Implant stability and bone remodeling up to 84 days of implantation with an initial static strain. An in vivo and theoretical investigation

Subsequent to implant placement, the healing process is initiated which eventually leads to formation of new bone immediately adjacent to the implant surface. Directly after implant installation, the implant stability depends solely on the mechanical contact between the surrounding bone (old bone) and the implant. During successful healing new bone formation enhances the implant stability over time (Cochran 2006; Coelho & Jimbo 2014). The degree of initial implant stability (degree of resistance to micromotion) is affected by the implant macrodesign and its relation to the osteotomy preparation. Micromotion of the implant gives rise to interfacial tissue deformations [i.e. strain] that in turn, affect the type of tissue that is formed [Morgan & Einhorn 2013; Coelho et al. 2015]. Thus, it is important that sufficient implant stability is obtained to achieve osseointegration. Szumkler-Moncler et al. [1998] suggested that the micro motions should be below 50-150 μm to avoid fibrous tissue encapsulation of the implant. Several theories have been developed to explain the mechanisms that control bone morphogenesis and tissue generation that is based on interfragmentary strain (Pauwels 1960; Perren & Cordey 1980; Carter et al. 1998). Perren & Cordey [1980] suggested that strain levels of less than 0.02 is a prerequisite for bone formation while Carter et al. [1998] proposed that the type of tissue that is formed depends on the hydrostatic pressure and by the tensile strain history. Naturally the bone-implant interfacial shear strength and implant stability depend on the type of tissue developed. Barewal et al. [2003] measured the implant stability by means of a resonance frequency method for various qualities of bone and reported that implants placed in bone of poor quality presented a decrease in resonance

Key words: bone condensation, implant stability, in vivo, remodeling, static strain

Abstract

Objectives: When implants are inserted, the initial implant stability is dependent on the mechanical stability. To increase the initial stability, it was hypothesized that bone condensation implants will enhance the mechanical stability initially and that the moderately rough surface will further contribute to the secondary stability by enhanced osseointegration. It was further hypothesized that as the healing progresses the difference in removal torque will diminish. In addition, a 3D model was developed to simulate the interfacial shear strength. This was converted to a theoretical removal torque that was compared to the removal torque obtained in vivo.

Material and methods: Condensation implants, inducing bone strains of 0.015, were installed into the left tibia of 24 rabbits. Non-condensation implants were installed into the right tibia. All implants had a moderately rough surface. The implants had an implantation time of 7, 28, or 84 days before the removal torque was measured. The interfacial shear strength at different healing time was estimated by the means of finite element method.

Results: At 7 days of healing, the condensation implant had an increased removal torque compared to the non-bone-condensation implant. At 28 and 84 days of healing, there was no difference in removal torque. The simulated interfacial shear strength ratios of bone condensation implants at different implantation time were in line with the in vivo data.

Conclusions: Moderately rough implants that initially induce bone strain during installation have increased stability during the early healing period. In addition, the finite element method may be used to evaluate differences in interlocking capacity.
frequency after 2–3 weeks. It was suggested that this phenomenon could be interpreted as a decrease of the implant stability. A plethora of research has been conducted to reduce the micromotion of the implant by modification of the Implant surface at the micro level in order to promote and enhance osseointegration (Coelho et al. 2015). As of today, the moderately roughened oral implants dominate the market, based on the scientific evidence that these surfaces provide rapid and strong bone response (Albrektsson & Wennergberg 2004; Wennergberg & Albrektsson 2009). A general trend in the in vivo experiments is that increased Sa value results in increased interfacial shear strength [Loberg et al. 2010]. By use of the finite element method Halldin et al. (2015) estimated the interfacial shear strength for different surface structures. It was found that a surface with a Sa value of 1.51 theoretically had a 45% increased interfacial shear strength at 12 weeks of healing compared to a surface with an Sa value of 0.91. Using linear regression between in vivo removal torque and Sa values presented by Loberg et al. (2010) resulted in an increase in removal torque of 48% for a Sa value of 1.51 compared to 0.91. In previously conducted in vivo studies by Halldin et al. (2011, 2014b), turned implants that induced bone strains of 0.015 (moderate) presented an increased removal torque at 3, 13, and 24 days compared to implants that did not induce bone strain [Halldin et al. 2011, 2014b]. The removal torque for this moderate bone condensation implant decreased over time due to reduced pressure on the surface [Halldin et al. 2014b]. The reduced pressure over time was suggested to be caused by an initially dominating viscoelastic relaxation and later dominating bone remodeling [Halldin et al. 2014b]. However, the removal torque of the moderate bone condensation implants seems to merge with that of the non-bone-condensation implants at ~30 days of healing [Halldin et al. 2014b]. To simulate the viscoelastic relaxation and remodeling behavior, Halldin et al. (2014a) developed a viscoelastic constitutive model. It was shown that with appropriate model parameters, the constitutive model captures the relaxation and remodeling behavior of bone which means that the model can be used to estimate the change of pressure on the implant surface over time. Thus, based on these theoretical findings and the previous studies with turned implants [Halldin et al. 2011, 2014b], it was hypothesized that bone condensation implants will enhance the mechanical stability initially and that the moderately rough surface will further contribute to the secondary stability by enhanced osseointegration.

Thus, as the healing progresses the difference in implant stability, as measured by removal torque between bone condensation implants and non-bone-condensation implants, will diminish. In order to test this hypothesis bone condensation implants and non-bone-condensation implants with a moderately rough surface were inserted in rabbit tibiae. The implant stability over time was compared by the means of removal torque measurement. Furthermore, it was hypothesized that by using the finite element method the interfacial shear strength of a bone condensation implants may be estimated.

### Material and methods

#### Implant design

Specially designed screw-shaped implants of titanium (grade 4) were manufactured with tight tolerances to induce controlled bone strains. The designs of the implants were identical to the implants in the study by Halldin et al. (2014b). The test implants comprised of a condensation portion that had an increased diameter of 0.05 mm compared to the cutting portion. These bone condensation implants induced moderate bone strains of 0.015. The control implants had no diametrical increase. All implants had the same surface as the commercially available OsseoSpeed™ implants (DENTSPLY Implants, Mölndal, Sweden).

#### Interferometry

To confirm that both implant groups had the same surface topography the implant surfaces were characterized by use of optical light interferometry (MicroXAM, ADE Phase shift Technology, Inc., Tucson, AZ, USA). In brief, six randomly selected implants (three controls and three tests) were selected. Each implant was measured on three thread peaks, three valleys, and three flanks (scan area of 264 x 200 μm, vertical measurement range of 100 μm). Data evaluation was performed with the MountainMaps software (Digital Surf, Besançon, France). WAViness and form were filtered with a 50 x 50 μm Gaussian filter. The surfaces were characterized with the surface roughness parameters S5 [μm], Sdr (%), and Sdr [1/μm²] (definition of them can be found elsewhere [Stout 2000; Thomas 1999]).

#### Animal study

Twenty-four mature female New Zealand white rabbits (weight 3–4 kg) were used in this study. The study was approved by the French MINISTÈRE DE L’ENSEIGNEMENT SUPERIEUR ET DE LA RECHERCHE and performed at the ÉcoleNationaleVétérinaire’ Alfort [Maisons-Alfort, Val-de-Marne, France, approval number, 00391-01]. Prior to surgery, 250 μg/kg of medetomidine [Domitor®, Zoetis, France], 20 mg/kg ketamine [Imalgène 1000®, Merial, Sanofi, France], and 1 mg/kg of diazepam [Valium®, Roche, France] was injected intramuscularly to provide general anesthesia. Subsequently, analgesics 30 μg/kg, buprenorphine [Buprécare, Animalcare, York, UK] was injected subcutaneously and 0.2 mg/kg meloxicam [Metacam®, Boehringer Ingelheim Vetmedica, Inc., St. Joseph, MO, USA] was injected intramuscularly. Thereafter, incision of the skin was made with a #15 blade and the muscle layers and periosteum were elevated and separated from the bone. Osteotomy was prepared in the proximal tibia by drilling with a sequence of burs, starting from a round bur, 2.8 mm drill and finally with a 3.3 mm drill, corresponding to the core diameter of the implants. The osteotomies were performed under constant irrigation with physiological saline solution. Each rabbit received two test implants in one leg (proximal and distal placement) and two control implants on the contralateral side. The implants were inserted with a rotation speed set to 25 revolutions/min using the W&H implant unit (Elcomed, W&H SA-310, Burmoos, Austria). The installation stopped when the superior thread was flush with the cortical bone surface. After implant insertion, the muscle layers were sutured with a resorbable suture (Vicryl3.0), and the skin with a 4-0 nylon suture (Ethicon, Auneau, France). Post-surgically, a patch of fentanyl [25 μg/h, Duragesic®; Janssen Pharmaceutica, Beerse, Belgium] was applied to each animal for 3 days. An antibiotic [enrofloxacin, 200 mg/l], Baytril®; Bayer Animal Health, Leverkusen, Germany] was administrated in the water supply for 5 days. The rabbits were kept in separate cages and were allowed to move and eat freely. At 7, 28, and 84 days after implant placement, eight rabbits each at time point were euthanized with an overdose injection of sodium pentobarbital [Euthasol, Virbac, Fort Worth, TX, USA]. Each tibia was disected and excessive soft and hard tissue around the implant square head was carefully removed.

#### Removal torque measurement

The bone samples were firmly secured with several individual bone pins and the removal torque of the implants was measured. The removal torque, time, and angle of rotation
were recorded with a calibrated torque measurement device (DR-2112 Lorenz Messtechnik GmbH, Alfdorf, Germany) using a constant rotational speed of 0.03 rpm. The torque device was calibrated in the range 50–2000 Nmm with an accuracy of 1 Nmm. The data were sampled with 100 Hz and thereafter analyzed in MATLAB software 2013b (Mathworks Inc., Natick, MA, USA). The data were initially smoothed with a running average over a set of 50 data points and the angle of rotation was set to 0 rad when the torque reached 10 Nmm. The fracture torque \( T_f \) was identified as the maximum torque.

**Theoretical interfacial shear strength**

In the present study, the test implant induced a pressure at the threads in the condensation region. The magnitude of pressure at the threads was simulated by a radial displacement of the thread profile of 0.025 mm as described in the 2D-axis symmetric model of Halldin et al. (2011). The 2D simulations were performed using bilinear mature bone material properties without hardening (Table 1). The reduction in pre-stress over time, due to relaxation and remodeling, of mature bone was simulated by use of the constitutive model, illustrated by the rheological model [Fig. 1], [Halldin et al. 2014a] with model parameter values according to Table 6. In brief: the parameters values obtained for simulation [Halldin et al. 2014a] of the Crowninshield & Pope (1974) experiment were used but recalibrated to represent rabbit bone. The stiffness of springs 1–3 [Fig. 1a] were adjusted to 5008, 2166 and 776 MPa, respectively, to represent Young's modulus of mature rabbit cortical bone (7950 MPa) [Isaksson et al. 2010]. Hence, by changing the spring stiffness values the viscosity was consequently adjusted to fit the relaxation behavior [Halldin et al. 2014a]. Thereafter, the initial strain was increased to achieve a pressure similar to the theoretical thread pressure, simulated in the 2D finite element method, of the test implant. Finally, the remodeling parameter was set to result in no pressure at 30 days which represents the expected theoretical time point when the initially induced bone strainshave vanished (Halldin et al. 2014b). The theoretical interfacial shear strength of the moderately rough surface was simulated using the finite element model described in Halldin et al. (2015). In brief, a representative patch of the implant surface was selected and a finite element model was developed according to Fig. 1b. The implant was moved until bone failure occurred which was assumed to represent the interfacial shear strength. In the current investigation the interfacial shear strength of a bone condensation implant was simulated with adding an external pressure (Fig. 1b). The pressure was reduced by use of the constitutive model [Halldin et al. 2014a] to represent relaxation and remodeling material behavior during healing. The control implant was consequently simulated with no pressure (Fig. 1b). In the present study the theoretical interfacial shear strength was simulated both with mechanical properties of healing bone and mature bone (Halldin et al. 2014a) (Table 1).

**Statistical analysis**

Pairwise differences in removal torque measurements between test and control implants that were assumed to be normally distributed were analyzed with Student's t-test. Pairwise

### Table 1. Mechanical properties of bone during healing derived from mineralization level during healing bone and the relationship between mineralization level and mechanical properties Halldin et al. (2014a,b)

| Healing time | Young's modulus \( E \) (MPa) | Ultimate strain \( \varepsilon_u \) | Yield strain \( \varepsilon_y \) (MPa) | Comment |
|-------------|-------------------------------|-------------------|-----------------|---------|
| 0.57        | 4                             | 2096              | 0.1153          | 28.4    | Material properties of healing bone (Halldin et al. 2015) |
| 1           | 7                             | 2200              | 0.1122          | 29.3    | Young's modulus of mature rabbit bone (Isaksson et al. 2010) |
| 2           | 14                            | 2738              | 0.1004          | 34.2    | and with the ultimate strain and yield strain (Halldin et al. 2015) |
| 4           | 28                            | 3161              | 0.0918          | 37.5    |  |
| 4.28        | 30                            | 3189              | 0.0914          | 37.8    |  |
| 12          | 84                            | 4005              | 0.0801          | 44.3    |  |
| 50          | 350                           | 7950              | 0.0506          | 71.3    |  |

Fig. 1. (a) Rheological model and the model parameters for the constitutive model developed by Halldin et al. (2014a). (b) FEA Model to simulate the interfacial shear strength of a rough surface exposed to a pressure. (c) Implant design to induce controlled static bone strain.
differences in measurements between test and control that were not assumed to be normally distributed were analyzed using Sign Test for Median. Pairwise ratios [test/control] for different healing times were analyzed using Mood Median Test to test the equality of medians from two populations. Interferometry measurements were analyzed using Kruskal–Wallis Test (assumed equal shape). Significance level was set to \( P < 0.05 \).

Results

**Interferometry**

There was no difference \( (P = 0.287) \) in \( S_a \) value of test implants [mean 1.55 \( \mu \text{m} \); std 0.21] compared to control implants [mean 1.49 \( \mu \text{m} \); std 0.19]. There was no difference \( (P = 0.441) \) in \( S_{vk} \) value of test implants [mean 109\%; std 44] compared to control [mean 103\%; std 49] and no difference \( (P = 0.337) \) in \( S_{ds} \) value of test implants [mean 0.061/\( \mu \text{m}^2 \); std 0.005] compared to control implants [mean 0.061/\( \mu \text{m}^2 \) 0.007]. Therefore, the surface structure can be assumed similar.

**Animal study**

The results of the removal torque measurement [Nmm], for the three different implantation times (7, 28, and 84 days) are presented in a boxplot (Fig. 2a) and Table 2. One sample [Tibia Proximal 28 days of healing] was unfortunately lost during carefully removing the excessive soft and hard tissue. The corresponding contralateral implant was consequently excluded in the analysis. The ratios between test and control, placed distally, of the present study [at 7, 28, and 84 days].

**Theoretical interfacial shear strength**

By use of the 2D axisymmetric model a theoretical average normal pressure on the thread profile of 39.5 MPa was obtained. By applying appropriate parameter values the relaxation behavior of mature rabbit bone is captured quite well [Fig. 3a]. Using the constitutive

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![Fig. 2](image_url)  
(a) Individual scatter plots of the removal torque for different implantation times. (b) Removal torque ratios of the present study and the study by Halldin et al. (2011, 2014b). \( P < 0.05 \) indicates a significant difference in ratios over time.

**Table 2.** Mean and median of removal torque values and the statistical analysis between the pairwise differences of test and control and the corresponding ratios

| Healing time (days) | Site     | # Samples | Test        | Control       | P-value | t-Test | Sign test for median | Mean pairwise ratio |
|---------------------|----------|-----------|-------------|---------------|---------|--------|----------------------|---------------------|
|                     |          |           | Mean torque (SD) Nmm | Median Nmm | Mean torque (SD) Nmm | Median Nmm | | | |
| 7                   | Proximal | 8         | 83 (50)     | 75            | 39 (18)  | 32     | 0.041$^1$            | 2.3                 |
| 7                   | Distal   | 8         | 136 (54)    | 122           | 47 (23)  | 50     | 0.002$^1$            | 4.3                 |
| 28                  | Proximal | 7         | 269 (72)    | 298           | 281 (135)| 294    | 0.453$^3$            | 1.2                 |
| 28                  | Distal   | 8         | 314 (120)   | 315           | 371 (134)| 300    | 0.249$^1$            | 0.9                 |
| 84                  | Proximal | 8         | 531 (150)   | 550           | 489 (139)| 468    | 0.289$^3$            | 1.1                 |
| 84                  | Distal   | 8         | 597 (235)   | 608           | 488 (80) | 492    | 0.093$^1$            | 1.3                 |

Subscription is commonly used to differentiate between different statistical methods.
model, with an initial strain to obtain an initial pressure of 39.5 MPa and a tuned remodeling parameter value that eliminated the pressure after 30 days, the decrease in pressure on the implant surface during healing was simulated (Fig. 3b). The parameter values used in the constitutive model to simulate the decrease in pressure over time are presented in Table 3. The theoretical interfacial shear strength, with mechanical properties of healing bone together with no pressure over time (represents control implants, simulation C), is presented in Fig. 4a. The theoretical interfacial shear strengths, with mechanical properties of healing bone together with decreased pressure over time (represents test implants, simulation B) and mature bone mechanical properties together with decreased pressure over time (represents test implants, simulation A), are presented in Fig 4a. The ratios between test and control of the in vivo study and the corresponding theoretical ratios are presented in Fig. 4b.

Discussion

An in vivo study was conducted to investigate how the implant stability was affected by a moderately rough implant that induced moderate bone strains of 0.015 at the time of implant installation. The implants were allowed to heal in the rabbit tibiae for 7, 28, and 84 days. It was hypothesized that bone condensation implants will enhance the mechanical stability initially and that the moderately rough surface will further contribute to the secondary stability by enhanced osseointegration. The bone condensation implants (i.e. test implants) presented an increased removal torque at 7 days of healing compared to the non-condensation implants (i.e. control implant), whereas at 28 and 84 days of healing no difference in removal torque between bone condensation implants and non-bone condensation implants (Fig 2b) was observed. In the previous in vivo studies by Halldin et al. (2011, 2014b) turned implants with the same moderate bone condensation level were used. The turned implants had a healing time of 3, 13, and 24 days and the removal torque values were measured with a removal torque device different to that of the one utilized in the current study. In the previous studies, the test (moderate bone condensation) implants presented significantly increased removal torque compared to control implants. The removal torque values of the two different removal torque devices cannot directly be compared. However, if the removal torque ratios (test/control) are calculated respectively for study, they may be compared. Interestingly, there was a similar trend of decreased ratio over time regardless

Table 3. Model parameters used in the constitutive model (Fig. 2) proposed by Halldin et al. (2014a,b) to simulate the change of pressure over time

| Model parameters | Value | Comments |
|------------------|-------|----------|
| $E_1$ [MPa] | 5009 | Represent mature bone Young’s modulus of 7950 MPa which was distributed according to ratios found in Halldin et al. (2014a,b) |
| $E_2$ [MPa] | 2166 | |
| $E_3$ [MPa] | 775 | According to the Crowninshield and Pope parameter values obtained in Halldin et al. (2014a,b) |
| $E_4$ [MPa] | 2065 | According to Crowninshield and Pope parameter values obtained in Halldin et al. (2014a,b) |
| $E_5$ [MPa] | 11,699 | |
| $v_2$ [MPa s] | 5.55E-08 | Calibrated to fit the relaxation behavior in Halldin et al. (2014a,b) with the Young’s modulus of 7950 MPa (fig. 6a) |
| $v_3$ [MPa s] | 7.48E-04 | The Young’s modulus of 7950 MPa (fig. 6a) |
| $v_4$ [MPa s] | 6.90E-02 | According to Crowninshield and Pope parameter values obtained in Halldin et al. (2014a,b) |
| $v_5$ [MPa s] | 1.60E-02 | |
| $H$ [MPa] | 114 | According to Crowninshield and Pope parameter values obtained in Halldin et al. (2014a,b) |
| $p^p$ | 13.5 | |
| $m_{2}$ [MPa s] | 46.6 | |
| $m_{3}$ [MPa s] | 3.0E+10 | |
| $m_{4}$ [MPa s] | 6.90E-02 | According to Crowninshield and Pope parameter values obtained in Halldin et al. (2014a,b) |
| $m_{5}$ [MPa s] | 1.60E-02 | |
| $H$ [MPa] | 114 | According to Crowninshield and Pope parameter values obtained in Halldin et al. (2014a,b) |
| $H$ [MPa] | 114 | |
| $R$ [s] | 2.0E-09 | Calibrated to eliminate the initial pressure after 30 days (fig. 6b) |
of the surface micro roughness (turned or moderately roughened) [Fig. 2b]. The ratios were considerably higher than 1 (or 100%) during the early healing times, which, in turn, means higher removal torque values for the test implants compared to the control implants. This suggests that the moderate bone condensation implants (test) enhances implant stability during the initial critical phases of healing. When the osseointegration consolidates (at 28 and 84 days) the ratios between test and control tend to reach the value 1 (Fig. 4b). This indicates that the affect moderate bone strains has on implant stability has diminished. It is interesting to observe that the ratio was higher of turned implants at 3 days compared to the ratio of the moderately rough implants at 7 days. These might be related to the fact that the controlled condensation is particularly effective for the retention the first few days of healing (3 days vs. 7 days) regardless of surface structure. The absolute removal torque values of moderately rough surface were increased during healing for both test and control implants, which attests to the favorable bone formation around all implants with a moderately rough surface structure.

An interesting reflection from the previous studies of turned implants is that the removal torque decreased over time for the test implants. This might reflect that the decreased pressure, due to relaxation and remodeling, had a more pronounced effect on removal torque than the interlocking of a turned surface structure. In the present study the removal torque increased over time. Therefore, it might be speculated that the increased interlocking capacity of the moderately rough surface had a more pronounced effect on the removal torque than the reduction in pressure, due to relaxation and remodeling, over time. It should be noted that the removal torque values, especially for the longer healing times, might be affected by bone growth into the cutting feature affecting the results. In addition, the potential grinding effect a structured surface has on the bone might influence the result. The present study used a rabbit model which comprises of a dense cortical bone layer of 3–4 mm and it does not incorporate the implant stability of a potential trabecular bone. The ratios of the present in vivo study were compared to the theoretical interfacial shear strength ratios simulated by the use of the 3D finite element model. Interestingly, it was found that the simulated ratios are in line with the in vivo ratios. In the 3D simulation the strength of the interfacial bone was increased over time, reflecting healing bone and increased mineralization, that increased the interlocking capacity [Halldin et al. 2015]. In the present simulation the interfacial shear strength was simulated both with constant mechanical properties of mature bone and mechanical properties representing healing bone. It may be speculated that mature bone has sporadic contact with the surface structure. Therefore the simulated shear strength of mature bone was assumed to provide the highest interlocking capacity that can be achieved during healing, thus reflecting only how a decreased pressure affects the interfacial shear strength. In the 3D finite element model the decreased pressure on the surface over time was obtained by use of the constitutive model simulating the relaxation and remodeling behavior of cortical bone (Halldin et al. 2014a). The characteristics of the actual mechanical properties of the in vivo bone during healing are unknown. Therefore, the simulation was based on the estimated mechanical properties of the interfacial bone obtained in the study by Halldin et al. [2015]. In the present study, the parameters of the constitutive model described in Halldin et al. [2014a] were recalibrated to fit the viscoelastic relaxation and remodeling behavior of rabbit mature bone. It is assumed that when analyzing the ratios these assumptions might have a limited impact. Despite these assumptions, regarding the healing bone mechanical properties and the relaxation and remodeling behavior of rabbit bone [Fig. 3a, b], the simulated ratios had the same trend as the ratios obtained the in vivo experiment [Fig. 4b]. This implies that finite element analysis, with appropriate material models, may be used to evaluate differences in structural interlocking capacity between two surfaces. In addition, it seems to be a more rapid decrease in ratios during the initial healing which is in line with the findings of Perren et al. (1969) and Cordey et al. (1976). The results of the study suggest that modification of the implant macro geometry in

**Fig. 4.** (a) Simulation of the interfacial shear strength for the test implants with constant mechanical properties of mature bone during healing time and mechanical properties of bone during healing time. Simulation of the interfacial shear strength for the control implants was performed with mechanical properties of bone during healing time. (b) The ratios of the present in vivo study and of the simulations.
relationship to the osteotomy preparation, to induce moderate bone strain levels of 0.015 during implant insertion does not result in reduction in stability of moderately rough implants. It is interesting to note that the ratios using finite element method were in line with the in vivo ratios. This is an indication that when the condensation level is controlled in vivo, the biological outcomes may be predicted, which could optimize implant design.

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**Conflicts of interests**

Mr. Halldin is employed at DENTSPLY Implants. The other authors have no conflict of interests.

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