ASCL1 and DLL3 expressions and their clinicopathological implications in surgically resected pure small cell lung cancer: A study of 247 cases from the National Cancer Center of China

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Abstract
Objective: Small cell lung cancer (SCLC) is one of the most aggressive malignancies characterized by neuroendocrine (NE) differentiation. The Delta-like protein 3 (DLL3), as a direct downstream target of ASCL1, is involved in NE differentiation and carcinogenesis of SCLC. This study aims to investigate the relationship between ASCL1 and DLL3 expressions and their clinicopathological implications in SCLC.

Methods: A total of 247 surgically resected pure SCLC samples with limited clinical stage and follow-up data were retrieved in this retrospective study. ASCL1 and DLL3 protein expression was detected by immunohistochemistry staining. The correlations between ASCL1 and DLL3 expressions, as well as their clinicopathological features, were analyzed by χ² tests. Disease-free survival (DFS) and overall survival (OS) in SCLC patients with ASCL1/DLL3 low and high expressions were compared by the Kaplan-Meier method and log-rank tests.

Results: ASCL1 high expression was detected in 105 (42.5%) patients. Its expression was positively correlated with the clinical stage (p=0.02) and nerve invasion (p=0.03). DLL3 high expression was observed in 188 (72.8%) patients and was correlated with vascular invasion (p=0.04). ASCL1 expression was positively associated with DLL3 expression (p=0.03). In addition, DLL3 expression has a strong correlation with the expression of thyroid transcription factor-1 (TTF1) and conventional NE markers.

Conclusion: ASCL1 and DLL3 were highly expressed in SCLC tumor samples, and a positive correlation between these two markers was observed. Co-analysis of ASCL1 and DLL3 may identify a distinct SCLC subgroup benefit from targeted therapy. Therefore, ASCL1 and DLL3 could be potential biomarkers served for the selection of related patients.

KEYWORDS
ASCL1, DLL3, SCLC small cell lung cancer

Chunfang Hu and Jiyan Dong contributed equally to this research.
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Small cell lung cancer (SCLC) is one of the most aggressive malignancies with neuroendocrine (NE) differentiation and poor prognosis, accounting for 15% of all lung cancers. For decades, traditional chemotherapy and localized radiotherapy have been the predominant therapies for SCLC. Although SCLC is sensitive to the initial chemotherapy and radiotherapy, it has a high rate of relapse and resistance to traditional drugs. Therefore, seeking other forms of treatment modalities is desperately necessary.

Emerging data revealed that SCLC was a considerable heterogeneous tumor in terms of histomorphology, molecular changes, and growth features. Several recent studies applying SCLC primary human tumors, xenografts, and cell lines have attempted to divide SCLC into distinct molecular subtypes, including ASCL1-high, NEUROD1-high, POU2F3, and YAP1 subtypes. The most well-established subgroup is the ASCL1-high group, representing the predominant subgroup of SCLC with high expression of NE markers. ASCL1 is expressed in most SCLC tumors and supposed to be closely correlated with NE differentiation.

Notch-ASCL1 signaling pathway plays an important role in the maintenance of NE phenotype and carcinogenesis of SCLC. DLL3 is an inhibitory ligand of the Notch pathway and results in the inactivation of Notch pathway. As a regulator, ASCL1 can enhance the expression of its downstream target Delta-like protein 3 (DLL3). Recently, DLL3-targeted antibody-drugs, rovalpituzumab tesirine (Rova-T), bispecific antibody, and chimeric antigen receptor-modified T cells (CAR-T) have been used in several clinical trials, demonstrating a potential prospect in SCLC treatment. Combined investigation of ASCL1 and DLL3 expression could be helpful for a selection of subgroup of patients who would benefit from these targeted therapies.

Analysis of both ASCL1 and DLL3 in SCLC patient samples has been only reported in one recent study, which investigated 95 surgically resected SCLC samples, including combined SCLC. However, our recently published studies revealed distinct prognostic impact factors and YAP1 protein expression between pure SCLC and combined SCLC. This suggests that combined SCLC and pure SCLC might be different entities and present different targetable oncogenic pathways. Therefore, in this study, we enrolled a large scale of 247 surgically resected pure SCLC samples (without combined non-small cell lung cancer component). All the samples were pure SCLC cases with limited clinical stages, and we aimed to clarify the relationship between ASCL1 and DLL3 expressions and their clinicopathological implications for SCLC patients.

METHODS

Sample selection

The study was approved by the ethics committee and institutional review board of Cancer Hospital, Chinese Academy of Medical Sciences (CHCAMS), and all patients were exempt from informed consent. A total of 247 surgically resected pure SCLC samples were retrieved in this retrospective study. They were primarily diagnosed in CHCAMS during the period of January 2005 and December 2016. Sample inclusion criteria were used as described in our previous study: (1) surgically radical excision specimen of SCLC with or without systemic lymph node dissection; (2) histologically confirmed to be pure SCLC without any combined component; and (3) clinically proven to be pulmonary primary tumor when metastasis was excluded. Tumors were staged and accessed according to the seventh edition of the American Joint Committee on Cancer (AJCC). General clinical information and follow-up data were obtained from the medical records system.

Reassessment of clinicopathological characteristics

All archival slides of hematoxylin and eosin (H&E) and immunohistochemical (IHC) staining from the 247 SCLC samples were retrieved and reevaluated by two senior thoracic pathologists. The diagnosis of pure SCLC was confirmed by pathological morphology and staining of several conventional NE biomarkers. Clinicopathological characteristics were also reevaluated and recorded, including invasion of bronchus, vessels, nerves and pleura, proportion of necrosis and fibrosis, regional lymph node metastasis, spread through air spaces (STAS), and tumor infiltrating lymphocytes (TILs).

IHC staining and scoring criteria

IHC staining for conventional NE markers including synaptophysin (SYN, SP11, MXB Biotechnologies), chromogranin A (CgA, LK2H10+PHE5, MXB Biotechnologies), CD56 (MX039, MXB Biotechnologies), and TTF1 (SP141, Roche) was performed on formalin-fixed paraffin-embedded (FFPE) tissues during routine clinical practices. ASCL1 (ab74065, Abcam) and DLL3 (E3j5R, Cell Signal Technology) staining was examined on SCLC tissue microarrays. The SCLC tissue microarrays of the 247 cases were constructed as described previously. All the staining was carried out on the fully automatic Roche IHC instruments (Roche Diagnosis) according to the manufacturer’s protocols.

ASCL1 expression was localized in the nucleus of SCLC tumor cells, whereas DLL3 expression was localized in the cytoplasm and membrane of tumor cells. For staining pattern of conventional NE markers, cytoplasm for SYN and CgA, membrane for CD56, and nucleus for transcription termination factor 1 (TTF1) in tumor cells were evaluated to be positive. Both the intensity and proportion of positive tumor cells were taken into account and the scoring criteria were used as described in the previous study. Percentages of stained tumor cells (0%–100%) and four-level intensity of
| Characteristics                  | Patients (N = 247) | ASCL1 expression | DLL3 expression |  
|---------------------------------|-------------------|------------------|-----------------|  
|                                 |                   | Low (%)          | High (%)        | p value | Low (%)           | High (%) | p value |  
| Gender                          |                   |                  |                 |         |                   |          |         |  
| Male                            | 175               | 101(71)          | 74(70)          | 1.00    | 44(75)            | 131(70)  | 0.52    |  
| Female                          | 72                | 41(29)           | 31(30)          | 1.00    | 15(25)            | 57(30)   | 1.00    |  
| Age                             |                   |                  |                 |         |                   |          |         |  
| ≤65                             | 202               | 115(81)          | 87(83)          | 0.74    | 48(81)            | 154(82)  | 1.00    |  
| >65                             | 45                | 27(19)           | 18(17)          | 1.00    | 11(19)            | 34(18)   | 1.00    |  
| Smoking                         |                   |                  |                 |         |                   |          |         |  
| Yes                             | 158               | 90(63)           | 68(65)          | 0.89    | 41(69)            | 117(62)  | 0.35    |  
| No                              | 89                | 52(37)           | 37(35)          | 0.37    | 18(31)            | 71(38)   | 0.37    |  
| Tumor location                  |                   |                  |                 |         |                   |          |         |  
| Left lung                       | 122               | 74(52)           | 48(46)          | 0.37    | 35(59)            | 87(46)   | 0.01    |  
| Right lung                      | 125               | 68(48)           | 57(54)          | 1.00    | 24(41)            | 101(54)  | 1.00    |  
| Clinical stage                  |                   |                  |                 |         |                   |          |         |  
| I                               | 78                | 47(33)           | 31(30)          | 0.02*   | 21(36)            | 57(30)   | 0.28    |  
| II                              | 68                | 47(33)           | 21(20)          | 0.74    | 19(32)            | 49(26)   | 0.52    |  
| III                             | 101               | 48(34)           | 53(50)          | 0.10    | 19(32)            | 82(44)   | 0.10    |  
| Vascular tumor thrombus         |                   |                  |                 |         |                   |          |         |  
| Yes                             | 125               | 76(54)           | 49(47)          | 0.31    | 35(59)            | 87(46)   | 0.10    |  
| No                              | 122               | 66(46)           | 56(33)          | 0.31    | 24(41)            | 101(54)  | 0.10    |  
| Pleural invasion                |                   |                  |                 |         |                   |          |         |  
| Yes                             | 170               | 91(64)           | 79(75)          | 0.07    | 42(71)            | 128(68)  | 0.75    |  
| No                              | 77                | 51(36)           | 26(25)          | 0.07    | 17(29)            | 60(32)   | 0.07    |  
| Lymph node metastasis           |                   |                  |                 |         |                   |          |         |  
| Yes                             | 143               | 75(53)           | 68(65)          | 0.07    | 28(47)            | 115(61)  | 0.07    |  
| No                              | 104               | 67(47)           | 37(35)          | 0.07    | 31(53)            | 73(39)   | 0.07    |  
| Bronchus invasion               |                   |                  |                 |         |                   |          |         |  
| Yes                             | 210               | 119(84)          | 91(87)          | 0.59    | 50(85)            | 160(85)  | 1.00    |  
| No                              | 37                | 23(16)           | 14(13)          | 0.59    | 9(15)             | 28(15)   | 0.59    |  
| Vascular invasion               |                   |                  |                 |         |                   |          |         |  
| Yes                             | 206               | 117(82)          | 89(85)          | 0.73    | 44(75)            | 162(86)  | 0.045*  |  
| No                              | 41                | 25(18)           | 16(15)          | 0.73    | 15(25)            | 26(14)   | 0.73    |  
| Nerve invasion                  |                   |                  |                 |         |                   |          |         |  
| Yes                             | 84                | 40(28)           | 44(42)          | 0.03*   | 18(31)            | 66(35)   | 0.64    |  
| No                              | 163               | 102(72)          | 61(58)          | 0.03*   | 41(69)            | 122(65)  | 0.64    |  
| STAS                            |                   |                  |                 |         |                   |          |         |  
| Yes                             | 182               | 104(73)          | 78(74)          | 0.89    | 40(68)            | 142(76)  | 0.24    |  
| No                              | 65                | 38(27)           | 27(26)          | 0.89    | 19(32)            | 46(24)   | 0.89    |  
| Necrosis proportion             |                   |                  |                 |         |                   |          |         |  
| ≤30%                            | 172               | 97(68)           | 75(71)          | 0.68    | 37(63)            | 135(72)  | 0.20    |  
| >30%                            | 75                | 45(32)           | 30(29)          | 0.68    | 22(37)            | 53(28)   | 0.68    |  
| Fibrosis proportion             |                   |                  |                 |         |                   |          |         |  
| ≤10%                            | 116               | 75(53)           | 41(39)          | 0.04*   | 30(51)            | 86(46)   | 0.55    |  
| >10%                            | 131               | 67(47)           | 64(61)          | 0.04*   | 29(49)            | 102(54)  | 0.55    |  
| TILs                            |                   |                  |                 |         |                   |          |         |  
| ≤30%                            | 212               | 124(87)          | 88(84)          | 0.46    | 49(83)            | 163(87)  | 0.52    |  
| >30%                            | 35                | 18(13)           | 17(16)          | 0.46    | 10(17)            | 25(13)   | 0.46    |  

*p value <0.05.
staining (0 = negative, 1 = weak, 2 = moderate, and 3 = strong) were recorded, respectively. A histoscore (H-Score) with a range of 0–300 was produced by multiplying the score of percentage (0–100) by the intensity of stained tumor cells (0, 1, 2, and 3). The expression level was further classified as high and low according to the best cut-off value determined by X-tile software.

Follow-up data

Overall survival (OS) was calculated during the time from the first diagnosis to death regardless of any cause. Disease-free survival (DFS) was defined during the time from start of treatment to documentation of any progression such as recurrence or metastasis. Follow-up was completed in February 2019. A total of 212 patients had complete follow-up data, and 35 patients (14.2%) were lost to follow-up.

Statistical analysis

The correlation between ASCL1 and DLL3 protein expressions, their expression and clinicopathological features as well as conventional NE scores were analyzed by $\chi^2$ tests and Fisher’s exact tests. OS and DFS in SCLC patients with ASCL1/DLL3 low and high expressions were compared by using the Kaplan-Meier method and log-rank test. Univariate and multivariate analysis was carried out in the Cox proportional hazards model. All differences were considered as

![Image](image.png)

**FIGURE 1** Representative slides of SCLC morphology and ASCL1/DLL3 expression level ($\times$ 200). (a) and (b) SCLC H&E staining and the corresponding ASCL1 high expression, respectively, (c) ASCL1 low expression; (d) and (e) SCLC H&E staining and the corresponding DLL3 high expression, respectively, (f) DLL3 low expression.
statistically significant if \( p \) value <0.05. Statistical analysis was performed using SPSS version 23.0 (SPSS).

**RESULTS**

**Patient and sample characteristics**

Among the entire cohort of the 247 surgically resected pure SCLC patients, 175 (70.9%) were male, 202 (81.8%) were <65 years, and 158 (64.0%) had a history of smoking. According to the 7th edition of AJCC Cancer Staging Manual, 78 (31.6%) were stage I, 68 (27.5%) were stage II, and 101 (40.9%) were stage III. A total of 143 (57.9%) patients showed regional lymph node metastasis (Table 1).

**Expression of ASCL1/DLL3 and correlation with clinicopathological features**

ASCL1 expression was localized in the nucleus of SCLC tumor cells (Figure 1(a)–(c)), and 105 (42.5%) patients showed high expression. ASCL1 high expression was associated with clinical stage (\( p = 0.02 \) and nerve invasion (\( p = 0.03 \)). DLL3 expression was localized in the cytoplasm and membrane of tumor cells (Figure 1(d)–(f)), and DLL3 high expression was observed in 188 (72.8%) patients. DLL3 high expression was correlated with vascular invasion (\( p = 0.04 \)). No significant difference was found in ASCL1/DLL3 expression and the remaining clinicopathological features. Co-expression of ASCL1/DLL3 was observed in 124 (50.2%) patients, and 87 of which showed co-high expression of these two markers. Subsequently, we analyzed the relationship between ASCL1 and DLL3 expression in SCLC, and a significantly positive association was observed (\( p = 0.035 \), Table 2).

**ASCL1/DLL3 expression and correlation with conventional NE markers**

Because ASCL1/DLL3 was highly expressed in SCLC tumor cells, we next examined the correlation between ASCL1/DLL3 expression and conventional NE markers (Syn, CgA, CD56, and TTF1). According to the H-Score of these markers, their expressions were divided into low and high expression groups. Expression levels of DLL3 had a strongly positive association with those of Syn (\( p = 0.01 \), CgA (\( p = 0.02 \), CD56 (\( p < 0.01 \), and TTF1 (\( p < 0.01 \), as shown in Table 3. However, expression of ASCL1 was only positively correlated with CgA (\( p = 0.031 \), but not correlated with Syn, CD56, and TTF1 (\( p > 0.05 \)).

**ASCL1/DLL3 expression and patients’ outcome**

All the 247 SCLC patients were followed up, with a median follow-up time of 48 months ranging from 0 to 167 months. Univariate analysis revealed that patients with ASCL1 high expression had a worse OS (\( p = 0.047 \), whereas ASCL1 expression was not associated with DFS (\( p > 0.05 \). No significant difference was found in DLL3 expression level and patients OS/DFS (\( p > 0.05 \)). Further multivariate cox regression analysis showed neither ASCL1 nor DLL3 expression was an independent prognostic factor (\( p > 0.05 \)).

**DISCUSSION**

SCLC is a highly aggressive cancer with poor prognosis. Although most patients are sensitive to the initial chemotherapy and radiotherapy, frequent relapse and resistance to traditional drugs will eventually result in treatment failure. Therapeutic options for recurrent patients were limited, and only a few survived more than 5 years from diagnosis.
Current advances in molecular biology established by using xenografts and cell lines proposed the concept of RNA-based molecular subtyping, which had great significance in exploration of therapeutic targets for distinct subgroups. Our study observed protein expressions of ASCL1 (an essential biomarker for SCLC-A subgroup) and DLL3 (a ROVA-T drug target), and analyzed their correlations with clinicopathological features, in an attempt to identify potential patients in SCLC-A subgroup who would benefit from ROVA-T treatment. In the current study, ASCL1 and DLL3 were found to be highly expressed in a large cohort of surgically resected SCLC samples ($n=247$), and a significantly positive correlation between these two markers were identified.

ASCL1 is one of the master transcription factors in SCLC and plays an important role in cell fate decisions during neurogenesis. Our previous study revealed that ASCL1 protein expression was strongly associated with ASCL1 mRNA expression and NE differentiation score, and ASCL1-high patients had a lower OS, suggesting that examination of ASCL1 expression by IHC staining could be an alternative option for SCLC subgrouping. In this study, ASCL1 expression was scored according to H-Score criteria, and then subdivided into ASCL1 low and high expression groups. ASCL1 was highly expressed in 42.5% (105/247) of SCLC. Compared to previous studies, it was consistent with one study as 64% of SCLC expressing ASCL1 in at least 5% tumor cells, whereas a bit lower than the other study as 80% of cases showing ASCL1 expression. As shown in the study by Baine et al., 60% cases were small biopsies and 80% of cases showing ASCL1 expression. Despite unfavorable results in some clinical studies, DLL3 remains a remarkable target in SCLC because of its high and homogeneous expression on the cell surface. DLL3 targeted therapy is expected to provide great advances in treatment of this aggressive tumor. Further studies are urgently needed to explore the molecular mechanism, to assess expression threshold for DLL3-high patients that benefit from targeted therapy, and to focus on selection of potential molecular markers for predicting treatment response.

Our previous work already showed ASCL1 was a NE related marker. As an inhibitory ligand of Notching pathway, DLL3 also plays an essential role in NE transformation and maintenance. We analyzed the relationship between DLL3 expression and several conventional NE markers, such as SYN, CgA, CD56, and TTF1 in this study. These markers were normally used for differential diagnosis of SCLC in routine clinical practices. We found a strong positive association between DLL3 and SYN, CgA, CD56, and TTF1 expressions, suggesting the NE function of DLL3. Our observation was consistent with other investigations. SYN, CgA, and CD56 have been classic markers for SCLC diagnosis, whereas DLL3 showed much wider coverage of expression in SCLC than SYN and CgA. This suggests DLL3 could be a candidate NE marker for SCLC diagnosis to remedy the deficiency of present markers. Moreover, considering the strong correction between DLL3 and TTF1 expression, several recent studies suggested that TTF1 expression could be a potential surrogate of DLL3 expression to identify patients who may respond to DLL3 targeted therapy.

Finally, we analyzed the impact of ASCL1 and DLL3 expression on prognosis of SCLC patients. Although univariate analysis revealed that patients with ASCL1 high expression had a worse OS, multivariate cox regression analysis showed neither ASCL1 nor DLL3 expression was an independent prognostic factor. These results were also supported by several other findings. ASCL1 and DLL3
expressions have limited impact on patient outcomes and may not be valuable indicators for prognosis prediction. Because the molecular mechanisms of SCLC are complicated, multiple signaling pathways are involved in the tumorigenicity and progression, including Hippo/Notch/EMT pathways. Therefore, signal indicator cannot predict prognosis, and analysis of ASCL1/DLL3 expression could be more significant in selection of patients suitable for target therapy. As mentioned above, ASCL1 positive phenotype was the most conventional and predominant subgroup of SCLC tumor. To some extent, DLL3 is elevated as a consequence of ASCL1 overexpression, and activities of DLL3 targeted therapy can be the highest in ASCL1-high and DLL3-high group tumors based on preferential expression of these markers in this subtype.

Our study has some advantages. First, we observed high expression levels of ASCL1/DLL3 and their positive correlations in the largest surgically resected samples. Second, co-analysis of ASCL1/DLL3 expression using routine IHC staining provides evidence for selection of potential subgroups, which is more applicable for advanced patients (accounting for 80% SCLC patients in clinical practice) because of insufficient sample amount for further examination. Third, unlike the study by Furuta et al., we enrolled pure SCLC samples for study (that is more representative of authentic SCLC features compared to combined SCLC) because both of our previous works based on clinicopathological analysis and YAP1 expression revealed the distinction between pure SCLC and combined SCLC, and the latter presented much more heterogeneity. Although the cell origin of combined SCLC remains unclear, SCLC components and non-SCLC components in combined SCLC have been reported to share almost 75% common mutations and show similar genetic background, suggesting that the SCLC components and non-SCLC components are derived from common precursors. Moreover, this also implies that one component of combined SCLC arises from the other with subsequent acquisition of oncogenic change and micro-environment in combined SCLC. In Furuta et al.’s study, 29 combined SCLC and 66 pure SCLC were included. Although DLL3 and ASCL1 scoring was only assessed in components of SCLC cells, but not in the components of histologically characterized as squamous cell carcinoma, adenocarcinoma, or large cell carcinoma, the SCLC components in combined SCLC is relatively different from that in pure SCLC. Combined with our previous studies, we believe that the combined SCLC as a distinct subtype should be considered as an important histological factor significantly affecting the efficacy and prognosis. Our research, with a much larger sample size, further confirmed the conclusion of the study by Furuta et al. that the expression of ASCL1/DLL3 has a significant positive correlation, and DLL3 was not an independent prognostic factor and may only benefit from targeted therapy.

However, this study also has some limitations. First, we only analyzed the correlation between ASCL1/DLL3 expressions and their clinical implications and no further study was carried out on exploration of the underlying molecular mechanisms. Second, all the clinical specimen used were retrospectively surgically resected samples, DLL3 targeted therapeutic effect cannot be straightly evaluated based on subgroups. Further studies are still needed to clarify the molecular mechanisms and to predict response of target therapy.

CONCLUSION

In this study, we analyzed ASCL1 and DLL3 expression in a large scale of surgically resected pure SCLC samples. ASCL1 and DLL3 were highly expressed in SCLC tumor cells, and their expression level was positively correlated. The patients with ASCL1 high expression level represent the dominant subgroup of SCLC with NE characteristics, and DLL3 high patients indicate a subgroup suitable for DLL3 targeted therapy. Therefore, we suppose that co-analysis of ASCL1 and DLL3 expression may contribute to selection of distinct SCLC subgroup that benefit most from DLL3 targeted therapy.

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CONFLICT OF INTERESTS

The authors declare that there are no conflicts of interest.

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