Pathway-based Analysis Tools for Complex Diseases: A Review

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Received 21 June 2014; revised 30 August 2014; accepted 4 September 2014
Available online 28 October 2014
Handled by Andreas Keller

Abstract Genetic studies are traditionally based on single-gene analysis. The use of these analyses can pose tremendous challenges for elucidating complicated genetic interplays involved in complex human diseases. Modern pathway-based analysis provides a technique, which allows a comprehensive understanding of the molecular mechanisms underlying complex diseases. Extensive studies utilizing the methods and applications for pathway-based analysis have significantly advanced our capacity to explore large-scale omics data, which has rapidly accumulated in biomedical fields. This article is a comprehensive review of the pathway-based analysis methods—the powerful methods with the potential to uncover the biological depths of the complex diseases. The general concepts and procedures for the pathway-based analysis methods are introduced and then, a comprehensive review of the major approaches for this analysis is presented. In addition, a list of available pathway-based analysis software and databases is provided. Finally, future directions and challenges for the methodological development and applications of pathway-based analysis techniques are discussed. This review will provide a useful guide to dissect complex diseases.

Introduction

The etiology for complex human disease is complicated, which involves numerous genes, environmental factors and their interactions [1]. Yet until recently, the genetic basis for most complex diseases has been largely unknown, with just a list of genes identified accounting for very little of the diseases in
the population [2]. Genetic approaches that explore the hereditary variants for complex human diseases have significantly changed from family-based linkage studies, which traditionally mapped Mendelian disorders, to population-based association studies, which were aimed at capturing both common and rare variants for the complex diseases. In the last decade, following the International HapMap Project [3], the development of industrial high-throughput genotyping platforms has led to large-scale genome-wide association studies (GWAS), which are now commonly used to determine the genetic basis for the complex human diseases [4,5].

The methods used to analyze large-scale genetic data are significantly behind the rapid advances in the industrial omics technology. Traditional genetic analysis to explore likely single genes or SNPs associated with the disease only identifies a small proportion of the susceptible genetic variants and contributes to a limited understanding of complex diseases. In addition, current popular single-point analysis of GWAS data suffers from the low replication and validation rate [1,6,7]. There is a growing consensus that genetic risk to complex disease is mostly contributed by multiple genes of small or moderate effect factors through their sophisticated interactions acting in a modular fashion, rather than by the mutations of individual genes [5,7]. Hence, to further interpret the underlying molecular mechanisms that cause complex diseases, systematic dissection of the interactions between the individual disease genes as well as their functionalities is essential [6,8].

Pathway-based analysis is an effective technique that overcomes the limitations of the current single-locus methods. This procedure provides a comprehensive understanding of the molecular mechanisms that cause complex diseases [2]. Principally, a pathway-based approach is similar to the Gene Ontology (GO) analysis [9]. However, the pathway analysis is more specific and detailed, and it tests the association between a pathway, which comprises a set of functionally-related genes, and a disease phenotype. Its capacity of capturing biological interaction among genes and improving power and robustness has been well recognized [10,11]. The early application of pathway-based approaches was extended directly from the Gene Set Enrichment Analysis (GSEA) in microarray data analysis [2,12] and now it has evolved in several directions [13,14]. Moreover, varieties of set-based methods with similar ideas have been developed, such as the gene set analysis (GSA) [15], SNP-ratio test (SRT) [16] and LRPath, a logistic regression-based method for pathway (or gene set) analysis [17]. Methods that focus on the original data instead of statistical results have also been developed and these techniques test the joint distribution of the multi-locus data or extract the principal components from the original data, such as in the linear combination test (LCT) [18] and supervised principal component analysis (SPCA) [19]. Recently, some topological methods to parse the internal information of pathway (e.g., signaling pathway impact analysis (SPIA) [20] and ChiPPER [21]) have also been developed. In short, pathway-based analysis has gradually become an advanced way to the analysis of complex diseases [22].

With the methodological advance, application of pathway-based analysis to unravel complex human diseases has also entered a new era [23,24]. Several studies have demonstrated that pathway-based analysis is superior when it is applied to large-scale genetic datasets for rheumatoid arthritis (RA) [18,24], type 2 diabetes (T2D) [25], schizophrenia [13], Parkinson’s disease [26], etc. In addition, tracing the shared pathways among several pathologies tends to be an ongoing interest of disease pleiotropism, for example, the study of genetic links between RA and systemic lupus erythematosus [27], schizophrenia and T2D [28].

This article is a comprehensive review of the pathway-based analysis methods. The general concepts and principles for the pathway-based analysis are introduced and then, a comprehensive review of the major approaches for this type of analysis is presented. In addition, a list of available pathway-based analysis software and databases is provided. Finally, future directions and challenges for the methodological development and applications of pathway-based analysis techniques are discussed.

**Pathway-based analysis: general concepts and principles**

Currently, there are a variety of pathway-based approaches, which correspond to different research designs and data types. In this article, we focus on SNP/GWAS-derived pathway analysis, but we also include some classical tools for analysis of microarray, as principally they can be easily extended to other data types. Despite some differences in methods for pathway prioritization or null hypotheses to be tested, the basic principle is largely the same, i.e., a pathway-based analysis relies on the use of a testing strategy that targets damaged functionalities, which can produce the outward disease phenotype. It is increasingly recognized that the genetic variations occurring at multiple loci often perturb signal transduction, regulatory and metabolic pathways, resulting in detrimental changes in phenotype [18]. Therefore, pathway-based methods are aimed at analyzing a predetermined aggregation of genes (or SNPs) (alternatively called a gene set) that are contained in a functional unit as defined by prior biological knowledge (e.g., Kyoto Encyclopedia of Genes and Genomes (KEGG), see http://www.genome.jp/kegg/). Depending on whether the individual genotype data or single-point SNP P values (often obtained by single-point association test) are used, varieties of methods, such as over-representation analysis (ORA), gene set analysis for ‘results’ data [29], principal components or regressions for the individual data [19,30] and topology-based analysis [20] (see next section for details), are proposed to combine information from multiple genetic loci within a pathway to assess its overall association with a phenotype.

Compared to single gene analysis methods, pathway-based approaches appear to be well suited for analysis of massive GWAS data, either from biological or statistical considerations [23]. First, since pathway-based approaches focus on sets of genes instead of individual genes, dimension reduction is automatically achieved. Consequently, pathway-based analysis unlikely suffers from the issue of the multiple-test corrections when a large number of SNPs are examined. Second, common diseases often arise from the joint action of multiple SNPs/genes within a pathway. Although each single SNP may confer only a small disease risk, their joint actions are likely to have a significant role in the development of disease. If one only considers the most significant SNPs, the genetic variants that jointly have significant risk effects but make only a small contribution if individually will be missed. Third, locus heterogeneity, in which alleles at different loci cause disease in different populations, will increase the difficulty in replicating
Pathway-based analysis methods

According to the strategies for handling the multivariate genetic data for pathways, we classify pathway-based analysis methods into four groups. Instead of individually reviewing each pathway analysis approach, our goal here is to illustrate the algorithms of the representative methods for each group, as shown in Table 1, and discuss their relative merits.

Over-representation analysis

ORA, often called functional enrichment analysis, is the earliest pathway-based approach to identify an over-represented pathway with a list of susceptible genes obtained by using traditional statistical tests for contingency tables (e.g., Fisher’s exact test, see Table 1) [32]. ORA for SNP data starts by selecting SNPs and mapping the interesting SNPs to the corresponding genes. This initial selection process is based on whether a SNP is mapped to the pathway or whether the SNP is susceptible to the disease [32]. Depending on the results, ORA builds a $2 \times 2$ contingency table to conduct a hypergeometric test [32]. The corresponding $P$ value of a given pathway ($k_i$) is computed by:

$$P(k_i) = 1 - \sum \frac{\binom{M}{n} \binom{N-M}{n-m}}{\binom{N}{n}}.$$

where $N$ is the number of all the studied genes, $n$ is the total number of the risk genes, $M$ is the total number of genes in a pathway $k_i$ and $m$ is the number of risk genes contained in pathway $k_i$.

ORA is the most widely used functional analysis method because it is easily performed. However, it has several limitations. First, ORA considers that each individual gene is of equal importance, which is often not so in biology. Second, the gene (or SNP) list for ORA is usually based on a stringent significant threshold, which can be a salient issue when the number of genes or SNPs analyzed is very large. For GWAS data, statistical power for identifying significant genes or SNPs is limited and therefore the list is often incomplete. Third, construction of the list of statistically significant genes based on the univariate analysis of individual genes does not permit results for genes in a pathway to reinforce each other for detecting an over-represented pathway.

Gene set-based scoring

Gene set-based scoring approaches cover a range of methods that are directly extended from ORA, in which each individual gene is not assumed to be equivalent, instead their importance is ranked by some statistics or $P$ values. Some non-parametric rank sum statistics like Kolmogorov–Smirnov statistic or the Wilcoxon rank sum is used to assess the overall effect of a gene set on a biological phenotype [2,12,33]. The abovementioned ORA is the simplest case in that an equal weight is assumed for all the genes included in a gene set.

The earliest application of a gene-set based scoring approach is the analysis of genome-wide expression profiles [12], in which Subramanian et al. described a powerful analytical method called GSEA for interpreting gene expression data. The method derives its power by focusing on gene sets, that is, groups of genes that share common biological function, chromosomal location or regulation. The rationale is: if a set of functionally-related genes (e.g., a module or a pathway) is correlated with a disease phenotype, there is a trend that the set enriches in a certain area of the ranked gene list according to their differential expression between the sample classes [12]. This idea can be directly borrowed for pathway-based analysis of GWAS data by using a ranked SNP/gene list according to their statistical significance in association with a disease phenotype. Then, similarly, an enrichment score (ES), a running-sum statistic, is calculated for a pathway-based gene set, by walking down the list. This statistic reflects the degree to which a gene

**Table 1 Algorithms and their applications in pathway-based analysis**

| Algorithm                      | Core method                  | Data types                  | Refs. |
|-------------------------------|------------------------------|-----------------------------|-------|
| Over-representation analysis   | Fisher’s exact test          | SNP                         | [32]  |
| Gene set-based scoring         | GSEA                         | Microarray/SNP              | [12]  |
|                               | GSA                          | SNP                         | [15]  |
|                               | SRT                          | SNP                         | [16]  |
|                               | L.Rpath                      | Microarray                  | [17]  |
| Multivariate approaches       | A two-stage approach         | SNP                         | [18]  |
|                               | SPCA                         | SNP                         | [19]  |
|                               | Logistic kernel machine regression | Microarray/SNP                  | [38,39] |
| Topological-based analysis     | SPIA                         | Microarray                  | [20,46] |
|                               | ChiPPER                      | Microarray                  | [21]  |

*Note: GSEA, gene set enrichment analysis; GSA, gene set analysis; SRT, SNP-ratio test; SPCA, supervised principal component analysis; SPIA, signaling pathway impact analysis.*
set is overrepresented at the extremes (top or bottom) of the entire ranked list. ES is the maximum deviation from zero encountered in the random walk; it corresponds to a weighted Kolmogorov–Smirnov-like statistic. Finally, we can assess significance of the analyzed pathway by using some permutation techniques and adjust for multiple testing, as described previously [34].

Later, Wang et al. [2] extended the aforementioned GSEA to analyze the GWAS data and explicitly formulate the ES computation. Supposed that $N$ genes, which are each represented by a SNP, have their statistical values ranked from the largest to smallest, and the list is denoted by $r_1, \ldots, r_N$. A weighted Kolmogorov–Smirnov-like running-sum statistic tests for the over-represented genes within a given gene set $S$ (e.g., a pathway composed of $N_H$ genes $G_j$) and is calculated by:

$$ES(S) = \max_{1 \leq c \leq N} \left( \sum_{j \in S; j \leq c} |r(j)|^p - \sum_{j \in S, j \geq c} \frac{1}{N - N_H} \right),$$

where $N_R = \sum_{j \in S} |r(j)|^p$ and $p$ is a parameter that gives more weight to the genes with extreme statistical values [2].

Evidently, in gene set-based scoring approach, each individual gene is no longer considered of equal importance and it uses more information than ORA to analyze pathways. Gene set-based scoring approaches (GSEA and its derivatives) differ from the previous ORA in two important aspects. First, it considers all of the genes/SNPs in an experiment, not only those above an arbitrary cutoff in terms of expression fold-change or association significance. Second, it assesses the significance by permuting the phenotype class labels, which preserves gene–gene correlations and, thus, provides a more accurate null model. Nevertheless, gene set-based scoring approach relies on the results of single-point analysis for each genetic locus, in essence it is a univariate analysis. It does not explicitly configure the sophisticated interplays between genes contained in a pathway.

**Multivariate approaches**

**A two-stage approach**

A two-stage approach, proposed by several scientists [18], is aimed at tackling several challenges inherent in the aforementioned ORA and gene set-based scoring approaches. The first challenge is how to represent a gene in GWAS. Wang et al. [2] suggested to choose the most significant SNP from each gene as a representative. But, in GWAS, a gene often contains a variable number of SNPs. The genes that contain a number of SNPs jointly having significant risk effects, but individually making only a small contribution, will be missed in such representation. The second challenge is how to deal with correlations among SNPs and genes. Owing to linkage disequilibrium (LD), there may be high correlations among some SNPs. The statistics that were used by Wang and colleagues [2] for testing association of a pathway with the disease do not take correlations among SNPs into account.

To solve these problems, Luo et al. [18] considered three basic units of association analysis—SNP, gene and pathway—and suggest a two-stage (gene and pathway) GWAS. In gene and pathway-based GWAS, each gene is represented by all SNPs of the gene, which are either located within the gene or are not > 500 kb away from the gene. Unlike the aforementioned ORA and gene set-based scoring approaches, in which one examines whether significantly-associated genes are overrepresented in the set of genes to be analyzed, the authors formulated the gene and pathway-based GWAS as the problem to jointly test for association of multiple SNPs within the gene or multiple genes within the pathway with disease. As the proposed two-stage analysis makes full use of the correlation structures between multiple SNPs within a gene or multiple genes within a pathway, it demonstrated better repeatability and reliability in identifying insightful pathways or gene groups related to the development of complex diseases in several large independent genome-wide association studies [18,19,29,35–37].

In order to combine a set of dependent $P$ values of SNPs into an overall significance level for a gene or a set of dependent $P$ values of genes into an overall significance level for a pathway, Luo et al. [18] proposed three novel statistics. These include LCT, quadratic test (QT) and the decorrelation test (DT). LCT takes a linear combination of $P$ values for all SNPs within the gene or a linear combination of statistics for testing association of the genes within the pathway; QT is based on a quadratic form and the test statistic thus follows asymptotically a central $\chi^2$ distribution; finally, in DT, to combine dependent $P$ values, these dependent variables are first transformed into independent variables and then independent variables are combined. For technical details of the three test statistics, readers can consult with the original publication [18].

**Supervised principal component analysis**

Similarly, to deal with the problems of multicollinearity encountered in a pathway-based analysis, Chen et al. [19] proposed to apply the SPCA model to pathway-based SNP association analysis to test the association between a group of SNPs and variation in disease outcome. The idea behind the SPCA model is that within a biological pathway, genetic variations in a subset of SNPs, each contributing a modest amount to disease predisposition, work together to disrupt normal biological processes. Given a gene set defined by a priori knowledge for pathways (e.g., KEGG database), SNPs on an array are first mapped to groups of genes within each pathway. Then a subset of SNPs that is most significantly associated with disease outcome is selected to estimate the latent variable through PCA of this subset. Finally, to identify pathways associated with disease outcome, the authors [19] proposed to test the association between the estimated latent variable and disease outcome using a linear model. In the proposed model, the estimated latent variable is an optimal linear combination of a selected subset of SNPs; therefore, the proposed SPCA model fully utilizes information from both disease-predisposing and disease-protective SNPs in a pathway.

**Logistic kernel machine**

The logistic kernel machine, first proposed by Liu et al. [38] for analysis of genome-wide expression profiles, may be the first unified approach for multi-dimensional parametric and non-parametric modeling of the pathway effect. This model links the disease to covariates parametrically, and to genes within a genetic pathway nonparametrically using kernel machines.
The nonparametric genetic pathway effect allows for possible interactions among the genes within the same pathway and a complicated relationship between the genetic pathway and the outcome. Later, Wu et al. [39] extended this semiparametric model to analyze GWAS data. This type of genome-wide regression-based analysis proceeds via a two-step procedure. First, SNPs are assigned to SNP sets based on certain meaningful biological criteria (e.g., KEGG pathway classifications or other genomic features). Then, tests for the association between each genomic feature and a disease phenotype are performed using a logistic kernel machine-based multimarker statistic as described below.

For a population-based case-control GWAS in which \( n \) independent subjects are genotyped, individual SNPs are first grouped into a SNP set belonging to a pathway (more accurately, genes annotated to these SNPs belong to a pathway). For a pathway-affiliated SNP set containing \( p \) SNPs, \( z_{i1}, z_{i2}, \ldots, z_{ip} \) are the genotyped values of SNPs for the \( i^{th} \) subject \((i = 1, \ldots, n)\), and \( y_i \) is the disease status for the \( i^{th} \) subject \((y_i = 0 \text{ or } 1 \text{ for controls and cases, respectively})\). Let \( x_{i1}, x_{i2}, \ldots, x_{im} \) denote the values of the covariates and \( z_{i0} = 0, 1 \) and \( 2 \), corresponding to homozygotes for the major allele, heterozygotes, and homozygotes for the minor allele, respectively. For the \( i^{th} \) individual, the semiparametric model is given by

\[
\logit\mathbb{P}(y_i = 1) = z_0 + z_1 x_{i1} + \cdots + z_m x_{im} + h(z_{i1}, z_{i2}, \ldots, z_{ip}),
\]

where \( z_0 \) is an intercept term and \( x_{i1}, \ldots, x_{im} \) are regression coefficients of the covariates. The general function \( h(\cdot) \) is arbitrary and is defined by a positive, semi-definite kernel function \( K(\cdot, \cdot) \), and \( y_i \) is influenced by \( z_{i0}, z_{i1}, z_{i2}, \ldots, z_{ip} \) through \( h(\cdot) [39] \).

Choosing an optimal kernel function is the key to analyzing \( h(\cdot) \). A desired model can be specified by changing the choice of \( K(\cdot, \cdot) \). Essentially, \( K(\cdot, \cdot) \) is a function that projects the genotype data from the original space to another space and then \( h(\cdot) \) is modeled linearly in this new space, such that if one considers \( h(\cdot) \) in the original space, it can be highly nonlinear. More intuitively, however, \( K(\cdot, \cdot) \) can be viewed as a function that measures the similarity between two individuals on the basis of the genotypes of the SNPs in the SNP set. The common choices for \( K(\cdot, \cdot) \) are the linear, Gaussian and identical-by-state (IBS) kernels. Finally, the genetic effect of the pathway specified by \( h(\cdot) \) is tested by a variance component score, which follows a scaled \( \chi^2 \) distribution.

**Topological-based analysis**

Recently a new group of methods for pathway-based analysis emerges as topological-based approaches, aiming at explicitly incorporating the dependent structure among genes highlighted by the topology of pathways. Although most methods of this category are developed for analyzing gene expression data, they virtually can be extended to other data types (e.g., GWAS) fairly easily. Unlike the aforementioned approaches that consider only the number of genes and their “expression” in a pathway, topological-based approaches combine the conventionally measured molecular data and also the structural information of the pathway provided by biological databases. A large number of publicly available pathway knowledge bases provide information beyond simple lists of genes for each pathway. These knowledge bases, including KEGG [40], Reactome [41], MetaCyc [42], RegulonDB [43], BioCarta (http://www.biocarta.com) and PantherDB [44], also provide information about gene products that interact with each other in a given pathway, how they interact (e.g., activation or inhibition) and where they interact (e.g., cytoplasm or nucleus).

Topological-based methods are essentially the same as the aforementioned multivariate approaches. The key difference between the two methodological groups is the use of pathway topology to compute gene-level statistics. Massa et al. [45] proposed to use graphical Gaussian models that exploit the graphical evidence of a pathway. This method converts a pathway into a graphical mode and then compares gene sets, which are defined by the pathway. The topological analysis is focused on the strength of the links among genes of a pathway between two phenotypic groups, which is analogous to the aforementioned logical kernel machine modeling. Thus, both the relationship between the genes (i.e., their strength) determined by their topology and experimental data (microarrays and GWAS data) are used to analyze this pathway. Later on, Martini et al. [21] proposed CliPPER, an empirical two-step method, for the identification of significant signal transduction paths within significantly-altered pathways. The initial step uses the aforementioned method to test the entire pathway, which identifies the subgroups of the genes (i.e., signal paths) that makes the entire structure different. The \( P \) value of the test determining whether two graphical Gaussian models are homoscedastic as the weight is collected to compute the relevance of each path. Then, a junction tree is reconstructed to identify related pathways with means or covariance matrices that are significantly different between biological statuses.

A recent impact factor analytic approach called SPIA was proposed by Tarca et al. [20,46], which attempts to capture several aspects of the data: changes in gene expression, the pathway enrichment and the topology of signaling pathways. This method considers the structure and dynamics of an entire pathway by incorporating a number of important biological factors, including changes in gene expression, types of interactions and the positions of genes in a pathway. In brief, SPIA models a signaling pathway as a graph, where nodes represent genes and edges represent interactions between them. Furthermore, it defines a gene-level statistic, called perturbation factor (PF) of a gene, as a sum of its measured change in expression and a linear function of the PFs of all genes in a pathway. The impact factor of a pathway (pathway-level statistic) is defined as a sum of PFs of all genes in a pathway.

In our own perception, topological-based analysis is superior to other methods because it also considers the internal structure of the pathway, reflecting its own property of the pathway, *i.e.*, it gains power from pathway topology. An additional advantage is that it has the merit of sound biological interpretation due to the very nature of this methodology. However, one obvious limitation of this analysis is that these methods are largely empirical, thus hard to prove. In addition, there is a dearth of available software and platforms for implementation, although some Bioconductor packages or web tools (e.g., graphite [47,48]) are released. Finally, current topological-based methods only handle the static properties of the network topology; thus, they have the inability for a dynamic systems model.

**Available resources for pathway-based analysis**

Pathway resources have been rapidly accumulated, which has in turn facilitated the development of pathway-based approaches. The term “pathway-based” refers to the basic
analysis unit that is not a gene or a SNP, but a pathway. A pathway is usually defined as a set of related functional genes. The composition of a pathway can be artificially defined or acquired from several public pathway databases. These public databases, include KEGG (website: http://www.genome.jp/kegg/pathway.html) [49], BioCarta and Pathway Interaction Database (PID, website: http://pid.nci.nih.gov), are aimed at providing different pathway repositories based on their functional categorizations (e.g., metabolic pathways and regulatory pathways). Generally, these databases can be grouped into four categories depending on what the researcher wants to emphasize [10]: the metabolic pathway databases, signal transduction pathway databases, protein–protein interaction pathway databases and transcriptional regulation pathway databases, respectively. More detailed descriptions about these categories and the long list of the corresponding databases are presented in Table 2.

In the last decade, software and web tools to utilize these databases for pathway-based analysis of omics data were rapidly developed with advances in database construction, and consequently have greatly promoted application of these public resources in the biomedical fields. Table 3 lists the common software and web platforms mainly used for analysis of microarray data, and Table 4 lists the available pathway-based analytical tools and web servers designed for pathway analysis on GWAS data. Also, their unique features, functionalities, and databases for annotations are given.

Challenges and future direction

Biological challenges

Pathway-based analysis has significantly enhanced our capacity to explore large-scale omic data, providing an invaluable tool for identifying the damaged functionalities involved in complex diseases. However, we should be cautious on several challenges and limits inherent in this knowledge-guided analysis, which can be divided into two broad categories: (i) knowledge biases and (ii) methodological challenges. Pathway-based analysis relies on acquiring knowledge from gene or pathway databases which provide us varieties of information about genes and how these genes interact with each other. Nevertheless, our knowledge about the existing genes or biological pathways is incomplete [22]. Although a few prominent pathways are well studied, our knowledge of majority of biological pathways (including the limited number of pathways documented in various pathway databases) remains largely fragmented. Because most of pathway-based approaches still rely on mapping multiple SNPs to a single gene, followed by gene-to-pathway mapping, it is often the case in a pathway-based association study that a large number of SNPs or genes fail to be mapped to their corresponding genes or pathways. Apparently, to make full use of biological knowledge to gain power for a pathway-based analysis, we need knowledge bases with high-resolution annotations, in particular for analysis of next-generation sequencing (NGS) data. In addition to the incomplete annotations, many of the existing annotations are of low quality and may be inaccurate. Annotations inferred from indirect evidence (e.g., computationally predicted) are considered to be of lower quality than those derived from direct experimental evidence. Finally, current diagrams for pathways are largely inferred from transcriptional relationships among genes or at protein level, and knowledge for many other regulatory factors (e.g., post transcriptional modifiers, epigenetic factors and environmental triggers) is generally lacking. Hence, for improved power of pathway-based approaches to find damaged functionalities, more integrated databases, which comprehensively review the link of genetic variants, pathways and the environmental triggers, and which jointly analyze their interplays and their contributions to disease, have been called [8,26,50,51]. The Human Genome Epidemiology Network (HuGENet) project is dedicated to this end [52].

Methodological challenges

Methodological challenges for pathway-based approaches in analysis of GWAS or transcriptome have been extensively reviewed in several previous reports [22,53–55]. Hence, we only discuss several key issues here. The first key issue is to improve statistical power for detecting damaged functionalities, which may be achievable by developing more proper weighing schemes for SNPs within a gene or genes within a pathway

| Table 2 | Pathway databases |
|---------|--------------------|
| Category | Description | Examples | URL |
| Metabolic pathways | Primarily contain a series of biochemical reactions, especially the chemical modifications of the small molecule substrates of enzymes | KEGG | http://www.genome.ad.jp/kegg/ |
| | | BioCyc | http://www.biocyc.org/ |
| | | BIOPATH | http://www.mol-net.de/databases/biopath.html |
| | | EMP | http://emp.mcs.anl.gov/ |
| Signal transduction pathways | Describe the information spread from one part or sub-process of the cell to another, generally through a series of covalent modifications of protein | CSNDB | http://geo.nih.go.jp/csndb/ |
| | | SPAD | http://www.grt.kyushu-u.ac.jp/spad/ |
| | | TransPath | http://transpath.gbf.de/ |
| Protein–protein interaction pathways | Focus on interactions between proteins, most of which are derived from various large-scale experimental methods | CYGD | http://mpqs.gsf.de/gene/proj/yeast/index.jsp |
| | | DIP | http://dip.doe-mbi.ucla.edu/ |
| | | GRID | http://biodata.ms.hri.on.ca/grid/perl/Pathways/Pathway/index.jsp |
| | | HPRD | http://www.hprd.org/ |
| Transcriptional regulation pathway | Mainly concern the relationships between transcription factors and the corresponding genes they regulate | STKE | http://www.stke.org/ |
| | | BTITTE | http://www.genome.ad.jp/brite/ |
| | | TRANSFAC | http://transfac.gbf.de/ |
| | | CST | http://www.cellsignal.com/ |
| Software          | Feature                                                                 | Annotations                     | URL                                                      |
|-------------------|-------------------------------------------------------------------------|---------------------------------|---------------------------------------------------------|
| PathMAPA          | A tool for displaying gene expression and performing statistical tests on metabolic pathways at multiple levels for Arabidopsis, based on expression data | Local databases                 | http://bioinformatics.med.yale.edu/pathmapa.htm         |
| MetaCore          | Based on a high-quality, manually-curated database, MetaCore is an integrated software suite for functional analysis of microarray, metabolic, SAGE, proteomics, NGS, copy number variation, siRNA, microRNA and screening data | MetaRodent, MetaLink, MetaSearch | http://www.genego.com/                                  |
| Ingenuity Pathway | A comprehensive software/database search tool for finding functions and pathways for specific biological states | GO, KEGG, BIND                  | https://www.ingenuity.com/                               |
| Analysis (IPA)    | An easy-to-use and powerful software for creating and managing illustrated 3D pathways for publications and presentations | eProtein, ePathway              | http://www.proteinlounge.com/epath3d/                    |
| ePath3D           |                                                                                   | GenBank, Uniprot/Swiss-Prot, TrEMBL, KEGG, ENZYME, etc. | http://www.pathwaybuilder.com/                          |
| Pathway Builder   | An online pathway drawing tool which is the fastest and easiest method of creating signal transduction pathways, enabling the users to design their own project or use pre-made pathway templates to help get them started | KEGG                           | https://pathwayexplorer.genome.tugraz.at                |
| Interactive Pathways | A web-based tool for the visualization, analysis and customization of various pathways maps from KEGG. The recently-released version 2 could deal with metabolic pathway, regulatory pathway and biosynthesis of secondary metabolites | KEGG                           |                                                         |
| Explorer (iPath)  | GSEA-P is a desktop application for Gene Set Enrichment Analysis, with a friendly graphic interface. R-GSEA is provided as a standalone R program. | MSigDB, Gene Set Cards, GEO     | http://www.broadinstitute.org/gsea/                      |
| GSEA-P & R-GSEA   |                                                                                   |                                 |                                                         |
| DAVID             | A tool for augmenting and integrating functional annotations from other databases | KEGG.GO                        | http://david.abcc.ncifcrf.gov/                          |
| MetaCyc           | Applications include serving as an encyclopedia of metabolism, providing a reference data set for the computational prediction of metabolic pathways in sequenced organisms, supporting metabolic engineering and helping to compare biochemical networks | KEGG, BioCyc, EcoCyc            | http://metacyc.org/                                     |
| Reactome          | Intuitive bioinformatics tools for the visualization, interpretation and analysis of pathway knowledge to support basic research, genome analysis, modeling, systems biology and education | KEGG                           | http://www.reactome.org/                               |
| GenMAPP           | Designed to visualize gene expression and other genomic data on maps representing biological pathways and groupings of genes | GenMAPP, GO                     | http://www.genmapp.org                                 |
| FunCluster        | An integrative tool for analyzing gene co-expression networks from microarray expression data; the analytic model implemented in the library involves two abstraction layers: transcriptional and functional (biological roles) | GO, KEGG                      | http://corneliu.henegar.info/FunCluster.htm             |
| Graphite web      | A novel web tool for pathway analyses, consisting of topological-based analysis and network visualization for gene expression data of both microarray and RNA-seq experiments | KEGG, Reactome                 | http://graphiteweb.bio.unipd.it/                        |
| Software     | Feature                                                                 | Annotations                     | URL                                      |
|--------------|--------------------------------------------------------------------------|---------------------------------|------------------------------------------|
| INRICH       | A pathway-based genome-wide association (GWA) analysis tool that tests for enriched association signals of predefined gene sets across independent genomic intervals | KEGG, GO                        | http://atgu.mgh.harvard.edu/inrich/      |
| GeSBAP       | A simple implementation tool of the GSA strategy for the analysis of GWA studies, provides a significant increase in the power testing for this type of studies | GO, BioCarta, KEGG              | http://bioinfo.cipf.es/gesbap/           |
| GSEA-SNP     | A program for the identification of disease-associated SNPs and pathways, the understanding of the underlying biological mechanisms, and the identification of markers with weak effects, undetectable in association studies without pathway consideration | dbSNP, GO, MSigDB, GeneCards     | http://gsa.muldas.org                    |
| GSA-SNP      | A standalone software that implements three widely-used GSA methods: Z-statistic method, Restandardized GSA and GSEA for pathway analysis, providing a fast computation and an easy-to-use interface | GO, MINT, IntAct, DIP, BioGRID, HPRD, MIPS/MPact | http://bioinfo.mc.vanderbilt.edu/dmGWAS.html |
| dmGWAS       | Designed to identify significant protein–protein interaction (PPI) modules and the candidate genes, specifically adapted for GWAS datasets, including data preparation, integration, searching and validation in GWAS permutation data | GO, MINT, IntAct, DIP, BioGRID, HPRD, MIPS/MPact | http://bioinfo.mc.vanderbilt.edu/dmGWAS.html |
| PLINK        | A whole genome association analysis toolset, designed to perform a range of basic, large-scale analyses in a computationally-efficient manner | NCBi, OMIM, KEGG, UCSC dbSNP, IIDB, PharmGKB, etc. | http://pngu.mgh.harvard.edu/purcell/plink/ |
| Path         | A valuable tool for investigating gene–gene interactions in large genetic association studies, designed to help researchers interface their data with biological information from several bioinformatics resources | KEGG                           | http://genapha.icapture.ubc.ca/PathTutorial |
| Pathway-PDT  | A new tool to perform pathway analysis using the framework of Pedigree Disequilibrium Test (PDT) for general family data, combining more information from raw genotypes in general nuclear families | KEGG                           | https://sourceforge.net/projects/pathway-pdt/ |
| ALIGATOR     | A program for testing for Gene Ontology categories over-represented on a list of significant SNPs from a GWA analysis, defining significant SNPs by prespecified P value cut-off and then counting significant genes in each pathway | GO, dbSNP, NCBI                 | http://x004.pyscm.uwcm.ac.uk/~peter/     |
| GenGen       | A suite of free software tools to facilitate the analysis of high-throughput genomics data sets | KEGG, BioCarta, GO               | http://www.openbioinformatics.org/gengen/ |
| SNP ratio test | Assess significance of enrichment of significant associations in GWA studies and can be applied to pathways such as KEGG/GO or user-defined pathways to test specific hypotheses | KEGG, GO                       | http://sourceforge.net/projects/snpratietest/ |
| i-GSEA4GWAS  | A tool for SNP label permutation, assign SNPs to genes, calculate modified GSEA enrichment score | KEGG, BioCarta, GO               | http://gsea4gwas.psych.ac.cn/            |
| ICSNPathway  | A tool for comprehensive interpretation of GWAS data by integrating LD analysis, functional SNP annotation and pathway-based analysis | KEGG, BioCarta, GO               | http://icsnpathway.psych.ac.cn/          |
by leveraging a priori biological information from multiple sources or integrating different omic data (e.g., GWAS, microarrays and proteomics). For a pathway-based GWAS, improving SNP coverage and, thus, the number of informative genes may also be a feasible solution. For multivariate approaches, imputation for un-typed SNPs or other covariates is an effective way to increase sample size for improved power. Better refined gene set definitions that group genes according to well-defined biological information or by identifying the signal paths within a pathway using the aforementioned topology-based approaches [20,21,45,46] may also be beneficial. The second key issue is to eliminate or reduce the biases introduced during various steps of a pathway-based analysis. For example, large genes containing many SNPs are more likely to contain significant SNPs by chance alone and so are large pathways containing large genes. Some permutation-based approaches that control for gene size by comparing the actual association data with the distribution of association statistics generated from randomly-permuted data sets are expected to reflect chance-based confounding effects, including biases introduced by gene size [53]. However, the permutation procedures per se, if not designed properly, can also introduce some bias [53]. For instance, permutation of SNPs, which is often used in P value-based approaches, can disrupt LD patterns between SNPs and may not generate the correct null distribution. For raw genotype-based approaches, permutation of phenotypes (binary traits or quantitative traits) may not generate the correct null distribution either, as explained previously [53]. Furthermore, no matter whether the SNPs or phenotypes are being permuted, the sampling units are assumed to be independent and identically distributed, which may not be the case, as gene–gene interactions may play an important role in disease susceptibility and study participants might be distantly related. The third key issue is to enhance robustness of pathway-based approaches or repeatability of the significance testing results obtained for pathways. One possible cause for poor repeatability of the results in pathway-based analysis is genetic heterogeneity, in which different variants may account for disease status or trait level in different patients. In addition, the different properties of statistical tests on a disease architecture with no major-effect [53] or arbitrary thresholds used to assess significance of a SNP, gene or gene set may also lead to poor repeatability [54]. In conclusion, although some scientists believe that results from testing gene sets rather than from individual markers would be more stable across different samples in the population and, thus, easier to replicate [56,57], enhancing robustness of pathway-based approaches or repeatability of the significance testing results in analysis of pathways still remains to be a difficult task.

Finally, it is worth noting that the recently-developed topological-based methods for analysis of microarrays have the potential to significantly advance the current methods for the pathway-based analysis for GWAS. The classical ORA [58] or GESA [11,12] pay little attention to the contribution of gene or pathway’s architecture on their biological activity [8,51], while the multivariate approaches developed so far only consider the statistical correlations between SNPs within a gene or between genes within a pathway. It can be imagined that the underlying genetic architecture for most complex phenotypes is far more complicated than any mathematical models could fully accommodate. For example, the commonly-used Pearson’s correlation metric only captures the linear dependence between genes in a pathway, while in reality the biological relationships between genes may not follow linearity. In our own perceptive, incorporating the internal structure or topological properties and environmental triggers into pathway-based analyses is essential for fully assessing the susceptibility of a pathway to a disease. In addition to the merits aforementioned, topology-based approaches have the potential to model and analyze dynamic responses or model effects of external stimuli [22]. All these factors would render topology-based approaches to be a next-generation benchmark methodology for pathway-based analysis of both microarrays and GWAS data.

Extension to analysis of NGS data

Although pathway-based approaches are initially developed for expression microarray data, and later extended to analysis of GWAS data, they may have good potential for a broader application to other omic data like NGS data. Recently, increasing attention is being placed on comprehensive exploration of the genomic variants as high-throughput sequencing has become a feasible solution in practice. The high resolution genome-wide sequence maps provided by these advanced sequencing technologies enable us to examine every detail in sequence variations including SNPs that is used for the traditional GWA studies. In spite of some specific challenges for pathway-based analysis of the massive but more informative sequence data (see the previous review by Wang et al. [53]), NGS data would provide golden opportunities to expand the capacity and power of pathway-based analysis or gene set analysis in general to formulate and test the global hypothesis on disease susceptibility. First, the NGS data could provide necessary coverage to capture all potential genomic subsets [59]. Such refined subsets of candidate genomic regions would allow us to examine all possible genomic structures and their biological roles. Second, the NGS data could provide clues for rare variant discovery. Rare variants and their interactions with common variants and the environment have been shown to contribute to the heterogeneity of several complex diseases [60]. These rare variants may help us to purify the population and to enhance power of a pathway-based analysis on the NGS data. Although the NGS data hold several extended promises, there are several outstanding statistical and practical considerations for performing a pathway-based (or a set-based) analysis of such huge data. To name a few, pathway-based analysis of NGS data requires more efficient statistical methods for detecting genome-wide rare variants, good computational capacity for handling large-scale datasets, improved genotype calling rate for sequencing data and finally the precise functional annotations. Nevertheless, with advent of 1000 Genomes Project [61] and improvements in both high-throughput sequencing techniques and data analytic tools, we believe that it will become feasible to perform sequencing-based GWA studies. It is delightful to see that the first methodological article for pathway analysis on NGS data is published this year. Zhao et al. [62] proposed a novel pathway analysis approach, called smoothed functional principal component analysis (SFPDA), for pathway-based association analysis on NGS data. We anticipate that more and more pathway-based methodologies for dealing with this new data type will appear in the near future. Also, we notice that a de novo sequencing technology, known as Single Molecule Real Time
(SMRT) DNA sequencing [63] is invented last year. It has super capacity in fast sequencing DNA, RNA and methylated DNA (10 times faster than current sequencing facilities). As a result, we can foresee that large amount of sequencing data will be available, providing unprecedented opportunities for fully exploring the power of pathway-based approaches for dissecting complex diseases.

Competing interests

The authors declare no conflict of interests.

Acknowledgements

This study was supported in part by the National Natural Science Foundation of China (Grant Nos. 31071166 and 81373085), Natural Science Foundation of Guangdong Province (Grant No. 8251008901000007), Science and Technology Planning Project of Guangdong Province (Grant No. 2009A030301004), Dongguan City Science and Technology Project (Grant No. 2011108101015) and the Guangdong Medical College Funds (Grant Nos. JB1214, XG1001, XZ1105 and STIF201122).

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