67 kDa laminin receptor (67LR) in normal and neoplastic hematopoietic cells: is its targeting a feasible approach?

Nunzia Montuori¹, Ada Pesapanè¹, Valentina Giudice², Bianca Serio², Francesca W Rossil, Amato De Paulis¹, Carmine Selleri²

¹Department of Translational Medical Sciences, University of Naples Federico II, Naples, Italy
²Department of Medicine and Surgery, University of Salerno, Salerno, Italy.

Abstract - The 67 kDa laminin receptor (67LR) is a non-integrin cell surface receptor for laminin (LM) that derives from a 37 kDa precursor (37LRP). 67LR expression is increased in neoplastic cells and correlates with an enhanced invasive and metastatic potential in many human solid tumors, recommending this receptor as a new promising target for cancer therapy. This is supported by in vivo studies showing that 67LR downregulation reduces tumor cell proliferation and tumor formation by inducing apoptosis. 67LR association with the anti-apoptotic protein PED/PEA-15 activates a signal transduction pathway, leading to cell proliferation and resistance to apoptosis.

However, the main function of 67LR is to enhance tumor cell adhesion to the LM of basement membranes and cell migration, two crucial events in the metastasis cascade. Thus, inhibition of 67LR binding to LM has been proved to be a feasible approach to block metastatic cancer cell spread.

Despite accumulating evidences on 67LR overexpression in hematologic malignancies, 67LR role in these diseases has not been clearly defined. Here, we review 67LR expression and function in normal and malignant hematopoietic cells, 67LR role and prognostic impact in hematological malignancies and first attempts in targeting its activity.

Keywords: laminin receptor, laminin, hematopoietic stem cells, leukemia, multiple myeloma

I. INTRODUCTION

The 67 kDa laminin receptor (67LR) is a non-integrin cell surface receptor for extracellular matrix (ECM) able to bind with high affinity laminin-1 (LM), the major glycoprotein of basement membranes [1]. The primary function of 67LR is to promote tumor cell adhesion and migration to LM, crucial steps in tissue invasion and metastasis, by binding LM with high affinity (Kd=2x10^-9 M). cDNA clones coding for human and mouse 67LR encode a protein with a molecular weight of 32kDa and an apparent electrophoretic mobility of about 37kDa. This polypeptide was identified as the precursor of 67LR, and named 37kDa laminin receptor precursor (37LRP) [2].

The mechanism by which 37LRP is converted into the mature 67LR is still unclear [3,4]. 37LRP is abundantly localized in the cytoplasm, where it acts as a multifunctional protein involved in the translational machinery and in ribosome assembly, and in the nucleus, tightly associated with nuclear structures [5].67LR, the mature form of the receptor, is localized in the cell membrane, from which it is internalized via early-endosomes and lysosomal-mediated degradation [6]. On the cell surface, 67LR is able to interact with α6β4 integrin; both receptors are co-expressed and physically associated in a complex that recognize different sites on LM, increasing the affinity of the binding [7]. Upon binding 67LR, LM interacts more efficiently with integrins and becomes more sensitive to proteolytic enzymes, releasing fragments endowed with chemotactic activity [8]. Three regions of 37LRP/67LR are involved in LM binding: (i) repeated sequences (TWEDS) at the C-terminal, (ii) a direct laminin binding region (amino acids 205-229) [9] and (iii) a heparan sulfate dependent LM binding region (amino acids 161-180), also called peptide G, and containing the palindromic sequence LMWWML, responsible for LM binding [10].

All the three LM-binding sites of the receptor bind the same minimal YIGSR region of the β1 chain of LM [11]. As a membrane receptor, 37LRP/67LR is also a receptor for viruses, bacteria and prions [5,12]. 67LR is overexpressed in neoplastic cells and correlates with an enhanced invasive and metastatic potential in many solid tumors [13-17]. 67LR role in metastatic diffusion is well documented and mostly rely in its ability to mediate adhesion to the LM of basement membranes of epithelia and endothelia and to mediate trans-endothelial migration of metastatic cancer cells [18]. 67LR role in hematological malignancies has not been clearly defined, even though many reports have been produced on its expression, function and inhibition on normal and malignant hematopoietic cells.
II. 37LRP/67LR FUNCTION AND TARGETING IN SOLID TUMORS

Given its importance in solid tumor progression, 67LR represents a suitable target for cancer therapy and different approaches have been used to inhibit its function, in order to contribute to metastasis prevention and/or treatment (reviewed in 19). Different strategies used against this receptor were able to reduce the invasive potential of HT1080 fibrosarcoma cells [20]. 37LRP/67LR is able to affect tumor progression also by promoting angiogenesis; indeed, a receptor specific antibody inhibited endothelial tube formation [21]. 37LRP/67LR is involved in the maintenance of cellular viability and reduction of its expression induced apoptosis of cancer cells [22]. The role of 37LRP/67LR in apoptosis was also confirmed by the finding of our group of a structural and functional association between 67LR and the anti-apoptotic protein PED/PEA-15 [23].

A recent study reveals a novel function of 37LRP/67LR: siRNA treatment of 37LRP/67LR resulted in a significant decrease of telomerase activity [24]. Recently, our group searched, by a virtual screening (VS) approach, small molecules able to specifically target 67LR. VS is a computational method that allows the identification of new therapeutics for a specific biological target from large chemical libraries [25]. This study led to the identification of a specific inhibitor of 37LRP/67LR, NSC47924. This compound specifically inhibited cell adhesion and migration to LM, as well as cell invasion. A subsequent hierarchical similarity search with NSC47924 allowed the refinement of this lead compound, identifying additional four compounds able to inhibit cell binding to LM and to block in vitro cancer cell invasion, exhibiting a lower IC50 as compared to NSC47924 [26].

These small molecules are cell-permeable and orally available, the most important advantage in respect to monoclonal antibodies. Moreover, they showed a short half-life, high specificity and low toxicity, thus may be of considerable clinical benefit in tailoring personalized target therapies in cancer.

III. 37LRP/67LR EXPRESSION AND FUNCTION IN NORMAL HEMATOPOIETIC STEM CELLS

HSCs normally resides within the bone marrow (BM) and can be mobilized into the circulation by chemotherapy or cytokine treatment [27]. The most common mobilizer is the granulocyte-colony stimulating factor (G-CSF). GCSF-mobilized HSCs are increasingly used in stem cell transplantation (SCT) for the relative ease of collection, the higher yield and the shorter time to engraftment than BM stem cells [28].

We have demonstrated that 67LR expression is increased in G-CSF-mobilized HSCs, as compared with BM HSCs, and significantly correlated with mobilization efficiency [29]. During G-CSF–induced HSC mobilization, the expression of laminin receptors switched from α6 integrins, which mediated LM-dependent adhesion of steady-state human BM HSCs, to 67LR, responsible for G-CSF–mobilized HSC migration toward LM. This switch in the expression of LM receptors also induced a change in the signal transduction pathway activated in response to LM binding. In vitro G-CSF treatment, alone or combined with exposure to marrow-derived endothelial cells, induced 67LR up-regulation in marrow HSCs; moreover, anti-67LR antibodies significantly inhibited transendothelial migration of G-CSF–stimulated marrow HSCs. Finally, G-CSF–induced mobilization in mice was associated with 67LR up-regulation both in circulating and marrow HSCs, and anti-67LR antibodies significantly reduced HSC mobilization.

Engraftment of HSCs to BM after transplantation is a key factor in SCT. G-CSF-induced 67LR overexpression on G-CSF mobilized HSCs, could play a crucial role also in HSC homing back to BM by mediating interactions with the basement membranes of vascular endothelia and subsequent cell migration. Indeed, 67LR also promoted homing to BM of transplanted HSCs, playing a key role in erythroid progenitor and precursor cells lodgment within the BM [30]. Thus, 67LR overexpression occurs in BM and circulating normal HSCs after cytokine stimulation and regulates HSC trafficking from and to BM. These findings further support a model in which HSC mobilization could represent a physiologic counterpart of leukemic and metastatic cell spread.

IV. 37LRP/67LR EXPRESSION AND TARGETING IN CHRONIC LYMPHATIC LEUKEMIA

B-cell chronic lymphocytic leukemia (CLL) is a heterogeneous group of diseases with various B-cell membrane markers expression and clinical course [31]. Despite the identification of genetic and phenotypic markers that correlate with prognosis, the biological basis of this clinical variability remains unclear [32].

In CLL, 37LRP/67LR is widely expressed and 37LRP is considered as an oncofetal antigen (OFA), thus often referred to as OFA/iLRP (oncofetal antigen/immature laminin receptor) [33]. Oncofetal antigens are conserved tumor-associated antigens or transplantation antigens expressed on the surface of human tumors and on fetal cells but not on normal adult tissues. OFAs are able to induce an immune response against tumors as well as a tolerogenic response, linked to cancer progression [34]. Dendritic cells (DCs) primed with OFA/iLRP or transfected with RNAs specific to OFA/iLRP induced a T-cell immune response against hematological malignancies, in particular acute myeloid leukemia (AML) and chronic lymphatic leukemia (CLL) cells. In a murine B-cell lymphoma model, treatment with syngeneic DCs transfected with OFA/iLRP-coding RNA resulted in powerful antitumor effect [35]. There is also
evidence of a humoral response against OFA/iLRP: pre-existing antibodies (Abs) to OFA/iLRP have been detected in sera of CLL patients. Patient Abs to OFA/iLRP were cytotoxic in vitro and individuals with an anti-OFA/iLRP humoral response had a more favorable prognosis. OFA/iLR Abs were cytotoxic and exerted also a role in the graft-vs-leukemia effect in CLL [36].

Confirming its nature of immune stimulating tumor associated antigen, high expression of the protein OFA/iLR correlated with mutated IGVH status and predicted for a favorable prognosis in CLL [37]. These results are in agreement with reports on the ability of anti-37LRP/67LR monoclonal antibodies (MoAbs) to block neoplastic B cell proliferation in vitro and in vivo. Two MoAbs, BV-15 and BV-27, showed anti-metastatic activity in the A20 B-cell leukemia model. Only BV-27 was growth-suppressive in vitro; however, both antibodies suppressed A20 cell attachment to LM [38]. Thus, inhibition of LM attachment seems crucial for the inhibitory effect, as reported with antibodies targeting both the immature and mature forms of the receptor or with treatments that down-regulate the expression of 37LRP/67LR in solid tumors [19,39]. These MoAbs could be used therapeutically even though it is not clear whether they exert their action by effector functions (Ab or complement dependent cytotoxicity) or by their action (cell growth inhibition and/or blocking of cell binding to LM).

Epigallocatechin-3-gallate (EGCG) is the major polyphenol of green tea; it is a small molecule that functions as an antitumor and antiangiogenic agent. EGCG induces cell death and cell cycle arrest and 67LR was identified as a receptor able to mediate its anti-cancer activity [40]. In contrast with previous results showing that 67LR inhibition, through MoAbs, blocked neoplastic B lymphocyte proliferation, a phase II clinical trial demonstrated that 67LR stimulation by oral Polyphenon™ was well tolerated and 29 of 43 CLL patients (67%) showed evidence of a biological response with decreased lymphadenopathy and/or absolute lymphocyte count [41]. Moreover, there was a significant correlation between EGCG susceptibility and 67LR expression in CLL cells and Vardenafil, a clinically available phosphodiesterase inhibitor, potentiated the killing effect of EGCG on CLL cells [42]. The molecular mechanism of Vardenafil action on EGCG-induced 67LR stimulation and tumor cell killing was better elucidated in multiple myeloma cells (see below).

V. 37LRP/67LR EXPRESSION AND FUNCTION IN MULTIPLE MYELOMA

Multiple myeloma (MM) represents a B cell malignancy, characterized by a monoclonal proliferation of malignant plasma cells. During disease evolution, terminally differentiated B cells preferentially accumulate in the BM. LN stimulated in vitro migration of human and murine MM cells, through its binding to 67LR,
overexpressed on MM cells. 67LR inhibition by the LM-derived peptide CDPGY1GSR, resulted in a decreased homing of MM cells to the BM in a murine in vivo model [43]. Thus, LM acts as a chemotactant for MM cells by interaction with 67LR and this interaction might be important during the trafficking of MM cells, as already demonstrated for normal HSCs [29,30]. 67LR is also involved in lymphoma cells homing to lymph nodes and in their trafficking to specific organs [44].

In MM, EGCG was able to induce inhibition of cell growth and apoptosis in vitro and in vivo. Silencing of 67LR resulted in abrogation of EGCG-induced apoptosis, confirming the role of 67LR in EGCG-mediated growth inhibition in MM cells [45]. EGCG induced apoptosis through 67LR by determining phosphorylation of PKCδ and activation of acid sphingomyelinase (aSMase). cGMP is a critical mediator of 67LR-dependent PKCδ/aSMase activation and MM apoptosis. EGCG induces nitric oxide (NO) production through 67LR-dependent activation of Akt and endothelial nitric oxide synthase (eNOS). NO increases the intracellular level of cGMP, that induces apoptosis by activating PKCδ/aSMase pathway. aSMase acts on sphingomyelin [46] to generate ceramide, which induced lipid rafts clustering, critical for apoptosis. Orally administered EGCG activated PKCδ and aSMase in a murine MM xenograft model [47]. In MM cells, phosphodiesterase 5 (PDE5), a major negative regulator of cGMP, is overexpressed and is able to reduce 67LR-mediated apoptosis induced by EGCG. Thus, Vardenafil, a PDE5 inhibitor, induced an enhancement of the EGCG-activated 67LR-dependent apoptosis, through amplification of the downstream effectors PKCδ and aSMase, and prolongation of the survival time in a mouse xenograft model [46,47].

VI. 37LRP/67LR EXPRESSION AND FUNCTION IN ACUTE MYELOID LEUKEMIA

Acute myeloid leukaemia (AML) is an aggressive blood cancer caused by the proliferation of immature myeloid cells. The genetic abnormalities underlying AML affect signal transduction pathways, transcription factors and epigenetic modifiers. The genetic landscape of AML cells could exert a direct effect on the anti-leukemic immune responses [48]. Thus, 67LR expression and function in AML could play a critical role in the evolution and prognosis of the disease.

We detected enhanced 67LR expression in 40% of 53 de novo AMLs, which frequently exhibited monocytic or myelomonocytic morphology. We did not detect 67LR expression in normal BM hematopoietic cells, in precursor-B acute lymphoblastic leukemia, in chronic lymphocytic leukemia, or in chronic myeloid leukemia in chronic phase. 67LR overexpression corresponded to a higher adhesion to LM. In contrast with 67LR behavior in solid tumors, no statistically significant difference was found between 67LR expression and any hematological characteristic of the disease at diagnosis, nor between 67LR expression and outcome of the disease as measured by complete remission rate, disease-free survival, or overall survival [49].

A more recent study demonstrated 67LR expression influenced the characteristics of AML cells toward an aggressive phenotype and increased the expression of GM-CSF receptor. Indeed, increased expression of 67LR was significantly related to elevation of white blood cell count, lactate dehydrogenase, and poorer survival among AML patients. Forced expression of 37LRP/67LR enhanced proliferation, cell-cycle progression, and antiapoptosis of AML cells associated with phosphorylation of STAT5, in the absence of stimulation LM. There was a significant relationship between the expression of 67LR and GM-CSFR in acute myeloid leukemia samples, with enhanced GM-CSF signaling [50].

This observation is not surprising; indeed, a previous work showed that 67LR was an interacting protein of both the alpha and beta subunits of GM-CSFR. Whereas GM-CSF functions by engaging the alpha and beta subunits into receptor complexes, 67LR inhibited GM-CSF-induced receptor complex formation. 67LR engagement by LM relieved the LR inhibition of GM-CSF. These findings provided a mechanistic basis for enhancing host defense cell responsiveness to GM-CSF at transendothelial migration sites, where 67LR is engaged by LM, while suppressing it in circulation [51].

EGCG-induced cell death through cGMP/aSMase axis activation and lipid raft clustering was described also in AML [52] and in chronic myeloid leukemia (CML) [53].

VII. CONCLUSIONS

Mobilization of HSCs into the blood following treatment with chemotherapy or cytokines mimics the enhancement of the physiologic stem-cell release in response to stress and inflammatory signals and results from changes in the adhesion profile of HSCs, facilitating their egress from BM. Cytokine-stimulated HSCs, leukemia and multiple myeloma cells, as metastatic cells from solid tumors, activate a 67LR-derived signaling pathway, which leads to cell dissemination and trafficking through the host.

36LRP/67LR is overexpressed in CLL and AML, but with a different prognostic impact. 37LRP/67LR overexpression in CLL has a positive impact, due the stimulation of an anti-leukemia effect. The same effect is not observed in AMLs in which 67LR upregulation is correlated, as in solid tumors, to increased aggressiveness and poorer response to treatments.

Biological agents inhibiting 67LR binding to LM, such as antibodies and peptides, could represent an efficient tool to target CLL and myeloma cells and the activity of EGCG in CLL has been already proved by an early clinical trial. Concurrently, many reports indicate that EGCG activity through 67LR may help in the design of new strategies to also treat AML and MM.
It is intriguing that a 67LR stimulating small molecule, such as EGCG, can exert the same antitumor effect of 67LR inhibitory molecules. Most probably, clarification of the molecular mechanism of action of new small molecules inhibiting LM binding to 67LR, such as NSC47924 and its analogs, will help to clarify this issue.

REFERENCES

[1] Montuori N, Sobel ME. The 67-kDa laminin receptor and tumor progression. Curr Top Microbiol Immunol 1996;213(Pt1):205-214.
[2] Rao CN, Castronovo V, Schmitt MC, Wewer UM, Claysmith AP, Liotta LA, Sobel ME. Evidence for a precursor of the high affinity metastasis-associated murine laminin receptor. Biochemistry 1989;28(18):7476-7486.
[3] Landowski TH, Dratz EA, Starkey JR. Studies of the structure of the metastasis-associated 67 kDa laminin binding protein: fatty acid acylation and evidence supporting dimerization of the 32 kDa gene product to form the mature protein. Biochemistr. 1995;34(35):11276-11287.
[4] Butô S, Tagliaabue E, Ardini E, Magnifico A, Ghirelli C, van den Brûle F, Castronovo V, Colnaghi MI, Sobel ME, Ménard S. Formation of the 67-kDa laminin receptor by acylation of the precursor. J Cell Biochem 1998;69(3):244-251.
[5] Di Giacomo V, Meruelo G. Looking into laminin receptor: critical discussion regarding the non-integrin 37/67-kDa laminin receptor/RPSA protein. Biol Rev Camb Philos Soc 2016;91(2):288-310.
[6] Sarnataro D, Pepe A, Altamura G, De Simone I, Pesapane A, Nitsch L, Lavecchia A, Zurzolo C. The 37/67 kDa laminin receptor (LR) inhibitor, NSC47924, affects 37/67 kDa LR cell surface localization and interaction with the cellular prion protein. Sci Rep 2016;6:24457.
[7] Ardini E, Tagliaabue E, Magnifico A, Butô S, Castronovo V, Colnaghi MI, Ménard S. Co-regulation and physical association of the 67-kDa monomeric laminin receptor and the α6β4 integrin. J Biol Chem 1997;272(4):2342-2345.
[8] Berno V, Porrini D, Castiglioni F, Campiglio M, Casalini P, Pupa SM, Balsari A, Menard S, Tagliaabue E. The 67 kDa laminin receptor increases tumor aggressiveness by remodeling laminin-1. Endocr Relat Cancer 2005;12(3):393-406.
[9] Kazmin DA, Hoyt TR, Taubner L, Teintze M, Starkey JR. Phage display mapping for peptide 11 sensitive sequences binding to laminin-1. J Mol Biol 2000;298(3):431-445.
[10] Castronovo V, Taraboletti G, Sobel ME. Functional domains of the 67kDa laminin receptor precursor. J Biol Chem 1991;266(30):20440-20446.
[11] Massia SP, Rao SS, Hubbell JA. Covalently immobilized laminin peptide Tyr-Ile-Gly-Ser-Arg (YIGSR) supports cell spreading and co-localization of the 67-kilo dalton laminin receptor with alpha-actinin and vinculin. J Biol Chem 1993;268(11):8053-8059.
[12] Omar A, Jovanovic K, Da Costa Dias B, Gonsalves D, Moodley K, Caveney R, Mbazima V, Weiss SF. Patented biological approaches for the therapeutic modulation of the 37 kDa/67 kDa laminin receptor. Expert Opin Ther Pat 2011;21(1):35-53.
[13] Menard S, Tagliaabue E, Colnaghi MI. The 67kDa laminin receptor as a prognostic factor in human cancer. Breast Cancer Res Treat 1998;52(1-3):137-145.
[14] Montuori N, Müller F, De Riu S, Fenzì G, Sobel RE, Rossi G, Vitale M. Laminin receptors in differentiated thyroid tumors: restricted expression of the 67-kilodalton laminin receptor in follicular carcinoma cells. J Clin Endocrinol Metab 1999;84(6):2086-2092.
[15] Song T, Choi CH, Cho YJ, Sung CO, Song SY, Kim TJ, Bae DS, Lee JW, Kim BG. Expression of 67-kDa laminin receptor was associated with tumor progression and poor prognosis in epithelial ovarian cancer. Gynecol Oncol 2012;125(2):427-432.
[16] Liu L, Sun L, Zhao P, Yao L, Jin H, Liang S, Wang Y, Zhang D, Pang Y, Shi Y, Chai N, Zhang H, Zhang H. Hypoxia promotes metastasis in human gastric cancer by up-regulating the 67-kDa laminin receptor. Cancer Sci 2010;101(7):1653-1660.
[17] Omar A, Reusch U, Knockmuss S, Little M, Weiss SF. Anti-LRP/LR-specific antibody IgG1-iS18 significantly reduces adhesion and invasion of metastatic lung, cervix, colon and prostate cancer cells. J Mol Biol 2012;419(1-2):102-109.
[18] Taraboletti G, Belotti D, Giavazzi R, Sobel ME, Castronovo V. Enhancement of metastatic potential of murine and human melanoma cells by laminin receptor peptide G: attachment of cancer cells to subendothelial matrix as a pathway for hematogenous metastasis. J Natl Cancer Inst 1993;85(3):235-240.
[19] Jovanovic K, Chetty CJ, Khumalo T, Da Costa Dias B, Ferreira E, Malindisa ST, Caveney R, Letsolo BT, Weiss SF. Novel patented therapeutic approaches targeting the 37/67 kDa laminin receptor for treatment of cancer and Alzheimer's disease. Expert Opin Ther Pat 2015;25(5):567-582.
[20] Zuber C, Knockmuss S, Zemora G, Reusch U, Vlasova E, Diehl D, Mick V, Hoffmann K, Nikles D, Fröhlich T, Arnold GJ, Brenig B, Wolf E, Lahm H, Little M, Weiss S. Invasion of tumorigenic HT1080 cells is impeded by blocking or downregulating the 37-kDa/67-kDa laminin receptor. J Mol Biol 2008;378(3):530-539.
[21] Khusai R, Da Costa Dias B, Moodley K, Penny C, Reusch U, Knockmuss S, Little M, Weiss SF. In vitro inhibition of angiogenesis by antibodies directed against the 37 kDa/67 kDa Laminin Receptor. PLoS One. 2013; 8(3):e58888.
[22] Moodley K, Weiss SF. Downregulation of the non-integrin laminin receptor reduces cellular viability by inducing apoptosis in lung and cervical cancer cells. PLoS One 2013; 8(3):e57409.
[23] Formisano P, Ragno P, Pesapane A, Alfano D, Alberobello AT, Rea VE, Giusto R, Rossi FW, Beguinot
F. Rossi G, Montuori N. PED/PEA-15 interacts with the 67 kD laminin receptor and regulates cell adhesion, migration, proliferation and apoptosis. J Cell Mol Med 2012;16(7):1435–1446.

[24] Naidoo K, Malindisa ST, Otaar TC, Bernert M, Da Costa Dias B, Ferreira E, Reusch U, Knackmuss S, Little M, Weiss SF, Letsolo BT. Knock-Down of the 37kDa/67kDa Laminin Receptor LRP/LR Impedes Telomerase Activity. PLoS One 2015;10(11): e0141618.

[25] Lavecchia A, Di Giovanni C. Virtual screening strategies in drug discovery: a critical review. Curr Med Chem 2013;20(23):2839–2860.

[26] Pesapan A, Di Giovanni C, Rossi FW, Alfano D, Formisano L, Ragno P, Selleri C, Montuori N, Lavecchia A. Discovery of new small molecules inhibiting 67 kDa laminin receptor interaction with laminin and cancer cell invasion. Oncotarget 2015;6(20):18116-18133.

[27] Pelus LM. Peripheral blood stem cell mobilization: new regimens, new cells, where do we stand. Curr Opin Hematol 2008;15(4):285-292.

[28] Battiwalla M, McCarthy PL. Filgrastim support in allogeneic HSCT for myeloid malignancies: a review of the role of G-CSF and the implications for current practice. Bone Marrow Transplant 2009;43(5):351-356.

[29] Selleri C, Ragno P, Ricci P, Visconte V, Scarpati N, Carriero MV, Rotoli B, Rossi G, Montuori N. The metastasis-associated 67-kDa laminin receptor is involved in G-CSF- induced hematopoietic stem cell mobilization. Blood 2006;108(7):2476-2484.

[30] Bonig H, Chang KH, Nakamoto B, Papayannopoulou T. The p67 laminin receptor identifies human erythroid progenitor and precursor cells and is functionally important for their bone marrow loddment. Blood 2006;108(4):1230-1233.

[31] Gentile M, Zirlik K, Ciolli S, Mauro FR, Di Renzo N, Mastrullo L, Angrilli F, Molica S, Tripepi G, Giordano A, Di Raimondo F, Selleri C, Coscia M, Musso M, Orsucci L, Mannina D, Rago A, Giannotta A, Ferrara F, Herzihanu Y, Shvidel L, Tachibana H, Shanafelt TD, Tachibana H. Vardenafil, a phosphodiesterase inhibitor for the treatment of oncofetal antigen/immature laminin receptor protein in pregnancy and cancer. J Cell Mol Biol Lett 2014;19(3):393-406.

[32] Giudice A, Musto P, De Feo V, Malindisa ST, Otgaar TC, Barsoum AL, Schwarzenberger PO. Oncofetal antigen/immature laminin receptor protein in pregnancy and cancer. J Cell Mol Biol Lett 2014;19(3):393-406.

[33] Siegel S, Wagner A, Kabelitz D, Marget M, Coggjin J Jr, Barsoum A, Rohrer J, Schmitz N, Zeis M. Induction of cytotoxic T-cell responses against the oncofetal antigen/immature laminin receptor for the treatment of hematologic malignancies. Blood 2003;102(13):4416-4423.

[34] Friedrichs B, Siegel S, Kloess M, Barsoum A, Coggjin J Jr, Rohrer J, Jakob I, Tienman M, Heidorn K, Schulte C, Kabelitz D, Steinmann J, Schmitz N, Zeis M. Humoral immune responses against the immature laminin receptor protein show prognostic significance in patients with chronic lymphocytic leukemia. J Immunol 2008;180(9):6374-6384.

[35] Friedrichs B, Siegel S, Reimer R, Barsoum A, Coggjin J Jr, Kabelitz D, Heidorn K, Schulte C, Schmitz N, Zeis M. High expression of the immature laminin receptor protein correlates with mutated IGHV status and predicts a favorable prognosis in chronic lymphocytic leukemia. Leuk Res 2011; 35(6): 721-729.

[36] McClintock SD, Warner RL, Ali S, Chekuri A, Wang MT, Attili D, Kribbs RK, Aslam MN, Sinkule J, Morgan AC, Barsoum A, Smith LB, Beer DG, Johnson KJ, Varani J. Monoclonal antibodies specific for oncofetal antigen/immature laminin receptor protein: Effects on tumor growth and spread in two murine models. Cancer Biol Ther 2015;16(5): 724-732.

[37] Chetty C, Khumalo T, Da Costa Dias B, Reusch U, Knackmuss S, Little M, Weiss SF. Anti-LRP/LR specific antibody IgG1-iS18 impedes adhesion and invasion of liver cancer cells. PLoS One 2014;9(5):e96268.

[38] Tachibana H, Koga K, Fujimura Y, Yamada K. A receptor for green tea polyphenol EGCG. Nat Struct Mol Biol 2004;11(4):380-381.

[39] Shanafelt TD, Call TG, Zent CS, Leis JF, LaPlant B, Bowen DA, Roos M, Laumann K, Ghosh AK, Lesnick C, Lee MJ, Yang CS, Jelinek DF, Erlichman C, Kay NE. Phase 2 trial of daily, oral Polyphenon E in patients with asymptomatic, Rai stage 0 to II chronic lymphocytic leukemia. Cancer 2013;119(2):363–370.

[40] Kumazoe M, Tsukamoto S, Lesnick C, Kay NE, Yamada K, Shanafelt TD, Tachibana H, Vardenafil, a clinically available phosphodiesterase inhibitor, potentiates the killing effect of EGCG on CLL cells. British J Haematol 2014;168(4):610-611.

[41] VandeBroek I, Vanderkerken K, De Greef C, Assouini G, Straetmans N, Van Camp B, Van Riet I. Laminin-1-induced migration of multiple myeloma cells involves the high-affinity 67 kD laminin receptor. Br J Cancer 2001;85(9):1387-1395.

[42] Chen A, Ganor Y, Rahimipour S, Ben-Aroya N, Koch Y, Levine M. The neuropeptides GnRH-II and GnRH-I are produced by human T cell and trigger laminin receptor gene expression, adhesion, chemotaxis and inducing immunogens on primary rodent and human cancers. Anticancer Res 1999; 19(6C): 5532-5542.
homing to specific organs. Nat Med 2002;8(12):1421-1426.

[45] Shammas MA, Neri P, Koley H, Batchu RB, Bertheau RC, Munshi V, Prabhala R, Fulciniti M, Tai YT, Treon SP, Goyal RK, Anderson KC, Munshi NC. Specific killing of multiple myeloma cells by (-)-epigallocatechin-3-gallate extracted from green tea: biologic activity and therapeutic implications. Blood 2006;108(8):2804-2810.

[46] Tsukamoto S, Hirotsu K, Kumazoe M, Goto Y, Sugihara K, Suda T, Tsurudome Y, Suzuki T, Yamashita S, Kim Y, Huang Y, Yamada K, Tachibana H. Green tea polyphenol EGCG induces lipid-raft clustering and apoptotic cell death by activating protein kinase Cδ and acid sphingomyelinase through a 67 kDa laminin receptor. Biochem J 2012;443(2):525-534.

[47] Kumazoe M, Sugihara K, Tsukamoto S, Huang Y, Tsurudome Y, Suzuki T, Suemasu Y, Ueda N, Yamashita S, Kim Y, Yamada K, Tachibana H. 67-kDa laminin receptor increases cGMP to induce cancer-selective apoptosis. J Clin Invest 2013;123(2):787-799.

[48] Austin R, Smyth MJ, Lane SW. Harnessing the immune system in acute myeloid leukaemia. Crit Rev Oncol Hematol 2016;103:62-77.

[49] Montuori N, Selleri C, Risitano AM, et al. Expression of the 67-kDa Laminin Receptor in acute myeloid leukemia cells mediates adhesion to laminin and is frequently associated with monocytic differentiation. Clin Cancer Res 1999;5(6):1465-1472.

[50] Ando K, Miyazaki Y, Sawayama Y, Tominaga S, Matsuo E, Yamasaki R, Inoue Y, Iwanaga M, Imanishi D, Tsushima H, Fukushima T, Imaizumi Y, Taguchi J, Yoshida S, Hata T, Tomonaga M. High expression of 67-kDa laminin receptor relates to the proliferation of leukemia cells and increases expression of GM-CSF receptor. Exp Hematol 2011;39(2):179-186.

[51] Chen J, Cárcamo JM, Bórquez-Ojeda O, Erdjument-Bromage H, Tempst P, Golde DW. The laminin receptor modulates granulocyte-macrophage colony-stimulating factor receptor complex formation and modulates its signaling. Proc Natl Acad Sci USA 2003;100(24):14000-14005.

[52] Kumazoe M, Kim Y, Bae J, Takai M, Murata M, Suemasu Y, Sugihara K, Yamashita S, Tsukamoto S, Huang Y, Nakahara K, Yamada K, Tachibana H. Phosphodiesterase 5 inhibitor acts as a potent agent sensitizing acute myeloid leukemia cells to 67-kDa laminin receptor-dependent apoptosis. FEBS Lett 2013;587(18):3052-3057.

[53] Huang Y, Kumazoe M, Bae J, Yamada S, Takai M, Hidaka S, Yamashita S, Kim Y, Won Y, Murata M, Tsukamoto S, Tachibana H. Green tea polyphenol epigallocatechin-O-gallate induces cell death by acid sphingomyelinase activation in chronic myeloid leukemia cells. Oncol Rep 2015;34(3):1162-1168.