Nicotinic-nAChR signaling mediates drug resistance in lung cancer

Wan-Li Cheng1, Kuan-Yuan Chen1,2, Kang-Yun Lee1,2,3, Po-Hao Feng2,3, Sheng-Ming Wu2,3

1. Graduate Institute of Clinical Medicine, College of Medicine, Taipei Medical University, Taipei 11031, Taiwan
2. Division of Pulmonary Medicine, Department of Internal Medicine, Shuang Ho Hospital, Taipei Medical University, New Taipei City 23561, Taiwan
3. Division of Pulmonary Medicine, Department of Internal Medicine, School of Medicine, College of Medicine, Taipei Medical University, Taipei 11031, Taiwan

Corresponding author: Dr. Sheng-Ming Wu, Division of Pulmonary Medicine, Department of Internal Medicine, School of Medicine, College of Medicine, Taipei Medical University, 250 Wuxing Street, Taipei 11031, Taiwan. Tel: +886 (0)22490088 ext. 8742. E-mail: chitosan@tmu.edu.tw.

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Abstract

Lung cancer is the leading cause of cancer death worldwide. Cigarette smoking is the most common risk factor for lung carcinoma; other risks include genetic factors and exposure to radon gas, asbestos, secondhand smoke, and air pollution. Nicotine, the primary addictive constituent of cigarettes, contributes to cancer progression through activation of nicotinic acetylcholine receptors (nAChRs), which are membrane ligand-gated ion channels. Activation of nicotine/nAChR signaling is associated with lung cancer risk and drug resistance. We focused on nAChR pathways activated by nicotine and its downstream signaling involved in regulating apoptotic factors of mitochondria and drug resistance in lung cancer. Increasing evidence suggests that several sirtuins play a critical role in multiple aspects of cancer drug resistance. Thus, understanding the consequences of crosstalk between nicotine/nAChRs and sirtuin signaling pathways in the regulation of drug resistance could be a critical implication for cancer therapy.

Key words: nicotinic acetylcholine receptor, drug resistance, mitochondria, sirtuin, lung cancer

Introduction

Globally, lung cancer is greatest cause of cancer-related deaths. Lung cancer accounts for 14% and 12% of all cancers in men and women, respectively, and represents 24.6% of all cancer-related deaths [1]. Because of its extraordinary disease burden and international variability in trends of population growth, aging, and smoking behaviors, the global epidemiology of lung cancer requires continual monitoring [2]. Two main subtypes of lung cancer are small-cell lung carcinoma (SCLC) and non-SCLC (NSCLC), respectively accounting for 15% and 85% of all lung cancers [3]. NSCLC is further classified into three types: squamous cell carcinoma (SCC), adenocarcinoma, and large-cell carcinoma. SCC is the subtype of NSCLC strongly correlated with cigarette smoking [4]. NSCLC is a complex heterogenous disease with interpatient, intratumor, and inter-/intrametastatic heterogeneity at the subtype level [5]. Both epithelial growth factor receptor (EGFR) mutation and gene amplification status may be notable in determining chemoresistance in NSCLC [6]. Several studies have revealed that never-smokers with lung cancer are more responsive to EGFR-tyrosine kinase inhibitor (EGFR-TKI) therapy than smokers [7, 8]. Tumor progression and chemoresistance mediated by nicotine and its metabolites have been partly attributed to phosphoinositide 3-kinase (PI3K)/AKT, nuclear factor-κB (NF-κB), and mitochondrial signaling pathways [7, 9, 10]. Therefore, resistance is an inevitable barrier limiting lung cancer therapy effectiveness and reducing enthusiasm, making it today’s pervasive challenge for long-term disease control.

http://www.jcancer.org
Figure 1: Tobacco-specific N-nitrosamines are formed by the N-nitrosation of nicotine. NNN (N'-nitrosonornicotine) and NNK (4-(methylamino)-1-(3-pyridyl)-1-butanone (NNK) and N-nitrosonornicotine (NNN), present in tobacco smoke, can activate nAChRs signaling and stimulate multiple cancer-promoting signaling [18, 19]. Total 4-(methylamino)-1-(3-pyridyl)-1-butanol (NNAL), an NNK metabolite, is associated with the lung cancer risk in smokers [20]. These oncogenic metabolites may induce the formation of DNA adducts that leads to mutations of tumor suppressor genes including Rb and p53 [21]. Nicotine or NNK signaling may contribute to cancer progression [18, 22]. Nicotine was implicated in promoting the self-renewal of stem-like side-population cells from lung cancers. The subpopulation of cancer stem-like cells was implicated in tumor initiation, generation of heterogeneous tumor populations, metastasis, dormancy, and drug resistance [23]. Furthermore, nicotine can inhibit apoptosis induced by opioids, etoposide, cisplatin, and UV irradiation in lung cancer cells [24-26]. Therefore, the activation of nicotine signaling might be associated with drug resistance in lung cancer.

The potential mechanisms of Nicotine on lung carcinogenesis

Cigarette smoke is associated with an increased risk of all histological types of lung cancer [27]. In general, the lungs retain on average 60%–80% of mainstream smoke particulate matter and 90%–100% of nicotine after cigarette smoke inhalation [28]. The nicotine-derived metabolites including NNK and NNN are potent carcinogens because they bind to α7nAChR [14]. These metabolites and DNA interactions may be the primary cause of lung cancer in smokers. The binding activity of NNK to α7nAChR was 1,300 times greater than that of nicotine [29]. Different nAChR subunits were expressed in NSCLC cells of smokers and nonsmokers [30]. Nicotine present in the plasma of average smokers enhanced α7nAChR expression in human SCC cells [31]. SCC was most strongly associated with cigarette smoke, and adenocarcinoma, the most common lung cancer in never-smokers, has the weakest association. The higher expression of all affected nAChR subunits (α7, dupα7, α5, and α9) in smokers than in nonsmokers indicated that tobacco components upregulate the expression of a few nAChR subunit genes in SCC histologic type [32]. The literature has also indicated that only two subunits (α7 and α5) demonstrated significantly increased expression in SCC with a poor
prognosis [32]. Furthermore, patients with SCC who died presented significantly higher α7nAChR expression than patients with SCC who survived. For early-stage lung cancers, smoking cessation was associated with a large reduction in mortality risk [33], α5-nAChR was associated with lung cancer risk and onset, with exposure to a primary cancer etiologic factor (smoking duration and amount), and with the effects of a preventive action (smoking cessation) [34-37]. Chronic exposure to nicotine and derivatives can lead to the upregulation of all nAChRs in smokers. Activation of nicotine–nAChR signaling promotes cancer progression [18]. Upregulation of α7nAChR in lung cancer cells involved in the nicotine-induced tumor progression [38]. Homomeric α7nAChR was implicated as the primary receptor facilitating nicotine- and NNK-mediated cell proliferation [21]. The nicotine–α7nAChR axis can initiate cell invasion and the epithelial-mesenchymal transition (EMT) in NSCLC [39]. nAChRs, β-AR, and EGFR often coexpress on human lung cancer cells and airway epithelial or endothelial cells that might lead to cancer progression [40]. Nicotine signaling triggers the production of β-AR ligands, such as adrenaline and noradrenaline, which contribute to the development of lung cancer [21].

Survival analysis of a Cancer Genome Atlas (TCGA) lung cancer dataset demonstrated that high expression of acetylcholine receptors (AChRs) gene family such as CHRM2, CHRM3, CHRNA1, CHRNA2, CHRNA6, CHRN8B, or CHRN5 is associated with favorable prognosis in NSCLC adenocarcinoma, but that of CHRNA5/α5nAChR or CHRNA7/α7nAChR is associated with an unfavorable prognosis [41]. Among these proteins, only α7nAChR in the AChR family could affect prognosis in both lung adenocarcinoma and SCC [41]. High CHRNA1 expression is associated with reduced survival in early-stage lung adenocarcinoma after complete resection [42]. Notably, the levels of α7nAChR expression in SCC are higher than those in adenocarcinoma among patients with lung cancer, particularly in smokers [32]. Thus, α7nAChR is a potential target for lung cancer treatment.

Single nucleotide polymorphisms (SNPs) located on chromosome 15q25, which contains the nAChR subunits encoding by the CHRNA5, CHRNA3, and CHRN8B4, are associated with lung cancer risk [43]. Lung cancer risk was more than fivefold higher among individuals with both a family history of lung cancer and two copies of high-risk alleles rs8034191 or rs1051730 located in the 15q24-25.1 locus [44]. The change in rs6495309T>C on 15q25 may affect the activity of OCT1 binding to the CHRNA3 promoter, which contributes to CHRNA3 overexpression. These results confirm that 15q25 is a susceptibility region for lung cancer in Chinese individuals [45]. The gene cluster CHRNA5–CHRNA3–CHRN8B4 contributes to nicotine dependence risk in African–American and European–American individuals [46]. The missense SNP rs16969968 of CHRNA5 is significantly associated with both nicotine dependence and increased risk of lung cancer [47, 48]. The genome-wide associations of CHRNA3 and CHRNA5 on 15q25.1 were confirmed in familial SCC [49]. The CHRNA3 polymorphism functions as a genetic modifier of lung adenocarcinoma risk in the Chinese population, particularly in nonsmoking women [50]. CHRNA5 polymorphism is associated with lung adenocarcinoma risk in a population-based series of lung adenocarcinoma patients and healthy controls. An analysis of a family-based series of nonsmoker lung cancer cases and healthy controls indicated a similar trend. In addition, the same D398N variation correlated with CHRNA5 mRNA levels in the lungs of patients with adenocarcinoma [51]. Acetylcholine/nAChRs and muscarinic acetylcholine receptors can promote cancer cell and lung fibroblasts or myofibroblast proliferation, respectively [52, 53]. Chronic nicotine inhalation or endogenous acetylcholine released in the lungs may play a role in lung disease including COPD and lung cancer. Nicotine/nAChRs stimulates several signaling pathways conferring lung cancer cell survival, as described in the next section (Fig. 2).

The potential mechanisms of Nicotine on lung cancer progression

Nicotine may induce α7nAChR expression in human SCLC cells via the Sp1/GATA regulation signaling pathway [31]. α7nAChR expression levels are elevated in SCC compared with adenocarcinoma of the lung, particularly in smokers [32, 54]. High α7nAChR expression levels in lung cancer cells may be involved in the nicotine-induced tumorigenesis [32, 54]. α7nAChR levels in patients with SCC who are active smokers are correlated with their smoking history [31]. The function of α7nAChR-mediated lung cancer progression including in proliferation [31, 54-65], angiogenesis [66], and metastasis [39, 67-69], has been revealed (Fig. 2). The tumor-promoting effect is mediated by α7nAChR signaling pathways described in subsequent subsections.

Cell proliferation

α7nAChR mediates the proliferative effects of nicotine in lung cancer cells [70]. Nicotine/α7nAChR signaling enhances NSCLC cell proliferation by scaffolding protein β-arrestin-mediated activation of the Src and Rb-RAF protooncogene serine/
threonine-protein kinase (RAF-1) pathways [55]. Moreover, continuous exposure to nicotine in SCC of the lungs results in α7nAChRs upregulation, which may enhance tumor growth [31]. Nicotine stimulates tumor growth and ERK activation in a murine orthotopic model of lung cancer [54]. A blockade of α7nAChRs suppresses nicotine-induced lung cancer cell growth and vimentin expression through the MEK/ERK signaling pathway [63]. Consistent with these results, nicotine increased expression of nAChR and stimulated proliferation of SCC cell line [65]. α7nAChR may mediate the proliferative activity of nicotine in poorly differentiated NSCLC [56]. Nicotine-induced α7nAChR and α9nAChR expression in NSCLC cells, along with p-CREB and p-ERK1/2 activation accompanied by increased noradrenaline, leading to cell proliferation [57]. Nicotine/α7nAChR promoted proliferation in human SCLC cells via the Sp1/GATA regulation signaling pathway [31]. NNK/α7nAChR may increase cell growth in SCLC cells via an influx of Ca²⁺ [60]. Nicotine stimulates NSCLC cell proliferation and PPARβ/δ expression through activation of PI3K/mTOR signals that suppress AP-2α binding activity to PPARβ/δ promoter [61]. α7nAChRs or α9nAChRs mediated nicotine-induced cell proliferation and activation of the AKT and ERK signaling pathways [64]. The activated cell-membrane α7nAChRs formed complexes with EGFR, whereas activated mitochondrial α7nAChRs was physically associated with the intramitochondrial protein kinases PI3K and Src that increased expression of cyclin D1 and activation of ERK1/2 to lead to lung cancer proliferation [71]. α-Cobratoxin, a high-affinity α7-nAChR antagonist reduced tumor growth in nude mice orthotopically engrafted with NSCLC cells [59]. An α7nAChR inhibitor (APS8) may suppress NSCLCs proliferative effects of nicotine [58]. These studies revealed that nicotine/α7nAChR signals mediate proliferation in lung cancer.

**Metastasis**

Cigarette smoke status and history are associated with lung cancer metastasis [72, 73]. α7nAChRs may mediate cancer cell growth depending on NSCLC differentiation status [56]. α7nAChR and heteromeric nAChRs can promote tumor invasion in NSCLC [56]. Nicotine/α7-nAChR can induce NSCLC cell migration and invasion via the MEK/ERK signaling pathway [39]. NNK promotes lung cancer cell migration and contactin-1 expression via the α7nAChR-mediated ERK signaling pathway [67]. Moreover, NNK enhances lung cancer cell migration and invasion via activation of the c-Src-PKCIota–FAK signaling axis [68]. β-Cryptoxanthin may repress lung cancer cell motility through the downregulation of α7nAChR/PI3K signaling [69]. Nicotine/α7nAChR signaling enhance migration and
the expression of SOX2 in NSCLC cell lines through the YAP-E2F1 signaling axis [23]. These studies suggest that α7nAChR can enhance lung cancer cell metastasis through the activation of different signaling pathways.

**Angiogenesis**

α7nAChR enhances angiogenesis via the PI3K/AKT pathway and NF-κB activation, which is partially dependent on vascular endothelial growth factor (VEGF) [74]. Nicotine/α7nAChR signaling mediates proangiogenic effects through angiogenesis and EMT [75]. MG624, an α7nAChR antagonist, reduces nicotine-induced early growth response gene 1 (Egr-1) binding activity to the fibroblast growth factor 2 (FGF2) promoter that inhibits angiogenic effects in SCLC [66]. Thus, α7nAChR may facilitate lung cancer progression including angiogenesis; however, its detailed effect requires investigation.

**Anti-Inflammation**

α7nAChR attenuates ventilator-induced lung injury and plays an anti-inflammatory role in several inflammatory diseases [76-78]. Activation of inflammation-related receptors, such as toll-like receptors, enhances NF-kB signaling pathways in both acute and chronic inflammation that can considerably increase cancer risk [79]. Choline/α7nAChR signaling modulates TNF release via the inhibition of NF-kB activation [80]. Moreover, nicotine/α7nAChR mediates anti-inflammatory action on macrophages via recruitment and activation of JAK2, initiating the STAT3 and SOCS3 signaling cascade [81]. Nicotine/α7nAChR suppresses TNF-α expression in human airway epithelial cells by inhibiting MyD88 and NF-kB activity [82]. An anti-inflammatory study revealed that α7nAChR signaling inhibits NLRP3 inflammasome activation by preventing mitochondrial DNA release [83]. Accumulating evidence suggests that the vagus nerve may modulate lung infection and inflammation through the α7nAChR signaling pathway [84]. Thus, α7nAChR may be a potential target for attenuating inflammatory cytokine production in lung diseases [85].

Exposure to nicotine adversely affects dendritic cells, a cell type that has an important role in anticancer immunosurveillance [86]. Nicotine may suppress anticancer immunity by increasing or reducing number of regulatory T cells and T helper 17 cells, respectively [87]. Moreover, nicotine inhibits the cytotoxic activity of natural killer (NK) cells, and the effect of nicotine on NK cells can be abolished by β2nAChR deficiency [88]. However, to understand immune regulation and progression in lung cancer, the roles of nicotine-mediated pathways in distinct immune cells warrant investigation.

**The potential mechanisms of sirtuins on cancer progression**

Sirtuins play diverse roles in controlling the cell cycle and proliferation in response to stress, thus promoting cell survival, apoptosis, or senescence [89]. Sirtuin 1 (SIRT1) has been involved in decision making over cellular senescence or apoptosis in lung diseases including COPD and cancer [53, 90]. Cellular senescence is caused by replicative and stress-related senescence with p53 and p16 activation, respectively, leading to p21 activation and cell-cycle arrest. SIRT1 can function as p53 deacetylase that blocks p53-dependent pathways, thereby regulating cell-cycle and inactivating apoptotic process [91]. SIRT1 and p53 play vital roles in maintaining genomic stability and integrity, which favor cell survival and protect against tumorigenesis. Increased DNA damage and cellular senescence may contribute to accelerated lung aging and COPD pathogenesis [53]. SIRT6 can significantly suppress TGF-β-induced senescence of human bronchial epithelial cells via p21 degradation [92]. Moreover, SIRT6 may play a pivotal role in inhibiting fibrosis. SIRT1 and SIRT6 prevent telomere dysfunction during DNA replication [93]. SIRT1 and SIRT6 loss may promote aging and arise from oxidative stress through PI3K-mTOR signaling activation [94].

Sirtuins are responsible for cellular metabolic reprogramming and drug resistance by inactivating cell death pathways and promoting uncontrolled proliferation [90]. SIRT1 overexpression is associated with poor prognosis in patients with lung cancer [95]. SIRT1-mediated survival of cancer cells contributes to chemoresistance in tumors. Furthermore, SIRT1 is considerably expressed in the brain metastatic tissues of patients with NSCLC [96]. SIRT1 may promote breast cancer progression through modulating AKT activity [97]. SIRT1 was significantly overexpressed in human prostate cancer cells, and SIRT1 inhibition contributes to suppress cancer cell growth [98]. SIRT1 limits prostatic intraepithelial neoplasia in SIRT1-knockout mice. In A/J mice, β-cryptoxanthin, strongly associated with reduced lung cancer risk, restores nicotine-reduced lung SIRT1 levels to that normal and inhibits nicotine-promoted lung tumorigenesis and emphysema [99]. In addition, SIRT1 levels decrease in some human cancers including glioma, bladder, and ovarian cancer [100]. SIRT1 also acts as a tumor suppressor via the c-Myc-SIRT1 feedback loop that regulates cell growth and transformation [101]. Thus, the tumor suppressor or promoter role of sirtuins in cancer progression may
depend on their tissue- and cancer-specific expression and the examined conditions. Similarly, SIRT6 plays functions of both tumor promoter and suppressor in the development of different cancer, potentially depending on specific tissue [90].

The SIRT1 inhibitor sirtinol induces senescence-like growth arrest through impaired activation of RAS-mitogen-activated protein kinase (MAPK) signaling in human lung cancer cells [102]. The combination of inauhzin (SIRT1 inhibitor) and cisplatin or doxorubicin can considerably reduce NSCLC cell growth in a p53-dependent manner [103]. Two SIRT2 inhibitors AEM1 and AEM2 can induce p53-dependent proapoptotic activity in NSCLC cells [104]. Moreover, AEM2 demonstrates similar inhibition of SIRT2 as AC-93253. SIRT1/2 inhibitor, salermide, can increase death receptor 5 expression via the ATF4–ATF3–CHOP axis and contribute to NSCLC cell apoptosis [105]. Therefore, sirtuins play notable roles in both lung cancer development and the nicotine/nAChR-regulated signaling (Fig. 3). Although several sirtuins can function as both tumor promoters and suppressors, SIRT1/3/5–7 blockade may aid in effective chemotherapy, as described in the next section.

The potential mechanisms of Nicotine and sirtuins on drug resistance

Nicotine may reduce the cytotoxic effects of chemotherapy and radiotherapy that cause poor therapeutic response [87, 106]. Nicotine-mediated tumor-promoting effects are apparently mediated by nAChRs expressed on cell membranes and by mitochondria [87]. Additionally, mitochondria are critical mediators of cancer progression, as this process requires flexibility to adapt to cellular and environmental alterations in addition to cancer therapies [107]. Nicotine-impaired metabolism and mitochondrial defects were critical in metabolic responses to cancer progression [108]. Activation of cell-membrane and mitochondrial nAChRs produces a combination of growth-promoting and antiapoptotic signals that implemented the tumor-promoting action of nicotine in lung cells [71]. Furthermore, nAChRs were identified to control either CaKMII or Src-dependent signaling pathways in mitochondria that protect cells from apoptosis [109].

Nicotine also permeated cells and activated mitochondrion–nAChRs coupling to inhibit...
mitochondrial permeability transition pore (mPTP) opening, preventing apoptosis [62]. Nicotine-induced survival may occur by a mechanism of multisite phosphorylation of BAD, which may lead to human lung cancer and/or chemoresistance development [110]. Activation of nicotine-α7nAChR signaling can trigger membrane depolarization, which activates voltage-gated calcium channels and subsequently activates the MAPK pathway, possibly increasing B-cell lymphoma-2 (Bcl-2) expression and apoptosis downregulation [111]. Nicotine prevents cisplatin-mediated apoptosis by regulating α5nAChR/AKT signaling and several mitochondria proteins including Bcl-2, Bax, survivin, and caspase 3 in gastric cancer cells [112]. Long-term nicotine exposure-induced chemoresistance is mediated by STAT3 activation and ERK1/2 downregulation via nAChR and β-AR in bladder cancer cells [113]. Emerging evidence suggests that feedback activation of STAT3 signaling is a common cause of drug resistance to receptor tyrosine kinase-targeted therapies and conventional chemotherapy [114]. Long-term exposure to NNK combined with arecoline activated EGFR/AKT signaling is involved in antiapoptosis, cancer stem cell properties, and cisplatin resistance in head and neck SCC (HNSCC) cells [115]. Nicotine/α9nAChR-PPMIF signaling can attenuate p-p53 (Ser-20)– and p-BAX (Ser-184)–induced proapoptotic pathways [116]. Therefore, nAChRs may be a promising molecular target to arrest lung cancer progression and re-open mitochondrial apoptotic pathways. Nicotine can induce erlotinib resistance via the crosstalk between α1nAChR and EGFR/AKT/ERK signaling pathways in NSCLC [117]. A recent study suggested that smoking containing nicotine causes resistance to erlotinib therapy in NSCLC [118].

Sirtuins can exert their capacity to respond to environmental changes and their expression is often altered in cancer [89]. SIRT1, SIRT3, SIRT4, and SIRT7 are strongly expressed in lung adenocarcinoma, whereas SIRT5 is highly expressed in SCC [119]. Analysis of the TCGA NSCLC dataset revealed that high expression levels of SIRT2/6 were associated with longer overall survival (OS) [119]. However, high SIRT6 expression was associated with poor OS in 98 patients with NSCLC [120]. Nicotine enhanced oxidative stress and activates NF-κB [121]. Several sirtuins are critical in inhibiting excessive, damaging levels of ROS that drive cancer drug resistance [122]. Nuclear SIRT1 promotes ROS stress resistance via the deacetylation of several transcriptional regulators, including p53, forkhead homeobox type O (FOXO) proteins, PGC-1α, heat shock factor protein 1 (HSF1), and nuclear erythroid factor 2-related factor 2 (NRF2), and this contributes to antioxidant production [123]. Studies have suggested that mitochondrial-sirtuins (SIRT3, SIRT4, and SIRT5) are members of a family of NAD+-dependent deacetylases and are implicated in the oxidative stress response through the regulation of mitochondrial metabolism and antioxidant mechanisms [124-126]. Mitochondrial SIRT3 may coordinate ROS; SIRT5 also limits ROS by activating SOD1 and NRF2 to maintain cellular redox homeostasis [122, 126, 127]. Thus, sirtuins may promote cancer cell survival by limiting ROS that would lead to cancer drug resistance. Several recent studies have supported the presence of sirtuins-mediated drug resistance with human cancers (Table 1). Some sirtuins regulate lung cancer progression, as described in the next subsection, and signaling molecules are associated with drug resistance.

**SIRT1**

Nicotine can upregulate SIRT1 expression in a time- and concentration-dependent manner [128]. BaP, a carcinogen in cigarette smoke, can induce SIRT1 in human bronchial epithelial cells [129]. SIRT1 is involved in BaP-induced transformation associated with TNF-α-β-catenin axis activation and is a potential therapeutic target for lung cancer [129]. The SIRT1–PARP-1 axis plays a critical role in the regulation of cigarette smoke-induced autophagy and has notable implications for understanding the mechanisms of cigarette smoke-induced cell death and senescence [130]. α7nAChR-SIRT1 axis activation alleviates angiotensin II-induced VSMC senescence [131]. Recent studies have focused on the biological functions of SIRT1 in metabolic diseases, cancer, aging and cellular senescence, inflammatory signaling in response to environmental stress, and cell survival [132-135]. SIRT1 expression was a strong predictor for poor OS and progression-free survival in patients with NSCLC who underwent platinum-based chemotherapy [136]. Silencing of SIRT1 could significantly enhance the chemosensitivity of lung cancer cells to cisplatin treatment [136]. SIRT1 was negatively associated with proapoptotic factors BAD, BAX, and BID in TCGA NSCLC patients [119]. SIRT1 suppression sensitizes lung cancer cells to WEE1 inhibitor-induced DNA damage and apoptosis [137]. SIRT1 deacetylates and inactivates p53, allowing cells to bypass apoptosis [138]. The transcription factor FOXO3a may induce the expression of several antioxidant genes, including Mn superoxide dismutase (MnSOD), catalase, peroxiredoxins 3 and 5 (Prx3 and Prx5, respectively), thioredoxin 2 (Trx2), thioredoxin reductase 2 (TR2), and uncoupling protein 2 (UCP-2) [139]. SIRT1-mediated deacetylation of FOXO3a increases
cell survival in response to oxidative stress [140]. Moreover, SIRT1 may play a role in the acquisition of aggressiveness and chemoresistance in ovarian cancer and have potential as a therapeutic target for ovarian cancer [141].

Table 1. Effects of sirtuins on cancer drug resistance

| Types of sirtuin | Types of cancer | Drug | Effect of cancer drug resistance | Year of publication | Ref |
|-----------------|-----------------|------|---------------------------------|--------------------|-----|
| SIRT1           | CRC             | TRAIL| miR-128 suppressed SIRT1 expression to sensitize TRAIL-induced apoptosis (142) | 2018               | [177] |
| SIRT1           | Breast cancer   | Paclitaxel | A SIRT1-PBRX1-LK4-ALDH1 circuitry regulates breast cancer stemness and metastasis. KLF4 inhibitor Kemppaallein sensitizes breast cancer cells | 2018               | [178] |
| SIRT1           | Cervical cancer | Paclitaxel | Knockdown of SIRT1 promotes apoptosis of paclitaxel-resistant human cervical cancer cells | 2018               | [179] |
| SIRT1           | ATC             | Doxorubicin | 6-phosphogluconate dehydrogenase (6PGD) was critically involved in ATC resistance to doxorubicin. Decreased enzymatic activity of SIRT1 in response to 6PGD inhibition in doxorubicin-resistant ATC cells | 2018               | [180] |
| SIRT1           | Cervical cancer | Doxorubicin | β2-AR activation induces chemoresistance by modulating p33 acetylation through upregulating Sirt1 in cervical cancer cells | 2017               | [181] |
| SIRT1           | Gastric cancer  | Cisplatin | miR-132 regulated SIRT1/CREB/ABCG2 signaling pathway contributes to cisplatin resistance | 2017               | [182] |
| SIRT1           | HCC             | Oxaliplatin | LncRNA HULC triggered autophagy by stabilizing SIRT1 and attenuates chemosensitivity of HCC cells | 2017               | [183] |
| SIRT1/2         | Breast and Cervical cancer | Etoposide | Cancer cells with low SIRT1 levels maintained their resistance and survival by increasing SIRT3 expression | 2017               | [148] |
| SIRT1           | Prostate cancer | Docetaxel | The UCA1-miR-204-SIRT1 axis modulates docetaxel sensitivity of prostate cancer cells | 2016               | [184] |
| SIRT1           | Breast cancer   | Tamoxifen | Brachyury mediates tamoxifen resistance by regulating SIRT1 | 2016               | [185] |
| SIRT1           | Bladder cancer  | Cisplatin | Cisplatin inhibited multiple bladder cancer cells by inhibiting tumor-associated NADH oxidase (NNOX) and SIRT1 | 2016               | [186] |
| SIRT1           | ATL             | Etoposide | SIRT1 inhibition enhances chemosensitivity and survival of ATL cells by reducing DNA double-strand repair | 2015               | [187] |
| SIRT1 Endometrial carcinoma | Cisplatin and paclitaxel | SIRT1 overexpression significantly enhanced drug resistance. Selective SIRT1 inhibitor (EX327) significantly increased chemosensitivity | 2015               | [188] |
| SIRT1 CML       | Hep90 inhibitors (17-AAG and AU7922) | SIRT1 inhibitors (amuresin G and EX327) effectively potentiated sensitivity of Hep90 inhibitors | 2015               | [189] |
| SIRT1 ESCC      | Cisplatin | Overexpression of SIRT1 may cause resistance of ESCC cells to cisplatin through Noxa expression | 2015               | [190] |
| SIRT1 PC        | 5-fluorouracil (5-FU) and gemcitabine | Overexpression of miR-494 inhibited chemoresistance of PC by downregulating SIRT1 and e-Myc | 2015               | [191] |
| SIRT1 CRC       | 5-FU | SIRT1/P53- and P73-dependent increase in oxidative phosphorylation leads to CRC drug resistance | 2015               | [192] |
| SIRT1 CML       | Imatinib | Divalproex sodium enhances antileukemic effects of imatinib in CML through SIRT1 | 2015               | [193] |
| SIRT1 AML       | TKI (Quinazolinib, AC220) | Inhibition of SIRT1 by SIRT1 inhibitor Teno1n-6 (TV6) enhanced TKI-mediated sensitivity | 2014               | [194] |
| SIRT1 Thyroid cancer | Etoposide | SIRT1-Fox3 signaling confers drug resistance | 2014               | [195] |
| SIRT1 Breast cancer | TRAIL | Metformin mediates miR-34a to suppress the SIRT1/PGC-1α/NRF2 pathway and increases drug sensitivity | 2014               | [196] |
| SIRT1 CML       | 5kDa (imatinib, nilotinib or dasatinib) | All-trans-retinoic acid (ATRA) effectively blocked acquisition of BCR-ABL mutations and resistance. ATRA inhibited NAD+ dependent SIRT1 deacetylase via CD38 expression | 2014               | [197] |
| SIRT2 RCC       | 5-FU | SIRT2+ cells mediates RCC drug resistance | 2018               | [198] |
| SIRT2 AML       | Dexamethasone, arabinocytidine | SIRT2 mediates multidrug resistance in AML cells via ERK1/2 signaling pathway | 2016               | [199] |
| SIRT2 Melanoma  | Doxorubicin | AC-9325, a SIRT2 inhibitor increases drug sensitivity | 2015               | [200] |
| SIRT3 Synovial sarcoma | Pazopanib | Knockdown of SIRT3 confers increased resistance to chemotherapeutic agents | 2018               | [201] |
| SIRT3 HCC       | Sorafenib | SIRT3 protein expression was significantly higher in patients treated with metformin | 2017               | [202] |
| SIRT3 Glioma    | Linalool | Overexpression of SIRT3 significantly inhibited a finalolinduced increase of mitochondrial ROS production and apoptotic cell death | 2017               | [203] |
| SIRT3 Breast cancer | Cisplatin | SIRT3 silencing sensitizes breast cancer cells to cytotoxic treatments through ROS production | 2017               | [134] |
| SIRT4 CRC       | 5-FU | SIRT4 increased the sensitivity of CRC cells to 5-FU | 2016               | [204] |
| SIRT5 NSCLC     | CDDP, 5-FU or bleomycin | SIRT5 facilitates cancer cell growth and drug resistance in NSCLC cells | 2014               | [127] |
| SIRT6 NSCLC     | Doxorubicin | SIRT6 increased doxorubicin resistance via FOXO3 activity | 2018               | [205] |
| SIRT6 NSCLC     | Gefitinib | Astragaloside IV sensitizes NSCLC cells to gefitinib partially via regulation of SIRT6 | 2017               | [206] |
| SIRT6 PC        | Gemcitabine | Quinazolinedione SIRT6 inhibitors sensitize cancer treatment | 2015               | [207] |
| SIRT6 NSCLC     | Paclitaxel | SIRT6 knockdown NSCLC cells improved drug sensitivity | 2015               | [120] |
| SIRT6 Breast cancer | Trastuzumab (Herceptin) | MDMA-mediated degradation of SIRT6 phosphorylated by AKT1 promotes drug resistance | 2014               | [208] |
| SIRT7 NSCLC     | Gemcitabine | Depletion of SIRT7 promoted drug sensitivity | 2018               | [151] |
| SIRT7 Breast cancer, osteosarcoma, and ovarian cancer | Cisplatin, Doxorubicin | SIRT7 inhibition significantly increases stress resistance and modulates insulin/IGF-1 signaling pathways | 2018               | [209] |

CRC, Colorectal cancer; ATC, Anaplastic thyroid carcinoma; HCC, Hepatocellular carcinoma; ATL, Adult T-cell leukemia-lymphoma; CML, Chronic myeloid leukemia; ESCC, Esophageal squamous cell carcinoma; PC, Pancreatic cancer; AML, Acute myeloid leukemia; RCC, Renal cell carcinoma; and NSCLC, Non–small-cell lung carcinoma.
Figure 4: Schematic of mediation of tumor-promoting actions by nicotine/nAChR. Nicotine interacts with nAChR and stimulates activation and crosstalk with β-AR and EGFR downstream, signaling to promote cancer progression. Activation of nAChRs and β-AR mediates EGF secretion to further transactivate EGFRs. In cancer cells, the signaling pathways downstream of nAChRs promote drug resistance and antiapoptosis by activating the transcription factors including STAT, NF-κB, Jun/Fos, and E2F through JAK, PI3K/AKT, RAS, RAF, and the MAPK signaling cascade. Mitochondrial nAChRs trigger phosphatidyl-inositol-3-kinase (PI3K) and AKT signaling pathways that prevent mPTP opening and cytochrome c release. Nicotine-induced antiapoptosis and drug resistance may include several mechanisms involved in overexpression of sirtuin proteins, phosphorylation of BAD, and blockade of BAX translocation, leading to tumor cell development. SIRT3 and SIRT5 are mitochondrial proteins. SIRT6 and SIRT7 are localized in the nucleus. SIRT1-mediated deacetylation of FOXO3a can induce expression of antioxidant enzymes including MnSOD and catalase that increase cell survival during cellular oxidative stress. Consequently, nicotine/nAChR mediates antipapoptotic pathways and concurrently crosstalks with β-AR or EGFR signaling activation may lead to cancer progression. N: nicotine; Ac: acetylation; NNK: 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone.

SIRT2–SIRT7

SIRT3, located in mitochondria, is correlated with NSCLC malignancy [142]. α7nAChRs activation inhibits platelet-derived growth factor-induced cells migration by activating the mitochondrial deacetylase SIRT3, implying a critical role for α7nAChRs in mitochondrial biology and PDGF-related diseases [143]. The activity of SIRT3 can protect cancer cells from chemotherapy-induced oxidative stress [144, 145]. Additionally, SIRT3 promotes the activation of AKT signaling pathways in NSCLC [142]. SIRT3 promoted p53 degradation in PTEN-deficient NSCLC cell lines via the ubiquitin–proteasome pathway [146]. SIRT3 can also mediate FOXO3a nuclear translocation that activates MnSOD and catalase expression [147]. Notably, SIRT1 knockdown cells can increase SIRT3 expression and cell survival and have relatively high resistance to H2O2 or etoposide treatment [148]. SIRT3 might be a therapeutic target for breast cancer, improving the effectiveness of cisplatin and tamoxifen treatments [124]. SIRT5 prevents cigarette smoke extract-induced apoptosis in lung epithelial cells via FOXO3 deacetylation [127]. Besides, SIRT5 knockdown makes lung cancer cells more sensitive to drug (cisplatin, 5-fluorouracil or bleomycin) treatment [127]. SIRT5 depletion suppresses the expression of NRF2 and its downstream drug-resistance genes [127].

Patients with high cytosol expression but low nuclear expression of SIRT6 can have poor clinical outcomes of lung cancer [120]. Furthermore, SIRT6 knockdown in NSCLC cell lines can improve paclitaxel sensitivity by reducing NF-κB and Beclin1 (autophagy mediator) levels [120]. Cigarette smoke increases SIRT6 expression in vivo (mice) and in vitro (rheumatoid arthritis synovial fibroblasts) [149]. Reduced SIRT6 expression mediates the augmentation of radiation-induced apoptosis via cAMP signaling in lung cancer cells [150]. A recent report revealed that SIRT7 depletion promotes
gemcitabine-induced cell death [151]. Functioning as an oncogene, SIRT7 can be suppressed by miR-3666, which could increase NSCLC cell apoptosis [152].

Thus, the aforementioned studies together have demonstrated tumor progression modulated by the SIRT1, SIRT3, and SIRT5-7, along with the tumor-suppressive effects of SIRT2 and SIRT4. SIRT2 mediates the ROS production and p27 levels, leading to lung cancer cell apoptosis and cell-cycle arrest [153]. SIRT2 overexpression increases NSCLC cells’ sensitivity to cisplatin treatment [153]. Moreover, recent findings suggest that SIRT4 inhibits lung cancer progression through mitochondrial dynamics mediated by the ERK-Drp1 pathway [154]. At present, one clinical trial (NCT02416739) is studying the combinatorial effects of the human sirtuin inhibitor (nicotinamide) and EGFR-TKI in NSCLC. The discovery of specific SIRT regulation and EGFR-TKI treatment would help elucidate the roles of sirtuins in lung cancer development. Although sirtuin clearly is critical in carcinogenesis, the crucial mechanisms by which the nicotine-mediated signaling or specific sirtuin pathways in different cell context lead to drug resistance require elucidation.

Cell-membrane nAChRs implement upregulation of proliferative and survival genes [62]. Nicotine can promote oral precancerous growth through suppression of apoptosis by upregulating α7nAChR and peroxiredoxin [155]. α7nAChR-mediated cell protection, through JAK2/PI3K/AKT/signal transducer and activator of transcription 3(STAT3)/NF-κB activation, leads to Bcl-2 production [156]. Nicotine binds to nAChRs and stimulates secretion several factors including epidermal growth factor (EGF), VEGF, and neurotransmitters [157]. Nicotine/nAChRs mediates EGF secretion and subsequent EGFR signaling activation, thus contributing to antiapoptosis [18]. Nicotine and NNK also bind to β-ARs and promote survival signaling cascades [18, 30]. Moreover, tissue-specific expression of α7β2, α3β2, α3β4, and α4β2 nAChRs located in the mitochondria outer membrane with anion channels that regulate the release of proapoptotic cytochrome c or ROS production has been observed [78, 158, 159]. nAChR signaling in mitochondria is stimulated and engages PI3K/AKT kinases, similar to those activated by plasma membrane nAChRs. Nicotine contributes to progression and erlotinib resistance in an NSCLC xenograft model through the nAChR-EGFR cooperation [117]. The nicotine-mediated α5nAChR/AKT signaling pathway prevents cisplatin-induced cancer cell apoptosis [112]. Blockade of α7nAChRs inhibited nicotine-induced tumor growth and vimentin expression in NSCLC through the RAS-RAF-MAPK kinase (MEK)-extracellular signal-regulated kinase (ERK) signaling pathway [63]. The nicotine and derivatives may mediate oncogenic signaling via nAChR, β-AR, and EGFR and combined with the effects of antiapoptosis in mitochondria that contribute to cancer progression (Fig. 4). The nicotine/nAChR signaling crosstalk with SIRT1/3/5-7 may contribute to cancer drug resistance.

Conclusions and Future Directions

Genome-wide association studies have indicated a strong link between nicotine/nAChRs and lung cancer risk [34, 38]. Nicotine might lead to suppressed apoptosis and cisplatin resistance via α5nAChR/AKT signaling [112]. In addition, α7nAChR may be implicated in the NAD+/SIRT1 pathway, which promotes chemotherapeutic drug resistance [131]. Nicotine/α9nAChR signaling can reduce apoptotic pathways [116]. Nicotine-mediated cancer progression is mediated by nicotine/nAChRs signaling. By contrast to the tumor-suppressing role of SIRT2/4, the roles of SIRT1/6/7 and mitochondrial SIRT3/5 are strongly associated with drug resistance in lung cancer [119, 120, 124, 135, 142, 144, 146, 150-154]. However, SIRT1/2 inhibitor, sirtinol, can increase anticancer potential and chemosensitivity in several cancer cells [102, 160, 161]. SIRT1 may have contradictory roles in promoting or suppressing in different cell contexts or types of tumor development [162]. Moreover, shikonin causes apoptosis in some lung cancer cell lines via the FOXO3a/EGR1/SIRT1 signaling pathway activation [163]. In contrast to the tumor-suppressing role of SIRT1, the role of shikonin involves promoting liver cancer cell death by downregulating the SIRT1-MDR1/P-gp signaling pathway [164]. However, emerging evidence suggests that SIRT1 can promote cancer drug resistance (Table 1). Thus, an investigation to unravel cancer cellular contexts in which ROS are beneficial or harmful, with sirtuins having tumor-suppressing or promoting functions, would contribute to the literature. Thus, a series of more comprehensive studies are necessary to validate the underlying molecular mechanisms of the nicotine-mediated pathway with specific sirtuin and related targets in different types of cancer.

Nicotine activates α7nAChR and β2nAChR, and these two factors are associated with EGF and VEGF receptors, respectively. However, activated mitochondrial α7nAChRs and β4nAChR interacted with the PI3K and Src [62]. This interaction leads to increased cell proliferation and ROS-mediated apoptosis resistance through ERK signaling and mPTP inhibition, respectively [62]. The same group of researchers demonstrated a nicotine-mediated growth-promoting effect via interaction of α7nAChR with that EGF, α3nAChR with VEGF, α4nAChR with
insulin–like growth factor I (IGF-I) and VEGF, whereas α7nAChR was with EGF, IGF-I, and VEGF. Similar to the binding affinity of cell-membrane nAChRs, nicotine can bind to mitochondrial nAChRs. Mitochondrial nAChRs coupled with the inhibition of mPTP opening causes tumor progression [71]. These results indicate that nicotine-mediated activation of the cell-membrane or mitochondrial nAChRs leads to tumor-promoting and antiapoptotic signals in tumor development. Therefore, nAChRs may be a potential molecular target to suppress lung cancer progression and trigger mitochondrial apoptotic pathways.

An accumulating body of evidence is demonstrating that long noncoding RNAs (lncRNAs) have various biological functions, including modulation of growth, cell differentiation, drug resistance, and cancer progression [165-167]. MALAT-1 is highly expressed and mediates poor progression in lung cancer [168]. LncRNA-SCAL1 (smoke and cancer-associated lncRNA1) can alleviate CS-mediated oxidative stress in airway epithelial cells [169]. Although lncRNAs were to be dysregulated in various human diseases, how lncRNAs connect with environmental exposures remains largely unknown. GAS5 may be proapoptotic and increase sensitivity to cell death due to environmental stressors [170, 171]. HOTAIR mediates EMT because of cigarette smoke extracts [172], and similar studies have indicated that MALAT-I is involved in cigarette smoke extract-induced EMT and malignant transformation [172]. Circulating extracellular vesicles (EVs) can transfer biomolecules, including ligands, cytokines, and genetic information, to recipient cells to enact functional changes [173]. EVs can contain relatively stable RNA species, including small noncoding RNAs and lncRNAs [174]. LncARSR is highly expressed in sunitinib-resistant renal cancer cells, and these drug-resistant cancer cells can transfer lncARSR to drug-sensitive recipient cells and acquire chemoresistance via exosomes [175]. Exosomes mediated transfer of lncRNA UCA1 can considerably increase tamoxifen resistance in estrogen receptor (ER)-positive breast cancer cells [176]. Thus, lncRNAs may cause cancer-acquired chemoresistance via exosomal pathway. However, whether nicotine promotes cancer drug resistance via the tumor environment communication remains unclear. Thus far, nicotine-mediated drug resistance induced by lncRNAs in lung cancer remains unclear. Therefore, nicotine-mediated drug resistance through lncRNA regulation in NSCLC warrants further exploration.

Over the past few years, nAChRs have been found to be selectively overexpressed in various cancers, including lung cancer. Targeting nAChRs signaling pathways can significantly attenuate nicotine-associated drug resistance. Several α7nAChR antagonists are potential anticancer drugs [38]. Collectively, studies have suggested that nAChR-induced antiapoptotic effects may play notable roles in drug resistance in lung cancer. Elucidation of the consequences of nicotine/nAChRs signaling crosstalk with other pathways and specific sirtuins may be crucial to therapeutic implications. These studies may facilitate the design of useful therapeutic strategies to reverse chemoresistance in patients with different types of cancer.

**Implication**

Nicotine and derived metabolites are associated with lung cancer risk in smokers. The α7nAChR is significantly upregulated in NSCLC and correlates with its unfavorable prognosis. Activation of nicotine/α7nAChR signaling leads to lung cancer progression. Studies have suggested that nicotine/nAChR axis and SIRT1/3/5-7 mediates cancer drug resistance. Blockade of the signaling mediated by nicotine/nAChR and specific sirtuins may enhance the efficacy of chemotherapy.

**Abbreviations**

SCLC: small-cell lung carcinoma; NSCLC: non-small-cell lung carcinoma; EGFR: epithelial growth factor receptor; EGFR-TKI: EGFR-tyrosine kinase inhibitor; NF-κB: nuclear factor-κB; PI3K: phosphoinositide 3-kinase; BaP: benzo[a]pyrene; PAH: polycyclic aromatic hydrocarbons; nAChRs: nicotinic acetylcholine receptors; β-AR: β-adrenergic receptors; NNK: 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanol; NNN: N-nitrosonornicotine; NNAL: 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanol; SNP: single nucleotide polymorphism; RfA-1: proto-oncogene serine/threonine-protein kinase; MEK: mitogen-activated protein kinase; ERK: extracellular signal–regulated kinase; MAPK: mitogen-activated protein kinase; EMT: epithelial-to-mesenchymal transition; SOX2: sex-determining region Y-box 2; VEGF: vascular endothelial growth factor; Egr-1: growth response gene 1; FGFR2: fibroblast growth factor 2; STAT3: signal transducer and activator of transcription 3; NLRP3: nucleotide-binding domain and leucine-rich repeat containing protein 3; ASC: adult stem cell; ROS: reactive oxygen species; NAC: N-acetyl-cysteine; mPTP: mitochondrial permeability transition pore; Bcl2: B-cell lymphoma-2; HNSCC: head and neck squamous cell carcinoma; SIRT1: Sir2; FOXO3: Forkhead box O3; OS: overall survival; RFS: recurrence-free survival; and NFR2: nuclear factor erythroid-2-related factor.
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Authors’ Contributions

WLC, KYC, KYL, PHF and SMW contributed to writing the manuscript and revising it critically for important intellectual content. All authors read and approved the manuscript.

Ethics Approval and Consent to Participate

Not applicable.

Competing Interests

The authors have declared that no competing interest exists.

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