ARLTS1 polymorphisms and basal cell carcinoma of the skin

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Abstract

Polymorphisms in the ARLTS1 gene, a member of the Ras super-family, have been associated with susceptibility in different cancer types. The involvement of the gene in apoptotic signalling motivated us to study the role of ARLTS1 polymorphic variations in basal cell carcinoma of the skin (BCC). In a case-control study, 529 cases diagnosed with BCC and 533 controls from Hungary, Romania and Slovakia were genotyped for the S99S (297G>A), P131L (392C>T), L132L (396G>C), C148R (442T>C) and W149X (446G>A) polymorphisms in the ARLTS1 gene. No significant association between any of the single nucleotide polymorphisms (SNP) and risk of BCC (S99S, odds ratio (OR) 0.96, 95% confidence interval (CI) 0.60-1.53; P131L, OR 1.31 95%CI 0.74-2.31; L132L, OR 0.50, 95%CI 0.02-7.07; C148R, OR 0.50, 95%CI 0.69-1.18; and W149X, OR 1.01, 95%CI 0.37-2.79) was detected. Furthermore, no significant difference in the distribution of haplotypes due to five polymorphisms in the ARLTS1 gene was found between the BCC cases and controls. Our data rule out an association between variants in ARLTS1 and risk of BCC in the investigated population.

Introduction

Basal cell carcinoma (BCC) is the most common skin neoplasm, accounting for over 75% of all skin cancers [1, 2]. Though BCC occurs mainly sporadically, multiple tumours with an early onset are associated with rare genetic disorders like Gorlin syndrome and xeroderma pigmentosum [3]. BCC tumours grow slowly and seldom metastasize; the recurrence and tissue destruction cause extensive morbidity [4]. UV radiation is a major environmental factor associated with increased incidence of BCC due to increased leisure sun exposure [5, 6]. In specific instances arsenic exposure has also been associated with increased risk of non-melanoma skin cancers, including BCC [7]. The aetiology of BCC ultimately involves an interplay between genetic and environmental factors like UV radiation.

BCC mainly originates through UV-mediated DNA damage. The consequent mutations in the critical genes result in growth advantage to the affected cells leading to clonal expansion. Mechanistically apoptosis prevents carcinogenesis by eliminating cells with excessive damage that escape the DNA repair machinery [8]. We hypothesized that variants in the genes involved in apoptosis can potentially modulate disease susceptibility. Genetic variants in critical genes can potentially modify susceptibility to BCC through subtle influences on critical cellular processes like DNA repair, cell cycle and apoptosis.
ADP-ribosylation factor-like tumour suppressor gene 1 (ARLTS1) is a member of the Ras superfamily, involved in apoptotic signalling. Previously this gene has been suggested to be associated with susceptibility of multiple cancer types [9]. Therefore in this study we investigated the role of variants in the ARLTS1 gene on BCC susceptibility in cases and matching controls recruited from areas of Hungary, Romania and Slovakia.

Materials and methods

Study population: Cases and controls were recruited as part of a large study on risk of various cancers due to environmental exposures in Hungary, Romania and Slovakia between 2002 and 2004. The recruitment was carried out in the counties of Bacs, Bekes, Csongrad and Jasz-Nagykun-Szolnok in Hungary; Bihor and Arad in Romania; and Banska Bistrica and Nitra in Slovakia. 529 skin cancer cases, mean age 63.5 (median 66; range 2-85), were invited on the basis of histopathological examinations by pathologists. 533 hospital-based controls, mean age 60 (median 61; range 28-82), were included in the study, subject to fulfilment of a set of criteria. Those controls included general surgery, orthopaedic and trauma patients with conditions such as appendicitis, abdominal hernias, duodenal ulcers, cholelithiasis and fractures; patients with malignant tumours, diabetes and cardiovascular diseases were excluded. They were also broadly matched with cases for age, gender, country of residence and ethnicity.

Genotyping

DNA was isolated from blood samples from cases and controls using Qiagen mini-preparation kits and genotyped for variants in the ARLTS1 gene by direct DNA sequencing. A 237-bp product containing exon 2 of the ARLTS1 gene was amplified and sequenced bidirectionally using primers 5'-GAT ATC CTC GTG TAC GTG CTG (forward) and 5'-GAG CAA AGA TAT GCT GCT CTG T (reverse). The conditions used for PCR and sequencing reactions were as described previously [10]. The sequencing reaction products were analyzed on an ABI prism 3100 genetic analyzer and primary sequence data checked using sequence analysis software (Applied Biosystem, Forster City, CA).

Statistical Analysis

Genotype frequencies in cases and controls for all SNPs were tested for deviation from the Hardy-Weinberg equilibrium using chi^2 or Fisher's exact test. Age, gender and nationality adjusted odds ratio (OR), 95% confidence intervals (CI) for risk of BCC associated with each genotype and variant allele of five SNPs were calculated with logistic regression using SAS version 9.1 (SAS Institute, Cary, NC). Relation between genotype and BCC risk were summarized as global P-values. Linkage disequilibrium was calculated with Haploview software ([www.broad.mit.edu/mpg/haplovieview/documentation.php](http://www.broad.mit.edu/mpg/haplovieview/documentation.php)). Power calculation was carried out using Power and Sample Size calculation software version 2.1.31.

Results

The genotyping of a 237 bp-fragment of exon 2 of the ARLTS1 resulted in detection of 5 single nucleotide polymorphisms in 529 BCC cases and 533 controls. The sample size of this study was sufficient to detect under the dominant model the effect of polymorphisms with a minor allele frequency higher than 0.2 and an associated OR equal to or higher than 1.5 with a power of 80%. Two SNPs (297G>A and 396G>C) were synonymous, 2 non-synonymous SNPs (392C>T and 442T>C) resulted in P131L and C148R amino acid changes, and the G446A SNP resulted in W149X truncating polymorphism. Genotype distribution for all five polymorphisms based on allele frequencies were in accordance with Hardy-Weinberg equilibrium. The minor allele frequencies for five polymorphisms ranged from 0.01 (396G>C) to 0.45 (442 T>C) (Table 1).

OR and corresponding 95% CI intervals associated with each genotype and variant allele are detailed in Table 1. For none of the five polymorphisms was any statistical significant difference observed for genotype and allele frequencies between BCC cases and controls. The analysis of resultant haplotypes due to 5 SNPs in the gene showed presence of 8 out of 32 possible haplotypes. In controls, however, only 4 haplotypes were detected. The major haplotype GCCGT consisted of common alleles of all 5 polymorphisms and together with GCCGG haplotype accounted for over 90 percent of all cases and controls. However, no differences in distribution of haplotypes between cases and controls were observed (Table 2). Linkage disequilibrium was observed between 392 C>T and 442T>C (D' = 1.0), and between 442T>C and 446G>A (D' = 1.0) (Fig. 1). However, the r^2 values for these linkages were only 0.02 and 0.007. The variant T-allele for the P131L (392C>T) polymorphism occurred only in the presence of one or two T-alleles of the C148R (442T>C) polymorphism. Similarly, A-allele for the W149X (446G>A) polymorphism was detected only in the presence of the T-allele of the C148T (442T>C) polymorphism. Consequently, the variant allele for the W149X (446G>A) polymorphism occurred together only with the common allele genotype of the P131L (392C>T) polymorphism.
Discussion

In this study we evaluated the association of polymorphisms in the ARLTS1 gene with modulation of BCC risk. Our results showed that none of the coding sequence polymorphisms in the ARLTS1 gene was associated with risk modulation of BCC. While data showed linkage disequilibrium between three of the polymorphisms, the resultant haplotypes did not show differential distribution between cases and controls. In the initial study the truncating W149X (446G>A) polymorphism in the ARLTS1 gene was reported to be associated with increased risk of familial breast cancer; in latter studies C148R (442T>C) polymorphism in the gene was also associated with marginal risk of familial breast cancer and melanoma [9-11]. At the same time

| Genotype/Allele | Cases* | Controls | OR (95%CI) | P-value |
|-----------------|--------|----------|------------|---------|
| S995 (297G>A)   |        |          |            |         |
| GG             | 487 (92.2) | 490 (91.9) | 1.0 (referent) |         |
| GA             | 41 (7.7) | 43 (8.1) | 1.1 (0.7-1.7) | 0.81** |
| G-allele | 1015 (96.1) | 1023 (96.0) | 1.0 (referent) |         |
| A-allele | 41 (3.9) | 43 (4.0) | 1.1 (0.7-1.6) | 0.81 |
| P131L (392C>T) |        |          |            |         |
| CC             | 496 (93.9) | 508 (95.3) | 1.0 (referent) |         |
| CT             | 32 (6.1) | 25 (4.7) | 1.3 (0.8-2.3) | 0.30 |
| C-allele | 1024 (97.0) | 1041 (97.7) | 1.0 (referent) |         |
| T-allele | 32 (3.0) | 25 (2.3) | 1.3 (0.8-2.3) | 0.31 |
| L132L (396G>C) |        |          |            |         |
| GG             | 527 (99.8) | 531 (99.6) | 1.0 (referent) |         |
| GC             | 1 (0.2) | 2 (0.4) | 0.4 (0.1-4.0) | 0.40 |
| G-allele | 1055 (99.9) | 1064 (99.8) | 1.0 (referent) |         |
| C-allele | 1 (0.1) | 2 (0.2) | 0.4 (0.1-4.0) | 0.40 |
| C148R (442T>C) |        |          |            |         |
| TT             | 165 (31.3) | 155 (29.1) | 1.0 (referent) |         |
| TC             | 250 (47.4) | 258 (48.4) | 0.9 (0.7-1.2) |         |
| CC             | 113 (21.0) | 120 (22.5) | 0.9 (0.6-1.2) | 0.64 |
| T-allele | 580 (54.9) | 568 (53.3) | 1.0 (referent) |         |
| C-allele | 476 (45.1) | 498 (46.7) | 0.9 (0.8-1.1) | 0.36 |
| W149X (446G>A) |        |          |            |         |
| GG             | 519 (98.3) | 524 (98.3) | 1.0 (referent) |         |
| GA             | 9 (1.7) | 9 (1.7) | 1.5 (0.6-4.0) | 0.41 |
| G-allele | 1047 (99.2) | 1057 (99.2) | 1.0 (referent) |         |
| A-allele | 9 (0.8) | 9 (0.8) | 1.5 (0.5-3.5) | 0.41 |

*The number of cases and controls included in the study was 529 and 533, respectively. The sequencing for genotyping failed for one sample among cases.

**Global P-values calculated from chi^2 test for genotype as a single event and allele effects were adjusted for age, gender and nationality.
no association was observed with risk of various sporadic cancers [12-14].

ARLTS1 is a member of the Ras superfamily and resides on chromosome 13q14, which is frequently deleted in a variety of haematopoietic and solid tumours [15, 16]. Promoter hypermethylation of ARLTS1 and consequent down regulation of expression has been reported in B-cell chronic lymphocytic leukaemia and lung cancers [9]. Moreover the restoration of ARLTS1 in negative ovarian cells has been reported to reduce tumorigenic potential in nude mice. Functionally, the W149X polymorphism has been shown to induce apoptosis in the infected cells at a significantly reduced level rather than complete ablation [17]. This was hypothesized as the potential reason for retention of a rather drastic polymorphism in the population and could probably also explain the lack of associations with many cancer types including the BCC in the present study.

In conclusion, our data show lack of an association between polymorphisms and resultant haplotypes in the ARLTS1 gene and risk of BCC. The hypothesis of association of multiple cancer types with ARLTS1 variants is not supported by the data in this study.

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