Differential outcomes of novel plant-herbivore associations between an invading planthopper and native and invasive Spartina cordgrass species

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Differential outcomes of novel plant-herbivore associations between an invading planthopper and native and invasive Spartina cordgrass species

Claire Harkin1 · Alan J. A. Stewart1

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Abstract
Non-native plants may benefit, briefly or permanently, from natural enemy release in their invaded range, or may form novel interactions with native enemy species. Likewise, newly arrived herbivores may develop novel associations with native plants or, where their hosts have arrived ahead of them, re-establish interactions that existed previously in their ancestral ranges. Predicting outcomes from this diversity of novel and re-established interactions between plants and their herbivores presents a major challenge for invasion biology. We report on interactions between the recently arrived invasive planthopper Prokelisia marginata, and the multi-ploidy Spartina complex of four native and introduced species in Britain, each representing a different level of shared evolutionary history with the herbivore. As predicted, S. alterniflora, the ancestral host, was least impacted by planthopper herbivory, with the previously unexposed native S. maritima, a nationally threatened species, suffering the greatest impacts on leaf length gain, new leaf growth and relative water content. Contrary to expectations, glasshouse trials showed P. marginata to preferentially oviposit on the invasive allododecaploid S. anglica, on which it achieved earlier egg hatch, faster nymphal development, larger female body size and greatest final population size. We suggest P. marginata is in the process of rapid adaptation to maximise its performance on what is now the most abundant and widespread host in Britain. The diversity of novel and re-established interactions of the herbivore with this multi-ploidy complex makes this a highly valuable system for the study of the evolutionary ecology of plant–insect interactions and their influence on invasion dynamics.

Keywords Prokelisia marginata · Biological invasions · Plant–insect interactions · Polyploidy · Enemy release

Introduction
Biological invasions are recognised as one of the primary drivers of biodiversity loss, responsible for significant ecological and economic costs worldwide (Mack et al. 2000; IPBES 2019). Most ecological communities now contain at least one non-native species, with invaders already representing over a fifth of many countries’ flora (Mooney and Cleland 2001). At least 13,168 species of vascular plant are known to have become naturalised outside their native range, with almost 5000 of them causing harm to the environment, the economy or human health (RBG Kew 2016).

A popular explanation for the success of invasive plants is that they often arrive in their new range without the full suite of natural enemies (herbivores, fungi and other pathogens) with which they have co-evolved (Maron and Vila 2001; Keane and Crawley 2002). Newly arrived plants may benefit from such natural enemy release, briefly or permanently, or they may form novel interactions with native enemy species in the new range. Likewise, newly arrived herbivores may develop novel associations with native plants or, where their hosts have arrived ahead of them, re-establish interactions that existed previously in their ancestral ranges. Predicting the outcomes from such a diversity of novel and re-established interactions between plants and their herbivores presents a major challenge for invasion biology (Chun et al.
One frequently encountered circumstance of particular interest concerns where a non-native introduced plant is reunited with its herbivore after temporarily benefitting from natural enemy escape in its new range. This may happen because the herbivore arrives naturally or anthropogenically, the latter being either accidental or as a deliberate attempt at biocontrol. The strength and character of the re-established association between plant and herbivore may differ from that found in their native range due to the influence of a suite of biotic and abiotic factors (Mitchell et al. 2006). As the plant and herbivore share a long evolutionary history prior to their introduction into the new range, the prediction is that the plant in its non-native environment will be less severely affected by the herbivore than native congeners with no prior exposure. Indeed, some studies show reassociation with historic enemies can result in levels of plant defence greater than those displayed in their shared native range (Zangerl and Berenbaum 2005; Lu and Ding 2012), although evidence that greater defence results in greater comparative performance remains equivocal (Chun et al. 2010). Due to the greater length of shared evolutionary time in which reciprocal adaptations have been able to develop, the herbivore is predicted to perform better on its coevolved host compared to on congeneric natives. Coevolved hosts have been shown to support a greater abundance and diversity of insect herbivores, with significant host discrimination by phloem-feeding insects persisting despite the assumed palatability of novel alternatives (Burghardt and Tallamy 2013). Such preferences are frequently correlated with greater performance outcomes (Gripenberg et al. 2010).

A further complication may arise if ploidy levels differ amongst sympatric congeneric species. Polyploidy is widespread in plants (Ramsey and Schemske 1998), especially grasses (Stebbins 1956), and in particular occurs at elevated frequencies amongst invasive plants compared to angiosperms in general (Prentis et al. 2008; Pandit et al. 2011; te Beest et al. 2012). Fitness differences between populations with different ploidy levels have been reported in some species complexes (Solits and Solits 2000; Prentis et al. 2008; Pandit et al. 2014), but the effects of polyploidy on plant-animal interactions remains relatively unexplored (Thompson et al. 2004; Munzbergoa 2006; Munzbergoa et al. 2015). Ploidy has been shown to affect the level of damage that herbivores exert on con specifics (Lou and Baldwin 2003). In many systems, higher-level cytotypes are subject to increased attack, however this is not universal and the preferred ploidy level can differ between even very closely-related herbivore species (Munzbergoa 2006; Hal verson et al. 2008; Segraves and Anneberg 2016). Reciprocal effects may also be evident, with consequent impacts on herbivore success resulting in further uncertainty in predicting the outcome of biological invasions mediated by plant–herbivore interactions (Hull-Sanders et al. 2009). This may necessitate system-by-system investigations to inform management interventions.

The introduction of the cordgrass Spartina alterniflora Loiseleur to Britain from North America, its hybridization with a threatened native congeneric species, including a chromosomal doubling event, and subsequent re-connection in the new range with its ancestral herbivore, the planthopper Prokelisia marginata Van Duzee, provides a unique opportunity to test these predictions. Here, we compare the impact of exposure to P. marginata on four species of Spartina in Britain with different histories of co-occurrence with the herbivore: the introduced ancestral host S. alterniflora, the native and previously unexposed S. maritima, the homoplod hybrid of these two species, S. x townsendii, and the allo dodecaploid S. anglica which arose from a chromosomal doubling of S. x townsendii. Also, we investigate whether P. marginata makes a preferential choice between the host species, and the impact that these host species have on P. marginata performance and fitness outcomes. We hypothesise that: (1) P. marginata will preferentially select S. alterniflora, the species with which it has the longest shared evolutionary history, for feeding and oviposition; (2) host plant species will have a significant impact on P. marginata life history traits, with the planthopper achieving the greatest performance outcomes when raised on species with which it has the longest shared evolutionary history; (3) exposure to P. marginata feeding and oviposition will have a deleterious impact on all species of Spartina, but the severity of impact will be related to the extent of shared evolutionary history, i.e. least for S. alterniflora and greatest for S. maritima; and (4) the level of impact will not differ significantly between S. x townsendii and S. anglica, despite their differing ploidy levels, because they share identical evolutionary histories with the planthopper and because S. anglica originated from a chromosomal doubling of S. x townsendii without the introduction of additional genetic material.

Materials and methods

Study system

Spartina is a genus of perennial rhizomatous polyploid C₄ grasses containing around fifteen species, including a number of hybrids (Ainouche et al. 2009; Strong and Ayres 2013; Bortolus et al. 2019). Most Spartina species are primary colonists of intertidal mud flats and have been intentionally introduced to many parts of the world due to their ability to trap sediment and thereby stabilise eroding shorelines, reclaim land and provide defence against extreme coastal weather events (Callaway and Josselyn 1992).
Four species of *Spartina* are present in Britain, only one of which, *S. maritima* (Curtis) Fernald, is ancestrally native. Populations of the non-native *S. alterniflora* are presumed to have been established from seeds transported in shipping ballast from the eastern coast of North America in the early nineteenth century (Thompson 1991). Interspecific hybridization of *S. alterniflora* with *S. maritima* resulted in the sterile homoploid hybrid *Spartina x townsendii* Groves, first described in 1880 from samples collected at Hythe, Hampshire on the south coast of England (Groves and Groves 1880). Fertile plants were first recorded in nearby Lymington in 1892 which appeared to have resulted from chromosome doubling in *S. x townsendii* (Marchant 1967) and were later described as the new fertile allododecaploid species *S. anglica* Hubbard (Hubbard et al. 1968) (Fig. 1). *S. anglica* rapidly colonised British coasts through natural dispersal both of seeds and rhizomes and by deliberate introduction for saltmarsh reclamation (Thompson 1991), and is now the dominant structuring species in a quarter of Britain’s lower saltmarsh communities (Gray et al. 1997). The three progenitor species all still occur in extremely small and localised populations, but *S. maritima* is listed as a species “of principal importance for the purpose of conserving biodiversity” under section 41 of the UK Natural Environment and Rural Communities Act 2006 and a priority species under the UK Biodiversity Action Plan (Joint Nature Conservation Committee 2007). Understanding the potential impacts of a newly arrived specialist herbivore is therefore of great importance for the conservation management of this species.

The planthopper *P. marginata* is native to the Gulf and Atlantic coasts of North America where it is the most abundant herbivore of *S. alterniflora*, frequently reaching densities exceeding 1000 adults/m² and 100,000 nymphs/m² (Denno et al. 1986). Despite their high densities, *P. marginata* have been found to have only weakly adverse effects on *S. alterniflora* where the native ranges of both species coincide (Gustafson et al. 2006; Roberts and Pullin 2008) as well as in invaded ranges where both have coexisted for several decades (Daehler and Strong 1995). However, *S. alterniflora* populations lacking a recent history of co-occurrence with *P. marginata* suffered significant reductions in growth and survival when re-exposed to the herbivore (Daehler and Strong 1997; Wu et al. 1999; Garcia-Rossi et al. 2003).

Across its North American range, *P. marginata* has been reported to feed only on *S. alterniflora*, *S. foliosa* and their hybrids, including the introduced *S. anglica*, whilst avoiding the sympatrically-occurring *S. patens*, *S. cynosuroides* and *S. bakeri* (Denno et al. 1996), the latter occupying a more distantly related clade (Baumel et al. 2002b). No-choice host specificity tests showed that *P. marginata* consistently achieved the greatest survival rates on *S. alterniflora* compared to the other three *Spartina* hosts and was unable to complete a full life cycle on any of twenty other plant species, including other *Spartina* spp., closely- and...
distantly-related monocotyledons and one dicotyledonous species (Grevstad et al. 2003). nymphs raised on more nutritious host plants eclose as significantly larger adults, which in females is positively correlated with daily fecundity. In contrast, high population density has been shown to reduce survivorship, decrease body size and delay nymphal development, consequently increasing the age of first reproduction and reducing lifetime realized fecundity (Denno and McCloud 1985). Nymphal emergence, development and survivorship are negatively impacted by poor quality or unsuitable hosts (Garcia-Rossi et al. 2003; Grevstad et al. 2003), whilst highly nutritious hosts moderate the fitness-reducing impacts of crowding (Denno et al. 1986).

*P. marginata* is presumed to have arrived in Britain sometime since 2000, with preliminary studies suggesting it is in the early stages of successful invasion (Harkin and Stewart 2020). Across its introduced continental European range, *P. marginata* has been recorded primarily on *S. anglica* or *S. x townsendii* (de Blauwe 2011), with a single Slovenian population recorded on *S. maritima* (Seljak 2004). Prior to the work presented here, *S. anglica* was the only recorded host for *P. marginata* in its British range (Badmin 2013; Harkin and Stewart 2020).

*Spartina* spp. and *Prokelisia marginata* experimental material

*Spartina* spp. source populations were identified at four sites along the south coast of England: *S. alterniflora*—Hythe (50°86'N, 1°39'W); *S. anglica*—Pagham (50°77'N, 0°78'W); *S. maritima*—Hayling Island (50°83'N, 0°97'W); and *S. x townsendii*—Beaulieu Estate (50°77'N, 1°40'W). Due to the extremely sparse and localised distribution of all populations apart from *S. anglica*, it was not possible to collect sufficient quantities of more than one species from the same site. *Spartina* spp. plants were grown from sampled rhizome material that had been washed, cut to approximately 12 cm lengths including at least one node and planted in 10 cm (then later transferred to 15 cm) diameter pots containing horticultural grade silver sand. Pots were watered with fresh water and kept continually wet but not inundated (following Denno et al. 2000), with the addition of 100% Hoagland nutrient solution (Hoagland and Arnon 1950) fortnightly. Plants were grown under glasshouse conditions with supplementary lighting (100 W Supanova LED grow lights, 8:2 light ratio comprising 660 nm Red and 430 nm Blue) on an 18:6 h light:dark cycle. Plants were acclimated to glasshouse conditions for 16 weeks prior to the start of experiments.

*P. marginata* individuals used in glasshouse experiments were drawn from a breeding culture maintained on clusters of potted *S. anglica* plants grown under glasshouse conditions. The culture was initiated using *S. anglica* plants removed from Hythe showing brown markings indicative of *P. marginata* oviposition. New plants were added to the culture as required to maintain a consistent supply of host plant material. Second generation glasshouse-reared insects were utilised for the experiments.

**P. marginata oviposition choice in Spartina sp. field assay**

Ten potted plants of each of the four *Spartina* species were transported to the *Spartina*-dominated marsh at Hythe, a site previously shown to have an established *P. marginata* population (Harkin and Stewart 2020). *Hythe* is considered to be the site of origin for both *Spartina x townsendii* and *Spartina anglica* (Raybould et al. 1991), although it is no longer possible to locate the former at the site. *S. alterniflora* is still present in a monospecific stand of approximately 125 m$^2$, with the remainder of the marsh populated by *S. anglica* (Renny-Byfield et al. 2010). Experimental plants were arranged in ten groups, with each group containing one individual of each species. Groups were randomly distributed in an area of established saltmarsh dominated by *S. anglica* measuring 20 m × 30 m, with a minimum of 1.5 m between each group. Within each group, plants were maintained in separate pots, arranged 10 cm apart in a 2 × 2 grid. Each group of pots was buried so that the tops were level with the surrounding substrate. After 24 days, all leaf material of the experimental plants was removed, measured for leaf length and examined under a dissecting microscope for *P. marginata* eggs. *P. marginata* egg density in each plant was expressed as the number per cm of combined lengths of all leaves.

**P. marginata oviposition choice between Spartina sp. under glasshouse conditions**

Ten plants of each *Spartina* species were randomly assigned to one of ten groups, each containing a single plant of each species rooted in separate pots. Each group was enclosed in a cylindrical PET polyester cage as before, and eight female and four male adult *P. marginata* were introduced to the centre of each cage. After 14 days, leaf measurements and a count of *P. marginata* eggs were used to calculate the number of eggs per cm of combined leaf length for each *Spartina* species.

**Impact of Spartina sp. on P. marginata development times under glasshouse conditions**

Two male and two female newly-emerged adult *P. marginata* were caged on each of twelve plants of each of the four *Spartina* species. Cages were monitored for 64 days to determine the date of first egg hatch, the date of first adult emergence and the date by which all adults had emerged. At the end
of the experimental period, all *P. marginata* remaining in each cage were counted and the above-ground material of all plants was harvested by cutting at ground level, dried and weighed.

**Impact of Spartina sp. and crowding densities on *P. marginata* body size under glasshouse conditions**

For each of the four *Spartina* species, five caged plants were randomly assigned to each of three planthopper crowding treatment levels: inoculation with ten (low), thirty (medium) or fifty (high) first-instar *P. marginata* nymphs. After 64 days, above-ground plant material was harvested, dried and weighed, and the body size of all *P. marginata* adults was measured as the distance from the anterior margin of the head to the tip of the abdomen.

**P. marginata impact on Spartina species**

Twenty plants of each *Spartina* species were randomly assigned in equal numbers to ‘herbivore’ and ‘control’ treatments. The following starting metrics were recorded for each plant: number of leaves; overall height; length of each leaf. Each plant was enclosed by a transparent cylindrical cage constructed from 175 μm PET polyester film, 13 cm in diameter, 50 cm tall and with a nylon gauze lid and a 5 × 7 cm gauze-covered ventilation window positioned 18 cm above the base of the cage. Replicates allocated to the herbivore treatment were inoculated with 30 s instar *P. marginata* nymphs, whilst those in the control group were maintained free of planthoppers. Individual plants were arranged in a randomized block design and maintained within a glasshouse for 8 weeks. At the end of the experimental period, all *P. marginata* adults and nymphs were individually removed and counted. Repeat metrics were recorded for each plant. Plants were then weighed to an accuracy of 0.01 g using a Precisa 125A balance, dried for 72 h at 70 °C in a Gallenkamp OV-420 drying oven and finally re-weighed to establish relative water content.

**Statistical analyses**

Statistical analyses were performed with R version 3.1.3 (R Core Team 2015) using the nlme (Pinheiro et al. 2012), lme4 (Bates et al. 2012), effects (Fox 2003) and multcomp (Hothorn et al. 2008) packages. Appropriate models were determined by the nature of the response variables. For the impact of *P. marginata* exposure on continuous plant measurements, ANOVA was used with change in the plant metric as the response variable and treatment, species and their interaction as explanatory variables. Oviposition choice data were analysed using a linear-mixed effect model (LMM). Other data were analysed with GLMMs using a Poisson distribution for count data and a binomial distribution for binary response variables. Each analysis began by fitting all relevant explanatory variables, interactions and random factors (block or plant IDs) in a maximal model. Model simplification then proceeded by a backwards deletion of non-significant terms until further removals led to a significant (*p* < 0.05) increase in deviance. This was assessed by comparing the model with and without the term in question using log-likelihood ratio tests for LMMs and *χ*² values for GLMMs. Plots of model residuals against fitted values were visually inspected for normal distribution, homogeneity of variance and the presence of influential outliers. Results showing significant treatment effects were further investigated using Tukey HSD (Honest Significant Differences) post-hoc tests to identify differences between treatment means.

**Results**

Oviposition rates varied significantly among host plant species in both the field (*L* = 7.87, *p* = 0.049) and glasshouse (*L* = 26.85, *p* < 0.001) experiments (Fig. 2). Under field conditions, 75% fewer eggs were laid on *S. maritima* than *S. alterniflora*, but there were no significant differences in any other two-species comparisons. In the glasshouse experiment, *P. marginata* laid significantly more eggs in *S. anglica* compared to all other species.

In glasshouse choice experiments, plant species had a significant effect on the final number of *P. marginata* adults per plant (*L* = 15.58, *p* = 0.001), with the mean number of individuals on *S. anglica* between two and five times greater than on any other species. The mean number of *P. marginata* per gram of dry *Spartina* biomass was also much greater on *S. anglica*, although the overall difference between host plant species was not statistically significant (*L* = 5.26, *p* = 0.153) (Fig. 3).

There was a significant effect of host plant species on mean time to first adult emergence (*χ*² = 8.29, *p* = 0.04), which was at least 3 days shorter for individuals reared on *S. anglica* compared to those on *S. alterniflora* or *S. maritima*. Time to first egg hatch was also shortest for eggs laid on *S. anglica*, although the overall effect of host plant species was not significant (*χ*² = 1.37, *p* = 0.714) (Table 1).

Both host plant species (*χ*² = 30.43, *p* < 0.001) and levels of crowding (*χ*² = 12.07, *p* = 0.002) had a significant impact on the body size of female, but not male, *P. marginata*. The mean body length of females raised in the high crowding treatment was 6.4% lower than that of females raised in the low crowding treatment across all host species. The interaction between host plant species and crowding level was not significant (*χ*² = 9.6, *p* = 0.143), however post-hoc tests showed that females reared on *S. alterniflora* and *S. anglica*
were significantly larger than those reared on the other species (Fig. 4).

Feeding by *P. marginata* significantly reduced height and leaf length relative growth rates and relative above ground water content for all host species, but, with the exception of relative water content, the impact was least severe for *S. alterniflora*. *S. maritima* was most severely impacted by *P. marginata* exposure in terms of relative leaf length gain (reduced by 65% in comparison to control means) and relative water content (reduced by 70%), but not for relative height gain, for which *S. anglica* suffered the greatest comparative reduction (79%). Planthopper

Fig. 2 Mean number of *P. marginata* eggs laid per cm of *Spartina* spp. leaf length in i field and ii glasshouse experiments. Species abbreviations refer to: *S. alterniflora* (SAlt), *S. anglica* (SAng), *S. maritima* (SMar) and *S. x townsendii* (SxTo). Treatments sharing lower case letters are not significantly different from each other (Tukey HSD post-hoc test, using 95% CI). Error bars show means ± 1 S.E.M.

Fig. 3 Mean number of *P. marginata* i per plant and ii per gram of *Spartina* dry biomass reared from four species of *Spartina* under glasshouse conditions. Species abbreviations as per Fig. 1. Error bars show means ± 1 S.E.M. Treatments sharing lower case letters are not significantly different from each other (Tukey HSD post-hoc test, using 95% CI)
exposure also reduced the number of new leaves by as much as 66%. This metric did not vary significantly between plant species, however the impact was greatest for *S. maritima* and least severe for *S. alterniflora* (Table 2).

**Discussion**

Previous studies have examined the impact of *P. marginata* herbivory only on *S. alterniflora*, its native host, as well as *S. anglica* populations that were introduced to Puget Sound, Washington, in 1961. *S. alterniflora* populations that had been separated from *P. marginata* for over 100 years and *S. anglica* populations with no prior experience of the planthopper suffered significant detrimental impacts when exposed to the herbivore (Daehler and Strong 1997; Wu et al. 1999; Garcia-Rossi et al. 2003; Grevstad et al. 2003). Our work extends investigation of the planthopper’s impacts to a unique species complex of native and introduced host species in the novel geographical context of Britain: an introduced species (*S. alterniflora*), a native species (*S. maritima*), a sterile homoploid hybrid between the two (*S. x townsendii*) and a fertile allododecaploid species arising from the last of these due to a chromosomal doubling event (*S. anglica*).

Our first prediction was that *P. marginata* would preferentially oviposit on *S. alterniflora*. Planthopper oviposition was found to be significantly affected by plant species, but results differed between experiments done under field compared to glasshouse conditions. As predicted, field planthopper populations showed a significant preference for *S. alterniflora* over *S. maritima*, although there were no other significant pairwise differences. Under controlled glasshouse conditions, treatments sharing lower case letters are not significantly different from each other (Tukey HSD post-hoc test, using 95% CI).
conditions, *P. marginata* unexpectedly showed a significant preference for *S. anglica* over all other species. It should be noted, however, that all planthoppers used in the experiments were cultured on *S. anglica*, raising the possibility of prior conditioning to this host species (Coaker and Cheah 1993). However, as *S. anglica* is now the most abundant, widespread and dominant *Spartina* species in Britain, this is increasingly likely to reflect the reality of field conditions (Harkin and Stewart 2020).

Our second hypothesis predicted that *P. marginata* would achieve the greatest performance outcomes on *S. alterniflora*, the host to which it is expected to be best adapted by virtue of their shared evolutionary history, and on which it has previously been shown to achieve the greatest survival in no-choice host specificity tests (Denno et al. 1986; Grevstad et al. 2003). Empirical evidence from our no-choice host tests offers some support for this prediction; however, overall performance outcomes were again unexpectedly greatest on *S. anglica*, the host species most favoured by ovipositing females under controlled glasshouse conditions. Nymphal development was significantly faster for individuals raised on *S. anglica* than for those raised on *S. alterniflora* and *S. maritima*, and eggs laid on *S. anglica* hatched earlier than those laid on any other species (although differences here

### Table 2: Effect of *P. marginata* exposure, *Spartina* species and their interactions on plant performance measured as: height; total leaf length; and relative shoot water content

|                      | d.f | Control (C) mean (SEM) | Treatment (T) mean (SEM) | T:C ratio | Test statistic | p     |
|----------------------|-----|------------------------|--------------------------|-----------|----------------|-------|
| Relative height gain |     |                        |                          |           |                |       |
| Treatment            | 1   | 58.74*                 | < 0.001***               |           |                |       |
| Species              | 3   | 30.33*                 | < 0.001***               |           |                |       |
| *S. alterniflora*    |     | 0.45 (0.07)            | 0.33 (0.05)              | 0.73      |                |       |
| *S. anglica*         |     | 0.28 (0.09)            | 0.06 (0.02)              | 0.21      |                |       |
| *S. maritima*        |     | 0.94 (0.08)            | 0.48 (0.07)              | 0.51      |                |       |
| *S. x townsendii*    |     | 0.59 (0.09)            | 0.14 (0.02)              | 0.23      |                |       |
| Treatment x species  | 3   | 3.08*                  | 0.033*                   |           |                |       |
| Relative leaf length gain |     |                        |                          |           |                |       |
| Treatment            | 1   | 100.88*                | < 0.001***               |           |                |       |
| Species              | 3   | 12.21*                 | < 0.001***               |           |                |       |
| *S. alterniflora*    |     | 1.69 (0.11)            | 1.08 (0.09)              | 0.64      |                |       |
| *S. anglica*         |     | 1.16 (0.10)            | 0.60 (0.07)              | 0.51      |                |       |
| *S. maritima*        |     | 2.59 (0.30)            | 0.91 (0.11)              | 0.35      |                |       |
| *S. x townsendii*    |     | 1.96 (0.19)            | 0.89 (0.12)              | 0.45      |                |       |
| Treatment x species  | 3   | 4.31*                  | 0.007**                  |           |                |       |
| Relative water content |     |                        |                          |           |                |       |
| Treatment            | 1   | 303.24*                | < 0.001***               |           |                |       |
| Species              | 3   | 5.96*                  | 0.001***                 |           |                |       |
| *S. alterniflora*    |     | 1.82 (0.04)            | 1.02 (0.11)              | 0.46      |                |       |
| *S. anglica*         |     | 1.63 (0.09)            | 0.89 (0.07)              | 0.55      |                |       |
| *S. maritima*        |     | 1.84 (0.07)            | 0.56 (0.03)              | 0.30      |                |       |
| *S. x townsendii*    |     | 1.61 (0.06)            | 0.58 (0.11)              | 0.36      |                |       |
| Treatment x species  | 3   | 4.85*                  | 0.004**                  |           |                |       |
| No. new leaves gained |     |                        |                          |           |                |       |
| Treatment            | 1   | 78.37b                 | < 0.001***               |           |                |       |
| Species              | 3   | 6.27b                  | 0.099                    |           |                |       |
| *S. alterniflora*    |     | 8.2 (0.55)             | 4.5 (0.82)               | 0.55      |                |       |
| *S. anglica*         |     | 7.5 (0.58)             | 2.7 (0.58)               | 0.36      |                |       |
| *S. maritima*        |     | 7.9 (0.84)             | 2.7 (0.50)               | 0.34      |                |       |
| *S. x townsendii*    |     | 9.3 (0.98)             | 4.1 (0.60)               | 0.44      |                |       |
| Treatment x species  | 3   | 3.44b                  | 0.329                    |           |                |       |
| Residuals (all models) | 72 |                        |                          |           |                |       |

Relative metrics indicate ratios of post- to pre-experiment measurements. Mean values are given for each host species ± 1 S.E.M. Treatment refers to 30 *P. marginata* individuals added; Control refers to no *P. marginata*. Test statistics are (a) *F* values for two-way ANOVAs and (b) *χ*² for GLMs. Significance levels indicated by: * ≤ 0.05; ** ≤ 0.01; *** ≤ 0.001
were not statistically significant). Faster development times are expected to confer fitness benefits because individuals more rapidly escape the elevated mortality risks associated with the vulnerable egg and nymph stages (Stiling and Strong 1982), whilst potentially increasing their lifetime realised fecundity as a consequence of achieving first reproduction earlier (Denno 1985). *P. marginata* eclosed as larger adults when raised on nutritionally superior hosts (Denno et al. 1986) and there is a positive correlation between body size and the average daily fecundity of females (Denno and McCloud 1985). In the experiments reported here, females were significantly larger when raised on *S. alterniflora* and *S. anglica*, although host plant species did not appear to mitigate the significant negative effects of crowding on body size. Greater fecundity and survivorship were realised on *S. anglica* with significantly larger final populations achieved per host plant. In concert, the data provide evidence of *P. marginata* achieving relatively small, but significant, improvements in fitness-related performance when utilising *S. anglica* as its host plant. Further investigation is needed to determine the mechanisms underlying this outcome, however the apparently rapid pre-conditioning suggested by our oviposition choice tests indicates that *P. marginata* can quickly develop a preference for the most abundant suitable host and enhanced performance on it. It is possible therefore, that *P. marginata* has undergone, or is in the process of, rapid adaptation to maximise its performance on *S. anglica* in the 15–20 years since its arrival because *S. anglica* is the most abundant and widespread potential host species in Britain (Lacambra et al. 2004; Harkin and Stewart 2020). Further studies comparing British and North American populations of *P. marginata* would be instructive in testing this hypothesis.

The founding population of *S. alterniflora* in Britain can be considered to have shared a long, unbroken, evolutionary history with *P. marginata* prior to its arrival. This founding population would initially have had high levels of defence against the herbivore, as suggested by previous studies in North America (Daehler and Strong 1995; Gustafson et al. 2006), although this may subsequently have eroded due to a lack of herbivore selection pressure in the ~200 years since its arrival. In contrast, *S. maritima* which is native to Europe had no exposure to *P. marginata* prior to the planthopper’s arrival and there are no known equivalent specialist *Spartina* herbivores native to Britain (Payne 1973). Therefore, *S. maritima* might be expected to display a lack of defence against *P. marginata* consistent with that reported for previously unexposed congeners (Wu et al. 1999).

Exposure to *P. marginata* had a significant negative impact on all four species of *Spartina* for all performance metrics, but subtle differences were evident in the severity of impacts between species. Three of the plant traits showed strong effects of exposure to planthopper feeding: relative height and leaf length growth rates, and relative water content. These traits reflect a composite metric of the cost of herbivory, plant compensatory growth and photosynthetic potential, indicative of differential impacts of *P. marginata* herbivory on the four species (Strauss and Agrawal 1999). Due to the sparse and extremely localised distribution of all but *S. anglica*, species identity and collection site are necessary to confound in this study. However, we suggest the abiotic effects associated with collection site will have been minimised by the lengthy period of acclimation to glasshouse conditions prior to the start of experiments.

Our third hypothesis, that the impact of *P. marginata* herbivory would be least severe for *S. alterniflora*, was supported by the results. Even though British populations of the grass are likely to have been separated from the herbivore for ~200 years, *S. alterniflora* remained the least severely impacted of all four species examined. Additionally, we predicted that *S. maritima* would suffer the greatest detrimental impact of exposure to *P. marginata* herbivory and oviposition because it shares no evolutionary history with the planthopper, nor with any other specialist herbivore (Payne 1973). This prediction was only partially supported. *S. maritima* suffered significant negative impacts of exposure on all performance metrics, however the relative impact in comparison to the other *Spartina* species was variable, being the most severely impacted for some traits, but not for others. It is interesting to note that *S. maritima* plants in the control group (i.e. no herbivory) performed substantially better than the control group for any other species, displaying the greatest relative height and leaf length mean growth rates, and the highest relative shoot water content.

In its current British distribution, *S. maritima* is extremely localised, sparsely populated and routinely out-competed by *S. anglica* (Lacambra et al. 2004); in our experiments, plants were grown individually in separate pots and hence freed from competition. *P. marginata* is currently in the early stages of invasion in the UK (Harkin and Stewart 2020). The results presented here suggest that its continued population growth and spread may pose a significant additional threat to the future survival of *S. maritima* across its remaining British distribution. Further experiments directly examining the comparative impacts of herbivory, competition and apparent competition on the interaction between *P. marginata*, *S. anglica* and *S. maritima* would be valuable to inform future management interventions.

We found no support for our final hypothesis: that the negative effect of *P. marginata* exposure would not differ between the allododecaploid *S. anglica* and its hexaploid progenitor *S. x townsendii*, as a result of their shared level of evolutionary history with the herbivore. Treatment means for all traits apart from relative height gain differed between these species, although further work would be needed to determine the role that ploidy levels may
have played in this result. The differences may result from divergent evolutionary changes due to selective adaptation or random processes such as genetic drift (Schluter 2001). *S. anglica* was formed as a separate species ~130 years ago (Gray et al. 1991), but recent studies have shown that significant adaptive evolution can occur in a range of invasive plant species within twenty or fewer generations (Prentis et al. 2008). There is very little inter-individual genetic variation in *S. anglica* populations, consistent with a severe genetic bottleneck due to the unique event which resulted in the origin of the species (Baumel et al. 2001), and very little structural change has been observed in the genomes of either *S. x townsendii* or *S. anglica* (Baumel et al. 2002a). However, considerable epigenetic alteration (Salmon et al. 2005) as well as changes to the transcriptome (Chelaifa et al. 2010) arose from both the hybridization event that led to the formation of *S. x townsendii*, and in the subsequent chromosome doubling which resulted in *S. anglica*. These epigenetic and regulatory changes are thought to account for the high levels of phenotypic plasticity documented for *S. anglica* (Renny-Byfield et al. 2010), and may similarly offer an explanation for the different responses to *P. marginata* that we found in the two species.

Whilst polyploidy remains relatively understudied, it has the potential to significantly impact the diversity and outcomes of plant–herbivore interactions (Segraves and Anneberg 2016). In this context, the multi-ploidy *Spartina* species complex present in Britain represents an excellent model for the study of the impact of ploidy levels on the invasiveness of plant species. The allododecaploid *S. anglica* is of recent origin, the hexaploid progenitors are all still extant, and there is wide agreement on the likely timings of speciation events based on historical records (Gray et al. 1991). The recent imposition of *P. marginata* herbivory on these species, additionally facilitating the study of a diversity of novel and re-connected interactions, further adds to the research value of this system, providing a tractable model for the study of the impact of ploidy levels on the evolutionary ecology of plant–insect interactions, and of how the complexity of these interactions may influence the outcome of invasion dynamics.

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**Author contribution statement** CH and AJAS contributed equally to project design and manuscript preparation. CH led on data collection and analysis.

**Data availability** The data that support the findings of this study are available in the Figshare repository https://doi.org/10.25377/sussex.13034399

**Declarations**

**Ethical approval** All applicable institutional and national guidelines for the care and use of animals were followed.

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