Voltammetric Determination of Ascorbic Acid in Pharmaceutical Formulations Using Modified Iodine-Coated Platinum Electrode

Determinación voltamétrica del ácido ascórbico en formulaciones farmacéuticas utilizando un electrodo de platino modificado recubierto de yodo

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Abstract

Background: Despite the high reactivity of the platinum electrode, the iodine-coated platinum electrode shows obvious inertness toward adsorption and surface processes. For that, iodine-coated platinum electrodes accommodate themselves to interesting voltammetric applications.

Objectives: This study reports using the modified iodine-coated polycrystalline platinum electrode as a voltammetric sensor for ascorbic acid determination in pharmaceutical formulations.

Methods: The developed voltammetric method based on recording cyclic voltammograms of ascorbic acid at iodine-coated electrode. The optimized experimental parameters for the determination of ascorbic acid were using 0.1 M KCl as a supporting electrolyte with a scan rate of 50mV/s.

Results: The anodic peak related to ascorbic acid oxidation was centered at nearly 0.28V. An excellent and extended linear dependence of the oxidative peak current on the concentration of ascorbic acid was observed in the range 2.84x10⁻³ - 5.68 mM. The limit of detection (LOD) and limit of quantitation (LOQ) were 1.0 µM and 3.01 µM, respectively, attesting to the method’s sensitivity. The investigation for the effect of potential interference from multivitamin tablet ingredients (vitamins B1, B6, B12, folic acid, citric acid, sucrose, glucose, and zinc) indicated specific selectivity toward ascorbic acid and the absence of any electrochemical response toward these components. Recovery results in the range 98.93±2.78 - 99.98±5.20 for spiked standard ascorbic acid in pharmaceutical formulations further confirmed the potential applicability of the developed method for the determination of ascorbic acid in real samples.

Conclusions: The developed method was successfully applied to the analysis of ascorbic acid (vitamin C), and the obtained results were in good agreement with the
labeled values; besides, the statistical tests indicated no significant difference at p=0.05 with a 95% confidence level.

**Keywords:** ascorbic acid analysis, pharmaceutical formulation, voltammetric analysis, iodine-coated platinum electrode, modified platinum electrode.

**Introduction**

L-ascorbic acid (C₆H₈O₆) is the trivial name of Vitamin C, scheme 1, considered one of the essential water-soluble vitamins for human health. It is found in various biological systems and fresh foodstuff (1). The human body required ascorbic acid for normal physiological functions such as the synthesis and metabolism of tyrosine, folic acid, and tryptophan (2). At the same time, ascorbic acid deficiency is associated with many diseases such as anemia infections and scurvy (3).
Scheme 1. The chemical structure of ascorbic acid

Additionally, synthetic ascorbic acid is available in several types of supplements such as tablets, capsules, chewable tablets, crystalline powder, effervescent tablets, and liquid forms (4). Several methods have been reported in the literature for the quantitative determination of ascorbic acid in various matrices. These methods include chromatography (5,6), titration (7), spectroscopy (8-12), fluorimetry (13), and flow injection analysis (14). However, some of the reported methods are time-consuming, and some are expensive and need skilled personnel. Alternatively, electrochemical methods are considered a promised methods because of the short time response, low cost, sensitivity, and simplicity of instrumentation (15). The modification of electrode surface is a quest to render electrochemical function that is not possible or difficult to achieve by using conventional electrodes. The goals of the improvement process include increasing selectivity, sensitivity, chemical and electrochemical stability, large usable potential window, and improving resistance to fouling (contaminating) (16). Therefore, the avenues have opened toward the modification of solid electrodes (17).

For instance, adsorbed iodine on platinum electrode surface enhances voltammetry’s reproducibility and simplifies background behavior (18). Also, coating of solid electrodes surface alters the kinetics and mechanisms of reactions run at the electrode surface. Iodine is one of the anions adsorbed to an electrode surface. The chemisorption process is achieved in two ways; from solution or vacuum to form stable chemisorbed monolayers; subsequently, the iodine-coated electrode is rinsed or evacuated (19). The iodine is adsorbed at potential 0.2 V vs. Ag/AgCl or SCE reference electrodes (20), which is the double layer potential. At the surface of the polycrystalline platinum electrode, the reaction of the iodine anions from the solution leads to spontaneous chemisorption of iodide anion to form stable neutral iodine atoms accompanied by an evolution of hydrogen gas. The adsorbed iodine is less reactive toward electrochemical oxidation than the free iodine anion in solution (21) and depends on the electrode potential (22). The chemisorbed iodine could be desorbed from the platinum electrode surface if the potential scanned is lower than -0.2 V, reducing of hydrogen ions and hydrogen gas generation (21). Also, the rate of iodine desorption from the electrode surface increases as the potential becomes more negative (22). In the positive direction, the chemisorbed iodine begins to desorb at a potential of 1.0 V (20). Studies have shown that carbon monoxide (CO) completely desorbed iodine from platinum electrode surface at potentials lower than 0.35 V, while at higher potentials, the desorption is incomplete (23).
Iodine-coated platinum electrode has been applied for the electrochemical determination of organic and non-organic species in many studies (24-32). Iodine-coated platinum electrode is characterized by the simplicity in preparation, application, and use of environmentally friendly chemical reagents. The simplicity of the method’s instrumentation stimulates our interest in this research. This work develops a simple method for ascorbic acid determination in pharmaceutical formulations.

Experimental

Materials and apparatus

A potentiostat (PAR Model 362, EG & G) interfaced to a computer via a GPIB interface (IEEE) data acquisition was used. Locally modified Labview® (IEEE) software was used for data acquisition. A one-compartment electrochemical cell with one inlet/outlet for gas purging and blanketing with oxygen-free nitrogen was used. The working electrode was a 0.5 mm polycrystalline platinum wire purchased from Aldrich (99.99% minimum purity certified reagent). The immersed end of the platinum electrode was curved at the end to a U-shape to mark for a constant surface area of the immersed part of the electrode. A silver/silver chloride was used as a quasi-reference electrode (QRE). The auxiliary electrode was a 0.5 mm polycrystalline platinum wire (Aldrich, certified 99.99% minimum purity). All reagents used were analytical grade and used as received from the suppliers without further purification. Sulfuric acid (95-97%) was supplied from Merck, L-ascorbic acid (99%) was purchased from AnalR, potassium iodide was purchased from Sigma-Aldrich. Ultra-pure water, Millipore-MilliQ system was used for the preparation of all solutions. The N\textsubscript{2} gas was a five grade, 99.999% minimum purity supplied from the International Jordanian Gases Company (Amman, Jordan).

Preparation of iodine-coated platinum electrode

The polycrystalline platinum electrode was cleaned with a freshly prepared chromic acid (H\textsubscript{2}CrO\textsubscript{4}), followed by rinsing with Millipore-Q water and sonicated for 10 minutes. After that, the platinum electrode was placed in contact with a supporting electrolyte solution of 0.5 M H\textsubscript{2}SO\textsubscript{4} and conditioned between -0.25V and 1.3V until obtaining a reproducible cyclic voltammogram of a polycrystalline platinum electrode, which manifests the cleanliness of the electrode surface and electrochemical cell contents (Fig.1).

After cleaning the platinum electrode, the electrode was immersed in a supporting electrolyte containing 0.5 M H\textsubscript{2}SO\textsubscript{4} + 0.01 M KI for five minutes under open-circuit conditions to complete the coating of the platinum electrode surface with iodine. Then the electrode was rinsed with water and 0.5M H\textsubscript{2}SO\textsubscript{4} solution extensively. After the coating step, the electrode was cycled in a supporting electrolyte solution between -
0.2V and +0.8V at a scan rate of 50 mV/s (Fig.1). The absence of oxygen and hydrogen adsorption/desorption features provides clear evidence for the complete coverage of the platinum electrode surface with a monolayer of iodine.

Sample preparation

The pharmaceutical formulation samples were purchased from local Jordanian pharmacies in the form of tablets and capsules. Three brands of pharmaceutical preparations were analyzed for their ascorbic acid content. The capsules of each sample were dissolved in 20 mL of 0.1 M KCl and sonicated for 10 min and left to equilibrate for 30 min. The solution was transferred to a 100 mL volumetric flask and filled to the mark with 0.1 M KCl. The solution was diluted to a concentration that matches the established calibration curve. A tablet of each sample was treated separately. The tablet of each brand was powdered using porcelain mortar and dissolved in 100.00 mL of the supporting electrolyte, 0.1 M KCl; the solution of the prepared samples was sonicated for 5 min and left to equilibrate for 5 min. A 5 mL aliquot of this solution was diluted to 50.00 mL with 0.1 M KCl to match the constructed calibration curve at different concentration ranges. A 10.00 mL of the diluted solution was placed in the electrochemical cell. The solution was bubbled with nitrogen gas (5G purity) and kept under a nitrogen gas atmosphere during the electrochemical experiment. The voltammetric analysis was conducted for ascorbic acid at the modified iodine-coated electrode within a potential window started at -0.2V and finished at 0.6V, where the adsorbed iodine is stable.

Results and discussions

Initially, a reproducible cyclic voltammogram for the polycrystalline platinum electrode, which indicates the cleanness of the electrochemical system, was obtained (Fig1-A). The process led to a successful coating process; the cyclic voltammogram of the iodine-coated platinum electrode between potential limits of -0.2V and 0.6V was displayed (Fig 1-B), where the adsorbed iodine was stable within this potential range. The complete absence of H₂ and O₂ oxidation-reduction features was the main indicator of a successful coated step.
Fig. 1. Cyclic voltammogram curves of (A) polycrystalline platinum electrode and (B) the same electrode after adsorption of iodine from 0.01 M KI in 0.5 M H₂SO₄ solution.

The effect of varying supporting electrolytes on the anodic peak current of ascorbic acid oxidation was investigated. A 0.5 M H₂SO₄ (pH=0.3), phosphate buffer of pH=3.5, and 0.1 M KCl (pH=7) solution were used. As displayed in Figure 2, various oxidation peak current was obtained for ascorbic acid oxidation with different supporting electrolyte solutions. The highest oxidation peak current was obtained in 0.5 M KCl (the highest pH value), 45.47±0.09 mV. Therefore, 0.1 M KCl solution was considered a supporting electrolyte in the following study.
Fig. 2. Effect of type of supporting electrolyte on the electrochemical signal of ascorbic acid at iodine-coated platinum electrode, Ascorbic acid: 0.57mM. Supporting electrolytes: H2SO4 0.5M (pH=0.3); phosphate buffer, pH=3.5; KCl 0.1M (pH=7) n=3; scan rate, 50 mV/sec. Numbers above the bars = mean±SD.

The effect of scan rate on the obtained anodic peak current of ascorbic acid was studied. As presented in Fig.3, there is a linear relationship between the square root of scan rate and oxidation peak current of ascorbic acid over the range of 10-100 mV/s, which suggested a diffusion-controlled irreversible oxidation process of ascorbic acid at the iodine-coated platinum electrode.
Cyclic voltammograms of iodine-coated platinum electrode in 0.1 M KCl and 50ppm of ascorbic acid recorded at 10, 20, 50, and 100 mv/s. b) The least square line for the ascorbic acid oxidation peak current vs. the square root of scan rate.

The obtained cyclic voltammograms for the iodine-coated platinum electrode in a series of ascorbic acid standard solutions show that the oxidation current increased steadily with ascorbic acid concentration (Fig.4). Three voltammograms were recorded for each standard solution. The anodic peak current was extracted for each cyclic voltammogram.

Plotting the anodic peak current variation against ascorbic acid concentration gave a straight and extended dynamic range with concentrations ranging between 2.84 μM - 5.68 mM. The calibration curve displayed in Fig 5 shows that all the variability of the
response data around its mean; $R^2=0.9969$, and the calibration equation is given by

$$I_{\mu A} = 65.248C_{\text{ascorbic acid}} + 3.06$$

Where $I$ represents the anodic peak current which attributed to the ascorbic acid oxidation as it is shown in the following equation:

Ascorbic acid $\rightarrow$ dehydro-ascorbic acid $+2H^+ + 2e^-$

The precision, that is, the repeatability of the method, was assessed by extracting the anodic peak current of the recorded cyclic voltammograms for a solution containing 0.28 mM ascorbic acid. The achieved coefficient of variation for 10 successive measurements was 1.36%, indicating the high precision of the developed method.

The limit of detection based on the formula $LOD=3.3\sigma/S$, and the limit of quantitation based on the formula $LOQ=10\sigma/S$, where $\sigma$ represents the blank signal (background current), and $S$ means the sensitivity of the calibration curve was calculated. The estimated limits were 1.0 $\mu$M and 3.01 $\mu$M, respectively. Thus, acceptable sensitivity of the applied voltammetric method with high precision was obtained. Higher sensitivity can be achieved by applying a more sensitive technique like differential pulse voltammetry (DPV). However, differential pulse voltammetry was not attempted because cyclic voltammetry provides satisfactory sensitivity for ascorbic acid determination in pharmaceutical formulations.

Fig. 5. An extended calibration curve shows the relationship between ascorbic acid concentration in ppm and the oxidation peak current measured from cyclic voltammograms for ascorbic acid in 0.1 M KCl at iodine-coated platinum electrode. Scan rate=50 mV*s$^{-1}$.

Potential interference

The influence of vitamins B1, B6, B12, folic acid, citric acid, sucrose, glucose, and zinc were investigated in order to verify the existence of matrix effects of vitamin C capsules and multivitamins tablets on ascorbic acid determination using cyclic voltammetry. The recorded cyclic voltammograms for each of these compounds show
the absence of any electrochemical response of iodine-coated platinum electrodes toward these compounds. Figure 6 shows the recorded voltammograms for a solution of Multivitamin sample (control) and after each addition of a known concentration of the ascorbic acid standard solution. The result proved the absence of any possible interference with ascorbic acid despite the various components included in the Multivitamin sample, Vitamin E, B1, B2, B6, Folic acid, Pantothenic acid, Biotin, and Niacin.

**Fig.6.** Cyclic voltammograms of iodine-coated platinum electrode in 0.1 M KCl containing Multivitamin tablet solution with addition of 0.1, 0.2, and 0.3 mM of ascorbic acid standard solution. Scan rate=50mV/s.

**Recovery**

The recovery experiment can be taken as evidence for the absence of interference. The feasibility of the developed voltammetric method for ascorbic acid determination was tested for three pharmaceutical formulation samples. Ascorbic acid standards of known concentration, 50 ppm and 60 ppm, were spiked into samples of tablet solutions in order to evaluate the percentage recovery for each brand of ascorbic acid. As listed in Table 1, the recovery values were found between 98.93±2.78 and 99.98±5.20 for all samples of ascorbic acid brands, showing the appropriateness of the iodine-coated platinum electrode for the quantitative analysis of ascorbic acid in pharmaceutical formulations.

**Table 1. Recoveries of ascorbic acid from spiked pharmaceutical formulations obtained by the developed method**

| Samples            | Content of ascorbic acid (mM) | Spiked ascorbic acid (mM) | Detected ascorbic acid after addition (mM), | % Recovery     |
|--------------------|-------------------------------|---------------------------|------------------------------------------|----------------|
| Vitamin C® plus     | 0.28                          | 0.227                     | 0.51                                     | 99.98±5.20     |
The developed voltammetric method was applied to analyze ascorbic acid in three brands of the pharmaceutical formulation, multivitamin tablets, and two brands of vitamin C capsules (Vito+ multivitamin, vitamin C plus, and vitamin C 1000). The standard addition method was applied to a diluted sample analysis to avoid the matrix effect. The evaluation of ascorbic acid concentration was found to be more suitable with the aid of a calibration graph. The results for the analysis of these pharmaceutical formulations with the developed voltammetric method are given in Table 2.

The obtained results by applying a cyclic voltammetry technique at iodine-coated platinum electrode were compared with the labeled values claimed by manufacturers. The data displayed in table 1 show that all nominal values are within the 95% confidence interval, which indicates the evident absence of errors in the results. The relative errors of the analysis of the three types of pharmaceutical formulations were lower than 5%, which attests to the accuracy of the developed method. The measured coefficient of variation values (0.55-2.19%) was considered obvious evidence of the precision of the developed method. The paired t-test was used to examine the significant difference at 95% confidence level between the labeled values and the obtained results determined by the developed voltammetric method. Comparing the calculated t value (0.0039) with the critical t value (4.30 at p=0.05) (33), it is shown that this result supported the null hypothesis and indicate no significant difference between the values determined by the voltammetric method and the nominal value obtained from manufacturers.

### Table 2 Ascorbic acid (Vitamin C) content in pharmaceutical formulations collected from the Jordanian local pharmacies as determined by cyclic voltammetry at iodine-coated platinum electrode.

| Pharmaceutical preparation | Nominal mass (in mg of ascorbic acid/capsule or tablet) | Average of determined mass (in mg) of ascorbic acid/capsule or tablet (n=3) | Standard deviation | 95% confidence limits | Relative error | Coefficient of variation |
|-----------------------------|--------------------------------------------------------|--------------------------------------------------------------------------------|--------------------|----------------------|---------------|-------------------------|
| Vitamin C plus®             | 500                                                    | 497.42                                                                           | 3.18               | 497.42±7.90          | 0.516         | 0.64%                  |
| Multivitamin+®              | 60                                                     | 60.32                                                                           | 1.32               | 60.31±3.28          | 0.53         | 2.19%                  |
| Vitamin C1000®             | 1000                                                   | 997.53                                                                           | 5.48               | 997.531±13.61       | 0.247         | 0.55%                  |

A comparison between the developed voltammetric method and some of the common analytical and voltammetric methods for ascorbic acid determination in terms of detection limit and the linear range was displayed in Table 2. As shown, the iodine-
coated platinum electrode exhibited a lower detection limit than that of other voltammetric methods (29-31) (34-36). In contrast, the obtained linear range was convenient and extended compared to other voltammetric methods. Also, the developed method has the advantages of simplicity in sample preparations and analysis, side by side with a short time of analysis and the low price of instrumentations compared with other methods (Table 3).

Table. 3 A comparison of analytical performance of developed voltammetric method using iodine-coated platinum electrode with other analytical methods reported in literature

| Method                    | Linear range         | Detection Limit(ppm) | Reference |
|---------------------------|----------------------|----------------------|-----------|
| HPLC                      | 56.78 µM-0.57 mM     | 1.7x10⁻⁷ mM          | [5]       |
| Spectrophotometry         | Method A:3.69 µM-63.59 µM Method B:2.9 µM-90.85 µM | Method A=0.85 µM, Method B=1.1 µM | [4] |
| Fluorometry               | 0.7 µM-6.02 µM       | 0.23 µM              | [13]      |
| Cyclic voltammetry        | 0.01 mM-0.101 mM     | 1.76 µM              | [34]      |
| Differential pulse voltammetry | 19.0 µM-0.21 mM              | 19.0 µM              | [35]      |
| Square-wave voltammetry   | -                    | 1.87 µM              | [36]      |
| Voltammetry               | 2.84 µM - 5.68 mM    | 0.96 µM              | This work |

Conclusion

In this work, a successive use of an iodine-coated platinum electrode to determine ascorbic acid was achieved. The developed method excludes any sophisticated procedures. In contrast, it is considered an applicable method for simplicity of analysis procedures. The reported extended dynamic range 2.84x10⁻³ - 5.68 mM of ascorbic acid supports the applicability of the voltammetric method for ascorbic acid analysis in pharmaceutical products. Based on the recovery experiment, the absence of any interference from the other ingredients of pharmaceutical formulations is considered an evident indicator of the selectivity of the developed method. The statistical analysis of the results showed no significant difference between the values obtained from the voltammetric method and the labeled values claimed by the manufacturers.

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Conflict of Interest

The authors declare that there is no conflict of interest

References

1. Pisoschi AM, Danet AF, Kalinowski S. Ascorbic acid determination in commercial fruit juice samples by cyclic voltammetry. J Autom Methods Manag Chem. 2008; 2008:1-8 https://doi.org/10.1155/2008/937651

2. Chambial S, Dwivedi S, Shukla KK, John PJ, Sharma P. Vitamin C in disease prevention and cure: An Overview. Indian Journal of Clinical Biochemistry. 2013; 28(4), 314–328. https://doi.org/10.1007/s12291-013-0375-3

3. Perry M, Page N, Manthey D, Zavitz J. Scurvy: Dietary Discretion in a developed country. Clinical Practice and Cases in Emergency Medicine. 2018; 2(2): 147–150. https://doi.org/10.5811/cpcem.2018.1.36860

4. Naidu KA. Vitamin C in human health and disease is still a mystery? An overview. Nutrition Journal. 2003; 2:1-10. https://doi.org/10.1186/1475-2891-2-7.

5. Hu L, Li L, Luo Z, Yang J, Liu W. Determination of trace vitamin C by ion-pair HPLC with UV detection in calcium gluconate and vitamin C compound oral solution. Journal of Chromatographic Science. 2012; 50(2):102–107. https://doi.org/10.1093/chromsci/bmr035

6. Gazdik Z, Zitka O, Petrolova J, Adam V, Zehnalek J, Horna A, Reznicek V, Beklova M, Kizek, R. Determination of vitamin C (Ascorbic Acid) using high performance liquid chromatography coupled with electrochemical detection. Sensors 2008; 8(11): 7097–7112. https://doi.org/10.3390/s8117097

7. Shrestha N, Shrestha S, Bhattarai A. Determination of ascorbic acid in different citrus fruits of Kathmandu Valley. Journal of medical and Biological Science Research. 2016; 2(1):9-14.

8. Anal PD, Shuchi, D. UV spectroscopic method for determination of vitamin C (ascorbic acid) content in different fruits in south Gujarat Region. International Journal of Environmental Sciences & Natural Resources. 2019; 21(2):41-44. https://doi.org/10.19080/IJESNR.2019.21.556056.

9. Mirsad S, Amra S. Spectrophotometric determination of L-ascorbic Acid in pharmaceutical based on its oxidation by potassium peroxymonosulfate and hydrogen Peroxide. Croatica Chemica Acta. 2015; 88:73-79. https://doi.org/10.5562/cca2551.

10. Zanini DJ, Silva MH, Aguiar-Oliveira E, Mazalli MR, Kamimura ES, Maldonado RR. Spectrophotometric analysis of vitamin C in different matrices utilizing potassium permanganate. European International Journal of Science and Technology. 2018; 7(1):70-84.

11. Arya SP, Mahajan M, Jain P. Photometric methods for the determination of vitamin C. Analytical Sciences. 1998; 14:889-895.

12. Lau O-W, Luk S-F, Wong K-S. Determination of ascorbic acid in pharmaceuticals using direct ultraviolet spectrophotometry. Analyst. 1987; 112:1023-1025.

13. Dilgin Y, Nisli G. Fluorimetric determination of ascorbic acid in vitamin C tablets using methylene blue. Chem. Pharm. Bull. 2015; 53:1251-1254. https://doi.org/10.1248/cpb.53.1251.
14. Ensafi AA, Rezaei B. Flow injection analysis determination of ascorbic acid with spectrofluorimetric detection. Analytical Letters. 1998; 31:333-342. https://doi.org/10.1080/00032719808002049.

15. Gazdik Z, Zitka O, Petrolova J, Adam V, Zehnalek J, Horna A, Reznicek V, Beklova M, kizek R. Determination of vitamin C (ascorbic acid) using high performance liquid chromatography coupled with electrochemical detection. Sensors. 2008; 8(11):7097-7112. https://doi.org/10.3390/s8117097.

16. Clucu AA. Chemically modified electrodes in biosensing. J Biosens Bioelectron. 2014; 5(3):1-10. https://doi.org/10.4172/2155-6210.1000154.

17. March G, Nguyen TD, Piro B. Modified electrodes used for electrochemical detection of metal ions in environmental analysis. Biosensor. 2015; 5(2):241-275. https://doi.org/10.3390/bios5020241.

18. Cox JA, Kulesza PJ. Oxidation and determination of nitrite at modified electrodes. J Electroanal. Chem. 1984; 175(1-2): 105-118. https://doi.org/10.1016/S0022-0728(84)80349-6

19. Felter TE, Hubbard AT. L.E.E.D. and electrochemistry of iodine on Pt (100) and Pt (111) single-crystal surfaces. J Electroanal. Chem. 1979, 100:473-491. https://doi.org/10.1016/S0022-0728(79)80179-5.

20. Shu ZX, Bruckenstein S. Iodine Adsorption Studies at Platinum. J. Electroanal. Chem. 1991, 317:263-277. https://doi.org/10.1016/0022-0728(91)85019-L.

21. Mebrahtu T, Rodriguez JF, Bravo BG, Soriaga MP. Hydrogenative/cathodic stripping of iodine chemisorbed on smooth polycrystalline platinum electrode. J. Electroanal. Chem. 1987, 219, 327-333. https://doi.org/10.1016/0022-0728(87)85050-7.

22. Thomas AE, Wieckowski A. Surface diffusion limited desorption of iodine on a platinum electrode?. Journal of Electroanalytical Chemistry. 1995; 399:207-212. https://doi.org/10.1016/0022-0728(95)04226-1.

23. Podlovchenko BI, Kolyadko EA. Adsorption of carbon monoxide on platinized platinum electrode with preadsorbed iodine and iodide Anions. Russian Journal of Electrochemistry. 2003; 39: 823-827.

24. Hourani MK. Determination of silver (I) by cyclic voltammetry at iodine-coated electrodes. Analyst. 1994; 119: 1975-1978. https://doi.org/10.1039/AN9941901975.

25. Lane RF, Hubbard AT, Fukunaga K, Blanchard RJ. Brain catecholamines: detection in vivo by means of differential pulse voltammetry at surface-modified platinum electrodes. Brain Research. 1976; 114(2): 346–352. https://doi.org/10.1016/0006-8993(76)90678-8.

26. Hourani M, Jarar A, Arar S. Atmospheric SO2 determination by voltammetric analysis at an iodine-coated platinum electrode. Electroanalysis. 1999; 11(9):637-640. https://doi.org/10.1002/(SICI)1521-4109(199907)11:9<637::AID-ELAN637>3.0.CO;2-R.

27. Hourani Mk, Hijaz B. Voltammetric Analysis of Hydroquinone and Catechol at Iodine-Coated Polycrystalline Platinum Electrode. Journal of Natural and engineering Science. 2014; 8(2): 25-29.

28. Amayreh M, Hourani MK. Determination of iron in dietary supplements by voltammetric analysis at an iodine-coated polycrystalline platinum electrode. Int.J. Electrochem. Sci. 2018; 13: 975-983. https://doi.org/10.20964/2018.01.81.
29. Amayreh M, Hournai MK. Direct determination of hemoglobin in blood using iodine-coated platinum polycrystalline electrode. Analytical and Bioanalytical Chemistry Research. 2019; 6(1): 59-68. https://doi.org/10.22036/ABCR.2018.125953.1198.

30. Hournai MK, Amayreh M, Hourani W. A Voltammetric sensor based on iodine-coated platinum electrode for determination of iron in blood serum. Anal.Bioanal.Electrochem. 2018; 10(12):1620-1628.

31. Amayreh M, Hourani M. Determination of Iron in Spinach Using Sweep Voltammetry at Iodine-Coated Platinum rotating Disk Electrode. Journal of AOAC International. 2019; 102(2):666-668. https://doi.org/10.5740/jaoacint.18-0267.

32. Amayreh M, Hourani W, Hourani MK. Anodic Stripping Voltammetric Determination of Copper in Multivitamin-Mineral Formulations using Iodine-Coated Platinum Electrode. Methods Objects Chem. Anal.2021,16(1),48-56. https://doi.org/10.17721/moca.2021.48-56.

33. Miller JN, Miller JC Statistics and Chemometrics for Analytical Chemistry.6th ed. Pearson, England;2010.43-45p.

34. Bitew Z, Amare M. Electrochemical determination of ascorbic acid in pharmaceutical tablets using carbon paste electrode. Organic & Medicinal Chemistry International. 2019; 8(5):1-9.https://doi.org/10.19080/OMCIJ.2019.08.555749

35. Lourenção BC, Medeiros RA, Rocha-Filho RC, Fatibello-Filho O, Simultaneous differential pulse voltammetric determination of ascorbic acid and caffeine in pharmaceutical formulations using a boron-doped diamond electrode. Electroanalysis.2010; 22(15):1717-1723. https://doi.org/10.1002/elan.200900612

36. Vedenyapina MD, Kazakova MM, Skundin AM. Voltammetric determination of ascorbic acid in pharmaceutical formulation on a boron doped diamond electrode. Russ. J. Phys. Chem. 2019; 93: 1178-1181. https://doi.org/10.1134/S0036024419060335

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