Dehydroepiandrosterone improves follicular fluid bone morphogenetic protein-15 and accumulated embryo score of infertility patients with diminished ovarian reserve undergoing in vitro fertilization: a randomized controlled trial

Huan H Zhang1, Ping Y Xu1, Juan Wu1, Wei W Zou2, Xia M Xu2, Xia Y Cao1 and Lian Z Wei1*

Abstract

Objective: To evaluate the effect of dehydroepiandrosterone (DHEA) on infertility patients with diminished ovarian reserve undergoing in vitro fertilization.

Methods: This is a prospective study. Ninety-five patients with diminished ovarian reserve were included in this study. Of them, 42 patients were randomly allocated to the DHEA group, who received DHEA 75 mg daily for three consecutive menstrual cycles prior to IVF cycles, and 53 patients were allocated to the control group, who entered IVF cycles directly. All patients were treated with the same ovarian stimulation protocol. Follicular fluid samples from both groups were collected for bone morphogenetic protein-15 (BMP-15) and growth differentiation factor-9 (GDF-9). Fluid from the first aspirated follicle without any visible blood contamination was carefully collected. In addition, day 3 Blood samples were collected pre- and post-treatment of DHEA for serum anti-Mullerian hormone (AMH), follicle stimulating hormone (FSH) and estradiol (E2) in the DHEA group.

Results: The level of BMP-15 in follicular fluid samples from the DHEA group was significantly higher than that of the control samples (P=0.000). Patients after DHEA treatment demonstrated a significantly higher level of AMH and a significantly lower level of FSH, E2 compared to themselves prior to DHEA therapy (P=0.015; P=0.036; P=0.002; respectively). Moreover, the accumulated score of embryos was significantly higher in the DHEA group (P=0.033).

Conclusions: These observations confirm the beneficial effect of DHEA for infertility patients with diminished ovarian reserve.

Trial registration: ChiCTR-TRC-14005002

Keywords: Dehydroepiandrosterone, Diminished ovarian reserve, BMP-15, GDF-9, AMH
paired analysis of before and after IVF parameters of twenty-five patients with DOR. Subsequently, Barad et al. [5] got a conclusion that addition of DHEA gave rise to pregnancy rate meanwhile other reported a significantly reduction in Miscarriage rate [6].

However, the early reports on DHEA supplement to the patients with DOR were still controversial because there was lack of large-scale, well-designed confirmatory studies [4,7,8]. To further evaluate the effect of DHEA, ovarian reserve markers such as AMH, inhibin-B and antral follicle count were detected and significant changes were observed by N. Yilmaz et al. [9]. However, a small-sized randomized double-blinded placebo-controlled trial observed no significant changes in serum AMH and FSH levels [10].

Given this, studies should be continued to assess the utilization of DHEA in patients with DOR undergoing IVF. There are still no ideal markers predicting the quality of oocytes and embryos though there are numerous markers predicting response to ovarian induction. To our knowledge, follicular fluid contains many vital cytokines in reaction of oocyte quality. BMP-15, GDF-9, members of the transforming growth factor-β super family, are known to be of great importance for follicle growth and female fertility [11,12]. GDF-9 deficient female mice were blocked at primary one-layer follicle stage during follicular development which eventually leads to complete infertility [13]. High BMP-15 level in follicular fluid has been demonstrated to associate with high quality oocyte, high cleavage rate and good morphology embryos [14].

Therefore, to gain further insights on the effect of DHEA, here in our study, BMP-15, GDF-9 in follicular fluid were studied to obtain more evidence. The specific aims of the present study were: 1) To explore the levels of BMP-15 and GDF-9 in follicular fluid and compare them between the DHEA group and the control; 2) To confirm whether there are significant changes in ovarian reserve markers such as AMH, FSH, E2 after DHEA supplementation in the study group; 3) To compare the outcome of IVF cycles between the two groups.

Materials and methods

Patients

The study recruited patients with primary or secondary infertility for diminished ovarian reserve at the Assisted Reproductive Center, The Affiliated Hospital of Anhui Medical University between March 2013 and May 2014. Despite there is no established definition of DOR, predictive criteria such as abnormal ovarian reserve tests (day 3 FSH or AMH level) and previous ovarian response number of oocytes retrieved after a standard-dose ovarian stimulation are two of the most frequent criteria adopted on the topic today [15]. In this study the criteria for defining DOR was defined as follow: 1) an elevated day 3 FSH level ≥ 10 mIU/mL or FSH/LH > 3; 2) the number of antral follicle count less than five; 3) a previous poor ovarian response: retrieval of fewer than five oocytes or cycle cancellation due to poor response to ovarian stimulation. A diagnosed with DOR was reached if they fulfill any of the above three. Patients were excluded if they had any of the following: 1) a history of ovarian cystectomy or oophorectomy; 2) a diagnosis of endometriosis; 3) a history of DHEA supplementation or hormonal replacement therapy.

Methods

This prospective study was approved by the Clinical Research Ethics Committee of The First Affiliated Hospital of Anhui Medical University. Before the trial, every participant was fully explained about the present study. A total of 105 patients fulfilled the selection criteria were recruited in our present study. According to a computer-generated randomization list by a nurse, 52 patients were randomly allocated to the DHEA group who received DHEA 25 mg three times per day for three consecutive menstrual cycles prior to IVF cycles and 53 patients to the control group who entered IVF cycles directly. On the first menstrual cycle and the fourth menstrual cycle, day 3 blood samples were collected for serum AMH, FSH, E2 in the DHEA group.

Everyone were given a dose of 225 IU human menopausal gonadotropin (HMG, Lizhu, China) combined with Clomiphene Citrate (CC, Lizhu, China) 100 mg daily from day 3 of the cycle to induce follicle growth. When at least one mature follicle of ≥ 18 mm in mean diameter was seen on ultrasound, a dose of 10,000 IU of Human chorionic gonadotropin (HCG, Lizhu, China) was administrated followed by oocyte retrieval 36 hour later.

Clean follicular fluid was aspirated from leading follicle by trans-vaginal ultrasound-guided puncture for BMP-15 and GDF-9 detection. Fluid samples from the first aspirated follicle and without any visible blood contamination were carefully collected. Samples were centrifuged at 2000 r/min for 15 minutes and stored at –80°C until to be tested.

Outcome measures

The primary outcome measures were follicular fluid BMP-15, GDF-9 and serum AMH, FSH, E2. Secondary outcome measures were the number of oocytes retrieved, MII oocytes, and embryos transferred; the accumulated score of embryos.

Sample detection

The concentration of BMP-15, GDF-9, AMH were determined by enzyme-linked immunosorbent assay (ELISA) [BMP-15: BMP-15 Gen II ELISA, USCN, USA; GDF-9: GDF-9 Gen II ELISA, USCN, USA; AMH: AMH Gen II ELISA, ASNL, USA]. FSH, E2 in serum
samples were determined by a radioimmunoassay kit (Beckman Coulter).

**Embryo grade**

On the third day of incubation, embryos were graded from one to four, based on percent fragmentation and cell counts: grade 4, equal-sized symmetrical cells with no fragmentation; grade 3, equal-sized symmetrical cells with 10% fragmentation; grade 2, non-symmetrical blastomeres with 10-50% fragmentation and grade 1, 50% fragmentation [16]. Up to three, best-quality embryos were transferred on day 3 or day 5. Accumulated embryo score was calculated by summing the score of each embryo produced by each patient on the third day of development. For example, a patient with a grade 4 embryo and two grade 3 embryos would be assigned a score of 10. Patients with cycle cancellation would be assigned a score of 0.

**Statistical analyses**

SPSS for Windows, Standard version 17.0 was utilized for data analysis. Quantitative variables were presented as mean ± standard deviation (SD) and analyzed by student’s t-test. Qualitative variables were presented as proportions and analyzed by χ2 test. For all analysis, the significant was set at P<.05.

**Results**

Between March 2013 and May 2014, a total of 147 patients with primary or secondary infertility for diminished ovarian reserve were screened at the Assisted Reproductive Center, The Affiliated Hospital of Anhui Medical University. Of them, 105 patients were recruited and randomized. Excluded 10 patients discontinued intervention; there were 42 patients in the DHEA group and 53 in the control. (See in Figure 1).

Patients enrolled in the DHEA group and the control group were aged 37.17 (±5.22) and 37.43 (±4.33) years respectively. As is shown in Table 1, all descriptive parameters were comparable between the DHEA group and the control group in terms of age, body mass index (BMI), duration of infertility, primary or secondary infertility (P=.785, P=.549, P=.756, P=.126 respectively). Moreover, serum AMH, FSH, E2, the total number of antral follicle were all matched in this two groups thus signifying they shared comparable ovarian reserve (P=.440; P=.421; P=.835; P=.764 respectively).

Characteristics of IVF cycles regarding with the number of oocytes, MII oocytes, embryos transferred, and the score of embryos were also analyzed between the DHEA group (n = 42) and the control group (n = 53). As is shown in Figure 2, the accumulated score of embryos was significantly higher in patients treated with DHEA compared to the control group (4.24 ± 3.39 vs. 2.87 ± 2.79, P=.033). However, no significantly changes were observed in the number of oocytes retrieved, MII oocytes and embryos transferred (P=.526, P=.289, P=.076 respectively).

After standardized ovarian stimulation, 26 follicular fluid samples were collected from the DHEA group, with 6 cycles canceled for poor responses and 10 follicular fluid samples unqualified for visible blood contamination. In the control group, we sampled 35 samples with 9 cycles canceled and 9 unqualified respectively.

Basic characters of the DHEA follicular fluid (FF) group and the control follicular fluid (FF) group are presented in Table 2. No significant differences were found in terms of serum AMH, FSH, E2, and the total number of antral follicle between the two groups (P=.496; P=.406; P=.653; P=.486 respectively). The mean levels of BMP-15 in the DHEA FF group (n = 26) and the control FF group (n = 35) were 0.81 (±0.25) ng/ml and 0.35 (±0.23) ng/ml respectively. And a statistically significant difference was found between the two FF groups (P=.000). No significant difference was detected in the level of GDF-9 in the DHEA FF group compared to the control (7.91 ± 3.77 vs. 6.82 ± 2.46, P=.203).

A comparison of ovarian reserve markers was made between before and after treatment in the 42 patients treated with DHEA. The mean duration of DHEA supplementation was 83.67 ± 6.45 days. The levels of DHEA-S and testosterone were significantly higher after treatment (1.24 ± 0.74 μg/mL vs. 5.50 ± 3.48 μg/mL, p=.000; 0.74 ± 0.41 nmol/L vs. 2.25 ± 1.28 nmol/L, p=.000). And there was a significant increase of AMH (1.01 ± 0.77 ng/ml vs. 1.29 ± 1.09 ng/ml, P=.015) and a significant decrease of FSH (11.68 ± 6.62 IU/L vs. 9.45 ± 5.09 IU/L, P=.036) and E2 (186.58 ± 142.19 pg/ml vs. 110.79 ± 68.28 pg/ml, P=.002) after about 12 weeks of DHEA supplementation. However no significant change was found in the count of antral follicle (2.95 ± 1.38 vs. 3.21 ± 1.22; P=.054). (See in Table 3).

In these 42 patients received DHEA treatment, eight patients conceived after the IVF cycles, with a pregnancy rate of 19.05%. In the control, seven patients conceived with a pregnancy rate of 13.21%. And there was no significant difference in pregnancy rate between them (P=.816).

During this trial, no major adverse effects were reported. Only one patient complained of dizziness and three patients complained of acne.

**Discussion**

This is the first report concerning the relationship between the DHEA supplementation for patients with DOR and the level of BMP-15 and GDF-9 in follicular fluid. The present study found a significantly higher level of BMP-15 in follicular fluid of patients treated with DHEA prior to IVF cycles than patients entered IVF cycles directly.
Human data demonstrated that high BMP-15 level in follicular fluid is associated with high quality oocyte and good morphology embryos [11]. Moreover, poor responders with a high level of BMP-15 in follicular fluid demonstrated higher implantation rate and pregnancy rate than those in the low BMP-15 group [17]. Here in our study, the higher level of BMP-15 observed in DHEA treated group predicted better ovarian reserve and outcome of IVF cycles.

Additionally, some other ovarian reserve markers such as serum AMH, FSH, E2 and the count of antral follicle were measured before and after the supplementation of DHEA in this article. There was a significant higher level of AMH, a significantly lower level of FSH and E2 after about 12 weeks treatment of DHEA. However, no significant change was observed in the count of antral follicle in our study. Research also noted that DHEA supplementation improved serum AMH, inhibin B and antral follicle count in patients with DOR. Serum level of AMH increases in parallel with length of DHEA supplementation [18]. Subsequently, other study observed a significant increase in AFC and ovarian volume but no significant differences were found in the levels of AMH and FSH [10].

Furthermore, main IVF cycle characteristics were compared between the two groups in our study. The accumulated score of embryos were significantly higher in patients treated with DHEA. Thus signified a better quality of embryos and strengthened the observations in ovarian reserve markers above. And there was a trend
for increased number of oocytes retrieved, MII oocytes and embryos transferred in the presence of DHEA, though this effect was not statistically significant.

In agreement with our study, Barad et al. [4] reported that DHEA treatment gave rise to a significant increase in oocyte, embryo numbers and better embryo grades through paired analysis of before and after IVF parameters of twenty five patients with DOR. In a late follow up, the first prospectively randomized clinical trial was published which suggested that addition of DHEA increased embryo quality and live birth rates in patients with DOR [16]. However, this paper has not accepted well as for its low number of 33 subjects with 17 in the DHEA group and 16 in the control. Subsequently in 2012, a study enrolled 133 DOR patients with 67 used DHEA and 63 without DHEA was reported. It demonstrated that peak E2, endometrial thickness, number of oocytes retrieved, number of embryos transferred in the study group were significantly higher than the control [19].

All of these observations raise a question, what may probably be the mechanism by which DHEA exert its effect on ovarian reserve? Recent data mostly focus on androgens receptor (AR), hold the opinion that androgens primarily affect granulose cells (GC) with ligand-activated AR [20]. AR-knockout (ARKO) female mice model revealed low

| Parameters                  | The DHEA group (n = 42) | The control group (n = 53) | P-value |
|-----------------------------|-------------------------|---------------------------|---------|
| Age (years)                 | 37.17 ± 5.22            | 37.43 ± 4.33              | .785    |
| BMI (kg/m²)                 | 22.13 ± 2.92            | 22.46 ± 2.42              | .549    |
| Duration of infertility(years) | 6.64 ± 4.54          | 6.34 ± 4.85               | .756    |
| Primary infertility (%)     | 42.86 %                 | 26.42%                    | .126    |
| Secondary infertility (%)   | 57.14 %                 | 73.58%                    |         |
| AMH (ng/ml)                 | 1.01 ± 0.77             | 1.12 ± 0.84               | .440    |
| FSH(mIU/mL)                 | 11.68 ± 6.62            | 10.81 ± 2.41              | .421    |
| Estradiol (pg/ml)           | 186.58 ± 142.19         | 193.06 ± 156.16           | .835    |
| Antral follicle count        | 2.95 ± 1.38             | 3.04 ± 1.38               | .764    |

Data are presented as mean ± SD (standard deviation) or percentage as appropriate. BMI: body mass index. P<.05 was considered as statistically significant.

![Figure 2](http://www.ovarianresearch.com/content/7/1/93)

Figure 2 Comparison of IVF cycle outcomes between the DHEA group and the control.
number of antral follicles, fewer ovulated oocytes, significantly higher rates of GC and follicles apoptosis, poor responses to ovarian induction with gonadotropins [21]. ARKO model demonstrated that ARs are crucial important for normal follicle genesis and development. Higher concentration of AR mRNA and AR protein were detected in pre-antral and early antral follicles which suggested the peak effect of androgens [22].

To gain further light into the effect of androgens, preovulatory granulose cells in follicular fluid were collected from patients undergoing IVF treatment, and cultured with the absence or presence of a low concentration of DHEA-S by ElBeltagy [23]. Increased expression of AR was detected after treatment of DHEAS compared to cells absence of DHEAS. This study explained that DHEAS may exert its effect on follicle development through up-regulating AR.

Human GC and follicular fluid from small antral follicles were obtained from ovaries surgically removed for fertility preservation by Nielsen et al. [24], who noted the expression of AR mRNA in GC and concentrations of androgens in follicular fluid were correlated with the expression of FSH receptor mRNA. Moreover, the expression of AR mRNA also showed a positive association with AMH receptor in their research. This indicated that proper androgens may potentially increase FSH receptor and AMH receptor.

In one word, androgens appear most modulate FSH responsibility through AR at stages of pre-antral and antral follicles [25]. Addition of DHEA could increase FSH receptor at early stages of folliculogenesis. Therefore DHEA could beneficially affect follicle recruitment.

Additionally, the secondary effect of DHEA on estrogen receptor could not be neglected. According to the two cell/two gonadotrophin theory, androgens play an essential role in ensuring adequate follicular steroidogenesis in human. DHEA is a crucial precursor steroid to human sex steroid synthesis and is converted to androgens or estrogens. Estrogens promote ovarian follicular growth and granulosa cell proliferation [26].

DHEA promotes the growth of small follicles, especially preantral and early-antral follicles. Since AMH is secreted by them, it is not difficult to understand the increase of AMH. However, the increase of AFC did not reach statistically difference. Why? Pre-antral and early-antral follicles could not be seen on ultrasound. We can only monitor the count of antral follicles, not the total number of pre-antral, early-antral and antral follicles. And AMH may reflect not only the quantity but also the quality of small follicles. The increase of AMH may be caused by the increased small follicles or the improved follicle quality or the both.

Here in our study, a significantly higher level of BMP-15 in follicular fluid of patients treated with DHEA was also observed. We speculated that DHEA may probably affect the level of BMP-15 in circulation via androgen receptors (AR). It has been demonstrated that multiple regulatory factors such as tyrosine kinases by Ligand (KITL), BMP-15, GDF-9 and so on are under the

Table 3 Ovarian reserve markers in patients before and after DHEA treatment

| Parameters          | Pre-treatment | Post-treatment | P-value |
|---------------------|---------------|----------------|---------|
| AMH (ng/ml)         | 1.01 ± 0.77   | 1.29 ± 1.09   | .015    |
| FSH (mIU/mL)        | 11.68 ± 6.62  | 9.45 ± 5.09   | .036    |
| Estradiol (pg/ml)   | 186.58 ± 142.19 | 110.79 ± 68.28 | .002    |
| DHEA-S (μg/mL)      | 1.24 ± 0.74   | 5.50 ± 3.48   | .000    |
| Testosterone (nmol/L) | 0.74 ± 0.41   | 2.25 ± 1.28   | .000    |
| Antral follicle count | 2.95 ± 1.38   | 3.21 ± 1.22   | .054    |

Data are presented as mean ± SD (standard deviation). P<.05 was considered as statistically significant.
regulation of AR [27]. And there exists an exchange loop between oocytes and granulose cells.

It has been confirmed that both GDF-9 and BMP-15 stimulate FSH receptor and increase the sensitivity to FSH but only BMP-15 was observed to up-regulate AMH expression at mRNA and protein level in GC [28]. What is more, recent research indicated that high level of FSH suppressed the expression of BMP-15, and there is a negatively correlation between them [29]. It may be inferred that here exist a feedback loop between BMP-15 and FSH. Therefore in present study, it is reasonable for us to assume that DHEA supplementation may potentially enhance FSH sensitivity and increase the expression of BMP-15 in follicular fluid. However, no significant difference was found in the level of GDF-9. It is likely that there exists some difference in the paths of regulating oocyte growth between BMP-15 and GDF-9 though they make function in a synergistic way. And the relationships between BMP-15 and androgens need further research to clarify.

**Strengths and limitations**

Strengths of this study were: 1) It was the first study concerning the relationship between DHEA supplementation and BMP-15, GDF-9 in follicular fluid. 2) Follicular fluid samples were collected distinguished serum samples in former studies assessed the effect of DHEA; 3) This research was conducted in a prospective way which ensured the consistency and comparability of the study group and the control.

Limitations of the study: It is not a placebo-controlled trial for patients with DOR were eager to receive treatment other than take placebo pills because of limited time to fertility.

**Conclusions**

In summary, DHEA supplementation improves follicular fluid BMP-15, serum AMH and accumulated embryo score of patients with DOR. Herein, addition of DHEA prior to IVF-cycles is an effective option for patients with DOR.

**Abbreviations**

DHEA: Dehydroepiandrosterone; DOR: Diminished ovarian reserve; NF: In vitro fertilization; BMP-15: Bone morphogenetic protein-15; GDF-9: Growth differentiation factor-9; AMH: Anti-Mullerian hormone; FSH: Follicle stimulating hormone; E2: Estradiol.

**Competing interests**

The authors declare that they have no competing interests.

**Authors’ contributions**

JW and YC collected patients; HZ assigned patients and wrote the article; WZ and MX collected blood and follicular fluid samples; YX made statistics; JW and YC collected patients; HZ assigned patients and wrote the article; GDF-9: Growth differentiation factor-9; AMH: Anti-Mullerian hormone; FSH: Follicle stimulating hormone; E2: Estradiol.

**Funding**

This research was funded by National Science and Technology Support Program (2012BAI3B01) and Anhui Provincial Natural Science Foundation (1208085MH161).

**Author details**

1. Department of Reproductive Endocrinology, the Reproductive Medicine Center, The Affiliated Hospital of Anhui Medical University, Anhui, People’s Republic of China.

2. Laboratory of the Reproductive Medicine Center, The Affiliated Hospital of Anhui Medical University, Anhui, People’s Republic of China.

**Acknowledgments**

Supported by National Science and Technology Support Program (2012BAI3B01), Anhui Provincial Natural Science Foundation (1208085MH161), People’s Republic of China.

**Received**

2 August 2014

**Accepted**

30 September 2014

**Published online**

21 October 2014

**References**

1. Frattarelli JL, Hill MJ, McWilliams GD, Miller KA, Bergh PA, Scott RT Jr: A luteal estradiol protocol for expected poor-responders improves embryo number and quality. Fertil Steril 2008, 89(1):118–122.

2. Uyg U, Ben-Shlomo I, Turan E, Eden HF, Alman MA, Rehacci M: Conception rates following assisted reproduction in poor responder patients: a retrospective study in 300 consecutive cycles. Reprod Biomed Online 2003, 6:489–493.

3. Casson PR, Lindsay MS, Piatak MD, Carson SA, Buster JE: Dehydroepiandrosterone supplementation augments ovarian stimulation in poor responders: a case series. Hum Reprod 2000, 15:2129–2132.

4. Barad DH: Effect of dehydroepiandrosterone on oocyte and embryo yields, embryo grade and cell number in IVF. Hum Reprod 2006, 21:2845–2849.

5. Barad D, Brill H, Gleicher N: Update on the use of dehydroepiandrosterone supplementation among women with diminished ovarian function. J Assist Reprod Genet 2007, 24:629–634.

6. Gleicher N, Ryan E, Weghofer A, Blanco-Mejia S, Barad DH: Miscarriage rates after dehydroepiandrosterone (DHEA) supplementation in women with diminished ovarian reserve: a case control study. Reprod Biol Endocrinol 2009, 7:108.

7. Gleicher N, Barad DH: Androgen priming before ovarian stimulation for IVF. Hum Reprod 2008, 23:2688–2670.

8. Barad D, Weghofer A, Gleicher N: Dehydroepiandrosterone treatment of ovarian failure. Fertil Steril 2009, 91:e14.

9. Yilmaz N, Uygur D, Inal H, Gorkem U, Cicek N: Mollamahmutoglu: Dehydroepiandrosterone supplementation improves predictive markers for diminished ovarian reserve: serum AMH, inhibin B and antral follicle count. Eur J Obstet Gynecol Reprod Biol 2013, 169:257–260.

10. Yeung TW, Li RH, Lee VC, Ho PC, Ng EH: A Randomized Double-Blinded Placebo-Controlled Trial on the Effect of Dehydroepiandrosterone for 16 Weeks on Ovarian Response Markers in Women with Primary Ovarian Insufficiency. J Clin Endocrinol Metab 2013, 98:380–388.

11. Orisaka M, Jiang JY, Orisaka S, Kotsuji F, Tsang BK: Growth Differentiation Factor 9 promotes rat pre-antral follicle growth by up-regulating follicular androgen biosynthesis. Endocrinology 2009, 150:2740–2748.

12. Enomoto C, Sugita K: Role of oocyte-derived paracrine factors in follicular development. Annu Rev Cell Dev Biol 2014, 30:627–633.

13. Dong J, Albertini DF, Nishimori K, Kumar TR, Lu N, Matzuk MM: Growth differentiation factor-9 is required during early ovarian folliculogenesis. Nature 1996, 381:531–535.

14. Wu YT, Tang L, Cai J, Lu YE, Xu J, Zhu XM, Luo Q, Huang HF: High bone morphogenetic protein-15 level in follicular fluid is associated with high quality oocyte and subsequent embryonic development. Hum Reprod 2007, 22:1526–1531.

15. Ferrari AP, La Marca A, Fauser BC: ESHRE consensus on the definition of poor response to ovarian stimulation for in vitro fertilization: the Bologna criteria. Hum Reprod 2011, 26:1616–1624.

16. Wiser A, Gonen O, Ghereti Y: Addition of dehydroepiandrosterone (DHEA) for poor-responder patients before and during IVF treatment improves the pregnancy rate: A randomized prospective study. Hum Reprod 2010, 25:2496–2500.

17. Wu YT, Wang TT, Chen XL, Zhu XM, Dong MY, Sheng J, Xu CM, Huang HF: Bone morphogenetic protein-15 in follicle fluid combined with age may differentiate between successful and unsuccessful poor ovarian responders. Reprod Biol Endocrinol 2012, 26:116.

18. Gleicher N, Weghofer A, Barad DH: Improvement in diminished ovarian reserve after dehydroepiandrosterone (DHEA) supplementation. Reprod Biol Online 2010, 21:360–365.

19. Moawad A, Shearer M: Long term androgen priming by use of DHEA improves IVF outcomes in poor responders: A randomized control study. Middle East Fertil Soc J 2012, 17:268–274.
20. Gleicher N, Weghofer A, Barad DH. The role of androgens in follicle maturation and ovulation induction: friend or foe of infertility treatment? Reprod Biol Endocrinol 2011, 9:116.

21. Sen A, Hammes SR. Granulosa-cell specific androgen receptors are critical regulators of ovarian development and function. Mol Endocrinol 2010, 24:1395–1403.

22. Weil SS, Vendola K, Zhou J, Adesanya OO, Wang J, Okafor J, Bondy CA. Androgen receptor gene expression in the primate ovary: cellular localization, regulation, and functional correlations. J Clin Endocrinol Metab 1998, 83:2479–2485.

23. Elbeltagy K, Honda K, Ozaki K, Tokuyama O, Kimura M, Kira Y, Ishiko O. In vitro effect of dehydroepiandrosterone sulfate on steroid receptors, aromatase, cyclooxygenase-2 expression, and steroid hormone production in preovulatory human granulosa cells. Fertil Steril 2007, 88(Suppl 4):1135–1142.

24. Nielsen ME, Rasmussen IA, Kristensen SG, Christensen ST, Mølgaard K, Wreford Andersen E, Byskov AG, Yding Andersen C. In human granulosa cells from small antral follicles, androgen receptor mRNA and androgen levels in follicular fluid correlate with FSH receptor mRNA. Mol Hum Reprod 2011, 17:63–70.

25. Lerie S, Smitz J. Functional AR signaling is evident in an in vitro mouse follicle culture bioassay that encompasses most stages of folliculogenesis. Biol Reprod 2000, 60:665–665.

26. Sunkara SK, Coomarasamy A, Arlt W, Bhattacharya S. Should androgen supplementation be used for poor ovarian response in IVF. Hum Reprod 2012, 27:637–640.

27. Hickey TE, Marrocco DL, Amato F, Ritter LJ, Norman RJ, Gilchrist RB, Armstrong DT. Androgens augment the mitogenic effects of oocyte-secreted factors and growth differentiation factor 9 on porcine granulosa cells. Biol Reprod 2005, 73:825–832.

28. Ogura-Nose S, Yoshino O, Osuga Y, Shi J, Hikoi H, Yano T, Taketani Y. Anti-Mullerian hormone (AMH) is induced by bone morphogenetic protein (BMP) cytokines in human granulosa cells. Eur J Obstet Gynecol Reprod Biol 2012, 164:44–47.

29. Thomas FH, Ether JF, Shimasaki S, Vanderhyden BC. Follicle-Stimulating Hormone Regulates Oocyte Growth by Modulation of Expression of Oocyte and Granulosa Cell Factors. Endocrinology 2005, 146:941–949.

doi:10.1186/s13048-014-0093-3
Cite this article as: Zhang et al. Dehydroepiandrosterone improves follicular fluid bone morphogenetic protein-15 and accumulated embryo score of infertility patients with diminished ovarian reserve undergoing in vitro fertilization: a randomized controlled trial. Journal of Ovarian Research 2014 7:93.