SHORT COMMUNICATION

Peripheral blood lymphocyte number and phenotype prior to therapy correlate with response in subcutaneously applied rIL-2 therapy of renal cell carcinoma

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Summary The phenotype of peripheral blood lymphocytes of 27 renal cell carcinoma patients before and at the end of subcutaneously given rIL-2 therapy was determined by two colour flow cytometry. Therapy induced changes in peripheral blood leucocyte composition and phenotypes were comparable to those reported for intravenously given rIL-2. The present paper shows a correlation between the 'activation status' of the patient before therapy and eventual response.

The therapeutic effects of recombinant interleukin-2 (rIL-2) therapy look promising in the case of renal cell carcinoma (RCC) and melanoma (Rosenberg et al., 1987; West et al., 1987).

Most of the interleukin-2 therapies so far, involved the intravenous administration of high dose rIL-2, causing severe side effects. In our clinic, rIL-2 was administered subcutaneously (s.c.) which avoided severe side effects, whereas clinical results were comparable to the ones involving intravenous (i.v.) administration of rIL-2 (Sleijfer et al., 1990; 1992). The immunomodulatory effects of subcutaneously administered rIL-2 still remain to be investigated.

The present paper gives data on the 'activation status' of the main peripheral blood lymphocyte subpopulations (helper T cells, cytotoxic T cells and NK cells) in 27 RCC patients by determining the expression of the activation marker HLA-DR on these lymphocytes before and after s.c. rIL-2 therapy (day 35 and day 40).

Materials and methods

Patients and therapeutic protocol

All patients (27; 16 male and 11 female) had evaluable metastatic renal cell carcinoma (RCC). Their average age was 59 (range 41–74). Patients participated in a phase II study of treatment with subcutaneous rIL-2 (Sleijfer et al., 1990; 1992), when informed consent was obtained. Patients received a 3-day cycle of Cetus rIL-2 (EuroCetus, Amsterdam, The Netherlands) every week for 6 consecutive weeks. During the first 5-day cycle 18 million IU rIL-2 were given once daily; in the following cycles the dose in the first 2 days was reduced to 9 million units. Treatment results were: two complete remissions (CR), four partial remissions (PR), seven patients had progressive disease (PD), and 14 patients showed stable disease (SD). A complete response required disappearance of all evidence of tumour for a minimum of 4 weeks, a partial response was registered when a 50% or greater decrease in the sum of the products of all diameters of evaluable lesions was reached; patients with a response less than partial or an increase of less than 25% were classified as stable disease. Progression was defined as an increase of more than 25% or the development of new lesions.

Monoclonal antibodies

All monoclonal antibodies (Becton Dickinson, Mountain View, CA, USA) were directly conjugated with phycoerythrin (PE) or fluorescein-isothiocyanate (FITC). Monoclonal antibodies (mAb) used were aleu4 (CD3), aleu3 (CD4), aleu2 (CD8), aleu19 (CD56), and aHLA-Dr.

Immunostaining of cells and flow cytometry

Peripheral blood lymphocytes of RCC patients were analysed before (day 0) and at the end of rIL-2 therapy (days 35 and 40). One hundred µl of EDTA blood was resuspended in 20 µl of mAb preparation (containing 10 µl of each mAb) and incubated at room temperature (RT) for 15 min. Two ml of FACS lysing solution (Becton Dickinson) was added and the cells were incubated for an additional 10 min. Subsequently, the solution was centrifuged at 1000 g for 2 min and the cells were washed in 2 ml of phosphate buffered saline supplemented with 15 U ml⁻¹ heparin (PBS/heparin) and resuspended in 150 µl of the same solution. The samples were immediately analysed on a FACStar (Becton Dickinson) with the laser tuned at 488 nm. Lymphocytes were gated using standard FSC/SSC settings.

Statistics

Statistical significant differences were determined using the Wilcoxon test or the distribution free sign test as indicated. P values of ≤ 0.05 were considered significant.

Results

Hematological changes during sc rIL-2 treatment

In all patients, in each cycle, during rIL-2 administration the absolute number of lymphocytes decreased, whereas during the 2 days without rIL-2 a rapid and large increase of the absolute number of lymphocytes was found. During therapy all patients developed highly elevated numbers of eosinophils (from 0.28*10⁶ ± 0.19*10⁶ per ml on day 0 to a peak of 7.9*10⁶ ± 4.0*10⁶ per ml on day 19).

Changes in peripheral blood lymphocyte composition

The changes in peripheral blood composition of 20 patients was determined. Ten patients were analysed on day 0 and day 40, seven on day 0 and day 35, and three on day 0, 35

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Received 24 February 1992; and in revised form 22 June 1992.
and 40. These patients were ordered in two groups. Group I consisted of 13 patients analysed on day 0 and day 40 (lymphopenic phase) and group II consisted of 10 patients analysed on day 0 and day 35 (rebound phase preceding the last rIL-2 cycle). Table I shows that, at the end of therapy, both the relative and absolute amounts of CD56+ cells had increased significantly, both on day 35 ($P < 0.01$, sign test) as well as on day 40 ($P < 0.01$) when compared with day 0 within each group. In contrast, the relative amount of CD3+ cells had decreased significantly ($P < 0.01$) in both groups when compared to day 0. This decrease was due to decreases in both the relative amounts of CD8bright+ cells and CD4+ cells. Still, the absolute numbers of CD3+ cells had increased significantly ($P < 0.05$) from day 0 to day 35 (predominantly due to an increase of the CD4+ cells, Table I), while there was no significant change in the absolute amount of CD3+ cells on day 40 compared to day 0.

'Activation status' of the subpopulations

Table II shows that the absolute numbers of various subpopulations expressing HLA-Dr had increased during rIL-2 therapy. Table II also shows that the activation marker HLA-Dr became predominantly expressed on CD56+ cells. Both on day 35 and day 40 the relative and absolute numbers of CD56+ HLA-Dr+ cells had increased significantly compared to day 0 within each group.

Lymphocyte 'activation status' and response to therapy

Two out of the 27 patients showed complete remission and four showed a partial remission. Seven patients had progression of disease, whereas 14 were qualified as stable. To determine a possible correlation between clinical outcome and immunological changes, patients were grouped according to these three categories (responders, progressive and stable) and immunological data were compared.

No correlation was found when data obtained at the end of the therapy were compared with each other. However, the six patients with remission showed significantly higher absolute and relative numbers of lymphocytes just before the first rIL-2 administration (Table III) than the patients with no remission ($P < 0.01$; Wilcoxon test). In addition, they had significantly higher absolute amounts of CD8bright+ HLA-Dr+ ($P < 0.01$), CD4+ HLA-Dr+ ($P = 0.01$), and CD56+ HLA-Dr+ ($P < 0.02$) cells than the patients who did not respond (SD, PD, see Table III).

**Discussion**

Many groups have examined the immunomodulatory effects in RCC patients and melanoma patients (Lotze et al., 1987; Ellis et al., 1988; Weil-Hillman et al., 1989; Urbá et al., 1990; Favrot et al., 1990) receiving rIL-2 intravenously. In the present paper the immunomodulatory effects of rIL-2 therapy in renal cell carcinoma patients receiving rIL-2 subcutaneously were studied.

Each time a new cycle of treatment was started, the absolute lymphocyte count decreased rapidly, whereas a quick increase (rebound lymphocytosis) was found when administration of rIL-2 was stopped between day 5–7 of each cycle. All patients showed an increase in the absolute eosinophil count in the peripheral blood. These results, obtained with subcutaneously applied rIL-2, extend the findings of others (Sondel et al., 1988) monitoring patients treated intravenously with rIL-2. So, the results might be taken as an indication that subcutaneously given rIL-2, despite its considerably lower toxicity as compared to intravenously given rIL-2, has an effect on the immune system comparable to i.v. given rIL-2.

The NK population showed the highest increase in relative and absolute numbers. This is in agreement with the results reported for intravenously administered rIL-2 (Weil-Hillman et al., 1989; Ellis et al., 1988), although the increase in the relative and absolute numbers of the NK population during s.c. administration was less pronounced. In addition, the relative number and absolute number of NK cells (CD56+) expressing HLA-Dr had increased significantly.

Until now, no good correlation between phenotypical changes in the peripheral blood and response to rIL-2 therapy has been found (Favrot et al., 1990). One study concerning i.v. administered rIL-2, however, showed a positive correlation between remission and lymphocytosis (West, 1989).

The study presented here extends the number of possible parameters correlated with response. It appears that high numbers of lymphocytes and high numbers of cells expressing the activation marker HLA-Dr prior to therapy are a prognostically favorable parameter (Table III). Interestingly, the two patients with complete remissions showed considerably higher numbers of CD8bright and CD56 positive cells expressing HLA-Dr. These high numbers of activated CD8bright cells might reflect a more immunogenic tumour in these patients. Related to the study presented here shows that subcutaneously administered rIL-2 induces immunological changes comparable to intravenously administered.

### Table I

| Day of treatment | CD3+ | CD4+ | CD8bright+ | CD56+CD2+ |
|------------------|------|------|------------|------------|
|                  | %    | × 10^6 ml⁻¹ | %    | × 10^6 ml⁻¹ | %    | × 10^6 ml⁻¹ | %    | × 10^6 ml⁻¹ |
| Group I (n = 13) |      |           |      |           |      |           |      |           |
| 0                | 71 ± 9 | 906 ± 359 | 48 ± 11 | 605 ± 295 | 24 ± 10 | 291 ± 159 | 13 ± 7 | 163 ± 105 |
| 40               | 47 ± 11 | 831 ± 362 | 34 ± 13 | 504 ± 248 | 14 ± 5 | 218 ± 135 | 38 ± 12 | 578 ± 238 |
| Group II (n = 10)|      |           |      |           |      |           |      |           |
| 0                | 70 ± 8 | 974 ± 561 | 43 ± 10 | 606 ± 408 | 29 ± 9 | 392 ± 236 | 12 ± 4 | 168 ± 111 |
| 35               | 54 ± 14 | 2351 ± 1999 | 39 ± 13 | 1696 ± 1422 | 17 ± 8 | 714 ± 638 | 36 ± 13 | 1628 ± 1327 |

*Significant increase compared to day 0, $P < 0.05$, $^b P < 0.02$, $^c P < 0.01$. $^d$Significant decrease compared to day 0, $P < 0.05$, $^e P < 0.02$, $^f P < 0.01$. Statistical significance determined by distribution free sign test.

### Table II

| Day of treatment | CD4+ HLA-Dr+ | CD8bright+ HLA-Dr+ | CD56+ HLA-Dr+ |
|------------------|--------------|-------------------|--------------|
|                  | % × 10^6 ml⁻¹ | % × 10^6 ml⁻¹ | % × 10^6 ml⁻¹ |
| Group I (n = 13) |      |           |      |           |
| 0                | 4 ± 2 | 50 ± 36 | 2 ± 2 | 34 ± 22 |
| 35               | 6 ± 4 | 95 ± 72 | 3 ± 2 | 45 ± 39 |
| Group II (n = 10)|      |           |      |           |
| 0                | 4 ± 2 | 61 ± 51 | 3 ± 4 | 65 ± 98 |
| 35               | 8 ± 5 | 316 ± 204 | 4 ± 3 | 206 ± 183 |

**Table** Changes in the activation status of various subpopulations. For statistical significance, see Table II.
rIL-2. We have already shown that s.c., in contrast to i.v., administered rIL-2, induces no severe side effects, whereas clinical responses are comparable (Sleijfer et al., 1990; 1992). Most importantly, this report shows that the 'activation status' of the patient prior to therapy is related to the outcome of therapy.

Table III Absolute and relative number of lymphocytes and lymphocyte subpopulations expressing HLA-Dr as determined on day 0

| Patient | Absolute number of lymphocytes | Relative number of lymphocytes | CD8bright* | CD4* | CD56* |
|---------|--------------------------------|--------------------------------|------------|------|-------|
| 1 CR    | 2.8a                           | 29b                            | 154c       | 113d | 84e   |
| 2 CR    | 2.1                            | 24                            | 248        | 144  | 165   |
| 3 PR    | 1.7                            | 35                            | 51         | 85   | 17    |
| 4 PR    | 1.8                            | 33                            | 95         | 73   | 52    |
| 5 PR    | 1.3                            | 21                            | 52         | 52   | 22    |
| 6 PR    | 1.3                            | 30                            | ND         | ND   | ND    |
| Average | 1.8 ± 0.6^6                   | 28.7 ± 5.3^6                   | 120.0 ± 83.0^6 | 93.4 ± 31.9^6 | 68.8 ± 59.8^6 |
| 7 PD    | 0.6                            | 14                            | 24         | 26   | 13    |
| 8 PD    | 0.5                            | 7                             | 10         | 25   | 28    |
| 9 PD    | 1.4                            | 31                            | 42         | 28   | 14    |
| 10 PD   | 1.3                            | 10                            | 13         | 26   | 13    |
| 11 PD   | 1.0                            | 13                            | ND         | ND   | ND    |
| 12 PD   | 1.1                            | 10                            | 44         | 22   | 13    |
| 13 PD   | 2.1                            | 22                            | ND         | ND   | ND    |
| Average | 1.1 ± 0.5                     | 15.3 ± 8.4                    | 26.6 ± 15.9 | 25.4 ± 2.2 | 20.2 ± 9.6 |
| 14 SD   | 1.1                            | 21                            | 88         | 33   | 33    |
| 15 SD   | 1.4                            | 27                            | 14         | 14   | 14    |
| 16 SD   | 1.4                            | 18                            | 70         | 126  | 14    |
| 17 SD   | 1.2                            | 16                            | ND         | ND   | ND    |
| 18 SD   | 0.6                            | 7                             | 18         | 24   | 30    |
| 19 SD   | 1.2                            | 21                            | 36         | 36   | 12    |
| 20 SD   | 1.2                            | 12                            | 24         | 60   | 12    |
| 21 SD   | 0.9                            | 12                            | 18         | 27   | 18    |
| 22 SD   | 1.4                            | 17                            | ND         | ND   | ND    |
| 23 SD   | 0.7                            | 7                             | ND         | ND   | ND    |
| 24 SD   | 0.9                            | 18                            | 54         | 54   | 18    |
| 25 SD   | 0.5                            | 7                             | 15         | 15   | 10    |
| 26 SD   | 2.4                            | 29                            | 48         | 96   | 48    |
| 27 SD   | 0.5                            | 20                            | 5          | 20   | 10    |
| Average | 1.1 ± 0.5                     | 15.4 ± 7.7                    | 35.5 ± 26.5 | 45.9 ± 36.0 | 19.9 ± 11.0 |

*Millions ml⁻¹; ^aPercentage of all leucocytes; ^bThousands ml⁻¹. Average numbers were calculated for each group of patients. ^cSignificantly higher than non-responders (P < 0.01); ^dSignificantly higher than non-responders (P < 0.02). Statistical significance was calculated using the Wilcoxon test (two-sided).

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