Umbilical cord serum concentrations of perfluorooctane sulfonate, perfluorooctanoic acid, and the body mass index changes from birth to 5 1/2 years of age

Takanobu Horikoshi1,2,3*, Tomoko Nishimura2,3, Yoko Nomura2,4,5, Toshiki Iwabuchi2,3, Hiroaki Itoh6, Takumi Takizawa1 & Kenji J. Tsuchiya2,3*

Prenatal exposure to perfluoroalkyl substances (PFAS) has been reported to affect body weight from birth to childhood, but the results remain inconclusive. We investigated whether umbilical cord blood concentrations of perfluorooctane sulfonate (PFOS) and perfluorooctanoic acid (PFOA) are associated with children’s risk trajectory for obesity. 600 children were randomly selected from the Hamamatsu Birth Cohort for Mothers and Children (HBC study) and their umbilical cord serum PFAS concentrations were quantified. Participants underwent BMI measurements at ages 1, 4, 10, 18, 24, 32, 40, 50, and 66 months. Growth curve modeling with random intercept was performed with standardized BMI as outcome variable. PFOS was negatively associated with standardized BMI (β = −0.34; p = 0.01), with a marginally significant interaction with the child’s age (β = 0.0038; p = 0.08). PFOA was negatively associated with standardized BMI (β = −0.26, 95% CI −0.51, 0; p = 0.05), with a significant interaction with the child’s age (β = 0.005; p = 0.01). Stratified analysis by sex revealed that these effects were significant only among girls. Prenatal exposure to PFAS initially was associated with lower standardized BMI during infancy, but this effect dissipated over time and reversed in direction during later childhood. The effects of prenatal PFAS on higher standardized BMI is stronger in girls.

Perfluoroalkyl substances (PFAS), including perfluorooctane sulfonate (PFOS) and perfluorooctanoic acid (PFOA), are a group of synthetic chemicals which have a totally fluorinated carbon chain and a terminal acid1. PFAS have some unique properties, including water and oil repellency and extremely high thermal and chemical stability2. Owing to these properties, PFAS have been used for a variety of products such as carpets, textiles, clothes, cookware, food packaging, and fire-fighting foam since the 1950s3,4. In the 1990s, PFOS and PFOA were detected in the blood of the general human population5. In 2000, the 3 M company, a principal manufacturer of PFAS, announced it would discontinue producing PFOS and related compounds6. In 2009, PFOS was included in Annex B of the Stockholm Convention on Persistent Organic Pollutants, indicating the initial recognition of PFOS as toxic, and it was subsequently banned. Despite the ban, PFOS remained in human serum due to its high persistency in the environment and long serum elimination half-lives (5.4 years for PFOS and 3.8 years for PFOA)7. PFAS accumulate in the blood primarily by binding to albumin8, and can cross the placental barrier9. They act as endocrine disrupters, affecting estrogen, thyroid hormone, and glucocorticoids10–13, and may influence the peroxisome proliferator-activated receptor (PPAR) α that mediates lipid metabolism in adipocytes14. Studies in rodents have shown that prenatal exposure to PFOA causes neonatal death, developmental delay, and postnatal growth deficits15,16.

Thus far, reported associations between PFAS and body weight from birth to childhood are still elusive. Many epidemiologic studies on human subjects have demonstrated an association between prenatal PFAS exposure and

1Department of Pediatrics, Gunma University Graduate School of Medicine, Maebashi, Japan. 2Research Center for Child Mental Development, Hamamatsu University School of Medicine, Hamamatsu, Japan. 3United Graduate School of Child Development, Hamamatsu University School of Medicine, Hamamatsu, Japan. 4The City University of New York, New York, NY, USA. 5The Icahn School of Medicine at Mount Sinai, New York, NY, USA. 6Department of Obstetrics and Gynecology, Hamamatsu University School of Medicine, Hamamatsu, Japan. *email: Hotaka1001@gmail.com; tsuchiya@hama-med.ac.jp
either a null or small reduction in birthweight\textsuperscript{17–20}. During childhood, studies have shown that increased prenatal exposure to PFAS was associated with a greater risk of obesity or weight gain\textsuperscript{21–24}, while others have shown no notable associations\textsuperscript{25,26}. A recent meta-analysis found that exposure to PFOA in early life is associated with an increased risk for childhood adiposity\textsuperscript{27}. Chen et al. showed that higher umbilical cord plasma PFOS concentration was associated with lower birthweight and height, but these effects diminished as children aged\textsuperscript{28}. They further showed the effects were more obvious among girls. However, as the authors acknowledged, considerable attritions during the follow-up period and their analytical plan which lacks adjusting for gestational age or parity prevented them from drawing a clear conclusion. To clarify and validate this line of investigation, capitalizing on the available longitudinal and frequent assessments and very low attrition rate in the Hamamatsu Birth Cohort for Mothers and Children (HBC Study)\textsuperscript{29}, we set out to evaluate the association between prenatal exposure to two most widely studied PFAS, PFOS and PFOA, and the trajectory of BMI from birth to early childhood (i.e., 5 1/2 years) with adjustment of a priori determined important potential confounders.

**Methods**

**Study population.** The study population is derived from a birth cohort study in Hamamatsu, Japan, the HBC Study, with data from mothers (n = 1138) and their children (n = 1258) born between December 2007 and March 2012. A majority (98.8%) of enrolled mothers were Japanese. All mothers were enrolled at either one of the two research sites (Hamamatsu University Hospital and Kato Maternity Clinic) between November 2007 and March 2011 during the first or second trimester of pregnancy and gave birth at Hamamatsu University Hospital. As previously reported\textsuperscript{30}, enrolled mothers and children were representative of the general Japanese population with regard to age and socioeconomic status of the families, and the child's birthweight and gestational age at birth, compared to Japanese government's national statistics\textsuperscript{29,31}. The details of the parent study have been described elsewhere\textsuperscript{29}.

Of the 1258 live births, umbilical cord venous blood was collected from 1244. Among the 1244 available samples, constrained by available funds, 600 were randomly selected for assays. Of those 600, 1 case was excluded due to the child having developed serious developmental delays (Fig. 1). Two more were excluded as the assay values were beyond the limit of detection (LOD) of the assay used to measure PFOS and PFOA concentrations (see below), leaving 597 samples.

**Exposure.** We used umbilical cord serum PFOS and PFOA concentrations as the measurement of prenatal exposure. Umbilical cord blood (UCB) was collected immediately after birth and placed at room temperature for 30 min. It was then centrifuged, and the serum layer was transferred to Eppendorf tubes and stored at −80 °C freezer until analysis. PFOS and PFOA concentrations were analyzed at Shimadzu Techno-Research, Inc. (Kyoto, Japan) using a high-performance liquid chromatography (HPLC) (Promincence; Shimadzu Corporation) coupled with a tandem mass spectrometer (MS/MS) (API 4000; AB Sciex) followed by solid phase extraction. Detailed methods are described in supplemental document.

---

**Figure 1.** Flow chart of the participant selection.
Outcomes. We used BMI to evaluate children's physical status. Between January 2008 and September 2017, the participating children were invited for assessments at Hamamatsu University Hospital at age 1, 4, 10, 18, 24, 32, 40, 50, and 66 months. At each assessment, the children's weight was measured to the nearest 0.1 kg and height to the nearest 0.1 cm. BMI was transformed into standard deviation scores (SDS) against the data from the Japanese Society for Pediatric Endocrinology. This standardized BMI (BMI SDS) was used as our outcome variable.

Statistical analysis. Statistical analyses were performed using STATA version 14.2 (StataCorp LP, Texas, USA) and Mplus 7.4 (www.statmodel.com). PFOS and PFOA concentrations were log10 transformed to reduce the influence of outliers and right-skewed distribution. Growth curve modeling with random intercept was used to analyze association of cord serum PFAS levels with a series of BMI SDS records over time as a best-fit line for each participant, considering individual variation of BMI SDS at birth (i.e. at baseline). Similarly, interaction between PFOS or PFOA concentration and months of age was calculated and entered into the models to estimate the slope of the best-fit line of BMI SDS over time. Thus, estimated intercept parameters represent BMI SDS at birth and estimated slope parameters indicate change in PFOS or PFOA concentration per month throughout the observation period. PFOS and PFOA concentrations were tested both dimensionally and categorically (i.e., tertiles) to assess potential non-linear association between PFAS concentration and child body weight. Based on our prior research findings, variables that were associated with both PFAS concentrations and child body weight were included as covariates in the adjusted model, including maternal age at delivery, maternal BMI before pregnancy, maternal education, household income, maternal smoking during pregnancy, parity, gestational age, and duration of breast feeding. Since a recent study suggests that the magnitude of the association between PFAS and BMI differed between males and females, we conducted an additional analysis by sex to see whether the associations under investigation can be observed for both sexes.

Of the 597 infants included, 30 (5.0%) infants had 5 or more missing data on BMI, and 18.8% had at least one missing BMI data across 9 observations between 1 and 66 months. We leveraged the central methodological strength of growth curve modeling and generated estimates for the missing data using maximum likelihood (ML) method. ML involves the computation of a case-wise likelihood function using all observed variables for a particular case, while including partially complete cases to estimate parameters for the missing data. Monte Carlo studies have shown that ML involves less restrictive assumptions about patterns of missing information and yields unbiased parameter estimates, increases the efficiency of parameter estimates, and eliminates bias in estimation arising from listwise or pairwise deletion and mean substitution of cases.

Ethical issues. This study was approved by the Hamamatsu University School of Medicine and University Hospital Ethics Committee. Written informed consent was obtained from all mothers for her own and her child's participation. All research was performed in accordance with the relevant guidelines and regulations.

Results
534 mothers and 597 children participated in this study (Table 1). There were no significant demographic differences between those included and those excluded except for in birth weight (p = 0.05) and parity (p = 0.009) (see Supplementary Table 2 and Supplementary Table 3, Supplementary Information).

Mean cord serum concentrations (ng/mL) of PFOS and PFOA were 1.38 and 1.39, respectively (Table 2). In the analysis using a dimensional measure of log10-transformed concentrations of PFOS and PFOA, there was a significant effect of PFOS concentration on the intercept of BMI SDS in both unadjusted and adjusted analyses,
indicating that a higher PFOS concentration was associated with a lower BMI SDS at birth (Table 3). Meanwhile, the interaction between PFOS concentration and child’s age, representing the slope of the individual best-fit line, was only marginally significant. Associations were clearer with PFOA. Specifically, higher PFOA concentration was associated with lower BMI SDS at birth in the adjusted model. Furthermore, there was a significant positive interaction between PFOA concentration and child’s age on BMI SDS, representing the positive slope (increase in the BMI SDS as they grow), in both unadjusted and adjusted model. In the analysis by sex, only among girls higher PFOS and PFOA concentrations were associated with lower BMI SDS at birth and had significant positive interactions with child’s age on BMI SDS (positive slope) indicating that greater levels of PFOS and PFOA are associated with greater increased in the BMI SDS as they grow.

To identify the high risk group by PFAS exposure levels, we repeated the analysis above with categorical measures (low, medium, and high) of PFOS and PFOA concentrations, with the low tertile group set as a reference. Only the high PFOS concentration was associated with lower BMI SDS at birth significantly (Table 3, Fig. 2a). There was a significant positive interaction between the high PFOS concentration and child’s age on BMI SDS, again indicating the greater BMI SDS as they grow (positive slope). Consequently, BMI SDS for the high PFOS

| Table 2. | Measured concentrations of cord serum PFOS and PFOA (ng/mL). NB: PFOS denotes perfluorooctane sulfonate; PFOA denotes perfluorooctanoic acid. |
| --- | --- |
| **PFOS** | **PFOA** |
| Mean | 1.38 | 1.39 |
| Standard deviation | 0.21 | 0.22 |
| Range | 0.21, 7.10 | 0.22, 10.00 |
| Low tertile | 0.21, 0.99 | 0.22, 0.91 |
| Middle tertile | 1.00, 1.50 | 0.92, 1.50 |
| High tertile | 1.60, 7.10 | 1.60, 10.00 |

| Table 3. | Results of the analysis treating PFOS and PFOA concentrations as dimensional and categorical measures. Controlling covariates: maternal age at delivery, maternal BMI before pregnancy, maternal education, household income, maternal smoking during pregnancy, parity, gestational age, and duration of breast feeding. NB: SE denotes standard error, PFOS denotes perfluorooctane sulfonate, PFOA denotes perfluorooctanoic acid. |
| --- | --- |
| Intercept | Interaction with age |
| β (SE) p value | β (SE) p value |
| Dimensional | | |
| PFOS | | |
| Overall | −0.27 (0.14) 0.05 | −0.35 (0.14) 0.01 | 0.0038 (0.002) 0.08 | 0.0038 (0.002) 0.09 |
| Boys | 0.001 (0.18) >0.99 | −0.095 (0.19) 0.62 | 0.0011 (0.003) 0.71 | 0.0011 (0.003) 0.72 |
| Girls | −0.67 (0.2) 0.001 | −0.69 (0.2) 0.001 | 0.0081 (0.003) 0.15 | 0.0082 (0.003) 0.14 |
| PFOA | | | | |
| Overall | −0.22 (0.12) 0.07 | −0.26 (0.13) 0.05 | 0.0051 (0.002) 0.01 | 0.0051 (0.002) 0.01 |
| Boys | 0.15 (0.18) 0.4 | 0.16 (0.19) 0.41 | −0.001 (0.003) 0.72 | −0.001 (0.003) 0.74 |
| Girls | −0.59 (0.17)<0.001 | −0.62 (0.18)<0.001 | 0.011 (0.003)<0.001 | 0.011 (0.003)<0.001 |
| Categorical | | | | |
| PFOS | | | | |
| Overall | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) |
| Boys | 0.042 (0.071) 0.55 | 0.022 (0.07) 0.75 | −0.0001 (0.0012) 0.91 | −0.0001 (0.0012) 0.92 |
| Girls | −0.16 (0.075) 0.03 | −0.175 (0.076) 0.02 | 0.0029 (0.0012) 0.02 | 0.0029 (0.0012) 0.02 |
| PFOA | | | | |
| Overall | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) |
| Boys | 0.14 (0.10) 0.16 | 0.11 (0.1) 0.28 | −0.0022 (0.0017) 0.2 | −0.0022 (0.0017) 0.2 |
| Girls | −0.021 (0.11) 0.84 | −0.03 (0.11) 0.75 | 0.0012 (0.0017) 0.49 | 0.0012 (0.0017) 0.49 |
| PFOA | | | | |
| Overall | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) | 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) 0.00 (Reference) |
| Boys | 0.0049 (0.10) 0.96 | 0.026 (0.1) 0.8 | 0.0029 (0.0017) 0.08 | 0.0029 (0.0017) 0.08 |
| Girls | 0.044 (0.10) 0.67 | 0.03 (0.11) 0.79 | 0.0003 (0.0017) 0.88 | 0.0003 (0.0017) 0.86 |

indicating that a higher PFOS concentration was associated with a lower BMI SDS at birth (Table 3). Meanwhile, the interaction between PFOS concentration and child’s age, representing the slope of the individual best-fit line, was only marginally significant. Associations were clearer with PFOA. Specifically, higher PFOA concentration was associated with lower BMI SDS at birth in the adjusted model. Furthermore, there was a significant positive interaction between PFOA concentration and child’s age on BMI SDS, representing the positive slope (increase in the BMI SDS as they grow), in both unadjusted and adjusted model. In the analysis by sex, only among girls higher PFOS and PFOA concentrations were associated with lower BMI SDS at birth and had significant positive interactions with child’s age on BMI SDS (positive slope) indicating that greater levels of PFOS and PFOA are associated with greater increased in the BMI SDS as the child grows.

To identify the high risk group by PFAS exposure levels, we repeated the analysis above with categorical measures (low, medium, and high) of PFOS and PFOA concentrations, with the low tertile group set as a reference. Only the high PFOS concentration was associated with lower BMI SDS at birth significantly (Table 3, Fig. 2a). There was a significant positive interaction between the high PFOS concentration and child’s age on BMI SDS, again indicating the greater BMI SDS as they grow (positive slope). Consequently, BMI SDS for the high PFOS
concentration was lower than that of the low PFOS concentration from birth to infancy, but gradually increased over time as the child grew and exceeded it at approximately 63 months of age. A similar pattern was found for PFOA. Only the high, but not the middle, PFOA concentration was associated with significantly lower BMI SDS at birth. There was a significant positive interaction between the middle and the high PFOA concentration and child’s age. Here again predected BMI SDS increase in these groups (middle and high) relative to low group as they grow. Furthermore, the predicted BMI SDS was lower in children with the high PFOA concentration than children with the lower concentration in infancy but it increased over time and exceeded it at about 47 months of age among children exposed to high PFOA. In the analysis by sex, only among girls, higher PFOS and PFOA concentrations were associated with lower BMI SDS at birth and had significant positive interactions with child’s age on BMI SDS (i.e., the rate of the increase on BMI SDS is positive as they grow) in both unadjusted and adjusted analyses (Fig. 2b). At about 65 months for PFOS and 50 months for PFOA, BMI SDS of girls with high level of exposure exceeded than that of girls with low level of exposure.

**Discussion**

**Principal findings.** We prospectively examined the trajectory of BMI SDS in 597 children over 66 months (5½ years) through an examination of the PFAS exposure in UCB serum. There were three main findings. First, prenatal exposure to PFAS was associated with lower BMI SDS during infancy. Second, it was associated with an increase in BMI SDS in later childhood. Although the effect sizes of the interactions between PFAS concentration and child’s age (i.e. the rate of growth in BMI SDS over time) are relatively small in early ages, the actual impact on BMI SDS at 66 months of age is not negligible. For example, an effect size of 0.001 for the interaction (slope) means that 10-time increase in PFAS concentration (due to the log-10 transformation) resulting in 0.66 SD increase in BMI at 66 months of age, which is substantial. Third, girls may be more vulnerable than boys to prenatal exposures to PFAS. These findings are consistent with previous research and support the hypothesis that PFAS may have the initial BMI-lowering effect in early infancy, but the effect could reverse during early childhood and become possible obesity risks.
Interpretation. Our findings of PFOA are consistent with other studies that reported an association between higher prenatal PFAS exposure and an elevated risk of obesity at 8 years and 20 years of age\cite{21,26,30}. However, our study ameliorated the limitation presented by Chen et al.\cite{28} that did not include gestational age in covariates and further provided evidence that prenatal exposure to PFOS was associated with lower BMI SDS in infancy and then upward trajectory of BMI SDS development as the offspring grow. However, unlike the results by Chen and colleagues, in our study, PFOA was associated with greater BMI SDS later in childhood, especially among girls. Low birth weight itself would lead to obesity in adulthood\cite{37}.

It is important to consider adiposity rebound (AR) when interpreting our findings. AR is generally explained by the BMI increasing rapidly after birth, then starts to decrease at around 1 year, reaching its lowest concentration at 5 to 7 years of age\cite{38}. Our results suggest that higher UCB serum PFAS concentrations may be associated with an earlier onset of AR, which is a well-known predictor of an elevated risk of obesity and metabolic syndrome in adulthood\cite{39–42}. Since reducing child obesity is an ultimate form of prevention for future diseases and reduction in the related social costs, one possible effective prevention can be to increase our surveillance of PFAS in the environment especially during pregnancy and offering early interventions targeted toward those children at a higher risk of obesity by virtue of prenatal PFAS exposure\cite{41}. It is also noteworthy among 599 samples we investigated, in 597 (99.5%) samples we found PFAS residues in UCB, indicating the prenatal exposures to the substances are pervasive. While our findings are preliminary and still require replication, they are alarming and deserve further investigation.

PFAS have an effect on a child’s body weight through different potential mechanisms. First, PFAS bind to PPAR, which are ligand-activated transcription factors and involved in lipid metabolism. The PPAR family was reported to regulate adipogenesis and weight gain, and is affected by chemical compounds such as tributyltin\cite{44}. Both PFOS and PFOA significantly activate PPARα\cite{14}, which regulates lipid metabolism in the liver. Second, PFAS may act as an endocrine disrupter which have effects similar to estrogen\cite{45,46}. Prenatal exposure to environmental estrogens may affect the expression of genes related to obesity via modifying the programming of various estrogen target tissues during early development\cite{37}. In line with this theory, two prior studies reported that the association between prenatal exposure to PFAS and the risk of obesity was stronger among girls than boys, which is consistent with our findings\cite{24,36}. Third, it is possible that prenatal exposure to PFAS affects body weight via a change in thyroid function\cite{10}, although there are a limited number of studies, and the results are inconclusive. As hypothyroidism is known to decrease prenatal weight gain\cite{46}, it can be an underlying mechanism for decreased BMI during infancy. However, this does not explain the increase in BMI SDS during later childhood. Last, maternal serum concentrations of PFAS in pregnancy were reported to be inversely associated with UCB glucocorticoid, and may lead to lower birthweight\cite{11}. However, we found no notable associations between these two variables in our samples.

In our study, the participants’ year of birth (from 2007 to 2012) was associated with decreased concentrations of PFAS (β = −0.28, 95% CI −0.35, −0.21 for PFOS, β = −0.26, 95% CI −0.33, −0.20 for PFOA). This is consistent with previous studies, which reported that concentrations of PFOS and PFOA in human serum peaked c. 2000 and subsequently continued to decrease\cite{49}. This would be attributable to the global phase-out of production of these compounds in the early 2000s. However, the results remained unchanged after adding the year of birth in the model. As stated above, almost all of our participants (99.5%) were exposed to PFAS despite that they were born later than 2000. This strongly urges policy makers and researchers to keep monitoring the PFAS levels in the environment.

Strengths of the study. The current study had several strengths. First, the study is a longitudinal study of our established cohort, with prospective observation and repeated measures from birth to 66 months of age with low attrition rates over time. With such a study design, the valid evaluation of longitudinal changes of BMI in infancy and then upward trajectory of BMI SDS development as the offspring grow is possible. Having multiple assessment points further allows us to use more advanced and sophisticated statistical methods that help uncover the complexity of the outcome among children at greater risk for obesity in childhood – the group that did not show initial problems with excessive weight gain during infancy. Moreover, using umbilical cord serum, not maternal serum, samples enables direct assessment of prenatal PFAS exposure among offspring.

Limitations of the study. The study also has several limitations. First, we have only assessed the cohort for their BMI up to 66 months (5½ years) of age. Optimally, we would have longitudinal observation beyond 5½ years of age because the mean age of AR is around 6 years of age. Second, measuring equipment for weight and height was not unified across each measurement. Also, measurement errors could have been minimized if the measuring equipment had been unified. Third, the children’s exposure to PFAS might not have been limited to exposure in the prenatal period. Postnatal exposures to PFAS, such as through breast milk, drinking water, diet, and dust inhalation, could also be possible. Last, although BMI in childhood can be affected by dietary habits\cite{50}, no variable reflecting dietary habits was incorporated in our analyses.

Conclusions

Our findings demonstrate that the effect of prenatal exposure to PFAS is a possible risk for obesity through an increase in BMI SDS, especially for girls. As PFAS are widely detected in lifestyle choices and the environment, it is important to understand the health effects of PFAS as well as their pathways and mechanisms to suboptimal health outcomes during childhood.
35. McArdle, J. J. & Hamagami, F. Multilevel models from a multiple group structural equation perspective. In: Advanced structural equation modeling techniques (eds Marcoulides, G. & Schumacker, R.) 89–124 (Lawrence Erlbaum Associates, Hillsdale, NJ, 1996).
36. Halldorsson, T. I. et al. Prenatal exposure to perfluorooctanoate and risk of overweight at 20 years of age: A prospective cohort study. Environ Health Perspect. 120, 668–673. doi:10.1289/ehp.1104034 (2012).
37. Barker, D. The developmental origins of chronic adult disease. Acta Paediatr. 93, 26–33. https://doi.org/10.1111/j.1651-2227.2004.00023.x (2004).
38. Rolland-Cachera, M. F. et al. Adiposity rebound in children: A simple indicator for predicting obesity. Am. J. Clin. Nutr. 39, 129–135. https://doi.org/10.1093/ajcn/39.1.129 (1984).
39. Whitaker, R. C., Pepe, M. S., Wright, J. A., Seidel, K. D. & Dietz, W. H. Early adiposity rebound and the risk of adult obesity. Pediatrics 101, E5 (1998).
40. Williams, S. M. & Goulding, A. Patterns of growth associated with the timing of adiposity rebound. Obesity (Silver Spring) 17, 335–341. https://doi.org/10.1038/oby.2008.547 (2009).
41. Eriksson, J. G., Forsen, T., Tuomilehto, J., Osmond, C. & Barker, D. J. Early adiposity rebound in childhood and risk of Type 2 diabetes in adult life. Diabetologia 46, 190–194. https://doi.org/10.1007/s00125-002-1012-5 (2003).
42. Koyama, S. et al. Adiposity rebound and the development of metabolic syndrome. Pediatrics 133, e114–119. https://doi.org/10.1542/pedi.2013-0966 (2014).
43. Pigeot, I., de Henauw, S. & Baranowski, T. The IDEFICS (Identification and prevention of Dietary- and lifestyle-induced health effects in children and infants) trial outcomes and process evaluations. Obes. Rev. 16(Suppl 2), 2–3. https://doi.org/10.1111/obr.12328 (2015).
44. Grun, E. et al. Endocrine-disrupting organotin compounds are potent inductors of adipogenesis in vertebrates. Mol. Endocrinol. 20, 2141–2155. https://doi.org/10.1210/me.2005-0367 (2006).
45. Zhao, Y. et al. Perfluorooctanoic acid effects on ovaries mediate its inhibition of peripubertal mammary gland development in Balb/c and C57Bl/6 mice. Reprod Toxicol 33, 563–576. https://doi.org/10.1016/j.reprotox.2012.02.004 (2012).
46. Liu, C., Du, Y. & Zhou, B. Evaluation of estrogenic activities and mechanism of action of perfluorinated chemicals determined by vitellogenin induction in primary cultured tilapia hepatocytes. Aquat. Toxicol. (Amsterdam, Netherlands) 85, 267–277. https://doi.org/10.1016/j.aquatox.2007.09.009 (2007).
47. Newbold, R. R., Padilla-Banks, E., Jefferson, W. N. & Heindel, J. J. Effects of endocrine disruptors on obesity. Int. J. Androl. 31, 201–208. https://doi.org/10.1111/j.1365-2605.2007.00858.x (2008).
48. Karakosta, P. et al. Thyroid dysfunction and autoantibodies in early pregnancy are associated with increased risk of gestational diabetes and adverse birth outcomes. J. Clin. Endocrinol. Metab. 97, 4464–4472. https://doi.org/10.1210/jc.2012-2540 (2012).
49. D’Eon, J. C. & Mabury, S. A. Is indirect exposure a significant contributor to the burden of perfluorinated acids observed in human serum? Environ Sci Technol 45, 7974–7984. https://doi.org/10.1021/es200171y (2011).
50. Emmett, P. M. & Jones, L. R. Diet, growth, and obesity development throughout childhood in the Avon Longitudinal Study of Parents and Children. Nutr. Rev. 73(Suppl 3), 175–206. https://doi.org/10.1093/nutrit/nuv054 (2015).

Acknowledgements

The authors would like to thank Dr. Tetsuo Kato of the Kato Maternity Clinic, as well as Drs. K. Sugihara, M. Sugimura, K. Takeuchi, K. Suzuki, Y. Murakami, Y. Kohmura, Y. Miyaide, K. Hirai, Y. Nakamura, R. Koizumi, H. Murakami, Y. Kobayashi-Kohmura, K. Muramatsu-Kato, Prof. N. Kanayama, and all attending obstetricians of Hamamatsu University School of Medicine, for their full support with participant enrolment in the study. The authors also thank the chief midwife, Ms. Kiyomi Hinoki, and all midwives and staff at the maternity clinic of Hamamatsu University School of Medicine, for their support with participant enrolment. The authors appreciate the technical support of Ms. R. Gonda and E. Hatano, and of Drs F. Daigo and M. Yasojima, Shimadzu Techno-Research, Inc. in measuring cord blood PFAS. The authors thank Mr. C. Davey for his helpful comments with technical writing in English. The HBC Study Team includes E. Hijashimoto, N. Kodera, A. Nakamura, C. Nakayasu, Y. Nishimura-Amma, H. Suzuki, Y. Kugizaki, Y. Suzuki, R. Takabayashi, M. Honda, H. Muraki, M. Narumiya, E. Sato, Mr. R. Nakahara, Drs. D. Choi, T. Harada, A. Okumura, M. Tsujii, K. Wakusawa, Y. Kamenoh, D. Kurita, H. Kuwabara, K. Takebayashi, M. Yokokura, T. Wakuda, R. Asano, T. Ismail, K. Iwata, Y. Iwata, E. Kawai, M. Kawai, Y.K. Kuroda, K. Matsumoto, H. Matsuzuki, T. Mori, K. Nakazumi, K. Nakamura, A.A. Pillai, Y. Seno, C. Shimmura, S. Suda, G. Sugihara, T. Sugiyama, K. Suzuki, K. Yamada, S. Yamamoto, Y. Yoshihara, Y. Endoh, K. Hirano, T. Suzuki, Prof. H. Yasumase, Prof. N. Takei, Prof. H. Matsuzaki, Prof. N. Mori.

Author contributions

T.H., T.N., Y.N., T.T. and K.J.T. were involved in study design and data interpretation. T.H., T.N., T.I., H.I. and K.J.T. were involved in the data collection and analysis. All authors critically revised the report, commented on drafts of the manuscript, and approved the final manuscript as submitted and agreed to be accountable for all aspects of the work.

Funding

This study was supported by JSPS KAKENHI Grant Number JP16H05374 and 19H03582 (K.J. Tsuchiya). The funding source had no involvement in the study design, the data collection, the data interpretation and discussion.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1038/s41598-021-99174-3.

Correspondence and requests for materials should be addressed to T.H. or K.J.T.
