Abstract. Previous studies have revealed that fibrosis may affect the biological behavior of tumors, however associated research on papillary thyroid microcarcinoma (PTMC) is rare. The aim of the present study was to explore the association between interstitial fibrosis (IF) and the biological behavior of PTMC. In the present study, a total of 511 consecutive cases of PTMC were evaluated for the presence of IF and its association with clinical parameters and pathologic biomarkers. IF was identified in 340 of the 511 consecutive cases and it was significantly associated with the age (P=0.033), tumor diameter (P=0.017) and lymphocytic metastasis (P<0.001) of the patient. There were significantly more female in the fibrosis group than in fibrosis‑absent group (P=0.024). In the analysis of clinical biomarkers using immunohistochemical staining, IF was significantly associated with cytokeratin 19 (P=0.008) and galectin‑3 (P=0.022). Analysis of patient outcomes indicated that IF was an independent prognostic factor of recurrence (hazard ratio = 2.181; 95% confidence interval = 1.163‑4.090; P=0.015). These findings suggest that the combined effect of a patient's age, sex and tumor size may potentially contribute to fibrotic lesions and IF was a factor contributing to poor prognosis in patients with PTMC.

Introduction

Papillary thyroid carcinoma (PTC) represents ~90% of differentiated thyroid carcinomas in Japan (2004), which are the most common malignant tumors of the endocrine system (1). Papillary thyroid microcarcinoma (PTMC) is defined as PTCs with a diameter of <10 mm, and account for 39.5% of all PTCs in Italy (between 1993 and 2002) (2). Certain factors including radiation exposure, iodine deficiency and a family history of PTCs are associated with tumorigenesis and prognosis; however, the expression status of specific biomarkers, including galectin-3 and cytokeratin 19 (CK19) may also be used to predict tumor progression (3,4).

PTMC grows slowly without obvious symptoms, however due to the increased ultrasound detection rate, the incidence rate of PTMC has been rising gradually in recent years (5). The majority of PTMCs are identified by accident or are combined with other thyroid diseases (6). PTMCs may be standardly diagnosed by removing thyroid lesions and observing their pathological features and by immunohistochemistry staining of biomarkers (for example galectin-3, CK‑19 and TTF‑1) (7‑9). Surgical treatment may lead to a favorable prognosis for PTMC; however, due to the low risk of papillary thyroid cancer, various postoperative treatments have little effect on prognostic outcome, and to the best of our knowledge no previous studies have identified which therapeutic method is the best option (10). Certain PTMCs are more malignant compared with others and have a poor prognosis with early‑stage lymphocytic metastasis (11,12). These highly malignant tumors may be associated with multilocus‑tumors, tumor size and dilation of a thyroid cyst (2,13‑15).

Interstitial fibrosis (IF), which is composed of fibroblasts and a variable number of collagen fibers, has been observed in numerous malignant cancers (16‑20). This suggests that IF may be a factor associated with tumor prognosis. In previous studies (21‑23), fibrosis appears to be associated with an increased cancer recurrence rate and mortality. Previous observations have suggested that dense fibrosis may be another vital indicator for the diagnosis of PTC (24), however, few studies have focused on the associations between IF and PTC. Therefore, the aim of the present study is to perform a retrospective analysis of the clinical parameters and biomarkers of PTC and to determine the association and interactions between IF and PTMC.

Materials and methods

Patients. A total of 511 patients were recruited into the present study from the First Hospital of China Medical University (Shenyang, China), between January 2009 and December 2013; 340 patients were diagnosed with PTMC with IF and 171 patients were diagnosed with PTMC without
IF. In total, 78.86% (304) of patients were female, 21.14% were male. The age range was 20-75 years, with an mean age of 46.83±10.69. All patients underwent a thyroidectomy and met the following criteria: i) Diagnosed with PTMC according to pathological standards (25); ii) did not receive steroids or drugs that may induce fibrosis; and iii) did not present with any other disease than PTMC. Patients who had PTMC combined with thyroid inflammation or systemic diseases were excluded from the study. The present study was approved by the Institutional Review Board of China Medical University and by the First Hospital of China Medical University Medical Research Ethics Committee (Shenyang, China). Written informed consent was obtained from all patients prior to their inclusion within the present study.

All patients were divided into 2 groups. The experimental group was composed of patients with IF (n=340) and the control group was composed of patients without IF (n=171). Patients with IF were defined as having a fibrotic area composed of fibroblasts and collagen fibers within the tumor, which occupied the central part of the tumor and formed a radially expanding fibro-sclerotic core. The patient’s baseline characteristics, including age, sex, calcification, serum thyroid-stimulating hormone (TSH) level, tumor size, tumor node and metastasis stage, tumor multiplicity, bilaterality, extrathyroidal extension (ETE), subtype of papillary microcarcinoma and lymphocytic metastasis, were evaluated. The thyroid tumor stage was classified in accordance with the American Joint Committee on Cancer (26). Postoperative follow-up ended on 1 September 2016.

Histopathology. Each specimen slice was stained with hematoxylin and eosin for observing the pathological form of the cancer tissue. Freshly resected specimens, ~1 cm in thickness, were fixed in 10% neutral formaldehyde for 12 h and then dehydrated using graded series of analytical pure ethanol at room temperature as follows: 60% (1 h), 75% (30 min), 80% (15 min), 95% (10 min), 100% (5 min), then the specimens were placed in xylene at room temperature (10 min x 2), and immersed in paraffin (efficient slice paraffin, melting point 56-58°C) at 59°C for 1 h, embedded in paraffin and cooled at room temperature. Paraffin specimens were then cut into 4 µm slices, dewaxed using xylene (15 min x 2) and hydrated using graded series of analytical pure ethanol at room temperature as follows: 100% (5 min x 2), 95, 85 and 75% ethanol (3 min each). Specimens were then stained with hematoxylin for 8 min at room temperature, followed by washing with 1% hydrochloric acid alcohol for a few seconds and then washed with tap water for 10 min. Specimens were then stained with eosin for 5 min at room temperature, and then specimens were dehydrated using a graded series of ethanol at room temperature as follows: 75, 85 and 95% (2 min each), 100% (1 min x 2); transparent in xylene at room temperature (5 min x 2); neutral gum (cat. no., G8590; Solarbio, Beijing, China) sealed slice. Light microscopy was used to observe specimens (magnification, x100). The immunohistochemistry of all slices was performed by a minimum of 2 pathologists from the Pathology Department of the First Hospital of China Medical University and each experiment was repeated 3 times to confirm results. All cases were classified according to multiple clinical biomarkers, including cellular tumor antigen p53, proliferation marker protein Ki-67 (Ki-67), CK19, thyroglobulin (Tg), thyroid transcription factor I (TTF-1) and galectin-3. These biomarkers are used for the common diagnosis of PTMC, p53 and Ki67 positive samples may exhibit the proliferation of tumor cells, while CK19, TTF-1, Tg and galectin-3 positive samples may predict that the origin of the malignant cells is the thyroid epithelium.

Immunohistochemistry. Paraffin embedded sections (4 µm-thick) were dewaxed and hydrated as aforementioned. Specimens were then washed with phosphate-buffered saline (PBS), citric acid heat antigen repair for 2 min, incubated with 3% hydrogen peroxide for blocking endogenous peroxidase for 10 min at room temperature, washed with PBS, then incubated with primary antibodies at room temperature for 60 min. Negative control specimens were incubated with PBS instead of the primary antibody. All antibodies were produced by Fuzhou Maixin Biotech Co., Ltd. (Fujian, China), and were supplied at ready-to-use dilutions. The primary antibodies (100 µl) used were as follows: Galectin-3 (cat. no. MAB-0572), CK19 (cat. no. Kit-0030), TTF-1 (cat. no. MAB-0599), Ki-67 (cat. no. Kit-0005), p53 (cat. no. Kit-0010) and Tg (cat. no. MAB-0161). Sections were then washed with PBS 3 times and incubated with enzyme-labeled goat anti-mouse/rabbit immunoglobulin G polymer at room temperature (cat. no. KIT-5030) for 15 min, and then washed with PBS. Following this, DAB (cat. no. DAB-1031; Fuzhou Maixin Biotech Co., Ltd.) was added for 3-5 min at room temperature for color development to assist in light microscopy analysis (magnification, x100), and then washed with tap water for 10 min. Finally, specimens were stained with hematoxylin for 8 min at room temperature, washed with tap water, and the procedure of dehydration and transparency was the same as aforementioned. Neutral gum was used to seal the slice. Specimens were observed using light microscopy. IF was determined using 2 pathologists who were blinded to the study aims. Each experiment was repeated three times.

Statistical analysis. Data analysis was performed using SPSS (version 20; IBM Corp., Armonk, NY, USA). Continuous data were presented as the mean ± standard deviation. Dichotomous variables were compared using the χ² test or Fisher’s exact test, while the Student’s t-test was used to compare continuous variables. The survival data were analyzed using Kaplan-Meier curves and the log-rank test and Cox proportional hazard models. P<0.05 was considered to indicate a statistically significant difference.

Results

Differences between PTMC with IF and without IF. There were 511 cases of PTMC examined in the present study. Patients with IF accounted for 66.54% (340 cases), while patients without IF accounted for 33.46% (171 cases) of the cases. The differences between PTMC and normal thyroid tissue in a fibrotic formation may be observed in Fig. 1. In Fig. 1A, the normal follicular structure interrupted by squashed cancer cells, accompanied by the proliferation of fibroblasts and collagen fibers. Fig. 1B demonstrates the complete thyroid follicular structure in the nodular goiter tissue with IF, collagen fibers are positioned next to the follicles. Diffuse collagen fibers appear next to the nodular goiter and there is almost no fibroblast proliferation.
Association of IF with clinical parameters. The details and clinical characteristics of all patients included in the present study are presented in Table I. Older patients (≥45 years) had a significantly increased incidence of IF (P=0.012) compared with patients <45 years. In addition, significantly more females presented with IF (P=0.024) compared with males that

| Characteristic                  | PTMC with IF (%) | PTMC without IF (%) | Total (%) | P-value |
|---------------------------------|------------------|----------------------|-----------|---------|
| Age (years)                     |                  |                      |           |         |
| Mean ± SD                       | 47.81±10.44      | 44.88±10.94          | 46.83±10.69 | 0.0331  |
| Range                           | 21-75            | 20-68                | 20-75     |         |
| <45                             | 120 (35.29)      | 80 (46.78)           | 200 (39.13) |         |
| ≥45                             | 220 (64.71)      | 91 (53.22)           | 311 (60.87) | 0.012   |
| Sex                             |                  |                      |           |         |
| Female                          | 278 (81.77)      | 125 (73.10)          | 403 (78.86) | 0.0236  |
| Male                            | 62 (18.23)       | 46 (26.9)            | 108 (21.14) |         |
| Calcification                   |                  |                      |           |         |
| Present                         | 261 (76.76)      | 137 (80.12)          | 398 (77.89) | 0.3889  |
| Absent                          | 79 (23.24)       | 34 (19.88)           | 113 (22.11) |         |
| TSH (mmol/l)                    |                  |                      |           | 0.863   |
| Mean ± SD                       | 1.76±1.49        | 1.78±1.343           | 1.77±1.44 |         |
| Range                           | 0.002-10.43      | 0.0020-7.61          | 0.002-10.44 |         |
| Tumor size (mm)                 |                  |                      |           | 0.0136  |
| Mean ± SD                       | 4.93±2.46        | 4.21±2.27            | 4.69±2.42 |         |
| Range                           | 0.5-10           | 0.6-9                | 0.5-10    |         |
| Diameter ≥5 mm                  | 201 (59.12)      | 82 (47.95)           | 283 (55.38) | 0.0166  |
| Diameter <5 mm                  | 139 (40.88)      | 89 (52.05)           | 228 (44.62) |         |
| Multiplicity                    |                  |                      |           | 0.2595  |
| Absent                          | 262 (77.06)      | 124 (72.51)          | 386 (75.54) |         |
| Present                         | 78 (22.94)       | 47 (27.49)           | 125 (24.46) |         |
| Bilaterality                    |                  |                      |           | 0.6463  |
| Absent                          | 301 (88.53)      | 149 (87.13)          | 450 (88.06) |         |
| Present                         | 39 (11.47)       | 22 (12.87)           | 61 (11.94) |         |
| ETE                             |                  |                      |           | 0.1322  |
| Absent                          | 290 (85.29)      | 154 (90.06)          | 444 (86.89) |         |
| Present                         | 50 (14.71)       | 17 (9.94)            | 67 (39.18) |         |
| Subtype                         |                  |                      |           | 0.7377  |
| Classic papillary               | 326 (95.88)      | 165 (96.49)          | 491 (96.07) |         |
| Follicular                      | 14 (4.12)        | 6 (3.51)             | 20 (3.93) |         |
| Lymph metastasis                |                  |                      |           | 0.0003  |
| LM present                      | 116 (34.18)      | 32 (18.71)           | 148 (28.96) |         |
| LM absent                       | 224 (65.72)      | 139 (81.29)          | 363 (65.17) |         |
| Lymph node status               |                  |                      |           | 0.0058  |
| N0                              | 226 (66.47)      | 137 (80.12)          | 363 (71.04) |         |
| N1a                             | 74 (21.76)       | 22 (12.87)           | 96 (18.78) |         |
| N1b                             | 40 (11.77)       | 12 (7.02)            | 52 (30.41) |         |
| TNM stage                       |                  |                      |           | 0.0067  |
| I                               | 278 (81.76)      | 159 (92.98)          | 437 (85.52) |         |
| III                             | 37 (10.88)       | 8 (4.68)             | 45 (8.81) |         |
| IV                              | 21 (5.33)        | 4 (2.34)             | 25 (4.89) |         |

PTMC, papillary thyroid microcarcinoma; TSH, thyroid-stimulating hormone; ETE, extrathyroidal extension; TNM, tumor-node-metastasis; SD, standard deviation.
presented with IF. The median tumor size was 5.0 mm (range, 1.0-10 mm). Using the median size as the cut-off point, a tumor ≥5.0 mm was revealed to be associated with a significantly increased possibility of fibrotic formation compared with PTMC without IF (P=0.017). Fibrosis was revealed to be significantly associated with lymph node metastasis.
However, there was no significant association between calcification, serum TSH levels, tumor multiplicity, bilaterality, ETE or subtype of PTMC and IF.

**Association of IF with pathological biomarkers.** Common biomarkers were observed and analyzed to determine whether they were associated with the presence of IF in tumors. Galectin-3 and CK19 revealed a significant association with IF (P=0.022 and P=0.008, respectively; Table II). There were no other significant associations observed between the presence of Tg, TTF-1, p53 or Ki67 and the cases of IF.

**Results of survival data.** Due to the decreased mortality rate of patients with PTMC, and since only 4 patients suffered mortality during the follow-up period, recurrence-free survival was used as the outcome of interest. A lack of recurrence outcome due to a lack of follow-up, loss to the follow-up period and non-thyroid-cancer mortality was defined as censored data. The censoring rate was 93.0 and 81.2% in the IF and no IF groups, respectively. The number of loss to follow-up period patients was 33 in the IF group and 52 in the no IF group. Each group had 1 patient who suffered from non-thyroid-cancer mortality. The median follow-up time was 46 months (range, 5.0-90 months) and the total recurrence rate was 14.87% (76 cases). The fibrosis group demonstrated an 18.82% (64 cases) recurrence rate and the no-fibrosis group had a 7.02% (12 cases) recurrence rate. Multivariate analysis by Cox's proportional hazard model revealed that the presence of IF (hazard ratio = 2.18195%; confidence interval = 1.163-4.090; P=0.015) was an independent factor for the prediction of a poor prognosis in patients with PTMC (Fig. 2).

Kaplan-Meier survival curves revealed the same outcome (log rank, P=0.002) (Fig. 3). At 32 months, the two groups became significantly different as analyzed by Cox's proportional hazard model (hazard ratio = 4.23795%; confidence interval = 1.287-13.950; P=0.018). Other poor prognostic indicators as presented in Table III were age (>45 years; P=0.49), extrathyroidal extension (P<0.001) and lymphocytic metastasis (P<0.001). Therefore, IF is a poor indicator of prognosis of PTMC.

**Discussion**

Cancer-associated fibroblasts have been described in multiple types of cancer; they appear primarily as abnormal IF and have been extensively studied (27-30). Fibrotic density, when there are no superior clear parameters, may be considered as a vital index of prognosis (24), however, few previous studies have focused on the correlation between IF and PTMC (31) and to the best of our knowledge the association between fibrosis and specific biomarkers has not been previously reported.

IF has not been clearly defined within the medical community. In general, IF is considered to occupy the central section of the tumor and often consists of radiating and expanding fibrous bands (32). Rebecchini et al (33) revealed that a characteristic feature of PTC was that interstitial fibers occupied 40-60% of the central section of the tumors. In a study performed by Isarangkul (24), 33 of 37 PTC specimens

| Variable          | Odds ratio | 95% confidence interval | P-value |
|-------------------|------------|-------------------------|---------|
| Sex               | 1.656      | 1.001-2.739             | 0.049   |
| Age (>45 years)   | 0.483      | 0.198-1.180             | 0.110   |
| Multiplicity      | 1.758      | 0.628-4.922             | 0.283   |
| Bilaterality      | 3.735      | 2.261-6.169             | <0.001  |
| ETE               | 1.289      | 0.793-2.094             | 0.305   |
| IF                | 2.227      | 1.186-4.181             | 0.013   |
| Lymph metastasis  | 3.560      | 2.164-5.858             | <0.001  |

PTMC, papillary thyroid microcarcinoma; ETE, extrathyroidal extension; IF, interstitial fibrosis.
presented interstitial changes consisting of dense collagen and fibroblasts. It was suggested that PTC was significantly associated with fibrosis when compared with the follicular subtype (odds ratio = 37.95; P<0.01). It is clear that there are correlations between tumorigenesis and fibrosis in many tumors, including breast, ovarian, esophageal, colorectal, pancreatic and prostate cancers (16-20). Interstitial changes influence tumor infiltration, metastasis, prognosis and neovascularization (34). In the present study observations under a light microscope demonstrated that the majority of IF cases were accompanied by an incomplete fibrotic boundary of the tumor, as with the fibrosis from the central section of the tumor, which disturbs the formation of the fiber encapsulation on the tumor. It was also revealed that IF is a poor indicator of prognosis; therefore, the formation of fibrotic lesions may be referred to as a more aggressive behavior of the tumor.

In the present study, tumors with fibrosis were significantly associated with a patient's age (P=0.0331) and tumor size (P=0.0166), as previously reported (35). Female patients were more likely to have fibrotic changes compared with male patients. Whether an estrogen receptor (ER) wound contributed to this change requires further study. It has been previously reported that fibrotic focus has an association with ERs in breast cancer (34).

Although certain previous studies have considered that the grade of IF may predict tumor recurrence and metastasis (34), there is little evidence for this in thyroid carcinoma. However, De Matos et al (36) detected the expression of galectin-3 in three types of thyroid tissue, the results of which revealed that the expression ratios in 84 cases of PTC and 38 follicular carcinoma cases were 72.6 and 21.0%, respectively, whereas it was nearly zero in non-tumor cases. Additionally, galectin-3 is considered as serving a key role in cellular adhesion and interactions between cells and their matrix, which may reflect tumor metastasis conditions (37). Certain previous studies have reported that galectin-3 may be viewed as a potential tumor marker of metastasis and poor prognosis (3). In the present study, it was revealed that a higher expression level of galectin-3 was significantly associated with fibrosis formation. Therefore, patients who are galectin-3 positive with PTMC with IF may be considered as evidence that the formation of IF predicts prognosis, alternatively IF formation may be as a result of the expression of galectin-3. Given the difficulties of diagnosing thyroid tumors, specimens stained to determine IF and the expression of galectin-3 may increase the sensitivity and specificity of a diagnosis and the prediction of a prognosis, particularly when PTMC is determined.

It was previously reported that the expression of CK19 was decreased in normal thyroid tissue; however, CK19 may be viewed as a vital biomarker for cancer when carcinoma cells of PTMC are positive, compared with normal thyroid tissue (38). Although there has been no evidence that CK-19 is a predictor of prognosis, the association between IF and CK19 is positive in the present study, it also can be speculated that the IF has an effect on the PTMC, additional studies are required to determine the association between IF and CK19.

To conclude, it was revealed that there is an association between IF and PTMC, how the underlying mechanism behind this association remains unclear. The present study demonstrated that IF combined with specific biomarkers may be an important diagnostic tool for PTMC. Additional studies are required to determine the course of the development of IF and whether it is associated with tumorigenesis and prognosis.

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Cancer-associated fibroblasts neutralize β2+ MicroRNA-26a and -26b inhibit lens fibrosis

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ONCOLOGY LETTERS 15: 4937-4943, 2018

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