Abstract. Mounting evidence has shown that miR-23b-3p, which is associated with cell proliferation, invasion, and apoptosis, acts as a biomarker for diagnosis and outcomes in numerous cancers. However, the clinicopathological implication of miR-23b-3p in hepatocellular carcinoma (HCC) remains unclear. Our study evaluated the role of miR-23b-3p in HCC and investigated its potential application as a marker for preliminary diagnosis and therapy in HCC. High-throughput data from the NCBI Gene Expression Omnibus (GEO) and The Cancer Genome Atlas (TCGA) were collected and analyzed. One hundred and one tissue sections of HCC were paired with adjacent non-cancerous HCC as further supplements. miR-23b-3p expression was detected using quantitative real-time PCR. Additionally, the relationship between miR-23b-3p expression and HCC progression and Time-to-recurrence (months) was explored. Ten algorithms were applied to predict the prospective target genes of miR-23b-3p. Next, we conducted bioinformatics analysis for further study. miR-23b-3p expression was pronouncedly decreased in HCC tissues in contrast with their paired adjacent non-cancerous HCC (P<0.001) with RT-qPCR. In total, 405 targets, acquired with consistent prediction from at least five databases, were used for the bioinformatics analysis. According to the Gene Ontology (GO) analysis, all targets were classified into biological processes, cellular components and molecular functions. In the pathway analysis, targets of miR-23b-3p were primarily enriched in the signaling pathways of renal cell carcinoma, hepatitis B and pancreatic cancer (corrected P-value <0.05). In the protein-protein interaction (PPI) network for miR-23b-3p, a total of 8 targets, including SRC, AKT1, EGFR, CTNNB1, BCL2, SMAD3, PTEN and KDM6A, were located in the key nodes with high degree (>35). In conclusion, this study provides impressive illumination of the potential role of miR-23b-3p in HCC tumorigenesis and progression. Furthermore, miR-23b-3p may act as a predictor of HCC and could be a new treatment target.

Introduction

Hepatocellular carcinoma (HCC) is a malignant tumor that has one of the highest morbidity and mortality rates, and its tumorigenesis is closely related to HBV and HCV infection (1). There were estimated 782,500 hepatic carcinoma cases and 745,500 related deaths globally in 2012, according to Torre et al (2). Moreover, estimated 70-90% of primary liver cancers are HCC. Despite advances in chemotherapy, radiotherapy and surgery, the low 5-year survival and quality of life observed in HCC patients remain intractable issues. Many etiological factors, such as alcohol consumption and aflatoxin B1 exposure, are widely accepted to facilitate HCC occurrence (3,4); however, the precise molecular mechanisms remain unknown. Thus, there is an urgent need for a reliable biomarker that can be used to predict HCC progression and prognosis.

MicroRNAs (miRNAs), which are limited to a length of 19-22 nucleotides, are small non-coding RNAs that post-transcriptionally influence gene expression (5). A vast array of oncogenes and anti-oncogenes are potentially regulated by miRNAs, which act as gene regulators by conjugating the 3'-untranslated region (3'-UTR) of their mRNAs. In regulating genes, miRNAs play a crucial biological role in tumor development, especially during initiation, proliferation, differentiation, invasion and metastasis (6). Thus, miRNAs may serve as promising diagnostic and prognostic indexes. Validating a potential target of miRNA is arduous and time-consuming due to the tremendous amounts of target
sites in miRNA. However, predicting the targets of miRNA, which is a vital step in performing miRNA-target interactions, contributes to narrowing down prospective target sites and promoting experimental verification. Following the principle of sequence complementarity, many algorithms were manipulated to figure out the predicted miRNA targets.

miR-23b-3p was identified as a tumor suppressor that showed a tendency toward downregulated expression in different classes of human malignant tumors such as prostate cancer, renal cell carcinoma, acute myeloid leukemia and osteosarcoma (7-10). Nevertheless, increasing evidence has indicated that the upregulation of miR-23b-3p promoted cell proliferation and invasion in glioma, gastric cancer and breast cancer (11-13). miR-23b-3p was also found to be involved in liver stem cell differentiation, and its reduced expression may contribute to liver regeneration after partial hepatectomy (14,15). However, to our knowledge, only one study (16) has explored the relationship between miR-23b-3p expression and HCC but did not comment on the clinicopathological significance and prognosis. Therefore, further inquiry is urgently needed.

Our objectives were as follows: to study the pathophysiologic expression quantity of miR-23b-3p in HCC tissue through comparison with matching adjacent tissues as well as to elucidate the correlation between miR-23b-3p expression and HCC clinicopathological parameters. Additionally, we aimed to identify the probable target genes of miR-23b-3p and determine the potential role of miR-23b-3p in HCC development and progression.

Materials and methods

miR-23b-3p expression in HCC based on GEO datasets

Data acquisition and exclusion criteria. Twenty-one microarray datasets were obtained from the GEO database (http://www.ncbi.nlm.nih.gov/geo/), and 11 were eliminated after screening. The search strategy was formulated as follows: (malignant OR cancer OR tumor OR tumour OR neoplas* OR carcinoma) AND (hepatocellular OR liver OR hepatic OR HCC). The last dataset search was on April 2016. The exclusion criteria were as follows: i) datasets without information on miR-23b-3p; ii) datasets without complete data for analysis; iii) samples based on cell lines; iv) not all subjects of the included studies were human; or v) miR-23b-3p was determined in the HCC patients without a comparison. To identify the clinical relevance of miR-23b-3p in HCC, we collected data on miR-23b-3p expression from the 6 datasets and analyzed their association with the clinicopathological characteristics of HCC. The clinical features, which could be obtained from more than two microarray datasets, were used for the meta-analysis.

Statistical analysis. The meta-analysis was carried out with Stata 12.0 (StataCorp LP, College Station, TX, USA). A standard mean difference (SMD) and a 95% confidence interval (CI) were utilized to measure continuous outcomes, including age, sex, HBV infection, invasion and metastasis. Fixed or random effects models were applied to pool the effect sizes. Cochran's Q test (Chi-square test; Chi2) (17) and inconsistency (I2) (18) were conducted to assess heterogeneity. A P<0.05 or I2>50% indicates significant heterogeneity (19), and a random effects model (20) was applied. Otherwise, the fixed effects model would be adopted. Begg's funnel plot for asymmetry and Egger's funnel plot for quantitation were generated to evaluate publication bias. In addition, sensitivity analysis was performed to evaluate the reliability of results by elimination of a study each time.

miR-23b-3p expression in HCC based on TCGA dataset

Retrieval of public data. Altogether, 377 anonymized HCC tissues and 50 normal tissues were retrieved from the TCGA database (http://cancergenome.nih.gov/publications/publicationguidelines). The non-HCC samples or samples with data deficiency were excluded; 361 HCC patients were finally included in this study. Additionally, a total of 50 normal liver tissues were retrieved for comparison. The clinicopathological features of HCC patients in TCGA are available in Table II.

Statistical analysis. The statistics were analyzed with SPSS 22.0 software (IBM Corp., Armonk, NY, USA). The final data after calculation are shown as the mean ± standard deviation (SD). Student's t-test was conducted for a comparative analysis of two independent groups. A one-way analysis of variance (ANOVA) test was utilized to evaluate the association between miR-23b-3p expression and other clinicopathological parameters. The relationship between miR-23b-3p and recurrence was obtained with the Kaplan-Meier survival approach along with a log-rank test. To differentiate between controls and HCC tissues, a diagnostic value was identified using a receiver operator characteristic (ROC) curve. A P-value <0.05 denoted a statistically significant difference. At least 5 individual tests were applied without ambiguity.
Statistical analysis. The statistical software and methods that were used to analyze the data from clinical patient files downloaded from TCGA were also used to identify the relationship between miR-23b-3p expression and HCC based on qPCR data.

Meta-analysis for GEO datasets, TCGA datasets and PCR verification in-house
Data acquisition. We next added the data from TCGA datasets and PCR verification in-house into meta-analysis to expand the sample size and enhance the credibility. The expression of hsa-miR-23b-3p in HCC tissues and adjacent non-cancerous tissues were extracted in meta-analysis.

Statistical analysis. As for statistical analysis of meta-analysis of GEO datasets, the same statistical methods and software were used to perform the meta-analysis of GEO datasets, TCGA datasets and qPCR data.

Biological information analysis
Prediction targets of miR-23b-3p collection. Ten algorithms, including those from TargetScan (http://www.targetscan.org), microRNA.org (http://www.microrna.org), RNA22 (https://cm.jefferson.edu/rna22/Precomputed), PicTar-vert (http://pictar.mdc-berlin.de/cgi-bin/PicTar_ vertebrate.cgi), miRDB (http://mirdb.org/miRDB), PolymiRTS Database (http://compbio.utexas.edu/miRSNP/), PITA (http://genie.weizmann.ac.il/pubs/mir07/mir07_dyn_data.html#), TargetMiner (http://www.isical.ac.in/~bioinfo_miu/targetminer20.htm), TarBase (http://diana.mis.athenainnovation.gr/DianaTools/index.php?r=tarbase), and miRTarBase (http://diana.imis.athenainnovation.gr/DianaTools/index.php?r=tarbase), and miRTarBase (http://pictar.mdc-berlin.de/cgi-bin/PicTar_ vertebrate.cgi). The validation targets of hsa-miR-23b-3p were divided into three primary classes: targets of miR-23b-3p were integrated with prediction targets after combination.

Validation targets of miR-23b-3p collection. We searched the PubMed database using the following search strategy: (MIRN23 OR microRNA23 OR microRNA-23 OR miR-23 OR hsa-mir-23 OR miR-23b OR microRNA-23b OR miRNA-23b OR 'miR23b' OR 'miRNA23b' OR 'microRNA23b' OR miR-23b-3p OR microRNA-23b-3p OR microRNA-23b-3p and target'). The validation targets of hsa-miR-23b-3p were recorded from the identified articles by two authors (P-R Wu and X-L Xiang), and the third author resolved controversies. These targets were integrated with prediction targets after combination.

Bioinformatics analysis of miR-23b-3p. Gene Ontology (GO) analysis. GO analysis was conducted using the Database for Annotation, Visualization and Integrated Discovery (DAVID) (http://David.abcc.ncifcrf.gov/). The targets of miR-23b-3p were divided into three primary classes: biological processes, cellular components and molecular functions.

Pathway analysis. For pathway analysis, we downloaded pathway data from the Kyoto Encyclopedia of Genes and Genomes (KEGG) (http://www.genome.ad.jp/KEGG) to investigate gene interactions. miR-23b-3p targets were mapped to the KEGG pathway database using KOBAS 2.0 (http://www.genome.jp/kegg/pathway.html), and P-values [and a corresponding false discovery rate (FDR)] were applied to estimate each enriched pathway.

Network analysis. The Search Tool for the Retrieval of Interacting Genes (STRING), which provides information for experimental and predicted interactions, is an online database. STRING was applied to search and to determine an interaction network of targets of miR-23b-3p via confidence score calculation.

Results

miR-23b-3p expression in HCC based on GEO datasets
Characteristics of the included datasets. The characteristics of the GEO datasets included are presented in Table I. A total of 10 datasets, including GSE6857 (USA, 2013) (21), GSE10694 (China, 2008) (22), GSE21362 (Japan, 2010) (23), GSE12717 (China, 2008) (24), GSE54751 (USA, 2014) (25), GSE57555 (Japan, 2015) (26), GSE41874 (Japan, 2013) (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE41874), GSE67138 (USA, 2015) (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE67138), GSE69580 (China 2015) (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE69580), and GSE67139 (USA, 2015) (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE67139), were identified to meet the standards. All of the datasets were derived from tissues.

Value of miR-23b-3p as a biomarker for HCC. In 7 datasets presenting miR-23b-3p expression with comparable groups, 414 patients with HCC and 343 healthy subjects were recruited. The expression of miR-23b-3p was lower in HCC patients than healthy controls (Fig. 1A); however, no statistical significance was found (SMD=-0.801; 95% CI, -0.216 to 0.054; P=0.239). A fixed effects model was selected when we eliminated the study of GSE6857, suggesting that this study may influence the pooled SMD and 95% CI. The forest plot after the removing of GSE6857 is shown in Fig. 2A; however, no statistical significance was found (SMD=-0.101 (95% CI, -0.203 to 0.405, P=0.515) (Fig. 3A), -0.266 (95% CI, -1.293 to 0.762, P=0.613) (Fig. 3C), -0.186 (95% CI, -1.093 to 1.465, P=0.775) (Fig. 3D), and -0.288 (95% CI, -0.787 to 0.210, P=0.257) (Fig. 3E), respectively. As a consequence, no publication bias was discovered. According to sensitivity analysis shown in Fig. 1D, the result was altered when we eliminated the study of GSE6857, suggesting that this study may influence the pooled SMD and 95% CI. The forest plot after the removing of GSE6857 is shown in Fig. 2A; however, no statistical significance was found (SMD=-0.101 (95% CI, -0.203 to 0.405, P=0.515) (Fig. 3A), -0.266 (95% CI, -1.293 to 0.762, P=0.613) (Fig. 3C), -0.186 (95% CI, -1.093 to 1.465, P=0.775) (Fig. 3D), and -0.288 (95% CI, -0.787 to 0.210, P=0.257) (Fig. 3E), respectively. None of the five parameters were associated with the level of miR-23b-3p expression.

Six datasets were enrolled to assess the association between miR-23b-3p expression and the clinical aspects of HCC. However, five parameters, including invasion (83 patients), age (90 patients), sex (8 patients), HBV infection (66 patients) and metastasis (27 patients) were estimated in this study. The pooled SMD were -1.302 (95% CI, -3.055 to 0.451, P=0.146) (Fig. 3A), 0.101 (95% CI, -0.203 to 0.405, P=0.515) (Fig. 3B), -0.266 (95% CI, -1.293 to 0.762, P=0.613) (Fig. 3C), -0.186 (95% CI, -1.093 to 1.465, P=0.775) (Fig. 3D), and -0.288 (95% CI, -0.787 to 0.210, P=0.257) (Fig. 3E), respectively. None of the five parameters were associated with the level of miR-23b-3p expression.

miR-23b-3p expression in HCC based on TCGA dataset
The expression level of miR-23b-3p was downregulated in HCC samples. To explore the relationship between miR-23b-3p and HCC, we further obtained data for 361 HCC
tissues and 50 normal liver tissues from TCGA. The clinicopathological features are presented in Table II. According to the TCGA data, the expression level of miR-23b-3p was substantially downregulated in HCC (12.6722±0.978150) being compared to normal liver tissues (13.4039±0.51072, P<0.001) (Fig. 4A).

**Relationships between miR-23b-3p and clinicopathological features.** Due to the downregulated miR-23b-3p expression observed in HCC, the association between clinicopathological features and miR-23b-3p was studied (Table II and Fig. 5). The data indicated that a higher level of miR-23b-3p was closely related with hepatitis B (12.8831±0.96070), in contrast to that without hepatitis B infection (12.5999±0.97009, P=0.013). miR-23b-3p was also downregulated in smoking patients (12.2025±1.11847) compared to those without smoking history (12.7127±0.96181, P=0.035). The results also gave a demonstration that the downregulated expression of miR-23b-3p was closely associated with vascular invasion (12.5905±0.978150) being compared to normal liver tissues (13.4039±0.51072, P<0.001) (Fig. 4A).

**Figure 1.** Meta-analysis of miR-23b-3p expression in hepatocellular carcinoma (HCC) patients and healthy controls retrieved from Gene Expression Omnibus (GEO) datasets. (A) Forest plot. The center dot and the horizontal line represent the standard mean difference (SMD) and 95% confidence interval (CI), respectively. The pooled SMD and its corresponding 95% CI are presented below the list of studies; (B) Begg's funnel plot. Hazard ratios are presented on a logarithmic scale; (C) Egger's publication bias plot. Hazard ratios are presented on a logarithmic scale; (D) sensitivity analysis.
applied when correlating miR-23b-3p with clinicopathological parameters, including hepatitis B (r=0.134, P=0.013), smoking (r=-0.014, P=0.035), vascular invasion (r=-0.122, P=0.032), and N status (r=-0.158, P=0.003). There were no other positive correlations with the remaining clinical features.

The diagnostic accuracy of miR-23b-3p in HCC tissues. To further determine miR-23b-3p as a biomarker in diagnosing HCC, we performed ROC for verification. As shown in Fig. 4B, the AUC of miR-23b-3p in HCC and its counterpart normal tissues was 0.737 (95% CI, 0.680 to 0.794; P<0.001). The cut-off value reached 0.429; the sensitivity and specificity were 82.0 and 60.9%, respectively. The result suggested that miR-23b-3p may be treated as a reliable biomarker in diagnosing HCC. As shown in Fig. 6, the AUC to judge hepatitis B was 0.597 (95% CI, 0.532 to 0.662; P=0.004). The cut-off value for miR-23b-3p was 0.174; the sensitivity and specificity were 65.1 and 52.3% respectively. The AUC to judge smoking was 0.626 (95% CI, 0.485 to 0.767; P=0.080). The cut-off value for miR-23b-3p was 0.259; the sensitivity and specificity were 78.8 and 47.1%, respectively. The AUC to judge vascular invasion was 0.566 (95% CI, 0.499 to 0.633, P=0.057). The cut-off value for miR-23b-3p was 0.125; the sensitivity and specificity were 45.2 and 67.3%, respectively. The AUC to judge N status was 0.601 (95% CI, 0.538 to 0.664, P=0.002). The cut-off value for miR-23b-3p was 0.125; the sensitivity and specificity were 63.2 and 55.8%, respectively.

miR-23b-3p expression in surviving HCC. According to the data from TCGA, 359 of the 361 patients were followed up. The Kaplan-Meier analysis in Fig. 7 showed that 176 patients had a lower miR-23b-3p expression (lower than the mean expression.

Figure 2. Meta-analysis of removing a single study from Gene Expression Omnibus (GEO) datasets. (A) Forest plot. The center dot and the horizontal line represent the standard mean difference (SMD) and 95% confidence interval (CI), respectively. The pooled SMD and its corresponding 95% CI are presented below the list of studies; (B) Begg’s funnel plot. Hazard ratios are presented on a logarithmic scale. (C) Egger’s publication bias plot. Hazard ratios are presented on a logarithmic scale; (D) sensitivity analysis. Pooled SMDs and 95% CIs by eliminating each study.
level of 12.6797), whereas the remaining 183 patients had a higher level (higher than the mean expression level of 12.6797). In contrast, the lower group had an average survival time of 1,617.668±151.722 days, whereas the higher group had a survival time of 1,944.626±160.587 days. However, the survival times of the high and low miR-23b-3p expression groups were not significantly different (Chi-square =3.351, \( P=0.061 \)).

**miR-23b-3p expression in HCC tissues from PCR verification in-house**

The expression of miR-23b-3p was downregulated in HCC. To study the expression level of miR-23b-3p, a quantitative real-time PCR method was performed in 101 HCC tissues and counterpart adjacent non-cancerous HCC. The clinicopathological features of the patients, obtained from medical records, are available in Table III. The expression of miR-23b-3p was obviously downregulated in HCC cases (2.7376±1.99328) in contrast to their counterpart adjacent non-cancerous HCC (2.7376±1.57739, \( P<0.001 \)) (Fig. 8A). The TCGA and PCR data provided similar results.

![Figure 4. miR-23b-3p expression in hepatocellular carcinoma (HCC) tissues from The Cancer Genome Atlas (TCGA). (A) The difference in relevant miR-23b-3p expression between HCC and adjacent non-cancerous HCC; (B) ROC curve to distinguish HCC from normal tissues. Error bars represented standard deviation (SD).](image)

**miR-23b-3p expression in HCC tissues from PCR verification in-house**

The expression of miR-23b-3p was downregulated in HCC. To study the expression level of miR-23b-3p, a quantitative real-time PCR method was performed in 101 HCC tissues and counterpart adjacent non-cancerous HCC. The clinicopathological features of the patients, obtained from medical records, are available in Table III. The expression of miR-23b-3p was obviously downregulated in HCC cases (2.7376±1.99328) in contrast to their counterpart adjacent non-cancerous HCC (2.7376±1.57739, \( P<0.001 \)) (Fig. 8A). The TCGA and PCR data provided similar results.

**The relationships between miR-23b-3p and clinicopathological features.** As noted above, the expression of miR-23b-3p was remarkably lower in the HCC tissues than their counterpart adjacent non-cancerous HCC, which indicated that miR-23b-3p may perform the function of a feasible tumor suppressive miRNA in HCC. We investigated the relationships between miR-23b-3p expression and clinicopathological features. The results shown in Table III and Fig. 9 validated that the downregulated level of miR-23b-3p was positively related to patients without metastasis (3.3959±1.57506), in contrast with those with metastasis (2.1173±1.31762, \( P<0.001 \)). Subsequently, the miR-23b-3p level was significantly positively associated with multiple tumor nodes (3.0158±1.66606) compared with single nodes (2.3773±1.66606, \( P=0.043 \)). The presence of a portal
Venous tumor embolus showed a lower expression of miR-23b-3p (1.9500±1.19055) than did its absence (3.1029±1.60797, \(P<0.001\)). Furthermore, we used Spearman's method to record the correlations between the expression of miR-23b-3p and clinicopathological parameters, including tumor nodes (\(r=-0.255, P=0.010\)), metastasis (\(r=-0.447, P<0.001\)), and portal vein embolus (\(r=-0.340, P=0.001\)). Nonetheless, we failed to gain any positive correlations between miR-23b-3p expression and other clinical features.

The diagnostic value of the miR-23b-3p level in HCC tissues. To verify the diagnostic value of the miR-23b-3p level in HCC tissues, a ROC curve was utilized. As shown in Fig. 8B, the area under the curve (AUC) of miR-23b-3p in HCC and adjacent non-cancerous HCC was 0.765 (95% CI, 0.700 to 0.830, \(P<0.001\)). At 0.396 (the cut-off value of miR-23b-3p), the sensitivity and specificity was 88.2 and 55.4%, respectively. In Fig. 10, the AUC to judge tumor nodes was 0.648 (95% CI, 0.538 to 0.758, \(P=0.011\)). The cut-off value for miR-23b-3p was 0.272; the sensitivity and specificity was 52.6 and 68.2%, respectively. The AUC to judge metastasis was 0.758 (95% CI, 0.664 to 0.851, \(P<0.001\)). The cut-off value for miR-23b-3p was 0.468; the sensitivity and specificity was 77.6 and 69.2%, respectively. The AUC to judge portal vein tumor embolus was 0.711 (95% CI, 0.601 to 0.820, \(P=0.001\)). The cut-off value for miR-23b-3p was 0.29; the sensitivity and specificity was 88.4 and 40.6%, respectively.

**Table I.** Characteristics of hsa-miR-23b-3p gene expression profiling datasets included in meta-analysis.

| Citation          | Country | Data source | HCC patients | Healthy controls | Platform         |
|-------------------|---------|-------------|--------------|-----------------|-----------------|
| Budhu et al 2013  | USA     | GEO: GSE6857| 240          | 241             | GPL4700         |
| Li et al 2008     | China   | GEO: GSE10694| 75           | 87              | GPL6542         |
| Sato et al 2011   | Japan   | GEO: GSE21362| 73           | 73\(^a\)        | GPL10312        |
| Su et al 2009     | USA     | GEO: GSE12717| 5            | 3               | GPL7274         |
| Shen et al 2015   | USA     | GEO: GSE54751| 10           | 10\(^a\)        | GPL18262        |
| Murakami et al 2015| Japan | GEO: GSE57555| 5            | 5\(^a\)+11       | GPL16699        |
| Morita 2013\(^b\) | Japan   | GEO: GSE41874| 6            | 4               | GPL7722         |
| Hung 2015\(^b\)   | China   | GEO: GSE69580| 5            | 5\(^a\)         | GPL10850        |
| Barry 2015\(^b\)  | USA     | GEO: GSE67138| 23           | 34              | GPL8786         |
| Barry 2015\(^b\)  | USA     | GEO: GSE67139| 60           | 60              | GPL8786         |

\(^a\)Cases from adjacent tumor tissues; \(^b\)no relevant references. HCC, hepatocellular carcinoma.

**Figure 5.** The relationship between miR-23b-3p expression and clinicopathological parameters of hepatocellular carcinoma (HCC) patients from The Cancer Genome Atlas (TCGA). (A) hepatitis B; (B) smoking history; (C) vascular invasion; (D) N status. Error bars represent standard deviation (SD).
HE et al: miR-23b-3p, A NOVEL BIOMARKER FOR HEPATOCELLULAR CARCINOMA

Among the 76 patients, 45 had a lower expression of miR-23b-3p (lower than the median level of 2.600), whereas 31 had higher miR-23b-3p expression. The lower expression group had 54.257±2.943 months of survival time without recurrence; the higher expression group had 54.509±3.962 months. Thus, there were no significant differences in survival time between high and low miR-23b-3p expression (Chi-Square =0.706, P= 0.401).

Value of miR-23b-3p as a biomarker for HCC. The pooled SMD and its 95% CI (Fig. 12A) indicated that people with lower expression of miR-23b-3p had a significant risk for HCC (SMD=-0.368; 95% CI, -0.689 to -0.048; P=0.024). A random effects model was selected to pool the effect variables with heterogeneity (P=0.000, I2=81.3%). The results of Begg’s and Egger’s test were 0.754 and 0.687, respectively (Fig. 12B and C); however, no publication bias was discovered. The sensitivity analysis showed that the results were altered when we removed the study of TCGA dataset (Fig. 13D).

Meta-analysis for GEO datasets, TCGA datasets and PCR verification in-house

Characteristics of included studies. Ten GEO datasets, TCGA dataset and the data derived from medical records, as mentioned above with 886 HCC patients and 587 normal persons in total, were included in the new meta-analysis. The characteristics of the studies were previously described (which were shown in sections ‘Characteristics of the included datasets’, ‘The expression level of miR-23b-3p was downregulated in HCC samples’ and ‘The expression of miR-23b-3p was downregulated in HCC’) in detail.

follow-up. A Kaplan-Meier analysis is presented in Fig. 11. Among the 76 patients, 45 had a lower expression of miR-23b-3p (lower than the median level of 2.600), whereas 31 had higher miR-23b-3p expression. The lower expression group had 54.257±2.943 months of survival time without recurrence; the higher expression group had 54.509±3.962 months. Thus, there were no significant differences in survival time between high and low miR-23b-3p expression (Chi-Square =0.706, P=0.401).
Bioinformatics analysis

Target genes of miR-23b-3p collection and integration. Using the 10 algorithms, 14,810 target genes were predicted to be modulated potentially by miR-23b-3p. The number of miR-23b-3p prediction targets of the 10 algorithms varied, ranging from 74 to 6,192 with an average of 2,375. Only targets that appeared more than or equal to five times among all 10 algorithms were finally applied for further analysis. In total, 357 targets from the algorithms were included in the next step.

After screening 203 articles from PubMed, we ultimately gathered 69 validation targets and found that 21 targets (ATG12, CA2, CFL2, CHUK, GLS, HMGB2, IL6R, KIAA1467, MET, NOTCH2, PLAU, PNRC2, PRDM1, PTEN, PTK2B, RRAS2, SEMA6D, SPRY2, TAB3, VHL and WBP2) were duplicated with prediction targets. By combining targets from algorithms and articles, we finally used 405 targets to conduct the bioinformatics analysis.

Bioinformatics analysis of miR-23b-3p targets. Using GO analysis, 405 targets regulated by miR-23b-3p were classified as biological processes, cellular components, and molecular functions (Table IV and Fig. 14). In the pathway analysis, targets of miR-23b-3p were primarily enriched in the signaling pathways of renal cell carcinoma, hepatitis B (shown in http://www.genome.jp/kegg-bin/show_pathway?hsa05161/hsa:6777%09red/hsa:4318%09red/hsa:8503%09red/hsa:353376%09red/hsa:596%09red/hsa:317%09red/hsa:1147%09red/hsa:5728%09red/hsa:6714%09red/hsa:4214%09red/hsa:355%09red/hsa:4088%09red/hsa:4089%09red/hsa:2185%09red/hsa:1869%09red/...).
Table II. Characteristics of the HCC patients included in TCGA.

| Clinicopathological features                  | N     | miR-23b-3p relevant expression ($2^{-\Delta Cq}$) | Correlation |
|-----------------------------------------------|-------|-------------------------------------------------|-------------|
|                                               |       | Mean ± SD                                       | t-value     | P-value | r-value | P-value |
| Tissue                                        |       |                                                 |             |         |         |         |
| Normal                                        | 50    | 13.4039±0.51072                                 | -8.286      | <0.001  | 0.246   | <0.001  |
| HCC                                           | 371   | 12.6722±0.97815                                 |             |         |         |         |
| Age (years)                                   |       |                                                 |             |         |         |         |
| <60                                           | 166   | 12.7336±0.97231                                 | 0.85        | 0.396   | -0.045  | 0.396   |
| ≥60                                           | 194   | 12.6464±0.9000                                 |             |         |         |         |
| Sex                                           |       |                                                 |             |         |         |         |
| Male                                          | 246   | 12.6581±0.93294                                 | -0.613      | 0.54    | 0.032   | 0.54    |
| Female                                        | 115   | 12.7259±1.0132                                 |             |         |         |         |
| Neoadjuvant treatment                         |       |                                                 |             |         |         |         |
| No                                            | 359   | 12.6798±0.97924                                 | 0.036       | 0.972   | -0.002  | 0.972   |
| Yes                                           | 2     | 12.6550±1.07244                                 |             |         |         |         |
| Radiation therapy                             |       |                                                 |             |         |         |         |
| No                                            | 231   | 12.6570±0.95382                                 | 0.998       | 0.319   | -0.065  | 0.319   |
| Yes                                           | 4     | 12.1762±1.03021                                 |             |         |         |         |
| Pharmaceutical treatment                      |       |                                                 |             |         |         |         |
| No                                            | 218   | 12.6475±0.96107                                 | 0.041       | 0.967   | -0.003  | 0.967   |
| Yes                                           | 12    | 12.6358±0.90350                                 |             |         |         |         |
| Alcohol consumption                           |       |                                                 |             |         |         |         |
| -                                             | 226   | 12.7394±0.99963                                 | 1.374       | 0.17    | -0.074  | 0.17    |
| +                                             | 117   | 12.5870±0.92032                                 |             |         |         |         |
| Hepatitis B                                   |       |                                                 |             |         |         |         |
| -                                             | 237   | 12.5999±0.97009                                 | -2.506      | 0.013   | 0.134   | 0.013   |
| +                                             | 106   | 12.8831±0.96070                                 |             |         |         |         |
| Hepatitis C                                   |       |                                                 |             |         |         |         |
| -                                             | 289   | 12.6963±0.97544                                 | 0.388       | 0.698   | -0.021  | 0.698   |
| +                                             | 54    | 12.6401±0.97811                                 |             |         |         |         |
| Non-alcoholic fatty liver                     |       |                                                 |             |         |         |         |
| -                                             | 324   | 12.6763±0.98327                                 | -0.874      | 0.383   | 0.047   | 0.383   |
| +                                             | 19    | 12.8774±0.81188                                 |             |         |         |         |
| Smoking                                       |       |                                                 |             |         |         |         |
| -                                             | 326   | 12.7127±0.96181                                 | 2.115       | 0.035   | -0.114  | 0.035   |
| +                                             | 17    | 12.2025±1.11847                                 |             |         |         |         |
| Cirrhosis                                     |       |                                                 |             |         |         |         |
| -                                             | 337   | 12.6956±0.95014                                 | 0.564       | 0.597   | -0.063  | 0.248   |
| +                                             | 6     | 12.2312±2.01359                                 |             |         |         |         |
| Grade                                         |       |                                                 |             |         |         |         |
| GI-II                                         | 23    | 12.6505±0.76988                                 | 1.493       | 0.147   | -0.214  | 0.256   |
| GIII-IV                                       | 7     | 12.0712±1.26499                                 |             |         |         |         |
| Vascular invasion                             |       |                                                 |             |         |         |         |
| No                                            | 199   | 12.8350±0.94514                                 | 2.149       | 0.032   | -0.122  | 0.032   |
| Yes                                           | 107   | 12.5905±0.95709                                 |             |         |         |         |
| Recurrence after treatment                    |       |                                                 |             |         |         |         |
| No                                            | 166   | 12.5805±0.98050                                 | -0.589      | 0.556   | 0.037   | 0.556   |
| Yes                                           | 94    | 12.6537±0.92925                                 |             |         |         |         |
| Survival status                               |       |                                                 |             |         |         |         |
| Dead                                          | 86    | 12.6050±1.01283                                 | -0.811      | 0.418   | 0.043   | 0.418   |
| Alive                                         | 275   | 12.7031±0.96776                                 |             |         |         |         |
hsa:207%09red) and pancreatic cancer (corrected P<0.05). In the protein-protein interaction (PPI) network for miR-23b-3p (Fig. 15), 338 target genes were identified to be involved in 1,074 nodes. A total of 8 targets, including SRC, AKT1, EGFR, CTNNB1, BCL2, SMAD3, PTEN and KDM6A, were located in the key nodes with high degree (>35).

Table II. Continued.

| Clinicopathological features | N   | miR-23b-3p relevant expression (2^{-ΔCq}) | Correlation |
|------------------------------|-----|------------------------------------------|-------------|
|                              |     | Mean ± SD | t-value | P-value | r-value | P-value |
| T status                     |     |            |         |         |         |         |
| T1                           | 177 | 12.7653±0.93707 | F=1.605\(^a\) | 0.202\(^a\) | -0.083 | 0.117 |
| T2-T4                        | 181 | 12.5926±1.01626 |           |         |         |         |
| TX                           | 1   | 12.0451±0.00000 |           |         |         |         |
| N status                     |     |            |         |         |         |         |
| N0                           | 247 | 12.7780±0.96867 | F=5.116\(^a\) | 0.006\(^a\) | -0.158 | 0.003 |
| N1                           | 3   | 11.7684±0.75393 |           |         |         |         |
| NX                           | 110 | 12.4733±0.96540 |           |         |         |         |
| M status                     |     |            |         |         |         |         |
| M0                           | 262 | 12.7389±0.96998 | F=1.778\(^a\) | 0.170\(^a\) | -0.101 | 0.055 |
| M1                           | 4   | 12.6180±0.12946 |           |         |         |         |
| MX                           | 95  | 12.5190±1.00762 |           |         |         |         |

\(^a\)One-way analysis of variance (ANOVA) test was utilized. t, Student’s t-test; SD, standard deviation. HCC, hepatocellular carcinoma; T, tumor; N, nodes; M, metastasis. TCGA, The Cancer Genome Atlas.
HE et al: miR-23b-3p, A NOVEL BIOMARKER FOR HEPATOCELLULAR CARCINOMA

Discussion

HCC is one of the most prevalent forms of liver cancer, and nearly one million cases of HCC occur annually. Thus, it is urgent to explore the mechanism of HCC tumorigenesis and progression and to determine treatment and prevention strategies. Exploiting a new self-assembled cell microarray, Zhang et al carried out high-throughput screening for miRNAs that are involved in cell transplantation. They discovered miR-23b-3p, which functioned as a tumor suppressor in human colon cancer (27). Since then, substantial research has studied the functions of miR-23b-3p in many physiological and pathological processes, especially tumor progression. As noted above, several studies showed that miR-23b-3p played a dual role in various cancers; these studies
also investigated the mechanisms of miR-23b-3p. This study focused on determining the role of miR-23b-3p in HCC and its effect on patient prognosis.

In the meta-analysis of data downloaded from GEO datasets, the pooled SMD indicated that miR-23b-3p expression showed no remarkable difference between HCC patients and normal subjects. No heterogeneity was noted. Additionally, based on GEO datasets, we investigated the association between miR-23b-3p and the clinicopathological aspects of HCC and observed that miR-23b-3p was significantly correlated with

| Term                                      | Count | P-value  | FDR   |
|-------------------------------------------|-------|----------|-------|
| Biological process                        |       |          |       |
| Regulation of transcription               | 111   | 3.87E-10 | 6.73E-07 |
| Regulation of transcription from RNA polymerase II promoter | 47    | 1.98E-09 | 3.44E-06 |
| Regulation of transcription, DNA-dependent | 83    | 2.93E-09 | 5.10E-06 |
| Regulation of RNA metabolic process       | 83    | 8.33E-09 | 1.45E-05 |
| Transcription                             | 90    | 3.70E-08 | 6.44E-05 |
| Positive regulation of transcription from RNA polymerase II promoter | 29    | 1.03E-07 | 1.80E-04 |
| Negative regulation of gene expression    | 34    | 2.12E-07 | 3.68E-04 |
| Positive regulation of gene expression    | 36    | 6.48E-07 | 0.001128 |
| Positive regulation of transcription      | 35    | 9.33E-07 | 0.001624 |
| Regulation of cell motion                 | 19    | 1.07E-06 | 0.001868 |
| Protein kinase cascade                    | 27    | 1.19E-06 | 0.002065 |
| Positive regulation of transcription, DNA-dependent | 31    | 1.79E-06 | 0.003114 |
| Positive regulation of RNA metabolic process | 31    | 2.12E-06 | 0.003697 |
| Positive regulation of nitrogen compound metabolic process | 37    | 2.57E-06 | 0.004477 |
| Regulation of cell proliferation          | 42    | 2.93E-06 | 0.005101 |
| Regulation of cell migration              | 17    | 3.52E-06 | 0.006119 |
| Positive regulation of macromolecule biosynthetic process | 37    | 3.65E-06 | 0.006356 |
| Positive regulation of cellular biosynthetic process | 38    | 4.10E-06 | 0.007132 |
| Positive regulation of macromolecule metabolic process | 44    | 4.31E-06 | 0.007507 |
| Regulation of locomotion                  | 18    | 4.36E-06 | 0.007589 |
| Response to hypoxia                       | 15    | 4.57E-06 | 0.007949 |
| Positive regulation of biosynthetic process | 38    | 5.68E-06 | 0.009878 |
| Negative regulation of transcription      | 29    | 7.16E-06 | 0.012463 |
| Response to oxygen levels                 | 15    | 8.29E-06 | 0.014432 |
| Positive regulation of nucleobase, nucleoside, nucleotide and nucleic acid metabolic process | 35    | 8.58E-06 | 0.014931 |
| Negative regulation of macromolecule biosynthetic process | 32    | 1.01E-05 | 0.01756 |
| Response to hormone stimulus              | 25    | 1.05E-05 | 0.018246 |
| Positive regulation of cell differentiation | 19    | 1.21E-05 | 0.021016 |
| Response to endogenous stimulus           | 26    | 1.84E-05 | 0.032008 |
| Embryonic organ development               | 16    | 1.91E-05 | 0.033319 |
| Cell morphogenesis                        | 24    | 1.96E-05 | 0.034027 |
| Positive regulation of cell motion        | 12    | 2.46E-05 | 0.042821 |
| Negative regulation of biosynthetic process | 32    | 2.51E-05 | 0.04366 |

| Term                                      | Count | P-value  | FDR   |
|-------------------------------------------|-------|----------|-------|
| Cellular component                        |       |          |       |
| Nuclear lumen                             | 54    | 1.14E-05 | 0.015418 |
| Nuclear envelope                          | 16    | 1.76E-05 | 0.023821 |

| Term                                      | Count | P-value  | FDR   |
|-------------------------------------------|-------|----------|-------|
| Molecular function                        |       |          |       |
| Transcription regulator activity          | 70    | 6.80E-07 | 9.94E-04 |
| DNA binding                               | 94    | 2.12E-06 | 0.0031  |
| Transcription factor activity             | 48    | 1.39E-05 | 0.020262 |

FDR, false discovery rate.
miR-23b-3p, A NOVEL BIOMARKER FOR HEPATOCELLULAR CARCINOMA

We identified miR-23b-3p as a HCC biomarker by analyzing TCGA data. miR-23b-3p expression was obviously down-regulated in HCC tissues compared to normal liver tissues. Additionally, miR-23b-3p expression was lower (P<0.05) in patients with hepatitis B, smoking history, vascular invasion and lymphatic metastasis. However, only hepatitis B history and lymphatic metastasis of HCC revealed diagnostic values. HCC patients with high miR-23b-3p expression presented longer survival; however, this finding was non-significant.

According to the results above, we propose that miR-23b-3p may play a crucial part in HCC tumorigenesis and invasion. We failed to prove the relevance of miR-23b-3p and other clinical parameters. Therefore, diagnosis and targeted therapy based on the miR-23b-3p expression level required further research.

Figure 11. Kaplan-Meier curve for recurrence-free survival in hepatocellular carcinoma (HCC) patients grouped by the level of miR-23b-3p expression. There was no significant association between miR-203 expression and recurrence free survival in patients with HCC (P=0.401). Error bars represented standard deviation (SD).

Figure 12. Meta-analysis of miR-23b-3p expression in hepatocellular carcinoma (HCC) patients and healthy controls retrieved from Gene Expression Omnibus (GEO) datasets, The Cancer Genome Atlas (TCGA) datasets and PCR data. (A) Forest plot. The center dot and the horizontal line represent the standard mean difference (SMD) and 95% confidence interval (CI), respectively. The pooled SMD and its corresponding 95% CI are presented below the list of studies; (B) Begg's funnel plot. Hazard ratios are presented on a logarithmic scale. (C) Egger's publication bias plot. Hazard ratios are presented on a logarithmic scale; (D) sensitivity analysis. Pooled SMDs and 95% CIs by eliminating each study.
development. Therefore, we recruited 101 fresh HCC tissues and counterpart adjacent non-cancerous HCC to inspect the expression of miR-23b-3p in HCC tissues and its association with the clinicopathological aspects and prognosis of HCC patients.

However, in contrast with the counterpart adjacent non-cancerous HCC, we observed that miR-23b-3p was aberrantly downregulated in HCC tissue (P<0.001). This consequence was supported by a statistical analysis of TCGA data but in contrast to the microarray meta-analysis. Additionally, HCC patients with multiple tumor nodes showed a lower level of miR-23b-3p expression than those with single tumor nodes (P<0.05). In addition to other parameters that indicated HCC progression, the deterioration of metastasis and the portal vein tumor embolus were strongly associated with a decrease in miR-23b-3p (P<0.001). In line with the aforementioned findings, miR-23b-3p was closely linked to HCC aggressiveness and metastasis. Additionally, Salvi et al transfected miR-23b-3p into HCC cells (SKHeplC3), which resulted in a reduction in motility and proliferative potential; they also found that miR-23b-3p was deregulated in HCC (28). To some extent, our study also revealed that increased miR-23b-3p was associated with an increased time-to-recurrence (months) in HCC patients. However, there was no obvious alteration
between the two groups concerning the levels of miR-23b-3p. The lack of a study population may explain this result. Thus, a larger cohort of patients is required to study this in the near future. Additionally, only tumor nodes, metastasis portal vein tumor embolus and EGFR expression of HCC were found to correlate with miR-23b-3p expression and presented diagnostic values.

Altogether, the data derived from GEO datasets, TCGA datasets and qPCR were included to perform a comprehensive meta-analysis. The pooled SMD indicated that pronounced downregulation of miR-23b-3p expression was noted in HCC patients as compared to healthy subjects, which indicated that low level of miR-23b-3p may play an important role in HCC tumorigenesis and diagnosis. Nevertheless, high heterogeneity indicated that this conclusion was less reliable. According to sensitivity analysis, the data from TCGA and qPCR contributed to the high heterogeneity. The difference of source of specimen and the detection methods of miR-23b-3p expression may be the major factors.

Ten prediction algorithms (utilizing different matching criteria and computational algorithms) were employed in miR-23b-3p target gene prediction. In contrast to the algorithms that provided validated targets and literature extraction, the prediction algorithms could not predict with accuracy. Some factors, such as varying approaches and rules for miRNA targeting, can affect the target prediction results. To reduce the defects associated with target prediction, we combined the results of these algorithms and stipulated that only the genes that appeared no fewer than five times would be involved in the next analysis.

In addition to targets based on experimental validation, prediction targets combined with algorithms were enrolled for GO analysis, pathways analysis and network interaction. A bioinformatics analysis predicted that miR-23b-3p was involved in extensive target regulation. Because these targets took part in multiple phases of tumorigenesis and tumor progression, we speculated that miR-23b-3p was dysregulated in various tumors and led to changes in biological characteristics by regulating a series of target genes in different phases. In a GO analysis, the targets of miR-23b-3p were involved in biological processes (like the regulation of transcription and transcription), cellular components (such as the nuclear lumen and nuclear envelope) and molecular functions (such as transcription regulator activity and DNA binding). These targets play a vital role in major cell processes, and a number of them participate in the carcinogenesis of various cancers, including HCC. According to KEGG analysis, miR-23b-3p participated in the hepatitis B pathway, which was recognized as a key factor of hepatic cirrhosis. Moreover, hepatic cirrhosis could result in hepatic carcinoma. We also found that miR-23b-3p was correlated with hepatitis B in the HCC patients who were included in TCGA. Thus, we speculated that miR-23b-3p may play an important role in HCC tumorigenesis.

A network analysis was ultimately conducted to predict the interaction networks of potential targets. Highly connected genes were regarded to play critical roles in stabilization, interaction, and gene network regulation. A connectivity analysis showed the highest connectivity between SRC and AKT1. SRC is a proto-oncogene encoding a tyrosine-protein kinase that belongs to Src family kinases, which are considered vital to cell proliferation, differentiation, apoptosis and invasion (29,30). Src family kinases have been shown to play a vital role in several malignancies, including breast cancer (30), colon cancer (31), ovarian cancer (32) and HCC (33), as well as melanoma, glioma, and various types of sarcoma (34). Akt1 or protein kinase B (PKB)α, which is regularly activated in human cancers, can phosphorylate downstream molecules and cope with a wide range of cell processes (35). Moreover, Akt1 was directly or indirectly controlled by Src kinase activity (35,36). We conjectured that Src kinase and Akt1 may function in the same signal transduction pathways that result in carcinogenesis and development. This suggests that
researchers should consider factors related to the hub genes for improving diagnosis and treatment.

Inquiries in various cancers indicated that miR-23b-3p, functioning as an oncogenic miRNA, can accelerate tumor processes such as proliferation and invasion. Jin et al reported that miR-23b/27b expression was elevated in breast cancer and that it indicated a poor prognosis (12). Nischarin partially participated in reducing the level of miR-23b/27b expression by inhibiting NF-κB phosphorylation and disturbing cancer aggressiveness in vivo (2). Additionally, the AKT/NF-κB pathway, which responds to TNFα and EGF, is the pivotal signaling cascade for Her2/neu-dependent miR-23b/27b in vitro (2). Li et al demonstrated that Fas mRNA, which enhances the cell multiplication and apoptosis rates, was directly silenced by miR-23b-3p upregulation in thymic lymphoma cells (37).

Many studies also noted that the role of miR-23b-3p as a tumor suppressor was downregulated in several cancers. Majid et al discovered that miR-23b-3p was downregulated in prostate cancer, and its decreasing function of directly inhibiting proto-oncogene Src kinase may lead to the acceleration of neoplasm growth (38). The same author in another study stated that miR-23b-3p played a key role in migration of bladder cancer cells via posttranscriptionally directly regulating at Zeb1, which could modulate the epithelial-to-mesenchymal transition (EMT) (39). All of the mechanisms by which miR-23b-3p acts as a tumor suppressor may contribute to HCC aggressiveness; further studies are required.

Furthermore, extensive study suggested that miR-23b-3p was associated with other clinicopathological features in HCC. Bisio et al confirmed the cis-mediated regulation of miR-23 by p53 in the MCF7 cell line (40). However, we
detected the reverse (upregulated miR-23b-3p) in p53-positive tissue (P>0.05). There were two possible reasons for this. First, miR-23b-3p served as oncogenic miRNA in the MCF7 cell line but acted as a tumor suppressor in HCC. Notably, the specific mechanism remained unclear. Secondly, our data was less trustworthy due to its lack of significance. Chen et al reported that the downregulated expression of miR-23b-3p inhibited HIF-1α/VEGF and β-catenin/Tcf4 signaling by enhancing the level of VHL (41). The quantity of VEGF in our study was also identical with miR-23b-3p expression; however, this was without significance (P>0.05). Patients with occult hepatitis virus B infection (OBI), who are generally negative for HBsAg, were discovered to have a differential expression of miR-23b-3p (42). Notably, as Salvi et al reported, miR-23b-3p overexpression contributed to a decrease in urokinase-type plasminogen (uPA) and c-met. This was determined by identifying the 3'-UTR of uPA and c-met in two and four sites, respectively, as well as by observing a reduction in cell proliferation and metastasis in HCC (28). The present study partially supports the above study. To our knowledge, the present study is also the first one to explore the clinicopathological significance of miR-23b-3p and contributed to a more comprehensive understanding of the correlation between miR-23b-3p and HCC progression.

Expression of miR-23b-3p was studied in HCC and their relationship explored. A preliminary analysis of the biological characteristics and function of miR-23b-3p was performed via bioinformatical methods. Despite existing limitations, miR-23b-3p was thought to be a tumor suppressor affecting the carcinogenesis and aggressiveness of HCC and may represent a predictive biomarker and therapeutic target for HCC. These results may contribute to a theoretical basis for HCC diagnosis and therapy.

Acknowledgements

The authors thank GEO and TCGA for the public available data.

Funding

This study was supported in part by the Natural Science Foundation of Guangxi, China (2015GXNSFFA139157 and 2017GXNSFAA198026) and Youth Science Foundation of Guangxi Medical University (WLXSZX18001).

Availability of data and material

All data generated or analyzed during this study are included in this published article.

Authors' contributions

RQH and PRW designed the study, did the experiments, analyzed and interpreted the data, and wrote the manuscript. They contributed equally to this work. XLX, XY and HWL performed the statistical analysis, designed and filled-out the figures and tables. XHQ, LHY, ZGP recruited the specimens, carried out the miRNA isolation and real-time RT-qPCR. LHY, ZGP and GC participated in designing the study, supervised all experiments and corrected the manuscript. All the authors read and approved the final manuscript.

Ethics approval and consent to participate

All experiments referred to patient tissues were authorized by the Ethics Committee of the First Affiliated Hospital of Guangxi Medical University (no. 2016-KY-NSFC-094). This study was a sub-study of the project mentioned in ethics approval. Method and participants are utilized in the same way.

Consent for publication

The consents for publication were obtained from all the patients.

Competing interests

The authors declare that they have no competing interests.

References

1. de Martel C, Maucourt-Boulech D, Plummer M and Franceschi S: World-wide relative contribution of hepatitis B and C viruses in hepatocellular carcinoma. Hepatology 62: 1190-1200, 2015.
2. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J and Jemal A: Global cancer statistics, 2012. CA Cancer J Clin 65: 87-108, 2015.
3. Yun EH, Lim MK, Oh JK, Park JH, Shin A, Sung J and Park EC: Combined effect of socioeconomic status, viral hepatitis, and lifestyle on hepatocellular carcinoma risk in Korea. Br J Cancer 103: 741-746, 2010.
4. M’Bengue AK, Dombia M, Denomaran SR, Ouattara DN, Adouabi I and Pineau D: A major shift of viral and nutritional risk factors affects the hepatocellular carcinoma risk among Ivorian patients: A preliminary report. Infect Agent Cancer 10: 18, 2015.
5. O’Harra SP, Mott JL, Splinter PL, Gores GJ and LaRusso NF: MicroRNAs: Key modulators of posttranscriptional gene expression. Gastroenterology 136: 17-25, 2009.
6. Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, Sweet-Cordero A, Ebert BL, Mak RH, Ferrando AA, et al: MicroRNA expression profiles classify human cancers. Nature 435: 834-838, 2005.
7. Francis P, Moon SY, Bilke S, Zhu YJ and Meltzer PS: Role of the microRNA-23-27 24 clusters in osteosarcoma. Cancer Res 72 (Suppl 8): 1113, 2012.
8. Goto Y, Nishikawa R, Kojima S, Sakamoto S, Kamakura K, Imamoto T, Chiyomaru T, Enokida H, Kinoshita T, Naya Y, et al: The functional significance and its regulated molecular targets of microrna-23b/27b/24-1 cluster in prostate cancer. J Urol 191: e456-e457, 2014.
9. Ishihara T, Chiyomaru T, Inoguchi S, Enokida H, Seki N and Nakagawa M: The clustered microRNA-23b-27b function as tumor suppressors and useful prognostic markers in renal cell carcinoma. J Urol 191: e243, 2014.
10. Jiang W, Min J, Sui X, Qian Y, Liu Y, Liu Z, Zhou H, Li X and Gong Y: MicroRNA-26a-5p and microRNA-23b-3p up-regulate peroxiredoxin III in acute myeloid leukemia. Leuk Lymphoma 56: 460-471, 2015.
11. Han L, Chen L, Zhang K, Shi Z, Zhang J, Zhang A, Wang Y, Song Y, Zheng Y, Jiang T, et al: MicroRNA-23b expression is regulated by VHL and effects on glioma cell survival and invasion. Cancer Res 72: 8, 2012.
12. Jin L, Wessel O, Marcussen EG, Ivan C, Calin GA and Alahari SK: Prooncogenic factors miR-23b and miR-27b are regulated by Her2/Neu, EGF, and TNF-α in breast cancer. Cancer Res 73: 2884-2896, 2013.
13. Ma G, Dai W, Song A, Yang X and Gao C: Upregulation of microRNA-23a/b promotes tumor progression and confers poor prognosis in patients with gastric cancer. Int J Clin Exp Pathol 7: 8833-8840, 2014.
14. Rogler CE, Levoci L, Ader T, Massimi A, Tchaikovskaya T, Norel R and Rogler LE: CE R: MicroRNA-23b cluster microRNAs regulate transforming growth factor-beta/bone morphogenetic protein signaling and liver stem cell differentiation by targeting Smads. Hepatology 50: 575-584, 2009.

15. Yuan B, Dong R, Shi D, Zhou Y, Zhao Y, Miao M and Jiao B: Down-regulation of miR-23b may contribute to activation of the TGF-β1/Smad3 signalling pathway during the termination stage of liver regeneration. FEBS Lett 585: 927-934, 2011.

16. Salvi A, Sabelli C, Moncini S, Venturin M, Arici B, Riva P, Portolani N, Giulini SM, De Petro G and Barlati S: MicroRNA-23b mediates urokinase and c-met downmodulation and a decreased migration of human hepatocellular carcinoma cells. FEBS J 276: 2966-2982, 2009.

17. Lau J, Ioannidis JP and Schmid CH: Quantitative synthesis in systematic reviews. Ann Intern Med 127: 820-826, 1997.

18. Higgins JP, Thompson SG, Deeks JJ and Altman DG: Measuring inconsistency in meta-analyses. BMJ 327: 557-560, 2003.

19. Zamora J, Abraira V, Muriel A, Khan K and Coomarasamy A: ing of miR-23b as a pleiotropic modulator suppressing cancer metastasis. Cell Mol Biol 82: 263-274, 2004.

20. Khan N: Meta-analysis: A quantitative approach of data pooling. Pak Oral Dental J 20: 214-221, 2000.

21. Badhu A, Roessler S, Zhao X, Yu Z, Forgues M, Ji J, Karody E, Qin LX, Ye QH, Jia HL, et al: Integrated metabolite and gene expression profiles identify lipid biomarkers associated with progression of hepatocellular carcinoma and patient outcomes. Gastroenterology 144: 1066-1075.e1, 2013.

22. Li W, He X, Li J, Tu K, Wei L, Wu J, Guo Y, Ma X, Zhang P, et al: Diagnostic and prognostic implications of microRNAs in human hepatocellular carcinoma. Int J Cancer 123: 1616-1622, 2008.

23. Sato F, Hatanou E, Kitamura K, Myamoto A, Fujitake T, Takizawa S, Tsujiya S, Usui K and Shimizu K: MicroRNA profile predicts recurrence after resection in patients with hepatocellular carcinoma within the Milan Criteria. PLoS One 6: e16435, 2011.

24. Su H, Yang JR, Xu T, Huang J, Xu L, Yuan Y and Zhuang SM: Down-regulation of miR-23b may contribute to activation of the TGF-β1/Smad3 signalling pathway during the termination stage of liver regeneration. FEBS Lett 585: 927-934, 2011.

25. Shen J, LeFave C, Iwaisako K, Ikeda K, Kawada N, Ochiya T and Taguchi YH: Comprehensive analysis of transcriptome and metabolome in metanephric adenoma with occult hepatitis B virus infection. Med Microbiol Immunol 201: 389-395, 2012.

26. Chen L, Han L, Zhang K, Shi Z, Zhang J, Zhang A, Wang Y, Song Y, Li Y, Jiang T, et al: VHL regulates the effects of miR-23b on glioma survival and invasion via suppression of HIF-1α/VEGF and β-catenin/Tcf-4 signaling. Neuror Oncol 14: 1026-1036, 2012.

27. Chen L, Han L, Zhang K, Shi Z, Zhang J, Zhang A, Wang Y, Song Y, Li Y, Jiang T, et al: VHL regulates the effects of miR-23b on glioma survival and invasion via suppression of HIF-1α/VEGF and β-catenin/Tcf-4 signaling. Neuro Oncol 14: 1026-1036, 2012.

28. Dehm SM and Bonham K: SRC gene expression in human cancer: The role of transcriptional activation. Biochem Cell Biol 82: 263-274, 2004.