HIV Coinfection Is Associated with Low-Fitness $rpoB$ Variants in Rifampicin-Resistant Mycobacterium tuberculosis

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ABSTRACT We analyzed 312 drug-resistant genomes of Mycobacterium tuberculosis isolates collected from HIV-coinfected and HIV-negative TB patients from nine countries with a high tuberculosis burden. We found that rifampicin-resistant M. tuberculosis strains isolated from HIV-coinfected patients carried disproportionally more resistance-conferring mutations in $rpoB$ that are associated with a low fitness in the absence of the drug, suggesting these low-fitness $rpoB$ variants can thrive in the context of reduced host immunity.

KEYWORDS HIV-TB coinfection, Mycobacterium tuberculosis, drug resistance, fitness cost, rifampicin

Tuberculosis (TB), caused by members of the Mycobacterium tuberculosis complex, is a leading cause of death worldwide, killing more people than any other infectious disease. Among the many factors driving the global TB epidemics, two factors stand out as particularly important: antibiotic resistance and HIV coinfection (1). Although the impact of both of these factors individually is well recognized, the interaction between them is less clear and likely depends on the particular epidemiologic setting (2). HIV coinfection and drug-resistant TB often coexist in severe epidemics, which indicates
spread of drug-resistant *M. tuberculosis* strains from immunocompromised patients (3–5). The propensity of drug-resistant *M. tuberculosis* strains to spread is influenced by the fitness cost associated with drug resistance determinants (6). Specifically, bacterial strains that have acquired drug resistance-conferring mutations may be less transmissible than their susceptible counterparts, although this fitness cost can be ameliorated by compensatory mutations (7–10). Moreover, the effect of different resistance-conferring mutations on fitness can be heterogeneous (11). In the clinical setting, there is a selection for high-fitness and/or compensated drug-resistant *M. tuberculosis* strains in TB patients (12). However, in immunocompromised hosts, such as HIV-coinfected patients, even strains with low-fitness resistance mutations might propagate efficiently (13–15), which could partially explain why drug-resistant TB has been associated with HIV coinfection (16, 17). However, to date, no evidence directly supports the notion that the immunological environment created by HIV coinfection modifies the fitness of drug-resistant *M. tuberculosis* (5, 18, 19).

In this study, we tested the hypothesis that resistance-conferring mutations with low fitness in *M. tuberculosis* are overrepresented among HIV-coinfected TB patients. We focused our analysis on isoniazid and rifampicin, the two most important first-line anti-TB drugs, for which resistance-conferring mutations have been shown to differ in their fitness effects when measured in the laboratory (11). In addition, the frequency of the resistance alleles found in a clinical setting correlates well with the *in vitro* fitness of strains (12, 20). To explore the association between HIV coinfection and the fitness effect of different drug resistance-conferring mutations in *M. tuberculosis*, we compiled a collection of drug-resistant strains using the global International Epidemiology Databases to Evaluate AIDS (IeDEA, http://www.iedea.org) consortium (21, 22) as a platform. For this study, 312 strains were collected from HIV-coinfected and HIV uninfected TB patients originating from nine countries on three continents: Peru, Thailand, South Africa, Kenya, Côte d’Ivoire, Botswana, Democratic Republic of the Congo, Nigeria, and Tanzania (Fig. 1; see also Table S1 in the supplemental material). The association between the fitness of isoniazid resistance-conferring mutations and HIV coinfection was tested in a univariate analysis (Fig. S1). Isoniazid resistance-conferring mutations were divided into three groups, as previously described (23): *katG* S315T mutation, *katG* mutations other than S315T, and *inhA* promoter mutations only. The S315T substitution in *katG* causes high-level isoniazid resistance while retaining some catalase/peroxidase functions (24). Conversely, the *inhA* promoter mutation does not affect KatG activity. Other substitutions/deletions in *katG* have been associated with a lower fitness in the laboratory and are observed only rarely among clinical isolates (23, 25, 26). In the case of rifampicin, the association between the fitness of *rpoB* variants and HIV coinfection was tested in both a univariate and multivariate analysis (Table 1). Resistance-conferring variants in *rpoB* were classified into two groups based on their fitness effects documented previously (11, 20, 27). The mutation *rpoB* S450L was considered high fitness, since this mutation was previously shown to confer a low fitness cost in the laboratory (11) and is generally the most common in clinical strains (28). Any other resistance-conferring variant affecting *rpoB* was considered low fitness (11). The multivariable logistic regression model with outcome of low-fitness *rpoB* variants was adjusted for host-related factors (history of TB, country of isolation, sex, and age) (29) and bacterial factors (*M. tuberculosis* lineage, presence of an *rpoA-C* compensatory mutation, clustering of the genome inferred by genetic relatedness). Seventy-six patients from Tanzania and Botswana were excluded from the model due to missing or unknown clinical data (see the supplemental methods file).

Out of 312 patients, 113 (36.2%) were HIV coinfected, 120 (38.5%) were women, 115 (36.9%) were newly diagnosed TB cases (therefore, treatment naive), 276 (88.5%) harbored isoniazid resistance-conferring mutations, with or without additional resistance, and 282 (90.4%) harbored rifampicin resistance-conferring mutations, with or without additional resistance. In total, 78.8% (*n* = 246) of the strains were classified as being at least multidrug resistant, defined as resistance to isoniazid and rifampicin with or without additional resistance to second-line drugs. Among the 113 HIV-coinfected
individuals, 34 (30%) were on antiretroviral therapy (ART), 26 (23%) were not, and 53 (47%) had an unknown ART start date. Four of the eight known \textit{M. tuberculosis} lineages were represented in the following proportions: 11 L1 (3.5%), 57 L2 (18.3%), 38 L3 (12.2%), and 206 L4 (66.0%). After dividing a total of 276 isoniazid-resistant strains into the three groups of isoniazid resistance-conferring mutations defined above, we found similar proportions in HIV-coinfected and HIV-uninfected patients (chi-square test, $P = 0.54$; Fig. S1), and, as expected, the \textit{katG} S315T mutation was the most frequent mutation in both categories (overall, found in 80% of isoniazid-resistant strains). In the case of rifampicin resistance, a univariate and multivariate analysis of 203 strains with complete clinical records indicated that HIV-coinfected TB patients carried a higher proportion of low-fitness \textit{rpoB} resistance variants than HIV-negative patients (72.3%
The univariate analysis showed higher odds of having a low-fitness rpoB variant in HIV-coinfected patients (odds ratio, 2.46 [95% confidence interval, 1.30 to 4.66], \( P = 0.006 \)) (Table 1). Our multivariable regression analysis confirmed these results and showed an association between low-fitness rpoB variants and HIV coinfection while controlling for other factors (odds ratio, 4.58 [95% confidence interval, 1.69, 12.44], \( P = 0.003 \)) (Table 1). This association can be explained in at least two ways. First, HIV-coinfected patients are thought to have fewer lung cavities on average and lower sputum bacillary load (30, 31). The resulting smaller M. tuberculosis population size would lead to fewer replication events, possibly reducing the number of mutations available for selection to act upon. In other words, low-fitness variants and high-fitness variants would co-occur less often in an HIV-coinfected patient, such that competition between them would be less likely. This scenario would be relevant for de novo acquisition of low-fitness drug-resistant variants within an HIV-coinfected patient. Second, following the transmission of a drug-resistant strain with low fitness to a host with reduced immunity, weaker immune pressure acting on this strain might lead to better bacterial survival. The association between low-fitness rpoB variants and HIV coinfection remained significant even after adjusting for the different epidemiologic settings (i.e., countries) and the strain genetic background (i.e., M. tuberculosis lineages). We also observed that strains carrying the rpoB S450L resistance-conferring mutation

### Table 1

Results of the univariate and multivariate analysis showing host and bacterial factors associated with low fitness rpoB variants in 203 TB patients

| Parameter for fitness of rpoB variants | No. (%) of patients by fitness level | Univariable | Multivariable |
|----------------------------------------|-------------------------------------|-------------|--------------|
|                                        | Low                    | High        | OR (95% CI) | \( P \) value | OR (95% CI) | \( P \) value |
| HIV status                             |                        |             |             |             |             |
| HIV                                    | 71 (51.4)              | 67 (48.6)   | Reference   | Reference   |             |             |
| HIV \( ^\ast \)                        | 47 (72.3)              | 18 (27.7)   | 2.46 (1.30–4.66) | 0.006       | 4.58 (1.69–12.44) | 0.003       |
| Presence of a compensatory mutation in rpoA-C |                   |             |             |             |             |
| No                                     | 117 (71.3)             | 47 (28.7)   | Reference   | Reference   |             |             |
| Yes                                    | 1 (2.6)                | 38 (97.4)   | 0.01 (0.00–0.08) | < 0.0001    | 0.01 (0.00–0.06) | < 0.0001    |
| M. tuberculosis lineage                |                        |             |             |             |             |
| 2                                      | 16 (44.4)              | 20 (55.6)   | Reference   | Reference   |             |             |
| 4                                      | 99 (61.5)              | 62 (38.5)   | 2.00 (0.96–4.14) | 0.06       | 3.10 (0.94–10.21) | 0.06       |
| Other (L1 or L3)                       | 3 (50.0)               | 3 (50.0)    | 1.25 (0.22–7.05) | 0.80       | 0.97 (0.11–8.31) | 0.98       |
| Clustering of the genome               |                        |             |             |             |             |
| No                                     | 109 (59.6)             | 74 (40.4)   | Reference   | Reference   |             |             |
| Yes                                    | 9 (45.0)               | 11 (55.0)   | 0.56 (0.22–1.41) | 0.21       | 1.05 (0.28–3.90) | 0.94       |
| Country of isolation                   |                        |             |             |             |             |
| South Africa                           | 29 (55.8)              | 23 (44.2)   | Reference   | Reference   |             |             |
| Democratic Republic of Congo           | 11 (37.9)              | 18 (62.1)   | 0.48 (0.19–1.23) | 0.13       | 0.39 (0.12–1.34) | 0.14       |
| Côte d’Ivoire                          | 35 (79.5)              | 9 (20.5)    | 3.08 (1.24–7.70) | 0.02       | 2.04 (0.58–7.23) | 0.27       |
| Kenya                                  | 4 (66.7)               | 2 (33.3)    | 1.59 (0.27–9.44) | 0.61       | 0.94 (0.10–8.42) | 0.96       |
| Nigeria                                | 20 (58.8)              | 14 (41.2)   | 1.13 (0.47–2.72) | 0.78       | 1.00 (0.29–3.40) | 0.99       |
| Peru                                   | 16 (53.3)              | 14 (46.7)   | 0.91 (0.37–2.23) | 0.83       | 1.49 (0.33–6.70) | 0.60       |
| Thailand                               | 3 (37.5)               | 5 (62.5)    | 0.48 (0.10–2.20) | 0.34       | 0.42 (0.07–2.65) | 0.36       |
| Age                                    |                        |             |              |             |             |
| Mean (SD)                              | 32.5 (10.4)            | 34.3 (12.3) | 0.99 (0.96–1.01) | 0.25       | 0.97 (0.94–1.01) | 0.10       |
| Sex                                    |                        |             |              |             |             |
| Female                                 | 47 (59.5)              | 32 (40.5)   | Reference   | Reference   |             |             |
| Male                                   | 71 (57.3)              | 53 (42.7)   | 0.91 (0.51–1.62) | 0.75       | 0.77 (0.34–1.71) | 0.52       |
| History of TB disease                  |                        |             |              |             |             |
| No                                     | 35 (52.2)              | 32 (47.8)   | Reference   | Reference   |             |             |
| Yes                                    | 83 (61.0)              | 53 (39.0)   | 1.43 (0.79–2.58) | 0.23       | 0.96 (0.34–2.73) | 0.94       |

\(^{a}\)Number of observations in model, 203; CI, confidence interval. The odds ratios and \( P \) values were obtained from the regression model.
were more likely to also carry a compensatory mutation in rpoA-C (97.4% versus 2.6%) (Table 1). Even though this phenomenon seems counterintuitive, it has been described multiple times (7, 9, 32–34) and, thus, might point to different mechanisms of compensation in strains carrying resistance mutations other than rpoB S450L. In addition, in our study, L4 strains were associated with low-fitness rpoB variants compared to L2 (odds ratio, 3.10 [95% confidence interval, 0.94, 10.21], P = 0.06) (Table 1), indicating that the strain genetic background plays a role in shaping the cost of resistance, as was previously shown for other bacterial species (35) and for other drugs (36). In the regression analysis, we had several categorical variables with only a few observations. Therefore, statistical power, especially for country of isolation, was low, and the results should be interpreted with care.

HIV-coinfected TB patients are generally thought to have a reduced potential for TB transmission (30, 37), because these patients have reduced formation of lung cavities, more extrapulmonary disease, and a shorter period of infectiousness due to earlier diagnosis or higher mortality, especially in the absence of antiretroviral treatment and if antibiotic resistance is already present (4). Based on the overrepresentation of low-fitness rpoB mutations in the context of HIV coinfection, one would expect a further reduction of the transmission potential of drug-resistant TB in this context. However, outbreaks of drug-resistant TB in HIV-coinfected patients have been reported (3). Such outbreaks might be explained by (i) a higher risk of M. tuberculosis infection and reinfection due to diminished host immunity, (ii) on-going transmission of drug-resistant M. tuberculosis from a larger pool of immunocompetent TB patients to immunocompromised patients, (iii) transmission occurring in conducive environments, such as health care settings, where both HIV-coinfected individuals and drug-resistant TB patients are more likely to coexist, and (iv) M. tuberculosis strains carrying high-fitness drug resistance mutations.

In summary, using a global sample of drug-resistant M. tuberculosis clinical strains from HIV-coinfected and HIV-negative TB patients, we showed that low-fitness rpoB variants were overrepresented in HIV-coinfected patients, and that this association was independent from other potential confounding factors. Taken together, our results provide new insights into how HIV coinfection can impact the fitness of drug-resistant M. tuberculosis.

Data availability. The M. tuberculosis whole-genome sequences from the patients are available on NCBI under several project identifiers. The accession number for each genome is indicated in Supplemental Table S1.

SUPPLEMENTAL MATERIAL
Supplemental material is available online only.
SUPPLEMENTAL FILE 1, PDF file, 0.1 MB.
SUPPLEMENTAL FILE 2, XLSX file, 0.03 MB.

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