Intraspecific variation in the cochleae of harbour porpoises (*Phocoena phocoena*) and its implications for comparative studies across odontocetes

Maria Clara Iruzun Martins Corresp., 1, 2, Travis Park 2, 3, Rachel Racicot 4, 5, 6, 7, Natalie Cooper 2

1 Division of Biosciences, University College London, University of London, London, United Kingdom
2 Department of Life Sciences, Natural History Museum, London, United Kingdom
3 Department of Earth Sciences, University of Oxford, Oxford, United Kingdom
4 Forschungsinstitut und Naturkundemuseum, Senckenberg der SNG, Frankfurt am Main, Germany
5 The Dinosaur Institute, Natural History Museum of Los Angeles County, Los Angeles, California, United States of America
6 Department of Earth and Environmental Sciences, Vanderbilt University, Nashville, Tennessee, United States of America
7 Department of Zoology, Smithsonian Museum of Natural History, Washington, District of Columbia, United States of America

Corresponding Author: Maria Clara Iruzun Martins
Email address: maria.martins.15@ucl.ac.uk

In morphological traits, variation within species is generally considered to be lower than variation among species, although this assumption is rarely tested. This is particularly important in fields like palaeontology, where it is common to use a single individual as representative of a species due to the rarity of fossils. Here, we investigated intraspecific variation in the cochleae of harbour porpoises (*Phocoena phocoena*). Intraspecific variation of cochlear morphology is well characterised among odontocetes (toothed whales) because of the importance of the structure in echolocation, but generally these studies use only a single cochlea to represent each species. In this study we compare variation within the cochleae of 18 specimens of *P. phocoena* with variations in cochlear morphology across 51 other odontocete species. Using both 3D landmark and linear measurement data, we performed Generalised Procrustes and principal component analyses to quantify shape variation. We then quantified intraspecific variation in our sample of *P. phocoena* by estimating disparity and the coefficient of variation for our 3D and linear data respectively. Finally, to determine whether intraspecific variation may confound the results of studies of interspecific variation, we used multivariate and univariate analyses of variance to test whether variation within the specimens of *P. phocoena* was significantly lower than that across odontocetes. We found low levels of intraspecific variation in the cochleae of *P. phocoena*, and that cochlear shape within *P. phocoena* was significantly less variable than across odontocetes. Although future studies should attempt to use multiple cochleae for every species, our results suggest that using just one cochlea for each species should not strongly influence the conclusions of comparative studies if our results are consistent.
across Cetacea.
Intraspecific variation in the cochleae of harbour porpoises (Phocoena phocoena) and its implications for comparative studies across odontocetes.

Maria Clara Iruzun Martins¹, ²*, Travis Park², ³, Rachel Racicot⁴, ⁵, ⁶, ⁷ and Natalie Cooper²

¹ Division of Biosciences, University College London, University of London, Gower Street, Bloomsbury, London, WC1E 6BT, UK.
² Department of Life Sciences, Natural History Museum London, Cromwell Road, London, SW7 5BD, UK.
³ Department of Earth Sciences, University of Oxford, 3 South Parks Road, Oxford, OX1 3AN, UK.
⁴ Forschungsinstitut und Naturkundemuseum, Senckenberg der SNG, Senckenberanganlage 25, 60325 Frankfurt am Main, Germany
⁵ The Dinosaur Institute, Natural History Museum of Los Angeles County, Los Angeles, CA 90007, USA.
⁶ Department of Earth and Environmental Sciences, Vanderbilt University, Nashville, TN 37240, USA.
⁷ Department of Zoology, Smithsonian Museum of Natural History, Washington, DC 20560, USA.

*Corresponding author: maria.martins.15@ucl.ac.uk
In morphological traits, variation within species is generally considered to be lower than variation among species, although this assumption is rarely tested. This is particularly important in fields like palaeontology, where it is common to use a single individual as representative of a species due to the rarity of fossils. Here, we investigated intraspecific variation in the cochleae of harbour porpoises (*Phocoena phocoena*). Interspecific variation of cochlear morphology is well characterised among odontocetes (toothed whales) because of the importance of the structure in echolocation, but generally these studies use only a single cochlea to represent each species. In this study we compare variation within the cochleae of 18 specimens of *P. phocoena* with variations in cochlear morphology across 51 other odontocete species. Using both 3D landmark and linear measurement data, we performed Generalised Procrustes and principal component analyses to quantify shape variation. We then quantified intraspecific variation in our sample of *P. phocoena* by estimating disparity and the coefficient of variation for our 3D and linear data respectively. Finally, to determine whether intraspecific variation may confound the results of studies of interspecific variation, we used multivariate and univariate analyses of variance to test whether variation within the specimens of *P. phocoena* was significantly lower than that across odontocetes. We found low levels of intraspecific variation in the cochleae of *P. phocoena*, and that cochlear shape within *P. phocoena* was significantly less variable than across odontocetes. Although future studies should attempt to use multiple cochleae for every species, our results suggest that using just one cochlea for each species should not strongly influence the conclusions of comparative studies if our results are consistent across Cetacea.

**Introduction**

Intraspecific variation, the diversity of genotypes and phenotypes within a single species, is a key component of adaptation and evolution by natural selection (O’Dell & Rajakaruna, 2011;
Des Roches et al., 2018). Intraspecific variation includes variation related to size, allometry, sex, and differences related to environment or genetics. It can influence community structure and ecosystem function as much as interspecific variation (Des Roches et al., 2018). Within species, there can be variation in predator defences, parasite resistance, resource manipulation and many more, all of which can alter the number and strength of interactions, shifting the dynamics of an ecosystem (Bolnick et al., 2011). Intraspecific variation also allows local adaptation and the formation of ecotypes leading to phenotypic or genotypic divergence (Ishikawa, Onoda & Hikosaka, 2007; O’Dell & Rajakaruna, 2011).

For most morphological traits, it is assumed that degrees of intraspecific variation are generally smaller than those of interspecific variation. Nevertheless, in many studies this assumption is not (or cannot be) tested because of logistical constraints, for example, where using multiple specimens for each species is not feasible due to time or cost restrictions, or because samples are limited. Palaeontological studies, for example, commonly use a single individual as representative of a whole population or species because of the rarity of fossils (Ekdale & Racicot, 2015; Ekdale, 2016; Park et al., 2017a; Gonzales, Malinzak & Kay 2018; Marx et al., 2018; Racicot et al., 2019).

One field where this assumption has been made is the study of cetacean cochleae. The cochlea, the organ of hearing, is a complex structure with a vital role in species survival (Cantos et al., 2000). Odontocetes (toothed cetaceans) are reliant on their hearing abilities using echolocation for feeding, communicating and navigating their habitat, among other functions (Ketten, 1992a; Ketten, 1992b). Most cross-species studies of cetacean cochlear morphology use only one cochlea for each species (e.g. Luo & Marsh, 1996; Ekdale & Racicot, 2015; Park, Fitzgerald & Evans, 2016; Churchill et al., 2016) due to the rarity of specimens in natural history collections and because μCT scanning them takes time and money. Many studies also focus on
using the cochlea to study interspecific variation among mammal groups (Ekdale & Rowe, 2011; Ekdale 2013, Benoit et al., 2015; Billet, Hautier & Lebrun, 2015; Ekdale, 2016; Costeur, et al., 2018; Racicot, et al. 2016; Racicot, Darroch & Kohno, 2018) rather than just within cetaceans.

However, increasing numbers of studies are using cochlear morphology (e.g. number of turns, shape of endolymphatic sac) for phylogenetic purposes using geometric morphometrics (e.g. Spoor et al., 2007; Billet, Hautier & Lebrun, 2015; Mennecart & Costeur, 2016; Costeur et al., 2018), which could have implications in comparative studies (Ekdale, 2010; Ekdale, 2015; Thean, Kardjilov & Asher, 2017).

Here, we tested whether intraspecific variation is a major confounding factor in cross-species analyses by comparing morphological variation among the cochleae of 18 specimens of harbour porpoise (Phocoena phocoena) with those across 51 other species of odontocete. Specifically we investigate whether variation within P. phocoena cochleae is significantly lower than variation across all odontocete cochleae. Intraspecific variation has not been quantified relative to interspecific variation in odontocete cochleae. Previous studies have either briefly noted clustering of taxa in morphospace, differences between left and right ears (Racicot, Darroch & Kohno, 2018), or only focused on population-level differences within a single species (Schnitzler et al., 2017). Our 18 conspecific specimens represent a large advance in our ability to quantify intraspecific variation in this feature.

Material and Methods

DATA COLLECTION

We chose the harbour porpoise (Phocoena phocoena) for this study because of the large number of specimens in natural history collections making it possible to obtain a large sample size for the species, something that is not available for most odontocetes. We assumed that
results from *P. phocoena* can be generalised across odontocetes, but were unable to test this assumption with the available data.

3D shape data

TP collected the majority of the 3D shape data for an earlier paper (Park et al., 2019). 18 cochleae of *P. phocoena* were added to this study by MCIM (Table S1). For all cochleae we obtained μCT scans of the periotics - the bone containing the inner ear - of 52 species (comprising 94% of extant genera) of odontocetes by imaging osteological specimens from museum collections. Scan parameters for cochleae of *P. phocoena* are available in Table S2 and scan parameters for previously collected data on the other species are available from Park *et al.* (2019). For this study we obtained periotics from 18 specimens of *P. phocoena*. Of these 15 are from the left-hand side of the skull, and three from the right-hand side; 17 are from the UK coast, and one is from the USA coast; four are female, five are male, and nine are unsexed (see Table S3). All specimens are adult animals.

Using these data, we reconstructed 3D models of the inner ears using the segmentation and thresholding editors in Avizo 9.0 (Visualization Sciences Group—a FEI Company, 2016), and then cleaned the resulting 3D models using Geomagic Wrap® (3D Systems, 2017). Next we landmarked the digital models with 40 sliding semilandmark curves comprising a total of 361 landmarks (Figure 1), using IDAV Landmark (Wiley, 2005). The position of these curves followed the protocols of Costeur *et al.* (2018), using only the curves from the cochlea because the semi-circular canals are not phylogenetically or ecologically informative in odontocetes (Costeur *et al.*, 2018). Landmarks were placed by a single investigator (TP) to avoid multi-user bias in placement. Finally we exported coordinates from the landmarked models as .pts files from IDAV Landmark (Wiley, 2005).
Geometric morphometrics

We performed all geometric morphometric analyses in R version 3.4.3 (R Core Team, 2019), using the R package GEOMORPH (Adams, Collyer & Kaliontzopoulou, 2018). First, we used Generalised Procrustes Analysis (GPA) to remove the effects of position, scale and orientation. The semilandmarks were ‘slid’ along their tangent vectors between adjacent semilandmarks until their positions minimised the shape difference between specimens (using the Procrustes distance criterion), to reduce the effect of their initially arbitrary placement (Bookstein et al., 1999; Gunz, Mitteroecker & Bookstein 2005; Adams, Rohlf & Slice, 2013). We then performed a principal component analysis (PCA) on the resulting Procrustes coordinates using the ‘plotTangentSpace’ function; and used these principal components (PCs) in further analyses.

Linear measurements

One of us (MCIM) took the following measurements of the cochlea (Figure 2) in Avizo 9.2.0 using the Measure and Spline Probe tools and following protocols from Park, Fitzgerald & Evans (2016): (1) cochlear height; (2) cochlear width; (3) number of turns of cochlear canal; (4) cochlear volume; (5) cochlear canal length; (6) length of secondary spiral lamina (SSL); (7) basal ratio (cochlear height/cochlear width); (8) axial pitch (cochlear height/number of turns); and (9) percentage of extent of secondary spiral lamina ((secondary spiral lamina length/cochlear canal length)*100). We also measured (10) width of basal turn perpendicular to cochlear width (W2); (11) maximum distance between turns of the cochlea (ITD); and (12) area of fenestra cochlearis (FC) following Churchill et al. (2016). Where possible (n = 7) for the specimens of P. phocoena, we also collected condylobasal length (skull length in mm) as a proxy for body size as body mass data was not available for all specimens.
We chose these measurements because they relate to auditory function in cetaceans. Cochlear height (1) and width (2, 10) relate to the auditory abilities of odontocetes (West, 1985; Ketten & Wartzok, 1990). The number of turns of the cochlear canal (3) also relates to auditory abilities and differs among mammal groups (West, 1985; Ketten & Wartzok, 1990; Ekdale & Rowe, 2011). Cochlear volume (4) is used as an estimate of size which relates to body mass and length (Racicot et al., 2016). Cochlear canal length (5) correlates with body size in odontocetes and is used as a quantitative measurement for comparison (Ketten, 1992a). Length of secondary spiral lamina (6) was measured to calculate percentage of extent of secondary spiral lamina (9), an indicator of the stiffness of the basilar membrane that is associated with high frequency hearing (Ekdale & Racicot, 2015; Park et al., 2017a). Basal ratio (7) and axial pitch (8) are two quantitative ways to examine cochlear shape and size for comparative studies (Racicot et al., 2016). Both measurements are negatively correlated with frequency of hearing (Ketten & Wartzok, 1990). Taxa that use high frequency hearing also tend to have a larger inter-turn distance (11; Ekdale & Racicot, 2015). The area of the fenestra cochleae (12) tends to be larger in low frequency species, although beaked whales and sperm whales also have large areas.

We then performed a principal component analysis (PCA) on the scaled and centred linear measurements listed above using the ‘prcomp’ function in R; and used these principal components (PCs) in further analyses.

All data used in this study are available on the NHM Data Portal (data.nhm.ac.uk). The 3D shape data for cochleae of *P. phocoena*, linear measurements of all cochleae, and other data used in this paper are deposited in Martins, Park & Cooper (2019). 3D shape data for cochleae of other odontocetes were collected for Park *et al.* (2019) and are deposited in Park *et al.* (2018).
All analyses were carried out in R version 3.4.3 (R Core Team, 2019), and reproducible R scripts are available on GitHub (github.com/nhcooper123/intraspecific-porpoise; Cooper, Martins & Park, 2020).

Quantifying intraspecific variation in Phocoena phocoena

To quantify intraspecific variation in cochleae of *P. phocoena* we estimated disparity for all principal components of the 3D shape data and linear measurements separately, using the R package dispRity (Guillerme, 2018). We used the sum of variances and median centroid distance metrics as our disparity metrics. For each linear measurement we also calculated the coefficient of variation (CV), i.e. (standard deviation/mean)*100. Generally values of CV below 30% represent low levels of variation (Brown, 1998), and previous studies that found low levels of intraspecific variation in cochleae find values of CV below 20% (Ekdale & Rowe, 2011; Mennecart & Costeur, 2016; Racicot, Darroch & Kohno, 2018). We also calculated values of CV for our measurements using data from previous studies of cochleae intraspecific variation in odontocetes so we could compare the values to those of *P. phocoena*. Where these studies contained right and left cochleae from one individual we first took the mean for the individual, then calculated CV across individuals.

Comparing intraspecific variation and interspecific variation

Quantifying the amount of intraspecific variation alone does not tell us whether intraspecific variation may confound the results of studies of interspecific variation. To investigate this we tested whether there were significantly higher levels of variation in cochlear shape among *P. phocoena* and the other odontocetes than within the specimens of *P. phocoena*. For the 3D
shape data, we first conducted a Procrustes MANOVA (multivariate analysis of variance) using
the ‘proc.lm’ function from GEOMORPH using the Procrustes aligned coordinates as the
response variable, and the grouping variable (P. phocoena vs. other odontocetes) as the
explanatory variable. Next we fitted a standard MANOVA using the ‘manova’ function in R, with
the first 26 PC axes scores (comprising 95% of the variance in cochlear shape) as the response
variable. To identify which PCs, if any, were most important in driving any differences among
the cochleae of P. phocoena and other odontocetes we also used analysis of variance (ANOVA)
to test each PC separately. To account for multiple testing we Bonferroni corrected our p values.
Finally, we repeated the latter two analyses, i.e. MANOVA and ANOVAs, using the first six PC
axes (comprising 95% of the variance) extracted from the linear measurements.

We visualised specimen position in 2D morphospace across combinations of PCs 1, 2, and 3
(3D shape data) and PCs 1-4 (linear measurement data) as these PCs were significant in our
ANOVAs (see Results). To aid the interpretation of our results we also created morphospace
plots of PC1 and PC2 where species were coloured based on taxonomic family and a selection
of ecological variables; specifically habitat (riverine = > 50% of time spent in river, nearshore = >
50% of time spent in coastal waters, oceanic = > 50% of time spent in pelagic waters), diet
(generalist = consumes a wide variety of prey types, fish = > 50% of diet is fish, cephalopods =
> 50% of diet is cephalopods), feeding mode (raptorial = prey capture occurs mainly via
snapping or ram feeding, suction = prey capture occurs mainly via suction, i.e. obligate suction
feeders), and dive type (shallow = maximum estimated dive depth less than 100m; middle depth
= maximum estimated dive depth ~500m; deep = maximum estimated dive depth ~ 1000m; very
deep = maximum estimated dive depth greater than 1000m) and hearing type (NBHF = known
or presumed to use narrow band high-frequency sonar; Type 2 = do not use NBHF). Some taxa
fall into several of these categories so we assigned these to composite categories e.g. Tursiops
is classed as nearshore/oceanic. Details of the categories for each species, and the references these were taken from, are in Tables S4 and S5. We also tested whether these variables were correlated with overall cochlear shape using MANOVAs.

Sensitivity analyses

Because we have 18 specimens of *P. phocoena* and only one specimen of each of the other species, cochleae of *P. phocoena* will have a stronger effect on the principal components scores than other cochleae, distorting the morphospace. Ideally we would collate an equal number of cochleae for each species, but these specimens simply do not exist in these numbers for the majority of species. Specimens of *P. phocoena* are the most readily available because they are common and often strand (Coombs et al., 2019). Their skulls are also small enough to store in bulk within natural history collections. To determine whether this distortion problem influenced our conclusions, we sampled two specimens of *P. phocoena* from the 18 and then repeated the GPA, PCA, and MANOVA analyses described above for the 3D shape dataset. We repeated this for every possible combination of two specimens of *P. phocoena*, and then for every combination of three specimens and so on until reaching every combination of 17 specimens.

To determine whether our results apply only to broad-scale taxonomic comparisons, i.e. *P. phocoena* vs. other odontocetes, we also repeated our GPA, PCA and MANOVA analyses comparing *P. phocoena* with other species of *Phocoena* only (*P. dioptrica*, *P. sinus*, *P. spinipinnis*). 3D shape analyses used PC1 - PC14, and linear measurement analyses used PC1 - PC7 as these were the PCs required to account for 95% of the variation in cochlear shape in each case.
Finally, variation within cochleae of *P. phocoena* may be explained by the differences among the specimens (Table S3) in terms of whether the cochleae came from the left or right side of the animal, their sex, origin or body size. We therefore tested whether variation in cochleae of *P. phocoena* was significantly correlated with the side of the head the cochlea came from, or the sex, origin (UK vs. USA) or skull length (as a proxy for body size) of the specimens. We repeated our GPA and PCA analyses for the specimens of *P. phocoena* only, and then used the resulting Procrustes coordinates or PCs as response variables in Procrustes ANOVAs or MANOVAs with either side, sex, origin or log skull length as predictors. 3D shape analyses used PC1 - PC16, and linear measurement analyses used PC1 - PC7 as these were the PCs required to account for 95% of the variation in cochlear shape in each case. For 3D shape analyses where sex or skull length were the predictors we used only PC1 - PC6 (accounting for 75% of the variation on cochlear shape) because there were not enough observations to fit MANOVAs for all 16 PCs.

**Results**

**3D shape variation**

The first 26 principal components (PC) axes comprised 95% of the variance in cochlear shape. PC1 accounted for 34.08% of the cochlear shape variation (Figure 3), and represents cochleae that are positively correlated with having a less oval-shaped and more circular fenestra vestibuli. PC2 accounted for 12.15% of the shape variation and represents cochleae that are negatively correlated with having a radial expansion of the scala tympani (i.e. a tympanal recess, *sensu* Park et al., 2017b; Park, Fitzgerald & Evans, 2017c; Figure 3), and PC3 accounted for 8.956% of the shape variation (Figure 3), and represents cochleae that are positively correlated with having cochlear canals that do not overlap the basal turn.
Linear measurements

The first six PC axes comprised 95% of the variance in cochlear linear measurements. PC1 accounted for 54.28% of the variation (Figure 4), which is best described by a combination of cochlea width, $W_2$, ITD and cochlea height. PC2 accounted for 18.65% of the variation best described by basal ratio and extent of secondary spiral lamina (Figure 4). PC3 (8.579% of the variation) and PC4 (6.355% of the variation) are best described by secondary spiral lamina extent and number of turns respectively (Figure 4).

Quantifying intraspecific variation in Phocoena phocoena

The disparity of $P.\ phocoena$ in the 3D shape data (sum of variances = 0.014, median centroid distance = 0.100) is lower than in the linear measurement data (sum of variances = 1.487, median centroid distance = 1.091), although the morphospace occupied is larger compared with other odontocetes in the 3D shape data (Figures 3 and 4). Values of CV vary across the linear measurements (Table 1) but all are below 30% and only area of fenestra cochlearis (FC) exceeds 20%. This reflects low levels of intraspecific variation in cochleae of $P.\ phocoena$.

Values of CV for all measurements except cochlear volume, axial pitch and ITD are higher than those found in previous studies (Table 1; Table S6).

Comparing intraspecific variation and interspecific variation

There was significantly more variation among odontocete species than within the specimens of $P.\ phocoena$ (Table 2; Figures 3 and 4) when the entire shape of the cochlea was considered, whether we used Procrustes coordinates or PCs from the 3D shape data or linear measurements.
When we investigated individual PC axes (Table S7), for the 3D shape data we found that only PC1 and PC3 of the 26 PCs significantly differed between *P. phocoena* and other odontocetes, and for linear measurements only PC1, PC2 and PC4 (only PC1 and PC2 are significant after Bonferroni correction) showed significant differences among the two groups.

Figures S1 and S2 show how species are spread across PC1/PC2 morphospace according to taxonomic family, habitat, diet, feeding mode, dive type and hearing type. All of these variables except feeding mode were also significantly correlated with cochlear shape (Table S8).

**Sensitivity analyses**

In our sensitivity analyses, MANOVAs for 3D shape data remained significant provided at least four specimens of *P. phocoena* were included in the analyses (Figures S3 and S4).

When the specimens of *P. phocoena* were compared to other *Phocoena* species only, there was still significantly less variation within the specimens of *P. phocoena* using Procrustes coordinates from the 3D shape data (but not for PCs) and linear measurements (Table S9).

Variation in cochleae of *P. phocoena* was not significantly correlated with the side of the head the cochlea came from, the sex, or skull length of the specimens for either 3D shape data or linear measurements (Table S10). Origin (UK vs. USA) was not significantly correlated with variation in cochleae of *P. phocoena* for the linear measurements, but was significantly correlated with variation in the 3D shape data (*p = 0.038*; after Bonferroni correction *p = 0.152*). This is driven by the large negative value of PC1 for the USA specimen. To ensure this did not influence our intraspecific variation results (see above) we repeated our 3D shape analyses (including the GPA and PCA steps) removing this specimen (NHMUK_1873.6.3.45). The results are qualitatively identical to those including the USA specimen (Table S11).
In this study, we tested whether intraspecific variation is a major confounding factor in cross-species analyses by comparing variation within the cochleae of the harbour porpoise (*Phocoena phocoena*) with those across a broad sample of other odontocete species. Levels of intraspecific variation in cochleae of *P. phocoena* were lower than the levels of variation seen across other odontocetes. *P. phocoena* showed higher values of coefficient of variation (CV) overall (ranging from 3.394 - 26.80%; Table 1) compared to other odontocetes (ranging from 0 - 23.33%; Table S6), however, for most cochlear measurements the values of CV were similar. The greatest difference in values of CV was for the fenestra cochlearis, which may be artificially variable due to a lack of standardised protocol for measuring it (Racicot et al., 2016; Churchill et al., 2016). Intraspecific variation in *P. phocoena* might be expected to be higher than in other odontocetes because of their large geographic range which stretches across almost all shallow cool temperate to subpolar waters of the northern hemisphere, within which they encounter many different kinds of habitats (Jefferson, 2014). This suggests that *P. phocoena* should generally have a higher level of intraspecific variation compared to other odontocetes with more limited geographic ranges and habitats. We cannot test this here, due to the paucity of studies containing more than one specimen for each species, but future research should compare the values of CV found for *P. phocoena* here with those of another similarly widespread species, for example *Tursiops truncatus* (common bottlenose dolphin) or *Delphinus delphis* (short-beaked common dolphin).

Although there was significantly more variation among odontocete species than among the specimens of *P. phocoena*, for at least some principal components, specimens from other species overlapped with *P. phocoena*. Cochlear morphology was also related to habitat, diet,
mode of echolocation, and dive type (Table S8), suggesting that species identity alone is not
even enough to explain variation across all principal components. In addition, not all of the specimens
of *P. phocoena* cluster together tightly (Figure 3). This could reflect differences in cochlear
morphology among populations (although this does not appear to be the case for these data as
only one specimen is from the USA and the rest are from the UK), or variability within the
species. Cochleae also exhibit convergent evolution (Park et al., 2019), so individuals or
species that live in similar acoustic environments (regions where variables such as water
temperature, pressure, and salinity result in sound travelling through the water at similar
velocities) as *P. phocoena* may have similar cochleae. The relatively low levels of intraspecific
variation found in *P. phocoena* adds further support for the acoustic environment being a
constraint on cochlear morphology. This is additionally demonstrated in the morphospace plot of
PC1 and PC2 (Figure 3), the area occupied by *P. phocoena* is shared with taxa such as *P.
sinus* (vaquita), *Cephalorhynchus commersonii* (Commerson’s dolphin), and *Inia geoffrensis*
(Amazon river dolphin), all of which also live in shallow waters. However, other taxa that spend
more time in deeper waters, for example *Pseudorca crassidens* (false killer whale) and *Tursiops
truncatus*, also occupy this region of morphospace, indicating that additional factors are also in
play, although they may also frequent shallower waters or have coastal ecotypes (Jefferson,
Webber & Pitman, 2015). Previous studies have also suggested that constraints on cochlear
morphology are strong, due to several factors including frequency propagation (Manoussaki et
al., 2008), spatial constraints (Pietsch et al., 2017), and its vital importance for survival in
odontocetes (Ketten, 1992a). As such the capacity for elaboration or innovation in cochlear
shape may be limited, which could force all odontocete cochleae to fall within the same limited
morphospace. Other sources of error, such as mislabelling of specimens in collections or errors
in µCT scanning or landmarking, are unlikely due to the care taken during these stages, and
assistance from expert curators in museum collections.
Although, as described above, the specimens of *P. phocoena* did not cluster together closely in our PC plots, we note that PC plots only show two dimensions of the data, not all of the 26 PC axes that comprised 95% of the variance in cochlear shape. Our MANOVA results, however, showed that there was significantly more variation in cochlear shape among odontocete species than within *P. phocoena* specimens. This highlights that clustering (or not) in several PC axes, does not necessarily equate to clustering across the entire morphospace, although limitations in how we visualise more than three dimensions mean that PC plots are still valuable visualisation tools. In addition, although PCAs on different datasets are not directly comparable, shape variation in the cochleae of odontocetes appears to be spread across a much higher number of PCs than that seen in most studies involving skulls (e.g. Cardini & Polly, 2013; McCurry et al., 2017; Page & Cooper 2017; Randau, Sanfelice & Goswami, 2019).

**Conclusion**

The 18 specimens of *P. phocoena* used in this study represent the highest number of records for one species used in intraspecific studies of cochlear morphology (the next highest is 12 individual sperm whales; Schnitzler et al., 2017) in cetaceans. Overall, our results show that intraspecific variation of cochlea shape in *P. phocoena* is lower than interspecific variation of cochlea shape across odontocetes. Assuming that interspecific variation of *P. phocoena* is representative of patterns across odontocetes, this suggests we can still be reasonably confident in our conclusions when using *n* = 1 for comparative studies of cochlea morphology, as done in most previous studies (Ekdale & Racicot, 2015; Ekdale, 2016; Park, Fitzgerald & Evans, 2016; Park et al., 2017a; Park et al., 2017b). It is possible that *P. phocoena* is not representative of patterns across odontocetes. If *P. phocoena* is less variable than other odontocetes this would reduce our confidence that using just one specimen for comparative studies is sufficient. However, as discussed above, we believe this is unlikely because *P.*
*Phocoena* has a broad geographic range, and shows variability in habitat use and trophic ecology across that range (Bjørge 2003; Jefferson 2014). As such we speculate that *P. phocoena* is instead likely to be more variable than most species making our conclusions more conservative. Ideally, future studies should use multiple cochleae for each species, but where this is impossible due to the rarity of specimens, or expense of specimen loans and/or µCT scanning, this should not influence the conclusions of comparative studies, assuming results from *P. phocoena* can be generalised across odontocetes.
References

Adams, D.C., Rohlf, F.J. and Slice, D.E., 2013. A field comes of age: geometric morphometrics in the 21st century. Hystrix, 24(1), 7.

Adams, D.C., Collyer, M.L. and Kaliontzopoulou, A., 2018. Geomorph: Software for geometric morphometric analyses. R package version 3.0.6.

Benoit, J., Lehmann, T., Vatter, M., Lebrun, R., Merigeaud, S., Costeur, L. and Tabuce, R., 2015. Comparative anatomy and three-dimensional geometric-morphometric study of the bony labyrinth of Bibymalagasia (Mammalia, Afrotheria). Journal of Vertebrate Paleontology, 35(3), e930043.

Billet, G., Hautier, L. and Lebrun, R., 2015. Morphological diversity of the bony labyrinth (inner ear) in extant xenarthrans and its relation to phylogeny. Journal of Mammalogy, 96(4), 658-672.

Bolnick, D.I., Amarasekare, P., Araújo, M.S., Bürger, R., Levine, J.M., Novak, M., Rudolf, V.H., Schreiber, S.J., Urban, M.C. and Vasseur, D.A., 2011. Why intraspecific trait variation matters in community ecology. Trends in Ecology & Evolution, 26(4), 183-192.

Bookstein, F., Schäfer, K., Prossinger, H., Seidler, H., Fieder, M., Stringer, C., Weber, G.W., Arsuaga, J.L., Slice, D.E., Rohlf, F.J. and Recheis, W., 1999. Comparing frontal cranial profiles in archaic and modern Homo by morphometric analysis. The Anatomical Record: An Official Publication of the American Association of Anatomists, 257(6), 217-222.
Bjørge, A. 2003. The harbour porpoise (*Phocoena phocoena*) in the North Atlantic: Variability in habitat use, trophic ecology and contaminant exposure. *NAMMCO Scientific Publications*, 5, 223-228.

Brown, C.E., 2012. *Applied multivariate statistics in geohydrology and related sciences*. Springer Science & Business Media.

Cantos, R., Cole, L.K., Acampora, D., Simeone, A. and Wu, D.K., 2000. Patterning of the mammalian cochlea. *Proceedings of the National Academy of Sciences*, 97(22), 11707-11713.

Cardini, A. and Polly, P.D., 2013. Larger mammals have longer faces because of size-related constraints on skull form. *Nature Communications*, 4, 2458.

Churchill, M., Martinez-Caceres, M., de Muizon, C., Mnieckowski, J. and Geisler, J.H., 2016. The origin of high-frequency hearing in whales. *Current Biology*, 26(16), 2144-2149.

Coombs, E. J., Deaville, R., Sabin, R. C., Allan, L., O’Connell, M., Berrow, S., Smith, B., Brownlow, A., Doeschate, M.T., Penrose, R., Williams, R., Perkins, M.W., Jepson, P.D. and Cooper, N., 2019. What can cetacean stranding records tell us? A study of UK and Irish cetacean diversity over the past 100 years. *Marine Mammal Science*, 35, 1527-1555.

Cooper, N., Martins, M.C.I., and Park, T., 2020. GitHub: nhcooper123/intraspecific-porpoise: code for the paper. Zenodo. DOI: 10.5281/zenodo.3693950.
Costeur, L., Grohé, C., Aguirre-Fernández, G., Ekdale, E., Schulz, G., Müller, B. and Mennecart, B., 2018. The bony labyrinth of toothed whales reflects both phylogeny and habitat preferences. *Scientific Reports*, 8(1), 7841.

Des Roches, S., Post, D.M., Turley, N.E., Bailey, J.K., Hendry, A.P., Kinnison, M.T., Schweitzer, J.A. and Palkovacs, E.P., 2018. The ecological importance of intraspecific variation. *Nature Ecology & Evolution*, 2(1), 57.

Ekdale, E.G., 2010. Ontogenetic variation in the bony labyrinth *Monodelphis domestica* (Mammalia: Marsupialia) following ossification of the inner ear cavities. *The Anatomical Record: Advances in Integrative Anatomy and Evolutionary Biology*, 293(11), 1896-1912.

Ekdale, E.G. and Rowe, T., 2011. Morphology and variation within the bony labyrinth of zhelestids (Mammalia, Eutheria) and other therian mammals. *Journal of Vertebrate Paleontology*, 31(3), 658-675.

Ekdale, E.G., 2013. Comparative anatomy of the bony labyrinth (inner ear) of placental mammals. *PLoS One*, 8(6), e66624.

Ekdale, E.G. and Racicot, R.A., 2015. Anatomical evidence for low frequency sensitivity in an archaeocete whale: comparison of the inner ear of *Zygorhiza kochii* with that of crown Mysticeti. *Journal of Anatomy*, 226(1), 22-39.

Ekdale, E.G., 2016. Morphological variation among the inner ears of extinct and extant baleen whales (Cetacea: Mysticeti). *Journal of Morphology*, 277(12), 1599-1615.
Galatius, A., Olsen, M.T., Steeman, M.E., Racicot, R.A., Bradshaw, C.D., Kyhn, L.A. and Miller, L.A., 2019. Raising your voice: evolution of narrow-band high-frequency signals in toothed whales (Odontoceti). *Biological Journal of the Linnaean Society*, 126(2), 213-224.

Gonzales, L.A., Malinzak, M.D. and Kay, R.F., 2019. Intraspecific variation in semicircular canal morphology - A missing element in adaptive scenarios? *American Journal of Physical Anthropology*, 168(1), 10-24.

Guillerme, T., 2018. dispRity: a modular R package for measuring disparity. *Methods in Ecology and Evolution*, 9(7), 1755-1763.

Gunz, P., Mitteroecker, P. and Bookstein, F.L., 2005. Semilandmarks in three dimensions. In *Modern Morphometrics in Physical Anthropology* (pp. 73-98). Springer: Boston, MA.

Hocking DP, Marx FG, Park T, Fitzgerald EMG, Evans AR. 2017. A behavioural framework for the evolution of feeding in predatory aquatic mammals. *Proceedings of the Royal Society B: Biological Sciences*. 284, 20162750.

Hocking DP, Marx FG, Park T, Fitzgerald EMG, Evans AR. 2017. Reply to comment by Kienle et al. 2017. *Proceedings of the Royal Society B: Biological Sciences*. 284, 20171836.

Ishikawa, K., Onoda, Y. and Hikosaka, K., 2007. Intraspecific variation in temperature dependence of gas exchange characteristics among *Plantago asiatica* ecotypes from different temperature regimes. *New Phytologist*, 176(2), 356-364.
Jefferson, T.A., 2014. Family Phocoenidae (Porpoises). In Wilson D.E., Mittermeier R.A. eds. *Handbook of the Mammals of the World. Vol 4. Sea Mammals* (pp. 528-547). Lynx Edicions, Barcelona.

Jefferson T.A., Webber M.A., and Pitman R.L. 2015. *Marine mammals of the world: A Comprehensive guide to their identification (second edition)* (pp. 326-329). Academic Press, London.

Ketten, D.R. and Wartzok, D., 1990. Three-dimensional reconstructions of the dolphin ear. In *Sensory Abilities of Cetaceans* (pp. 81-105). Springer, Boston, MA.

Ketten, D.R., 1992a. The cetacean ear: form, frequency, and evolution. In *Marine Mammal Sensory Systems* (pp. 53-75). Springer, Boston, MA.

Ketten, D.R., 1992b. The marine mammal ear: specializations for aquatic audition and echolocation. In *The Evolutionary Biology of Hearing* (pp. 717-750). Springer, New York, NY.

Luo, Z. and Marsh, K., 1996. Petrosal (periotic) and inner ear of a Pliocene kogiine whale (Kogiinae, Odontoceti): implications on relationships and hearing evolution of toothed whales. *Journal of Vertebrate Paleontology, 16*(2), 328-348.

Marx, F.G., Park, T., Fitzgerald, E.M. and Evans, A.R., 2018. A Miocene pygmy right whale fossil from Australia. *PeerJ, 6*, e5025.
McCurry, M.R., Evans, A.R., Fitzgerald, E.M., Adams, J.W., Clausen, P.D. and McHenry, C.R., 2017. The remarkable convergence of skull shape in crocodilians and toothed whales. *Proceedings of the Royal Society B: Biological Sciences*, 284(1850), 20162348.

Manoussaki, D., Chadwick, R.S., Ketten D.R., Arruda, J., Dimitriadis, E.K. and O’Malley, J.T., 2008. The influence of cochlear shape on low-frequency hearing. *Proceedings of the National Academy of Sciences*, 105 (16), 6162-6166.

Martins, M.C.I., Park, T. and Cooper, N., 2019. Dataset: Intraspecific variation in harbour porpoise cochleae. Natural History Museum Data Portal (data.nhm.ac.uk). https://doi.org/10.5519/0091362.

Mennecart, B. and Costeur, L., 2016. Shape variation and ontogeny of the ruminant bony labyrinth, an example in Tragulidae. *Journal of Anatomy*, 229(3), 422-435.

Mourlam, M.J. and Orliac, M.J., 2017. Infrasonic and ultrasonic hearing evolved after the emergence of modern whales. *Current Biology*, 27(12), 1776-1781.

O’Dell, R.E. and Rajakaruna, N., 2011. Intraspecific variation, adaptation, and evolution. In *Serpentine: Evolution and Ecology in a Model System* (pp. 97-137). University of California Press, Berkeley.

Page, C.E. and Cooper, N., 2017. Morphological convergence in ‘river dolphin’ skulls. *PeerJ*, 5, e4090.
Park, T., Fitzgerald, E.M. and Evans, A.R., 2016. Ultrasonic hearing and echolocation in the earliest toothed whales. *Biology Letters*, 12(4), 20160060.

Park, T., Marx, F.G., Fitzgerald, E.M. and Evans, A.R., 2017a. The cochlea of the enigmatic pygmy right whale *Caperea marginata* informs mysticete phylogeny. *Journal of Morphology*, 278(6), 801-809.

Park, T., Evans, A. R., Gallagher, S. J. and Fitzgerald, E. M., 2017b. Low-frequency hearing preceded the evolution of giant body size and filter feeding in baleen whales. *Proceedings of the Royal Society B: Biological Sciences*, 284, 20162528.

Park, T., Fitzgerald, E.M. and Evans, A.R., 2017c. The tympanal recess of the cetacean cochlea: Function and evolution. *Acoustics Australia*, 45(2), 273-278.

Park, T., Mennecart, B., Costeur, L., Grohé, C. and Cooper, N., 2018. Dataset: Convergent evolution in toothed whale cochleae. Natural History Museum Data Portal (data.nhm.ac.uk). [https://doi.org/10.5519/0082968.](https://doi.org/10.5519/0082968)

Park, T., Mennecart, B., Costeur, L., Grohé, C. and Cooper, N., 2019. Convergent evolution in toothed whale cochleae. *BMC Evolutionary Biology*, 9, 195.

Pietsch, M., Dávila, L.A., Erfurt, P., Avci, E., Lenarz, T. and Kral, A., 2017. Spiral form of the human cochlea results from spatial constraints. *Scientific Reports*, 7(1), 7500.

R Core Team, 2019. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. [http://www.R-project.org/.](http://www.R-project.org/)
Racicot, R.A., Gearty, W., Kohno, N. and Flynn, J.J., 2016. Comparative anatomy of the bony labyrinth of extant and extinct porpoises (Cetacea: Phocoenidae). *Biological Journal of the Linnean Society*, 119(4), 831-846.

Racicot, R.A., Darroch, S.A. and Kohno, N., 2018. Neuroanatomy and inner ear labyrinths of the narwhal, *Monodon monoceros*, and beluga, *Delphinapterus leucas* (Cetacea: Monodontidae). *Journal of Anatomy*, 233(4), 421-439.

Racicot, R.A., Boessenecker, R.W., Darroch, S.A. and Geisler, J.H., 2019. Evidence for convergent evolution of ultrasonic hearing in toothed whales (Cetacea: Odontoceti). *Biology Letters*, 15(5), 20190083.

Randau, M., Sanfelice, D. and Goswami, A., 2019. Shifts in cranial integration associated with ecological specialization in pinnipeds (Mammalia, Carnivora). *Royal Society Open Science*, 6(3), 190201.

Schnitzler, J.G., Frédérich, B., Früchtnicht, S., Schaffeld, T., Baltzer, J., Ruser, A. and Siebert, U., 2017. Size and shape variations of the bony components of sperm whale cochleae. *Scientific Reports*, 7, 46734.

Thean, T., Kardjilov, N. and Asher, R.J., 2017. Inner ear development in cetaceans. *Journal of Anatomy*, 230(2), 249-261.

Visualization Sciences Group-a FEI Company, 2016. Avizo: 3D analysis software for scientific and industrial data, standard edition.
West, C.D., 1985. The relationship of the spiral turns of the cochlea and the length of the basilar membrane to the range of audible frequencies in ground dwelling mammals. *The Journal of the Acoustical Society of America*, 77(3), 1091-1101.

Wiley, 2005. Landmark Editor 3.0. Institute for Data Analysis and Visualization.

3D Systems, 2017. Geomagic Wrap.

**Acknowledgements**

We thank: Oliver Lambert (IRNSB), Gabriel Aguirre Fernandez (UZH), Brett Clark and Vincent Fernandez (NHM), Alistair Evans (MU) and Loïc Costeur (UB) for scanning several of the specimens in this study; Richard Sabin and Roberto Portela Miguez (NHM), Erich Fitzgerald and Karen Roberts (NMV), Neil Duncan and Eileen Westwig (AMNH) for access to specimens.

**Data and code availability**

All data are available on the NHM Data Portal (data.nhm.ac.uk). 3D data for cochleae of *P. phocoena*, linear measurements of all cochleae, and other data used in this paper are in Martins, Park & Cooper (2019; doi.org/10.5519/0091362). 3D data for cochleae of the other odontocete species are available from Park *et al.* (2018; doi.org/10.5519/0082968).

Reproducible R scripts are available on GitHub (github.com/nhcooper123/intraspecific-porpoise; Cooper, Martins & Park, 2020).
Table 1: Coefficients of variation (CV) of 12 linear measurements for 18 Phocoena phocoena specimens, along with the mean CV of these measurements from previous odontocete studies (see Table S3 for raw data), the number of studies featuring each measur

SSL = length of secondary spiral lamina; W2 = Width of basal turn perpendicular to cochlear width; ITD = Inter-turn distance, maximum distance between turns; FC = area of fenestra cochlearis (FC). No previous studies measured intraspecific variation in SSL or extent of SSL.
Table 1: Coefficients of variation (CV) of 12 linear measurements for 18 Phocoena phocoena specimens, along with the mean CV of these measurements from previous odontocete studies (see Table S3 for raw data), the number of studies featuring each measurement (N), and the maximum number of individuals in each of these previous studies. SSL = length of secondary spiral lamina; W2 = Width of basal turn perpendicular to cochlear width; ITD = Inter-turn distance, maximum distance between turns; FC = area of fenestra cochlearis (FC). No previous studies measured intraspecific variation in SSL or extent of SSL.

| measurement                              | This study CV (%) | Previous studies Mean CV (%) | N | maximum number of individuals |
|------------------------------------------|-------------------|-----------------------------|---|-------------------------------|
| cochlea height                           | 7.783             | 6.793                       | 8 | 12                           |
| cochlea width                            | 4.842             | 3.364                       | 4 | 12                           |
| number of turns of the cochlear canal    | 3.394             | 2.478                       | 7 | 5                            |
| cochlear volume                          | 8.015             | 10.23                       | 1 | 2                            |
| cochlear canal length                    | 6.189             | 3.401                       | 4 | 12                           |
| SSL                                      | 6.619             | -                           | - | -                            |
| basal ratio                              | 8.846             | 0.000                       | 1 | 2                            |
| axial pitch                              | 8.688             | 8.736                       | 5 | 4                            |
| extent of SSL | 3.089 | - | - | - |
| W2           | 7.612 | 2.544 | 2 | 5 |
| ITD          | 11.93 | 12.622 | 2 | 5 |
| FC           | **26.80** | 9.838 | 2 | 5 |
Table 2 (on next page)

Table 2. Results of Procrustes MANOVA and standard MANOVA analyses using either Procrustes aligned coordinates or principal components (PCs) accounting for 95% of the variance as the response variable, and whether a specimen was Phocoena phocoena

3D shape data analyses used PCs 1 to 26; linear measurements analyses used PCs 1 to 6.
Table 2. Results of Procrustes MANOVA and standard MANOVA analyses using either Procrustes aligned coordinates or principal components (PCs) accounting for 95% of the variance as the response variable, and whether a specimen was *Phocoena phocoena* or another odontocete species as the explanatory variable. 3D shape data analyses used PCs 1 to 26; linear measurements analyses used PCs 1 to 6.

| Test                  | df | F/approx F | Pillai | p     |
|-----------------------|----|------------|--------|-------|
| Procrustes ANOVA      | 1,67 | 6.005     | NA     | 0.001 |
| MANOVA                | 1,67 | 8.623     | 0.842  | < 0.001 |

Linear measurements

| MANOVA                | 1,67 | 9.090     | 0.468  | < 0.001 |
Figure 1

Endocast of NHMUK_SW1934.31 in anterior, lateral, dorsal and ventral views showing semi-landmark curves.

A-anterior view, B-lateral view, C-dorsal view, D-ventral view. Dotted lines show the semi-landmark curves.
Figure 2

*Line drawing of Phocoena phocoena (NHMUK_SW1934.31) cochlea in vestibular and posterior view, illustrating key measurements.*

A-vestibular view, B-posterior view
Figure 3

Principal components (PC) plots for 3D cochlea shape data for PCs 1-3.

Purple points and convex hulls are Phocoena phocoena specimens; green points are all other odontocete species. Numbers in brackets show the percentage variance explained for each PC.
Manuscript to be reviewed

A

PC2 (12.15%)

0.1

-0.1

-0.4 -0.2 0.0 0.2

PC1 (34.08%)

B

PC3 (8.956%)

0.1

-0.1

-0.4 -0.2 0.0 0.2

PC1 (34.08%)

C

PC3 (8.956%)

0.1

-0.1

-0.2 -0.1 0.0 0.1 0.2

PC2 (12.15%)
Figure 4

Cochleae of taxa near the extremes of the morphospace shown in figure 3 (not to scale)

A: Mesoplodon mirus (PC1 max); B: Globicephala melas (PC1 min); C: Sousa teuszii; D: Tasmacetus shepherdi.
A  
Mesoplodon  
\textit{mirus}

B  
Globicephala  
\textit{melas}

C  
Sousa  
\textit{teuszii}

D  
Tasmacetus  
\textit{shepherdii}