Clear-cuts are temporary habitats, not matrix, for endangered grassland burnet moths (Zygaena spp.)

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Abstract
Burnet moths (Zygaena spp.) are day-flying Lepidoptera considered indicative of species-rich grasslands. In the present study, our aim was to clarify whether clear-cuts are habitat, supporting habitat or matrix for three species of Zygaena. We did so by sampling these species with sex pheromones on 48 clear-cuts, varying in amount of host and nectar plants, in southern Sweden. To compare the efficiency of such sampling, we also conducted transect walks on these clearcuts. Overall, host-plants on clear-cuts best explained the abundance of Zygaena spp. recorded, better than nectar-plants or connectivity with nearby grasslands. These results indicate that clear-cuts with an abundance of host plants are used as a fully functional habitat, and not a supporting habitat in the sense of only providing nectar. There is no support in these results for considering clear-cuts as an inert matrix. With about half the work-effort, pheromone traps recorded 100 times more Zygaena spp. as transect walks. The poor correspondence between observations during transects walks and pheromone trap catches suggest Zygaena spp. being difficult to monitor by transect walks. In contrast to grasslands, clear-cuts are short-term in nature requiring repeated recolonization, indicating the importance of permanent grasslands. However, clear-cuts are important temporary insect habitats due to their great acreage, and suitable management can increase the time they remain a habitat.

Keywords Clear-cut · Day-flying moth · Forestry · Landscape · Zygaena

Introduction
Traditional agricultural landscapes are among Europe’s most species-rich areas. Substantial areas were historically subject to fodder production, mowing or grazing, and these semi-natural areas are today biodiversity hotspots (Poschlod and WallisDeVries 2002; Habel et al. 2013). However, the rapid change in agriculture has led to abandonment of low-productive areas and more intensified use of other areas (Ihse 1995; Eriksson et al. 2002). As a consequence, a severe decline in species-rich grassland and biodiversity of agricultural areas has been repeatedly reported in Europe (Kearns et al. 1998; Krebs et al. 1999; Bengtsson et al. 2000; Maes and Van Dyck 2001; Robinson and Sutherland 2002; Shrub 2003; Foley et al. 2005).

A large proportion of European plants and insects prefer sun-exposed conditions and thrive in traditional agricultural landscapes (Ellenberg et al. 1991; Lindhe et al. 2005; Horák and Rébl 2013). An interesting debate is whether these sun-loving species assemblages were recruited from natural semi-open habitats created by large herbivores that are now extinct (e.g. Owen-Smith 1989; Vera 2000; van Vuure 2005; Feurdean et al. 2018; Ohwaki 2018), from grasslands originating from natural disturbances in the forests as floods, fires, storms or beavers (Ellenberg 1988; Pykälä 2000; Svenning 2002) or whether they have adapted to the features of the agricultural systems that evolved (Zopfi 1991, 1998; Lennartsson 1997). Whatever the origin, it has become increasingly clear that these species can sometimes
be supported by other habitats that surrounds remaining grazed semi-natural grassland fragments (e.g. Berg et al. 2016; Bušek and Reif 2017; Lampinen et al. 2018; Bergman et al. 2018), often referred to as a matrix. The matrix is often neglected in butterfly studies; in a systematic review studying fragmented landscapes, 60% of the papers excluded the matrix (Sweaney et al. 2014). However, the matrix may contribute to survival and persistence of populations by providing resources (Dennis et al. 2006; Shreeve and Dennis 2011) like food (Dennis 2004; Brady et al. 2011), facilitating dispersal between patches (Jauker et al. 2009; Kuefler et al. 2010) or by decreasing negative edge effects (Lindenmayer and Fisher 2006; Franklin and Lindenmayer 2009).

Several studies of grassland butterflies have shown that a forest matrix decreases the negative effects of fragmentation of grasslands (Bergman et al. 2008, 2018; Öckinger et al. 2012; Villeney et al. 2015). Forest edges and clear-cuts in temperate climates may provide adult Lepidoptera with nectar sources and favourable microclimate for larval development (Dennis et al. 2004; Kuusaari et al. 2007; Van Halder et al. 2010; Ibbe et al. 2011; Jonason et al. 2014; Blixt et al. 2015; Korpela et al. 2015; Viljur and Teder 2016; Ohwaki et al. 2018a). Together with more permanent openings (Berg et al. 2013, 2016; Ohwaki et al. 2018b), a forest matrix may be important for conservation of grassland Lepidoptera by adding to the network of patches in metapopulation dynamics. One might even ask whether clear-cuts should be considered a “matrix” or a primary habitat for some presumed grassland species (Blixt et al. 2015).

To be able to successfully conserve species traditionally seen as agricultural grassland species there is a need to identify and characterize “alternative habitats”. Burnet moths (Zygaenidae) seem restricted to specific types of habitat (Ravenscroft and Young 1996; Crispin and Warrington 1997), and may be abundant if habitat is suitable (Bourn 1995; Naumann et al. 1999). The reduction of suitable habitats has affected the burnet moths severely (Wenzel et al. 2006) and five out of Sweden’s six species are red-listed (Ahrné et al. 2015). Furthermore, they are considered good indicators of species-rich semi-natural grasslands (Franzén and Ranius 2004). This makes them suitable to use when studying the importance of temporary habitats and land use history for conservation of burnet moths in Sweden. Recently, the identification and synthesis of sex pheromones has opened up a new frontier when it comes to investigations of rare insects (e.g. Musa et al. 2013; Andersson et al. 2014; Burman et al. 2016; Larsson 2016). Zygaena moths use sex pheromones and pheromones have been synthesised (Priesner et al. 1984; Subchev 2014; Oleander et al. 2015) for Zygaena osterodensis, Z. viciae and Z. filipendulae.

In the present study, we sampled these three species with pheromone traps in clear-cuts—an assumed alternative habitat—and used attributes of the clear-cuts to explain differences in occurrence and abundance. More specifically, we wanted to evaluate the relative importance of:

1. connectivity of clear-cuts to species-rich grasslands
2. nectar sources on clear-cuts
3. larval host plants on clear-cuts

We hoped to assess whether clear-cuts should be considered a habitat (mainly 2 and 3 important), a supportive habitat (mainly 1 and 2 important), or a matrix (mainly 1 important).

It is known that clear-cuts on land that was previously meadow 150 years ago, are richer in plants, grassland plants, and butterflies compared with areas of continued forest cover (Ibbe et al. 2011; Blixt et al. 2015; Jonason et al. 2014, 2016; Milberg et al. 2019). It is also known that the flora of clearcuts change rapidly in their first years (e.g. Schoonmaker and McKee 1988; Pykälä 2004). Therefore, the range of the two variables nectar and host plant abundances were maximized by choosing clear-cuts of different age (2–8 years) and land-use history (meadow or continuous forest).

Finally, as the clear-cuts had also been surveyed using conventional transect count methods in the same season as pheromone traps were used (Blixt et al. 2015), we wanted to compare the outcomes to evaluate the relative efficacy of the pheromone traps to transect counts.

### Material and methods

#### Study species

*Zygaena* is a genus of moths in the family Zygaenidae. They are brightly coloured, day-flying and restricted to the western Palearctic. They prefer open and sunny biotopes and *Zygaena* larvae feed mainly on plants of the Fabaceae family, and the adults frequently use red and violet Dipsacaceae and Asteraceae flowers as nectar sources (Naumann et al. 1999; Sarin and Bergman 2010). Sweden has six species, of which five are on the red-list (Artdatabanken 2019a). Sex pheromones have been identified and synthesized for the species included in this study: *Z. osterodensis, Z. viciae and Z. filipendulae*. These species often co-occur, and their larvae feed mainly on one or a few species of Fabaceae (Söderström 2006).
Study area and selection of sites

The study was performed in southern Sweden in the province of Östergötland (N57°43′–58°15′; E15°00′–15°40′), in a landscape dominated by coniferous forest. The selection of clear-cuts is described in Blixt et al. (2015). Half of the clear-cuts had a management history as meadow and half as forest according to land use maps from the 1870s (Häradsekonomiska kartan, Jansson 1993; Runborg 1994). We selected clear-cuts between 1.5 and 7.0 ha in size (Table 1). We also selected clear-cuts according to the time since the cut (Table 1). Furthermore, we selected clear-cuts that were located at least 300 m from nearest semi-natural grasslands (a distance longer than most reported average dispersal distances of Zygaena, but shorter than maximum dispersal distance, according to a review by Franzen and Nilsson 2007). Finally, the distance between two clear-cuts also needed to be at least 300 m. No other considerations were made regarding the surroundings of the clear-cuts, that was dominated by coniferous forest of different age. Butterfly data from the clear-cuts have previously been reported by Blixt et al. (2015) while vegetation data were reported by Jonason et al. (2014).

Sex pheromone lures

Lures were prepared from grey rubber septa (PheroNet, Sweden) according to the methods of Burman et al. (2016), using compounds obtained from PheroBank, The Netherlands. The pheromone used for Z. filipendulae was a blend of Z7-12:Ac, Z9-14:Ac, Z5-12:Ac in the proportion of 100/10/3 µg per septa and for Z. viciae the same substances in the proportion of 100/10/10 µg, as published by Priesner et al. (1984). The pheromone blend used for Z. osterodensis was Z7-12:Ac, Z9-14:Ac in the proportion of 100/100 µg per septa, based on unpublished data by Ernst Priesner and Nils Ryrholm.

Sampling of burnet moths

Three sticky traps (transparent plastic delta traps; Csalomon, Budapest, Hungary) with one pheromone lure each were distributed at each clear-cut and left for one week. Traps were hung from a shrub, small tree or logging debris, at about breast height. The placement of traps on a clear-cut aimed at (i) selecting trap locations that were representative for that particular clear-cut, and (ii) the three trap locations being as similar as possible. Traps for Z. osterodensis were put up in the first week of July 2013, while the other traps were put up two weeks later, reflecting the flight period of the species.

Transect walks were conducted on three occasions during 2013, following standard procedures for this methodology (full details given in Blixt et al. 2015). Only two of the transect walks (June 17 to July 11; and July 17 to Aug. 3) occurred during the flight period of Zygaena species sampled. Walks were conducted between 09:00 and 17:00 (UTC +2) at temperatures above 17 °C and under predominantly sunny conditions, with winds of up to level 4 on the Beaufort scale. Transects were walked at a constant pace of 50 m/min. Transect lines were 25 m apart, and all specimens within an area of 5 m in front, 5 m to each side and 5 m up in the air were identified to species level. In this way 40% of each clear-cut was covered.

Connectivity

Species-rich grassland in the study area was identified using the TUVA database, administered by the Swedish Board of Agriculture. The database is the result of field inventories searching species-rich grasslands. Connectivity (Hanski 1994) for a clear-cut i, to nearby species-rich grasslands, was calculated using $C_i = \sum \exp\left(-d_j/\alpha\right)A_j$, where the areas of neighboring grasslands ($A_j$) are summed after scaling with the distance from i to j ($d_j$) and a scaling factor ($\alpha = 1.4$). The latter corresponds to “average dispersal distance”, and 1.4 was chosen based on Franzen and Nilsson (2007) who report such distances for Z. viciae being 1.1 and 1.8 km (two different years). Connectivity estimates were numerically skewed and therefore square-root transformed before analyses.

Vegetation sampling

In 2013, plant presence was recorded within 100 circular sample plots (radius 1 m) placed evenly throughout each clear-cut along transects spaced 25 m apart. The number of sample plots in which a species was present was taken as a measure of its frequency. The vegetation data has been presented elsewhere (Jonason et al. 2014, 2016).

Table 1 Background data on the 48 clear-cuts sampled for Zygaena spp. by pheromone traps

|                   | Average | SD   | Min | Max |
|-------------------|---------|------|-----|-----|
| Area (ha)         | 3.57    | 1.38 | 1.54| 7.00|
| Time since cutting (years) | 4.90    | 2.20 | 2   | 8   |
| Connectivity     | 194.8   | 129.8| 20.8| 530.3|
| Nectar index     | 23.2    | 26.7 | 0   | 130 |
| Host plant index (HPI) |         |      |     |     |
| HPI Z. osterodensis | 18.96   | 21.46| 0   | 73  |
| HPI Z. viciae     | 36.35   | 38.81| 0   | 138 |
| HPI Z. filipendulae | 4.62    | 5.44 | 0   | 21  |
| Specimens of Zygaena | 22.90   | 24.92| 2   | 123 |
Nectar sources

Adult *Zygaena* use several plant species for nectar, but some species are of particular importance. Based on literature (Naumann et al. 1999; Sarin and Bergman 2010) and the occurrence of species in our vegetation sample, we calculated an index that was the sum of frequencies of the following species on a clear-cut: *Knautia arvensis, Cirsium* spp., *Rubus fruticosus* coll. and *Trifolium* spp. As *Knautia arvensis* seems to be very important (Lack 1982; Holbeck et al. 2000; Sarin and Bergman 2010; unpublished data), we arbitrarily doubled its frequency when calculating the sum. As this nectar index was numerically skewed, it was square-root transformed before analyses.

Host plants

Although larvae of *Zygaena* seem to use only, or mainly, species of Fabaceae, there also seem to be different preferences among *Zygaena* species (e.g. Söderström 2006) within this plant family. For these reasons we calculated one host plant index per species, all being the sum of all frequencies on a clear-cut of the plant species that Artdatabanken lists as hostplants (http://artfakta.artdatabanken.se/taxon/1000572):

1. *Z. osterodensis*: *Vicia cracca, Vicia sylvatica* and *Lathyrus pratensis*
2. *Z. viciae*: *Lotus corniculatus, Lathyrus pratensis, Vicia* spp. and *Trifolium* spp.
3. *Z. filipendulae*: *Lotus corniculatus*

These host plant indices were all numerically skewed, and was therefore square-root transformed before analyses.

Statistical analysis

Species-wise generalized linear models (GLM with Negative binomial distribution and log-link) were used to assess the relative importance of three clear-cut attributes (explanatory variables) for the number of *Zygenae* trapped: connectivity, nectar and host plants. As the three variables were correlated (Table 2), one analysis was conducted per explanatory variable and species. Analyses were conducted with the software Statistica 13 (TIBCO Software Inc.).

Results

In total, 1075 *Zygaena* individuals were caught, of which 45.9%, 33.4%. 20.7% were of *Z. osterodensis*, *Z. viciae* and *Z. filipendulae*, respectively. *Zygaena* specimens were recorded on all clear-cuts, from a minimum of 2 to a maximum of 123 (divided among three traps per clear-cut).

| Correlation between connectivity, nectar index, and host plant index (HPI, one each for the three species of *Zygaena*) in data from the 48 clear-cuts sampled |
|---------------------------------------------------------------|
| **Square-root connectivity** | **Square-root nectar index** |
| Square-root nectar index | 0.442 | – |
| Square-root HPI *Z. osterodensis* | 0.389 | 0.848 |
| Square-root HPI *Z. viciae* | 0.422 | 0.856 |
| Square-root HPI *Z. filipendulae* | 0.454 | 0.437 |

*Zygaena osterodensis, Z. viciae* and *Z. filipendulae* were not recorded on 7, 5 and 7 clearcuts, respectively.

*Zygaena osterodensis* traps caught only the intended species, which is not surprising as it has an earlier flight period than the other species (n.b. its traps were put up two weeks earlier than the other traps). For the traps that were up simultaneously, targeting species with similar flight period, 26.5% of *Z. viciae* were caught in a trap targeting *Z. filipendulae*, and 30.5% of the *Z. filipendulae* were recorded in a trap targeting *Z. viciae*. Low specificity was expected due to the similarity of the pheromone blends.

Most *Zygaena* were caught by pheromone trapping in the clear-cuts that had previously been meadows (72.5%), which were also richer in nectar and host plants, but did not differ in connectivity from clear-cuts with a history as forest (Fig. 1).

Abundance of *Zygaena*

Overall, host plant index was the most important explanatory variable for the abundance of two of the three species while there were substantial differences among the species (Table 3). The abundance of *Z. osterodensis* could be explained by host plants (highest Wald, significant) and nectar plants. For *Z. viciae*, all three variables were highly significant, with nectar plants being the best model, followed by host plant (Table 3). Only host plants could explain *Z. filipendulae* abundance (Table 3).

Comparing transect walk data and pheromone trap catches

During transect walks, in total 10 specimens of two species of *Zygaena*, were recorded compared with 1075 specimens of three species in pheromone traps. *Zygaena viciae* was not recorded during transect walks but made up 33% of the trap catches and was recorded at 43 of the 48 clear-cuts. Only a single *Z. filipendulae* was seen in transects while contributing to 21% of trap catches being recorded on 41 of the clear-cuts.

In seven clear-cuts, a total of nine specimens of *Z. osterodensis* were seen during walks. In contrast, this species made
up 46% of the trap catch, being recorded in 41 clear-cuts. The numerical correspondence between observations and trap catches was low with, e.g., observations on 2 of the 7 clear-cuts lacking trap catches (Fig. 2).

### Discussion

#### Clear-cuts: habitat, supportive habitat, or matrix?

The perfect answer to our key question would emerge from studies of population dynamics. Such studies are unlikely to be feasible, however, given the mobility of the species involved, and the need to record fecundity and juvenile survival on different host plants and areas, etc. An alternative approach for these insects is to explore the relative importance of occurrence in the field, of (i) distance to presumed main habitat, (ii) nectar plants and (iii) host plants. The abundance of host plants was the strongest explanatory factor for the abundance of two species and the close second for the third, with consistently larger explanatory power than connectivity. Connectivity had only one case of significance (Z. viciae). These results suggest that clear-cuts are used as a fully functional habitat, and not a supporting habitat in the sense of only providing nectar. There is no support in these results for considering clear-cuts as an inert matrix, in which case only connectivity should have explanatory power. If clear-cuts are habitat, a grassland-based connectivity estimate is not meaningful for Zygaena, and a relevant estimate should include also clear-cuts.

Of the three species, Z. osterodensis was least affected by connectivity (lowest Wald values). This confirm previous reports stating that this species frequently use glades, forest edges and forest roads. Although a species occurring also in semi-natural grasslands in agricultural landscapes, our
results indicate that it is more of a forest species than the other two species (Artdatabanken 2019b).

Even if *Zygaena* spp. use clear-cuts as a habitat, clear-cuts themselves are temporary with a lifespan of about a decade (Jonason et al. 2016) after which a tree canopy is forming and shading increases. This overgrowth makes the habitat deteriorate as nectar sources and host plants decreases. Hence, in the longer time perspective, *Zygaena* spp. will have to repeatedly colonize new clear-cuts from nearby habitats, in a manner similar to that documented for *Mellicta athalia* and its occurrence in coppice woodland, a habitat that is open and suitable for a limited number of years forcing constant re-colonisation (Warren 1987a, b). Nearby habitats might constitute grasslands, road verges and powerlines—that are permanent habitats—and clear-cuts that are temporary (cf Wahlberg et al. 2002). According to classical metapopulation understanding, there would be a lower limit to the connectivity or number of patches, below which long-term survival is unlikely (Hanski 1998). In fact, this does seem to be the case, as some *Zygaena* species are absent from forest-dominated areas in southern Sweden (Sarin and Bergman 2010), suggesting that permanent habitats like grasslands become important on the larger landscape scale (Bergman et al. in prep). Wahlberg et al. (2002) made a similar conclusion when studying the population dynamics of the butterfly *Euphydryas aurinia* occurring in a landscape with a mix of clear-cuts and permanent grasslands. Their simulations indicated that clear-cuts on their own were not enough for long-term survival of the species; permanent grasslands were needed for continued presence in the landscape.

Previous studies have shown that historical management influenced butterfly and plant species in clear-cuts in the study areas. It is difficult to isolate the causal chain as connectivity, history, and plant species composition are interrelated. Still, *Zygaena* spp. do use clear-cuts as habitat, and rely on certain plants (nectar, hostplant) that are more likely to be encountered on clear-cuts on land affected by previous agricultural practices. To sum up, due to their prevalence in many landscapes, clear-cuts are important habitats likely affecting the geographic distribution of many *Zygae- nae*. Furthermore, landuse history influence the vegetation on clear-cuts (Jonason et al. 2014, 2016) and can thereby locally boost *Zygaena* populations in a way similar to that documented for butterflies (Ibbe et al. 2011; Berg et al. 2011; Blixt et al. 2015; Viljur and Teder 2016; Ohwaki et al. 2018a).

**Pheromones and monitoring**

Pheromone traps recorded 100 times more *Zygaena* specimens than did the transect walks. The time invested in fieldwork depends on logistics and, in the case of transect walks, weather. The current pheromone trapping needed four visits to a site (to set up and take down traps targeting the early-flying *Z. osterodensis*, then traps for the other two species; using slightly prolonged periods, the pheromone monitoring could have been done with three visits). In contrast, the transect walks needed only three visits. On the other hand, transect walks are more time-consuming and weather-dependent. Our two field workers sampled on average 6 clear-cuts per day using transect walks (SD 3.5; range 1–15), needing in total 48 man-days in the field. Assuming a person can visit 10 clear-cuts in a day in our study area, it would take less than 20 man-days to manage the pheromone trapping. Hence, with less than half the work effort, pheromone traps caught 100 times more *Zygaena* than were recorded in transect walks in the present study, which points to the potential importance of pheromone trapping for monitoring. It is worth pointing out that we used transect walk methodology developed for day-flying butterflies, and methods specifically developed for *Zygaena* would probably result in more reliable data.

Given the efficacy of traps, it is important to adjust the catching effort not to negatively affect populations. For example, refining the size and number of the traps, and limiting the area of the sticky surface in a trap are ways to minimize potential negative effects of using pheromones for monitoring *Zygaena*. Also, the days in the field can be adjusted during fieldwork and thereby fine-tuned to achieve an appropriate catch. Finally, it is worth pointing out the prospect of using live traps in monitoring (e.g. Andersson et al. 2014; Oleander et al. 2019), a hitherto unexplored option for monitoring of *Zygaena* spp. A drawback with live traps, however, is the need to empty them frequently (preferably daily for *Zygaena*), which limits the scale of sampling. But if monitoring is conducted as ‘citizen science’, there would be less constraints on labour. An interesting alternative to both sticky and live traps is if game cameras can be fine-tuned for burnet moths and baited with pheromones.

There was very poor correspondence between observations during transects walks and pheromone trap catches. Theoretically, this could be ascribed to transect walks underreporting and/or pheromone traps over-reporting. There is some indication that *Zygaena* turn up during transect walks to less extent than butterflies, as the former seem to preferentially fly in the afternoon (Franzén and Nilsson 2007; Wikström et al. 2009), suggesting they are sensitive to low temperatures prevailing earlier in the day. There might be a risk that pheromone traps, if highly efficient or targeting a very mobile species, attract specimens from outside of the intended sampling area. *Zygaena* spp. seems to be relatively mobile, with mark-recapture studies recording dispersals up to 5600 m, and with 8% of recaptures in a different habitat patch (Franzen and Nilsson 2007, 2012). Nevertheless, the evidence suggests that direct odour-guided attraction to pheromone sources only occurs within tens to a few hundred
meters (e.g., Schlyter 1992) which seems well-suited to the size of our sampled areas (clear-cuts of a few ha). Another line of argument, for pheromone trapping not being overly efficient, is that the traps failed to catch Z. filipendulae on two clear-cuts where this species had been seen during transect walks. Finally, in two catch-and-release trials involving Z. filipendulae in the UK, 58 individuals were recaptured after releasing from increasing distances from a centroid pheromone trap, of which only 2 returned from 100 and 120 m of the release point respectively (Burman et al. unpublished).

**Forestry management implications and conservation**

The current study adds to the growing evidence that clear-cuts are important, temporary Lepidoptera habitats, including for many red listed species. It has previously been proposed (Ibbe et al. 2011; Jonason et al. 2014, 2016; Blixt et al. 2015; Milberg et al. 2019) that such habitats can be boosted, at least on land with a grassland legacy, by planting of deciduous trees (more light to ground compared with conifers like Picea abies), increasing planting distances (that would delay the effect of tree canopy closure), or leaving some areas for free development (i.e. creating future glades in plantations). A more extreme measure would be to introduce forest grazing, a currently subsidized practice (Westin and Lennartsson 2018).

An open question remains regarding clear-cuts: if the presence of Zygaena hostplants on forested land does reflect previous grassland land use (Jonason et al. 2014, 2016; Milberg et al. 2019), or previous open, grazed forests (a practice that ceased during the early 1900s), then how much of this botanical legacy will remain after an additional forestry cycle? Considering that the next forestry cycle will be much denser than the previous one (Hedwall and Brunet 2016; Bergstedt et al. 2017; Pettersson et al. 2019), the legacy seems unlikely to prevail long-term, unless some measures are taken (see above).

**Conclusions**

This study suggests that clear-cuts in boreal forests are a fully functional habitat for three species of Zygaena studied. Given how prevalent clear-cuts are, they constitute a very significant habitat, albeit temporary, that likely affect the geographic distribution of many Zygaenae. Furthermore, clear-cuts on land that were used as meadow 150 years ago were particularly rich in Zygaena. Using sex pheromones to sample Zygaena proved very efficient compared with transect walks, and especially if live or camera traps can be developed, seems promising for monitoring.

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**Compliance with ethical standards**

**Conflicts of interest** There are no conflicts of interest in this study.

**Research involving animals** This study involves trapping insects for which no permit is needed in Sweden.

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