Chronic lymphadenopathy caused by a Brazilian strain of *Bartonella henselae*

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**ABSTRACT**

*Bartonella henselae* is a relevant causative agent of bartonelloses in humans. We described an immunocompetent patient with clinical manifestation of chronic cervical lymphadenopathy after a cat-scratch in her forearm. This case shows *B. henselae* infection persistence even after prolonged antibiotic treatment.

**KEYWORDS:** Bartonella. Cat-scratch disease. Lymph nodes.

**INTRODUCTION**

*Bartonella* spp. are microorganisms that can cause diseases in humans and animals¹. The most involved species in human infections are *Bartonella bacilliformis*, *Bartonella quintana* and *Bartonella henselae*. Bartonellosis was formerly characterized as Carrión’s disease that is caused by *B. bacilliformis*. The species *B. quintana*, the agent of trench fever, was named for its hallmark five-day fever episodes during infection. Bacillary Angiomatosis (BA), Endocarditis (E), and Cat-Scratch Disease (CSD) are other classical bartonelloses caused by a different *Bartonella* spp., the *B. henselae* which was described in 1992 and may cause BA, E and CSD. This species is an important agent of bartonelloses in humans and have been related to many other clinical manifestations²,³.

In Brazil, seroprevalence rates for *B. henselae* can range from 13.7 to 34.4% and 38.4% in healthy and in HIV seropositive asymptomatic individuals, respectively⁴⁻⁶. *B. henselae* DNA has been detected in 10.6% and 4.5% of cats in the Southern and Northeastern regions of Brazil, respectively⁶⁻⁷. The lack of proper control of ectoparasites, especially cat fleas, and the close interactions between pets and humans may support the transmission of these pathogens to humans in Brazil⁸, as has been suggested in other countries⁵.

**CASE REPORT**

A 10-year-old female girl from the countryside of Caldas, Minas Gerais State (21°55’26”S 46°23’9”W), Southeast region of Brazil, was presented to her pediatrician with a two-month history of a non-painful right cervical nodule of 3 cm in diameter. She reported a fever of 38 ºC during the initial days. At the time of presentation, no fever, weight loss or other clinical signs were reported. Epidemiological data revealed that the patient had frequent contact with cats and, two months prior to presentation, she was scratched on the right forearm by a kitten from her farm. The
patient denied taking any prescription, over-the-counter medication or illicit drugs. The patient’s past medical and travel history were unremarkable. No other person in the household had similar clinical signs. Physical examination revealed an enlarged left cervical lymph node of 2 cm in diameter without armpit adenopathy. No other abnormalities were detected. A biopsy of her right mass revealed chronic granulomatous adenitis suggesting CSD. Tuberculosis test (PPD) and paracoccidioidomycosis serology were negative. She was properly treated for six weeks with 200 mg/day of doxycycline with lymph node regression.

Immunofluorescence assays (IFA) detected IgG titers of 1:256 and IgM titers of 1:64 for *B. henselae* and IgG titers of 1:256 and IgM titers of 1:32 for *Bartonella quintana* (IgG cut-off 1:64 and IgM 1:16 for both species). Two years later, she had a new right cervical lymph node enlargement episode with no other concurrent symptoms. No laboratory abnormalities were detected, including protein electrophoresis and immunoglobulin analysis. Doxycycline treatment was initiated at the same previously used dose, and clinical improvement of the lymphadenopathy was obtained after a complete six-week treatment.

Approximately four years after her initial presentation, the patient was referred to and hospitalized at the University of Campinas (UNICAMP) medical teaching hospital, State of São Paulo, Brazil, due to a new mass on the left cervical region. Physical examination revealed a 4 cm painless, soft, freely moveable mass on the left anterior cervical region of the patient. Cervical scars (Figure 1) from previous suppuration and biopsies were also evident. No other abnormalities were found during physical examination. Cell blood count and serum chemistry were unremarkable. A computed tomographic (CT) evaluation of the neck revealed anterior and posterior cervical lymphadenomegaly, with central necrosis and calcification, while CT evaluation of the abdomen exhibited mesenteric lymphadenomegaly and splenomegaly, while no thoracic lymphadenopathy was observed. IFA-IgG for *B. henselae* and *B. quintana* were positive at 1:128. Histopathology of the lymph node and surrounding skin indicated the presence of a chronic granulomatous inflammatory process and under special stains (Giemsa and Warthin-Starry) structures suggestive of *Bartonella* sp were found. A fine needle aspirate of the affected cervical lymph node was also performed. Standard and *Bartonella*-specific lymph node pus culture presented no isolate in solid agar supplemented with 5% sheep blood and grown in a 5% CO2 atmosphere. An aliquot of this material was submitted for Real Time PCR to test for the presence of *Bartonella* DNA using sequence-specific primers: 16s-23s rRNA internal transcribed spacer (ITS) region and phage-associated protein gene (pap31) followed by DNA sequencing for bacterial identification, as previously described10. The ITS and Pap31 DNA sequences amplified from the patient were 100% identical to four and six DNA sequences of the *B. henselae* strain Brazil-1 previously described in dogs, cats and a human from Brazil, respectively (Figure 2). A definitive diagnosis of chronic lymphadenopathy due to bartonellosis was undertaken. The patient was treated with intravenous gentamicin (160 mg/day) for two weeks and oral erythromycin (2 g/day) for six weeks. After having been discharged, she continued on azithromycin (500 mg/day) treatment for one year.

The patient has repeatedly sought medical attention due to spontaneous recurrent episodes of cervical lymphadenomegaly for the past two years with partial clinical resolution during new azithromycin/doxycycline treatments or even without treatments. Despite the prolonged antibiotic therapy, cervical lymphadenomegaly became persistent and has never completely regressed making the possibility of re-infections unlikely. Nowadays, she is 20 years old and is still followed up at the UNICAMP outpatient clinic because she maintains bilateral cervical lymph nodes enlargement. We have recently reported that, in the same geographic region of this case report, contact with cats is a major risk factor for *Bartonella* infection4. In this case report, we describe a patient with an initial clinical manifestation of chronic cervical lymphadenopathy after a cat-scratch in her forearm. Medical literature refers to patients with chronic lymphadenopathy as atypical CSD, but since the beginning of the clinical manifestation, she has had bilateral lymph nodes enlargement. Later, mesenteric lymphadenopathy was also documented, similar to previous reports as reviewed by Choi et al.11. Despite the early diagnosis of bartonellosis, the infection failed to resolve even upon prolonged and diverse antibiotic therapies, similar to a report by Simonton and Rupar12.

![Figure 1 - Right and left cervical regions of the neck, respectively, showing site scarring from suppuration and lymphadenopathy](image)
We detected *B. henselae* DNA, so that IFA positive results to *B. henselae* and *B. quintana* should represent cross-reactions in the serologic diagnosis as described before. The phylogenetic analysis indicated that this patient was infected with a strain of *B. henselae* different from the majority of strains that have been previously identified infecting humans in the United States, Europe and Brazil (Houston-1, SA2, Marseille, 90-615, URBHLIE 9, among others). The DNA fragments of the ITS region and Pap31 gene of *B. henselae* obtained from this patient showed 100% identity with the strain Brazil-1 (Figure 2), which has been described multiple times from infected cats and dogs in Brazil. Furthermore, a growing number of human cases in Brazil were shown to be infected with the Brazil-1 strain, including blood donors, suggesting that this strain may be endemic in that country. Nonetheless, the same DNA sequence of ITS and Pap31 present in this patient have also been reported in other *B. henselae* strains from cats, wild felids, and humans from the United States, Europe and Southeast Asia. Three of our *B. henselae* Brazil-1 isolates from cats have been deposited into the Instituto Adolfo Lutz Culture Collection and are available to the scientific community. Future genome-wide sequencing analysis will be able to further characterize the relationship of this isolate with other *B. henselae* strains reported in Brazil and in other countries around the world.

**CONCLUSIONS**

In summary, this report documents the detection of *B. henselae* Brazil-1 strain DNA from the lymph node of an immunocompetent patient with recurring bartonellosis episodes for several years that has persisted even after diverse antimicrobial therapies. Similar cases of recurring isolation of *Bartonella* sp. despite antibiotic therapy have also been reported in the veterinary literature. In our case, the patient history did not support the occurrence of frequent re-infection episodes, although we cannot exclude this possibility. Nonetheless, this case should raise the awareness of the medical community for the persistence of *Bartonella* sp. infections in humans.

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