Fixed Dose Oral Dispersible Tablet of Bitter Drug Using Okra Mucilage: Formulation and Evaluation

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ABSTRACT

Background: The solid oral dosage forms containing bitter drugs need improved palatability for administration. Formulation scientists have given attention to the improvement of taste masking technologies and utilised various strategies.

Objective: The present work aimed to mask the bitter taste of Promethazine Hydrochloride by formulating Oral Dispersible Tablets using Okra mucilage as a taste-masking agent.

Methods: The Okra mucilage was extracted from Okra by the aqueous extraction process. An emulsion solvent diffusion technique was used for masking the bitter taste of Promethazine Hydrochloride by using Okra mucilage. The Oral Dispersible Tablet was prepared by the wet granulation method. The mucilage and the formulation were characterized and evaluated by standard methods and protocols.

Results: Taste masking of the bitter drug was successfully achieved by Okra mucilage. The DSC and FTIR study revealed that the drug molecule was compatible with okra mucilage and drug entrapment efficacy was found to be 94.76%. The palatability test asserted that masking of the bitter taste of the drug. The in vitro drug release study showed that the F7 tablet batch has a better drug release rate and followed non-fickian mechanism of drug release.

Conclusion: Thus, taste masking with Okra mucilage was successful and this opens opportunities for application of common edible substances in formulation development.

Keywords: Fast disintegrating tablet; Natural polymer; Mouth dissolving tablet; Promethazine Hydrochloride; Taste masking

INTRODUCTION

The progress of Oral Dispersible Tablets (ODTs) has acquired more considerable attention among researchers, formulation scientist as well as pharmaceutical industries since the last decade. The ODT formulations are formulated to dissolve or disintegrate quickly at the buccal cavity on exposure to saliva without any addition of water in contrast to the conventional tablet dosage forms. The different names of ODTs are known as quick disintegrating tablets, mouth dissolving tablets, fast disintegrating tablets, rapid-dissolving tablets and porous tablets etc. The novel ODTs are designed for patients having trouble in swallowing or chewing and also suitable for quick medication for critical patients or traveling patients (motion sickness), who are unable to access water for easy administration. The ODTs are put inside the buccal cavity; consequently, tablets are allowed to rapidly disintegrate owing to salivary penetration via the pores of the tablet core without chewing by the patients. Mostly, the disintegration time of ODT is less than one minute (DT<1 minute). However, the actual disintegration time varies from 5 to 30 seconds in different patients. Thus, the active agent can rapidly dissolve in the saliva and be absorbed through whatever membrane it encounters, during deglutition, unless it is protected from pre-gastric absorption. The tablets disintegrate in the mouth in less than 30 seconds by the mechanism of high-swellability, chemical reaction or capillary action. The resulting suspension is swallowed without the need for water, making this dosage form a convenient. So, ODTs are potentially more effective as an alternative to conventional solid dosage forms.

The advantages of ODTs include, it can be given to the patients who cannot swallow (older people, stroke victims, bedridden patients, patients with esophageal problems etc.),
thus improve patient compliance and increased bioavailability. Moreover, ODTs can provide rapid absorption of drugs through the pre-gastric absorption site. The risk of choking or suffocation during oral administration of conventional formulations due to physical obstruction is avoided, thus provides more safety. The disadvantages include its hygroscopicity in nature. So, it must be stored in a dry place and also required special packaging for proper stabilization & safety of the product. 6-8.

The Okra mucilage is a natural polymer, which is a non-starch, a linear water-soluble polysaccharide extracted from the raw unripe Okra fruit (Abelmoschus esculentus (L) Moench, Family Malvaceae). It has a property like swelling, binding, suspending, taste masking activities on bitter drugs. Okra mucilage could mask the taste of the drug sufficiently long enough in the oral cavity for fast dissolving tablets. 9,10.

Taste masking is done for bitter and unpalatable drugs to increase palatability. It is done by various physical and chemical methods that prevent the interaction of taste buds with drugs. Numerous taste-masking approaches are commonly used to overcome bad taste of the drug. For instance, by giving a physical barrier in-between the drug and the taste buds across the drug uptake by granulation, encapsulation, coating by incorporating the sweetener and flavouring agents, that may change the human taste perception; by reduction of drug solubility in saliva, where a balance between reduced solubility and bioavailability must be achieved; or to alter the ability of the drug to interact with taste receptor via chemical modification (Prodrug and salt forms) or by making a complex with an ion exchange resin (e.g. Tulsion335), solid dispersion method, drug particle coating technique etc. that are utilized to successfully mask the bitter taste of the drug in all types of formulations. 11-13.

Considering the above information, it is thought to be worthy of masking the taste of the drug with Okra mucilage and of formulating ODTs for better therapeutic efficacy. In the present study, Promethazine Hydrochloride i.e. (RS) - dimethyl (2-phenothiazine-10-y-propyl) amine hydrochloride (PMZ) is used as a model drug. PMZ is a first-generation H1 receptor antagonist. It is used as an antihistaminic as well as an antiemetic agent. 14,15. In conventional dosage forms, it undergoes the first-pass metabolism, where the oral bioavailability is reduced to 27% from 88% 16. In the present study, an attempt was made to formulate Oral disintegrating tablets (ODTs) of PMZ using Okra mucilage as a taste-masking agent to mask the intensely bitter taste of the above drug.

**MATERIALS AND METHOD**

Promethazine Hydrochloride (PMZ) was obtained as a gift sample from Quality Pharma Pvt. Ltd., Dibrugarh, Assam and India. The fresh Okra pods were purchased from local market Dibrugarh, Assam and India. Microcrystalline cellulose (MCC), Starch 1500, Polyvinyl-pyrrolidone (PVP K-30), Croscarmellose sodium, Lactose, Sodium starch glycollate (SSG) Polyvinyl alcohol (PVA), Aerosil-200, Vanilla flavour were purchased from Yarrow Chem. Products, Mumbai, India. Sodium starch glycollate, sorbitol were purchased from Himedia Ltd., Mumbai, India.

**Extraction of Okra mucilage**

The natural Okra mucilage was extracted by aqueous extraction technique in which water was used as the menstruum under the following consideration 17.

In the aqueous extraction process, the following steps were processed on raw materials to obtain mucilage: 18.

**Extraction of mucilage**

In this step, the raw Okra pods were initially washed with tap water, followed by rinsing with double distilled water to remove all adhering dirt and dried. Then Okra pods were chopped to remove the seeds and homogenized. Then deionized water was added to homogenized Okra pieces in conical flasks at the ratio of 1:1.5 % w/w 19. The flasks were sealed and kept on a constant heating bath at 60°C for around 4-5 hours with agitation. Throughout the extraction processes, the evaporation rate was asserted to be less than 0.05%. After the specified duration of time, the flasks were kept separately for 1 hour at room temperature to complete removal of the mucilage in water 20.

The marc was separated from the extract by straining through a tea mesh. The crude mucilage (filtrate) was kept aside in the refrigerator for overnight to the sediment of soluble soils. The supernatant part of crude mucilage was decanted into a clean and dry flask and concentrated on a water bath at 50-60°C. Three volumes of acetone were used to wash the concentrated samples. The collected precipitate was washed with acetone. Finally, the precipitate was dried at 50-60°C in a hot air oven for 4 hours and passed through sieve nos. 120 & 150 20.

**Characterization of Okra mucilage**

After the extraction process, mucilage solutions were tested to ascertain the presence of carbohydrates by performing the Molisch’s test and Ruthenium red test respectively. Besides, pH, solubility, swelling index, density and viscosity measurements were carried out on dry powder mucilage 21.

**Drug-excipient compatibility studies**

Fourier Transform Infrared spectroscopy and Differential Scanning Calorimetry studied the possible interaction between the drug and the excipients.

**Fourier Transform Infrared spectroscopy (FT-IR) analysis**

The FT-IR spectra of PMZ, Okra mucilage, mucilage coated PTZ and physical mixture of mucilage along with PMZ were analyzed in KBr pellets using the Bruker Alpha FT-IR spectrophotometer, Germany to distinguish the involvement of any interaction between the drug and polymers.

**Differential Scanning Calorimetry (DSC) analysis**

The DSC thermogram of PMZ, Okra mucilage, mucilage coated PTZ, and physical mixture of mucilage were analyzed separately by utilizing a Perkin Elmer JADE DSC (USA) instrument. The samples were placed in an aluminum pan and scanned at a speed of 10°C min⁻¹ in the temperature range of 20-300°C under the inert nitrogen gas atmosphere and thermograms obtained were observed for any interaction among the components present in the formulation 22.

**Methods for masking the bitter taste of PMZ**

**Identification of threshold bitterness concentration PMZ**

A group of ten healthy human volunteers (age 20–25 years) were selected. A series of solutions of PMZ in phosphate buffer of pH 6.8 with concentrations 10, 20, 30, 40, 50, 60, 70, 80, 90, 100 μg/ml were prepared. The volunteers were asked to hold 10 ml of each solution in the oral cavity for 30 seconds. The taste of PMZ was recorded on a numerical scale from 0 to 4, where 0, 1, 2, 3 and 4 were represented with no bitterness, threshold bitter, bitter, moderate bitterness and strong bitterness respectively. Based on the opinion of the volunteers, the threshold bitterness concentration of PMZ was determined.
Taste masking by emulsion solvent diffusion Technique

In this technique, PMZ and powdered Okra mucilage were taken in a ratio of (1: 1, 1: 2, 1: 3, 1: 4, 1: 5 and 1: 6) to obtain an emulsion. The homogenous mixture of PMZ and Okra mucilage powder was first dissolved in a mixture of methanol, acetone and dichloromethane (1:2:3). The above solution was slowly poured in a 1% aqueous Polyvinyl Alcohol (PVA) solution maintained at 40-50°C with continuous stirring at 2000 rpm for 2-3 hours to form an emulsion. The emulsion was kept in a deep freezer at -35°C to solidify. Finally, solidification of the formulation was carried out by Lyophilization technique 24.

Characterization of taste-masked PMZ

Evaluation of In vitro taste-masking:

A required amount of taste masking substance (equivalent to 25 mg of PMZ) was dissolved in six volumetric flasks containing 10 ml of phosphate buffer of pH 6.8. The solutions were shaken for 0, 15, 30, 60, 120 and 300 seconds and filtered. The amount of PMZ content in each filtrate was determined at the respective time. The PMZ content at the end of 120 seconds was considered as the significance of taste masking. It should be less as compared to the threshold bitterness concentration of the PMZ 13.

Drug loading

The drug content in the taste-masked product was determined in triplicate by UV Spectrophotometric method by following formula (1) 25.

\[
\text{Percentage of drug loading} = \frac{\text{Amount of drug in substance recovered}}{\text{Total weight of substance}} \times 100 \quad \text{Eq. (1)}
\]

Drug encapsulation efficiency

The percentage of drug encapsulation efficacy of coated PMZ was determined by using of following formula (2) 25.

\[
\text{Percentage of drug encapsulation efficiency} = \frac{\text{Amount of drug in substance recovered}}{\text{The total amount of drug added}} \times 100 \quad \text{Eq. (2)}
\]

Preparation of Oral Dispersible Tablets of PMZ Using Okra Mucilage

Total 10 batches of ODT of PMZ were formulated by the wet granulation method. The preparation of the tablets was carried out according to three different steps. The steps were given as follows:

Preparation of fast melt granule

All the granulating materials (Table 1) were accurately weighed and sifted through a sieve separately. Then SSG, sorbitol, Starch 1500, Lactose were mixed (Dry mix) and PVPK-30 was dissolved in a binder solution. The binder solution was slowly poured in the dry mixed powder to obtain wet mass. The granules were prepared by passing the wet mass through a sieve no 12# and dried at a temperature of 40-50°C.

Preparation of final blend and tablet compression

The disintegrate, glidants, lubricant materials were sifted through a different type of sieves separately, such as coated substances and fast melt granule through 40#, sorbitol, vanilla flavour, SDS, talc, aerosol-200 through 60#, MCC, CCS and SSG through 30#. All the sifted material except aerosil-200 were mixed in a polyethylene bag for 15 minutes. Finally, aerosil-200 was added to it and mixed for 3 minutes. The compression of tablets was carried out using Shakti 10 station compression machine.

Powder fineness

Five different mesh sizes of sieves were taken, for instance, 20#, 40#, 60#, 80# and 100#. All the sieves were arranged in descending order, i.e., sieve of larger particle size (20#) was at the top and the smallest one at the bottom (100#). The bottom sieve was attached to the receiving pan. Accurately weighed quantity of the sample was placed on the top and agitated for 5 minutes. Then each sieve was carefully removed and the weight of material on each sieve was determined. The fineness of the powder may be expressed as a percentage (% w/w) passing through the sieve(s). Percentage of fineness below 60# mesh size was calculated by using the following formula:

\[
\text{Percentage of fineness below 60#} = \frac{\text{Weight of fines Collected below 60# (gm)}}{\text{Initial Weight of Lubricated granules (gm)}} \times 100 \quad \text{Eq. (3)}
\]

### Table 1: Compositions used in tablet formulation

| Sl. No. | Name of ingredients | Formulation Quantity ingredients (in mg) per Tablet |
|--------|---------------------|--------------------------------------------------|
| 1      | Coated Substances   | F1  175  175  175  175  175  175  175  175  175 |
| 2      | Fast Melt Granule   | F2  150  150  150  150  150  150  150  150  150 |
| 3      | Sorbitol            | F3  2    2    2    2    2    2    2    2    2 |
| 4      | MCC                 | F4  33   31   29   27   33   31   29   27   30 |
| 5      | CCS                 | F5  3    5    7    9    -    -    -    -    -  |
| 6      | SSG                 | F6  -    -    -    -    3    5    7    9    3   |
| 7      | Vanilla flavour     | F7  2    2    2    2    2    2    2    2    2   |
| 8      | SDS                 | F8  2    2    2    2    2    2    2    2    2   |
| 9      | Talc                | F9  3    3    3    3    3    3    3    3    3   |
| 10     | Aerosil-200         | F10 5    5    5    5    5    5    5    5    5   |

Total 375 375 375 375 375 375 375 375 375
Evaluation of blends

The final mixed blend was evaluated by determination of various parameters such as Bulk Density (BD), Tapped Density (TD), Compressibility Index (CI), Hausner’s Ratio (HR), Particle size distribution (PSD), Loss on drying (LOD), angle of repose, percentage (%d) of yield for the final mixed blend was calculated by using the following formula:

\[
\text{Percentage yield} = \frac{\text{The actual Weight of blend}}{\text{Theoretical Weight of blends}} \times 100 \quad \text{Eq. (4)}
\]

Evaluation of formulated tablets

Tablets were evaluated for weight variation, tablet thickness and diameter, hardness, friability, disintegration time, waiting time, the water absorption ratio as per standard methods.

In vitro drug disintegration

One tablet from each batch was introduced into six cylindrical tubes of the disintegration test apparatus. The tubes were moved upward and downward at a rate of 29 to 32 strokes per minute in 900 ml distilled water at 37±2°C. The time taken to pass all the fragments of the tablet through the mesh of the cylindrical tubes was considered as disintegration time.

In vitro drug release study

In vitro drug release test was performed in phosphate buffer solution (PBS) of pH 6.8: The paddle method (USP Apparatus II) was used to carry out the dissolution test of the formulation of all batches (n=10) in 900 ml of PBS. The study medium was kept in a thermostatically controlled water bath, maintained at 37±0.5°C. The pre-weighed tablet was then introduced into the dissolution jar and the paddle was rotated at the rate of 50 rpm. The 5 ml of sample solution was withdrawn from the dissolution compartment at regular intervals (0, 15, 30, 45, 60, 75, 90, 105, 120, 135, 150, 165, 180, 195, 215, 240, 255, 285 and 300 minutes) and immediately replaced with 5 ml of dissolution media to maintain a constant volume. The samples were filtered through Whatman filter paper and the absorbance was determined by UV-Visible Spectrophotometer at 249.6 nm (Shimadzu UV-1800)25,27,28. The cumulative percentages of drug released were calculated.

Wetting time

A tissue paper folded twice was positioned in a small petri dish (internal diameter = 5 cm) having 6 ml of distilled water. A tablet was placed on the folded tissue paper. Finally, the time needed for completion of the wetting of the tablet was measured.

Water absorption ratio

A tissue paper folded twice was positioned in a small petri dish (internal diameter = 6.5 cm) having 5 ml of distilled water. A tablet was placed on the folded tissue paper. The wetted tablet was weighed. The test was done in triplicate. The water absorption ratio (R) was determined according to the following equation:

\[
\text{Water absorption ratio (R)} = \frac{W_a - W_b}{W_a} \times 100 \quad \text{Eq. (5)}
\]

\(W_a\) = weight of the tablet before the test

\(W_b\) = weight of the tablet after water absorption.

Characterization of formulated tablets

The physical characterization of the final formulation was carried out similarly as performed in the case of pure drug and mucilage by using FT-IR, DSC.

Drug release kinetics of ODTs

The kinetics of drug release from ODTs were determined by different in vitro kinetics models such as zero-order (Eq. 6), first-order (Eq. 7), Higuchi (Eq. 8) and Korsmeyer-peppas model (Eq. 9).

\[
F = K_0 t \quad \text{Eq. (6)}
\]

\[
\ln (1 - F) = -K_1 t \quad \text{Eq. (7)}
\]

\[
F = K_2 t^{1/2} \quad \text{Eq. (8)}
\]

\[
M_t / M_b = K_n t^n \quad \text{Eq. (9)}
\]

Where the F is a fraction of drug release in time t and K0, K1, K2 and Kn are rate constants for zero order, first order, Higuchi model and Korsmeyer-Peppas model respectively. M0 is the amount of drug release at time t, M∞ is the amount of drug release at infinity and n is diffusion constant.29 By using the above kinetic model equation, correlation coefficient \(R^2\) values and n were determined. Based on \(R^2\) and n value the suitable drug release model as well as the release mechanism of the formulation was identified.

RESULTS

Physicochemical properties of mucilage:

From Okra pods 0.48 % w/w of mucilage was collected, as shown in Table 2. The presence of mucilage in Okra was confirmed by the development of Purple to the violet colour ring and pink colour (Positive) upon the treatment of Molisch’s reagents and Ruthenium red test respectively (Table 3). The \(\lambda_{max}\) of the Okra mucilage was observed at 201.00 nm in UV-VIS Spectrum analysis17.

Table 2: Percentage yield of Okra mucilage

| Sl. No. | Batches | Quantity of Okra pods (gm) | Weight of mucilage after drying (gm) | Percentage Yield (%) |
|--------|---------|---------------------------|-------------------------------------|---------------------|
| 1.     | Batch 1 | 5000                      | 26                                  | 0.52                |
| 2.     | Batch 2 | 5000                      | 23                                  | 0.46                |
| 3.     | Batch 3 | 5000                      | 28                                  | 0.56                |
| 4.     | Batch 4 | 5000                      | 22                                  | 0.44                |
| 5.     | Batch 5 | 5000                      | 21                                  | 0.42                |
Table 3: Results of the identification test for mucilaginous substances

| Sl. No. | Description     | Observation                  | Results          |
|---------|-----------------|------------------------------|------------------|
| 1       | Molisch’s test  | Purple to violet color ring  | Presence of carbohydrate. |
| 2       | Ruthenium test  | Pink color                   | Present of mucilage |

**Threshold bitterness concentration of PMZ**

The threshold bitterness concentration of PMZ was reported 100 µg/ml by the majority of volunteers (Table 4).

**Evaluation of In vitro taste-masking**

It was found that the amount PMZ present in a buffer solution (pH 6.8) was less than from the threshold concentration at the end of 120 seconds (showed in table 5).

Table 4: For threshold bitterness concentration of PMZ

| Sl. No | Volunteer Code | Rate of Mouth Feel in different concentration PMZ (µg/ml) |
|--------|----------------|----------------------------------------------------------|
|        |                | 10  | 20  | 30  | 40  | 50  | 60  | 70  | 80  | 90  | 100 |
| 1      | A              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 1   |
| 2      | B              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 1   | 1   |
| 3      | C              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   |
| 4      | D              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 1   | 1   |
| 5      | E              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 1   |
| 6      | F              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 1   | 1   |
| 7      | G              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 1   | 1   |
| 8      | H              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 1   |
| 9      | I              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   |
| 10     | J              | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 1   |

Table 5: Amount of PMZ in a different time interval

| Volumetric Flask No. | Shaking Time in Second | Concentration in µg/ml |
|----------------------|------------------------|------------------------|
| 1                    | 0                      | 0                      |
| 2                    | 15                     | 0.16                   |
| 3                    | 30                     | 1.39                   |
| 4                    | 60                     | 1.77                   |
| 5                    | 120                    | 2.05                   |
| 6                    | 300                    | 2.19                   |
The drug loading percentage in coated substance was found to be 13.71%, according to equation (2). It might be due to the loss of some excipients during the process.

The drug encapsulation efficiency of mucilage coated substance was found to be 94.76%. Moreover, the $\lambda_{\text{max}}$ of the UV spectrum of the mucilage coated PMZ solution in water was found to be at 249.4 nm, which was similar to the $\lambda_{\text{max}}$ of pure PMZ at PBS pH 6.8. Thus, the drug molecule might be entrapped by the mucilage of the coated formulation.

**Interpretation of FT-IR spectrum**

All the typical bands of Okra mucilage were present in the spectrum as in our previous study report (shown in figure 1), where $\text{-NH}_2$ group (at 3266.51 cm$^{-1}$), C-H aromatic bond (2937.37 cm$^{-1}$), C=O aldehyde bond (at 1722.63 cm$^{-1}$), C=N group (at 1595.78 cm$^{-1}$), C-H$_2$ group (at 1414.51 cm$^{-1}$) and C-H$_3$ group (at 1370.86 cm$^{-1}$) peaks were observed.

The FT-IR spectra of PMZ, excipients mixture, pure Okra mucilage and coated substances were shown in figure 2 (a, b, c & d). The pure PMZ spectrum showed sharp characteristic peaks of C-H stretching, aromatic C=C stretches, C-H$_3$ and C-H$_2$ bending, C-N is stretching of the tertiary amine band at 2878.33 cm$^{-1}$, 1589.92 cm$^{-1}$, 1453.23 cm$^{-1}$ and 1332.16 cm$^{-1}$ respectively.
Figure 2 (b): Physical Mixture (Drug with all excipients)

Interpretation of DSC thermograms

The DSC thermogram of Okra mucilage demonstrated a sharp endothermic peak at 105.79°C. The DSC thermogram of coated substances showed a sharp endothermic peak at 108.21°C, which is almost similar to the Okra mucilage. The DSC thermograms of pure drug and drug-excipients mixture were shown in figure 3. The DSC thermogram of PMZ showed (curve S1) a sharp endothermic peak at 236.5°C. The physical blend of PMZ and excipients showed (curve S13) a peak of PMZ at 240.1°C.
Characterization of blends and tablets

The compatibility study report of drug–excipients indicated that PMZ and Okra mucilage were compatible with each other. The Cars’ index (CI), Hausen’s Ratio (HR) and Angle of repose of powdered blend were observed in between 12.11% (F3) to 13.66% (F6), 1.14 to 1.15 and 31.86 (F7) to 33.21 (F1) respectively (Table 6). The ODTs were prepared by compression of the powder blend. It was observed that all the ODTs were a white colour, round in shape, flat on both sides with brake line on one side and passed the weight variation test (Table 7). The hardness of all the ODTs was found to be 3.5 to 5.5 kg cm\(^{-2}\) (Table 7). In the dissolution study, the cumulative percentage of drug release of formulating ODTs (F5, F7, F8) was found to be 50% within 90 minutes (Figure 4).

Table 6: Bulk Density (BD), Tapped Density (TD), Compressibility Index (CI), Hausen’s Ratio (HR), Particle size distribution (PSD), Loss on drying (LOD) and Angle of repose of a powdered blend of batches

| Sl. No. | Test | F1  | F2  | F3  | F4  | F6  | F7  | F8  | F9  | F10 |
|--------|------|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 1.     | % Yield | 97.28 | 98.33 | 98.26 | 96.85 | 97.11 | 97.86 | 96.32 | 98.21 | 97.11 |
| 2.     | LOD  | 1.56 | 1.62 | 1.36 | 1.51 | 1.44 | 1.46 | 1.49 | 1.54 | 1.52 |
| 3.     | BD (gm/ml) | 0.416 | 0.423 | 0.421 | 0.433 | 0.436 | 0.422 | 0.435 | 0.419 | 0.422 |
| 4.     | TD (gm/ml) | 0.478 | 0.485 | 0.479 | 0.496 | 0.505 | 0.482 | 0.501 | 0.483 | 0.486 |
| 5.     | CI  | 12.97 | 12.78 | 12.11 | 12.70 | 13.66 | 12.45 | 13.17 | 13.25 | 13.17 |
| 6.     | HR  | 1.14 | 1.15 | 1.14 | 1.15 | 1.16 | 1.14 | 1.15 | 1.15 | 1.15 |
| 7.     | Angle of Repose (θ) | 33.21 | 32.86 | 32.36 | 32.12 | 32.65 | 31.86 | 33.25 | 33.11 | 32.44 |
| 8.     | PSD | 46 | 48 | 51 | 43 | 44 | 48 | 48 | 52 | 54 |

Table 7: Weight Variation, Thickness, Diameter, Hardness and Friability of formulation batches

| Sl. No. | Form. Code | Weight Variation Avg.(mg) ± SD | Thickness Avg(mm) ± SD | Diameter Avg(mm) ± SD | Hardness Avg(kg/cm\(^2\)) ± SD | Friability (%) ± SD |
|--------|------------|-------------------------------|------------------------|-----------------------|-------------------------------|---------------------|
| 1.     | F1         | 376.01±1.61                  | 4.1±0.25               | 10.0±0.01             | 4.5±0.62                      | 0.61±0.12           |
| 2.     | F2         | 375.03±1.25                  | 4.0±0.018              | 10.0±0.01             | 4.0±0.53                      | 0.54±0.31           |
| 3.     | F3         | 377.02±1.65                  | 4.1±0.13               | 10.0±0.00             | 3.9±0.51                      | 0.94±0.11           |
| 4.     | F4         | 376.05±1.33                  | 4.1±0.08               | 10.0±0.02             | 4.9±0.63                      | 0.69±0.08           |
| 5.     | F5         | 376.07±1.57                  | 4.3±0.14               | 10.0±0.00             | 5.2±0.21                      | 0.55±0.09           |
| 6.     | F6         | 377.06±1.69                  | 4.2±0.18               | 10.0±0.01             | 5.5±0.36                      | 0.66±0.12           |
| 7.     | F7         | 375.02±0.63                  | 4.0±0.02               | 10.0±0.00             | 4.7±0.11                      | 0.34±0.02           |
| 8.     | F8         | 376.04±1.15                  | 4.3±0.11               | 10.0±0.02             | 4.5±0.52                      | 0.49±0.10           |
| 9.     | F9         | 376.05±0.95                  | 4.3±0.19               | 10.0±0.02             | 3.8±0.41                      | 0.51±0.14           |
| 10.    | F10        | 375.01±1.11                  | 4.1±0.09               | 10.0±0.02             | 3.5±0.14                      | 0.45±0.21           |
The ODTs of PMZ were successfully prepared with various quantity of super disintegrants (MCC, CCS and SSG) by wet granulation technique. The pre-compression characterization of mixed blends was done for the determination of mass-volume relationships as well as flow properties. The present ODTs of PMZ, the taste-masking, was achieved within the 2 minutes that indicates the PMZ was not released from the tablet core or the released amount was below the human threshold level for noticing its bad taste. Therefore, it might be evidenced that the taste masking was successfully achieved by Okra mucilage. Aside from all the characteristic peaks of PMZ also appeared in the same wavenumber of the spectrum in the physical mixture and the coated formulation. From the comparative study, it was observed that these two spectra were almost the same. It indicated the absence of interaction between the drug and the excipients. The peak shape of DSC thermogram and the melting point is slightly changed from pure Okra mucilage due to entrapment of PMZ. The DSC thermograms of (Figure 3) of PMZ (curve S1), the physical blend of PMZ and all excipients (curve S13) has minor changes in the melting endotherm of the PMZ could be due to the mixing of drug with the impurities present in the excipients. The endothermic peak of individual excipient as well as a physical mixture of excipients with the drug clearly describes the integrity and compatibility of excipients with PMZ.

The powdered bland has an excellent flow property. Tablets have a superior mechanical strength indicating friability below 1%. In the dissolution study (Figure 8), it was observed that the drug release property was significantly increased with the increasing concentration of super disintegrant SSG due to the porous structure SSG is facilitating rapid water uptake into the tablets.

**DISCUSSION**

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Korsemeyer-Peppas model (Eq. 9) was used to understand the release mechanisms, where n is the diffusion exponent. The value of n was estimated by plotting log (M/M₀) of drug release from the tablets versus log t. The value of n was found to be 0.81, which can be ascertained by the release of PMZ from ODTs. It follows the anomalous (Non-fickian) mechanism (n value in between 0.45 < n <0.89) 29.

Table 9: Correlation coefficient (R²) and diffusional exponent (n) of optimum formulation batch.

| Formula Code | Zero Order model | First Order model | Higuchi model | Korsemeyer-Peppas model |
|--------------|------------------|-------------------|---------------|------------------------|
|              | K₀                | R²                | K₁            | R²                     | Kᵦ            | R²                | n               | R²             |
| F7           | 0.497             | 0.996             | 1.0917        | 0.5704                 | 9.4253        | 0.9613            | 0.81            | 0.9843        |

CONCLUSION

The oral dispersible tablets of a bitter drug by using Okra mucilage as a taste-masking agent are beneficial as well as commercially applicable. Moreover, ODTs were rapidly dispersed in the buccal cavity and released the drug immediately. From the dissolution profile, it was observed that the drug release property was significantly increased with the increasing concentration of super disintegrant SSG. The formulation has followed an anomalous (Non-fickian) mechanism of drug release from ODTs.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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