Factors associated with the clinical outcome of patients with relapsed/refractory CD19+ acute lymphoblastic leukemia treated with ARI-0001 CART19-cell therapy

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The prognosis of patients with relapsed/refractory (R/R) acute lymphoblastic leukemia (ALL) remains poor, particularly for those relapsing after allogeneic hematopoietic cell transplantation (alloHCT).1 Novel agents such as inotuzumab ozogamicin or blinatumomab achieve increased response rates, but these are generally transient unless followed by alloHCT. Chimeric antigen receptors (CAR) targeting CD19 have shown promising results in R/R ALL, and one of these products (tisagenlecleucel) has been approved for the treatment of patients with R/R ALL up to 25 years of age.2

In 2013, we designed our own CAR19 construct, comprising a single chain variable fraction sequence from the A381 hybridoma and the 4-1BB/CD3z signaling domains (online supplemental figure 1). After scaling up both lentiviral and cell production,3,4 the Spanish Medicines Agency (AEMPS) approved our product (ARI-0001 cells) and the CART19-BE-01 trial.5 In October 2019, once the recruitment was completed, the AEMPS approved a compassionate use program (CUP) for patients fulfilling the same inclusion/exclusion criteria. The trial’s preliminary results were published elsewhere.3 Here, we present the long-term results of all consecutive patients with R/R ALL recruited into the CART19-BE-01 trial and the subsequent CUP. Detailed information on ARI-0001 cell manufacturing, inclusion/exclusion criteria, primary and secondary endpoints and assessment criteria can be found elsewhere.5

Before ARI-0001 cell infusion, patients received fludarabine at 30 mg/m²/day plus cyclophosphamide at 300 mg/m²/day on days −6, −5, and −4 followed by ARI-0001 cells. The first 15 patients received a single intravenous infusion of 0.5–1×10⁶ ARI-0001 cells/kg (adults) or 5×10⁶ ARI-0001 cells/kg (children) on day 0. The following 38 patients (23 from the CART19-BE-01 trial and 15 from the CUP) received 1×10⁶ ARI-0001 cells/kg regardless of age: the first fraction (10%) on day 0, followed by the second (30%) and third (60%) fraction. The second fraction was administered 24–48 hours after the first, and the third 24–48 hours after the second, only if the patient had no signs or symptoms of CRS (online supplemental figure 2). The reason for this amendment to the protocol were three cases of fatal toxicity (two patients, aged 11 and 19, who died of refractory CRS, and one patient, aged 35, who died of pseudomembranous colitis as a complication of grade 4 CRS).5 The entire patients’ disposition is depicted in online supplemental figure 3.

Adverse events and response rates are presented with 95% exact Clopper-Pearson CIs. The possible association between CRS
(all grades and grade ≥3), tumor burden at screening (<5% vs ≥5% blasts in the bone marrow (BM)) and type of administration (single dose vs fractionated) was assessed using Fisher’s exact test. We also analyzed the impact on progression-free survival (PFS) and overall survival (OS) of the following variables: age (<25 vs ≥25 years), type of administration, tumor burden and loss of B-cell aplasia (BCA), the latter as a time-dependent covariate. PFS/OS curves were plotted using the Kaplan-Meier method for time-fixed covariates and the Simon-Makuch method for BCA loss. Landmark analyses were performed to identify the most appropriate timepoint for BCA loss. Univariate Cox regression was used to evaluate the impact of these covariates on PFS/OS, and those with Benjamini-Hochberg adjusted p values lower than 0.1 were introduced into a multivariate Cox regression. Schönfeld residuals were used to check the proportional hazards assumption. The trial was registered at clinicaltrials.gov (NCT03144583).

Fifty-three patients with R/R ALL received therapy with ARI-0001 cells: 38 in the context of the CART19-BE-01 trial and 15 as part of the CUP. The median age was 30 years (range, 3–68), while 24 (45%) patients were female. Baseline characteristics of the entire population are displayed in online supplemental table 1). The data cut-off date was March, 2021, when all infused patients had a minimum follow-up of 100 days or had experienced disease relapse or death before that date.

Patients received ARI-0001 cells a median of 55.5 days (range, 27–216) after inclusion, and the median vein-to-vein time (from apheresis to infusion) was 43 days (range, 21–190). The original target dose was infused to all except 3 (5.7%) patients who received 0.1–0.4×10^6 ARI-0001 cells/kg due to CRS. CRS was reported in 56.6% (95% CI 42.3%–70.2%) of patients, being grade ≥3 in 11.3% (95% CI 4.3% to 23%) and requiring treatment with tocilizumab and steroids in 20.7% and 11.3% of patients, respectively. Patients with ≥5% lymphoblasts in the BM had a higher incidence of CRS (any grade: 82% vs 39%, p=0.0022; grade ≥3: 27% vs 0%, p=0.0036) compared with those with <5% lymphoblasts. Moreover, the incidence and severity of CRS was also associated with single dose versus fractionated administration of ARI-0001 cells (any grade: 87% vs 45%, p=0.0064; grade ≥3: 27% vs 5%, p=0.047). Neurotoxicity was observed in 13.2% (95% CI 5.5% to 25.3%) of patients, with one self-limited grade ≥5 occurrence (1.9%). No new second malignancies have been reported apart from a previously notified case of myelodysplasia (1/53; 1.9%).

The safety profile of ARI-0001 cells was comparable to that of similar products, with grade ≥3 CRS/neurotoxicity rates lower than 5%. Moreover, both fractionated administration and tumor burden were significantly associated with the incidence of CRS, in keeping with similar studies. Of note, two patients enrolled in the CUP experienced grade ≥3 CRS with the first fraction (0.1×10^6 cells/kg), but successfully recovered after treatment with tocilizumab. Both patients had a high tumor burden (>90% blasts in the BM) at study inclusion, and yet they could receive therapy while avoiding irreversible toxicity.

The measurable residual disease (MRD)-negative CR rate was 88.6% (95% CI 77.0% to 95.7%) at day +28 and 79.2% (95% CI 65.9% to 89.2%) at day +100. All three patients who received less than 1×10^6 ARI-0001 cells/kg due to toxicity achieved an MRD-negative CR. All evaluable patients (n=50) developed absolute BCA that lasted for a median of 4.2 months (95% CI 3.32 to 7.53 months). PFS was 50.9% (95% CI 38.4% to 67.4%) and 32.9% (95% CI 20.6% to 52.6%) at one and 2 years, respectively, while the 1-year and 2-year OS were 70.2% (95% CI 58.1% to 84.8%) and 53.9% (95% CI 40.5% to 71.8%). Progressive disease has occurred in 27 (50.9%; 95% CI 36.8 to 64.9%) patients at a median of 5.3 (range, 0.2–23.1) months. Tumor cells expressed CD19 in 24 (89%) of these relapses, while three (11%) were CD19-negative. ARI-0001 cells served as a bridge to alloHCT in three (6%) patients. Second ARI-0001 infusions were documented in nine patients (three more than previously reported): four due to CD19 +relapse and five in patients with early BCA loss. These resulted in transient responses and brief periods of BCA, but one of these responses allowed the patient to receive a second alloHCT.

Subgroup analyses are depicted in figure 1 and online supplemental table 2. By univariate analysis, only two variables had a potential impact on PFS: tumor burden (<5% vs ≥5% lymphoblasts in the BM at screening), with a 2-year PFS of 52.5% (95% CI 36.4% to 75.7%) vs 10.7% (95% CI 2.1% to 54.4%) and an HR of 2.14 (95% CI 1.04 to 4.42) for patients with 5% or more blasts (adjusted p=0.077). On the other hand, loss of BCA had an HR of 4.41 (95% CI 1.59 to 12.2), adjusted p=0.0172. Both variables (tumor burden and loss of BCA) were also confirmed in the multivariate model, with an HR of 2.05 (95% CI 1.004 to 4.17) for patients with 5% or more blasts at screening (p=0.0484) and an HR of 4.32 (95% CI 1.57 to 11.86) for patients with loss of BCA (p=0.0045). Regarding OS, none of the covariates evaluated had sufficient impact to justify a multivariate analysis. Seeing that BCA loss had such an impact on PFS, we performed a series of landmark analyses to identify the most appropriate cut-off for clinical practice. We chose 3 and 6 months as potential landmark times because the median time to BCA loss was 4.2 months in our series. According to these analyses, the 3-month time point was the closest to statistical significance (HR 1.83; 95% CI 0.82 to 4.11; p=0.15, (online supplemental table 3).

With more patients and longer follow-up, we identified two covariates of potential predictive value: tumor burden in the BM and loss of BCA. This contrasts with our previous report, where this potential effect was not fully evident. The adverse impact of tumor burden has been documented for other therapies, including alloHCT and CART19-cells and is therefore not surprising. The importance of BCA loss is, on the other hand, more controversial. Our strategy was associated with a relatively short in vivo ARI-0001 cell survival (median 4.2 months)
and most relapses had a CD19-positive phenotype, all in keeping with other studies. Consequently, further immune manipulation may be justified in patients with a short-lived BCA, perhaps less than 3 months, although this potential timepoint needs further validation.

In conclusion, both tumor burden and BCA loss appeared to have a significant impact on PFS and could guide clinicians in the management of patients after cell infusion. The validity of the fractionated cell administration was also confirmed and will be explored in a phase 2 trial. The results of the CART19-BE-01 trial led to the approval by the AEEMPS of ARI-0001 cells for the treatment of patients with R/R ALL older than 25 years of age (Hospital Exemption). To the best of our knowledge, this is the first purely academic CART19-cell product approved in any European country for any indication. This also makes Spain the only country in Europe where patients with R/R ALL have at least one approved CART19-cell product regardless of age (tisagenlecleucel for younger patients; ARI-0001 cells for older patients).

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