Gene Co-Expression Network Analysis Confirms Four Independent Predictors of Survival In Ovarian Cancer Patients Under Optimal Debulking Surgery

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Primary research

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Abstract

Background: Serious ovarian cancer (OvCa) is the most common histological type of epithelial OvCa with poor prognosis. Despite received optimal cytoreduction and standard chemotherapy, a large proportion of patients are forced to recurrence or death within three years. To identify exact prognostic biomarkers associated with overall survival (OS) is urgent requirements of exploring rapid tumor progression mechanisms and developing novel strategies for immunotherapy.

Methods: The gene expression profiles of GSE49997, GSE9891 and TCGA were screened through rigorous criteria using R software and Bioconductor package. Weighted gene co-expression network analysis (WGCNA) was constructed to figure out gene clusters associated with OS. Protein-protein interaction (PPI) networks were built through STRING website. Prognostic values of potential biomarkers were validated using forest map and Kaplan-Meier analysis.

Results: According to screening criteria, 788 samples and 10402 genes were reserved as the modeling dataset. We detected five modules related to OS and intersected 108 genes through WGCNA after random sampling. PPI network analysis, Gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis revealed potential mechanisms of above biomarkers.

Conclusions: Four exact biomarkers (CANT1, P4HB, DUS1L and SIRT7) were confirmed as independent predictors of survival in OvCa patients with success of debulking surgery, which might provide promising biomarkers for prognostic judgement in ovarian cancer.

Background

Ovarian cancer is the most lethal gynecological malignancy, with 239,000 new cases diagnosed and 152,000 deaths worldwide annually[1]. It is also a high malignancy with insidious onset, consequently, patients with advanced stage (FIGO III/IV) account for up to three-quarters of all diagnosed cases[2]. Despite satisfactory tumor reduction surgery and adequate chemotherapy, 60% of patients face peritoneal recurrence and only 29% survive till 5-year[3, 4]. Among all the pathological types, high grade serous ovarian cancer (HGSOC) is the one with the worst prognosis. Prediction of HGSOC prognostic correlation biomarkers has great therapeutic significance and urgent requirement for clinical transformation.

The identification of ovarian cancer molecular subtype by Tothill promotes the application of integrated genomic research[5]. Previous study by Verhaak analyzed HGSOC individually which provided a prognostic model using 100-gene signature [6]. To reduce economic costs and improve repeatability, we need more precise genetic biomarkers. Weighted gene co-expression network analysis (WGCNA) is proposed to reconstruct robust gene modules in terms of large-scale gene expression profiles and the distinction of hub genes that drive key cellular signaling pathways.

In our present study, we have identified prognosis-associated genes for esophageal cancer[7] and grade predictive genes for serous ovarian cancer[8] through WGCNA analysis. On the basis of previous research, we conducted gene co-expression networks and clustered overall survival related genes based on three public microarray datasets (GSE49997, GSE9891, and TCGA), which included 464 HGSOC samples and 10647 genes.
This approach identified 108 co-expression genes and four validated hub genes significantly related to OS. Our study might be useful to develop clinical transformation of prognostic biomarkers and immunotherapy drugs.

**Materials And Methods**

**Preparation of HGSOC datasets**

The HGSOC datasets were screened out from the ‘NormalizerVcuratedOvarianData’ Bioconductor package[9], which included both gene microarray expression profile data and curated clinical data of ovarian cancer cohorts. Strict screening strategy was developed to reduce tumor heterogeneity: all high grade serious ovarian cancer patients that undergo optimal debulking surgery and with complete follow-up information were included. Accordingly, three microarray platforms datasets (GSE4997, GSE9891 and TCGA) were merged into the modeling set and 10647 common genes among all 3 modeling datasets were remained[10]. As results, 464 samples (87 patients, 75 patients and 302 patients) were used to do data pre-processing and batch effect adjustment through surrogate variable analysis [11](Figure 1A).

**Random sampling and WGCNA construction**

464 patients were regarded as the population. In order to increase accuracy and repeatability, we selected individual samples randomly to form different subsets on a pro-rata basis. Select 90%, 80%, 70% and 60% of the population and repeat 10 times, furthermore, select 90% samples and repeat 100 times. As mentioned above, five subsets were used to construct WGCNA separately (Figure 1B).

R package ‘WGCNA’ [12] was used to perform gene co-expression network separately. First, a matrix of pairwise correlations between all pairs of genes across all samples was constructed. Second, we chose soft-thresholding power 3 to which co-expression similarity is raised to achieve consistent scale free topology in multiple datasets. Third, we performed automatic network construction and module detection with min Module Size of 50, max Block Size of 10000, deep Split of 2, and merge Cut Height of 0.25. This procedure comprised calculation of network adjacencies and topological overlap dissimilarities, followed by scaling of topological overlap matrices and calculation of consensus topological overlap. Then, we built hierarchical clustering dendrograms of gene expression data for each dataset, and similar expression genes were merged into potential modules (Figure 2). Correlations between gene expressions and prognostic traits (vital time and vital status) were calculated among each dataset.

**Calculation of eigengene expression and identification of prognostic related gene modules**

The first principal component which represented the highest percent of variance for expression values of all genes in a module was regarded as module eigengene (ME). We used the expression profile of module eigengenes to discuss the correlation of module genes expression with overall survival (OS). Cox regression model was used for survival analysis using ‘OptimalCutpoints’ [13]and ‘survival’ Bioconductor packages[14].

**Construction of protein–protein interaction network**

We extracted a subnetwork with module genes from the STRING protein interaction database[15]. Since the STRING database weights and integrates information from numerous sources, including computational
prediction methods, experimental repositories, and public text collections, we set minimum required interaction score as high confidence (0.700), max number of interactions to show no more than 5, hoping to get a convincing interaction subnetwork of our module genes.

**Gene Ontology and Kyoto Encyclopedia of Genes and Genomes analysis**

The Database for Annotation, Visualization and Integrated Discovery (DAVID, https://david.ncifcrf.gov/) [16] was applied to perform Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) analyses of hub genes and their most relevant cooperators. The human genome (Homo sapiens) was selected as the background variables. Enrichment terms were considered statistically significant when the FDR were less than 0.05.

**Statistical analysis and verification**

All statistical analyses were performed using R 3.4.1 software. Kaplan–Meier survival plots were generated with survival curves through Kaplan-Meier Plotter website[17]. For all analyses, differences were considered statistically significant if the P values were less than 0.05.

**Results**

**Clinical characteristics in the HGSOC training set**

Three high quality datasets, respectively, GSE49997, GSE9891, and TCGA, were adopted from the ‘NormalizerVcuratedOvarianData’ Bioconductor package and were aggregated as the modeling dataset. In total, 464 samples and 10647 genes were included after data preprocessing. Among them, 429 were late-stage patients (FIGO III-IV). Clinical information statistically analysis and univariate cox regression analysis were performed. As shown in Table 1, there was significantly statistical difference between FIGO stage with recurrence (p = 0.001). Moreover, age more than 60 years old (p = 0.001), late FIGO stage (p = 0.049) and recurrence (p = 0.005) were also associated with the death consequence. These Data trends were consistent with clinical and epidemiological distributions.
Table 1

| Clinical features | Sample n = 464 | Non-recurrence n = 212 | Recurrence n = 252 | χ² | P value | Living n = 260 | Death n = 204 | χ² | P value |
|-------------------|----------------|------------------------|-------------------|----|---------|----------------|--------------|----|---------|
| age < 60          | 248            | 118                    | 130               | 3.67 | 0.055  | 157           | 91           | 10.815 | 0.001   |
| age ≥ 60          | 216            | 94                     | 122               |     |         | 103           | 113          |        |         |
| FIGO I            | 15             | 14                     | 1                 | 13.933 | 0.003 | 12            | 3            | 4.821 | 0.185   |
| FIGO II           | 20             | 11                     | 9                 |     |         | 15            | 5            |        |         |
| FIGO III          | 370            | 160                    | 210               |     |         | 201           | 169          |        |         |
| FIGO IV           | 59             | 27                     | 32                |     |         | 32            | 27           |        |         |
| Early stage(FIGO I-II) | 35         | 25                     | 10                | 11.279 | 0.001 | 27            | 8            | 3.885 | 0.049   |
| Late stage(FIGO III-IV) | 429       | 187                    | 242               |     |         | 233           | 196          |        |         |

Non-recurrence 212 - - - - 170 42 7.787 0.005
Recurrence 252 - - - - 90 162 - -

Construction of gene co-expression network and screening overall survival correlated modules

WGCNA was performed to identify gene co-expression network associated with HGSOC prognosis. A soft threshold of 3 was implemented, resulting in the detection of 27 significant gene modules (Fig. 2E). For each module, we calculated correlations between gene expressions and clinical features such as tumor stage, age, recurrence time, vital time, recurrence status, and vital status. In the present study, we focused on the risk factors of death. Survival analysis indicated five correlated modules, their names were greenyellow (138 genes), midnightblue (126 genes), cyan (126 genes), darkturquoise (70 genes) and white (56 genes)(Fig. 3A).

In order to test the reproducibility and stability of gene co-expression networks, we constructed gene co-expression network among ten random sampling datasets each containing 90% of 464 samples. In parallel, the sampling ratio gradually diminished to 80%, 70% and 60%. According to survival analysis, 2 out of 20 modules and 1 out of 24 modules were correlated with OS in 90% and 80% sampling groups. No prognostic module was found in 70% or 60% sampling groups. Eventually, we built WGCNA among 100 random sampling datasets each containing 90% of 464 samples and screened unique overall survival correlated cyan* module (113 genes) (Fig. 1B). As expected, there were 108 intersect genes between greenyellow module and cyan* module. Hence, we identified cyan* module genes for subsequent analysis (Fig. 3).

GO and pathway enrichment analysis of cyan* module genes

To explore the biological functions of cyan* module genes, we performed Gene Ontology (GO) analysis included biological process (GOBP), cellular component (GOCC) and molecular functions (GOMF). The top five enriched
terms in GOBP were protein modification by small protein removal (FDR=0.014), protein deubiquitination (FDR=0.03), ubiquitin-dependent protein catabolic process (FDR=0.011), post-translational protein modification (FDR=0.014) and cellular protein catabolic process (FDR=0.011). The significantly related KEGG pathway were proteasome (hsa03050) and thermogenesis (hsa04714), while Reactome pathway was apoptosis (FSA-109581).

**PPI construction of cyan* module genes**

We extracted a protein-protein interaction subnetwork with 118 nodes and 68 edges from the high quality STRING protein interaction database. PPI enrichment p-value is 0.0183. PSMA4, PSMC4, PSMD8, PSMA3, PSCMC5 and PSMD12 interacted with each other directly and were involved in the proteasome pathway (FDR=6.89e-05)(Figure 4). Meanwhile, ATP5D, RPS6KB1, COX11, ATP5H, GRB2, ATP5G1 and SMARCD2 were involved in thermogenesis KEGG pathway (hsa04714). RPS6KB1, RAC3, GRB2 and PRKCA were involved in choline metabolism in cancer (hsa05231).

**Validation of ten independent predictors of overall survival**

As mentioned above, there were 108 intersect genes between greenyellow module and cyan* module. Aiming to test the prognostic association with OS, we built forest plots of the expression of each gene using another 7 datasets (E.MTAB.386, GSE17260, GSE26712, GSE30161, GSE32062.GPL6480, PMID17290060 and TCGA.RNASeqV2) and illustrated the top ten independent predictors (C17orf62, CANT1, DUS1L, FN3KRP, GRB2, NARF, NUP85, P4HB, SIRT7 and STRA13) in table 2. The p values for the overall HR were between 4.42e-08 to 6.48e-05 (Figure 5). In addition, we demonstrated gene descriptions and other prognostic related cancers.

**Validation of four candidate predictors in optimal debulking ovarian cancer patients**

We selected 801 optimal debulking ovarian cancer patients as the validation dataset through online tool. Four out of ten predictors were filtered with absolutely significant differences. As demonstrated in Figure 6, low mRNA expression levels of CANT1 (p=7.6e-05), P4HB (p=9.8e-07), DUS1L (p=6.4e-06) and SIRT7 (p=9.5e-08) were associated with worse OS in OvCa patients.
| Gene name  | P value     | Prognostic related cancer | Gene description                                      |
|------------|-------------|----------------------------|-------------------------------------------------------|
| C17orf62   | 6.48e-05    | renal/ liver               | Chromosome 17 open reading frame 62                   |
| CANT1      | 3.74e-05    | renal/ lung                | Calcium activated nucleotidase 1                      |
| DUS1L      | 4.42e-08    | renal                      | Dihydrouridine synthase 1 like                        |
| FN3KRP     | 1.51e-05    | glioma                     | Fructosamine 3 kinase related protein                 |
| GRB2       | 2.73e-05    | liver                      | Growth factor receptor bound protein 22              |
| NARF       | 7.29e-07    | renal/cervical             | Nuclear prelamin a recognition factor                 |
| NUP85      | 3.57e-05    | renal/ liver               | Nucleoparin 85                                        |
| P4HB       | 2.21e-05    | renal                      | Prolyl 4-hydroxylase subunit beta                     |
| SIRT7      | 6.04e-05    | renal/ liver               | Sirtuin 7                                             |
| STRA13     | 5.84e-06    | pancreatic                 | Basic helix-loop-helix family member e40             |

**Discussion**

The advantage of bioinformatics analysis is the integration of large amounts of data. In this study, we integrated large-scale transcriptional profiling to identify robust co-expression gene modules associated with ovarian cancer overall survival. The filtering condition of modeling dataset determines whether the results are logical and worthy of generalization. Through epidemiological investigation and clinical observation, we confirmed HGSOC patients with optimal debulking surgery in survival analysis not only because their high incidence, but also rarely effectiveness of aftertreatment. Our long-term goal was to provide insights into genetic targets of rapid tumor progression and improve clinical prognosis.

In general, 464 samples and 10647 genes were included to construct WGCNA and 801 samples were used in validation in silico. Four predictors, namely, CANT1, P4HB, DUS1L and SIRT7 were associated with OS in OvCa patients with optimal debulking surgery. Reports of associations between these genes and ovarian cancer are rare. However, their link to other cancers are revealed increasingly.

The androgen-regulated Calcium-Activated Nucleotidase 1 (CANT1) is widely expressed in various organs. Previous study has shown a commonly overexpressed in prostate carcinomas and prostatic intraepithelial neoplasia[18]. Protein level of CANT1 is significantly higher in clear cell renal cell carcinoma tissues than in adjacent normal tissues. CANT1 silencing suppressed cell proliferation, migration, and invasion in renal cancer[19]. Mutation of CANT1 is identified as a common pathogenic change for Desbuquois dysplasia type 1[20].

Autophagy-related gene prolyl 4-hydroxylae, beta polypeptide (P4HB) has been validated as a promising diagnosis and prognosis biomarker in kidney renal clear cell carcinoma[21] and bladder cancer[22]. Besides, P4HB contributes to cell proliferation, migration or invasion in gastric cancer[23], breast cancer[24] and colon cancer[25] through multiple pathways.
Sirtuin 7 (SIRT7) encodes a member of the sirtuin family of NAD+ dependent protein deacetylases and acts as a key mediator of multiple cellular activities. Protein expression is linked to oncogenic activity and regulation of ribosome biogenesis[26]. It also antagonizes transforming growth factor β signaling and inhibits lung metastases in breast cancer[27]. SIRT7 depletion inhibits cell proliferation, androgen-induced autophagy, and invasion in prostate cancer[28]. The expression pattern of SIRT superfamily and their prognostic values in serous ovarian cancer patients has been analyzed using GSE10971 and GSE30587 datasets. Increased expression of SIRT3, SIRT5, and SIRT7 is associated with better overall survival by univariable analysis which is consistent with our findings[29]. Dihydrouridine synthase 1 like (DUS1L) expresses ubiquitous in prostate, spleen and 25 other tissues with unclear gene function.

Multiple integrated bioinformatics analysis are attempting to identify potential biomarkers in ovarian cancer[30, 31]. Despite the insightful findings, limitations still exist in likeness study. Firstly, the allocation ratio between modeling and validation datasets may affect the prediction results. Secondly, multi-center follow up data requires significant time, economic and labor costs. The predicted biomarkers need to be verified through biology experiments and clinical evidences.

**Conclusions**

Overall, our results predicted and validated valuable prognostic biomarkers in ovarian cancer patients with optimal debulking surgery. We obtained four promising genes (CANT1, P4HB, DUS1L and SIRT7) through WGCNA analysis and survival analysis in silico. Further molecular mechanism studies on prognostic biomarkers may provide insights into the new targets for progressive tumor treatment.

**Abbreviations**

OC: ovarian cancer; SOC: serous ovarian cancer; GEO: Gene Expression Omnibus database; TCGA: The Cancer Genome Atlas database; OS: overall survival; KEGG: Kyoto Encyclopedia of Genes and Genomes; GO: Gene Ontology; GOBP: Gene Ontology biological processes; GOMF: Gene Ontology molecular function; 95% CI: 95 confidence intervals; HR: Hazard ratio.

**Declarations**

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Not applicable.

**Authors’ contributions**

CZ and TZ have equal contributions to this study. QS and CZ conceived the study and participated in the study design, performance, coordination. QS, CZ and TH carried out the assays and analysis. CZ draft the manuscript. QS and TZ revised the manuscript. All authors reviewed the manuscript. All authors read and approved the final manuscript.

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**Availability of data and materials**

All data are included in the article.

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

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**Competing interests**

The authors declare that they have no competing interests.

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**References**

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394–424.

2. Peres LC, Cushing-Haugen KL, Kobel M, Harris HR, Berchuck A, Rossing MA, Schildkraut JM, Doherty JA. Invasive Epithelial Ovarian Cancer Survival by Histotype and Disease Stage. J Natl Cancer Inst. 2019;111(1):60–8.

3. Azais H, Estevez JP, Foucher P, Kerbage Y, Mordon S, Collinet P. Dealing with microscopic peritoneal metastases of epithelial ovarian cancer. A surgical challenge. Surg Oncol. 2017;26(1):46–52.

4. Reid BM, Permuth JB, Sellers TA. Epidemiology of ovarian cancer: a review. Cancer Biol Med. 2017;14(1):9–32.

5. Tothill RW, Tinker AV, George J, Brown R, Fox SB, Lade S, Johnson DS, Trivett MK, Etemadmoghadam D, Locandro B, et al. Novel molecular subtypes of serous and endometrioid ovarian cancer linked to clinical outcome. Clin Cancer Res. 2008;14(16):5198–208.

6. Verhaak RG, Tamayo P, Yang JY, Hubbard D, Zhang H, Creighton CJ, Fereday S, Lawrence M, Carter SL, Meremel CH, et al. Prognostically relevant gene signatures of high-grade serous ovarian carcinoma. J Clin Invest. 2013;123(1):517–25.

7. Zhang C, Sun Q. Weighted gene co-expression network analysis of gene modules for the prognosis of esophageal cancer. J Huazhong Univ Sci Technolog Med Sci. 2017;37(3):319–25.

8. Sun Q, Zhao H, Zhang C, Hu T, Wu J, Lin X, Luo D, Wang C, Meng L, Xi L, et al. Gene co-expression network reveals shared modules predictive of stage and grade in serous ovarian cancers. Oncotarget.
9. Ganzfried BF, Riester M, Haibe-Kains B, Risch T, Tyekucheva S, Jazic I, Wang XV, Ahmadifar M, Birrer MJ, Parmigiani G, et al: *curatedOvarianData*: clinically annotated data for the ovarian cancer transcriptome. *Database (Oxford)* 2013, 2013:bat013.

10. Pils D, Hager G, Tong D, Aust S, Heinze G, Kohl M, Schuster E, Wolf A, Sehouli J, Braicu I, et al. Validating the impact of a molecular subtype in ovarian cancer on outcomes: a study of the OVCAD Consortium. Cancer Sci. 2012;103(7):1334–41.

11. Lee S, Sun W, Wright FA, Zou F. An improved and explicit surrogate variable analysis procedure by coefficient adjustment. Biometrika. 2017;104(2):303–16.

12. Langfelder P, Horvath S. WGCNA: an R package for weighted correlation network analysis. BMC Bioinformatics. 2008;9:559.

13. Perkins NJ, Schisterman EF. The inconsistency of "optimal" cutpoints obtained using two criteria based on the receiver operating characteristic curve. Am J Epidemiol. 2006;163(7):670–5.

14. Gentleman RC, Carey VJ, Bates DM, Bolstad B, Dettling M, Dudoit S, Ellis B, Gautier L, Ge Y, Gentry J, et al. Bioconductor: open software development for computational biology and bioinformatics. Genome Biol. 2004;5(10):R80.

15. Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, Simonovic M, Doncheva NT, Morris JH, Bork P, et al. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 2019;47(D1):D607–13.

16. Dennis G Jr, Sherman BT, Hosack DA, Yang J, Gao W, Lane HC, Lempicki RA. DAVID: Database for Annotation, Visualization, and Integrated Discovery. Genome Biol. 2003;4(5):P3.

17. Gyorffy B, Lanczky A, Szallasi Z. Implementing an online tool for genome-wide validation of survival-associated biomarkers in ovarian-cancer using microarray data from 1287 patients. Endocr Relat Cancer. 2012;19(2):197–208.

18. Gerhardt J, Steinbrech C, Buchi O, Behnke S, Bohnert A, Fritzsche F, Liewen H, Stenner F, Wild P, Hermanns T, et al. The androgen-regulated Calcium-Activated Nucleotidase 1 (CANT1) is commonly overexpressed in prostate cancer and is tumor-biologically relevant in vitro. Am J Pathol. 2011;178(4):1847–60.

19. Liu X, Yang Z, Luo X, Luo J, Fu W, Fang Z, Xia D, Li L, Xu J. Calcium-activated nucleotidase 1 silencing inhibits proliferation, migration, and invasion in human clear cell renal cell carcinoma. J Cell Physiol. 2019;234(12):22635–47.

20. Wang HD, Guo LJ, Feng ZQ, Zhang DW, Zhang MT, Gao Y, Chen CL, Zhu BF. Cloning, expression and enzyme activity delineation of two novel CANT1 mutations: the disappearance of dimerization may indicate the change of protein conformation and even function. Orphanet J Rare Dis. 2020;15(1):240.

21. Xie L, Li H, Zhang L, Ma X, Dang Y, Guo J, Liu J, Ge L, Nan F, Dong H, et al. Autophagy-related gene P4HB: a novel diagnosis and prognosis marker for kidney renal clear cell carcinoma. Aging. 2020;12(2):1828–42.

22. Wu Y, Peng Y, Guan B, He A, Yang K, He S, Gong Y, Li X, Zhou L. P4HB: A novel diagnostic and prognostic biomarker for bladder carcinoma. Oncol Lett. 2021;21(2):95.

23. Zhang J, Guo S, Wu Y, Zheng ZC, Wang Y, Zhao Y. P4HB, a Novel Hypoxia Target Gene Related to Gastric Cancer Invasion and Metastasis. Biomed Res Int. 2019;2019:9749751.
24. Yang W, Wu X, Zhou F. Collagen Type X Alpha 1 (COL10A1) Contributes to Cell Proliferation, Migration, and Invasion by Targeting Prolyl 4-Hydroxylase Beta Polypeptide (P4HB) in Breast Cancer. Med Sci Monit. 2021;27:e928919.

25. Zhou Y, Yang J, Zhang Q, Xu Q, Lu L, Wang J, Xia W. P4HB knockdown induces human HT29 colon cancer cell apoptosis through the generation of reactive oxygen species and inactivation of STAT3 signaling. Mol Med Rep. 2019;19(1):231–7.

26. Blank MF, Grummt I. The seven faces of SIRT7. Transcription. 2017;8(2):67–74.

27. Tang X, Shi L, Xie N, Liu Z, Qian M, Meng F, Xu Q, Zhou M, Cao X, Zhu WG, et al. SIRT7 antagonizes TGF-beta signaling and inhibits breast cancer metastasis. Nat Commun. 2017;8(1):318.

28. Ding M, Jiang CY, Zhang Y, Zhao J, Han BM, Xia SJ. SIRT7 depletion inhibits cell proliferation and androgen-induced autophagy by suppressing the AR signaling in prostate cancer. J Exp Clin Cancer Res. 2020;39(1):28.

29. Li J, Yue H, Yu H, Lu X, Xue X. Development and validation of SIRT3-related nomogram predictive of overall survival in patients with serous ovarian cancer. J Ovarian Res. 2019;12(1):47.

30. Liu J, Meng H, Li S, Shen Y, Wang H, Shan W, Qiu J, Zhang J, Cheng W. Identification of Potential Biomarkers in Association With Progression and Prognosis in Epithelial Ovarian Cancer by Integrated Bioinformatics Analysis. Front Genet. 2019;10:1031.

31. Chen J, Cai Y, Xu R, Pan J, Zhou J, Mei J. Identification of four hub genes as promising biomarkers to evaluate the prognosis of ovarian cancer in silico. Cancer Cell Int. 2020;20:270.

**Figures**

**Figure 1**

Flow chart of WGCNA construction. A. The gene expression profiles of HGSOC patients with optimal debulking were filtered using three step screening strategy through 'NormalizeVcuratedOvarianData' Bioconductor
package. GSE49997, GSE9891 and TCGA were used as modeling datasets in data pre-processing. B. Weighted gene co-expression networks were constructed and 5 out of 27 modules were related with overall survival. Select 90%, 80%, 70% and 60% of the population and repeat 10 times, furthermore, select 90% samples and repeat 100 times to build another five parallel modeling datasets. As results, 1 out of 23 modules was related with overall survival in 90% random sampling *100 times group.

**Figure 2**

Clustering dendrograms of genes were based on dissimilarity topological overlap and module colors. A. 90% samples repeat 10 times; B. 80% samples repeat 10 times; C. 70% samples repeat 10 times; D. 60% samples repeat 10 times; E. 100% samples; F. 90% samples repeat 100 times.
Figure 3

Identification of relevant modules associated with OvCa clinical traits. Detection of co-expression networks. A. Heatmap of the correlation between five prognostic related module eigengenes and clinical traits in 100% samples group. B. Heatmap of the correlation between unique prognostic related module eigengenes and clinical traits in 90% samples*100 times group. C. Venn diagram intersected 108 genes between cyan* module and greenyellow module.
Figure 4

The PPI network of hub cyan* genes constructed by STRING. 113 hub genes was constructed in high quality STRING website.
Figure 5

Top ten genes with significant p-value less than 0.00005 were shown in the forest plot.
Figure 6

The prognostic values of four genes in validation dataset. Patients were divided into a high-expression group and a low-expression group according to the median gene expression. The prognostic values of A. CANT1; B. P4HB; C. DUS1L; D. SIRT7 expression in predicting OS in OvCa patients.