Prenatal diagnosis of a novel pathogenic variation in the ACAN gene presenting with isolated shortening of fetal long bones in the second trimester of gestation: a case report

Paolo Toscano 1,2, Lavinia Di Meglio 2,3*, Fortunato Lonardo 4, Letizia Di Meglio 2, Laura Letizia Mazzarelli 1,2, Carmine Sica 2 and Aniello Di Meglio 2

Abstract

Background: Heterozygous mutations of the ACAN gene are a major cause of different evolutive growth defects in the pediatric population, but were never described as a cause of fetal skeletal dysplasia.

Case presentation: A G1 at 21w + 3d came to our institution for the second-trimester ultrasound and a skeletal dysplasia with prevalent involvement of limb’s rhizomelic tracts was suspected. Amniocentesis followed by CGH-array was performed, with normal results. An examination by NGS of some genes associated with skeletal dysplasias showed a novel pathogenic variant of the ACAN gene: c.2677delG.

Conclusion: Sequence variations of ACAN were never described as a possible cause of fetal skeletal anomalies to date. In this case report, we describe the first prenatal diagnosis of skeletal dysplasia associated with a pathogenic variant of ACAN.

Keywords: Prenatal diagnosis, Skeletal dysplasia, ACAN, Aggrecan, Case report

Background

The prenatal finding of fetal short long bones represents a challenging condition for the physician and the patient, due to its association with skeletal dysplasia, chromosomal abnormalities, and genetic syndromes [1–3]. In the postnatal life, heterozygous pathogenic sequence variations of the ACAN gene (coding for the proteoglycan aggrecan) were associated with evolutive growth defects, ranging from mild idiopathic short stature to severe skeletal dysplasias. ACAN mutations were considered responsible for 1.4% of cases of idiopathic short stature and 13.8% of cases of short stature and advanced bone age in the pediatric population [4–6]. A literature review on PUBMED and EMBASE did not identify reports of ACAN mutations in the antenatal period as a cause of fetal short limbs [4–6].

Here we report the first in-utero diagnosis of a novel heterozygous sequence variation in the ACAN gene in a fetus with severe fetal growth restriction with prevalent reduction of the rhizomelic bones biometry.
Case presentation
A 41-year-old woman (G1) turned to our second-level prenatal ultrasound diagnostic center to undergo the second-trimester screening exam at 21w + 3d of gestation. Besides maternal age > 35 years, in vitro fertilization (IVF) was the only relevant risk factor. Gestational age errors were excluded since this pregnancy was obtained by IVF. Fetal growth was overall ten days smaller than the one attended based on gestational age, with a fetal growth corresponding to 19w + 6d of gestation. In particular, growth’s delay was mostly due to fetal bones: femur was under the third centile, humerus corresponded to the fifth centile and mesomelic bones were all > the 5° centile, with the only exception of the tibia (1.7° centile) (Table 1 and Fig. 1). Fetal anatomy was regular, except for a slight prominence of the forehead with a low nasal bridge and mild nasal hypoplasia, with nasal bone at 17° centile (Fig. 2). No other skeletal anomaly and in particular no polydyctyly or bone fractures were detected. The uterine, umbilical, and middle cerebral artery pulsed-wave Doppler measurements were all normal, except for a slight prominence of the forehead. The amniotic fluid was regular. Infective diseases were ruled out based on laboratory tests. Considering the ultrasound findings, a skeletal dysplasia with prevalent involvement of the proximal limbs was suspected.

Amniocentesis was performed to exclude chromosomal, genomic, and genetic anomalies, with particular regard to some genes associated with skeletal dysplasias.

Table 1  Biometry report

| GA: 21w + 3d | Biometry | Gestational age | Media + – 2DS | Percentile |
|--------------|----------|----------------|--------------|------------|
| BPD          | 44.2 mm  | 19w            | 51.7 + – 5.0 | 6.7        |
| HC           | 183.9 mm | 20w + 6d       | 189.3 + – 29.6 | 42.8   |
| CEREB        | 22.1 mm  | 21w + 1d       | 22.4 + – 2   | 44.0       |
| PV           | 6.7 mm   |                |              |            |
| AC           | 146.0 mm | 19w + 2        | 173.7 + – 26 | 16.1       |
| HL           | 26.0 mm  | 19w            | 32.4 + – 4.0 | 5.5        |
| ULNA         | 25.3 mm  | 19w + 3 s      | 30.9 + – 4.0 | 8.1        |
| RAD          | 22.1 mm  | 19w + 3d       | 27.9 + – 4.0 | 7.4        |
| FL           | 27.2 mm  | 18w + 5d       | 34.9 + – 4.0 | 2.7        |
| TIB          | 24.5 mm  | 19w + 1d       | 30.9 + – 3.0 | 1.7        |
| FIB          | 25.1 mm  | 19w + 5d       | 29.9 + – 3.0 | 5.5        |
| FOOT         | 31.0 mm  | 19w + 2d       | 37.3 + – 4.0 | 5.8        |
| NB           | 6.5 mm   | 19w + 4d       | 7.2 + – 0.7  | 17.7       |
| EFW          | 306 g    |                |              | < 2.5      |

G4 gestational age, BPD biparietal diameter, HC head circumference, CEREB cerebellum, PV posterior ventricle, AC Abdominal circumference, HL Homeurs length, RAD radial length, FL Femur length, TIB Tibial length, FIB Fibula length, NB Nasal bone, EFW estimated fetal weight

Array-based Comparative Genomic Hybridization (CGH-array) and Next Generation Sequencing (NGS) were conducted in parallel. The following databases were used to select the genes that needed to be analyzed: Human Phenotype Ontology, OMIM and/o GeneReviews. CGH-array was performed at the Centro Poldiagnostico s.r.l., Naples, Italy, on a CGH Array platform SurePrint G3 ISCA V2 CGH 8X60K, resolution: 250 kb, Software: Agilent Cytogenomics 5.0.2.5. NGS was performed at Bambino Gesù Pediatric Hospital, Rome, Italy, on a NovaSeq6000 Illumina platform; Genes analyzed: NM_000142.4 (FGFR3) and NM_001135.3 (ACAN). The NGS revealed a heterozygous c.2677delG variant in the ACAN gene, which determines the p.Gly893AspfsTer52 protein variation. The mutation is located in exon 12 of the gene and in the first chondroitin sulfate (CS1) attachment domain of the protein. The mutation arose de novo (it wasn’t present in both parent’s DNA). Based on the clinical features associated with ACAN mutations a diagnosis of osteochondrodysplasia with short limbs and a possible future involvement of the spine and joints were done. The couple opted for voluntary interruption of pregnancy at 25 weeks of gestational age. No autopsy was performed.

Discussion
Growth’s delay occurs in almost 10% of pregnancies; the major causes are maternal, placental, fetal, or genetic factors; DNA sequence variations are responsible for almost one-third of growth’s delays [7]. If a growth’s delay is suspected, it is mandatory to collect a complete maternal and familial history, to evaluate previous ultrasound scans, to track fetal growth, to assess gestational age from both the date of the last menstrual period and the first-trimester ultrasound, to evaluate all biometry, to rule out placental problems and infective diseases [7]. In our case report, we describe a fetus that presented 10 days growth delay, mostly due to small fetal rhizomelic bones. Fetal skeletal anomalies occur in 5:1000 pregnancies and, when the gynecologist/fetal radiologist suspects a skeletal anomaly, a genetic consultation should be offered and fetal DNA may be analyzed [3]. In almost 50% of cases, the gene responsible for that anomalies is known, however, in 15% of cases, a variant of uncertain significance is detected, making it difficult for both the geneticist and the gynecologist/fetal radiologist to answer to the couple’s questions [2, 3].

In our case report, amniocentesis followed by CGH-array and NGS was performed and the fetus was found out to be a de novo carrier of a heterozygous pathogenic sequence variation of the ACAN gene, which codes for aggrecan. Aggrecan is a proteoglycan component in the extracellular matrix largely present in the articular cartilage and the growth plate [8]. Aggrecan is responsible for

---

Toscano et al. BMC Pregnancy and Childbirth (2021) 21:459
the load-bearing capacity of the joints. The glycosaminoglycan side chain attracts counter-ions and water and creates the osmotic swelling pressure that is crucial for biomechanical properties of cartilage under compressive load [8–10]. Moreover, aggrecan is also involved in the autocrine and paracrine regulation of chondrogenesis and osteogenesis [8–10]. Studies conducted on animals with homozygous mutations of the ACAN gene showed a premature vascular invasion and ossification of the growth plate with a premature maturation and block of

![Fig. 1 Fetal estimated body weight. Fetal estimated body weight under the 5°centile](image)

![Fig. 2 Ultrasounds skeletal findings. A. Femur and tibia< 5° centile B. Prominence of the forehead with mild nasal hypoplasia. C. Humerus 5° centile](image)
bone elongation [11]. Very interestingly, the tissue in which ACAN is expressed at by far the highest levels is the tibial artery (Data Source: GTEx Analysis Release V8, dbGaP Accession phs000424.v8.p2), and in our case, the most compromised bone is the tibia, whose size corresponds to 1.7° centile.

At least 50 pathogenic ACAN variations have been identified in patients with highly variable phenotypes of syndromic or non-syndromic short stature (Table 2); no genotype-phenotype relationship was apprised [4–6, 8, 9, 11]. However, in all of these different diseases, the phenotype is characterized by short bones and major involvement of the joints and especially of the spine.

Small trials involving GH therapy after birth have been conducted in children with ACAN mutations and the therapy resulted in a possible height improvement [6].

In this case, NGS revealed a heterozygous c.2677delG variation in the ACAN gene sequence, which resulted in the protein variation p.Gly89eAspfsTer52. The following databases were analyzed to interpret the variant: VarSome, ClinVar, Human Gene mutation database, Leiden Open Variation Database; gnomAD was used to assess the variant’s frequency in the general population (variant not found). The null (frameshift) variant c.2677delG can be classified according to the ACMG guidelines as a Pathogenic variant, since it meets a Very Strong (PVS1), a Moderate (PM2), and a Supporting (PP3) criterion [12]. No mutation involving the FGFR3 gene was detected.

Despite the variability of the clinical features related to ACAN mutations, the genetic variation detected in this case, resulted in a rare form of osteochondrodysplasia with short limbs and possible future involvement of the spine and joints.

From a literature review, ACAN mutations have not been reported in the antenatal period, although in the pediatric field they are routinely screened in children with short stature [11].

In this case report, we, therefore, describe a genetic diagnosis never made before in the antenatal period of a pathogenic sequence variation not so far described in the literature. More studies involving fetuses with short limbs are advocated to better understand the impact of ACAN mutations in the prenatal age.

**Conclusions**

Since ACAN gene mutations are among the main causes of genetic short stature in the pediatric population, based on our findings, we believe that these should be considered also in the prenatal differential diagnosis of fetal growth’s delay in a fetus with short limbs. The presence of short limbs, even if not all below the pathological value of the 3rd centile, should suggest the possibility of an underlying genetic condition. In case of suspicion of skeletal dysplasia, the ACAN gene should be considered. An early diagnosis is particularly important in cases in which, after birth, it is possible to practice without delay a therapy that can positively influence the clinical picture and, ultimately, the quality of life of the patient.

**Abbreviations**

IVF: in vitro fertilization; CGH-array: Array-based Comparative Genomic Hybridization; NGS: Next Generation Sequencing

**Acknowledgments**

not applicable.

**Authors’ contributions**

ADM, CS ideated the manuscript; LaDM, PT wrote the original draft. FL, LM, LeDM reviewed critically the manuscript and edited the final draft. FL and LeDM performed the revision of the draft. All authors approved the final version.

**Funding**

The authors declare that no funding were used.

**Availability of data and materials**

All data generated or analysed during this study are included in this published article.

**Declarations**

**Ethics approval and consent to participate**

The study was conducted in accordance with the Helsinki Declaration (Fortaleza revision 2013), the Good Clinical Practice Standards (CPMP/ICH/135/95), and the current Decree-Law 196/2003 regarding personal data and all the requirements set out in the European regulations on this subject.

**Consent for publication**

Written informed consent was obtained from the patient for publication of this Case report and any accompanying images. A copy of the written consent is available for review by the Editor of this journal.

**Competing interests**

The authors declare that they have no competing interests.

**Author details**

1 Department of Neuroscience, Reproductive Sciences and Dentistry, School of Medicine and Surgery Federico II of Naples, University of Naples Federico II, Naples, Italy. 2 Diagnostica Ecografica e Prenatale di A. Di Meglio, Via dei Fiorentini n.21, Naples, Italy. 3 Department of Obstetrics and Gynecology, H. Buzzi, University of Milan, Milan, Italy. 4 Department of Medical Genetics, A.O.R.N. “San Pio”, Benevento, Italy.

**Received:** 23 March 2021 **Accepted:** 10 June 2021

**Published online:** 29 June 2021

**References**

1. Kajimura M, Ulander VM, Ryynanen M, Stefanovic V. Risk of Adverse Outcomes in Euploid Pregnancies With Isolated Short Fetal Femur and

---

**Table 2 Diseases associated with ACAN mutations**

| Isolated Short Stature |
|------------------------|
| Spondyloepiphyseal dysplasia, aggrecan type (OMIM 612813) |
| Spondyloepiphyseal dysplasia, Kimberley type (OMIM 608361) |
| Short stature and advanced bone age with or without early-onset osteoarthritis and/or osteochondritis dissecans (OMIM 165800) |
1. Humerus on Second-Trimester Sonography. J Ultrasound Med. 2016;35(12). https://doi.org/10.7863/ultra.16.01086.

2. Krakow D, Lachman RS, Rimoin DL. Guidelines for the prenatal diagnosis of fetal skeletal dysplasias. Genet Med. 2009;11(2). https://doi.org/10.1097/GIM.0b013e3181971ccbb PMID: 19265753; PMCID: PMC2832520.

3. Schwarz M, Mommens H. Fetal Skeletal Disorders. Ultraschall Med. 2018;39(6). https://doi.org/10.1055/a-0660-9417.

4. Gibson BG, Briggs MD. The aggrecanopathies: an evolving phenotypic spectrum of human genetic skeletal diseases. Orphanet J Rare Dis. 2016;11(1).86. https://doi.org/10.1186/s13023-016-0459-2.

5. Hauer NN, Stich H, Boppudi S, Büttner C, Kraus C, Trautmann U, et al. Genetic screening confirms heterozygous mutations in ACAN as a major cause of idiopathic short stature. Sci Rep. 2017;22(1):7(1). https://doi.org/10.1038/s41598-017-12465-6.

6. van der Steen M, Pfundt R, Maas SJWH, Bakker-van Waarde WM, Odink RJ, Hokken-Koelega ACS. ACAN Gene Mutations in Short Children Born SGA and Response to Growth Hormone Treatment. J Clin Endocrinol Metab. 2017;102(5). https://doi.org/10.1210/jc.2016-2941.

7. Sharma D, Shastri R, Farahbakhsh N, Sharma P. Intrauterine growth restriction - part 1. J Matern Fetal Neonatal Med. 2016;29(24). https://doi.org/10.3109/14767058.2016.1152249 Epub 2016 Mar 7. PMID: 26856409.

8. Lauing KL, Cortes M, Domowicz MS, Henry JG, Batta AT, Schwartz NB. Aggrecan is required for growth plate cytoarchitecture and differentiation. Dev Biol. 2014;396(2). https://doi.org/10.1016/j.ydbio.2014.10.005.

9. Day JM, Olin AI, Murdoch AD, Canfield A, Sasaki T, Timpl R, et al. Alternative splicing in the aggrecan G3 domain influences binding interactions with tenascin-C and other extracellular matrix proteins. J Biol Chem. 2004;279(13). https://doi.org/10.1074/jbc.M400242200.

10. Roughley PJ, Mott JS. The role of aggrecan in normal and osteoarthritic cartilage. J Exp Orthop. 2014;1(1):8. https://doi.org/10.1186/s40634-014-0008-7.

11. Dateki S. ACAN mutations as a cause of familial short stature. Clin Pediatr Endocrinol. 2017;26(3). https://doi.org/10.1297/cpe.26.1.19.

12. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. ACMG Laboratory Quality Assurance Committee. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17(5). https://doi.org/10.1038/gim.2015.30.

Publisher's Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.