The role of intraoperative cell salvage for musculoskeletal sarcoma surgery

Raja Bhaskara Rajasekaran a,⇑, Antony J.R. Palmer a, Duncan Whitwell a, Thomas D.A. Cosker a, David Pigott b, Orosz Zsolt c, Robert Booth d, M.R.J.P Gibbons a, Andrew Carr e, Collaborators 1

a Nuffield Department of Orthopaedics, Rheumatology and Musculoskeletal Science (NDORMS), Nuffield Orthopaedic Centre, Windmill Road, Oxford OX3 7LD, UK
b Consultant Anaesthetist, Oxford University Hospitals NHS Foundation Trust, Oxford, UK
c Consultant Pathologist, Oxford University Hospitals NHS Foundation Trust, Oxford, UK
d Transfusion Practitioner, Oxford University Hospitals NHS Foundation Trust, Oxford, UK
e Botnar Research Centre, University of Oxford, Oxford OX3 7LD, UK

Article info

Article history:
Received 8 June 2021
Revised 4 September 2021
Accepted 6 September 2021
Available online 16 September 2021

Keywords:
Cell salvage
Transfusion
Musculoskeletal oncology
Reinfusion
Sarcoma

Abstract

Background: The efficacy and safety of cell salvage for musculoskeletal sarcoma surgery have not been reported, and concerns over re-infusion of tumour cells remain. This study aims to i) describe the intra-operative blood loss and cell salvage reinfusion volumes for lower limb sarcoma and pelvic sarcoma procedures ii) and explore whether there is evidence of tumour cells in reinfused blood.

Methods: Retrospective analysis of 109 consecutive surgical procedures for biopsy-proven sarcoma or bone metastasis performed between 1 July 2015 and 30 October 2019. Salvaged blood was processed and reinfused when intraoperative blood loss exceeded 500 ml. Primary bone tumour (n = 86(79%)) and metastasis (n = 23(21%)) constituted the study group and surgeries were classified under hemipelvectomy (n = 43(39%)), lower limb endoprostheses replacement (LLE) (n = 50(46%)) and wide excision surgery (WE) (n = 16(15%)). Microscopic examination of imprint cytology of leuco-depletion (LDF) filters, and peripheral smear examination was performed for reinfused blood.

Results: Median (IQR) intra-operative blood loss was 1750 (600–3000) ml for hemipelvectomy, 850 (600–1200) ml for LLE, and 1000 (550–2000) ml for WE. Salvaged blood was re-infused in 102 of 109 (94%) patients. The mean (SD) volume of re-infusion was 445(425) ml for hemipelvectomy, 206(131) ml for LLE, and 184(106) ml for WE. In total, 64 of 109 (59%) patients received an allogeneic red blood transfusion within 72 h of surgery. Cytology analysis of imprints taken from the filtered blood available in 95 (87%) patients and peripheral smear examination of reinfused blood available in 32(29%) patients did not reveal evidence of tumour cells on microscopic examination of any samples.

Conclusion: Our study demonstrates that musculoskeletal sarcoma surgery is associated with significant blood loss, and cell salvage permits reinfusion of autologous blood in most patients. The cytological analysis did not reveal evidence of tumour cells in reinfused blood, consistent with other studies where cell salvage is used for cancer surgery.

© 2021 The Author(s). Published by Elsevier GmbH. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).
studies have shown the use of ICS to save in urologic oncology surgery [10] and also in gynaecologic oncology [11].

The Association of Anaesthetists guidelines in 2018 stated that there is no conclusive evidence the ICS induces metastasis or affects cancer prognosis, but recommend considering the use of leucodepletion filters during re-infusion of salvaged blood in cancer surgery [12].

The aims of this study are to describe the intra-operative blood loss and cell salvage reinfusion volumes for musculoskeletal sarcoma procedures and explore whether there is evidence of tumour cells in reinfused blood.

2. Methods

We performed a retrospective cohort study of consecutive patients who underwent lower limb surgery for musculoskeletal sarcoma and received ICS for their surgery at the Nuffield Orthopaedic Centre, Oxford University Hospitals NHS Foundation Trust, Oxford, UK, between 1 July 2015 and 30 October 2019. Indication for surgery was classified as primary bone tumour or metastasis. Procedures performed were classified as hemipelvectomy, lower limb endoprosthesis (LLE) replacement, and wide excision surgery (WE). Hemipelvectomy included internal hemipelvectomy (type I, type II, and type III) and external hemipelvectomy cases; LLE included patients who needed prosthesis replacement after tumour excision or in metastatic cases/fractures; WE included patients where wide excision was performed that did not require reconstruction. Finding are reported in accordance with the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) statement. The study received approval from the local Research Ethics Committee, and the reference number granted was 6649.

All patients had their diagnosis and treatment plan finalized at the multidisciplinary team (MDT) meeting. The procedures were performed by two surgeons (DW and MG). All patients received general anaesthetics with or without neuraxial anaesthesia. Intravenous 1 g tranexamic acid was administered to all patients before skin incision. Routine thromboprophylaxis of low-molecular-weight heparin was commenced 6 h post-surgery. During the study period, there was no change in routine anaesthesia or surgical technique.

ICS was used in all musculoskeletal sarcoma surgeries from 1 July 2015, where intraoperative blood loss was expected to exceed 500 ml. The anaesthetist explained to all patients about the potential risks and benefits and a specific informed consent was obtained prior to surgery. Cell salvage was performed using Sorin XtraTM machine (LivaNova, London, UK), and the size of the bowl was selected based on the volume of salvaged blood (55–225 ml) (Fig. 1). Leucocyte depletion filter (LDF) (40 μm) was used in all cases. Wound irrigation was performed using saline. All procedures were commenced with the ICS system in ‘collect only’ mode. The salvaged blood was also then processed to re-suspend the red blood cells with a haematocrit of 60% once the estimated blood loss exceeded 500 ml.

Data were collected from local electronic health records and cell salvage databases. The collected data included: patient demographics (age, sex, BMI, ASA grade); diagnosis made at MDT; procedure performed; cell salvage records (calculated intra-operative blood loss, re-infused blood volume and haematocrit (Hct) measurements); and perioperative Hb measurements. Records were also reviewed for reinfusion hypotension in the postoperative period. Patients with incomplete record details were excluded from the study (Fig. 2). In line with national guidelines [13], the haemoglobin red blood cell transfusion threshold was 70 g/l in our hospital, except in patients with acute coronary syndrome where the threshold was 80 g/l. Transfusions of red blood cells on the day of surgery were defined as those taking place before midnight.

Estimated intra-operative blood loss was calculated according to the formula [14]:

\[
\text{Blood loss (ml)} = \text{Estimated blood volume (ml)} \times \left[ \left( \frac{\text{Hct procedure start} \% - \text{Hct pretransfusion} \%}{\text{Hct procedure start} \% + \text{Hct pretransfusion} \%} \right) \times 2 \right]
\]

Re-infusion volume was collected from the cell salvage records. The proportion of estimated blood loss re-infused was calculated as [15]:
Cell salvage efficiency (%) = \(\frac{\text{Re-infusion volume (ml) \times Re-infusion Hct (%)}}{\text{Hct procedure start (%) \times Estimated blood loss (ml)}}\).

The volume of salvaged blood was standardised to haematocrit 60%.

To analyse whether there were malignant tumour cells in the re-infused blood, touch imprint samples were taken from the plastic case on the outflow portion of the filter (Fig. 3). Imprint cytology is a well-recognised simple technique for preparation of specimen for assessment [16]. The sample which needs analysis is pressed onto a glass slide, which is then fixed and stained for assessment. Samples of blood for re-infusion were taken and two peripheral smears were prepared. Two touch imprints were taken on 2–2 slides from the central and peripheral portion of the filter. The slides were air-dried, fixed in ether-alcohol solution(1:1), and stained with haematoxylin and eosin. Microscopic examination of the slides was performed by a histopathologist (OZ).

3. Results:

A total of 111 patients of biopsy proven sarcoma of the pelvis or lower limb underwent surgery using ICS during the study period. Two patients had incomplete cell salvage data and were excluded from the study (Fig. 2). The indication for surgery was primary bone tumour in 86 cases (79%) and metastasis in 23 cases (21%). Surgery consisted of hemipelvectomy in 43 patients (39%), lower limb endoprosthesis replacement (LLE) in 50 patients (46%) and wide excision surgery (WE) in 16 patients (15%). Intraoperative blood loss was more than 500 ml in 96%(105/109) of patients. A sufficient volume of blood was salvaged for processing and...
re-infusion in 94%(102/109) of procedures. An estimated one-fourth of intra-operative blood volume loss was returned to the patient (Table 1) through ICS. In total, 64 of 109 (59%) patients received an allogenic red blood transfusion within 72 h of surgery (Fig. 4).

Cytology analysis of imprints taken from the LD filter, which were available in 95(87%) patients, and peripheral smears of re-infused bloods which were available in 32 (29%) of patients, did not reveal evidence of tumour cells on microscopic examination of any samples (Fig. 5 & Fig. 6). As per records of the 102 patients who received re-infusion, no patient was reported to have had reinfusion hypotension.

4. Discussion

Our study demonstrates that musculoskeletal sarcoma surgery involving the pelvis and lower limb is associated with significant blood loss, and ICS permits reinfusion of autologous blood in the majority of the patients. In cases where sufficient blood was processed for re-infusion, it equated to approximately two units of red blood cells per patient who underwent a hemipelvectomy and nearly one unit in LLE and WE surgeries. Almost a quarter of intraoperative blood loss was salvaged and re-infused. Cytological analysis of imprints of filtered blood did not show any evidence of malignant cells.

Our reported estimated total blood loss was comparable to previously published studies. Our median estimated total blood loss was 1750 ml for hemipelvectomy. A review of 137 pelvic resections for sarcoma demonstrated nearly 45% of patients have blood loss exceeding 3000 ml[2]. Another study of 28 pelvic resections for tumours, reported the average blood loss to be 4793 ml[1]. As per the recent recommendation by the association of anaesthetists [17], ICS should be considered for all procedures with anticipated intraoperative blood loss exceeding 500 ml of blood loss, which accounts for 96% patient in our cohort. The proportion of patients with sufficient blood salvaged for re-infusion in our cohort was higher for musculoskeletal sarcoma surgery that is reported for other orthopaedic surgeries where ICS is employed[14,17]. Studies analysing re-infusion rates following ICS in cohorts involving revision arthroplasty patients demonstrated sufficient volume for salvage in 76%[15] and 54%[18] of patients, respectively. Nearly 28% of intra-operative blood loss was re-infused after salvage in our cohort. This is lower than a similar study performed at our institution of ICS in revision hip arthroplasty patients, where an estimated 35% was salvaged for re-infusion [15]. One probable reason is difficulty containing blood loss during sarcoma surgery since the surgical fields can be larger compared with revision hip arthroplasty.

Our reported blood loss for different musculoskeletal sarcoma procedures can guide clinicians as to the estimated blood that

---

**Table 1**

|                      | Hemipelvectomy | Lower Limb Endoprosthesis replacement | Wide Excision |
|----------------------|----------------|---------------------------------------|---------------|
| **No: of Patients**  | 43 (39.4%)     | 50 (45.9%)                            | 16 (14.7%)    |
| **Age (in years)**   | 53.8 (18.4)    | 64.4 (14.6)                           | 60.5 (26.4)   |
| **Sex (%Female); %** | 32.5%          | 56%                                   | 37.5%         |
| **Body Mass Index (BMI)** | 26.9 (5.1) | 26.3 (5.7)                             | 28.4 (5.5)    |
| **Intra-operative blood loss (in ml)** | 1750 (600–3000) | 850 (600–1200)                       | 1000 (550–2000) |
| **Volume of re-infused blood (in ml)** | 445 (425) | 206 (131)                              | 184 (106)     |
| **Proportion of blood loss salvaged; %** | 30.5 (11.5) | 29.7 (12.3)                           | 23.2 (10.8)   |

* Excludes all cases where inadequate blood was available for re-infusion. Standardized to hematocrit 60%.
can be salvaged using ICS and the blood products that may be required. There has been uncertainty around the safety of ICS in cancer surgery, and debate regarding their safety still exists. Studies have demonstrated that circulating tumour cells are often present in cancer patients who undergo surgery, irrespective of ICS use. Moreover, very few of these are thought to be capable of causing metastasis [19]. The use of LDF reduces the number of malignant cells with no adverse effect on the final quality of the blood available for re-infusion [19] and recent guidelines advocate using LDF
in ICS for cancer surgery [12,18]. Our histological analysis of slides of reinfused blood from 95 patients with a range of sarcoma pathologies who had their blood filtered through an LDF, did not show any evidence of any malignant cells. This is reassuring to surgeons and clinicians regarding the use of ICS in sarcoma surgery. An observational study of flow-cytometric evaluation of filtered blood through LDF in spine tumour surgery showed the tumour cell count to be zero in 8 of the 11 samples. It demonstrated LDF to be an effective tool in removing tumour cells [20]. The safety of ICS in cancer surgery has been supported by studies of hematopoietic carcinoma surgery [21] where ICS use did not show an increased rate of metastasis or recurrence. National Institute for Health and Care Excellence (NICE) guidelines recommend using ICS in patients undergoing radical prostatectomy and cystectomy for cancer [21]. Recently, encouraging results of ICS use in surgery involving urology oncology [10] and metastasis [21,22] strongly suggest that ICS can be strongly considered in cancer surgery, including musculoskeletal tumours. Another major advantage of the use of ICS is may reduce complication associated with allogeneic transfusion [12].

Cost-effectiveness of ICS has not been considered in this study, but is improved by using the cell salvage machine in ‘collect only’ mode until adequate blood has been salvaged for reinfusion. ICS has demonstrated cost-effectiveness in high bleeding risk surgical settings, with clear economic benefits of cell salvage evident over the use of peri-operative allogeneic blood transfusion [23].

Our study has limitations. The use of ICS for the patients in our cohort was at the discretion of the surgeon and the anaesthetist, when they estimated blood loss to exceed 500 ml. The volume of blood loss and cell salvage efficiency are estimations that do not account for intra-operative fluid therapy and blood transfusions. Retrospective studies have an inherent bias of few missing data, which was also evident in our study. The surgical technique and anaesthesia technique may have influenced blood loss, which was not assessed in this study. Musculoskeletal sarcoma patients are a broad heterogeneous group, unlike many other cancers, making it challenging to draw generalised conclusions. We adopted imprint cytology of filtered blood to look for malignant cells, whereas flow cytometry has been shown to have higher sensitivity. Long-term data in larger cohorts is required to examine the rate of disease recurrence. This will be invaluable in determining whether there is a longer-term association between the use of ICS and disease recurrence.

5. Conclusion

Our study demonstrates that musculoskeletal sarcoma surgery is associated with significant blood loss and use of ICS permits reinfusion of autologous blood in most patients. Cytological analysis did not reveal evidence of tumour cells in reinfused blood, consistent with other studies where cell salvage is used for cancer surgery. Future studies should aim to analyse if any increased risk of metastasis associated with use of ICS in sarcoma surgery and also analyse the cost-effectiveness of ICS.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

[1] W. Guo, D. Li, X. Tang, Y. Yang, T. Ji, Reconstruction with modular hemipelvic prostheses for periacetabular tumor, Clin. Orthop. Relat. Res. 461 (2007) 180–188. https://doi.org/10.1097/BLO.0b013e31803f65d7.
[2] X. Tang, W. Guo, R. Yang, S. Tang, T. Ji, Evaluation of blood loss during limb salvage surgery for pelvic tumours, Int. Orthop. 33 (3) (2009) 751–756. https://doi.org/10.1007/s00264-008-0695-9.
[3] A. Kawai, H. Kadota, U. Yamaguchi, Y. Morimoto, T. Ozaki, Y. Beppu, Blood loss and transfusion associated with musculoskeletal tumor surgery, J. Surg. Oncol. 92 (1) (2005) 52–58, https://doi.org/10.1002/jso.20375.
[4] W.H. Ou, H.L. Wu, M.S. Mandell, M.Y. Tsou, K.Y. Chang, The association of allogeneic blood transfusion and the recurrence of hepatic cancer after surgical resection, Anaesthesia 75 (4) (2020) 464–471, https://doi.org/10.1111/anae.20462.
[5] T.S. Schiergens, M. Bentsch, M.S. Kasparek, K. Frenes, W.K. Jauch, W.E. Thasler, Impact of perioperative allogeneic red blood cell transfusion on recurrence and overall survival after resection of colorectal liver metastases, Dis. Colon Rectum 58 (1) (2015) 74–82, https://doi.org/10.1097/DCR.0000000000000233.
[6] A. Shah, A. Palmer, A.A. Klein, Strategies to minimize intraoperative blood loss during major surgery, Br. J. Surg. 107 (2) (2020) e26–e38, https://doi.org/10.1002/bjs.11391.
[7] S.A. Esper, J.H. Waters, Intra-operative cell salvage: a fresh look at the indications and contraindications, Blood transfusion = Transfusione del sangue 9 (2) (2011) 139–147, https://doi.org/10.2450/2011.0081-10.
[8] P.B. Yaw, M. Sentry, W.J. Link, W.M. Wahle, G.Glover, L. J. Tumor cells carried through autotransfusion: Contraindication to intraoperative blood recovery?, JAMA 231 (5) (1975) 490–491.
[9] Autologous blood transfusions. Council on Scientific Affairs. (1986). JAMA, 256, 966-970. https://doi.org/10.3909/riu0721.
[10] M.C. Ferroni, A.F. Correa, T.D. Lyon, B.J. Davies, M.C. Ost, The use of intraoperative cell salvage in urologic oncology. Rev. Urol. 19 (2) (2017) 89–96, https://doi.org/10.3939/juot118.
[11] N.P. Nagarsheth, T. Sharma, A. Shander, A. Awan, Blood salvage use in gynecologic oncology. Transfusion 49 (10) (2009) 2048–2053, https://doi.org/10.1111/j.1537-2995.2009.02256.x.
[12] A.A. Klein, C.R. Bailey, A.J. Charlton, E. Evans, M. Guckian-Fisher, R. McCrossan, A.F. Nimmo, S. Payne, K. Shreeve, J. Smith, F. Torella, Association of Anaesthetists guidelines: cell salvage for peri-operative blood conservation 2018, Anaesthesia 73 (9) (2018) 1141–1150, https://doi.org/10.1111/anae.14331.
[13] NICE guideline [NG24]. Recommendations, Blood transfusion. Guidance; NICE; 2015. Available from: https://www.nice.org.uk/guidance/ng24/chapter/Recommendations. [Accessed 13 May 2021].
[14] J.B. Gross, Estimating allowable blood loss: corrected for dilution, Anesthesiology 58 (3) (1983) 277–280, https://doi.org/10.1097/00000542-198303000-00016.
[15] A.J.R. Palmer, T.D. Lloyd, V.N. Gibbs, A. Shah, P. Dhiman, R. Booth, M.F. Murphy, A.H. Taylor, B.J.L. Kendrick, A McGill, A Alvand, A.J. Carr, The role of intra-operative cell salvage in patient blood management for revision hip arthroplasty: a prospective cohort study, Anaesthesia 75 (4) (2020) 479–486, https://doi.org/10.1095/j.anae.2019.05.111404.119499.
[16] Z. Bell, I. Cameron, J.S. Dace, Imprint cytology predicts auxillary node status in breast cancer. Expert Med. 79 (2010) 119–122.
[17] C. Carroll, F. Young, Intraoperative cell salvage, Br. J. Anaesth. 108 (1) (2012) 95–101, https://doi.org/10.1093/bja/bae326.
[18] J.P. Cata, H. Wang, V. Gottumukkala, J. Reuben, D.J. Sessler, Inflammatory response, immunosuppression, and cancer recurrence after perioperative blood transfusions, Br. J. Anaesth. 110 (5) (2013) 690–701, https://doi.org/10.1093/bja/aet068.
[19] N. Kumar, R. Lam, A.S. Zaw, R. Malhotra, J. Tan, G. Tan, T. Setiobudi, Flow cytometric evaluation of the safety of intraoperative salvaged blood filtered with leukocyte depletion filter in spine tumour surgery, Ann. Surg. Oncol. 21 (13) (2014) 4330–4335, https://doi.org/10.1245/sj0134-014-3950-9.
[20] Fabrice Muscari, Bertrand Suc, Dominique Vigouroux, Jean-Pierre Duffas, Isabelle Miquere, Anne Mathieu, Laurence Lavayssiere, Lionel Rostaing, Gilles Fourtainer, Blood salvage autotransfusion during transplantation for hepatocarcinoma: does it increase the risk of neoplastic recurrence?, Transplant Int Off. J. Eur. Soc. Organ Transplantat. 18 (11) (2005) 1236–1239, https://doi.org/10.1111/j.1398-2735.2005.00027.x.
[21] N. Kumar, R. Narkumar, J. Tan, K. Akbarry, R.S. Patel, R. Kannan. Current Status of the Use of Salvaged Blood in Metastatic Spine Tumor Surgery Neurosurgery, 15 (3) (2018) 206-215 DOI: 10.1247/sns.1836140.70.
[22] G. Lim, V. Melnyk, F.L. Facco, J.H. Waters, K.J. Smith, Cost-effectiveness Analysis of Intraoperative Cell Salvage for Obstetric Hemorrhage, Anesthesiology 128 (2) (2018) 328–337, https://doi.org/10.1097/ANES.0000000000001981.