Antibody Response Patterns to *Bordetella pertussis* Antigens in Vaccinated (Primed) and Unvaccinated (Unprimed) Young Children with Pertussis

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In a previous study, it was found that the antibody response to a nonvaccine pertussis antigen in children who were vaccine failures was reduced compared with the response in nonvaccinated children who had pertussis. In two acellular pertussis vaccine efficacy trials in Sweden, we studied the convalescent-phase enzyme-linked immunosorbent assay (ELISA) geometric mean values (GMVs) in response to pertussis toxin (PT), filamentous hemagglutinin (FHA), pertactin (PRN), and fimbriae (FIM 2/3) in vaccine failures and controls with pertussis. In Germany, the antibody responses to *Bordetella pertussis* antigens PT, FHA, PRN, and FIM-2 were analyzed by ELISA according to time of serum collection after onset of illness in children with pertussis who were vaccine failures or who were previously unvaccinated. Antibody values were also compared by severity of clinical illness. In Sweden, infants who had received a PT toxin vaccine and who were vaccine failures had a blunted response to the nonvaccine antigen FHA compared with the response in children who had received a PT/FHA vaccine. Similarly, infants who had pertussis and who had received a PT/FHA vaccine had a blunted response to the nonvaccine antigens PRN and FIM 2/3 compared with the response in children who were vaccine failures and who had received a PT, FHA, PRN, and FIM 2/3 vaccine. In Germany, in sera collected from 0 to 15 days after pertussis illness onset, the GMVs for all 4 antigens (PT, FHA, PRN, and FIM-2) were significantly lower in an unvaccinated group than in children who were diphtheria-tetanus-acellular pertussis (DTaP) vaccine failures. In the unvaccinated group, the GMV of the PT antibody rose rapidly over time so that it was similar to that of the DTaP vaccine recipients at the 16- to 30-day period. In contrast, the antibody responses to FHA, PRN, and FIM-2 at all time periods were lower in the diphtheria-tetanus vaccine (DT) recipients than in the DTaP vaccine failures. In both Sweden and Germany, children with less severe illness had lower antibody responses than children with typical pertussis. Our findings indicate that upon exposure and infection, previous vaccinees have more-robust antibody responses to the antigens contained in the vaccine they had received than to *Bordetella* antigens that were not in the vaccine they had received. In addition, over time the antibody responses to FHA, PRN, and FIM-2 were greater in children with vaccine failure (primed subjects) than in unvaccinated children (unprimed subjects) whereas the responses to PT were similar in the primed and unprimed children, as determined from sera collected after 15 days of illness. Our findings lend support to the idea that DTaP vaccines should contain multiple antigens.

In a previous study, it was observed that children who were diphtheria-tetanus-acellular pertussis (DTaP) vaccine failures had a minimal antibody response to the nonvaccine antigen adenylate cyclase toxin (ACT), whereas unvaccinated children had a vigorous response to this antigen (4). Specifically, the convalescent-phase enzyme-linked immunosorbent assay (ELISA) antibody geometric mean value (GMV) in response to ACT in 20 unvaccinated children with pertussis was 872 ELISA units (EU)/ml whereas the convalescent-phase GMV in 10 DTaP vaccine failures was only 49 EU/ml. This observation of a blunted antibody response to a nonvaccine antigen in children who were DTaP vaccine failures led us to do a broader retrospective study of patterns of antibody responses to vaccine and nonvaccine antigens in children who were vaccine failures in two vaccine efficacy trials in Sweden (1, 5–9, 22, 24). The initial analysis of data from these two trials led us to do further retrospective analyses of antibody response patterns in diphtheria-tetanus-pertussis (DTP) and DTaP vaccine failures (primed subjects) and in diphtheria-tetanus vaccine (DT) recipients (unprimed subjects). We have examined the convalescent-phase GMVs at various times from illness onset in children in a DTaP vaccine efficacy trial in Germany, and we also have examined convalescent-phase GMVs in the German trial and one of the Swedish trials by severity of pertussis illness in vaccine failures and in unvaccinated children (DT recipients) (5–9, 22, 24).

In addition to the studies presented here, ELISA results for the long-term kinetics of antibodies to pertussis toxin (PT) and fimbriae (FIM 2/3) following infection and vaccination in Swedish children have recently been presented (7, 8).
MATERIALS AND METHODS

Studies done in Sweden. Convalescent-phase serum samples were obtained from 107 children with pertussis who were participants in a vaccine efficacy trial done in Sweden in the 1980s (1). There were 25 children who had received an acellular pertussis (aP) vaccine containing PT toxoid and filamentous hemagglutinin (FHA) (JNIH-6), 33 children who had received an aP vaccine containing PT toxoid (JNIH-7), and 49 children who had received a placebo. Antibody responses to PT and FHA were measured by ELISA (23). JNIH-6 contained 3.8 μg protein nitrogen each of the PT antigen and the FHA antigen per dose. JNIH-7 contained 6.0 μg protein nitrogen of PT antigen per dose. Eight-month-old infants received 2 doses of vaccine 2 to 3 months apart. Postvaccination GMVs to 2 components (PT and FHA) DTaP vaccine (DTaP-2), 13 children who had received a 5-component (PT, FHA, pertactin [PRN], and FIM 2/3) DTaP vaccine (DTaP-5), and 33 children who had received a diphtheria–tetanus–whole-cell pertussis vaccine (DTPw). DTaP-2 contained 25 μg (each) PT and FHA antigens per dose. DTaP-5 contained 10 μg PT, 5 μg FHA, 5 μg PRN 2/3, and 3 μg PRN per dose. DTPw contained 5.7 protective units per dose. Infants received 3 doses of a vaccine, at 2, 4, and 6 months of age. The ELISA antibody data collected 1 month after the third dose are expressed as reversed distribution curves and not mean values (6). The 50% frequency values for antibodies against the PT antigen were 40 EU/ml for DTwP; and those for antibodies against the FIM 2/3 antigen were 100 EU/ml for FHA. The PRN antigen were 1000 EU/ml for FHA and 164 EU/ml for PT and <2 EU/ml for FHA for the JNIH-6 group and ≤164 EU/ml for PT and <2 EU/ml for FHA for the JNIH-7 group.

In a second pertussis vaccine efficacy trial, done in Sweden in the 1990s, convalescent-phase serum samples were obtained from 137 children in a household-contact substudy who had laboratory-confirmed pertussis (6, 24). There were 53 children who had received DT, 38 children who had received a 2-component (PT and FHA) DTwP vaccine (DTwP-2), 13 children who had received a 5-component (PT, FHA, pertactin [PRN], and FIM 2/3) DTwP vaccine (DTwP-5), and 33 children who had received a diphtheria–tetanus–whole-cell pertussis vaccine (DTPw). DTwP-2 contained 25 μg (each) PT and FHA antigens per dose. DTwP-5 contained 10 μg PT, 5 μg FHA, 5 μg PRN 2/3, and 3 μg PRN per dose. DTPw contained 5.7 protective units per dose. Infants received 3 doses of a vaccine, at 2, 4, and 6 months of age. The ELISA antibody data collected 1 month after the third dose are expressed as reversed distribution curves and not mean values (6). The 50% frequency values for antibodies against the PT antigen were 40 EU/ml for DTwP, 100 EU/ml for FHA, 50 EU/ml for PRN, and <1 EU/ml for DTPw; those for antibodies against the FHA antigen were ~200 EU/ml for DTwP-2, ~40 EU/ml for DTwP-5, and ~10 EU/ml for DTPw; those for antibodies against the PRN antigen were ~1 EU/ml for DTwP-2, ~100 EU/ml for DTwP-5, and ~60 EU/ml for DTPw; those for antibodies against the FIM 2/3 antigen were ~1 EU/ml for DTwP-2, ~400 EU/ml for DTwP-5, and ~30 EU/ml for DTPw. The median time interval from cough onset to collection of the convalescent-phase serum sample was 73 days. This time interval did not vary significantly among the four vaccine groups. Antibody titers to PT, FHA, PRN, and FIM 2/3 were measured by ELISA.

In the first Swedish trial, convalescent-phase IgG GMVs against PT and FHA were examined by vaccine group (JNIH-6, JNIH-7, and placebo). In the second trial, convalescent-phase IgG GMVs against PT, FHA, PRN, and FIM 2/3 were examined by vaccine group (DTwP, DTwP-2, DTwP-5, and DTPw). In addition, in the second Swedish trial the GMVs against each antigen by vaccine group were analyzed by illness severity. The severity of illness is one DTAp2 vaccine was not available. "Typical" illness was determined according to the World Health Organization (WHO) case definition (established by an ad hoc committee), which required at least 21 consecutive days of paroxysmal cough and a positive culture for Bordetella pertussis, a significant increase in IgA or IgG antibody to PT, FHA, or PRN, or a culture-confirmed household contact (2b). Less severe cases included all children with laboratory confirmation or a culture-confirmed household contact but with less severe cough illnesses.

ELISA antibody values in the 2 Swedish studies should not be compared because the studies were done during different time periods, using different sources of antigens and different reference sera. Similarly, antibody data between the Swedish and German studies should not be compared. All ELISAs were performed at the Swedish Institute for Infectious Disease Control, Stockholm, Sweden, by using the reference line method (6, 18, 23, 24). All cases were laboratory confirmed and had at least 1 day of cough illness.

The 1980s trial was approved by the ethics committees at the Karolinska Institute in Stockholm. The JNIH-6 group was approved by the Ethics Committee at the Karolinska Institute in Stockholm. For the 1990s trial, informed consent was obtained from the parents of all participants (6). The trial was approved by the Ethics Committee at the Karolinska Institute in Stockholm.

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RESULTS

Studied done in Sweden. The GMVs of IgG antibody to PT and FHA in convalescent-phase sera from children with pertussis who were previous recipients of an aP vaccine containing PT and FHA (JNIH-6), an aP vaccine containing PT (JNIH-7), or a placebo who had pertussis due to B. pertussis infection formed consent was obtained from the parents of all participants (6). The trial was approved by the Ethics Committee at the Karolinska Institute in Stockholm.

Studies done in Germany. From May 1991 to December 1994, a controlled vaccine efficacy trial that included a DTaP vaccine (which contained PT toxoid, FHA, PRN, and FIM-2), DTwP, and a DT control group was carried out (5, 9, 22). Infants in this trial received 4 doses of DTaP vaccine, at 3, 4½, 6, and 15 months of age. The DTaP vaccine contained 3.2 μg PT, 34.4 μg FHA, 1.6 μg PRN, and 0.8 μg FIM-2. Antibody GMVs after the third dose were 13.4 EU/ml for PT, 58.1 EU/ml for FHA, 69.1 EU/ml for PRN, and 8.0 EU/ml for FIM-2. In this trial, there were 238 children with laboratory-confirmed B. pertussis illnesses. Of this group, 84 had a case definition consistent with the WHO criteria (26) and 154 had less severe respiratory illness (≥7 days of cough), with the same laboratory and household-contact criteria. For each pertussis group (DT, DTaP, and DTwP), GMVs of IgG antibody to PT, FHA, PRN, and FIM-2 were compared between cases consistent with the WHO definition and those consistent with the definition of less severe illness.

Of the 238 children with pertussis, 231 had “acute-phase” sera available. Of this group, we compared the GMVs for the 4 antigens during four time periods from illness onset between cases in the DT group and the DTaP vaccine group. We could not compare the DT group with the DTwP group because the DTwP group did not contain enough cases. Antibody values were determined by ELISA performed at Wyeth-Lederle Vaccines and Pediatrics by a modification of the parallel line method (2, 3, 9, 10, 13, 15, 22). All cases were laboratory confirmed and had at least 7 days of cough illness.

This study was approved by the ethics committee of the University of Erlangen (Erlangen, Germany) (9, 22).

Statistics. Geometric mean values (GMVs) were determined using logarithmic-transformed data, and all values are expressed as GMV in EU/ml or as log GMV. GMVs were estimated and compared using a 2-way analysis of variance (ANOVA) model, with vaccine group and time interval as fixed effects. In one comparison, the paired t test was used.

| TABLE 1. GMVs of IgG antibody to PT and FHA in convalescent-phase sera from previous recipients of JNIH-6, JNIH-7, or placebo who had pertussis due to B. pertussis infection |
|---|---|---|
| Group (no. of subjects) | GMV, EU/ml (95% CI)* | PT | FHA |
| JNIH-6 (25) | 107.2 (74.7–153.8) | 87.1 (60.7–125.0)** | 129.0 (8.9–18.6)** |
| JNIH-7 (33) | 131.8 (90.6–191.9) | 74.0 (14.7–32.5)** |
| Placebo (49) | 195.0 (137.2–276.9) |

*a 95% CI, 95% confidence interval.
** JNIH-6 versus JNIH-7, P < 0.0001.
*** JNIH-7 versus placebo, P = 0.23.
convalescent-phase sera from children with pertussis who were previous recipients of DTaP-2, DTaP-5, DTwP, or DT in the second Swedish trial are presented in Fig. 1. As noted, the DTaP-5 and DTwP vaccine failures had vigorous ELISA antibody responses to all four vaccine antigens (PT, FHA, PRN, and FIM 2/3). In contrast, the DTaP-2 vaccine failures had similar vigorous responses to only the two vaccine antigens (PT and FHA) for which they were primed. The controls (DT recipients) had mod-

### FIG. 1. Convalescent-phase GMVs of IgG antibody to PT, FHA, PRN, and FIM 2/3 in 13 DTaP-5, 38 DTaP-2, and 33 DTwP vaccine failures and in 53 controls (DT recipients) in the second Swedish study.

| Vaccine group | p values vs vaccine group | p value |
|---------------|---------------------------|---------|
| DTaP-2        | DTwP                      | <.0001  |
| DTaP-2        | DT                        | 0.0002  |
| DTwP          | DTaP-5                    | <.0001  |
| DTwP          | DTaP-5                    | 0.0879  |
| DT            | DTaP-5                    | <.0001  |

| Vaccine group | p values vs vaccine group | p value |
|---------------|---------------------------|---------|
| DTaP-2        | DTwP                      | <.0001  |
| DTaP-2        | DT                        | 0.0002  |
| DTaP-2        | DTaP-5                    | <.0001  |
| DTwP          | DT                        | <.0001  |
| DTwP          | DTaP-5                    | 0.0879  |
| DT            | DTaP-5                    | <.0001  |

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**A.**

**B.**

**C.**

**D.**
It is of interest to note that the antibody response to PT in the unprimed DT vaccinees was significantly greater than that in the DTaP-2 or DTaP-5 vaccine failures (\( P < 0.0001 \)).

Also of note is the antibody response to FHA in the unprimed group (DT recipients) compared to the responses in the three primed groups (DTaP-2, DTwP, and DTaP-5 recipients). The vaccine failures who were primed with DTaP-2 or DTwP had significantly greater GMVs of antibody to FHA than the unprimed group (DT recipients); however, the FHA antibody responses were similar in the DTaP-5 vaccine failures and the placebo recipients (DT vaccinees).

Presented in Table 2 are IgG GMVs against the 4 \( B. pertussis \) antigens, assorted by vaccine group and analyzed by illness severity. For DT, DTaP-2, and DTaP-5, the GMVs in the typical cases tend to be higher than the GMVs in the children with less severe illness. However, in only 2 of the 12 comparisons was a \( P \) value of \( \leq 0.5 \) reached. In the DTwP group, the GMVs appear to be lower in the infants with typical pertussis than in those with less severe illness (\( P > 0.05 \) in all comparisons).

**Studies done in Germany.** Presented in Fig. 2 are the GMVs of antibody to PT, FHA, PRN, and FIM-2 during 4 time periods from illness onset in DTaP vaccine failures and in nonvaccinated children (DT recipients).

For sera collected from 0 to 15 days after illness onset, the GMVs for all four antigens were significantly lower in the unvaccinated group than in children who had received the DTaP

| Vaccine group | Antigen | GMV, EU/ml | \( P \) value |
|---------------|---------|------------|--------------|
| DT            | PT      | 311.3      | 0.53         |
|               | FHA     | 58.9       | 0.18         |
|               | PRN     | 2.6        | 0.16         |
|               | FIM 2/3 | 6.2        | 0.71         |
| DTaP-2        | PT      | 74.7       | 0.06         |
|               | FHA     | 298.1      | 0.28         |
|               | PRN     | 0.6        | 0.02         |
|               | FIM 2/3 | 2.5        | 0.95         |
| DTaP-5        | PT      | 76.8       | 0.06         |
|               | FHA     | 84.1       | 0.22         |
|               | PRN     | 135.7      | 0.05         |
|               | FIM 2/3 | 304.9      | 0.15         |
| DTwP          | PT      | 313.0      | 0.94         |
|               | FHA     | 294.8      | 0.10         |
|               | PRN     | 103.8      | 0.56         |
|               | FIM 2/3 | 228.3      | 0.24         |

\( a \) Typical, criteria established by an \( ad hoc \) committee from WHO, which required at least 21 consecutive days of paroxysmal cough and a positive culture for \( B. pertussis \), a significant increase in IgA or IgG antibody to PT, FHA, or FIM, or a culture-confirmed household contact (26); less severe, laboratory confirmation or culture-confirmed household contact but with cough illness that did not include 21 consecutive days of paroxysmal cough. For the DT group, 12 subjects had less severe symptoms and 41 had typical symptoms; for the DTaP-2 group, 15 subjects had less severe symptoms and 22 had typical symptoms; for the DTaP-5 group, 6 subjects had less severe symptoms and 7 had typical symptoms; and for the DTwP group, 10 subjects had less severe symptoms and 23 had typical symptoms.
vaccine. In the unvaccinated group, the GMV of the PT antibody rose rapidly over time so that it was similar to that for the DTaP vaccine recipients at the 16- to 30-day period (32.1 versus 36.8 EU/ml, respectively [P = 0.8]), and at the 31- to 50-day and >50-day periods it appears higher than the GMV of the DTaP vaccine group (92.2 versus 53.9 EU/ml [P = 0.5] and 106.3 versus 45.8 EU/ml [P = 0.2], respectively). The FHA antibody pattern rise was less marked than that of PT in the unvaccinated subjects, and its peak was lower than that for the DTaP vaccinees. With that for the DTaP vaccinees.

**DISCUSSION**

In a previous study, it was observed that children who were DTaP vaccine failures had a blunted antibody response to the nonvaccine antigen ACT, whereas unvaccinated children with pertussis had a vigorous antibody response to this antigen (4). This observation led us to perform an additional retrospective study in Sweden and two retrospective studies of German children who participated in a DTaP vaccine efficacy trial (5, 6, 9, 22).

The results of the two retrospective analyses in Sweden indicated that children who were vaccine failures had brisk antibody responses to antigens present in the vaccines they had previously received, whereas their antibody responses to nonvaccine antigens were minimal. In the first Swedish trial, there was a suggestion that the antibody response to the nonvaccine antigen FHA in vaccine failures was lower than the response to the same antigen in unvaccinated controls with pertussis. In the second Swedish trial, the antibody responses to the two nonvaccine antigens PRN and FIM 2/3 in vaccine failures were significantly lower than the responses to these antigens in the children with pertussis who were DT recipients (controls).

The blunted antibody response to nonvaccine antigens in the children with vaccine failure might be explained simply by the effect of priming versus nonpriming (11). However, this does not explain the fact that the response to the nonvaccine antigens in the vaccine failures was lower than that which occurred in previously unvaccinated children with pertussis. There are perhaps two mechanisms which could explain the minimal response to the nonvaccine antigens in the children who were vaccine failures.

One explanation might be that illness in vaccine failures tends to be less severe, which might result in a lowered antibody response. To shed light on this hypothesis, we carried out the two analyses presented in Tables 2 and 3. Although the findings in both data sets suggest that more severe illness is associated with a more vigorous antibody response, the differences overall are not striking.

A second hypothesis, which we favor, is that the findings can be explained by linked epitope suppression caused by preferential responses of memory B cells following secondary exposure to vaccine components (16). Memory B cells and circulating antibodies are readily available to respond to additional exposures. They outcompete naïve B cells for access to the *Bordetella* epitopes, as they are more numerous and their receptors exhibit a higher antigen affinity. Higher expression levels of cell surface major histocompatibility complex class II (MHC-II) and costimulatory molecules also allow memory B cells to preferentially interact with T helper cells required for further propagation of the immune response. Linked epitope suppression applies as the immune response to the new epitopes is suppressed by the strong response to the original vaccine components.

We believe that the blunted antibody response that we have demonstrated is an important consideration in the choice of DTaP vaccines for general use and for the development of new DTaP vaccines. For example, as data from our two previous studies (5, 24) of serologic correlates of immunity indicate, children who received only a PT toxoid or PT/FHA vaccine have enhanced and continued susceptibility to *B. pertussis* infections compared to that of children immunized with multi-component vaccines. As modest PRN antibody values are most important for exposed children, failure to include PRN in vaccines likely results in subsequent suppression of PRN responses. The original exposure essentially “locks in” the immune response to certain epitopes and inhibits the response to linked epitopes even following subsequent exposures. The only
way to break this pattern is to expose the individual to new epitopes that are unlinked to the epitopes to which the individual had been exposed in prior encounters. Thus, if immunization against additional epitopes is desired, these epitopes have to be introduced separately in a vaccine to take effect, because in combination vaccines, the generation of antibodies against them is always suppressed due to the preferential response to previously introduced antigens.

Because the antibody responses to PRN and FIM 2/3 in primary infections in controls (DT recipients) were modest in the second study in Sweden, we performed the time-related-response study of the children in the German efficacy trial; we thought that the inferior responses in the controls might be due to the time of collection of the convalescent-phase serum samples. However, this does not appear to be the case (Fig. 2). Of interest are the differences in the patterns of antibody response to PT in the vaccine failures and controls compared with the patterns of response to the other antigens. In the >50-day time period, the GMV response to PT in the control group was similar to or perhaps higher than that in the vaccine failures (106.3 versus 45.8 EU/ml, respectively [P = 0.2]). In a study in Senegal, Simon et al. (21) noted similar findings with children who were vaccine failures or who were unvaccinated. In contrast, in our study of the >50-day period, the responses to FHA, PRN, and FIM-2 were significantly lower in the controls than in the vaccine failures.

The DTaP vaccine failure group has a memory response against specific Bordetella antigens, while the DT group is considered naïve. Memory responses are characterized primarily as faster, not necessarily higher, at all time points (11). Therefore, we expect the antibody responses to these Bordetella antigens to initially be higher in the DTaP vaccine group than in the DT group but for the DT group’s antibody values to eventually catch up. This result occurred with antibody to PT but not with antibody to FHA, PRN, or FIM-2. This difference in the response to PT compared with the responses to FHA, PRN, and FIM-2 might be explained by the uniqueness of PT. Specifically, PT is unique in nature, with no homologues, whereas the other antigens have many homologues in other bacterial species which infect children (14, 25).

Therefore, the DT group could have prior exposure to other bacteria that express FHA-, PRN-, and FIM-like proteins that could bias the immune response away from the B. pertussis detectable form of each antigen. This again could be a manifestation of linked epitope suppression. Specifically, the children would have a major antibody response to the proteins like FHA, PRN, and FIM from other bacteria and a lower response to PT in the vaccine failures and controls compared with the patterns of response to the other antigens. In five trials DTaP vaccines were more efficacious than DTaP vaccines (9, 12, 17, 19, 20, 22), we should reexamine the antibody responses of DTaP recipients to other antigens (such as lipopolysaccharide [LPS], BrkA, SphB1, Vag8, Bsp22, and TcfA) and explore whether one or more of these antigens as well as ACT should be added to DTaP vaccines.

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