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Synthesis of New N,N'-Bis(5-arylidene-4-oxo-4,5-dihydrothiazolin-2-yl)piperazine Derivatives Under Microwave Irradiation and Preliminary Biological Evaluation

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Abstract

New N,N'-bis(5-arylidene-4-oxo-4,5-dihydrothiazoline-2-yl)diamine derivatives 5 were prepared in two steps from rhodanine and piperazine, or 1,4-bis(3-amino-propyl)piperazine, under microwave reaction conditions with retention of configuration. Some of these compounds were tested for in vitro antiproliferative activities and for their kinase inhibitory potencies towards six kinases (CDK5/p25, GSK3α/β, DYRK1A, DYRK2, CLK1, and CLK2). The compound 5d showed nanomolar activity towards DYRK1A kinase (IC50 = 0.041 μM).

Keywords

5-Arylidene rhodanine • 2,2'-(Piperazine-1,4-diyl)bis(5-benzylidene-1,3-thiazol-4(5H)-one) • Microwave irradiation • Cytotoxicity • Protein kinase • Diamines
Introduction

The 2-amino-5-arylidene-5H-thiazol-4-ones and their 5-arylidene rhodanine precursors are a class of five-membered heterocyclic rings (FMHRs) considered as "privileged scaffolds" in the medicinal chemistry community [1]. Considerable work has been published over decades about their chemistry and biology. A number of compounds containing the 2-amino-5-arylidene-thiazol-4(5H)-one moiety have been shown to exhibit anti-inflammatory [2], antimicrobial [3], and antitumor [4] effects. Among these compounds, Darbufelone® A [5] (Figure 1) is orally active in animal models of inflammation [6] and DBPT B is under clinical trials for colon cancer [7]. 5-Arylidene rhodanines have also proven to be attractive for the discovery of new candidates. A series of rhodanine-based hits C (Figure 1) were found as potent and selective inhibitors of the "atypical" dual-specificity phosphatase (DSP) family member-JNK-stimulating phosphatase-1 (JSP-1). Compounds of this class may be useful for the treatment of inflammatory and proliferative disorders [8]. As the last example, epalrestat D reduced the symptoms of diabetic neuropathy [9].

Fig. 1. Structures of Darbufelone (A), DBPT (B), inhibitor of DSP (C), and epalrestat (D).

Protein kinases are the enzymes which control the phosphorylation of protein in cellular life [10] which is frequently deregulated in human diseases. For the pharmaceutical industry, the protein kinases represent interesting targets for new therapeutic agents [11] and this interest was boosted by the approval of the first marketed inhibitor Gleevec™ used in myeloid leukemia [12]. Due to the biological activity associated with the 2-amino-thiazolidinone moiety, we decided in this paper to explore the synthesis of N,N'-bis(5-arylidene-4-oxo-4,5-dihydrothiazolidin-2-yl)diamines derived from piperazine or 1,4-bis(3-aminopropyl)piperazine as linkers, and to study their effects on cells and protein kinases.
Results and Discussion

Chemistry

The strategy used for the synthesis of the symmetric derivatives 5 is outlined in Scheme 1. The reactions have been realized under microwave irradiation [13]. The main benefits of microwave irradiation technology are the significant rate-enhancements and sometimes elevated product yields enabling the rapid synthesis of molecules of potential value in medicinal chemistry [14]. The synthesis started by the solution-phase Knoevenagel condensation of aryl aldehydes 2 and commercial rhodanine 1a. The expected compounds 3a–c were prepared in yields ranging from 65 to 88% with a reaction time of 10 min. under microwave irradiation (MWI) at 65°C in the presence of sodium acetate. In a similar approach, the 5-arylidene rhodanine propanoic derivatives 3d–g were easily synthesized using solvent-less reaction conditions under microwave irradiation (130°C, 10 min.). The geometric double bond of 3 was attributed as being Z by the shielding effect of the carbonyl group C-4 on the olefinic proton H-5 (δ 7.5 ppm) in the ¹H NMR spectra.

\[
\begin{align*}
\text{R}^1 \quad \text{R}^2 \quad \text{R}^3
\end{align*}
\]

Sch. 1. a) for 3a–c: MeOH, AcONa 3 eq., MWI, 65°C, 10 min. and for 3d–g: MWI, 130°C, 10 min. b) MWI, 80–120°C, 30 min.
Transformation of 5-arylidene rhodanine 3 into 2-amino-5-arylidene-5\(\text{H}\)-thiazol-4-one after addition of a primary amine [15] usually involves activation of the C=S bond of rhodanine via the thioether intermediate that subsequently undergoes a thioalkyl/nitrogen displacement. In order to be able to carry out such sulfur/nitrogen displacement in a faster and more efficient way – avoiding the preparation of the thioether intermediate – we examined the influence of microwave irradiation on the reaction between compound 3 and the symmetric diamino linkers 4\(a,b\). The experiments revealed that optimal reaction conditions were obtained at 80–120°C after 30 minutes. It is noteworthy that sulfur/nitrogen displacement reactions at 120°C under microwave irradiation have been realized in solution with hexane to avoid decomposition of compound 5. The desired compounds 5\(a\)-e were prepared in poor to moderate yields (13–26%) and their structures were substantiated by \(^1\)H, \(^{13}\)C NMR, and HRMS analyses and only the more thermodynamically stable Z-isomers were obtained.

**Biology**

To evaluate the potency of the compounds 3\(a\)–c, 5\(a\), and 5\(c\) for their *in vitro* antiproliferative activities, we used six representative tumor cell lines of liver (Huh7), colon (Caco2, HCT 116), breast (MDA-MB 231), prostate (PC3), lung (NCI), and one normal cell line (fibroblasts) and measured survival. Results are reported in Table 1. The 5-arylidene rhodanines 3\(a\)–c and two compounds 5\(a\) and 5\(c\) showed measurable, albeit very poor, cytotoxic activity. No clear tendency for higher activity of these compounds can be deduced from these results. Since the log P values of all tested compounds are in the same range, influences of lipophilicity on cytotoxic activity cannot be deduced as well.

| Cpd. | Cell lines IC\(_{50}\) (μM) |
|------|------------------|
|      | Huh7 | Caco2 | MDA-MB 231 | HCT 116 | PC3 | NCI | Fibroblasts | Log P\(_{\text{calc.}}\) |
| 3a   | > 25  | > 25  | > 25       | > 25    | > 25| > 25| > 25       | 2.0     |
| 3b   | > 25  | > 25  | > 25       | > 25    | > 25| > 25| > 25       | 1.9     |
| 3c   | > 25  | > 25  | > 25       | > 25    | > 25| > 25| > 25       | 1.8     |
| 5a   | > 25  | 25    | > 25       | > 25    | > 25| > 25| > 25       | 4.8     |
| 5c   | > 25  | 20    | > 25       | > 25    | 40  | > 25| 40         | 5.3     |
|      | 10    | 10    | 15          | 8       | 8   | 20  | > 25       | –       |
| Roscovitine | 10 | > 25 | > 25 | > 25 | > 25 | > 25 | > 25 | – |
| DMSO | > 25  | > 25  | > 25       | > 25    | > 25| > 25| > 25       | –       |

The kinase inhibitory potencies of compounds 3\(a\)–c and 5\(d\) were evaluated as IC\(_{50}\) values towards six protein kinases (CDK5/p25, GSK3\(\alpha/\beta\), DYRK1A, DYRK2, CLK1 and CLK2) and the results are reported in Table 2. The 5-arylidene rhodanines 3\(a,b\) appeared to be inactive towards these six protein kinases, but interesting results were obtained with compounds 5\(d\) and 3\(c\). The 1,4-bis[(5\(Z\))-5-(4-methoxybenzylidene)methylene-4-oxo-4,5-dihydrothiazol-2-yl]piperazine 5\(b\) and the 5-arylidene rhodanine 3\(c\) are very active on DYRK1A [16] and a noteworthy IC\(_{50}\) = 41 nM was measured for 5\(b\). The compound 5\(d\) has also shown submicromolar inhibition potencies towards DYRK2 (IC\(_{50}\) = 0.6 μM) and CLK1 (IC\(_{50}\) = 0.5 μM). Regarding these results, the potential of the (5\(Z\)) 5-arylidene-4-oxo-4,5-...
dihydro-thiazolidine-2-yl moiety appended on 1,4-bis(3-aminopropyl)piperazine 4b as a linker could be highly interesting in the development of a new class of inhibitors of DYRK1A kinase which is known to be involved in Alzheimer's disease/Down syndrome [17].

Tab. 2. CDK5/p25, GSK3α/β, DYRK1A, DYRK2, CLK1, CLK3 inhibitions.

| Cpd. | CDK5/p25 (μM) | GSK3α/β (μM) | DYRK1A (μM) | DYRK2 (μM) | CLK1 (μM) | CLK2 (μM) |
|------|---------------|--------------|-------------|-------------|------------|------------|
| 3a   | > 10          | > 10         | > 10        | > 10        | > 10       | > 10       |
| 3b   | > 10          | > 10         | > 10        | > 10        | > 10       | > 10       |
| 3c   | > 10          | 8.5          | 0.070       | –           | –          | –          |
| 5d   | > 10          | > 10         | 0.041       | 0.6         | 0.5        | 7.7        |

Conclusion

In conclusion, we worked out a short and practical synthesis under microwave irradiation of \(N,N'-(5\text{-arylidene-4-oxo-4,5-dihydrothiazolin-2-yl})\)diamines 5a–e derived from piperazine 4a [18, 19] and 1,4-bis(3-aminopropyl)piperazine 4b. The \textit{in vitro} anti-proliferative activities are extremely weak and this could be due to a lower cellular penetration of compounds 3 and 5 or a lower interaction with the cellular targets. Surprisingly, the compound 5d has shown nanomolar inhibition potency towards DYRK1A and this interesting inhibition led us to expand our efforts in the synthesis of diversely disubstituted \(N,N'\)-bispiperazine derivatives with the 5-arylidene thiazolidinone moiety as new potential inhibitors of this kinase. Work is in progress to gain deeper insight into the structure-activity relationships (SAR) of this new interesting class of diamines.

Experimental

\textit{General}

Elemental analysis: Flash EA1112 CHN/O Thermo Electron; HRMS (MS/MS ZABSpec Tof Micromass, EBE TOF geometry, IP 8 eV); NMR: BRUKER AC 300P (\(^1\text{H}: 300 \text{ MHz}, \ ^{13}\text{C}: 75 \text{ MHz}\)); melting points: Leica System Kofler VMHB Melting Point apparatus (not corrected); microwave reactor: Monowave 300 Anton-Paar (850 W), monowave software package.

\textit{General procedure I: Preparation of 5-arylidene-2-thioxo-thiazolidine-4-one 3a–c under microwave irradiation.}

A mixture of rhodanine 1a (1 g, 7.5 mmol), aldehyde 2 (8.95-9 mmol), sodium acetate (1.85 g, 22.55 mmol), and methanol (5 ml) was placed in a borosilicate glass vial (10 ml) with a Teflon® magnetic stir bar and sealed with a snap cap. The glass tube was then introduced into the Monowave 300 Anton-Paar microwave cavity (P = 850 Watt) and the stirred mixture was irradiated at 65°C (with a power of 10 Watt) for 30 min. After microwave dielectric heating, the crude reaction mixture was allowed to cool down to room temperature, and then was filtered on a Buchner funnel and the insoluble compound 3 was washed with methanol (2x5 ml). The crude compound 3 was purified by recrystallization.
from methanol and further dried under high vacuum (10^{-2} Torr) for 1 hour, which gave the desired compound 3 as a powder.

**General procedure II: Preparation of 3-(5-arylidene-4-oxo-2-thioxo-thiazolidine-3-yl)propanoic acid 3d–g under solvent-free microwave irradiation.**

A mixture of rhodanine 1b (0.5 g, 2.44 mmol) and aldehyde 2 (2.44 mmol) was placed in a borosilicate glass vial (10 ml) with a Teflon® magnetic stir bar and sealed with a snap cap. The glass tube was then introduced into the Monowave 300 Anton-Paar microwave cavity (P = 850 Watt) and the stirred mixture was irradiated at 130°C for 10 min. After microwave dielectric heating, the crude reaction mixture was allowed to cool down to room temperature and 5 ml of a mixture of ethanol/hexane (1:1) were added directly to the glass vial. The resulting precipitated product 3 was filtered on a Buchner funnel and the insoluble compound 3 was washed with the same mixture (2x5 ml). The crude compound 3 was further dried under high vacuum (10^{-2} Torr) for 1 hour, which gave the desired compound 3 as a powder.

**General procedure III: Synthesis of N,N’-bis(5-arylidene-4-oxo-4,5-dihydro-thiazolidine-2-yl)piperazine derivatives 5a–e under microwave irradiation.**

A mixture of 5-arylidene-2-thioxo-thiazolidine-4-one 3 (4 mmol), piperazine 4a (172 mg, 2 mmol) or 1,4-bis(3-aminopropyl)piperazine 4b (400 mg, 2 mmol) was placed in a borosilicate glass vial (10 ml) with a Teflon® magnetic stir bar and sealed with a snap cap. The glass tube was then introduced into the Monowave 300 Anton-Paar microwave cavity (P = 850 Watt) and the stirred mixture was irradiated at 80–120°C (with a power of 5–200 Watt) for 30 min. After microwave dielectric heating, the crude reaction mixture was allowed to cool down to room temperature and 10 ml of cooled ethanol (4°C) were added directly in the glass vial. The resulting precipitated product 5 was filtered off, washed with 2 x 5 ml of ethanol and dried under high vacuum (10^{-2} Torr) at room temperature for 1 hour. After ^1H NMR analysis, the product 5 was purified by recrystallization from methanol, which gave the desired compound 3 as a powder.

**(5Z)-5-Benzylidene-2-thioxo-1,3-thiazolidin-4-one (3a)**

Prepared following General procedure I from benzaldehyde 2a (0.95 g, 8.95 mmol) to give 1.08 g (65%) as an orange powder. Mp = 208–210°C. ^1H NMR (300 MHz, DMSO-d_6) δ (ppm) = 7.44–7.55 (m, 5H, Ar), 7.57 (s, 1H, =CH), 13.12 (br s, 1H, NH). ^13C NMR (75 MHz, DMSO-d_6) δ (ppm) = 126.88 (C-1), 129.20 (C-2), 130.30 (C-3), 130.37 (C-4), 130.45 (C=, C-5), 133.22 (=CH), 171.28 (C=S, C-2); 196.64 (C=O, C-4). HRMS, m/z = 243.9868 found (calculated for C_{10}H_{7}NONaS_{2}, [M+Na]^+ requires 243.9867). Anal. Calcd. for C_{10}H_{7}NOS_{2}: C, 54.27; H, 3.19. Found: C, 54.23; H, 3.16.

**(5Z)-5-(4-Methoxybenzylidene)-2-thioxo-1,3-thiazolidin-4-one (3b)**

Prepared following General procedure I from methoxybenzaldehyde 2b (1.23 g, 9 mmol) to give 1.36 g (74%) as an orange powder. Mp = 250–252°C. ^1H NMR (300 MHz, DMSO-d_6) δ (ppm) = 3.82 (s, 3H, OCH_3), 7.10 (d, J = 8.8Hz, 2H, H-3, Ar), 7.55 (d, J = 9.1 Hz, 2H, H-2, Ar), 7.57 (s, 1H; =CH), 13.76 (br s, 1H, NH). ^13C NMR (75 MHz, DMSO-d_6) δ (ppm) = 55.45 (OCH_3), 114.92 (C-3’), 124.90 (C=, C-5), 126.02 (C-1’), 129.71 (=CH), 132.28 (C-2’), 160.82 (C-4’), 173.024 (C=S, C-2), 197.25 (C=O, C-4). HRMS, m/z = 295.9792 found.
(calculated for C₁₁H₉NO₂Na₂S₂, [M-H+2Na]⁺ requires 295.9792). Anal. Calcd. for C₁₁H₉NO₂S₂: C, 52.57; H, 3.61. Found: C, 52.65; H, 3.67.

(5Z)-5-(1,3-Benzodioxol-5-ylmethylidened)-2-thioxo-1,3-thiazolidin-4-one (3c)
Prepared following General procedure I from piperonaldehyde 2c (1.35 g, 9 mmol) to give 1.74 g (88%) as an orange powder. Mp > 260°C. ¹H NMR (300 MHz, DMSO-d₆) δ (ppm) = 6.13 (s, 2H; OCH₂O), 7.11–7.45 (m, 3H; H-5', H-6', Ar), 7.52 (s, 1H, =CH), 7.55 (s, 1H, H-2'; Ar), 13.74 (br s, 1H, NH). ¹³C NMR (75 MHz, DMSO-d₆) δ (ppm) = 102.10 (OCH₂O), 109.26 (C=, C-5), 109.45 (C-6'), 123.02 (C-1'), 126.65 (C-5'), 127.16 (=CH), 131.81 (C-2'), 148.27 (C-4'), 149.60 (C-3'), 169.58 (C=S, C-2), 195.51 (C=O, C-4). HRMS, m/z = 287.9765 found (calculated for C₁₁H₇NO₃NaS₂, [M+Na]⁺ requires 287.9765). Anal. Calcd. for C₁₁H₇NO₃S₂: C, 49.80; H, 2.66. Found: C, 49.91; H, 2.69.

3-[(5Z)-5-Benzylidene-4-oxo-2-thioxo-1,3-thiazolidin-3-yl]propanoic acid (3d)
Prepared following General procedure II from benzaldehyde 2a (0.256 g, 2.44 mmol) to give 3d in 60% yield as a brown powder. Mp = 224–226°C. ¹H NMR (300 MHz, DMSO-d₆) δ (ppm) = 2.33 (m, 2H, CH₂), 4.15 (m, 2H, CH₂), 7.51–7.65 (m, 5H, H-2, H-3, H-4, Ar), 7.80 (s, 1H, =CH), 13.16 (br s, 1H, OH). ¹³C NMR (75 MHz, DMSO-d₆) δ (ppm) = 33.30 (CH₂), 41.78 (CH₂), 122.6 (C=, C-5), 129.46 (C-2'), 130.57 (C-3'), 130.84 (C-4'), 132.63 (=CH), 133.02 (C-1'), 166.76 (C=O, C-4), 171.94 (CO₂H), 193.15 (C=S, C-2). HRMS, m/z = 337.9904 found (calculated for C₁₃H₁₀NO₃Na₂S₂, [M+Na]⁺ requires 337.9898).

3-[(5Z)-5-(4-Methoxybenzylidene)-4-oxo-2-thioxo-1,3-thiazolidin-3-yl]propanoic acid (3e)
Prepared following General procedure II from 4-methoxybenzaldehyde 2b (0.332 g, 2.44 mmol) to give 3e in 60% as a brown powder. Mp > 260°C. ¹H NMR (300 MHz, DMSO-d₆) δ (ppm) = 2.24 (t, 2H, J = 8.2 Hz, CH₂), 3.83 (s, 3H, OCH₃), 4.12 (t, 2H, J = 8.2 Hz, CH₂), 7.10 (d, 2H, J = 8. 8 Hz, H-3, Ar), 7.64 (d, 2H, J = 8.8 Hz, H-2, Ar), 7.86 (s, 1H, =CH), 13.26 (br s, 1H, OH). ¹³C NMR (75 MHz, DMSO-d₆) δ (ppm) = 34.11 (CH₂), 42.20 (CH₂), 55.56 (OCH₃), 115.11 (C-3'), 119.26 (C=, C-5), 125.58 (C-1'), 125.90 (=CH), 132.84 (C-2'), 161.41 (C-4'), 166.86 (C=O, C-4), 171.98 (CO₂H), 192.86 (C=S, C-2). HRMS, m/z = 346.0185 found (calculated for C₁₄H₁₃NO₄Na₂S₂, [M+Na]⁺ requires 346.0184).

3-[(5Z)-5-(1,3-Benzodioxol-5-ylmethylidene)-4-oxo-2-thioxo-1,3-thiazolidin-3-yl]propanoic acid (3f)
Prepared following General procedure II from piperonaldehyde 2c (0.366 g, 2.44 mmol) to give 3f in 63% as a brown powder. Mp = 220–224°C. ¹H NMR (300 MHz, DMSO-d₆) δ (ppm) = 2.61 (t, 2H, J = 3.9 Hz, CH₂), 4.21 (t, 2H, J = 4.7 Hz, CH₂), 6.15 (s, 2H, OCH₂O), 7.09–7.23 (m, 3H, H-2, H-5, H-6, Ar), 7.74 (s, 1H, =CH), 12.50 (br s, 1H, OH). ¹³C NMR (75 MHz, DMSO-d₆) δ (ppm) = 30.74 (CH₂), 40.30 (CH₂), 102.21 (OCH₂O), 109.35 (C-2'), 109.61 (C-5'), 119.71 (C=, C-5), 126.98 (C-6'), 127.14 (C-1'), 133.21 (=CH), 148.35 (C-4'), 149.88 (C-3'), 166.67 (C=O, C-4), 171.72 (CO₂H), 192.97 (C=S, C-2). HRMS, m/z = 346.0185 found (calculated for C₁₄H₁₃NO₄Na₂S₂, [M+Na]⁺ requires 346.0184).

3-[(5Z)-5-(2,3-Dihydro-1,4-benzodioxin-6-ylmethylidene)-4-oxo-2-thioxo-1,3-thiazolidin-3-yl]propanoic acid (3g)
Prepared following General procedure II from 1,4-benzodioxane-6-carboxaldehyde (2d)
(0.401 g, 2.44 mmol) to give 3g in 54% yield as a brown powder. Mp = 226–228°C. $^1$H NMR (300 MHz, DMSO-$d_6$) $\delta$ (ppm) = 2.55 (t, 2H, J = 7.3 Hz, CH$_2$), 4.18 (t, 2H, J = 7.4 Hz, CH$_2$), 4.30 (m, 4H, OCH$_2$CH$_2$O), 7.02 (d, 1H, J = 8.6 Hz, H-5, Ar), 7.12 (d, 1H, J = 2.2 Hz, H-6, Ar), 7.15 (s, 1H, H-2, Ar), 7.70 (s, 1H, =CH), 12.46 (br s, 1H, OH). $^{13}$C NMR (75 MHz, DMSO-$d_6$) $\delta$ (ppm) = 31.30 (CH$_2$), 40.30 (CH$_2$), 64.01 (OCH$_2$CH$_2$O), 118.22 (C-2'), 119.47 (C-5'), 119.79 (C=, C-5), 124.46 (C-6'), 126.31 (C-1'), 132.96 (=CH), 143.82 (C-4'), 146.25 (C-3'); 166.67 (C=O, C-4), 171.80 (CO$_2$H), 192.93 (C=S, C-2). HRMS, m/z = 374.0138 found (calculated for C$_{15}$H$_{13}$NO$_5$NaS$_2$, [M+Na]$^+$ requires 374.0133).

$(5Z,5'Z)$-2,2'-(Piperazine-1,4-diyl)bis(5-benzylidene-1,3-thiazol-4(5H)-one) (1,4-Bis[(5Z)-5-benzylidene-4-oxo-4,5-dihydro-1,3-thiazol-2-yl]piperazine, 5a)

Prepared following General procedure III from 5-benzylidene-2-thioxothiazolidin-4-one (3a) (885 mg, 4 mmol) and piperazine 4a (172 mg, 2 mmol) in hexane (2 ml) at 120°C to give 5a in 13% yield as a brown powder. Mp > 260°C. $^1$H NMR (300 MHz, DMSO-$d_6$) $\delta$ (ppm) = 3.34 (br s, 8H, N(CH$_2$)$_4$N), 7.28-7.7 (m, 12H, H-2', H-3', H-4', Ar), 7.16 (s, 2H, =CH), 7.43 (d, J = 8.7 Hz, 4H, H-2', Ar). $^{13}$C NMR (75 MHz, DMSO-$d_6$) $\delta$ (ppm) = 42.45 (N(CH$_2$)$_4$N), 129.08 (C-2'), 129.94 (C-1'), 130.41 (C=N), 159.56 (C=O). HRMS, m/z = 483.0918 found (calculated for C$_{24}$H$_{20}$N$_4$O$_2$NaS$_2$, [M+Na]$^+$ requires 483.0925). Anal. Calcd. for C$_{24}$H$_{20}$N$_4$O$_2$S$_2$: C, 62.59; H, 4.38. Found: C, 62.56; H, 4.41.

$(5Z,5'Z)$-2,2'-(Piperazine-1,4-diyl)bis[5-(4-methoxybenzylidene)-1,3-thiazol-4(5H)-one] (1,4-Bis[(5Z)-5-(4-methoxybenzylidene)-4-oxo-4,5-dihydro-1,3-thiazol-2-yl]piperazine, 5b)

Prepared following General procedure III from 5-(4-methoxybenzylidene)-2-thioxothiazolidin-4-one (3b) (1.005 g, 4 mmol) and piperazine 4a (172 mg, 2 mmol) at 80°C to give 5b in 36% yield as a yellow powder. Mp > 260°C. $^1$H NMR (300 MHz, DMSO-$d_6$) $\delta$ (ppm) = 2.99 (s, 8H, N(CH$_2$)$_4$N), 3.79 (s, 6H, OCH$_3$), 7.04 (d, J = 5 Hz, 4H, H-3', Ar), 7.16 (s, 2H, =CH), 7.43 (d, J = 8.7 Hz, 4H, H-2', Ar). $^{13}$C NMR (75 MHz, DMSO-$d_6$) $\delta$ (ppm) = 42.99 (N(CH$_2$)$_4$N), 55.27 (OCH$_3$), 114.54 (C-3'), 114.73 (C-1'), 124.13 (=CH), 127.55 (C=, C-5), 131.29 (C-2'), 131.38 (C-4'), 132.14 (C=N, C-2), 159.56 (C=O). HRMS, m/z = 521.1317 found (calculated for C$_{26}$H$_{25}$N$_4$O$_4$S$_2$, [M+H]$^+$ requires 521.1310). Anal. Calcd. for C$_{26}$H$_{24}$N$_4$O$_4$S$_2$: C, 59.98; H, 4.65. Found: C, 59.87; H, 4.64.

$(5Z,5'Z)$-2,2'-(Piperazine-1,4-diyl)bis[5-(1,3-benzodioxol-5-ylmethylene)-1,3-thiazol-4(5H)-one] (1,4-Bis[(5Z)-5-(1,3-benzodioxol-5-ylmethylene)-4-oxo-4,5-dihydro-1,3-thiazol-2-yl]piperazine, 5c)

Prepared following General procedure III from 5-(1,3-benzodioxol-5-ylmethylene)-2-thioxothiazolidin-4-one (3c) (1.061 g, 4 mmol) and piperazine 4a (172 mg, 2 mmol) in hexane (2 ml) at 120°C to give 5a in 15% yield as a brown powder. Mp > 260°C. $^1$H NMR (300 MHz, DMSO-$d_6$) $\delta$ (ppm) = 3.02 (s, 8H, N(CH$_2$)$_4$N), 6.11 (s, 4H, OCH$_2$O), 7.02–7.18 (m, 6H, H-2', H-5', H-6', Ar), 7.58 (s, 2H, =CH). $^{13}$C NMR (75 MHz, DMSO-$d_6$) $\delta$ (ppm) = 52.72 (N(CH$_2$)$_4$N), 101.88 (OCH$_2$O), 108.71 (C-2'), 125.13 (C-5'), 126.04 (C=, C-5), 127.86 (C-1'), 130.41 (C-6'), 144.21 (=CH), 146.02 (C=N, C-2), 148.11 (C-4'), 148.78 (C-3'), 182.22 (C=O, C-4). HRMS, m/z = 571.0726 found (calculated for C$_{26}$H$_{20}$N$_4$O$_6$NaS$_2$, [M+Na]$^+$ requires 571.0722). Anal. Calcd. for C$_{26}$H$_{20}$N$_4$O$_6$S$_2$: C, 56.92; H, 3.67. Found: C, 56.96; H, 3.71.
Prepared following General procedure III from 5-(4-methoxybenzylidene)-2-thioxothiazolidin-4-one (3b) (1.005 g, 4 mmol) and 1,4-bis(3-aminopropyl)piperazine (4b) (0.4 g, 2 mmol) in hexane (2 ml) at 120°C to give 5d in 25% yield as a yellow powder. Mp = 220–222°C. 1H NMR (300 MHz, DMSO-d6) δ (ppm) = 1.68 (quint, J = 6.9 Hz, 4H, NCH2CH2CH2N(CH2)4N), 2.37 (t, J = 6.7 Hz; 4H, NCH2CH2CH2N(CH2)4N), 2.57 (s, 8H, NCH2CH2CH2N(CH2)4N), 2.84 (t, J = 7.1 Hz, 4H; NCH2CH2CH2N(CH2)4N), 3.79 (s, 6H, OCH3), 7.02 (d, J = 8.8 Hz, 4H, H-3', Ar), 7.18 (s, 2H; =CH), 7.44 (d, J = 8.8 Hz, 4H, H-2', Ar). 13C NMR (75 MHz, DMSO-d6) δ (ppm) = 23.39 (NCH2CH2CH2N(CH2)4N), 37.41 (NCH2CH2CH2N(CH2)4N), 52.03 (NCH2CH2CH2N(CH2)4N), 54.42 (NCH2CH2CH2N(CH2)4N), 114.58 (C-3'), 125.04 (=CH), 127.28 (C-1'), 130.99 (C=), 131.44 (C-2'), 131.44 (C-1'), 135.77 (C-4'), 201.53 (C=O, C-4). HRMS, m/z = 635.2396 found (calculated for C32H39N6O4S2, [M+H]+ requires 635.1819). Anal. Calcd. for C32H38N6O4S2: C, 60.54; H, 6.03. Found: C, 60.57; H, 6.07.

Prepared following General procedure III from 5-(1,3-benzodioxol-5-ylmethylene)-2-thioxothiazolidin-4-one (3c) (1.061 g, 4 mmol) and 1,4-bis(3-aminopropyl)piperazine (4b) (0.4 g, 2 mmol) in hexane (2 ml) at 120°C to give 5e in 15% yield as a brown powder. Mp = 222–224°C. 1H NMR (300 MHz, DMSO-d6) δ (ppm) = 1.69 (quint, J = 6.9 Hz, 4H, NCH2CH2CH2N(CH2)4N), 2.37 (t, J = 6.6 Hz, 4H, NCH2CH2CH2N(CH2)4N), 2.37 (s, 8H, NCH2CH2CH2N(CH2)4N), 2.83 (t, J = 7.1 Hz, 4H; NCH2CH2CH2N(CH2)4N), 5.07 (s, 4H, OCH2O), 6.07 (s, 4H, OCH2O), 7.02 (m, 6H, H-2', H-5', H-6', Ar), 7.14 (s, 2H, =CH). 13C NMR (75 MHz, DMSO-d6) δ (ppm) = 23.47 (NCH2CH2CH2N(CH2)4N), 37.45 (NCH2CH2CH2N(CH2)4N), 52.22 (NCH2CH2CH2N(CH2)4N), 54.54 (NCH2CH2CH2N(CH2)4N), 55.27 (OCH3), 101.52 (OCH2O), 108.74 (C-2'), 108.82 (C-5'), 124.21 (C-6'), 124.84 (=CH), 147.72 (C-4'), 147.89 (C-3'), 182.66 (C=, C-5), 202.14 (C=O, C-4). HRMS, m/z = 663.1981 found (calculated for C32H35N6O6S2, [M+H]+ requires 663.7790). Anal. Calcd. for C32H34N6O6S2: C, 57.99; H, 5.17. Found: C, 57.92; H, 5.16.

In vitro kinase inhibition assays

Buffer A: 10 mM MgCl2, 1 mM EGTA, 1 mM DTT, 25 mM Tris-HCl pH 7.5, 50 µg heparin/ml. Buffer C: 60 mM β-glycerophosphate, 15 mM p-nitrophenylphosphate, 25 mM Mops (pH 7.2), 5 mM EGTA, 15 mM MgCl2, 1 mM DTT, 1 mM sodium vanadate, 1 mM phenylphosphate. Kinase activities were assayed in Buffer A or C, at 30°C, at a final ATP concentration of 15 µM. Blank values were subtracted and activities expressed in % of the maximal activity, i.e. in the absence of inhibitors. Controls were performed with appropriate dilutions of DMSO. The kinase peptide substrates were obtained from Millegen (Labege, France). DYRK1A and DYRK2 (human, recombinant, expressed in E. coli as a GST fusion protein) were purified by affinity chromatography on glutathione-agarose and assayed in buffer A (+ 0.5 mg BSA /ml) using Woodtide (KKISGRLSPIMTEQ) (1.5 µg/assay) as a
substrate, in the presence of 15 µM [γ-33P] ATP (3,000 Ci/mmol; 10 mCi/ml) in a final volume of 30 µl. After 30 min incubation at 30°C, the reaction was stopped by harvesting onto P81 phosphocellulose papers (Whatman) using a FilterMate harvester (Packard) and were washed in 1% phosphoric acid. Scintillation fluid was added and the radioactivity was measured in a Packard counter. CLK1 and CLK3 (human, recombinant, expressed in E. coli as GST fusion proteins) were assayed in buffer A (+ 0.15 mg BSA /ml) with RS peptide (GRSRSRSRSRSR) (1µg/assay). CDK5/p25 (human, recombinant) was prepared as previously described [26]. Its kinase activity was assayed in buffer C, with 1 µg histone H1 /ml. GSK-3α/β (porcine brain, native) was assayed in Buffer A and using a GSK-3 specific substrate (GS-1: YRRAAVPPSPSLRSRHSSPHQSpEDEEE) (pS stands for phosphorylated serine) [20].

**Cell culture and survival assays**

Skin diploid fibroblastic cells were provided by BIOPREDIC International Company (Rennes, France). Caco-2 cells and Huh7 cells were obtained from the ECAC collection. Cells were grown according to ECAC recommendations. The RLEC-F1 clone was derived from an established rat biliary epithelial cell line as previously described [21]. The toxicity test of the compounds on these cells was as follows: 4 x 10³ cells were seeded in 96 multiwell plates and left for 24 h for attachment, spreading, and growing. Then they were exposed for 48 h to increasing concentrations of the compounds, ranging from 0.1 to 25 µL in a final volume of 80 µL of culture medium. They were then fixed with 4% paraform-aldehyde solution and the nuclei were stained with Hoechst 3342 and counted using automated imaging analysis (Simple PCI software).

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**Authors’ Statement**

**Competing Interests**

The authors declare no conflict of interest.

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