Short Report: Genetics

Analysis of cell-free fetal DNA for non-invasive prenatal diagnosis in a family with neonatal diabetes

E. De Franco1,*, R. Caswell1,*, J. A. L. Houghton1, V. Iotova2, A. T. Hattersley1 and S. Ellard1

1Institute of Biomedical and Clinical Science, University of Exeter Medical School, Exeter, UK and 2University Hospital ‘St. Marina’, Varna, Bulgaria

Accepted 27 June 2016

Abstract

Aims An early genetic diagnosis of neonatal diabetes guides clinical management and results in improved treatment in ~ 40% of patients. In the offspring of individuals with neonatal diabetes, a prenatal diagnosis allows accurate estimation of the risk of developing diabetes and, eventually, the most appropriate treatment for the baby. In this study, we performed non-invasive prenatal genetic testing for a fetus at risk of inheriting a paternal KCNJ11 p.R201C mutation causing permanent neonatal diabetes.

Methods A droplet digital polymerase chain reaction assay was used to detect the presence of the mutation in cell-free circulating DNA (cfDNA) extracted from maternal plasma at 12 and 16 weeks’ gestation.

Results The mutation was not detected in the cfDNA samples, suggesting that the fetus had not inherited the KCNJ11 mutation. The fetal DNA fraction was estimated at 6.2% and 10.7%, which is above the detection limit of the assay. The result was confirmed by Sanger sequencing after the baby’s birth, confirming that the baby’s risk of developing neonatal diabetes was reduced to that of the general population.

Conclusions We report the first case of non-invasive prenatal testing in a family with neonatal diabetes. A prenatal diagnosis in families at high risk of monogenic diabetes informs both prenatal and postnatal management. Although the clinical impact of this novel technology still needs to be assessed, its implementation in clinical practice (including cases at risk of inheriting mutations from the mother) will likely have a positive impact upon the clinical management of families affected by monogenic diabetes.

Diabet. Med. 34, 582–585 (2017)

Introduction

Neonatal diabetes is diagnosed before 6 months of age and affects ~ 1 in 100 000 live births [1]. Almost 40% of people affected with neonatal diabetes have mutations in the genes encoding for the subunits of the ATP-sensitive potassium channel, ABCC8 and KCNJ11. Approximately 20% of individuals with a mutation in a KATP channel gene have additional neurological features, most commonly developmental delay and, in the more severe cases, early-onset epilepsy [2,3]. A recent study suggested that all individuals with neonatal diabetes caused by mutations in ABCC8 or KCNJ11 have some degree of neurological impairment [4].

Most of these individuals can be successfully treated with sulfonylurea instead of insulin, leading to improved glycaemic control and neurological function [5,6]. Current evidence indicates that the earlier sulfonylurea treatment is started, the better the outcome for neurological function [7,8]. An early genetic diagnosis of neonatal diabetes is, therefore, essential to guide treatment decisions, with benefits for the management of both diabetes and additional complications [9].

Analysis of the cell-free fetal DNA (cffDNA) present in maternal blood provides a non-invasive means of testing pregnancies at high risk of genetic conditions. It has transformed prenatal genetic testing for Down syndrome [10]. cffDNA is present in the maternal circulation from the first trimester at a proportion of ~ 10% of the total circulating DNA [11]. The implementation of highly sensitive techniques such as massively parallel deep sequencing [12] and droplet digital polymerase chain reaction (ddPCR) [13] allows the detection of variants that are present in the

Correspondence to: Sian Ellard. E-mail: sian.ellard@nhs.net
*Joint senior authors.
This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.
What’s new?

- This article describes the first case of non-invasive prenatal diagnosis using cell-free fetal DNA in a family with neonatal diabetes.
- The test showed that the baby had not inherited the neonatal diabetes mutation, providing reassurance to the parents at mid-gestation that this baby was not at high risk of neonatal diabetes and informing management of the pregnancy, delivery and neonate accordingly.

Patients and Methods

Informed consent was obtained from the couple. Cell-free DNA (cfDNA) was extracted from maternal plasma collected at 12 and 16 weeks’ gestation using the Chemagic Circulating NA kit (Perkin-Elmer, Waltham, MA, USA). Detection of the KCNJ11 c.601C>T, p.R201C mutation (NM_000525.3) was carried out by ddPCR. A custom Life Technologies (Carlsbad, CA, USA) TaqMan assay was designed to detect the C (reference) and T (mutated) alleles: forward PCR primer, CGGCCGCCTCTGCTT; reverse PCR primer, TCATGCTTGCCGAGGTC; probe 1 (C allele), VIC-ACCCACACATAGCATG-NFQ; probe 2 (T allele), FAM-CCCCACACATAGCATG-NFQ. This assay was used in reactions for droplet generation, PCR and detection using the Bio-Rad (Hercules, CA, USA) QX200 ddPCR system, as recommended by the manufacturers, with an annealing/extension temperature during PCR of 55°C. All data were analysed using Bio-Rad QuantaSoft software, with merging of data for multiple wells where relevant.

Assays were performed on paternal genomic DNA (gDNA) (known genotype C/T), maternal gDNA (known genotype C/C) and cfDNA extracted from maternal plasma (genotype and gender unknown). To establish the sensitivity of the assay for the T allele in a background of maternal gDNA, control samples were prepared in which paternal gDNA was spiked into maternal gDNA at concentrations of 10%, 1% and 0.1% of total DNA.

To establish the proportion of cfDNA in the cfDNA samples from maternal plasma, parental genomic DNA samples were assayed by ddPCR using a panel of common single nucleotide polymorphisms to identify one for which the fetus would be an obligate heterozygote. The single nucleotide polymorphism selected was rs4536103 (NEUROG3, NM_020999.3:c.596T>C) at which mother and father were homozygous for the A or G alleles, respectively. Thus, the presence of the paternally inherited G allele in the cfDNA sample allowed accurate estimation of the percentage of cfDNA in the 12- and 16-week samples.

Results

Counts of the C (reference) and T (mutated) alleles for the KCNJ11 c.601C>T mutation in paternal and maternal gDNA using ddPCR were consistent with the known genotypes of these samples (father heterozygous C/T, mother homozygous C/C), with paternal alleles present at a 50:50 ratio (within 95% Poisson confidence intervals) as expected (Fig. 1a).

The sensitivity of the assay was assessed using maternal gDNA spiked with paternal gDNA at various concentrations. The fraction of fetal DNA in maternal cfDNA is typically in the range 2–20% [11], and we were able to readily detect the paternal T allele when spiked into maternal gDNA at a concentration of 1% (data not shown). We set a conservative limit for reliable detection of the T allele as 10 positive droplets across all wells analysed (26 881 droplets in total for the 1% spiked sample), which equates to ~18 copies of amplifiable DNA. No droplets positive for the T allele were observed in negative (no template) controls or in maternal gDNA samples. Analysis of cfDNA extracted at either 12 or 16 weeks’ gestation showed that although the C allele was abundant, the T allele was not reliably detected. Pooled data from replicate wells gave an apparent concentration of 0.016 copies/µl in the 12-week sample (one positive droplet observed of 74 635 analysed), whereas no positive droplets for the T allele were observed in the 16-week sample (Fig. 1a). These results suggest that the fetus had not inherited the KCNJ11 p.R201C mutation.

Estimation of the fetal DNA fraction in the two cfDNA samples using rs4536103 (for which the fetus was an obligate heterozygote) confirmed the presence of cfDNA at a relative amount of 6.2% in the 12 weeks’ gestation sample and 10.7% in the 16 weeks’ sample, consistent with previous reports [11] (Fig. 1b).

Given the fetal fractions measured using rs4536103, had the fetus inherited the paternal KCNJ11 c.601T allele this would have been expected at an abundance of ~173 copies or ~114 copies in the ddPCR assay. Because these values are well above the limit of sensitivity, this confirms that absence of the mutant allele was due to non-inheritance and not a failure to detect the allele as a result of low fetal fraction. The risk of the baby developing neonatal diabetes was, therefore, reduced to that of the population, and non-inheritance of the KCNJ11 p.R201C mutation was confirmed by direct Sanger sequencing of a DNA sample extracted from cord blood soon after the baby’s birth.

Discussion

We describe the application of a ddPCR assay for non-invasive prenatal genetic testing in the offspring of a father.
heterozygous for a KCNJ11 p.R201C mutation causing neonatal diabetes. Our assay showed that the fetus had not inherited the mutation and this result was confirmed after birth. This is the first report of prenatal diagnosis using cfDNA in a family with neonatal diabetes.

Early diagnosis of neonatal diabetes is essential to provide the most appropriate treatment, because most individuals with mutations in the potassium channel genes achieve improved glycaemic control upon switching from insulin to sulfonylurea therapy [5,6]. Furthermore, early treatment with high-dose sulfonylureas is likely to improve neurological development in the 20% of individuals with neurological features. In the case we report, the fetus had not inherited the mutation and we could reassure the parents that the risk of developing neonatal diabetes was reduced to that of the population. This couple had sought prenatal testing in order to be prepared for the birth of a baby likely to develop diabetes during the first weeks of life. If the fetus had inherited the mutation, the baby’s blood glucose would have been closely monitored and sulfonylurea therapy initiated promptly.

In this case, the father was affected with neonatal diabetes, so cfDNA analysis of the KCNJ11 mutation was focused on detection of an allele that was not present in the maternal circulating DNA. Detection of mutations that are maternally...
inherited is more challenging because of the background maternal genome confounding the analysis. Improvement of cfDNA prenatal testing using highly sensitive techniques such as ddPCR may overcome this difficulty, allowing reliable testing for maternally inherited mutations. Prenatal testing is particularly relevant for the optimal management of diabetes in pregnant women with a mutation in one of the genes ( KCNJ11 or ABCC8) encoding the potassium channel subunits. Previous studies have shown that sulfonylureas can cross the placenta and influence fetal growth, potentially resulting in macrosomia and neonatal hyperinsulinaemia in babies who do not inherit the mutation [14].

Knowledge of the fetal genotype will influence both prenatal and postnatal management of pregnancies in which one parent is affected with neonatal diabetes. We illustrate how non-invasive prenatal testing using cfDNA allowed accurate and timely diagnosis for a pregnancy at risk of neonatal diabetes due to a paternal mutation. Although the clinical impact of this novel technology needs to be formally assessed, we anticipate that its implementation in clinical practice (including cases at risk of inheriting maternal mutations) will likely provide clinical benefits for families affected by monogenic diabetes.

Funding sources

ATH and SE are the recipients of a Wellcome Trust Senior Investigator award and ATH is employed as a core member of staff within the NIHR funded Exeter Clinical Research Facility. EDF is a Naomi Berrie Fellow in Diabetes Research.

Competing interests

None declared.

Acknowledgements

The authors thank the family for participating in this study.

References

1 Iafusco D, Massa O, Pasquino B, Colombo C, Iughetti L, Bizzarri C et al. Minimal incidence of neonatal/infancy onset diabetes in Italy is 1:90,000 live births. Acta Diabetol 2012; 49: 405–408.

2 Babenko AP, Polak M, Cave H, Busiah K, Czernichow P, Scharfmann R et al. Activating mutations in the ABCC8 gene in neonatal diabetes mellitus. N Engl J Med 2006; 355: 456–466.

3 Gloyn AL, Pearson ER, Antcliff JF, Proks P, Bruining GJ, Slingerland AS et al. Activating mutations in the gene encoding the ATP-sensitive potassium-channel subunit Kir6.2 and permanent neonatal diabetes. N Engl J Med 2004; 350: 1838–1849.

4 Busiah K, Drunat S, Vavrre-Douret L, Bonnefond A, Simon A, Flechtern I et al. Neuropsychological dysfunction and developmental defects associated with genetic changes in infants with neonatal diabetes mellitus: a prospective cohort study [corrected]. Lancet Diabetes Endocrinol 2013; 1: 199–207.

5 Pearson ER, Flechtner I, Njolstad PR, Malecki MT, Flanagan SE, Larkin B et al. Switching from insulin to oral sulfonylureas in patients with diabetes due to Kir6.2 mutations. N Engl J Med 2006; 355: 467–477.

6 Rafiq M, Flanagan SE, Patch AM, Shields BM, Ellard S, Hattersley AT. Effective treatment with oral sulfonylureas in patients with diabetes due to sulfonylurea receptor 1 (SUR1) mutations. Diabetes Care 2008; 31: 204–209.

7 Shimomura K, Horster F, de Wet H, Flanagan SE, Ellard S, Hattersley AT et al. A novel mutation causing DEND syndrome: a treatable channelopathy of pancreas and brain. Neurology 2007; 69: 1342–1349.

8 Slingerland AS, Hurkx W, Noordam K, Flanagan SE, Jukema JW, Meiners LC et al. Sulphonylurea therapy improves cognition in a patient with the V59M KCNJ11 mutation. Diabetic Med 2008; 25: 277–281.

9 De Franco E, Flanagan SE, Houghton JA, Lango Allen H, Mackay DJ, Temple IK et al. The effect of early, comprehensive genomic testing on clinical care in neonatal diabetes: an international cohort study. Lancet 2015; 386: 957–963.

10 Norton ME, Jacobsson B, Swamy GK, Laurent LC, Randini AC, Brar H et al. Cell-free DNA analysis for noninvasive examination of trisomy. N Engl J Med 2015; 372: 1589–1597.

11 Lun FM, Chiu RW, Chan KC, Leung TY, Lau TK, Lo YM. Microfluidics digital PCR reveals a higher than expected fraction of fetal DNA in maternal plasma. Clin Chem 2008; 54: 1664–1672.

12 Chiu RW, Chan KC, Gao Y, Lau VY, Zheng W, Leung TY et al. Noninvasive prenatal diagnosis of fetal chromosomal aneuploidy by massively parallel genomic sequencing of DNA in maternal plasma. Proc Natl Acad Sci USA 2008; 105: 20458–20463.

13 Lun FM, Tsui NB, Chan KC, Leung TY, Lau TK, Charoenwan P et al. Noninvasive prenatal diagnosis of monogenic diseases by digital size selection and relative mutation dosage on DNA in maternal plasma. Proc Natl Acad Sci USA 2008; 105: 19920–19925.

14 Myngheer N, Allegaert K, Hattersley A, McDonald T, Kramer H, Ashcroft FM et al. Fetal macrosomia and neonatal hyperinsulinemic hypoglycemia associated with transplacental transfer of sulfonylurea in a mother with KCNJ11-related neonatal diabetes. Diabetes Care 2014; 37: 3333–3335.