A critical reappraisal of dietary practices in methylmalonic acidemia raises concerns about the safety of medical foods. Part 2: cobalamin C deficiency

Irini Manoli, MD, PhD1, Jennifer G. Myles, MS, RD2, Jennifer L. Sloan, PhD, MS1, Nuria Carrillo-Carrasco, MD1,3, Eva Morava, MD, PhD4, Kevin A. Strauss, MD5, Holmes Morton, MD5 and Charles P. Venditti, MD, PhD1

Purpose: Cobalamin C (cblC) deficiency impairs the biosynthesis of 5′-deoxyadenosyl-adenosyl- and methyl-cobalamin, resulting in methylmalonic acidemia combined with hyperhomocysteinemia and hypomethioninemia. However, some patients with cblC deficiency are treated with medical foods, devoid of methionine and high in leucine content, that are formulated for patients with isolated propionate oxidative defects. We examined the effects of imbalanced branched-chain amino acid intake on growth outcomes in cblC-deficient patients.

Methods: Dietary intake was correlated with biochemical, anthropometric, and body composition measurements and other disease parameters in a cohort of 28 patients with early-onset cblC deficiency.

Results: Protein-restricted diets were followed by 21% of the patients, whereas 32% received medical foods. Patients on protein-restricted diets had lower height-for-age Z-score (P = 0.034), whereas patients consuming medical foods had lower head circumference Z-scores (P = 0.037), plasma methionine concentrations (P = 0.001), and predicted methionine influx through the blood–brain barrier Z-score (−1.29 vs. −0.0617; P = 0.007). The combination of age at diagnosis, a history of seizures, and the leucine-to-valine dietary intake ratio best predicted head circumference Z-score based on multiple regression modeling (R² = 0.945).

Conclusions: Patients with cblC deficiency treated with medical foods designed for isolated methylmalonic acidemia are at risk for iatrogenic methionine deficiency that could adversely affect brain growth and development.

Key Words: cobalamin C (cblC) deficiency; dietary guidelines; medical foods; methionine; methylmalonic acidemia

INTRODUCTION

Combined methylmalonic acidemia and hyperhomocysteinemia, cobalamin C (cblC) type, results from impaired biosynthesis of 5′-deoxyadenosyl- and methyl-cobalamin, cofactors for the methylmalonyl-CoA mutase and methionine synthase enzymes, respectively.1–3 Although the biochemical phenotype of this disorder shares elevated methylmalonic acid concentrations with the classic mut subtype of isolated methylmalonic acidemia (MMA), the propionate-derived metabolic perturbations are of lesser magnitude, whereas the clinical findings and disease pathophysiology are distinct. CblC-deficient patients display significantly increased homocysteine and decreased methionine concentrations, and patients with early onset present with failure to thrive, visual impairment due to macular degeneration/pigmentary retinopathy, hemolytic uremic syndrome and thromboembolic/microangiopathic complications, cardiopulmonary findings, neurocognitive delay, seizures, and other neurological manifestations.4–6 Decreased methionine synthesis and secondary impairments of the remethylation cycle, methyltransfer reactions, homocysteine–thiolactone metabolism, and increased oxidative stress have been implicated in disease pathophysiology.5,8–11 The mechanisms underlying the multisystemic manifestations of cblC deficiency remain unknown, in part because of the lack of viable animal models.12

Treatment remains particularly challenging in cblC-deficient patients because symptoms such as intrauterine growth retardation, microcephaly, mild craniofacial dysmorphism, and congenital heart disease can develop in utero.13–16 Moreover, despite early diagnosis by newborn screening, clinical manifestations, including the characteristic ocular and neurological complications, are at best incompletely responsive to conventional therapies.6,7,9 The mainstay of treatment involves high-dose intramuscular hydroxocobalamin injections that can ameliorate the biochemical perturbations, resulting in a decrease of homocysteine and methylmalonic acid concentrations and an increase in methionine synthesis. Betaine supplementation also enhances remethylation of homocysteine to methionine via the betaine–homocysteine methyltransferase reaction. Folinic

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1Organic Acid Research Section, Genetics and Molecular Biology Branch, National Human Genome Research Institute, National Institutes of Health, Bethesda, Maryland, USA; 2Nutrition Department, Clinical Center, National Institutes of Health, Bethesda, Maryland, USA; 3Therapeutics for Rare and Neglected Diseases, National Center for Advancing Translational Sciences, National Institutes of Health, Bethesda, Maryland, USA; 4 Tulane University Medical School, New Orleans, Louisiana, USA; 5 Clinic for Special Children, Strasburg, Pennsylvania, USA. Correspondence: Charles P. Venditti (venditti@mail.nih.gov)

Submitted 3 March 2015; accepted 22 June 2015; advance online publication 13 August 2015. doi:10.1038/gim.2015.107
or folic acid and other supplements (carnitine, methylcobalamin, pyridoxine, methionine) have been used, yet longitudinal efficacy studies of disease-related clinical outcomes remain uncertain.9

Protein restriction can be considered to decrease the pro¬piogenic amino acid load (valine, isoleucine, threonine, and methionine) and help lower methylmalonic acid production.17 However, methylmalonic acid concentrations in cblC-deficient patients are typically orders of magnitude less than those seen in isolated patients with mut MMA, consistent with the observations that metabolic decompensations of the “intoxication” type seen in classic isolated MMA are very rare in patients with cblC deficiency.18,19 Furthermore, protein restriction carries the risk of inducing iatrogenic methionine, and methyl-donor (S-adenosylmethionine) deficiency, resulting in the impairment of multiple biological transmethylation reactions such as creatine, phosphatidylcholine, and sphingomye¬lin biosynthesis, as well as DNA methylation.9,20–22 This could adversely affect growth and neurodevelopmental outcomes, as evidenced by the severe neurological phenotypes variably seen in patients with other remethylation disorders, including 5′,10′-methylene-tetrahydrofolate reductase deficiency, functional methionine synthase (cblE/G) deficiencies, or other intra¬cellular cobalamin biosynthesis disorders (cblD, F, I).12,23,24

Over the course of several years of evaluating cblC-deficient patients via a natural history protocol, we observed a highly variable approach to the dietary management of cblC deficiency. Dietary practices ranged from following a completely unrestricted diet to prescribing strict protein restriction, supplemented by large quantities of medical foods. The surpris¬ingly large number of cblC-deficient patients who are managed with medical foods designed for isolated MMA and propionic acidemia (PA) in our study and the published literature6 led us to critically reappraise the dietary management of cblC deficiency. We find that the use of medical foods in cblC-deficient patients is associated with iatrogenic methionine and essential branched-chain amino acid deficiencies, impaired growth, and abnormal body composition.

**METHODS**

**Study population**

Patient studies were approved by the National Human Genome Research Institute Institutional Review Board and performed in compliance with the Helsinki Declaration (ClinicalTrials.govNCT00078078). Patients were enrolled between 2004 and 2014 from centers across the United States and Canada. Long-term dietary management was dictated by each patient’s metabolic center. Study participants had no clinical symp¬toms or laboratory markers of metabolic instability during their National Institutes of Health evaluations. The diagnosis of cblC deficiency was confirmed using cellular biochemistry (in the laboratory of Dr. David S. Rosenblatt, Division of Medical Genetics, McGill University, Montreal, Canada) and/or through molecular genetic analysis of the MMAHC gene (GeneDx, Gaithersburg, MD).25

**Laboratory studies**

The National Institutes of Health Clinical Center laboratory performed routine laboratory investigations, including total homocysteine and vitamin B12 concentrations. Total homocys¬teine was determined by high-performance liquid chromatog¬raphy between 2004 and 2013 and by an enzymatic assay (Cobas 6000 analyzer; Roche Diagnostics Indianapolis, IN) from 2013 to present. Plasma and urine methylmalonic acid and amino acid concentrations were determined by liquid chromatogra¬phy–tandem mass spectrometry (Mayo Medical Laboratories Rochester, MN). Samples were obtained in the fasting state, or 2–4 hours after a meal, whenever possible.

**Dietary regimens and daily amino acid intake calculations**

Dietary analysis was performed using the patient’s medical foods (specialized formula) prescriptions and a detailed diet history obtained by a research dietitian. A 3-day food record completed by the families was available for 16 of 28 patients.

Dietary analyses and calculations were performed using Nutrition Data System for Research software versions 2007–2012 developed by the Nutrition Coordinating Center (University of Minnesota, Minneapolis, MN).26 Medical food composition information was obtained from the respective manufacturers. Recommended dietary allowance (RDA) for protein and amino acids were based on the dietary refer¬ence intakes for protein and amino acids.27 Protein intake was recorded as complete protein from natural protein sources and incomplete/deficient protein-equivalent intake (grams/kilogram/day) in the form of specialized MMA/PA medical foods, such as Propimex1/2 (Abbott Laboratories, Columbus, OH); XMTVI Analog; Maxamaid or Maxamum (Nutricia North America, Cedar Knolls, NJ); and MMA/PA Express (VitaFlo USA, Alexandria, VA). Amounts of individual pro¬piogenic amino acids (valine, isoleucine, methionine, and threonine), as well as leucine, were recorded for patients with detailed dietary records (n = 13), whereas estimated deficient versus complete protein intake (grams/kilogram/day) was doc¬umented for all patients.

Patients received intramuscular hydroxocobalamin therapy with different therapeutic dosages, ranging between 0.01 and 0.61 mg/kg/day. All patients received betaine at dosages rang¬ing from 31 to 338 mg/kg/day, as well as folate or folic acid supplementation, whereas a minority were prescribed aspirin, fish oil, docosahexaenoic acid (DHA), or other dietary supple¬ments. These data are provided for each patient, along with additional nutritional information, in Supplementary Table S1 online.

**Predicted cerebral amino acid influx calculations**

Cerebral influx of substrates for the cationic (y+; lysine, arginine, ornithine) and large neutral amino acid transport systems (LAT1; glutamine, histidine, isoleucine, leucine, methionine, phenylalanine, threonine, tryptophan, tyrosine, and valine) was estimated as previously described for patient cohorts with 5′,10′-methylene-tetrahydrofolate reductase deficiency, maple
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syrup urine disease, and glutaric acidemia. These calculations were based on the principle that the cerebral uptake of each amino acid is influenced by the ambient (plasma) concentrations of its competitors. Substrate competition is expressed by an apparent \( K_m \) called \( K_{app} \) (\( \mu \text{mol} / \text{L} \)), calculated for each amino acid according to the equation: \( K_{app} = K_m [1 + \frac{1}{n} (C/K_i) ] \). \( K_m \) is the classical Michaelis–Menten affinity constant for the single amino acid of interest, \( C \) is the plasma concentration (micromoles/l) for each of \( n \) competitors, and \( K_i \) is the classical affinity constant of that competitor (micromoles/l). The \( K_{app} \) value then was used to calculate the brain influx (nanomoles/minute/gram of brain tissue) of each amino acid in the competing group, according to the equation: Influx = \( \frac{V_{max}}{K_{app} + C} \), where \( V_{max} \) and \( C \) are the maximal transport velocity (nanomoles/gram/minute) and plasma concentration (micromoles/l), respectively, of each amino acid. Estimated brain influx Z-score values were calculated using mean values observed in a control population of 52 children not affected by disorders of amino acid, folic acid, or cobalamin metabolism from the Clinic for Special Children (Strasburg, PA).

Anthropometry, body composition, and neurocognitive evaluations

Anthropometric measurements were expressed as age- and sex-specific Z-scores using the epidemiological software package Epi Info (http://www.cdc.gov/epiinfo/), based on the Centers for Disease Control and Prevention 2002 reference database (Atlanta, GA) for patient under 20 years of age. Whole-body composition in grams of fat or fat mass, and fat-free (or lean) body mass, were measured using dual-energy X-ray absorptiometry (Delphi A; Hologic, Bedford, MA). For patients between 5 and 23 years old, a height-adjusted Z-score was calculated using the Bone Mineral Density in Childhood Study online calculator (http://www.bmdcspublic.com/zscore.htm).

Data analysis

The results are presented as means (±SD). Significance was set at \( P < 0.05 \). Statistical manipulations were performed using SPSS version 21.0 (IBM, Chicago, IL) or GraphPad Prism software version 6.0 (GraphPad Software Inc., San Diego, CA). An independent Student t-test was used to compare values between patients who had or had not been receiving medical foods. Pearson correlation coefficient and multiple linear regression analyses were used to evaluate correlations between variables. Dummy variables were generated for dichotomous categorical variables such as sex, seizures, or congenital microcephaly to be used in the regression model.

RESULTS

Anthropometric characteristics and body composition

We studied 28 patients with early-onset cblC deficiency (18 male, 10 female; age range: 2.03–27.04 years; mean age ± SD: 10.2 ± 7.61 years). Age at diagnosis varied widely, from the first week of life (\( n = 4 \)) to 24 months (mean ± SD: 2.62 ± 4.44 months; median: 1.5 months). Seventeen of 28 patients (60%) were homozygous for the common c.271dupA, p.R91KfsX14 mutation in the MMACHC gene, which is associated with a severe early-onset phenotype (Table 1). Height, weight, body mass index, and head circumference (occipitofrontal circumference (OFC))–for–age Z-scores (mean ± SD) are presented (Table 1). Mean Z-score for height was –1.04 ± 1.33 (range: 1.8 to −3.5); weight, –0.66 ± 1.83 (range 3.00 to −6.8); body mass index, 0.13 ± 1.47 (range: 2.6 to −4.4); and OFC, −1.57 ± 1.23 (range: 0.32 to −4.40; median: −1.06). Height and OFC Z-scores less than −2.00 were found in six and seven patients, respectively. Of note, three of the patients had a history of congenital microcephaly, whereas 9/28 (32.14%) had a history of seizures, 2 of whom had a diagnosis of infantile spasms (Table 1). Growth outcomes were not different between male and female subjects, or between patients homozygous for the c.271dupA mutation versus others.

Whole-body composition analysis based on dual-energy X-ray absorptiometry imaging studies was available for 18 patients (12 males and 6 females). Mean ± SD percentage of fat mass was 31.56 ± 11.2%, with 27.9 ± 8.8% in males and 38.11 ± 12.85% in females (\( P = 0.07 \)). Mean bone mineral density height-adjusted Z-score was −1.9 ± 1.55, with −1.67 ± 0.93 for males and −2.46 ± 2.77 for females (\( P = 0.057 \)).

Correlations of growth outcomes with protein restriction and medical foods utilization

Six of 28 (21%) of the patients received complete protein below 85% of the RDA for healthy children. Nine of the 28 patients (32.14%) received medical foods designed for MMA. Additional protein-free (\( n = 5 \)) or elemental amino acid formulas (\( n = 2 \)) were used in a minority of patients, whereas one patient required valine supplementation for persistently low valine plasma concentrations (Table 1).

Comparing growth and biochemical parameters between patients following a protein-restricted diet or receiving medical foods versus those who were not, it became evident that protein restriction or chronic medical food use were associated with poorer growth parameters. Patients following a diet with less than 85% the RDA for complete protein had lower height-for-age (−2.16 ± 1.04 vs. −0.72 ± 1.24; \( n = 5 \) vs. 17; \( P = 0.030 \)) and head circumference (OFC)–for–age Z-scores (−2.3 ± 1.30 vs. −1.2 ± 1.11; \( P = 0.062 \)) and a higher percentage of fat mass (and hence lower lean or skeletal muscle mass) by dual-energy X-ray absorptiometry analysis (43.1 ± 10.8 vs. 26.7 ± 7.3%; \( n = 12 \) and 5; \( P = 0.002 \)). Similarly, a lower mean weight-for-age (−1.7 ± 2.28 vs. −0.09 ± 1.3; \( P = 0.038 \); \( n = 15 \) and 8, respectively), height-for-age (−1.72 ± 1.01 vs. −0.73 ± 1.37; \( P = 0.075 \)), and head circumference–for–age Z-score (−2.48 ± 1.43 vs. −1.15 ± 0.88; \( P = 0.037 \)) was observed in the subset of patients consuming medical foods (Figure 1a).

Patients with a history of seizures had a significantly lower height and OFC Z-score than those without seizure history (\( P = 0.004 \) and <0.001, respectively) (Figure 1b). Growth outcomes (height or OFC Z-scores) in patients who carried no, one, or two copies of the common c.271dupA mutation were
Table 1: Clinical, molecular and biochemical features of subjects with cblC deficiency

| Age at evaluation (years) | Age at diagnosis (months) | MMAACHC | Z-scores | Complete protein %RDA | Deficient protein (g/kg/day) | Plasma Met (μmol/l) | Predicted LAT1 Met influx Z-score |
|--------------------------|---------------------------|---------|----------|-----------------------|-----------------------------|-------------------|----------------------------------|
| 2.03                     | 4                         | c.271dupA, p.R91KfsX14 | —        | 118.9                 | 0.20                        | 19                | -0.36                            |
| 2.04                     | 0.3⁴                      | c.271dupA, p.R91KfsX14 | —        | 61.0                  | 1.42                        | 2                 | -2.89                            |
| 2.18                     | 0.5                       | c.271dupA, p.R91KfsX14 | 3        | 0                     | 41                          | 1.69              |                                  |
| 2.63                     | 1.5                       | c.331C>T, p.R111X    | -2.2     | -2.1                  | -4.4⁴                       | 133.4             | 1.50 n/a                         |
| 2.67                     | 0.1¹                      | c.271dupA, p.R91KfsX14 | 0.8      | 0.2                   | 0.32                        | 369.7             | 0                                |
| 3.14                     | 0.5                        | c.271dupA, p.R91KfsX14 | 1.7      | -0.1                  | -0.07                       | 153.2             | 0                                |
| 3.20                     | 0.3⁴                      | c.271dupA, p.R91KfsX14 | -1.2     | -1.5                  | n/a                         | 100.4             | 0                                |
| 3.25                     | 0.25³                     | c.619dupG, p.D207GfsX38| -0.16    | 0.55                  | -0.88                       | 315.9             | 0                                |
| 3.76                     | 0.8⁵                      | c.271dupA, p.R91KfsX14 | -2.48    | -2.1                  | -3.75⁵                       | 56.1⁴             | 0.45                            | -1.68                            |
| 4.18                     | 1.1                       | c.271dupA, p.R91KfsX14 | 0.3      | 0.4                   | -1.69                       | 271.3             | 0                                |
| 4.49                     | 0.6²                      | c.471G>A, p.W157X    | -0.9     | -0.9                  | -0.97                       | 263.5             | 0.61                            |
| 5.25                     | 4                          | c.271dupA, p.R91KfsX14 | -0.9     | -1.4                  | 0.23                        | 134.6             | 0                                |
| 7.25                     | 1.5                       | c.3G>G, p.M17       | 0.1      | -2.9                  | -3.4⁶                       | 56.2              | 0.54                            |
| 8.90                     | 0.1³                      | c.271dupA, p.R91KfsX14 | -1.3     | -2.4                  | -1.25                       | 104.4             | 0                                |
| 9.62                     | 2.5                       | c.271dupA, p.R91KfsX14 | -1.2     | -1.4                  | -0.83                       | 106.7             | 0 n/a                           |
| 9.83                     | 3                          | c.331C>T, p.R111X    | -0.6     | -1.3                  | 0.23                        | 118.9             | 0                                |
| 10.85                    | 1.75                      | c.271dupA, p.R91KfsX14 | -1.9     | -3.5                  | -2.94⁷                       | 86.1              | 0                                |
| 11.11                    | 0.3                       | c.271dupA, p.R91KfsX14 | -0.9     | -0.9                  | -0.57                       | 115.2             | 0                                |
| 12.07                    | 1.2                       | c.271dupA, p.R91KfsX14 | 0.6      | -1.2                  | -0.97                       | 145.8             | 0                                |
| 13.77                    | 0.8                       | c.271dupA, p.R91KfsX14 | 0.8      | 0                     | -0.81                       | 102.6             | 0                                |
| 15.35                    | 2                          | c.271dupA, p.R91KfsX14 | -0.4     | -1.2                  | 0.23                        | 83.8              | 0                                |
| 15.52                    | 24                         | c.271dupA, p.R91KfsX14 | 0.9      | n/a                   | -3.22⁸                       | 74.9              | 0.60                            |
| 19.38                    | 5                          | c.271dupA, p.R91KfsX14 | -6.8     | -2.8                  | -2.4⁹                       | 104.1             | 0.34                            |
| 20.33                    | 1.5                       | c.271dupA, p.R91KfsX14 | —        | —                    | -1.8                        | 90.7              | 0                                |
| 20.79                    | 1.8                       | c.271dupA, p.R91KfsX14 | —        | —                    | 24.8                        | 128.8             | 0.28                            |
| 22.08                    | 5.5                       | c.600G>A, p.W200X    | —        | —                    | ab                          | 132.1             | 0                                |
| 23.04                    | 3                          | c.271dupA, p.R91KfsX14 | —        | —                    | b                           | 129.6             | 0                                |
| 24.10                    | 2.25                      | c.271dupA, p.R91KfsX14 | —        | —                    | —                           | 79.3              | 0                                |

Age at the time of patients' evaluations and age at diagnosis, as well as the molecular genetic analysis of the MMAACHC gene and major variables for the study, are provided for each of the patients with early-onset cblC deficiency. Patients with the lowest methionine influx scores and head circumference (occipitofrontal circumference (OFC)) Z-scores are in bold.

%RDA, percentage of recommended daily allowance; n/a, data not available. Met, methionine; LAT1, large neutral amino acid transporter type 1.

⁴Patients with congenital microcephaly. ⁵Patients with a history of seizures. ⁶Patients with infantile spasms. ⁷A patient requiring valine supplementation for persistently low plasma valine concentrations. ⁸Patients diagnosed by newborn screening.
not significantly different ($P = 0.876$ and $0.591$, respectively, one-way analysis of variance), suggesting nongenetic effects have a significant contribution to the variance observed. Similarly, no significant difference was observed in the percentage RDA of natural protein intake or the use of medical foods (grams/kilograms/day) between patients with early-onset cblC deficiency who have different genotypes.

Complete protein intake (percentage RDA) showed a positive correlation with the height-for-age $Z$-score ($r = 0.705$). Given that MMA/PA medical foods contain no propiogenic amino acid, but a normal to increased amount of leucine, ratios of leucine to methionine or valine dietary intake were significantly elevated in patients consuming medical foods ($P = 0.015$ and $0.003$, respectively). A higher leucine-to-valine dietary intake showed a negative correlation to the height-for-age $Z$-scores ($r = -0.673$; $P = 0.033$; $R^2 = 0.453$) and OFC-for-age $Z$-scores ($r = -0.840$; $P = 0.001$; $R^2 = 0.705$).

**Figure 1** Growth parameters, serum metabolites and predicted amino acid influx through the blood brain barrier. Association with medical food vs. complete protein intake, history of seizures. (a) A lower weight-for-age ($−1.7 ± 2.28$ vs. $−0.09 ± 1.3$; $P = 0.038$; $n = 15$ and $8$, respectively) and head circumference–for–age $Z$-score ($−2.48 ± 1.43$ vs. $−1.15 ± 0.88$; $P = 0.037$) and a trend towards a lower mean height-for-age ($−1.72 ± 1.01$ vs. $−0.73 ± 1.37$; $P = 0.075$) and body mass index–for–age $Z$-score ($−0.60 ± 0.08$ vs. $0.48 ± 0.99$; $P = not significant$) was observed in the group consuming medical foods compared with patients who did not at the time of evaluation. (b) Patients with a history of seizures had a significantly lower height and OFC $Z$-scores than those without seizure history ($P = 0.004$ and $<0.001$, respectively). (c) Complete protein intake as a percentage of the recommended daily allowance (RDA) correlated positively with height-for-age $Z$-score ($r = 0.575$; $P = 0.008$; $R^2 = 0.31$). (d) Patients consuming medical foods had slightly lower total homocysteine concentrations ($48.7 ± 27.7$ vs. $59.8 ± 25.2$; $P = not significant$), significantly lower plasma methylmalonic acid ($2.75 ± 3.27$ vs. $13.9 ± 16.4$; $n = 9$ vs. $18$; $P = 0.012$) and methionine concentrations ($14.5 ± 6.92$ vs. $25.8 ± 8.8$; $P = 0.004$), as expected; and lower valine and isoleucine concentrations ($P < 0.001$ and $0.026$, respectively; data not shown), compared with patients not consuming medical foods. (e) Predicted influx through the blood–brain barrier transporter LAT1 for the propiogenic amino acids and leucine is provided as a $Z$-score from mean values derived from plasma amino acid concentrations in a control population of 52 children from the Clinic for Special Children. Patients consuming medical foods had mean predicted influx $Z$-scores lower than those not consuming special formulas (valine: $−2.11 ± 0.92$ vs. $−0.47 ± 0.56$; $P = 0.001$; isoleucine: $−1.41 ± 1.09$ vs. $−0.45 ± 0.56$; $P = 0.045$; methionine: $−1.29 ± 0.90$ vs. $−0.61 ± 0.89$; $P = 0.004$; and threonine: $−1.33 ± 0.72$ vs. $0.06 ± 0.87$; $P = 0.001$). (f) The leucine-to-methionine plasma concentration ratio correlated negatively with the predicted cerebral methionine influx ($r = −0.749$; $R^2 = 0.541$; $P < 0.001$) as well as the head circumference–for–age $Z$-scores ($r = −0.553$; $R^2 = 0.306$; $P = 0.008$ *$P<0.05$, ** $P<0.01$, ***$P<0.001$).
Intakes of other nutrients that can affect growth and cognitive function, including iron, zinc, and omega 3 fatty acids, were also assessed (Supplementary Table S1 online). All subjects well surpassed the RDA for iron from dietary intake plus supplements. Three subjects had intake of zinc below the RDA, but none of these subjects had growth failure. Nine subjects had alpha-linolenic acid below the adequate intake. Two of the nine were taking DHA or fish oil supplements to compensate; the remaining seven had negligible DHA intake.

**Biochemical indexes and cerebral amino acid influx calculations**

Since protein restriction and medical food supplementation are both used to improve metabolic control, we looked at the effects of the dietary management on plasma metabolite concentrations. Patients who received medical foods had significantly lower plasma methyalmalic acid and methionine concentrations (Figure 1d), as well as lower valine and isoleucine (data not shown). Total homocysteine was not significantly different. The higher ratio of leucine over methionine, valine, and isoleucine dietary intake also resulted, predictably, in elevated leucine/methionine (P = 0.05) and leucine/valine (P = 0.002) plasma amino acid concentrations (similarly, elevated ratios of tyrosine or phenylalanine to any of the restricted amino acids were observed).

These skewed plasma amino acid ratios are predicted to alter amino acid transport across cells and physiological barriers through competitive inhibition at shared transporters/transport systems, such as the leucine–preferring large neutral amino acid transporter type 1, LAT1 (SLC7A5). To investigate whether these skewed ratios would affect the influx of amino acids at the blood–brain barrier (BBB) via the LAT1 transporter, we applied the cerebral influx model using plasma amino acid concentrations from our cblC-deficient patient cohort.30 We document Z-scores for methionine influx ranging from 1.69 to −2.89, with 18/28 (64%) having a negative Z-score, and 5/27 (18%) having a score <−1.00 (Table 1). The range of values for valine (0.37 to −3.97), isoleucine (0.51 to −3.42), and threonine (1.71 to −2.65) are also reported. These Z-scores were significantly lower in patients receiving medical foods compared with those who did not (Figure 1e), whereas methionine (and valine) BBB influx Z-scores was significantly lower among patients with a history of seizures (for methionine: −0.035 for no seizure history (n = 16) vs. −1.087 in patients with seizures (n = 10); P = 0.012). The leucine-to-methionine plasma concentration ratio correlated negatively with the predicted cerebral methionine influx (P < 0.001) as well as the head circumference–for–age Z-scores (P = 0.008) (Figure 1f).

We present an illustrative case study of an infant, who presented with severe encephalopathy and hyperammonemia and was therefore treated with protein restriction and MMA/PA medical foods, along with daily intramuscular hydroxyocobalamin and adequate doses of betaine and carnitine. The infant showed a decline in head size and length centiles, from the 33rd to the 2nd centile for the head circumference and from the 60th to the 8th for length, with no significant improvement in growth until the age of 5 months. This trend was reversed upon increasing the relative ratio of natural to deficient protein intake and changing briefly to a maple syrup urine disease formula (containing minimal amounts of branched-chain amino acids but normal concentrations of methionine) (Supplementary Figure S1 online).

Stepwise multiple regression modeling showed that the combination of age at diagnosis (β coefficient = −0.295; P = 0.031), history of seizures (β coefficient = −0.324; P = 0.034), and leucine-to-valine intake ratio (β coefficient = −0.754; P < 0.001) best predicted OFC Z-score, with a model R2 of 0.944. The addition of the dietary component (leucine-to-valine intake ratio) improves the R2 of the model from 0.4 to 0.944, suggesting that diet contributes significantly to the variance observed in the OFC Z-score.

**DISCUSSION**

This study detailed the growth outcomes and dietary management of a large cohort of cblC-deficient patients evaluated as part of a dedicated, single-center, natural history protocol (ClinicalTrials.gov NCT00078078). The ability to enroll patients treated at a large number of metabolic clinics in North America afforded a broad representation of the varied therapeutic practices used and the outcomes observed, and provided a large data set of current management approaches.

Protein restriction in cblC deficiency is intended to lower the amino acid load into the propionate oxidation pathway that is partially blocked at the methylmalonyl-CoA mutase (MUT) step as a result of impaired biosynthesis of the cofactor for MUT, 5′-deoxyadenosylcobalamin. Providing an appropriate dose of parenteral hydroxocobalamin usually leads to a significant decrease in plasma methyalmalic acid concentrations, which are generally 10- to 100-fold less than those observed in patients with mut deficiency. Despite the minimal effects on influencing metabolites in the face of hydroxocobalamin therapy, additional protein restriction and administration of propiogenic amino acid–free medical foods, containing minimal amounts of valine, isoleucine, threonine, and, importantly, methionine, were frequently used to treat the patients. In this study we documented that a number of cblC-deficient patients follow a diet restricted in protein (21%) and/or supplemented with medical foods designed for MMA/PA (32%; Table 1). The use of MMA/PA medical foods, although not universally adopted, seems to have gained support in some metabolic centers, as evidenced by the fact that 76% of cblC-deficient patients identified by newborn screening were prescribed medical foods in one study.6 Our data support the concept that protein restriction (and/or medical foods use) to lower the plasma methyalmalic acid concentration has little effect on total homocysteine concentration, but, importantly, it can lower methionine and essential branched-chain amino acid concentrations.

Hypomethioninemia is considered a critical component of cblC disease pathophysiology, and maximizing methionine synthesis, and the downstream provision of methyl groups, is
the aim of the two major therapeutic modalities for the disorder, specifically high-dose vitamin B12 (hydroxocobalamin) and betaine. Brain white matter demyelination and subacute combined degeneration have been observed in several case studies/series of cblC deficiency,\textsuperscript{6,23,31,32} resembling findings in animal models with B12 deficiency. The animals’ symptoms and pathology could be mitigated by methionine supplementation.\textsuperscript{33} Moreover, methionine supplementation was claimed to improve photoreceptor sensitivity (on electroretinography) and retinal function in a single patient, but it had no effect on the characteristic macular degeneration of the disease.\textsuperscript{34} The pathophysiology of cblC deficiency is certainly complex because it cannot be reversed or prevented simply by methionine supplementation or any currently available therapies.\textsuperscript{3,7,9,23} Furthermore, decreased transmethylation products, such as choline, creatine, or guanidinoacetate, have not been consistently aberrant in magnetic resonance spectroscopy or serum/urine biochemical studies in cblC-deficient patients,\textsuperscript{6,35} and a paradoxical increase in S-adenosylmethionine has been noted, postulated to result from secondary inhibition of methylases through adenosylhomocysteine or other metabolites.\textsuperscript{22,35}

Despite the unresolved underlying pathophysiology of cblC deficiency, and the lack of evidence in favor of additional methionine supplementation, a basic therapeutic goal remains to avoid essential amino acid deficiencies, including methionine, especially during periods of brain development. Brain methionine concentrations depend on uptake through the circulation, as well as from de novo methionine synthesis.\textsuperscript{36} Given that MMA/PA medical foods contain no methionine, they may exacerbate methionine deficiency, given the already impaired de novo methionine synthesis that is a feature of cblC deficiency. Therefore, there is no theoretical rationale or experimental support for the practice of administering medical foods to patients with cblC deficiency or any disorder that impairs the remethylation of homocysteine. Furthermore, MMA/PA medical foods contain normal to increased amounts of leucine, which, when administered out of proportion to the other branched-chain amino acids, carries the risk of inducing iatrogenic amino acid deficiencies, probably as a result of alpha ketosocaproic acid (KIC)-induced branched-chain ketoacid dehydrogenase kinase inhibition and increased valine and isoleucine oxidation rates.\textsuperscript{37} Moreover, branched-chain amino acid imbalances exert a multitude of effects in different organ systems, as detailed in our related article (“A critical reappraisal of dietary practices in methylmalonic acidemia raises concerns about the safety of medical foods—Part 1: isolated methylmalonic acidemias”), and summarized in extensive reviews of branched-chain amino acid metabolism.\textsuperscript{28,37,38} The complexity of leucine effects and the tight coupling with isoleucine and valine metabolism have been overlooked in the design of MMA/PA medical foods, perhaps because leucine does not generate propionyl-CoA and is therefore considered “safe” in patients with MMA and PA. In the case of cblC deficiency, however, the imbalanced intake of leucine and other large neutral amino acids relative to methionine is predicted to result in competitive inhibition for the uptake of dietary methionine at the BBB, and further accentuate methionine deficiency in the brain/central nervous system of patients. Using the plasma amino acid concentrations observed in our cohort of cblC-deficient patients as the data set for the predictive equations used to study maple syrup urine disease, we indeed document a lower calculated influx for methionine into the central nervous system among patients on medical foods compared with those not consuming special formulas (Figure 1e). Moreover, the leucine-to-methionine ratio correlated negatively with the head circumference Z-scores (Figure 1f). While the cerebral amino acid influx model takes into account only two of the multiple transport systems at different physiological barriers, such as the gastrointestinal system and BBB, the principle has been used with success in other disorders.\textsuperscript{21,28,29} Manipulations of brain amino acid influx have gained experimental support in animal studies of glutaric aciduria type 1, where lysine uptake in the brain through the y+ transport system (SLC7A1–3) was modified with homoarginine or arginine supplementation, leading to lower lysine concentrations in brain homogenates, as well as improved disease-related outcomes.\textsuperscript{39,40} Moreover, modulation of brain methionine and S-adenosylmethionine concentrations with dietary methionine modifications has been demonstrated in animal models,\textsuperscript{36} whereas increased methionine synthesis mediated via betaine supplementation was shown to increase methionine and S-adenosylmethionine concentrations in the cerebrospinal fluid, reduce cellular Hcy-thiolactone formation, and improve brain growth in a cohort of patients with 5,10'-methylene tetrahydrofolate reductase deficiency.\textsuperscript{23} Although LAT1 and y+ are discussed here from a proof-of-concept perspective, there are multiple additional transport systems that control amino acid flux between the gastrointestinal system, kidney, systemic circulation, and brain compartment. For example, B0AT1 (SLC 6A19) and ATB\textsuperscript{8a} (SLC6A14) are transporters of large aliphatic amino acids, including leucine, valine, isoleucine, and methionine, in the kidney, gastrointestinal tract, and the BBB, respectively.\textsuperscript{41–43} The complex interactions among these various systems and how metabolic foods deficient in selected amino acids may interfere with amino acid flux between different organs remains largely unexplored.

A delay in symptomatic diagnosis, a history of seizures, and a high relative leucine-to-valine dietary intake were the main predictors of a lower head circumference Z-score in our multiple regression modeling. Although assigning a cause-and-effect relationship between the different variables is not possible with the cross-sectional approach used here, the strong correlation observed between dietary parameters and plasma methionine concentrations, together with the considerations regarding the leucine competition for transport of methionine across the BBB, suggests that these medical foods could contribute to a poor neurocognitive outcome and provide a strong theoretical argument against the use of medical foods with high leucine and absent methionine in patients with cblC deficiency. If protein restriction is deemed necessary, for example, at presentation or
during a rare metabolic crisis with hyperammonemia, medical foods designed for maple syrup urine disease—e.g., Ketonex (Abbott Nutrition) and MSUD Analog (Nutricia)—that contain no branched-chain amino acids (no leucine, valine, or isoleucine) but have a regular amount of methionine are a safer choice for managing patients with cblC deficiency. Further studies need to be designed to estimate the effects of methionine or other methyl-donor supplementation on disease outcome.

In addition to the protein restriction considerations outlined in detail here, other aspects of the dietary needs in this population remain largely understudied, such as essential fatty-acid intake. Alpha-linolenic acid and DHA intake were low to negligible in 9/16 (56%) of the patients for whom sufficient data were available. Erythrocyte phospholipid and plasma content of DHA and amino acids may be lower in inborn errors of metabolism secondary to reduced intake on restricted diets and/or interference with conversion of alpha-linolenic acid to DHA and amino acids secondary to toxic metabolites. Six of seven subjects with inadequate alpha-linolenic acid and DHA intake followed a restricted diet related to prescribed protein restriction or had poor intake for other reasons. It is well recognized that omega-3 fatty acids are important for optimal cognitive function. Furthermore, omega-3 fatty-acid deficiency can also affect platelet aggregation, an important consideration in patients with hyperhomocysteinemia who are prone to thromboembolic complications. Monitoring of dietary intake and plasma concentrations should be incorporated in the management of cblC-deficient patients.

Our study emphasizes the need to develop disease-specific guidelines under the American College for Medical Genetics and Genomics ACT sheet and algorithm for the follow-up of increased propionylcarnitine (C3) detected by newborn screening to more clearly distinguish isolated MMA from early cobalamin biosynthetic defects, because the dietary management should be distinct. In addition, the studies presented here highlight the paucity of experimental and clinical trial support underlying dietary management practices commonly used to treat patients with varied forms of MMA and underscore the community need for a more rigorous scientific and clinical study of medical foods in the treatment of patients with inborn errors of metabolism.

SUPPLEMENTARY MATERIAL

Supplementary material is linked to the online version of the paper at http://www.nature.com/gim

ACKNOWLEDGMENTS

The authors thank all the patients and their families for their participation in our natural history protocol and donation of blood/tissues for our studies; referring physicians, nurses and dietitians for their help with patient's evaluations; Isa Bernardini and Roxanne Fischer for processing patient samples; the nurses, research dietitians of the National Institutes of Health Clinical Research Center, and clinical fellows of the National Human Genome Research Institute genetics fellowship program for their help with patient care and dedication to clinical research. I.M, J.L.S., and C.P.V were supported by the Intramural Research Program of the National Human Genome Research Institute, Bethesda, MD. J.M. was supported by the National Institutes of Health Clinical Center. The authors dedicate this manuscript to the memory of Harvey S. Mudd, who inspired us to study the role of methionine, methylation, and transsulfuration cycles in human physiology and disease. His stimulating discussions contributed significantly to our understanding of dietary effects in the management of cblC-deficient patients.

DISCLOSURE

The authors declare no conflict of interest.

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