Genetic pattern and demographic history of cutlassfish (*Trichiurus nanhaiensis*) in South China Sea by the influence of Pleistocene climatic oscillations

Sui Gu1, Yun-Rong Yan1,2,3, Mu-Rong Yi1,2, Zhi-Sen Luo1, Hui Wen3, Chang-Ping Jiang1, Hung-Du Lin4 & Xiong-Bo He1,3*  

*Trichiurus nanhaiensis* is one of the most important commercial fish species in the South China Sea. This study aimed to investigate the level of genetic variation and population genetic structure of *T. nanhaiensis* in the South China Sea for the first time, using 281 individuals collected from seven locations along the coast of mainland China, Taiwan, and Hainan Island. A high level of haplotype diversity and low nucleotide diversity were detected in the mitochondrial DNA cyt b gene and nuDNA RYR 3 gene. The overall expected heterozygosity (He = 0.693) among the seven populations ranged from 0.681 to 0.706 in microsatellite DNA data, which revealed high levels of genetic diversity. Significant genetic differentiation was found in Taidong populations in Taiwan, revealing the prevention of gene flow caused by the Kuroshio Current. Two major lineages based on the cyt b gene suggested that the Taiwan Strait acted as a geographic barrier for *T. nanhaiensis* during the glacier periods in the late Pleistocene. The Bayesian skyline plot also revealed that population demographic expansion of *T. nanhaiensis* was estimated to have occurred in 0.1 Mya. Our results indicated that all populations of *T. nanhaiensis* had experienced a recent genetic bottleneck following recent expansion based on ABC analysis.

Because of the rise in overfishing, degradation, pollution, and industrialization, understanding the genetic diversity and population structure of economically important marine species is one major crucial objective of species conservation strategies and fisheries management. The population genetic pattern and phylogeographic structure in marine species are contingent on historical factors, including Pleistocene climatic oscillations, glacial vicariance, ocean current systems, and life-history traits for organisms. With a wide distribution, as well as extensive larval and adult dispersal, some marine fish are characterized by weak genetic structure and inferred connectivity. Several important studies have revealed that species with broad regions, as expected from larval dispersal, showed extensive gene flow and genetic homogeneity. Furthermore, in many cases, this extensive gene flow and genetic homogeneity might be attributed to fewer barriers to gene flow as well as large population sizes. Many previous molecular studies have shown that many marine fish with high mobility exhibit low genetic structure in the northwestern Pacific, such as in the Chinese beard eel (*Cirrhimuraena chinensis*) and ponyfish (*Nuchequula mammusella*). Increasing phylogeographic evidence indicates that cyclical glacial fluctuations shaped the distribution of species, demographic expansions, and genetic divergence of marine fish species. During the Pleistocene glacial-interglacial cycles, the area and configuration of marginal seas in the northwestern Pacific changed dramatically. The emergence of the Taiwan Strait associated with sea-level falls measuring approximately 120–140 m during glacial periods might act as phylogeographic barriers that impede the dispersal of marine fish between both sides of the strait. Recent mtDNA studies indicate that there are two distinct lineages due to

1College of Fisheries, Guangdong Ocean University, No.1 Haida Road, Mazhang District, Zhanjiang 524088, China. 2Southern Marine Science and Engineering Guangdong Laboratory (Zhanjiang), Zhanjiang 524000, China. 3Guangdong Provincial Engineering and Technology Research Center of Far Sea Fisheries Management and Fishing of South China Sea, Guangdong Ocean University, Zhanjiang 524088, China. 4The Affiliated School of National Tainan First Senior High School, No. 1, Sec. 1, Mintzu Rd., Tainan 701, Taiwan. *email: varicorhinus@hotmail.com; xiongbo98@163.com
vicariance events in some marine fish, such as in Chinese black sleeper (Bostrychus sinensis)\(^1\), large yellow croaker (Larimichthys crocea)\(^1\), Lepturacanthus savala\(^1\), and yellow seabream (Acanthopagrus latus)\(^1\). Understanding the genetic characteristics of marine fish, including their genetic diversity, genetic structure, and population historical demography, provides reference opinions for the long-term management and protection of species germplasm resources\(^1\).

Cutlassfishes (of the family Trichiuridae) belonging to the suborder Scombroidei of Perciformes is a diverse group of demersal fishes, including three subfamilies, eight genera, and twelve species in mainland China. The fisheries of cutlassfishes showed declining trends over recent years on the coast of mainland China and the third most marine catches in the South China Sea (SCS). Trichiurus nanhaiensis, Wang and Xu 1992 featuring a very elongated and highly compressed body with a deeply forked caudal fin, is a benthic fish of the family Trichiuridae that inhabit the continental shelves and shallow seas (20–200 m) of tropical and temperate waters, and is wide geographical distribution in the Indo-Pacific Ocean, including the South China Sea. The Taiwan Strait is thought to be the northernmost limit of its range in the Western Pacific. Trichiurus nanhaiensis is one of the most important commercial fish species in the South China Sea, accounting for 13.7–14.2% of the total annual fishing catch in the South China Sea between 2019 and 2020. Previous studies on T. nanhaiensis have focused mainly on its mitogenome taxonomy\(^2\), DNA barcoding\(^3\), characterization of microsatellite DNA\(^4\), reproduction\(^5\) and demographic history\(^6,7\). Understanding the genetic structure of T. nanhaiensis populations is necessary and required to formulate strategies for the effective protection and utilization of marine fishery resources\(^8\). In addition, previous population genetic studies of T. nanhaiensis using limited genetic data were used to delineate population structure, and discrepancies among studies have to some extent hindered the effectiveness of fisheries management and conservation\(^9,10\).

Nuclear DNA is inherited from two parents, and recombination of genes from the parents in every generation, whereas mtDNA is haploid and only maternally inherited, reflecting changes in population structure faster due to its lower effective population size and therefore rapid coalescence time. Microsatellites are simple repetitive sequences and codominant throughout the eukaryote nuclear genome, showing high resolution at the population level\(^11\). Direct comparisons among mtDNA, nuDNA and microsatellite loci can be very informative for population diversity and genetic structure, as they provide valuable information for evaluation and conservation issues. This study aimed to investigate the level of genetic variation and population genetic structure of T. nanhaiensis in the South China Sea (including mainland China, Taiwan, and Hainan Island) for the first time, and it used mitochondrial, nuclear, and microsatellite DNA markers to reveal the distribution of genetic variation in T. nanhaiensis. Additionally, our results provide a workable and responsible stock for scientific and conservation purposes to maintain long-term sustainability and resilience to the fishing pressure of T. nanhaiensis resources.

### Results

**Mitochondrial and nuclear DNA.** A total of 181 and 57 haplotypes were identified by sequencing 1141 bp and 882 bp of the complete mtDNA cyt b gene and nuclear RYR 3 gene from 281 T. nanhaiensis individuals, respectively (Table 1, Fig. 1). The most common haplotypes of the mtDNA cyt b gene (Hm8, Hm18, and Hm27) (Supplementary Table S1) and nuclear RYR 3 gene (Hn1, Hn2, Hn6, and Hn8) were shared by specimens from seven and all populations, respectively (Supplementary Table S2). The average haplotype diversity of the mtDNA cyt b gene was high (0.989), ranging from 0.981 (ST) to 0.995 (ZJ), and the average nucleotide diversity (\(\theta_\pi\)) was

| Population | Code | Coordinates | Sample size | mtDNA | nuDNA | Microsatellite |
|------------|------|-------------|-------------|-------|-------|--------------|
|            |      |             |             | N of h | H     | \(\theta_s\) (%) | \(\theta_\pi\) (%) | N of h | H     | \(\theta_s\) (%) | Aa | AR | HO | HE | FIS | HWE |
| Mainland China |      |             |             |        |       |               |                |        |       |               |    |     |     |    |     |    |
| 1. Shantou  | ST   | 23.02° N 116.93° E | 63          | 45    | 0.981 | 0.451          | 0.987           | 16     | 0.740 | 0.149          | 0.398 | 10.6 | 7.789 | 0.563 | 0.685 | 0.172 | 0.160 |
| 2. Shenzhen | SZ   | 21.86° N 114.15° E | 40          | 30    | 0.985 | 0.434          | 0.867           | 11     | 0.650 | 0.100          | 0.183 | 9.4  | 7.907 | 0.601 | 0.690 | 0.109 | 0.274 |
| 3. Yangjiang| YJ   | 21.31° N 112.19° E | 36          | 33    | 0.994 | 0.528          | 0.889           | 10     | 0.680 | 0.127          | 0.211 | 8.5  | 7.537 | 0.614 | 0.685 | 0.100 | 0.422 |
| 4. Zhanjiang| ZJ   | 20.90° N 110.88° E | 30          | 28    | 0.995 | 0.381          | 0.665           | 13     | 0.702 | 0.129          | 0.195 | 8.2  | 7.672 | 0.607 | 0.702 | 0.131 | 0.352 |
| 5. Beihai   | BH   | 21.23° N 109.24° E | 39          | 34    | 0.988 | 0.422          | 0.851           | 13     | 0.697 | 0.123          | 0.253 | 9.4  | 8.113 | 0.628 | 0.706 | 0.103 | 0.185 |
|             |      |             |             |        |       |               |                |        |       |               |    |     |     |    |     |    |
| Taiwan Island |      |             |             |        |       |               |                |        |       |               |    |     |     |    |     |    |
| 6. Taidong  | TD   | 22.72° N 121.23° E | 25          | 22    | 0.990 | 0.511          | 0.698           | 12     | 0.693 | 0.123          | 0.202 | 8.5  | 8.453 | 0.555 | 0.702 | 0.191 | 0.189 |
|             |      |             |             |        |       |               |                |        |       |               |    |     |     |    |     |    |
| Hainan Island |     |             |             |        |       |               |                |        |       |               |    |     |     |    |     |    |
| 7. Sanya    | SY   | 17.89° N 109.60° E | 48          | 40    | 0.988 | 0.475          | 1.029           | 25     | 0.812 | 0.185          | 0.419 | 8.8  | 7.309 | 0.590 | 0.681 | 0.127 | 0.245 |
| Mean        |      |             |             | 0.9893 | 0.465 | 1.724          | 0.724           | 0.139 | 0.673 | 9.1  | 7.826 | 0.594 | 0.693 |

Table 1. Samples used for analysis, location, code and summary statistics. a Number of haplotypes; b Haplotype diversity; c Average nucleotide diversity; d Nucleotide diversity; e Average number of alleles; f Mean allelic richness; g Mean observed heterozygosity; h Mean expected heterozygosity; i Mean level of inbreeding observed; j Hardy–Weinberg equilibrium.
The pairwise $F_{ST}$ values ranged from 0.011 (between SY and YJ) to 0.084 (between TD and SZ), with a mean value of 0.023 in the mtDNA cyt $b$ gene. The pairwise $F_{ST}$ values between the populations from TD and other populations were differentiated, except for the YJ population in the mtDNA cyt $b$ gene (Supplementary Table S3). The pairwise $F_{ST}$ values ranged from 0.001 (between ST and BH) to 0.060 (between YJ and ZJ), with a mean value of 0.020 in the nuDNA RYR 3 gene. No significant correlation was found between genetic distance and geographic distance by using IBD analyses ($r = 0.438,$ $p = 0.087$).

Hierarchical analyses of molecular variance (AMOVA) from the probable factors shaping genetic structure demonstrated that most of the genetic variation was among individuals within populations in the mtDNA cyt $b$ gene and nuclear RYR 3 gene, i.e., one group (97.79%, 98.42%), two groups (Scenario I, 94.47%, 97.46%), two groups (Scenario II, 99.00%, 98.74%), and three groups (Scenario III, 97.30%, 98.32%) (Table 2). However, only 2.21%, 4.19%, 1.87%, and 0.97% of the total variation was found among the groups in one group, two groups (Scenario II), two groups (Scenario II), and three groups (Scenario III) in the mtDNA cyt $b$ gene, respectively (Table 2). Phylogenetic trees and haplotype networks based on the cyt $b$ gene support the formation of two major lineages, and genetic clusters do not correspond to geographical sampling sites (Figs. 2, 3). However, the phylogenetic relationship and networks based on the nuDNA fragments showed no evidence of significant geographical structure corresponding to the sampling locations (Supplementary Figs. S1, S2). Neutrality tests, mismatch distribution analysis, and Bayesian skyline plots were used to analyze signatures of historical demographic events. The significant negative values of Tajima’s $D$ and Fu’s $Fs$ tests demonstrated deviation from the neutrality test, suggesting past population expansion for $T.$ nanhaiensis (Tajima’s $D = −2.221,$ $P < 0.001$ and Fu’s $Fs = −358.690,$ $P < 0.000$). Likewise, the same results were supported by the presence of a star-like topology in the cyt $b$ gene tree and by the results of mismatch distribution analysis, which showed unimodal distributions (Supplementary Fig. S3). The Bayesian skyline plot showed that $T.$ nanhaiensis populations seem to have experienced demographic expansion starting approximately 100,000 years ago and a flat population history during the Pleistocene (Fig. 4). The current ($θω$) and historical ($θπ$) genetic diversity was 0.017 and 0.0046, respectively. When the current genetic diversity was larger than the historical genetic diversity, the $T.$ nanhaiensis populations revealed a pattern of decline.22

**Microsatellite DNA.** No indication of genotyping errors due to stutter bands, large allele dropout, or null alleles was revealed using a microchecker25. A total of 118 alleles were detected for the 13 polymorphic markers, and the number of alleles per marker ranged from 3 to 27, with an average of 9.0, in 281 individuals, with 72 unique alleles in the total population. The sample size, number of alleles ($N_a$), number of allelic richness ($A_d$) per population, expected heterozygosity ($H_e$), observed heterozygosity ($H_o$), inbreeding coefficient ($F_i$), and deviation from Hardy–Weinberg equilibrium (HWE) for each population are shown in Table 1. The average number of alleles and allelic richness for each population ranged from 8.200 (ZJ) to 10.600 (ST) (average = 9.050) and from 7.530 (YJ) to 8.450 (TD) (average = 7.820), respectively. The mean observed and expected heterozygosity ($H_e$) was 0.594 and 0.693, ranging from 0.244 (Tna1) to 0.759 (Tna14), and ranging from 0.100 (YJ) to 0.191 (TD) (Table 1). The average number of alleles ($N_a$) and allelic richness ($A_d$) per locus were 9.0 and 7.8, ranging from 3 (Tna3) to 27 (Tna16) and from 2.771 (Tna3) to 22.406 (Tna16), respectively. The observed heterozygosity ($H_o$) and the expected heterozygosity ($H_e$) were 0.594 and 0.693, ranging from 0.244 (Tna1) to 0.759 (Tna14), and ranging from 0.001 (SZ) to 0.002 (SY) (Table 1).

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Table 2. Analysis of molecular variance (AMOVA) for *Trichiurus nanhaiensis* populations based on three molecular markers. Significant scores (P < 0.05) are shown in bold face.
populations for each locus showed populations TD clustered at the top of the heatmap, while the remaining populations belonged to the other group. The results of DAPC analyses indicated that the populations of *T. nanhaiensis* could be split into two groups (Fig. 7). Populations TD belonged to one group, and the remaining populations belonged to the other group.

**Approximate Bayesian computation.** Among the five different plausible demographic scenarios (see Fig. 8I) tested in our DIYABC simulations, Scenario 5 (DECINC model) had the highest PP and 95% CI (posterior probability = 0.998 [0.995,1.000]) compared with the other scenarios. The results of a logistical model comparing posterior probability of each scenario with the number of simulations used to calculate it (see Fig. 8II). The prior distribution of parameters corresponding to the five scenarios tested (see Fig. 8III). This scenario indicated that *T. nanhaiensis* experienced a reduction in the effective population size in the past, followed by a single instantaneous increase in population size.

**Figure 2.** Phylogenetic trees reconstructed from mitochondrial sequences of the cyt b gene in *Trichiurus nanhaiensis*. The values above the branches are bootstrap values for the NJ, BI, and ML analyses.
Discussion

Genetic diversity. Monitoring genetic diversity is an integral part of conservation biology, and maximizing the maintenance of genetic variation is necessary to dissect the integrated effects of evolutionary and ecological processes following environmental changes\(^2\). Here, we provided an assessment of molecular markers (mitochondrial, nuclear, and microsatellite DNA) to analyze the genetic diversity of *T. nanhaiensis*, which is of importance to biodiversity conservation and the sustainable use of biological resources. High haplotype diversity \((h = 0.989)\) and low nucleotide diversity \((0.005)\) were found in *T. nanhaiensis* populations based on the mitochondrial cyt *b* gene. High haplotype diversity was similar to the haplotype diversity of most marine fishes on the coastlines of mainland China, suggesting large effective population sizes over time \([e.g., Pampus argenteus\(^2\) \((h = 0.736); Terapon jarbua\(^2\) \((h = 0.857); Acanthogobius ommaturus\(^2\) \((h = 0.923); Scatophagus argus\(^2\) \((h = 0.870)\)]\). The nucleotide diversity of *T. nanhaiensis* was higher than the nucleotide diversity of *L. savala*\(^8\) but lower than the nucleotide diversity of *T. japonicas* in the family Trichiuridae along the coastlines of mainland China\(^1\). Population size is considered to be an important factor in the maintenance of genetic variation, and
small populations should show lower levels of genetic variability than large populations. The current effective population sizes of *T. japonicas*, the dominant cutlassfish species, were larger than the current effective population sizes of *T. nanhaiensis* throughout Taiwan and the coast of mainland China. The values of the average number of alleles per population (9.0) of *T. nanhaiensis* were relatively lower than the average number of alleles per population of the relative species *L. savala* and other marine fish populations along the coast of mainland China (e.g., *N. albiflora* $N_a = 12.02031$; *Lateolabrax maculatus* $N_a = 28.72732$; *Larimichthys polyactis* $N_a = 24.95033$). In this study, the average value of expected heterozygosity (0.693) of *T. nanhaiensis* was relatively lower than the average value of expected heterozygosity of marine fish (average $H_E = 0.790$) with many marine fish populations. In this study, the positive inbreeding coefficient ($F_{IS} = 0.145$) of *T. nanhaiensis* indicates heterozygosity deficiency in all populations, which might be attributed to inbreeding. Previous studies have reported that low observed heterozygosity points to a deficiency of heterozygotes and high levels of inbreeding in the waters off mainland China (e.g., *Larimichthys polyactis*; *Collichthys lucidus*; *Lateolabrax maculatus*), as well as in other related species (e.g., *L. savala*). In addition, the loss of genetic diversity may reflect past historical events associated with current evolutionary forces, such as fishing pressure or habitat loss.

**Figure 5.** Clustering of individuals by structure at $K = 2$. Individuals are represented by vertical bars. Each vertical column represents one individual, and the separation of the column into two colors represents the estimated probability of belonging to one population or the other. Different colors in the same individual indicate the percentage of the genome shared with each cluster according to the admixture proportions. The y-axis represents the probability of belonging to a certain cluster, while each population (code name given in Table 1) delimited by a black solid vertical line is reported on the x-axis.

**Figure 6.** Heat map representing allele frequency for 13 loci in all 7 populations as the gradual increase in color density from lighter color (blue) to darker color (red) means the increase in the allele frequency as the red color, which indicates the highest value of allele frequency.
Loss of genetic diversity is most often associated with reduced population adaptive capacity fitness through inbreeding depression, the maintenance of the high levels of genetic variability in natural populations has been recommended for conservation.

**Phylogeography and population structure of *Trichiurus nanhaiensis*.** Many previous studies suggested that the major drop in sea level in the Taiwan Strait acted as a biogeographic barrier during major falls in sea level in the Pleistocene and might have cut off migration on either side of the strait and presented two lineages in marine fishes, such as the genera *Helice*[^1], *B. sinensis*[^2], and *L. savala*[^3]. Our results revealed that haplotype networks and phylogenetic trees support the formation of two major lineages based on the cyt b gene, and genetic clusters do not correspond to geographical groups. During the Pleistocene, the East China Sea and the South China Sea were semienclosed marginal seas and separated by the Taiwan Strait. We suggested that the Taiwan Strait acted as a geographic barrier for *T. nanhaiensis* during the glacier periods in the late Pleistocene and gene flow due to the absence of physical barriers during glacial retreat, while more recent secondary contacts could have played a major role[^4]. The results of the AMOVA test based on mitochondrial, nuclear, and microsatellite DNA data indicated that there were no significant differences at all hierarchical levels. All genetic markers, including mitochondrial, nuclear, and microsatellite markers, congruently showed a similar genetic pattern, revealing overall low genetic differentiation among *T. nanhaiensis* populations, by previous studies on other marine fishes among the geographical stocks in the South China Sea (e.g., *L. savala*[^5]; *Nemipterus bathybius*[^6]).

The results from STRUCTURE indicated the presence of two clusters after independent clustering of all populations based on microsatellite DNA data, which may be explained by the high dispersal capacity of *T. nanhaiensis*. The scenario of evolutionary diversification of lineages followed by postglacial expansion facilitating secondary contacts was observed in previous studies (e.g., *S. macrorhynchos*[^7]; *Scatophagus argus*[^8]; *L. savala*[^9]). A previous study in *Trichiurus* species suggests a genetic well mixture and a lack of distribution barrier for *T. nanhaiensis*[^10]. Ocean currents, which change with the monsoon and are highly variable in the SCS, play an important role in transporting eggs, larvae, or juveniles of *T. nanhaiensis*[^11,12]. We suggested that the high dispersal ability of marine fish may be appeared by gene flow following divergence, indicating that divergent mitochondrial lineages arose within a panmictic population. The results from DAPC and the heatmap of allele distribution and frequency across populations revealed that population TD was distinctly separated from the other geographic populations. Population TD was located in southern Taiwan, and samples were collected from the Pacific Ocean. We suggest that population TD belonging to peripheral populations may always have low genetic variability due to isolation and founder effects, but the differentiation of the populations is high owing to genetic drift and reduced gene flow under the influence of the Kuroshio Currents[^13].
Demographic history and DIY-ABC. Both mismatch distribution analyses and neutrality indices of Tajima’s $D$ and Fu’s $Fs$ tests suggested that $T. nanhaiensis$ populations indicate recent demographic expansion. Moreover, the star-like structure of the haplotype network shows consistent evidence for past recent population bottlenecks followed by population expansion. The Bayesian skyline plot also revealed that population demographic expansion of $T. nanhaiensis$ was estimated to have occurred in 0.1 Mya. The presence of a consistent signal of recent population expansion gives a similar estimated expansion time to other previous studies in $T. nanhaiensis$, which suggests that the population expansion event took place in the late Pleistocene$^{18,19}$. Furthermore, the results of our comparison to the previous study of marine fish in the South China Sea thus provide considerable evidence of population growth and expansion in $T. nanhaiensis$ (e.g., $S. macrorhynchos$$^{39}$; $Tridacna maxima$$^{43}$; $Scatophagus argus$$^{28}$; $Nemipterus bathybius$$^{38}$; $L. savala$$^{10}$). However, a higher $\theta_w$ than $\theta_p$ indicated population decline, and all seven populations appeared to experience genetic bottlenecks based on the microsatellite markers in $T. nanhaiensis$. The demographic scenario of $T. nanhaiensis$ illustrates an even more complex history. Our best-fitting ABC model [Scenario 5 (DECINC model)] was identified with recent expansion preceded by a bottleneck. Our results support previous studies showing that the population of $T. nanhaiensis$ steadily declined during 45–165 kya and exhibited a pattern of glacial growth$^{18}$.

Materials and methods
Sample collection, DNA extraction, and microsatellite genotyping. A total of 281 $T. nanhaiensis$ were collected from seven locations in the South China Sea in 2019 (Table 1, Fig. 1). All studies in animals were conducted by ethical committee guidelines and approved by the Animal Research and Ethics Committee of College of Fisheries, Guangdong Ocean University (China), and the study was carried out in compliance with the ARRIVE guidelines. Fin tissues were stored in 95% alcohol for DNA extraction after morphological identification of all samples. Total DNA was extracted using a DNA extraction kit (Sangon Biotech, Shanghai, China).

The mitochondrial cytochrome $b$ (Cyt $b$) gene was amplified using the gsCY4-F (5’-TCCCTCATTGACCTTCCA-3’)/gsCY4-R (5’-GGTTGCGGTTCACTGTGAG-3’) primer pair, and the nuclear DNA gene (its protein-coding sequence) for ryanodine receptor 3 (RyR3) was amplified using the primers RYR3-F22 (5’-TCGGTA
Data analysis. The Cyt b and RyR 3 sequence alignments for all 281 samples were separately performed using Clustal X 2.0 software. The PHASE algorithm in DnaSP v5.0 software was used to resolve the haplotype phases of RyR3 sequences that contained more than one ambiguous site and analyze summary statistics such as the number of haplotypes (Nh), haplotype diversity (h), and nucleotide diversity (θS, θO) used to analyze the phylogeographic relationship of T. nanhaiensis. Bayesian inference (BI) and maximum likelihood (ML) were performed using PhyloSuite v1.2.2. The best-fit partitioning strategy and nucleotide substitution model for BI and ML tree were selected by ModelFinder installed in Phylosuite v1.2.2 with the greedy algorithm and BIC (Bayesian information criterion) criteria. Haplotype networks for Cyt b and RyR 3 were computed using the minimum-spanning network method (minspnet in Arlequin 3.5). In addition, the neutrality test (Tajima’s D and Fu’s Fu test) and mismatch distributions were evaluated to test the hypothesis of demographic expansion using DnaSP v5.0 software. Bayesian skyline plots (BSPs) were implemented in BEAST v1.8.2 for T. nanhaiensis to determine the effective population size changes over time. The MCMC chains were run three times each for 100 million generations, sampling every 10,000th generation for each data set. In the BSPs analysis, the effective sample size (ESS) values for each parameter was larger than 200, and the 95% highest posterior density (HPD) intervals were reported. Bayesian skyline plots (BSPs) were analyzed and drawn using the tracer v1.6 program. A 2.03%/MY divergence rate has been calibrated for the mtDNA cyt b genes in Trichturus fishes for population expansion.

Pairwise FST between all pairs of populations and analysis of molecular variance (AMOVA) were executed in Arlequin 3.5. Seven populations were grouped with three scenarios based on geographical barriers: (1) Scenario I: two independent groups divided by the Taiwan Strait (TD; ST, SZ, YJ, ZJ, BH, SY); (2) Scenario II: two independent groups divided by the Qiongzhou Strait (BH; ST, SZ, YJ, ZJ, SY, TD); (3) Scenario III: three independent groups that included the Taiwan island group, the Taiwan island group and the Hainan island group (ST, SZ, YJ, ZJ; TD; SY, BH). In addition, an isolation-by-distance Mantel test was performed to examine correlations between genetic differentiation and geographic distance by IBD 1.52 for Windows. The Google Earth database (http://earth.google.com) was used to measure the geographic distance in kilometres from GPS locations between sampling sites.

Microsatellite DNA analysis. All loci were tested for possible genotyping errors, the presence of null alleles, and allelic dropout using MICRO-CHECKER 2.2.3 software. In this study, Arlequin 3.5 was used to calculate the number of alleles (Na), mean observed heterozygosity (Ho), mean expected heterozygosity (He), and deviation from Hardy–Weinberg equilibrium (HWE) for each population. Allelic richness (Ar) and inbreeding coefficient (FIS) were estimated using FSTAT version 2.9.3. Population differentiation of T. nanhaiensis was studied by estimation of the FST and RST using Arlequin 3.5 and AMOVA procedures as mitochondrial DNA data (mentioned above).

STRUCTURE v.2.2.3 was used to delineate the optimum number of homogenous groups of sampled individuals (K). A series of 10 independent runs for each K (ranging from 2 to 5) was performed using 100,000 iterations of the Markov chain Monte Carlo (MCMC) with a burn-in period of 20,000 chains. Structure Harvester Web 0.6.94 was used to determine that the optimum number of genetic clusters within the data were predicted using the delta K method and Evanno method. GenAIEx version 6.5.03 was used to calculate allelic frequency and the R package “pheatmap” was used for data visualization and allele frequency heatmap construction. Moreover, to investigate the genetic relationship between populations, discriminant analysis of principal components (DAPC) was performed using the “adegenet 2.0.1” package in R 3.2.2 software.

Approximate Bayesian computation (ABC) analysis. The approximate Bayesian computation (ABC) method is incorporated into different historical and demographic evolutionary models that can be suited to complex problems that arise in scenarios in population genetics. To explore the possible historical demography of T. nanhaiensis, we used a coalescence analysis implemented in the program DIYABC v.2.0.4 to obtain relevant and detailed information on the population history. We separately tested the five scenarios of possible demographic changes for T. nanhaiensis based on Cyt b and RyR3 sequences and microsatellite data (Fig. 8). The HKY was the best evolutionary model for our sequence dataset. The following scenarios were used: In scenario 1 (CON model), assumed constant population size of T. nanhaiensis; In scenario 2 (DEC model), assumed populations of T. nanhaiensis decline and had experienced a bottleneck event; In scenario 3 (INC model), assumed populations of T. nanhaiensis expansion recently; In scenario 4 (INCDEC model), assumed populations of T. nanhaiensis expanded followed by a single instantaneous decrease in population size; and in scenario 5 (DECINC model), assumed populations of T. nanhaiensis bottleneck followed by re-expansion. To obtain robust results, a total of 3,000,000 simulations were performed as recommended by DIYABC for all five demographic scenarios (Fig. 8). We calculated all the summary statistics included in the software DIYABC. The intervals of the prior
distribution of applied parameters were all 10–100,000, and those of mean mutation were 10−9–10−7. The best-supported scenario was obtained by the highest posterior probability, which were calculated by performing a polychotomous weighted logistic regression on 1% of the simulated datasets closest to the observed dataset64. Type I and II errors were calculated to evaluate the confidence of the selected scenario for each approach65.

Data availability
The sequences of dataset were deposited in GenBank (accession numbers: OM303509-OM303789 for Cytb and OM388568-OM389129 for RYR3). Voucher specimens are housed at College of Fisheries, Guangdong Ocean University, Zhanjiang, China.

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References
1. Smouse, P. E. & Peakall, R. O. D. Spatial autocorrelation analysis of individual multiallele and multilocus genetic structure. *Heredity* **82**(5), 561–573 (1999).
2. Liu, J. X., Gao, T. X., Wu, S. F. & Zhang, Y. P. Pleistocene isolation in the Northwestern Pacific marginal seas and limited dispersal along the marine fish, *Chelon haematochilus* (Temminck & Schlegel, 1845). *Mol. Ecol.* **16**(2), 275–288 (2007).
3. Ding, S., Mishra, M., Wu, H., Liang, S. & Miyamoto, M. M. Characterization of hybridization within a secondary contact region of the inshore fish, *Bostryx sinusia*, in the East China Sea. *Heredity* **120**(1), 51–62 (2018).
4. Ashrafzadeh, M. R. et al. Assessing the origin, genetic structure and demographic history of the common pheasant (*Phasianus colchicus*) in the introduced European range. *Sci. Rep.* **11**(1), 1–14 (2021).
5. Caccavo, J. A. et al. Along-shelf connectivity and circumpolar gene flow in Antarctic silverfish (*Pleuragramma antarctica*). *Sci. Rep.* **8**(1), 1–18 (2018).
6. Otwoma, L. M., Reuter, H., Timm, J. & Meyer, A. Genetic connectivity in a herbivorous coral reef fish (*Acanthurus leucosternon* Bennet, 1833) in the Eastern African region. *Hydrobiologia* **806**(1), 237–250 (2018).
7. Li, H., Lin, H., Li, J. & Ding, S. Phylogeography of the Chinese beard eel, *Cirrhimuraena chinensis* Kaup, inferred from mitochondrial DNA: A range expansion after the last glacial maximum. *Int. J. Mol. Sci.* **15**(8), 13564–13577 (2014).
8. Gao, B., Song, N., Li, Z., Gao, T. & Liu, L. Population genetic structure of *Nucheaquula mannsuella* (Perciformes: Leioagnathidae) population in the Southern Coast of China inferred from complete sequence of mtDNA Cyt b gene. *Pak. J. Zool.* **51**(4), 1527–1535 (2019).
9. Qiu, F., Li, H., Lin, H., Ding, S. & Miyamoto, M. M. Phylogeography of the inshore fish, *Bostryx sinusia*, along the Pacific coastline of China. *Mol. Phylogenet. Evol.* **96**, 112–117 (2016).
10. Gu, S. et al. Genetic diversity and population structure of cutlassfish (*Leptura canthus savala*) along the coast of mainland China, as inferred by mitochondrial and microsatellite DNA markers. *Reg. Stud. Mar. Sci.* **43**, 101702 (2021).
11. Liu, Q. et al. Genetic variation and population genetic structure of the large yellow croaker (*Larimichthys crocea*) based on genome-wide single nucleotide polymorphisms in farmed and wild populations. *Fish. Res.* **232**, 105718 (2020).
12. Song, P. et al. Genetic characteristics of yellow seabream *Acanthopagrus latus* (Houttuyn, 1782) (*Teleostei: Sparidae*) after stock enhancement in southeastern China coastal waters. *Reg. Stud. Mar. Sci.* **48**, 102065 (2021).
13. Ward, R. D. Genetics in fisheries management. *Hydrobiologia* **420**, 191–201 (2000).
14. Liu, X., Guo, Y., Wang, Z. & Liu, C. The complete mitochondrial genome sequence of *Trichirias nanhaiensis* (Perciformes: Trichiridae). *Mitochondrial DNA* **24**(5), 516–517 (2013).
15. Wang, H. Y., Dong, C. A. & Lin, H. C. DNA barcoding of fisheries catch to reveal composition and distribution of cutlassfishes along the Taiwan coast. *Fish. Res.* **187**, 103–109 (2017).
16. Guo, Y. S., Liu, X. M., Wang, Z. D., Lu, H. S. & Liu, C. W. Isolation and characterization of microsatellite DNA loci from Naihai cutlassfish (*Trichiura nanhaiensis*). *J. Genet.* **93**(1), 109–112 (2014).
17. Kwok, K. Y. & Ni, I. H. Reproduction of cutlassfishes *Trichirius* spp. from the South China Sea. *Mar. Ecol. Prog. Ser.* **176**, 39–47 (1999).
18. He, L. et al. Demographic response of cutlassfish (*Trichirius japonicus* and *T. nanhaiensis*) to fluctuating palaeo-climate and regional oceanographic conditions in the China seas. *Sci. Rep.* **4**(1), 1–10 (2014).
19. Lin, H. C., Tsai, C. J. & Wang, H. Y. Variation in global population distribution, populations structures and demographic history for four *Trichirius* cutlassfishes. *Peer. P.* e12639 (2021).
20. Funk, W. C., McKay, J. K., Hohenlohe, P. A. & Allendorf, F. W. Harnessing genomics for delineating conservation units. *Trends Ecol. Evol.* **27**(9), 489–496 (2012).
21. Tautz, D. Hypervariability of simple sequences as a general source for polymorphic DNA markers. *Nucleic Acids Res.* **17**, 6463–6471 (1989).
22. Templeton, A. R. The, “Eve” hypotheses: A genetic critique and reanalysis. *Am. Anthropol.* **95**, 51–72 (1993).
23. Van Oosterhout, C., Hutchinson, W. F., Wills, D. P. & Shipley, P. MICRO-CHECKER: Software for identifying and correcting genotyping errors in microsatellite data. *Mol. Ecol. Notes* **4**(3), 535–538 (2004).
24. Franklin, C., E. Challenges and opportunities of genetic approaches to biological conservation. *Biol. Conserv.* **143**(9), 1919–1927 (2010).
25. Sun, P., Shi, Z., Yin, F. & Peng, S. Population genetic structure and demographic history of *Pampus argenteus* in the Indo-West Pacific inferred from mitochondrial cytochrome b sequences. *Biochem. Syst. Ecol.* **43**, 54–63 (2012).
26. Liu, S. Y. V. et al. Genetic stock structure of *Terapon jarbua* in Taiwanese waters. *Mar. Coast. Fish.* **8**(1), 464–473 (2015).
27. Song, C. Y., Sun, Z. C., Gao, T. X. & Song, N. Structure analysis of mitochondrial DNA control region sequences and its applications for the study of population genetic diversity of *Acanthogobius ommatius* *Russ. J. Mar. Biol.* **46**(4), 292–301 (2020).
28. Yan, Y. R. et al. Cryptic diversity of the spotted scat *Scatophagus argus* (Perciformes: Scatophagidae) in the South China Sea: Pre-or post-production isolation. *Mar. Freshw. Res.* **71**(12), 1640–1650 (2020).
29. Hartl, D. L., Clark, A. G., & Clark, A. G. *Principles of Population Genetics*, vol. 116 (Sinauer Associates, 1997).
30. Wu, R. et al. Study on the nomenclature and taxonomic status of hairtail *Trichiura japonicus* from the Chinese coastal waters. *Genom. Appl. Biol.* **37**(9), 3782–3791 (2018).
31. Xu, D. et al. Genetic diversity and population differentiation in the yellow drum *Nibea albiflora* along the coast of the China Sea. *Mar. Biol. Res.* **13**(4), 456–462 (2017).
32. Wang, W. et al. Genetic diversity and population structure analysis of *Lateolabrax maculatus* from Chinese coastal waters using polymorphic microsatellite markers. *Sci. Rep.* **11**(1), 1–11 (2021).
33. Cheng, Q., Chen, W. & Ma, L. Genetic diversity and population structure of small yellow croaker (*Larimichthys polyactis*) in the Yellow and East China seas based on microsatellites. *Aquat. Living Resour.* **32**, 16 (2019).
34. DeWoody, J. A. & Avise, J. C. Microsatellite variation in marine, freshwater and anadromous fishes compared with other animals. J. Fish Biol. 56(3), 461–473 (2000).
35. Song, N., Yin, L., Sun, D., Zhao, L. & Gao, T. Fine-scale population structure of Collichthys lucidus populations inferred from microsatellite markers. J. Appl. Ichthyol. 35(3), 709–718 (2019).
36. Yin, W. et al. Species delimitation and historical biogeography in the genus Helice (Brachyura: Varunidae) in the Northwestern Pacific. Zool. Sci. 26(7), 467–475 (2009).
37. Hewitt, G. The genetic legacy of the Quaternary ice ages. Nature 405(6789), 907–913 (2000).
38. Yi, M. R. et al. Genetic structure and diversity of the yellowbelly threafdn bream Nemipterus bathybius in the Northern South China Sea. Diversity 13(7), 324 (2021).
39. Chen, X., Wang, J. J., Ai, W. M., Chen, H. & Lin, H. D. Phylogeography and genetic population structure of the spadenoce shark (Scylliodon macrorynchos) from the Chinese coast. Mitochondrial DNA Part A 29(7), 1100–1107 (2018).
40. Huang, W. et al. Genetic diversity and large-scale connectivity of the scleractinian coral Porites lutea in the South China Sea. Coral Reefs 37(4), 1259–1271 (2018).
41. Hou, G. et al. Identification of eggs and spawning zones of hairtail fishes Trichiurus (Pisces: Trichiuridae) in Northern South China Sea, using DNA barcoding. Front. Environ. Sci. 9, 703029 (2021).
42. Tamaguchi, K., Nakajima, M. & Taniguchi, N. Loss of genetic variation and increased population differentiation in geographically peripheral populations of Japanese char Salvelinus leucomaenis. Aquaculture 30520–S27 (2010).
43. Neo, M. L., Liu, L. L., Huang, D. & Soong, K. Thriving populations with low genetic diversity in giant clam species, Tridacna maxima and Tridacna noae, at Dongsha Atoll, South China Sea. Reg. Stud. Mar. Sci. 24, 278–287 (2018).
44. Jeannotte, P., Thompson, J., Goyou, M., Higgins, D. & Gibson, T. Multiple sequence alignment with Clustal X. Trends Biochem. Sci. 23, 403–405 (1998).
45. Librado, P. & Rozas, J. DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. Bioinformatics 25(11), 1451–1452 (2009).
46. Kumar, S., Stecher, G., Li, M., Knyaz, C. & Tamura, K. MEGA X: Molecular evolutionary genetics analysis across computing platforms. Mol. Biol. Evol. 35, 1547–1549 (2018).
47. Zhang, D. et al. PhyloSuite: An integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetic studies. Mol. Ecol. Resour. 20(4), 348–355 (2020).
48. Excoffier, L. & Lischer, H. E. Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. Mol. Ecol. Resour. 10(3), 564–567 (2010).
49. Tajima, F. Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. Genetics 123, 585–595 (1989).
50. Fu, Y. X. Statistical tests of neutrality of mutations against population growth, hitchhiking and background selection. Genetics 147(2), 915–925 (1997).
51. Drummond, A. J., Suchard, M. A., Xie, D. & Rambaut, A. Bayesian phylogenetics with BEAUti and the BEAST 1.7. Mol. Biol. Evol. 29(8), 1969–1973 (2012).
52. Rambaut, A., Suchard, M. A., Xie, D., & Drummond, A. J. Tracer v1.6. (2014). http://tree.bio.ed.ac.uk/software/tracer/ (accessed 23 Jan 2018).
53. Bohonak, A. J. IBD (isolation by distance): A program for analyses of isolation by distance. J. Hered. 93(2), 153–154 (2002).
54. Weir, B. S. & Cockerham, C. C. Estimating F-statistics for the analysis of population structure. Genetics 148(2), 951–962 (1998).
55. Goudet, J. fstat. html (2001).
56. Pritchard, J. K., Stephens, M. & Donnelly, P. Inference of population structure using multilocus genotype data. Genetics 155(2), 945–959 (2000).
57. Earl, D. A. & VonHoldt, B. M. STRUCTURE HARVESTER: A website and program for visualizing STRUCTURE output and implementing the Evanno method. Conserv. Genet. Resour. 4(2), 359–361 (2012).
58. Peakall, R. O. D. & Smouse, P. E. GENALEX 6: Genetic analysis in Excel. Population genetic software for teaching and research. Mol. Ecol. Notes 6(1), 288–295 (2006).
59. Jombart, T., Devillard, S. & Balloux, F. Discriminant analysis of principal components: A new method for the analysis of genetically structured populations. BMC Genet. 11(1), 1–15 (2010).
60. R Core Team. R: A Language and Environment for Statistical Computing (R Foundation for Statistical Computing, 2015).
61. Cornuet, J. M. et al. DIYABC v2.0: A software to make approximate Bayesian computation inferences about population history using single nucleotide polymorphism, DNA sequence and microsatellite data. Bioinformatics 30(8), 1187–1189 (2014).
62. Cornuet, J. M., Ravigne, V. & Estoup, A. Inference on population history and model checking using DNA sequence and microsatellite data with the software DIYABC (v1.0). BMC Bioinform. 11, 401 (2010).

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Author contributions
Y.-R.Y., X.-B.H. and H.D.L. conceived and designed the experiments; M.-R.Y., Z.-S.L., H.W., and C.-P.J. contributed to the sample collection. S.G., M.-R.Y., and Z.-S.L. contributed to the specimen examination; S.G., M.-R.Y. and H.-D.L. analyzed the data; S.G., Y.-R.Y., H.-D.L. and X.-B.H. wrote the manuscript.

Competing interests
The authors declare no competing interests.

Additional information
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Correspondence and requests for materials should be addressed to H.-D.L. or X.-B.H.

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