Establishment of accurate axillary lymph node status is of essential importance in determining both prognosis and the potential need for adjuvant therapy in patients with invasive breast cancer. Axillary lymph node heterotopias can in some cases result in overdiagnosis of metastatic disease. Nodal endosalpingiosis is perhaps the least commonly reported type of axillary lymph node heterotopia. We herein illustrate a case in which second opinion pathologic interpretation combined with ancillary immunohistochemical studies allowed for a specific diagnosis of axillary nodal müllerian-type inclusions, confirming ypN0 staging and resulting in appropriate disease management and prognostication.

1. Introduction

Axillary lymph node status is one of the most important prognostic indicators for patients with invasive breast carcinoma, and intraoperative pathologic examinations of sentinel lymph nodes are performed at many centers to accurately stage patients as well as help determine the need for completion axillary dissections [1–4]. Various benign epithelioid cellular mimics of metastatic breast carcinoma in lymph nodes have been reported. These include but are not necessarily limited to heterotopic benign mammary epithelium, displaced cutaneous adnexal epithelial structures, nevus cell clusters, squamoid inclusions, mesothelial cell rests, and, most recently, müllerian-type epithelial inclusions (endosalpingiosis) [5–18]. All of these entities can be overinterpreted as metastases involving sentinel lymph nodes on intraoperative studies and/or in final pathologic evaluations of permanent sections. Such overcalls can result in falsely elevated nodal staging. Overdiagnosis of heterotopias as metastases may result in incorrect prognostic information and can potentially lead to overtreatment with surgical, radiation, and systemic therapies. Accurate interpretations are of paramount importance as misinterpretations of microscopic nodal structures may result in disproportionately large treatment changes. The histopathologic changes of endosalpingiosis are relatively commonly encountered in pelvic lymph nodes of adult females undergoing staging for gynecologic malignancies; however, such changes are perhaps the least commonly encountered type of benign epithelial inclusion in axillary nodes of women with breast carcinoma [19–28]. We herein report a case of endosalpingiosis involving an axillary lymph node in an adult female with a known diagnosis of invasive ipsilateral breast carcinoma.

2. Case Presentation

The patient was a 70-year-old female with a known history of invasive ductal carcinoma of the left breast. Her primary breast tumor was diagnosed via histologic assessment of an image-guided core biopsy at another facility. The invasive carcinoma was reported to measure more than 5 cm (clinical
stage T3) by imaging studies. She was advised to undergo pre-operative neoadjuvant chemotherapy. Upon completion of her neoadjuvant therapy, 6 months after the time of her original core biopsy diagnosis of invasive disease, the patient underwent left mastectomy and sentinel lymph node biopsies at the outside facility. No residual carcinoma was identified in the mastectomy specimen (complete histologic response to neoadjuvant therapy/ypT0); however, glandular epithelial structures were identified within one of the sentinel lymph nodes, and the outside pathologist forwarded the slides and tissue blocks from the lymph nodes and the original diagnostic core biopsy sample to our center for expert extradepartmental consultation. The best diagnosis for the glandular elements in the lymph nodes was in question.

The outside slides were reviewed, and the original core biopsy diagnosis of invasive ductal carcinoma, nuclear grade 2, was confirmed (Figure 1). The invasive tumor was 70% estrogen receptor positive by immunohistochemistry and was HER2 nonreactive/negative by IHC, scaled score 1+. The slides from one of the small, postneoadjuvant, excised sentinel lymph nodes demonstrated multiple (at least nine) distinct nests of glandular epithelium within the lymph node capsule (Figure 2). Some of these epithelial nests abutted the subcapsular sinus. These nodal capsular glandular cell groups were discontinuous and were seen to encircle approximately one-half of the circumference of the 3.5 mm lymph node. The histologic differential diagnosis for these glandular structures within the lymph node included metastatic disease versus benign/reactive heterotopic epithelium.

Histomorphologic comparisons between the capsular-associated glandular structures in the node and the patient’s primary breast carcinoma core biopsy slides confirmed disparate appearances. The primary tumor showed greater nuclear pleomorphism and less overt tubule formation. In addition, the primary tumor was mitotically active, and no mitoses were seen in the nodal capsular epithelioid nests. Ancillary immunohistochemical (IHC) studies performed in our laboratory confirmed that the glandular cell groups in the periphery of the patient's sentinel node were immunoreactive (positive) for PAX8 (Figure 3) and WT1 (Figure 4) and negative for GATA3 (Figure 5). Slides from the original diagnostic breast core biopsy block were also prepared in our laboratory, and the invasive tumor was found to be positive for GATA3 (Figure 6) and negative for PAX8 and WT1, confirming a pattern of protein expression opposite of the glandular structures in the sentinel lymph node capsule. On 100x oil-immersion microscopy, rare singly scattered ciliated cells could be identified in the capsular nodal rests (Figure 7). A diagnosis of nodal endosalpingiosis (benign lymph node
capsular müllerian inclusions) was rendered, and the patient’s disease was then able to be accurately defined as ypT0, ypN0.

### 3. Discussion

On histologic examination, Müllerian-type glandular inclusions lined by cuboid to columnar epithelial cells that are reminiscent of fallopian tube type lining or coelomic type lining are commonly encountered within the capsules and/or peripheral cortices of pelvic lymph nodes from adult females. Case reports of endosalpingiosis specific to axillary lymph nodes are rare and have begun to appear within the literature in the last decade [22–26, 28]. The origins of gynecologic type epithelial and stromal heterotopias in lymph nodes are attributed by some to a theory of “müllerianosis” in which embryonic müllerian tissues that are displaced during organogenesis can result in the formation of at least four benign disease states, namely, developmental adenomyosis, endometriosis, endosalpingiosis, and endocervicosis [29]. In addition to a few previously reported cases of axillary nodal endosalpingiosis, one report of axillary nodal endocervicosis also exists [27]. All of the conditions included in the broader category of müllerianiasis have been described as choristomal organoid structures composed of müllerian rests incorporated into other normal tissues/organs. All müllerian heterotopias are considered to be benign and can be definitively surgically managed [29]. The medical/scientific literature is uncertain as to whether or not endosalpingiosis can represent a precursor of malignancy. In addition to lymph nodes, heterotopic inclusions of tubal type epithelium have been reported in the peritoneal lining (by both biopsy and cytology), in the female reproductive organs including the cervix/uterus/ovaries, and in the urinary bladder, as well as in the soft tissues of the mediastinum and in the spleen [29–32]. A mouse model of morphologic ovarian endosalpingiosis exists with the describing authors suggesting that ovarian cortical epithelial cysts may be derived from the highly plastic ovarian surface epithelium resulting in endosalpingiosis-like ovarian inclusions [33].

The epithelial cells of endosalpingiosis are cytologically bland and are not seen in association with tissue invasion or associated desmoplasia. Ciliation, when present, is a reassuring feature, helping to confirm a benign diagnosis; however, many cases (such as our index case) present with a cuboidal cell morphology either completely lacking cilia or with only rare ciliated cells. Nonciliated forms of müllerianiasis can be especially challenging to separate from metastatic adenocarcinomas, and in some instances endosalpingiosis may undergo hyperplastic changes making the distinction from malignancy even more difficult. In such cases, ancillary IHC testing may be of great value. While estrogen receptor (ER) IHC testing may be of value in differentiating metastatic breast carcinoma from some heterotopic rests, such as mesothelial rests or squamoid inclusions, ER is not a useful marker for distinguishing true mammary heterotopias and/or endosalpingiosis in lymph nodes, as these entities may also be ER positive. Certain commercially available IHC markers such as PAX8 and WT1, transcription factors/regulators involved in the development of the female reproductive system, are good markers of müllerian origin. Immunoreactivity (positivity) is commonly seen by IHC analysis in both fallopian tube epithelium and ovarian surface epithelium as well as in neoplastic proliferations derived from these cell types [31, 34–38]. These markers are typically nonreactive in breast epithelium and in neoplasms derived from breast epithelium. Alternatively, GATA3, a transcription
factor important in breast luminal epithelial development, is immunoreactive by IHC studies in breast epithelium and in most tumors derived from breast epithelium [39, 40]. Some authors have shown sensitivities as high as 95% for GATA3 in labeling of mammary adenocarcinomas [41]. GATA3 IHC testing generally shows nonreactive (negative) results in müllerian epithelium and in epithelial lesions derived from the gynecologic tract. PAX8, WT1, and GATA3 IHC studies were of value in this case and helped to confirm the patient's ypN0 nodal status.

Pathologists should be aware that not all epithelioid glandular-appearing cell groups within lymph nodes and their capsules represent metastatic disease. Availing themselves of morphologic correlations between known primary tumor slides and subsequent nodal sampling slides is a good starting point when questions arise. IHC testing can be of great adjunctive value, and careful attention to cellular details, such as recognition of true ciliation, can prove immensely helpful. Seasoned breast clinicians who are the consumers of pathology reports and descriptions/images in multidisciplinary conferences should also be aware of axillary endosalpingiosis as an entity and should raise questions when appropriate. Unnecessary completion axillary lymph node dissections can be associated with arm movement disorders, chronic pain, paresthesias, lymphedema, and decreased quality of life. A correct diagnosis of axillary lymph node müllerianosis rests on a combination of awareness of existence and attention to light microscopic details. The goal of accurate classification is to avoid overdiagnosis of benign inclusions, müllerian and otherwise, so that staging and therapy are optimized for every patient.

Competing Interests

The authors declare that there are no competing interests regarding the publication of this paper.

References

[1] M. Gipponi, C. Bassetti, G. Canavese et al., “Sentinel lymph node as a new marker for therapeutic planning in breast cancer patients,” Journal of Surgical Oncology, vol. 85, no. 3, pp. 102–111, 2004.
[2] P. G. Roy, S. M. Chan, V. Ng, B. M. Smith, H. Umeh, and S. P. Courtney, “Risk stratification of patients with early breast cancer,” Clinical Breast Cancer, vol. 14, no. 1, pp. 68–73, 2014.
[3] R. Hong, Z. Dai, W. Zhu, B. Xu, and P. Chu, “Association between lymph node ratio and disease specific survival in breast cancer patients with one or two positive lymph nodes stratified by different local treatment modalities,” PLoS ONE, vol. 10, no. 10, Article ID e0138908, 2015.
[4] U. Veronesi, G. Viale, G. Paganelli et al., “Sentinel lymph node biopsy in breast cancer: ten-year results of a randomized controlled study,” Annals of Surgery, vol. 251, no. 4, pp. 595–600, 2010.
[5] R. Garret and A. E. Ada, “Epithelial inclusion cysts in an axillary lymph node: report of a case simulating metastatic adenocarcinoma,” Cancer, vol. 10, pp. 173–178, 1957.
[6] D. W. Edlow and D. Carter, “Heterotopic epithelium in axillary lymph nodes: report of a case and review of the literature,” American Journal of Clinical Pathology, vol. 59, no. 5, pp. 666–673, 1973.
[7] S. Longo, “Benign lymph node inclusions,” Human Pathology, vol. 7, no. 3, pp. 349–354, 1976.
[8] D. R. Turner and R. R. Millis, “Breast tissue inclusions in axillary lymph nodes,” Histopathology, vol. 4, no. 6, pp. 631–636, 1980.
[9] P. J. Holdsworth, J. M. Hopkinson, and S. H. Leveson, “Benign axillary epithelial lymph node inclusions—a histological pitfall,” Histopathology, vol. 13, no. 2, pp. 226–228, 1988.
[10] C. J. Fisher, S. Hill, and R. R. Millis, “Benign lymph node inclusions mimicking metastatic carcinoma,” Journal of Clinical Pathology, vol. 47, no. 3, pp. 245–247, 1994.
[11] P. Argani and J. Rosai, “Hyperplastic mesothelial cells in lymph nodes: report of six cases of a benign process that can simulate metastatic involvement by mesothelioma or carcinoma,” Human Pathology, vol. 29, no. 4, pp. 339–346, 1998.
[12] L. J. Layfield and E. Mooney, “Heterotopic epithelium in an intramammary lymph node,” Breast Journal, vol. 6, no. 1, pp. 63–67, 2000.
[13] E. Resetkova, S. A. Hoda, J. L. Clarke, and P. P. Rosen, “Benign heterotopic epithelial inclusions in axillary lymph nodes. Histological and immunohistochemical patterns,” Archives of Pathology & Laboratory Medicine, vol. 127, no. 1, pp. 25–27, 2003.
[14] E. Maiorano, G. M. Mazzarol, G. Pruner et al., “Ectopic breast tissue as a possible cause of false-positive axillary sentinel lymph node biopsies,” American Journal of Surgical Pathology, vol. 27, no. 4, pp. 513–518, 2003.
[15] S. Jaffer and I. J. Bleiweiss, “Regional metastases, artifacts and mimics in axillary lymph nodes,” Seminars in Breast Disease, vol. 8, no. 2, pp. 62–70, 2005.
[16] L. E. Norton, I. K. Komenaka, R. E. Emerson, C. Murphy, and S. Badve, “Benign glandular inclusions a rare cause of a false positive sentinel node,” Journal of Surgical Oncology, vol. 95, no. 7, pp. 593–596, 2007.
[17] Y. Peng, R. Ashfaq, G. Ewing, A. M. Leitch, and K. H. Molberg, “False-positive sentinel lymph node in breast cancer patients caused by benign glandular inclusions: report of three cases and review of the literature,” American Journal of Clinical Pathology, vol. 130, no. 1, pp. 21–27, 2008.
[18] G. Fellegara, M. L. Carcangiu, and J. Rosai, “Benign epithelial inclusions in axillary lymph nodes: report of 18 cases and review of the literature,” The American Journal of Surgical Pathology, vol. 35, no. 8, pp. 1123–1133, 2011.
[19] L. A. Karp and B. Czernobilsky, “Glandular inclusions in pelvic and abdominal para-aortic lymph nodes. A study of autopsy and surgical material in males and females,” American Journal of Clinical Pathology, vol. 52, no. 2, pp. 212–218, 1969.
[20] V. Maaßen and K. Hiller, “Glandular inclusions in lymph nodes: pattern of distribution and metaplastic transformation,” Archives of Gynecology and Obstetrics, vol. 255, no. 1, pp. 1–8, 1994.
[21] L.-C. Horn and K. Bilek, “Frequency and histogenesis of pelvic retroperitoneal lymph node inclusions of the female genital tract: an immunohistostochmic study of 34 cases,” Pathology Research and Practice, vol. 191, no. 10, pp. 991–996, 1995.
[22] A. D. Corben, T. Nehhozina, K. Garg, C. E. Vallejo, and E. Brogi, “Endosalpingiosis in axillary lymph nodes: a possible pitfall in the staging of patients with breast carcinoma,” The American Journal of Surgical Pathology, vol. 34, no. 8, pp. 1211–1216, 2010.
[23] A. H. Salehi, G. Omeroglu, Y. Kanber, and A. Omeroglu, “Endosalpingiosis in axillary lymph nodes simulating metastatic...
breast carcinoma: a potential diagnostic pitfall,” International Journal of Surgical Pathology, vol. 21, no. 6, pp. 610–612, 2013.

[24] M. Welsher and K. L. Snook, “Endosalpingiosis in an axillary lymph node with synchronous micro-metastatic mammary carcinoma,” Pathology, vol. 46, no. 7, pp. 665–667, 2014.

[25] J. V. Groth, S. Prabhu, and E. Wiley, “Coexistent isolated tumor cell clusters of infiltrating lobular carcinoma and benign glandular inclusions of müllerian (endosalpingiosis) type in an axillary sentinel node: case report and review of the literature,” Applied Immunohistochemistry & Molecular Morphology, vol. 24, no. 2, pp. 144–148, 2016.

[26] E. Carney, A. Cimino-Mathews, C. Argani, J. Kronz, R. Vang, and P. Argani, “A subset of nondescript axillary lymph node inclusions have the immunophenotype of endosalpingiosis,” American Journal of Surgical Pathology, vol. 38, no. 12, pp. 1612–1617, 2014.

[27] P. Mukonoweshuro and W. G. McCluggage, “Endocervicosis involving axillary lymph nodes: first case report,” International Journal of Gynecological Pathology, vol. 33, no. 6, pp. 620–623, 2014.

[28] U. Ozerdem and S. A. Hoda, “Endosalpingiosis of axillary sentinel lymph node: a mimic of metastatic breast carcinoma,” Breast Journal, vol. 21, no. 2, pp. 194–195, 2015.

[29] R. E. Batt and J. Yeh, “Müllerianosis: four developmental (embryonic) müllerian diseases,” Reproductive Sciences, vol. 20, no. 9, pp. 1030–1037, 2013.

[30] J. Park, T. H. Kim, H. H. Lee, S. Chung, and D. Jeon, “Endosalpingiosis in postmenopausal elderly women,” Journal of Menopausal Medicine, vol. 20, no. 1, pp. 32–34, 2014.

[31] N. Sneige, M. A. Dawlett, T. L. Kologinczak, and M. Guo, “Endosalpingiosis in peritoneal washings in women with benign gynecologic conditions: thirty-eight cases confirmed with paired box-8 immunohistochemical staining and correlation with surgical biopsy findings,” Cancer Cytopathology, vol. 121, no. 10, pp. 582–590, 2013.

[32] K. Maeda, F. Kojima, M. Ishida, M. Iwai, A. Kagotani, and A. Kawauchi, “Müllerianosis and endosalpingiosis of the urinary bladder: report of two cases with review of the literature,” International Journal of Clinical and Experimental Pathology, vol. 7, no. 7, pp. 4408–4414, 2014.

[33] S. K. Bristol-Gould, C. G. Hutten, C. Sturgis, S. M. Kilen, K. E. Mayo, and T. K. Woodruff, “The development of a mouse model of ovarian endosalpingiosis,” Endocrinology, vol. 146, no. 12, pp. 5228–5236, 2005.

[34] D. Tacha, D. Zhou, and L. Cheng, “Expression of PAX8 in normal and neoplastic tissues: a comprehensive immunohistochemical study,” Applied Immunohistochemistry and Molecular Morphology, vol. 19, no. 4, pp. 293–299, 2011.

[35] Y. Wang, Y. Wang, J. Li et al., “PAX8: a sensitive and specific marker to identify cancer cells of ovarian origin for patients prior to neoadjuvant chemotherapy,” Journal of Hematology and Oncology, vol. 6, article 60, 2013.

[36] L. Liliac, M. L. Carcangiu, S. Canevari et al., “The value of PAX8 and WT1 molecules in ovarian cancer diagnosis,” Romanian Journal of Morphology and Embryology, vol. 54, no. 1, pp. 17–27, 2013.

[37] C. Bárcena and E. Oliva, “WT1 expression in the female genital tract,” Advances in Anatomic Pathology, vol. 18, no. 6, pp. 454–465, 2011.

[38] G. Acs, T. Pasha, and P. J. Zhang, “WT1 is differentially expressed in serous, endometrioid, clear cell, and mucinous carcinomas of the peritoneum, fallopian tube, ovary, and endometrium,” International Journal of Gynecological Pathology, vol. 23, no. 2, pp. 110–118, 2004.

[39] H. Kouros-Mehr, J.-W. Kim, S. K. Bechis, and Z. Werb, “GATA3 and the regulation of the mammary luminal cell fate,” Current Opinion in Cell Biology, vol. 20, no. 2, pp. 164–170, 2008.

[40] S. M. Wendroth, M. J. Membrikoski, and M. R. Wick, “GATA3 expression in morphologic subtypes of breast carcinoma: a comparison with gross cystic disease fluid protein 15 and mammaglobin,” Annals of Diagnostic Pathology, vol. 19, no. 1, pp. 6–9, 2015.

[41] R. Asch-Kendrick and A. Cimino-Mathews, “The role of GATA3 in breast carcinomas: a review,” Human Pathology, vol. 48, pp. 37–47, 2016.