Comparing Antimicrobial Susceptibilities among *Mycoplasma pneumoniae* Isolates from Pediatric Patients in Japan between Two Recent Epidemic Periods

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**ABSTRACT** We compared the antimicrobial susceptibility of *Mycoplasma pneumoniae* isolates from pediatric patients in Japan in 2011–2012 and 2015–2016, when epidemics occurred. The antimicrobial activity of macrolides and tetracyclines against *M. pneumoniae* infection tended to be restored in 2015–2016. There was no change in the antimicrobial activity of quinolones against *M. pneumoniae* infection.

**KEYWORDS** antimicrobial susceptibility, children, epidemic, Japan, *Mycoplasma pneumoniae*

*Mycoplasma pneumoniae* is an important pathogen that causes human respiratory tract infection, particularly in children and young adults. Epidemics of *M. pneumoniae* infection occur in 3- to 5-year cycles. In 2011–2012 and 2015–2016 in Japan, the number of patients increased by 2-fold the number in a typical year (1).

Macrolides are the first-line treatments for respiratory tract infections caused by *M. pneumoniae* (2). However, macrolide-resistant (MR) *M. pneumoniae* isolates were detected in Japanese pediatric patients in 2001 for the first time worldwide and have become widespread in Japan (3). The rate of MR *M. pneumoniae* infection was as high as 80% among pediatric patients in Japan in 2009 to 2011 (4). We also investigated the prevalence of MR *M. pneumoniae* since 2008 (5) and reported that the prevalence of MR *M. pneumoniae* among pediatric patients decreased from 74.6% to 49.5% between 2011 and 2015 in Japan (6).

Tetracyclines or quinolones are recommended for treatment of MR *M. pneumoniae* infection. Second-line treatments, such as tetracycline and quinolones, are increasingly used because of the increase in MR *M. pneumoniae* cases in Japan (2).

It is important to conduct surveillance of the susceptibilities of *M. pneumoniae* isolates to tetracyclines, quinolones, and macrolides. We previously reported that quinolones exhibited potent antimicrobial activity against both MR and macrolide-sensitive (MS) *M. pneumoniae* isolates from pediatric patients in 2009 to 2011 (7). However, there are no recent reports of antimicrobial activity against *M. pneumoniae* infection.

We investigated the antimicrobial susceptibility of *M. pneumoniae* isolates from pediatric patients in Japan in 2011 to 2016 and compared the cumulative distributions of the MICs of macrolides, quinolones, and tetracyclines in 2011–2012 and 2015–2016.

We enrolled all pediatric patients with acute respiratory tract infections at 85 institutions located in 8 areas throughout Japan (20 institutions in Kyushu, 25 in Chugoku, 3 in Shikoku, 11 in Kinki, 7 in Chubu, 3 in Kanto, 2 in Tohoku, and 3 in Hokkaido) in 2011 to 2016.
Pediatricians at the facilities collected samples from patients with suspected *M. pneumoniae* infections. Informed consent was obtained from the parents of all patients. The Ethics Committee at Kawasaki Medical School, Kurashiki, Japan, approved the study protocol on 15 October 2018 (no. 3119-1).

*M. pneumoniae* isolates were obtained by cultivation of specimens. The medium used for isolation and determination of the MIC was pleuropneumonia-like organism broth (PPLO) (Oxoid, Hampshire, UK) supplemented with 0.5% glucose (FUJIFILM Wako Pure Chemical Corporation, Osaka, Japan), 20% mycoplasma supplement G (Oxoid), and 0.0025% phenol red (Sigma-Aldrich, St. Louis, MO).

The MICs of antimicrobial agents for the isolated strains were determined with microdilution methods (8). Briefly, medium containing 10⁵ to 10⁶ CFU/ml of *M. pneumoniae* was added to 96-well microplates and incubated at 37°C for 6 to 8 days.

MIC was defined as the lowest concentration of antimicrobial agent at which the metabolism of the organism was inhibited, which was evidenced by the lack of a color change in the medium 3 days after the drug-free control first showed a color change.

The reference strain FH was used as a drug-susceptible control. The antimicrobial agents used for MIC determination were erythromycin, clarithromycin, azithromycin, clindamycin, minocycline, tetracycline, tosufloxacin, garenoxacin, and levofloxacin.

### Table 1: In vitro antimicrobial activity against clinical isolates of *Mycoplasma pneumoniae* strains

| Organism (no. of strains) | Antimicrobial agent* | MIC (µg/ml) | Range | MIC₅₀ | MIC₉₀ |
|---------------------------|----------------------|-------------|-------|-------|-------|
| *Mycoplasma pneumoniae* (1,256) | TFLX                  | 0.0625 to 0.5 | 0.25  | 0.25  |
|                           | GRNX                  | 0.0078 to 0.125 | 0.0313 | 0.0313 |
|                           | LVFX                  | 0.25 to 1     | 0.5   | 0.5   |
|                           | TC                    | 0.125 to 1    | 0.5   | 0.5   |
|                           | MINO                  | 0.125 to 4    | 1     | 2     |
|                           | CLDM                  | 0.25 to >128  | 64    | 128   |
|                           | EM                    | 0.001 to >128 | >128  | >128  |
|                           | CAM                   | 0.00025 to >128 | >128  | >128  |
|                           | AZM                   | 0.0000313 to >128 | 32    | 64    |
| Macrolide-susceptible *M. pneumoniae* (383) | TFLX                  | 0.0625 to 0.5 | 0.25  | 0.5   |
|                           | GRNX                  | 0.0156 to 0.125 | 0.0313 | 0.0625 |
|                           | LVFX                  | 0.25 to 1     | 0.5   | 0.5   |
|                           | TC                    | 0.125 to 1    | 0.5   | 0.5   |
|                           | MINO                  | 0.125 to 4    | 1     | 2     |
|                           | CLDM                  | 0.25 to 4     | 1     | 1     |
|                           | EM                    | 0.001 to 2    | 0.0039 | 0.0078 |
|                           | CAM                   | 0.00025 to 0.5 | 0.002 | 0.0039 |
|                           | AZM                   | 0.0000313 to 0.0313 | 0.00025 | 0.0005 |
| Macrolide-resistant *M. pneumoniae* (873) | TFLX                  | 0.0625 to 0.5 | 0.25  | 0.25  |
|                           | GRNX                  | 0.0078 to 0.063 | 0.0313 | 0.0313 |
|                           | LVFX                  | 0.25 to 1     | 0.5   | 0.5   |
|                           | TC                    | 0.125 to 1    | 0.5   | 0.5   |
|                           | MINO                  | 0.125 to 4    | 1     | 2     |
|                           | CLDM                  | 4 to >128     | 128   | 128   |
|                           | EM                    | 8 to >128     | >128  | >128  |
|                           | AZM                   | 0.25 to >128  | 32    | 64    |

*TFLX, tosufloxacin; GRNX, garenoxacin; LVFX, levofloxacin; TC, tetracycline; MINO, minocycline; EM, erythromycin; CAM, clarithromycin; AZM, azithromycin.*
than those of the other antimicrobials. All quinolones, particularly garenoxacin, showed potent antimicrobial activity against MR *M. pneumoniae*, with MIC<sub>50</sub>/MIC<sub>90</sub> values of 0.0313/0.0313 µg/ml. These values were equal to those of MS *M. pneumoniae* isolates. Tosufloxacin, the only quinolone approved for treatment of pneumonia in pediatric patients in Japan, also showed good activity against MR and MS *M. pneumoniae* isolates, with MIC<sub>50</sub>/MIC<sub>90</sub> values of 0.25/0.25 and 0.25/0.5 µg/ml, respectively. Tetracyclines, such as tetracycline and minocycline, showed comparably good activity against MR and MS *M. pneumoniae* isolates.

Figure 1 shows the MIC distribution of macrolides, quinolones, and tetracyclines in 2011–2012 and 2015–2016 and statistical analysis of the differences in each MIC value between the two periods by the Wilcoxon rank-sum test. The resistance rate of erythromycin, clarithromycin, and azithromycin decreased from 75%, 74%, and 71.9% in 2011–2012 to 54.2%, 54.2%, and 53.1% in 2015–2016, respectively. The MIC values of macrolides and tetracyclines in 2015–2016 were significantly lower than those in 2011–2012. The antimicrobial activity of quinolones remained potent in 2016. Strains resistant to these agents were not detected in this study.

In comparing the two periods when *M. pneumoniae* epidemics occurred (2011–2012 and 2015–2016), the antimicrobial activities of all macrolides and tetracyclines against *M. pneumoniae* isolates were restored significantly in 2015–2016. The sensitivity to macrolides may have been restored because of a decrease in *M. pneumoniae* isolates with specific point mutations in domain V of the 23S rRNA gene (6).

We considered two reasons for recovery of the sensitivity to macrolides. One is the appropriate use of tosufloxacin for treating *M. pneumoniae* infection, and the other is a shift in the P1 type.

First, tosufloxacin was approved in 2010 in Japan as treatment for pediatric patients and is recommended for use in patients with suspected MR *M. pneumoniae* infection as a second-line drug under various guidelines (2). Specifically, tosufloxacin is recommended for cases with *M. pneumoniae* infection in which fevers are not reduced by 48 to 72 h after the initiation of macrolide treatment. Ouchi et al. (10) reported that tosufloxacin was significantly more effective than clarithromycin in eradicating MR *M. pneumoniae*. Additionally, total oral antimicrobial use of macrolides decreased, whereas that of quinolones, including tosufloxacin, increased from 2011 to 2013 in children (age, 0 to 14 years), based on analysis of health insurance claim data in the national database (11). Miyashita et al. (12) reported lower macrolide resistance rates of *M. pneumoniae* infection in adults to whom macrolides, tetracyclines, or respiratory quinolones were commonly administered than in children to whom only macrolides or tetracyclines were administered in 2008 to 2011. Thus, because tosufloxacin was used appropriately for *M. pneumoniae* infections, the development of MR *M. pneumoniae* was prevented.

Second, a type shift in p1 may explain the recovery of sensitivity to macrolides. At the surface of the attachment organelle is the 170-kDa adhesin protein P1, which is densely clustered and plays a major role in binding to the receptor molecule of host epithelial cells (13). Two major subtypes of p1 (subtypes 1 and 2) are known that form some minor variants (subtype 1, 2a, 2b, and 2c).

A type-shift phenomenon occurs in Japan every 8 to 10 years. A major subtype of p1 was subtype 2 in 1995 to 2001. Thereafter, subtype 1 reached a level of 90% in 2005, whereas subtype 2 decreased from 2001 to 2005. Recently, it was reported that a type shift from subtype 1 to subtype 2 occurred in 2013 to 2015 in Yamagata Prefecture, Japan (14). It was presumed that because this subtype had few opportunities to be exposed to macrolides since 2000, isolates of subtype 2 may have been more sensitive to macrolides than isolates of subtype 1. Furthermore, correlations of P1 with multilocus variable-number tandem-repeat analysis (MLVA), which is one of the methods for typing, have been described (15, 16). As revealed by a previous MLVA-4 analysis, almost all isolates of 4/5/7/2 or 4/5/7/3 strains belonged to subtype 1 of p1, whereas almost all of the 3/5/6/2 or 3/6/6/2 strains belonged to subtype 2 of p1. We did not perform MLVA, and we hope to address this aspect in the future.

Next, we discuss the reason that the MIC values of tetracyclines against *M. pneum-
were restored significantly in 2015–2016 compared with 2011–2012. Okubo et al. (17) investigated the trends of use in practice patterns on pediatric *M. pneumoniae*-related respiratory infections. They reported that the usage of tetracyclines against pediatric *M. pneumoniae*-related respiratory infections decreased after the

**FIG 1** MIC distribution of antimicrobial agents for *M. pneumoniae* isolates in two recent epidemics in 2011–2012 and 2015–2016. TFLX, tosufloxacin; LVFX, levofloxacin; TC, tetracycline; MINO, minocycline; EM, erythromycin; CAM, clarithromycin; AZM, azithromycin.

*moniae* were restored significantly in 2015–2016 compared with 2011–2012. Okubo et al. (17) investigated the trends of use in practice patterns on pediatric *M. pneumoniae*-related respiratory infections. They reported that the usage of tetracyclines against pediatric *M. pneumoniae*-related respiratory infections decreased after the
pandemic of *M. pneumoniae* infections in 2011–2012. Although they did not investigate the cases in 2015–2016, we suggest that the use of tetracycline in 2015–2016 might not have increased as much as in 2011–2015 because of the recommendation of quinolones against pediatric *M. pneumoniae* infections. In other words, because quinolones were not recommended in 2011–2012, some cases of children (<8 years old) suspected to have MR *M. pneumoniae* infections were prescribed tetracyclines. If quinolones were prescribed instead of tetracyclines in these cases in 2015–2016, the chances of prescribing tetracyclines may have decreased.

In summary, quinolones and tetracyclines exhibited potent antimicrobial activities against MS and MR *M. pneumoniae* infection in 2011–2012 and 2015–2016, when *M. pneumoniae* epidemics occurred. The antimicrobial activities of macrolides and tetracyclines were restored significantly in 2015–2016 compared with 2011–2012.

The antimicrobial susceptibility of *M. pneumoniae* isolates should continue to be surveyed in Japan and other countries.

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