Title:
X-ray stereo microscopy for investigation of dynamics in soil

Author:
Gleber, S.-C.

Publication Date:
11-20-2008

Permalink:
http://escholarship.org/uc/item/5j35p54h

Copyright Information:
All rights reserved unless otherwise indicated. Contact the author or original publisher for any necessary permissions. eScholarship is not the copyright owner for deposited works. Learn more at http://www.escholarship.org/help_copyright.html#reuse
X-ray stereo microscopy for investigation of dynamics in soils

S-C Gleber\textsuperscript{1}, J Sedlmair\textsuperscript{1}, M Bertilson\textsuperscript{3}, O von Hofsten\textsuperscript{3}, S Heim\textsuperscript{4}, P Guttmann\textsuperscript{1,4}, H Hertz\textsuperscript{3}, P Fischer\textsuperscript{2}, and J Thieme\textsuperscript{1}

\textsuperscript{1} Institut für Röntgenphysik, Georg-August-Universität Göttingen, Friedrich-Hund-Pl.1, 37077 Göttingen, Germany
\textsuperscript{2} Center for X-Ray Optics, Lawrence Berkeley National Laboratory, 1 Cyclotron Road, Berkeley CA 94720, USA
\textsuperscript{3} Biomedical and X-ray Physics, Department of Applied Physics, Royal Institute of Technology, AlbaNova, SE-10691 Stockholm, Sweden
\textsuperscript{4} Berliner Elektronenspeicherring Gesellschaft für Synchrotronstrahlung m.b.H., Albert-Einstein-Str. 15, 12389 Berlin, Germany

E-mail: sgleber@gwdg.de

Abstract. The here presented combination of stereo imaging and elemental mapping with soft X-ray microscopy reveals the spatial arrangement of naturally aqueous colloidal systems, e.g. iron oxides in soil colloid clusters. Changes in the spatial arrangement can be induced by manipulating the sample mounted to the X-ray microscope and thus be investigated directly.

1. Introduction
Due to its high spatial and spectral resolution, X-ray microscopy is an important instrument for investigation of colloidal systems, e.g. from environmental sciences \cite{1}. Taking advantage of the natural contrast mechanism, sample preparation is not necessary, it can be imaged in transmission within its original aqueous media up to 10\,\mu m thickness with a resolution in the range of 20-50\,nm. In soil science, the element distribution within soil colloid clusters is of great interest \cite{2}. With images taken above and below an absorption edge, distribution maps of the corresponding element are achieved. Thus, the iron distribution in colloidal samples from the environment has been determined. By tilting the object, stereo pairs of images have been taken. The here presented combination of elemental mapping with X-ray microscopy and stereo imaging provides a tool for that and reveals the spatial arrangement of e.g. iron oxides in soil colloid clusters. Changing the chemical conditions of aqueous medium leads to changes in the spatial arrangement, which can be done directly in the X-ray microscope. Since many of the so induced morphological changes are much slower than the time required even for taking stereo pairs, these changes can be investigated. Since many of the so induced morphological changes are much slower than the time required even for taking stereo pairs, these changes can be investigated. Theory and results of such experiments at three different setups will be presented briefly.

2. Parallax equation based stereo reconstruction
Stereo calculations are based on the parallax equation \cite{3}. It relates the parallax $\Delta Y$ to the vertical distance $h$ and the tilt angle $2\theta$ ($\theta$ is the stereo angle). Thus, for a tilt by $\theta_2 - \theta_1 = \theta$
around the x-axis, the z-coordinate of a structure can be calculated by the difference between the y-coordinates \((y_1 \text{ and } y_2)\) in the respective images projecting the structure onto the x-y-plane (Fig.1). The spatial coordinates become:

\[
X = x_1 = x_2 \\
Y = \frac{y_1 + y_2}{2} \\
Z = \frac{y_1 - y_2}{2 \sin \theta}
\]

A program has been written in IDL to mark or select significant structures in a set of tilted x-ray images and get information about the 3-d configuration, distances and lengths. Matching points recognizable in both projections are marked manually. In the 3-d plot, features like curvatures are displayed.

3. Experiments with XM-1, ALS
The soft X-ray microscope XM-1 is a full field microscope at a bending magnet [4]. For the experiments, a 25 nm micro zone plate was used to take high resolution images below and above the Fe II edge at 707 eV. The magnification was about 2800, the spectral resolution \(E/\Delta E \approx 500\).

Two kinds of tiltable sample holders for aqueous samples were used: glass capillaries of 1 mm diameter with tips of about 3 \(\mu\)m diameter, and flat holders with two \(\text{Si}_3\text{N}_4\) foils of 100 nm thickness each. Both holders were sealed, so the samples kept aqueous over several hours.

In Fig. 2, images of an aqueous flock of montmorillonite with hematite between two \(\text{Si}_3\text{N}_4\) foils taken at 704 eV (left) and 707 eV (right) are shown. The hematite particles are clearly appearing at the Fe II absorption edge. Fig. 3 presents a detail of the montmorillonite sample shown in Fig. 2 viewed with an angular difference between both images of 14° around a horizontal tilt axis. Both images are taken at \(E = 707\) eV. Lines, points, and edges are marked for analysis. The results from xstereo are given on Fig. 4: two views of a 3-d plot of points and edges marked in Fig. 3. The unit of the axis is pixel, where 11 pixel relate to 100 nm. Distances had been determined e.g. between structure 1 (hematite) and 7 (clay edge) to 180 nm, or between structure 4 (hematite) and 11 (clay edge) to 630 nm, revealing results about the interrelation between pH value of the solution and the corresponding edge charge of the clay platelets [5].

4. Experiments with the compact X-ray microscope, KTH
The compact X-ray microscope at the KTH, Stockholm, Sweden, provides a sub-40 nm resolution in the lab. It is based on a liquid-nitrogen-jet laser-plasma source and two zone plates [6]. The images were taken at \(\lambda = 2.48\) nm wavelength with exposure times between 4 and 8 minutes. The image diameter is 20 \(\mu\)m. The vertical tilt was given by the stereo mount to be 14°, which is ideal for human stereo vision. Two pairs of images are shown in Fig. 5 and 6.
5. Experiments with the STXM, BESSY

The Scanning Transmission X-ray Microscope at the Undulator U41 at BESSY scans the sample with a 50 nm spot, at best resolution a region of up to 40x40 µm² [7]. The sample stage has been adapted for tilting capillaries.

Fig. 7 shows montmorillonite in a capillary. Two pairs of images were taken of the same sample region with equal image parameters, one before (Fig. 7 up) and one after (Fig. 7 below) adding carbon nanotubes. Both image pairs were taken at 400 eV within 30 minutes, respectively, with a horizontal tilt of 15° of a sample region of 8x7 µm² onto 160x140 pixel² with 16 ms exposure time per pixel. The time between the pairs was 5 hours, resulting in a big change in structure.

6. Conclusions

The suitability of a TXM, a STXM, and a TXM with a laboratory source for the investigation of samples from the environment in ambient conditions in combination with stereo imaging and elemental mapping could be shown. Manipulating the sample and tilting the sample was possible even under observation. Therefore, studies of the dynamical behavior of samples will be performed as a next step using both, stereo imaging and elemental mapping, to obtain a complete description of the system under investigation.

Acknowledgments

This work is supported by the DFG within the Collaborative Research Center SFB 755 Nanoscale Photonic Imaging and under contract numbers Th445/8-1 and Th445/8-2. The ALS microscope is supported by the Director, Office of Science, Office of Basic Energy Sciences, Materials Sciences and Engineering Division, of the U.S. Department of Energy. The KTH microscope is supported by the Swedish Science Research Council, the Göran Gustafsson Foundation, and the Wallenberg Foundation. We thank the staff of ALS and BESSY for providing excellent working conditions.

References

[1] Thieme J, McNulty I, Vogt S and Paterson D Environm. Sc. and Tecn. 41
[2] Mitrea G, Thieme J, Guttmann P, Heim S and Gleber S C 2008 J. Synchr. Rad. 15 26
[3] Gleber S C, Knoechel C, Thieme J, Rudolph D and Schmahl G 2003 X-Ray Microscopy 2002 (J. Physique IV Proc. vol 104) ed Susini J, Joyeux D and Polack F (EDP Sciences, Les Ulis) p 639
[4] Fischer P, Kim D H, Chao W, Liddle J A, Anderson E H and Attwood D Mat. Today 9
[5] Gleber S C, Thieme J, Chao W and Fischer P J. Micr. submitted
[6] Takman P, Stollberg H, Johannson G, Holmberg A, Lindblom M and Hertz H 2007 J. Micr. 226 175
[7] Wiesemann U 2003 The Scanning Transmission X-Ray Microscope at BESSY II Ph.D. thesis Goettingen