**Gene Polymorphisms and Susceptibility to Functional Dyspepsia: A Systematic Review and Meta-Analysis**

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Functional dyspepsia (FD) is a common chronic gastrointestinal disorder with a complex, undefined mechanism. Clustering of patients with FD in families highlights the role of genetic factors in the pathogenesis of FD [7, 8]. Furthermore, the presence of family history of abdominal pain or family history of indigestion increases the likelihood of developing FD [9]. Gene association studies to evaluate gene polymorphisms encoding neuromodulatory and immunomodulatory proteins related to gastrointestinal motility and visceral hypersensitivity, important in the pathogenesis of FD, have been performed [10]. Although previous studies identified several genes associated with FD susceptibility, the results are inconsistent. Therefore, we conducted a systematic review and meta-analysis to critically evaluate existing literature to determine whether specific genetic polymorphisms are associated with FD susceptibility and also stratified by EPS and PDS subtypes.

1. Introduction

Functional dyspepsia (FD) is a common gastrointestinal disorder affecting 20% of the global population [1]. FD is a complex, multifactorial disorder with possible etiologic factors including visceral hypersensitivity, brain-gut dysfunction, immune activation, Helicobacter pylori (H. pylori) infection, and delayed gastric emptying [2–6]. Patients with FD can be further categorized into epigastric pain syndrome (EPS) and postprandial distress syndrome (PDS) subtypes according to symptom characteristics [2].

Emerging studies demonstrate that susceptibility to FD is influenced by hereditary factors. Clustering of patients with FD in families highlights the role of genetic factors in the pathogenesis of FD [7, 8]. Furthermore, the presence of family history of abdominal pain or family history of indigestion increases the likelihood of developing FD [9]. Gene association studies to evaluate gene polymorphisms encoding neuromodulatory and immunomodulatory proteins related to gastrointestinal motility and visceral hypersensitivity, important in the pathogenesis of FD, have been performed [10]. Although previous studies identified several genes associated with FD susceptibility, the results are inconsistent. Therefore, we conducted a systematic review and meta-analysis to critically evaluate existing literature to determine whether specific genetic polymorphisms are associated with FD susceptibility and also stratified by EPS and PDS subtypes.

2. Materials and Methods

2.1. Literature Search. In order to search for all relevant studies investigating association of gene polymorphisms and FD susceptibility, we conducted a systematic literature search with PubMed, EMBASE, the Cochrane Library, and HuGE database through April 2018 using the following keywords and subject terms: “functional dyspepsia” or “dyspepsia” and “polymorphism,” “mutation,” or “variant.” Detailed
information of search strategy can be found in the supplementary file (available here).

2.2. Eligibility Criteria. The inclusion criteria for studies were as follows: (1) case-control studies or cohort studies assessing the association between any gene polymorphism and FD, (2) sufficient data available to obtain genotypic frequencies to calculate odds ratio (OR) and 95% confidence interval (CI), and (3) studies in adult population. Non-English manuscripts, review articles, conference abstracts, or studies with insufficient demographic data were excluded.

2.3. Data Extraction. Two investigators (L.D., H.R.) independently extracted data. Data on the author, publication year, demographic characteristics, FD diagnostic criteria, genotyping method, and distribution of genotypes were collected. The quality of the studies was assessed by using the Newcastle-Ottawa Scale (NOS) based on three components: selection, comparability, and ascertainment of outcome [11]. From a range of 1 to 9 stars, studies with higher stars were considered to be higher quality. The values of the gene polymorphisms for those with at least three or more available studies were pooled to perform a meta-analysis designed a priori. An additive model that assumes the contribution of each allele to the relative risk was used to prevent multiple testing of differences between each pair of genotypes [12].

2.4. Statistical Analysis. The primary outcome was the OR of a specific gene polymorphism in patients with FD compared to the control population evaluated in three or more published studies. The presence of selection bias in control participants was evaluated by calculating the Hardy-Weinberg equilibrium (HWE), and genotypes frequencies of the control participants were compared using the chi-square test. ORs with 95% CI were calculated to assess the strength of the associations between gene polymorphisms and FD. When studies demonstrated significant heterogeneity, subgroup and sensitivity analyses were performed. Otherwise, a fixed effects model was used. Publication bias was evaluated by Begg’s and Egger’s tests. Two-sided P value <0.05 was considered significant. All statistical analyses were conducted using Stata 13.0 and Review manager 5.3.

3. Results

3.1. Literature Search and Study Characteristics. The initial literature search yielded 1,362 citations, of which 912 remained after removing duplicates. After screening titles and abstracts of studies, 768 were not relevant to the study aim and one was performed in the pediatric population. Furthermore, 70 review articles, 22 case reports, and 16 animal studies were excluded.

Finally, 35 case-control studies met the inclusion criteria for the systematic review. Thirteen studies evaluated the G-protein beta 3 subunit gene (GNB3) 825C>T in 1,390 FD and 3,058 control participants. Four studies evaluated the SCL6A4 serotonin transporter protein (5HTTLPR) in 326 FD and 1,285 control participants. Four studies evaluated the cholecystokinin receptor (CCK-1R) 779T>C in 521 FD and 677 control participants.

Other gene polymorphisms associated with the pathophysiology of FD have been studied in one (serotonin-1A (5HT1A), 5HT3A, 5HT4A, tumor necrosis factor-alpha (TNF-α), interleukin-10 (IL-10), IL-17, IL-1b, regulated upon activation normal T cell expressed and secreted (RANTES), sodium channel Na (SCN10A), neuronal nitric oxide synthase (nNOS), transient receptor potential vanilloid 1 receptor (TRPV1), cytochrome P450 (CYP1A), alpha 2A adrenergic receptor (ADRA2A), glutathione-S-transferases (GSTP1), CD14, catechol-o-methyltransferase (COMT), fatty acid amide hydrolase (FAAH), macrophage migration inhibitory factor (MIF), Toll-like receptor-2 (TLR2), cyclooxygenase-1 (COX-1), ghrelin, and p22PHOX) or two (5HT2A) studies for susceptibility to FD [31–33].

Meta-analysis was performed for the following three genes (GNB3 825C>T, SCL6A4 5HTTLPR, and CCK-1R 779T>C) with three or more studies available meeting the study criteria for meta-analysis. Quality scores of the selected studies ranged from seven to nine indicating moderate to high quality. The study selection process is summarized in Figure 1. Among the eligible studies, all were case-control studies. Only two studies demonstrated selection bias in the control group when calculating HWE [32, 33]. However, the association was not significantly changed when the two studies were excluded from the meta-analysis. Detailed characteristics of the studies included in the meta-analyses are shown (Table 1).

3.2. GNB3 825C>T Polymorphism and FD Susceptibility. The additive model was used to evaluate the association between GNB3 825C>T polymorphism and FD susceptibility (Figure 2, Table 2) [32–44]. Carriers of the minor allele (T) failed to demonstrate susceptibility to FD (OR = 1.15, 95% CI 0.99–1.34, P = 0.07). Given the presence of substantial heterogeneity (I² = 53%), the random effects model was utilized.

Subgroup analyses were further conducted by FD subtypes. Minor allele (T) was associated with increased susceptibility to the EPS subgroup (OR = 1.34, 95% CI 1.10–1.63, P = 0.003) and a trend towards increased susceptibility to the PDS subgroup (OR = 1.19, 95% CI 0.99–1.43, P = 0.07). Further subgroup analyses demonstrated that studies with N > 200 (OR = 1.22, 95% CI 1.01–1.49, P = 0.04) demonstrated increased susceptibility to FD but not in studies with sample size <200 (OR = 1.05, 95% CI 0.86–1.28, P = 0.65). Finally, no increased susceptibility was observed when the studies were stratified by studies from Asian (OR = 1.18, 95% CI 0.82–1.69, P = 0.38) or Western (OR = 1.10, 95% CI 0.97–1.25, P = 0.13) population.

In the sensitivity analysis removing one study at a time, no single study substantially influenced the pooled ORs. Similarly, there was little change in the estimated pooled ORs after excluding studies by Holtmann et al. [32] and Chung et al. [33], whose genotype distribution of the control group deviated from HWE. Finally, no evidence of publication bias was observed according to the Begg’s test (Table 3).
3.3. SCL6A4 5HTTLPR Polymorphism and FD Susceptibility.
SCL6A4 5HTTLPR polymorphism as a predictor of FD susceptibility was examined (Figure 3, Table 2) [34, 35, 43, 45]. No association was observed between minor allele (S) and FD susceptibility (OR = 0.92, 95% CI 0.75-1.12, P = 0.40). Three studies were available for subgroup analysis evaluating susceptibility to FD subtypes. No association was observed in minor allele (S) and susceptibility to either FD subgroups: EPS (OR = 0.73, 95% CI 0.52-1.04, P = 0.08) or PDS (OR = 0.75, 95% CI 0.54-1.04, P = 0.08).

3.4. Other Gene Polymorphisms and FD Susceptibility. CCK-1R 779T>C polymorphism failed to demonstrate susceptibility to FD (OR = 0.86, 95% CI 0.72-1.03, P = 0.09) (Figure 4 and Table 2) [34–36, 46]. Meta-analysis by FD subtype was not possible due to an insufficient number of studies.

In terms of individual studies, nNOS, CD14, MIF, and TRPV1 gene polymorphisms demonstrated increased susceptibility to FD [22, 42]. Ghrelin was associated with feeling of hunger in FD patients in one study [27]. Furthermore, a single study showed that p22PHOX, IL-1b-31CC genotype, and SCN10A were associated with decreased susceptibility to FD [16, 24, 28]. IL-10, IL-17, TNF-α, and 5HT3A were not associated with FD in a single study, respectively [16, 25, 36, 40]. Two studies exploring 5HT2A did not show any association [34, 40]. For studies evaluating FD subtypes, one study suggested that RANTES promoter -28G carrier was associated with a reduced susceptibility to PDS especially in patients with H. pylori infection [15]. COX-1 was associated with susceptibility to EPS in one study [31].

4. Discussion
In the present meta-analysis, we used additive genetic model to assess and measure the associations of the most extensively studied gene polymorphisms and susceptibility to FD. Carriers of the minor allele in genes GNB3 825C>T, SCL6A4 5HTTLPR, and CCK-1R 779T>C in
Table 1: Characteristics of studies included in the meta-analyses of GNB3 825C>T, SCL6A4 5HTTLPR, and CCK-1R 779T>C polymorphisms and FD susceptibility.

| Author (year) | Country     | Diagnostic criteria | Genotyping method | Age (year) (SD) | Gender (n) (M/F) | Genotype, CC/CT/TT, or LL/SL/SS (n) | HWE | NOS |
|---------------|-------------|---------------------|-------------------|-----------------|------------------|--------------------------------------|-----|-----|
| **GNB3 825C>T polymorphism** |             |                     |                   |                 |                  |                                      |     |     |
| Holtmann (2004) | Germany     | Rome II             | PCR               | 46.4 (1.9)      | 44.4 (1.3)       | 20/36/40                     | 34/18/4 46/62/4 0.002 8       |     |     |
| Camilleri (2006) | America     | Rome II             | PCR               | 55.8 (13.0)     | 59.5 (12.7)      | 27/23/17                   | 32/10/8 17/21/1 0.07 7         |     |     |
| Tahara (2008)   | Japan       | Rome II             | PCR               | 60.1 (13.1)     | 61.1 (13.1)      | 33/56/55                   | 20/38/31 23/48/23 0.84 7         |     |     |
| Van Lelyveld (2008) | Netherlands | Rome II             | PCR               | 42.3 (10.6)     | 41.9 (18.3)      | 32/80/94                    | 48/54/10 180/126/30 0.25 9        |     |     |
| De Vries (2009) | Netherlands | Rome II             | PCR               | 49.7 (12.3)     | 40.1 (12.3)      | 62/66/120                  | 60/56/12 199/138/36 0.10 7       |     |     |
| Oshima (2010)   | Japan       | Rome III            | PCR               | 44.5 (13.3)     | 48.0 (18.6)      | 25/43/325                   | 17/29/22 191/368/202 0.37 8       |     |     |
| Shimpuku (2011) | Japan       | Rome III            | PCR               | 59.2 (14.2)     | 37.2 (9.1)       | 36/38/57                   | 14/44/16 17/28/19 0.32 9         |     |     |
| Kim (2012)      | Korea       | Rome III            | PCR               | 49.0 (15.0)     | 47.0 (15.0)      | 62/105/167                 | 52/76/39 112/215/107 0.85 8       |     |     |
| Chung (2014)    | Korea       | Rome III            | PCR               | 46.8 (15.7)     | 50.5 (11.1)      | 21/29/41                   | 5/27/18 15/51/15 0.02 8           |     |     |
| Hwang (2014)    | Korea       | Rome III            | PCR               | 50.3 (18.2)     | 53.3 (19.3)      | 35/77/112                  | 20/62/29 64/132/73 0.77 7        |     |     |
| Yamawaki (2015) | Japan       | Rome III            | PCR               | N/A             | N/A               | 39/35/34                   | 14/46/14 16/29/19 0.46 9          |     |     |
| Singh (2016)    | India       | Rome III            | PCR-RFLP          | 38.4 (12.0)     | 37.3 (11.5)      | 173/64/152                 | 125/86/26 143/95/12 0.45 9       |     |     |
| Triantafyllou (2016) | Greece    | Rome III            | PCR-RFLP          | 50.2 (15.0)     | 53.7 (12.0)      | 61/113/68                  | 99/60/15 137/38/6 0.11 7         |     |     |
| **SCL6A4 5HTTLPR polymorphism** |             |                     |                   |                 |                  |                                      |     |     |
| Camilleri (2006) | America     | Rome II             | PCR               | 55.8 (13.0)     | 59.5 (12.7)      | 27/23/17                   | 14/24/12 14/19/6 0.91 7           |     |     |
| Van Lelyveld (2008) | Netherlands | Rome II             | PCR               | 42.3 (10.6)     | 41.9 (18.3)      | 32/80/94                    | 37/50/25 108/170/58 0.52 9       |     |     |
| Toyoshima (2011) | Japan       | Rome III            | PCR               | 51.8 (17.6)     | 46.8 (17.1)      | 26/27/326                   | 3/20/30 24/174/448 0.17 7        |     |     |
| Hwang (2014)    | Korea       | Rome III            | PCR               | 50.3 (18.2)     | 53.3 (19.3)      | 35/77/112                  | 3/42/66 8/71/185 0.71 7           |     |     |
| **CCK-1R 779T>C polymorphism** |             |                     |                   |                 |                  |                                      |     |     |
| Camilleri (2006) | America     | Rome II             | PCR               | 55.8 (13.0)     | 59.5 (12.7)      | 27/23/17                   | 0/11/39 0/9/30 0.42 7            |     |     |
| Tahara (2009)   | Japan       | Rome III            | PCR-RFLP          | 58.0 (14.0)     | 59.6 (13.1)      | 57/67/71                   | 12/57/55 11/55/53 0.54 7          |     |     |
| Hwang (2014)    | Korea       | Rome III            | PCR               | 50.3 (18.2)     | 53.3 (19.3)      | 35/77/112                  | 4/50/56 13/107/149 0.26 7         |     |     |
| Singh (2016)    | India       | Rome III            | PCR-RFLP          | 38.4 (12.0)     | 37.3 (11.5)      | 173/64/152                 | 19/124/94 46/122/82 0.96 9       |     |     |

M: male; F: female; HWE: Hardy-Weinberg equilibrium; NOS: Newcastle-Ottawa Scale; PCR: polymerase chain reaction; RFLP: restriction fragment length polymorphism; N/A: not available.
| Study or subgroup | FD Events | Control Events | Total | Weight | Odds ratio M-H, Fixed, 95% CI | Year |
|------------------|-----------|----------------|-------|--------|-----------------------------|------|
| Holtmann 2004    | 26        | 112            | 70    | 224    | 0.67 (0.39, 1.12)           | 2004 |
| Camilleri 2006   | 26        | 100            | 23    | 78     | 0.84 (0.43, 1.63)           | 2006 |
| Van Lelyveld 2008| 74        | 224            | 186   | 672    | 1.29 (0.93, 1.79)           | 2008 |
| Tahara 2008      | 100       | 178            | 94    | 188    | 1.28 (0.85, 1.93)           | 2008 |
| de Vries 2009    | 80        | 256            | 210   | 746    | 1.16 (0.85, 1.58)           | 2009 |
| Oshima 2010      | 73        | 136            | 772   | 1522   | 1.13 (0.79, 1.60)           | 2010 |
| Shimpuku 2011    | 76        | 148            | 66    | 128    | 0.99 (0.62, 1.59)           | 2011 |
| Kim 2012         | 154       | 334            | 429   | 868    | 0.88 (0.68, 1.13)           | 2012 |
| Hwang 2014       | 120       | 222            | 278   | 538    | 1.10 (0.80, 1.51)           | 2014 |
| Chung 2014       | 63        | 100            | 81    | 162    | 1.70 (1.02, 2.83)           | 2014 |
| Yamawaki 2015    | 74        | 148            | 67    | 128    | 0.91 (0.57, 1.46)           | 2015 |
| Trintafyllou 2016| 90        | 348            | 50    | 362    | 2.18 (1.48, 3.19)           | 2016 |
| Singh 2016       | 138       | 474            | 119   | 500    | 1.31 (0.99, 1.75)           | 2016 |

Total (95% CI) 2780 6116 100.0% 1.15 (0.99, 1.34)

(a)

| Study or subgroup | EPS Events | Control Events | Total | Weight | Odds ratio M-H, Fixed, 95% CI | Year |
|------------------|------------|----------------|-------|--------|-----------------------------|------|
| Holtmann 2004    | 8          | 32             | 70    | 224    | 0.73 (0.31, 1.71)           | 2004 |
| Camilleri 2006   | 0          | 6              | 6     | 23     | 0.18 (0.01, 3.36)           | 2006 |
| Tahara 2008      | 29         | 50             | 30    | 188    | 1.38 (0.74, 2.59)           | 2008 |
| Oshima 2010      | 53         | 86             | 367   | 1522   | 1.56 (1.00, 2.44)           | 2010 |
| Chung 2014       | 66         | 128            | 394   | 538    | 1.00 (0.68, 1.46)           | 2014 |
| Hwang 2014       | 15         | 32             | 15    | 162    | 0.88 (0.41, 1.89)           | 2014 |
| Trintafyllou 2016| 59         | 234            | 293   | 362    | 2.15 (1.41, 3.28)           | 2016 |
| Singh 2016       | 18         | 52             | 119   | 500    | 1.70 (0.92, 3.11)           | 2016 |

Total (95% CI) 616 3574 100.0% 1.34 (1.10, 1.63)

(b)

| Study or subgroup | PDS Events | Control Events | Total | Weight | Odds ratio M-H, Fixed, 95% CI | Year |
|------------------|------------|----------------|-------|--------|-----------------------------|------|
| Holtmann 2004    | 17         | 80             | 70    | 224    | 0.53 (0.32, 0.99)           | 2004 |
| Camilleri 2006   | 8          | 42             | 23    | 78     | 0.56 (0.23, 1.40)           | 2006 |
| Tahara 2008      | 18         | 32             | 94    | 188    | 1.29 (0.60, 2.73)           | 2008 |
| Oshima 2010      | 43         | 80             | 772   | 1522   | 1.13 (0.72, 1.77)           | 2010 |
| Hwang 2014       | 32         | 54             | 81    | 162    | 1.00 (0.78, 1.27)           | 2014 |
| Chung 2014       | 67         | 116            | 278   | 538    | 1.28 (0.41, 1.92)           | 2014 |
| Singh 2016       | 36         | 142            | 50    | 362    | 2.12 (1.31, 3.43)           | 2016 |
| Trintafyllou 2016| 29         | 110            | 119   | 500    | 1.15 (0.72, 1.84)           | 2016 |

Total (95% CI) 656 3574 100.0% 1.19 (0.99, 1.43)

(c)

Figure 2: Forest plot of studies evaluating GNB3 825C>T and FD susceptibility using the additive genetic model.
FD failed to demonstrate susceptibility to FD. In the subgroup analysis, only minor allele (T) of GNB3 825C>T was associated with increased susceptibility to the EPS subtype.

Guanine nucleotide-binding proteins (G-proteins) play an integral role in the function of stimulus-response coupling of membrane receptors that are linked to intracellular effector system [47]. Hormones, neurotransmitters, and inflammatory stimuli involved in the pathophysiology of FD exert effect on cells probably by binding to G-protein-coupled receptors (GPCRs) [43]. GNB3 is the most widely studied G-protein in various disease processes including depression, cardiovascular disease, obesity, and irritable bowel syndrome [48, 49]. In FD, 825C>T variation-induced signal transduction is more closely linked to pain sensory rather than motility abnormalities. However, the observation of an increased susceptibility to FD in studies with sample size >200 but not in studies with sample size <200 suggested that heterogeneity present smaller studies may have impacted the effect estimate. Furthermore, the effect size of minor allele (T) on FD susceptibility in EPS subgroup was modest and should be interpreted with caution.

5-HT is the primary neurotransmitter involved in the regulation of psychological processes [53]. 5-HT plays a key role in the pathogenesis of both mood disorders and functional gastrointestinal disorders including FD [54, 55]. Psychiatric comorbidities including anxiety disorder and depression are more common in patients with FD compared to the control population [56]. Furthermore, a large body of neurobiological research have demonstrated that psychological factors impact gut physiology such as heightened pain sensitivity to gastric distention in patients with FD [57]. Clinical studies have also demonstrated that 5-HT3 receptor antagonism leads to relief of dyspeptic and anxiety symptoms [58, 59]. Given that 5-HT transporter (SERT) is the key role in the pathogenesis of both mood disorders and functional gastrointestinal disorders including FD [54, 55]. Psychiatric comorbidities including anxiety disorder and depression are more common in patients with FD compared to the control population [56]. Furthermore, a large body of neurobiological research have demonstrated that psychological factors impact gut physiology such as heightened pain sensitivity to gastric distention in patients with FD [57]. Clinical studies have also demonstrated that 5-HT3 receptor antagonism leads to relief of dyspeptic and anxiety symptoms [58, 59]. Given that 5-HT transporter (SERT) is the principle regulator of 5-HT levels by facilitating the reuptake, SERT gene-linked polymorphic region (5HTTLPR) may be involved in the pathogenesis of FD [60, 61]. More specifically, the short (S) allele has been associated with lower transcription and impaired reuptake of 5-HT compared to the long (L) allele [62]. However both overall and subgroup analyses in our study did not show that the carrier of S allele is associated with susceptibility to FD. However,
Impaired gastric emptying is one of the major pathophysiologic mechanisms in FD. CCK-1, secreted by the neuroendocrine cells of the duodenal mucosa, has a physiologic function of delaying gastric emptying and inducing satiety. Furthermore, CCK-1 receptor belongs to the family of

![Forest plot of studies evaluating SCL6A4 5HTTLPR and FD susceptibility using the additive genetic model.](image1)

![Forest plot of studies evaluating CCK-1R 779T>C and FD susceptibility using the additive genetic model.](image2)

Bias affecting results are possible given that data on S allele status to conduct subgroup analysis was not consistently provided by the studies. Additional studies with larger sample size specifically evaluating S allele are needed to validate our findings.
Patients are needed to clarify the role of genetic polymorphisms associated with larger sample size and detailed characterization of the susceptibility independent of H. pylori infection. Several other candidate genes have been explored for FD susceptibility. Genes that play important roles in the regulation of enteric primary afferents and brain-gut interaction (5-HT receptor genes, TRPV1, COMT, and SCN10), induction of inflammatory response (CD14, MIF, TNF-α, IL-17, IL-10, IL-1b, and RANTES), and mediation of gastric accommodation or relaxation (NOS) were hypothesized to be associated with FD susceptibility [24, 30, 65–70]. However, a meaningful meta-analysis was not able to be performed given the sparse number of studies. Furthermore, H. pylori is a potential confounder of FD. Previous studies have demonstrated that homozygous GNBD 825C>T may be associated with dyspeptic symptom among H. pylori-negative subjects [41]. However, the lack of data on H. pylori status in majority of the studies precluded a subgroup analysis to examine the association of genetic polymorphisms on FD susceptibility independent of H. pylori infection.

The present meta-analysis has limitations. First, the language of the manuscripts was restricted to English which may have excluded eligible studies in other languages. Furthermore, variable definition of dyspepsia, controls, and methodologies for measuring SNPs among studies led to study heterogeneity that may have affected the validity of the meta-analysis. In addition, the lack of data on long-term follow-up and treatment response precluded the assessment of the impact of gene polymorphisms on clinical application in FD. Finally, the lack of data on H. pylori status did not allow evaluation of genetic polymorphisms on FD susceptibility independent of H. pylori infection.

In conclusion, carriers of the minor allele in genes GNBD 825C>T, SCL6A4 SHTTLPR, and CCK-1R 779T>C were not associated with susceptibility to FD. In a subgroup analysis, only minor allele (T) of GNBD 825C>T was associated with an increased susceptibility to EPS subtype. The potential role of utilizing gene polymorphisms to decide diagnostic strategy and therapeutic interventions in FD is appealing, but robust evidence is lacking. Additional studies with larger sample size and detailed characterization of the patients are needed to clarify the role of genetic polymorphisms in FD.

Conflicts of Interest
No competing interests to declare.

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Supplementary Materials
A detailed search strategy is provided (using PubMed as functional dyspepsia an example). (Supplementary Materials)

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