Scrub Typhus Involving Central Nervous System, India, 2004–2006

To the Editor: Scrub typhus, caused by Orientia tsutsugamushi, is one of the most common infectious diseases of rural southern Asia, south-eastern Asia, and the western Pacific. The disease is transmitted to humans by the bite of larvae of trombiculid mites harboring the pathogen. The disease often appears as a nonspecific febrile illness. The clinical picture of scrub typhus is typically associated with fever, rash, myalgia, and diffuse lymphadenopathy (1). Immunofluorescence assay (IFA) is the test of choice for serodiagnosis of rickettsial diseases (2). Scrub typhus has been reported from northern, eastern, and southern India, and its presence has been documented in at least 11 Indian states (3–7).

Our study’s goal was to retrospectively analyze data of patients with scrub typhus involving the central nervous system. Scrub typhus was suspected on the basis of clinical signs such as febrile illness or fever with rash or eschar. The fever workup profile (Widal agglutination test, peripheral smear, blood, and urine culture) was noncontributory. Blood samples were obtained after patients gave informed consent. All patients with clinically suspected scrub typhus received azithromycin or doxycycline.

Table. Clinical features and laboratory investigations of patients who had scrub typhus with central nervous system involvement, India, 2004–2006a

| Clinical and laboratory features | Patient 1 | Patient 2 | Patient 3 | Patient 4 |
|----------------------------------|-----------|-----------|-----------|-----------|
| Age, y/sex                       | 52/M      | 50/F      | 30/F      | 47/F      |
| Date of hospital admission       | 2004 Aug 22 | 2004 Aug 30 | 2005 Sep 9 | 2005 Sep 15 |
| Fever duration before admission, d† | 12       | 12        | 9         | 16        |
| Chills                           | +         | +         | +         | _         |
| Chills                           | +         | +         | +         | _         |
| Headache                         | +         | +         | +         | _         |
| Myalgia                          | +         | +         | +         | _         |
| Abdominal pain                   | +         | +         | +         | _         |
| Seizure                          | _         | +         | +         | _         |
| Altered sensorium                | +         | +         | +         | _         |
| Conjunctival suffusion           | +         | _         | +         | +         |
| Jaundice                         | +         | +         | +         | _         |
| Eschar                           | _         | + Axilla  | +         | _         |
| Lymphadenopathy                  | + Generalized | + Cervical | + Cervical | _         |
| Meningeal signs                  | +         | +         | +         | +         |
| Urea, mg/dL                      | 104       | 96        | 84        | 143       |
| Creatinine, mg/dL                | 3.9       | 1.5       | 1.6       | 2.7       |
| Bilirubin, mg/dL                 |           |           |           |           |
| Total                            | 3.5       | 2.7       | 4.6       | 3.0       |
| Conjugated                       | 1.0       | 1.7       | 3.6       | 2.6       |
| Aspartate aminotransferase, IU   | 167       | 160       | 166       | 30        |
| Alanine aminotransferase, IU     | 139       | 198       | 185       | 38        |
| Alkaline phosphatase, IU         | 80        | 1,000     | 1,000     | 782       |
| Proteinuria, mg/dL               | +         | +         | +         | +         |
| CSF cytology                     |           |           |           |           |
| Lymphocytes, 54 cells/mm³        | 125       | 69        | 34        | 118       |
| Lymphocytes, 14 cells/mm³        | 34        | 44        | 33        | 48        |
| Lymphocytes, 68 cells/mm³        | <64       | <64       | Not done  | Not done  |
| Neutrophils, 38 cells/mm³        |           |           |           |           |
| Proteinuria, mg/dL               |           |           |           |           |
| Glucose, mg/dL                   |           |           |           |           |
| IFA titer§                       |           |           |           |           |
| IgG                              | 512       | 512       | Not done  | Not done  |
| IgM                              | <64       | <64       |           |           |
| Drug treatment                   | Azithromycin, doxycycline | Ceftriaxone, doxycycline | Ceftriaxone, doxycycline | Ceftriaxone, doxycycline |
| Outcome                          | Died      | Improved  | Improved  | Left hospital against advice |

aCSF, cerebrospinal fluid; IFA, immunofluorescence assay; Ig, immunoglobulin.
†Fever defined as AM temperature >98.9°F or PM temperature >99.9°F.
‡IFA significant titer: IgG >128; IgM >64. The single serum samples, obtained at admission to hospital, were subjected to IFA by using a panel of 11 rickettsial antigens comprising spotted fever group rickettsiae (Rickettsia japonica, R. helvetica, R. slovaca, R. conorii subsp. indica, R. honei, R. helongjiangensis, and R. felis), R. typhi, and Orientia tsutsugamushi (Gilliam, Kato, and Kawasaki strains).

Emerging Infectious Diseases • www.cdc.gov/eid • Vol. 16, No. 10, October 2010 1641
or azithromycin) empirically. IFA and PCR of blood samples were performed to confirm scrub typhus following standard protocol (3). DNA was extracted from the blood sample (buffy coat) by using QIAamp DNA Mini Kit (QIAGEN GmbH, Hilden, Germany) according to the manufacturer’s instructions. A standard PCR specific for the 56-kDa protein with forward and reverse primers (OtsuF: 5′-AATTGCTAGTGCAATGTCTG-3′ and OtsuR: 5′-GGCATTATAGTGCTGCTG-3′) was performed (3). PCR products were purified by using the QIAquick PCR Purification Kit (QIAGEN) according to the manufacturer’s instructions. Sequencing reactions were done by using a DNA sequencing kit, dRhodamine Terminator Cycle Sequencing Ready Reaction Mix (Applied Biosystems). The obtained sequences were identified by comparison with sequences available in GenBank by using the BLAST software (http://blast.ncbi.nlm.nih.gov) (3).

During 2004–2006, scrub typhus was confirmed in 27 patients; 4 had features of central nervous system involvement. All 4 had fever with altered sensorium and meningeal signs; 2 had seizures. No neurologic focal deficit was noted, but all showed cerebrospinal fluid abnormalities. One patient had an eschar, but none had a rash. Serum of 2 patients was subjected to IFA; both samples showed high titers (Table), and PCR for blood was positive for O. tsutsugamushi for all patients. Serum was not subjected to examination for leptospirosis. Patients were treated mainly on the basis of clinical grounds because results of serology were not available immediately. Some clinical features of scrub typhus and leptospirosis are similar, and dual infections have been reported (8); therefore, antimicrobial drugs active against both leptospirosis and scrub typhus were included in treatment regimens. One patient received doxycycline and azithromycin, and the remaining 3 received ceftriaxone in addition to doxycycline. Two patients improved, 1 died, and 1 left hospital against medical advice. The clinical and laboratory details, treatments, and outcomes of all patients are given in the Table.

O. tsutsugamushi is an obligate intracellular parasite of professional and nonprofessional phagocytes that invades the central nervous system as part of systemic infection and is found in endothelial cells of blood vessels and in circulating phagocytes. A severe headache occurs almost invariably and has been used as a key clinical criterion for identifying suspected cases. Severe features of central nervous system involvement, such as neck stiffness, neurologic weakness, seizures, delirium, and coma, have been reported. Meningismus or meningitis has been found in 5.7%–13.5% of patients (9). The greatest degree of central nervous system involvement in rickettsial diseases occurs in Rocky Mountain spotted fever and epidemic typhus, followed closely by scrub typhus. The meninges are more commonly involved by O. tsutsugamushi than by other rickettsial infections, and the overall histologic picture in the central nervous system is best described as a meningoencephalitis (9). An exhaustive study of 200 cases of scrub typhus showed central nervous system involvement in most patients. However, focal central nervous system damage was rare, and during the encephalitis stage, few objective neurologic signs were apparent, other than those suggesting more generalized cerebral involvement, such as confusion, tremor, and restlessnes (10).

Now that it is established that O. tsutsugamushi does invade cerebrospinal fluid, scrub typhus should be considered a cause of mononuclear meningitis in areas in which it is endemic. In our study 1 patient died despite treatment with doxycycline and azithromycin, suggesting the possibility of resistance to these antimicrobial drugs as recently posited in a study conducted in southern India (6). Scrub typhus in these regions should be further investigated in prospective studies, and clinical isolates should be obtained to evaluate susceptibility to antimicrobial drugs.

Sanjay K. Mahajan, Jean-Marc Rolain, Anil Kanga, and Didier Raoult

Author affiliations: Indira Gandhi Medical College, Shimla, India (S.K. Mahajan, A. Kanga); and Université de la Méditerranée, Marseille, France (J.-M. Rolain, D. Raoult)

DOI: 10.3201/eid1610.100456

References

1. Raoult D. Scrub typhus. In: Mandell GL, Bennet JE, Dolin R, editors. Principles and practice of infectious diseases. 6th ed. Philadelphia: Churchill Livingstone; 2004. p. 230–10.
2. Blacksell SD, Bryant NJ, Paris DH, Doust AJ, Sakoda Y, Day NPJ. Scrub typhus serologic testing with indirect immunofluorescence method as a diagnostic gold standard: a lack of consensus leads to lot of confusion. Clin Infect Dis. 2007;44:391–401. DOI: 10.1086/510585
3. Mahajan SK, Rolain JM, Kashyap R, Bakshe D, Sharma V, Prasher BS, et al. Scrub typhus in Himalayas. Emerg Infect Dis. 2006;12:1590–2.
4. Mahajan SK, Kashyap R, Kanga A, Sharma V, Prasher BS, Pal LS. Relevance of Weil-Felix test in diagnosis of scrub typhus in India. J Assoc Physicians India. 2006;54:619–21.
5. Chaudhry D, Garg A, Singh L, Tandon C, Saini R. Rickettsial diseases in Haryana: not an uncommon entity. J Assoc Physicians India. 2009;57:334–7.
6. Mathai E, Rolain JM, Verghes GM, Abraham OC, Mathai D, Mathai M, et al. Outbreak of scrub typhus in southern India during the cooler months. Ann N Y Acad Sci. 2003;990:359–64.
7. Prabagaravaranthan R, Harish BN, Parija SC. Typhus fever in Pondicherry. J Commun Dis. 2008;40:159–60.
8. Lee CH, Liu JW. Coinfection with leptospirosis and scrub typhus in Taiwanese patients. Am J Trop Med Hyg. 2007;77:525–7.
Pandemic (H1N1) 2009 and HIV Co-infection

To the Editor: We report a case of pandemic (H1N1) 2009 infection in a man with serologic evidence of HIV-1 infection. The clinical course was complicated by lung and brain involvement (respiratory failure and lethargy), severe leukopenia, and thrombocytopenia, but complications resolved after treatment with oseltamivir (150 mg 2×/d).

In November 2009, a 47-year-old man who had received a diagnosis of hepatitis C infection 8 months earlier sought treatment at Ospedale Santa Maria Nuova, Reggio Emilia, Italy. He had a 3-day history of fever, dry cough, and drowsiness. Eight days before being admitted, the man had resided in the hospital’s inpatient detoxification unit, in which at least 10 inflammatory cases had been recorded. While in the detoxification unit, he had received methadone, 50 mg 1×/d. Computed tomography images of the brain and radiographs of the chest were normal; ultrasound examination showed upper lobe consolidation of the left lung. Hematochemistry showed high creatine phosphokinase levels, leukocyte count 1,380 cell/mm³ (reference range 4,000–10,000 cells/mm³), thrombocyte count 34,000 cells/mm³ (reference range 150,000–450,000 cells/mm³), partial pressure of oxygen 56 mm Hg, and partial pressure of carbon dioxide 53 mm Hg. Urinalysis results were negative for heroin, cocaine, and alcohol; cerebrospinal fluid (CSF) analysis results were within normal limits. Thrombocyte count returned to reference range after 2 days, and leukocyte count improved but remained <3,500 cells/mm³ for 3 weeks. After admission to the hospital, the man became lethargic and received noninvasive continuous positive airway pressure ventilation and treated with oseltamivir (150 mg 2×/d for 5 d), as well as with ceftriaxone, and levofloxacin. Reverse transcription–PCR on a throat swab confirmed influenza subtype H1N1 infection; blood cultures and urine were negative for pneumococcus, and Legionella spp. antigens. In addition, PCR of CSF for enterovirus and herpesvirus had negative results. The patient needed respiratory support for 4 days, after which his mental status and blood gases returned to reference levels. He was discharged from the hospital 2 weeks later.

On day 3 after admission, a nurse was accidentally exposed to the patient’s urine through her eye. An ELISA was positive for HIV infection. Negative results for confirmatory Western blot tests on days 5, 15, and 23 showed the p24 and p41 bands; HIV RNA was >6 million copies/mL, CD4 lymphocytes 51% (reference range 29%–59%). Reverse transcription–PCR for influenza subtype H1N1 performed 2 months later on a stored CSF sample gave a negative result; PCR for HIV of the same sample indicated 25,000 copies/mL. In mid-December, because of a further drop in CD4 lymphocytes to 17% (214 cells/mm³) and blood HIV RNA of 2.8 million copies/mL, the patient started highly active antiretroviral therapy and is being followed up as an outpatient.

Influenza (H1N1) and primary HIV infection share many signs and symptoms, such as fever, cough, sore throat, joint or limb pain, and diarrhea. The infections also share uncommon complications of the central nervous system (CNS); e.g., drowsiness, coma, and seizures. We cannot confirm that CNS involvement in the patient reported here was caused primarily by pandemic (H1N1) 2009, as suggested by influenza-like symptoms and the apparent effect of oseltamivir. Nor can we attribute CNS involvement to primary infection with HIV-1 (1); CSF results within normal limits and PCR negative for influenza subtype H1N1 do not rule out a causal relationship with pandemic (H1N1) 2009. In fact, the few cases of pandemic (H1N1) 2009 encephalopathy described show similar characteristics among children and adults (2–4). Alternatively, some authors have attributed HIV in CSF to brain inflammation and damage (5,6). The severe leukocytopenia and thrombocytopenia in our patient have not been described, even in complicated influenza subtype H1N1 infections (7). Because lymphopenia and mild thrombocytopenia are the usual findings, we believe that they probably resulted from HIV-1 or the effect of both viruses.

HIV seroconversion may initially occur during an acute febrile illness resembling influenza, and CNS involvement can complicate both infections. During an epidemic, acute HIV infection should also be considered (8). Less frequently, as in the patient described above, the 2 infections can occur simultaneously. History of recent risk behavior for blood exposure and severe leukocytopenia and thrombocytopenia should alert clinicians to other causes and prompt them to offer an HIV test to the patient.

Enrico Barchi, Francesca Prati, Maria Parmeggiani, and Maria Luisa Tanzi