Mental Disorders Linked to Crosstalk between The Gut Microbiome and The Brain

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INTRODUCTION

The microbiome is the sum of “microbe” and “biome,” which is defined as microorganisms coexisting with host organisms and their theater of activity, including genetic information. The human microbiome is found in various areas of the body, such as the skin, placenta, uterus, vagina, oral cavity, and gut. Approximately 95% of the human microbiome is found in the digestive system, including the large intestine \cite{1}. This is called the gut microbiome. Recent studies have shown that the gut microbiome is important for maintaining brain homeostasis in host organisms. For example, compositional changes in the gut microbiome are closely associated with several mental disorders, which can be improved by controlling the gut microbiome composition \cite{2-4}.

Classically, mental disorders are known to be caused by structural abnormalities or dysfunction of the brain. For example, structural, molecular, or physiological changes in the mesolimbic dopamine system, which is primarily involved in rewarding and motivational behaviors, are the main pathophysiological mechanisms of depression \cite{5}. For more than a decade, however, accumulating evidence suggests that another participant, the gut microbiome, is involved in maintaining normal mental health and the pathogenesis of mental disorders.

In this review, we will focus on the mechanism by which the gut microbiome affects mental health and diseases. Furthermore, we discuss the potential use of probiotics as therapeutic agents for psychiatric disorders.

Key words: Gastrointestinal microbiome, Brain, Mental disorders, Probiotics

Often called the second brain, the gut communicates extensively with the brain and vice versa. The conversation between these two organs affects a variety of physiological mechanisms that are associated with our mental health. Over the past decade, a growing body of evidence has suggested that the gut microbiome builds a unique ecosystem inside the gastrointestinal tract to maintain the homeostasis and that compositional changes in the gut microbiome are highly correlated with several mental disorders. There are ongoing efforts to treat or prevent mental disorders by regulating the gut microbiome using probiotics. These attempts are based on the seminal findings that probiotics can control the gut microbiome and affect mental conditions. However, some issues have yet to be conclusively addressed, especially the causality between the gut microbiome and mental disorders. In this review, we focus on the mechanisms by which the gut microbiome affects mental health and diseases. Furthermore, we discuss the potential use of probiotics as therapeutic agents for psychiatric disorders.

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mechanism by which they improve mental disorders.

**GUT MICROBIOME AND MENTAL DISORDERS**

Recent studies have revealed that controlling the gut microbiomes of the experimental animals confirmed various behavioral changes, including social, emotional and cognitive functions. This suggests that the gut microbiome affects the pathogenesis of many mental disorders. In this section, we summarize the changes in the gut microbiome found in several kinds of mental disorders.

**Major depressive disorder (MDD)**

Major depressive disorder (MDD), commonly known as depression, is a common but serious mental disorder with low mood, feelings of hopelessness, and anxiety. The cause of MDD is unclear, but it is widely accepted that severe and/or prolonged stress is the leading cause of this disorder. Stress affects many physiological states, such as the synthesis or secretion of stress hormones including corticosterone or neurotransmitters related to mood, including dopamine and serotonin. Changes in the structure or function of the brain reward system or immune system are also influenced by stress [5]. Collectively, these changes are thought to cause MDD.

Many preclinical and clinical studies have reported that compositional changes in the gut microbiome are highly linked to these disorders. Germ-free (GF) animals usually exhibit antidepressant-like behaviors [6-8], but antibiotic-induced gut dysbiosis elicited depression-like behaviors [9, 10]. Compositional changes in the gut microbiome were observed in several animal models of stress-induced depression. Chronic social defeat stress (CSDS) increased Proteobacteria and Verrucomicrobia, but decreased Chloroflexi in the gut [11]. Another study using the CSDS protocol reported that Bifidobacterium was increased in resilient, but not susceptible, animals. Of note, the depressive phenotypes induced by this stress protocol were recovered by administrating these bacteria [12]. Different stress protocol – chronic unpredictable stress (CUS) – also altered the composition of microorganisms in the gut. For example, Lactobacillus was decreased in CUS-induced depressive animals, and the treatment of probiotic L. reuteri rescued depressive behaviors induced by the CUS protocol [13]. Recently, it was reported that delivering the gut microbiome from stressed animals to healthy controls induced depression-like phenotypes in recipient animals such as despair-like behavior and decreased neurogenesis in the hippocampus [14].

Conflicting results have also been reported in human studies. One group reported that the diversity of the gut microbiome was higher in patients with depression than in the healthy group [15]. However, other reports showed that this diversity was not observed in patients with MDD [16, 17]. Some clinical studies showed that Bifidobacterium, Lactobacillus and Bacteroidetes were reduced [18, 19], but Actinobacteria, Firmicutes, and Faecalibacterium were increased in patients with MDD compared to healthy controls [18, 20-22]. We summarized the main findings that report gut dysbiosis in patients with depression in Table 1. These results suggest a high correlation between gut dysbiosis and depression in animal and human studies. However, more detailed research is needed on the causal relationship between changes in the gut microbiome and depression for treating or preventing this disorder by regulating the gut microbiome.

**Anxiety disorder**

Anxiety disorder is a mental disorder in which people feel abnormal and excessive anxiety that makes daily life difficult. It is categorized by symptoms such as generalized anxiety disorder, phobia, panic disorder and post-traumatic stress disorder (PTSD). Anxiety symptoms also accompany many chronic diseases such as cancer, chronic fatigue syndrome (CFS), and irritable bowel syndrome. Anxiety and the gut microbiome are closely related. GF and antibiotic-treated animals usually showed anxiolytic behaviors, such as increased time in the center zone during OFT or in open arms during EPM [6, 23-30]. Some clinical reports have shown that the administration of probiotics improved anxiety symptoms in healthy humans [31-33] or patients with CFS [34]. These results suggest that anxiety disorders can be improved through the control of the gut microbiome using probiotics, but further research to reveal the detailed mechanisms is needed.

**Autism spectrum disorder (ASD)**

Autism spectrum disorder (ASD) is a neurodevelopmental disease characterized by repetitive behaviors and impairment of social or cognitive function. Some human studies have reported the relationship between the gut microbiome and ASD. Many patients with ASD have gut trouble, such as bloating and diarrhea, and the treatment of vancomycin, broad-spectrum antibiotics, improved behavioral deficits in patients with ASD [35, 36]. Several studies have reported that patients with ASD have a different composition of the gut microbiome compared to healthy controls: levels of the advantageous bacteria Bifidobacterium were decreased, but the pathogenic bacteria Desulfovibrio and Clostridia were increased in the gut of patients with ASD [37]. It was also reported that Firmicutes/Bacteroidetes ratio is increased in patients with ASD [38-41]. Table 2 shows the major changes in the gut microbiome of patients with ASD.

Sometimes, deficits in sociability, the most representative behavioral change in ASD, were observed in GF animals [25, 42-45]
and in antibiotic-treated animals [10, 30]. Many animal models of autism have shown gut dysbiosis. For example, shank 3 (autism candidate gene) transgenic mice had differences in the composition of the gut microbiome with reduced Veillonella, Prevotella and Lactobacillus. Autistic behavioral phenotypes observed in shank 3 transgenic mice were ameliorated by the administration of L. reuteri [46, 47]. In utero exposure to valproic acid (VPA), another animal model for ASD, also induced gut dysbiosys similarly observed in ASD patients [48].

### Table 1. Gut dysbiosis observed in human patients with MDD

| Gut dysbiosis | References | Etc. |
|---------------|------------|------|
| ↓ Bacteroides | 15 A-MDD   |      |
| Dialister     |            |      |
| Faecalibacterium |        |      |
| Prevotella    |            |      |
| ↑ Alistipes   |            |      |
| Blautia       |            |      |
| Clostridium XIX |        |      |
| Lachnocpracea incertae sedis | |      |
| Megamonas     |            |      |
| Parabacteroides |          |      |
| Parasutterella |          |      |
| Phascolarctobacterium | |      |
| Oscillibacter |            |      |
| ↓ Escherichia/Shigella | R-MDD |      |
| Oscillibacter |            |      |
| Dialister     |            |      |
| Faecalibacterium |        |      |
| Prevotella    |            |      |
| Ruminococcus  |            |      |
| ↑ Alistipes   |            |      |
| Bacteroides   |            |      |
| Parabacteroides |          |      |
| Phascolarctobacterium | |      |
| Roseburia     |            |      |
| ↓ Howardella  | 17 MDD-F   |      |
| Sutterella    |            |      |
| Pyramydobacter |          |      |
| ↑ Actinomyces  |            |      |
| Biidobacterium |            |      |
| Asaccharobacter |          |      |
| Atopobium     |            |      |
| Eggertella    |            |      |
| Gordonibacter  |          |      |
| Olsenella     |            |      |
| Eubacterium   |            |      |
| Anaerostipes  |            |      |
| Blautia       |            |      |
| Roseburia     |            |      |
| Faecalibacterium |        |      |
| Desulfovibrio |            |      |
| ↓ Anaerovorax  | MDD-M      |      |
| Gordonibacter  |          |      |
| Pyramydobacter |          |      |
| ↑ Bacteroides  |            |      |
| Erysipelotrichaceae incertae sedis | |      |
| Veillonella   |            |      |
| Atopobium     |            |      |
| ↑ phylum Firmicutes | 20 |      |
| ↓ phylum Bacterodetes | |      |
| ↑ Prevotella   |            |      |
| Klebsiella    |            |      |
| Streptococcus |            |      |
| Clostridium XI |           |      |
| a ↓, decreased compared to control; ↑, increased compared to control. |
| b A-MDD, active-major depressive disorder; R-MDD, responded-major depressive disorder; MDD-F, female patients with MDD; MDD-M, male patients with MDD. |
| c Duplicate bacteria are indicated in bold. |

### Table 2. Gut dysbiosis observed in other human mental disorders

| Disorders                  | Gut dysbiosis | References |
|----------------------------|---------------|------------|
| Autism spectrum disorder (ASD) | ↓ phylum Bacteroidetes | 40 |
|                           | ↑ phylum Firmicutes |            |
|                           | Streptococcus |            |
|                           | Veillonella |            |
|                           | Escherichia |            |
|                           | Actinomyces |            |
|                           | Parvimonas |            |
|                           | Bulleidia |            |
|                           | Peptoniphilus |        |
|                           | ↓ Akkermansia | 41 |
|                           | Bacteroides |            |
|                           | Biidobacterium |        |
|                           | Parabacteroides |        |
|                           | Enterococcus |            |
|                           | Escherichia coli |        |
|                           | Bacteroides |            |
|                           | Biidobacterium |        |
|                           | Faecalibacterium |        |
|                           | Lactobacillus |        |
| Alcoholism                 | ↑ phylum Proteobacteria | 52 |
|                           | Sutterella |            |
|                           | Holdemania |            |
|                           | Clostridium |            |
|                           | Faecalibacterium |        |
|                           | ↓ phylum Firmicutes | 69 |
|                           | phylum Actinobacteria |        |
|                           | Biidobacterium |        |
|                           | Dialister |            |
|                           | Clostridium |            |
|                           | Turicibacter |            |
|                           | Adlercreutzia |        |
|                           | ↑ phylum Bacteroidetes |        |
|                           | Blautia |            |
|                           | Bacteroides |            |
|                           | Alistipes |            |
|                           | Phascolactobacterium |        |
|                           | Bilophila |            |
|                           | Gemella |            |
| Alzheimer’s disease (AD)    | ↓ phylum Firmicutes |        |
|                           | Biidobacterium |        |
|                           | Clostridium |            |
|                           | Turicibacter |            |
|                           | Adlercreutzia |        |
|                           | ↑ phylum Bacteroidetes |        |
|                           | Blautia |            |
|                           | Bacteroides |            |
|                           | Alistipes |            |
|                           | Phascolactobacterium |        |
|                           | Bilophila |            |
|                           | Gemella |            |
| a ↓, decreased compared to control; ↑, increased compared to control. |

Addiction

Addiction is a chronic brain disease defined as compulsive substance use despite negative consequences. Recent studies have
revealed that the gut microbiome is involved in this disorder. Alcohol, the most common addictive substance, changes the gut microbiome both in humans and in experimental animals [49–52]. We summarized the main findings that report gut dysbiosis in patients with alcoholism in Table 2.

Rats susceptible to compulsive alcohol-seeking behavior showed increased Firmicutes and decreased Actinobacteria [53]. Alcoholic hepatitis was recovered by the treatment of L. rhamnosus GG [54]. Several animal studies have shown that gut dysbiosis is also induced by opioids [55, 56], cannabis [57], and tobacco [58, 59]. Interestingly, gut dysbiosis caused by the treatment of antibiotics increased conditioned place preference and behavioral sensitization induced by low (5 mg/kg), but not high (10 mg/kg), concentrations of cocaine [60].

**Alzheimer’s disease (AD)**

Alzheimer’s disease (AD) is a typical neurodegenerative disease that induces dementia. It progressively worsens cognitive function, including memory. Although the exact mechanisms and causes of this disease are not fully known, it is thought that various complex pathological mechanisms, including the accumulation of beta-amyloid (Aβ), hyperphosphorylation of the tau protein, and immune responses are involved [61]. Some reports indicate that microorganisms are involved in the pathogenesis of AD. For example, amyloid, one of the main molecules related to AD, was generated from the gut microbiome [62–64], and acted as an antimicrobial peptide [65, 66]. Other studies have shown that Aβ pathology is modulated by the gut microbiome. APP/PS1 transgenic mice, an AD mouse model with human transgenes for APP bearing the Swedish mutation and L166P-mutated PSEN1, lacking gut microbiome showed a decreased Aβ pathology compared to the same transgenic mice with a normal gut microbiome [67]. Similarly, antibiotic treatment in young APP/PS1 transgenic mice resulted in decreased Aβ deposition [68]. Clinically, alterations in the composition of the gut microbiome in patients with AD, such as a reduction of Firmicutes and an enhancement of Bacteroidetes, have been reported [69]. Table 2 shows the major changes in the gut microbiome of patients with AD.

Of note, probiotics can affect AD pathology. A probiotic cocktail decreased oxidative stress, one of AD pathologies, in a triple transgenic mouse model (3xTg), another AD mouse model containing three mutations related to familial AD (APP Swedish, MAPT P301L, and PSEN1 M146V) [70]. Another preclinical study revealed that cognitive impairment in AD animals induced by direct injection of Aβ in the hippocampus was recovered by another probiotic mixture with different combinations [71]. Cognitive deficits in patients with AD were also recovered by administering a mixture of probiotics, including L. acidophilus, L. casei, L. fermentum and B. bifidum [72]. Not only the probiotic treatment but also FMT ameliorated AD pathogenesis. FMT from healthy animals to AD animal models ameliorated the cognitive deficits and immune responses of recipient AD animals [73]. More studies are needed to reveal how the regulation of the gut microbiome improves AD.

**COMMUNICATION BETWEEN GUT MICROBIOME AND THE BRAIN**

The gut, sometimes called the ‘second brain,’ interacts with the brain despite the distance between these two organs. While the gut microbiome exists in the gastrointestinal (GI) tract, it is involved in various normal behaviors and mental disorders. This raises the question of how the gut microbiome, gut and brain communicate?

**Enteric nervous system (ENS)**

In contrast to other peripheral organs, the GI tract has an intrinsic nervous system with numerous neurons (approximately 500 million cells in humans) and glial cells called the enteric nervous system (ENS). This system is a mesh-like neural network distributed from the esophagus to the anus. The ENS is composed of two ganglionated plexi; the myenteric and submucosal plexuses. They are connected by the central nervous system (CNS) and directly control the movement of the GI tract and the secretion of gut hormones and gastric acid [74, 75]. ENS neurons produce various neurotransmitters, including dopamine and serotonin, to communicate with other neurons inside or outside this system [76, 77]. Communication by ENS neurotransmitters is also affected by the gut microbiome and its metabolites because they are anatomically adjacent. For example, the gut microbiome affects ENS function via the recognition of microbial molecules by toll-like receptors (TLRs) [78].

**Vagus nerve**

The vagus nerve, a part of the parasympathetic nervous system that regulates involuntary body functions such as digestion, respiration, and heartbeat, is the tenth cranial nerve that connects the gut and the brain. It gathers information from various organs such as the GI tract and respiratory and cardiovascular systems. For example, vagal afferents sense physical changes in the gut, such as stretch and tension, or detect metabolites from the gut microbiome or gut hormones. Their cell bodies are located in the nodose ganglia and are connected to many nuclei of the brainstem. In particular, vagal fibers connected to the GI tract are linked to the nucleus of the solitary tract (NTS). Excitatory synapses from the NTS are connected to the paraventricular hypothalamic nuclei.
(PVN) and parabrachial nucleus (PBN). These areas project to the midbrain, including the substantia nigra pars compacta (SNc) and ventral tegmental area (VTA), two major brain regions to make dopamine. Thus, manipulating the vagus nerve may regulate motivational and reward behaviors by affecting dopamine release to the striatum, medial prefrontal cortex (mPFC), or other brain regions [79-81].

Two therapeutic methods, vagotomy and vagus nerve stimulation, provide insight into the role of the vagus nerve in communicating between the gut and the brain. Interestingly, damage to the vagus nerve caused by gastrectomy to treat peptic ulcers increases some psychiatric disorders [82]. Vagotomy in adult mice reduces brain-derived neurotrophic factor (BDNF) expression and neurogenesis in the hippocampus [83]. Comparably, anxiolytic phenotypes were observed in vagotomized rats [84]. Vagus nerve stimulation is usually used to treat depression and chronic pain [85, 86]. In contrast to vagotomy in rodents, the electrical stimulation of the vagus nerve increases cell proliferation and BDNF expression in the hippocampus [87]. These results suggest that communication between the gut and brain through the vagus nerve is important for maintaining brain homeostasis.

**Spinal cord**

The gut and brain are physically connected through not only the vagus nerve but also the spinal cord. Although the vagus nerve relays the physiological information from the gut, it is well-established that noxious information, including pain and injury, is conveyed through the spinal cord. For example, vagotomy, but not spinal transection, inhibits the delivery of information about nutrition sensing in the gut or mechanical stretch of the stomach [88, 89]. However, visceral pain occurs when the sensory neurons in the gut are sensitized, and the ascending spinal neurons are subsequently over-activated [90, 91]. In addition, spinal cord injury results in GI dysfunction, including enhanced intestinal permeability and gut dysbiosis, microbial imbalance or maladaptation on or inside the body. The imbalance of the gut microbiome induced by spinal cord injury also affects neurological function [92, 93]. These results indicate that the spinal cord is important for health and behaviors through monitoring and controlling the gut.

**Immune system**

Various microorganisms that exist in the gut do not directly affect the GI tract because there is a physical barrier between these two systems. Viscous mucus is secreted from goblet cells in the gut and inhibits direct contact between the gut microbiome and gut tissues. Despite this physical barrier, many immune cells located in the gut can interact with gut bacteria. Other cell types, such as enterocytes that synthesize and secrete cytokines and chemokines, also exist in the gut epithelium [94]. In addition to goblet cells releasing mucus in the gut, enteroendocrine cells in the GI tract produce various neuroendocrine molecules such as ghrelin and somatostatin [95]. Antigens present in the gut bacteria, such as peptidoglycans and polysaccharides, are recognized by epithelial pattern recognition receptors, and the immune system is activated [96]. On the other hand, hormones from the brain, such as cortisol secreted from the hypothalamus-pituitary-adrenal (HPA) axis, affect gut immune function and gut permeability [97].

**Neurotransmitters**

Among the byproducts produced by the gut microbiome, neurotransmitters act directly on the nervous system. Serotonin (also called 5-hydroxytryptophan or 5-HT), a neurotransmitter known as a happiness hormone because of its major function in experiencing happiness by regulating mood, is mainly synthesized in enterochromaffin cells in the gut. Intriguingly, the production of 5-HT is affected by the gut microbiome. For example, *Clostridium perfringens*, one of the gut microbiomes in humans and rodents, is involved in the synthesis of 5-HT in the gut [76, 77, 98]. The gut microbiome also affects the production of another neurotransmitter, dopamine, or its precursor molecule L-DOPA (L-3,4-dihydroxyphenylalanine), also known as levodopa. Tyrosine decarboxylases from *Enterococcus faecalis* found in the GI tract of mammals convert L-DOPA to dopamine in the intestine, regulating the level of L-DOPA in the serum [99, 100]. Oral administration of another gut microbiome, *Lactobacillus reuteri*, in adult mice increased serum dopamine [101]. Moreover, a major inhibitory neurotransmitter, gamma-aminobutyric acid (GABA), is synthesized by lactic acid bacteria [102]. These neurotransmitters produced in the gut are thought to be utilized for communicating with the brain, but a detailed mechanism remains unknown. However, it is certain that serotonin and dopamine produced in the gut cannot transport to the brain directly because these molecules do not pass the blood-brain barrier. These neurotransmitters are thought to act directly on neurons or other cell types in the GI tract. Another possibility is that the precursors of these neurotransmitters, such as tryptophan or L-DOPA, produced in the gut are delivered to the brain for conversion into neurotransmitters. More research is needed on the mechanism by which neurotransmitters produced in the gut regulate brain function and affect behaviors.

We summarized previously described pathways to communicate between the gut and the brain and the relationship of gut dysbiosis and mental disorders in Fig. 1.

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HOW TO STUDY THE ROLE OF THE GUT MICROBIOME ON MENTAL HEALTH AND DISEASES

Many recent studies have revealed a high correlation between human mental disorders and changes in the gut microbiome. However, this correlation does not prove a causal relationship in the onset of mental disorders following changes in the gut microbiome. To explain this causality, it is necessary to observe whether behavioral changes are induced by artificially controlling gut microbiomes using experimental animal models. Table 3 shows several types of behavioral changes induced by the alteration of the gut microbiome, as described below.

Germ free (GF) animals

The surest way to identify the role of an element is to observe what changes occur when the element is removed. We can know how the gut microbiome affects our mental health and behavior through GF animals who do not have microorganisms in or on their bodies. GF animals display different physiology and behavior compared to animals containing a normal microbiome. Many groups reported that the basal locomotor activity of GF animals was similar to that of control animals [23, 26, 28, 103; but see ref. 24, 25, 27]. Anxiolytic behaviors such as increased time and movement in the center zone during the open field test (OFT) and increased time in the open arm during the elevated plus maze (EPM) test were also observed in GF animals [6, 23-28; but see ref. 42, 103]. Social behavior was also decreased in GF animals [25, 42-45; but see ref. 27]. Some anti-depressive behavioral phenotypes were also observed in GF animals; GF mice showed reduced immobile

Table 3. Various behavioral changes observed in GF animals or by antibiotics-treated gut dysbiosis

| Behaviors        | Changes | Animal models | Behavioral methods (references) |
|------------------|---------|---------------|---------------------------------|
| Basal locomotion | ↔       | GF            | Open field test (23, 26, 28, 28, 103) |
|                  |         | ABX           | Open field test (9, 30, 105)    |
| Anxiety          | ↑       | GF            | Open field test (24, 25*, 27)   |
|                  | ↓       | GF            | Open field test (23, 24, 27, 28) |
|                  |         | Noveltly suppressed feeding test (23) |
|                  |         | ABX           | Light-dark test (6, 24)         |
|                  |         |               | Elevated plus maze test (25, 26) |
| Depression       | ↓       | GF            | Tail suspension test (6, 8)     |
|                  |         | ABX           | Forced swim test (7, 8)         |
|                  |         |               | Sucrose preference test (8)     |
| Social interaction| ↓     | GF            | 3-chamber social interaction test (25, 43, 44, 45) |
|                  |         | ABX           | Social interaction test (42)    |
|                  | ↑       | GF            | 3-chamber social interaction test (10**, 30) |
| Cognition        | ↓       | GF            | Nobel object recognition test (25, 104) |
|                  |         | ABX           | Nobel object recognition test (29, 105, 106) |
|                  |         |               | Morris water maze test (9)     |
|                  | ↔       | ABX           | Barnes maze test (105)         |

* ↔, no changes; ↑, increase; ↓, decrease; GF, germ-free animals; ABX, antibiotics.
*b female only; **normal sociability, but reduced social novelty.
time during behavioral despair tests such as the forced swimming test (FST) or tail suspension test (TST), and increased sucrose preference, suggesting hedonic behavior [6-8]. In addition, cognitive impairment in GF animals has been observed in some studies. Recognition memory and working memory were impaired in GF mice, as shown by performance in the novel object recognition test and T-maze spontaneous alternation test [25, 104]. Taken together, the gut microbiome affects many aspects of behaviors such as locomotive, emotional, social and cognitive behaviors, but not in a consistent manner.

**Antibiotics-induced gut dysbiosis**

Antibiotics are usually used to treat bacterial infections by inhibiting or killing the growth of bacteria. Antibiotic-induced gut dysbiosis is another good model to study the role of the gut microbiome in mental health and disease. It has the advantage of being able to control some microbes by using selective antibiotics or their combination at specific times. Gut microbiome studies through the use of antibiotics may explain abnormalities in brain and behavior caused by antibiotic prescriptions observed in humans. Many behavioral changes were observed in antibiotic-treated animals, similar to those observed in GF animals. Antibiotic-treated animals usually showed anxiolytic behaviors such as increased time in the light chamber during the light-dark test or increased time in the open arms during EPM [29, 30; but see ref 9, 105]. Impairment of social behavior was also observed in antibiotic-treated animals [10, 30]. Gut dysbiosis by antibiotics induced cognitive impairment: object recognition memory was reduced in antibiotic-treated animals compared to control [29, 105, 106]. In contrast to GF animals showing anti-depressive behaviors, antibiotic-treated animals showed depression-like behaviors such as increased immobility time during TST or FST [9, 10]. On the other hand, locomotion was not changed by antibiotics [9, 30, 105]. These results indicate that gut dysbiosys induced by administering antibiotics induce several behavioral changes similar to those observed in GF animals.

**Fecal microbiota transplant (FMT)**

Fecal microbiota transplant (FMT), as known as stool transplant, is a process of transferring the gut microbiome from one individual to another. Known as an effective treatment for Clostridiodoses difficile infection (CDI) [107], it is also used to study the function of the gut microbiome in experimental animals. Some studies have shown that FMT from depressed patients to normal experimental animals delivered depressive-like physiological and behavioral phenotypes [18, 108]. In addition, transplanting the gut microbiome from obese mice chronically consuming a high-fat diet to normal mice induced anxiety-like behavior, memory impairment, and increased body fat [109, 110]. These results suggest that the gut microbiome contains information about the mental health of the host. Decoding information from the gut microbiome would be helpful in understanding the etiology and pathophysiology of psychiatric disorders.

**PROBIOTICS FOR THE PREVENTION OR THE TREATMENT OF MENTAL DISORDERS**

Probiotics are living microorganisms that provide benefits to our body when ingested. Among them, bacteria that are beneficial to mental health are called psychobiotics. We previously confirmed that symptoms of various mental disorders were improved by several kinds of bacteria or their combinations [12, 13, 31-34, 70-72]. *Bifidobacteria* and *Lactobacillus* families are the most commonly used probiotics for many disorders. Some strains of these families make GABA and serotonin, which are involved in regulating cognition and emotion and are the potential target mechanisms to improve many symptoms of mental disorders [111, 112]. The effects of *Bifidobacteria* and *Lactobacillus* have been confirmed in some animal experiments. Stress-induced depression-like behaviors observed in experimental animals were recovered by the administration of these bacteria [12, 13]. In addition, *L. reuteri* ameliorated the impaired social behavior observed in ASD animal models [46, 47]. Anxiety- and depression-like behaviors observed in early life stressed (ELS) animals were recovered by the treatment of *L. plantarum* PS128, which reduced inflammation and corticosterone and increased dopamine and serotonin [113, 114]. Another study reported that behavioral changes induced by chronic restraint stress were rescued by *L. helveticus* NS8, which increased serotonin and BDNF in the hippocampus [115]. The effectiveness of probiotics has been proven not only in animal research, but also in human studies. For example, chronic administration of *B. longum* 1714 relieved stress and increased cognitive function in healthy humans [116]. These two families also recovered cognitive deficits in AD animal models [70, 71]. The representative probiotics used to treat or to ameliorate mental disorders are summarized in Table 4. Various bacteria can be used as probiotics. However, it is unknown how they positively and precisely affect people’s health and treat several diseases. To use probiotics more effectively, it is necessary to know what kinds of metabolites or byproducts of probiotics are produced and how they work.

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CONCLUSIONS

Recent studies have revealed the relationship between the gut microbiome and mental health conditions. The GI tract, which contains the gut microbiome, communicates with the brain via several pathways such as the vagus nerve, neurotransmitters, and immune system. Through this communication, the gut microbiome plays an important role in the maintenance of brain homeostasis. Studies using GF animals, antibiotic-induced gut dysbiosis, and FMT provide critical information about the role of the gut microbiome in several emotional and cognitive behaviors in psychiatric conditions. Using these methodologies, the correlation between various mental disorders and the gut microbiome has been clarified. In particular, there are ongoing attempts to treat or

| Disorders or models | Subjects | Probiotics | Suggested mechanisms | Ref. |
|--------------------|----------|------------|----------------------|------|
| Major depressive disorder (MDD) | Human | Lactobacillus helveticus R0052 and Bifidobacterium longum R0175 | Regulating tryptophan-kynurenine pathway metabolites | 12 |
| | Mouse (CSDS) | Lactobacillus reuteri | Reducing serum corticosterone and inflammatory cytokines, increasing anti-inflammatory cytokines, modulating DA and 5-TH in PFC | 114 |
| | Mouse (CUS) | Lactobacillus plantarum PS128 | | |
| | Mouse (ELS) | Lactobacillus plantarum PS128 | | |
| Rat (CRS) | Lactobacillus helveticus NS8 | Reducing plasma stress hormones, regulating cytokines, and increasing hippocampal BDNF | 115 |
| Anxiety | Human | Lactobacillus helveticus R0052 and Bifidobacterium longum R0175 | Reducing plasma pro-inflammatory cytokines and increasing plasma anti-inflammatory cytokines, enhancing the serotonin pathway | 31 |
| | Human | Streptococcus thermophilus, Lactobacillus bulgaricus, Lactococcus lactis, Lactobacillus acidophilus, Streptococcus thermophilus, Lactobacillus plantarum, Bifidobacterium lactis, Lactobacillus reuteri | | |
| | Human | Lactobacillus plantarum DR7 | Reducing serum corticosterone and inflammatory cytokines, increasing anti-inflammatory cytokines, modulating DA and 5-TH in PFC | 114 |
| | Human (patients with chronic fatigue syndrome [CFS]) | Lactobacillus casei strain Shirota (LcS) | Reducing plasma pro-inflammatory cytokines and increasing plasma anti-inflammatory cytokines, enhancing the serotonin pathway | 33 |
| | Mouse (ELS) | Lactobacillus plantarum PS128 | Reducing plasma pro-inflammatory cytokines and increasing plasma anti-inflammatory cytokines, enhancing the serotonin pathway | 34 |
| | Rat (CRS) | Lactobacillus helveticus NS8 | Reducing plasma stress hormones, regulating cytokines, and increasing hippocampal BDNF | 115 |
| Autism spectrum disorder (ASD) | Mouse (shank3 KO) | Lactobacillus reuteri | Regulating GABA receptor and oxytocin expression | 46 |
| | Mouse (shank3 KO, BTBR) | Lactobacillus reuteri | Redulating oxytocin expression | 47 |
| Alzheimer's disease (AD) | Human | Lactobacillus acidophilus, Lactobacillus casei, Bifidobacterium bifidum, Lactobacillus fermentum | Reducing oxidative stress by activating SIRT-1 dependent mechanisms | 70 |
| | Mouse (3xTg) | SLAB51 (Streptococcus thermophilus, Bifidobacterium longum, B. breve, B. infantis, Lactobacillus acidophilus, L. plantarum, L. paracasei, L. delbrueckii subsp. bulgaricus, L. brevis) | | |
| Social stress | Human | Lactobacillus acidophilus, L. fermentum, Bifidobacterium lactis, B. longum | | |

CSDS, chronic social defeat stress; CUS, chronic unpredictable stress; ELS, early life stress; CRS, chronic restraint stress.
prevent mental disorders by regulating the gut microbiome using probiotics. Although various studies on gut dysbiosis and mental disorders have been conducted, these studies are limited in that they show some correlation, but not a causal relationship, between the gut microbiome and mental health.

Future research should focus on elucidating the details of how changes in the gut microbiome are involved in mental illness. For example, it will be necessary to look at what byproducts or metabolites are produced by changes in the gut microbiome and how they change brain function. Preclinical studies using animal models to study human mental disorders should be conducted under consistent conditions such as use of experimental animals of the same strain, sex and age to analyze the changes in gut microbiome and metabolites. In addition, clinical studies require analysis of a sufficient population of race, residential area, and sex. There is a need to develop advanced gut microbiome analysis methods. It is better to sample directly in the gut under endoscope (for humans) or gut tissues (for experimental animals) than to use samples from feces [117]. Through these approaches, it is necessary to establish a database of gut microbiome and metabolites for each animal model of mental disorders or for patients with mental disorders. In addition, more research is needed to study how gut dysbiosis observed in mental disorders affects the transmission of signals from the gut to the brain through the vagus nerve or spinal cord. Finally, it would be necessary to investigate the therapeutic specificity of probiotics to symptoms of mental illness for the discovery of new, precise treatments to prevent or improve mental disorders toward the goal of more personalized treatments. It is not easy to study how the changes in gut microbiome observed in mental disorders or by the treatment of probiotics affects the environment in the gut or the communication between the gut and the brain. Main reason is there was no in vitro model to study the gut microbiome and probiotics in the past. However, recently developed 3D human intestinal organoids or other in vitro models will accelerate this research [118]. To understand the mechanism of the improvement of mental disorders by probiotics, it will be helpful to elucidate molecular or physiological changes in neural circuitries of gut-brain axis using cutting-edge techniques such as next generation sequencing (NGS) or in vivo electrophysiology or calcium imaging.

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