Role of Macrophages in Immunity

Tissue macrophages (MΦs) and blood-recruited monocytes have remarkable immune plasticity, with the ability to sense and adapt to the local milieu. MΦ phenotype is defined by the surrounding environment, which ultimately is controlled by the host. These environmental contributions to MΦ plasticity, in turn, contribute to the efficiency and potency of their involvement in inflammation and host homeostasis including host defense, the initiation and resolution of inflammation, tissue repair and the removal of dead cells and tissue debris. The plasticity and phenotypic response to the environment has led to the description of M1 MΦs and an entire series of M2 MΦs with variable surface expression of scavenger receptors, pathogen recognition receptors, autoregulatory machinery and secretion of inflammatory/anti-inflammatory molecules [1].

In the lung, 2 major MΦ populations have been described: alveolar (AMs), which are located in the airway lumen, and interstitial, which reside in the lung parenchyma. The AMs, distinguished by their unique expres-
sion pattern (CD11c-pos; CD11b-neg), are the sentinels of the lung. They maintain immunological and physiological homeostasis (e.g. the removal of debris and the recycling of surfactant molecules) and provide a first line of host defense [1]. They are inherently suppressive, in order to protect the lung from inflammation due to environmental perturbations. Once the MΦs are activated, they rapidly amplify the host response via the secretion of antimicrobials and proinflammatory mediators, by recruiting specialized phagocytes (e.g. neutrophils) and by communicating with other lung cells (e.g. alveolar cells [1]) for a rapid amplified response. The interstitial MΦs (CD11c-low; CD11b-pos) are less characterized, being highly heterogeneous, with the potential for regulatory control. Indeed, it has been suggested that these MΦs are major contributors to the production of cytokines (IL-10) associated with the adaptive immune response. During infections, activation of the proinflammatory process and release of neutrophil granule proteins (e.g. azurocin, LL-37 and cathepsin G) will trigger the recruitment of monocytes from the circulation, aiding in the fight against infection, the removal of dead cells and, eventually, the resolution of the ensuing lung inflammatory response [2].

MΦs express a variety of receptors (plasma membrane or intracellular) and proteins which make them capable of sensing changes in the environment. These receptor interactions allow the MΦs to rapidly respond to the presence of microorganisms (e.g. bacteria, viruses and fungi), changes in physiology (e.g. changes in pH, hypoxia and osmolarity), metabolite concentrations (e.g. ATP, fatty acids, heme, etc.) and extracellular matrix alterations (e.g. collagen degradation products and hyaluronic acid). When tissue homeostasis is perturbed, these receptors are activated, leading to a specific signaling transduction and the expression of mediators that will allow cross-talk with neighboring cells and the recruitment of immune cells which, together, will cooperate in reestablishing the lung status quo [3].

MΦs sense microorganisms by recognizing pathogen-associated molecules or PAMPs, e.g. lipopolysaccharide (LPS), dsRNA and flagellin, through a repertoire of receptors called pattern-recognition receptors (PRRs). PRRs are expressed on immune and structural cells, including airway epithelium. They are divided into 4 major families: Toll-like receptors (TLRs), nucleotide oligomerization (NOD) receptors, c-type lectin receptors and retinoic acid-inducible gene 1 (RIG-I) receptors. Each of these receptor systems recognizes unique and diverse pathogen-associated molecules. PRRs are also activated by damaged cells that dump their cytoplasmic and nuclear components (e.g. HMGB1, ATP and adenosine) into the extracellular milieu; these are the ‘damage-associated molecular patterns’ (DAMPs) of the inflammatory response. Once activated, PRRs, through a tightly regulated signal transduction mechanism, initiate the inflammatory response by producing inflammatory mediators, e.g. proinflammatory cytokines, reactive oxygen species (ROS), nitric oxide, carbon monoxide and antimicrobials, which lead to pathogen elimination, inflammation resolution and, eventually, the reestablishment of tissue homeostasis. Abundance, location, turnover and signal transduction regulation of these receptors will determine the quality, intensity and duration of the immune response [4].

In addition to external effector functions, MΦs are also professional phagocytes, comprising the major mechanisms associated with the immune-regulated removal of pathogens, dead/dying cells and debris, and, in the case of AMs, are involved in surfactant homeostasis. Phagocytosis is a complex mechanism that involves an extraordinary cytoskeleton and plasma membrane reorganization, which allows MΦ chemotaxis and receptor-mediated binding to the inciting material to be phagocytized. Once internalized, elimination of the ingested material will be mediated by the fusion of phagosome with lysosomes. Thus, the MΦ ability to quickly remove microorganisms will depend on the activation of a complex transcriptional response, which encompasses cytoskeleton dynamics and vesicle trafficking/fusion, the production of lysosomal enzymes and the proton-pump acidification of lysosomal compartments. MΦ PRRs and other plasma membrane receptors, such as complement Fc and scavenger receptors, contribute to binding and responding to invading microorganisms [2].

The extensive role of MΦs in the pulmonary host response makes it an essential participant in lung homeostasis which, when dysfunctional, contributes to several human lung diseases including obstructive lung disease, asthma and allergic airways disease and fibrotic lung diseases [5].

**Monocyte/MΦ Alterations in CF Are due to both Intrinsic and Acquired Factors**

The hypothesis that monocytes/MΦs may contribute to CF lung disease was first proposed in 1982 [6]. CF MΦ dysfunction was associated with altered activation defined by metabolic hyperactivity, with elevated production of proinflammatory cytokines [7], elastase [8] and tissue-damage mediators (termed ciliary dyskinesia sub-
stances). These mediators impact airway epithelial-cell ciliary movements, contributing to the accumulation of mucus secretion [9]. Not until a few years later, with the discovery of the CFTR gene, was CFTR expression documented in MΦs [10].

Several descriptive studies using MΦs from CF patients have demonstrated that the MΦ phenotype changes during CF pathogenesis, as a result of plasticity. As already hypothesized 3 decades ago [11, 12], the CF lung environment (e.g. mucus, airway surface dehydration, increased protease activity, ROS, cytokines, etc.) plays a significant role in defining the phenotype of monocytes/MΦs, such that their ability to properly regulate the inflammatory response, clear bacteria and favor lung tissue repair is altered. Furthermore, it has been documented that there is an increase in the absolute numbers of MΦs in CF airways in the later stages of fetal development [13] and in young children with CF without detectable infection [14]. This increase has been correlated with elevated levels of the monocyte chemotactic protein 1 (MCP-1, also called CCL2) in the bronchoalveolar lavage fluid (BALF) and induced sputum from CF patients [14] and with lung exacerbations in patients with CF [15].

The remarkable ability of tissue MΦs to adapt to the environment and carry out different functions led to their broad classification as either classically activated M1 MΦs, implicated in initiating and sustaining inflammation, or alternatively activated M2 MΦs, associated with anti-inflammatory, immunoregulatory and tissue-repair properties. M2 MΦs can also contribute to fibrotic pathology and allergic conditions [1]. In CF, the contribution of MΦ polarization to the lung disease is still unclear. In in vitro M2 cells, polarization is highly dependent on IL-4/IL-13 signaling and the production of high levels of arginase. Hartl et al. [16] reported that BALF from patients with CF infected with Pseudomonas aeruginosa had higher IL-4 and IL-13 concentrations and lower levels of IFN-γ (an environment that favors M2 MΦ polarization) compared with uninfected patients. Further, BALF levels of IL-4 and IL-13 correlated inversely with FEV1 [16]. Arginase activity, which has been postulated to be an important mediator of airway remodeling and lung fibrosis, has been shown to be elevated in CF lungs [17], with a concurrent, increased expression of mannose receptor (CD206). These markers of lung fibrosis have been correlated with a decline in pulmonary function in P. aeruginosa-infected CF patients [18]. Increased arginase activity has also been identified in the lung and airways of Cfr-deficient mice that is further augmented by infection with P. aeruginosa [19]. The CF lung environment is highly complex, concurrently demonstrating a proinflammatory phenotype, with high levels of IL-8, IL-6 and TNF-α and low levels of IL-10, an environment that would favor M1 MΦ polarization instead [20, 21]. Furthermore, the expression of CD206 and other scavenger receptors (discussed later) has been found to be down-regulated in sputum-derived MΦs from CF patients [22], suggesting differences in the milieu of the lung and sputum. Animal studies have suggested that alveolar and peritoneal MΦs from F508del CF mice exposed to LPS have elevated levels of both M1and M2 MΦs compared to control Cfr-sufficient animals [23]. Thus, defining the contribution of MΦ polarization in CF lung disease may be challenging, since these cells dynamically adapt to the tissue environment and can vary with age, the status of lung disease, the bacterial flora and the therapeutic regimen that are unique to each patient.

The plasticity of the MΦ has made it difficult to completely appreciate the mechanisms associated with the changes in CF MΦs [10]. The CFTR protein has been documented in murine [24–26], ferret [27] and human [28, 29] monocytes/MΦs and associated with CFTR-like Cl− conductance abnormalities [28, 30, 31]. Treatment of MΦs with specific CFTR inhibitors have been shown to change the MΦ phenotype to resembles CF MΦs, with an increased secretion of proinflammatory cytokines [24, 30]. LPS-induced hypersecretion of IL-8 has also been demonstrated utilizing peripheral blood (PB) monocytes isolated from subjects heterozygous for the F508del CFTR mutation compared to non-CF controls [32], further supported by the same observations with MΦs isolated from heterozygous Cfr mice [26]. Thus, a single allelic CFTR mutation is sufficient to augment proinflammatory activation in response to LPS in CF, implying CFTR-dependent defects in CF MΦs.

**CF Monocytes/MΦs Are Hyperinflammatory**

Cytokines such as TNF-α, IL-1β, IL-6 and IL-8 are elevated in the lungs of patients with CF compared with healthy controls, while the secretion of cytokines involved in resolution of inflammation, such as IL-10, is reduced [20, 21]; this correlates with the expression in AMs and MΦs. Further, treatments that improve clinical parameters in patients with CF, such as the antibiotic, azithromycin, have been found to reduce the proinflammatory phenotype in AMs [33], and PB monocytes respond to CFTR potentiator therapy (ivacaftor) in patients carrying the G551D mutation [34–36].
The proinflammatory behavior of MΦs may be directly associated with chronic bacterial infection, which is constitutive once established in the lung of CF patients. In addition, the altered CF lung environment, which is rich in inflammatory mediators such as ROS, HMGB1, neutrophil proteases and cellular matrix proteolytic products (e.g. proline-glycine-proline and hyaluronan fragments) [37], may persistently activate the PRRs in MΦs (and other cell types in the lung) to optimize pathogen interaction and sensitivity as well as to stimulate proinflammatory pathways (e.g. NF-κB and MAPK). Mucus obstruction and changes in airway surface hydration [38], the hallmark of CF lung disease, may also affect the ability of MΦs to properly respond to inflammatory triggers or efficiently phagocytose the pathogen invader.

In recent studies, it has been demonstrated that the robust production of inflammatory mediators in human CF AMs may be due to the activation of the inositol-requiring enzyme 1 (IRE1)-α-dependent X-box-binding protein-1 (XBP-1) arm of the unfolded protein response (UPR). These studies demonstrate higher mRNA levels of XBP-1 in CF when compared to non-CF AMs. This response seems to reflect an adaptation to the infectious/inflammatory environment of CF airways rather than the loss of CFTR in CF MΦs [39]. The factor in the CF lung milieu that contributes to this dysregulation is unknown.

Another observation, justified by a wealth of research, is that CF MΦs are intrinsically hyperinflammatory. In support of this hypothesis, studies have demonstrated that in vitro cultures of MΦs isolated from the PB [30, 32, 40, 41] of CF patients have an exaggerated inflammatory response to several inflammatory mediators. Consistent with the in vitro human data, MΦs from CF mice and ferrets have also been shown to be proinflammatory [23, 25–27, 30, 41–43], with an increased production of proinflammatory cytokines when exposed to bacteria such as P. aeruginosa [42] and Burkholderia cepacia [40, 44, 45] and PAMPs such as LPS [23, 25, 26] and flagellin [41]. Furthermore, studies in which bone marrow (BM) chimeras were made by transplanting wild-type and CF mice with either wild-type or CF BM, demonstrated the enhanced secretion of various proinflammatory cytokines after exposure to LPS. This demonstrates that the proinflammation may be related to MΦs lacking functional CFTR rather than on the resident epithelial cells [26]. These data were consistent with studies on a model of P. aeruginosa infection using myeloid-specific Cfr knockout mice [42] and in G551D/G551D CF mice [46]. Thus, studies on animal models suggest that a loss of CFTR in MΦs/monocytes contributes to lung hyperinflammation in CF.

Since the identification of CF differences in response to pathogens, the search for how CFTR alters MΦ function and activity has been ongoing. One potential hypothesis relates to the altered plasma membrane expression of immune receptors (i.e. TLR4 [30, 47] and TLR5 [48]) involved in immune signaling. In particular, the increased expression of TLR4, which is activated by LPS and also by DAMPs such as HMGB1, have been reported in murine [30] and human [30, 47] MΦs. Furthermore, increased TLR4 levels lead to the sustained signal transduction and activation of the NF-κB and MAPK pathways, which induces a robust transcription of proinflammatory cytokines. The changes in TLR4 presence on the plasma membrane of CF MΦs may be due to an abnormal internalization of the receptor in the endosomal compartment and the trafficking to the lysosomal compartment, where it is degraded, thus terminating signaling [30]. The endosomal defects contributing to the inefficient trafficking of TLRs may be related to observed abnormalities in the reduced expression of Rab proteins [30, 49] as well as the rate of microtubule formation [50], both of which contribute to vesicle trafficking, docking and maturation. CF cells, including MΦs, have altered levels of sphingolipids (e.g. ceramide) and cholesterol [51–53], which are key for shaping the signaling platform at the plasma membrane, called the lipid raft. CF MΦs challenged with LPS also have impaired expression/distribution of the plasma membrane scaffolding-protein caveolin-1 [43], which facilitates the cellular transport of cholesterol to the plasma membrane in MΦs [54]. Thus, low levels of caveolin-1 in CF MΦs may affect the organization of lipid rafts during activation.

All these dysfunctions may have a tremendous impact on the regulation of immune receptor signaling.

The mechanisms mediating negative feedback of immune receptor signaling are also altered in CF MΦs. CF MΦs have abnormal expression of nuclear receptors, such as peroxisome proliferator-activated receptors (PPARs) and liver X receptors (LXR), which mediate fatty acid metabolism, a negative regulator of inflammation. MΦs isolated from Cfr−/− mice have low basal levels of PPARγ expression and attenuated LPS-driven induction of PPARδ and LXRα [25]. Cellular distribution of the heme-oxygenase 1 (HO-1) protein, a key stress response protein involved in balancing cellular redox status and inflammation (including TLR4-negative regulation) is also altered in murine and human CF MΦs exposed to LPS. In particular, HO-1 translocates to lipid rafts in a caveolin 1-dependent manner and catalyzes the local production of.
carbon monoxide (CO), which favors destabilization between TLR4 and its adaptor protein MyD88 (thus signaling termination), and this negative feedback mechanism is blunted in CF MΦs [43]. In addition, in response to TLR-MyD88 activation, CF MΦs display blunted PI3K/AKT signaling [41], which normally plays a key role in regulating immune function [55] and in downregulating levels of microRNAs that amplify the TLR4 signaling (e.g., miR-155, let7e, miR-125b and miR199a-5p) in MΦs.

Previous literature has suggested that both the mouse models of CF and the human disease have deficient autophagy mechanisms in their epithelial cells [56]. Autophagy is a conserved mechanism by which cells manage intracellular damage in order to maintain sustainability and efficiency. Furthermore, the process of autophagy allows for the recycling of membranes and cytosolic components for reuse. Deficiency in effective autophagy has been linked to a variety of diseases which have a part in the pathophysiology of chronic inflammation, including Crohn’s disease and neurodegeneration [57]. MΦ autophagy has been well established as a mechanism of sustaining homeostasis [58], and when homeostasis is defective, this is linked to ineffective MΦ function [57]. Importantly, a proper autophagy flux in activated MΦs has negative regulatory effects on TLR signaling, by controlling the signal transduction, trafficking and degradation of the receptor [57]. This autophagy dysfunction predisposes the CF MΦs to the elevated production of proinflammatory cytokines during *Burkholderia cepacia* infection [40].

Finally, a lack of CFTR in monocytes affects the Rho-small GTPase inside-out signaling triggered by monocyte/MΦ chemoattractant cytokines (e.g., MCP-1). This CFTR-dependent defect has been found to impair β-1 and β-2 integrin-mediated monocyte adhesion and chemotaxis. As a consequence, monocytes lacking CFTR accumulated in the lung parenchyma and displayed blunted transmigration into the BAL space of wild-type mice intranasally treated with MCP-1 [35]. This incapability of CF monocytes to localize appropriately in different lung compartments in response to chemoattractants may very well affect both the inflammatory response and the host defense. The mechanisms that could contribute to CF MΦs hyperinflammatory behavior are summarized in figure 1a.

**CF MΦs Display Inefficient Management of Infection**

Monocytes from CF patients have relatively low expression of the complement receptors urokinase-type plasminogen-activator receptor (uPAR) and CD11b, both important in facilitating the binding and phagocytosis of opsonized *P. aeruginosa* [29]. CF lungs also have pronounced expansion of small MΦs with low expression of the scavenger receptors, CD206 and MARCO, involved in the binding and internalization of unopsonized particles as well as microbes [22]. Caveolin-1, which mediates *P. aeruginosa* internalization [59], is expressed at low levels in activated CF MΦs [41, 43]. These observations suggest that monocytes/MΦs in CF may have an impaired ability to properly phagocytose bacteria [29].

The CF environment also impacts the capability of CF MΦs to recognize and internalize bacteria [11] by altering signaling [38, 60] and cleaving the receptors necessary to orchestrate the adequate host-pathogen interaction. In particular, it has been found that elevated levels of neutrophil elastase in CF lungs cleave plasma membrane receptors/proteins such as complement proteins (e.g., C3, C5 and C3bi), complement receptors (e.g. CR1) and lymphocyte receptors (CD4 and CD8) [12]. Fick et al. [61], in 1981, suggested that specific *P. aeruginosa* IgG in the serum and sputum of CF patients functioned in an inhibitory fashion, decreasing the efficiency of *P. aeruginosa* phagocytosis and intracellular killing. These early studies suggested that the Fab and Fc portions of the CF immunoglobulin molecule are impaired in their attachment to the alveolar MΦ membrane Fc-γ receptors, thus decreasing internalization. The phagocytic function of the complement receptors has also been shown to be defective in CF, with active elastase cleaving CR3 off the surface of phagocytes, contributing to inefficient infection resolution [62].

MΦs isolated from CF animal models (mice and ferrets) and CF patients also display an impaired capability for killing internalized bacteria, including *P. aeruginosa* and *Burkholderia cenocepacia*. Consistent with early studies that suggested a potential role for CFTR in maintaining a differential pH in the trans-Golgi, endosome and lysosome [63], Di et al. [24] proposed a mechanism by which a loss of CFTR in MΦs is associated with the alkalization of the phagosomal lumen, which impairs *P. aeruginosa* killing. The working hypothesis is that CFTR-mediated Cl⁻ entry functions as counter-ion conductance to balance H⁺ influx through V-ATPase in the lysosome, thereby maintaining an acidified lysosomal environment. These data were corroborated in MΦs isolated from CF
Mechanisms altered in CF MØs
1) Elevated levels of DAMPS in CF airways lead to persistent immune receptor signalling.
2) Abnormal lipid raft composition (e.g., altered distribution of cholesterol, sphingolipids and scaffolding proteins, which perturbs normal signal transduction regulation.)
3) Impaired autophagy affects TLR4 signal transduction.
4) Suboptimal vesicle trafficking, docking and maturation that impairs temporal and spatial localization of immune receptors, thus altering signalling intensity and duration. Altered Rab and SNARE protein expression and microtubule polarization may exacerbate this defect in CF.
5) Elevated activation of XBP-1 arm of the unfolded protein response (UPR).
6) PPARs – Dysregulation of PPAR results in constitutive NF-κB activation and cytokine production.
7) Altered chemotaxis affects MØ localization in the lung.

**OUTPUT IN CF LUNG**
- Increased secretion of proinflammatory mediators
- Sustained hyperinflammation
- Increase responsiveness to external stimuli
- Inefficient waste management creating accumulation of damages and dysfunctional cellular products

Mechanisms altered in CF MØs
1) A high level of proteases in CF airway cleaves receptors (e.g., complement, Fc or scavenger receptors) on MØs or on corpus to be phagocytes (e.g., complement, find/eat-me signals, IgG).
2) Abnormal lipid raft composition perturbs engulfment of phagocytic corpus.
3) Impaired autophagy affects phagosome maturation.
4) Suboptimal vesicle trafficking, docking and maturation may alter phagolysosome formation and/or autophagy flux, which are both necessary for bacterial killing.
5) Altered lysosomal function (e.g., pH regulation and ROS formation).
6) Decreased production of bactericidal molecules (e.g., CO, lysozyme, etc.).

**OUTPUT IN CF LUNG**
- Increased secretion of proinflammatory mediators
- Bacterial survival and proliferation
- Impaired initiation of the anti-inflammatory program
- Impaired capacity for directing the resolution of either infection or inflammation
patients [64] and in AMs from CF ferrets [27]. More recently, vesicle alkalinization also been correlated with the failure of CF MΦs to activate the acid sphingomyelinase enzyme, which is necessary for the formation of ceramide-enriched membrane platforms [52]. The ceramide facilitates gp91phox-mediated oxidative burst in response to *P. aeruginosa* infection, promoting killing [52]. An altered mechanism of vesicle acidification in CF MΦs was also proposed to favor intracellular survival of *B. cenocepacia* [44].

The major difficulty in all of these studies involving CFTR in lysosomal acidification has been the reproducibility of the observations, which has led to controversy about the theory of the dysfunctional lysosomal contribution of MΦs to CF pathophysiology. It is likely that the differences are not a matter of right or wrong, but more likely due to the complexity of MΦs and the exquisite sensitivity of the cell to minute changes in the environment of culture or purification [65, 66]. While this controversy has not yet been resolved, an accurate comparison of studies suggests that defective vesicle acidification in CF MΦs may depend on the complex cell signaling that is activated by MΦs in response to live bacteria, which cannot be fully recapitulated when phagocytosis is mimicked with opsonized beads [44]. In any case, whether or not CFTR directly controls lysosomal pH, a recent study suggests that drugs able to promote phagosome acidification, via potententiating the lysosomal activity of the transient receptor potential canonical-6 (TRPC6) calcium-permeable channel, can sufficiently restore microbicidal function in CF alveolar MΦs [67].

As discussed previously, autophagy, a mechanism first described to be defective in CF epithelia [56], has more recently been directly linked to the ineffective bactericidal function of MΦs in CF. Indeed, deficient autophagy prevents destruction of *B. cenocepacia* in murine CF MΦs [45, 68], and autophagy stimulation with rapamycin alleviates some of the dysfunction resulting in improved bacterial clearance of *B. cenocepacia*, *P. aeruginosa* and *Staphylococcus aureus* [45, 69, 70]. Unfortunately, rapamycin therapeutics are counter indicated for use in CF due to significant side effects, opening the door for therapeutic development targeting autophagy utilizing different drugs.

CF MΦs might be defective in releasing bactericidal mediators that contribute to the extraordinary role of keeping the lungs sterile. For example, as discussed in the previous section, the plasma membrane trafficking of HO-1 is blunted in activated CF MΦs [43]. HO-1 catalytic products have strong immune-modulatory effects, and, importantly, by producing CO, facilitate the killing of bacteria [71]. MΦs produce high levels of antimicrobial enzymes (e.g. lasozyme) and a loss of CFTR may also interfere with this basic MΦ function. The mechanisms that compromise the bactericidal function of CF MΦs are summarized in figure 1b.

**CF MΦs Have Reduced Scavenger Ability**

MΦ scavenger function is altered by the CF environment (e.g. mucus, increased proteases, ROS, etc.). Apoptosis and efferocytosis are in tandem in maintaining tissue homeostasis, with disruption in interactions contributing to inefficient inflammation resolution and tissue destruction [72]. Alveolar MΦs have been suggested to have deficient efferocytosis processes in a variety of diseases including CF [73]. The efficiency of the efferocytosis process is defined by MΦ phenotype, with the M1 phenotype having little or no capacity for the process while the M2 phenotype has a high capacity for efferocytosis [74]. Much of the regulator process of efferocytosis goes back to the inverse relationships between PPARγ and NF-κB again regulating the phenotypic characteristic of the contribution of MΦs to the inflammatory process. PPARγ drives the M2 phenotype whereas NF-κB drives the M1 phenotype [75]. The PPARs as retinoic acid-based lipid scavenger receptors are important in the process associated with surfactant and lipid metabolism [76]. Treatment of *Cftr*–/– AMs with endogenous PPARγ/α ligands, including rosiglitazone (PPARγ ligand) or WY14643 (PPARα ligand) decreases the LPS-induced TNF-α response [25, 77]. As PPARγ has been shown to be important for the maturation and phagocytic capacity of MΦs [78], these data would imply that the AM phenotype has the capacity to control proinflammatory cytokines via PPARγ in CF. The implication is that the change in the membrane lipids alters the functionality of the MΦs, which translates into inefficiency in both bacterial clearance and inflammation resolution. The role of these scavenger receptors is thought be important in the removal of foreign substances and waste materials that utilize extensive ligand specificity [79]. The absence of effective ‘clean-up’ and ‘removal’ systems can ultimately result in the accumulation of biologic waste, which interferes with homeostatic mechanisms. Therapeutic intervention aimed at improving scavenger receptor activity may provide support for the self-management of the unique milieu of the CF lung.

Removal of biologic waste in MΦs has been termed efferocytosis, which is the mechanism by which the MΦs
recognize phagocytes and dispose of apoptotic cells from the airway. However, elevated proteases in the CF lung alter efferocytosis efficiency. Proteases such as active elastase cleave MΦ phosphatidylylserine receptors; this impairs the capacity of MΦs in the recognition and phagocytosis of apoptotic cells in CF lungs [12, 80] (fig. 1b). Removal of dying apoptotic cells is fundamental for the resolution of the inflammatory response [81]. Neutrophil phagocytosis by MΦs triggers the production of mediators that curbs neutrophil migration and induces an anti-inflammatory transcriptional program in the MΦs (e.g. the expression of IL-10, lipoxins, resolvins, etc.). In addition, it favors a high level of expression of scavenger receptors (e.g. CD206 and MARCO) that play a pivotal role in the binding and internalization of particles in the absence of opsonization, which reduces the responsiveness to TLR ligands [2]. In CF, small MΦs with defects in the expression of CD206 and MARCO have been shown to be present in the CF airways (BALF and induced sputum) [22, 82]. The lower expression of these receptors on MΦs might contribute to an inability to properly clear inflammatory glycoproteins, oxidized lipids and inhaled particles that may be abundant in the damaged CF lung, thus enhancing the inflammation and tissue damage that is observed in the CF airways.

The MΦ products of the inflammatory response not only manage the infection but often contribute to tissue damage due to the potency of the agents such as reactive oxygen radicals, matrix metalloproteinases, cytokines and other protein-modifying agents [83]. Many of these products are produced through redirecting MΦ phenotype. Further recent data suggest that many of these changes can alter the MΦ activation process, impacting inflammatory regulator pathways, activity and phenotype via epigenomic modification [75, 84]. Once the lung manages the intrusion, the healthy lung starts the reparative process, which includes reducing the inflammation by ‘ramping up’ anti-inflammatory molecules and protease-neutralizing agents as well as products potentially generated by the alveolar macrophage. Thus, MΦ abnormality in CF may perpetuate lung tissue scarring due to the inefficient activation of anti-inflammatory and repairing pathways.

The CF Phagocyte and Adaptive Communication

MΦs, which can also function as professional antigen-presenting cells (APC), have been suggested to participate in redirecting downstream adaptive responses in CF [85, 86]. In these studies, shifting costimulatory molecules like CD80 and CD86 are suggested to alter T-cell reactivity and, potentially, the management of both infection and inflammation in CF. In this case, although the primary defect in the epithelial cells establishes the abnormal airway surface environment and initiates the pathophysiology that leads to progressive lung disease in CF, changes in both adaptive and innate immunity exacerbate the disease pathophysiology associated with infection and inflammation resolution. The inability to resolve infection and attenuate inflammation downstream of infection plays a major role in the morbidity and mortality of the disease. Furthermore, improving the ability to manage exposure to pathogens and the relenting inflammatory process appears to be individualized, even within families that have the same CFTR mutation variant [87] or monozygotic twins [88]. The concept of genomic and environmental modifiers contributing to the complexity to CF pathophysiology provides a significant insight for therapeutic development and the understanding of the variability of the disease and its subsequent response to treatment.

Genome-wide association studies have begun to demonstrate that although the CFTR gene is the causative factor for the development of CF pathophysiology, there are certain associated genes that tend to correlate with the severity of CF disease [89, 90]. Table 1 highlights some genes that are particularly relevant, specifically for how MΦs may ultimately regulate the downstream adaptive control of inflammation and infection resolution. Alpha-1 antitrypsin plays a role in innate immunity and has been the focus of studies in CF [91]. Bioactive mediators such as IL-10, C3, MIF, TGF-β, IFN-γ and TNF-α have also been shown to correlate with some aspects of lung function or clinical exacerbations according to the comprehensive review by Weiler and Drumm [90].

The control of pathogen response is based upon effective immune defenses. The immune system is composed of multiple cell types which, together, improve the resistance against infections. Communication between APC or phagocytes such as MΦs, their environment and downstream adaptive immunity is essential for an effective immune response. The studies of the CF genome-wide association project have shown that there is a significant correlation between MHC class II polymorphisms in the HLA-DR and HLA-DQ regions and overall disease severity [89, 92], implicating differences in the capacity to communicate to T cells and other adaptive immune cells. This is not unique to CF since there have been several instances of the association between specific MHC class II alleles and increased susceptibility or severity of inflam-
Inflammatory diseases, including allergic bronchopulmonary aspergillosis, multiple sclerosis, rheumatoid arthritis, sarcoidosis, asthma and diabetes [93, 94]. The interaction between T cells and MHC class II, along with the surrounding milieu, is crucial for defining the phenotype and success of the inflammatory response to infection. The host response to bacterial infection requires communication between APC and T cells [95]. This communication is relayed through MHC class II antigen presentation to helper T cells followed by adaptive T-cell or B-cell responses. Furthermore, the impact of HLA expression on MΦs has the capacity to change downstream adaptive antigen presentation-dependent events, and decreased expression of HLA-DQ and HLA-DR is described in CF monocytes/MΦs [96]. In traditional immunology, antigen presentation by APC, such as MΦs and dendritic cells, results in adaptive networks including T-cell immunity and B-cell activation [93]. Recently, it has become increasingly evident that B cells are not only responders to T-cell help, but, in exchange, are important programmers of the CD4 T-cell response including the priming and induction of T-cell memory [97]. In patients with CF, there is significant evidence supporting the concept of inefficient immune adaptive functions including the effector functions of NK cells [98], B cells [99] and T-cell abnormalities [86, 100] ultimately also contributing to the ineffective management of CF pathophysiology, which is directed by downstream communication circuits relayed through MΦs and dendritic cells.

The interactions of MΦs with other cells of the immune system also implicate them in changing downstream immune events. Dysfunctional epithelial cell function, the hallmark of CF, and changes in MΦ phenotype, can be complicated by the cross-talk between MΦs and epithelial

| Gene with polymorphism | Function                      | p value          | Clinical correlation                                      |
|------------------------|-------------------------------|------------------|----------------------------------------------------------|
| MHC                    | Antigen presentation          | ≤0.04 onset of colonization | Onset of pathogen colonization frequency of colonization |
| Alpha-1 antitrypsin    | Protease inhibition           | 0.04             | FEV1 % pred                                              |
| ADRB2                  | Alternative vs. classical MΦs | ≤0.05             | FEV1 % pred; 5-year decline in pulmonary function        |
| CD14                   | Pathogen interactions         | No association   | Disease severity                                          |
| DCTN4                  | Microtubule function          | ≤0.05             | Age at onset of chronic infection; Age at first infection |
| HLA                    | Adaptive communication        | ≤0.05             | FEV1 % pred; Age at onset of chronic infection           |
| IFN-γ                  | MΦ activation                 | ≥0.09             | Chronic infection: *P. aeruginosa* age at death; Age that FEV1 50% pred |
| HMOX1                  | Transcriptional regulation    | 0.01              | FEV1 % pred; Pulmonary function decline                  |
| IL-10                  | Immunoregulation              | ≥0.02             | Age of *P. aeruginosa* infection; Age at death           |
| NOS                    | Immunoregulation              | ≥0.02             | Age at colonization; Decline in pulmonary function       |
| TGF-β                  | Immunoregulation              | ≤0.04             | Age that FEV1 ≤50% pred; Impairment of lung function     |
| TLR4                   | Pathogen interactions         | ≥0.10             | Rate of change of FEV1; Age at first infection           |
| TNF-α                  | Inflammatory response         | ≥0.02             | Mean FEV1 pred; Mean Shwachman score; Age at first infection |

Recently, significant progress in genome-wide association studies has generated considerable interest in understanding other genes that might either enhance the CFTR-deficient phenotype or improve outcomes. This table provides a summary of the genes associated with MΦ function and clinical outcomes in CF patients.
cells [1]. MΦs and respiratory epithelial cells can communicate via direct contact (the formation of gap junction channels), via cell surface receptors (e.g. CD200R and TGFBR) and via paracrine communication mediated by cytokines and microvesicles [1]. No knowledge is available regarding the potential dysregulation on MΦs and respiratory epithelium cross-talk in CF. However, as highlighted in this review, the combined effects of immune cross-talk and the cellular immune artillery amplifies the role of MΦs in the CF immune response to infection. Figure 2 demonstrates the hypothesis related to the diversity of the MΦ phenotype and the impact that this may have not only on MΦ function but also on the function of the adaptive immune cells, which ultimately will relay the MΦ message regarding the appropriate host response.

Concluding Remarks: MΦ Function and CF

In summary, the innate immune response is altered in CF lungs, and monocytes/MΦs are key contributors in orchestrating this process. The studies discussed in this
review highlight that inherited (the loss of CFTR), as well as acquired factors (CF lung environment) affect the function of monocytes/MΦs, so that they do not properly handle inflammatory triggers, they struggle to resolve inflammation and they fail to clear bacterial infection. In addition, MΦs may improperly communicate with the other cells of the immune system, thus harming the adaptive immune response. Furthermore, the remarkable plasticity of MΦs and the different MΦ sub-populations that can coexist complicate how these cells participate in CF lung pathophysiology, and our understating of these mechanisms is still in its infancy. Altogether, these studies support the notion that MΦs contribute to CF lung pathology concomitantly with bronchial epithelium dysfunction.

Importantly, immune dysregulation represents a hallmark of the multiorgan manifestations in CF, such that hyperinflammation contributes to the destruction of the exocrine pancreas [101] and the tissue integrity of the gastrointestinal tract [102]. Thus, alterations in monocyte/MΦ function may contribute to CF manifestations beyond the lung disease.

As a prospective for the future, an effective, long-term therapy for CF should also modulate monocyte/MΦ function and the tissue integrity of the airways [34–36].

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Disclosure Statement

The authors declare no conflict of interest.
Macrophages in CF Lung Disease

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