Mendelian randomization analysis identified genes pleiotropically associated with central corneal thickness

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Abstract

Objective: To prioritize genes that were pleiotropically or potentially causally associated with central corneal thickness (CCT).

Methods: We applied the summary data-based Mendelian randomization (SMR) method integrating summarized data of genome-wide association study (GWAS) on CCT and expression quantitative trait loci (eQTL) data to identify genes that were pleiotropically associated with CCT. We performed separate SMR analysis using CAGE eQTL data and GTEx eQTL data. SMR analyses were done for participants of European and East Asian ancestries, separately.

Results: We identified multiple genes showing pleiotropic association with CCT in the participants of European ancestry. CLIC3 (ILMN_1796423; \( P_{SMR} = 4.15 \times 10^{-12} \)), PTGDS (ILMN_1664464; \( P_{SMR} = 6.88 \times 10^{-9} \)) and C9orf142 (ILMN_1761138; \( P_{SMR} = 8.09 \times 10^{-9} \)) were the top three genes using the CAGE eQTL data, and RP11-458F8.4 (ENSG00000273142; \( P_{SMR} = 5.89 \times 10^{-9} \)), LCNL1 (ENSG00000214402.6; \( P_{SMR} = 5.67 \times 10^{-9} \)), and PTGDS (ENSG00000107317.7; \( P_{SMR} = 1.92 \times 10^{-7} \)) were the top three genes using the GTEx eQTL data. No genes showed significantly pleiotropic association with CCT in the participants of East Asian ancestry after correction for multiple testing.

Conclusion: We identified several genes pleiotropically associated with CCT, some of which represented novel genes influencing CCT. Our findings provided important leads to a better understanding of the genetic factors influencing CCT, and revealed potential therapeutic targets for the treatment of primary open-angle glaucoma and keratoconus.

Keywords: Corneal thickness, Expression quantitative trait loci, Summary Mendelian randomization, Genome-wide association study

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Introduction
The cornea is a highly collagenous and transparent tissue through which light reaches the interior structures of the eye. Previous studies highlighted the importance of central corneal thickness (CCT) in relation to several ocular and non-ocular conditions. For example, decrease in CCT is significantly associated with intraocular pressure (IOP) [1]. Thinner CCT has been demonstrated as an important feature of keratoconus and a risk factor for primary open-angle glaucoma (POAG) in patients with ocular hypertension [2–7]. Keratoconus is the leading cause of corneal transplants worldwide, with an estimated prevalence of 0.14% [8], while POAG is the most common cause of irreversible blindness worldwide, accounting for approximately 70% of all the cases of glaucoma [9].

Epidemiologic studies have shown that CCT differs among ethnic groups, with Europeans having higher CCT values than Africans, and Asians showing a larger variation in CCT [10]. CCT is highlyheritable, with an estimated heritability ranging from 68 to 95% [11–13]. Previous genome-wide association studies (GWAS) identified a number of CCT-associated loci in Europeans and Asians, such as genetic loci in ZNF469, FOXO1, LRRK1 and IBTK [14–20]. Recent genetic studies revealed additional novel loci associated with CCT, some of which conferred relatively high risks of keratoconus and POAG, highlighting the potential involvement of CCT-associated genes underlying the pathogenesis of keratoconus and POAG [21, 22].

Mendelian randomization (MR) uses genetic variants as the proxy to randomization and is a promising tool to explore pleiotropic/potentially causal effect of an exposure (e.g., gene expression) on the outcome (e.g., CCT) [23]. MR could reduce confounding and reverse causation that are commonly encountered in traditional association studies, and has been successful in identifying gene expression probes or DNA methylation loci that are pleiotropically/potentially causally associated with various phenotypes, such as neuropathologies of Alzheimer’s disease and severity of COVID-19 [24, 25].

In this paper, we applied the summary data-based MR (SMR) method integrating summarized GWAS data for CCT and cis-eQTL (expression quantitative trait loci) data to prioritize genes that are pleiotropically/potentially causally associated with CCT.

Methods
Data sources
eQTL data
In the SMR analysis, cis-eQTL genetic variants were used as the instrumental variables (IVs) for gene expression. We performed SMR analysis using gene expression in blood due to unavailability of eQTL data for the eye. Specifically, we used the CAGE eQTL summarized data for blood [26], which included 2765 participants, and the V7 release of the GTEx eQTL summarized data for blood [27], which included 338 participants. The eQTL data can be downloaded at https://cnsgenomics.com/data/SMR/eQTLsummarydata.

GWAS data for corneal thickness
The GWAS summarized data for CCT were provided by a recent genome-wide association meta-analysis of CCT [21]. The results were based on meta-analyses of 1000 genomes phase 1 imputed GWASs on CCT, with a total of 19 cohorts from the International Glaucoma Genetics consortium (IGGC). Specifically, the meta-analysis for participants of European ancestry included 14 cohorts with a sample of size of 17,803, and the meta-analysis for participants of East Asian ancestry included 5 cohorts with a sample size of 8107. All participating studies assumed an additive genetic model, adjusting for age, sex and at least the first five principal components. The GWAS summarized data can be downloaded at https://datashare.is.ed.ac.uk/handle/10283/2976.

SMR analysis
We conducted the SMR analysis with cis-eQTL as the IV, gene expression as the exposure, and CCT as the outcome. The analysis was done using the method as implemented in the software SMR. Detailed information regarding the SMR method was reported in a previous publication [28]. In brief, SMR applies the principles of MR to jointly analyze GWAS and eQTL summary statistics in order to test for pleiotropic association between gene expression and a trait due to a shared and potentially causal variant at a locus. In SMR, the top cis-eQTL was used to estimate the effect of gene expression on the outcome. SMR analysis relies on the validity of the genetic variants as IVs to obtain consistent estimates. A genetic variant has to satisfy three assumptions to be a valid IV: 1) it is associated with the risk factor (gene expression); 2) it is not associated with any confounder of the risk factor-outcome association; and 3) it is conditionally independent of the outcome given the risk factor and confounders. It was found that if the P-value in a linear regression of the eQTL analysis for each variant is less than $1 \times 10^{-5}$, then weak instrument bias was small [29]. In the SMR analysis, we adopted the default threshold of $P_{eQTL} = 5 \times 10^{-8}$ to select the top associated cis-eQTL (with a default window size of 2000 Kb) for the SMR analysis, minimizing the risk of weak IV. By default, we removed SNPs with allele frequency difference > 0.2 between any pairwise data sets, including the LD reference sample data, the eQTL summary data and the GWAS summary data. We also conducted the heterogeneity in dependent instruments (HEIDI) test to
evaluate the existence of linkage in the observed association. Rejection of the null hypothesis (i.e., $P_{\text{HEIDI}} < 0.05$) indicates that the observed association could be due to two distinct genetic variants in high linkage disequilibrium with each other. In conducting the HEIDI test, we adopted the default settings: removing SNPs in very strong linkage disequilibrium [$LD, r^2 > 0.9$] with the top associated eQTL, and SNPs in low LD or not in LD [$r^2 < 0.05$] with the top associated eQTL; $P_{\text{eQTL}} < 1.5 \times 10^{-3}$; number of cis-SNPs $\geq 3$ for a HEIDI test loses power if cis-SNPs $< 3$; and maximum eQTLs in a HEIDI test $= 20$. We used false discovery rate (FDR) to adjust for multiple testing.

Data curation and statistical/bioinformatical analysis was performed using R version 4.0.3 (https://www.r-project.org/), PLINK 1.9 (https://www.cog-genomics.org/plink/1.9/) and SMR (https://cnsgenomics.com/software/smr/).

Results

Basic information of the summarized data
The number of participants of the CAGE eQTL data is much larger than that of the GTEx eQTL data, so is the number of eligible probes. The sample size of the GWAS data for the European ancestry is much large than that for the East Asian ancestry, so is the number of eligible genetic variants. The detailed information was shown in Table 1.

SMR analysis in the European population
In participants with European ancestry, we identified multiple genes showing pleiotropic association with CCT after correction for multiple testing using FDR (Table 2). Specifically, using the CAGE eQTL data, our SMR analysis identified 12 genes that were pleiotropically/potentially causally associated with CCT, with $CLIC3$ (ILMN_1796423; $P_{\text{SMR}} = 4.15 \times 10^{-12}$), $PTGDS$ (ILMN_1664464; $P_{\text{SMR}} = 6.88 \times 10^{-9}$) and $C9orf142$ (ILMN_1761138; $P_{\text{SMR}} = 8.09 \times 10^{-9}$) being the top three genes (Fig. 1).

Using the GTEx eQTL data, our SMR analysis identified six genes that were pleiotropically/potentially causally associated with CCT, with $RP11-458F8.4$ (ENSG00000273142.1; $P_{\text{SMR}} = 5.89 \times 10^{-9}$; Fig. 2), $LCNL1$ (ENSG00000214402.6; $P_{\text{SMR}} = 5.67 \times 10^{-8}$; Fig. 3), and $PTGDS$ (ENSG00000107317.7; $P_{\text{SMR}} = 1.92 \times 10^{-7}$; Fig. 3) being the top three genes. It should be noted that the gene $PTGDS$ was the top gene showing significant pleiotropic association in both SMR analyses.

SMR analysis in the east Asian population
In participants of East Asian ancestry, we identified no genes showing significant pleiotropic association with CCT after correction for multiple testing using FDR (Table 3). Specifically, using the CAGE eQTL data, we found that two genes, $PASK$ (ILMN_1754858, $P_{\text{SMR}} = 7.98 \times 10^{-5}$; ILMN_1667022, $P_{\text{SMR}} = 8.24 \times 10^{-5}$) and $HIATL1$ (ILMN_1703229, $P_{\text{SMR}} = 4.30 \times 10^{-4}$; ILMN_1737964, $P_{\text{SMR}} = 5.84 \times 10^{-4}$), each of which was tagged by two probes, were among the top hits in the SMR analysis. However, none of the genes survived multiple comparison. Using the GTEx eQTL data, we found that several genes overlapped with the top genes in the SMR analysis using CAGE eQTL data, including $MAP7D1$, $HIATL1$, $POLA2$ and $ACADM$. Again, none of the genes survived multiple comparison.

Discussion
In the present study, we integrated summarized data of GWAS on CCT and eQTL data in the MR analysis to explore putative genes that showed pleiotropic/potentially causal association with CCT. In the participants of European ancestry, we identified multiple genes showing significantly pleiotropic association with CCT, some of

| Data Source | Total # of participants | Number of eligible genetic variants or probes |
|-------------|------------------------|---------------------------------------------|
| European eQTL data | | |
| CAGE | 2765 | 16,363 |
| GTEx | 338 | 9765 |
| GWAS data | 17,803 | 815,426 |
| Asian eQTL data | | |
| CAGE | 2765 | 12,034 |
| GTEx | 338 | 6901 |
| GWAS data | 8107 | 435,923 |

Table 1 Basic information of the eQTL and GWAS data

CAGE Consortium for the Architecture of Gene Expression, eQTL expression quantitative trait loci, GTEx Genotype-Tissue Expression, GWAS genome-wide association studies
Table 2 Significant genes showing pleiotropic association with CCT in participants of European ancestry

| eQTL data | Gene ID | Gene | CHR | Top SNP | P(eQTL) | P(GWAS) | Beta | SE | P(SMR) | P(HEIDI) | Q value |
|-----------|---------|------|-----|---------|---------|---------|------|----|--------|----------|---------|
| CAGE      | ILMN_1796423 | CLIC3 | 9   | rs908839 | 8.48 × 10^{-7} | 6.88 × 10^{-14} | 6.171 | 0.890 | 4.15 × 10^{-12} | 2.59 × 10^{-1} | 6.79 × 10^{-8} |
|           | ILMN_1664464 | PTGDS | 9   | rs885070 | 7.83 × 10^{-3} | 3.43 × 10^{-11} | 8.474 | 1.463 | 6.88 × 10^{-9} | 9.97 × 10^{-4} | 4.41 × 10^{-5} |
|           | ILMN_1761138 | C9orf142 | 9   | rs4542025 | 1.08 × 10^{-2} | 4.93 × 10^{-13} | 11.896 | 2.063 | 8.09 × 10^{-9} | 1.02 × 10^{-4} | 4.41 × 10^{-5} |
|           | ILMN_2198408 | MFF | 2   | rs7560053 | ~0 | 1.37 × 10^{-7} | -1.834 | 0.351 | 1.71 × 10^{-7} | 2.36 × 10^{-3} | 6.99 × 10^{-4} |
|           | ILMN_1714108 | TP53INP1 | 8   | rs896853 | 2.29 × 10^{-3} | 4.09 × 10^{-8} | -6.619 | 1.226 | 5.96 × 10^{-7} | 1.42 × 10^{-1} | 1.95 × 10^{-3} |
|           | ILMN_1802894 | VKORC1L1 | 7   | rs2460421 | 4.93 × 10^{-4} | 3.57 × 10^{-7} | -5.016 | 1.046 | 1.64 × 10^{-6} | 6.08 × 10^{-5} | 4.46 × 10^{-3} |
|           | ILMN_1766859 | MSL2 | 3   | rs13433683 | 2.00 × 10^{-4} | 1.17 × 10^{-6} | 4.751 | 1.035 | 4.40 × 10^{-6} | 1.97 × 10^{-1} | 0.010 |
|           | ILMN_1781560 | ST3GAL6 | 3   | rs2439225 | 1.80 × 10^{-5} | 2.04 × 10^{-6} | 4.043 | 0.887 | 5.23 × 10^{-6} | 1.83 × 10^{-2} | 0.011 |
|           | ILMN_1716651 | RUNX2 | 6   | rs1200428 | 8.94 × 10^{-10} | 5.21 × 10^{-6} | 2.142 | 0.476 | 6.92 × 10^{-6} | 1.87 × 10^{-1} | 0.013 |
|           | ILMN_1680171 | MFF | 2   | rs7572491 | 3.84 × 10^{-16} | 1.47 × 10^{-7} | -9.712 | 2.199 | 1.00 × 10^{-5} | 3.13 × 10^{-1} | 0.016 |
|           | ILMN_1785635 | BRD3 | 9   | rs457420 | 5.98 × 10^{-47} | 4.23 × 10^{-6} | -4.571 | 1.043 | 1.18 × 10^{-5} | 8.29 × 10^{-3} | 0.018 |
|           | ILMN_2188204 | ATG12 | 5   | rs6872406 | 4.42 × 10^{-61} | 8.61 × 10^{-6} | 3.638 | 0.847 | 1.74 × 10^{-5} | 8.65 × 10^{-2} | 0.024 |
| GTEx      | ENSG00000273142.1 | RP11-458F8.4 | 7   | rs4718424 | 3.54 × 10^{-25} | 2.02 × 10^{-12} | 6.017 | 1.034 | 5.89 × 10^{-9} | 3.58 × 10^{-5} | 5.76 × 10^{-5} |
|           | ENSG00000214402.6 | LCNL1 | 9   | rs11145951 | 1.84 × 10^{-13} | 9.02 × 10^{-16} | 10.562 | 1.946 | 5.67 × 10^{-8} | 4.65 × 10^{-1} | 2.77 × 10^{-4} |
|           | ENSG00000107317.7 | PTGDS | 9   | rs868102 | 8.79 × 10^{-20} | 2.18 × 10^{-10} | 9.142 | 1.756 | 1.92 × 10^{-7} | 8.55 × 10^{-2} | 6.24 × 10^{-4} |
|           | ENSG00000226824.2 | RP4-756H11.3 | 7   | rs6979488 | 3.99 × 10^{-30} | 4.92 × 10^{-8} | 2.943 | 0.509 | 8.66 × 10^{-7} | 9.24 × 10^{-5} | 0.002 |
|           | ENSG00000170175.6 | CHRN8 | 17  | rs12600861 | 3.47 × 10^{-17} | 9.03 × 10^{-7} | 4.382 | 1.033 | 2.20 × 10^{-5} | 1.89 × 10^{-1} | 0.037 |
|           | ENSG00000164938.9 | TP53INP1 | 8   | rs2340586 | 1.17 × 10^{-10} | 1.95 × 10^{-8} | - | 4.694 | 2.30 × 10^{-5} | 6.53 × 10^{-1} | 0.037 |

*The GWAS summarized data were provided by the study of Iglesias et al. and can be downloaded at https://datashare.is.ed.ac.uk/handle/10283/2976. The CAGE and GTEx eQTL data can be downloaded at https://cnsgenomics.com/data/SMR/eQTLsummarydata. P(eQTL) is the P-value of the top associated cis-eQTL in the eQTL analysis, and P(GWAS) is the P-value for the top associated cis-eQTL in the GWAS analysis. Beta is the estimated effect size in SMR analysis, SE is the corresponding standard error, P(SMR) is the P-value for SMR analysis and P(HEIDI) is the P-value for the HEIDI test. FDR was calculated at P = 10^{-5} threshold.

**Bold font** means statistical significance after multiple testing using FDR.

| CAGE Consortium for the Architecture of Gene Expression, CCT central corneal thickness, CHR chromosome, eQTL expression quantitative trait loci, GTEx Genotype-Tissue Expression, HEIDI heterogeneity in dependent instruments, SNP single-nucleotide polymorphism, SMR summary data-based Mendelian randomization, FDR false discovery rate, GWAS genome-wide association studies.

which represented novel genes associated with CCT. Our findings provided important leads to a better understanding of the genetic factors influencing CCT, and revealed potential therapeutic targets for the treatment of POAG and keratoconus.

We found that PTGDS (prostaglandin D2 synthase) showed significantly pleiotropic association with CCT in the participants of European ancestry using both CAGE and GTEx eQTL data. PTGDS encodes the gluthione-independent prostaglandin D2 synthase which catalyzes the conversion of prostaglandin H2 (PGH2) to prostaglandin D2 (PGD2), an important marker for keratocytes [30, 31]. A genetic polymorphism in PTGDS (rs11145951) was reported to be associated with CCT in the European population (P = 9.20 × 10^{-12}) but not in the Asian population (P = 2.30 × 10^{-2}) [15]. The association of this polymorphism with CCT did not reach genome-wide significance in the Latino population (P = 1.15 × 10^{-5}), suggesting ethnic-specific effect of this genetic polymorphism on CCT [32]. PTGDS was found to be highly expressed in corneal endothelial cells (CECs) [33], ranked the among the top 50 most highly expressed genes in CECs [34]. When using a novel dual media culture approach for the in vitro expansion of primary human corneal endothelial cells (hCECs), the expression of PTGDS increased by 12.64 folds following exposure of cultivated hCECs from proliferative (M4) to maintenance (M5) medium [35]. Given that thinner CCT was an important feature for keratoconus and a potential risk factor for POAG, these findings, together with ours, demonstrated the important role of PTGDS in influencing CCT and highlighted the potential of this
gene as a promising target for the prevention and treatment of keratoconus and POAG.

RP11-458F8.4 showed the most significantly pleiotropic association with CCT using GTEx eQTL data in the participants of European ancestry (Table 2). RP11-458F8.4 is a long intergenic noncoding RNA (lincRNA). LincRNAs exercise various tissue-specific functions such as remodeling chromatin and genome architecture, RNA stabilization and transcription regulation \cite{36}. RP11-458F8.4 was reported to be associated with various types of malignant tumors. For example, it was upregulated in late-stage colon cancers, and its expression may be involved in the progression of colon cancer \cite{37}. It was found to be a prognostic lincRNA in high-grade serous epithelial ovarian cancer \cite{38}, and was differently expressed in patients with breast cancer \cite{39}. No studies have reported the association of this gene with CCT. As a result, further investigation is needed to elucidate the exact functions of this gene and examine its role in influencing CCT.

\textit{VKORC1L1} (vitamin K epoxide reductase complex subunit 1 like 1) was pleiotropically associated with CCT in the participants of European ancestry using CAGE eQTL data. \textit{VKORC1L1} can mediate vitamin K-dependent intracellular antioxidant functions \cite{40}. A genetic variant in \textit{VKORC1L1} (rs11763147) was found to be associated with CCT in a combined meta-analysis of the European and the Asian samples \cite{15}. Another genetic variant (rs10563220) located upstream of \textit{VKORC1L1} was found to be significantly associated with intraocular pressure in a GWAS of 8552 Chinese participants \cite{41}. More studies are needed to elucidate the exact functions of \textit{VKORC1L1} in association with CCT, and explore whether/how it is involved in the pathogenesis of POAG and keratoconus.

\textit{RUNX2} (runt-related transcription factor 2) showed significantly pleiotropic association with CCT in the participants of European ancestry using the CAGE eQTL data (Table 2). \textit{RUNX2} is a member of the RUNX family of transcription factors and encodes a nuclear protein, a
key transcription factor of osteoblast differentiation [42, 43]. A genetic variant in \textit{RUNX2} (rs13191376) was found to be associated with CCT in a cross-ancestry GWAS [21]. Another genetic variant in \textit{RUNX2} (rs1755056) was found to be significantly associated with IOP and weakly associated with POAG in a GWAS combing participants from the UK Biobank and published data from International Glaucoma Genetic Consortium [44]. In a transient transfection experiment using MG-63 cells and primary bovine corneal keratocytes, \textit{RUNX2} transcription factors affected the expression of several small leucine rich proteoglycans (SLRP) including mimecan, biglycan and keratocan [45], which were shown to be important for the development and maintenance of corneal transparency [46–48]. In another study using the rabbit corneal epithelial cell line RCE1(5 T5), RNAseq based transcriptome analysis showed that \textit{RUNX2} exhibited the highest expression in terminally differentiated corneal epithelial cells [49]. In breast cancer, it was found that \textit{RUNX2} functioned through the androgen receptor to regulate prolactin-induced protein (PIP) [50], a new biomarker for keratoconus [51]. These findings suggested that \textit{RUNX2} likely plays an important role in affecting CCT and the susceptibility of POAG and keratoconus.

Of the multiple genes showing significantly pleiotropic association with CCT in the participants of European ancestry, none was significant in the SMR analysis for the participants of the East Asian ancestry. And only \textit{RP11-458F8.4}, which was pleiotropically associated with CCT in the participants of European ancestry using GTEx data, appeared among the top-hit genes in the SMR analysis for the participants of East Asian ancestry using GTEx data (Tables 2 and 3). These findings implied possible ethnicity-specific genetic mechanisms underlying CCT, leading to anatomic changes (e.g., cornea and optic nerves) [52–54] and differences in the susceptibility of CCT-related disorders/diseases. For example, the prevalence of POAG was reported to be high in Afro-Caribbeans and African Americans, intermediate in Latinos and Asians, and low in non-Hispanic whites [55]. Moreover, compared with Asians,
Europeans have a higher rate of keratoconus incidence and an older age-of-onset of keratoconus [56, 57]. The findings of our study revealed genes harboring potential biomarkers for tailored screening and treatment of POAG and keratoconus in subjects of different ethnicities.

Our study has some limitations. The number of probes used in our SMR analysis was limited, especially in the SMR analysis of the participants of East Asian ancestry. Moreover, the sample size of eQTL analysis is limited, especially for the GTEx data, which may lead to reduced power in the eQTL analysis. As a result, we may have missed some important genes. Moreover, the sample size and the number of eligible genetic variants for GWAS in the participants of East Asian ancestry is limited, compared to GWAS in the participants of European ancestry, which may affect the power of our SMR analysis and contribute to the insignificant findings in the participants of East Asian ancestry. The HEIDI test was significant for some of the identified genes (Table 2-3). As results, the possibility of horizontal pleiotropy, i.e., the identified association might be due to two distinct genetic variants in high linkage disequilibrium with each other, could not be ruled out. We only used eQTL data in the blood due to the unavailability of eQTL data from the eye. Our findings need to be validated in the future when eQTL data from the eye are available. We used the same eQTL data in the SMR analysis for both European and Asian ancestry. It is possible that eQTL pattern could vary with ethnicity. Ethnicity-specific eQTL data with larger sample size are needed in future SMR research.

**Conclusions**

We identified several genes pleiotropically associated with CCT, some of which represented novel genes influencing CCT. Our findings provided important leads to a better understanding of the genetic factors influencing
Table 3  The top ten genes identified in the SMR analysis of CCT in participants of East Asian ancestry

| eQTL data | Gene ID | Gene | CHR | Top SNP | P_eQTL | P_GWAS | Beta | SE | P_SMR | P_HEIDI | Q value |
|-----------|---------|------|-----|---------|--------|--------|------|----|-------|----------|--------|
| CAGE      | ILMN_1754858 | PASK | 2   | rs16843242 | 7.23 x 10^-116 | 6.20 x 10^-3 | 2.865 | 0.726 | 7.98 x 10^-3 | 3.73 x 10^-1 | 0.400 |
|           | ILMN_1667022 | PASK | 2   | rs16843242 | 1.61 x 10^-102 | 6.20 x 10^-5 | 3.056 | 0.776 | 8.24 x 10^-5 | 3.90 x 10^-1 | 0.400 |
|           | ILMN_1670870 | ALCAM | 3   | rs9810954  | 2.67 x 10^-54 | 6.92 x 10^-3 | -5.006 | 1.299 | 1.16 x 10^-4 | 4.93 x 10^-1 | 0.400 |
|           | ILMN_1777906 | MAP7D1 | 1   | rs11263861 | 3.51 x 10^-100 | 1.05 x 10^-4 | -3.348 | 0.876 | 1.33 x 10^-4 | 2.01 x 10^-5 | 0.400 |
|           | ILMN_1778104 | ACADM | 1   | rs2133134  | 7.36 x 10^-72 | 1.85 x 10^-4 | 6.341 | 1.733 | 2.52 x 10^-4 | 1.15 x 10^-3 | 0.575 |
|           | ILMN_1765578 | TIPARP | 3   | rs344050  | 1.98 x 10^-9 | 6.46 x 10^-6 | 10.924 | 3.030 | 3.12 x 10^-4 | 3.42 x 10^-2 | 0.575 |
|           | ILMN_1654065 | ATOH8 | 2   | rs10187314 | 6.31 x 10^-93 | 3.22 x 10^-4 | 3.393 | 0.958 | 3.97 x 10^-4 | 1.95 x 10^-1 | 0.575 |
|           | ILMN_1696713 | POLA2 | 11  | rs487989  | 1.06 x 10^-30 | 2.03 x 10^-4 | 6.068 | 1.716 | 4.07 x 10^-4 | 1.68 x 10^-3 | 0.575 |
|           | ILMN_1703229 | HIATL1 | 9   | rs56987829 | 8.47 x 10^-26 | 1.86 x 10^-4 | 4.510 | 1.281 | 4.30 x 10^-4 | 4.26 x 10^-2 | 0.575 |
|           | ILMN_1737964 | HIATL1 | 9   | rs117447155 | 2.16 x 10^-128 | 5.12 x 10^-4 | 1.601 | 0.465 | 5.84 x 10^-4 | 7.80 x 10^-3 | 0.634 |
| GTEx      | ENSG00000273142.1 | RP11-45BF8.4 | 7   | rs4718424  | 3.54 x 10^-25 | 1.89 x 10^-6 | 5.550 | 1.282 | 1.50 x 10^-4 | 2.50 x 10^-2 | 0.103 |
|           | ENSG00000226824.2 | RP4-756H11.3 | 7   | rs6979488  | 3.99 x 10^-30 | 2.67 x 10^-5 | 3.156 | 0.801 | 8.11 x 10^-4 | 3.92 x 10^-4 | 0.280 |
|           | ENSG00000116871.11 | MAP7D1 | 1   | rs11263861 | 3.60 x 10^-30 | 1.05 x 10^-4 | -7.011 | 1.892 | 2.11 x 10^-4 | 4.71 x 10^-3 | 0.484 |
|           | ENSG00000148110.11 | HIATL1 | 9   | rs56074891 | 3.46 x 10^-33 | 1.95 x 10^-4 | 4.615 | 1.297 | 3.73 x 10^-4 | 3.17 x 10^-2 | 0.644 |
|           | ENSG00000241258.2 | CRCP | 7   | rs1643388  | 1.34 x 10^-34 | 2.99 x 10^-4 | -4.654 | 1.342 | 5.24 x 10^-4 | 1.81 x 10^-1 | 0.723 |
|           | ENSG00000238268.2 | RP11-229P13,19 | 9   | rs3814499  | 4.82 x 10^-36 | 4.58 x 10^-4 | 4.691 | 1.390 | 7.38 x 10^-4 | 1.65 x 10^-1 | 0.849 |
|           | ENSG00000014138.4 | POLA2 | 11  | rs1858783  | 3.68 x 10^-14 | 2.52 x 10^-4 | 7.358 | 2.233 | 9.82 x 10^-4 | 6.70 x 10^-4 | 0.941 |
|           | ENSG000000203363.2 | ACO12454.4 | 2   | rs1561285  | 1.15 x 10^-19 | 5.29 x 10^-4 | 3.674 | 1.133 | 1.21 x 10^-3 | 8.10 x 10^-1 | 0.941 |
|           | ENSG00000117054.9 | ACADM | 1   | rs51799967 | 1.32 x 10^-12 | 3.69 x 10^-4 | 14.740 | 4.631 | 1.46 x 10^-3 | 4.90 x 10^-3 | 0.941 |
|           | ENSG00000168528.7 | SERINC1 | 1   | rs7541464  | 4.49 x 10^-25 | 8.34 x 10^-4 | 3.282 | 1.032 | 1.48 x 10^-3 | 7.00 x 10^-2 | 0.941 |

*The GWAS summarized data were provided by the study of Ilegyas et al. and can be downloaded at https://datashare.is.ed.ac.uk/handle/10283/2976. The CAGE and GTEx eQTL data can be downloaded at https://cnsgenomics.com/data/SMR/eQTLsummarydata. P_eQTL is the P-value of the top associated cis-eQTL in the eQTL analysis, and P_GWAS is the P-value for the top associated cis-eQTL in the GWAS analysis. Beta is the estimated effect size in SMR analysis, SE is the corresponding standard error, P_SMR is the P-value for SMR analysis and P_HEIDI is the P-value for the HEIDI test. FDR was calculated at P = 10^-3 threshold. CAGE Consortium for the Architecture of Gene Expression, CCT central corneal thickness, CHR chromosome, eQTL expression quantitative trait loci, GTEx Genotype-Tissue Expression, HEIDI heterogeneity in dependent instruments, SNP single-nucleotide polymorphism, SMR summary data-based Mendelian randomization, FDR false discovery rate, GWAS genome-wide association studies. CCT, and revealed potential therapeutic targets for the treatment of POAG and keratoconus.

Supplementary Information
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Additional file 1.

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Not applicable.

Authors’ contributions
ZY, JY and WY designed and registered the study. DL and JY analyzed data and performed data interpretation. ZY, DL and JY wrote the initial draft, and JY and WY contributed to subsequent versions of the manuscript. All authors reviewed the study findings and read and approved the final version before submission.

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Availability of data and materials
All data generated or analyzed during this study are publicly available as specified in the methods section of this paper. Specifically, the eQTL data can be downloaded at https://cnsgenomics.com/data/SMR/eQTLsummarydata, and the GWAS summarized data can be downloaded at https://datashare.is.ed.ac.uk/handle/10283/2976.
Declarations

Ethics approval and consent to participate
Not applicable.

Consent for publication
Not applicable.

Competing interests
No potential conflicts of interest were disclosed by the authors.

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