White matter microstructure relates to motor outcomes in myotonic dystrophy type 1 independently of disease duration and genetic burden

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Deficits in white matter (WM) integrity and motor symptoms are among the most robust and reproducible features of myotonic dystrophy type 1 (DM1). In the present study, we investigate whether WM integrity, obtained from diffusion-weighted MRI, corresponds to quantifiable motor outcomes (e.g., fine motor skills and grip strength) and patient-reported, subjective motor deficits. Critically, we explore these relationships in the context of other potentially causative variables, including: disease duration, elapsed time since motor symptom onset; and genetic burden, the number of excessive CTG repeats causing DM1. We found that fractional anisotropy (a measure of WM integrity) throughout the cerebrum was the strongest predictor of grip strength independently of disease duration and genetic burden, while radial diffusivity predicted fine motor skill (peg board performance). Axial diffusivity did not predict motor outcomes. Our results are consistent with the notion that systemic degradation of WM in DM1 mediates the relationship between DM1 progression and genetic burden with motor outcomes of the disease. Our results suggest that tracking changes in WM integrity over time may be a valuable biomarker for tracking therapeutic interventions, such as future gene therapies, for DM1.

Myotonic dystrophy type 1 (DM1) is a trinucleotide repeat disorder, classically characterized by prolonged muscle contractions (myotonia), progressive muscle wasting, and weakness1. Histopathology and molecular studies of DM1 show that the neuromuscular dysfunction is due to a primary abnormality in muscle function. The transcribed mutant DMPK mRNA is toxic and leads to a ‘spliceopathy’ resulting in shifts to multiple alterations in proteins involved with skeletal muscle function in DM12. However, it is also well established that the brain is significantly affected in DM13. Although most of the CNS findings have been focused on linking the brain findings to cognitive and behavioral issues, the extent that brain abnormalities may play a role in neuromuscular dysfunction in DM1 is less well established.

Microstructural white matter (WM) pathology in the central nervous system is one of the most robust and reproducible CNS observations in DM14. Fractional Anisotropy (FA), is obtained with diffusion-weighted magnetic resonance imaging, and is an index of non-uniform movement of water molecules ranging from 0 (unrestricted diffusion as in cerebrospinal fluid) to 1 (directionally restricted diffusion as in well-formed white matter fiber bundles). In DM1, FA has been shown to be significantly reduced throughout the cerebral cortex, with little regional specificity15–14. Statistically significant and widespread FA decrements are seen in samples as small as nine and ten subjects illustrating the large effect size of FA reduction in DM17,11. Several of these studies have shown correlations between FA and muscle impairment as defined by clinical scales such as the...
Health Index (MDHI) is a disease-specific, patient-reported outcome measure for DM1. It is composed of 114 items broken down into 16 individual subscales that collectively measure multi-factorial patient-reported burden of disease. Subjective changes of deterioration, or more importantly, ratings that indicate subjective improvement may be important measures in the context of clinical trials aimed at DM1. It has not been evaluated whether subjective measures of muscle or motor dysfunction also are related to measures of brain pathology.

A major limitation of human brain imaging studies and motor dysfunction studies is that relationships are not representative of true mechanistic relationships – correlations do not prove causality. In evaluating how changes in WM microstructure may drive motor dysfunction, there needs to be careful consideration of other confounding factors such as disease duration and CTG repeat length. In a progressive neurodegenerative disorder such as DM1, muscle symptoms worsen over time. FA has also been shown to be directly correlated with disease duration. Therefore, a correlation between FA and motor dysfunction may be due to temporal coordination of independent disease processes rather than any mechanistic or causal relationship. As with disease duration, the genetic burden of longer CTG repeats may result in spurious correlations between measures of central and peripheral disease symptoms. Both brain FA and muscle dysfunction have shown to be directly related to CTG repeats, suggesting underlying mechanisms links. CTG repeat length and disease duration are related, but independent phenomena. Disease progression may start earlier with longer CTG repeats, while disease manifestation becomes worse over time regardless of at what age it started. Accordingly, no previous report has adjusted for disease duration and CTG repeat length when correlating cross-sectional changes in FA and muscle function.

The present study has three aims: (1) to replicate and extend prior observations of WM degradation in a larger sample of DM1 participants compared to a sample of healthy adults; (2) to replicate and extend prior observations of motor impairment in DM1 by using quantitative measures of motor function as well as subjective measures of perceived motor impairments; and (3) explore the relationships between WM degradation and motor impairments, while controlling for CTG repeat number and DM1 duration.

**Results**

Sample characteristics are summarized in Table 1. Briefly, the sample included 69 unaffected individuals and 50 individuals with a positive genetic diagnosis of DM1. There were significant differences between groups in terms of distribution of sex or mean age. There were 2 individuals with DM1 and 3 unaffected individuals who reported prior diagnosis of Attention Deficit/Hyperactivity disorder (ADHD), but this was unrelated to all of the motor outcomes we measured (all p’s > 0.15) except for potentially self reports of myotonia, however the small number of individuals reporting ADHD precludes testing the possibility that ADHD is a contributing factor. Neuropsychological variables are summarized in Table 2. DM1 and unaffected individuals differed on several measures, where individuals with DM1 had higher depression (t(63.07) = –6.32, CI = –7.38 to –3.84, p = 2.9e–08), anxiety (t(72.47) = –4.62, CI = –7.14 to –2.84, p = 1.6e–05), apathy (t(83.51) = –4.09, CI = –7.44 to –2.57, p = 9.9e–05), and lower full-scale IQ scores (t(105.85) = 4.07, CI = 4.69–13.59, p = 9.1e–05), visuospatial memory (t(102.74) = 1.97, CI = 0.02 to 7.49, p = 0.051), word association (t(107.97) = 2.19, CI = 0.40–8.09, p = 0.031), trail-making (t(84.76) = –4.03, CI = –24.62 to –8.34, p = 0.0012), judgement of line orientation (t(96.20) = 2.42, CI = 0.30–3.05, p = 0.018), and Wisconsin card sorting performance (t(71.91) = 3.07, CI = 0.31–1.46, p = 0.003). As follow-up analyses, we explored whether these cognitive variables were related to motor outcomes and whether these potential relationships were mediated by white matter integrity.

**Muscle/motor outcomes.** We compared muscle/motor outcomes in healthy adults and individuals affected by DM1 using linear mixed effects (LME) models, including family as a random effect and fixed effects of group, age, and sex (p’s corrected for False Discovery Rate, R² represents marginal variance explained or the variance explained by fixed effects only). DM1 is associated with objective, quantitative decline in motor performance as well as subjective ratings of motor impairments. Individuals affected by DM1 were significantly impaired relative to healthy adults on all quantitative muscle/motor outcomes, including peg board (β = 9.08, CI[95%] = 3.69–14.5, t[94.81] = 3.31, p[FW] = 0.00456, R² = 0.0869), finger tapping (β = –9.32, CI[95%] = 12.8 to –6.28, t[110.21] = –5.71, P[FW] = 3.35 × 10⁻⁷, R² = 0.222), grip strength (β = –10.5, CI[95%] = 14.5 to –6.61, t[107.33] = –5.25, P[FW] = 1.82 × 10⁻⁶, R² = 0.346). DM1-affected individuals, likewise, reported significant burden of myotonia (β = 18.8, CI[95%] = 12.4–
We conducted a set of follow-up mediation analyses to explore how well each of these causal pathways contributed to motor outcomes or may be mediated by WM microstructure; the relationships between pathways. For instance, genetic burden (see Fig. 6D) and disease duration (see Fig. 6E) may make independent contributions to new data (see Supplementary Data for all cross-validation results). Critically, neither disease duration nor genetic burden were significant predictors of muscle/motor function independently of cerebral WM FA (all $p > 0.05$). This is consistent with the notion that WM FA is a proximal mediator of muscle and motor deficits in DM1.

**WM microstructure.** Individuals affected by DM1 had significantly degraded WM microstructure, revealed by LME models predicting WM measures with the random effect of family and fixed effects of group, age, and sex. WM microstructure was impacted by DM1 in a systemic rather than regional fashion (see Figs. 1, 2 and Tables 3, 4, 5 for full results); most regions exhibited a decrease in fractional anisotropy (e.g., Cerebral WM FA: $\beta = -0.0339, CI_{95\%} = -0.0431$ to $-0.0285, t_{(106.64)} = -4.7, p_{FDR} = 4.7 \times 10^{-5}, R^2 = 0.468$), axial diffusivity (e.g., Cerebral WM AD: $\beta = 5.56 \times 10^{-5}, CI_{95\%} = 3.49 \times 10^{-5}$ to $7.62 \times 10^{-5}, t_{(102.16)} = 7.84, p_{FDR} = 2.3 \times 10^{-10}, R^2 = 0.202$), and radial diffusivity (e.g., Cerebral WM RD: $\beta = 6.19 \times 10^{-5}, CI_{95\%} = 4.63 \times 10^{-5}$ to $7.73 \times 10^{-5}, t_{(108.27)} = 8.84, p_{FDR} = 1.38e^{-10}, R^2 = 0.178$).

**Relationships between WM microstructure and motor outcomes in DM1.** WM integrity predicted several motor outcomes in LME models after controlling for fixed effects of disease duration, genetic burden, age, and sex. Results are consistent with the notion that cerebral WM FA mediated between disease duration, genetic burden, and grip strength. It is possible that other ‘causal’ pathways remain (see Fig. 6A for a diagram of all causal pathways). For instance, genetic burden (see Fig. 6D) and disease duration (see Fig. 6E) may make independent contributions to motor outcomes or may be mediated by WM microstructure; the relationships between genetic burden and WM microstructure (see Fig. 6B) and motor outcomes (see Fig. 6C) may be mediated by disease duration given that earlier onsets are associated with longer CTG repeat lengths; and/or the direction of causality could potentially be inverted, where changes in motor abilities result in changes in cerebral WM (see Fig. 6F). We conducted a set of follow-up mediation analyses to explore how well each of these causal pathways match our data. According to the criteria for mediation\textsuperscript{17}, we consider mediation pathways significant if: (1)
there is a relationship between the predictor (e.g., genetic burden) and the outcome (e.g., grip strength), (2) there is a relationship between the mediator (e.g., cerebral WM FA) and the predictor, and (3) there is a relationship between the mediator and the outcome while controlling for the predictor. Furthermore, if (4) the predictor does not predict the outcome after considering the mediator a fully mediated “causal” pathway is indicated.

First, mediation analyses suggest that ePAL is not the proximal cause of decreases in motor performance in DM1, as cerebral WM FA mediated the effect between ePAL and grip strength ($\text{Indirect Effect (} \delta \text{)} = -6.59$, $\text{CI}_{95\%} = -16.12$ to $0.09$, $p = 0.06$; $\text{Direct Effect (} \zeta \text{)} = -3.09$, $\text{CI}_{95\%} = -14.33$ to $7.73$, $p = 0.62$) but disease duration did not mediate this effect ($\delta = -3.61$, $\text{CI}_{95\%} = -9.33$ to $1.59$, $p = 0.22$; $\zeta = -5.23$, $\text{CI}_{95\%} = -16.88$ to $7.98$, $p = 0.40$). Second, disease duration mediated between ePAL and cerebral WM FA ($\delta = -0.015$, $\text{CI}_{95\%} = -0.027$ to $0$, $p \sim 0$; $\zeta = -0.01$, $\text{CI}_{95\%} = -0.03$ to $0.01$, $p = 0.52$). Third, cerebral WM FA mediated between disease duration and grip strength ($\delta = -0.646$, $\text{CI}_{95\%} = -1.36$ to $-0.21$, $p = 0$; $\zeta = 0.24$, $\text{CI}_{95\%} = -0.442$ to $0.87$, $p = 0.60$). Together these results suggest that the underlying genetic cause of the disease (CTG repeat expansion) results in progressive changes in cerebral WM microstructure that in turn result in decreased motor performance (i.e., reduced grip strength).

![Figure 1. Group differences in WM microstructure measures.](https://www.nature.com/scientificreports/) Fractional anisotropy values (left panel) were lower in DM1 relative to healthy adults on most regions evaluated (colored estimates), indicating reduced WM integrity. Axial and radial diffusivity (middle and right panels, respectively) were higher in DM1 relative to healthy adults, indicating decreased directional restriction of water molecules in DM1-affected WM. Red indicates volumes of interest (VOIs) where FDR-corrected $p < 0.05$, blue indicates FDR-corrected $p < 0.1$, gray indicates uncorrected $p < 0.05$, and white indicates uncorrected $p > 0.05$. Error bars represent the 99.99% confidence limits, which approximates the FDR-corrected $p = 0.0081$. Data were plotted using R (version 3.5.0, https://www.r-project.org/).
There is some evidence of regional contributions of WM FA to motor performance. Peg board performance was related to WM FA in the cerebral peduncles ($\beta = -263$, CI$_{95\%} = -416$ to $-111$, $t_{(19.32)} = -3.39$, $p = 0.00304$, $pFDR = 0.0551$, $R^2 = 0.141$); finger tapping was related to WM FA in the anterior limb of the internal capsule ($\beta = 157$, CI$_{95\%} = 60.9$–254, $t_{(39.00)} = 3.2$, $p = 0.00274$, $pFDR = 0.074$, $R^2 = 0.191$), and grip strength was related to WM FA in the anterior corona radiata ($\beta = 170$, CI$_{95\%} = 69.6$–270, $t_{(39.00)} = 3.32$, $p = 0.00197$, $pFDR = 0.0531$, $R^2 = 0.203$) and hippocampal portion of the cingulum ($\beta = 214$, CI$_{95\%} = 85.4$–343, $t_{(39.00)} = 3.26$, $p = 0.00232$, $pFDR = 0.0574$, $R^2 = 0.197$) (see Fig. 5A). Cross-validation demonstrates these models have low bias ($<|0.074|$ SD), error measures are less than 1 SD, and the ratio of error measures between training and test sets varies from 1.272 to 2.216. This suggests that regional measures are less reliable than more global measures of cerebral WM FA, which may in part be due to more individual variation in smaller brain regions compared to an individual average across many brain regions. WM FA did not tend to be a predictor of subjective ratings of motor impairment (all uncorrected $p’s > 0.05$).

Axial diffusivity did not tend to be a significant predictor of motor function, except for a trend where WM AD in the superior cerebellar peduncle predicted peg board performance ($\beta = 4.55 \times 10^4$, CI$_{95\%} = 1.81 \times 10^4$–7.3 $\times 10^4$, $t_{(31.96)} = 3.25$, $p = 0.00271$, $pFDR = 0.0733$, $R^2 = 0.163$), see Supplementary Data for full results. Radial diffusivity in the external capsule significantly predicted peg board performance ($\beta = 3.95 \times 10^5$, CI$_{95\%} = 2.07 \times 10^5$–5.84 $\times 10^5$).

Figure 2. Voxelwise Group Differences in WM microstructure in DM1. WM microstructure in DM1 relative to healthy adults is characterized by decreased fractional anisotropy (panel A), and increased axial (panel B) and radial diffusivity (panel C). This decreased WM integrity in DM1 appears to be systemic rather than regionally specific, though the size of effects varies. Cold colors indicate beta estimates for the group comparison in LME models thresholded at $p_{uncorrected} < 0.05$; hot colors indicate estimates thresholded at $p_{uncorrected} < 0.01$. Neuroimages were created using ITK-SNAP (version 3.8.0, http://www.itksnap.org/)26. Inkscape (version 0.1, https://inkscape.org/) was used to add labels and color bars to the figure.
Effects of disease duration and genetic burden on motor outcomes. While WM FA, AD and RD values did not tend to be significant predictors of subjective ratings of motor impairments, disease duration tended to be a non-significant predictor of subjective measures of motor impairments in mobility, myotonia, and swallowing, but not upper extremity function (all uncorrected p's > 0.05). All non-significant effects were in the same direction, where longer disease durations were associated with higher subjective ratings of motor impairment. By contrast, genetic burden, while controlling for disease duration and WM microstructure did not significantly predict motor impairments (all uncorrected p's > 0.05).

Neuropsychological and cognitive contributors to motor outcomes. Intellectual ability and neuropsychological impairments can potentially impact performance on motor tasks, e.g., grip strength and cognitive declines both accompany increasing age and are potentially but not necessarily causally related14. For example, deficits in motor performance may be manifestations of lack of attention to the task or lack of motivation to complete the task to the best of one's ability. Likewise, mood disturbances may impact one's ability to successfully engage with tasks, thus hindering performance. As a follow-up analysis, we explored whether neuropsychological measures predicted motor performance in participants with DM1. We limited this follow-up analysis to grip strength, finger-tapping, and peg board performance as these measures exhibited significant (or trending) relationships to cerebral WM FA. Furthermore, if the relationships between neuropsychological measures and

| β | t(df) | Puncorrected | Pcorr | CI99.9% | R² |
|---|-------|--------------|------|---------|----|
| Cerebral WM | −0.0359 | −9.69 (105.69) | 2.89 × 10⁻⁶ | 3.9 × 10⁻⁵ | −0.0431 to −0.0285 | 0.468 |
| Cerebellar WM | −0.0279 | −6.43 (88.30) | 6.32 × 10⁻⁹ | 8.53 × 10⁻⁸ | −0.0363 to −0.0192 | 0.272 |
| Corpus Callosum—genu | −0.0695 | −5.54 (93.68) | 2.77 × 10⁻⁷ | 3.75 × 10⁻⁶ | −0.0938 to −0.0451 | 0.216 |
| Corpus callosum—body | −0.0584 | −6.75 (90.36) | 1.35 × 10⁻⁸ | 1.83 × 10⁻⁷ | −0.0753 to −0.0411 | 0.292 |
| Corpus callosum—splenium | −0.0575 | −6.34 (99.79) | 6.88 × 10⁻⁷ | 9.29 × 10⁻⁶ | −0.0752 to −0.0399 | 0.269 |
| Cerebral peduncle | −0.0203 | −2.36 (109.00) | 0.0203 | 0.183 | −0.037 to −0.00357 | 0.067 |
| Corticospinal tract | −0.0161 | −1.92 (81.84) | 0.0583 | 0.524 | −0.0326 to 0.000232 | 0.145 |
| Medial lemniscus | −0.0399 | −3.68 (90.87) | 0.00391 | 0.00528 | −0.0618 to −0.0181 | 0.112 |
| Pontine crossing tract | −0.0323 | −3.46 (108.97) | 0.00767 | 0.0104 | −0.0511 to −0.0121 | 0.099 |
| Middle cerebellar peduncle | −0.0332 | −6.45 (109.00) | 3.2 × 10⁻⁴ | 4.32 × 10⁻³ | −0.0432 to −0.0232 | 0.271 |
| Inferior cerebellar peduncle | −0.0245 | −3.33 (105.86) | 0.0012 | 0.0162 | −0.0389 to −0.0102 | 0.094 |
| Superior cerebellar peduncle | −0.0259 | −2.74 (109.00) | 0.00725 | 0.0653 | −0.0456 to −0.00751 | 0.117 |
| Anterior limb inner capsule | −0.0245 | −4.37 (109.00) | 3.68 × 10⁻⁸ | 0.00331 | −0.0356 to −0.0134 | 0.194 |
| Posterior limb inner capsule | −0.0164 | −2.77 (108.50) | 0.00659 | 0.0593 | −0.0279 to −0.00482 | 0.092 |
| Retrolenticular inner capsule | −0.0269 | −5.22 (104.45) | 9.03 × 10⁻⁵ | 1.22 × 10⁻⁴ | −0.037 to −0.0168 | 0.203 |
| Anterior corona radiata | −0.0502 | −8.83 (107.34) | 2.23 × 10⁻¹⁵ | 3.01 × 10⁻¹⁴ | −0.0612 to −0.039 | 0.421 |
| Superior corona radiata | −0.0174 | −3.35 (108.34) | 0.00115 | 0.0134 | −0.0279 to −0.00674 | 0.094 |
| Posterior corona radiata | −0.0388 | −5.91 (105.32) | 4.35 × 10⁻⁴ | 5.84 × 10⁻³ | −0.0521 to −0.0255 | 0.246 |
| Posterior thalamic radiation | −0.0497 | −7.5 (95.66) | 3.23 × 10⁻¹⁰ | 4.35 × 10⁻⁹ | −0.0626 to −0.0367 | 0.341 |
| Sagittal stratum | −0.0426 | −6.26 (106.23) | 8.32 × 10⁻⁴ | 1.12 × 10⁻³ | −0.056 to −0.0289 | 0.268 |
| External capsule | −0.0221 | −4.88 (109.00) | 3.68 × 10⁻⁸ | 4.96 × 10⁻⁷ | −0.0309 to −0.0133 | 0.176 |
| Cingulum—cingulate | −0.0635 | −6.13 (109.00) | 1.43 × 10⁻⁸ | 1.93 × 10⁻⁷ | −0.0837 to −0.0434 | 0.252 |
| Cingulum—hippocampus | −0.0233 | −3.92 (109.00) | 0.00157 | 0.00212 | −0.0348 to −0.0117 | 0.121 |
| Superior longitudinal fasc | −0.0349 | −6.91 (104.66) | 3.99 × 10⁻¹⁰ | 5.38 × 10⁻⁹ | −0.045 to −0.0245 | 0.309 |
| Superior frontooccipital fasc | −0.0133 | −1.9 (102.07) | 0.0597 | 0.537 | −0.0269 to 0.000401 | 0.051 |
| Uncinate fasc | −0.0399 | −3.91 (96.05) | 0.000169 | 0.00228 | −0.0607 to −0.0186 | 0.125 |
| Tapetum | −0.0521 | −5.47 (108.97) | 2.89 × 10⁻⁸ | 3.91 × 10⁻⁷ | −0.0708 to −0.0356 | 0.216 |

Table 3. Group differences in fractional anisotropy.
motor outcomes were significant, we conducted a causal mediation analysis to explore whether cerebral WM FA mediated these relationships.

Grip strength was significantly related to full-scale IQ ($\beta = 0.35$, CI95% = −0.03 to 0.72, $t(36.00) = 2.12$, $p = 0.041$) and this effect was indeed mediated by cerebral WM FA ($\delta = 0.41$, CI95% = 0.08−0.78, $p = 0.02$, $\zeta = 0.02$, CI = −0.05 to 0.21, $p = 0.09$). These results are consistent with the notion that cerebral WM FA fully mediates the relationship between full-scale IQ and grip strength, according to the criteria for mediation\cite{7}. All other neuropsychological measures were not significant predictors of grip strength.

Finger tapping was significantly predicted by full-scale IQ ($\beta = 0.33$, CI95% = 0.04−0.66, $t(34.33) = 2.51$, $p = 0.017$), COWA scores ($\beta = 0.34$, CI95% = 0.02−0.66, $t(32.96) = 2.44$, $p = 0.02$), and Trails B performance ($\beta = 0.15$, CI95% = −0.29 to 0.01, $t(35.74) = −2.66$, $p = 0.011$). Moreover, these relationships were fully mediated by cerebral WM FA: full-scale IQ ($\delta = 0.24$, CI95% = 0.06−0.48, $p = 0$, $\zeta = 0.17$, CI = −0.08 to 0.47, $p = 0.32$), COWA ($\delta = 0.09$, CI95% = −0.03 to 0.21, $p = 0.08$, $\zeta = 0.24$, CI95% = −0.03 to 0.49, $p = 0.1$), and Trails B ($\delta = 0$−0.09, CI95% = −0.17 to 0, $p = 0.02$, $\zeta = −0.07$, CI = −0.21 to 0.03, $p = 0.24$).

Peg board performance was significantly predicted by self-reported apathy ($\beta = 1.25$, CI95% = 0.10−2.44, $t(36.00) = 2.62$, $p = 0.013$), full-scale IQ ($\beta = 0.95$, CI95% = −1.56 to −0.37, $t(33.43) = −3.69, p = 0.0076$), Trails B performance ($\beta = 0.39$, CI95% = 0.05−0.71, $t(33.33) = 3.38$, $p = 0.0018$), judgement of line orientation performance ($\beta = −1.61$, CI95% = −3.56 to 0.27, $t(33.00) = 2.04$, $p = 0.049$), Bender-Gestalt scores ($\beta = −1.03$, CI95% = −1.87 to −0.20, $t(36.07) = −3.40$, $p = 0.0019$), categories completed on the Wisconsin Card Sorting Task ($\beta = −4.78$, CI95% = 11.19 to 1.17, $t(33.93) = −2.40$, $p = 0.022$), and score on Benton Faces ($\beta = 2.55$, CI95% = 4.61 to 0.61, $t(36.00) = 3.45$, $p = 0.0014$). However, in contrast to grip strength and finger tapping, cerebral WM FA did not mediate any of these effects, which is consistent with our observation of a weaker relationship between peg board performance and FA in general (all $p$'s for indirect effects > 0.12).

**Discussion**

The current study replicates several other studies documenting the severity of cerebral white matter microstructural change in DM1. The study expands previous findings by showing white matter pathology was directly predictive of motor abnormalities such as grip strength, finger tapping, and peg board performance, even after controlling for factors such as disease duration and genetic burden (length of CTG repeat or EPAL). While there are primary genetic defects driving muscle pathology in DM1, our results support the notion that there may also be a role for brain pathology driving motor abnormalities.
FA and the MIRS13 assume that the numerical distance between adjacent categories is equal, which may or may not be accurate, thus results must be considered approximations. Our results support and extend the findings of this previous work, by recruiting a large sample of DM1 participants and using an enhanced statistical model accounting for the indirect path where these effects were mediated by WM FA), supporting the notion that this burden and grip strength (the direct path from genetic burden and disease duration was no longer significant after follow-up mediation analysis confirms that in our sample cerebral WM FA completely mediates between the ultimate causes of the disease and motor symptoms. Indeed, our follow-up mediation analysis confirms that in our sample cerebral WM FA completely mediates between disease burden and grip strength (the direct path from genetic burden and disease duration was no longer significant after accounting for the indirect path where these effects were mediated by WM FA), supporting the notion that this relationship is potentially causal with white matter damage leading to motor dysfunction. Our data and mediation analyses are not consistent with the notion that there is a strong retrograde effect whereby muscle changes lead to alterations in cerebral WM microstructure. Further research should target this relationship in a longitudinal sample where changes in WM FA can be explored in the context of DM1 progression. This relationship between cerebral WM FA and grip strength could prove to be crucial for translational research where disease progression in the context of treatment regimens must be measured.

Our data are consistent with the notion that WM degradation in DM1 is systemic rather than regionally specific. Additional support for this notion is that WM degradation mediates the relationships between cerebral WM FA and grip strength could prove to be crucial for translational research where disease progression in the context of treatment regimens must be measured.

### Table 5. Group differences in radial diffusivity.

| Region                        | β         | γ(df)     | Pcorrected  | pFDR  | CI99.99%  | R²       |
|-------------------------------|-----------|-----------|-------------|-------|----------|----------|
| Cerebral WM                   | 6.19 x 10⁻⁵ | 7.84 (104.27) | 3.4 x 10⁻¹² | 4.6 x 10⁻¹¹ | 4.63 x 10⁻⁵ - 7.73 x 10⁻⁵ | 0.355    |
| Cerebellar WM                 | 3.57 x 10⁻⁵ | 4.55 (106.18)  | 1.41 x 10⁻⁵  | 0.00019  | 2.05 x 10⁻⁴ - 5.1 x 10⁻⁴  | 0.163    |
| Corpus callosum—genu         | 0.000103   | 5.53 (103.04)  | 2.47 x 10⁻⁷  | 3.33 x 10⁻⁷ | 6.66 x 10⁻⁵ - 0.000139  | 0.222    |
| Corpus callosum—body         | 7.74 x 10⁻⁵ | 6.59 (108.79)  | 1.65 x 10⁻⁵  | 2.23 x 10⁻⁵ | 5.39 x 10⁻⁵ - 0.00001  | 0.283    |
| Corpus callosum—splenium     | 8.51 x 10⁻⁵ | 6.39 (108.46)  | 4.2 x 10⁻⁴   | 5.67 x 10⁻⁴ | 5.92 x 10⁻⁵ - 0.000111  | 0.275    |
| Cerebral peduncle             | 3.46 x 10⁻⁵ | 1.97 (99.84)   | 0.0511       | 0.453   | -1.49 x 10⁻⁵ to 7.05 x 10⁻⁵ | 0.0426   |
| Corticospinal TRACT           | 3.71 x 10⁻⁵ | 2.39 (92.83)   | 0.0187       | 0.168   | 6.96 x 10⁻⁶ - 6.73 x 10⁻⁴  | 0.0934   |
| Medial lemniscus              | 0.000101   | 3.67 (102.18)  | 0.00393      | 0.055   | 4.67 x 10⁻⁵ - 0.000155  | 0.104    |
| Pontine crossing tract        | 6.64 x 10⁻⁵ | 5.07 (106.06)  | 1.68 x 10⁻⁴  | 2.27 x 10⁻⁴ | 4.09 x 10⁻⁵ - 9.18 x 10⁻⁵ | 0.194    |
| Middle cerebellar peduncle    | 4.71 x 10⁻⁵ | 4.62 (109.00)  | 1.06 x 10⁻⁵  | 0.000134 | 2.73 x 10⁻⁵ - 6.69 x 10⁻⁵ | 0.161    |
| Inferior cerebellar peduncle  | 5.16 x 10⁻⁵ | 3.29 (108.31)  | 0.00136      | 0.0183  | 2.09 x 10⁻⁵ - 8.26 x 10⁻⁵ | 0.0913   |
| Superior cerebellar peduncle  | 7.91 x 10⁻⁵ | 2.88 (95.91)   | 0.0049       | 0.0662  | 2.49 x 10⁻⁵ - 0.000135  | 0.0719   |
| Anterior limb internal capsule| 3 x 10⁻⁵   | 4.93 (889.80)  | 9.6 x 10⁻⁶   | 1.3 x 10⁻⁵ | 1.82 x 10⁻⁵ - 4.19 x 10⁻⁵ | 0.173    |
| Posterior limb internal capsule| 2.38 x 10⁻⁵ | 3.45 (108.79)  | 0.000802     | 0.0108  | 1.04 x 10⁻⁵ - 7.2 x 10⁻⁵  | 0.0992   |
| Retrolenticular internal capsule| 4.6 x 10⁻⁵ | 6.08 (108.99)  | 1.8 x 10⁻⁴   | 2.42 x 10⁻⁴ | 3.1 x 10⁻⁵ - 6.07 x 10⁻⁵ | 0.253    |
| Anterior corona radiata       | 7.63 x 10⁻⁵ | 7.64 (108.37)  | 9.17 x 10⁻¹⁵ | 1.24 x 10⁻¹⁵ | 5.68 x 10⁻⁵ - 9.57 x 10⁻⁵ | 0.352    |
| Superior corona radiata       | 3.82 x 10⁻⁵ | 4.91 (107.09)  | 3.26 x 10⁻⁶  | 4.4 x 10⁻⁵ | 2.29 x 10⁻⁵ - 5.34 x 10⁻⁵ | 0.184    |
| Posterior corona radiata      | 6.67 x 10⁻⁶ | 6.34 (108.96)  | 5.27 x 10⁻⁴  | 7.11 x 10⁻⁴ | 4.58 x 10⁻⁵ - 8.74 x 10⁻⁵ | 0.27     |
| Posterior thalamic radiation  | 8.55 x 10⁻⁵ | 7.58 (108.11)  | 2.16 x 10⁻¹⁰ | 1.7 x 10⁻¹⁰ | 6.33 x 10⁻⁵ - 0.000108  | 0.348    |
| Sagittal stratum              | 7.63 x 10⁻⁵ | 6.9 (108.64)   | 3.68 x 10⁻⁵  | 4.97 x 10⁻⁵ | 5.35 x 10⁻⁵ - 9.85 x 10⁻⁵ | 0.301    |
| External capsule              | 4.71 x 10⁻⁵ | 7.93 (108.74)  | 2.09 x 10⁻¹² | 2.83 x 10⁻¹¹ | 3.54 x 10⁻⁵ - 5.88 x 10⁻⁵ | 0.368    |
| Cingulum—cingulate            | 6.83 x 10⁻⁵ | 7.2 (106.41)   | 9.12 x 10⁻¹⁴ | 1.23 x 10⁻¹⁴ | 4.91 x 10⁻⁵ - 8.72 x 10⁻⁵ | 0.326    |
| Cingulum—hippocampus          | 4.84 x 10⁻⁵ | 5.13 (105.46)  | 1.33 x 10⁻⁶  | 1.79 x 10⁻⁶ | 2.98 x 10⁻⁵ - 6.68 x 10⁻⁵ | 0.198    |
| Superior longitudinal fasc    | 5.73 x 10⁻⁵ | 7.15 (108.61)  | 1.07 x 10⁻⁷  | 1.44 x 10⁻⁷ | 4.13 x 10⁻⁵ - 7.31 x 10⁻⁵ | 0.316    |
| Superior frontooccipital fasc | 2.8 x 10⁻⁴   | 3.41 (108.63)  | 0.000923     | 0.0125  | 1.2 x 10⁻⁴ - 4.39 x 10⁻⁵ | 0.098    |
| Uncinate fasc                 | 4.97 x 10⁻⁴ | 4.4 (102.63)   | 2.66 x 10⁻⁵  | 0.000359 | 2.65 x 10⁻⁵ - 7.23 x 10⁻⁵ | 0.153    |
| Tapetum                       | 0.000139   | 3.47 (94.04)   | 0.000795     | 0.0107  | 6.07 x 10⁻⁵ - 0.000218  | 0.101    |
neuropsychological measures and motor outcomes. Specifically, our results are consistent with the notion that systemic WM decay results in cognitive impairments and contributes to motor decline. Importantly, WM degradation cannot be the only mechanism that is causing cognitive and muscle/motor decline. For example, our result whereby performance on the peg board task was not mediated by WM integrity suggests that there are likely other non-WM neurological factors impacted by DM1 that result in both cognitive and motor impairments. Despite most regions exhibiting some degree of degradation, the amount of degradation does vary regionally. Moreover, there appears to be at least some degree of localization of the relationship between WM FA decline and motor outcomes. For example, FA in the WM near the motor cortex predicts grip strength most strongly, peg board performance tended to be associated with WM measures that inputs and outputs for the cerebellum, and finger tapping with the anterior limb of the internal capsule near the basal ganglia. While these fluctuations in WM integrity likely relate to severity and variety of symptoms in a regionally specific way, aggregating FA values across the whole brain may provide a stronger, more consistent biomarker for translational research.

Despite the relationship between FA and muscle/motor impairments in DM1, the nature of deficits in WM structural integrity remains unclear and could be a fruitful topic for future research. Our results, specifically our observation that AD did not have any significant relationship with motor outcomes while RD did (albeit weaker than FA), may provide a hint of the underlying WM issues. These results are more consistent with the notion...
that while axons may not be damaged per se (no change in AD) there may be dys- or demyelination (increased RD)\textsuperscript{22,23}, for a review see: \textsuperscript{24}. Further research that includes histological examination of WM microstructure in DM1 could address the source of these deficits. This work may be limited by the sample size, where a larger sample may elucidate more specific changes in WM measures of AD and RD, which may point to specific WM pathologies.

We did not observe a relationship between patient-reported, subjective motor symptoms as measured by the MDHI and WM microstructure. These subjective reports may be confounded by patient apathy and/or insight. However, disease duration tended to be weakly related to self-reported myotonia, swallowing, and mobility. We speculate that WM degradation may go unnoticed in terms of symptomatology until some critical threshold is met; particularly considering our sample of patients with DM1 who have relatively mild symptoms. Further research, in a longitudinal setting would allow us to elucidate the time course of WM degradation and the patient experience of symptoms and establish the natural history of FA abnormalities and muscle pathology.

This work is potentially limited by sample size and its cross-sectional nature, as discussed above. Increased sample size and repeated measures over time would allow us to explore in more detail the underlying WM pathology (e.g., larger sample would give more power to AD and RD results) and a longitudinal approach would provide further evidence for mediation if motor and WM degradation occur in concert. Furthermore, this work is limited due to the nature of MRI studies where more scanning time could yield higher resolution images which could

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**Figure 4.** Fractional anisotropy predicting motor outcomes. The figure lists standardized betas listed on the x-axis for the association between regional FA (y-axis) and motor outcomes (top facets). Significant associations are marked by color, where red indicates VOIs where FDR-corrected $p < 0.05$, blue indicates FDR-corrected $p < 0.1$, gray indicates uncorrected $p < 0.05$, and white indicates uncorrected $p > 0.05$. Error bars represent the 95%. Data were plotted using R (version 3.5.0, \url{https://www.r-project.org/})\textsuperscript{25}.
yield better insight into specific WM pathology as well as possible spatial specificity within the brain. Finally, there may be other factors that contribute and/or mediate these effects that were not measured as a part of our study; with further research these unknown factors may come to light.

Given DM1 is a single gene disorder, there is a frenetic push towards development of gene knock-down and other therapies designed to treat or slow down disease progression. One important caveat is that although some therapies have been developed for delivery to muscle, our results are consistent with the notion that complimentary treatments delivered to the brain could also benefit motor function. Our results suggest that cerebral WM integrity may be a critical tool in evaluating these therapies.

Methods
Participants. The Iowa Brain DM1 study focuses on recruitment of individuals with adult-onset DM1 (those who exhibited disease-related symptoms after the age of 21 years old) as well as individuals who are at-risk for DM1 (those with a family history of DM1), but have not yet been genetically tested. Participants were recruited from our own multidisciplinary specialty clinic for DM1 at the University of Iowa and through the Myotonic Dystrophy Foundation. A control group consisting of healthy adults was recruited from partners of DM1 participants, and from the Iowa City area via advertisements. Exclusion criteria for all participants included: MRI contraindication, a history of serious head injury, or a chronic neurological disorder other than DM1. Healthy adults were additionally required to be without a history of substance abuse, psychiatric disease, or major medical disease, including: heart disease, sleep disorder, vascular disease, uncontrolled hypertension, cancer, diabetes mellitus, lung disease, and autoimmune conditions. The current sample included 119 individuals: 61 healthy adults, 45 individuals with confirmed DM1 and 13 individuals with a family history of DM1 who had not undergone confirmative testing. Participants underwent genetic testing for research purposes only. At-risk individuals who were determined to have CTG repeat length ≥ 50 were included in the DM1 group (N = 5); the remainder had CTG repeat length in the non-expanded range and were included in the group of healthy adults (N = 8). The final sample included 50 individuals with DM1 and 69 healthy adults. For the patients with DM1, disease duration was determined by the time at which they received a clinical diagnosis of DM1. This ranged from 0 (those who were at-risk, found to have the gene-expansion but no clinical symptoms) to 28.9 years with a mean of 8.88 years.
Research staff, clinicians, and scientists involved in this study remained blind to the genetic status of at-risk individuals. All data were de-identified, and all participants consented to non-disclosure of genetic results obtained as part of the study. All participants gave written, informed consent prior to enrolling in the protocol in accordance with the Declaration of Helsinki. The study was approved by the University of Iowa Institutional Review Board. All methods were performed in accordance with these guidelines and regulations.

**Estimated progenitor allele length (ePAL).** Genotyping of CTG repeat in DM1-affected participants, and at-risk individuals was completed by SP-PCR. For each patient, four reactions were completed, each using a 300 pg genomic DNA template derived from blood leukocytes. CTG repeat lengths were estimated by comparison against DNA fragments of known length and molecular weight markers, using CLIQS software (TotalLabs UK Ltd.). The lower boundary of the expanded molecules in SP-PCR was used to estimate the progenitor (inherited) allele length (ePAL). ePAL is a major determinant of age at symptom onset.
Estimation of CTG repeat length of the non-disease-causing allele(s) at-risk individuals and controls was determined by Illumina MiSeq sequencing, essentially as described for Huntington disease\textsuperscript{25}. Barcoded primers that contained all the sequences required for MiSeq sequencing, combined with gene-specific sequences flanking the CTG repeats, were used to generate the amplicon sequencing library. Open-source bioinformatic tools on the Galaxy platform\textsuperscript{31} were used to process sequence reads and align them against custom reference sequences comprising unique DM1-specific flanking sequences, separated by 0 to 100 CTG repeats. The reference sequence(s) with the highest number of aligned reads corresponded to the number of CTG repeats in the non-disease-causing allele(s).

**Motor/muscle function.** Fine motor skills were measured with the Lafayette Instruments finger tapping test and the grooved Pegboard. The finger tapping apparatus consists of a tapping key with a device for recording the number of taps. Participants completed five consecutive tap trials that were 1 s each. The dependent variable included the average number of taps across five trials using the dominant hand. The Grooved Pegboard test requires participants to insert keyed pegs into slots. The outcome measure of interest was time to completion in seconds using the dominant hand.

The Lafayette Instruments dynamometer was used to assess grip force in kilogram-force (kgf). While standing, participants squeezed the instrument as hard as they could six times (three trials using the dominant hand, three trials using the non-dominant hand). The dependent variable represents the average grip force of the dominant hand across three trials.

**Patient-reported motor/muscle function.** The Myotonic Dystrophy Health Index (MDHI) is a disease-specific patient-reported outcome measure for myotonic dystrophy type-1\textsuperscript{19,32}. It is composed of 114 items broken down within 16 individual subscales that together measure multi-factorial patient-reported burden of disease. Each item is rated on a 6-point Likert scale as to how much the item “impacts the participant’s life now.” The range of response options are “I don’t experience this” to “It affects my life severely.” Given the current emphasis on motor and muscle related outcomes, we utilized scores on only the relevant subscales, including: myotonia, mobility, upper extremity function, and swallowing.

**Magnetic resonance imaging.** Participants who participated before June 2016 (N = 49, 24 unaffected, 25 with DM1) were scanned using a 3 T Siemens TrioTIM scanner (Siemens AG, Munich, Germany; 12 channel head coil, software version: syngo B17). Those who participated after June 2016 were scanned using a 3 T General Electric Discovery MR750w scanner (GE Medical Systems, Chicago, IL; 16 channel head and neck coil, software versions: 25.0, 25.1, and 26.0) (N = 65, 43 unaffected, and 22 with DM1). Participants completed DWI acquisitions with either a single-shell (B1000, 64 directions), multi-shell (B1000 and B2000, 29–30 directions per shell), or both (details provided in ***eTables 1 and 1 in the Supplement). Diffusion-weighted images were collected using echo planar recovery magnitude sequences collected in the axial plane. Anatomical T1-weighted and T2-weighted images were collected and used for co-registration, normalization, and labelling purposes using acquisition parameters described previously\textsuperscript{33}.

**White matter FA.** Diffusion-weighted images were processed using standard procedures of the FMRIB Diffusion toolbox from the FSL software package (http://www.fmrib.ox.ac.uk/fsl), where phase encoding distortion and eddy current artifacts were removed using topup and eddy tools respectively\textsuperscript{34,35}. Following correction, diffusion tensor models were generated using dtifit, and from these tensors, scalar measures, including fractional anisotropy (FA), axial diffusivity (AD) and radial diffusivity (RD), were calculated. B0 maps were co-registered to T2-weighted images for each subject, which were in turn registered to their T1-weighted images, which were normalized to a standard space. All registrations consisted of rigid, affine, and nonlinear (symmetric normalization) components and were conducted using Advanced Normalization Tools\textsuperscript{36}. All steps in the registration sequence were combined into a single transform as necessary and applied simultaneously to scalar maps (FA, AD, RD) to avoid compounding interpolation errors. Data were normalized to an unbiased average of the brains from the Human Connectome Project\textsuperscript{37}, which itself was normalized to ICBM 2009b Nonlinear Asymmetric space\textsuperscript{38}.

Regions of interest were extracted using the BRAINSAutoWorkup pipeline which optimizes tissue classification through an iterative framework and produces robust parcellation of brain regions results in a multi-scanner setting\textsuperscript{39}. BRAINSAutoWorkup labels brain regions using a multi-atlas, similarity-weighted, majority-vote procedure (joint label fusion)\textsuperscript{30} using a set of expert-segmented templates adapted from the Desikan-Killiany atlas\textsuperscript{40}. White matter labels corresponding to cerebral and cerebellar regions were combined to generate regions of interest for each participant’s brain in its native space. For voxel-wise modelling and the JHU WM atlas\textsuperscript{42–44}, spatially normalized scalar values were used.

**Statistical analyses.** Statistics and figures (Figs. 1, 3, 4; not including neuroimaging) were generated using R 3.5.0\textsuperscript{25}. Neuroimaging figures (Figs. 2, 5) were created using ITKSnap\textsuperscript{39} for brain overlays and Inkscape (version 0.1, https://inkscape.org/) for panel layout and generation of colour bars.

Demographic characteristics of the DM1 group and the unaffected group were summarized and compared by chi-squared test for proportions, 2-sample t-test for means, or Wilcoxon’s rank-sum tests for medians.

First, batch effects associated with scanner vendor and software version were harmonized using ComBat harmonization\textsuperscript{45,46} implemented in R. Group differences in functional outcomes and WM scalar values were examined using a linear mixed effects (LME) framework, where the random effect of family (to control for the effects of familial relationships among some individuals in our sample) and fixed effects of group, age, and sex
were included in each model. Given the number of tests, models including functional outcomes and JHU regions were FDR-corrected using the Benjamini & Hochberg procedure. LME models exploring WM values were also applied voxelwise to explore potential regionally specific WM degradation using tools implemented in R.

Second, the LME framework was used to assess whether WM microstructure, disease duration, and/or genetic burden predicted functional outcomes in DM1 participants only. Family identifiers were included as random effects and WM measures, disease duration (years from diagnosis), genetic burden (natural log transformed ePAL), age, and gender were included as fixed factors. Critically, measures of WM microstructure (fractional anisotropy, axial diffusivity, and radial diffusivity), disease duration, and ePAL were included in each model, such that the effect of each of these variables could be considered while controlling for the potential effects of the others. Interaction terms were not included in LME models, as we are interested in exploring independent effects of each of these variables rather than conditional effects (or the effect of one variable on another). P-values were FDR-corrected. To visually explore regional variation in these relationships we ran these LME models voxel wise in addition to volume-of-interest-based measures. Given the exploratory goal of voxelwise models, these values were not FDR-corrected; rather a threshold of p < 0.001 and a cluster size of 25 contiguous voxels was used for visualization purposes.

**Ethical approval.** All methods were performed in accordance with the guidelines and regulations of the Declaration of Helsinki and the University of Iowa Institutional Review Board.

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**References**

1. Meola, G. & Cardani, R. Myotonic dystrophies: An update on clinical aspects, genetic, pathology, and molecular pathomechanisms. *Biochim. Biophys. Acta* 1852, 594–606 (2015).
2. Nakamori, M. et al. Splicing biomarkers of disease severity in myotonic dystrophy. *Ann. Neurol.* 74, 862–872 (2013).
3. Wozniak, J. R., Mueller, B. A., Lim, K. O., Hemmy, L. S. & Day, J. W. Tractography reveals diffuse white matter abnormalities in Myotonic Dystrophy Type 1. *J. Neurol. Sci.* 341, 73–78 (2014).
4. Oikkersen, K. et al. Brain imaging in myotonic dystrophy type 1: A systematic review. *Neurology* 89, 960–969 (2017).
5. Minnerop, M. et al. The brain in myotonic dystrophy 1 and 2: evidence for a predominant white matter disease. *Brain* 134, 3530–3546 (2011).
6. Caso, F. et al. Cognitive impairment in myotonic dystrophy type 1 is associated with white matter damage. *PLoS ONE* 9, e104697 (2014).
7. Serra, L. et al. How genetics affects the brain to produce higher-level dysfunctions in myotonic dystrophy type 1. *Front. Neurol.* 30, 21–31 (2015).
8. Baldanzi, S. et al. Relationship between neuropsychological impairment and grey and white matter changes in adult-onset myotonic dystrophy type 1. *NeuroimageClin.* 12, 190–197 (2016).
9. Zanigni, S. et al. Relationship of white and gray matter abnormalities to clinical and genetic features in myotonic dystrophy type 1. *Neuroimage: Clinical* 11, 678–683 (2016).
10. Cabada, T. et al. Brain involvement in myotonic dystrophy type 1: A morphometric and diffusion tensor imaging study with neuropsychological correlation. *Arch. Clin. Neuropsychol.* 32, 401–412 (2017).
11. Yoo, W. K., Park, Y. G., Choi, Y. C. & Kim, S. M. Cortical thickness and white matter integrity are associated with CTG expansion size in myotonic dystrophy type 1. *Yonsei Med. J.* 58, 807–815 (2017).
12. Park, J.-S. et al. Diffusion tensor imaging and voxel-based morphometry reveal corticospinal tract involvement in the motor dysfunction of adult-onset myotonic dystrophy type 1. *Sci. Rep.* 8, 15592 (2018).
13. Labayru, G. et al. Regional brain atrophy in gray and white matter is associated with cognitive impairment in Myotonic Dystrophy type 1. *NeuroimageClin.* 24, 102078 (2019).
14. van Dorst, M. et al. Structural white matter networks in myotonic dystrophy type 1. *NeuroimageClin.* 21, 101615 (2019).
15. Heatwole, C. et al. Myotonic dystrophy health index: initial evaluation of a disease-specific outcome measure. *Muscle Nerve* 49, 906–914 (2014).
16. Overend, G. et al. Allele length of the DMPK CTG repeat is a predictor of progressive myotonic dystrophy type 1 phenotypes. *Hum. Mol. Genet.* 28, 2245–2254 (2019).
17. Baron, R. M. & Kenny, D. A. The moderator–mediator variable distinction in social psychological research: Conceptual, strategic, and statistical considerations. *J. Pers. Soc. Psychol.* 51, 1173 (1986).
18. Zammit, A. R., Robitaille, A., Piccinin, A. M., Muniz-Terrera, G. & Hofer, S. M. Associations between aging-related changes in grip strength and cognitive function in older adults. *A systematic review. J. Gerontol. A Biol. Sci. Med. Sci.* 74, S19–S27 (2019).
19. Shields, R. K. et al. Myotonic dystrophy type 1 alters muscle twitch properties, spinal reflexes, and perturbation-induced trans-cortical reflexes. *Muscle Nerve* 61, 205–212 (2020).
20. Mathieu, J., Boivin, H., Meunier, D., Gaudreault, M. & Bégin, P. Assessment of a disease-specific muscular impairment rating scale in myotonic dystrophy. *Neurology* 56, 336–340 (2001).
21. Mateos-Aierdi, A. J. et al. Muscle wasting in myotonic dystrophies: a model of premature aging. *Front. Aging Neurosci.* 7, 125 (2015).
22. Song, S.-K. et al. Demyelination revealed through MRI as increased radial (but unchanged axial) diffusion of water. *Neuroimage* 17, 1429–1436 (2002).
23. Sun, S.-W. et al. Noninvasive detection of cuprizone induced axonal damage and demyelination in the mouse corpus callosum. *Magn. Reson. Med.* 53, 302–308 (2006).
24. Alexander, A. L., Lee, J. E., Lazar, M. & Field, A. S. Diffusion tensor imaging of the brain. *Neurotherapeutics* 4, 316–329 (2007).
25. R Core Team. R: A language and environment for statistical computing. (R Foundation for Statistical Computing, Vienna, Austria, 2016).
26. Yushkevich, P. A. et al. User-guided 3D active contour segmentation of anatomical structures: significantly improved efficiency and reliability. *Neuroimage* 31, 1116–1128 (2006).
27. Gomes-Pereira, M., Bidichandani, S. I. & Monckton, D. G. Analysis of unstable triplet repeats using small-pool polymerase chain reaction. *Methods Mol. Biol.* 277, 61–76 (2004).
28. Monckton, D. G., Wong, L. J., Ashizawa, T. & Caskey, C. T. Somatic mosaicism, germline expansions, germline reversions and intergenerational reductions in myotonic dystrophy males: small pool PCR analyses. *Hum. Mol. Genet.* 4, 1–8 (1995).

29. Morales, F. et al. Somatic instability of the expanded CTG triplet repeat in myotonic dystrophy type 1 is a heritable quantitative trait and modifier of disease severity. *Hum. Mol. Genet.* 21, 3558–3567 (2012).

30. Ciosi, M. et al. Library preparation and MiSeq sequencing for the genotyping-by-sequencing of the Huntington disease HTT exon one trinucleotide repeat and the quantification of somatic mosaicism. *Protocol Exchange* https://doi.org/10.1038/protx.2018.089 (2018).

31. Giardine, B. et al. Galaxy: a platform for interactive large-scale genome analysis. *Genome Res.* 15, 1451–1455 (2005).

32. Heatwole, C. et al. Myotonic dystrophy health index: Correlations with clinical tests and patient function. *Muscle Nerve* 53, 183–190 (2016).

33. van der Plas, E. et al. Brain structural features of myotonic dystrophy type 1 and their relationship with Cig repeats. *J. Neuromuscul. Dis.* https://doi.org/10.3233/JND-190397 (2019).

34. Andersson, J. L. R., Skare, S. & Ashburner, J. How to correct susceptibility distortions in spin-echo echo-planar images: application to diffusion tensor imaging. *Neuroimage* 20, 870–888 (2003).

35. Andersson, J. L. R. & Sotiropoulos, S. N. An integrated approach to correction for off-resonance effects and subject movement in diffusion MR imaging. *Neuroimage* 125, 1063–1078 (2016).

36. Avants, B. B. et al. A reproducible evaluation of ANTs similarity metric performance in brain image registration. *Neuroimage* 54, 2033–2044 (2011).

37. Van Essen, D. C. et al. The WU-Minn human connectome project: An overview. *Neuroimage* 80, 62–79 (2013).

38. Fonov, V. S., Evans, A. C., McKinstry, R. C., Almli, C. R. & Collins, D. L. Unbiased nonlinear average age-appropriate brain templates from birth to adulthood. *Neuroimage* 47(Supplement 1), S102 (2009).

39. Young Kim, E. & Johnson, H. J. Robust multi-site MR data processing: iterative optimization of bias correction, tissue classification, and registration. *Front. Neuroinform.* 7, 29 (2013).

40. Wang, H. et al. Multi-atlas segmentation with joint label fusion. *IEEE Trans. Pattern Anal. Mach. Intell.* 35, 611–623 (2013).

41. Desikan, R. S. et al. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *Neuroimage* 31, 968–980 (2006).

42. Mori, S., Wakané, S., van Zijl, P. C. M. & Nagae-Poetscher, L. M. MRI Atlas of Human White Matter (Elsevier, Amsterdam, 2005).

43. Wakané, S. et al. Reproducibility of quantitative tractography methods applied to cerebral white matter. *Neuroimage* 36, 630–644 (2007).

44. Hua, K. et al. Tract probability maps in stereotaxic spaces: analyses of white matter anatomy and tract-specific quantification. *Neuroimage* 39, 336–347 (2008).

45. Johnson, W. E., Li, C. & Rabinovic, A. Adjusting batch effects in microarray expression data using empirical Bayes methods. *Biostatistics* 8, 118–127 (2007).

46. Fortin, J.-P. et al. Harmonization of multi-site diffusion tensor imaging data. *Neuroimage* 161, 149–170 (2017).

47. Benjamini, Y. & Hochberg, Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing (J. R. Stat. Soc, 1995).

**Author contributions**

T.R.K. and E.v.d.P. drafted the manuscript and cleaned the data; T.R.K. undertook neuroimage processing and statistical analysis; L.G. administered the Muscle Impairment Rating Scale and edited the manuscript; D.M. and S.A.C. provided genetic phenotyping data; D.M., S.A.C., and R.K.S. edited the manuscript; V.M. supervised the statistical analysis; L.G. administered the Muscle Impairment Rating Scale and edited the manuscript; D.M. and R.K.S. supervised the neuroimaging component of the study and edited the manuscript; P.C.N. conceptualized the study; supervised data collection; and edited the manuscript.

**Competing interests**

DGM has been a scientific consultant and/or received an honoraria/stock options from AMO Pharma, Biogen Idec, Charles River, LoQuis23, Small Molecule RNA, Triplet Therapeutics and Vertex Pharmaceuticals and had research contracts with AMO Pharma and Vertex Pharmaceuticals. DGM has received research grants/contracts from the European Union, CHDI, European Huntington Disease Network, Huntington Disease Society of America, National Institute of Health, Muscular Dystrophy UK and the Myotonic Dystrophy Support Group. DGM is on the Scientific Advisory Board of Myotonic (formerly the Myotonic Dystrophy Foundation) and EuroDyMA (European Dystrophia Myotonica Association), is a scientific advisor to the Myotonic Dystrophy Support Group and is a vice president of Muscular Dystrophy UK. TRK, EvdP, LG, SAC, VM, RKS, and PCN have no competing interest.

**Additional information**

**Supplementary Information** The online version contains supplementary material available at https://doi.org/10.1038/s41598-021-84520-2.

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