Associations between Single-Nucleotide Polymorphisms in Corticotropin-Releasing Hormone-Related Genes and Irritable Bowel Syndrome

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Abstract

Irritable bowel syndrome (IBS) is a common functional disorder with distinct features of stress-related pathophysiology. A key mediator of the stress response is corticotropin-releasing hormone (CRH). Although some candidate genes have been identified in stress-related disorders, few studies have examined CRH-related gene polymorphisms. Therefore, we tested our hypothesis that single-nucleotide polymorphisms (SNPs) in CRH-related genes influence the features of IBS. Methods: In total, 253 individuals (123 men and 130 women) participated in this study. They comprised 111 IBS individuals and 142 healthy controls. The SNP genotypes in CRH (rs28364015 and rs6472258) and CRH-binding protein (CRH-BP) (rs10474485) were determined by direct sequencing and real-time polymerase chain reaction. The emotional states of the subjects were evaluated using the State-Trait Anxiety Inventory, Perceived Stress Scale, and the Self-rating Depression Scale. Results: Direct sequencing of the rs28364015 SNP of CRH revealed no genetic variation among the study subjects. There was no difference in the genotype distributions and allele frequencies of rs6472258 and rs10474485 between IBS individuals and controls. However, IBS subjects with diarrhea symptoms without the rs10474485 A allele showed a significantly higher emotional state score than carriers. Conclusions: These results suggest that the CRH and CRH-BP genes have no direct effect on IBS status. However, the CRH-BP SNP rs10474485 has some effect on IBS-related emotional abnormalities and resistance to psychosocial stress.

Introduction

Irritable bowel syndrome (IBS) is a widespread functional disorder of the lower gastrointestinal tract that is primarily characterized by abdominal pain and altered bowel habits (diarrhea and/or constipation) [1]. IBS is associated with impaired quality of life and excessive use of health care resources and has long been considered to be a stress-related disorder [2, 3]. Co-morbidity...
of psychological disorder and somatic co-morbidities have been reported in IBS [4, 5]. IBS patients reported more anxiety, depression, and perceived stress than healthy people [4]. The impact of stress and psychosocial factors on predisposition to IBS symptoms and their precipitation and maintenance highlights the importance of taking into consideration the effect of stress on the underlying molecular mechanisms [6, 7].

Some hypothalamic-pituitary-adrenal (HPA) axis-related genetic variation [8], including 11β-hydroxysteroid dehydrogenase type 1[9], glucocorticoid receptor [10], and mineralocorticoid receptor [11] gene polymorphisms, is known to be associated with the stress response. Corticotropin-releasing hormone (CRH), encoded by the CRH gene, is a key integrator of the stress response [12–14]. The response to stress in mammals is mediated through endocrine, autonomic, and behavioral systems via secretion of CRH by the paraventricular nucleus of the hypothalamus [15, 16]. This system is critical for survival but its chronic over-activity can lead to stress-related pathologies [17–19]. In this way, dysregulation of this system has been linked to a variety of stress-related disorders, including depression, post-traumatic stress disorder, suicidality, panic disorder, and alcohol consumption [20–27]. Components of the highly interrelated system of the HPA axis include not only CRH but also CRH receptors (CRH-R1/CRH-R2) and CRH-binding protein (CRH-BP) [14]. CRH-BP represents a passive ligand trap that binds to and neutralizes CRH, thereby terminating its biological actions, in contrast to its active receptor that initiates signal transduction on binding [28]. Furthermore, CRH-related peptides are expressed not only in the brain but also within the colon, where they activate enteric, endocrine, and immune cells and may be involved in colonic manifestations of IBS [1].

Recently, some candidate gene studies have identified an association between gene polymorphisms and IBS [29–32]. Most of the studies investigated the association of IBS with genetic variations in the serotonin (5-hydroxytryptamine) signaling system, immune system, or adhesion molecules [29–31]. Single-nucleotide polymorphisms (SNPs) in interleukin-10 (IL-10) and hydroxytryptamine receptor 2A were associated with diarrhea-predominant IBS in female patients [30]. IL-6, cadherin-1, and toll-like receptor-9 genes were associated with post-infectious IBS [29]. However, only a few gene studies have found positive evidence of CRH-related gene polymorphisms associated with IBS [32]. Polymorphisms in CRH-related peptide CRH-R1 and CRH-R1 haplotypes were suggested to moderate IBS and IBS bowel patterns in a previous study [32]. Although CRH-related peptides are clearly important in IBS, no study has clarified the role of gene polymorphisms in CRH and CRH-BP in IBS. Earlier studies suggested an association of CRH-BP gene polymorphisms with the antidepressant treatment response, major depressive disorder, and suicidal behavior in schizophrenia [33–35]. In rhesus macaques, a polymorphism in the CRH promoter region increases the risk of disorder in stress-exposed individuals [22].

Thus, the aim of this study was to investigate the associations between genetic variants of the CRH system and IBS. We had the following two hypotheses: 1) SNPs in the CRH and CRH-BP genes are different between IBS patients and healthy people; and 2) these selected SNPs in the CRH and CRH-BP genes influence the IBS endophenotype (bowel movement patterns and psychological abnormalities).

**Materials and Methods**

**Subjects**

The characteristics of the subjects are shown in Table 1. In total, 253 volunteers (130 women and 123 men) aged 22.0 ± 2.2 years (mean ± standard deviation [SD]) participated in the current study. They were all Japanese individuals recruited from universities in Sendai, Japan. IBS patients were diagnosed according to the Rome III criteria [36]. IBS patients were classified by
their predominant bowel pattern: constipation, diarrhea, or mixed. There was no significant difference in age, sex ratio, and IBS status between the groups. This study was approved by the Ethics Committee of Tohoku University Graduate School of Medicine. All participants gave their written informed consent.

Among the 253 participants, 190 were included in a study of the polymorphisms of the 5-hydroxytryptamine transporter [37] and CRH-R1 polymorphic region [32]. However, those genes were completely different from the genes targeted in this study. The hypotheses in this study and the previous studies are different. The previous analysis does not impact on this study.

### Psychometric Tests

The emotional states of the subjects were rated using the Self-rating Depression Scale (SDS) [38, 39], Perceived Stress Scale (PSS) [40, 41], and State-Trait Anxiety Inventory (STAI) [42, 43]. The SDS is a rating of affective, psychological, and somatic symptoms associated with depression, the PSS measures the perception of stress, and the STAI is a rating of two types of anxiety: state anxiety and trait anxiety.

### Genotyping

Whole blood (10mL) was sampled from the median cubital vein of all subjects. DNA was extracted from the white blood cells using a standard protocol [32]. SNPs in CRH (rs28364015 and rs6472258) [22, 33] and CRH-BP (rs10474485) [33] were selected by considering previous reports, and minor allele frequencies were determined from a database [NCBI: http://www.ncbi.nlm.nih.gov/snp/]. CRH is encoded by the CRH gene, which is located on chromosome 8 and spans 2.34 kb. rs28364015 and rs6472258 are also located on chromosome 8 (position 67253453 and 67259033, respectively). CRH rs28364015 is a 5’ untranslated region variant, and rs6472258 is an intergenic variant. Further, CRH-BP is encoded by the CRH-BP gene, which is located on chromosome 5 and spans 16.76 kb. CRH-BP rs10474485 is located on chromosome 5: (position 76306609). rs10474485 is also intronic and is a non-coding transcript variant.

The genotypes of the CRH and CRH-BP polymorphisms were determined by real-time polymerase chain reaction (PCR) (TaqMan assay). DNA samples from 40 participants were amplified by direct PCR to determine the positive controls. A 50-μL PCR reaction consisted of 0.2 μM of each primer, 1.25 U Primer STAR HS DNA polymerase, 200 μM deoxynucleoside triphosphate, and 1× Prime STAR buffer. After initial denaturation at 94°C for 4 min, amplification was performed using 35 cycles at 94°C for 1 min (denaturation), 60°C for 1 min (annealing), and 72°C for 1.5 min (elongation), followed by a final elongation step at 72°C for 7 min.
Then, the amplified PCR products were purified from a 2% agarose gel using a QIAquick Gel Extraction Kit (QIAGEN, Hilden, Germany). Amplimers were directly sequenced using an ABI PRISM dRhodamine™ Terminator Cycle Sequencing Ready Reaction Kit (PE Applied Biosystems, Foster City, CA), and excess dye terminators were removed using CENTRI-SEP columns (Princeton Separations, Adelphia, NJ). Automated sequencing was performed on an ABI 3130 Genetic Analyzer (PE Applied Biosystems). All procedures were carried out according to the instructions of the manufacturers.

Forward and reverse primers were used to sequence the PCR products. Using Primer3 Input version 0.4.0 (http://frodo.wi.mit.edu/primer3/), two CRH SNP (rs28364015 and rs6472258) and one CRH-BP SNP (rs10474485) oligonucleotide primer sets were designed. The forward primers were 5'-GCA GAA AGA TGG TGG GAC TC-3' (rs28364015), 5'-AGG AGA ATC GCT TGA ACC TG-3' (rs6472258), and 5'-GCA CCC AAA AGA GAG TTG TG-3' (rs10474485). The reverse primers were 5'-TCT CTT GAC AGC TCG ATT GC-3' (rs28364015), 5'-CTG GAT TGA ATT CCC TGT CC-3' (rs6472258), and 5'-GGA GAG TCA ACA GGG GAA TTG-3' (rs10474485).

Similarly, rs6472258 and rs10474485 TaqMan probe sets were designed, and the participants were genotyped by real-time PCR. However, the TaqMan assay failed for rs28364015 because of the frequency of the C allele (0%). The PCR reactions consisted of 200 nM of each primer, 200 nM fluorogenic probe, 1× SsoAdvanced Probes Supermix, and distilled water (up to 10 μL). Automated sequencing was performed on a CFX 96™ Real-Time PCR Detection System (Bio-Rad Laboratories, Inc., Tokyo, Japan).

Statistical Analysis

We used Haploview [44] to confirm the Hardy-Weinberg equilibrium. The effects of the genotypes or alleles of each SNP were compared between the IBS patients and controls using the χ² test and Odds Ratio (OR) (95% confidence interval [CI]). Two-way analysis of variance with a post hoc test was performed to determine the association between the SNPs and the psychometric tests. Results are expressed as the mean ± SD. We considered p < 0.05 to be statistically significant, whereas a p-value < 0.1 was considered to be tendentially but not statistically significant. Statistical analyses were performed using SPSS Statistic 22.0 (IBM, Inc., New York, NY).

Results

The genotype frequencies of the selected CRH and CRH-BP gene variations in the IBS patients and controls are shown in Table 2. There was no significant difference in the genotype frequencies of rs6472258 (p = 0.990, χ² test) and rs10474485 (p = 0.969, χ² test) between the patients and controls. Similarly, there was no significant difference in the allele frequencies of the rs6472258 T allele (p = 0.675, χ² test) or G allele (p = 0.890, χ² test), and rs10474485 C allele (p = 0.816, χ² test) or A allele (p = 0.986, χ² test) between the IBS patients and controls. Moreover, the results of the additional analysis (OR [95% CI]) in rs6472258, alleles are not associated with IBS, with an OR of 1.036 (95% CI, 0.629–1.71) for G allele and 1.015 (95% CI, 0.524–1.97) for T allele. Similarly, the allelic OR (95% CI) of rs10474485 were 1.000 (95% CI, 0.976–1.025) for A allele and 0.914 (95% CI, 0.427–1.956). There was no difference in statistical results between genotype or allele frequency and OR.

The psychometric scores of the IBS patients and controls are shown in Table 3. For the SDS, the score of the IBS patients was higher than that of the controls (p = 0.016). In addition, for the PSS, the score of the IBS patients was higher than that of the controls (p = 0.028). The STAI score was not significantly different between the two groups (State, p = 0.111; Trait, p = 0.115).
In the analysis of all subjects (IBS patients + controls), rs10474485 A allele non-carriers showed a higher PSS score than carriers (F$[1, 507] = 6.459$, p = 0.012). There was a significant group (IBS vs. control) effect with the SDS (p = 0.014) and PSS (p = 0.030) scores. The mean (± SD) PSS score of A allele carriers was 25.56 (± 8.359), whereas that of non-carriers was 28.48 (± 9.303). The other psychometric scores were not significantly different (SDS, p = 0.112; State, p = 0.140; Trait, p = 0.143). In contrast, we analyzed the associations between rs6472258 (genotype and allele) and the psychometric scores, but there was no significant difference.

Next, we analyzed IBS symptoms. In individuals with diarrhea symptoms, there was a significant difference in the PSS (F$[1, 363] = 6.186$, p = 0.018) and Trait (F$[1, 589] = 6.192$, p = 0.017) scores between rs10474485 A allele non-carriers and carriers (Fig 1A). The mean (± SD) PSS scores of A allele carriers vs. non-carriers were 24.75 (± 8.234) vs. 30.96 (± 7.252), and the mean Trait scores were 44.19 (± 6.804) vs. 52.09 (± 11.329), respectively. Also, in those with mixed symptoms, the mean (± SD) SDS score of rs10474485 A allele carriers was 39.75 (± 5.802), whereas that of non-carriers was 45.00 (± 6.585). This difference was significant (F$[1, 207] = 5.193$, p = 0.030) (Fig 1B). There was no significant difference for those with constipation symptoms. Moreover, the associations between rs6472258 (genotype and allele) and IBS subtypes were not significantly different.

In addition, in analysis of the male subjects alone, significant group (IBS/control) × rs10474485 genotype (F$[2, 167] = 3.180$, p = 0.045) (Fig 2A) and group (IBS/control) × rs10474485 A allele (F$[1, 309] = 5.821$, p = 0.017) (Fig 2B) interaction were observed in the SDS score. However, there was no difference in the emotional scores of the male subjects with respect to the A allele (Fig 3A). In the female subjects, significant differences were observed in the PSS (F$[1,360] = 4.931$, p = 0.028) and State (F$[1, 520] = 4.557$, p = 0.035) scores with respect

| dbSNP rs#  | Genotype | IBS (%) | Control (%) | p value |
|----------|----------|---------|------------|--------|
|          | Male (n = 47) | Female (n = 64) | Total (n = 111) | Male (n = 76) | Female (n = 66) | Total (n = 142) |
| rs6472258 | GG       | 9 (8.1) | 10 (9.0) | 15 (10.6) | 9 (6.3) | 0.990 |
|          | GT       | 12 (10.8) | 20 (18.0) | 21 (14.8) | 19 (13.4) | 0.969 |
|          | TT       | 26 (23.4) | 34 (30.6) | 40 (28.2) | 38 (26.8) | 0.969 |
| rs10474485 | AC   | 7 (6.3) | 6 (5.4) | 7 (4.9) | 11 (7.7) | 0.990 |
|          | CC       | 24 (21.6) | 41 (36.9) | 48 (33.8) | 35 (24.6) | 0.990 |

Abbreviations: dbSNP, Single Nucleotide Polymorphism Database; IBS, irritable bowel syndrome.

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Table 3. Psychometric Scores.

| Psychometric score, mean [SD] | IBS | Control | p value |
|-------------------------------|-----|---------|--------|
|                               | Male (n = 47) | Female (n = 64) | Total (n = 111) | Male (n = 76) | Female (n = 66) | Total (n = 142) |
| SDS                           | 40.2 [7.4] | 42.8 [6.9] | 41.7 [7.2] | 37.9 [7.4] | 40.8 [9.5] | 39.2 [8.6] | 0.016 |
| PSS                           | 27.4 [8.5] | 29.6 [7.9] | 28.7 [8.2] | 24.4 [9.3] | 28.2 [9.3] | 26.1 [9.5] | 0.028 |
| STAI (State)                  | 47.1 [9.0] | 44.8 [9.6] | 45.8 [9.4] | 42.6 [7.7] | 45.3 [11.8] | 43.9 [9.9] | 0.111 |
| STAI (Trait)                  | 47.8 [11.0] | 49.5 [10.0] | 48.8 [10.4] | 46.1 [9.2] | 47.2 [12.7] | 46.6 [10.9] | 0.115 |

Abbreviations: IBS, irritable bowel syndrome; SD, standard deviation; SDS, Self-rating Depression Scale; PSS, Perceived Stress Scale; STAI, State-Trait Anxiety Inventory.

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to the rs10474485 A allele (Fig 3B). The mean (± SD) PSS score of female A allele carriers was 26.87 (± 7.983), whereas that of non-carriers was 30.42 (± 8.843). The mean (± SD) State score of A allele carriers was 42.74 (± 10.091), whereas that of non-carriers was 46.72 (± 10.962). Analyses of rs28364015 did not indicate a significant difference in males or females.

As for the CRH SNP rs6472258, there was a nonsignificant trend for a difference in the SDS score between G allele carriers and non-carriers (all subjects, F[1, 244] = 3.837, p = 0.051; male *p < 0.05.

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**rs10474485 (Male)**

- **(a)** In male subjects, a significant group (IBS/control) × rs10474485 genotype interaction (p = 0.045) was observed. (b) Further, there was a significant group (IBS/control) × rs10474485 A allele interaction (p = 0.017). *p < 0.05.

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**rs10474485 (Female)**

- **(a)** In female subjects, a significant group (IBS/control) × rs10474485 genotype interaction (p = 0.047) was observed. (b) Further, there was a significant group (IBS/control) × rs10474485 A allele interaction (p = 0.017). *p < 0.05.

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**rs10474485 (All Sexes)**

- **(a)** In both sexes, a significant group (IBS/control) × rs10474485 genotype interaction (p = 0.044) was observed. (b) Further, there was a significant group (IBS/control) × rs10474485 A allele interaction (p = 0.017). *p < 0.05.
subjects, $F[1, 171] = 3.180, p = 0.077$). In the present study, all subjects ($n = 40$) whose DNA was analyzed with direct sequencing had the rs28364015 TT genotype and no subjects had the C allele. Therefore, we excluded rs28364015 from the analysis of the real-time PCR data.

**Discussion**

In the current study, we analyzed CRH-related peptide genetic variations in controls and patients with IBS, which is one of the representative disorders with stress-related pathophysiology. On the basis of previous studies and reports of the sexually dimorphic expression of CRH-related genes [45–48], sex differences would be expected in our study. Indeed, in the present study, the CRH-BP SNP rs10474485 had a significant effect on the PSS score, especially in female subjects. In addition, there was a significant gene × group interaction in the SDS score of males. It has been suggested that sex differences are important when considering genetic influences on IBS. Some previous studies suggested that the functions of hormone and genetic variations were connected to sex [45–49]. For example, a role for 5-hydroxytryptamine receptor 2B genotypes in impulsivity was shown in male subjects [45], and transcriptional regulation of the human CRH-BP gene promoter by estrogen receptors was reported [49]. Similarly, it is thought that the association between differences in sex hormones and gene polymorphisms are important. As CRH plays a dominant role in the HPA axis, the effect of sex differences on hormone and hypothalamic function must be investigated. For the CRH SNP rs6472258, a tendency for a difference in the SDS score between G allele carriers and non-carriers was found, which may have influenced the differences in the SDS score in male subjects. Significant results might have been observed with more subjects.

Some studies have suggested that CRH-related gene variations are not associated with IBS per se, but that the IBS subgroup or patients’ comorbid anxiety is associated with polymorphisms [50–52]. IBS subjects with diarrhea showed a higher response (namely, more inhibition) than healthy controls to a CRH receptor antagonist [53]. Therefore, IBS subjects with diarrhea might be more responsive to CRH or CRH-related peptides than healthy controls.
Here, the IBS subjects with diarrhea symptoms (diarrhea or a mix) showed significant differences in some emotion states according to the presence of the rs10474485 SNP. It is suggested that these CRH-BP polymorphisms might more strongly influence IBS patients with diarrhea symptoms. CRH is a major mediator to activate HPA axis in stress response. In contrast, CRH-BP reduces the effect of CRH [28]. If rs10474485 A allele make the CRH-BP effects increase, CRH function may be more interfered with. Thereby, it is suggested that HPA axis activity attenuated, and stress response decreased. As a result, it is thought that psychometric scores might decrease when individuals have A allele.

In the present study, the CRH G and CRH-BP A alleles were minor alleles. According to previous studies of stress-related disorders, CRH-related gene-specific polymorphisms appeared to protect against adult depressive symptoms [54, 55]. Subjects without the minor alleles of the selected SNPs showed higher psychometric scores than minor allele carriers. CRH exaggerates various stress responses, such as anxiety. Conversely, human CRH-BP binds to CRH and prevent activation of CRH receptors, thereby, reduce high activity with CRH [56]. In rodents, CRH-BP cells are concentrated within a hypothalamic circuit involved in mediating neuroendocrine and autonomic responses to stress [57]. CRH-BP expression downregulates the HPA axis by interfering with CRH, and the stress responses converge [28].

Here, CRH-BP rs10474485 was not directly related to IBS. However, the selected CRH-BP polymorphism was found to be in linkage disequilibrium with the CRH-BP SNPs rs10055255 (r² = 0.410, D’ = 0.928) and rs1875999 (r² = 0.410, D’ = 0.928) in the 1000 Genomes Project of Japanese individuals in Tokyo [the 1000 Genomes Browsers: http://browser.1000genomes.org/]. Homozygosity of the rs10055255 minor allele (TT) was associated with fewer incidences of post-traumatic stress disorder and depressive symptoms in post-intensive care units [58]. Under a condition of stress manipulation, individuals homozygous for the rs10055255 minor allele (TT) were noted to show high stress-induced alcohol craving, tension, and negative mood compared with their pre-stress condition. During the neutral imagery condition, there was no effect, but the post-stress rating was lower than that of pre-stress [59]. Thus, this SNP (rs10055255) may only show the differential effects of stress responses under a stressful situation. However, because our study was not conducted under stress conditions, the rs10055255 SNP was not selected. Nonetheless, it is suggested that CRH-BP polymorphisms influence stress responses during stress manipulation. In addition, the rs1875999 major allele (T) was more common in unipolar patients [60], and its minor allele (C) was associated with both heroin and cocaine addiction [61]. At present, the functionality of rs1875999 is unknown and the directionality of the role of the alleles in the stress response is not consistent. Accordingly, we did not select this SNP in this study. However, it is thought that this SNP has an inhibitory role in the development of stress-related neuropsychiatric disorders. Therefore, it is conceivable that a functional increase may occur in the CRH-BP produced by the CRH-BP rs10474485 minor allele and that the activity of the HPA axis may decrease. In other words, there may be protective effects of minor CRH/CRH-BP gene variants on the stress response.

Although identified in the rhesus monkey [22], the genetic variant of the rs28364015 SNP has not been identified in the Japanese population. The present study is the first to try to identify this polymorphism in Japanese individuals. However, the results were negative. Regardless of the limited number of subjects (40 individuals), the possibility of finding this variation is considered to be very low.

There are some limitations to this study. First, we did not have a sufficient number of samples to investigate SNPs. A future direction would be to collect more samples and to explore more SNPs in CRH, CRH-BP, CRH-R1, and CRH-R2. Moreover, because the average age of all subjects was low (22.0 ± 2.2 years), it is thought that the present data could not be extrapolated to the entire IBS population. However, a meta-analysis study showed that there was not
significant difference in the prevalence of IBS between different age bands [62]. Therefore, it is thought that the subjects used in the present study were representative as an IBS group. Second, IBS severity was not explored. In the future, the association between disease severity and polymorphisms should be explored. Third, the rs6472258 and rs10474485 SNPs selected in this study are intronic. The rs6472258 SNP is located upstream of CRH gene exon 1. Similarly, rs10474485 is located downstream of CRH-BP gene exon 7 (Ensembl Genome Browser: http://www.asia.ensembl.org/). The effects of these areas on the transcriptional activity and protein expression of the CRH/CRH-BP genes are not clear. However, in a previous study, CRH receptor-1 SNPs located in the intron around exon 2 were associated with IBS [32]. Therefore, there may be a possibility that intronic SNPs can regulate the function of the CRH/CRH-BP genes. In any future study, the role of CRH genes in the endophenotype of IBS should be explored.

In conclusion, our study suggests that CRH-BP gene polymorphisms had some effect on IBS-related psychological abnormalities. In contrast, we could not detect the associations between the CRH/CRH-BP genes and IBS status per se. Further evidence for a general role of CRH and CRH-BP gene polymorphisms in stress-related disorders is required.

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Author Contributions

Conceived and designed the experiments: SF. Performed the experiments: AS. Analyzed the data: AS N. Sato N. Suzuki MA. Wrote the paper: AS SF. Screened the subjects and took samples: M. Kano YT M. Kanazawa.

References

1. Fukudo S. Role of corticotropin-releasing hormone in irritable bowel syndrome and intestinal inflammation. J Gastroenterol 2007; 42 (Suppl 17): 48–51. PMID: 17238026
2. Talley NJ. Functional gastrointestinal disorder as a public health problem. Neurogastroenterol Motil 2008; 20 (Suppl 1): 121–129. doi: 10.1111/j.1365-2982.2008.01097.x PMID: 18402649
3. Whitehead WE. Assessing the effects of stress on physical symptoms. Health Psychol 1994; 13: 99–102. PMID: 8020462
4. Kanazawa M, Endo Y, Whitehead WE, Kano M, Hongo M, Fukudo S. Patients and nonconsulters with irritable bowel syndrome reporting a parental history of bowel problems have more impaired psychological distress. Dig Dis Sci 2004; 49: 1046–53. PMID: 15309899
5. Levy RL, Olden KW, Naliboff BD, Bradley LA, Franciscioni C, Drossman DA, et al. Psychosocial aspects of the functional gastrointestinal disorders. Gastroenterology 2006; 130: 1447–58. PMID: 16678558
6. Grover M, Drossman DA. Psychotropic agents in functional gastrointestinal disorders. Curr Opin Pharmacol 2008; 8: 715–723. doi: 10.1016/j.coph.2008.07.012 PMID: 18760380
7. Riedle A, Schmidtmann M, Stengel A, Goebel M, Wisser AS, Klapp BF, et al. Somatic comorbidities of irritable bowel syndrome: a systematic analysis. J Psychosom Res 2008; 64: 573–582. doi: 10.1016/j.jpsychores.2008.02.021 PMID: 18501287
8. Tsartsali L, Papadopoulous M, Lagona E, Papadimitriou A, Kanaka-Gantenbein C, Louizou E, et al. Association of hypothalamic-pituitary-adrenal axis-related polymorphisms with stress in asthmatic children on inhaled corticosteroids. Neuroimmunomodulation 2012; 19: 88–95.
9. Dekker M, J. H. J, Tiemeier H, Luijendijk H, J, Kunungas M, Hofman A, de Jong F. H, et al. The Effect of Common Genetic Variation in 11Β-Hydroxysteroid Dehydrogenase Type 1 on hypothalamic-pituitary-
10. Aleksandra S, Anna LR, Joanna P, Aleksandra RR, Monika DW, Monika W, et al. Glucocorticoid receptor polymorphisms is associated with major depression and predominance of depression in the course of bipolar disorder. Journal of Affective Disorders 2011; 134: 138–144. doi: 10.1016/j.jad.2011.06.020 PMID: 21764460

11. Bogdan R, Perlis RH, Fagerness J, Pizzagalli DA. The impact of mineralocorticoid receptor ISO/VAL genotype (rs5522) and stress on rewaris learning. Genes Brain Behav 2010; 9(6): 658–667. doi: 10.1111/j.1601-183X.2010.00600.x PMID: 20528958

12. Vale W, Spies J, Rivier J. Characterization of a 41-residue ovine hypothalamic peptide that stimulates secretion of corticotrophin and beta-endophin. Science 1981; 213: 1394–1397. PMID:6267699

13. Potter E, Sutton S, Donaldson C, Chen R, Perrin M, Lewis K, et al. Distribution of corticotrophin-releasing factor mRNA expression in the rat brain and pituitary. Proc Natl Acad Sci U S A 1994; 91: 8777–8781. PMID: 8090722

14. Behan DP, De Souza EB, Lowry PJ, Potter E, Sawchenko P, Vale W. Corticotropin releasing factor (CRF) binding protein: a novel regulator if CRF and related peptide. Front Neuroendocrinol 1995; 16:362–382. PMID: 8557170

15. Owens MJ, Nemeroff CB. Physiology and pharmacology of corticotropinreleasing factor. Pharmacol Rev 1991; 43: 425–473. PMID: 1775506

16. Costa SC, Kesterson RA, Heldwein KA, Stevens SL, Heard AD, Hollis JH, et al. Abnormal adaptations to stress and impaired cardiovascular function in mice lacking corticotropin-releasing hormone receptor-2. Nat Genet 2000; 24: 403–409. PMID: 10742107

17. Korte SM, Koolhaas JM, Wingfield JC, McEwen BS. The Darwinian concept of stress: Benefits of allostasis and costs of allostatic load and the trade-offs in health and disease. Neurosci Biobehav Rev 2005; 29: 3–38. PMID: 15652252

18. McEwen BS. Protective and damaging effects of stress mediators: Central role of the brain. Dialogues Clin Neurosci 2006; 8: 367–381. PMID: 17290796

19. Sapolsky RM. in Coping with the Environment: Neural and Endocrine Mechanisms, eds McEwen BS, Goodman HM (Oxford Univ Press, New York) 2001; pp 517–532.

20. de Kloet CS, Vermetten E, Geuze E, Lentjes EG, Heijnen CJ, Stalla GK, et al. Elevated plasma corticotrophin-releasing hormone levels in veterans with posttraumatic stress disorder. Prog Brain Res 2008; 167:287–291. PMID: 18037027

21. Southwick SM, Vythilingam M, Charney DS. The psychobiology of depression and resilience to stress: Implications for prevention and treatment. Annu Rev Clin Psychol 2005; 1:255–291. PMID: 17716089

22. Barr CS, Dvoskin RL, Gupte M, Sommer W, Sun H, Schwandt ML, et al. Functional CRH variation increases stress-induced alcohol consumption in primates. Proc Natl Acad Sci U S A 2009; 106 (34):14539–14598. PMID: 16815632

23. Liu Z, Zhu F, Wang G, Xiao Z, Wang H, Tang J, et al. Association of corticotropin-releasing hormone receptor1 gene SNP and haplotype with major depression. Neuroscience Letters 2006; 404: 358–362. PMID: 16815632

24. Licinio J, O’Kirwan F, Irizarry K, Merriman B, Thakur S, Jepson R, et al. Association of a corticotropin-releasing hormone receptor 1 haplotype and antidepressant treatment response in Mexican-Americans. Mol Psychiatry 2004; 9(12):1075–1082. PMID: 15365580

25. Wasserman D, Sokolowski M, Rozanov V, Wasserman J. The CRHR1 gene: a marker for suicidality in depressed males exposed to low stress. Genes Brain Behav 2008; 7(1):14–19. PMID: 17376150

26. Keck ME, Kern N, Erhardt A, Unschuld PG, Ising M, Salyakina D, et al. Combined effects of exonic polymorphisms in CRHR1 and AVPR1B genes in a case/control study for panic disorder. Am J Med Genet B Neuropsychiatr Genet 2008; 147B(7):1196–1204. doi: 10.1002/ajmg.b.30750 PMID: 18384079

27. Enoch MA, Shen PH, Ducci F, Yuan Q, Liu J, White KV, et al. Common genetic origins for EEG, alcoholism and anxiety: the role of CRH-BP. PLoS One 2008; 3(10):e3620. doi: 10.1371/journal.pone.0003620 PMID: 18974851

28. Jahn O, Radulovic J, Stiedl O, Tezval H, Eckart K, Spies S. Corticotropin-releasing factor binding protein: a ligand trap? Mini Rev Med Chem 2005; 5:953–960. PMID: 16250837

29. Villani A, Saito Y, Lemire M, Lemire M, Thabane M, Larson J, et al. Validation of genetic risk factors for post-infectious irritable bowel syndrome (IBS) in patients with sporadic IBS. Gastroenterology 2009; 296:G211–G218.
30. Kapeller J, Houghton L, Walsteb J, Boenisch H, Rappold G, Niesler B. A coding variant in the serotonin receptor 3C subunit is associated with diarrhea-predominant irritable bowel syndrome. Gastroenterology 2009; 136: A155–A156.

31. van der Veek PPJ, de Kroon YE, van den Berg M, Verspaget HW, Masclée AAM. Tumor necrosis factor-alpha and interleukin 10 gene polymorphisms in irritable bowel syndrome. Eur J Gastroenterol Hepatol 2005; 17(1): A26.

32. Sato N, Suzuki N, Sasaki A, Aizawa E, Obayashi T, Motoyori K, et al. Corticotropin-Releasing Hormone Receptor 1 Gene Variants in Irritable Bowel Syndrome. PLoS ONE 2012; 7(9): e42450. doi: 10.1371/journal.pone.0042450 PMID: 22957021

33. Binder EB, Owens MJ, Liu W, Deveau TC, Rush AJ, Trivedi MH, et al. Association of polymorphisms in genes regulating the corticotropin-releasing factor system with antidepressant treatment response. Arch Gen Psychiatry 2010; 67(4): 369–379. doi: 10.1001/archgenpsychiatry.2010.18 PMID: 20368512

34. Van Den Eede F, Venken T, Del Favero J, Norrback KF, Souery D, Nilsson LG, et al. Single nucleotide polymorphisms analysis of corticotropin-releasing factor-binding protein gene in recurrent major depressive disorder. Psychiatry Research 2007; 157:17–25.

35. De Luca V, Tharmalingam S, Zai C, Potapova N, Strauss J, Vincent J, et al. Association of HPA axis genes with suicidal behaviour in schizophrenia. Journal of Psychopharmacology 2010; 24(5):677–682. doi: 10.1177/0269881108097817 PMID: 18838498

36. Longstreth GF, Thompson WG, Chey WD, Houghton LA, Mearin F, Spiller RC. Functional bowel disorder. Gastroenterology 2006; 130: 1480–1491. PMID: 16678561

37. Fukudo S, Kanazawa M, Mizuno T, Hamaguchi T, Kano M, Watanabe S, et al. Impact of serotonin transporter gene polymorphisms on brain activation by colorectal distention. Neuroimage 2009; 47: 946–951. doi: 10.1016/j.neuroimage.2009.04.083 PMID: 19426812

38. Zung W. W. K. A self-rating depression scale. Arch Gen Psychiat 1965; 12: 63–70. PMID: 14221692

39. Fukuda K, Kobayashi S. A study on a self-rating depression scale. Seishin Shinkeigaku Zassi 1973; 75: 673–679.

40. Cohen S, Kamarck T, Mermelstein R. A global measure of perceived stress. J Health Soc Behav 1983; 24: 385–396. PMID: 668417

41. Iwahashi S, Tanaka Y, Fukudo S, Hongo M. The development of the Japanese Version of the Perceived Stress Scale. Shin-shin-Igaku (Jpn J Psychosom Med) 2002; 42: 459–466.

42. Spielberger CD, Gorsuch RL, Lushene RE. In: STAI manual. Palo Alto, California: Consulting Psychologist Press; 1970. p. 23–49.

43. Nakazato K, Shimokawa Y. The Japanese State-Trait Anxiety Inventory: age and sex differences. Percept Mot Skills 1989; 69: 611–617. PMID: 2813009

44. Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics 2005; 21(2): 263–265. PMID: 15297300

45. Bevilaqua L, Doly S, Kaprio J, Yuan Q, Tikkanen R, Paunio T, et al. A population-specific HTR2B stop codon predisposes to severe impulsivity. Nature 2011; 470(7334): 424.

46. Sánchez-Martín JR, Azurmendi A, Pascual-Sagastizabal E, Cardas J, Braza F, Braza P, et al. Androgen levels and anger and impulsivity measures as predictors of physical, verbal and indirect aggression in boys and girls. Psychoneuroendocrinology 2011; 36: 750–760. doi: 10.1016/j.psyneuen.2010.10.011 PMID: 21075538

47. Fujisawa TX, Nishitani S, Ishii S, Shinhara K. Differential modulation of impulsive behavior by loneliness and testosterone in adolescent females. Neuro Endocrinol Lett 2012; 32(6): 836–840.

48. Speert DB, McClennen SJ, Seasholtz AF. Sexually dimorphic expression of corticotropin-releasing hormone-binding protein in the mouse pituitary. Endocrinology 2002; 143: 4730–4741. PMID: 12446601

49. Van de Stolpe A, Slycke AJ, Reinders MO, Zomer AW, Goodenough S, Behl C, et al. Estrogen receptor (ER)-mediated transcriptional regulation of the human corticotropin-releasing hormone-binding protein promoter: difference effect of ERalpha and ERbeta. Molecular Endocrinology 2004; 18: 2908–2923. PMID: 15345745

50. Li Y, Nie Y, Xie J, Tang W, Liang P, Sha W, et al. The association of serotonin transporter genetic polymorphisms and irritable bowel syndrome and its influence on tagaserod treatment in Chinese patients. Dig Dis Sci 2007; 52: 2942–2949. PMID: 17394071

51. Niesler B, Kapeller J, Felli C, Atkinson W Möller D, Fischer C, et al. 5-HTTLPR and STin2 polymorphisms in the serotonin transporter gene and irritable bowel syndrome: effect of bowel habit and sex. Eur J Gastroenterol Hepatol 2009.

52. Sikander A, Rana S, Sinha S, Prasad K, Arora S, Sharma S, et al. Serotonin transporter promoter variant: Analysis in Indian IBS patients and control population. J Clin Gastroenterol 2009; 43: 902–904.
53. Sagami Y, Shimada Y, Tayama J, Nomura T, Sakata M, Endo Y, et al. Effect of a corticotropin releasing hormone receptor antagonist on colonic sensory and motor function in patients with irritable bowel syndrome. Gut 2004; 53: 958–964. PMID: 15194643

54. Bradley RG, Binder EB, Epstein MP, Tang Y, Nair HP, Liu W, et al. Influence of child abuse on adult depression: moderation by the corticotropin-releasing hormone receptor gene. Arch Gen Psychiatry 2008; 65: 190–200. doi: 10.1001/archgenpsychiatry.2007.26 PMID: 18250257

55. Polanczyk G, Caspi A, Williams B, Price T, Danese A, Sugden K, et al. Protective effect of CRHR1 gene variants on the development of adult depression following childhood maltreatment: replication and extension. Arch Gen Psychiatry 2009; 66: 978–985. doi: 10.1001/archgenpsychiatry.2009.114 PMID: 19736354

56. Seasholtz AF, Valverde RA, Denver RJ. Corticotropin-releasing hormone-binding protein: biochemistry and function from fishes to mammals. J Endocrinol 2002; 175(1): 89–97. PMID: 12379493

57. Henry BA, Lightman SL, Lowry CA. Distribution of corticotropin-releasing factor binding protein-immunoreactivity in the rat hypothalamus: association with corticotropin-releasing factor-, urocortin 1- and vimentin-immunoreactive fibres. J Neuroendocrinol. 2005; 17: 135–144. PMID: 15796765

58. Davydow DS, Kohen R, Hough CL, Tracy JH, Zatzick D, Katon WJ. A Pilot Investigation of the Association of Genetic Polymorphisms Regulating Corticotrophin-Releasing Hormone with Posttraumatic Stress and Depressive Symptoms in Medical-Surgical Intensive Care Unit Survivors. J Crit Care. 2014; 29(1):. doi: 10.1016/j.jcrc.2013.08.016

59. Ray LA. Stress-Induced and Cue-Induced Craving for Alcohol in Heavy Drinkers: Preliminary Evidence of Genetic Moderation by the OPRM1 and CRH-BP Genes. Alcohol Clin Exp Res 2011; 35(1): 166–174. doi: 10.1111/j.1530-0277.2010.01333.x PMID: 21039637

60. Claes S, Villafuerte S, Forsgren T, Sluijs S, Del-Favero J, Adolfsson R, et al. The Corticotropin-Releasing Hormone Binding Protein Is Associated with Major Depression in a Population from Northern Sweden. Biol Psychiatry 2003; 54: 867–872. PMID: 14573312

61. Levran O, Randesi M, Li Y, Rotrosen J, Ott J, Adelson M, et al. Drug Addiction and Stress-Response Genetic Variability: Association Study in African Americans. Ann Hum Genet 2014; 78(4): 290–298. doi: 10.1111/ahg.12064 PMID: 24766650

62. Lovell RM, Ford AC. Global prevalence of and risk factors for irritable bowel syndrome: a meta-analysis. Clin Gastroenterol Hepatol 2012; 10(7): 712–721. doi: 10.1016/j.cgh.2012.02.029 PMID: 2242687