Aquaporin-1 and aquaporin-9 gene variations in sudden infant death syndrome

Siri Hauge Opdal 1 · Linda Ferrante 1 · Torleiv Ole Rognum 2 · Arne Stray-Pedersen 1,2

Received: 22 June 2020 / Accepted: 17 December 2020 / Published online: 18 January 2021
© The Author(s) 2021

Abstract
Several studies have indicated that a vulnerability in the development and regulation of brain function is involved in sudden infant death syndrome (SIDS). The aim of this study was to investigate the genes encoding the brain aquaporins (AQPs) AQP1 and AQP9 in SIDS. The hypothesis was that specific variants of these genes are part of the genetic vulnerability predisposing infants to sudden unexpected death. The study included 168 SIDS cases with a median age of 15.5 (range 2–52) weeks and 372 adolescent/adult deceased controls with a median age of 44 (range 11–91) years. In the AQP1 gene, the rs17159702 CC/CT genotypes were found to be associated with SIDS ($p = 0.02$). In the AQP9 gene, the combination of a TT genotype of rs8042354, rs2292711 and rs13329178 was more frequent in SIDS cases than in controls ($p = 0.03$). In the SIDS group, an association was found between genetic variations in the AQP1 gene and maternal smoking and between the 3xTT combination in the AQP9 gene and being found lifeless in a prone position. In conclusion, this study adds further evidence to the involvement of brain aquaporins in SIDS, suggesting that specific variants of AQP9 genes constitute a genetic predisposition, making the infant vulnerable to sudden death together with external risk factors and probably other genetic factors.

Keywords Aquaporin-1 · Aquaporin-9 · Genetic predisposition · SIDS · Sudden infant death syndrome

Introduction
Sudden infant death syndrome (SIDS) is defined as the sudden unexpected death of an infant less than 1 year of age, with the onset of the fatal episode apparently occurring during sleep, and which remains unexplained after a thorough investigation, including a complete autopsy and a review of the circumstances of the death and the clinical history [1].

Several studies have indicated that a vulnerability in the development and regulation of brain function is involved in SIDS [2–5]. The most robust neurochemical abnormality involves the medullary serotoninergic (5-HT) system, and it has been proposed that an important subset of SIDS infants has serotonergic abnormalities, leading to a failure of protective brainstem responses [6, 7].

Aquaporins (AQPs) are a group of proteins that function as water channels; they facilitate the rapid transport of water and other solutes across the plasma membrane. The three best characterised aquaporins in the brain are AQP1, AQP4 and AQP9 [8]. AQP4 is the main water channel in the brain and spinal cord and it plays a role in brain water homeostasis and neural signal transduction [9, 10]. In mice, it has been reported that a lack of AQP4 expression is paralleled by abnormally high levels of 5-HT in various areas of the brain [11]. This may indicate that a well-functioning water balance is important for the regulation of neurotransmitters, suggesting a possible explanation for the 5-HT imbalances observed in SIDS [7].

In a previous study of the AQP4 gene in Norwegian SIDS, an association was found between rs2075575 CT/TT and SIDS [12], even though this was not replicated in a study of Swiss SIDS cases [13]. An association between a CC genotype (4xCC) in four SNPs (rs17375748, rs1130183, rs12133079 and rs1186688) in the gene encoding the potassium channel Kir4.1 and SIDS has also been reported [14]. This channel is co-expressed with AQP4, and together, they compose a multifactorial unit responsible for maintaining ion and water homeostasis in the brain. The findings in SIDS so far,
both concerning the serotonergic network and aquaporins, as well as the lack of clarity with regard to AQP4 and SIDS. AQP1, AQP4 and AQP9 have been shown to have altered levels of expression in several brain disorders in rodents and humans [15].

AQP1 is located in the apical membrane of the choroid plexus, where it is involved in the formation of cerebrospinal fluid but it is also expressed in the capillary endothelium in many organs and in the tubular epithelium of the kidneys [16]. AQP1 has been linked to hydrocephalus conditions and probably plays a role in the development of cytotoxic oedema [10]. Hypoxia and ischaemia have been shown to stimulate increased AQP1 expression both in vitro and in vivo [17]. AQP1 is reported to be upregulated in pathological states such as brain tumors and Alzheimer’s disease [18, 19].

AQP9 is the only aquaglyceroprotein in the brain; this AQP transports glycerol, amino acids and lactate in addition to water. In the brain, AQP9 is expressed in neurons in the brainstem and in the midbrain, and AQP9 is enriched in the inner mitochondrial membrane [20]. Its level of expression is under the control of insulin, and it is believed that AQP9 is involved in energy metabolism in the brain [21].

The purpose of the present study was to investigate genetic variations in the genes encoding AQP1 and AQP9 in SIDS cases and controls. Our hypothesis is that specific variants of these genes predispose infants to sudden unexpected death.

Materials and methods

Individuals

The individuals included in this study consisted of 168 SIDS cases and 372 adolescent/adult controls (Table 1). All cases were collected between 1988 and 2013, and all underwent autopsy at the Department of Forensic Medicine at Oslo University Hospital. The investigation protocol included an evaluation of the circumstances of death, a review of their medical and family history, a total skeletal radiographic examination and a thorough autopsy with extensive histologic and microbiologic examinations, including a neuropathological examination and toxicological analysis. Adolescent/adult controls were consecutively collected in 2001–2011 from medico-legal autopsies of cases originating from the same geographical area as the SIDS cases. All SIDS cases and controls included in this study were of Caucasian ethnicity.

The SIDS cases were classified according to the San Diego definition [1], applying the criteria used in the Nordic SIDS study [22, 23]. After the genetic analyses, three cases with medium-chain acyl-CoA dehydrogenase A985G mutations and six cases with mutations in long QT syndrome-related genes were excluded. A total of 135 cases were classified as pure SIDS, corresponding to category I SIDS according to the San Diego definition. Another 33 cases were classified as category II SIDS in the San Diego definition, corresponding to undetermined in the new classification recommendations [24]. In these cases, the post-mortem investigation revealed findings of slight infection (n = 19), insignificant findings in the brain (n = 8) or findings in skeletal muscle that might indicate metabolic disease (n = 6); however, these findings were not sufficient to fully explain the cause of death.

Information regarding the risk factors nicotine exposure, position of the infant when found lifeless and any known infection prior to death was obtained from the autopsy reports or directly from the parents using a questionnaire (Table 1). The parents received the questionnaire via mail one to several months after the infant’s death, and participation was voluntary [25]. Therefore, some information is missing for some of the cases.

Gene analyses

A total of 39 single nucleotide polymorphisms (SNPs) were selected for analysis in the genes encoding AQP1 (located at chromosome 7, 19 SNPs) and AQP9 (located at chromosome

| Table 1 | Survey of the individuals included in this study |
|---------|---------------------------------------------|
| Number of cases | 168 | 372 |
| Cause of death | SIDS | 169 violent death | 133 disease | 70 intoxication |
| Sex (male/female) | 108/64 | 265/107 |
| Age (median, range) | 15.5 weeks (2–52 weeks) | 44 years (11–91 years) |
| Nicotine exposure | 82/27/59a | 0/0/372a |
| Found dead prone | 89/58/21 | 0/0/372 |
| Infection prior to death | 68/77/23 | 0/0/372 |

a Number of cases: yes/no/unknown
15, 20 SNPs) (Figs. 1 and 2; online resource 1, 2). There were no interactions between the genes or any conjoined genes. The selection of SNPs was based on these criteria: tag SNPs, SNPs recognised to influence the function or production of the gene, and SNPs resulting in amino acid shifts. The online databases used were The International Genome Sample Resource (www.internationalgenome.org), The University of California Santa Cruz Genome Browser (genome.ucsc.edu) and the dbSNP database provided by the National Centre for Biotechnology Information, USA (www.ncbi.nlm.nih.gov). In addition, literature searches for relevant studies, including AQP1 and AQP9, were performed using PubMed, and four additional SNPs were included [26–28].

DNA was extracted from blood/spleen using standard methods, including phenol/chloroform extraction, ethanol precipitation and a QIAmp DNA Mini Kit using the BioRobotEZ (Qiagen, Hombrechtikon, Switzerland). All SNPs were genotyped using the Sequenom MassARRAY system, following the manufacturer’s protocols (Agena Bioscience, Hamburg, Germany). A multiplex assay was designed using Assay designer software version 2.0, which automatically designed the primers. Following polymerase chain reaction (PCR) and shrimp alkali phosphatase treatment, a primer extension reaction was performed to introduce mass differences between the different alleles. The PCR products were dispensed onto a SpectroCHIP with 384 patches. The mass difference was detected using matrix-assisted laser desorption time-of-flight mass spectrometry (MALDI-TOF MS), and the genotypes were determined using the type 3.1 software (Sequenom, Agena Bioscience, Hamburg, Germany). Cluster plots were made according to the signals of the different allele masses using the default parameters set in the software. An independent platform manager performed the scoring in a blinded fashion. The genotyping was performed at CIGENE, University of Life Sciences, Ås, Norway.

Statistical analyses

The Hardy-Weinberg equilibrium test was performed using a web-based calculator made available through the Online Encyclopedia for Genetic Epidemiology Studies (www.oeyege.org). The chi-square test was used for the first comparison of gene frequencies between SIDS cases and controls and for correlations between risk factors and the genotype. These analyses were implemented using SPSS version 20.0 (SPSS, Chicago, IL). The genotype distributions were further analysed using the Haplin statistical package, which is an open source R package for genetic association studies available at folk.uib.no/gjessing/genetics/software/haplin [29]. Power analysis was performed using the power calculator available at clinical.com/statistics/.

Results

AQP1

In the AQP1 gene, 19 SNPs were investigated (Fig. 1). Of these, 10 SNPs were homozygous for the most common allele in both SIDS cases and controls. Three of the SNPs showed little variation; only one or a few cases were heterozygous and the rest were homozygous for the most common allele. One SNP (rs28362692) was not in Hardy-Weinberg equilibrium in the controls and was therefore excluded. Five SNPs were subjected to further calculations, including rs1004317, rs17159702, rs765840, rs28362731 and rs1049305. The allele frequencies for each SNP are given in Online Resource 1. When investigating each SNP separately, a difference in genotype distribution between the SIDS cases and controls was found for one SNP, rs17159702 (p = 0.036) (Table 3). For this SNP, the C allele and the CC/CT genotypes were more common in SIDS cases than in the controls; 52.1 % of the SIDS cases had these genotypes, compared to 41.5 % of the controls (p = 0.02). The relative risk associated with this SNP was calculated to be 1.4 (1.1–1.9) for the C-allele (p = 0.01) and 2.1 (1.8–3.7) for the CC genotype (p = 0.01). The difference was confirmed when correcting for multiple testing using the Suest command in Haplin (p = 0.045 for alleles). This test computes a joint overall p value based on aggregating the individual p values and then correcting for multiple testing, and it was performed on the five SNPs selected for further studies. In addition, there was a tendency for the GG/
AG genotypes of rs1004317 to be more frequent in SIDS cases than in controls ($p = 0.047$) (Table 2).

When adding known risk factors, we found that the TT genotype of rs17159702 and the AA/AG genotypes of rs1004317 were more frequent in SIDS cases exposed to maternal smoking than in SIDS cases not exposed ($p = 0.014$ and $p = 0.004$, respectively) (Table 3). No other associations between SNPs in the AQP1 gene and the risk factors for SIDS were found.

### AQP9

In the AQP9 gene, 20 SNPs were investigated (Fig. 2). Amongst these SNPs, one SNP (rs17848098) was homozygous for the most common allele in both the SIDS cases and controls, whilst two SNPs, rs77284866 and rs78314281, showed little variation in the investigated populations. These SNPs were homozygous for the most common allele, with the exception of a few cases found to be heterozygous. All SNPs were in Hardy-Weinberg equilibrium in the controls, and 17 polymorphic SNPs were subject to further calculations. The allele frequencies for each SNP are given in Online Resource 2.

When investigating each SNP separately, the TT genotypes of rs8042354, rs2292711 and rs13329178 tended to occur more frequently in SIDS cases than in controls (Table 4). Combined, 56 % of the SIDS cases had a 3xTT genotype combination, compared to 23 of the 52 SIDS cases found lifeless supine or on their side (44 %) ($p = 0.006$). No other associations between SNPs in the AQP9 gene and risk factors for SIDS were uncovered.

### Power analysis

The number of SIDS cases and controls included in this study has the ability to detect a difference in the minor allele frequency of 15 % with a power between 70 and 80 % at the 5 % level of significance.

### Discussion

In this study, it was found that specific variants of the gene encoding AQP1 were more frequent in SIDS cases than in controls (Table 2). AQP1 is present at an early stage in the developing human foetus, and during the early foetal period (8–12 weeks), the third and fourth ventricles of the choroid plexus show strong apical AQP1 staining [30]. This indicates that AQP1 functions as a water channel starting in early foetal life and is thus of functional importance in the developing brain. We found that specific genotypes in two SNPs in the gene encoding AQP1 tended to be more frequent in SIDS cases than in controls (Table 3). Taking into account the essential role of a properly developing choroid plexus and cerebrospinal fluid in the foetal brain, one might speculate that AQP1 gene variations are contributing to a vulnerability to sudden death.

### Table 2: AQP1 allele variants associated with SIDS

| SNP           | Genotype | SIDS          | Controls       | $p$ value |
|---------------|----------|---------------|----------------|-----------|
| rs1004317     | GG/AG    | 114 (67.9 %) * | 216 (58.9 %)   | 0.047**   |
|               | AA       | 54 (32.1 %)   | 151 (41.1 %)   |           |
| rs17159702    | CC       | 17 (10.2 %)   | 23 (6.2 %)     | 0.036***  |
|               | CT       | 70 (41.9 %)   | 131 (35.3 %)   |           |
|               | TT       | 80 (47.9 %)   | 217 (58.5 %)   |           |

* Percentage of the total cases
** Chi-square test, $2 \times 2$ table
*** Chi-square test, $3 \times 2$ table

### Table 3: AQP1 genotype frequency in SIDS according to exposure to maternal smoking

| SNP           | Genotype | Exposed          | Not exposed     | $p$ value |
|---------------|----------|------------------|-----------------|-----------|
| rs1004317     | AA/AG    | 72 (87.8 %) *    | 17 (63 %)       | 0.004     |
|               | GG       | 10 (12.2 %)      | 10 (37 %)       |           |
| rs17159702    | CC/CT    | 38 (46.9 %)      | 20 (74.1 %)     | 0.014     |
|               | TT       | 43 (53.1 %)      | 7 (25.9 %)      |           |

* Percentage of the total cases
** Chi-square $2 \times 2$ table
Maternal smoking is a well-known risk factor for SIDS [31]. Nicotine affects the brain through nicotinic acetylcholine receptors (nAChRs), and it has been shown that these receptors are present in the human choroid plexus [32]. Thus, nicotine has the ability to exert site-specific effects on choroid plexus mechanisms and thereby modulate the function of this tissue [32]. A high incidence of histologic/immunohistochemical alterations in the fourth ventricle of the choroid plexus in SIDS has been reported, and these alterations are significantly related to maternal smoking [33]. APQ1 is located at the apical membrane of the choroid plexus, and it is interesting that the present study revealed an association between genetic variations in the AQP1 gene and maternal smoking in SIDS (Table 4). Our observations strengthen the theory by Lavezzi et al. that some SIDS cases have a vulnerability in the choroid plexus that could be associated with distortions in the cerebrospinal fluid volume, its content and its flow dynamics [33].

The present study found an association between two SNPs in the AQP1 gene and exposure to maternal smoking (Table 3). It is, however, intriguing that although the CT/TT genotypes of rs17159702 were more common in SIDS cases than in controls, the only genotype found to be associated with maternal smoking was the TT genotype. However, for SIDS to occur, it seems that multiple genetic and environmental risk factors must be present. One might therefore speculate that, as maternal smoking is a strong environmental risk factor for SIDS, the death of infants exposed to nicotine depends to a lesser extent on their genetic predisposition. Another explanation may be that the discovered variant acts as a phenotype modifier rather than being a strong genetic predisposing factor for SIDS.

The present study also found specific variants in the AQP9 gene that were more frequent in SIDS cases than in controls (Table 4). The expression of AQP9 has been described in several brain structures, including catecholaminergic neurons [34]. These neurons are known to be involved in energy balance, and the presence of AQP9 may facilitate the diffusion of glycerol that serves as an energy substrate. AQP9 is also present in astrocytes, and it has been shown that a reduction in AQP9 expression induces a decrease in glycerol uptake and changes in astrocyte energy metabolism [35]. In this study, it was found that the combination of a TT genotype in three SNPs in the AQP9 gene was more frequent in SIDS cases than in controls (Table 4). In addition, the 3xTT genotype combination was more frequent in SIDS cases found lifeless in a prone position compared to cases found lifeless supine. One might speculate that part of the risk for SIDS induced by prone sleeping is linked to energy metabolism and a low energy supply due to a lower feeding frequency during the nighttime than during the daytime.

In addition to prone sleeping and maternal smoking, a minor infection in the week prior to death is a risk factor for SIDS [25]. Amongst the cases included in this study, approximately half of the cases had a slight upper airway infection during the last week prior to death. This is more frequent than expected in a series of 100 infants seen in our department for paternity testing, where 26% either had signs of or had a history of a recent common cold [25]. However, we did not find any association between the investigated SNPs and signs of infection in the SIDS cases. This may indicate that a susceptibility to infection is more likely due to variations in genes encoding components of the immune response than in genes encoding brain aquaporins.

It is interesting that several SNPs in aquaporin genes are associated with known external risk factors for SIDS. Previously, rs2075575 CT/TT in the AQP4 gene was reported to be associated with maternal smoking, whilst a combination of 4xCC in rs17375748, rs1130183, rs12133079 and rs1186688 in the gene encoding Kir4.1 was reported to be associated with the prone sleeping position [12, 14]. The present study found that allelic variants of two SNPs in the AQP1 gene are associated with maternal smoking and that three SNPs in the AQP9 gene are associated with a prone sleeping position (Tables 3 and 4). Taken together, these findings fit with the theory of a fatal triangle in SIDS [36]. This theory states that infants may die from SIDS when three conditions occur at the same time: a genetic predisposition, a vulnerable developmental stage in the central nervous system and/or immune system and a triggering event such as maternal smoking or prone sleeping. Given that all three occur, the mechanism of death is a vicious circle starting with trigger events and ending with brain oedema, hypoxia, coma and death [37].

There are a vast number of papers investigating a possible genetic predisposition to SIDS. To date, genes involved in cardiac function, the regulation of the immune system, and brain development and regulation have emerged as the most important [38–40]. A recent study, performing next-generation sequencing of cardiac arrhythmia genes, reported likely pathogenic variants in 15% of the investigated SIDS cases [40]. With regard to the immune system, the most important genes seem to be those encoding interleukins, in

---

### Table 4: AQP9 allele variants found to be associated with SIDS

| SNP          | Genotype | SIDS          | Controls        | *p value*^** |
|--------------|----------|---------------|-----------------|--------------|
| rs2292711    | TT       | 99 (59.3 %)   | 189 (51.4 %)    | 0.09         |
|              | TC/CC    | 68 (40.7 %)   | 179 (48.6 %)    |              |
| rs13329718   | TT       | 101 (60.1 %)  | 190 (51.4 %)    | 0.06         |
|              | TA/AA    | 67 (39.9 %)   | 180 (48.6 %)    |              |
| rs8042354    | TT       | 101 (60.1 %)  | 188 (50.8 %)    | 0.045        |
|              | TA/AA    | 67 (39.9 %)   | 182 (49.2 %)    |              |
| Combined     | 3 x TT   | 94 (56.0 %)   | 171 (46.0 %)    | 0.032        |
|              | other    | 74 (46.0 %)   | 201 (54.0 %)    |              |

^** Chi-square 2 × 2 table
particular IL-6, IL-10 and TNFα, whilst in the serotonergic network, the most promising gene seems to be the gene encoding the serotonin transporter 5-HTT [3, 38]. It is interesting that TNFα-mediated signalling has been reported to upregulate AQ4P and that AQ4P participates in the regulation of serotonergic neurotransmission in different brain regions [11, 41].

In conclusion, this study adds further evidence to the involvement of brain aquaporins in SIDS, suggesting that genetic variations in AQ1P and AQ9P together with the AQ4P complex constitute a genetic predisposition making the infant vulnerable to sudden death in concert with external risk factors and probably other genetic factors.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00414-020-02493-9.

Acknowledgements Open Access funding provided by University of Oslo (incl Oslo University Hospital).

Authors’ contribution All authors contributed to the study conception and design. Case selection, data collection and data analysis were performed by Siri Hauge Opdal. Linda Ferrante was responsible for the selection of the included SNPs. The first draft of the manuscript was written by Siri Hauge Opdal. all authors commented on previous versions of the manuscript and all authors have read and approved the final version of the manuscript.

Funding This study was funded by the South-Eastern Norway Regional Health Authority (grant number 39487 HSØ pro 2012017) and The Norwegian SIDS and Stillbirth Society.

Data availability The datasets generated and analysed during the current study are available from the corresponding author on reasonable request.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethics approval The study was reviewed and approved by The National Committees for Research Ethics in Norway (REK 2011/2356).

Consent to participate The parents or guardians of the included infants were contacted and given the opportunity to decline inclusion in research. None of the included cases have declined. Regarding the deceased adolescent/adult controls, these are completely anonymous and may therefore be included without consent according to Norwegian guidelines. This study was approved by The National Committees for Research Ethics in Norway (REK 2016/2021 and 2010/55).

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

1. Krous HF, Beckwith JB, Byard RW, Rognum TO, Bajanowski T, Corey T, Cutz E, Hanzlick R, Keens TG, Mitchell EA (2004) Sudden infant death syndrome and unclassified sudden infant deaths: a definitional and diagnostic approach. Pediatrics 114(4): 234–238
2. Weese-Mayer DE, Berry-Kravis EM, Zhou L, Maher BS, Curran ME, Silvestri JM, Marazita ML (2004) Sudden infant death syndrome: case-control frequency differences at genes pertinent to early autonomic nervous system embryologic development. Pediatr Res 56(3):391–395. https://doi.org/10.1203/01.PDR.0000136285.91048.AA
3. Opdal SH, Vege A, Rognum TO (2008) Serotonin transporter gene variation in sudden infant death syndrome. Acta Paediatr 97(7): 861–865. https://doi.org/10.1111/j.1651-2227.2008.00813.x
4. Klintschar M, Reichenplader B, Saternus KS (2008) A functional polymorphism in the tyrosine hydroxylase gene indicates a role of noradrenergic signaling in sudden infant death syndrome. J Pediatr 153(2):190–193. https://doi.org/10.1016/j.jpeds.2008.02.032
5. Brownstein CA, Goldstein RD, Thompson CH, Haynes RL, Giles E, Sheidley B, Bainbridge M, Haas EA, Mena OJ, Lucas J, Schaber B, Holm IA, George AL, Kinney HC, Poduri AH (2018) SCN1A variants associated with sudden infant death syndrome. Epilepsia 59(4):e56–e62. https://doi.org/10.1111/epi.14055
6. Bright FM, Byard RW, Vink R, Paterson DS (2017) Medullary serotonin neuron abnormalities in an Australian cohort of sudden infant death syndrome. J Neuropathol Exp Neurol 76(10):864–873. https://doi.org/10.1093/jnen/nlx071
7. Kinney HC, Haynes RL (2019) The serotonin brainstem hypothesis for the sudden infant death syndrome. J Neuropathol Exp Neurol 78:765–779. https://doi.org/10.1093/jnen/niz062
8. Benga O, Huber VJ (2012) Brain water channel proteins in health and disease. Mol Asp Med 33(5-6):562–578. https://doi.org/10.1016/j.mam.2012.03.008
9. Manley GT, Binder DK, Papadopoulos MC, Verkman AS (2004) New insights into water transport and edema in the central nervous system from phenotype analysis of aquaporin-4 null mice. Neuroscience 129(4):983–991. https://doi.org/10.1016/j.neuroscience.2004.06.088
10. Filippidis AS, Carozza RB, Rekate HL (2016) Aquaporins in brain edema and neuropathological conditions. Int J Mol Sci 18(1). https://doi.org/10.3390/ijms18010055
11. Fan Y, Zhang J, Sun XL, Gao L, Zeng XN, Ding JH, Cao C, Niu L, Hu G (2005) Sex- and region-specific alterations of basal amino acid and monoamine metabolism in the brain of aquaporin-4 knockout mice. J Neurosci Res 82(4):458–464. https://doi.org/10.1002/jnr.20664
12. Opdal SH, Vege A, Stray-Pedersen A, Rognum TO (2010) Aquaporin-4 gene variation and sudden infant death syndrome. Pediatr Res 68(1):48–51. https://doi.org/10.1203/PDR.0b013e3181d7e7c
13. Shuder J, Bartsch C, Haas C (2014) Aquaporin-4 polymorphisms and brain/body weight ratio in sudden infant death syndrome (SIDS). Pediatr Res 76(1):41–45. https://doi.org/10.1038/pr.2014.59
14. Opdal SH, Vege A, Stray-Pedersen A, Rognum TO (2017) The gene encoding the inwardly rectifying potassium channel Kir4.1
may be involved in sudden infant death syndrome. Acta Paediatr 106(9):1474–1480. https://doi.org/10.1111/apa.13928

15. Badaut J, Fukuda AM, Jullienne A, Petry KG (2014) Aquaporin and brain diseases. Biochim Biophys Acta 1840(5):1554–1565. https://doi.org/10.1016/j.bbagen.2013.10.032

16. Longatti PL, Basaldella L, Orvieto E, Fiorindi A, Carteri A (2004) Choroid plexus and aquaporin-1: a novel explanation of cerebrospinal fluid production. Pediatr Neurosurg 40(6):277–283. https://doi.org/10.1159/000083740

17. Kaneko K, Yagui K, Tanaka A, Yoshibara K, Ishikawa K, Takahashi K, Jujo H, Sakurai K, Saito Y (2008) Aquaporin 1 is required for hypoxia-inducible angiogenesis in human retinal vascular endothelial cells. Microcirculation 15(3):297–301. https://doi.org/10.1080/0929101070123256

18. Oshio K, Binder DK, Liang Y, Bollen A, Feuerstein B, Berger MS, Manley GT (2005) Expression of the aquaporin-1 water channel in human glial tumors. Neurosurgery 56(2):375–381; discussion 375–381. https://doi.org/10.1227/01.neu.0000148904.57841.6b

19. Perez E, Barrachina M, Rodriguez A, Torrejon-Escribano B, Boada M, Hernandez I, Sanchez M, Ferrer I (2007) Aquaporin expression in the cerebral cortex is increased at early stages of Alzheimer disease. Brain Res 1128(1):164–174. https://doi.org/10.1016/j.brainsci.2006.09.109

20. Amiry-Moghaddam M, Lindland H, Zelenin S, Roberg BA, Bajanowski T, Vege A, Byard RW, Krous HF, Arnestad M, Bachs J, Banner J, Blair PS, Borthne A, Dettmeyer R, Fleming P, Gaustad K, Gidlund E, Helweg-Larsen K, Hirvonen J, Isaksen CV, Kock K, Lavezzi AM, Matturri L, Del Corno G, Johanson CE (2013) Vulnerability of fourth ventricle choroid plexus in sudden unexplained fetal and infant death syndromes related to smoking mothers. Int J Dev Neurosci 31(5):319–327. https://doi.org/10.1016/j.ijdevneuro.2013.04.006

21. Badaut J, Regli L (2004) Distribution and possible roles of aquaporin 9 in the brain. Neuroscience 129(4):971–981. https://doi.org/10.1016/j.neuroscience.2004.06.035

22. Badaut J, Brunet JF, Guerin C, Regli L, Pellerin L (2012) Alteration of glucose metabolism in cultured astrocytes after AQP9-silencing. Biochim Biophys Acta 1840(5):1554–1565. https://doi.org/10.1016/j.bbalip.2012.07.041

23. Badaut J, Brunet JF, Giacomini KM (2008) Genetic variation in regulatory elements of the AQP9 gene motif of aquaporin-1 found in a Colton-null phenotype. Blood 93(11):4021–4023

24. Fabrega E, Berja A, Garcia-Unzueta MT, Guerra-Ruiz A, Cobo M, Lopez M, Bolado-Carrancio A, Amado JA, Rodriguez-Rey JC, Pons-Romero F (2011) Influence of aquaporin-1 gene polymorphism on water retention in liver cirrhosis. Scand J Gastroenterol 46(10):1267–1274. https://doi.org/10.3109/030036552.2011.603161

25. Sorani MD, Manley GT, Giacomini KM (2008) Genetic variation in human aquaporins and effects on phenotypes of water homeostasis. Hum Mutat 29(9):1108–1117. https://doi.org/10.1002/humu.20762

26. Gessing HK, Lie RT (2006) Case-parent triads: estimating single- and double-dose effects of fetal and maternal disease gene haplotypes. Ann Hum Genet 70(Pr 3):382–396. https://doi.org/10.1111/j.1469-1809.2005.00218.x

27. Johansson PA, Dziegielewsk KM, Ek CJ, Hubagd M, Mollgard K, Potter A, Schuliga M, Saunders NR (2005) Aquaporin-1 in the choroid plexuses of developing mammalian brain. Cell Tissue Res 322(3):353–364. https://doi.org/10.1007/s00441-005-1120-x

28. Anderson TM, Lavista Ferres JM, Ren SY, Moon RY, Goldstein RD, Ramirez JM, Mitchell EA (2019) Maternal smoking before and during pregnancy and the risk of sudden unexpected infant death. Pediatrics 143(4):e20183325. https://doi.org/10.1542/peds.2018-3325

29. Lallai V, Grimes N, Fowler JP, Sequeira PA, Cartagena P, Limon A, Coutts M, Monuki ES, Bunney W, Demuro A, Fowler CD (2019) Nicotine acts on cholinergic signaling mechanisms to directly modulate choroid plexus function. eNeuro 6(2):ENEURO.0051–ENEU19.2019. https://doi.org/10.1523/ENEURO.0051-19.2019

30. Johansson PA, Dziegielewsk KM, Ek CJ, Hubagd M, Mollgard K, Potter A, Schuliga M, Saunders NR (2005) Aquaporin-1 in the choroid plexuses of developing mammalian brain. Cell Tissue Res 322(3):353–364. https://doi.org/10.1007/s00441-005-1120-x

31. Anderson TM, Lavista Ferres JM, Ren SY, Moon RY, Goldstein RD, Ramirez JM, Mitchell EA (2019) Maternal smoking before and during pregnancy and the risk of sudden unexpected infant death. Pediatrics 143(4):e20183325. https://doi.org/10.1542/peds.2018-3325

32. Badaut J, Regli L (2004) Distribution and possible roles of aquaporin 9 in the brain. Neuroscience 129(4):971–981. https://doi.org/10.1016/j.neuroscience.2004.06.035

33. Badaut J, Bruet JF, Guerin C, Regli L, Pellerin L (2012) Alteration of glucose metabolism in cultured astrocytes after AQP9 small interference RNA application. Brain Res 1473:19–24. https://doi.org/10.1016/j.brainres.2012.07.041

34. Badaut J, Brunet JF, Giacomini KM (2008) Genetic variation in regulatory elements of the AQP9 gene motif of aquaporin-1 found in a Colton-null phenotype. Blood 93(11):4021–4023

35. Badaut J, Brunet JF, Giacomini KM (2008) Genetic variation in regulatory elements of the AQP9 gene motif of aquaporin-1 found in a Colton-null phenotype. Blood 93(11):4021–4023

36. Badaut J, Brunet JF, Giacomini KM (2008) Genetic variation in regulatory elements of the AQP9 gene motif of aquaporin-1 found in a Colton-null phenotype. Blood 93(11):4021–4023

37. Badaut J, Brunet JF, Giacomini KM (2008) Genetic variation in regulatory elements of the AQP9 gene motif of aquaporin-1 found in a Colton-null phenotype. Blood 93(11):4021–4023

38. Badaut J, Brunet JF, Giacomini KM (2008) Genetic variation in regulatory elements of the AQP9 gene motif of aquaporin-1 found in a Colton-null phenotype. Blood 93(11):4021–4023

39. Badaut J, Brunet JF, Giacomini KM (2008) Genetic variation in regulatory elements of the AQP9 gene motif of aquaporin-1 found in a Colton-null phenotype. Blood 93(11):4021–4023

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.