ORIGINAL ARTICLE

Design and biomechanical evaluation of a rodent spinal fixation device

M Shahrokni1,2, Q Zhu1,2, J Liu3, W Tetzlaff2 and TR Oxland1,2

Study design: An in vitro and in vivo study in rats.

Objectives: To design a novel rat spinal fixation device and investigate its biomechanical effectiveness in stabilizing the spine up to 8 weeks post-injury.

Methods: A fixation device made of polyetheretherketone was designed to stabilize the spine via bilateral clamping pieces. The device effectiveness was assessed in a Sprague–Dawley rat model after it was applied to a spine with a fracture–dislocation injury produced at C5–C6. Animals were euthanized either immediately (n=6) or 8 weeks (n=9) post-injury and the C3-T1 segment of the cervical spine was removed for biomechanical evaluation. Segments of intact spinal columns (C3-T1) (n=6) served as uninjured controls. In these tests, anterior–posterior shear forces were applied to the C3 vertebra to produce flexion and extension bending moments at the injury site (peak 12.8 Nnm). The resultant two-dimensional motions at the injury site (that is, C5–C6) were measured using digital imaging and reported as ranges of motion (ROM) or neutral zones (NZ).

Results: Flexion/extension ROMs (average ± s.d.) were 18.1 ± 3.3°, 19.9 ± 7.5° and 1.5 ± 0.7°, respectively for the intact, injured/fixed, and injured/8-week groups, with the differences being highly significant for the injured/8-week group (P=0.0002). Flexion/extension NZs were 3.4 ± 2.8°, 5.0 ± 2.4°, and 0.7 ± 0.5°, respectively for the intact, injured/fixed, and injured/8-week groups, with the differences being significant for the injured/8-week group (P=0.04).

Conclusion: The device acutely stabilizes the spine and promotes fusion at the site of injury.

Spinal Cord (2012) 50, 543–547; doi:10.1038/sc.2011.185; published online 31 January 2012

Keywords: spinal cord injury; biomechanics; implant; fixation device; rat model; dislocation

INTRODUCTION

In vivo experimental SCI studies involve animal models with survival post-injury and recovery. In many of these models, the spinal column is substantially destabilized. Examples include single or double laminectomies for dorsal contusion injuries,1,2 septuple hemi-laminectomy is substantially destabilized. Examples include single or double laminectomies or anterior and posterior column damage after dorsal rhizotomies,3 or anterior and posterior column damage after a fracture–dislocation injury.4–5 Failure to stabilize the spinal column may complicate the progression of subsequent secondary damage at the site of injury and likely exacerbate pain; therefore it is important to properly stabilize the spinal column post-injury.

Different methods have been used to achieve stabilization of the spinal column post-injury in rat models.7–9 Spinal process wires as internal fixation of the vertebral column is a time-consuming technique and it is difficult to achieve a consistent degree of fixation due to the unknown tensile load applied to the wires. Transpedicular screws and rods are theoretically advantageous, but not feasible in the rat spine due to the small size of the rat vertebrae.7,8 Other methods have used casting of the spinal column with dental cement or tissue adhesive (Vet-bond).10 However, this method compromises circulation, introduces potentially neurotoxic chemicals (for example, methacrylate) and is prone to failure. There appears to be a need for a more repeatable and validated method of spinal fixation in the rat spine.

There is a well-established methodology to assess the biomechanical effectiveness of fixation devices in stabilizing the spinal column. Panjabi11 first described the basic principles and there are many examples of these being applied to a wide range of devices for the human spine.11–16 The main concept is that a fixation device is intended to decrease the relative motion between adjacent vertebrae. To assess this biomechanically, one applies known loads to the specimen and measures the resultant displacements, either immediately after injury or after a certain healing time. A substantially reduced magnitude of displacement is generally desirable, particularly in a situation where vertebral dislocation has been performed and fusion is a clinical goal. We are unaware of any studies that have evaluated a fixation device for the rat spine in such a manner.

The objectives of this study were to design a custom MRI-compatible rat cervical spinal stabilization device and to evaluate the biomechanical fixation provided by this device at 8 weeks post-injury. It is hypothesized that the stiffness of the stabilized spinal column is higher than the unstabilized spine immediately after the injury, and that the overall stiffness of the spinal column increases after 8 weeks owing to healing and scar tissue formation.

MATERIALS AND METHODS

Design

The fixation device for the rat spine met a specific set of design criteria: structural stiffness, ease of surgical handling, versatility, size and cost. Several proposed designs were considered, all of which consisted of two parts that...
clamp the vertebrae laterally. The two serrated edges on the vertebral clamps fit on the spine at the narrowing of the rat vertebrae that was anterior to the lateral masses and posterior to the transverse processes (Figure 1a). The device–spine attachment was a friction-fit; thus, it was essential to create a sufficiently high normal force at this interface. Two slots on the posterior aspect of the device (Figure 1b) accommodated attachment to different sizes of spinal columns and also enabled the user to apply a normal force to the clamps before fixation. The clamping force was applied to the outside faces of the fixation device using custom-designed instrumented forceps. A preliminary analysis of the device–structural stiffness predicted that it could provide adequate mechanical stability for the injured spine. The posterior-screw design was compact and easy for the surgeon to work within the microscopic field of vision.

The design was made of polyetheretherketone (PEEK), which is widely used in medical and clinical applications.17,18 PEEK is a rigid, bio-inert polymer with excellent biocompatibility and MRI-compatibility characteristics.19

**Injury production and spine stabilization**

All procedures were approved by the University of British Columbia Animal Care Committee in accordance with the guidelines published by the Canadian Council on Animal Care. Fixation of the device was biomechanically evaluated in a rat cervical spine model in two stages; first in vitro and second in vivo (Figure 2). The in vitro assessments were conducted on intact specimens (control) (n=6, mass=293±12 g) and specimens that were injured and fixed immediately post-injury (injured/fixed) (n=6, mass=293±12 g). The in vivo experiment was done in one group, injured and fixed with a survival time of 8 weeks (injured/8-week) (n=9, mass=291±11 g) (Figure 2). A small subgroup of animals was evaluated at 3 weeks post-injury and fixation (injured/3-week) (n=3).

In the injured groups, animals were anesthetized (2–4% isoflurane; 11/min O2) and prepared for surgery and administered a subcutaneous injection of lactated-Ringer’s (Hospira, Lake Forest, IL, USA) and Buprenorphine hydrochloride (72 mg kg \(^{-1}\)) and xylazine hydrochloride (9 mg kg \(^{-1}\)) at the corresponding time point. The spinal column was harvested from C3 to T1.

In the animals designated to the in vivo survival study, the muscle and the skin was sutured over the fixation clamp. The animals were allowed to recover normally, and appropriate post-operative procedures were taken. Specific quantitative behavioral evaluation was not conducted as it was outside the scope of this study.

At 8 weeks post injury, the in vivo group of animals was euthanized and the spinal columns were harvested similar to the in vitro group. Note that three additional injured animals were euthanized and tested at 3 weeks post-injury for a qualitative analysis at this time point. The number of specimens in this group was not statistically sufficient; therefore, the results at this time point were not used to compare the spine stabilization across the other groups at different time points.

**Biomechanical evaluation of the implant**

A custom-made experimental apparatus was designed to biomechanically evaluate the excised spinal column. Because of the small size of the rat spinal column, small and wireless visual markers were used to not obstruct or interfere with the spinal motion. The markers consisted of two small circular ‘+’ signs (3 mm diameter) glued to pins (Figure 3a), that were inserted in the C5 and C6 vertebrae. The spinal column (C3-T1) was mounted on a screw with a similar diameter as the spinal canal (Figure 3a) such that it did not interfere with the motion at C5/C6. A shorter screw was inserted into the cranial two vertebrae. The top screw was attached to a balancing weight (9.68 g) connected via a string to hold the column vertically. Another string attached to the top screw applied known shear forces at distance \(d\) (moment arm) from C5/C6 (Figure 3a). The shear forces of 0.24, 0.50, 0.74 and 0.98 N created bending moments of flexion or extension at C5/C6, depending upon the direction of the application (Figure 3b). The moment arm ‘\(d\)’ was (mean ± s.d.) 13 ± 1 mm, ensuring the application of a consistent clamping force. A small stretching load was applied on the spinal column to straighten it while implanting the device. While clamping the device on the vertebrae, strain measurements from the instrumented forceps were monitored and recorded. Once the appropriate range of force magnitude was reached, the screws were tightened and the fixation device was secured to the spinal column.

Following the injury and fixation, the in vitro group of animals were euthanized while deeply anesthetized by an over-dose of mixed ketamine hydrochloride (72 mg kg \(^{-1}\)) and xylazine hydrochloride (9 mg kg \(^{-1}\)) at the corresponding time point. The spinal column was harvested from C3 to T1.

The dislocation injury at C5/C6 was created with an electromagnetic linear actuator in a multimechanism SCI system at 364 mm s \(^{-1}\) velocity and 1.5 mm displacement.2 The displacement level of 1.5 mm was based on pilot study consistent injuries.21 The intact (control) group animals underwent identical surgical procedures and were secured within the stereotactic frame. The caudal skin was sutured over the fixation clamp. The animals were allowed to recover normally, and appropriate post-operative procedures were taken. Specific quantitative behavioral evaluation was not conducted as it was outside the scope of this study.

**Figure 2** Experiment time line showing the study groups, the interventions and the number of specimens in each group.
producing bending moments of 3.1, 6.5, 9.6 and 12.7 Nmm. Moment arm ‘d’ was measured when the spine was vertically oriented. Stepwise loads were applied from lowest to highest, per standard protocols for flexibility testing\textsuperscript{11–13} (Figure 4). The maximum load applied for the injured/fixed group was 0.74 N to avoid producing additional tissue damage in specimens that had been dislocated. Between each loading step, a time window of 30 s was given to allow the specimen to creep.\textsuperscript{11,13} To reduce cyclic viscoelastic effects, loading in flexion and extension was repeated three times and the last cycle was selected for the motion measurements (Figure 4).

The position of markers was tracked using camera snapshots (Phantom V9 camera, Vision Research, Wayne, NJ, USA). The change in angle between the markers placed into the two vertebrae represented their relative motion under different loads. The accuracy and precision of measuring 2D rotations using this motion capture method was found to be 0.5° and 0.2°, respectively.

Rotation–moment graphs for individual specimens were generated to find neutral zone (NZ) and range of motion (ROM) values. NZ was defined as the region within which the spinal motion was produced with no external load at the beginning of the third loading cycle.\textsuperscript{11} The entire motion measured from the neutral position was defined as ROM. These values were analyzed for both flexion and extension motion on the third cycle of loading to obtain the total NZ and ROM of the specimen (Figure 4).

**RESULTS**

Typical moment rotation curves for one specimen in the three experimental groups are shown in Figure 5, and the summary data for the study groups is shown in Table 1. The magnitudes of intervertebral motion for the three experimental groups are shown in Figure 6. The intact moment–rotation curve is highly nonlinear as is observed for human specimens (Figure 5). After injury and fixation with the custom device, the response was a similar motion magnitude to intact, which represents substantial stabilization given that this represents the response of a fixed joint after dislocation. The intact group was not different from the injured/fixed group for either NZ or ROM (NZ, $P=0.2$; ROM, $P=0.5$) (Figure 6).

After 8 weeks of healing, note that the motion of this joint was decreased substantially to less than 20% of the intact (Figure 5). For both NZ and ROM, the motions after 8 weeks were significantly less than the other two groups (NZ, $P=0.04$; ROM, $P=0.0002$) (Figure 6). Motion of the spine after the device was removed remained small (Table 1; NZ, 0.7deg, ROM, 1.5deg). For comparison, the injured/3-week specimen NZ and ROM values (with the device removed) were 1.8 ± 1.5 and 4.1 ± 3.1, respectively (Mean ± s.d.). Therefore, there was already substantial stabilization due to healing at this early timepoint.
DISCUSSION

This study highlights the design and biomechanical evaluation of a novel fixation device for rodents. The two overall objectives of this project were to design a custom MRI-compatible spinal stabilization device for the unstable cervical spinal column and to evaluate the biomechanical fixation provided by this device immediately after injury and at 8 weeks post-injury. The performance of the implant was evaluated by measuring the degree of biomechanical fixation that it provided to the injured spine. The results of this study demonstrated that the custom-designed fixation device was effective in stabilizing the dislocated rat cervical spine at both time points of observation.

In this study, a two-dimensional biomechanical analysis was implemented. To fully characterize the device, a three-dimensional assessment is required. However, the rat’s spine did stabilize within the 8-week healing period; thus, the initial sagittal plane analysis was deemed to be sufficient for device evaluation.

Currently, there is no study in the literature reporting the change of intervertebral motion in a rat model. There are many previous human and animal in vitro studies that report post-surgical spinal motion and generally, the spinal motion post-surgery is lower than the intact level.23 Obviously this result depends upon the injury model and the type of fixation device used. In an in vivo ovine model, a destabilizing procedure was performed in the lumbar spine followed by the application of a fixation device, and results showed that 16 and 43% reductions in ROM were achieved with the fixation device and with fixation along with the band compared with the intact.24 In a human cadaveric model, the C4/C5 ROM of the intact specimens and those with the arthroplasty device were similar and about 75% more than the specimens with the fusion construct.21 These studies show that a wide range of reduction in intervertebral motion is achieved immediately after fusion operation, up to 75% reduction compared with the intact motion, and indicate that the severity of the injury model is an important variable. The device design is clearly another determining factor in restabilizing the injured spine to the intact level.

The current study found the reductions in NZ and ROM at 8 weeks post-injury to be 79 and 92%. These values are comparable to the change in intervertebral motion in other animal models. In an in vivo rabbit model,25 the NZ and ROM of the fused specimens were significantly decreased from that of intact specimens in flexion and extension (60–80%). In a bovine model with 16 weeks of healing,26 it was found that after removing a fixation plate, subjects with allograft bone were about 30% stiffer than the subjects with a titanium cage implant, possibly indicating fusion at the construct. In a similar study on interbody fusion, results demonstrated a trend of increased fusion stiffness from 3 to 24 months.27

A novel spinal fixation device for rats was designed and its performance in providing biomechanical fixation to an injured spine confirmed that the device provided stabilization of the cervical spine post-injury, both acutely and after 8 weeks. This device is suitable for dorsal dislocation injury and enables long-term survival studies. Potential benefits of this device could include a reduction in secondary damage and the volume of scar tissue in the spinal cord post-injury.9 It will be used in future survival studies to further investigate mechanisms of spinal cord injury, characterize cord deformation using MRI, and study the effects of different treatment methods over time.

Table 1 Summary data for the study groups providing NZ and ROM values

| Study group          | NZ (deg.) | ROM (deg.) |
|----------------------|-----------|------------|
| Intact               | 3.4 ± 2.8 | 18.1 ± 3.3 |
| Injured/fixed        | 5.0 ± 2.4 | 19.9 ± 7.5 |
| Injured/8-week       |           |            |
| With device          | 0.3 ± 0.3 | 0.7 ± 0.4  |
| With device removed  | 0.7 ± 0.5 | 1.5 ± 0.7  |

Figure 5 Typical rotation–moment curves for a single specimen in each of the three experimental groups. The Intact curve (shown as triangles) depicts the classical nonlinear biomechanical behaviour seen for all human and animal spine columns. The Injured/fixed group (shown as squares) has a similar rigidity to the intact condition and only three load steps were applied in each direction owing to the presence of the dislocation injury. The Injured/8-week group (shown as diamonds) exhibited very little motion under the applied moments, demonstrating the significant rigidity of the tissues surrounding the spinal column after 8 weeks of healing and scar formation.

Figure 6 Statistical comparison of the three experimental groups is shown as means with 95% confidence intervals (ROM shown as diamonds and NZ as squares). Intact ROM and NZ were not significantly different, while the injured/8-week group exhibited significantly less ROM and NZ than the other two groups (*, **, and *** represent P-values of 0.0002, 0.006 and 0.04 respectively).
The device design may have utility in a distraction injury model, but further assessment is needed. We have identified that the sharpness of the teeth and the tightness of the screw fix on the PEEK material may be important factors in the effectiveness of the device in this injury model. It can be customized for use in the thoracic or lumbar regions of the spine, as well as for other strains of rats or mice depending on their size and vertebral structure.

This study furthers fundamental understanding of SCI mechanics in survival studies and helps facilitate future studies establishing a link between injury-specific observations and clinical treatment of human SCI.

**DATA ARCHIVING**

There were no data to deposit.

**CONFLICT OF INTEREST**

The authors declare no conflict of interest.

**ACKNOWLEDGEMENTS**

This work was supported by the Natural Sciences and Engineering Research Council of Canada and the Canadian Institutes for Health Research. The technical assistance of Peggy Assinck Femke Streijger, and Jennifer Douglas-Mills is gratefully acknowledged.