Pain Experience is Somatotopically Organized and Overlaps with Pain Anticipation in the Human Cerebellum

F. H. S. Michelle Welman, Albertine E. Smit, Joost L. M. Jongen, Dick Tibboel, Jos N. van der Geest, Jan C. Holstege

Published online: 26 February 2018
© The Author(s) 2018. This article is an open access publication

Abstract

Many fMRI studies have shown activity in the cerebellum after peripheral nociceptive stimulation. We investigated whether the areas in the cerebellum that were activated after nociceptive thumb stimulation were separate from those after nociceptive toe stimulation. In an additional experiment, we investigated the same for the anticipation of a nociceptive stimulation on the thumb or toe. For his purpose, we used fMRI after an electrical stimulation of the thumb and toe in 19 adult healthy volunteers. Following nociceptive stimulation, different areas were activated by stimulation on the thumb (lobule VI ipsilaterally and Crus II mainly contralaterally) and toe (lobules VIII-IX and IV-V bilaterally and lobule VI contralaterally), i.e., were somatotopically organized. Cerebellar areas innervated non-somatotopically by both toe and thumb stimulation were the posterior vermis and Crus I, bilaterally. In the anticipation experiment, similar results were found. However, here, the somatotopically activated areas were relatively small for thumb and negligible for toe stimulation, while the largest area was innervated non-somatotopically and consisted mainly of Crus I and lobule VI bilaterally. These findings indicate that nociceptive stimulation and anticipation of nociceptive stimulation are at least partly processed by the same areas in the cerebellum. This was confirmed by an additional conjunction analysis. Based on our findings, we hypothesize that input that is organized in a somatotopical manner reflects direct input from the spinal cord, while non-somatotopically activated parts of the cerebellum receive their information indirectly through cortical and subcortical connections, possibly involved in processing contextual emotional states, like the expectation of pain.

Keywords fMRI · Nociceptive stimulation · Anticipation · Electrical · Cerebellum · Pain matrix

Introduction

The experience of pain is produced by a complex neuronal system, which involves nociception, i.e., the signaling of tissue damage, as well as various cognitive and emotional components that are inherent to pain as a feeling [1]. One of the subcortical structures of the pain matrix, which is consistently activated after a peripheral nociceptive stimulus, is the cerebellum [2, 3]. Casey and coworkers were one of the first to identify cerebellar activity in the vermis after 50 °C heat stimuli of 5 s’ duration on the forearm using PET imaging [4]. Since then, many imaging studies have confirmed cerebellar activity after electrical, laser, capsaicin, or other types of nociceptive stimulation, mainly involving the vermis (lobules IV–V), the ipsilateral cortex (lobules IV–VI, Crus I), and the contralateral cortex (lobule VI, Crus I) of the cerebellum [3]. Animal studies that focused on the cerebellum in pain processing have suggested spinally projecting multisensory information from the skin, including tactile Aβ- and nociceptive Aδ- and C-fiber input [5–7]. However, the functional role of the cerebellum in pain processing remains largely unclear. The most widely accepted idea is that cerebellar activity is related
to the fine-tuning of the motor output when we experience pain, in order to protect it from further harm [8, 9]. However, a role in pain anticipation [10–12], in the inhibition of pain [13–17], and in perceiving pain induced in others [18] has also been suggested.

Taken together, the abovementioned data strongly indicate that the cerebellum is involved in pain perception, but there are still many questions about the precise nature of this involvement. One of these questions is whether the cerebellum is involved in modulating the processing of the incoming nociceptive signals or with the preparation and execution of a motor response to the nociceptive signal [19]. Another question regarding the role of the cerebellum in pain processing is whether it is instrumental in producing tasks that are localization independent, like pain inhibition and the production of warning signals, or whether it is involved in the precise localization and precise movement planning in response to the nociceptive signals. In the latter case, a more detailed processing of the pain signal will be necessary, that would require a precise somatotopical organization. The present fMRI study was initiated to investigate whether activation in the cerebellum after nociceptive stimulation is somatotopically organized. In addition, to learn more about the nature of cerebellar processing, we performed a separate study on the somatotopic activation pattern in the cerebellum during the anticipation of a nociceptive stimulus. Finally, we determined to which extend the cerebellar areas activated by nociceptive stimulation were separate from those activated by the anticipation of such stimuli.

Methods

The pain-only and pain anticipation experiments were approved by the medical ethics committee of Erasmus MC. All subjects had given written informed consent prior to the study.

Nociceptive Stimulation

Nociceptive stimuli were administered by transcutaneous electrical stimulation (5-Hz sine waves with adjustable intensity) using a Neurometer (Neurotron Inc., Baltimore, MD). The onset and offset of the stimuli were triggered manually.

Prior to scanning, subjects had to verbally rate the pain stimulation on a Numerical Rating Scale (NRS) from 1 (no pain) to 10 (unbearable pain). The intensity of the electrical stimulus was individually adjusted to elicit a stimulus that was rated as seven.

Pain-Only Experiment

Seventeen healthy subjects (10 males, 7 females; age range 18 to 29 years) participated in the pain study. One male subject was excluded due to technical problems.

We used a block design paradigm in which resting periods alternated with stimulation periods. During a stimulation period, the subject received a nociceptive stimulus to the right thumb and right big toe. Volunteers underwent five fMRI scans, each consisting of five blocks of nociceptive thumb stimulation and five blocks of nociceptive big toe stimulation. Stimulus blocks lasted 15 s; resting periods had three different durations: 10, 15, and 20 s. The order of the location of stimulation was random to limit a possible effect due to anticipation. Every scan started and ended with a resting period of 20 s.

Pain Anticipation Experiment

Seventeen healthy subjects (10 males, 7 females; age range 20 to 30 years) participated in the pain anticipation study. These included the same subjects that also took part in the pain-only experiment, except two that were replaced by two other volunteers. Three subjects were excluded (two males, one female) due to incomplete data.

We used a block design with three conditions: rest, nociceptive anticipation, and nociceptive stimulation. Identical to the pain-only experiment, the nociceptive stimulus was given on either the right thumb or the right big toe. Volunteers underwent four fMRI scans, each consisting of 4 cycles of blocks of rest and nociceptive anticipation, followed by nociceptive stimulation in both thumb and big toe. During nociceptive anticipation blocks, subjects were shown a red screen to indicate that nociceptive thumb stimulation was imminent and a blue screen to indicate the same for big toe stimulation. Resting periods had two different durations: 20 and 30 s; nociceptive stimulus blocks lasted 10 s; anticipation blocks had two different durations: 10 and 15 s. The order of the location of stimulation was random. Every scan started and ended with a resting period of 20 s.

Data Acquisition

Data were acquired on a 3T MRI scanner (HD platform, GE Healthcare, Milwaukee, WI) using a dedicated eight-channel head coil. For anatomical reference a 192-slice high-resolution three-dimensional inversion recovery (IR) fast spoiled gradient echo (FSPGR) T1-weighted image was acquired (parameters: slice thickness 1.6 mm with 0.8-mm overlap; repetition time (TR)/echo time (TE)/inversion time (TI) 10.3/2.0/300 ms; 18° flip angle; matrix 416 × 256 and field of view (FOV) 250 × 180 mm²). For the functional scans, a 32-slice single-shot T2*-weighted echo-planar imaging (EPI)
sequence sensitive to blood oxygenation level dependent (BOLD) contrast was used (parameters: slice thickness 3.0 mm and a 0.5-mm gap; TR/TE 2500/30 ms; 75° flip angle; 64 × 96 matrix with a FOV of 220 × 220 mm²; voxel sizes 3.0 × 3.4 × 2.9 mm³). Acquisition time of each scan was 6 min 50 s, including 10 s of dummy scans that were discarded.

**Data Analysis**

The functional imaging data were pre-processed and analyzed using the statistical parametric mapping toolbox (SPM 5, Wellcome Department of Cognitive Neurology, London, UK) run with MATLAB (version 7.8, Mathworks, Sherborn, MA). The anatomical scans were segmented into maps for white matter, grey matter, and cerebrospinal fluid. Normalization into the Montreal Neurological Institute (MNI) space was performed with parameters obtained during segmentation. The anatomical data were re-sliced into voxels of 1 × 1 × 1 mm³.

Functional scans were re-aligned, coregistered to the grey matter map, normalized with parameters obtained during segmentation, and re-sliced into 2 × 2 × 2-mm³ voxels and subsequently smoothed with a Gaussian kernel of 6-mm FWHM (full width at half maximum).

Single subject statistical analysis was performed with the general linear model. The fMRI time series was modeled as a series of event blocks convolved with a canonical hemodynamic response function. The two conditions of the pain study were nociceptive stimulation of the thumb and nociceptive stimulation of the big toe. There were four conditions of the anticipation experiment: anticipation of nociceptive stimulation of the thumb, anticipation of nociceptive stimulation of the big toe, actual nociceptive stimulation of the thumb, and actual nociceptive stimulation of the big toe. In addition, the time derivatives were modeled and movement parameters were included as regressors of no interest. The model was estimated with a high-pass filter with a cutoff period of 128 s. For each session, a T-contrast map was calculated for each condition, which was used in the second level, random effects analysis.

First, whole brain group results for thumb stimulation and toe stimulation were evaluated. For this analysis, an a priori statistical threshold of $p < 0.001$ at the voxel level (uncorrected) and family wise error rate (FWER) correction ($p < 0.05$) at the group level was used, resulting in a minimum cluster extent of 104 voxels. Anatomical structures were defined with the Talairach Daemon Labels atlas of the WFU Pick Atlas [20] in AAL (Automated Anatomical Labeling) [21], to aid with the description of the whole brain analysis results.

Further analysis focused on the cerebellum. The cerebellum was isolated and normalized to a cerebellum-specific template using the SUIT procedure, which provides a widely used analysis method for cerebellar fMRI data [9, 22, 23]. Subsequently, the unsmoothed functional data was modeled as described previous. Finally, the contrast images were smoothed with a Gaussian kernel of 6-mm FWHM. Anatomical structures were labeled according to the standard AAL atlas according to the MNI coordinates of the observed activation patterns.

To visualize differences in the activation induced by thumb and toe stimulation, activation maps were made for the thumb > toe and the toe > thumb stimulations. Differences in the activation induced by thumb and toe anticipation were visualized with activation maps of anticipation thumb > anticipation toe and anticipation toe > anticipation thumb. Conjunction analysis was performed to evaluate the overlap in activation after thumb and toe stimulation as well as thumb and toe anticipation. Comparisons used to study differences in activation included activation tables of thumb pain > thumb anticipation, thumb anticipation > thumb pain, toe pain > toe anticipation and toe anticipation > toe pain. Areas both active during anticipation of pain and sensation of pain were studied with conjunction analysis for thumb and toe. Overall conjunction was studied with a full factorial conjunction analysis with thumb pain, toe pain, thumb anticipation, and toe anticipation. For these analyses, an a priori threshold of $p < 0.005$ (uncorrected) and a minimum cluster extent of 24 voxels were used, based on a similar cerebellar fMRI study by Coombes and Misra [9], in which an empirical cluster-extent threshold of 192 mm³ was determined, corresponding to 24 2 × 2 × 2-mm voxels. The cluster-extent threshold of 24 voxels for the cerebellar subregions was based on the assumption that areas of true neural activity will tend to stimulate signal changes over contiguous voxels [24]. Anatomical structures were defined with the standard AAL atlas in AAL [21].

**Results**

All participants were stimulated on the right thumb and the right big toe. They generally described the stimulation as a painful sensation, a touch-like sensation was never reported. NRS pain scores were obtained after every session resulting in a mean score of 7.4 (± 0.7 SD) for the thumb and 7.7 (± 0.8 SD) for the big toe in the pain-only experiment and 7.5 (± 0.7 SD) for the thumb and 7.7 (± 0.7 SD) for the big toe in the pain anticipation experiment. The intensity of the stimulus remained constant during the experiment in most subjects. In a few occasions, when the subjective experience of pain increased, the stimulus intensity was lowered and vice versa.
General Brain Activation After Nociceptive Thumb or Toe Stimulation

A group analysis after stimulation of the thumb and big toe yielded activation in various areas of the brain, including the insula, the post central gyrus, and the cerebellum (Supplementary Table S1, Fig. 1). For the thumb, the post central gyrus was activated laterally on the contralateral side, while for the toe activation was found mainly medially. The insula and cingulate cortex were activated bilaterally. These findings are in general agreement with previous studies.

Cerebellar Activation During Nociceptive Thumb or Toe Stimulation

In the cerebellum, we found an activation during thumb and toe stimulation. In order to determine whether this activation was somatotopically organized, a comparison analysis was performed. In several areas, the activation was correlated significantly more with thumb stimulation, than with toe stimulation (thumb > toe, \( p < 0.005 \); Table 1, Fig. 2). These areas included a relatively large cluster in lobule VI ipsilaterally, in Crus II contralaterally, and a cluster in lobule VIII on the ipsilateral side of the stimulation. Activation that was correlated more with toe than with thumb stimulation (toe > thumb, \( p < 0.005 \); Table 1, Fig. 2) was located bilaterally in lobules VIII and IX and in lobules IV and V and contralaterally in lobule VI. We also examined which areas in the cerebellum were activated by thumb as well as toe stimulation, i.e., that were not somatotopically organized. For this purpose, we used a conjunction analysis to identify areas where the activation was significantly correlated with both toe and thumb stimulation (conjunction, \( p < 0.005 \); Table 1, Fig. 2). In this case, relatively strong activation was found in the vermis at the levels of lobule IV–VII and IX–X. In addition, activation was found bilaterally in Crus I.

Cerebellar Activation During Anticipation of Nociceptive Thumb or Toe Stimulation

In a separate experiment, we examined the activation in the cerebellum during the anticipation of a nociceptive stimulus to the thumb or the big toe. This experiment also included a nociceptive experience after the anticipation phase, which showed a similar activation pattern in the whole brain as we found in the first experiment (described previously), including activation clusters in the (contralateral) somatosensory cortex, the insula, the cingulate cortex, and the cerebellum (details not

---

Fig. 1 Group analysis maps overlaid on axial, coronal, and sagittal images from a standard MNI brain showing clusters of activation in the post central gyrus, insula, and cerebellum after nociceptive stimulation of the thumb and toe. Clusters are coded from dark red (\( T = 0 \)) to bright yellow (\( T = 10 \)) as indicated by the scale and L = left and R = right. A threshold of \( p = 0.001 \) was used (\( n = 16 \)).
| Cluster | $T$  | $x$  | $y$  | $z$  | Side | $N$ voxels | Structure            |
|---------|------|------|------|------|------|------------|----------------------|
| THUMB > TOE nociceptive stimulation |       |      |      |      |      |            |                      |
| 28      | 4.34 | −8   | −84  | −32  | Left | 28         | Cerebellum_Crus2     |
| 244     | 4.23 | 22   | −56  | −18  | Right | 227       | Cerebellum_6         |
|         |      |      |      |      | Right | 17        | Cerebellum_4_5       |
| 59      | 4.21 | −34  | −76  | −40  | Left  | 59         | Cerebellum_Crus2     |
| 10      | 3.51 | 46   | −76  | −46  | Right | 10         | Cerebellum_Crus2     |
| 44      | 3.44 | 22   | −62  | −56  | Right | 44         | Cerebellum_8         |
| 8       | 3.4  | 16   | −80  | −52  | Right | 7          | Cerebellum_7b        |
|         |      |      |      |      | Right | 1          | Cerebellum_Crus2     |
| 3       | 3.09 | 36   | −70  | −36  | Right | 3          | Cerebellum_Crus1     |
| 2       | 3.05 | 38   | −76  | −34  | Right | 2          | Cerebellum_Crus1     |
| TOE > THUMB nociceptive stimulation |       |      |      |      |      |            |                      |
| 33      | 4.35 | 12   | −56  | −58  | Right | 20         | Cerebellum_9         |
|         |      |      |      |      | Right | 13         | Cerebellum_8         |
| 47      | 4.01 | 24   | −38  | −30  | Right | 41         | Cerebellum_4_5       |
|         |      |      |      |      | Right | 6          | Cerebellum_3         |
| 31      | 3.92 | −34  | −42  | −34  | Left  | 23         | Cerebellum_6         |
|         |      |      |      |      | Left  | 8          | Cerebellum_Crus1     |
| 42      | 3.79 | −18  | −54  | −22  | Left  | 27         | Cerebellum_6         |
|         |      |      |      |      | Left  | 15         | Cerebellum_4_5       |
| 53      | 3.54 | −18  | −66  | −52  | Left  | 53         | Cerebellum_8         |
| 8       | 3.4  | −4   | −56  | −36  | –      | 5          | Vermis_9             |
|         |      |      |      |      | –      | 3          | Cerebellum_9         |
| CONJUNCTION nociceptive stimulation |       |      |      |      |      |            |                      |
| 175     | 3.84 | 2    | −66  | −14  | –      | 104        | Vermis_6             |
|         |      |      |      |      | –      | 38         | Vermis_4_5           |
|         |      |      |      |      | –      | 26         | Vermis_7             |
|         |      |      |      |      | Right  | 5          | Cerebellum_Crus1     |
|         |      |      |      |      | Right  | 2          | Cerebellum_6         |
| 82      | 3.83 | 0    | −54  | −32  | –      | 46         | Vermis_9             |
|         |      |      |      |      | –      | 23         | Vermis_10            |
|         |      |      |      |      | Left  | 7          | Cerebellum_9         |
|         |      |      |      |      | Right  | 6          | Cerebellum_9         |
| 4       | 3.48 | −10  | −34  | −10  | Left  | 4          | Cerebellum_4_5       |
| 70      | 3.42 | 30   | −78  | −30  | Right  | 52         | Cerebellum_Crus1     |
|         |      |      |      |      | Right  | 18         | Cerebellum_6         |
| 28      | 3.38 | 46   | −70  | −32  | Right  | 28         | Cerebellum_Crus1     |
| 27      | 3.18 | −28  | −70  | −32  | Left  | 27         | Cerebellum_Crus1     |
| 4       | 3.14 | −34  | −64  | −52  | Left  | 7          | Cerebellum_8         |
| 29      | 3.1  | −42  | −76  | −34  | Left  | 27         | Cerebellum_Crus1     |
|         |      |      |      |      | Left  | 2          | Cerebellum_Crus2     |
| 11      | 3.07 | −52  | −64  | −34  | Left  | 11         | Cerebellum_Crus1     |
| 4       | 2.84 | −50  | −60  | −24  | Left  | 4          | Cerebellum_Crus1     |
| 3       | 2.79 | 50   | −62  | −28  | Right  | 3          | Cerebellum_Crus1     |

Cluster size (Cluster) in voxels, $T_{\text{max}}$ ($T$) and its MNI coordinates ($x$, $y$, $z$), and side at which the activation occurs (Side) are given followed by the specification of the number of voxels ($N$ voxels) per structure (Structure) within that cluster. Significant areas with an uncorrected threshold of $p = 0.005$ and a cluster-extent of at least 24 voxels are shown in italic, while areas containing subthreshold clusters are shown in regular face ($n = 16$).
The detailed analysis of the cerebellar contrasts was similar to the results of the first experiment (compare Supplementary Table S2 with Table 1).

We then examined cerebellar activation during the anticipation phase. We found a bilateral activation in the cerebellum during the anticipation of nociceptive stimulation of the thumb or toe. In order to determine whether this organization was somatotopically organized, a comparison analysis was performed. We identified areas that were correlated significantly more with the anticipation of thumb stimulation, than with that of toe stimulation (thumb > toe, \( p < 0.005 \); Table 2, Fig. 3). The activated areas were in lobules VI and VIII ipsilaterally. No clusters were identified that were correlated significantly more with anticipation of nociceptive toe than thumb stimulation, after cluster-extent-based thresholding (toe > thumb, \( p < 0.005 \); Table 2, Fig. 3). The activated areas were in lobules VI and VIII ipsilaterally. No clusters were identified that were correlated significantly more with anticipation of nociceptive toe than thumb stimulation, after cluster-extent-based thresholding (toe > thumb, \( p < 0.005 \); Table 2, Fig. 3). The activated areas were in lobules VI and VIII ipsilaterally. No clusters were identified that were correlated significantly more with anticipation of nociceptive toe than thumb stimulation, after cluster-extent-based thresholding (toe > thumb, \( p < 0.005 \); Table 2, Fig. 3). The activated areas were in lobules VI and VIII ipsilaterally. No clusters were identified that were correlated significantly more with anticipation of nociceptive toe than thumb stimulation, after cluster-extent-based thresholding (toe > thumb, \( p < 0.005 \); Table 2, Fig. 3).

Comparing Cerebellar Activation During the Experience and the Anticipation of Nociceptive Thumb or Toe Stimulation

Since the anticipation experiment included a pain stimulus immediately following the anticipation phase, we were able to compare pain anticipation and pain experience for thumb and toe. Areas in the cerebellum that were activated by pain experience on the thumb but not by pain anticipation were limited in volume and found predominantly in lobule VIII (thumb pain > thumb anticipation, \( p < 0.005 \); Table 3). In contrast, the total area activated by pain anticipation but not by pain experience was much larger, including lobules IV–V, VI bilaterally, and the vermis (thumb anticipation > thumb pain, \( p < 0.005 \); Table 3). However, the largest area was found in the conjunction analysis of the thumb (Crus I and lobule VI on both sides (conjunction thumb pain and pain anticipation, \( p < 0.005 \); Table 3), showing that this area was activated by both pain experience and pain anticipation. For toe stimulation, similar findings were obtained: relatively small areas in the cerebellum were activated by nociceptive toe stimulation and anticipation specifically, while the conjunction analysis showed that a much larger area, located in Crus I and lobule VI bilaterally, was activated by both (\( p < 0.005 \); Table 4). Since the conjunction analysis of the thumb and toe showed very similar areas of activation, we performed an additional conjunction analysis (\( p < 0.005 \); Table 5) to identify the areas of the cerebellum that were activated by the nociceptive thumb stimulation, the nociceptive toe stimulation, the anticipation of nociceptive thumb stimulation, and the anticipation of nociceptive toe stimulation. This analysis confirmed that the same areas in Crus I and lobule VI, mostly on the contra-lateral side, were activated by each of these four types of stimulation. A schematic representation of the results is presented in Fig. 4.

Discussion

In this fMRI study, we have investigated the activation in the cerebellum after nociceptive stimulation of the thumb and the toe and after the anticipation of these stimuli. We found that

1. Nociceptive stimulation of the toe and the thumb activated separate areas in the cerebellum, while areas that were activated by toe as well as thumb stimulation were of similar size. This finding shows that the activation of the cerebellum is partly organized in a somatotopic fashion.
Table 2  Group analysis of differences and overlap in activation in the cerebellum after anticipation of nociceptive stimulation of thumb and toe

| Cluster | $T$  | $x$  | $y$  | $z$  | Side  | $N$ voxels | Structure                  |
|---------|------|------|------|------|-------|------------|-----------------------------|
| THUMB > TOE anticipation of pain | 2    | 6.38 | −18  | −42  | −4    | Left       | 2 Cerebellum_4.5            |
|         | 47   | 4.46 | 22   | −64  | −58   | Right      | 47 Cerebellum_8              |
|         | 105  | 4.13 | 38   | −48  | −28   | Right      | 105 Cerebellum_6             |
|         | 22   | 3.92 | 10   | −66  | −8    | Right      | 9 Cerebellum_6               |
|         |      |      |      |      | Right  | 6        | Cerebellum_4.5               |
|         |      |      |      |      | −5     | Cerebellum_4.5               |
|         |      |      |      |      | −2     | Cerebellum_6               |
|         | 4    | 3.54 | 12   | −50  | −12   | Right      | 4 Cerebellum_4.5             |
|         | 2    | 3.38 | 20   | −72  | −46   | Right      | 2 Cerebellum_8               |
| TOE > THUMB anticipation of pain | 8    | 3.82 | 52   | −72  | −36   | Right      | 8 Cerebellum_Crus1           |
|         | 1    | 3.56 | −32  | −42  | −44   | Left       | 1 Cerebellum_8               |
| CONJUNCTION anticipation of pain | 1036 | 4.99 | −50  | −60  | −28   | Left       | 831 Cerebellum_Crus1         |
|         |      |      |      |      | Left   | 196       | Cerebellum_6               |
|         | 126  | 4.08 | 34   | −80  | −24   | Right      | 114 Cerebellum_Crus1         |
|         |      |      |      |      | Right  | 12        | Cerebellum_6               |
|         | 51   | 3.73 | 34   | −40  | −26   | Right      | 39 Cerebellum_6             |
|         |      |      |      |      | Right  | 12        | Cerebellum_4.5             |
|         | 74   | 3.43 | 38   | −66  | −22   | Right      | 58 Cerebellum_6             |
|         |      |      |      |      | Right  | 16        | Cerebellum_Crus1           |
|         | 15   | 3.09 | 0    | −54  | −8    | −5     | 15 Vermis_4.5             |
|         | 15   | 2.97 | −2   | −48  | −22   | −5     | 5 Vermis_3               |
|         |      |      |      |      | Left   | 5         | Cerebellum_3               |
|         |      |      |      |      | −4     | Cerebellum_4.5               |
|         |      |      |      |      | −1     | Cerebellum_1.2               |
|         | 2    | 2.85 | 10   | −48  | −36   | Right      | 2 Cerebellum_9             |
|         | 1    | 2.82 | 14   | −30  | −24   | Right      | 1 Cerebellum_3               |

Cluster size (Cluster) in voxels, $T$ max ($T$) and its MNI coordinates ($x$, $y$, $z$), and side at which the activation occurs (Side) are given followed by the specification of the number of voxels ($N$ voxels) per structure (Structure) within that cluster. Significant areas with an uncorrected threshold of $p = 0.005$ and a cluster-extent of at least 24 voxels are shown in italic, while areas containing subthreshold clusters are shown in regular face ($n = 14$).

Fig. 3  Group analysis of cerebellar data displayed on the SUIT template. The upper two rows show stimulus specific differences in activation after anticipation of nociceptive stimulation of thumb or toe found with the thumb > toe contrast (first row) and toe > thumb contrast (second row). Conjunction results are shown in the third row and represent the overlap of activation found after nociceptive stimulation of thumb or toe. Slices are arranged from inferior ($Z = −56$) to superior ($Z = −8$) and L = left and R = right. A threshold of $p = 0.005$ was used ($n = 14$).
2. Anticipation of nociceptive stimuli on the thumb or toe mainly activated the same areas of the cerebellum. Much smaller areas were activated by the anticipation of thumb pain only and none by the anticipation of toe pain only.

3. When comparing the cerebellar areas involved in pain experience and the anticipation of that experience, we found a relatively large area, mainly in Crus I and lobule VI on the contralateral side, that was activated by

| Cluster | THUMB nociceptive stimulation > THUMB anticipation of pain |  |
|---------|-------------------------------------------------------------|---|
| 77      | 3.94 26 –66 –52 Right 77 Cerebellum_8                     |   |
| 23      | 3.83 16 –76 –54 Right 20 Cerebellum_8                     |   |
| 7       | 3.49 –8 –88 –30 Left 7 Cerebellum_Crusb2                  |   |
| 15      | 3.47 –22 –78 –50 Left 15 Cerebellum_Crusb2                |   |
| 3       | 3.11 20 –64 –20 Right 3 Cerebellum_6                      |   |
| 5       | 3.1 26 –58 –22 Right 5 Cerebellum_6                       |   |
| 1       | 3.03 16 –68 –18 Right 1 Cerebellum_6                      |   |

| Cluster | THUMB anticipation of pain > THUMB nociceptive stimulation |  |
|---------|-----------------------------------------------------------|---|
| 578     | 5.78 –24 –48 –28 Left 333 Cerebellum_4_5                 |   |
| 229     | 5.67 26 –36 –22 Right 71 Cerebellum_Crusb1               |   |
| 276     | 5.15 8 –42 –14 –115 Vermis_3                             |   |
| 31      | 4.53 –14 –44 0 Left 31 Cerebellum_4_5                    |   |
| 2       | 3.73 –8 –66 –32 Left 2 Cerebellum_8                      |   |
| 15      | 3.57 –12 –52 –56 Left 15 Cerebellum_9                    |   |
| 18      | 3.42 –2 –42 –36 –13 Vermis_10                            |   |
| 6       | 3.29 52 –60 –50 Right 6 Cerebellum_Crusb2                |   |
| 2       | 3.21 14 –30 –24 Right 2 Cerebellum_3                     |   |
| 4       | 3.14 –12 –32 –22 Left 4 Cerebellum_3                     |   |
| 3       | 3.12 52 –70 –40 Right 3 Cerebellum_Crusb2                |   |
| 1       | 3.03 14 –48 –50 Right 1 Cerebellum_9                     |   |

| Cluster | CONJUNCTION THUMB nociceptive stimulation and anticipation of pain |  |
|---------|-----------------------------------------------------------------|---|
| 269     | 5.04 38 –80 –24 Right 154 Cerebellum_Crusb1                    |   |
| 717     | 4.08 –42 –72 –20 Left 617 Cerebellum_Crusb1                   |   |
| 22      | 3.35 22 –78 –28 Right 22 Cerebellum_Crusb1                    |   |
| 6       | 2.84 2 –56 –6 –6 Vermis_4_5                                   |   |
| 1       | 2.8 36 –60 –22 Right 1 Cerebellum_6                           |   |

Cluster size (Cluster) in voxels, \( T \) \( x \) and \( y \) \( z \), and side at which the activation occurs (Side) are given followed by the specification of the number of voxels (\( N \) voxels) per structure (Structure) within that cluster. Significant areas with an uncorrected threshold of \( p = 0.005 \) and a cluster-extent of at least 24 voxels are shown in italic, while areas containing subthreshold clusters are shown in regular face (\( n = 14 \))
nociceptive stimulation as well as nociceptive anticipation, both for the thumb and for the toe. This indicates that the anticipation of a nociceptive stimulus is processed by the same cerebellar areas as the actual nociceptive stimulus and that this occurs for the thumb and toe alike, i.e., without a substantial somatotopic organization.

General Aspects of the Experimental Setup and Analysis

In this study, we have used electrical stimulation, applied with a Neurometer [25, 26]. In rats, transcutaneous stimulation with 5-Hz sine waves stimulates mainly C-fibers [27, 28]. This suggests that this type of stimulation preferentially induces pain and not touch. Our subjects, who all declared that the stimuli induced a clear sensation of pain rather than an innocuous sensation like touch, corroborated this notion. We therefore conclude that our stimulation protocol preferentially activated nociceptive fibers.

A potential weakness of our study is that somatotopy was inferred based on the stimulation of only two body parts. This is especially true, since nociceptive stimulation may evoke confounding factors like perceived threat and unpleasantness that may differ between thumb pain and toe pain.

It may be argued that the present statistical analysis, which was chosen beforehand based on literature describing similar experiments and using similar group sizes in the human somatosensory cortex and cerebellum [9, 29], might be too liberal. An uncorrected statistical threshold of 0.005 in the cerebellum could indeed lead to an increase in false positives (type I errors). Future studies might consider using a stricter uncorrected threshold and/or voxel-based thresholding. These alternatives have, however, the drawback of an increased number

| Cluster | \( T \) | \( x \) | \( y \) | \( z \) | Side | \( N \) voxels | Structure |
|---------|-----|------|------|------|-----|-----------|-----------|
| TOE nociceptive stimulation > TOE anticipation of pain | 75 | 5.78 | 10 | -66 | -6 | Right | 28 | Vermis_4_5 |
| | | | | | | Left | 14 | Cerebellum_4_5 |
| | 6 | 3.44 | -24 | -78 | -50 | Left | 6 | Cerebellum_7b |
| | 2 | 3.38 | 36 | -84 | -42 | Right | 2 | Cerebellum_Crus2 |
| | 4 | 3.21 | 0 | -54 | -6 | – | 4 | Vermis_4_5 |
| | 1 | 3.03 | 16 | -42 | -22 | Right | 1 | Cerebellum_4_5 |
| TOE anticipation of pain > TOE nociceptive stimulation | 112 | 5.04 | -22 | -40 | -30 | Left | 106 | Cerebellum_4_5 |
| | | | | | | Left | 6 | Cerebellum_6 |
| | 31 | 4.37 | -12 | -32 | -22 | Left | 16 | Cerebellum_3 |
| | | | | | | – | 15 | Vermis_1_2 |
| | 10 | 3.97 | 32 | -42 | -24 | Right | 10 | Cerebellum_6 |
| | 2 | 3.41 | 26 | -32 | -24 | Right | 2 | Cerebellum_4_5 |
| | 1 | 3.1 | -38 | -42 | -34 | Left | 1 | Cerebellum_Crus1 |
| CONJUNCTION TOE nociceptive stimulation and anticipation of pain | 679 | 4.84 | -26 | -80 | -20 | Left | 526 | Cerebellum_Crus1 |
| | | | | | | Left | 125 | Cerebellum_6 |
| | 138 | 4.12 | 46 | -62 | -24 | Right | 75 | Cerebellum_Crus1 |
| | | | | | | Right | 63 | Cerebellum_6 |
| | 102 | 3.89 | 34 | -80 | -24 | Right | 92 | Cerebellum_Crus1 |
| | | | | | | Right | 10 | Cerebellum_6 |
| | 10 | 3.61 | 26 | -38 | -30 | Right | 10 | Cerebellum_4_5 |
| | 14 | 3.33 | 14 | -80 | -24 | Right | 14 | Cerebellum_Crus1 |
| | 4 | 3.25 | 10 | -44 | -24 | Right | 4 | Cerebellum_3 |
| | 5 | 2.9 | 0 | -54 | -6 | – | 5 | Vermis_4_5 |

Cluster size (Cluster) in voxels, \( T \) \( \text{max} (T) \) and its MNI coordinates \((x, y, z)\), and side at which the activation occurs (Side) are given followed by the specification of the number of voxels \((N \text{ voxels})\) per structure (Structure) within that cluster. Significant areas with an uncorrected threshold of \( p = 0.005 \) and a cluster-extent of at least 24 voxels are shown in italic, while areas containing subthreshold clusters are shown in regular face \((n = 14)\).
of false negatives (type II errors), which might be a serious problem in experimental manipulations that affect the BOLD signal only slightly. Instead, we used a very conservative cluster-extent threshold of 24 voxels based on Coombes and Msra [9]. Alternatively, the present experiment may provide cerebellar regions that would allow future studies to use specific regions of interest rather than whole brain/whole cerebellum analyses, thereby drastically decreasing the number of voxels to be analyzed.

The Activity Pattern of the Cerebellum During Nociceptive Thumb and Toe Stimulation

Nociceptive stimulation of the thumb and toe resulted in a clear activation of the pain matrix in the brain, including the insula, post central gyrus, cingulate gyrus, and cerebellum, which is in agreement with previous studies (see [30, 31] for review). Specifically, in the primary somatosensory cortex, we found an activation in the hand area after thumb stimulation and in the foot area after toe stimulation, as shown previously for hand and foot stimulation with capsaicin [32], electrical stimuli [33], or nociceptive laser stimulation [34]. These findings validated our experimental setup, allowing for a detailed analysis of the activation patterns in the cerebellum.

We have combined thumb and toe stimulation within one experiment, in order to keep circumstances of the thumb and toe stimulation, like arousal, attention, and visceral activation. This will allow for a comparison analysis in which it can be determined for each voxel, whether its activity is correlated more with toe than with thumb activation, or with both. These comparisons indicate that there is indeed a somatotopic organization of the cerebellum for nociceptive stimuli originating from different body parts. At this point, it should be stressed that the somatotopic organization of the cerebellum for sensory and motor input has been extensively studied in the past century and is still under much debate (see [35, 36] for review). With respect to fMRI studies, like the present study, results are mapped to cerebellar lobules, while the cerebellum is functionally organized in longitudinal zones that run across the various lobules. These zones cannot be reliably identified with fMRI, which may lead to a fractured (patchy) somatotopy [35]. Furthermore, in functional studies, it is often difficult to disentangle activities related to sensory input and (subsequent) motor output. This makes it difficult to compare the results of our study with other studies in detailed anatomical terms. Nevertheless, our finding that some areas in the cerebellum are activated by both thumb and toe nociceptive stimulation, while other areas are separately activated, suggesting a somatotopic organization, remains valid and should be interpreted in general terms. One subdivision from the literature, stating that medially located parts of the cerebellum are preferentially involved in basic sensory-motor performances, while cognitive tasks tend to engage lateral cerebellar regions, i.e., lobule VI and lobule VII (Crus I, Crus II, and VIIb) [37], seems in general agreement with our findings, since we find activations both medially and laterally in the cerebellum, as is to be expected when using pain stimuli which, by their nature, have both sensory-motor as well as emotional-cognitive aspects.

While the thumb and toe areas are separate, they are to a large extent located in the same lobules of the cerebellum, i.e., lobules VI and VIII of the posterior cerebellum and lobules IV–V in the anterior cerebellum. Crus II of the cerebellum is an exception as it is activated only by

| Cluster | T   | x   | y   | z   | Side | N voxels | Structure       |
|---------|-----|-----|-----|-----|------|----------|----------------|
| CONJUNCTION nociceptive stimulation and anticipation of pain, thumb and toe |
| 586     | 3.84| −36 | −62 | −24 | Left  | 487      | Cerebellum_CrusI |
|         |     |     |     |     | Left  | 99       | Cerebellum_6    |
| 75      | 3.81| 36  | −80 | −24 | Right | 68       | Cerebellum_CrusI |
|         |     |     |     |     | Right | 7        | Cerebellum_6    |
| 37      | 3.47| 46  | −60 | −24 | Right | 22       | Cerebellum_CrusI |
|         |     |     |     |     | Right | 15       | Cerebellum_6    |
| 8       | 2.89| 24  | −74 | −28 | Right | 8        | Cerebellum_CrusI |
| 2       | 2.75| 2   | −54 | −6  | –     | 2        | Vermis_4_5      |
| 2       | 2.72| 40  | −52 | −30 | Right | 1        | Cerebellum_6    |
| 1       | 2.71| 18  | −80 | −26 | Right | 1        | Cerebellum_CrusI |

Cluster size (Cluster) in voxels, T max (T) and its MNI coordinates (x, y, z), and side at which the activation occurs (Side) are given followed by the specification of the number of voxels (N voxels) per structure (Structure) within that cluster. Significant areas with an uncorrected threshold of p = 0.005 and a cluster-extent of at least 24 voxels are shown in italic, while areas containing subthreshold clusters are shown in regular face (n = 14)
thumb stimulation. Activation of lobule VI has been reported by several other studies that have used nociceptive temperature stimulation [38, 39]. Lobules IV–VI have been described as being involved in sensory-motor processing, and lobule VIIIb in secondary sensory processing [40]. Thus, our findings suggest that the activation in the hemispheres of lobuli VI and VIII as well as lobules IV–V and Crus II is organized in a somatotopic manner, at least with respect to nociceptive stimulation of the thumb and the toe.

When the areas in the cerebellum that were activated by thumb as well as by toe stimulation were examined, we found that most clusters of activation were located in the vermis of lobuli IV–V, VI, VII, IX, and X; bilaterally in Crus I; and a small cluster in lobule VI ipsilaterally, i.e., mainly in the posterior cerebellum. This finding shows that nociceptive stimuli on the thumb and toe also activate areas of the cerebellum in a non-somatotopic manner. Activation of the vermis has also been shown in other studies [41] using peripheral nociceptive stimulation, although these studies have demonstrated
activation primarily in the vermis of the anterior cerebellum, while we found that the bulk of the activation was located in the vermis of the posterior cerebellum. The posterior vermis has been described as the limbic part of the cerebellum [42]. It makes sense for nociceptive input, which is a highly emotional type of information, to reach the limbic area of the cerebellum. Crus I, which is considered part of the cognitive cerebellum [42], is also substantially activated by both the toe and the thumb. Thus, it seems that the somatotopically organized input to the cerebellum after nociceptive stimulation preferentially activates sensory-motor areas of the posterior cerebellum, while the same input also activates cognitive and limbic parts of the cerebellum in a non-somatotopic manner.

The Activity Pattern of the Cerebellum During the Anticipation of Nociceptive Thumb and Toe Stimulation

Anticipation of thumb stimulation, but not anticipation of toe stimulation, activated limited areas of lobules VI and VIII ipsilaterally, as after the pain-only stimulation. Remarkably, the largest clusters of activation were found in the conjunction analysis, which seems to indicate that the anticipation of a nociceptive stimulus is a situation that the cerebellum is processing primarily in a non-somatotopic manner.

Comparing Pain and the Anticipation of Pain

Anticipation of a nociceptive stimulus was first shown to lead to cerebellar activation by Ploghaus et al. [41]. They found that nociceptive heat stimuli, when compared to a warmth stimulus, localized bilaterally around the midline in the anterior cerebellum, while the anticipation of nociceptive heat localized ipsilaterally in the posterior cerebellum, thus showing differential localization of pain and its anticipation within the cerebellum. In contrast, our conjunction analysis makes clear that the areas in the cerebellum that were activated by an actual pain stimulus are also activated by the anticipation of such a stimulus and that these areas, i.e., Crus I and lobule VI, were located bilaterally in the posterior cerebellum. This is in accordance with findings in other areas of the brain, both at the cortical and at the subcortical level, which showed that the anticipation of pain was found to activate both the same areas as activated during actual pain perception as well as other areas, possibly involved in preparing for the expected nociceptive stimulus [43]. Furthermore, a study [44] comparing activations in the cerebellum by aversive pictures with activations of a heat pain stimulus also showed that Crus I and lobule VI (and additionally lobule VIIb) were activated by both stimuli. Our finding that Crus I and lobule VI become activated by both the anticipation of a specific pain stimulus and the actual stimulus, irrespective of the localization of that stimulus, would fit very well with the idea that these cerebellar areas are involved in processing general aversive emotions, like pain. Interestingly, emotions like (the expectation of) reward, which are on the opposite side of aversion in the emotional spectrum, are also processed in the cerebellum [45, 46]. Recordings from granule cells in lobules V and VI of the mouse cerebellum showed an expectation of reward-related activity even independent of any (preparatory) motor activity [46].

In conclusion, many fMRI studies have shown activation of the cerebellum in pain processing [47]. In these studies, it remained unclear how the nociceptive information reaches the cerebellum. Anatomical and physiological data show that the cerebellum receives this information directly, through connections with the spinal cord, as well as indirectly through its connections with the cortex. It seems likely that the input that is organized in a somatotopical manner reflects the direct input from the spinal cord, while the non-somatotopically activated parts of the cerebellum receive more general contextual information, like (expected) nociception, indirectly through cortical and subcortical connections. These parts are possibly involved in processing general emotions, like aversion and reward, thus allowing (expected) emotional states to affect sensory-motor processing in the cerebellum. The findings in this study seem to support this notion.

Acknowledgements A.E. Smit is a recipient of a Grant from Pijnkennis Centrum Rotterdam.

The authors thank Dr. M. Smits, Dept. of Radiology Erasmus MC, for the advice on the analysis of the data.

Author Contributions Contributions from the individual authors are summarized as follows:
- MW has performed the experiments, has analyzed the data, and has written the paper.
- AS has analyzed the data.
- JJ has analyzed the data and has written the paper.
- DT has written the paper.
- JG has conceived the experiment, has analyzed the data, and has written the paper.
- JJ has conceived the experiment, has analyzed the data, and has written the paper.

Compliance with Ethical Standards

All experiments were in accordance with the ethical standards of the institutional research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. Subjects have given written informed consent prior to the study.

Conflict of Interest The authors declare that they have no conflicts of interest.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.
References

1. Tracey I, Mantyh PW. The cerebral signature for pain perception and its modulation. Neuron. 2007;55:377–91. https://doi.org/10.1016/j.neuron.2007.07.012.

2. Saab CY, Willis WD. The cerebellum: organization, functions and its role in nociception. Brain Res Brain Res Rev. 2003;42:85–95.

3. Moulton EA, Schmahmann JD, Becerra L, Borsook D. The cerebellum and pain: passive integrator or active participant? Brain Res Rev. 2010;65:14–27. https://doi.org/10.1016/j.brainresrev.2010.05.005.

4. Casey KL, Minoshima S, Berger KL, Koepp RA, Morrow TJ, Frey KA. Positron emission tomographic analysis of cerebellar structures activated specifically by repetitive noxious heat stimuli. J Neurophysiol. 1994;71:802–7.

5. Ekerot CF, Gustavsson P, Oscarsson O, Schouenborg J. Climbing fibres projecting to cat cerebellar anterior lobe activated by cutaneous A and C fibres. J Physiol. 1987;386:529–38.

6. Ekerot CF, Oscarsson O, Schouenborg J. Stimulation of cutaneous nociceptive C fibres causing tonic and synchronous activity in climbing fibres. J Physiol. 1987;386:539–46.

7. Wu J, Chen PX. Cerebellar evoked potential elicited by stimulation of C-fiber in saphenous nerve of cat. Brain Res. 1990;522:144–6.

8. Wiech K, SSPM, Driver J, et al. Modulation of pain processing in hyperalgesia by cognitive demand. NeuroImage. 2005;27:59–69. https://doi.org/10.1016/j.neuroimage.2005.03.044.

9. Coomes SA, Misra G. Pain and motor processing in the human cerebellum. Pain. 2016;157:117–27. https://doi.org/10.1097/j.pain.0000000000000337.

10. Ploghaus A, Tracey I, Clare S, Gati JS, Rawlins JN, Matthews PM. Learning about pain: the neural substrate of the prediction error for aversive events. PNAS. 2000;97:9281–6. https://doi.org/10.1073/pnas.160266497.

11. Ploghaus A, Becerra L, Borras C, Borsook D. Neural circuitry underlyning pain modulation: expectation, hypnosis, placebo. Trends Cogn Sci. 2003;7:197–200.

12. Keltner JR, Furst A, Fan C, Redfern R, Inglis B, Fields HL. Isolating the modulatory effect of expectation on pain transmission: a functional magnetic resonance imaging study. J Neurosci. 2006;26:4437–43. https://doi.org/10.1523/JNEUROSCI.4463-05.2006.

13. Siegel P, Wepsic JG. Alteration of nociception by stimulation of cerebellar structures in the monkey. Physiol Behav. 1974;13:189–94.

14. Dey PK, Ray AK. Anterior cerebellum as a site for morphine analgesia and post-stimulation analgesia. Indian J Physiol Pharmacol. 1982;26:3–12.

15. Saab CY, Willis WD. Cerebellar stimulation modulates the intensity of a visceral nociceptive reflex in the rat. Exp Brain Res. 2002;146:117–21. https://doi.org/10.1007/s00221-002-1107-8.

16. Hagains CE, Senapati AK, Huntington PJ, He JW, Peng YB. Inhibition of spinal cord dorsal horn neuronal activity by electrical stimulation of the cerebellar cortex. J Neurophysiol. 2011;106:2515–22. https://doi.org/10.1152/jn.00719.2010.

17. Rucheweeyh R, Kuhnel M, Filippopulos F, Blum B, Eggert T, Straube A. Altered experimental pain perception after cerebellar infarction. Pain. 2014;155:1303–12. https://doi.org/10.1016/j.pain.2014.04.006.

18. Moriguichi Y, Decety J, Ohnishi T, Maeda M, Mori T, Nemoto K, et al. Empathy and judging other’s pain: an fMRI study of alexithymia. Cereb Cortex. 2007;17:2223–34. https://doi.org/10.1093/cercor/bhl130.

19. Perini I, Bergstrand S, Morrison I. Where pain meets action in the human brain. J Neurosci. 2013;33:15930–9. https://doi.org/10.1523/JNEUROSCI.3135-12.2013.

20. Malgian JA, Laurienti PJ, Kraft RA, Burdette JH. An automated method for neuroatomic and cytarchitectonic atlas-based interrogation of fMRI data sets. NeuroImage. 2003;19:1233–9.

21. Tzourio-Mazoyer N, Landeau B, Papathanassiou D, Crivello F, Etard O, Delcroix N, et al. Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. NeuroImage. 2002;15:273–89. https://doi.org/10.1006/nimg.2001.0978.

22. Diedrichsen J. A spatially unbiased atlas template of the human cerebellum. NeuroImage. 2006;33:127–38. https://doi.org/10.1016/j.neuroimage.2006.05.056.

23. Diedrichsen J, Balsters JH, Flavell J, Cussans E, Ramnani N. A probabilistic MR atlas of the human cerebellum. NeuroImage. 2009;46:39–46. https://doi.org/10.1016/j.neuroimage.2009.01.045.

24. Masson EA, Boulton AJ. The Neuroemr: validation and comparison with conventional tests for diabetic neuropathy. Diabet Med. 1991;8:Spec No: S63–6.

25. Pitei DL, Watkins PJ, Stevens MJ, Edmonds ME. The value of the Neuroemr in assessing diabetic neuropathy by measurement of the current perception threshold. Diabet Med. 1994;11:872–6.

26. Masson EA, Boulton AJ. The Neuroemr: validation and comparison with conventional tests for diabetic neuropathy. Diabet Med. 1991;8:Spec No: S63–6.

27. Katims JJ, Naviasky EH, Ng LK, Rendell M, Bleecker ML. New screening device for assessment of peripheral neuropathy: J Occup Med. 1986;28:1219–21.

28. Koga K, Furue H, Rashid MH, Takaki A, Katafuchi T, Yoshimura M. Selective activation of primary afferent fibers evaluated by single-wave electrical stimulation. Mol Pain. 2005;1:13. https://doi.org/10.1016/j.molpain.2005.03.004.

29. Martuzzi R, van der Zwaag W, Farhoujt J, Gruetter R, Blanke O. Human finger somatotopy in areas 3b, 1, and 2: a 7T fMRI study using a natural stimulus. Hum Brain Mapp. 2014;35:213–26. https://doi.org/10.1002/hbm.22172.

30. Peyron R, Laurent B, Garcia-Larrea L. Functional imaging of brain responses to pain. A review and meta-analysis (2000). Neuropsychol Clin. 2000;30:263–88.

31. Apkarian AV, Bushnell MC, Treede RD, Zubieta JK. Human brain mechanisms of pain perception and regulation in health and disease. Eur J Pain. 2005;9:463–84. https://doi.org/10.1016/j.ejpain.2004.11.001.

32. Andersson JL, Lilja A, Hartvig P, Langstrom B, Gordh T, Handwerker H, et al. Somatotopic organization along the central sulcus, for pain localization in humans, as revealed by positron emission tomography. Exp Brain Res. 1997;117:192–9.

33. Kurth R, Illingringer K, Curio G, Wolf KJ, Krause T, Repentin J, et al. fMRI shows multiple somatotopic digit representations in human primary somatosensory cortex. Neuroreport. 2000;11:1487–91.

34. Bingel U, Lorenz J, Glauche V, Knab R, Glascher J, Weiller C, et al. Somatotopic organization of human somatosensory cortices for pain: a single trial fMRI study. NeuroImage. 2004;23:224–32. https://doi.org/10.1016/j.neuroimage.2004.05.021.

35. Manni E, Petrosini L. A century of cerebellar somatotopy: a debated organization. Front Syst Neurosci. 2014;8:227. https://doi.org/10.3389/fnsys.2014.00227.

36. Stoodley CJ. The cerebellum and cognition: evidence from functional imaging studies. Cerebellum. 2012;11:352–65. https://doi.org/10.1007/s12311-011-0260-7.
38. Helmchen C, Mohr C, Erdmann C, Petersen D, Nitschke MF. Differential cerebellar activation related to perceived pain intensity during noxious thermal stimulation in humans: a functional magnetic resonance imaging study. Neurosci Lett. 2003;335:202–6.

39. Borsook D, Moulton EA, Tully S, Schmahmann JD, Becerra L. Human cerebellar responses to brush and heat stimuli in healthy and neuropathic pain subjects. Cerebellum. 2008;7:252–72. https://doi.org/10.1007/s12311-008-0011-6.

40. Stoodley CJ, Schmahmann JD. Functional topography in the human cerebellum: a meta-analysis of neuroimaging studies. NeuroImage. 2009;44:489–501. https://doi.org/10.1016/j.neuroimage.2008.08.039.

41. Ploghaus A, Tracey I, Gati JS, Clare S, Menon RS, Matthews PM, et al. Dissociating pain from its anticipation in the human brain. Science. 1999;284:1979–81.

42. Stoodley CJ, Schmahmann JD. Evidence for topographic organization in the cerebellum of motor control versus cognitive and affective processing. Cortex. 2010;46:831–44. https://doi.org/10.1016/j.cortex.2009.11.008.

43. Palermo S, Benedetti F, Costa T, Amanzio M. Pain anticipation: an activation likelihood estimation meta-analysis of brain imaging studies. Hum Brain Mapp. 2015;36:1648–61. https://doi.org/10.1002/hbm.22727.

44. Moulton EA, Elman I, Pendse G, Schmahmann J, Becerra L, Borsook D. Aversion-related circuitry in the cerebellum: responses to noxious heat and unpleasant images. J Neurosci. 2011;31:3795–804. https://doi.org/10.1523/JNEUROSCI.6709-10.2011.

45. Volkow ND, Wang GJ, Ma Y, Fowler JS, Zhu W, Maynard L, et al. Expectation enhances the regional brain metabolic and the reinforcing effects of stimulants in cocaine abusers. J Neurosci. 2003;23:11461–8.

46. Wagner MJ, Kim TH, Savall J, Schnitzer MJ, Luo L. Cerebellar granule cells encode the expectation of reward. Nature. 2017;544:96–100. https://doi.org/10.1038/nature21726.

47. Duerden EG, Albanese MC. Localization of pain-related brain activation: a meta-analysis of neuroimaging data. Hum Brain Mapp. 2013;34:109–49. https://doi.org/10.1002/hbm.21416.