Micro-fragmented adipose tissue (mFAT) associated with arthroscopic debridement provides functional improvement in knee osteoarthritis: a randomized controlled trial

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Abstract

Purpose Current conservative treatments for knee OA provide limited benefits, with symptoms relief for a short amount of time. Regenerative medicine approaches such as the use of microfragmented adipose tissue (mFAT) showed promising results in terms of durable effects and the possibility to enhance tissue healing and counteract the progression of the pathology. Nevertheless, up to today, the large part of clinical data about mFAT use refers to uncontrolled studies, especially in the surgical setting. The purpose of this study was to evaluate the effectiveness of mFAT applied in association with arthroscopic debridement (AD) for the treatment of knee OA, in terms of symptoms relief and tissue healing.

Methods This study is a prospective, randomized controlled clinical trial. 78 patients affected by knee OA grade 3–4 according to KL classification were randomly assigned to AD or AD + mFAT treatment groups. Clinical, radiological and serological assessments were performed at 6 months after treatment. Additional clinical evaluation was performed at the end of the study with an average follow-up of 26.1 ± 9.5 months. VAS, KOOS, WOMAC and SF-12 were also collected at both timepoints, KSS only at 6 months.

Results Treatment with AD + mFAT improved functional scores at both 6 months (KOOS-PS: + 11.7 ± 20.2 vs + 24.4 ± 22.5, in AD and AD + mFAT, respectively, \(p = 0.024\); KSS: + 14.9 ± 15.9 vs + 24.8 ± 23.5, in AD and AD + mFAT, respectively, \(p = 0.046\)) and 24-month follow-ups (KOOS-PS Functional subscale: − 2.0 ± 3.5 vs − 4.7 ± 4.2, in AD and AD + mFAT, respectively, \(p = 0.012\)). Lower T2-mapping scores were obtained in AD + mFAT-treated group in medial and lateral condyle compartments \((p < 0.001)\). Slight increase was observed in the levels of a serum biomarker of cartilage deposition (PIIINP) in both groups at 6-month follow-up \((p = 0.037)\).

Conclusion mFAT improves functional outcome and MRI appearance when used in association with AD, therefore supporting its use in the treatment of knee OA in an arthroscopic setting.

Keywords Knee osteoarthritis · Arthroscopic debridement · Micro-fragmented adipose tissue · Regenerative medicine · Cartilage biomarkers

Abbreviations

ACL Anterior cruciate ligament
ANOVA Analysis of variance
BMI Body Mass Index
CTx-II Cross-linked C-Telopeptide of Type II collagen
AD Arthroscopic debridement
dGEMRIC Delayed gadolinium enhanced MRI of cartilage
GAG Glycosaminoglycan
KL Kellgren–Lawrence classification

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Introduction

Knee osteoarthritis (OA) is a common condition, and a variety of conservative solutions has been proposed to control the related symptomatology. Traditional conservative therapies, including but not limited to anti-inflammatory drugs and viscosupplementation, showed short-term benefits for the management of symptoms, but they have no effect on pathology progression or tissue restoration [10]. In this scenario, an increasing attention has been addressed to the development of treatments potentially targeting the degenerative processes underlying the pathology, including mesenchymal stromal cell (MSC)-based therapy [22, 40]. As an alternative to the use of in vitro expanded MSCs, that require extensive cell manipulation that make them advanced-therapy medicinal products (ATMPs), one step approaches based on MSCs/pericytes-derived products have been considered as promising for the treatment of knee OA [20, 25, 32].

Adipose tissue, in particular, represents an easy accessible source of MSCs and its micro-fragmentation (mFAT) allows to quickly harvest a relevant volume of a minimally manipulated tissue composed by clusters containing MSCs [38]. This procedure empowers tissue regeneration by improving MSCs secretion of cytokines and angiogenic factors [7, 20, 25, 32, 41]. Moreover, mFAT contains an inferior amount of leukocytes [3] and supra-adventitial-adipose stromal cells with respect to raw adipose tissue, along with an enrichment in endothelial progenitors [31], that have been described to sustain proliferation and differentiation in an interplay with tissue resident cells [17, 19]. The biological composition of mFAT suggests the ability of micro-fragmentation technology to reduce the presence of pro-inflammatory elements, while promoting the interaction between endothelial progenitors and MSCs/pericytes, in turn activating their anti-inflammatory and pro-regenerative potential [31]. Several clinical studies investigating the intra-articular injection of mFAT for the treatment of knee OA showed significant pain and articular stiffness reduction, along with an increased articular mobility [26], together with improvements in cartilage quality as assessed by dGEMRIC (delayed gadolinium enhanced MRI of cartilage) index at 12- and 24-month follow-up [4, 11, 12]. Cohort studies confirmed these results as well as the safety of the treatment [2, 23, 24, 29]. When used as surgical adjuvant in different surgical settings, including anterior cruciate ligament (ACL) and lateral collateral ligament (LCL) reconstruction, meniscectomy, osteotomy and debridement, mFAT demonstrated to be a safe and effective approach [33, 34]. Nevertheless, the lack of a control group in most studies did not allow drawing definitive conclusion up to today. In this scenario, RCTs are needed to reinforce these observations and shed light on the real potential of mFAT in the treatment of OA.

This work reports the results of a randomized controlled trial (RCT) comparing the efficacy of autologous mFAT (obtained using Lipogems®; Lipogems International SpA, Milan, Italy) in association with arthroscopic debridement (AD) with respect to arthroscopic debridement alone for the treatment of severe knee OA (Kellgren–Lawrence, KL 3–4). While arthroscopic debridement does not represent an effective treatment for severe knee OA [28], it has been selected as a convenient surgical control for the present study to standardize the joint basal condition in each patient.

The study hypothesis is that mFAT would improve the clinical outcomes in comparison to AD procedure alone.

Methods

Study design

The study is a single-center, interventional, prospective, randomized, controlled study. Patients were enrolled after approval of the study protocol by the relevant Ethics Committee (Approval number 150/INT/2016 released on 2017 March the 9th). The protocol complies with the current Declaration of Helsinki, the EN ISO 14155: 1 and EN ISO 141155: 22 standards and the Good Clinical Practice (GCP).

Subjects, homogeneous for sex, age and physical activity, were randomly assigned to two treatment groups (Arthroscopic Debridement or Arthroscopic Debridement + mFAT) using a computer-generated 1:1 table.
A total of 78 patients with knee osteoarthritis were divided into 2 groups: Group I (Arthroscopic Debridement, AD) consisting of 39 patients undergoing arthroscopic debridement alone and Group II (AD + mFAT) consisting of 39 patients undergoing arthroscopic debridement and concomitant injection of autologous microfragmented liposorbed tissue (Lipogems®) [30].

**Patient selection**

The study was proposed to any patient affected by knee OA (3–4 KL) who met the inclusion/exclusion criteria and came to the Galeazzi Orthopaedic Institute between 13/06/2017 and 04/12/2019. Inclusion criteria were: age range 45–75 years, knee osteoarthritis grade 3–4 according to Kellgren-Lawrence classification [37], Visual Analog Scale (VAS) joint pain ≥ 6 at the time of enrollment, ligament instability ≤ grade II, mechanical axis with axial deviation (varus or valgus) under load not exceeding 10°, normal coagulation parameters (PT (INR) < 1.5) and BMI between 18 and 30 kg/m².

Exclusion criteria included: previous treatment for traumatic injuries (tibial osteotomies, tibial plateau fractures), visscosupplementation, injection of cortisone or meniscal surgery within 3 months of the procedure, vascular necrosis/osteonecrosis of the knee previous articular infections. Other exclusion criteria were presence of autoimmune diseases, general infections within the previous 6 weeks, diabetes, malignant neoplasms, metabolic diseases, cardiac, pulmonary, neurological pathologies, bleeding disorders, pregnancy. Patients undergoing radiation therapy, chemotherapy, immunosuppression, steroid therapy within the previous 8 weeks were also excluded as well as drug and/or alcoholic abuser and heavy smokers.

**Surgical procedures**

The arthroscopic knee debridement was performed by two surgeons involved in the study. The procedure consists of abundant joint lavage aimed at removing joint debris, unstable chondral fragments, cartilaginous mobile bodies, osteophytes, parts of degenerated menisci and hypertrophic synovial membrane. Other procedures such as microfractures, massive meniscectomies or synovectomies not performed.

The adipose tissue was harvested by lipoaspiration from the abdominal or thigh subcutaneous adipose tissue, depending on the physical characteristics of the patient, during the same operating session by a plastic surgeon. The patients were anesthetized locally in the harvest area (250 ml of physiological solution + 2 ml marcaine vials + 1 ml adrenaline vial) using a spinal needle, diffusely in the subcutaneous layer. The adipose tissue was harvested through a 13G cannula, equipped with a tip specifically designed not to damage the tissue, and connected to a syringe to dose the suction pressure to reduce the traumatic action on the cells. Approximately 100–120 ml of adipose tissue were collected from each patient. The adipose tissue obtained was then processed using the 120 ml Lipogems® device (Lipogems International SpA, Milan, Italy), a closed, full-immersion, low-pressure cylindrical system, aimed to wash and resize the adipose tissue [3]. Briefly, the lipoaspirate was injected through a large filter placed on top of the cylinder that contained stainless steel beads and had been prefilled with saline before beginning the processing to avoid cell damage. When the whole amount of lipoaspirate tissue was transferred to the device, the cylinder was quickly shaken for about one minute to emulsify residues of lipid droplets which were subsequently removed together with contaminating blood components by a gravity counterflow of saline solution. The washed and resized adipose tissue migrated to the top of the Lipogems device and was collected by passing through a size reduction filter into 10-ml syringes connected to the upper opening of the device. On average, per each patient 1 syringe of approximately 10 ml of mFAT was obtained. At the end of the arthroscopic debridement, 6–8 ml of mFAT obtained was injected by an 18-gauge syringe.

The patients were advised to follow a simple rehabilitation protocol including exercises for flexion–extension recovery and quadriceps strengthening, while starting to walk with two crutches with partial weight bearing. Concerning the post-operative use of drugs, patients were prescribed Ibuprofen 600 mg two times/day and, if further needed, Paracetamol 1 g (max three times/day); Clexane 4000 UI for the first 12 days or until complete recovery of deambulation and, only for Lipogems-group patients, Cefixime 400 mg 1 cp for 5 days.

**Clinical–functional and imaging post-operative evaluation protocol**

The patients in both study groups were evaluated using an accurate clinical–functional and imaging post-operative evaluation protocol.

A clinical visit was performed before (T0) and 6 months after the treatment (T6). At the same time point, VAS [15], Knee injury and Osteoarthritis Outcome Score (KOOS)-PS (and its subscales activity (A), and function (F)) [8], Knee Society Score (KSS) (and the subscale KSS-F) [13], Western Ontario and McMaster Universities osteoarthritis index (WOMAC) [35], short form-12 (SF-12, physical and mental component subscales, PCS/MCS) [16] were also collected. In particular, higher values of KOOS-PS, SF-12 and KSS are representative of better clinical conditions; on the contrary, lower values of VAS, WOMAC and KOOS-PS subscales (Activity and Function) are representative of better clinical conditions.
At longer follow-up (13–42 months), the patients were reached out by phone and asked to fill the same patient-reported outcome measures (PROMs: VAS, KOOS, WOMAC, SF-12).

Radiological analysis

All the evaluations were performed by a senior radiologist who was blinded to the group allocation of the patients. All patients underwent routine pre-operative X-ray and magnetic resonance imaging evaluation (MRI) for disease assessment (T0), together with a 6-month MRI follow-up (T6). All the follow-up MRI examinations were executed on a 1.5-T MRI scanner (Avanto, Siemens Medical Solution, Erlangen, Germany), using: sagittal turbo spin-echo (TSE) T1-weighted sequence (time of repetition, TR = 600–610 ms, time of echo, TE = 8–15 ms), TSE T2-weighted (TR = 3900–5500 ms, TE = 80–110 ms) on the axial and coronal planes, and fat-saturated proton density sequences on the three planes (TR = 2800–3100 ms, TE = 30–40 ms). Field of view was 180 mm for the axial and coronal images and 160 mm for the sagittal images. The radiological assessments considered the femorotibial angle, % Weight-Bearing Line (WBL) and Outerbridge classification.

T2 mapping analysis

For T2 mapping, a sagittal multiecho sequence (TR = 3500 ms, TE = 100 ms, number of echoes = 10, matrix 384 × 384 pixels, acquisition voxel 0.7 × 0.7 × 4 mm, bandwidth 180 Hz; slice thickness 4 mm, slice gap 4.4 mm), with a field of view of 160 mm was added. MR images were evaluated by a senior musculoskeletal radiologist who manually drew four regions of interest (ROIs), to include the whole thickness of the cartilage of medial and lateral side, both at the level of femur condyles and tibia. As for previous studies [6], the analysis was performed using a dedicated software (Olea Sphere® 3.0 software, Olea Medical®, La Ciotat, France). Before the analysis, the software pre-processed all images with an automatic correction to remove possible motion artefacts.

Biochemical analysis

The blood samples were collected pre-operatively (T0) and at 6-month (T6) follow-up, allowing them to clot at room temperature before centrifugation (3000g, 10 min). The sera were aliquoted and stored at −80 °C until assayed. Cross-linked C-Telopeptide of Type II collagen (CTx-II, Cusabio Technologies, Houston, Texas, USA) and N-Terminal Procollagen III Propeptide (PIINP, Elabscience, Houston, Texas, USA) enzyme-linked immunosorbent assays (ELISA) kits were used to determine the concentration of CTx-II and PIINP following the manufacturer’s indications. The range of detection for CTx-II was 312–20,000 pg/ml, sensitivity of 78.0 pg/ml, whereas for PIINP it was 23.4–1500 pg/ml with a sensitivity of 14.1 pg/ml.

Sample size and statistical analysis

The sample size was calculated using KOOS-PS as the primary outcome. A difference of ten points at 6 months between the two different groups of patients was assumed (α = 0.05; β = 0.20). Considering an intra-group standard deviation of 15 points, 70 subjects were necessary to obtain the desired statistical power (35 per group). Considering a 10% dropout rate, 8 patients were added to this number, for a total of 78 subjects (39 per group).

Analyses were performed using R software v4.0.3 (R Core Team, Wien, Austria). Continuous variables were tested for normal distribution using Shapiro–Wilk test. According to the result of the test, between groups comparisons were performed with unpaired t test or Mann–Whitney test, while paired tests were used to compare baseline and follow-up results in the whole cohort. Changes in clinical scores were measured using the difference between follow-up and baseline values. Percentage change was calculated as the ratio between follow-up—baseline absolute change and baseline value * 100. Between groups differences in the categorical variables were tested using Fisher’s exact test. Multiple regression linear models were used to assess the relation of CTX-II and PIINP serum levels and the clinical scores after adjustment for age and gender. The contemporary influence of two variables were assessed by a two-way ANOVA test with Bonferroni post hoc test. P values < 0.05 were considered statistically significant.

Results

Patients’ demographics

Seventy-eight patients were enrolled in the study (34 females, 44 males), with a mean age of 60.7 ± 7.9 years old. Twelve (12) patients did not complete the 6-month follow-up, with no difference between study groups (AD 9; AD + mFAT 3; p = 0.114) (Fig. 1) No significant differences in terms of baseline PROMs were observed between patients who eventually dropout and patients who completed the study. The final analysis was conducted on 67 patients.

Patients in the two groups had similar age and gender distribution.

At baseline, the two groups showed similar values for what concern VAS, WOMAC, KSS-F, SF-12 MCS and SF-12 PCS, while the groups differ significantly considering...
KSS \((p < 0.001)\) and KOOS-PS \((p < 0.05)\) (Supplementary Table 1).

**Safety of the treatment**

Four Adverse Events (AEs), all in the AD + mFAT group, were reported and notified to the Ethics Committee as well as to the National Competent Authority. Three of them were classified as serious but not related to the procedure. In details, they included a case of breast cancer diagnosed 5 months after the procedure, a case of multiple contusions after an accidental fall 8 months after the procedure and a case of pulmonary embolism occurred three months after intervention and therefore, due to the time lag between intervention and event onset, considered as non-related to the procedure. A mild procedure-related adverse event also occurred at the site of collection of the adipose tissue (hematoma formation), that resolved spontaneously in few days.

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**Fig. 1** PRISMA flowchart. Flowchart of patients’ enrollment, treatment, follow-up and analysis
Clinical evaluations

In general, higher improvements were observed in patients treated with AD + mFAT and AD alone for all scores (Fig. 2), even if at the 6-month follow-up, statistical significant differences between groups were observed only for what concern KOOS-PS and KSS. The differences in KOOS-PS was mainly due to the functional subscale (F), which showed a significant improvement ($p=0.005$).

At the mean final follow-up of $26.1 \pm 9.5$ months, the PROMs of 55 patients (28 in the AD + mFAT group and 27 in the AD group) were collected. The mean follow-up was similar between the two groups (AD $25.6 \pm 9.9$ months; AD + mFAT $26.7 \pm 9.3$ months; $p=0.545$).

Considering changes from baseline at the last follow-up, there are no significant differences between groups, although a tendency was observed in KOOS-PS improvements ($p=0.064$) (Fig. 2). Again, the reduction in this parameter was influenced by functional subscale in particular ($p=0.012$) (Fig. 2D).

Overall, the significant intra-group improvements observed at 6 months with respect to baseline were

![Fig. 2](image)

Fig. 2 Changes in patients reported outcome measures. Changes at 6 and final follow-up (calculated as value at follow-up – value at baseline) for VAS (A), KOOS-PS (B), KOOS-PS Activity subscale (C), KOOS-PS Function subscale (D), WOMAC (E), SF-12 Physical Component Score (F) and SF-12 Mental Component Score (G). Concerning KSS (H) and its functional subscale (I) values were collected at baseline and 6-month follow-up only. *$p<0.05$, **$p<0.01$, ***$p<0.001$ vs baseline (within-group difference); $^*p<0.1$ (tendency), $^p<0.05$, $^{**}p<0.01$ vs AD group (between-group difference). Clinical improvements are represented as decreases in VAS, KOOS-PS Activity subscale, KOOS-PS Function subscale and WOMAC, and as increases in KOOS-PS, KSS, KSS Function subscale, SF-12 MCS and SF-12 PCS.
maintained in both groups at the final follow-up for all PROMs tested at the final follow-up (VAS, KOOS-PS, SF12-PCS, SF12-MCS, WOMAC) (Fig. 2). A multivariate analysis considering both the time of observation and the study group confirmed the significant increases in time for both groups in KOOS-PS, VAS, SF12-P and WOMAC \( (p < 0.001) \) each, as well as the significant influence of mFAT on KOOS-PS increment during time (group*time interaction: \( p = 0.006 \)). Values for each scores and changes with respect to baseline are reported in Supplementary Table 1.

### Changes in serum levels of CTx-II and PIIINP

Data about the serum levels of CTx-II and PIIINP were available for 56 patients, 28 in control group and 28 in treatment group. The two groups showed different values at baseline for both markers, even if these differences were not statistically significant. A slight and non-significant reduction was observed in the whole cohort for what concern the catabolic marker CTx-II \( (p = 0.527) \). Also analyzing the two groups separately, no significant differences emerged between DA and AD + mFAT \( (p = 0.724) \). In the whole cohort, PIIINP showed a slight increase with respect to baseline, which resulted statistically significant \( (p = 0.037) \). Nevertheless, the median percentage increase was very small (7%). No differences were observed comparing AD and AD + mFAT. Table 1 summarizes these findings.

### Relation between the clinical scores and the serum levels of CTX-II and PIIINP

Preoperative levels of serum CTX-II demonstrated a significant correlation with pre-operative KOOS-PS (Spearman’s \( r = -0.342, p = 0.009 \)), KSS score (Spearman’s \( r = -0.364, p = 0.006 \)) and KSS-F (Spearman’s \( r = -0.385, p = 0.003 \)). After adjustment for age and gender, elements that significantly influence these scores, only KSS and KSS-F maintain the significance \( (p = 0.014) \) and \( p = 0.018 \), respectively. Serum levels of PIIINP did not correlate with any score.

### Radiological evaluations

Patients in AD and AD + mFAT groups demonstrated similar Outerbridge classification before intervention in both medial and lateral tibio-femoral compartments (Table 2).

MRI evaluations were performed at 6-month follow-up. Representative images are reported in Figs. 3 and 4. Significant differences were observed between AD and AD + mFAT-treated patients in terms of T2 mapping score, in anterior/posterior medial condyle and in anterior/posterior lateral condyle compartments \( (p < 0.001) \). In all cases, lower values were observed in the AD + mFAT group compared to AD-treated patients, with medians > 30.

No significant differences were found concerning anterior or posterior medial tibia, while in the anterior lateral tibia compartment lower values were observed in AD + mFAT group compared to AD (Fig. 5C, D). Supplementary Table 2 reports median and range for all compartments.

### Discussion

The main finding of this prospective, controlled, randomized study is that the injection of microfragmented adipose tissue in the arthroscopic setting improved functional outcomes in patients affected by knee OA.

Indeed, the use of adipose tissue derivatives produced at the point of care represents a promising strategy for the

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**Table 1** CTX-II and PIIINP levels in patients’ serum before and after the procedures

|                      | Overall \((n = 56)\) | AD \((n = 28)\) | AD + mFAT \((n = 28)\) |
|----------------------|---------------------|----------------|------------------------|
|                      | Baseline            | 6 months \(\delta\) | Baseline | 6 months \(\delta\) | Baseline | 6 months \(\delta\) |
| CTX-II (pg/mL)       | 962 ± 881           | 882 ± 588        | -80 ± 521 | 805 ± 716 | 774 ± 535 | -89 ± 439 | 1119 ± 752 | 1047 ± 601 | -71 ± 600 |
| PIIINP (pg/mL)       | 1880 ± 1922         | 1957 ± 1814*     | 77 ± 499 | 2252 ± 2356 | 2261 ± 2165 | 8 ± 573 | 1508 ± 1299 | 1655 ± 1353 | 146 ± 412 |

*\(p < 0.05\) vs baseline. Data expressed as mean ± standard deviation

**Table 2** Outerbridge classifications in medial and lateral tibio-femoral compartments

| Compartment | AD\(^a\) | AD + mFAT\(^a\) | \(P\) value |
|-------------|---------|----------------|-------------|
| Medial      | Class 1: 1 | Class 1: 4 | 0.889 |
|             | Class 2: 7 | Class 2: 2 |   |
|             | Class 3: 3 | Class 3: 4 |   |
|             | Class 4: 9 | Class 4: 11 |   |
| Lateral     | Class 1: 4 | Class 1: 4 | 0.971 |
|             | Class 2: 9 | Class 2: 11 |   |
|             | Class 3: 6 | Class 3: 4 |   |
|             | Class 4: 1 | Class 4: 2 |   |

\(^a\)X-rays were available for 20 and 21 patients in AD and AD + mFAT group, respectively.
A large number of case series demonstrated the effectiveness of these products when used as an injective treatment alone up to 12 and 24 months [1]. The idea of using mFAT in association with surgical procedures would represent a further step to exploit the regenerative potential of this product to enhance the outcomes of the treatment. Few reports up to today demonstrate that patients would benefit from surgeries (mainly osteotomy and AD) augmented with mFAT injection, and that the procedure is characterized by a high safety profile.
This study confirmed the safety of the procedure, with a single procedure-related adverse event (a small hematoma at the harvesting site of the adipose tissue, in the thigh), which resolved spontaneously in few days. To the best of the authors' knowledge, this study represents the first randomized and controlled trial evaluating the effect of mFAT compared to the index surgery alone. According to the meta-analysis by O'Connor and colleagues published in 2022 [28], arthroscopic debridement is nowadays considered to provide little or no clinically important benefit in the treatment of severe knee OA. It is notable that mFAT was able to foster higher improvements compared to AD alone, reaching minimally clinical important difference (MCID). Indeed, changes in AD + mFAT compared to AD reached MCID for VAS [27] (MCID: 1.0; between-groups difference of 1.1 mm at both 6 months and final follow-up), KOOS-PS [39] (MCID: 10; between groups difference of 12.7 and 9.9 points at 6-month and final follow-up, respectively) and KSS [18] (MCID: 9; between-groups difference of 9.9 points at 6-month follow-up) (Supplementary Table 1). Unfortunately, the dropout rate was higher than expected, probably due to the influence of COVID-19 pandemic. This resulted in a reduced power of the statistical tests, especially at the final follow-up (68%). This may explain the lack of significance in the results despite the improvements in AD + mFAT group.

Beside subjective functional improvement, it is also interesting to notice that patients treated with AD + mFAT in this trial showed significantly better T2 mapping MRI results compared to those who underwent non-augmented AD. Indeed, previous reports demonstrated that the intra-articular injection of mFAT was able to improve the glycosaminoglycan (GAG) content of knee cartilage as observed by Delayed

![Fig. 5 T2 mapping scores. T2 mapping score measured by MRI in the anterior and posterior compartments in AD and AD + mFAT-treated patients. *p < 0.05, ***p < 0.001](image)
Gadolinium-Enhanced MRI of Cartilage (dGEMRIC) at 12 and 24 months from the procedure [4, 12]. Then, the results of this study are compatible with an improvement in cartilage tissue quality possibly due to the enhancement of GAG content.

The evaluation of subjective and objective elements represents a strength of this study. Indeed, even if non-conclusive due to the high inter-individual variability, the testing for blood markers of collagen type II synthesis (PIIINP) [14] and degradation (CTx-II) [9] showed at 6 months a trend for increase in the former and decrease in the latter, suggesting a correlation between this markers and the improvement of patients conditions. Previous reports identified an association between the stage of OA pathology and these markers [14], and in the present study, this seems to be maintained with patients in the AD + mFAT group, who demonstrated worst conditions at baseline, reported consistently reduced PIIINP and increased CTx-II compared to the patients in the AD group.

The main limitation of this work is the lack of blinding of patients and assessors. Indeed, the performance of an additional procedure (liposuction) made it impossible to maintain blinding, since the performance of sham liposuction would have been considered unethical both by Authors of the study and by the IRB. In addition, final follow-up was performed by phone interview, and thus clinician-reported score (KSS) and MRI evaluations were not available at this time-point. The unbalance in the two groups due to chance in the randomization process in terms of baseline scores and conditions represents a further limitation of this study. Suitable statistical methods were employed to balance this difference, but it is possible that this bias influenced the outcomes indirectly, thus not allowing for confounders control. Another limitation is represented by the lack of baseline T2 mapping scores that would have strengthened the significance of the observed improvements in the AD + mFAT group. Nevertheless, the fact that patients in the two groups reported similar profile in terms of Outerbridge classification allows to speculate that the better imaging outcomes of AD + mFAT group are not due to a pre-existing better condition at baseline, but it is the result of the additional treatment. Indeed, all other indexes (PROMs, blood markers) indicate that patients in the AD + mFAT group bear a worse condition at baseline compared to patients treated with AD alone, and thus it is unlikely they presented better conditions at baseline in terms of tissue quality measured by imaging techniques.

Conclusions

Overall, the results of this study demonstrate that mFAT can be effective in the treatment of knee OA in an arthroscopic setting, in terms of both subjectively reported symptoms and tissue quality observed by MRI T2 mapping at 6 months post-operatively. In addition, benefits of mFAT were still observable at ~24-month follow-up, showing overall better results compared to patients treated with AD only. This is the first randomized controlled trial investigating the effect of mFAT combined with a surgical procedure, and it confirms the favorable safety profile of this treatment as well as it supports the rationale of mFAT application in knee OA. Nevertheless, given the unblinded study design adopted, further evidences are needed to draw definitive conclusions.

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Author contributions Conceptualization, MU, GMP, LM and LdG; methodology, VM, MV, AC, MDML, NR, LO, CM and LMS; writing—original draft preparation, MU, VM, MV and AC; writing—review and editing, GMP, LM and LdG; supervision, GMP, LM and LdG; project administration, MU, GMP and LdG; funding acquisition, MU, GMP, and LdG. All authors have read and agreed to the published version of the manuscript.”

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Declarations

Conflict of interest Laura de Girolamo, Marco Viganò: paid consultant for Lipogems SpA; Luca Orlandini: paid consultant for Smith and Nephews.

Ethical approval Approval number 150/INT/2016 released on 2017 March the 9th; Ethic Committee San Raffale Hospital.

Informed consent Written informed consent was obtained for all participants before enrollment.

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References

1. Agarwal N, Mak C, Bojanic C, To K, Khan W (2021) Meta-analysis of adipose tissue derived cell-based therapy for the treatment of knee osteoarthritis. Cells 10:1365

Springer
2. Barford KW, Bland L (2019) Treatment of osteoarthritis with autologous and microfragmented adipose tissue. Dan Med J 66:5565

3. Bianchi F, Maioni M, Leonardi E, Olivi E, Pasquinelii G, Valente S, Mendez AJ, Ricordi C, Raffianni M, Tremolada C, Ventura C (2013) A new nonenzymatic method and device to obtain a fat tissue derivative highly enriched in pericyte-like elements by mild mechanical forces from human liposapristes. Cell Transplant 22:2063–2077

4. Boric I, Hudetz D, Rod E, Jeleč Ž, Vrdoljak T, Skelin A, Polašek O, Plečko M, Trbojević-Akačić I, Lauc G, Primorac D (2019) A 24-month follow-up study of the effect of intra-articular injection of autologous microfragmented fat tissue on proteoglycan synthesis in patients with knee osteoarthritis. Genes 10:1051

5. Cattaneo G, De Caro A, Napoli F, Chiapale D, Trada P, Camera A (2018) Micro-fragmented adipose tissue injection associated with arthroscopic procedures in patients with symptomatic knee osteoarthritis. BMC Musculoskelet Disord 19:176

6. Chianca V, Albano D, Cuocolo R, Messina C, Gitto S, Brunetti A, Sconifenza LM (2020) T2 mapping of the trapeziometacarpal joint and triangular fibrocartilage complex: a feasibility and reproducibility study at 1.5 T. Radiol med 125:306–312

7. Crisan M, Yap S, Castella L, Chen C-W, Corselli M, Park TS, Andriolo G, Sun B, Zheng B, Zhang L, Notorte C, Teng P-N, Traas J, Schugar R, Desay BM, Badylak S, Bühring H-J, Giacino J-P, Lazzari L, Huard J, Peault B (2008) A perivascular origin for mesenchymal stem cells in multiple human organs. Cell Stem Cell 3:301–313

8. Davis AM, Perruccio AV, Canizares M, Hawker GA, Roos EM, Mailefert J-F, Lohmander LS (2009) Comparative, validity and responsiveness of the HOOS-PS and KOOS-PS to the WOMAC physical function subscale in total joint replacement for osteoarthritis. Osteoarthritis Cartilage 17:843–847

9. for the IOF-IFCC Bone Marker Standards Working Group, Vasiček J, Dulhunty A, Cooper C, Forssell H, Insall JN, Dorr LD, Scott RD, Scott WN (1989) Rationale of prospective study. Clin Orthop Anaesthesiol 34:369–382

10. Hudetz D, Boric I, Rod E, Jeleč Ž, Kunovac B, Polašek O, Vrdoljak T, Plečko M, Skelin A, Polašek O, Lauc G, Primorac D (2019) Early results of intra-articular micro-fragmented liposapristes injection in patients with late stages knee osteoarthritis: a prospective study. Croat Med J 60:227–236

11. Hudetz D, Boric I, Rod E, Jeleč Ž, Radic A, Vrdoljak T, Skelin A, Lauc G, Trbojević-Akačić I, Plečko M, Polašek O, Primorac D (2017) The effect of intra-articular injection of autologous microfragmented fat tissue on proteoglycan synthesis in patients with knee osteoarthritis. Genes 8:270

12. Insall JN, Dorr LD, Scott RD, Scott WN (1989) Rationale of the Knee Society clinical rating system. Clin Orthop Relat Res 248:13–14

13. Jiao Q, Wei L, Chen C, Li P, Wang X, Li Y, Guo L, Zhang C, Wei X (2016) Cartilage oligomeric matrix protein and hyaluronic acid are sensitive serum biomarkers for early cartilage lesions in the knee joint. Biomarkers 21:146–151

14. Joyce CRB, Zutshi DW, Hruby V, Mason RM (1975) Comparison of fixed interval and visual analogue scales for rating chronic pain. Eur J Clin Pharmacol 8:415–420

15. Kodraliu P, Mosconi N, Groth GG (2001) Subjective health status assessment: evaluation of the Italian version of the SF-12 Health Survey. Results from the MiOS Project. J Epidemiol Biostat 6:305–316

16. Liang T, Zhu L, Gao W, Tong M, Ren J, Yao H, Wang K, Shi D (2017) Coculture of endothelial progenitor cells and mesenchymal stem cells enhanced their proliferation and angiogenesis through PDGF and Notch signaling. FEBS Open Bio 7:1722–1736

17. Liszár-Utrilla A, González-Parreño S, Martínez-Mendez D, Miralles-Muñoz FA, López-Prats FA (2020) Minimal clinically important differences and substantial clinical benefits for Knee Society Scores. Knee Surg Sports Traumatol Arthrosc 28:1473–1478

18. Lubić B, Binder A, Herrmann M, Dutttenhoefer F, Richards RG, Nerlich M, Alini M, Verrier S (2014) Direct cell-cell contact between mesenchymal stem cells and endothelial progenitor cells induces a pericyte-like phenotype in vitro. BioMed Res Int 2014:1–10

19. Lopa S, Colombini M, Moretti M, de Girolamo L (2019) Injjective mesenchymal stem cell-based treatments for knee osteoarthritis: from mechanisms of action to current clinical evidences. Knee Surg Sports Traumatol Arthrosc 27:2003–2020

20. Magnanelli S, Scepis D, Di Benedetto P, Natali S, Causero A, Zorzi C (2020) Open–wedge high tibial osteotomy associated with lipogems® intra-articular injection for the treatment of varus knee osteoarthritis—retrospective study. Acta Bio Medica Atenei Parmensis 91:e2020022

21. Maheshwer B, Polce EM, Paul K, Williams BT, Wolfson TS, Yanke A, Verma NN, Cole BJ, Chahla J (2021) Regenerative potential of mesenchymal stem cells for the treatment of knee osteoarthritis and chondral defects: a systemic review and meta-analysis. Arthroscopy 37:362–378

22. Malanga GA, Chirichella PS, Hogaboos NM, Capella T (2021) Clinical evaluation of micro-fragmented adipose tissue as a treatment option for patients with meniscus tears with osteoarthritis: a prospective pilot study. Int Orthop 45:473–480

23. Maunder K, Bowers R, Easley K, Fausel Z, Robinson R (2019) Functional outcomes following microfragmented adipose tissue versus bone marrow aspirate concentrate injections for symptomatic knee osteoarthritis. Stem Cells Transl Med 8:1149–1156

24. Moroni L, Fornasarli PM (2013) Human mesenchymal stem cells: A bank perspective on the isolation, characterization and potential of alternative sources for the regeneration of musculoskeletal tissues. J Cell Physiol 228:680–687

25. Moshref SKAM, Al-Hibshi AM, Jamal S (2014) Intra-articular injectionof autologous fat micro-graft for the treatment of knee osteoarthritis: preliminary experience. Life Sci 111:30–35

26. Myles PS, Myles DB, Galagher W, Boyd D, Chew C, MacDonald N, Dennis A (2017) Measuring acute postoperative pain using the visual analog scale: the minimal clinically important difference and patient acceptable symptom state. Br J Anaesth 118:424–429

27. O’Connor D, Johnston RV, Brignardello-Petersen R, Poolman RW, Cyril S, Vandvik PO, Buchbinder R (2022) Arthroscopic surgery for degenerative knee disease (osteoarthritis including degenerative meniscal tears). Cochrane Database Syst Rev. https://doi.org/10.1002/14651858.CD014328

28. Panchal J, Malanga G, Shinkop M (2018) Safety and efficacy of percutaneous injection of lipogems micro-fractured adipose tissue for osteoarthritic knees. Am J Orthop (Belle Mead NJ) 47:1–11

29. Peretti GM, Ulivi M, De Girolamo L, Meroni V, Lombardo MD, Mangiavini L (2018) Evaluation of the use of autologous microfragmented adipose tissue in the treatment of knee osteoarthritis: preliminary results of a randomized controlled trial. J Biol Regul Homeost Agents 32:193–199

30. Polancic Z, Hudetz B, Jeleč Ž, Vrdoljak S, Plečko T, Nogalo P (2019) Immunophenotyping of a stromal vascular fraction from microfragmented liposapristes used in osteoarthritis cartilage treatment and its liposapristes counterpart. Genes 10:474
32. Primorac D, Molnar V, Rod E, Jeleč Ž, Čukelj F, Matišić V, Vrdoljak T, Hudetz D, Hajsko H, Borić I (2020) Knee osteoarthritis: a review of pathogenesis and state-of-the-art non-operative therapeutic considerations. Genes 11:854
33. Russo A, Condello V, Madonna V, Guerriero M, Zorzi C (2017) Autologous and micro-fragmented adipose tissue for the treatment of diffuse degenerative knee osteoarthritis. J Exp Ortop 4:33
34. Russo A, Screpis D, Di Donato SL, Bonetti S, Piovan G, Zorzi C (2018) Autologous micro-fragmented adipose tissue for the treatment of diffuse degenerative knee osteoarthritis: an update at 3 year follow-up. J Exp Ortop 5:52
35. Salaffi F, Leardini G, Canesi B, Mannoni A, Fioravanti A, Caporali R, Lapadula G, Punzi L (2003) Reliability and validity of the Western Ontario and McMaster Universities (WOMAC) Osteoarthritis Index in Italian patients with osteoarthritis of the knee. Osteoarthritis Cartilage 11:551–560
36. Schiavone Panni A, Vasso M, Braile A, Toro G, De Cicco A, Viggiano D, Lepore F (2019) Preliminary results of autologous adipose-derived stem cells in early knee osteoarthritis: identification of a subpopulation with greater response. Int Orthop 43:7–13
37. Scott WW, Lethbridge-Cejku M, Reichle R, Wigley FM, Tobin JD, Hochberg MC (1993) Reliability of grading scales for individual radiographic features of osteoarthritis of the knee. The Baltimore longitudinal study of aging atlas of knee osteoarthritis. Invest Radiol 28:497–501
38. Shariatzadeh M, Song J, Wilson SL (2019) The efficacy of different sources of mesenchymal stem cells for the treatment of knee osteoarthritis. Cell Tissue Res 378:399–410
39. Singh JA, Luo R, Landon GC, Suarez-Almazor M (2014) Reliability and clinically important improvement thresholds for osteoarthritis pain and function scales: a multicenter study. J Rheumatol 41:509–515
40. Song Y, Zhang J, Xu H, Lin Z, Chang H, Liu W, Kong L (2020) Mesenchymal stem cells in knee osteoarthritis treatment: a systematic review and meta-analysis. J Orthop Translat 24:121–130
41. Vezzani B, Shaw I, Lesme H, Yong L, Khan N, Tremolada C, Pèault B (2018) Higher pericyte content and secretory activity of microfragmented human adipose tissue compared to enzymatically derived stromal vascular fraction. Stem Cells Transl Med 7:876–886

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