Cardiovascular magnetic resonance findings in non-hospitalized paediatric patients after recovery from COVID-19

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Abstract

Aims  Our study aimed to investigate the cardiac involvement with sensitive tissue characterization in non-hospitalized children with coronavirus disease 2019 (COVID-19) infection using cardiovascular magnetic resonance (CMR) imaging.

Methods and results  We prospectively enrolled children who recovered from mildly symptomatic COVID-19 infection between November 2020 and January 2021. Patients underwent CMR at 1.5 T (Achieva, Philips Healthcare, Best, the Netherlands) including cine images, native T1 and T2 mapping. Healthy children and paediatric patients with biopsy-proven myocarditis served as control groups. We performed CMR in 18 children with a median (25th–75th percentile) age of 12 (10–15) years, 38 (24–47) days after positive PCR test, and compared them with 7 healthy controls [15 (10–19) years] and 9 patients with myocarditis [10 (4–16) years]. The COVID-19 patients reported no cardiac symptoms. None of the COVID-19 patients showed CMR findings consistent with a myocarditis. Three patients (17%) from the COVID-19 cohort presented with minimal pericardial effusion. CMR parameters of COVID-19 patients, including volumetric and strain values as well as T1 and T2 times, were not significantly different from healthy controls, but from myocarditis patients. These had significantly reduced left ventricular (LV) ejection fraction ($P = 0.035$), LV global longitudinal strain, and left atrial strain values as well as elevated native T1 values compared with COVID-19 patients ($P < 0.001$, respectively).

Conclusions  There was no evidence of myocardial inflammation, fibrosis, or functional cardiac impairment in the studied cohort of children recently. CMR findings were comparable with those of healthy controls. Pericardial effusion suggests a mild pericarditis in a small subgroup. This is pointing to a minor clinical relevance of myocardial involvement in children after mildly symptomatic COVID-19 infections.

Keywords  COVID-19; Paediatric; Myocarditis; Inflammation; CMR

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Background

The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has led to the global pandemic of coronavirus disease 2019 (COVID-19). Recent cardiovascular magnetic resonance (CMR) studies have reported frequent cardiac injury in adults with COVID-19 infections.1–3 Data in paediatric patients are limited and mainly related to the occurrence of multisystem inflammatory syndrome in children associated with SARS-CoV-2 infection (MISc), which presents mostly with a severe onset.4–7 The prevalence and clinical role of cardiac injury in paediatric COVID-19 patients without systemic inflammation is not well studied using sensitive methods for myocardial tissue characterization.
Table 1  Demographics and cardiovascular magnetic resonance findings

|                               | COVID-19 n = 18 | Healthy n = 7 | Myocarditis n = 9 | COVID-19 vs. healthy | COVID-19 vs. myocarditis | Myocarditis vs. healthy |
|-------------------------------|----------------|--------------|------------------|----------------------|-------------------------|------------------------|
| **Demographics**              |                |              |                  |                      |                         |                        |
| Age, years                    | 12 (10;15)     | 15 (10;19)   | 10 (4;16)        | 0.270                | 0.463                   | 0.174                  |
| Sex male, n (%)               | 6 (33)         | 5 (71)       | 4 (44)           | 0.683                | 0.177                   | 0.358                  |
| BSA, m²                       | 1 (1;2)        | 2 (1;2)      | 1 (1;2)          | 0.657                | 0.900                   | 0.758                  |
| Time symptom onset—CMR, days | 42.0 (37.8–54.0) | n.a.        | 7.0 (5.0–16.0)   | n.a.                 | 0.013                   | n.a.                   |
| **Functional parameters**     |                |              |                  |                      |                         |                        |
| LV EDVi, mL/m²                | 79 (75;87)     | 80 (78;86)   | 93 (77;172)      | 0.574                | 0.053                   | 0.091                  |
| LV ESVi, mL/m²                | 32 (27;34)     | 31 (28;38)   | 48 (28;139)      | 0.574                | 0.053                   | 0.142                  |
| LV EF, %                      | 62 (58;67)     | 62 (54;67)   | 50 (19;64)       | 0.534                | 0.035                   | 0.142                  |
| RV EDVi, mL/m²                | 79 (75;83)     | 80 (71;86)   | 69 (58;99)       | 0.929                | 0.375                   | 0.606                  |
| RV ESVi, mL/m²                | 29 (25;34)     | 28 (27;37)   | 30 (21;41)       | 0.836                | 0.900                   | 0.758                  |
| RV EF, %                      | 63 (60;67)     | 64 (58;67)   | 58 (49;67)       | 0.883                | 0.322                   | 0.351                  |
| Pericardial effusion, n (%)   | 3 (17)         | 0 (0)        | 6 (67)           | 0.534                | 0.026                   | 0.011                  |
| Wall motion abnormalities, n (%) | 0 (0)         | 0 (0)        | 9 (100)          | n.a.                 | <0.001                  | <0.001                 |
| **Strain values**             |                |              |                  |                      |                         |                        |
| Endocardial LV longitudinal strain, % | –27 (–30;–25) | –25 (–31;–22) | –21 (–24;–12) | 0.220 | <0.001 | 0.054 |
| Myocardial LV longitudinal strain, % | –26 (–28;–25) | –25 (–29;–22) | –20 (–21;–12) | 0.297 | <0.001 | 0.014 |
| Endocardial LV circumferential strain, % | –30 (–34;–29) | –31 (–34;–25) | –21 (–32;–9) | 0.495 | 0.059 | 0.252 |
| Myocardial LV circumferential strain, % | –24 (–25;–22) | –22 (–23;–19) | –13 (–22;–8) | 0.034 | <0.001 | 0.005 |
| Endocardial RV longitudinal strain, % | –27 (–30;–23) | –25 (–29;–22) | –26 (–30;–10) | 0.458 | 0.461 | 0.867 |
| Myocardial RV longitudinal strain, % | –27 (–29;–22) | –25 (–28;–22) | –24 (–29;–11) | 0.615 | 0.397 | 0.779 |
| LA strain, %                  | 46 (41;62)     | 48 (37;56)   | 32 (22;–36)      | 1.000                | <0.001                  | 0.002                  |
| RA strain, %                  | 43 (33;53)     | 37 (24;57)   | 47 (17;–54); n = 7 | 0.458 | 0.574 | 0.805 |
| **Mapping**                   |                |              |                  |                      |                         |                        |
| T1 native, ms                 | 1034 (1005;1062) | 1050 (1031;1071) | 1151 (1090;1238) | 0.357 | <0.001 | 0.001 |
| T2, ms                        | 48 (47;50)     | 56 (49;70)   | n = 4            | n.a.                 | 0.118                   | n.a.                   |

BSA, body surface area; CMR, cardiovascular magnetic resonance imaging; EDVi, indexed end-diastolic volume; EF, ejection fraction; ESVi, indexed end-systolic volume; LA, left atrial; LV, left ventricular; n.a., not applicable; RA, right atrial; RV, right ventricular.

Values are given as n (%) or median with median (25th–75th percentile). For comparison of the continuous variables, Mann–Whitney U-test was used, and for categorical variables, Pearson’s χ² or Fisher’s test were used; a P-value < 0.05 was considered significant. For incomplete set of data, n represents the number of subjects included in the analysis. Bold P-values represent significant P-values.
Aims

Our study aimed to investigate the presence of cardiac involvement including functional impairment, myocardial inflammation, and fibrosis using CMR without contrast-enhancement and sensitive CMR methods for myocardial tissue characterization in non-hospitalized children with COVID-19.

Methods

We prospectively enrolled children who recovered from mildly symptomatic COVID-19 infection between November 2020 and January 2021. All infections were diagnosed by positive polymerase chain reaction (PCR). The presence of MISc was an exclusion criterion. No cardiac diagnostics have been performed before enrolment in relation to the COVID-19 infection. Patients underwent CMR at 1.5 T (Achieva, Philips Healthcare, Best, the Netherlands) including cine images, native T1 and T2 mapping. Images were analysed using commercially available software, mapping parameters by QMap RE Version 2.0, and global longitudinal strain (GLS) by QStrain (Medis Medical Imaging Systems, Leiden, the Netherlands).

Figure 1. CMR findings in a 12-year-old girl 2 months after positive SARS-CoV2 PCR. Upper row: Cine image of the left ventricle (LV) in radial long-axis (LAX) view with corresponding endomyocardial longitudinal strain (LS) in %. LV ejection fraction was 60%, LV LS – 25%. Middle row: Cine image of the right ventricle (RV) in LAX view with corresponding endomyocardial LS of – 24%. Lower row: Cine image of the LV in short-axis (SAX) view with corresponding T1 and T2 maps (T1, 1001 ms; T2, 47 ms).
In addition, children who underwent cardiomyopathy screening due to a family history of cardiomyopathy, but without any pathologies and pathogenic cardiomyopathy variants were included as controls. Paediatric patients with biopsy and CMR proven myocarditis enrolled within the MYKKE Registry served as a reference with myocardial inflammation. Parents or legal guardians gave written informed consent. Ethical approval was obtained from the responsible ethics committee.

Results

We performed CMR in 18 children recovered from COVID-19 infection with a median (25th–75th percentile) age of 12 (10–15) years, 38 (24–47) days after positive PCR test, and compared them with 7 healthy controls [15 (10–19) years] and 9 patients with myocarditis [10 (4–16) years]. CMR was performed significantly earlier after symptom onset in the myocarditis group compared with COVID-19 patients ($P = 0.013$, Table 1). Demographics and CMR parameters are presented in Table 1. The COVID-19 patients reported mild symptoms including fatigue (61%), fever (56%), respiratory symptoms (50%), loss of smell and taste (44%), gastrointestinal symptoms (39%), and dyspnoea (17%).

None of the COVID-19 patients showed CMR findings consistent with a myocarditis based on the updated Lake Louise Criteria. Findings of a 12-year-old female COVID-19 patient are displayed in Figure 1. Three patients (17%) from the COVID-19 cohort presented with minimal pericardial effusion. CMR parameters of COVID-19 patients, including volumetric and strain values as well as T1 and T2 times, were not significantly different from healthy controls. In contrast, myocarditis patients more often showed pericardial effusion (67% vs. 17%; $P = 0.026$) and wall motion abnormalities ($P < 0.001$) and had significantly reduced left ventricular (LV) ejection fraction ($P = 0.035$), LV GLS, and left atrial strain as well as elevated native T1 values compared with COVID-19 patients ($P < 0.001$, respectively; Table 1). See Figure 2 as an overview of different CMR findings between healthy controls, the COVID-19, and the myocarditis cohort.

Conclusions

In the studied cohort of children, recently recovered from mildly symptomatic COVID-19 infections, no evidence of myocardial inflammation, fibrosis, or functional cardiac impairment was found. CMR findings were comparable with those of healthy controls but clearly different to findings in myocarditis patients. With a longer time difference between...
symptom onset and CMR in COVID-19 patients, a cardiac involvement in the first 4 weeks cannot be ruled out. Especially myocardial oedema might have not been detected in our study more than 1 month after symptom onset. In children, the cardiac involvement after mild COVID-19 infections was lower compared with studies in adults with mild or moderate COVID-19 infections, where high frequencies of CMR manifestations (30–78%) as ongoing myocardial inflammation, positive late gadolinium enhancement, and LV dysfunction were reported. The minimal pericardial effusion might be explained by the postulated cytokine storm, which seems not to be necessary in this patient group.13

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Conflict of interest

None declared.

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