Abstract. Osteosarcoma is a common bone tumor and a frequently occurring cancer-associated threat to children. Notably, the prognosis of osteosarcoma is very poor when it is diagnosed with metastasis. A growing number of studies have indicated that various microRNAs (miRs) serve important regulatory roles in the pathogenesis of different types of cancer. However, the functions of miR-210 in osteosarcoma need to be elucidated comprehensively. The aim of the present study was to investigate the potential roles of miR-210 in osteosarcoma by targeting fibroblast growth factor receptor-like 1 (FGFRL1). Reverse transcription-quantitative polymerase chain reaction results revealed that the expression of miR-210 was highly elevated while FGFRL1 expression was reduced inversely in osteosarcoma tissues compared with matched normal tissues. The results of Transwell assays showed that miR-210 promoted osteosarcoma cell migration and invasion. Furthermore, the luciferase reporter assay results suggested that miR-210 could directly bind to FGFRL1 in osteosarcoma cells. In conclusion, the findings of the present study suggested that miR-210 exerted tumor carcinogenic functions in osteosarcoma by targeting FGFRL1. The findings of this study demonstrated that FGFRL1 was a direct target of miR-210 in osteosarcoma involved in the promoting functions mediated by miR-210 in the invasion and migration of osteosarcoma, suggesting that miR-210/FGFRL1 may be promising for discovering diagnostic and prognostic biomarkers for the therapies of osteosarcoma.

Introduction

Osteosarcoma, a kind of lethal malignant bone tumor, predominantly occurs among children as well as adolescents (1). Accumulating studies have showed that morbidity and mortality are both high in osteosarcoma. Currently, the major therapies for osteosarcoma are chemotherapy and surgery (2). With the development of combination including neoadjuvant chemotherapy and limb salvage surgery, the survival rates of osteosarcoma patients have significantly increased (3,4). However, osteosarcoma patients with metastasis often have a poor prognosis, in the meantime, overall clinical outcomes of osteosarcoma remain unsatisfactory (5). Therefore, it is necessary to understand the molecular mechanisms underlying the osteosarcoma development and progression. Moreover, finding out useful novel biomarkers for the diagnosis and treatment of osteosarcoma is one of the most important clinical problems. In recent years, emerging studies have indicated that miRNAs have important roles in different kinds of tumors.

miRNA is a kind of small non-coding RNA and plays crucial role in regulating gene expression, being considered as cancer biomarkers (6,7). Studies have indicated that the expressions of almost 30% human genes were controlled by different miRNAs (8,9). Briefly, miRNAs can regulate the expression of their mRNA targets by complete or partial binding to the 3'untranslated region (3'-UTRs) (10). Based on the characteristics of their target genes, they can inhibit or promote multiple biological processes of obvious cancers (11). For instance, miR-130b suppresses epithelial ovarian carcinoma by regulating runt-related transcription factor 3 (RUNX3) (12); miR-92b functions as an oncogene in hepatocellular carcinoma via regulating Smad7 (13); miR-143 functions as a suppressor in prostate cancer via targeting matrix metalloproteinase 13 (14). In addition, miR-210 has been reported to modulate the development of many malignancies, including glioblastoma (15), breast carcinoma (16) and pancreatic carcinoma (17). However, the function of miR-210 in osteosarcoma needs to be fully elucidated.

Fibroblast growth factor receptor-like 1 (FGFRL1) is one member of FGFR family and unlike the other members of
FGFR, FGFR1 has no intracellular kinase domain, instead, it contains a short intracellular domain (18,19). Increasing evidence has shown that the FGFR family has vital regulatory roles in proliferation, differentiation and migration of various cancers (20). FGFR1 has also been reported to exert multiple biological functions in different kinds of cancers. However, the specific functions of FGFR1 in osteosarcoma still remain unclear. Present study aimed to investigate the correlation between miR-210 and FGFR1 in osteosarcoma cell migration and invasion.

Materials and methods

Osteosarcoma cell lines and tissue specimens. Two osteosarcoma cell lines (MG63 and Saos-2) and one human normal osteoblast cell line hFOB 1.19 were purchased from Shanghai Institute for Biological Sciences (Shanghai, China). Both the osteosarcoma cells and the hFOB 1.19 were maintained in DMEM medium which contained penicillin-streptomycin and 10% FBS (all from Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) in an atmosphere with 5% CO₂ at 37°C.

A total of 54 osteosarcoma tissue specimens and the corresponding para-carcinoma tissues were collected from the People’s Hospital of Qingdao West Coast New District between 2015 and 2017 and snap-frozen at -80°C. The clinical characteristics of osteosarcoma patients were shown in Table I. All the patients involved in current study signed written informed consent. The present study was approved by the Ethics Committee of the People’s Hospital of Qingdao West Coast New Area (Shandong, China).

Cell transfection. Lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.) was used to transfec miR-210 mimics, inhibitor or scrambled negative miR-control (NC) as well as FGFR1 overexpression plasmid and the negative control vector (GenePharma Co., Ltd., Shanghai, China) into osteosarcoma cells. Plasmids were transfected into osteosarcoma cell lines by X-treme GENE HP DNA Transfection Reagent (Roche Diagnostics, Basel, Switzerland).

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). RT-qPCR was conducted to measure the miR-210 expressions and FGFR1 expressions in osteosarcoma cell lines and tissues. Firstly, the total RNA from osteosarcoma tissues and cells as well as normal tissues was isolated using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA). The total proteins were extracted by lysising cells with RIPA buffer which contained protease and phosphatase inhibitors (Beyotime Institute of Biotechnology, Haimen, China). Put the lysates onto ice for half an hour, then centrifuged them for 20 min at 12,000 x g. The protein concentrations were quantified with the BCA kit (Pierce; Thermo Fisher Scientific, Inc.). The protein lysates were conducted by SYBR® Premix Ex Taq™ (Takara Biotechnology Co., Ltd., Dalian, China) and qPCR assay was conducted by SYBR® Premix Ex Taq™ (Takara Biotechnology Co., Ltd.) on the system of ABI 7900 Sequence Detection System (Applied Biosystems; Thermo Fisher Scientific, Inc.). The miR-210 expression was normalized to U6 while the FGFR1 was normalized to GAPDH. The relative expression levels of genes were calculated by the 2−ΔΔCq method (21). The primers used are shown in Table II.

Western blot analysis. The total proteins were extracted by lysising cells with RIPA buffer which contained protease and...
separated by SDS-PAGE and subsequently transferred onto PVDF membrane which was blocked with 5% non-fat milk and incubated with the primary antibodies at 4°C overnight. Then, a secondary incubation step was performed with appropriate secondary antibody at room temperature for one hour. The proteins were detected by Chemoluminescence method. GAPDH (Abcam, Cambridge, UK) was used as an internal loading control.

Migration and invasion assays. The invasion and migration abilities of the treated osteosarcoma cells were assessed by Transwell assays using Transwell chambers (Corning Incorporated, Corning, NY, USA) with or without Matrigel (Clontech Laboratories, Inc., Mountainview, CA, USA). For the invasion assay, the Transwell chambers were pretreated with Matrigel. Firstly, osteosarcoma cell transfections were suspended in serum-free medium and seeded into the upper chamber. At the same time, DMEM including 10% FBS was joined into the lower chamber. Being cultivated for 48 h, cells on the upper chambers were removed with cotton swabs carefully, while the invasive cells on the lower chamber were subsequently fixed and stained with 4% formaldehyde and 0.1% crystal violet, respectively. The difference between the migration assay and the invasion assay was that there was no Matrigel in the Transwell chambers. An inverted microscope (Olympus Corporation, Tokyo, Japan) was used to measure and count the invasive and migratory cells in five randomly selected fields.

Luciferase reporter assay. The amplified FGFRL1-3'-UTR-WT and corresponding FGFRL1-3'-UTR-MUT were respectively cloned into pGL3 luciferase vector (Promega Corporation, Madison, WI, USA). Osteosarcoma cells were added into a 96-well plate, and after 24 h, FGFRL1-3'UTR-WT and miR-210 mimics or FGFRL1-3'UTR-MUT and miR-210 mimics were transfected into the treated osteosarcoma cells using Lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.). Subsequently, the Dual Luciferase Reporter Assay kit (Promega Corporation) was used to detect the relative luciferase activities 48 h after the transfections.

Statistical analysis. All the above assays were performed three times. Statistical analysis was performed using SPSS statistical software, version 17.0 (SPSS, Inc., Chicago, IL, USA) with Student's t-test and one-way ANOVA with Scheffe's post hoc test. Correlation between mRNA and miRNA were estimated using the Spearman's correlation method. Kaplan-Meier method with log-rank test were applied to estimate the survival rates and compare the survival curves respectively. The data was indicated as means ± SD. P<0.05 was considered to indicate a statistically significant difference.

Results

miR-210 expression is upregulated and FGFRL1 expression is downregulated in osteosarcoma. In this current study, the expressions of miR-210 and FGFRL1 in 54 paired
miR-210 accelerates osteosarcoma cell invasion and migration. Subsequently, we investigated the invasion and migration abilities of osteosarcoma cells which were transfected with miR-210 mimics or inhibitor. The osteosarcoma cells (MG63 and Saos-2) transfected with miR-210 mimics or inhibitor were used to detect the roles of miR-210 in osteosarcoma cell invasion and migration. The results of RT-qPCR assays demonstrated that the expression of miR-210 mimics was high in MG63 (P=0.0035) and Saos-2 (P=0.0041) cells (P<0.01; Fig. 2A). The results of Transwell assays indicated that miR-210 overexpression facilitated migration ability of osteosarcoma cells (P=0.0042 for MG63 cells and 0.0030 for Saos-2 cells). On the contrary, the transwell results also demonstrated a significant decrease in the migration of MG63 (P=0.0048) and Saos-2 (P=0.0035) cells transfected with miR-210 inhibitor in contrast to the control group (Fig. 2B and D). In addition, according to the Transwell assays, the consequences also demonstrated that miR-210 could promote osteosarcoma invasion ability (P=0.0021 for MG63 cells and 0.0019 for Saos-2 cells) (Fig. 2C and E).

miR-210 suppresses FGFR1L gene transcription in osteosarcoma by targeting its 3′-UTR. To investigate whether FGFR1L expression was associated with the expression of miR-210 and to better understand the mechanisms of miR-210 in osteosarcoma, Target Scan was used to find out the target sites in the FGFR1L sequence of miR-210 (Fig. 3A). To confirm the results, the luciferase reporter vector contained the wide-type
(WT) or mutant (Mut) FGFRL1 3’-UTR was constructed. Then, the luciferase reporter assays were performed to detect the fluorescence activities of the FGFRL1 3’UTR in MG63 cells (B) and Saos-2 cells (C) which were cotransfected with wild-type FGFRL1 3’UTR or mutational type FGFRL1 3’UTR and miR-210, respectively (**P<0.01). The results of luciferase reporter assays indicated that miR-210 remarkably reduced the FGFRL1 3’-UTR-WT luciferase activities in both MG63 and Saos-2 cells. However, miR-210 had no significant influence on the FGFRL1 3’-UTR-Mut luciferase activities (P=0.0039 for MG63 cells and 0.0027 for Saos-2 cells) (Fig. 3B and C), suggesting that FGFRL1 was direct target of miR-210. Furthermore, we also detected the FGFRL1 expressions at mRNA level as well as at protein level in osteosarcoma cell lines transfected with miR-210 mimics or inhibitor. The results of RT-qPCR and western blot both demonstrated that miR-210 could inhibit the expression of FGFRL1 in MG63 (P=0.0051 for miR-210 mimics and 0.0023 for miR-210 inhibitor) and Saos-2 cells (P=0.0036 for miR-210 mimics and 0.0036 for miR-210 inhibitor (Fig. 3D-F).

The roles of FGFRL1 in regulating miR-210 effects in osteosarcoma cell migration and invasion. We continued to investigate whether FGFRL1 was needed in regulation of the promoting functions mediated by miR-210 in osteosarcoma cell invasion and migration. Firstly, FGFRL1 overexpression vector and miR-210 mimics were transfected into osteosarcoma cells. Then, RT-qPCR and western blotting were performed to measure the mRNA and protein expression levels respectively and the results indicated that the FGFRL1 expression in osteosarcoma cells transfected with FGFRL1 was significantly increased in contrast to the control group (P=0.0056; Fig. 4A and B). The results of Transwell assays indicated that FGFRL1 deprived the promotion effect of miR-210 on cell migration and invasion, suggesting that FGFRL1 played important roles in the miR-210-mediated biological functions in osteosarcoma cells (P=0.0043 for migration and 0.0036 for invasion) (Fig. 4C and D).
Discussion

Osteosarcoma is one common bone tumor which affects more and more children globally. At present, the primary treatments for osteosarcoma are radiotherapy, chemotherapy and surgical resection (22). Although great efforts have been made to explore the underlying mechanisms of osteosarcoma carcinogenesis, the prognosis is still unsatisfied due to extremely-high metastatic rate as well as aggressive invasion into local tissues (23,24). Therefore, it is urgent to elucidate the molecular mechanism of osteosarcoma to explore the novel therapeutic approaches, such as diagnostic biomarkers and therapeutic targets. During the past decades, increasing studies have demonstrated that various miRNAs are involved in osteosarcoma proliferation, apoptosis and metastasis (25).

Growing evidence has demonstrated that miR-210 is aberrantly expressed in multiple tumors. Additionally, miR-210 can modulate multiple physiological processes, including cell differentiation, apoptosis and survival proliferation via regulating its target genes (26). For instance, miR-210 has been reported to accelerate proliferation, autophagy and angiogenesis of schwannoma cells (27); miR-210 has also been reported to induce endothelial cell apoptosis by directly targeting PDK1 in the setting of atherosclerosis (28); in addition, miR-210-3p can inhibit the bladder cancer growth and metastasis through regulating FGFR1 (29). As we all know, miR-210 has been demonstrated to play important roles in many tumors, but its role in osteosarcoma remains unclear. This study aimed to explore the function of miR-210 in osteosarcoma. The results in this study revealed that miR-210 was significantly upregulated in osteosarcoma. Recent studies have indicated that hypoxia is one of the hallmarks of cancer and cells within the tumor become hypoxic as the tumor mass increases, resulting in activation of HIF-1α to induce various malignant phenotypes (30,31). The potential mechanism underlying the miR-210 overexpression in osteosarcoma is that miR-210 is a hypoxia regulated miRNA and the induction
of miR-210 is a consistent characteristic of the hypoxic response (32). FGFR1L1, the fifth member of the FGFR family, plays important roles in the development of virtually all cell types such as the proliferation, differentiation, migration and apoptosis (33). FGFR1L1 is newly described and usually expressed in adult pancreas and embryonic bone (34). A study has demonstrated that FGFR1L1 overexpression in HEK 293 Tet-On cells suppresses the development of tumor in nude mice (35). For another example, FGFR1L1 can't enhance cell proliferation but induce cell adhesion (36). However, little is known about the potential functions of FGFR1L1 in osteosarcoma. In current study, we investigated the relationship between miR-210 and FGFR1L1 in osteosarcoma.

In conclusion, miR-210 was upregulated not only in osteosarcoma tissues but also in osteosarcoma cells and its expression was correlated with clinicopathological features. In addition, we found that the expressions of miR-210 were negatively correlated with the expression of FGFR1L1 in osteosarcoma tissues. miR-210 exerted tumor carcinogenic functions in osteosarcoma by targeting FGFR1L1 directly. In summary, all the results in this study suggested that miR-210-FGFR1L1 axis may be the novel biomarker for the diagnosis and treatment of osteosarcoma.

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Availability of data and materials

The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.

Authors' contributions

XL and CZ perform the analysis with constructive discussions. CW contributed to analysis and manuscript preparation. JS performed the data analyses and wrote the manuscript. DW and YZ contributed to the conception of the study. XX contributed to manuscript preparation and the conception of the study. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study obtained approval from the Ethics Committee of the People’s Hospital of Qingdao West Coast New Area (Shandong, China). Signed written informed consents were obtained from the patients.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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