REVIEW ARTICLE

Urinary tract infection in pediatrics: an overview∗†

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Urinary tract infection; CAKUT; Risk factors; Renal ultrasonography; Antibiotic prophylaxis; Chronic kidney disease

Abstract

Objective: This review aimed to provide a critical overview on the pathogenesis, clinical findings, diagnosis, imaging investigation, treatment, chemoprophylaxis, and complications of urinary tract infection in pediatric patients.

Source of data: Data were obtained independently by two authors, who carried out a comprehensive and non-systematic search in public databases.

Summary of findings: Urinary tract infection is the most common bacterial infection in children. Urinary tract infection in pediatric patients can be the early clinical manifestation of congenital anomalies of the kidney and urinary tract (CAKUT) or be related to bladder dysfunctions. E. coli is responsible for 80–90% of community-acquired acute pyelonephritis episodes, especially in children. Bacterial virulence factors and the innate host immune systems may contribute to the occurrence and severity of urinary tract infection. The clinical presentation of urinary tract infections in children is highly heterogeneous, with symptoms that can be quite obscure. Urine culture is still the gold standard for diagnosing urinary tract infection and methods of urine collection in individual centers should be determined based on the accuracy of voided specimens. The debate on the ideal imaging protocol is still ongoing and there is tendency of less use of prophylaxis. Alternative measures and management of risk factors for recurrent urinary tract infection should be emphasized. However, in selected patients, prophylaxis can protect from recurrent urinary tract infection and long-term consequences. According to population-based studies, hypertension and chronic kidney disease are rarely associated with urinary tract infection.

Conclusion: Many aspects regarding urinary tract infection in children are still matters of debate, especially imaging investigation and indication of antibiotic prophylaxis. Further longitudinal studies are needed to establish tailored approach of urinary tract infection in childhood.

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Introduction

Urinary tract infections (UTIs) are among the most common bacterial infections in children. Up to 8% of children will experience at least one UTI between the ages of 1 month and 11 years, and up to 30% of infants and children experience recurrent infections during the first six to 12 months after initial UTIs. In the United States, there are about 1.5 million pediatric ambulatory visits annually for UTIs. The overall US health care costs for management and treatment of UTI in 2013 was $630 million. UTIs cause short-term morbidity such as fever, dysuria, and flank pain, and may also result in long-term renal injury, such as permanent kidney scarring.

A fundamental issue in the topic of the management of UTI in children is that a single episode may be the sentinel event for an underlying renal abnormality and in 30% of children with congenital anomalies of the kidney and urinary tract (CAKUT), UTI can be the first sign. Therefore, since the 1960s, the management of UTI in children has been based on the conception that recurrent episodes, particularly with vesicoureteral reflux (VUR), increase the risk of chronic kidney disease (CKD), hypertension, and ultimately end-stage renal disease (ESRD). Consequently, the guidelines on the management of UTIs in children are elaborated on the assumptions that prompt diagnosis and treatment and comprehensive imaging investigation might prevent an unfortunate chain of deleterious events and long-term renal injury.

Over the last two decades, the scenario of the management of children with a febrile UTI has changed. The old model proposed that all children with UTI were to be investigated using ultrasound (US), a micturition cystourethrogram (MCUG), and some form of nuclear imaging, such as dimercaptosuccinic acid (DMSA). The aim of these investigations was to identify all children with CAKUT, especially those with VUR and renal scarring. In addition, children with a febrile UTI were hospitalized for intravenous antibiotics and children with VUR of any grade were treated with prophylactic antibiotics. Several randomized clinical trials and prospective cohort studies have called into question all these old paradigms. The results of this new body of knowledge led to a review of the existing guidelines once they failed to show any evidence of a change in clinical evolution driven by antibiotic prophylaxis or imaging tools. Moreover, improved prenatal US has revealed that major kidney damage in children is frequently related to the presence of congenital hypoplasia, associated with urologic abnormalities. Consequently, recent guidelines on the management of UTIs in children have shown a shift from aggressive imaging investigation and the indiscriminate use of pro-
phyllactic antibiotics to a more restrictive and targeted approach.\textsuperscript{16,17}

Despite these advances, the management of UTIs in the pediatric population remains challenging and controversial. Diagnosis, treatment, and follow-up of children with UTI are important issues for general pediatricians and involve multiple decisions.\textsuperscript{18} It is consensual that a correct diagnosis, appropriate treatment, and a subsequent selected imaging investigation in children with UTI is still pivotal because of the association between UTI, underlying urological abnormalities, and its consequences. Therefore, a prompt diagnosis and immediate initiation of treatment remain important in preventing long-term renal damage. However, it must be pointed out that establishing a suitable approach and identifying children with risk of renal parenchymal damage is not a simple task.

This review article discusses recent recommendations for the diagnosis, treatment, prophylaxis, and imaging of UTI in children based on evidence, and when this is lacking, based on expert consensus.

Source of data

Data were obtained independently by two authors who carried out a comprehensive and non-systematic search in the PubMed, Embase, LILACS, Cochrane, Scopus and Scielo databases. Search strategies included Medical Subject Heading terms for "urinary tract infection," "CAKUT," "renal scarring," "vesicoureteral reflux," "renal ultrasonography," "renal scintigraphy," "antibiotic prophylaxis," and "chronic kidney disease." No time or language restrictions were established. The search emphasized recent consensus statements, guidelines, meta-analyses, systematic reviews, randomized clinical trials, and prospective cohort studies. The publications were critically selected by the authors.

Summary of findings

Pathogenesis of UTI

The role of bacteria

The urinary tract is normally sterile, except for the distal part of the urethra. Physiologically, the periurethral area has bowel bacteria. In healthy young girls the predominant bacteria is \textit{Escherichia coli} (\textit{E. coli}), whereas, in boys, after the first 6 months of life, \textit{Proteus mirabilis} predominates. On the other hand, bowel bacteria do not usually form the periurethral flora of older children. It should be pointed out, however, that colonization with Gram-negative bacteria generally precedes the occurrence of UTI.\textsuperscript{19} In some occasions, the prescription of broad-spectrum antibiotics for other infections may produce changes in the normal flora.\textsuperscript{20}

\textit{E. coli} is responsible for 80–90% of community-acquired acute pyelonephritis episodes, especially in children. Less common uropathogenic bacteria include \textit{Proteus mirabilis}, \textit{Klebsiella} spp., and \textit{Staphylococcus saprophyticus}.\textsuperscript{21,22} Infectious agents of UTI acquired during hospitalization depend on the hospital environment and underlying host factors.\textsuperscript{21,22} Bacterial virulence factors and the innate host immune systems may contribute to the occurrence and severity of UTI.\textsuperscript{23-27}

UTI may occur via two routes: hematogenic and ascendant. The hematogenic route is typical in newborns, while the ascendant route characteristically develops after the neonatal period. In newborns, UTI may manifest as sepsis, largely with nonspecific clinical features, including anorexia, vomiting, poor sucking, irritability, lethargy, convulsions, pallor, hypothermia and, sometimes, jaundice.\textsuperscript{28} As with most infections, in this age group, there is high probability of bacteremia and high rate of mortality (around 10%) due to the spread of infection to other sites, leading to meningitis, for instance.\textsuperscript{28,29} The ascendant route comprises the migration, fixation, and proliferation of uropathogenic bacteria in the urinary tract. Uropathogenic bacteria may reside for long periods in the gastrointestinal tract before spreading to the periurethral area. After spreading via the perineum to the periurethral area, bacteria ascend the urinary tract against urine flow, and establish infection by means of several mechanisms. The main mechanisms include fimbrae that promote adhesion to urothelial cells, flagella-mediated motility, resistance to antibacterial defenses, and other adaptation strategies.\textsuperscript{23,26,27}

In this regard, the subtype of \textit{E. coli} strain that causes acute pyelonephritis in healthy children has genes that confer virulence, forming the so-called "pathogenicity islands".\textsuperscript{30,31} The sequential activation of these genes increases host tissue attack and bacterial survival. The presence of fimbrae promotes bacterial adhesion to the mucosa that facilitates tissue attack\textsuperscript{32} by increasing the exposure to other virulence factors, such as hemolysin and lipopolysaccharide (LPS). These toxins secreted by \textit{E. coli} may affect cellular functions or induce cell death. Uropathogenic strains of \textit{E. coli} can be identified by the presence of surface antigen expression (OKH serotypes) or of surface expression of P-fimbrae.\textsuperscript{33,34} Different types of fimbrae recognize different oligosaccharide receptor epitopes. Type 1 fimbrae bind to mannosylated epitopes present in the Tamm–Horsfall glycoprotein, in secretory immunoglobulin A (IgA), in bladder cell uroplakins, or in integrin molecules.\textsuperscript{35-37} S-fimbrae bind to receptors on sialylated glycoproteins and glycolipids, while P-fimbrae recognize Galα1-4Gal epitopes in the glycolipids, which are antigens in the P blood group system.\textsuperscript{38}

Other virulence factors are LPS, capsular polysaccharide, and hemolysin. LPS is an endotoxin of Gram-negative bacteria that contains lipid A anchored in the outer membrane, as the component responsible for the toxic effects including fever and acute phase response. Other components of LPS are the oligosaccharide core and the repeating oligosaccharide that determines the O-antigen. LPS activates toll-like receptor 4 (TLR4) signaling, after binding to soluble or cell surface-associated CD14.\textsuperscript{39,40} Capsular polysaccharides are formed from oligosaccharide polymers surrounding bacteria. Capsules confer to bacteria resistance against host defenses by counteracting lytic effects of complement and phagocytosis.\textsuperscript{41} Hemolysins are cytotoxic, pore-forming proteins that permeate the cell membrane. Hemolysin production was first observed in the 1940s in \textit{E. coli} causing acute pyelonephritis.

Besides mechanisms of virulence, uropathogenic bacteria may also compete with host cells for nutrients, such as iron. All uropathogenic strains express some molecules responsible for iron uptake. For example, enterobactin is expressed by nearly all \textit{E. coli} strains, but most \textit{E. coli} strains
causing acute pyelonephritis produce aerobactin, which is a high-affinity iron-binding protein, as well as other iron-sequestering proteins, including yersiniabactin, ChuA, and Iro.42-44

The role of host immune response

Host resistance to UTI depends for the most part on innate immune defenses, mainly during the acute phase of the disease. The response to uropathogenic E. coli is activated by P-fimbriae mediated adhesion to glycolipid receptors, leading to activation of TLRs, of which TLR4 has been considered the most important.45,46 Activation of TLR4 signaling results in the release of transcription factors such as IRF3, which trigger neutrophil recruitment and cytokine production in order to kill bacteria. These mechanisms determine the symptoms and signs of UTI. Urothelial cells produce interleukin-8 (IL-8), which attracts neutrophils to urinary tract leading to pyuria.24,25,47 Infection itself enhances the expression of IL-8 receptors, further stimulating neutrophil attraction and activation. Interleukin-6 (IL-6) is also secreted by urothelial cells. IL-6 activates C-reactive protein (CRP) production and stimulates the production of mucosal IgA.25

Another source of innate immune defense are the antimicrobial peptides (AMPs), which are natural antibiotics produced by nearly all organisms.48,49 AMPs are small cationic proteins expressed by phagocytic and epithelial cells, either constitutively or through induction by invading agents.48

Further supporting the role of innate immunity in UTI is the fact that genetic variation affecting innate immunity influences host susceptibility. For example, mutations in the TLR4 gene promoter lead to low expression of TLR4, which was detected in children with asymptomatic bacteriuria when compared to age-matched controls or children with acute pyelonephritis. In addition, single nucleotide polymorphisms (SNPs) in the gene promoter of the transcription factor IRF3 have been identified in about 80% of patients with recurrent episodes of acute pyelonephritis. Reduced expression of CXCR1, the IL-8 receptor, due to SNPs in the CXCR1 gene, was also found in children with frequent episodes of acute pyelonephritis.50-52 Individuals of blood group P lack functional receptors for P-fimbriae, while children with blood group P1 have an increased risk of acute pyelonephritis. Very few AMPs have been described in the human kidney and urinary tract, which include defensins, cathelicidin, hepcidin, and ribonuclease 7. Other proteins with antimicrobial activity present in the kidney and urinary tract are Tamm–Horsfall protein, lactoferrin, lipocalin, and secretory leukocyte proteinase inhibitor.48,49,53

It should also be mentioned that a specific immune response develops after three to seven days in patients with acute pyelonephritis. As an attempt to stimulate specific immune mechanisms, experimental vaccines against antigens of uropathogenic E. coli have been tested.64 Besides vaccines, other alternative methods and therapeutic strategies to prevent and/or control UTIs include receptor analogues, pilicides and curlicides, bacterial interference, or phagotherapy.64

The role of host urinary tract malformations

UTIs may be the sentinel event for underlying congenital anomalies of the kidney and urinary tract (CAKUT), although normal anatomy is more common.6 In 30% of children with CAKUT, UTI can be the first sign.7 If pediatricians fail to detect patients at risk of CAKUT, the upper urinary tract may be damaged.

Hypothetically, anatomical or functional alterations of normal urinary flux may certainly predispose to episodes of UTI, and these episodes probably occur in neonates or young infants. In this regard, the VUR has been associated with approximately 20% of neonatal cases of UTI, although the incidence of VUR is not significantly different between genders, birth weight, gestational age, or mode of delivery.55 In a study with infants less than 2 months of age from a neonatal intensive care unit, a rate of anatomic abnormalities in patients with UTI of less than 5% was detected. However, VUR was associated with a younger age at UTI presentation.56 In another study including 45 male infants with first UTI episode occurring early in life, renal ultrasound scan (RUS) and voiding cystourethrogram (VCUG) found CAKUT in half of the cases.57 The most common anomalies were VUR, duplicated collecting system, posterior urethral valves, ureteropelvic junction obstruction, and renal hypoplasia.57 The DMSA scan revealed renal scars in those with VUR grade 3 or higher.58 Similarly, renal anomalies were found in 47% of febrile infants less than 30 days of age at the first UTI episode.59 However, even in the absence of any abnormalities detected on the RUS or VCUG, infants with UTI can have an abnormal DMSA scan, indicating renal cortical damage. The question is if the renal cortical damage would be an effect rather than a cause of a UTI.6

Clinical findings

Early and prompt diagnosis of UTIs is paramount to initiating therapy and thereby limiting morbidity and renal damage. In children, however, the diagnosis is rarely straightforward. The clinical presentation of UTIs in children is highly heterogeneous, sometimes misleading, with symptoms that can be quite obscure. As a consequence, unfortunately many UTIs are likely either not diagnosed or diagnosed late.60 Therefore, it is important that the pediatrician or the primary care providers have a high index of suspicion for UTIs in children. The evaluation must include a thorough history and the importance of the physical exam in pediatric patients cannot be overstated.

The clinical manifestations of the UTIs are clearly related with the age of the children and the site of the infection. Smellie et al.,61 in a classic study of 200 children (3 days to 12 years of age) with UTI, demonstrated that the most common symptoms in the first 2 years of life were failure to thrive, feeding problems, vomiting, and fever. In the 2- to 5-year-old child, fever and abdominal pain were the most common symptoms, and after 5 years of age, the classic symptoms and signs of UTI (fever, dysuria, urgency, and costovertebral angle tenderness) predominated. While the history and physical examination represent the cornerstone of an accurate diagnosis, in nonverbal children the clinical
suspicion of UTI can be troublesome due to the nonspecific nature of symptoms. The classic presentations of dysuria, frequency, and flank pain in adults are unreliable when applied to pediatric UTI, particularly in infants. The presenting complaints in children tend to be vague, including fever, irritability, lethargy, poor feeding, failure to thrive, and gastrointestinal complaints. In addition, evidence of infection outside the urinary tract does not exclude the possibility of UTI. To make matters more complicated, symptoms and signs of respiratory or gastrointestinal infections are often present in febrile infants and in children with documented UTI.

Newborns and infants younger than 3 months may have, at onset, vague and nonspecific symptoms of illness that are difficult to interpret, including failure to thrive, diarrhea, irritability, lethargy, malodorous urine, hypothermia, fever, asymptomatic jaundice, and oliguria or polyuria. As with most bacterial infections in this age group, there is an elevated probability of bacteremia, sepsis, and high rate of mortality (around 10%) due to the spread of infection to other sites. In this age group, UTI may also present with less acute, insidious symptoms, such as food refusal, occasional vomiting, pallor, and jaundice. In fact, it has been recommended that testing for UTI be part of the evaluation of asymptomatic jaundice in infants younger than 8 weeks, especially those with elevated conjugated bilirubin levels. The American Academy of Pediatrics (AAP) recommends that infants with elevated direct bilirubin levels be screened for UTIs. However, those with elevated unconjugated bilirubin levels should not be excluded, especially if other concerning clinical features are present.

In infants between 3 months and 2 years old, fever is the main symptom, and often the only sign of infection. High temperatures associated with nonspecific manifestations like appetite loss, vomiting, abdominal pain, dehydration, and poor weight gain are commonly found in this age group. It can be rarely associated with specific signs or symptoms related to the urinary tract, such as urinary dysuria and foul-smelling urine. The pediatrician should consider investigating UTI in infants with unexplained fever. The prevalence of UTI in infants and young children with fever that is not localizable by history or physical examination is high. In a meta-analysis, Shaikh et al. have shown a pooled prevalence of 7.0% (CI: 5.5–8.4) of UTI among the 14 studies of febrile infants less than 24 months of age. Among males, prevalence rates were highest during the first 3 months of life and declined thereafter. In females, prevalence rates were highest during the first 12 months. According to the AAP guideline, the presence of UTI should be considered in neonates and children between 2 months to 2 years of age with unexplained fever (with strong evidence). In the same guideline, the experts pointed out that the two sexes are not affected equally. The prevalence of UTI in febrile girls with 2 months to 2 years is more than twice that in boys (relative risk, 2.27). The prevalence of UTI in girls younger than 1 year of age is 6.5%, while, in boys, it is 3.3%. The prevalence of UTI in girls between 1 and 2 years of age is 8.1%, whereas, in boys, it is 1.9%. In an updated guideline, the AAP recommended that if a clinician decides that a febrile infant with no apparent cause for the fever requires antimicrobial therapy to be administered because of ill appearance or another pressing reason, the clinician should ensure the collection of urine specimens for both culture and urinalysis before antibiotic administration. If a clinician assesses a febrile infant with no apparent source for the fever as not being so ill as to require immediate antimicrobial therapy, then the clinician should assess the likelihood of UTI.

In this regard, Shaikh et al. recently developed and validated a calculator for estimating the probability of UTI in young febrile children. Electronic medical records of febrile children aged 2–23 months who were brought to the emergency department of Children’s Hospital of Pittsburgh were reviewed. The authors created an independent training database with 1686 patients and a validation database with 384 patients. They tested five multivariable logistic regression models for predicting risk of UTI; one clinical model had only clinical variables and the remaining had laboratory results. The clinical model had lower accuracy than the laboratory models, indicating the nonspecific signs and symptoms of UTI in young children (clinical model area under the curve [AUC] of 0.80 [95% CI 0.77–0.82] vs. 0.97 [95% CI, 0.96–0.98] to 0.98 [95% CI, 0.98–0.99] for the laboratory models). Models including a Gram-stained smear had better accuracy than those that did not use this exam. The authors concluded that accurate diagnosis of UTI may decrease the delay in starting the treatment and may avoid unnecessary use of antibiotics.

Older children are better able to verbalize symptoms and, for this reason, specific symptoms of UTI are more commonly identified. Abdominal pain and fever are the most common presenting symptoms in children between 2 and 5 years of age. After 5 years, the classic urinary tract symptoms, including dysuria, frequency, suprapubic or flank discomfort, incontinence, and costovertebral angle tenderness are usually present. Less common signs such as secondary enuresis in a previously toilet-trained child or frank hematuria can also occur. The pediatrician must be aware, however, that sometimes even older children may be less able to describe localized symptoms.

Adolescent girls are more likely to present with typical cystitis symptoms including frequency, urgency, dysuria, cloudy urine, hematuria, and lower abdominal discomfort. The prevalence of UTI among adolescent boys is very low. Adolescents are better able to provide history and participate in physical exams. Sexual activity is a special issue for this population that requires additional attention. Sexually transmitted infections (STI) are an important differential diagnosis in adolescents with urinary symptoms. Adolescent girls with vaginitis or a sexually transmitted infection (STI) may present with symptoms similar to UTI. In addition, adolescent girls who are diagnosed with cystitis may have a concurrent vaginitis or STI.

Concerning the differentiation between pyelonephritis and cystitis, host and bacterial biomarkers were recently investigated in blood and urine samples from 61 children with febrile UTI. To detect children with pyelonephritis, a DMSA scan was performed within two weeks of UTI diagnosis. Inflammatory proteins were measured in blood and urine samples, and in children with UTI caused by E. coli, polymerase chain reaction for four previously identified virulence genes was also performed. The best urinary markers that differentiated pyelonephritis from cystitis were the chemokines CXCL1, CXCL9, and CXCL12, C-C motif chemokine ligand 2, INF-γ, and IL-15. The best blood marker
Box 1  Conditions to consider for the investigation of urinary tract infection in children.

- Any child and adolescent with urinary symptoms
- Any child, especially infants, with an unexplained fever
- Any newborn with signs or symptoms of bacteremia
- Any newborn with elevated conjugated serum bilirubin
- Any infant with unexplained failure to thrive

Box 2  Risk factors for urinary tract infection in children.

- Structural urinary tract anomalies
- Antenatal hydronephrosis
- Bowel and bladder dysfunction
- Neurogenic bladder
- Immunocompromised states, including newborns
- Gender
- Sexual activity
- Indwelling catheter
- Uncircumcised boys

for pyelonephritis was procalcitonin. However, *E. coli* virulence genes did not associate with pyelonephritis.

The physical examination of children with UTI can be nonspecific. Occasionally, an abdominal mass may be palpated secondary to an enlarged kidney. In older children, palpation of the flank or abdomen may provoke discomfort. Suprapubic palpation can reveal a palpable bladder. Examination of the external genitalia and perineum is important and may reveal the possible origin of symptoms like balanitis, meatal ulcer, or vulvovaginitis. Regardless of age, all children should have their sacral region examined for dimples, pits, or a sacral fat pad, because the presence of these signs is associated with neurogenic bladder. The magnitude of the temperature elevation might assist with the clinical evaluation. Low-grade fevers are more likely to implicate a lower tract infection, whereas temperatures greater than 39.0 °C are indicative of upper tract infection. Hypertension can be transitory in acute pyelonephritis. However, if elevated blood pressure persists, the suspicion of CAKUT or renal parenchyma lesions should be considered.

In summary, diagnosis of UTI in children, especially in younger infants, can be difficult and requires a high index of suspicion. Some key points must be considered by the pediatrician to prevent missing the diagnosis of UTI in children. Many guidelines and opinions of experts have recommended an investigation for possible UTI in some selected conditions (Box 1). In addition, it is a pivotal issue that the pediatrician recognizes children at risk for UTI (Box 2). For instance, possible urinary tract anomaly, voiding dysfunction, and constipation must be investigated during the history taking. In this regard, the pediatrician should inquire about fetal ultrasonography, as the majority of urinary tract anomalies, a well-known strong risk factor for UTI, are currently suspected in utero. Therefore, the pediatrician with a high index of suspicion, a thorough and accurate history, and a complete physical exam might be able to prevent the delay of UTI diagnosis and thus avoid kidney damage.

Diagnosis and imaging investigation

Urine collection

The main methods of urine collection include clean-catch, plastic bag, bladder catheterization, and suprapubic aspiration (SPA). These four methods have variable contamination rates and invasiveness.

Most commonly urine is obtained from clean-catch urine samples, especially for toilet-trained children. In newborns, and when voided specimens cannot be obtained, the best way to collect urine is still controversial. Clean-catch urine is also possible to obtain in non-toilet trained children. In these cases, the patient is placed in the lap of a parent or nurse holding a sterile foil bowl underneath the genitalia. A systematic review of five studies comparing clean voided urine specimens with bladder tap specimens reported a wide variation among studies, with sensitivity ranging between 75% and 100% and specificity varying between 57% and 100%. Conversely, Ramage et al. previously detected a good correlation between results of urine culture obtained by this method and by SPA. In regard to urine contamination, a study with 120 infants and children showed a 25% contamination rate with samples from clear-voided urine when compared to samples from SPA.

Collection in a sterile plastic bag attached to the cleaned genitalia is a technique often used in several centers. Although a culture-negative urine bag sample is reliable, this technique has a high rate of false-positive cultures because of contamination by periurethral flora. In a cross-sectional study of 303 non toilet-trained children under age 3 years at risk for UTI, sensitivity and specificity of the urinalysis collected by clean-voided bag were compared with catheter urine specimens using the catheter culture as the gold standard. The bag dipstick was more sensitive than the catheter dipstick for the entire study sample: 0.85 vs. 0.71, respectively. However, specificity was consistently lower for the bag specimens than for the catheter specimens: 0.62 vs. 0.97, respectively.

International guidelines generally recommend that urine should be collected by bladder catheterization or SPA under ultrasound control. SPA is the most sensitive method for obtaining an uncontaminated urine sample. When urine is collected by SPA, any colony count is considered to represent significant bacteriuria. All other methods of urine collection (clean catch, bladder catheterization, and plastic bag collections) require passage of urine through the urethra. SPA has been considered the standard method for obtaining urine that is uncontaminated by perineal flora. Variable success rates for obtaining urine have been reported, ranging from 23% to 90%. When RUS guidance is used, success rates improve. Despite invasiveness, the technique has limited risks when employed by expert physicians. SPA is recommended for boys with severe phimosis, girls with tight labial adhesions, and in case of external genital infection or presence of complex genital abnormalities. Bladder catheterization is considered an alternative to SPA, although with higher rates of contamination. Urine obtained through catheterization for culture has a sensitivity of 95% and a specificity of 99% in comparison to that obtained via SPA.
In conclusion, methods of urine collection in individual centers should be determined based on the accuracy of voided specimens.

**Urine culture**

Urine culture is still the gold standard for diagnosing UTI. In freshly voided urine, a growth of more than 10^8 colony-forming units (CFU) per liter (10^8 per mL) of a unique bacterium is regarded most frequently as the cutoff between contamination and UTI. However, what is not broadly understood is that CFU quantification is a semiquantitative test. The method is based on the microbiology technician unfailingly differentiating 10 and 100 separate CFUs on an agar plate, which has been streak plated with 1 mL of urine. Considering the intrinsic imprecision of the method, physicians should take into account signs and symptoms of UTI to treat a child. It must be mentioned that some children with UTI do not reach the traditional diagnostic threshold of 10^9 CFU/L.

As an example, Upadhyay et al., found that 20% of children with UTI based on SPA had CFU below 10^8/L (10^7/mL) on voided specimens. The CFU of these children with UTI was 10^9/L (10^8/mL) to 10^7/L (10^6/mL). Indeed, different cutoffs for significant bacteriuria are adopted for SPA or catheter specimens based on the lower risk of contamination in such specimens. The cutoff used for bladder catheter specimens is > 10^2 CFU/mL (> 10^3 CFU/mL), while any bacterial growth in urine obtained by SPA indicates a UTI.^

**Imaging studies for urinary tract infections**

Most children undergo one or more imaging studies following their first UTI aiming to identify abnormalities, which increases the risk of recurrent UTI or kidney damage. In the last decade, however, newer guidelines assume that imaging is only of value if subsequent management reduces the risk of UTI, kidney damage, and its long-term sequelae. These guidelines were released by the National Institute of Health and Care Excellence (NICE) in the UK, from the AAP, and from the Italian Society of Pediatric Nephrology (ISPN). All of them suggest limited investigations for children with UTI.

The NICE guidelines provide imaging recommendations for children of all ages, whereas the AAP guidelines apply to children aged 2–24 months, and the ISPN guidelines refer to children aged 2–36 months. The NICE guidelines recommend that children aged over 6 months with their first uncomplicated UTI require no investigations following the episode and that children under 6 months should have only RUS. The AAP and ISPN guidelines recommend that all infants aged 2–24 months with febrile UTIs should undergo RUS, although they recognize that prenatal US is likely to identify most serious urinary tract abnormalities. None of these recent guidelines recommend routine VCUG or DMSA scans, but they recommend further evaluation if the ultrasound is abnormal, if the child is critically ill and fails to respond promptly to antibiotics, and in case of recurrent infections. Some studies have evaluated the impact of fewer investigations and concluded that the recent guidelines are safe to follow. On the other hand, other authors consider that potentially important abnormalities will be missed if the newer guidelines are followed. It should also be mentioned that these guidelines assume that most serious urinary tract abnormalities would be identified in antenatal US and that high-quality RUSs interpreted by expert pediatric radiologists are always available. However, in many situations, those assumptions are not true.

In a spite of the imaging protocol adopted, RUS is generally considered the first-line investigation for urinary tract malformations, since the method is noninvasive and can identify structural anomalies including obstructive uropathies, kidney hypoplasia, and urinary tract dilatations. However, the main limitations of RUS are the dependence on the equipment and the operator, and the impossibility to obtain data on renal function. Compared with VCUG and DMSA scans, RUSs are poor predictors of the presence of VUR or of kidney damage, respectively.

The termed ‘bottom-up’ approach is traditionally adopted for imaging evaluation of UTI. RUS and VCUG are recommended after the first episode of UTI in all pediatric patients despite sex and age group. VCUG is regarded as the reference standard for identifying VUR and for providing information on the bladder and urethra. More recently, it has been debated whether the presence and severity of VUR on VCUG might influence decisions on the management of VUR. In this regard, subgroup analyses in randomized controlled trials have found no difference in the efficacy of antibiotics in preventing UTI between children with and without VUR or between mild (grades I or II) and severe (grades III or IV) VUR. Some authors consider that since the presence or severity of VUR does not influence the efficacy of treatment, routine VCUG following the first UTI is no longer justified. The remaining clear indication for a VCUG is to evaluate the bladder and urethra in children suspected of having obstructive uropathy, such as posterior urethral valves.

To avoid all children from having unnecessary VCUGs, some authors have suggested a ‘top-down’ approach to imaging studies, with RUSs and DMSA scans performed first and VCUG only performed if the DMSA scan shows acute kidney parenchymal injury. The DMSA scan is a sensitive test for detecting acute parenchymal injury following UTI, with a sensitivity of 86% and specificity of 91%. However, DMSA scans cannot differentiate between damage due to UTI and congenital kidney damage. In addition, most acute changes resolve over time regardless of whether antibiotic prophylaxis is used. The present authors conducted a retrospective cohort study with the aim to evaluate the diagnostic accuracy of DMSA scan and RUS in identifying high-grade VUR in 533 children after a first episode of UTI. The findings showed that if a negative diagnosis was established only when both test results were normal, sensitivity was 97% and the diagnostic odds ratio was 25. Only nine children (6.3%) with severe reflux would be missed by an absence of alterations in both tests. Nevertheless, a systematic review of 13 studies evaluating DMSA scans for the identification of dilating VUR (grades III–V) concluded that DMSA scans are poor predictors of dilating VUR, with considerable heterogeneity between studies. It should be also taken into account that, in many countries, there is limited availability of DMSA scans and that these are expensive for families.

Both VCUG and DMSA scans are associated with significant radiation (both equivalent to 40–50 chest X-rays or four
months of natural background radiation), and are unpleasant and time-consuming tests for the child and their families. In addition, a nationwide population-based retrospective cohort study in Taiwan found that the overall risk of cancer was 1.92-fold greater in children who had undergone VCUG compared with matched controls, with highest risk for genital and urinary system cancers.99

A recent line of investigation on the ideal imaging protocol is the use of machine learning algorithms to develop predictive models for the probability of recurrent UTI associated with VUR in children after first infection. In this regard, the Advanced Analytics Group of Pediatric Urology and the ORC Personalized Medicine Group evaluated 500 subjects, including 305 from the Randomized Intervention for Children with Vesico-Ureteral Reflux (RIVUR) and 195 from the Careful Urinary Tract Infection Evaluation (CUTIE) trials. Most subjects were females (90%) and they had a mean age of 21 ± 19 months. Recurrence of UTI occurred in 72 subjects, of whom 53 also exhibited VUR. The final predictive model included age, sex, race, weight, systolic blood pressure percentile, dysuria, urine albumin-to-creatinine ratio, prior antibiotic exposure, and current medication. The model predicted recurrent UTI related to VUR, with an AUC of 0.761 (95% CI: 0.714–0.808).100

The debate on the ideal imaging protocol is still ongoing, but experts do agree that longitudinal prospective studies are still needed to establish tailored imaging protocols for the approach of UTI in childhood. An interesting approach was proposed by Marks et al.10 They suggested that targeting investigations for a selected group of children (as opposed to protocol-based investigations of all children with UTI) would be clinically safe and effective, and would avoid the distress and cost of unnecessary invasive investigations. Certain clinical risk factors have been defined in the literature and can help identify which infants and children with febrile UTI have a high risk of having an abnormal urinary tract, and consequently warrant investigation (Box 3).

**Box 3** Features of high-risk children that warrant investigation for an abnormal urinary tract.

- Recurrent infections
- Clinical signs, including poor urinary stream or palpable kidneys
- Unusual organisms (those which are not Escherichia coli)
- Urinary tract infection associated with bacteremia or septicemia
- Prolonged clinical course, with failure to respond fully to antibiotic treatment within 48–72 h
- Unusual clinical presentation, such as in an older boy
- Known dilatation or abnormality on antenatal ultrasound screening of the urinary tract

Treatment and chemophrophylaxis

Therapy of UTI eradication

The aims of the management of children with UTI are (1) resolution of the acute symptoms of the infection; (2) prompt recognition of concomitant bacteremia, particularly in infants less than 2 months of age and (3) prevention of renal damage by eradication of the bacterial pathogen, identification of abnormalities of the urinary tract, and avoidance of recurrent infections.101 Prompt treatment of UTI in preschool children might prevent renal scarring. For instance, Coulthard et al.102 reported that treating children’s UTI in less than three days after the onset of the symptoms more than halves the risk of them acquiring kidney scars. Accordingly, Shaikh et al.103 showed that there was a significant association between delay in treatment of febrile UTI and permanent renal scarring. The authors analyzed data of 482 children, 90% females and 78% with VUR, and found, after adjusting other covariates, that a delay of 48 h or more would increase the odds of new renal scarring by about 47%.

The clinical management of UTIs in children should be tailored according to the age of the patient, severity of presentation, and infection location (cystitis vs. pyelonephritis). Antibiotic treatment is the cornerstone of treatment for acute UTI. The decision to initiate empiric treatment should be based on clinical suspicion of UTI that includes careful history and physical exam, and positive urine analysis in an appropriately collected urine specimen.10 The clinician should base the choice of agent on local antimicrobial sensitivity patterns (if available) and should adjust the choice according to sensitivity testing of the isolated uropathogen.69 Most patients can be treated in an outpatient basis with oral therapy, if the child has a nontoxic appearance, can tolerate oral medications, and the family complies with recommendations.72 On the other hand, inpatient parenteral therapy should be considered for acutely ill children, children who cannot tolerate oral therapy, or when adherence with the prescribed regimen is in question.6 The AAP currently recommends that parenteral antibiotic therapy and hospitalization be considered for children who appear to be severely ill or dehydrated and those who are unable to retain oral intake.69 Children with a renal or perinephric abscess should also be treated initially with parenteral therapy and surgical drainage should be considered. Parenteral therapy should also be considered in children who are immunocompromised and in those with indwelling devices.6

Infants aged 3 months or less with UTI should be treated initially with intravenous antibiotics due to the risk of urosepsis and the higher possibility of structural urinary tract anomaly, although data on prevalence of uropathies are imprecise.104–106 In addition, infants younger than 60–90 days are more likely to have their course of disease change abruptly because of their incompletely developed immune system.107,108 Broad coverage for group B streptococci and Enterobacteriaceae using the intravenous route is required during the first 12 weeks of life, pending results of blood and cerebrospinal fluid cultures. Once blood and cerebrospinal fluid cultures are confirmed as negative, the systemic signs have resolved, and the infant is afebrile, antimicrobial therapy may be completed using the oral route for a total duration of therapy of seven to 14 days.108 Antibiotic parenteral treatment regimens are detailed in Table 1.

Current treatment recommendations for children over the age of 3 months with clinical suspected pyelonephri-
This is described in five urinary tract infections. For instance, Hoberman et al. compared three days of intravenous cefotaxime followed by 11 days of oral cefixime vs. 14 days of oral amoxicillin alone. In a Cochrane review, this review, based on three randomized trials (960 children), provides good evidence that oral antibiotics are an effective treatment for acute febrile pyelonephritis. For instance, Hoberman et al. compared three days of intravenous cefotaxime followed by 11 days of oral cefixime vs. 14 days of oral cefixime alone in 306 children 1–24 months of age; there was no difference in outcome. A more recent study involving 502 children (>1 month and <7 years of age) had similar results. Five trials (including 534 children) using intravenous antibiotics for 48–72 h followed by oral antibiotics demonstrated no difference in DMSA abnormality or resolution of symptoms compared with seven to 14 days of intravenous antibiotics. Thus, it appears that oral antibiotics may be appropriate for the first febrile UTI in children older than 3 months of age. This review corroborates recent guidelines, which recommend that oral antibiotics should be given for seven to ten days unless the child is seriously unwell or unable to take oral antibiotics, in which case intravenous antibiotics are indicated.

Empiric oral antibiotics usually recommended are those that have proven to be effective against the usual uropathogens. The initial choice of antibacterial therapy is preferably based on the knowledge of the predominant uropathogens in the patient’s age group, antibacterial sensitivity patterns in the practice area, the clinical status of the patient, and the opportunity for close follow-up.

A common choice for treatment of UTI orally in the well-appearing child includes a sulfonamide-containing antimicrobial (trimethoprim-sulfamethoxazole [TMP-SMX]) or a cephalosporin. Antibiotics that are excreted in the urine, but fail to achieve therapeutically concentra tions in the bloodstream like nitrofurantoin, are not recommended for treatment of febrile infants or children in whom renal involvement is suspected. Of particular interest, Edlin et al. described resistance patterns in pediatric urinary isolates from 192 hospitals throughout the United States and found that up to 24% of E. coli cultured were resistant to TMP-SMX and 45% were resistant to amoxicillin. On the other hand, resistance was found in less than 10% of E. coli for cephalosporins, amoxicillin-clavulanate, ciprofloxacin, and nitrofurantoin. Likewise, a surveillance study of E. coli isolates from 967 children with UTI in a tertiary hospital from 1992 to 1994 revealed that 30% of 1636 isolates were resistant to TMP-SMX. Risk factors associated with TMP-SMX resistance were young age, multiple inpatient hospital admissions, and previous antimicrobial therapy for greater than four weeks in the past six months. In Brazil, Reis et al. recently described a similar pattern. The authors conducted a retrospective study in 1641 patients with community-acquired UTI for five years (2010–2014). Resistance to amoxicillin was observed in 55.9% of the isolated species, TMP-SMX showed 33.6% bacterial resistance, ciprofloxacin 18.4%, levofloxacin 18.0%, gentamicin 6.3%, cefepime 3.7%, and amikacin showed the lowest frequency, at 1.3%. Nevertheless, an important recent issue has emerged regarding the use of fluoroquinolones for the treatment of uncomplicated UTI in all age groups. Pharmacovigilance risk assessment committees from two leading international agencies, the United States Food and Drug Administration (FDA) and the European Medicines Agency (EMA), have released warnings that fluoroquinolones should not be prescribed for patients who have other treatment options for infectious diseases, including uncomplicated UTIs, because the risks outweigh the benefits in these patients and other antibiotics to treat these conditions are available.

There is one systematic review with meta-analysis of six randomized controlled trials (RCTs) including 523 children (aged 2 weeks to 16 years) with microbiologically proven UTI and acute clinical pyelonephritis. These RCTs made comparisons of different classes of antibiotics. Reported outcomes were persistence of bacteriuria at 48–72 h, resolution of clinical symptoms, symptomatic recurrence, and adverse effects. Three RCTs compared third generation cephalosporins with other antibiotics, including amoxicillin-clavulanate and TMP-SMX. There was no difference in the reduction of persistent bacteriuria at 48 h (two RCTs, RR 5.5, 95% CI, 0.30–1.28), recurrent or persistent UTI five to ten days after the end of therapy (three RCTs, RR 0.42, 95% CI, 0.03–6.23), or the incidence of gastrointestinal adverse effects (three RCTs, n = 108, RR 0.55, 95% CI, 0.10–3.16).

Alternative options for outpatient management include outpatient parenteral therapy for patients with acute pyelonephritis. Several studies have reported that once-daily parenteral administration of gentamicin or ceftriaxone in a day treatment center is safe, functional, and cost effective in children with UTI. Alternative options for afebrile children and adolescents with cystitis symptoms are based on many well-conducted trials and systematic reviews. These studies have shown that short duration therapy (three to four days) is as effective as standard therapy (seven to 14 days) in eradicating urinary bacteria. The NICE guidelines recommend three days of treatment, with the choice of antibiotic directed by local guidelines.

### Table 1 Parenteral antibiotics for treating acute pyelonephritis.

| Antibiotic       | Dose: mg/kg/day | Interval |
|------------------|-----------------|----------|
| Ampicillin       | 100 mg          | Every 6 h|
| Gentamicin       | 7.5 mg          | Every 8 h|
| Ceftriaxone      | 50–100 mg       | Every 12 h|
| Cefotaxime       | 100–200 mg      | Every 8 h|
| Ceftepine        | 100 mg          | Every 12 h|

### Table 2 Orally administered antibiotics for acute urinary tract infection.

| Antibiotic              | Dose: mg/kg/day | Interval |
|-------------------------|-----------------|----------|
| Trimethoprim-sulfamethoxazole | 40 mg (SMT)     | Every 12 h|
| Cefadroxil              | 30–50 mg        | Every 12 h|
| Cephalaxin              | 50–100 mg       | Every 6 h|
| Amoxicillin-clavulanate | 40 mg (amoxicillin) | Every 12 h|

 utis are based on eight randomized controlled trials and summarized in a Cochrane review. This review, based on three randomized trials (960 children), provides good evidence that oral antibiotics are an effective treatment for acute febrile pyelonephritis.
Prevention of the recurrent UTI and antibiotic prophylaxis

Prevention of recurrent UTI is a debatable issue in the pediatric setting. Patients with significant urinary tract abnormalities or frequent symptomatic UTIs may benefit from prophylactic antibiotics. The basis for this practice was established by Smellie et al., who examined the effect of antibiotic prophylaxis in children with recurrent UTI and structurally normal urinary tracts. In this study, 45 children with radiologically normal urinary tracts were given prophylactic doses of cotrimoxazole or nitrofurantoin, or no prophylaxis, after treatment of a symptomatic UTI. During the initial 10 months of the study, the 25 children on prophylaxis had significantly fewer episodes of UTI. Further studies have confirmed that nitrofurantoin, sulfonamides, and cotrimoxazole are effective in reducing the recurrence rate of infection in patients with normal urinary tracts as long as the drug is given.

More recently, in a multicenter clinical trial, Craig et al. demonstrated the efficacy of prophylaxis in predisposed children. The authors randomized 576 children to receive either daily TMP-SMX or placebo for 12 months. During the study, UTI developed in 36 of 288 patients (13%) in the group receiving TMP-SMX (antibiotic group) and in 55 of 288 patients (19%) in the placebo group (hazard ratio in the antibiotic group, 0.61; 95% CI, 0.40–0.93; p = 0.02).

Nevertheless, the use of long-term antimicrobial prophylaxis to prevent UTI in children with and without VUR remains controversial. One meta-analysis investigated antibiotic prophylaxis in children and included six RCTs with a total of 388 participants, predominantly girls younger than 14 years of age, who were identified as being at risk of recurrent UTI, but without any predisposing anatomic or neurologic abnormalities. Compared with placebo, four studies reported that the incidence of recurrence was reduced in the antibiotic-treated group, although with a wide range (21%–69%) in the recurrence of repeat positive cultures (RR 0.44, 95% CI, 0.19–1.00). However, when analysis was limited to high-quality studies, the results were not statistically significant. The authors concluded that more evidence in the form of properly randomized double-blinded trials is needed to support the routine use of antibiotic prophylaxis in preventing recurrent UTI in children. Another issue is the effect of prophylaxis on developing a multigrid-resistant recurrent UTI. Selekman et al. recently demonstrated in a meta-analysis that children with VUR treated with prophylaxis were more likely to have a multigrid-resistant infection (33% vs. 6%, p < .001) and were more likely to receive broad-spectrum antibiotics (68% vs. 49%, p = .004). Those receiving prophylaxis had 6.4 times the odds (95% confidence interval: 2.7–15.6) of developing a multigrid-resistant infection. In 2007, NICE published its recommendations that healthcare professionals in the United Kingdom should not use antibiotic prophylaxis routinely in infants and children following first time UTI, and only selectively in recurrent UTI.

Although the effectiveness of antimicrobial prophylaxis for the prevention of UTI has not been fully demonstrated, in the present authors’ experience, some selected patients with recurrent episodes may benefit from this approach by reducing the morbidity and possibly preventing kidney damage. First, following treatment of UTI, the present authors have used prophylactic antibiotic coverage for the young child until urinary tract abnormalities have been excluded by imaging studies. After, the decision regarding continuous antibiotic prophylaxis is based on the results of these imaging studies and/or clinical features, such as age and gender of the children. Concomitantly, it is important to establish fundamental interventions that might reduce recurrent UTI.

In this regard, the prompt identification and adequate treatment of urinary tract abnormalities – such as vesicoureteral reflux, posterior urethral valves, or ureteropelvic obstruction – are relevant. It is beyond the scope of this review to explore the myriad facets of the treatment of specific uropathies. Additional interventions that have been associated with a decrease in symptomatic UTI in children with recurrent UTI include treatment of constipation and management of voiding dysfunction. Obstacles to the effectiveness of antimicrobial prophylaxis are adherence to a daily regimen, side effects associated with the various agents, and the potential for emergence of antimicrobial resistance. According to AAP, to overcome these issues, evidence of effectiveness with a well-tolerated, safe product would be required, and parents would need adequate education to understand the value and importance of adherence.

Agents of choice for prophylaxis of recurrent UTI are nitrofurantoin and cotrimoxazole. In this regard, the earlier mentioned review also identified two RCTs that compared antibiotic classes in prophylaxis of UTI (nitrofurantoin versus trimethoprim, and nitrofurantoin versus cefixime). Nitrofurantoin was found to be superior to trimethoprim, but no different from cefixime in reducing the incidence of recurrent repeat-positive urine cultures. In turn, nitrofurantoin was three times more likely to be discontinued because of the adverse effects such as of nausea, vomiting, or stomachache.

In summary, there has been a tendency of less use of prophylaxis due to dispute about its efficacy, increasing bacterial resistance, and a propensity to low adherence. Alternative measures and management of risk factors for recurrent UTI should be emphasized. However, in selected patients carefully followed, prophylaxis can protect from recurrent UTI and long-term sequelae.

Complications and prognosis

The involvement of renal parenchyma in UTI may lead to an inflammatory reaction with risk of permanent damage. The long-term consequences of such damage include hypertension and impaired renal function, but the frequency of these complications is still poorly known. The main issue is the difficulty in following patients over several decades, which is needed to obtain reliable findings.

Earlier studies from specialized centers reported high rates of hypertension and chronic kidney disease (CKD) in children and young adults with kidney damage following UTI. However, a few years later, population-based studies of individuals with previous UTI did not find similar results. In this regard, Wennstrom et al. evaluated glomerular filtration rate (GFR) and 24 h ambulatory blood pressure
measured 16–26 years after first UTI in 57 (77%) of 74 people with renal parenchyma damage and in a matched group of 51 adults without kidney damage (control group) from the same cohort. The mean GFR of 99 mL/min/1.73 m² in the group with kidney damage did not differ significantly from that in the control group (102 mL/min/1.73 m²). Only eight patients (six in the kidney damage group and two in the matched group) exhibited GFR below 80 mL/min/1.73 m². Similarly, ambulatory blood pressure measurements did not differ between individuals with and without renal parenchyma damage. Blood pressure exceeded the reference values in 9% of those with kidney damage vs. 6% in those without.

However, more recently, a systematic review made an estimative for the risk of renal damage after childhood UTI of approximately 15%. Subsequently, 193 young people randomly sampled from a cohort of 1161 children were evaluated following their first childhood UTI and followed up six to 17 years later. Patients with congenital kidney dysplasia or obstruction (24 in total) were excluded. Twenty-two among 150 patients (15%), who were submitted to RUS, presented renal damage and/or decreased kidney growth. Recurrence of UTI and VUR grades III-V were more commonly detected in this subgroup. However, GFR and blood pressure remained within normal range in all participants of the study. In 2015, Gebäck et al. evaluated a population-based cohort of women followed for a median period of 35 years from their first UTI in childhood. GFR was estimated by Cr-educ acid clearance, while renal parenchyma damage was diagnosed by DMSA scan. Eighty-six among 111 women initially recruited completed the investigation; 58 with renal damage and 28 without. Of those with renal damage, one had stage 3 CKD, 14 had stage 2, and 43 had stage 1. Bilateral damage was positively associated with lower GFR. However, most women with UTI-associated renal damage had impressively well-preserved renal function.

Conclusions

Despite its high prevalence, UTI in pediatric patients still has many unsolved issues. First, in neonates and infants, signs and symptoms of UTI are often unspecific, delaying the diagnosis. Should a urine sample be collected in all cases of fever without a known cause? In the present authors’ opinion, this is the one the most important recommendations for a timely diagnosis in this age group. Second, which is the best way to collect urine in non toilet-trained children? On one hand, some guidelines recommend urine catheterization; on the other hand, some authors argue in favor of urine bag collection, despite the prevalence of false positive findings. Third, must the bacteria colony count for infants be equal or higher than \(10^5\) CFU per liter (\(10^6\) per mL)? Fourth, should all children after a single episode of UTI have their urinary tract investigated by imaging? If yes, which is the best protocol? Fifth, in regard to antibiotic prophylaxis, which subgroup of children will benefit from its long-term use? Which criteria should be taken into account to prescribe and to stop antibiotic prophylaxis? Sixth, is there a real risk for CKD and hypertension for children with UTI?

In conclusion, these and other questions are still not solved. However, some general advice must be taken into account by the pediatrician. Early diagnosis of UTI is very important, especially for neonates and infants. Risk factors for recurrence of UTI and for CAKUT should always be considered. The protocol of urine collection must consider the limitations and risks of each method, and also the local practice and feasibility. Imaging evaluation is critical to detect CAKUT, but always depends on the quality of equipment and the experience of the radiologist. In the present authors’ point of the view, a comprehensive RUS by a trained radiologist is advisable for all children who have had a confirmed episode of febrile UTI. Antibiotic prophylaxis is useful to prevent recurrence of UTIs in patients with obstructive uropathies and high-grade reflux. All decisions in regard to a child with UTI must be based on detailed clinical history and physical examination and careful clinical judgment to avoid, on one hand, unnecessary invasive exams and, on the other hand, future adverse outcome for renal function.

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Conflicts of interest

The authors declare no conflicts of interest.

References

1. Hoberman A, Chao HP, Keller DM, Hickey R, Davis HW, Ellis D. Prevalence of urinary tract infection in febrile infants. J Pediatr. 1993;123:17–23.
2. Marild S, Jodal U. Incidence rate of first-time symptomatic urinary tract infection in children under 6 years of age. Acta Paediatr. 1998;87:549–52.
3. Mangiarotti P, Pizzini C, Fanos V. Antibiotic prophylaxis in children with relapsing urinary tract infections: review. J Chemother. 2000;12:115–23.
4. Nuutinen M, Uhari M. Recurrence and follow-up after urinary tract infection under the age of 1 year. Pediatr Nephrol. 2001;16:69–72.
5. Copp HL, Shapiro DJ, Hersh AL. National ambulatory antibiotic prescribing patterns for pediatric urinary tract infection, 1998-2007. Pediatrics. 2011;127:1027–33.
6. Millner R, Becknell B. Urinary tract infections. Pediatr Clin North Am. 2019;66:1–13.
7. Wenerstrom M, Hansson S, Jodal U, Stokland E. Primary and acquired renal scarring in boys and girls with urinary tract infection. J Pediatr. 2000;136:30–4.
8. Arshad M, Seed PC. Urinary tract infections in the infant. Clin Perinatol. 2015;42:17–28, vii.
9. Sastre JB, Aparicio AR, Cotallo GD, Colomer BF, Hernandez MC. Grupo de Hospitales Castrillo. Urinary tract infection in the newborn: clinical and radio imaging studies. Pediatr Nephrol. 2007;22:1735–41.
10. Paintsil E. Update on recent guidelines for the management of urinary tract infections in children: the shifting paradigm. Curr Opin Pediatr. 2013;25:88–94.
11. Tullius K. What do the latest guidelines tell us about UTIs in children under 2 years of age. Pediatr Nephrol. 2012;27:509–11.
12. Coelho GM, Bouzada MC, Pereira AK, Figueiredo BF, Leite MR, Oliveira DS, et al. Outcome of isolated antenatal hydronephrosis: a prospective cohort study. Pediatr Nephrol. 2007;22:1727–34.

13. Montini G, Toffolo A, Zucchetta P, Dal’Amico R, Gobber D, Calderan A, et al. Antibiotic treatment for pyelonephritis in children: multicentre randomised controlled non-inferiority trial. BMJ. 2007;335:386.

14. Montini G, Tullus K, Hewitt I. Febrile urinary tract infections in children. N Engl J Med. 2011;365:239–50.

15. Penido Silva JM, Oliveira EA, Diniz JS, Bouzada MC, Vergara RM, Souza BC. Clinical course of prenatally detected primary vesicoureteral reflux. Pediatr Nephrol. 2006;21:86–91.

16. Marks SD, Gordon I, Tullus K. Imaging in childhood urinary tract infections: time to reduce investigations. Pediatr Nephrol. 2008;23:9–17.

17. Mori R, Lakanpaul M, Verrier-Jones K. Diagnosis and management of urinary tract infection in children: summary of NICE guidance. BMJ. 2007;335:395–7.

18. Williams GJ, Hodson EH, Isaacs D, Craig JC. Diagnosis and management of urinary tract infection in children. J Paediatr Child Health. 2012;48:296–301.

19. Bollgren I, Winberg J. The periurethral aerobic flora in girls highly susceptible to urinary infections. Acta Paediatr Scand. 1976;65:81–7.

20. Hansson S, Jodal U, Noren L, Bjure J. Untreated bacteriuria in asymptomatic girls with renal scarring. Pediatrics. 1989;84:964–8.

21. Okarsa-Napierala M, Wasilewska A, Kuchar E. Urinary tract infection in children: Diagnosis, treatment, imaging - comparison of current guidelines. J Pediatr Urol. 2017;13:567–73.

22. Simoes e Silva AC, Oliveira EA. Update on the approach of urinary tract infection in childhood. J Pediatr (Rio J). 2015;91:S2:10.

23. Nielubowicz GR, Mobley HL. Host-pathogen interactions in urinary tract infection. Nat Rev Urol. 2010;7:430–41.

24. Ragnarsdottir B, Lutay N, Gronberg-Hernandez J, Koves B, Svanborg C. Genetics of innate immunity and UTI susceptibility. Nat Rev Urol. 2011;8:449–68.

25. Ragnarsdottir B, Svanborg C. Susceptibility to acute pyelonephritis or asymptomatic bacteriuria: host-pathogen interaction in urinary tract infections. Pediatr Nephrol. 2012;27:2017–29.

26. Spencer JD, Schwaderer AL, Becknell B, Watson J, Hains DS. The innate immune response during urinary tract infection and pyelonephritis. Pediatr Nephrol. 2014;29:1139–49.

27. Svanborg C. Urinary tract infections in children: microbial virulence versus host susceptibility. Adv Exp Med Biol. 2013;764:205–10.

28. O’Brien K, Stanton N, Edwards A, Hood K, Butler CC. Prevalence of urinary tract infection (UTI) in sequential acutely unwell children presenting in primary care: exploratory study. Scand J Prim Health Care. 2011;29:19–22.

29. Stull TL, LiPuma JJ. Epidemiology and natural history of urinary tract infections in children. Med Clin North Am. 1991;75:287–97.

30. Eden CS, Hanson LA, Jodal U, Lindberg U, Akerlund AS. Variable adherence to normal human urinary-tract epithelial cells of Escherichia coli strains associated with various forms of urinary-tract infection. Lancet. 1976;1:490–2.

31. Oelschlaeger TA, Dobrindt U, Hacker J. Virulence factors of uropathogens. Curr Opin Urol. 2002;12:33–8.

32. Yamamoto S. Molecular epidemiology of uropathogenic Escherichia coli. J Infect Chemother. 2007;13:68–73.

33. Evans DJr, Evans DG. Classification of pathogenic Escherichia coli according to serotype and the production of virulence factors, with special reference to colonization-factor antigens. Rev Infect Dis. 1983;5:5692–701.

34. Orskov I, Orskov F, Jann B, Jann K. Serology, chemistry, and genetics of O and K antigens of Escherichia coli. Bacteriol Rev. 1977;41:667–710.

35. Pak J, Pu Y, Zhang ZT, Hasty DL, Wu XR. Tamm-Horsfall protein binds to type 1 fimbriated Escherichia coli and prevents E. coli from binding to uropakin Ia and Ib receptors. J Biol Chem. 2001;276:9924–30.

36. Wold AE, Mestecky J, Tomana M, Kobata A, Ohbayashi H, Endo T, et al. Secretory immunoglobulin A carries oligosaccharide receptors for Escherichia coli type 1 fimbrial lectin. Infect Immun. 1990;58:3073–7.

37. Xie B, Zhou G, Chan SY, Shapiro E, Kong XP, Wu XR, et al. Distinct glycan structures of uropakin Ia and Ib: structural basis for the selective binding of FimH adhesin to uropakin Ia. J Biol Chem. 2006;281:14644–53.

38. Leffer H, Svanborg C. Chemical identification of a glycosphin-golipid receptor for Escherichia coli attaching to human urinary tract epithelial cells and agglutinating human erythrocytes. FEBS Microbiol Lett. 2006;8:127–34.

39. Palsson-McDermott EM, O’Neill LA. Signal transduction by the lipopolysaccharide receptor, Toll-like receptor-4. Immunology. 2004;113:153–62.

40. Poltorak A, He X, Smirnova I, Liu MY, Van Huffel C, Du X, et al. Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in Tlr4 gene. Science. 1998;282:2085–8.

41. Horwitz MA, Silverstein SC. Influence of the Escherichia coli capsule on complement fixation and on phagocytosis and killing by human phagocytes. J Clin Invest. 1980;65:82–94.

42. Garcia EC, Brumbaugh AR, Mobley HL. Redundancy and speci-ficity of Escherichia coli iron acquisition systems during urinary tract infection. Infect Immun. 2011;79:1225–35.

43. Johnson JR, Stell AL. Extended virulence genotypes of Escherichia coli strains from patients with urosepsis in relation to phylogeny and host compromise. J Infect Dis. 2000;181:261–72.

44. Kanamaru S, Kurazono H, Ishitoya S, Terai A, Habuchi T, Nakano M, et al. Distribution and genetic association of putative uropathogenic virulence factors Iroh, rha, kpsMT, ompT and usp in Escherichia coli isolated from urinary tract infections in Japan. J Urol. 2003;170:2490–3.

45. Ragnarsdottir B, Fischer H, Godaly G, Gronberg-Hernandez J, Gustafsson M, Karpman D, et al. TLR- and CXCRI-dependent innate immunity: insights into the genetics of urinary tract infections. Eur J Clin Invest. 2008;38:12–20.

46. Samuelsson P, Hang L, Wüllt B, Irjala H, Svanborg C. Toll-like receptor 4 expression and cytokine responses in the human urinary tract mucosa. Infect Immun. 2004;72:3179–86.

47. Cheng CH, Lee YS, Tsau YK, Lin TY. Genetic polymorphisms and susceptibility to parenchymal renal infection among pediatric patients. Pediatr Infect Dis J. 2011;30:309–14.

48. Ali AS, Townes CL, Hall J, Pickard RS. Maintaining a sterile urinary tract: the role of antimicrobial peptides. J Urol. 2009;182:21–8.

49. Zasloff M. Antimicrobial peptides, innate immunity, and the normally sterile urinary tract. J Am Soc Nephrol. 2007;18:2810–6.

50. Frendrup B, Godaly G, Hang L, Karpman D, Lundstedt AC, Svanborg C. Interleukin 8 receptor deficiency confers susceptibility to acute experimental pyelonephritis and may have a human counterpart. J Exp Med. 2000;192:881–90.

51. Lundstedt AC, Leijonhufvud I, Ragnarsdottir B, Karpman D, Andersson B, Svanborg C. Inherited susceptibility to acute pyelonephritis: a family study of urinary tract infection. J Infect Dis. 2007;195:1227–34.

52. Lundstedt AC, McCarthy S, Gustafsson MC, Godaly G, Jodal U, Karpman D, et al. A genetic basis of susceptibility to acute pyelonephritis. PLoS One. 2007;2:e825.
53. Ohlsson S, Ljungkvart I, Ohlson K, Segelman M, Wieslander J. Novel distribution of the secretory leucocyte proteasine inhibitor in kidney. Mediators Inflammm. 2001;10:347–50.
54. Zalewska-Platek BM, Platek RJ. Alternative treatment approaches of urinary tract infections caused by uropathogenic Escherichia coli strains. Acta Biochim Pol. 2019;66:129–37.
55. Cleper R, Krause I, Eisenstein B, Davidovits M. Prevalence of vesicoureteral reflux in neonatal urinary tract infection. Clin Pediatr (Phil). 2004;43:619–25.
56. Bonadio W, Maida G. Urinary tract infection in outpatient febrile infants younger than 30 days of age: a 10-year evaluation. Pediatr Infect Dis J. 2014;33:342–4.
57. Goldman M, Laha E, Strauss S, Reisler G, Livne A, Gordin L, et al. Imaging after urinary tract infection in male neonates. Pediatrics. 2000;105:1232–5.
58. Bauer R, Kogan BA. New developments in the diagnosis and management of pediatric UTIs. Urol Clin North Am. 2008;35:47–58, vi.
59. Smellie JM, Hodson CJ, Edwards D, Normand IC. Clinical and radiological features of urinary infection in childhood. Br Med J. 1964;2:1222–6.
60. Bauchner H, Philipp B, Dashefsky B, Klein JO. Prevalence of bacteriuria in febrile children. Pediatr Infect Dis J. 1987;6:239–42.
61. Chang SL, Shortliffe LD. Pediatric urinary tract infections. Pediatr Clin North Am. 2006;53:379–400, vi.
62. Ginsburg CM, McCracken GH Jr. Urinary tract infections in young infants. Pediatrics. 1982;69:409–12.
63. Saadeh SA, Mattek TK. Managing urinary tract infections. Pediatr Nephrol. 2011;26:1967–76.
64. Garcia FJ, Nager AL. Jaundice as an early diagnostic sign of urinary tract infection in infancy. Pediatrics. 2002;109:846–51.
65. Bell LE, Mattek TK. Update on childhood urinary tract infection and vesicoureteral reflux. Semin Nephrol. 2009;29:349–59.
66. Feld LG, Mattek TK. Urinary tract infections and vesicoureteral reflux in infants and children. Pediatr Rev. 2010;31:451–63.
67. Shaikh N, Morone NE, Bost JE, Farrell MH. Prevalence of urinary tract infection in childhood: a meta-analysis. Pediatr Infect Dis J. 2008;27:302–4.
68. Practice parameter: the diagnosis, treatment, and evaluation of the initial urinary tract infection in febrile infants and young children. American Academy of Pediatrics. Committee on Quality Improvement. Subcommittee on Urinary Tract Infection. Pediatrics. 1999;103:843–52.
69. Urinary tract infection: clinical practice guideline for the diagnosis and management of the initial UTI in febrile infants and children 2 to 24 months. Pediatrics. 2011;128:595–610.
70. Shaikh N, Hoberman A, Hum SW, Alberty A, Muniz G, Kurs-Lasky M, et al. Development and validation of a calculator for estimating the probability of urinary tract infection in young febrile children. JAMA Pediatr. 2018;172:550–6.
71. Chon CH, Lai FC, Shortliffe LM. Pediatric urinary tract infections. Pediatr Clin North Am. 2001;48:1441–59.
72. Evans JH. Investigation of urinary tract infection in children. Curr Pediatr. 2006;16:248–53.
73. Schmidt B, Copp HL. Work-up of pediatric urinary tract infection. Urol Clin North Am. 2015;42:519–26.
74. Shaikh N, Martin JM, Hoberman A, Skae M, Milkovich L, Nowalk A, et al. Host and bacterial markers that differ in children with cystitis and pyelonephritis. J Pediatr. 2019;209:146–53, e1.
75. Dias CS, Silva JM, Diniz JS, Lima EM, Marciano RC, Lana LG, et al. Risk factors for recurrent urinary tract infections in a cohort of patients with primary vesicoureteral reflux. Pediatr Infect Dis J. 2010;29:139–44.
76. Quirino IG, Diniz JS, Bouzada MC, Pereira AK, Lopes TJ, Paixão GM, et al. Clinical course of 822 children with prenatally detected nephropathies. Clin J Am Soc Nephrol. 2012;7:444–51.
77. Stein R, Dogan HS, Hoebeca P, Koçvara R, Nijman RJ, Radmay C, et al. Urinary tract infections in children: EAU/ESPU guidelines. Eur Urol. 2015;67:546–58.
78. Ammenti A, Cataldi L, Chimenz R, Fanos V, La Manna A, Marra G, et al. Febrile urinary tract infections in young children: recommendations for the diagnosis, treatment and follow-up. Acta Paediatr. 2012;101:451–7.
79. Whiting P, Westwood M, Bjølke L, Palmer S, Richardson G, Cooper J, et al. Clinical effectiveness and cost-effectiveness of tests for the diagnosis and investigation of urinary tract infection in children: a systematic review and economic model. Health Technol Assess. 2006;10:1–154, iii-iv, xi-xiii.
80. Ramage IJ, Chapman JP, Hollman AS, Elbassass M, McColl JH, Beattie TJ. Accuracy of clean-catch urine collection in infancy. J Pediatr. 1999;135:765–7.
81. Aronson AS, Gustafson B, Svenningsen NW. Combined suprapubic aspiration and clean-voided urine examination in infants and children. Acta Paediatr Scand. 1973;62:396–400.
82. Wald ER. To bag or not to bag. J Pediatr. 2005;147:418–20.
83. McGillivray D, Mok E, Mulrooney E, Kramer MS. A head-to-head comparison: clean-voidbag versus catheter urinalysis in the diagnosis of urinary tract infection in young children. J Pediatr. 2005;147:451–6.
84. Buys H, Pead L, Hallett R, Maskell R. Suprapubic aspiration under ultrasound guidance in children with fever of undiagnosed cause. BMJ. 1994;308:690–2.
85. Kramer MS, Tange SM, Drummond KN, Mills EL. Urine testing in young febrile children: a risk-benefit analysis. J Pediatr. 1994;125:6–13.
86. Upadhyay J, McLorie GA, Baldcù S, Bagli DJ, Khoury AE, Farhat W. Natural history of neonatal reflux associated with prenatal hydrourephrosis: long-term results of a prospective study. J Urol. 2003;169:1837–41.
87. Kocer D, Sarıguzel FM, Karakukcu C. Cutoff values for bacteria and leukocytes for urine sediment analyser FUS200 in culture-positive urinary-tract infections. Scand J Clin Lab Invest. 2014;74:414–7.
88. Berry CS, Vander Brink BA, Koff SA, Alpert SA, Jayanthi VR. Is VCUG still indicated following the first episode of urinary tract infection in boys? Urology. 2012;80:1351–5.
89. Deader R, Tiboni SG, Malone PS, Fairhurst J. Will the implementation of the 2007 National Institute for Health and Clinical Excellence (NICE) guidelines on childhood urinary tract infection (UTI) in the UK miss significant urinary tract pathology? BJU Int. 2012;110:454–8.
90. Pennesi M, L’Erario I, Traván L, Ventura A. Managing children under 36 months of age with febrile urinary tract infection: a new approach. Pediatr Nephrol. 2012;27:611–5.
91. Cowlthard MG, Lambert HJ, Vernon SJ, Hunter EW, Keir MJ. Guidelines to identify abnormalities after childhood urinary tract infections: a prospective audit. Arch Dis Child. 2014;99:448–51.
92. Craig JC, Simpson JM, Williams GJ, Lowe A, Reynolds GJ, McCaggart SJ, et al. Antibiotic prophylaxis and recurrent urinary tract infection in children. N Engl J Med. 2009;361:1748–59.
93. Hoberman A, Greenfield SP, Matteo TK, Keren R, Mathews R, Pohl HG, et al. Antimicrobial prophylaxis for children with vesicoureteral reflux. N Engl J Med. 2014;370:2367–76.
94. Preda I, Jodal U, Sixt R, Stokland E, Hansson S. Normal dimercaptosucin acid scintigraphy makes voiding cystourethrography unnecessary after urinary tract infection. J Pediatr. 2007;151(581-1), 4 e1.
95. Craig JC, Irwig L, Ford M, Willis NS, Howman-Giles RB, Uren RF, et al. Reliability of DMSA for the diagnosis of renal parenchymal abnormality in children. Eur J Nucl Med. 2000;27:1610–6.
96. Montini G, Rigon L, Zucchetta P, Fregonesi F, Toffolo A, Gobber D, et al. Prophylaxis after first febrile urinary tract infection
in children? A multicenter, randomized, controlled, noninferiority trial. Pediatrics. 2008;122:1064–71.

97. Quirino IG, Silva JM, Diniz JS, Lima EM, Rocha AC, Simões e Silva AC, et al. Combined use of late phase dimercaptopro-mercaptopurin acid renal scintigraphy and ultrasound as first line screening after urinary tract infection in children. J Urol. 2011;185:258–63.

98. Mantadakis E, Vouloumanou EK, Georghiati GG, Tsikalidis A, Chatzimichael A, Falagas ME. Acute Tc-99m DMSA scan for identifying dilating vesicoureteral reflux in children: a meta-analysis. Pediatrics. 2011;128:e169–79.

99. Liao YH, Lin CL, Wei CC, Tsai PP, Shen WC, Sung FC, et al. Subsequent cancer risk of children receiving post voiding cystouretrography: a nationwide population-based retrospective cohort study. Pediatr Nephrol. 2014;29:885–91.

100. Advanced Analytics Group of Pediatric Urology and ORC. Personalized Medicine Group. Targeted Workup after initial febrile urinary tract infection: using a novel machine learning model to identify children most likely to benefit from voiding cystourethrogram. J Urol. 2019;202:144–52.

101. Schlager TA. Urinary tract infections in infants and children. Infect Dis Clin North Am. 2003;17:353–65, ix.

102. Coulthard MG, Lambert HJ, Vernon SJ, Hunter EW, Keir MJ, Matthews JN. Does prompt treatment of urinary tract infection in preschool children prevent renal scarring: mixed retrospective and prospective audits. Arch Dis Child. 2014;99:342–7.

103. Shaikh N, Mattoo TK, Keren R, Ivanova A, Cui G, Moxey-Mims M, et al. Early antibiotic treatment for pediatric febrile urinary tract infection and renal scarring. JAMA Pediatr. 2016;170:848–54.

104. Hsiao AL, Chen L, Baker MD. Incidence and predictors of serious bacterial infections among 57- to 180-day-old infants. Pediatrics. 2006;117:1695–701.

105. Navarro M, Espinosa L, de las Heras JA, Garcia Meseguer MC, et al. Symptomatic urinary infection in infants less than 4 months old: outcome in 129 cases. An Esp Pediatr. 1984;21:564–72.

106. Pantell RH, Newman TB, Bernzweig J, Bergman DA, Takayama JI, Segal M, et al. Management and outcomes of care of fever in early infancy. JAMA. 2004;291:1203–12.

107. Hans LA. Esch. coli infections in childhood. Significance of bacterial virulence and immune defence. Arch Dis Child. 1976;51:737–43.

108. Littlewood JM. 66 infants with urinary tract infection in first month of life. Arch Dis Child. 1972;47:218–26.

109. Downs SM. Technical report: urinary tract infections in febrile infants and young children. The Urinary Tract Subcommittee of the American Academy of Pediatrics Committee on Quality Improvement. Pediatrics. 1999;103:354.

110. Hodson EM, Willis NS, Craig JC. Antibiotics for acute pyelonephritis in children. Cochrane Database Syst Rev. 2007;(4):CD003772.

111. Hoiberg A, Wald ER, Hickey RW, Baskin M, Charron M, Majd M, et al. Oral versus initial intravenous therapy for urinary tract infections in young febrile children. Pediatrics. 1999;104:79–86.

112. Edlin RS, Shapiro DJ, Hersh AL, Copp HL. Antibiotic resistance patterns of outpatient pediatric urinary tract infections. J Urol. 2013;190:222–7.

113. Allen LD, MacDonald N, Fuite L, Chan F, Stephens D. Risk factors for resistance to first-line antimicrobials among urinary tract isolates of Escherichia coli in children. CMJ. 1999;160:1436–40.

114. Reis AC, Santos SR, Souza SC, Saldanha MG, Pitanga TH, Oliveira RR. Ciprofloxacin resistance pattern among bacteria isolated from patients with community-acquired urinary tract infection. Rev Inst Med Trop. 2016;58:53.

115. Food and Drug Administration [cited 22 Oct 2019]. Available from: FDA reinforces safety information about serious low blood sugar levels and mental health side effects with fluoroquinolone antibiotics: requires label changes; 2018 https://www.fda.gov/media/114192/download.

116. European Medicines Agency [cited 22 Oct 2019]. Available from: Fluoroquinolone and quinolone antibiotics: PRAC recommends new restrictions on use following review of disabling and potentially long-lasting side effects; 2018 https://www.ema.europa.eu/en/news/fluoroquinolone-quinolone-antibiotics-prac-recommends-new-restrictions-use-following-review.

117. Masson P, Matheson S, Webster AC, Craig JC. Meta-analyses in prevention and treatment of urinary tract infections. Infect Dis Clin North Am. 2009;23:355–85. Table of Contents.

118. Baskin MN, O’Rourke EJ, Fleisher GR. Outpatient treatment of febrile infants 28 to 89 days of age with intramuscular administration of ceftriaxone. J Pediatr. 1992;120:22–7.

119. Lieu TA, Baskin MN, Schwartz JS, Fleisher GR. Clinical and cost-effectiveness of outpatient strategies for management of febrile infants. Pediatrics. 1992;89:1135–44.

120. Keren R, Chan E. A meta-analysis of randomized, controlled trials comparing short- and long-course antibiotic therapy for urinary tract infections in children. Pediatrics. 2002;109:E70–0.

121. Michael A, Hodson EM, Craig JC, Martin S, Moyer VA. Short versus standard duration oral antibiotic therapy for acute urinary tract infection in children. Cochrane Database Syst Rev. 2003;(1):CD003966.

122. Tran D, Muchant DG, Aronoff SC. Short-course versus conventional length antimicrobial therapy for uncomplicated lower urinary tract infections in children: a meta-analysis of 1279 patients. J Pediatr. 2001;139:93–9.

123. Williams GJ, Lee A, Craig JC. Long-term antibiotics for preventing recurrent urinary tract infection in children. Cochrane Database Syst Rev. 2001;(4):CD001534.

124. Lohr JA, Nunley DH, Howards SS, Ford RF. Prevention of recurrent urinary tract infections in girls. Pediatrics. 1979;59:562–5.

125. Smellie JM, Katz G, Gruneberg RN. Controlled trial of prophylactic treatment in childhood urinary-tract infection. Lancet. 1978;2:175–8.

126. Gruneberg RN, Smellie JM, Leakey A, Atkin WS. Long-term low-dose co-trimoxazole in prophylaxis of childhood urinary tract infection: bacteriological aspects. Br Med J. 1976;2:206–8.

127. Smellie JM, Gruneberg RN, Leakey A, Atkin WS. Long-term low-dose co-trimoxazole in the management of urinary tract infection in children. J Antimicrob Chemother. 1976;2:287–91.

128. Williams GJ, Wei L, Lee A, Craig JC. Long-term antibiotics for preventing recurrent urinary tract infection in children. Cochrane Database Syst Rev. 2006;(3):CD001534.

129. Selekmán RE, Shapiro DJ, Boscardin J, Williams G, Craig JC, Brandström P, et al. Urophathogen resistance and antibiotic prophylaxis: a meta-analysis. Pediatrics. 2018:142.

130. Baumer JH, Jones RW. Urinary tract infection in children, National Institute for Health and Clinical Excellence. Arch Dis Child Educ Pract Ed. 2007;92:189–92.

131. Loening-Baucke V. Urinary incontinence and urinary tract infection and their resolution with treatment of chronic constipation in childhood. Pediatrics. 1997;100:1282–8.

132. Brandström P, Hansson S. Long-term, low-dose prophylaxis against urinary tract infections in young children. Pediatr Nephrol. 2015;30:425–32.

133. Gebäck C, Hansson S, Himmelmann A, Sandberg T, Sixt R, Jodal U. Twenty-four-hour ambulatory blood pressure in adult women with urinary tract infection in childhood. J Hypertens. 2014;32:1658–64.
134. Jacobson SH, Eklof O, Eriksson CG, Lins LE, Tidgren B, Winberg J. Development of hypertension and uraemia after pyelonephritis in childhood: 27 year follow up. BMJ. 1989;299:703–6.
135. Silva JM, Diniz JS, Silva AC, Azevedo MV, Pimenta MR, Oliveira EA. Predictive factors of chronic kidney disease in severe vesicoureteral reflux. Pediatr Nephrol. 2006;21:1285–92.
136. Silva JM, Santos Diniz JS, Marino VS, Lima EM, Cardoso LS, Vasconcelos MA, et al. Clinical course of 735 children and adolescents with primary vesicoureteral reflux. Pediatr Nephrol. 2006;21:981–8.
137. Simoes e Silva AC, Silva JM, Diniz JS, Pinheiro SV, Lima EM, Vasconcelos MA, et al. Risk of hypertension in primary vesicoureteral reflux. Pediatr Nephrol. 2007;22:459–62.
138. Smellie JM, Prescod NP, Shaw PJ, Risdon RA, Bryant TN. Childhood reflux and urinary infection: a follow-up of 10–41 years in 226 adults. Pediatr Nephrol. 1998;12:727–36.
139. Martinell J, Claesson I, Lidin-Janson G, Jodal U. Urinary infection, reflux and renal scarring in females continuously followed for 13–38 years. Pediatr Nephrol. 1995;9:131–6.
140. Wennerstrom M, Hansson S, Hedner T, Himmelmann A, Jodal U. Ambulatory blood pressure 16–26 years after the first urinary tract infection in childhood. J Hypertens. 2000;18:485–91.
141. Wennerstrom M, Hansson S, Jodal U, Sixt R, Stokland E. Renal function 16 to 26 years after the first urinary tract infection in childhood. Arch Pediatr Adolesc Med. 2000;154:339–45.
142. Shaikh N, Ewing AL, Bhatnagar S, Hoberman A. Risk of renal scarring in children with a first urinary tract infection: a systematic review. Pediatrics. 2010;126:1084–91.
143. Hannula A, Perhomaa M, Venhola M, Pokka T, Renko M, Uhari M. Long-term follow-up of patients after childhood urinary tract infection. Arch Pediatr Adolesc Med. 2012;166:1117–22.
144. Gebäck C, Hansson S, Martinell J, Sandberg T, Sixt R, Jodal U. Renal function in adult women with urinary tract infection in childhood. Pediatr Nephrol. 2015;30:1493–9.