Antibody Repertoires Identify β-Tubulin as a Host Protective Parasite Antigen in Mice Infected With Trypanosoma cruzi

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Few studies investigate the major protein antigens targeted by the antibody diversity of infected mice with Trypanosoma cruzi. To detect global IgG antibody specificities, sera from infected mice were immunoblotted against whole T. cruzi extracts. By proteomic analysis, we were able to identify the most immunogenic T. cruzi proteins. We identified three major antigens as pyruvate phosphate dikinase, Hsp-85, and β-tubulin. The major protein band recognized by host IgG was T. cruzi β-tubulin. The T. cruzi β-tubulin gene was cloned, expressed in E. coli, and recombinant T. cruzi β-tubulin was obtained. Infection increased IgG reactivity against recombinant T. cruzi β-tubulin. A single immunization of mice with recombinant T. cruzi β-tubulin increased specific IgG reactivity and induced protection against T. cruzi infection. These results indicate that repertoire analysis is a valid approach to identify antigens for vaccines against Chagas disease.

Keywords: Trypanosoma cruzi, antibody repertoires, beta-tubulin, lymphocyte activation, Chagas disease

INTRODUCTION

Chagas disease is caused by the protozoan parasite Trypanosoma cruzi and imposes a heavy burden on human health in Latin America. Infection with T. cruzi leads to parasitemia and spread of the parasite to host tissues. Control of T. cruzi infection depends on cells of innate and acquired immunity, which produce cytokines, inflammatory mediators, and antibodies (1, 2). However, tissue infection persists indefinitely at low levels (1, 2). Several studies indicate the importance of antibodies in protection against T. cruzi infection (3, 4, 5, 6), but the precise role of humoral immunity in host defense remains incompletely understood.

Purified T. cruzi proteins induce protection in mice challenged with live parasites. These antigens include cruzipain (7), trans-sialidase (8), members of mucin-associated surface protein family (9), and excretory-secretory antigens (10). However, most studies amplify detection by using recombinant antigens, or probing with antibodies from immunized animals. New studies have characterized novel parasites antigens that could be targeted in vaccine studies (11, 12). An alternative approach to the testing of randomly purified molecules is the identification of T. cruzi antigens which are...
targeted by the host antibody diversity in the course of infection. Antibody diversity can be analyzed by an immunoblot technique which detects global antibody reactivity against whole protein extracts (13). This method detects autoantibodies produced in autoimmune diseases (14, 15) and identifies repertoire changes linked to resistance against T. cruzi infection (16).

Microtubules are cytoskeletal structures composed of α/β tubulin heterodimers that are found in eukaryotic cells and are abundant in trypanosomatid parasites from the order Kinetoplastida (17). These structures have important functions in cell division, maintenance of cellular morphology, motility, intracellular transport, and signal transduction (18). In trypanosomes microtubules have two α-tubulin isofoms and one β-tubulin isoform (19) and are found underneath the plasma membrane (the subpellicular microtubules), in the flagellum, and as a component of mitotic spindle apparatus (20). The microtubules function as a perfect target for many compounds with trypanocidal activity, blocking tubulin activity (21). Therefore, a microtubule component is a suitable target to be considered as an effective vaccine candidate to protect from trypanosomais infections (22, 23). It was previously shown that mice vaccinated with native tubulin purified from Trypanosoma brucei were protected against T. brucei, Trypanosoma congoense, and Trypanosoma rhodesiense infection (22). Previous report showed that mice vaccinated with the microtubule-associated protein (MAP) p52 of T. brucei, together with aldolase, GAPDH, and MAP p15, were protected after challenge with a homologous infection (24). Furthermore, rabbit antibodies to tubulin-rich fractions from T. brucei inhibit the growth of trypanosomes in culture (25).

Given the paucity of information regarding the subject, this work addresses the identification of prominent T. cruzi antigens targeted by IgG antibodies during infection. Here, we describe that acute and chronic infection of BALB/c mice induced limited changes in the antibody diversity. Using a proteomic approach, we identified T. cruzi β-tubulin (TcβTUB) as one of the major antigens targeted by antibodies. The T. cruzi β-tubulin gene was isolated, cloned, and expressed. Recombinant TcβTUB was recognized by sera from infected mice, and immunization of naïve mice with a single dose of recombinant TcβTUB induced protection against infection. These results indicate the importance of selecting candidate vaccines antigens from analysis of unbiased Ab reactivities from infected mice.

**MATERIALS AND METHODS**

**Mice, Parasite, and Infection**

Male wild-type (WT) BALB/c and Fas-L mutant BALB/c.gld (gld) mice aging 6–8 weeks, weighing 25–30 g were from the Oswaldo Cruz Institute Animal Care facility, Rio de Janeiro. BALB/gld mice (26) were produced at the National Institutes of Health, Bethesda, MD, USA by serially backcrossing the gld gene onto a BALB/c background for 15 generations.

All mouse studies followed the guidelines set by the National Institutes of Health, United States. The study was approved by the Research Ethics Committee of Federal University of Rio de Janeiro (protocol 062/14). Protocols for animal were approved by the Institutional Ethical Committees in accordance with international guidelines. All animal experimentation was performed in accordance with the terms of the Brazilian guidelines for the animal welfare regulations.

Mice were infected with intraperitoneal injection (i.p.) with 10⁷ chemically induced metacyclic forms of T. cruzi clone Dm28c (17, 18). Chemically induced and insect-derived metacyclic forms induce a similar infection as demonstrated previously (18). Acute infection was evaluated after 23–33 days of infection, while chronic infection was evaluated after 150 days.

**Recombinant T. cruzi β-Tubulin**

Genomic DNA was extracted from T. cruzi Dm28c epimastigotes (1 x 10⁶; Rapidprep isolation kit, Pharmacia) and used as template for amplification of T. cruzi β-tubulin gene (19) by touchdown PCR (27). Touchdown PCR was carried out in 50 µL of 20 mM Tris–HCl (pH 8.8), 2 mM MgSO₄, 10 mM KCl, 10 mM (NH₄)₂SO₄, 0.1% (v/v) Triton X-100, 0.1 mg/mL nuclease-free BSA, 100 ng DNA, 0.5 mM dNTPs, 0.4 µM of primers TcβTubBF (5’-ATCATGCGTGGATTGGTGGCG-3’) and TcβTubBR (5’-ATGAAATTGTAGTACTGCTCCCTC-3’), and a mixture of 2.0 U PuF (Fermentas) and 0.25 U Taq (Biotools) DNA polymerases. The resulting 1,342 bp fragment was cloned into pTZ57R/T (Fermentas), sequenced and sub-cloned, yielding pET-28a-TcβTUB expression plasmid. pET-28a-TcβTUB was introduced into BL21 (DE3) E. coli strain, and transfectants were induced with 0.5 mM isopropyl 1-thio-β-d-galactopyranoside (Invitrogen, USA) for 18 h at 20°C. Cells were suspended in 300 µg/mL lysozyme (Sigma) in 50 mM Tris–HCl (pH 7.5), 200 mM NaCl, 5% (w/v) glycerol, 1 mM DTT, and protease inhibitor mix (Sigma), and left 30 min on ice. After addition of 10 U DNAsc I (Fermentas) and 5 mM MgCl₂, the suspension was incubated for 30 min on ice and lysed by sonication. The lysate was centrifuged at 30,900 g for 40 min at 4°C, and SDS-PAGE indicated the presence of recombinant T. cruzi β-tubulin (TcβTUB). The pellet was solubilized in 2% Triton X-100, 2 M Urea, 100 mM Tris–HCl (pH 7.5), 5 mM NaCl, 0.5% (w/v) glycerol, 1 mM DTT, and protease inhibitor mix (Sigma), and left 30 min on ice. After addition of 10 U DNAsc I (Fermentas) and 5 mM MgCl₂, the suspension was incubated for 30 min on ice and lysed by sonication. The lysate was centrifuged at 30,900 g for 40 min at 4°C, and SDS-PAGE indicated the presence of recombinant T. cruzi β-tubulin (TcβTUB). The pellet was solubilized in 2% Triton X-100, 2 M Urea, 100 mM Tris–HCl (pH 7.5), 30% NaCl, 5% (w/v) glycerol, 1 mM DTT, and protease inhibitor mix (Sigma), and left 30 min on ice. After addition of 10 U DNAsc I (Fermentas) and 5 mM MgCl₂, the suspension was incubated for 30 min on ice and lysed by sonication. The lysate was centrifuged at 30,900 g for 40 min at 4°C, and SDS-PAGE indicated the presence of recombinant T. cruzi β-tubulin (TcβTUB). The pellet was solubilized in 2% Triton X-100, 2 M Urea, 100 mM Tris–HCl (pH 7.5), 5 mM NaCl, 0.5% (w/v) glycerol, 1 mM DTT, and protease inhibitor mix (Sigma), and left 30 min on ice. After addition of 10 U DNAsc I (Fermentas) and 5 mM MgCl₂, the suspension was incubated for 30 min on ice and lysed by sonication. The lysate was centrifuged at 30,900 g for 40 min at 4°C, and SDS-PAGE indicated the presence of recombinant T. cruzi β-tubulin (TcβTUB). The pellet was solubilized in 2% Triton X-100, 2 M Urea, 100 mM Tris–HCl (pH 7.5), 5 mM NaCl, 0.5% (w/v) glycerol, 1 mM DTT, and protease inhibitor mix (Sigma), and left 30 min on ice. After addition of 10 U DNAsc I (Fermentas) and 5 mM MgCl₂, the suspension was incubated for 30 min on ice and lysed by sonication. The lysate was centrifuged at 30,900 g for 40 min at 4°C, and SDS-PAGE indicated the presence of recombinant T. cruzi β-tubulin (TcβTUB). The pellet was solubilized in 2% Triton X-100, 2 M Urea, 100 mM Tris–HCl (pH 7.5), 0.5 M NaCl containing 5 mM imidazole (wash buffer). Recombinant TcβTUB was eluted with wash buffer containing 0.25 M imidazole, analyzed for purity on SDS-PAGE, and quantified by the method of Bradford (28) using BSA as standard.

**ELISA**

For determination of serum IgG, microplates (Greiner BiOne) were coated with 2 µg/mL goat anti-mouse IgG (Southern Biotechnology) for 16 h at 4°C. Concentrations were calculated based on standard curves generated with purified mouse IgG (Southern). In addition, microplates were coated with dsDNA from calf thymus, histone from calf thymus, myosin from rabbit heart, KLH (5 µg/mL; all from Sigma), or T. cruzi antigen diluted at 1:1,000. T. cruzi antigen was obtained by lysing epimastigotes in buffer containing 200 mM Tris, 400 mM NaCl, 40 mM EDTA, 20 mM iodoacetamide, and 1 mM PMSF. ELISA tests for IgG against recombinant TcβTUB (0.5 µg/mL) were performed. Plates
were blocked with PBS-1% gelatin (Vetec, Brazil) for 1 h and washed with PBS containing 0.05% Tween-20. Sera were kept at −20°C until use. Data show the results with 1:100 serum dilution. Binding was determined following incubation with alkaline phosphatase-conjugated goat anti-mouse IgG (Southern). Reaction was developed with PnPP substrate (Southern) diluted in Tris–MgCl₂ buffer. Absorbance was read at 405 nm.

Immunoblots

For antibody repertoire analysis, mouse hearts were lysed in extraction buffer (2% SDS, 5% 2-mercaptoethanol, and 62.5 mM Tris, pH 6.8) on ice, without protease inhibitors, at a proportion of 1 g/10 mL buffer. *T. cruzi* epimastigotes (clone Dm28c) were lysed in 10 mL extraction buffer (5 × 10⁶/mL). Extracts were sonicated for 10 min, boiled for 10 min, centrifuged at 1,000 g, and then at 10,000 g. Supernatants were stored at −20°C. IgG reactivities against heart and *T. cruzi* polypeptides were identified by a modified immunoblot technique (13). Briefly, extracts (600 µg/mL) were subjected to SDS-PAGE, and proteins were transferred to nitrocellulose. Membranes were blocked for 18 h with PBS-0.2% Tween-20 at RT and incubated for 4 h with sera adjusted for 100 µg/mL IgG concentration, using the Cassette Miniblot System (Immunetics, Cambridge, MA, USA). Alkaline phosphatase-conjugated secondary goat anti-mouse IgG antibody (Southern) was added for 90 min. After washing, immunoreactivities were revealed with nitroblue-tetrazolium/5-bromo-chloro-indolyl-phosphate (NBT/BCIP; Promega) and phosphatase-conjugated goat anti-mouse IgG (Southern). Binding was determined following incubation with alkaline phosphatase-conjugated goat anti-mouse IgG (Southern). Reaction was developed with PnPP substrate (Southern) diluted in Tris–MgCl₂ buffer. Absorbance was read at 405 nm.

Mass Spectrometry

*T. cruzi* extracts were subjected to SDS-PAGE in parallel mirror lanes. One lane was blotted and reacted with serum to identify the major bands. Following precise location, the mirror lane was excised in the gel. Excision and processing of protein spots were performed as previously described (31). Digestions were done with sequencing grade porcine trypsin (Promega, Madison, WI, USA) at 37°C for 16 h. Tryptic peptides (0.5 mL) were mixed with a saturated solution of CHCA matrix in 50% ACN, 1% TFA (0.5 mL), spotted onto an MALDI sample plate, and allowed to crystallize at room temperature. MALDI/TOF-TOF peptide sequencing, operated in reflectron-delayed extraction mode with high resolution for the 800–4,000 Da range, was performed by precursor ion fragmentation, using N₂ gas in the collision cell at 2.8 × 10⁻⁴ torr, in a 4700 Explorer Proteomics Analyzer (Applied Biosystems). Protein identification was carried out against the NCBI database using the MASCOT software (www.matrixscience.com) and the following parameters: cys-carbamidomethylation as fixed modification; methionine oxidation as variable modification; one missing trypsin cleavage, monoisotopic masses, peptide, and ion tolerances at 0.2 Da. Under these conditions, a probability based score 81 was considered significant (p < 0.05). The other criteria for identification were a minimum of 20% of protein coverage and four peptides with hits in the database (for PMF data) or two peptide sequence tags (for MS/MS).

Immunization Assays

BALB/c mice were injected with PBS, BSA, or recombinant TcβTUB (both 20 µg/animal) emulsified in Complete Freund Adjuvant (CFA, Thermo Scientific) in the hind footpads. After 14 days, mice were infected with 10³ metacyclic trypomastigotes as above. Parasitemia was evaluated by tail vein puncture; viable parasites were counted in a Neubauer chamber. Mice were sacrificed at day 32 of infection, and individual spleens were weighted. Individual sera were collected and analyzed by western blotting for reactivity against *T. cruzi* and mouse tubulins.

Statistical Analysis

Data were analyzed by Student’s t-test for independent samples and ANOVA test, using SigmaPlot™ for Windows. Data from parasitemia were normalized by log transformation before statistical testing. Data from densitometric analysis were analyzed by Mann–Whitney test, using Graph Pad InStat 3.01 for Windows. Differences with a p value < 0.05 or lower were considered significant.

RESULTS

Antibody Reactivities Against Exogenous and Autologous Antigens in *T. cruzi* Infection

We infected BALB/c mice with chemically induced metacyclic forms of *T. cruzi* clone Dm28c (17, 18). Chemically induced and insect-derived metacyclic forms induce a similar infection (18). Acute and chronic infection with *T. cruzi* increased the concentrations of serum IgG (data not shown). We measured IgG reactivities against a panel of exogenous and autologous antigens (*T. cruzi*, KLH, myosin, dsDNA, and histone) by ELISA in the sera of control and infected mice. Only IgG reactivities against *T. cruzi* antigens increased after acute (Figure 1A, left) or chronic...
infection (Figure 1B, left). As a positive control for autoreactivity, we used sera from lupus-prone gld mice for comparison (Figures 1A,B, right). Our results show that a highly focused Ab response against T. cruzi antigens, and not autoreactivity, is the final outcome following infection.

Global Analysis of IgG Reactivities Against T. cruzi and Cardiac Polypeptides

We analyzed global IgG antibody reactivities with a semi-quantitative immunoblot technique against T. cruzi epimastigote extracts (13–16). Western blots of individual sera from chronic infection showed that IgG reactivities were directed to a restricted set of antigens (Figure 2A, right). The strongest reactivity was directed to a 50–55 kDa band, which was detected with sera from all infected mice (Figure 2A). This reactivity colocalized with the most abundant band present on Coomassie blue-stained gels of T. cruzi extracts (Figure 2B, left). We quantified the immunoblots from all animals at both acute and chronic infection and showed the average optical density profiles of the IgG reactivities against T. cruzi proteins (Figures 3A,B, left). Global IgG reactivity against T. cruzi was focused on a few bands of the parasite extract (Figures 3A,B, left). The most prominent antibody reactivity against T. cruzi was represented by a band located at positions 600 and 800 in the blots of acute and chronic stages, respectively, depending on conditions of the electrophoresis assay (arrows on Figures 3A,B, left). Reacting T. cruzi extract with a mixture of acute and chronic sera on the same membrane yielded a single band, suggesting that it was the same reactivity (not shown). Acutely infected gld mice gave the same prominent IgG reactivity (Figure S1 in Supplementary Material). In addition, experiments with trypomastigote enriched extracts indicated the presence of an intense 50–55 kDa IgG reactivity (data not shown).

Infection with T. cruzi results in myocarditis (18). We therefore analyzed IgG antibody reactivities against autologous heart polypeptides (13–16). Global IgG reactivity against cardiac polypeptides was not significantly increased following acute or chronic infection with T. cruzi (Figures 3A,B, right).
Identification of T. cruzi β-Tubulin as a Major Antigenic Target for Antibodies in Infection

To identify the 50–55 kDa band reacting with sera from infection, the band was located in the gel with the help of the blot of a mirror lane, excised and subjected to mass spectrometry analysis. As shown in Table 1 and Figure 3C, the 50–55 kDa band was identified as T. cruzi β-tubulin (TcβTUB). We also identified two additional major T. cruzi antigens targeted by antibodies. The 100 kDa band was identified as pyruvate phosphate dikinase, and the 85 kDa band was identified as Hsp-85 (Figure 3C and data not shown).

The β-tubulin coding region (19) was PCR amplified from T. cruzi Dm28c genomic DNA. The resulting fragment was cloned, sequenced and sub-cloned into pET-28a-TcβTUB expression plasmid. Recombinant TcβTUB was expressed and purified (Figure S2 in Supplementary Material). Following reaction with anti-β-tubulin monoclonal antibody, recombinant TcβTUB gave a single band compatible with the major band expressed in T. cruzi extracts (Figure 2B right). Analysis by ELISA confirmed that chronic infection with T. cruzi increased the production of IgG reactive with TcβTUB (Figure 4).

Immunization With T. cruzi β-Tubulin Induced Protection Against Infection

Naïve BALB/c mice were immunized with a single dose of recombinant TcβTUB in CFA and challenged with T. cruzi after 2 weeks. Control groups were immunized with PBS/CFA and BSA/CFA. All mice were sacrificed after 32 days of infection. Parasitemia was markedly reduced in mice immunized with TcβTUB/CFA, compared with controls (Figure 5A). Spleen cellularity, which correlates with parasite burden in T. cruzi infection (32), was reduced in mice immunized with TcβTUB/CFA, compared with controls (Figure 5B). As expected, mice immunized with TcβTUB/CFA before infection produced higher amounts of anti-TcβTUB IgG, compared with controls (Figure 5C).

IgG reactivities of control and immunized mice were compared by western blotting against mouse tubulin (MBE), recombinant TcβTUB, and native T. cruzi tubulin (Figure 6). IgG from mice immunized with BSA/CFA before infection reacted...
TABLE 1 | Mass spectrometry analysis of peptide fragments of the major 50–55 kDa band of Trypanosoma cruzi extract corresponding to T. cruzi β-tubulin.

| Start | End | Miss | Peptide sequence | Mr (expt)b | Mr (calc) | Delta | m/zc |
|-------|-----|------|------------------|------------|-----------|--------|-------|
| 310   | 318 | 0    | R.YLTASALFR.G    | 1,040.5700 | 1,040.5655 | 0.0046 | 1,041.5773 |
| 253   | 262 | 0    | R.KAVNLYVPR.P.R  | 1,124.6709 | 1,124.6706 | 0.0003 | 1,125.6782 |
| 242   | 251 | 0    | R.FPGQLNSDL.R.K  | 1,145.5862 | 1,145.5829 | 0.0033 | 1,146.5935 |
| 381   | 390 | 0    | R.VGEQFTAMFR.R   | 1,200.5578 | 1,200.5598 | -0.0019 | 1,201.5651 |
| 381   | 390 | 0    | R.VGEQFTAMFR.R   | 1,273.6710 | 1,273.6779 | -0.0068 | 1,274.6783 |
| 253   | 262 | 0    | R.KAVNLYVPR.P.R  | 1,356.6472 | 1,356.6609 | -0.0136 | 1,357.6545 |
| 63    | 77  | 0    | R.AVLIDLEPGT.M.S.V.R.A | 1,614.8089 | 1,614.8287 | -0.0198 | 1,615.8162 |
| 63    | 77  | 0    | R.AVLIDLEPGT.M.S.V.R.A | 1,653.7986 | 1,653.8160 | -0.0174 | 1,654.8059 |
| 263   | 276 | 0    | R.LHFFMGMFAPLTS.R.G | 1,669.7885 | 1,669.8109 | -0.0224 | 1,670.7958 |
| 263   | 276 | 0    | R.LHFFMGMFAPLTS.R.G | 1,685.7819 | 1,685.8058 | -0.0239 | 1,686.7892 |
| 337   | 350 | 0    | R.KNSYSREWPNPKS   | 1,723.8311 | 1,723.8569 | -0.0258 | 1,724.8384 |
| 47    | 62  | 1    | R.INVYFDEATGORYR.P.A | 1,856.9165 | 1,856.9217 | -0.0052 | 1,857.9267 |
| 363   | 379 | 0    | R.AGDYQGQFRPONIFGQSGAGNNWAK.G | 2,811.2385 | 2,811.2974 | -0.0589 | 2,812.3563 |
| 104   | 122 | 0    | R.AGPYGQIFRPDNFIFGQSGAGNNWAK.G | 2,811.2385 | 2,811.2974 | -0.0589 | 2,812.3563 |

>Peptide sequences and their corresponding "start" and "end" positions, as well as the number of missing cleavages are indicated.

>Expected and calculated molecular weight (in Da) for each peptide are shown; deltas are differences between expected and calculated values.

>Mass/charge ratio observed in MS spectrum (m/z).

The Mascot score for protein identification and the % of protein sequence coverage given by the identified peptides are also indicated.

T. cruzi β-tubulin, and identified the expected 2 kDa molecular weight shift due to an additional histidine tail in recombinant TcβTUB (Figures 6C,D). Taken together, our data on repertoire analysis indicated that TcβTUB is an immunodominant antigen in T. cruzi infection and that previous immunization with TcβTUB elicited a protective host immune response against infection.

**DISCUSSION**

Several proteins purified from T. cruzi induce protective immune responses. However, few studies identify the major protein antigens targeted by the antibody response of infected hosts. Here we investigated global changes in the antibody diversity of infected mice with T. cruzi. We employed a non-biased immunoblot technique against total parasite extracts. Under these selective conditions, the infection induced significant antibody reactivity against a range of T. cruzi antigens, which could be identified by proteomic analysis of the target proteins. We identified the major protein band recognized by IgG as TcβTUB. We cloned and expressed T. cruzi TcβTUB, and demonstrated that a single immunization with recombinant TcβTUB induced protection against T. cruzi infection. Therefore, our results indicated that repertoire analysis is a valid approach to identify new candidate antigens for a vaccine against Chagas disease.

Infection with T. cruzi can lead to polyclonal B lymphocyte activation, hypergammaglobulinemia, and production of autoantibodies (33, 34). We compared parasite-specific versus non-specific antibody reactivities elicited by infection. Since FasL-deficient mice undergo hypergammaglobulinemia, lymphoproliferation, and autoantibody production (35), we also compared antibody production by infected WT- and FasL-deficient gld mice.
mice. We used a limited set of exogenous and autologous antigens, and this approach did not reveal any evidence of autoreactivity or polyclonal lymphocyte activation. Our results might also reflect the use of Dm28c isolate, as parasite genetic diversity can influence the profile of immunoglobulins produced (36). On the other hand, age-matched FasL-deficient mice showed high levels of natural antibodies against T. cruzi and dsDNA, which did not increase following acute infection. Therefore, lymphocyte activation elicited by T. cruzi is weaker than that induced by the gld mutation. Chronic infection of gld mice increased the production of antibodies against T. cruzi, dsDNA and histone. The reason for increased responses to nuclear autoantigens is unknown but could be related to increased lymphoproliferation precipitated by T. cruzi infection. Production of antibodies against nuclear autoantigens correlates with increased apoptosis (37). Infection with T. cruzi exacerbates lymphocyte apoptosis (38, 39), which is partially dependent on the Fas/FasL death pathway (40–43), but apoptosis is also increased in gld mice (40).

Studies with purified T. cruzi proteins amplify the ability of antibodies to detect antigen by employing recombinant antigens or serum from immunized hosts (44). Instead, our approach employed immunoblots of whole T. cruzi extracts to detect antibody diversity of animals infected with T. cruzi. In this way, we identified by proteomic analysis the most immunogenic T. cruzi proteins serving as targets for the antibody response. A very limited number of protein bands reacted with the sera. The most prominent reactivity was a 50–55 kDa band which colocalized with the most abundant protein band of the T. cruzi extract. This band was excised, digested, analyzed by mass spectrometry, and identified as TcβTUB. All sera from infected mice reacted strongly with TcβTUB. In addition, in spite of already detectable immunological abnormalities, all sera from acutely infected gld mice reacted strongly with TcβTUB. These results suggest a robust response. Two additional bands recognized by host IgG antibodies were identified as T. cruzi pyruvate phosphate dikinase (100 kDa) and T. cruzi Hsp-85 (85 kDa).

We investigated IgG reactivity against autologous heart extracts. Global IgG antibody reactivity against heart polypeptides did not differ between control and infected sera. We detected increased or induced IgM reactivities against a set of cardiac polypeptides were following infection (data not shown). These cardiac polypeptides deserve further characterization.

To investigate the immunogenicity of TcβTUB, we cloned the β-tubulin gene from T. cruzi Dm28c genomic DNA and expressed recombinant TcβTUB in E. coli. Recombinant TcβTUB reacted with monoclonal antibodies against β-tubulin, but not α-tubulin. Chronic infection increased IgG reactivity to TcβTUB, compared with uninfected mice. Immunization of mice with a single dose of TcβTUB in CFA protected against T. cruzi infection, as seen by markedly reduced parasitemia, compared with animals immunized with PBS or BSA. Although our data show a wide range in the levels of anti-β tubulin antibody title in the immunized group, we observed a correlation of host protective response with higher...
IgG reactivity against recombinant TcβTUB. Mice immunized with TcβTUB before infection also gave increased reactivity against parasite and mouse tubulins. Although T. cruzi tubulin reacts with antibodies during infection, to our knowledge it has not been tested previously as an immunogen. Interestingly, immunizations of mice with native T. brucei tubulin or recombinant Trypanosoma evansi beta-tubulin confer broad spectrum protection against infection by African trypanosomes (22, 23, 25). Humoral and cellular mechanisms leading to immunoprotection following immunization with TcβTUB are unknown. TcβTUB is expressed both intracellularly and at the surface of live parasites (45). The majority of monoclonal antibodies reacting against the membrane of live T. cruzi parasites recognize a 50/55 kDa antigen related to tubulin (46). In addition, antibodies reactive against parasite tubulin cross-react with host tubulin (47). Infection with T. cruzi increases the amount and the affinity of naturally occurring antibodies against parasites recognize a 50/55 kDa antigen related to tubulin (47). The majority of monoclonal antibodies reacting against the membrane of live T. cruzi parasites recognize a 50/55 kDa antigen related to tubulin (46). In addition, antibodies reactive against parasite tubulin cross-react with host tubulin (47). Infection with T. cruzi increases the amount and the affinity of naturally occurring antibodies against autologous tubulin and induces novel specificities against tubulin fragments (48, 49). We found slightly increased reactivity against mouse tubulin in the serum of mice immunized with TcβTUB. Additional studies are required to determine whether increased humoral reactivity against autologous tubulin plays any deleterious effect in the host. Taken together, our results indicate that non-biased identification of immunodominant parasite antigens through analysis of antibody diversity of infected hosts is a valid approach to identify candidate antigens for vaccines against Chagas disease.

ETHICS STATEMENT

This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health (USA). The protocol was approved by the Committee on the Ethics of Animal Experiments of the Health Science Center of the Federal University of Rio de Janeiro (CEUA-CCS, Permit Number: IBCCF 062/14), and all efforts were made to minimize suffering.

AUTHOR CONTRIBUTIONS

Conceived and designed the experiments: CGF-de-L, GADR, FM, and DON. Performed the experiments: FM, DON, GADR, RV, CGF-de-L, and MN. Contributed reagents/materials/analysis tools: LML, PMB, RV, LF-de-L, AM, MFL, SMT. AM, NH, GD, and CGF-de-L. Wrote the manuscript: GD, MN, CGF-de-L, and FM.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at https://www.frontiersin.org/articles/10.3389/fimmu.2018.00671/full#supplementary-material.

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