AlnC: An extensive database of long non-coding RNAs in angiosperms

Ajeet Singh, A. T. Vivek, Shailesh Kumar

Bioinformatics Lab, National Institute of Plant Genome Research (NIPGR), Aruna Asaf Ali Marg, New Delhi, India

Abstract

Long non-coding RNAs (lncRNAs) are defined as transcripts of greater than 200 nucleotides that play a crucial role in various cellular processes such as the development, differentiation and gene regulation across all eukaryotes, including plant cells. Since the last decade, there has been a significant rise in our understanding of lncRNA molecular functions in plants, resulting in an exponential increase in lncRNA transcripts, while these went unannounced from the major Angiosperm plant species despite the availability of large-scale high throughput sequencing data in public repositories. We, therefore, developed a user-friendly, open-access web interface, AlnC (Angiosperm lncRNA Catalogue) for the exploration of lncRNAs in diverse Angiosperm plant species using recent 1000 plant (1KP) transcriptomes data. The current version of AlnC offers 10,855,598 annotated lncRNA transcripts across 682 Angiosperm plant species encompassing 809 tissues. To improve the user interface, we added features for browsing, searching, and downloading lncRNA data, interactive graphs, and an online BLAST service. Additionally, each lncRNA record is annotated with possible small open reading frames (sORFs) to facilitate the study of peptides encoded within lncRNAs. With this user-friendly interface, we anticipate that AlnC will provide a rich source of lncRNAs for small-and large-scale studies in a variety of flowering plants, as well as aid in the improvement of key characteristics in relevance to their economic importance. Database URL: http://www.nipgr.ac.in/AlnC

1. Introduction

Angiosperms are flowering plants that constitute an exceptionally large group of plants that grow in a wide variety of habitats. It comprises of more than 3,000,000 recorded species worldwide, encompassing one of the most diverse group within the plant kingdom [1, 2]. Most angiosperms are a main source of consumer goods like textile fibres, herbs and spices, fuel and pharmaceuticals, as well as a major source of food. Most model plants belonging to this group have been intensively studied to understand flowering and other major mechanisms. Consequently, research in Angiosperms exploded with the advent of next generation sequencing (NGS) resulting in an improved picture of transcriptome, especially from the point of non-coding RNAs (ncRNAs). In recent years, IncRNAs are a major RNA class of greater research
interest to study alongside miRNAs. LncRNAs are typically more than 200 nucleotides with no or less protein coding capacity, as several recent studies suggest the presence of sORFs with a potential for translating into micropeptides [3–5]. Further, there is compelling evidence that lncRNAs role in multiple plant biochemical pathways in recent years [6–8]. With the rise of transcriptome data in public repositories, thousands of plant lncRNAs were identified and maintained in a number of lncRNA dedicated databases in the last decade [9–14]. However, despite being in the spotlight, lncRNAs still need to be annotated in a variety of plant species, given the availability of lncRNA databases focused primarily on model plants and major crops. As information on lncRNA in many Angiosperms is still scarce, the advancement of lncRNA research in these plants is largely hindered. This research gap can be potentially addressed by the use of large volumes of RNA sequencing (RNA-seq) data available in public databases as it provides enormous opportunities to discover and classify potential lncRNAs [15, 16]. Various lncRNA investigations in plant species were undertaken using independent bioinformatics pipelines for genome-wide annotation and further, archiving in databases, but several plants are still unexplored due to a complete lack of complete genome sequences [15, 17, 18]. In order to fill this potential gap, various lncRNA methods are available and are increasingly being developed to improve lncRNA identification and annotation from de novo assembled transcripts [19, 20].

In this post-genomics era/advanced genomics era, it is crucial to develop comprehensive methods for lncRNA annotation across various plant species to improve our understanding of the complex interplay of lncRNAs across plants. More importantly, the creation and maintenance of a stable lncRNA data repository is equally necessary if lncRNA biology is to be understood [17, 21]. In this research work, we use large-scale transcriptome data to identify potential lncRNAs with three major goals. First, we anticipate to provide information on most potential lncRNAs of plant species with no available genome sequence. The second goal is to mediate the importance of unused large-scale datasets as an annotation source for thousands of lncRNAs. Last, to develop a database that can act as a catalyst to promote lncRNA research in angiosperms and to provide a one-stop data access platform. Here, we capitalised on large-scale transcriptomic data of 682 angiosperms from the 1000 plants (1KP) project as this enabled us to annotate 1,08,55,598 lncRNAs. The results of the study were organised and stored in a user-friendly web-interface, the AlnC) with a plan to update periodically on the basis of new knowledge and an expansion in the number of species of Angiosperm in the future.

2. Materials and methods

Data collection and systematic lncRNA identification

To find potential lncRNAs, we used de novo assembled transcripts of 682 Angiosperm plant species from the 1KP project (http://www.onekp.com/public_data.html) [22]. For each species, lncRNAs were identified from each sample relying on a bioinformatics pipeline previously exploited by Singh et al., 2017, and transcripts longer than 200 bp were retained that are not overlapping with protein-coding gene models (Fig 1(A)) [23]. First, we excluded potential coding transcripts from the assembled transcripts set for that species (translated proteins of matched orthogroups derived from annotated plant genomes) [24]. Further, protein-coding transcripts were discarded using PLncPRO (python prediction.py -p plncpro -result-file -i sequence.fa -m models/<monocot or dicot>.model -o plncpro-out -d lib/blastdb/swissprotDB -t 15 -r) on the basis of a BLAST approach to a manually curated list of Swissprot proteins [25]. In the final filtering step, high-confident lncRNA transcripts were extracted by setting a minimum length threshold of 200 nt length and a non-coding probability score of 0.8 (python predstoseq.py -f sequence.fa -o output-file -p plncpro-result-file -l 0 -s 0.8—min 200).
Fig 1. Overview of lncRNAs in AlnC. (A) Systematic workflow adopted to annotate potential lncRNAs of flowering plants available from the 1KP project. (B) Lengthwise distribution of AlnC annotated lncRNAs across clades. (C) Pie chart represents the percentage of lncRNA entries in AlnC. (D) Percentage composition of AU content in protein-coding transcripts and lncRNAs in AlnC.

https://doi.org/10.1371/journal.pone.0247215.g001
Database construction and implementation

AlnC is operating on a Linux, Apache, MySQL and PHP stack as of now. The current database framework is built on the Apache server and the AlnC web interface has been designed using HTML, CSS, and JavaScript. All AlnC IncRNAs and other related data annotations is handled by a relational database set up with MySQL. In addition, AlnC is integrated with stand-alone BLAST (v2.11.0) for online similarity search, ViennaRNA (v2.4.16) for the visualisation of secondary structure and ORFfinder (v0.4.3) for the exploration of IncRNA containing sORFs [26–28].

3. Results and discussion

Data content in AlnC

We organised and compiled a collection of 1,08,55,598 lncRNAs from 809 samples available in the 1 KP project, which functions as a comprehensive IncRNA catalogue of 682 flowering plants stored in the AlnC platform ('Statistics' section of webpage). No other data repositories on IncRNAs on this scale exist, and most IncRNAs of the species included in AlnC belong to poorly studied taxa, rendering AlnC of wide interest among plant researchers (Fig 1(B) and 1(C)). In AlnC, the newly identified IncRNAs of flowering plants across clades ranged in size from 200 to 7633 nt with an average length of 405 nt (Fig 1(B)). We found the median length of identified IncRNAs is smaller than the median length of the coding sequences (Figs 1(B) and 4(E)). Moreover, most IncRNAs (66%) were less than 400 bp in length whereas only 2.9% IncRNAs were more than 1000 bp in length. We found the AU content of AlnC IncRNAs varied from 50–90% with an average of 75% in comparison to the coding transcripts which ranged 30–80% with an average of 60% (Fig 1(D)). Most IncRNAs contained more than 75% AU content, and the analysis implies the richness of AU than that in coding sequences [29–31].

Fig 1 and Table 1 provide a brief description of IncRNA entries in AlnC. Our attempts to identify ortholog relationships using BLASTn search (>70% identity, ± 50nt alignment length of subject/query IncRNA sequence, and e-value cutoff 0.01) resulted in significant hits, however, we could only identify a moderate number of AlnC annotated IncRNAs identical to those stored in two major databases, NONCODE (v6.0) and PLncDB (v2.0) (Fig 2) [13, 32]. This could be apparent differences in the number of IncRNAs in these database, but it also suggests certain conservation across species. The list of significant hits to NONCODE IncRNAs (384 hits), and experimentally validated IncRNAs available in Plncdb (6 hits) is tabulated in S1 Table. Further, we recognised the relationship between 1KP total transcripts, coding transcripts and AlnC annotated IncRNAs across clades using ternary graphs on the basis of size attributes to each axis separately (1 KP total transcripts, coding transcripts and lncRNAs in Fig 3A–3C, respectively). The ternary plot of AlnC annotated IncRNAs to 1 KP assembled transcripts and protein-coding transcripts enabled us to identify two distinct clusters of monocot and dicots, respectively (Fig 3). We observed proportional discovery rate of lncRNAs with respect to the 1KP assembled transcripts and also found out that the ratio of total transcripts to coding transcripts is smaller in dicot species whereas the reverse trend in case of monocots.

AlnC query and search platform

Search options. Current release of AlnC provides two query interfaces—(a) Simple Search, and (b) Advanced Search (Fig 4(A)). ‘Simple search’ allows users to perform quick searches for IncRNAs based on taxonomic rank (Clade, Order, Family, Species) and non-coding probability score (min: 0.8; max: 1.0) while ‘Advanced search’ provides enhanced query functionality using logical operators (AND/OR/\geq/\leq). Consequently, a list of potential
In lncRNAs will be displayed in the result page with the options to download and save search results. The search results display the entries of the chosen species covering the basic meta details of the 1KP sample code with links describing the sample information, including sample preparation, sample supplier, sample extractor and tissue type, NCBI ID showing experimental sample library and run data, source transcript ID from which lncRNA is annotated, lncRNA length and non-coding probability score (Fig 4(B)). The users can take advantage of AlnC ID link to view and navigate to a detailed record of a lncRNA providing primary lncRNA features, secondary structure and other accessory information.

**InRNA details webpage.** Each ‘lncRNA details’ page of AlnC enables access to lncRNA and its structure information (Fig 4(C)). This detail webpage is divided into two parts: the first section contains basic details of lncRNA sequence (including species name of annotated lncRNA, source transcript ID, length, GC content, and coding/non-probability), and particulars of secondary structure in dot-bracket notation with a provision for the user to view and download the structure; second section displays ORFs and conceptual translation products of lncRNA sequence. Further, these peptides can be checked for possible functional activity with the BLAST option to the PlantPepDB database (contains 3,848 unique entries categorized into 9 major functional categories) [33]. All the associated annotations of a lncRNA entry can be downloaded in a tabular form.

**BLAST module.** BLAST feature was incorporated into AlnC web user interface to find regions of similarity between the user input and AlnC lncRNAs using BLASTn option. The default BLAST search enables searching for lncRNA transcript models of all angiosperm
species in AlnC, and besides searches can also be limited by specifying species/clade/order/family and/or e-value cut-offs. The BLAST search output includes pairwise alignment, a report with BLAST hits based upon alignment scores and other measures of statistical significance.

**LncRNA vs mRNA module.** This module allows the interactive visualisation and comparison of the length-wise distribution as well as the AU percent of the annotated IncRNA transcripts relative to the protein-coding transcripts in multiple samples for each species (Fig 4(D) and 4(E)). This webpage also enables users with options to download these comparison plots in multiple image file formats.

**Download AlnC data.** Sequences can be searched and downloaded from the AlnC archive, and the full AlnC collection can also be found on the download page. The download webpage in AlnC provides access to both the hierarchical bulk download and the species-wise download in FASTA file format.
Fig 4. Screenshots of AlnC web interface. (A) Interface of simple search and advanced search modules. (B) Results page showing a table view of all lncRNA entries of the plant species *Aextoxicon punctatum*. (C) Sequence features including primary sequence information, secondary structure and possible sORFs as well peptides are displayed for lncRNA entry AlnC_6143890. (D) Percentage composition of AU content IncRNAs relative to mRNAs. (E) Pie chart representation of length-wise distribution of *Aextoxicon punctatum* IncRNAs and mRNAs.

https://doi.org/10.1371/journal.pone.0247215.g004
As there are sizeable researchers working on several flowering plants, we created a user form to submit any information or data with regard to angiosperm species lncRNAs. This will allow us to develop, upgrade and manage AlnC on a regular basis. The related lncRNA data, research results and publications can be submitted using the form provided on the 'submit data' section of webpage. If found to be relevant, the received data will be curated manually and appended to AlnC. All submitted lncRNA data will be processed in compliance with the AlnC standard pipeline mentioned in Fig 1(A) and manually curated. We encourage submissions to AlnC curators as this will drive our plans to include additional species in the future.

### Conclusion and future prospects

In this research work, we used transcriptome data from 682 flowering plants, most of which had no genomic information and/or no documented lncRNA studies prior to this work. This, in our view, is a key feature of AlnC as it was developed with the primary goal of facilitating lncRNA studies in various Angiosperm organisms. We used an analysis workflow tailored for plants, but it could also be used for RNA-seq-based lncRNA identification in non-plants species. The workflow provides a machine learning (ML)-based bioinformatics pipeline for identifying high-confidence lncRNAs across Angiosperm organisms, which differs from other annotation methods used in already available lncRNA databases (such as CATATAdb) [10]. This method produced a large number of putative lncRNA transcripts, which were then organised and catalogued in the AlnC web interface. AlnC covers information from 1,08,55,598 lncRNA loci derived from 809 samples and provides a user-friendly platform for browsing, searching and accessing all annotated lncRNAs via simple and interactive web pages. AlnC includes lncRNAs with evidence of non-coding RNA probability score and allows further exploration of sORFs alongside other primary lncRNA features, thus providing researchers with functional capability to leverage AlnC data and information on their individual projects through our web interface. With this research work, we attempted to develop a first-ever database covering the largest number of confident lncRNA entries of wide-range plant species, including those with no information on lncRNA of any sort. Although it is clear that several plants belonging to Angiosperms are still to be discovered and transcriptomes are waiting to be studied by individual research groups, AlnC will continue to be updated. At the same time, AlnC will strive to periodically search freely accessible databases, and other forms of documentation to gather useful information for annotated lncRNAs, and add additional functionality to enhance user engagement. It is our intention that AlnC will move forward to provide new

### Table 1. Summary of annotated lncRNAs across higher-level clades in AlnC database.

| Clade                        | Number of Species | Number of Samples | Number of lncRNAs |
|------------------------------|-------------------|-------------------|-------------------|
| Basal Eudicots               | 33                | 55                | 835185            |
| Basalmost angiosperms        | 8                 | 8                 | 281514            |
| Chloranthales                | 2                 | 2                 | 46074             |
| Core Eudicots                | 95                | 116               | 1828555           |
| Core Eudicots/Asterids       | 217               | 242               | 3656812           |
| Core Eudicots/Rosids         | 201               | 250               | 2627218           |
| Magnoliids                   | 26                | 27                | 383010            |
| Monocots                     | 61                | 64                | 829121            |
| Monocots/Commelinids         | 39                | 45                | 568109            |
| Total                        | 682               | 809               | 10855598          |

https://doi.org/10.1371/journal.pone.0247215.t001
databases representing additional species as well as to fine-tune and optimise the annotations currently available. We will also aim to focus on the new pipeline for the development of lncRNA annotations as the lncRNA biology research progresses. All in all, AlnC will strive to continue to be in line with the lncRNA community, and remain to serve useful lncRNA data in future.

Supporting information

S1 Table. List of significant hits of AlnC annotated lncRNAs to lncRNAs available at NON-CODE and PlncDB databases.

(XLSX)

Acknowledgments

The authors are thankful to the Department of Biotechnology (DBT)-eLibrary Consortium, India, for providing access to e-resources. A.S and A.T.V are thankful to the Council of Scientific and Industrial Research and the DBT for research fellowships, respectively. Also, the authors acknowledge the Bioinformatics Center at the National Institute of Plant Genome Research (NIPGR).

Author Contributions

Conceptualization: Shailesh Kumar.
Data curation: Ajeet Singh, A. T. Vivek.
Methodology: Shailesh Kumar.
Project administration: Shailesh Kumar.
Resources: Shailesh Kumar.
Supervision: Shailesh Kumar.
Writing – original draft: Ajeet Singh.
Writing – review & editing: A. T. Vivek, Shailesh Kumar.

References

1. Thorne RF. How many species of seed plants are there? Taxon. 2002; 51: 511–512. https://doi.org/10.2307/1554864
2. Christenhusz MJM, Byng JW. The number of known plants species in the world and its annual increase. Phytotaxa. 2016; 261: 201–217. https://doi.org/10.11646/phytotaxa.261.3.1
3. Cech TR, Steitz JA. The noncoding RNA revolution—Trashing old rules to forge new ones. Cell. Cell Press; 2014. pp. 77–94. https://doi.org/10.1016/j.cell.2014.03.008 PMID: 24679528
4. Nelson BR, Makarewicz CA,Anderson DM, Winders BR, Troupes CD, Wu F, et al. A peptide encoded by a transcript annotated as long noncoding RNA enhances SERCA activity in muscle. Science (80-.). 2016; 351: 271–275. https://doi.org/10.1126/science.aad4076 PMID: 26816378
5. Kopp F, Mendell JT. Functional Classification and Experimental Dissection of Long Noncoding RNAs. Cell. Cell Press; 2018. pp. 393–407. https://doi.org/10.1016/j.cell.2018.01.011 PMID: 29373828
6. Chen L, Zhu QH, Kaufmann K. Long non-coding RNAs in plants: emerging modulators of gene activity in development and stress responses. Planta. Springer Science and Business Media Deutschland GmbH; 2020. p. 92. https://doi.org/10.1007/s00425-020-03480-5 PMID: 33099688
7. Chekanova JA. Long non-coding RNAs and their functions in plants. Current Opinion in Plant Biology. Elsevier Ltd; 2015. pp. 207–216. https://doi.org/10.1016/j.pbi.2015.08.003 PMID: 26342908
8. Yu Y, Zhang Y, Chen X, Chen Y. Plant noncoding RNAs: Hidden players in development and stress responses. Annual Review of Cell and Developmental Biology. Annual Reviews Inc.; 2019. pp. 407–431. https://doi.org/10.1146/annurev-cellbio-100818-125218 PMID: 31403819

9. Xuan H, Zhang L, Liu X, Han G, Li J, Li X, et al. PLNIncRbase: A resource for experimentally identified IncRNAs in plants. Gene. 2015; 573: 328–332. https://doi.org/10.1016/j.gene.2015.07.069 PMID: 26211629

10. Szczęśniak MW, Bryzghalov O, Ciomborowska-Basheer J, Makałowska I. CANTA TAdb 2.0: Expanding the Collection of Plant Long Noncoding RNAs. Methods in Molecular Biology. Humana Press Inc.; 2019. pp. 415–429. https://doi.org/10.1007/978-1-4939-9045-0_26 PMID: 30945201

11. Xuan H, Zhang L, Liu X, Han G, Li J, Li X, et al. PLNIncRbase: A resource for experimentally identified IncRNAs in plants. Gene. 2015; 573: 328–332. https://doi.org/10.1016/j.gene.2015.07.069 PMID: 26211629

12. Gallart AP, Pulido AH, De Lagnaren IAM, Sanseverino W, Cigliano RA. GREENC: A Wiki-based database of plant IncRNAs. Nucleic Acids Res. 2016; 44: D1161–D1166. https://doi.org/10.1093/nar/gkv1215 PMID: 26578586

13. Jin J, Lu P, Xu Y, Li Z, Yu S, Liu J, et al. PLncDB V2.0: a comprehensive encyclopedia of plant long non-coding RNAs. Nucleic Acids Res. 2020 [cited 10 Dec 2020]. https://doi.org/10.1093/nar/gkaa910 PMID: 33079992

14. Cheng Quek X, Thomson DW, Maag JL V, Bartonicek N, Signal B, Clark MB, et al. lncRNA db v2.0: expanding the reference database for functional long noncoding RNAs. Nucleic Acids Res. 2015; 43. https://doi.org/10.1093/nar/gku988 PMID: 25332994

15. Cao H, Wahlestedt C, Kapranov P. Strategies to Annotate and Characterize Long Noncoding RNAs: Advantages and Pitfalls. Trends in Genetics. Elsevier Ltd; 2018. pp. 704–721. https://doi.org/10.1016/j.tig.2018.06.002 PMID: 30017313

16. Mattick JS, Rinn JL. Discovery and annotation of long noncoding RNAs. Nature Structural and Molecular Biology. Nature Publishing Group; 2015. pp. 5–7. https://doi.org/10.1038/nsmb.2942 PMID: 30945201

17. Vivek AT, Kumar S. Computational methods for annotation of plant regulatory non-coding RNAs using RNA-seq. Brief Bioinform. 2020; 2020: 1–24. https://doi.org/10.1093/bib/bbaa322 PMID: 33333550

18. Budak H, Kaya SB, Cagirici HB. Long Non-coding RNA in Plants in the Era of Reference Sequences. Frontiers in Plant Science. Frontiers Media S.A.; 2020. p. 276. https://doi.org/10.3389/fpls.2020.00276 PMID: 3226437

19. Ayachit G, Shaikh I, Sharma P, Jani B, Shukla L, Sharma P, et al. De novo transcriptome of Gymnema sylvestre identified putative IncRNA and genes regulating terpenoid biosynthesis pathway. Sci Rep. 2019; 9: 1–13. https://doi.org/10.1038/s41598-018-37186-2 PMID: 30626917

20. Li A, Zhang J, Zhou Z. PLEK: A tool for predicting long non-coding RNAs and messenger RNAs based on an improved k-mer scheme. BMC Bioinformatics. 2014; 15: 311. https://doi.org/10.1186/1471-2105-15-311 PMID: 25239026

21. Yotsukura S, du Verle D, Hancock T, Natsume-Kitatani Y, Mamitsuka H. Computational recognition for long non-coding RNA (IncRNA): Software and databases. Brief Bioinform. 2017; 18: 9–27. https://doi.org/10.1093/bib/bbv114 PMID: 26839320

22. Carpenter EJ, Matasci N, Ayyampalayam S, Wu S, Sun J, Yu J, et al. Access to RNA-sequencing data from 1,173 plant species: The 1KP. Gigascience. 2019; 8: 1–7. https://doi.org/10.1093/gigascience/giz126 PMID: 31644802

23. Singh U, Khemka N, Singh Rajkumar M, Garg R, Jain M. PLncPRO for prediction of long non-coding RNAs (IncRNAs) in plants and its application for discovery of abiotic stress-responsive IncRNAs in rice and chickpea. Nucleic Acids Res. 2017; 45: 183. https://doi.org/10.1093/nar/gxk666 PMID: 29036354

24. Wickett NJ, Mirarb S, Nguyen N, Warnow T, Carpenter E, Matasci N, et al. Phylotranscriptomic analysis of the origin and early diversification of land plants. Proc Natl Acad Sci U S A. 2014; 111: E4859–E4868. https://doi.org/10.1073/pnas.1323926111 PMID: 25355905

25. Consortium TU. UniProt: a worldwide hub of protein knowledge. Nucleic Acids Res. 2019; 47: D506–D515. https://doi.org/10.1093/nar/gky1049 PMID: 30395287

26. Rombel IT, Sykes KF, Rayner S, Johnston SA. ORF-FINDER: A vector for high-throughput gene identification. Gene. 2002; 282: 33–41. https://doi.org/10.1016/s0378-1119(01)00819-8 PMID: 11814675

27. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. J Mol Biol. 1990; 215: 403–410. https://doi.org/10.1016/S0022-2836(05)80360-2 PMID: 2231712

28. Lorenz R, Bernhart SH, Höner zu Siederdissen C, Tafer H, Flammini C, Stadler PF, et al. ViennaRNA Package 2.0. Algorithms Mol Biol. 2011; 6: 26. https://doi.org/10.1186/1748-7188-6-26 PMID: 22115189
29. Liu J, Jung C, Xu J, Wang H, Deng S, Bernad L, et al. Genome-wide analysis uncovers regulation of long intergenic noncoding RNAs in arabidopsis. Plant Cell. 2012; 24: 4333–4345. https://doi.org/10.1105/tpc.112.102855 PMID: 23136377

30. Zhang W, Han Z, Guo Q, Liu Y, Zheng Y, Wu F, et al. Identification of Maize Long Non-Coding RNAs Responsive to Drought Stress. Scaria V, editor. PLoS One. 2014; 9: e98958. https://doi.org/10.1371/journal.pone.0098958 PMID: 24892290

31. Zhang YC, Liao JY, Li ZY, Yu Y, Zhang JP, Li QF, et al. Genome-wide screening and functional analysis identify a large number of long noncoding RNAs involved in the sexual reproduction of rice. Genome Biol. 2014; 15: 512. https://doi.org/10.1186/s13059-014-0512-1 PMID: 25517485

32. Zhao Y, Li H, Fang S, Kang Y, Wu W, Hao Y, et al. NONCODE 2016: An informative and valuable data source of long non-coding RNAs. Nucleic Acids Res. 2016; 44: D203–D208. https://doi.org/10.1093/nar/gkv1252 PMID: 26586799

33. Das D, Jaiswal M, Khan FN, Ahamad S, Kumar S. PlantPepDB: A manually curated plant peptide database. Sci Rep. 2020; 10: 1–8. https://doi.org/10.1038/s41598-019-56847-4 PMID: 31913322