Neural Pharmacodynamic State Space Modeling

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Abstract
Modeling the time-series of high-dimensional, longitudinal data is important for predicting patient disease progression. However, existing neural network based approaches that learn representations of patient state, while very flexible, are susceptible to overfitting. We propose a deep generative model that makes use of a novel attention-based neural architecture inspired by the physics of how treatments affect disease state. The result is a scalable and accurate model of high-dimensional patient biomarkers as they vary over time. Our proposed model yields significant improvements in generalization and, on real-world clinical data, provides interpretable insights into the dynamics of cancer progression.

1. Introduction
Clinical biomarkers capture snapshots of a patient’s evolving disease state as well as their response to treatment. However, these data can be high-dimensional, exhibit missingness, and display complex nonlinear behaviour over time as a function of time-varying interventions. Good unsupervised models of such data are key to discovering new clinical insights. This task is commonly referred to as disease progression modeling (Alaa & van der Schaar, 2019; Elibol et al., 2016; Liu et al., 2015; Schulam & Saria, 2016; Severson et al., 2020; Venuto et al., 2016; Wang et al., 2014).

Reliable unsupervised models of time-varying clinical data find several uses in healthcare. One use case is enabling practitioners to ask and answer counterfactuals using observational data (Bica et al., 2020a; Pearl et al., 2009; Rubin, 1974). Other use cases include guiding early treatment decisions based on a patient’s biomarker trajectory, detecting drug effects in clinical trials (Mould et al., 2007), and clustering patterns in biomarkers that correlate with disease sub-type (Zhang et al., 2019b). To do these tasks well, understanding how a patient’s biomarkers evolve over time given a prescribed treatment regimen is vital, since a person’s biomarker profile is often the only observed proxy to their true disease state. Like prior work (Alaa & van der Schaar, 2019; Krishnan et al., 2017; Severson et al., 2020), we frame this problem as a conditional density estimation task, where our goal is to model the density of complex multivariate time-series conditional on time-varying treatments.

Representation learning exposes a variety of techniques for good conditional density estimation (Che et al., 2018; Choi et al., 2016; Miotto et al., 2016; Suresh et al., 2017). For sequential data, a popular approach has been to leverage black-box, sequential models (e.g. Recurrent Neural Networks (RNNs)), where a time-varying representation is used to predict clinical biomarkers. Such models are prone to overfitting, particularly on smaller clinical datasets. More importantly, such models often make simplistic assumptions on how time-varying treatments affect downstream clinical biomarkers; for example, one choice is to concatenate treatments to the model’s hidden representations (Alaa & van der Schaar, 2019; Krishnan et al., 2017). The assumption here is that the neural network learns how treatments influence the representation. We argue that this choice is a missed opportunity and better choices exist. Concretely, we aim to encourage neural models to learn representations that encode a patient’s underlying disease burden by specifying how these representations evolve due to treatment. We develop a new disease progression model that captures such insights by using inductive biases rooted in the biological mechanisms of treatment effect.

Inductive biases have been integral to the success of deep learning in other domains such as vision, text and audio. For example, convolutional neural networks explicitly learn representations invariant to translation or rotation of image data (Jaderberg et al., 2015; LeCun, 2012; Veeling et al., 2018), transformers leverage attention modules (Bahdanau et al., 2014; Vaswani et al., 2017) that mimic how human vision pays attention to various aspects of an image, and modified graph neural networks can explicitly incorporate laws of physics to generalize better (Seo & Liu, 2019). For some learning problems, the physics underlying the domain are often known, e.g. the laws of motion, and may be leveraged in the design of inductive biases (Anderson et al., 2019; Ling...
Covariates

models to characterize the progression of chronic diseases. Much work has been done across machine learning, pharmacology, statistics and biomedical informatics on building architectures that specify how representations respond to treatments. Patient Data (Bottom): Illustration of data from a chronic disease patient. Baseline (static) data typically consists of genomics, demographics, and initial labs. Longitudinal data typically includes laboratory values (e.g. serum IgG) and treatments (e.g. lenalidomide). Baseline data is usually complete, but longitudinal measurements are frequently missing at various time points.

Contributions: We present a new attention-based neural architecture, $\text{PK-PD}_{\text{Neural}}$, that captures the effect of drug combinations in representation space (Figure 1 [top]). It learns to attend over multiple competing mechanistic explanations of how a patient’s genetics, past treatment history, and prior disease state influence the representation to predict the next outcome. The architecture is instantiated in a state space model, SSM$_{\text{PK-PD}}$, and shows strong improvements in generalization compared to several baselines and prior state of the art. We demonstrate the model can provide insights into multiple myeloma progression. Finally, we release a disease progression benchmark dataset called ML-MMRF, comprising a curated, pre-processed subset of data from the Multiple Myeloma Research Foundation CoMMpass study (US National Institutes of Health, and others). Our model code can be found at https://github.com/clinicalml/ml_mmrf, and the data processing code can be found at https://github.com/clinicalml/ief.

2. Related Work

Much work has been done across machine learning, pharmacology, statistics and biomedical informatics on building models to characterize the progression of chronic diseases. Gaussian Processes (GPs) have been used to model patient biomarkers over time and estimate counterfactuals over a single intervention (Futoma et al., 2016; Schulam & Saria, 2017; Silva, 2016; Soleimani et al., 2017). In each of these cases, the focus is either on a single intervention per time point or on continuous-valued interventions given continuously, both strong assumptions for chronic diseases. To adjust for biases that exist in longitudinal data, Bica et al. (2020a); Lim et al. (2018) use propensity weighting to adjust for time-dependent confounders. However, they concatenate multi-variate treatments to patient biomarkers as input to RNNs; when data is scarce, such approaches have difficulty capturing how the hidden representations respond to treatment.

State space models and other Markov models have been used to model the progression of a variety of chronic diseases, including Cystic Fibrosis, scleroderma, breast cancer, COPD and CKD (Alaa & van der Schaar, 2019; Perotte et al., 2015; Schulam & Saria, 2016; Taghipour et al., 2013; Wang et al., 2014). There has also been much research in characterizing disease trajectories, subtypes, and correlations between risk factors and progression for patients suffering from Alzheimer’s Disease (Goyal et al., 2018; Khatami et al., 2019; Marinescu et al., 2019; Zhang et al., 2019a). Like us, the above works pose disease progression as density estimation but in contrast, many of the above models do not condition on time-varying interventions.

3. Background - State Space Models (SSMs)

SSMs are a popular model for sequential data and have a rich history in modeling disease progression.

Notation: $B \in \mathbb{R}^J$ denotes baseline data that are static, i.e. individual-specific covariates. For chronic diseases, these data comprise a $J$-dimensional vector, including patients’ age, gender, genetics, race, and ethnicity. Let $U = \{U_0, \ldots, U_{T-1}\}$; $U_t \in \mathbb{R}^L$ be a sequence of $L$-dimensional interventions for an individual. An element of $U_t$ may be binary, to denote prescription of a drug, or real-valued, to denote dosage. $X = \{X_1, \ldots, X_T\}$; $X_t \in \mathbb{R}^M$ denotes the sequence of real-valued, $M$-dimensional clinical biomarkers. An element of $X_t$ may denote a serum lab value or blood count, which is used by clinicians to measure organ function as a proxy for disease severity. $X_t$ frequently contains missing data. We assume access to a dataset $D = \{(X^1, U^1, B^1), \ldots, (X^N, U^N, B^N)\}$. For a visual depiction of the data, we refer the reader to Figure 1. Unless required, we ignore the superscript denoting the index of the datapoint and denote concatenation with $[]$.

Model: SSMs capture dependencies in sequential data via a time-varying latent state. When this latent state is discrete, SSMs are also known as Hidden Markov Models (HMM).
In our setting, we deal with a continuous latent state. The generative process is:

\[ p(X|U, B) = \int \prod_{t=1}^{T} p_{\theta}(Z_t|Z_{t-1}, U_{t-1}, B)p_{\theta}(X_t|Z_t)dZ \]

\[ Z_t|\sim N(\mu_{\theta}(Z_{t-1}, U_{t-1}, B), \Sigma_{\theta}(Z_{t-1}, U_{t-1}, B)), \]

\[ X_t|\sim N(\kappa_{\theta}(Z_t), \Sigma_{\theta}(Z_t)) \]  

(1)

We denote the parameters of a model by \( \theta \), which may comprise weight matrices or the parameters of functions that index \( \theta \). SSMs make the Markov assumption on the latent variables, \( Z_t \), and we assume that relevant information about past medications are captured by the state or contained in \( U_{t-1} \). We set \( \Sigma_{\theta}, \Sigma_{\theta}', \kappa_{\theta} \) to be functions of a concatenation of their inputs, e.g. \( \Sigma_{\theta}(\cdot) = \text{softplus}(W[Z_{t-1}, U_{t-1}, B] + b) \). \( \Sigma_{\theta}, \Sigma_{\theta}' \) are diagonal matrices where the softplus function is used to ensure positivity.

**Learning:** We maximize \( \sum_{i=1}^{N} \log p(X^i|U^i, B^i) \) with respect to \( \theta \). For a nonlinear SSM, this function is intractable, so we learn via maximizing a variational lower bound on it.

To evaluate the bound, we perform probabilistic inference using a structured inference network (Krishnan et al., 2017). The learning algorithm alternates between predicting variational parameters using a bi-directional recurrent neural network, evaluating a variational upper bound, and making gradient updates jointly with respect to the parameters of the generative model and the inference network.

When evaluating the likelihood of data under the model, if \( X_t \) is missing, it is marginalized out. Since the inference network also conditions on sequences of observed data to predict the variational parameters, we use forward fill imputation where data are missing.

### 4. Attentive Pharmacodynamic State Space Model

To make the shift from black-box models to those that capture useful structure for modeling clinical data, we begin with a discussion of PK-PD models and some of the key limitations that practitioners may face when directly applying them to modern clinical datasets.

#### 4.1. Limitations of Pharmacokinetic-Pharmacodynamic Modeling

Pharmacology is a natural store of domain expertise for reasoning about how treatments affect disease. We look specifically at pharmacokinetics (PK), which deals with how drugs move in the body, and pharmacodynamics (PD), which studies the body’s response to drugs. Consider a classical pharmacokinetic-pharmacodynamic (PK-PD) model used to characterize variation in tumor volume due to chemotherapy (Norton, 2014; West & Newton, 2017). Known as the log-cell kill model, it is based on the hypothesis that a given dose of chemotherapy results in killing a constant fraction of tumor cells rather than a constant number of cells. The original model is an ordinary differential equation but an equivalent expression is:

\[ S(t) = S(t-1) \cdot \left( 1 + \rho \log(K/S(t-1)) - \beta_c C(t) \right) \]  

(2)

\( S(t) \) is the (scalar) tumor volume, \( C(t) \) is the (scalar) concentration of a chemotherapeutic drug over time, \( K \) is the maximum tumor volume possible, \( \rho \) is the growth rate, and \( \beta_c \) represents the drug effect on tumor size. Besides its bespoke nature, there are some key limitations of this model that hinder its broad applicability for unsupervised learning:

- **Single intervention, single biomarker:** The model parameterizes the effect of a single, scalar intervention on a single, scalar, time-varying biomarker making it impossible to apply directly to high-dimensional clinical data. Furthermore, the quantity it models, tumor volume, is unobserved for non-solid cancers.

- **Misspecified in functional form:** The log-cell-kill hypothesis, by itself, is not an accurate description of the drug mechanism in most non-cancerous chronic diseases.

- **Misspecified in time:** Patients go through cycles of recovery and relapse during a disease. Even if the hypothesis holds when the patient is sick, it may not hold when the patient is in recovery.

In what follows, we aim to mitigate these limitations to build a practical, scalable model of disease progression.

#### 4.2. Latent Representations of Disease State

Tackling the first limitation, we use nonlinear SSMs in order to model longitudinal, high-dimensional data. Even though tumor volume may not be observed in observational clinical datasets, various proxies (e.g. lab values, blood counts) of the unobserved disease burden often are. We conjecture that the time-varying latent representation, \( Z_t \), implicitly captures such clinical phenotypes from the observations.

To ensure that the phenotypes captured by \( Z_t \) vary over time in a manner akin to clinical intuition, we focus the efforts of our design on the transition function, \( \mu_{\theta}(Z_{t-1}, U_{t-1}, B) \), of the state space model. This function controls the way in which the latent state \( Z_t \) in an SSM evolves over time (and through it, the data) when exposed to interventions, \( U_t \); this makes the transition function a good starting point for incorporating clinical domain knowledge.

#### 4.3. Neural Attention over Treatment Effect Mechanisms

In order to design a good transition function, we first need to address the second limitation that we may not know the
exact mechanism by which drugs affect the disease state. However, we often have a set of reasonable hypotheses about the mechanisms that underlie how we expect the dynamics of the latent disease state to behave.

Putting aside the specifics of what mechanisms we should use for the moment, suppose we are given \( d \) mechanism functions, \( g_1, \ldots, g_d \), each of which is a neural architecture that we believe captures aspects of how a representation should vary as a response to treatment. How a patient’s representation should vary will depend on what state the patient is in. e.g. sicker patients may respond less well to treatment than healthier ones.

To operationalize this insight, we make use of an attention mechanism (Bahdanau et al., 2014) to attend to which choice of function is most appropriate.

Attending over mechanisms of effect. Attention mechanisms operate by using a “query” to index into a set of “keys” to compute a set of attention weights, which are a distribution over the “values”. We propose a soft-attention mechanism to select between \( g_1, \ldots, g_d \). At each \( t \), for the query, we have \( q = Z_{t-1} W_q \).

\[
\hat{K} = [g_1(Z_{t-1}, U_{t-1}, B); \ldots; g_d(Z_{t-1}, U_{t-1}, B)]^T W_k \\
\hat{V} = [g_1(Z_{t-1}, U_{t-1}, B); \ldots; g_d(Z_{t-1}, U_{t-1}, B)]^T W_v.
\]

Note that \( W_q, W_k, W_v \in \mathbb{R}^{Q \times Q} \) and that \( q \in \mathbb{R}^{Q} \), \( \hat{K} \in \mathbb{R}^{Q \times d} \), and \( \hat{V} \in \mathbb{R}^{Q \times d} \). Then, we have the following,

\[
\mu_\theta(Z_{t-1}, U_{t-1}, B) = \left( \sum_{i=1}^{d} \text{softmax} \left( \frac{q \odot \hat{K}}{\sqrt{Q}} \right) \odot \hat{V}_i \right) W_v.
\]

4.4. Lines of Therapy with Local and Global Clocks

Here, we address a third limitation of classical PK-PD models: a proposed drug mechanism’s validity may depend on how long the patient has been treated and what stage of therapy they are in. Such stages, or lines of therapy, refer to contiguous plans of multiple treatments prescribed to a patient. They are often a unique structure of clinical data from individuals suffering from chronic diseases. For example, first line therapies often represent combinations prioritized due to their efficacy in clinical trials; subsequent lines may be decided by clinician preference. Lines of therapy index treatment plans that span multiple time-steps and are often laid out by clinicians at first diagnosis. We show how to make use of this information within a mechanism function.

To capture the clinician’s intention when prescribing treatment, we incorporate line of therapy as a one-hot vector in \( U_t[; K] \forall t \) (\( K \) is the maximal line of therapy). Lines of therapy typically change when a drug combination fails or causes adverse side effects. By conditioning on line of therapy, a transition function (of the SSM) parameterized by a neural network can, in theory, infer the length of time a patient has been on that line. However, although architectures such as Neural Turing Machines (Graves et al., 2014) can learn to count occurrences, they would need a substantial amount of data to do so.

To enforce the specified drug mechanism functions to capture time since change in line of therapy, we use clocks to track the time elapsed since an event. This strategy has precedent in RNNs, where Che et al. (2018) use time since the last observation to help RNNs learn well when data is missing. Koutnik et al. (2014) partition the hidden states in RNNs so they are updated at different time-scales. Here, we augment our interventional vector, \( U_t \), with two more dimensions. A global clock, \( gc \), captures time elapsed since \( T = 0 \), i.e. \( U_t[K] = gc = t \). A local clock, \( lc \), captures time elapsed since a line of therapy began; i.e.
We are the first to make use of lines-of-therapy information. Neural Log-Cell Kill: This architecture is inspired by the classical log cell kill model of tumor volume in solid cell tumors (West & Newton, 2017) but unlike the original model, scales to high-dimensional representations and takes into account lines of therapy via the local clock. This allows the model to effectively reset every time a new line of therapy begins. The functional form of the model is,

\[ g_2(Z_{t-1}, U_{t-1}, B) = Z_{t-1} \cdot (1 - \rho \log(Z_{t-1}^-)) - \beta \exp(-\delta \cdot l_{c_{t-1}})), \]

where \( \beta = \tanh(W_{b1}U_{t-1} + b_{lc}) \), \( W_{lc} \in \mathbb{R}^{Q \times L} \), \( b_{lc} \in \mathbb{R}^{Q} \), \( \delta \in \mathbb{R}^{Q} \) and \( \rho \in \mathbb{R}^{Q} \) are learned. While diseases may not have a single observation that characterizes the state of the organ system (akin to tumor volume), we hypothesize that representations, \( Z_{t} \), of the observed clinical biomarkers may benefit from mimicking the dynamics exhibited by tumor volume when exposed to chemotherapeutic agents. We emphasize that unlike Equation 2, the function in Equation 5 operates over a vector valued set of representations that can be modulated by the patient’s genetic markers.

Neural Treatment Exponential: Xu et al. (2016) develop a Bayesian nonparametric model to explain variation in creatinine, a single biomarker, due to treatment. We design an architecture inspired by their model that scales to high dimensional representations, allows for the representation to vary as a function of the patient’s genetics, and makes use of information in the lines of therapy via the clocks.

\[ g_3(Z_{t-1}, U_{t-1}, B) \]

where \( b_{lin} \in \mathbb{R}^{Q} \), \( W_{lin} \in \mathbb{R}^{Q \times (L+J)} \). Here, the effects on the representation are bounded (via the tanh function) but depend on the combination of drugs prescribed and the patient’s baseline data, including genetics.

Modeling slow, gradual relapse after treatment: One of the defining features of many chronic diseases is the possibility of a relapse during active therapy. In cancer, a relapse can happen due to cancerous cells escaping the treatment or a variety of other bio-chemical processes, such as increased resistance to treatment due to mutations. The relapse can result in bio-markers reverting to values that they held prior to the start of treatment; for an example of this, see Figure 2 (right). We design the following neural architectures to capture such patterns in a latent representation.

Neural Log-Cell Kill: This architecture is inspired by the classical log cell kill model of tumor volume in solid cell tumors (West & Newton, 2017) but unlike the original model,
is used as the transition function in an SSM, we refer to the
resulting model as SSM_PK_PD, a heterogeneous state space
model designed to model the progression of diseases.

5. Evaluation

5.1. Datasets

We study SSM_PK_PD on three different datasets – two here,
and on a third semi-synthetic dataset in the appendix.

**Synthetic Data:** We begin with a synthetic disease progres-
sion dataset where each patient is assigned baseline covari-
ates \( B \in \mathbb{R}^6 \). \( B \) determines how the biomarkers, \( X_t \in \mathbb{R}^2 \),
behave in the absence of treatment. \( U_t \in \mathbb{R}^3 \) comprises the
line of therapy (\( K = 2 \)), the local clock, and a single binary
variable indicating when treatment is prescribed. To mimic
the data dynamics described in Figure 1, the biomarkers
follow second-order polynomial trajectories over time with
the underlying treatment effect being determined by the
Neural Treatment Exponential (see Equation 6). Biomarker
1 can be thought of as a marker of disease burden, while
biomarker 2 can be thought of as a marker of biological
function. The full generative process for the data is in the
supplementary material. To understand generalization of
the model as a function of sample complexity, we train on
100/1000 samples and evaluate on five held-out sets of size
50000.

**ML-MMRF:** The Multiple Myeloma Research Foundation
(MMRF) CoMMpass study releases de-identified clinical
data for 1143 patients suffering from multiple myeloma, an
 incurable plasma cell cancer. All patients are aligned to the
start of treatment, which is made according to current stan-
dard of care (not random assignment). With an oncologist,
we curate demographic and genomic markers, \( B \in \mathbb{R}^{16} \),
clinical biomarkers, \( X_t \in \mathbb{R}^{16} \), and interventions, \( U_t \in \mathbb{R}^{9} \),
with one local clock, a three dimensional one-hot encoding
for line of therapy, and binary markers of 5 drugs. Our
results are obtained using a 75/25 train/test split. To select
hyperparameters, we perform 5-fold cross validation on the
training set. Finally, there is missingness in the biomarkers,
with 66% of the observations missing. We refer the reader
to the appendix for more details on the dataset.

5.2. Setup

We learn via: \((\text{arg min}_\theta - \log p(X|U, B; \theta))\) using ADAM
(Kingma & Ba, 2014) with a learning rate of 0.001 for
15000 epochs. L1 or L2 regularization is applied in one of
two ways: either we regularize all model parameters (in-
cluding parameters of inference network), or we regularize
all weight matrices except those associated with the attention
mechanism. We search over regularization strengths of
0.01, 0.1, 1, 10 and latent dimensions of 16, 48, 64 and
128. We do model selection using the negative evidence
lower bound (NELBO); Appendix B contains details on the
derivation of this bound. We use a single copy of \( g_1, g_2, \) and
\( g_3 \) in the transition function. Multiple copies of each
function as well as other “mechanistic” functions can be
used, highlighting the flexibility of our approach. However,
this must be balanced with potentially overfitting on small
datasets.

5.3. Baselines

**SSM_{Linear}** parametrizes \( \mu_{\theta}(Z_t, U_{t-1}, B) \) with a linear
function. This model is a strong, linear baseline whose
variants have been used for modeling data of patients suffer-
ing from Chronic Kidney Disease (Perotte et al., 2015).

**SSM_{NL}:** Krishnan et al. (2017) use a nonlinear SSM to cap-
ture variation in the clinical biomarkers of diabetic patients.
We compare to their model, parameterizing the transition
function with a 2-layer MLP.

**SSM_{Attn.Hist}:** We use an SSM whose transition function is pa-
rameterized via a Mixture-of-Experts (MoE) architecture
(Jacobs et al., 1991; Jordan & Jacobs, 1994); i.e. \( g_1, g_2, g_3 \)
are each replaced with a multi-layer perceptron. This base-
line does not incorporate any domain knowledge and tests
the relative benefits of prescribing the functional forms via
mechanisms versus learning them from data.

**SSM_{Attn.Hist}:** We implement a variant of the SSM in Alaa &
van der Schaar (2019), a state-of-the-art model for disease
progression trained via conditional density estimation. The
authors use a discrete state space for disease progression
modeling making a direct comparison difficult. However,
SSM_{Attn.Hist} preserves the structural modeling assumptions
they make. Namely, the transition function of the model at-
tends to a concatenation of previous states and interventions
at each point in time. We defer specifics to Appendix B.

In addition, we run two simpler baselines, a First Order
Markov Model (FOMM) and Gated Recurrent Unit (GRU)
(Cho et al., 2014), on the synthetic data and ML-MMRF but
defer those results to Appendix E.

5.4. Evaluation Metrics

**NELBO** On both the synthetic data and ML-MMRF data,
we quantify generalization via the negative evidence lower
bound (NELBO), which is a variational upper bound on
the negative log-likelihood of the data. A lower NELBO
indicates better generalization.

**Pairwise Comparisons** For a fine-grain evaluation of our
models on ML-MMRF, we compare held-out NELBO under
SSM_{PK_PD} versus the corresponding baseline for each pa-

tient. For each held-out point, \( \Delta_i = 1 \) when the NELBO of
that datapoint is lower under SSM_{PK_PD} and \( \Delta_i = 0 \) when
it is not. In Table 1 (bottom), we report \( \frac{1}{N} \sum_{i=1}^{N} \Delta_i \), the
proportion of data for which $SSM_{PK-PD}$ yields better results.

**Counts** To get a sense for the number of patients on whom $SSM_{PK-PD}$ does much better, we count the number of held-out patients for whom the held-out negative log likelihood (computed via importance sampling) is more than 10 nats lower under $SSM_{PK-PD}$ than the corresponding baseline (and vice versa for the baselines).

### 5.5. Results

We investigate three broad categories of questions.

#### 5.5.1. Generalization under different conditions

$SSM_{PK-PD}$ generalizes better in setting with few ($\sim 100$) samples. Table 1 (top) depicts NELBOs on held-out synthetic data across different models, where a lower number implies better generalization. We see a statistically significant gain in generalization for $SSM_{PK-PD}$ compared to all other baselines. $SSM_{NL}$, $SSM_{MOE}$ overfit quickly on 100 samples but recover their performance when learning with 1000 samples.

$SSM_{PK-PD}$ generalizes well when it is misspecified. Because we often lack prior knowledge about the true underlying dynamics in the data, we study how $SSM_{PK-PD}$ performs when it is misspecified. We replace the Neural Treatment Exponential function, $g_3$, from $PK-PD_{Neural}$ with another instance of $g_1$. The resulting model is now misspecified since $g_3$ is used to generate the data but no longer lies within the model family. We denote this model as (SSM $PK-PD$ w/o TExp). In Table 1 (top), when comparing the sixth column to the others, we find that we outperform all baselines and get comparable generalization to $SSM_{PK-PD}$ with the Neural Treatment Exponential function. This result emphasizes our architecture’s flexibility and its ability to learn the underlying (unknown) intervention effect through a combination of other, related mechanism functions.

$SSM_{PK-PD}$ generalizes well on real-world patient data. A substantially harder test of model misspecification is on the ML-MMRF data where we have unknown dynamics that drive the high-dimensional (often missing) biomarkers in addition to combinations of drugs prescribed over time. To rigorously validate whether we improve generalization on ML-MMRF data with $SSM_{PK-PD}$, we study model performance with respect to the three metrics introduced in Section 5.4. We report our results in Table 1 (bottom). First, we consistently observe that a high fraction of patient data in the test set are explained better by $SSM_{PK-PD}$ than the corresponding baseline (pairwise comparisons). We also note that out of 282 patients in the test set, across all the baselines, we find that the $SSM_{PK-PD}$ generalizes better for many more patients (counts). Finally, $SSM_{PK-PD}$ has lower NELBO averaged across the entire test set compared to all baselines.

#### 5.5.2. Model complexity & generalization

The improvements of $SSM_{PK-PD}$ are consistent taking model sizes into account. We show in Table 1 (bottom) the number of parameters used in each model. We find that more parameters do not imply better performance. Models with the most parameters (e.g. $SSM_{NL}$) overfit while those with the lowest number of parameters underfit (e.g. $SSM_{Linear}$) suggesting that the gains in generalization that we observe are coming from our parameterization. We experimented with increasing the size of the $SSM_{Linear}$ model (via the latent variable dimension) to match the size of the best PK-PD model. We found that doing so did not outperform the held-out likelihood of $SSM_{PK-PD}$.

When data are scarce, a Mixture of Experts architecture is difficult to learn: How effective are the functional forms of the neural architectures we develop? To answer this question, we compare the held-out log-likelihood of $SSM_{PK-PD}$ vs $SSM_{MOE}$ in the third column of Table 1 (bottom). In the ML-MMRF data, we find that the $SSM_{PK-PD}$ outperforms the $SSM_{MOE}$. We suspect this is due to the fact that learning diverse “experts” is hard when data is scarce and supports the hypothesis that the judicious choice of neural architectures plays a vital role in capturing biomarker dynamics.

Can $PK-PD_{Neural}$ be used in other model families? In the supplement, we implement $PK-PD_{Neural}$ in a first-order Markov model and find similar improvements in generalization on the ML-MMRF dataset. This result suggests that the principle we propose of leveraging domain knowledge from pharmacology to design mechanism functions can allow other kinds of deep generative models (beyond SSMs) to also generalize better when data are scarce.

#### 5.5.3. Visualizing patient dynamics

In Figure 4 (right), to further validate our initial hypothesis that the model is using the various neural PK-PD effect func-
Neural Pharmacodynamic State Space Modeling

Table 1. Top: Synthetic data: Lower is better. We report the test NELBO with std. dev. for each SSM model to study generalization in the synthetic setting. Bottom: ML-MMRF: Pairwise Comparison: each number is the fraction (with std. dev.) of test patients for which $\text{SSM}_{\text{PK-PD}}$ has a lower NELBO than an SSM that uses a different transition function. Counts: We report the number of test patients (out of 282) for which an SSM model (PK-PD or otherwise) has a greater than 10% difference in negative log likelihood compared to the alternative model. NELBO: We report the test NELBO of each model. Note that we label the metrics associated with $\text{SSM}_{\text{PK-PD}}$ and $\text{SSM}_{\text{Linear}}$ in the first column, but drop these labels in subsequent columns.

Figure 4. ML-MMRF: (Left two) We visualize the TSNE representations of each test patient’s latent state, $Z_t$, at the start of treatment and three years in. (Right) For $\text{SSM}_{\text{PK-PD}}$, we visualize the attention weights, averaged over all time steps and all patients, on each of the neural effect functions across state space dimensions.

How do the clocks help model patient dynamics? Figure 3 shows samples from three SSMs trained on synthetic data. $\text{SSM}_{\text{PK-PD}}$ captures treatment response accurately while $\text{SSM}_{\text{Linear}}$ does not register that the effect of treatment can persist over time. To study the impact of clocks on the learned model, we perform an ablation study on SSMs where the local clock in $U_t$, used by $\text{PK-PD}_{\text{Neural}}$, is set to a constant. Without clocks ($\text{PK-PD}$ w/o lc), the model does not capture the onset or persistence of treatment response.

$\text{SSM}_{\text{PK-PD}}$ learns latent representations that reflect the patient’s disease state: In ML-MMRF, we restrict the patient population to those with at least $T = 36$ months of data. At two different points during their treatment of the disease, we visualize the result of TSNE (Maaten & Hinton, 2008) applied to their latent representations in Figure 4 (left).

Early in their treatment, the latent representations of these patients appear to have no apparent structure. As time progresses, we find that the dimensions split into two groups. One group, for the most part, is still being treated, while the other is not being treated. A deeper dive into the untreated patients reveals that this cohort has a less severe subtype of myeloma (via a common risk assessment method known as ISS staging). This result suggests that the latent state of $\text{SSM}_{\text{PK-PD}}$ has successfully captured the coarse disease severity of patients at particular time points.

Visualizing patient samples from $\text{SSM}_{\text{PK-PD}}$: Figure 5 shows the average of three samples from $\text{SSM}_{\text{Linear}}$ and $\text{SSM}_{\text{PK-PD}}$ trained on ML-MMRF. We track two biomarkers used by...
clinicians to map myeloma progression. SSM\textsubscript{PK-PD} better captures the evolution of these biomarkers conditioned on treatment. For serum IgG, SSM\textsubscript{PK-PD} correctly predicts the relapse of disease after stopping first line therapy, while SSM\textsubscript{Linear} does not. On the other hand, for serum lambda, SSM\textsubscript{PK-PD} correctly predicts it will remain steady.

6. Discussion

PK-PD\textsubscript{Neural} leverages domain knowledge from pharmacology in the form of treatment effect mechanisms to quantitatively and qualitatively improve performance of a representation-learning based disease progression model. Bica et al. (2020b) note the potential for blending ideas from pharmacology with machine learning: our work is among the first to do so.

We highlight several avenues for future work. For example, in machine learning for healthcare, the model we develop can find use in practical problems such as forecasting high-dimensional clinical biomarkers and learning useful representations of patients that are predictive of outcomes. Doing so can aid in the development of tools for risk stratification or in software to aid in clinical trial design. Applying the model to data from other chronic diseases such as diabetes, congestive heart failure, small cell lung cancer and rheumatoid arthritis, brings opportunities to augment PK-PD\textsubscript{Neural} with new neural PK-PD functions tailored to capture the unique biomaker dynamics exhibited by patients suffering from these diseases.

Finally, we believe PK-PD\textsubscript{Neural} can find use in the design of parameteric environment simulators for different domains. In pharmacology, such simulation based pipelines can help determine effective drug doses (Hutchinson et al., 2019). Our idea of attending over multiple dynamics functions can find use in the design of simulators in domains such as economics, where multiple hypothesized mechanisms are used to explain observed market phenomena (Ghosh et al., 2019).

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Supplementary Material

The supplementary material contains the following sections. For each section, we highlight the key findings about the experiments we conduct.

A. Learning Algorithms: This section expands upon the learning algorithm for SSM\textsubscript{PK-PD} in the main paper. We also describe two additional sequential models – a First Order Markov Model (FOMM) and a Gated Recurrent Neural Network (GRU).

B. Synthetic Dataset: This section provides an in-depth description of the generative process that underlies the synthetic dataset used in the experimental section.

C. The Multiple Myeloma Research Foundation CoMMpass Study: This section provides details on data extraction, pre-processing and construction of the ML-MMRF dataset.

D. FOMM and GRU Experiments: We study how incorporating a variant of PK-PD\textsubscript{Neural} into a FOMM and GRU improves model generalization. The key take-away from this section, with supporting evidence in Table 2, is that SSM\textsubscript{PK-PD} improves generalization not just in state space models, but also in other popular choices of neural network based models of sequences such as FOMMs and GRUs.

E. Semi-synthetic Experiments: We introduce a semi-synthetic dataset that we use to further evaluate SSM\textsubscript{PK-PD}. The key take-away from this section, with supporting evidence in Table 3, is that SSM\textsubscript{PK-PD} improves generalization on a new dataset whose sequential patterns mimic real-world multiple myeloma data. These improvements are confirmed in a model misspecification scenario.

F. Additional Experiments: This section details additional experiments to interpret the model we develop and understand the relative utility of its various parts.

F1. Patient Forecasting - We explore different ways in which SSM\textsubscript{PK-PD} may be used to forecast patient trajectories given some initial data. When conditioning on different lengths of patient history and then sampling forward in time, we see a qualitative improvement in samples from SSM\textsubscript{PK-PD} compared to one of the best performing baselines.

F2. Visualizing Disease Progression - We extend our analysis of the SSM\textsubscript{PK-PD}’s latent states to studying how they evolve over the entire disease course. We find that clustering patients based on the latent state reveals subgroups that, due to differences in disease severity, have been assigned different treatment regimens. This result suggests that the latent representation has encoded the patient’s underlying disease state.

F3. Per-feature Breakdown - We perform a per-feature analysis of how well SSM\textsubscript{PK-PD} and SSM\textsubscript{Linear} model different clinical biomarkers, finding that SSM\textsubscript{PK-PD} does particularly well for important markers of progression, such as serum IgA.

F4. Ablation Analysis - We study which treatment mechanism function yields the most benefit for modeling the ML-MMRF dataset. Our analysis finds that the Neural Treatment Exponential function provides the most differential gains in NELBO and that the time-varying treatments are crucial for accurately modeling the dynamics of serums IgA, IgG, and Lambda.
A. Learning Algorithms

We implement all the models that we experiment with in PyTorch (Paszke et al., 2019).

State Space Models Recall that the generative process is:

\[ p(X|U, B) = \int \prod_{t=1}^{T} p(Z_t|Z_{t-1}, U_{t-1}, B; \theta) p(X_t|Z_t; \theta) dZ \]

where the transition function, \( \mu_\theta \), and variational parameters, \( \phi \), we use forward fill imputation where \( X \) is missing, then it depends on their entire history of latent states and interventions. Thus, we have,

\[ p(X|U, B) = \]

Maximum Likelihood Estimation of \( \theta \): Since the log likelihood \( p(X|U, B) \) is difficult to evaluate and maximize directly due to the high-dimensional integral, we resort to a variational learning algorithm that instead maximizes a lower bound on the log-likelihood to learn the model parameters, \( \theta \). We make use of a structured inference network (Krishnan et al., 2017) that amortizes the variational approximation, \( q_\phi(Z|X) \), to the posterior distribution, \( p_\theta(Z|X) \), of each datapoint.

\[
\log p(X|U, B; \theta) \geq \mathcal{L}(X; (\theta, \phi)) \tag{7}
\]

\[
= \sum_{t=1}^{T} \mathbb{E}_{q_\phi(Z_t|X, U, B)}[\log p_\theta(X_t|Z_t)] \\
- \text{KL}(q_\phi(Z_1|X, U, B)|p_\theta(Z_1|B)) \\
- \sum_{t=2}^{T} \mathbb{E}_{q_\phi(Z_{t-1}|X, U, B)}[ \text{KL}(q_\phi(Z_t|Z_{t-1}, X, U)|p_\theta(Z_t|Z_{t-1}, U_{t-1}, B))] \\
\]

The lower bound on the log-likelihood of data, \( \mathcal{L}(X; (\theta, \phi)) \), is a differentiable function of the parameters \( \theta, \phi \) (Krishnan et al., 2017), so we jointly learn them via gradient ascent. As mentioned in the main paper, if \( X_t \) is missing, then it is marginalized out when evaluating the likelihood of the data under the model. Since the inference network also conditions on sequences of observed data to predict the variational parameters, we use forward fill imputation where data are missing.

Hyperparameters: We present the results of the hyperparameter search on the datasets that we study. Please see the evaluation section of the main paper for the specific ranges that we searched over.

- **SSM\text{Linear}**
  1. Synthetic: State space dimension 48, L2 regularization on all parameters with strength 0.01
  2. ML-MMRF: State space dimension 16, L2 regularization on all parameters with strength 0.01

- **SSM\text{NL}**
  1. Synthetic: State space dimension 48, hidden layer dimension 300, L1 regularization on all parameters with strength 0.1
  2. ML-MMRF: State space dimension 48, hidden layer dimension 300, L2 regularization on all parameters with strength 0.1

- **SSM\text{PK-PD}**
  1. Synthetic: State space dimension 48, L1 regularization on subset of parameters with strength 0.01
  2. ML-MMRF: State space dimension 48, L1 regularization on all parameters with strength 0.01

- **SSM\text{MOE}**
  1. Synthetic: State space dimension 16, hidden layer dimension 300, L1 regularization on all parameters with strength 0.01
  2. ML-MMRF: State space dimension 48, hidden layer dimension 300, L1 regularization on all parameters with strength 0.01

**SSM Attn. Hist.**

1. Synthetic: State space dimension 16, hidden layer dimension 300, L1 regularization on all parameters with strength 0.01
2. ML-MMRF: State space dimension 48, hidden layer dimension 300, L1 regularization on all parameters with strength 0.01

**SSM Attn. Hist. Baseline:** We provide details on the SSM architecture proposed by (Alaa & van der Schaar, 2019) for disease progression modeling. The generative process of their architecture differs from a normal state space model in that the transition function, \( \mu_\theta \), assumes that the patient’s latent state at time \( t \) depends on their entire history of latent states and interventions. Thus, we have,

\[
\int \prod_{t=1}^{T} p(Z_t|Z_{t-1}, U_{1:t-1}, B; \theta)p(X_t|Z_t; \theta) dZ \\
Z_t| \sim N(\mu_\theta(Z_{1:t-1}, U_{1:t-1}, B), \Sigma_\phi(Z_{1:t-1}, U_{1:t-1}, B)) \\
X_t| \sim N(\kappa_\theta(Z_t), \Sigma_\phi(Z_t)) 
\]
Neural Pharmacodynamic State Space Modeling

latent states to compute the current latent state. These attention weights are a function of a patient’s entire clinical lab and treatment history. Therefore, the transition function that we use to capture their modeling assumptions is as follows:

$$
\mu_\theta(Z_{1:t-1}, \alpha_{1:t-1}) = W_h \left( \sum_{i=1}^{t-1} \alpha_i \odot Z_i \right) + b_h,
$$

where $$\alpha_{1:t-1} = A_t(X_{1:t-1}, U_{1:t-1})$$ via an attention mechanism, $$A_t$$. We use a bi-directional recurrent neural network for the inference network, as opposed to the authors’ proposed attentive inference network. We argue that the bi-RNN is just as expressive, since the variational parameters are a function of all past and future observations. Moreover, our goal is to study the effect of altering the generative model in this work.

We also experiment with First Order Markov Models (FOMM) and Gated Recurrent Units (GRU) (Chung et al., 2014), which we detail below.

**First Order Markov Models**  FOMMs assume observations are conditionally independent of the past given the previous observation, intervention and baseline covariates. The generative process is:

$$
p(X|U, B) = \prod_{t=1}^{T} p(X_t|X_{t-1}, U_{t-1}, B);
$$

$$
X_t|\sim N(\mu_\theta(X_{t-1}, U_{t-1}, B), \Sigma_\theta(X_{t-1}, U_{t-1}, B)),
$$

where the transition function, $$\mu_\theta$$, differs akin to the transition function of SSM models, as described in the main paper. Here, we will experiment with FOMM_{Linear}, FOMM_{NL}, FOMM_{MDOE}, & FOMM_{PK-PD}

**PK-PD_{Neural} for FOMM_{PK-PD}:** We will use a simpler variant of the PK-PD_{Neural} formulation introduced in the main paper as a proof of concept. Namely, we have,

$$
\mu_\theta(X_{1:t-1}, U_{1:t-1}, B) = \sum_{i=1}^{d} \sigma(\delta)_i \odot g_i(S_{t-1}, U_{t-1}, B),
$$

where each $$\delta$$ is a learned vector of weights and $$\sigma$$ refers to a softmax on the weights. Note that the PK-PD_{Neural} introduced in the main paper is a generalization of Equation 10; the primary difference is that the attention mechanism allows the weights to be a function of the prior state, which enables the weights to change over time.

**Maximum Likelihood Estimation of $$\theta$$:** We learn the model by maximizing $$\max_\theta \log p(X|U, B)$$. Using the factorization structure in the joint distribution of the generative model, we obtain:

$$
\log p(X|U, B) = \sum_{t=1}^{T} \log p(X_t|X_{t-1}, U_{t-1}, B).
$$

Each $$\log p(X_t|X_{t-1}, U_{t-1}, B)$$ is estimable as the log-likelihood of the observed multi-variate $$X_t$$ under a Gaussian distribution whose (diagonal) variance is a function $$\Sigma_\theta(X_{t-1}, U_{t-1}, B)$$ and whose mean is given by the transition function, $$\mu_\theta(X_{t-1}, U_{t-1}, B)$$. Since each $$\log p(X_t|X_{t-1}, U_{t-1}, B)$$ is a differentiable function of $$\theta$$, its sum is differentiable as well, and we may use automatic differentiation to derive gradients of the log-likelihood with respect to $$\theta$$ in order to perform gradient ascent. When any dimension of $$X_t$$ is missing, that dimension’s log-likelihood is ignored (corresponding to marginalization over that random variable) during learning.

**Hyperparameters:** We present the results of the hyperparameter search on the datasets that we study.

- **FOMM_{Linear}**
  1. **Synthetic:** L1 regularization on all parameters with strength 0.1
  2. **ML-MMRF:** L1 regularization on all parameters with strength 0.1

- **FOMM_{NL}**
  1. **Synthetic:** Hidden layer dimension 200, L1 regularization on all parameters with strength 0.1
  2. **ML-MMRF:** Hidden layer dimension 300, L1 regularization on all parameters with strength 0.1

- **FOMM_{PK-PD}**
  1. **Synthetic:** L1 regularization on subset of parameters with strength 0.1
  2. **ML-MMRF:** L1 regularization on subset of parameters with strength 0.1

**Gated Recurrent Neural Network (GRUs):** GRUs (Cho et al., 2014) are auto-regressive models of sequential observations i.e. $$p(X|U, B) = \prod_{t=1}^{T} p(X_t|X_{<t}, U_{<t}, B)$$. GRUs use an intermediate hidden state $$h_t \in \mathbb{R}^H$$ at each time-step as a proxy for what the model has inferred about the sequence of data until $$t$$. The GRU dynamics govern how $$h_t$$ evolves via an update gate $$F_t$$, and a reset gate $$R_t$$:

$$
F_t = \sigma(W_z \cdot [X_t, U_t, B] + V_z h_{t-1} + b_z),
$$

$$
R_t = \sigma(W_r \cdot [X_t, U_t, B] + V_r h_{t-1} + b_r)
$$

$$
h_t = F_t \odot h_{t-1} + (1 - F_t) \odot \tanh(W_h \cdot [X_t, U_t, B] + V_h (R_t \odot h_{t-1}) + b_h)
$$

$$
\theta = \{ W_z, W_r, W_h, V_z, V_r, V_h \in \mathbb{R}^{H \times (M + L + J)}; V_z, V_r, V_h \in \mathbb{R}^{H \times H}; b_z, b_r, b_h \in \mathbb{R}^H \}$$ are learned parameters and $$\sigma$$ is the sigmoid function. The effect of interventions may be felt in any of the above time-varying representations and so the “transition function” in the GRU is distributed across
Hyperparameters: We present the results of the hyperparameter search on the datasets that we study.

- **GRU**
  1. **Synthetic**: Hidden layer dimension 500, L2 regularization on all parameters with strength 0.1
  2. **ML-MMRF**: Hidden layer dimension 250, L2 regularization on all parameters with strength 0.1

- **GRU**
  1. **Synthetic**: Hidden layer dimension 500, L2 regularization on subset of parameters with strength 0.01
  2. **ML-MMRF**: Hidden layer dimension 500, L2 regularization on subset of parameters with strength 0.01

B. Synthetic Dataset

Below, we outline the general principles that the synthetic data we design is based on:

- We sample six random baseline values from a standard normal distribution.
- Two of the six baseline values determine the natural (untreated) progression of the two-dimensional longitudinal trajectories. They do so as follows: depending on which quadrant the baseline data lie in, we assume that the patient has one of four subtypes.
- Each of the four subtypes typifies different patterns by which the biomarkers behave such as whether they both go up, both go down, one goes up, one goes down etc. To see a visual example of this, we refer the reader to Figure 6 (left).

**Baseline** The generative process for the baseline covariates is \( B \sim N(0; I) \), \( B \in \mathbb{R}^6 \).

**Treatments (Interventions)**: There is a single drug (denoted by a binary random variable) that may be withheld (in the first line of therapy) or prescribed in the second line of therapy. For each patient \( d_t \sim \text{Unif.}[0, 18] \) denotes when the single drug is administered (and the second line of treatment begins). \( d_t \) is the point at which the local clock resets. We can summarize the generative process for the treatments as follows:

\[
\begin{align*}
    d_t & \sim \text{Unif.}[0, 18] \\
    U_t = 0 & \text{ if } t < d \text{ and } 1 \text{ otherwise} \\
    \text{line}_t[0] = 1 & \text{ if } t < d \text{ and } 0 \text{ otherwise} \\
    \text{line}_t[1] = 0 & \text{ if } t < d \text{ and } 1 \text{ otherwise} \\
    \end{align*}
\]

(12)

where \( \text{line}_t[0] \), \( \text{line}_t[1] \) denote the one-hot encoding for line of therapy. Next, we use the Neural Treatment Exponential (Equation 6 in the main paper) to generate a treatment response in the data. The functional form of the Treatment Exponential function (TE) is re-stated below for convenience,

\[
\text{TE}(\text{lc}_t) = \begin{cases}
    b_0 + \alpha_1/[1 + \exp(-\alpha_2(\text{lc}_t - \frac{\gamma_1}{2}))], & \text{if } 0 \leq \text{lc}_t < \gamma_1 \\
    b_1 + \alpha_0/[1 + \exp(\alpha_3(\text{lc}_t - \frac{3\gamma_1}{2}))], & \text{if } \text{lc}_t \geq \gamma_1 \\
\end{cases}
\]

(13)

The parameters that we use to generate the data are: \( \alpha_2 = 0.6, \alpha_3 = 0.6, \gamma_1 = 2, b_1 = 3 \), and \( \alpha_1 = [10, 5, -5, -10] \), which we vary based on patient subtype. We set \( \alpha_0 = (\alpha_1 + 2b_0 - b_1)/(1 + \exp(-\alpha_3\gamma_1)/2) \) to ensure that the treatment effect peaks at \( t = \text{lc}_t + \gamma_1 \) and \( b_0 = -\alpha_1/(1 + \exp(\alpha_2 \cdot \gamma_1/2)) \) for attaining \( \text{TE}(0) = 0 \).
Neural Pharmacodynamic State Space Modeling

Figure 6. Visualization of synthetic data: Left: A visualization of "patient" s baseline data (colored and marked by patient subtype). Right four plots: Examples of patient’s longitudinal trajectories along with treatment response. The blue and green longitudinal data denote two different patient biomarkers. The solid blue and green lines are what the trajectories would be with no intervention whereas the dotted blue and green lines are what the trajectories are with an intervention. Gray-dotted line represents an intervention. The subtypes may, optionally, be correlated with patient outcomes as highlighted using the values of $y$.

**Biomarkers:** We are now ready to describe the full generative process of the longitudinal biomarkers.

$$
B_1\ldots 6 \sim \mathcal{N}(0; I),
$$

$$
f_d(t) = 2 - 0.05t - 0.005t^2, \quad f_u(t) = -1 + 0.0001t + 0.005t^2,
$$

$$
X_1(t); X_2(t) = \begin{cases} 
  f_d(t) + \text{TE}(lc_1) + \mathcal{N}(0, 0.25); & f_d(t) + \text{TE}(lc_1) + \mathcal{N}(0, 0.25), B_1 \geq 0, B_2 \geq 0 \text{ if subtype 1} \\
  f_d(t) + \text{TE}(lc_1) + \mathcal{N}(0, 0.25); & f_u(t) + \text{TE}(lc_1) + \mathcal{N}(0, 0.25), B_1 \geq 0, B_2 < 0 \text{ if subtype 2} \\
  f_u(t) + \text{TE}(lc_1) + \mathcal{N}(0, 0.25); & f_u(t) + \text{TE}(lc_1) + \mathcal{N}(0, 0.25), B_1 < 0, B_2 \geq 0 \text{ if subtype 3} \\
  f_u(t) + \text{TE}(lc_1) + \mathcal{N}(0, 0.25); & f_u(t) + \text{TE}(lc_1) + \mathcal{N}(0, 0.25), B_1 < 0, B_2 < 0 \text{ if subtype 4} 
\end{cases}
$$

Intuitively, the above generative process captures the idea that without any effect of treatment, the biomarkers follow the patterns implied by the subtype (encoded in the first two dimensions of the baseline data). However the effect of interventions is felt more prominently after the $d$, the random variable denoting time at which treatment was prescribed.

**C. The Multiple Myeloma Research Foundation CoMMpass Study**

Here, we elaborate upon the data made available by the Multiple Myeloma Research Foundation in the IA13 release of data. We will make code available to go from the files released by the MMRF study to numpy tensors that may be used in any machine learning framework.

**Inclusion Criteria:** To enroll in the CoMMpass study, patients must be newly diagnosed with symptomatic multiple myeloma, which coincides with the start of treatment. Patients must be eligible for treatment with an immunomodulator or a proteasome inhibitor, two of the most common first line drugs, and they must begin treatment within 30 days of the baseline bone marrow evaluation (US National Institutes of Health, and others).

**C.1. Features**

**Genomic Data:** RNA-sequencing of CD38+ bone marrow cells was available for 769 patients. Samples were collected at initiation into the study, pre-treatment. For these patients, we used the Seurat package version 2.3.4 (Butler et al., 2018) in R to identify variable genes, and we then limit downstream analyses to these genes. We use principal component analysis (PCA) to further reduce the dimensionality of the data. The projection of each patient’s gene expression on to the first 40 principal components serves as the genetic features used in our model.

**Baseline Data:** Baseline data includes PCA scores, lab values at the patient’s first visit, gender, age, and the revised ISS stage. The baseline data also includes binary variables detailing the patient’s myeloma subtype, including whether or not they have heavy chain myeloma, are IgG type, IgA type, IgM type, kappa type, or lambda type. Additionally, several labs are measured at baseline, as well as longitudinally at subsequent visits. We detail these labs in the next sub-section. The genetic and baseline data jointly comprise $B$.

**Longitudinal Data:** Longitudinal data is measured approximately every 2 months and includes lab values and treatment information.

The biomarkers are real-valued numbers whose values evolve over time. They include: absolute neutrophil count ($10^3/\text{l}$), albumin (g/l), blood urea nitrogen (mmol/l), calcium (mmol/l), serum creatinine (umol/l), glucose (mmol/l),
hemoglobin (mmol/l), serum kappa (mg/dl), serum m protein (g/dl), platelet count $10^9$/l, total protein (g/dl), white blood count $10^9$/l, serum IgA (g/l), serum IgG (g/l), serum IgM (g/l), serum lambda (mg/dl).

Treatment information includes the line of therapy (we group all lines beyond line 3 as line 3+) the patient is on at a given point in time, and the local clock denoting the time elapsed since the last line of therapy. We also include the following treatments as (binary, indicating prescription) features in our model: lenalidomide, dexamethasone, cyclophosphamide, carfilzomib, bortezomib. The aforementioned are the top five drugs by frequency in the MMRF dataset. This dataset has significant missingness, with $\sim 66\%$ of the longitudinal markers missing. In addition, there is right censorship in the dataset, with around 25% of patients getting censored over time.

C.2. Data Processing

Longitudinal biomarkers $X$: Labs are first clipped to five times the median value to correct for outliers or data errors in the registry. They are then normalized to their healthy ranges (obtained via a literature search) as (unnormalized labs - healthy maximum value), and then multiplied by a lab-dependent scaling factor to ensure that most values lie within the range $[-8, 8]$. Missing values are represented as zeros, but a separate mask tensor, where 1 denotes observed and 0 denotes missing, is used to marginalize out missing variables during learning.

Baseline $B$: The biomarkers in the baseline are clipped to five times their median values. Patients without gene expression data (in the PCA features) are assigned the average normalized PCA score of their five nearest neighbors, using the Minkowski distance metric calculated on FISH features, ISS stage, and age.

D. FOMM and GRU Experiments

PK-PD$_{\text{Neural}}$ improves generalization in both FOMM and GRU models on synthetic data: Table 2 (bottom) depicts negative log-likelihoods on held-out synthetic data across different models, where a lower number implies better generalization. The non-linearity of the synthetic data makes unsupervised learning a challenge for FOMM$_{\text{Linear}}$ at 50 samples, allowing FOMM$_{PK-PD}$ to easily outperform it. In contrast, FOMM$_{NL}$ can capture non-linearities in the data, making it a strong baseline even at 50 samples. Yet, FOMM$_{PK-PD}$ outperforms it, emphasizing the utility of PK-PD$_{\text{Neural}}$ in when data is scarce. At 1000 samples, FOMM$_{NL}$ is able to learn enough about the dynamics to improve its performance relative to FOMM$_{PK-PD}$. GRU is a strong model on this dataset, but at both sample sizes, the GRU$_{PK-PD}$ improves generalization.

FOMM and GRU generalization performance on ML-MMRF: We observe improvements in generalization across FOMMs with the use of PK-PD$_{\text{Neural}}$. However, we do not see discernible gains from GRU$_{PK-PD}$, perhaps due to missingness in the data, which also results in the GRU's generalizing worse than SSM models based on negative log likelihood (Table 2 (top)). Overall, the GRU, due to the distributed nature of its "transition function", is a harder model to use PK-PD$_{\text{Neural}}$ in, although figuring how to do so effectively is a valuable direction for future work.
Neural Pharmacodynamic State Space Modeling

| Dataset      | Held-out Neg Log Likelihood | FOMM Linear       | FOMM Nonlinear     | FOMM PK-PD       | FOMM MOE        | GRU PK-PD       | SSM PK-PD (NELBO) |
|--------------|-----------------------------|-------------------|-------------------|-----------------|----------------|-----------------|------------------|
| ML-MMRF      | 92.80                       | 97.53             | 90.26             | 97.26           | 89.89          | 99.98           | 61.54            |

| Dataset      | FOMM PK-PD vs. FOMM Linear | FOMM PK-PD vs. FOMM NL | FOMM PK-PD vs. FOMM MOE | GRU PK-PD vs. GRU | ——               |
|--------------|-----------------------------|------------------------|-------------------------|------------------|------------------|
| ML-MMRF      | 0.792 (0.405)               | 0.668 (0.457)          | 0.510 (0.490)           | 0.406 (0.489)     | ——               |

| Synthetic (50 samples)     | 71.06 +/- .03 | 58.80 +/- .03 | 56.81 +/- .04 | 56.65 +/- .11 | 53.49 +/- .04 |
| Synthetic (1000 samples)   | 62.93 +/- .03 | 57.16 +/- .03 | 57.81 +/- .02 | 31.09 +/- .02 | 29.27 +/- .01 |

Table 2. Generalization of FOMM and GRU models on ML-MMRF and Synthetic Data: Top: Lower is better. We report negative log-likelihood averaged across five folds of held-out data. Bottom: ML-MMRF: We report pairwise comparisons of models trained on ML-MMRF. Higher is better. Each number is the fraction (with std. dev.) of held-out patients for which the model that uses PK-PD is better than a model in the same family that uses a different transition function. Synthetic: Lower is better. We report held-out negative log likelihood with std. dev. on FOMM and GRU to study generalization in the synthetic setting.

E. Experiments on Semi-synthetic Dataset

In this section, we cover how to generate the semi-synthetic dataset. We then provide experimental results on generalization performance as well as a result in a model misspecification scenario.

Semi-synthetic data: We train the SSM model on the ML-MMRF dataset and generate samples from the model. For each sequence of treatments, we generate 30 random samples per training data point resulting in a dataset of size 14000. Then we uniformly at random sample 1000 samples from that pool to form our training set. We perform a similar procedure to generate several held-out sets (size 87000 samples each). This semi-synthetic dataset allows us to ask questions about generalization on data with statistics similar to ML-MMRF.

Generalization and Model Misspecification SSM continues to generalize well even when it is misspecified: We run a similar experiment to what we ran on ML-MMRF, where we take out the Neural Treatment Exponential mechanism function from $\mu_0$ and instead opt for using a linear function. We see that SSM w/o TExp performs comparably to a SSM with the Neural Treatment Exponential mechanism, providing further evidence that our architecture can recover the unknown intervention effect in the data via a combination of related mechanism functions.

Table 3. Held-out NELBO on semi-synthetic data: Trained on 1K or 20K samples and evaluated on 87K samples, we show the mean test NELBO and standard deviation across five test sets.

| Sample Size | SSM Linear | SSM NL | SSM MOE |
|-------------|------------|--------|---------|
| 1000        | -211.54 +/- 49.61 | -192.86 +/- 25.77 | -266.37 +/- 10.42 |
| 20000       | -294.00 +/- 10.25 | -295.44 +/- 8.18 | —— |

Hyperparameters: We present the best hyperparameters for each model at each sample size. We search over the ranges as described in the main paper; however, at 20000 samples, we train for 1000 epochs instead of 15000 epochs, which we found to be a more stable training configuration.

- **SSM**
  1. 1000 samples: State space dimension 64, L2 regularization on all parameters with strength 0.01
  2. 20000 samples: State space dimension 128, L2 regularization on all parameters with strength 0.01
F. Additional Analyses

This section presents experimental results that provide an additional qualitative lens onto the PK-PD Neural.

F.1. Exploring different strategies of sampling patient data using SSM_{PK-PD} on ML-MMRF

In the main paper (Figure 5), we show samples from SSM models trained on ML-MMRF, conditioned on a patient’s first two years of data and the sequence of interventions they were prescribed. In each case, we additionally condition on the patient’s baseline covariates.

Here, we experiment with different conditioning strategies. Let $C$ denote the point in time until which we condition on patient data and $F$ denote the number of timesteps that we sample forward into the future. We limit our analysis to the subset of patients for which $C + F < T$ where $T$ is the maximum number of time steps for which we observe patient data.

The samples we display are obtained as a consequence of averaging over three different samples, each of which is generated (for the SSM) as follows:

$$ Z \sim q_\phi(Z_C|Z_{C-1}, X_{1:C}, U_{0:C-1}) $$
$$ Z_k \sim p_\theta(Z_k|Z_{k-1}, U_{k-1}, B) \quad k = C + 1, \ldots, C + F $$
$$ X_k \sim p_\theta(X_k|Z_k) \quad k = C + 1, \ldots, C + F $$

We study the following strategies for simulating patient data from the models.

1. Condition on 6 months of patient data, and then sample forward 2 years,

2. Condition on 1 year of patient data and then sample forward 1 year,

3. Condition on the baseline data of the patient and then sample forward 2 years.

In Supp. Figure 7, we show additional samples from SSM_{PK-PD} when conditioning on differing amounts of data. Overall, in all three cases, SSM_{PK-PD} models capture treatment response better than one of the best performing baselines (i.e. SSM_{Linear}). For 1. (Figure 7a)), we see that SSM_{PK-PD} correctly captures that the serum IgA value goes up, while SSM_{Linear} predicts that it will stay steady. For 2. (Figure 7b)), SSM_{PK-PD} does well in modeling down-trends, as in serum IgA and serum lambda. For 3. (Figure 7c)), we similarly see that SSM_{PK-PD} captures the up-trending serum IgG and serum lambda.
Figure 7. **Forward samples from learned SSM models with differing conditioning strategies**: We visualize samples from $SSM_{PK-PD}$ (o) and $SSM_{linear}$ (x). Each row corresponds to a single patient, whereas each column represents a different biomarker for that patient. a): We condition on 6 months of patient data and forward sample 2 years. b): We condition on 1 year of patient data and forward sample 1 year. c): We condition on a patient’s baseline data and forward sample 2 years. As in the main paper, blue circles denote ground truth, and the markers above the trajectories represent treatments prescribed across time.
F.2. Analyzing the Latent State learned by SSM\textsubscript{PK-PD} over Time

In Supp Figure 8, we show the latent state of each held-out patient (reduced down to two dimensions via TSNE (Maaten & Hinton, 2008)) over multiple time points, expanding on the two time points that were shown in Figure 4 of the main paper. As we saw before, early in the treatment course, the latent representations of the patients have no apparent structure. However, as time goes on, we find that the latent representations separate based on whether treatment is administered or not.

F.3. Deep Dive into SSM\textsubscript{PK-PD} vs SSM\textsubscript{Linear} on ML-MMRF

We are also interested in the absolute negative log likelihood measures and predictive capacity of the models at a per-feature level. In Supp. Figure 11a), we use importance sampling to estimate the marginal negative log likelihood of SSM\textsubscript{Linear} and SSM\textsubscript{PK-PD} for each covariate across all time points. Namely, we utilize the following estimator,

\[ p(X) \approx \frac{1}{S} \sum_{s=1}^{S} p(X|Z^{(s)}) p(Z^{(s)}) q(Z^{(s)}|X), \]

akin to what is used in (Rezende et al., 2014). SSM\textsubscript{PK-PD} has lower negative log likelihood compared to SSM\textsubscript{Linear} for several covariates, including neutrophil count, albumin, BUN, calcium, and serum IgA. This result is corroborated with the generated samples in Supp. Figure 7, which often show that the PK-PD model qualitatively does better at capturing IgA dynamics compared to the Linear model. In general, although there is a some overlap in the estimates of the likelihood under the two models for some features, it is reassuring to see that SSM\textsubscript{PK-PD} does model the probability density of vital markers like serum IgA (which is often used by doctors to measure progression for specific kinds of patients), better than the baseline.

In Supp. Figure 11b), c), and d), we show the L1 error of SSM\textsubscript{PK-PD} and SSM\textsubscript{Linear} when predicting future values of each covariate. We do so under three different conditioning strategies: 1) condition on 6 months of patient data, and predict 1 year into the future; 2) condition on 6 months of patient data, and predict 2 years into the future; 3) condition on 2 years of patient data, and predict 1 year into the future. Observing 1) and 2) (Supp. Figure 11b) and c)), we see that prediction quality expectedly degrades when trying to forecast longer into the future. Additionally, we find that when increasing the amount of data we condition on to two years (i.e. forward sampling later on in a patient’s disease course) (Supp. Figure 11d)), the prediction quality is similar to that of conditioning only on six months of data (Supp. Figure 11b)) [barring serum M-protein and glucose]. This result reflects the ability of our model to generate accurate samples at multiple stages of a patient’s disease. Finally, we also report L1 error for forward samples taken from SSM\textsubscript{PK-PD} and SSM\textsubscript{Linear} over 6-month time windows in
Neural Pharmacodynamic State Space Modeling

Figure 9. L1 error for 6-month forward samples from PK-PD and Linear models: We report L1 error for forward samples over a 6-month time window conditioned only on baseline data.

Supp. Figure 9. We see that for some biomarkers, such as serum IgA, the PK-PD model has lower L1 error, while for others, such as Calcium, both models do very well.

Figure 10. Feature ablation of conditioning set for subset of biomarkers: We report the per-biomarker mean-squared error (MSE), averaged over all patients and all time points, of SSM_{PK-PD} models trained on an increasing subset of baseline and treatment features.

F.4. Ablation Studies for SSM_{PK-PD}

We report an ablation experiment in Supp. Table 4, where we assess the effect of adding each mechanism function to SSM_{PK-PD} on held-out NELBO. We see that the Neural Log-Cill Kill function gives a modest improvement, while the addition of the Neural Treatment Exponential function gives most of the improvements.

Secondly, in Figure 10, we show a feature ablation experiment to determine the importance of baseline and treatment features in forecasting several multiple myeloma markers. We train SSM_{PK-PD} models on subsets of features, while tuning the latent variable size ([16, 48, 64, 128]) on a validation set for each subset. Then, we evaluate the mean-squared error (MSE), averaged over all examples and time points, of each trained model on a separate held-out set. Our results are shown in Figure 10. We focus on serums IgA, IgG, and lambda, three biomarkers that are commonly tracked in multiple myeloma to evaluate response to treatment and overall progression of disease (Group, 2003; Larson et al., 2012).

We find that for serums IgA and Lambda, adding the treatment signal intuitively leads to a reduction in the MSE. For serum IgG, while the treatment signal helps with predictive performance, the baseline features, such as the genomic and myeloma type features, also seem to play a role.
**Neural Pharmacodynamic State Space Modeling**

| Dataset         | Held-out NELBO | Fold 1  | Fold 2  | Fold 3  | Fold 4  | Fold 5  |
|-----------------|----------------|---------|---------|---------|---------|---------|
| ML-MMRF linear  | 85.26          | 73.25   | 70.61   | 59.86   | 71.11   |         |
| linear + log-cell | 84.42         | 68.11   | 66.69   | 58.83   | 71.48   |         |
| linear + log-cell + te | 65.06    | 57.20   | 66.73   | 53.37   | 58.12   |         |

Table 4. **Ablation Experiment on ML-MMRF Dataset**: We study the effect of adding each mechanism function to $SSM_{PK-PD}$. We report held-out bounds on negative log likelihood.

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**Figure 11.**

- **a)** **NLL estimates via Importance Sampling [top left]**: We estimate the NLL of $SSM_{PK-PD}$ and $SSM_{Linear}$ for each feature, summed over all time points and averaged over all patients.
- **b)** **Condition on 6 months, forward sample 1 year [top right]**: We show L1 Prediction Error for forward samples over a 1 year time window conditioned on 6 months of patient data. At each time point, we compute the L1 error with the observed biomarker and sum these errors (excluding predictions for missing biomarker values) over the prediction window. We employ this procedure for each patient.
- **c)** **Condition on 6 months, forward sample 2 years [bottom left]**: We report L1 error for forward samples over a 2 year window conditioned on 6 months of patient data.
- **d)** **Condition on 2 years, forward sample 1 year [bottom right]**: Finally, we report L1 error for forward samples over a 1 year time window conditioned on 2 years of patient data.