Interpretation and application of carbon isotope ratios in freshwater diatom silica

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ABSTRACT: Carbon incorporated into diatom frustule walls is protected from degradation enabling analysis for carbon isotope composition ($\delta^{13}$C$_{\text{diatom}}$). This presents potential for tracing carbon cycles via a single photosynthetic host with well-constrained ecophysiology. Improved understanding of environmental processes controlling carbon delivery and assimilation is essential to interpret changes in freshwater $\delta^{13}$C$_{\text{diatom}}$. Here relationships between water chemistry and $\delta^{13}$C$_{\text{diatom}}$ from contemporary regional data sets are investigated. Modern diatom and water samples were collected from river catchments within England and lake sediments from across Europe. The data suggest dissolved, biogenically produced carbon supplied proportionately to catchment productivity was critical in the rivers and soft water lakes. However, dissolved carbon from calcareous geology overwhelmed the carbon signature in hard water catchments. Both results demonstrate carbon source characteristics were the most important control on $\delta^{13}$C$_{\text{diatom}}$ with a greater impact than productivity. Application of these principles was made to a sediment record from Lake Tanganyika. $\delta^{13}$C$_{\text{diatom}}$ co-varied with $\delta^{13}$C$_{\text{bulk}}$ through the last glacial and Holocene. This suggests carbon supply was again dominant and exceeded authigenic demand. This first systematic evaluation of contemporary $\delta^{13}$C$_{\text{diatom}}$ controls demonstrates that diatoms have the potential to supply a record of carbon cycling through lake catchments from sediment records over millennial timescales.

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KEYWORDS: carbon cycling; diatom frustule carbon; Lake Tanganyika; palaeoclimate; stable carbon isotopes.

Introduction

Stable isotope analyses of the siliceous cell walls (frustules) of diatoms provide insights into a broad range of environmental processes tracked from the perspective of a single, ecologically well-constrained organism. To date, most diatom-based stable isotope studies have focused on the stable oxygen and silicon isotope composition ($\delta^{18}$O$_{\text{diatom}}$ and $\delta^{30}$Si$_{\text{diatom}}$) of diatoms from lacustrine and marine sediments (Leng and Barker, 2006; Swann and Leng, 2009; Leng and Henderson, 2013). Changes in $\delta^{18}$O$_{\text{diatom}}$ are used as a proxy record of water source and hydrological balance in palaeolimnology (Barker et al., 2001; Rioual et al., 2001; Shemesh et al., 2001) and global ice volume, temperature and local effects in palaeoceanography (Shemesh et al., 1995, 2002; Hodell et al., 2001). $\delta^{30}$Si$_{\text{diatom}}$ in freshwater is used to understand changes in climate, weathering and soil processes through the balance of silicon supply and demand (De La Rocha et al., 2000; Ding et al., 2004; Street-Perrott et al., 2008). Within marine environments, utilization of dissolved silica can be reconstructed through the ratio of silicic acid uptake by diatoms to initial dissolved concentrations (De La Rocha et al., 1998; Varela et al., 2004; Cardinal et al., 2005).

Diatom frustules are also a host for carbon isotopes measured on organic molecules occluded within diatom frustule walls ($\delta^{13}$C$_{\text{diatom}}$). This occluded organic matter comprises proteins and long-chain polyamines (Kröger and Poulsen, 2008) and represents a source of carbon potentially protected from degradation over geological timescales (Singer and Shemesh, 1995; Crosta and Shemesh, 2002). During cell formation diatoms source this carbon via photosynthetic uptake from the surrounding water body. The fraction available for photosynthesis is dissolved inorganic carbon (DIC), which diatoms take up preferentially as dissolved CO$_2$ or as bicarbonate under conditions of high carbon demand (Giordano et al., 2005). $\delta^{13}$C$_{\text{diatom}}$ provides a record of changes in this carbon pool, overcoming issues of sample heterogeneity and potential diagenesis associated with investigations of stable carbon isotopes of bulk organic material ($\delta^{13}$C$_{\text{bulk}}$) from sediments.

Use of $\delta^{13}$C$_{\text{diatom}}$ as a palaeoenvironmental proxy is already well established within palaeoceanography where $\delta^{13}$C$_{\text{diatom}}$ is usually interpreted as a record of pelagic primary productivity as discrimination against $^{13}$C by diatoms is reduced during periods of high carbon demand (Singer and Shemesh, 1995; Crosta and Shemesh, 2002; Schneider-Mor et al., 2005). However, care is required in interpretation of $\delta^{13}$C$_{\text{diatom}}$ records, as further biological variables with potential to impact carbon fractionation and isotope composition are yet to be fully constrained (Jacot Des Combes et al., 2008). These variables include diatom species assemblage, carbon availability and carbon source.

Two key factors that determine the degree of fractionation during photosynthetic carbon uptake are the balance between internal and external CO$_2$ concentrations and discrimination by the enzyme RuBisCO (Jacot Des Combes et al., 2008). The impact of these factors is compounded by species-specific ’vital effects’, including cell growth rate, geometry and

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growth environment. For example, high growth rates reduce the internal to external CO₂ ratio (Hill et al., 2008). Discrimination by RubisCO to 13C is theoretically proportional to this ratio (Korb et al., 1996; Hill et al., 2008), and fractionation of carbon will subsequently be less in faster growing cells. Conversely, where cell geometry maximizes the surface area to volume ratio, CO₂ is more efficiently absorbed leading to a relative increase in carbon fractionation (Popp et al., 1998). Planktonic species have also been associated with lower δ13C values compared with benthic varieties, attributed to the more turbulent growth environment of the former, which reduces the impact of boundary layer thickness on carbon uptake (France, 1995; France and Cattaneo, 1998; Wang et al., 2013).

Confinement of carbon isotopic analysis to the initial protein matrix established during cell formation represents a carbon source less likely to be affected by such species-specific effects. In fact, tests of the impact of different diatom species composition on δ13C diatom from freshwater Lake Challa, Mount Kilimanjaro, were within analytical error (Hurrell et al., 2011). To reduce the risk of any vital or species-specific effects, it is recommended within the more established field of palaeoceanography to sieve to the <20-μm fraction as it is here that most diatom material is found and assemblages tend to be dominated by fewer species (Crost and Shemesh, 2002).

Availability of DIC is also a key determinant of carbon fractionation, as slow diffusion of dissolved CO₂ through water risks transport limitation. To prevent this, carbon concentrating mechanisms (CCMs), which manifest as active uptake of dissolved CO₂ and/or bicarbonate, are believed to take place in almost all diatoms (Giordano et al., 2005). Utilization of bicarbonate by diatoms can result in a further increase in δ13C of the photosynthate by approximately 9‰ (Finlay, 2004). Whilst interpreting δ13C diatom records it is therefore important to consider whether an increase in value has been enhanced by carbon transport limitation.

Of particular relevance to interpretation of freshwater δ13C diatom is the carbon isotope composition of DIC sources (δ13C DIC). A far greater variety of carbon sources is found within terrestrial environments compared with the open ocean. The associated δ13C DIC is in turn diverse, ranging from 0 to +1‰ for carbonate bedrock and from −26 to −20‰ for soil carbon in C4 landscapes, for example (Clark and Fritz, 1997). The expression of these different origins is then modified by mixing and fractionation during carbon phase transformation and species changes in transfer from the catchment to the water body, or because of internal aquatic processing (Finlay, 2003).

As a possible consequence of greater complexity introduced by more diverse carbon sourcing, there have been far fewer studies of δ13C diatom in palaeclimatology compared with palaeoceanography. Interpretations reached have also been inferential rather than reconstructions grounded in, and constrained by, modern environmental data (Hernández et al., 2011, 2013; Barker et al., 2013). For example, in Lago Chungará, Chile, diatoms deposited over the late glacial to early Holocene period have high δ13C diatom values (−27 ± 5 to −22.6‰) during arid stages compared with those of wetter, humid periods (−30.3 to −25.4‰) when greater input of 13C depleted dissolved biogenic carbon to the lake from the catchment was likely (Hernández et al., 2011, 2013). Similarly, a sediment core from Lake Challa, Mount Kilimanjaro, displayed positive correlation between δ13C diatom and δ13C DIC during dry intervals as high diatom productivity depleted the lake DIC pool. This correlation largely broke down during wetter periods and it was hypothesized that increased catchment carbon loading satisfied demand from primary productivity (Barker et al., 2013). These lake sediment records demonstrate an application for δ13C diatom in tracing catchment carbon cycling, and the importance of testing the down-core changes against contemporary environmental relationships.

Here we (i) explore relationships between water chemistry and δ13C diatom from contemporary data sets to more precisely determine the environmental controls of δ13C diatom, and (2) apply the understanding gained to assess the utility of δ13C diatom in unravelling temporal carbon dynamics. Firstly, diatom epilithon (diatoms extracted from submerged biofilm comprising other algae, bacteria, fungi and the products they secrete) from river reaches in north-west England were sampled to provide an understanding of detailed carbon dynamics at the catchment scale. Rivers were investigated as they represent active hydrological pathways, connecting a lake with its catchment. Secondly, δ13C diatom from sediment samples of lakes situated across central, north-west and northern Europe were analysed. These sediments were collected as part of a broader sampling campaign designed to study the relationship between methane concentrations, δ13C of DIC and methane, and the carbon isotopic composition of aquatic invertebrate fossils and other sediment components (e.g. Schilder et al., 2015; Stötter et al., unpublished data). The lakes are geographically dispersed, spanning different climate zones ranging from temperate to boreal and incorporating diverse catchment geology with examples of calcareous and non-calcareous lithology (Rinta et al., 2015). Thirdly, we applied the method to core material from Lake Tanganyika, East African Rift Valley. This lake has a well-established palaeoenvironmental history combined with deposition of sediments in anoxic conditions allowing for ideal organic carbon preservation. Methodological refinements were made to reduce and assess any impact of species and vital effects on the resulting δ13C diatom values.

Study sites

Contemporary UK river sites

Sampling of the north-west England river catchments for epilithon and spot water chemistry samples took place in May 2012 over consecutive days to minimize hydroclimate variability. Collection from riffles in 20 river reaches captured the late spring diatom bloom. Six major river catchments were targeted: the Wyre, Ribble, Lune, Derwent, Leven and Eden, situated in North Lancashire and Cumbria (Fig. 1). Land use was largely agricultural with land proximal to the river and stream sites dominated by rough and improved pasture for grazing. The geology underlying the sampling sites consisted of combinations of sedimentary rocks (sandstones, siltstones, mudstones and gritstones) with volcanic rocks at the most easterly sites.

Contemporary European lake sites

The European lake study consisted of surface sediments analogous to core tops, and spot water chemistry samples from 30 relatively small (0.3–303 ha) lakes situated in five countries; Switzerland, the Netherlands, Germany, Sweden and Finland (Fig. 2). Water samples were collected 0.5 m below the surface in late summer before the breakdown of water stratification in the autumn (Rinta et al., 2015). The Swiss, Dutch and German lakes are located in the temperate zone, whereas the Finnish and Swedish lakes are in the hemiboreal to boreal zones. The underlying lithology of the Swiss, Dutch and German lakes is dominated by Quaternary sediments and older limestones. In
contrast, most of the Finnish and Swedish lakes are underlain by non-calcareous Precambrian bedrock covered by Quaternary deposits.

Lake Tanganyika sediments

The Lake Tanganyika down-core study comprised 14 sediment samples taken from Kullenberg piston core NP04-KH04-4A-1K collected in 2004 as part of the Nyanza Project (Felton et al., 2007). The pelagic zone of meromictic Lake Tanganyika is highly sensitive to catchment changes that alter the carbon and nutrient concentrations. The core was taken from the Kalya Horst, which is a structural high within the southern basin of Lake Tanganyika (Fig. 3). The coring location was situated below the oxycline, the anoxic state providing ideal conditions for organic carbon preservation. Sediments were dated to the last ca. 34 000 years by correlation to a second, directly radiocarbon-dated core (NP04-KH04-3A-1K) (Tierney et al., 2008) using 20 age/depth control points.

Methods

Pretreatment and measurement of $\delta^{13}C_{\text{diatom}}$

Successful determination of $\delta^{13}C_{\text{diatom}}$ relies on the removal of all sources of both inorganic and organic carbon external to the frustule inclusions. To produce clean diatom material from the samples with variable organic carbon content we adapted the method described by Hurrell et al. (2011), which was based on that of Singer and Shemesh (1995). All samples were first passed through a 1-mm gauge sieve then heated to

Figure 1. Location of the 20 UK river sites in this study. Lakes and watercourses are also shown.

Figure 2. Location of the 30 European lake sites in this study.

Figure 3. Location of the coring site in Lake Tanganyika, situated in the East African Rift Valley.
70°C in 10% HCl for 2 h to remove inorganic carbon. For the Lake Tanganyika material a sub-sample was removed at this stage for δ13Corg.

For the determination of δ13Cdiatom, organic carbon (excluding the occluded material) was removed through oxidation by heating samples in 30% H2O2 for 15 h at 70°C and a further 2 h at 100°C. Persistent organic carbon was eliminated through heating samples to 70°C for 1 h in concentrated HNO3. Large mineral grains were separated by differential separation and discarded. Clay and silt particles with similar densities to diatoms were reduced by sieving to 20 μm. Samples with no more than 1% carbon content were considered free from contamination following Hurrell et al. (2011). Sieving to 20 μm was also completed to reduce potential influence of species effects attributed to cell size and geometry (Crosta and Shemesh, 2002). Permanent slides were made of the processed samples and outline diatom counts were made based on 150 valves. For the contemporary samples, diatom genera were categorized by growth habitat as planktonic, benthic or colonial after Bellinger and Sigee (2010) as a further check for possible confounding vital effects.

δ13C/12C ratios for diatom material and bulk organic material from sediments were determined using an Elemental vario PYRO cube elemental analyser linked to an IsoPrime100 isotope ratio mass spectrometer at Lancaster University for contemporary UK river samples and Lake Tanganyika sediments, and at Isoprime UK in Cheadle for contemporary European lake samples. Analysis was by combustion within tin capsules at 950°C. 13C/12C ratios were corrected against VPDB using within-run analysis of standards (IAEA-CH-6 (sucrose), Low Organic Content Soil Standard OAS and High Organic Content Soil Standard OAS (assuming δ13C values of -20.45‰ (International Atomic Energy Agency, 2011), -27.46 and -26.27‰ (Elemental Microanalysis, 2011), respectively). Data are reported in the usual delta notation; within-run replication of standard materials was <0.2‰ (1 SD, n=10). To ensure consistency between laboratories and conditions of analysis, external precision was monitored by use of a standard material analysed between all run sequences <0.2‰ (1 SD, n=164). Precision of sample analysis was <0.2‰ (1 SD, n=3). Where n=2, sample replicates did not vary by more than 0.5‰.

Water analysis

In-stream spot measurements of river site pH and electrical conductivity (EC) were taken using a WTW Multi 340i multiparameter water meter. Measurement accuracy was to 0.03 pH units and 1 μS cm⁻¹. Analysis of river water samples was completed at Lancaster University. Total phosphorus (TP) was measured following an acid-persulfate digest (O’Dell, 1993) using a Seal Analytical AQ2+ discrete colorimetric analyser (Seal Analytical, 2005). Total dissolved nitrogen (TDN) was measured using an Analytical Sciences Thermalox analyser (BS EN, 2003). Detection limits (standard deviation of blanks multiplied by 3) for TP and TDN analysis were 0.005 and 0.13 mg L⁻¹, respectively.

For determination of δ13C of the DIC pool (δ13CDIC) at the river sites, 10 μL of river water was injected into 12 mL pre-evacuated extainers containing 150 μL of de-gassed, concentrated phosphoric acid after Waldron et al. (2007). 13C values of the product CO2 were measured at the NERC Centre for Ecology and Hydrology, Lancaster, using a GV Instruments Tracegas Pre-concentrator coupled to an IsoPrime isotope ratio mass spectrometer. The isotope ratio of the resultant CO2 was compared with pulses of known reference CO2 and expressed relative to VPDB. Data are reported in the usual delta notation; within-run standard replication (1 SD) was better than or equal to ±0.15‰.

Lake water spot samples and measurements were taken in the deepest part of each lake basin using a 5-litre water sampler approximately 0.5 m below the surface as described in detail by Rinta et al. (2015). pH and EC were measured in the field using a pHScan 2 and WTW LF 330 device with TetraCon conductivity measuring cell, respectively. TP, total nitrogen (TN) and δ13CDIC were determined using laboratory methods as described by Rinta et al. (2015).

Results

Contemporary UK river sites

At least 90% of each assemblage consisted of the same nine benthic diatom genera. "Achnanthidium" was present in all assemblages, generally as A. minutissimum, and was typically dominant alongside Gymnophona, Cocconeis and Cymbella species. No systematic correlation was found between δ13Cdiatom and species composition in these data.

River water pH values ranged from 6.1 to 8.5 and EC from 16 to 331 μS cm⁻¹ (Table 1). The nutrient concentrations confirmed these streams to have low-to-moderate trophic status (EA, 1998), with several TP measurements below detection and maximum TP and TDN values of 0.052 and 1.10 mg L⁻¹, respectively. This is consistent with low-intensity farming practices that dominated the sampled area of northwest England.

δ13CDIC of the UK river waters ranged between -11.5 and -2.9‰. In contrast, δ13Cdiatom had a smaller range, lying between -28.9 and -25.3‰. Correlation analysis using Spearman’s rank correlation coefficient (IBM SPSS) identified a significant positive relationship between δ13CDIC and δ13Cdiatom (rs = 0.70, P < 0.01) (Fig. 4). Significant negative relationships were also identified between TDN concentrations and δ13CDIC (rs = -0.77, P < 0.01) in addition to TDN concentrations and δ13Cdiatom (rs = -0.73, P < 0.01) (Fig. 5).

Contemporary European lake sites

The lake sediment diatom assemblages comprise planktonic life forms alongside benthic and colonial examples making them more diverse than the river assemblages. Dominant genera included planktonic Cyclotella, Aulacoseira and Stephanodiscus species. Despite this diversity, no systematic correlation was found between δ13Cdiatom, the species data or the proportions of different life forms.

In comparison with the UK river study set, larger ranges in water chemistry values were measured in the lake waters. The pH values ranged between 5.4 and 8.9 and EC values from 24 to 462 μS cm⁻¹ (Table 2). Trophic conditions varied from ultra oligotrophic to hypertrophic (OECD, 1982), reflecting a wide range of nutrients with maximum TP and TN values of 0.12 and 2.30 mg L⁻¹, respectively. δ13CDIC values varied from -23.6 to -2.7‰, and two distinct clusters were observed: the Swiss, Dutch and German lakes had δ13CDIC > -10‰ (group 1), and the Swedish and Finnish lakes δ13CDIC < -10‰ (group 2). These groupings corresponded to differences in catchment lithology, with lakes situated in hard water catchments containing calcareous bedrock (group 1) associated with δ13CDIC > -10‰.

The range in δ13Cdiatom values from -33.4 to -25.4‰ is lower than the range in δ13CDIC. Statistical comparison using a Mann–Whitney test (IBM SPSS) found the δ13Cdiatom values of each group of lakes differed significantly (U = 42.5, z = -2.85, P < 0.01, r = -0.52). Generally, lakes with calcareous
catchments (group 1) had more positive $\delta^{13}$C\textsubscript{diatom} (median: $-26.7\%_\text{e}$) compared with lakes in group 2 situated in non-calcareous catchments (median: $-27.8\%_\text{e}$). In addition a smaller range in values of 2.3\%e was present in group 1 compared with 7.3\%e in group 2.

As seen within the UK river data, a significant positive relationship was present between European lake $\delta^{13}$C\textsubscript{DIC} and $\delta^{13}$C\textsubscript{diatom} ($r_s = 0.59$, $P < 0.01$) (Fig. 6). As was found for $\delta^{13}$C\textsubscript{DIC}, two groupings of $\delta^{13}$C\textsubscript{diatom} values emerged, with the Swiss, Dutch and German data (group 1) forming a cluster of higher isotope values, and the Swedish and Finnish data points (group 2) spread along a linear gradient of lower isotope values. There was a strong relationship between $\delta^{13}$C\textsubscript{DIC} and $\delta^{13}$C\textsubscript{diatom} in group 2 ($r_s = 0.63$, $P < 0.01$) but not between the equivalent values for lakes in group 1 ($r_s = -0.25$, $P = 0.41$). No significant relationships were identified between $\delta^{13}$C\textsubscript{diatom} and either TP ($r_s = 0.13$, $P = 0.49$) or TN ($r_s = 0.23$, $P = 0.22$) concentrations.

### Table 1.
Spot sample water chemistry and $\delta^{13}$C\textsubscript{diatom} values determined for each of the 20 UK river sites in this study. Median, maximum, minimum, range and the standard deviation for each parameter are included.

| Catchment   | Stream/river | pH | EC (µS cm\(^{-1}\)) | TP (mg L\(^{-1}\)) | TDN (mg L\(^{-1}\)) | $\delta^{13}$C\textsubscript{DIC} (% VPDB) | $\delta^{13}$C\textsubscript{diatom} (% VPDB) |
|-------------|--------------|----|----------------------|---------------------|----------------------|------------------------------------------|------------------------------------------|
| Ribble/Wyre | Grizedale Brook | 6.8 | 186 | 0.013 | 0.90 | -11.0 | -27.8 |
|             | River Dunlop | 7.8 | 127 | 0.010 | 0.38 | -7.4 | -26.3 |
|             | River Wyre | 7.8 | 174 | 0.052 | 0.76 | -8.6 | -28.4 |
|             | Marshaw Wyre | 7.6 | 97 | 0.004 | 0.36 | -7.5 | -27.4 |
|             | Tarbrook Wyre | 7.8 | 77 | 0.009 | 0.65 | -7.0 | -27.8 |
| Lune South  | Keasden Beck | 7.9 | 197 | 0.013 | 0.39 | -5.9 | -27.3 |
|             | River Roeburn | 7.9 | 118 | 0.007 | 0.39 | -5.4 | -26.2 |
|             | River Hindburn | 8.0 | 161 | 0.011 | 0.59 | -6.5 | -28.2 |
|             | River Hindburn | 8.1 | 148 | 0.012 | 0.48 | -6.9 | -26.5 |
|             | River Wenning | 8.5 | 331 | 0.026 | 1.10 | -8.5 | -28.6 |
| Lune North  | River Rawthey | 7.5 | 177 | <0.005 | 0.51 | -8.0 | -27.3 |
|             | Crossdale Beck | 6.5 | 79 | 0.010 | 0.33 | -6.9 | -27.8 |
|             | Chapel Beck | 7.0 | 91 | <0.005 | 0.39 | -6.2 | -25.3 |
|             | River Lune | 7.8 | 227 | 0.006 | 0.57 | -7.9 | -27.4 |
|             | Trout Beck | 6.6 | 103 | <0.005 | 0.67 | -11.5 | -28.7 |
|             | River Rothay | 6.1 | 16 | 0.005 | 0.63 | -9.3 | -28.3 |
|             | Unnamed | 6.4 | 47 | <0.005 | 0.44 | -6.1 | -27.8 |
|             | Unnamed | 6.7 | 58 | <0.005 | 0.30 | -2.9 | -26.0 |
| Eden        | River Lowther | 7.7 | 200 | 0.006 | 0.61 | -8.5 | -28.4 |
|             | River Lowther | 8.2 | 192 | 0.007 | 0.59 | -8.0 | -28.9 |
| Median      | 7.7 | 138 | n/a | 0.54 | -7.5 | -27.8 |
| Max.        | 8.5 | 331 | 0.052 | 1.30 | -2.9 | -25.3 |
| Min.        | 6.1 | 16 | <0.005 | 0.30 | -11.5 | -28.9 |
| Range       | 2.3 | 315 | n/a | 0.80 | 8.5 | 3.6 |
| SD          | 0.7 | 74 | n/a | 0.20 | 1.9 | 1.0 |

Figure 4. Significant positive relationship between $\delta^{13}$C\textsubscript{DIC} and $\delta^{13}$C\textsubscript{diatom} values determined for the contemporary UK river data set ($r_s = 0.70$, $P < 0.01$). Relationship identified using Spearman’s rank correlation coefficient.

Figure 5. Significant negative relationships between TDN concentrations and $\delta^{13}$C\textsubscript{DIC} (open circles) ($r_s = -0.77$, $P < 0.01$) and $\delta^{13}$C\textsubscript{diatom} (shaded circles) ($r_s = -0.73$, $P < 0.01$) values determined for the contemporary UK river data set. Relationships identified using Spearman’s rank correlation coefficient.
Table 2. Spot sample water chemistry and δ13Cdiatom values determined for each of the 30 European lake sites in this study. Median, maximum, minimum, range and the standard deviation for each parameter are included. Data from Rinta et al. (2015).

| Country | Lake         | pH  | EC (μS cm⁻¹) | TP (mg L⁻¹) | TN (mg L⁻¹) | δ¹³Cdish (%) VPDB | δ¹³Cdiatom (% VPDB) |
|---------|--------------|-----|--------------|-------------|-------------|------------------|---------------------|
| Switzerland | Lauenensee   | 6.6 | 462          | 0.007       | 0.53        | −7.9             | −26.1              |
|          | Schwarzees   | 6.9 | 386          | 0.015       | 0.42        | −9.0             | −25.8              |
|          | Hinterburgsee | 8.9 | 137          | 0.011       | 0.80        | −6.0             | −27.5              |
|          | Gerzensee    | 7.0 | 289          | 0.016       | 0.68        | −4.4             | −27.7              |
|          | Rotsee       | 8.8 | 183          | 0.034       | 0.81        | −5.4             | −27.1              |
|          | Borgschicsee | 8.6 | 275          | 0.014       | 1.20        | −5.6             | −27.3              |
|          | Seelapsee    | 8.7 | 150          | 0.010       | 0.55        | −6.6             | −25.8              |
|          | Hasensee     | 7.7 | 329          | 0.036       | 1.20        | −5.1             | −27.5              |
|          | Hüttwilsee   | 8.5 | 316          | 0.015       | 1.10        | −4.3             | −26.7              |
|          | Nussbaumersee| 8.1 | 346          | 0.024       | 1.20        | −6.6             | −26.7              |
| Netherlands | De Waay      | 8.0 | 368          | 0.120       | 2.30        | −9.1             | −26.7              |
|          | Holzschees   | 8.2 | 328          | 0.036       | 0.86        | −5.5             | −26.5              |
|          | Schohsee     | 8.2 | 247          | 0.017       | 0.57        | −2.7             | −25.4              |
| Sweden   | Glimmingen   | 6.9 | 57           | 0.008       | 0.33        | −19.0            | −26.6              |
|          | Kisaajon     | 7.7 | 127          | 0.018       | 0.49        | −18.9            | −27.7              |
|          | Hargsjons    | 6.7 | 214          | 0.045       | 1.17        | −19.3            | −28.6              |
|          | Skottenesjons | 6.9 | 139         | 0.038       | 0.65        | −20.3            | −27.5              |
|          | Essjon       | 6.3 | 55           | 0.018       | 0.66        | −19.7            | −29.3              |
|          | Illersjon    | 7.6 | 311          | 0.025       | 0.48        | −19.6            | −27.8              |
|          | Marn         | 7.7 | 132          | 0.027       | 1.05        | −19.7            | −26.5              |
|          | Storafanesem | 5.7 | 78           | 0.009       | 0.46        | −19.3            | −27.7              |
|          | Lillsjong    | 7.7 | 41           | 0.018       | 0.61        | −20.8            | −31.1              |
|          | Skargolens   | 7.7 | 48           | 0.012       | 0.35        | −18.4            | −27.9              |
|          | Grissjon     | 6.5 | 27           | 0.011       | 0.39        | −18.9            | −29.9              |
| Finland  | Lovojarvi    | 7.1 | 123          | 0.027       | 0.84        | −13.4            | −26.5              |
|          | Syjunalunen  | 6.1 | 59           | 0.003       | 0.20        | −21.6            | −29.4              |
|          | Nimeton      | 5.6 | 68           | 0.010       | 0.48        | −23.6            | −33.4              |
|          | Meklojaivei  | 5.4 | 44           | 0.011       | 0.61        | −14.0            | −26.9              |
|          | Vakeka-Kotinen | 5.9 | 24           | 0.011       | 0.57        | −22.8            | −30.4              |
|          | Jyvajarvi    | 6.1 | 75           | 0.025       | 0.62        | −17.6            | −26.1              |
| Median   | 7.1          | 138 | 0.017        | 0.62        | −15.8        | −27.4            |                    |
| Max.     | 8.9          | 462 | 0.120        | 2.30        | −2.7         | −25.4            |                    |
| Min.     | 5.4          | 24  | 0.003        | 0.20        | −23.0        | −33.4            |                    |
| Range    | 3.5          | 439 | 0.117        | 2.10        | 20.9         | 8.0              |                    |
| SD       | 1.0          | 130 | 0.021        | 0.41        | 7.0          | 1.8              |                    |

Figure 6. Plot of contemporary European lake data set δ¹³Cdish and δ¹³Cdiatom values. A significant positive relationship was present across the whole data set (rₛ = 0.29, P < 0.01). When split and reanalysed no significant relationship was present between group 1 data (shaded symbols) (rₛ = −0.25, P = 0.41), but a stronger relationship was seen within group 2 (open symbols) (rₛ = 0.63, P < 0.01). Relationships identified using Spearman’s rank correlation coefficient.

Lake Tanganyika sediments

Diatom communities reconstructed from the sediments were dominated by planktonic taxa including Cyclotella, Aulacoceira and Stephanodiscus species. δ¹³Cdiatom varied from −30.0 to −22.4‰, equating to a range of 7.6‰ (Fig. 7a). In comparison, δ¹³Cbulk values were higher with a range of 7.3‰ from −28.2 to −20.9‰ (Fig. 7b). The offset between the two data sets varied from 0.4 to 4.2‰ with a median value of 2.7‰ (Fig. 7c). A significant positive relationship was present between δ¹³Cdiatom and δ¹³Cbulk (rₛ = 0.73, P < 0.01). At this coarse millennial scale, lowest values for both δ¹³Cdiatom and δ¹³Cbulk occurred between 14.8 and 5.5 ka, the period broadly recognized as the African Humid Period (deMenocal et al., 2000) (light shading in Fig. 7). Conversely, both isotope proxies had their maximum values in sediments dating to the end of the last glacial period and again in the late Holocene. Here also the smallest offset between the two records was measured. The corresponding trends in δ¹³Cdiatom and δ¹³Cbulk closely track that of higher plant leaf waxes (δ¹³Cwax) (Fig. 7d), a terrestrial vegetation change proxy extracted from Lake Tanganyika sediments by Tierney et al. (2010).

Discussion

Previous studies of freshwater environments have identified (i) δ¹³C of carbon sources, (ii) the relative contributions of these
sources and (iii) $^{13}$C enrichment by preferential $^{12}$C uptake by lake primary producers (including diatoms) as key variables determining the $\delta^{13}$CDIC available to diatoms for assimilation as $\delta^{13}$Cdiatom. Investigation into environmental controls of contemporary $\delta^{13}$Cdiatom over the different spatial and temporal scales reported here pinpoints the relative influence of carbon supply and demand factors, aiding the interpretation of palaeolimnological records as well as contemporary carbon cycling.

Translation of the carbon cycling history

Positive relationships identified between $\delta^{13}$CDIC from waters and contemporary $\delta^{13}$Cdiatom represent the primary control of catchment carbon source on the carbon isotopes in the diatom frustules. Principal carbon sources within a freshwater catchment have both geological and biotic origins and are associated with differing carbon isotopic ranges. Weathering of calcareous rock releases bicarbonate, which has a relatively high $\delta^{13}$C value of 0 to $+1\%e$ (Clark and Fritz, 1997). In contrast, CO$_2$ released into soils from plant root respiration and vegetation decay has a lower $\delta^{13}$C value, ranging from $-26$ to $-20\%e$ in C$_3$ and $-12$ to $-6\%e$ in C$_4$ landscapes, respectively (Mook et al., 1974; O’Leary, 1988). Oxidation of methane, associated with $\delta^{13}$C values between $-80\%e$ and $-50\%e$, provides a carbon source that is even further $^{13}$C depleted (Whiticar, 1999).

These catchment carbon signatures are transported via infiltrating flows to freshwater bodies where they mix with dissolved carbon from autochthonous sources, including macrophytes with recorded $\delta^{13}$C values of $-50$ to $-11\%e$ (Keeley and Sandquist, 1992) and phytoplankton with bulk values ranging between $-42$ and $-26\%e$ (Leng and Marshall, 2004). Oxidation of such materials during decomposition enables further release of $^{13}$C depleted CO$_2$, but if waters are stratified this process is slowed and the potential biotic carbon source is stored within anoxic sediments (Leng and Marshall, 2004).

The resulting $\delta^{13}$CDIC signature of a water body is further impacted by atmospheric exchange leading to the preferential loss of $^{12}$C. In addition, primary productivity (including diatoms) results in discriminatory uptake of $^{12}$C. The pH of a water body is also significant as it determines the proportioning of different DIC species; each of which has a contrasting carbon isotope signature. At pH 8 the percentage of DIC present as dissolved CO$_2$ is close to 0, and the hydration and disassociation of dissolved CO$_2$ into bicarbonate causes an increase in $\delta^{13}$C of approximately $9\%e$ (Clark and Fritz, 1997). $\delta^{13}$CDIC is therefore a record of catchment carbon cycling history reflecting sourcing and further changes to the isotope value related to fractionation during carbon phase and species changes. Final translation of the $\delta^{13}$CDIC signature to $\delta^{13}$Cdiatom is dependent on the photosynthetic pathway used, and any species-specific or vital effects that determine the degree of fractionation on uptake. The positive correlation identified between $\delta^{13}$CDIC and $\delta^{13}$Cdiatom in the UK rivers (Fig. 4) and European lakes (Fig. 6) suggests that even in sites of highly varying environmental characteristics, $\delta^{13}$CDIC is a significant control in the determination of $\delta^{13}$Cdiatom.

The role of catchment productivity

Within the contemporary UK rivers sampled, DIC probably had a biotic origin as no significant areas of calcareous geology were present in any of the catchments. The negative relationship identified between $\delta^{13}$CDIC and TDN concentrations (Fig. 5) supports this conclusion and indicated that river DIC pool characteristics were probably controlled by catchment productivity at this scale. This relationship showed that
dissolved biotic carbon supply was enhanced within more productive catchments and aquatic primary productivity did not lead to relative enrichment in $^{13}$C. This agrees with the findings of Maberly et al. (2013) who found a greater availability of DIC associated with more productive catchments in the English Lake District, increased loadings of dissolved biogenic carbon with a relatively low $^{13}$C signature were attributed to catchment land use, with greater availability of nutrients resulting in enhanced dissolved carbon release within the catchment.

The absence of a correlation between $^{13}$C$_{\text{diatom}}$ and species composition suggests sieving to control for species-specific and vital effects was successful, or at least limited any significant impact on diatom isotopic value. In addition $^{13}$C$_{\text{diatom}}$ values were consistently lower than $^{13}$C$_{\text{DIC}}$ and fell within a small range of 3.6‰. This suggests possible uptake of bicarbonate via CCMS has not had a pronounced effect on the resulting diatom carbon values. These findings show that $^{13}$C$_{\text{diatom}}$ can be used to investigate catchment productivity, highlighting the close coupling between a water body and its catchment conceptualized as the balance between carbon supply and demand. For specific catchments the strength of this relationship is dependent on the multiple and interacting controls on $^{13}$C$_{\text{DIC}}$ in lake waters and its translation to the diatoms.

The role of catchment geology

In comparison with the UK river epilithon, the European lake sediments represented a potentially more integrated temporal record of carbon cycling, with seasonal variability masked by sediment accumulation during several annual cycles. In addition, the much broader geographical range was clearly manifest in the heterogeneity of water chemistry variables and diatom assemblage compared with that of the rivers. Most striking was the identification of two groupings in $^{13}$C$_{\text{diatom}}$ values, which coincided with both $^{13}$C$_{\text{DIC}}$ and major geological differences in catchment carbon source characteristics.

At the continental scale of analysis, no relationships were identified between $^{13}$C$_{\text{diatom}}$ and productivity indicators TP or TN, either in the complete data set or within sub-groups. It is likely that highly varied catchment carbon processing attributed to the diverse climate, land use and geology represented obscured any record of pelagic carbon demand differences between the lakes. As seen with the UK rivers, the impact of species effects on $^{13}$C$_{\text{diatom}}$ appears to have been limited successfully through sieving to <20 µm. Instead, DIC characteristics appear to be the principal environmental controls of $^{13}$C$_{\text{diatom}}$ at this scale. Higher $^{13}$C$_{\text{diatom}}$ signatures (−27.7 to −25.4‰) in lakes with calcareous catchments (group 1) compared with those without (−33.4 to −26.1‰) (group 2) reflected the contribution of carbonate geology (associated with $^{13}$C between 0 and +1‰) to respective lake carbon pools. This demonstrated the significance of DIC sourcing to the production of an initial carbon signature, which is transferred to $^{13}$C$_{\text{diatom}}$. Also influential was the relative availability of DIC for uptake by diatoms. The absence of a relationship between $^{13}$C$_{\text{DIC}}$ and $^{13}$C$_{\text{diatom}}$ in group 1 is probably a result of high background levels of geologically sourced dissolved carbon. In addition, the influence of geological sourcing on $^{13}$C$_{\text{diatom}}$ in group 1 may have been compounded by likely enhanced uptake of bicarbonate via CCMS due to the near 0% contribution of CO$_2$ to DIC at pH values over 8. It is only in the absence of significant geological carbon sources (group 2) where the transfer of a dissolved biogenic carbon signature reflecting catchment productivity can be determined in $^{13}$C$_{\text{diatom}}$.

Interpretation of palaeoenvironmental records

Advancement of the findings by Maberly et al. (2013) concerning lake catchment productivity to include rivers has important implications for interpretation of palaeoenvironmental records. The close coupling between freshwater networks and catchment carbon cycling, and in particular the relative availability of dissolved biogenic carbon in response to land use, has been clearly demonstrated. Analysis of $^{13}$C$_{\text{diatom}}$ from highly varied lake sites demonstrated the difficulties associated with developing a universal model of catchment and water productivity relationships. Nevertheless, successful extraction of $^{13}$C$_{\text{diatom}}$ from lake sediments highlights the potential for obtaining palaeoenvironmental archives of changes in catchment carbon cycles from lake sediment cores. In particular, the fundamental principal of a carbon supply and demand balance can be applied to the interpretation of freshwater $^{13}$C$_{\text{diatom}}$ extracted from lakes situated in contrasting environmental and climatic settings.

The sediments from Lake Tanganyika provided an opportunity to test these conclusions on a lake with a well-established palaeoenvironmental history (e.g. Gasse et al., 1989; Scholz et al., 2003; Talbot et al., 2006) that would be expected to respond to changes in carbon cycling at the landscape scale. Within the Lake Tanganyika sediments there is a close coupling between $^{13}$C$_{\text{diatom}}$ and $^{13}$C$_{\text{bulk}}$ throughout the 34 000-year record with an offset no greater than 4.2‰ (Fig. 7). Diatom values were consistently lower than bulk carbon, suggesting diatoms were using the lighter isotope from dissolved carbon inputs. Using the conceptual relationships developed above, these isotope changes are thought to indicate that the lake carbon pool principally reflected changes in the quantity and nature of carbon supplied from the catchment with modifications by lake primary productivity as a secondary factor. This finding is attributed to the great size of Lake Tanganyika and its catchment where, particularly during wet periods, dissolved and particulate biogenic carbon produced in the catchment would have significantly contributed to the lake carbon pool.

Even during dry intervals of the last glacial period (Barker and Gasse, 2003) and the late Holocene (Haberyan and Hecky, 1987), signified by high $^{13}$C signatures in both $^{13}$C$_{\text{diatom}}$ (−24.7 to −22.4‰) and $^{13}$C$_{\text{bulk}}$ (−21.6 to −20.9‰), maintenance of a correlation and a constant offset indicates primary productivity did not significantly deplete the carbon pool. This is despite a probable decrease in carbon delivery from the catchment and potential enhancement of lake mixing processes leading to nutrient recycling (Scholz et al., 2003). In addition to variability in carbon loading, the coinciding measurement of high $^{13}$C signatures for diatoms and bulk sediments suggests a change in carbon source. Corresponding high $^{13}$C$_{\text{bulk}}$ values (−29.0 to −26.4‰) at this time (Fig. 7d) indicate increased prevalence of C$_4$-dominated savanna grassland within the Lake Tanganyika catchment (Tierney et al., 2010). Because of these ecosystem changes, greater contributions of dissolved carbon with higher $^{13}$C entered the lake during dry periods, and were thus translated into higher $^{13}$C$_{\text{diatom}}$ and $^{13}$C$_{\text{bulk}}$ values. The discovery of pervasive terrestrial supply domination over aquatic demand suggests large lakes are likely to have been substantial carbon sources to the atmosphere over centennial to millennial timescales. This contrasts with smaller lakes such as Lake Challa on the flank of Kilimanjaro where diatom and bulk carbon isotope records became periodically decoupled by enhanced in-lake productivity (Barker et al., 2013).

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The Lake Tanganyika study demonstrates that comparison of $^{13}C_{\text{diatom}}$ with $^{13}C_{\text{bulk}}$ extracted from sediment cores enables catchment and lake carbon cycles to be disentangled, overcoming inherent ambiguities in the interpretation of bulk $^{13}C$. If geological carbon sources of lakes can be assumed to be constant and modifications to the soil carbon pool from vegetation changes can be understood, individual site histories can be reconstructed. Lake sediment $^{13}C_{\text{diatom}}$ therefore represents a largely under-exploited resource with the potential to provide highly insightful carbon cycling chronologies over millennial timescales.

Conclusions

Stable isotopes analysis of diatom organic molecules, occluded in silica, constrains uncertainties associated with measurements of undifferentiated sedimentary carbon. The occluded organic matter also provides a carbon archive largely protected from degradation, oxidation and diagenesis. The application of the $^{13}C_{\text{diatom}}$ method in freshwaters requires adjustments to the standard inferences concerning pelagic productivity established by early marine studies. In freshwaters the controls of $^{13}C_{\text{diatom}}$ are more complex due to the high degree of connectivity between terrestrial vegetation, soils, bedrock and aquatic ecosystems. Within these environments carbon supply-side characteristics, including relative abundance of DIC from differing sources and associated $^{13}C_{\text{DIC}}$, are important controls that change as a function of catchment characteristics at various spatial scales.

The concept of inorganic carbon supply and demand offers a useful framework through which to develop the interpretation of $^{13}C_{\text{diatom}}$. Analysis of contemporary $^{13}C_{\text{diatom}}$ from UK river epilithon demonstrates the close linkages between carbon cycling in freshwater networks and their catchments. In support of findings by Maberly et al. (2013), more productive catchments are associated with greater availability of dissolved biogenic carbon. Investigation of contemporary European lake surface sediments confirms palaeoecological inferences concerning catchment control of lake carbon supply made by Barker et al. (2013) and Hernández et al. (2011, 2013). Consequently variations within a single site may be readily interpreted in terms of land use as the lithological template is held constant. It is presently not possible to produce a globally relevant quantitative relationship between $^{13}C_{\text{diatom}}$ and specific environmental variables. However, as demonstrated by the Lake Tanganyika study, great potential lies in the use of $^{13}C_{\text{diatom}}$ to inform interpretation of lake sediment records. Palaeoenvironmental interpretation could be further enhanced by modelling the transfer of carbon through specific catchments. Of particular significance is improved understanding of dissolved carbon cycling from diatom frustules, independent of particulate carbon compositional changes normally associated with lake sediment $^{13}C_{\text{bulk}}$ analysis, to evaluate changes in freshwater ecosystems and palaeoenvironments.

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Abbreviations. CCM, carbon concentrating mechanism; DIC, dissolved inorganic carbon; EC, electrical conductivity; TDN, total dissolved nitrogen; TN, total nitrogen; TP, total phosphorus.

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