CD207+/langerin positive dendritic cells in invasive and in situ cutaneous malignant melanoma

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Abstract

Introduction: Dendritic cells are crucial for cutaneous immune response. Their role in melanoma progression is however a matter of controversy.

Material and methods: The number of dendritic cells within epidermis and in peri- and intratumoral location was analyzed using CD207 immunostain in 17 cases of in situ and 25 case of invasive melanoma.

Results: Average peritumoral CD207+ cells count was 22.88 for all cases, 17.94 for in situ lesions and 26.24 for invasive cases. Average epidermal CD207+ cells count was 164.47 for all cases, 183.00 for in situ lesions and 150.78 for invasive cases. In case of invasive melanomas, peritumoral CD207+ cells count was positively correlated with Breslow stage ($R = 0.59$) mitotic activity within the tumor ($R = 0.62$). Invasive cases with regression showed higher intratumoral and epidermal CD207+ cells count than the ones without (275.00 vs. 95.32 and 173.20 vs. 148.35) but lower peritumoral CD207+ cells count (17.60 vs. 27.26). Invasive cases with ulceration showed higher intratumoral and peritumoral CD207+ cells count than the ones without ulceration (220.08 vs. 55.67 and 44.17 vs. 9.69).

Conclusions: CD207+ cells play a role in both progression and regression of melanoma but their exact role needs further studies.

Key words: malignant melanoma, dendritic cells.

Introduction

The incidence of cutaneous malignant melanoma (CMM) in Europe has steadily increased over the last decades [1–3]. In 2012, CMM ranked the seventh among the most frequently diagnosed malignancies in the European Union, constituting 3% of all new cancer cases (excluding non-melanoma skin cancers) [4]. Moreover, in many EU countries the mortality of CMM has been continuously increasing [5] and despite developments in melanoma treatment, including BRAF or MEK inhibitors as well as anti-CTLA4 monoclonal antibodies [6, 7], clinicians are still powerless in the most advanced cases. Consequently, there is still much to be discovered and scientists should go further in searching for new prognostic, predictive and therapeutic tools.

In recent years, many authors have demonstrated that cancer development and progression is, to some extent, controlled by the host’s immune system [8–11], which suggests that immunological biomarkers might be good indicators of patient’s outcome. As CMM is considered to be a highly immunogenic tumour [12], the utilisation of immune cells as prognostic determinants seems to represent a very promising approach. Recently, there has also been a growing interest in therapeutic potential of immune cell-based vaccines against various human malignancies [13–16], including CMM [17, 18]. Especially dendritic cells (DCs) have been widely investigated, since they are believed to be capable of inducing tumour-specific immunity [19] and, thus, could be used as prognostic biomarkers as well as potential therapeutic targets.

Dendritic cells are the most potent, highly specialised antigen-presenting cells (APCs), which links innate and adaptive immunity. The DCs constitute a complex system of cells found in peripheral tissues as well as immunological organs, and their presence was also documented in a vast range of human malignant tumours [20–25]. Response to cancer cells involves the capture of foreign antigens by immature DC subsets, their entailing maturation, migration to the lymph nodes and presentation of antigen peptides to T cells via major histocompatibility
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Complex (MHC) molecules. However, depending on the biological context, DCs can either boost an effective host reaction to cancer cells or maintain local tissue tolerance that facilitates tumour growth [26, 27]. Distinct DC subtypes, each having a different phenotype, location and function can be identified on the basis of cell surface markers and intracellular molecules. The normal human skin contains epidermal Langerhans cells (LCs) and dermal DCs, while under pathological conditions, plasma-cytoid as well as inflammatory dendritic epidermal cells happen to be detected [28].

Despite many studies on the issue of dendritic cells in various cancers, the underlying tumour-host interactions involving certain subsets has not yet been clearly elucidated, which prevents the translation of these immune factors into clinical use. Additionally, in the light of the importance of DCs, there is a very limited number of studies investigating the correlation between their density and histological characteristics of CMM [29–31].

**Aim**

The objective of the present study was to evaluate the infiltration of DCs expressing CD207/langerin, which is a C-type lectin displaying mannose-binding specific-ity, involved in ligand internalisation [32]. The study was conducted on a subset of 42 cases, which were obtained from patients diagnosed with melanoma in situ or invasive malignant melanoma of the skin. The number of DCs was analysed in relation to a degree of CMM progression and spatial location of DC infiltration (intratumoral, peritumoral, and epidermal). Evaluation of these parameters might be helpful in explaining the prognostic and predictive value of DCs in the progression of CMM as well as their potential implementation in immunotherapeutical protocols.

**Material and methods**

**Material**

We used cutaneous tissue samples excised from 42 patients who were diagnosed with melanoma in situ or invasive cutaneous melanoma of the skin between 2009 and 2013. The paraffin-embedded samples were obtained from the archives of the Department of Pathomorphology, while data concerning the subtype of lesion, melanoma stage (described using Breslow depth), presence of ulceration, mitotic activity and regression features were collected from the database of the Department.

**Immunohistochemistry**

Immunohistochemistry was performed using the anti-human CD207 (Langerin) monoclonal antibody (clone 12D6, Novocastra, Leica Biosystems, Germany), dilution 1 : 100, incubation time 60 min. Cutaneous tissue samples were stained manually and processed according to the protocol used on a routine basis in the laboratory of the Department of Pathomorphology. The selected paraffin-embedded tissue blocks were cut into 4 μm thick sections, mounted on SuperFrost glass slides (ThermoScientific, USA) and dried in an incubator for 12 h at 34°C. The obtained slides were deparaffinized, dehydrated and then incubated in 3% H2O2 solution for 10 min to block endogenous peroxidase activity. Antigen retrieval was performed by immersing the slides in citrate buffer (pH 6.0; 0.01 M) and subjecting them to 97°C in a water bath for 40 min. Polyclonal secondary antibodies conjugated to horseradish peroxidase (Ultra Vision Quanto Detection System, Lab Vision, ThermoScientific, USA), using DAB (3,3'-diaminobenzidine) as chromogen, were applied to visualize the obtained antigen-antibody complexes. Cell nuclei were stained with hematoxylin to enhance contrast in tissue sections.

**Evaluation of immunostaining**

For each case quantitative assessment of CD207/langerin-positive cells was performed under light microscopy. The positively stained cells (brown membranous staining) were counted at 400× magnification. For evaluation of intratumoral, peritumoral and intraepidermal cells, the CD207/langerin-positive cells were counted in tumour mass, along the tumour border and in overlying epidermis, respectively, within the whole section; then the cell count was recalculated per mm² of tissue (depending on the number of investigated fields). The obtained results were entered into an Excel spreadsheet (Microsoft Corporation, USA).

**Statistical analysis**

Statistical data analysis was performed using Statistica 10 software (StatSoft, USA). The difference between the number of CD207/langerin-positive intratumoral, peritumoral or intraepidermal dendritic cells and type of lesion (invasive vs. in situ) were evaluated by using the Mann-Whitney U-test. Spearman rank and gamma correlation tests were performed to assess correlations between the numbers of CD207/langerin-positive cells (intratumoral, peritumoral and intraepidermal) and tumour stage (Breslow depth), presence of ulceration, mitotic activity and regression features. To evaluate the relationships between the content of CD207/langerin-positive cells and type of lesion, presence of ulceration or features of tumour regression, the t-test was performed. P-values < 0.05 were considered significant.

**Results**

Among 42 cases investigated in our study, 25 (59.5%) were diagnosed as invasive melanoma and 17 (40.5%) as melanoma in situ.
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For all cases, average peritumoral CD207/langerin-positive cell count was 22.88 (SD = 28.15) and average epidermal CD207/langerin-positive cell count was 164.47 (SD = 101.15) (Figures 1 and 2).

There were statistically significant correlations between the number of peritumoral CD207/langerin-positive cells and Breslow depth ($R = 0.60; p = 0.002$), mitotic activity ($R = 0.62; p = 0.001$) as well as the presence of ulceration ($R = 0.58; p = 0.002$) in invasive melanoma cases. There was a significantly ($p = 0.002$) higher number of peritumoral CD207/langerin-positive cells in ulcerated invasive melanoma samples (mean: 44.17, SD = 35.22) in comparison with invasive melanomas without ulceration (mean: 9.69, SD = 9.40).

There was no statistically significant ($p > 0.05$) difference in peritumoral or intraepidermal CD207/langerin-positive cell count between invasive and in situ melanoma groups. Nonetheless, it is noteworthy that we observed a higher number of CD207/langerin-positive cells in the peritumoral area of invasive (mean: 26.24, SD = 30.36) than of in situ lesions (mean: 17.94, SD = 24.59).

Contrary to the above results, the number of intraepidermal CD207/langerin-positive cells in invasive melanomas was lower (mean: 150.78, SD = 114.18) than in in situ lesions (mean: 183.00, SD = 79.85) (Figures 3 and 4).

Among invasive melanomas, the average number of intratumoral CD207/langerin-positive cells (mean: 275.00, SD = 483.84) and intraepidermal cells (mean: 173.20, SD = 156.57) was higher in tumours with regres-
sion than in tumours without regression (mean: 95.32, SD = 134.34 – for intratumoral, mean: 148.35, SD = 106.41 – for intraepidermal) (Figures 5 and 6).

By contrast, the peritumoral CD207/langerin-positive cell count was lower in invasive melanomas with regression (mean: 17.60, SD = 35.05) than in cases without regression (mean: 27.26, SD = 30.05). However, for all cases the differences were not statistically significant.

Discussion

Langerin (CD207) is a C-type lectin, binding glycosylated ligands in a calcium-dependent manner, and has many roles, such as in cell adhesion, endocytosis, natural killer-cell target recognition and dendritic-cell activation [33]. So far, it was generally assumed that Langerhans cells are the only cells expressing langerin [32, 34, 35], but now there is a growing body of evidence indicating that langerin expression is not confined to LCs exclusively as it is also found in dermal DCs [36–39] and other organs, such as lungs, liver, spleen and lymph nodes [36, 40]. In mouse skin, the population of DCs comprises CD207/langerin-positive LCs, forming a network in epidermis, as well as CD207/langerin-positive and CD207/langerin-negative DC subsets residing in the dermis [37–39]. As far as human dermis is concerned, one study confirming the existence of CD207/langerin-positive dermal DCs has been published recently [36]. One of the main reasons why previous studies have evaluated dermis as langerin-negative compartment [32, 41] might be the fact that CD207/langerin-positive DCs found in the steady state in dermis are rare, and if they were ever noticed, they were regarded as LCs in transit through the dermis on their way to the lymph nodes [39, 42]. In addition, some in vitro studies have demonstrated that the production of langerin in human progenitor cells and monocytes can be induced by a variety of factors, such as transforming growth factor β (TGF-β), granulocyte-macrophage colony-stimulating factor (GM-CSF), tumor necrosis factor α (TNF-α) and bone morphogenetic protein 7 (BMP7) [43–45], which also supports the view that langerin might be expressed by distinct dermal DC subtype, not related to epidermal LCs. Although these observations need to be clarified, they have to be considered in the interpretation of the presented results.

Tumour thickness, mitotic rate and ulceration are the most dominant prognostic factors for clinical outcome of malignant melanoma [46]. Our study revealed a statistically significant positive correlation between each recognised prognostic feature and the infiltration of CD207/langerin-positive DCs in the peritumoral area of invasive CMM. Since the increased tumour thickness, higher mitotic rate and presence of ulceration are indicators of decline in survival rates, our findings may suggest that a higher number of CD207/langerin-positive DCs at the border of the tumour favours disease progression and is associated with worse prognosis. This conclusion would be consistent with the observation that CD207/langerin-expressing dendritic cells in human tissues are related to immature CD1a/CD1c-positive cells [36], and thus presumably play a role in tolerance rather than in immunity, as immature DCs are unable to effectively present processed foreign antigens to T cells. It is, however, still unknown whether the higher infiltration of immature DCs in peritumoral area is mostly the consequence of their impaired maturation or is solely caused by the excessive migration to the pathologically altered site, and the progression of the disease should be rather ascribed to the inadequate functioning of mature dendritic cells. If a case of massive migration was considered to be more probable, it would be recommended to investigate local factors, which may possibly support the recruitment of
CD207+/langerin-positive dendritic cells in CMM, as e.g. CCL20 chemokine was proposed to be crucial for CD207+/langerin-positive DCs migration to EBV-associated tumours [47]. There are also substantial ambiguities resulting from the lack of sufficient information about the pathophysiological processes involved in CMM progression, which makes the speculation about the role of DCs in the disease progression even more complicated; for instance, very little is known about the underlying biological mechanisms of CMM ulceration [48]. Literature data concerning the significance of CD207/langerin-positive cells in CMM are very scarce and findings vary considerably between papers. In a study conducted on 185 primary melanomas, Gerber et al. did not show any significant influence of CD207/langerin expression on the clinical outcome [49]. Moreover, given the observation that in human tissues CD207/langerin-positive DCs were found to be closely related to CD1a-positive subsets, as described above, it is justified to mention that infiltration of CD1a-positive DC peritumoral area of CMM was observed to correlate positively with decreased tumour thickness [50], which might be interpreted as being contrary to our findings. In addition, we have found a significant inverse association between the number of CD207/langerin-positive DCs and the presence of melanoma regression features, whose presence is traditionally regarded as an indicator of poor prognosis. Nonetheless, the impact of tumour regression is still unclear, as some studies revealed that this feature does not have any independent prognostic value [51], while others demonstrated a higher metastatic rate in CMMs with extensive regression [52]. Taken together, there are still many unresolved questions, which prevents us from drawing valid conclusions on whether CD207/langerin-positive cells play any specific role in inducing or regulating an immune response in the progression of CMM, and thus their correlations with established prognostic factors must be interpreted with caution.

We found a substantially higher average number of intratumoral and epidermal CD207/langerin-positive DCs in invasive melanomas with regression. With regard to the differences in the spatial distribution of CD207/langerin-positive DCs we can conclude that despite the enhanced intranestal recruitment of DCs in more advanced stages, the immune interactions are ineffective to elicit an appropriate antitumor response. It is suggested that some melanoma-derived factors, such as TGF-β and interleukin 10 (IL-10), found in melanoma cell nests, might adversely influence migration of dendritic cells, their differentiation and function [53–55]. Neagu et al. demonstrated that the presence of LCs within melanoma tumour mass correlates with the Breslow index but not with ulceration; yet the general intratumoral LC content was described to be rather scarce [8]. Regarding other human malignancies, the infiltration of CD207/langerin-positive DCs was also observed in Hodgkin lymphoma, nasopharyngeal carcinoma, oral squamous cell carcinoma and breast cancer [47, 56, 57]. However, in the latter case the infiltration by immature CD207/langerin-positive DCs was found in only one-third of the breast cancer tissues and did not correlate with clinico-pathological data. In our cohort of melanoma samples, we have also observed a higher intratumoral langerin expression compared to the peritumoral area in invasive CMM. Assuming again that langerin is mostly present on immature dendritic cells and is related to CD1a-positive subsets, this observation is in agreement with the results obtained by other authors investigating various human malignancies, as it is generally suggested that immature dendritic cells are located within tumour nests, while mature subsets reside in peritumoral compartment [29, 58]. Conversely, the intraepidermal infiltration of CD207/langerin-positive DCs was more pronounced in non-invasive lesions of our study, which is in concert with the findings of other authors showing a reduced number of intraepidermal DCs in invasive melanoma tissues [53, 59]. The possible explanations of the depletion of DCs in the epidermis during the evolution of CMM include necrosis, regression and scarring in more advanced stages of the tumour [53] as well as the presence of basement membrane along with the lack of epidermal own blood vessels. We represent the view that in order to better understand the role of CD207/langerin-positive DCs in CMM evolution, the issue of their uneven distribution between invasive and non-invasive melanomas should be revisited on a larger sample of cases.

Conclusions

Our results may shed light on the relevance of CD207/langerin-positive DCs at different stages of CMM and clarify some of their associations with traditional risk prediction based on histopathological characteristics. In interpreting these findings we should, however, remember that the evaluation of density and location of DCs relied on staining for CD207/langerin exclusively, thus, we were unable to analyse the whole spectrum of DC subsets and their interactions with other immune cells involved in melanoma carcinogenesis. In order to draw a complete picture of the role of DCs in melanoma control, the thorough analysis of the dendritic cell family, including numerous markers, as well as other presumably contributing microenvironmental factors, such as cytokines, is required. Moreover, the inclusion of melanoma metastases might also bring some added value to the evaluation of DCs in progression of the disease. In addition, we should also bear in mind that melanoma patient’s survival can vary significantly among individuals within the same tumour stage and widely accepted classifications (e.g. Breslow depth, TNM) may not utterly correspond with the ongoing immunological processes. Thus, only a prospective analysis of the patient’s clinical
outcome will allow us to better understand the predictive and prognostic significance of CD207/langerin-positive tumour-infiltrating DCs. Although the role of DCs infiltrating CMM demands further elucidation, we believe that our study gives a valuable insight into the role of CD207/langerin-positive DCs in the progression of CMM and might prove useful in incorporating immune scoring in clinical settings.

Conflict of interest

The authors declare no conflict of interest.

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