Effects of Desflurane and Sevoflurane anesthesia on regulatory T cells in patients undergoing living donor kidney transplantation: a randomized intervention trial

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Abstract

Background: Volatile anesthetic agents used during surgery have immunomodulatory effects which could affect postoperative outcomes. Recognizing that regulatory T cells (Tregs) play crucial roles in transplant tolerance and high peripheral blood Tregs associated with stable kidney graft function, knowing which volatile anesthetic agents can induce peripheral blood Tregs increment would have clinical implications. This study aimed to compare effects of desflurane and sevoflurane anesthesia on peripheral blood Tregs induction in patients undergoing living donor kidney transplantation.

Methods: A prospective, randomized, double-blind trial in living donor kidney transplant recipients was conducted at a single center, tertiary-care, academic university hospital in Thailand during August 2015 – June 2017. Sixty-six patients were assessed for eligibility and 40 patients who fulfilled the study requirement were equally randomized and allocated to desflurane versus sevoflurane anesthesia during transplant surgery. The primary outcome included absolute changes of peripheral blood CD4⁺CD25⁺FoxP3⁺Tregs which measured by flow cytometry and expressed as the percentage of the total population of CD4⁺ T lymphocytes at pre-exposure (0-h) and post-exposure (2-h and 24-h) to anesthetic gas. P-value < 0.05 denoted statistical significance.

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Background

Kidney transplantation is the best option for renal replacement therapy in patients with end-stage renal disease (ESRD), often restoring quality of life in ESRD patients. Allograft rejection, an immune-mediated process, is a common cause of transplant failure [1–4]. Evidence indicates CD4+CD25+FoxP3+ cells, commonly known as regulatory T cells (Tregs), play a critical role in preventing graft rejection by suppression of recipient alloimmune response [5–8]. In healthy subjects, Tregs represent up to 5% of peripheral CD4+ T cells [9–11]. In kidney transplant patients, high peripheral blood Tregs were associated with stable graft function. Low peripheral blood Tregs was associated with allograft rejection [12–17].

Currently, adoptive transfer of ex vivo expanded Tregs is a promising strategy to induce transplant tolerance and control graft rejection in kidney transplant recipients [18, 19]. It has been investigated for safety and feasibility in phase I trials, i.e., the ONE (NCT02091232) and TRACT (NCT02145325). Identifying Treg-friendly agents from pharmacologic choices in multiple steps of kidney transplant management may also offer an attractive therapeutic strategy [19]. Characterization of Tregs under various treatment conditions may help refine current preventive measures or identify novel therapeutic targets.

Volatile anesthetic agents are widely used for general anesthesia during kidney transplantation. A growing body of evidence from ex vivo and clinical studies [20–26] suggest desflurane and sevoflurane (halogenated ether inhaled agents) exhibit immunomodulatory effects (e.g., cell proliferation, activation, migration, cytokine production) on neutrophils, macrophages, natural killer cells, B and T lymphocytes. These effects may be mediated via activation of volatile anesthetic receptors (i.e., γ-aminobutyric acid type A receptor, nicotinic acetylcholine receptor, serotonin receptor and non-canonical β2-integrins) or via binding to surface adhesion molecules such as integrin leukocyte function associated antigen-1, which express differentially on peripheral blood leukocytes [25, 26]. However, effects of desflurane and sevoflurane on Treg immunomodulation is surprisingly overlooked and has only rarely been investigated. A better understanding of these effects would have translational potential. For example, the early Treg immunomodulation by anesthetic agents may help mitigating the initiation of alloimmune responses during the 24-h perioperative period, and works in conjunction with the standard immunosuppressive regimen to seamlessly maintain the graft survival in LDKT patients.

This interventional trial aims to compare the immunomodulatory effects of desflurane and sevoflurane anesthesia on peripheral blood Treg induction in patients undergoing living donor kidney transplantation (LDKT). Several plasma cytokines were measured as the surrogate outcomes of the volatile anesthetic effects on anti- and pro-inflammatory responses. Evidence from this study would support future investigation of volatile anesthetic agents as part of perioperative management with an aim to improve transplant outcomes.

Methods

Trial design and patient enrollment

This prospective, double-blind, randomized intervention trial was approved by the Ethical Clearance Committee on Human Rights Related to Research Involving Human Subjects, Faculty of Medicine Ramathibodi Hospital, Mahidol University (protocol ID 045823) and the protocol was registered to ClinicalTrials.gov (identifier NCT02559297) on September 22, 2015. Patients aged ≥18 years old who received their first living donor kidney transplantation at Ramathibodi Hospital were included in the study. Patients were excluded for hyperacute graft rejection, currently on immunosuppressive drugs due to underlying diseases, receiving blood products during 24-h perioperative period, or patient refusal to participate in the study at any time point. Informed consent was obtained from all subjects. No interim analysis was performed during the trial. This study followed the CONSORT reporting guideline [27].

Results: Demographic data were comparable between groups. No statistical difference of peripheral blood Tregs between desflurane and sevoflurane groups observed at the baseline pre-exposure (3.6 ± 0.4% vs. 3.1 ± 0.4%; p = 0.371) and 2-h post-exposure (3.0 ± 0.3% vs. 3.5 ± 0.4%; p = 0.319). At 24-h post-exposure, peripheral blood Tregs was significantly higher in desflurane group (5.8 ± 0.5% vs. 4.1 ± 0.3%; p = 0.008). Within group analysis showed patients receiving desflurane, but not sevoflurane, had 2.7% increase in peripheral blood Treg over 24-h period (p < 0.001).

Conclusion: This study provides the clinical trial-based evidence that desflurane induced peripheral blood Tregs increment after 24-h exposure, which could be beneficial in the context of kidney transplantation. Mechanisms of action and clinical advantages of desflurane anesthesia based on Treg immunomodulation should be investigated in the future.

Trial registration: ClinicalTrials.gov, NCT02559297. Registered 22 September 2015 - retrospectively registered

Keywords: Clinical trial, Inhalation agent, Kidney transplant, Tregs, Volatile anesthesia
Randomization
Randomization was generated in a 1:1 allocation with a block size of 8 and the random number was put in a sealed envelope. Patients were randomly assigned to either desflurane or sevoflurane intervention by drawing a sealed envelope. Randomization took place on the day of surgery just prior to initiation of anesthesia.

Blinding
Subjects and outcome assessors (including laboratory technicians and all investigators except the designated research coordinator) were blinded to group allocation throughout surgery, laboratory investigation and data collection. Blinding was uncovered at the time of data analysis.

Interventions
Patients were randomly assigned to receive desflurane or sevoflurane for the maintenance phase of anesthesia. In addition to the randomized inhalation agents, patients received the same regimen of 1–2 mg of midazolam for premedication and intravenous anesthetic agents including 1–2 mcg kg$^{-1}$ of fentanyl, 1–2 mg kg$^{-1}$ of propofol and 0.5–0.6 mg kg$^{-1}$ of atracurium for induction of anesthesia and intubation. A balance anesthesia technique was used for maintenance phase. The inhalation agent (sevoflurane or desflurane) was used in conjunction with 50% nitrous oxide in oxygen. Ventilation was adjusted to maintain normocarbia. End-tidal anesthetic gas monitoring was used to ensure 1.0–1.5 minimum alveolar concentration (MAC) of the inhalation agent during maintenance phase in both groups.

During anesthesia, blood pressure, heart rate, oxygen saturation, ETCO$_2$, and temperature were monitored and recorded. Blood pressure was maintained within 20% of baseline values. Hypotension was managed by intravenous fluid and ephedrine IV bolus as needed. Total doses of intravenous medications were recorded. All patients in both interventions were transferred to the kidney transplant unit for postoperative care.

Blood sample collection
Venipuncture was performed at three time-points; pre-exposure (0-h) and post-exposure (2-h, and 24-h) to inhalation agents. Two tubes of 0.5-ml EDTA blood were collected at each time point, one for Treg enumeration and the other for cytokine measurement.

Outcome measures
The primary outcome was the absolute change in number of peripheral blood CD4$^+$CD25$^+$FoxP3$^+$Tregs, which was measured by flow cytometry and expressed as the percentage of the total population of CD4$^+$ T lymphocytes at pre-exposure (0-h) and post-exposure (2-h and 24-h) to anesthetic gas. A secondary outcome was the plasma level of anti-inflammatory cytokine IL-10 (the major cytokine produced by Tregs), TGF-β1 (anti-inflammatory cytokines produced by many types of cells and required for Tregs differentiation), and pro-inflammatory cytokines produced by T helper (Th) 1/Th2, i.e., GM-CSF, IFN-γ, IL-2, IL-4, IL-5, IL-12, IL-13 and TNF-α, which measured by multiplex immunoassay. All measurements were performed in triplicate.

Treg enumeration by flow cytometry
Peripheral blood mononuclear cells (PBMC) were isolated by density gradient centrifugation. Approximately 5 × 10$^5$ cells were suspended in 20 μL phosphate buffer saline (PBS) in the presence of cell surface marker antibodies (APC-CD4, PE-Cy7-CD25) (#MHCD0404, #25–0259-41; ThermoFisher, Florence, KY), mixed well and incubated at room temperature for 15 min. Thereafter, cells were permeabilized and intracellularly stained using FoxP3-FITC antibody (#11–4776-42; ThermoFisher). Flow cytometry (BD FACSVerse with BD FACSuite software; BD Bioscience, San Jose, CA) was used to measure the number of Tregs, expressed as a percentage of CD4$^+$CD25$^+$FoxP3$^+$ T cells among the CD4$^+$ cell population. The estimated number of CD4$^+$ cells and Treg were calculated by determining the ratio of CD4$^+$ cell count and CD4$^+$CD25$^+$FoxP3$^+$ cell count, respectively, to the total count in the flow cytometry, and then multiplied by the number of white blood cells measured from the complete blood count (CBC) which ordered at the pre-operative and post-operative evaluations.

Cytokine measurement by multiplex immunoassay
Multiplex cytokine immunoassay was performed by BioPlex-200 system (Bio-Rad, Hercules, CA). GM-CSF, IFN-γ, IL-2, IL-4, IL-5, IL-10, IL-12, IL13, and TNF-α were detected by BioPlex Pro human cytokine Th1/Th2 assay (Bio-Rad), and TGF-β1 was measured by single-plex custom assay (Bio-Rad) as the manufacturer’s instruction.

Sample size estimation and statistical analysis
There was no data related to sevoflurane and desflurane anesthesia on Treg immunomodulation available at the initiation of the study. Nevertheless, Pirbudak Cocelli L et al. [21], showed that sevoflurane and desflurane anesthesia caused a significant difference in total lymphocyte count at 2-h post-induction in patients undergoing abdominal surgery. Since Treg is a subset of lymphocytes, our study then adopted mean difference under the randomized inhalation agents. Two tubes of 0.5-ml EDTA blood were collected at each time point, one for Treg enumeration and the other for cytokine measurement.

Outcome measures
The primary outcome was the absolute change in number of peripheral blood CD4$^+$CD25$^+$FoxP3$^+$Tregs, which was measured by flow cytometry and expressed as the percentage of the total population of CD4$^+$ T lymphocytes at pre-exposure (0-h) and post-exposure (2-h and 24-h) to anesthetic gas. A secondary outcome was the plasma level of anti-inflammatory cytokine IL-10 (the major cytokine produced by Tregs), TGF-β1 (anti-inflammatory cytokines produced by many types of cells and required for Tregs differentiation), and pro-inflammatory cytokines produced by T helper (Th) 1/Th2, i.e., GM-CSF, IFN-γ, IL-2, IL-4, IL-5, IL-12, IL-13 and TNF-α, which measured by multiplex immunoassay. All measurements were performed in triplicate.
Statistical analysis was performed by Excel and R packages. Data were reported in number, percentage, mean ± SD (or SEM) or median [IQR] as appropriate. Parametric and non-parametric tests were used, as appropriate, to determine difference between groups. ANOVA with Tukey post-hoc test was performed for multiple comparison. P-value < 0.05 was considered to be statistically significant.

Results

Baseline characteristics

Figure 1 shows the flow of the participants in this study. A total of 66 patients were assessed for eligibility and 46 patients who met inclusion criteria were recruited during August 10, 2015 to June 3, 2017 for randomization and allocation to the intervention. Six patients who met exclusion criteria after allocation due to receiving perioperative blood products were excluded. Table 1 shows demographic and clinical data for the 40 patients enrolled in this study. Most variables, including recipient factors, donor factors, protocol immunosuppressive regimens, intraoperative parameters, the dosage of intravenous anesthesia were not significantly different between the intervention groups. Donor age and the estimated blood loss were slightly lower in patients receiving desflurane anesthesia. Factors contributing to ischemic-reperfusion injury, i.e., cold and warm ischemic time, were comparable between groups. Also, anesthesia time was not different between groups (279 ± 42 min vs. 303 ± 45 min, p = 0.098). Given that the depth of sevoflurane and desflurane anesthesia was maintained at 1.0–1.5 MAC for each arm, this finding supported that patients were exposed to inhalation agents equally (Table 1).

Effects of Desflurane and Sevoflurane anesthesia on Tregs in LDKT recipients

Figure 2a demonstrates the gating strategy of flow cytometry and Fig. 2b shows the effects of sevoflurane and desflurane anesthesia on CD4+CD25+FoxP3+ Tregs in peripheral blood of LDKT recipients (n = 20 per group). No significant difference of peripheral blood Tregs (mean ± SEM) was observed at pre-exposure (3.6 ± 0.4% vs. 3.1 ± 0.4%; p = 0.371) and 2-h post-exposure (3.0 ± 0.3% vs 3.5 ± 0.4%; p = 0.319) between desflurane and sevoflurane, respectively. However, at 24-h post-exposure, desflurane group had significantly higher peripheral blood Tregs as compared to sevoflurane group (5.8 ± 0.5% vs. 4.1 ± 0.3%; p = 0.008) (Fig. 2b). Within-group analysis showed that the patients receiving desflurane had 2.7% increase in Tregs over 24-h period (p <
0.001) (Fig. 2b), while this effect was not observed in sevoflurane group.

Although peripheral blood Treg are commonly presented in the literatures as the percentage of CD25⁺FoxP3⁺cells in the CD4⁺cell population, one argument was that the increment in Treg percentage might be corresponding to the global changes of leukocytes or CD4⁺ T cells in response of surgical procedure and postoperative inflammation but not the influence of inhalation agents. To address this issue, the absolute number of white blood cells (as measured by the complete blood count), and the calculated numbers of CD4⁺ T cells and CD4⁺CD25⁺FoxP3⁺ Tregs (details in the

| Characteristics                          | Sevoflurane (n = 20) | Desflurane (n = 20) | P-value |
|------------------------------------------|----------------------|---------------------|---------|
| Recipient                                |                      |                     |         |
| Age (year), mean ± SD                    | 37.6 ± 10.5          | 42.3 ± 10.6         | 0.163   |
| Gender, male                             | 13 (65)              | 15 (75)             | 0.490   |
| BMI (kg/m²), median [IQR]                | 21.8 [19.2, 24.4]    | 22.3 [21.5, 25.1]   | 0.301   |
| Hypertension                             | 18 (90)              | 19 (95)             | 0.999   |
| Diabetes mellitus                        | 4 (20)               | 0 (0)               | 0.106   |
| ADPKD                                    | 1 (5)                | 0 (0)               | 0.999   |
| Hemodialysis                             | 18 (90)              | 15 (75)             | 0.407   |
| Peritoneal dialysis                      | 2 (10)               | 5 (25)              | 0.407   |
| Baseline creatinine (mg/dL), median [IQR]| 7.7 [5.4, 11.8]      | 7.2 [5.2, 12.9]     | 0.797   |
| Pretransplant PRA (%), median [IQR]      | 0 [0, 0]             | 0 [0, 0]            | 0.999   |
| HLA-A+B+DR mismatch, median [IQR]        | 1 [1, 1.5]           | 3 [1, 3]            | 0.371   |
| White blood cell count (cells/mL), mean ± SD | 6130 ± 2157         | 5838 ± 2072         | 0.665   |
| Lymphocyte (%), mean ± SD                | 21.4 ± 6.7           | 22.8 ± 9.2          | 0.586   |
| Donor                                    |                      |                     |         |
| Age (year), mean ± SD                    | 44.8 ± 12.8          | 36.1 ± 11.3         | 0.029   |
| Gender, male                             | 7 (35)               | 6 (30)              | 0.736   |
| Donor warm ischemic time (min), median [IQR] | 3.0 [2.0, 4.0]       | 3.0 [2.0, 5.3]      | 0.519   |
| Cold ischemic time (min), median [IQR]   | 26.5 [18.0, 31.5]    | 26.5 [19.5, 40.8]   | 0.432   |
| Protocol immunosuppressive drug          |                      |                     |         |
| Tacrolimus                                | 8 (40)               | 11 (55)             | 0.342   |
| Cyclosporin A                             | 3 (15)               | 2 (10)              | 0.999   |
| Tacrolimus plus basiliximab               | 7 (35)               | 5 (25)              | 0.490   |
| Cyclosporin A plus basiliximab            | 2 (10)               | 2 (10)              | 0.999   |
| Intraoperative variable                   |                      |                     |         |
| Recipient warm ischemic time (min), median [IQR] | 37.5 [31.5, 44.5]    | 35.5 [30.0, 41.3]   | 0.647   |
| Estimated blood loss (mL), mean ± SD     | 348 ± 124            | 250 ± 129           | 0.020   |
| Intravenous fluid (mL), mean ± SD        | 3716 ± 869           | 3968 ± 979          | 0.396   |
| Intravenous ephedrine (mg), median [IQR]  | 0.0 [0.0, 7.5]       | 0.0 [0.0, 12.8]     | 0.701   |
| Intravenous anesthesia                    |                      |                     |         |
| Propofol (mg), median                     | 175.0 [127.5, 200.0, 170.0, 0.26] |  | 2 |
| Fentanyl (mcg), median                    | 100.0 [100.0, 100.0] | 100.0 [100.0, 0.76] | 7 |
| Morphine (mg), median                     | 5.5 [3.0, 7.3]       | 6.0 [3.0, 10.0]     | 0.678   |
| Midazolam (mg), median                    | 2.0 [0.0, 2.1]       | 2.0 [0.0, 2.5]      | 0.650   |
| Atracurium (mg), median                   | 90.0 [80.0, 110.0]   | 92.5 [85.0, 101.3]  | 0.643   |
| Anesthesia time (min)                     | 279 ± 42             | 303 ± 45            | 0.098   |

Abbreviations: ADPKD Autosomal dominant polycystic kidney disease, BMI Body mass index, HLA Human leukocyte antigen, PRA Panel reactive antibodies
Method section), were shown in Fig. 2c. Postoperative leukocytosis was observed in both groups as expected (Fig. 2c, the left panel), while the number of CD4+ T cells were not significantly changed during 24-h perioperative period (Fig. 2c, the center panel). Consistently, the number of Tregs (median [IQR]) were comparable between groups at baseline (7 [6, 11] vs. 8 [4, 9] cells/mM; \( p = 0.780 \)) and were significantly higher in desflurane group (12 [9, 16] vs. 8 [4, 12] cells/mM; \( p = 0.033 \)) at 24-h post-exposure (Fig. 2c, the right panel). This finding suggested the effect of desflurane anesthesia on the peripheral blood Treg induction during 24-h postoperative period.

Plasma cytokine levels were measured by multiplex immunoassay as a surrogate outcome of immunomodulation possibly influenced by inhalation agents (\( n = 26, 12 \) sevoflurane and 14 desflurane). Although there was no statistically significant difference between groups in any cytokine, a trend of increased IL-10 was observed in desflurane group (12 [9, 16] vs. 8 [4, 12] cells/mM; \( p = 0.033 \)) at 24-h post-exposure (Fig. 2c, the right panel). This finding suggested the effect of desflurane anesthesia on the peripheral blood Treg induction during 24-h postoperative period.

An increased trend of plasma IL-10 in the desflurane group was in line with previous results (Fig. 2) and suggested that desflurane anesthesia was associated with IL-10-producing Tregs induction in LDKT patients during the perioperative period. Matched-pair data of Tregs and plasma IL-10 levels in 26 patients (14 desflurane, 12 sevoflurane) were analyzed to observe this immunophenotypic response. Scatter plot showed a positive relationship between Tregs and IL-10 fold changes over 24-h period (Fig. 4a), in which the proportion of patients with increased Tregs and IL-10 immunophenotypic response was higher in the desflurane group (Fig. 4b). Taken together, our findings revealed that desflurane anesthesia induced IL-10-producing Tregs in LDKT recipients over 24-h postoperative period.

**Discussion**

Increasing evidence suggests that volatile anesthetic agents exhibit immunomodulatory effects linked to innate and...
adaptive immunity via induction and suppression of neutrophils, macrophages, NK cells and B and T lymphocytes [25, 26]. However, their effects on Treg have remained unknown. This study, for the first time, showed that desflurane, but not sevoflurane, increased Treg frequency in peripheral blood of LDKT recipients during 24-h perioperative period. Selection of desflurane anesthesia in kidney transplantation may have additional benefits to kidney graft outcome, particularly preventing allograft rejection.

Studies showed that kidney transplant patients who maintained a high level of peripheral blood Tregs were associated with better outcomes [14, 15, 28]. San Segundo D, et al. [14], reported that among 90 kidney transplant recipients, patients who maintained high levels (above 70th percentile) of peripheral blood Tregs at both 6 and 12 months had a better prognosis in the aspect of long-term graft survival after 4 and 5 years follow-up. Liu L, et al. [15], compared peripheral blood Treg levels between 42 patients with stable kidney graft function and ten patients who suffered from chronic rejection. The results showed that Treg levels were significantly higher in the stable group than the chronic rejection group. Alberu J, et al. [28], investigated the association between Treg levels and de novo donor-specific HLA-antibody (DSA) production in 53 kidney transplant patients. Although early development of DSA was not associated with Treg numbers, at 12 months after kidney transplant DSA-negative patients had higher number of peripheral blood Treg.

The mechanisms for which higher peripheral blood Tregs help prevention of allograft rejection and maintenance of transplant tolerance meet the same concept of peripheral regulation in autoimmune reaction [18, 29–32]. On a cellular basis, Tregs utilize four modes of action in peripheral regulation including [29–32]: i) secretion or generation of inhibitory cytokines (e.g., IL-10, IL-35, TGF-β and adenosine); ii) direct killing of targets through Granzyme A/B and perforin-dependent cytotoxicity; iii) IL-2 consumption through high IL-2R expression which leads to cytokine-mediated deprivation and

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**Fig. 3** Plasma cytokines were measured by multiplex cytokine immunoassay. Box plots exhibited plasma levels of anti-inflammatory cytokines IL-10 and TGF-β, and pro-inflammatory cytokines GM-CSF, IFN-γ, IL-2, IL-4, IL-5, IL-12, IL-13 and TNF-α (n = 26; 14 desflurane, 12 sevoflurane). IL-10 showed an increased trend over 24-h period in patients receiving desflurane anesthesia. Des, desflurane; Sevo, sevoflurane.
apoptosis of effector cells; and iv) direct interaction with 
CTLA-4, LAG-3 and PD1 molecules. Although accumu-
lation evidence would favor contact-dependent mechan-
isms over non-contact/secretory component alone, these different mechanisms should work in concert to 
control various immune effector cells and regulate dif-
ferent inflammatory settings [29–32]. Given that breadth 
of regulatory function on autoimmunity and self-
tolerance, changes in peripheral blood Tregs in kidney 
transplant recipients may shift the balance between allo-
graft rejection and transplant tolerance.

According to these lines of evidence, the increment of 
Tregs and IL-10 (Figs. 2, 3 and 4) after exposure to des-
flurane anesthesia should be beneficial to graft outcomes 
in LDKT recipients. In fact, several drugs routinely used 
in general anesthesia (besides volatile anesthetic agents) 
have immunomodulatory properties [25, 26]. It is pos-
sible that some of them have positive effects on Tregs. 
Synergistic effects of multiple Treg-modulated agents 
may provide a better transplant outcome. Understanding 
how anesthetic agents exhibit varied effects on the im-
une system, particularly on Tregs, is important for fu-
ture development of perioperative medicine in kidney 
transplantation.

This study was associated with several limitations. 
First, clinical outcomes, such as short-term and long-
term graft survival, that were associated with Treg 
immunomodulation of desflurane anesthesia were not 
investigated. Graft survival is influenced by various fac-
tors (such as adequacy and toxicity of immunosuppres-
sive drugs, presence of donor-specific antigen, PRA 
levels, numbers of HLA mismatch). These factors, to-
gether with Treg immunomodulation of desflurane, 
should be taken into account in future studies. Secondly, 
plasma cytokine levels were measured in 26 out of 40 
patients (2/3 of total population in this study) due to the 
limited budget. A non-significant difference of cytokines 
between intervention groups may be due to a lack of 
statistical power, but at least, the upward trend of 
plasma IL-10 was observed, given the supportive evi-
dence that desflurane anesthesia induced peripheral 
blood Tregs with potential IL-10 production. Third, the 
mechanisms of action (MoA) that drive desflurane-Treg 
immunomodulation were not defined and not the focus 
of this study. Further studies to characterize receptors 
and downstream signaling pathways that are responsible 
for desflurane-Treg effects would give some insight into 
a new MoA class of volatile anesthetic agent or a new 
biological process that facilitates transplant tolerance in LDKT recipients.

Conclusion
In summary, desflurane had the advantage over sevoflur-
ane as the inhalation anesthetic in LDKT patients 
regarding the increment of peripheral blood Tregs. Fur-
ther research focused on clinical outcomes and pharma-
cological actions of desflurane on Treg 
immunomodulation has translational potential, which 
could eventually benefit LDKT recipients.

Supplementary information
Supplementary information accompanies this paper at https://doi.org/10. 
1186/s12871-020-01130-7.

Additional file 1: Table S1. Effects of sevoflurane and desflurane 
anesthesia on plasma cytokine levels of LDKT patients.

Abbreviations
ADPKD: Autosomal dominant polycystic kidney disease; BMI: Body mass 
index; ESRD: End-stage renal disease; HLA: Human leukocyte antigen; 
LDKT: Living donor kidney transplantation; MoA: Mechanisms of action; 
PRA: Panel reactive antibodies; Th1: T helper 1; Th2: T helper 2; 
Tregs: Regulatory T cells

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Authors’ contributions
AC and SC initiated the conception. AC, NOA, GG, WI, VS, SC developed 
the design. SP performed patient recruitment, randomization, and allocation. AC 
and WI performed anesthesia intervention. NUA, GG, VS provided medical 
care and surgical operation. NP, SS, NK, NOA, SH, SC performed Treg 
enumeration and cytokine measurement. AC, SP, SC analyzed data, prepared 
figures and tables. AC and SP wrote the manuscript. NP, SS, NK, NOA, GG, 
NUA, SH, WI, VS, SC revised the manuscript. All authors read and approved 
the final version to be published.

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manuscript.

Availability of data and materials
The datasets used and/or analyzed during the current study are available 
from the corresponding author on reasonable request.

Ethics approval and consent to participate
This study was approved by the Ethical Clearance Committee on Human 
Rights Related to Research Involving Human Subjects, Faculty of Medicine 
Ramathibodi Hospital, Mahidol University (protocol ID 045823) and the 
protocol was registered to ClinicalTrials.gov (identifier NCT02559297). All 
patients provided their written informed consents to participate in this study.

Consent for publication
Not applicable.

Competing interests
The authors declare no conflict of interests.

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