Dietary supplementation of chromium for finishing pigs

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ABSTRACT: Increasing fat deposition and fixed conversion ratio over the days in finishing phase directly influence pork quality and productive profitability. Nonetheless, the slaughter of heavier pigs can result in benefits for the slaughterhouse due to dilution of production and processing costs, as well as economic benefits for the pig producer resulting from the dilution of production costs. Therefore, dietary supplementation of chromium for finishing pigs of high lean-genotypes is a strategy to increase lean tissue accretion and minimize fat deposition, reflecting positively on growth performance. This review discussed recent studies results and mechanisms of action of this modifier of performance and carcass traits in finishing pigs. Chromium increases the insulin action, facilitating insulin binding to the receptors on cell membrane. As a result, insulin-sensitive cells uptake more glucose, which will be later converted into energy. This additional energy is use to increase protein synthesis, thereby increasing the amount of lean tissue and reducing fat content in the carcase, without altering protein intake by pigs. Chromium also reduces lipid oxidation rate maintaining meat quality for longer period. In conclusion, dietary Cr supplementation for finishing pigs have shown that 0.2 mg/kg of organic Cr sources for pigs from ~60 kg until the slaughter can improve growth performance, lean gain and reduce fat content in carcase. However, the development of nanotechnology has allowed the use of inorganic Cr source at 0.2 mg/kg of inclusion, leading to improve the growth performance and carcass traits finishing pigs.

Key words: carcass traits, growth performance, mineral, nutrition, swine.

Suplementação de cromo na dieta de suínos em termoacção

RESUMO: O aumento na deposição de gordura e da conversão alimentar durante a fase de terminação são fatores que implicam diretamente na qualidade da carne e também na rentabilidade da cadeia produtiva. Apesar disso, o abate tardio dos suínos pode resultar em benefícios para o frigorífico, pela redução dos custos operacionais por suíno abatido e melhor utilização dos equipamentos, bem como vantagens econômicas para o produtor resultantes da diluição dos custos de produção. Portanto, a suplementação de cromo na ração de suínos de linhagens com maior potencial para deposição de músculo, é uma estratégia para aumentar a deposição de tecido magro e minimizar a deposição de gordura, refletindo positivamente no desempenho. Em vista disso, objetivou-se com este trabalho, apresentar e discutir os mecanismos de ação, finalidades e resultados recentes de estudos na literatura sobre a suplementação desse modificador de carcaça na ração de suínos em terminação. O principal papel do cromo é potencializar a ação da insulina, facilitando a ligação entre a insulina e os seus receptores na membrana celular. Com isso, as células sensíveis à insulina captam maior quantidade de glicose, que posteriormente serão convertidas em energia. Essa energia adicional pode ser utilizada para aumentar a síntese proteica, aumentando assim, a quantidade de carne magra e reduzindo o teor de gordura na carcaça, sem alterar o consumo de proteína pelos animais. O cromo também diminui a taxa de oxidação lipídica, mantendo a qualidade da carne por mais tempo. Em conclusão, a suplementação dietética de Cr para suínos em terminação mostra que 0.2 mg/kg de fontes orgânicas de Cr para suínos de ~60 kg até o abate podem melhorar o desempenho, ganho de massa magra e reduzir o teor de gordura na carcaça. No entanto, o desenvolvimento da nanotecnologia tem permitido o uso de fonte inorgânica de Cr na dose de 0.2 mg/kg de inclusão, melhorando o desempenho e as características de carcaça de suínos em terminação. Palavras-chave: características da carcaça, desempenho, minerais, nutrição, suínos.

INTRODUCTION

Pork is the most consumed meat in the world, reaching a total of 112,433 thousand tons in 2018 (USDA, 2018), highlighting the great importance of pig industry in the world meat market, providing consumers with food of high biological value. Over the years, with advances in breeding programs, genetic strains with less backfat thickness...
and greater potential for muscle deposition have been selected (WOOD & WHITTEMORE, 2006).

In the finishing phase is where the greatest change in the carcass composition occurs. The increase of subcutaneous fat deposition and feed conversion ratio over the days directly affect carcass quality and pig production profitability (VALENTE JÚNIOR et al., 2019).

Heavier pigs result in benefits for the abattoir due to dilution mainly of operational costs per slaughtered pig and better use of equipment, as well as economic advantages resulted from higher carcass yield and commercial cuts (WOOD & WHITTEMORE, 2006).

Conversely, as pigs become heavier, feed efficiency is reduced due to increase in fat deposition, which cause a decrease in growth performance and an increase in production costs, in addition to the risk of not meeting the carcass quality requirements imposed by the abattoir (MARCOLLA et al., 2017). Thus, one of the main challenges of pig industry is to slaughter pigs over 100 kg with higher lean gain and less fat deposition (MADEIRA et al., 2016).

Therefore, dietary supplementation of carcass modifying additives for finishing pigs of high lean-genotypes can contribute to increase lean gain and reduce fat deposition (MARCOLLA et al., 2017). This additional energy is used to improve the use of nutrients by increasing lean gain and reducing fat content in the carcass, without altering the protein intake (WANG et al., 2007; JACKSON et al., 2009; SALES & JANCÍK, 2011). Consequently, this contributes to reduce the environmental impact caused by nitrogen excretion into environment (LINDEMANN, 1999).

It has known that muscle deposition is more efficient compared to adipose tissue deposition, since less dietary energy is required to deposit muscle tissue in comparison with adipose tissue; thereby, dietary Cr supplementation for finishing pigs can improve feed efficiency and reduce feed costs (LINDEMANN & LU, 2019).

In this way, dietary supplementation of chromium for finishing pigs of high lean-genotypes is a strategy to increase lean tissue accretion and minimize fat deposition, reflecting positively on growth performance, reported in some studies (Table 1).

Evaluating Cr and its different sources (Cr propionate, Cr picolinate, Cr yeast and Cr methionine) on growth performance of castrated and female finishing pigs, LINDEMANN et al. (2008) reported that dietary Cr yeast (0.5 mg/kg) improved average daily gain (ADG) compared to the control diet.

LI et al. (2013) evaluated the effects of supplementing diets for finishing barrows (75 to 100 kg) with CrMet (0, 0.3, 0.6, and 0.9 mg/kg) and observed a linear increase in ADG, average daily feed intake (ADFI) and feed conversion ratio (FCR). There was also a linear increase in loin muscle area (LMA) and a decrease in backfat thickness (BF) of pigs fed CrMet diet.

In a study with different sources of dietary fat, JACKSON et al. (2009) reported the inclusion of 0.2 mg/kg Cr propionate to finishing gilts did not improve growth performance. However, dietary supplementation of Cr propionate decreased BF in 10th rib and increased the percentage of muscle in carcass. ZHANG et al. (2011) evaluated the effects of 0, 0.2 and 0.4 mg/kg Cr picolinate (CrPic) in the
Dietary supplementation of chromium for finishing pigs.

During growth phase, no differences were observed on growth performance. However, during the finishing phase, pigs fed 1.61 mg/kg CrPic had better ADG. During the entire period, pigs fed with CrPic in the diet had higher ADG (6.08%) and lower FCR (3.30%). Regarding carcass quality, an increase in muscle:fat ratio and lMA were observed in pigs fed with 1.61 mg/kg CrPic.

Similarly, SILVA et al. (2019) observed an increase in ADG and ADFI, and an improvement in FCR with a dietary supplement composed by L-carnitine (50 mg/kg), Cr propionate (0.2 mg/kg) and organic selenium (0.3 mg/kg) for finishing pigs.

LI et al. (2017) also observed an increase in ADG and ADFI of growing and finishing pigs supplemented with 0.2 mg/kg Cr nanoparticle in the diet. In addition, the pigs that received Cr in the diet had a reduction in BF.

CARAMORI JÚNIOR et al. (2017) reported that dietary supplementation of Cr-yeast (0.8 mg/kg) and Se-yeast (0.6 mg/kg) to barrows from 70 to 130 kg increased Longissimus dorsi depth without affecting pigs’ final body weight. In addition, HUNG et al. (2015) observed that 0.4 mg/kg Cr picolinate in the diet for finishing gilts promoted an increase in carcass weight and loin muscle depth, and a reduction in BF. The studies above evidenced the effects of Cr in reducing fat and increasing protein deposition, which improves the growth performance of finishing pigs.

Cr can also improve meat quality, through decreasing lipid oxidation rate and; consequently, increasing shelf life. AGEs (Advanced Glycation End-products) are complexes formed during glycation between protein and sugar molecules that occurs mainly in high levels of blood glucose. These complexes form free radicals, thus triggering oxidative stress that increases meat lipid oxidation (BIERHAUS et al., 1998; JAKUS & RIE TBROCK, 2004). Cr contributes to decrease the generation of free radicals (PREUSS et al., 1997), increasing glucose uptake by cells and decreasing blood sugar levels (EVANS & BOWMAN, 1992), which reduces the susceptibility to meat lipid oxidation.

In this sense, TIAN et al. (2014; 2015) and XU et al. (2017) observed an improvement in

**Table 1 - Studies assessing the effects of different sources and levels of chromium in the diets for growing and finishing pigs on growth performance and carcass traits.**

| Source                  | Level mg/kg | Animals          | Weight     | Main results                                                                 | Reference   |
|-------------------------|-------------|------------------|------------|------------------------------------------------------------------------------|-------------|
| Cr Nanocomposite        | 0.2         | Finishing barrows| 65 to 90 kg| Reduced FCR. Decreased BF in 10th rib and increased LMA.                      | WANG et al. (2007) |
| Cr Yeast                | 0.5         | Crossbred pigs   | 48 to 114 kg| Improved ADG and FCR.                                                        | LINDEMANN et al. (2008) |
| Cr Propionate           | 0.2         | Finishing gilts  | 29 to 109 kg| Decreased BF in 10th rib and increased the percentage of muscle in carcass.  | JACKSON et al. (2009) |
| Cr Picolinate           | 0, 0.2 and 0.4 | Growing-finishing pigs | 35 to 100 kg | Higher ADG and lower FCR. Increase in muscle:fat ratio and LMA.              | ZHANG et al. (2011) |
| Cr Methionine           | 0, 0.3, 0.6, and 0.9 | Finishing barrows | 75 to 100 kg | Linear increase in ADG, ADFI, FCR and LMA. Decrease in BF.                  | LI et al. (2013) |
| Cr Methionine           | 0.2         | Barrows          | 60 to 107 kg| Reduced 72h post-mortem meat lipid oxidation. Better results of ADG and FCR. | PERES et al. (2014) |
| Cr Nanoparticles        | 0, 0.1, 0.2, and 0.4 | Finishing pigs | 66 to 93 kg | Linear increase in feed efficiency and a linear reduction in carcass fat percentage and BF. | WANG et al. (2014) |
| Cr Picolinate           | 0.4         | Finishing gilts  | 52 to 91 kg | Increase in carcass weight and loin muscle depth, and a reduction in BF.     | HUNG et al. (2015) |
| Cr Nanoparticles        | 0.2         | Growing-finishing pigs | 31 to 120 kg | Increase in ADG and ADFI. Reduction in BF.                                  | LI et al. (2017) |
| Cr Methionine           | 0.2         | Growing-finishing pigs | 32 to 110 kg | Reduced FCR and increased LMA.                                              | XU et al. (2017) |

ADG = average daily gain; ADFI = average daily feed intake; FCR = feed conversion ratio; LMA = loin muscle area and BF = backfat thickness.
antioxidant status of growing and finishing pigs supplemented with Cr methionine up to 0.4 mg/kg in the diet. However, higher inclusion of Cr methionine (0.8 mg/kg) decreased the serum antioxidant capacity (TIAN et al., 2014; 2015).

PERES et al. (2014) observed that dietary supplementation of 0.2 mg/kg CrMet for barrows (60 to 107 kg) reduced the 72h post-mortem meat lipid oxidation, suggesting the inclusion of CrMet in diets for finishing pigs maintain the meat quality for longer period. In addition, CrMet supplementation provided better results of ADG and FCR compared to control diet.

However, there is a need for further investigation to better understand the mechanisms of Cr and its different sources and levels in reducing meat lipid oxidation.

**Chromium on activation of glucose tolerance factor (gtf) and chromodulin**

Higher protein deposition and lower fat deposition in the carcass in response to dietary Cr supplementation may be related to carbohydrate metabolism, acting as part of the mechanism of insulin signaling amplification, improving glucose and amino acids uptake by muscle tissue cells (EVANS & BOWMAN, 1992; VINCENT et al., 2000). In this sense, Chromium may increase the cells glucose uptake by activating the glucose tolerance factor (GTF) and chromodulin.

Cr is a biological active compound of an essential trace that participates in the formation of the molecule GTF. GTF is a low molecular weight organic complex, composed of Cr, nicotinic acid, glycine, cysteine, glutamic acid and calcium, which increases cell membrane fluidity and enhances insulin action, facilitating the binding of insulin with its receptor and increasing the cell’s sensitivity to glucose (MIRSKY et al., 1981; MERTZ, 1993). In this way, in vitro studies indicated that GTF can increase glucose transport in yeast cells (MIRSKY et al., 1981; MIRSKY & BERDICEVSKY, 1994), adipocytes (TOKUDA et al., 1987), and cardiomyocytes (FISCHER et al., 1992).

In addition, MIRSKY et al. (2016) observed that diabetic rats treated with GTF had lower blood glucose levels than the diabetic ones not treated with GTF (252 mL/DL vs 450 mL/DL, respectively), suggesting a positive influence of GTF in hyperglycemia control.

GTF regulates blood glucose levels, potentiating insulin activity. When insulin binds to its receptor on the cell membrane, blood glucose levels are reduced due to glucose uptake by cells. Then, glucose is used as an energy source for protein synthesis, regulated by growth hormone (GH) and insulin-like growth factor (IGF-1) (LINDER, 1991). GTF also increases the conversion of thyroxine (T4) to triiodothyronine (T3), hormones that accelerate metabolic rate and increase protein synthesis capacity by muscle cells (BURTON, 1995). Therefore, Cr is of a great importance since without it GTF molecule is inactive (HOSSAIN et al., 1998).

Cr is an active compound of another molecule called chromodulin, which is considered a biologically active form of Cr (ALMEIDA et al., 2010). Therefore, activation of chromodulin could be another pathway of Cr action on carbohydrate metabolism.

Chromodulin is an oligopeptide composed of Cr\(^{3+}\) and amino acid residues of glycine, cysteine, glutamate and aspartate (YAMamoto et al., 1987), which has the function of potentiating insulin signaling (VINCENT, 2001).

Adapted chromodulin activation model is shown in figure 1. Chromodulin is stored as protein in insulin sensitive cells. Cr is transported in bloodstream mainly by transferrin and when there is an increase in circulating insulin there will be a greater mobilization of chromodulin to tissue cells, and greater mobilization of transferrin receptors from intracellular vesicles to cells membrane. Posteriorly, trasferrin saturated with Cr molecules binds to its specific receptors and the complex formed is internalized by endocytosis. The acidic pH in intravesicular space causes digestion of this complex, releasing Cr to cytosol. Apochromodulin binds to four ions of Cr\(^{3+}\) becoming active as chromodulin form, which in turn binds to cells insulin receptor amplifying the insulin signal (VINCENT, 1994).

As blood glucose levels rise insulin is released quickly into bloodstream. The binding of insulin to its membrane receptors causes a change in the conformation of these receptors that autophosphorylates tyrosine residues transforming the receptor into an active kinase (SALTIEL, 1994).

These changes in insulin receptors promote translocation of glucose transporter type 4 (GLUT 4) to plasma membrane, potentiating the activity of tyrosine kinase, thus increasing membrane sensitivity and improving glucose uptake (VINCENT, 1994; SALTIEL & KAHN, 2001).

In this sense, TIAN et al. (2014) observed a decrease in glucose and insulin serum levels in pigs supplemented with different levels of CrMet (0.1, 0.2, 0.4 or 0.8 mg/kg), indicating greater uptake of glucose by cells.
In a study with dietary supplementation of nanocomposite chromium (0.2 mg/kg) for finishing barrows (65 to 90 kg), WANG et al. (2007) reported an increase of IGF-1 and total protein concentrations in serum, and a reduction in glucose and insulin levels. Indicating the mechanism that Cr exerts on protein synthesis, resulting in increased LMA and reduced body and BF in that study. In addition, the authors also observed a reduction in FCR (3.30%) in pigs fed with the diet supplemented with Cr.

Similar results were reported by WANG et al. (2014), who evaluated the supplementation of Cr nanoparticles (0, 0.1, 0.2, and 0.4 mg/kg) for finishing pigs. The authors reported a linear increase in feed efficiency and a linear reduction in carcass fat percentage and BF. In addition, pigs fed Cr in the diet had an increase in serum IGF-1 concentrations and a reduction in plasma insulin.

Chromium on protection of the cortical actin filament (f-actin)

The structural dynamics of the actin filament (F-actin) is a fundamental characteristic of glucose transport in skeletal muscle cells (BROZINICK JR et al., 2004; MCCARTHY et al., 2006). F-actin is also essential for GLUT4, which plays an important role in glucose uptake by skeletal muscle cells as well as in regulation of blood glucose levels (TSAO et al., 2000). Therefore, compromising the structure of F-actin can decrease glucose uptake by cells, causing metabolic disorders.

HABEGGER et al. (2012) observed in muscle tissues of humans and animals resistant to insulin, a reduction of F-actin due to increase in cholesterol in the skeletal muscle membrane, suggesting the increase in cholesterol deposition in the membrane may compromise the F–actin structure. According to CHEN et al. (2006), Cr\(^{3+}\) can increase GLUT4 activity and glucose transport in 3T3-L1 adipocytes by reducing cholesterol in the cell membrane.

The mechanisms involved in reduction of cholesterol in the cell membrane by Cr are not clear. In vitro and in vivo studies have reported that Cr\(^{3+}\) increases the activity of 5’ adenosine monophosphate-activated protein kinase (AMPK) (WANG et al., 2009;
HOFFMAN et al., 2014). This leads to an increase of fatty acids β-oxidation and to a decrease of muscle tissue lipogenesis, indicating the effects of chromium on protection of F-actin, as well as on carcass traits (MUOIO et al., 1999). For this, phosphorylated AMPK inhibits 3-Hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase, which is one of the main enzymes in endogenous cholesterol synthesis (PEDERSEN, 1994). Thus, the increase in AMPK activity protects the cell membrane against the accumulation of excess cholesterol, reducing the damage on F-actin (HABEGGER et al., 2012).

However, studies evaluating the action of Cr on serum cholesterol levels have shown inconsistent results, ranging from lack of effect (MATTHEWS et al., 2001; BURGOS et al., 2016), increase (SHELTON et al., 2003) or reduction in serum cholesterol (LIEN et al., 2001; PARK et al., 2009) in pigs supplemented with CrPic in the diet. Evaluating the effects of dietary supplementation of 0.2 mg/kg CrPic for finishing gilts, BURGOS et al. (2016) did not observe differences in serum total cholesterol levels. However, these authors reported that cholesterol concentrations in Longissimus thoracis muscle and in adipose tissue were considerably lower in pig supplemented with CrPic compared to those not supplemented.

**Chromium on regulation of gene expression**

In addition to enzymes/proteins already discussed in this review that act positively on glucose uptake by the cells, several other factors are involved in regulation of glucose metabolism. The increase in expression of some genes, such as glycogen synthase (GS) and uncoupling protein-3 (UCP3), may play an important role in physiological function of Cr (QIAO et al., 2009).

GS regulates glycogen synthesis and also plays a role in glucose storage (HUPPERTZ et al., 2001). However, its activation is regulated by insulin, which promotes dephosphorylation and activation of GS by inactivation of glycogen synthase kinase 3 (GSK3) through phosphorylation by Akt (BOUSKILA et al., 2008).

The UCP3 stimulates glucose transport and GLUT4 translocation to the surface of skeletal muscle cells, by activating a phosphoinositide 3 kinase-dependent pathway (HUPPERTZ et al., 2001).

QIAO et al. (2009) reported that Cr positively regulates the mRNA expression of insulin receptor, GLUT4, GS and UCP3 in skeletal muscle cells, which potentialize the uptake and metabolism of insulin-stimulated glucose.

Hormone sensitive lipase (HSL) is considered as a key enzyme in fatty acid mobilization, mainly hydrolyzing triacylglycerol, diacylglycerol, and monoaoylglycerol into free fatty acid (LAFONTAN & LANGIN, 2009). Fatty acid synthetase (FAS) plays a key role in de novo lipogenesis of long-chain fatty acids (LIU et al., 2010). Therefore, higher and lesser activity of HSL and FAS, respectively, in the adipose tissue may reduce the carcass fat content. In this sense, dietary Cr supplementation may improve the carcass lean: fat ratio by upregulate HSL and downregulate FAS activity.

In this way, evaluating the effects of dietary Cr nanoparticles supplementation (0, 0.1, 0.2, and 0.4 mg/kg) for barrows on lipid metabolism, WANG et al. (2014) observed a reduction in the enzyme FAS and an increase in HSL and free fatty acids in adipose tissue, resulting in greater lipid oxidation and less lipogenesis in pigs supplemented with Cr. Therefore, these pigs had less fat and greater muscle deposition in carcass.

**CONCLUSION**

The results of dietary Cr supplementation for finishing pigs have shown that 0.2 mg/kg of organic Cr sources, such Cr Methionine, Cr Picolinate and Cr Yeast for pigs from ~60kg until the slaughter can improve growth performance, lean gain and reduce fat content in carcass. However, the development of nanotechnology has allowed the use of inorganic Cr source at 0.2 mg/kg of inclusion, based on the increase in Cr bioavailability, leading to improve the growth performance and carcass traits of finishing pigs.

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**DECLARATION OF CONFLICT OF INTERESTS**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**AUTHORS’ CONTRIBUTIONS**

The authors contributed equally to the manuscript.

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