Crescentic Glomerulonephritis: Pathogenesis and Therapeutic Potential of Human Amniotic Stem Cells

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Chronic kidney disease (CKD) leads to significant morbidity and mortality worldwide. Glomerulonephritis (GN) is the second leading cause of CKD resulting in end stage renal failure. The most severe and rapidly progressive type of GN is characterized by glomerular crescent formation. The current therapies for crescentic GN, which consist of broad immunosuppressive drugs, are partially effective, non-specific, toxic and cause many serious side effects including infections, cancer, and cardiovascular problems. Therefore, new and safer therapies are needed. Human amniotic epithelial cells (hAECs) are a type of stem cell which are isolated from the placenta after birth. They represent an attractive and novel therapeutic option for the treatment of various inflammatory conditions owing to their unique and selective immunosuppressive ability, as well as their excellent safety profile and clinical applicability. In this review, we will discuss the immunopathogenesis of crescentic GN, issues with currently available treatments and how hAECs offer potential to become a new and harmless treatment option for this condition.

Keywords: chronic kidney disease, crescentic glomerulonephritis, human amniotic stem cells, inflammation, immunity

INTRODUCTION

The immune system and the kidneys are closely linked, and chronic kidney disease (CKD) often results from various auto(immune) disorders. CKD is due to slowly progressive, chronic deterioration of kidney function. Crescentic glomerulonephritis (GN) is a chronic immune-mediated disease which causes severe glomerular inflammation and injury, and often leads to irreversible kidney failure. It is a common cause of morbidity and mortality worldwide. GN is a major contributor to the escalating health burden associated with CKD.

Almost all current therapeutic concepts in autoimmune diseases are based on the systemic suppression of immune functions and are not curative. Currently used immunosuppressive therapies for crescentic GN are only partially effective, toxic and provide broad, non-specific immunosuppression, thus producing significant adverse effects. These including mainly severe infections, but also cancer and cardiovascular events. It is these treatment-caused side effects which cause the majority of patient deaths. Therefore, new and safer therapies are much needed.
Human amniotic epithelial cells (hAECs) are a type of stem cell which are isolated from the human placenta after birth. They are an attractive and novel therapeutic option for the treatment of crescentic GN due to their (i) ethical, non-invasive, and speedy isolation from the amniotic membrane of the placenta which results in an abundance of readily available cells, (ii) potent immunosuppressive capacity, (iii) low immunogenicity (ability to activate the immune system) and (iv) their ability to fight microbes and cancer and protect against cardiovascular disease. hAECs have attenuated various inflammatory diseases in mouse models, without being rejected, producing tumors or causing any major side effects. They are currently being tested in several clinical trials to treat different conditions including lung and liver disease and stroke. The purpose of this review is to discuss and link some of the major immune and inflammatory mechanisms to the progression of crescentic GN, discuss the common therapies and their limitations, and outline the therapeutic potential of hAECs in safely reducing glomerular injury.

**CRESCENTIC GLOMERULONEPHRITIS**

The term glomerulonephritis (GN) refers to immune-mediated inflammation of the renal glomeruli. GN is diagnosed based on clinical presentation, etiology, histopathology or pathogenesis. Most patients present with hematuria, proteinuria, and impaired glomerular filtration rate (Chadban and Atkins, 2005). This pathological condition can be acute or chronic (developing over several months to years) based on the timing of clinical presentation (Mejia-Vilet and Parikh, 2019). Acute GN that develops into rapidly progressive disease most often results from conditions that involve an abnormal immune reaction. Sometimes, acute GN does not resolve, and instead becomes long lasting (chronic) (Vinen and Oliveira, 2003).

Crescentic GN is a severe form of glomerulonephritis characterized by the destruction of the renal glomeruli that often lead to end-stage renal disease over a relatively short period of time (days, weeks, or months) (Jennette and Thomas, 2001; Jennette, 2003). It is characterized morphologically by extensive crescent formation, defined as two or more cell layers in Bowman’s or urinary space (Jennette and Thomas, 2001; Parmar and Bashir, 2019). Crescents are formed by infiltrating and proliferating immune and local cells, along with deposited fibrin. This occurs after the disruption of the glomerular structure, which allow for circulating cells, inflammatory cytokines, and blood proteins to pass through the blood vessel wall into the Bowman space. The major components in the glomerular crescent are procoagulant factors, macrophages, T cells, fibroblasts, and parietal and visceral epithelial cells (Karras, 2018; Tsui et al., 2018).

On the basis of immunopathological findings, crescentic GN can be classified into three major categories: anti-glomerular basement membrane (GBM) antibody disease (Goodpasture’s syndrome), immune complex GN (e.g., lupus nephritis), and pauci-immune GN which is often associated with anti-neutrophil cytoplasmic autoantibodies (ANCA) and is thus also known as ANCA-associated vasculitis (AAV) (Jennette and Thomas, 2001; Parmar and Bashir, 2019). Anti-GBM autoantibodies are highly specific for Goodpasture’s disease, in which they are generally directed against the non-collagenous (NC1) domain of the alpha 3 chain of type IV collagen [α3(IV)NC1] (Gulati and McAdoo, 2018). These immunoglobulins cause glomerular capillary wall damage by local complement activation and neutrophils. T-cells also play a distinct pathogenic role in driving cell-mediated destruction of the glomeruli in this disease (Dean et al., 2005; McAdoo and Pusey, 2017). Environmental factors, like smoking, hydrocarbons and exposure to high oxygen, are thought to increase the likelihood of developing anti-GBM (McAdoo and Pusey, 2017). Anti-GBM GN accounts for around 10–15% of all cases of crescentic GN (McAdoo and Pusey, 2017). Immune complex GN, which comprises 25–30% of all cases crescentic GN, is characterized by a granular pattern of immune complex deposition in glomeruli (Naik and Shawar, 2020). AAV is associated with ANCA specific for neutrophil proteins, predominantly myeloperoxidase (MPO) or proteinase 3 (PR3) (Dey et al., 2016). This is the most common form of crescentic GN, contributing to about 65-70% of all cases (Naik and Shawar, 2020), and it refers to a necrotizing/crescentic GN with few or no immune deposits in glomeruli, as detected by immunofluorescence (Syed et al., 2015).

**Pre-clinical Models of Crescentic Glomerulonephritis**

Experimental evidence for the pathogenesis of crescentic GN comes mainly from pre-clinical models, including lupus nephritis, experimental autoimmune GN (EAG; model of Goodpasture’s disease), MPO-ANCA vasculitis and nephrotoxic nephritis (NTN).

**Nephrotoxic Nephritis**

A large body of evidence about the immunopathogenesis of crescentic GN has come from one of the most widely used and best-characterized models, called nephrotoxic nephritis (NTN), also known as autologous anti-GBM globulin GN. In this model, rodents such as rats, mice and rabbits are passively injected with foreign polyclonal antibodies (globulins) targeting the mouse GBM (Odobasic et al., 2014). It comprises of two distinct phases. The first phase is called the heterologous phase and is associated with transient glomerular injury and inflammation due to the binding of injected foreign antibodies in a linear fashion to the GBM, before the development of an adaptive immune response (Odobasic et al., 2014). This phase is characterized by a neutrophil influx which peaks around 2 h after the injection of transferred antibodies, and proteinuria, peaking within the first 24 h (Schrijver et al., 1990; Tipping et al., 1994; Odobasic et al., 2014). Neutrophils contribute to glomerular damage by producing several inflammatory mediators including reactive oxygen species (ROS), proteases (Johnson et al., 1988; Couser, 1998), and MPO (Odobasic et al., 2007). Accumulation of neutrophils in inflamed glomeruli is dependent on intraglomerular expression of adhesion molecules, P-selectin, and intercellular adhesion molecule-1 (ICAM-1) (Tipping et al., 1994; Nomura et al., 1996; Saleem et al., 1998). The pathogenic role of neutrophils in glomerulonephritis is based...
on the production of several inflammatory mediators including ROS and protease such as elastase, proteinases and cathepsin G (Johnson et al., 1988; Couser, 1998).

The second phase is called the autologous phase and tends to begin about 6 to 7 days later. It is caused by an adaptive immune response to the foreign anti-GBM globulin (Odagard et al., 2014; Ougaard et al., 2018). This phase is characterized by more severe injury and crescent formation due to the presence of cellular effectors including CD4+ T cells, macrophages, neutrophils and fibrin (Huang X.-R. et al., 1997; Huang X. et al., 1997).

In mice, an accelerated model of NTN can be induced by pre-immunizing mice with normal sheep globulin (NSG) (Odagard et al., 2018). In accelerated NTN, the use of adjuvants which contain dead mycobacteria are included in the pre-immunization to improve the immune response (Odagard et al., 2018). This pre-immunization produces a strong T and B cell response to sheep globulin and gives rise to an immediate autologous phase once sheep anti-GBM antibodies are administered (Odagard et al., 2014). For animal models without pre-immunization, injury is less acute compared to the pre-immunized accelerated model and termed as a non-accelerated model (Odagard et al., 2014).

**Experimental Models of MPO-AAV, Lupus Nephritis, and Goodpasture’s**

Other models of crescentic GN exist and they have been also widely used to reveal pathogenetic mechanisms of disease. MPO-AAV can be induced in mice by immunizing animals with MPO which results in the development of active anti-MPO autoimmunity, followed by neutrophil lodgment in glomeruli and deposition of the autoantigen for subsequent recognition by MPO-specific T cells (Ruth et al., 2006; Odobasic et al., 2019). Alternatively, glomerular injury can be initiated by passive transfer of anti-MPO antibodies (Xiao et al., 2002; Ooi et al., 2014). Experimental autoimmune GN (EAG), a model of Goodpasture’s disease, can be induced in mice by repeated immunization with α3(IV)NC1 (Ooi et al., 2009). Several models of lupus nephritis exist in which susceptible mice such as MRL/lpr and NZB/NZW1 spontaneously develop disease (Richard and Gilkeson, 2018). Although none of these models fully recapitulate human disease, they all (including NTN) closely resemble crescentic GN seen in patients, both immunologically and pathologically, and are therefore invaluable pre-clinical tools to study disease pathogenesis and test new potential therapies.

**Mechanisms of Pathogenesis of Crescentic Glomerulonephritis**

Multiple immune mechanisms contribute to the pathogenesis of crescentic GN. A summary of the main immune pathways which positively or negatively regulate the development of crescentic GN is shown in Figure 1A.

**The Role of Innate Immunity – Neutrophils and Macrophages**

Innate and adaptive immune system activation are a common underlying mechanism for several forms of crescentic GN (Imig and Ryan, 2013). Studies in the NTN model using neutrophil depletion have shown that these cells contribute to glomerular injury during the early autologous phase of disease (Disteldorf et al., 2015). Macrophages are the dominant effector cells present in the kidney which mediates glomerular crescent formation and kidney damage in all models of crescentic GN (Lan et al., 1997; Odobasic et al., 2014; Rousselle et al., 2017). Their accumulation within glomeruli has been blocked by administration of polyclonal sheep anti-rabbit macrophage serum in models of anti-GBM disease, leading to prevention of the development of GN (Holdsworth et al., 1981). Moreover, a micro-encapsulated approach in which macrophages are depleted by clodronate has resulted in reducing renal damage in anti-GBM GN (D’Souza et al., 1999). Further studies have shown that macrophages can induce glomerular injury by several mechanisms including deposition of fibrin (which impairs glomerular filtration) and production of various proinflammatory mediators including ROS, reactive nitrogen species such as nitric oxide (NO) and cytokines such as IL-1, TNF-α, and macrophage migration inhibitory factor (MIF) (Imig and Ryan, 2013).

**The Role of Adaptive Immunity – T Cells and B Cells**

Numerous studies in various models of crescentic GN have shown that CD4+ T cells play a critical role in this disease (Tipping et al., 1998; Dean et al., 2005; Ruth et al., 2006). CD4+ T helper cells (Th) can be divided into several subsets, with the major and best-understood ones being Th1, Th17 and Th2, mainly characterized by the expression of IFNγ, IL-17A, and IL-4, respectively. Several studies have indicated that crescentic anti-GBM globulin GN is driven by Th1 cells, while Th2 cells reduce the severity of disease. For instance, GN induced in mice with a predominant Th1 response (C57BL/6) shows severe crescentic formation with prominent glomerular T cell and macrophage infiltration and fibrin deposition which are associated with elevated IFN-γ and weak IL-4 production (Huang X.-R. et al., 1997). On the other hand, in Th2-prone (BALB/c) mice, crescent formation as well as glomerular T cell and macrophage influx were relatively low (Huang X.-R. et al., 1997). In addition, mice lacking endogenous IFN-γ developed less severe GN than genetically normal C57BL/6 mice, whereas IL-4-deficient mice developed more severe crescentic GN associated with increased accumulation of T cells and macrophages in glomeruli (Kitting et al., 1998, 1999). While IFNγ is protective in EAG (Kitting et al., 2004), the Th1 pathway has also been shown to promote glomerular crescent formation and kidney injury in experimental lupus nephritis and MPO-AAV (Richards et al., 2001; Summers et al., 2011).

Moreover, it has become clear that the Th17 pathway is important in the development of crescentic GN. In experimental MPO-AAV, mice lacking IL-17A are protected from early glomerular injury (Gan et al., 2010). Similarly, several lines of evidence, coming from using IL-17A-deficient mice or infusion of Th17 cells, have shown that the Th17 pathway promotes early glomerular injury in NTN (Paust et al., 2009; Summers et al., 2009; Odobasic et al., 2011), in line with results from experimental autoimmune uveitis (Luger et al., 2008). On the contrary, the Th1 pathway acts later in the disease process and produces more severe GN (Odobasic et al., 2011). Interestingly, local IL-17A production is protective in crescentic
FIGURE 1 | The major immune pathways involved in the pathogenesis of crescentic GN and mechanisms by which hAECs may attenuate glomerular injury. (A) Effector immune mechanisms in crescentic GN. Macrophages contribute by secreting nitric oxide (NO) and TNF-α, while neutrophils contribute by producing reactive oxygen species (ROS) and releasing myeloperoxidase (MPO) enzyme. IFN-γ produced by Th1 cells mediates crescent formation. IL-17A (a Th17 cytokine) promotes early stage of kidney injury, but attenuates established disease by inhibiting Th1 responses. B cells, which act as antigen-presenting cells and produce autoantibodies, also promote glomerular injury. On the contrary, IL-4 producing Th2 and IL-10-producing regulatory T cells (Tregs) play a protective role in this disease. Arrows indicate glomerular crescent formation. (B) Potential mechanisms by which hAECs may reduce inflammation and damage in glomeruli. hAECs suppress activation and/or infiltration of pro-inflammatory T cells (Th1 and Th17), B cells, macrophages and neutrophils, while promoting inhibitory cells such as Tregs and Th2. They exert their effects by producing anti-inflammatory mediators such as TGF-β, IL-10 and PGE2, and releasing exosomes, to restrict kidney injury.

GN (Hamour et al., 2015), and in later stages of the disease (which is mediated by Th1), IL-17A can attenuate injury by systemically inhibiting Th1 responses (Odobasic et al., 2011).

Tregs are a specialized inhibitory subset of CD4⁺ T cells characterized by expression of CD25 and Foxp3. They play a key role as negative regulators of pathogenic immunity in crescentic GN. Studies using Foxp3-GFP reporter mice explored the functional role of T-regulatory cell in inhibiting anti-GBM nephritis (Ooi et al., 2011). Foxp3 protected against renal damage in anti-GBM GN induced in antigen-primed Foxp3-transgenic mice, via boost of Treg numbers and activity, and inhibition of Th immune responses at the systemic level and at sites of tissue injury (Yang et al., 2017). Similarly, Tregs are protective in other models of crescentic GN including MPO-AAV and lupus nephritis (Humrich et al., 2010; Tan et al., 2013; Odobasic et al., 2019). Further studies have shown that Tregs attenuate crescentic GN by releasing a potent anti-inflammatory cytokine, IL-10 (Ostmann et al., 2013).
In addition to T cells, B cells and autoantibodies have also been demonstrated to play a major role in the development of crescentic GN. B cells and/or antibodies against various autoantigens including MPO in AAV, α3(IV)NC1 in Goodpasture's and nuclear antigens in lupus, are pathogenic in experimental crescentic GN (Xiao et al., 2002; Dean et al., 2005; Richard and Gilkeson, 2018). In contrast, evidence coming from μ-chain knockout mice, which lack mature B cells and cannot produce immunoglobulin, has shown that glomerular antibody deposition is not essential for crescent formation in response to the planted glomerular antigen in the NTN model (Li et al., 1997). B cells are well known to promote immune-mediated injury by several mechanisms including acting as antigen-presenting cells to activate T cells and differentiating into autoantibody-producing plasma cells.

**Pathogenic Mechanisms in Human Crescentic Glomerulonephritis**

In human studies, strong evidence from patients' kidney biopsies, and blood samples has accumulated to support the role of immune cells and antibody in crescentic GN, with findings similar to those in animal models. While neutrophils are prominent in crescentic GN (Kaplan, 2013), a considerable body of evidence has emerged to show that CD4+ T cells and macrophages are critically involved in all patterns of human crescentic glomerulonephritis (Stachura et al., 1984; Bolton et al., 1987; Nolasco et al., 1987; Müller et al., 1988; Cunningham et al., 1999).

Although many limitations have constrained the assessment of nephritogenic responses in human GN, a number of studies have indicated that crescentic glomerulonephritis is a manifestation of a Th1 predominant delayed type hypersensitivity (DTH)-mediated immune response associated with predominant infiltration of macrophages and deposition of fibrin (Kitching et al., 2000). Furthermore, patients with crescentic GN have increased serum levels of IL-17A in comparison to healthy individuals, suggesting a role for the Th17 pathway in disease pathogenesis (Lu et al., 2017; Herrnstadt and Steinmetz, 2020). Human studies in AAV, SLE, and Goodpasture's syndrome have revealed that dysregulation of Treg homeostasis and function is also associated with the development of crescentic GN (Herrnstadt and Steinmetz, 2020). Finally, B cells and autoantibodies against various endogenous targets in crescentic GN are pathogenic in human disease (Falk and Jennette, 1988; McAdoo and Pusey, 2017).

**Current Therapies for Crescentic Glomerulonephritis**

In general, crescentic GN therapy is often classified into two phases. First is the activation of remission phase during the acute period, followed by the subsequent phase of maintenance therapy to control the underlying long-term immunopathology (Parmar and Bashir, 2017). The induction treatment is mainly composed of high-dose corticosteroids and intravenous (i.v.) pulse cyclophosphamide with the purpose of blocking the active inflammation and reducing the cellular and humoral immune response (Jennette, 2003; Moroni and Ponticelli, 2014). In addition, therapeutic plasma apheresis (plasma exchange) is a form of treatment that is frequently used to treat almost all types of crescentic GN by removing pathogenic autoantibodies (Flossmann et al., 2011; Kallenberg, 2014; Prenderci and Pusey, 2019).

Corticosteroids, synthetic drugs which closely resemble cortisol hormone, have been used for decades to modulate inflammation therapeutically by decreasing the movement of neutrophils to the inflammatory sites and inducing a transient lymphocytopenia (Oines et al., 2016). This medication inhibits the expression and action of most proinflammatory cytokines, adhesion molecules, and suppresses MHC expression (Ponticelli and Locatelli, 2018). Cyclophosphamide, a cytotoxic drug, is one of the oldest anti-cancer drugs and is widely used in therapy of crescentic GN by reducing the activity of the immune system (Jhaveri et al., 2013). Cyclophosphamide therapy significantly reduces total neutrophil, macrophage and lymphocyte counts (Jhaveri et al., 2013; Ménétrier-Caux et al., 2019).

Combination therapy with pulse cyclophosphamide plus pulse methyl prednisolone therapy significantly reduce proteinuria, serum creatinine, the level cellular crescents in crescentic GN (Tumlin et al., 2003), and present a greater reduction in the risk for ESRD.

However, induction therapy (corticosteroids with cyclophosphamide) is only partially effective, non-specific and produces many toxicities and serious side effects. Therefore, there is a major unmet need for safer, effective therapies. For example, in AAV, this treatment induces remission in 70–90% of patients, but the incidence of dialysis or death at 5 years is still high (~30%), with the majority of early deaths caused by drug-related side effects, mainly infections caused by broad immunosuppression (Flossmann et al., 2011; Kallenberg, 2014; King and Harper, 2017). In lupus nephritis, remission is achieved in 50–60% of patients at best (Mene et al., 2018), relapse occurs in up to 25% of cases (Mene et al., 2018; Anders et al., 2020) and despite optimal care, many patients (up to 20%) develop kidney failure which requires renal transplantation or dialysis for survival (Mene et al., 2018; Anders et al., 2020). The unwanted adverse effects caused by these drugs include predominantly severe infections, as well as increased risk of malignancy, bone disease, dysglycemia, obesity, hypertension, mental problems, gastrointestinal bleeding, cataracts, and long-term risks of developing cardiovascular disease (King and Harper, 2017; Jefferson, 2018). More recently, rituximab, a pan B cell depleting anti-CD20 monoclonal antibody, has been approved for use instead of cyclophosphamide as a second line of treatment, particularly in AAV. However, although it is not inferior to cyclophosphamide in reducing disease, it induces similar rates of adverse effects, mainly infections, most likely due to hypogammaglobulinaemia and late onset neutropenia (Jones et al., 2015; Santos et al., 2020).

Adalimumab, an anti-TNFα monoclonal antibody, has been used as an adjunct therapy to standard immunosuppression. It can be an effective therapy for the induction of remission in AAV and may permit reduced prednisolone dosing, thus decreasing therapy-related toxicity (Laurino et al., 2010). However, other anti-TNFα agents, infliximab and etanercept,
failed to demonstrate a benefit for remission maintenance in patients with AAV (Booth et al., 2004; Wegener’s Granulomatosis Etanercept Trial (WGET) Research Group, 2005).

Mycophenolate mofetil (MMF), is a salt form of the immunosuppressive drug mycophenolic acid. It inhibits the releasing of proinflammatory cytokines, nitric oxide, and LDH in macrophages and suppresses proliferation and infiltration of both T and B lymphocytes (Allison and Eugui, 2000). Recent studies, indicate that MMF may be as or even more effective in reducing proteinuria and hematuria and less toxic compared with cyclophosphamide in LN (Hu et al., 2002).

STEM CELL-BASED THERAPY FOR AUTOIMMUNE AND INFLAMMATORY DISEASES

Stem cell-based therapy is an attractive approach to ameliorate a broad range of human diseases and injuries, and it has been proven to be safe and effective in a wide range of immune-mediated diseases (Jin et al., 2014). Several types of stem cells, including mesenchymal, induced pluripotent and embryonic (Ryu et al., 2020), have reduced organ injury in models of immune diseases and their potential side effects and efficiency have been assessed in clinical trials (Mûzes and Sipos, 2019).

Mesenchymal, Embryonic, and Induced Pluripotent Stem Cells

Mesenchymal stem cells (MSCs) are multipotent adult stem cells that exist in various locations including umbilical cord, bone marrow and adipose tissue (Murray and Péault, 2015). MSCs have been successfully applied in treating a vast array of inflammatory and autoimmune conditions such as graft-versus-host disease (GVHD), multiple sclerosis (MS), type 1 diabetes (T1D), inflammatory bowel diseases (IBD), systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), lung fibrosis, liver, and pancreatic fibrosis (Ryu et al., 2020). These cells modulate the immune reaction directly through regulating various immune cells including dendritic cells (DCs), macrophages, B cells, T cells and neutrophils, and by producing suppressive cytokines such as IL-10, TGF-β, and IL-35, as well as expressing inhibitory ligands and receptors (e.g., PD-L1 and PD-1) (Jiang and Xu, 2020). MSCs can attenuate the development of crescentic GN (Thakkar et al., 2017) by decreasing neutrophil and macrophage recruitment to the kidney and promoting the phenotypic switching of renal macrophages to immunoregulatory cells (Furuhashi et al., 2013). In addition, the beneficial effects of MSCs in GN appear to be mediated by modification of the Th1/Th2 and/or Th17/Treg balance and inhibition of B-cell activation, as well as stimulation of IL-10, IL-4, foxp3, prostaglandin E2 (PGE2), and TGF-β production (Ma et al., 2013; Suzuki et al., 2013). Hence, MSCs have demonstrated capacity to attenuate experimental crescentic GN, but they also have a few disadvantages and carry some risks. For example, some methods of MSC isolation are invasive (e.g., obtaining MSCs from the bone marrow or adipose tissue). MSCs also require long-term culture (weeks) to expand adequate cell numbers for infusion. This increases the cost and risk of in vitro mal-transformation. It is also not optimal for the treatment of diseases such as crescentic GN in which early treatment is required to stop the progression of rapidly progressing deterioration of kidney function.

Embryonic and induced pluripotent stem cells (ESCs and iPSCs) are pluripotent stem cells. ESCs are isolated from the inner cell mass of blastocysts, while iPSCs are produced from adult somatic cells that are genetically reprogrammed to an ESC-like state by transcription factors (Ryu et al., 2020). It has been reported that the ESC-loaded gelatin microcryogels on rats slowed down the progression of CKD and alleviated glomerular injury (Geng et al., 2016). Likewise, administration of iPSCs in models of CKD preserved residual renal function by decreasing macrophage infiltration, upregulating TGF-β, inhibiting apoptosis and regulating cell proliferation and death signaling (Caldas et al., 2017; Sheu et al., 2020). Thus, ESCs and iPSCs may be able to reduce glomerular damage and retard the progression of CKD, but they also have some disadvantages. For instance, isolation of ESCs poses obvious ethical concerns, while iPSCs carry the risk of tumor development (Caldas et al., 2017).

Human Amniotic Epithelial Cells

One particular stem cell type, hAECS, have attracted much attention in the recent years as an ideal therapeutic option for the treatment of autoimmune and inflammatory disorders due to their immunosuppressive ability and their superior safety and clinical applicability over other stem cell types. hAECs are a heterogeneous epithelial population that originates from the lining of the inner membrane of the placenta which provides an abundant cellular source for stem cell-based therapy (Miki et al., 2005). In comparison to other sources of stem cells, gestational tissue gives great advantages including easy collection without the need for invasive methods (Qiu et al., 2020). Large numbers of hAECS can be isolated from the placenta after birth, thus bypassing ethical barriers and resulting in an abundance of immediately available, primary (non-cultured) cells to be used therapeutically. These epithelial stem cells are pluripotent and have the capability of self-renewal and differentiating into all three germ layers, including the ectoderm, mesoderm, and endoderm (Miki et al., 2005). hAECs have immense potential to safely reduce the burden of many serious diseases and injuries to different organs, including the kidney, due to their unique properties (Ren et al., 2020). They possess some degree of plasticity, immune privilege, non-tumorigenicity, anti-infection/cancer properties, and lack of ethical concerns and paracrine properties that are essential to their potential therapeutic applications in immune-mediated diseases (Qiu et al., 2020).

Low Immunogenic Profile

Human Leukocyte antigens (HLA), encoded by the major histocompatibility complex (MHC) gene complex in humans, are the major molecules that initiate graft rejection (Mahdi, 2013).
However, low HLA class-I (HLA-A, HLA-B, and HLA-C) expression and HLA class-II (HLA-DR) on hAEC surface have been identified, resulting in a low immunogenic profile upon transplantation (Hori et al., 2006). hAECs express non-classical HLA-G, which is thought to protect the fetal semi-allograft from maternal immune system rejection (Lefebvre et al., 2000). Expression of HLA-G confers a degree of immune privilege by suppressing natural killer cells, inducing apoptosis of activated CD8+ T cells and inhibiting CD4+ T cell proliferation (Banás et al., 2008).

**Low Risk of Tumor Formation**

Tumorigenicity is a common obstacle for cell-based therapies, since some cells may cause formation of tumors due to being immortal. Unlike ESCs and iPSCs (Ben-David and Benvenisty, 2011), hAECs do not express telomerase reverse transcriptase (Miki et al., 2005), which is a catalytic subunit of the telomerase enzyme playing a central role in tumorigenesis (Daniel et al., 2012). Therefore, hAECs do not promote tumor formation after transfer into recipients.

**Immunosuppressive Capacity**

Consistent with the role of the placenta to protect the fetus from being attacked by the maternal immune system during pregnancy, hAECs are immunosuppressive. Similar to MSCs, the beneficial effects of hAECs are primarily mediated via their paracrine actions and not by their differentiation into target cells (Tógel et al., 2005; Wang et al., 2014). hAECs have blocked the immune system by suppressing effector T cells, switching macrophage polarization from M1 to anti-inflammatory M2 phenotype, and inhibiting neutrophils (Li et al., 2005; Tan J.L. et al., 2018). In addition, several studies in animal models have shown that they can exert immunomodulatory effects by inducing other immunosuppressive cells, in particular Tregs and Bregs (Manuelpillai et al., 2016; Liu et al., 2012; Tan et al., 2014, 2015; Evans et al., 2018; Li et al., 2018; Tan B. et al., 2018). Moreover, in vitro studies showed similar inhibitory effects of hAECs on human cells. For instance, hAECs suppressed human CD4+ T cell proliferation, induced a Th2 cytokine profile, suppressed production of Th1 and Th17 cytokines and promoted differentiation of naïve CD4+ T cells into Tregs (Wolbank et al., 2007; Motedayyen et al., 2018).

Human amniotic epithelial cells have been reported to inhibit immunity by secreting various immunosuppressive mediators. For example, they produce prostaglandin E2 (PGE2), which has several immunosuppressive properties such as inhibition of T cell proliferation. hAECs also secrete TGFβ, a T cell growth inhibitor and a powerful immunosuppressive molecule (Liu et al., 2012). In addition, the suppressive activity of hAECs has been demonstrated through an increased secretion of the anti-inflammatory cytokine IL-10 (Charles-Henri and Ekaterine, 2020). IL-10 inhibits proinflammatory cytokine production, as well as Th1 and macrophage activation (Howes et al., 2014).

**Anti-infection and Anti-cancer Properties**

Unlike the current therapies for crescentic GN, hAECs have anti-infection properties and protect against cancer development. Human β-defensins, small proteins which promote microbial death, are primarily expressed by epithelial and immune cells at mucosal surfaces (Dorin et al., 2015). These natural antimicrobial molecules have been reported to be secreted by human placenta cells including hAECs and by the fetal membrane during human pregnancy in order to protect the uterus from infection (King et al., 2007; Nemr et al., 2017). hAECs also produce type I interferons (IFNs) in response to viruses in vitro (Uchide et al., 2002; Nemr et al., 2017). This group of proteins, which is made up mainly of IFNα and IFNβ, initiate intracellular anti-microbial systems and influence innate and adaptive immune responses (Ivashkiv and Donlin, 2014). They are secreted by infected cells and are important for host protection against viruses through the induction of anti-viral effector molecules (Uchide and Toyoda, 2007). Moreover, hAECs suppress cancers directly by inducing apoptosis and reducing motility of malignant cells (Niknejad et al., 2014). They also suppress tumor development indirectly by promoting anti-tumor cytotoxic T cell immunity in vivo, as shown in a mouse model of colon adenocarcinoma (Tabatabaei et al., 2018).

**Protection From Cardiovascular Disease**

In contrast to the current drugs used to treat crescentic GN, hAECs also protect against cardio-vascular disease. They have decreased areas of myocardial infarction in athymic nude rats (Fang et al., 2012). In addition, administration of hAECs has reduced brain injury in a murine and non-human primate model of ischemic stroke (Evans et al., 2018).

**Human Amniotic Epithelial Cells in Preclinical Studies**

**Human Amniotic Epithelial Cells in Autoimmune Diseases**

Administration of hAECs has ameliorated immune-mediated organ damage in models of several autoimmune diseases. hAEC infusion in experimental autoimmune thyroiditis (EAT) and SLE maintained organ function, minimized inflammation and modified the immune balance (Tan B. et al., 2018). In EAT, hAECs reduced disease severity by inhibiting infiltration of inflammatory cells in thyroid glands, as well as suppressing Th17 responses. Moreover, hAECs improved the local cytokine environment in both EAT and SLE mice, by suppressing the levels of IFN-γ and enhancing TGF-β (Tan B. et al., 2018). In SLE mice specifically, hAEC administration promoted Tregs and decreased the levels of pathogenic autoantibodies (Tan B. et al., 2018).

Human amniotic epithelial cell also potently attenuated disease severity in experimental autoimmune encephalomyelitis (EAE), and a mouse model of multiple sclerosis (MS) (Liu et al., 2012). T cell and macrophage infiltration were significantly reduced by hAEC treatment. It was reported that hAECs utilized PGE2 and TGF-β for their immunosuppressive effects (Liu et al., 2012). In another relapsing model of MS, hAECs significantly ameliorated disease progression, while promoting Tregs and augmenting Th2 responses (McDonald et al., 2015).

In an autoimmune uveitis (EAU) rat model, hAECs treatment ameliorated the pathological progression of disease and maintained the retinal structural organization (Li et al., 2018).
Infiltration of macrophages and T cells was suppressed after hAEC administration. The stem cells regulated the balance of T cell subsets by decreasing Th17 cells and boosting IL-10-producing Tregs in the spleen and lymph nodes. Furthermore, hAEC treatment changed the ocular chemokine and cytokine environment in EAU rats, indicated by decreased levels of monocyte chemoattractant protein-1, IL-17 and IFN-γ levels, and enhancement of IL-10 (Li et al., 2018).

The immunomodulatory effect of hAECs has also been investigated in mice with autoimmune ovarian disease (AOD) (Zhang Q. et al., 2019). The outcomes showed that hAEC injection improved ovarian function. This was associated with a significant increase in the number of Tregs in the spleen of AOD mice (Zhang Q. et al., 2019).

Human Amniotic Epithelial Cells in Other Inflammatory Diseases

Transplantation of both hAEcs and their soluble factors have shown beneficial effects in animal models of hepatic fibrosis. hAEcs given to mice with induced liver fibrosis reduced hepatocyte apoptosis and decreased hepatic inflammation and fibrosis (Manuelpillai et al., 2010). This study showed that intact cells expressing human-specific markers, inner mitochondrial membrane protein and HLA-G were found in mouse liver 2 weeks following hAEC injection, without evidence of host rejection of the transplanted cells (Manuelpillai et al., 2010).

In another study using the same model, hAEcs significantly decreased liver fibrosis, in line with reduced hepatic levels of the pro-fibrogenic cytokine TGF-β1, increased expression of the anti-inflammatory mediator IL-10 and decreased hepatic T cell infiltration (Manuelpillai et al., 2012). Furthermore, hAEcs administration reduced hepatic macrophage numbers and induced an anti-inflammatory M2 macrophage phenotype (Manuelpillai et al., 2012).

In a rat model of ischemic stroke, hAEcs were administered by intracerebral injection, after which they reduced the infarct volume and cerebral apoptosis (Liu et al., 2008). A further study in mice found that hAEcs injected 1.5 h after stroke migrated to the ischemic brain and spleen, and limited functional deficit, infarct volume and brain inflammation (Evans et al., 2018).

In a rat model of intracerebral hemorrhage, Liang et al. reported that hAEcs reduced the levels of proinflammatory cytokines TNF-α and IL-1β in microglia culture medium (Liang et al., 2014). Similarly, in preterm fetal sheep models of brain injury, inflammation was reduced in fetuses that received hAEcs (Yawno et al., 2013).

Many studies have explored the immunomodulatory effect of hAEcs on lung fibrosis. A study using the bleomycin-induced model of pulmonary fibrosis showed that hAEcs can modulate the host inflammatory response, decrease fibrosis and preserve lung function (Murphy et al., 2011). hAEcs reduced expression of the pro-inflammatory cytokines TNF-α, IFN-γ and IL-6, and reduced inflammatory cell infiltration (Murphy et al., 2011). A further study using the same model demonstrated that hAEC administration significantly decreased macrophage recruitment into the lung and promoted the majority of alveolar macrophages toward the M2 phenotype (Tan et al., 2014). Moreover, hAEcs treatment increased Treg numbers in the injured lungs. It was found that hAEcs require Tregs to polarize macrophages toward an M2 phenotype and that hAEcs promote Treg cells via TGF-β (Tan et al., 2015).

A recent study in a mouse model of renal ischemia-reperfusion injury, which leads to acute kidney injury, has shown that systematically administered hAEcs effectively regulated the kidney immune response (Ren et al., 2020). hAEC infusion attenuated tubular cell death and endothelial necrosis and increased cell proliferation in the injured kidney. The stem cells reprogramed macrophages to shift from a pro-inflammatory M1 to anti-inflammatory M2 phenotype. In addition, hAEcs enhanced levels of IL-4 and IL-13 and decreased levels of TNFα and IFNγ, which in turn helped to minimize the inflammatory response.

Secretome Derived From Human Amniotic Epithelial Cells

The secreted factors (also named secretome) exist in the medium where the stem cells are cultured. Numerous studies on stem cell-derived secreted factors showed that these mediators alone, without the stem cell itself, affect the maturation, migration, polarization and function of immune cells, and thus influence the strength and duration of immune responses. The hAEC secretome contains metabolites, lipids, free nucleic acids, cytokines, growth factors and extracellular matrix proteins. These are all known to play crucial roles in cell-cell communication, acting proximally as well as systemically.

The application of cell-free therapy confers some advantages over stem-cell based applications. Usage of soluble factors bypasses a number of safety concerns potentially linked to the administration of living cell populations including post-transfer mal-transformation, embolism, and transmission of infections. In addition, hAEC secretome may be evaluated for safety, dosage and potency in an approach similar to that used for traditional medications (Vizoso et al., 2017). hAEC-conditioned media (CM) can be also manufactured, packaged, and transported more easily than hAEcs themselves (Kay et al., 2017). Therefore, stem cell-derived secretomes have a promising prospect to be used as pharmaceuticals for immune diseases. hAEC CM has shown beneficial effects in reducing a range of conditions by modulating immune responses including liver fibrosis, AOD, inflammatory bowel disease and diabetic wound healing (Hodge et al., 2014; Kuk et al., 2018; Zheng et al., 2018; Zhang Q. et al., 2019). In general, to prepare hAEC CM, hAEcs are cultured in chemically defined, serum-free ultraculture medium for 4 days at 37°C in a humidified chamber containing 5% CO2, after which conditioned media is harvested and secretome obtained by serial centrifugation (Alhomrani et al., 2017).

Human amniotic epithelial cells also mediate their effects by secreting exosomes. Exosomes are nano-sized biovesicles secreted by various cell types including stem cells under both normal and pathophysiological conditions. They are characterized by a diameter of 50–100 nm and a density of 1.09–1.18 g/mL (Zhang Y. et al., 2019). Exosome cargo is diverse
and a pool of exosomes can demonstrate all cellular elements including protein, nucleic acids and lipids (Spada, 2020). They represent a novel manner of intercellular communication, which may play a central role in many cellular processes such as the immune response, signal transduction and antigen presentation (Zhang Y. et al., 2019). Exosomes derived from hAECs produce potent immunomodulatory, anti-fibrotic and pro-regenerative effects and have been successfully used as a cell-free therapy in inflammatory conditions (Alhomrani et al., 2017; Tan J.L. et al., 2018). hAEC-derived exosomes can suppress many immune cells such as T cells, macrophages, and neutrophils in vitro. In addition, they have been highly effective in delivering protection from organ damage in a range of disease models including pulmonary and liver fibrosis, as well as acute kidney injury (Alhomrani et al., 2017; Tan J.L. et al., 2018; Ren et al., 2020). Thus, hAEC-derived soluble factors and exosomes may represent a potential cell-free therapy in crescentic GN.

Human Amniotic Epithelial Cells in Clinical Trials

Based on their immunomodulatory properties, the amniotic membrane and hAECs have been safely used for several years as a therapy for wounds and ocular injuries (Parmar et al., 2006; Jirsova and Jones, 2017). hAECs have also entered clinical trials as a treatment for several conditions including liver fibrosis, stroke and lung injury in premature babies (Table 1; Lim et al., 2017, 2018; Phan et al., 2018; Malhotra et al., 2020). So far, these trials have shown that hAECs are very safe and well-tolerated in humans.

CONCLUSION

In summary, there is a requirement for alternative, safer treatments for crescentic GN as the only effective therapies currently available are broadly immunosuppressive drugs which cause many serious side effects (mainly infections, cancer and cardiovascular problems) and patient deaths. hAECs, as well as hAEC-CM and hAEC-derived exosomes, exert a protective effect in models of various immune-driven conditions, with minimal side effects. They mediate their effects through multiple immunomodulatory and anti-inflammatory mechanisms, and their safety has been proven in clinical trials. Thus, hAEC-based therapy offers promise to be a safe, feasible and effective treatment for crescentic GN. This is due to their (i) ethical and speedy isolation from the placenta which results in an abundance of readily available cells, (ii) unique and selective immunosuppressive capacity, (iii) low immunogenicity, and (iv) ability to fight microbes and cancer and protect against cardiovascular conditions. If proven to be effective in preclinical models of crescentic GN, hAECs have the potential to change clinical practice in this disease and provide immense advantages to patients by alleviating their risk of death and complications from unwanted symptoms caused by the existing treatments.

AUTHOR CONTRIBUTIONS

AA, JO, and DO wrote the article and agreed with the final version of the manuscript. All authors contributed to the article and approved the submitted version.

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TABLE 1 | Completed and active clinical trials utilizing hAECs to treat immune-related diseases*.

| Category       | Registration number | Disease                                      | Phase | Age          | Country       |
|----------------|---------------------|----------------------------------------------|-------|--------------|---------------|
| Neurology      | ACTRN12618000076279 | Ischemic stroke                              | 1     | 18–85 years  | Australia     |
| Ophthalmology  | NCT00344708         | Corneal epithelial dystrophy                 | N/A   | 18–88 years  | United States |
| Pneumology     | ACTRN12614000174684 | Bronchopulmonary dysplasia                   | 1     | 36 weeks     | Australia     |
| Pneumology     | ACTRN1261800092029| Bronchopulmonary dysplasia, extremely preterm birth | 1     | 14–18 days   | Australia     |
| Orthopedics    | NCT03031509         | Non-union fracture                           | 1     | 18–80 years  | China         |
| Gynecology     | NCT02027412         | Premature ovarian failure                     | N/A   | 18–40 years  | China         |
| Others         | ACTRN12616000437460 | Cirrhosis, liver fibrosis                    | 1     | 18–70 years  | Australia     |
| Others         | ACTRN12618001883202 | Crohn’s disease, perianal fistulas            | N/A   | 18–80 years  | Australia     |

These data were collected from the clinical trial database (ClinicalTrials.gov and anzctr.org.au).
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