Review Article

Bioactive Natural Products against Systemic Arterial Hypertension: A Past 20-Year Systematic and Prospective Review

Maisa Gomes da Silva,1 Sara Léa Fortes Barbosa,1,2 Diego Santos Silva,1 Isadora Basílio Meneses Bezerra,1 Érika Alves Bezerra,1,2 Angélica Gomes Coelho,1 Ilmar Cecília Pinheiro da Silva Morais,1 Luis Mário Rezende-Júnior,1 Iolanda Souza do Carmo,2 José de Sousa Lima-Neto,3 Simón Gabriel Comerma-Steffensen,4,5 Antônia Maria das Graças Lopes Cító,2 and Daniel Dias Rufino Arcanjo1

1Department of Biophysics and Physiology, Federal University of Piauí, Teresina, Piauí, Brazil
2Department of Chemistry, Federal University of Piauí, Teresina, Piauí, Brazil
3Faculty of Pharmacy, Federal University of Piauí, Teresina, Piauí, Brazil
4Pulmonary and Cardiovascular Pharmacology, Department of Biomedicine, Aarhus University, Aarhus, Denmark
5Department of Biomedical Sciences/Animal Physiology, Veterinary Faculty, Central University of Venezuela, Maracay, Aragua, Venezuela

Correspondence should be addressed to Daniel Dias Rufino Arcanjo; daniel.arcanjo@ufpi.edu.br

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Background. Systemic arterial hypertension is one of the most common cardiovascular risks, corresponding to 45% of deaths involving CVDs. The use of natural products, such as medicinal plants, belongs to a millennial part of human therapeutics history and has been employed as an alternative anti-hypertensive treatment. Objectives. The present review aims to prospect some natural products already experimentally assayed against arterial hypertension through scientific virtual libraries and patent documents over the past 20 years. Search strategy. This is a systematic review of the adoption of the PRISMA protocol and a survey of the scientific literature that synthesizes the results from published articles between 2001 and 2020 concerning the use of medicinal plants in the management of hypertension, including which parts of the plant or organism are used, as well as the mechanisms of action underlying the anti-hypertensive effect. Furthermore, a technological prospection was also carried out in patent offices from different countries in order to check technologies based on natural products claimed for the treatment or prevention of hypertension. Inclusion criteria. Scientific articles where a natural product had been experimentally assayed for anti-hypertensive activity (part of plants, plant extracts, and products derived from other organisms) were included. Data extraction and analysis. The selected abstracts of the articles and patent documents were submitted to a rigorous reading process. Those articles and patents that were not related to anti-hypertensive effects and claimed potential applications were excluded from the search. Results. Eighty specimens of biological species that showed anti-hypertensive activity were recovered, with 01 representative from the kingdom Fungi and 02 from the kingdom Protista, with emphasis on the families Asteraceae and Lamiaceae, with 6 representatives each. Leaves and aerial parts were the most used parts of the plants for the extraction of anti-hypertensive products, with maceration being the most used extraction method. Regarding phytochemical analyses, the most described classes of biomolecules in the reviewed works were alkaloids, terpenes, coumarins, flavonoids, and peptides, with the reduction of oxidative stress and the release of NO among the mechanisms of action most involved in this process. Regarding the number of patent filings, China was the country that stood out as the main one, with 813 registrations. Conclusion. The anti-hypertensive activity of natural products is still little explored in Western countries. Besides, China and India have shown more results in this area than other countries, confirming the strong influence of traditional medicine in these countries.
1. Introduction

Arterial hypertension (AH) is a complex, multifactorial, and polygenic disease dependent on diet, demographic, and genetic factors, resulting from the imbalance of several systems, considered a public health problem and a risk factor for cardiovascular diseases (CVD), promoting heart failure, kidney failure, and stroke. Defined by blood pressure levels, AH is characterized by persistent and sustained elevation of blood pressure (BP), that is, systolic BP (SBP) greater than or equal to 140 mm-Hg and/or diastolic BP (DBP) greater than or equal to 90 mm-Hg [1].

CVDs are the leading cause of death, hospitalizations, and outpatient care worldwide, including in developing countries such as Brazil. In 2017, complete and revised data from DATASUS showed the occurrence of 1,312,663 deaths in total, with a percentage of 27.3% for CVD, with AH associated with 45% of these cardiac deaths [2, 3]. Recently, a study led by Imperial College London in collaboration with the World Health Organization (WHO) showed that the number of adults with hypertension between 30 and 79 years of age has increased from 650 million to 1.28 billion over the past 30 years, mainly in developing countries. The study revealed that the prevalence of AH decreased in high-income countries (Canada, Peru, and Switzerland), while in low-income countries (Dominican Republic, Jamaica, Paraguay, Hungary, and Poland), there was a significant increase. The factors involved in this increase would be the aging of the population and greater exposure to other risk factors [4]. Projections show that by 2030, 41.4% of US adults will have hypertension, an increase of 8.4% from 2012 estimates [5].

Determined by the product of cardiac output (CO) and peripheral vascular resistance (PVR), blood pressure is regulated by neural, renal, humoral, endothelial, and local control mechanisms of cardiovascular and renal functions. In this way, SAH can develop from abnormalities in any homeostatic control mechanisms of PVR and/or CD [6]. Thus, the pathophysiology of AH involves changes in its different mechanisms (baroreflex dysfunction, increased sympathetic activation, alterations in the renin-angiotensin-aldosterone system, increased NAD(P)H oxidase activity, oxidative stress, and endothelial dysfunction) [7], whose common trait is endothelial dysfunction, characterized by the low availability of nitric oxide (NO) and the consequent local imbalance between factors of relaxation and constriction of arterioles [8].

Classically, the treatment of AH consists of the use of anti-hypertensive therapy, which, associated or not with other methods, such as lifestyle modifications, can effectively reduce morbidity and mortality related to this condition [7]. This information becomes of great relevance for both the academic community and the scientific community, as a way of designing new intervention strategies, so that the individual with this disease can achieve greater success in its control and treatment. Thus, pharmacological and non-pharmacological measures protect against endothelial dysfunction by helping to preserve cardiovascular function through the reduction of oxidative stress and other mechanisms [9]. Figure 1 summarizes some physiological mechanisms towards therapeutic approaches that can be addressed.

Although more than 50% of existing medicines are synthesized from substances extracted from plants and herbs, the search for active ingredients present in plants, thus creating the first medicines with the characteristics that we know today, began only in the twentieth century—nineteenth century, according to historical records [10]. Among the drugs used in clinical practice whose origin comes from natural products, we can mention ephedrine (from *Ephedra sinica*), aspirin (from *Salix alba*), lovastatin (from *Monascus purpureus*), and reserpine (from *Rauwolfia serpentina*), for example [11].

Based on such knowledge, the objective of this study was to develop a systematic evaluation of the research carried out over the past 20 years on natural products with anti-hypertensive activity, extracting relevant information from scientific articles and patents.

2. Material and Methods

Through the adoption of the PRISMA protocol, a systematic review was carried out using scientific articles that addressed the anti-hypertensive activity of natural products. Following the guideline proposed by Sampaio and Mancini [12], the following question was formulated: “How many natural products have proven anti-hypertensive activity and been described in scientific articles published over the past 20 years?” Thus, a search was carried out in May 2020 in the main virtual libraries: Capes Periodical Portal, SciELO (Scientific Electronic Library Online), and portal BVS (Virtual Health Library), using the following terms: anti-hypertensive OR hypertension AND “natural product” OR products have proven anti-hypertensive activity and been described in scientific articles published over the past 20 years?”

Based on such knowledge, the objective of this study was to develop a systematic evaluation of the research carried out over the past 20 years on natural products with anti-hypertensive activity, extracting relevant information from scientific articles and patents. A search was carried out from databases associated with the INPI (National Institute
How many natural products have proven antihypertensive activity and described in scientific articles published over the last 20 years?

Data base: "Capes Periódicos" Portal; SciELO; VHL

Descriptors: Antihypertensive OR hypertension AND "natural product" OR "medicinal plant"
Languages: English, Portuguese and Spanish
Period: 2001 until 2020

Inclusion criteria: scientific articles in which a natural product has been experimentally tested for antihypertensive activity
Exclusion criteria: articles that were not intended to assess the antihypertensive function of natural products; which had no antihypertensive response; which evaluated synthetic or semi-synthetic products.

Reading the abstract of the articles independently by 02 researchers

Extraction of information about the species used (family, chemical constituents, and mechanisms of action)

Figure 1: Main mechanisms and signaling pathways involved in blood pressure control.

Figure 2: Process of the research and treatment protocol of scientific production articles about natural products that possess anti-hypertensive activity.
of Industrial Property), the USPTO (United States Patent and Trademark Office), the EPO (European Patent Office), the WIPO (World Intellectual Property Organization), and the LATIPAT, using the keywords: medicinal plant(s) and herb(s) associated with the words hypertension, anti-hypertensive, and hypotensive activity. In the case of duplication, the patent of the database other than WIPO was recorded. After an exploratory reading of the titles and abstracts, those that were in accordance with the objective of the study were selected and fully analyzed.

3. Results and Discussion

The search for the terms associated with hypertension, anti-hypertensive drugs, natural products, and medicinal plants in three selected virtual libraries yielded a total of 219,519 scientific articles published between the years 2001 and 2020. Of this total, 3,813 came from the search on the Capes Periodicals Portal, 3,203 from SciELO and 212, 503 from the VHL portal. Ninety-five articles were selected after applying the inclusion and exclusion criteria. Within these works, 80 specimens of biological species that showed anti-hypertensive activity were identified, with 01 representative of the Fungi kingdom and 02 of the Protista kingdom (Table 1). Species from the most different families were found, especially the Asteraceae and Lamiaceae families, with 6 representatives each (Figure 3).

The Asteraceae family is one of the largest of the angiosperms group, with about 180 genera, being considered one of the most important sources of plant species of therapeutic interest [13]. In Brazil, it covers the phytogeographic domains of the Caatinga, Amazon, Pantanal, Pampa, Atlantic Forest, and Cerrado [14]. It bears great importance for the composition of the vegetation of different places, being one of the richest families [15]. Its plants can produce a wide variety of secondary compounds; the most common compounds are phenolic acids, polyacetylenes, flavonoids, coumarins, benzofuran, and terpenoids such as monoterpenes, diterpenes, triterpenes, sesquiterpenes, and especially sesquiterpene lactones [16].

In turn, the Lamiaceae family, with 295 genera and about 7,775 species, another great representative of angiosperms [17], is a group with a cosmopolitan distribution, occurring mainly in open savannas and mountainous regions with a tropical to subtropical climate [18]. Being represented in Brazil by 34 genera and 498 species, the species of this family produce a wide variety of secondary metabolites [19] and accumulate substances with great structural diversity, such as steroids, flavonoids, iridoids, and terpenoids, including triterpenes. The latter are known to have anti-tumor, anti-HIV, anti-inflammatory, anti-oxidant, anti-bacterial, and anti-fungal activities, among others [20]. Thus, other members of these families deserve to be studied.

Popular observations on the use and effectiveness of medicinal plants significantly contribute to the dissemination of the therapeutic virtues of plants, frequently prescribed, for the medicinal effects they produce, despite not having their known chemical constituents. Indirectly, this type of medicinal culture draws the interest of researchers in studies involving multidisciplinary areas, such as botany, pharmacology, and phytochemistry, which together enrich the knowledge about the world flora [21].

In general, the choice of a particular medicinal plant is made through the ethnopharmacological approach. Once the plant species to be studied is defined, the place of the collection is also defined, as well as the part of the plant that will be investigated (root, stem bark, stem, branches, leaves, flowers, and fruits) for carrying out the phytochemical study. Thus, in a project that links phytochemistry with pharmacology, the part of the plant that is used in folk medicine should be chosen for collection [21]. During the survey, it was observed that the leaves and aerial parts, roots, and seeds were the most used parts of the plants for the extraction of anti-hypertensive products, as shown in Figure 4(a).

The search for bioactive compounds of natural origin has increased considerably over the past two decades, mainly due to their preventive potential and in the treatment of cardiovascular, chronic, and neurodegenerative diseases [22]. In this sense, finding efficient extractive methods as well as the characterization of bioactive compounds from natural sources is a great challenge for researchers. Extractive methods for obtaining plant extracts include maceration, infusion, percolation, decocion, continuous hot extraction (Soxhlet), countercurrent extraction, microwave-assisted extraction, ultrasound, supercritical fluid, and turbolysis. In addition to the extractive methods, there are several factors that influence the extraction, such as the part of the plant material used, its origin, the degree of processing, the particle size, the solvent used, the extraction time, temperature, polarity, solvent concentration [23], and how communities do it [24]. Maceration was the most used extraction method in the research reviewed, followed by infusion/decocion and parallel Soxhlet. Other techniques such as percolation and critical superfluid extraction occurred less frequently (Figure 4(b)).

In order to quantify and qualify the chemical constituents of plant extracts, whose beneficial effects of some substances of certain species act as a key factor for the development of research bases for future applications of these bioactives [25], a preliminary phytochemical investigation is carried out to recognize the chemical constituents and/or assess their presence in the species being studied [26]. Specifically, phytochemical screening makes it possible to carry out preliminary tests to identify the presence of chemical compounds in certain plant species and thus link them to possible biological activities.

For example, alkaloids have anti-bacterial, anti-fungal, anti-plasmodic, and anti-tumor properties [27, 28] due to the ability to destabilize biological membranes. They also have the ability to inhibit the synthesis of DNA and RNA by binding to nucleic acids and intercalating into the double helix [29]. Examples of alkaloids currently used are morphine (analgesics), scopalamine (anti-cholinergic), theophylline (diuretics), vincristine (anti-tumours), and codeine (anti-tussives) [30].

Flavonoids are the most numerous compounds in angiosperms and have anti-inflammatory, anti-allergic, anti-ulcerogenic, anti-viral, anti-proliferative, anti-oxidant,
| Species                  | Family       | Used part (s) | Chemical constituents/classification                                                                 | Mechanisms of action                                                                                                                                                      | Study model                                                                                     | Reference |
|-------------------------|--------------|---------------|-----------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------|-----------|
| Agelanthus dodoneifolius| Loranthaceae | No data       | Dodonein (lactone)                                                                                   | Blockade of the L-type calcium channels and inhibition of carbonic anhydrase in smooth muscle cells                                                                                                                                 | Ex vivo assays for vasodilation in rat aortic rings; in vitro assay by the culture of vascular smooth muscle cells and determination of messenger RNA of carbonic anhydrase isozyme A in smooth muscle cells | [33]      |
| Allium cepa             | Amaryllidaceae| Rhizome       | Diallyl thiosulphinate, methyl allyl thiosulphinate, allylmethyl thiosulphinate, protocatechuic acid,  | Inhibition of angiotensin-converting enzyme                                                                                                                                                                                          | In vitro angiotensin-converting enzyme inhibitory assay                                                                                                     | [34]      |
| Allium sativum          | Amaryllidaceae| Rhizome       | S-allyl cysteine                                                                                      | Inhibition of angiotensin-converting enzyme                                                                                                                                                                                          | In vivo assays with mice with fructose-induced hypertension Ex vivo tests of isolation of the superior mesenteric artery of rats | [35]      |
| Alpinia zerumbet        | Zingiberaceae | Leaves        | Routine and kaempferol-3-O-β-D-glucuronide (flavonoids)                                               | Stimulates NO/cGMP pathway                                                                                                                                                                                                           | Ex vivo tests of isolation of the superior mesenteric artery of rats                                                                                       | [36]      |
| Annona muricate         | Annonaceae    | Leaves        | Roseoside, isolariciresinol 9-O-β-D-xylloside, massonianoside B, icariside E4, and nicotiflorin         | Anti-oxidant, anti-inflammatory, and anti-vascular remodeling properties and reduced AT1 receptor expression                                                                                                                 | In vitro assay in angiotensin II (Ang II) stimulated H9C2 cells                                                                                              | [37]      |
| Angelica dahurica       | Apiaceae      | Root          | Imperatorin                                                                                          | Reduction of oxidative stress and prevention of hypertension-related renal injury                                                                                                                                                     | In vivo assay in rats with renovascular hypertension and ex vivo assays that evaluate the cellular redox state                                                   | [38]      |
| Angelica decursiva      | Apiaceae      | Root          | Decursin and nodakenin                                                                               | Opening of the potassium channels Reduction of renal fibrosis; reduction of oxidative stress; decreased levels of TNF-α, IL-6, and NF-κB                                                                                       | Assays in rat aortic arteries                                                                                                                             | [39]      |
| Apium graveolens        | Apiaceae      | Seed          | 3-n-butylphthalide                                                                                   | In vivo assays with spontaneously hypertensive rats                                                                                                                                                                      |                                                                                                                                                | [40]      |
| Species                  | Family       | Used part (s)          | Chemical constituents/ classification      | Mechanisms of action                                                                 | Study model                                                                 | Reference |
|-------------------------|--------------|------------------------|--------------------------------------------|--------------------------------------------------------------------------------------|-------------------------------------------------------------------------------|-----------|
| *Arbutus andrachne*     | Ericaceae    | Root, leaves, and fruit| Phenols, flavonoids, tannins, and anthocyanins | Reduction of oxidative stress                                                         | *Ex vivo* tests for vasodilation in rat aorta rings with intact endothelium; *ex vivo* assays that evaluate the cellular redox state | [41]      |
| *Arbutus unedo*         | Ericaceae    | Root                   | Tannins and flavonoid (quercetin and tannic acid) | Stimulation of the endothelial nitric oxide synthase and activation of muscarinic receptors; Activation of muscarinic receptors in the heart, reducing the heart rate and increasing peripheral resistance | *Ex vivo* tests for vasodilation in rat aorta rings with intact endothelium | [42]      |
| *Azadirachta indica*    | Meliaceae    | Leaves                 | No data                                    | Stimulation of the endothelial nitric oxide synthase and activation of muscarinic receptors; Activation of muscarinic receptors in the heart, reducing the heart rate and increasing peripheral resistance | *In vivo* assay in rats with hypertension induced by DOCA-salt injection       | [43]      |
| *Berberis vulgaris*     | Berberidaceae| Fruit                  | No data                                    | Activation of the l-arginine-nitric oxide pathway                                     | *In vivo* assay in rats with hypertension induced by DOCA-salt injection, *in vitro* studies in aortic rings, and *in vitro* studies in the isolated perfused mesenteric beds | [44]      |
| *Bidens pilosa*         | Asteraceae   | Leaves                 | Alkaloids, saponins, flavonoids, polyacetylenes and triterpenes, phenylheptatriyne, linoleic acid, and linolic acid | Stimulation of the endothelial nitric oxide synthase and activation of muscarinic receptors; Activation of muscarinic receptors in the heart, reducing the heart rate and increasing peripheral resistance | *Ex vivo* assays for vasodilation in rat aortic rings                          | [45]      |
| *Boerhavia diffusa*     | Nyctaginaceae| Root                   | Culubin (diterpenoid)                      | Blocking of calcium channels                                                         | *In vivo* assay in rats with hypertension caused by obesity induced by a lipid-rich diet | [46]      |
| *Cassia tora*           | Fabaceae     | Seed                   | Chrysofanol, Aurantium Obtusine, alaternine, and chrysothymbain (anthraquinones) | Stimulation of the endothelial nitric oxide synthase and activation of muscarinic receptors; Activation of muscarinic receptors in the heart, reducing the heart rate and increasing peripheral resistance | *In vitro* assays                                                             | [47]      |
| *Cecropia pachystachya* | Urticaceae   | Leaves                 | Ambaina and ambainina, long-chain carboxylic acids, and β-sitosterol                  | Stimulation of the endothelial nitric oxide synthase and activation of muscarinic receptors; Activation of muscarinic receptors in the heart, reducing the heart rate and increasing peripheral resistance | *In vivo* assay in normotensive Wistar rats through cannulation of internal carotid artery | [48]      |
| *Cleistanthus collinus* | Phyllanthaceae| Leaves                 | Cleistantin A and B (glycosides)           | Stimulation of the endothelial nitric oxide synthase and activation of muscarinic receptors; Activation of muscarinic receptors in the heart, reducing the heart rate and increasing peripheral resistance | *In silico* molecular interaction                                              | [49]      |
| Species                      | Family       | Used part (s) | Chemical constituents/ classification | Mechanisms of action                                                                                      | Study model                                                                 | Reference |
|------------------------------|--------------|---------------|----------------------------------------|----------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------|-----------|
| Crataegus tanacetifolia      | Rosaceae     | Leaves        | Hyperoside                             | Increase in kidney NOS activity, diuretic activity, and efflux of water and sodium, preventing hyperlipidemia and decrease in body weight | *In vivo* assay in normal male Wistar albino and L-NAME-induced hypertensive rats | [50]      |
| Codonopsis lanceolata        | Campanulaceae| Rhizome       | Lancemaside A                         | Increase in NO levels by eNOS (inducible NO synthase)                                                  | *In vitro* assay in human umbilical vein endothelial cells                  | [51]      |
| Coffea                       | Rubiaceae    | Fruit         | Chlorogenic acids                     | Stimulation of the endothelial nitric oxide synthase                                                   | A double-blind, randomized, placebo-controlled study in humans             | [52]      |
| Coix larchrymajoji           | Poaceae      | Seed          | Glutelin hydrolyzate                  | Inhibition of angiotensin-converting enzyme                                                              | *In vivo* assays in hypertensive rats                                       | [53]      |
| Cordyceps sinensis*          | Clavicipitaceae | Entire organism | Mannose, glucose, and galactose (polysaccharide fraction) | Increase in NO levels and decrease of the levels of endothelin-1, epinephrine, noradrenaline, angiotensin II, and TGF-β1 | *In vivo* assays with spontaneously hypertensive rats                      | [54]      |
| Coriandrum sativum           | Apiaceae     | Fruit         | Camphor, camphene, carvone, cineole, cimene, coriandrine, limonene, linoleic acid, myrcene, myristic acid, oleic acid, palmitic acid, α-phenyltriene, β-phenylandrene, and α-terpinene, among others | Blockade of calcium channels, interaction with muscarinic receptors and diuretic effect                   | *In vivo* assays in normotensive mice and *ex vivo* assays in isolated tissue preparations | [55]      |
| Crocus sativus               | Iridaceae    | Flower        | Crocin, crocetina, and Safranal       | Release of nitric oxide, reduction of oxidative stress, and modulation of the renin-angiotensin system  | *In vivo* assay in rats through cannulation of arteries and femoral veins of rats with hypertension induced by Ang-II | [56]      |
| Croton schiedeanus           | Euphorbiaceae| Aerial parts (stem and leaves) | Flavonoids, diterpenoids, and phenylbutanoids | Stimulation of NO/cGMP pathway                                                                        | *In vivo* assays in mice with hypertension by chronic inhibition of nitric oxide and *ex vivo* assay in isolated tissue preparations | [9]       |
| Cucurbita pepo               | Cucurbitaceae| Seed          | Cucurbitacins (triterpenes); lutein, carotene, and beta carotene(carotenoids); unsaturated linoleic and oleic acids | Increase of NO levels                                                                                   | *In vivo* assays in mice with chronic inhibition of nitric oxide and *in vitro* assays | [57]      |
| Species             | Family           | Used part (s) | Chemical constituents/ classification                                                                 | Mechanisms of action                                                                                                                                                       | Study model                                      | Reference |
|---------------------|------------------|---------------|-------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--------------------------------------------------|-----------|
| *Curcuma spp.*      | Zingiberaceae    | Rhizome       | Curcumin, demethoxycurcumin, and bisdemethoxycurcumin                                                   | Blocking of calcium channels and the partial inhibition of β-adrenergic receptors                                                                                               | Ex vivo vasodilation assay on intact endothelium pigs basilar arteries pre-contracted                   | [58]      |
| *Cyclocarya paliurus* | Juglandaceae     | Leaves and seeds | Polysaccharides                                                                                         | Reduction of oxidative stress                                                                                                                                               | In vitro and *in vivo* assays using hypertensive rats                                               | [59]      |
| *Dendranthema indicum* | Asteraceae       | Flower        | Linarin                                                                                               | Modulation of the Renin-angiotensin system                                                                                                                                 | *In vivo* assays with spontaneously hypertensive rats                                                 | [60]      |
| *Dicksonia sellowiana* | Dicksoniaceae    | Leaves        | Polyphenols                                                                                           | Reduction of oxidative stress, activation of the pathway PI3K/Akt/eNOS                                                                                                       | *Ex vivo* tests on isolated tissues; *in vitro* assay on pig endothelial cell culture; *in vivo* tests with spontaneously hypertensive rats | [61]      |
| *Dioscorea opposita* | Dioscoreaceae    | Rhizome       | Saponins, starch, mucopolysaccharides, protein, amino acids, mucilage, and polyphenols                   | Inhibition of angiotensin II converting enzyme, inhibition of endothelin-1 and reduction of oxidative stress                                                                 | *Ex vivo* assay in rats with renovascular hypertension and *ex vivo* assays that evaluate the cellular redox state | [62]      |
| *Eclipta alba*      | Asteraceae       | Aerial parts  | Culubin (diterpenoid)                                                                                   | Diuresis due to increase in sodium excretion                                                                                                                                 | *Ex vivo* assay in rats with hypertension caused by obesity induced by a lipid-rich diet                | [46]      |
| *Eucommia ulmoides* | Eucommiaceae     | Stem bark     | Wogonin (flavonoid)                                                                                     | Inhibition of the intracellular release of Ca^{2+} and the extracellular influx of Ca^{2+}                                                                                   | *Ex vivo* testing on isolated tissue preparations                                                      | [63]      |
| *Ficus deltoidea*   | Moraceae         | Leaves        | β-amyrin, lupeol, β-amyrin cinnamate and bergapten, tanacetene, β-elemene, stigmasterol, β-sitosterol, lupenone, and αβ-amyrenone, as well as alkaloids, saponin, phenols, flavonoids, and tannins | Modulation of the renin-angiotensin-aldosterone system, antioxidant and endothelial system                                                                                | *In vivo* assays with spontaneously hypertensive rats                                                  | [64]      |
| *Gardenia jasminoides* | Rubiaceae        | Fruit         | Crocetin (carotenoid)                                                                                   | Increase in NO levels by eNOS and iNOS (inducible NO synthase)                                                                                                               | *Ex vivo* vasodilation assay on intact endothelium mouse aorta rings, and *in vitro* assays           | [65]      |
| *Glycine Max*       | Fabaceae         | Seed          | Equol (flavonoid)                                                                                       | Diuresis by an increase in sodium excretion and increases transcription of the enzyme eNOS                                                                               | A double-blind, randomized, placebo-controlled study in humans                                       | [66]      |
| Species                     | Family         | Used part(s) | Chemical constituents/ classification | Mechanisms of action                                                                 | Study model                        | Reference |
|-----------------------------|----------------|--------------|---------------------------------------|---------------------------------------------------------------------------------------|-------------------------------------|-----------|
| Gomphrena celosioides      | Amaranthaceae  | Aerial parts | Phenolic acids and flavonoids         | Increased levels of bradykinin, prostaglandins, and NO. Increase in NO by activation of PI3K/Akt/eNOS pathway and activation of potassium channels | In vivo assays in hypertensive animals; Ex vivo rat assays in isolated tissue preparations | [67]      |
| Hibiscus sabdariffa        | Malvaceae      | Flower       | Anthocyanins                          | Inhibition of angiotensin-converting enzyme                                            | In vivo assays in hypertensive adult rats | [68]      |
| Inula viscosa              | Asteraceae     | Leaves       | Phenolic compounds and flavonoids     | Anti-oxidative and lipid-lowering effect                                              | In vivo assays with hypertensive rats induced by oral administration of ethanol | [69]      |
| Leersia hexandra           | Poaceae        | Aerial parts | Not identified                        | Activation of calcium-activated potassium channels and increase in cAMP and and cytosolic cGMP | In vivo assays in mice with hypertension by chronic inhibition of nitric oxide | [70]      |
| Lippia origanoides         | Verbenaceae    | Aerial parts | Naringenin and pinocembrina (flavonoids), quercetin (flavonol), and luteolin (flavones) | Activation of renin-angiotensin-aldosterone system and reduction of oxidative stress | In vivo assays with spontaneously hypertensive rats and normotensive rats; in vitro assays | [71]      |
| Lithocarpus polystachys     | Fagaceae       | Leaves       | florizine, flavonol, quercetin, dihydrochalcone-20-b-D-glucopyranoside, luteolin, and quercetin (Flavonoids) | Modulation of the renin-angiotensin-aldosterone system and reduction of oxidative stress | In vivo assays in spontaneously hypertensive rats; ex vivo testing on isolated tissue preparations; molecular interaction in silico | [72]      |
| Lonchocarpus xuul           | Fabaceae       | Root         | Dihydrospinocchalcone-A and isocardoin | Activation of potassium channels and activation of NO/sCG/PKG pathway | In vivo assay in spontaneously hypertensive rats; ex vivo assay on isolated tissue preparations; molecular interaction in silico | [73]      |
| Lycopersicon esculentum    | Solanaceae     | Fruit        | α-tocopherol and the carotenoids: lycopene, β-carotene, phytoene, and phytofluene | Attenuation of inflammatory signaling by the inhibition of the NF-κB transcription factor in endothelial cells | A double-blind, randomized, placebo-controlled study in humans; in vitro assay on human endothelial cell culture | [74]      |
| Mentha x villosa           | Lamiaceae      | Leaves       | No data                               | Active vascular relaxation                                                             | In vivo assay in rats with hypertension induced by DOCA-salt injection | [75]      |
| Mesona procumbens          | Lamiaceae      | Leaves       | Caffeic acid (polyphenol)             | Reduction of oxidative stress                                                         | In vivo assay in spontaneously hypertensive rats and ex vivo assay evaluating the cellular redox state | [63]      |
| Species                        | Family     | Used part(s) | Chemical constituents/classification                                                                 | Mechanisms of action                                                                 | Study model                                           | Reference |
|-------------------------------|------------|--------------|------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------|-------------------------------------------------------|-----------|
| Mimosa caesalpiniifolia       | Fabaceae   | Inflorescences| Gallic acid, rutin, quercetin, and vicenine (flavonoids)                                              | Activation of the muscarinic and ganglionic pathways and blockade of the transmembrane calcium influx | *In vivo* assay in normotensive mice; *ex vivo* testing on isolated tissue preparations | [76]      |
| Mitragyna ciliata             | Rubiaceae  | Stem, Bark   | Alkaloids (mitragynine, mitraphylline, and rhynophylline) and/or flavonoid                            | Blocking of calcium channels                                                        | *Ex vivo* rat assays in guinea pig and rat isolated aortic rings                   | [77]      |
| Mixture containing Pine densiflora | Pinaceae | Leaves       | Roseoside, isolariciresinol 9-O-β-D-xylloside, massonianoside B, icariside E4, and nicotiflorin       | Anti-oxidant, anti-inflammatory, and anti-vascular remodeling properties and reduced AT1 receptor expression | *In vitro* assay in Angiotensin II (Ang II)-stimulated H9C2 cells               | [37]      |
| Momordica charantia           | Cucurbitaceae | Leaves      | Roseoside, isolariciresinol 9-O-β-D-xylloside, massonianoside B, icariside E4, and nicotiflorin       | Anti-oxidant, anti-inflammatory, and anti-vascular remodeling properties and reduced AT1 receptor expression | *In vitro* assay in Angiotensin II (Ang II) stimulated H9C2 cells               | [37]      |
| Morinda citrifolia            | Rubiaceae  | Root         | Alkaloids, phenolic compounds, sterols, flavonoids, tannins, coumarins, and anthraquinoes         | Blocking of calcium channels and release of intracellular calcium                   | *Ex vivo* rat assays in tissue preparations isolated from rats                   | [78]      |
| Moringa oleifera              | Moringaceae | Leaves       | Nitrile, glucosinolates and thio carbamate glycosides, flavonoids, phenolic acids, tannins, quercetin-3-O-glucoside, kaempferol-3-O-glucoside, Niazicin-A, Niazimin-A, and Niaziminin-B | Alleviation of vascular dysfunction and oxidative stress, blunted adrenergic-mediated vasoconstriction, promoted endothelium-dependent vasorelaxation; inhibition of angiotensin-converting enzyme   | *In vivo* assay in L-NAME-treated rats; *in vitro* angiotensin-converting enzyme inhibitory assay; *in silico* molecular interaction | [79–81]  |
| Moringa stenopetala (Baker f.) | Moringaceae | Leaves       | Alkaloids, flavonoids, and saponins                                                                | Inhibition of carbonyl anhydrase Reduction of oxidative stress and increase in NO by activation of pathway PI3K/Akt/eNOS | *In vivo* assay on mice                                                           | [82]      |
| Musa sapientum                | Musaceae   | Fruit peel   | (±)-7, 8-Dihydroxy-3-methyl-isochromanone-4 (polyphenol)                                            | Inhibition of carbonyl anhydrase Reduction of oxidative stress and increase in NO by activation of pathway PI3K/Akt/eNOS | *In vivo* assay in spontaneously hypertensive rats                                | [83]      |
| Species                  | Family            | Used part(s) | Chemical constituents/classification                                                                 | Mechanisms of action                                      | Study model                                      | Reference   |
|--------------------------|-------------------|--------------|------------------------------------------------------------------------------------------------------|-----------------------------------------------------------|-------------------------------------------------|-------------|
| *Nardostachys jatamansi* | Caprifoliaceae    | Rhizome      | Jatamansone, calarene, spirojatamol, aristolone, valencene and patchouli alcohol, α-pinene, and β-maaliene (E)-1-oxo-3, 4-dihydro-1-H-isochromen-7-yl-3-(3, 4-dihydroxyphenyl) acrylate | Inhibition of angiotensin-converting enzyme                | *In vitro* angiotensin-converting enzyme inhibitory assay | [84]        |
| *Onopordum acanthium*    | Asteraceae        | Seed         | 2, 3, 5-trimethoxy-9-(12, 14, 15-trimethoxybenzyl)-1H-indene and pellicidin A                         | Inhibition of angiotensin-converting enzyme                | Molecular interaction *in silico in vitro* assays    | [85]        |
| *Orthosiphon stamineus*  | Lamiaceae         | Leaves       | No data                                                                                               | Modulation of α1-adrenergic receptors and AT1 and increase in levels of NO | A parallel-group, randomized, placebo-controlled study in humans; rings of aorta of spontaneously hypertensive rats | [86]        |
| *Panax notoginseng*      | Araliaceae.       | Root         | Ginsenoside Rg1 and Rb1                                                                                   | NO/sGC/cGMP pathway and β2-adrenergic receptors            | *Ex vivo* rat assays in isolated tissue preparations (aortic ring model) | [87]        |
| *Peperomia pellucida*    | Piperaceae        | Leaves       | 2, 3, 5-trimethoxy-9-(12, 14, 15-trimethoxybenzyl)-1H-indene and pellicidin A                         | Inhibition of angiotensin-converting enzyme                | *In vitro* angiotensin-converting enzyme inhibitory assay | [88]        |
| *Phaseolus vulgaris*     | Fabaceae          | Seed         | Catechins, flavonoids, and γ-aminobutyric acid (GABA)                                                   | Inhibition of angiotensin-converting enzyme and modulation of pressure via GABA. | *In vitro* assays                                | [89]        |
| *Phoenix dactylifera*    | Areaceae          | Fruits       | Squalene, lauric acid, palmitic acid, caprate, stearate, vitamin E, β-sitosterol, phytol, linolenic acid, isosorbide, coumarins, and taurine | Inhibition of angiotensin-converting enzyme                 | *In vitro* enzyme inhibition assays               | [90]        |
| *Piper nigrum*           | Piperaceae        | Seed         | Piperine (alkaloid)                                                                                     | Reduces oxidative stress                                   | *In vivo* assay in rats with hypertension caused by obesity induced by a lipid-rich diet | [91]        |
| *Prunus persica*         | Rosaceae          | Aerial parts | Amygdalin, cyanogenic glycosides, prunasin, caffic acid, chlorogenic acid, kaempferol, p-coumaric acid, prussic acid, quercetin, quercitrin, quinic acid, tannin, and ursolic acid Reserpine, ajmalicine, serpentine, ajmalidine, ajmaline, rescinnami-dine, rescinnamine, reserpine, serpentine, indobidine, yohimbine, and deserpidine | NO-sGC-cGMP, vascular prostacyclin, and muscarinic receptor transduction pathway | *Ex vivo* rat assay in isolated tissue preparations (aortic ring model) | [92]        |
| *Rauvolfia serpentina*   | Apocynaceae       | Roots        | Protecting the liver and renal architectures                                                             | *In vivo* assay in rats with hypertension induced by high salt diet | *In vivo* assays                                | [93]        |
| Species            | Family       | Used part(s) | Chemical constituents/classification                          | Mechanisms of action                                                                 | Study model                        | Reference |
|--------------------|--------------|--------------|---------------------------------------------------------------|----------------------------------------------------------------------------------------|-------------------------------------|-----------|
| *Rubus rosifolius* | Rosaceae     | Leaves       | Escaphic acid, flavonoids, and triterpenes                    | Diuretic effect                                                                        | *In vivo* assay in hypertensive male rats | [94]      |
|                    |              | Root         | Lithospermic acid B                                           | Inhibition of angiotensin-converting enzyme                                            | *Ex vivo* assays for vasodilation in rat aortic rings | [95]      |
| *Salvia miltiorrhiza* | Lamiaceae   | Root         | *Tanshinato of magnesium B*                                    | Increase in NO levels                                                                  | *In vivo* assay in rats with phenylephrine-induced hypertension | [96]      |
| *Salvia scutellarioides* | Lamiaceae | Aerial parts | Alkaloids, triterpenes, lignans, and flavonoids                | Inhibition of angiotensin-aldosterone system, and increase in concentrations of epinephrine and vasopressin | *In vivo* assay in L-NAME-treated rats | [97]      |
| *Sargassum siliquastrum*** | Sargassaceae | Entire organism | Sargachromenol D                                            | Induced depolarization                                                                 | *In vivo* assay in rat basilar arteries | [98]      |
| *Sceletium tortuosum* | Mesembrythemaecae | Leaves | Mesembrine (alkaloid)                                          | Inhibition of aldosterone synthesis                                                    | *In vitro* assay on the culture of human adrenocortical carcinoma cells | [99]      |
| *Solanum donianum* | Solanaceae   | Leaves       | Unreported                                                    | Inhibition of angiotensin-converting enzyme                                              | *In vitro* assay inhibitory assay     | [100]     |
| *Spirulina maxima*** | Cyanophyceae | It has no true tissues | Phycocyanin                                                    | Increases transcription of the enzyme eNOS                                              | Cohort study with humans             | [101]     |
| *Taraxacum officinale* | Asteraceae | Leaves and root | Saponins, alkaloids, phenols, flavonoids, tannins, and glycosides | Increase in NO levels by eNOS (inducible NO synthase)                                  | *In vivo* assay in L-NAME-treated rats and with spontaneously hypertensive rats | [102]     |
| *Taxus chinensis var. mairei* | Taxaceae Gray | Leaves | Palmitic acid, 9-octadecenate of hexadecanil, and octan-3-ol | Reduction of the level of angiotensin II and increase in NO levels                     | *In vivo* assays with mice with hypertension by chronic nitric oxide inhibition and *in vitro* assays | [103]     |
| *Terminalia superba* | Combretaceae | Stem bark    | Saponins, glycosides, flavonoids, and chalcones               | Reduction of oxidative stress                                                         | *In vivo* assays that evaluate the cellular redox state | [104]     |
hepatoprotective, anti-thrombic, and anti-carcinogenic activities [29, 30]. Tannins are phenolic compounds that have the property of complexing with metal ions and macromolecules such as proteins and polysaccharides, so they play the role of anti-oxidant and protector against herbivores and microorganisms. They are used as anti-septics, astringents, anti-diarrheals, wound healing, burns, and inflammation due to their ability to precipitate proteins [29–31]. They also have the ability to stimulate phagocytic cells [30].

Terpenes make up some essential oils and, therefore, act to attract pollinators. They also have insecticidal, anti-microbial, hepatoprotective, analgesic, anti-inflammatory, anti-microbial, and hemolytic action, among others [29, 31]. Triterpenes have anti-inflammatory, analgesic, cardiovascular, and anti-tumor effects [32].

Saponins have the ability to decrease the surface tension of water and, in vitro, cause erythrocyte hemolysis. They alter membrane permeability by lipophilic action and complexation with lipids and cell membrane proteins, which causes cell destruction. Therefore, they have toxic characteristics [31]. They also perform molluscicidal, anti-fungal, anti-microbial, anti-parasitic, anti-viral, cytotoxic, and anti-tumor functions [30].

Phenolic compounds have the ability to neutralize free radicals, inhibiting the risk of cardiovascular disease, diabetes, tumors, and inflammatory processes. Coumarins are used for dermatoses, psoriasis, vitiligo, and other skin diseases; they are also anti-coagulants and laxatives, such as anthraquinones. Catechins are anti-oxidants, thermogenic, anti-inflammatory, and anti-carcinogenic. Steroids have cardiotonic functions, activators of anabolism, precursors of vitamin D, and contraceptives [30].

Regarding phytochemical analyses, the most described classes of biomolecules in the reviewed works were alkaloids, terpenes, coumarins, flavonoids, and peptides (Table 1). Thus, when relating them to anti-hypertensive activity, the focus of the prospective studies, we observed that alkaloids, such as reserpine and alstonine, reduce the availability of norepinephrine and, therefore, act as vasodilators. Flavonoids such as quercetin and rutin are primarily active in the myocardium and reduce cardiac output. Linoleic acid inhibits atherosclerosis-generating deposits of cholesterol and

| Species                  | Family       | Used part(s) | Chemical constituents/classification | Mechanisms of action                                                                 | Study model                              | Reference |
|--------------------------|--------------|--------------|-------------------------------------|-------------------------------------------------------------------------------------|------------------------------------------|-----------|
| Ulmus wallichiana        | Ulmaceae     | Stem bark    | Flavonoids analogous to quercetin   | Modulation of the renin-angiotensin-aldosterone system and stimulation of NO/cGMP pathway | In vivo assay in spontaneously hypertensive rats and assays in rats with salt and mineralocorticoid-induced hypertension, and with rats with chronic inhibition of nitric oxide Ex vivo assays in isolated Langendorff perfused rat heart and vasodilation in rat aortic rings A double-blind, randomized, placebo-controlled study in humans A double-blind, randomized, placebo-controlled study in humans Ex vivo tests on preparations of tissues isolated from rats In vivo assay in spontaneously hypertensive rats and in vitro assays | [105]     |
| Urtica dioica            | Urticaceae   | Aerial parts | No data                             | An important bradycardia, which is independent of cholinergic and 1-adrenergic receptors | Ex vivo assays in isolated Langendorff perfused rat heart and vasodilation in rat aortic rings A double-blind, randomized, placebo-controlled study in humans A double-blind, randomized, placebo-controlled study in humans Ex vivo tests on preparations of tissues isolated from rats In vivo assay in spontaneously hypertensive rats and in vitro assays | [42]      |
| Vaccinium virgatum       | Ericaceae    | Fruit        | Anthocyanins and polyphenols        | Stimulation of NO/cGMP pathway                                                    | A double-blind, randomized, placebo-controlled study in humans | [106]     |
| Vaccinium corymbosum     | Ericaceae    | Fruit        | Anthocyanins and polyphenols        | Stimulation of NO/cGMP pathway                                                    | A double-blind, randomized, placebo-controlled study in humans | [106]     |
| Vitex cienkowskii        | Lamiaceae    | Stem bark    | Tetra-acetyl jugasterone C          | Stimulation of NO/cGMP pathway and blockade of transmembrane calcium influx       | Ex vivo testing on preparations of tissues isolated from rats | [107]     |
| Zea mays                 | Poaceae      | Seed         | Corn peptide                        | Inhibition of angiotensin-converting enzyme                                       | In vivo assay in spontaneously hypertensive rats and in vitro assays | [108]     |

* Fungus species and ** species of seaweed.
triglycerides [109]. Phenolic compounds are anti-oxidants responsible for scavenging free radicals, capable of minimizing the harmful effects of ROS, and considered potential for the prevention of cardiovascular diseases [36].

The control of BP is made through two main mechanisms: neural and humoral. The neural mechanism is made by the autonomic nervous system composed of the sympathetic and parasympathetic systems, which act by increasing or decreasing heart rate as well as acting on peripheral vascular resistance. Humoral control is carried out by several substances that directly interfere with peripheral vascular resistance. Thus, the increase in vasodilating substances such as NO can contribute to an improvement in SAH. On the other hand, the renin-angiotensin system (RAS) plays a fundamental role due to its vasoconstrictor action, mainly through angiotensin II (Ang II) [110].

The classic view of the RAS is given by the production of angiotensinogen by the liver, being released into the circulation, where it is found in high concentrations. In the circulation, angiotensinogen undergoes the action of renin, a glycoproteolytic enzyme of renal origin [111]. After being synthesized and released into circulation, renin promotes the conversion of angiotensinogen into Angiotensin I (Ang I) [112], and this is converted into angiotensin II (Ang II) by the catalytic action of angiotensin-converting enzyme (ACE) [113]. This conversion occurs almost exclusively in the vessels of the lungs, catalyzed by the ACE present in the endothelium of the pulmonary vessels.

The effects of Ang II are mediated by two distinct types of receptors: AT₁ and AT₂, and the greatest interaction of this peptide occurs via the AT₁ receptors, causing vasoconstrictor action, arrhythmogenic effect, cell proliferation, thrombosis, coagulation, inflammation, and hypertrophy of vascular smooth muscle [114, 115]. This Ang II signaling pathway with AT₁ receptors is carried out by the activation of the G protein, with consequent activation of phospholipase C-β and formation of 1,4,5-triphosphate and diacylglycerol, which in turn increases the intracellular concentration of calcium leading to vasoconstriction [116].

In addition to these effects, it is known that q Ang II via AT₁ receptors stimulates aldosterone secretion by the zona glomerulosa of the adrenal cortex [117]. Contrary to this, the interaction of Ang II with the AT₂ receptor has an

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**Figure 3:** Distribution of the main families used in scientific research on natural products with anti-hypertensive activity published in the virtual libraries portal BVS, CAPES, and SciELO from 2001 to 2020.

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**Figure 4:** Percentages of the main chosen plant parts (a) and the main extractive methods (b) used to obtain the different natural products with anti-hypertensive activity retrieved from the virtual libraries BVS, CAPES, and SciELO over the period between 2001 and 2020.
Activation of potassium channels
Blocking of calcium channels
Reduction of oxidative stress
Release of nitric oxide (NO)
Increasing of natriuresis
Inhibition of NF-κB
Inhibition of ACE
Activation of potassium channels
Other mechanisms

Figure 5: Distribution of the anti-hypertensive mechanisms elucidated in the scientific research of natural products with anti-hypertensive activity published in the virtual libraries BVS Portal, CAPES, and SciELO in the period of 2001 to 2020.

antagonistic effect on the action of the Ang II axis–AT₁ receptor, resulting in the formation of NO, and consequent vasodilation [118].

In addition to the physiological effects of controlling cardiovascular function, Ang II is also involved in the pathophysiology of cardiovascular diseases since this peptide induces the formation of reactive oxygen species (ROS) in the endothelium and vascular smooth muscle [119]. This process occurs via AT₁ receptors and consequent activation of the enzyme NAD (P) H oxidase [120], which reduces the oxygen molecule, forming O₂⁻. The latter is dismutated to H₂O₂ by the action of the enzyme superoxide dismutase (SOD) or reacts with NO to form peroxynitrite (ONOO⁻)

Regarding the mechanisms of action used in research cataloged to support the anti-hypertensive effect of the species evaluated, NO release, reduction of oxidative stress, ACE inhibition, Ca²⁺ channel block, RAAS modulation, activation of K⁺ channels, inhibition of nuclear kappa transcription factor (NF-κB), and increase in natriuresis can be observed (Figure 5). Among the possible study models, the research was carried out in in vivo, ex vivo, in vitro, and in silico studies, including studies in humans (Table 1).

As for the number of patents deposited in the databases according to the keywords used, it was observed that the WIPO database markedly recovered 925 documents, followed by the EPO and USPTO, with only 6 and 2 documents, respectively. In the others, INPI and LATIPAT, no patents were found. This may suggest a lack of interest on the part of research centers or industries to innovate in anti-hypertensive products, even though hypertension is one of the main causes of death in the world and 25% of currently available drugs originate from medicinal plants [126].

China was the country with the highest number of patent filings with 813 registrations. The other countries and their respective patent offices showed very low values when compared to China: the Republic of Korea with 75, the United Kingdom with 15, the World Intellectual Property Organization with 12, the European Patent Office with 7, Japan with 6, the United States with 2, and Canada, Russia, and France with 1 each (Figure 6). Although Brazil is a country rich in biodiversity and one that develops a lot of research on medicinal plants, research in the patent databases revealed a lack of interest in the development of technologies with market potential related to anti-hypertensive herbal medicines.

The superiority in the number of patents filed by China is related to the economic and technological position of this country in relation to the world scenario, as China has been increasingly establishing itself as a producer of knowledge and technological development, mainly through the work of pharmaceutical multinationals, electronics, and food, as well as the implementation of scholarship programs to encourage research. At the same time, traditional Chinese medicine is
accompanied by a vast agricultural experience, which favors the study and development of technological alternatives that take advantage of the therapeutic potential of different plant species, especially regarding such promising applicability, for example, prevention and treatment of hypertension [127].

As for the temporal evolution of the number of patent filings (Figure 7), the sharp increase in the years 2014 and 2015 is noticeable. Regarding this increase, it is important to mention that in 2010 in the National Patent Development Strategy in China, the government defined benchmarks for future performance by 2015, setting the number of patent applications to reach two million, which would quadruple the number of applications in 2010, so that by 2015, China would be among the top two countries in a number of invention patents granted to national applicants. Such targets may have greatly influenced the significant increase in the number of patents registered in 2014 and 2015 [128].

4. Conclusion

From this perspective, the anti-hypertensive activity of natural products is still little explored, especially in Western countries. In this sense, China and India have shown more results in this area than other countries, confirming the strong influence of traditional medicine in these countries.
Leaves and aerial parts were the main fractions of plants with potential exploitation. The maceration technique was the most used in obtaining the extract. The Fabaceae family was the most cited, which may indicate that more plant species belonging to this family should be studied regarding the anti-hypertensive potential. The largest number of patents related to anti-hypertensive herbs is deposited in WIPO. China is the country that invests most in research with medicinal plants for the treatment and prevention of hypertension. This study was able to provide theoretical subsidies for future research with medicinal plants on the use of natural products as a coadjuvant in the treatment of systemic arterial hypertension.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

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