The Burden of Carbapenem-Resistant Acinetobacter baumannii in ICU COVID-19 Patients: A Regional Experience

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Abstract: Since the beginning of the COVID-19 pandemic, the impact of superinfections in intensive care units (ICUs) has progressively increased, especially carbapenem-resistant Acinetobacter baumannii (CR-Ab). This observational, multicenter, retrospective study was designed to investigate the characteristics of COVID-19 ICU patients developing CR-Ab colonization/infection during an ICU stay and evaluate mortality risk factors in a regional ICU network. A total of 913 COVID-19 patients were admitted to the participating ICUs; 19% became positive for CR-Ab, either colonization or infection (n = 176). The ICU mortality rate in CR-Ab patients was 64.7%. On average, patients developed colonization or infection within 10 ± 8.4 days from ICU admission. Scores of SAPS II and SOFA were significantly higher in the deceased patients (43.8 ± 13.5, p = 0.006 and 9.5 ± 3.6, p < 0.001, respectively). The mortality rate was significantly higher in patients with extracorporeal membrane oxygenation (12; 7%, p = 0.03), septic shock (61; 35%, p < 0.001), and in elders (66 ± 10; p < 0.001). Among the 176 patients, 129 (73%) had invasive infection with CR-Ab: 105 (60.7%) Ventilator-Associated Pneumonia (VAP), and 46 (26.6%) Bloodstream Infections (BSIs). In 22 cases (6.5%), VAP was associated with concomitant BSI. Colonization was reported in 165 patients (93.7%). Mortality was significantly higher in patients with VAP (p = 0.009). Colonized patients who did not develop invasive infections had a higher survival rate (p < 0.001). Being...
colonized by CR-Ab was associated with a higher risk of developing invasive infections \( (p < 0.001) \). In a multivariate analysis, risk factors significantly associated with mortality were age (OR = 1.070; 95% CI (1.028–1.115) \( p = 0.001 \)) and CR-Ab colonization (OR = 5.463 IC95% 1.572–18.988, \( p = 0.008 \)). Constant infection-control measures are necessary to stop the spread of \textit{A. baumannii} in the hospital environment, especially at this time of the SARS-CoV-2 pandemic, with active surveillance cultures and the efficient performance of a multidisciplinary team.

**Keywords:** \textit{Acinetobacter baumannii}; \textit{Acinetobacter} infections; intensive care unit; COVID-19; SARS-CoV-2; nosocomial infections; carbapenems; multidrug resistance; antimicrobial drug resistance; critical care

### 1. Introduction

The Gram-negative aerobic bacillus \textit{Acinetobacter baumannii} (\textit{A. baumannii}) primarily causes hospital-acquired infections in especially fragile patients with prolonged hospitalization and with long-term exposition to broad-spectrum antibiotic treatment. It is characterized by disinfection resistance, and its high pathogenicity is increased by the production of a polysaccharide capsule and by the ability to form biofilms [1]. Furthermore, due to the acquisition of multiple antimicrobial resistance, especially to carbapenems, it has been recognized as a major public health concern [2] and considered as “priority 1” (critical) in the World Health Organization (WHO)’s first list of “priority pathogens” resistant to antibiotics, including the 12 families of bacteria most dangerous for human health and for which new antimicrobials are urgently required [3].

It is well known that \textit{A. baumannii} exhibits a wide variety of mechanisms of resistance to antibiotic agents, as differential clones had been isolated in Europe [4]. \textit{A. baumannii} includes several mechanisms of resistance such as lipopolysaccharide expression disorders, permeability alterations due to porins, and the production of active efflux pumps. In particular, resistance to carbapenems is related to numerous beta-lactamases with carbapenemase activity, including type OXA carbapenemases—both constitutive or acquired. Moreover, a transmissible resistance mechanism of colistin, called mobile colistin resistance (MCR), was discovered. Up to ten families with MCR and more than 100 variants of Gram-negative bacteria have been reported worldwide. Even though few have been reported from \textit{Acinetobacter} spp. and \textit{Pseudomonas} spp., it is important to closely monitor the epidemiology of MCR genes in these pathogens [1,4].

\textit{A. baumannii} can survive for long periods on surfaces, including human skin and dry surfaces (up to 33 days) [5], and this ability might facilitate its persistence in intensive care units (ICUs), rightly considered as the epicenters of carbapenem-resistant \textit{A. baumannii} (CR-Ab) infections [6,7]. Some specific factors may increase the potential of cross-transmission: the heavy colonization of the patient, the colonization of the surfaces surrounding the patients, and the total number of patients colonized in the unit at the same time [8]. CR-Ab also has a further important feature, namely its tendency to generate outbreaks, generally transmitted through the hands of healthcare workers, contaminated equipment, and the healthcare environment [7,9,10].

The most frequently reported risk factors for CR-Ab infections are prior colonization, the severity of illness, the need for mechanical ventilation (particularly in case of prolonged duration), immunosuppression, malignancies, chronic pulmonary diseases, respiratory failure on admission, previous antimicrobial therapy, previous sepsis in ICU, previous use of carbapenems and third generation cephalosporins, long ICU stay [11], and a consequent greater degree of exposure to infected or colonized patients in the hospital environment [12,13].

Overall, CR-Ab is accountable for more than 12% of the cases of hospital-acquired bloodstream infections (BSI) in the ICU, with wide geographic variations: it is frequent in Southern Europe, Middle Eastern countries, Asia, and South America, whereas it is rare in Northern European countries and Australia [14]. CR-Ab is a common cause of ICU-acquired pneumonia, particularly late-onset pneumonia [15].
Since the beginning of the COVID-19 pandemic, the impact of superinfections in ICU patients has progressively increased and many studies have shown that the rate of BSIs [16] and Ventilator-Associated Pneumonia (VAP) [17] is raised compared to that observed in non-COVID-19 patients [18–21]. It has also been reported that the prevalence of Gram-negative multi-drug resistant organisms, especially \textit{A. baumannii}, known to increase mortality, seems to have escalated [22,23].

In Italy, various experiences of multidrug-resistant (MDR) bacterial infection in COVID-19 and its impact on patient outcomes have been published, [24] but few describe specifically \textit{A. baumannii} cases. Several studies showed that MDR infection arose after a median time of 8 [4–11] days and the incidence rate ratio of MDR infection in ICU increased in the COVID-19 period [25,26].

Despite the above evidence and the interest in superinfections from multidrug-resistant pathogens, particularly CR-Ab in ICU patients with COVID-19 [27], to date, no multicenter study has been conducted with the aim of better describing the phenomenon.

The present multicenter retrospective study was designed to investigate the characteristics of COVID-19 ICU patients who developed CR-Ab colonization or infection during their ICU stay and evaluate risk factors for ICU mortality in a regional ICU network.

2. Materials and Methods

2.1. Study Design and Population

This was an observational, multicenter, and retrospective study. Nineteen COVID-19 ICUs in the Piedmont Region, Italy, were invited to participate in an observational, multicenter, retrospective study to describe the incidence of colonization and infection with CR-Ab in SARS-CoV2 pneumonia patients admitted to ICUs between 1 July and 31 December 2021.

The data sources were the hospital administrative records and the Microbiology Laboratory database. Data acquisition and analysis were performed in accordance with the protocols approved by the local Ethics Committee (Ethics Committee: Comitato Etico Interaziendale A.O.U. Città della Salute e della Scienza di Torino—A.O. Ordine Mauriziano—A.S.L. Città di Torino; ethics approval number 0031285). Written informed consent was waived according to Italian regulations due to the retrospective nature of this study. The study was conducted according to the guidelines of the Declaration of Helsinki.

All consecutive adult (≥18 years) patients admitted to the ICU and presenting CR-Ab colonization or infection during the study period were enrolled. All patients were followed up until the hospital discharge to compute: ICU, 28-day and overall mortality, length of ICU and hospital stay.

2.2. Context

During the study period, several infection control programs were active in the ICUs involved, with specific leadership and scope. Surveillance cultures (tracheal aspirate, rectal swab, urinary culture) were performed weekly; universal screening for carbapenem-producing Enterobacteriales (CPE) and \textit{A. baumannii} using rectal swabs was performed upon admission to the ICU and then once a week. In mechanically ventilated patients, the surveillance of respiratory samples (tracheal aspirates or bronchoalveolar lavage) was also performed at least once a week, with some differences between the different centers. Blood cultures or bronchoalveolar lavage cultures were performed on clinical decision.

2.3. Definitions

Pneumonia by Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) was defined based on real-time polymerase chain reaction (RT-PCR) on at least one low respiratory tract specimen [28].

The occurrence of colonization and/or infection with \textit{A. baumannii} was assessed from the date of ICU admission to ICU discharge. It was considered only once at the time of the first incidence of a positive sample. Colonization was defined as bacterial isolation without
clinical signs or symptoms suggestive of infection. Infection was defined according to the Centers for Disease Control and Prevention (CDC) criteria [29]. Carbapenem resistance was defined according to the EUCAST criteria [30].

All episodes of VAP and/or BSI, as well as the development of septic shock with the requirement of vasoactive drugs [31], were registered according to the European Centre for Disease Prevention and Control (ECDC) current definitions [32].

2.4. Microbiology

*A. baumannii* and CPE strains from blood, respiratory, and rectal samples were collected in accordance with active surveillance screening and following local guidelines. Rectal swabs were collected from hospitalized patients and screened for CPE by combining culture-based detection and the identification of carbapenemase type.

We identified CR-Ab according to the European Committee on Antimicrobial Susceptibility Testing (EUCAST) criteria of carbapenem resistance. Cultures were analyzed with the BD BACTECTM FX system (Becton Dickinson) according to EUCAST breakpoint tables. The identification of microorganisms was conducted with mass spectrometry MALDI-TOF (Matrix-Assisted Laser Desorption Ionization Time-of-Flight) and VITEK®, whereas susceptibility to antibiotic molecules was tested using VITEK 2 (VITEK® according to EUCAST breakpoint tables). The whole-genome sequencing of CR-Ab isolates collected from blood cultures and respiratory samples was not available in the pandemic context. The clonal relationship of CR-Ab isolates was currently not investigated.

2.5. Statistical Analysis

Data were entered and analyzed using SPSS version 27. Statistical significance was defined as less than 0.05. Descriptive analysis was reported as frequencies, percentages, means, and standard deviations. Categorical variables, demographics, and clinical characteristics were compared against mortality using the Chi-squared test. Continuous variables were tested for normality by the Kolmogorov–Smirnov test. Non-normally distributed variables were evaluated using the Mann–Whitney test.

Significant values in the univariate analysis were evaluated with a multivariate model: a logistic regression model for mortality to assess independent predictors. The odds ratio was reported with corresponding 95% confidence intervals.

3. Results

Sixteen ICUs joined the data collection. Four of them had no cases of CR-Ab in COVID-19 patients. The first data collection was completed in May 2021. The review of the data by independent investigators was completed in the months of June–September 2021.

During the study period, 913 COVID-19 patients were admitted to the participating ICUs. Of them, 19% became positive for CR-Ab, either colonization or infection (*n* = 176).

The ICU mortality rate in patients with *A. baumannii* was as high as 64.7% (*n* = 112) (Table 1).

**Table 1.** Demographic and general characteristics of COVID-19 ICU patients with CR-Ab.

| Variable | Total (n; %) | Survived (n; %) | Dead (n; %) | p-Value |
|----------|-------------|----------------|-------------|---------|
| **Demographics** | | | | |
| Males | 136 (78.6) | 48 (67.6%) | 88 (78.6%) | 0.986 |
| Age | 65.35 ± 10.3 | 62.84 ± 10.7 | 66.44 ± 10 | <0.001 |
| BMI | 30.8 ± 7.3 | 31.33 ± 7.4 | 30.83 ± 7.36 | 0.858 |
| Ex-smoker | 8 (4.5) | 4 (6.5) | 4 (3.5) | 0.372 |
| Smoker | 8 (4.5) | 3 (4.9) | 5 (4.5) | 0.892 |
| Obese | 52 (29.5) | 20 (32.8) | 32 (28.6) | 0.563 |
Table 1. Cont.

| Variable                      | Total (100%) | Survived (35.3%) | Dead (64.7%) | p-Value |
|-------------------------------|--------------|------------------|--------------|---------|
| **Comorbidities**             |              |                  |              |         |
| Cardiovascular disease        | 118 (67)     | 38 (62.3)        | 80 (71.4)    | 0.218   |
| Diabetes                      | 39 (22.1)    | 12 (19.7)        | 27 (24.1)    | 0.505   |
| Hematologic disease           | 2 (1.1)      | 1 (1.6)          | 1 (0.9)      | 0.661   |
| Chronic pulmonary disease     | 22 (12.5)    | 6 (9.8)          | 16 (14.3)    | 0.401   |
| Renal failure                 | 15 (8.5)     | 5 (8.1)          | 10 (8.9)     | 0.870   |
| Active neoplasm               | 7 (4)        | 2 (3.3)          | 5 (4.5)      | 0.705   |
| Autoimmune disease            | 18 (10.2)    | 6 (9.8)          | 12 (10.7)    | 0.857   |
| Immunodepression              | 4 (2.3)      | 2 (3.3)          | 2 (1.8)      | 0.532   |
| **Clinical characteristics**  |              |                  |              |         |
| ICU length of stay            | 24.27 ± 17.9 | 25.7 ± 20.58     | 24.1 ± 18.22 | 0.930   |
| Days to infection/colonization from hospital admission | 17.31 ± 13.3 | 17.2 ± 13.44 | 17.31 ± 12.34 | 0.718   |
| Days to infection/colonization from ICU admission | 10.69 ± 8.4 | 10.63 ± 8.38 | 10.69 ± 8.42 | 0.585   |
| Referral                      | 54 (30.7)    | 17 (27.9)        | 37 (33)      | 0.483   |
| ECMO                          | 13 (7.4)     | 1 (1.6)          | 12 (10.7)    | 0.031   |
| SAPS II                       | 42.28 ± 13.37| 41.6 ± 13        | 43.88 ± 13.5 | 0.006   |
| SOFA                          | 8.3 ± 3.7    | 6 ± 2.6          | 9.5 ± 3.6    | <0.001  |
| ARDS on admission             | 165 (93.2)   | 59 (96.7)        | 106 (94.6)   | 0.534   |
| Septic shock                  | 67 (38.1)    | 6 (9.8)          | 61 (54.5)    | <0.001  |
| Colistin sensitive            | 159 (90.3)   | 53 (86.9)        | 106 (94.6)   | 0.074   |
| Colistin resistant            | 14 (7.9)     | 8 (13.1)         | 6 (5.3)      |         |
| Carbapenem-resistant          | 122 (69.3)   | 46 (75.4)        | 76 (67.8)    | 0.479   |
| **Invasive infections**       |              |                  |              |         |
| CR-Ab VAP                     | 105 (59.6)   | 29 (47.5)        | 76 (67.8)    | 0.009   |
| CR-Ab BSI                     | 46 (41.1)    | 14 (22.9)        | 32 (28.6)    | 0.424   |
| CR-Ab + co-infection          |              |                  |              |         |
| K. pneumoniae—KPC             | 29 (16.5)    | 11 (18)          | 18 (16.1)    | 0.726   |
| MRSA                          | 8 (4.5)      | 3 (4.9)          | 5 (4.5)      | 0.892   |
| VRE                           | 7 (4)        | 3 (4.9)          | 4 (3.6)      | 0.668   |
| Enteric pathogens             | 55 (31.2)    | 18 (29.5)        | 37 (33)      | 0.634   |
| **Colonization**              |              |                  |              |         |
| CR-Ab                         | 165 (93.7)   | 58 (95)          | 104 (92.8)   | 0.567   |
| Cp-K.pneumoniae               | 14 (7.9)     | 4 (6.5)          | 10 (8.9)     | 0.585   |
| VRE                           | 1 (0.6)      | 1 (1.6)          | 0            | 0.174   |
| E.coli                        | 2 (1.1)      | 2 (3.2)          | 0            | 0.054   |
| Candida spp                   | 8 (4.5)      | 3 (4.9)          | 5 (4.5)      | 0.892   |
| MRSA                          | 5 (2.8)      | 1 (1.6)          | 4 (3.6)      | 0.469   |
| Other                         | 74 (42)      | 28 (45.9)        | 46 (41.1)    | 0.587   |
| **Combination treatment with colistin** |         |                  |              |         |
| Total colistin treatment      | 100 (56.8)   | 33 (54)          | 67 (59.8)    | 0.466   |
| Colistin monotherapy          | 10 (5.7)     | 1 (1.6)          | 9 (8)        | 0.085   |
| Meropenem                     | 37 (21)      | 16 (26.2)        | 21 (18.7)    | 0.252   |
| Ampicillin sulbactam          | 32 (18.1)    | 9 (14.7)         | 23 (20.5)    | 0.349   |
| Rifampicin                    | 30 (17)      | 9 (14.7)         | 21 (18.7)    | 0.507   |
| Tigecycline                   | 15 (8.5)     | 2 (3.2)          | 13 (11.6)    | 0.063   |
| Vancomycin                    | 7 (4)        | 3 (4.9)          | 4 (3.6)      | 0.668   |
| Ceftazidime-avibactam         | 7 (4)        | 1 (1.6)          | 6 (5.3)      | 0.236   |

**Only colonized/infected vs. mortality**

| CR-Ab colonized (without infection) | 47 (26.7) | 25 (40.9) | 22 (19.6) | <0.001 |
| CR-Ab infected (without colonization) | 11 (6.2) | 3 (4.9) | 8 (7.1) | 0.567 |

List of abbreviations: intensive care unit, ICU; carbapenem-resistant Acinetobacter baumannii, CR-Ab; number, n; Body Mass Index, BMI; extracorporeal membrane oxygenation, ECMO; Simplified Acute Physiology Score, SAPS; Sequential Organ Failure Assessment, SOFA; Acute Respiratory Distress Syndrome, ARDS; Ventilator-Associated Pneumonia, VAP; Bloodstream infection, BSI; K. pneumoniae producing KPC; methicillin-resistant Staphylococcus aureus, MRSA; vancomycin-resistant Enterococcus, VRE. bold was used for *p < 0.05.*
The majority of patients were males (136; 78.6%), with a median age of 65 ± 10.3 years. The average Simplified Acute Physiology Score (SAPS) II and Sequential Organ Failure Assessment (SOFA) scores were 42 ± 13.37 and 8.3 ± 3.7, respectively. Leading comorbidities were cardiovascular diseases (118 patients, 67%), obesity (52 patients, 29.5%), diabetes (39 patient, 22.1%), and chronic pulmonary disease (22 patients, 12.5%). Around 31% of patients were transferred from one hospital to another; 93.2% of them presented acute respiratory distress syndrome upon ICU admission. The mean length of stay in the ICU was 24 ± 18 days. On average, patients developed colonization or infection within 10 ± 8.4 days from ICU admission.

The scores of SAPS II and SOFA were significantly higher in the deceased patients (43.8 ± 13.5, p = 0.006 and 9.5 ± 3.6, p < 0.001, respectively). Furthermore, the mortality rate was significantly higher in patients with extracorporeal membrane oxygenation (ECMO; 12; 7%, p = 0.03), septic shock (61; 35%, p < 0.001), and in elders (66 ± 10, p < 0.001) (Table 1).

Among the 176 patients enrolled in the study, 129 (73%) had invasive infection with CR-Ab, distributed as follows: 105 (60.7%) VAP and 46 (26.6%) BSI. In 22 cases (6.5%), VAP was associated with concomitant BSI. Colonization was reported in 165 patients (93.7%). Of note, 118 patients previously colonized by CR-Ab developed invasive infections, while 11 patients developed infection without any known previous colonization. Mortality was significantly higher in patients with VAP (p = 0.009). Colonized patients who did not develop invasive infections had a higher survival rate (p < 0.001; Table 1). Being colonized by CR-Ab was associated with a higher risk of developing invasive infections (p < 0.001).

In the present multicentric study, conducted on 16 ICUs in the Piedmont region during the COVID-19 pandemic, it was found that 19% of ICU COVID-19 patients became positive for CR-Ab, either colonization or infection, during an ICU stay. Although the whole-genome sequencing of CR-Ab isolates was not available in the pandemic context and the clonal relationship of CR-Ab isolates was currently not investigated, this elevated percentage and some epidemiological factors deserve very high attention. Furthermore,
the mortality rate in patients with CR-Ab was as high as 64.7%, significantly higher than the overall mortality in critically ill COVID-19 patients [36].

To the best of our knowledge, this is the first multicenter regional study reporting the impact of CR-Ab colonization and severe infection in ICUs during the COVID-19 pandemic. Interestingly, our analysis refers to the so-called Italian “second-wave” of the pandemic, when the global emergency scenario of the first months of the pandemic had extensively changed. A recent multicenter, cross-sectional study compared the rates of colonization and infection with carbapenemase-producing Enterobacteriales (CPE) and/or CR-Ab in two study periods, pre and during the COVID-19 pandemic. No significant change in either incidence rate ratios and weekly trends in CPE colonization and infection was observed, while the incidence rate ratios of colonization and infection with CR-Ab increased by 7.5- and 5.5-fold, respectively, during the COVID-19 period. A clonal lineage was demonstrated and appointed for the occurrence of horizontal transmission [26].

Other authors previously highlighted that, during the first wave of the COVID-19 pandemic, several factors could have favored the emergence and spread of antimicrobial-resistant bacteria in hospitals [25], such as the overload of hospitalized patients, especially in intensive care, favoring patient-to-patient transmission [37]; the initial overuse of antibiotics for suspected bacterial co/super-infections [38]; the possible delay in providing microbiological culture and sensitivities results due to the COVID-19 overload [39]. During the first months of the pandemic, in several countries, including Italy, a lack of appropriate protective personal equipment and health personnel hired on an emergency basis to respond to the COVID-19 pandemic, sometimes impeding adequate training in infection prevention and control, were common. However, that may not be completely true in the period of our study, when the first pandemic phase with its need for reorganization was already over.

Other factors may have contributed to the described spread of CR-Ab infections. First of all, the need for the referral of critically ill patients (e.g., requiring ECMO [40]) and the high number of patients transferred from one hospital to another may have facilitated the dissemination of cases at the regional level. Even the structural characteristics of ICUs (new, re-opened, or already functioning before the COVID-19 pandemic) may also have played a role, in terms of spaces dedicated to patients and workstations, devices, and hospital pathways between departments (e.g., emergency department, radiology). In fact, CR-Ab cross-transmission between equipment (ventilators, infusion pumps, hemodialysis machines, ultrasound devices) and COVID-19 patients may also partly explain the onset of this outbreak.

Focusing on the identification and characterization of Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumannii, Pseudomonas aeruginosa, and Enterobacter spp. (ESKAPE) bacteria and their possible clonal spread in medical devices, patients, and medical personnel in the ICU, a recent work [41] has shown that 91% of the analyzed sites were colonized by bacteria (pathogenic and commensal), where S. aureus and A. baumannii MDR showed a high incidence, and A. baumannii MDR showed a clonal distribution in surfaces, patients, and health personnel.

It is in fact known that even when there is the scrupulous protection of medical personnel to avoid the transmission of SARS-CoV-2 from patients to health personnel, the transmission of other pathogens such as ESKAPE bacteria is not automatically avoided. In a previous study in ICUs, it was shown that the bacterial recontamination of contact surfaces occurred after 4 h after standard cleaning with detergents with chlorine-releasing agents, isopropyl alcohol, and sodium hypochlorite [42]. Moreover, COVID-19 critically ill patients often require prolonged hospitalizations, and it is known that staying in an intensive care setting for a long time—as well as immunosuppression, the need for prolonged previous antibiotic therapies, and the invasiveness of care—are known risk factors for infections with multidrug-resistant pathogens.
In our analysis, the median ICU length of stay was high (24.27 ± 17.9 days), with a time lag before the development of colonization or the onset of invasive infection of 17.31 ± 13.3 days of hospital stay and 10.69 ± 8.4 days of ICU stay, respectively. Some other factors must be taken into account in the analyzed population. Certainly, patient severity had an impact on mortality, with statistical significance for the need for ECMO support, higher SAPS and SOFA scores, and the presence of septic shock as infection presentation. Similar to other settings, the use of steroids might be related to a higher risk of developing MDR infection [43]. Concerning the impact of VAP in CR-Ab infected patients, the diagnosis of VAP may have been made difficult by the presence of the radiological and clinical signs of COVID-19 pneumonia, which made it even more difficult to apply the classical criteria and the consequent definition of VAP.

The presence of colonization preceding the infection represented, in our series, a risk factor with respect to mortality. It is well known from the literature that colonization does not require any "pre-emptive" therapy if the patient has no clinical signs of infection, but these data confirmed the finding that colonization remains one of the main risk factors for invasive infections and represent a "wake up call" regarding the frailty of our patients. Therefore, implementing an early pre-emptive therapy in cases of known colonization, at the time of clinical worsening, is one of the main steps to improve survival in this setting.

As previously reported in the literature, the role of combination therapy is widely debated in the absence of definitive evidence [44,45]. The data are insufficient for a more completed analysis, but the unmet need for new and effective therapies is of paramount importance considering the mortality of these critically ill patients.

The presence of multi-bacterial co-infections is a further interesting fact, able to describe not only the fragility of the patients but also the delicate hospital ecology and to reinforce the need for effective and strict control measures. In particular, the combination of various Gram-negative pathogens describes the context of our ICUs and may be the consequence of the high use of empiric broad-spectrum antibiotic therapies used in COVID patients not only at home but also in the early stages of hospitalization.

Our study has several limitations. First, the retrospective nature of the study and therapeutic management on the risk of A. baumannii infection. Secondly, the lack of data on the total number of COVID-19 ICU patients did not allow a comparison of risk factors and outcomes. Third, as the clonal relationship was not investigated, it is impossible to define the common origin of the burden of infections or a relationship, at least in the high number of referral patients. Moreover, it was not possible to obtain a cumulative antibiogram for antibiotic classes to show the overall sensibility of different strains. Finally, the local epidemiology and the need to re-organize the capacity, spaces, and staff of our ICUs during the pandemic could limit the generalizability of our results.

5. Conclusions

The need to not neglect antimicrobial stewardship principles during the COVID-19 pandemic has already been recently underlined [46], as well as the importance of enhancing infection control activities directed against antimicrobial resistance. In continuity with this message, our study remarks on the need to pursue antimicrobial stewardship principles during the COVID-19 pandemic, and infection control activities targeted against the spread of antimicrobial resistance inside and between hospitals.

During a pandemic, not only in the first phases, but especially later in the time course, infection control activities should be revised and eventually re-modulated according to the new organizational structures. Constant infection-control measures are necessary to stop the spread of A. baumannii in the hospital environment, prevent outbreaks, and lower mortality rates, especially at this time of the SARS-CoV-2 pandemic. Stricter barrier measures need to be implemented, increasing the effectiveness of screening and surveillance for A. baumannii, especially when resistant to carbapenems. The active surveillance culture and efficient performance of a multidisciplinary team will be highly important in detecting and controlling the CR-Ab outbreak in COVID-19 ICUs.
References

1. Garnacho-Montero, J.; Timsit, J.F. Managing Acinetobacter baumannii infections. *Curr. Opin. Infect. Dis.* 2019, 32, 69–76. [CrossRef] [PubMed]

2. Cassini, A.; Högberg, L.D.; Plachouras, D.; Quattrocchi, A.; Hoxha, A.; Simonsen, G.S.; Colomb-Cotinat, M.; Kretzschmar, J.; et al. Attributable deaths and disability-adjusted life-years caused by infections with antibiotic-resistant bacteria in the EU and the European Economic Area in 2015: A population-level modelling analysis. *Lancet Infect. Dis.* 2019, [19], 56–66. [CrossRef]

3. Tacconelli, E.; Carrara, E.; Savoldi, A.; Harbarth, S.; Mendelson, M.; Monnet, D.L.; Pulcini, C.; Kahlmeter, G.; Kluytmans, J.; et al. WHO Pathogens Priority List Working Group. Discovery, research, and development of new antibiotics: The WHO priority list of antibiotic-resistant bacteria and tuberculosis. *Lancet Infect. Dis.* 2018, 18, 318–327. [CrossRef]

4. Fitzpatrick, M.A.; Ozer, E.A.; Hauser, A.R. Utility of Whole-Genome Sequencing in Characterizing Acinetobacter Epidemiology and Analyzing Hospital Outbreaks. *J. Clin. Microbiol.* 2016, 54, 593–612. [CrossRef]

5. Howard, A.; O’Donoghue, M.; Feeney, A.; Sleator, R.D. Acinetobacter baumannii: An emerging opportunistic pathogen. *Virology* 2012, 3, 243–250. [CrossRef]

6. Jawad, A.; Seifert, H.; Snelling, A.M.; Heritage, J.; Hawkey, P.M. Survival of Acinetobacter baumannii on dry surfaces: Comparison of outbreak and sporadic isolates. *J. Clin. Microbiol.* 1998, 36, 1938–1941. [CrossRef]

7. Meschiari, M.; López-Lozano, J.M.; Di Pilato, V.; Gimenez-Esparza, C.; Vecchi, E.; Bacca, E.; Orlando, G.; Franceschini, E.; Sarti, M.; Pecorari, M.; et al. A five-component infection control bundle to permanently eliminate a carbapenem-resistant *Acinetobacter baumannii* spreading in an intensive care unit. *Antimicrob. Resist. Infect. Control.* 2021, 10, 123. [CrossRef] [PubMed]
9. Munoz-Price, L.S.; Arheart, K.; Nordmann, P.; Boulanger, A.E.; Cleary, T.; Alvarez, R.; Pizano, L.; Namias, N.; Kett, D.H.; Poirel, L. Eighteen years of experience with Acinetobacter baumannii in a tertiary care hospital. *Crit. Care Med.* 2013, 41, 2733–2742. [CrossRef]

10. Escudero, D.; Cofíño, L.; Forcelledo, L.; Quindós, B.; Calleja, C.; Martín, L. Control of an Acinetobacter baumannii multidrug resistance endemic in the ICU. Recalling the obvious. *Med. Intensiv.* 2017, 41, 497–499. [CrossRef]

11. Garnacho-Montero, J.; Dimopoulos, G.; Poulakou, G.; Akova, M.; Cisneros, J.M.; De Waele, J.; Petrosillo, N.; Seifert, H.; Timsit, J.F.; Vila, J.; et al. Task force on management and prevention of Acinetobacter baumannii infections in the ICU. *Intensive Care Med.* 2015, 41, 2057–2075. [CrossRef] [PubMed]

12. Lee, H.Y.; Chen, C.L.; Wu, S.R.; Huang, C.W.; Chiu, C.H. Risk factors and outcome analysis of Acinetobacter baumannii complex bacteremia in critical patients. *Crit. Care Med.* 2014, 42, 1081–1088. [CrossRef] [PubMed]

13. Huang, H.; Chen, B.; Liu, G.; Ran, J.; Lian, X.; Huang, X.; Wang, N.; Huang, Z. A multi-center study on the risk factors of infection caused by multi-drug resistant Acinetobacter baumannii. *BMC Infect. Dis.* 2018, 18, 11. [CrossRef] [PubMed]

14. Tabah, A.; Koulenti, D.; Laupland, K.; Misset, B.; Valles, J.; Bruzzi de Carvalho, F.; Paiva, J.A.; Cakar, N.; Ma, X.; Eggimann, P.; et al. Characteristics and determinants of outcome of hospital-acquired bloodstream infections in intensive care units: The EUROBACT International Cohort Study. *Intensive Care Med.* 2012, 38, 1930–1945. [CrossRef]

15. Koulenti, D.; Tsigou, E.; Rello, J. Nosocomial pneumonia in 27 ICUs in Europe: Perspectives from the EU-VAP/CAP study. *Eur. J. Clin. Microbiol. Infect. Dis.* 2017, 36, 1999–2006. [CrossRef]

16. Ripa, M.; Galli, L.; Poli, A.; Oltolini, C.; Spagnuolo, V.; Mastrangelo, A.; Muccini, C.; Monti, G.; De Luca, G.; Landoni, G.; et al. COVID-19 infection in secondary infections in patients hospitalized with COVID-19: Incidence and predictive factors. *Clin. Microbiol. Infect.* 2021, 27, 451–457. [CrossRef]

17. Giacobbe, D.R.; Battaglini, D.; Enrie, E.M.; Dentone, C.; Vena, A.; Robba, C.; Ball, L.; Bartoletti, M.; Coloretti, I.; Di Bella, S.; et al. Incidence and Prognosis of Ventilator-Associated Pneumonia in Critically Ill Patients with COVID-19: A Multicenter Study. *J. Clin. Med.* 2021, 10, 555. [CrossRef]

18. Sharifipour, E.; Shams, S.; Esmkhani, M.; Khodadadi, J.; Fotouhi-Ardakani, R.; Koohpaei, A.; Doostiz, E.; Golzarli, S. Evaluation of bacterial co-infections of the respiratory tract in COVID-19 patients admitted to ICU. *BMC Infect. Dis.* 2020, 20, 646. [CrossRef]

19. Gottesman, T.; Fedorovsky, R.; Erushalmi, R.; Lellouche, J.; Nutman, A. An outbreak of carbapenem-resistant *Acinetobacter baumannii* in a COVID-19 dedicated hospital. *Infect. Prev. Pract.* 2021, 3, 100113. [CrossRef]

20. Shinohara, D.R.; Dos Santos Saalfeld, S.M.; Martinez, H.V.; Altafini, D.D.; Costa, B.B.; Fedrigo, N.H.; Tognin, M.C.B. Outbreak of carbapenem-resistant *Acinetobacter baumannii* in a coronavirus disease 2019 (COVID-19)-specific intensive care unit. *Infect. Control. Hosp. Epidemiol.* 2021, 43, 815–817. [CrossRef]

21. Nasir, N.; Rehman, F.; Omair, S.F. Risk factors for bacterial infections in patients with moderate to severe COVID-19: A case-control study. *J. Med. Virol.* 2021, 93, 4564–4569. [CrossRef] [PubMed]

22. Bardi, T.; Pintado, V.; Gomez-Rojo, M.; Escudero-Sanchez, R.; Azzam Lopez, A.; Diez-Remesal, Y.; Martinez Castro, N.; Ruiz-Garbajosa, P.; Pestaña, D. Nosocomial infections associated to COVID-19 in the intensive care unit: Clinical characteristics and outcome. *Eur. J. Clin. Microbiol. Infect. Dis.* 2021, 40, 495–502. [CrossRef] [PubMed]

23. Liu, Y.; Wang, Q.; Zhao, C.; Chen, H.; Li, H.; Wang, H.; CARES Network. Prospective multi-center evaluation on risk factors, clinical characteristics and outcomes due to carbapenem resistance in *Acinetobacter baumannii* complex bacteriaemia: Experience from the Chinese Antimicrobial Resistance Surveillance of Nosocomial Infections (CARES) Network. *J. Med. Microbiol.* 2020, 69, 949–959. [PubMed]

24. Montucchio, G.; Corcione, S.; Sales, G.; Curtoni, A.; De Rosa, F.G.; Bragazzi, L. Carbapenem-resistant Klebsiella pneumiaeia in ICU-admitted COVID-19 patients: Keep an eye on the ball. *J. Glob. Antimicrob. Resist.* 2020, 23, 398–400. [CrossRef] [PubMed]

25. Karruli, A.; Bocca, F.; Gagliardi, M.; Patauner, F.; Ursi, M.P.; Sommese, P.; De Rosa, R.; Murino, P.; Ruocco, G.; Corcione, A.; et al. Multidrug-Resistant Infections and Outcome of Critically Ill Patients with Coronavirus Disease 2019: A Single Center Experience. *Microb. Drug Resist.* 2021, 27, 1167–1175. [CrossRef]

26. Pascale, R.; Bussini, L.; Gaibani, P.; Bovo, F.; Fornaro, G.; Lombardo, D.; Ambretti, S.; Pensaline, G.; Appolloni, L.; Bartoletti, M.; et al. Carbapenem-resistant bacteria in an intensive care unit during the coronavirus disease 2019 (COVID-19) pandemic: A multicenter before-and-after cross-sectional study. *Infect. Control. Hosp. Epidemiol.* 2021, 43, 461–466. [CrossRef]

27. Rangel, K.; Chagas, T.P.G.; De-Simone, S.G. *Acinetobacter baumannii* Infections in Times of COVID-19 Pandemic. *Pathogens* 2021, 10, 1006. [CrossRef]

28. World Health Organization. Laboratory Testing for 2019 Novel Coronavirus (2019-nCoV) in Suspected Human Cases. Available online: https://www.who.int/publications-detail/laboratory-testing-strategy-recommendations-for-covid-19-interim-guidance (accessed on 10 July 2022).

29. Horan, T.C.; Andrus, M.; Dudeck, M.A. CDC/NHSN surveillance definition of health care-associated infection and criteria for specific types of infections in the acute care setting. *Am. J. Infect. Control.* 2008, 36, 309–332. [CrossRef]

30. EUCAST. Breakpoint Tables for Interpretation of MICs and Zone Diameters. Version 11.0. 2020. Available online: http://www.eucast.org/clinical_breakpoints/ (accessed on 10 July 2022).

31. Singer, M.; Deutchman, C.S.; Seymour, C.W.; Shankar-Hari, M.; Annane, D.; Bauer, M.; Bellomo, R.; Bernard, G.R.; Chiche, J.D.; Coopersmith, C.M.; et al. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 2016, 315, 801–810. [CrossRef]
32. Plachouras, D.; Lepape, A.; Suetens, C. ECDC definitions and methods for the surveillance of healthcare-associated infections in intensive care units. *Intensive Care Med.* 2018, 44, 2216–2218. Erratum in *Intensive Care Med.* 2018, 44, 2020. [CrossRef] [PubMed]

33. Grasselli, G.; Cattaneo, E.; Florio, G. Secondary infections in critically ill patients with COVID-19. *Crit. Care* 2021, 25, 317. [CrossRef] [PubMed]

34. Montrucchio, G.; Lupia, T.; Lombardo, D.; Stroffolini, G.; Corcione, S.; De Rosa, F.G.; Brazzi, L. Risk factors for invasive aspergillosis in ICU patients with COVID-19: Current insights and new key elements. *Ann. Intensive Care* 2021, 11, 136. [CrossRef]

35. European Centre for Disease Prevention and Control (ECDC) ECDC. Surveillance of Antimicrobial Resistance in Europe 2018. 2019. Available online: [https://www.ecdc.europa.eu/en/publications-data/surveillance-antimicrobial-resistance-europe-2018](https://www.ecdc.europa.eu/en/publications-data/surveillance-antimicrobial-resistance-europe-2018) (accessed on 10 July 2022).

36. Oliveira, E.; Parikh, A.; Lopez-Ruiz, A.; Carrilo, M.; Goldberg, J.; Ceasars, M.; Fernainy, K.; Andersen, S.; Mercado, L.; Guan, J.; et al. ICU outcomes and survival in patients with severe COVID-19 in the largest health care system in central Florida. *PLoS ONE* 2021, 16, e0249038. [CrossRef] [PubMed]

37. Grasselli, G.; Pesenti, A.; Cecconi, M. Critical Care Utilization for the COVID-19 Outbreak in Lombardy, Italy: Early Experience and Forecast During an Emergency Response. *JAMA* 2020, 323, 1545–1546. [CrossRef]

38. Langford, B.J.; So, M.; Raybardash, S.; Leung, V.; Westwood, D.; MacFadden, D.R.; Soucy, J.R.; Daneman, N. Bacterial co-infection and secondary infection in patients with COVID-19: A living rapid review and meta-analysis. *Clin. Microbiol. Infect.* 2020, 26, 1622–1629. [CrossRef]

39. Vaughn, V.M.; Gandhi, T.N.; Petty, L.A.; Patel, P.K.; Prescott, H.C.; Malani, A.N.; Ratz, D.; McLaughlin, E.; Chopra, V.; Flanders, S.A. Empirc Antibacterial Therapy and Community-onset Bacterial Coinfection in Patients Hospitalized with Coronavirus Disease 2019 (COVID-19): A Multi-hospital Cohort Study. *Clin. Infect. Dis.* 2021, 72, e533–e541. [CrossRef]

40. Montrucchio, G.; Sales, G.; Urbino, R.; Simonetti, U.; Bonetto, C.; Cura Stura, E.; Simonato, E.; Fuoco, G.; Fanelli, V.; Brazzi, L. ECMO Support and Operator Safety in the Context of COVID-19 Outbreak: A Regional Center Experience. *Membranes* 2021, 11, 334. [CrossRef]

41. Durán-Manuel, E.M.; Cruz-Cruz, C.; Ibáñez-Cervantes, G.; Bravata-Alcantará, J.C.; Sosa-Hernández, O.; Delgado-Balbuena, L.; León-García, G.; Cortés-Ortiz, I.A.; Cureño-Díaz, M.A.; Castro-Escarpulli, G.; et al. Clonal dispersion of Acinetobacter baumannii in an intensive care unit designed to patients COVID-19. *J. Infect. Dev. Ctries.* 2021, 15, 58–68. [CrossRef]

42. Wilson, A.P.; Smyth, D.; Moore, G.; Singleton, J.; Jackson, R.; Gant, V.; Jeanes, A.; Shaw, S.; James, E.; Cooper, B.; et al. The impact of enhanced cleaning within the intensive care unit on contamination of the near-patient environment with hospital pathogens: A randomized crossover study in critical care units in two hospitals. *Crit. Care Med.* 2011, 39, 651–658. [CrossRef]

43. Fanelli, V.; Montrucchio, G.; Sales, G.; Simonetti, U.; Bonetto, C.; Rumbolo, F.; Mengozzi, G.; Urbino, R.; Pizzi, C.; Richiardi, L.; et al. Effects of Steroids and Tocilizumab on the Immune Response Profile of Patients with COVID-19-Associated ARDS Requiring or Not Veno-Venous Extracorporeal Membrane Oxygenation. *Membranes* 2021, 11, 603. [CrossRef]

44. Liu, J.; Shu, Y.; Zhu, F.; Feng, B.; Zhang, Z.; Liu, L.; Wang, G. Comparative efficacy and safety of combination therapy with high-dose sulbactam or colistin with additional antibacterial agents for multiple drug-resistant and extensively drug-resistant *Acinetobacter baumannii* infections: A systematic review and network meta-analysis. *J. Glob. Antimicrob. Resist.* 2021, 24, 136–147. [PubMed]

45. Russo, A.; Bassetti, M.; Ceccarelli, G.; Carannante, N.; Losito, A.R.; Bartoletti, M.; Corcione, S.; Granata, G.; Santoro, A.; Giacobbe, D.R.; et al. ISGRI-SITA (Italian Study Group on Resistant Infections of the Società Italiana Terapia Antinfettiva) Bloodstream infections caused by carbapenem-resistant Acinetobacter baumannii: Clinical features, therapy and outcome from a multicenter study. *J. Infect.* 2019, 79, 130–138. [CrossRef] [PubMed]

46. Huttner, B.D.; Catho, G.; Pano-Pardo, J.R.; Pulcini, C.; Schouten, J. COVID-19: Don’t neglect antimicrobial stewardship principles! *Clin. Microbiol. Infect.* 2020, 26, 808–810. [CrossRef] [PubMed]