Abstract. The present study performed a retrospective observational study in order to investigate the relationship between the interleukin family gene polymorphisms and risk of multiple myeloma (MM), based on sixteen case-control studies that contained 2,597 patients with MM and 3,851 controls. The results demonstrated that the genotypes IL-6 and IL-1 GG increased the risk of MM by approximately 40.8 and 80.2% compared with the genotypes AA and CC [odds ratio (OR)=1.14, 95% confidence interval (CI), 0.88-1.47, and OR=1.16, 95% CI, 0.61-2.19; respectively]. The results also revealed a significant association between T:C polymorphism of the IL-6 and IL-10 and the risk of MM (TC/CC: OR=1.37, 95% CI, 0.88-2.16 and TT/CC: OR=1.26, 95% CI, 0.77-2.06, respectively). Additionally, no significant association was identified between the C:A polymorphisms of the IL-6 (rs8192284) and IL-10 (rs1800872) receptors and the overall risk of MM (P>0.05). G:C polymorphisms of the IL-1β1464G>C and IL-6572G>C significantly increased the risk of MM (P<0.05). However, it has been determined that there is a significant association between the C:T polymorphism of the IL-1α-889C>T and IL-1β-3737C>T and the risk of MM (P<0.001). Subgroup analysis revealed that the detection of G:A polymorphisms in the IL-6 promoter (OR=1.05, 95% CI, 0.78-1.44) is more accurate in MM samples of the Asian population (OR=1.24, 95% CI, 0.92-1.74). In addition, no significant association was identified between the IL family gene polymorphisms in MM samples categorized by ethnicity and the IL family type (P=0.27). These single nucleotide polymorphism loci may be the appropriate gene markers for gene screening and a promising therapeutic strategy in the prognostics of patients with MM.

Introduction

Multiple myeloma (MM) is a common neoplastic disorder (1). It is the second most diagnosed hematological malignancy in China with the incidence rate of ~1.1/100,000 that accounts for 2.1% of all new cancer cases in 2016 (2). Typically, MM is an incurable cancer of the plasma cells, which is identified by the infiltration and clonal proliferation of antibody-secreting post-germinatal center plasma cells in the bone marrow that lead to renal insufficiency, bone disease and anemia (3,4).

Recent articles have indicated that age, male sex, obesity and ionizing radiation exposure are the most common risk factors for MM (5,6). Nevertheless, the exact cause of MM is still unknown (6). Epidemiological studies have shown an increasing incidence rate of MM in Caucasian countries. Although African populations have the highest incidence rate of MM, while the lowest ones belong to the Asian and Mediterranean populations (7,8).

The pathology of MM is speculated to involve multiple factors including genetic, immune system and environmental causes. Since MM is a heterogeneous disease, genetic factors play an important role in its etiology (9). Gene polymorphisms of inflammatory factors may be involved in the progression of MM by unbalancing pro- and anti-inflammatory cytokines profile (9,10).

Interleukin (IL) family that are involved in the immune response and inflammatory processes of MM, consist of a group of lymphatic factors with a multiplicity of biological activities (11,12). Many experimental evidences have revealed specific associations between the metastatic risk of MM and activation of interleukin (12).
interleukin family, IL-6 with its receptor IL-6R is the major interleukin for the T helper (Th) mediated inflammation and it is important in maintaining the Th1/Th2 balance in the inflammatory stage of MM [13,14]. These observations indicate that human IL-6 and IL-10 polymorphisms might act as a biomarker for monitoring the clinical course of MM [15,16]. Consequently, the prognostic value of genetic factors are potentially limited in genotype-related interleukin, specific interleukin loci and individual phenotype [14].

Recently, a multivariable analysis was used to determine if interleukin family polymorphisms are a prognostic factor for survival in MM patients [17-19]. It has been widely reported that many different polymorphisms in different loci are related to poor survival and prognosis of MM [19,20]. The polymorphism of interleukin family occurs in MM patients, and it is associated with statistically increased of MM risk [20-22]. Interleukin production is regulated by the polymorphisms of cytokine genes in promoter regions [20,23]. The high level of pro-inflammatory interleukin level is a significant predictor of both progression-free survival (PFS) and overall survival (OS) in Caucasian MM patients [24,25].

Although, the results of these studies are controversial or inconclusive due to their limited genuine heterogeneity, stage of MM and sample size. Therefore, we conducted a quantitative systematic review along with a comprehensive meta-analysis to resolve inconsistent and often ambiguous findings. Hence, this paper attempted to investigate the associations between the interleukin family gene polymorphisms and MM patients.

Materials and methods

Ethics statement. The present meta-analysis was approved by an independent Ethics Committee/institutional review board at Southwest Medical University, Department of Hematology in Luzhou, China. This investigation was carried out by following recommendations of the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) [26].

Search strategy and study identification. A comprehensive literature search was conducted in MEDLINE electronic databases of PubMed, Embase, Wiley Online Library, Web of Science, Science Direct and VIP- Google Scholar Database. All databases were searched without using language restrictions to assess the prognostic value of interleukin family polymorphism in MM patients prior to July 05, 2018, with no lower date limit. The search string was conducted by using MeSH terms and following the main heading term or word (both the US and UK variants). Based on the research question, the combinations of the keywords or main headings were determined as follows: 'IL' OR 'interleukin', OR 'IL-' AND 'polymorphism' OR 'polymorphisms' OR 'interleukin polymorphisms' OR 'single nucleotide polymorphism' OR 'SNP', AND 'myeloma' OR 'multiple myeloma'. We have repeated these terms for each of the investigated interleukins, including: IL-1α/β, -4, -6, -10, -17, -21, -23, -26 and their receptors. Alternative synonyms and spellings were incorporated using Boolean 'OR' and the main terms were linked with Boolean 'AND'.

Inclusion/exclusion criteria. The following inclusion standards were used to select potential studies for this meta-analysis: i) Evaluated association between the individual IL and/or the IL receptor polymorphism with MM risk; ii) Human genotypes involvement iii) Use of a case-control design; iv) Contained raw genotype frequencies for at least one IL promoter and/or IL receptor polymorphism; v) Publication in English. All case reports, retrospective studies, editorials and review articles, family-based studies, unrelated articles, studies without available genotype frequencies, studies that only investigated the impact of IL polymorphisms in response to therapy or drug resistance and genome-wide association studies were excluded. When an author had published several articles with data obtained from the same patient population, only the newest or the most informative article was selected.

Data collection. The titles and abstracts of all selected articles were analyzed according to the PICO principle by two independent investigators (MNS and II) [27]. Any disagreement or discrepancy was adjudicated through debate or consultation; if a consensus was not reached then a third investigator (SH) was consulted. The electronic study was supplemented by a hand-search of relevant articles from reference lists to ensure that all relevant research were identified. Synchronously, references of review articles were checked for any relevant articles and bibliographies. The following key components of all qualified studies were recorded: First author's name, year of publication, ethnicity, country of origin, genotyping methods, sample size, characteristics of controls and matching criteria, study design, TNM stage, clerk stage, reference control, total number of cases and controls that were stratified by genotype frequencies (homozygous wild-type, heterozygous and homozygous mutant), P-value for Hardy-Weinberg equilibrium (HWE) of controls and true and false positives and negatives. We also e-mailed the corresponding authors of the selected articles to obtain additional and/or any missing information, as well as the copies of the original data required for the meta-analysis. Finally, if the above information was not mentioned in the original study or we did not receive any response via email, the item was preserved as 'not reported (NR)'.

Quality assessment. The quality of the included studies were assessed by two authors (CY and CH), independently according to the Newcastle-Ottawa Scale (NOS) [28] and Quality Assessment of Diagnostic Accuracy Studies 2 (QUADAS-2) [29]. Low-quality studies with scores of 4 or lower were excluded. Additionally, the risk of bias was calculated according to criteria from the Cochrane Collaboration's tool (Cochrane Handbook for Systematic Reviews of Interventions Version 5.1.0.).

Statistical analysis. The systematic search was performed using Review Manager Software version 5.2 [The Nordic Cochrane Centre, Copenhagen, Denmark]. The RevMan version 5.2 was used to combine data (free software was downloaded from http://www.cochrane.org, The Cochrane Collaboration, The Nordic Cochrane Centre, Copenhagen, Denmark, 2012). Data were presented as means ± standard deviation (SD) or median (range), and a description of qualitative variables as
number and percentage. HWE was checked by \( \chi^2 \) test (30). The results of the meta-analysis were reported as odds ratios (ORs) with 95% confidence intervals (CIs). The ORs were pooled for each allele comparison in the three models, the dominant model, the recessive model and heterozygote model. The Chi-square-based Q-test was applied to testify between-study heterogeneity. Hazard ratio (HR) was calculated by the fixed effects model when \( P \) (heterogeneity) > 0.05, as a secondary analysis. Otherwise, the random-effects model was used. According to our hypothesis and inclusion criteria, HR > 1 implies poor prognosis for any IL polymorphisms. We used pooled HRs with 95% CIs to find the relationship between IL single nucleotide polymorphisms (SNPs) and MM susceptibility. The Hazard ratios and 95% CIs were calculated by Tierney's method if the data were not reported in the original paper. Furthermore, subgroup analysis was conducted for the type of each allele polymorphisms and ethnicity, genotyping methods, IL type and reference control. Publication bias was evaluated by Begg's funnel plots and Egger's regression test (31). A value of ‘Pr>|z|’ less than 0.05 was considered as potential publication bias (32). Moreover, we also conducted a sensitivity analysis by precluding a single study to observe whether the pooled HRs changed. Statistical analyses were conducted using Meta-DiSc version 1.4 and R software Packages (version 3.3.1), included ‘mada’ (for sensitivity and specificity analysis), \( P < 0.05 \) and \( I^2 > 50\% \) was considered as statistically significant.

**Results**

**Study selection.** A detailed PRISMA flowchart of the study identification, screening and exclusion process are shown in Fig. 1. A total of 431 studies were retrieved by database searching, with 340 potentially eligible studies that met the inclusion and exclusion criteria and 1 record attained through manual search. By screening the titles, 268 studies were excluded due to being duplicated. After carefully reviewing abstracts, 126 studies were excluded including conference studies, review articles, thesis compilations, those that were not either in English or Chinese, cell culture and animal studies data. Of the remaining 37 full-text candidate articles, 21 potential studies were excluded involving insufficient data, other cancer studies and unrelated studies. Finally, 16 studies were selected and presented in the current meta-analysis that attempts to find a relationship between interleukin polymorphism and risk of MM.

**Study and SNP characteristics.** The main characteristics of the included studies are shown in Table I. The published dates of all research studies varied from 2000 to 2018. Most of the research was conducted within Caucasian communities: 10 studies in Caucasian populations (62.5%) (20,22,23,25,33-38), 3 within Asian populations (18.75%) (17,39,40) and 3 in Mixed Afro-American populations (18.75%) (24,41,42). Of note, the quantitative TaqMan method (TaqMan PCR) was often used

Figure 1. Flowchart of the study selection in the meta-analysis (following PRISMA guidelines, n=number of studies).
to detect the SNP state with control references (11 studies, 68.75%) (20,22-25,36-39,41,42). Also, polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method was used in 5 studies (31.25%) (17,33-35,40). The variables from sixteen relevant case-control studies were contained 2,597 cases and 3,851 controls. Source controls were mainly selected by population-based method with age/sex-matched (12 studies, 75%) (17,20,23-25,33,34,36,38,40-42). Regarding the NOS methodological quality, all included studies were of high quality with ≥7 out of 10 (mean 8.02 point).

The genotype susceptibility of MM and interleukin family gene polymorphisms are shown in Table SI. In Total, 16 published studies had detected interleukin polymorphism of IL-1β, IL-1α, IL-4, IL-6, IL-6R, IL-10, IL-10R, IL-17 and IL-23 in the prediction of MM patients. In more details, 2 studies had identified the relationship between the SNP of IL-1α/β (22,36), one study of IL-1α (17,20,23-25,33,34,36,38,40-42). Regarding the NOS methodological quality, all included studies were of high quality with ≥7 out of 10 (mean 8.02 point).

The genotype susceptibility of MM and interleukin family gene polymorphisms are shown in Table SI. In Total, 16 published studies had detected interleukin polymorphism of IL-1β, IL-1α, IL-4, IL-6, IL-6R, IL-10, IL-10R, IL-17 and IL-23 in the prediction of MM patients. In more details, 2 studies had identified the relationship between the SNP of IL-1α/β (22,36), one study of IL-1α (17,20,23-25,33,34,36,38,40-42). Regarding the NOS methodological quality, all included studies were of high quality with ≥7 out of 10 (mean 8.02 point).

Quality assessment. All 16 selected papers were methodologically assayed by NOS and QUADAS-2 quality evaluation standards of the Cochrane Reviewers' Handbook. The detailed quality assessment of eligible studies, according to the NOS score, was summarized in Table SII. Overall, all studies included in the current meta-analysis were judged to be at moderate to high risk of bias, with scores ≥7 points (Table SII). The average NOS score was 8.01 out of 10, that was relatively classified in the high quality. Many studies provided sufficient information about study design and execution. Also, QUADAS-2 results confirmed that significant bias were not present in this meta-analysis. Fig. 2 shows all parameters of QUADAS-2 assessment, regarding bias risk and applicability concerns. Most studies had an acceptable range with regard to completeness of outcome data (attrition bias) and other sources of bias. More than half of the included studies were rated as low risk for most parameters of the bias risk (48.84%) and applicability concerns (62.5%). As shown in Fig. 2, no signification bias (Fig. 2A) and applicability concerns (Fig. 2B) were found in any of the selected studies.

The outcome of the meta-analysis. The present meta-analysis was performed in the both homozygous and heterozygous allele genetic model. Based on our systematic approach, we tried to find the associations between the MM risk and SNP of G:A, G:C and T:C in IL-1β, IL-1α, IL-4, IL-6, IL-6R, IL-10,

| First author | Year | Population (ethnicity) | Genotyping method | Sample size (case/control) | Source of control | Sex | NOS score | Refs. |
|--------------|------|------------------------|-------------------|--------------------------|------------------|-----|-----------|-------|
| Zheng        | 2000 | Sweden (C)             | PCR-RFLP          | 73/129                   | PB               | Not matched | 7     | (33)    |
| Zheng        | 2001 | Sweden (C)             | PCR-RFLP          | 73/109                   | PB               | Not matched | 7     | (34)    |
| Mazur        | 2005 | Poland (C)             | Taq-PCR           | 54/50                    | PB               | Matched     | 8     | (23)    |
| Cozen        | 2006 | USA (C, M)             | Taq-PCR           | 146/125                  | PB               | Matched     | 8     | (24)    |
| Duch         | 2007 | Brazil (C, M)          | Taq-PCR           | 52/60                    | PB               | Matched     | 9     | (41)    |
| Abazis-Stamboulieh | 2007 | Greece (C)            | Taq-PCR          | 74/160                   | HB               | Not matched | 8     | (22)    |
| Aladzisy     | 2009 | Hungary (C)            | Taq-PCR and PCR-RFLP | 100/99                   |                 |              | 8     | (35)    |
| Birmann      | 2009 | USA (C)                | Taq-PCR           | 82/164                   | PB               | Not matched | 8     | (20)    |
| Martino      | 2012 | Italy and Germany (C)  | Taq-PCR           | 201/234                  | PB               | Matched     | 9     | (38)    |
| Vangsted     | 2012 | Denmark (C)            | Taq-PCR           | 348/1,700                | PB               | Matched     | 8     | (36)    |
| Stephens     | 2012 | USA (C, M)             | Taq-PCR           | 626/44                   | PB               | Not matched | 8     | (42)    |
| Iakupova     | 2003 | Russia (C)             | Taq-PCR           | 69/102                   | PB               | Not matched | 9     | (25)    |
| Chakraborty  | 2017 | India (A)              | Taq-PCR           | 103/117                  | HB               | Matched     | 8     | (39)    |
| Kasamatsu    | 2017 | Japan (A)              | PCR-RFLP          | 128/202                  | PB               | Matched     | 9     | (40)    |
| Nielsen      | 2017 | Denmark (C)            | Taq-PCR           | 348/355                  | HB               | Not matched | 7     | (37)    |
| Kasamatsu    | 2018 | Japan (A)              | PCR-RFLP          | 120/201                  | PB               | Matched     | 8     | (17)    |

PCR, polymerase chain reaction; PCR-RFLP, polymerase chain reaction-restriction fragment length polymorphism; Taq-PCR, Taq man Real time polymerase chain reaction; PB, population-based; HB, hospital-based; C, Caucasian; A, Asian; M, Mixed Afro-Americans; NOS, Newcastle-Ottawa scale.
IL-10R, IL-17 and IL-23 polymorphisms. Also, the association between the type of each allele polymorphisms of G:A, G:C, T:C and ethnicity, genotyping methods, IL type and control reference were measured as subgroup analysis.

**G:A polymorphisms and MM susceptibility.** Table II shows the results of the meta-analysis for G:A and MM in the three different genotypes GG vs. AA, GG vs. AG and AA vs. GA. The combined analysis of 14 studies indicated that GG vs AA polymorphism was associated with a statistically significant improvement of 40.8% in MM, when compared with the control group (OR=1.14, 95% CI, 0.88-1.47, P<0.05); suggesting that the over-expression of GG vs AA polymorphism is a prognostic factor for MM (Fig. 3A). Also, the subtotal OR of GG vs AG and AA vs GA were 1.18 [95% CI, 0.94-1.4; P=0.27 (Fig. 3B)] and 0.98 [95% CI, 0.76-1.27; P=0.005 (Fig. 3C)], respectively. No significant coloration was found between IL-17Ars2275913 and IL-10Rαrs2228055 polymorphism (OR=0.64, 95% CI, 0.48-1.33, P=0.26 and OR=0.72, 95% CI, 0.62-1.83, P=0.43, respectively). Strikingly, the OR of GG vs AA was notably different compared with other polymorphisms. Subgroup analyses was conducted according to ethnicity, genotyping methods, IL type and control reference (Fig. S1). Furthermore, the G:A polymorphism detected in the IL-6 promoter (OR=1.05, 95% CI, 0.78-1.44) is more accurate in MM samples of the Asian population (OR=1.24, 95% CI, 0.92-1.74). The PCR-RFLP based methods are more valuable methods for the detection of the G:A polymorphism in MM samples (OR=1.18, 95% CI, 0.94-1.62, P=0.002).

**G:C polymorphisms and MM susceptibility.** As shown in Fig. 4, based on heterogeneity, the dominant model GG/CC has an appropriate effect model in the detection of the MM. The results in Fig. 4 clearly show that the GG genotype for the IL-6 promoter and IL-1β would increase MM risk by approximately 81% compared with the CC genotype. Correspondingly, G:C polymorphism for IL-1βI464G>C and IL-657G>C would increase MM risk (Table III). Meanwhile, the sub-analysis findings suggest that hospital-based samples were more accurate in the detection of the G:C polymorphism in MM patients.
Although, other subgroup analyses such as ethnicity, genotyping methods and IL type did not have any significant effect on MM prognosis and therefore they were ignored (Fig. S2). T:C polymorphisms and MM susceptibility. Data from 12 studies on IL-1, -6 and -10 polymorphisms were pooled and analyzed for the detection of the T:C polymorphisms and MM susceptibility (Table IV). A high risk of MM was found in the heterozygous model (Fig. 5A) and dominant model (Fig. 5B) of the T:C polymorphisms (TC/CC: OR=1.37, 95% CI, 0.88-2.16, P=0.001 and TT/CC: OR=1.07, 95% CI, 0.77-2.06, P=0.007). Moreover, sub-analysis results of the T:C polymorphisms revealed a significant correlation between IL-1β polymorphism (OR=1.18, 95% CI, 0.66-1.89, P=0.005) in hospital-based source samples (OR=1.82, 95% CI, 0.54-2.06, P=0.073) and MM risk (Fig. S3). There was no significant correlation between other subgroup analysis and MM risk in the T:C polymorphisms (data not shown).

C:A polymorphisms and MM susceptibility. Table V and Fig. 6 demonstrate the meta-analysis results of C:A polymorphisms. No significant association was identified between the overall MM risk and three polymorphism models, CC/AA, CC/AC and AC/AA in IL-6 (rs8192284) and IL-10 (rs1800872) receptors (P>0.05). In the stratified analyses by ethnicity, genotyping

| Allele          | Subgroup                | Number | study | OR   | 95% CI     | P-value |
|-----------------|-------------------------|--------|-------|------|------------|---------|
| GG vs. AA       | Ethnicity (C)           | 10     | 0.98  | (0.77-1.25) | 0.352 |
|                 | (A)                     | 3      | 1.17  | (0.82-1.68) | 0.465 |
| Method Taq      | 10                      | 1.17   | (0.80-1.70) | 0.017 |
| Method PCR      | 4                       | 1.17   | (0.84-1.63) | 0.675 |
| Source PB       | 12                      | 1.25   | (0.92-1.71) | 0.083 |
| Source HB       | 2                       | 0.86   | (0.63-1.17) | 0.398 |
| Gene Interleukin-6 promoter | 4 | 1.43 | (0.69-2.97) | 0.006 |
| Interleukin-6 receptor | 3 | 1.37 | (0.42-4.50) | 0.038 |
| Interleukin-10  | 3                       | 1.05   | (0.72-1.50) | 0.893 |
| Interleukin-10 receptor | 2 | 1.33 | (0.87-2.02) | 0.631 |
| Overall         | 14                      | 1.14   | (0.88-1.47) | 0.056 |
| GG vs. AG       | Ethnicity (C)           | 10     | 1.09  | (0.92-1.29) | 0.626 |
|                 | (A)                     | 3      | 1.41  | (1.03-1.92) | 0.960 |
| Method Taq      | 10                      | 1.03   | (0.84-1.26) | 0.202 |
| Method PCR      | 4                       | 1.37   | (1.03-1.81) | 0.956 |
| Source PB       | 12                      | 1.16   | (0.95-1.41) | 0.223 |
| Source HB       | 2                       | 1.00   | (0.78-1.27) | 0.504 |
| Gene Interleukin-6 promoter | 4 | 0.84 | (0.62-1.14) | 0.150 |
| Interleukin-6 receptor | 3 | 1.53 | (1.03-2.26) | 0.767 |
| Interleukin-10  | 3                       | 1.12   | (0.83-1.50) | 0.949 |
| Interleukin-10 receptor | 2 | 1.36 | (0.92-2.02) | 0.949 |
| Overall         | 14                      | 1.11   | (0.94-1.43) | 0.273 |
| GA vs. AA       | Ethnicity (C)           | 11     | 1.08  | (0.88-1.32) | 0.373 |
|                 | (A)                     | 3      | 1.18  | (0.83-1.67) | 0.318 |
| Method Taq      | 11                      | 0.89   | (0.62-1.26) | 0.002 |
| Method PCR      | 4                       | 1.15   | (0.85-1.55) | 0.492 |
| Source PB       | 13                      | 0.92   | (0.66-1.29) | 0.002 |
| Source HB       | 2                       | 1.16   | (0.88-1.53) | 0.732 |
| Gene Interleukin-6 promoter | 5 | 0.60 | (0.30-1.180) | 0.001 |
| Interleukin-6 receptor | 3 | 1.14 | (0.50-2.90) | 0.077 |
| Interleukin-10  | 3                       | 1.07   | (0.77-1.50) | 0.972 |
| Interleukin-10 receptor | 2 | 1.03 | (0.71-1.49) | 0.631 |
| Overall         | 15                      | 0.98   | (0.76-1.27) | 0.005 |

GG, AG, AA, homozygotes for the common allele, heterozygotes, and homozygotes for the rare allele, respectively. Controls deviate from Hardy-Weinberg equilibrium (χ², P<0.05). Effect after correction for Hardy-Weinberg equilibrium deviation. C, Caucasian; A, Asian; PCR, polymerase chain reaction; Taq, Taqman-polymerase chain reaction; PB, population-based; HB, hospital-based.
Figure 3. Forest plot of the association between G:A gene polymorphism and MM risk in the three difference genotype models: (A) GG vs. AA, (B) GG vs. AG, and (C) AA vs. GA model. The size of each square is proportional to the percentage weight that each study contributed to the pooled odds ratio. Weights are from random effects analysis. OR, Odds ratio; CIs, confidence intervals.

Figure 4. Forest plot of the association between G:C gene polymorphism and MM risk in the three difference genotype models: (A) GC vs. CC, (B) GC vs. CC, and (C) GC vs. GG model. The size of each square is proportional to the percentage weight that each study contributed to the pooled odds ratio. Weights are from random effects analysis. OR, Odds ratio; CIs, confidence intervals.
methods, IL type and control reference no significant association were detected between C/A polymorphism and MM (Fig. S4). In the overall analysis, no significant association was detected between T:A and T:G polymorphism and MM under all three genetic models (data not shown).

Publication bias. Begg's and Egger's tests were used to estimate the publication bias of each allele polymorphism. The analysis was carried out by precluding a single study at a time (Fig. 7) (43). The shape of funnel plot and Egger's test provided no statistical evidence for publication bias of the G:A \([t=-0.92, \ P=0.38, 13\text{ study (Fig. 7A)}]\), G:C \([t=-2.02, \ P=0.0.069, 14\text{ study (Fig. 7B)}]\), TC \([t=-1.51, \ P=0.162, 12\text{ study (Fig. 7C)}]\) and G:A \([t=-1.23, \ P=0.31, 5\text{ study (Fig. 7D)}]\). Likewise, Table SIII shows detailed publication bias of all investigated allele in three different models. P-values were revealed in Table SIII and no publication bias was found. Hence, there is no noticeable evidence for significant publication bias in our meta-analysis, which signifies that our meta-analysis results were stable and credible.

Discussion

To the best of our knowledge, this is the first comprehensive systematic review and meta-analysis study that has been conducted to identify the prognostics accuracy of all interleukin family gene polymorphisms in advanced MM patients. Data were extracted from a total of 16 publications that contained individuals carrying the GG genotype for IL-6 and IL-1 with an increased risk of MM by approximately 40.8 and 80.2% compared with the AA and CC genotypes. The results implied that a high MM risk was similar for IL-1 as well as IL-6; and revealed a significant association between their T:C polymorphism and MM risk. Furthermore, this study showed that patients with the heterozygous and dominant genotype of the T:C polymorphism have the highest MM risk. Notably, no significant association was found between the MM risk and C:A polymorphism in the IL-6 (rs8192284) and IL-10 (rs1800872) receptors (Fig. 6). Furthermore, hospital-based samples detected by PCR-RFLP methods are more attributable in identifying the interleukin family gene polymorphisms in MM patients. Keeping in mind the mentioned background, we tried to find significant correlations between ethnicity, IL type and MM sensitivity. We examined an overall sub-analysis of 2,597 patients and ORs for MM. Our results indicated that there was no significant association between IL gene polymorphism of the myeloma samples, ordered by ethnicity and type of IL family; which adversely affect cancer survival. Remarkably, we could not find any evidence about MM samples associated with sex and IL type. Our results clearly show that GG genotype for IL-6 promoter and IL-1β would increase MM risk by approximately 81% compared with the CC genotype \((R=1.02, 95\% \ CI, 0.62-1.42, P=0.001)\).

It is well established that interleukin family polymorphisms may be a prognostic factor for survival in MM patients (20-23). Single-nucleotide polymorphisms in the promoter region of

| Allele | Subgroup | Number study | OR    | 95% CI | P-value |
|--------|----------|--------------|-------|--------|---------|
| GG vs. CC | Ethnicity (C) | 9 | 1.21 | (0.50-2.93) | 0.001 |
|         | (C, M) | 3 | 1.17 | (0.74-1.86) | 0.96 |
|         | Method Taq | 11 | 1.21 | (0.57-2.57) | 0.001 |
|         | Source PB | 10 | 1.19 | (0.51-2.80) | 0.001 |
|         | HB | 3 | 1.25 | (0.88-1.78) | 0.89 |
|         | Gene Interleukin-6 promoter | 11 | 1.24 | (0.97-1.58) | 0.9 |
|         | Overall | 13 | 1.16 | (0.61-2.19) | >0.001 |
| GC vs. CC | Ethnicity (C) | 9 | 0.96 | (0.31-2.92) | 0.001 |
|         | (C, M) | 3 | 1.11 | (0.63-1.92) | 0.97 |
|         | Method Taq | 11 | 0.92 | (0.34-2.47) | 0.001 |
|         | Source PB | 10 | 0.94 | (0.30-2.93) | 0.001 |
|         | HB | 3 | 1.15 | (0.83-1.59) | 0.67 |
|         | Gene Interleukin-6 promoter | 11 | 1.18 | (0.94-1.48) | 0.97 |
|         | Overall | 13 | 0.96 | (0.41-2.24) | >0.001 |
| GG vs. GC | Ethnicity (C) | 9 | 0.77 | (0.45-1.33) | 0.001 |
|         | (C, M) | 3 | 0.92 | (0.63-1.34) | 0.96 |
|         | Method Taq | 11 | 0.74 | (0.48-1.13) | 0.001 |
|         | Source PB | 10 | 0.76 | (0.46-1.28) | 0.001 |
|         | HB | 3 | 0.90 | (0.69-1.18) | 0.58 |
|         | Gene Interleukin-6 promoter | 11 | 0.94 | (0.78-1.13) | 0.98 |
|         | Overall | 13 | 0.81 | (0.55-1.18) | >0.001 |

**Table III. Meta-analysis results for G:C gene polymorphism.**

GG, GC, CC, homozygotes for the common allele, heterozygotes, and homozygotes for the rare allele, respectively. Controls deviate from Hardy-Weinberg equilibrium \((\chi^2, P<0.05)\). Effect after correction for Hardy-Weinberg equilibrium deviation. C, Caucasian; M, Mixed Afro-Americans; PB, population-based; HB, hospital-based.
IL-1, -6 and -10 are associated with multiple inflammatory and immune-mediated diseases, including cancer, Crohn's disease, ulcerative colitis and type 1 diabetes (44,45). As a major pro-inflammatory cytokine, IL-6 and IL-1 could mediate chemokine, adhesion molecule expression, recruit monocytes and macrophages to release a large number of growth factors and cytokines (20). Many studies show that overexpression of the pro-inflammatory interleukin is a significant prognostic biomarker in patients with MM (25). In the present meta-analysis, the G:C polymorphisms (mainly GG genotype) in IL-6-572G>C and IL-6-174G>C are associated with an increased susceptibility to MM. Therefore, both tagging SNPs of IL-6rs1800795 and IL-6rs1800797 are most reliable and could be used for the prediction of MM progression (23,24). These SNPs located in the promoter region of the IL-6 gene may regulate IL-6 production and correlate with the susceptibility of some other cancers (23,25). Recently, Li et al., (2017) reported that IL-6 may be a potential biomarker for MM diagnosis (18). It suggested that IL-6 SNPs are very useful prognostic biomarkers in clinical usage and could be truly specific in prognosis of a particular cancer type (18). These results are completely consistent with other investigations (46-48).

IL-1α and IL-1β, two members of IL-1, have a common receptor with shared SNPs (IL-1α-889C>T and IL-1β-3737C>T) (49). Our results show that C:T polymorphisms of the IL-1α-889C>T and IL-1β-3737C>T were associated with MM risk. IL-10 is one of the well-investigated pro-inflammatory cytokines (34). It is documented that IL-10-592G/A and IL-10-1082G>A SNPs are located in the negative and positive regulatory sequence of the promoter region, respectively (34). The IL-10 gene with -592 (C) or -1082 (G) is associated with a high expression of IL-10, and it has a low expression with -592 (A) or -1082 (A). Our results indicate that the C allele of IL-10-592 has no significant association with MM susceptibility under the three allele model, CC/AA, CC/AC and AC/AA (18). In contrast, our cumulative results indicate that IL-10-1082G>A is found to be associated with MM risk (R=1.18, 95% CI, 0.94-1.3).

Recently, Sultana et al., (2018) (48) indicate that the C allele of IL-1α-889 is higher in MM patients than in controls. Similarly, Ziakas et al., (2013) (50) show that the G allele of IL-1β-1464G>C was significantly associated with MM in an Asian population. Of course, the discrepancies between the mentioned studies may also be due to small sample size and ethnic differences.

Despite these competent studies, the conventional formation and balancing of interleukin family gene polymorphisms in the interconnecting angiogenesis of the human MM neoplasms are not well-defined yet (45). Location variety and heterogenic morphology of MM tumors that have a close relationship with the interleukin family SNPs, represented noteworthy challenge for oncologist. With these presuppositions, the present investigation allows us to acquire a better understanding of the prognostic role of interleukin family gene SNPs in MM patients by using statistical approaches. Conversely, the correlation

### Table IV. Meta-analysis results for G:C gene polymorphism.

| Allele       | Subgroup | Number study | OR     | 95% CI   | P-value |
|--------------|----------|--------------|--------|----------|---------|
| TT vs. CC    | Source   |              |        |          |         |
|              | PB       | 7            | 0.95   | 0.74-1.23| 0.19    |
|              | HB       | 5            | 0.92   | 0.43-1.99| 0.002   |
| Gene         | Interleukin-1β | 4    | 0.91   | 0.67-1.25| 0.174   |
|              | Interleukin-1α | 2    | 0.49   | 0.09-2.58| 0.003   |
|              | Interleukin-6 receptor | 3   | 1.06   | 0.64-1.75| 0.161   |
|              | Interleukin-10 | 2    | 1.26   | 0.52-3.04| 0.280   |
| Overall      |          | 12           | 0.98   | 0.73-1.30| 0.007   |
| TT vs. TC    | Source   |              |        |          |         |
|              | PB       | 6            | 0.72   | 0.46-1.13| 0.025   |
|              | HB       | 5            | 2.39   | 1.46-3.90| 0.157   |
| Gene         | Interleukin-1β | 4    | 1.28   | 0.59-2.75| 0.001   |
|              | Interleukin-1α | 2    | 2.39   | 1.16-4.95| 0.214   |
|              | Interleukin-6 receptor | 3   | 0.90   | 0.24-3.33| 0.006   |
|              | Interleukin-10 | 2    | 0.99   | 0.34-2.89| 0.180   |
| Overall      |          | 12           | 1.26   | 0.77-2.06| 0.001   |
| TC vs. CC    | Source   |              |        |          |         |
|              | PB       | 6            | 0.768  | 0.63-0.93| 0.391   |
|              | HB       | 5            | 2.693  | 1.15-6.33| 0.001   |
| Gene         | Interleukin-1β | 4    | 1.342  | 0.71-2.54| 0.001   |
|              | Interleukin-1α | 2    | 5.181  | 1.99-13.43| 0.078   |
|              | Interleukin-6 receptor | 3   | 0.81   | 0.37-1.78| 0.090   |
|              | Interleukin-10 | 2    | 0.851  | 0.63-1.14| 0.001   |
| Overall      |          | 11           | 1.37   | 0.88-2.13| 0.001   |

TT, TC, CC, homozygotes for the common allele, heterozygotes, and homozygotes for the rare allele, respectively. Controls deviate from Hardy-Weinberg equilibrium ($\chi^2$, P<0.05). Effect after correction for Hardy-Weinberg equilibrium deviation. PB, population-based; HB, hospital-based.
Table V. Meta-analysis results for C:A gene polymorphism.

| Allele            | Subgroup         | Number study | OR       | 95% CI       | P-value |
|-------------------|------------------|--------------|----------|--------------|---------|
| CC vs. AA         | Source           |              |          |              |         |
|                   | PB               | 4            | 1.79     | (1.17-2.75) | 0.462   |
|                   | HB               | 2            | 1.35     | (0.72-2.53) | 0.743   |
| Gene              | Interleukin-6 receptor | 3        | 2.11     | (1.29-3.46) | 0.730   |
|                   | Interleukin-10   | 3            | 1.26     | (0.76-2.10) | 0.759   |
| Ethnicity         | (C)              | 5            | 1.74     | (1.18-2.57) | 0.614   |
| Method            | Taq              | 4            | 1.80     | (1.17-2.75) | 0.462   |
| Overall           |                  | 6            | 1.64     | (1.15-2.33) | 0.666   |
| CC vs. AC         | Source           |              |          |              |         |
|                   | PB               | 4            | 1.56     | (0.80-3.04) | 0.062   |
|                   | HB               | 2            | 1.06     | (0.57-1.97) | 0.816   |
| Gene              | Interleukin-6 receptor | 3        | 2.05     | (1.28-3.28) | 0.373   |
|                   | Interleukin-10   | 3            | 0.95     | (0.49-1.84) | 0.211   |
| Ethnicity         | (C)              | 5            | 1.50     | (0.86-2.59) | 0.094   |
| Method            | Taq              | 4            | 1.56     | (0.80-3.04) | 0.062   |
| Overall           |                  | 6            | 1.40     | (0.87-2.25) | 0.107   |
| AC vs. AA         | Source           |              |          |              |         |
|                   | PB               | 4            | 0.93     | (0.72-1.21) | 0.362   |
|                   | HB               | 2            | 0.79     | (0.54-1.14) | 0.869   |
| Gene              | Interleukin-6 receptor | 3        | 0.94     | (0.66-1.34) | 0.7     |
|                   | Interleukin-10   | 3            | 0.82     | (0.59-1.16) | 0.228   |
| Ethnicity         | (C)              | 5            | 0.91     | (0.72-1.14) | 0.463   |
| Method            | Taq              | 4            | 0.93     | (0.72-1.21) | 0.362   |
| Overall           |                  | 6            | 0.89     | (0.72-1.09) | 0.578   |

AA, AC, CC, homozygotes for the common allele, heterozygotes, and homozygotes for the rare allele, respectively. Controls deviate from Hardy-Weinberg equilibrium ($\chi^2$, P<0.05). Effect after correction for Hardy-Weinberg equilibrium deviation. PB, population-based; HB, hospital-based; C, Caucasian; Taq, Taqman- polymerase chain reaction.

Figure 5. Forest plot of the association between T:C gene polymorphism and MM risk in the three difference genotype models: (A) TC vs. CC, (B) TT vs. CC, and (C) TT vs. TC model. The size of each square is proportional to the percentage weight that each study contributed to the pooled odds ratio. Weights are from random effects analysis. OR, Odds ratio; CIs, confidence intervals.
Figure 6. Forest plot of the association between C:A gene polymorphism and MM risk in the three difference genotype models: (A) CC vs. AA, (B) CC vs. AC, and (C) AC vs. AA model. The size of each square is proportional to the percentage weight that each study contributed to the pooled odds ratio. Weights are from random effects analysis. OR, Odds ratio; CIs, confidence intervals.

Figure 7. Contour-enhanced funnel plots for the detection of a publication bias of the G:A (A); G:C (B); T:C (C); and G:A (D). All enrolled 16 studies represent by each point for the specified association, individually. These plots indicate that some studies were in significant areas where P<0.01 (solid lines) and where P=0.01-0.05 (dashed lines); other studies were in the non-significant area (the area between the two dashed lines). Solid triangles refer to included studies and X's refer to filled studies. The vertical axis represents standard error of logarithmic HR and the horizontal axis represents the HRs limits. HR, hazard ratio.
between the interleukin gene SNPs and the survival of cancer patients are controversial or inconclusive. On the other hand, several published meta-analyses have been concerned with the evaluation of the dissimilarity in interleukin family gene polymorphisms related to the prognosis of cancer (20-25,51). The overall findings help us describe the possible prognostic role of the interleukin polymorphism in MM progress, and identify a novel therapeutic target of MM progression. We tried to include all the possible literature on the relationship between interleukin polymorphism and MM risk and to obtain the maximum sample size and comprehensive results. Although, it should be noted that many future studies are needed for the description of the molecular mechanisms, because at the moment the published proof is scant. Unavoidably, there are still some limitations: i) A few studies and a small sample size have demonstrated correlations between interleukin gene family polymorphism and MM prognosis; ii) we only included the papers in English language, while published papers in other languages, especially Chinese and Russian were ignored; iii) poor homogeneous distribution and population based on subgroup parameters existed in some studies; iv) the occurrence and development of MM are on the basis of many hereditary and environmental factors as well as the interaction between them, but these factors and the pathological states of MM were not considered in this meta-analysis; and v) the low quality of some included studies led to meta-analysis results that were based on unadjusted estimates, because some studies did not provide detailed information to calculate the adjusted estimates. Besides, a lot of confounding factors were not controlled or reported in biased statistical results. We like to point out that future well-accepted clinical studies with larger samples sizes, systematized protocols and more homogenized populations would be needed to fully investigate the prognostic potential of each SNP in interleukin gene family of MM patients.

In conclusion, the present meta-analysis shows that G:C polymorphism of the IL-1β1464G>C and IL-6572G>C would increase MM risk. However, it has been determined that there is a significant association between MM risk and C:T polymorphism in IL-1α-889C>T and IL-1β-3737C>T. Additionally, hospital-based source samples are more accurate and attributable in the detection of interleukin family gene polymorphisms in Asian patients with MM, by using the PCR-RFLP method. These SNP loci may be appropriate biomarkers for gene screening and an effective adjuvant therapeutic strategy in prognostics of MM patients.

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Availability of data and materials
The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.

Authors’ contributions
CH and MNS were in charge of the idea and study designation. II, SN, and CY searched and collected data. FL and SL performed data analysis. MNS and II wrote the manuscript and were in charge of language revision. SN edited the paper. All authors read and approved the final manuscript.

Ethics approval and consent to participate
The present meta-analysis was approved by an independent Ethics Committee/institutional review board at Southwest Medical University, Department of Hematology (Luzhou, China). This investigation was carried out by following recommendations of the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA).

Patient consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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