Recent advances in ankylosing spondylitis: understanding the disease and management [version 1; referees: 2 approved]

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Abstract
The term spondyloarthritis refers to a group of immune-mediated diseases characterised by inflammation of the axial skeleton, peripheral joints, and entheses. Ankylosing spondylitis (AS) is the most common and characteristic of these entities and even though it was first described over two centuries ago, the understanding of the underlying disease mechanism remains incomplete. It is known that around 40% of patients with AS have subclinical bowel inflammation, suggesting that the origin of the disease could be in the gut. Also, more genes and new molecules have demonstrated a role in the pathogenesis of AS. In this review, we analyse the latest therapies for spondyloarthritis and the most relevant discoveries over the last three years, together with their implications for different aspects of the disease.

Keywords
Ankylosing spondylitis, spondyloarthritis, spondyloarthropathy, management, pathogenesis

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Introduction

Ankylosing spondylitis (AS) is a chronic immune-mediated inflammatory arthritis included in the so-called group of spondyloarthritis (SpA). It typically develops in males in their third decade of life and affects mainly the axial skeleton and the sacroiliac joints. Although the oldest descriptions date from the time of Galen, it was not until the 19th century that the disease could be accurately diagnosed on the basis of reports by Vladimir Bekhterev, Adolph Strümpell, and Pierre Marie. The HLA-B27 allele is known to have a strong association with the disease; however, other genes play a part in its development. The discovery of several inflammatory pathways led to the era of the biologic therapies, which meant a revolution in the treatment and prognosis of AS. Tumour necrosis factor inhibitors (TNFis) were the first ones to be approved, but in the last few years the interleukin-17 (IL-17)/IL-23 axis has gained relevance, catalyzing in the licence of new biological disease-modifying antirheumatic drugs (bDMARDs) blocking IL-17.

In spite of all of these advances, the disease mechanisms underlying the disease are not fully understood and new information concerning the pathogenesis, triggers, and the outcome of new treatments continues to appear. In this review, we will focus on the advances of the last three years regarding the above aspects of AS.

Genetics

AS is considered an inherited disease, as over 90% of the risk for its development relies on genes1. However, the HLA-B27 allele accounts for only 20% of the genetic effect1. Other alleles, especially HLA-B, are thought to play an important role in the disease: HLA-B*13:02, HLA-B*40:01, HLA-B*47, and HLA-B*51 are some examples1. The most significant discovery of the last three years has been the interaction of ERAP1, the protein endoplasmic reticulum aminopeptidase 1, with the HLA-B alleles, resulting in a higher risk of developing AS. The main variant of the gene (rs30187, K528R) interacts only with the HLA-B27 allele, and in patients who are HLA-B27 negative, ERAP1 interacts with the HLA-B40 allele1. The mechanism underlying the increased risk remains unclear; nevertheless, it is known that the presence of this gene is not related to the radiographic severity of the disease1.

A recent study reported that a lower copy number of the TLR7 gene was a marker for susceptibility in AS in males but behaves as a protective factor in women1. Ruan et al.7 found that genetic polymorphisms of IL-12B (rs6871626) and IL-6R (rs4129267) were associated with an increased risk of AS independently of gender and also could function as biomarkers for diagnosis and prognosis.

Genome-wide association studies have shown that the T helper 17/23 (Th17/23) axis and its multiple genetic polymorphisms are involved not only in AS but also in inflammatory bowel disease (IBD) and psoriasis, supporting the hypothesis that there is a common underlying pathogenic mechanism and that the microbiome seems to be implicated in the development of the diseases.

Pathogenesis

How HLA-B27 initiates AS is unknown, and, after many years, some of the earliest hypotheses are still being investigated. The original hypothesis, called the ‘arthritogenic peptide theory’, suggests that the presentation of either bacterial peptides by HLA-B27 or self-mimicking HLA-B27-binding peptides from certain bacteria could initiate a cell-mediated immune reaction leading to AS8. The second one is the ‘unfolded protein response’ hypothesis, which suggests that HLA-B27 tends to misfold and accumulate in the endoplasmic reticulum, triggering a stress response that results in the release of IL-239–11. However, a new study12 has questioned these two theories, claiming that the arthritogenic peptide theory should be reassessed in terms of quantitative changes concerning self-peptide presentation and T-cell selection. It also stated that the absolute binding preferences of HLA-B27 allotypes are not sufficient to explain the association of the disease.

The third hypothesis is the ‘HLA-B27 homodimer model’13, which supports the view that HLA-B27 homodimers have an abnormal interaction with natural killer (NK) and CD4 T cells. Unlike the heterodimeric form of HLA-B27, the homodimer is able to bind to certain killer cell immunoglobulin-like receptors (KIRs), which are expressed on NK cells and T cells, causing the release of IL-1714–16. Ridley et al.17 have proven that CD4 T cells upregulate the expression of KIR-3D1L2 on the cell surface and that the binding of this receptor to HLA-B27 potentiates T-cell survival and Th17 cell differentiation. Th17 cells are a type of T-helper lymphocyte which produces IL-1718, a cytokine able to increase T-cell priming and stimulate immune cells such as fibroblasts and macrophages promoting the release of IL-6, TNF-α, and other chemokines19.

Oppmann et al. found that IL-23 is one of the triggers of the Th17 response20. This molecule is a pro-inflammatory cytokine that seems to play an important part in stabilising Th17 cell phenotype through the transcription factor Blimp-1 (Prdm1)21, which is associated with Crohn’s disease22. Th17 cells are commonly found in the intestinal lamina propria of the gut23 and their instability can be induced by exposure to certain bacteria, leading to the transition of Th17 cells into regulatory T (Treg) cells24. Maxwell et al.25 also suggested that IL-23 promotes the accumulation of Treg cells in the bowel, some of which probably were Th17 previously26,27.

Microbiome

Gut mucosal inflammation is estimated to be present in 70% of patients with AS28, progressing to clinical IBD in 5% of cases. Crohn’s disease and ulcerative colitis are characterised by having gut dysbiosis (a qualitative or quantitative microbial imbalance) which is also seen in AS29. The Ghent inflammatory arthritis and spondylitis cohort (GIANIT) studies strongly support the concept that there is a relationship between gut inflammation and the pathogenesis of AS30–32. One of the theories to explain it is that a constant antigenic stimulation can activate T cells and this might be responsible for chronic bowel inflammation31. Other studies suggest that patients with AS (and their first-degree relatives) have a high gut permeability,
which increases their exposure to gut microbes. In addition, animal studies proved that HLA-B27 alone was insufficient to develop AS, since transgenic rats that had been raised in a germ-free environment did not develop features of SpA. In the last few years, investigators have focussed on detecting pathogens as triggers for AS. *Klebsiella pneumoniae* was the first to be reported; this bacterium is thought to carry an antigen that resembles a molecule coded by the *HLA-B27* gene; however, the mechanism is not fully understood and other studies have stated that its involvement in AS is unlikely. Other relevant families of bacteria that have been associated with the development of AS are *Lachnospiraceae, Prevotellaceae, Rikenellaceae, Porphyromonadaceae*, and *Bacteroidaceae*. Non-gut bacteria are also thought to be involved; periodontal disease has become a possible target as anti-*Porphyromonas gingivalis* and anti-*Prevotella intermedia* antibodies are detected in high titres in patients with SpA. Some studies even suggest that chronic periodontitis is associated with severe spinal dysmobility in AS. Nevertheless, it is still an area of investigation, as recently published results contradict these findings.

**Gender differences**

Evidence gained in the last few years suggests that AS affects men and women differently. Landi et al. analysed a sample of 2,044 patients with AS and reported that disease commences earlier in men but that the diagnosis is usually more delayed than in women. Men have lower disease activity measured by Bath Ankylosing Spondylitis Disease Activity Index (BASDAI) and Assessment of SpondyloArthritis international Society (ASAS)-endorsed Disease Activity Score (ASDAS) and a better quality of life (Ankylosing Spondylitis Quality of Life Questionnaire, or ASQol) but have worse spinal mobility (Bath Ankylosing Spondylitis Metrology Index, or BASMI) and a more serious radiologic progression. New treatments and strategies have paralleled the description of new pathogenic mechanisms. **Table 1** summarises the most relevant drugs and their mechanism of action; some are still under investigation.

| Therapy             | Reference | Mechanism of action | Current status                  |
|---------------------|-----------|---------------------|---------------------------------|
| CT-P13 (infliximab biosimilar) | 43        | Anti-TNF            | Approved for use in AS         |
| SB4 (etanercept biosimilar) | 44        | Anti-TNF            | Approved for use in AS         |
| Secukinumab         | 45        | Anti-IL-17A         | Approved for use in AS         |
| Ixekizumab          | 46        | Anti-IL-17A         | Under investigation. Primary endpoints met. |
| Brodalumab          | 47        | Anti-IL-17R         | Not approved owing to safety issues |
| Ustekinumab         | 48,49     | Anti-IL-12/IL-23    | Not approved owing to lack of efficacy |
| ABT-122             | 50        | Anti-IL-17A/TNF-α   | Under investigation. Successful phase II clinical trials |
| COVA322             | 51        | Anti-IL-17A/TNF-α   | Not approved owing to safety issues |
| CBP30               | 52        | CBP/p300 bromodomain inhibition | Future trials announced |
| Sarilumab           | 53        | Anti-IL-6Rα         | Not approved owing to lack of efficacy |
| Tofacitinib         | 54        | JAK inhibitor       | Under investigation. Successful phase II clinical trials |

AS, ankylosing spondylitis; IL, interleukin; JAK, Janus kinase; TNF, tumour necrosis factor.

**Therapies**

Since 2015, there has been remarkable progress concerning the therapeutic management of AS. New treatments and strategies have paralleled the description of new pathogenic mechanisms. Table 1 summarises the most relevant drugs and their mechanism of action; some are still under investigation.

**Tumour necrosis factor inhibitors**

The first bDMARDs were approved several years ago. As the patents of some of these drugs were closer to expiration, pharmaceutical companies focused their interest on developing more affordable drugs. CT-P13, the infliximab biosimilar, was the first one to be released, followed by Benepali (SB4), the etanercept biosimilar, in 2016. Multiple trials endorse the similar safety and efficacy of CP-P13 and infliximab not only in patients with AS but also in the treatment of rheumatoid arthritis (RA). The PLANETAS study reported comparable results for patients with AS regarding ASAS20 and ASAS40. The extension of the study showed that switching from infliximab to its biosimilar did not imply negative effects on safety or efficacy. These findings were also confirmed in the PLANETRA study, carried out in patients with RA. In regard to Benepali (SB4), the most important efficacy and safety studies have been carried out in RA, showing results similar to those obtained with Enbrel. As far as therapy switching is concerned, the
safety and efficacy are maintained after switching from Enbrel to Benepali in patients with RA\textsuperscript{46}, and a study is taking place in Germany to assess this transition in patients with AS\textsuperscript{45}.

**Interleukin inhibitors**

The approval of secukinumab, a fully human monoclonal antibody able to neutralise IL-17A, has been a major advance in the treatment of AS\textsuperscript{46}. Patients with AS are known to have high levels of this interleukin\textsuperscript{46}, which is critically involved in the pathogenesis of the disease. IL-23 is secreted by antigen-presenting cells and stimulates Th17 cells, which are defined by their production of IL-17 cytokines\textsuperscript{44}.

Two double-blind, placebo-controlled, phase III clinical trials—MEASURE 1\textsuperscript{65} and MEASURE 2\textsuperscript{66}—reported a significant improvement in disease activity in patients with AS with effects sustained over 2 years; this was combined with a good safety profile. Pavelka et al.\textsuperscript{83} recently published the results of MEASURE 3, a randomised, double-blind, phase III trial in patients with active AS. In accordance with previous studies, a significant improvement was reported in the 16-week follow-up compared with placebo and this was sustained until week 52. The patients initially on placebo, who were randomly assigned on week 16, also experienced improvement through week 52. During this time, no major new adverse events were noted. In addition, MEASURE 5, another randomised, double-blind, phase III trial is ongoing. It is also comparing the safety and tolerability of secukinumab in patients with active AS versus placebo\textsuperscript{46}.

Nevertheless, there are other drugs targeting IL-17. COAST-W\textsuperscript{69} is a phase III, randomised, double-blind, placebo-controlled trial assessing the effect of ixekizumab on radiographic axial SpA. Ixekizumab is also a fully human monoclonal antibody that binds to IL-17A. The first results show that the drug met all of the primary and secondary endpoints. Brodalumab, a monoclonal antibody neutralising IL-17R, proved its efficacy for psoriatic arthritis; however, development was paused after a higher incidence of suicidal ideation was noted\textsuperscript{47}.

IL-23 is directly related to the development of enthesitis\textsuperscript{79}, and the IL-23/IL-17 axis occupies a central place in the pathogenesis of SpA. Ustekinumab is a human monoclonal antibody targeting the p40 subunit of IL-12 and IL-23. It is considered one of the most effective treatments for psoriasis\textsuperscript{71}; however, its efficacy in AS has not been as expected, and results have been contradictory: TOPAS, a prospective, open-label, single-arm, proof-of-concept clinical trial\textsuperscript{72}, reported a reduction of signs and symptoms in patients with active AS. In contrast, a phase III, multicentre, randomised, double-blind, placebo-controlled study\textsuperscript{80} evaluating the efficacy and safety of ustekinumab in the treatment of non-radiographic axial SpA had to end prematurely after a related study\textsuperscript{80} did not achieve the key points.

In regard to future approaches, ABT-122, an immunoglobulin molecule targeting both IL-17A and TNF-α, has recently demonstrated its efficacy in phase I and II trials for RA and psoriatic arthritis\textsuperscript{50,53}; however, given the importance of IL-17 in the pathogenesis of AS, it is expected that trials in patients with AS will commence soon. COVA322\textsuperscript{74} is another dual agent, a fusion protein antibody able to bind TNF-α and IL-17A. It was thought to be a promising therapy for AS; however, owing to safety issues in a trial for psoriatic arthritis, investigations have not gone further\textsuperscript{81}. CBP30\textsuperscript{75} is a selective inhibitor of CBP/p300 bromodomains able to suppress the production of cytokines by Th17 cells in patients with AS and healthy controls. Future trials with this molecule have been announced. Sarilumab, a fully human monoclonal antibody that blocks the α-receptor of IL-6, demonstrated a lack of efficacy for the treatment of AS in the ALIGN study\textsuperscript{53}.

**Janus kinase inhibitors**

Tofacitinib, a Janus kinase (JAK) inhibitor, is also able to interfere in the inflammatory cascade of IL-17, IL-21, and IL-23. In a 16-week clinical trial (with 12 weeks of treatment and 4 weeks of washout period), van der Heijde et al. reported the superiority of the 5 mg dose twice a day versus placebo: 63% and 40% of patients, respectively, achieved ASAS20 after 12 weeks\textsuperscript{44}.

An important aspect for biologic therapies is the cost. As more patients receive bDMARDs, healthcare systems have been forced to find new ways to cope financially: one cost-saving strategy is tapering. The 2016 update of ASAS-European League Against Rheumatism (ASAS-EULAR) recommendations for the management of axial AS\textsuperscript{80} suggested that patients in sustained remission are candidates to taper the dose of the drug. However, there is controversy regarding this approach because of conflicting results. One single-centre prospective study\textsuperscript{86} compared the evolution of two groups of patients: one receiving TNFis at the standard dose and another group with down-titration. After one year of treatment, the clinical outcomes were similar in the two arms but costs were significantly reduced in the latter group. Plasencia et al.\textsuperscript{77} previously reported similar results in a retrospective study comparing patients in Spain on tapering strategy versus patients on standard dose from the Netherlands. Even though the proportion of patients who maintained remission was similar, those in the tapering group had more flares than the patients receiving the standard dose. Regarding radiographic progression, Park et al.\textsuperscript{78} found that patients who had syndesmophytes at baseline had a more rapid radiographic progression if they received a tapering regime.

**Other therapies**

bDMARDs are not the only therapies that have been developed within the last few years. Fattahi et al.\textsuperscript{79} reported the results of a randomised, placebo-controlled trial of a new non-steroidal anti-inflammatory drug (NSAID): B-D-mannuronic acid. The 12-week ASAS response was similar to that obtained with naproxen, and the safety profile was considerably better; there were no renal side effects and gastrointestinal tolerability was good.

There has also been controversy regarding the administration of NSAIDs. Wanders et al.\textsuperscript{80} reported that continuous administration of NSAIDs reduced radiographic progression in comparison with on-demand therapy, whereas more recentlySieper et al.\textsuperscript{81} found that continuous treatment with diclofenac
for 2 years did not reduce radiographic progression compared with an on-demand administration.

**Biomarkers**

C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) are two acute-phase reactants that classically have been used to assess the presence of inflammation in patients. Unfortunately, they are not specific and the sensitivity is also low\(^{82}\), especially in patients with non-radiographic axial SpA\(^{83}\) where acute-phase proteins remain within normal limits most of the time.

Owing to the lack of a reliable test, many studies have tried to find a biologic marker capable of predicting a clinical outcome or disease activity in AS. After the discovery of new pathological pathways, interleukins became targets. IL-6 was in the spotlight for some years; but, whereas some studies reported an association between IL-6 and disease activity\(^{84-86}\), others produced opposite results\(^{87,88}\).

Calprotectin is a dimer of calcium-binding proteins used as a surrogate marker for gut inflammation. Its main application is monitoring disease activity in IBD\(^{89}\). Both Crohn’s disease and ulcerative colitis are known to be associated with AS. The pathway is not clear; there are discrepancies concerning whether gut inflammation is the cause or the consequence of musculoskeletal disease. Given this, studies have tried to evaluate calprotectin for the management of AS. Duran et al.\(^{90}\) found that faecal calprotectin was associated with a higher disease activity and is a better predictor of bowel involvement than CRP, ESR, BASDAI, and Bath Ankylosing Spondylitis Functional Index (BASFI). Klingberg et al.\(^{91}\) confirmed these results and also suggested that calprotectin could be used to identify patients with AS at high risk of developing IBD. In accordance with this, another study\(^{82}\) demonstrated that exercise could decrease the levels of calprotectin in patients with AS and that this was associated with a reduction of disease activity.

**Bone involvement**

A typical feature of AS is bone formation co-existing with bone resorption. As with many other inflammatory diseases, there is an imbalance between these processes, which results in the appearance of syndesmophytes and bone erosions in patients with AS\(^{92}\). IL-17, one of the most active cytokines in SpA, has a major role promoting osteoclastogenesis directly and through the activation of receptor activator of nuclear factor kappa B (RANK)\(^{94-97}\). However, it does not exert its action alone; it works in combination with TNF-α. The latter molecule triggers bone destruction through the RANK-RANK ligand (RANK-RANKL) system and inhibits bone formation via the overexpression of Dickkopf-related protein 1 (DKK1)\(^98\), which suppresses the WNT bone pathway\(^{99,100}\). WNT/b-catenin signalling is a regulator of osteogenesis and its high levels promote the formation of osteoblasts and reduce osteoclastogenesis (and therefore bone resorption)\(^{100}\). In contrast to TNF-α, IL-17 has a dual effect because it can promote not only bone destruction by acting complementarily with TNF-α\(^99\) but also bone formation at sites of inflammation or exposed to mechanical stress\(^{101}\). An international study\(^{102}\) reported contradictory results, as they found that patients with AS had decreased indicators of osteoclast formation and bone resorption, suggesting a reduced capacity of osteoclast precursors to differentiate into osteoclasts and resorb, probably because of low RANKL/OPG and CD51/CD61 expression.

Ranganathan et al.\(^{103}\) found that macrophage migration inhibitory factor (MIF) was able to act on osteoblasts, promoting inflammation and new bone formation. They also suggested that the main source of MIF-producing cells was in the gut and that certain pathogens can induce its release. In addition, the level of MIF was predictive of progressive spinal damage.

More cytokines are being implicated in the pathogenesis of AS. El-Zayadi et al.\(^{104}\) recently reported that IL-22 regulates the function of mesenchymal cells, inducing proliferation, migration, and osteogenesis, in an inflammation-dependent context. IL-32γ has been shown to be elevated in the joints and tissues of patients with AS and is able to induce osteoblast differentiation and atypical new bone formation\(^{105}\). Another interleukin, IL-37, is also elevated in patients with osteoporosis in addition to AS, and its levels correlate with disease activity and bone mineral density\(^{106}\). This is consistent with Kwon et al.\(^{107}\), who found that osteoblast-lineage cells are increased in patients with AS and are reduced after therapy with infliximab. They also reported that the drug allows mature osteoblast differentiation in late inflammation.

Machado et al.\(^{108}\) stated that fat deposition in the vertebral corners is associated with radiographic progression, and they postulated that there should be a ‘window of opportunity’, as fat lesions can be preceded by inflammatory lesions. Interestingly, they also found that even though the absence of bone marrow oedema and fat deposition was negatively associated with radiographic progression, there was still bone formation in those areas, implying that there must be other pathways stimulating osteoproliferation.

**Imaging**

The assessment of structural damage is valuable in patients with AS. The modified Stoke Ankylosing Spondylitis Spinal Score (mSASSS) (which consists of the sum of scores of the lumbar and cervical spine from a lateral view, ranging from 0 to 72) is a commonly used method for detecting changes in plain radiographs\(^{109}\); however, X-rays are not a very sensitive tool and they are not able to properly differentiate squaring of vertebral bodies. Kim et al.\(^{110}\) evaluated the squaring of the first sacral vertebra (S1) by using a method proposed by Ralston et al.\(^{111}\): squaring was associated with mSASSS and changes in magnetic resonance imaging (MRI) and therefore suggested that this assessment could be used to predict early axial involvement of the spine in AS. Regarding other predictors of progression, Dougados et al.\(^{112}\) found that HLA-B27-negative patients with normal CRP and normal MRI of the sacroiliac joints have a 1.2% likelihood of progressing to radiological axial SpA within 5 years. This likelihood of progression increased to 18.4% in HLA-B27-positive patients, with raised CRP and
bone marrow oedema in the MRI of the sacroiliac joints. In HLA-B27-positive patients, smoking also seems to be associated with a proliferative metabolic pattern of the cartilage, which could justify its outgrowth and the development of enthesopathic lesions\textsuperscript{[11]}. Althoff et al. were the first to suggest that contrast was not needed to assess axial activity in AS\textsuperscript{[12]}. However, Zhao et al.\textsuperscript{[13]} confirmed these results by using a large sample of patients, showing that the combination of short tau inversion recovery (STIR) and diffusion weighted imaging (DWI) in MRI is sufficient to monitor disease activity, avoiding the unnecessary use of contrast with its risks. Conversely, Bradbury et al.\textsuperscript{[14]} found that DWI has a moderate utility for assessing disease activity or monitoring response to treatment; however, it seems to be a great tool to distinguish axial SpA and non-inflammatory back pain.

Another MRI finding has been the discovery of the ‘backfill sign’. It was initially described by Weber et al.\textsuperscript{[15]}, who defined it as a high signal intensity filling the sacroiliac joint space on T1-weighted images which could represent fat metaplastic tissue filling excavations in subchondral bone; however, the histopathological origin has not been assessed yet. Recently, studies confirmed that the ‘backfill sign’ had a high specificity for SpA (between 95.8\% and 98\%\textsuperscript{[16]} but the sensitivity was only 59\%\textsuperscript{[17]}. Therefore, its presence could be used to support the diagnosis of axial SpA, but its absence should not be used to exclude it.

**Summary**

The numerous investigations and studies carried out within the last three years have improved the understanding of the pathogenesis of AS. This has facilitated the development of new treatment strategies with the consequent improvement of the quality life of patients with SpA. As more is known about the disease, the greater the complexity that is revealed, emphasizing the need to continue investigation to achieve even more efficient control of the disease.

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**References**

1. Chen B, Li J, He C, et al.: Role of HLA-B27 in the pathogenesis of ankylosing spondylitis (Review). Mol Med Rep. 2017; 15(4): 1943–51. Published Abstract | Publisher Full Text | Free Full Text
2. Reveille JD: The genetic basis of ankylosing spondylitis. Curr Opin Rheumatol. 2000; 12(4): 332–41. Published Abstract | Publisher Full Text | Free Full Text
3. Rubin LA, Amos CI, Wade JA, et al.: Investigating the genetic basis for ankylosing spondylitis. Immunogenetics. 1995; 43(1–2): 271–287. Published Abstract | Publisher Full Text
4. Cortes A, Pult SL, Leo PJ, et al.: Major histocompatibility complex associations of ankylosing spondylitis are complex and involve further epistasis with ERAP1. Nat Commun. 2016; 7: 1146. Published Abstract | Publisher Full Text | Free Full Text
5. Ozen G, Denz R, Eren F, et al.: Association of ERAP1, IL23R and PTGER4 Polymorphisms with Radiographic Severity of Ankylosing Spondylitis. Open Rheumatol. 2017; 11: 1–9. Published Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
6. Wang M, Xu S, Zhang X, et al.: Association of TLR7 gene copy number variations with ankylosing spondylitis in a Chinese population: a case control study. Clin Exp Rheumatol. 2018. Published Abstract | F1000 Recommendation
7. Ruan WF, Xie JT, Jin Q, et al.: The Diagnostic and Prognostic Role of Interleukin 12B and Interleukin 6R Gene Polymorphism in Patients With Ankylosing Spondylitis. J Clin Rheumatol. 2018; 24(1): 18–24. Published Abstract | F1000 Recommendation
8. Costello ME, Elewaut D, Kenna TJ, et al.: Microbes, the gut and ankylosing spondylitis. Arthritis Res Ther. 2013; 15(2): 214. Published Abstract | Publisher Full Text | Free Full Text
9. Benjamin R, Parham P: HLA-B27 and disease: a consequence of inadvertent antigen presentation? Rheum Dis Clin North Am. 1992; 18(1): 11–21. Published Abstract
10. Mear JP, Schreiber KL, Münz C, et al.: Misfolding of HLA-B27 as a result of its B pocket suggests a novel mechanism for its role in susceptibility to spondyloarthropathies. J Immunol. 1999; 163(12): 6665–70. Published Abstract
11. Smith JA, Colbert RA: Review: The interleukin-23/interleukin-17 axis in spondyloarthropathies: Th17 and beyond. Arthritis Rheumatol. 2014; 66(2): 231–41. Published Abstract | Publisher Full Text | Free Full Text
12. Schittenhelm RB, Sian TC, Wilmann PG, et al.: Revisiting the arthritogenic peptide theory: quantitative not qualitative changes in the peptide repertoire of HLA-B27 allelotypes. Arthritis Rheumatol. 2015; 67(3): 702–13. Published Abstract | Publisher Full Text
13. Allen RL, O’Culannagh CA, McMichael AJ, et al.: Cutting edge: HLA-B27 can form a novel beta 2-microglobulin-free heavy chain homodimer structure. J Immunol. 1999; 162(8): 5045–8. Published Abstract
14. Kollberger S, Bird L, Sun MY, et al.: Cell-surface expression and immune receptor recognition of HLA-B27 homodimers. Arthritis Rheum. 2002; 46(11): 2972–82. Published Abstract | Publisher Full Text
15. Kollberger S, Chan A, Sun MY, et al.: Interaction of HLA-B27 homodimers with KIR3DL1 and KIR3DL2, unlike HLA-B27 heterotrimers, is independent of the sequence of bound peptide. Eur J Immunol. 2007; 37(5): 1313–22. Published Abstract | Publisher Full Text
16. Payeli SK, Kollberger S, Marroquin Belaunzaran O, et al.: Inhibiting HLA-B27 homodimer-driven immune cell inflammation in spondylarthritides. Arthritis Rheum. 2012; 64(10): 3139–49. Published Abstract | Publisher Full Text
17. Ridley A, Hatano H, Wong-Basza I, et al.: Activation-Induced Killer Cell Immunoglobulin-like Receptor 3DL2 Binding to HLA-B27 Licenses Pathogenic T Cell Differentiation in Spondyloarthritides. Arthritis Rheumatol. 2016; 68(4): 901–14. Published Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
18. Dong C, T,17 cells in development: an updated view of their molecular identity and genetic programming. Nat Rev Immunol. 2008; 8(5): 337–48. Published Abstract | Publisher Full Text
19. Yan D, Cheung J, Scheeren H, et al.: IL-23 is essential for T cell-mediated colitis and promotes inflammation via IL-17 and IL-6. J Clin Invest. 2006; 116(5): 1310–6. Published Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
20. Oppmann B, Lesley R, Blom B, et al.: Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. Immunity. 2000; 13(5): 715–25. Published Abstract | Publisher Full Text
21. Jain R, Chen Y, Kanno Y, et al.: Interleukin-23-Induced Transcription Factor Blimp-1 Promotes Pathogenicity of T Helper 17 Cells. Immunity. 2016; 44(1): 131–42. Published Abstract | Publisher Full Text | F1000 Recommendation
22. Ellinghaus D, Zhang H, Zeissig S, et al.: Association between variants of PRDM1 and NOD2 and Crohn’s disease, based on exome sequencing and functional
studies. Gastroenterology. 2013; 145(2): 339–47. PubMed Abstract | Publisher Full Text | Free Full Text

23. Ghoreschi K, Laurence A, Yang XP, et al.: Generation of pathogenic T,17 cells in the absence of TGF-β signalling. Nature. 2010; 467(7318): 987–71. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation

24. Gagliani N, Amezcuca Vesely MC, Iseppon A, et al.: Th17 cells. Transdifferentiate into regulatory T cells during resolution of inflammation. Nature. 2015; 523(7559): 221–5. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation

25. Maxwell JR, Zhang Y, Brown WA, et al.: Differential Roles for Interleukin-23 and Interleukin-17 in Intestinal Immunoregulation. Immunity. 2016; 44(3): 739–50. PubMed Abstract | Publisher Full Text

26. Ommacht C, Park JH, Cording S, et al.: MUCOSAL IMMUNOLOGY. The microbiota regulates type 2 immunity through RORC+ T cells. Science. 2015; 349(6251): 91–93. PubMed Abstract | Publisher Full Text | Free Full Text

27. Fasching P, Stradner M, Granner W, et al.: Therapeutic Potential of Targeting the Th17/Treg Axis in Autoimmune Disorders. Molecules. 2017; 22(1): ei134. PubMed Abstract | Publisher Full Text

28. COSTELLO ME, CICcia F, Wilner D, et al.: Brief Report: Intestinal Dysbiosis in Ankylosing Spondylitis. Arthritis Rheumat. 2015; 67(3): 686–91. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation

29. Ciccia F, Rizzo A, Triolo G: Arthritis/spondylitis. Curr Rheumatol Rep. 2016; 18(5): 87–92. PubMed Abstract | Publisher Full Text | Free Full Text

30. Jacques P, Elewaut D, Mielants H: Subclinical gut inflammation in ankylosing spondylitis and their healthy relatives. Arthritis Care Res (Hoboken). 2013; 65(11): 1823–8. PubMed Abstract | Publisher Full Text | Free Full Text

31. van der Heijde D, Deodhar A, Wei JC, et al.: Sarilumab for the treatment of ankylosing spondylitis: Results of a Phase II, randomised, double-blind, placebo-controlled study (ALIGN). Ann Rheum Dis. 2016; 75(10): 1501–7. PubMed Abstract | Publisher Full Text | Free Full Text

32. van der Heijde D, Deodhar A, Wei JC, et al.: Tofacitinib in patients with ankylosing spondylitis: a phase II, 16-week, randomised, placebo-controlled, dose-ranging study. Ann Rheum Dis. 2017; 76(7): 1340–7. PubMed Abstract | Publisher Full Text | Free Full Text

33. Sieper J, Braun J, Kay J, et al.: Subclinical gut inflammation in ankylosing spondylitis: The 5-year data of the DESIR cohort. Ann Rheum Dis. 2017; 76(1): 1823–8. PubMed Abstract | Publisher Full Text | Free Full Text

34. Cho IH, Lee N, Song D, et al.: Evaluation of the structural, physicochemical, and biological characteristics of SB4, a biosimilar of etanercept. Mabs. 2016; 8(6): 1136–55. PubMed Abstract | Publisher Full Text | Free Full Text

35. Park W, Yoo DH, Min J, et al.: Efficacy and safety of switching from reference infliximab to CT-P13 compared with maintenance of CT-P13 in ankylosing spondylitis: 102-week data from the PLANETAS extension study. Arthritis Rheumatol. 2017; 79(3): 346–54. PubMed Abstract | Publisher Full Text | Free Full Text

36. Park W, Yoo DH, Min J, et al.: Efficacy and safety of switching from reference infliximab to CT-P13 compared with maintenance of CT-P13 in ankylosing spondylitis: 102-week data from the PLANETAS extension study. Arthritis Rheumatol. 2017; 79(3): 346–54. PubMed Abstract | Publisher Full Text | Free Full Text

37. Sieper J, Kay J, Braun M, et al.: Subclinical gut inflammation in ankylosing spondylitis: The 5-year data of the DESIR cohort. Ann Rheum Dis. 2017; 76(1): 1823–8. PubMed Abstract | Publisher Full Text | Free Full Text

38. Cho IH, Lee N, Song D, et al.: Evaluation of the structural, physicochemical, and biological characteristics of SB4, a biosimilar of etanercept. Mabs. 2016; 8(6): 1136–55. PubMed Abstract | Publisher Full Text | Free Full Text

39. Efficacy, safety, and tolerability of secukinumab in patients with rheumatoid arthritis continuing on SB4 or switching from reference etanercept to SB4. Ann Rheum Dis. 2017; 76(2): iii17–21. PubMed Abstract | Publisher Full Text | Free Full Text

40. Emery P, Vencovský J, Sylwestrzak A, et al.: Long-term efficacy and safety in patients with rheumatoid arthritis continuing on SB4 or switching from reference etanercept to SB4. Ann Rheum Dis. 2017; pii: arhms-2017-211591. PubMed Abstract | Publisher Full Text | Free Full Text

41. Liu W, Wu YH, Zhang L, et al.: Elevated serum levels of IL-6 and IL-7 may associate with the development of ankylosing spondylitis. Int J Clin Exp Med. 2015; 8(10): 17362–76. PubMed Abstract | Publisher Full Text | Free Full Text

42. Yago T, Naruke Y, Kawamoto M, et al.: IL-23 and Th17 Disease in Inflammatory Arthritis. J Clin Med. 2017; 6(9): pii: E81. PubMed Abstract | Publisher Full Text | Free Full Text

43. Barakalluo X, Kivitz AJ, Deodhar AA, et al.: Long-term effects of interleukin-17A inhibition with secukinumab in active ankylosing spondylitis: 3-year efficacy and safety results from an extension of the Phase 3 MEASURE 1 trial. Clin Exp Rheumatol. 2016; 34(1): 50–5. PubMed Abstract | F1000 Recommendation

44. Marzo-Ortega H, Sieper J, Kivitz A, et al.: Secukinumab and sustained improvement in signs and symptoms of patients with active ankylosing spondylitis. Arthritis Rheumatol. 2017; pii: arthriminds-2017-211591. PubMed Abstract | Publisher Full Text | F1000 Recommendation

45. Pavelka K, Kivitz A, Dokoupilova E, et al.: Efficacy, safety, and tolerability of...
inflammation followed by fat deposition is the strongest contributor to the development of new bone at the same vertebral corner: a multilevel longitudinal analysis in patients with ankylosing spondylitis. Ann Rheum Dis. 2016; 75(8): 1486–93.

108. Wanders AJ, Landewé RB, Spoorenberg A, et al.: What is the most appropriate radiologic scoring method for ankylosing spondylitis? A comparison of the available methods based on the Outcome Measures in Rheumatology Clinical Trials filter. Arthritis Rheum. 2004; 50(8): 2822–32.

109. Kim JY, Lee S, Joo KB, et al.: Loss of anterior concavity of the first sacrum can predict spinal involvement in ankylosing spondylitis. Rheumatol Int. 2016; 36(1): 161–5.

110. Ralston SH, Urquhart GD, Brzeski M, et al.: A new method for the radiological assessment of vertebral squaring in ankylosing spondylitis. Ann Rheum Dis. 1992; 51(3): 330–3.

111. Munk HL, Guddmann NS, Christensen AF, et al.: Cartilage collagen type II seromarker patterns in axial spondyloarthritis and psoriatic arthritis: associations with disease activity, smoking and HLA-B27. Rheumatol Int. 2016; 36(4): 541–9.
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