Knockdown circTRIM28 enhances tamoxifen sensitivity via the miR-409-3p/HMGA2 axis in breast cancer

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Abstract

**Background:** Tamoxifen (TAM) is a frequently-used treatment for breast cancer (BC). But the TAM resistance seriously affects the patient therapeutic effect. Previous research indicated that circular RNAs (circRNAs) might participate in the regulatory processes of BC. Here, we discovered the parts of circular RNA tripartite motif-containing 28 (circTRIM28) in BC.

**Methods:** CircTRIM28, microRNA-409-3p (miR-409-3p), and high mobility group AT-hook 2 (HMGA2) levels were perceived by qRT-PCR and western blot. Moreover, the biological functions of the cells were examined. Furthermore, dual-luciferase report was employed to reconnoiter the targeted relationship between miR-409-3p and circTRIM28 or HMGA2.

**Results:** CircTRIM28 and HMGA2 were augmented, and the miR-409-3p was repressed in BC. Silencing circTRIM28 enhanced tamoxifen sensitivity and cell apoptosis, whereas hampered cell development in BC cells. In mechanism, circTRIM28 could sponge miR-409-3p to increase HMGA2. In addition, silencing circTRIM28 impeded tumor growth.

**Conclusion:** CircTRIM28 facilitated the BC via miR-409-3p/HMGA2.

Highlights

1. The expression of circTRIM28 was upregulated in BC.
2. Silencing circTRIM28 enhanced tamoxifen sensitivity and cell apoptosis, whereas curbed cell proliferation, cell migration and invasion in BC cells.
3. CircTRIM28 expedited BC by targeting miR-409-3p.
4. MIR-409-3p repressed BC by retarding HMGA2.

**Keywords:** Breast cancer, circTRIM28, miR-409-3p, HMGA2, Tamoxifen

Introduction

Breast cancer (BC) is common cancer worldwide. Hormone receptor (HR) positive patients accounted for three-quarters of all BC patients [1]. In the current clinical treatment, tamoxifen (TAM) is an important treatment for patients with HR-positive, especially for premenopausal patients [2, 3]. However, patients who develop resistance to TAM can make treatment difficult [4, 5]. Therefore, we urgently need to determine the drug resistance mechanism of TAM.

Circular RNAs (circRNAs) are a kind of non-coding RNAs that employ a key role in numerous illnesses [6, 7].
Hsa_circ_001783 regulates cell proliferation and invasion in BC [8]. Circ-ABCB10 promotes BC proliferation by regulating miR-1271 [9]. Besides, circ-DNMT1 activates autophagy in BC [10]. In addition, circ_103809 regulates the PI3K/AKT signaling pathway in BC progression [11]. However, the definite monitoring mode of circTRIM28 in BC is not clear.

MicroRNAs (miRNAs) are a class of RNAs that affects the cellulate processes [12, 13]. MiR-409-3p inhibits the evolution of tongue squamous cell carcinoma [14]. Besides, miR-409-3p participates in regulating the cell proliferation of osteosarcoma [15]. Moreover, miR-4732-5p regulates BC progression via TSPAN13 [16]. CircDENND4C regulates glycolysis in BC [17]. All the same, the identification of miR-409-3p in BC was not yet testified. Previous research has reported that the high mobility group AT-hook 2 (HMGA2) is connected with many cancer developments, like differentiation and angiogenesis. HMGA2 has been shown to be an effective carcinogen [18]. Yet, the influence of HMGA2 in BC cells is blurry.

Up to date, increasing attention has been paid to the competing endogenous RNA (ceRNA) mechanism, which proposed that endogenous RNA transcripts with microRNA (miRNA) response elements (MREs) might derepress target mRNAs expression via sponging miRNA [19, 20]. Of note, the mechanism has been considered the major regulation manner for circRNAs [21, 22]. Herein, the online bioinformatics tools exhibited that there were some binding sites between circTRIM28 and miR-409-3p for the first time. Therefore, our purpose is to analyze the function of circTRIM28/miR-409-3p in BC and reconnoiter possible governing network basics on these effects.

### Materials and methods

#### Clinical tissues
The test was approved by Jing Men No. 2 People’s Hospital. Sixty-four BC tissues (BC-S: 30 cases were sensitive to tamoxifen; BC-R: 34 cases were resistant to tamoxifen) and normal tissue ($n = 64$) were collected from Jing Men No. 2 People’s Hospital. All volunteers wrote the informed consent. Besides, the clinicopathological features of these patients were shown in Table 1.

#### Cell lines
The BC cell lines MCF7 and MDA-MB-231 with MCF10A as control. MCF7 cells a human breast cancer cell line with estrogen, progesterone and glucocorticoid receptors. When grown in vitro, the cells are capable of forming domes and the epithelial-like cells grow in monolayers. MDA-MB-231 cells were chosen as an ER negative cell line with invasive phenotype in vitro, and have epithelial-like morphology and appears phenotypically as spindle-shaped cells. All cells were purchased from the American type culture collection (ATCC, Manassas, VA, USA) and used at between three and ten passages. Cells were grown in RPMI 1640 supplemented with 10% fetal bovine serum (FBS) and penicillin/streptomycin at 37°C in a 5% CO2 and 95% humidified atmosphere at a frequency of 2–3 times/week. According to the [https://web.expasy.org/cellosaurus/CVCL_0023](https://web.expasy.org/cellosaurus/CVCL_0023) website, the cells have no cross-infection. MCF7 and MDA-MB-231 cells were treated with 0.1 μM 4-hydroxytamoxifen (4-OH-TAM) (Sigma, St. Louis, MO, USA) for at least 6 months to produce anti-TAM BC cells, named MCF7/R and MDA-MB-231/R. The TAM-resistant phenotype of

### Table 1  Relationship between circTRIM28 expression and clinicopathologic features of BC patients

| Characteristics | circTRIM28 expression | $P$ value |
|-----------------|-----------------------|-----------|
|                | Low ($n = 32$) | High ($n = 32$) |
| Age (years)    |                    |           |
| $\leq 50$      | 28                  | 15        |
| $> 50$         | 36                  | 17        |
| TNM grade      |                    |           |
| I + II         | 29                  | 21        |
| III            | 35                  | 11        |
| Lymph node metastasis | | |
| Positive       | 38                  | 13        |
| Negative       | 26                  | 19        |
| Tumor size     |                    |           |
| $\leq 2$ cm    | 25                  | 20        |
| $> 2$ cm       | 39                  | 12        |

TNM tumor-node-metas-tasis; *$P < 0.05$
BC cells was maintained using the same medium supplemented with 0.5 μM TAM.

**QRT-PCR**
Whole RNAs were utilized by Trizol reagent (Takara, Tokyo, Japan) and reverse transcription as cDNA. Afterward, the cDNA level was quantified by SYBR Green kit (Takara). Successively, the data were assessed by the $2^{-\triangle\triangle C_t}$ method and standardized by U6 or GADPH. Primers were listed in Table 2.

### Western blot
The method described by Hou et al. [23]. The antibodies were listed: anti- HMGA2 (ab207301; 1:1000 dilutions; 0.693 μg/mL; Abcam, Cambridge, MA, USA), anti-PCNA (ab92552; 1:1000 dilutions; 0.164 μg/mL; Abcam), anti-Cleaved-caspase 3 (ab32042; 1:500 dilutions; 3.088 μg/mL; Abcam), anti-MMP9 (ab76003; 1:10,000 dilutions; 0.186 μg/mL; Abcam), and anti-β-actin (ab8227; 1:1000 dilutions; 0.1 μg/mL; Abcam).

### Cell transfection
The sh-circTRIM28 (sh-circTRIM28#1, sh-circTRIM28#2, sh-circTRIM28#3), the control (sh-NC), miR-409-3p mimics, miR-409-3p inhibitors (anti-miR-409-3p) and controls, Bio-NC, Bio-miR-409-3p, Oligo probe, circTRIM28 probe, HMGA2 overexpression (HMGA2) and control plasmid (vector) were also fabricated by Ribobio (Guangzhou, China). Lipofectamine 2000 (Sigma) was employed in transfection.

### RNase R and act D assay
The circ_0047921 and CD226 mRNA were treated with RNase R (Sigma). Similarly, actinomycin D (Act D, 2 mg/mL) or DMSO (Sigma) was added to the medium as control, and the tests were performed respectively. Finally, the expression level of circ_0047921 and CD226 mRNA were uncovered by qRT-PCR.

### CCK8 assay and cytotoxicity assay
After post-transfection, BC cells (2.0 \times 10^3/well) were seeded in 96-well plates. The cells in each well were exposed to different doses of tamoxifen (0, 5, 10, 20, 30 or 40 μM) for 48 h. Next, CCK8 (Sigma) was supplemented and the half-maximal inhibitory concentration (IC$_{50}$) with assessed. A similar model was enforced to measure cell viability.

### Cell proliferation assay
BC cells were seeded into 96-well plates. Then, the EdU Apollo In Vitro Imaging Kit (Sigma) was employed along with the guide. Generally, tumor cells were incubated with 50 μM EdU for 2 h at 37°C and fixed in 4% formaldehyde. After being stained with Apollo reaction cocktail and DAPI (identify the nuclei) for 30 min, the proliferation-positive cells were photographed and counted under a microscope.

### Colony formation assay
In short, 1 × 10^3 BC cells were plated into 6-well plates and maintained for 2 weeks. Finally, the colonies were fixed with methanol for 20 min and dyed with 0.1%
crystal violet (Sigma) at room temperature. Finally, cell colonies were counted and photographed.

**Flow Cytometry assay**

BC cells with transfection were seeded in 6-well plates. The apoptotic was assessed by Annexin V-FITC/PI kit (Sigma) with a flow cytometer. In short, tumor cells were trypsinized and washed three times in PBS, followed by re-suspending in binding buffer. After being dual-stained with 5 μL Annexin V-FITC and 10 μL PI solution in the dark for 10 min, the apoptotic cells were identified according to a flow cytometer.

**Transwell assay**

BC cells were plated into a transwell with a pore polycarbonate membrane (BD Bio-sciences, Bedford, MA, USA). In brief, $4 \times 10^5$ transfected BC cells were planted into the top chamber. Then the lower chamber of the transwell contains 500 μL of DMEM and 10% FBS. Succeeding, the inferior surface of the membrane cells was stained. The same method was enforced to detect the invasion ability, but the transwell chamber was precoated with matrigel (BD Biosciences). Eventually, a light microscope was performed to validate the count of cells.

**Dual-luciferase reporter assay**

The targeted combination relationship of miR-409-3p and circTRIM28 or HMGA2 was predicted by circbank (http://www.circbank.cn/), starbase (http://starbase.sysu.edu.cn), and circinteractome (https://circinteractome.nia.nih.gov). Then, the circTRIM28 and HMGA2 wild and mutant were synthesized by Ribobio (circTRIM28-WT, HMGA2–3’UTR-WT or circTRIM28-MUT, HMGA2–3’UTR-MUT). Finally, luciferase activity was quantified.

**RNA pull-down**

Bio-miR-409-3p and Bio-NC were synthesized from Ribobio. The RNA pull-down assay was applied as beforehand reported [24]. Finally, circTRIM28 and miR-409-3p contents were assessed.

**RIP assay**

The Magna RNA immunoprecipitation kit (Millipore, Billerica, MA, United States) was enforced to carry out RIP assay in accordance with the guide. To be brief, tumor cells at 80% confluency were harvested, followed by lysing in complete RIP lysis buffer. Then, the obtained cell lysates were incubated with anti-AGO2 or anti-IgG for 4 h at 4°C before treating magnetic protein A/G beads for 2h. Subsequently, the beads were washed, and the extracted total RNA was subjected to qRT-PCR analysis.

**In vivo assay**

All-female BALB/C nude mice (6-week-old, 18–22 g) were bought from Shanghai Laboratory Animal Company (SLAC, Shanghai, China) under a pathogen-free environment at a temperature of 25°C and relative air humidity between 45 and 50% with a 12/12 h day/night cycle. The in vivo work was approved by the guidance of the Animal Care and Use Committee of Jing Men No. 2 People’s Hospital. MDA-MB-231/R cells ($1 \times 10^6$) with or without tamoxifen (5 mg/kg) transfected with the circTRIM28 knockdown vector (sh-circTRIM28) or sh-NC were vaccinated into mice ($n=6$ group) under a specific-pathogen-free environment. The volume ($mm^3$) = length × width$^2$/2. Mice were sacrificed on day 35, the tumors were used for deeper level research.

**IHC assay**

In short, formalin-fixed and paraffin-embedded mice tumor tissue specimens were cut into 4 μm sections, followed by staining with hematoxylin and eosin for histopathology under a microscope. To analyze cell proliferation, the Ki67 (ab92742; 1:1000 dilutions; Abcam) contents in the tumor were detected by IHC assay. The unambiguous assessment scheme as per the description of Ma et al. [25]. Ultimately, the slides were photographed.

**Statistical assay**

These statistics were assembled from no less than 3 groups of repeats. The Student's $t$-test or ANOVA was enforced to measure the difference in GraphPad Prism 7. P-value < 0.05 was significant.

**Results**

The circTRIM28 was augmented in BC

Hence, to further explore the role of circTRIM28 in BC, qRT-PCR was applied. CircTRIM28 was amplified in BC tissues. Besides, the expression of circTRIM28 was greatly increased in tamoxifen-resistance tissues (BC-R, $n=34$) relative to tamoxifen-sensitive tissues (BC-S, $n=30$) (Fig. 1A). Furthermore, our information proposed that circTRIM28 was augmented in BC cell lines (MCF7 and MDA-MB-231) than in the control cell line (MCF10A). In addition, circTRIM28 was significantly upregulated in tamoxifen-resistant BC cell lines (MCF7/R and MDA-MB-231/R) in comparison to the parental cell lines (MCF7 and MDA-MB-231) (Fig. 1B and C). Meanwhile, patients with higher circTRIM28 expression levels had significantly lower post-operative survival compared to those with lower circTRIM28 expression levels (Fig. 1D). RT-PCR usually uses two pairs of primers to exclude gene recombination, one pair of divergent primers to detect circRNA and recombinant genes, and the other pair of
convergent primers to detect mRNA and circRNA [26]. Figure 1E showed that cDNA and genomic DNA (gDNA) were used as templates for detection. When cDNA was used as the template, both pairs of primers had products; when gDNA was used as the template, only convergence primers had products, indicating that the detected RNA was not produced by gene recombination and the structure pattern was circular RNA. Meanwhile, the GAPDH structure pattern was linear RNA. To further confirm the structure of circTRIM28, the RNase R enzyme and Act D treatment assay were applied to detect the structure of circTRIM28. As shown in Fig. 1F and G, the circTRIM28 was resistant to RNase R and Act D treatments, while GAPDH was impeded. In addition, the expression of circTRIM28 in the nucleus was higher than that in the cytoplasm (Fig. 1H). These outcomes exposed the circTRIM28 was upregulated in BC, and is more significantly upregulated in tamoxifen-resistant tissues and cells, which also could take effect in tamoxifen-resistant BC. Additionally, circTRIM28 structure pattern was manifested in circular RNA.

Silencing circTRIM28 enhanced tamoxifen sensitivity and cell apoptosis, whereas inhibited cell development in BC cells

MCF7/R and MDA-MB-231/R cells were transfected with sh-circTRIM28 (sh-circTRIM28#1, sh-circTRIM28#2, sh-circTRIM28#3), with sh-NC as control. The results designated that circTRIM28 was constrained in MCF7/R and MDA-MB-231/R cells by sh-circTRIM28 (Fig. 2A). Among them, sh-circTRIM28#1 had the most obvious inhibiting effect, so it was selected for subsequent tests. Functionally, the downregulated circTRIM28 diminished the cell vitality (Fig. 2B). In addition, the results showed that the knockdown of circTRIM28 decreased the IC50 value of tamoxifen (Fig. 2C and D). Besides, the EdU assay confirmed that sh-circTRIM28#1 transfection diminished the cell proliferation (Fig. 2E). Furthermore, downregulation of circTRIM28 constrained the number of colonies (Fig. 2F). Additionally, silencing circTRIM28 promoted cell apoptosis in MCF7/R and MDA-MB-231/R cells (Fig. 2G). Subsequently, the circTRIM28 deficiency inhibited invasion and migration of MCF7/R and MDA-MB-231/R cells (Fig. 2H and I). The PCNA, Cleaved-caspase 3, and MMP9 are related to cell proliferation, apoptosis, and invasion. The knockdown of circTRIM28 significantly repressed the level of PCNA and MMP9, whereas increased the expression of Cleaved-caspase 3 (Fig. 2J). Our results indicated that circTRIM28 deficiency enhanced tamoxifen sensitivity and cell apoptosis, but repressed cell proliferation, cell migration and invasion in BC cells.

MiR-409-3p targeted circTRIM28

CircBank, starbase and circinteractome was enforced to foresee the target miRNA of circTRIM28. Among them, four miRNAs overlapped, namely miR-217, miR-409-3p, miR-524-3p, and miR-525-3p (Fig. 3A). Furthermore, we revealed that miR-409-3p was amplified by circTRIM28 probe, but there were no significant changes in other miRNAs (Fig. 3B). Therefore, miR-409-3p was used
as the target miRNA for subsequent tests. Besides, the miR-409-3p level in BC tissues (n = 64) was significantly decreased than tumor-adjacent normal tissues (n = 64). Meanwhile, miR-409-3p was greatly diminished in tamoxifen-resistant tissues (BC-R, n = 34) relative to tamoxifen-sensitive tissues (BC-S, n = 30) (Fig. 3C). Moreover, our numbers also displayed that miR-409-3p was downregulated in BC cell lines (MCF7 and MDA-MB-231) relative to the control cell line (MCF10A). In addition, miR-409-3p was retarded in tamoxifen-resistant BC cell lines (MCF7/R and MDA-MB-231/R) in comparison to the parental cell lines (MCF7 and MDA-MB-231) (Fig. 3D and E). Furthermore, miR-409-3p was amplified by miR-409-3p mimics (Fig. 3F). Figure 3G showed the targeted binding site of miR-409-3p and circTRIM28. Results showed that the luciferase activity was lessened in circTRIM28-WT and that miR-409-3p co-transfected in MCF7/R and MDA-MB-231/R cells versus miR-NC groups even though no alteration was observed between circTRIM28-MUT co-transfection groups (Fig. 3H). The RIP and RNA pull-down assay confirmed the targeted relationship of miR-409-3p and circTRIM28 (Fig. 3I and J).

CircTRIM28 expedited BC by binding miR-409-3p
MiR-409-3p content was limited in MCF7/R and MDA-MB-231/R cells by anti-miR-409-3p (Fig. 4A). MCF7/R and MDA-MB-231/R cells were transfected with

Fig. 2 CircTRIM28 deficiency impeded BC. A The contents of sh-circTRIM28 were validated. B-D The cell viability and the IC50 of tamoxifen were detected. E and F The cell proliferation was examined. G The apoptosis of BC cells was measured. H and I The cell migration and invasion were assessed. J The PCNA, Cleaved-caspase 3 and MMP9 were measured. ***P < 0.001
sh-NC + anti-miR-409-3p and sh-circTRIM28#1 + anti-miR-409-3p, with sh-NC + anti-NC and sh-circTRIM28#1 + anti-NC as the control. We revealed that knockdown miR-409-3p could enhance the cell vitality and IC₅₀ value of tamoxifen, and reverted the effect of sh-circTRIM28#1 (Fig. 4B and C). For further research, we found that anti-miR-409-3p promoted cell proliferation and reverted the effect of silencing circTRIM28 (Fig. 4D and E). Additionally, the anti-miR-409-3p hindered the cell apoptosis, and reverted the effect of sh-circTRIM28#1 (Fig. 4F). Besides, we detected that anti-miR-409-3p promoted cell invasion and migration and reverted the effect of silencing circTRIM28 (Fig. 4G and H). Afterward, the anti-miR-409-3p could increase the level of PCNA and MMP9 and diminished the Cleaved-caspase 3 level, and reverted the effect of sh-circTRIM28#1 (Fig. 4I).

**MiR-409-3p bound to HMGA2 in MCF7/R and MDA-MB-231/R cells**

The bound sites of miR-409-3p in HMGA2 3’UTR were enforced by StarBase (Fig. 5A). Luciferase activity declined when BC cells with miR-409-3p mimics
and HMGA2–3’UTR-WT co-transfected. But the luciferase activity of BC cells that contained the HMGA2–3’UTR-MUT was not altered by miR-409-3p (Fig. 5B). Additionally, HMGA2 was notably decreased by miR-409-3p (Fig. 5C). Furthermore, western blot assay was performed to illustrate that HMGA2 was amplified by anti-miR-409-3p, whereas decreased by sh-circTRIM28#1. The anti-miR-409-3p reverted the effect of sh-circTRIM28#1 on the levels of HMGA2 (Fig. 5D).

Besides, HMGA2 in BC tissues (n=64) was upregulated compared with tumor-adjacent normal tissues (n=64). Meanwhile, the expression of HMGA2 was greatly increased in tamoxifen-resistance tissues (BC-R, n=34) relative to tamoxifen-sensitive tissues (BC-S, n=30) (Fig. 5E and F). What is more, our information demonstrated that HMGA2 was enhanced in BC cell lines (MCF7 and MDA-MB-231) than MCF10A cells. In addition, HMGA2 was significantly higher in tamoxifen-resistant BC cell lines (MCF7/R and MDA-MB-231/R) in comparison to the parental cell lines (MCF7 and MDA-MB-231) (Fig. 5G and H).

CircTRIM28 facilitated the progression of BC by regulating HMGA2

The results indicated that HMGA2 expression was memorably increased in MCF7/R and MDA-MB-231/R cells transfected with HMGA2 compared to the vector group (Fig. 6A). Then, MCF7/R and MDA-MB-231/R cells were transfected with sh-NC+HMGA2 and sh-circTRIM28+HMGA2, with sh-NC+vector or sh-circTRIM28+vector as the control. We found the HMGA2 could enhance the cell vitality and IC50 value of tamoxifen, and reverted the effect of sh-circTRIM28#1 (Fig. 6B and C). Besides, we established that HMGA2 stimulated cell proliferation and reverted the effect of silencing circTRIM28 (Fig. 6D and E). In addition, the HMGA2 overexpression could inhibit the cell apoptosis, and reverted the effect of sh-circTRIM28#1 (Fig. 6F). Moreover, we observed that HMGA2
overexpression promoted cell invasion and migration and reverted the effect of silencing circTRIM28 (Fig. 6G and H). Afterward, the HMGA2 overexpression could increase the level of PCNA and MMP9 and diminished Cleaved-caspase 3 level and reverted the effect of sh-circTRIM28#1 (Fig. 6I). In conclusion, our findings demonstrated that circTRIM28 deficiency inhibited the progression of BC by regulating HMGA2.

CircTRIM28 knockdown restricted tumor growth in vivo
MDA-MB-231/R cells were transfected with sh-circTRIM28#1 and sh-circTRIM28#1+ tamoxifen, with sh-NC and sh-NC+ tamoxifen as the control. As shown in Fig. 7A and B, tamoxifen treatment repressed tumor volume and weight. Meanwhile, circTRIM28 silencing could also restrain tumor volume and weight. The repression impact is more pronounced when tamoxifen acted synergistically with circTRIM28 downregulation. The Ki67 level was lower after the sh-circTRIM28#1 treatment. And tamoxifen treatment also repressed the expression level of Ki67. When tamoxifen and sh-circTRIM28#1 act synergistically, the inhibitory effect is enhanced (Fig. 7C). This suggests that silencing circTRIM28 enhances the inhibitory effect of tamoxifen on tumor development. The tumor tissues examined the impacts of circTRIM28 knockdown and tamoxifen treatment on circTRIM28, miR-409-3p, HMGA2, PCNA, Cleaved-caspase 3, and MMP9 expression. The results showed that circTRIM28 knockdown or tamoxifen treatment both repressed the expression of circTRIM28, miR-409-3p, HMGA2, PCNA, Cleaved-caspase 3, and MMP9 expression. The results showed that circTRIM28 knockdown or tamoxifen treatment both repressed the expression of circTRIM28, HMGA2, PCNA, and MMP9, but amplified the miR-409-3p and Cleaved-caspase 3 levels. The tamoxifen and sh-circTRIM28#1 act synergistically to enhance these effects (Fig. 7D and E). These outcomes signposted that circTRIM28 deficiency repressed tumor growth via miR-409-3p/HMGA2 axis. Besides, circTRIM28 knockdown could improve the drug sensitivity of BC tumors to tamoxifen.
Discussion

As an anti-estrogen, Tamoxifen has verified its value in the treatment of ER-positive BC. However, current research suggested that resistance to tamoxifen continues to be one of the primary causes of the treatment failure [5, 27]. Previous studies have shown that there are many mechanisms of TAM acquired resistance, among which epigenetic modification is of great importance [28]. Interestingly, this modification could alter gene expression without altering DNA sequence, containing non-coding RNAs. It has been confirmed that circRNAs could perform vital roles in TAM resistance and malignant biological behaviors in BC. For example, a series of circRNAs, such as circ_0025202 and circBMPR2 have been proved to partake in the regulation of tumor development and tamoxifen sensitivity in BC [29, 30]. Furthermore, preceding reporting has been discovered that the under-expression of circCDK1 might repress BC cell development and enhance the sensitivity of tamoxifen [31]. Herein, our research inspected the character of circTRIM28. In the present experiment, circTRIM28, a newly identified circRNA with typical characteristics, was aberrantly upregulated in tamoxifen-resistant BC tissues and cells. Moreover, we found a negative correlation between circTRIM28 high expression and poor prognosis of BC patients, further supporting the potential involvement of circTRIM28 in BC progression. In the functional experiment, our outcomes signposted that the downregulated circTRIM28 enhanced tamoxifen sensitivity and cell apoptosis, whereas inhibited cell proliferation, cell migration, and invasion in BC cells. Apart from that, the chemo-resistance of circTRIM28 was confirmed on BC xenografts in nude mice. These findings indicated that circTRIM28 appears to play a signification role in resistance to tamoxifen in vitro and in vivo.

Currently, one hypothesis is that circRNAs could function as the competing endogenous RNA (ceRNA) to sequester miRNAs away from their target mRNAs [21, 22]. Previous papers suggested that the regulatory networks were also applicable to the study of BC
progression and drug resistance [32]. In this study, circTRIM28 was witnessed to increase the speed of BC development by regulating miR-409-3p, which is parallel to preceding outcomes. MiR-409-3p regulated human ovarian cancer, colon cancer, and gastric cancer [33–35]. This evidence disclosed that miR-409-3p took part in human diseases development. Herein, we elucidated that miR-409-3p adjusted the evolution of BC by repressing HMGA2. The upshots confirmed that miR-409-3p could participate in BC.

HMG2 correlates with the progression of many human cancers. For example, HMG2 could regulate the cell metastasis of lung cancer [36]. Besides, HMG2 could modulate the Hippo-YAP signaling pathway to promote BC metastasis [37]. In this work, the expression of HMG2 was neutralized in BC. Besides, miR-409-3p upregulation repressed BC, but this effect was lessened by HMG2. These upshots backup the adjusting character of the circTRIM28/miR-409-3p/HMG2 in BC cells.

**Conclusion**
CircTRIM28 and HMG2 were intensified and miR-409-3p was constrained in BC. Furthermore, our study manifested that circTRIM28 knockdown enhanced tamoxifen sensitivity and cell apoptosis, whereas
inhibited cell proliferation, cell migration, and invasion in BC cells via miR-409-3p/HMGA2 axis. This paper manifests a new and feasible treatment idea in BC patients with chemoresistance.

Abbreviations
TAM: Tamoxifen; BC: breast cancer; circRNAs: circular RNAs; circ-TRIM28: containing 28; miR-409-3p: microRNA-409-3p; HMGA2: high mobility group box AT-hook 2; qRT-PCR: quantitative real-time reverse transcription-polymerase chain reaction; HR: Hormone receptor; IHC: Immunohistochemistry.

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Authors’ contributions
Conceptualization and Methodology: Changwu Zou and Yuxin Li; Formal analysis and Data curation: Xianguo Yang, Wei Liu and Guannan Zhang; Validation and Investigation: Nina Lu, Shiyong Yang and Changwu Zou; Writing – original draft preparation and Writing – review and editing: Shiyong Yang, Changwu Zou, and Yuxin Li; Approval of final manuscript: all authors.

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Availability of data and materials
The analyzed data sets generated during the present study are available from the corresponding author on reasonable request.

Declarations
Ethics approval and consent to participate
The present study was approved by the ethical review committee of Jing Men No. 2 People's Hospital. Written informed consent was obtained from all enrolled patients.

Consent for publication
Patients agree to participate in this work.

Competing interests
The authors declare that they have no competing interests.

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References
1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer. 2015;136(5):E359–86.
2. International Breast Cancer Study G, Colleoni M, Gelber S, Goldhirsch A, Aebi S, Castiglione-Gertsch M, et al. Tamoxifen after adjuvant chemotherapy for premenopausal women with lymph node-positive breast cancer: international breast cancer study group trial 13-93. J Clin Oncol. 2006;24(9):1332–41.
3. Jiang Q, Zheng S, Wang G. Development of new estrogen receptor-targeting therapeutic agents for tamoxifen-resistant breast cancer. Future Med Chem. 2013;5(9):1023–35.
4. Early breast cancer Trialists’ collaborative G: effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: an overview of the randomised trials. Lancet. 2005;365(9472):1687–717.
5. Musgrove EA, Sutherland RL. Biological determinants of endocrine resistance in breast cancer. Nat Rev Cancer. 2009;9(9):631–43.
6. Greene J, Baird AM, Brady L, Lim M, Gray SG, McDermott R, et al. Circular RNAs: biogenesis, function and role in human diseases. Front Mol Biosci. 2017;4:38.
7. Sheng JQ, Liu L, Wang MR, Li PY. Circular RNAs in digestive system cancer: potential biomarkers and therapeutic targets. Am J Cancer Res. 2018;8(7):1142–56.
8. Liu Z, Zhou Y, Liang G, Ling Y, Tan W, Tan L, et al. Circular RNA hsa_circ_001783 regulates breast cancer progression via sponging miR-200c-3p. Cell Death Dis. 2019;10(2):55.
9. Liang Hf, Zhang KZ, Liu BG, Ja GT, Li WL. Circular RNA circ-ABC10 promotes breast cancer proliferation and progression through sponging miR-127-1. Am J Cancer Res. 2017;7(7):1566–76.
10. Du WW, Yang W, Li X, Awan FM, Yang Z, Fang L, et al. Circular RNA circ-DNMT1 enhances breast cancer progression by activating autophagy. Oncogene. 2018;37(44):5829–42.
11. Qiu X, Wang Q, Song H, Shao D, Xue J. circ_103809 promotes breast cancer progression by regulating the PI3K/AKT signaling pathway. Oncol Lett. 2020;19(6):3725–30.
12. Ambros V. The functions of animal microRNAs. Nature. 2004;431(7006):350–5.
13. Garzon R, Marcucci G, Croce CM. Targeting microRNAs in cancer: rationales, strategies and challenges. Nat Rev Drug Discov. 2010;9(10):775–89.
14. Chen H, Dai J. miR-409-3p suppresses the proliferation, invasion and migration of tongue squamous cell carcinoma via targeting RDX. Oncol Lett. 2018;16(1):543–51.
15. Wu L, Zhang Y, Huang Z, Gu H, Zhou K, Yin X, et al. MiR-409-3p inhibits cell proliferation and invasion of osteosarcoma by targeting zinc-finger E-box-binding Homeobox-1. Front Pharmacol. 2019;10:1137.
16. Wang YW, Zhao S, Yuan XY, Liu Y, Zhang K, Wang J, et al. miR-4732-5p promotes breast cancer progression by targeting TSPO1. J Cell Mol Med. 2019;23(4):2549–57.
17. Ren S, Liu J, Feng Y, Li Z, He L, Li E, et al. Knockdown of circDENND4C inhibits glycolysis, migration and invasion by up-regulating miR-200b/c in breast cancer under hypoxia. J Exp Clin Cancer Res. 2019;38(1):388.
18. Krahn N, Meier M, To V, Booy EP, Mcleney K, O’Neil JD, et al. Nanoscale assembly of high-mobility group AT-hook 2 protein with DNA replication fork. Biophys J. 2017;113(12):2609–20.
19. Ergun S, Oztuçu S. Oncocers: ceRNA-mediated cross-talk by sponging miRNAs in oncogenic pathways. Tumour Biol. 2015;36(5):3129–36.
20. Qi X, Zhang DH, Wu N, Xiao JH, Wang X, Ma W, ceRNA in cancer: possible functions and clinical implications. J Med Genet. 2015;52(10):710–8.
21. Panda AC. Circular RNAs act as miRNA sponges: Adv Exp Med Biol. 2018;1087:67–79.
22. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Fahrenkrog B, Damgaard CK, et al. Natural RNA circles function as efficient microRNA sponges. Nature. 2013;495(7441):384–8.
23. Hou W, Zhang Y. Circ_0025033 promotes the progression of ovarian cancer by activating the expression of LSM4 via targeting miR-184. Pathol Res Pract. 2021;217:153275.
24. Zheng X, Huang M, Xing L, Yang R, Wang X, Jiang R, et al. The circRNA circSEPT9 mediated by EZF2 and EID4A3 facilitates the carcinogenesis and development of triple-negative breast cancer. Mol Cancer. 2020;19(1):73.
25. Ma Y, Zha J, Yang X, Li Q, Zhang Q, Yin A, et al. Long-chain fatty acyl-CoA synthetase 1 promotes prostate cancer progression by elevation of lipidogenesis and fatty acid beta-oxidation. Oncogene. 2021;40(10):1806–20.
26. Li Z, Huang C, Bao C, Chen L, Lin M, Wang X, et al. Exon-intron circular RNAs regulate transcription in the nucleus. Nat Struct Mol Biol. 2015;22(3):256–64.
27. Viedma-Rodriguez R, Baiza-Gutman L, Salamanca-Gomez F, Diaz-Zaragoza M, Martinez-Hernandez G, Ruz Esparraga-Garrido R, et al. Mechanisms associated with resistance to tamoxifen in estrogen receptor-positive breast cancer (review). Oncol Rep. 2014;32(1):3–15.
28. Raja P, Thomas S, Munster PN. Epigenetic modulation: a novel therapeutic target for overcoming hormonal therapy resistance. Epigenomics. 2011;3(4):451–70.
29. Liang Y, Song X, Li Y, Ma T, Su P, Guo R, et al. Targeting the circ8WPR2/miR-553/USP44 Axis as a potent therapeutic approach for breast Cancer. Mol Therapy Nucleic Acids. 2019;17:347–61.
30. Sang Y, Chen B, Song X, Li Y, Liang Y, Han D, et al. circRNA_c002502 regulates Tamoxifen sensitivity and tumor progression via regulating the miR-182-5p/FOXD3a Axis in breast Cancer. Mol Therapy. 2021;29(12):3525–7.
31. Liu D, Zhou Z, Guo Y, Du Q, Li L. CircCDK1 knockdown reduces CDK1 expression by targeting miR-489-3p to suppress the development of breast cancer and strengthen the sensitivity of Tamoxifen. Anti-Cancer Drugs. 2022;33(3):286–99.
32. Gao L, Shen K, Yin N, Jiang M. Comprehensive Transcriptomic analysis reveals Dysregulated competing endogenous RNA network in endocrine resistant breast Cancer cells. Front Oncol. 2020;10:600487.
33. Li Y, Chen L, Zhang B, Ohno Y, Hu H. miR-409-3p inhibits the proliferation and migration of human ovarian cancer cells by targeting Rab10. Cell Mol Biol (Noisy-le-grand). 2020;66(7):197–201.
34. Tan S, Shi H, Ba M, Lin S, Tang H, Zeng X, et al. miR-409-3p senstizes colon cancer cells to oxaliplatin by inhibiting Beclin-1-mediated autophagy. Int J Mol Med. 2016;37(4):1030–8.
35. Wang Y, Zhang J, Chen X, Gao L. Circ_0001023 promotes proliferation and metastasis of gastric Cancer cells through miR-409-3p/PHF10 Axis. Onco Targets Ther. 2020;13:4533–44.
36. Gao X, Dai M, Li Q, Wang Z, Lu Y, Song Z. HMGA2 regulates lung cancer proliferation and metastasis. Thorac Cancer. 2017;8(S1):S01–10.
37. Xu J, Fang X, Long L, Wang S, Qian S, Lyu J. HMGA2 promotes breast cancer metastasis by modulating hippo-YAP signaling pathway. Cancer Biol Ther. 2021;21(1):5–11.

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