Brown adipose tissue is associated with healthier body fat distribution and metabolic benefits independent of regional adiposity

Graphical abstract

Highlights

- Brown adipose tissue is associated with more subcutaneous and less visceral fat
- Brown adipose tissue is associated with health independent of fat distribution
- Brown adipose tissue is associated with less liver fat and type 2 diabetes
- Brown adipose tissue is most beneficial in individuals with central obesity

Authors

Andreas G. Wibmer, Tobias Becher, Mahmoud Eljalby, ..., Roger Vaughan, Heiko Schöder, Paul Cohen

Correspondence

schoderh@mskcc.org (H.S.), pcohen@rockefeller.edu (P.C.)

In brief

Wibmer et al. report that brown adipose tissue is associated with a healthier body fat distribution and less central obesity and also associated independently with surrogate parameters of metabolic health and decreased liver fat accumulation. These observations are most prominent in individuals with central obesity.

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Brown adipose tissue is associated with healthier body fat distribution and metabolic benefits independent of regional adiposity

Andreas G. Wibmer,1,7 Tobias Becher,2,3,4,7 Mahmoud Eljalby,2,6 Audrey Crane,2 Pamela Causa Andrieu,1 Caroline S. Jiang,5 Roger Vaughan,5,6 Heiko Schöder,1,6,8* and Paul Cohen2,8,9,*

1Department of Radiology, Memorial Sloan Kettering Cancer Center, New York, NY, USA
2Laboratory of Molecular Metabolism, The Rockefeller University, New York, NY, USA
3DZHK (German Centre for Cardiovascular Research), Partner Site Heidelberg/Mannheim, Mannheim, Germany
4First Department of Medicine (Division of Cardiology), University Medical Center Mannheim, Mannheim, Germany
5Center for Clinical and Translational Science, The Rockefeller University, New York, NY, USA
6Weill Cornell Medicine, New York, NY, USA
7These authors contributed equally
8These authors contributed equally
9Lead contact

*Correspondence: schoderh@mskcc.org (H.S.), pcohen@rockefeller.edu (P.C.)

SUMMARY

The association of brown adipose tissue (BAT) and body fat distribution and their combined effects on metabolic health in humans remains unknown. Here, we retrospectively identify individuals with and without BAT on 18F-fluorodeoxyglucose (18F-FDG) positron emission tomography (PET)/computed tomography (CT) and assemble a propensity score-matched study cohort to compare body fat distribution and determine its role in mediating the benefits of brown fat. We find that BAT is associated with lower amounts of visceral adipose tissue and higher amounts of subcutaneous adipose tissue, resulting in less central obesity. In addition, BAT is independently associated with lower blood glucose and white blood cell count, improved lipids, lower prevalence of type 2 diabetes mellitus, and decreased liver fat accumulation. These observations are most prominent in individuals with central obesity. Our results support a role of BAT in protection from visceral adiposity and improved metabolic health.

INTRODUCTION

Obesity affects over 40% of American adults1 and is expected to reach a prevalence of 48.9% by 2030.2 Moreover, over 1 billion adults worldwide are predicted to be obese by 2030. These trends are a major public health concern because obesity and its sequelae contribute to increased morbidity, mortality, and healthcare costs.3

White adipose tissue (WAT), found in subcutaneous and visceral depots, stores excess triglycerides and accounts for increased body weight in states of positive energy balance.4 Although subcutaneous WAT (SAT) is thought to be relatively benign5,6 or even beneficial,7 visceral WAT (VAT) has been implicated in metabolic dysfunction and insulin resistance.8 Even more so than excess total body weight, visceral adiposity confers increased risk of coronary heart disease, type 2 diabetes mellitus (T2DM), certain types of cancer, and overall mortality.9–14

Recently, positron emission tomography (PET) and computed tomography (CT) imaging with 18F-fluorodeoxyglucose (18F-FDG) have been applied widely to identify brown adipose tissue (BAT) and quantify its activity in human adults.15–18 BAT is characterized by expression of uncoupling protein 1 (UCP1), which is located in the inner mitochondrial membrane and facilitates a futile cycle, resulting in energy dissipation in the form of heat.19–21 This exothermic process is fueled by increased glucose and fatty acid metabolism, suggesting that activation of BAT may result in decreased storage of calories in WAT and might therefore mitigate obesity and its harmful sequelae.22–24 Individuals with BAT exhibit improved measures of glucose and lipid metabolism and a lower prevalence of cardiometabolic disease.25–28 However, it still remains unclear how BAT mediates these beneficial effects.

Previous studies found an association between BAT and body fat distribution,29–32 suggesting the possibility that healthier WAT distribution may mediate improved cardiometabolic outcomes in individuals with BAT. However, interpretation of these data is challenging because of small sample sizes and comparisons with unmatched controls, limiting the ability to make conclusions about whether BAT is linked to white fat distribution. It is also possible that BAT may improve metabolic outcomes independent of body fat distribution. In this study, we addressed this question in a large, propensity score-matched human cohort. Our findings suggest a more salutary
pattern of body fat distribution in individuals with BAT as well as improved metabolic outcomes independent of effects on body fat distribution.

RESULTS AND DISCUSSION

Study cohort selection

We previously showed that BAT is associated with beneficial cardiometabolic effects,25 however, the extent to which these effects might depend on WAT distribution is unknown. To assess the relationship between BAT and WAT distribution, we examined all 19,019 18F-FDG PET/CT scans performed at Memorial Sloan Kettering Cancer Center (MSKCC) in 2016 (Figure 1). These scans were performed to determine cancer diagnosis and stage as well as to monitor treatment response. It is the protocol at MSKCC to comment on BAT status in each study, and we have previously validated the accuracy of this reporting.25 For individuals with BAT (BAT+ group), the first study revealing the presence of BAT was selected as the index scan. If an individual did not have any detectable BAT on any of their scans (BAT/C0 group), then the earliest scan was designated the index scan.25 Only the index scan was used for further analysis, excluding a total of 7,238 studies that represented multiple scans of the same individual (Figure 1). An additional 23 scans did not meet the Brown Adipose Reporting Criteria in Imaging Studies (BARCIST 1.0) criteria for BAT activity (standardized uptake value [SUV] > 1.5 g/mL)33 and were excluded (Figure 1).

Of the remaining 11,758 individuals who met inclusion and exclusion criteria, 10,886 (92.6%) were BAT/C0, and 872 (7.4%) were BAT+ (Figure 1). To minimize baseline differences between BAT− and BAT+ individuals, we applied 1:1 propensity score matching using age, gender, body mass index (BMI), and ambient temperature in the month of the scan as matching variables to derive a study cohort for analysis (Figure S1). Each of these variables has been shown previously to be associated with brown fat prevalence and activity.25,34,35 We then measured body fat distribution in the propensity score-matched study cohort. For this purpose, the CT scan that was obtained as part of 18F-FDG PET/CT imaging was used to determine SAT and VAT distribution at the L3/L4 lumbar level. Analysis of the CT scans revealed that 42 subjects in the propensity score-matched study cohort had scans with significant technical artifacts (e.g., ascites or soft tissue edema) that interfered with quantification of abdominal WAT, resulting in their exclusion and leaving a final cohort of 856 BAT+ and 846 BAT− individuals for analysis (Figure 1).

Cohort characteristics and body fat distribution

After propensity score matching, there were no significant differences between BAT− and BAT+ individuals in age (49.5 years versus 49.0 years, respectively; p = 0.4527), gender (77.2% versus 76.6% females, respectively; p = 0.8319), BMI (24.7 kg/m² versus 24.9 kg/m², respectively; p = 0.6204), or ambient temperature in the month of the scan (12.5°C versus 12.5°C, respectively; p = 0.9867) (Table 1).

On the same PET/CT scans that were examined for BAT status, we quantified waist circumference and SAT and VAT areas and determined the SAT:VAT area ratio as a measure of central adiposity (Figure 2A). To validate our measurements against the commonly used girth measurement,36,37 we assessed available physical girth measurements against the CT-derived waist circumference and observed a strong and significant correlation between the two metrics (r = 0.8257, p < 0.0001) (Figure S2).
the SAT:VAT ratio \( r = -0.0241, p = 0.3212 \) (Figures 2B and 2C). The distribution of the SAT:VAT ratio was right-skewed (Figure 2D), as described previously.\(^38,39\) We observed statistically significant but diminished correlations between VAT area and blood glucose \( r = 0.2435, p < 0.0001 \), HDL \( r = 0.2402, p = 0.0005 \), triglycerides \( r = 0.2799, p < 0.0001 \), and WBC count \( r = 0.1795, p < 0.0001 \) but not with total cholesterol or LDL levels (Table S1).\(^40\) In contrast, SAT area was correlated significantly but weakly with blood glucose \( r = 0.07041, p = 0.0039 \) and WBC count \( r = 0.0738, p = 0.0028 \) but not with total cholesterol, triglycerides, LDL, or HDL levels (Table S1). To adjust for known confounders that may underpin these correlations, we used a multivariable linear regression model adjusted for SAT and VAT area in addition to age, gender, and BMI. Consistent with the literature, VAT area was identified as an independent positive predictor of blood glucose, triglycerides, and WBC count and negative predictor of HDL (Table S2), whereas SAT area was an independent negative predictor of blood glucose only (Table S2).\(^40\) Our findings confirm previous reports that VAT, much more than SAT, is the main driver of comorbid metabolic disease.\(^6,8\)

**BAT is associated with a reduction in central adiposity**

We then investigated whether BAT status is associated with body fat distribution. Although we observed no difference between the matched BAT− and BAT+ groups in BMI or waist circumference (86.9 versus 87.7 cm, respectively; \( p = 0.6740 \)), BAT+ status was associated with a significantly decreased VAT area (74.3 versus 68.2 cm\(^2\), \( p = 0.0307 \)) and increased SAT area (176.0 versus 188.0 cm\(^2\), \( p = 0.0036 \)) and, thus, a higher

### Table 1. Characteristics of individuals with and without brown fat on \(^{18}\)F-FDG PET/CT after propensity score matching

| Parameter | BAT (BAT−) \( n = 846 \) | BAT (BAT+) \( n = 856 \) | Value |
|-----------|--------------------------|--------------------------|-------|
| **Clinical characteristics** | | | |
| Age, mean (SD) | 49.5 (15.3) | 49.0 (15.8) | 0.4527 |
| Gender, no. (%) | | | 0.8319 |
| Female | 653 (77.2) | 656 (76.6) | 0.6204 |
| Male | 193 (22.8) | 200 (23.4) | |
| BMI, median (IQR) | 24.7 (21.7–28.4) | 24.9 (22.0–28.0) | |
| Ambient temperature in °C in month of scan, mean (SD) | 12.5 (6.4) | 12.5 (6.4) | 0.9867 |
| **Race, no. (%)** | | | 0.0240 |
| Asian | 88 (10.4) | 58 (6.8) | 0.6204 |
| African American | 78 (9.2) | 99 (11.6) | |
| Caucasian | 596 (70.4) | 603 (70.4) | |
| Other or unknown | 84 (9.9) | 96 (11.2) | |
| **Ethnicity, no. (%)** | | | 0.0511 |
| Hispanic | 70 (8.3) | 89 (10.4) | |
| Non-Hispanic | 769 (90.9) | 751 (87.7) | |
| Unknown | 7 (0.8) | 16 (1.9) | |
| **Fat distribution, median (IQR)** | | | 0.0511 |
| Waist circumference on CT, cm | 86.9 (78.6–96.3) | 87.7 (79.3–96.2) | 0.6740 |
| VAT, area cm\(^2\) | 74.3 (39.5–128.8) | 68.2 (39.5–111.0) | 0.0307 |
| SAT, area cm\(^2\) | 176.0 (113.3–257.0) | 188.0 (127.0–272.3) | 0.0036 |
| SAT/VAT ratio | 2.4 (1.5–3.4) | 2.7 (1.9–4.1) | <0.0001 |
| **Laboratory values, median (IQR)** | | | |
| Glucose, mg/dL | 90.0 (82.3–97.8) | 87.0 (80.0–96.0) | <0.0001 |
| LDL, mg/dL | 108.5 (80.0–142.5) | 110.0 (84.5–137.5) | 0.7605 |
| HDL, mg/dL | 47.0 (34.0–63.0) | 53.5 (43.0–73.0) | 0.004 |
| Cholesterol, mg/dL | 192.0 (152.0–223.5) | 196.0 (160.0–220.0) | 0.5081 |
| Triglycerides, mg/dL | 129.0 (87.0–177.5) | 98.0 (75.0–136.0) | 0.0027 |
| WBC count, 10\(^3\) cells/L | 6.7 (5.3–8.2) | 5.8 (4.4–7.5) | <0.0001 |
| **Metabolic outcomes** | | | |
| Liver density, HU, median (IQR) | 58.6 (53.4–62.9) | 60.3 (55.7–64.3) | <0.0001 |
| T2DM, no. (%) | 88 (10.4) | 52 (6.1) | 0.0012 |

BAT, brown adipose tissue; BMI, body mass index; HU, Hounsfield unit; SAT, subcutaneous adipose tissue; SD, standard deviation; IQR, interquartile range, VAT, visceral adipose tissue.
SAT:VAT ratio (2.4 versus 2.7, p < 0.0001) (Figures 3A–3C; Table 1). These BAT-associated differences in WAT distribution were noted even independent of age, gender, and BMI (β-estimate −10.42, 95% confidence interval [CI] −14.60 to −6.25, p < 0.0001) for VAT; β-estimate 14.18, 95% CI 8.99–19.38, p < 0.0001 for SAT; Table S3) and were most pronounced at higher BMI values (β-estimate 0.03, 95% CI 0.00–0.06, p = 0.0210; Figures 3D and 3E).

Previous work reported that, in healthy individuals subjected to cold exposure before 18F-FDG PET/CT, BAT activity was inversely correlated with visceral and, to a lesser extent, subcutaneous adiposity.32 To assess whether the quantity or glucose uptake activity of BAT is associated with body fat distribution, we examined whether the number or FDG uptake intensity of BAT depots on PET/CT were correlated with WAT distribution. Interestingly, the number of active BAT depots was weakly inversely correlated with VAT area (r = −0.1895, p < 0.0001) and positively correlated with SAT:VAT ratio (r = 0.2149, p < 0.0001) but not significantly correlated with SAT area (r = −0.0205, p = 0.4999) (Figures 3F–3H). Peak BAT activity was negatively predictive of VAT area (β-estimate −2.78, 95% CI −4.02 to −1.55, p < 0.0001) and positively predictive of SAT:VAT ratio (β-estimate 0.14, 95% CI 0.10–0.18, p < 0.0001) but not of SAT area (β-estimate 2.22, 95% CI −0.16 to 4.61, p = 0.0677; Figures 3I and 3J). Our results thus support previous observations regarding associations between BAT activity and body fat distribution. Our work illustrates that, even without previous cold exposure, BAT activity correlates inversely with visceral

Figure 2. Illustration of 18F-FDG PET/CT measurements and WAT distribution
(A) Schematic of WAT quantification on transverse CT slides at the L3/L4 level and detection and quantification of BAT by 18F-FDG PET.
(B) Scatterplot depicting the correlation between BMI and VAT and SAT areas. Solid lines represent fitted curves derived from second-order polynomial equations, and shaded areas are 95% CIs. Correlation was assessed by calculating Spearman’s rank correlation coefficient. Fitted curves are shown up to a BMI of 50 kg/m², and individual data points are depicted for 1,701 individuals.
(C) Scatterplot depicting the correlation between BMI and SAT:VAT ratio. The solid line represents the fitted curve derived from second-order polynomial equations, and shaded areas are 95% CIs. Correlation was assessed by calculating Spearman’s rank correlation coefficient. The fitted curve is shown up to a BMI of 50 kg/m², and individual data points are depicted for 1,701 individuals.
(D) Scatterplot depicting the relative distribution of SAT:VAT ratio in our cohort. The solid blue line represents the fitted curve derived from second-order polynomial equations. The fitted curve is shown up to a SAT:VAT ratio of 7.8. For (C) and (D), fitted curves were derived from measurements of 1,702 individuals.
adiposity. The discrepancy between our data and the data reported by Saito et al.32 regarding the correlation of BAT activity and SAT area may be related to the differences in the study population and between cold-induced and retrospective identification of BAT on18F FDG PET/CT.

Prior studies have shown that age is correlated with decreased prevalence and activity of BAT34,35 and increased quantity of VAT.41–45 We thus used two multivariable analysis models to test whether SAT and VAT areas (model 1) or SAT:VAT ratio (model 2) can predict the number of BAT depots or BAT activity. These models were adjusted for age, gender, and BMI. We found no independent association between SAT, VAT, or SAT:VAT ratio and the number of BAT depots (Table S4). However, BAT activity was weakly negatively associated with VAT area (β-estimate −0.09, 95% CI −0.14 to −0.04, p = 0.0004) and positively associated with SAT:VAT ratio (β-estimate 0.25, 95% CI 0.13–0.36).

Figure 3. BAT status and activity are associated with body fat distribution
(A) Distribution of BMI by BAT status. Lines represent the 10th percentile, median, and 90th percentile. The p value was calculated based on a Mann-Whitney U test.
(B) Distribution of waist circumference on CT by BAT status. Lines represent the 10th percentile, median, and 90th percentile. The p value was calculated based on a Mann-Whitney U test.
(C) Distribution of SAT:VAT ratio by BAT status. Lines represent the 10th percentile, median, and 90th percentile. The p value was calculated based on a Mann-Whitney U test. The depicted data show measurements for 1,662 individuals.
(D) Best fit curves representing the relationship between BMI and SAT area (solid lines) or VAT area (dashed lines), stratified by BAT status, were calculated using linear regression analysis. The shaded areas represent the 95% CIs. The p values were calculated for assessing statistically significant differences between the slopes of these two lines. Best fit curves are shown up to a BMI of 40 kg/m².
(E) Best fit curves representing the relationship between BMI and SAT:VAT ratio, stratified by BAT status, were calculated using linear regression analysis. The shaded areas represent the 95% CIs. β-Estimates and p values correspond to the linear regression model. Best fit curves are shown up to a BMI of 40 kg/m².
(F) Boxplot of the total VAT area against the number of active BAT depots. Boxes represent the 25th percentile, median, and 75th percentile, and whiskers illustrate the 10th and 90th percentiles. Correlations were assessed by calculating Spearman’s rank correlation coefficient.
(G) Boxplot of the total SAT area against the number of active BAT depots. Boxes represent the 25th percentile, median, and 75th percentile, and whiskers illustrate the 10th and 90th percentiles. Correlations were assessed by calculating Spearman’s rank correlation coefficient.
(H) Boxplot of the SAT:VAT ratio against the number of active BAT depots. Boxes represent the 25th percentile, median, and 75th percentile, and whiskers illustrate the 10th and 90th percentiles. Correlations were assessed by calculating Spearman’s rank correlation coefficient.
(I) Best fit curves of SAT and VAT areas as a function of BAT activity on PET/CT were calculated using linear regression analysis. The shaded areas represent the 95% CIs. β-Estimates and p values correspond to the linear regression model. Data are shown up to a BAT activity of 25 SUVs.
(J) Best fit curve of SAT:VAT ratio as a function of BAT activity on PET/CT was calculated using linear regression analysis. The shaded areas represent the 95% CIs. β-Estimate and p value correspond to the linear regression model. Data are shown up to a BAT activity of 25 SUVs.

For (A) and (B), data are shown for measurements of 1,702 individuals. For (D) and (E), fitted curves were derived from measurements of 1,702 individuals. For (F)–(H), data are shown for measurements of 856 patients. For (I) and (J), fitted curves are derived from measurements of 856 individuals.
p < 0.0001) (Table S4). A trend toward a positive association between BAT activity and SAT area (β-estimate 0.04, 95% CI 0.00–0.08, p = 0.0640) was also noted (Table S4). These data suggest that BAT may mitigate central adiposity by decreasing VAT and increasing SAT in a dose-dependent manner.

BAT is associated with an improved metabolic profile, specifically in individuals with central obesity

We have previously shown that BAT is associated with improved cardiometabolic health.25 Here we show that BAT is associated with improved body fat distribution, offering one possible explanation for our previous findings. To better characterize the extent to which BAT and WAT distribution conjointly affect metabolic function, we first examined the effect of these fat depots on laboratory metrics of metabolic health. We found that VAT was associated with higher blood glucose, WBC count, and triglycerides and lower HDL levels, whereas SAT was not associated or only weakly associated with these parameters (Figures S3A–S3D). The presence of BAT was associated with lower blood glucose, WBC count, and triglycerides and higher HDL (Figures 4A–4D). When stratified by SAT:VAT ratio, these associations were more pronounced in individuals with a lower SAT:VAT ratio (Figures 4E–4H). Last, the presence of BAT was associated independently with lower blood glucose, WBC count, and triglycerides and higher HDL after adjusting for age, gender, BMI, and SAT:VAT ratio (Figures 4I–4L). These data indicate that the beneficial metabolic effects associated with BAT can be explained partially by healthier body fat distribution and that BAT is also associated with additional benefits independent of body fat distribution.

BAT is associated with decreased liver fat and T2DM

To determine whether the observed associations between fat depots and laboratory metrics of metabolic health are associated with improved disease status, we next assessed liver fat accumulation on CT (Figure 5A). Lower liver parenchymal density on unenhanced CT, as quantified by lower Hounsfield units (HU), is an established surrogate for hepatic lipid accumulation.46 Multivariable analysis showed that increased VAT is associated with lower liver density, indicating more liver fat, whereas SAT was associated with liver density, but this association was lost when adjusted for age, gender, and BMI (Table 2; Figure S3E). A lower SAT:VAT ratio was significantly associated with decreased liver density (β-estimate 0.73, 95% CI 0.49–0.97, p < 0.0001; Figure 5B). These results corroborate prior observations that central obesity is associated with increased liver fat accumulation.47 Additionally, individuals with BAT+ scans exhibited higher liver density compared with BAT− individuals (Figure 5C). This association was most pronounced in individuals with a low SAT:VAT ratio (Figure 5D) and remained significant after adjusting for age, gender, BMI, and SAT or VAT area (Table 2). These findings suggest that BAT is associated with attenuated liver fat accumulation, particularly in individuals with central adiposity, and that this observation is not solely explained by improved body fat distribution.
BAT– (OR 0.59, 95% CI 0.47–0.72, p < 0.0001) or BAT+ (OR 0.82, 95% CI 0.67–0.97, p = 0.0288) individuals (Figures 5F, 5G, and S3F). The effect of BAT on this association was greatest at lower SAT:VAT ratios (Figure 5G). These data suggest that BAT has its most potent metabolic effects at unfavorably low SAT:VAT ratios (indicating a relative increase of VAT area compared with SAT area).

Last, we used a multivariable logistic regression model to adjust for the potential interaction between BAT, SAT, and VAT areas (in addition to other potential confounders, such as age, gender, and BMI) when determining the association between these factors and T2DM. Our results showed that BAT is associated with lower prevalence of T2DM (OR 0.60, 95% CI 0.41–0.86, p = 0.0066), whereas age (OR 1.04, 95% CI 1.03–1.06, p < 0.0001) and VAT area (OR 1.04, 95% CI 1.01–1.08, p = 0.0119) were associated with an increased OR of T2DM prevalence (Table 2). These results indicate that BAT has an effect on metabolic health independent of its association with a beneficial body fat distribution.

Conclusions
In conclusion, our data reveal that BAT is associated with decreased central adiposity, as indicated by a decrease in VAT and an increase in SAT and SAT:VAT ratio. This change in body fat distribution is associated with improved metabolic health\(^6,7,14\) and may hence contribute to the metabolic benefits associated previously with BAT.\(^25,48\) In addition, increased BAT activity and number of depots are associated with improvements in metabolic parameters, suggesting that not only the quantity but also the function of BAT may be of importance for WAT distribution.

Importantly, we observe that BAT retains its association with improved beneficial effects on blood glucose, LDL and HDL levels, liver fat accumulation, and T2DM independent of WAT distribution. Furthermore, these observations seem to be most pronounced in individuals with central adiposity. Although energy expenditure at rest has been linked previously to changes in body fat distribution,\(^49,50\) the association of BAT with metabolic health independent of WAT distribution suggests that additional mechanisms may underlie the observed findings. As an example, potential endocrine factors secreted by BAT have been implicated as a mechanism that may explain how BAT contributes to metabolic health.\(^51\) An intriguing association is that of BAT status and liver fat accumulation. Although a number of studies in rodents have linked BAT function to protection from hepatic steatosis,\(^52–54\) this connection is less well established in humans.\(^48\) The underlying mechanisms may include modulation of circulating fatty acid levels through BAT,\(^55\) resulting in decreased lipotoxicity; BAT-secreted factors, such as neuregulin 4, that directly modulate hepatic metabolism;\(^52\) or endocrine...
interactions between BAT and other organs that indirectly influence hepatic lipid storage.

Finally, our findings suggest that BAT status and increased BAT activity in humans may ameliorate the severity of metabolic disease associated with obesity, particularly visceral adiposity. Our observation that not only the presence but also the activity of BAT are associated with decreased visceral adiposity supports a relationship between BAT and WAT distribution. Further research into the mechanisms underlying the observed associations between BAT and WAT distribution as well as a deeper understanding of how BAT modulates metabolic outcomes is necessary if we are to harness the therapeutic potential of this tissue to treat obesity and comorbid diseases.1,2

Limitations of study
This is a retrospective, observational study that reports associations. The study design does not allow one to make conclusions about cause-effect relationships between BAT and body fat distribution or metabolic health. Individuals in our study received 18F-FDG PET/CT scans for cancer-associated diagnoses, treatment, or surveillance. Thus, it cannot be excluded that the underlying cancer type, stage, and treatment may have confounded our results. Furthermore, individuals were not cold stimulated prior to their 18F-FDG PET/CT scans, resulting in sub-maximal BAT activation and, subsequently, detection. Impaired systemic insulin sensitivity may also reduce 18F-FDG uptake in BAT and, thus, its detection on PET/CT scans. Moreover, body fat distribution was derived from one CT slice only, and this may not fully capture body fat distribution characteristics. Last, unenhanced CTs and laboratory parameters were only available for subgroups of the study cohort, which might have introduced bias (Table S5).

STAR METHODS

Detailed methods are provided in the online version of this paper and include the following:

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SUPPLEMENTAL INFORMATION

Supplemental information can be found online at https://doi.org/10.1016/j.xcrm.2021.100332.

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AUTHOR CONTRIBUTIONS

P.C., H.S., A.G.W., and T.B. designed and conceived the study. A.G.W., T.B., and P.C.A. acquired the data. A.G.W., T.B., M.E., A.C., C.S.J., R.V., H.S., and P.C. analyzed and interpreted the data. T.B., A.G.W., M.E., A.C., H.S., and P.C. wrote the manuscript with input from all authors.

DECLARATION OF INTERESTS

T.B. is currently an employee of Roche Diagnostics, Rotkreuz, Switzerland.

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STAR METHODS

KEY RESOURCES TABLE

| REAGENT or RESOURCE | SOURCE | IDENTIFIER |
|---------------------|--------|------------|
| Software and algorithms |        |            |
| Prism (version 9.0.0) | GraphPad Software | https://www.graphpad.com/scientific-software/prism/ |
| SAS (version 9.4) | SAS Institute | https://www.sas.com/en_us/software/viya.html |
| Adobe Illustrator CC (2019) | Adobe | https://www.adobe.com/ |
| Aquarius iNtuition (version 4.4.13) | TeraRecon | https://www.terarecon.com/ |

RESOURCE AVAILABILITY

Lead contact
Requests for additional information, resources and reagents should be directed to the Lead Contact, Paul Cohen (pcohen@rockefeller.edu).

Materials availability
No new reagents or materials were generated in this study.

Data and code availability
Anonymized clinical data related to this study are available upon request. All requests are subject to an internal review by T.B., P.C., A.G.W. and H.S. and completion of a data sharing agreement, in accordance with the Rockefeller University and Memorial Sloan Kettering Cancer Center Institutional Review Board and Institutional Guidelines.

METHOD DETAILS

Study design, data collection and patient selection
The current study was designed as a retrospective, single-center cross-sectional case/control study conducted at Memorial Sloan Kettering Cancer Center (MSKCC). The requirement for informed consent was waived due to its retrospective nature, and the study followed institutional guidelines and was approved by the Institutional Review Boards of The Rockefeller University and MSKCC.

Demographic and clinical characteristics, laboratory tests, and diagnosis codes based on the International Classification of Disease (9th and 10th editions) were extracted from electronic medical records using a standardized data collection method, as previously described. The laboratory tests included blood glucose measured at the day of the 18F-FDG-PET/CT after a fasting interval of at least 4-6 hours, white blood cell count (obtained if measured within three months of the PET/CT) and lipid profile (obtained if measured within one year of the PET/CT). ICD codes were used to categorize relevant diseases as previously described.

A propensity-score matched cohort was established as described below. Of these 1,702 individuals, lab data were available as follows: blood glucose levels were available for 1,683 (98.88%) individuals, LDL and HDL levels were available for 209 (12.28%) individuals, and total cholesterol for 229 (13.45%) individuals (Table S5). White blood cells (WBC) count was available for 1,645 (96.65%) individuals overall and for 1,221 (96.29%) of 1,268 individuals without a hematologic malignancy (Table S5). Girth measurements were available for 74 individuals (4.35%) (Table S5). Liver density measurements were performed for 1,298 (76.26%) individuals and omitted on 350 (20.56%) contrast-enhanced CT scans and 54 (3.17%) scans due to imaging artifacts or prior major liver resections (Table S5).

18F-FDG-PET/CT examinations
18F-FDG-PET/CT examinations were performed on a hybrid PET/CT scanner (Discovery series; GE Healthcare, Milwaukee, WI) after intravenous injection of approximately 444 MBq of 18F-FDG. The radiotracer was obtained from a commercial source (IBA Molecular, Reston, VA) and calibrated by the MSKCC in-house radiopharmacy. 18F-FDG was administered after an approximately 6-hour-long fasting period, and the time interval between tracer injection and imaging was approximately 60 minutes. For anatomic correlation and attenuation correction, free-breathing low-dose CT was performed with a tube voltage of 120–140 kV, a tube current of 80 mA, a pitch factor of 0.75-1.75, and a slice thickness of 3.75 mm.
Standardized uptake value (SUV) was used to quantify FDG uptake of BAT as a surrogate of its glucose uptake activity and was calculated as the ratio of a voxel’s radioactivity concentration [kBq/mL] and the decay-corrected and body weight adjusted injected radiotracer activity [kBq/g] and reported as g/mL.

**Identification of BAT and measurement of BAT activity**

Identification of BAT and measurement of BAT activity was performed as previously described. Briefly, 6 adipose regions of interest (ROIs) — namely cervical, supraclavicular, axillary, mediastinal, paraspinal and abdominal — were defined based on anatomical landmarks and CT characteristic of adipose tissue. An SUV threshold of ≥ 1.5 g/mL, normalized to body mass, was used to identify BAT. BAT activity was reported as peak SUV value, measured as the average SUV within a 1 cm³ sphere centered on the voxel with the highest ¹⁸F-FDG uptake in each of the six adipose depots.

**Measurement of body fat distribution**

Image analyses were performed using commercially available software (Aquarius iNtuition version 4.4.13.; TeraRecon; Foster City, CA). Areas of VAT and SAT (CT density range: –195 to –45 HU) as well as the outer body circumference were semi-automatically measured in the transverse plane between the vertebral bodies L3/4. Two-dimensional measurements at this level were previously shown to be most strongly correlated with abdominal fat volumes and associated with cardiometabolic risk factors in a cohort of the Framingham Heart Study.

**Assessment of liver density on CT**

For liver density measurements, circular regions of interest (approximate area: 5 cm²) were placed at the level of the porta hepatis in the right posterior, right anterior, and left lateral hepatic segments, respectively (Figure 5A). Lower liver density on unenhanced CT, as quantified by HU, is a reliable surrogate of hepatic fat content. Liver density was not measured on contrast-enhanced scans (n = 350) or cases with imaging artifacts or prior major liver resections (n = 54).

**QUANTIFICATION AND STATISTICAL ANALYSIS**

Continuous variables with non-normal distributions were expressed as median [IQR]. Categorical variables were expressed as number and percentage (%). Comparisons between groups were performed with Mann-Whitney U test for nonparametric variables and Fisher’s exact test or chi-square test for categorical variables. Correlations between BMI and VAT or SAT area were assessed using Spearman’s rank correlation coefficient. Non-linear regression curves were derived from second order polynomial curve fitting. Propensity scores were estimated using a non-parsimonious multivariable logistic-regression model with brown adipose tissue status (presence or absence) as the dependent variable and age, gender, BMI, and temperature at time of scan as covariates. Matching was performed using a greedy nearest neighbor matching algorithm without replacement with a caliper width set to 0.2 and a matching ratio of 1:1. Standardized difference means were used to assess balance before and after matching (Figure S1). Matching was conducted in SAS (version 9.4, SAS Institute, Cary, NC) using the `psmatch` function. All WBC count analyses were done using values from individuals without hematologic malignancies. A two-sided p value of less than 0.05 was used to define statistical significance for all tests. Data were analyzed in Prism (version 8.4.3, GraphPad Software, La Jolla, CA, USA) and SAS (version 9.4, SAS Institute, Cary, NC, USA).