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To cite this version:
Célestine Atyame Nten, Nicole Pasteur, Emilie Dumas, Pablo Tortosa, Michaël L. Tantely, et al.. Cytoplasmic Incompatibility as a Means of Controlling Culex pipiens quinquefasciatus Mosquito in the Islands of the South-Western Indian Ocean. PLoS Neglected Tropical Diseases, Public Library of Science, 2011, 5 (12), pp.e1440. 10.1371/journal.pntd.0001440. hal-01274608

HAL Id: hal-01274608
https://hal.univ-reunion.fr/hal-01274608
Submitted on 14 Jun 2018

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Cytoplasmic Incompatibility as a Means of Controlling *Culex pipiens quinquefasciatus* Mosquito in the Islands of the South-Western Indian Ocean

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Abstract

The use of the bacterium *Wolbachia* is an attractive alternative method to control vector populations. In mosquitoes, as in members of the *Culex pipiens* complex, *Wolbachia* induces a form of embryonic lethality called cytoplasmic incompatibility, a sperm-egg incompatibility occurring when infected males mate with uninfected females or with females infected with incompatible *Wolbachia* strain(s). Here we explore the feasibility of the Incompatible Insect Technique (IIT), a species-specific control approach in which field females are sterilized by inundative releases of incompatible males. We show that the *Wolbachia* wPip(Is) strain, naturally infecting *Cx. p. pipiens* mosquitoes from Turkey, is a good candidate to control *Cx. p. quinquefasciatus* populations on four islands of the south-western Indian Ocean (La Réunion, Mauritius, Grande Glorieuse and Mayotte). The wPip(Is) strain was introduced into the nuclear background of *Cx. p. quinquefasciatus* mosquitoes from La Réunion, leading to the LR[wPip(Is)] line. Total embryonic lethality was observed in crosses between LR[wPip(Is)] males and all tested field females from the four islands. Interestingly, most crosses involving LR[wPip(Is)] females and field males were also incompatible, which is expected to reduce the impact of any accidental release of LR[wPip(Is)] females. Cage experiments demonstrate that LR[wPip(Is)] males are equally competitive with La Réunion males resulting in demographic crash when LR[wPip(Is)] males were introduced into La Réunion laboratory cages. These results, together with the geographic isolation of the four south-western Indian Ocean islands and their limited land area, support the feasibility of an IIT program using LR[wPip(Is)] males and stimulate the implementation of field tests for a *Cx. p. quinquefasciatus* control strategy on these islands.

Introduction

The last few years have witnessed an increasing interest in the alpha-proteobacterium *Wolbachia* (Rickettsiiales) for the biological control of insect pest populations [for reviews see [1–5]]. *Wolbachia* is the most common intracellular bacterium yet described [6,7], present in more than 65% of insect species and found in all major insect families [8]. Some medically important mosquitoes are naturally infected by *Wolbachia*, such as the common house mosquito *Aedes aegypti* [9,10] and the Asian tiger mosquito *Aedes albopictus* [11], or can otherwise be artificially infected, such as the yellow fever mosquito *Ae. aegypti* [12–14]. *Wolbachia* is vertically inherited from a female host to its progeny through the egg cytoplasm, males being a dead end in terms of transmission [4,15]. *Wolbachia* is usually termed a ‘reproductive parasite’ in the sense that it optimizes its transmission by manipulating its host’s reproductive biology [15,16]. In mosquitoes, *Wolbachia* induces a form of embryonic death called cytoplasmic incompatibility (CI) [9]. This phenomenon results from sperm-egg incompatibility occurring when *Wolbachia*-infected males mate with uninfected females or females infected with an incompatible *Wolbachia* strain [17]. Therefore, CI has been investigated as a mechanism to control field populations [1,18,19,20–22], or to drive transgenes into field populations [2,3,10,23]. In addition, recent investigations showed that...
Wolbachia strains (here arbitrarily named far [39–43]. When the largest variation of CI crossing types observed in arthropods thus different the complex are the subspecies Wolbachia B group of Wolbachia as \( w \)[10,36,37]. Recent multi-loci typing approaches revealed that the is high in natural populations with currently expanding in the Indian Ocean [34,35]. This species also transmits the Rift Valley Fever (RVF) virus, thousands of cases have been identified in the last decade [32,33] Mediterranean countries [30,31] and in the United States where case, for example, of the West Nile Virus (WNV), recrudescent in known vector for many arboviruses worldwide [29]. This is the authors), representing the southern and northern mosquito populations, which could be implemented on the SWIO islands. First, the regional genetic diversity of \( Cx. p. pipiens \) and the medfly \( Ae. polynesiensis \) [49] and the medfly Ceratitis capitata [19].

We focused on natural populations of \( Cx. p. quinquefasciatus \) collected on five islands in the south-western Indian Ocean (SWIO): La Réunion, Mauritius, Mayotte, Madagascar and Grande Glioreuse. Prior studies have demonstrated that \( Cx. p. quinquefasciatus \) lines from La Réunion are infected with closely related \( w \)Pip strains which express complete CI (ca. 100% embryo mortality) with \( Cx. pipiens \) lines from distant geographic areas and infected by genetically different \( w \)Pip strains [43]. The \( Cx. p. pipiens \) Is line from Turkey, infected by the \( w \)Pip(Is) strain, is of particular interest: all crosses between Is males and females from La Réunion are incompatible and almost all reciprocal crosses are incompatible as well [43]. This complete bidirectional CI makes the \( w \)Pip(Is) strain a good candidate for an IIT program.

In this study, we obtained robust data that encourage the use of \( w \)Pip(Is)-infected \( Cx. p. quinquefasciatus \) males in an IIT program which could be implemented on the SWIO islands. First, the regional genetic diversity of \( w \)Pip infections is low as all identified \( w \)Pip strains belong to the \( w \)Pip-I group; this indicates that immigration of mosquitoes into the controlled area is unlikely to introduce a new \( w \)Pip strain compatible with \( w \)Pip(Is)-infected males. Second, the \( w \)Pip(Is) strain, from the \( w \)Pip-IV group, was introduced into the nuclear background of \( Cx. p. quinquefasciatus \) mosquitoes, leading to a line (LR[\( w \)Pip(Is)]) expressing complete CI with wild females sampled from all 5 SWIO Islands. Last, CI properties expressed by this line are optimal as (i) there is no effect of males ageing on CI expression, (ii) LR[\( w \)Pip(Is)] males show similar body size and longevity as males from La Réunion Island, suggesting good competitiveness of incompatible males vs. wild males, which was further confirmed in cage confrontations and (iii) LR[\( w \)Pip(Is)] mosquitoes are mainly bidirectionally incompatible with La Réunion, Mauritius, Mayotte and Grande Glioreuse field mosquitoes: this lowers the risk of \( Wolbachia \) replacement possibly induced by accidental releases of LR[\( w \)Pip(Is)] females.

Materials and Methods

Mosquito collections

Two laboratory lines of \( Cx. p. pipiens \) mosquitoes naturally infected by \( Wolbachia \) were used in the experiments: the isofemale line Is, a \( Cx. p. pipiens \) line from Turkey infected by the \( w \)Pip(Is) strain, and the \( Cx. p. quinquefasciatus \) LR line, infected by the \( w \)Pip(LR) strain, and established from several hundred field-caught larvae in La Réunion Island (Table 1 and Figure 1). In addition, one uninfected line, LR-TG, was generated by curing \( Wolbachia \) of mosquitoes from the LR line with antibiotic, following the protocol described in [50]. Briefly, ca. 5,000 LR larvae were reared for three
generations in a solution containing tetracycline hydrochloride at concentrations of 10^{-1}, 2 \times 10^{-4} and 4 \times 10^{-4} M for the first, second and third-instar larvae, respectively. Mosquitoes from LR-TC were next reared for at least two generations in the absence of tetracycline before experiments, to prevent any possible side-effects of the treatment.

Field Cx. p. quinquefasciatus larvae and pupae were collected during the summers 2007–2011 in 29 natural breeding sites on five islands of the Indian Ocean: La Réunion (16 populations), Mauritius (four populations), Mayotte (three populations) Madagascar (five populations) and Grande Glorieuse (one population) (Table 1 and Figure 1). Specimens were brought to the laboratory for emergence and identification. Individuals were either directly stored in 70% EtOH for molecular analyses or kept alive for crossing experiments. All mosquitoes were reared in 65 dm³ cages kept at ca. 25 ± 2°C with 12 h/12 h light/dark cycle. Larvae were fed ad libitum with a mixture of shrimp powder and rabbit pellets, and adults with a honey solution.

Table 1. Mosquito collections.

| Laboratory lines | Culex pipiens taxon | Origin | Number of screened field specimens | Year of collection | Reference |
|------------------|---------------------|--------|------------------------------------|--------------------|-----------|
| Istanbul (Is)    | pipiens             | Istanbul (Turkey) | –                                  | 2003               | [34]      |
| LR               | quinquefasciatus    | Etang Salé, Saint Pierre and Saint Louis (La Réunion) | –                                  | 2009               | This study |
| LR-TC            | quinquefasciatus    | derived from LR  | –                                  | –                  | This study |
| LR(Pip(Is))      | quinquefasciatus    | derived from LR and Is  | –                                  | –                  | This study |

Natural populations

| #1, Tana1    | quinquefasciatus | Antananarivo (Madagascar) | 23 | 2010 | This study |
| #2, Tana2    | quinquefasciatus | Antananarivo (Madagascar) | 22 | 2010 | This study |
| #3, Itaoys1  | quinquefasciatus | Antananarivo (Madagascar) | 20 | 2010 | This study |
| #4, Itaoys2  | quinquefasciatus | Antananarivo (Madagascar) | 20 | 2010 | This study |
| #5, Mada     | quinquefasciatus | Antananarivo (Madagascar) | 20 | 2011 | This study |
| #6, Grande Glorieuse | quinquefasciatus | Grande Glorieuse | 24 | 2011 | This study |
| #7, Acoua    | quinquefasciatus | Acoua (Mayotte) | 24 | 2011 | This study |
| #8, Tsoundzou| quinquefasciatus | Tsoundzou (Mayotte) | 21 | 2010 | This study |
| #9, M’Tsamoudou | quinquefasciatus | M’Tsamoudou (Mayotte) | 24 | 2011 | This study |
| #10, Saint Leu| quinquefasciatus | Saint Leu (La Réunion) | 24 | 2007 | [41] |
| #11, Etang Salé| quinquefasciatus | Etang Salé (La Réunion) | 24 | 2009 | [41] |
| #12, Saint Louis | quinquefasciatus | Saint Louis (La Réunion) | 23 | 2009 | [41] |
| #13, Saint Pierre | quinquefasciatus | Saint Pierre (La Réunion) | 24 | 2009 | [41] |
| #14, Saint Joseph | quinquefasciatus | Saint Joseph (La Réunion) | 19 | 2007 | [41] |
| #15, Plaine des Cafres | quinquefasciatus | Plaine des Cafres (La Réunion) | 24 | 2009 | [41] |
| #16, Plaine des Palmistes | quinquefasciatus | Plaine des Palmistes (La Réunion) | 22 | 2007 | [41] |
| #17, Sainte Rose | quinquefasciatus | Sainte Rose (La Réunion) | 24 | 2007 | [41] |
| #18, Saint Benoît | quinquefasciatus | Saint Benoît (La Réunion) | 24 | 2007 | [41] |
| #19, Bras Panon | quinquefasciatus | Bras Panon (La Réunion) | 32 | 2007 | [41] |
| #20, Saint André | quinquefasciatus | Saint André (La Réunion) | 24 | 2007 | [41] |
| #21, Sainte Suzanne | quinquefasciatus | Sainte Suzanne (La Réunion) | 24 | 2007 | [41] |
| #22, Sainte Marie | quinquefasciatus | Sainte Marie (La Réunion) | 24 | 2009 | [41] |
| #23, Saint Denis | quinquefasciatus | Saint Denis (La Réunion) | 24 | 2007 | [41] |
| #24, Samuel   | quinquefasciatus | Saint Denis (La Réunion) | 24 | 2010 | [41] |
| #25, La Possession | quinquefasciatus | La Possession (La Réunion) | 24 | 2007 | [41] |
| #26, Beau Bassin | quinquefasciatus | Beau Bassin – Rose Hill (Mauritius) | 42 | 2010 | This study |
| #27, Salines  | quinquefasciatus | Les Salines (Mauritius) | 7  | 2010 | This study |
| #28, Port Louis | quinquefasciatus | Port Louis (Mauritius) | 19 | 2010 | This study |
| #29, Cap Malheureux | quinquefasciatus | Cap Malheureux (Mauritius) | 23 | 2010 | This study |

*, the LR line was established from several hundred of field-caught larvae from three natural populations, i.e. Etang Salé, Saint Pierre and Saint Louis.

**, LR-TC is Wolbachia-uninfected line generated by antibiotic exposure of specimens from the LR line.

***, the LR(Pip(Is)) line combined the Is cytoplasm, including the wPip(Is) strain, and the LR nuclear genome.

****, number of field specimens examined for wPip genetic diversity.
Molecular typing

Mosquito DNA was extracted using a CetylTrimethylAmmonium Bromide (CTAB) protocol [51]. The \textit{wPip} infections were characterized through the analysis of one \textit{Wolbachia} marker, the ankyrin domains encoding gene, \textit{ank2} [52] (primers are listed in Table S1). This marker differentiated \textit{wPip} strains from groups I and IV on the basis of the size of the PCR amplified fragments: 313 bp and 511 bp fragment for group I and IV, respectively. For field samples, the \textit{ank2} PCR products from two specimens per sample site were sequenced to confirm their identity with La Réunion \textit{ank2} allele [Genbank AM397068; [43]].

The examination of the mosquito nuclear genome was assessed by PCR/RFLP tests based on \textit{Cx. pipiens ace-2} and \textit{Estere} genes (primers are in Table S1). The \textit{ace-2} gene is located on chromosome I and encodes acetylcholinesterase 2 (AChE2) [53]. The \textit{Estere} gene is located on chromosome II and encodes a carboxylester hydrolase [54]. A PCR/RFLP test on \textit{ace-2} using the \textit{ScaI} restriction enzyme \textit{(37°C, 3 hours; see [55])} allows the discrimination between the Is (two fragments: 230 and 470 bp) and the LR (three fragments: 120, 230 and 350 bp) nuclear genomes. We developed a PCR/RFLP test on \textit{Estere} using the \textit{AvaII} enzyme \textit{(37°C, 3 hours)} that also generated different restriction fragments for the Is (three fragments: 37, 519 and 544 bp) and LR (four fragments: 91, 176, 313 and 520 bp) nuclear genomes.

All PCRs were performed with ca. 20 ng of genomic DNA solution in a 40 µl final volume reaction for 35 cycles (94°C, 5 min; 94°C, 30 sec; 52°C, 30 sec; 72°C, 1 min). Direct sequencing of PCR products was performed on an ABI Prism 3130 sequencer using the BigDye Terminator Kit (Applied Biosystems) after purification with the QIAquick gel extraction kit (QIAGEN, Valencia, CA). Sequence alignment and analyses were done using MEGA software [56].

Backcrossing

The cytoplasm of the Is line, including the \textit{wPip(Is)} strain, was introduced into the LR nuclear background through eight generations of backcrossing, a procedure that should result in at least 99% genome replacement of the Is line by the LR nuclear genome. A first cross was performed using 200 virgin Is females and 250 LR-TC males. For the following generations, 200 hybrid females were backcrossed with 250 LR-TC males. Using this protocol, we obtained the LR[\textit{wPip(Is)}] line which carries the LR nuclear genome and the \textit{wPip(Is)} strain.

Crossing experiments

We examined the crossing relationships between mosquito lines through crossing experiments. Mass crosses were carried out using 35–200 two-day-old males and an equivalent number of females.
that had been individually separated at the pupal stage (age was assessed from the emergence of adults; day 0 = emergence). We also tested the effect of male aging on CI by comparing crossing relationships of young males (two-day-old) to that of older males (24-day-old). For all crosses, females were allowed to blood feed 5 days after caging. Egg-rafts were collected and stored separately until hatching at 25°C ± 2°C. Hatching rates (HR) were scored 72 h after egg-raft collection to determine the CI phenotype. All unhatched egg-rafts were checked for fertilization through observation of embryonic development following the procedure of [57].

Male performance

The longevity of the LR[wPip(Is)] and LR males was compared. We obtained males from larvae reared in standardized laboratory conditions at ca. 25°C ± 2°C. For each line, three containers containing 300 first-instar larvae with 1 L of water were set up. The water of each container was changed every two days and food provided ad libitum. Pupae were randomly sampled from the three containers to minimize possible rearing bias. Pupae were placed separately in 5 mL vials for emergence. Freshly-emerged males were kept in their vials until they died, and mortality was checked twice a day. No food was provided to the adults but they had access to the water in their tube. Survival data were fitted to the Cox proportional hazards model (coxph, survival package) [58] and a ratio for each line was estimated as their instantaneous risk of death relative to each other. These analyses were performed using R software (www.r-project.org). One posterior leg was removed on dead specimens and the tibia was measured with a micrometer (NIKON Digital Counter CM-6S).

Four cages were set up to compare the mating performance of both LR[wPip(Is)] and LR males. Each cage contained an equal number of two-day-old virgin LR females and LR males (1:1), as well as different numbers of two-day-old virgin LR[wPip(Is)] males so that different ratios of the three types of mosquitoes could be tested (1:1:0, 1:1:1, 1:1:5 and 1:1:10). Thus the total number of adults for each of these confrontations was 200, 300, 350 and 600, respectively. For each confrontation, all the mosquitoes were introduced into the cage at the same time. Females were allowed to blood feed five days after caging and their egg-rafts were collected daily to score HR. To assess the stability of the expression of CI over the mosquito lifespan, a second blood meal was given to females 15 days after the first one, and new collections of egg-rafts were then made.

Results

Only one wPip group is present in the south-western Indian Ocean islands

We first examined the genetic diversity of wPip strains found in natural populations of Cx. p. quinquefasciatus from La Réunion, Mauritius, Mayotte, Madagascar and Grande Glorieuse. The main purpose of this investigation was to assess the possibility of controlling mosquito populations in each of these four islands with wPip-infected males.

We examined 650 Cx. p. quinquefasciatus field specimens from 29 populations: La Réunion (16 populations, n = 384 individuals), Mauritius (4 populations, n = 91 individuals), Mayotte (3 populations, n = 69 individuals), Madagascar (5 populations, n = 105 individuals) and Grande Glorieuse (1 population, n = 24) (Table 1 and Figure 1). The genotyping of wPip infections in these samples was performed using only the ank2 gene which was recently shown to discriminate wPip strains into five distinct phylogenetic groups (referred as wPip-I to wPip-V) [38].

PCR assays using ank2 indicated the occurrence of wPip infection in all Cx. p. quinquefasciatus field specimens, as observed in other geographic areas for this species [10,36,37], and all shared the same ank2 allele as indicated by the length of ank2 PCR products (313 bp). This similarity was further confirmed by sequencing the ank2 gene of two individuals per population from Mauritian, Mayotte, Madagascar and Grande Glorieuse.

All sequences were found to be strictly identical to that found in the wPip strains infecting all 10 laboratory isofemale lines from La Réunion and to other wPip strains belonging to the wPip-I group [38]. This result shows that wPip strains from La Réunion, Mauritius, Mayotte, Madagascar and Grande Glorieuse are genetically closely related and are genetically different from the wPip(Is) strain belonging to the wPip-IV group.

Establishment of the LR[wPip(Is)] line

Males from the Is line belong to Cx. p. pipiens subspecies and may not be optimally adapted to the tropical environment of the Indian Ocean where Cx. p. quinquefasciatus is found. More specifically the two subspecies known to differ by behavioral and physiological characters including mating behavior [27]. To circumvent this problem, we introduced the wPip(Is) strain into the Cx. p. quinquefasciatus nuclear background from La Réunion. First a LR line was established from a large number (>5,000) of field-caught Cx. p. quinquefasciatus from three localities of La Réunion in order to have a good representation of the local genetic diversity. This line was then cured of its Wolbachia by tetracycline treatment of larvae during three generations (LR-TC line). Finally wPip(Is) from the Is line was introduced into the nuclear background of the LR-TC line by successive backcrossing. The LR[wPip(Is)] line thus created shares the same nuclear genetic background as the LR line but is infected by the wPip(Is) strain (Figure S1). This was verified by PCR/RFLP tests on ace-2 and Estere7 Cx. pipiens nuclear genes (Figure S2A and S2B) and by analyzing the allelic profiles of the ank2 gene of the infecting Wolbachia (Figure S2C).

Crossing experiments between LR[wPip(Is)] and Is lines were conducted to check that Cx. p. quinquefasciatus nuclear background has not altered the CI phenotype of the wPip(Is) strain. This aspect needs to be investigated since the host nuclear genome has been reported to affect the penetration of the CI phenotype induced by a Wolbachia strain [59–61]. Our data show that both lines behave similarly: LR[wPip(Is)] and Is showed bidirectional CI with LR while LR[wPip(Is)] and Is were mutually compatible (Table 2). The intensity of CI was very high, with 98–100% of the embryos that did not hatch in incompatible crosses. In addition, crosses between infected and uninfected lines showed unidirectional CI: males from all infected lines (LR[wPip(Is)], Is and LR ) induced complete CI (100% embryo mortality) when crossed with uninfected females (LR-TC), the reverse crosses (i.e. uninfected males and infected females) being always compatible. Overall, no significant difference of hatching rate (HR) was found when the LR[wPip(Is)] and Is lines were compared (Wilcoxon test; all P > 0.14). This shows that the CI phenotype of the wPip(Is) strain was not altered by the LR genetic background, and that the CI phenotype is controlled by the wPip infection rather than by nuclear genes, which is in accordance with most studies involving species of the Cx. pipiens complex [43,62].

The effect of male ageing on CI intensity was also tested as, in a few host species including some mosquitoes, CI intensity has been shown to decrease with male ageing [63–67]. Such an effect could impede the use of LR[wPip(Is)] males to sterilize field females. To investigate this aspect, we crossed two-day and 24-day old LR[wPip(Is)] males with two-day old LR females. No viable embryo was obtained in incompatible crosses with both young and
old LR[µPip(Is)] males (Table 3). Thus CI is expressed with the same intensity throughout the LR[µPip(Is)] males’ lifespan, a result also observed in diverse Cx. pipiens laboratory lines [10,68].

LR[µPip(Is)] males sterilize field females from islands of the SWIO

LR[µPip(Is)] males were crossed with field females from five populations: Samuel (La Réunion; n = 75 females), Salines (Mauritius; n = 37), Tsoundzou (Mayotte; n = 75); Mada (Madagascar; n = 44) and Grande Glorieuse (Grande Glorieuse; n = 97 females). All crosses were incompatible, displaying >99% embryo mortality (Table 4). This shows that LR[µPip(Is)] males express high CI intensity with field females from the four islands, as observed with females of the LR line.

Crossing relationships between LR[µPip(Is)] females and field males were also investigated to determine how the LR[µPip(Is)] line may evolve in Cx. p. quinquefasciatus field populations in the case of accidental release of LR[µPip(Is)] females. LR[µPip(Is)] females were incompatible with all males from Samuel (n = 36 males), Salines (n = 37) and Grande Glorieuse (n = 40) (Table 4). This shows that LR[µPip(Is)] expresses bidirectional CI with field specimens from these populations. However, males from Tsoundzou (n = 16) were polymorphic for their CI properties, the majority (n = 14) expressing complete CI with LR[µPip(Is)] females and a few (n = 2) being compatible (HR = 0.895±0.035) (Table 4). This shows that LR[µPip(Is)] expresses either bidirectional CI or unidirectional CI with field specimens from Tsoundzou. Thus, two crossing types coexist in Mayotte, but it is likely that the bidirectional CI crossing type is the most frequent one. Males from Mada were also polymorphic for their CI properties but, in contrast to Tsoundzou males, most Mada males were compatible with LR[µPip(Is)] females (n = 18, HR = 0.804±0.283) while only two males expressed CI. So the unidirectional CI type was the most frequent in the Mada population.

LR[µPip(Is)] and LR males show similar mating performances

Inferior competitive ability of LR[µPip(Is)] males compared with field males may limit the efficiency of an IIT program. Thus, the performances of LR[µPip(Is)] and LR males, reared in standardized conditions, were examined for different life history traits. Longevity of LR[µPip(Is)] and LR males (n = 154 and n = 238, respectively) was investigated in conditions where males had to survive by metabolizing nutritional reserves accumulated during their larval life (see material and methods) [69]. No significant difference was found (χ^2 = 0.04, P = 0.84; Figure 2), suggesting that the infection by µPip(Is) did not alter mosquito metabolism. There was also no significant difference between LR[µPip(Is)] and LR males' lifespan, a parameter known to be positively correlated with mosquitoes’ adult size and reproductive success [70]. This suggests that LR[µPip(Is)] and LR males most probably exhibit similar mating performance.

To further test this assumption, mating competition between LR[µPip(Is)] and LR males was investigated in laboratory cages. Four cages containing different ratios of LR females to LR males to LR[µPip(Is)] males (1:1:10, 1:1:1, 1:1:5 and 1:1:10) were set up. Note that as CI occurring between LR[µPip(Is)] males and LR females is complete, it was easy to distinguish egg-rafts produced from compatible (LR males×LR females) or incompatible (LR[µPip(Is)] males×LR females) crosses. Two successive collections of egg-rafts were obtained for each cage by giving females two distinct blood meals. There was no significant variation in the proportion of incompatible egg-rafts between the first and the second series of egg-rafts (Fisher exact test, all P>0.57).

As expected, when only LR males were present, all the egg-rafts were compatible (Table 5). In the other cages, no significant

### Table 2. Comparisons of CI properties of different mosquito lines.

| Females | Males | Is | LR | LR-TC |
|---------|-------|----|----|-------|
| LR[µPip(Is)] | 0.919±0.027 (2298; 14)^a | 0.967±0.010 (1405; 12)^a | 0.000±0.000 (>6000; 47)^b | 0.915±0.070 (2953; 19)^a |
| Is | 0.958±0.016 (1414; 12)^a | 0.951±0.061 (1402; 10)^a | 0.021±0.021 (>6000; 47)^b | n.d. |
| LR | 0.000±0.000 (>3500; 31)^b | 0.000±0.000 (>8000; 68)^b | 0.930±0.031 (1423; 10)^a | n.d. |
| LR-TC | 0.000±0.000 (>2000; 21)^b | 0.000±0.000 (>1200; 15)^b | 0.000±0.000 (>3500; 31)^b | 0.905±0.028 (1423; 10)^a |

For each cross, mean hatching rate ± standard error, number of eggs and egg-rafts are reported.

^a and ^b represent statistical groups (Wilcoxon two-sided test with Bonferroni’s adjustment for multiple comparisons).

This cross is the same as shown in Table 2.

### Table 3. Effect of LR[µPip(Is)] males ageing on CI phenotype.

| Crosses | Hatching rate |
|---------|---------------|
|         | 2-day old males | 24-day old males |
| LR[µPip(Is)]×LR[µPip(Is)] | 0.919±0.027 (2298; 14)^a | 0.940±0.030 (900; 8)^a |
| LR[µPip(Is)]×LR | 0.000±0.000 (>3500; 31)^b | 0.000±0.000 (>1000; 9)^b |

For each cross, mean hatching rate ± standard error, number of eggs and egg-rafts are reported.

^a and ^b represent statistical groups (Wilcoxon two-sided test with Bonferroni’s adjustment for multiple comparisons).
difference between LR[wPip(Is)] and LR males’ mating capacity was found. Indeed, the number of incompatible egg-rafts observed was not significantly different from expected values assuming an equal competitiveness of LR[wPip(Is)] and LR males and random mating (Binomial test, all $P > 0.18$; Table 5). For instance, with an identical ratio of LR[wPip(Is)] and LR males (1:1), ca. 50% of the egg-rafts produced by LR females were incompatible. When the LR[wPip(Is)] males’ ratio was higher than that of LR males, i.e. at 1:5 and at 1:10, we observed ca. five and ten times more incompatible egg-rafts than compatible ones. Taken together, these results showed that LR[wPip(Is)] males are as fit as LR males, at least in our laboratory conditions. These experiments also established that LR females cannot discriminate between compatible LR males and incompatible LR[wPip(Is)] males, a result consistent with previous observations of random mating between Cx. pipiens mosquitoes infected by incompatible Wolbachia strains [37, 48, 71].

### Discussion

The recent expansion of the Rift Valley Fever (RVF) virus [34, 35] combined with high frequencies of insecticide resistance genes in Cx. p. quinquefasciatus populations in the SWIO [72] encourage the development of new research to reduce mosquito population densities. Among these approaches, the most promising is the use of Wolbachia in an ‘Incompatible Insect Technique’ (IIT), a species-specific control approach in which inundative releases of incompatible males sterilize field Cx. p. quinquefasciatus females and possibly lead to the reduction of mosquito population densities.

The present study was undertaken to explore the feasibility of the IIT strategy on the islands of SWIO. We first acquired genetic diversity of wPip strains infecting Cx. p. quinquefasciatus mosquitoes from five islands including La Réunion, Mauritius, Mayotte, Madagascar and Grande Glorieuse. All wPip strains from these islands are genetically closely related, belonging to the wPip-I
group, which indicates that the \( \text{uPip} \) diversity is relatively low over this region. However, the variability of crossing types found in Mayotte and Madagascar shows that genetically close \( \text{Wolbachia} \) strains can exhibit distinct CI properties, as observed in some \( \text{Cx.} \text{pipiens} \) populations \[43,52\] and also in \( \text{Drosophila} \) spp. \[73\].

Next, we constructed a \( \text{Cx.} \text{p. quinquefasciatus} \) LR[\( \text{uPip(Is)} \)] line that is stably infected with the \( \text{uPip(Is)} \) strain – a \( \text{uPip} \) strain previously known to induce bi-directional CI with most La Réunion \( \text{uPip} \) strains \[43\]. Care was taken to have a nuclear genetic variability of this line as representative of La Réunion \( \text{uPip} \) strain, and that is stably infected with the \( \text{uPip(Is)} \) strain – a \( \text{uPip} \) strain previously known to induce bi-directional CI with most La Réunion \( \text{uPip} \) strains \[43\]. Care was taken to have a nuclear genetic variability of this line as representative of La Réunion \( \text{uPip} \) strain, and aging did not affect their CI properties. Finally, the sterilizing capacity of LR[\( \text{uPip(Is)} \)] males was tested with field females from each of the five islands: LR[\( \text{uPip(Is)} \)] males were incompatible with all tested field females with very high embryonic mortality (>99%). Overall, these findings demonstrate the feasibility of an IIT program using LR[\( \text{uPip(Is)} \)] males and encourage field tests for a \( \text{Cx.} \text{p. quinquefasciatus} \) elimination strategy in islands of the Indian Ocean.

The geographical isolation of the four islands is an attractive situation for developing an IIT strategy: they are at least 170 km apart from one another and more than 400 km from continental Africa. Thus natural migration is quite unlikely to occur, which should facilitate a local control approach, and minimize the reestablishment of mosquito populations. However, the variability of crossing types found in Mayotte and Madagascar shows that genetically close \( \text{Wolbachia} \) strains can exhibit distinct CI properties, as observed in some \( \text{Cx.} \text{pipiens} \) populations \[43,52\] and also in \( \text{Drosophila} \) spp. \[73\]. Another positive aspect is the small size of La Réunion \( (2,511 \text{ km}^2) \), Mauritius \( (2,040 \text{ km}^2) \), Mayotte \( (374 \text{ km}^2) \) and Grande Glorieuse \( (7 \text{ km}^2) \), facilitating an exhaustive follow-up of an IIT strategy; this will obviously not be possible on Madagascar because of its size \( (587,000 \text{ km}^2) \).

However, it must be noted that the success of an IIT strategy could be affected by the accidental release of LR[\( \text{uPip(Is)} \)] females which might lead to \( \text{uPip(Is)} \) fixation in natural populations \[1,3,75\]. An efficient sexing system producing only LR[\( \text{uPip(Is)} \)] males is thus required. Several methods including biological, genetic and transgenic methods have been developed for sex separation of insects \[20,76\]. For instance, a biological method consisting of visual separation has been used to hand-select \( \text{Cx.} \text{p. quinquefasciatus} \) males \[48\], but this method is of very limited interest in the context of the large numbers of males needed. Concerning genetic methods, a sex ratio distorter allele, linked to the dominant male-determining gene, has been described in \( \text{Cx.} \text{pipiens} \) \[77\] leading to >80% males in broods. For transgenic methods, a sexing strategy based on the use of Y-linked transgenes expressing fluorescent proteins may be considered, as shown for sexing larvae and pupae in the medfly \( \text{Ceratitis capitata} \) \[78\]. However, while distorter alleles or transgenes should maximize the production of males for releases, it remains to verify that they do not alter male fitness. An alternative method is the combination of irradiation with CI. Although several studies showed that irradiation can affect male fitness, this scheme was recently tested on \( \text{Ae. polynesiensis} \) by \[79\] who determined an irradiation dose sufficient to cause sterility of females without sterilising the males or harming their fitness.

In this paper, we present a simple diagnostic PCR based test to genotype \( \text{uPip} \) infections using the \( \text{ank2} \) marker that could be used

![Tibia size of the LR and LR[uPip(Is)] males.](image)

Thirty individuals have been measured for each line. a represents statistical group (Wilcoxon two sided-test).

doi:10.1371/journal.pntd.0001440.g003

### Table 5. Competition cages with different ratio of LR[uPip(Is)] males.

| First blood meal | Number of adults (number of LR) | Number of egg-rafts (number of eggs) | Observed frequency of infertile egg-rafts (n) | Expected frequency of infertile egg-rafts | P-value* |
|------------------|---------------------------------|------------------------------------|-----------------------------------------------|------------------------------------------|----------|
| 1:0              | 200 (100, 100, 0)               | 72 (>7500)                         | 0.00 (72)                                     | 0.00                                     | 0.99     |
| 1:1              | 300 (100, 100, 100)             | 90 (>9000)                         | 0.52 (47)                                     | 0.50                                     | 0.75     |
| 1:5              | 350 (50, 50, 250)               | 43 (>4500)                         | 0.91 (39)                                     | 0.83                                     | 0.22     |
| 1:10             | 600 (50, 50, 500)               | 45 (>4600)                         | 0.98 (44)                                     | 0.91                                     | 0.18     |

| Second blood meal | Number of adults (number of LR) | Number of egg-rafts (number of eggs) | Observed frequency of infertile egg-rafts (n) | Expected frequency of infertile egg-rafts | P-value* |
|-------------------|---------------------------------|------------------------------------|-----------------------------------------------|------------------------------------------|----------|
| 1:0               | _                              | 38 (>7500)                         | 0.00 (38)                                     | 0.00                                     | 0.99     |
| 1:1               | _                              | 42 (>9000)                         | 0.45 (19)                                     | 0.50                                     | 0.64     |
| 1:5               | _                              | 12 (>4500)                         | 0.92 (11)                                     | 0.83                                     | 0.70     |
| 1:10              | _                              | 14 (>4600)                         | 1.00 (14)                                     | 0.91                                     | 0.63     |

* comparisons between the observed and expected frequencies of infertile egg-rafts through exact binomial test.

doi:10.1371/journal.pntd.0001440.t005
to regularly monitor the accidental introduction of the \textit{aPip(Is)} strain in wild populations. Such a presence would be monitored by analysing aPip strain diversity in mixtures of larvae from natural breeding sites. In the case of the presence of field \textit{aPip(Is)}-infected individuals, LR[\textit{aPip(Is)}] male releases would have to be suspended until the elimination of \textit{aPip(Is)} individuals in the controlled area. Indeed, the bidirectional CI between LR[\textit{aPip(Is)}] line and field mosquitoes will prevent the establishment of \textit{aPip(Is)} infected individuals in these islands.

Conclusion

The study presented here supports the feasibility of an IIT strategy using the LR[\textit{aPip(Is)}] males and targeting field \textit{C. p. quinquefasciatus} populations, a species of medical and veterinary concern in the SWIO islands. This method now needs to be further tested in semi-field conditions in order to optimize several key parameters, i.e. the number of males to be released as well as the timing of releases. Recently, new semi-field cages were developed to measure the impact of the life-shortening \textit{Wolbachia} \textit{wMelPop} strain on populations of \textit{Aedes aegypti} [80]. Such cages provide a realistic transitional platform between laboratory and field conditions. The risk of accidental releases of females needs also to be limited by developing an efficient sexing method to prevent any unintentional \textit{Wolbachia} replacement.

Supporting Information

Figure S1 Backcrossing procedure. Mosquito nuclear backgrounds are indicated by colours: black represents \textit{C. p. quinquefasciatus} nuclear background (LR and LR-TC lines) and red represents \textit{C. p. pipiens} nuclear background (Is line). \textit{Wolbachia} infection types are indicated by \textit{w}-labelled symbols: black-filled symbols represent the \textit{aPip(LR)} strain and red-filled symbols the \textit{aPip(Is)} strain. Note that the LR[\textit{aPip(Is)}] line carries the LR nuclear background and the \textit{aPip(Is)} infection and could be used to produce incompatible males for field release; LR-TC is an uninfected mosquito line. M, molecular weight markers; kb, kilo bases. (TIF)

Table S1 Genes and primers of \textit{Wolbachia} and \textit{Culex pipiens}.

(DOC)

Acknowledgments

We are grateful to Arnaud Berthomieu, Patrick Makounoud and Sandra Unal for technical assistance. All sequence and morphological data were obtained on the Environmental Genomic Platform and on the Morphometry Platform of the IFR Montpellier-Environnement-Biodiversité, respectively. Contribution 2011-159 of the Institut des Sciences de l’Évolution de Montpellier (UMR 5534 CNRS — Université Montpellier 2).

Author Contributions

Conceived and designed the experiments: CMA MW OD. Performed the experiments: CMA ED OD. Analyzed the data: CMA ED OD. Contributed reagents/materials/analysis tools: CMA ED OD PT MLT Unal for technical assistance. All sequence and morphological data were

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www.plosntds.org 9 December 2011 | Volume 5 | Issue 12 | e1440

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