Water Triggered Synthesis of Highly Stable and Biocompatible 1D Nanowire, 2D Nanoplatelet, and 3D Nanocube CsPbBr₃ Perovskites for Multicolor Two-Photon Cell Imaging

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ABSTRACT: Two-photon imaging in the near-infrared window holds huge promise for real life biological imaging due to the increased penetration depth. All-inorganic CsPbX₃ nanocrystals with bright luminescence and broad spectral tunability are excellent smart probes for two-photon bioimaging. But, the poor stability in water is a well-documented issue for limiting their practical use. Herein, we present the development of specific antibody attached water-resistant one-dimensional (1D) CsPbBr₃ nanowires, two-dimensional (2D) CsPbBr₃ nanoplatelets, and three-dimensional (3D) CsPbBr₃ nanocubes which can be used for selective and simultaneous two-photon imaging of heterogeneous breast cancer cells in the near IR biological window. The current manuscript reports the design of excellent photoluminescence quantum yield (PLQY), biocompatible and photostable 1D CsPbBr₃ nanowires, 2D CsPbBr₃ nanoplatelets, and 3D CsPbBr₃ nanocubes through an interfacial conversion from zero-dimensional (0D) Cs₄PbBr₆ nanocrystals via a water triggered strategy. Reported data show that just by varying the amount of water, one can control the dimension of CsPbBr₃ perovskite crystals. Time-dependent transition electron microscopy and emission spectra have been reported to find the possible pathway for the formation of 1D, 2D, and 3D CsPbBr₃ nanocrystals from 0D Cs₄PbBr₆ nanocrystals. Biocompatible 1D, 2D, and 3D CsPbBr₃ nanocrystals were developed by coating with amine−poly(ethylene glycol)−propionic acid. Experimental data show the water-driven design of 1D, 2D, and 3D CsPbBr₃ nanocrystals exhibit strong single-photon PLQY of ~66−88% as well as excellent two-photon absorption properties ($\sigma_2$) of $\sim 8.3 \times 10^5−7.1 \times 10^4$ GM. Furthermore, reported data show more than 86% of PL intensity remains for 1D, 2D, and 3D CsPbBr₃ nanocrystals after 35 days under water, and they exhibit excellent photostability of keeping 99% PL intensity after 3 h under UV light. The current report demonstrates for the first time that antibody attached 1D and 2D perovskites have capability for simultaneous two-photon imaging of triple negative breast cancer cells and human epidermal growth factor receptor 2 positive breast cancer cells. CsPbBr₃ nanocrystals exhibit very high two-photon absorption cross-section and good photostability in water, which are superior to those of commonly used organic probes ($\sigma_2 = 11$ GM for fluorescein), and therefore, they have capability to be a better probe for bioimaging applications.

KEYWORDS: water triggered synthesis, 1D, 2D, and 3D CsPbBr₃ nanocrystals, biocompatible CsPbBr₃ perovskites, two photon absorption properties, two photon luminescence imaging of cancer cells, triple negative breast cancer cell identification

1. INTRODUCTION

All-inorganic cesium lead halide (CsPbX₃, X = Cl, Br, and I) perovskites exhibit excellent photoluminescence quantum yield (PLQY), IP luminescence emission, and large two-photon (2P) absorption cross sections, which allow them to be a promising candidate for next-generation bioimaging materials.1−10 However, the major obstacle for future bioimaging material applications for perovskites is the poor stability in water due to their ionic nature.11−43 As we and others have reported, CsPbX₃ perovskites decompose fast when exposed to water or kept in a moist environment.28−40 As a result, enhancing the long-time stability of CsPbX₃ perovskites is the most important parameter to promote commercial applications.30−44

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As shown in Figure 1, for the design of water-resistant lead halide perovskite-based two-photon bioimaging is rare. As a result, it is highly desirable to develop a method which can produce water resistant, biocompatible, and photostable 1D, 2D, and 3D perovskite, and these nanocrystals will have capability for two-photon bioimaging applications. As we and others reported, two-photon absorption (2PA) is a nonlinear process where near-infrared light (NIR) in a biological window can be used, which features deep penetration depths, higher spatial resolution, and smaller sample damage for bioimaging and photodynamic therapy.\(^{55-64}\) Driven by the need, herein, we report the development of high water resistance, excellent photoluminescence quantum yield (PLQY), biocompatible and photostable lead halide perovskite, a PEG ligand has been used to anchor a biospecific antibody so as to decorate them with moisture-tolerant molecules.\(^{34}\) Despite these great efforts, reports on 1D, 2D, and 3D lead halide perovskite-based two-photon bioimaging is rare. As a result, it is highly desirable to develop a method which can produce water resistant, biocompatible, and photostable 1D, 2D, and 3D perovskite, and these nanocrystals will have capability for two-photon bioimaging applications. Applications for water-resistant lead halide perovskites include selective and simultaneous two-photon imaging of TNBC cells and HER-2 positive breast cancer cells. In the two-photon imaging process, CsPbBr\(_3\) perovskites simultaneously absorb two monochromatic infrared photons in the biological window and emit a shorter wavelength photon, which offers significant advantages for bioimaging.\(^{55-68}\)

### 2. Results and Discussion

#### 2.1. Water Triggered Synthesis of 3D CsPbBr\(_3\) Nanocubes from 0D Cs\(_4\)PbBr\(_6\) Nanocrystals Using 0.2 mL of Water

Initially, we developed 0D Cs\(_4\)PbBr\(_6\) NCs using the hot injection method as we and others have reported before\(^{25-40}\) (see more details in the experimental section in the Supporting Information). The freshly prepared 0D Cs\(_4\)PbBr\(_6\) was then separated by centrifugation for 15 min at 6000 rpm, followed by resuspension in hexane at 4 °C for further use. We determined the element molar ratios using inductively coupled mass spectrometry (ICP-MS) data. As reported in Figure 1, the transmission electron microscopy (TEM) image from freshly prepared 0D Cs\(_4\)PbBr\(_6\) perovskite is highly monodisperse with size of 30 ± 3 nm and has no luminescence before immersing in water. The HRTEM image, as inserted in Figure 1, shows clear lattice fringes with interplanar spacing (\(d\)) of ~0.56 nm corresponding to the (110) crystal plane of Cs\(_4\)PbBr\(_6\). The X-ray powder diffraction (XRD) data from 0D Cs\(_4\)PbBr\(_6\), as reported in Figure 2A, show that it retains the rhombohedral phase, where main peaks are assigned to be (120), (113), (300), (020), (301), (223), (214), (314), (324), and (311) planes.\(^{26-40}\) As shown in Figure 1 and Table 1, in the next step, nanowires, nanosheets, and nanocubes were obtained through an interfacial conversion from 0D Cs\(_4\)PbBr\(_6\) nanocrystals by water induction.

For the development of 3D CsPbBr\(_3\) nanocubes, we used 1 mM of Cs\(_4\)PbBr\(_6\) solution in hexane. We also added 0.2 mL of water and kept it undisturbed for several hours at room temperature. This process where near-infrared light (NIR) in a biological window can be used, which features deep penetration depths, higher spatial resolution, and smaller sample damage for bioimaging and photodynamic therapy.

![Figure 1. TEM image, inserted HRTEM image, inserted SEM image, and photograph of water resistant 1D, 2D, and 3D CsPbBr\(_3\) perovskite nanocrystals derived through an interfacial conversion from 0D Cs\(_4\)PbBr\(_6\) nanocrystals using different amounts of water.](https://dx.doi.org/10.1021/jacsau.0c00038)
temperature. Upon contact with water, colorless Cs₄PbBr₆ solution rapidly became greenish, which indicates the formation of 3D CsPbBr₃ nanocubes. To understand the evolution of the process from zero dimensional Cs₄PbBr₆ nanocrystals to 3D CsPbBr₃ nanocubes, we monitored the time-dependent emission spectra during the synthesis process. As shown in Figure 2C, a green emission peak appears with luminescence maximum at 528 nm even within 25 min and continually increases as the reaction proceeds.

During this process, stripping of CsBr occurs because of the ionic nature of Cs₄PbBr₆ and the very high solubility of CsBr in water. The above process helps the decomposition of Cs₄PbBr₆ and the formation of the simple cubic structure of 3D CsPbBr₃. When the CsPbBr₃ nanocubes are formed, they diffused to the top, and as a result, we observed the greenish color at the water surface. It is now well-documented that the capping oleic acid (OA) and olamine (OAm) ligands are mostly coordinating at Pb sites.

In the presence of minor amounts of water, H₂O molecules can partly ionize into H₃O⁺ and OH⁻ with the help of OA and OAm. In the next step, OH⁻ can partially replace OA or OAm on the surface. In this condition, the structure became loose, which allowed it to form decoupled [PbBr₆]₄⁻ octahedrons. These octahedral monomers help to form bigger hexagonal particles, which allows them to develop 3D CsPbBr₃ nanocubes, and they are protected by the hydroxy (OH) group.

After the full transformation, CsPbBr₃ nanocrystals were isolated by centrifuging at 8000 rpm for 5 min to obtain high-quality nanocubes. TEM image from freshly prepared 3D CsPbBr₃, as reported in Figure 1, and dynamic light scattering (DLS) data, as reported in Table 2, show that the perovskites...
water for 3D CsPbBr₃ developed using the water-triggered protection by the surface photoluminescent quantum yield for 3D CsPbBr₃ was (200), (211), and (220) planes of the cubic lattice. 1 The trigger synthesis. We also observed peaks for the C−OH groups.

TEM image from freshly prepared PEG coated 3D CsPbBr₃ collected through sonicated for a few minutes, and PEG coated nanocrystals were measured to be ∼68%. Figure 3E shows that the stability in filtration. As reported in Figure S1A, B, the polymers. FTIR spectra as reported in Figure 2B shows vibrational peaks from PEG coated 3D CsPbBr₃, which is mainly due to the coating of PEG. The XRD data, reported in Figure 2A, 3D CsPbBr₃ is retained the cubic phase, where main peaks are assigned to be the (100), (110), (111), (200), (211), and (220) planes of the cubic lattice. ¹ The photoluminescent quantum yield for 3D CsPbBr₃ was measured to be ∼68%. Figure 3E shows that the stability in water for 3D CsPbBr₃ developed using the water-triggered process is much better than that of 3D CsPbBr₃ developed using the hot injection method, which can be attributed to the protection by the surface −OH groups.

To understand how the water amount variation can change the morphology of the product, we performed the same experiment by varying the water amount from 0.05 to 0.2 mL and keeping the Cs₄PbBr₆ concentration the same. As reported in Table 1, when we used 0.05 mL of water, we obtained 60% 0D Cs₄PbBr₆ and 40% 3D CsPbBr₃. As we increased the amount of water from 0.05 to 0.1 mL, we obtained 20% 0D Cs₄PbBr₆ and 80% 3D CsPbBr₃. At the end, when we used 0.2 mL water, we obtained 100% 3D CsPbBr₃.

In the next step for the development of biocompatible 3D CsPbBr₃ nanocrystals, we redispersed the nanocrystal in water keeping the Cs₄PbBr₆ concentration the same. As reported in Table 1, when we used 0.05 mL of water, we obtained 60% 0D Cs₄PbBr₆ and 40% 3D CsPbBr₃. As we increased the amount of water from 0.05 to 0.1 mL, we obtained 20% 0D Cs₄PbBr₆ and 80% 3D CsPbBr₃. At the end, when we used 0.2 mL water, we obtained 100% 3D CsPbBr₃.

Table 1. Variation of the % of 0D Cs₄PbBr₆, 1D, 2D, and 3D CsPbBr₃ NCs in the Presence of Different Amount of Water<sup>a</sup>

| amount of water (mL) | % of 0D Cs₄PbBr₆ | % of 3D CsPbBr₃ nanocubes | % of 2D CsPbBr₃ small nanosheets | % of 2D CsPbBr₃ nanoplatelets | % of 2D CsPbBr₃ big nanosheets | % of 1D CsPbBr₃ nanowires |
|----------------------|------------------|---------------------------|---------------------------------|-----------------------------|-------------------------------|--------------------------|
| 0                    | 100              | 0                         | 0                               | 0                           | 0                             | 0                        |
| 0.05                 | 60               | 40                        | 0                               | 0                           | 0                             | 0                        |
| 0.1                  | 20               | 80                        | 0                               | 0                           | 0                             | 0                        |
| 0.2                  | 0                | 100                       | 0                               | 0                           | 0                             | 0                        |
| 0.5                  | 0                | 80                        | 20                              | 0                           | 0                             | 0                        |
| 1.0                  | 0                | 50                        | 50                              | 0                           | 0                             | 0                        |
| 3.0                  | 0                | 0                         | 35                              | 65                          | 0                             | 0                        |
| 5.0                  | 0                | 0                         | 0                               | 100                         | 0                             | 0                        |
| 7.0                  | 0                | 0                         | 40                              | 60                          | 0                             | 0                        |
| 10.0                 | 0                | 0                         | 5                               | 70                          | 25                            | 0                        |
| 13.0                 | 0                | 0                         | 0                               | 30                          | 70                            | 0                        |
| 15.0                 | 0                | 0                         | 0                               | 0                           | 100                           | 0                        |

<sup>a</sup>The percentages of different shapes were calculated from the TEM image. For this purpose, we used 10 TEM pictures and averaged them.

Table 2. Relationship between the Amount of Water and the Morphology of 1D, 2D, and 3D CsPbBr₃ NCs<sup>a</sup>

| amount of water (mL) | morphology | size (nm, measured by TEM) | size (nm, measured by DLS) | emission maxima (nm) | IP PLQY (%) | 2P absorption cross section |
|----------------------|------------|---------------------------|---------------------------|----------------------|-------------|-----------------------------|
| 0.2                  | nanocubes  | $L = 25 \pm 5$            | $L = 28 \pm 6$           | 528                  | ~68         | $8.1 \times 10^4$           |
| 5.0                  | nanoplatelet | $L = 20 \pm 4$           | $20 \pm 5$               | 484                  | ~86         | $4.8 \times 10^3$           |
| 15.0                 | nanowire   | $L = 200 \pm 50$          | $220 \pm 70$             | 534                  | ~76         | $2.3 \times 10^5$           |

<sup>a</sup>Size distribution for 1D, 2D, and 3D CsPbBr₃ NCs measured by TEM/SEM and DLS. Single-photon and two-photon optical properties for 1D, 2D, and 3D CsPbBr₃ NCs.

are highly monodispersed with size of 25 ± 5 nm. The HRTEM image as inserted in Figure 1 shows clear lattice fringes with interplanar spacing ($d$) of ∼0.55 nm corresponding to the (110) crystal plane of the CsPbBr₃ cubic phase. As shown in Figure 2A, 3D CsPbBr₃ is retained the cubic phase, where main peaks are assigned to be the (100), (110), (111), (200), (211), and (220) planes of the cubic lattice.¹ The photoluminescent quantum yield for 3D CsPbBr₃ was measured to be ∼68%. Figure 3E shows that the stability in water for 3D CsPbBr₃ developed using the water-triggered process is much better than that of 3D CsPbBr₃ developed using the hot injection method, which can be attributed to the protection by the surface −OH groups.

To understand how the water amount variation can change the morphology of the product, we performed the same experiment by varying the water amount from 0.05 to 0.2 mL and keeping the Cs₄PbBr₆ concentration the same. As reported in Table 1, when we used 0.05 mL of water, we obtained 60% 0D Cs₄PbBr₆ and 40% 3D CsPbBr₃. As we increased the amount of water from 0.05 to 0.1 mL, we obtained 20% 0D Cs₄PbBr₆ and 80% 3D CsPbBr₃. At the end, when we used 0.2 mL water, we obtained 100% 3D CsPbBr₃.

In the next step for the development of biocompatible 3D CsPbBr₃ nanocrystals, we redispersed the nanocrystal in water with PEG under stirring for 40 min. Finally, the suspension was sonicated for a few minutes, and PEG coated nanocrystals were collected through filtration. As reported in Figure S1A, B, the TEM image from freshly prepared PEG coated 3D CsPbBr₃ shows that the size variation is within 35 ± 5 nm. The increase in size from 25 ± 5 to 35 ± 5 nm is mainly due to the coating of PEG. FTIR spectra as reported in Figure 2B shows −OH vibrational peaks from PEG coated 3D CsPbBr₃, which is mainly due to hydroxy coating originated during water triggered synthesis. We also observed peaks for the C−O stretch, −NH stretch, and −NH bend, which are due to the propionic acid−PEG−NH₂ coating. Other observed vibrational peaks are mainly due to OA and OAme. The XRD data, reported in Figure 2D, show that PEG coated 3D CsPbBr₃ does retain the cubic phase after it was encapsulated with polymers.

Amine−PEG−propionic acid (M₉, 1100) coated CsPbBr₃ nanocrystals were highly dispersed in water for several weeks. We did not observe any aggregation even after 21 days. To understand how the variation of PEG concentration affects the water dispersity and single- and two-photon luminescence behavior of CsPbBr₃ nanocrystals, we performed the PEG coating experiment with the change in concentration of PEG from 0.5 to 10 mg/mL. From the experimental observation, we found that the water dispersity increases as we increase concentration of PEG from 0.5 to 5 mg/mL, and after that, it remains constant with the increase in PEG concentration.

On the other hand, single- and two-photon luminescence behavior remains the same as we increase the concentration of PEG from 0.5 to 3 mg/mL. After that, the PLQY decreases with the increase in concentration of PEG. As a result, for our experiment, we used 3 mg/mL PEG.

The photograph reported as Figure S1C, D and the luminescence spectra as reported in Figure 2E show that the luminescence color and spectra remain the same after polymer coating. The photoluminescence quantum yield for 3D CsPbBr₃ nanocubes is measured using eq 1.¹⁵⁻³⁰

\[
\text{PLQY} = \frac{N_{\text{emit}}}{N_{\text{absorb}}} \tag{1}
\]

where $N_{\text{emit}}$ and $N_{\text{absorb}}$ are numbers of emitted and absorbed photons, respectively, for 3D CsPbBr₃. From the experimental measurement, we estimated that the photoluminescent quantum yield for PEG coated 3D CsPbBr₃ is ~66%.

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Figure 3C and D shows that the stability in water for PEG coated 3D CsPbBr$_3$ is excellent, and our data indicate that more than 86% IP luminescence intensity remains after 35 days under water. On the other hand, as reported in Figure 3E, luminescence intensity became zero after 8 days under water for 3D CsPbBr$_3$ without PEG and was developed using water triggered method. All the above experimental data clearly show that for long-term stability, the PEG coating is very important.

The linear absorption cross-section ($\sigma_{\text{lin}}$) and molar extinction coefficients ($\varepsilon$) for the PEG coated 3D CsPbBr$_3$ nanocubes were determined using inductively coupled plasma mass spectrometry (ICP-MS) combined with UV−vis, as we and others have reported before. From the elemental analysis data, we obtained $\sigma_{\text{lin}} = 1.6 \times 10^{-14}$ cm$^2$ PEG coated 3D CsPbBr$_3$ nanocubes at 400 nm. We also confirmed the values using one-photon induced ground state bleaching (GSB) data derived from femtosecond-transient absorption spectroscopy. From the transient absorption spectroscopy data, we obtained $\sigma_{\text{lin}} = 1.2 \times 10^{-14}$ cm$^2$ at 400 nm. From the $\sigma_{\text{lin}}$ value, we determined the $\varepsilon$ for PEG coated 3D CsPbBr$_3$ nanocubes, which was $\sim 3.1 \times 10^6$ L cm$^{-1}$ mol$^{-1}$ at 400 nm. Experimental data match quite well with the reported values.

It is now well-documented that the photostability of CsPbX$_3$ perovskites is a very important issue for their practical applications in optical devices. To determine the photostability of the PEG coated CsPbX$_3$ perovskites, we exposed perovskites with 360 nm UV-light irradiation. For this purpose, we exposed 30 mW/cm$^2$ power UV light for several hours. We monitored how PL intensity changes during the exposure to UV light. Figure 3F shows very good photo stability for PEG coated 3D CsPbBr$_3$ nanocubes when they are placed under UV light for several hours.
2.2. Water Triggered Synthesis of 2D CsPbBr\textsubscript{3} Nanoplatelets from 0D Cs\textsubscript{4}PbBr\textsubscript{6} Nanocrystals Using 5 mL of Water

For the development of 2D CsPbBr\textsubscript{3} nanoplatelets, we used 1 mM of CsPbBr\textsubscript{3} solution in hexane and added 5 mL of water. We kept the solution undisturbed for several hours at room temperature. It is very interesting to note that just by varying the amount of water, we can vary the dimension of CsPbBr\textsubscript{3} perovskite crystals. To understand the evolution process from 0D Cs\textsubscript{4}PbBr\textsubscript{6} nanocrystals to 2D CsPbBr\textsubscript{3} nanoplatelets, we performed a time dependent TEM study, as reported in Figure 4A. TEM data show that within 20 min, nanocubes start forming, and as a result, we observed a mixture of 0D Cs\textsubscript{4}PbBr\textsubscript{6} and 3D CsPbBr\textsubscript{3} nanocubes after 20 min of the water triggered synthesis process. After 80 min of the water triggered synthesis process, we observed the combination of 3D CsPbBr\textsubscript{3} nanocubes, 2D nanosheets, and 2D nanoplatelets. As shown in Figure 4A, after 260 min of the water triggered synthesis process, we observed mainly 2D nanoplatelets. As we have discussed before, during the water triggered synthesis process, dropping of CsBr into water happens through the interface. It is possible that when we use 0.2 mL of water, the amount of water is enough to form only cubic crystals. In the presence of more water (5 mL in this case), more and more capping ligands are partially detached from the surface, and a higher amount of [PbBr\textsubscript{6}]\textsuperscript{4−} octahedrons are formed in the system. As a result, the stability of the initial hexagons became lower. In this condition, formation of 2D CsPbBr\textsubscript{3} nanoplatelets occurs through a self-organization process, as shown in Figure 4. The XRD spectra reported in Figure 2F show only two peaks which correspond to (100) and (200). XRD results and TEM images reported in Figures 1 and 4 evidently reveal that the final product is 2D CsPbBr\textsubscript{3} nanoplatelets. Reported TEM data and DLS data in Table 2 indicate that the size of nanoplatelets is ∼20 ± 4 nm and the thickness is ∼4 nm. The HRTEM image, as inserted in Figure 1, shows clear lattice fringes with interplanar spacing (d) of ∼0.29 nm corresponding to the (200) crystal plane of CsPbBr\textsubscript{3}.

To understand how varying the water amount can change the morphology of the product, we performed the same experiment by varying the water amount from 0.5 to 5 mL and keeping Cs\textsubscript{4}PbBr\textsubscript{6} concentration the same. As reported in Table 1, when we used 0.5 mL of water, we obtained 80% 3D CsPbBr\textsubscript{3} nanocubes and 20% 2D small nanosheets. As we increased the amount of water from 0.5 to 3 mL, we obtained 35% 2D small nanosheets and 65% 2D CsPbBr\textsubscript{3} nanoplatelets. At the end, when we used 5 mL of water, we obtained 100% 2D CsPbBr\textsubscript{3} nanoplatelets.

The photoluminescent quantum yield for 2D CsPbBr\textsubscript{3} nanoplatelets was measured to be ∼86% using eq 1, which is mainly due to the quantum confining effect.5−20 Because the thickness of 2D CsPbBr\textsubscript{3} nanoplatelets (4 nm) is much less than the Bohr diameters for CsPbBr\textsubscript{3} (7 nm),1−10 one can expect excellent quantum confinement. In the next step, we developed PEG coated nanocrystals using the method we have discussed before. The XRD data, as reported in Figure 2F, show that PEG coated 2D CsPbBr\textsubscript{3} nanoplatelets does retain the same phase with 2D CsPbBr\textsubscript{3} nanoplatelets before encapsulation. The photoluminescent quantum yield for PEG coated 2D CsPbBr\textsubscript{3} nanoplatelets was measured to be ∼84%. Figure 3D show that the stability in water for PEG coated 2D CsPbBr\textsubscript{3} nanoplatelets is excellent and experimental data show that more than 80% IP luminescence intensity remain after 35 days under water.

From the elemental analysis data, we obtained σ\textsubscript{in} = 4.2 × 10−14 cm\textsuperscript{2} for PEG coated 2D CsPbBr\textsubscript{3} nanoplatelets at 400 nm. From σ\textsubscript{in} value we determined the ε for 2D CsPbBr\textsubscript{3} nanocrystals, which was ∼9.2 × 10\textsuperscript{6} L cm\textsuperscript{−1} mol\textsuperscript{−1} at 400 nm. Experimental data match quite well with the reported values.50−52

2.3. Water Triggered Synthesis of 1D CsPbBr\textsubscript{3} Nanowires from 0D Cs\textsubscript{4}PbBr\textsubscript{6} Nanocrystals Using 15 mL of Water

For the development of 1D CsPbBr\textsubscript{3} nanowires, we used 6 mM of CsPbBr\textsubscript{3} solution in cyclohexane and we added 15 mL of water. We kept it undisturbed for several hours at room temperature. To understand the possible mechanism, we performed time dependent TEM study as reported in Figure 4B. TEM data show that within 60 min of the water triggered synthesis process, big size 2D-CsPbBr\textsubscript{3} large nanosheets are formed. After 210 min of the water triggered synthesis process, we observed the combination if 2D nanosheets and 1D nanowires.
As shown in Figure 4B, after 510 min of the water triggered synthesis process, we observed 100% of 1D nanowires. As we have discussed before, water, as a polar solvent, can help to activate the nanocrystal surface by decreasing the surface density of the capping ligand.\(^{30-40}\) The above process also increases \([\text{PbBr}_6]^{4-}\) octahedron concentration by redissolving some of the already formed nanocubes, which will yield a higher growth rate\(^ {1a-i}\). Because both \(\text{H}_3\text{O}^+\) and \(\text{OH}^-\) can act as surface ligands with higher activity as compared to OA and OAm, in the presence of water, it changes the perovskite orientation growth to form nanowires, which could lower the surface energy.\(^ {30-44}\) The XRD spectra reported in Figure 3B shows only two peaks which correspond to (100) and (200). XRD result and TEM image reported in Figure 1 and Figure 5 evidently reveal that the final product is 1D \(\text{CsPbBr}_3\) nanowires. Reported TEM data in Figures 1 and 5 and DLS data as reported in Table 2, indicate that the length of nanowires is \(\sim 200 \pm 50\) nm and diameter is \(\sim 10\) nm. The HRTEM image, as inserted in Figure 1, shows clear lattice fringes with interplanar spacing \((d) \sim 0.55\) nm corresponding to the (100) crystal plane of \(\text{CsPbBr}_3\).

To understand how the water amount variation can change the morphology of the product, we performed the same experiment by varying the water amount from 7 to 15 mL and keeping \(\text{Cs}_4\text{PbBr}_6\) concentration the same. As reported in Table 1, when we used 7 mL water, we obtained 40% 2D \(\text{CsPbBr}_3\) nanoplatelets and 60% 2D \(\text{CsPbBr}_3\) big nanosheets. As we increased the amount of water from 7 to 13 mL, we obtained 30% 2D \(\text{CsPbBr}_3\) big nanosheets and 70% 1D \(\text{CsPbBr}_3\) nanowires. At the end when we used 15 mL water, we
obtained 100% 1D CsPbBr₃ nanowires. The photoluminescent quantum yield for 1D CsPbBr₃ nanowires was measured to be \( \sim 76\% \) using eq 1. The obtained high PLQY is mainly due to the quantum confinement effect to some extent. Because the thickness of 1D CsPbBr₃ nanowires (8 nm), which is slightly higher than Bohr diameters for CsPbBr₃ (7 nm), one can expect quantum confinement to some extent. In the next step, we developed PEG coated nanocrystals using the method we have discussed before.

The XRD data, reported in Figure 3B, show that PEG coated 1D CsPbBr₃ nanowires do retain the same phase with 1D CsPbBr₃ nanowires before encapsulation. The photoluminescent quantum yield for PEG coated 2D CsPbBr₃ nanoplatelets was measured to be \( \sim 74\% \). Figure 3D shows that the stability in water for PEG coated 1D CsPbBr₃ nanowires is excellent, and experimental data show that more than 85% IP luminescence intensity remains after 35 days under water.

From the elemental analysis data, we obtained \( \sigma_{\text{in}} = 3.4 \times 10^{-14} \) cm² for PEG coated 1D CsPbBr₃ nanowires at 400 nm. From the \( \sigma_{\text{in}} \) value, we determined \( \epsilon \) for 1D CsPbBr₃ nanowires, which was \( \sim 7.3 \times 10^6 \) L cm⁻¹ mol⁻¹ at 400 nm. Experimental data match quite well with the reported values.⁵⁰⁻⁵²

Figure 3F shows very good photostability for PEG coated 1D CsPbBr₃ nanorods when they are place under UV light for several hours. Strong PLQY and excellent photostability and water resistance make PEG coated 1D, 2D, and 3D-CsPbBr₃ nanocrystals very good candidates for bioimaging applications.

### 2.4. Two-Photon Absorption Properties for 1D, 2D, and 3D CsPbBr₃ Nanocrystals

We used TPL spectroscopy and Z-scan technique for the determination of the two-photon absorption properties for PEG coated 1D, 2D, and 3D CsPbBr₃ nanocrystals.⁴⁵⁻⁵⁴ Experimental details have been reported before and are discussed in the Supporting Information.⁴⁷,⁵³,⁵⁸⁻⁶⁰,⁶⁸⁻⁷¹ In brief, for the measurement, we used an 80 MHz Ti:sapphire laser as an excitation source with 100 fs pulse width. ⁴⁷,⁵³,⁵⁸⁻⁶⁰ We used an optical parametric amplifier to generate 800 nm excitation wavelength. ³⁴,⁴⁷,⁵³,⁵⁸,⁶⁰ The laser beam was focused down to a radius of \( \sim 30 \) μm on the sample.

To determine the photostability of the PEG coated CsPbX₃ perovskites during 2P absorption and Z-scan experiments, we determined how PL intensity changes during the exposure to 800 nm NIR light. For this purpose, we used 100 mW 800 nm light. As shown in Figure S2A in the Supporting Information, PL intensity for PEG coated 1D CsPbBr₃ nanocrystals remains about same during the exposure to 800 nm NIR light for 15 min. Because in the 2P and Z-scan experiments 800 nm light exposure time is less than 10 min, the photostability for CsPbBr₃ nanocrystals will remain the same. We also performed an XRD experiment before and after the 2P experiment. XRD data reported in Figure S2B indicate that CsPbBr₃ nanocrystals are retained in the same phase, which indicates no optical damage within our experimental time range.

Figure 5A shows the normalized 1P and 2P luminescence spectra from 1D CsPbBr₃ nanowires using 400 and 800 nm excitation light, which indicates that one- and two-photon absorption are very identical. It may be because the nanocrystals excited by either 1P or 2P absorption will relax to the same lowest excited state.⁴⁵⁻⁵⁴ To understand better, we also performed the excitation intensity dependent 2P luminescence measurements. As shown in Figures 5B–E, the quadratic dependence of the luminescence intensity from 3D CsPbBr₃ nanocubes and 2D CsPbBr₃ nanoplatelets clearly confirms the 2P processes. The absolute two-photon absorption cross sections for PEG coated 1D, 2D, and 3D CsPbBr₃ nanocrystals were determined using fluorescein as a reference, whose 2PA is known in literature.¹⁻³ Using the reference, 2PA cross-section for PEG coated 1D CsPbBr₃ nanowires was determined using eq 2.⁴⁴⁻⁵⁴

\[
\sigma_{\text{nannowires}} = \sigma_{\text{FL}}(F_{\text{nannowires}}/F_{\text{FL}})(\Phi_{\text{FL}}/\Phi_{\text{nannowires}}) (C_{\text{FL}}/C_{\text{nannowires}}) \tag{2}
\]

where \( F \) is the observed fluorescence intensity from fluorescein and PEG coated 1D CsPbBr₃ nanowires. Similarly, \( \Phi \) is the quadratic dependence of the luminescence intensity from 3D CsPbBr₃ nanocubes, and \( C \) is the concentration of fluorescein and PEG coated 1D CsPbBr₃ nanowires. For the TPL experiment, we used \( \sim 100 \) nM CsPbBr₃ nanocrystals. Using experimental data, we determined the two-photon absorption cross sections for 1D CsPbBr₃ nanowires to be \( 5.1 \times 10^5 \) GM at 800 nm excitation. We also determined the 2P cross-section using the Z-scan technique,⁴⁵⁻⁵⁴ as shown in Figure 5F. The transmission for open aperture Z-scan was measured using eq 3.⁴⁴⁻⁵⁴ for PEG coated 1D, 2D, and 3D CsPbBr₃ nanocrystals,

\[
T_{\text{open}}(Z) = 1 - 1/2\sqrt{2[(\alpha_{\text{eff}}L_{\text{eff}})]/[1 + (z/z_0)^2]} \tag{3}
\]

where \( L_{\text{eff}} = (1 - e^{-aL})/a_{\text{0}} \) and \( L \) is the sample thickness for PEG coated 1D, 2D, and 3D CsPbBr₃ nanocrystals. Similarly, \( L_{\text{fl}} \) is the on-axis peak intensity. \( z \) is the longitudinal displacement of the sample from the focus, and \( z_0 \) is the Rayleigh diffraction length. After subtraction of the solvent contribution to the measured two-photon signal, we determined the 2PA absorption coefficients for PEG coated 1D, 2D, and 3D CsPbBr₃ nanocrystals. Using Z-scan data, we determined 2PA coefficients of PEG coated 1D, 2D, and 3D nanocrystals, and these are \( \sigma_2 \sim 2.3 \times 10^5 \) GM for 1D nanowires, \( \sigma_2 \sim 4.8 \times 10^7 \) GM for 2D nanoplatelets, and \( \sigma_2 \sim 8.1 \times 10^6 \) GM for 3D nanocubes (1 GM = \( 1 \times 10^{-36} \) cm²·s·photon⁻¹), which are several orders of magnitude higher than that of organic chromophores (>100 GM). Reported 2P cross-sections for 2D and 3D nanocrystals match very well with the reported data.⁴⁴⁻⁵⁴

Because the volumes of 1D, 2D, and 3D nanocrystals are different, we calculated volume-normalized 2PA cross sections (in GM nm⁻³⁻¹) for 1D, 2D, and 3D nanocrystals, and these are \( \sigma_2 \sim 28.5 \) (GM nm⁻³⁻¹) for 1D nanowires, \( \sigma_2 \sim 430 \) (GM nm⁻³⁻¹) for 2D nanoplatelets, and \( \sigma_2 \sim 9.2 \) (GM nm⁻³⁻¹) GM for 3D nanocubes. Our observed highest VN 2PA cross-section for 2D nanoplatelets can be due the higher transition dipole moment attributed to stronger confinement effect than that of 3D nanocubes and 1D nanowires. ⁵⁰,⁵³,⁵⁴,⁵⁸ To understand whether the PEG coating enhances the 2PA cross-section for 1D, 2D, and 3D CsPbBr₃ nanocrystals, we also measured the 2PA cross-section for 1D, 2D, and 3D CsPbBr₃ nanocrystals with PEG. Using Z-scan data, we determined 2PA coefficients of 1D, 2D, and 3D nanocrystals, and these are \( \sigma_2 \sim 2.1 \times 10^4 \) GM for 1D nanowires, \( \sigma_2 \sim 4.9 \times 10^4 \) GM for 2D nanoplatelets, and \( \sigma_2 \sim 8.5 \times 10^4 \) GM for 3D nanocubes, which are very similar to the 2PA properties for PEG coated 1D, 2D, and 3D CsPbBr₃ nanocrystals.
2.5. Two-Photon Imaging of Cancer Cells Using 1D, 2D, and 3D CsPbBr3 Nanocrystals

Inspired by the excellent two-photon absorption coefficients, very good water resistance capability, and good photostability, we attempted to explore the use of anti-AXL antibody attached CsPbBr3 nanoplatelet and anti-HER-2 antibody attached nanowire probes for tracking TNBC and HER-2(+) SK-BR-3 cells. Initially, we determined the biocompatibility for nanocrystals. For this purpose, we incubated PEG coated 2D nanoplatelets with 2.6 × 10^5 cells per mL normal HaCaT skin cells, HER-2(+) SK-BR-3 cells, MDA-MB-231 TNBC cells, and LNCaP human prostate cancer cells separately for 24 h. After that, the numbers of live cells were measured using an MTT test. As reported in Figure 6A, PEG coated 2D nanoplatelets show excellent biocompatibility. We have also performed a concentration dependent biocompatibility experiment for PEG coated 2D nanoplatelets using normal HaCaT skin cells, HER-2(+) SK-BR-3 breast cells, MDA-MB-
231 TNBC cells, and LNCaP human prostate cancer cells separately. As reported in Figures 5A–D, our experimental data show that PEG coated 2D nanoplatelets show excellent biocompatibility even at the concentration of 50 μg/mL. The observed very good biocompatibility is mainly due to the presence of the PEG coating, which helps CsPbBr3 perovskites not to decompose when exposed to water.

After that, the antibody was attached to the PEG coated 2D nanoplatelets and 1D nanowires via noncovalent interaction through PEG. Next, different concentrations of antibody-conjugated nanocrystals were mixed with cancer cells for 30 min. In the next step, unbound antibody-conjugated nanocrystals were separated using centrifugation. For the two-photon cancer imaging experiment, we used a Nikon multiphoton microscope (FV1000MPE), as we reported before.55,58,60,65,66 Figure 6B shows that anti-HER-2 antibody conjugated nanocrystals are attached to the surface of HER-2(+) SKBR-3 breast cancer cells. Figure 6C shows that anti-AXL antibody conjugated 2D nanoplatelets can be used for blue color two-photon imaging of TNBC cells. Similarly, Figure 6D shows that anti-HER-2 antibody conjugated 1D nanowires can be used for green color two-photon imaging of HER-2 (+) SK-BR-3 cells. The bright field images are reported in Figures S4A–C in the Supporting Information.

To find out the selectivity, we performed the experiment for HER-2(−) TNBC cells using anti-HER-2 antibody conjugated 1D nanowires. As reported in Figure 6E, we did not observe any luminescence image because anti-HER-2 antibody conjugated 1D CsPbBr3 nanowires do not bind with HER-2(−) MDA-MB-231 TNBC cells. The above data indicate that antibody attached 1D, 2D, and 3D CsPbBr3 nanocrystals are highly selective for targeted cancer cells. Next, we determined whether a mixture of anti-HER-2 antibody attached 1D nanowires and anti-AXL antibody conjugated 2D nanoplatelets can be used for the simultaneous identification of MDA-MB-231 TNBC cells and HER-2(+) breast cancer cells together. Figure 6F shows a multicolor blue/green two-photon luminescence image which shows that HER-2(+) SK-BR-3 cells and MDA-MB-231 TNBC cells can be simultaneously imaged using 2D nanoplatelets and 1D nanowires.

3. CONCLUSIONS

In conclusion, our findings reveal that water resistant, biocompatible, and photostable 1D CsPbBr3 nanowires, 2D CsPbBr3 nanoplatelets, and 3D CsPbBr3 nanocubes can be designed through an interfacial conversion from 0D Cs4PbBr6 nanocrystals via a water triggered strategy using water and PEG. Reported data show that just by varying the amount of water one can control the dimension of CsPbBr3 perovskite crystals. Time dependent TEM and emission data show the possible pathway for the formation of 1D CsPbBr3 nanowires, 2D CsPbBr3 nanoplatelets, and 3D CsPbBr3 nanocubes from 0D Cs4PbBr6 nanocrystals. Our experimental results find that these nanocrystals exhibit excellent IP luminescence PLQY and 2PL absorption coefficients. Reported data also show that more than 86% of IP luminescence intensity from PEG coated nanocrystals remained after 35 days under water. Experimental results show that selective and simultaneous two-photon imaging of TNBC cells and HER-2 (+) SKBR3 breast cancer cells in a near-IR biological window can be performed using specific antibody attached 1D and 2D perovskites.

As our reported data show, 1D CsPbBr3 nanowires and 2D CsPbBr3 nanoplatelets exhibit very high PLQY and good photostability in water, which are much superior to that of commonly used organic probes; after proper engineering design, they will be better probes for bioimaging application. In addition, lead halide perovskite nanocrystals exhibit high absorption coefficients for two-, three- and five-photon processes, which is rare for commonly used organic probes that are used as fluorescence labels. These excellent properties provide advantages for all-inorganic cesium lead halide perovskite-based nanocrystals to be useful in the field of bioimaging. However, there are many issues that need to be addressed before 1D CsPbBr3 nanowires and 2D CsPbBr3 nanoplatelets can be used for real life applications, and these are stability in physiological conditions; possible toxicity due to degradation; interaction with DNA, RNA, and other biomolecules during circulation; epigenetic interaction, etc.

ASSOCIATED CONTENT

# Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/jacsau.0c00038.

Detailed synthesis, characterization of antibody attached nanowires, and other experiments (PDF)

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Notes

The authors declare no competing financial interest.

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