Diagnostic value of \( \text{BRAF}^{\text{V600E}} \)-mutation analysis in fine-needle aspiration of thyroid nodules: a meta-analysis

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Abstract: Fine-needle aspiration (FNA) is a reliable method for preoperative diagnosis of thyroid nodules; however, about 10%–40% nodules are classified as indeterminate. The \( \text{BRAF}^{\text{V600E}} \) mutation is the most promising marker for thyroid FNA. This meta-analysis was conducted to investigate the diagnostic value of \( \text{BRAF}^{\text{V600E}} \) analysis in thyroid FNA, especially the indeterminate cases. Systematic searches were performed in PubMed, Web of Science, Scopus, Ovid, Elsevier, and the Cochrane Library databases for relevant studies prior to June 2015, and a total of 88 studies were ultimately included in this meta-analysis. Compared with FNA cytology, the synergism of \( \text{BRAF}^{\text{V600E}} \) testing increased the diagnostic sensitivity from 81.4% to 87.4% and decreased the false-negative rate from 8% to 5.2%. In the indeterminate group, the mutation rate of \( \text{BRAF}^{\text{V600E}} \) was 23% and varied in different subcategories (43.2% in suspicious for malignant cells [SMC], 13.77% in atypia of undetermined significance/follicular lesion of undetermined significance [AUS/FLUS], and 4.43% in follicular neoplasm/suspicious for follicular neoplasm [FN/SFN]). The sensitivity of \( \text{BRAF}^{\text{V600E}} \) analysis was higher in SMC than that in AUS/FLUS and FN/SFN cases (59.4% vs 40.1% vs 19.5% respectively), while specificity was opposite (86.1% vs 99.5% vs 99.7% respectively). The areas under the summary receiver-operating characteristic curve also confirmed the diagnostic value of \( \text{BRAF}^{\text{V600E}} \) testing in SMC and AUS/FLUS rather than FN/SFN cases. Therefore, \( \text{BRAF}^{\text{V600E}} \) analysis can improve the diagnostic accuracy of thyroid FNA, especially indeterminate cases classified as SMC, and select malignancy to guide the extent of surgery.

Keywords: thyroid cancer, fine-needle aspiration, \( \text{BRAF}^{\text{V600E}} \) mutation, meta-analysis

Introduction

Thyroid cancer is the most common endocrine malignancy, with favorable outcome after early detection and treatment.1,2 Fine-needle aspiration (FNA) guided by ultrasound is a routine and reliable approach for preoperative evaluation of thyroid nodules. Approximately 10%–40% of FNA specimens yield indeterminate results, and the majority of them turn out to be benign after diagnostic surgery, and thus a sizable portion of indeterminate specimens lead to unnecessary thyroidectomy.3–7 The Bethesda System for Reporting Thyroid Cytopathology divides indeterminate nodules into three subgroups: atypia of undetermined significance/follicular lesion of undetermined significance (AUS/FLUS), follicular neoplasm/suspicious for follicular neoplasm (FN/SFN), and suspicious for malignant cells (SMC).8 The indeterminate thyroid nodule is the most intractable problem in clinical management, which highlights the urgency to develop effective ancillary testing to identify cancerous nodules for timely and appropriate management.
Great progress has been achieved in the understanding of molecular mechanisms of thyroid cancer, and various mutations have been identified in the early stage of thyroid cancer, such as \textit{BRAF}, \textit{RAS}, \textit{PI3K}, and \textit{PTEN}.\textsuperscript{7} These genetic alterations are excellent candidates for disease hallmarks, since 60%–70% of thyroid cancers harbor at least one genetic mutation.\textsuperscript{9} The \textit{BRAF}\textsubscript{V600E} mutation appears to be the most promising biomarker specific for papillary thyroid cancer (PTC),\textsuperscript{9} which aberrantly activates the tumor-initiating MAPK pathway and drives the carcinogenesis and progression of thyroid cancer.\textsuperscript{9,10}

Whether \textit{BRAF}\textsubscript{V600E} analysis could be routinely used in clinical practice is still controversial. Numerous researchers have proved that \textit{BRAF}\textsubscript{V600E}-mutation testing is an effective diagnostic approach for thyroid FNA,\textsuperscript{11} while others believe that its utility is limited by low prevalence of \textit{BRAF}\textsubscript{V600E} mutation in indeterminate nodules.\textsuperscript{12} Therefore, we conducted a structured meta-analysis to estimate the additional diagnostic yield of \textit{BRAF}\textsubscript{V600E}-mutation analysis in thyroid FNA, and further evaluated the malignancy rate, \textit{BRAF}\textsubscript{V600E}-mutation frequency, and diagnostic value of \textit{BRAF}\textsubscript{V600E} testing in different categories of indeterminate nodule.

Materials and methods

Search strategy and selection criteria

Systematic searches were performed in the PubMed, Web of Science, Scopus, Ovid, Elsevier, and Cochrane Library databases for relevant articles prior to June 2015. The search terms were: [(thyroid cancer] or [thyroid neoplasm] or [thyroid tumor], \textit{BRAF}, and [FNA] or [fine needle aspiration]). The references of available articles were also reviewed. Study selection consisted of initial screening of titles or abstracts and second screening of full texts. Studies were included if they met the following criteria: 1) research article rather than review, system review, case report, editorial, or comments; 2) the material for \textit{BRAF}\textsubscript{V600E}-mutation analysis was obtained by FNA; 3) the final diagnosis was based on a definite gold standard, such as surgical histology, unequivocal histocytology, or reliable clinical follow-up; 4) the data were available to construct 2×2 tables or analyze malignancy rate or \textit{BRAF}\textsubscript{V600E}-mutation prevalence.

Data extraction and quality assessment

The following items were extracted: study by author name(s), country, number of centers, enrollment period, study design, mean age of patients, mean diameter of nodules, reference standard of final diagnosis, and genotyping method. Most research classified cytological results according to the Bethesda system\textsuperscript{9} or the British Thyroid Association,\textsuperscript{13,14} as shown in Table 1. In this meta-analysis, FNA cases classified as AUS/FLUS (Thy3a) and FN/SFN (Thy3f) were regarded as cytologically negative and lesions diagnosed as SMC (Thy4) were cytologically positive. Final diagnosis was based on histopathologic examination after surgery or a combination of cytological examination and clinical follow-up. Then, patient numbers for true-positive, false-positive, false-negative, and true-negative results were extracted to construct the 2×2 tables.

The methodological quality of studies eligible for diagnostic analysis of FNA cytology and/or \textit{BRAF}\textsubscript{V600E} testing was assessed according to the Quality Assessment of Diagnostic Studies 2, which comprises four domains: patient selection, index test, reference standard, and flow and timing.\textsuperscript{15} A series of questions was used to judge the risk of bias and applicability concerns as low, high, or unclear risk.

Statistical analysis

The threshold effect was calculated by the Spearman correlation coefficient, and \(P<0.05\) indicated the existence of a threshold effect. Nonthreshold heterogeneity was assessed by the Cochran \(Q\) test and inconsistency index (\(I^2\)). \(I^2>50\%\) suggested significant heterogeneity, and a random-effect model (DerSimonian–Laird method) was chosen.\textsuperscript{16,17} Metaregression analysis was used to identify the possible sources of nonthreshold heterogeneity. The following covariates were considered in the metaregression analysis: country, number of centers (single or multiple), sample size (<100, 100–500, 500–1,000, or >1,000), study design (prospective or retrospective), reference standard (histology or cytology plus clinical follow-up), and genotyping method. If \(P<0.05\), the covariate was to be regarded as the source of nonthreshold heterogeneity.

The pooled sensitivity, specificity, positive likelihood ratio (PLR), negative likelihood ratio (NLR), and diagnostic

| Table 1 Comparison between the British and Bethesda systems for classification of thyroid cytopathology |
|-----------------------------------------------------|
| **Bethesda** | **British** |
| Nondiagnostic or unsatisfactory | Thy1 (nondiagnostic) |
| Benign | Thy2 (nonneoplastic) |
| AUS/FLUS (atypia of undetermined significance/follicular lesion of undetermined significance) | Thy3a (neoplasm possible, atypia/nondiagnostic) |
| FN/SFN (follicular neoplasm/suspicious for follicular neoplasm) | Thy3f (neoplasm possible, suggesting follicular neoplasm) |
| SMC (suspicious for malignancy) | Thy4 (suspicious of malignancy) |
| Malignant | Thy5 (malignant) |
odds ratio (DOR) with 95% confidence interval (CI) were computed to estimate diagnostic accuracy. DOR combined the data of sensitivity and specificity into a single indicator ranging from 0 to infinity, reflecting the discriminatory performance of testing. The summary receiver-operating characteristic (SROC) curve was a mathematical model for the plot of sensitivity (1 − specificity). The $Q$ index indicated the point at which sensitivity was equal to specificity. The areas under the SROC curve (AUCs) calculated the inherent capacity of the diagnostic test. If the DOR closed to 1, the diagnostic method was thought to be perfect.

The threshold effect, pooled diagnostic features, and metaregression were calculated by Meta-Disc (version 1.4; Ramony Cajal Hospital, Madrid, Spain). Pooled rates of malignancy and $BRAF^{V600E}$ mutation were calculated by R statistical software (version 3.2.1; R Foundation for Statistical Computing, Vienna, Austria). Quality assessment was conducted using Review Manager (version 5.2; Cochrane Collaboration). $P<0.05$ was considered statistically significant.

**Results**

**Search results and quality assessment**

The search process is shown in Figure 1. A total of 1,261 articles were initially identified, and 1,130 of these were excluded after reviewing titles and abstracts. The remaining 131 articles were investigated in detail. In accordance with the selection criteria mentioned in the Materials and methods section, 43 articles were excluded after reading the full texts. Finally, 88 studies published from 2004 to 2015 were included in this meta-analysis. Among these, 51 studies were included in the analysis of diagnostic accuracy, and at the same time 37 studies and 62 studies were available for analysis of malignancy rate and $BRAF^{V600E}$-mutation rate, respectively.

The characteristics of studies eligible for diagnostic analysis of FNA cytology and $BRAF^{V600E}$ testing are summarized in Table 2. As shown in Figure 2, about a third of studies had a high risk of bias in patient selection, because 14 of them did not enroll the samples consecutively or at random and eleven excluded a number of patients inappropriately. Twelve studies did not receive the same reference standard, since some patients were diagnosed by histopathology and others by FNA cytology plus clinical follow-up. Also, 17 studies did not include all patients, due to the unsatisfactory FNA or failure of $BRAF^{V600E}$ testing. As a result, nearly half of the studies harbored a high risk of bias in flow and timing. Fortunately, the risk of bias in the index test and reference standard was relatively low.

![Figure 1](image1.png)

**Figure 1** Flowchart of study-selection process.

**Abbreviation:** FNA, fine-needle aspiration.
Table 2 Characteristics of studies eligible for the diagnostic analysis of FNA cytology and BRAF<sup>V600E</sup> testing

| Study          | Country | Centers, n | Enrollment period | Design | Mean age, years | Mean diameter, cm | Final diagnosis | Genotyping method                                      |
|----------------|---------|------------|-------------------|--------|-----------------|-------------------|-----------------|--------------------------------------------------------|
| Cohen et al<sup>18</sup> | USA     | 1          | Jan 2001–Jan 2003 | Retro<sup>a</sup> | –                | –                 | A               | Direct sequencing + mutector assay                     |
| Xing et al<sup>19</sup> | USA     | 1          | –                 | Pro<sup>b</sup>   | –                | –                 | B               | Direct sequencing + colorimetric method                |
| Domingues et al<sup>20</sup> | Portugal | 1          | –                 | Retro       | –                | –                 | A               | PCR-RFLP                                               |
| Pizzolani et al<sup>21</sup> | Italy    | 1          | Sep 2005–Jun 2006 | Pro         | –                | –                 | B               | Real-time AS-PCR                                       |
| Sapio et al<sup>22</sup> | Italy    | 2          | –                 | Retro       | –                | –                 | B               | Direct sequencing                                      |
| Kim et al<sup>23</sup> | South Korea | 1          | Aug 2005–Jul 2006 | Retro     | –                | –                 | A               | Pyrosequencing                                         |
| Bentz et al<sup>25</sup> | USA      | 1          | 1994–2004         | Retro     | 40.9             | –                 | A               | LCPCR + FMCA                                           |
| Jo et al<sup>26</sup>   | South Korea | 1          | June 2006–Dec 2006 | Pro       | –                | 1                 | A               | Pyrosequencing                                         |
| Marchetti et al<sup>27</sup> | Italy    | 1          | 1996–2008         | Retro     | –                | –                 | A               | Direct sequencing                                      |
| Sapio et al<sup>28</sup> | Italy    | 2          | –                 | Pro      | –                | –                 | B               | LCPCR + FMCA                                           |
| Kim et al<sup>29</sup> | South Korea | 1          | Oct 2008–Dec 2009 | Pro       | 50.7             | 1.1               | A               | DHPLC + direct sequencing                              |
| Cantara et al<sup>30</sup> | Italy    | 1          | –                 | Pro      | 51.2             | –                 | A               | Direct sequencing                                      |
| Girlande et al<sup>31</sup> | Italy    | 1          | –                 | Pro      | –                | –                 | A               | Direct sequencing                                      |
| Kim et al<sup>32</sup> | South Korea | 1          | Mar 2008–Jun 2008 | Pro      | 45.6             | 1.17              | A               | DPO-based multiplex PCR + direct sequencing            |
| Moses et al<sup>33</sup> | USA      | 1          | Jun 2006–Jul 2008 | Pro       | 51               | –                 | B               | DPO-based multiplex PCR                                |
| Musholt et al<sup>35</sup> | Germany  | 6          | Jan 2008–Jul 2009 | Pro      | –                | –                 | A               | Direct sequencing                                      |
| Adeniran et al<sup>36</sup> | USA      | 1          | Sep 2009–Nov 2010 | Pro       | 52.6             | –                 | A               | SSCP analysis                                          |
| Lee et al<sup>37</sup> | South Korea | 1          | Oct 2008–Dec 2009 | Pro       | 50.7             | 1.1               | A               | Pyrosequencing                                         |
| Lee et al<sup>38</sup> | South Korea | 1          | July 2007–Dec 2009 | Pro      | 50.3             | 1.46              | A               | Pyrosequencing                                         |
| Moon et al<sup>39</sup> | South Korea | 1          | Sep 2008–May 2009 | Retro     | 49.4             | 0.95              | B               | Direct sequencing                                      |
| Pelizzo et al<sup>40</sup> | Italy    | 1          | Oct 2008–Sep 2009 | Pro      | 47.8             | –                 | A               | Direct sequencing + MASA                              |
| Smith et al<sup>41</sup> | USA      | 1          | –                 | Retro     | –                | –                 | A               | MCA                                                    |
| Yeo et al<sup>42</sup> | South Korea | 1          | Jul 2009–Jan 2010 | Pro      | 51.27            | 1.3               | B               | Pyrosequencing                                         |
| Cañadas-Garre et al<sup>43</sup> | Spain    | 1          | Jun 2006–Dec 2009 | Pro      | 49.8             | –                 | A               | PCR-RFLP                                               |
| Kang et al<sup>44</sup> | South Korea | 1          | Apr 2008–Jul 2009 | Pro      | –                | –                 | A               | AS-PCR + direct sequencing                             |
| Kwak et al<sup>45</sup> | South Korea | 1          | Jun 2009–Oct 2010 | Retro     | 48               | 0.92              | A               | DPO-PCR + real-time PCR                                |
| Lee et al<sup>46</sup> | South Korea | 1          | Aug 2008–Mar 2011 | Pro      | 49.5             | –                 | A               | MEMO-PCR + direct sequencing                           |
| Mancini et al<sup>47</sup> | Italy    | 1          | –                 | Pro      | 55.1             | 2.38              | A               | High-resolution melting analysis                       |
| Rossi et al<sup>48</sup> | Italy    | 1          | –                 | Pro      | 52               | –                 | B               | Direct sequencing                                      |
| Tomei et al<sup>49</sup> | Italy    | 1          | –                 | Retro     | –                | –                 | A               | Pyrosequencing                                         |
| Brahma et al<sup>50</sup> | Indonesia | 1          | Aug 2010–Jun 2011 | Pro      | 46               | >1                | A               | PCR-RFLP                                               |
| Di Benedetto et al<sup>51</sup> | Italy    | 1          | –                 | Pro      | –                | –                 | A               | Direct sequencing                                      |
| Koh et al<sup>52</sup> | South Korea | 1          | Jan 2009–Oct 2010 | Pro      | 48.6             | 1.05              | B               | DPO-PCR                                                |
| Park et al<sup>53</sup> | South Korea | 1          | Jan 2011–May 2011 | Retro     | –                | –                 | B               | Real-time PCR + pyrosequencing                         |
| Beaudenon-Huibrege et al<sup>54</sup> | USA      | 5          | Jul 2010–Oct 2012 | Pro      | –                | –                 | A               | Multiplex PCR                                           |
| Crescenz et al<sup>55</sup> | Italy    | 1          | –                 | Pro      | –                | –                 | A               | Real-time sequencing                                   |
| Ezlinger et al<sup>56</sup> | Germany  | 1          | 1995–2009         | Retro     | –                | –                 | A               | High-resolution melting PCR + pyrosequencing           |
Synthesis of analysis results

Diagnostic value of FNA cytology, \(BRAF^{V600E}\)-mutation analysis, and combined strategy in all the thyroid FNA specimens

Spearman correlation coefficients for FNA cytology, \(BRAF^{V600E}\) testing and combined strategy were 0.032 (\(P=0.826\)), 0.254 (\(P=0.078\)), and 0.064 (\(P=0.661\)), respectively; therefore, no threshold effect existed in the analysis. However, there was substantial nonthreshold heterogeneity (\(I^2=50\%\), \(P=0.05\)), so the random-effect model was chosen to pool the diagnostic features. A total of 51 studies were included in this part of the analysis, but one was excluded because it had no false-positive or true-negative case to calculate the diagnostic index (Table 3).

Based on the feasible FNA cytology results from 50 studies, pooled sensitivity, specificity, PLR, NLR, and DOR were 0.814 (95% CI 0.803–0.824), 0.981 (95% CI 0.978–0.985), 23.868 (95% CI 14.139–40.293), 0.216 (95% CI 0.172–0.273), and 127.73 (95% CI 75.082–217.28) (Table 4). The AUC of the SROC curve was 0.9551 (standard error [SE] 0.0127), with a \(Q\)-value of 0.8975 (SE 0.0178) (Figure 3A). Data for the \(BRAF^{V600E}\)-mutation test were unavailable in one study, and 49 studies with 9,361 patients were finally analyzed. Pooled sensitivity, specificity, PLR, NLR, and DOR were 0.619 (95% CI 0.605–0.633), 0.997 (95% CI 0.995–0.998), 34.982 (95% CI 23.801–51.415), 0.433 (95% CI 0.384–0.489), and 96.570 (95% CI 63.932–145.87) (Table 4). The AUC of the SROC was 0.9207 (SE 0.0233), with a \(Q\)-value of 0.8542 (SE 0.0268) (Figure 3B).

Also, the positive predictive value of \(BRAF^{V600E}\) testing was 99.5% (2,886 of 2,900). After \(BRAF^{V600E}\) analysis was combined with FNA cytology, sensitivity increased to 0.874 (95% CI 0.865–0.884), the DOR and AUC improved to 187.92 (95% CI 110.24–320.35) and 0.9744 (SE 0.0062), respectively, with a \(Q\)-value of 0.9271 (SE 0.0107) (Table 4, Figure 3C). The synergism between FNA cytology and \(BRAF^{V600E}\) testing also decreased the false-negative rate from 8% in FNA cytology to 5.2%, but increased the false-positive rate from 3% to 5% at the same time.

Diagnostic value of \(BRAF^{V600E}\)-mutation analysis in indeterminate cases (Bethesda categories III–V)

There were 43 studies included in the diagnostic analysis of \(BRAF^{V600E}\) testing in the indeterminate thyroid nodules (Table 5). Our data showed that 23% of indeterminate nodules harbored the \(BRAF^{V600E}\) mutation. No threshold effect was detected, so the random-effect model was chosen to pool the diagnostic
Figure 2  Methodological quality of studies included, assessed by the Quality Assessment of Diagnostic Studies 2 criteria.

Table 3 Diagnostic analysis of FNA cytological examination and BRAFV600E-mutation analysis in all the FNA specimens.

| Study               | Year | FNA     |       |       | BRAF     |       |       | FNA + BRAF |
|---------------------|------|---------|-------|-------|----------|-------|-------|------------|
|                     |      | TP      | FP    | FN    | TN       | TP    | FP    | FN         | TN         |
| Cohen et al         | 2004 | 25      | 0     | 34    | 32       | 23    | 0     | 36         | 32         |
| Xing et al          | 2004 | 10      | 0     | 19    | 12       | 8     | 0     | 22         | 14         |
| Domingues et al     | 2005 | 10      | 0     | 3     | 11       | 3     | 0     | 10         | 11         |
| Pizzolanti et al    | 2007 | 13      | 0     | 4     | 32       | 11    | 0     | 6          | 32         |
| Sapio et al         | 2007 | 24      | 23    | 2     | 95       | 10    | 0     | 16         | 118        |
| Sapio et al         | 2007 | 6       | 0     | 2     | 67       | 4     | 0     | 4          | 123        |
| Kim et al           | 2008 | 60      | 0     | 21    | 22       | 63    | 0     | 18         | 22         |
| Bentz et al         | 2009 | 22      | 0     | 18    | 5        | 17    | 0     | 20         | 5          |
| Jo et al            | 2009 | 30      | 0     | 9     | 58       | 30    | 0     | 10         | 58         |
| Marchetti et al     | 2009 | 88      | 2     | 4     | 17       | 59    | 0     | 32         | 19         |
| Nikiforov et al     | 2009 | 27      | 2     | 21    | 36       | 18    | 0     | 30         | 38         |
| Zatelli et al       | 2009 | 66      | 5     | 24    | 373      | 48    | 0     | 42         | 378        |
| Cantara et al       | 2010 | 46      | 8     | 16    | 112      | 33    | 0     | 45         | 157        |
| Girlando et al      | 2010 | 38      | 0     | 22    | 2        | 41    | 0     | 19         | 2          |
| Kim et al           | 2010 | 251     | 2     | 6     | 690      | 221   | 5     | 47         | 688        |
| Kwak et al          | 2010 | 108     | 10    | 1     | 10       | 87    | 0     | 22         | 20         |
| Moses et al         | 2010 | 71      | 13    | 30    | 337      | 23    | 0     | 78         | 95         |
| Musholt et al       | 2010 | 19      | 13    | 11    | 50       | 9     | 0     | 21         | 63         |
| Adeniran et al      | 2011 | 47      | 0     | 13    | 12       | 40    | 0     | 20         | 12         |
| Kim et al           | 2011 | 146     | 0     | 27    | 21       | 154   | 1     | 19         | 20         |
| Lee et al           | 2011 | 127     | 0     | 70    | 29       | 174   | 1     | 24         | 28         |
| Moon et al          | 2011 | 98      | 0     | 10    | 191      | 57    | 0     | 51         | 191        |
| Pelizzo et al       | 2011 | 133     | 5     | 6     | 117      | 98    | 0     | 59         | 113        |
| Smith et al         | 2011 | 10      | 0     | 5     | 5        | 10    | 0     | 5          | 5          |
| Yeo et al           | 2011 | 183     | 1     | 9     | 709      | 99    | 0     | 93         | 710        |
| Cañadas-Garre et al | 2012 | 12      | 0     | 31    | 132      | 17    | 0     | 31         | 160        |
| Kang et al          | 2012 | 289     | 1     | 15    | 8        | 226   | 2     | 78         | 7          |
| Kwak et al          | 2012 | 318     | 0     | 33    | 86       | –     | –     | –          | –          |
| Lee et al           | 2012 | 382     | 1     | 47    | 33       | 342   | 0     | 87         | 34         |
| Mancini et al       | 2012 | 13      | 1     | 10    | 32       | 12    | 0     | 11         | 33         |
| Marchetti et al     | 2012 | 85      | 0     | 5     | 0        | 63    | 0     | 22         | 0          |
| Rossi et al         | 2012 | 159     | 3     | 73    | 1,621    | 114   | 0     | 172        | 93         |
| Tomei et al         | 2012 | 44      | 0     | 5     | 38       | 28    | 0     | 21         | 38         |
| Brahma et al        | 2013 | 23      | 0     | 26    | 21       | 17    | 0     | 32         | 21         |
| Di Benedetto et al  | 2013 | 15      | 1     | 3     | 239      | 13    | 0     | 5          | 240        |
| Koh et al           | 2013 | 277     | 0     | 27    | 194      | 176   | 3     | 141        | 198        |
| Park et al          | 2013 | 71      | 5     | 8     | 31       | 44    | 1     | 37         | 35         |
| Beaudenon-Huibregts et al | 2014 | 36     | 4     | 18    | 49       | 21    | 0     | 35         | 53         |
| Crescenzi et al     | 2014 | 20      | 0     | 1     | 9        | 8     | 0     | 13         | 9          |
| Ezlinger et al      | 2014 | 57      | 0     | 28    | 225      | 22    | 0     | 43         | 188        |
| Guo et al           | 2014 | 55      | 1     | 8     | 19       | 41    | 0     | 22         | 20         |
| Johnson et al       | 2014 | 31      | 3     | 19    | 44       | 16    | 0     | 28         | 42         |

(Continued)
features: sensitivity 0.442 (95% CI 0.416–0.468), specificity 0.997 (95% CI 0.994–0.999), PLR 12.267 (95% CI 8.175–18.406), NLR 0.613 (95% CI 0.551–0.683), and DOR 23.939 (95% CI 15.388–37.242) (Table 6; Figure 4A and B). The AUC of the SROC was 0.8711 (SE 0.0414), with a Q-value of 0.8015 (SE 0.0410) (Figure 4C).

To evaluate the diagnostic value of BRAFV600E testing in different categories of indeterminate nodules, we separated the indeterminate cases into three different and more specific categories according to the Bethesda system. Studies with sample sizes fewer than ten were excluded to avoid potential bias. The malignancy rates of FN/SFN and AUS/FLUS were 30.55% and 34.99%, while 90.35% of SMC cases turned out to be malignant (Table 7). Besides that, the BRAFV600E-mutation rate varied among these groups: it existed in 43.2% of SMC cases, but only 13.77% in AUS/FLUS and 4.43% in FN/SFN patients (Table 7). Furthermore, the sensitivity of BRAFV600E testing was higher in SMC (0.594, 95% CI 0.556–0.631) than AUS/FLUS (0.401, 95% CI 0.328–0.477) and FN/SFN (0.195, 95% CI 0.128–0.278), while specificity was higher in the AUS/FLUS (0.995, 95% CI 0.982–0.999) and FN/SFN (0.997, 95% CI 0.983–1.000) groups than the SMC group (0.861, 95% CI 0.784–0.918) (Table 6). The AUC of the SROC was 0.7674 (SE 0.0564) with a Q-value of 0.7079 (SE 0.0474) in the SMC group, and 0.7999 (SE 0.0897) with a Q-value of 0.7358 (SE 0.0783) in the AUS/FLUS group, but was not significant in FN/SFN cases, since the lower limit of the AUC was less than 0.5 (Figure 5).

Heterogeneity test

Heterogeneity was present in our meta-analysis, and Spearman correlation coefficients suggested no significant threshold effect. To explore sources of heterogeneity, we assessed multiple variables by metaregression, including country, number of centers, sample size, study design, reference standard, and genotyping method. The results indicated that country and sample size were possible sources of heterogeneity (data not shown). Other covariates that may have caused heterogeneity, such as enrollment period, age, sex, nodule diameter, size of needle, use of blinding method, and

Table 4 Results of meta-analysis for diagnostic value of FNA cytology, BRAFV600E-mutation analysis, and the combined strategy in all FNA specimens

| Parameter | FNA | B BRAF | FNA + BRAF |
|-----------|-----|--------|------------|
|           |     |        |            |
|           | Result | 95% CI | Heterogeneity, $I^2$ | Result | 95% CI | Heterogeneity, $I^2$ | Result | 95% CI | Heterogeneity, $I^2$ |
| Pooled sensitivity | 0.814 | 0.803–0.824 | 93.5% | 0.619 | 0.605–0.633 | 93% | 0.874 | 0.865–0.884 | 92.5% |
| Pooled specificity | 0.981 | 0.978–0.985 | 86.4% | 0.997 | 0.995–0.998 | 14.1% | 0.968 | 0.963–0.972 | 92.5% |
| Pooled LR, + | 23.868 | 14.139–40.293 | 87.7% | 34.982 | 23.801–51.415 | 19.5% | 22.353 | 13.027–38.355 | 93.1% |
| Pooled LR, – | 0.216 | 0.172–0.273 | 94.2% | 0.433 | 0.384–0.489 | 91.8% | 0.146 | 0.111–0.192 | 93% |
| Pooled DOR | 127.73 | 75.082–217.28 | 76.1% | 96.570 | 63.932–145.87 | 21.4% | 187.92 | 110.24–320.35 | 76.4% |
| SROC | | | |
| AUC | 0.9551 | | 0.9207 | 0.9744 |
| $Q^*$ | 0.8975 | | 0.8542 | 0.9271 |

Note: The $Q$ index indicates the point at which sensitivity is equal to specificity.

Abbreviations: FNA, fine-needle aspiration; CI, confidence interval; LR, likelihood ratio; DOR, diagnostic odds ratio; SROC, summary receiver-operating characteristic; AUC, area under the curve.
differences in operating protocol, were not analyzed here, due to the loss of partial data.

**Discussion**

Thyroid cancer is on a rapid increase these days, partially due to advancing diagnostic methods. The majority of cases have an excellent prognosis, with 30-year survival rate exceeding 90% after thyroidectomy and/or radioiodine ablation. Preoperative diagnosis is of indisputable value in distinguishing thyroid cancer from benign nodules. FNA biopsy is a conventional technique to identify malignant thyroid nodules preoperatively and effectively, which has also been demonstrated in our meta-analysis. However, the extensive use of this approach is influenced by its inherent limitations, such as size or location of nodule, quantity and quality of obtained material, technical skill of the cytopathologist, and the overlap of cytomorphological features between malignant and benign nodules. Therefore, a fraction of cases are classified as nondiagnostic or indeterminate, and about 15%–30% of them get malignant pathology after diagnostic surgery. Since the occurrence of malignancy is too high for just watchful waiting, numerous patients with indeterminate diagnosis accept unnecessary surgical intervention. $BRAF^{V600E}$ mutation is the most promising marker for thyroid nodules. A similar meta-analysis conducted by Jia et al of 16 studies suggested that $BRAF^{V600E}$ analysis had diagnostic value in indeterminate thyroid nodules, but another analysis of eight eligible studies found a low $BRAF^{V600E}$-mutation rate within indeterminate
Table 5 Diagnostic analysis of BRAF V600E mutation analysis for indeterminate cases

| Year | Study | TP | FP | FN | TN |
|------|-------|----|----|----|----|
| 2007 | liu et al | 0 | 0 | 0 | 0 |
| 2008 | Johnson et al | 0 | 0 | 0 | 0 |
| 2009 | Park et al | 0 | 0 | 0 | 0 |
| 2010 |Poller et al | 0 | 0 | 0 | 0 |
| 2011 | li Mercier et al | 0 | 0 | 0 | 0 |
| 2012 | Mancini et al | 0 | 0 | 0 | 0 |
| 2013 | seo et al | 0 | 0 | 0 | 0 |
| 2014 | seo et al | 0 | 0 | 0 | 0 |
| 2015 | brahma et al | 0 | 0 | 0 | 0 |
| 2016 | brahma et al | 0 | 0 | 0 | 0 |
| 2017 | brahma et al | 0 | 0 | 0 | 0 |
| 2018 | brahma et al | 0 | 0 | 0 | 0 |
| 2019 | brahma et al | 0 | 0 | 0 | 0 |
| 2020 | brahma et al | 0 | 0 | 0 | 0 |
| 2021 | brahma et al | 0 | 0 | 0 | 0 |

Table 6 Results of meta-analysis for diagnostic value of BRAF V600E mutation in indeterminate cases

| Parameter | Indeterminate | SMC | AUS/FLUS | FN/SFN |
|-----------|---------------|-----|-----------|--------|
| Pool sensitivity | 0.442 | 0.416–0.468 | 0.594 | 0.556–0.631 | 0.401 | 0.328–0.477 | 0.195 | 0.128–0.278 |
| Pool specificity | 0.997 | 0.994–0.999 | 0.861 | 0.784–0.918 | 0.995 | 0.982–0.999 | 0.997 | 0.983–1.000 |
| Pool LR+ | 12.267 | 8.175–18.406 | 3.434 | 1.625–7.259 | 7.001 | 3.336–14.691 | 9.573 | 3.611–25.379 |
| Pool LR− | 0.613 | 0.551–0.683 | 0.542 | 0.462–0.637 | 0.694 | 0.576–0.835 | 0.733 | 0.522–1.030 |
| Pool DOR | 23.939 | 15.388–37.242 | 7.588 | 3.944–14.598 | 14.469 | 6.100–34.320 | 14.808 | 4.966–44.156 |

Note: "The Q index indicates the point at which sensitivity is equal to specificity. "−" indicates the AUC of the SROC was not significant in FN/SFN cases, since the lower limit of the AUC was less than 0.5.

Abbreviations: FNA, fine-needle aspiration; CI, confidence interval; LR, likelihood ratio; DOR, diagnostic odds ratio; SROC, summary receiver-operating characteristic; AUC, area under the curve.
Consistent with previous research, our meta-analysis showed that BRAF\textsuperscript{V600E} analysis had high specificity and positive predictive value. As a rule-in test, a positive result of BRAF\textsuperscript{V600E} analysis indicates a high probability of malignancy so that therapeutic surgery is recommended, but the negative result cannot exclude malignancy, and further evaluations, such as follow-up ultrasound or repeat FNA, are needed. When we combined BRAF\textsuperscript{V600E}-mutation testing with FNA cytological examination, sensitivity increased by 6% and the false-negative rate decreased from 8% to 5.2%, while the...
false-positive rate increased from 3% to 5% at the same time. However, BRAF<sup>V600E</sup> testing had relatively low sensitivity of 44.2% in the indeterminate group. Also, the yield and usefulness of BRAF<sup>V600E</sup> analysis can be greatly varied with the prevalence of BRAF<sup>V600E</sup> mutation in different subcategories of indeterminate nodules. BRAF<sup>V600E</sup> mutation was present in 43.2% of SMC cases regarded as cytologically positive in our meta-analysis, but only 13.77% in AUS/FLUS and 4.43% in FN/SFN cases. Therefore, it was reasonable that BRAF<sup>V600E</sup> analysis did best in SMC lesions (sensitivity 59.4%, specificity 86.1%) and also had certain diagnostic value in AUS/FLUS nodules (sensitivity 40.1%, specificity 99.5%), but no

| Category     | Malignancy rate | BRAF<sup>V600E</sup>-mutation rate |
|--------------|-----------------|-------------------------------------|
|              | n   | Event | Pooled | 95% CI      | Heterogeneity, I² | n     | Event | Pooled | 95% CI      | Heterogeneity, I² |
| SMC          | 1,214 | 1,067 | 0.9035 | 0.8769–0.9301 | 83.62%          | 2,382 | 1,074 | 0.4320 | 0.3340–0.5299 | 98.22%          |
| FN/SFN       | 509   | 158   | 0.3055 | 0.2394–0.3715 | 54.6%           | 1,758 | 101  | 0.0443 | 0.0292–0.0594 | 64.02%          |
| AUS/FLUS     | 594   | 198   | 0.3499 | 0.2956–0.4042 | 83.01%          | 2,304 | 310  | 0.1377 | 0.0989–0.1765 | 95.93%          |

Abbreviations: CI, confidence interval; SMC, suspicious for malignant cells; FN/SFN, follicular neoplasm/suspicious for FN; AUS/FLUS, atypia of undetermined significance/follicular lesion of undetermined significance.

Figure 5 Summary receiver-operating characteristic (SROC) curve and area under the curve (AUC) of SMC cases (A), AUS/FLUS cases (B) and FN/SFN cases (C). Note: *The Q index indicates the point at which sensitivity is equal to specificity.

Abbreviations: SMC, suspicious for malignant cells; AUS/FLUS, atypia of undetermined significance/follicular lesion of undetermined significance; FN/SFN, follicular neoplasm/suspicious for FN; SE, standard error.
significant benefit in the FN/SFN group, which needs other diagnostic approaches with high sensitivity.

\(BRAF^{V600E}\) mutation is specific to PTC or anaplastic thyroid cancer arising from PTC, and more common in conventional and tall-cell PTC than follicular-variant PTC (FVPTC), which results in the discrepancy of \(BRAF^{V600E}\) test in different indeterminate subgroups. The FN/SFN category is mainly constituted of FVPTC, follicular thyroid cancer (FTC), adenomatoid hyperplasia, and follicular adenoma, which harbors low prevalence of \(BRAF^{V600E}\) mutation and is hard for \(BRAF^{V600E}\) testing to determine malignancy, so FVPTC and FTC may be the main source of false-negative results. The molecular profiles of FVPTC and FTC are similar, with frequent \(RAS\) and rare \(BRAF\) mutation.\(^{78,76}\) \(RAS\) mutation, mutually exclusive with \(BRAF\) mutation, is the most frequent genetic mutation in indeterminate nodules, and provides important diagnostic information for \(BRAF^{V600E}\) negative nodules.\(^{69,77}\) An et al reported that single \(RAS\)-mutation analysis had a sensitivity of 93.3% and specificity of 75.0% in indeterminate nodules, and the combination of \(RAS\) and \(BRAF\) mutation provided additional diagnosis value for 60%–70% indeterminate thyroid nodules.\(^{78}\) Other genetic alterations, such as \(RET/PTC\) and \(PAX8/PPARG\), also contribute to the definite diagnosis of indeterminate nodules.\(^{69,79,80}\) Therefore, an expanded panel can be more effective, which is also recommended by the revised American Thyroid Association management guidelines.\(^{73}\) As some mutations also present in benign nodules, the accompanying increase in false-positive rate should not be neglected. For instance, \(RAS\) mutation and \(PAX8/PPARG\) translocation are also found in follicular adenoma.\(^{79,81}\) Additionally, some thyroid cancer does not have definitive molecular mutation, and other efficient rule-out testing with high negative predictive value should be explored.

The clinical management decision is directly based on the malignant risk, ranging from repeat FNA to diagnostic lobectomy to total thyroidectomy. Uncertain diagnosis may lead to delayed treatment or unnecessary intervention. Based on the Bethesda classification, malignancy rates for FN/SFN and SMC nodules are 15%–30% and 60%–75%, respectively, and are much more variable in AUS/FLUS cases (7%–48%).\(^{8}\) In our analysis, the malignancy rate of the SMC group was higher than that recorded in the Bethesda classification, and this discrepancy might have resulted from continuous improvement in FNA technique, since the data for the Bethesda system were collected several years ago. \(BRAF^{V600E}\) mutation is a strong indicator for malignancy, and total thyroidectomy should be proposed as the first-line treatment for \(BRAF^{V600E}\)-positive nodules to decrease the recurrence and avoid complications caused by standard two-stage surgery. Nevertheless, \(BRAF^{V600E}\) testing is relatively insufficient for AUS/FLUS and even has no effect in FN/SFN patients, due to the low prevalence of \(BRAF^{V600E}\) mutation, but their malignant occurrence (30.55% and 34.99%) was too high to perform clinical observation. Other approaches, such as core-needle biopsy and immunohistochemistry, are also required to confidently guide the management. Several multicenter studies have reported that \(BRAF^{V600E}\) mutation is associated with aggressive clinicopathological characteristics and predicts recurrence and mortality for PTC patients.\(^{82–89}\) Therefore, more aggressive surgery, such as prophylactic central lymph-node dissection and closer follow-up, should be considered in the management of \(BRAF^{V600E}\)-positive thyroid cancer.

Despite its achievements, our meta-analysis had several limitations. Firstly, there was significant nonthreshold heterogeneity, partly caused by country and sample size of different studies, but other possible covariates were unable to be analyzed due to the paucity of data. The heterogeneity from country may be due to the different \(BRAF^{V600E}\) prevalence in worldwide populations, eg, it is up to 80% in South Korea, which is much higher than other regions.\(^{24}\) Secondly, about a third of the studies had a high risk of bias in patient selection, and nearly half had a high risk of bias in flow and timing, which may affect the reliability of our results.

### Conclusion

This meta-analysis demonstrated that \(BRAF^{V600E}\) analysis using residual material obtained from routine FNA could improve diagnostic accuracy and reduce false-negative rates. Besides, \(BRAF^{V600E}\) analysis had certain diagnostic value in SMC and AUS/FLUS cases, especially the SMC group, selecting cases with high malignancy possibility and guiding intraoperative or postoperative management, though its value in FN/SFN cases was doubtful, and expanded panels containing other diagnostic markers are recommended. Therefore, more studies of high quality are needed to balance the advantages and disadvantages of \(BRAF^{V600E}\) testing for patients and to select the most suitable population for this diagnostic method.

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Disclosure
The authors report no conflicts of interest in this work.

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