Research Article

Biogeographic Patterns of Finnish Crane Flies (Diptera, Tipuloidea)

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Species richness of terrestrial and freshwater biota generally decreases with increasing latitude. Some taxa, however, show an anomalous species richness pattern in a regional or global scale. The aim of this study was to examine (i) regional variation in species richness, (ii) faunistic composition, (iii) occupancy, and (iv) proportions of different distribution types of Finnish crane flies. Analyses were based on incidence data pooled into 20 biogeographical provinces. Finnish crane fly fauna consists of 335 species; the provincial richness varies from 91 to 237. The species richness of all species and saproxylic/fungivorous species decreased with increasing latitude; mire-dwelling crane flies displayed a reversed pattern (Spearman’s correlations). Thirty-one species occupied a single province and 11 species were present in all provinces. Provincial assemblages showed a strong latitudinal gradient (NMS ordination) and faunistic distance increased with increasing geographical distance (Mantel test). Nearly half (48%) of the Finnish crane flies are Trans-Palaearctic, roughly one-third (34%) are West Palaearctic, and only 16 and 2% are Holarctic and Fennoscandian, respectively. Endemic Fennoscandian species are discussed in detail; most likely there are no true endemic crane flies in this region.

1. Introduction

1.1. Species Richness Gradient. In general, species richness decreases with increasing latitude. The tropics harbor far more animal and plant species than temperate or arctic zones (e.g., [1–3]). The most probable explanations for this pattern are related to productivity and biomass that are determined by the amount of available energy (sun light) and evapotranspiration (moisture) [4, 5]. Moreover, historical factors such as glaciations have shaped local flora and fauna: retreating and advancing glaciers during the Pleistocene totally eradicated fauna and flora from most high latitude areas, while the tropics probably experienced less severe climatic stress [6]. In addition, the tropics have the largest geographical land area, and larger areas invariably support more species than smaller areas [7, 8]. In smaller spatial scales, isolation, interspecific interactions, disturbance, and environmental heterogeneity also influence species richness (e.g., [9]).

Despite the preponderance of the general trend, some taxa show a reversed latitudinal pattern in species richness. For example, sawflies [10] and aphids [11] are richer in species in the north boreal and temperate zones than in the tropics. Within northwestern Europe (Fennoscandia, that is, Nordic countries, Russian Karelia, and Kola peninsula), species richness of stone flies [12, 13], waders [14], and mire-dwelling bird communities [15, 16] increases with increasing latitude. It is hypothesized that availability of resources best explains the reversed patterns. For instance, environmental complexity, total area of flark fens, and abundance of invertebrate food explain the high species richness of mire-dwelling birds and waders in north Fennoscandia [14, 16]. In a similar vein, exceptional diversity and abundance of Salix species, the most important food plant for sawfly larvae, account for the reversed pattern of sawfly richness [10, 17]. Parasitoid ichneumonid wasps have long been assumed to have the highest species richness in midlatitudes (e.g., [18, 19]), but this notion is likely an artifact due to the poor sampling and premature taxonomy of the superfamily [20–22].

1.2. Biogeography of Finland. Finland is geographically part of Fennoscandia, being located in northernmost Europe. Current Fennoscandian biota in general, and Finnish in
Table 1: Finnish biogeographical provinces, their abbreviations and full names, locations, sizes, and number of Malaise trapping sites during the years 2000–2011.

| Abbr. | Provinces                     | Name               | Coordinates¹ | Size² | Malaise |
|-------|-------------------------------|--------------------|---------------|-------|---------|
| Al    | Alandia                       | 6698 3107          | 1.5           | 19    |
| Ab    | Regio aboensis                | 6714 3278          | 11.5          | 41    |
| N     | Nylandia                      | 6698 3385          | 8             | 16    |
| Ka    | Karelia australis             | 6727 3505          | 3             | 0     |
| St    | Satakunta                     | 6841 3262          | 14.5          | 25    |
| Ta    | Tavastia australis            | 6849 3357          | 25.5          | 24    |
| Sa    | Savonia australis             | 6846 3547          | 22.5          | 2     |
| Oa    | Ostrostettia australis        | 6995 3267          | 14            | 35    |
| Tb    | Tavastia borealis             | 6946 3410          | 19            | 28    |
| Sb    | Savonia borealis              | 6983 3531          | 21            | 24    |
| Kb    | Karelia borealis              | 6990 3642          | 24            | 28    |
| Om    | Ostrostettia media            | 7186 3404          | 25            | 3     |
| Ok    | Ostrostettia kajanense        | 7133 3570          | 24.5          | 12    |
| Oba   | Ostrostettia borealis pars australis | 7244 3477 | 15            | 32    |
| Obb   | Ostrostettia borealis pars borealis | 7388 3412 | 24.5          | 22    |
| Ks    | Regio kuusamoensis            | 7327 3598          | 18            | 63    |
| Lkoc  | Lapponia kemensis pars occidentalis | 7514 3426 | 13            | 30    |
| Lkor  | Lapponia kemensis pars orientalis | 7532 3534 | 21            | 10    |
| Le    | Lapponia enontekiensis        | 7636 3299          | 8.5           | 34    |
| Li    | Lapponia inariensis           | 7708 3518          | 23            | 28    |

¹ National coordinate system grid 27° E; N coordinate refers to the distance from the equator (in km). Coordinates are given with an accuracy of one km and refer to the midpoints of the provinces. ² ×1000 km².

particular, is strongly influenced by the legacy of Pleistocene glaciations [23]. The latest glacial maximum (Weichselian) took place about 18 000 years ago and only a small part of Fennoscandia was free of ice some 11 000 years ago [6, 23]. Based on the distribution of plant taxa [24], proportions of short-winged carabids (Coleoptera) [25, 26], and fossil evidence [27], it has been suggested that the highest mountain tops (nunataks) and islands along the Norwegian coast were free of ice and some taxa were able to “overwinter” there during the Weichselian maximum. However, within Finnish borders, all terrestrial biota must have colonized the area during the last circa 10 000 years. Because of this recent origin of Finnish biota, endemic species (i.e., taxa only present in Fennoscandia and nowhere else) are exceptions (e.g., [28]). In general, Finnish biota could perhaps be classified into three different colonization groups. Firstly, European, or West Palaearctic, species currently present in Finland are descendants of populations from separated glacial refugia [29–31]. Secondly, in addition to southern biota, Finnish fauna has a strong taiga element, that is, boreal species of eastern origin, and thirdly, an arctic element, that is, circumpolar species (e.g., [32–35]).

Finland is part of the Holarctic biome called the boreal zone, that is, a belt of coniferous forests. Southernmost parts of the country are hemiboreal, with mixed broad-leaved and coniferous stands, whilst northermost Finland is characterized by mountain birch (*Betula pubescens* ssp. *czerepanovii*) forests and treeless fells. Because of this latitudinal variation, Finland is an optimal region for biogeographic studies. In addition, such large-scale studies are made possible by the long faunistic and floristic tradition practiced there. For over 100 years, Finnish fauna and flora have been mapped according to 21 biogeographical provinces (e.g., [36–38], Table 1, Figure 1). Despite the fact that the boundaries of the provinces follow historical and/or current Finnish political municipalities, occurrence data mapped as such has been successfully analyzed in several biogeographic studies (e.g., [13, 39]). One of the advantages of provincial approach to biodiversity studies is that old data from the literature or museum specimens with inaccurate labeling can be taken into account. It allows one to examine large-scale trends in species richness and distribution of taxa, but finer scale variation will of course be hidden. To conclude, the composition and species richness of Finnish fauna and flora are mainly driven by latitudinal variation and local environmental factors [39–44]. The impact of latitude is not surprising, given the long, over 1100 km, gradient from south to north. To some extent, plant assemblages reflect increase of continentality from western to northeastern Fennoscandia [45, 46], but among insects longitude is a poor biogeographic predictor in NW Europe [39].

1.3. Crane Flies (Diptera, Tipuloidea), Finnish Fauna Emphasized. Crane flies are very speciose and ecologically diverse. Over 15 000 valid species are currently known, 3175 of these are from the Palaearctic region [47]. Most crane fly
larvae prefer moist environments and eat detritus or prey upon other invertebrates [46]. There are also saproxylic, fungivorous, and herbivorous crane flies [48–50]. Adults are mainly short lived and have nonbiting mouthparts. Globally, the highest species richness of crane flies is encountered in the tropics and perhaps a notable number of tropical tipuloids still await their description. However, crane flies, based on data pooled into biogeographical provinces. (such as Molophilus bihamatus, Prionocera turcica), but they are generally core species of peatlands. Subjective assessment of the bulk of this data is extracted from one of the largest Malaise trap samples ever collected (see Section 2). I also studied proportions of four main distribution types (Trans-Palaearctic, West Palaearctic, Holarctic, Pannonian) within ecoregions (hemi, and south boreal, middle boreal, and north boreal). Species richness patterns were examined for all species and for two ecological groups, mire-dwellers and saproxylic/fungivorous species.

2. Material and Methods

2.1. Study Area and Data Sets. Finland is located between 59°30′N and 70°05′N and 19°07′E and 31°35′E, being part of the boreal zone, that is, zone of coniferous forests. Finland is divided into four major ecoregions or vegetation zones, namely, (from south to north) hemiboreal, south boreal, middle boreal, and north boreal and these zones are further divided into subzones (Figure 1). This zonation is mainly controlled by climate (e.g., decreasing mean annual temperature towards the north, differences in the length of the growing season, duration of snow cover, and continentality in a northeast-southwest gradient) and also topographic relief. Differences in vegetation structure between neighboring zones are not clear-cut but gradual changes take place along a latitudinal gradient (e.g., change of mire massif types from peat bogs to aapamires across the border of the southern and middle boreal regions). Finnish bedrock is mainly composed of acidic siliceous rocks, intermediate (e.g., mica schist, amphibolite) or calcareous (marbles, dolomite) rocks are generally rare. For further information, see for example [45, 64, 65].

A traditional way to map species’ occurrences in Finland is to use biogeographical provinces (see, e.g., [37], Figure 1). There are a total of 21 such provinces, their surface areas range from 1500 to 25 500 km² (Table 1). In the present study, the very small province Kl (Karelia ladogensis) was merged to Sa (full names of the provinces are given in Table 1). Boundaries of the provinces mainly follow borderlines of Finnish municipalities and are thus administrative in nature. Because the provinces were unequal in size, the number of species in each province was corrected using formula:

\[ S_{cor} = \frac{S_{obs}}{A^2} \]

where \( S_{cor} \) is the corrected number of species, \( S_{obs} \) the observed number of species in a province, \( A \) is the area of a province, and \( z \) is a constant taken from the species-area relationship (0.15 was chosen, see [13, 40]).

Observed and corrected provincial species numbers were calculated for (i) all species, (ii) mire-dwelling species, and (iii) saproxylic/fungivorous species. Ombrotrophic bogs and poor-rich fens, mostly open or sparsely wooded, are the principal habitats for the mire-dwelling species as defined here. Spring-dwelling taxa were neglected, although most springs fall to mire types in Finnish mire ecology [65]. Some of the mire dwellers may also occur in swampy lake shores (such as Molophilus bihamatus, Prionocera turcica), but they are generally core species of peatlands. Subjective assessment
is unavoidable here, but it is based on careful consideration by the author. Less subjective is the classification of crane flies into saproxylic/fungivorous species. Some genera are strictly fungivorous (e.g., Metalimmobia, Ula) or dependent on decaying wood (e.g., Gnaphomyia, Lipsothrix) [66–68]. Classification of some species as saproxylic is based on the author's personal observations (Limononia badia, Tipula pseudorororata, and T. stenostyla). Because some species are both saproxylic and fungivorous (i.e., larvae feeding on wood-decaying polyporous fungi), their combination here is justified. A total of 51 species were classified as mire dwellers and 42 as saproxylic/fungivorous (Table 6).

The occurrence of crane flies in Finnish provinces mainly follows Salmela [56]. However, after that publication, three species have been recorded as new for the regional fauna (Dicranomyia klefbecki, Ormosia hederae, and Tipula paudl), one species was described as new for science (Tipula recondata, [69]) and some provincial occurrences were added and corrected. The Finnish list of crane flies now consists of 335 species, of which only one (Tipula peliostigma, doubtful species) lacks provincial data (see [56]).

Biogeographical provinces were further classified into three groups, roughly corresponding to ecoregions or vegetation zones: (i) hemiboreal and south boreal (Al, Ab, N, Ka, Ta, Tb, Sa, St, and Sb), (ii) middle boreal (Oa, Kh, Ok, Om, Oba, and Obb), and (iii) north boreal (Ks, Lkoc, Lkor, Le, Li). For each zone, (a) total species richness of crane flies, (b) number of species present in only one of the zones, (c) number of species present in all three zones, and (d) numbers of species representing four different distribution types were calculated. The distribution types used are Holarctic, Trans-Palaearctic, West Palaearctic, and Fennoscandian [47]. Holarctic species occur in both Nearctic and Palaearctic realms, Trans-Palaearctic species are recorded from both the eastern and the western part of the Palaearctic region, West Palaearctic species occur west of Ural mountains, and Fennoscandian species are not recorded outside Finland, Sweden, Norway, Kola Peninsula, and Russian Karelia. Further, numbers of crane flies that are absent from Central Europe (occurrence in the Baltic countries was allowed) were recorded for the three zones.

All available data dealing with the Tipuloidea fauna of Finland was compiled for the first time in 2006. Data from the literature, Finnish museum specimens, and the author's personal observations were entered into a database, which includes locality data and ecological information, if available. This database has since been updated and by the end of March 2012 it included 14 782 entries for the families Limoniidae, Tipulidae, Pediciidae, and Cylindrotomidae (entry = data from a museum specimen or an observation from a single locality). Between the years 2000 and 2011, crane flies have been collected quite intensively in Finland, especially by Malaise traps and sweep netting. Most of this material has been collected by me, but also material collected by several other persons was identified and tabulated. In total, 476 Malaise trapping sites, circa 1670 Malaise trapping months, form the core of this new material; this collecting effort yielded 101081 crane fly specimens and 301 species and is perhaps the largest Malaise trapping so far performed. One trap per locality was used in the majority of study sites, rarely three or more (maximum 15 traps in a study site). Malaise trapping was performed across a wide spatial scale, ranging from Aland Islands to the northernmost Finland (Table 1). Further, important material was collected from decaying trees using trunk-emergence traps (e.g., Halme et al. submitted ms). However, it must be stressed that some provinces are far better sampled than others (Table 1) and some habitats are rather well (headwater streams, and springs, northern aapamires), some rather poorly (meadows, shores of large rivers, and Baltic coastal meadows south of Oba) represented in the Malaise trapping material. In spite of that, collecting effort has not been substantially different in the south, middle, and north boreal zones and the variation in species richness and faunistic composition should reflect real phenomena, not artifacts due to differing sample sizes (see below for details).

2.2. Statistical Methods. The faunistic composition of provincial crane fly assemblages was examined using non-metric multidimensional scaling (NMS) ordination. NMS is an ordination method, in which the original ranked distances (based on distance measure) of the sample units in the p-dimensional species space are forced to a reduced, k-dimensional ordination [70, 71]. The Jaccard coefficient was used as a distance measure. Spearman's correlation coefficient was calculated between the ordination's coordinates of the provinces and latitudinal and longitudinal coordinates. McCune and Grace [71, pages 107-108] questioned whether it is appropriate to present P values in this connection because coordinate points of the sampling units along the dimensions are not independent variables. By calculating correlation, however, it is possible to interpret the geographical variation of provincial assemblages.

The Mantel test was used to examine the relationship between the faunistic dissimilarity and the geographical distance of the provinces. The Mantel test is used to test the null hypothesis of no relationship between two distance matrices, that is, the test evaluates linear correlation between two distance matrices. Each matrix is calculated from a different set of variables, measured for the same sample units (here provinces) [70, 71]. The test value rM is analogous to the Pearson correlation coefficient (range −1 and 1). Statistical significance is calculated by permutation (9000 permutations were used). The Jaccard coefficient was used as a distance measure for crane flies and Euclidean distance for geographic coordinates of the provinces.

Occupancy of crane flies in the provinces was calculated (i.e. number of species present in one, two, three,.., 20 provinces). No statistical fitting of occupancy frequency distribution was applied (e.g., [72]), and the shape of the distribution was assessed based on visual examination.

In order to analyze relationship with latitude and species richness (observed and corrected richness, see above), Spearman's (R_s) correlations were calculated. However, because observed species richness correlated positively with Malaise trapping effort (R_s = 0.54, P = 0.014), a partial correlation was also applied. This method can be defined as the correlation of the residuals after regression on the controlling
Table 2: Observed and corrected species richness of crane flies (Diptera, Tipuloidea) in Finnish biogeographical provinces for all species, mire-dwellers, and saproxylic/fungivorous species.

|         | All species | Mire species | Saproxylic species |
|---------|-------------|--------------|--------------------|
|         | Observed    | Corrected    | Observed           | Corrected    | Observed | Corrected |
| Al      | 117         | 110.1        | 19                 | 17.9         | 14       | 13.2      |
| Ab      | 237         | 164.3        | 30                 | 20.8         | 31       | 21.5      |
| N       | 204         | 149.3        | 22                 | 16.1         | 33       | 24.2      |
| Ka      | 95          | 80.6         | 10                 | 8.5          | 13       | 11.0      |
| St      | 152         | 101.8        | 18                 | 12.1         | 21       | 14.1      |
| Ta      | 230         | 141.5        | 30                 | 18.5         | 35       | 21.5      |
| Sa      | 140         | 87.8         | 13                 | 8.1          | 22       | 13.8      |
| Oa      | 162         | 109.0        | 26                 | 17.5         | 20       | 13.5      |
| Tb      | 188         | 120.9        | 32                 | 20.6         | 30       | 19.3      |
| Sb      | 176         | 111.5        | 29                 | 18.4         | 28       | 17.7      |
| Kb      | 168         | 104.3        | 25                 | 15.5         | 30       | 18.6      |
| Om      | 91          | 56.2         | 19                 | 11.7         | 10       | 6.2       |
| Ok      | 139         | 86.0         | 23                 | 14.2         | 26       | 16.1      |
| Oba     | 145         | 96.6         | 29                 | 19.3         | 16       | 10.7      |
| Obb     | 120         | 74.3         | 32                 | 19.8         | 16       | 9.9       |
| Ks      | 178         | 115.4        | 40                 | 25.9         | 21       | 13.6      |
| Lkoc    | 160         | 108.9        | 44                 | 29.9         | 22       | 15.0      |
| Lkor    | 95          | 60.2         | 32                 | 20.3         | 7        | 4.4       |
| Le      | 127         | 92.1         | 34                 | 24.7         | 10       | 7.3       |
| Li      | 136         | 85.0         | 34                 | 21.2         | 13       | 8.1       |

Table 3: Spearman’s correlations between provincial species richness (observed and corrected) and latitude (N coordinates of the provinces). Correlation coefficients and associated P values are given for all species, mire-dwelling species, and saproxylic/fungivorous species. Partial correlation coefficients and P values are given for observed species richness and latitude, controlling for per province Malaise trapping effort (see Table 1).

|            | Rs     | P      | Partial |                   |
|------------|--------|--------|---------|--------------------|
| All species| -0.352 | 0.128  | -0.628  | 0.004              |
|            | -0.521 | 0.018  |         |                    |
| Mire species| 0.705  | 0.001  | 0.731   | <0.001             |
|            | 0.557  | 0.011  |         |                    |
| Saproxylic species| -0.522 | 0.018  | -0.646  | 0.003              |
|            | -0.614 | 0.004  |         |                    |

3. Results

Species richness of all species is the highest in the southern provinces Ab, Ta, and N (237–204 spp., Table 2), and both observed and corrected species richness were in negative relation with latitude (Table 3). The correlation is statistically significant for corrected species richness and partial correlation between latitude and observed species richness (controlling for Malaise trapping effort) (Table 3). Saproxylic/fungivorous species richness also follows the same pattern as that of all species, but mire-dwellers species have a reversed species richness pattern, that is, increasing number of species with increasing latitude (Tables 2 and 3). A total of 31 species occupied a single province and 11 species were present in all provinces (Figure 2); mean occupancy per species was 9.2 (SD 5.9). The obtained occupancy frequency distribution (Figure 2) resembles unimodal with a satellite mode.

NMS ordination (Figure 3) indicated that provincial crane fly assemblages are separating along a latitudinal gradient. Distribution of provinces along the first NMS axis is positively correlated with latitude ($R_S = 0.904, P < 0.0001$), but neither the first nor the second axis is correlated with longitude ($R_S = 0.354–0.099, P = 0.125–0.677$). In parallel to the NMS ordination, the faunistic similarity of the provinces decreases with increasing geographic distance (Mantel test $M_R = 0.737, P < 0.001$, Figure 4). Pairwise
Almost half (48%) of the Finnish crane fly fauna is composed of wide-ranging, Trans-Palaearctic species (Table 4). Roughly one-third (34%) of the species are West Palaearctic and only 16 and 2% are Holarctic and Fennoscandian, respectively. Considering regional faunae (i.e., hemi- and south, middle, and north boreal), proportions of different distribution types remain roughly similar (Table 4). There is, however, a trend that the proportion of West Palaearctic species decreases from south to north. Correspondingly, the proportion of Holarctic and Trans-Palaearctic species increases from south to north (Table 4). The proportion of Fennoscandian species is low, roughly 1 or 2%, in each zone.

Poole species richness is the highest in the hemi- and south boreal zone (278 spp.), being lower, but of similar magnitude in the middle boreal (244) and north boreal (235) zones (Figure 5). Based on rarefied Malaise trapping data, ecoregions are ranked similarly as raw species richness (Figure 6(a)). Rarefied richness at the level of 101 trapping sites (the number of trap sites in the middle boreal ecoregion) is 220 (SD ± 4.9) for the hemi- and south boreal region, 195 (SD ± 4.8) and 186 (SD ± 4.2) for the middle and north boreal zones, respectively. However, standard deviations of the two latter zones are overlapping, indicating similar level of species richness (Figure 6(b)).

The hemi- and south boreal zone harbors 45 species that are only recorded there, only six species are restricted to the middle boreal zone, and 30 species occur only in the north boreal zone. A total of 170 species are far ranging, known from all boreal zones within Finland. 56 (17% out of total) Finnish crane fly species are absent from Central Europe and the number of such species clearly increases from the hemi- and south boreal (20 spp.) to the north boreal zone (47 spp.) (Figure 5).

4. Discussion

4.1. Regional Species Richness and Its Variation. In general, the species richness and assemblage composition of Finnish crane flies correlated strongly with latitude, a result that is highly concordant with studies on other insects [41, 44], plants [40], and birds [74]. The diversity gradients are not explained by latitude itself, but environmental variables correlated with it [3]. In Fennoscandian scale, those variables

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Table 4: Total numbers and proportions of different distribution types among crane flies (Diptera, Tipuloidea) in Finland and in the ecoregions (hemi and south boreal, middle boreal, and north boreal).

| Ecoregion       | FIN  | Hemi- and south bor. | Middle bor. | North bor. |
|-----------------|------|----------------------|-------------|------------|
| Holarctic (HOL) | 54 (16%) | 37 (13%)          | 40 (16%)    | 41 (17%)   |
| Trans-Palaearctic (Tr-PAL) | 161 (48%) | 137 (49%)        | 124 (51%)   | 124 (53%)  |
| West Palaearctic (W PAL) | 113 (34%) | 102 (37%)        | 77 (31%)    | 65 (28%)   |
| Fennoscandian (FENS) | 6 (2%)   | 2 (1%)             | 3 (1%)      | 5 (2%)     |

1Holarctic, 2Trans-Palaearctic, 3West Palaearctic, 4Fennoscandian.
are mostly associated with climate [75–77], for example, length of growing season and monthly mean temperatures. In addition to climate, historical and ecological factors may also account for the observed patterns [75, 78]. Within vascular plants, for example, north of latitude 64°N the species richness of plants ceases to decrease; a potential explanation is the ancient, postglacial colonization of the land area from both northern and southern species pools [75]. Colonization in general is faster in favorable climatic conditions and slower in the ecological tolerance limits of a species [75]. Hence, at least some species, initially expending their ranges either from the south or the north, are now in their climatic equilibrium. It still must be stressed that recent range extensions of Finnish crane flies are unknown, because of the lack of long-term monitoring.

Firstly, I found that richness of all species and saproxylic/fungivorous species decreased with increasing latitude. Most species rich provinces lie in southwestern Finland (Ab, Ta, and N), and most species poor in southern, central, and northern Finland (Ka, Om, and Lkor). With no doubt, this result is partly explained by unequal sampling efforts between the provinces: it is highly unlikely that neighboring provinces N and Ka differ almost 50% in their species richness. There are differences in the recent provincial sampling efforts (Table 1), but there are also historical differences. Although there has been modest and nonsystematic sampling prior to the 2000s in Finland [56], some regions have gained more faunistic interest than others. For example, the southwestern part of the province Ta was sampled by T. Brander and others in the 1960s [79, 80] yielded numerous new records for that province. The northeastern province Ks also received special attention [81], as did Li [82]. The southern provinces Ab and N are the most densely populated in Finland and have also harbored most professional and amateur entomologists during past decades; even occasional collecting by these persons has accumulated the number of observed species in south Finland (see, e.g., [83] for a bias in mapping data). However, regional differences in species richness are not solely explained by sampling artifacts. Local species richness is usually correlated with regional richness (type I relationship, [84]). That is, local assemblages embedded in species rich regions harbor more species than those in less diverse regions. Salmela [85] compared three sampling localities, which were considered the most species rich crane fly sites so far studied in southern and northern Finland. Those sites, headwater streams with similar sampling effort, were situated in the provinces N, Ta, and Li. Using raw data and individual-based rarefaction, it was noted that species richness was the highest in the south (N, Ta) and the lowest in the north (Li). Even though the data set was small, this may be used as implicit evidence of the general latitudinal gradient. In parallel, sample-based rarefaction of the nation-wide Malaise trapping data also indicated the decrease in species richness from south to north, although middle boreal and north boreal ecoregions seemed to harbor similar level of rarefied species richness.

Considering saproxylic and fungivorous species, the observed pattern was expected since the number of tree
species decreases toward the north (e.g., [45]). In northernmost Finland, large districts are treeless fells or mainly covered by one species, mountain birch, whereas there is a much richer assortment of deciduous and coniferous trees in southeastern Finland. It should be noted that some of the saproxylic crane flies, such as Elephantomyia edwardsi and Gnophomyia acheron, are confined to, or at least preferring, old-growth forests. Due to the human influence, such forests are rare especially in southern and western Finland, being isolated patches within a matrix of managed stands [64]. Reflecting this habitat degradation, many forest-dwelling beetles have vanished from southern and western parts of the country [86]. Furthermore, restoration success, measured as numbers of pyrophilous and saproxylic species after prescribed burning, differs between eastern and western Finland [87]. Thus, regional differences in the management history of Finnish forests have already affected species’ ranges, but there is no data to evaluate regional extinctions among forest-dwelling crane flies.

Secondly, even though a general decrease of provincial species numbers from south to north was observed, this decline is not very large. Pooling the provinces into three zones, roughly corresponding to the boreal ecoregions (Figure 4), an absolute difference of 43 species between hemi- and south boreal and north boreal zones was found. Furthermore, there is also a true turnover in the Finnish fauna; this is evidenced by the provincial assemblage variation (see below for further discussion) and also by the numbers of species restricted to respective zones. Over 40 species are truly southern, not occurring in the middle or north boreal zones, and 30 north boreal species display an opposite pattern. The middle boreal zone harbors few (6 spp.) species recorded only there, which implies that this zone is a mixture of northern and southern elements. In other words, there is true turnover, not just a gradient created by regional differences in alpha diversity (e.g., [88]).

Thirdly, there was a reversed latitudinal species richness pattern among mire-dwelling crane flies. A similar increase in species numbers has been observed among the avifauna of Finnish mires [16]. In general, Finland is a land of mires: about one-third of the land area was originally covered by peatlands [65]. Mires prevail in the Finnish landscape in central and northern Finland, especially in provinces such as Om, Oba, Lkoc, and Lkor [65]. Finnish mires can be roughly divided into two major types, ombrotrophic (raised bogs) and minerotrophic (aapamires, fens). The former are typical in the south boreal zone, the latter dominate in the middle and north boreal zones [65]. Compared to bogs, aapamires are rather heterogeneous in their surface patterns and vegetation. By definition, bogs receive only rain water and are thus poor in cations and characterized by low pH values. Aapamires vary from poor to rich fens and have diverse plant communities driven by pH and availability of nutrients [89]. Within aapamires, there is a tendency that shallow, inundated pools called flarks are more common in the north compared to the south [16, 65]. The large surface area of flark fens in the north provides plentiful insect food for waders and other insectivorous birds [15], partly explaining the reversed gradient among peatland birds. It is likely that sheer area effect (total surface area of mires) and environmental heterogeneity (amount of flark fens, presence of calcareous bedrocks) account for the high species richness of mire-dwelling crane flies in the northern provinces. As with the forests discussed above, human impact on Finnish mires is higher in the south than in the north. About 60% of Finnish mires have been drained in order to improve timber growth, and most of the pristine or lightly affected mires lie in the north boreal zone [65]. In a nationwide scale, the importance of this large-scale deterioration in habitat quality and diminishing area is practically unknown for crane flies.

Fourthly, occupancy of crane fly species in the provinces was dominated by rare species, and the obtained occupancy frequency distribution resembles unimodal with a satellite mode [72]. However, no statistical distribution was fitted to the observed one. If an ad hoc threshold value close to 25% (quartile) is applied here (see [90]), 104 Finnish crane fly species are known from four or fewer provinces (≤20%) and could be regarded as rare. On the other end, 52 species are common, known from 17 to 20 provinces.

4.2. Provincial Variation of Crane Fly Assemblages. Provincial crane fly assemblages were highly correlated with latitude. That is, crane fly assemblages differentiate along a south-north gradient. This result is in accordance with the result that faunistic similarity decreases with increasing geographic distance. Based on visual inspection of the NMS ordination (Figure 3), there are no clear clusters of provinces; rather, we see a continuum-like faunal differentiation along the latitudinal gradient. It must be noted that the distance measure used, 1-Jaccard coefficient, is based on incidence data only. The pair-wise provincial similarities take no information on abundances of species, which may have effects to the results obtained [91]. I hypothesize that differences between provinces would be greater if abundance data had been used. For example, Tipula excisa is among the most numerous species in northernmost Finland, in Li and Le (e.g., [61]), becoming a low-abundance species south of the subalpine fell district. The range of this species extends as far south as Oba (the record from Oa is dubious, [56]) and, despite differences in abundance, the species has the same weight in all pair-wise calculations. However, good abundance data is hard to achieve [92], and such data for crane flies is far beyond the horizon. Even though I have Malaise trapping data that is spatially extensive, this trapping method seriously underestimates abundances of certain species (J. Salmela, unpublished).

The notion that provincial assemblages are greatly affected by latitude is not surprising [39, 76]. As noted above (Sections 1 and 2), zonation of ecoregions is principally controlled by the climate. Along this climatic gradient habitats gradually change, partly explaining faunal differences. Perhaps more importantly, however, climate itself must play a crucial role as a determinant of species’ ranges, regulating, for example, respiration and life cycles of ectothermic animals (e.g., [93]). Among Fennoscandian insects, longitude is a poor predictor of species composition [39], but it explains floristic composition better [40, 46].
As evidenced by diversity change of species compositions, there seemingly are zones′ vegetation zones [13, 44]. However, despite the gradual of species composition is gradual, not strictly defined by the apices of the latitudinal gradient, but the transition invertebrate assemblages in Finland are rather di (incidence, not abundance) for such purposes. It seems that based on the occurrence of crane fly species, mainly due to their area of occupancy, that is, the number of grid cells occupied, and thus Trans-Palaearctic (and Holarctic) species may be anything from common to rare.

Western Palaearctic species total about one-third (34%) of the Finnish crane fly fauna, and the proportion of these species is clearly higher in the south (37%) than in the north (28%). It is very likely that species of European lowlands, that are mainly associated with deciduous broad-leaved forests [96], are in Fennoscandia restricted by habitat and climate to the southern areas. Nevertheless, the group of western Palaearctic species also encompasses crane flies that are boreo-alpine or boreo-montane [97], that is, species not associated with nemoral forests (e.g., Tipula pallidicosta and T. cinereocincta). Holarctic species, covering 16% of the fauna, are chiefly northern species with a circumpolar range (e.g., Arctoconopa forcipata, Prionocera ringdahlii). Some of the Holarctic species are “western arctic”, that is, only known from the Nearctic region and Fennoscandia (e.g., Dicranomyia intricata, D. moniliformis). However, as noted above, not too much weight should be given to the current faunistic knowledge of crane flies, since occurrences of species are still poorly mapped. In addition, there may still be taxonomic problems and some presently Holarctic species may actually consist of separate species in the Nearctic and Palaearctic realms, or vice versa. According to Mikkola et al. [32], truly Holarctic noctuid species are mainly arctic (inhabitants of tundra) and species living in the taiga (boreal forests) are mainly Nearctic/Palaearctic species pairs. This pattern is apparently caused by the legacy of the last glaciation (Beringian refugia of Holarctic species) and an earlier (6 mya) split of Holarctic forests [32].

4.3. Distribution Types of Finnish Crane Flies. Almost half of the Finnish crane flies are Trans-Palaearctic species. Proportion of these species is the highest in the north boreal zone (53%), the lowest in the hemi- and south boreal zone (49%). The difference, however, is not drastic. This group of Trans-Palaearctic species contains crane flies that are ubiquitous and wide spread, such as Nephrotoma scurra [95]. On the other hand, there are also species such as Tipula kaisilai and Tipula subexcisa that are rare, disjunct species known from northern Fennoscandia and the mountains of the eastern Palaearctic region [47]. In general, due to the rather poor knowledge of crane flies outside Europe, the designation of species here as wide ranging does not refer to their area of occupancy, that is, the number of grid cells occupied, and thus Trans-Palaearctic (and Holarctic) species may be anything from common to rare.

In this study, no significant correlations between ordination axes and longitude were found. However, in Fennoscandian scale, some species are indeed eastern taiga species and have not been found from Sweden or Norway (e.g., Phylidorea umbraeum, Gnophomyia acheron, and Tipula octomaculata). Western, or oceanic, crane fly species may be hard to be distinguished from southern species.

As discussed above, there is a turnover of crane fly species from south to north, not merely a decreasing number of species along this gradient. The observed patterns lead to the following predictions: (i) the composition of local communities is partly determined by latitude and (ii) local communities should be richer in species in the south than in the north (open bogs and fens showing opposite pattern). I refrain from delineating any biogeographic zones than in the north (open bogs and fens showing opposite pattern). I refrain from delineating any biogeographic zones.

| Occurrence | Taxonomy | True range |
|------------|----------|------------|
| Finland1, Sweden2 | OK | Palaearctic, boreal, and hemiarctic? |
| Limonia mesaurea m. | FIN, SWE | Palaearctic, boreal? |
| Dicranomyia lulensis | FIN, SWE | Arctic? |
| Rhabdomastix parva | FIN, SWE, NOR3, ICE4 | Parthenogenetic |
| Symplectia lindrothi | FIN, SWE | OK |
| Tipula fendleri | FIN | Perhaps a syn. of T. nigrolamina |
| Cylindrotoma borealis | FIN, NOR, RKar5 | Perhaps intraspecific variation of C. distinctissima |

1Finland, 2Sweden, 3Norway, 4Iceland, 5Russian Karelia.
Table 6: List of Finnish crane fly (Diptera, Tipuloidea) species, supplied with information on regional and global range size. Saproxylic/fungivorous and mire-dwelling species are indicated. Nomenclature follows Oosterbroek [47].

| Range | Prov. | FIN | Mal% | Glob | No C EUR | Sx | Mi |
|-------|-------|-----|------|------|-----------|----|----|
| Limoniidae |
| Achyroliomonia decemmaculata | 8 | Sb, Mb | 0.5 | W PAL | | 1 |
| Adelphomyia punctum | 3 | Sb, Mb | 1.2 | Tr-PAL | | |
| Antocha vitripennis | 4 | Sb, Mb, Nb | n.p. | Tr-PAL | | |
| Arctoconopa forcipata | 1 | Nb | n.p. | HOL | 1 |
| A. obscuripes | 1 | Nb | n.p. | HOL | 1 |
| A. zonata | 11 | Sb, Mb, Nb | 2.9 | Tr-PAL | | |
| Atypophthalmus inustus | 6 | Sb | 2.1 | Tr-PAL | 1 |
| Austrolimnophila (Archilimnophila) harperi | 5 | Mb, Nb | 1.9 | HOL | 1 |
| Cheliotrichia (Cheliotrichia) imbuta | 10 | Sb, Nb | 11.9 | HOL | 1 |
| C. (Empeda) arcorelata | 8 | Mb, Nb | 5.7 | HOL | | 1 |
| C. (Empeda) cinerascens | 15 | Sb, Mb, Nb | 24.7 | W PAL | | |
| C. (Empeda) neglecta | 10 | Sb, Mb | 3.3 | W PAL | | |
| Chionia (Chionea) araneoides | 6 | Sb, Mb, Nb | 0.5 | W PAL | | |
| C. (C.) crassipes | 3 | Mb, Nb | n.p. | Tr-PAL | 1 |
| C. (Sphaeconomus) lutescens | 6 | Sb, Mb, Nb | 0.2 | W PAL | | |
| Crypteria limnophiloides | 11 | Sb, Mb, Nb | 2.6 | Tr-PAL | | |
| Dicranomyia (D.) aperta | 6 | Sb, Mb, Nb | 5.2 | W PAL | | |
| D. (D.) autumnalis | 13 | Sb, Mb, Nb | 4.3 | W PAL | | |
| D. (D.) consimilis | 11 | Sb, Mb, Nb | 3.6 | Tr-PAL | | |
| D. (D.) didyma | 11 | Sb, Mb, Nb | 3.3 | Tr-PAL | | |
| D. (D.) distendens | 18 | Sb, Mb, Nb | 41.6 | HOL | | 1 |
| D. (D.) frontalis | 17 | Sb, Mb, Nb | 4.8 | HOL | | |
| D. (D.) halterata | 9 | Sb, Mb, Nb | 1.7 | HOL | | |
| D. (D.) handlirschi | 5 | Sb, Mb | 0.5 | Tr-PAL | | |
| D. (D.) hylalinata | 12 | Sb, Mb, Nb | 6.4 | HOL | | 1 |
| D. (D.) longipennis | 3 | Mb, Nb | 0.7 | HOL | | 1 |
| D. (D.) midis | 12 | Sb, Nb | 1.2 | Tr-PAL | | |
| D. (D.) modesta | 20 | Sb, Mb, Nb | 26.1 | HOL | | |
| D. (D.) moniliformis | 2 | Nb | 0.5 | HOL | 1 |
| D. (D.) omissinervis | 3 | Mb, Nb | 1.0 | Tr-PAL | | 1 |
| D. (D.) patens | 13 | Sb, Mb, Nb | 2.4 | Tr-PAL | | |
| D. (D.) radegastii | 1 | Sb | 0.2 | W PAL | | |
| D. (D.) sera | 4 | Sb, Mb | 1.2 | HOL | | |
| D. (D.) terraenovae | 17 | Sb, Mb, Nb | 18.8 | HOL | 1 |
| D. (D.) ventralis | 11 | Sb, Mb, Nb | 5.5 | Tr-PAL | | 1 |
| D. (D.) zernyi | 4 | Sb, Mb | 0.5 | Tr-PAL | 1 |
| D. (G.) liberta | 5 | Sb, Mb | n.p. | HOL | 1 |
| D. (G.) tristis | 6 | Sb, Nb | 1.0 | Tr-PAL | | |
| D. (Idiopyga) danica | 3 | Sb | n.p. | Tr-PAL | | |
| D. (I.) esbeni | 1 | Mb | 0.2 | Tr-PAL | 1 |
| D. (I.) halterella | 15 | Sb, Mb, Nb | 14.0 | HOL | | |
| D. (I.) intricata | 4 | Mb, Nb | 1.7 | HOL | 1 |
| D. (I.) klefbecki | 1 | Sb | n.p. | HOL | 1 |
| D. (I.) lulensis | 3 | Nb | 2.9 | FENNIS | | 1 |
| D. (I.) magnicaua | 11 | Sb, Mb, Nb | 0.5 | HOL | 1 |
| D. (I.) murina | 3 | Nb | 0.5 | HOL | 1 |
| D. (I.) ponojensis | 9 | Sb, Mb, Nb | 11.4 | HOL | 1 |
| **Species**                                                                 | **Prov.** | **FIN** | **Mal%** | **Glob** | **No C. EUR** | **Sx** | **Mi** |
|----------------------------------------------------------------------------|-----------|---------|----------|----------|---------------|--------|--------|
| *D. (I.) stigmatica*                                                       | 15        | Sb, Mb, Nb | 20.0     | Tr-PAL   | 1             |        |        |
| *D. (Melanolimonia) caledonica*                                           | 3         | Nb       | 2.9      | Tr-PAL   |               |        |        |
| *D. (M.) morio*                                                           | 15        | Sb, Mb, Nb | 1.7      | Tr-PAL   |               |        |        |
| *D. (M.) occida*                                                          | 5         | Mb, Nb   | 2.9      | Tr-PAL   | 1             |        |        |
| *D. (M.) rufiventris*                                                     | 15        | Sb, Mb, Nb | 26.4     | Tr-PAL   | 1             |        |        |
| *D. (M.) stylifera*                                                       | 3         | Nb       | 0.7      | W PAL    |               |        |        |
| *D. (Numantia) fusca*                                                     | 8         | Sb, Mb   | 4.8      | HOL      |               |        |        |
| *Dicranopycha cinerascens*                                                | 2         | Sb       | n.p.     | W PAL    |               |        |        |
| *D. fuscescens*                                                           | 2         | Sb       | n.p.     | Tr-PAL   |               |        |        |
| *Dicranophragma (Brachylinnophila) separatum*                             | 20        | Sb, Mb, Nb | 52.3     | W PAL    |               |        |        |
| *Discobola annulata*                                                      | 16        | Sb, Mb, Nb | 12.8     | HOL      | 1             |        |        |
| *D. caesarea*                                                             | 14        | Sb, Mb, Nb | 5.9      | Tr-PAL   | 1             |        |        |
| *Elephantomyia (E.) edwardsi*                                             | 3         | Sb       | 0.5      | W PAL    | 1             |        |        |
| *E. (E.) krivosheinae*                                                    | 11        | Sb, Mb, Nb | 1.4      | Tr-PAL   | 1             |        |        |
| *Eloeophila maculata*                                                     | 15        | Sb, Mb, Nb | 16.2     | Tr-PAL   |               |        |        |
| *E. submarmorata*                                                         | 4         | Sb, Mb   | 1.7      | W PAL    |               |        |        |
| *E. trimaculata*                                                          | 18        | Sb, Mb, Nb | 19.2     | W PAL    |               |        |        |
| *E. verralli*                                                             | 1         | Sb       | n.p.     | W PAL    |               |        |        |
| *Epiphragma (E.) ocellare*                                                | 12        | Sb, Mb   | 10.2     | HOL      | 1             |        |        |
| *Eriocnopa diurna*                                                        | 12        | Sb, Mb, Nb | 10.2     | W PAL    | 1             |        |        |
| *E. trivialis*                                                            | 13        | Sb, Mb   | 1.9      | W PAL    |               |        |        |
| *Erioptera (E.) beckeri*                                                  | 13        | Sb, Mb, Nb | 2.6      | Tr-PAL   | 1             |        |        |
| *E. divisa*                                                               | 6         | Sb, Mb   | 2.1      | W PAL    |               |        |        |
| *E. (E.) flavata*                                                         | 20        | Sb, Mb, Nb | 24.5     | Tr-PAL   | 1             |        |        |
| *E. (E.) griseipennis*                                                    | 2         | Sb       | n.p.     | W PAL    |               |        |        |
| *E. (E.) lutea*                                                           | 19        | Sb, Mb, Nb | 32.3     | Tr-PAL   |               |        |        |
| *E. nielseni*                                                             | 12        | Sb, Mb, Nb | 4.3      | W PAL    | 1             |        |        |
| *E. (E.) pederi*                                                          | 4         | Sb       | 1.2      | Tr-PAL   |               |        |        |
| *E. (E.) sordida*                                                         | 19        | Sb, Mb, Nb | 17.6     | Tr-PAL   |               |        |        |
| *E. (E.) squallida*                                                       | 6         | Sb, Mb, Nb | 1.9      | W PAL    |               |        |        |
| *E. (E.) tordi*                                                           | 1         | Sb       | 0.2      | Tr-PAL   | 1             |        |        |
| *Euphylidorea dispar*                                                     | 5         | Sb       | 1.7      | W PAL    |               |        |        |
| *Eugonomyia (G.) abscondita*                                              | 14        | Sb, Mb, Nb | 12.4     | W PAL    | 1             |        |        |
| *E. (G.) meigenii*                                                        | 20        | Sb, Mb, Nb | 28.5     | W PAL    |               |        |        |
| *Eutonia barbipes*                                                        | 4         | Sb       | 0.2      | W PAL    |               |        |        |
| *Gnaphomyia acheron*                                                      | 2         | Sb, Mb   | 0.2      | Tr-PAL   | 1             | 1      | 1      |
| *G. lugubris*                                                             | 5         | Sb, Mb   | 0.5      | Tr-PAL   | 1             |        |        |
| *G. viridipennis*                                                         | 3         | Sb       | 0.2      | Tr-PAL   | 1             |        |        |
| *Genempeda flavia*                                                        | 1         | Sb       | 0.2      | W PAL    |               |        |        |
| *Genomyia (G.) abscondita*                                                | 6         | Sb, Mb   | 1.9      | W PAL    |               |        |        |
| *G. (G.) bifida*                                                          | 2         | Sb       | 0.2      | W PAL    |               |        |        |
| *G. (G.) dentata*                                                         | 5         | Sb, Mb   | 0.5      | Tr-PAL   |               |        |        |
| *G. (G.) simplex*                                                         | 7         | Sb, Mb   | 1.9      | W PAL    |               |        |        |
| *G. (G.) stackelbergi*                                                    | 7         | Mb, Nb   | 8.6      | Tr-PAL   | 1             |        |        |
| *G. (G.) tenella*                                                         | 3         | Sb       | 0.2      | W PAL    |               |        |        |
| *G. (Teuchogonomyia) edwardsi*                                            | 2         | Sb       | n.p.     | Tr-PAL   |               |        |        |
| *Helius (H.) flavus*                                                      | 6         | Sb       | 1.2      | Tr-PAL   |               |        |        |
| *H. (H.) longirosstri*                                                    | 17        | Sb, Mb, Nb | 7.4      | W PAL    |               |        |        |
| *H. (H.) pallirostris*                                                    | 1         | Sb       | n.p.     | Tr-PAL   |               |        |        |
| *Hexatoma (H.) fuscipennis*                                               | 7         | Mb, Nb   | 0.2      | W PAL    |               |        |        |
| Prov. | FIN | Range | Glob | No C EUR | Sx | Mi |
|-------|-----|-------|------|-----------|----|----|
| **Hoplolabis (Parlilisia) areolata** | 4 | Sb, Nb | 0.2 | W PAL |    |    |
| *H. (P.) vicina* | 9 | Sb, Mb, Nb | 1.0 | Tr-PAL |    |    |
| *Idiopera linei* | 15 | Sb, Mb, Nb | 12.8 | Tr-PAL | 1 |    |
| *I. pulchella* | 19 | Sb, Mb, Nb | 23.5 | Tr-PAL |    |    |
| *Libnotes (Afrolimonia) ladogensis* | 3 | Sb, Mb | 0.5 | Tr-PAL | 1 |    |
| *Limophila (L.) pictipennis* | 1 | Sb | n.p. | Tr-PAL |    |    |
| *L. (L.) schranki* | 16 | Sb, Mb, Nb | 8.3 | Tr-PAL |    |    |
| *Limonia badia* | 6 | Sb, Mb | n.p. | HOL | 1 | 1 |
| *L. flavipes* | 13 | Sb, Mb, Nb | 12.1 | W PAL |    |    |
| *L. macrostigma* | 18 | Sb, Mb, Nb | 11.9 | Tr-PAL |    |    |
| *L. maculicosta* | 1 | Nb | n.p. | HOL | 1 |    |
| *L. mesauraea* | 1 | Mb | n.p. | FENNSC | 1 |    |
| *L. tuberculosa* | 8 | Sb, Mb | 1.0 | HOL |    |    |
| *L. phragmitidis* | 15 | Sb, Mb, Nb | 6.2 | Tr-PAL |    |    |
| *L. stigma* | 4 | Sb, Mb | 0.2 | W PAL |    |    |
| *L. sylvicola* | 16 | Sb, Mb, Nb | 10.5 | Tr-PAL |    |    |
| *L. trivittata* | 17 | Sb, Mb, Nb | 5.0 | Tr-PAL |    |    |
| *Lipsothrix ecucullata* | 11 | Sb, Mb, Nb | 9.5 | W PAL | 1 |    |
| *L. errans* | 1 | Sb | 0.5 | W PAL | 1 |    |
| *Metalimnobia (M.) bifasciata* | 19 | Sb, Mb, Nb | 22.1 | Tr-PAL | 1 |    |
| *M. (M.) charlesi* | 11 | Sb, Mb, Nb | 4.3 | W PAL | 1 |    |
| *M. (M.) quadrirufilateralia* | 20 | Sb, Mb, Nb | 11.9 | HOL | 1 |    |
| *M. (M.) quadrinotata* | 20 | Sb, Mb, Nb | 33.5 | Tr-PAL | 1 |    |
| *M. (M.) tenua* | 12 | Sb, Mb, Nb | 11.4 | Tr-PAL | 1 |    |
| *M. (M.) zetterstedti* | 20 | Sb, Mb, Nb | 50.1 | Tr-PAL | 1 |    |
| *Molophilus (M.) appendiculatus* | 12 | Sb, Mb, Nb | 8.3 | Tr-PAL |    |    |
| *M. (M.) ater* | 18 | Sb, Mb, Nb | 12.8 | Tr-PAL |    |    |
| *M. (M.) biphastus* | 4 | Sb, Nb | 0.7 | W PAL |    |    |
| *M. (M.) cinereifrons* | 7 | Sb, Mb | 2.6 | W PAL |    |    |
| *M. (M.) corniger* | 7 | Sb, Mb | 6.9 | W PAL |    |    |
| *M. (M.) crassipygus* | 18 | Sb, Mb, Nb | 17.3 | W PAL |    |    |
| *M. (M.) flavus* | 19 | Sb, Mb, Nb | 50.4 | W PAL |    |    |
| *M. (M.) griseus* | 11 | Sb, Mb, Nb | 4.0 | W PAL |    |    |
| *M. (M.) medius* | 8 | Sb, Mb | 6.7 | W PAL | 1 |    |
| *M. (M.) obscurus* | 1 | Sb | n.p. | W PAL |    |    |
| *M. (M.) occulus* | 1 | Sb | 0.2 | W PAL | 1 |    |
| *M. (M.) ochraceus* | 11 | Sb, Mb | 5.9 | W PAL |    |    |
| *M. (M.) propinquus* | 16 | Sb, Mb, Nb | 9.0 | Tr-PAL |    |    |
| *M. (M.) pullus* | 2 | Sb | 0.7 | W PAL |    |    |
| *Neolimnomyia (N.) batava* | 3 | Sb | 1.0 | W PAL |    |    |
| *Neolimnophila carteri* | 8 | Sb, Mb | 6.4 | W PAL |    |    |
| *N. placida* | 8 | Sb, Mb | 0.2 | Tr-PAL |    |    |
| *Neolimonia dumetorum* | 11 | Sb, Mb | 8.1 | W PAL | 1 |    |
| *Orimarga (O.) attenuata* | 8 | Sb, Mb, Nb | 8.6 | Tr-PAL | 1 |    |
| *O. (O.) juvenilis* | 2 | Mb, Nb | 0.2 | W PAL | 1 |    |
| *Ormosia (Oreophila) sootryeni* | 8 | Sb, Mb, Nb | 2.4 | Tr-PAL | 1 |    |
| *O. (Ormosia) brevinervis* | 1 | Nb | n.p. | W PAL | 1 |    |
| *O. (O.) clavata* | 6 | Sb, Mb | 5.9 | W PAL |    |    |
| *O. (O.) depilata* | 15 | Sb, Mb, Nb | 33.7 | W PAL |    |    |
| Taxon | Prov. | FIN | Range | Gloc | No C. EUR | Sx | Mi |
|-------|-------|-----|-------|------|-----------|----|----|
| O. (O.) fascipennis | 6 | Sb, Nb | 0.7 | HOL |
| O. (O.) hederae | 1 | Sb | n.p. | Tr-PAL |
| O. (O.) lineata | 12 | Sb, Mb | 10.0 | W PAL |
| O. (O.) laxia | 4 | Sb | 1.2 | W PAL |
| O. (O.) pseudosimilis | 16 | Sb, Mb, Nb | 13.3 | W PAL |
| O. (O.) ruficauda | 20 | Sb, Mb, Nb | 67.7 | W PAL |
| O. (O.) staegeriana | 18 | Sb, Mb, Nb | 11.6 | W PAL |
| Paradelphomyia (Oxynhiza) fuscata | 9 | Sb, Mb | 14.7 | W PAL |
| P. (O.) nigrina | 3 | Sb, Nb | 1.9 | W PAL |
| P. (Macrolabina) nigronotata | 2 | Sb, Nb | n.p. | Tr-PAL |
| P. (Paraphylidorea) fulvoversa | 19 | Sb, Mb, Nb | 41.1 | Tr-PAL |
| Phylidorea (P.) abdominalis | 16 | Sb, Mb, Nb | 7.4 | W PAL |
| P. (P.) bicolor | 14 | Sb, Mb, Nb | 3.6 | W PAL |
| P. (P.) ferruginea | 14 | Sb, Mb, Nb | 10.2 | Tr-PAL |
| P. (P.) heterogyna | 16 | Sb, Mb, Nb | 11.4 | W PAL |
| P. (P.) longicornis | 18 | Sb, Mb, Nb | 11.9 | Tr-PAL |
| P. (P.) nervosa | 7 | Sb, Mb | 1.7 | W PAL |
| P. (P.) squalens | 19 | Sb, Mb, Nb | 37.8 | Tr-PAL |
| P. (P.) umbrarum | 4 | Nb | 5.9 | Tr-PAL |
| Phyllolabis macroura | 4 | Sb, Nb | 1.7 | W PAL |
| Pilaria decolor | 19 | Sb, Mb, Nb | 8.6 | W PAL |
| P. discicollis | 13 | Sb, Mb, Nb | 2.6 | W PAL |
| P. meridiana | 18 | Sb, Mb, Nb | 12.4 | HOL |
| P. nigropunctata | 4 | Sb, Mb | 0.5 | W PAL |
| P. scutellata | 3 | Sb | 0.2 | Tr-PAL |
| Pseudolimnophila (P.) lucorum | 4 | Sb, Mb | 1.2 | Tr-PAL |
| Rhabdomastix (R.) borealis | 1 | Nb | n.p. | HOL |
| R. (R.) lacta | 8 | Sb, Mb, Nb | 3.1 | Tr-PAL |
| R. (unplaced) parva | 1 | Nb | 0.5 | FENNSC |
| Rhipidia (R.) maculata | 20 | Sb, Mb, Nb | 25.7 | HOL |
| R. (R.) uniseriata | 16 | Sb, Mb, Nb | 3.3 | Tr-PAL |
| Rhypophilus haemorrhoidalis | 16 | Sb, Mb, Nb | 13.1 | W PAL |
| R. varius | 5 | Sb | 1.9 | W PAL |
| Scleroprocta pentagonalis | 2 | Mb | 1.0 | Tr-PAL |
| S. sororcula | 16 | Sb, Mb, Nb | 16.6 | W PAL |
| Symplecta (Psiloconopa) lindrothi | 7 | Sb, Mb, Nb | 1.0 | FENNSC |
| S. (P.) meigeni | 7 | Mb, Nb | 3.1 | Tr-PAL |
| S. (P.) stictica | 8 | Sb, Mb | 1.7 | Tr-PAL |
| S. (S.) chosenensis | 1 | Sb | n.p. | Tr-PAL |
| S. (S.) hybrida | 16 | Sb, Mb, Nb | 4.5 | HOL |
| S. (S.) scotica | 5 | Sb, Mb, Nb | 0.5 | HOL |
| S. (S.) mabelana | 2 | Nb | 0.2 | HOL |
| S. (Trimicra) pilipes | 2 | Sb | n.p. | HOL |
| Tastocera (Dasymolophilus) exigua | 10 | Sb, Mb, Nb | 10.5 | W PAL |
| T. (D.) fusescens | 2 | Sb | 1.2 | W PAL |
| T. (D.) murina | 4 | Sb, Nb | 1.7 | W PAL |

**Tipulidae**

Angarotipula tumidicornis | 7 | Sb, Mb, Nb | 6.2 | Tr-PAL |
Ctenophora (C.) flaveolata | 2 | Sb | n.p. | W PAL |
C. (C.) guttata | 11 | Sb, Mb | 0.2 | Tr-PAL |
Table 6: Continued.

| Prov.² | FIN² | Mal%³ | Glob⁴ | No C EUR⁵ | Sx⁶ | Mi⁷ |
|--------|------|-------|-------|-----------|------|-----|
| C. (C.) nigriceps | 1 | Mb | 0.2 | W PAL | 1 |
| C. (C.) pectinicornis | 2 | Sb | n.p. | W PAL | 1 |
| Dictenidia bimaculata | 18 | Sb, Mb, Nb | 12.4 | Tr-PAL | 1 |
| Dolichopeza (D.) albipes | 9 | Sb, Mb, Nb | 2.9 | W PAL | 1 |
| D. (D.) bifida | 4 | Sb, Mb, Nb | 1.0 | Tr-PAL | 1 |
| Nephrotoma aculeata | 12 | Sb, Mb | 1.7 | Tr-PAL | 1 |
| N. analis | 10 | Sb, Mb | 4.0 | Tr-PAL | 1 |
| N. appendiculata | 9 | Sb, Mb | 0.7 | W PAL | 1 |
| N. cornicina | 14 | Sb, Mb | 1.2 | HOL | 1 |
| N. crocata | 11 | Sb, Mb | n.p. | Tr-PAL | 1 |
| N. dorsalis | 10 | Sb, Mb, Nb | 1.7 | Tr-PAL | 1 |
| N. flavescens | 11 | Sb, Mb | 1.4 | HOL | 1 |
| N. lundbecki | 1 | Nb | n.p. | HOL | 1 |
| N. lunulicornis | 11 | Sb, Mb | 3.1 | Tr-PAL | 1 |
| N. pratensis | 5 | Sb | n.p. | Tr-PAL | 1 |
| N. quadrirstrata | 7 | Sb, Nb | n.p. | Tr-PAL | 1 |
| N. relictica | 1 | Nb | n.p. | Tr-PAL | 1 |
| N. scurra | 19 | Sb, Mb, Nb | 1.4 | Tr-PAL | 1 |
| N. submaculosa | 1 | Mb | n.p. | W PAL | 1 |
| N. tenuipes | 12 | Sb, Mb, Nb | 1.4 | Tr-PAL | 1 |
| Nigrotipula nigra | 14 | Sb, Mb, Nb | 1.4 | Tr-PAL | 1 |
| Phoroctenia vidtata | 6 | Sb, Mb, Nb | 1.0 | Tr-PAL | 1 |
| Prionocera abscondita | 2 | Nb | 1.0 | Tr-PAL | 1 |
| P. chosenicola | 11 | Sb, Mb, Nb | 3.3 | HOL | 1 |
| P. pubescens | 16 | Sb, Mb, Nb | 14.0 | HOL | 1 |
| P. recta | 4 | Nb | 1.2 | HOL | 1 |
| P. ringdahli | 6 | Mb, Nb | 3.6 | HOL | 1 |
| P. serricornis | 7 | Sb, Nb | 5.5 | Tr-PAL | 1 |
| P. subsericornis | 19 | Sb, Mb, Nb | 13.5 | HOL | 1 |
| P. turcica | 18 | Sb, Mb, Nb | 13.5 | HOL | 1 |
| P. woodorum | 4 | Mb, Nb | 1.4 | HOL | 1 |
| Tanyptera (T.) atrata | 19 | Sb, Mb, Nb | 10.9 | Tr-PAL | 1 |
| T. (T.) nigricornis | 14 | Sb, Mb, Nb | 3.3 | Tr-PAL | 1 |
| Tipula (Acutipula) fulvipennis | 13 | Sb, Mb, Nb | 6.4 | Tr-PAL | 1 |
| T. (A.) maxima | 5 | Sb | 1.7 | W PAL | 1 |
| T. (Arctotipula) salicetorum | 4 | Nb | 0.5 | Tr-PAL | 1 |
| T. (Beringotipula) unca | 17 | Sb, Mb, Nb | 4.0 | Tr-PAL | 1 |
| T. (Dendrotipula) flavolineata | 5 | Sb | 1.2 | Tr-PAL | 1 |
| T. (Emodotipula) obscuriventris | 3 | Mb, Nb | 1.0 | W PAL | 1 |
| T. (Lindnerina) bistilata | 9 | Sb, Mb, Nb | 0.2 | Tr-PAL | 1 |
| T. (L.) subexcisa | 5 | Nb | 0.5 | Tr-PAL | 1 |
| T. (Lunatipula) affinis | 12 | Sb, Mb, Nb | 0.5 | Tr-PAL | 1 |
| T. (L.) circumdata | 14 | Sb, Mb, Nb | 1.7 | Tr-PAL | 1 |
| T. (L.) fascipennis | 13 | Sb, Mb | 4.5 | W PAL | 1 |
| T. (L.) humilis | 9 | Sb, Mb, Nb | 0.7 | Tr-PAL | 1 |
| T. (L.) laetabilis | 12 | Sb, Mb, Nb | 1.9 | Tr-PAL | 1 |
| T. (L.) limitata | 16 | Sb, Mb, Nb | 11.2 | Tr-PAL | 1 |
| T. (L.) lunata | 15 | Sb, Mb, Nb | 3.1 | Tr-PAL | 1 |
| T. (L.) recticornis | 2 | Sb | 0.2 | Tr-PAL | 1 |
| T. (L.) selene | 9 | Sb, Mb | n.p. | W PAL | 1 |
| Speciation | Prov. | FIN | Mal% | Glob | No C. EUR | Sx | Mi |
|------------|-------|-----|------|------|-----------|----|----|
| T. (L.) trispinosa | 5 | Mb, Nb | 5.9 | Tr-PAL | 1 |
| T. (L.) vernalis | 10 | Sb, Mb, Nb | 0.5 | W PAL | |
| T. (Odonatisca) nodicornis | 10 | Sb, Mb, Nb | 0.2 | Tr-PAL | |
| T. (Platytipula) luteipennis | 18 | Sb, Mb, Nb | 8.6 | Tr-PAL | |
| T. (P.) melanoceros | 19 | Sb, Mb, Nb | 20.7 | Tr-PAL | |
| T. (P.) cinereocincta | 4 | Sb | 0.5 | W PAL | |
| T. (P.) crassicornis | 1 | Mb | n.p. | HOL | |
| T. (P.) irrorata | 16 | Sb, Mb, Nb | 17.3 | Tr-PAL | |
| T. (P.) jutlandica | 1 | Sb | n.p. | Tr-PAL | |
| T. (P.) kaisilai | 1 | Nb | n.p. | Tr-PAL | |
| T. (P.) laetibasis | 5 | Sb, Mb, Nb | 0.7 | Tr-PAL | |
| T. (P.) luridorostris | 4 | Sb, Nb | 1.0 | Tr-PAL | |
| T. (P.) matsumariana pseudohortensis | 3 | Sb | 0.5 | W PAL | |
| T. (P.) mutila | 13 | Sb, Mb, Nb | 1.4 | Tr-PAL | |
| T. (P.) octomaculata | 3 | Sb, Mb, Nb | 0.5 | W PAL | |
| T. (P.) pauli | 1 | Sb | 0.2 | W PAL | |
| T. (P.) pseudoirrorata | 5 | Sb, Mb, Nb | 1.2 | W PAL | |
| T. (P.) recondita | 1 | Nb | 0.2 | Tr-PAL | |
| T. (P.) submarmorata | 16 | Sb, Mb, Nb | 2.4 | W PAL | |
| T. (P.) stenostyla | 4 | Sb, Nb | 0.7 | Tr-PAL | |
| T. (P.) truncorum | 13 | Sb, Mb, Nb | 1.0 | Tr-PAL | |
| T. (P.) wahlsgreni | 7 | Sb, Mb, Nb | 2.4 | Tr-PAL | |
| T. (P.) winthemi | 8 | Sb, Mb, Nb | 1.7 | Tr-PAL | |
| T. (Savsthenkia) alpium | 2 | Sb | 0.2 | HOL | |
| T. (S.) benesignata | 6 | Sb, Mb, Nb | 1.0 | Tr-PAL | |
| T. (S.) confusa | 6 | Sb, Mb | 0.7 | W PAL | |
| T. (S.) gimmerthali | 6 | Mb, Nb | 11.2 | W PAL | |
| T. (S.) grissescens | 17 | Sb, Mb, Nb | 27.3 | Tr-PAL | |
| T. (S.) interserta | 11 | Sb, Mb, Nb | 4.5 | Tr-PAL | |
| T. (S.) invenusta | 4 | Nb | 6.2 | HOL | |
| T. (S.) limbata | 15 | Sb, Mb, Nb | 16.9 | Tr-PAL | |
| T. (S.) obsoleta | 6 | Sb, Nb | 0.2 | W PAL | |
| T. (S.) pagana | 9 | Sb, Mb, Nb | 0.7 | W PAL | |
| T. (S.) signata | 10 | Sb, Mb, Nb | 0.7 | Tr-PAL | |
| T. (S.) subnodicornis | 18 | Sb, Mb, Nb | 43.7 | Tr-PAL | |
| T. (Schummelia) variicornis | 18 | Sb, Mb, Nb | 48.7 | Tr-PAL | |
| T. (Tippula) paludosa | 13 | Sb, Mb | 2.6 | HOL | |
| T. (T.) subcunctans | 12 | Sb, Mb, Nb | 1.2 | Tr-PAL | |
| T. (Vestiplex) excisa | 8 | Mb, Nb | 13.5 | Tr-PAL | |
| T. (V.) fororum | 4 | Sb | 0.2 | W PAL | |
| T. (V.) laccata | 4 | Nb | 0.5 | Tr-PAL | |
| T. (V.) montana verberneae | 4 | Nb | 2.6 | Tr-PAL | |
| T. (V.) nubeculosa | 18 | Sb, Mb, Nb | 24.2 | Tr-PAL | |
| T. (V.) pallidicosta | 4 | Mb, Nb | n.p. | W PAL | |
| T. (V.) scripta | 18 | Sb, Mb, Nb | 14.7 | Tr-PAL | |
| T. (V.) sintenisi | 11 | Sb, Mb, Nb | 4.8 | Tr-PAL | |
| T. (V.) tchukchi | 2 | Nb | 1.0 | Tr-PAL | |
| T. (Yamatotipula) chonsaniana | 3 | Mb, Nb | 1.2 | Tr-PAL | |
Table 6: Continued.

| Species                        | Prov. | FIN | Range     | Glob    | No C EUR | Sx | Mi    |
|--------------------------------|-------|-----|-----------|---------|----------|----|-------|
| T. (Y.) coerulescens           | 13    | Sb, Mb, Nb | 3.6 | W PAL   |          |    |       |
| T. (Y.) couckei                | 10    | Sb, Mb, Nb | 1.0 | Tr-PAL  |          |    |       |
| T. (Y.) fendleri               | 4     | Nb   | 1.9 | FENNSC  | 1        |    |       |
| T. (Y.) freyana                | 4     | Mb, Nb | 1.9 | Tr-PAL  | 1        |    |       |
| T. (Y.) lateralis              | 14    | Sb, Mb, Nb | 2.6 | Tr-PAL  |          |    |       |
| T. (Y.) marginella             | 7     | Sb, Mb | 0.2 | Tr-PAL  |          |    |       |
| T. (Y.) moesta                 | 3     | Nb   | 4.8 | Tr-PAL  | 1        |    |       |
| T. (Y.) montium                | 9     | Sb, Mb, Nb | 1.2 | Tr-PAL  |          |    |       |
| T. (Y.) pierrei                | 13    | Sb, Mb, Nb | 0.5 | Tr-PAL  |          |    |       |
| T. (Y.) pruinosa               | 15    | Sb, Mb, Nb | 4.5 | Tr-PAL  |          |    |       |
| T. (Y.) quadrivittata          | 9     | Sb, Mb, Nb | 1.4 | W PAL   |          |    |       |

Pediciidae

| Species                        | Prov. | FIN | Range     | Glob    | No C EUR | Sx | Mi    |
|--------------------------------|-------|-----|-----------|---------|----------|----|-------|
| Dicranota (D.) bimaculata      | 15    | Sb, Mb, Nb | 18.3 | Tr-PAL  |          |    |       |
| D. (D.) crassicauda            | 1     | Nb   | n.p. | Tr-PAL  | 1        |    |       |
| D. (D.) guerini                | 17    | Sb, Mb, Nb | 22.3 | Tr-PAL  |          |    |       |
| D. (Paradicranota) gracilipes  | 11    | Sb, Mb, Nb | 5.0  | W PAL   |          |    |       |
| D. (P.) pavida                 | 16    | Sb, Mb, Nb | 8.6  | W PAL   |          |    |       |
| D. (P.) robusta                | 4     | Nb   | 1.2 | W PAL   |          |    |       |
| D. (P.) subtilis               | 3     | Nb   | 0.5 | W PAL   |          |    |       |
| D. (Rhaphidolabis) exclusa     | 17    | Sb, Mb, Nb | 16.6 | Tr-PAL  |          |    |       |
| Pedicia (Crunobia) straminea   | 13    | Sb, Mb, Nb | 24.0 | W PAL   |          |    |       |
| P. (P.) rivosa                 | 20    | Sb, Mb, Nb | 55.8 | Tr-PAL  |          |    |       |
| Tricyphona (T.) immaculata     | 19    | Sb, Mb, Nb | 77.2 | Tr-PAL  |          |    |       |
| T. (T.) livida                 | 13    | Sb, Mb, Nb | 18.5 | W PAL   |          |    |       |
| T. (T.) schummeli              | 17    | Sb, Mb, Nb | 13.1 | W PAL   |          |    |       |
| T. (T.) unicolor               | 17    | Sb, Mb, Nb | 26.6 | W PAL   | 1        |    |       |
| Ula (U.) bolitophila           | 11    | Sb, Mb, Nb | 4.3  | Tr-PAL  | 1        |    |       |
| U. (U.) kiushiuensis           | 9     | Sb, Mb, Nb | 2.9  | Tr-PAL  | 1        |    |       |
| U. (U.) mixta                  | 16    | Sb, Mb, Nb | 28.7 | W PAL   | 1        |    |       |
| U. (U.) mollissima             | 7     | Sb, Mb, Nb | 2.4  | W PAL   | 1        |    |       |
| U. (U.) sylvatica              | 19    | Sb, Mb, Nb | 35.9 | HOL     | 1        |    |       |

Cylindrotomidae

| Species                        | Prov. | FIN | Range     | Glob    | No C EUR | Sx | Mi    |
|--------------------------------|-------|-----|-----------|---------|----------|----|-------|
| Cylindrotoma borealis          | 7     | Sb, Mb, Nb | 0.7  | FENNSC  | 1        |    |       |
| C. distinctissima              | 17    | Sb, Mb, Nb | 16.2 | HOL     |          |    |       |
| C. nigriventris                | 6     | Sb, Mb | 0.7 | Tr-PAL  | 1        |    |       |
| Diogma caudata                 | 10    | Sb, Mb, Nb | 7.6  | Tr-PAL  | 1        |    |       |
| D. glabrata                    | 13    | Sb, Mb, Nb | 14.3 | Tr-PAL  |          |    |       |
| Phalacrodera replicata          | 19    | Sb, Mb, Nb | 16.9 | HOL     | 1        |    |       |
| Triogma trisulcata             | 8     | Sb, Mb, Nb | 2.9  | Tr-PAL  | 1        |    |       |

1Number of occupied provinces in Finland. 2Range in Finland, Sb: hemi- and south boreal, Mb: middle boreal, and Nb: north boreal zone. 3Occupancy frequency in Finnish Malaise trapping data (out of 421 sites; some closely laying sites were combined to the sites × species matrix, that is why a smaller figure than 476 is given in the text), n.p.: not present. 4Global range, HOL: Holarctic, Tr-PAL: Trans-Palaearctic, W PAL: West Palaearctic, and FENNSC: Fennoscandian. 5Species not occurring in Central Europe. 6Saproxylix and fungivorous species. 7Mire-dwelling species.

(J. Salmela, pers. obs.). It is likely that the species occurs east of Fennoscandia, even though so far it has not been recorded there. Limonia messaurea messaurea is known from Sweden and Finland [56], apparently only from two localities. The species has a subspecies L. m. boreoorientalis that is known from Kamchatka, Russia [98]; this eastern Palaearctic subspecies differs from the nominotypical form only in color pattern, not by male genitalia [98]. Both subspecies are perhaps associated with pine bogs [98, 99]. Given the ambiguous nature of subspecies in crane fly taxonomy [100], it is likely that L. messaurea is a wide-ranging but disjunct Palaearctic species, not a Fennoscandian endemic. Rhabdomastix parva is a parthenogenetic species, males are unknown [101, 102]. The species is arctic, dwelling in northernmost Fennoscandia...
and Iceland. Evaluation of the status of the species is difficult because taxonomy of crane flies is heavily based on male genitalia. Molecular data and comparison with other Holarctic species would perhaps be fruitful in order to clarify the taxonomy of the species. *Symplecta lindrothi* is known from Sweden and Finland, in the latter country mainly around small lotic waters from south to north [56]. Due to its large range in Finland, it is perhaps a species occurring in but not yet collected from areas east of Fennoscandia. *Tipula fenderi* is known only from Finland, collected in the vicinity of springs and cold headwater streams [56]. Some of the collecting sites are very close to Sweden and Russia, thus the species is likely to be present also outside of the Finnish borders. The species may be a synonym of *T. nigrolamina*, a species known from Russian Far East [103] and Altay. Finally, *Cylindrotoma borealis*, originally described as a subspecies of Holarctic *C. distinctissima*, is known from Norway, Finland, and Russian Karelia [105]. This is the only case (see summary in Table 5).

5. Conclusions

The current number of Finnish crane fly species is 335. In practically all cases, the spatial scale being local or regional, the observed number of species is lower than estimated "true" richness (e.g., [107]). It is likely that there are crane fly species living within Finnish borders which are not yet collected and recorded. A total of 356 species are known from Sweden, a neighbor of Finland, and at least some Swedish (and Norwegian) species should occur in Finland too. However, geographic area of Sweden is larger than that of Finland, and it extends farther south than Finland, to the nemoral zone (more area, more heterogeneity = more species). Despite the potential occurrence of undetected species, it may be concluded that the general trends reported here are most likely genuine and will not change if additional species are discovered.

Finnish crane fly fauna is strongly influenced by latitude. Thus, one may predict that both species composition and richness of local assemblages are in a large scale determined by geographical position alone. Local environmental factors, such as bedrock composition, vegetation type, and moisture then finally filter the inhabitants from the regional pool of species (e.g., [61]).

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References

[1] A. G. Fischer, “Latitudinal variations in organic diversity,” *Evolution*, vol. 14, pp. 64–81, 1960.
[2] E. R. Pianka, “Latitudinal gradients in species diversity: a review of concepts,” *American Naturalist*, vol. 100, pp. 33–46, 1966.
[3] K. J. Gaston, “Global patterns in biodiversity,” *Nature*, vol. 405, no. 6783, pp. 220–227, 2000.
[4] M. Mönkkönen, J. T. Forsman, and F. Bokma, “Energy availability, abundance, energy-use and species richness in forest bird communities: a test of the species-energy theory,” *Global Ecology and Biogeography*, vol. 15, no. 3, pp. 290–302, 2006.
[5] A. Clarke, “Climate and diversity: the role of history,” in *Scaling Biodiversity*, D. Storch, P. A. Marquet, and J. H. Brown, Eds., pp. 225–245, Cambridge University Press, 2007.
[6] B. C. Cox and J. A. Moore, *Biogeography. An Ecological and Evolutionary Approach*, Blackwell Publishing, 7th edition, 2005.
[7] M. L. Rosenzweig, *Species Diversity in Space and Time*, Cambridge University Press, 1995.
[8] K. J. Gaston and T. Blackburn, *Pattern and Process in Macroecology*, Blackwell Science, 2000.
[9] M. Begon, J. L. Harper, and C. R. Townsend, *Ecology: Individuals, populations and communities*, Blackwell Science, Oxford, UK, 3rd edition, 1996.
[10] J. Kouki, P. Niemelä, and M. Viitasari, “Reversed latitudinal gradient in species richness of sawflies (Hymenoptera, Symphyla),” *Annales Zoologici Fennici*, vol. 31, no. 1, pp. 83–88, 1994.
[11] A. E. G. Dixon, P. Kindlmann, J. Leps, and J. Holman, “Why there are so few species of aphids, especially in the tropics,” *American Naturalist*, vol. 129, no. 4, pp. 580–592, 1987.
[12] A. Lillehammer, “Zoogeographical studies on Fennoscandian stoneflies (Plecoptera),” *Journal of Biogeography*, vol. 12, no. 3, pp. 209–221, 1985.
[13] J. Heino, “Concordance of species richness patterns among multiple freshwater taxa: a regional perspective,” *Biodiversity and Conservation*, vol. 11, no. 1, pp. 137–147, 2002.
[14] O. Järvinen and R. A. Väisänen, “Ecological zoogeography of North European waders, or why do so many waders breed in the North?” *Oikos*, vol. 30, pp. 495–507, 1978.
[15] O. Järvinen and L. Sammalisto, “Regional trends in the avifauna of Finnish peatland bogs,” *Annales Zoologici Fennici*, vol. 13, pp. 31–43, 1976.
T. Lindholm and R. Heikkilä, “Why do Tipulomorpha (Diptera, Insecta) succeed in the arctic conditions?” *Acta Zoologica Cracoviensia*, vol. 35, no. 1, pp. 193–197, 1992.

S. F. MacLean, “Life cycle and growth energetics of the Arctic crane-fly *Pedicia hannai antennata*,” *Oikos*, vol. 24, pp. 434–443, 1973.

M. J. Petersen, M. A. Bertone, B. M. Wiegmann, and G. W. Courtney, “Phylogenetic synthesis of morphological and molecular data reveals new insights into the higher-level classification of Tipuloidae (Diptera),” *Systematic Entomology*, vol. 35, pp. 526–545, 2010.

B. M. Wiegmann, M. D. Trautwein, I. S. Winkler et al., “Episodic radiations in the fly tree of life,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 14, pp. 5690–5695, 2011.

J. Salmela, “Phylogeny and classification of Tipulomorpha, with special emphasis on the family Liposyrphidae,” *Acta Zoologica Cracoviensia*, vol. 35, pp. 11–36, 1992.

J. Salmela, “Annotated list of Finnish crane flies (Diptera: Liposyrphidae, Ptychopteridae & Cylindrotomidae),” *Entomologica Fennica*, vol. 22, pp. 219–242, 2011.

O. Autio and J. Salmela, “A survey on the nematoceran (Diptera) communities of southern Finnish wetlands,” *Memoranda Societatis Pro Fauna et Flora Fennica*, vol. 86, pp. 43–53, 2010.

J. Salmela, “Adult craneflies (Diptera: Nematocera) around springs in southern Finland,” *Entomologica Fennica*, vol. 12, no. 3, pp. 139–152, 2001.

J. Salmela, “Semiaquatic flies (Diptera, Nematocera) of three mires on the southern boreal zone, Finland,” *Memoranda Societatis Pro Fauna et Flora Fennica*, vol. 80, no. 1, pp. 1–10, 2004.

J. Salmela, “Semiaquatic fly (Diptera, Nematocera) fauna of fens, springs, headwater streams and alpine wetlands in the northern boreal ecoregion, Finland,” *W-Album*, vol. 6, pp. 3–63, 2008.

J. Salmela, “The semiaquatic nematoceran fly assemblages of three wetland habitats and concordance with plant species composition, a case study from subalpine Fennoscandia,” *Journal of Insect Science*, vol. 11, article 35, 2011.

J. Salmela and J. Ilmonen, “Cranefly (Diptera) communities of southern Finnish wetlands,” *Memoranda Societatis Pro Fauna et Flora Fennica*, vol. 83, no. 2, pp. 33–47, 2007.

Metsien suojelun tarve Etelä-Suomessa ja Pohjanmaalla. J. Salmela, “The semiaquatic nematoceran fly assemblages of southern Finnish wetlands,” *W-Album*, vol. 6, pp. 3–63, 2008.

J. Salmela, “Semiaquatic flies (Diptera, Nematocera) of three mires on the southern boreal zone, Finland,” *Memoranda Societatis Pro Fauna et Flora Fennica*, vol. 80, no. 1, pp. 1–10, 2004.

J. Salmela, “Semiaquatic fly (Diptera, Nematocera) fauna of fens, springs, headwater streams and alpine wetlands in the northern boreal ecoregion, Finland,” *W-Album*, vol. 6, pp. 3–63, 2008.

J. Salmela, “The semiaquatic nematoceran fly assemblages of three wetland habitats and concordance with plant species composition, a case study from subalpine Fennoscandia,” *Journal of Insect Science*, vol. 11, article 35, 2011.

J. Salmela and J. Ilmonen, “Cranefly (Diptera: Tipulidae) fauna of a boreal mire system in relation to mire trophic status: implications for conservation and bioassessment,” *Journal of Insect Conservation*, vol. 9, no. 2, pp. 85–94, 2005.

J. Salmela, O. Autio, and J. Ilmonen, “A survey on the nematoceran (Diptera) communities of southern Finnish wetlands,” *Memoranda Societatis Pro Fauna et Flora Fennica*, vol. 83, no. 2, pp. 33–47, 2007.

“Metsien suojelun tarve Etelä-Suomessa ja Pohjanmaalla. Etelä-Suomen ja Pohjanmaan metsien suojelun tarve -työryhmän mietintö,” in *Suomen Ympäristö*, vol. 11, article 35, 2011.

T. Lindholm and R. Heikkilä, *Finland—Land of Mires*, vol. 23, The Finnish Environment, Helsinki, Finland, 2006.

E. B. Yakovlev, *Palaearctic Diptera Associated with Fungi and Myxomycetes*, Karelian Research Center, Russian Academy of Sciences, Petrozavodsk, Russia, 1994.

G. E. Hancock, S. M. Hewitt, A. Godfrey et al., “Thoracic spiracular gill structure of Liposyrphus (Diptera, Liposyrphidae) in Britain described from scanning electron micrographs,” *Zoosymposia*, vol. 3, pp. 77–87, 2009.
