Deep Sequencing Uncovers Caste-Associated Diversity of Symbionts in the Social Ant *Camponotus japonicus*

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**ABSTRACT** Symbiotic microorganisms can have a profound impact on the host physiology and behavior, and novel relationships between symbionts and their hosts are continually discovered. A colony of social ants consists of various castes that exhibit distinct lifestyles and is, thus, a unique model for investigating how symbionts may be involved in host eusociality. Yet our knowledge of social ant-symbiont dynamics has remained rudimentary. Through 16S rRNA gene deep sequencing of the carpenter ant *Camponotus japonicus* symbiont community across various castes, we here report caste-dependent diversity of commensal gut microbiota and lineage divergence of "*Candidatus Blochmannia*," an obligate endosymbiont. While most prevalent gut-associated bacterial populations are found across all castes (*Alphaproteobacteria*, *Gammaproteobacteria*, *Bacteroidetes*, and *Cyanobacteria*), we also discovered uncultured populations that are found only in males (belonging to *Corynebacteriales*, *Alkanindiges*, and *Burkholderia*). Most of those populations are not detected in laboratory-maintained queens and workers, suggesting that they are facultative gut symbionts introduced via environmental acquisition. Further inspection of "*Ca. Blochmannia*" endosymbionts reveals that two populations are dominant in all individuals across all castes but that males preferentially contain two different sublineages that are diversified from others. Clearly, each caste has distinct symbiont communities, suggesting an overlooked biological aspect of host-symbiont interaction in social insects.

**IMPORTANCE** Social animals, such as primates and some insects, have been shown to exchange symbiotic microbes among individuals through sharing diet or habitats, resulting in increased consistency of microbiota among social partners. The ant is a representative of social insects exhibiting various castes within a colony; queens, males, and nonreproductive females (so-called workers) show distinct morphologies, physiologies, and behaviors but tightly interact with each other in the nest. However, how this social context affects their gut microbiota has remained unclear. In this study, we deeply sequenced the gut symbiont community across various castes of the carpenter ant *Camponotus japonicus*. We report caste-dependent diversity of commensal gut microbial community and lineage divergence of the mutualistic endosymbiont "*Candidatus Blochmannia*." This report sheds light on the hidden diversity in microbial populations and community structure associated with guts of males in social ants.

**KEYWORDS** ant, gut microbiota, social insect
social interactions (e.g., through sharing diet or habitats), resulting in increased consistency between microbiota among social partners (3–5). However, disentangling how those social contexts affect the diversity of gut microbiota remains challenging.

Ants are eusocial insects that can serve as model organisms for social interactions given their clear social hierarchy. An ant colony is, in principle, composed of reproductive castes (queens and males) and a nonreproductive caste (workers), each of which shows different morphologies, physiologies, and behaviors (6). For carpenter ants of the *Camponotus* species, newly emerged queens and males mate in the nuptial flight. The inseminated queens construct a nest to lay eggs, and most of the newly hatched ants are raised as workers, consisting of a nonreproductive female caste showing task allocations distributed in an age-dependent and environment-dependent manner (6, 7). While workers forage outside and then feed nest mates, queens and males remain inside the nest until the nuptial flight. These castes also display differences in longevity—queens can live for over 20 years (6, 8), workers live for between a month and a few years, and males die immediately after the mating flight. Given the diversity among castes, *Camponotus* species can be a suitable model to explore how social contexts, particularly castes and colonies, affect gut microbiota.

The forms of diversity in the commensal gut microbiota remain unclear due to the high abundance of endosymbiotic *“Candidatus Blochmannia”* in gut-associated cells (9–11) overwhelming those of commensal gut symbionts in DNA-based analyses (e.g., more than 95% of the reads of *Camponotus* gut metagenome were assigned to *“Ca. Blochmannia”* [12, 13]). While *“Ca. Blochmannia”* undergoes maternal transmission to offspring and interacts mutualistically with the host (14), whether host-endosymbiont interactions differ between castes also remains unknown.

Here, we explored and compared the gut microbial communities of various castes (virgin or mated queen, worker, and male) of the carpenter ant *C. japonicus* through deep sequencing of the bacterial 16S rRNA gene. We collected a total of 39 individual ants in Tsukuba, Japan, in the following categories: 7 virgin queens, 4 mated queens, 7 workers, and 10 males from eight field colonies and 2 mated queens and 9 workers from two laboratory-reared colonies (see Text S1 in the supplemental material). The amplicon sequencing of individual gut samples yielded 101,961,345 sequences (average, $2.6 \times 10^6$ sequences per sample) associated with 756 operational taxonomic units (OTUs). A major portion of the sequences (100,843,490) were affiliated with endosym-
Phylogeny, distribution, and relative abundance of symbionts across various castes of *C. japonicus*. (a) Relative abundance of commensal gut microbes. OTUs were grouped based on 97% sequence similarity (CD-HIT-EST v4.8.1). The relative abundance of each OTU group in non-Enterobacteriaceae in each individual is shown along with each individual’s caste (VQ = virgin queen, MQ = mated queen, W = worker, M = male) and colony source (colonies A to K, with unknown colonies indicated as “?”). Individuals belonging to colonies maintained in the laboratory are highlighted in gray. Only OTU groups with.

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biotic Enterobacteriaceae, most (99.999%) of which primarily consisted of 2 dominant "Ca. Blochmannia" populations (46.1 million reads for CJS630 and 54.1 million reads for CJS647) and 261 other minor "Ca. Blochmannia" populations (0.6 million reads). Using results of quantitative PCR (qPCR) analyses of 16S rRNA gene copies (see Fig. S1 in the supplemental material), we found that the estimated abundance of "Ca. Blochmannia" was much lower in males than in other castes (P < 0.05; only field samples compared), while the abundances of commensal gut microbiota (i.e., non-Enterobacteriaceae) were statistically indistinguishable among castes (P > 0.1) (Fig. 1A).

Our deep sequencing allowed detection of 1,117,855 sequences affiliated with commensal gut microbes spanning 421 OTUs (Fig. S2). We performed beta-diversity analysis of the commensal microbial communities to examine differences in their structures among castes (Text S1). The community structures indeed differed within colonies, whereas no obvious differences were observed between colonies (Fig. 1B; see also Fig. S3). The relative contribution of caste to the community structure was obvious: while queens tend to have consistent communities, workers and males had distinctive communities that were highly variable among individuals (Fig. 1B). All castes in the field had gut microbes belonging to Gammaproteobacteria, Alphaproteobacteria, Cyanobacteria, and Bacteroidetes (Fig. 2A), most of which were not detected in laboratory-reared samples. Acetobacteraceae populations (CJS181 and CJS184) frequently detected in field samples (12) were also undetected, indicating that most commensal microbial populations are facultative symbionts acquired from the environment. Several Gammaproteobacteria species were still visible in laboratory-born workers, suggesting the potential of vertical inheritance. Interestingly, while no clades were exclusive to queens or workers, three taxa were associated only with males (Alkanindiges, Burkholderia, and Corynebacteriales), suggesting that the male gut environment or behavior may harbor unique features that select for such organisms. As of yet, detailed studies on the behavior of males in the nest are still lacking. Currently conceivable routes of exposure to exogenous microbes include nest soil or trophallaxis with nest mates (6), but both are available to other castes. The male-specific microbes might thus be provided by further unknown mechanisms, such as specific diet or infection for males. The uncultured Corynebacteriales population (CJS031) was found to be present across nearly all male samples (9 of 10) and belongs to an uncharacterized lineage containing moderately related (≥96.2% sequence similarity) symbionts from Australian weaver ants (Polyrhachis rosonbi) (Fig. S4) (15). The Corynebacteriales order contains a variety of environmental bacteria often degrading hydrocarbons (16) and secreting various metabolites (17, 18). Given the characteristics of these bacteria, we suspect that the male-specific microbes may be involved in the production of cuticular hydrocarbons, profiles of which often act as sensory cues to distinguish species, sexes, or social status (19–21).

“Ca. Blochmannia” populations were diverse in individual ants (Fig. 2B; see also Fig. S5), which is in agreement with the general aspects of endosymbiont genome evolution (22, 23). We found that CJS647 (53.43% of total microbial community on average), CJS630 (45.58%), and CJS623 (0.04%) were ubiquitous across castes and colonies in the field. Given that all "Ca. Blochmannia" genomes sequenced had a single copy of a 16S rRNA gene (24), each detected OTU likely represents distinct "Ca.

FIG 2 Legend (Continued)

at least 1% abundance in at least two individuals are included. For each OTU group, a representative OTU, phylum, class, and the most specific classification beyond class (genus, family, or order; if defined) are shown. OTU groups with the same classification are indicated with hyphens. OTUs associated only with males are highlighted in purple. OTUs are depicted in red letters if (i) the OTUs were detected across at least 70% of the individuals in a specific caste and (ii) the average abundance across that caste greater was than 5.6%—the lowest detection limit among queens—based on the data from the queen with the lowest nonzero number of reads for non-Enterobacteriaceae symbionts (18 reads for virgin queen G1; leftmost in figure). (b) Abundance and phylogenetic tree of "Ca. Blochmannia." Only OTUs with at least 0.01% abundance in two or more individuals are shown. Sequences were aligned using SINA and SILVA v132. The phylogenetic tree was constructed using RaML v8 with generalized time reversal (GTR) model, 4 discrete GAMMA categories, and 100 bootstrap iterations (see complete tree in Fig. S5). Bootstrap values are shown as colored circles on each branch (black = ≥90, gray = ≥75, and white = ≥50) none are shown for those with values of <50). Note that the OTUs are not intracellular variations of 16S rRNA gene sequences because "Ca. Blochmannia" has a single copy of a 16S rRNA gene. The abundance of each OTU in each individual is shown as described for panel a. Those found in statistically significantly higher abundance in males than in other castes (P < 0.05) are marked in purple.

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“Ca. Blochmannia” populations and not variation in 16S rRNA sequences in a single genome. This is further supported by unequal abundances of two dominant populations (CJS647 and CJS630) (Fig. S6). The two dominant populations were present even in all laboratory-reared samples, suggesting that they are fundamentally important for their symbiotic relationships with the host. While the presence of two different genotypes of “Ca. Blochmannia” has been previously reported at the colony level of polygyne species where the two genotypes are derived from two queens (25), this is the first report that two dominant “Ca. Blochmannia” phylotypes transmitted by a single queen cocolonized individual ants in monogyne colonies. More importantly, the community structures of the minor Enterobacteriaceae symbionts (excluding the two dominant populations CJS630 and CJS647) were found to differ between the male caste and other castes (Fig. 1C). While most “Ca. Blochmannia” populations were sporadically present across different castes and colonies, two specific sublineages containing CJS502 or CJS572 were found in all males at a higher relative abundance than in other castes (P < 0.05), indicating that those lineages are associated with males rather than representing random inheritance due to the small inoculum size in males. In addition, one of those containing CJS502 showed a relatively high level of phylogenetic deviation from the others. The higher abundances of these populations in males than in other castes (7.1× to 12.4× and 4.4× to 6.3× higher than queens and workers, respectively) are unlikely to have a significant impact on the host given their relatively low abundance compared to the two dominant CJS647 and CJS630 populations. However, the abundance of these male-associated “Ca. Blochmannia” endosymbionts can collectively reach up to 151% higher than that of the remaining non-Enterobacteriaceae commensal gut symbionts. The pervasiveness and pronounced abundance of the minor “Ca. Blochmannia” populations suggest that they may be enriched through host development and/or may increase their fitness more in males than females to play potential roles in male-specific functions.

Males in social insect seldom show social tasks and thus have not been the focus of analyses of social behavior or physiology. However, considering their central role in reproduction, one can expect that males have unique systems or life cycles to maximize their success in the nuptial flight. Our report provides the first glimpse into the uniqueness of microbial populations and community structure associated with male guts in social insects.

Data availability. The 16S rRNA gene amplicon data sets generated during this study have been deposited and are available in the Sequence Read Archives of the National Center for Biotechnology Information (NCBI), European Bioinformatics Institute (EBI), and DNA Data Bank of Japan (DDBJ) under accession no. SRR10569543 to SRR10569587.

SUPPLEMENTAL MATERIAL
Supplemental material is available online only.

TEXT S1, DOCX file, 0.03 MB.
FIG S1, TIF file, 1.5 MB.
FIG S2, TIF file, 0.3 MB.
FIG S3, TIF file, 0.3 MB.
FIG S4, TIF file, 0.3 MB.
FIG S5, PDF file, 1.9 MB.
FIG S6, TIF file, 0.1 MB.

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We declare that we have no conflict of interest.
A.K. and R.M. designed research and performed experiments; M.K.N. and R.M. analyzed data; all of us wrote and approved the paper.

REFERENCES

1. Cani PD, Plovier H, Van Hul M, Geurts L, Delzenne NM, Druart C, Everard A. 2016. Endocannabinoids–at the crossroads between the gut microbiota and host metabolism. Nat Rev Endocrinol 12:133–143. doi.org/10.1038/nrendo.2015.211.

2. Sommer F, Backhed F. 2013. The gut microbiota–masters of host development and physiology. Nat Rev Microbiol 11:227–238. https://doi.org/10.1038/nrmicro2974.

3. Dill-McFarland KA, Tang ZZ, Kerby RL, Chen G, Palloni A, Sorenson T, Rey FE, Herd P. 2019. Close social relationships correlate with human gut microbiota composition. Sci Rep 9:703. https://doi.org/10.1038/s41598-018-37298-9.

4. Koch H, Schmidt-Hampel P. 2011. Socially transmitted gut microbiota protect bumble bees against an intestinal parasite. Proc Natl Acad Sci U S A 108:19288–19292. https://doi.org/10.1073/pnas.110474108.

5. Wilson E, Hölldobler B. 1990. The ants. Springer, Berlin, Germany.

6. Wilson E, Hölldobler B. 1990. The ants. Springer, Berlin, Germany.

7. Cani PD, Plovier H, Van Hul M, Geurts L, Delzenne NM, Druart C, Everard A.K. and R.M. designed research and performed experiments; M.K.N. and R.M. wrote and approved the paper.

11. Schroder D, Deppisch H, Obermayer M, Krohne G, Stackebrandt E, Koto et al. 2015. Posting date. Deep divergence analyzed data; all of us wrote and approved the paper.

12. Brown BP, Wernegreen JJ. 2017. Species-specific signatures of human gut microbiota composition. Rep 9:703. https://doi.org/10.1038/s41598-018-37298-9.

13. Ramalho MO, Bueno OC, Moreau CS. 2017. Social networks predict gut microbiome composition in wild bumble bees against an intestinal parasite. Proc Natl Acad Sci USA 115:E226–E235. doi.org/10.1073/pnas.1533499100.

14. Schroder D, Deppisch H, Obermayer M, Krohne G, Stackebrandt E, Koto et al. 2015. Posting date. Deep divergence analyzed data; all of us wrote and approved the paper.

15. Russell JA, Funaro CF, Giraldo YM, Goldman-Huertas B, Suh D, Kronauer DJ, Moreau CS, Pierce NE. 2012. A veritable menagerie of heritable bacteria from ants, butterflies, and beyond: broad molecular surveys and a systematic review. PLoS One 7:e51027. https://doi.org/10.1371/journal.pone.0051027.

16. Mikolash A, Hammer E, Schauer F. 2003. Synthesis of imidazol-2-yl amino acids by cells using from alkane-oxidizing bacteria. Appl Environ Microbiol 69:1670–1679. https://doi.org/10.1128/AEM.69.3.1670-1679.2003.

17. Becker J, Wittmann C. 2015. Advanced biotechnology: metabolically engineered cells for the bio-based production of chemicals and fuels, materials, and health-care products. Angew Chem Int Ed 54:3328–3350. https://doi.org/10.1002/anie.201409033.

18. Vonshak MAS. 2009. A Camponotus fellah queen sets a record for Israeli ant longevity. Isr J Entomol 39:165–168.

19. Chernenko A, Holman L, Helantera H, Sundstrom L. 2012. Cuticular chemistry of males and females in the ant Formica fusca. J Chem Ecol 38:1474–1482. https://doi.org/10.1007/s10886-012-0217-4.

20. Smith AA, Vanderpool W, Millar JG, Hanks LM, Suarez AV. 2014. Converged male-specific cuticular hydrocarbon patterns in the trap-jaw ant Odontomachus brunneus. Chemoecology 24:29–34. https://doi.org/10.1007/s10886-013-0143-0.

21. Chung H, Carroll SB. 2015. Wax, sex and the origin of species: dual roles of insect cuticular hydrocarbons in adaptation and mating. Bioessays 37:822–830. https://doi.org/10.1002/bies.201500014.

22. Lukasik P, Nazario K, Van Leuven JT, Campbell MA, Meyer M, Michalik A, Pessaq P, Simon C, Veloso C, McCutcheon JP. 2018. Multiple origins of interdependent endosymbiotic complexes in a genus of cicadas. Proc Natl Acad Sci U S A 115:E226–E235. https://doi.org/10.1073/pnas.1712321115.

23. Woyke T, Tighe D, Mavromatis K, Clum A, Copeland A, Schackwitz W, Cheng JF. 2010. One bacterial cell, one complete genome. PLoS One 5:e10314. https://doi.org/10.1371/journal.pone.0010314.

24. Williams LE, Wernegreen JJ. 2015. Genome evolution in an ancient bacteria-ant symbiosis: parallel gene loss among Blochmannia spanning the origin of the ant tribe Camponotini. PeerJ 3:e881. https://doi.org/10.7717/peerj.881.

25. Williams LE, Wernegreen JJ. 2012. Purifying selection, sequence composition, and context-specific indel mutations shape intraspecific variation in a bacterial endosymbiont. Genome Biol Evol 4:44–51. https://doi.org/10.1093/gbe/evr128.