Cellular Senescence in the Treatment of Ovarian Cancer

Zehua Wang, MD,*† Haiou Liu, PhD,*† and Congjian Xu, PhD*‡‡

Objective: This review aimed to update the research and development of cellular senescence in the treatment of ovarian cancer. We discussed the current mechanisms of senescence and the major biomarkers of senescence, especially the methods of cellular senescence in the treatment of ovarian cancer.

Materials and Methods: We collected all relevant studies in PubMed from 1995 to 2017. The search terms included senescence and cancer, senescence and ovarian cancer, senescence-associated secretory phenotype, ovarian cancer and chemotherapy, radiotherapy, or biotherapy. PubMed search with the key words senescence and ovarian cancer lists approximately 85 publications. After excluding the duplicated articles, we selected 68 articles most relevant to senescence and ovarian cancer in this review.

Results: Cellular senescence plays a key role in various biological processes of ovarian cancer, which is closely related with the occurrence, development, and treatment of ovarian cancer. Cellular senescence on the one hand can reduce the dose of chemotherapy in ovarian cancer; on the other hand, it also can solve the problem of tumor resistance to apoptosis. Therefore, cellular senescence has been shown to be the third intracellular mechanism of ovarian cancer prevention followed by cellular DNA repair and apoptosis.

Conclusions: In the near future, cellular senescence therapy could be a powerful tool for ovarian cancer treatment.

Key Words: Senescence, Ovarian cancer, Chemotherapy, Radiotherapy

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Ovarian cancer is the most lethal gynecologic malignancy at present. Despite the recent new diagnostic possibilities and medical advances, the 5-year survival rate of patients with advanced ovarian cancer is only 25% to 35%.1 Prognosis is usually very poor because of the late diagnosis. More than 70% of patients diagnosed with this cancer have reached late stage (stage III–IV), when cancer has already spread beyond the ovaries.2 Thus, there is an urgent need to develop effective therapies for ovarian cancer.

Cellular senescence is a process characterized by normal human cells losing their proliferative capacity and departing from the cell cycle into a relatively stable state, also named replicative senescence. In recent years, some studies have found that the phenotype of cellular senescence induced by factors such as DNA damage, oxidative stress, and oncogene stress is basically the same with replicative senescence, but differs in the specific mechanism. This form of cellular...
senescence is unrelated with the shortening of telomeres and the 
generations of proliferation; therefore, it is called premature 
senescence.3 Premature senescence has been shown to be the 
third intracellular mechanism of cancer prevention followed by 
cellular DNA repair and apoptosis, which is closely related with 
the occurrence, development, and treatment of tumors.4 
Therefore, cellular senescence has been suggested as a novel 
therapy for ovarian cancer. In this article, we focus on the 
current research and development of cellular senescence in 
the treatment of ovarian cancer.

Cellular Senescence

The Mechanism of Cellular Senescence

Normal human cells have limited life under the condi-
tions of in vitro culture. This phenomenon was first observed in 
 fibroblasts cultured in vitro human cells. According to 
Haiflick and Moorhead’s5 findings in 1961, the continuous 
culture of human diploid fibroblast cells would enter a state of 
senescence on 50 to 70 generations of in vitro proliferations. 
Under such circumstances, cells had lost the ability of contin-
uous proliferation but were still alive. Even if the most suitable 
culture conditions were provided, the cells could not escape 
the fate of limited proliferation. This proliferative limit of cells is 
sometimes referred to as the Haiflick limit. The phenomenon 
that the normal human cells have limited potential of in vitro 
division is known by Haiflick as cellular senescence, or more 
precisely, replicative senescence. The hypothesis of telomere 
shortening is commonly recognized among the hypothesis of 
the replicative senescence mechanism. Muller6 proposed telo-
mere in 1938 for the first time and found that it was easily 
connected between the ends of injured and ruptured chromo-
somes in Drosophila, thereby forming the various types of 
chromosome aberrations, such as the dissolved end and the 
formation of annularity or dicentric chromosomes. But the natural 
end of chromosomes seems neither connected to the ends of 
ruptured chromosomes nor combined with the natural end of 
chromosomes, just like a hat to maintain the stability of the end 
of the chromosome. Therefore, the fragment located at both 
ends of the chromosome was named telomere by Muller.6 The 
hypothesis of telomere shortening refers to the correlation be-
tween the division ability and the telomere length of the fi-
broblast and more division times of cells with longer telomeres 
than those with shorter telomeres. Studies have shown that the 
presence of telomere shortening is seen in a variety of primary 
human cells during culture. These results suggest that telomere 
shortening plays a direct role in the number of division times 
that may be experienced by primary cells and may serve as a 
marker of cellular senescence. Therefore, telomere shortening 
or telomere structural destruction is considered as the main 
mechanism of senescence.7

With the deepening of this study, the researchers found 
that telomere shortening was not the only factor to induce the 
senescence phenotype (irreversible growth arrest, apoptosis, 
and changes in cell functions). Some of the stimuli that were 
unrelated with telomere, including DNA damage and the ex-
pression of some oncogenes, could also induce normal cell 
growth arrest and exhibit a senescence phenotype. The inducers 
other than telomeres had one thing in common that they had the 
potential to cause or promote tumorigenesis. Cellular senescence 
seems to be a safe mechanism to prevent tumorigenesis through 
inhibition of cell proliferation. Cellular senescence involves the 
signal regulation factor of cell cycle and complex approach.8 
When cellular DNA encounters damage attacks, it can activate 
proteins such as p16, p21, p53, and retinoblastoma (RB) protein 
related to G1/S arrest, and proliferation arrest can occur to 
cells.9,10 Among them, the signaling pathways of p53-p21-
pRB and p16-pRB are the most important. The tumor sup-
pressor gene p53 in induction of cell cycle arrest is realized by 
induction of high expression of p21. And high-expression p21 
can inhibit the binding of cyclin-dependent kinase (CDK) 2 
and type E cell cycle protein, thereby inhibiting the phos-
phorylation of the RB. The RB that is in the form of nonphosphorylation or low phosphorylation can be combined 
with several transcription factors to inhibit their transcription 
activation functions and further inhibit the expression of 
downstream genes needed from S phase into G1 phase, which 
causes growth arrest so as to control the cell cycle progression and 
differentiation. The expression of p16 gene plays an important role 
in the maintenance of cellular senescence. Overexpression of p16 
can inhibit the binding of CDK4/6 and type D cell cycle protein 
to block the phosphorylation of CDK4/6 on RB and prevent 
cells from entering the S phase for induction of cellular se-
nescence (Fig. 1). Whether p16-pRB or p53-p21-pRB pathway 
activation is involved in the induction of senescence seems to 
de on the cell type and origin of the species.11

Detection Biomarkers of Cellular Senescence

At present, there are many ways to detect cellular senescence. In addition to the morphological changes of senescent 
cells observed by using an optical microscope, senescence-
associated β-galactosidase (SA-β-gal) staining is a common 
way to show the senescence cells. In 1995, Dimri et al12 found 
that when the pH value of culture medium was 6, the positive 
rate of β-galactosidase staining for the diploid fibroblasts 
cultured in vitro was gradually increased with the increase of 
the age. This neutral β-galactosidase was defined by them as 
SA-β-gal, namely the SA-β-gal. The β-galactosidase produced 
by senescent cells or tissues can catalyze the substrate X-Gal 
to form a dark blue product, which is easily observed under 
an optical microscope. The SA-β-gal can also be found to be 
increased with the increase of the age in the stratum corneum 
cells of human epidermis. In addition, SA-β-gal can differentiate 
during DNA cells and quiescent cells, which are independent of 
the replication process. The SA-β-gal is a kind of biological 
marker that can be used in vivo and in vitro. Because the method 
of detecting SA-β-gal is simple and easy to use, it has been 
widely applied in the detection of senescent cells. At present, 
more than 2300 articles have applied this method. But as we 
know, there is no single biomarker that can robustly identify 
senescent cells. Recently, multiple studies proved that 
several classes of biomarkers can be identified as potential 
senescence indicators, which are summarized in Table 1.13–25

Cellular Senescence in the Treatment of 
Ovarian Cancer

In the recent researches, it has been confirmed that 
inducing tumor cellular senescence to prevent the unlimited
proliferation of tumor cells can be an important treatment. Normal cells accumulate the alterations from DNA damage, oxidative stress, and oncogene stress that trigger an initial phase of aberrant cell proliferation. This aberrant proliferation can give rise to preneoplastic lesions; to parallel this aberrant situation, ovarian cancer cell intrinsic fail-safe mechanisms such as senescence and apoptosis are activated. During ovarian cancer progression, the accumulation of damage is acquired to override these protective mechanisms, giving rise to ovarian cancer. Experimental researches have confirmed that conventional treatments such as chemotherapy and radiotherapy can induce cellular senescence; at the same time, research about the treatment of target biological treatment on ovarian cancer has also made certain progress. Chemotherapy-, radiotherapy- and biotherapy-induced cellular senescence permanently arrests growth. In cancer cells, chemotherapy-induced senescence is an important alternative cell fate to apoptosis. The study reported an apoptosis-independent function to p53 in cancer and showed that murine lymphomas overexpressing Bcl-2 respond to chemotherapy by engaging a senescence program controlled by p53 and p16INK4a rather than apoptosis. Conversely, upregulating the expression of DNA methyltransferase (DNMT) DNMT3a in colorectal cancer cells subject to DNA damage induced by doxorubicin switches senescence to apoptosis. When senescent cells were challenged with p53-dependent apoptotic stimuli, they underwent necrosis in response to the treatments. Muñozespín et al proposed that cellular senescence, followed by macrophage-dependent clearance of

![FIGURE 1. Overview of replicative senescence and premature senescence. A, Telomere shortening–induced replicative senescence. B, Accumulation stress–induced premature senescence.]

| Biomarker Category                        | Senescence Biomarker | Reference         |
|------------------------------------------|----------------------|-------------------|
| Secretion and surface                    | SA-β-Gal             | [13],2010; [14],2014 |
|                                          | SASP                 | [15],2016         |
| DNA damage and repair                    | γH2AX                | [16],2010         |
|                                          | Lamin B1             | [17],2012         |
|                                          | SAHF                 | [18],2015         |
| Cell cycle related                       | p16INK4a             | [19],2012; [20],2015; [21],2016 |
|                                          | p21WAF1              | [22],2013; [23],2014; [24],2015 |
|                                          | p15INK4B             | [25],2014         |
|                                          | p14ARF               | [21],2016         |

γH2AX indicates gammaH2AX; and SAHF, senescence-associated heterochromatic foci.
senescent cells or by the overgrowth of nearby cells, alters cellularity, resulting in tissue remodeling. Interestingly, the activation of senescence in ovarian cancer cells was associated with a strong senescence-associated secretory phenotype (SASP), which can attract and activate immune system cells to alter the ovarian cancer microenvironment. On immune dysfunction, the SASP's proinflammatory nature can recruit immune cells, which can kill and clear senescent cells or other surviving cancer cells. All in all, chemotherapy-, radiotherapy- and biotherapy-induced cellular senescence can be explored as alternative or complement treatments to ovarian cancer (Fig. 2).

Cellular Senescence and Ovarian Cancer Chemotherapy

Therapies such as surgical adjuvant chemotherapy are the most commonly used methods in treatment of ovarian cancer, which are difficult to significantly prolong the 5-year survival rate (approximately 20%). In particular, the chemotherapy resistance and strong ability of invasion and metastasis of ovarian cancer result in severe reduction of the long-term curative effect and survival rate for ovarian cancer patients. The chemotherapy of ovarian cancer is intended to induce the apoptosis of ovarian cancer cells. However, the tumor cells have a barrier in apoptosis generally, as a consequence of which apoptosis resistance is seen as a difficult problem in the treatment of ovarian cancer. As we know, the tumor cells have the ability of proliferation without restriction, and it was once thought that the tumor cells had lost the ability to enter the senescent state. However, with the deepening of research, Studies have confirmed that chemotherapy drugs can induce cellular senescence in tumor cells. Senescent cells are in permanent growth arrest and, after repairing DNA damage, still undergo cell cycle arrest. Chemotherapy can induce cell cycle arrest and senescence, but cell cycle arrest is not a synonym of senescence. One type of quiescence is simple quiescence, a reversible arrest. Cells are arrested because of lack of growth factors, and addition of growth factors causes proliferation. Another type of quiescence is locked quiescence, an irreversible arrest. Differentiated cells are put on the brakes, and excessive stimulation induces senescence. Studies have found that low doses of chemotherapy drugs can cause tumor cellular senescence and further proved that induction of tumor cellular senescence is a feasible strategy to reverse drug resistance. Senescence-inducing chemotherapy, on the one hand, can reduce the dose of chemotherapy; on the other hand, it also can solve the problem of tumor resistance to apoptosis. Studies have shown that low doses of chemotherapy drugs can induce A2780 ovarian carcinoma cellular senescence, which resulted in prolonged cell cycle arrest predominately in the G1 phase of the cell cycle. In addition, according to Schmitt's findings in the study of chemotherapy in mice, chemotherapy drugs can cause senescence changes in tumor cells, and cellular senescence pathway directly affects the effect of chemotherapy, which confirms that the expression rate of senescent cells is closely related to prognosis. At present, it is believed that the cycle arrest of cell proliferation induced by chemotherapeutic agents is related to DNA damage. If the DNA damage caused by low-dose chemotherapy cannot be repaired, ovarian cancer cells tend to undergo senescence rather than apoptosis. Therefore, senescence plays an important role in the in vivo response to chemotherapy.

FIGURE 2. Overview of cellular senescence in the treatment of ovarian cancer.
Cellular Senescence and Ovarian Cancer Radiotherapy

The property of photon radiation (x-ray, γ-ray) and special ray (α, neutron, proton, and electron) that can lead to ionization effect is made full use of by radiotherapy to destroy the genetic material structure of tumor cells and kill tumor cells. Compared with surgical treatment, radiotherapy can avoid the traumatic injury of organ tissue, ensure the integrity of organ structure, and significantly prolong the survival time for cancer patients.

In ovarian cancer, debulking surgery and adjuvant chemotherapy are still fundamental treatments. Compared with debulking surgery and chemotherapy, the application of radiotherapy is relatively less in the treatment of ovarian cancer, but it still has a certain position. Ovarian cancer is a radiosensitive tumor; although radiotherapy is not the main treatment for ovarian cancer, it can be used as an adjuvant therapy after surgery and palliative therapy of advanced and recurrent lesions. In 2010, International Federation of Gynecology and Obstetrics implemented a phase I clinical trial, which confirmed the clinical feasibility of intensity-modulated whole abdominal radiotherapy for patients with advanced ovarian carcinoma after surgery and adjuvant chemotherapy. Phase 2 clinical trial was conducted in 2011, further suggesting that intensity-modulated whole abdominal radiotherapy can be used as an effective consolidation treatment for the advanced ovarian carcinoma patients after adjuvant chemotherapy. Until now many researches have confirmed that radiotherapy has a therapeutic effect on ovarian cancer. In epithelial ovarian cancer, radiotherapy can be used to relieve the symptoms of pain, bleeding, or oppression caused by advanced ovarian cancer. Yahara et al selected 27 patients with limited recurrence after complete remission and assessed the efficacy and toxicity of definitive radiotherapy for the recurrence of epithelial ovarian cancer. They found that radiotherapy for limited recurrence of epithelial ovarian cancer could achieve a better local control rate without severe toxicity.

As we know, the key to successful radiotherapy is to improve the sensitivity of tumor cells to radiotherapy. Some data demonstrate that large-scale radiation (x-ray) can induce tumor cellular senescence and further alter x-ray sensitivity in tumor cells. Penha et al reported that different dose and time of radiation exposure dictated different impacts on tumor cells. In particular, low-dose ionizing radiation-induced senescence might be a promising therapeutic target to induce growth inhibitory response in both early and late steps of carcinogenesis and might offer an opportunity to reduce the toxicity of radiotherapy. As senescence plays a more and more important role in anti-cancer strategies, senescence-inducing radiotherapy has been used widely in thyroid cancer, lung cancer, breast cancer, head and neck cancer, etc. Although senescence-inducing radiotherapy is quickly developed, this new therapy has not yet been used widely in ovarian cancer in the hope that this new therapy can provide a useful tool in treatment of ovarian cancer.

Cellular Senescence and Ovarian Cancer Biotherapy

The traditional tumor therapies such as radiotherapy and chemotherapy mainly achieve the treatment goals through causing tumor cell death. Owing to the advantage of good short-term therapeutic effect, the tumor mass shrinks rapidly. However, occurrence of the adverse conditions such as metastasis and recurrence results in less satisfactory therapeutic effect of the traditional tumor treatment mode. Biotherapy can significantly prolong patients’ survival time without great adverse reactions, unlike traditional chemotherapy and radiotherapy. Therefore, it is a promising new therapeutic target to restore the senescence pathway and induce senescence of tumor cells. There are some researches that provide evidence for senescence features not only in ovarian cancer cell lines, but also specimens. The SA-β-gal and p16 have been identified as cellular senescence markers; in Konecny et al’s study, they found that the overexpression of p16 in primary clinical ovarian cancer specimens could drive senescence. Mikulapietraski et al showed that the expression of SA-β-Gal was significantly increased in ovarian cancer metastases specimens which had been induced senescence. In recent years, the researchers have made useful attempts in this direction, which are summarized in Table 2.

In the aspect of gene therapy, Bitler et al were the first to show that Wnt5a activation induced senescence of human epithelial ovarian cancer cells by promoting the histone cell cycle regulator/promyelocytic leukemia senescence pathway. It is worth noting that the different Wnt5a expression correlates with different tumor stage and overall survival in epithelial ovarian cancer patients. These data were consistent with the idea that Wnt5a signaling to drive senescence of human epithelial ovarian cancer cells was a potentially novel strategy for developing ovarian cancer therapeutics. Pan et al confirmed that Daxx deletion could accelerate mouse ovarian surface epithelium cells senescence in a p53/p21-dependent manner and induced the high expressions of DNA damage-related proteins (p-H2AX and p-CHK2) and cell cycle–related gene (p21 and p27). These results suggest that DAXX may play a role in epithelial ovarian carcinoma tumorigenesis and may be an anticancer target.

Aird et al found that ribonucleotide reductase M2 (RRM2) expression was significantly higher in epithelial ovarian cancer cell lines and specimens than normal controls, and knockdown of RRM2 expression inhibited the growth of human epithelial ovarian cancer cells through a cellular senescence mechanism. These data suggest that inhibition of RRM2 to induce senescence is a novel therapeutic strategy for epithelial ovarian cancer patients. Özeş et al...
| Category                  | Target Gene/miRNA | Cell Line         | Specimens (n) | Reference |
|---------------------------|-------------------|-------------------|---------------|-----------|
| Gene therapy              | Wnt5a             | OVCAR5 (EOC)      | EOC (130)     | [60],2011 |
|                           | DAXX              | mOSE (EOC)        | EOC (130)     | [61],2013 |
|                           | RRM2              | OVCAR5 (EOC), PEO1 (EOC), SKOV3 (EOC) | EOC (105)     | [62],2014 |
|                           | HOTAIR            | A2780 (EOC), SKOV3 (EOC), HEY2 (EOC), OV90 (EOC), IOSE (EOC), IGROV (EOC), OVMUNA (EOC) | EOC (105)     | [63],2016 |
| RNA interference          | EZH2              | SKOV3/DDP(EOC)    | SOC (92)      | [64],2016 |
| miRNA                     | miRNA-506         | HeyA8 (EOC), SKOV3 (EOC), OVCA432 (EOC), HeLa (CC) | SOC (92)      | [65],2014 |
|                           | miRNA-433         | A2780 (HGSOC), PEO1 (HGSOC), PEO4 (HGSOC) | SOC (92)      | [66],2015 |

**TABLE 2. Senescence-inducing factors as targets of ovarian cancer biotherapy**

| Category                  | Target Gene/miRNA | Cell Line         | Specimens (n) | Reference |
|---------------------------|-------------------|-------------------|---------------|-----------|
| Gene therapy              | Wnt5a             | OVCAR5 (EOC)      | EOC (130)     | [60],2011 |
|                           | DAXX              | mOSE (EOC)        | EOC (130)     | [61],2013 |
|                           | RRM2              | OVCAR5 (EOC), PEO1 (EOC), SKOV3 (EOC) | EOC (105)     | [62],2014 |
|                           | HOTAIR            | A2780 (EOC), SKOV3 (EOC), HEY2 (EOC), OV90 (EOC), IOSE (EOC), IGROV (EOC), OVMUNA (EOC) | EOC (105)     | [63],2016 |
| RNA interference          | EZH2              | SKOV3/DDP(EOC)    | SOC (92)      | [64],2016 |
| miRNA                     | miRNA-506         | HeyA8 (EOC), SKOV3 (EOC), OVCA432 (EOC), HeLa (CC) | SOC (92)      | [65],2014 |
|                           | miRNA-433         | A2780 (HGSOC), PEO1 (HGSOC), PEO4 (HGSOC) | SOC (92)      | [66],2015 |

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demonstrated the key role that HOTAIR in DNA damage contributes to cellular senescence. They reported that the overexpression of HOTAIR could alter the tumor microenvironment and induced the tumor growth slow. They suggested that HOTAIR may represent a target for a novel therapeutic strategy of drug resistance in ovarian cancer.

In the aspect of RNA interference inhibitor, Sun et al found that increased enhancer of zeste homolog 2 (EZH2) expression could induce SKOV3/DDP cell cycle arrest in the G0/G1 phase; at the same time, some expressions of the senescence signaling proteins (p14ARF, p16INK4a, p53, and p21) were significantly increased, and other proteins (CDK1, CDK2, and H3K27me3) were lower expressed. These findings suggest that interfering with EZH2 expression can be a new candidate that could possibly reverse the cisplatin resistance of ovarian cancer.

In the aspect of miRNA: Liu et al demonstrated that overexpression of miR-506 could inhibit the CDK4/CDK6-FOXM1 signaling pathway, consequently decrease proliferation, and promote senescence in ovarian cancer cells and specimens. This newly recognized miR-506-CDK4/6-FOXM1 axis provides further insight into the pathogenesis of ovarian cancer, and miR-506 could be a potential therapy for ovarian cancer. Weinergorzel et al showed that the aberrant expression of miR-433 in ovarian cancer cells could result in the induction of cellular senescence and affect intracellular signaling to mediate chemoresistance. They guessed that the high miR-433 expression might act as a protective mechanism to affect the progression of the cancer and patient survival.

**CONCLUSIONS AND PERSPECTIVES**

In conclusion, cellular senescence can effectively prevent the uncontrolled cell growth and carcinogenesis caused by cytokine DNA damage or oncogene; therefore it is an important tumor suppression mechanism. The research of cellular senescence in ovarian cancer treatment is mainly related to chemotherapy, radiotherapy, and biotherapy, etc. Chemotherapy/radiotherapy–induced cellular senescence is not intended to kill the ovarian cancer cells; the required medication/radiation dose and their toxic reactions are likely to be less than traditional chemotherapy and radiotherapy. What’s more, chemotherapy/radiation–induced cellular senescence is not dependent on telomere length and telomerase activity; the related study will possibly promote the advances in tumor treatment. Biotherapy mainly includes gene therapy, RNA interference, and medication miRNA treatment, etc, which can further clarify the molecular mechanism of ovarian cancer cellular senescence. At present, the research of cellular senescence in ovarian cancer treatment is still in progress; the mechanism of cellular senescence is unclear, and many senescence-related genes have not been studied. As a promising ovarian cancer therapy, induction of ovarian cancer cellular senescence is facing great opportunities and challenges.
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