Ginkgo suppresses atherosclerosis through downregulating the expression of connexin 43 in rabbits

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Abstract

Introduction: Ginkgo biloba extract (GBE) EGb761 is widely used for cardiovascular prevention. Here, we investigated the effects of GBE on atherosclerotic lesion development in rabbits with a high-fat diet.

Material and methods: Forty New Zealand white male rabbits were randomly divided into four groups. The first two were the normal diet group (C) and the high-fat group (HF). The remaining two groups were those who received a high cholesterol diet supplemented with either the standard drug (simvastatin 2 mg/kg/day) or GBE (3 mg/kg/day). At 12 weeks, histopathological and chemical analyses were performed.

Results: Plasma lipid measurement showed that GBE inhibited high-fat diet-induced increase of serum triglyceride (TG), total cholesterol (TC), and low density lipoprotein cholesterol (LDL-C) by 59.1% (0.9 ±0.24 mmol/l vs. 2.2 ±0.4 mmol/l), 18.2% (31.1 ±1.4 mmol/l vs. 38.0 ±0.4 mmol/l) and 15% (28.9 ±1.3 mmol/l vs. 34.0±1.0 mmol/l), respectively, at 12 weeks (p < 0.01). The en face Sudan IV-positive lesion area of the aorta in the GBE group (51.7 ±3.1%) was significantly lower compared with that in the HF group (88.2 ±2.2%; p < 0.01). The mean atherosclerotic lesion area of the GBE group was reduced by 53.2% compared with the HF group (p < 0.01). Immunohistochemistry and western blot analysis showed that GBE markedly suppressed high-fat diet-induced upregulation of connexin 43 (Cx43) in rabbits (p < 0.01).

Conclusions: Thus, our study revealed that GBE prevented atherosclerosis progress through modulating plasma lipid, suppressing atherosclerotic lesion development, and attenuating the expression of Cx43 protein.

Keywords: atherosclerosis, Ginkgo biloba extract, connexin 43, plasma lipid, high-fat diet.

Introduction

Atherogenesis is a multi-step process, which includes the accumulation of oxidized low density lipoprotein or low-density lipoprotein (LDL) within the blood vessel wall and an ensuing inflammatory cascade, leading to later stages of plaque development and thrombosis that are difficult to reverse [1]. Moreover, atherosclerosis is a chronic disease, and undetected in the early stage [1]. Thus, management of atherosclerosis, especially at the early stage, proves difficult. Even though treatments for clinical
manifestations of atherosclerosis are available, they tend to be plagued by several inherent drawbacks. Many pharmaceutical candidates and theoretically cardioprotective drugs fall short in a clinical setting, because of their off-target effects and low efficacy at tolerated doses [2-4]. Statins, one of the cholesterol lowering drugs, are proven to decrease the mortality of coronary heart disease. However, statins fail to rescue from acute ischemic events and prevent cellular uptake of oxidatively modified LDL [3-6]. Anti-cytokine antibodies and cytokine secretion inhibitors can diminish inflammatory responses which deteriorate atherosclerosis [7, 8], but trials have not shown efficacy in reducing clinical endpoints [9, 10]. Anti-chemokine therapy can reduce macrophage recruitment and block inflammation initiation. But these effects are limited by the interaction between chemokines and receptors [11]. Anti-platelet therapies prevent clot formation and aid clot breakup, but only address the latter stages of cardiovascular disease when clots are more likely to form [12, 13]. Current status of drug therapy for atherosclerosis demonstrates the need for the development of alternative strategies and platforms [14, 15].

*Ginkgo biloba* extract (GBE), from *Ginkgo biloba* leaves, has been widely used as a therapeutic agent for cardiovascular and neurological disorders [16]. Clinical studies showed that GBE can reduce atherosclerotic nanoplaque formation and size in cardiovascular high-risk patients [17, 18]. Experimental studies showed that GBE can inhibit nuclear factor κB (NF-κB) activation induced by hydrogen peroxide (H$_2$O$_2$) [19]. *Ginkgo biloba* extract can diminish cytokine-stimulated endothelial adhesiveness by downregulating intracelluar reactive oxygen species formation, NF-κB and activator protein 1 (AP-1) activation, and adhesion molecule expression in human aortic endothelial cells (HAECs) [20]. It also inhibits production of pro-inflammatory cytokines interleukin 1β (IL-1β) and tumor necrosis factor α (TNF-α), but up-regulates the production of anti-inflammatory cytokines [21]. All of these studies support the notion that GBE may have potential implications in clinical atherosclerosis disease. GBE-mediated inhibition of ox-LDL-induced lipoperoxide levels in endothelial cells and patients appears to be an important mechanism by which GBE exhibits the atherosclerosis inhibiting effect [17, 18, 22]. *Ginkgo biloba* extract is also documented to downregulate vascular endothelial growth factor (VEGF) expression, inhibit VEGF-mediated permeability, and suppress monocyte/macrophage-derived foam cell formation by a novel heme oxygenase-1 (HO-1)-dependent regulation of cholesterol homeostasis in macrophages [23-26]. In the present study, we analyzed the effects of GBE on atherosclerosis in rabbits with a high-fat diet.

**Material and methods**

**Animal model**

Forty New Zealand white male rabbits (4 months old, weighing 2.2-2.5 kg) were provided by the Laboratory Animal Centre of Shanghai Songlian (Shanghai, China) and individually caged in an air-conditioned room. There was an adaptation period of 7 days for animals before experiments. These rabbits were randomly divided into four groups: control group animals (C; n = 10) were fed a regular diet for 12 weeks; atherosclerosis group animals (HF; n = 10) were fed a high fat diet supplemented with 1% (w/w) cholesterol, 8% lard (w/w) and 0.05% cholate (w/w); simvastatin group animals (S; n = 10) were fed with the same high fat diet plus simvastatin (2 mg/kg/day); and GBE group animals (GBE; n = 10) were fed with the same high fat diet plus GBE (3 mg/kg/day). Each rabbit consumed about 120 g of food daily. Rabbits were caged individually with water ad libitum for 12 weeks, and maintained on a 12-h day/night cycle. Animal procedures were performed in accordance with guidelines set by the Animal Experiment Committee of Fudan University, Shanghai, China.

**Evaluation of atherosclerotic plaques**

To measure the atherosclerotic plaques, two complementary approaches were used: en face whole and histological section analyses [27]. The en face approach was employed to analyze the distribution and extent of atherosclerosis in the aorta, whereas microscopic analysis was used to evaluate lesion complexity as previously described [28]. In brief, the entire aorta was opened longitudinally and stained with Sudan IV-Herxheimer’s solution (Sigma). Lesion area was determined by calculating the proportion of aortic intimal surface area occupied by the red stain in the descending thoracic aorta, and abdominal aorta with the use of a video-based image analysis program (MCID, Imaging Research, St Catharines, Ontario, Canada).

Next, all aortic segments were embedded in paraffin, and 4-µm thick cross-sectional serial sections were obtained [29]. To quantify luminal cross-sectional area involved by atherosclerotic plaque, 10 sections obtained every 20 µm from the arteries were stained with hematoxylin and eosin. Lesion areas were quantitated by light microscopy (Olympus BX40). Lesion surface area and total aortic cross-sectional area were measured using Image Pro Plus version 6 (Media Cybernetics, Bethesda, MD).

**Plasma lipid analysis**

Plasma collected from 8 rabbits from each group was used for analysis of lipid compositions. Plasma total cholesterol (TC), triglyceride (TG), and low-den-
sity lipoprotein cholesterol (LDL-C) were analyzed by enzyme-linked immunosorbent assay (ELISA) [30]. The concentrations were subsequently determined using a spectrophotometer (Roche/Hitachi analyzer, Roche Diagnostics, Indianapolis, IN, USA).

**Immunohistochemistry**

Four sections from eight rabbits per group were analyzed by immunohistochemical staining. Paraffin-embedded artery sections were deparaffinized by immersion in xylene, followed by a series of alcohol treatments. Endogenous peroxidase activity was quenched by immersing the slides in 0.3% hydrogen peroxide in methanol for 15 min. The sections were rinsed in PBS three times for 5 min each and were blocked with 5% normal goat serum for 30 min. Primary antibodies were incubated for 90 min (goat anti-rabbit connexin 43 (Cx43), USCNLIFE, China, 1:200 dilution). Sections were washed and incubated with HRP-conjugated secondary antibodies for 30 min. Staining was detected using the Alkaline Phosphatase Standard ABC Kit (Vector Labs, Burlingame, CA). The slides were counterstained by immersion in xylene, followed by a series of alcohols.

**Western blot analysis**

Total protein was isolated from arteries, and equal amounts (20 µg) of protein samples were resolved by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and transferred onto a nitrocellulose membrane. The membrane was blocked for 1 h in 5% milk in TBS-T (Tris buffered saline and 0.1% Tween 20) and then incubated with primary antibody against Cx43 (1:200; USCNLIFE) or β-actin (1:200; USCNLIFE) as an internal control overnight at 4°C, followed by incubation with horse-radish peroxidase-conjugated secondary antibody (1:10,000; USCNLIFE). After washing 3 times with TBS, the membrane was developed using an enhanced chemiluminescent detection system (Amersham, Piscataway, NJ, USA). Signal intensities were quantitated using the Quantity One software (Bio-Rad, Hercules, CA).

**Statistical analysis**

All statistical analyses were carried out using the SPSS 11 software. If not otherwise stated, means of at least three independent experiments ± SD are shown. Statistical evaluations were done using analysis of variance (ANOVA) with the Tukey post hoc test. Values of p less than 0.05 were considered significant.

**Results**

**Effects of GBE on serum lipid composition in rabbit**

Before treatment, the baseline serum lipid levels (TG, TC, and LDL-C) were comparable among all experimental groups and there were no obvious changes of serum lipid levels (TG, TC, and LDL-C) in the control group after 6 and 12 weeks (data not shown). However, after the indicated treatment for 6 weeks, GBE lowered high-fat diet-induced increase of serum TC by 24.4% (32.3 ±1.6 vs. 42.7 ±2.6 mmol/l; p < 0.01; Table I). Moreover, GBE decreased high-fat diet-induced increase of serum TG, TC, and LDL-C by 59.1% (0.9 ±0.2 mmol/l vs. 2.2 ±0.4 mmol/l), 18.2% (31.1 ±1.4 mmol/l vs. 38.0 ±0.4 mmol/l) and 15% (28.9 ±1.3 mmol/l vs. 34.0 ±1.0 mmol/l), respectively, at 12 weeks (p < 0.01; Table I).

**Effects of GBE on development of aortic atherosclerotic lesions with high-fat diet in rabbits**

To examine the effect of GBE on atherosclerotic lesion development in rabbits, we used GBE to treat the rabbits (GBE group) with a high-fat diet. Simvastatin-treated rabbits (S group) were used as a positive control. The en face Sudan IV-positive lesion areas of the aorta in the rabbits with a high-fat diet (HF group, 88.2 ±2.4%) were significantly larger than those in the rabbits with a normal-fat diet (control group, C; 0) (p < 0.01; Figure 1), indicating that HF-induced atherosclerosis was successfully set up. The lesion areas in both S (40.8 ±2.4%) and GBE (51.7 ±3.1%) groups were significantly decreased relative to those in the HF group (88.2 ±2.2%; p < 0.01; Figure 1). Atherosclerotic

| Parameter | 6 weeks | 12 weeks |
|-----------|---------|----------|
|           | HF      | S        | GBE      | HF      | S        | GBE      |
| TG        | 0.8 ±0.2 | No change* | 0.9 ±0.1 | 2.2 ±0.4 | No change* | 0.9 ±0.2* |
| TC        | 42.7 ±2.6 | 22.3 ±1.4* | 32.3 ±1.6* | 38.0 ±0.4 | 20.2 ±2.0* | 31.1 ±1.4* |
| LDL-C     | 28.1 ±1.8 | 18.4 ±0.5* | 27.4 ±0.2 | 34.0 ±1.0 | 15.7 ±1.6* | 28.9 ±1.3* |

n = 10. *p < 0.01 compared to changes of HF group. No change: the change of TG level is less than 0.1 mmol/l. C – normal diet control group, HF – high-fat diet group, S – simvastatin, GBE – Ginkgo biloba extract, TC – total cholesterol, TG – triglyceride, LDL-C – low-density lipoprotein cholesterol.

Table I. Effect of Ginkgo biloba extract on serum lipid composition changes in rabbits (mmol/l)
lesion development in the aorta was also analyzed by H + E staining (Figure 2). Mean atherosclerotic lesion area in the GBE group was reduced by 53.2% compared with the HF group (25.7 ±3.8% vs. 54.9 ±4.0%; p < 0.01; Figure 2).

**Figure 1.** Effects of GBE on plaque formation in rabbits. Representative examples of en face dissection of thoracic and abdominal aorta, stained with Sudan IV, showed atherosclerotic lesions (red). Atherosclerotic involvement for each rabbit was quantitatively determined in en face sections by measuring the proportion of aortic intimal surface area occupied by Sudan IV positive lesions in total aorta

C – normal diet control group, HF – high-fat diet group, S – simvastatin treated group, GBE – Ginkgo biloba extract treated group; results were expressed as mean ± SD; *p < 0.01 relative to normal diet control (C), # p < 0.01 relative to high-fat diet group (HF).

**Figure 2.** GBE inhibits atherosclerotic lesion development in the aorta of rabbits with high-fat diet. Representative hematoxylin and eosin stained histological cross sections from aorta of indicated groups

Abbreviations – see Figure 1. Results were expressed as mean ± SD; *p < 0.01 relative to normal diet control (C), # p < 0.01 relative to high-fat diet group (HF). Magnification 100×

Ginkgo suppresses atherosclerosis through downregulating the expression of connexin 43 in rabbits

**GBE treatment reduced the expression of Cx43**

Gap junction protein Cx43 plays a key role in atherogenesis [31-34]. To explore the mechanism of GBE-induced suppression of atherosclerotic development, we analyzed the effect of GBE on the expression of Cx43 by immunohistochemistry in the rabbits with a high-fat diet. Figure 3 shows that Cx43 expression in the HF group was significantly augmented. GBE significantly diminished high-fat diet-induced Cx43 expression. Western blot analysis was also performed to measure the expression of Cx43 (Figure 4). In line with immunohistochemistry results, high-fat diet-induced Cx43 expression level was also downregulated in GBE groups.

**Discussion**

*Ginkgo biloba* extract EGb761 has antioxidant and antiplatelet aggregation effects indicating that it might protect against atherosclerosis [35]. Our results documented that GBE significantly decreased TC, LDL-C and TG levels, and attenuated atherosclerotic lesion development in the aorta, indicating potential anti-hyperlipidemic and anti-atherosclerotic effects of GBE. Although it seems that the anti-hyperlipidemic and anti-atherosclerotic properties of GBE are not as efficiency as simvastatin, GBE is extracted from *Ginkgo biloba* leaves with less side effects and has been widely used as a therapeutic agent for cardiovascular disorders [16]. Moreover, GBE has been suggested to be used in
arteriosclerosis prophylaxis in patients [18]. All of the studies indicate GBE as a promising anti-atherosclerotic drug.

It has been suggested that a distinctive spatial and temporal pattern of Cx43 expression between smooth muscle cells accompanies development of the atherosclerotic plaque [36]. Moreover, reduced Cx43 expression inhibits atherosclerotic lesion formation in low-density lipoprotein receptor-deficient mice [37]. Thus, Cx43 is a cardiovascular risk marker and therapeutic target [38]. Statins have been suggested as potent connexin modulators, synergizing with its lipid-lowering effect to reduce atherosclerosis mortality [39]. To investigate the mechanism of GBE-mediated suppression of atherosclerotic progress, we measured the expression of Cx43. We found that GBE treatment suppressed Cx43 expression, indicating another mechanism of GBE-mediated inhibition of atherosclerosis. In contrast to the suppressive effect of GBE on the expression of Cx43 in atherosclerosis, GBE could exert its neuroprotective effects by improvement of Cx43 expression and gap junction intercellular communication (GJIC) induced by hypoxia/ischemia-reoxygenation/reperfusion injury [40]. The different effects of GBE on the expression of Cx43 indicate that GBE regulates Cx43 through multiple pathways.

Although we found that GBE significantly inhibits high-fat diet-induced atherosclerosis via suppressing the expression of Cx43, there are still some open questions. One would be how GBE inhibits Cx43 expression in animal models of high-fat diet-induced atherosclerosis. Another one is whether the downregulation of Cx43 is sufficient for GBE-
mediated protective effects against high-fat diet-induced atherosclerosis. More attention will be focused on the molecular mechanisms of how GBE prevents the development of atherosclerotic plaques in the near future.

In conclusion, our study documents that GBE prevents atherosclerosis progress through modulating plasma lipid, suppressing atherosclerotic lesion development and Cx43 expression.

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Jian Ming Wei and Xin Wang contributed equally to this work.

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