A new species in the genus *Acartia* Dana, 1846 (Crustacea, Copepoda, Calanoida, Acartiidae) from the South Pacific coastal waters of Nadi Bay, Fiji

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Abstract

A new species in the genus *Acartia*, *Acartia nadiensis* sp. nov., is described from Fijian coastal waters. This species belongs to the subgenus *Odontacartia* based on the following morphological features: presence of a rostral filaments, a pointed process on the last prosomite, a serrated terminal spine on female P5, and the absence of a protrusion on the basis of the male right P5. This new species can be differentiated from its congeners by the combination of the absence of a spine on the first segment of the antennules, the short outer seta of female P5, and a medial spine on the exp-2 of the left male P5. Phylogenetic analyses using mitochondrial COI partial sequences show that the new species is distinct from its congeners.

Keywords

*Odontacartia*, planktonic copepod, mtCOI, taxonomy, South Pacific

Introduction

The genus *Acartia* Dana, 1846 is one of the most dominant groups of planktonic copepods and has a worldwide distribution in estuarine, coastal, and even oceanic waters (Bradford 1976; Walter and Boxshall 2019). Sixty-five species in this genus have been
reported from various locations ranging from tropical to polar regions (Soh et al. 2013; Razouls et al. 2019; Srinui et al. 2019), and these species have been allocated to six subgenera: Acartiura Steuer, 1915, Euacartia Steuer, 1915, Hypoacartia Steuer, 1915, Acanthacartia Steuer, 1915, Odontacartia Steuer, 1915, and Acartia (= Plankacartia) Dana, 1846 (Steuer 1915, 1923). Among these subgenera, the subgenus Odontacartia contains 13 species: Acartia amboinensis Carl, 1907; A. australis Farran, 1936; A. bispinosa Carl, 1907; A. bowmani Abraham, 1976; A. centura Giesbrecht, 1889; A. edentata Srinui, Ohtsuka & Metillo, 2019; A. erythraea Giesbrecht, 1889; A. japonicus Mori, 1940; A. lilljeborgi Giesbrecht, 1889; A. mertoni Steuer, 1917; A. ohtsukai Ueda & Bucklin, 2006; A. pacifica Steuer, 1915; and A. spinicauda Giesbrecht, 1889.  

During a survey of the diversity of planktonic copepods in Fijian waters, we collected an undescribed species of the genus Acartia that clearly belongs to the subgenus Odontacartia. In this study, we describe the morphological characters of the new species from Nadi Bay, Fiji. Partial mtCOI sequences were also obtained and compared with related species to determine if this new species is also genetically distinct from its congeners.

**Materials and methods**

**Sample collection and identification**

Specimens were collected from Nadi Bay, Fiji, using a 100 μm mesh plankton net having a 30 cm diameter mouth, and then preserved in 99% ethanol. Specimens were dissected in lactic acid, and mounted on slides with lactophenol. Preparations were sealed with transparent nail varnish. All drawings were prepared using a drawing tube attached to an Olympus BX51 differential interference contrast microscope. For scanning electron microscope (SEM) preparation, specimens were dehydrated in a series of graded ethanol solutions, then placed in isoamyl acetate, critical point dried, mounted on stubs, coated in platinum, and observed under a Hitachi S4700 field-emission electron microscope at Eulji University, Seoul, Korea. Descriptive terminology was adopted from Huys and Boxshall (1991).

**DNA extraction and amplification**

For DNA extraction, ethanol was removed from fixed specimens (99% EtOH) by washing with distilled water, and DNA was extracted using a tissue DNA purification kit (COSMO GENETECH, Co. Ltd, Korea). DNA was extracted from individual specimens. mtCOI DNA was amplified in 20 μl reaction volumes containing extracted tissue DNA and primers LCO-1490 (5’-GGT CAA CAA ATC ATA AAG ATA AAG ATA TTG G-3’) and HCO-2198 (5’-TAA ACT TCA GGG TGA CCA AAA AAT CA-3’) (Folmer et al. 1994). PCR conditions comprised initial denaturation at 94 °C for 5 min, followed by 40 cycles of denaturation at 94 °C for 1 min, annealing at 46 °C for 2 min, and extension at 72 °C for 3 min. This was followed by a final extension
step at 72 °C for 10 min. PCR products were evaluated by electrophoresing amplification products on 1% agarose gel containing ethidium bromide. Purification of amplified products was performed using a PCR purification kit (COSMO GENETECH Co. Ltd, Korea), and both strands were sequenced using an ABI 3730XL sequencer (COSMO GENETECH Co. Ltd, Korea).

**Phylogenetic analysis**

Sequences were aligned and edited using CLUSTAL W (Thompson et al. 1994) within MEGA6 (Tamura et al. 2013). For the phylogenetic analysis, three Acartia species (A. erythraea, A. japonica, and A. ohtsukai) belonging to the subgenus Odontacartia were collected from South Korea and Japan for this study (Table 1). Sequences of A. pacifica and A. spinicauda were obtained from the NCBI database for comparison. Phylogenetic analysis and pairwise distance analysis were conducted using MEGA6 software using neighbor-joining and minimum-evolution algorithms, respectively, and the Tamura-Nei model of sequence evolution was applied (Tamura and Nei 1993). Codon positions were set as follows: 1st + 2nd + Noncoding. All positions containing gaps and missing data were eliminated. Acartia (Acartiura) omorii Bradford, 1976 was used as outgroup.

**Systematics**

Order Calanoida G. O. Sars, 1903  
Family Acartiidae G. O. Sars, 1900  
Genus Acartia Dana, 1846  
Subgenus Odontacartia Steurer, 1915

*Acartia nadiensis* sp. nov.  
http://zoobank.org/DD2852BB-7AAE-4B65-85F1-4A741FD85F7F  
Figures 1–8

**Type locality.** Coastal water (17°45.848’S, 177°22.348’E), Nadi Bay, Fiji.  
**Materials examined.** All specimens have been deposited in the Marine Biodiversity Institute of Korea (MABIK). Holotype 1♀ (MABIK CR00246502) and Allotype 1♂ (MABIK CR00246503) undissected and preserved in 70% ethanol. Paratype: 2♀♀ (MABIK CR00246504-CR00246505) dissected on 13 and 10 slides, respectively; 2♂♂ (MABIK CR00246506-CR00246507) dissected on 14 and 8 slides, respectively; 10♀♀ (MABIK CR00246508-CR00246517) and 4♂♂ (MABIK CR00246518-CR00246521) undissected and preserved in 70% ethanol. 4♀♀ and 4♂♂ dried, mounted on stub, and coated with platinum for SEM. All specimens are from the type locality and were collected by S. Lee on 10 October 2013. The illustrations are based on the paratypes (♀, MABIK CR00246504; ♂, MABIK CR00246506).
Etymology. The specific name refers to the type locality of Nadi bay, Fiji.

Description of female. Total body length 975–1050 μm (mean ± SD = 1018 ± 26 μm, n = 10, holotype 1015 μm) as measured from anterior margin of cephalosome to posterior margin of the caudal rami. Body surface armed with some sensillae (Fig. 1A). Prosome:urosome length ratio = 3.52:1.

Prosome 5-segmented (Fig. 1A, B), cephalosome and first pedigerous somite completely separate; fourth and fifth pedigerous somite fused. Posterior corners of fifth pedigerous somite rounded, each with three spines. Rostral filaments thick and short (Figs 2A, 7A).

Urosome 3-segmented (Figs 1A–C, 7D–H, 8A), genital double somite slightly swollen anterolaterally, with paired gonopores ventromedially, each gonopore covered with pointed operculum; first and second urosomites each with four spines on posterdorsal margin. Caudal rami bearing short hairs on lateral margin. Proportional lengths of urosomites and caudal rami as 38:23:17:22 = 100.

Antennule incompletely 18-segmented (Fig. 2A, B), fourth to seventh segments partly fused on dorsal surface; ninth to eleventh segment each with one row of setules, twelfth segment with three rows of setules, thirteenth and seventeenth segment each with one row of setules; segmentation and setation patterns as follows: (1) I-1, (2) II-VI-[5+ae], (3) VII-[1+ae], (4) VIII-XI-[4(1spiniform)+ae], (5) XII-[0], (6) XIII-[0], (7) XIV-XV-[2+ae], (8) XVI-[1+ae], (9) XVII-XVIII-[2+ae], (10) XIX-[1], (11) XX-[1], (12) XXI-[1+ae], (13) XXII-[1], (14) XXIII-[1], (15) XXIV-[2(1+1)], (16) XXV-[2(1+1)+ae], (17) XXVI-[2(1+1)], (18) XXVII-XXVIII-[4+ae].

Antenna (Fig. 2C): coxa with seta; basis and first endopodal segment fused to form elongated allobasis bearing eight setae medially and one seta terminally along inner marin, and spinular row on distal area; second endopodal segment elongated, with seven setae, rows of spinules on lateral margin; third exopodal segment short, with seven setae. Exopod 4-segmented; setation formula 1, 2, 3.

Mandible: (Fig. 3A) coxa with well developed gnathobase bearing eleven teeth; basis with seta and row of setules on lateral and posterior margins; endopod 2-segmented, first endopodal segment with two setae, second segment with seven setae; exopod 5-segmented, setation formula as 1, 1, 1, 1, 2.

Table 1. List of species analyzed for molecular comparison.

| Species                  | Locality          | GenBank no.        | References              |
|-------------------------|-------------------|--------------------|-------------------------|
| A. (Odontacartia) erythraea | Mokpo, Korea     | MN603769–MN603773  | Present study           |
| A. (Odontacartia) japonica | Okinawa, Japan   | MN603774          | Present study           |
| A. (Odontacartia) nadiensis | Nadi Bay, Fiji   | MN603766–MN603768 | Present study           |
| A. (Odontacartia) ohtsukai  | Busan, Korea     | MN603777–MN603777 | Present study           |
| A. (Odontacartia) pacifica | Nakajima Island, Japan | KC287267           | Bucklin and Blanco-Bercial 2014 |
| A. (Odontacartia) spinicauda | Nakajima Island, Japan | DQ871177          | Ueda and Bucklin 2006   |
| A. (Acartiura) omorii    | Xiamen waters, China | DQ665253–DQ665254 | Liu et al. 2006         |
| A. (Acartiura) omorii    | Gwangyang Bay, Korea | MN603778         | Present study           |
Figure 1. *Acartia nadiensis* sp. nov. female. A Habitus, dorsal B habitus, lateral C urosome, ventral. Scale bars: in μm.
Maxillule: (Fig. 3B) precoxa and coxa incompletely fused, praecoxal arthrite with eight setae; coxal endite with three setae; one short seta and eight long setae on coxal epipodite; basal endite with one seta; basal exite with one seta; 1-segment exopod with two setae laterally and five setae terminally; endopod absent.
Maxilla: (Fig. 3C) precox and coxa incompletely fused, setation formula of endites 4, 2, 2, 3; basal endite with a seta and row of spinules on distal margin; endopod 3-segmented, with setation formula 2, 2, 3.
Maxilliped (Fig. 3D) comprising syncoxa with six setae; basis with spiniform seta; endopod 2-segmented, first segment with three setae, second segment with two setae.

Legs 1–4 (Fig. 4A–D) biramous, each with 3-segmented exopod and 2-segmented endopod, and spinules along inner and outer margins as illustrated. Intercoxal sclerites well developed. Spine and setal formulae as follows:

|   | Coxa | Basis | Exopod | Endopod |
|---|------|-------|--------|---------|
| P1 | 0-0  | 0-0   | 1-1; 1-1; II, 1, 4 | 0-1; 1, 2, 3 |
| P2 | 0-0  | 0-0   | 0-1; 0-1; 0, 1, 5 | 0-2; 1, 2, 4 |
| P3 | 0-0  | 0-0   | 0-1; 0-1; 0, 1, 5 | 0-2; 1, 2, 4 |
| P4 | 0-0  | 1-0   | 0-1; 0-1; 0, 1, 5 | 0-3; 1, 2, 3 |

P5 (Figs 2D, 7B, C) symmetrical, 3-segmented; basis ovate, with outer seta; exopod tapering, thick, bent at midlength, distal portion serrated, base slightly swollen.

**Description of male.** Total body length 910–952 μm (mean ± SD = 931 ± 16 μm, n = 5, allotype 930 μm) measured from anterior margin of cephalosome to posterior margin of caudal rami. Body surface armed with some sensilla (Fig. 5A, B). Prosome:urosome length ratio = 3.12:1.

Prosome (Fig. 5A, B) 5-segmented. Rostral filaments thin (Figs 5A, B, 8B). Fifth prosomite with six spines on posterior margin.

Urosome (Figs 6C, D, 8C–H) 5-segmented. Second urosomite with four spines on posterodorsal margin and two spines on posteroventral margin; pair of sensillae on dorsal surface. Third and fourth urosomites each with four spines on posterodorsal margin. Caudal rami bearing short hairs on lateral margin. Length proportions of urosomites to caudal rami as 16:31:21:7:12:14 = 100.

Left antennule 22-segmented (Fig. 6A). Segmentation and setation pattern as follows: (1) I-[1], (2) II-VII-[3+ae], (3) VIII-[2], (4) IX-[1+ae], (5) X-[2(1spiniform)], (6) XI-[2+ae], (7) XII-[0], (8) XIII-[0], (9) XIV-[2(1spiniform)+ae], (10) XV-[1], (11) XVI-[1+ae], (12) XVII-[1], (13) XVIII-[1+ae], (14) XIX-[1], (15) XX-[1], (16) XXI-[1+ae], (17) XXII-[1], (18) XXIII-[1], (19) XXIV-[2(1+1)+ae], (20) XXV-[2(1+1)+ae], (21) XXVI-[2(1+1)], (22) XXVII-XXVIII-[4+ae]. Right antennule 18-segmented (Fig. 6B), with geniculation with fourteenth and fifteenth segments. Segmentation and setation pattern as follows: (1) I-[1], (2) II-VII-[3+ae], (3) VIII-[2], (4) IX-[1+ae], (5) X-XI-[3(1spiniform)+ae], (6) XII-[0], (7) XIII-[0], (8) XIV-[2(1spiniform)+ae], (9) XV-[1], (10) XVI-[1+ae], (11) XVII-[1], (12) XVIII-[1+ae], (13) XIX-[1], (14) XX-[1], (15) XXI-XXII-[3+ae], (16) XXIV-XXV-[4(2+2)+ae], (17) XXVI-[2(1+1)], (18) XXVII-XXVIII-[4+ae].

Other mouthparts and P1–P4 as in female. P5 asymmetrical (Fig. 5C); intercoxal sclerite distinct. Left leg 4-segmented; basis armed with posterolateral seta and rounded lobe on posterior surface; exopod 2-segmented, exp-1 unarmed; exp-2 with hairs, and one spine with teeth on medial margin and one small spine distally. Right leg 5-segmented, basis armed with posterolateral seta. Exopod 3-segmented, exp-1 with long slender seta; exp-2 with oblong inner lobe bearing one spine on distal margin; exp-3 with one spine on medial margin and one spine distally.
Figure 4. *Acartia nadiensis* sp. nov. female. A P1 B P2 C P3 D P4. Scale bars: in μm.
Figure 5. *Acartia nadiensis* sp. nov. male. **A** Habitus, dorsal **B** habitus, lateral **C** P5. Scale bars: in μm.
Figure 6. *Acartia nadiensis* sp. nov. male. A Antennule (left) B antennule (right) C urosome, dorsal D urosome, ventral. Scale bars: in μm.
Figure 7. *Acartia nadiensis* sp. nov. Scanning electron micrographs. A Female, rostrum B female, P5 C female, P5, terminal spine D female, genital double-somite E female, genital field F female, 1st urosomite, dorsal view G female, 2nd urosomite, lateral view H female, caudal rami, dorsal view. Scale bars: in μm.
Figure 8. *Acartia nadiensis* sp. nov. Scanning electron micrographs. **A** Female, urosome and caudal rami, ventral view **B** male, rostrum **C** male, 1st urosomite **D** male, 2nd–4th urosomite, dorsal view **E** male, 1st urosomite, lateral view **F** male, 2nd urosomite, lateral view **G** male, 5th urosomite and caudal rami, dorsal view **H** male, 4th urosomite and caudal rami, ventral view. Scale bars: in μm.
Molecular analysis

A 581 bp partial region of mtCOI was sequenced from five species: *A. nadiensis* sp. nov., *A. erythraea*, *A. japonica*, *A. ohtsukai*, and *A. omorii*. Sequences of two species (*A. pacifica* and *A. spinicauda*) were obtained from NCBI and also included in the analysis. All species belong to the subgenus *Odontacartia* except *A. omorii*, which belongs to the subgenus *Acartiura* and was used as the outgroup. The mtCOI sequences of *A. nadiensis* differed in a 24.1% from *A. japonica*, and in up to 29.0% from *A. pacifica* (Table 2). Neighbor joining and minimum evolution phylogenetic analyses using the Tamura-Nei model showed that *A. nadiensis* was clearly distinct from its congeneric species (Fig. 9).

Discussion

The new species, *Acartia nadiensis* sp. nov., clearly belongs to the subgenus *Odontacartia*. This subgenus displays the following diagnostic characters compared to the other five subgenera of *Acartia*: presence of rostral filaments, symmetrical pointed process on the posterior corner of the last prosomite, minutely serrated terminal spine on female P5, and absence of protrusion on the basis of male right P5 (Steuer 1915; Ueda and Bucklin 2006; Soh 2010). The new species can be distinguished from other *Odontacartia* species by several distinctive characters (Table 3). First, *Odontacartia* species, including the new
Table 2. Genetic variation within the subgenus *Odontacartia* species based on mtCOI sequence comparison including *A. (Acartiura) omorii* as outgroup.

|                | 1  | 2  | 3  | 4  | 5  | 6  | 7  | 8  | 9  | 10 | 11 | 12 | 13 | 14 | 15 | 16 |
|----------------|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
| *A. (Odontacartia) nadiensis* 1 |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) nadiensis* 2 | 0.000 |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) nadiensis* 3 | 0.000 | 0.000 |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) erythraea* 1 | 0.247 | 0.247 | 0.247 |    |    |    |    |    |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) erythraea* 2 | 0.247 | 0.247 | 0.247 | 0.000 |    |    |    |    |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) erythraea* 3 | 0.247 | 0.247 | 0.247 | 0.000 | 0.000 |    |    |    |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) erythraea* 4 | 0.247 | 0.247 | 0.247 | 0.000 | 0.000 | 0.000 |    |    |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) erythraea* 5 | 0.247 | 0.247 | 0.247 | 0.000 | 0.000 | 0.000 | 0.000 |    |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) japonica* 1 | 0.241 | 0.241 | 0.241 | 0.244 | 0.244 | 0.244 | 0.244 | 0.244 |    |    |    |    |    |    |    |    |
| *A. (Odontacartia) japonica* 2 | 0.277 | 0.277 | 0.277 | 0.258 | 0.258 | 0.258 | 0.258 | 0.258 | 0.258 |    |    |    |    |    |    |    |
| *A. (Odontacartia) ohtsukai* 1 | 0.275 | 0.275 | 0.275 | 0.266 | 0.266 | 0.266 | 0.266 | 0.266 | 0.266 | 0.269 | 0.028 |    |    |    |    |    |
| *A. (Odontacartia) ohtsukai* 2 | 0.278 | 0.278 | 0.278 | 0.260 | 0.260 | 0.260 | 0.260 | 0.260 | 0.267 | 0.002 | 0.030 |    |    |    |    |    |
| *A. (Odontacartia) pacifica* 1 | 0.288 | 0.288 | 0.288 | 0.282 | 0.282 | 0.282 | 0.282 | 0.282 | 0.282 | 0.267 | 0.249 | 0.249 | 0.252 |    |    |    |
| *A. (Odontacartia) pacifica* 2 | 0.290 | 0.290 | 0.290 | 0.287 | 0.287 | 0.287 | 0.287 | 0.287 | 0.278 | 0.254 | 0.254 | 0.254 | 0.257 | 0.009 |    |    |
| *A. (Odontacartia) spinicauda* 1 | 0.266 | 0.266 | 0.266 | 0.254 | 0.254 | 0.254 | 0.254 | 0.254 | 0.292 | 0.183 | 0.203 | 0.181 | 0.252 | 0.262 |    |    |
| *A. (Odontacartia) spinicauda* 2 | 0.263 | 0.263 | 0.263 | 0.259 | 0.259 | 0.259 | 0.259 | 0.259 | 0.287 | 0.183 | 0.203 | 0.181 | 0.247 | 0.257 | 0.003 |    |
| *A. (Acartiura) omorii* | 0.292 | 0.292 | 0.292 | 0.251 | 0.251 | 0.251 | 0.251 | 0.251 | 0.270 | 0.297 | 0.290 | 0.299 | 0.296 | 0.304 | 0.268 | 0.270 |
Table 3. Morphological differences among species within the subgenus *Odontacartia* (Calanodia: Acartiidae: *Acartia*).

|                         | A. nadiensis *sp. nov.* | A. amboinensis | A. australis | A. bispinosa | A. bowmani | A. centura | A. advena | A. erythraea | A. japonica | A. lilljeborgi | A. mertoni | A. ohtsukai | A. pacifica | A. spinicauda |
|-------------------------|-------------------------|----------------|--------------|--------------|------------|------------|-----------|--------------|-------------|----------------|------------|-------------|-------------|--------------|
| **Female**              |                         |                |              |              |            |            |           |              |             |                |            |             |             |              |
| Body length             | 975–1050               | 1340–1510     | 1290–1400    | 1200–1300    | 1350–1400  | 1400       | 1350–1410 | 1350–1400    | ND          | 1190–1230     | 1190–1210 | 1250        |              |              |
| Spine on 1st seg of antennules | absent             | present       | present      | present      | absent     | absent     | absent    | present      | present     | absent         | absent     | absent      | absent      | absent       |
| **P5**                  |                         |                |              |              |            |            |           |              |             |                |            |             |             |              |
| Basis length/width ratio | 2                     | 4             | 2.5          | 2.4          | 1.5        | 1.6        | 1.4       | 2.3          | 2.1         | 1.4            | 2          | 1.8         | 1.4         | 1.4          |
| Length ratio of P5 outer seta/terminal spine | 0.4                  | 1.8           | 1.2          | 1.4          | 0.9        | 1.6        | 1.3       | 1.6          | 0.7         | 1.5            | 1          | 1           | 1.8         | 1.2          |
| **Urosome**             |                         |                |              |              |            |            |           |              |             |                |            |             |             |              |
| Dorsal spines on 1st urosomite | 4                    | 2             | 2            | 2            | 0          | 2          | 0         | 2            | 2           | (small spinules) | 2          | 2           | 2           | 2            |
| Dorsal spines on 2nd urosomite | 4                    | 4             | 0            | 0            | 2          | 2          | 2         | 2            | (small spinules) | (small spinules) | 2          | 2           | 2           | 2            |
| Caudal rami length/width ratio | 1.8                  | 1.3           | 1.1          | 1.8          | 2          | 1.7        | 3         | 1.4          | 1.2         | 1.5            | 2          | 3           | 2.5         | 3            |
| **Male**                |                         |                |              |              |            |            |           |              |             |                |            |             |             |              |
| Body length             | 910–952                | ND            | 1170–1230    | 1070–1160    | 1100       | 1250–1280 | 1080–1150 | ND           | 1190–1240   | 1100          | ND         | 1030–1050  | 1030–1160  | ND           |
| Left P5                 |                         |                |              |              |            |            |           |              |             |                |            |             |             |              |
| Length ratio of medial process/segment on 2nd exopodite | 0.5                  | ND            | 0.7          | 0.4          | 0.9        | 0.8        | 2         | 0.4          | 1           | 0.5            | 3.5        | 1.4         | 1.6         | 0.9          |
| Type of medial process on 2nd exopodite | Spine with teeth | ND            | Spine       | Spine with fine setae | Spine | Spine | Long seta | Spine | Spine with teeth | Spine | Long seta | Long seta | Long seta | Spine |
| **References**          | This study             | Tanaka 1965    | Ueda 1986    | Nishida 1985; El-Sherihny and Al-Addaros 2014 | Abraham 1976 | Abraham 1976 | Srinui et al. 2019 | Mori 1964 | Ueda 1986 | Giesbrecht 1892 | Steuer 1923; Ueda and Bucklin 2006 | Ueda and Bucklin 2006 | Ueda and Bucklin 2006 | Giesbrecht 1982; Mori 1964 |
A new Acartia from Fiji

Acartia species, can be divided into two groups based on the presence of a spine on the first segment of antennules (Steuer 1923; Srinui et al. 2019). Species with this spine include *A. amboinensis*, *A. australis*, *A. bispinosa*, *A. erythraea*, *A. japonica*, and *A. lilljeborgi*. Species lacking spine include *A. bowmani*, *A. centura*, *A. mertoni*, *A. ohtsukai*, *A. pacifica*, and *A. spinicauda*. *Acartia nadiensis* sp. nov. also lacks a spine on the first segment of antennules. Second, the outer seta of the female P5 of *A. nadiensis* sp. nov. is much shorter than the terminal spine, and the length ratio of the outer seta/terminal spine is 0.4. Most species of *Odontacartia* have an outer seta that is longer than the terminal spine in female P5. There are two species (*A. bowmani* and *A. japonica*) that have a short terminal seta on female P5, and the length ratio of outer seta/terminal seta are 0.9 and 0.7, respectively. Third, the male P5 of *A. nadiensis* is clearly distinguishable from the rest of species based on its length and the type of medial process on the exp-2 of the left leg. Furthermore, the new species shows other minor differences compared to the other 13 *Odontacartia* species, such as the number of dorsal spines on the urosomite, the length/width ratio of the female P5 basis, and the length/width ratio of caudal rami.

To supplement the morphological evidences, we conducted molecular phylogenetic analyses using partial mtCOI sequences of six *Odontacartia* species, including the new species. The mtCOI gene is widely used to identify sibling species due to its higher evolutionary rate than 16s and 18s rDNA (Knowlton and Weight 1998; Hebert et al. 2003; Schindel and Miller 2005; Karanovic et al. 2018). In previous studies of calanoid copepods, mtCOI sequence divergence between species have been shown to range from 13.0–22.0% (Bucklin et al. 1999), 17.6–26.7% (Eyun et al. 2007), and 21.0–23.0% (Soh et al. 2013). The mtCOI partial sequence of *A. nadiensis* sp. nov. differed by 24.1–29.0% from the sequences of congeneric species, which is greater than the range of interspecific differences reported in previous studies.

The length ratio of the outer seta/terminal spine of the female P5 is the most diagnostic morphological feature in *Odontacartia* species. However, this character is also used to determine the subgenus *Euacartia* (Soh et al. 2013). This confusion between subgenus systems has been documented previously (Madhupratap and Haridas 1994). Barthélémy (1999) compared female genital structure of 25 species of Acartiidae using light and scanning electron microscopy and concluded that there is no support for the current subdivision of *Acartia* into subgenera. Although the new species *A. nadiensis* belongs to the subgenus *Odontacartia* based on the current identification system, the validity subgeneric taxa, as proposed by Steuer (1915, 1923), within *Acartia* should be reevaluated.

Key to species of the subgenus *Odontacartia* Steuer, 1915

1 Presence of spine on 1st to 2nd segments of female antennule.......................2
– Absence of spine on 1st to 2nd segments of female antennule.......................5
2 Small spinule row present on dorsal surface of female 1st urosomite..........
  ..........................................................................................................................*A. lilljeborgi*
– Strong spines present on dorsal surface of female 1st urosomite...............3
3. Absence of processes (spines and spinules) on dorsal surface of female 2nd urosomite

- Small spine row present on dorsal surface of female 2nd urosomite
  \[A. japonica\]
- 2 strong spines present on dorsal surface of female 2nd urosomite
  \[A. erythraea\]
- 4 strong spines present on dorsal surface of female 2nd urosomite
  \[A. amboinensis\]

4. Length-width of female caudal rami are almost similar; medial process on 2nd exopodite of male left P5 as spine
  \[A. australis\]
- Female caudal rami almost twice longer than wide; medial process on 2nd exopodite of male left P5 as spine with fine seta
  \[A. bispinosa\]

5. Dorsal surface of female 1st urosomite devoid of processes (spines and spinules)

- Spine present on dorsal surface of female 1st urosomite
  \[A. bowmani\]
- Female caudal rami twice longer than wide; medial process and 2nd exopodite segment of male left P5 almost similar in length
  \[A. edentata\]
- Female caudal rami three times longer than wide; medial process of male left P5 twice longer than 2nd exopodite segment
  \[A. nadiensis sp. nov.\]
- Four strong spines on dorsal surface of female 1st and 2nd urosomite
  \[A. mertoni\]

6. Female caudal rami three times longer than wide

- Female P5 outer seta is longer than terminal spine
  \[A. ohtsukai\]

7. Length of female P5 outer seta and terminal spine similar

- Female P5 outer seta is longer than terminal spine
  \[A. centaura\]

8. Length-width ratio of female caudal rami as 1.7; medial process on 2nd exopodite male left P5 as spine
  \[A. pacifica\]
- Length-width ratio of female caudal rami as 2.5; medial process on 2nd exopodite of male left P5 as long seta
  \[A. spinicauda\]

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