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Role of viral bioaerosols in nosocomial infections and measures for prevention and control

Bing-Yuan\(^{a,b,1}\), Yun-Hui Zhang\(^{b,1}\), Nancy H.L. Leung\(^{c}\), Benjamin J. Cowling\(^{c,*}\), Zi-Feng Yang\(^{b,**}\)

\(^a\) State Key Laboratory of Respiratory Disease, National Clinical Research Center for Respiratory Disease, First Affiliated Hospital of Guangzhou Medical University, No. 151, Yanjiangxi Road, Yuexiu District, Guangzhou, Guangdong 510120, China

\(^b\) Department of Respiration, First People’s Hospital of Yunnan Province, 157, Jinbi Road, Kunming, Yunnan 650032, China

\(^c\) WHO Collaborating Centre for Infectious Disease Epidemiology and Control, School of Public Health, Li Ka Shing Faculty of Medicine, The University of Hong Kong, Patrick Manson Building (North Wing), 7 Sassoon Road, Pokfulam, Hong Kong SAR, China

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ABSTRACT

The presence of patients with diverse pathologies in hospitals results in an environment that can be rich in various microorganisms including respiratory and enteric viruses, leading to outbreaks in hospitals or spillover infections to the community. All hospital patients are at risk of nosocomial viral infections, but vulnerable groups such as older adults, children and immuno-compromised/-suppressed patients are at particular risk of severe outcomes including prolonged hospitalization or death. These pathogens could transmit through direct or indirect physical contact, droplets or aerosols, with increasing evidence suggesting the importance of aerosol transmission in nosocomial infections of respiratory and enteric viruses. Factors affecting the propensity to transmit and the severity of disease transmitted via the aerosol route include the biological characteristics affecting infectivity of the viruses and susceptibility of the host, the physical properties of aerosol particles, and the environmental stresses that alter these properties such as temperature and humidity. Non-specific systematic and individual-based interventions designed to mitigate the aerosol route are available although empirical evidence of their effectiveness in controlling transmission of respiratory and enteric viruses in healthcare settings are sparse. The relative importance of aerosol transmission in healthcare setting is still an on-going debate, with particular challenge being the recovery of infectious viral bioaerosols from real-life settings and the difficulty in delineating transmission events that may also be a result of other modes of transmission. For the prevention and control of nosocomial infections via the aerosol route, more research is needed on identifying settings, medical procedures or equipment that may be associated with an increased risk of aerosol transmission, including defining which procedures are aerosol-generating; and on the effectiveness of systematic interventions on aerosol transmission of respiratory and enteric viruses in healthcare settings.

1. Introduction

Infections that spread predominantly in healthcare settings pose potential threats to public health, and cause considerable...
The purpose of this article is to review the burden of nosocomial infections of respiratory and enteric viruses, studies and methodologies employed to investigate the role of aerosol transmission in nosocomial infections, and prevention and control measures that could mitigate viral aerosols in healthcare settings. We also discuss the challenges and research gaps in studying the importance of aerosol transmission of respiratory and enteric viruses in healthcare settings.

2. Nosocomial infections of respiratory and enteric pathogens in hospitals

2.1. Definition of nosocomial infections

Health care-associated infection (HCAI), also known as nosocomial infection, is defined as an infection occurring in a patient during the process of care in a hospital or other health-care facility which was not present or incubating at the time of admission. This includes infections acquired in the hospital, but appearing after discharge, and also occupational infections among staff of the facility (Benenson, 1995). It is traditionally believed that most HCAIs become apparent at least 48 h after admission to the healthcare facilities (Horan, Andrus, & Dudeck, 2008; World Health Organization, 2011), but this definition may not apply to viral infections that have longer mean incubation periods for example measles (12.5 days), adenoviruses (5.6 days), respiratory syncytial virus (RSV, 4.4 days), SARS (4.0 days) and other human coronaviruses (3.2 days), and parainfluenza virus (2.6 days) (Lessler et al., 2009). Theoretically patients (hospitalized or outpatient), healthcare workers (HCWs) and visitors such as the family members of hospitalized patients can all acquire nosocomial infection, but it is more difficult to ascertain nosocomial infections in outpatients and visitors as they could also acquire the infection from the community (Aitken & Jeffries, 2001). Therefore, studies of nosocomial infections mostly focus on infections developed in hospitalized patients or HCWs.

The most frequent HCAIs include urinary tract infections (UTI), surgical site infections (SSI), bloodstream infections (BSI) and hospital-acquired pneumonia (Allegrenzi et al., 2011; World Health Organization, 2011) that are device-associated and acquired independently during medical procedures, although transmission of pathogens between patients in the hospital is also considered to contribute considerably to the burden of nosocomial infections (Valenti, Menegus, Hall, Pincus, & Douglas, 1980) with occasional outbreaks of different respiratory and enteric viruses in hospitals (Bobo & Dubberke, 2010). Note that for the group of patients who may acquire pneumonia by attending a hemodialysis clinic or long-term care facilities, which was previously denoted as healthcare-associated pneumonia, it has now been denoted as community-acquired pneumonia by the latest patient management guidelines from the Infectious Diseases Society of America (IDSA) and the American Thoracic Society (ATS) (American Thoracic Society, 2005; Kalil et al., 2016).

2.2. Burden and significance of nosocomial respiratory and enteric viral infections

In a detailed report by the World Health Organization on the worldwide burden of endemic HCAIs, the worldwide HCAI prevalence was 15.5% and HCAIs were more than twice as common in low- and middle-income countries (LMICs) compared to high-income countries (Ling, Apisarnthanarak & Madriaga, 2015; World Health Organization, 2011). In both high-income and LMIC settings, there is a considerable burden in hospital-acquired pneumonia (HAP) and ventilator-associated pneumonia (VAP), and patients admitted to emergency and intensive care unit (ICU) and those with impaired functional status are associated with more nosocomial infections (World Health Organization, 2011). In LMICs, infants less than one year of age or patients with multiple comorbidities are also at higher risk of nosocomial infections (World Health Organization, 2011). In China, it has been reported that 16.2% of ICU patients developed ICU-acquired pneumonia with a mortality rate of 37% (Zhang et al., 2014).

Viruses are considered as the most common cause of infectious diseases acquired within indoor environments (Brankston, Gitterman, Hirji, Lemieux, & Gardam, 2007), and many nosocomial infections are due to respiratory and enteric infections of viruses (Belliot, Lopman, Ambert-Balay, & Pother, 2014; Bruijning-Verhagen, Quach, & Bonten, 2012; Kambhampati, Koopmans, & Lopman, 2015; Rhinehart, Walker, Murphy, O'Reilly, & Leeman, 2012). Recent examples of particular concern are emerging infectious diseases such as SARS outbreaks in 2003, the outbreak of EVD in West Africa in 2014-15, and the on-going MERS outbreaks in the Middle East since 2012, where many HCWs were infected and acted as the amplifiers for the spread to the community (Ansumana et al., 2017; Ho et al., 2003; Ki, 2015; Shears & O’Dempsey, 2015). For human avian influenza viruses, although no strong evidence of human-to-human transmission has been reported for the highly pathogenic avian influenza (HPAI) A(H5N1) virus, there have been reports suggesting nosocomial transmission of avian influenza A(H7N9) virus, potentially through aerosols or droplets (Chen et al., 2016; Fang et al., 2015). Infections among HCWs also significantly impeded outbreak controls (Chan, 2014). On the other hand, endemic respiratory and enteric viruses also cause hospital outbreaks regularly. Human seasonal influenza viruses are a common cause of respiratory tract infections both in the community and in hospitals. Salgado, Farr, Hall, and Hayden (2002) reviewed 12 influenza...
outbreaks in hospitals, and reported the patient infection rates ranged from 3% to 50% in wards affected by the outbreaks compared to 0.7–20% throughout the hospital. Outbreaks of influenza B virus have also been reported in hospitalized older adults (Seale et al., 2009; Van Voris, Belshe, & Shaffer, 1982). Other respiratory viruses including adenovirus, rhino/enterovirus, metapneumovirus and RSV could also cause nosocomial infections in pediatric and adult patients (Chow & Mermel, 2017; La Rosa, Fratini, Della Libera, Iaconelli, & Muscillo, 2013; Sakata et al., 1998; Silva Cde et al., 2012; Yang et al., 2014). We previously reported an outbreak of human metapneumovirus in a hospital ward with bedridden patients with severe motor-and-intellectual disabilities (Yang et al., 2014). For enteric infections, based on the data captured by a German surveillance system, 49% of the norovirus-related and 14% of rotavirus-related hospitalizations were acquired nosocomially, with 65% and 41% in older adults, and 16% and 16% in infants less than one year of age for nosocomial norovirus and rotavirus infections respectively (Lindsay, Wolter, De Coster, Van Damme, & Verstraeten, 2015; Spackova, Altmann, Eckmanns, Koch, & Krause, 2010). Nosocomial outbreaks of rotaviruses and noroviruses also happened frequently (Bruining-Verhagen et al., 2012; Kambhampati et al., 2015; Rhinehart et al., 2012). In addition to those mentioned above, other respiratory and enteric viruses such as measles virus and varicella-zoster virus can also cause epidemics in hospitals through airborne transmission, which often leads to serious consequences (Aitken & Jeffries, 2001; Herfst et al., 2016).

Apart from a higher risk of infection, vulnerable groups such as older adults, children, cancer patients, patients undergoing major surgery, and immunocompromised or immunosuppressed patients are susceptible to more severe disease after acquiring nosocomial infection, as the infection may cause deterioration of the patients’ primary disease including prolonged hospital stay, complications and even death. For example, the mortality due to influenza was 33–60% in transplant or ICU patients compared to 16% in patients in acute-care facilities and geriatric hospitals, and pneumonia and death were common complications of influenza in children, transplant and cancer patients undergoing immunosuppressive treatment (Salgado et al., 2002). Álvarez-Lerma et al. (2017) reported ICU patients with hospital-acquired influenza A (H1N1)pdm09 virus infection was associated with higher risk of death than those acquired the infection from the community. A prospective observational cohort study of ventilated children with RSV infection showed that there was an increase in mortality of children patients that acquired RSV infection in pediatric ICUs (26%) or hospital wards (29%), compared to 4% in ventilated children with community-acquired RSV infection (Thorburn, Eisenhut, & Riordan, 2012).

3. Aerosol transmission of respiratory and enteric infections in healthcare settings

3.1. Modes of transmission in healthcare settings

Pathogenic microorganisms can be transmitted through direct or indirect physical contact, or through the air in respiratory droplets and aerosols (Siegel, Rhinehart, Jackson, & Chiarello, 2007), leading to cross-infections or outbreaks in hospitals. Different transmission routes could also interplay with each other. For example, on one hand, inanimate objects contaminated with pathogens from patients’ secretions such as saliva, nasal fluid and feces (fomites) can serve as vehicles for the spread of pathogens directly, contributing to indirect contact transmission (Mubareka et al., 2009); on the other hand, disturbance to fomites (e.g. by toilet flushing) leads to resuspension and generation of contaminated aerosols (La Rosa et al., 2013), contributing to aerosol transmission. In general, viruses infected with aerosol exposure seem to have greater replication efficiency in vivo, such as influenza viruses (Zhang et al., 2017).

Aerosol transmission refers to the dissemination of either droplet nuclei or small particles (aerosols) in the respirable size range containing pathogenic microorganisms that remain infective over time and distance (Siegel et al., 2007). In contrast, droplet transmission occurs when larger particles containing pathogenic microorganisms travel through the air in short distances (unusually suggested as within 1 m / 3 feet) (World Health Organization, 2014) to the mouth, nasal mucosa, ocular conjunctiva or other parts of the susceptible population. To classify between droplet and aerosol transmission, respiratory droplets are typically defined as particles with an aerodynamic diameter > 5 µm, and aerosols as those ≤ 5 µm, since the latter is believed to be deposited in the alveolar region of human lung by settlement (Duguid, 1946; Siegel et al., 2007). Larger inhaled particles (mainly droplets) will deposit in the head airway or trachea-bronchial regions of the respiratory tract, whereas smaller particles (mainly aerosols or droplet nuclei) will primarily deposit in the alveoli (Hinds, 1999; Jones & Brouseau, 2015). However, the proportion of particles that are inhaled and deposit in different regions of the respiratory tract (and therefore classified as via the droplet or aerosol route accordingly) are determined by particle aerodynamic diameter as well as lung morphology and breathing characteristics (Nazaroff, 2016). Separately, some define aerosols as those which can remain suspended in the air for prolonged periods of time with potentially a higher risk of transmission through the air. Particles with aerodynamic diameter around 20 µm take 4 min to fall for a vertical distance of 3 m and those around 5 µm take an hour to fall for the same height (Knight, 1980; Tellier, 2009). Furthermore, respiratory droplets < 20 µm at expulsion could transform into droplet nuclei by evaporation and shrinks to a little less than half of the initial diameter (Nicas, Nazaroff, & Hubbard, 2005; Tellier, 2009; Xie, Li, Chwang, Ho, & Soto, 2007). Especially particles with aerodynamic diameter < 5 µm that can remain suspended in the air for prolonged periods (La Rosa et al., 2013). Bioaerosols are usually defined as aerosols or particulate matter of microbial, plant or animal origin that is often used synonymously with organic dust (Dowes, Thorne, Pearce, & Heederik, 2003; Tellier, 2009), which generally include bacteria, fungi, viruses and their derivatives such as endotoxin, glucans and mycotoxin (Guan and Yao, 2010) with particle size varying based on the composition of the aerosols, and could range from sub-micron for those composed of viral particles to as large as 1 mm for those composed of pollen grains (Haig, Mackay, Walker, & Williams, 2016).

In healthcare settings, infection prevention and control measures are often designed to act on particular modes of transmission, and pathogen-specific measures are recommended based on the understood mode(s) of transmission for the particular pathogen (Table 1) (World Health Organization, 2014). Apart from the transmission results directly from other patients, the presence of
Table 1
Outline of the four different types of infection prevention and control (IPC) precautions in healthcare settings as recommended by the World Health Organization (2014).

| IPC precautions | Rationale | Measures | Examples |
|-----------------|-----------|----------|----------|
| Standard        | To minimize spread of infection associated with healthcare, via avoiding direct contact with patients’ blood, body fluids, secretions and non-intact skin | Hand hygiene, PPE, respiratory hygiene, environmental control, waste management and prevention of needle-stick/sharps injuries | Routine for all patients |
| Contact         | Transmitted through contact particularly by hand contamination and self-inoculation into conjunctival or nasal mucosa | PPE (disposable gloves and gowns), specific patient placement and limited patient movement | Parainfluenza, Respiratory syncytial virus (RSV) |
| Droplet         | Transmitted through large droplets which typically remain suspended in the air for a limited period of time and settle within 3 feet of the source | Use of face mask if working within 3 feet of the patients, specific patient placement (cohorting) and limited patient movement | Adenovirus, Avian influenza A(H5N1), Human influenza SARS-CoV |
| Airborne*       | Transmitted through inhalation of droplet nuclei that remain infectious over a long distance (e.g. over 3 feet) | Requires special air handling including the use of respirators (e.g. N95), specific patient placement (in ventilated isolation rooms) and limited patient movement | SARS-CoV, Human influenza during aerosol-generating procedures (AGPs) |

* This WHO infection control guideline defines airborne pathogens as those ‘transmitted through inhalation of droplet nuclei that remain infectious over a long distance (e.g. > 1 m), and require special air handling’, and therefore the measures of this article can be interpreted as ‘aerosol’.

...patients with diversity in pathologies in hospital environments causes the floors, walls, surfaces of medical equipment and the air to be frequently contaminated with a range of pathogenic microorganisms including respiratory and enteric viruses, namely, MERS-CoV, rhinovirus, Toque teno virus, human adenovirus, rotaviruses, noroviruses et al. (Akhter, al-Hajjar, Myint, & Qadri, 1995; Ansari, Springthorpe, Sattar, Rivard, & Rahman, 1991; Gallimore et al., 2008; Kim et al., 2016; Morter et al., 2011; Verani, Bigazzi, & Carducci, 2014). In one instance, viruses could be recovered from 82% of the surfaces and 85% of air samples collected in a hospital (Verani et al., 2014).

In the paragraphs below, we will focus on the aerosol and droplet transmission of respiratory and enteric viruses in healthcare settings. Much of the discussion is illustrated using studies on influenza virus since it is one of the respiratory viruses most studied, however we expect the general principles also apply to other respiratory and enteric viruses.

3.2. Aerosol and droplet transmission of respiratory and enteric viruses

The presence of microorganisms in the air can affect the health of humans and animals. These pathogenic microorganisms are often adsorbed on the solid or liquid particles suspended in the air to form aerosols or droplets, whereas very small amounts of pathogenic microorganisms in the air are usually sufficient to cause infection compared to other modes of transmission (Alford, Kasel, Gerone, & Knight, 1966). For many respiratory viruses, the relative importance of different modes in transmission are controversial if not unknown, and can even vary within the same type of virus depending on the various viral, host and environmental factors (Branch-Elliman, Savor Price, McGeer, & Perl, 2015; Carlson, Budd, & Perl, 2010; Gautret et al., 2014). However, in general, respiratory viruses considered to be transmitted mainly through droplets include SARS coronavirus (SARS-CoV), influenza virus, adenovirus, rhinovirus, RSV and parainfluenza virus, and viruses considered to be transmitted mainly through aerosols include measles (rubeola virus) and chickenpox (varicella-zoster virus) (Hall, 2000, 1982; Tang, Li, Eames, Chan, & Ridgway, 2006; Xie et al., 2007), in addition to other modes such as direct or indirect contact (fomites) transmission (Boone & Gerba, 2007; Pica & Bouvier, 2012). One study showed that a large community outbreak in SARS in Hong Kong could be explained by aerosol transmission (Yu et al., 2004), while another study showed that environmental surfaces in the hospitals could be contaminated with SARS-CoV and therefore contact transmission was considered possible (Dowell et al., 2004). It has been suggested that respiratory transmission of Ebola viruses could occur, although this is controversial (Osterholm et al., 2015; Vetter et al., 2016). Several experimental studies indicate that EBOV can be transmitted through aerosols in various animal species (Jaax et al., 1995; Weingartl et al., 2012). In addition to respiratory viruses, some enteroviruses can also be spread by airborne transmission through aerosol particles. For example, the main symptom of noroviruses infections is vomiting, a fine mist of virus particles passes into the air because of projectile vomiting. Droplets being inhaled by anyone in the immediate vicinity can be deposited in the upper respiratory tract, and subsequently be swallowed along with respiratory mucus (Bonifait et al., 2015). The evidences supporting the relative importance of different modes in transmission for respiratory and enteric viruses are summarized in Table 2. Additional discussion on modes of transmissions of these viruses can be found in the review by La Rosa et al. (2013).

3.3. Factors affecting airborne transmission in nosocomial settings

Aerosol transmission of viruses can be affected by many factors, such as the mass, diameter and the shape of the aerosol particles (Hinds, 2012). In addition to these physical parameters of aerosol particles, the properties of the viruses, environmental factors and susceptibility of the host can also affect the probability of infection being established from aerosol transmission (Table 3). On the other hand, one could also hypothesize which factors may be involved and their effects on aerosol transmission by looking at their...
and replication) and genome, and could also affect microscopically by altering the structure of viral proteins (including proteins that are involved in viral entry and replication) and genome, and could also affect macroscopically by setting up large exchange air flows in space due to temperature difference (Tang, 2009). The influence of humidity on virus survival is mainly related to whether the virus has lipid envelopes or not (Yang & Marr, 2012). But it is important to note that humidity and temperature can interact to affect the survival of viruses in aerosols, viruses decay rapidly in dry and hot environments, and only 4.7% survival over 60 min procedure (Pyankov Oleg, Pyankova Olga, & Agranovski Igor, 2012).

3.4. Methods for sampling and analysis of viral bioaerosols in healthcare settings

The size of the bioaerosol that contains viruses determines the time for which the pathogen is suspended in the air and where it deposits in the respiratory tract (La Rosa et al., 2013; Nazaroff, 2016). Many studies have been conducted with the aim of detecting levels of viral aerosols in the air, so as to inform whether infection control strategies targeting aerosol transmission (Table 1) is needed. At present, the methods used for detection of viruses in bioaerosols include impaction, impingement, cyclone sampling, electrostatic precipitation and filtration (Ghosh, Lal, & Srivastava, 2015; Haig et al., 2016; Hinds, 2012).

The principle of impaction is to collect microorganism and particles in the air, the impaction sampler draws in air and forces to change its direction which causes particles with high inertia to get impacted over collecting surfaces (Ghosh et al., 2015; Henningson et al., 2012); 2) viruses can invade the target cell; and 3) there is sufficient virus infective dose. The minimal viral load resulting in infection (i.e. minimal infectious dose) varies between different viruses, for example, the viral load of smallpox virus is 10–100 organisms; however, about 50% of H3N2 viral particles were still alive even after 90 min (Pyankov Oleg, Pyankova Olga, & Agranovski Igor, 2012).

Table 2
Relative importance of aerosol, droplet and indirect contact transmission for common respiratory and enteric viruses transmitted in healthcare settings.

| Nosocomial respiratory & enteric viruses | Mode of transmission | References |
|------------------------------------------|----------------------|------------|
|                                         | Aerosol | Droplet | Indirect contact (fomites) |
| Influenza virus                           | +       | + +     | +                     |
| Respiratory syncytial virus (RSV)         | +       | + +     | +                     |
| Adenovirus                               | +       | + +     | +                     |
| Rhinovirus                               | +       | + +     | +                     |
| Coronaviruses (CoVs), incl. SARS & MERS  | + +     | + + +   | +                     |
| Noroviruses                              | +       | + +     | +                     |
| Enteroviruses                            | + +     | + +     | +                     |
| Rubeola virus (measles)                   | + +     | +       | +                     |
| Varicella-zoster virus (chickenpox)       | + +     | +       | +                     |
| Mumps virus                              | +       | + +     | +                     |
| Ebola virus                              | + +     | + +     | +                     |
| References                               | La Rosa et al. (2013); Tellier (2009); Blachere et al. (2009); Tellier (2006). | |
|                                          | La Rosa et al. (2013); Lindsay et al. (2010), Hall (2000). | |
|                                          | La Rosa et al. (2013). | |
|                                          | Myatt et al. (2004); Jennings, Dick, Mink, Wartgow, and Inhorn (1988). | |
|                                          | La Rosa et al. (2013); Seto et al. (2003). | |
|                                          | La Rosa et al. (2013); Nazaroff (2011); Barker and Jones (2005). | |
|                                          | La Rosa et al. (2013); Chang et al. (2004); Couch, Douglas, Lindgren, Gerone, and Knight (1970). | |
|                                          | Bischoff et al. (2016); Lakssono, de Vries, McQuaid, Duprex, and de Swart (2016); Bloch et al. (1985). | |
|                                          | Garner (1996); Josephson and Gambert (1988); Gustafson et al. (1982); Leclair, Zaia, Levin, Congdon, and Goldmann (1980). | |
|                                          | Hvid, Rubin, and Mühlemann (2008). | |
|                                          | Vetter et al. (2016); Ostecholm et al. (2015). | |

Direct contact is not included in this table since most of the research is focused on the possibilities of transmission through these other mechanisms requiring an intermediate medium. Perceived relative importance of the different routes of transmission is indicated for each virus (+ + +, most important; +, least important).

Other modes of transmission include bloodborne, fecal-oral, waterborne and foodborne. Summarized from the review done by La Rosa et al. (2013) and additional references.
Table 3
Factors affecting risk of infection and severity of disease in airborne transmission of respiratory and enteric viruses in healthcare settings. In the table, we use influenza as a model to describe factors that could affect the transmissibility and severity of disease of aerosol transmission.

| Factors                      | Roles in transmission                                                                                                                                                                                                 | References                                                                                      |
|------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------|
| **Virus**                    |                                                                                                           |                                                                                               |
| Molecular structure          | Balance of different viral surface proteins affect transmission efficiency via different routes. Survival of enveloped viruses is longer at lower relative humidity (20% – 30%), while survival of non-enveloped viruses is longer at higher relative humidity (70% – 90%). | Yen et al. (2011); Tang (2009)                                                                 |
| Dose                         | Minimal infectious doses required to initiate infection are different across different viruses.                                                             | Franz et al. (1997)                                                                             |
| **Host**                     |                                                                                                           |                                                                                               |
| Behavior                     | As source, difference in frequency of coughing or sneezing could affect the total viral load released. On the other hand, it is hypothesized that bioaerosols generated during sneezing mostly originated from the nasopharynx region, while those from coughing originated from the lower respiratory tract. | Hatagishi et al. (2014)                                                                         |
| Sit of virus release         | As source, viral titers in throat or nose may not correlate with that in exhaled breath and cough. The viral loads in the laryngopharynx region and lower airway were significantly different in patients with HPAI H7N9. Under the condition where the virus was negative in laryngopharynx region, 10^5 to 10^6 copies/ml of viruses were still detected in lower airway. | Yang et al. (2015); Hatagishi et al. (2014); Milton, Fabian, Cowling, Grantham, and McDevitt (2013) |
| Site of infection            | As recipient, the expression of viral receptors at different sites determines the location of pathology (and therefore affecting severity) upon infection and susceptibility to different viruses. | Nicholls, Bourne, Chen, Guan, and Peiris (2007)                                                   |
| Immunity                     | As recipient, prior infections or vaccinations can induce production of antibodies which confers protection to subsequent infection or reduce symptom severity even if infected. | Thomas, Keating, Hulse-Post, and Doherty (2006); Hobson, Curry, Beare, and Ward-Gardner (1972) |
| Environment                  |                                                                                                           |                                                                                               |
| Temperature                  | As temperature rises, survival of viruses decreases. Temperatures higher than 30 °C can block airborne transmission of influenza virus. At high temperatures, DNA viruses are more stable than RNA viruses. Low temperatures can suppress host immunity in the respiratory tract, which can easily lead to viral infection. | Lowen, Steel, Mubareka, and Palese (2008); Lowen, Mubareka, Steel, and Palese (2007); Harper (1961) |
| Humidity                     | Survival of enveloped viruses is longer at lower relative humidity (20% – 30%), while survival of non-enveloped viruses is longer at higher relative humidity (70% – 90%). | Yang and Marr (2012); McDevitt, Rudnick, First, and Spengler (2010); Tang (2009); Lowen et al. (2007) |
| Ultraviolet radiation        | Viral bioaerosols could be disinfected with ultraviolet radiation (affected by relative humidity).                                                           | McDevitt, Rudnick, and Radonovich (2012); Walker and Ko (2007); Jordan (1961)                  |
| Organic/ inorganic contents  | Contents in blood, feces and sputum e.g. salt content produced by the host, or the materials of the surfaces of which the viral particles are attached to, can reduce/increase the stress from the environment to viral particles. | Yang and Marr (2012); Tang (2009)                                                               |

4. Prevention and control of aerosol transmission in healthcare settings

Hospital environments are complex and the exposure of health care workers, patients, and visitors to airborne virus infections remains a substantial problem (McDevitt et al., 2012; Rudnick, McDevitt, First, & Spengler, 2009). Susceptible patients, health care workers and visitors can acquire infections from the hospital environment, through droplet or aerosol transmission from viruses carried by the movement of air, or indirect contact transmission that involves physical contact with contaminated surfaces. The
potential risk of nosocomial infections is very high, but there is still lack of effective standard control measures to prevent the outbreak of nosocomial infection, for example, many studies identified the marked lack of evidence for neonatal HAI outbreak management globally (Birt et al., 2016). Understanding the route of transmission is critical for implementing the best control strategies (McDevitt et al., 2012). A range of interventions can be applied for the prevention and control of viral aerosols in hospitals (Fig. 1) as described in the following subsections.

4.1. Engineering solutions for prevention

The purpose of air filtration and purification in hospitals is to reduce the concentrations of airborne pathogens so that it is lower than their infectious dose. This could be an effective intervention to block aerosol transmission where the following methods can be applied:

First, mixing the contaminated air with uncontaminated air to dilute the contaminated air. This can reduce the peak concentrations of droplet nuclei in the contaminated indoor air (Jensen, Lambert, Iademarco, & Ridzon, 2005). However, the average concentration of droplet nuclei in contaminated indoor air will increase over time unless air filtration systems are adopted.

Second, usage of displacement ventilation. This can increase the air change rate (ac/h) of indoor air so that the contaminated air can be replaced (Jensen et al., 2005). However, the use of displacement ventilation in hospital wards can increase the risk of aerosol transmission because a patient lying in bed may exhale air plumes that can spread over long distances due to differences in air temperature and density (Qian et al., 2006). Existing guidelines recommend that the airflow should be from the room ceiling to the healthcare workers’ region, then to the patients’ region, and lastly expelled through the exhaust vents that are located at a lower level (Jensen et al., 2005). In practice, mixing ventilation and displacement ventilation are commonly used to remove contaminant particles.

Third, installation of air filtration and purification systems. Methods such as air filtration system, ionization and high voltage field, ultraviolet radiation, photocatalytic oxidation, dielectric barrier discharge and others have been adopted to purify or decontaminate indoor air (Alonso, Raynor, Davies, Morrison, & Torremorell, 2016; Dee, Batista, Deen, & Pijoan, 2006; Kozlova et al., 2010; McDevitt et al., 2012; Zhao, Aarnink, & Xin, 2014; Hyun et al., 2017). At present, national standards and related technical specifications of air purification are lacking. Medical air purifiers used should meet the following requirements: 1) Safety, should not create secondary air pollution; 2) Convenience, low noise operation, and portable; 3) Effectiveness, can effectively kill different types of pathogens. Air purification technology as a means to reduce or eliminate airborne particles or pathogenic microorganism levels has been reported previously in various fields. Traditionally, ultraviolet radiation is widely used to sanitize air (First, Rudnick, Banahan, Vincent, & Brickner, 2007), Several previous studies have indicated that UV air disinfection protects humans from infectious diseases caused by airborne microorganism (such as influenza virus, pox viruses, rhinovirus and coronavirus), and those experiments also show that virus aerosols are less susceptible to UV at higher RH than at lower RH (McDevitt et al., 2012; McDevitt, Lai, Rudnick, Houseman, & First, 2007; McDevitt, Milton, Rudnick, & First, 2008; Scarpino, Jensen, Jensen, & Ward, 1998; Walker & Ko, 2007), but some exceptions exist (Walker & Ko, 2007). In addition, the combination of UV-irradiation and other measures for air purification cannot handle large volumes air due to their limited capacity in buildings (Schulz, Bao, Claus, & Hartung, 2013).

Air filtration system is another common purification technique, which allows the air to pass through a series of filters of decreasing pore size through positive pressure ventilation. Dee et al. (2006) showed that high-efficiency particulate air filtrations which are capable of blocking the passage of particles 0.3 µm or more in diameter can significantly reduce PRRSV transmission. In addition, some filters treating with special material (e.g., dialdehyde starch) can disinfect airborne viruses simultaneously (Woo et al., 2012). Similarly, low-cost filtering systems can also filter PRRSV viruses effectively. An electrostatic particle ionization technology that generates a high negative ion output which results in the electrification of airborne particles. The ionized airborne particles are attracted toward opposite charges and may be cleared from the air by adhesion to the walls or other charged surfaces (Alonso et al.,
challenges remain for new prevention and control measures, as well as applied research: 1) The importance of respiratory pathogens. However, cross infection of viral infectious diseases, some possibly via aerosols, still exists and occurs in the hospital. A number of to improve aerosol management and surface sterilization, including engineering solutions, hand hygiene and wearing face masks.

aerosol virus particles, including those that are generated from the resuspension from the surface of contaminated objects in the humidity. In view of the possibilities of a large number of respiratory viruses and other pathogenic microorganisms that may exist as particles physical properties, virus characteristics and host factors; or extrinsic factors, including environmental temperature and numbers of respiratory and enteric viruses. Aerosol transmission of viruses can be a

infections in healthcare and community settings are needed to better de

5. Conclusions and future prospects

Pathogenic airborne respiratory and enteric viruses are important causes of nosocomial infection. These infections are a significant problem for hospitalized patients, which can lead to major burden on health and healthcare costs, particularly in the elderly, children and those with underlying diseases. In this article, we review, from the standpoints of epidemiologists and virologists, the burden and significances of nosocomial infections of respiratory and enteric viruses, the possible role of aerosol transmission in these nosocomial infections, and prevention and control measures of aerosol transmission of respiratory and enteric virus infections in healthcare settings. There is increasing evidence in the scientific literature that aerosol transmission is an important route of a number of respiratory and enteric viruses. Aerosol transmission of viruses can be affected by intrinsic factors, which include aerosol particles physical properties, virus characteristics and host factors; or extrinsic factors, including environmental temperature and humidity. In view of the possibilities of a large number of respiratory viruses and other pathogenic microorganisms that may exist as aerosol virus particles, including those that are generated from the resuspension from the surface of contaminated objects in the hospital, much prevention and control measures have been adopted to mitigate nosocomial infections via the aerosol route which aim to improve aerosol management and surface sterilization, including engineering solutions, hand hygiene and wearing face masks. However, cross infection of viral infectious diseases, some possibly via aerosols, still exists and occurs in the hospital. A number of challenges remain for new prevention and control measures, as well as applied research: 1) The importance of respiratory pathogens in interpersonal transmission, including the range and form of transmission; 2) the potential of aerosols generated by medical
instruments or equipment, as well as during medical procedures; 3) the actual contributions of physical contact, droplets and aerosols in transmission of respiratory pathogens; 4) the efficacy of different types of face masks, efficiency of air purification system in reducing the transmission of respiratory viral influenza, as well as the combined forms and effects with hand hygiene and personal protective measures. At the same time, relevant clinical medical research can also be carried out, such as controlled studies of the impact on these measures on biologically relevant measures (e.g. viral loads) and patient outcomes.

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References

AintablianZhao, N., Walpita, P., & Sawyer, M. H. (1998). Detection of Bordetella pertussis and respiratory syncytial virus in air samples from hospital rooms. *Infection Control & Hospital Epidemiology*, 19(12), 918–923 [PubMed: 9872529].

Aitken, C., & Jeffries, D. J. (2001). Nosocomial spread of viral disease. *Clinical Microbiology Reviews*, 14(3), 528–546 [PubMed: 11432812].

Akhter, J., al-Hajar, S., Mynit, S., & Quadri, S. M. (1995). Viral contamination of environmental surfaces on a general paediatric ward and playroom in a major referral center in Riyadh. *European Journal of Epidemiology*, 11(5), 587–590 [PubMed: 8549735].

Alford, R. H., Kael, J. A., Gerone, P. J., & Knight, V. (1966). Human influenza resulting from aerosol inhalation. *Proceedings of the Society for Experimental Biology and Medicine*, 122(3), 800–804 [PubMed: 5918954].

Allegranzi, B., BagheriNejad, S., Combescure, C., Graafmans, W., Attar, H., Donaldson, L., & Pittet, D. (2011). Burden of endemic health-care-associated infection in developing countries: systematic review and meta-analysis. *Lancet*, 377(9761), 228–241 [PubMed: 2146207].

Alonso, C., Raynor, P. C., Davies, P. R., Morrison, R. B., & Torremorell, M. (2016). Evaluation of an electrostatic particle ionization technology for decreasing airborne pathogens in pigs. *Aerosol Science*, 32(3), 405–419 [PubMed: 27316181].

Alvarez-Lerma, F., Marin-Corral, J., Vilà, C., Mascrans, J. R., Loeches, I. M., Barbadillo, S., ... Rodriguez, A. (2017). H1N1 GETGAG/SEMICYUC group study. Characteristics of patients with hospital-acquired influenza A (H1N1)pdm09 virus admitted to the intensive care unit. *Journal of Hospital Infection*, 95(2), 200–206 [PubMed: 28135360].

American Thoracic Society (2005). Infectious diseases society of America. Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. *American Journal of Respiratory and Critical Care Medicine*, 171(4), 388–416 [PubMed: 15699079].

Ansari, S. A., Springthorpe, V. S., Sattar, S. A., Rivard, S., & Rahman, M. (1991). Potential role of hands in the spread of respiratory viral infections: studies with human parainfluenza virus type 3 and rhinovirus 14. *Journal of Clinical Microbiology*, 29(10), 2115–2119 [PubMed: 1658033].

Ansarman, R., Keitel, S., Roberts, G. M., Nisun, F., Petersen, E., Ippolito, G., & Zumla, A. (2017). Impact of infectious disease epidemics on tuberculosis diagnostic, management, and prevention services: experiences and lessons from the 2014-2015 Ebola virus disease outbreak in West Africa. *International Journal of Infectious Diseases*, 56, 101–104 [PubMed: 27818362].

Barker, J., & Jones, M. V. (2005). The potential spread of infection caused by aerosol contamination of surfaces after flushing a domestic toilet. *Journal of Applied Microbiology*, 99(2), 339–347 [PubMed: 16033465].

Belliot, G., Lopman, B. A., Ambert-Balay, K., & Pothier, P. (2014). The burden of norovirus gastroenteritis: an important foodborne and healthcare-related infection. *Clinical Microbiology and Infection*, 20(8), 724–730 [PubMed: 24943671].

Bender, B. S., Johnson, M. P., & Small, P. A. (1991). Influenza in senescent mice: impaired cytotoxic T-lymphocyte activity is correlated with prolonged infection. *Immunity*, 72(4), 514–519 [PubMed: 2037313].

Benenson, S. A. (1995). Control of communicable diseases manual. (16th ed.). Washington: American Public Health Association.

Birx, J., Le Deaux, C., Lawn, J., Heath, P. T., & Sharland, M. (2016). Lack of evidence for the efficacy of enhanced surveillance compared to other specific interventions to control neonatal healthcare-associated infection outbreaks. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 110(2), 98–106 [PubMed: 26822602].

Bischoff, W. E., Reid, T., Russell, G. B., & Peters, T. R. (2011). Transcural entry of seasonal influenza-attenuated virus aerosols and the efficacy of n95 respirators, surgical masks, and eye protection in humans. *Journal of Infectious Diseases*, 204(2), 193–199 [PubMed: 21673029].

Bischoff, W. E., Swett, K., Leng, I., & Peters, T. R. (2013). Exposure to influenza virus aerosols during routine patient care. *Journal of Infectious Diseases*, 207(7), 1037–1046 [PubMed: 23372182].

Bischoff, W. E., McNall, R. J., Blevins, M. W., Turner, J., Lopareva, E. N., & Rota, P. A. (2016). Detection of measles virus RNA in air and surface specimens in a hospital setting. *Journal of Infectious Diseases*, 213(4), 600–603 [PubMed: 26386428].

Blachere, M. F., Lindsley, W. G., Pearce, T. A., Anderson, S. E., Fisher, M., Khakoo, R., et al. (2009). Measurement of airborne in-fluenza virus in a hospital emergency department. *Clinical Infectious Diseases*, 48(4), 438–440 [PubMed: 19133798].

Bloch, A. B., Orenstein, W. A., Ewing, W. M., Spain, W. H., Mallison, G. F., Herrmann, K. L., & Hinnan, A. R. (1985). Measles outbreak in a pediatric practice: airborne transmission in an office setting. *Pediatrics*, 75(4), 676–683 [PubMed: 3982900].

Bobo, L. D., & Dubberke, E. R. (2010). Recognition and prevention of hospital-associated enteric infections in the intensive care unit. *Critical Care Medicine*, 38(8 Suppl), S324–S334 [PubMed: 20647790].

Bonfil, I., Charlebois, R., Vimont, A., Turgeon, N., Veillette, M., Longtin, Y., ... Duchaine, C. (2015). Detection and quantification of airborne norovirus during outbreaks in healthcare facilities. *Clinical Infectious Diseases*, 61(3), 299–304 [PubMed: 25900175].

Boone, S. A., & Gerba, C. P. (2007). Significance of fomites in the spread of respiratory and enteric viral disease. *Aerosol Science*, 36(6), 1687–1696 [PubMed: 17220247].

Branch-Elliman, W., Savor Price, C., McGeer, A., & Perl, T. M. (2015). Protecting the frontline: designing an infection prevention platform for preventing emerging respiratory viral illnesses in healthcare personnel. *Infection Control & Hospital Epidemiology*, 36(3), 336–345 [PubMed: 25695176].

Branch-Elliman, W., Savor Price, C., Perl, T. M. (2015). Control of influenza in healthcare settings: early lessons from the 2009 pandemic. *Current Opinion in Infectious Diseases*, 28(4), 293–299 [PubMed: 20592530].

Chan, M. (2014). Ebola virus disease in West Africa—no early end to the outbreak. *New England Journal of Medicine*, 371(13), 1183–1185 [PubMed: 25140856].

Chang, L. Y., Tsao, K. C., Hsia, S. H., Shih, S. R., Huang, C. G., Chan, W. K., ... Lin, T. Y. (2004). Transmission and clinical features of enterovirus 71 infections in household contacts in Taiwan. *JAMA*, 291(2), 222–227 [PubMed: 14722149].

Chen, H., Liu, S., Liu, J., Chai, C., Mao, H., Yu, Z., ... Chen, E. (2016). Nosocomial co-transmission of avian influenza A(H7N9) and A(H1N1)pdm09 viruses between 2 patients with hematologic disorders. *Emerging Infectious Diseases*, 22(4), 598–607 [PubMed: 26982379].

Chow, E. J., & Mermel, L. A. (2017). Hospital-acquired respiratory viral infections: incidence, morbidity, and mortality in pediatric and adult patients. *Open Forum*
