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Npp1 promotes atherosclerosis in ApoE knockout mice

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Abstract

Ecto-nucleotide pyrophosphatase/phosphodiesterase 1 (NPP1) generates inorganic pyrophosphate (PPi), a physiologic inhibitor of hydroxyapatite deposition. In a previous study, we found NPP1 expression to be inversely correlated with the degree of atherosclerotic plaque calcification. Moreover, function-impairing mutations of ENPP1, the gene encoding for NPP1, are associated with severe, artery tunica media calcification and myointimal hyperplasia with infantile onset in human beings. NPP1 and PPi have the potential to modulate atherogenesis by regulating arterial smooth muscle cell (SMC) differentiation and function, including increase of pro-atherogenic osteopontin (OPN) expression. Hence, this study tested the hypothesis that NPP1 deficiency modulates both atherogenesis and atherosclerotic intimal plaque calcification. Npp1/ApoE double deficient mice were generated by crossing mice bearing the ttw allele of Enpp1 (that encodes a truncation mutation) with ApoE null mice and fed with high-fat/high-cholesterol atherogenic diet. Atherosclerotic lesion area and calcification were examined at 13, 18, 23 and 28 weeks of age. The aortic SMCs isolated from both ttw/ttw ApoE<sup>+/−</sup> and ttw+/ApoE<sup>+/−</sup> mice demonstrated decreased Opn expression. The 28-week-old ttw/ttw ApoE<sup>+/−</sup> and ttw+/ApoE<sup>+/−</sup> had significantly smaller atherosclerotic lesions compared with wild-type congenic ApoE<sup>+/−</sup> mice. Only ttw/ttw but not ttw+/ mice developed artery media calcification. Furthermore in ttw/+ mice, there was a tendency towards increased plaque calcification compared to ApoE<sup>+/−</sup> mice without Npp1 deficiency. We conclude that Npp1 promotes atherosclerosis, potentially mediated by Opn expression in ApoE knockout mice.

Keywords: atherosclerosis • plaque calcification • osteopontin • inorganic pyrophosphate

Introduction

Plaque calcification is a biomarker of early atherosclerosis [1] and correlates with complication, severity and plaque stability [2, 3]. Whereas arterial calcification in aging, diabetes mellitus and end-stage renal disease is primarily confined to the medial layer, calcification in atherosclerosis occurs in the intimal plaque. Both intima and media calcification appear to be mediated by ectopic cartilage and bone formation, a consequence of the now well-documented transdifferentiation potential of vascular smooth muscle cells (SMCs) [4, 5] and also of osteochondral differentiation by other cells in the artery, including adventitial myofibroblasts [6], stem cells and pericytes [7–9]. The differentiation to ‘bone-forming’ chondrocyte- or osteoblast-like cells is accompanied by the expression of tissue-nonspecific alkaline phosphatase, increased fibrillar collagen type I expression [10] and certain matrix proteins, e.g. osteopontin (OPN), osteocalcin (OC), collagen type II and type X [11–13]. Recent studies in human beings and animal models have demonstrated that (as in mineralizing bone) arterial calcification is inhibited by physiological function of resident cells (including SMCs and endothelial cells) [14, 15]. Specifically, it was shown that SMCs normally express inhibitors of mineralization, such as inorganic pyrophosphate (PPi) and matrix gla protein (Mgp), respectively, and that lack of these molecules leads to spontaneous vascular calcification and increased mortality [16, 17].

One of these inhibitors is the nucleotide pyrophosphatase/phosphodiesterase (NPP) family member NPP1, because severe inborn deficiency of NPP1 is associated with calcification of arteries in mice and human beings [18, 19].

NPP1 is a major generator of extracellular PPi in cartilage and a variety of other tissues, including heart, kidney, liver, pancreas and skeletal muscle [20], PPi potently inhibits hydroxyapatite crystal deposition and growth and regulates chondrogenesis, collagen I expression and synthesis [21] and other cell differentiation processes [22]. These include promotion of the expression of

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OPN by PPi, [19, 23], which in turn promotes atherogenesis via recruitment of monocyte macrophages, and also inhibits arterial calcification by effects that include suppression of hydroxyapatite crystal growth [24].

The physiological role of NPP1-mediated PPi generation in tissue calcification has come to light recently. In human beings, inactivating homozygous or inactivating compound heterozygous mutations of ENPP1 are associated with generalized arterial calcification of infancy (GACI), a frequently lethal form of disseminated arterial media calcification and myointimal proliferation [25]. In Enpp1 knockout mice, hydroxyapatite crystals are found in the aortic medial layer at the level of the internal elastic lamina [18]. These mice additionally develop abnormal calcification of perivascular tissues [23]. A similar phenotype was found in the tw/tw mouse, harbouring a homozygous truncation mutation in the Enpp1 gene [26, 27].

The role of NPP1 in atherogenesis and intimal calcification is unknown. Previous studies have shown that the lack of Npp1 leads to decreased expression of Opn in osteoclasts [23]. Opn is a known chemoattractant for macrophages, the most prominent cell type in atherosclerotic lesions [28]. Additionally, we have shown that NPP1 expression is down-regulated in calcified plaques compared to uncalcified atherosclerotic lesions in human carotid endarterectomy samples [29]. We suggested that atherosclerotic lesion progression is attenuated and calcification in the atherosclerotic plaque is enhanced due to a total or partial deficiency of Npp1. To test our hypotheses, we crossed Npp1-deficient tw/tw mice with ApoE-deficient mice to generate Npp1 and ApoE double deficient mice. ApoE is a structural component of all lipoprotein particles except low-density lipoprotein (LDL). It serves as a high-affinity ligand for lipoprotein receptors and thus allows the uptake of ApoE-containing particles by the liver [30]. Mice deficient in ApoE show increased serum cholesterol and triglycerides leading to atherosclerotic plaque development [31].

Our studies revealed that on the ApoE knockout background, both partial and severe Npp1 deficiency (heterozygous and homozygous tw phenotype) slow down atherosclerotic lesion progression. Furthermore, we found that partial Npp1 deficiency was associated with a tendency towards greater intimal plaque calcification. Thus, our results suggest that in the atherosclerotic plaque NPP1 suppresses calcification but promotes atherogenesis.

Materials and methods

Mice

The creation of the Npp1-deficient tw/tw mouse [a breeding colony of a tw mouse stock with ICR (Institute of Cancer Research) background in the Central Institute for Experimental Animals (Kanagawa, Japan)] used in this study has been described previously [27]. In this study, heterozygous mice are designated as tw/+ Enpp1 wild-type as wt. C57BL/6 ApoE−/− mice (Jackson Laboratory, Bar Harbour, ME, USA) were bred to tw/+ mice. Homozygous double-knockout tw/tw ApoE−/− mice were generated by brother–sister mating of tw/+ ApoE−/− mice on a ICR/C57BL/6 background (1:1).

Enpp1 wt, Enpp1 heterozygous and Enpp1 null mice among ApoE null mice were designated as wt ApoE−/−, tw/+ ApoE−/− and tw/tw ApoE−/−, respectively. Enpp1 and ApoE genotyping was performed by the polymerase chain reaction analysis of tail DNA as described elsewhere ([27]; JAX® Mice database).

The study was approved by the local committee for animal studies (Reg. No.: G21/2004).

Diet and experimental design

The pups were weaned at three weeks of age and then maintained on normal chow diet. At eight weeks of age male littermates (18 tw/tw ApoE−/−, 18...
ttw/+ ApoE−/− and 21 wt ApoE−/−) were set on high-fat/high-cholesterol diet with 16% coconut oil, 1% cholesterol and 0.5% sodium cholate for 5, 10, 15 or 20 weeks. Mice were killed at 13, 18, 23 or 28 weeks of age. Blood samples (0.5–0.8 ml) were obtained by retro-orbital bleed to assess lipid profiles. The arterial tree was fixed by perfusion through the left ventricle at physiological pressure using warmed (37°C) 0.9% NaCl for 4 min. followed by 4% (wt/vol) paraformaldehyde (pH 7.5) for 6 min. The aortic arch, including its main branchpoints (brachiocephalic trunk, left common carotid artery and left subclavian artery) were removed and fixed in 4% paraformaldehyde. All vessels were processed and embedded longitudinally in paraffin. The entire aortic arch was then cut into approximately 30 sections, and a series of 15 sections of 4 μm thickness, which represented the central area of the aortic arch with an intact morphology of the complete arch, was used for analysis.

Plasma cholesterol and triglycerides

Whole blood obtained was centrifugated (5000 rpm, 5 min.) for separating serum and stored at ~20°C. Total serum cholesterol and serum triglyceride levels were determined in duplicate using commercially available enzymatic colorimetric assays (cholesterol CHOD-PAP and triglycerides GPO-PAP; ProDiagnostics, Freiburg, Germany).

Histology and morphometry

For histological analysis, four sections (16 μm apart), stained with Elastica von Giesson were used for the morphometrical analysis. Elastica von Giesson staining, which stains the elastic laminae, was used to delineate the internal elastic lamina, to permit an easier distinction between media and intima. The aortic arch region was defined as the aortic root to approximately 1.5 mm caudal from the left subclavian artery branchpoint. Main branchpoints were also included into the measurements: the brachiocephalic trunk, left common carotid artery and left subclavian artery. Plaque area, determining the underlying media, was defined for each individual plaque as the number of pixels using Adobe Photoshop CS3 extended (San Jose, CA, USA) [32]. Plaque area was converted to square micrometres. Calcified areas were detected in von Kossa stainings in the same serial sections and in the same manner used to determine total plaque area; the relative calcification content was determined by dividing calcified area by plaque area. Opn- and macrophages (MΦ)-containing areas were detected by immunohistochemistry and determined in the same manner in which relative calcification content was estimated. Haematoxylin and eosin standard stain demonstrated general morphology.

Immunohistochemistry

A goat antimouse antibody reacting against OPN, was used in this study (R&D Systems GmbH, Wiesbaden-Nordenstadt, Germany). A monoclonal antibody, reacting against monocytes and macrophages (clone MOMA-2) was purchased from Serotec, Tokyo, Japan. The sections were stained with OPN and MOMA-2 (1:50) followed by biotin-conjugated goat anti-rat IgG followed by Streptavidin-biotin peroxidase complex (ABC Vectastain Elite Kit; Vector Laboratories, Burlingame, CA, USA) and counterstained with nuclear fast red or haematoxylin and eosin.

Isolation and culture of mouse SMCs

To isolate primary SMCs, the aorta of 6-week-old wt ApoE−/−, ttw/+ ApoE−/− and ttw/ttw ApoE−/− mice was explanted, and cultured in Dulbecco’s minimal Eagle medium (DMEM) containing 2% L-glutamine, 100 U/ml penicillin, 50 μg/ml streptomycin and 20% foetal calf serum (FCS) Gold (Lonza, Verviers, Belgium). The sprouted SMC were passaged and cultured in DMEM containing 10% FCS Gold. Only cultures containing more than 90% SMC specific α-actin (using a monoclonal antibody HHH35, 1:500; Dako Corporation, Carpenteria, CA, USA) and SM22α (using a monoclonal antibody 1B8, 1:250; Abcam plc, Cambridge, UK) positive stained cells in passage 3 were used for total RNA isolation.

RT-PCR analyses

Total RNA of SMC was isolated using RNeasy® Mini Kit (Qiagen GmbH, Hilden, Germany). One microgram of total RNA was reverse-transcribed after an initial denaturation step at 65°C for 10 min. in a total volume of 20 μl using 10 U/μl of Superscript™ III reverse transcriptase (Invitrogen™ Life Technologies, Paisley, UK), 1 U/μl Ribo Lock™ ribonuclease inhibitor (MBI Fermentas GmbH, St. Leon-Rot, Germany), 25 ng/μl oligo(dT)12–18 (Invitrogen™ Life Technologies), 0.5 mM/ml each of 2’-desoxynucleoside-5’triphosphate (dNTP), 5 mM/ml dithiothreitol, 1× first strand buffer (250 mM/ml Tris-HCl (pH 8.4), 375 mM/ml KCl, 15 mM/ml MgCl2) freshly diluted from 5× stock (Invitrogen™ Life Technologies) at 37°C for 60 min. Samples were heated at 95°C for 5 min. to terminate reverse transcriptase activity. Total reverse transcription (RT) products were subsequently used for PCR amplification. mRNAs of SMC specific SMC marker (using a monoclonal antibody HHF35, 1:500; Dako Corporation, Carpenteria, CA, USA) and the housekeeping gene Gapdh were assessed in aortic SMC.

The RT products (100 ng) were brought to a volume of 25 μl containing 1× Qiagen GmbH PCR buffer, 2.5 mM MgCl2, 0.1 mM of each dNTP, 1.5 U of Taq Polymerase (Qiagen GmbH) and 0.4 μM of each primer. Primers were designed across intron/exon boundaries. Table S1 summarizes the individual sequences of the primers, annealing temperatures and cycle numbers used for each primer pair and the PCR product sizes. After denaturation at 95°C for 5 min. amplification was carried out in an Eppendorf Mastercycler Gradient (Eppendorf AG, Hamburg, Germany) using the following temperature and time profile: denaturation at 95°C for 30 sec.; primer annealing at the respective temperatures for 60 sec.; primer extension at 72°C for 60 sec. and a final extension of 72°C for 10 min. 10 μl of the PCR reaction were analysed using standard agarose gel electrophoresis. To evaluate the relative expression, the agarose gels were scanned using a video documentation system and analysed using ImageJ software (version 1.40g; ImageJ, imagej.nih.gov/ij/). The absorption units were corrected for Gapdh mRNA levels.

Statistical analysis

Statistical analysis for comparison of multiple groups used one-way ANOVA followed by the Tukey’s post hoc test, performed with GraphPad Prism software (version 4.0, GraphPad software; San Diego, CA, USA). Results were expressed as mean ± S.E.M. Probability values <0.05 were considered statistically significant.
For serum cholesterol and triglycerides samples, blood was obtained by retro-orbital venipuncture from mice with high-fat/high-cholesterol diet (high-fat diet) and from mice fed with normal chow diet (control diet). Data are presented as mean ± S.D. for n = 4–6 mice. One-way ANOVA was used to determine significant differences with Tukey’s post-hoc analysis comparing results from different genotypes. 

*P < 0.05 versus mice with high-fat/high-cholesterol diet, #P < 0.01 and ##P < 0.005, both versus wt ApoE<sup>−/−</sup> and ttw/+ ApoE<sup>−/−</sup> mice.

### Results

#### Npp1 deficiency decreases body weight without affecting serum lipid levels in ApoE null mice

Feeding a high-fat/high-cholesterol diet to wt ApoE<sup>−/−</sup>, ttw/+ ApoE<sup>−/−</sup> and ttw/ttw ApoE<sup>−/−</sup> mice caused hyperlipidemia reflected by elevated serum cholesterol and triglycerides compared to mice with normal chow diet (Table 1). There was no significant difference between genotypes. Body weight measurement revealed no difference between wt ApoE<sup>−/−</sup> and ttw/+ ApoE<sup>−/−</sup> mice, but weights of these genotypes were significantly higher than in ttw/ttw ApoE<sup>−/−</sup> mice. High-fat/high-cholesterol diet had no effect on body weight of all three genotypes compared to mice fed with normal chow diet (Table 1).

#### Opn expression is decreased in Npp1-deficient ApoE null aortic SMC

RNA preparations of cultured aortic SMC of wt ApoE<sup>−/−</sup>, ttw/+ ApoE<sup>−/−</sup> and ttw/ttw ApoE<sup>−/−</sup> littermate mice, demonstrated equal levels of expression of the SMC-specific marker sm22α as well as mgp (Fig. 1). The expression of Opn differed between the genotypes. In cultured ttw/+ ApoE<sup>−/−</sup> SMC Opn expression was down-regulated by approximately 14% compared to wt ApoE<sup>−/−</sup> SMC (P < 0.05), whereas Opn expression in ttw/ttw ApoE<sup>−/−</sup> SMC was decreased by approximately 30% compared to wt ApoE<sup>−/−</sup> SMC (P < 0.001).

Opn was mainly present in areas of the plaque containing macrophages (Fig. 2). These areas were also the major sites of Enpp1 expression in the plaque (data not shown). After 28 weeks, the relative Opn content in atherosclerotic lesions was significant decreased in ttw/ttw ApoE<sup>−/−</sup> mice compared to wt ApoE<sup>−/−</sup> mice (23.7% ± 11.6% <i>versus</i> 47.0% ± 9.9%, P < 0.05, Figs 2A, E and S1). The relative Opn-containing area was lower by trend in ttw/+ ApoE<sup>−/−</sup> mice compared with wt ApoE<sup>−/−</sup> mice (34.7% ± 11.1% <i>versus</i> wt ApoE<sup>−/−</sup> mice, P < 0.1; Figs 2A, C, E and S1).

The macrophages-containing area expressed as percentage of total lesion area was smaller by trend in ttw/ttw ApoE<sup>−/−</sup> mice compared with ttw/+ ApoE<sup>−/−</sup> and wt ApoE<sup>−/−</sup> mice (22.2% ± 14.2% for ttw/+ ApoE<sup>−/−</sup> and wt ApoE<sup>−/−</sup> mice, P < 0.1; Figs 2B, D, F and S1).
Absence of Npp1 attenuates atherosclerosis in ApoE null mice

Mice of the three genotypes were analysed at 13, 18, 23 or 28 weeks of age. At the age of 13 weeks, 60% of ttw/ttw ApoE\(^{-/-}\) mice and 80% of ttw/+ ApoE\(^{-/-}\) and 100% of wt ApoE\(^{-/-}\) mice developed atherosclerotic plaques in the aortic arch (Table 2). One hundred percent of older mice of all genotypes showed plaque development. The atherosclerotic lesion area in the aortic arch was carefully determined on 57 mice. Individual data points plotted by genotype and age are presented in Figure 4. There were no significant differences in the mean lesion area in the aortic arch between groups at 13, 18 and 23 weeks of age (Figs 3A–C, 4A–C and Table 2). However, in 28-week-old mice, ttw/+ ApoE\(^{-/-}\) and ttw/ttw ApoE\(^{-/-}\) mice had significantly smaller atherosclerotic lesions compared with wt ApoE\(^{-/-}\) mice (*P < 0.05 for ttw/ttw ApoE\(^{-/-}\) versus wt ApoE\(^{-/-}\); **P < 0.01 for ttw/+ ApoE\(^{-/-}\) versus wt ApoE\(^{-/-}\); Figs 3D–F, 4D and Table 2). This finding was
consistent when normalizing the lesion areas to the respective body weights of the animals (Fig. S2).

**Npp1 deficiency increases both media and atherosclerotic plaque calcification in ApoE null mice**

von Kossa staining demonstrated aortic media calcifications in 60% of 13-week-old, 80% of 18-week-old and 100% of older ttw/ttw ApoE−/− mice, whereas ttw/+ ApoE−/− and wt ApoE−/− mice were totally devoid of medial calcification (Fig. 3G–I and Table 3). At the age of 28 weeks, plaque calcification developed in all genotypes, in 60% of wt ApoE−/− and 75% of ttw/+ ApoE−/− and ttw/ttw ApoE−/− mice (Fig. 3D–F). There was a non-significant increase in mean size of calcified areas in the intimal layer of ttw/ttw ApoE−/− and ttw/+ ApoE−/− compared with wt ApoE−/− mice (1332 μm² ± 763.2 for ttw/ttw ApoE−/− and 1286 μm² ± 621.1 for ttw/+ ApoE−/− versus 598 μm² ± 290.7 for wt ApoE−/− mice, \( P = 0.36 \) and \( P = 0.32 \); Fig. 5C and Table 3). When expressed as percentage of total lesion area, the calcified lesion area was greater by trend in ttw/+ ApoE−/− mice compared with wt ApoE−/− mice (39.7% ± 19.2% for ttw/+ ApoE−/− versus 6.4% ± 2.8% for wt ApoE−/− mice, \( P < 0.1 \); Fig. 5D and Table 3).

**Discussion**

Our study was consistent with prior findings of media calcifications in the aorta of severely Npp1-deficient mice in the absence
### Table 3: Summary of frequency of medial and intimal calcifications, mean calcified areas and calcified area expressed as percentage of total lesion area

| Genotype       | Region   | 13 weeks | 18 weeks | 23 weeks | 28 weeks |
|----------------|----------|----------|----------|----------|----------|
|                |          |          |          |          |          |
| wt ApoE<sup>-/-</sup> | Media    | 0% (0/4) | 0% (0/6) | 0% (0/6) | 0% (0/5) |
|                | Intima   | 0% (0/4) | 17% (1/6) | 33% (2/6) | 60% (3/5) |
| ttw/+ ApoE<sup>-/-</sup> | Media    | 0% (0/5) | 0% (0/4) | 0% (0/5) | 0% (0/4) |
|                | Intima   | 0% (0/5) | 0% (0/4) | 0% (0/5) | 75% (3/4) |
| ttw/ttw ApoE<sup>-/-</sup> | Media    | 60% (3/5) | 80% (4/5) | 100% (4/4) | 100% (4/4) |
|                | Intima   | 0% (0/5) | 0% (0/5) | 25% (1/4) | 75% (3/4) |

Mean area of intimal calcifications in µm<sup>2</sup>

| Genotype       | Region   | 13 weeks | 18 weeks | 23 weeks | 28 weeks |
|----------------|----------|----------|----------|----------|----------|
|                |          |          |          |          |          |
| wt ApoE<sup>-/-</sup> |          | 0        | 113 ± 113 | 516.5 ± 347.3 | 598 ± 290.7 |
| ttw/+ ApoE<sup>-/-</sup> |          | 0        | 0        | 0        | 1286 ± 621.1 |
| ttw/ttw ApoE<sup>-/-</sup> |          | 0        | 0        | 50.25 ± 50.25 | 1332 ± 763.2 |

Calcified area as percentage of total lesion area

| Genotype       | Region   | 13 weeks | 18 weeks | 23 weeks | 28 weeks |
|----------------|----------|----------|----------|----------|----------|
|                |          |          |          |          |          |
| wt ApoE<sup>-/-</sup> |          | 0%       | 12.6% ± 12.6% | 16.1% ± 10.5% | 6.4% ± 2.8% |
| ttw/+ ApoE<sup>-/-</sup> |          | 0%       | 0%       | 0%       | 39.7% ± 19.2%<sup>a</sup> |
| ttw/ttw ApoE<sup>-/-</sup> |          | 0%       | 0%       | 1.8% ± 1.8% | 30.4% ± 15.9% |

Values represent the mean ± S.E.M. Numbers in parenthesis indicate number of mice in each group. One-way ANOVA was used to determine significant differences with Tukey’s post-hoc analysis.

<sup>a</sup>P < 0.1 versus wt ApoE<sup>-/-</sup> mice.

Fig. 4 Mean atherosclerosis lesion area in individual mice sorted by genotype and age. Mean lesion area in µm<sup>2</sup> was calculated in 13 (A), 18 (B), 23 (C) and 28-week-old mice (D). Bars represent the data mean. One-way ANOVA was used to determine significant differences with Tukey’s post-hoc analysis.

<sup>*</sup>P < 0.05 versus wt ApoE<sup>-/-</sup> mice, <sup>**</sup>P < 0.01 versus wt ApoE<sup>-/-</sup> mice.
of atherosclerosis [18, 26] (and own unpublished results). Interestingly, we did not observe any medial calcification in ttw/H11001 mice with atherosclerosis, suggesting that one functional copy of Enpp1 transcript is sufficient to suppress media calcification. Instead, advanced plaques of 28-week-old ttw/H11001 ApoE/H11002/H11002 mice showed a tendency of an increase in the calcified to total lesion area ratio compared to wt ApoE/H11002/H11002 mice, suggesting that function- or expression-impairing mutations on one allele of ENPP1 may lead to increased calcified areas in late stages of atherogenesis. However this result was not statistically significant, which may be due to limited power of our study. Our results suggest that expression of NPP1 in the arterial wall is necessary to suppress intimal plaque calcification. This finding is in line with our previous study showing that NPP1 expression is down-regulated in calcified plaques compared to uncalcified atherosclerotic lesions in human carotid endarterectomy samples [29]. Interestingly, several studies on the K173Q genotype of ENPP1 in adults associated this with coronary calcification, aortic stiffness, myocardial infarction and earlier onset of peripheral arterial disease [33, 34]. Accordingly, we previously demonstrated a small decrease in enzymatic activity of the K173Q variant of NPP1 in vitro [16]; however, levels of enzymatic NPP1 activity in patients carrying this variant on one allele have not been studied yet.

In our study we show that, although one copy of ENPP1 is sufficient to suppress media calcification, it is not sufficient to suppress intima calcification in the context of atherosclerosis in mice. We would therefore speculate that those patients carrying the K173Q polymorphism of ENPP1 have an increased risk of intima calcification instead of media calcification. Conversely these patients might have a decreased atherosclerotic lesion burden compared to the general population.

In the current study, we show that in ApoE-deficient mice with total absence as well as with the partial loss of Npp1 Opn expression is reduced, leading to reduced Opn-containing areas in atherosclerotic lesions. Previous studies showed decreased Opn levels in Enpp1+/− mice osteoblast and arterial SMC [18, 23, 35]. Studies on different proteins involved in PP i metabolism, e.g. NPP1, the multiple-pass transmembrane protein ANK and the tissue non-specific alkaline phosphatase, determined that OPN expression is directly regulated by extracellular PPi levels [23, 36]. That reduced OPN levels correlate with reduced plaque size has been shown previously. Matsui et al. described 30–40% smaller atherosclerotic lesion areas in 36-week-old female Opn-deficient atherosclerotic (Opn/ApoE null) mice compared with ApoE null mice, due to less macrophages and lymphocytes in the lesion [37]. Also, in female 34-week-old Opn/ApoE/ LDL receptor triple knockout mice, atherosclerotic lesion area is reduced compared with ApoE/ LDL receptor deficient mice [38]. Furthermore, it has been shown that transgenic mice that overexpress Opn are more prone to develop fatty streaks than wild-type mice [39]. We also found that the infiltration of macrophages in atherosclerotic lesions in Npp1-/ApoE-deficient mice was less extensive compared with ApoE-deficient mice by trend. These results are consistent with previous findings that OPN is a chemoattractant for macrophages [28]. We speculate that decreased Opn expression associated with Npp1 deficiency is a likely mediator of decreased atherogenesis in our mouse model.
Taken together, our study revealed that absolute NPP1 deficiency caused by a homozygous null mutation as well as partial deficiency caused by heterozygosity in the ENPP1 locus promote intimal plaque calcification and attenuate atherosclerotic lesion progression. Thus, although mutations on both ENPP1 alleles lead to GACI, mutations on one allele might be protective against atherosclerosis. Decreased OPN expression associated with NPP1 deficiency is a likely mediator of decreased atherogenesis but also likely promotes atherosclerotic plaque calcification.

Limitations of the study

Our study was limited by the short lifespan of Npp1-deficient mice, which is caused by the severity of the skeletal phenotype resulting in a feeding disorder. We had to accelerate atherosclerotic lesion development in Npp1 and ApoE double deficient mice by feeding a cholesterol-rich diet, because Npp1-deficient mice usually survive no longer than 30 weeks. For the atherosclerotic ApoE-deficient mice on chow diet, Rattazzi et al. described small areas of plaque calcification apparent between 45 and 60 weeks of age at the earliest [40]. In our study, because the atherogenesis was accelerated by a high-fat/high-cholesterol diet, we were not able to investigate effects of Npp1 deficiency on ‘natural’ plaque development in ApoE null mice.

Conclusions

Based on the results of our current study and previous work [29], we propose a novel model for modulation of NPP1 expression and the role of NPP1 in promotion of atherogenesis and suppression of intimal plaque calcification (Fig. 6). NPP1 generates PPi and thereby promotes OPN expression that promotes progression of atherogenesis. However, a shift within lesions to decreased TGF-β responsiveness [45] and increased inflammation including IL-1β expression [43, 44] could account for the previously described drop NPP1 expression in atherosclerotic lesion cells including SMCs. This effect, linked with decreased PPi generation and inhibition of OPN expression, promotes plaque calcification. We observe increased atherosclerotic plaque calcification in Npp1/ApoE-deficient lesions in vivo in this study.

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Conflict of interest

The authors confirm that there are no conflicts of interest.

Supporting Information

Additional Supporting Information may be found in the online version of this article.

Fig. S1 Opn- and macrophages-containing areas in individual 28-week-old mice, sorted by genotype. Area containing Opn (A) and macrophages (B) expressed as percentage of total lesion area in 28-week-old mice. Bars represent the data mean. One-way ANOVA was used to determine significant differences with Tukey’s post hoc analysis. *P < 0.5, aP < 0.1.

Fig. S2 Mean lesion area normalized on body weight, sorted by genotype. Values represent mean lesion data in μm² pro g body weight. One-way ANOVA was used to determine significant differences with Tukey’s post hoc analysis. *P < 0.05 versus wt ApoE−/− mice, **P < 0.01 versus wt ApoE−/− mice.

Table S1 Primer and PCR characteristics

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