Phenotypic characterization and susceptibility of gram negative bacteria from surgical site infections in a tertiary care hospital

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ABSTRACT

Background: Gram negative bacteria are the common isolates among the cases of Surgical Site Infections (SSI). Resistant and specially Multi Drug Resistant (MDR) Gram negative isolates are a serious challenge for the treatment to clinicians. Present study was undertaken for phenotypic characterization and susceptibility pattern of Gram negative bacterial isolates from cases of surgical site infections in a tertiary care institute.

Methods: This descriptive cross sectional hospital based study was conducted in a tertiary care teaching hospital over a period of one and half year from January 2012 to June 2013. Centers for disease control and prevention (CDC) SSI case definitions were used to label a case as SSI. Only culture proven cases, out of clinically suspected was included in the study for evaluation.

Results: During the study period a total of 5949 patients were operated and screened for SSI in the wards under surveillance. Out of which 556 were clinically suspected as a case of SSI. With 408 culture proven cases of SSI the rate of SSI in present study was (6.86%). Rates of SSI were more in dirty (22.54%) and contaminated (13.78%) type of wounds. Among Gram negative bacterial isolates (n=343) from SSI, E. coli (45.18%) was the commonest followed by Pseudomonas aeruginosa (16.03%) and Klebsiella pneumoniae (13.42%). Unpleasant trend in antimicrobial resistance observed during study is a serious concern.

Conclusions: The data presented in this study clearly indicate the continuous need of surveillance of SSI. This will clearly help health care personnel in curtailing down the incidences of SSI.

Keywords: Klebsiella pneumoniae, Multi drug resistant organism, Pseudomonas aeruginosa, Surgical site infections

INTRODUCTION

Hospitalized patients encountered by the most common complications during their hospital stay are Health care-associated infections (HAIs).¹ Despite of remarkable advances in infection control methods, HAIs remain a major public health problem and a significant cause of increased morbidity and mortality among hospitalized patients. In addition, HAIs are also responsible for inconvenience to the patients because of prolonged hospital stay that constitute an economic burden on patients and health care facility too.²⁻⁴ India has estimated prevalence of HAIs varies from 10-30% in comparison of HAIs prevalence worldwide varies from 3.8% to 18.6% depending upon the population surveyed.⁵ SSI contributes around a quarter of all health care associated infections. SSI have a remarkable impact on cost of therapy, which are associated with prolonged hospital stay, a high mortality and increased economic burden.⁶⁻⁷ Higher rates of infection often leads to increased rates of antimicrobial prescription which in turn contribute to increased antimicrobial resistance.⁶ Treatment of multi-drug resistant pathogens often requires costly second-and third-line antimicrobials which may be associated with increased adverse reactions, increased risk of complications, prolonged illness, morbidity and mortality, as well as economic load on the patient.⁶
Microorganisms both from endogenous and exogenous sources are responsible for SSI. Both gram positive and gram negative organisms are responsible for HAI. The Enterobacteriaceae family being the most commonly identified group overall among gram negative organisms responsible for SSI. Multidrug-resistant organisms are increasingly being reported.

The present study was designed to study the phenotypic characterization and susceptibility of gram negative bacteria from Surgical Site Infections (SSI) in this geographic area. The purpose of the Antimicrobial Susceptibility Test (ABST) was:

- To guide the choice of the antimicrobial for the treatment.
- To provide surveillance data to monitor the resistance trend.

METHODS

This descriptive cross sectional hospital based study was conducted in a tertiary care teaching hospital over a period of one and half year from January 2012 to June 2013. The study was carried out in surgical ICU, general surgery, and obstetrics/gynaecology wards. Patients admitted in hospital and fitting in the definition of HAI were included in the study. Patients after one month of discharge and post transplant patients were not included in the study. In this study a HAI was an infection which occur ≥48 h or more after admission to hospital and which was not present or incubating on admission and was suspected when the patient had signs and symptoms according to one of the CDC definitions, or had one or more signs or symptoms included in one of the CDC definitions. CDC’s SSI case definitions were used to label a case as SSI. Only culture proven cases, out of clinically suspected was included in the study for evaluation. Samples were collected from suspected SSI patients, 48 hours after admission and fulfilling the criteria for different types of SSI. Both active and passive surveillance methods were used for sample collection. Active surveillance was done by visiting various study areas daily along with infection control nurse. Passive surveillance was done by following the positive culture results obtained in microbiology laboratory to retrospective wards in the hospital and tried to look for HAI. Details of samples and patients were filled appropriately in predesigned Daily HAI Surveillance forms. The labelled specimens were transported to microbiology laboratory within 30 min of collection. Specimen were inoculated on appropriate culture media including blood agar, MacConkey agar, chocolate agar and incubated for 16-18 hrs at 35-37°C by using standard laboratory techniques.

Identification of bacteria was based on the colony characteristics of the organism i.e. colony morphology, hemolysis on blood agar, changes in the physical appearance of the differential media and enzyme activities of the organisms, Gram staining and biochemical tests.

Antimicrobial sensitivity was performed on Muller Hinton agar plates by Kirby-Bauer disk diffusion method as per CLSI guidelines. Antibiotic discs were procured from HiMedia laboratories, Mumbai, India. Isolates were labeled susceptible, resistant and intermediate on the basis of CLSI disc zone interpretative criterion. All Gram-negative isolates were tested for ESBL and MBL production as per CLSI guidelines. ESBL positive Klebsiella pneumonia ATCC 700603 and ESBL negative Escherichia coli ATCC 25922 were included in the study for quality control of ESBL tests. Pseudomonas aeruginosa 27853 was used as the control for MBL tests. All the media and reagents were procured from Himedia, Mumbai, India.

RESULTS

A total of 5949 patients were operated and screened for SSI in the wards under surveillance during study period. The age range of operated patients was 10-94 years (Mean: 44.63 yrs). Out of which 556 were clinically suspected as a case of SSI. With 408 culture proven cases of SSI the rate of SSI in present study was (6.86%) (Table 1). Females had more rates (4.34%) of SSI in comparison with males (2.52%) (Table 1). Rates of SSI were more in dirty (22.54%) and contaminated (13.78%) type of wounds (Table 2). Out of total 408 cases of culture proven cases of SSI, superficial surgical site infections (75.74%) were more in comparison with deep surgical site infections (24.26%) (Table 3). Rates of SSI were more in emergency surgeries (13.01%) in comparison with elective surgeries (5.16%) (Table 2). Among Gram negative bacterial isolates (n=343) from SSI, E. coli (45.18%) was the commonest followed by Pseudomonas aeruginosa (16.03%) and Klebsiella pneumoniae (13.42%) (Table 4).

Table 1: Overall and gender wise distribution of SSI.

| Clinically suspected SSI | Culture positive SSI | Rate of SSI (%) |
|--------------------------|----------------------|-----------------|
| 556                      | 408                  | 6.86            |
| M=216                    | M=150                | M=2.52          |
| F=340                    | F=258                | F=4.34          |

Table 2: Rates of SSI according to type of wounds and timing of surgeries.

| Type of wound         | Total | Culture proven SSI | SSI rate (%) |
|-----------------------|-------|--------------------|--------------|
| Clean                 | 2929  | 89                 | 3.04         |
| Clean contaminated    | 2593  | 245                | 9.45         |
| Contaminated          | 254   | 35                 | 13.78        |
| Dirty                 | 173   | 39                 | 22.54        |
| Elective              | 4666  | 241                | 5.16         |
| Emergency             | 1283  | 167                | 24.26        |

E. coli was the most common isolates among Gram negative bacteria isolated from the cases of SSIs, around Overall (96.77%) of E. coli were resistant to ampicillin.
followed by (91.61%) to cefuroxime and (90.96%) to amoxycillin+ clavulanic acid. While none of the E. coli isolates were resistant to imipenem. Among aminoglycosides group resistance rates to gentamicin (42.58%) were more in comparison with amikacin (13.54). Resistance rates to ciprofloxacin (76.77%) were high.

Table 3: Distribution of different categories of SSI (n=408).

| Type of SSI                  | Culture positive SSI (n=408) | (%) |
|-----------------------------|-----------------------------|-----|
| Superficial surgical site infections | 309                          | 75.73 |
| Deep surgical site infections | 99                           | 24.26 |
| Organ/ space                | -                            | -   |

Table 4: Gram negative bacterial isolates from SSI (n=343).

| Microorganisms              | Number (n=343) | (%) |
|-----------------------------|----------------|-----|
| E. coli                     | 155            | 45.18 |
| Pseudomonas aeruginosa      | 55             | 16.03 |
| Klebsiella pneumoniae       | 46             | 13.42 |
| Citrobacter freundii        | 20             | 5.84  |
| Citrobacter koseri          | 18             | 5.25  |
| Enterobacter aerogenes      | 12             | 3.49  |
| Klebsiella oxytoca          | 10             | 2.92  |
| Pseudomonas stutzeri        | 9              | 2.62  |
| Proteus mirabilis           | 8              | 2.34  |
| Proteus vulgaris            | 3              | 0.87  |
| Pseudomonas putida          | 3              | 0.87  |
| Acinetobacter baumannii     | 2              | 0.58  |
| Morganella morganii         | 2              | 0.58  |

Table 5: Resistance pattern of common Gram negative bacterial isolates from SSI (% resistance).

| Antibiotics                  | E. coli (n=155) | Pseudomonas aeruginosa (n=55) | Klebsiella pneumoniae (n=46) | Citrobacter freundii (n=20) | Citrobacter koseri (n=18) | Enterobacter aerogenes (n=12) | Proteus spp. (n=11) |
|------------------------------|----------------|------------------------------|-----------------------------|-----------------------------|---------------------------|-----------------------------|---------------------|
| Ampicillin                   | 150 (96.77)    | ND                           | 45 (97.82)                  | 18 (90)                     | 16 (88.88)                | 12 (100)                    | 11 (100)            |
| Amoxycillin + Clavulanic acid| 141 (90.96)    | ND                           | 42 (91.30)                  | 18 (90)                     | 16 (88.88)                | 12 (100)                    | 10 (90.90)          |
| Piperacillin                 | 104 (67.09)    | 10 (18.18)                   | 37 (80.43)                  | 15 (75)                     | 15 (83.33)                | 11 (91.66)                  | 05 (45.45)          |
| Piperacillin + Tazobactum    | 36 (23.22)     | 06 (10.90)                   | 19 (41.30)                  | 12 (60)                     | 07 (38.88)                | 05 (41.66)                  | 03 (27.27)          |
| Cefazolin                    | 128 (82.58)    | ND                           | 41 (89.13)                  | 17 (85)                     | 14 (77.77)                | 09 (75)                     | 10 (90.90)          |
| Cefepime                     | 94 (60.64)     | 19 (34.54)                   | 36 (78.26)                  | 14 (70)                     | 12 (66.66)                | 09 (75)                     | 08 (72.72)          |
| Cefotaxime                   | 118 (76.12)    | ND                           | 41 (89.13)                  | 17 (85)                     | 15 (83.33)                | 12 (100%)                   | 09 (81.81)          |
| Cefoxitin                    | 54 (34.83)     | 16 (29.09)                   | 27 (58.69)                  | 16 (80)                     | 13 (72.22)                | 11 (91.66)                  | 06 (54.54)          |
| Ceftazidime                  | 105 (67.74)    | 11 (20)                      | 39 (84.78)                  | 16 (80)                     | 14 (77.77)                | 11 (91.66)                  | 09 (81.81)          |
| Cefoparazone                 | ND             | 11 (20)                      | ND                          | ND                          | ND                        | ND                          | ND                  |
| Cefuroxime                   | 155 (91.61)    | ND                           | 43 (93.47)                  | 18 (90)                     | 15 (83.33)                | 12 (100)                    | 11 (100)            |
| Imipenem                     | 00 (00)        | 00 (00)                      | 01 (2.17)                   | 00 (00%)                    | 00 (00)                   | 00 (00)                     | 01 (9.09)           |
| Amikacin                     | 21 (13.54)     | 15 (27.27)                   | 20 (43.47)                  | 09 (45)                     | 07 (38.88)                | 05 (41.66)                  | 04 (36.36)          |
| Gentamicin                   | 66 (42.58)     | 19 (34.54)                   | 31 (67.39)                  | 11 (55)                     | 13 (72.22)                | 09 (75)                     | 08 (72.72)          |
| Ciprofloxacin                | 119 (76.77)    | 24 (43.63)                   | 40 (86.95)                  | 15 (75)                     | 13 (72.22)                | 11 (91.66)                  | 07 (63.63)          |
| Trimethoprim/ Sulfamethoxazole| 41 (26.45)     | ND                           | 26 (56.52)                  | 14 (70)                     | 10 (55.55)                | 07 (58.33)                  | 07 (63.63)          |
*Pseudomonas aeruginosa* was the second most common isolates and it shows around (43.63%) resistance to Ciprofloxacin followed by (34.54%) to cefepime and (20%) to cefoparazone and cefazidime.

While none of the *Pseudomonas aeruginosa* isolates were resistant to imipenem. Among aminoglycosides group resistance rates to gentamicin (34.54%) were more in comparison with amikacin (27.27%) (Table 5). Overall (40%) isolates of *E. coli* and (54.34%) isolates of *Klebsiella pneumoniae* were ESBL producers, (2.17%) isolates of *Klebsiella pneumoniae* were MBL producers and (74.84%) of *E. coli*, (98.21%) of *Klebsiella pneumoniae* and (21.81%) of *Pseudomonas aeruginosa* were considered as MDR (Figure 1).

![Figure 1: ESBL, MBL and MDR among Gram negative bacterial isolates of SSI.](image)

**DISCUSSION**

The surgical site infection rates reported by different workers have considerable difference. The overall infection rate in the present study was (6.86%) and compares favorably with other reported rates ranging from (2.5%) to (41.9%). while the rate is much higher than the overall average SSI rate of (2.61%), published by the NNISS, USA.  

The higher infection rate may be because of poor awareness about the HAI and ineffective infection control practices. Number of studies carried out in India indicates an overall infection rate of 4.04% to 30% for clean surgeries and 10.06% to 45% for clean contaminated surgeries. Findings in the present study showed that there was a significant rise in infection with increased degree of operative contamination; rate of infection from clean wounds was (3.04%) while in dirty wounds it was (22.54%). Similar findings were reported by with Li et al.  

Emergency operations were more likely, than routine to be infected. In present study the rate of SSI (13.01%) in emergency surgeries were higher than the rate of SSI (5.16%) in elective surgeries. Other studies have also demonstrated higher infection rate in the emergency operations, this might be because of inappropriate pre-operative preparation and the severity of underlying condition that required emergency procedure. Present study shows that *E. coli* was the leading cause of SSI followed by *Pseudomonas aeruginosa* and *Klebsiella pneumonia* (Table 4). These results are in accordance with few studies while differ from some other studies.

A total of (96.77%) of *E. coli* isolates were resistant to ampicillin the present study. While Sikka et al, reported (98.6%) resistance to ampicillin. Thus we can say that the ampicillin which was previously sensitive to many Gram-negative bacteria is now resistant to almost all. Difference between the percent resistance rate of amikacin and gentamicin (13.54% vs 42.58%) is in accordance with the result of sikka et al (58.3% vs 95.8%). Resistance pattern of *E. coli* in this study as compared to Sikka et al, were for ciprofloxacin (76.77 vs 93.1%), imipenem (0% vs 4.2%), amoxicillin/ clavulanic acid (90.96% vs 84.7%), cefazidime (67.74% vs 91.6%), cefotaxime (76.12% vs 94.4%) and cefepime (60.64% vs 91.6%). So, increased antibiotic resistance pattern noted for *E. coli* suggests its importance for hospital acquired infection. Resistance pattern of *K. pneumoniae* in this study as compared to Sikka et al, were for ampicillin (97.82% vs 100%), ciprofloxacin (86.95% vs 88.2%), imipenem (2.17% vs 0%), amoxicillin/ clavulanic acid (91.30% vs 88.2%), cefazidime (84.78% vs 100%), cefotaxime (89.13% vs 94.1%), cefepime (78.26% vs 82.3%), amikacin (43.47% vs 76.4%) and gentamicin (67.39% vs 94.1%). Resistance pattern of *Pseudomonas aeruginosa* in this study as compared to Sikka et al, were for piperacillin/tazobactum (10.90% vs 50%), imipenem (00% vs 0%), cefazidime (20% vs 87.5%), cefepime (34.54% vs 87.5%), amikacin (27.27% vs 75%) and gentamicin (34.54% vs 100%). Resistance to *Pseudomonas aeruginosa* isolates from SSI in present study was lower in comparison to Sikka et al. Susceptibility pattern of common isolates in present study varied from the results of other observers.

Among the species *E. coli* and *K. pneumonia* the development of resistance to extended-spectrum cephalosporins is becoming a serious trend worldwide. Such resistance is most often due to the production of extended-spectrum β-lactamases (ESBLs). The data from the Tigecycline Evaluation and Surveillance Trial (TEST) global surveillance database shows that the rate of the ESBL production was the highest among the *K. pneumoniae* isolates.

Previous studies which have documented the prevalence of ESBL in India, as 24.8-63.8% among the *E.coli* isolates, at 10.1-76.2% among the *Klebsiella pneumoniae* and at 14.4-70.5% among *Proteus mirabilis*. In the present study the rate of ESBL producers among *E. coli* isolated from cases of SSI were (40%) and *K. pneumoniae* (54.34%). Analysis of data from National Nosocomial Infection Surveillance (NNIS) system demonstrate that only 2.3%
Enterobacteriaceae isolates tested nonsusceptible to imipenem. However, rate of Carbapenem Resistant Enterobacteriaceae (CRE) is increasing worldwide. In the Meropenem Yearly Susceptibility Test Information Collection Program, meropenem resistance among clinical isolates of *Klebsiella pneumoniae* increased significantly from 0.6% in 2004 to 5.6% in 2008. Among isolates reported NHSN in 2009-2010, carbapenem resistance was up to (1%) to *K. pneumoniae*/*oxytoca* and (1%) to *E. coli*. In the present study the rate of MBL producers among *K. pneumoniae* isolates were (2.17%). Results are comparable with the NHSN 2009-2010 data. Indian studies have documented the prevalence rate ranged from 1-11% among Enterobacteriaceae.

No isolates of *E. coli* and *Pseudomonas aeruginosa* were found resistant to Carbapenem. Though the rate is low in comparison to other studies but still the emergence of resistance for carbapenem group of antibiotics in present study warranted the judicious use which will help in, to prevent further development of resistance.

Isolates resistant to three or more antimicrobial classes were considered Multi Drug Resistant (MDR) in the present study. Results of current study documented that (74.84%) of *E. coli* isolates and (98.21%) *K. pneumoniae* isolates were MDR. MDR in *P. aeruginosa* is usually defined as resistance to three or more of the following antimicrobial agents: antipseudomonal penicillins (e.g., piperacillin), antipseudomonal cephalosporins (e.g., ceftazidime), fluoroquinolones (e.g., ciprofloxacin), carbapenems (imipenem, meropenem, and doripenem), and the aminoglycosides (gentamicin, tobramycin, or amikacin). The prevalence of MDR *Pseudomonas aeruginosa* in various parts of the world ranged from (1.6-50%). In the present study the prevalence rate of MDR *Pseudomonas aeruginosa* were (21.81%). These results are comparable with the data from the different parts of the world.

So, this study finding supports the unpleasant trends in antibiotic resistance, indicating decreasing efficacy of various antibiotic classes against gram negative bacilli. The increased prevalence of extended-spectrum beta-lactamases may contribute to the finding of multidrug resistance. Hospital set up, injudicious use of antimicrobials and case materials determines the rate of SSI in any hospital. Antibiogram might be varying depending on the study group and hospital set up.

Overall, the pattern of antimicrobial resistance among Gram negative isolates from SSI in the present study emphasized the need for:

- Provide a healthcare set up that is safe for patients.

**CONCLUSION**

The data presented in this study clearly indicate the gravity of antibiotic resistance. Surveillance of SSI and antibiotic resistance in SSI causing pathogens is a need of future. Hence the data generated will help to treating physician and/or surgeon in getting the knowledge of current trends of SSI in this setup that ultimately help them in the selection of appropriate antimicrobial therapy, thereby decreases length of hospital stay, cost of therapy, morbidity and mortality. Appropriate and adequate antimicrobial treatment of SSI cases based on microbiological culture report is need of time.

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