Over a hundred billion bacteria are found in human intestines. This has emerged as an environmental factor in metabolic diseases, such as obesity and related diseases. The majority of these bacteria belong to two dominant phyla, *Bacteroidetes* and *Firmicutes*. Since the ratio of *Firmicutes* to *Bacteroidetes* increases in people with obesity and in various animal models, it has been assumed that phylum composition causes the increase in occurrence of metabolic diseases over the past decade. However, this assumption has been challenged by recent studies that have found even an opposite association of phylum composition within metabolic diseases. Moreover, the gut microbiota affects host energy metabolism in various ways including production of metabolites and interaction with host intestinal cells to regulate signaling pathways that affect energy metabolism. However, the direct effect of gut bacteria on host energy intake, such as energy consumption by the bacteria itself and its effects on intestinal energy absorption, has been underestimated. This review aims to discuss whether increased ratio of *Firmicutes* to *Bacteroidetes* is associated with the development of metabolic diseases, and whether energy competition between the bacteria and host is a missing part of the mechanism linking gut microbiota to metabolic diseases.

**Keywords:** Energy metabolism; Etiology; Gastrointestinal microbiome; Intestinal absorption; Metabolic diseases

**INTRODUCTION**

“Microbiota” is used to refer to all microorganisms that inhabit the human body, including bacteria, fungi, protozoa, and viruses. Among these microorganisms, gut bacteria are the largest population of microbiota, and weigh roughly 1.5 kg of cells in human [1]. The human body carries approximately $3.9 \times 10^{13}$ bacterial cells, with the largest number of cells residing in the large intestine, with $10^{11}$ bacteria cells g$^{-1}$ of wet stool [2,3]. Moreover, 10 million microbial genes have been identified in the human gut, and this number is over 100-fold higher than the number of genes in human genome [4]. The majority of bacteria belong to two dominant phyla, *Bacteroidetes* and *Firmicutes* [2]. Since the ratio of *Firmicutes* to *Bacteroidetes* has increased in obese people and in various animal models [5], phylum composition has been considered to be a potential cause of metabolic disease over the past decade [6]. However, this assumption has been challenged by recent studies that have identified either no, or even a negative association between phylum composition and metabolic diseases in human studies [7-10].

Excessive energy accumulation in the host is a major common factor in metabolic disease pathogenesis (e.g., in obesity and type 2 diabetes mellitus) [11]. Considering that the number of bacterial cells in the human gut, it is not surprising that gut bacteria can alter host energy metabolism and affect meta-
bolic disease pathogenesis. Indeed, many researchers have underlined the role of gut microbiota for host health. Furthermore, the gut microbiota has emerged as an important environmental factor for metabolic disease including obesity and type 2 diabetes mellitus [6,12,13]. Most of these studies have focused on the metabolites produced by bacteria (e.g., short-chain fatty acids [SCFAs] and bile acids) and the signaling pathways that can affect host energy metabolism. However, the direct effects of gut bacteria on host energy intake (e.g., energy consumption by bacteria themselves and its effects on intestinal energy absorption) have been greatly underestimated. Given that such a large number of bacteria live in the gastrointestinal tract [2,3], it is surprising that the number of studies over the past 20 years regarding bacterial energy consumption and the effect on host energy harvest have been so few and far between. This is likely due to the long-held belief that the gut microbiota aids host energy harvest by breaking down indigestible carbohydrates into SCFAs that can be used as energy substrates by the host [6,14]. This energy harvest has been known to supply up to approximately 10% of daily calories when the diet is rich in fiber [15,16]. However, in today’s world we have been faced with the Western diet which is low in fiber [17]. Therefore, it is doubtful whether SCFAs contribute to energy harvest in a meaningful way. Therefore, this review focuses on two issue, firstly, whether bacterial phylum composition is associated with metabolic disease and, secondly, whether energy competition between bacteria and host is another missing piece of the puzzle that is the mechanism(s) of metabolic diseases.

**COMPOSITION OF BACTERIAL PHYLUM AND METABOLIC DISEASES**

Until the late 1990s, the gut microbiota had mostly been studied for its role in influencing the mucosal immune system [18] and increasing energy harvest for the host [14,19]. In the early 2000s, Backhed et al. [20] linked the gut bacterial SCFAs to host energy harvest and suggested the increased ratio of Firmicutes to Bacteroidetes was associated with obesity in leptin-deficient (ob/ob) mice [21,22]. As followed Table 1, the altered composition of bacterial phylum has emerged as a factor which regulates the host metabolism [5,22-24], because germ-free mice which were transplanted with cecal microbiota from

### Table 1. Association between the bacterial phylum composition and metabolic disease in human and rodents

| Disease   | Ratio of Firmicutes to Bacteroidetes | Description                                                                                  | Species | Reference |
|-----------|-------------------------------------|-----------------------------------------------------------------------------------------------|---------|-----------|
| Obesity   | Increase                            | Increase of Bacillus class at the phylum Firmicutes in ob/ob mice                              | Mouse   | [22]      |
| Obesity   | Increase                            | Increase of Clostridiales at the phylum Firmicutes and decrease of Bacteroidales at the phylum Bacteroidetes on high-fat diet | Mouse   | [23]      |
| Obesity   | Increase                            | Fecal microbiota of 12 obese people according to body-weight change                           | Human   | [5]       |
| Obesity   | Decrease                            | Increase of family Prevotellaceae (most of genus Prevotella) at the phylum Bacteroidetes in obese patients | Human   | [30]      |
| Obesity   | No association                      | No association with BMI                                                                      | Human   | [7]       |
| Obesity   | Decrease                            | Decrease of M. smithii and B. animalis but no association with Firmicutes and obese           | Human   | [8]       |
| Obesity   | No association                      | No relationship between the relative populations of gut microbiota and BMI                    | Human   | [9]       |
| NAFLD     | Increase                            | Decrease of Bacteroidetes in NASH patients with obesity (BMI > 30 kg/m²)                     | Human   | [24]      |
| NAFLD     | Decrease                            | Decrease of SCFAs-producing bacteria at the phylum Firmicutes and increase of LPS-producing bacteria in non-obese people with NAFLD | Human   | [10]      |
| NAFLD     | Decrease                            | Increase of genus Prevotella at the phylum Bacteroidetes and genus Escherichia at the phylum Proteobacteria in NASH patients | Human   | [30]      |
| T1DM      | Decrease                            | Increase of Proteobacteria and decrease of Firmicutes with NAFLD                             | Human   | [31]      |
| T2DM      | Decrease                            | Increase of butyrate-producing bacteria in heathy people                                     | Human   | [32]      |
| T2DM      | Decrease                            | Decrease of Clostridia class at the phylum Firmicutes in T2DM patients                      | Human   | [33]      |

* M. smithii, Methanobrevibacter smithii; B. animalis, Bifidobacterium animalis; NAFLD, non-alcoholic fatty liver disease; BMI, body mass index; NASH, non-alcoholic steatohepatitis; SCFA, short-chain fatty acid; LPS, lipopolysaccharide; T1DM, type 1 diabetes mellitus; T2DM, type 2 diabetes mellitus.
obese mice, become obese [25]. Additionally, a series of publications support the association between bacterial phylum composition, obesity, and obesity-related metabolic diseases in a variety of animal models including in humans [4,5] and pigs [26]. Gut microbial community affect to host energy metabolism via digestion of dietary nutrients, for instance, bacteria belonging to phylum Bacteroidetes possess a large number of carbohydrate-active enzyme (CAZyme) [27,28]. The derivatives by CAZyme-producing bacteria such as SCFAs can use energy sources in enterocyte and peripheral tissues; regulate gene expression of host’s metabolism [16,29]. Thus, the association between obesity and bacterial phylum composition has emerged as a major factor underlying metabolic disease pathogenesis.

However, there are some concerns regarding the old hypothesis of energy harvest and phylum composition in the pathogenesis of metabolic diseases. Recently there have been an increasing number of studies which have shown no association between phylum composition and obesity in human stool samples [7-9], and some studies even show an opposite association in patients with other major metabolic diseases, such as non-alcoholic fatty liver disease (NAFLD) [10,30,31], type 1 diabetes mellitus [32], and type 2 diabetes mellitus (Table 1) [33]. The ratio of Firmicutes to Bacteroidetes is decreased in NAFLD patients [10,30,31], in type 1 and the type 2 diabetes mellitus [32,33]. An intake of saccharin induces glucose intolerance in human and in mice, and associated with the decreased ratio of Firmicutes to Bacteroidetes and increased fecal SCFAs content [34]. Furthermore, Akkermansia muciniphila, a member of not Firmicutes nor Bacteroidetes but Verrucomicrobia, is related with reduced adiposity [35] and glucose homeostasis [36] in mice. Taken together, these studies suggest that at the phylum-level of bacterial composition does not consistently indicate the direction of metabolic diseases in animal and human studies in relation to metabolic diseases such as NAFLD and diabetes.

**THE EFFECT OF GUT MICROBIAL METABOLITES IN HOST ENERGY METABOLISM**

Gut bacteria can affect host energy metabolism via its microbial products and metabolites. In this section, we discuss how the gut microbiota and its metabolites affect host metabolism and diseases progression (Fig. 1).

**Short-chain fatty acids**

As mentioned previously, SCFAs produce metabolites by fermentation of carbohydrates. These metabolites include acetate, propionate, and butyrate [6,14]. Moreover, these SCFAs can influence host energy metabolism through the regulation of metabolic pathways or by directly used as an energy source after absorption into host’s body [16]. In this section, we summarize the effect of respective SCFAs on the host energy metabolism. Firstly, the absorbed acetate suppresses weight gain and liver lipid accumulation under a high-fat diet, upregulating the gene expression of fatty acid oxidation through activating AMP-activated protein kinase/peroxisome proliferator-activated receptor α (AMPK/PPARα)-pathway [37,38]. Acetate also inhibits lipolysis in adipose tissue through activating G protein-coupled receptor 3 (GPR43) [39]. Furthermore, acetate decreases the circulating-free fatty acids in plasma by regulating GPR43 in the host [40]. Secondly, the absorbed propionate in the liver raises hepatic glucose production and the tricarboxylic acid cycle [41]. Propionate fermented from soluble fiber activates intestinal gluconeogenesis (IGN) as a substrate of glucose and a gut-brain transmission axis via free fatty acid receptor 2 (FFAR2, also known as GPR43) in the portal vein. They confirmed that absence of IGN using intestinal glucose-6-phosphatase knockout mice is not effective for metabolic effect of soluble fiber [42]. Furthermore, propionate regulates host glucose homeostasis via induction of IGN [42]. Thirdly, butyrate increases skeletal muscle size, inducing transformation into oxidative fiber and mitochondrial function in mice. Moreover, the mice treated with butyrate also have increased adaptive thermogenesis in cold environments via stimulation of the AMPK-peroxisome proliferator-activated receptor gamma coactivator 1α (PGC-1α) pathway, and the mice are protected from insulin resistance and high-fat diet-induced obesity [43,44]. It has also been reported that butyrate decreases the size of adipose tissue in mice by activating adrenergic β3 receptor (ARβ3) and hormone-sensitive lipase (HSL) [45]. Together, the SCFAs which are gut microbiota-derived metabolites can regulate energy metabolism in several tissues in the host including liver, adipose tissue, skeletal muscle, and the brain (the specific part of brain will be discussed in the next section).

**Gut-brain axis: modulation of appetite**

Gut microbiota-derived metabolites can also contribute to modulation of appetite and satiety in the brain through regula-
Fig. 1. Gut microbiota contribute to host energy absorption through the direct/indirect ways. Food intake alters the composition of the gut microbiota and the gut microbiota influences host energy absorption directly or indirectly while the nutrients are traveling in the intestine. The nutrients, which are obtained from a meal, undergo one of three routes during the digestive processes in the intestine, in face of energy metabolism, or the remains of them are excreted from the body. First, the nutrients such as carbohydrates, proteins, and lipids are digested and degraded with digestive enzymes and then absorbed into the host. Second, gut microbiota directly consume nutrients as their energy source. Lastly, the nutrients can be converted into metabolites such as short-chain fatty acids (SCFAs; acetate, butyrate, and propionate), trimethylamine N-oxide (TMAO), and indole propionate (IPA) by the gut microbiota. The gut microbiota producing metabolites are absorbed and circulated in the host. Acetate reduces the fat accumulation by stimulating the AMPK-peroxisome proliferator-activated receptor α (PPARα) pathway and G-protein-coupled receptor 43 (GPR43) in the liver and white adipose tissue (WAT), respectively [37,38]. Butyrate induces intestinal gluconeogenesis (IGN) via cAMP signal and decreases hepatic fat accumulation by GPR43 and releases phosphorylated hormone-sensitive lipase (p-HSL) by adrenergic β3 receptor (ARβ3). Moreover, butyrate activates thermogenesis in brown adipose tissue (BAT) and transformation to oxidative fibers in the skeletal muscle via the AMPK-peroxisome proliferator-activated receptor gamma coactivator 1α (PGC1α) pathway. Propionate, a precursor of glucose in tissues, stimulates IGN and hepatic gluconeogenesis. Gastrointestinal microbiome-producing SCFAs release gut hormones (glucagon like peptide-1 [GLP-1], peptide YY [PYY], cholecystokinin [CCK]) from enteroendocrine cells by stimulating GPR43. The gut hormones regulate appetite and satiety. Besides producing SCFAs, the gut microbiota can also produce TMAO, which is a major risk factor of cardiovascular disease. The IPA, which is produced from tryptophan by the gastrointestinal microbiome, improves insulin secretion. Lipopolysaccharide (LPS), produced by gram-negative bacteria, promote metabolic disease thought induction of inflammation as an endotoxin. Succinate, which fermented from dietary fiber by gut bacteria, activates IGN, resulting improving metabolic disease such as glucose homeostasis. Additionally, gut microbiota can convert primary bile acids into secondary bile acids in the intestine. The bile acids bind G protein-coupled bile acid receptor 1 (TGR5) upon the enteroendocrine cell and function as farnesoid X receptor (FXR) agonist. FFA, free fatty acid; AA, amino acid.
As a result, activation of TGR5 and FXR by bile acid can regulate intestinal signaling for host energy metabolism. Furthermore, the composition of bile acid is associated with gut microbiota and diet.

**Other metabolites contributing host energy metabolism**

Various metabolites other than SCFAs and bile acids can be produced by gut microbiota. Trimethylamine N-oxide (TMAO), which is converted from carnitine or lecithin by gut microbiota, is a risk factor for cardiovascular disease [60]. The lipopolysaccharide (or endotoxin), which is produced from gram-negative bacteria, promotes adiposity resulting obesity via low-grade inflammation [61]. The absorbed TMAO induces hyperglycemia by activating protein kinase R-like endoplasmic reticulum kinase (PERK) [62]. Methane which produced by methanogens such as *Methanobrevibacter smithii* may modulate host energy balance via conversion of hydrogen to methane gas [63]. In addition, the high concentration of plasma indolepropionic acid (IPA) which is produced from tryptophan by gut microbiota, is associated with the lower risk factor of type 2 diabetes mellitus in human [64], and plasma glucose is decreased in IPA-fed mice [65]. Moreover, succinate, which is one of gut microbial metabolites from dietary fiber, improves host glucose homeostasis via induction of IGN as a glucose precursor [66]. Therefore, gut microbiota-producing metabolites can be absorbed through the intestines, which can have an influence on the metabolism of the whole body.

**Bile acid**

Bile acid is known to be facilitator for dietary lipid absorption, and can be converted into secondary bile acids by gut bacteria [54]. The concentration of bile acid in human stool is increased by a high-fat diet [55]. Moreover, cholic acid, which is the major component of primary bile acids, augments the ratio of *Firmicutes* to *Bacteroidetes* of cecal contents in rats [56]. Taken together, bile acid can be affected by the intestinal environment and can alter gut microbiota composition. Moreover, bile acids can regulate intestinal cell signaling by activating receptors such as G protein-coupled bile acid receptor 1 (TGR5) and farnesoid X receptor (FXR) [6], which can modulate host energy metabolism [57]. The regulation of intestinal signaling is considered to be an example of host lipid metabolism [6,58] and the FXR agonist lowers plasma glucose levels by sensitization to insulin [59]. In addition, there is a publication that shows that energy expenditure is increased by bile acid administration in brown adipose tissue and white adipose tissue [57]. As a result, activation of TGR5 and FXR by bile acid can regulate intestinal signaling for host energy metabolism. Furthermore, the composition of bile acid is associated with gut microbiota and diet.

**THE COMPETITION OF BACTERIA AND HOST FOR ENERGY HARVEST**

Gut microbiota consume nutrients for energy first because the gut microbiota and the host share nutrients (Fig. 1). Consequently, the gut microbiota and the host can be in competition for absorption of the same energy sources (e.g., sugar, fat, and protein). This means that the energy absorption into the host can be restricted by specific bacteria even though the appropriate nutritional content is supplied by the hosts diet. Moreover, metabolic diseases are linked to increased intestinal permeability and translocation of bacteria [67]. Diet composition is also highly linked with the proportion of gut microbiota and the diet source can directly change gut microbiota composition in human [68,69], and the altered microbiota contribute to an enhanced harvest of energy from the hosts diet in patients with obesity and metabolic diseases [12,70].
Host and microbiota competition in dietary energy harvest

Recently, we reported a study suggesting that an increased *Firmicutes* to *Bacteroidetes* ratio is not necessarily associated with an increase in intestinal energy harvest and obesity [69]. This study was designed to address the question of whether the increased *Firmicutes* is the cause or the consequence of obesity. By using the common probiotic bacteria, *Lactobacillus rhamnosus* GG (LGG), and a quantitative tracing method with isotope tracers, we showed that the anti-obesity effect of the probiotic bacteria is due to the bacterial-host competition for intestinal energy sources rather than the phylum composition. Furthermore, the increase of these specific gut bacteria, which consume dietary substrates results in a decrease in the remaining nutrients in which, in turn, can reduce the amount of nutrients that can be absorbed by the host. *Lactobacillus* bacteria consume fatty acids during cultivation and delay the intestinal absorption of oleic acids in high-fat diet-fed mice [71]. Chronic oral administration of *Lactobacillus* reduced weight gain, body fat mass, and hepatic lipid accumulation during high-fat diet feeding, without altering dietary calorie intake or fecal calorie excretion [71]. Pyrosequencing data show that the ratio of *Firmicutes* to *Bacteroidetes* is unchanged by LGG treatment, despite the dramatic changes in metabolic phenotypes [71]. Another study also suggests a similar result and shows that *Lactobacillus* strains reduce intestinal free fatty acids, resulting in a loss of body weight [72]. It is important to note that the *Lactobacillus* species are the most widely used probiotics and belong to the phylum *Firmicutes* [73]. Probiotics are thought to confer health benefits against metabolic diseases including obesity [74] and NAFLD [75,76]. Moreover, *Lactobacillus* strains can use exogenous fatty acids to increase their survival in acidic conditions [77]. Therefore, the metabolic phenotypes from probiotic *Lactobacillus* belonged to *Firmicutes* would be worsen by colonizing these bacteria to intestine if the metabolic disease status can simply be estimated by the increased ratio of *Firmicutes* to *Bacteroidetes*.

Furthermore, *Bacteroidetes* is known to be the major bacterium producing acetate (C2) and propionate (C3) in the intestine, which can induce glucose-stimulated insulin secretion and insulin resistance through activation of the parasympathetic nerve system and lipogenesis [50,78]. *Firmicutes* strains are known to mainly produce butyrate (C4), which improves insulin resistance by inhibiting histone deacetylase [79,80]. Given the health benefits of *Lactobacillus* in relation to metabolic diseases, an increase in *Firmicutes* due to a high-fat diet, particularly the increase in *Lactobacillus*, is likely to shift substrate-preferences secondary to the increased dietary lipid supply rather than cause obesity. Taken together, all above studies suggest that the assumption of bacterial contribution in host metabolic diseases has no consistency yet in the view of bacterial phylum composition, and further studies should be elucidated.

CONCLUSION

Since the initial findings of an association between bacterial phylum composition and obesity, a number of studies have examined the phylum-level patterns in relation to metabolic diseases including obesity, NAFLD, and diabetes. However, the results are not ubiquitous, and debate has remained whether alterations in the composition of the gut microbiota are the cause of consequence of metabolic diseases. More specifically, over the phylum-level, individual species level of gut bacteria should be studied to investigate which specific bacteria can affect the development of metabolic diseases. Moreover, gut bacteria are highly diverse due to their ability to relocate and their metabolic responses to different environments (e.g., diet, cold temperature, and antibiotics). Thus, the bacteria themselves need to be studied in further detail including their response to the environment as well as the profiles of metabolites produced and the characteristics of substrate preference regarding energy metabolism.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

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