Procalcitonin for diagnosis of bacterial pneumonia in critically ill patients during 2009 H1N1 influenza pandemic: a prospective cohort study, systematic review and individual patient data meta-analysis

Roman Pfister1*, Matthias Kochanek2†, Timo Leygeber1, Christian Brun-Buisson3, Elise Cuquemelle3, Mariana Benevides Paiva Machado4, Enrique Piacentini5, Naomi E Hammond6,7, Paul R Ingram8,9 and Guido Michels1

Abstract

Introduction: Procalcitonin (PCT) is helpful for diagnosing bacterial infections. The diagnostic utility of PCT has not been examined thoroughly in critically ill patients with suspected H1N1 influenza.

Methods: Clinical characteristics and PCT were prospectively assessed in 46 patients with pneumonia admitted to medical ICUs during the 2009 and 2010 influenza seasons. An individual patient data meta-analysis was performed by combining our data with data from five other studies on the diagnostic utility of PCT in ICU patients with suspected 2009 pandemic influenza A(H1N1) virus infection identified by performing a systematic literature search.

Results: PCT levels, measured within 24 hours of ICU admission, were significantly elevated in patients with bacterial pneumonia (isolated or coinfection with H1N1; \( n = 77 \)) (median = 6.2 μg/L, interquartile range (IQR) = 0.9 to 20) than in patients with isolated H1N1 influenza pneumonia (\( n = 84 \); median = 0.56 μg/L, IQR = 0.18 to 3.33). The area under the curve of the receiver operating characteristic curve of PCT was 0.72 (95% confidence interval (CI) = 0.64 to 0.80; \( P < 0.0001 \)) for diagnosis of bacterial pneumonia, but increased to 0.76 (95% CI = 0.68 to 0.85; \( P < 0.0001 \)) when patients with hospital-acquired pneumonia and immune-compromising disorders were excluded. PCT at a cut-off of 0.5 μg/L had a sensitivity (95% CI) and a negative predictive value of 80.5% (69.9 to 88.7) and 73.2% (59.7 to 84.2) for diagnosis of bacterial pneumonia, respectively, which increased to 85.5% (73.3 to 93.5) and 82.2% (68.0 to 92.0) in patients without hospital acquired pneumonia or immune-compromising disorder.

Conclusions: In critically ill patients with pneumonia during the influenza season, PCT is a reasonably accurate marker for detection of bacterial pneumonia, particularly in patients with community-acquired disease and without immune-compromising disorders, but it might not be sufficient as a stand-alone marker for withholding antibiotic treatment.

* Correspondence: roman.pfister@uk-koeln.de
† Equal contributors
1 Department of Internal Medicine III, University of Cologne, Kerpener Str. 62, 50937 Cologne, Germany
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Introduction

In 2009, the pandemic influenza A(H1N1) virus infection caused more than 18,000 deaths and was associated with more than 200,000 estimated respiratory deaths worldwide [1]. Although most H1N1 patients in 2009 and 2010 presented with mild febrile illnesses and cough, sore throat and occasional gastrointestinal symptoms, 7% to 25% of patients developed severe respiratory distress requiring admission to ICUs [2-5]. Bacterial coinfection in ICU patients with 2009 H1N1 influenza infection is reported to be 18% to 34% and is associated with significantly increased mortality [6,7]. Importantly, the initial clinical features of isolated H1N1 infection cannot be distinguished from additional bacterial coinfection [6]. Antibiotic treatment is not recommended in patients with H1N1 infections, but early antibiotic treatment in patients with community-acquired pneumonia is associated with decreased mortality [8]. A rapid diagnosis at admission of bacterial coinfection or isolated bacterial infection is crucial to early initiation or withdrawal of antibiotic treatment. However, obtaining the results cultures for bacterial pathogens may take several days. As a consequence, researchers in several studies have analyzed the predictive value of biomarkers to distinguish bacterial from isolated viral pneumonia [9,10].

Procalcitonin (PCT) may be a clinically useful marker for the diagnosis of bacterial infections [11,12] and may be a useful adjunct for reducing antibiotic usage [13-15]. However, data on the diagnostic value of PCT, including comparisons with other biomarkers such as C-reactive protein (CRP), in critically ill patients in the setting of 2009 H1N1 influenza infection are rare, and the reported results are inconsistent [16-20]. Earlier results of studies on PCT in critically ill patients might not be transferable to pandemic H1N1 influenza, because the latter is associated with a remarkable release of proinflammatory cytokines such as interleukin 6 and a defective T-cell response, both of which might affect PCT release and hence discriminatory performance of PCT in diagnosing bacterial infection [21,22]. Conflicting results of studies of PCT in critically ill patients with suspected 2009 H1N1 influenza are most likely due to the small sample sizes and substantial differences in populations studied. More powerful analyses are needed to address the role of potential confounders of the diagnostic accuracy of PCT, such as antibiotic pretreatment, underlying impairment of the immune system and hospital-acquired infections.

Our aims in the present study were to test the utility of PCT in the diagnosis of bacterial pneumonia in patients admitted to the ICU during the 2009 H1N1 influenza pandemic, including its performance in important clinical subgroups, and to compare its performance with CRP. We performed a prospective cohort study in patients admitted to a 28-bed ICU of a tertiary care hospital with pneumonia during the 2009 and 2010 influenza seasons and then combined our data with individual patient data derived from published studies identified by conducting a systematic literature search and performed a meta-analysis.

Material and methods

Prospective cohort study

The study population was patients admitted to a medical ICU at a tertiary care hospital in Cologne, Germany, during the pandemic influenza A(H1N1) season between November 2009 and January 2010 as well as those admitted between December 2010 and May 2011. All adult patients (older than 18 years of age) with community-acquired pneumonia and visual evidence of a pulmonary infiltrate on a chest X-ray or computed tomography scan were included. In addition, we included patients with hospital-acquired pneumonia admitted to the ICU during a local outbreak of H1N1 influenza within our hospital during the study time periods. Hospital-acquired pneumonia was defined as disease that manifested later than 48 hours after hospital admission. H1N1 status was assessed by PCR in every patient using nasopharyngeal swabs or bronchoalveolar lavage fluid upon admission to the ICU. Standard diagnostic tests for bacterial pathogens were performed as part of the routine clinical assessment universally applied to all consecutive patients. Bacterial pneumonia was defined by detection of a bacterial pathogen compatible with pneumonia infection using blood cultures, urinary antigen tests (for Pneumococcus and Legionella), PCR (for Chlamydia and Mycoplasma) and/or cultures of respiratory tract secretions (tracheobronchial aspirate or bronchoalveolar lavage). PCT and CRP were measured using the ELECSYS B•R•A•H•M•S PCT electrochemiluminescence assay (Fisher Scientific, Schwerte, Germany) and the TinaQuant C-Reactive Protein Gen 3 assay (Roche Diagnostics, Mannheim, Germany) according to the treating physician’s discretion. Impairment of the immune system was defined by the presence of active leukemia or lymphoma or if myelotoxic chemotherapy was applied. The study was approved by the ethics committee of the medical faculty of the University of Cologne, and informed consent was waived because all data assessed in the study were part of routine clinical practice and retrieved from patient records without any study intervention.

Systematic review

In accordance with PRISMA recommendations (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) [23], we performed an electronic literature search using PubMed to identify studies reporting the performance of PCT in differentiating 2009 H1N1 influenza
from bacterial pneumonia in adult ICU patients. Restrictions regarding language and publication year were not applied, and publications up until June 2013 were included. The following search terms were used: “procalcitonin” in combination with “H1N1,” “influenza,” “swine flu” or “avian flu.” Two independent reviewers (TL and RP) screened the electronic search results for eligible articles by reading the titles and abstracts. Inclusion criteria for in-depth examinations were the evaluation of PCT alone or in comparison with other laboratory markers, such as CRP, to diagnose H1N1 influenza or bacterial pneumonia in adult patients with respiratory tract infections admitted to ICUs. References to other sources within the selected articles were researched for other eligible articles.

For all eligible articles, we contacted the authors by e-mail and invited them to share their data in this collaborative project. Data sets needed to include at least the following variables: anonymous patient identifiers and patient characteristics obtained from clinical history (gender, age, antibiotics given before ICU admission, community- or hospital-acquired pneumonia, underlying impairment of the immune system and results of testing for H1N1 and bacteria). Because individual studies used different risk scores to assess the severity of critical illness and mortality, we defined predicted high risk using cutoffs of respective risk scores which might be associated with an in-hospital mortality rate greater than 15%: Sequential Organ Failure Assessment score higher than 7, Simplified Acute Physiology Score II (SAPS II) higher than 35, SAPS III score above 47 and Acute Physiology and Chronic Health Evaluation II score higher than 15.

All individual studies were approved by the respective local ethics committees where appropriate (Ethics Review Board of the Société de Réanimation de langue Française [16], the ethical committee of Hospital Universitario Mútua de Terrassa [20], the Prince Charles Hospital Human Research Ethics Committee [17] and the ethics committee of Universidade Federal de Minas Gerais [19]). Approval was not required in the study by Ingram et al. [18].

Statistical analysis
We merged the data of our cohort and the previously published cohorts into a summary database if variables were compatible. Incompatible data were first recoded and then added to the summary database. We included into the analysis only patients with PCT measured within the first 24 hours after admission to the ICU and with either confirmed H1N1 influenza or microbiologically confirmed bacterial pneumonia, or both. If more than one measure of PCT or CRP was available within 24 hours after admission, we used the first measure performed after admission for analysis. Continuous variables are expressed as median and interquartile range (IQR) or as mean ± standard deviation (SD) as appropriate. Categorical variables are presented as absolute and relative frequencies. Pairwise comparisons for categorical variables were performed by using a χ² test. For continuous variables, we used a nonparametric Mann–Whitney U test. The diagnostic discrimination of inflammatory markers for diagnosis of microbiologically confirmed bacterial pneumonia (combining patients with isolated bacterial pneumonia with patients with mixed bacterial and H1N1 pneumonia) from isolated H1N1 pneumonia is expressed by receiver operating characteristic curve (ROC) data. Sensitivity, specificity, positive predictive value and negative predictive value were calculated. Because distinct population-specific issues might influence the discriminatory performance of PCT, we performed sensitivity analyses in subgroups without impairment of the immune system, with community-acquired pneumonia, with typical bacterial pathogens (excluding Mycoplasma, Chlamydia and Legionella infections), without extracorporeal membrane oxygenation (ECMO) and renal replacement therapy and without antibiotics given before ICU admission. P-values less than 0.05 were considered to be statistically significant. All reported P-values are two-sided, except for comparison of areas under the curve (AUCs). Statistical analyses were performed with Stata/SE 12.1 for Windows software (StataCorp, Austin, TX, USA) and SPSS Statistics version 21 software (IBM SPSS, Chicago, IL, USA).

Results
Prospective cohort study
Table 1 shows the baseline characteristics of our cohort. The mean age (±SD) of the patients was 56 (±17) years, and 50% of the patients were male. Sixty-one percent of the patients had bacterial pneumonia, 33% were being treated with antibiotics before ICU admission and 57% were positive for H1N1. Overall, 67% of the patients had hospital-acquired pneumonia and 35% had immune-compromising disorders. All patients had at least one measure of PCT within the first 24 hours after admission to the ICU.

Systematic review
A flowchart depicting the literature search process and inclusion of studies for our meta-analysis is shown in Figure 1. We identified 30 potentially relevant titles on MEDLINE that reported on utilizing PCT in adult patients with suspected influenza pneumonia. From among these 30 articles, 16 articles were excluded after the abstract was read because they were reviews without original data (n = 7), were comments without original data (n = 3) or were focused on other topics (for example, pediatric patients and outcomes without information on bacterial infection or vaccination; n = 6). Upon reading
the full text of the remaining 14 studies, we found that 8 studies were on non-ICU patients and one did not provide data on bacterial infection; therefore, they were excluded, leaving us with 5 studies for analysis. Checks of for cross-references of selected articles identified no other applicable studies. The authors of all five included studies responded after we contacted them by e-mail, and they sent us their primary data. The characteristics of these five studies are shown in Table 2. The authors in all five of the included studies examined patients with community-acquired pneumonia, whereas we examined mixed community- and hospital-acquired pneumonia patients in our study.

When we combined our cohort with the data derived from the other five studies, we had a total of 242 patients. We excluded 60 patients with missing PCT measured within the first day of ICU admission and 21 patients with neither confirmation of H1N1 influenza infection nor confirmation of a bacterial pathogen causative of infection. Thus we had 161 patients remaining for inclusion in the analyses. Patients who were excluded (n = 81) did not differ significantly regarding age, immune-compromising disorder, H1N1 status, bacterial infection rate or CRP level (all P > 0.05); however, compared to patients who were included, the excluded group had a higher ratio of women, with borderline statistical significance (60% vs. 46%; P = 0.03), and a lower ratio of antibiotics given before ICU admission (19% vs. 38%; P = 0.004).

Table 1 Characteristics of our prospective cohort study

| Characteristics | Patient data (n = 46) |
|-----------------|----------------------|
| Gender (male)   | 23 (50%)            |
| Age (years), mean ±SD | 56 ± 17          |
| H1N1 status     | 26 (57%)            |
| Death in ICU    | 15 (33%)            |
| TISS-28, median (IQR) | 21 (19 to 25) |
| SAPS II, median (IQR) | 51 (41 to 60) |
| APACHE II, median (IQR) | 22 (17 to 26) |
| SOFA, median (IQR) | 9 (7 to 11)     |
| Asthma          | 2 (4.3%)            |
| COPD            | 11 (24%)            |
| Smokers         | 9 (20%)             |
| Alcoholic disease | 5 (11%)         |
| Diabetes mellitus | 12 (26%)        |
| Immune-compromising disorder | 16 (35%)   |
| Hospital-acquired pneumonia | 31 (67%)   |
| Bacterial pathogen | 28 (61%)      |
| Pneumococcus    | 6                   |
| Chlamydia/Mycoplasma | 3               |
| Legionella      | 2                   |
| Gram-negative bacilli (Pseudomonas aeruginosa, Klebsiella, Morganella morgan) | 9 |
| Staphylococcus aureus | 4               |
| Other gram-positive cocci (Enterococci, coagulase-negative Staphylococci) | 4 |
| Antibiotics given during ICU | 31 (67%) |
| Antibiotics given before ICU | 15 (33%) |
| Mechanical ventilation | 34 (74%) |
| Length of ventilation (days), median (IQR) | 7 (1 to 11) |
| ECMO            | 5 (11%)             |
| Renal replacement therapy | 9 (19.6%) |
| Vasopressor therapy | 30 (65%)          |
| Length of stay (days), median (IQR) | 9 (5 to 16) |
| Procalcitonin (µg/L) at admission (n = 30) | 0.85 (0.3 to 4.1) |
| Procalcitonin (µg/L) within day 1 (n = 30) | 0.9 (0.2 to 6) |
| C-reactive protein (mg/L) at admission (n = 40) | 159 (61 to 253) |
| C-reactive protein (mg/L) within day 1 (n = 30) | 101 (62 to 208) |

aAPACHE II, Acute Physiology and Chronic Health Evaluation II; COPD, Chronic obstructive pulmonary disease; ECMO, Extracorporeal membrane oxygenation; IQR, Interquartile range; SAPS II, Simplified Acute Physiology Score II; SD, Standard deviation; SOFA, Sequential Organ Failure Assessment; TISS-28, 28-item Therapeutic Intervention Scoring System. bAssessed at ICU admission.
| Study                  | Year | PCT available n/N (assay) | Study design | Inclusion criteria                          | H1N1-positive (%)<sup>b</sup> | Bacterial infection (%)<sup>b</sup> | Hospital-acquired pneumonia (%)<sup>b</sup> | Antibiotics before admission (%)<sup>b</sup> | Immune disorder (%)<sup>b</sup> |
|------------------------|------|--------------------------|--------------|---------------------------------------------|-----------------------------|--------------------------------------|--------------------------------------------|---------------------------------|-----------------------------|
| Piacentini et al. [20] | 2010 | 22/22 (not reported)     | Prospective  | Adults, ICU, community-acquired pneumonia   | 45                          | 50 (cultures, antigen)               | 0                                          | 100                             | 5                           |
| Hammond et al. [17]    | 2011 | 17/17 (VIDAS BRAHMS PCT) | Retrospective | Adults, ICU, respiratory tract infection    | 82                          | 41 (cultures)                       | 0                                          | n.a.                            | 0                           |
| Cuquemelle et al. [16] | 2011 | 52/103 (BRAHMS KRYPTOR PCT) | Prospective  | Adults, ICU, community-acquired pneumonia, H1N1-positive | 100                        | 37 (cultures, antigen)               | 0                                          | 0                               | 6                           |
| Ingram et al. [18]     | 2010 | 16/25 (VIDAS BRAHMS PCT) | Retrospective | Adults, ICU, community-acquired pneumonia, positive for H1N1 or bacterial infection | 63                         | 38 (cultures, seroconversion)        | 0                                          | 94                              | 6                           |
| Paiva et al. [19]      | 2012 | 29/29 (VIDAS BRAHMS PCT) | Prospective  | Adults, ICU, pneumonia, suspected H1N1     | 41                          | 21 (cultures)                       | 0                                          | 41                              | 14                          |

<sup>a</sup>PCT, Procalcitonin. <sup>b</sup>Referring to patients with available procalcitonin.
Table 3 shows the characteristics of the study population divided by bacterial infection status. Eighty-four patients had isolated H1N1 infections and seventy-seven had bacterial infections (sixty-nine with typical bacterial pathogens and eight with atypical bacterial pathogens), with thirty-seven patients having isolated bacterial infections and forty having mixed H1N1 and bacterial infections. One hundred forty-three patients also had CRP measured within the first 24 hours after admission. The baseline characteristics did not differ significantly between patients with vs. without bacterial infections, except for H1N1 status, high-risk of mortality predicted on the basis of ICU risk scores and the inflammatory markers PCT and CRP.

Figure 2A shows PCT and CRP levels by status of bacterial infection. Patients with bacterial pneumonia had significantly elevated PCT levels (6.2 μg/L; IQR = 0.9 to 20) compared to those with isolated H1N1 pneumonia (0.56 μg/L; IQR = 0.18 to 3.3, P = 0.0001). There was no significant difference in PCT levels between patients with isolated bacterial pneumonia and mixed bacterial and H1N1 pneumonia (P = 0.14), as shown in Additional file 1. Both isolated bacterial pneumonia patients (3.63 μg/L; IQR = 0.63 to 18.2, P = 0.002) and mixed bacterial and H1N1 pneumonia patients (11.36 μg/L; IQR = 1.07 to 41.14, P = 0.0001) had significantly elevated PCT levels compared to those with isolated H1N1 pneumonia (0.56 μg/L; IQR = 0.18 to 3.33) (see Additional file 1).

Patients with bacterial pneumonia had significantly elevated CRP levels (214 mg/L; IQR = 81 to 335) compared with patients with isolated H1N1 pneumonia (108 mg/L; IQR = 50 to 213, p = 0.003) (Figure 2B). Neither PCT levels (2.21 μg/L; IQR = 0.55 to 12.6 vs. 1.2 μg/L; IQR = 0.3 to 11.9, P = 0.34) nor CRP levels (175 mg/L; IQR = 77 to 311 vs. 142 mg/L, IQR = 58 to 278, P = 0.14) were significantly different between patients who died during their hospital stay and those who survived.

Figure 3 shows the ROC curves and Table 4 shows the discriminatory performance of PCT for detection of bacterial pneumonia. The AUC of the ROC curve of PCT in the total population was 0.72 (95% CI = 0.64 to 0.80; P < 0.0001). The results were similar when only PCT levels measured immediately upon admission to the ICU were considered (data not shown). When we restricted the analysis to bacterial infections with typical pathogens, excluding Mycoplasma, Chlamydia and Legionella, the AUC was 0.73 (95% CI = 0.64 to 0.81; P < 0.0001). Exclusion of patients who received antibiotic treatment before...
ICU admission or patients with ECMO or renal replacement therapy did not change the discriminatory performance of PCT substantially. When we excluded patients with hospital-acquired pneumonia and patients with immune-compromising disorders, the AUC increased to 0.76 (95% CI = 0.68 to 0.85; \( P < 0.0001 \)), which was not significantly different from the overall population (\( P \)-value for comparison = 0.24). The discriminatory performance of CRP assessed according to the AUC (0.64; 95% CI = 0.55 to 0.73) was lower than that of PCT, which was of borderline statistical significance (\( P = 0.10 \)).

A cutoff PCT level of 0.5 μg/L showed a sensitivity and negative predictive value of 80.5% (95% CI = 69.9 to 88.7) and 73.2% (95% CI = 59.7 to 84.2), respectively, for detection of a bacterial pneumonia (Table 5). The accuracy improved when we excluded patients with hospital-acquired pneumonia or immune-compromising disorder, with a respective sensitivity and negative predictive value of 85.5% (95% CI = 73.3 to 93.5) and 82.2% (95% CI = 68.0 to 92.0).

**Discussion**

By utilizing an individual patient data meta-analytic approach in this study, we show that, among patients admitted to the ICU with pneumonia during the H1N1 pandemics in 2009 and 2010, PCT levels were significantly increased in patients with bacterial pneumonia compared to those with isolated H1N1 pneumonia. The overall discriminatory performance of PCT for identification of bacterial pneumonia was moderate, with an

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**Table 4** Discriminatory accuracy of procalcitonin and C-reactive protein for detection of bacterial pneumonia

| Characteristics                                      | AUC     | 95% CI      | \( P \)-values | \( P \)-value vs. total population |
|-------------------------------------------------------|---------|-------------|-----------------|----------------------------------|
| Total (N = 161)                                       | 0.72    | 0.64 to 0.80| <0.0001         | –                                |
| Without antibiotics before admission (n = 91)         | 0.73    | 0.62 to 0.83| <0.0001         | 0.47                             |
| Without ECMO or renal replacement therapy (n = 110)   | 0.74    | 0.64 to 0.83| <0.0001         | 0.40                             |
| Without hospital-acquired pneumonia (n = 131)         | 0.76    | 0.68 to 0.84| <0.0001         | 0.25                             |
| Without immune-compromising disorder (n = 140)        | 0.74    | 0.65 to 0.82| <0.0001         | 0.40                             |
| Without hospital-acquired pneumonia and               | 0.76    | 0.68 to 0.85| <0.0001         | 0.24                             |
| Without immune-compromising disorder (n = 124)        |         |             |                 |                                  |
| CRP (total population, n = 143)                       | 0.64    | 0.55 to 0.73| 0.003           | 0.10                             |

*\( AUC \), Area under the curve; CI, 95% confidence interval; CRP, C-reactive protein; ECMO, extracorporeal membrane oxygenation.
| Cutoff      | Sensitivity, % (95% CI) | Specificity, % (95% CI) | Positive PV, % (95% CI) | Negative PV, % (95% CI) | Positive likelihood ratio (95% CI) | Negative likelihood ratio, % (95% CI) |
|------------|------------------------|-------------------------|------------------------|------------------------|-----------------------------------|----------------------------------------|
| Total      |                        |                         |                        |                        |                                   |                                        |
| >0.25 µg/L | 87.0 (77.4 to 93.6)    | 28.6 (19.2 to 39.5)     | 52.8 (43.7 to 61.7)    | 70.6 (52.5 to 84.9)    | 1.22 (1.04 to 1.43)               | 0.45 (0.23 to 0.89)                   |
| >0.5 µg/L  | 80.5 (69.9 to 88.7)    | 48.8 (37.7 to 60.0)     | 59.1 (49.0 to 68.6)    | 73.2 (59.7 to 84.2)    | 1.57 (1.24 to 1.99)               | 0.40 (0.24 to 0.66)                   |
| >1 µg/L    | 71.4 (60.0 to 81 to 2) | 59.5 (48.3 to 70.1)     | 61.8 (50.9 to 71.9)    | 69.4 (57.5 to 79.8)    | 1.76 (1.31 to 2.37)               | 0.48 (0.32 to 0.71)                   |
| Without hospital-acquired pneumonia or immune-compromising disorder | | | | | | |
| >0.25 µg/L | 90.9 (80.1 to 97.0)    | 31.9 (21.2 to 44.2)     | 51.6 (41.2 to 61.8)    | 81.5 (61.9 to 93.7)    | 1.33 (1.11 to 1.60)               | 0.29 (0.12 to 0.70)                   |
| >0.5 µg/L  | 85.5 (73.3 to 93.5)    | 53.6 (41.2 to 65.7)     | 59.5 (47.9 to 70.4)    | 82.2 (68.0 to 92.0)    | 1.84 (1.40 to 2.43)               | 0.27 (0.14 to 0.53)                   |
| >1 µg/L    | 74.6 (61.0 to 86.3)    | 60.9 (48.4 to 72.4)     | 60.3 (47.7 to 72.0)    | 75.0 (61.6 to 85.6)    | 1.91 (1.37 to 2.66)               | 0.42 (0.26 to 0.68)                   |

*Ci, Confidence interval; PV, Predictive value.
AUC of 0.72, and seemed to be negatively affected by the presence of hospital-acquired infections and immune-compromising disorders. The AUC increased to 0.76 when these conditions were excluded from the analysis, and PCT at a cutoff of 0.5 μg/L had a sensitivity and negative predictive value of 85.5% (95% CI = 73.3 to 93.5) and 82.2% (95% CI = 68.0 to 92.0), respectively.

Patients with 2009 H1N1 infection and potential bacterial coinfection admitted to an ICU are at high risk for mortality [3-6,24]. Early differentiation of isolated viral infection from isolated or associated bacterial infection might help in the decision to initiate appropriate antimicrobial treatment or avoid unnecessary treatment. To date, studies evaluating PCT in ICU patients with suspected or confirmed 2009 H1N1 influenza have included 16 and 52 patients, and, accordingly, they have major inherent limitations [16-20]. Owing to small sample sizes, differences in inclusion criteria, patient selection and outcome definitions, the discriminatory capacity of PCT appears to be unclear [17,19], with researchers in some studies reporting good discriminatory performance of PCT [16,18,20]. Authors of a recent meta-analysis examined the value of PCT for the detection of bacterial coinfection in patients with influenza [25]. Of note, only two of the six studies included in that meta-analysis exclusively comprised ICU patients. Both of these studies were included in our analyses [16,18]. Additionally, the authors of that meta-analysis did not use individual patient data, but instead reported only meta-analysis of summary statistics such as AUC, sensitivity and specificity.

Our collaborative approach yielded a sample of 161 patients, which is three times larger than any earlier study on this ICU collective and allowed us to estimate discriminatory performance more robustly than other studies, not only in the overall population but also in important subgroups. We demonstrate improved accuracy when patients with compromised immune systems and hospital-acquired disease are excluded, albeit that this difference did not reach statistical significance, most likely because of the limited sample size. The first finding might be explained by a trend toward overall lower PCT levels in immune-compromised patients [26]. Regarding the latter point, PCT is known to have weaker diagnostic utility in patients with nosocomial pneumonia because hospitalized patients generally have higher exposure to bacterial infections independently of potential pneumonia [27,28]. Furthermore, we provide a set of cutoffs for PCT for detection or exclusion of bacterial infection in ICU patients with potential 2009 H1N1 influenza. Three of the earlier studies did not report any cutoff, owing to weak overall discrimination of PCT or small sample size, and the two largest studies reported a cutoff of 0.8 μg/L. This cutoff is in between the generally accepted levels in ICU patients with respiratory tract infections [29], and the generalizability might be limited by the small and selected derivation sample. We show that recommended rule-out and rule-in cutoffs (0.25, 0.5 and 1 μg/L) for bacterial infection and, as a consequence, for antibiotic treatment discriminate moderately in patients with suspected 2009 H1N1 influenza infection. In patients with community-acquired disease without immune-compromising disorder, PCT might be helpful in ruling out bacterial infection or coinfection, with a negative predictive value of 82.2% and a negative likelihood ratio of 0.27. However, this negative predictive value still seems too low to use PCT as a stand-alone marker for withholding antibiotic therapy in such critically ill patients. Serial changes in PCT might further improve discrimination of bacterial pneumonia, but we had serial PCT measures in only 40% of our population, which is not sufficient for analysis. Thus, further study is needed to evaluate the clinical benefit of PCT testing and to define its role in an algorithm for antimicrobial therapy in ICU patients with suspected H1N1 influenza pneumonia.

In our patient population, PCT showed better discrimination than CRP for bacterial infection, albeit the difference was of borderline significance because of the small sample size. This finding is in line with a body of evidence supporting the superiority of PCT over CRP for detection of bacterial infection [30]. PCT is upregulated through stimulation by cytokines released in response to bacterial infection, and, in contrast to CRP, is inhibited through interferon γ, which is released in viral infections, thus exhibiting a strong specificity for bacterial infections [31].

Our study has some limitations. Despite the combination of six cohorts, our total sample size is still limited and thus subgroup analyses must be interpreted cautiously. Furthermore, half of the studies were retrospective and the study populations were variably selected, which limit the generalizability of our results and might have introduced bias. For instance, one-third of patients did not have PCT measured within the first 24 hours after admission. However, we show that these patients with missing PCT data were not significantly different from the patients included in the analysis regarding most baseline characteristics. A major limitation, which applies to almost all observational studies evaluating PCT, is that there is no accurate gold standard for the outcome definition of bacterial infection. The intensity of diagnostics have differed across studies and was clinically driven in all studies but ours and in the one by Piacentini et al. [20], who performed systematic microbiological testing on blood, urinary and tracheobronchial secretions in all patients. Nonetheless, microbiological cultures of bacterial pathogens are generally inaccurate, particularly in cases of respiratory infections [32,33]. Thus, underdiagnosing of bacterial infection might be common and thus might attenuate the resulting accuracy of PCT as a marker. Hence,
to definitively assess the value of PCT in patients with possible H1N1 infection, a randomized trial on the clinical outcome of PCT-guided antimicrobial treatment is needed. Furthermore, our findings are not generalizable to other clinical scenarios, such as in the emergency department and cases of interpandemic pneumonia or other viral strains causing pneumonia, because the inflammatory response, including PCT, might differ, depending on clinical severity and the underlying viral pathogen [34]. We do not have information on the time from symptom onset to admission to the ICU, which might affect not only the risk of bacterial coinfection but also absolute PCT levels because the kinetics of PCT differ between viral and bacterial infections [20]. Hence the discriminatory performance of PCT might vary over time since the onset of infection. This factor might lead to underestimation of overall PCT accuracy.

Conclusions

In patients admitted to the ICU with suspected H1N1 pneumonia, PCT is a sensitive marker with good negative predictive value for the identification of bacterial infection and is superior to CRP. The accuracy of PCT was highest in patients with community-acquired disease and in patients who are not immunocompromised. However, a negative predictive value of 82% might not be sufficient to use PCT as a stand-alone marker for withholding antibiotic treatment, and thus further studies are needed to define the clinical benefit of PCT testing in ICU patients during H1N1 epidemics.

Key messages

- PCT shows good sensitivity and negative predictive value for bacterial pneumonia in critically ill patients during H1N1 influenza epidemics.
- The discrimination accuracy of PCT was best in patients with community-acquired pneumonia and in patients without immune-compromising disorders.
- The negative predictive value of PCT for bacterial pneumonia might not be sufficient to use it as a stand-alone marker for withholding antibiotics.

Competing interests

The authors declare that they have no competing interests.

Authors’ contributions

RP, MK and GM conceived of the study, participated in its design and coordination and drafted the manuscript. MK and TL designed the prospective cohort study in Cologne, screened patients and acquired and analyzed the data. RP, MK, TL and GM performed statistical analyses. CBB, EC, MBPM, EP, NH and PI acquired and analyzed data from the individual cohorts. TL, CBB, EC, MBPM, EP, NH and PI critically revised the manuscript for important intellectual content. All authors read and approved the final manuscript.

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Author details

1Department of Internal Medicine III, University of Cologne, Kerpener Str. 62, 50937 Cologne, Germany. 2Department of Internal Medicine I, University of Cologne, Cologne, Germany. 3Medical Intensive Care Unit, Hôpitaux Universitaires Henri Mondor, Assistance Publique-Hôpitaux de Paris, Université Paris-Est Créteil, Créteil, France. 4Hospital das Clínicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil. 5Department of Critical Care Medicine, Hospital Universitari Mutua Terrassa, Terrassa, Spain. 6Department of Intensive Care Medicine, Royal North Shore Hospital, Sydney, Australia. 7The George Institute for Global Health, Sydney, Australia. 8Department of Microbiology and Infectious Diseases, Royal Perth Hospital, Perth, Australia. 9School of Pathology and Laboratory Medicine, University of Western Australia, Perth, Australia.

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References

1. Dawood FS, Iuliano AD, Reed C, Meltzer MI, Shay DK, Cheng PY, Bandaranayake D, Breiman RF, Brooks WA, Buchy P, Feikin DR, Fowler KB, Gordon A, Hien NT, Horby P, Huang QS, Katz MA, Krishnan A, Lal R, Montgomery JM, Melbak K, Pebody R, Presanis AM, Razun H, Steens A, Tinoco YO, Wallinga J, Yu H, Vong S, Breshe E, et al. Estimated global mortality associated with the first 12 months of 2009 pandemic influenza A H1N1 virus circulation: a modelling study. Lancet Infect Dis 2012, 12:687–695. A published erratum appears in Lancet Infect Dis 2012, 12:655.

2. Cao B, Li XW, Mao Y, Wang J, Lu HZ, Chen YS, Liang ZA, Liang L, Zhang SJ, Zhang B, Gu L, Lu LH, Wang YF, Wang C, National Influenza A Pandemic H1N1 2009 Clinical Investigation Group of China: Clinical features of the initial cases of 2009 pandemic influenza A (H1N1) virus infection in China. N Engl J Med 2009, 361:2507–2517.

3. Domínguez-Cherit G, Lapinsky SE, Macias AE, Pinto R, Espinosa-Perez L, de la Torre A, Poblano-Morales M, Baltazar-Torres JA, Bautista E, Martínez A, Rivera E, Valdez R, Ruiz-Palacios G, Hernández M, Stewart TE, Fowler RA: Critically ill patients with 2009 influenza A(H1N1) in Mexico. JAMA 2009, 302:1880–1887.

4. Jain S, Kaminoto L, Bramley AM, Schmitz AM, Benoit SR, Louie J, Sugerman DE, Druckenmiller JK, Ritter KA, Chugh R, Juska S, Deutscher M, Chen S, Walker JD, Duchin JS, Lert S, Sulkova S, Wells EV, Swerdlow D, Uyeki TM, Fiore AE, Olzen SJ, Fry AM, Bridges CB, Finelli L, Pandemic Influenza A (H1N1) 2009 Hospitalizations Investigation Team: Hospitalized patients with 2009 H1N1 influenza in the United States, April–June 2009. N Engl J Med 2009, 361:1935–1944.

5. Rello J, Rodríguez J, Ibañez P, Socas L, Cebrian J, Marques A, Guerrero J, Ruiz-Santana S, Marquez E, Del Nogal-Saez F, Alvarez-Lerma F, Martinez A, Ferrer M, Avellanas M, Granada R, Marani-Poma A, Albert P, Sierra R, Vidaur L, Ortiz P, Prieto Del Portillo I, Galian B, Léon-Gil C, H1N1 SEMICYUC Working Group: Intensive care adult patients with severe respiratory failure caused by influenza A (H1N1) in Spain. Crit Care 2009, 13:R184.

6. Rice TW, Robinson L, Uyeki TM, Vaughn FL, John BB, Miller RR 3rd, Higgs E, Randolph AG, Smoot BE, Thompson BT, the NHLBI ARDS Network: Critical illness from 2009 pandemic influenza A virus and bacterial coinfection in the United States. Crit Care Med 2012, 40:1487–1498.

Abbreviations

CRP: C-reactive protein; PCT: Procalcitonin; ROC: Receiver operating characteristic curve.

Additional file

Additional file 1: Boxplot showing procalcitonin levels in isolated H1N1, isolated bacterial pneumonia and mixed bacterial and H1N1 pneumonia. Procalcitonin levels are significantly increased in patients with isolated bacterial pneumonia and mixed bacterial and H1N1 pneumonia compared to patients with isolated H1N1 pneumonia, with no significant difference between patients with isolated bacterial pneumonia and mixed bacterial and H1N1 pneumonia.
7. Webb SAR, Pettivia V, Seppelt I, Belloso R, Bailey M, Cooper DJ, Cretikos M, Davies AR, Finfer S, Harrigan PWJ, Hart G, Howe B, Iredell JR, McArthur C, Mitchell I, Morrison S, Nichol AD, Paterson DL, Peake S, Richards B, Stephens D, Turner A, Yung M, the Australian and New Zealand Intensive Care Society (ANZICS) Influenza Investigators: Critical care services and 2009 H1N1 influenza in Australia and New Zealand. N Engl J Med 2009, 361:1925–1934.

8. Houck PM, Bratzer DW, Na W, Ma A, Bartlett JG: Antibiotic administration in community-acquired pneumonia. Chest 2004, 126:320–321.

9. Niederman MS: Biological markers to determine eligibility in trials for community-acquired pneumonia: a focus on procalcitonin. Clin Infect Dis 2008, 47:5127–5132.

10. Simon L, Gauvin F, Amre DK, Saint-Louis P, Lacroix J: Procalcitonin: an antibiotic marker for sepsis: a systematic review and meta-analysis. Clin Infect Dis 2004, 39:206–217.

11. Boussekey N, Leroz O, Georges H, Devos P, d’Escrivan T, Guery B: Diagnostic and prognostic values of admission procalcitonin levels in community-acquired pneumonia in an intensive care unit. Infection 2005, 33:257–263.

12. Wacker C, Prkno A, Brunkhorst FM, Schlattmann P: Antibiotic therapy decisions: a systematic review of randomized controlled trials and recommendations for clinical algorithms. Arch Intern Med 2011, 171:1322–1331.

13. Christ-Crain M, Stolz D, Bingisser R, Müller C, Miedinger D, Huber PR, Zimmerli W, Harbarth S, Tamim M, Müller B: Procalcitonin guidance of antibiotic therapy in community-acquired pneumonia: a randomized trial. Am J Respir Crit Care Med 2006, 174:84–93.

14. Schuetz P, Chiappa V, Briel M, Greenwald JL: Procalcitonin algorithms for antibiotic therapy decisions: a systematic review of randomized controlled trials and recommendations for clinical algorithms. Arch Intern Med 2011, 171:1322–1331.

15. Nobre V, Harbarth S, Graf JD, Rohner P, Pugin J: Use of procalcitonin to shorten antibiotic treatment duration in septic patients: a randomized trial. Am J Respir Crit Care Med 2008, 177:498–505.

16. Cuquerelle E, Soulis F, Villers D, Roche-Campf F, Ara Sornho S, Fartoukh M, Koutchet A, Mouvier B, Dellamonica J, Picard W, Schmidt M, Boulain T, Brun-Buisson C, A/H1N1 REVA-SRLF Study Group: Can procalcitonin help identify associated bacterial infection in patients with severe influenza pneumonia? A multicentre study. Intensive Care Med 2011, 37:976–800.

17. Hammond NE, Cotley A, Fraser JF: The utility of procalcitonin in diagnosis of H1N1 influenza in intensive care patients. Ann Intern Med 2011, 135:299–301.

18. Ingram PR, Inglis T, Moxon D, Speers D: Procalcitonin and C-reactive protein in severe 2009 H1N1 influenza infection. Intensive Care Med 2010, 36:528–532.

19. Paiva MBS, Botoni FA, Teixeira AL Jr, de Miranda AS, de Oliveira CR, de Oliveira Abraham J, Faria GM, Nobre V: The behavior and diagnostic utility of procalcitonin and five other inflammatory molecules in critically ill patients with respiratory distress and suspected 2009 influenza A H1N1 infection. Clinics (Sao Paulo) 2012, 67:327–334.

20. Paciementi E, Sánchez B, Araujo V, Calbo E, Cuchi E, Nava JM: Procalcitonin levels are lower in intensive care unit patients with H1N1 influenza A virus pneumonia than in those in community-acquired bacterial pneumonia: a pilot study. J Crit Care 2011, 26:201–205.

21. Almansa R, Bememojo-Martin IF, Ontz de Lejarazu Leonardo R: [Immunopathogenesis of 2009 pandemic influenza] [in Spanish]. Enferm Infecc Microbiol Clin 2012, 30:18–24.

22. Lee N, Wong CK, Chan PKS, Chan MCW, Wong RYK, Lun SWM, Ngai KKL, Lui GCY, Wong BCK, Lee SKW, Choi KW, Hui DSC: Cytokine response patterns in severe pandemic 2009 H1N1 and seasonal influenza among hospitalized adults. PLoS One 2011, 6:e26305.

23. Moher D, Liberati A, Tetzlaff J, Altman DG, the PRISMA Group: Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med 2009, 6:e1000097.

24. Dawood FS, Jain S, Finelli L, Shaw MW, Lindstrom S, Garten RJ, Gubareva LV, Xu X, Bridges CB, Uyeki TM, the Novel Swine-Origin Influenza A (H1N1) Virus Investigation Team: Emergence of a novel swine-origin influenza A (H1N1) virus in humans. N Engl J Med 2009, 360:2604–2615. A published erratum appears in N Engl J Med 2009, 360:102.

25. Wu MH, Lin CC, Huang SL, Shih HM, Wang CC, Lee CC, Wu JY: Can procalcitonin tests aid in identifying bacterial infections associated with influenza pneumonia? A systematic review and meta-analysis. Influenza Other Respir Viruses 2013, 7:549–555.