Potentiometric titration for the high precision determination of active components in six types of chemical disinfectants

Jiansheng Liang*, Junsheng Zhu, Lin Gong, Xiaoli Liu, Bin Wang

Department of Disinfection and Pest control, Wuhan Centers for Disease Prevention and Control, Hubei, China

* wh-ljs@sohu.com

Abstract

Chemical disinfectants effectively kill pathogenic microorganisms, eliminating routes of transmission for infectious diseases. Accurate quantification of the active ingredients can help make a more effective use of disinfectants. In this study, the active ingredients in six different types of chemical disinfectants were systematically quantified with great precision and accuracy using potentiometric titration. The coefficient of variations fell in the range of 0.04%-0.46%. The recovery rates of samples were all above 95% and the extended uncertainty was 0.32g/L. This method can be broadly applied to the analysis of disinfectants in the future.

Introduction

Chemical disinfectants effectively kill or remove pathogenic microorganisms, blocking disease transmission routes[1]. Commonly used chemical disinfectants include chlorine compounds, iodine, oxidants, aldehydes, alcohols, and quaternary ammonium salts[2]. The efficacy of these chemical disinfectants to eliminate microbes can be affected by many factors[3], of which the content of the active component of the disinfectant is clearly the most critical one. The most commonly used techniques for determining the content of chemicals include volumetric analysis, spectrophotometry, gas chromatography, and high-performance liquid chromatography [4–7]. Nevertheless, using any of the above methods for systematic measurement of the components in a chemical disinfectant can be problematic due to certain limitations of the techniques, such as large error range, difficulty of operation, susceptibility to interference, and the different properties of the various chemical components contained within each disinfectant. In this article, potentiometric titration[8] is used to determine the effective content of general chemical disinfectants from six different categories. The method offers many advantages, including simplicity, speed, easier end-point determination and higher accuracy. The testing and results of potentiometric titration are discussed in the following sections.

Materials and methods

Instruments and reagents

A. Instruments. Titrations were performed using an 809 Titrando automatic potentiometric titrator, a burette drive and a magnetic stirrer.
B. Test samples. The test samples included chlorine compounds (Dezhou Gelijie Disinfection Products Limited Company, lot number:20150722, available chlorine: 4.5%-5.5%), iodine (Wuhan Lianhua Sanitary Supplies Factory, lot number:20161004, available iodine: 1.0%-1.2%), hydrogen peroxide (Wuhan Lianhua Sanitary Supplies Factory, lot number:20160902, available hydrogen peroxide: 2.8%-3.5%), glutaraldehyde (Wuhan Donghuxing Technology Limited Company, lot number:20151103, available glutaraldehyde: 2.0%-2.2%), chlorhexidine (Dezhou Gelijie Disinfection Products Limited Company, lot number:20161115, available chlorhexidine acetate: 94%-96%) and benzalkonium bromide (Wuhan Donghuxing Technology Limited Company, lot number:20160727, available benzalkonium bromide: 0.9%-1.1%). These samples were all used prior to the expiration date.

C. Reagents. The following aqueous solutions were prepared: sulfuric acid (lot number: T20140412, 2mol/L,), potassium iodide (lot number:F20150311, 100g/L), manganese sulfate (lot number:F20150616, 100g/L), triethanolamine (lot number:F20130616, 65g/L), hydrochloric acid (lot number:20140208, 10g/L), sodium hydroxide (lot number:20120919, 43g/L) and acetic acid (lot number:T20130207, 36%). Glacial acetic acid (lot number:20130202, ≥ 99.5%) and acetone (lot number:T20141103, ≥ 99.5%) were also used. They were all purchased from Sinopharm Chemical Reagent Limited Company. A neutral hydroxylamine hydrochloride solution was prepared by adding 17.5g of hydroxylamine hydrochloride to 75mL of distilled water and diluting to 500mL with isopropyl alcohol. Next, 15mL of a bromophenol blue/ethanol solution (0.4g/L) was added, and finally triethanolamine (65g/L) was added to the solution until a blue-green color was obtained.

Electrode measurements
Composite platinum (Pt) electrodes or a monomer platinum (Pt) indicator electrode and reference electrode were used to measure available chlorine, available iodine and hydrogen peroxide content. A composite water phase pH electrode (containing a pH indicator electrode and a Ag/AgCl reference electrode) or an aqueous phase pH-indicating electrode and reference electrode monomer were used to measure glutaraldehyde content. A composite non-aqueous phase pH electrode (containing a pH indicator electrode and a Ag/AgCl reference electrode) was used to measure the chlorhexidine content. Finally, a surfactant aqueous phase titration indicator electrode and a reference electrode were used to measure benzalkonium bromide content.

Test method
A. Ambient temperature and humidity. The ambient temperature was 20˚C-25˚C, and the relative humidity was 45%-85% for these studies.

B. Principle. The indicator electrode and the reference electrode (or the reference and an indicator electrode included as a composite electrode) were immersed in the same solution, in which the reference electrode was maintained at a constant; then, the indicator electrode was immersed in the test substance. When the titration approached the equivalence point, small changes in the activity of the test substance solution elicited a dramatic change to the indicator electrode, and the largest change detected in indicator electrode potential was considered the end point of titration.

C. Titration mode, control program design, and installation. The Tiamo operating procedure software was installed before installing and debugging the automatic potentiometric titrator. Prior to using the instrument, titration mode was selected, and the standard detection method for test items and parameters were entered into the control program. The test project and the indicators for samples were transferred to the method bar and bar usage before conducting the titration, allowing the database to generate results automatically.
Method for determination

A. Capacity Analysis. This method was performed in accordance with the Ministry of Health of the People’s Republic of China’s "Disinfection Technical Specifications" (2002) [9].

B. Automatic potentiometric titration (dx.doi.org/10.17504/protocols.io.rkqd4vw). This method was performed as described in GB/T 9725–2007 “Chemical Reagent Potentiometric Titration General Principles” (ISO6353-1:1982) [10] and in accordance with DB/T 801–2012, "Determination of the Active Ingredient in the Chemical Disinfectant Automatic Potentiometric Titration" [11].

Method details: (1) The electrode and the titration mode were chosen according to the type of sample. (2) Sample pretreatment: 10 times the amount of solid (powder, tablet) chemical disinfectant required for analysis was obtained, and the appropriate amount of sample for accurate determination after grinding was weighed. The liquid chemical disinfectant was shaken until reaching a uniform state and then analyzed with or without dilution. (3) The sample information, standard titrant concentration and formula were entered as the input into the device, and the content of the effective component of the sample was measured. (4) The database then generated the results automatically.

C. Determination of the titration equivalence point. A peak maximum appears during the titration procedure when the threshold value is exceeded, and the jump point, or the titration equivalence point, is identified by taking the first derivative of the curve. The titration equivalence point corresponding to the volume of titrant recorded (a titration curve example is shown in Fig 1) is transferred to the formula for further calculations.

D. Calculation of results. The content of the test substance was determined using the chemical reaction formula and the amount of titrant consumed according to formulas (1) and (2). The result was averaged across six experiments.

Fig 1. Potentiometric titration curve. The solid curve represents the change in electric potential and its first derivative is plotted as the dashed curve.

https://doi.org/10.1371/journal.pone.0203558.g001
Liquid chemical disinfectant effective content

$$X(\text{g/L}) = \frac{c \times V_{st} \times k}{V} \times 1000$$  \hspace{1cm} (1)$$

Solid chemical disinfectant effective content

$$X(\%) = \frac{c \times V_{st} \times k}{m} \times 100\%$$  \hspace{1cm} (2)$$

For formulas (1) and (2): $X =$ substance content (g/L or %), $C =$ standard titration solution concentration (mol/L), $V_{st} =$ corrected volume for the standard titration (mL), $k =$ coefficient (g), $V =$ test substance sampling volume (mL), and $m =$ test substance sampling mass (g).

**Evaluation and methods**

**A. Precision assessment.** The measurements were compared with those obtained by the manual chemical titration method (i.e., the volumetric analysis) The precision of the method was assessed by the coefficient of variation (CV) according to formulas (3) and (4), which should not be greater than 1.0%.

$$S = \sqrt{\frac{\sum (x_i - x_t)^2}{n - 1}} \quad (1 \leq i \leq n)$$  \hspace{1cm} (3)$$

$$CV = \frac{S}{x_t} \times 100\%$$  \hspace{1cm} (4)$$

_in which_ $S =$ standard deviation, $X_i =$ single measured value, $X_t =$ average of measurements, $\Sigma =$ the sum of absolute values, $CV =$ coefficient of variation, and $n =$ number of measurements.

**B. Linearity test.** For the six types of chemical disinfectants that were analyzed, available chlorine (3.35–42.97 g/L), available iodine (2.29–11.43 g/L), hydrogen peroxide (10.48–34.94 g/L), glutaraldehyde (4.57–19.99 g/L), benzalkonium bromide (0.83–8.50 g/L), and chlorhexidine acetate (7.74%–93.41%) were determined by titration with sodium thiosulfate (0.1 mol/L), potassium permanganate (0.02 mol/L), sulfuric acid (0.25 mol/L), perchloric acid (0.1 mol/L), and sodium tetraphenylborate (0.02 mol/L), respectively. The measured chemical disinfectant composition was plotted as the horizontal axis, and the consumption of titration liquid volume was plotted as the vertical axis. A linear regression analysis was performed to evaluate the reproducibility of the method. The linear regression equation: $y = a + bx$ ($y$: titrant consumption, $a$: constant, $b$: slope, $x$: content or sample volume).

**C. Accuracy assessment.** The accuracy of the method was measured by the recovery of standard substance added ($P$) as calculated by Eq (5), which should be $>95\%$.

$$P = \frac{c_2 - c_1}{c_3} \times 100\%$$  \hspace{1cm} (5)$$

_in which_ $P =$ spiked recoveries, $c_1 =$ sample background concentration, $c_2 =$ sample spiked concentration, and $c_3 =$ plus scalar.

**D. Uncertainty assessment.** The main sources of uncertainty of this analysis included sample volume ($\mu_1$), standard solution preparation ($\mu_2$), repeated measurements ($\mu_3$) and interpolation using the standard curve ($\mu_4$). Glutaraldehyde disinfectant was selected as a substance to estimate the upper bound of the uncertainty of this method because of the complexity of its composition.
Mathematical model
\[ X(g/L) = \frac{c \times V_{st} \times 0.1001}{V} \times 1000 \]  \hspace{1cm} (6)

in which \( x \) = Glutaraldehyde content(g/L), \( c \) = concentration of sulfuric acid titrant(mol/L), \( V_{st} \) = corrected volume of sulfuric acid titrant(ml), and 0.1001 is the conversion factor.

Statistical analysis
The data from the experiments were analyzed with the SAS9.0 statistics software. Results from two groups were compared using independent sample T-test and correlation analysis. The difference was considered to be statistically significant with a \( P \leq 0.05 \).

Results
Accuracy and precision of potentiometric titration
The results of potentiometric titration and direct titration were shown in Table 1. There were no significant differences between the two methods (\( P > 0.05 \)). This indicated that the analysis of chemical disinfectants by potentiometric titration agrees with the current National Standard (direct titration). The CVs for the effective content in chemical disinfectants determined by potentiometric titration were lower than those obtained using the other method, except for the bromogeramine disinfectant, suggesting that potentiometric titration has a superior precision direct titration.

Linearity potentiometric titration tests
Table 2 lists the linearity test results for the effective contents of six different chemical disinfectants measured by potentiometric titration. The results for each disinfectant and its neutralizer are shown in Figs 2–7 along with the linear regression models. These equations were as follows: for available chlorine, \( y = 0.2723x + 0.039 \) and the \( R^2 \) (determinate coefficient) = 1.000; for available iodine, \( y = 1.9033x - 0.0127 \), \( R^2 = 1.000 \); for hydrogen peroxide, \( y = 0.5851x - 0.004 \), \( R^2 = 1.000 \); for glutaraldehyde, \( y = 0.3993x + 0.0046 \), \( R^2 = 1.000 \); for chlorhexidine

Table 1. Results and CVs of the effective content of six different chemical disinfectants by potentiometric titration and direct titration.

| Experimental projects (unit) | Experimental methods* | Each experimental result | Mean | Standard deviation | CV(%) | P value |
|-----------------------------|-----------------------|-------------------------|------|-------------------|-------|---------|
|                             | A                     | 1 2 3 4 5 6             |      |                   |       |         |
| Available chlorine concentration (g/L) | A | 43.34 43.44 43.36 43.35 43.38 43.41 | 43.38 | 0.035 | 0.081 | 0.1013  |
|                             | B                     | 43.16 43.18 43.13 43.17 43.16 43.17 | 43.17 | 0.019 | 0.044 |         |
| Available iodine concentration (g/L) | A | 11.41 11.43 11.41 11.43 11.43 11.43 | 11.42 | 0.010 | 0.083 | 0.4956  |
|                             | B                     | 11.42 11.43 11.43 11.42 11.43 11.43 | 11.43 | 0.005 | 0.043 |         |
| Hydrogen peroxide concentration (g/L) | A | 35.04 34.92 34.95 34.96 34.96 34.96 | 34.97 | 0.036 | 0.104 | 0.1116  |
|                             | B                     | 34.90 34.93 34.94 34.94 34.93 34.94 | 34.94 | 0.018 | 0.052 |         |
| Glutaraldehyde concentration (g/L) | A | 19.84 19.71 19.89 19.71 19.84 19.75 | 19.79 | 0.070 | 0.354 | 0.1274  |
|                             | B                     | 20.02 20.04 20.01 20.03 20.01 20.03 | 20.03 | 0.013 | 0.064 |         |
| Chlorhexidine acetate concentration (%) | A | 94.47 95.05 94.62 95.03 94.67 94.57 | 94.74 | 0.224 | 0.236 | 0.6414  |
|                             | B                     | 94.87 94.96 94.84 94.56 94.87 94.65 | 94.79 | 0.139 | 0.147 |         |
| Benzalkonium bromide concentration (g/L) | A | 8.43 8.40 8.49 8.43 8.46 8.49 | 8.45 | 0.033 | 0.392 | 0.0795  |
|                             | B                     | 8.48 8.54 8.51 8.54 8.43 8.47 | 8.50 | 0.040 | 0.465 |         |

* A = direct titration, B = potentiometric titration.
acetate, \( y = 0.0479x + 0.1192 \), \( R^2 = 0.9998 \), and for benzalkonium bromide, \( y = 3.1069x + 0.0734 \), \( R^2 = 0.9999 \). The above coefficients were all greater than 0.999, suggesting a strong linearity of the potentiometric titration method in analyzing the active ingredients of all these disinfectants.

### Recovery tests of standard substance (sample) addition

The accuracy of potentiometric titration was assessed by the recovery rates of standard substance (sample). If the recovery was between 90% and 110%, the method was considered reliable. The data in Table 3 show that the recovery rates for all of the disinfectants were in the range of 95%-104%, demonstrating the high reliability of this method.

![Fig 2. Linear regression diagram for available chlorine and its neutralizer.](https://doi.org/10.1371/journal.pone.0203558.g002)
Uncertainty measurements for glutaraldehyde disinfectant

A. Relative uncertainty caused by sampling volume ($\mu_1$). By sampling 10L with a 10mL single scale line straw, the relative uncertainty ($\mu_1$) caused by the sampling volume ($v_1$) was the result of a reading value variation from the scale line($\mu_{1,1}$) and the volume variation of the scale due to a temperature change($\mu_{1,2}$).

1. The allowable error in a 10mL single scale line straw was 0.1mL, with a uniform distribution of $\kappa = \sqrt{3}$, then:

$$\mu_{1,1} = \frac{\mu_{1,1}}{\kappa} = \frac{0.1}{\sqrt{3}} = 0.577\%$$
2. Set temperature variation was 3°C ($\Delta T = 3$, expansion coefficient of water $\alpha = 2 \times 10^{-4}$/°C, with a uniform distribution of $\kappa = \sqrt{3}$, then:

$$\mu_{1,2} = \frac{\Delta T \times \alpha}{\kappa} = \frac{3 \times 2 \times 10^{-4}}{\sqrt{3}} = 0.0346\%$$
3. Relative uncertainty ($\mu_1$) caused by sampling volume ($v_1$)

$$\mu_1 = \sqrt{\mu_{1,1}^2 + \mu_{1,2}^2} = \sqrt{0.577^2 + 0.0346^2} = 0.578\%$$

B. Relative uncertainty caused by preparation of the standard titration solution ($\mu_2$).

1. The allowable error in a 10mL single scale line straw was 0.1mL, with a uniform distribution of $\kappa = \sqrt{3}$, then:

$$\mu_{2,1} = \frac{v_{1,1}}{V_1} = \frac{0.1}{10} = 0.010\%$$

2. Uncertainty due to the burette ($\mu_{2,2}$)

The value of a scale division in a 50mL acid burette was 0.1 mL, and the maximum permissible error of which was ±0.05mL, with a uniform distribution of $\kappa = \sqrt{3}$ and a half width $\alpha = 0.05$ mL, then:

$$\mu_{2,2} = \frac{2}{V_2} = \frac{0.05}{50} = 0.001\%$$

3. The concentration uncertainty of calibrated standard titrant solution by means of repetitive measurements ($\mu_{2,3}$).
The same standard solution was titrated 6 times, and the results are shown in Table 4.

\[
\bar{x} = \frac{1}{n} \sum_{i=1}^{6} x_i = 0.2530
\]

\[
S(x) = \sqrt{\frac{\sum_{i=1}^{6} (x_i - \bar{x})^2}{n-1}} = 0.0000392 = 0.00392\%
\]

\[
\mu_{2,3} = \frac{S(x)}{\sqrt{n}} = \frac{0.0000392}{2.4494897} = 0.0016\%
\]

Table 3. The recovery test results of standard substance (sample) addition in six types of chemical disinfectants.

| Disinfectants | Grouping (g/L) | Measurement results (Active ingredient content of each preparation solution (g/L)) | Mean (g/L) | Recoveries(%) |
|---------------|----------------|-----------------------------------------------------------------------------|------------|---------------|
|               | 1   | 2   | 3   | 4   | 5   | 6   |               |              |               |
| Chlorine      | Background | 9.34 | 9.33 | 9.34 | 9.36 | 9.31 | 9.34 | 9.34 | /              |
|               | 4.30 | 13.62 | 13.59 | 13.63 | 13.62 | 13.61 | 13.58 | 13.61 | 99.30          |
|               | 8.60 | 17.91 | 17.94 | 17.89 | 17.92 | 17.91 | 17.91 | 17.91 | 99.65          |
|               | 12.89 | 22.25 | 22.20 | 22.20 | 22.22 | 22.20 | 22.19 | 22.21 | 99.84          |
| Iodine        | Background | 1.16 | 1.15 | 1.16 | 1.17 | 1.14 | 1.17 | 1.16 | /              |
|               | 2.29 | 3.53 | 3.52 | 3.51 | 3.52 | 3.50 | 3.51 | 3.52 | 103.0          |
|               | 4.58 | 5.90 | 5.93 | 5.89 | 5.91 | 5.89 | 5.88 | 5.90 | 103.5          |
|               | 9.16 | 10.35 | 10.34 | 10.32 | 10.33 | 10.34 | 10.31 | 10.33 | 100.1          |
| Hydrogen peroxide | Background | 11.45 | 11.44 | 11.44 | 11.46 | 11.44 | 11.44 | 11.45 | /              |
|               | 3.49 | 14.95 | 14.96 | 14.96 | 14.95 | 14.96 | 14.96 | 14.96 | 100.6          |
|               | 6.99 | 18.42 | 18.41 | 18.41 | 18.42 | 18.43 | 18.41 | 18.42 | 99.71          |
|               | 10.48 | 21.90 | 21.91 | 21.94 | 21.94 | 21.89 | 21.90 | 21.91 | 99.81          |
| Glutaraldehyde | Background | 9.88 | 9.85 | 9.81 | 9.79 | 9.83 | 9.87 | 9.84 | /              |
|               | 2.02 | 11.87 | 11.81 | 11.79 | 11.85 | 11.81 | 11.83 | 11.83 | 98.51          |
|               | 4.03 | 13.85 | 13.96 | 13.91 | 13.87 | 13.95 | 13.89 | 13.91 | 101.0%         |
|               | 10.08 | 19.97 | 19.85 | 19.85 | 19.95 | 19.92 | 19.88 | 19.90 | 99.80%         |
| Chlorhexidine  | Background | 0.0968 | 0.0960 | 0.0972 | 0.0960 | 0.0972 | 0.0960 | 0.0965 | /              |
|               | 0.0186 | 0.1145 | 0.1144 | 0.1147 | 0.1146 | 0.1145 | 0.1147 | 0.1146 | 97.31          |
|               | 0.0284 | 0.1234 | 0.1237 | 0.1234 | 0.1238 | 0.1234 | 0.1231 | 0.1235 | 95.07          |
|               | 0.0379 | 0.1336 | 0.1339 | 0.1341 | 0.1336 | 0.1339 | 0.1342 | 0.1339 | 98.68          |
| Benzalkonium bromide | Background | 0.85 | 0.84 | 0.85 | 0.84 | 0.84 | 0.84 | 0.84 | /              |
|               | 1.7 | 2.51 | 2.49 | 2.52 | 2.48 | 2.49 | 2.49 | 2.49 | 97.06          |
|               | 3.4 | 4.22 | 4.26 | 4.23 | 4.22 | 4.21 | 4.23 | 4.23 | 99.71          |
|               | 6.8 | 7.66 | 7.68 | 7.62 | 7.65 | 7.68 | 7.63 | 7.65 | 100.1          |

Table 4. Calibration results for titration of standard sulfuric acid solution.

| Calibration number | x_i (mol/L) | \(\bar{x}\) (mol/L) | S(x) (%) | \(\mu_{2,3}\) (%) | n-1 |
|--------------------|------------|--------------------|--------|----------------|-----|
| 1                  | 0.2521     | 0.2530             | 0.00392| 0.0016        | 5   |
| 2                  | 0.2537     |                    |        |               |     |
| 3                  | 0.2536     |                    |        |               |     |
| 4                  | 0.2525     |                    |        |               |     |
| 5                  | 0.2529     |                    |        |               |     |
| 6                  | 0.2532     |                    |        |               |     |
4. Relative uncertainty caused by the preparation of standard titration solutions ($\mu_2$)

$$\mu_2 = \sqrt{\mu_{2,1}^2 + \mu_{2,2}^2 + \mu_{2,3}^2} = \sqrt{0.577\%^2 + 0.0577\%^2 + 0.0016\%^2} = 0.580\%$$

C. Relative uncertainty caused by the repeatability of the results ($\mu_3$). Samples were measured 6 times in parallel, and the results for glutaraldehyde content are listed in Table 1:

$$\mu_3 = \frac{S}{\sqrt{n \times \bar{x}}} = 0.0265\%$$

D. Relative uncertainty caused by the standard curve ($\mu_4$). From Fig 5, the glutaraldehyde content curve was analyzed by linear regression, and the linear regression equation was:

$$y = 0.3993x + 0.0046, R = 1.000.$$  

1. The standard deviation of the regression line ($\mu_{4,1}$)

$$\mu_{4,1} = \sqrt{\frac{\sum_{i=1}^{m} \sum_{j=1}^{n} \left[Y_{ij} - (a + bc_i)\right]^2}{m \times n - 2}} = \sqrt{\frac{3.16754 \times 10^{-5}}{1 \times 5 - 2}} = 3.249 \times 10^{-3}$$

in which $n$ = the concentration point ($n = 5$), $m$ = the repeat measurement frequency for each concentration point ($m = 1$)

2. The glutaraldehyde concentration sum of squares (Ss)

$$SS = \sum (c_i - \bar{c})^2 = 145.591$$

3. Relative uncertainty caused by the standard curve ($\mu_4$)

$$\mu_4 = \frac{\mu_{4,1}}{C_X \times b} \sqrt{\frac{1}{m} + \frac{1}{n} + \frac{(c_i - \bar{c})^2}{SS}}$$

$$= \frac{3.249 \times 10^{-3}}{20.03 \times 0.3993} \times \sqrt{\frac{1}{1} + \frac{1}{5} + \frac{(20.03 - 13.242)^2}{145.591}}$$

$$= 0.05\%$$

in which concentration of sample $C_X = 20.03\text{g/L}$

E. Synthetic relative uncertainty ($\mu_w$). Each relative uncertainty was independent; therefore, the synthetic relative uncertainty could be calculated as follows:

$$\mu_w = \sqrt{\mu_1^2 + \mu_2^2 + \mu_3^2 + \mu_4^2}$$

$$= \sqrt{0.578\%^2 + 0.580\%^2 + 0.0265\%^2 + 0.05\%^2}$$

$$= 0.8208\%$$
The synthetic standard uncertainty $\mu(w)$:

$$\mu(w) = \bar{x} \times \mu_\sigma = 20.03 \times 0.008208 = 0.16g/L$$

F. Expanded uncertainty (U) and the result of sample concentration (C). Setting the coverage factor $k = 2$, the extended uncertainty:

$$U = k \times \mu(w) = 2 \times 0.16g/L = 0.32g/L$$

The result is expressed as follows:

$$CC = \bar{x} \pm UU = (20.03 \pm 0.32)g/L$$

G. By analyzing and calculating the uncertainties for the glutaraldehyde disinfectant, all of the partial uncertainties were very low (<0.60%), and the value for the expanded uncertainty was 0.32g/L. This suggests that potentiometric titration measurements are of high quality.

**Discussion**

In classical chemical titration (volumetric analysis), the endpoint is based on the color change of an indicator. If the sample being tested is colored or turbid, it is difficult to estimate the titration endpoint with a reliable indicator. In contrast, potentiometric titration relies on abrupt changes in electrode potential to determine the endpoint. The ion concentration being evaluated often varies by orders of magnitude and generates abrupt changes in the electrode potential when the endpoint is approached. The content of the samples can then be calculated from titrant consumption.

In this article, potentiometric titration was adopted to quantify the effective contents of 6 different chemical disinfectants. Compared to standard chemical titration, this method offers many advantages, including simplicity, speed, ease of end-point determination, accuracy, stability, and the ability to overcome a number of confounding factors. Therefore, potentiometric titration is suitable for the fast and accurate determination of 1) effective chlorine/effective iodine/hydrogen peroxide content using the selective composite platinum (Pt) electrode or monomer platinum (Pt) indicator electrode combined with reference electrode, 2) glutaraldehyde content using the composite aqueous phase acid-base electrode (containing pH indicator electrode and Ag/AgCl Reference electrode) or aqueous phase acid-base monomer indicator electrode combined with reference electrode, 3) chlorhexidine acetate using the composite non-aqueous phase acid-base electrode (containing pH indicator electrode and Ag/AgCl Reference electrode), and 4) benzalkonium bromide using the surfactant aqueous phase titration indicator electrode and the reference electrode. This method is particularly adaptable to determining the content of certain bactericidal active ingredients in compound chemical disinfectants.

This method has robust precision for all the disinfectants tested, with the CVs ranging from 0.04% to 0.46%. Furthermore, our tests also show that the potentiometric titration method yields results that agree well with the current National Standard (direct titration) while having outstanding accuracy as indicated by high the recovery rates (greater than 95% for all samples tested).

The results of the linearity tests shows that potentiometric titration maintains an excellent linearity for a wide concentration range of the analyte. Moreover, the results also demonstrate good robustness. The value for expanded uncertainty was only 0.32g/L. This work represents the first analysis of various chemical disinfectants for systematic determination of the effective
contents using potentiometric titration. The robustness and simplicity of this method will prepare it for a broad application in the analysis of disinfectants in the future.

Acknowledgments

We would like to express our gratitude and appreciation to Huangshi CDC and Yichang CDC for their assistance in the testing analysis and to Metrohm (China) Limited Company for providing the samples.

Author Contributions

Conceptualization: Jiansheng Liang, Xiaoli Liu.
Data curation: Junsheng Zhu, Lin Gong, Bin Wang.
Formal analysis: Lin Gong, Xiaoli Liu.
Resources: Lin Gong.
Software: Xiaoli Liu.
Supervision: Xiaoli Liu.
Writing – original draft: Jiansheng Liang.
Writing – review & editing: Jiansheng Liang.

References

1. Banach JL, Sampers I, Van Haute S, van der Fels-Klerx HJ. Effect of disinfectants on preventing the cross-contamination of pathogens in fresh produce washing water. Int J Environ Res Public Health. 2015; 12: 8658–8677. https://doi.org/10.3390/ijerph120808658 PMID: 26213953
2. Fleurette J. Methods of in vitro study and classification of antiseptics and disinfectants. Rev Prat. 1980; 30: 2167–2168, 2173–2166, 2181–2162. PMID: 7444262
3. Chen X, Liang T, Wen K, Lan ZX. Research progress in influencing factors of effect of iodine disinfectants. Med Recapitulate. 2015; 21: 1061–1063.
4. Chen G, Gao Q, Li S, Song J, Yi L, Zhu YK. Uncertainty assessment of effective iodine content measurement in volumetric analysis. Chinese Journal of Disinfection. 2014; 31: 349–352.
5. Gao J, Chen J, Tie J, Wang CD. Determination of polyhexamethylene biguanidine hydrochloride using ultraviolet spectrophotometry. Chinese Journal of Disinfection. 2013; 30: 608–610.
6. Yun M, Fang Q. Observation on the accuracy of gas chromatography method to determine glutaraldehyde content. Chinese Journal of Disinfection. 2010; 27: 687–689.
7. Zheng R, Zheng B, Yang Y, Hua YY, Lin LW, Huang HN. Study on accuracy of polyhexamethylene biguanide in compound disinfectant by high performance liquid chromatography. Chinese Journal of Disinfection. 2015; 32: 10–12.
8. Granholm K, Sokalski T, Lewenstam A, Ivaska A. Ion-selective electrodes in potentiometric titrations; a new method for processing and evaluating titration data. Anal Chim Acta. 2015; 888: 36–43. https://doi.org/10.1016/j.aca.2015.05.056 PMID: 26320956
9. National Health and Family Planning Commission of the People's Republic of China, Department of Oversight Division. Disinfection technical specifications 2002. Available from: http://www.nhfpc.gov.cn/zhjy/s9139/200804/86d017920ad84e64a90806717719624f.shtml
10. The State Administration of Quality Supervision, Inspection and Quarantine of the People’s Republic of China, Standardization Administration of China. General Principles of Potentiometric Titration in Chemical Reagents. 2008; ISO 6353–1:1982. Available from: http://www.tsinfo.js.cn/inquiry/gbtdetails.aspxA100.
11. Hubei Bureau of Quality and Technical Supervision. Determination of the Active Ingredient in the Chemical Disinfectant, Automatic Potentiometric Titration. 2012; DB/T 801–2012. Available from: http://www.hbdlsw.com/SearchInfo/SeniorList.aspx/searchType.