Histopathological and Follicular Atresia Assessment of Rat’s Ovarian Tissue Following Experimental Chronic Spinal Cord Injury

Sam Zarbakhsh, Mohammadhasan Tabrizi Amjad, Behpour Yousefi, Mohammadreza Aldaghi, and Hamidreza Sameni

1Research Center of Nervous System Stem Cells and Department of Anatomical Sciences, School of Medicine, Semnan University of Medical Sciences, Semnan, Iran

Abstract

Background: One of the important side effects in spinal cord injuries in both genders is sexual dysfunction. This study aimed at investigating histopathological changes of ovaries in the chronic phase after spinal cord injury.

Methods: Animals (adult female rats) were divided to the following groups, Control (Co), Sham (Sh), and Spinal Cord Injury (SCI); each group contained 18 rats. The spinal cord of SCI animals was transected by the bilateral laminectomy method on level T9, while the rats of the sham group were incised in the same location without any spinal cord injury. Animals were sacrificed at 7th, 14th and 21st day postsurgical intervention, then the body weight, and the weight and volume of ovaries were measured. The slices were stained by hematoxylin and eosin and periodic acid Schiff methods. The histomorphometric changes of the diameter of the follicle and ovum, and the thickness of granulosa layer were measured in different kinds of follicles including, Unilaminar Primary Follicle (UPF), Multilaminar Primary Follicle (MPF), Secondary Follicle (SF), and Tertiary Follicle (TF).

Results: The results showed that the animal weights were decreased in three SCI groups (P = 0.018). In the SCI groups, the diameter of follicle and ovum and the thickness of granulosa layer were significantly decreased in different kinds of follicles (P = 0.012). Also the thickness of zona pellucida and theca interna were significantly decreased in UPF, SF, and TF in the SCI groups on 14th and 21st day (P = 0.024). The histopathologic examination revealed widespread ovarian follicle atresia in the SCI groups on the 14th and 21st day, including numerous cell debris and inflammatory cells in the antrum atretic follicles. The ovarian stroma showed edema, fibrosis, hypercellularity, and vasodilation in the SCI group, compared to the sham or control groups.

Conclusions: The histopathologic data indicated that after spinal cord injury many histologic parameter changes occurred and heterogeneity of ovarian structure increased. These changes may be caused by dysfunction of the autonomic system and the modification in amount of the ovarian neurotransmitters.

Keywords: Ovary, Spinal Cord Injury, Denervation, Histopathology, Rat

1. Background

Sexual and reproductive health is an important quality of life component, which can have a major impact on patient satisfaction. Spinal pathology arising from trauma, deformity, and degenerative disease processes may be harmful to sexual and reproductive function (1).

Natural fertility of females is a consequence of regular and coordinated biological processes. The neurohormonal system plays a very important and vital role in regulation and stability of the reproductive system (2-4).

Sexual dysfunction is an important complication of spinal cord injury (SCI). Similar to many other complications of SCI, sexual function is also affected by the integrity of neural pathways, motor, sensory and autonomic, as well as psychological and social factors (5).

Sexual dysfunction in patients with SCI is a product of 2 major factors: the physiological changes resulting from the neurological deficit and the psycho-social consequences of the disability. Sexual disturbances in females after SCI are less understood and described than those occurring in post-SCI males (6, 7).

Recent findings have shown that in mamalian ovaries content aggregation of neuron perikaryons occurs, especially in close proximity with theca folliculi layer, interestitial glands, and vessels. Some of these neurons are sympathetic and have an important role on regulation of ovarian function (8, 9). Other evidences have shown that the ovary is innervated by external adrenergic nerves in 2 ways; superior ovarian nerve and ovarian plexus around the ovarian artery (10, 11). Previous studies have shown that superior ovarian nerve transection decreases ovarian progesterone level and effects the maturation of its steroids (12-14). Some studies stated that in females with spinal transection, Follicle stimulating hormone (FSH) level decreased and es-
trogen had little effect on vaginal mucosa (15). In animals with bilateral transection of ovarian nerve, decreases were observed at the intra-ovarian norepinephrine level. Also these findings suggested that hormonal regulation of the ovaries is directly controled by central nervous system via specific neuronal pathways (10-13, 15).

Other studies reported that ovarian nerves participate in differentiation of primary follicles, coexisting of FSH receptors, via neurotransmitter connection to cyclic Adenosine Monophosphate (cAMP) productive system. Therefore, as follicles are grown in parts of the ovary including higher density of neurons, they could be more rapidly regulated by gonadotropins (14). On the other hand, it has been reported that CNS includes specific cells in parapyramidal nuclei, caudal rapha, paraventricles, and lateral hypothalamus that play an effective role on ovarian function via sympathetic or vagus nerves (16). There are evidences that prove the neuro-hormonal effects on ovarian physiology, specially follicular maturation and ovulation (16).

Though ovaries in mammals are innervated by spinal nerves (T10-L2) and abdominal vagal branches, yet, progressive signaling occurs only through the spinal cord. Therefore, the SCI disconnects oncoming signaling transport from gonads to the brain, which cut off the liberation of gonadotropin from the hypothalamus and lead to hypogonadism of patients with SCI (17, 18). Although the ovaries are transmitted with both the vagus nerve and the spinal cord branches, the continuation of usual ovarian action after SCI could be mainly due to the preservation of neuronal tissue of the spinal cord. Hence, interruptions in signaling following SCI changes ovarian physiology via inflammation and apoptosis (17, 18).

Spinal cord injury is one of the complications of medicines that effect organs of the extremities and several systems of the human body. One of the most important complications of spinal cord injury is sexual disorders in both genders that may lead to major psychological and mental problems. Findings of earlier studies and our results have demonstrated that male rats with acute and chronic spinal cord injury, showed extensive histopathological changes, including spermatogenesis arrest, decrease of seminiferous epithelial thickness, and decrease of prostate and seminal vesicle epithelium, etc. These structural changes lead to sexual dysfunctions of reproductive organs such as testis, prostate gland, seminal vesicle, and genital ducts (2, 19-21).

Although the frequency of females with spinal cord injury is low, yet, sexual dysfunction in this gender is more complicated and unclear (2, 3). Thus, findings of some reports about spinal cord transection effects on females reproductive system is impossible and there are only a few studies about the physiological and hormonal outcomes of these patients.

The goal of this research was to study the structural and histopathological alterations in ovaries following chronic spinal cord transection in wistar rats.

2. Methods

2.1. Experimental Animals

Overall, 54 adult female wistar rats (180 ± 20 g) were included in this interventional experimental study. The rats were maintained in 12:12 light and dark photoperiod, relative humidity of 45% to 55%, and temperature of 24 ± 2°C and free access to standard diet and water. All experimental protocols were approved by the Institutional animal ethics committee of Semnan University of Medical Sciences.

2.2. Spinal Cord Injury Procedure

Following a week of acclimation, the rats were randomly divided to 3 groups (n = 18 for each group): Spinal Cord Injury group (SCI, animals with their spinal cord transected), Sham group (Sh, animals only had an incision without their spinal cord transected) and Control group (Co, without incision). Animals were examined on 7th, 14th and 21st day after the procedure. In the SCI group, animals were anesthetized by administering ketamine/xylazine (80/20 mg/kg), intraperitoneally, Then, T9 vertebrae laminae were cut by bilateral laminectomy and then transverse transection of the spinal cord was made (20, 21). The rats in the sham group were incised in the same location without spinal cord transected. No intervention was implemented for the control group. The location of the surgery in the SCI and sham groups was disinfected and muscles, fascia, and skin were sutured. In the SCI group, in order to prevent probable mortality due to lower limb paralysis and numbness, special care was applied.

2.3. Animals Special Cares

The place in which the rats were kept was clean, and the temperature was regulated. Neurogenic bladder due to spinal cord injury was evacuated regularly. In this study, handling of the bladder was achieved by the Linsenmeyer method (22). External genitalia was disinfected with 70% alcohol after bladder evacuation. All animals in sham and SCI groups received 5 mg/kg of gentamycin once a day for 5 days post-surgery. For a possible bladder infection control, 2.5 mg/kg of ketoprofen once a day for 2 days was administered to reduce post-surgical pain (20, 22, 23). To avoid autophagia, heterophagia and following mortality rate, paralytic extremities care was important; thus, the lower limbs were plased in sterile polyethylene tubes.
2.4. Tissue Processing and Histomorphometric Analysis

All the animals were weighted on 7th, 14th and 21st day after spinal cord transection. The rats were then sacrificed and ovaries were removed and their weight and volume were measured immediately and fixed in Boin’s and 10% formaldehyde solution. After fixation and processing of specimens, paraffin blocks were prepared and were serially sectioned with micrometre (5 µm). The slides were stained by Hematoxylin and Eosin (H and E) and periodic acid schiff (PAS) methods (24).

For the purpose of an exact evaluation of ovarian quantitative changes, follicles were categorized, according to the criteria listed in the Nomina Histologica, as the following types: Perimordial Follicle (PF, monolayer of flattened follicular cells), Unilaminar Primary Follicle (UPF, monolayer of cuboidal granulosa cells), Multilaminar Primary Follicle (MPF, multilayer of cuboidal granulosa cells), Secondary Follicle (SF, oocyte surrounded by 2 or more granulosa cell layer, without an antrum) and graafian follicle or Tertiary Follicle (TF, oocyte surrounded by a stratified epithelium of granulosa cells with follicular antrum) (25).

From each ovary, a transverse section at 10 fields was randomly selected at a × 400 magnification. Slides were studied by a light microscope equipped with a computerized image analysis system (Motic Images plus 2 Co., LTD). In each instance of the ovary, follicle and ovum diameter, thickness of zona pellocida, granulosa layer, and theca interna in all kinds of follicles were calculated.

2.5. Follicular Atresia

In this study, diagnoses of atretic follicles on the basis of the morphological criteria of the method were described by Greenwald and Roy (1994) in H and E stained serial sections of the ovary (26). The earliest sign of atresia was presence of 5% pyknotic granulosa cells in the largest cross section of the follicle.

2.6. Statistical Analysis

All data were expressed as means ± standard error of the mean (SEM). The quantitative data were analyzed using the SPSS software (Version 20). Data from experiments with more than 2 independent variables were analyzed using Analysis of Variance (ANOVA) followed by the Tukey-Kramer post-hoc tests. Significant differences were considered at P values of < 0.05.

3. Results

3.1. General Characteristic of Animals

Animals weight, mean volume, and weight of ovaries declined in spinal cord injury (SCI) rats in comparison with the sham and control groups. Decrease in animals weight was statistically significant (P < 0.05), (Table 1).

| Animal Groups | Body Weight, g | Ovarian Weight, mg | Ovarian Volume, cm³ |
|---------------|---------------|--------------------|---------------------|
| Co | 223.8 ± 14.8 | 56.2 ± 3.3 | 0.059 ± 0.002 |
| Sh | 225.4 ± 12.4 | 55.9 ± 4.7 | 0.061 ± 0.005 |
| SCI | 208.5 ± 13.6b | 55.5 ± 2.7 | 0.058 ± 0.003 |

| Animal Groups | Body Weight, g | Ovarian Weight, mg | Ovarian Volume, cm³ |
|---------------|---------------|--------------------|---------------------|
| Co | 227.7 ± 8.4 | 60.7 ± 4.8 | 0.063 ± 0.005 |
| Sh | 219.2 ± 11.3 | 61.3 ± 3.9 | 0.062 ± 0.004 |
| SCI | 198.2 ± 10.9b | 59.3 ± 4.2 | 0.061 ± 0.003 |

Abbreviations: Co, Control; Sh, Sham; SCI, Spinal Cord Injury.

3.2. Histopathological Assessment

In the ovaries of the SCI group there were follicles at different stages of development and a number of corpus luteum consisting of irregular and heterogenous figures. Also, atretic follicles increased in SCI animals, specially on the 14th and 21st day (Figure 1A-F).

Almost all oocytes of SCI rats were pale stained and their nuclei were euchromatic, while the nucleoli were indistinguishable. In granulosa cells, the nucleus was spherical, centric, and euchromatic, and the cells had decreased cytoplasmic volume and eosinophilia (Figure 1C-F).

Degenerative changes including, pyknotic nucleus and vacuolated cytoplasm, were observed in some of the granulosa and theca interna cells. The amount of these cells increased specially on 14th and 21st day (Figure 1C-F). Cellular density, edema, fibrosis, and inflammation increased in ovarian stroma of SCI rats. Dilated and congested vessels were obvious specially in central stroma of the ovary and between theca interna and granulosa layer of early antral follicle and pre-ovulatory follicle (Figure 1C to F).
3.3. Histomorphometric Assessment

The mean diameter of the follicle and ovum and the mean thickness of granulosa layer were significantly decreased in all types of follicles of the SCI group, specially on the 14th and 21st day after the operation compared with the control and sham groups (P < 0.05), (Figure 2A-C).

The mean thickness of theca interna in SF and TF and the mean thickness of zona pellocida in MPF, SF, and TF were significantly decreased in the SCI rats, specially on the 14th and 21st day after the operation compared with control and sham groups (P < 0.05), (Figure 2D and E).

3.4. Follicular Atresia

Atreatic Unilaminar Primary Follicles (UPF) number were significantly more in SCI rats on 7th, 14th and 21st day than the control groups (Table 2). Atreatic Multilaminar Primary Follicles (MPF) number and Secondary Follicles (SF) were significantly more in SCI rats on 14th and 21st day than the control group. Also atreatic Tertiary Follicles (TF) number was significantly more in SCI rats on 21st day than the control groups (Table 2).
4. Discussion

According to the results, animals weight, ovarian weight, and volume of the SCI rats decreased in comparison with Co and Sh rats, that corresponded with other studies. However, decrease in ovarian weight and volume during the acute phase after surgery was not significant.

Since gonadotropins have an important role in growth and development of ovarian follicles and corpus luteum, alteration in levels of gonadotropins after spinal cord transection, may lead to a decrease in ovarian weight and volume (16, 22, 24).

In the rats of the SCI group, severe changes in ovarian stroma, specially on the 21st day after the operation were observed, and the inflammation due to lack autonomic function and hyperprolactinemia after spinal cord transection may have caused these changes. Following the inflammation and edema in ovary, then stromal fibroblasts are activated by transforming growth factor beta (TGF-β), secreted by mast cells, thus collagen production increases and then fibrosis ensues in stroma. On the other hand, alteration in the amount of neurotransmitters and increase in prolactin and LH levels following the spinal cord transection may increase vascular permeability that in turn leads to edema and increase in stromal volume (14, 27).

This study proved that in all of the 3 subdivisions of the SCI group, the oocytes in all types of follicles were less dyeable, their nuclei were euchromatic, and the nucleoli were indistinguishable. Eosinophilia and cytoplasmic volume in granulosa layer and theca interna decreased and their nuclei became pale. Also, some of the cells with dense, small, and sometimes fusiform nuclei (possibly pyknotic cells) were visible.

Previous studies have shown that in addition to hypothalamus-pituitary-ovary axis, other factors includ-
Table 2. Mean Number of Atretic Follicles in Control, Sham and Spinal Cord Injury Groups on the 7th, 14th, and 21st Day After Spinal Cord Transaction (N = 6)³

| Animal Groups | Mean Number of Atretic Follicles / Ovary ± SEM |
|---------------|-----------------------------------------------|
|               | UPF  | MPF  | SF   | TF   |
| 7 days after surgery |     |      |      |      |
| Co            | 13.30 ± 0.84 | 8.40 ± 0.53 | 7.20 ± 0.58 | 4.60 ± 1.12 |
| Sh            | 14.10 ± 0.54 | 9.10 ± 0.45 | 7.80 ± 0.84 | 4.30 ± 0.94 |
| SCI           | 19.40 ± 0.63³ | 14.80 ± 0.78 | 11.20 ± 0.36 | 6.40 ± 0.82 |
| 14 days after surgery |     |      |      |      |
| Co            | 12.40 ± 0.78 | 9.20 ± 0.44 | 7.30 ± 0.82 | 5.40 ± 1.04 |
| Sh            | 12.90 ± 0.66 | 9.80 ± 0.79 | 6.80 ± 0.58 | 4.20 ± 0.93 |
| SCI           | 20.10 ± 0.76³ | 15.60 ± 0.74³ | 10.30 ± 0.47³ | 7.30 ± 0.38 |
| 21 days after surgery |     |      |      |      |
| Co            | 15.60 ± 0.58 | 8.70 ± 0.48 | 6.50 ± 0.44 | 6.30 ± 0.57 |
| Sh            | 14.80 ± 0.94 | 9.10 ± 0.54 | 7.10 ± 0.86 | 5.80 ± 0.82 |
| SCI           | 23.20 ± 1.07³ | 14.30 ± 0.72³ | 12.10 ± 0.92³ | 9.30 ± 0.38³ |

Abbreviations: Co, Control; MPF, Multilaminar Primary Follicle; SCI, Spinal Cord Injury; SF, Secondary Follicle; Sh, Sham; TF, Tertiary Follicle; UPF, Unilaminar Primary Follicle.

³Data are shown as Mean ± SEM, ³P < 0.05 as compared with the control and sham groups.

ing catecholamines and neurotransmitters via stimulation of β-adrenergic receptors in follicular cells and so increase of cAMP, interfere in regulation of follicular growth and development. Furthermore, ovarian nerves (superior ovarian nerve and ovarian plexus) in conjunction with neurotransmitters to cAMP connection and ovarian cholinergic neurons, consisting of Y neuropeptide, intervene in this process (8-11, 15, 28).

Following spinal cord transection, the regulatory role of nerves and neurotransmitters on growth and integrity of ovums, granulosa layer, and theca interna may be disrupted; thus, severe changes, with inhibitory affect on development and differentiation of granulosa layer and theca interna, follow. Finally, these changes lead to pyknosis and increased ovarian heterogeneity.

This study proved that the main parameters of all kinds of follicles such as, the diameter of the follicle and ovum, thickness of granulosa layer, theca interna, and zona pellucida in the SCI group were exposed to severe changes. Increase in diameter of all types of follicles and thickness of granulosa layer, specially in graafian and primary follicles, was more severe in comparison with other parameters.

Since the CNS directly effects hormone regulation and ovarian development via spsific descending and ascending pathways and reasonable levels of gonadotropins are related to impulses from ovarian adrenergic nerves, it may be possible that spinal cord transection disrupts these connections and deranges synthesis and maturation of steroids hormones and also deranges regulation of ovarian hormones and as a result leads to histological and structural changes in all types of follicles (1, 8-10, 13, 29, 30).

4.1. Conclusion

The results suggest that spinal cord transaction severely effects the ovaries and alters histology and structure of ovumary and increases their heterogeneity. These changes are due to disorder of gonadotropin regulation that is a result of autonomic function loss, alteration of catecholamines, and ovarian neurotransmitters level following spinal cord transection.

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Footnotes

Authors’ Contribution: Hamidreza Sameni (Corresponding author) and Sam Zarbakhsh, developed the ideas, performed the research, collected the data, and wrote the manuscript; Behpour Yousefi was the technical assistant.
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