Research Article

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Enzymatic degradation study of PLA-based composite scaffolds

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Abstract: Disadvantages in the use of polylactic acid (PLA) as a base material for Tissue Engineering applications include the low osteoconductivity of this biomaterial, its acidic degradation and the deficient cellular adhesion on its surface. In order to counteract these drawbacks, calcium carbonate (CaCO₃) and β-tricalcium phosphate (Ca₃(PO₄)₂, β-TCP) were proposed in this work as additives of PLA-based support structures. Composite scaffolds (PLA:CaCO₃:β-TCP 95:2.5:2.5) manufactured by fused deposition modeling (FDM) were tested under enzymatic degradation using proteinase K enzymes to assess the modification of their properties in comparison with neat PLA scaffolds. The samples were characterized before and after the degradation test by optical microscopy, scanning electron microscopy, compression testing and thermogravimetric and calorimetric analysis. According to the results, the combination of the PLA matrix with the proposed additives increases the degradation rate of the 3D printed scaffolds, which is an advantage for the application of the composite scaffold in the field of Tissue Engineering. The higher degradation rate of the composite scaffolds could be explained by the release of the additive particles and the statistically higher microporosity of these samples compared to the neat PLA ones.

Keywords: polylactic acid (PLA); bone tissue engineering; proteinase K

1 Introduction

Polylactic acid (PLA) is a widely used biomaterial in Bone Tissue Engineering because of its biocompatibility, its suitable mechanical properties and ease to be processed [1]. However, there are some disadvantages which limit its efficiency as a support material for bone ingrowth, such as the low osteoconductivity of this biomaterial, its acidic degradation and the deficient cellular adhesion on its surface [2]. Furthermore, the degradation of PLA is deemed to be too slow to enhance the replacement of the material by the new bone tissue [3, 4]. In order to counteract these drawbacks, calcium carbonate (CaCO₃) and β-tricalcium phosphate (Ca₃(PO₄)₂, β-TCP) are proposed in this work as additives of PLA-based support structures intended for bone regeneration. β-TCP is a biodegradable and biocompatible ceramic material that has been extensively used in the field of Bone Tissue Engineering due to its osteoconductivity and its ability of complete bioreabsorption [5]. The use of the CaCO₃ as an additive of PLA responds to the need of counteracting the release of acidic products during the degradation of the base material, maintaining the pH around 7.4 by buffer effect [6]. The composite blend developed was used to manufacture scaffolding structures by the method of additive manufacturing, under the category of "material extrusion" (ISO/ASTM 52900:2015), commonly known as fused deposition modeling (FDM). Additive manufacturing techniques provide the possibility of controlling the porosity of the scaffolds to be used in Tissue Engineering and personalizing their design according to the patients’ needs [7].

In this work, the assessment of the degradation rate modification due to the presence of CaCO₃ and β-TCP has been carried out. For this purpose, degradation tests of PLA and composite (PLA:CaCO₃:β-TCP 95:2.5:2.5) scaffolds catalyzed by proteinase K enzymes were carried out. The enzymatic degradation test was designed with the aim of accelerating the degradation of PLA and composite scaffolds, since it can take more than 6 months to obtain significant weight losses when the experiment is carried out using PBS as degradation medium [8, 9]. Several examples are found in the literature about the use of this en-
zyme to degrade PLA fibres, films or scaffolds, obtaining significant differences in terms of weight loss or mechanical properties in days or even hours [10–12]. Sheng et al. [13], for example, obtained weight losses of between 30-50% in only 5 days by studying the degradation of PLA scaffolds using proteinase K enzymes. Although these working conditions do not rigorously simulate the conditions for the degradation of the material in vivo, this experiment design allows us to compare the composite scaffolds manufactured with the neat PLA ones. The degradation study was complemented with the morphological, thermogravimetric and mechanical characterization of the samples, before and after degradation, in order to assess the modification of the PLA properties due to the incorporation of the additives.

2 Experimental

PLA L105 in powder form was kindly supplied by Corbion Purac. Commercial grade calcium carbonate 0179-500G with a particle size of 30 µm was kindly supplied from VWR, while β-tricalcium phosphate (β-TCP) with a mean particle size of 45 µm was kindly provided by the 3B’s Research Group of Universidade do Minho. These three materials were mixed to obtain the following mixture (wt:wt): PLA:CaCO$_3$:β-TCP 95:2.5:2.5.

PLA and composite filaments were obtained using a lab prototype extruder with an 8 mm screw, a 1.6 mm diameter nozzle tip and an L/D cylinder ratio of 10. The working parameters included a rotating speed of 7 rpm and a temperature set at 245°C for the thermal resistance (the measured temperature in the proximity of the nozzle tip was equal to 180°C). The filaments needed to print the 3D structures were obtained after a final cooling stage using compressed air. A BQ Hephestos 2 3D printer (Spain) was used to manufacture scaffolds with a rectangular 0/90° pattern, a diameter of 9.8 mm and a height of 7 mm, resulting in a theoretical porosity of 50%. A nozzle diameter of 0.4 mm was used to print the scaffolds and the printing parameters included a temperature of 215°C and a deposition speed of 40 mm/s.

For the enzymatic degradation study, PLA and composite scaffolds were tested for time periods up to 5 and 10 days. Four replicas per group and time period were used. After measuring the weight of the scaffolds using an analytical balance (±0.1 mg, A&D Scales Gemini Series, GR-200, Germany), the samples were placed individually in a 24 well-plate and immersed in 2 mL of 0.05 M pH 8.6 Tris–HCl buffer solution containing 0.2 mg/mL of proteinase K from Tritirachium album (Merck, Darmstadt, Germany) and 0.2 mg/mL of sodium azide (Merck, Darmstadt, Germany). The degradation study was carried out at 37°C and the buffer-enzyme solution was replaced daily in order to maintain a high enzymatic activity. PLA and composite scaffolds incubated without enzymes in Tris–HCl solution were also evaluated as control samples. After the time periods studied, the scaffolds were weighed again to calculate the mass loss.

Before and after the degradation test, the surface morphology of the scaffolds was evaluated by scanning electron microscopy (SEM; Hitachi TM 3030 at an acceleration voltage of 15 kV). In addition, the pore size of these structures was assessed as the distance between filaments, using an Olympus BX51 optical microscope for that purpose. The pore size was calculated as the average of 40 measures per group of samples. Furthermore, the following equation was used in order to estimate the porosity of the 3D printed scaffolds [14, 15]:

\[
\%\text{porosity} = 100 \cdot \left(1 - \frac{\rho_{ap}}{\rho_{bulk}}\right),
\]

where $\rho_{ap}$ is the apparent density of the structure and $\rho_{bulk}$ is the density of the bulk material. The first one was calculated using the mass and dimensions of the 3D printed scaffolds of each group of samples studied. The density of the bulk material was determined by measuring the dimensions and mass of samples (n=8) extracted from the PLA and composite extruded filaments.

Samples extracted from the degraded scaffolds were subjected to thermogravimetric analysis in a TGA/DSC 1 Mettler Toledo device. PLA in powder form as well as non-degraded PLA and composite scaffolds were also analysed. The samples (n=4) were placed in aluminium crucibles and heated up to 385°C at a rate of 10°C/min. The thermal cycle was performed while working with a nitrogen flow of 10 mL/min. During the TGA testing and using the same thermal cycle, calorimetric data (heat flow curve) of each type of sample was obtained, from which the melting temperature and the melting enthalpy were estimated. The melting enthalpy values obtained were used to calculate the crystallinity of each group of samples according to the following equation [16]:

\[
\%X_c = 100 \cdot \left(1 - \frac{\Delta H_f}{\Delta H^0_{f \cdot W_{PLA}}}\right),
\]

where $X_c$ is the degree of crystallinity, $\Delta H_f$ is the enthalpy of fusion of the sample, $\Delta H^0_f$ corresponds to the heat of fusion of 100% crystalline PLA and $W_{PLA}$ is the net weight fraction of the PLA in the sample tested. The value used for $\Delta H^0_f$ was 93.7 J/g [17].

Regarding the mechanical characterization, the scaffolds degraded were tested by compression on an LIYI
testing machine in displacement control mode. Crosshead speed was set at 1 mm/min. The compressive modulus was calculated according to ASTM D695-15 using the initial steepest straight-line portion of the load-strain curve. These results were compared with the ones obtained by testing non-degraded PLA and composite scaffolds. Four replicas were used per group of samples.

Statistical analysis was performed using MATLAB software (MATLAB and Statistics Toolbox Release 2017a, The MathWorks, Inc., Natick, USA). The data obtained during this study were analysed by the Wilcoxon two-sided rank sum test when comparing two groups and by the Kruskal-Wallis test when more than two groups were compared. The significance level was set to *p < 0.05, **p < 0.01 and ***p < 0.001 for statistically significant, highly statistically significant and very highly statistically significant differences, respectively. All the figures show the mean values of each group and their standard deviations are represented with error bars.

### 3 Results and Discussion

After 5 days of enzymatic degradation, the mean weight loss of the PLA scaffolds group was 8.0%, while this value was equal to 13.4% for the group of composite samples. At the end of the experiment (day 10), the PLA and composite samples were degraded up to a 17.6% and a 22.7%, respectively. As showed in Figure 1, a statically significant difference (p < 0.05) in terms of weight loss was obtained when comparing each type of samples at the two time periods studied. The PLA and composite samples used as control (immersed in Tris-HCl buffer solution with sodium azide but without enzymes) showed no weight loss during the degradation test. According to these results, the degradation rate of both groups between days 5 and 10 was very similar, being 9.6% and 9.3% for the PLA and composite samples, respectively. Therefore, the greater weight loss obtained for the composite scaffolds can be attributed to an increased degradation rate for these samples during the first steps of the experiment.

SEM images of each group of scaffolds showed mesostructures with well-defined square shaped pores in an interconnected network, as presented in Figure 2. Unlike PLA samples, which showed a translucent appearance, scaffolds manufactured using composite filaments exhibited a whitish colour due to the presence of CaCO$_3$ and β-TCP particles. No relevant differences in relation to the surface morphology of the filaments printed by FDM were observed when comparing PLA and composite samples before enzymatic degradation (Figures 2a and 2d). Both groups of samples showed filaments with a smooth surface but a slightly variable diameter. From the figures of the samples after 5 and 10 days of degradation (Figure 2b, 2c for PLA scaffolds and Figures 2e and 2f for the composite ones), it is evident how the diameter of the filaments is significantly reduced during the experiment, which led to an increase in the pore size and the porosity of the samples. These figures suggest different degradation profiles for the PLA and the composite samples, since for the latter group the presence of a large number of holes over the surface of the filaments can be easily observed; these voids could be attributed to the release of the additive particles during the degradation of the structure. This could be a reason for the higher rate of degradation observed in the composite scaffolds after 5 and 10 days. In the case of the PLA samples, some fractures on the surface of the filaments were found, from which the degradation of the structures progresses. We expect that the release of the additives during the degradation of the structure will have a positive effect on maintaining the pH at an appropriate level for the surrounding tissue.

Results concerning variations in porosity and pore size of the samples are presented in Table 1. Composite scaffolds showed a statistically higher porosity (p < 0.05) in comparison with the neat PLA scaffolds before the degradation test. This result could be related to the greater varia-
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**Table 1:** Porosity and pore size values of the 3D printed scaffolds before and after degradation.

| Sample        | Porosity (%) | Pore size (µm) |
|---------------|--------------|----------------|
| PLA           | 57±2         | 476±52         |
| PLA 5D        | 57±4         | 531±51         |
| PLA 10D       | 65±3         | 527±62         |
| COMPOSITE     | 62±2         | 482±37         |
| COMPOSITE 5D | 67±4         | 476±58         |
| COMPOSITE 10D| 68±4         | 518±59         |

The results obtained from the thermogravimetric and calorimetric analyses are shown in Table 2, in which \( T_{\text{onset}} \) represents the temperature for the start of the thermal degradation of the material and \( T_{\text{peak}} \) is the temperature of maximum degradation rate. The comparison between the results of PLA powder and 3D printed PLA samples showed a statistically significant difference (\( p < 0.05 \)) in terms of \( T_{\text{onset}} \), but not regarding the degree of crystallinity of the samples. One important trend observed (non-statistically significant difference) is the increase of the crystallinity due to the introduction of the additives into the PLA matrix, as the CaCO\(_3\) and β-TCP particles act as nucleation points [17].

On the other hand, after 10 days of enzymatic degradation PLA and composite samples showed, respectively, a statistically significant (\( p < 0.05 \)) and a highly statistically significant (\( p < 0.01 \)) decrease in terms of degree of crystallinity when compared to the non-degraded PLA and composite samples. A statistically significant (\( p < 0.05 \)) decrease was also obtained for the composite scaffolds after 10 days regarding the degradation and melting temperatures. The decrease of the samples’ crystallinity was an unexpected result, as early enzymatic degradation preferentially occurs in the amorphous region of PLA [19], so
Table 2: Values determined from TGA and calorimetric analysis.

| Sample          | $T_{\text{onset}}$ (°C) | $T_{\text{peak}}$ (°C) | Melting temperature (°C) | Enthalpy of fusion (J/g) | Xc (%)  |
|-----------------|--------------------------|-------------------------|--------------------------|--------------------------|---------|
| PLA powder      | 350±1                    | 370±1                   | 174±2                    | 52.3±0.9                 | 55.9±1.0|
| PLA             | 353±1                    | 369±1                   | 175±3                    | 53.6±1.8                 | 57.2±1.9|
| PLA 5D          | 353±1                    | 369±1                   | 174±3                    | 38.5±2.9                 | 41.1±1.4|
| PLA 10D         | 350±2                    | 369±2                   | 174±3                    | 37.4±1.3                 | 40.0±1.4|
| COMPOSITE       | 353±1                    | 370±1                   | 177±3                    | 58.4±2.0                 | 59.3±2.0|
| COMPOSITE 5D    | 348±1                    | 368±1                   | 173±3                    | 35.4±1.2                 | 35.9±1.2|
| COMPOSITE 10D   | 349±1                    | 367±2                   | 171±2                    | 28.6±4.0                 | 29.0±1.0|

Figure 3: Mechanical properties of the 3D printed scaffolds under compression testing (*p<0.05).

the crystallinity percentage should have increased during the experiment. However, according to the literature [4], when the molecular weight of the PLA samples under enzymatic degradation decreased down to a value around 30,000 g/mol, the hydrolytic degradation of crystalline regions of the material could be also enhanced. This can be a reason for the significant reduction of the crystallinity in both groups of samples, although further research is required to clarify this extent.

Regarding the mechanical characterization, the values of the compressive modulus for non-degraded samples were 52±10 MPa for the PLA scaffolds and 61±12 MPa for composite ones (Figure 3). After 5 days of degradation, these values were decreased down to 31±7 MPa and 31±13 MPa, respectively. At the end of the experiment, the compression modulus was 26±5 MPa for PLA scaffolds and 22±1 MPa for the composite samples, both results being statistically significantly lower (p<0.05) than the initial values for each type of sample.

4 Conclusions

The higher rate of degradation observed in the composite scaffolds after 5 and 10 days could be explained by the release of the additive particles and the statistically higher microporosity of composite scaffolds compared to the neat PLA samples, which could enhance the degradation rate at the early steps of the test. Regarding the compression test, a statistically significant decrease of the elastic modulus after 10 days of enzymatic degradation was confirmed for both PLA and composite scaffolds, with results still in the range of values reported for cancellous bone (20–500 MPa) [20].
The combination of the PLA matrix with the proposed additives increases the degradation rate of the 3D printed scaffolds, which is an advantage for the application of the composite scaffold in the field of tissue engineering, taking into account the large amount of time necessary for the degradation of PLA structures. The ratio of use of these additives could be adjusted to the degradation rate required to match the growth rate of new bone tissue.

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