphyletic genera, such as *Henneguya*, *Sphaerospora*, *Myxidium*, *Chloromyxum*, *Kudoa*, and *Ceratomyxa*, to name a few (Fiala, 2006; Lom and Dyková, 2006; Fiala et al., 2015a). In fact, most myxosporean genera where molecular sequences are available were resolved as non-monophyletic. Such great discrepancies between traditional classification and DNA sequence-based phylogenies have resulted in a number of taxonomic revisions (Whipp et al., 2004; Gleeson and Adlard, 2012; Bartošová et al., 2013) and have motivated taxonomists to consider both morphological and genetic data when classifying myxosporeans.

The characters used to establish many myxosporean genera centered around morphological features of the myxospore (polar capsule number and orientation, overall shape, number of valves, etc.). For example, what we now classify as *Thelohanellus* species were originally placed in the genus *Myxobolus* Bütschli, 1882 based on the presence of idoinophilous vacuole in the sporoplasm, and other general features of...
the spore (pyriform, two valves, no tails). Subsequently, Kudo (1933) established genus *Thelohanellus* to delineate species with spores containing one polar capsule from those of *Myxobolus* which possess two polar capsules. Currently, *Thelohanellus* comprises 108 nominal species, most of which are plasmodia-forming parasites infecting various tissues of freshwater fishes, and some of which are pathogenic, causing economical losses in aquaculture (Zhang et al., 2013). The monophyly of genus *Thelohanellus* is not supported by phylogenetic analyses, with *Thelohanellus* species clustered in several *Myxobolus* clades (Griffith and Goodwin, 2011; Shin et al., 2014; Yuan et al., 2015; Liu et al., 2016; Ye et al., 2017). This suggests that the difference in polar capsule number between *Thelohanellus* and *Myxobolus* is not a character that reflects a single evolutionary divergence, but has evolved several times. To investigate this further, we explored the phylogenetic relationships of *Thelohanellus* species and traced the evolutionary origins of the single polar capsule within the larger myxobolid clade.

2. Materials and methods

2.1. Myxospore samples and morphological observation

Fresh spores of two *Thelohanellus* species were collected from their hosts in Hubei Province in China during 2015–2016 (Table 1). Spores were photographed using an Olympus BH2 microscope equipped with the Olympus DP73 camera and measured following the guidelines of Lom and Arthur (1989). All measurements are given in micrometers (μm).

2.2. DNA extraction, amplification and sequencing

Total genomic DNA was extracted from fresh spores using General All Gen Kit (animal tissue protocol; CoWin Biosciences, China) following manufacturer’s protocol. Small subunit ribosomal DNA (SSU rDNA) was amplified using primers pair 18e of Hills and Dixon (1991) and 18R of Whipp et al. (2003). Polymerase chain reactions (PCRs) were performed in a 50-μl reaction solution comprising approximately 200 ng of extracted genomic DNA (2 μl), 25 μl 2× Es Taq MasterMix (CWBio, China), and 10 pmol each primer. The reactions were run on an Eppendorf Mastercycler® nexus GX2 gradient thermocycler (Germany) with cycling parameters as follows: an initial denaturation at 94 °C for 7 min, followed by 35 cycles of 94 °C for 45 s, 55 °C for 45 s, and 72 °C for 60 s, and a terminal extension at 72 °C for 10 min. The PCR products were purified using the Gel Extraction Kit (CWBio, China) and sequenced in both directions on the ABI PRISM 3730XL DNA sequencer (Applied Biosystems Inc., Foster City, CA, USA). The SeqMan utility of the Lasergene software package (DNAStar, USA) was used to assemble contiguous sequences which were subsequently uploaded to NCBI with accessions MHN29614 and MHN29616.

2.3. Phylogenetic analyses

Only species with molecular sequence and morphological records provided by the same peer reviewed paper were included in the present study. In addition, most sequences used were selected with a length threshold set above 1300 bp. These included mostly *Myxobolus*, *Henneguya* and *Thelohanellus* species, but also *Cardimyoxobolus*, *Dicauda*, *Triangula*, *Unicauda* and *Hennegoides*. Three species from the family Myxidiidae were used as outgroup. Alignments of SSU rDNA sequences were performed in MAFFT v7.306b (Katoh and Standley, 2013) with L–INS–I method which is optimized for sequence with multiple conserved domains and long gaps, such as rDNA genes. Gblocks 0.91b (Castresana, 2000) was used to remove ambiguous regions and highly variable sections (parameters were: t = d, -b1 = 65, -b2 = 67, -b3 = 10, -b4 = 5, -b5 = a), 71% positions of the original data set were saved. Substitution saturation was evaluated using DAMBE 5 (Xia and Xie, 2001). Maximum likelihood (ML) analysis was performed using RAxML 8.2.9 (Stamatakis, 2014) with a GTR GAMMA model as recommended by the author of the program. One thousand bootstrap replicates were performed to assess clade support. The above analyses were performed again after removing ambiguous species identified by RogueNaRok algorithm (Aberer et al., 2012), Bayesian Inference (BI) analysis was done in MrBayes v3.2.6 (Ronquist et al., 2012) with GTR + G + I model indicated by evaluation with jModelTest 2.1.10 (Darriba et al., 2012) under Akaike Information Criterion (AIC). Four parallel MCMC runs of 1 million generations were conducted with a burn-in of 25% of the samples and sampled every 100 generations. A 50% majority-rule consensus tree, with mean branch lengths, and posterior probabilities of clades was generated to summarize the results of the post-burn-in posterior distribution of trees. Convergence of the Markov chains was confirmed in Tracer v.1.6 (Rambaut and Drummond, 2007) by a combined effective sample size (ESS) > 200.

2.4. Testing alternative phylogenetic hypotheses

Phylogenetic trees that constrained the involved genera to be monophyletic and per-site log likelihood scores for unconstrained and constrained ML trees were created using RAxML v8.2.9 with the GTR GAMMA model mentioned above. We calculated approximately unbiased (AU) values using CONSEL v1.20 (Shimodaira and Hasegawa, 2001) to determine statistical significance of differences in the likelihood scores. The alternative tree topologies could be rejected at the 5% confidence level.

2.5. Ancestral state reconstruction

In order to investigate the origin of *Thelohanellus* species, we traced the evolutionary lineage of the polar capsule number with the following possible three states: 1) single polar capsule; 2) two unequal polar capsules; 3) two equal polar capsules. In some cases of *Myxobolus* species, it is difficult to determine whether the two polar capsules are equal or not (Lom and Dyková, 2006) because this relies largely on prior authors’ opinions. In these cases, where polar capsules were reported as slightly different, almost equal, similar, etc., we classified them as equal (Supplementary Table 1). In addition, to avoid mismatches of molecular data and morphology brought by misidentification of species, morphological descriptions in present study were referred to articles that provided the molecular data rather than original records. The SSU-based ML trees were chosen as basis for reconstruction of ancestral states. A standard categorical matrix of polar capsule number (2 - two equal polar capsules, 1 - single polar capsule, 0 - two unequal polar capsules) was created to map on the tree. Mesquite v3.20 (Maddison, 2008) was used to trace the evolutionary history of this character. Reconstruction of character states at ancestor nodes was done by likelihood method with Markov k-state 1 parameter model. Any change in character state is equally probable within this model.

Table 1

| Myxosporean species | Host species | Dates of collection | Location | Site of infection | Accession No. |
|---------------------|--------------|---------------------|----------|------------------|---------------|
| *Thelohanellus kitauei* | *Cyprinus carpio* | March 2015 | Baishazhou Fish Market, Wuhan City, Hubei Province, China | Intestine | MHN29614 |
| *Thelohanellus wuhanensis* | *Carassius auratus gibelio* | Jun 2016 | Honghu Lake, Hubei Province, China | Skin | MHN29616 |
3. Results

3.1. Atypical spore morphology

Two *Thelohanellus* species (*Thelohanellus kitauei* and *Thelohanellus wuhanensis*) showed two morphotypes (Fig. 1) in a plasmodium: typical spores with one polar capsule and atypical spores with two polar capsules. Besides the polar capsule number, they were different in spore shape, spore length and spore width. The atypical spore and typical spore of *T. kitauei* (Fig. 1 A) are contrasted as follows: oval versus prolonged pyriform shape, 22.62 versus 29.34 ± 1.34 (26.49–32.08) (n = 30) in spore length, and 16.00 versus 9.52 ± 0.88 (8.34–11.42) (n = 30) in spore width. The atypical spore and typical spore of *T. wuhanensis* (Fig. 1 B, C) are contrasted as follows: oval versus prolonged pyriform shape, 19.22 versus 22.86 ± 0.77 (21.63–24.69) (n = 30) in spore length, and 19.73 versus 12.05 ± 0.71 (11.01–13.58) (n = 30) in spore width. Despite the differences in overall spore shape, the polar capsules of the atypical spore and typical spore of *T. kitauei* (Fig. 1 A) and *T. wuhanensis* (Fig. 1 B, C) were similar in shape whether there was one or two. The proportion of atypical spores in the plasmodia of both present *Thelohanellus* species was exceedingly low, approximately 0.1% (n = 1000).

3.2. Phylogenetic relationships among myxobolid species

Identification of rogue taxa was implemented on the complete data set (138 taxa) using RogueNaRok, showed that 9 taxa (Supplementary Table 2), significantly increased the nodal support and did not affect the topology of the tree when removed from the analysis. The subsequent phylogenetic analyses were performed on the pruned data set, with the ingroup consisting of 126 myxobolid species covering 8 genera (*Myxobolus*, *Thelohanellus*, *Henneeguya*, *Unicauda*, *Dicauda*, *Henneegoides*, *Cardimyxoabulus*, *Triangula*). Tree topologies retrieved from ML and BI analyses were identical among main clades, both resulting in 5 clades 

![Fig. 1. Photomicrograph of fresh spores of *Thelohanellus kitauei* and *Thelohanellus wuhanensis*. A - the spore of *T. kitauei* with two polar capsules (arrow); B, C - the spores of *T. wuhanensis* with two polar capsules (arrows). Scale bars indicate 20 μm for A, B and C.](image)

except the central part of the tree. However, clade E, comprising all of *Thelohanellus* species, clustered stably as one of the most diverged branches free from the impact of different phylogenetic methods.

3.3. Hypothesis testing

We performed the AU tests to further test the hypothesis that most of the genera are non-monophyletic group (Table 2). The monophyly of *Thelohanellus*, *Myxobolus*, *Unicauda* and *Henneegoides* was significantly rejected (p < 0.05). However, the monophyly of *Henneegoides* was not rejected (p = 0.541).

3.4. Reconstruction of ancestral polar capsule number

We conducted an ancestral state reconstruction analysis based on the preferred ML topology (Fig. 2) to investigate myxoboloid evolution, particularly focused on the transitional events of the polar capsule number (Fig. 3). The analysis assigned high probability for most nodes. The ancestor of all myxobolid species was reconstructed to have two equal polar capsules with a probability 89%. The polar capsule number of the most recent common ancestors of clade E and clade A, B, C, D had...
Fig. 2. SSU rDNA based maximum likelihood tree of selected myxobolid species. The table containing detail information of taxa is shown behind taxa names; Bootstrap supports and posterior probabilities are given beside the nodes, respectively. Asterisks represent values = 100/1.00. Dashes represent values < 60/0.60.
not shifted, and were suggested to be two equal polar capsules with a probability > 95%. Within clade E, there were evidences for six different evolutionary lineages of the polar capsule morphology (number and relative size): 1) Species with two equal polar capsules evolved to species with two unequal polar capsules, which was the most common shift, forming species like *Myxobolus orissae*, *Myxobolus kingchowensis*, and *Myxobolus csabai*. 2) Species with two equal polar capsules transferred to species with single polar capsule, resulting in species like *T. wuhanensis*, *T. kitauei*, and *T. zahraeae*. 3) Species with two equal polar capsules evolved to species with two unequal polar capsules, and then transformed to species with single polar capsule, leading to *T. qadri*. 4) Species with two equal polar capsules evolved to species with single capsule, transformed to species with two unequal polar capsules subsequently, transformed to species with single capsule finally, resulting in *T. caudatus* and *T. habibpuri*. 5) Species with two equal polar capsules converted into species with single polar capsule, and then became species with two unequal polar capsules, evolved to species with two equal polar capsules eventually, resulting in species like *Myxobolus macrocapsularis*, *Myxobolus bliccae*, and *Myxobolus shaharomae*. 6) Species with two equal polar capsules evolved to species with single polar capsule, then transferred to species with two unequal polar capsules.

Table 2

Results of approximately unbiased tests comparing the best maximum likelihood tree with the best constrained trees obtained under alternative hypotheses of monophyletic genera. Results in which P-values < 0.05 are shown in bold.

| Alternative hypothesis       | P-value       |
|------------------------------|---------------|
| unconstrained tree           | 0.459         |
| monophyly of Thelohanellus   | $5 \times 10^{-120}$ |
| monophyly of Myxobolus       | $6 \times 10^{-12}$ |
| monophyly of Henneguya       | $8 \times 10^{-65}$ |
| monophyly of Dicauda         | $2 \times 10^{-12}$ |
| monophyly of Hennegoides     | 0.541         |

Fig. 3. Ancestral polar capsule number reconstruction for myxobolid species based on the preferred ML tree (Fig. 2). Pie charts show the marginal probability of the ancestral state at each node. Green circle represents species with single polar capsule, black circle represents species with two equal polar capsules, white circle represents species with two unequal polar capsules. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)
became species with two equal polar capsules subsequently, transformed to species with two unequal polar capsules finally, leading to *Myxobolus diversicolor*.

To further evaluate the effect of topological instability on the results of reconstruction, we also reconstructed the ancestral state based on the five alternative tree topologies (Supplementary Fig. 2). There is no difference from the evolutionary lineages documented above according to the ancestral state reconstruction based on topology 2, topology 4, topology 5 and topology 6. However, the phylogenetic position of *T. caudatus* and *T. habibpuri* altered in topology 3, making the origins of *T. caudatus* and *T. habibpuri* obscure.

4. Discussion

Myxosporean classification was established based mainly on spore morphology, and currently there are 16 established families (Fiala et al., 2015a). Of them, the family Myxobolidae has the most nominal species and is split into 14 genera based largely on the number of polar capsules and projections, and shape of sutural line. However, the monophyly of some genera of Myxobolidae was not supported by phylogenetic analyses (Kent et al., 2001; Fiala, 2006; Fiala and Bartošová, 2010; Moreira et al., 2014; Rocha et al., 2015; Capodifoglio et al., 2016). Our phylogenetic analyses included eight myxosporean genera, which were also shown as paraphyletic or polyphyletic groups except *Dicauda*, *Cardimyobolus* and *Triangula*, for which there is only one SSU rDNA sequence included. Members of *Myxobolus* were distributed throughout the phylogenetic tree, indicating the close interrelatedness with the species of other genera, such as *Henneguya*, *Dicauda*, *Thelohanellus*, and *Unicauda*. In fact, spore features that distinguish these genera from *Myxobolus*, such as the spore projections of *Henneguya* or single polar capsule of *Thelohanellus*, likely arose from *Myxobolus* ancestors (Fiala and Bartošová, 2010). The genus *Hennegoides* was paraphyletic but both species are within the same clade including a *Unicauda* species. Still, the monophyly of *Hennegoides* was not rejected by AU tests. Consistent with the results of Rosser (2016), the genus *Unicauda* was resolved as non-monophyletic group in present study. *Henneguya* species nested with *Myxobolus* species in clade B and D, but formed a monophyletic assemblage in clade B in the present study. The polyphyly of *Henneguya* is well noted as early as the first broad phylogenetic analysis of Myxosporea (Kent et al., 2001). All members of *Thelohanellus* clustered in clade E, but didn’t form a monophyletic group, consistent with previous studies (Griffin and Goodwin, 2011; Yuan et al., 2015; Liu et al., 2016). The present study highlights the general non-monophyly of genera within Myxobolidae, indicating that taxonomic features used to establish different genera may be plastic and unstable. Gain or loss of these features probably occurs several times in their evolutionary history.

According to polar capsule number, the family Myxobolidae was divided into genera with two polar capsules (ten genera) and genera with single polar capsule (four genera) (Lom and Dykova, 2006; Sarkar, 2009). Among these two groups, *Myxobolus* and *Thelohanellus* were the first established genera characterized by 2-PCs and one-PC, respectively. Based on the reconstruction of myxozoan morphotypes, Fiala and Bartošová (2010) proposed that the ancestor of myxobolids had two polar capsules, which was supported by our results. Moreover, we found that the two polar capsules of the ancestor of myxobolids were equal in size. Consideration of polyphyly of genus *Thelohanellus* and the placement of *Thelohanellus* species within several *Myxobolus* clades, Yuan et al. (2015) speculated that ancestors of *Thelohanellus* underwent several times of loss or gain of polar capsule in the evolutionary history. The ancestral state reconstruction in present study revealed that the formation of single polar capsule of *Thelohanellus* happened four times after the most recent common ancestor of *Thelohanellus* infecting cyprinid fish. However, the possibility of ancestral state of the clade rooted by *T. catleae* to be single polar capsule (0.61) is not very high. The situation that species within this clade evolved with an independent origin of a single polar capsule could not be excluded, which makes us conclude that *Thelohanellus* evolved at least four times. These transitional events could be classified in three different ways according to the evolutionary process: (1) the direct type: single polar capsule formed by two equal polar capsules losing one polar capsule directly; (2) the gradual type: single polar capsule formed by one of the two polar capsules gradually degenerating and finally missing; and (3) the devious type: single polar capsule formed by losing one of the two equal polar capsules firstly, gaining one polar capsule subsequently, and losing one polar capsule eventually. However, this evolutionary process of the devious type might be changeable due to the unstable phylogenetic position of *T. caudatus* and *T. habibpuri*.

Surprisingly, the *Thelohanellus* spores observed in the present study sometimes possessed two polar capsules, which indicated *Thelohanellus* species have the genetic capacity to express two polar capsules. In addition, Desser et al. (1983) observed two capsulogen cells in the young sporoblast of *Thelohanellus nikolskii* and its single polar capsule was formed by abortion of the second capsulogen cell, supporting the idea that ancestor of *Thelohanellus* species had two polar capsules.

A single polar capsule is the only feature that discriminates *Thelohanellus* from *Myxobolus*, yet this character is of limited taxonomic significance as its multiple evolutionary routes and the polyphyletic character of *Thelohanellus*. A possible course of action would be to synonymize *Thelohanellus* with *Myxobolus*. This may be the ultimate course of action, but we recognize that there is utility in maintaining separate genera for the ease of identification, particularly with pathogenic species.

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Declarations of interest

None.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.ijppaw.2018.11.006.

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