Smoking and antidepressants pharmacokinetics: a systematic review

Pedro Oliveira¹, Joana Ribeiro¹, Helena Donato² and Nuno Madeira¹*

Abstract

Background: Despite an increasingly recognized relationship between depression and smoking, little is known about how smoking influences antidepressant response and treatment outcomes. The aim of this study was to systematically review the evidence of the impact of smoking on new-generation antidepressants with an emphasis on the pharmacokinetic perspective.

Methods: We present a systematic review of clinical trials comparing the serum levels of new-generation antidepressants in smokers and nonsmokers. Data were obtained from MEDLINE/PubMed, Embase, and other sources. Risk of bias was assessed for selection, performance, detection, attrition, and reporting of individual studies.

Results: Twenty-one studies met inclusion criteria; seven involved fluvoxamine, two evaluated fluoxetine, sertraline, venlafaxine, duloxetine or mirtazapine, and escitalopram, citalopram, trazodone and bupropion were the subject of a single study. No trials were found involving other common antidepressants such as paroxetine or agomelatine. Serum levels of fluvoxamine, duloxetine, mirtazapine and trazodone were significantly higher in nonsmokers compared with smokers.

Conclusions: There is evidence showing a reduction in the concentration of serum levels of fluvoxamine, duloxetine, mirtazapine and trazodone in smoking patients as compared to nonsmokers. The evidence regarding other commonly used antidepressants is scarce. Nonetheless, smoking status should be considered when choosing an antidepressant treatment, given the risk of pharmacokinetic interactions.

Keywords: Antidepressant agents, Pharmacokinetics, Depressive disorder, Smoking

Background

One-quarter of the general population, 40–50% of people with depression and 70–80% of those with schizophrenia smoke [1, 2]. Major depressive disorder (MDD) is a common psychiatric disease and a major public health problem. It has been projected to become the leading cause of disability and the second leading contributor to the global burden of disease and overall mortality by the year 2020; according to the recent epidemiologic data, about 10% of world population suffers with depression [3]. The advent of antidepressants (AD), starting with imipramine in 1958, has revolutionized the treatment of depression; yet this first generation of AD had an unfavorable adverse effect profile that has improved significantly with the advent in the late 1980s of selective serotonin reuptake inhibitors (SSRI) and, some years later, of serotonin and noradrenaline reuptake inhibitors (SNRI) and others like mirtazapine, trazodone, agomelatine and bupropion [4]. Given their favorable adverse effects profile, new-generation antidepressants are the first-line treatment of MDD. The mechanism of action and efficacy is similar between drugs of the same pharmacological class. However, there are considerable differences among interindividual responses to a given drug. Many theories tried to explain this variation but the most accepted is that clinical response is related to serum levels of the antidepressants. For many SSRI and other new-generation antidepressant drugs, a relationship between plasma concentrations and clinical effects is not reported [5]. However, serum levels
below therapeutic range could compromise the clinical response [5]. The serum levels of antidepressants are essentially dependent on two variables: the intake dose of drug and the rate of elimination. The rate of elimination of antidepressants is almost exclusively dependent on their hepatic metabolization. The hepatic metabolism of AD is made by cytochrome P450 (CYP) which is a group of many enzymes that exists in different amounts in human liver. Different antidepressants are metabolized by different subtypes of CYP and the amount of each CYP varies from person to person [6]. Thus, it is now accepted that the difference found in the response to an antidepressant could be correlated with serum concentrations. With the exception of tricyclic antidepressants, the correlation between plasmatic levels and the clinical outcome is still not consensual [7]. However, AGNP Consensus Guidelines from 2011 attribute a level 2 of recommendation for therapeutic drug monitoring for SSRI (except paroxetine) and SNRI [7]. Despite the availability of techniques capable of quantifying the activity of CYP isozymes such as 1A2, 2B6, 2C9 and 3A4 [8], they are not commonly used in clinical practice. So, given the absence of clinical indicators as relating to the higher or lower individual activity of each CYP, often the choice is made based on clinical experience with the particular drug and previous response of the individual to antidepressants in the past. Such imprecision can lead to the inappropriate use of drugs, not only wasting time until an adequate remission of depression but also causing iatrogenic harm, jeopardizing patient trust in antidepressant treatments.

There are 4000 chemical compounds found in cigarette smoke and 43 have been identified to be carcinogenic. Cigarette smoke constituents have been shown to stimulate or induce hepatic CYP isozymes, which play a central role in drug metabolism. Polycyclic aromatic hydrocarbons (PAH) from cigarette smoke are responsible for the induction of CYP isozymes. PAHs have been shown to induce CYP1A1, CYP1A2 and CYP2E1 [9].

Given the possibility that some antidepressants are metabolized by CYP induced or inhibited by substances in tobacco, their identification can be a guide for drug initial choice in smoking patients, allowing a more accurate antidepressant selection and consequently improving the pharmacologic treatment of depression in smokers.

Cigarette smoking can affect the clinical management of patients with psychiatric disorders because of the pharmacokinetic and pharmacodynamical changes; it can cause to various psychotropic drugs. This article reviews the impact of smoking on new-generation antidepressants with an emphasis on the pharmacokinetic perspective. It also seeks to provide critical information on whether such variations should influence antidepressant choice in this population.

Methods

This review was performed according to the PRISMA guidelines [10], thus providing a comprehensive framework which objectively assesses indicators of quality and risk of biases of included studies.

All original studies investigating the difference between the levels of any new-generation antidepressive agents (SSRI, SNRI, trazodone, mirtazapine, bupropion or agomelatine) and the smoking status were eligible for this systematic review. Further criteria adopted were: (1) publication date between January 1970 and June 2016, (2) empirical study, (3) written in English or Portuguese language, (4) published in a scholarly peer-reviewed journal, (5) studies that determined serum levels within a steady state, and (6) comparison of serum levels between 2 groups—smokers and nonsmokers. Additionally, studies were excluded from review if they were: (1) single-case report, (2) review articles, (3) repeated study population, (4) comparisons involving combinations of drugs, (5) animal studies, and (6) trials involving only metabolites.

Studies were identified by searching relevant papers via PubMed/MEDLINE (http://www.ncbi.nlm.nih.gov/pubmed), Cochrane Library and EMBASE using the following keywords: (“antidepressive agents”) AND (smok*). Finally, reference lists of retrieved studies were hand searched to identify any additional relevant studies. Keywords and combination of keywords were used to search the electronic databases and were organized following the population intervention comparison outcome (PICO) model (Fig. 1). In this model, the search strategy can be organized based on the topics: population (P), intervention (I), control group (C), and outcome (O) and several searches in the aforementioned databases.

After performing the initial literature searches, each study title and abstract was screened for eligibility by the first author. Full text of all potentially relevant studies were subsequently retrieved and further examined for eligibility. The PRISMA flow diagram (Fig. 1) provides more detailed information regarding the selection process of studies. Information from the included studies was then analyzed and recorded in an electronic spreadsheet designed by the first author. Different types of data were extracted from each study including: (a) country in which the data were collected and participants’ characteristics, (b) number of subjects, (c) number of smokers, (d) age average, (e) percentage of males, (f) main results (g) intervention protocol, (h) risk of bias in individual studies, and (i) limitations among others. The Cochrane Collaboration’s tool for assessing risk of bias was adopted to evaluate the risk of bias in individual studies [11]. The following risk of biases was analyzed: (1) selection bias, (2) performance bias, (3) detection bias, (4) attrition bias, and (5) reporting bias.
Results
Twenty-one articles were included in this review: seven are about fluvoxamine (FLV) [18–24], two about fluoxetine (FLX) [12, 13], sertraline [14, 15], venlafaxine (VEN) [25, 26], duloxetine [27, 28] and mirtazapine [30, 31], one about escitalopram [16], citalopram [17], trazodone [29] and bupropion [32]. No studies were found about paroxetine, milnacipran or agomelatine.

Eight studies were from Sweden, six were from Japan, four from Germany, two were from the United States of America (USA), and one from France and Switzerland. The studies reviewed included 2375 participants of which 733 were smokers. In terms of gender distribution, the vast majority of the studies reviewed recruited more female participants (64.31%) than male participants (35.69%). The average age of the subjects included in the studies is 45.53 years. A summary of results is given in Table 1 and the risk of bias in individual studies based on Cochrane Collaboration’s tool for assessing risk of bias is given in Table 2. As shown in Table 2, selection and reporting bias are the most frequent, with seven of twenty-one studies assessed with high risk for selection bias and four with high risk of reporting bias. No detection and attrition bias were found, although the risk of bias was not always clear.

Discussion
As mentioned above, tobacco interferes with drug metabolism essentially by the action of PAH that have effects through the induction of CYP 1A1, CYP1A2 and CYP2E1 [9].

All the antidepressants evaluated are metabolized in the liver by different types of cytochromes. With respect to SSRIs, citalopram is metabolized by CYP 2C19 and 3A4 [33], fluoxetine by 2D6, 3A4 and 2C9 [33], fluvoxamine by 1A2 and 2D6 [33], escitalopram by 2C19, 2D6 and 3A4 [16], and sertraline by 2D6, 3A4, 2C9 and 2C19 [33]. Regarding SNRI, venlafaxine is metabolized by CYP 2D6, 3A4 and 2C9 [33], and duloxetine by 2D6 and 1A2 [27]. Trazodone is metabolized by CYP 2D6 [29], mirtazapine by 1A2, 2D6 and 3A4 [31], and bupropion by 2B6 [32].

SSRI, except fluvoxamine, have little research on the effects of tobacco consumption in their serum levels. The trials performed with sertraline, escitalopram and citalopram showed no influence of smoking on their pharmacokinetics. However, the study on citalopram was based on a population in a restricted age group (all subjects were younger than 21 years) and studies regarding sertraline and escitalopram were not randomized and had many important limitations like the possibility of interactions with other drugs. The fluoxetine concentrations
| Study            | Drug      | Country | N total/N smokers | Age | Men (%) | Main findings                                                                 | Study limitations                                                                 |
|------------------|-----------|---------|-------------------|-----|---------|--------------------------------------------------------------------------------|----------------------------------------------------------------------------------|
| Lundmarck et al. [12] | Fluoxetine | Sweden  | 291/138           | 43  | 39      | No significant correlation between serum levels of FLX and smoking habits was found | Possibility of interactions with other medications                               |
| Koelch et al. [13] | Fluoxetine | Germany | 65/13             | 14.6| 37.0    | Serum concentrations of the active moiety and NORFLX were significantly correlated with smoking status | Possibility of interactions with other medications                               |
|                  |           |         |                   |     |         | Serum levels of the active moiety of FLX were 44% lower in smokers than in nonsmokers | Due to the ethnic polymorphism of CYP2D6 [6], the current results are not transferable to races other than Caucasians |
| Lundmarck et al. [14] | Sertraline | Sweden  | 319/89            | 54.4| 31      | Smokers had significantly lower concentration-to-dose (C/D) mean ratios of serum sertraline and its main metabolite desmethylsertraline than nonsmokers | Possibility of interactions with other medications                               |
| Taurines et al. [15] | Sertraline | Germany | 85/5              | 14.8| 45.1    | No significant correlation between serum levels of Sertraline and smoking habits was found | Possibility of interactions with other medications                               |
|                  |           |         |                   |     |         | No standardization of timing of blood withdrawal and length of treatment as well as patient compliance | Low number of subjects taking high doses of sertraline                           |
| Reis et al. [16]  | Escitalopram | Sweden  | 130/31            | 51  | 32      | No significant correlation between serum levels of Escitalopram and smoking habits was found | 76% of the patients took one or more drugs in addition to escitalopram             |
| Reis et al. [17]  | Citalopram | Sweden  | 19/10             | <21 | 18.8    | No significant correlation between serum levels of Citalopram and smoking habits was found | Possibility of interactions with other medications                               |
| Spigset et al. [18] | Fluvoxamine | Sweden  | 24/12             | 36.7| 58.3    | Cmax, and AUC were significantly lower in the smokers than in the nonsmokers | Only 50 mg/day of FLV was tested                                                  |
|                  |           |         |                   |     |         | There were no group differences in elimination half-life                           | Small sample size                                                                 |
| Carrillo et al. [19] | Fluvoxamine | Sweden  | 14/6              | 33.9| 50      | Among extensive metabolizers (CYP1A2 e CYP 2D6) there was no difference in FLV kinetics between smokers and nonsmokers | Only 50 mg/day of FLV was tested                                                  |
| Yoshimura et al. [20] | Fluvoxamine | Japan   | 30/11             | 52  | 36.7    | Serum levels of FLV were significantly higher in nonsmokers than smokers            | Small sample size                                                                |
| Gerstenberg et al. [21] | Fluvoxamine | Japan   | 49/15             | 49.9| 69.0    | No significant difference between nonsmokers and smokers in theCss of FLV and FLA and FLA/FLV ratio was found | Limited to extensive metabolizers                                               |
| Sugahara et al. [22] | Fluvoxamine | Japan   | 49/13             | w.i.| w.i.    | The mean C/D ratio of FLV in smokers was reduced by more than 30% in comparison with that in nonsmokers | Some subjects also took benzodiazepines                                           |
|                  |           |         |                   |     |         | No information about age or sex of patients                                       | Thirty-nine patients also took benzodiazepines                                   |

Table 1 Impact of smoking on main antidepressants (SSRI and SNRI) pharmacokinetics
| Study                  | Drug       | Country          | N total/N smokers | Age | Men (%) | Main findings                                                                 | Study limitations                                      |
|-----------------------|------------|------------------|-------------------|-----|---------|--------------------------------------------------------------------------------|--------------------------------------------------------|
| Katoh et al. [23]     | Fluvoxamine| Japan            | 32/6              | 39.0| 46.9    | The steady-state plasma C/D ratio of FLV in patients who smoked was significantly lower than that in nonsmoker patients | Possibility of interactions with other medications. Small sample size |
| Suzuki et al. [24]    | Fluvoxamine| Japan            | 87/22             | 36.6| 65.5    | Heavy smokers had significantly lower FLV concentration than nonsmokers in the FLV 50 mg/d dose group. At 150 mg/day and 200 dose groups, no significant differences in FLV concentration were observed between nonsmokers and heavy smokers | Possibility of interactions with other medications. Small sample size |
| Reis et al. [25]      | Venlafaxine| Sweden           | 141/58            | 61.3| 33      | The steady-state plasma C/D ratio of ODV and DDV in patients who smoked was significantly lower than that in nonsmoker patients. No differences in C/D VEN values or in any of the metabolite/VEN ratios were found | Possibility of interactions with other medications. |
| Unterecker et al. [26]| Venlafaxine| Germany          | 227/87            | 49.1| 36.4    | In smokers, mean serum levels of ODV were 21% lower than in nonsmokers. No differences in C/D VEN values between two groups | Possibility of interactions with other medications. |
| Fric et al. [27]      | Duloxetine | Germany          | 23/8              | 47.3| 75      | Smokers show significantly lower duloxetine serum level than nonsmokers | Possibility of interactions with other medications. Small sample size |
| Lobo et al. [28]      | Duloxetine | USA              | 594/123           | 48.8| 26      | Serum levels of duloxetine were significantly lower in smokers than in nonsmokers. Nonsmokers have a 43% higher Css than smokers | Possibility of interactions with other medications. |
| Ishida et al. [29]    | Trazodone  | Japan            | 43/16             | 43  | 44.2    | Smokers show significantly lower duloxetine serum level than nonsmokers | Small sample size                                       |
| Lind et al. [30]      | Mirtazapine| Sweden           | 56/36             | 50  | 34      | Smokers show significantly lower S-mirtazapine and R-N-desmethylmirtazapine serum levels than nonsmokers | No information about the number of cigarettes consumed daily |
| Sirot et al. [31]     | Mirtazapine| France/Switzerland| 45/17            | 51  | 18.9    | Smokers show significantly lower mirtazapine, S-mirtazapine and R-N-desmethylmirtazapine serum levels than nonsmokers | Possibility of interactions with other medications. |
| Hsu et al. [32]       | Bupropion  | USA              | 34/17             | 26.2| 52.9    | No significant correlation between serum levels of bupropion and smoking habits was found | Almost every subjects are caucasian. Small sample size. Only evaluates 150 mg daily. |

Table 1 continued
did not differ between the two study groups but the levels of its active metabolite, norfluoxetine, were higher in the group of smokers. Since norfluoxetine is an active metabolite, such association may impact the response of patients to take fluoxetine as well as implications with increased half-life of this molecule, which is already long, for example, by drug bioaccumulation and the ability to induce serotonin syndromes. Fluvoxamine has the greater evidence of decreased serum levels when associated with tobacco consumption. Although not proved, most studies suggest that there may be an association between the biotransformation of fluvoxamine and the activity of CYP1A2. It could not be excluded, however, that other factors may account for such a difference. Reasons include a possible association between smoking and fluvoxamine absorption, as well as between smoking and the elimination of fluvoxamine by other metabolic pathways. Further studies are, therefore, needed to clarify the role of CYP1A2 and other specific CYP in fluvoxamine metabolism and to elucidate which of the various metabolic steps could be dependent on CYP1A2. The recommended therapeutic reference range of fluvoxamine is 60–230 ng/mL [5]. Several studies have shown relations between plasma concentrations and clinical effects [5]. As a possible bias of these studies we highlight that most data involves Japanese individuals. This may influence the effect size, given, that this population presents quantitative differences, sometimes substantial, the various cytochrome P450 enzymes.

Assays for the SNRIs provide more consistent results than those involving SSRIs. Research on venlafaxine and duloxetine has similar designs, making result comparison easier and strengthening results. In both studies with venlafaxine, serum levels of both study groups showed no significant differences. Both pointed to a significant decrease in ODV levels. However, the pharmacodynamic effects of this are not completely understood, though many studies point to considerably weaker inhibition of serotonin and noradrenaline reuptake pumps when compared with venlafaxine itself [34]. For duloxetine, available data suggest a decrease in serum concentrations caused by tobacco consumption. In this drug, the available evidence is strong, based on multicenter randomized trials, involving a broad number of evaluated subjects. The effects of smoking status on duloxetine bioavailability can be attributed to the mechanism of duloxetine metabolism primarily by CYP1A2 enzyme. Smoking increases the expression of CYP1A2, which may explain the lower duloxetine bioavailability noted in smokers. The recommended therapeutic reference range of duloxetine is 30–120 ng/mL [5]. So far there is only a single retrospective analysis on plasma concentrations and
clinical effects that has shown concentration-dependent improvement [5].

Data suggest a decrease in serum concentrations of trazadone caused by tobacco consumption and no influence in serum concentrations of trazadone’s active metabolite m-chloro-phenylpiperazine (mCPP). This reduction could be due to the enhancement of hydroxylation and N-oxidation of trazadone caused by PAH from cigarette smoke [29]. A concentration–response relationship for trazadone has not been established [29]; however, one study has suggested the presence of a linear relationship [35].

The two studies that evaluated the effect of smoking on mirtazapine’s pharmacokinetics show significantly lower mirtazapine and R-N-desmethylmirtazapine serum levels in smokers than in nonsmokers. In vitro tomography study has shown that CYP1A2 is involved in 8-hydroxylation and possibly N-oxidation of mirtazapine [30]. Additionally, uridine diphosphate glucuronosyltransferases, another enzyme involved in the metabolism of mirtazapine, are inducible by smoking [30]. The recommended therapeutic reference range of mirtazapine is 30–80 ng/mL [5]. In a study on patients with depression, responders to mirtazapine treatment presented higher plasma concentrations than non-responders [36].

In the study with bupropion serum levels, both study groups showed no significant differences [32]. This study only evaluated daily doses of 150 mg of bupropion, reporting nothing on higher doses of drug.

Most antidepressants adverse effects are dose dependent, and some arise only when serum antidepressant levels reach a certain value [5]. Given that inhibition of CYP1A2 by tobacco smoke may decrease serum levels of some drugs, smoking cessation in heavy smokers taking such medication might lead to increased serum levels. Such an increase may cause adverse effects hitherto absent.

As mentioned above, the most frequent bias found was related with selection bias, which can lead to an over- or underestimation of the obtained results. Increasing the sample size and the use of control groups are recommended strategies to decrease this risk in future studies.

Conclusions
Despite numerous limitations in most studies, available evidence indicates a reduction in the concentration of serum levels of fluvoxamine, duloxetine, trazadone and mirtazapine in smoking patients when compared to nonsmokers. These differences raise the possibility of a semi-directed choice in antidepressant treatments, adapting the dose of these drugs and being aware of possible appearances of side effects after smoking cessation.

A personalized pharmacological treatment of depression could be made possible in a nearby future, guided by increasingly common and less expensive genotyping tools. For now, treatment personalization could be based on identifying phenotypes or external variables that influence antidepressant response or side effects. Further research is needed to improve our knowledge on the influence of smoking in depression pharmacological treatment.

Authors’ contributions
PO, JR, HD and NM conceived the study and participated in its design and coordination. PO drafted the manuscript. NH, HD and JR revised the manuscript. PO, JR and NM revised it critically for important intellectual content. All authors read and approved the final manuscript.

Acknowledgements
We would like to thank the anonymous peer reviewers for their insightful comments on this paper.

Competing interests
The authors declare that they have no competing interests.

Availability of data and materials
The datasets used and/or analyzed during the current study available from the corresponding author on reasonable request.

Received: 30 November 2016 Accepted: 24 February 2017
Published online: 06 March 2017

References
1. Goff DC, Henderson DC, Amico E. Cigarette smoking in schizophrenia: relationship to psychopathology and medication side effects. Am J Psychiatry. 1992;149(9):1189–94.
2. Winterer G. Why do patients with schizophrenia smoke? Curr Opin Psychiatry. 2010;23:112–9.
3. Bromet E, Andrade LH, Hwang I, Sampson NA, Alonso J, de Girolamo G, et al. Cross-national epidemiology of DSM-IV major depressive episode. BMJ. Med. 2011;9:90.
4. Bastos H, Polido F, Saraiva C. História da Psiquiatria. In: Saraiva C, Cerejeira J, editors. Psiquiatria fundamental. Coimbra: Livraria de Técnicas, Coimbra; 2011.
5. Haji EO, Hiemke C, Pfuhlmann B. Therapeutic drug monitoring for antidepressant drug treatment. Curr Pharm Des.. 2012;18(36):5818–27.
6. Ma Q, Lu AY. Pharmacogenetics, pharmacogenomics, and individualized medicine. Pharmacol Rev. 2011;63:437–59.
7. Hiemke C, Baumann P, Bergemann N, Conca A, Dietmaier O, Egberts K, et al. AGNP consensus guidelines for therapeutic drug monitoring in psychiatry: update 2011. Pharmacopsychiatry. 2011;44(6):195–235.
8. Kim YH, Bae YJ, Kim HS, Cha HJ, Yun JS, Shin JS, et al. Measurement of human cytochrome P450 enzyme induction based on mesalazine and mosapride citrate treatments using a luminessent assay. Biomed Ther. 2015;23(5):486–92.
9. Zévin S, Benowitz N. Drug interactions with tobacco smoking: an update. Clin Pharmacokin. 1999;36(6):425–38.
10. Moher D, Liberati A, Tetzlaff J, Altman DG. The PRISMA Group; preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. Open Med. 2009;3:123–30.
11. Higgins J, Green S. Cochrane handbook for systematic reviews of interventions Version 5.1.0. The Cochrane Collaboration. 2011. http://www.cochrane-handbook.org. Accessed June 2016.
12. Lundmark J, Margareta R, Finn B. Serum concentrations of fluoxetine in the clinical treatment setting. Ther Drug Monit. 2001;23:139–47.
13. Koelch M, Pfalzer A, Kliegl K, Rothenhofer S, Ludolph AG, Fegert JM, et al. Therapeutic drug monitoring of children and adolescents treated with fluoxetine. Pharmacopsychiatry. 2012;45:72–6.
14. Lundmark J, Margareta R, Finn B. Therapeutic drug monitoring of sertraline: variability factors as displayed in a clinical setting. Ther Drug Monit. 2000;22:446–54.
15. Taurines R, Burger R, Wewetzer C, Pfuhlmann B, Mehler-Wex C, Gerlach M, et al. The relation between dosage, serum concentrations, and clinical outcome in children and adolescents treated with sertraline: a naturalistic study. Ther Drug Monit. 2013;35:84–91.
16. Reis M, Cherma M, Carlsson B, Bengtsson F. Therapeutic drug monitoring of escitalopram in an outpatient setting. Ther Drug Monit. 2007;29:758–66.
17. Reis M, Olson G, Carlsson B, Bengtsson F. Serum levels of citalopram and its main metabolites in adolescent patients treated in a naturalistic clinical setting. J Clin Psychopharmacol. 2002;22:406–13.
18. Spigset O, Carleborg L, Hedenmalm K, Dahlqvist R. Effect of cigarette smoking on fluvoxamine pharmacokinetics in humans. Clin Pharmacol Ther. 1995;4:399–403.
19. Carrillo J, Dahl M, Svensson J, Alm C, Rodriguez I, Bertilsson L. Disposition of fluvoxamine in humans is determined by the polymorphic CYP2D6 and also by the CYP1A2 activity. Clin Pharmacol Ther. 1996;2:183–90.
20. Yoshimura R, Ueda N, Jun Nakamura J, Eto S, Matsushita M. Interaction between fluvoxamine and cotinine or caffeine. Neuropsychobiology. 1993;25:908–14.