Guayule, a shrub native to the Chihuahuan Desert that produces a variety of secondary compounds, is currently being investigated as a source of hypoallergenic natural rubber latex (Cornish et al., 2001; Ray et al., 2005). Although guayule has been known as a source of high-quality rubber since the beginning of the twentieth century, research on guayule has been very intermittent (Ray et al., 2005). Due to these intermittent research efforts, improvement of guayule has been slow (Dierig et al., 2001; Ray et al., 2005; Thompson and Ray, 1989).

Multiple strategies to improve guayule have been used during the sporadic periods of intensive research. Sexual diploids and a few naturally occurring sexual polyploids of guayule have been used in breeding programs for generation of intraspecific and interspecific hybrids (Esau, 1944; Gardner, 1946; Hashemi et al., 1989; Kuruvadi et al., 1997a; Naqvi et al., 1984). However, the most successful methods have used mass, family, or single plant selections (Ray et al., 1995, 1999, 2005; Thompson and Ray, 1989). A more recent tool for improvement of guayule is transformation (Pan et al., 1996) to insert genes from the rubber biosynthetic pathway into the plant in an attempt to increase rubber concentration and rubber yield (Veatch et al., 2005). This can be problematic because many of these products are also used in other plant pathways (Oh et al., 2000).

The economics of guayule production will improve significantly if higher-yielding lines can be developed using reliable and rapid methods of selecting plants with the best possible traits that will be passed faithfully to their progeny. One prediction method is to examine the linear correlation between different morphological traits. Correlations between traits aid in selection by using nondestructive measurements, such as plant height or width, to indirectly select for traits like rubber and resin content, which would otherwise involve destructive sampling of the plant. For example, rubber yield has been found to be positively correlated with fresh weight, dry weight, height, and width (De Rodriguez et al., 2001; Dierig et al., 1989b; Ray et al., 1993; Thompson and Ray, 1989). On the other hand, rubber concentration generally has a low positive or even a negative correlation with the traits that are positively correlated with rubber yield (Dierig et al., 1989b; Ray et al., 1993).

Successful breeding programs focus on characteristics with high heritabilities, indicating low environmental effects. Heritability estimates of the traits associated with rubber concentration and rubber yield have been variable. In a study by Dierig et al. (2001), variance from measured traits of clones represented environmental variance and were compared with variances from traits of open-pollinated progeny, representing the phenotypic variance (G × E). They found that height, resin concentration, and rubber concentration had high broad-sense (BS) heritabilities at 2 years of growth, but by the third year, BS heritability was almost zero as environmental effects compounded. In another study, using path-coefficient analysis, all traits measured at 1 year were found to be highly heritable in California but much less heritable in Arizona (Estilai et al., 1992). In a third study, when progeny and parents from single plant selections were compared when they were 2 and 3 years of age, respectively, no significant regressions were found between parents and progeny, which is indicative of low heritabilities (Ray et al., 1993). Additionally, resin and rubber concentration have been difficult to predict from year to year, possibly because of large environmental effects (Coffelt et al., 2005; Dierig et al., 1989a).

Improvement of guayule is further complicated because of its relatively long generation time (2–5 years) and its complex genetic and reproductive system (Ray et al., 2005; Thompson and Ray, 1989). Guayule has a haploid chromosome number...
of 18, with a natural ploidy series ranging from diploid to tetraploid or higher (Bergner, 1944; De Rodriguez et al., 1993; Powers, 1945; Thompson and Ray, 1989). In native populations ≈95% of the accessions surveyed are polyploid, with the majority being tetraploid (Kuruvadi et al., 1997a). Diploid guayule plants reproduce sexually, but tetraploid guayule reproduces predominantly by facultative apomixis, which is embryo development without fertilization. However, even in apomictic plants pollination is required for endosperm development (Esau, 1944; Ray et al., 1990). The exact frequency of apomixis in guayule has yet to be determined, but it is known to vary by line and season (Keys et al., 2002; Thompson and Ray, 1989). Sexually reproducing plants are self-incompatible with a sporophytic system of self-incompatibility (Gerstel, 1950; Ray et al., 1993). This system of enforced outcrossing promotes variation within cultivated and native populations.

Most of the guayule germplasm, upon which the present University of Arizona and USDA-ARS breeding program is based, comes from only a few plants (Thompson and Ray, 1989). Despite this apparently narrow genetic base, there appears to be a large amount of variation in cultivated guayule (Ray et al., 2005), not only among lines but also within lines (Dierig et al., 1989a; Naqvi, 1985; Ray et al., 1990). There is also a great deal of unexploited variation available in native populations (De Rodriguez et al., 1993; Kuruvadi et al., 1997b). This variation could be a valuable tool in guayule improvement; however, methods that determine the genetic contribution to this variation and subsequent response to selection need to take into account guayule’s unusual mode of reproduction. The objective of this study was to estimate narrow sense heritability of this variation and subsequent response to selection need to vary by line and season (Keys et al., 2002; Thompson and Ray, 1989). Sexually reproducing plants are self-incompatible with a sporophytic system of self-incompatibility (Gerstel, 1950; Ray et al., 1993). This system of enforced outcrossing promotes variation within cultivated and native populations.

**Materials and Methods**

Open-pollinated seeds, assumed to be a mix of apomictic and sexual progeny, were collected individually from 21 tetraploid parent plants during the first year and a half of growth and used to generate 92 progeny plants. The parent plants consisted of eight plants from line AZ 101, a putative interspecific hybrid of *P. argentatum* × *P. tomentosum* DC; four plants from a germplasm release AZ-2 (PI 599675), selected for interspecific hybrid of *P. argentatum* × *P. tomentosum* DC; four plants from a germplasm release AZ-2 (PI 599675), selected for vigorous growth; and nine plants from a breeding line N6-5. All parents were transgenic and generated from tissue culture. The parents were transgenic for one of three pyrophosphate synthase genes (*farnesyl pyrophosphate synthase, geranylgeranyl pyrophosphate synthase, or hexa-heptaprenyl pyrophosphate synthase*) and neomycin phosphotransferase II as the selectable marker (Veatch et al., 2005). The parents were grown from May 2001 through March 2003 at the University of Arizona Maricopa Agricultural Center (MAC), in Maricopa, Ariz.

The number of progeny germinated from each parent ranged from 1 to 16. The AZ 101 parents had an average number of progeny in each family of 6.8, with 54 total progeny plants. The AZ-2 parents had an average family size of 6.5, with 26 total progeny plants. The N6-5 parents had an average family size of 1.3, with 12 total progeny plants.

Both the parents and the progeny were established in a greenhouse for 3 months before transplantation in the field, so that they were about the same morphological age at the time of transplantation. The progeny were grown at the University of Arizona Maricopa Agricultural Center, Maricopa, from May 2003 through March 2005 in the same field where the parents had grown, in a sandy loam soil. Irrigation of both the parents and progeny was on the same schedule of every 14 d from March to October and every 42 d from October to March. During the growing season the average high and low temperatures were, respectively, 37.7 and 19.4 °C for the parents and 37.6 and 19.3 °C for the progeny. The average high and low temperatures during the dormant period were, respectively, 22.8 and 4.8 °C for the parents and 21.7 and 5.7 °C for the progeny. The parent field was not randomized (Veatch et al., 2005), but the progeny field was set up in a completely randomized design.

Height and width were measured at 1 and 2 years of growth for both experimental sets. Width measurements were taken at the widest point of the plant in two directions perpendicular to each other, and the width measurements were averaged. After 1 year of growth, resin, rubber, and guayulins A and B were extracted from two branches harvested from each plant. At the end of the second year of growth, the aboveground biomass was harvested, the fresh weight was measured, and the plants were run through a chipper. Dry weights were determined by drying either the branches (year 1) or a representative sample of the chipped aboveground biomass (year 2) for 2 d at 60 °C to a constant weight. Dried samples were ground in a coffee grinder, and secondary compounds were extracted as described below. Resin and rubber yields were calculated using their respective concentrations multiplied by the dry weight of each plant.

Resin and rubber concentrations were determined by a modification of the gravimetric method of Black et al. (1983). A 0.5-g sample of the ground plant material was homogenized in 20 mL of acetone for 30 s with a homogenizer at a speed of 27,000 rpm to extract resin. The homogenizer was rinsed with an additional 10 mL of acetone, and the sample was centrifuged at 974g, for 12 min. The supernatant was poured through a solvent-wetted filter paper into a preweighed aluminum dish. The procedure was repeated twice more, and the supernatant from all three extractions was combined. The acetone in the aluminum dish evaporated overnight, and any residual liquid was removed in a vacuum oven for 30 min at 60 °C. After the acetone from the sample residue evaporated overnight, rubber was extracted from it with cyclohexane using the procedure described above for acetone. Resin and rubber concentrations were calculated by subtracting the initial pan weight from the final pan weight all divided by the original sample weight.

Secondary products, guayulin A and B, were extracted from dried resin samples that were made up into ≈10 mg·mL⁻¹ solutions with absolute ethyl alcohol (EtOH) before being analyzed by high-performance liquid chromatography (HPLC). Guayulin quantification on the HPLC was done with HPLC-grade degassed acetonitrile and HPLC-grade H₂O. HPLC was performed using a Hitachi HPLC system (Hitachi, Ltd., Tokyo). Separation was done using a reverse-phase C18 column (5 μm, 250 × 4.6 mm i.d.; Microsorb-MV, Varian, Palo Alto, Calif.), at ambient temperature, with retention times of 24 and 25 min for guayulin B and guayulin A, respectively. Ultraviolet (UV) detection was at 262 nm. Guayulin concentration in the resin was determined by the following formula:

\[
\% \text{guayulin A or B} = \frac{AC \times RF \times mL \text{EtOH}}{mg \text{resin} \times 100,000,000}
\]

where AC = area counts and RF = response factor (RF guayulin A = 1.235 ng/1000 AC, RF guayulin B = 0.867 ng/1000 AC).
Area counts were determined by the Hitachi HPLC System Manager software (version 4.0).

Due to the unequal number of progeny and unequal variance among families, the relationship between parents and progeny was examined using weighted regression where the regression coefficient was calculated using the following formula (Steel et al., 1997):

\[ b = \frac{\sum w_i (X_i - \bar{X}_i)(Y_i - \bar{Y}_i)}{\sum w_i (X_i - \bar{X}_i)^2} \]

where \( X_i \) represents the progeny values, \( Y_i \) represents the progeny values, and \( w_i \) is the weighting factor (1/\( \sigma^2 \)). The weighting factor used was based on the variance of individuals within the same line transformed with the same gene. Regression analysis was done in the Fit Y by X platform of JMP 4.0 academic software (Sall et al., 2001). Rubber concentration at year 2 was \( \log_{10}(\sqrt{1/\% \text{rub}} + 1) \) transformed and guayulin A concentration at year two was \( \log_{10}(\% \text{guayulin A} + 1) \) transformed for regression analysis, to normalize the data. All other data were not transformed for analysis as it was already normally distributed. Narrow-sense heritability was calculated from the regression of the progeny on the maternal parent using a modification of the formula \( h^2 = 2b \) (Falconer and Mackay, 1996). The formula \( (2 - p)b, \) used for calculating heritability, incorporated the frequency of apomixis, where \( p = \) proportion of apomixis and \( b = \) regression coefficient. Upper and lower heritability values were determined using values of \( p = 0 \) (complete sexuality) and \( p = 1 \) (complete apomixis), respectively. Correlations among traits within the progeny and also among the traits of the progeny and the parents were calculated using Spearman’s rank correlation in the Multivariate platform (Sall et al., 2001). A \( P \) value \( \leq 0.05 \) was considered significant throughout.

Table 1. Means ± SE, range, and Spearman’s rank correlation coefficient of traits measured on parent and progeny plants of transgenic guayule at 1 and 2 years of growth.

| Measurement | Parent \((n = 21)\) | Progeny \((n = 92)\) | Parent–progeny correlation |
|-------------|-------------------|-------------------|---------------------------|
| Yr 1        |                   |                   |                           |
| Height (cm) | 54 ± 4 (24–81)    | 53 ± 2 (17–74)    | 0.27*                     |
| Width (cm)  | 54 ± 4 (25–90)    | 56 ± 2 (14–84)    | 0.18                      |
| Resin concn (%) | 5.5 ± 0.3 (3.8–8.6) | 5.6 ± 0.1 (4.1–7.7) | 0.42*                   |
| Rubber concn (%) | 2.3 ± 0.2 (0.9–4.3) | 2.0 ± 0.1 (0.7–3.3) | 0.02                    |
| Guayulin A concn (%) | 0.91 ± 0.39* (0.04–7.34) | 0.21 ± 0.03 (0–1.10) | 0.03                    |
| Guayulin B concn (%) | 0.19 ± 0.05* (0–0.91) | 0.06 ± 0.01 (0–0.29) | –0.03                   |
| Yr 2        |                   |                   |                           |
| Height (cm) | 79 ± 4 (56–114)   | 78 ± 3 (18–120)   | 0.32*                     |
| Width (cm)  | 92 ± 4 (58–132)   | 91 ± 3 (15–133)   | 0.39*                     |
| Resin concn (%) | 7.2 ± 0.4* (5.1–11.1) | 6.5 ± 0.1 (4.7–8.1) | 0.55*                 |
| Rubber concn (%) | 3.4 ± 0.1 (2.3–4.8) | 3.1 ± 0.1 (3.0–5.3) | 0.08                    |
| Guayulin A concn (%) | 0.38 ± 0.09 (0.03–1.55) | 0.43 ± 0.05 (0–3.67) | 0.22*                 |
| Guayulin B concn (%) | 0.06 ± 0.01 (0–0.19) | 0.10 ± 0.01 (0–0.44) | 0.13                    |
| Fresh wt (kg/plant) | 3.2 ± 0.4* (0.9–8.5) | 1.9 ± 0.2 (0.03–6.8) | 0.34*                |
| Dry wt (kg/plant) | 1.5 ± 0.2* (0.4–3.8) | 0.8 ± 0.1 (0.01–2.7) | 0.34*               |
| Resin yield (g/plant) | 114 ± 19* (26–325) | 55 ± 4 (0.7–159) | 0.37*                   |
| Rubber yield (g/plant) | 48 ± 5* (19–106) | 26 ± 2 (0.2–75) | 0.23*                 |

*Parent and progeny means significantly different at \( P \leq 0.05 \) based on ANOVA, and Spearman’s rank correlation coefficient significant at \( P \leq 0.05 \).
Table 2. Regression coefficients \((b) \pm SE\) and range of heritability values of parent and progeny plants of transgenic guayule at 1 and 2 years of growth.

| Measurement | Regression ± SE | Range of heritabilities* |
|-------------|-----------------|--------------------------|
| Yr 1        |                 |                          |
| Height (cm) | 0.30 ± 0.09*    | 0.30–0.60                |
| Width (cm)  | 0.19 ± 0.08*    | 0.19–0.38                |
| Resin concn (%) | 0.13 ± 0.04*   | 0.13–0.26                |
| Rubber concn (%) | −0.04 ± 0.04  | 0–0                      |
| Guayulin A concn (%) | −0.04 ± 0.04 | 0–0                      |
| Guayulin B concn (%) | −0.03 ± 0.03 | 0–0                      |
| Yr 2        |                 |                          |
| Height (cm) | 0.85 ± 0.13*    | 0.85–1.00                |
| Width (cm)  | 0.94 ± 0.16*    | 0.94–1.00                |
| Resin concn (%) | 0.22 ± 0.04*   | 0.22–0.44                |
| Rubber concn (%) | 0.29 ± 0.18    | 0.29–0.58                |
| Guayulin A concn (%) | 0.20 ± 0.15   | 0.20–0.40                |
| Guayulin B concn (%) | 0.18 ± 0.16   | 0.18–0.36                |
| Fresh wt (kg/plant) | 0.40 ± 0.07*  | 0.40–0.80                |
| Dry wt (kg/plant) | 0.41 ± 0.06*  | 0.41–0.82                |
| Resin yield (g/plant) | 0.33 ± 0.04*  | 0.33–0.66                |
| Rubber yield (g/plant) | 0.32 ± 0.08*  | 0.32–0.64                |

*Estimates of heritability were calculated as \((2 – p)b\), with \(p\) = proportion of apomixis. The lower estimate assumed complete apomixis with \(P = 1.0\), and the upper estimate assumed complete sexuality with \(P = 0\).

**Both upper and lower estimates of heritability are zero because the regression coefficient was negative.**

*Regression coefficient significant at \(P \leq 0.05\). The regression coefficient was calculated from a weighted regression using \((1/\sigma^2)\) as the weighting factor.

Discussion

Many crops such as citrus (\(Citrus\ L\).), apple (\(Malus\ Mill\).), and forage grasses, such as \(Eragrostis\ curvula\) Schrad. and \(Panicum\ maximum\) Jacq., share characteristics with guayule that make accurate prediction of heritability difficult, such as long generation time, polyploidy, apomixis, or a combination of these factors (Campbell et al., 1991; Di Renzo et al., 2003; Elisario et al., 1999; Khan and Grosser, 2004; Resende et al., 2004). The two main factors that can affect the accuracy of heritability estimates in guayule are autotetraploidy and apomixis. In allotetraploids, which genetically act as diploids, narrow sense heritability is simply the additive variance \((V_A)\) divided by the phenotypic variance \((V_P)\) and can be estimated using parent–progeny regression, but in an autotetraploid heritability estimates also need to include a fraction of the dominance variance \((V_D)\) (Bernardo, 2002). Swanson et al. (1974) showed that parent–progeny regression provides the best estimate of narrow sense heritability in an autotetraploid, and this method has been used to estimate heritability in other autotetraploids, such as alfalfa (\(Medicago\ sativa\) L.) (Pecetti and Piano, 2005).

Generally, when progeny values are regressed on one parent, narrow-sense heritability can be calculated by multiplying the regression coefficient \((b)\) by 2 (Falconer and Mackay, 1996; Lynch and Walsh, 1998; Nyquist, 1991; Stratton, 1991). In the case of an apomictic species, the frequency of apomixis needs to be included in the heritability calculation. Here we have used the formula \(h^2 = \frac{2 – \pi}{\pi}\), with \(\pi = \) the proportion of apomixis. If the population is completely sexual, then \(p = 0\) and \(h^2 = 2b\); however, if the population is completely apomictic, then \(p = 1\) and \(h^2 = b\), which is the same as parent–progeny regression on the midparent value (Falconer and Mackay, 1996). If the contribution of apomixis is not taken into account heritability, values will be overestimated.

Unfortunately, the actual frequency of apomixis displayed by different guayule lines and the effect of different seasons of the year are unknown (Keys et al., 2002; Ray et al., 1993). Therefore, heritability of traits in guayule is more accurately presented as a range of possible values falling somewhere between the single parent and midparent regression values. Because guayule is assumed to be more apomictic than sexual (Keys et al., 2002; Thompson and Ray, 1989), heritability is likely to be closer to the midparent–progeny regression value.

There are distinct differences in heritability estimates among the different traits examined in this study. Height and width appear to be the most heritable traits. Heritability of fresh and dry weight, resin and rubber yield, and resin concentration is low to moderate (Table 2). The heritability estimates for rubber concentration are in the same range as rubber yield, but the regression was not significant, so it is likely that the heritability of rubber concentration is lower than estimated. Rubber yield is a function of rubber concentration and plant biomass, and our data show a stronger relationship between biomass and rubber yield than between rubber concentration and rubber yield (Tables 2 and 3). Research has shown that there is a substantial effect of the environment on rubber production (Coffelt et al., 2005; Dierig et al., 2001; Veatch-Blohm et al., 2006). It may be that traits with lower heritabilities, such as resin and rubber concentration (Table 2), are sensitive enough to even slight environmental differences that the environmental variance masks the genetic variance of the population.

In addition to the moderate heritabilities estimated for resin and rubber yield in this environment (Table 2), significant correlations of parent height and width with progeny resin and rubber yields were found (Table 4). Because height and width appear to be the most heritable characteristics based on regression (Table 2), it may be possible to increase resin and rubber yields indirectly by selecting for height and width. Another effect of selecting for increased width in the parents is a probable decrease in the progeny of guayulin A concentration, indicated by the negative correlation between these traits.

Table 3. Rank correlation of rubber and resin concentration (%) and rubber and resin yield to various morphological traits at 2 years of growth of progeny plants of transgenic guayule.

| Measurement | Resin yield (%) | Rubber yield (%) | Resin (%) | Rubber (%) |
|-------------|-----------------|------------------|-----------|------------|
| Height      | 0.77*           | 0.77*            | 0.07      | 0.11       |
| Width       | 0.94*           | 0.91*            | 0.07      | 0.14       |
| Fresh wt    | 0.99*           | 0.98*            | 0.06      | 0.19       |
| Dry wt      | 0.99*           | 0.97*            | 0.04      | 0.17       |
| Resin yield | 1.00            | 0.97*            | 0.16      | 0.21*      |
| Rubber yield| 0.97*           | 1.00             | 0.13      | 0.35*      |
| Resin (%)   | 0.16            | 0.13             | 1.00      | 0.41*      |
| Rubber (%)  | 0.21*           | 0.35*            | 0.41*     | 1.00       |
| Guayulin A (%) | −0.06           | 0.01             | 0.07      | 0.32*      |
| Guayulin B (%) | 0.04            | 0.11             | 0.17      | 0.27*      |

*Spearman’s rank correlation coefficient significant at \(P \leq 0.05\).
Table 4. Parent–progeny correlation among all traits in transgenic guayule measured at 2 years of growth.

| Parent measurement | Height | Width | Fresh wt | Dry wt | Resin yield | Rubber yield | Resin (%) | Rubber (%) | Guayulin A (%) | Guayulin B (%) |
|--------------------|--------|-------|----------|--------|-------------|--------------|-----------|-----------|----------------|----------------|
| Height             | 0.32*  | 0.38* | 0.41*    | 0.41*  | 0.44*       | 0.38*        | 0.36*     | 0.10      | –0.10          | –0.04          |
| Width              | 0.38*  | 0.39* | 0.43*    | 0.43*  | 0.45*       | 0.37*        | 0.29*     | 0.01      | –0.20*         | –0.13          |
| Fresh wt           | 0.30*  | 0.32* | 0.34*    | 0.34*  | 0.34*       | 0.26*        | 0.11      | –0.11     | –0.22*         | –0.19          |
| Dry wt             | 0.29*  | 0.32* | 0.34*    | 0.34*  | 0.34*       | 0.26*        | –0.10     | –0.10     | –0.21*         | –0.18          |
| Resin yield        | 0.27*  | 0.32* | 0.34*    | 0.34*  | 0.37*       | 0.29*        | 0.30*     | 0.01      | –0.16          | –0.11          |
| Rubber yield       | 0.21*  | 0.27* | 0.29*    | 0.30*  | 0.32*       | 0.23*        | 0.24*     | –0.03     | –0.16          | –0.12          |
| Resin (%)          | 0.26*  | 0.30* | 0.30*    | 0.31*  | 0.37*       | 0.34*        | 0.55*     | 0.21*     | 0.01           | 0.10           |
| Rubber (%)         | –0.41* | –0.44*| –0.44*   | –0.44* | –0.42*      | –0.40*       | –0.02     | 0.08      | 0.19           | 0.10           |
| Guayulin A (%)     | –0.26* | –0.20*| –0.20*   | –0.14  | –0.10       | –0.11        | 0.14      | 0.15      | 0.22*          | 0.16           |
| Guayulin B (%)     | –0.06  | 0.01  | 0.06     | 0.06   | 0.11        | 0.08         | 0.31*     | 0.17      | 0.15           | 0.13           |

*Spearman’s rank correlation coefficient significant at \( P \leq 0.05 \).

(Table 4). This is important because guayulin A is a potential cause of contact dermatitis (Schloman et al., 1983).

Heritability can be a difficult value to calculate, as its value can change with the environment, and its accuracy may be affected when the reproductive system within the population in question is disregarded. Guayule, many citrus and apple cultivars, and forage grasses reproduce through facultative apomixis. If the population is assumed to be a completely sexual population, heritability estimates may be inflated, which makes it more difficult to predict response to selection and may result in very little improvement in the population overall. When the effect of apomixis is considered, especially when the frequency of apomixis is accurately known, breeders can implement selection strategies with more realistic probability for population improvement.

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