Keywords: Haematoxylin and Eosin; Periodic acid schiff; High iron diamine; Alcian blue

Introduction

Mucin is a high molecular weight glycoprotein that is synthesized, stored and secreted by the epithelial mucus cells, especially the goblet cells [1]. Mucins are expressed by various epithelial cell types that exist in relatively harsh environments [2]. Mucins' key characteristic is its ability to form gels; therefore they are a key component in most gel-like secretions, serving functions such as lubrication, cell signaling and forming chemical barriers [3]. Histochemically, the mucins are classified into neutral mucins and acidic mucins which include sulpho and sialo mucins. Many reviews on their histochemical classification and identification have been put forward to explain the intricacies and forming chemical barriers [3]. Histochemically, the mucins are classified into neutral mucins and acidic mucins which include sulpho and sialo mucins. Many reviews on their histochemical classification and identification have been put forward to explain the intricacies of mucins. The simplest, yet a lucid method to identify mucins by routine light microscopy were employed in the present study [4]. Their general structure and biochemical composition provides protection for the cell surface and specific molecular structures regulate the local microenvironment near the cell surface. In addition, mucins also communicate the information of the external environment to the epithelial cells via cellular signalling through membrane-anchored mucins [5,6]. Mucus provides a protective barrier against pathogens and toxins and contributes to the innate defensive system in mucosal immunity [7]. It seems that mucins play a role in the processes of tumour progression, invasion and metastasis and also in tumour cell survival and protection against the host immune response [8]. Increased mucin production occurs in many adenocarcinoma, including cancers of pancreas, lung, breast, ovary, colon and other tissues [9]. Mucinous tumours represent a subgroup of carcinomas exhibiting large amounts of mucous, grossly visible during microscopic examination. This morphological definition applies with some modifications to about 10–20 per cent of colonic, 5 per cent of breast, 3 per cent of ovarian, and 1 per cent of pancreatic carcinomas. The colonic mucinous carcinomas are most precisely defined in this group: according to the WHO definition, at least 50 per cent of the microscopically evaluated area in these tumours must be filled with mucus [10].

Materials and Methods

Paraffin embedded sections were prepared using automatic tissue processor, followed by preparation of paraffin block using our embedding station. The sections were stained with H&E stain to determine the histological diagnosis for selecting tissues with mucin production. Slides were stained with PAS, Alcian blue, high iron diamine-Alcian blue, Meyer’s mucicarmine and Alcian blue-PAS to demonstrate the mucin production and to identify types of mucins.

Results

We included 34 cases from different organs i.e colon (n=16), ovaries (n=13) and lung (n=5). Mucin histochemistry can effectively determine the types of mucins.

Conclusion

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(n=13) and lungs (n=5). They were divided into the following groups on the basis of mucin content in the whole section which was marked from ‘+’ to ‘+++’ as follows: mild (<20%) = +, moderate (20-40%) = ++, marked (40-60%) = ++++. Sections from normal colon were taken as controls. We observed the location of mucins which are extra-cellular or intra-cellular or both extra-cellular and intra-cellular mucins on the basis of H&E staining. We observed that 50% cases showed both extra-cellular and intra-cellular mucins that was dominant in our study and 38.2% cases showed extra-cellular mucins and only 11.8% showed intra-cellular mucins. After H&E staining we performed PAS stain for mucins. In PAS stain we could locate the mucins better than H&E staining but in this stain we did not differentiate the types of mucins. In PAS staining we observed that 70.60% cases showed mild mucin, 17.60% cases contained moderate mucin and 11.80% cases showed marked mucin in tissues. All the cases of mucinous adenocarcinoma of lung showed only a mild mucin but in ovary and colon we found different results from mild to marked mucins. After mucicarmine staining we differentiated the acidic mucins but not the other types of mucins. In 61.8% cases we observed mild mucin, 35.3% cases showed moderate mucin and 2.9% cases contained only marked mucin. However, after mucicarmine stain we could not differentiate the types of mucins. In Alcian blue staining 73.5% cases showed mild mucin, 8.8% cases showed moderate mucin, 8.8% cases showed marked mucin and 8.8% cases showed no mucin. The Alcian blue staining differentiated the acidic mucins from other types of mucins. On the other hand Alcian blue–PAS stain differentiated both neutral and acidic mucins. The Alcian blue–PAS staining showed 62.6% cases with mild acidic mucin, 20.6% cases with moderate acidic mucin in 5.9% cases marked mucin was present and in 2.9% cases we observed mild neutral mucin and in 2.9% cases we observed moderate neutral mucins. In High Iron Diamine-Alcian blue staining we could differentiate the sulphomucin in which we observed black brown colour and the sialomucin in which we found shades of blue colour. On the basis of mucin content in High Iron Diamine-Alcian blue staining 50.6% cases showed mild sulphomucin, 17.6% cases showed moderate sulphomucin, in 2.9% cases they showed marked sulphomucin, 5.9% cases contained mild sialomucin were present, 5.9% cases showed moderate sialomucin and in 17.6% cases there were both sulphomucin and sialomucin (Tables 1, 2 and Figures 1-7).

| Tissue organs | Acidic mucin | Neutal mucin |
|---------------|--------------|--------------|
|               | Mild | Moderate | Marked | Mild | Moderate | Marked |
| Colon         | 9    | 6        | 1      | 0    | 0        | 0      |
| Ovary         | 10   | 0        | 1      | 1    | 1        | 0      |
| Lung          | 4    | 1        | 0      | 0    | 0        | 0      |
| Total         | 23   | 7        | 2      | 1    | 1        | 0      |
| Percent%      | 67.6%| 20.6%    | 5.9%   | 2.9% | 2.9%     | 0      |

Mild ++, Moderate ++++, Marked++++

Table 1: Shows staining results of Alcian blue–PAS in different organs.

| Tissue organs | Sulphomucin | Sialomucin | Both (sulphomucin & sialomucin) |
|---------------|-------------|------------|-------------------------------|
|               | Mild | Moderate | Marked | Mild | Moderate | Marked |
| Colon         | 7    | 2        | 1      | 1    | 2        | 0      | 3 |
| Ovary         | 6    | 4        | 0      | 1    | 0        | 0      | 2 |
| Lung          | 4    | 0        | 0      | 0    | 0        | 0      | 1 |
| Total         | 17   | 6        | 1      | 2    | 2        | 0      | 6 |
| Percent%      | 50.6%| 17.6%    | 2.9%   | 5.9% | 5.9%     | 0      | 17.6% |

Mild ++, Moderate ++++, Marked++++

Table 2: shows staining results of high Iron Diamine-Alcian blue in different organs.

Discussion

Mucins are complex carbohydrates secreted by epithelial and connective tissue cells. Mucin glycoproteins are thought to play an important role in protecting the intestine from chemical or physical injury but the mechanisms of protection and the possible relationship...
Compared with nonmucinous carcinoma, histological variant that accounts for 5% to 15% of cases of primary others find no significant difference. Mucinous carcinoma is a mucinous neoplasm occurring more common in males and Western series. There may be a difference in this percentage prognosis. Mucinous adenocarcinomas account for 10–20% in most the use of staging as superior to histological grading as a criterion for as compared to tumours with no mucin content, the authors advocate the higher tumour grade than non-mucinous adenocarcinoma. In other studies mucinous adenocarcinoma showed a higher incidence of mucinous adenocarcinoma. The PAS technique is perhaps the most versatile and widely used of the techniques for the demonstration of glycoproteins, carbohydrates and mucins. The PAS technique is particularly sensitive to the detection of neutral mucins as well as acid mucins that contain significant quantities of sialic acid. The combined alcian blue AB/PAS technique is widely used for the detection and characterization of mucosubstances in tissue sections. The Alcian blue-PAS technique is a simple procedure and appears to differentiate sharply between acid mucins and neutral mucins. This might be of value in demonstrating small amounts of acid mucin. The high iron diamine-alcian blue staining technique, which permits the simultaneous recognition of sulfated and non-sulfated sialomucins. Different series define mucinous carcinoma as the presence of at least 50%–60% mucin in the extracellular matrix. In some cases there is an admixture of extracellular and intracellular mucin, the latter resulting in signet ring configuration. Although it is not clearly established some variations exist in the amount of extracellular mucin for the definition of mucinous colorectal carcinoma, which range between 50% and 80% recent studies however have reported a higher incidence of mucinous adenocarcinoma. In the present study the incidence of extracellular mucin and combined extracellular and intracellular mucins are similar to those reported. In some cases there is an admixture of extracellular and intracellular mucin, the latter resulting in signet ring configuration. Recent studies however have reported a higher incidence of mucinous adenocarcinoma. The PAS technique is perhaps the most versatile and widely used of the techniques for the demonstration of glycoproteins, carbohydrates and mucins. The high iron diamine-alcian blue sequence stains sulphomucins dark brown to black and sialomucins stain blue. Another study revealed that the HID/AB stain highlighted the predominance of acid mucins. The acid mucins being the predominant type (>90% of cases) They noticed that AB/PAS stain revealed the prevalence of acid mucins. The high-iron diamine-alcian blue sequence stains sulphomucins dark brown to black and sialomucins stain blue. Another study revealed that the HID/AB stain highlighted the predominance of sulphomucins contrary to these we found different results i.e the predominant mucin in our study was sulphomucin followed by both sulphomucin and sialomucin whereas sialomucin was observed only in 4 cases. However the number of cases in our study was rather small found abundant neutral and acidic mucins in approximately equal amounts in ovarian tissue. A slight predominance of sulphomucins was found. In our study we observed predominance of acid mucins over neutral mucins. In addition in these cases we observed sulphomucin predominating over the sialomucin. We also found that in mucinous adenocarcinoma of lung sulphomucins were present and in one case both sulphomucins and sialomucins were detected whereas we could not find any histochemical studies on mucinous adenocarcinoma. The PAS technique is perhaps the most versatile and widely used of the techniques for the demonstration of glycoproteins, carbohydrates and mucins. This might be of value in demonstrating small amounts of acid mucin. The high iron diamine-alcian blue staining technique, which permits the simultaneous recognition of sulfated and non-sulfated sialomucins.
Conclusion
Mucin histochemistry can effectively determine the presence and types of mucins. The PAS, mucicarmine and Alcian blue are localizing stains for mucins but could not differentiate the types of mucins. In mucin histochemistry we observed better contrast of alcian blue/PAS stains for mucins but could not differentiate the types of mucins. The PAS, mucicarmine and Alcian blue are localizing carbohydrates. J Histochem Cytochem 13: 211-234.

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Mucinous adenocarcinomas need to be further investigated at molecular level to elucidate the biological significance of mucin in carcinoma.

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References
1. Kim YS, Gum JR Jr, Byrd JC, Toribara NW (1991) The structure of human intestinal apomucins. Am Rev Respir Dis 144: S10-14.
2. Forstner JF (1978) Intestinal mucins in health and disease. Digestion 17: 234-263.
3. Marin F, Luquet G, Marie B, Medakovic D (2008) Moltuscan shell proteins: primary structure, origin, and evolution. Curr Top Dev Biol 80: 209-276.
4. Stanley S, Rapheal SS (1997) Staining of Carbohydrates and Connective Tissue around substance: Fibrin and Amyloid. In: Lynch Medical Laboratory Technology. 3rd edn. Philadelphia: W.B. Saunders Company: 963-979.
5. Moniaux N, Escande F, Porcher N, Aubert JP, Batra SK (2001) Structural organization and classification of the human mucin genes. Front Biosci 6: D1192-1206.
6. Hollingsworth MA, Swanson BJ (2004) Mucins in cancer: protection and control of the cell surface. Nat Rev Cancer 4: 45-60.
7. Forstner JF (1978) Intestinal mucins in health and disease. Digestion 17: 234-263.
8. Marin F, Luquet G, Marie B, Medakovic D (2008) Moltuscan shell proteins: primary structure, origin, and evolution. Curr Top Dev Biol 80: 209-276.
9. Stanley S, Rapheal SS (1997) Staining of Carbohydrates and Connective Tissue around substance: Fibrin and Amyloid. In: Lynch Medical Laboratory Technology. 3rd edn. Philadelphia: W.B. Saunders Company: 963-979.
10. Moniaux N, Escande F, Porcher N, Aubert JP, Batra SK (2001) Structural organization and classification of the human mucin genes. Front Biosci 6: D1192-1206.
11. Hollingsworth MA, Swanson BJ (2004) Mucins in cancer: protection and control of the cell surface. Nat Rev Cancer 4: 45-60.
12. Forstner JF (1978) Intestinal mucins in health and disease. Digestion 17: 234-263.
13. Marin F, Luquet G, Marie B, Medakovic D (2008) Moltuscan shell proteins: primary structure, origin, and evolution. Curr Top Dev Biol 80: 209-276.
14. Stanley S, Rapheal SS (1997) Staining of Carbohydrates and Connective Tissue around substance: Fibrin and Amyloid. In: Lynch Medical Laboratory Technology. 3rd edn. Philadelphia: W.B. Saunders Company: 963-979.
15. Moniaux N, Escande F, Porcher N, Aubert JP, Batra SK (2001) Structural organization and classification of the human mucin genes. Front Biosci 6: D1192-1206.
16. Hollingsworth MA, Swanson BJ (2004) Mucins in cancer: protection and control of the cell surface. Nat Rev Cancer 4: 45-60.
17. Forstner JF (1978) Intestinal mucins in health and disease. Digestion 17: 234-263.
18. Marin F, Luquet G, Marie B, Medakovic D (2008) Moltuscan shell proteins: primary structure, origin, and evolution. Curr Top Dev Biol 80: 209-276.
19. Stanley S, Rapheal SS (1997) Staining of Carbohydrates and Connective Tissue around substance: Fibrin and Amyloid. In: Lynch Medical Laboratory Technology. 3rd edn. Philadelphia: W.B. Saunders Company: 963-979.
20. Moniaux N, Escande F, Porcher N, Aubert JP, Batra SK (2001) Structural organization and classification of the human mucin genes. Front Biosci 6: D1192-1206.
21. Hollingsworth MA, Swanson BJ (2004) Mucins in cancer: protection and control of the cell surface. Nat Rev Cancer 4: 45-60.