Screening trematodes for novel intervention targets: a proteomic and immunological comparison of *Schistosoma haematobium*, *Schistosoma bovis* and *Echinostoma caproni*

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SUMMARY

With the current paucity of vaccine targets for parasitic diseases, particularly those in childhood, the aim of this study was to compare protein expression and immune cross-reactivity between the trematodes *Schistosoma haematobium*, *S. bovis* and *Echinostoma caproni* in the hope of identifying novel intervention targets. Native adult parasite proteins were separated by 2-dimensional gel electrophoresis and identified through electrospray ionisation tandem mass spectrometry to produce a reference gel. Proteins from differential gel electrophoresis analyses of the three parasite proteomes were compared and screened against sera from hamsters infected with *S. haematobium* and *E. caproni* following 2-dimensional Western blotting. Differential protein expression between the three species was observed with circa 5% of proteins from *S. haematobium* showing expression up-regulation compared to the other two species. There was 91% similarity between the proteomes of the two *Schistosoma* species and 81% and 78·6% similarity between *S. haematobium* and *S. bovis* versus *E. caproni*, respectively. Although there were some common cross-species antigens, species-specific targets were revealed which, despite evolutionary homology, could be due to phenotypic plasticity arising from different host-parasite relationships. Nevertheless, this approach helps to identify novel intervention targets which could be used as broad-spectrum candidates for future use in human and veterinary vaccines.

Key words: *Schistosoma*, *S. bovis*, *S. haematobium*, *Echinostoma caproni*, trematode, proteomics, immunology, DIGE, DIA, vaccine development.

INTRODUCTION

Schistosomes are important blood-fluke parasites of humans and domestic livestock (Rollinson et al. 1997). These trematodes are divided into 4 main groups: *Schistosoma mansoni* group, *S. haematobium* group, *S. indicum* group and *S. japonicum* group (Secor and Colley, 2005). Echinostomes are also trematodes but, unlike schistosomes, they develop and are restricted to the intestinal lumen of the definitive host and do not have a tissue invasive phase (Toledo and Fried, 2005; Toledo et al. 2009).

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Human schistosomiasis is a neglected tropical disease and a major public health concern in Africa, the Middle East, Asia and South America. Some 200 million people are infected with schistosomes, with a further 700 million at risk of infection in tropical and subtropical regions (Engels et al. 2002). As there is currently no available vaccine for this disease in people (Bergquist et al. 2008), the foundation of control is based upon provision of chemotherapy to afflicted communities, in particular mass drug administration of the anthelmintic praziquantel (Doenhoff et al. 2009). However, the search for an effective vaccine continues to be a key priority (Secor and Colley, 2005).

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Urinary schistosomiasis, caused by *Schistosoma haematobium*, is the most prevalent form of schistosomiasis in Africa and the Middle East. Children carry the heaviest burden of infection with as many as 100% of primary school children infected in areas such as our study sites in Zimbabwe (Midzi et al. 2008). Children younger than school-age can also be infected and begin to exhibit disease (Garba et al. 2010). As a result, schistosome-related morbidities include both non-immunological forms (blood in the urine, pain during urination, anaemia, growth retardation, poor cognition and memory) and immunemediated forms (tissue damage and organomegaly) (Midzi et al. 2008). Immuno-pathological reactions against schistosome eggs trapped in the tissues leads to inflammatory and obstructive disease in the bladder, ureter and kidney as well as fibrosis. Urinary schistosomiasis likely predisposes to bladder cancer and HIV infection (Stoever et al. 2009).

To develop vaccines protective against infection and/or pathology based on natural immune responses against schistosomiasis, there is ongoing research both in humans and animals in a context of experimental and natural schistosomiasis (Hagan et al. 1991; Dunne et al. 1992; Demeure et al. 1993; Grogan et al. 1997; Mutapi et al. 1998). Several studies have demonstrated similarities between different *Schistosoma* species in terms of life-histories and immunological aspects (Verjovski-Almeida et al. 2003; Capron et al. 2005; Berriman et al. 2009; Zhou et al. 2009), but little is known about molecular phenotypic differences that may be involved in host adaptation which might affect the efficacy of future vaccines. Despite the demonstration that antibody-mediated responses can protect against schistosome infection in experimental models, current human schistosome vaccine research, based on antibody-mediated protection, has stalled with the failure of many of the vaccine candidate antigens to enter Phase III clinical trials (Hagan and Sharaf, 2003). Limitations in our current understanding of the development of protective anti-schistosome responses against specific antigenic proteins, as well as the parasite’s biology (particularly antigen expression patterns) may be contributing to the slow development of effective anti-schistosome vaccines.

To shed light on these issues, comparison of the protein expression of *S. haematobium* adult worms with other trematode parasites could be illuminating especially in reference to *S. bovis* which is a ‘molecular’ analogue of *S. haematobium* and an experimental model for vaccine research (Capron et al. 2005). Comparison with other more distantly-related trematodes, e.g. *Echinostoma caproni*, is also useful by providing inferences into putative responses to different life history tracts, i.e. echinostomes do not have a tissue phase in the definitive host, and present an opportunity to investigate host-related adaptations in protein expression patterns. Although *S. bovis* and *S. haematobium* differ in their definitive hosts and in their niches within the host vasculature (Vercruysse and Gabriel, 2005), being sufficiently closely related in terms of evolutionary distance (Bowles et al. 1995; Webster et al. 2006), they have an ability to hybridise (Huyse et al. 2009). As *S. bovis* is much easier to keep in laboratory passage in rodents (Agnew et al. 1989) as well as eliciting similar cross-immunogenic profiles (Losada et al. 2005), makes study of *S. bovis* particularly informative. However, significant differences are known: for example, early studies of the *S. haematobium* vaccine candidate glutathione-S-transferase (28 kDa GST) showed inter-species variation in the coding regions of *S. haematobium* vs. *S. bovis* vs. *S. japonicum* 28 kDa GST. This variation gives rise to phenotypic differences associated with host immunity (Trottein et al. 1992).

To date, several studies using proteomic approaches have compared protein expression patterns between different helminth life stages (Curwen et al. 2004; Jolly et al. 2007; Wang et al. 2010), including parasites of different sexes and parasite development in different hosts (Toledo et al. 2004; Cheng et al. 2005). There have been no comparative proteomic studies on different trematode species which could lead to novel intervention targets with broader spectra and a better understanding of parasite-related host immune modulation (Harnett and Harnett, 2010). Previous evolutionary and ecological studies have been carried out using genetic techniques such as micro-array (transcriptome) or genome sequencing (Cieslak and Ribera, 2009) and these have given important insights into the biology of the parasites. These techniques do not take into account post-transcriptional regulation of protein expression (López, 2007; Schrimp and Hengartner, 2010) and cannot determine the degree of epitope cross-reactivity between parasite species. Moreover, the proteomic approach is particularly useful in non-model organisms (López, 2007; Ramm et al. 2009). Comparative proteomic approaches have been successfully used in other more general molecular studies: for example, assessing the divergence between different rodent species (Aquadro and Avise, 1981).

In this study, we have used a proteomic approach to compare phenotypic differences between the three different parasite species in terms of protein expression and immunogenicity. We compared protein expression patterns and immune cross-reactivity between *S. haematobium*, *S. bovis* and *E. caproni* which may indicate proteins involved in the adaptation to different hosts and different niches potentially warranting further scrutiny as potential vaccines targets for schistosomiasis as well as several other trematode diseases.
Parasites and experimental infections

The techniques used for the maintenance of *Echinostoma caproni* in the laboratory have been described in detail elsewhere (Toledo et al. 2004). Briefly, encysted metacercariae of *E. caproni* were removed from the kidneys and pericardial cavities of experimentally infected *Biomphalaria glabrata* snails and used to infect golden hamsters (Mesocricetus auratus). Outbred male golden hamsters, weighing 45–60 g, were infected through a stomach tube with 75 metacercariae each of *E. caproni*. The worm egg release by each animal was monitored daily as described previously (Toledo et al. 2003). Soluble adult worm antigens (SWAP) were prepared from adult worms collected from the intestine of hamsters 6 weeks post-infection with 100 metacercariae of *E. caproni* following previously published protocols (Toledo et al. 2003). For *S. haematobium* infections used for the serological studies, parasite eggs obtained from urine of *S. haematobium*-infected children in Zanzibar (Stothard et al. 2004) were hatched and used to infect *Bulinus* species snails with 3 miracidia per snail. Upon infection patency 150 cercariae were pooled from these shedding snails and used to infect golden hamsters by the paddling technique; all experiments were in accordance with ethical principles in animal research and Home Office (UK) approvals.

Adult *S. haematobium* SWAP was obtained freeze dried from the Theodor Bilharz Institute (Giza, Egypt). To prepare this fraction, worms were perfused in saline buffer from hamsters, washed in PBS (pH 7.4), homogenized, centrifuged to obtain the soluble fraction and freeze-dried in aliquots (5 mg/mL). These were reconstituted with distilled water as required. Freeze-dried adult *S. bovis* SWAP from sheep was prepared as previously described in detail elsewhere (Oleaga and Ramajo, 2004). SWAP preparations were prepared following similar protocols to reduce proteome variations due to different preparation approaches.

Rodent sera

For the immunological cross-reactivity assays, the antigen recognition patterns of sera from hamsters infected with *S. haematobium* and *E. caproni* were determined. For *E. caproni*, a pool was made from sera collected at 5, 6 and 10 weeks post-infection (hamsters normally make parasite-specific antibodies from 5 weeks) from 5 hamsters. After clotting overnight at 4°C, serum was separated from the clot by centrifugation. All the sera and the antigens were stored at −20°C until use. For sera from schistosome infected hamsters, Syrian golden hamsters were infected with 150 cercariae by paddling and bled 12 weeks post-infection. After clotting, blood collected from each hamster was centrifuged at 1400 g for 5 min to collect sera which were snap frozen in liquid nitrogen for long-term storage in liquid nitrogen. A pool of sera was made from 5 hamsters for use in this study. There were no experiments of hamsters infected with *S. bovis* parasites.

Preparations for CyDye labelling for DIGE

CyDye DIGE Fluor minimal dyes (GE Healthcare) were reconstituted following the manufacturer’s instructions. 50μg protein of each sample were labelled with either Cy3 or Cy5. The sample volumes were adjusted to 18μL with labelling buffer (7M urea, 2M thiourea, 4% CHAPS, (w/v), 25 mM Tris Base; pH 8.5), followed by addition of 1μL dye (400 pmol) to each individual sample. The samples were left on ice for 30 minutes in the dark, followed by adding 1μL of 10 mmol/L lysine to stop the reaction.

Two-dimensional differential in gel electrophoresis (2D-DIGE)

To compare the parasite proteomes in 2D-DIGE assays, three gels were run—one for each pair of samples. Differentially labelled samples were mixed into the same tube with 210μl of rehydration buffer (7M urea, 2M thiourea, 4% CHAPS, 5% DTE (dithioerythritol), 0.8% IPG buffer 3–10 pH and bromophenol blue). Thereafter, the first dimension i.e. isoelectric focusing (IEF) and second dimension were run following previously described protocols (Mutapi et al. 2005) using the IEF protocol for 13 cm IPG strips; rehydration for 14 h at 20 V, 500 V for 1 h, 1000 V for 1 h and 8000 V for 3 h and performing the second dimension using 12% polyacrylamide gels with SDS buffer. Images from these gels were subsequently analysed as described below.

Image analysis and mass spectrometry

Gels were scanned on a Typhoon spectrophotometer (GE Healthcare) at the appropriate excitation/emission wavelength for each fluorophore Cy3 (532/580 nm) and Cy5 (633/670 nm) at 50 microns resolution. The images were analyzed using the Difference In-gel Analysis (DIA) module of Decyder software version 7.0 (GE Healthcare). The protein spots showing greater than 5-fold differences in relative abundance between parasite preparations were considered as differentially expressed proteins. The 5-fold difference was used to reduce the likelihood of detecting spurious differences. Proteins from the different trematodes were identified by comparing DIGE images with the proteomic map of *S. haematobium* (Mutapi et al. 2005) and *E. caproni* (Sotillo et al. 2010) since there is no complete genome
Fig. 1. 2D-DIGE images comparing pairs of different helminth species’ adult worm proteomes.

A. *S. haematobium* (green spots) vs. *S. bovis* (red spots).
B. *S. haematobium* (red spots) vs. *E. caproni* (green spots).
or protein sequence available for any of the three species studied. *S. haematobium* protein identities on the proteome map were obtained from a Coomassie Blue-stained reference gel which had been prepared and processed to obtain MS/MDS data which were submitted for an MS/MS ion search via the Mascot search engine (Matrix Science), and non-redundant National Center for Biotechnology Information (NCBI) database (Mutapi et al. 2005). Briefly, plugs of 1·4 mm were excised from the reference Coomassie Blue-stained gel and subjected to in-gel trypsin digestion in an Ettan Spot Handing Workstation (GE Healthcare), in accordance with standard protocols (Amersham).

The resulting tryptic peptides were solubilized in 0·5% formic acid and were fractionated by nanoflow high-performance liquid chromatography on a C18 reverse phase column (GE Healthcare), and elution was performed with a continuous linear gradient of 40% acetonitrile for 20 min. The eluants were analyzed by online electrospray tandem MS (MS/MS) by use of a Qstar Pulsar mass spectrometer (Applied Biosystems). A 3 sec survey scan preceded each MS/MS data-collection cycle of 4 product ion scans of 3 sec each, and this gave a duty cycle of 15 sec. Data were submitted for an MS/MS ion search via the Mascot search engine (Matrix Science), and both locally established databases for *S. mansoni* EST sequences and the present non-redundant National Center for Biotechnology Information (NCBI) database were searched.

**Two-dimensional electrophoresis and Western blotting**

In order to determine cross-reactive antigens, 2D gel electrophoresis (2DE) was conducted on 7 cm gels as above, with some modifications. 100 μg each protein were solubilised in rehydration buffer (7 M urea, 2 M thiourea, 4% CHAPS (w/v), 65 mM DTE and trace bromophenol blue) and 0·8% IPG buffer (pH 3–10) to make a total volume of 125 μL. Each protein preparation was then added to a 7 cm linear pH 3–10 IPG strip and the IEF was performed following the following protocol (1) passive rehydration for 14 h; (2) 500 V for 30 min; (3) 1000 V for 30 min; (4) 8000 V for 4 h followed by equilibration in 2 mL of 1% DTE for 15 min and 2 mL of 4% iodoacetamide in equilibration buffer containing 6 M urea, 0·375 M Tris pH 8·8, 2% SDS and 20% glycerol. The second dimension was performed using 10% polyacrylamide precast gels from Invitrogen. Proteins from SDS-PAGE were stained with Coomassie blue or transferred onto nitrocellulose membranes in 25 ml 20X transfer buffer (Invitrogen), methanol 10% (v/v).

After confirming transfer by staining with 0·1% Ponceau S (Sigma), membranes were blocked with TBS Start Block buffer T20 (Invitrogen) for 1 h at room temperature. After washing with TBS containing 0·05% Tween-20 (TBST), blots were incubated overnight at 4°C with a pool of 10 serum samples of *E. caproni*-infected hamsters, or *S. haematobium*-infected hamsters or negative control sera at 1:200 dilution in TBS Start Block buffer. The membrane was then washed three times for 10 min each time in TBS, 0·05% Tween 20, 0·5% Triton-X100 (TBS/TT). Bound antibodies were detected by incubating blots for 1 h at RT with horseradish peroxidase (HRP)-conjugated rabbit anti-Syrian hamster IgG (Abcam), in blocking buffer. After washing four times for 10 min each time in TBS/TT and once in TBS alone, recognised antigens were visualized using ECL Plus (Amersham) following the manufacturer’s instructions, and exposed to X-OMAT film (Kodak) for 10 sec. Images from Western blotting and Coomassie blue staining were digitalised and matched by using ImageMaster software (GE Healthcare).

**RESULTS**

**Proteome comparisons**

The 2D-DIGE gels were run comparing the 3 proteomes as shown in Fig. 1. DIA analysis of the gels showed both quantitative and qualitative differences. There was more similarity between the two schistosome species than between *Echinostoma* and *Schistosoma*. On the first gel comparing *S. haematobium* and *S. bovis*, 1701 spots representing different proteins (including different isoforms) were detected, with 91% showing similar expression levels (Fig. 2A). 5·4% of the proteins showed increased expression in *S. haematobium* by our criteria of 5-fold or greater difference in abundance on the gel while 3·6% showed increased expression in *S. bovis*. On the second gel, comparing *S. haematobium* vs. *E. caproni*, 1967 spots were detected with 81% showing similar expression levels. 8·4% of the protein spots showed increased expression in *S. haematobium* and 10·6% showed increased expression in *E. caproni* while 81% were present in similar amounts on both gels (Fig. 2B).

On the final gel, comparing *S. bovis* vs. *E. caproni*, 1757 spots were detected with 78·6% showing similar expression levels. 9·1% of the protein spots showed increased expression in *S. bovis* and 12·3% showed increased expression in *E. caproni* (Fig. 2C).

It was possible to identify some of the proteins present on the adult worm proteomes by comparing DIGE images with the proteome maps of

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C. *S. bovis* (red spots) vs. *E. caproni* (green spots). Identified proteins are indicated by solid arrows (for *S. haematobium*), dashed arrows (for *E. caproni*) and encircled (for both).
Fig. 2. Pair-wise comparison of protein expression patterns in adult worm proteomes of pairs of parasite species from DIA plug-in analysis. This analysis co-detects the spots from the image and, after normalization, compares the volume
S. haematobium and E. caproni. Details of S. haematobium protein identities from mass spectrometry of proteins have already been published (Mutapi et al. 2005) while those from E. caproni have not previously been published and are given in Table 1 and annotated in Fig. 3.

The expression patterns of pairs of proteins spots between different parasite species are compared in Table 2. The heat shock protein HSP70 was more highly expressed in S. haematobium compared to S. bovis or E. caproni. Between the Schistosoma species, there were similar expression levels of metabolic enzymes, signal transduction molecules and detoxification enzymes, but expression levels of these proteins differed between the latter species and E. caproni. Three homologous proteins were identified in S. haematobium and E. caproni; protein disulfide isomerase, enolase and GST, but the gel migration showed that they differed in molecular weights between the two species.

Immune cross-reactivity

Using sera from hamsters infected with S. haematobium and E. caproni, we performed 2D Western-blot analyses. As expected, homologous pairs of sera and antigen showed the highest levels of recognition (Fig. 4). Interestingly, heterologous sera also detected spots in the gels, confirming cross-reactivity among these trematode species. In this context, S. haematobium sera recognised more antigenic spots in the S. bovis proteome than in E. caproni. There was some cross-reactivity, between E. caproni and S. haematobium. Three spots in the E. caproni proteome which were identified as isoforms of GADPH reacted with sera from both E. caproni-infected and S. haematobium-infected hamsters. However, sera from E. caproni-infected hamsters did not react against GADPH in the S. haematobium proteome.

DISCUSSION

Trematodes are an evolutionarily distinct group of parasites of importance to both human and veterinary medicine in the diseases that they cause. Understanding similarities and differences in their phenotypic molecular biology is important in several areas such as drug target discovery, vaccine design and development of helminth-derived therapeutic agents for immune disorders; also in the context of when infections are acquired during childhood and beyond. Here, a comparative analysis of the proteome of three species of trematode: S. haematobium,

Table 1. Proteins identified in the adult soluble worm preparation of Echinostoma caproni using MASCOT search engine (Matrix Science)

| Sample | NCBI accession # | Protein (Species) | MW  | Hit_score |
|--------|------------------|------------------|-----|-----------|
| 1      | gi|3891573         | Chain A, Fasciola hepatica Glutathione S-transferase isoform 1 in complex with glutathione | 25217 | 77       |
| 2      | gi|3891573         | Chain A, Fasciola hepatica Glutathione S-transferase isoform 1 in complex with glutathione | 25217 | 83       |
| 3      | gi|16406594        | Glyceraldehyde phosphate dehydrogenase (Fasciola hepatica) | 23687 | 247      |
| 4      | gi|16406594        | Glyceraldehyde phosphate dehydrogenase (Fasciola hepatica) | 23687 | 201      |
| 5      | gi|226475754       | Aldolase (Schistosoma japonicum) | 39800 | 134      |
| 6      | gi|29841453        | Similar to GenBank Accession Number AF026805 fructose bisphosphate Aldolase (S. mansoni and S. japonicum) | 32063 | 162      |
| 7      | gi|268535422       | Hypothetical protein CBG15039 (Caenorhabditis briggsae) | 20029 | 58       |
| 8      | gi|112950027       | Enolase (Echinostoma caproni) | 46568 | 588      |
| 9      | gi|226489288       | Tubulin beta-2C chain (Schistosoma japonicum) | 50056 | 666      |
| 10     | gi|3392892         | Protein disulphide isomerase (Fasciola hepatica) | 55587 | 69       |

of a spot from the two samples as a volume ratio. Spots were detected as different if the volume ratio threshold difference was ≥ 5 fold. Each gel image shows different expression patterns on individual species gels.

A. Gels showing the DIA analyses of S. haematobium vs. S. bovis on the gels from each of the two species. Green marks represent proteins over-expressed in S. haematobium. Red marks represent proteins over-expressed in S. bovis. Blue represents spots expressed to similar levels in the two species.

B. Gels showing the DIA analyses of S. haematobium vs. E. caproni on the gels from each of the two species. Green marks represent proteins over-expressed in S. haematobium. Red marks represent proteins over-expressed in E. caproni. Blue represents spots expressed to similar levels in the two species.

C. Gels showing the DIA analyses of S. bovis vs. E. caproni on the gels from each of the two species. Green marks represent proteins over-expressed in E. caproni. Red marks represent proteins over-expressed in S. bovis. Blue represents spots expressed to similar levels in the two species. The histograms represents spot frequencies plotted against logarithm volume ratio.
S. haematobium and S. bovis are closely related species and can undergo hybridisation (Huyse et al. 2009), the results obtained here show that there are also significant proteomic differences, even among conserved proteins. These species-specific differences could be due to phenotypic plasticity arising from different host-parasite relationships (Schrimpf and Hengartner, 2010). Evolutionary and phylogenetic studies have demonstrated that highly expressed genes tend to evolve more slowly (Hirsh and Fraser, 2001; Schrimpf and Hengartner, 2010), nonetheless here we demonstrate that some of these conserved proteins differ in molecular weight, most likely due to post-translational modifications which should be explored further particularly as vaccine candidates.

It was possible to identify some of the proteins present in the proteomes by searching public databases. Table 2 shows the proteins differentially expressed between the different trematodes identified on the 2-dimensional gel in Differential in Gel Electrophoresis (DIGE).

### Table 2. Proteins differentially expressed between the different trematodes identified on the 2-dimensional gel in Differential in Gel Electrophoresis (DIGE)

#### S. haematobium vs. S. bovis

| INCREASED in *S. haematobium* | NCBI accession # | MW   |
|-------------------------------|------------------|------|
| Heat shock protein /chaperone | gi|10168 | 68331 |
| 70,000 mol wt antigen/hsp70   | gi|312018 | 44508 |
| Metabolic enzyme              | gi|312018 | 44508 |
| Triosephosphate isomerase     | gi|605647 | 39963 |
| Fructose 1,6 bisphosphate aldolase | gi|312018 | 54463 |
| Protein disulfide isomerase   | gi|556413 | 44508 |
| Phosphoglycerate kinase       | gi|3023710| 47421 |
| Heat shock protein /chaperone | gi|21634531| 58740 |
| Heat shock protein HSP60      | gi|21634531| 58740 |
| Signal transduction           | gi|6649234 | 28754 |
| Detoxification protein        | gi|161013 | 2407 |
| 28 kDa glutathione-S transferase | gi|161013 | 2407 |

#### SIMILAR

| NCBI accession # | MW   |
|------------------|------|
| gi|1351281 | 28447 |
| gi|605647 | 39963 |
| gi|10168  | 68331 |
| gi|312018 | 44508 |
| gi|3023710| 47421 |
| gi|21634531| 58740 |

#### E. caproni vs. S. bovis

| INCREASED in *E. caproni* | NCBI accession # | MW   |
|----------------------------|------------------|------|
| Metabolic enzyme           | gi|3392892 | 55587 |
| Protein disulfide isomerase | gi|112950027| 46568 |
| Enolase                    | gi|16406594| 23687 |
| Glutathione S-Transferase  | gi|3891573 | 25217 |

### Table 2. Proteins differentially expressed between the different trematodes identified on the 2-dimensional gel in Differential in Gel Electrophoresis (DIGE)

#### Similar

| NCBI accession # | MW   |
|------------------|------|
| gi|226475754 | 39800 |

#### S. haematobium vs. E. caproni

| INCREASED in *S. haematobium* | NCBI accession # | MW   |
|-------------------------------|------------------|------|
| Heat shock protein /chaperone | gi|10168 | 68331 |
| 70,000 mol wt antigen/hsp70   | gi|312018 | 44508 |
| Metabolic enzyme              | gi|312018 | 44508 |
| Triosephosphate isomerase     | gi|605647 | 39963 |
| Fructose 1,6 bisphosphate aldolase | gi|312018 | 54463 |
| Protein disulfide isomerase   | gi|556413 | 44508 |
| Phosphoglycerate kinase       | gi|3023710| 47421 |
| Heat shock protein /chaperone | gi|21634531| 58740 |
| Heat shock protein HSP60      | gi|21634531| 58740 |
| Signal transduction           | gi|6649234 | 28754 |
| Detoxification protein        | gi|161013 | 2407 |
| 28 kDa glutathione-S transferase | gi|161013 | 2407 |

#### Similar

| NCBI accession # | MW   |
|------------------|------|
| gi|3392892 | 55587 |
| gi|112950027| 46568 |
| gi|16406594| 23687 |
| gi|3891573 | 25217 |

#### S. bovis and E. caproni using 2D-DIGE was conducted. *S. haematobium* and *S. bovis* are closely related species and can undergo hybridisation (Huyse et al. 2009), the results obtained here show that there are also significant proteomic differences, even among conserved proteins. These species-specific differences could be due to phenotypic plasticity arising from different host-parasite relationships (Schrimpf and Hengartner, 2010). It was possible to identify some of the proteins present in the proteomes by searching public databases.
databases, but due to the limited sequence information available on these three trematodes, a large number of the proteins remain unidentified (Nowak and Loker, 2005). Comparing the proteomes of the two schistosome species, only HSP70 identified from the Coomassie Blue-stained reference gel showed increased expression in *S. haematobium* despite the 10% difference in protein expression levels detected by the DIA analysis. The DIGE analysis can detect much lower concentrations of proteins than Coomassie staining. Thus, a large number of proteins present on the DIGE gel were present at a concentration too low to be detected from the Coomassie Blue-stained gel for mass spectrometry. These proteins accounted for some of the 10% differences between the two schistosomes. Our results showing differential expression of HSP70 are consistent with those from a different trematode genus, *Fasciola* where *F. hepatica* and *F. gigantica* show different levels of HSP70 expression (Smith et al. 2008). Furthermore, we have previously reported differences in HSP70 expression in *E. caproni* parasites from low vs. high compatible hosts (Higón et al. 2008). Therefore, expression levels of HSP70 seem to depend on the host environment and this could be a common mechanism used by different parasites in order to adapt to different hosts. The sequencing of the genome and subsequent identification of all proteins present in the proteome of all 3 species compared in this study will greatly strengthen such comparative approaches as they will allow more robust comparison of identified proteins as well as comparisons of the number of isoforms and the relative abundance of each isoform to the compared.

There were more differentially expressed proteins between the two different genera. Most proteins identified in both *E. caproni* and *S. haematobium* (with known identities) are homologues (protein disulfide isomerase, enolase and GST). However, these homologues have different molecular weights. It is likely that this difference is due to post-translational modifications rather than changes in the gene sequence, since these proteins are highly conserved (Ramajo-Hernández et al. 2007a,b; Sotillo et al. 2008). Furthermore, these proteins are important for the host-parasite relationship (E/S products, immunogenic properties), so these modifications could be involved in the host-parasite surface interaction. Protein disulfide isomerase (PDI) catalyses the formation (oxidation), breakage (reduction) and rearrangement (isomerisation) of disulfide bonds within proteins, thereby permitting their proper folding in the endoplasmic reticulum and transit through the secretory pathway (Ellgaard and Ruddock, 2005). PDI has been identified in the E/S products of adult *E. caproni*, *E. friedi* and *F. hepatica* worms, suggesting that it may be important in host-parasite interactions (Salazar-Calderon et al. 2003; Bernal et al. 2006; Sotillo et al. 2010). Moreover, PDI is immunogenic in human *S. haematobium* infections (Mutapi et al. 2005) and experimental *F. hepatica* (Moxon et al. 2010) and it has been shown to be immunologically protective against the hookworm, *Ancylostoma* (Epe et al. 2007). Differences in PDI

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**Fig. 3.** Coomassie blue-stained 2-dimensional *E. caproni* gel showing spots excised, and identified by MASCOT. Corresponding protein identities are given in Table 1.
Fig. 4. Antigen recognition profile of sera from trematode-infected hamsters.

A. *S. bovis* SWAP antigen recognition by sera from *E. caproni*-infected hamsters.
B. *S. bovis* SWAP antigen recognition by sera from *S. haematobium*-infected hamsters.
molecular weight between *S. haematobium* and *E. caproni* could be due to post-translational modifications, akin to the PDI glycosylation reported in *Trypanosoma brucei* where it is related to parasite defence (Rubotham et al. 2005).

The main function of glutathione S-transferase (GST) is detoxification of oxygen and endogenous free radicals (Torres-Rivera and Landa, 2008). It is present in *Echinostoma* spp. and *S. bovis* tegument and E/S products (Bernal et al. 2006; Perez-Sanchez et al. 2006; Sotillo et al. 2010). It is also the leading schistosome vaccine candidate (Capron et al. 2005; McManus and Loukas, 2008). There is a difference in the theoretical and observed molecular weights for GST. Ramajo-Hernandez et al. (2007a) reported no glycosylation of GST in *S. bovis*. Enolase is a multifunctional glycolytic enzyme (Pancholi, 2001), also present in E/S products (Bernal et al. 2006; Perez-Sanchez et al. 2006; Sotillo et al. 2010). In *S. bovis* as well as *E. caproni*, enolase has been identified as a human plasminogen-binding protein; this protein may be involved in preventing blood clotting during feeding in *Schistosoma* (Ramajo-Hernández et al. 2007b) or in mucosal erosion in *Echinostoma* (Marcilla et al. 2007).

To investigate some of the biological differences arising from differences in the proteomes, the immunogenicity of the adult worm antigens was compared. There was cross-reactivity between the three trematode species, but the intensity and antigen pattern recognition patterns differed. The most immune cross-reactivity occurred between the two schistosomes which is consistent with the DIGE results. We have identified a novel antigen for *E. caproni*, 3 isoforms of GADPH. *E. caproni* GAPDH was also recognized by sera from *S. haematobium*-infected hamsters. Interestingly, *S. haematobium* GADPH antigen was not recognized by sera from *E. caproni*-infected hamsters. GAPDH’s immunogenicity has been reported from other studies and is one of the World Health Organisation’s human schistosome vaccine candidates (Bergquist et al. 2002; El Ridi et al. 2010). Nevertheless it has not previously been reported as an antigen in *Echinostoma* spp. Toledo et al. (2004) discovered an immunogen of 37 kDa, 6 weeks post *Echinostoma* infections in rats, but the intensity of this response declined during the infection, suggesting that the protein could be released in the juvenile stages of the parasites. This immunogen is likely to be GAPDH and this present study and that of Toledo et al. (2004) suggest that the kinetics of antigen release and antibody production against GAPDH require further investigation, especially in the future context of screening against human sera from infected people.

Schistosomiasis continues to be a major public health problem in several tropical and sub-tropical countries. There are now several studies (e.g. Garba et al. 2010), showing that children as young as 1 year old are infected and can harbour levels of infection comparable to those in the adults in their communities and the search for an effective vaccine continues to be a key priority (Secor and Colley, 2005). One promising approach being pursued is to treat people, children in particular, repeatedly with praziquantel to induce immune-mediated resistance to re-infection (Black et al. 2010a). However, studies using this protocol indicate that the number of PZQ treatments required to reduce re-infection is significantly variable and can take several rounds of PZQ treatment (Black et al. 2010b). This suggests that an integrated approach using treatment and a recombinant vaccine as proposed by the World Health Organisation (Bergquist, 2004) might lend predictability and consistency as well as improved efficacy to future schistosome control programmes. Thus the molecular phenotypic differences shown in this study, particularly those which appear to be post-translational, may influence the development and production of recombinant vaccines (e.g. bacterial expression systems may not process the proteins appropriately after translation) and affect the efficacy of future vaccines.

Overall this study has demonstrated that, despite several biological and phylogenetic similarities between the three trematode species *S. haematobium*, *S. bovis* and *E. caproni*, there are quantitative and qualitative differences in protein expression patterns in their adult worm proteomes. The differences could be due to phenotypic plasticity arising from different host-parasite relationships. Some of these differences translate to differences in immunogenicity. Further studies characterizing the differentially expressed proteins will be important in determining the identity of proteins involved in host-parasite adaptation and the nature of the interaction between the host and parasite. This is particularly important for identifying vaccine candidates and predicting the effects vaccination, especially in childhood, would have on the parasite population structure.

C. *S. haematobium* SWAP antigen recognition by sera *E. caproni* from infected hamsters.
D. *S. haematobium* SWAP antigen recognition by sera from *S. haematobium*-infected hamsters.
E. *E. caproni* SWAP antigen recognition by sera from *E. caproni*-infected hamsters.
F. *E. caproni* SWAP antigen recognition by sera from *S. haematobium*-infected hamsters.
A novel *E. caproni* antigen GAPDH is encircled.
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REFERENCES

AGnew, A. M., Murave, H. M., Lucas, S. B. and Doenhoff, M. J. (1989). Schistosoma bovis as an immunological analogue of S. haematobium. Parasite Immunology 11, 329–340.

Aquardo, C. F. and Avise, J. C. (1981). Genetic divergence between rodent species assessed by using two-dimensional electrophoresis. Proceedings of the National Academy of Sciences, USA 78, 3784–3788.

Bergquist, R. (2004). http://www.who.int/dtrld/publications/dtrnews/news71/schisto.htm. Accessed 26 January 2011.

Bergquist, R., Al-Sherbini, M., Barakat, R. and Olds, R. (2002). Blueprint for schistosomiasis vaccine development. Acta Tropica, 82, 183–187.

Bergquist, R., Utzinger, J. and McManus, D. P. (2008). Trick or Treat: the role of integrative studies in the control of human schistosomiasis. PLoS Neglected Tropical Diseases 2, e244. doi:10.1371/journal.pntd.0000244.

Bernal, D., Carpena, I., Espert, A. M., De la Rubia, J. E., Esteban, J. G., Toledo, R. and Marcella, A. (2006). Identification of proteins in excretory/secretory extracts of Echinostoma friedii (Trematoda) from chronic and acute infections. Proteomics 6, 2835–2843.

Berriman, M., Haas, B. J., LoVerde, P. T., Wilson, R. A., Dillen, G. P., Graser, G., Capron, A., Riveau, G., Capron, M. and Trottein, F. (2008). A molecular phylogeny of Schistosoma species assessed by using two-dimensional electrophoresis. International Journal for Parasitology 38, 391–399.

Black, C. L., Muok, E. M., Mwinzi, P. N., Karanja, D. M., Carter, J. M., Kariuki, C. R., Coghlan, A., Coulson, R., Day, T. A., Delcher, A., D’Souza, R. F., Djibos, A., El Ridi, R., Tallima, H., Mahana, N. and Dalton, J. P. (2010). In vitro immunogenicity and in vivo protective potential of Schistosoma mansoni lung schistosomula excretory-secretary candidate vaccine antigens. Microbes and Infection 12, 780–799.

Engels, D., Chitsulo, L., Montresor, A. and Savioli, L. (2002). The global epidemiological situation of schistosomiasis and new approaches to control and research. Acta Tropica 82, 139–146.

Epe, C., Behrens, C., Strube, C. and Schneider, T. (2007). Evaluation of the translation level of the protein diagnostic isomerase in different stages from Ancylostoma caninum with a real-time PCR assay. Parasitology Research 101, 1589–1595.

Garba, A., Barkier, N., Djibo, A., Lamine, M. S., Sofo, B., Galvani, A. P., Bosque-Oliva, E., Nosten, F., Utzinger, J. and Fenwick, A. (2010). Schistosomiasis in infants and preschool-aged children: infection in a single Schistosoma haematobium and a mixed S. haematobium-S. mansoni foci of Niger. Acta Tropica 115, 212–219.

Grogan, J., Kremers, P. J., Van Dam, G. J., Deelder, A. M. and Vazquez-Anadon, A. M. (1997). Anti-schistosome IgG4 and IgE at 2 years after chemotherapy: infected versus uninfected individuals. Journal of Infectious Diseases 176, 1344–1350.

Hagan, P., Blumenthal, U. J., Dunne, D., Simpson, A. J. D. and Wilkins, A. H. (1991). Human IgE, IgG4 and resistance to reinfection with Schistosoma haematobium. Nature 349, 234–245.

Hagenaar, P. and Sharaf, O. (2003). Schistosomiasis vaccines. Expert Opinion in Biological Therapy, 3, 1271–1278.

Harnett, W. and Harnett, M. M. (2010). Helminth-derived immunomodulators: can understanding the worm produce the pill? Nature Reviews Immunology 10, 278–284.

Higón, M., Monteagudo, C., Fried, B., Esteban, J. G., Toledo, R. and Marcella, A. (2008). Molecular cloning and characterization of Echinostoma caproni caproni heat shock protein-70 and differential expression in the parasite derived from low- and high-compatible hosts. Parasitology 135, 1469–1477.

Hirsh, A. E. and Fraser, H. B. (2001). Protein dispensability and rate of evolution. Nature 411, 1046–1049.

Huyse, T., Webster, B. L., Geldof, S., Stothard, J. R., Diaw, O.T., Polman, K. and Rollinson, D. (2009). Bidirectional introgressive hybridization between a cattle and human schistosome species. PLoS Pathogens 5, e1000571. doi: 10.1371/journal.ppat.1000571.

Jolly, E. R., Chin, C. S., Miller, S., Bahgat, M. M., Lim, K. C., DeRisi, J. and McKerrow, J. H. (2007). Gene expression patterns during adaptation of a helminth parasite to different environmental niches. Genome Biology 8, R65.

López, J. L. (2007). Two-dimensional electrophoresis in proteome expression analysis. Journal of Chromatography: B, 849, 190–202.

Losada, S., Chacón, N., Colmenares, C., Bermúdez, H., Lorenzo, A., Pointer, J. E., Theron, A., Alarcón de Noya, B. and Noya, O. (2005). Schistosoma Cross-reactivity and antigenic community among different species. Experimental Parasitology 111, 182–190.

Marcella, A., Perez-Garcia, A., Espert, A., Bernal, D., Muñoz-Antoli, C., Esteban, J. G. and Toledo, R. (2007). Schistosoma caproni caproni: identification of enolase in excretory/secretory products, molecular cloning, and functional expression. Experimental Parasitology 117, 56–74.

McManus, D. P. and Lokouas, A. (2008). Current status of vaccines for schistosomiasis. Clinical Microbiology Reviews 21, 225–242.

Midi, N., Sangweme, D., Zinowera, S., Mapingure, M., Bewoorer, K. C., Munatsi, A., Mutapi, F., Mudzori, J., Kumar, N., Woelk, G. and Mdlulza, T. (2008). The burden of polyparasitism among primary schoolchildren in rural and farming areas in Zimbabwe. Transactions of the Royal Society of Tropical Medicine and Hygiene 102, 1059–1064.

Moxon, J. V., Flynn, A. R., Golden, A. O., Hamilton, J. V., Mulcahy, G. and Brophy, M. P. (2010). Immune responses directed at egg proteins during experimental infection with the liver fluke Fasciola hepatica. Parasite Immunology 32, 111–124.

Mutapi, F., Burchmore, R., Mdlulza, T., Foucher, A., Harcus, Y., Nicoll, G., Mutapi, F., Turner, C. M. and Maizels, R. M. (2005). Praziquantel treatment of individuals exposed to Schistosoma haematobium
enhances serological recognition of defined parasite antigens. *Journal of Infectious Diseases* 192, 1108–1118.

Mutapi, F., Ndlovu, P. D., Hagan, P., Spicer, J. T., Mduluza, T., Turner, C. M., Chandiwana, S. K. and Woolhouse, M. E. J. (1998). Characterization accelerates the development of acquired immune responses to *Schistosoma haematobium* infection. *Journal of Infectious Diseases* 178, 289–293.

Nowak, T. S. and Loker, E. S. (2005). *Echinostoma paraense*: differential gene transcription in the sporocyst stage. Experimental *Parasitology* 109, 91–95.

Oleaga, A. and Ramajo, V. (2004). Efficiency of the oral, intramuscular and subcutaneous routes for the experimental infection of hamster and sheep with *Schistosoma bovis*. *Veterinary Parasitology* 124, 53–61.

Pancholi, V. (2001). Multifunctional alpha-ence: its role in diseases. *Cellular and Molecular Life Science*. CMLS 58, 902–920.

Pérez-Sánchez, R., Ramajo-Hernández, A., Ramajo-Martín, V. and Oleaga, A. (2006). Proteomic analysis of the tegument and excretory-secretory products of adult *Schistosoma bovis* worms. *Proteomics* 6, S226–S230.

Ramajo-Hernández, A., Oleaga, A., Ramajo-Martín, V. and Pérez-Sánchez, R. (2007a). Carbohydrate profiling and protein identification of tegumental and excreted/secreted glycoproteins of adult *Schistosoma bovis* worms. *Veterinary Parasitology* 144, 45–60.

Ramajo-Hernández, A., Pérez-Sánchez, R., Ramajo-Martín, V. and Oleaga, A. (2007b). *Schistosoma bovis*: Plasminogen binding in adults and the identification of plasminogen-binding proteins from the worm tegument. *Experimental Parasitology* 115, 83–91.

Ramm, S. A., McDonald, L., Hurst, J. L., Beynon, R. J. and Sommer, F. (2000). Comparative proteomic study of rodent seminal de isomerases. *Journal of Molecular and Biochemical Parasitology* 115, 203–226.

Salazar-Calderón, M., Martín-Alonso, M. J., Castro, A. M. and *Journal of Biological Chemistry* 280, 26201–26208.

Sato, D. (2008). Carbohydrate profiling and protein identification of tegumental and excreted/secreted glycoproteins of adult *Schistosoma bovis* worms. *Veterinary Parasitology* 144, 45–60.

Toida, R., Esteban, J. G. and Fried, B. (2003). Recent advances in the biology of *echinostomes*. *Parasite Immunology* 25, 147–294.

Toida, R. and Fried, B. (2005). *Echinostomes* as experimental models for interactions between adult parasites and vertebrate hosts. *Trends in Parasitology* 21, 251–254.

Torres-Rivera, A. and Landa, A. (2008). Cooperative kinetics of the recombinant glutathione transferase of *Taenia solium* and characterization of the enzyme. *Archives of Biochemistry and Biophysics* 477, 372–378.

Trottein, F., Godin, C., Pierce, R. J., Sellin, B., Taylor, M. G., Gorillot, I., Silva, M. S., Lecoq, J. P. and Capron, A. (1992). Inter-species variation of *schistosome* 28-kDa glutathione S-transferases. *Molecular and Biochemical Parasitology* 52, 115–121.

Vercruysse, J. and Gabriel, S. (2005). Immunity to schistosomiasis in animals: an update. *Parasite Immunology* 27, 289–295.

Verjovski-Almeida, S., DeMarco, R., Martins, E. A., Guimaraes, P. E., Ojopi, E. P., Paquola, A. C., Piazza, J. P., Nishiyama, M. Y. Jr, Krajina, J. P., Adamson, R. E., Ashton, P. D., Bonaldo, M. F., Couplon, P. S., Dillon, G. P., Farias, L. P., Gregorio, S. P., Ho, H. L., Leite, R. A., Malaquias, L. C., Marques, R. C., Miyasato, P. A., Nascimento, A. L., Ohlweller, F. P., Reis, E. M., Ribeiro, M. A., Sa, R. G., Stukart, G. C., Soares, M. B., Gardonio, C., Kawano, T., Rodrigues, V., Madeira, A. M., Wilson, R. A., Menck, C. F., Setubal, J. C., Leite, L. D. and Dias-Neto, E. (2003). Transcriptome analysis of the acelomate human parasite *Schistosoma mansoni*. *Nature Genetics* 35, 148–157.

Wang, Z., Abubucker, S., Martin, J., Wilson, R. B., Hawdon, H. J. and Mitreva, M. (2010). Characterizing *Ancylostoma caninum* transcriptome and exploring nematode parasitic adaptation. *BMC Genomics* 11, 307. doi: 10.1186/1471-2164-11-307

Webster, B. L., Southgate, V. R. and Littlewood, D. T. (2006). A revision of the interrelationships of *Schistosoma* including the recently described *Schistosoma guineensis*. *International Journal for Parasitology* 36, 947–955.

Zhou, Z., Zheng, H., Chen, Y., Zhang, L., Wang, K., Guo, J., Huang, Z., Zhang, B., Huang, W., Jin, K., Dou, T., Hasegawa, M., Wang, L., Zhang, Y., Zhou, J., Yao, Z., Li, Y., Yin, H., Bao, L., Chen, Y., Yu, M., Wu, H., Wang, Z., Zhang, Q. H., Song, H. D., Chen, S., Xu, X., Xu, B., Ju, C., Huang, Y., Brindley, P. J., McManus, D. P., Feng, Z., Han, Z. G., Lu, G., Ren, S., Wang, Y., Wu, K., Kang, H., Chen, J., Chen, X., Chen, S., Wang, L., Yan, J., Wang, B., Lu, P., Zhang, X., Zhang, W., Hu, Q., Zhu, G., Wang, J., Yu, J., Wang, J., Yang, H., Ning, Z., Beriman, M., Wei, C. L., Ruan, Y., Zhao, G., Wang, S., Liu, F., Zhou, Y., Wang, Z., Liu, G., Zheng, H., Brindley, P. J., McManus, D. P., Blair, D., Zhang, Q. H., Zhong, Y., Wang, S., Han, Z. G., Chen, Z., Wang, S., Han, Z. G. and Chen, Z. (2009). The *Schistosoma japonicum* genome reveals features of host-parasite interplay. *Nature* 460, 345–351.