The effect of concurrent elevation in CO$_2$ and temperature on the growth, photosynthesis, and yield of potato crops

Yun-Ho Lee, Wan-Gyu Sang, Jae-Kyeong Baek, Jun-Hwan Kim, Pyeong Shin, Myung-Chul Seo*, Jung-Il Cho*  

Crop Production and Physiology Division, National Institute of Crop Science, Rural Development Administration, Wanju, Jeollabuk-do, Republic of Korea  
* jungilcho@korea.kr (JIC); mcseo@korea.kr (MCS)

Abstract

Global climate change accompanied by continuous increases in atmospheric carbon dioxide (CO$_2$) concentration and temperature affects the growth and yield of important crops. The present study investigated the effect of elevated temperature and CO$_2$ concentrations on the growth, yield, and photosynthesis of potato (Solanum tuberosum L. cv. Superior) crops using Korean Soil-Plant-Atmosphere-Research chambers that allow the regulation of temperature and CO$_2$ concentration under daylight conditions. Based on the average temperature from 1991 to 2010 in the Jeonju area, South Korea, potato plants were exposed to four different conditions: ambient weather (400 μmol mol$^{-1}$, aCaT), elevated temperature (+4°C, aCeT), elevated CO$_2$ concentration (800 μmol mol$^{-1}$, eCaT), and concurrently elevated CO$_2$ concentration and temperature (eCeT). Under aCeT conditions, the temperature exceeded the optimal growth temperature range towards the late growth phase that decreased stomatal conductance and canopy net photosynthetic rate and subsequently reduced biomass and tuber yield. Stomatal conductance and chlorophyll concentration were lower under eCaT conditions than under aCaT conditions, whereas late-growth phase biomass and tuber yield were greater. Compared to other conditions, eCeT yielded a distinct increase in growth and development and canopy net photosynthetic rate during tuber initiation and bulking. Consequently, biomass and canopy net photosynthesis increased, and tuber yield increased by 20.3%, which could be attributed to the increased tuber size, rather than increased tuber number. Elevated CO$_2$ reduced chlorophyll, magnesium, and phosphorus concentrations; reducing nitrogen concentration (by approximately 39.7%) increased the C:N ratio. The data indicate that future climate conditions will likely change nutrient concentration and quality of crops. The present study shows that while elevated temperature may negatively influence the growth and yield of potato crops, especially towards the late-growth phase, the concurrent and appropriate elevation of CO$_2$ and temperature could promote balanced development of source and sink organs and positively effect potato productivity and quality.
Introduction

Global climate change has accelerated since the beginning of the 21st century. For example, the atmospheric CO$_2$ concentration in 1880 was 280 μmol mol$^{-1}$ but it has increased continuously since the industrial revolution and is currently at least 400 μmol mol$^{-1}$. The average annual temperature has increased by 0.8˚C since 1880 [1]. Based on the Representative Concentration Pathway (RCP) Scenario 8.5 of the Intergovernmental Panel on Climate Change [2], atmospheric temperature could rise by as much as 4.8˚C by around 2100 and the CO$_2$ concentration could reach 940 μmol mol$^{-1}$. These predicted increases in atmospheric CO$_2$ concentration and temperature are anticipated to affect crop productivity and quality [3–5]. However, each crop variety exhibits different response patterns to changes in temperature and CO$_2$ concentration [6–8].

Potato (*Solanum tuberosum* L.) is one of the five major global food crops along with rice, wheat, corn, and soybean [9]. It is important to analyze the effect of climate change on the production of potato plants to secure the future of the global food supply. Modern potato cultivars generally grow well under moderate temperatures around 20˚C, and the optimum temperature for the growth of above-ground parts, such as leaves and stems, and the initiation and bulking of the underground tubers are known to be 20–25˚C and 15–20˚C, respectively [10–13]. The rise in temperature affects almost all biological processes of potato plants including photosynthesis, respiration, and enzyme activities [14, 15]. Long-term exposure to high temperatures has been reported to increase photorespiration rather than photosynthetic rate [16] and has been reported to reduce photosystem II (PS II) activity, chlorophyll concentration, and enzyme activities [10, 17–19]. Multiple studies have reported that the rise in temperature above the optimum level decreased the leaf area and biomass, delayed tuber initiation and decreased the tuber yield through the inhibition of carbon assimilation and its subsequent translocation to tuber [20–24].

In contrast, the increase in atmospheric CO$_2$ concentration is generally known to exert a positive influence on crop photosynthesis, growth, and yield [8, 25–29]; potato has been reported to exhibit high yields under high CO$_2$ concentrations. The increased yield can be attributed to the increase in the rate of foliar photosynthesis and the facilitated allocation of assimilates to tubers [30]. Recent studies have reported that if CO$_2$ concentration increases by two-fold, the biomass and tuber yield of potato plants could increase by as much as 49% [30–33]; some studies have reported that increases in atmospheric CO$_2$ levels could also improve water and nitrogen use efficiency [34–36].

Studies have also reported that elevated CO$_2$ levels could lead to a decrease in stomatal conductance and chlorophyll concentrations [31, 37–42], as well as to changes in essential mineral concentrations and carbon to nitrogen (C:N) ratios in plants [43–47]. Upon exposure to elevated CO$_2$ concentrations for a long period, the carboxylase activity of ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) decreases while the C:N ratio increases due to the high carbon status [36, 48]. Such an increase in the C:N ratio with lower nitrogen levels leads to lower protein content, and it eventually deteriorates the quality of the crop.

Carbon dioxide and temperature are important factors in the growth and development of potato crops. The recent global climate changes have affected significantly the growth, yield, and photosynthesis of the crop. These two factors together influence the growth and photosynthesis of the crop, not independent of each other [49]. It is important to investigate how crops are affected by elevated temperature and CO$_2$ concentration, separately and in combination. Most studies on the effect of elevated temperature or CO$_2$ concentrations have been conducted using closed-top fields (CTFs), open-top chambers (OTCs), or free-air carbon dioxide enrichment (FACE) [26, 47, 50–52] and have generally neglected to investigate the effect of
concurrent elevations in CO\textsubscript{2} concentration and temperature. The SPAR facility is known to be optimized equipment to analyze the growth and development of crops at the canopy-level under precisely controlled environmental conditions that can control temperature and CO\textsubscript{2} concentration under daylight conditions [53]. Using the Korean Soil-Plant-Atmosphere-Research (KSPAR) chambers, the present study aimed to investigate the effect of long-term exposure to elevated CO\textsubscript{2} concentration and temperature on potato plants, with the goals of identifying interactions between i) growth, development, and yield; ii) photosynthetic responses; and iii) the concurrent elevation of CO\textsubscript{2} concentration and temperature.

Materials and methods

Experimental site and Korean Soil-Plant-Atmosphere-Research (KSPAR) chamber

The present study was conducted at the National Institute of Crop Science, Wanju, Jeollabuk-do, Korea (35˚84’ 34" N, 127˚04’ 84" E). The Korean Soil-Plant-Atmosphere-Research (KSPAR) chamber was a slightly modified version of the SPAR chamber of the USA Agricultural Research Service (ARS) [53]. The most significant feature of the equipment is the double chamber composed of inner and outer chambers and designed to reduce CO\textsubscript{2} leakage and temperature rise due to radiant-heat generation (Fig 1).

The CO\textsubscript{2} exchange rate can be measured without the use of N\textsubscript{2}O gas [54]. The inner chamber consists of a structure covered by 10 mm thick Plexiglas (Evonik Industries, Essen, Germany) measuring 2.00 m $\times$ 0.95 m $\times$ 2.25 m (length $\times$ width $\times$ height). The lower part of the inner chamber contains a steel soil bin measuring 2.00 m $\times$ 0.78 m $\times$ 1.42 m (length $\times$ width $\times$ height). The upper part of the outer chamber is a 4 mm thick clear laminated glass measuring 2.30 m $\times$ 2.64 m $\times$ 2.50 m (length $\times$ width $\times$ height) with a light transmissivity of approximately 93–95%.

The inner chamber includes an air-temperature and relative-moisture sensor (TRH-300, Rixen Technology, Taipei, Taiwan) that collects data at 30-sec intervals. The soil bin has a soil temperature and moisture sensor (Drill & Drop, Sentkek Technologies, Stepney, Australia) that collects data at 1-h intervals (CEM20, Thermo Fisher Scientific Inc., Scoresby, Australia).

![Fig 1. Korean Soil-Plant-Atmosphere-Research (KSPAR) chamber. (A) Schematic diagram, (B) Experimental site in Wanju-gun, Jeollabuk-do, Korea.](https://doi.org/10.1371/journal.pone.0241081.g001)
Photosynthetically active radiation was measured using a quantum sensor (SQ-326, Apogee Instruments Inc., UT, USA) at 10-min intervals.

The CO₂ supply to the SPAR chamber was maintained at a steady level using a gas-pressure sensor (Model 200 IM, Seowon, Yongin, Korea) and a flowmeter. All chambers were equipped with two non-dispersive infrared (NDIR) CO₂ analyzers (LI-820, LI-COR Biosciences Inc., NE, USA) to monitor the concentration of CO₂ in the air flowing in and out of the inner chamber. Before the experiment, all CO₂ analyzers were calibrated for the zero and span values, according to the instructions given in the LI-820 manual.

**Experimental design**

Before emergence, the day and night air temperatures were maintained at 20 and 15°C, respectively, and the CO₂ concentration was maintained at 400 μmol mol⁻¹. Starting temperature and CO₂ concentration was based on the ambient climate conditions or the projected 2100 climate (RCP Scenario 8.5), depending on the treatment [2].

During the experiment, the CO₂ concentration was maintained at either 400 or 800 μmol mol⁻¹; the temperature was maintained at either the daily mean temperature of the Jeonju area (1991–2010) or 4°C warmer. The treatments involved the following conditions: ambient CO₂ and ambient temperature (aCaT), ambient CO₂ and elevated temperature (aCeT), elevated CO₂ and ambient temperature (eCaT), and concurrently elevated CO₂ and temperature (eCeT). All treatment conditions were arranged in completely randomized design [28].

During the day (05:00–19:00), the CO₂ concentrations for the two ambient and two elevated CO₂ conditions were maintained at 400 μmol mol⁻¹ and 800 μmol mol⁻¹, respectively. The CO₂ concentration during the night (19:00–05:00) was left uncontrolled and fluctuated within the 504–650 μmol mol⁻¹ range owing to the plants’ respiration that releases CO₂ (Table 1).

The temperature setting of the chamber was adjusted by the Unit of the week (once every week) based on the mean temperature data of the past 20 years (1991–2010) in Jeonju area and the daily air temperature was adjusted to follow a diurnal curve pattern similar to the natural environment; the error range among the chambers was ±0.3°C (Table 1 and S1 Fig). The daily mean solar radiation during the period of cultivation was 9.97 MJ m⁻² (S2 Fig).

Table 1. Treatment details of experiments conducted in the Korean Soil-Plant-Atmosphere-Research (KSPAR) chamber.

| Treatment | aCaT | eCaT | aCeT | eCeT |
|-----------|------|------|------|------|
| Daily mean, day | 20.9 ± 0.18 | 20.9 ± 0.18 | 24.9 ± 0.18 | 24.9 ± 0.18 |
| Daily mean, night | 14.6 ± 0.15 | 14.6 ± 0.15 | 18.6 ± 0.15 | 18.6 ± 0.15 |
| CO₂ (μmol mol⁻¹) concentration | | | | |
| Daily mean (day/night) | 417/504 | 771/508 | 425/518 | 804/650 |
| Soil temperature (°C) | | | | |
| Daily mean | 16.4 ± 0.28 | 17.3 ± 0.30 | 19.4 ± 0.35 | 18.7 ± 0.35 |
| Soil water content (% soil volume) | | | | |
| Daily mean | 36.9 ± 0.31 | 33.1 ± 0.44 | 33.2 ± 0.44 | 33.9 ± 0.32 |

aCaT: Ambient 400 μmol mol⁻¹ CO₂, ambient average temperature, eCaT: Elevated 800 μmol mol⁻¹ CO₂, ambient average temperature, aCeT: Ambient 400 μmol mol⁻¹ CO₂, ambient average-temperature increases of 4°C, eCeT: Elevated 800 μmol mol⁻¹ CO₂, ambient average-temperature increases of 4°C. Values represent means ± SE.
Plant materials and growth conditions

The potato (Solanum tuberosum L. cv. Superior) cultivar is widely cultivated in Korea. Seed potatoes of this cultivar were obtained from the Highland Agriculture Research Institute (Pyeongchang, Korea) and stored at 10–12˚C.

The quantity of chemical fertilizer was based on the topdressing by N-P\textsubscript{2}O\textsubscript{5}-K\textsubscript{2}O (54-30-42 g/chamber) before seeding. The soil composition in the soil bin was sandy loam (72.8% sand, 22.9% silt, and 4.3% clay). On March 23, 2018, 30 sprouted tubers were planted into each chamber with the row space of 20 cm and the plant space of 15 cm, at ~10 cm depth, and the soil moisture content was maintained at $\geq 30\%$ (v/v) using the micro-tube-pipe of pressure-compensated drippers for the watering at 08:00 and 17:00.

Plant measurements

Plant height, number of branches, and biomass were measured for six plants from each chamber at 35 days after emergence (DAE), a mid-phase in tuber initiation, and at 58 DAE, during tuber bulking. At the same time, the leaf area was measured using a leaf-area meter (LI-3100C, LI-COR Biosciences Inc., NE, USA), and the leaf, stem, and tuber biomass was measured after drying for 4 d at 75˚C.

On June 18, 2018, the remaining potatoes were harvested and divided into leaf, stem, and tuber. The potatoes were classified into size groups ($< 30$ g, 30–80 g, and $> 80$ g), and a marketable yield was calculated based on the $> 80$ g class. The plant materials were oven-dried for 4 d at 75˚C to obtain the biomass of each plant part.

Stomatal conductance and chlorophyll concentration

The stomatal conductance and chlorophyll concentration of five potato leaves from each chamber was measured at 34, 43, 50, 56, and 64 DAE using a portable leaf porometer (SC-1, Mater Group Inc., USA) and a chlorophyll-content meter (CCM-300, Opti-Sciences Inc., Hudson, USA), respectively, following the manufacturer’s guidelines.

Organic carbon, nitrogen, and mineral nutrients

To measure the concentration of carbon, nitrogen, phosphorus, potassium, and magnesium, potato leaves were collected from three potato plants of each chamber at 35, 43, 50, 58, and 64 DAE, dried at 75˚C for 4 d, finely ground using a stainless steel grinder, and then sieved through 2-mm mesh.

Carbon and nitrogen were measured using the Dumass method, analyzing 0.2 g of each ground-leaf sample using an elemental analyzer (Elementar, vario MAX cube, GmbH, Germany) [55]. Phosphorus, potassium, and magnesium were measured by decomposing 0.5 g ground-leaf sample in 5 ml H\textsubscript{2}SO\textsubscript{4} and 20 ml H\textsubscript{2}O\textsubscript{2} for 4 h at 220˚C, diluting the resulting solution to 50 ml with distilled water, and then analyzing 3 ml using inductively coupled plasma-mass spectrophotometry (ICP Integra XL, GBC Scientific Equipment Pty Ltd., Braeside, Australia) [56].

CO\textsubscript{2} gas exchange

The KSPAR chamber is a semi-closed chamber that is equipped with two CO\textsubscript{2} analyzers and fans. Therefore, the CO\textsubscript{2} exchange rate (CER; mol CO\textsubscript{2} m\textsuperscript{-2} s\textsuperscript{-1}) was estimated using the concentrations of CO\textsubscript{2} flowing in and out of the chamber and the following equation:

$$CER = V \times P \times (\Delta CO_{2}) / (R \times T)$$

(1)
where $V$ is wind velocity (m s$^{-1}$), $P$ is the absolute pressure, $\Delta CO_2$ is the difference in CO$_2$ concentration between the inlet and outlet (ppm), $R$ is the absolute atmospheric pressure (L·atm·K$^{-1}$·mol$^{-1}$), and $T$ is the absolute temperature (273.15 K).

The net photosynthetic rate ($\mu$mol CO$_2$ m$^{-2}$ s$^{-1}$) for the CO$_2$ exchange rate was determined during the day (05:00–19:00) when CO$_2$ was supplied. The night respiration rate ($\mu$mol CO$_2$ m$^{-2}$ s$^{-1}$) was determined from 19:00–05:00. The CO$_2$ exchange rates of all the chambers were collected at 30-sec intervals, and the measured data were stored as the mean of the values measured during 5 min.

The accumulation of carbon in the plant body was calculated as follows:

$$A_T = \left( \sum_{05:00}^{19:00} P_N \times 12 \right)$$

where $A_T$ is the overall accumulated C in the plant body (g C plant$^{-1}$), $P_N$ (mol CO$_2$ m$^{-2}$ s$^{-1}$) is the mean per plant body for 5 min calculated by Eq (1), and 12 is the molecular weight of carbon. The actual dark respiration rate was not used.

Statistical analysis

The data were evaluated by analysis of variance (ANOVA) performed using statistical analysis systems 9.2. The mean values of plant traits in response to applied treatments were compared using Tukey’s tests at $P < 0.05$.

Results

Plant growth and biomass dynamics

To examine the effect of the CO$_2$ concentration and temperature elevation on the growth of potato plants, the plant height, number of branches, and leaf area measured on 35 and 58 DAE were analyzed (Fig 2 and S1 Table).

At 35 DAE, plant height was significantly higher under the aCeT and eCeT conditions compared to aCaT and eCaT conditions ($P < 0.001$), whereas neither the number of branches nor leaf area was significantly affected. At 58 DAE, neither plant height nor the number of branches was significantly affected, whereas the leaf area was drastically reduced under the aCeT conditions.

To examine the growth characteristics of each organ relative to the potato leaf, stem, and tuber, the total biomass and the percent allocations of biomass were determined (Fig 3 and S1 Table).

At 35 DAE, total biomass was significantly higher under the aCeT and eCeT conditions than under the aCaT and eCaT conditions ($P < 0.001$). The proportion of biomass allocation to leaves and stems was lower under eCaT and eCeT conditions than under the other conditions whereas the proportion of biomass allocation to tubers was higher. At 58 DAE, the total biomass was significantly higher under the eCaT and eCeT conditions, and the total biomass of plants grown under the eCaT and eCeT conditions was significantly higher by 25.6% and 59.2%, respectively, compared to the aCaT condition ($P < 0.001$). The proportion of biomass allocation was the lowest for the leaves of plants grown under aCeT condition due to the deviation from the optimal growth-temperature range during the late growth phase.

Canopy net photosynthesis and carbon balance

Both CO$_2$ and temperature affected canopy net photosynthetic rates between 35 DAE and 58 DAE (Fig 4).
Fig 2. Growth parameters of potato plants at 35 DAE and 58 DAE under different conditions of CO$_2$ and temperature. Values are mean ± SE ($n = 6$). Bars showing different letters indicate significant differences among treatments at $P < 0.05$ according to the Tukey test. DAE: Days after emergence.

https://doi.org/10.1371/journal.pone.0241081.g002
The canopy net photosynthetic rates at 35 and 58 DAE increased rapidly between 08:00 a.m. and 11:00 a.m. At 35 DAE, the canopy net photosynthetic rate and light-response curve were highest under eCeT conditions followed by the aCeT, aCaT, and eCaT conditions. At 58 DAE, the canopy net photosynthetic rate and light-response curve were highest under eCaT conditions; however, there was no difference between values for plants grown under the aCaT and eCeT conditions even though lower values were observed under aCeT conditions. To compare daily canopy net photosynthesis levels between each condition of CO₂ concentration and temperature during the growth period, the box-plot analysis was carried out (Fig 5).

Net photosynthesis was significantly higher under the eCeT than under the other conditions ($P < 0.001$) and there was no significant difference in the values observed under the aCaT and eCaT conditions. However, the lowest value was obtained under aCeT conditions. The close association ($r^2 = 0.873$) between total carbon gain and total dry biomass was
determined using Eq (2) (Fig 6). Since the most of plant’s dry biomass (>90%) is derived from photosynthesis and the CER is closely related to plant’s dry biomass production, CER is an excellent indicator of plant growth and it has a close relationship between total carbon gain and above-ground total dry biomass [24, 34]. The slope (0.468) of the linear regression indicated that the carbon fixed from net photosynthesis accounted close to 46.8% (or 0.468 g C g⁻¹ dry biomass) of the total above-ground dry biomass; in potatoes, it is generally known to be 0.4–0.46 [24, 34].

**Leaf stomatal conductance and chlorophyll concentration**

The stomatal conductance and chlorophyll concentration changed significantly during the growth period depending on CO₂ concentration and temperature (Fig 7).

Stomatal conductance on 34 DAE was higher under aCeT conditions than under the other conditions (Fig 7A). From 42 DAE, an average reduction of 15% in stomatal conductance was observed under the eCaT and eCeT compared to the aCeT and aCaT conditions. A sharp decline in stomatal conductance was observed under aCeT conditions during the late-growth
Fig 5. Comparison of daily canopy net photosynthesis levels from 25 DAE to 64 DAE for potato plants grown under four different conditions of CO₂ and temperature. Means labeled with different letters differ significantly at \( P < 0.05 \).

https://doi.org/10.1371/journal.pone.0241081.g005

Fig 6. The relationship between the total carbon gain and total above-ground dry biomass of potato plants grown under different conditions of CO₂ and temperature. Potato plants were collected at 35 DAE, 58 DAE, and the final harvest growth stages.

https://doi.org/10.1371/journal.pone.0241081.g006
phase. The chlorophyll concentration on 34 DAE was unaffected by either temperature or CO$_2$ concentration. However, from 50 DAE, the chlorophyll concentration decreased by 9.4% under the eCaT and eCeT compared to the aCeT and aCaT conditions (Fig 7B).

### Organic carbon and mineral nutrients

The changes in the concentration of carbon, nitrogen, phosphorus, potassium, and magnesium in leaves according to the CO$_2$ concentration and temperature during the growth period were analyzed (Fig 8).

Carbon concentrations showed statistically significant differences depending on the growth stage and treatment conditions but not as much as nitrogen concentrations (Fig 8A). The nitrogen concentrations were on average 24% lower under the eCaT and eCeT compared to the aCeT and aCaT conditions (Fig 8B). These differences were also reflected in the C:N ratio (Fig 8C). The concentrations of phosphorus, potassium, and magnesium were generally lower (16% on average) under the eCaT and eCeT compared to the aCeT and aCaT conditions (Fig 8D–8F and S2 Table).

### Tuber yield and biomass

At harvest time, the tuber size, biomass, yield, and marketable yield of potatoes cultivated under varying CO$_2$ concentrations and temperatures were analyzed (Table 2).

There were significant differences in the number of small (< 30 g), medium (30–80 g), and large (> 80 g) potatoes. The tuber size was greater under eCeT than under the eCaT, eCaT, and aCeT conditions and was the smallest under aCeT conditions. There was no significant difference in the total number of tubers per plant among the aCaT, eCaT, and eCeT conditions but was lower at the aCeT conditions.

Biomass was significantly affected by the treatment conditions, with the highest leaf, stem, and tuber biomass achieved under eCeT conditions. There was no significant difference in the
Fig 8. Carbon (A), nitrogen (B), C:N ratio (C), phosphorus (D), potassium (E), and magnesium (F) in potato leaves at 35, 43, 50, 58, and 64 DAE under different CO$_2$ and temperature conditions. Values are mean ± SE ($n=3$). Bars indicate significant differences among treatments at $P < 0.05$ according to Tukey’s test. DAE: Days after emergence.

https://doi.org/10.1371/journal.pone.0241081.g008
biomass observed under the aCaT and eCaT conditions. The biomass was much lower under aCeT conditions.

Tuber yield and marketable yield were significantly affected by the treatment conditions and were 604 and 352 g, respectively, under eCeT conditions. Similar to the observed biomass, there was no significant difference in either tuber yield or marketable yield under the aCaT and eCaT conditions. Both the tuber and marketable yields were lower under the aCeT conditions (342 and 195 g, respectively).

The harvest index varied significantly across the treatment conditions, with the greatest index observed under eCaT conditions. ANOVA indicated that tuber yield was significantly affected by both the CO\textsubscript{2} concentration and the CO\textsubscript{2} × temperature interaction, whereas the harvest index was affected only by temperature.

### Discussion

#### The main effects of elevated temperature

During the early growth phase, plant height, biomass, and canopy net photosynthetic rate were higher under the aCeT and eCeT conditions (Figs 2, 3, and 4). Under the aCeT condition, however, leaf stomatal conductance, canopy net photosynthetic rate, and leaf area all declined sharply as the conditions deviated from the optimal growth temperature at the late-growth phase. As a result, the biomass and yield of the harvested tubers were significantly lower than those obtained under the other treatment conditions.

The harvest index varied significantly across the treatment conditions, with the greatest index observed under eCaT conditions. ANOVA indicated that tuber yield was significantly affected by both the CO\textsubscript{2} concentration and the CO\textsubscript{2} × temperature interaction, whereas the harvest index was affected only by temperature.

### Table 2. Numbers of different sized and total tubers, biomass, tuber yield, and harvest index of potato plants under different treatment conditions of CO\textsubscript{2} and temperature at the final harvest.

| Treatment | No. tubers (plant\textsuperscript{-1}) | Total no. tubers (plant\textsuperscript{-1}) | Biomass (g DW plant\textsuperscript{-1}) | Yield (g FW plant\textsuperscript{-1}) | Harvest Index |
|-----------|----------------------------------------|--------------------------------------------|----------------------------------------|----------------------------------------|---------------|
|           | <30 g | 30–80 g | >80 g | Leaf | Stem | Tuber | Tuber | Marketable |               |
| aCaT      | 4.8\textsuperscript{a} | 2.9\textsuperscript{b} | 2.3\textsuperscript{b} | 10.2\textsuperscript{a} | 10.4\textsuperscript{a} | 7.1\textsuperscript{b} | 106.4\textsuperscript{b} | 502\textsuperscript{b} | 273\textsuperscript{b} | 0.87\textsuperscript{a} |
| eCaT      | 5.2\textsuperscript{a} | 3.9\textsuperscript{a} | 2.4\textsuperscript{a} | 11.5\textsuperscript{a} | 9.8\textsuperscript{b} | 6.2\textsuperscript{b} | 110.3\textsuperscript{b} | 527\textsuperscript{b} | 269\textsuperscript{b} | 0.89\textsuperscript{a} |
| aCeT      | 2.0\textsuperscript{a} | 2.8\textsuperscript{b} | 1.9\textsuperscript{b} | 6.4\textsuperscript{a} | 7.6\textsuperscript{b} | 6.0\textsuperscript{b} | 76.9\textsuperscript{a} | 342\textsuperscript{b} | 195\textsuperscript{b} | 0.84\textsuperscript{a} |
| eCeT      | 3.9\textsuperscript{b} | 4.1\textsuperscript{a} | 2.9\textsuperscript{a} | 10.8\textsuperscript{a} | 14.4\textsuperscript{a} | 9.3\textsuperscript{a} | 127.5\textsuperscript{a} | 604\textsuperscript{a} | 352\textsuperscript{b} | 0.82\textsuperscript{b} |

ANOVA: CO\textsubscript{2} × Temp: ns. All other effects were significant (*** P < 0.001; ** P < 0.01; * P < 0.05).

Means followed by the same letters in each column are not significantly different at P < 0.05 according to Tukey’s test.

*Significant at the 0.05 probability level;
**Significant at the 0.01 probability level;
***Significant at the 0.001 probability level;
NS—not significant; Tuber size: <30 g Small, 30–80 g Medium, >80 g Large; Marketable: weight > 80 g; Harvest index: tuber dry weight / total dry weight; Temp: Temperature.

https://doi.org/10.1371/journal.pone.0241081.t002
Rubisco enzyme for CO₂ decreases compared to the specificity for O₂ and the solubility of O₂ decreases less than that of CO₂ making it a more favorable environment for oxygenation reactions [49]. The accelerated loss of assimilated CO₂ consequently reduces the photosynthetic rate [48, 60] and ultimately leads to a decrease in the weight and yield of the tubers [12, 15, 19, 24].

The main effects of increased CO₂ concentration. In the present study, potato plants grown under the elevated CO₂ condition showed a relatively lower leaf area and biomass at the early growth stage than the other conditions. This outcome partially agrees with the results of previous OTC and FACE studies that reported that elevated CO₂ concentrations could reduce the above-ground growth during the early growth phase [30, 50–52]. Towards the late-growth phase, the canopy net photosynthetic rate and biomass were relatively higher under the eCaT than under the aCaT conditions but the final slightly increased tuber yield was not statistically different from that obtained under the aCaT conditions. Previous studies conducted in a growth chamber, OTC, and FACE have demonstrated that although the effect varies depending on the experimental conditions, elevated CO₂ concentrations have beneficial effects on the photosynthetic rate, growth, and yield of potatoes [27, 28, 30, 50–52, 61]. However, in this study, the eCaT condition with only an elevated CO₂ did not lead to a distinct increase in the yield of potatoes; this outcome may be related to the photosynthesis acclimation observed upon long-term exposure to elevated CO₂ concentrations or the relatively densely grown environments.

Studies have reported that stomatal conductance and chlorophyll concentrations decrease at CO₂ concentrations two-times higher than the current atmospheric levels [11, 62–65]. The findings of the present study concur that both stomatal conductance and chlorophyll concentrations decrease rapidly towards the late-growth phase when plants are grown under elevated CO₂ concentrations (Fig 7B). Stomatal conductance is known to decrease under elevated CO₂ conditions, and the responses vary according to the diversity of plant species and the environment [7]. A particularly steep decrease occurs upon long-term rather than short-term exposure [39, 45]. On the other hand, long-term exposure to elevated CO₂ concentrations causes photosynthesis acclimation that is accompanied by a reduced stomatal conductance, a decrease in leaf chlorophyll concentration, and a reduction in the Rubisco activation state in the photosynthetically active source leaves [36].

The interaction of temperature and CO₂
Many of the effects of elevated CO₂ on plant growth and carbon metabolism are offset or influenced by elevated temperatures [49]. For example, the inhibition of photorespiration by elevated CO₂ is temperature-dependent and the benefits of increased CO₂ in photosynthesis are greater at higher temperatures. To realistically analyze how climate change affects crops, these climate-change factors need to be investigated together. A recent study in rice has shown that the interactive effects of elevated CO₂ and temperature are advantageous for photosynthesis but not for the growth and yield of rice [4]. It was found that the high-temperature condition causes yield loss regardless of whether it is a high temperature alone or combined with an elevated CO₂ concentration.

In the present study, potato plants grown under eCeT conditions exhibited the greatest height, leaf area, total biomass, and canopy net photosynthetic rates during the early growth phase. Towards the late-growth phase, the canopy net photosynthetic rate became lower than that of plants grown under eCaT conditions (Fig 4). As a result, during the early growth phase, the photosynthetic rate was high under the eCeT condition owing to the elevated temperature. However, during the late growth phase, the elevated CO₂ and temperature are known to affect
the assimilates and cause them to accumulate in mesophyll cells or chlorophylls, thereby reducing the photosynthetic rate [11, 62]. Generally, the net photosynthetic rate decreases more rapidly than the respiration rate when the temperature is elevated and the net assimilation has been reported to decrease above certain temperature levels [63, 66]. Nevertheless, under the eCeT condition, the biomass, yield, and marketable yield of potato tuber was the highest among the treatment conditions at the final harvest because starch, as an assimilate, was accumulated in the tubers for storage. Correspondingly, the accumulated ratio of assimilates produced by the net photosynthesis under the elevated CO₂ concentrations is relatively higher, whereby the proportion of tubers in the total biomass increases [27]. The high tuber yield in this study was attributed to the increase in tuber size rather than in tuber number. The findings agree with previous studies showing that the increase in tuber yield under elevated CO₂ conditions is mainly due to an increase in the size of already initiated tubers rather than an increase in the number of tubers [27, 30, 47, 50, 67].

Elevated CO₂ concentrations can help restore PS II activity after it is reduced by an elevated temperature [66, 68]. Even though PS II activity was not measured in the present study, we found that the concurrent elevation of temperature and CO₂ enhanced plant thermostability and reduced the damaging effect of elevated temperature.

Changes in organic carbon, nitrogen, and mineral concentrations
The balance between carbon and nitrogen is a critical factor in plant growth and development; in general, the concentrations of carbon and nitrogen in the plant biomass are 45% and 5%, respectively [69]. In the present study, the elevation of CO₂ concentration had negative effects on the concentrations of nitrogen, as well as phosphorus, potassium, and magnesium (Fig 8); this is consistent with the previous FACE, OTC, CTC, glasshouse, and growth-chamber studies showing that the elevation of CO₂ concentration can reduce nitrogen concentration by 7–18% depending on the crop species and variety [41, 70–73]. However, the decrease observed in potatoes was relatively large compared to the losses observed in other crop species.

The photosynthesis of C₃ plants is mediated via CO₂ fixation in Calvin’s cycle and CO₂ is fixed by the enzyme Rubisco that catalyzes the carboxylation of ribulose-1,5-bisphosphate, the first major step of carbon fixation. Some studies have reported that prolonged exposure to elevated CO₂ lowers the content and activity of Rubisco enzyme and causes an increase in the C:N ratio with lower N levels resulting in lower protein levels [26, 36, 41]. Increased levels of assimilates resulting from the elevated CO₂ concentration have been reported to dilute the nitrogen concentration of plants [72].

Overall, elevated CO₂ conditions reduce stomatal conductance, inhibit the rubisco-activating enzyme, and increase the C:N ratio. However, the efficiency of nitrogen use has been reported to increase despite reductions in nitrogen concentration [35]. The level of magnesium contained in chlorophyll has been reported to decrease by 1–8% under elevated CO₂ conditions [40, 74]. Elevated CO₂ can cause the accumulation of assimilates in photosynthetically active source leaves when the photosynthetic rate exceeds the export capacity or the capacity of sink organs, thereby reducing the photosynthetic rate [11, 62]. Thus, source-sink imbalance may occur under the elevated CO₂ condition and the delayed transport or accumulation of assimilates may drive the reduction in photosynthetic rate during the late-growth phase.

Conclusions
The study examined the effects of CO₂ concentration and temperature on the growth, yield, and photosynthesis of potato crops using a KSPAR chamber. At elevated temperature, both stomatal conductance and canopy net photosynthetic rate decreased sharply; as the
temperature deviated from the effective range towards the late-growth phase, both biomass and tuber yield were reduced. At an elevated CO\textsubscript{2} level, the area and biomass of leaves that create photosynthetic products increased during the late-growth phase. However, the tuber yield did not change significantly compared to that obtained under the aCaT conditions. The concurrent elevation of CO\textsubscript{2} and temperature caused a distinct difference in the growth, development, and canopy net photosynthetic rate of potato plants compared to independent elevation. This increased biomass and tuber yield was due to the enlarged tuber size rather than to an increase in the number of tubers.

The source and sink organs of crop plants can change according to the external environment and the influence of the CO\textsubscript{2} and temperature is particularly significant. Our study indicates that excessive increases in only one of these parameters will prevent the balanced development of the source and sink organs of potatoes. However, in conditions of concurrently elevated CO\textsubscript{2} and temperature, the source and sink organs are likely to exhibit balanced development that may have positive effects on crop growth and quality. The elevation of CO\textsubscript{2} and temperature within the effective range will exert positive effects on the growth, yield, and photosynthesis of potato crops. To elucidate the conditions of concurrent elevation in a more systematic way, molecular-biology experiments should be conducted.

**Supporting information**

S1 Fig. Temperature setting for KSPAR chambers programmed to maintain ambient Temperature (aT) or ambient temperature plus 4˚C (eT) conditions. DAE: days after emergence; Average: daily mean temperature of the Jeonju area (1991–2010).

(TIF)

S2 Fig. Mean daily solar radiation for the Days After Emergence (DAE) at the National Institute of Crop Science (35˚84’34” N, 127˚04’84” E).

(TIF)

S1 Table. ANOVA results for the growth parameters of the potato plants grown under the four different treatment conditions.

(DOCX)

S2 Table. ANOVA results of C, N, C: N, P, K, and Mg parameters at 35, 43, 50, 58, 64 Days After Emergence (DAE).

(DOCX)

**Author Contributions**

**Conceptualization:** Wan-Gyu Sang, Myung-Chul Seo, Jung-II Cho.

**Formal analysis:** Yun-Ho Lee, Wan-Gyu Sang, Jae-Kyeong Baek, Jun-Hwan Kim, Pyeong Shin, Jung-II Cho.

**Funding acquisition:** Myung-Chul Seo.

**Investigation:** Yun-Ho Lee, Jae-Kyeong Baek, Jun-Hwan Kim, Pyeong Shin, Myung-Chul Seo.

**Project administration:** Jung-II Cho.

**Writing – original draft:** Yun-Ho Lee, Jung-II Cho.

**Writing – review & editing:** Yun-Ho Lee, Jung-II Cho.
References

1. NASA. Global climate change. 2019 [cited 2020 January]. Vital signs of planet. http://climate.nasa.gov/vital-signs/carbon-dioxide

2. IPCC (Intergovernmental Panel on Climate Change) Climate change 2014. Synthesis report. contribution of working groups I, II and III to the fifth assessment report of the intergovernmental panel on climate change, the core writing team, R. K. Pachauri, and L. A. Meyer. Geneva: IPCC; 40–54.

3. Yubi Y, Jun L, Haiyang N, Xiuyun Z. Collaborative influence of elevated CO$_2$ concentration and high temperature on potato biomass accumulation and characteristics. Open Chem. 2019; 17: 728–737.

4. Wang W, Gai C, He J, Gu J, Zhu G, Zhanf W, et al. Yield, dry matter distribution and photosynthetic characteristics of rice under elevated CO$_2$ and increased temperature conditions. Field Crops Research. 2019; 248, 107605.

5. Liu S, Waqas MA, Wang S, Xiong X, Wan Y. Effects of increased levels of atmospheric CO$_2$ and high temperatures on rice growth and quality. PLoS ONE. 2017; 12: e0187724. https://doi.org/10.1371/journal.pone.0187724 PMID: 29145420

6. Kimball BA. Carbon dioxide and agricultural yield: an assemblage and analysis of 430 prior observations. J. Agron. 1983; 75: 779–788.

7. Bunce J. Direct and acclamatory responses of stomatal conductance to elevated carbon dioxide in four herbaceous crop species in the field. Global Change Biol. 2001; 7: 323–331.

8. Jablonski LM, Wang X, Curtis PS. Plant reproduction under elevated CO$_2$ conditions: a meta-analysis of reports on 79 crop and wild species. New Phytol. 2002; 156: 9–26.

9. Navarre R.; Pavek MJ. The potato botany, production and uses. 3 Potato utilization and markets. CABI. 2014. pp 29–34.

10. Hammes PS, Jager JA. Net photosynthetic rate of potato at high temperature. Potato Res. 1990; 33: 515–520.

11. Prasad PVV, Allen LH Jr, Boote KJ. Crop responses to elevated carbon dioxide and interaction with temperature. J. Crop Imp. 2005; 13: 113–155.

12. Rykaczewska K. The impact of high temperature during growing season on potato cultivar with different response environmental stresses. Am. J. Plant Sci. 2013; 4: 2386–2393.

13. Tang R, Niu S, Zhang G, Chen G, Haroon M, Yang Q, et al. Physiology and growth responses of potato cultivars to heat stress. Botany. 2018; 96: 897–912.

14. Hatfield JL, Prueger JH. Temperature extremes: Effect on plant growth and development. Weather. Clim. Extrem. 2015; 10: 4–10.

15. Kim YU, Lee BW. Differential mechanisms of potato yield loss induced by high day and night temperatures during tuber initiation and bulking: photosynthesis and tuber growth. Front. Plant Sci. 2019; 10: 300. https://doi.org/10.3389/fpls.2019.00300 PMID: 30923532

16. Yamori W, Hikosaka K, Way DA. Temperature response of photosynthesis in C$_3$, C$_4$, and CAM plant: temperature acclimation and temperature adaptation. Photosynth Res. 2014; 119: 101–117. https://doi.org/10.1007/s11120-013-9874-6 PMID: 23901717

17. Prange RK, McRae KB, Midmore DJ, Deng R. Reduction in potato growth at high temperature: role of photosynthesis and dark respiration. Am. J. Potato Res. 1990; 67: 357–369.

18. Barták M, Nijs I, Impens I. The susceptibility of PS II of Lolium perenne to a sudden fall in air temperature- response of plant grown in elevated CO$_2$ and/or increased air temperature. Environ. Exp. Bot. 1998; 39: 85–95.

19. Aienl A, Kheterpal S, Pal M. Photosynthetic characteristics of potato cultivars grown under high temperature. J. Agri. Environ. Sci. 2011; 11: 633–639.

20. Menzel CM. Tuberization in potato at high temperatures: interaction between temperature and irradiance. Ann. Botany. 1985; 55: 35–39.

21. Wolf S, Oleinski AA, Marani A. Effect of high temperature on photosynthesis in potatoes. Ann. Bot. 1990; 65: 179–185.

22. Cao WX, T_pubbitts TW. Phasic temperature change patterns affect growth and tuberization in potatoes. J. Am. Soc. Hort. Sci. 1994; 119: 775–778.

23. Fleisher DH, Timlin DJ. Modeling expansion of individual leaves in the potato canopy. Agric. For. Meteorol. 2006; 139: 84–93.

24. Timlin D, Rahman Lutfur SM, Baker J, Reddy VR, Fleisher D, Quebedaux B. Whole plant photosynthesis, development, and carbon partition in potato as a function of temperature. J. Agron. 2006; 98: 1195–1203.

25. Rogers HH, Dahlman RC. Crop responses to CO$_2$ enrichment. Vegetatio. 1993; 104/105: 117–131.
26. Long SP, Ainsworth EA, Rogers A, Ort DR. Rising atmospheric carbon dioxide: plants FACE the future. Annu. Rev. Plant Biol. 2004; 55: 591–628. https://doi.org/10.1146/annurev.arplant.55.031903.141610 PMID: 15377233

27. Finnan JM, Donnelly A, Jones MB, Burke I. The effect of elevated levels of carbon dioxide on Potato crops. J. Crop Imp. 2005; 13: 91–110.

28. Fleisher DH, Timlin DJ, Yang Y, Reddy VR, Reddy KR. Uniformity of Soil-Plant-Atmosphere-Research chambers. Am. Soc. Agric. Eng. 2009; 52: 1721–1731.

29. Reddy RA, Rasineni KG, Raghavendra AS. The impact of global elevated CO2 concentration on photosynthesis and plant productivity. Curr. Sci. 2010; 99: 46–55.

30. Miglietta F, Magliulo V, Bindi M, Cerio L, Vaccafi FP, Peressottis A. Free air CO2 enrichment of potato (Solanum tuberosum L.) development, growth and yield. Global Change Biol. 1998; 4: 163–172.

31. Heagle AS, Miller JE, Pursley WA. Atmospheric pollutants and trace gases. Technical Reports. 2003; 32: 1603–1610.

32. Jeffery SC, Cochran VL. Response of potato (Solanum tuberosum L) to elevated atmospheric CO2 in North American Subarctic. Agric. Ecosys. Environ. 2006; 112: 49–57.

33. Chen CT, Setter TL. Response of potato dry matter assimilation and partitioning to elevated CO2 at various stages of tuber initiation and growth. Environ. Exp. Bot. 2012; 80: 27–34.

34. Fleisher DH, Timlin DJ, Reddy VR. Elevated carbon dioxide and water stress effects on potato canopy gas exchange, water use, and productivity. Agric. For. Meteorol. 2008; 148: 1109–1122.

35. Leakey ADB, Ainsworth EA, Bernacchi CJ, Roger A, Long SP, Ort DR. Elevated CO2 effects on plant carbon, nitrogen, and water relations: six important lessons from FACE. J. Exp. Bot. 2009; 60: 2859–2876. https://doi.org/10.1093/jxb/erp096 PMID: 19401412

36. Kant S, Seneweerapu R, Rodin J, Materne M, Burch D, Rothstein SJ, et al. Improving yield potential in crops under elevated CO2: integrating the photosynthetic and nitrogen utilization efficiencies. Plant Sci. 2012; 3: 162. https://doi.org/10.3389/fpls.2012.00162 PMID: 22837349

37. Woodward FI, Kelly CK. The influence of CO2 concentration on stomatal density. New Phytol. 1995; 131: 311–327.

38. Wheeler RM, Mackowiak CL, Yorioand NC, Seger JC. Effects of CO2 on stomatal conductance: Do stomata open at very high CO2 concentrations? Ann. Bot. (Lond). 1999; 83: 243–251.

39. Sicher RC, Bunce JA. Adjustments of net photosynthesis in Solanum tuberosum in response to reciprocal change in ambient and elevated growth CO2 partial pressures. Physiol. Plant. 2001; 112: 55–61. https://doi.org/10.1043/j.1399-3054.2001.1120108.x PMID: 11319015

40. Fangmeier A, Temmerman De, Black C, Persson K, Vorne V. Effects of elevated CO2 and/or ozone on nutrient concentrations and nutrient uptake of potatoes. Eur. J. Agron. 2012; 3: 162. https://doi.org/10.3389/fpls.2012.00162 PMID: 22837349

41. Ainsworth EA, Long SP. What have we learned from 15 years of free-air CO2 enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO2. New Phytol. 2005; 165: 351–372. https://doi.org/10.1111/j.1469-8137.2004.01224.x PMID: 15720649

42. Xu Z, Jiang Y, Jia B, Zhou G. Elevated-CO2 response of stomatal and its dependence on environmental factors. Front. Plant Sci. 2016; 7: 657. https://doi.org/10.3389/fpls.2016.00657 PMID: 27242658

43. Ziska L, Crimmins A. The impacts of climate change on human health in the united states food safety, nutrition, and distribution. US. Global Change Research Program. 2016; 190–216.

44. Gifford RM, Barrett DJ, Lutze JL. The effects of elevated [CO2] on the C: N and C: P mass rations of plant tissues. Plant. Soil. 2000; 224: 1–4.

45. Schapendonk AHCM, Oijen Mv, Dijkstra P, Sander PC, Jordi W.JRM, Stoopen GM. Effect of elevated CO2 concentration on photosynthetic acclimation and productivity of two potato cultivars grown in open-top chambers. Aust. J. Plant Physiol. 2000; 27: 1119–1130.

46. Ainsworth EA, Rogers A. The response of photosynthesis and stomatal conductance to rising [CO2]: mechanisms and environmental interactions. Plant Cell Environ. 2007; 30: 258–270. https://doi.org/10.1111/j.1365-3040.2007.01641.x PMID: 17263773

47. Kumari S, Agrawal M. Growth, yield and quality attributes of tropical potato variety (Solanum tuberosum L.cv Kufri chandramukhi) under ambient and elevated carbon dioxide and ozone and their interaction. Ecotoxicol. Environ. Safety. 2014; 101: 146–156. https://doi.org/10.1016/j.ecoenv.2013.12.021 PMID: 24507140

48. Xu Z, Jiang Y, Zhou G. Response and adaptation of photosynthesis, respiration, and antioxidant systems to elevated CO2 with environmental stress in plants. Front. Plant Sci. 2015; 6: 701. https://doi.org/10.3389/fpls.2015.00701 PMID: 26442017
Dusenge ME, Duarte AG, Way DA. Plant carbon metabolism and climate change: elevated CO$_2$ and temperature impacts on photosynthesis, photorespiration and respiration. New Phytol. 2019; 221, 32–49. https://doi.org/10.1111/nph.15283 PMID: 29983005

Donnelly A, Craigo J, Black CR, Colls JJ, Landon G. Effects of elevated CO$_2$ increases biomass and tuber yield in potato even at high ozone concentrations. New Phytol. 2001; 149: 265–274.

Hacour A, Craigo J, Vandermerrien K, Ojanperä K, Pleijel H, Danielsson H, et al. CO$_2$ and ozone effects on canopy development of potato crops across Europe. Eur. J. Agron. 2002; 17: 257–272.

Hög P, Fangmeier A. Atmospheric CO$_2$ enrichment affects potatoes: 1. Aboveground biomass production and tuber yield. Eur. J. Agron. 2009; 30: 78–84.

Reddy KR, Hodges HF, Red JJ, Mckinion JM, Baker JT, Trapley L, et al. Soil-plant-atmosphere-research (SPAR) facility: A tool for plant research and modeling. Biotronics. 2001; 30: 27–50.

Baker JT, Kim SH, Gitz DC, Timlin D, Reddy VR. A method for estimating carbon dioxide leakage rates in controlled-environment chambers using nitrous oxide. Environ. Exp. Bot. 2004; 51: 103–110.

Frank B. Fundamentals of chemistry: Laboratory studies. 3rd edition. Elsevier Science. 2012. pp. 75.

Hansen TH, Laursen KH, Persson DP, Pedas P, Hustend S, Schjoerring JK. Micro-scale high-throughput digestion of plant tissue samples for multi-elemental analysis. Plant Meth. 2009; 5: 1–11.

Kim YU, Seo BS, Choi DH, Ban HY, Lee BW. Impact of high temperatures on the marketable tuber yield and related traits of potato. Eur. J. Agron. 2017; 89: 46–52.

Fleisher DH, Timlin DJ, Reddy V.R. Temperature influence on potato leaf and branch distribution and canopy photosynthetic rate. J. Agron. 2006; 98: 1442–1452.

Kaushal N, Bhandari K, Siddique KHM, Nayyar H. Flood crops face rising temperatures: An overview of responses, adaptive mechanisms, and approaches to improve heat tolerance. Cogent food & agriculture. 2013; 2: 1134380.

Chen WR, Zheng JS, Li YQ, Guo WD. Effects of high temperature on photosynthesis chlorophyll fluorescence, chloroplast ultrastructure, and antioxidant activities in fingered citron. Russ. J. Plant Physiol. 2012; 59: 732–740.

Wheeler RM, Mackowiak CL, Sager JC, Knott WM. Growth of soybean and potato at high CO$_2$ partial pressures. Adv. Space. Res. 1994; 14: 251–255. https://doi.org/10.1016/0273-1177(94)90305-0 PMID: 11540190

Lawson T, Craigo J, Tulloch AM, Black CR, Colls JJ, Landon G. Photosynthetic responses to elevated CO$_2$ and O$_3$ in field-grown potato (Solanum tuberosum). J. Plant Physiol. 2001a; 158: 309–323

Lawson T, Craigo J, Black CR, Colls JJ, Tulloch AM, Landon G. Effects of elevated carbon dioxide and ozone on the growth and yield of potatoes (Solanum tuberosum) grown in open-top chambers. Environ. Pollut. 2001b; 111: 479–491. https://doi.org/10.1016/s0269-7491(00)00080-4 PMID: 11202753

Kumari K, Agrawal M, Singh A. Effects of ambient and elevated CO$_2$ and ozone on physiological characteristics, antioxidative defense system and metabolites of potato in relation to ozone flux. Environ. Exp. Bot. 2015; 109: 276–287.

Lahijani MJA, Kafi M, Nezami A, Nabat J, Mehrjerdi MZM, Shahkoomahally S, et al. Variation in assimilation rate, photo assimilate translocation, and cellular fine structure of potato cultivars (Solanum Tuberum L.) exposed to elevated CO$_2$. Plant Physiol. Biochem. 2018; 130: 303–313. https://doi.org/10.1016/j.plaphy.2018.07.019 PMID: 30036859

Morison JIL, Lawlor DW. Interaction between increasing CO$_2$ concentration and temperature on plant growth. Plant Cell Environ. 1999; 22: 659–682.

Graigon J, Fangmeier A, Jones M, Donnelly A, Bindi M, Temmerman LD, et al. Growth and marketable-yield responses of potato to increased CO$_2$ and ozone. Eur. J. Agron. 2002; 17: 273–289.

Taub DR, Seemann JR, Coleman J. Growth in elevated CO$_2$ protects photosynthesis against high-temperature damage. Plant Cell Environ. 2000; 23: 649–656.

Marschner H. Mineral nutrition of higher plants, 2nd Edn. San Diego, CA: Academic Press. 1995. pp. 229–239.

Cao WX, Tibbits TW. Starch concentration and impact on specific leaf weight and element concentrations in potato leaves under aird carbon dioxide and temperature. J. Plant Nutr. 1997; 20: 887–881.

Cotrufo MF, Ineson P, Scott A. Elevated CO$_2$ reduces the nitrogen concentration of plant tissues. Global Change Biol. 1998; 4: 43–54.

Taub DR, Miller B, Allen H. Effects of elevated CO$_2$ on the protein concentration of food crops: a meta-analysis. Global Change Biol. 2008; 14: 565–575.
73. Dong J, Gruda N, Lam SK, Li L, Duan Z. Effects of elevated CO$_2$ on nutritional quality of vegetables: a review. Front. Plant Sci. 2018; 9: 924. https://doi.org/10.3389/fpls.2018.00924

74. McGrath J, Lobell D. Reduction of transpiration altered nutrient allocation contribute to nutrient decline of crops grown in elevated CO$_2$ concentrations. Plant Cell Environ. 2013; 36: 697–705. https://doi.org/10.1111/pce.12007 PMID: 22943419