Using the otolith sulcus to aid in prey identification and improve estimates of prey size in diet studies of a piscivorous predator

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**Abstract**

Diet studies are fundamental for understanding trophic connections in marine ecosystems. In the southeastern US, the common bottlenose dolphin *Tursiops truncatus* is the predominant marine mammal in coastal waters, but its role as a top predator has received little attention. Diet studies of piscivorous predators, like bottlenose dolphins, start with assessing prey otoliths recovered from stomachs or feces, but digestive erosion hampers species identification and underestimates fish weight (FW). To compensate, FW is often estimated from the least affected otoliths and scaled to other otoliths, which also introduces bias. The sulcus, an otolith surface feature, has a species-specific shape of its ostium and caudal extents, which is within the otolith edge for some species. We explored whether the sulcus could improve species identification and estimation of prey size using a case study of four sciaenid species targeted by fisheries and bottlenose dolphins in North Carolina. Methods were assessed first on otoliths from a reference collection (\(n = 421\)) and applied to prey otoliths (\(n = 5,308\)) recovered from 120 stomachs of dead stranded dolphins. We demonstrated in reference-collection otoliths that cauda to sulcus length (CL:SL) could discriminate between spotted seatrout (*Cynoscion nebulosus*) and weakfish (*Cynoscion regalis*) (classification accuracy = 0.98). This method confirmed for the first time predation of spotted seatrout by bottlenose dolphins in North Carolina. Using predictive models developed from reference-collection otoliths, we provided evidence that digestion affects otolith length more than sulcus or cauda length, making the latter better predictors. Lastly, we explored scenarios of calculating total consumed biomass across degrees of digestion. A suggested approach was for the least digested otoliths to be scaled to other otoliths iteratively from within the same stomach, month, or season as samples allow. Using the otolith sulcus helped overcome challenges of species identification and fish size estimation, indicating their potential use in other diet studies.

**KEYWORDS**

diet, digestive erosion, estimated prey size, otolith sulcus, prey identification, *Tursiops truncatus*
1 | INTRODUCTION

Understanding trophic connections in marine ecosystems is integral to ecosystem-based fisheries management (Bowen, 1997; Kenney, Scott, Thompson, & Winn, 1997; Morissette, Kaschner, & Gerber, 2010; Pikitich et al., 2004; Sissenwine & Mace, 2003; Smith, Link, Cadrin, & Palka, 2015; Spitz, Ridoux, Trites, Laran, & Authier, 2018). Piscivorous marine mammals, with their large metabolic energy demands (Williams, Haun, Davis, Fuiman, & Kohin, 2001), are significant consumers of many fish species (Overholtz & Link, 2006; Santos, Saavedra, & Pierce, 2014). Along most of the US Atlantic coast (New Jersey to Florida) and the entire Gulf of Mexico coast, the coastal form of the common bottlenose dolphin *Tursiops truncatus* (hereafter, bottlenose dolphin, or dolphin) (Figure 1) is the predominant marine mammal in estuarine and nearshore coastal waters (Hayes et al., 2018; Hoelzel et al., 1998; Mead & Potter, 1995). Studies on bottlenose dolphin diet indicate that their primary prey are often valuable recreational and commercial fish species (Gannon & Waples, 2004; Pate & McFee, 2012); however, their role as a top predator has received little attention in terms of the effects of their predation on ecosystem dynamics.

For bottlenose dolphins, like many other cetaceans, diet studies often rely on the recovery and identification of hard parts such as fish otoliths (usually the sagitta pair) and cephalopod beaks from stomachs to identify prey and estimate prey size (length and weight) (Fitch & Brownell, 1968; Jobling & Breiby, 1986; Pierce & Boyle, 1991). Otoliths are susceptible to digestive erosion, which can hamper species identification, particularly for closely related species (see Pierce & Boyle, 1991). For bottlenose dolphins along the US Atlantic and Gulf coasts, species in the family Sciaenidae comprise a significant portion of the diet, including at least four species in the genus *Cynoscion* (Barros, 1993; Barros & Odell, 1990; Barros & Wells, 1998; Bowen, 2011; Gannon & Waples, 2004; Pate & McFee, 2012). Otoliths of weakfish (*Cynoscion regalis*) and spotted seatrout (*Cynoscion nebulosus*) look similar. Both species can have crenulated nodules on the distal side of sagittal otoliths (Chao, 1978; Mohsin, 1981), but the nodules have been proposed as more common and pronounced in spotted seatrout than weakfish (Simons, 1986). Nodules may not be a strong predictor of species because of variability in nodules among individuals and age (Simons, 1986). As a result, discriminating between the two species in diet studies often relies on differences in the shape of their sulcus, a surface feature on the proximal side of otoliths that serves as the attachment point of the sensory epithelium (Platt & Popper, 1981). The sulcus in weakfish is slightly longer and narrower compared to spotted seatrout (N. B. Barros pers. comm. ~ June 2009); however, the difference is subtle, making misidentification possible. Misidentification could explain reports of weakfish as dolphin prey outside of its known range (Bowen, 2011; Mercer, 1989) or the lack of reports of spotted seatrout as dolphin prey where both *Cynoscion* species overlap (Gannon & Waples, 2004; Robins & Ray, 1986).

Although the predictive relationship between otolith size and fish size allows for the estimation of original prey size, digestive erosion reduces otolith size resulting in underestimates of fish size, and complete otolith digestion biases the number and species composition of recovered otoliths (Bowen & Iverson, 2013; Harvey, 1989; Pierce & Boyle, 1991). This erosion has cumulative effects on estimates of overall consumption biomass of important prey species for a given predator. As a result, there have been efforts to compensate for the erosion and reduce bias in estimates of prey size. Correction factors have been developed for a variety of prey from many pinniped species in experimental studies (Bowen & Iverson, 2013; Harvey, 1989; Orr & Harvey, 2001; Tollit et al., 1997). The variability in correction factors even for a single predator and prey species limits the application of correction factors across species (Bowen & Iverson, 2013). Additionally, pinniped diet studies typically use hard parts recovered from feces not stomachs, further limiting the applicability of experimentally derived correction factors to cetaceans. Similar experimental studies have not been performed on cetaceans (Bowen & Iverson, 2013). In vitro experiments mimicking digestion in marine mammals have been performed on otoliths to estimate erosion rates (Christiansen, Ganst Moen, Hansen, & Nilsen, 2005; Wijnsma, Pierce, & Santos, 1999), but their application is not straightforward (Andreassen et al., 2017).

In the absence of correction factors, past studies have simply not accounted for digestion erosion (MacLeod, Santos, Lopez, & Pierce, 2006; Santos, Fernandez, López, Martinez, & Pierce, 2007; Santos et al., 2004, 2014; Spitz, Rousseau, & Ridoux, 2006) or have minimized erosion bias by sorting otoliths by the degree of digestion and using the least eroded to estimate fish size (Börjesson, Berggren, & Ganning, 2003; Gannon & Waples, 2004; Recchia & Read, 1989). For the latter, the mean estimated weight of a given prey species from the least eroded otoliths within or across all stomach samples is then applied to the number of otoliths with moderate to severe erosion to estimate overall consumption. This method assumes that original prey sizes are similar between the least and most eroded otoliths. This assumption may not be valid as

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**FIGURE 1** Common bottlenose dolphin (*Tursiops truncatus*) surfacing for air in North Carolina. Photograph taken by staff at the National Marine Fisheries Service, Southeast Fisheries Science Center, Beaufort, NC, under Marine Mammal Protection Act Scientific Research Permit 779-1633-00
highly eroded otoliths may consist of originally larger prey whose
otoliths require longer to digest (Jobling & Breiby, 1986; Johnstone,
Harris, Wanless, & Graves, 1990; Tollit et al., 1997; Wijnsma et al.,
1999). Also, otoliths in the same stomach with differing degrees
of digestion may represent more than one feeding event (Shippee,
2014) or a single feeding event on schools of differently sized fish
(Bowen & Iverson, 2013; DeBlois & Rose, 1996; Tollit et al., 1997;
Tollit et al., 2004). Furthermore, the assumption that original prey
sizes are similar between the least and most eroded otoliths across
samples may not be valid due to variability among individual ani-
mals and among sizes for a given prey (Tollit et al., 1997), and to
seasonal growth patterns in fish that may influence the size and
availability of consumed prey (Axenrot & Hansson, 2004; Claridge,
Potter, & Hardisty, 1986). As a result, excluding the more eroded
otoliths can introduce bias.

In North Carolina (NC), the sciaenids weakfish, Atlantic croaker
Micropogonias undulatus (hereafter “croaker”), and spot Leisostomus
xanthurus were found to be the three most important prey species
for the coastal form of bottlenose dolphins (Gannon & Waples,
2004). These species have important commercial and recreational fisheries in
the state (NC Division of Marine Fisheries [NCDMF], 2019). Recently,
there has been interest from fisheries managers on the drivers and lev-
els of natural mortality due to predation on the weakfish stock along
the US Atlantic, which has been decreasing despite decades of harvest
restrictions (Atlantic States Marine Fisheries Commission [ASMFC],
2016). To that end, the current study considered ways to ensure ac-
curate discrimination of weakfish and spotted seaturt otoliths recov-
ered from stranded bottlenose dolphins and to improve estimates of
prey size.

The sulcus showed promise as a way to aid in species identifi-
cation and limit the effects of erosion bias. Not only has the sul-
cus been used to predict fish size (Aguirre, 2003), the shape of its
ostium and cauda sections has been used to differentiate species
and stocks (Chao, 1978; Torres, Lombarte, & Morales-Nin, 2000;
Tuset, Lombarte, & Assis, 2008). Therefore, we explored whether
the sulcus could improve diet studies using a case study of bottlene-
bose dolphins and their prey croaker, spot, and weakfish, and the
conger to weakfish, the spotted seaturt. For these species, the peri-
meter of the sulcus is within the otolith edge perhaps mak-
ing its length less affected by digestion, thereby reducing bias of
estimated prey size (Wijnsma et al., 1999). Using pristine otoliths
from a reference collection and otoliths recovered from stomachs
of dead stranded bottlenose dolphins, we had four objectives: (a)
determine if a quantitative analysis of sulcus shape can distinguish
between spotted seaturt and weakfish; (b) develop model equa-
tions to predict fish length and weight from otolith, sulcus, and
cauda lengths for spotted seaturt, weakfish, croaker, and spot; (c)
assess if digestive erosion affects lengths of the otolith, sulcus, and
cauda differently; and (d) examine differences in estimated prey
consumption biomass for a given prey across three scenarios that
exclude or include measurements of otoliths with moderate to se-
vere erosion.

## 2 | METHODS

### 2.1 | Sample collection and processing

Sagittal otoliths were obtained from two sources: a reference col-
lection (hereafter “reference-collection otoliths”) and stomachs of
bottlenose dolphins (hereafter “prey otoliths”). Reference-collection
otoliths were accessed from a collection of otoliths and squid beaks
archived at the National Oceanic and Atmospheric Administration
laboratory in Beaufort, NC. The reference collection comprises pris-
tine otoliths removed from fresh or previously frozen fish of species
identified using reference guides (Kells & Carpenter, 2011; Robins &
Ray, 1986). Reference specimens were collected in a variety of gears
(gillnets, hook and line, cast nets, and trawls) across multiple seasons
and years primarily in NC estuarine and state coastal waters (<5.6 km
from shore). The collection has an accompanying dataset that includes
lengths and weights for most specimens. Additional weakfish speci-
mens were loaned by the NCDMF and NC State University (NCSU)
to improve sample size. The left and right otolith for each fish were
not always available because one side may have broken during ex-
traction or, for samples collected by the NCDMF and NCSU, the left
otolith was sectioned and used for aging. Otoliths used for measure-
ments, therefore, were primarily right otoliths unless only the left
otolith was available. We assumed measurements between sides
did not differ because asymmetry in otolith length between left and
right otoliths of round fishes has been demonstrated to be negli-
ble (Lychakov, Rebane, Lombarte, Fuiman, & Takabayashi, 2006; Mille,
Mahe, Villanueva, Pontual, & Ernande, 2015).

Prey otoliths were recovered from stomach dissections of 213
dead bottlenose dolphins found stranded in NC between 1996 and
2012. For each dolphin, date of stranding and length measurements
was recorded. Date was used to assign dolphins to season: Winter
(December–February), Spring (March–May), Summer (June–August),
and Fall (September–November). Length was used to assign dol-
phins to age-class categories: Young-of-year (YOY, <184 cm), older
calves (184–211 cm), subadults (212–240 cm), and adults (>240 cm)
(Byrd & Hohn, 2017; Fernandez & Hohn, 1998; Read, Wells, Hohn,
& Scott, 1993). Extraction of the digestive tract (esophagus to du-
odenal ampulla) and contents followed established methods (see
Gannon & Waples, 2004); however, contents from most strandings
were kept separate among digestive compartments (i.e., esophagus,
forestomach, main stomach, pyloric stomach, duodenal ampulla).
We restricted this study to contents removed from the forestom-
ach and esophagus (Recchia & Read, 1989) because contents from
the esophagus are likely regurgitated from the forestomach during
death (Harrison, Johnson, & Young, 1970; Pierce & Boyle, 1991),
and contents from the forestomach represent the most recent feeding event(s). Also, the continuation of digestion in the main
and pyloric stomachs would disproportionately affect otoliths com-
pared to squid beaks and increase the chances that small otoliths
would be completely digested (Bowen & Iverson, 2013). This study
was intended to include only the coastal form or “morphotype” of
bottlenose dolphins, which is genetically distinct from (Hoelzel et al., 1998, Rosel, Hansen, & Hohn, 2009) and has different diet than (Mead & Potter, 1995) the offshore morphotype. As such, samples were excluded from bottlenose dolphins genetically determined to be the offshore morphotype (see Byrd et al., 2014) or thought to be offshore morphotype by the presence of parasites (e.g., *Monorhgamma* and *Phyllolothrium*) not found in the coastal morphotype (Mead & Potter, 1995).

Prey otoliths were identified by comparison to the reference collection and published otolith guides (Baremore & Bethea, 2010; Campana, 2004). To enumerate prey for each dolphin, the left and right otolith of each prey species was counted. The side with the highest count was used to estimate the number of prey consumed for that species (Orr & Harvey, 2001). Afterward, otoliths from that side were graded according to digestive erosion from grade 0 to 5 (adapted from Recchia & Read, 1989; Tollit et al., 1997). Otoliths with no to slight erosion were graded separately: 0, undamaged; 1, barely degraded, sulcus fully visible; and 2, slightly degraded, somewhat chalky appearance, sulcus fully visible (Figure 2). Otoliths with moderate to severe erosion were combined into a single category, Grade 3–5, with no attempt to individually assign grade number. These otoliths ranged from moderate erosion where intricacies of otolit shape showed obvious wear and distance between otolith and sulcus edges was decreased, to severe erosion where the sulcus extent was obviously compromised or the otolith was broken (Figure 2). There were indications that digestive fluids may have entered into fish skulls, which were often found separated from the bodies and in various stages of digestion. As such, we graded all otoliths removed from fish skulls rather than automatically scoring them as grade 0 (undamaged) as has been reported in past studies (Gannon & Waples, 2004; Recchia & Read, 1989). All otoliths graded individually as 0–2 then were combined into one category (Grade 0–2) for analyses due to low sample sizes for grade 0 and grade 1 otoliths.

Reference-collection otoliths and prey otoliths from croaker, spot, spotted seatrout, and weakfish (Figure 3) were measured to the nearest 0.01 mm using a dissecting microscope (Olympus BZ61) at 6.7–10x magnification and imaging software (Olympus cellSens). We took three steps to reduce measurement error. First, the imaging software was set to collect images at the highest resolution (2,448 × 1,920 pixels). The "helper lines" tool was also used for measurements whereby perpendicular lines are shown at the beginning and end of the measured line to ensure correct placement and longitudinal alignment. Lastly, the same person measured all otoliths. Measurements included length of the otolith (OL) and the features sulcus (SL) and cauda (CL) (Figure 3). For these species, CL was chosen as a predictor over ostium length because the cauda was further from the otolith margin. The CL was measured between the inferior point of the ostium and inferior point of the cauda. Because of the large number of prey otoliths graded 3–5, a subset was measured for croaker (*n* = 120) and spot (*n* = 101). In case prey size is influenced by the size (thus, age) of the dolphin predator, the subset was first chosen by proportional representation across dolphin age-classes and the unknown age-class for dolphins without total lengths. Within each age-class category, dolphins and measured otoliths were chosen at random to best represent the sizes contained therein. Any otolith obviously compromised (e.g., broken or sulcus extent missing) was not measured. To assess the direction of erosion bias on SL and CL, otoliths from each species were examined by eye and using the dissecting microscope to assess the angle of sulcus walls (i.e., outward angle would overestimate prey size, whereas inward angle would underestimate prey size with erosion). In each case, the otoliths were examined across the longitudinal plane of the otolith.

### 2.2 | Discrimination between spotted seatrout and weakfish

We assessed two otolith characteristics on reference-collection otoliths to discriminate between spotted seatrout and weakfish prey otoliths: sulcus shape and nodules. For both sample types (reference collection and prey otoliths), we quantified the difference in sulcus shape between the two species by using the ratio CL:SL. We qualitatively assessed the presence and relative prominence of nodules by categorizing reference collection and prey otoliths as having nodules present, nodules reduced (in prominence), or nodules absent (Figure 4). We had a similar size range in the reference collection for both species, but not the largest sizes recorded for either species (Figure 5) (Robins & Ray, 1986). Nevertheless, bottlenose dolphins are unlikely to prey on fish at the upper size limit of these species (>400 mm, Gannon & Waples, 2004; Robins & Ray, 1986) so we would not expect to find those older fish in stomachs. Using only data from reference-collection otoliths, we conducted a recursive partition analysis (i.e., Classification and Regression Tree [CART] (SAS JMP 12) that produces binary splits of observations with similar response values into clusters (Breiman, Friedman, Stone, & Olshen, 1984). We used two candidate covariates, CL:SL and nodule category, to predict response variable spotted seatrout or weakfish. With this approach, all possible splits of candidate covariates create an optimal decision tree with the minimum number of misclassifications (maximizing homogeneity) while also minimizing the complexity, in this case using 10-fold cross-validation. The results from the CART analysis were used to assign spotted seatrout or weakfish prey otoliths to species. Comparisons were made between visual identification and assigned identification from the CART analysis. When species identification differed, two readers verified prey identification based on quantitative diagnostics of CL:SL or visual comparison with reference-collection otoliths.

### 2.3 | Estimating prey size

Using reference-collection otoliths for croaker, spot, spotted seatrout, and weakfish, linear and curvilinear (Gompertz, Logistic)
regression models were fitted to examine the relationship between the predictors (OL, SL, CL) and two measures of fish size, length (total length, mm), and weight (g) (SAS JMP 12). We assumed that the broad range in fish sizes available for each species would account for any differential growth among the predictors with fish growth (Aguirre, 2003). The best fit was determined by the lowest Akaike information criterion (AIC) values with two caveats. First, we used the simpler linear model when its ΔAIC ≤ 2 from the lowest AIC (Burnham & Anderson, 2004). Second, for consistency, if one model fit better for two of three predictors for a species, we used that model for all predictors. For the final models, we assessed how the variance among predictors affected estimated prey size for each species. First, we subtracted the measured fish size from the predicted fish size per fish for each of the predictors, OL, SL, and CL (referred to as measured–predicted). For each species, we used ANOVAs to test for differences in the mean measured–predicted across OL, SL, and CL for length, and separately for weight.

2.4 | Assessing differential erosion rates

Detecting differential erosion rates across the predictors, otolith, sulcus, and cauda lengths, is confounded because it is not possible to know their initial lengths before a fish was eaten. From uneroded (i.e., reference collection) otoliths, however, we can model the expected relationship between pairs of predictors (baseline relationships), and determine if those relationships are maintained for eroded prey otoliths. We used two metrics to assess if digestive erosion affected lengths of the predictors equally. First, we calculated baseline relationships from reference-collection otoliths for all pairs of predictors using linear regressions: OL to CL, OL to SL, and SL to CL. We then repeated the regressions for prey otoliths by grade and compared the direction and variability of the relationships to the baseline. For example, we overlaid the measured OLs from prey otoliths against the measured CLs from prey otoliths by grade category with the comparable baseline relationships. This series of plots was repeated for SL against CL and for OL against SL.
Given the strong relationship between each predictor and fish size, the second metric was to determine if differential rates of erosion on the three predictors significantly affected estimated fish size (length and weight). The equations from the best-fitting models per predictor from reference-collection otoliths were applied to estimate length and weight of fish from prey otoliths. We tested whether estimated prey size differed among measured predictors and between grade categories (Grade 0–2 and Grade 3–5) using a nested repeated measures analysis of variance (ANOVA) that incorporates a linear mixed-effects model (JMP 12 with add-in available from https://community.jmp.com/t5/JMP-Add-Ins/Full-Factorial-Repeated-Measures-ANOVA-Add-In/ta-p/23904). Repeated-measures tests take into account that there were three measurements from each otolith, OL, SL, and CL. The resulting three estimated prey sizes were nested within their corresponding grade category to account for variation between grade categories. These analyses were performed separately for length and weight, and separately for croaker, spot, and weakfish. Data from spotted seatrout were not included because few otoliths were recovered from stomachs (Table 1). Tukey HSD post hoc tests were used when ANOVAs were significant.

2.5 Methods of estimating consumption biomass

One of the assumptions of applying the mean estimated fish length or weight from otoliths Grade 0–2 across all samples to otoliths Grade 3–5 is that there are no seasonal differences in prey

**FIGURE 4** Spotted seatrout (*Cynoscion nebulosus*) and weakfish (*Cynoscion regalis*) otoliths showing prominence of nodules: present (top), reduced (middle), and absent (bottom). For weakfish, the top and bottom otoliths are from the same fish. FTL = fish total length

**FIGURE 5** Distribution of fish total length (FTL) measured from reference-collection fish samples and estimated from prey otoliths recovered from dolphin stomachs. Otoliths measured from reference-collection fish were used in regressions to predict fish size from otolith length (OL), sulcus length (SL), and caudal length (CL), for the shape analysis (Classification and Regression Tree [CART]), or both. For prey otoliths, estimated FTL was from measurement of otoliths Grade 0–2 using prediction equations from reference-collection otoliths. Sulcus length was used to estimate FTL of croaker (*Micropogonias undulatus*); cauda length was used to estimate FTL of spot (*Leiostomus xanthurus*), spotted seatrout (*Cynoscion nebulosus*), and weakfish (*Cynoscion regalis*). Note x-axes scaled differently.
We tested this assumption using weight because it is used to examine consumption biomass by a predator. First, we estimated weights of fish represented by prey otoliths using species-specific predictors (OL, SL, or CL) depending on results from the nested repeated measures ANOVA outlined above. We then tested for differences in mean weight across seasons within a given species and grade category using a nonparametric Kruskal–Wallis test and Tukey–Kramer HSD post hoc test because parametric test assumptions were violated (unequal variances, \( p < .05 \)). For example, differences in mean weight among seasons for croaker prey otoliths Grade 0–2 were tested separately from differences in mean weight among seasons for croaker prey otoliths Grade 3–5. Similarly, these tests were repeated for spot and weakfish.

Lastly, we examined differences in consumption biomass of a given prey species for each dolphin across three scenarios that exclude or include measurements of otoliths Grade 3–5:

### 2.5.1 Standard scenario

Only fish weights estimated from otoliths Grade 0–2 were used. Mean estimated fish weight was calculated from otoliths Grade 0–2 across all dolphins and then applied to the number of otoliths Grade 3–5 for each dolphin.

\[
\text{Biomass}_{\text{St}} = \sum W_{\text{Grade}0–2} + N_{\text{Grade}3–5} \left( \frac{\sum W_{\text{Grade}0–2}}{N_{\text{Grade}0–2}} \right);
\]

where Biomass is the weight of all fish of a given species found within an individual dolphin’s stomach \( (St) \), \( W \) is the estimated weight from recovered otoliths, \( N \) is the number of otoliths, and subscripts refer to grade categories (i.e., \( \text{Grade}0–2 \) and \( \text{Grade}3–5 \)).

### 2.5.2 Iterative scenario

Only fish weights estimated from otoliths Grade 0–2 were used. In contrast to Scenario A, the mean estimated fish weight calculated from otoliths Grade 0–2 was applied to otoliths Grade 3–5 in an iterative process. When possible, the mean estimated weight from otoliths Grade 0–2 within a stomach was applied to otoliths Grade 3–5 within that same stomach (Gannon & Waples, 2004).

\[
\text{Biomass}_{\text{St}} = \sum W_{\text{Grade}0–2} + N_{\text{Grade}3–5} \left( \frac{\sum W_{\text{Grade}0–2}}{N_{\text{Grade}0–2}} \right);
\]

where subscripts refer to month.

If there were no otoliths Grade 0–2 within the stomach, the mean estimated weight of otoliths Grade 0–2 from stomachs of dolphins that stranded within the same month was applied to otoliths Grade 3–5.

\[
\text{Biomass}_{\text{St}} = \sum W_{\text{Grade}0–2} + N_{\text{Grade}3–5} \left( \frac{\sum W_{\text{Grade}0–2}}{N_{\text{Grade}0–2}} \right);
\]

where subscripts refer to season.

If there were no otoliths Grade 0–2 within the same season, the mean estimated weight of otoliths Grade 0–2 from stomachs of dolphins that stranded within the same season was applied to otoliths Grade 3–5.

\[
\text{Biomass}_{\text{St}} = \sum W_{\text{Grade}0–2} + N_{\text{Grade}3–5} \left( \frac{\sum W_{\text{Grade}0–2}}{N_{\text{Grade}0–2}} \right);
\]

where subscripts refer to season.

If there were no otoliths Grade 0–2 within the same season, the mean estimated weight of otoliths Grade 0–2 from stomachs across the study was applied to otoliths Grade 3–5 (essentially the Standard Scenario).

### 2.5.3 Inclusive scenario

In this analysis, estimated fish weights from all otoliths, Grade 0–2 and Grade 3–5, were used. The mean estimated fish weight was calculated from otoliths (all grades) in an iterative process similar to the Iterative Scenario to apply to unmeasured otoliths (i.e., broken or not subsampled for measurement).

\[
\text{Biomass}_{\text{St}} = \sum W_{\text{measured}} + N_{\text{Unmeasured}} \left( \frac{\sum W_{\text{measured}}}{N_{\text{measured}}} \right);
\]
where subscripts refer to all measured otoliths (\textsubscript{measured}), or unmeasured otoliths (\textsubscript{unmeasured}). Using the estimated consumed biomass of each prey species across the three scenarios for each dolphin, we tested for differences using a repeated measures ANOVA and Tukey HSD post hoc test (JMP 12). These analyses were performed for all species except spotted seatrout because of a low sample size.

3 | RESULTS

3.1 | Sampling

The reference collection included croaker, spot, spotted seatrout, and weakfish specimens that were all collected in NC. Specimens were distributed across the size range reportedly consumed by bottlenose dolphins in NC (Figure 5) (Gannon & Waples, 2004). Weight data were missing for some spotted seatrout and weakfish specimens resulting in lower samples sizes used in the regression analyses than the CART analysis. Croaker had the lowest sample size (\(n = 53\)) and most constrained size range.

Out of 213 stranded bottlenose dolphins, 169 met criteria for inclusion in the analysis. Of those, 120 had identifiable contents in the forestomach, esophagus, or both. All age-class categories were represented: YOY (\(n = 6\)), older calves (\(n = 33\)), subadults (\(n = 34\)), and adult (\(n = 36\)). Age-class category was unknown for 11 dolphins because length could not be determined, primarily because their flukes were removed during human interactions (see Byrd et al., 2014). The 120 dolphins were distributed across all seasons: Spring, \(n = 44\); Summer, \(n = 16\); Fall, \(n = 25\); Winter, \(n = 35\). Croaker otoliths were found in 52% of the 120 stomachs, spot in 62%, spotted seatrout in 10%, and weakfish in 47%. These four species comprised 74% (\(n = 5,308\) of 7,120) of fish represented by otoliths (Table 1). The majority of prey otoliths were Grade 3–5 (89%) (Table 1). Few prey otoliths removed from fish skulls (\(n = 386\) of 5,308) were undamaged (Grade 0, \(n = 4\)) or barely degraded (Grade 1, \(n = 39\)) because most skulls had been partially digested. The majority of otoliths recovered from such skulls were Grade 2 (\(n = 159\)) or Grade 3–5 (\(n = 184\)).

3.2 | Discrimination between spotted seatrout and weakfish

The CART analysis using reference-collection otoliths indicated that one split produced the optimal decision tree using the predictor variable CL:SL at a cut-point of .49. With one split, the \(r^2 = .89\) and the classification accuracy was .98 for both species. All otoliths with CL:SL < .49 were assigned to spotted seatrout (\(n = 164\)). All but six otoliths with CL:SL > .49 were assigned to weakfish (\(n = 173\)). Forcing an additional split showed that the uncertainty occurred when CL:SL was between .49 and .50, which included six spotted seatrout and 25 weakfish (\(r^2 = .93\)) (Table 2). The contribution of nodules was not a candidate in either the optimal split or the forced extra split.

### TABLE 2 Predicted species, spotted seatrout (Cynoscion nebulosus) or weakfish (Cynoscion regalis), assignment from a recursive partition analysis of Cauda Length (CL): Sulcus Length (SL) from reference-collection otoliths. With this analysis, probabilities are never zero (\(p ≠ .0\)) even when the assignment number (\(n\) is 0

| Species                | CL:SL ≤ 0.48 | CL:SL > 0.49 < 0.50 | CL:SL ≥ 0.51 |
|------------------------|--------------|----------------------|--------------|
| Spotted seatrout       | 164 .997     | 6 .202               | 0 .003       |
| Weakfish               | 0 .003       | 25 .798              | 142 .997     |

The cut-points of < .49 and > .50 were applied to unbroken prey otoliths initially identified as 125 spotted seatrout and 914 weakfish, resulting in a predicted distribution of 17 spotted seatrout (< .49), 1,001 weakfish (> .50), and 21 fish that fell between the two cut-points. All 17 spotted seatrout were initially correctly identified. Of the 1,001 fish predicted to be weakfish, 99 were originally identified as spotted seatrout. Of the 21 fish with CL:SL values from .49 to .50, initial identifications were 9 spotted seatrout and 12 weakfish. The 21 were re-examined by visual comparison to reference-collection otoliths of similar size resulting in changes to the identification (6 spotted seatrout, 15 weakfish), which is slightly different than applying the CART analysis probability of .80 (4 spotted seatrout, 17 weakfish). Although CL:SL could not be measured and calculated for broken otoliths initially identified as seven spotted seatrout and 80 weakfish, they were visually compared to reference-collection otoliths for the second time. Five of the seven otoliths originally as spotted seatrout were changed to weakfish resulting in a final count of two spotted seatrout and 85 weakfish. Final identifications for measured and broken otoliths were 25 spotted seatrout (23 measured and 2 broken) and 1,101 weakfish (1,015 measured and 85 broken). Nine of the 12 dolphins that ate spotted seatrout also ate weakfish.

Nodules on reference-collection otoliths were present for 96% of spotted seatrout (108 of 113) and 34% of weakfish (37 of 110) (Figure 4). When present, nodules were categorized as reduced for 31% of spotted seatrout and 57% of weakfish. Despite digestive erosion, nodules were present and not reduced on most prey otoliths of spotted seatrout (present = 23, absent = 1, not recorded/broken = 2). Weakfish showed the opposite trend (present = 190, reduced = 97, absent = 734, not recorded/broken = 80).

3.3 | Estimating prey size

Using data from reference-collection otoliths, the best-fitting model to estimate prey size differed among species and between predicting length versus weight (Table A1). The \(\Delta AIC\) values indicated that nonlinear models often provided the best fit across the three predictive features (Figures A1–A4, Table A1). For length models of spotted seatrout, the best-fitting models differed among the three predictors; models from OL and SL were linear (\(\Delta AIC = 0\),
but the ∆AIC value for the linear model from CL was 12.5. As a compromise, we selected the logistic model for which the CL model ∆AIC = 0, and the OL and SL model ∆AIC values were slightly over 2.0 (2.1 and 2.3, respectively). Of the best models for each species and predictor, r² values were high, ranging from .88 to .98 (Table 3). Differences in r² values among predictors were negligible in most cases; however, for croaker the r² values of models predicting length and weight from CL (length = .88, weight = .89) were .08 lower than from OL (length = .96, weight = .97) (Table 3). Visual inspection of the residuals showed no trends meaning lower r² values represented higher variances around estimated fish size, but no bias in the direction of that variance. For each species, there was no significant difference in the mean measured–predicted values for length and for weight based on OL, SL, and CL (croaker TL: $df = 2$, $F$ ratio < .0001, $p = 1.0$; croaker FM: $df = 2$, $F$ ratio = .0013, $p = 1.0$; spot TL: $df = 2$, $F$ ratio = .0002, $p = 1.0$; spot FM: $df = 2$, $F$ ratio = .0085, $p = .99$; spotted seatrout TL: $df = 2$, $F$ ratio = .0004, $p = 1.0$; spotted seatrout FM: $df = 2$, $F$ ratio = .0083, $p = .99$; weakfish TL: $df = 2$, $F$ ratio < .0001, $p = 1.0$, weakfish FM: $df = 2$, $F$ ratio = .0015, $p = 1.0$).

### Table 3. Best-fitting models used to predict the relationship between otolith length (OL), sulcus length (SL), and cauda length (CL) (mm) to fish total length (FTL, mm) and fish weight (FW, g) for croaker (Micropogonias undulatus), spot (Leiostomus xanthurus), spotted seatrout (Cynoscion nebulosus), weakfish (Cynoscion regalis). Gompertz and Logistic models have three parameters. Regression = Regress. EXP = inverse natural logarithm

| Species          | Regress. | Model       | $r^2$ | Prediction Model                                                                 |
|------------------|----------|-------------|-------|-----------------------------------------------------------------------------------|
| Croaker          | OL-FTL   | Linear      | .96   | FTL = (−25.51784) + (24.410412*OL)                                                |
|                  | SL-FTL   | Linear      | .95   | FTL = (−20.75988) + (29.028549*SL)                                                |
|                  | CL-FTL   | Linear      | .88   | FTL = (1.2494378) + (61.30351*CL)                                                 |
|                  | OL-FW    | Logistic    | .97   | FW = 286.38443/(1 + EXP(−0.6077569*(OL−14.004909)))                               |
|                  | SL-FW    | Logistic    | .94   | FW = 262.50792/(1 + EXP(−0.7774777*(SL−8.3679234)))                               |
|                  | CL-FW    | Logistic    | .89   | FW = 261.82446/(1 + EXP(−1.6664716*(CL−3.5895675)))                               |
| Spot             | OL-FTL   | Logistic    | .98   | FTL = 380.79763/(1 + EXP(−0.3929095*(OL−6.705528)))                               |
|                  | SL-FTL   | Logistic    | .98   | FTL = 353.24343/(1 + EXP(−0.4677786*(SL−5.3485423)))                               |
|                  | CL-FTL   | Logistic    | .95   | FTL = 350.7459/(1 + EXP(−1.056006*(CL−2.5360011)))                                |
|                  | OL-FW    | Gompertz    | .97   | FW = 85.841.563*(EXP(−EXP(−0.085671*(OL−16.16622)))                             |
|                  | SL-FW    | Gompertz    | .96   | FW = 29.777.77*(EXP(−EXP(−0.1101145*(SL−21.640302)))                             |
|                  | CL-FW    | Gompertz    | .92   | FW = 483.412.11*(EXP(−EXP(−0.1733285*(CL−15.095524)))                           |
| Spotted seatrout | OL-FTL   | Logistic    | .97   | FTL = 793.22148/(1 + EXP(−0.1702467*(OL−14.004909)))                              |
|                  | SL-FTL   | Logistic    | .97   | FTL = 659.58373/(1 + EXP(−0.5267355*(CL−5.3660654))                               |
|                  | CL-FTL   | Logistic    | .95   | FTL = 2042.2502/(1 + EXP(−0.3516577*(OL−19.132198))                              |
|                  | OL-FW    | Logistic    | .95   | FTL = 2029.3521/(1 + EXP(−0.4105468*(CL−16.059515))                              |
|                  | SL-FW    | Logistic    | .90   | FTL = 1654.0488/(1 + EXP(−1.1770936*(CL−6.5814683))                              |
| Weakfish         | OL-FTL   | Gompertz    | .98   | FTL = 2,477.0097*(EXP(−EXP(−0.0394058*(CL−33.676056)))                         |
|                  | SL-FTL   | Gompertz    | .97   | FTL = 3,409.5556*(EXP(−EXP(−0.0379836*(OL−36.322885)))                         |
|                  | CL-FTL   | Gompertz    | .97   | FTL = 2,957.7632*(EXP(−EXP(−0.0793614*(CL−17.141034)))                         |
|                  | OL-FW    | Logistic    | .93   | FW = 5,861.9572/(1 + EXP(−0.2409874*(CL−27.556653))                             |
|                  | SL-FW    | Logistic    | .93   | FW = 5,941.1188/(1 + EXP(−0.2737339*(OL−24.319011))                             |
|                  | CL-FW    | Logistic    | .92   | FW = 5,377.982/(1 + EXP(−0.5400035*(CL−12.275319))                             |

### 3.4 Assessing differential erosion rates

Relationships between predictors (OL, SL, and CL) measured from prey otoliths showed deviation from the baseline relationships, although analyses of covariance (ANCOVA) tests could not be used because slopes of the regression lines were unequal ($p < .05$). For spot and weakfish, regressions of measured OL to measured CL and measured SL to measured CL were negatively biased relative to the expected 1:1 baseline relationship (Figure 6). Grade category did not affect the degree of negative bias for spot or SL to CL for weakfish, but did for OL to CL for weakfish. Otolith length to SL, however, showed no bias between measured and baseline relationships.

Regression results from croaker otoliths differed from spot and weakfish. For croaker, regressions of measured OL to measured CL for grade category 0–2 became positively biased with increased measured CL relative to the baseline relationship (Figure 6). A similar pattern was seen for regression lines of SL to CL for both grade categories. Finally, regressions of measured OL to measured SL were negatively biased relative to the baseline relationship, more so for Grade 3–5 otoliths than Grade 0–2 otoliths.
The differential erosion rates seen in the afore-mentioned baseline model comparisons were evident in the estimated fish sizes across the three predictors, OL, SL, and CL. For estimating original fish size from prey otoliths, the most common pattern was that estimated fish size (length and weight) was generally larger from CL than SL and/or OL (Table 4, Figure 7). For spot, there was no significant difference in estimated length and weight between grade categories, but CL did estimate larger prey size than OL and SL. Across
both grade categories, estimated size from CL was ~2.5% larger than from OL for length and ~9% larger for weight. For weakfish, there was a significant difference in estimated length and weight among predictors and across grade categories. Weakfish CL always estimated significantly larger fish than SL and OL \((p < .05)\) and across grade categories. Weakfish CL always estimated significantly larger fish than otoliths Grade 3–5 \((p < .05)\). In fact, estimated length from otoliths Grade 0–2 was 3% larger using CL rather than OL, and estimated weight from otoliths Grade 0–2 were 8% larger using CL rather than OL. The difference between the two predictors was even larger for otoliths Grade 3–5 (~8% for length and 20% for weight).

The results for croaker were less consistent between estimated length and weight. For estimated length of croaker, there was a significant difference \((p < .05)\) among predictors and across grade categories (Table 4). Croaker CL and SL estimated larger lengths than OL, but there was no difference in estimated length between SL and CL. Mean estimated length was generally larger from otoliths Grade 0–2 than Grade 3–5, but this was most pronounced for the significantly lower estimated length from OL. For estimated weight of croaker, there was no significant difference among predictors, but otoliths Grade 0–2 estimated significantly larger weights (25%) than otoliths Grade 3–5 \((p < .05)\).

The differential digestive erosion rates across the three predictors could be caused by erosion decreasing OL relative to CL or SL, or erosion increasing CL or SL relative to the other predictors. Our inspection of reference-collection otoliths found that for all three species the sulcus walls were nearly parallel except for a slight angle inward at the sulcus floor where the transition to the walls was smooth. The shape of the sulcus walls means that erosion would not increase CL and SL; thus, erosion was greater for OL where the edges are abraded than for CL or SL, as the latter two were contained within the margin of the otolith. For spot and weakfish, for which the ostium portion of the sulcus is close to the otolith edge, the effect of digestion was greater for SL than for CL.

### Table 4

| Source                  | df | F Ratio | \(p > F\)  |
|-------------------------|----|---------|------------|
| **Total length**        |    |         |            |
| Croaker (Grade 0–2 = 284, Grade 3–5 = 120) |       |         |            |
| Grade cat.              | 1  | 7.48    | .0065      |
| Pred.                   | 2  | 30.02   | <.0001     |
| Grade cat. * Pred.      | 2  | 5.53    | .0041      |
| **Weight**              |    |         |            |
| Croaker (Grade 0–2 = 284, Grade 3–5 = 120) |       |         |            |
| Grade cat.              | 1  | 4.57    | .0332      |
| Pred.                   | 2  | 2.47    | .0855      |
| Grade cat. * Pred.      | 2  | 1.6     | .2025      |

| Source                  | df | F Ratio | \(p > F\)  |
|-------------------------|----|---------|------------|
| **Spot (Grade 0–2 = 195, Grade 3–5 = 101)** |       |         |            |
| Grade cat.              | 1  | 0.39    | .5341      |
| Pred.                   | 2  | 36.01   | <.0001     |
| Grade cat. * Pred.      | 2  | 1.31    | .2713      |

| Source                  | df | F Ratio | \(p > F\)  |
|-------------------------|----|---------|------------|
| **Weakfish (Grade 0–2 = 117, Grade 3–5 = 876)** |       |         |            |
| Grade cat.              | 1  | 55.44   | <.0001     |
| Pred.                   | 2  | 441.69  | <.0001     |
| Grade cat. * Pred.      | 2  | 51.59   | <.0001     |

| Source                  | df | F Ratio | \(p > F\)  |
|-------------------------|----|---------|------------|
| **Weight**              |    |         |            |
| Spot (Grade 0–2 = 195, Grade 3–5 = 101) |       |         |            |
| Grade cat.              | 1  | 0.55    | .4607      |
| Pred.                   | 2  | 30.56   | <.0001     |
| Grade cat. * Pred.      | 2  | 0.05    | .9469      |

| Source                  | df | F Ratio | \(p > F\)  |
|-------------------------|----|---------|------------|
| **Weakfish (Grade 0–2 = 117, Grade 3–5 = 876)** |       |         |            |
| Grade cat.              | 1  | 42.28   | <.0001     |
| Pred.                   | 2  | 205.30  | <.0001     |
| Grade cat. * Pred.      | 2  | 11.32   | <.0001     |

#### 3.5 Methods of estimating consumption biomass

There were significant differences in mean estimated weight across seasons for otoliths Grade 0–2 and Grade 3–5 from croaker, spot, and weakfish (Figure 8). Mean estimated weights of croaker and spot were significantly larger during fall than during other seasons for otoliths Grade 0–2 (croaker: \(df = 3, F Ratio = 32.85, p < .0001\); spot: \(df = 3, F Ratio = 42.36, p < .0001\)) and otoliths Grade 3–5 (croaker: \(df = 3, F Ratio = 19.05, p < .0001\); spot: \(df = 3, F Ratio = 14.62, p < .0001\)). For weakfish otoliths Grade 0–2, estimated fish weight was significantly greater during winter than during spring and summer (\(df = 3, F Ratio = 9.56, p < .0001\)). For weakfish otoliths Grade 3–5, estimated fish weights during winter and fall were significantly greater than during spring and summer (\(df = 3, F Ratio = 42.72, p < .0001\)).

Estimated weights across the three scenarios for estimating consumption biomass were significantly different for weakfish (\(df = 2, F Ratio = 6.90, p = .0015\)), but no significant difference was found for croaker (\(df = 2, F Ratio = 0.81, p = .45\)) or spot (\(df = 2, F Ratio = 2.66, p = .07\)) (Figure 9). For weakfish, weights calculated in the Inclusive Scenario (using actual measurements from otoliths Grade 3–5) were significantly lower than the Standard and Iterative Scenarios. For the iterative selection approach (Iterative and Inclusive Scenarios), we found otoliths Grade 0–2 within the stomach, month, or season. Most stomachs containing weakfish (38 of 57) had otoliths Grade 0–2 within the stomach to scale to the otoliths Grade 3–5, while less than half of the stomachs containing croaker (30 of 63) and spot (32 of 75) had otoliths Grade 0–2 within the same stomach with otoliths Grade 3–5. Many stomachs had otoliths Grade 3–5 that could be scaled with otoliths Grade 0–2 from other stomachs in the same month; relatively few stomachs had otoliths Grade 3–5 that could only be scaled with otoliths Grade 0–2 from the same season (croaker = 5, spot = 2, weakfish = 13).
Use of the sulcus provided benefits of producing an easily repeatable way to distinguish between species with similar otoliths and of producing estimates of fish size with a reduced influence of digestion. Qualitative and quantitative assessments of the sulcus have been used in nondiet studies to distinguish among genera, species, and stocks (Chao, 1978; Torres et al., 2000; Tuset et al., 2008). The quantitative approach used in the current study was simple compared to more involved otolith shape analyses used in fish studies (Campana & Casselman, 1993; Colura & King, 1995; Škeljo & Ferri, 2012). Otolith chemistry is an alternative approach that has been reported for distinguishing among fish stocks in fish-focused studies (e.g., Campana, Chouinard, Hanson, Fréchet, & Brattey, 2000) and among species in a pinniped diet study (Ferenbaugh, Strauss, Tollit, Chen, & Diamond, 1995; Johnson, Ross, McKenna, & Lewis, 2006; Pierce & Boyle, 1991; Tollit et al., 1997). In the current study, comparisons of measured predictors from prey otoliths to baseline relationships between predictors indicated differential erosion rates for OL, SL, and CL. These differences were borne out in the significant differences for spot and weakfish in estimated fish length and weight among the predictors, and for croaker in estimated fish length among predictors. For all species, where significant differences occurred, estimated fish size from OL was smaller than CL, but often the same as SL. The models to estimate fish size from predictors had similar and high explanatory powers and did not account for differences in estimated prey size from these predictors in prey otoliths. As such, differences in estimated size across predictors indicate that erosion differentially affected OL compared to CL and sometimes SL. Although digestive erosion would affect otoliths in all dimensions, the margins of the sulcus (i.e., sulcus and cauda lengths) in the three species we examined are not expected to change much with erosion given that the sulcus walls are nearly parallel and not angled away from the otolith edges. The significantly larger estimates of fish size

**FIGURE 7** For prey otoliths removed from stranded bottlenose dolphins (*Tursiops truncatus*), mean estimated fish total length and weight from predictors otolith length (OL), sulcus length (SL), and cauda length (CL) nested within grade categories, Grade 0–2 (no to slight digestive erosion) and Grade 3–5 (moderate to severe digestive erosion). Beneath x-labels are letters denoting significant differences identified from Tukey HSD post hoc comparisons for significant nested repeated measures ANOVAs. Levels with different letters are significantly different. Standard error bars are shown. Note that axes for weakfish (*Cynoscion regalis*) are different from croaker (*Micropogonias undulatus*) and spot (*Leiostomus xanthurus*)

### 4 | DISCUSSION

where the two species overlap (Atlantic) and where weakfish has been reported outside of its known range (Gulf of Mexico) (Bowen, 2011). Additionally, the approach could be adapted to incorporate other congers of weakfish and spotted seatrout or adapted to other groups of species.

Understanding diets of piscivorous predators and the connection to predation effects on prey is influenced by the digestive erosion of otoliths that affects the size and, for fragile otoliths especially, number recovered (Bowen & Iverson, 2013; Casaux, Favero, Barrera-Oro, & Silva, 1995; Johnson, Ross, McKenna, & Lewis, 2006; Pierce & Boyle, 1991; Tollit et al., 1997). In the current study, comparisons of measured predictors from prey otoliths to baseline relationships between predictors indicated differential erosion rates for OL, SL, and CL. These differences were borne out in the significant differences for spot and weakfish in estimated fish length and weight among the predators, and for croaker in estimated fish length among predators. For all species, where significant differences occurred, estimated fish size from OL was smaller than CL, but often the same as SL. The models to estimate fish size from predictors had similar and high explanatory powers and did not account for differences in estimated prey size from these predictors in prey otoliths. As such, differences in estimated size across predictors indicate that erosion differentially affected OL compared to CL and sometimes SL.
from CL or SL, therefore, are likely to be closer to the original size of the fish consumed.

In the absence of correction factors, the current study demonstrated that using alternatives to OL, such as CL or SL, to estimate fish size is a reasonable and time-efficient method to limit erosion bias. For spot, weakfish, and spotted seatrout, the ostium edge is close to the otolith edge making CL a better predictor of fish size than SL. In fact, the CL for weakfish was visibly affected in only the most eroded (Grade 5) otoliths (Figure 2). For croaker, there was notable variability in sulcus shape especially at the inferior point of the ostium (Figure A5), increasing variability for the relationship between CL and the other predictive features, and decreasing the difference in estimated fish size between the SL and CL. Results for croaker may also be influenced by the lower sample size compared to spot and weakfish. The percent size difference between using OL and an alternative predictor for a given species and grade category in this study was less than from captive pinniped experiments (Reviewed in Bowen & Iverson, 2013). Although digestive erosion may not be directly comparable between otoliths recovered from pinnipeds (feces) and cetaceans (stomachs), this comparison indicates that our findings are within a reasonable range. The percent differences were small between estimated size from CL or SL compared to OL; however, the differences could translate into large effects on a fish stock when estimated consumed biomass is applied to, in this case, the total number of dolphins that seasonally occur in NC (>10,000; Hayes et al., 2018) (Laake, Browne, DeLong, & Huber, 2002).

It is still unknown how well the alternatives to OL compensate for erosion of otoliths with moderate to severe erosion (Grade 3–5). Although the mean estimated sizes for croaker and weakfish from otoliths Grade 3–5 were smaller than otoliths Grade 0–2, the opposite was true for spot, regardless of the predictor. Thus, it cannot be assumed that the mean size of otoliths Grade 0–2 is representative of otoliths Grade 3–5. For example, bottlenose dolphins along the US mid-Atlantic are known to feed multiple times in a single day (Shippee, 2014). In this case, the stomach could have otoliths of different sizes from schools of differently sized fish. Otoliths from smaller fish of even the same species are likely to erode faster (Phillips & Harvey, 2009; Tollit et al., 1997), which could result in smaller fish being under-represented by otoliths with little to no erosion. Without grade-specific correction factors (Tollit et al., 1997), should intact otoliths with moderate to severe erosion be excluded altogether? On a dolphin by dolphin basis, a difference in estimated consumed biomass among scenarios was only significant for weakfish with the Inclusive Scenario resulting in smaller biomass estimates than the other scenarios. Although estimated size may be more accurate for otoliths with less erosion, studies often recover few in that condition. The resulting sample size limitation may mean that the least eroded otoliths are not representative of fish sizes across an entire study, especially given seasonal changes in size classes of fish recovered in this study and elsewhere (Johnson et al., 2006). The Iterative Scenario to estimate weight of prey species would account for the
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The chosen approach (i.e., scenario), therefore, may depend on a re-
by minimizing erosion bias. No approach is without bias, however.

Some or all otoliths of moderate to severe erosion (Inclusive Scenario)
measurements to OL, such as CL for weakfish, may allow inclusion of
a preferable alternative to the Standard Scenario. Using alternative
understanding of diet composition of predators and of natural
size. Applying these approaches has the potential to improve the
liths provides the most representative true composition of prey
fish (Tollit et al., 1997), perhaps using all but severely eroded oto-
part differences in estimated prey size between grade categories are only
result of digestive erosion or feeding bouts of differently sized
fish (Tollit et al., 1997), perhaps using all but severely eroded oto-
liths provides the most representative true composition of prey
size. Applying these approaches has the potential to improve the
understanding of diet composition of predators and of natural
mortality (consumption) of prey species in trophic studies and im-
prove ecosystem-based management.

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CONFLICT OF INTEREST
None declared.

AUTHORS’ CONTRIBUTIONS
BLB and AAH conceived the ideas and designed methodology; BLB,
JRK, and AAH collected the data; BLB and AAH analyzed the data.
All authors contributed to the writing of the manuscript and gave
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All data used in this study are archived and available from the US
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APPENDIX 1

**FIGURE A1** For croaker, Linear (green line), Logistic (blue line), and Gompertz (orange line) models were compared using ΔAIC Information Criterion (ΔAIC) and $r^2$ values to determine the best fit between otolith length (OL), sulcus length (SL), and cauda length (CL), to fish total length (Fish TL, top row) and fish weight (bottom row). See Supplemental Tables A1 & A2 for ΔAIC and $r^2$ values.
FIGURE A2  For spot, Linear (green line), Logistic (blue line), and Gompertz (orange line) models were compared using ∆Akaike Information Criterion (ΔAIC) and $r^2$ values to determine the best fit between otolith length (OL), sulcus length (SL), and cauda length (CL), to fish total length (Fish TL, top row) and fish weight (bottom row). See Supplemental Tables A1 & A2 for ΔAIC and $r^2$ values.
For spotted seatrout, Linear (green line), Logistic (blue line), and Gompertz (orange line) models were compared using \( \Delta \text{Akaike Information Criterion (AIC)} \) and \( r^2 \) values to determine the best fit between otolith length (OL), sulcus length (SL), and cauda length (CL), to fish total length (Fish TL, top row) and fish weight (bottom row). See Supplemental Tables A1 & A2 for \( \Delta \text{AIC} \) and \( r^2 \) values.
FIGURE A4  For weakfish, Linear (green line), Logistic (blue line), and Gompertz (orange line) models were compared using $\Delta$Akaike Information Criterion ($\Delta$AIC) and $r^2$ values to determine the best fit between otolith length (OL), sulcus length (SL), and cauda length (CL), to fish total length (Fish TL, top row) and fish weight (bottom row). See Supplemental Tables A1 & A2 for $\Delta$AIC and $r^2$ values.
TABLE A1 For otoliths from a reference collection, sample size, ΔAIC (Akaike Information Criterion) score, and $r^2$ values of models used to predict fish total length (FTL, mm) and fish weight (FW, g) from otolith length (OL), sulcus length (SL), and cauda length (CL) (mm) for croaker (n = 53), spot (n = 104), spotted seatrout (n = 133), and weakfish (n = 131). Gompertz and Logistic models have 3 parameters. See Figs. A1-A4 for model plots.

| Regression | Model   | ΔAIC | $r^2$ | ΔAIC | $r^2$ | ΔAIC | $r^2$ | ΔAIC | $r^2$ |
|------------|---------|------|-------|------|-------|------|-------|------|-------|
| OL-FTL     | Gompertz| 0.6  | 0.97  | 0.7  | 0.98  | 1.8  | 0.97  | 0.0  | 0.98  |
|            | Linear  | 0.7  | 0.96  | 8.3  | 0.98  | 0.0  | 0.97  | 23.3 | 0.98  |
|            | Logistic| 0.0  | 0.97  | 0.0  | 0.98  | 2.1  | 0.97  | 4.4  | 0.98  |
| SL-FTL     | Gompertz| 0.7  | 0.95  | 0.2  | 0.98  | 2.0  | 0.97  | 0.0  | 0.98  |
|            | Linear  | 1.6  | 0.95  | 4.1  | 0.98  | 0.0  | 0.97  | 29.6 | 0.97  |
|            | Logistic| 0.0  | 0.95  | 0.0  | 0.98  | 2.3  | 0.97  | 3.0  | 0.98  |
| CL-FTL     | Gompertz| 1.6  | 0.88  | 1.1  | 0.95  | 3.2  | 0.97  | 0.0  | 0.97  |
|            | Linear  | 0.0  | 0.88  | 2.8  | 0.95  | 12.5 | 0.93  | 21.9 | 0.96  |
|            | Logistic| 1.1  | 0.88  | 0.0  | 0.95  | 0.0  | 0.94  | 1.8  | 0.97  |
| OL-FW      | Gompertz| 2.3  | 0.96  | 0.0  | 0.97  | 13.7 | 0.95  | 0.0  | 0.93  |
|            | Linear  | 15.0 | 0.95  | 180.9| 0.83  | 79.2 | 0.92  | 123.5| 0.82  |
|            | Logistic| 0.0  | 0.97  | 7.8  | 0.97  | 0.0  | 0.95  | 0.1  | 0.93  |
| SL-FW      | Gompertz| 3.4  | 0.94  | 0.0  | 0.97  | 16.1 | 0.94  | 0.4  | 0.93  |
|            | Linear  | 9.4  | 0.93  | 165.6| 0.84  | 73.7 | 0.91  | 130.7| 0.81  |
|            | Logistic| 0.0  | 0.94  | 9.8  | 0.97  | 0.0  | 0.95  | 0.0  | 0.93  |
| CL-FW      | Gompertz| 1.9  | 0.89  | 0.0  | 0.93  | 12.6 | 0.89  | 0.0  | 0.92  |
|            | Linear  | 6.5  | 0.87  | 102.1| 0.80  | 37.4 | 0.86  | 112.4| 0.81  |
|            | Logistic| 0.0  | 0.89  | 3.2  | 0.92  | 0.0  | 0.90  | 0.0  | 0.92  |

FIGURE A5 Croaker otoliths removed from bottlenose dolphin stomachs (top: grade 2, bottom: grade 1). Notice variation in the shape and inferior position of the ostium (arrows).