Research Article
Identification of Two Novel Mutations of *ABCD1* Gene in Pedigrees with X-Linked Adrenoleukodystrophy and Review of the Literature

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Background. X-linked adrenoleukodystrophy (ALD) is an inherited peroxisomal metabolism disorder, resulting from the loss-of-function mutation of ATP-binding cassette protein subfamily D1 (*ABCD1*) gene. The dysfunction of ALD protein, a peroxisomal ATP-binding cassette transporter, results in the excessive saturated very long-chain fatty acids (VLCFAs) accumulation in organs including the brain, spine, and adrenal cortex. X-ALD is characterized as the childhood, adolescent, adult cerebral ALD, adrenomyeloneuropathy (AMN), adrenal insufficiency, and asymptomatic phenotypes, exhibiting a high variety of clinical neurological manifestations with or without adrenocortical insufficiency. Results. In this study, we reported two cases of X-ALD, which were first diagnosed as adrenal insufficiency (Addison’s disease) and treated with adrenocortical supplement. However, both of the cases progressed as neurological symptoms and signs after decades. Elevated VLCFAs level, brain MRI scan, and genetic analysis confirmed final diagnosis. In addition, we identified two novel mutations of *ABCD1* gene, NM_000033.3 (*ABCD1*): c.874_876delGAG (p.Glu292del) and NM_000033.3 (*ABCD1*): c.96_97delCT (p.Tyr33Profs∗161), in exon 1 of *ABCD1* gene. Sanger sequencing confirmed that the proband’s mother of the first case was heterozygous carrying the same variant. Adrenal insufficiency-only type is very rare; however, it may be the starting performance of X-ALD. In addition, we summarized reported mutation sites and clinical manifestations to investigate the correlationship of phenotype-genotype of X-ALD. Conclusions. The early warning manifestations should be noticed, and the probability of X-ALD should be considered. This report could be beneficial for the early diagnosis and genetic counseling for patients with X-ALD.

1. Background

X-linked adrenoleukodystrophy (X-ALD) (OMIM#300100) is a X-linked inherited peroxisome metabolic neurodegenerative disorder, resulted from the defects in ATP-binding cassette protein subfamily D1 (*ABCD1*) gene. *ABCD1* gene locates in chromosome Xq28, contains 10 exons, and codes the adrenoleukodystrophy protein (ALDP), a member of the ABC transporter superfamily. ALDP expresses in the peroxisome membrane, containing 745 amino acids. It plays a key role in VLCFAs transporting into peroxisome for β-oxidation and degradation [1]. The defective ALDP results in the impaired degradation of saturated very long-chain fatty acids (VLCFAs). Excessive VLCFAs accumulate in plasma and tissues including adrenal cortex, cerebral white matter, or spinal cord [2] to exhibit clinical manifestations. Till now, there are more than three thousand variants reported (X-ALD mutation database, http://www.x-ald.nl/) [3].

Based on the age of onset and the extent of organs affected, X-ALD is classified into seven clinical subtypes: childhood cerebral, adolescent cerebral, adult cerebral, adrenomyeloneuropathic (AMN), cerebellar, adrenal insufficiency-only (Addison’s disease), and asymptomatic or presymptomatic types. X-ALD exhibits a high variety of clinical characteristics [4]. Childhood cerebral ALD...
(CCALD) and adult-onset AMN are the two most common phenotypes [5]. The clinical symptoms include progressive dysfunction of central and peripheral nervous systems, such as apraxia, behavioral disturbance, abnormalities of sphincter control, sensory loss, or paresthesia, with or without adrenal insufficiency. Brain MRI shows a typical pattern of symmetrical enhanced T2WI signal in the bilateral white matter around the lateral ventricle, cerebellum, and the genu of the corpus callosum [6]. Significantly elevated level of VLCFAs is the important diagnostic clue for the individuals with ALD. There are three parameters of VLCFAs to analyze the concentration of C26:0, the ratio of C24:0/C22:0, and the ratio of C26:0/C22:0 [5]. Final diagnosis relays on the genetic analysis to detect ABCD1 mutation.

Adrenal insufficiency-only type is rare, especially in females less than 1% [7,8]. Some individuals were diagnosed as Addison’s disease without symptoms of cerebral or myelopathy ALD for decades. However, the early onset Addison’s disease may gradually progress to exhibit neuropsychiatric symptoms, showing a time-dependent follow-up [9].

In this study, we reported two pedigrees of X-ALD patients first diagnosed as Addison’s disease for decades and gradually exhibited variable clinical neuropsychiatric manifestations in their late twenty’s and showed rapid progression and poor prognosis. In addition, we identified two novel mutations of ABCD1 gene, NM_000033.3 (ABCD1): c.874_876delGAG (p.Glu292del) and NM_000033.3 (ABCD1): c.96_97delCT (p.Tyr33Profs*161), in the two pedigrees of X-ALD. Further study of genotype-phenotype correlation analysis is needed to provide deeper insights into X-ALD.

2. Methods and Materials

2.1. Patients’ Recruitment. The diagnosis of X-ALD depends on clinical symptoms, biochemical parameters, neurological imaging, and genetic analysis. This study was approved by the Ethics Committee of the Affiliated Hospital of Qingdao University.

2.2. Genetic Analysis and Prediction of Pathogenicity. Genomic DNA was extracted from peripheral blood sample using the QIAamp Blood DNA Mini Kit (QIAGEN, USA) according to the manufacturer’s protocol. After amplification using 2X polymerase chain reactions (PCR) MasterMix polymerase (Tiangen, China) by ABI9700 PCR (Life technology, USA), the products were captured and purified with Panel probe (illumine Inc. USA) and then directly sequenced on the ABI 3500 automated DNA sequencer (Life Technology, USA). Mutations of ABCD1 were detected using next generation sequencing (NGS) and subsequently confirmed using Sanger sequencing. The databases including ESP6500, 1000 Genomes, and dbSNP and UniProt were used for elimination of polymorphisms. Prediction of mutations was performed with PROVEAN (Protein Variation Effect Analyzer, http://provean.jcvi.org/) and Mutation Taster (http://www.mutationtaster.org) to predict the possible impacts of amino acid substitution on the structure and function. The PROVEAN score represented the normalized probability that the amino acid change is deleterious. The cutoff of PROVEAN score was –2.5. The PROVEAN score < –2.5 was predicted to be deleterious. In addition, the following forty genes associated with congenital adrenal hypoplasia or glucocorticoid deficiency were investigated for Case 2, including MC2R, MRAP, AAAS, AIP, PRKAR1A, PDE11A, PDE8B, PRKACA, MEN1, RET, GNAS, ARMC5, NFI, NR3C1, LMNA, SOX3, PCSK1, PROP1, LHX3, HSD11B1, CYP21A2, CYP17A1, CYP11B1, HSD3B2, STAR, MCM4, TBX9, POR, POMC, AIRE, NR5A1, and so on.

2.3. Three-Dimensional Structure Prediction of Protein and the Potential Dysfunction. We identified the alteration of the ABCD1-encoded ALD protein structure and potential dysfunction using the SWISS-MODEL workspace (http://swiss-model.expasy.org).

2.4. Case Presentations and Biochemistry Results

Case 1. A 27-year-old male was presented in hospital with weakness, fatigue, poor academic performance, exhibiting skin, and mucous membrane pigmentation, especially in areolae, joints, and axilla for two decades. The cortisol at 8 am was reduced 3.58 nmol/L (range 171–536 nmol/L), and adrenocorticotropic hormone (ACTH) was increased 499.30 pg/mL (range 7.2–63.6 pg/mL). After diagnosed as primary adrenal insufficiency (Addison’s disease), he was administrated with glucocorticoid replacement (hydrocortisone 20 mg per day) for one year, and the skin pigmentation and weakness improved. In recent half a year, the patient acted as mild cognitive impairment, slurred speech, ataxia, and rapidly progressed as walking instability, incontinence, and paroxysmal blurring vision. Physical examination detected scanty scalp hair, V-grade muscle strength, hypomyotonia, tendon hyperreflexia (+++), and positive Babinski’s sign and Gordon’s sign. The light and deep reflexes were normal. The heel-to-knee test, alternating movement test, and Romberg’s test were negative. The respiratory, cardiovascular, and abdominal examinations were unremarkable. Brain MRI showed symmetrical distribution of hyperintensity long T1 and long T2 signal in bilateral splenium of corpus callosum and widened fissures and sulci (Figure 1(a)). Serum VLCFA levels were significantly increased in the proband and also elevated in the proband’s mother (Table 1, Figure 2(a), II-10). Until now, his mother did not show manifestations of cerebral disorders, myeleterosis, or adrenal insufficiency. His maternal grandmother (Figure 2(a), I-2) died at her 55-year-old due to encephalopathy. However, family members were unable to provide valid information, and no blood samples could be obtained for testing. Other siblings of his mother did not show neuropsychiatric manifestations. Genetic analysis revealed variants c.874_876delGAG of ABCD1 gene in exon 1. According
to the clinical manifestations and genetic analysis, a pedigree of X-ALD was constructed. The patient was administrated with hydrocortisone 20mg per day, mecobalamine, and compound vitamin B. During the follow-up, the proband showed rapidly progressed neuropsychiatric manifestations including blurred vision, instability of gait, and incontinence within six months.

Case 2. A 31-year-old male was presented with adrenal crisis, exhibiting severe fatigue, anorexia, nausea, and vomiting five years ago. The patient had significantly

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Figure 1: Brain magnetic resonance imaging (MRI) scan of patients. (a) Symmetric bands with high T2 low T1 abnormal signal in bilateral genu of the corpus callosum detected in case one. (b) MRI scan of case two revealed long T1 and long T2 signals in white matter of bilateral frontal lobes, around the anterior of lateral ventricle, and splenium of the corpus callosum. (c) Computed tomography (CT) scan of the adrenal gland of case one to show bilateral adrenal atrophy. Red arrow shows lesions.
generalized skin pigmentation especially in the gingiva, areolae, and creases of the hands. Laboratory examination revealed severe hyponatremia (serum sodium 108–125 mmol/L, normal range 145–155 mmol/L), markedly decreased cortisol level (24.83–45.40–35.13 mmol/L) and increased ACTH level (1522–1071–1750 pg/mL) at 8 am–4 pm–0 am, respectively. The adrenal CT scan showed bilateral adrenal atrophy. At first, the patient was diagnosed as adrenal insufficiency, without exhibiting central or peripheral nephropathy. After therapy with hydrocortisone, symptoms of fatigue and vomiting improved. The patient showed incorrugible severe hyponatremia, only corrected into normal by treated combination with fludrocortisone (0.05 mg per day). The serum sodium levels were maintained in normal range for five years. However, in recent eight months, the patient showed significantly aggravated walking instability, cognitive dysfunction, intellectual disabilities, emotional disturbance, memory loss, and incontinence.

Neurological physical examination revealed positive Babinski’s sign and Hoffman’s sign, hyperactive tendon reflexes, and abnormal heel-to-knee-to-toe test. He spoke unclearly with dysarthria, while vision and hearing were normal. Brain MRI revealed long T1 and long T2 signals in white matter of bilateral frontal lobes and around the anterior of lateral ventricle and corpus callosum (Figure 1(b)). The patient was sick and showed poor learning ability during childhood. His brother (Figure 2(c), III-3) also showed sickness in childhood and died at 8-year-old. His maternal uncle (Figure 2(c), IL-5) was weak and fatigued in his youth and showed disturbance of consciousness from 25-year-old and died at 28-year-old. None of the other family members exhibited neuropsychiatric symptoms and signs of X-ALD. To confirm the diagnosis, genetic sequencing of the proband detected deletion mutation c.96_97delCT in ABCD1 gene. Thus, the pedigree of X-ALD was suspected. During the follow-up, the patient was treated with hydrocortisone and fludrocortisone, and serum sodium level was maintained in normal range. However, he showed rapidly progressed symptoms of incontinence and consciousness dysfunction.

2.5. Genetic Analysis and Prediction of Pathogenicity. Genetic sequencing of ABCD1 gene was performed on the probands of two cases and the familial relatives of the proband in case one. In case one, genetic sequencing detected heterozygous mutation of NM_000033.3 (ABCD1): c.874_876delGAG (p.Glu292del) in ABCD1 gene exon 1 (Figures 2(a) and 2(b)), leading to the deletion of translation product glutamate and accumulation of VLCFAs (Table 1).

Sanger sequencing of ABCD1 gene was performed on his mother and confirmed the heterozygous point mutation for the same variant. In case two, gene sequencing showed a deletion mutation c.96_97delCT in ABCD1 gene (Figures 2(c) and 2(d)), leading to the frameshift and premature transcription termination of amino acid p.Tyr33Profs*161. His immediate family members (parents and his sister) refused to perform genetic sequencing.

The frequency of those variants was examined in the reference database. Both novel mutations were not detected in the database including ESP6500, 1000 Genomes, or dbSNP (Table 2). The potential pathogenicity of both mutations was investigated using prediction bioinformatics, suggesting both mutations were possible to be disease causing (Table 2). The pathogenicity evaluation was analyzed according to the criteria of American College of Medical Genetics (ACMG) guideline and given in Table 2.

2.6. Three-Dimensional Structure Prediction of Protein and Potential Dysfunction. The ABCD1 gene-encoded ALD protein contains transmembrane domains and three segments on the peroxisome side. We identified the predicted alteration of the ABCD1 protein structure and its potential dysfunction induced by mutations NM_000033.3 (ABCD1): c.874_876delGAG (p.Glu292del) and NM_000033.3 (ABCD1): c.96_97delCT (p.Tyr33Profs*161) in silico (Figure 3). In both of the mutations, p.Glu292del and p.Tyr33Profs*161, the amino acid changes in ALD protein due to those deletion mutations of ABCD1 gene caused more than three visible differences in protein structure, might affect function, leading to the impaired degradation of VLCFAs and inflammation environment.

3. Discussion

X-ALD is a demyelinating and neurodegenerative disorder due to the loss-of-function mutation of ABCD1 gene. ABCD1 gene codes the peroxisome transporter protein ALDP, which plays an important role in regulating peroxisome oxidation and degradation of VLCFAs [5]. The

### Table 1: Clinical characteristics and genetic analysis results of the patients.

| No. | Sex | Family history | C24: C22 | C26: C22 | Cortisol (nmol/L) | ACTH (pg/mL) | Sodium (mmol/L) | Potassium (mmol/L) | Corticoid dosage | Mutation | Exon |
|-----|-----|----------------|----------|----------|------------------|-------------|-----------------|--------------------|----------------|----------|------|
| Case 1 | M | + | 1.75 | 0.089 | 3.58 | 499.3 | 128 | 5.1 | Hydrocortisone, 20 mg/d | c.874_876delGAG | 1 |
| Case 1 | F | + | 1.06 | 0.033 | ND | ND | 138 | 4.8 | None | c.874_876delGAG | 1 |
| Case 2 | M | – | ND | ND | 45.4 | 1071 | 125 | 5.3 | Hydrocortisone, 30 mg/d | c.96_97delCT | 1 |

Reference range: <1.39 <0.023 171-536 7.2-63.6 135-145 3.5-5.5 Fludrocortisone
Figure 2: Continued.
dysfunction of ALDP, a fatty acid transporter, results in the impaired VLCFA oxidation and excessive VLCFAs accumulation in tissues [5, 10]. Excessive VLCFAs in cytoplasm lead to mitochondrion dysfunction and ER stress and finally caused cell death. In addition, accumulated VLCFAs arouse inflammation, resulting in demyelinating and neurodegeneration.

In this study, we reported two Chinese X-ALD pedigrees. Both of the probands were diagnosed as adrenal insufficiency at beginning without neuropsychiatric symptoms for decades, but developed into neurological disorders at their late 20s and showed rapid progression and poor prognosis. Two novel mutations in exon 1 of \textit{ABCD1} gene were detected and suggested to be associated with X-ALD.

According to the previous reports, the majority is cerebral ALD among those seven types of ALD [11]. CCALD is early onset with a feature of rapidly progressed neuro-inflammatory of cerebral demyelination. Neurological symptoms of AMN commonly occur from the ages of 20s, with manifests as a chronic progressive paraparesis, accompanied with sensory and motor disturbances [12]. Due to the variable clinical manifestations, some are difficult to make the diagnose at the early stage [13]. Adrenocortical insufficiency was reported as the first clinical manifestation of 38% X-ALD, greater than the group of spinal cord 25% and cerebral disorders 14.5% [9]. To be noticed, some of those male patients diagnosed as adrenal insufficiency may develop with cerebral or myeloneuropathy ultimately and show rapid progressive neurological symptoms and poor prognosis. On the contrast, female ALD patients rarely show adrenocortical insufficiency or cerebral ALD. Most of female patients suffer from lately onset myelopathy [10, 14]. We reported two cases; both were first diagnosed as Addison’s disease, and then, the therapy focused on the adrenocortical replacement. However, after the neurological symptoms and signs become apparent, both of the cases were rapidly progressed. The therapy including fatty acid restriction or neurotrophic treatment did not show the therapeutic effect. Therefore, at diagnosis of congenital adrenal insufficiency, the probability of ALD should be fully considered, especially for male patients [8, 15]. To make the early diagnosis, family history, MRI scan of brain or spinal cord, VLCFAs measurement, and genetic screening may be helpful for those patients with subtle early neurological manifestations.

We identified two novel point mutations in exon 1 of \textit{ABCD1} gene in the pedigree of first case NM_000033.3 (ABCD1): c.874_876delGAG (p.Glu292del) and in second case NM_000033.3 (ABCD1): c.96_97delCT (p.Tyr33Profs∗161). The genetic sequencing demonstrated nucleotides deletion, resulting in those frameshift mutations. Based on the clinical symptoms and positive neurologic signs, family history, imaging, biochemical characteristics, and the results of pathogenicity prediction using bioinformatics tools, we suggest those mutations as pathogenic variants. However, the biopsy of the brain, spinal cord, and adrenal gland was not done. Based on the elevated plasma VLCFAs, the deleterious change of ALDP impaired the cellular beta-oxidation of fatty acid [16].

There are more than three thousand mutations reported in the ALD database (http://www.x-ald.nl/). Among them, over 900 variant sites are nonrecurrent unique. Most of them are pathogenic mutations. Among more than thousand mutation sites (combination of 913 nonrecurrent mutations, 120 synonymous variants, and 247 variants of unknown significance), 67.57% of them are pathogenic mutations, 9.65% are synonymous, 2.16% are benign, 18.65% are VUS (variant of undetermined significance), and 1.87% are status unknown mutations detected during the screening of newborns (Figure 4(a)). More than 77.5% variant sites are point mutations; other types include 12.93% deletion, 1.97% del-insertion, 4.22% insertion, and 3.37% duplication (Figure 4(b)). Most of the mutations locate in the exon (92.94%) and others in the intron (6.18%), 5’UTR (0.61%), and 3’UTR (0.26%), respectively [4]. In the database, nearly half of the mutations were reported anchoring in exon 1 (Figure 4(c)), which encodes the transmembrane domain (TMD) of the protein, contains ligand-specific binding sites, and plays an important role in the localization of ALDP [17]. Another clustering of mutations occurs in exon 6, which encodes the ATP-binding domain of ALDP [18]. ALDP locates in the peroxisome membrane to transport VLCFAs from cytosol into the peroxisome for degradation. Pathogenic \textit{ABCD1} mutations may induce defective stability of ALDP transmembrane structure region and ATPase activity.
| No. | Nucleotide mutations | Amino acid variants | Exon | Variation type | Inheritance | ESP6500 | 1000G | dbSNP | PROVEAN score | Mutation taster | Pathogenicity evaluation |
|-----|----------------------|---------------------|------|----------------|-------------|---------|-------|-------|---------------|----------------|--------------------------|
| Case 1 | c.874_876delGAG | p.Glu292.del | 1 | Deletion | Familial | ND | ND | ND | −13.987 (deleterious) | Disease causing | Likely pathogenic (PM2 + PM4, PP3 + PP4 + PP5) |
| Case 2 | c.874_876delGAG | p.Tyr33Profs *161 | 1 | Deletion | Familial | ND | ND | ND | −6.7 (deleterious) | Disease causing | Pathogenic (PVS1, PM2 + PM4, PP3 + PP4 + PP5) |

ND, not detected. Pathogenicity evaluation of American College of Medical Genetics (ACMG) guideline. PROVEAN score (cutoff = −2.5)
to affect the localization of VLCFAs to the peroxisome correctly. As a result, VLCFAs cannot be transported into peroxisomes for $\beta$-oxidation and then accumulate mainly in the white matter, neuron axons of the central nervous system, and adrenal cortex. Excess VLCFAs accumulation leads to destructive progression by triggering oxidative stress and mitochondrial dysfunction, chronic inflammation, lipid-induced neuron apoptosis, and degenerative changes in the nervous system, and finally lead to neurodegeneration and adrenal insufficiency [3, 19]. Though some variations may not affect protein translation or function, they still represent polymorphisms. The genotype-phenotype correlationship has not been identified clearly [20, 21]. To further investigate the relationship between genotype and phenotype, we searched the database of UniProt and reported references and then summarized the reported mutation sites and clinical manifestations including childhood and adolescent cerebral ALD, adult ALD, AMN, adrenal insufficiency-only, asymptomatic or presymptomatic types, and complicated ALD which cannot be classified (Figure 5). X-ALD has high genetic heterogeneity. The clinical manifestations are highly variable even in the pedigree carrying same variant. The main phenotypes differ from different periods of the disease. The same variants can exhibit

\[ \text{Health control} \]
\[ \text{p.Glu292del} \]
\[ \text{p.Tyr33Profs}^{\star161} \]

**Figure 3:** Three-dimensional structure prediction of adrenoleukodystrophy (ALD) protein. (a) The modeled structure of health control. The visible alteration of ALD protein structure induced by mutations (b) c.874_876delGAG (p.Glu292del) and (c) c.96_97delCT (p.Tyr33Profs $^{\star161}$) are indicated in circles. It indicates that those mutations might lead to alteration of the protein structure and potential dysfunction.
Figure 4: *ABCD1* gene reported from the ALD mutation database. (a) Mutation sites types of variants of *ABCD1* gene among reported nonrecurrent and other mutation sites, including pathogenic, synonymous, benign, variants of undetermined significance (VUS), and screening of newborns state of unknown (UN). (b) The content of *ABCD1* mutations, including points mutations, deletion, del-insertion, insertion, and duplication (http://www.x-ald.nl/). (c) The distribution of mutations, including point mutation, deletion, del-insertion, insertion, and duplication. Most of the mutations harboring in exon 1. The lower panel shows the mutation rate in each exon. Bold shows the location of two novel mutations indicated in this study.
Figure 5: Summary of the reported mutation sites and clinical manifestations from the database of UniProt (https://www.uniprot.org/uniprot/) and reported references [22–35]. The clinical types include childhood and adolescent cerebral ALD (CCALD and adoles ALD), adult ALD, AMN (adrenomyeloneuropathy), adrenal insufficiency-only (AIO), complicated ALD which cannot be classified, and asymptomatic or pre-symptomatic types.
different clinical features. Thus, further effects may be needed to clarify the diversity in clinical manifestations and the correlation of genotype-phenotype.

Therapeutic strategies include adrenocortical replacement; some of the adrenal insufficiency patients may require both glucocorticoid and mineralocorticoid to maintain water electrolyte balance. For severe adrenal insufficiency or even adrenal crisis, timely initiation of steroid replacement is essential. Nutrition intervention, for instance, low-fat diet and Lorenzo’s oil may reduce VLCFAs level [36]. In this way, the risk of development with cerebral disorders may be reduced. However, they lack the evidence to improve the symptoms and progression. Another efficiency approach is hematopoietic stem cell transplantation (HSCT), especially in the early stage of the disease [37]. Transplantation with the autologous bone marrow transplanted in vitro with ABCD1 gene modification has been performed [38, 39]. Bone marrow transplant has been reported in CCALD therapy, and most of the patients achieved improved prognosis. However, the preexisted severe symptoms may influence the outcome after transplantation [40]. In animal experiments, Abcd1 mice show reduction of the mitochondrial biogenesis driven by the PGC-1α/PPARγ pathway [41]. Oxidative stress is involved in the physiopathological mechanism of neurodegenerative cascade and mitochondrial depletion [42]. Pioglitazone, a PPARγ/PGC-1α axis activator, may modulate the anti-inflammatory and antioxidant response. Recently, pioglitazone is under phase II clinical trials for adrenoleukodystrophy patients, revealing a potential for clinical translation [43]. Therapy options are limited; in addition, some therapeutic approaches relay on the early diagnosis and intervention. Therefore, gene sequencing analysis may be an important tool for newborn screening and genetic counseling.

4. Conclusions

In this study, we reported two novel mutations of ABCD1 gene in men with X-ALD. These two cases were both considered as adrenal insufficiency at first and finally diagnosed with X-ALD when neurological manifestations occurred after several years. In these two pedigrees, we identified two novel mutations of ABCD1 gene, NM_000033.3(ABCD1): c.874_876delGAG (p.Glu292del) and NM_000033.3(ABCD1): c.96_97delCT (p.Tyr33Profs∗161). The screening test for ALD, such as VLCFA concentration and brain MRI scan, should be considered in the early onset Addison’s disease patients with the family history to screen X-ALD early.

Abbreviations

X-ALD: X-linked adrenoleukodystrophy
ABCD1: ATP-binding cassette subfamily D member 1
ALDP: Adrenoleukodystrophy protein
VLCFAs: Very long-chain fatty acids
AMN: Adrenomyeloneuropathy
ACTH: Adrenocorticotrophic hormone
MRI: Magnetic resonance imaging

Data Availability

The data used to support the findings of this study are included within the article.

Ethical Approval

All procedures performed in this study involving human participants were in accordance with the Ethical Standards of the Affiliated Hospital of Qingdao University with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Consent

The clinical data, images, and individual details were obtained from the probands and their family members. All the participants gave their written consent for publication.

Disclosure

A preprint has previously been published in Research Square [44]. The funders had no role in this study.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Bingzi Dong and Wenshan Lv carried out the molecular genetic studies, the design of this study, and drafted the manuscript. Lili Xu and Zhongchao Wang participated in the sequence alignment and coordination of this study. Yuhang Zhao, Xiaofang Sun, Zhengjiu Fu, and Bingfei Cheng provided helpful suggestion. Yangang Wang conceived the study and took responsibility of this study. All authors read and approved the final manuscript. Bingzi Dong and Wenshan Lv contributed equally to this work as co-first authors.

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References

[1] S. Kemp, I. C. Huffnagel, G. E. Linthorst, R. J. Wanders, and M. Engelen, “Adrenoleukodystrophy-neuroendocrine pathogenesis and redefinition of natural history,” Nature Reviews Endocrinology, vol. 12, no. 10, pp. 606–615, 2016.
[2] J. Berger, S. Forss-Petter, and F. S. Eichler, “Pathophysiology of X-linked adrenoleukodystrophy,” Biochimie, vol. 98, pp. 135–142, 2014.
[3] S. Kemp, A. Pujol, H. R. Waterham et al., “ABCD1 mutations and the X-linked adrenoleukodystrophy mutation database:
role in diagnosis and clinical correlations,” Human Mutation, vol. 18, no. 6, pp. 499–515, 2001.

[4] M. Engelen, S. Kemp, M. de Visser et al., “X-linked adrenoleukodystrophy (X-ALD): clinical presentation and guidelines for diagnosis, follow-up and management,” Orphanet Journal of Rare Diseases, vol. 7, pp. 51, 2012.

[5] S.-S. Chu, J. Ye, H.-W. Zhang et al., “Eight novel mutations in the ABCD1 gene and clinical characteristics of 25 Chinese patients with X-linked adrenoleukodystrophy,” World Journal of Pediatrics, vol. 11, no. 4, pp. 366–373, 2015.

[6] H. W. Moser, D. J. Lloes, E. R. Melhem et al., “X-linked adrenoleukodystrophy: overview and prognosis as a function of age and brain magnetic resonance imaging abnormality. A study involving 372 patients,” Neuropediatrics, vol. 31, no. 5, pp. 227–239, 2000.

[7] M. Engelen, M. Barbier, I. M. Dijkstra et al., “X-linked adrenoleukodystrophy in women: a cross-sectional cohort study,” Brain: A Journal of Neurology, vol. 137, no. Pt 3, pp. 693–706, 2014.

[8] M. A. Horn, M. M. Erichsen, A. S. B. Wolff et al., “Screening for X-linked adrenoleukodystrophy among adult men with Addison’s disease,” Clinical Endocrinology, vol. 79, no. 3, pp. 316–320, 2013 Sep.

[9] I. C. Huffnagel, F. K. Laheji, R. Azir-Bose et al., “The natural history of adrenal insufficiency in X-linked adrenoleuko- dystrophy: an international collaboration,” Journal of Clinical Endocrinology & Metabolism, vol. 104, no. 1, pp. 118–126, 2019.

[10] M. Engelen, S. Kemp, and B.-T. Poll-The, “X-linked adreno- leukodystrophy: pathogenesis and treatment,” Current Neurology and Neuroscience Reports, vol. 14, no. 10, p. 486, 2014.

[11] M.-y. Jiang, Y.-n. Cai, C.-l. Lianget al., “Clinical, biochemical, neuroimaging and molecular findings of X-linked Adreno- leukodystrophy patients in South China,” Metabolic Brain Disease, vol. 30, no. 6, pp. 1439–1444, 2015.

[12] C. Tran, J. Patel, H. Stacy et al., “Long-term outcome of patients with X-linked adrenoleukodystrophy: a retrospective cohort study,” European Journal of Paediatric Neurology, vol. 21, no. 4, pp. 600–609, 2017.

[13] S. Paláu-Hernández, I. Rodríguez-Leyva, and J. M. Shiguemori-Medina, “Late onset adrenoleukodystrophy: a review related clinical case report,” eNeurologicalSci, vol. 14, pp. 62–67, 2019.

[14] S. Hsieh and P. C. White, “Presentation of primary adrenal insufficiency in childhood,” Journal of Clinical Endocrinology & Metabolism, vol. 96, no. 6, pp. E925–E928, 2011.

[15] L. Eng and M. O. Regelmann, “Early onset primary adrenal insufficiency in males with adrenoleukodystrophy: case series and literature review,” The Journal of Pediatrics, vol. 211, pp. 211–214, 2019.

[16] I. C. Huffnagel, M.-C. van de Beek, A. L. Showers et al., “Comparison of C26:0-carnitine and C26:0-lysophosphatidylcholine as diagnostic markers in dried blood spots from newborns and patients with adrenoleukodystrophy,” Molecular Genetics and Metabolism, vol. 122, no. 4, pp. 209–215, 2017.

[17] E. Procko, M. L. O’Mara, W. F. D. Bennett, D. P. Tieleman, and R. Gaudet, “The mechanism of ABC transporters: general lessons from structural and functional studies of an antigenic peptide transporter,” The FEBS Journal, vol. 23, no. 5, pp. 1287–1302, 2009.

[18] Y.-F. Niu, W. Ni, and Z.-Y. Wu, “ABCD1 mutations and phenotype distribution in Chinese patients with X-linked adrenoleukodystrophy,” Gene, vol. 522, no. 1, pp. 117–120, 2013.

[19] B. R. Turk, C. Theda, A. Fatemi, and A. B. Moser, “X-linked adrenoleukodystrophy: pathology, pathophysiology, diagnostic testing, newborn screening, and therapies,” International Journal of Developmental Neuroscience: The Official Journal of the International Society for Developmental Neuroscience, vol. pii, no. 19, pp. 30133–30139, 2019.

[20] C. Wiesinger, F. S. Eichler, and J. Berger, “The genetic landscape of X-linked adrenoleukodystrophy: inheritance, mutations, modifier genes, and diagnosis,” The Application of Clinical Genetics, vol. 8, pp. 109–121, 2015.

[21] C. Wang, H. Liu, B. Han, H. Zhu, and J. Liu, “A novel ABCD1 gene mutation causes adrenomyeloneuropathy in a Chinese family,” Brain and Behavior, vol. 9, no. 10, Article ID e01416, 2019.

[22] M. B. R. Lachtermacher, H. N. Seunze, A. B. Moser, H. W. Moser, and K. D. Smith, “Determination of 30 X-linked adrenoleukodystrophy mutations, including 15 not previously described,” Human Mutation, vol. 15, no. 4, pp. 348–353, 2000.

[23] M. Wichers, W. Köhler, W. Brennemann et al., “X-linked adrenomyeloneuropathy associated with 14 novel ALD-gene mutations: no correlation between type of mutation and age of onset,” Human Genetics, vol. 105, no. 1-2, pp. 116–119, 1999.

[24] C. Perusi, M. Gomez-Lira, M. Mottes et al., “Two novel missense mutations causing adrenoleukodystrophy in Italian patients,” Molecular and Cellular Probes, vol. 13, no. 3, pp. 179–182, 1999.

[25] S. Fuchs, C. O. Sarde, H. Wedemann, E. Schwinger, J. L. Mandel, and A. Gal, “Missense mutations are frequent in the gene for X-chromosomal adrenoleukodystrophy (ALD),” Human Molecular Genetics, vol. 3, no. 10, pp. 1903–1905, 1994.

[26] E. W. Krasemann, V. Meier, G. C. Korenke, D. H. Hunneman, and F. Hanefeld, “Identification of mutations in the ALD-gene of 20 families with adrenoleukodystrophy/adrenomyelo- neuropathy,” Human Genetics, vol. 97, no. 2, pp. 194–197, 1996.

[27] G. Montagna, A. Di Biase, M. Cappa et al., “Identification of seven novel mutations in ABCD1 by a DHPLC-based assay in Italian patients with X-linked adrenoleukodystrophy,” Human Mutation, vol. 25, p. 222, 2005.

[28] Y. Wang, R. Busin, C. Reeves et al., “X-linked adrenoleu- kodystrophy: ABCD1 novo mutations and mosaicism,” Molecular Genetics and Metabolism, vol. 104, no. 1-2, pp. 160–166, 2011.

[29] L. Dvoráková, G. Štorkánová, G. Unterrainer et al., “Eight novel ABCD1gene mutations and three polymorphisms in patients with X-linked adrenoleukodystrophy: the first polymorphism causing an amino acid exchange,” Human Mutation, vol. 18, no. 1, pp. 52–60, 2001.

[30] C. P. Guimarães, M. Lemos, I. Menezes, T. Coelho, C. Sá-Miranda, and J. E. Azevedo, “Characterisation of two mutations in the ABCD1 gene leading to low levels of normal ALDP,” Human Genetics, vol. 109, no. 6, pp. 616–622, 2001.

[31] M. G. Lira, M. Mottes, P. F. Pignatti et al., “Detection of mutations in the ALD gene (ABCD1) in seven Italian families: description of four novel mutations,” Human Mutation, vol. 16, no. 3, p. 271, 2000.

[32] P. Shukla, N. Gupta, S. Gulati et al., “Molecular analysis of ABCD1 gene in Indian patients with X-linked adrenoleu- kodystrophy,” Clinica Chimica Acta, vol. 412, no. 23-24, pp. 2289–2295, 2011.
[33] A. Braun, H. Ambach, S. Kammerer et al., “Mutations in the gene for X-linked adrenoleukodystrophy in patients with different clinical phenotypes,” *The American Journal of Human Genetics*, vol. 56, pp. 854–861, 1995.

[34] L. X. Liu, K. Janvier, V. Berteaux-Lecellier, N. Cartier, R. Benarous, and P. Aubourg, “Homo- and heterodimerization of peroxisomal ATP-binding cassette half-transporters,” *Journal of Biological Chemistry*, vol. 274, no. 46, pp. 32738–32743, 1999.

[35] G. C. Korenke, E. Krasemann, V. Meier, W. Beuche, D. H. Hunneman, and F. Hanefeld, “First missense mutation (W679R) in exon 10 of the adrenoleukodystrophy gene in siblings with adrenomyeloneuropathy,” *Human Mutation*, vol. 11, no. S1, pp. S204–S206, 1998.

[36] P. Aubourg, C. Adamsbaum, M.-C. Lavallard-Rousseau et al., “A two-year trial of oleic and erucic acids (“Lorenzo’s Oil”) as treatment for adrenomyeloneuropathy,” *New England Journal of Medicine*, vol. 329, no. 11, pp. 745–752, 1993.

[37] F. Eichler, C. Duncan, P. L. Musolino et al., “Hematopoietic stem-cell gene therapy for cerebral adrenoleukodystrophy,” *New England Journal of Medicine*, vol. 377, no. 17, pp. 1630–1638, 2017.

[38] N. Cartier, S. Hacein-Bey-Abina, C. C. Bartholomae et al., “Hematopoietic stem cell gene therapy with a lentiviral vector in X-linked adrenoleukodystrophy,” *Science*, vol. 326, no. 5954, pp. 818–823, 2009.

[39] J. S. Kühl, J. Kupper, H. Baqué et al., “Potential risks to stable long-term outcome of allogeneic hematopoietic stem cell transplantation for children with cerebral X-linked adrenoleukodystrophy,” *JAMA Network Open*, vol. 1, no. 3, Article ID e180769, 2018.

[40] W. P. Miller, S. M. Rothman, D. Nascene et al., “Outcomes after allogeneic hematopoietic cell transplantation for childhood cerebral adrenoleukodystrophy: the largest single-institution cohort report,” *Blood*, vol. 118, no. 7, pp. 1971–1978, 2011.

[41] J.-F. Lu, A. M. Lawler, P. A. Watkins et al., “A mouse model for X-linked adrenoleukodystrophy,” *Proceedings of the National Academy of Sciences*, vol. 94, no. 17, pp. 9366–9371, 1997.

[42] J. M. Powers, Z. Pei, A. K. Heinzer et al., “Adreno-leuko-dystrophy: oxidative stress of mice and men,” *Journal of Neuropathology and Experimental Neurology*, vol. 64, no. 12, pp. 1067–1079, 2005.

[43] A. Pujol, “Novel therapeutical targets and drug candidates for modifying disease progression in adrenoleukodystrophy,” *Endocrine Development*, vol. 30, pp. 147–160, 2016.

[44] B. Dong, W. Lv, L. Xu et al., “Identification of two novel mutations of ABCD1 gene in pedigrees with X-linked adreno-leukodystrophy,” impress.