Melissopalynological Characterization of Honey Samples from Southeastern, Nigeria

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The identification of plants in which the bees forage is key in establishing bee farms and increasing honey production. In this study pollen analysis of honey samples from the southeastern part of Nigeria was carried out to ascertain their floral sources and ecological origin. The honey samples were acetolyzed and microscopically studied to determine the pollen types. A total of seventy-one pollen types belonging to forty-one families of plants were identified. The honey samples were dominated by pollen grains from the families of Arecaceae, Euphorbiaceae, Myrtaceae, Irvingiaceae, Fabaceae, Combretaceae/Melastomataceae, and phyllanthaceae. Some of the dominant pollen grain identified include Elaeis guineensis, Alchornea cordifolia, Hymenocardia acida, Ocimum gratissimum, Syzygium guineense, Nauclea latifolia, and Afzelia africana. Out of the six samples studied, Njokoka sample was monofloral having Mimoso pigra as predominant pollen while Ayamelemu, Ekwusigo, Nsukka, Ezeagu, and Udenu samples are multifloral containing Elaeis guineensis, Phyllanthus sp., Pilostigma reticulatum, Irvingia sp., Alchornea cordifolia, and Lannea sp. as major secondary pollen. All the samples analyzed have Elaeis guineensis in common except Ezeagu, indicating that these plants are present in all five locations. These results can also be used as a tool in the geographical identification of Southeastern Nigeria honey from other geopolitical zones.

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Introduction

The practice of beekeeping in the production of honey is an old agricultural system in southeastern Nigeria, it has helped in improving the standard of living of people both in rural and urban areas. Although, pests and diseases have been reported to cause a about 15% decline in honey bee colony establishment, as well as the regular absconding and aggressiveness of the honeybees (Adekanmbi et al., 2019). Honey’s medicinal, therapeutics, and nutritional properties have made it a sought after commodity both in domestic and international markets, by providing employment and room for the adulteration of honey through the demand and supply gap. There is a need for honey products to be subjected to these parameters; floral type, precise place of origin, and quality to check their authenticity in Nigeria. Most markets sell adulterated honey Made of brown sugar, which could affect our health and increase the sugar level in humans.

The composition of honey varies according to the source of flowers used by bees, the harvest period, and the geo-climatic conditions of the regions concerned (Yédomonhan, 2009). Studies of pollen analysis will assist in bee management and in the development of beekeeping. It provides reliable information on floral and geographical sources of honey along with the relative preference of the bees among the diverse assemblage of plant species flowering at the same period. The wind and insect-pollinated taxa found in a honey sample will often produce a pollen spectrum that is unique for the specific geographical region where it was produced. Pollen analysis of honey has important commercial value because honey made from some plants commands a premium price.

According to Ige and Obasanmi, (2014), analysis of pollen in honey dates as far back as the nineteenth century, starting with the pioneering work of Pfister (1895). Since this period, a lot of studies (Agwu and Njokuocha, 2004; Atanassova et al., 2004; Fagundez and Caccavarl, 2006; Sadia et al., 2008; Adekanmbi and Ogundipe, 2009; Forcone, 2014; Moar; 2014; Aino, 2016; Njokuocha et al.,...
2019) have been carried out by several researchers around the world to examine the pollen contents of honey from various countries. In Nigeria, available literature on the pollen contents of honey from various parts of Nigeria has all revealed the floral sources utilized by bees in honey production. Fifty-six (56) honey plants which were characteristic flora of tropical rainforest and mosaic of Low land rainforest taxa were recorded by Nnamani and Uguru (2013) from the study of honey samples collected from Southern Nigeria. Emuobosa (2017) recorded various honey plants from the comparative study of the pollen content of honey collected from the apiary and open markets in Nigeria and the Bénin republic. Similarly, Njokuocha et al. (2019) determined the pollen spectrum of Apis mellifera honey from different locations in Nigeria.

However, in Enugu and Anambra state where honey production is a popular business in many communities, apart from the works of (Agwu et al., 1989; Agwu and Abaee, 1991; Njokuocha and Nnamani, 2009; Njokuocha and Ekweozor, 2007; Njokuocha and Osayi, 2015; Njokuocha et al., 2019) on the pollen content of Nsukka honey, reliable information on floral sources of honey produced in this area until now are limited. These studies help to differentiate monofloral honey from multiflora and specific types of honey which are of high commercial value. The aim of this research, therefore, was to examine the pollen grains contained in honey from these States to provide more information on the botanical and geographical origins of the honey.

Materials and Methods

**Honey Sample Collection**

The honey samples used were collected from the beekeepers in the Ayamelum, Ekwusigo, Ezeagu, Njikoka, Nsukka, and Udenu in Anambra and Enugu States, South-eastern, Nigeria (Figure 1).

**Results and Discussion**

The detailed pollen record of each of the samples is presented in (Table 1a,b). Microscopic examination of honey samples from Anambra and Enugu revealed a total of twenty-seven thousand four hundred and seventy pollen grains (27,470). A total of 71 pollen types belonging to 41 families of plants were recorded in the honey samples. The identified species belong to varying genera of herbs, shrubs, grass, and trees. The colours of the samples after dilution were amber, golden yellow, and yellowish-brown for the honey samples collected from Ayamelum, Ekwusigo, and Njikoka Local Government Areas in Anambra State as well as dark amber, light brown, and amber for the samples from Ezeagu, Nsukka, and Udenu Local Government Areas in Enugu State (Table 2). Amongst all the samples analyzed, one was found to contain predominant pollen type (> 45%) in occurrence (Table 3a,b). In Anambra State, the honey sample collected from Njikoka Local Government Area was dominated by pollen of the *Mimosas pigra* (Table 4). All the honey samples from Enugu State were multi-floral honey (Table 4). Generally, *Elaeis guineensis* was present in all the honey samples. Also, present in the samples as secondary pollen types (16-45%) were *Lannea alchornea*, *Alchornea cordifolia*, *Phyllanthus sp.*, *Piliostigma reticulatum*, *Irvingia sp.*, *Syzygium guineense*, and Combretaceae/Melastomataceae. The pollen types were classified as predominant (>45%), secondary pollen (16-45%), important minor (3-15%), and minor (<3%) (Jones and Bryant, 2004).

**Pollination Analysis**

Polliniferous residue was mounted on glycerine jelly and observed under a compound microscope with 400X magnification. The pollen grains were identified with the help of descriptions and photomicrographs in books and Journals (Y’bert, 1979; Bonnefille and Riollet, 1980; Agwu and Akanbi, 1985; Gosling et al., 2013). They were also compared with reference slide collections in the Palynology Laboratory, Department of Plant Science and Biotechnology, University of Nigeria, Nsukka.

**Pollen Count**

The characterization of pollen was based on percentages of each pollen type: the pollen grains were placed into one of the following pollen frequency classes: Predominant (> 45% of the total pollen grains counted); Secondary (16% - 45%); Important Minor (3% - 15%) and Minor pollen types (<3%) (Jones and Bryant, 2004).
| Sn | Pollen types                      | Anambra State (%) | Enugu State (%) |
|----|-----------------------------------|-------------------|-----------------|
|    |                                   | Ayamelum | Njikoka | Ekwusigo | Nsukka | Eziagu | Udenu |
| 1  | Amaranthaceae                     | 0.2      | 0.4     | 0.4      | 1.2    | -      | -     |
| 2  | Ampelidaceae                      | -        | -       | -        | -      | -      | -     |
| 3  | *Cissus doerignii* Gilg. & Brandt.| -        | 0.2     | -        | 0.3    | -      | -     |
| 4  | Anacardiaceae                     | -        | -       | -        | -      | -      | -     |
| 5  | *Anacardium occidentale* Linn.    | 0.2      | -       | 3.7      | -      | -      | -     |
| 6  | *Mangifera indica* Linn.          | -        | 1.3     | -        | 0.9    | 26.2   | -     |
| 7  | *Lannea sp.*                      | 1.9      | -       | -        | -      | -      | -     |
| 8  | Annonaceae                        | -        | -       | -        | -      | -      | -     |
| 9  | *Monodora sp.*                    | -        | -       | -        | -      | -      | -     |
| 10 | Apioaceae                         | -        | 0.6     | 0.4      | -      | -      | -     |
| 11 | Apocynaceae                       | -        | -       | -        | -      | -      | -     |
| 12 | Arecalesaceae                     | -        | -       | -        | -      | -      | -     |
| 13 | *Irvingia*                        | 23.3     | 35.1    | 26.3     | 41.0   | -      | 27.8  |
| 14 | *Elaeis guineensis* Jacq.         | -        | -       | -        | -      | -      | -     |
| 15 | *Asteraceae*                      | 1.7      | -       | 4.6      | 0.5    | -      | -     |
| 16 | Bombacaceae                       | -        | -       | -        | -      | -      | -     |
| 17 | *Bombax buonopozense* P. Beauv.   | 0.2      | -       | -        | -      | -      | -     |
| 18 | Boraginaceae                      | -        | -       | -        | -      | -      | -     |
| 19 | Cordia sp.                        | -        | 0.2     | -        | 0.9    | 0.3    | -     |
| 20 | Heliotropium indicum* L.          | 0.4      | -       | -        | -      | -      | -     |
| 21 | Burseraceae                       | -        | -       | -        | -      | -      | -     |
| 22 | *Canarium schweinfurthii* Engl.   | -        | 1.6     | -        | -      | -      | -     |
| 23 | *Commiphora* sp.                 | 9.0      | -       | -        | -      | -      | -     |
| 24 | *Capparidaceae*                   | 0.3      | -       | -        | -      | -      | -     |
| 25 | *Cadaba* sp.                     | -        | -       | -        | 0.6    | -      | -     |
| 26 | *Celastraceae*                    | -        | -       | -        | -      | -      | -     |
| 27 | *Hippocratea africana* (Wild.) Loes | 0.7      | -       | -        | -      | -      | -     |
| 28 | *Combretaceae/Melastomataceae*    | -        | -       | -        | -      | -      | -     |
| 29 | *Combretum* sp.                   | -        | -       | -        | -      | -      | -     |
| 30 | *Cyperaceae*                      | -        | -       | -        | -      | -      | -     |
| 31 | *Ebenaceae*                       | -        | -       | -        | -      | -      | -     |
| 32 | *Diospyros* sp.                   | 0.2      | -       | -        | -      | -      | -     |
| 33 | *Ephorbiaceae*                    | 2.7      | -       | -        | -      | -      | -     |
| 34 | *Acalypha* sp.                    | -        | -       | 1.4      | -      | -      | -     |
| 35 | *Achyroclinea cordifolia* (Shum. & Thonn.) Mull. Arg. | - | 4.8 | 25.3 | 1.8 | 9.0 | 23.6 |
| 36 | *Securinea virosa* (Rosb. Ex Wild.) Baill. | 3.8 | - | - | - | - | - |
| 37 | *Antidesma* sp.                   | -        | -       | -        | 0.3    | -      | -     |
| 38 | *Fabaceae*                        | -        | -       | -        | -      | -      | -     |
| 39 | *Caesalpinioideae*                | -        | -       | -        | -      | -      | -     |
| 40 | *Afzelia africana* Sm.            | 0.2      | -       | -        | -      | -      | -     |
| 41 | *Albizia* sp.                     | -        | -       | -        | 0.1    | -      | -     |
| 42 | *Cassia* sp.                      | 0.5      | -       | 0.3      | 0.4    | 4.2    | -     |
| 43 | *Delonix regia* (Boj. Ex Hook.) Raf. | - | - | - | 0.4 | 4.2 | - |
| 44 | *Dialium guineense* Wild.          | -        | -       | 0.3      | -      | -      | -     |
| 45 | *Piliostigma reticulatum* (Dc.) Hochst | 21.7 | - | - | - | - | - |
| 46 | *Mimosoidae*                      | -        | -       | -        | -      | -      | -     |
| 47 | *Cleome* sp.                      | -        | -       | -        | 0.2    | -      | -     |
| 48 | *Mimosa Pigra* Linn.              | -        | 53.5    | -        | -      | -      | -     |
| 49 | *Faboidae*                        | -        | -       | -        | -      | -      | -     |
| 50 | *Crotalaria* pycnostachya Benth.   | -        | -       | 0.6      | -      | -      | -     |
| 51 | *Puerocarpus* sp.                 | -        | -       | -        | 0.4    | 0.4    | -     |
| 52 | *Hymenocardiae*                   | -        | -       | -        | -      | -      | -     |
| 53 | *Hymenocardia acida* Tul.         | -        | 0.3     | 0.2      | 7.9    | 13.0   | -     |
| 54 | *Irvingiaceae*                    | -        | -       | -        | -      | -      | -     |
| 55 | *Irvingia* sp.                    | 1.8      | 0.4     | 27.5     | -      | -      | -     |
| 56 | *Lamiaceae*                       | -        | -       | -        | -      | -      | -     |
| 57 | *Ocimum gratissimum* L.           | 6.3      | -       | -        | -      | -      | -     |
| 58 | *Liliaceae*                       | 4.5      | 1.0     | 0.3      | 0.3    | -      | -     |
| 59 | *Loganiaceae*                     | -        | -       | -        | -      | -      | -     |
| 60 | *Anthocelesia vogelii* Planch.    | -        | -       | -        | 0.3    | -      | -     |
| 61 | Loranthaceae                      | 0.2      | -       | -        | -      | -      | -     |
| 62 | *Meliaceae*                       | -        | -       | -        | -      | -      | -     |
| 63 | *Trichilia* sp.                   | 0.2      | -       | 0.4      | -      | -      | -     |
| 64 | *Khaya senegalensis* (Desr.) A Juss | - | - | - | 1.3 | - | - |
| 65 | *Moraceae*                        | 0.4      | 0.1     | 0.2      | 0.4    | -      | -     |
These are plants that are either cultivated or conserved specially for their economic benefits as commercial or subsistence crops in the areas where the honey was produced. This result agrees with the findings of Njokuocha et al. (2019).

The fact that the samples were collected from the wild gives the honey bee access to many plant species which may have contributed to the high pollen diversity. Ige and Apo (2007), are of the view that the more the source of nectar/pollen available to the bees for collection, the more pollen type and this automatically means the more richness of the honey. The age of the bees is also an important factor when it comes to pollen diversity and abundance. Adeonipekun (2012), observed that an old and defensive colony of bees recorded a higher abundance of pollen grains, while a young colony recorded lesser pollen grains but had higher diversity a reflection of the difference in their experience and nature.

These honey samples generally rich in pollen displayed a vivid landscape of the bee foraged plants growing in the area where the honey samples were collected. Even though bees are species-specific, they still collect pollen from readily available flowers. The pollen types came from nectariferous and non-nectariferous plants.

The dominance of *Elaeis guineensis* pollen in Ayamelum, Njikoka, Ekwusigo, Nsukka, and Udenu honey samples means that the oil palm is readily available and these could be attributed to the fact that the palm is used in commercial agriculture in the production of palm oil. It is a non-nectariferous plant, but the honey bees’ feeds on the juice of their fruits in the absence of nectar may be due to their high-calorie level. It also serves as a haven for their preferred choice of nectar/pollen and this may have contributed to the high pollen diversity. Adeonipekun (2012), observed that an old and defensive colony of bees recorded a higher abundance of pollen grains, while a young colony recorded lesser pollen grains but had higher diversity a reflection of the difference in their experience and nature.

In Ayamelum honey samples *Piliostigma reticulatum* was also found to be dominant, but they were absent in Njikoka, Ekwusigo, Nsukka, Ezeagu, and Udenu samples (Figure 2), there is an indication of its shelterbelt in homesteads when in full foliage, which serves as a haven for bees.

![Table 1b. Percentage composition of honey samples collected from Anambra and Enugu States](image-url)
Table 2. The weight of pollen sediment and colour of the honey samples after dilution

| S.n. | Sample    | Colour           | Weight |
|------|-----------|------------------|--------|
| 1    | Ayamelum  | Amber            | 2.56   |
| 2    | Ekwsigo   | Golden yellow    | 1.88   |
| 3    | Njikoka   | Yellowish brown  | 3.45   |
| 4    | Ezeagu    | Dark amber       | 4.23   |
| 5    | Nsukka    | Light brown      | 4.44   |
| 6    | Udenu     | Amber            | 1.91   |

Table 3a. Frequency class of pollen types in the honey samples

| Sn | Pollen types                          | Anambra State | Enugu State |
|----|---------------------------------------|---------------|-------------|
|    |                                       | Ayamelum (%)  | Ejisu (%)   |
|    |                                       | Njikoka (%)   | Ekwsigo (%) |
|    |                                       | Nsukka (%)    | Ezeagu (%)  |
| 1  | Amaranthaceae                         | M             | M           |
| 2  | Ampelidaceae                          | -             | -           |
| 3  | Cissus doeringii Gilg. and Brandt.    | -             | M           |
|    | Anacardiaceae                         | -             | M           |
|    | Anacardium occidentale Linn.          | M             | -           |
|    | Mangifera indica Linn.                | -             | IM          |
|    | Lannea sp.                            | S             | S           |
|    | Spondias mombin Linn.                 | M             | -           |
| 4  | Annonaceae                            | -             | -           |
| 5  | Monodora sp.                          | -             | M           |
| 6  | Apicaceae                             | -             | M           |
| 7  | Areaceae                              | -             | -           |
|    | Elaeis guineensis Jacq.               | S             | S           |
|    | Asteraceae                            | M             | IM          |
|    | Bombacaceae                           | -             | -           |
|    | Bombax buonopozense P. Beav.          | M             | -           |
|    | Boraginaceae                          | -             | -           |
|    | Cordia sp.                            | -             | M           |
|    | Heliotropium indicum L.               | M             | -           |
|    | Burseraceae                           | -             | -           |
|    | Canarium schweinfurthii Engl.         | -             | M           |
|    | Commiphora sp.                        | M             | -           |
| 12 | Capparidaceae                         | M             | -           |
| 13 | Gadabu sp.                            | -             | -           |
|    | Hippocratea africana (Wild.) Loes     | M             | -           |
|    | Combretaceae/Melastomataceae          | -             | M           |
|    | Combretum sp.                         | -             | -           |
|    | Cyperaceae                            | -             | M           |
|    | Ebenaceae                             | -             | -           |
|    | Diospyros sp.                         | M             | -           |
|    | Ephrbiaceae                           | M             | -           |
|    | Acalypha sp.                          | -             | -           |
|    | Alchornea cordifolia (Shum. & Thonn) Mull. Arg. | IM | S | M | IM | S  |
|    | Securinega virosa (Rostb. Ex Wild.) Baill. | IM | - | - | - | -  |
|    | Antidesma sp.                         | IM | - | - | - | -  |
| 14 | Fabaceae                              | -             | -           |
|    | Caesalpinioideae                      | -             | -           |
|    | Afzelia africana Sm.                  | M             | -           |
|    | Albizia sp.                           | -             | -           |
|    | Cassia sp.                            | M             | -           |
|    | Delonix regia (Boj. Ex Hook.) Raf.    | -             | -           |
|    | Dialium guineense Wild.               | M             | -           |
|    | Pilostigma reticulatum (Dr.) Hochst.  | S             | -           |
|    | Mimmosoideae                          | -             | -           |
|    | Cleome sp.                            | -             | -           |
|    | Mimoso Pigna Linn.                    | P             | -           |
|    | Faboideae                             | -             | -           |
|    | Croton alismifolia                    | -             | -           |
|    | Pterocarpus sp.                       | -             | M           |
| 19 | Hymenocardieae                        | -             | -           |
|    | Hymenocardia acida Tul.               | M             | M           |
| 20 | Irvingiaceae                          | M             | M           |
|    | Irvingia sp.                          | M             | S           |
Table 3b. Frequency class of pollen types in the honey samples

| Sn | Pollen types | Anambra State (%) | Enugu State (%) |
|----|--------------|--------------------|----------------|
| 21 | Lamiaceae    | -                  | -              |
|    | Ocimum gratissimum L | IM                | -              |
| 22 | Liliaceae    | IM M - - - - IM M - |
| 23 | Loganiaceae  | -                  | -              |
|    | Anthocleista vogelii Planch. | - | - |
| 24 | Loranthaceae | -                  | -              |
| 25 | Meliaceae    | -                  | -              |
|    | Trichilia sp. | M - - M - - M - |
|    | Khaya senegalensis (Desr.) A. Juss | - | - |
| 26 | Moraceae     | M M M M M -       |
| 27 | Myrtaceae    | -                  | -              |
|    | Syzygium guineense Engl. | - M IM S M IM |
|    | Psidium guajava L. | - - M - |
| 28 | Ochnaceae    | -                  | -              |
|    | Lophira lanceolata Van Tiegh. Ex Keay | - | - |
| 29 | Passifloraceae | M - - - - - - |
| 30 | Phyllanthaceae | M - - M - - S - |
|    | Phyllanthus sp. | - M - - S - |
|    | Bridelia ferruginea Benth. | IM - - M M - |
| 31 | Poaceae      | M M M M IM -       |
| 32 | Proteaceae   | -                  | -              |
|    | Protea angolensis Welw. | - - - - M - |
| 33 | Rhamnaceae   | -                  | -              |
|    | Ziziphus sp. | -                  | -              |
| 34 | Rubiaceae    | -                  | -              |
|    | Nauclea latifolia Sm. | IM - M - M - |
|    | Crossospermyx febrifuga (Afzel. Ex G. Don) Benth. | - - - - S - |
|    | Mussaenda erythrophylla Schum & Thonn. | - IM M - |
| 35 | Rutaceae     | -                  | -              |
|    | Fagara xanthoxyloides (Lam.) | - - M - - |
| 36 | Sapindaceae  | -                  | -              |
|    | Blihia sapind Konig | M - - - - |
|    | Paulinia pinnata Linn. | M - - - - |
| 37 | Sapotaceae   | -                  | -              |
|    | Mimosop andogensis Hiern. | - - - M - |
| 38 | Scrophulariaceae | - - - - - - |
| 39 | Sterculiaceae | -                  | -              |
|    | Sterculia tragacantha Lindl. | - - - - M - |
| 40 | Solanaceae   | -                  | -              |
|    | Solanum sp.  | -                  | -              |
|    | Ulmacea      | -                  | -              |
|    | Celtis sp.   | -                  | -              |

M: Minor, S: Secondary, IM: Important minor, P: Predominant

Apart from their medicinal properties, they are also prolific producers of pollen grains which makes the bees seek after them. The extant plants of *Mimosas pigra* were found to be dominant in the Njikoka community due favourably weather conditions, they are an important pollen source for *Apis* foragers since they are prolific producers of pollen grains (Figure 2). Their pollen concentration in the honey sample was very high due to the proximity between the beehives and where the plant was growing.

Pollens of *Irvingia* sp and *Ocimum gratissimum* was dominant in the honey sample from Ekwusigo and Ayamelum, these an indication that the bees foraged cultivated areas within the forest zone. These plants could only be found in farmland or conserved area. *Alchornea cordifolia* was also found to be dominant in Ekwusigo, Ezeagu, Njikoka, and Udenu Honey samples which is indicative of forest regrowth.

*S. guineense* pollen was slightly dominant in Ekwusigo, Nsukka, and Udenu Honey samples. While Asteraceae and Poaceae were dominant in honey samples from Ezeagu and Nsukka. This may be an indication of an increase in deforestation and expansion of agricultural landscapes which promoted increase and extension in agricultural weeds. Pollen of *Phyllanthus* sp., *Combretaceae/Melastomataceae*, *Lannea* sp. and *Crossospermyx febrifuga* were found to be common in Ezeagu and Udenu honey samples. Since these plants are prolific producers of pollen and would reduce competition during the foraging period among bees when its flowering makes them a preferred plant for foraging.

These pollen types are comparable to the ones identified in the present study. The pollen from both wind and insect-pollinated taxa present in a honey sample will often create a pollen spectrum that is unique for the specific geographical region or micro-vegetation area where it was produced (Ige and Obasanmi, 2014). Honey sample from Ayamelum,
Ekwusigo, Nsukka, Ezeagu, Udenu were classified as multiflora as dominant pollen types were within (6.3-23.3%), (6.9-27.5%), (4.2-41.0%), (9-24.4%), (4.7-27.8%) respectively. While, pollen types from Njikoka honey sample were found within (4.8-53.5%), which was classified as monofloral with *Mimosa pigra* dominating the sample with a record high of 53.5% (Table 4). The pollen spectrum revealed the common plant species foraged by the honey bees in this zone for pollen and nectar. Generally, the honey samples were dominated by plant species that reflect the forest savannah mosaic of vegetation.

### Table 4. Predominant pollen types, percentage occurrence and classification of honey samples collected from Anambra and Enugu States

| Location  | Class of honey | Pollen types               | Percentage occurrence (%) |
|-----------|----------------|-----------------------------|----------------------------|
| Ayamelum  | Multi floral   | *Elaeis guineensis* Jacq.   | 23.3                       |
|           |                | *Piliostigma reticulatum* (Dc.) Hochst | 21.7                      |
|           |                | *Bridelia ferruginea* Benth. | 10.3                      |
|           |                | *Commiphora* sp.            | 9.0                       |
|           |                | *Nauclea latifolia* Sm.     | 7.3                       |
|           |                | *Ocimum gratissimum* L      | 6.3                       |
| Njikoka   | Monofloral     | *Elaeis guineensis* Jacq.   | 35.1                      |
|           |                | *Mimosa pigra*              | 53.5                      |
|           |                | *Alchornea cordifolia* (Shum. & Thonn) Mull. Arg. | 4.8                      |
| Ekwusigo  | Multi floral   | *Elaeis guineensis* Jacq.   | 26.3                      |
|           |                | *Irvingia* sp.              | 27.5                      |
|           |                | *Alchornea cordifolia* (Shum. & Thonn) Mull. Arg. | 25.3                      |
|           |                | *Syzygium guineense* Engl.  | 6.9                       |
| Nsukka    | Multi floral   | *Elaeis guineensis* Jacq.   | 41.0                      |
|           |                | *Syzygium guineense* Engl.  | 18.0                      |
|           |                | *Hymenocardia acida*Tul.    | 7.9                       |
|           |                | *Asteraceae*                | 4.6                       |
|           |                | *Sapindaceae*               | 4.2                       |
| Ezeagu    | Multi floral   | *Phyllanthus* sp.           | 24.4                      |
|           |                | *Combretaceae/Melastomataceae* | 19                       |
|           |                | *Crossopteryx febrifuga* (Afzel. Ex G. Don) Benth. | 16.7                      |
|           |                | *Hymenocardia acida*Tul.    | 13.0                      |
|           |                | *Alchornea cordifolia* (Shum. & Thonn.) Mull. Arg. | 4.2                       |
|           |                | *Poaceae*                   | 9.0                       |
| Udenu     | Multi floral   | *Elaeis guineensis* Jacq.   | 27.8                      |
|           |                | *Lannea* sp.                | 26.2                      |
|           |                | *Alchornea cordifolia* (Shum. & Thonn.) Mull. Arg. | 23.6                      |
|           |                | *Syzygium guineense* Engl.  | 8.9                       |
|           |                | *Ziziphus* sp.              | 4.7                       |

**Conclusion**

The pollen spectra of the honey samples revealed the plants utilized by the bees for honey production, which indicates that the honeys were pure and not adulterated and provides possibility of utilizing this rich bee flora of the region for the development of apiculture and increased honey production of Nigeria. The pollen contents of studied honeys revealed the characteristic floristic composition of the ecological regions of the source areas.

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![Figure 2](image-url)
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