Serotypes, virulence genes, and PFGE profiles of *Escherichia coli* isolated from pigs with postweaning diarrhoea in Slovakia

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Abstract

**Background:** Postweaning diarrhoea (PWD) in pigs is usually the main infectious problem of large-scale farms and is responsible for significant losses worldwide. The disease is caused mainly by enterotoxigenic *E. coli* (ETEC) and Shiga-toxin producing *E. coli* (STEC). In this study a total of 101 *E. coli* isolated from pigs with PWD in Slovakia were characterized using phenotypic and genotypic methods.

**Results:** These 101 isolates belonged to 40 O:H serotypes. However, 57% of the isolates belonged to only six serotypes (O9:H51, O147:H-, O149:H10, O163:H-, ONT:H-, and ONT:H4), including two new serotypes (O163:H- and ONT:H4) not previously found among porcine ETEC and STEC isolated in other countries. Genes for EAST1, STb, STa, LT and Stx2e toxins were identified in 64%, 46%, 26%, 20%, and 5% of isolates, respectively. PCR showed that 35% of isolates carried genes for F18 colonization factor, and further analyzed by restriction endonuclease revealed that all of them were F18ac. Genes for F4 (K88), F6 (P987), F17, F5 (K99), F41, and intimin (eae gene) adhesins were detected in 19%, 5%, 3%, 0.9%, 0.9%, and 0.9% of the isolates, respectively. The study of genetic diversity, carried out by PFGE of 46 representative ETEC and STEC isolates, revealed 36 distinct restriction profiles clustered in eight groups. Isolates of the same serotype were placed together in the dendrogram, but high degree of polymorphism among certain serotypes was detected.

**Conclusion:** Seropathotype O149:H10 LT/STb/EAST1/F4 (14 isolates) was the most commonly detected followed by O163:H- EAST1/F18 (six isolates), and O163:H4 STa/STb/Stx2e/F18 (five isolates). Interestingly, this study shows that two new serotypes (O163:H- and ONT:H4) have emerged as pig pathogens in Slovakia. Furthermore, our results show that there is a high genetic variation mainly among ETEC of O149:H10 serotype.
Background
Postweaning diarrhoea (PWD) is usually the main infectious problem of large-scale farms and is responsible for significant losses worldwide [1,2]. The disease is caused mainly by enterotoxigenic *E. coli* (ETEC), and Shiga-toxin producing *E. coli* (STEC), also called verotoxin-producing *E. coli* (VTEC) [2-7]. Porcine pathogenic *E. coli* involved in PWD typically belong to serogroups O8, O138, O139, O141, O147, O149 and O157, of which O149 seems to be the predominant serogroup in most countries [1,5,7,8]. ETEC can cause severe diarrhoea in newborn and weaned piglets by the production of heat-labile enterotoxin (LT) and/or heat-stable enterotoxins (Sta or Stb). These enterotoxins are extracellular proteins or peptides, which are able to cause diarrhoea by changing the water and electrolyte balance of the small intestine [5]. Porcine STEC produce the edema enterotoxin (VTe), also named Shiga toxin 2e (Stx2e), which damages the vascular endothelium of the small intestine, subcutis and brain and ultimately leads to subcutaneous edema and neurological disorders [9]. ETEC and STEC implicated in PWD in pigs most frequently produce either the F4 (K88) or F18 fimbrial adhesins [10,11]. Two variants of the F18 fimbrae exist: F18ab (F107) and F18ac (2134P) [11,12]. F18ac is associated with diarrhoea while F18ab is involved in edema disease [11]. In addition to F4 (K88) and F18, other fimbrial colonization antigens such as F5 (K99), F6 (P987), and F41 have also been associated with postweaning diarrhoea, but less frequently [4,13-15].

Porcine attaching and effacing *E. coli* (AEEC) induce intestinal lesions similar to those produced by enteropathogenic *E. coli* (EPEC) in humans. These *E. coli* carry *eae* gene encoding a 94 kDa outer membrane protein (intimin) which is responsible for intimate attachment to epithelial cells. However, the pathogenic significance of porcine *eae*-positive isolates in weaned pigs is unclear [16,17]. A new category of the diarrhoeagenic *E. coli* family, named enteroaggregative *E. coli* (EAEC), has been recognized. EAEC elaborate a low-molecular-weight, partially heat-stable, plasmid-encoded enterotoxin named enteroaggregative *E. coli* heat-stable enterotoxin 1 (EAST1). The gene (*astA*) encoding the production of EAST1 has been detected in several groups of diarrhoeagenic *E. coli* (EAEC, EPEC, ETEC, and STEC) isolated from humans and from pigs. The pathogenic significance of EAST1 in diarrhoea in pigs is not known [7,8,18-20].

Although, PWD is frequently observed in Slovakia, there is a lack of information about the prevalence of serogroups, serotypes, and virulence factors of porcine pathogenic *E. coli*. Thus, the aim of this study was to determine the distribution of serogroups, serotypes, and virulence genes, and to study the genetic relatedness among *E. coli* isolated from pigs with PWD. This is the first study in Slovakia of a large collection of pathogenic *E. coli* isolated from PWD.

Results

**Serogroups and serotypes**

The 101 porcine isolates belonged to 24 O serogroups and 40 O:H serotypes. However, 54% were of one of these eight serogroups (O8, O9, O45, O54, O141, O147, O149, and O163) and 57% of the isolates belonged to only six serotypes, including two new (O163:H- and ONT:H4) serotypes not previously found among porcine pathogenic *E. coli*. The most common serotypes were: O149:H10 (16 isolates), ONT:H- (13 isolates), O163:H- (11 isolates), O9:H51 (nine isolates), ONT:H4 (five isolates), and O147:H- (four isolates) (Table 1).

**Toxin genes**

Amplification of the toxin genes by PCR showed that 77% of isolates possessed genes for production of five types of toxins: LT, Sta, Stb, Stx2e, and EAST1. The gene encoding for EAST1 toxin (65 isolates) was the most prevalent, followed by the STb (47 isolates), Sta (27 isolates), and LT (20 isolates) genes. The Stx2e gene was detected in five isolates, which also carried genes for Sta and Stb (Table 1). Genes encoding Stx1, Stx1c, Stx1d, Stx2, Stx2c, Stx2d, and Stx2g toxins were not detected in any of the 101 porcine isolates studied.

**Adhesin genes**

The PCR analysis of all 101 isolates of *E. coli* showed that 61 (60%) carried at least one fimbrial or intimin gene. The most prevalent fimbrial adhesin was F18, detected in 35 isolates. Analysis by restriction endonucleases of PCR F18-positive products revealed that all 35 F18-positive isolates showed the F18ac variant. Three of these 35 F18 isolates were also positive for either F4 or F17 genes. The gene encoding F4 was identified in 19 isolates. F6, F5, F41, and F17 genes were detected in five, one, one, and three isolates, respectively. The *eae* gene (intimin type β1) was detected in only one isolate (0.9%) of serotype O45:H- (Table 1).

Of the 19 *E. coli* F4-positive, 15 isolates belonged to O149:H10 serotype. The remaining four isolates belonged to O8:H19, O8:HNT, O118:H9, and ONT:H19. The F18 isolates were distributed in a wide range of serotypes, however, 29 of 35 isolates belonged to four predominant including O163:H- (11 isolates), ONT:H- (nine isolates), ONT:H4 (five isolates), and O147:H- (four isolates). Also F6 isolates were widespread among serotypes. Isolates carrying F17 gene belonged to O141:H25, O147:H-, and ONT:H- serotypes, and the isolate carrying both F5 and F41 genes belonged to ONT:H9 serotype.
Most isolates showing genes for fimbrial adhesins also possessed genes for toxin production, and the most common associations were: LT/STb/EAST1/F4 (18 isolates), EAST1/F18 (13 isolates), STa/Stb/F18 (seven isolates), STa/Stb/EAST1/F18 (five isolates), STa/Stb/Stx2/F18 (five isolates), and STa/Stb/EAST1/F6 (five isolates).

Seropathotypes
Although the 101 porcine E. coli isolates belonged to 57 different seropathotypes (association between serotypes and virulence genes), only seven accounted for 39% of isolates. Seropathotype O149:H10 LT/STb/EAST1/F4 (18 isolates), EAST1/F18 (13 isolates), STa/Stb/F18 (seven isolates), STa/Stb/EAST1/F18 (five isolates), STa/Stb/Stx2/F18 (five isolates), and STa/Stb/EAST1/F6 (five isolates) were the most common, followed by O163:H- EAST1/F6 (five isolates), STa/Stb/STa/STb/EAST1/F18 (two isolates), and STa/Stb/EAST1/F17/F18 (two isolates).

Haemolytic activity
Haemolytic activity on blood agar plates was detected in 72 (71%) of the 101 E. coli isolates. All F4 and F18 isolates were haemolytic. Regarding the serotypes, all isolates of O149:H10, O163:H- and ONT:H4 were haemolytic (Table 1).

Macrorestriction fragment analysis by pulsed-field gel electrophoresis (PFGE)
A representative group of 46 isolates (45 ETEC and/or STEC) were selected to be analyzed by PFGE: O149:H10 (15 isolates), O163:H- (11 isolates), O147:H- (four isolates), ONT:H4 (five isolates), O141 (two isolates), and O60:H- (one isolate). The study revealed 36 distinct restriction profiles, considering as significant a difference of a single band (Fig. 1). In the dendrogram produced by the UPGMA algorithm, the isolates were clustered in eight groups (I to VIII; 1 to 13 isolates per group) of 70% similarity according to the Dice similarity index, with 35 isolates clustering in nine subgroups of closely related (similarity > 85%) PFGE profiles. E. coli isolates of the same serotype were placed together in the dendrogram, but high degree of polymorphism among certain serotypes was detected. Thus, the 15 O149:H10 isolates were clustered in three groups (I-III, 70% similarity) with only three small subgroups of closely related profiles (similarity > 85%; five, two and two isolates, each). Genetic distance among O149:H10 isolates was as considerable as similarity < 66%. Group V clustered 13 isolates (all 11 O163:H- and two ONT:H4 isolates; similarity > 71%) with three subgroups (two of them clustering five isolates each, similarity > 85%). Curiously, the highest homogeneity (similarity > 92%) was observed among a group of 10 isolates (group VI) belonging to serotypes O147:H- (four isolates) and ONT:H- (six isolates). E. coli isolates of serotypes O141:H- and O141:H34 were clustered in group VII (similarity > 97%). And all five isolates of serotype ONT:H4 clustered in group VIII showing a similarity > 81%.

Discussion
It is widely accepted that specific serotypes and pathotypes of ETEC and STEC are responsible for the major part...
| Target gene coding for virulence factors | Oligonucleotide sequences of primers | Amplified Products (bp) | Primer Coordinates | Accession number | Reference |
|----------------------------------------|-------------------------------------|------------------------|--------------------|------------------|-----------|
| LT                                    | 5'-ATT TAC GGC GTT ACT ATC TGT CGG TCA CAT ATC-3' 5'-TTT TGG TCT TGG TCA CAT ATC-3' | 281 | 27–47 287–307 | S60731 | Osek et al. [35] |
| STa*                                 | 5'-TCC GTG AAA CAA CAT GAC GGC GGC-3' 5'-ATA ACA TCC AGC ACA GCC AGG-3' | 244 | 267–286 492–510 | M58746 | Ojeniyi et al. [34] |
| STb                                   | 5'-GCC TAT GCA TCT ACA CAA GAC GTC GGT-3' 5'-TGA GAA ATG GAC AAT GTC CGG-3' | 279 | 515–534 773–793 | AY028790 | Ojeniyi et al. [34] |
| Stx1aP                                | 5'-CGG TGA ATG TCA TCT GTG CTC GTG CTC-3' 5'-CGT GGT ATA GTA GCT ATG GTC ACC-3' | 302 | 113–134 394–414 | M17358 | Blanco et al. [32] |
| Stx2aP                                | 5'-CTG TGG TAT CCT ATT CCC GGC-3' 5'-CTG TGG TGA CAG GCA AAC GGC-3' | 516 | 50–69 543–565 | M59432 | Blanco et al. [32] |
| STx2e                                 | 5'-ATG GAG ATG ATT ATA ATG GAG-3' 5'-TCA GGT AAA CTT CAC CGG CTT-3' | 264 | 1176–1196 1419–1439 | M56727 | Osek et al. [35] |
| EAST1                                 | 5'-CCA TCA ACA CAG TAT ATC CGA-3' 5'-GCC GCT GAG TGA CGG CTT TGT CGG-3' | 111 | 2–24 94–114 | S81691 | Yamamoto and Nakazawa [18] |
| F4 (K88)d                             | 5'-GCT GCA TCT GCT GTA TGT GAT G-3' 5'-CCA CAT TAC ATG GTG CTC GC-3' | 792 | 31–54 798–822 | M29374 | This study |
| F5 (K99)                              | 5'-TGC TGC TAC CAA TCA TGC TGC-3' 5'-ATG CCA CCA TTA GAC GGA GC-3' | 450 | 45–64 475–494 | M35282 | Ojeniyi et al. [34] |
| F6 (P987)                             | 5'-TCT CCT GCT TTA AAA GCT ACT GCT GGC-3' 5'-AAC TCC ACC TGG TTA ATG AGG-3' | 333 | 193–212 506–525 | M35257 | Ojeniyi et al. [34] |
| F17e                                  | 5'-GGG GTG ACA GAG GAG GTG GGG-3' 5'-GCC GGC GAC AAC GAC TTT ATC ACC GGC-3' | 411 | 289–310 677–699 | AF055313 | This study |
| F18f                                  | 5'-GTG AAA AGA CTA GTG TTT ATT TCA-3' 5'-CTT GTT AGT ACC CGG CTG AGG-3' | 510 | 1–23 490–510 | M61713 | Imberechts et al. [33] |
| F41                                   | 5'-GAG GGA CTT TCA TCT TTA AGT CGA CCA TCC TGT TTA TAG GC-3' | 431 | 154–173 565–584 | X14354 | Ojeniyi et al. [34] |
| eae*                                  | 5'-GGG ACA GCA GAC GAT TGT CGG CAG-3' 5'-GCC GCT CAT CAT AGT GGT TTT TCA-3' | 775 | 1441–1460 2193–2215 | AF022236 | Blanco et al. [32] |

*Detects STaP and STaH variants.
*Detects STa and the variants STa1 and STa2.
*Detects STa2 and the variants STa2c, STa2d, STa2e, and STa2g.
*Detects K88b, K88b, and K88d.
*Detects F17a, F17b, F17c and F17d.
*Detects F18a (F107) and F18c (134P).
*Detects all types of eae variants (Primer pair with homology to the 5' conserved region of eae).
of PWD in piglets. However, the distribution and frequencies of serotypes and pathotypes can vary considerably from region to region and over time in a given region. The majority of the virulence factors are controlled by transferable genetic elements (plasmids and transposons) and thus, common pathogenic seropathotypes may be replaced by previously uncommon types emerging as new pig pathogenic *E. coli*.

This is the first study in Slovak Republic of a large collection of pathogenic *E. coli* isolated from PWD. In the present study, although 101 isolates belonged to 40 different O:H serotypes, more than a half of ETEC and STEC belonged to only five serotypes: O147:H-, O149:H10, O163:H-, ONT:H-, and ONT:H4. Most isolates of these five serotypes possessed either the F4 or the F18 genes. ETEC of O147:H-, O149:H10 and ONT:H- serotypes are also frequently detected in pigs from other countries, especially O149:H10 [5,8]. The seropathotype O149:H10 LT/Stb/EAST1/F4 (14 isolates) was the most prevalent in the present study, and the reason for its predominance is not known. A possible explanation could be that the virulence factor association of these isolates makes them especially adapted to propagation in swine populations and their environment [8].

The main discovery of this study was the identification of two new serotypes (O163:H- and ONT:H4) not previously detected among porcine ETEC and STEC isolated in other geographical zones. All 11 isolates of O163:H- serotype harbored the F18ac fimbiae gene and five of them were positives for Sta and Stb enterotoxins. Interestingly, all five Stx2e-positive isolates identified in the current study belonged to the new serotype ONT:H4. In previous studies the Stx2e production was associated mainly with O138:H14, O138:H-, O139:H1, O141:H4, O147:H6 and O157:H19 serotypes [5,12,21]. Thus, this study reports two new serotypes (O163:H- and ONT:H4) emerging as pig pathogens in Slovakia.

Although haemolysin does not seem to play an essential role in the virulence of porcine ETEC and STEC, most of the typical PWD *E. coli* are haemolytic [22]. In this study, 71% of isolates were haemolytic, and 66 isolates of those 72 possessed other virulence genes. Furthermore, all 32 isolates belonging to the three serotypes (O149:H10, O163:H- and ONT:H-) most frequently detected in this study showed haemolytic activity. Thus, the haemolytic activity is a very good marker for pig pathogenic *E. coli*.

F4 and F18 are the most important fimbrial adhesins of ETEC and STEC causing PWD [10,11]. In the present study using PCR analysis, it was shown that 35% of *E. coli* isolated from PWD carried genes for F18 colonization antigen. Our findings are in accordance with those of others [1,7,10,23,24]. Based on the studies of Rippinger et al. [25], the F18-family of fimbriae were divided into two variants F18ab and F18ac. The *E. coli* expressing F18ab cause edema disease, whereas the isolates with F18ac cause PWD [11]. In the present study, after digestion of PCR products of F18 isolates, all 35 F18-positive showed the F18ac variant, and more than a half of these 35 isolates belonged to O141, O147 and O163 serogroups. We found that 19 (19%) of 101 isolates from pigs with PWD carried the F4 gene. Several studies have demonstrated that the O serogroups associated with the fimbria F4 are mainly: O8, O149, and O157 [1,5,7,13,15]. Our results confirm these findings as 17 of 19 of Slovak F4 isolates belonged to O8 and O149 serogroups.

Zhu *et al.* [16] demonstrated that the *eae* gene is associated with A/E activity of O45 *E. coli* isolated from swine PWD. However, the AEEC are less commonly associated with PWD than ETEC. In the present study, we found only one
O45:H- isolate (0.9%) positive for the eae gene. In accordance with our results, Frydendahl [7] and Osek [26] also found only 1% (3 of 219) and 3% (6 of 224) of Danish and Polish PWD isolates carrying the eae gene, respectively. F17-producing E. coli are commonly isolated from calves with or without diarrhea [27]. In this study, the F17 gene was detected in three E. coli, two of which were also positive for F18, enterotoxins, and EAST1 toxin. Similarly, Osek [6] found that only four (1%) of 372 isolates from PWD in Poland were positive for F17.

The role of EAST1 toxin in swine colibacillosis has not been demonstrated, however, the gene encoding EAST1 toxin is commonly found in isolates associated with PWD [7,8,19,20]. Our results confirm these observations as we found that 65 of 101 isolates harbored astA gene and all F4 isolates were astA positive. The high frequency of the astA gene suggests the necessity of further studies to investigate the significance of this toxin in porcine PWD.

Genotyping methods such as multilocus enzyme electrophoresis (MLEE) and pulsed-field gel electrophoresis ( PFGE) have been used for differentiation and epidemiological characterization of E. coli isolated from pigs with PWD and edema disease. PFGE is a powerful tool to reveal inter- and intra-serotype specific genetic differences among porcine pathogenic E. coli [12,28,29]. However, there are few studies reporting genetic relatedness of E. coli isolated from diarrhoea in pigs. Osek [29] used the PFGE technique to analyze 82 E. coli from pigs with PWD, isolated from geographically separated farms in the western part of Poland. The 82 isolates belonging to four serogroups (O138, O139, O141, and O149) showed 13 different PFGE profiles and although a high degree of polymorphism among different serotypes was observed, isolates belonging to the same serological group showed a close relationship. Thus, the 25 isolates of serotype O149:K91 generated only two PFGE types. In our study, a representative group of 46 ETEC and STEC isolates revealed 36 distinct restriction profiles. Although isolates of the same serotype were placed together in the dendrogram, high degree of polymorphism among certain serotypes was detected. Thus, 13 distinct PFGE profiles resulted from 15 O149:H10 isolates analyzed, in spite of the fact that 14 E. coli of those 15 carried the same virulence genes (LT/ STb/EAST1/F4). Similarly results have been found in Spain among isolates of the most prevalent serotype (O157:H- LT/STb/F4) nowadays (unpublished data). Further studies are necessary to know if some of these clusters have appeared recently, and if so, analyze its evolution, as well as if there is any relationship with pathogenicity in farms.

Conclusion
Our results indicate that in Slovakia, as described in other countries, pathogenic E. coli isolates from PWD belong to a restricted number of serotypes and pathotypes. The ETEC serotype O149:H10 seems to be predominant, but also two new serotypes (O163:H- and ONT:H4) not previously described in porcine ETEC and STEC isolated in other countries are common. The F18ac and F4 fimbriae were the most prevalent colonization factors detected in postweaning E. coli in Slovakia. Macrorestriction analysis showed that, although isolates of the same serotype and virulence markers mainly share the same PFGE group, there is a high genetic variation, especially among ETEC of O149:H10 serotype.

Methods
E. coli isolates
One hundred and one E. coli isolated from the same number of pigs with PWD were investigated in this study. The 101 post-weaning isolates came from 20 farms located in different parts of Slovakia. Five isolates were selected randomly from each farm, except one farm from Zemplinska Teplica, from which six isolates were collected. Of these 20 farms, nine farms were located in East part of Slovakia, seven farms in Central part of Slovakia, and the remaining four farms were in West part of Slovakia. The E. coli were isolated from the intestinal contents of carcasses of postweaning pigs with diarrhea at the Department of Bacteriology (State Veterinary Institutes in Bratislava, Nitra, and Zvolen, Slovakia) and the Department of Food Hygiene and Technology (Institute of Microbiology and Immunology, University of Veterinary Medicine, Kosice, Slovakia) between 2001 and 2003. The fecal samples were plated onto MacConkey agar (Oxoid, UK) and the E. coli isolates were identified by standard biochemical procedures. After isolation, the E. coli were stored in Luria-Bertani broth containing 20% glycerol at 70°C for further characterization studies.

Reference strains
The E. coli strains used as a control were: 298 (F4/K88), 329 (F5/K99), 318 (F6/987P), 320 (F41), 216 (F18 and Stx2e), 281 (LT), 256 (Sta and STb), EDL933 (Stx1, Stx2 and eae-γ1), G491(F4/K88ac), P201 (F4/K88ad), 5138 (F18ab), 8813 (F18ac), 253KH85 (F17), 226KH85 (F17), 960205 (EAST1), 022206 (EAST1), and E. coli C600 (as negative control). Some of the control strains were kindly supplied by Dr. J. Osek (National Veterinary Research Institute, Pulawy, Poland), Dr. P. Alexa (Veterinary Research Institute, Brno, Czech Republic), Dr. P.F. Lintemans (Institut National de Recherches Veterinaires, Bruxelles, Belgium), and Dr. C. Chae (Department of Veterinary Pathology, College of Veterinary Medicine, Seoul National University, Republic of Korea).
Serotyping
The determination of O and H antigens was carried out by the method described by Guinée et al. [30] employing all available O (O1–O181) and H (H11–H56) antisera in the LREC (Lugo). All antisera were obtained and absorbed with the corresponding cross-reacting antigens to remove the nonspecific agglutinins. The O antisera were produced in the LREC (Lugo, Spain) and the H antisera were obtained from the Statens Serum Institut (Copenhagen, Denmark).

Haemolysin activity
The isolates were inoculated on blood agar base supplemented with 5% sheep blood (Oxoid, UK) and incubated at 37 °C for 18h. β-Haemolysis was evident as a zone of lysis surrounding the bacterial growth.

Detection of virulence genes by PCR
The polymerase chain reaction (PCR) for detection of toxins (LT, Sta, STb, Stx1, Stx2, Stx2e, and EAST1) and adhesins (F4, F5, F6, F17, F18, F41, and eae) was carried out as described by Vu-Khac et al. [31] and Blanco et al. [32]. Base sequences and predicted sizes of the amplified products for the specific oligonucleotide primers used in this study are shown in Table 2 [31-35]. Typing of eae ( intimin) gene detected in one positive isolate identified in this study was carried out by PCR as described elsewhere [32].

Digestion of PCR products with restriction endonuclease
After amplification, the PCR products of F18-positive isolates were digested with restriction enzyme NgoMIV (formerly NgoMI) to distinguish genes encoding F18ab and F18ac [33]. The master mix was prepared with 0.15 U l volume containing 5 l of PCR products (after purifying with Wizard PCR Preps [Promega]; 10 l 1x MULTI-CORE™ buffer [Promega]; and 1 unit of enzyme NgoMIV. After incubation at 37 °C for one hour, the DNA digestion was analyzed by electrophoresis in 2 % agarose gel.

Pulsed-field gel electrophoresis
PFGE was performed in a CHEF MAPPER system (Bio-Rad, Hemel Hempstead, United Kingdom) at 14 °C in 0.5XTBE by the Internet proposed standard-protocol for PFGE [36]. Cleavage of the agarose-embedded DNA was achieved with 0.2–0.8 U/µl XbaI (Roche) according to instructions of the manufacturer. Run times and pulse times were 2.20 to 54.0s for 22 h with linear ramping. PFGE was used to establish clonal relatedness and diversity among a representative group of 46 isolates. To perform the comparison of the PFGE pulsotypes, TIFF files were analyzed with BioNumerics software (Applied Maths, Sint-Martens-Latem, Belgium). Cluster analysis of the Dice similarity indices based on the unweighted pair group method using aritmetic averages (UPGMA) was done to generate a dendrogram describing the relationship among EPEC pulsotypes. A difference of at least one restriction fragment in the profiles was considered the criterion for discriminating between clones.

Authors’ contributions
H. Vu-Khac, E. Holoda, and E. Pilipeinec isolated the E. coli and performed the detection of virulence genes by PCR, M. Blanco, G. Dahbi and E. A. González also participated in the PCR study, J.E. Blanco did the serotyping of the isolates, A. Mora and C. López were responsible for the study of genetic diversity carried out by PFGE, and H. Vu-Khac, and I. Blanco designed the study and drafted the manuscript. All authors read, commented on and approved the final manuscript.

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