Prognostic Relevance of BRCA1 Expression in Survival of Patients With Cervical Cancer

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Objective: BRCA1 expression can be lost by a variety of mechanisms including germline or somatic mutation and promotor hypermethylation. Given the potential importance of BRCA1 loss as a predictive and prognostic biomarker in several cancers, the objective of this study was to investigate BRCA1 expression using immunohistochemistry (IHC) in cervical cancer and its possible prognostic relevance.

Methods: Seventy patients with cervical cancer were enrolled in this study. Samples from each tumor were stained for BRCA1 and reviewed independently by gynecologic pathologists blinded to the BRCA status. Kaplan–Meier methods were used to estimate overall survival according to BRCA1 expression. Differentially expressed genes (DEGs) by BRCA1 expression were selected using GSE44001 dataset, which included 300 samples treated with radical hysterectomy. In addition, cox regression analysis with backward elimination was performed to select independent prognostic markers. Gene set enrichment analysis (GSEA) was done using these DEGs.

Results: BRCA1 IHC was positive in 62.9% (44/70) of cases. Patients with BRCA1 expression showed better overall survival (100% vs. 76.2%, HR 0.20, 95% CI 0.04 – 0.99, p = 0.028) than those without BRCA1 expression. Analysis of gene expression profiles according to BRCA1 expression identified 321 differentially expressed mRNAs. Gene set enrichment analysis results showed two dysregulated pathways (VEGF_A_UP.V1_DN and E2F1_UP.V1_UP). Of these DEGs, alterations of 20 gene signatures were found to be independently associated with survival outcomes of patients.

Conclusions: BRCA1 expression in cervical cancer tissue is associated with survival. In addition, the identification of specific gene alterations associated with BRCA1 expression could help to provide individualized prediction in these patients.

Keywords: BRCA1, gene signature, cervical cancer, immunohistochemistry, prognosis
INTRODUCTION

Cervical cancer is the fourth most frequent malignancy in women and the seventh most common cancer overall worldwide (1). In Korea, it is one of common gynecologic cancers, accounting for 9.8% of newly diagnosed malignancies in women (2). To treat invasive cervical cancer, surgical resection and/or radiation and cisplatin-based chemotherapy are commonly used. After recurrence, there are only a few feasible treatment options. The prognosis of patients with recurrence of cervical cancer is generally poor (3, 4). Most cervical cancer patients become resistant to treatment. The resistance is either intrinsic or acquired during treatment (5). Chemoradiotherapy may induce DNA double-strand break (DSB), which is considered a lethal form of DNA damage. DNA damage can induce a series of molecular responses responsible for the maintenance of genome integrity (6). Deficiencies in DSB response and repair could lead to intrinsic resistance which will affect the prognosis of patients.

Loss of BRCA1 function in cancer cells can lead to absence of intact homologous recombinant DNA repair, and result in cells being more sensitive to agents that cause DSBs. BRCA1 loss is known to be associated with the loss of heterozygosity (7) or promoter hypermethylation (8, 9). It has been shown that miRNA regulation of BRCA1 mRNA stability might contribute to BRCA1 silencing (10, 11). A method for identifying BRCA1 expression would be useful for prognostication of cervical cancer patients. However, studies on genomic profiles of cervical cancer patients regarding BRCA1 expression are insufficient.

Despite recent decrease in cost and improved efficiency of sequencing, genetic testing is still expensive and time-consuming. Another alternative for assessing genetic implications in cancer is by identifying pathological features based on protein expression (8). In this study, we performed BRCA1 expression analysis using immunohistochemistry (IHC), an inexpensive and widely available technique, in cervical cancer patients for its prognostic significance, and determined the correlation between its mRNA and protein expression. In addition, we identified genes associated with BRCA1 expression in cervical cancer to determine its clinical significance.

MATERIALS AND METHODS

Patients

We analyzed data of 70 cervical cancer patients treated at the Department of Gynecologic Oncology, Samsung Medical Center from 2002 to 2009. Patients with rare histology or an advanced stage treated primarily with radiotherapy or chemotherapy were excluded. The tissue specimens and medical records were collected after we obtained informed consent from included patients and approval from the institutional review boards (IRB) of Samsung Medical Center, Seoul, Republic of Korea (approval no. 2009-09-002-002 and 2015-07-122). All of included patients had undergone radical hysterectomy with or without pelvic/para-aortic lymph node dissection as primary treatment. According to pathologic reports, patients with risk factors such as positive resection margin, lymph node metastasis, parametrial involvement, and stromal invasion of more than half of the cervix received adjuvant radiation treatment with or without a concurrent chemotherapy. After primary and adjuvant treatment, patients were routinely followed up every 3 months for the first 2 years, every 6 months for the next 3 years, and every 12 months thereafter. Progression-free survival (PFS) was defined as the time from the initial surgical treatment to recurrence or last follow-up. Overall survival (OS) was defined as the time from the initial surgical treatment to death or last follow-up.

Immunohistochemistry (IHC) and Quantitative Evaluation

IHC was performed for FFPE 4-μm thick tissue sections using a standard protocol. Tissue blocks were used for routine pathological evaluations. Original archived hematoxylin-eosin–stained slides were reviewed by a pathologist. Following rehydration and deparaffinization of tissue sections, antigen retrieval for 20 minutes using 0.01 M citrate buffer (pH 6.0) and a boiling process (pressure cooker) was performed. Endogenous peroxidase block was done with H2O2 (3%). Incubation of sections was done with primary antibodies at 20°C (room temperature) for 30 minutes in a humid chamber. IHC staining was done using a primary monoclonal antibody for BRCA1 (BioVision Inc., OH, USA, clone#3364-100) at a dilution of 1:400. Sections were sensitized and then incubated with a secondary antibody. The peroxidase reaction development was done using diaminobenzidine tetrachloride (DAB) as a chromogen. Counterstain of sections was done with hematoxylin.

Staining results included the percentage of positive cells and intensities of stained cells in the nucleus and cytoplasm. IHC score was graded visually by pathologists. The staining intensity grade was divided to 4 groups (negative, weak, moderate, or strong). For avoidance of bias, the IHC score grading was performed independently by two pathologists, and quality-control was done by score comparison for equivalence (14). Moderate or strong intensity grade was considered as positive for BRCA1 expression (Figure 1). In case of discrepancy in judgement, conclusion was drawn by discussion using a multi-head microscope.

In Silico Analysis for GSE44001 for the Selection of Differentially Expressed Genes According to BRCA1

To examine the prognostic significance of BRCA1 expression, the Gene Expression Omnibus ( GEO) data were analyzed (15, 16). From a total of 300 available patient samples of GSE44001 (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE44001), forty-nine samples were used in the IHC analysis of current study. For mRNA analysis, cDNA mediated annealing, selection, extension, and ligation (DSL) assay data were used (15). Obtained data were dichotomized by the distribution of expression of genes.
To identify differentially expressed genes according to BRCA1 expression, normalized expression levels were used. Differences in expression were featured by correlation coefficient (> 0.5 or < -0.5) and associated p-value (FDR-adjusted p < 0.05) in Pearson correlation analysis.

Gene Set Enrichment Analysis (GSEA)
To determine pathways associated with BRCA1 expression, gene set enrichment analysis (GSEA) was performed using R package (GSEA.1.0.R) (17, 18). Curated oncogenic signatures v5.2 (including 189 gene sets) (https://www.gsea-msigdb.org/gsea/msigdb/genesets.jsp?collection=C6) were chosen to perform enrichment analysis among the two groups. Gene sets were first preprocessed to exclude those with < 10 or > 500 genes. The phenotype label was set as BRCA-high vs. BRCA-low. The t-statistic mean of genes was computed for each pathway using a permutation test with 1000 replications. For discovery, gene sets with a normalized p-value < 0.05 were chosen as significantly enriched.

Statistical Analysis
For all statistical analysis, R software, version 3.1.3 (R Foundation, Vienna, Austria; http://www.R-project.org) was used. Student’s t-test or Mann–Whitney U-test was used for analysis of expression levels of proteins according to clinicopathological characteristics. Analysis using Spearman’s rho coefficient was performed for assessing correlations between proteins and mRNA expression levels. Survival distributions were estimated using the Kaplan–Meier method. The log-rank test was used for analyzing the relation of survival and each parameter. A Cox proportional hazards model was used for identification of independent predictors of survival. A p-value of < 0.05 was considered as significant.

RESULTS
Clinicopathological Characteristics of Patients (Patient and Tumor Characteristics)
We analyzed 70 patients with cervical cancer. In this patient group, thirteen cases developed recurrence, and eight patients died within a mean follow-up time of 52 months (range, 1–96 months). The clinicopathological characteristics of these 70 patients according to BRCA1 protein expression status are presented in Table 1. The median age was 44 (range, 41–48) years for BRCA1 negative patients and 47.5 (range, 40–59.5) years for BRCA1 positive patients. Most patients were at stage IB1 (69.2% for BRCA1 negative, and 84.1% for BRCA1 positive patients). Most patients were at stage IB1 (69.2% for BRCA1 negative, and 84.1% for BRCA1 positive patients). There were no significant differences in clinicopathological characteristics of patients between the two groups except for histologic cell type. Higher proportion of squamous cell carcinoma (SCC, 93.2%) was observed in the BRCA1 positive group than in the BRCA1 negative group (69.2%). Six (23.1%) patients in the BRCA1 negative group and 2 (4.5%) patients in the BRCA1 positive group died.
We performed immunohistochemistry staining for BRCA1 using cervical cancer tissues. BRCA1 expression was mainly seen in the cytoplasm or the nucleus. Representative examples of positive and negative staining of BRCA1 are shown in Figure 1.

We examined correlations between BRCA1 protein and mRNA expression levels by using Spearman’s rho coefficient. There was a positive correlation (r = 0.245, p = 0.089, Figure 2) between the BRCA1 protein and mRNA expression levels, although the correlation was not statistically significant. In Figure 2, red dots indicate adenocarcinoma and black dots imply squamous cell carcinoma. Adenocarcinoma was mostly observed in area of high BRCA1 mRNA expression and low BRCA1 IHC expression. It can be inferred that mRNA expression and protein expression have a different association with cancer prognosis and further research is needed in this regard.

Kaplan-Meier curves of cervical cancer patients with positive or negative BRCA1 expression are shown in Figure 3. The BRCA1 negative group showed inferior survival outcomes, including worse PFS (Figure 3A) and OS (Figure 3B). However, significant worse survival outcome was only found for the OS (HR: 0.20, 95% CI: 0.04 – 0.99, p = 0.028), not the PFS (HR: 0.46, 95% CI: 0.15 – 1.37, p = 0.151). The prognostic significance of BRCA1 expression showed a trend in multivariate analysis (HR: 0.21, 95% CI: 0.04 – 1.11) (Table 2). However, it was not statistically significant (p = 0.06).

Identification of Molecular Signatures Associated With BRCA1
To further examine the prognostic significance of BRCA1 expression, we analyzed the GEO database (GSE44001). Analysis of gene expression profiles by BRCA1 expression identified 320 genes whose expression levels were correlated with BRCA1 expression (correlation coefficient > 0.5 or < -0.5 and p-value < 0.05) (Supplementary Table S1). Of these, 178 genes were positively correlated while 142 genes were negatively correlated with BRCA1 expression.

Of these 320 genes, Cox regression with backward elimination was performed to select genes as independent prognostic markers. Twenty genes (PGK1, FBLN5, CCR6, CEP170, FZD1,}

### TABLE 1 | Clinical characteristics of patients by BRCA1 protein expression.

|                          | BRCA1 negative (n = 26) | BRCA1 positive (n = 44) | p-value |
|--------------------------|-------------------------|-------------------------|---------|
| Age, years, median (range) | 44 (41–48)              | 47.5 (40–59.5)          | 0.374   |
| Stage, n (%)             |                         |                         | 0.245   |
| IB1                      | 18 (69.2)               | 37 (84.1)               |         |
| IIA                      | 8 (30.8)                | 7 (15.9)                |         |
| SCC Ag, ng/mL, median (range) | 1.6 (0.9–4.2)          | 1.6 (1.1–5.1)           | 0.484   |
| Cell type                |                         |                         | 0.020   |
| SCC                      | 18 (69.2)               | 41 (93.2)               |         |
| AC                       | 8 (30.8)                | 3 (6.8)                 |         |
| Size, cm, median (range) | 3.5 (2.2–5.0)           | 3.1 (2.1–4.5)           | 0.368   |
| Parametrial invasion, n (%) |                      |                         | 0.732   |
| No                       | 25 (96.2)               | 40 (90.9)               |         |
| Yes                      | 1 (3.8)                 | 4 (9.1)                 |         |
| Resection margin with tumor, n (%) |          |                         | 0.789   |
| No                       | 25 (96.2)               | 44 (100.0)              |         |
| Yes                      | 1 (3.8)                 | 0                       |         |
| LVSI, n (%)              |                         |                         | 0.252   |
| No                       | 5 (33.3)                | 19 (55.9)               |         |
| Yes                      | 10 (66.7)               | 15 (44.1)               |         |
| Deep tumor invasion, n (%) |                      |                         | 0.288   |
| ≤1cm                     | 14 (56.0)               | 17 (39.5)               |         |
| >1cm                     | 11 (44.0)               | 26 (60.5)               |         |
| LN metastasis, n (%)     |                         |                         | 0.714   |
| No                       | 14 (53.8)               | 27 (61.4)               |         |
| Yes                      | 12 (46.2)               | 17 (38.6)               |         |
| Recurrence, n (%)        |                         |                         | 0.288   |
| No                       | 19 (73.1)               | 38 (86.4)               |         |
| Yes                      | 7 (26.9)                | 6 (13.6)                |         |
| Death, n (%)             |                         |                         | 0.049   |
| No                       | 20 (76.9)               | 42 (95.5)               |         |
| Yes                      | 6 (23.1)                | 2 (4.5)                 |         |

**SCC**, Squamous cell carcinoma; **AC**, Adenocarcinoma; **LVSI**, Lympho-vascular space invasion; **LN**, Lymph node.

Bold values mean they are statistically significant.

**BRCA IHC Expression and Prognostic Significance**
We performed immunohistochemistry staining for BRCA1 using cervical cancer tissues. BRCA1 expression was mainly seen in the cytoplasm or the nucleus. Representative examples of positive and negative staining of BRCA1 are shown in Figure 1. We examined correlations between BRCA1 protein and mRNA expression levels by using Spearman’s rho coefficient. There was a positive correlation (r = 0.245, p = 0.089, Figure 2) between the BRCA1 protein and mRNA expression levels, although the correlation was not statistically significant. In Figure 2, red dots indicate adenocarcinoma and black dots imply squamous cell carcinoma. Adenocarcinoma was mostly observed in area of high BRCA1 mRNA expression and low BRCA1 IHC expression. For further investigation, when patient’s overall survival information was applied (data not shown), death events also showed a tendency to be associated with high BRCA1 mRNA expression and low BRCA1 IHC expression. It can be inferred that mRNA expression and protein expression have a different association with cancer prognosis and further research is needed in this regard.

Kaplan-Meier curves of cervical cancer patients with positive or negative BRCA1 expression are shown in Figure 3. The BRCA1 negative group showed inferior survival outcomes, including worse PFS (Figure 3A) and OS (Figure 3B). However, significant worse survival outcome was only found for the OS (HR: 0.20, 95% CI: 0.04 – 0.99, p = 0.028), not the PFS (HR: 0.46, 95% CI: 0.15 – 1.37, p = 0.151). The prognostic significance of BRCA1 expression showed a trend in multivariate analysis (HR: 0.21, 95% CI: 0.04 – 1.11) (Table 2). However, it was not statistically significant (p = 0.06).
SNORD35A, COL1A2, RECK, SNORA75, SNORA2B, SERINC5, SNORA11D, MAPK14, SNORD21, SDHAP3, TMSB4X, LAMB1, SNORD8, SCARNA5, and SNORA50A) were selected. Univariate analysis results using these selected 20 genes are shown in Table 3. The Kaplan-Meier survival curve is shown in Supplementary Figure S1.

**Gene Set Enrichment Analysis (GSEA)**

Gene set enrichment analysis (GSEA) results showed one up-regulated pathway and one down-regulated pathway in cervical cancer (Supplementary Table S2 and Supplementary Figure S2). VEGF A-regulated gene pathway was downregulated, and E2F1-regulated gene pathway was upregulated. E2F1-regulated genes modulate the transition from quiescence into DNA synthesis, or have roles in apoptosis, signal transduction, membrane biology, and transcription repression. The other dysregulated pathway without statistical significance were PRC2_EZH2_UP.V1_UP (p = 0.067), CSR_LATE_UP.V1_UP (p = 0.055), HOXA9_DN.V1_DN (p = 0.081), TBK1.DF_DN (p = 0.059), GCNP_SHH_UP_LATE.V1_UP (p = 0.214), CSR_EARLY_UP.V1_UP (p = 0.152),

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**FIGURE 2** | Correlation between BRCA1 mRNA and protein expression (IHC score) in cervical cancer tissues shown by Spearman’s rho coefficient. (Red dots indicate adenocarcinoma, and black dots indicate squamous cell carcinoma).

**FIGURE 3** | Kaplan-Meier graph showing progression-free survival (A) and overall survival (B) according to BRCA1 expression in patients with cervical cancer.
TABLE 2 | Multivariate cox regression analysis for the overall survival.

| HR     | 95% CI     | p-value |
|--------|------------|---------|
| BRCA1 positive | 0.20     | (0.04 – 1.11) | 0.066 |
| Stage   | 4.12      | (0.85 – 19.93) | 0.078 |
| LN metastasis | 0.75     | (0.15 – 3.74) | 0.730 |
| Cell type | 1.86      | (0.38 – 9.12) | 0.442 |
| Parametrial invasion | 3.32     | (0.49 – 22.45) | 0.219 |

HR, hazard ratio; CI, confidence interval.

TABLE 3 | Univariate analysis for overall survival of 20 selected genes correlated with BRCA1.

| Gene | HR     | 95% CI     | p-value |
|------|--------|------------|---------|
| PGK1 | 5.42   | (2.01 – 14.58) | 0.001 |
| FBLN6 | 0.37   | (0.14 – 1) | 0.049 |
| CCR6 | 0.33   | (0.15 – 0.72) | 0.006 |
| CEP170 | 0.26 | (0.07 – 0.92) | 0.036 |
| FZD1 | 0.23   | (0.03 – 1.65) | 0.143 |
| SNORD35A | 0.48 | (0.28 – 0.84) | 0.01 |
| COL1A2 | 0.08   | (0.02 – 0.42) | 0.002 |
| RECK | 3.49   | (1.04 – 11.67) | 0.042 |
| SNORA75 | 0.39  | (0.14 – 1.05) | 0.062 |
| SNORA2B | 0.09   | (0.01 – 0.61) | 0.013 |
| SERINC5 | 0.34  | (0.09 – 1.3) | 0.116 |
| SNORA11D | 2.62 | (0.88 – 7.77) | 0.084 |
| MAPK1 | 3.57   | (1.02 – 12.53) | 0.047 |
| SNORD21 | 0.28  | (0.06 – 1.36) | 0.116 |
| SDHAP3 | 0.46   | (0.19 – 1.14) | 0.095 |
| TMSB4X | 6.79   | (1.06 – 43.35) | 0.043 |
| LAMB1 | 3.8    | (1.13 – 13.1) | 0.035 |
| SNORD8 | 7.31   | (1.18 – 45.49) | 0.033 |
| SCARN5 | 5.89   | (1.12 – 30.93) | 0.036 |
| SNORA50A | 5.29  | (1.02 – 27.41) | 0.047 |

HR, hazard ratio; CI, confidence interval.

ERB2_UP.V1_DN (p = 0.246), TBK1_DF_UP (p = 0.599), G.N.P_S.H.U.P._E.A.R.L.Y.V1_UP (p = 0.620), CAMP_UP.V1_DN (p = 0.765), and LTE2_UP.V1_DN (p = 0.889).

DISCUSSION

This study was initiated with hypothesis that BRCA1 expression may be associated to prognosis in cervical cancer. In the current study, we investigated the prognostic relevance of expression of BRCA1 in cervical cancer by IHC analysis of BRCA1 in cervical cancer tissue for comparison of overall survival according to BRCA1 expression. Additional analysis was done to find genes associated with BRCA1 expression using Gene set enrichment analysis (GSEA) method. This study showed that alteration associated to BRCA1 expression was associated with survival outcomes of patients in uterine cervical cancer.

In current study, low BRCA1 expression was associated with inferior prognosis in cervical cancer. As similar results, a previous study on BRCA expression in cancer has revealed that promoter hypermethylation can inactivate genes in the Fanconi Anemia (FA)-BRCA pathway, including BRCA1 and BRCA2 (19). Narayanan et al. (20) have reported that BRCA1 promoter hypermethylation is present in 6.1% of cervical cancer patients. Promoter hypermethylation of FANCF gene could disrupt the FA-BRCA pathway, resulting in cisplatin resistance in cervical cancer (21). Interestingly, other studies have shown different results, as BRCA-deficient cells are inefficient in repairing DNA damage through homologous recombination (HR) (22, 23) and that they are more sensitive to chemotherapeutic drugs. Also, Balacescu et al. (24) have shown that overexpression of BRCA1 and BRCA2 in patients with advanced cervical cancer is associated with treatment failure. In a recent in vitro study, Wen et al. (25) have also suggested that BRCA1 overexpression might be a mechanism involved in the enhanced chemoradiation resistance of cervical squamous cell carcinoma. Although previous studies tried to reveal prognostic relevance of BRCA expression, these studies did not show consistent results. The reason for showing different results may be that in addition to the known relationship between gene mutation and cancer prognosis, it can be inferred that mRNA expression and protein expression have a different association with cancer prognosis caused by mechanism such as post-transcriptional modifications. Further research is needed in this regard.

Determining BRCA1 protein expression by IHC can theoretically detect BRCA1 loss-of-function tumors. Previously, studies have shown BRCA1 IHC is effective method to screen for genetic BRCA1 alterations (12, 26), and BRCA1 IHC testing has been performed for ovarian cancer with excellent correlation with survival (12, 26–29). It can be inferred that these results will be similar for uterine cervical cancer as well but further study is warranted. Currently, BRCA1 germline testing and Next Generation Sequencing (NGS) testing are available options for BRCA assessment. However, these genetic tests remain expensive and time-consuming. As the clinical significance of somatic mutations and promoter hypermethylation is further ascertained, BRCA1 IHC testing can be an effective method to identify BRCA1 expression through both genetic and epigenetic mechanisms.

In this study, GSEA result showed that VEGF A-regulated gene pathway downregulation was associated with BRCA1 expression. VEGF plays a role in tumor angiogenesis which is essential for tumor growth, invasion, and metastasis (30). However, previous studies on the relationship between angiogenesis marker and prognosis of cervical cancer have reported conflicting results. Zijlmans et al. (31) reported that VEGF mRNA expression correlated with CD31 expression, which is a marker for microvessel density (MVD) analysis, and CD31 expression was associated with lymphovascular space invasion and lymph node metastasis. With same context, a study in stage IB cervical cancer showed that patients with high tumor MVD has poor survival (5-year survival 63% vs 90%) (32). These are consistent with our finding, assuming that BRCA1 positive tumors with VEGF A pathway downregulation, leading to low angiogenesis and better prognosis. On the contrary, there are reports that decreased level of tumor angiogenesis, which was represented as CD31 MVD, is associated with poor survival in high-risk, early stage cervical cancer (33, 34). They hypothesized that survival advantage in
high MVD may owe to improved chemoradiation response in well-vascularized tumors. Further study is required since the relationship between tumor angiogenesis and prognosis has not yet been elucidated.

To our best knowledge, this study is first to show association of BRCA1 IHC expression and prognosis in uterine cervical cancer patients. However, this study has some limitations. First, we used a small number of samples for analysis. Second, although the relationship between IHC results and prognosis was shown, a comparative NGS study was not conducted to determine whether gene expression in cervical cancer was consistent with the BRCA1 IHC results. Previous studies have used an additional validation with microarray data for associated gene alterations but further study is needed. Also, we were not able to show a mechanism to explain the relationship between low BRCA1 expression and inferior survival in current analysis. Further studies with larger sample sizes are warranted to determine whether BRCA1 expression is an important prognostic factor that determines poor response to radiation and chemotherapy for subgroups of patients with cervical cancer.

CONCLUSIONS

In conclusion, low expression of BRCA1 in cervical cancer has potential prognostic and therapeutic significance. Our results indicated that BRCA1 IHC may be effective method to identify BRCA1 expression through both genetic and epigenetic mechanisms to identify these patients in the clinic.

DATA AVAILABILITY STATEMENT

Publicly available datasets were analyzed in this study. This data can be found here: https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE44001, https://www.gsea-msigdb.org/gsea/msigdb/genesets.jsp?collection=C6.

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ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the institutional review boards (IRB) of Samsung Medical Center, Seoul, Republic of Korea. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

Conceptualization, CHC and Y-YL. Methodology, CHC and S-JB. Formal Analysis, EP. Investigation, CK. Data Curation, YC and SO. Writing – Original Draft Preparation, C-SC and EP. Writing – Review & Editing, EP and C-SC. Supervision, T-JK. Funding Acquisition, J-WL and B-GK. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2021.770103/full#supplementary-material

Supplementary Figure 1 | Kaplan-Meier graph showing overall survival of BRCA1 correlated genes.

Supplementary Figure 2 | Gene Set Enrichment Analysis of cervical cancer.
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