Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.
Environmentally Transmitted Pathogens

Charles P. Gerba

22.1 ENVIRONMENTALLY TRANSMITTED PATHOGENS

Although humans are continually exposed to a vast array of microorganisms in the environment, only a small proportion of these microbes are capable of interacting with the host in such a manner that infection and disease will result. Disease-causing microorganisms are called pathogens. Infection is the process in which the microorganism multiplies or grows in or on the host. Infection does not necessarily result in disease since it is possible for the organism to grow in or on the host without producing an illness (see Chapter 29). In the case of enteric infections (i.e., diarrhea) caused by Salmonella, only half of the individuals infected develop clinical signs of illness. A frank pathogen is a microorganism capable of producing disease in either normal healthy or immunocompromised persons. Opportunistic pathogens are usually capable of causing infections only in immunocompromised individuals (burn patients, patients taking antibiotics, those with impaired immune systems, elderly patients with diabetes, etc.). Opportunistic pathogens are common in the environment and may be present in the human gut or skin, but normally do not cause disease.

To cause illness, the pathogen must usually first grow within or on the host. The time between infection and the appearance of clinical signs and symptoms such as diarrhea, fever or rash, is the incubation time (Table 22.1). This may range from as short as 6–12 h in the case of Norwalk virus diarrhea to 30–60 days for hepatitis A virus, which causes liver disease. At any time during infection, the pathogen may be released into the environment by the host through feces, urine, or respiratory secretions. Although the maximum release may occur at the height of the disease, it may also precede the first signs of clinical illness. In the case of hepatitis A virus, the maximum excretion in the feces occurs before the onset of signs of clinical illness. The concentration of organisms released into the environment varies with the type of organism and the route of transmission (Table 22.2). The concentration of enteric viruses during gastroenteritis may be as high as $10^{10}$–$10^{12}$ per gram of feces.

Pathogenic microorganisms usually originate from an infected host (either human or other animal) or directly from the environment. Many human pathogens can be transmitted only by direct or close contact with an infected person or animal. Examples include the herpesvirus, Neisseria gonorrhoeae (gonorrhea), and Treponema pallidum (syphilis).
This is because their survival time outside the host is very brief. Pathogens transmitted through the environment may survive from hours to years outside the host, depending on the organism and the environment. Pathogens may exit a host in respiratory secretions from the nose and mouth or be shed on dead skin or in feces, urine, saliva, or tears. Thus they may contaminate the air, water, food, or inanimate objects (fomites). When contaminated air is inhaled or food consumed, the organisms are effectively transmitted to another host, where the infection process begins again. Airborne transmission can occur via release from the host in droplets (i.e., coughing) or through natural (surf at a beach) or human activities (cooling towers, showers) (see Chapter 5). Some organisms may be carried great distances, hundreds of meters or miles (e.g., Legionnaires’ disease and foot-and-mouth disease).

Virus transmission by the airborne route may be both direct and indirect. Infection of a host may be by direct inhalation of infectious droplets or through contact with fomites on which the airborne droplets have settled. Hand or mouth contact with the organism on the surface of a fomite results in the transfer of the organism to the portal of entry, that is, nose, mouth, or eye.

Microorganisms transmitted by the fecal–oral route are usually referred to as enteric pathogens because they infect the gastrointestinal tract. They are characteristically stable in water and food and, in the case of enteric bacteria, are capable of growth outside the host under the right environmental conditions.

Waterborne diseases (Table 22.3) are those transmitted through the ingestion of contaminated water that serves as the passive carrier of the infectious agent. The classic waterborne diseases, cholera and typhoid fever, which frequently ravaged densely populated areas throughout

---

**TABLE 22.1 Incubation Time for Common Enteric Pathogens**

| Agent                | Incubation period | Modes of transmission                          | Duration of illness |
|----------------------|------------------|------------------------------------------------|--------------------|
| Adenovirus           | 8–10 days        | Fecal–oral–respiratory                          | 8 days             |
| *Campylobacter jejuni* | 3–5 days        | Food ingestion, direct contact                  | 2–10 days          |
| *Cryptosporidium*    | 2–14 days        | Food or water ingestion, direct and indirect contact | Weeks to months    |
| *Escherichia coli*   |                  |                                                 |                    |
| ETEC                 | 16–72 h          | Food or water ingestion                         | 3–5 days           |
| EPEC                 | 16–48 h          | Food or water ingestion, direct and indirect contact | 5–12 days          |
| EHEC                 | 72–120 h         | Food/ingestion, direct or indirect contact      | 2–15 days          |
| *Giardia lamblia*    | 7–14 days        | Food or water ingestion, direct and indirect contact | Weeks to months    |
| Norovirus            | 24–48 h          | Food or water ingestion, direct and indirect contact, aerosol? | 1–2 days          |
| Rotavirus            | 24–72 h          | Direct and indirect contact                     | 4–6 days           |
| Hepatitis A          | 30–60 days       | Hepatitis                                       | 2–4 weeks          |
| *Salmonella*         | 16–72 h          | Food ingestion, direct and indirect contact      | 2–7 days           |
| *Shigella*           | 16–72 h          | Food or water ingestion, direct and indirect contact | 2–7 days          |
| *Yersinia enterocolitica* | 3–7 days     | Food ingestion, direct contact                   | 1–3 weeks          |

**TABLE 22.2 Concentration of Enteric Pathogens in Feces**

| Organism                | Per gram of feces |
|-------------------------|-------------------|
| Protozoan parasites      | 10^6–10^7         |
| *Ascaris*               | 10^6–10^5         |
| Helminths               |                   |
| *Enteric viruses*       | 10^3–10^7         |
| Enteroviruses           | 10^7              |
| Rotavirus               | 10^10             |
| Adenovirus              | 10^11             |
| *Enteric bacteria*      | 10^4–10^10        |
| *Salmonella* spp        | 10^4–10^10        |
| *Shigella*              | 10^3–10^8         |
| Indicator bacteria      |                   |
| Coliform                | 10^7–10^9         |
| Fecal coliform          | 10^9              |
human history, have been effectively controlled by the protection of water sources and by treatment of contaminated water supplies. In fact, the control of these classic diseases illustrated the importance of water supply treatment and played an important role in the reduction of infectious diseases. Other diseases caused by bacteria, viruses, protozoa, and helminths may also be transmitted by contaminated drinking water. However, it is important to remember that waterborne diseases are transmitted by the fecal–oral route, from human to human or animal to human, so that drinking water is only one of several possible sources of infection.

**Water-washed diseases** are those closely related to poor hygiene and improper sanitation. In this case, the availability of a sufficient quantity of water is generally considered more important than the quality of the water. The lack of water for washing and bathing contributes to diseases that affect the eye and skin, including infectious conjunctivitis and trachoma, as well as to diarrhea illnesses, which are a major cause of infant mortality and morbidity in developing countries. Diarrheal diseases may be directly transmitted through person-to-person contact or indirectly through contact with contaminated foods and utensils used by persons whose hands are fecally contaminated. When enough water is available for hand washing, the incidence of diarrheal diseases has been shown to decrease, as has the prevalence of enteric pathogens such as *Shigella*.

**Water-based diseases** are caused by pathogens that either spend all (or essential parts) of their lives in water or depend on aquatic organisms for the completion of their life cycles. Examples of such organisms are the parasitic helminth *Schistosoma* and the bacterium *Legionella*, which cause schistosomiasis and Legionnaires’ disease, respectively.

**Water-related diseases**, such as yellow fever, dengue, filariasis, malaria, onchocerciasis, and sleeping sickness, are transmitted by insects that breed in water (e.g., mosquitoes that carry malaria) or live near water (e.g., the flies that transmit the filarial infection onchocerciasis). Such insects are known as *vectors*.

### 22.2 BACTERIA

#### 22.2.1 Salmonella

In the United States, the concept of waterborne and foodborne disease was poorly understood until the late nineteenth century. During the Civil War (1860–1865), encamped soldiers often disposed of their waste upriver but drew drinking water from downriver. This practice resulted in widespread dysentery. In fact, dysentery, together with its sister disease typhoid fever (*Salmonella typhi*), was the leading cause of death among soldiers of all armies until the twentieth century. It was not until the end of the nineteenth century that this state of affairs began to change. At that time, the germ theory was generally accepted, and steps were taken to treat wastes properly and protect drinking water and food supplies.

In 1890, more than 30 people out of every 100,000 in the United States died of typhoid. But by 1907, water filtration was becoming common in most U.S. cities, and in 1914 chlorination was introduced. Because of these new practices, the national typhoid death rate in the United States between 1900 and 1928 dropped from 36 to 5 cases per 100,000 people. The lower death toll was largely the result of a reduced number of outbreaks of waterborne diseases. In Cincinnati, for instance, the yearly typhoid rate of 379 per 100,000 people in the years 1905–1907 decreased to 60 per 100,000 people between 1908 and 1910, following the inception of sedimentation and filtration treatment of drinking water. The introduction of chlorination after 1910 decreased this rate even further.

*Salmonella* is a very large group of rod-shaped, gram-negative bacteria comprising more than 2000 known serotypes that are members of the family *Enterobacteriaceae*. All these serotypes are pathogenic to humans and can cause a range of symptoms from mild gastroenteritis to severe illness or death. *Salmonella* are capable of infecting a large variety of both cold- and warm-blooded animals. Typhoid fever, caused by *S. typhi*, and paratyphoid fever, caused by *S. paratyphi*, are normally found only in

### TABLE 22.3 Classification of Water-Related Illnesses Associated with Microorganisms

| Class         | Cause                                                                                     | Example                        |
|---------------|-------------------------------------------------------------------------------------------|--------------------------------|
| Waterborne    | Pathogens that originate in fecal material and are transmitted by ingestion                | Cholera, typhoid fever          |
| Water-washed  | Organisms that originate in feces are transmitted through contact because of inadequate sanitation or hygiene | Trachoma                        |
| Water-based   | Organisms that originate in the water or spend part of their life cycle in aquatic animals and come in direct contact with humans in water or by inhalation | Schistosomiasis, Legionellosis   |
| Water-related | Microorganisms with life cycles associated with insects that live or breed in water        | Yellow fever                    |
humans, although \textit{S. paratyphi} is found in domestic animals on rare occasions. In the United States, salmonellosis is due primarily to foodborne transmission because the bacteria infect beef and poultry and are capable of growing in these foods. Salmonellosis is the second leading cause of foodborne illness in the United States. Since the route of transmission is fecal–oral, any food or water contaminated with feces may transmit the organism to a new host.

It is estimated that 1.4 million cases of salmonellosis (diarrhea) occur in the United States annually (Voetsch et al., 2004). All age groups are susceptible, but symptoms are most severe in the elderly, infants, and the infirm. The onset time is usually between 12 and 36 h after ingestion of contaminated food or water. Intestinal disease occurs with the penetration of \textit{Salmonella} organisms from the gut lumen into the lining of the intestines, where inflammation occurs and an enterotoxin is produced. The immediate symptoms include nausea, vomiting, abdominal cramps, diarrhea, fever, and headache. Acute symptoms may last for one to two days or may be prolonged, again depending on host factors, and individual strain characteristics.

\textit{Salmonella typhi} and \textit{S. paratyphi} A, B, and C produce typhoid and typhoid-like (paratyphoid) fever in humans. Any of the internal organs may be infected. The fatality rate of untreated cases of typhoid fever is 10%, compared with less than 1% for most forms of salmonellosis. \textit{Salmonella} septicemia (bacteria multiplying in the blood) has been associated with the subsequent infection of virtually every organ system.

Typhoid fever presents a very different clinical picture than salmonellosis. The onset of typhoid fever (one to three weeks) is usually insidious, with fever, headache, anorexia, enlarged spleen, coughing, and constipation being more common than diarrhea in adults. Intestinal hemorrhage and perforation occur in about 1% of the cases. Paratyphoid fever presents a similar clinical picture but tends to be milder. A carrier state may follow infection (<1.0 to 3.9% of the population). A carrier is a person who is permanently infected and may transmit the organism, but does not demonstrate any signs or symptoms of disease. A chronic carrier state is most common among individuals infected during middle age, especially females. The organism is usually carried in the gallbladder and secreted in the bile.

### 22.2.2 \textit{Escherichia coli} and \textit{Shigella}

\textit{Escherichia coli} is a gram-negative rod found in the gastrointestinal tract of all warm-blooded animals and is usually considered a harmless organism. However, several strains are capable of causing gastroenteritis; among these are the enterotoxigenic (ETEC), enteropathogenic (EPEC), enteroinvasive (EIEC), or enterohemorrhagic (EHEC) \textit{E. coli}. They are grouped by their mechanisms of pathogenesis (Fig. 22.1). All of the \textit{E. coli} are spread by the fecal–oral route of transmission.

The enterotoxigenic \textit{E. coli} are a major cause of traveler’s diarrhea in persons from industrialized countries who visit less developed countries, and it is also an important cause of diarrhea in infants and children in less developed countries (Table 22.4). Following an incubation period of 10–72 h, symptoms including cramping, vomiting, diarrhea (may be profuse), prostration, and dehydration. The illness usually lasts less than three to five days. Disease is caused by two toxins, one the heat-labile toxin and the other the heat-stable toxin. The ETEC \textit{E. coli} are usually species specific; that is, humans are the reservoir for the strains causing diarrhea in humans. Only a few outbreaks have been documented in the United States. One resulted from the consumption of water contaminated with human sewage, another from the consumption of food prepared by an infected food handler, another in a hospital cafeteria, and the last aboard a cruise ship.

The EPEC is the oldest recognized category of diarrhea-causing \textit{E. coli}. Diarrheal disease caused by this group of \textit{E. coli} is virtually confined to infants less than one year of age. It is associated with infant summer diarrhea, outbreaks of diarrhea in nurseries, and community epidemics of infant diarrhea. Symptoms include watery diarrhea with mucus, fever, and dehydration. The diarrhea can be severe and prolonged with a high fatality rate (as high as 50%). Since the 1960s, EPEC has largely disappeared as an important cause of infant diarrhea in North America and Europe. However, it remains a major agent of infant diarrhea in South America, Africa, and Asia. Humans, cattle, and pigs can be infected by this organism. Thus, foods implicated in outbreaks are raw beef and pork.

The disease caused by \textit{EIEC} closely resembles that caused by \textit{Shigella}. Illness begins with severe abdominal cramps, watery stools, and fever. Disease is usually self-limiting with no known complications. Any age group is susceptible. This type of \textit{E. coli} carries a plasmid that enables it to invade cells lining the gastrointestinal tract, resulting in a mild form of dysentery. EIEC infections are endemic in less developed countries, although occasional
cases and outbreaks are reported in industrialized countries. Outbreaks also have been associated with hamburger and unpasteurized milk.

The EHEC *E. coli* were described in 1982 when a multi-state epidemic of hemorrhagic colitis occurred in the United States and was shown to be due to a specific serotype known as *E. coli* O157:H7. This organism produces two toxins called verotoxins I and II. These toxins are closely related to or identical to the toxin produced by *Shigella dysenteriae*. The toxin production depends on the presence of a prophage. A prophage is a bacterial virus that inserts its DNA into a bacterial chromosome. A plasmid codes for a novel type of fimbriae (hairlike projections) that enable the organism to adhere to the intestinal lining and initiate disease.

EHEC infections are now recognized to be an important problem in North America, Europe, and some areas of South America. The illness usually includes severe cramping and diarrhea, which is initially watery but becomes grossly bloody. The illness is usually self-limiting and lasts for an average of eight days. However, some victims, particularly the very young, develop hemolytic-uremic syndrome (HUS), resulting in renal failure and hemolytic anemia (Fig. 22.2). This disease can result in permanent loss of kidney function. In older individuals, HUS plus two other symptoms, fever and neurological symptoms, constitutes thrombotic thrombocytopenic purpura (TTP). This illness can have a mortality rate in the elderly as high as 50%. Most outbreaks are associated with undercooked or raw hamburger. Raw milk, unpasteurized fruit juices, and vegetables contaminated with cow dung have also been implicated. Waterborne outbreaks involving both nondisinfected groundwater and recreational waters have also occurred.

*Shigella* is closely related to *E. coli*. Four species have been described: *S. dysenteriae*, *S. flexneri*, *S. boydii*, and *S. sonnei*. *S. dysenteriae* causes the most severe disease and *S. sonnei* causes the mildest symptoms. Fortunately, *S. sonnei* is the serotype most often found in the United States (Lee et al., 1991). *Shigella* very rarely occurs in animals. It is principally a disease of humans and rarely of other primates such as monkeys and chimpanzees. The organism is often found in water polluted with human sewage and is transmitted by the fecal-oral route. *Shigella* has been a common cause of waterborne recreational outbreaks in the United States. However, most cases of shigellosis are the result of person-to-person transmission through the fecal-oral route. Secondary attack rates are 20–40% of household contacts. An estimated 440,000 cases of shigellosis occur annually in the United States (Gupta et al., 2004). *Shigella* is associated with certain foods such as salads, raw vegetables, milk and dairy products, and poultry. After an onset time of 12 h to one week, symptoms of abdominal pain, cramps, diarrhea, and vomiting may occur. The organisms multiply in the cells of the gastrointestinal tract and spread to neighboring cells, resulting in tissue destruction. Some infections are associated with ulceration, rectal bleeding, drastic dehydration, and fatality rates as high as 10–15%. Infants, the elderly, and the infirm are most susceptible.

### 22.2.3 Campylobacter

*Campylobacter jejuni* is a gram-negative curved rod. It is relatively fragile and sensitive to environmental stress.

| Pathogenic E. coli | Site of infection | Associated disease | Incidence | Target population | Significant transmission route |
|--------------------|-------------------|-------------------|-----------|-------------------|-------------------------------|
| ETEC               | Small intestine   | Travelers’ diarrhea, chronic childhood diarrhea (in developing countries) | 16 U.S. outbreaks (1996–2003); prevalence 1.4% in patients with diarrhea; 79,420 cases of travelers’ diarrhea each year (in the U.S.) | International travelers and children in developing countries | Food (raw produce, street vendors) and water |
| EPEC               | Small intestine   | Infant diarrhea   | Hundreds of thousands of deaths worldwide | Children in developing countries | Water, infant formula |
| EHEC               | Large intestine   | Hemorrhagic colitis, HUS | 110,000 cases and 61 deaths annually in the U.S. | All ages | Food (beef, produce), person-to-person, water, animals |
| EIEC               | Large intestine   | Dysentery         | Low in developed countries | Children in developing countries | Water (rare), person-to-person |

**TABLE 22.4 Summary of Pathogenic E. coli Incidence and Epidemiology**
(including an oxygen content of 21%, drying, heating, contact with disinfectants, or acidic conditions). Before 1972, when methods were developed for its isolation from feces, it was believed to be primarily an animal pathogen causing abortion and enteritis in sheep and cattle. Recent surveys have shown that *C. jejuni* is a leading cause of bacterial diarrheal illness in the United States, with an estimated 2,000,000 cases per year (Samuel et al., 2004). Although *C. jejuni* is not carried by healthy individuals in the United States or Europe, it is often isolated from healthy cattle, chickens, birds, and even flies. It is sometimes present in nonchlorinated water sources such as streams and ponds.

*Campylobacter jejuni* infections cause diarrhea with fever, abdominal pain, nausea, headache, and muscle pain. The illness usually occurs 2–5 days after the ingestion of the contaminated food or water. Illness generally lasts 7–10 days, and relapses are not uncommon. Although anyone can be infected with *C. jejuni*, children under five years of age and young adults (15–29 years old) are more frequently afflicted than other age groups.

Surveys show that 20 to 100% of retail chickens are contaminated with *C. jejuni*. This is not surprising because many healthy chickens carry these bacteria in their intestinal tracts. Non-chlorinated water may also be a source of infection. However, properly cooking chicken, pasteurizing milk, and chlorinating water will kill the bacteria (Blaser et al., 1986).

*Campylobacter jejuni* has been isolated from 22% of coastal and estuary water samples in concentrations ranging from 10 to 230 campylobacters per 100 ml and from 28% of river samples in concentrations of 10 to 36 cells per 100 ml. Carter et al. (1987) found *Campylobacter* in 10–44% of pond water samples. However, it is thought that virtually all surface waters contain *Campylobacter*. Recovery rates from surface waters are highest in the fall and winter months and lowest during the spring and summer months. *Campylobacter* density does not show a significant correlation with the isolation of indicator bacteria such as fecal or total coliforms (Carter et al., 1987).

This bacterium dies off rapidly in stream water at 37°C, showing a 9 log decrease within 3–12 days (Rollins and Colwell, 1986). However, the organism can remain viable for extended periods at cooler temperatures, surviving over 120 days at 4°C in stream water.

### 22.2.4 Yersinia

*Yersinia enterocolitica* and *Y. pseudotuberculosis* are small rod-shaped gram-negative bacteria. Both organisms have
often been isolated from animals such as pigs, birds, beavers, cats, and dogs. Only *Y. enterocolitica* has been detected in environmental and food sources such as ponds, lakes, meats, ice cream, and milk. To date, no foodborne outbreaks caused by *Y. pseudotuberculosis* have been reported in the United States, but human infections transmitted via contaminated water and foods have been reported elsewhere (Nuorti et al., 2004).

Symptoms usually begin 24 to 48 h after ingestion of contaminated food or drink. Yersiniosis is frequently characterized by such symptoms as gastroenteritis with diarrhea and/or vomiting. However, fever and abdominal pain are the hallmark symptoms. Yersinia infections mimic appendicitis, but the bacteria may also cause infections of other sites such as wounds, joints, and the urinary tract. Yersiniosis is more common in Northern Europe, Scandinavia, and Japan than in the United States. The greatest incidence of disease is seen during the cold season.

Although strains of *Y. enterocolitica* can be found in meats, oysters, fish, and raw milk, the exact cause of food contamination is unknown. However, the prevalence of this organism in soil and water and in animals offers ample opportunities for it to enter our food supply. Poor and improperly sanitized techniques of food handlers, including improper storage, cannot be overlooked as contributing to contamination. In addition, *Y. enterocolitica* is able to grow at refrigeration temperatures and under microaerophilic conditions posing an increased risk if uncooked meat stored in evacuated plastic bags is undercooked.

Waterborne outbreaks that have occurred in the United States but such documented outbreaks are very rare. One of these outbreaks occurred at a Montana ski resort from December 6, 1974, to January 17, 1975 (Craun, 1979). Approximately 750 cases of gastroenteritis caused by *Y. enterocolitica* occurred among 1550 guests and 350 employees. A significant association was found between drinking water and illness. Two 60-foot-deep wells developed in sand and gravel supplied water to the resort. A sewer line was found to pass near the wells, and samples collected from the wells after the outbreak yielded *Y. enterocolitica* and coliform organisms. Routine bacteriological surveillance during the previous three years had not detected coliform contamination of the wells. Chlorination of the wells stopped the outbreak.

### 22.2.5 *Vibrio*

London’s Dr. John Snow (1813–1858) was one of the first to make a connection between certain infectious diseases and drinking water contaminated with sewage. In his famous study of London’s Broad Street pump, published in 1854, he noted that people afflicted with cholera were clustered in a single area around the Broad Street pump, which he determined was the source of the infection. When, at his insistence, city officials removed the handle of the pump, Broad Street residents were forced to obtain their water elsewhere. Subsequently, the cholera epidemic in that area subsided.

The gram-negative genus *Vibrio* contains more than one member that is pathogenic to humans. The most famous member of the genus is still *V. cholerae*. Cholera is transmitted through the ingestion of fecally contaminated food and water. Cholera remains prevalent in many parts of Central America, South America, Asia, and Africa.

*Vibrio cholerae* serogroup O1 includes two biovars, cholerae (classical) and El Tor, each of which includes organisms of the Inaba and Ogawa serotypes. A similar enterotoxin is elaborated by each of these organisms, so the clinical pictures are similar. Asymptomatic infection is much more common than disease, but mild cases of diarrhea are also common. In severe untreated cases, death may occur within a few hours and the fatality rate without treatment may exceed 50%. This is due to a profuse watery diarrhea referred to as rice-water stools. The rice-water appearance is due to the shedding of intestinal mucosa and epithelial cells. With proper treatment, the fatality rate is below 1%. Humans are the only known natural host. Thus, the reservoir for *V. cholerae* is human, although environmental reservoirs may exist, apparently in association with copepods or phytoplankton.

Vibrios that are biochemically indistinguishable, but do not agglutinate in *V. cholerae* serogroup O1 antisera, were formerly known as nonagglutinable vibrios (NAGs) or noncholera vibrios (NCVs). They are now included in the species *V. cholerae*. Some of these strains produce enterotoxin but most do not. Thus reporting of non-O1 *V. cholerae* infections as cholera is inaccurate and leads to confusion. *V. mimicus* is a closely related species that can cause diarrhea, of which some strains elaborate an enterotoxin indistinguishable from that produced by *V. cholerae*.

The following example illustrates the possible devastating impacts of this disease. A cholera epidemic caused by *V. cholerae* O1 began in January 1991 in Peru and spread to Central America and South America. A total of 1,041,422 cases occurred with 9642 deaths (MMWR, 1995). The epidemic was believed to have been initiated by inadequate chlorination of drinking water. A second example is an outbreak in southern Asia, an epidemic caused by the newly recognized strain *V. cholerae* O139, which began in late 1992 and continued to spread to at least 11 countries. This latter strain of *V. cholerae* also produces severe watery diarrhea and dehydration that is indistinguishable from the illness caused by *V. cholerae* O1.

*Vibrio cholerae* is a native marine organism and its potential for transmission to humans is related to a complex ecology that controls its occurrence and concentration in the marine food chain (Lipp et al., 2002). Warmer temperatures in combination with elevated pH and plankton blooms can influence *V. cholerae* attachment, growth, and multiplication in the aquatic environment, particularly in association with copepods (Fig. 22.3). Thus, factors such
as climate change, climate variability (El Niño/Southern Oscillation), and monsoons can influence the *V. cholerae* concentration in marine waters and exposure via the marine food chain to humans (see also Ch. 18).

Another species, *V. parahaemolyticus*, is usually transmitted by contaminated food. The ingestion of inadequately cooked seafood or any food cross-contaminated by handling raw seafood or by rinsing with contaminated seawater may transmit this disease. A period of time at room temperature is generally necessary to allow multiplication of the organisms. Disease symptoms include watery diarrhea and abdominal cramps in the majority of cases, sometimes with nausea, vomiting, fever, and headache. Usually the disease is self-limiting and lasts one to seven days. Most cases are reported during the warmer months, and marine coastal environments are a natural habitat. The organisms have been found in marine silt, in coastal waters, and in fish and shellfish.

Of all foodborne infectious diseases, infection with *V. vulnificus* is one of the most severe; the fatality rate for *V. vulnificus* septicemia exceeds 50% (Tacket et al., 1984). Cases are most commonly reported during warm-weather months (April–November), and are often associated with eating raw oysters. Many patients are found to have had a preexisting liver disease, usually associated with alcohol use or viral hepatitis. All of these latter patients had eaten raw oysters one to two days before the onset of symptoms. These symptoms include thrombocytopenia, bulbous skin lesions, hypotension, and shock. Cases have also been reported that arose through the contamination of a wound by seawater or seafood drippings.

**22.2.6 Helicobacter**

In 1982, a physician in Australia cultured a gram-negative, spiral-shaped bacterium observed in biopsied tissue of a stomach ulcer. Initially called *Campylobacter pylori* based on biochemical and morphological characteristics, the organism is now named *Helicobacter pylori* (Fig. 22.4). The stomach mucosa contains cells that secrete proteolytic enzymes and hydrochloric acid. Other specialized cells produce a layer of mucus that protects the stomach itself from digestion. If this mucous layer is disrupted, the ensuing inflammation leads to an ulcer. *H. pylori* has been shown to bind to O– blood group antigens on the gastric epithelial cells. People of this blood group are twice as likely to develop gastric ulcers. *H. pylori* produces large amounts of urease, an enzyme that converts urea to the alkaline product ammonia. This results in a high pH in the local region.
Humans are thought to be a natural host for \textit{H. pylori}. Person-to-person spread is probably the dominant source of transmission, although controversy exists over whether the fecal–oral or oral–oral route of transmission predominates. \textit{H. pylori} has been isolated from feces (Thomas \textit{et al.}, 1992) and detected in saliva (Nguyen \textit{et al.}, 1993). Infection may also be spread by ingesting contaminated food or water. Several studies have shown a greater risk of acquiring \textit{H. pylori} infections from drinking untreated well water (Klein \textit{et al.}, 1991; Reavis, 2005; Rolle-Kampczyk \textit{et al.}, 2004). Nonculturable \textit{H. pylori} organisms have been detected in river water and groundwater in the United States (Heegarty \textit{et al.}, 1999).

### 22.2.7 Legionella

\textit{Legionella} is the causative agent of Legionnaires’ disease and Pontiac fever. Both syndromes are characterized initially by anorexia, malaise, myalgia, and headache. Within 24h, a fever ensues with chills. A nonproductive cough may occur and abdominal pain and diarrhea are seen in many patients. In Legionnaires’ disease, chest radiographs show areas of consolidation indicative of pneumonia and indeed, respiratory failure may occur. This disease has a 15% fatality rate in hospitalized cases. Pontiac fever is not associated with pneumonia or death. Patients recover spontaneously in two to five days.

\textit{Legionella} are poorly staining gram-negative bacilli that require cysteine and other specific nutrients when grown on artificial laboratory media. At least 40 species of \textit{Legionella} have been shown to cause disease, but the most prominent pathogenic species is \textit{L. pneumophila}, which first received extensive attention after an outbreak in 1976 in Philadelphia. The disease is found worldwide, with most sporadic cases occurring during the summer and fall months. The reservoir is primarily aquatic. Hot water systems, air-conditioning cooling towers, and evaporative condensers have all been implicated in outbreaks, as have decorative fountain and retail store misters (Fig. 22.5). The organism has also been

![FIGURE 22.5](A) Sources of \textit{Legionella} in the environment. (B) Cooling towers. Outbreaks of Legionnaires’ disease have been commonly traced to cooling towers.

(A)

(B)
found in creeks and ponds and the soil from their banks. The bacterium survives for months in tap and distilled water. The primary route of transmission is thought to be through the inhalation of aerosols. Exposure is not uncommon, as reflected in serologic assays that show that 1–20% of the general population have antibodies to \textit{L. pneumophila}.

Concentrations of $1.4 \times 10^3$ to $1.7 \times 10^5$ cells per liter of \textit{Legionella} spp. have been detected in raw drinking water sources using direct fluorescent antibody (DFA) techniques (Tison and Seidler, 1983). The concentrations of \textit{Legionella} found in distribution water samples by DFA were as follows (colony-forming units per liter): chlorinated water, $<8 \times 10^3$ to $1.4 \times 10^4$; water treated by slow sand filtration and chlorination, $<5.4 \times 10^3$ to $4.6 \times 10^3$; water treated by flocculation, filtration, and chlorination, $<8 \times 10^3$ to $2.2 \times 10^4$. Zacheus and Martikainen (1994) found that 30% of hot water systems in apartment buildings contained \textit{Legionella}. The mean number of \textit{L. pneumophila} was $2.7 \times 10^3$ CFU/l with a range of $<50$ to $3.2 \times 10^5$ CFU/l. For all positive hot water systems, \textit{Legionella} was also isolated from the hot water tap and shower head. \textit{Legionella pneumophila} was isolated from 12.5, 29.0, and 37.5% of the hot water distribution systems receiving chlorinated groundwater, unchlorinated groundwater, and chlorinated surface water, respectively.

A great deal of work has been carried out on the survival and growth of legionellae in potable water distribution systems and plumbing in hospitals and homes. Legionellae appear to be more resistant to chlorine than \textit{E. coli}. For example, \textit{Legionella} has been shown to exist in potable water systems even when exposed to 0.75 to 1.5 ppm free chlorine residual. Such protection is afforded by intracellular growth in protozoa such as \textit{Tetrahymena pyriformis} and \textit{Acanthamoeba castellani} (Fields et al., 1984; Moffat and Tompkins, 1992).

\textit{Legionella} survives well at 50°C and environmental isolates are able to grow in tap water at temperatures as high as 42°C (Fig. 22.6). The enhanced survival and growth in these systems have been linked to stagnation stimulated by rubber fittings in the plumbing system (Colbourne et al., 1988) and trace concentrations of metals such as iron, zinc, and potassium (States et al., 1989). Sediment promotes the growth of \textit{Legionella}, as does stagnation in the hot water tank. \textit{Legionella} can be removed from hot water heaters by raising the temperature to over 60°C near the heating element and to over 50°C at outlets, combined with regular flushing (Meenhorst et al., 1985).

The overall attack rate of pneumonia in the United States is 12–15 cases per 1000 persons per year, resulting in approximately 3,957,000 cases annually. Pneumonia is the sixth leading cause of death in the United States, with an estimated annual cost of $23 billion. \textit{L. pneumophila} causes 4.1 to 20.1% of community-acquired cases, many of which result in hospitalization (Marrie, 1994). These data suggest that \textit{L. pneumophila} is a major cause of serious cases of pneumonia.

### 22.2.8 Opportunistic Bacterial Pathogens

An opportunistic pathogen is one that usually causes disease only in those whose immune system is compromised. The weakened immune system may be due to very young or old age, pregnancy, cancer therapy, immunosuppressive drugs, human immunodeficiency virus (HIV), and other causes. Opportunistic pathogens are numerous in the environment. Many opportunistic bacterial pathogens are found in surface and drinking waters.

Concern has been generated in the drinking water industry regarding the health effects of heterotrophic bacteria that are found in tap water, bottled water, and other sources of potable water. Heterotrophic bacteria are those that require organic carbon rather than carbon dioxide as a carbon source. All human bacterial pathogens are heterotrophic.

Most of the heterotrophic bacteria in drinking water are not human pathogens. However, some of the genera, including \textit{Legionella}, \textit{Mycobacterium}, \textit{Pseudomonas}, \textit{Acinetobacter}, \textit{Stenotrophomonas}, and \textit{Aeromonas}, include species that are opportunistic pathogens. See Section 22.2.7 for a discussion of the genus \textit{Legionella}.

The most important opportunistic pathogen in the genus \textit{Pseudomonas} is \textit{P. aeruginosa}, which is primarily a nosocomial (hospital acquired) pathogen responsible for 9.9% of nosocomial infections. Human disease is often associated with water-related reservoirs such as swimming pools, whirlpools, hot tubs, and contact lens solutions. The source of community-acquired infections of cystic fibrosis patients has not been clarified. It is important as the ultimate cause of death in these patients is often lung infections by \textit{P. aeruginosa}. Although \textit{P. aeruginosa} is found in drinking water, it is not ubiquitous. Results of surveys showed that 2% of bottled water and 2–3% of tap water samples contain \textit{P. aeruginosa} at concentrations between 1 and 2300 organisms/ml (Allen and Geldreich, 1975).
Acinetobacter is the second most frequently isolated nonfermentative gram-negative rod in the clinical laboratory. However, it is generally considered to be of low virulence. Up to 25% of healthy adults carry this organism in the respiratory tract. The human skin is the likely source for most outbreaks of hospital infections. As an opportunistic pathogen, Acinetobacter is involved in nosocomial urinary tract infections, bacteremia, wound infections, and pneumonia. Acinetobacter can also be a cause of community-acquired pneumonia and urinary tract infections. Acinetobacter has been isolated from 97% of natural surface water samples with a concentration of 0.1–100 cells/ml (Baumann, 1968). Bifulco et al., (1989) isolated Acinetobacter from 38% of groundwater supplies at an arithmetic mean density of 8 CFU/100ml in the positive samples.

Acinetobacter has been isolated from 5 to 92% of distribution water samples (LeChevallier et al., 1980; Bifulco et al., 1989). It comprised 1.0 to 5.5% of the heterotrophic plate count (HPC) flora in drinking water samples (Payment et al., 1988) with concentrations of 6–21 CFU/ml. It has also been found in 5 to 35% of bottled water samples, at concentrations of 2–30 CFU/ml.

Pseudomonas maltophilia has been reclassified as Stenotrophomonas maltophilia and is the third most commonly isolated nonfermentative gram-negative rod in clinical laboratories. This organism can colonize the body and cause disease. Risk factors for infection include stays in intensive care units, mechanical ventilation, antibiotic treatment, and cancer. Diseases it can cause include septicemia, pneumonia, wound infections, and more rarely meningitis and endocarditis.

Stenotrophomonas maltophilia constituted 5.7% of the HPC found in raw surface water samples and 0–1.2% of the flora in distribution water samples (LeChevallier et al., 1980). Distribution water is drinking water that has exited the water treatment plant and is being distributed in pipes to the consumer. The bacterium has also been found in 2% of bottled water samples at concentrations of 2–22 CFU/ml and in <5% of cooler water samples at 34 CFU/ml.

Aeromonas hydrophila, A. sobria, and A. caviae are very similar and were all referred to as A. hydrophila until 1976, when they were first split into separate species. Papers written before 1985 may use the term A. hydrophila including all three species and biochemical variants. The three gram-negative species are biochemically very similar and have all been implicated as diarrheal agents in humans. The exact mechanism for diarrhea has not been elucidated.

A strong association has been found between drinking untreated water and the occurrence of diarrhea with the isolation of Aeromonas species. Higher counts of A. hydrophila in distribution water correlate with greater frequency of diarrheal isolates; however, no waterborne outbreaks have been documented.

Children and the elderly tend to be affected most often. As with many diarrheal agents, outbreaks of diarrhea, in which Aeromonas has been implicated, have been associated with day care centers. Aeromonas is frequently found in environmental water samples. It has been recovered from 0.6 to 18.2% of natural freshwater samples at a concentration range of 0.1–3600 CFU/ml. It has also been recovered from 0.9 to 27% of distribution water samples at an average concentration of 0.022 CFU/ml. However, even at large oral doses of up to 10^{10} CFU, A. hydrophila failed to produce diarrhea in human volunteers (Morgan et al., 1985).

The Mycobacterium avium complex (MAC) consists of 28 serovars of two distinct species, M. avium and M. intracellulare. It also included three serovars of M. scrofulaceum in the past, but its inclusion is no longer appropriate due to recent advances in mycobacterial systematics. Pulmonary disease caused by MAC is now as common as pulmonary tuberculosis in many areas of the United States. Evidence suggests the prevalence of this disease is growing. Predisposing factors include age, chronic lung disease, bronchogenic carcinoma, previous gastrectomy, and AIDS. MAC can also cause pulmonary disease, osteomyelitis, and septic arthritis in people with no known predisposing factors (Jones et al., 1995). In the United States, many infections are asymptomatic and occur early in life: 12% of the population has been infected by MAC (von Reyn et al., 1993a). However, disease by MAC can be lethal and is difficult to treat because of resistance to many antimycobacterial agents.

Despite extensive investigation, the precise mode of transmission of MAC remains undetermined. Infection with MAC is thought to occur from colonization of the gastrointestinal and/or respiratory tract (Chin et al., 1994). This suggests that exposure occurs by inhalation or ingestion.

Epidemiological investigations have associated water sources with infections by atypical mycobacteria (Burns et al., 1991). These bacteria can multiply in water that is oligotrophic (Falcao et al., 1993) or essentially free of nutrients, including dialysis water, and they are relatively resistant to disinfection by chlorination and chloramines (Collins et al., 1984). Atypical mycobacteria are widespread in water environments. Mycobacteria have been isolated from 11 to 38% of raw water samples at concentrations of <0.1–48 CFU/ml. It has also been found in up to 50% of municipal and private drinking water samples at concentrations of 0.01–5.2 CFU/ml.

Hospital water systems often harbor MAC and may be a source of nosocomial infection (von Reyn et al., 1994). M. avium has primarily been associated with hot water systems; von Reyn et al., 1993b). In some cases, a hot water system may be persistently colonized by the same strain of M. avium (von Reyn et al., 1994).

22.2.9 Blue-Green Algae

Blue-green algae or cyanobacteria occur in an enormous diversity of habitats, freshwater and marine, as plankton
(free floating), mats, and periphyton (attached to surfaces). Hot spring mats of some Oscillatoria develop up to temperatures of 62°C (Fig. 22.7). They have many beneficial functions such as nitrogen fixation and cycling of nutrients in the food chain.

Despite their beneficial roles in the environment, cyanobacteria sometimes become problematic. Occasionally, they increase rapidly resulting in cyanobacterial blooms (Fig. 22.8). Blooms are associated with eutrophic water, especially with levels of total phosphorus >0.01 mg/l and levels of ammonia- or nitrate-nitrogen >0.1 mg/l. Optimal temperatures for blooms are 15–30°C, and optimal pH is 6–9. Calm or mild wind conditions sometimes allow blooms to cover the water surface, but the highest concentrations of cyanobacteria may occur at depths ranging from 2 to 9 m, which will not be visible from the shore. The offending bacteria may also grow in the sediment. These blooms can impart an off-taste and odor to the water and/or result in the production of toxins.

The most common complaints related to such blooms are of taste and odor. Geosmin and 2-methylisoborneol (MIB) can produce odors at levels as low as 1.3–10 and 6.3–29 ng/l, respectively (Young et al., 1996). The odor produced by geosmin is described as earthy and that of MIB as musty or camphoraceous smelling. Concentrations of MIB and geosmin are usually highest in summer and fall. Several compounds produced by cyanobacteria can cause off-tastes and odors (Table 22.5).

Geosmin is produced by several cyanobacteria including Oscillatoria, Anabaena, Lyngbya, Phormidium, Symploca (Narayan and Nunez, 1974), Aphanizomenon, and Fischerella (Wu and Juttner, 1988). Lyngbya, Oscillatoria, and Phormidium (Izaguirre, 1992) are the most common genera producing MIB. Some strains of Diplocystis and Schizothrix can also cause off-tastes and odors. Microcystis release some odorous sulfur compounds, especially when they decay.

Many cyanobacteria found in algal blooms can produce toxins that cause liver damage, neural damage, and gastrointestinal (GI) disturbances. This has been well documented

![Figure 22.7](image1.png) Oscillatoria—a blue-green alga. Courtesy of Michael Clayton, 2006.

![Figure 22.8](image2.png) Cyanobacterial bloom. Courtesy of C.P. Gerba.

| Compound                  | Odor                  | Taste               |
|---------------------------|-----------------------|---------------------|
| Geosmin                   | Earthy, musty, grassy | Musty, earthy, stale|
| MIB                       | Musty, earthy, peaty  | Musty, earthy, stale|
| Isobutylmethoxypyrazine   | Woody, stale, musty   | Creosote, stale, dusty|
| Isopropylmethoxypyrazine  | Sooty, dusty, cabbage | Musty, vegetable water|
| Octa-1,3-diene            | Musty                 |                     |
| Hexanal                   | Green apple-like      |                     |
| Octan-1-ol                | Rancid                |                     |
| β-Cyclocitral             | Tobacco               |                     |
in many wild animal and livestock cases and implicated in human cases as well. *Microcystis* is the number one offender worldwide. Other toxin producing genera include *Anabaena, Aphanizomenon, Alexandrium, Cylindrospermopsis, Nodularia, Nostoc,* and *Oscillatoria* (Turner et al., 1990). Different types of toxins produced are shown in Tables 22.6 and 22.7. Most toxic species are associated with temperate rather than tropical climates.

In livestock and wild animals, the hepatotoxins cause weakness, anorexia, and liver damage. They can be lethal within minutes to a few days. Neurotoxins can cause twitching, muscle contraction, convulsions, and death. Signs and symptoms in humans associated with the ingestion of water with algal blooms are dizziness, headaches, muscle cramps, nausea, vomiting, gastroenteritis, and pneumonia (Phillip et al., 1992). Long-term exposure to toxins is associated with liver cancer as well (Carmichael, 1994).

In mice, the median lethal dose or LD$_{50}$ (with intraperitoneal injection) of toxins varies. The LD$_{50}$ of *Anabaena circinalis* neurotoxin is 36±1.4 mg/kg mouse (Falconer et al., 1989). The hepatotoxin of *Microcystis* had an LD$_{50}$ of 14.8±1.1 mg/kg mouse.

### 22.3 PARASITOLOGY

The study of parasitology embodies a large diversity of eucaryotic organisms. This group includes organisms that are unicellular, multicellular, and multinucleate; aerobic and anaerobic; motile and nonmotile; sexual and asexual. For this chapter parasites will be grouped into two categories: protozoa and helminths (Table 22.8). Protozoa are unicellular microorganisms in the kingdom Protista and are classified according to their means of locomotion: flagella, cilia, pseudopodia, or no locomotion. Some are parasitic, although the majority are nonparasitic. Some undergo a sexual stage, whereas others reproduce by asexual means: fission, budding, or schizogony. Helminths belong to the kingdom Animalia and include roundworms, flatworms, tapeworms, and flukes. They are multicellular, complex organisms containing organs and tissue. They usually develop in the soil or in intermediate hosts to complete their life cycle and have elaborate life cycles, including larvae, eggs, and adult stages. Parasitic protozoa and helminths can be acquired from the environment—water, soil, and contaminated food—and have had great health and economic impacts in many developing and developed countries. In the United States, there are two protozoa that are especially of concern: *Giardia* and *Cryptosporidium*. They form hardy cysts and oocysts that can survive water treatment disinfection and are one of the biggest concerns of water utilities today. Some of the characteristics of environmentally transmitted parasites are summarized in Table 22.9.

#### Table 22.6 Cyanobacterial and Types of Toxins Produced

| Genus       | Toxins produced                  |
|-------------|----------------------------------|
| *Anabaena*  | Anatoxin a, hepatotoxins         |
| *Aphanizomenon* | Saxitoxin, neosaxitoxin, hepatotoxins |
| *Alexandrium* | Saxitoxin                     |
| *Cylindrospermopsis* | Hepatotoxin      |
| *Nodularia* | Nodularins                      |
| *Oscillatoria* | Neurotoxins, hepatotoxins      |
| *Microcystis* | Microcystins                   |

#### Table 22.7 Characterization of Cyanobacterial Toxins

| Toxin       | Characterization of toxin      |
|-------------|--------------------------------|
| Anatoxins   | Neurotoxins                    |
| Microcystins| Hepatotoxins                   |
| Nodularins  | Hepatotoxins                   |
| Saxitoxins  | Neurotoxins                    |

#### Table 22.8 Classification of Some Environmentally Transmitted Protozoa and Helminths

**Protozoa**

- Phylum *Apicomplexa*
  - *Cyclospora cayetanensis*
  - *Cryptosporidium parvum*
  - *Toxoplasma gondii*
- Phylum *Microspora*
  - *Enteroctozena bieneusi*
  - *Encephalitozoon cuniculi*
  - *Encephalitozoon hellem*
  - *Encephalitozoon intestinalis*
- Phylum *Sarcocystigophora*
  - *Entamoeba histolytica*
  - *Giardia lamblia*
  - *Naegleria fowleri*

**Helminths**

- Phylum *Nematoda*
  - *Ascaris lumbricoides*
  - *Necator americanus*
  - *Trichuris trichiura*
- Phylum *Platyhelminthes*
  - Class *Cestoidea*
    - *Taenia saginata*
  - Class *Trematoda*
    - *Schistosoma mansoni*
22.3.1 Protozoa

22.3.1.1 *Giardia lamblia*

*Giardia lamblia* was first described in 1681 by Antonie van Leeuwenhoek, who found them in his own feces (Table 22.9). He called the trophozoites “animalcules.” In 1859, Vilerm Lambl rediscovered *Giardia* by finding the trophozoites in stools of young children with diarrhea. It was not until the early twentieth century that physicians began associating diarrhea with the presence of *Giardia* in stools. In 1954 Robert Rendtorff confirmed infectivity in human volunteers with oral administration of *Giardia* cysts.

*Giardia* is the most frequently identified intestinal parasite in the United States (Adam, 1991). In the United States, the prevalence of *G. lamblia* infections has been estimated to range from <1% in Midwestern middle-class...
adults (McHenry et al., 1987) to as high as 10–13% in Oregon adults (Skeels et al., 1986). In some settings, such as day care, the incidence can be as high as 33% in children (Ginsberg et al., 1994). In the United States, it is one of the most frequent identified causes of waterborne disease (Craun et al., 2006).

Humans become infected with *G. lamblia* by ingesting the environmentally resistant stage, the cyst (Figs. 22.9 and 22.10). Once ingested, it passes through the stomach and into the upper intestine. The increase in acidity via passage through the stomach stimulates the cyst to excyst, which releases two trophozoites into the upper intestine. The trophozoites attach to the epithelial cells of the small intestine. It is believed that the trophozoites use their sucking disks to adhere to epithelial cells. The adherence to the cells flattens the villi, causing malabsorption and diarrhea by not allowing adsorption of water and nutrients across the intestine. It can cause both acute
and chronic diarrhea within one to four weeks of ingestion of cysts resulting in foul-smelling, loose, and greasy stools. Once the trophozoites detach from the epithelial cells and travel down the intestine, cholesterol starvation is believed to stimulate the trophozoites to encyst and pass back into the environment as a cyst. Giardidiasis can be treated with metronidazole (Flagyl). Although in some cases the symptoms will spontaneously disappear without treatment, in most cases without treatment, the symptoms will wax and wane for many months. In symptomatic patients, more trophozoites than cysts are excreted into the feces, which cannot withstand the harshness outside the human body. In asymptomatic humans, mostly cysts are passed in stools; therefore Giardia carriers can serve as a source of cysts in the environment.

When Giardia cysts enter the environment they can survive for prolonged periods. G. lamblia cysts have been documented to survive for up to 77 days at 8°C and 4 days at 37°C in distilled water (Bingham et al., 1979). In another study, cysts of Giardia muris (a species that infects mice but is often used as a model for G. lamblia) were suspended in lake and river water and found to survive 28 days in lake water at a depth of 15 feet at 19.2°C. At a 30-foot depth at a temperature of 6.6°C, the cysts remained viable for 56 days.

It is still controversial whether animals such as beavers produce a G. lamblia strain infectious for humans. Studies have shown that beavers can be infected with G. lamblia from humans, but the reverse has not been demonstrated (Erlandsen et al., 1988). In one study 40 to 45% of beavers in Colorado were found to be infected with Giardia and shedding up to $1 \times 10^8$ cysts/animal/day, making them a major source of Giardia in the environment (Hibler and Hancock, 1990). Other animals that may contribute to Giardia in the environment are muskrats, where 95% of the population is infected. Various other animals found to be infected are cattle, goats, sheep, pigs, cats, and dogs (Erlandsen, 1995). To date, no infections with G. lamblia in humans have been directly linked to an animal host, but there is evidence which suggests that animal-source Giardia could potentially infect humans. Studies based on isoenzyme analysis and pulsed field gel electrophoresis (PFGE) banding patterns did not find a difference between cysts from beaver hosts and human hosts (Isaac-Renton et al., 1993).

### 22.3.1.2 Cryptosporidium parvum

Cryptosporidium was first described by Tyzzer in 1907 when he identified the organism in the intestinal epithelium of a mouse. It was not identified as a human pathogen until 1976, when it was described in the stools of immunocompromised hosts. Since that time, there have been numerous waterborne outbreaks, the most notable being the Milwaukee outbreak in April 1993, which infected over 400,000 people (MacKenzie et al., 1994) and killed more than 50 (Case Study 22.1).

Cryptosporidium parvum has a complex life cycle involving both sexual and asexual stages (Fig. 22.11). The host ingests sporulated oocysts (ranging from 3 to 6 μm in diameter) from contaminated water, food, or direct contact (Fig. 22.12). In the small intestine, the oocysts encyst, releasing four sporozoites, which attach to the epithelial cells of the mucosa. The sporozoite becomes enveloped by the microvilli, which fuse and elongate to cover the

---

**Case Study 22.1 Cryptosporidiosis in Milwaukee**

Early in the spring of 1993, heavy rains flooded the rich agricultural plains of Wisconsin. These rains produced an abnormal runoff into a river that drains into Lake Michigan, from which the city of Milwaukee obtains its drinking water. The city’s water treatment plant seemed able to handle the extra load: it had never failed before, and all existing water quality standards for drinking water were properly met. Nevertheless, by April 1, thousands of Milwaukee residents came down with acute watery diarrhea, often accompanied by abdominal cramping, nausea, vomiting, and fever. In a short period of time, more than 400,000 people developed gastroenteritis, and more than 100—mostly elderly and infirm individuals—ultimately died, despite the best efforts of modern medical care. Finally, after much testing, it was discovered that Cryptosporidium oocysts were present in the finished drinking water after treatment. These findings pointed to the water supply as the likely source of infection, and on the evening of April 7, the city put out an urgent advisory for residents to boil their water. This measure effectively ended the outbreak. All told, direct costs and loss of life are believed to have exceeded $150 million.

The Milwaukee episode was the largest waterborne outbreak of disease ever documented in the United States. But what happened? How could such a massive outbreak occur in a modern U.S. city in the 1990s? And how could so many people die? Apparently, high concentrations of suspended matter and oocysts in the raw water resulted in failure of the water treatment process—a failure in which Cryptosporidium oocysts passed right through the filtration system in one of the city’s water treatment plants, thereby affecting a large segment of the population. And among this general population were some whose systems could not withstand the resulting illness. In immunocompetent people, Cryptosporidiosis is a self-limiting illness; it is very uncomfortable, but it goes away of its own accord. However, in the immunocompromised, Cryptosporidiosis can be unrelenting and fatal.
Consumption of contaminated water or fecal-oral transmission are common routes of infection. Many animals especially cattle are reservoirs of infection. Excystation of oocysts occurs when the oocyst is expelled from the cell surface. Attachment of sporozoites to epithelial cells initiates the infection process. Type II meront (schizont) formation occurs, and four second-generation merozoites are released. These merozoites attach and form either micro or macrogametocytes. A micro and macrogamete join to form a zygote, which differentiates into a new oocyst. Sporozoite is enveloped by microvilli and matures into type I meront. A sexual reproduction results in the formation of eight merozoites which can reinfect or move into sexual reproduction. Oocyst can sporulate in the intestines and reinfect the host.

Within 3–10 days after ingestion of oocysts, nonbloody, voluminous watery diarrhea begins and lasts for 10–14 days in most immunocompetent hosts. There is no medical treatment and the disease is self-limiting; however, immunocompromised hosts (e.g., AIDS patients) can succumb, and about 10–15% of AIDS patients die of complications related to cryptosporidiosis. The prevalence of cryptosporidiosis in the United States is 0.3–4.3% (Ungar, 1990). The oocysts are very infectious and the presence of low numbers of oocysts in water or food poses a health threat (Messner et al., 2001).

Cryptosporidium oocysts can enter the environment via human and animal wastes. They have been found in marine environments.
water and bathing beaches in the vicinity of a nearby sewage outfall (Johnson et al., 1995). Cryptosporidiosis has been reported in many domestic animals, especially cattle. An infected calf can excrete $10^{10}$ oocysts per day. In a study by Kemp et al. (1995), farm drains were found to contain 0.06 to 19.4 oocysts per liter. This results in agricultural land runoff that can contaminate surface water.

Cryptosporidium parvum forms an extremely hardy oocyst that survives chlorine disinfection as commonly practiced at conventional water treatment plants. It has also been found to survive for weeks in surface waters (Johnson et al., 1997).

22.3.1.3 Entamoeba histolytica

Entamoeba histolytica was discovered in 1873 by F. Lösch in St. Petersburg, Russia, although its life cycle was not determined until Dobell did so in 1928. It causes amebic dysentery (bloody diarrhea) and is the third most common cause of parasitic death in the world. The world prevalence exceeds 500 million infections with more than 100,000 deaths each year. There are two sizes of cysts, small (5–9 μm) and large (10–20 μm), with each cyst producing eight trophozoites in the host. Only the larger cyst has been associated with disease; the smaller cyst tends to be associated with a commensal lifestyle (the organism benefits from the host, while the host is unaffected). About 2–8% of people infected develop invasive amebic dysentery in which the trophozoites actively invade the intestinal wall, bloodstream, and liver. It is unknown why this occurs. This organism is generally a problem in developing countries where sanitation is substandard and is transmitted via contaminated food and water. Humans are the main reservoir, although pigs, monkeys, and dogs have also been found to serve as reservoirs. No waterborne outbreaks have occurred in the United States for more than 30 years. Entamoeba is not as resistant to disinfectants as Giardia and Cryptosporidium.

22.3.1.4 Naegleria fowleri

Naegleria fowleri is an amoeboflagellate, changing between a cyst, amoeba, and flagellate with the amoeba stage dominant. The free-living protozoa are ubiquitous and found throughout the world in freshwaters (John, 1982). Cysts are usually present in low numbers, but when the water temperature exceeds 35°C (hot springs and warm stagnant waters), the amoeba transforms to the flagellated form quite rapidly, which enables the microorganism to swim. Infections are usually associated with children swimming in natural springs or warm waters although they are rare in the United States. The flagellate swims into the nose of a host and sheds its flagella (or it may be forced into the nose via diving). The amoeba then follows the nerves to the brain, producing a toxin that liquefies the brain. The organisms do not form cysts in the host. Primary amoebic meningoencephalitis (PAM) develops, causing severe, massive headaches. Death usually follows four to six days later. Diagnosis is most frequently postmortem on brain examination. There is treatment (amphotericin B) if the diagnosis is made quickly enough, although permanent brain damage may already have occurred.

22.3.1.5 Cyclospora sp.

Cyclospora cayetanensis is an emerging waterborne and foodborne protozoan pathogen. It was first identified in the intestine of a mole in 1870 by Eimer. It was not recognized as a human pathogen until the early 1980s, and then it was believed to be a cyanobacterium (Soave and Johnson, 1995). It is a coccidian protozoan in the phylum Apicomplexa. It produces a round spherical oocyst (Fig. 22.13) measuring $8 \times 10 \mu m$ which contains two sporozoites, each containing two sporozoites. Its sporulation life cycle is typical of a coccidian protozoan parasite. Desiccation of the organism kills it, and therefore it must be in an aquatic environment during maturation.

Cyclosporiasis causes voluminous, explosive, watery, nonbloody diarrhea in addition to abdominal cramps, nausea, and fatigue. Illness associated with Cyclospora averages 43 days in the immunocompetent host which can be compared to cryptosporidiosis that has a duration of only 10 to 14 days (Ortega et al., 1993). However, unlike cryptosporidiosis, it can be treated with Bactrim (trimethoprim–sulfamethoxazole), which appears to eliminate the parasite (Knight, 1995).

Acquisition of Cyclospora is not completely understood, as the host range and reservoirs are not known. However, water and food are believed to be a major route of infection, because for the organism to be infectious it must mature (sporulate) in the environment for two weeks (Ortega et al., 1993). Water can potentially become contaminated with sporulated oocysts, but most infections have

![FIGURE 22.13 Oocyst of Cyclospora cayetanensis. Magnification 100× under DIC. Courtesy H. Smith.](image-url)
been associated with contaminated produce. Outbreaks related to imported contaminated raspberries, snow peas, basil, and mesclun lettuce have been implicated in various outbreaks of cyclosporiasis in the United States (CDC, 2004). Two suspected waterborne outbreaks have been documented, one in Chicago and one in Nepal (Huang et al., 1995; Raebold et al., 1994). Infections have also been associated with individuals who live in or visit the Caribbean Islands, Central America and South America, Southeast Asia, and Eastern Europe (Knight, 1995).

22.3.1.6 Microsporidia

Microsporidia, the nontaxonomic name to describe organisms belonging to the phylum Microspora, were first described in 1857, when Nägeli identified Nosema bombycis, a microsporidium responsible for destruction of the silkworm industry. To date, over 1000 species of microsporidia infecting insects, invertebrates, and all five phyla of vertebrate hosts have been described. Microsporidia are for the most part considered to be opportunistic pathogens in humans. There were only a handful of documented cases before the advent of the AIDS epidemic. Since then there have been hundreds of documented cases in immunocompromised patients. However, there have also been cases documented among the immunocompetent. Five genera have been associated with the majority of human infections: Enterocytozoon bieneusi, Encephalitozoon hellem, Encephalitozoon cuniculi, Encephalitozoon intestinalis, Pleistophora spp., and Nosema corneum. The first four have the potential to be waterborne because they are shed in feces and urine. E. bieneusi, E. hellem, and E. intestinalis are the most common cause of microsporidian infections in patients with AIDS (Curry and Canning, 1993). In addition, they are much smaller (0.8 × 1.5 μm depending on species) than other parasites and potentially more difficult to remove by water treatment filtration.

The microsporidian spore has the potential of being transmitted by water. The life cycle of microsporidia contains three stages: the environmentally resistant spore, merogony, and sporogony. The spore is ingested by a host or possibly inhaled in some cases. Once in the body, it infects cells and goes through merogony followed by sporogony, which results in production of resistant infective spores (Fig. 22.14). The spores are then shed via bodily fluids such as urine and excreta. Once in the environment they have a strong potential to enter water sources. E. intestinalis spores have been identified in sewage, surface, and groundwaters, supporting the notion of environmental transmission. The spores are highly resistant to heat inactivation and drying. Waller (1979) found that E. cuniculi survived 98 days at 4°C and six days at 22°C.

Enterocytozoon bieneusi, Encephalitozoon hellem, Encephalitozoon cuniculi, and Encephalitozoon intestinalis cause a variety of illnesses. E. bieneusi causes diarrhea and wasting disease. It is the most important cause of microsporidiosis in AIDS patients. Several surveys have determined that 7 to 30% of AIDS patients who have unexplained chronic diarrhea are infected with E. bieneusi (Weber et al., 1994). E. intestinalis is similar to E. bieneusi in that it infects the intestines and causes diarrhea, but it can also infect kidneys and bronchial and nasal cells. It can infect macrophages, which allows it to disseminate throughout the body. It is secreted in feces and urine, which supports the notion of water transmission. E. cuniculi is not an intestinal parasite but, it can be shed in urine (Zeman and Baskin, 1985), and therefore environmental transmission is a possibility. It has also been described infecting many different mammals, which means that there could be many animal reservoirs that can contaminate the environment. E. hellem has been recognized and shown to cause eye infections (keratoconjunctivitis) and disseminated infections such as urteritis and pneumonia. It does not invade the intestine, but it can be shed in the urine (Schwartz et al., 1994).

22.3.1.7 Toxoplasma gondii

Toxoplasma gondii, the causative agent of toxoplasmosis, is an intestinal coccidium of felines with a very wide range of intermediate hosts. T. gondii causes a wide range of clinical conditions including brain damage in children, lymphadenopathy, ocular disease, and encephalitis. In the immunocompetent adult it causes symptoms that could be mistaken for influenza. It is estimated that 13% of the world’s population is infected with T. gondii (Hughes, 1985). Acquisition of the organism can be through contact with infected undercooked meat containing a bradyzoite (tissue cyst) and through contact with the environmental stage, the oocysts. The oocyst is excreted only in cat feces (domestic and wild) (Fig. 22.15). Congenital infection occurs when a pregnant woman becomes infected for the first time. Once infected, tachyzoites can cross the placenta and infect the fetus. Of the fetuses that become infected, 15% have severe complications. The oocysts can persist in soil and water, which can serve as a route of transmission. One study in England found that 100% of wild cats had antibodies to T. gondii (McOrist et al., 1991). Various studies of cats in the United States have found a seropositive prevalence of approximately 40% (Dubey and Beattie, 1988). The high prevalence of T. gondii in the cat population
demonstrates that they can be a significant source of oocysts in the environment. The oocyst can survive 18 months in the soil (−20 to 33°C) in Kansas (Frenkel et al., 1975) and over 410 days in water (Yilmaz and Hopkins, 1972). Its hardiness in the environment also suggests the possibility of environmental transmission.

To date, there have been only three well-documented drinking water outbreaks of toxoplasmosis. Several epidemiological studies have associated increase risk of infection with the consumption of unfiltered drinking water (Jones et al., 2005). *T. gondii* caused a waterborne outbreak in British Colombia, Canada, where more than 110 people including 12 newborns were infected after exposure to an unfiltered water supply (Mullens, 1996). In a later study of the area, four of seven domestic cats found in the watershed had antibodies to *T. gondii*, potentially being the initial source of contamination (Stephen et al., 1996).

To date, no one has studied the resistance of *T. gondii* to disinfectants, other than iodine. It survived for 3 h in 2% iodine (Dubey and Beattie, 1988). The cysts can be inactivated in meat by either freezing −12°C or cooking to an internal temperature of 67°C (Dubey, 1996).
Step 1. Egg containing developed juvenile is ingested

Step 2. Egg hatches in small intestine and penetrates the mucosa, entering the blood stream

Step 3. Larvae travel through blood stream to lungs, entering alveoli where they can cause pneumonitis. Once in the lung they migrate up the bronchioles to the trachea where they are swallowed. In the small intestines they develop into adults

Step 4. Adults mate and produce eggs in small intestine

Step 5. Egg passes in feces and develops in soil

FIGURE 22.16 Life cycle of *Ascaris lumbricoides*.

FIGURE 22.17 *Ascaris lumbricoides* ova. Photo courtesy P. Watt.

22.3.2 Nematodes

Nematodes are nonsegmented roundworms belonging to the phylum Nematoda. The majority of roundworms are free living in soil and fresh and salt water. The typical nematode has a flexible outer cuticle that protects the worm. They move via a muscular system and most lay eggs.

22.3.2.1 Ascaris lumbricoides

*Ascaris lumbricoides* is probably the most prevalent parasitic infection, with over 1.4 billion affected or about 22% of the world’s population. The prevalence is quite high in some regions of the world. Infection percentages range from 40 to 98% in Africa, 73% in Southeast Asia, 45% in Central America and South America, and 2% in the United States (Freedman, 1992; Kappus et al., 1994) of embryonated eggs (Figs. 22.16 and 22.17). There are no known animal reservoirs. The eggs are swallowed and the larvae hatch in the small intestine (Fig. 22.17). The larvae develop into second-stage larvae, which penetrate the lumen and enter into the bloodstream and capillaries. They travel via pulmonary circulation to the liver and heart, where the larvae develop into third-stage organisms that can lodge in the alveolar space (Fig. 22.18). This migration through the lungs can cause pneumonitis (Loeffler’s syndrome). The immature worms leave the lungs and travel through the bronchi, trachea, and epiglottis. They are then swallowed and arrive in the small intestine. There they undergo two molts, mate, and produce eggs. There is medical treatment against the adult intestinal worm: mebendazole or pyrantel pamoate. Symptoms usually correspond to the worm load, and a heavy
worm load can lead to intestinal blockage. The adult worm can reach more than 30 cm in size. Although most infections are mild, more than 20,000 people die annually with complications caused by intestinal blockage (Freedman, 1992).

An adult worm can produce more than 200,000 eggs per day. In 2–4 weeks after deposition in soil, they embryonate if the soil conditions are suitable (humid and warm) and are infectious. The eggs can survive months before embryonation if soil conditions are not appropriate. The eggs can survive freezing, chemicals, disinfectants, and sewage treatment. Because of their large size (35 × 55 μm), they accumulate in sewage sludge. This is especially a concern with sewage sludge-amended soils. In one study, 31–53% of eggs that had been deposited in soil were still viable 10 years later (Brudastov et al., 1971).

### 22.3.2.2 Trichuris trichiura

*Trichuris trichiura* is a worm that measures about 30–50 mm in length and is referred to as a whipworm as the worm’s shape resembles a whip. Incidence around the world ranges from 0.5 to 40% with the greatest being in developing countries. It is the third most common nematode infection. It is common in the southeastern United States as the weather conditions are ideal for egg survival in the soil (Fig. 22.19). The egg must be deposited in the soil and requires 21 days in moist, shady, warm soil to embryonate. In one study, 20% of ova deposited in soil were viable after 18 months (Burden et al., 1976). Infection occurs in humans via ingestion of contaminated water or soil. The worms can survive for years in a host, causing disease symptoms of diarrhea or constipation, anemia, inflamed appendix, vomiting, flatulence, and insomnia. The infection is diagnosed by identification of worms or eggs in the stool. The infection can be treated successfully with mebendazole. To prevent transmission, education on hand washing and sanitary feces disposal is necessary.

### 22.3.2.3 Necator americanus and Ancylostoma duodenale

There are two major hookworm species that infect humans: *Necator americanus* (New World hookworm) and *Ancylostoma duodenale* (Old World hookworm). The species are differentiated by mouth parts of adults and body size. There are no known reservoirs. They inhabit the small intestine and feed on intestinal mucosa and blood. They secrete an anticoagulant, causing great blood loss and anemia. They are the leading cause of iron deficiency in the tropics.

*Necator americanus* have round cutting teeth and are 7–10 mm in length. They lay an average of 10,000 eggs a day and enter humans via skin penetration. Each worm consumes 0.03 ml of blood per day. *A. duodenale* are larger (10–12 mm in length) and cause even greater blood loss. Each worm consumes 0.26 ml of blood per day. *A. duodenale* also lay more eggs (28,000) and are orally infective as well as able to penetrate the skin. In addition, *A. duodenale* have sharp cutting teeth.

The eggs embryonate once they are passed into the small intestine (Fig. 22.20). The eggs further develop into the rhabditiform larvae within 48 h in warm, moist sandy or loamy soil. The larvae feed, grow, and molt twice and then transform to filariform larvae. The filariform larvae do not eat. They seek out the highest point in the surroundings (e.g., top of grass blade) waiting for a host. On contact with skin, they penetrate the tissue and pass through a hair follicle or cut. They burrow through the subcutaneous tissue, then through the capillaries to the lungs. In the lungs they break out of the alveolar capillaries and migrate up the bronchi and trachea, where they are swallowed and enter the stomach and small intestine. They can live an average of five years but have been found to survive up to 15 years. The larvae can survive up to six weeks in moist, shady sandy or loamy soil. They do not survive well in clay soil, dry conditions, or at temperatures below freezing or greater than 45°C.
22.3.3 Cestodes (Taenia saginata)

Cestodes are tapeworms consisting of a flat segmented body and a scolex (head) containing hooks and/or suckers and grooves for attachment. The segments are called proglottids and pregnant segments are called gravids. The adults are parasitic and live in the intestinal lumen of many vertebrates.

*Taenia saginata* is transmitted by infected beef products and is the most common tapeworm found in humans. It is present in every country where beef is consumed (Fig. 22.21). Cattle become infected from eating grass or soil contaminated with human waste containing gravid proglottids. This occurs in areas where night soil (human waste) is used as fertilizer. The proglottids can survive in the environment for weeks. One study found that they could survive 71 days in liquid manure, 16 days in untreated sewage, and up to 159 days on grass (Jepsen and Roth, 1952). The eggs hatch in the duodenum, and hexacanths (tapeworm embryos containing six pairs of hooklets) are released. The hexacanths penetrate the mucosa, enter the intestine, and travel throughout the body. They then enter muscle and form a cysticercus (larval tapeworm enclosed in a cyst), which becomes infective in a few months. Humans become infected when they eat undercooked beef containing such cysticerci. The cysticerci can be inactivated at 56°C or by freezing at −5°C for 1 week (Schmidt and Roberts, 1989). Within 2–12 weeks the worm begins shedding gravid proglottids. The average worm length is 10–15 feet. Symptoms of infection are abdominal pain, headache, nausea, diarrhea, intestinal blockage, and loss of appetite (contrary to the belief that tapeworm infections cause an increase in appetite). The infection can be diagnosed by examining a gravid proglottid or scolex. The best methods for prevention are sanitation (proper disposal of human wastes) and thorough cooking of beef.

22.3.4 Trematodes (Schistosoma mansoni)

Trematodes, or flukes, are bilaterally symmetric worms that have two deep suckers and flame cells (for excretion). The suckers are used for both attachment and locomotion.
The life cycles are complex, with trematodes being either hermaphrodites (adults have both female and male gonads) or schistosomes (separate sexes). Trematodes require an intermediate host (snail) to complete their life cycle; the human is the definitive host, excreting eggs in the feces.

Three species of *Schistosoma* are medically important. In the past, *S. japonicum* and *S. haematobium* were the main causes of schistosomiasis, but now *S. mansoni* is recognized to be the most widespread of the three. The genus *Schistosoma* is responsible for more than 200 million infections worldwide and causes up to 200,000 deaths annually (Hopkins, 1992). More than 400,000 of those infected live in the United States (West and Olds, 1992). However, all these cases are imported as the intermediate host, the freshwater snail (*Biomphalaria* sp.), is not present in the United States. *S. mansoni* is distributed through most of Africa and the Middle East but is also found in parts of Central America and South America as well as some Caribbean Islands (Savioli *et al.*, 1997).

The larvae of schistosomes of bird and mammals can penetrate human skin, causing what is known as “swimmer’s itch.” These schistosomes do not mature in humans. This occurs in the Great Lakes region and along the coast of California.

The life cycle of *S. mansoni* is complicated, and human infection begins with the cercariae penetrating human skin. In addition, each parasitic stage involves different organs and thus different medical symptoms. The cercariae from infected snails are found in freshwater bodies. Once the cercariae penetrate the skin, they transform into the...
22.4 VIRUSES

22.4.1 Enteric Viruses

Viruses are a leading cause of gastroenteritis, in particular in infants and young children, in which they are a major cause of mortality worldwide. Four major groups of human gastroenteritis viruses have been identified: rotavirus, enteric adenovirus, caliciviruses (norovirus and sapporovirus), and astrovirus (Table 22.10). Of these, norovirus is of note because it has become the enteric virus most commonly associated with water- and foodborne illness worldwide. Although endemic viral gastroenteritis can be controlled in some areas; the Caribbean, excluding Puerto Rico, and Brazil. Strategies that have been successful are chemotherapeutics, health education, water supply treatment, and sanitation. Very little success has been achieved by eradicating the intermediate host, the snail, with either molluscicides or predator fish.

Diseases caused by enteric viruses range from trivial to severe or even fatal. The viruses that are detected most often in polluted water are the enteroviruses. However, recent studies suggested that adenoviruses occur in the greatest numbers of all viruses in wastewater. Waterborne outbreaks caused by enteric viruses are difficult to document because many infections by these agents are subclinical; that is, the virus may replicate in an individual, resulting in virus shedding but without overt disease. Therefore, an individual with waterborne infection but without overt disease may infect others, who in turn may become ill, spreading the infection throughout the community. In addition, epidemiological techniques lack the sensitivity to detect low-level transmission of viruses through water. Recreational activities in swimming pools have sometimes resulted in waterborne outbreaks caused by norovirus, hepatitis A virus, Coxsackie virus, echovirus, and adenoviruses. Enteric viruses from infected individuals may contaminate recreational waters by direct contact or by fecal release.

22.4.1.1 Astroviruses

Astroviruses were first observed by electron microscopy in diarrheal stools in 1975. These agents are icosahedral viruses with a starlike appearance and with a diameter of approximately 28 nm. Astroviruses have a single-stranded RNA (ssRNA) genome. The sequencing of the astrovirus genome allowed the establishment of its own virus family, the Astroviridae, with Astrovirus as the only genus. Serology assays and sequence analysis have resulted in the identification of seven distinct serotypes (Lee and Kurtz, 1994).

Astrovirus type 1 seems to be the most prevalent strain in children. Type 4 has been associated with severe gastroenteritis in young adults. Astrovirus-like particles have been found in feces of a number of animals suffering from a mild self-limiting diarrheal infection, but no antigenic cross-reactivity has been found between these agents and human astroviruses. Astrovirus infections occur throughout the year, with a peak during the winter–spring seasons in temperate zones. In warm climates, however, the highest incidence of astrovirus infection has been observed in May (Cruz et al., 1992).

Astroviruses cause a mild gastroenteritis after an incubation period of 3 to 4 days. Overt disease is common in
1- to 3-year-old children. However, adults and young children are also affected. Outbreaks of astrovirus infection have been associated with oysters and drinking water (Kurtz and Lee, 1987).

### 22.4.1.2 Adenoviruses

Adenoviruses are double-stranded DNA (dsDNA) icosahedral viruses approximately 70 nm in diameter with protruding spikes called pentons (Fig. 22.22). At least 50 human adenovirus types have been identified. Although there are many avian and mammalian adenovirus types, they are species specific. Adenoviruses can replicate in the respiratory tract, the eye mucosa, the intestinal tract, the urinary bladder, and the liver.

Most adenovirus human illness is associated only with one-third of adenovirus types. Although many adenovirus infections are subclinical, these viruses may cause acute respiratory disease (types 1–7, 14, and 21), conjunctivitis (types 3, 7, 8, 11, 14, 19, and 37), acute hemorrhagic cystitis (11 and 21), acute respiratory disease (ARD) of military recruits (types 3, 4, 7, 14, and 21), and gastroenteritis (types 31, 40, and 41) (Table 22.11). Adenovirus type 36

![Adenovirus](image)

**FIGURE 22.22 Adenovirus. Copyright Russell Kightley Media, reproduced with permission.**

| Subgenus | Serotype | Human illness |
|----------|----------|---------------|
| A        | 12       | Meningoencephalitis |
|          | 18, 31   | Diarrhea |
| B        | 3        | Acute febrile pharyngitis; adenopharyngo-conjunctival fever; pneumonia; follicular conjunctivitis; fatal infection in neonates |
|          | 7        | Acute febrile pharyngitis; adenopharyngo-conjunctival fever; acute respiratory disease with pneumonia; fatal infection in neonates; meningoencephalitis |
|          | 11       | Follicular conjunctivitis; hemorrhagic cystitis in children |
|          | 21       | Hemorrhagic cystitis in children; fatal infection in neonates |
|          | 14, 16   | Acute respiratory disease with pneumonia |
|          | 34, 35   | Acute and chronic infection in patients with immunosuppression and AIDS |
| C        | 1, 2, 6  | Acute febrile pharyngitis; pneumonia in children |
|          | 5        | Acute febrile pharyngitis; pertussis-like syndrome; acute and chronic infection in patients with immunosuppression and AIDS |
| D        | 8, 19, 37| Epidemic keratoconjunctivitis |
|          | 9, 10, 13, 15, 17, 42 | |
|          | 19, 20, 22–29 | |
|          | 30       | Fatal infection in neonates |
|          | 32, 33, 36, 38 | Asymptomatic (36 obesity) |
|          | 39, 42–47| Acute and chronic infection in patients with immunosuppression and AIDS |
| E        | 4        | Respiratory infection |
| F        | 40, 41   | Diarrhea |
has been associated with obesity in humans and animals (Atkinson et al., 2005).

Adenoviruses gain access to susceptible individuals through the mouth, the nasopharynx, or the conjunctiva. Although initial infection may occur by the respiratory route, fecal–oral transmission accounts for most adenovirus infections in young children because of the prolonged shedding of viruses in feces (Horwitz, 1996).

Nose, throat, and eye infections caused by adenoviruses have been associated with improperly disinfected swimming pool water. The enteric adenoviruses 40 and 41 have been recognized as the second most important etiologic agents of viral gastroenteritis in children. These viruses, in contrast to other adenoviruses, are not shed in respiratory secretions; thus their transmission is limited to the oral–fecal route. They have been associated with several outbreaks involving drinking water (Divizia et al., 2004).

Adenoviruses are common in primary sewage sludge, where they have been found in concentrations 10 times greater than that of the enteroviruses. In addition, a greater number of adenoviruses than enteroviruses have been consistently found in raw sewage around the world. Results of a comparative study of cytopathogenicity, immunofluorescence, and in situ DNA hybridization as methods for the detection of adenoviruses from water suggest that 80% of infectious adenoviruses in raw sewage may be enteric adenoviruses (Hurst et al., 1988).

Contaminated inanimate surfaces may play a significant role in adenovirus transmission because of its stability to drying. At room temperature, adenovirus 2 survives for eight and 12 weeks at low (7%) and high (96%) relative humidity, respectively, being more resistant than poliovirus 2, vaccinia virus, Coxsackie virus B3, and herpesvirus (Mahl and Sadler, 1975). The longer survival of adenoviruses has also been observed in water. Adenovirus type 5 survives longer in tap water, at either 4 or 18°C, than either poliovirus 1 or echovirus 7 (Bagdasar’yan and Abieva, 1971), and enteric adenoviruses 40 and 41 survive longer than poliovirus 1 and HAV in tap water and seawater (Enriquez et al., 1995b). The increased survival of the enteric adenoviruses in tap water and seawater, and the faster inactivation in sewage, may indicate that these viruses are inactivated by different mechanisms than those affecting the enteroviruses. The enteric adenoviruses are more thermally stable than polio 1, which is inactivated faster at temperatures above 50°C (Enriquez et al., 1995a). In addition, enteric adenoviruses 40 and 41 are more resistant to ultraviolet (UV) light disinfection than poliovirus type 1 and coliphage MS-2, which has been suggested as a model for enteric virus disinfection (Meng and Gerba, 1996).

The increased resistance showed by enteric adenoviruses, compared with other enteric viruses, may be associated with the double-stranded nature of their DNA, which, if damaged, may be repaired by the host cell DNA repair mechanisms. This mechanism would not be effective with ssRNA genome viruses such as polio 1 or HAV. It has been suggested that the longer survival of the enteric adenoviruses in tap water and seawater may be associated with DNA damage, and the faster inactivation in sewage may result from protein capsid damage.

22.4.1.3 Enteroviruses and Paraechoviruses

The enteroviruses are members of the family Picornaviridae, which are among the smallest ribonucleic acid (RNA) viruses. “Picornavirus” means small RNA virus. Enteroviruses and paraechoviruses are icosahedral viruses approximately 27 to 32 nm in diameter. Enteroviruses are divided into five groups (Table 22.12). The nucleic acid of enteroviruses consists of ssRNA. These are the viruses most often detected in sewage-polluted water. However, their apparent higher prevalence may be associated, in part, with available cell lines for their propagation, because many pathogenic enteric viruses such as HAV, enteric adenoviruses, rotavirus, norovirus, and other small round viruses are difficult to grow in conventional cell lines.

Although viruses belonging to the genera Enterovirus and Paraechovirus are capable of causing a wide variety of clinical conditions, from asymptomatic to disabling, or even fatal infections, they often do not cause overt disease (Table 22.11).

Although there are bovine, porcine, simian, and murine enteroviruses, it is believed that humans are the only natural host of human enteroviruses. Enteroviruses replicate primarily in the gastrointestinal tract and may be shed in large numbers (approximately 10⁹/g feces). The most common forms of transmission include the fecal–oral and respiratory routes (Fig. 22.23). Waterborne transmission may be considered a form of fecal–oral transmission in which the responsible vehicle is water instead of hands or fomites. Although enteroviruses are readily found in fecally contaminated drinking or recreational waters, waterborne enterovirus infection has been only occasionally documented. Waterborne outbreaks related to enteroviruses are difficult to document because many infections by these agents are subclinical. Therefore, an individual with waterborne infection without overt disease may infect others, who in turn may become ill, spreading the infection further. Attack rates of enteroviruses vary depending on the virus and the host age. Asymptomatic infections by poliovirus outnumber symptomatic disease (10:1), whereas symptomatic infection by echovirus 9 is relatively high (10:7) (Mores and Pallansch, 1995).

Both disease and infection caused by polioviruses are age related. Generally, infection is more common in infants, but adults and older children are more severely affected. However, some exceptions exist. Coxsackie-virus B virus infection is usually more severe in newborns than in older children and adults, often causing fulminant myocarditis, encephalitis, hepatitis, and death. Coxsackie-viruses are the most prevalent nonpolio enteroviruses (Mores and
**TABLE 22.12** Human Enteroviruses and Paraechoviruses and Clinical Illness

| Virus           | Serotypes                     | Clinical illness                                      |
|-----------------|-------------------------------|------------------------------------------------------|
| Poliovirus      | 3 types                        | Paralysis, aseptic meningitis, febrile illness        |
| **Enterovirus A** |                               |                                                      |
| Coxsackievirus  | A 2–7                         | Paralysis, aseptic meningitis                        |
|                 | A 8–16                         | Hand, foot, and mouth disease, encephalitis          |
| Enterovirus     | 71                             | Herpangina, exanthema, diarrhea                      |
|                 | 76                             |                                                      |
| **Enterovirus B** |                               |                                                      |
| Coxsackievirus  | B1–B6, A9                      | Aseptic meningitis, paralysis                        |
| Echovirus       | 1–9, 11–21, 24–33              | Exanthema, respiratory diseases                      |
| Enterovirus     | 69, 73–91                     | Pericarditis, myocarditis, febrile illness           |
| **Enterovirus C** |                               |                                                      |
| Coxsackievirus A| 1, 11, 13, 15, 17–22, 24       | Paralysis, aseptic meningitis                        |
| **Enterovirus D** |                               |                                                      |
| Enterovirus     | 68, 70                        | Pneumonia, acute hemorrhagic conjunctivitis          |
| Paraechoviruses | 1–3                           | Pericarditis, herpangina, respiratory disease        |

**FIGURE 22.23** Infection by Coxsackie viruses.
Pallansch, 1995). Coxsackie viruses are also the most common nonpolio enteroviruses isolated from water and wastewater. These viruses have been associated with several serious illnesses (Table 22.12). Coxsackievirus B5 infection has been associated with recreational water. In an outbreak at a boys’ summer camp, this virus was isolated from lake water; however, person-to-person contact appeared to be the main form of transmission (Hawley et al., 1973).

It is believed that almost all enteroviruses can be transmitted by the fecal-oral route; however, it is not clear if all of them can be transmitted by the respiratory route as well. Airborne transmission of enterovirus might include aerosol spread or direct exposure to respiratory secretions. Fecal-oral transmission may predominate in areas with poor sanitary conditions, whereas respiratory transmission may occur more often with better sanitation. In temperate climates, enteroviruses are more common during the summer season. In the United States, most enterovirus isolations (82%) occur from June to October. However, vaccine strains of poliovirus are isolated year-round because of routine vaccination of children. In contrast, in tropical and semitropical areas, enteroviruses do not show seasonality. Transmission of enteroviruses within a household is usually started by young children; then the infection spreads quickly to other family members, especially in larger families living under crowded conditions with poor hygiene. Ironically, paralytic poliomyelitis and perhaps some nonpolio enteroviral diseases are more often observed in developing countries, where sanitary conditions are improving. With poor hygiene, most individuals are infected at a very early age, when infection rarely results in overt disease and maternal immunity limits infection. This early exposure to the virus elicits a protective immune response on reexposure to these viruses later in life. In contrast, when early exposure is prevented or delayed as a result of better sanitation, an initial poliovirus infection is likely to occur at an older age, when maternal immunity has waned and the possibility of developing a more severe clinical condition is greater.

22.4.1.4 Hepatitis A Virus (HAV)

HAV is a picornavirus morphologically indistinguishable from other members of the same family. This agent was formerly classified as member of the Enterovirus genus (enterovirus 72). However, differences in nucleotide and amino acid sequences resulted in its classification as the only member of the hepatovirus group.

Hepatitis A was the most frequent viral waterborne disease in the United States from 1946 to 1994, with 68 outbreaks and 2297 cases. The average HAV incubation is approximately 30 days, but it may vary from 10 to 50 days, a variation associated with the dose. Infection with very few particles results in longer incubation periods and vice versa. The period of communicability extends from early in the incubation period to about a week after the development of jaundice. The greatest danger of spreading the disease to others occurs during the middle of the incubation period, well before the first presentation of symptoms. During this period the patient remains asymptomatic; however, active shedding of the virus is the norm. Therefore, it is during the incubation time that the infected individual has the highest potential for spreading HAV.

Hepatitis A is usually a mild illness, which almost always results in complete recovery. Severity and disease manifestation are age related. An estimated 80 to 95% of infected children younger than 5 years of age do not develop overt disease, whereas clinical manifestations are observed in approximately 75 to 90% of infected adults. The mortality rate in children 14 years old or younger is 0.1%; this rate rises to 0.3% in individuals between the ages of 15 and 39 years and 2.1% in those older than 40 years.

Hepatitis A is characterized by sudden onset of fever, malaise, nausea, anorexia, and abdominal discomfort, followed in several days by jaundice. In contrast to hepatitis B, HAV infection is not chronic. HAV is excreted in feces of infected people and can produce clinical disease when susceptible individuals consume contaminated water or foods. Water, shellfish, and salads are the most frequent sources. Contamination of foods by infected workers in food processing plants and restaurants is not uncommon.

HAV is very stable in the environment. It is more resistant to high temperatures than poliovirus. HAV can survive at 60°C for 1 h, and temperatures up to 85 to 95°C are needed to inactivate it in shellfish. Poor sanitation and crowding facilitate HAV transmission. Therefore, outbreaks of hepatitis A are common in institutions, crowded house projects, prisons, and military forces. In developing countries, the incidence of disease in adults is relatively low because of exposure to the virus in childhood. Most individuals 18 years and older possess an immunity that provides lifelong protection against reinfection. In the United States, the percentage of adults with immunity increases with age (10% for those 18–19 years of age to 65% for those over 50).

The survival of HAV on hands and its transfer to hands or inanimate surfaces were studied by Mbithi et al. (1992). They found that approximately 20% of the initial HAV inoculated on hands remained infectious for at least 4 h, and that inoculated onto a stainless steel surface survived for 2 h. They also determined that exerting higher pressure and friction between HAV-contaminated hands or fomites resulted in more efficient transfer of this virus to clean hands.

With better sanitation, HAV infection shifts to older individuals, and the incidence of overt disease increases. In more developed countries, low levels of HAV transmission occur. However, disease outbreaks are relatively common in most of these countries as a significant segment of the population is susceptible to HAV infection. Finally, in a few industrialized countries, hepatitis A outbreaks are uncommon, and nearly all HAV infections occur among individuals who have visited areas where HAV is highly prevalent.
22.4.1.5 Hepatitis E Virus (HEV)

The hepatitis E virus is the leading cause of acute viral hepatitis among young and middle-aged adults in developing countries. HEV has a diameter of 32–34 nm and an ssRNA genome. Serologically related smaller (27–30 nm) particles are often found in feces of patients with hepatitis E and are presumed to represent degraded viral particles. It has been suggested that the higher prevalence of HEV in adults may be due to a silent infection early in life, with subsequent waning of immunity after 10 to 20 years, when they again become susceptible to infection by HEV (Bradley, 1992). Important epidemiological features of HEV infection are the frequent occurrence of outbreaks associated with consumption of sewage-polluted water (Case Study 22.2) and its severity, particularly among pregnant women, in whom the case-fatality rate may be as high as 25% (Balayan, 1993). HEV virus also infects swine and other animals and has been transmitted by the consumption of undercooked meat (Tamada et al., 2004). To date, no outbreak has occurred in the United States.

Hepatitis E is clinically indistinguishable from hepatitis A. It is characterized by jaundice, malaise, anorexia, abdominal pain, arthralgia, and fever. The incubation period for hepatitis E varies from two to nine weeks. The disease is most often seen in young to middle-aged adults (15–40 years old). The disease is usually mild and resolves in two weeks, leaving no long-term effects, and with a relatively low fatality rate (0.1–1%). There is no evidence of immunity against HEV in the population that has been exposed to this virus (Margolis et al., 1997).

22.4.1.6 Rotavirus

Rotaviruses are classified within the family Reoviridae. These viruses have a characteristic genome consisting of 11 dsRNA segments surrounded by a distinctive two-layered protein capsid. Particles are approximately 70 nm in diameter. Six serological groups (A–F) have been identified; groups A, B, and C infect both humans and animals, and D, E, and F have been detected only in animals. Group A rotaviruses have been associated with the majority of infantile acute gastroenteritis cases, group B with severe diarrhea epidemics in adults in China, and group C with sporadic cases of diarrhea in children, but their clinical importance has not been determined. Within each group, rotaviruses are classified into serotypes.

Rotaviruses are the most important agents of infantile gastroenteritis around the world. Group A rotavirus is endemic worldwide. It is the leading cause of severe diarrhea among infants and children and accounts for about half of the cases requiring hospitalization. In temperate areas, it occurs primarily in the winter, but in the tropics it occurs throughout the year. Group B rotavirus, also called adult diarrhea rotavirus, has caused major epidemics of severe diarrhea affecting thousands of persons of all ages in China. Group C rotavirus has been associated with rare and sporadic cases of diarrhea in children in many countries.

Rotaviruses are shed in large numbers, up to $10^{10}$ viral particles per gram of feces (White and Fenner, 1994). Names applied to the infection caused by the most common and
widespread group A rotavirus include acute gastroenteritis, infantile diarrhea, winter diarrhea, acute nonbacterial infectious gastroenteritis, and acute viral gastroenteritis.

Rotaviruses are transmitted by the fecal–oral route. Person-to-person spread by contaminated hands is probably one of the most important routes by which rotaviruses are transmitted. Institutions or close communities such as pediatric wards, day care centers, and family homes are usually most affected by outbreaks of gastroenteritis caused by rotaviruses. Because of high infectivity, rotavirus-infected food handlers may contaminate foods that require handling but do not require further cooking, such as cakes, salads, and fruits. Among adults, multiple foods served at banquets were implicated in two outbreaks, and an outbreak related to contaminated municipal water occurred in Colorado in 1981. Several large outbreaks of group B rotavirus involving millions of persons as a result of sewage contamination of drinking water supplies have occurred in China since 1982. Although to date outbreaks caused by group B rotavirus have been confined to mainland China, seroepidemiological surveys have indicated lack of immunity to this group of viruses in the United States. Rotaviruses are quite stable in the environment and have been found in estuary samples at levels as high as one to five infectious particles per gallon (FDA, 1992). Sanitary measures adequate for bacteria and parasites seem to be ineffective in endemic control of rotavirus, as similar incidences of rotavirus infection are observed in countries with both high and low health standards.

Rotavirus gastroenteritis is a self-limiting disease, which can be mild to severe. It is characterized by vomiting, watery diarrhea, and mild fever. Asymptomatic rotavirus excretion has been well documented and may play a role in perpetuating endemic disease. The incubation period ranges from 1 to 3 days. Symptoms often start with vomiting followed by 4–8 days of diarrhea. As with other viral gastroenteritis, rotavirus gastroenteritis treatment consists of fluid and electrolyte replacement; if it is untreated, severe diarrhea with dehydration and death may occur. Individuals of all ages are susceptible to rotavirus infection. Premature infants, children 6 months to 2 years of age, the elderly, and the immunocompromised are particularly prone to develop more severe symptoms. In adults, rotaviral infection is usually subclinical. However, rotavirus gastroenteritis outbreaks have occurred in army recruits, geriatric patients, and hospital staff. Over 3 million cases of rotavirus gastroenteritis occur annually in the United States. It has been estimated that rotaviruses, in the United States alone, cause more than 1 million cases of severe diarrhea and up to 150 deaths per year. Worldwide, close to 1 million infants and young children die from rotavirus infection each year. Rotavirus infection does not result in an efficient or long-lasting immunity. Therefore, rotavirus infection in the same child often occurs up to six times during childhood.

Some animal rotaviruses, such as SA-11, are readily propagated in cell culture; the human rotaviruses, however, are rather fastidious. These viruses have not been grown efficiently in any conventional tissue culture system.

22.4.1.7 Human Caliciviruses

The use of electron microscopy (EM) since the early 1970s for the examination of fecal specimens from individuals suffering from nonbacterial gastroenteritis has shown many previously unknown viruses. Among these, viruses such as adenoviruses and rotaviruses are larger and well defined; thus, they are relatively easy to identify by EM. However, many smaller (20–40 nm) viruses without a distinctive morphology are often present in fecal samples of patients suffering from gastroenteritis (Fig. 22.24). Norwalk virus was the first to be described (Kapikian et al., 1972). Subsequently, other small round viruses were observed in diarrheal stools, namely, Montgomery County, Hawaii, Wollan, Ditchling, the Parramata, the Cockle agents, and minirota-virus. Norwalk-like viruses have emerged as a major cause of food- and waterborne disease. Several epidemiological studies have shown that Norwalk viruses are the major cause of foodborne illness, causing as much as 67% of the foodborne illness in the United States (Gerba and Kayed, 2003). However, the identity of the Norwalk virus was not defined until Jiang et al. (1990) sequenced the Norwalk virus genome, allowing its classification in the family Caliciviridae. Subsequently, Norovirus was approved as the official genus name for the Norwalk-like viruses. These viruses nonenveloped the virus with a diameter of approximately 26 to 35 nm and a positive-sense ssRNA genome. The human Caliciviridae have been divided into two genera: Norovirus and Sapporovirus.

Immunity to noroviruses is poorly understood. However, infectivity studies with volunteers have shown that individual susceptibility is more important than acquired immunity. It has been suggested that genetically determined factors are the primary determinants of resistance to norovirus infection, perhaps at the level of cellular receptor sites (i.e., blood group antigens expressed in the gut). In developing countries, antibodies to norovirus are acquired early in life, and the peak incidence of illness may also occur among younger age groups than in developed nations. Norovirus and related viruses usually produce a mild and brief illness, lasting 1 to 2 days. It is characterized by nausea and abdominal cramps, followed commonly by vomiting in children and diarrhea in adults.

22.4.2 Respiratory Viruses

Worldwide, respiratory illnesses are the most common illnesses in humans and most have a viral etiology. Respiratory disease is associated with a large number of
viruses, including rhinoviruses, coronaviruses, parainfluenza viruses, respiratory syncytial virus (RSV), influenza virus, and adenovirus. These viruses, when they infect the upper respiratory tract, can cause acute viral rhinitis or pharyngitis (common cold); when the primary site of infection is the lower respiratory tract, they can cause laryngotracheitis (croup), bronchitis, or pneumonia.

Mortality related to acute respiratory disease may be especially significant in children and in the elderly. In adults, temporary disability results in important economic loss. Respiratory infection often results from self-inoculation, when virus-contaminated hands or fingers rub the eyes or when viruses are introduced into the mouth or nose. Another important form of transmission of respiratory viruses is inhalation of contaminated aerosols. The mode of transmission of viruses causing respiratory disease is influenced by the type of virus. For example, fragile enveloped viruses such as parainfluenza virus, RSV, and influenza virus are usually transmitted by close contact with infected individuals, whereas the more stable nonenveloped viruses such as adenoviruses, rhinoviruses, and coronaviruses are commonly transmitted by self-inoculation by contaminated hands.

22.4.2.1 Rhinoviruses

Rhinoviruses (Latin rhino, nose), belong to the family Picornaviridae. Two important characteristics differentiate these two types of viruses: stability at low pH values and temperature. Whereas the rhinoviruses are inactivated at pH below 6, the enteroviruses are stable at low pH values. In contrast, rhinoviruses are stable at temperatures (50°C) that would inactivate most enteroviruses (Couch, 1996). Human rhinoviruses have an icosahedral morphology, containing a single-stranded RNA genome. The diameter of these viruses is approximately 25 to 30 nm. Although no etiologic agent is identified in half of the acute upper respiratory illnesses, it has been estimated that 30 to 50% are caused by rhinoviruses. In fact, rhinovirus infection is probably the most common type of human acute infection (Gwaltney, 1997). The optimal temperature for rhinovirus growth is 33 to 35°C, which corresponds to the normal temperature of the nasal mucosa. This may explain why these viruses propagate most efficiently in the upper respiratory tract. These viruses can be propagated in monkey and human cell lines. There are 100 different human rhinovirus serotypes. Although these viruses are species specific, an equine rhinovirus can infect other species, including humans. Experimentally, chimpanzees and gibbons have been infected with human rhinoviruses, but these animals do not develop disease. Volunteer studies have shown that infection can be started with less than one tissue culture median infectious dose (TCID50).

Rhinovirus infection, after one to four days of incubation, is characterized by nasal obstruction and discharge, sneezing, scratchy throat, mild cough, and malaise. The relatively high rate of rhinovirus infection may be associated...
with the large number of rhinovirus serotypes and the fact that the same serotype can infect an individual more than once. In addition, immunity to rhinovirus infection is short-lived.

Although rhinovirus infection occurs throughout the year, in temperate climates its frequency increases during colder months and in the tropics the peak incidence occurs during the rainy season. Experiments with volunteers have failed to associate exposure to low temperatures with an increased susceptibility to rhinovirus infection. Therefore, the cause of this seasonality remains unclear. Nevertheless, it has been suggested that colder temperatures or rain may increase the survivability of these viruses and/or promote crowding conditions, in which the virus may propagate more efficiently.

During rhinovirus infection, the highest concentration of viruses is found in the nasal and pharyngeal mucosa. In contrast, in saliva, rhinoviruses are found in low numbers and inconsistently. This suggests that aerosols may not play an important role in the transmission of these agents, as sneezing and coughing generate aerosols composed mainly of saliva (Fig. 22.25). In fact, studies with volunteers have shown that rhinovirus transmission by hand contact and self-inoculation of the eye or nasal mucosa is much more efficient than through aerosols (Hendley et al., 1973).

The high concentration of rhinoviruses in nasal secretions easily leads to hand contamination. Rhinoviruses have been isolated from 40 to 90% of hands of ill persons and from 6 to 15% of environmental objects such as doorknobs, dolls, coffee cups, and glasses. It is believed that contamination of objects in the environment may play a significant role in the transmission of rhinoviruses. This and the ability of rhinoviruses to survive on human skin and on inanimate objects may explain the more prevalent self-inoculation form of transmission. Both finger-to-eye and finger-to-nose contact are part of normal human behavior. Therefore, experimental rhinovirus transmission has been effectively interrupted by hand disinfection.

Rhinoviruses are relatively stable on inanimate surfaces. They can survive at least 3h on human skin, nonporous materials such as plastic surfaces, Formica, stainless steel, and hard synthetic fabrics such as nylon and Dacron. In porous materials such as paper tissue and cotton fabric, rhinoviruses can survive for 1h (Hendley et al., 1973).

22.4.2.2 Paramyxovirus

Parainfluenza viruses and respiratory syncytial viruses (RSV) belong to the family Paramyxoviridae. These viruses are spherical enveloped particles of heterogeneous size, ranging from 125 to 250nm. The paramyxovirus viral particle has a lipid envelope covered with spikes of about 10nm. The nucleic acid of these agents consists of a single-stranded RNA molecule (Ginsberg, 1990).

RSV and parainfluenza viruses do not survive well in the environment. If suspended in a protein-free medium at 4°C, 90 to 99% of their infectivity is lost within 4h. Organic solvents and detergents rapidly inactivate these viruses by dissolving their lipid envelopes. Both RSV and parainfluenza viruses cause a variety of illnesses, primarily in infants and young children. The most important are common cold, bronchitis, bronchiolitis, laryngotracheobronchitis (croup), and pneumonia.

22.4.2.3 Parainfluenza Viruses

The parainfluenza viruses constitute two of the four genera of the family Paramyxoviridae: Paramyxovirus (parainfluenza virus types 1 and 3) and Rubulavirus (parainfluenza virus types 2 and 4). Parainfluenza viruses infect most people during childhood. Types 1 and 2 are often associated with croup in infants and type 3 with bronchiolitis and pneumonia; type 4 seldom causes illness. Although parainfluenza infections in children and infants can result in serious disease, most infections are subclinical. In adults these viruses can cause a mild cold. Survival studies have shown that parainfluenza virus type 3 remains infectious in aerosol particles for at least 1h (Miller and Artenstein, 1967).

Disease caused by parainfluenza viruses is observed throughout the year. Infection by parainfluenza virus types 1 and 2 occurs endemically, but small epidemics caused by these agents are observed every two years. Parainfluenza virus type 3 infects approximately 60% of infants during the first two years of life, reaching 80% by 4 years of age, whereas infection by types 1 and 2 does not reach 80% until 10 years of age. In children, an incubation period of two to four days has been estimated for parainfluenza virus illness. Studies have shown that a child infected with parainfluenza type 3 may shed viruses an average of eight days, but shedding viruses for up to four weeks has been documented (Frank et al., 1981).

FIGURE 22.25 Source of rhinovirus dispersion. Data from Buckland et al., 1964.
22.4.2.4 Respiratory Syncytial Virus

RSV belongs to the family Paramyxoviridae, as the only member of the genus Pneumovirus. The characteristics of RSV have already been described. This virus is the most important respiratory pathogen during infancy and early childhood. It causes approximately half of the cases of bronchiolitis and 25% of cases of pneumonia in infants. Approximately 90,000 hospital admissions and 4500 deaths are associated each year with RSV in the United States in both infants and young children. In approximately half of infants younger than eight months, the infection spreads to the lower respiratory tract, resulting in life-threatening bronchitis, bronchiolitis, bronchopneumonia, and croup (Ginsberg, 1990). Furthermore, RSV is the most common cause of nosocomial infections in pediatric wards and the most important cause of middle ear infection in children. Approximately 1% of all infants suffer from a severe RSV infection that may require hospitalization, and about 1% of those die. Asymptomatic first infection with RSV is rare; almost 100% of infected children develop disease. Because of poor protective immunity, reinfection is very common, but the disease is not as severe. Inoculation of adult volunteers with RSV has shown that the nose and eye mucosae are the most important portals of entry of this virus. Efficient transmission of RSV by large droplets or by touching occurs when susceptible individuals are in close contact with children shedding the virus but not when they are exposed to small-particle aerosol (McIntosh, 1997).

The incubation period of RSV respiratory disease is 4 to 5 days. Replication of RSV in the upper respiratory tract can reach concentrations of $10^4$ to $10^6$ TCID$_{50}$/ml of secretion, with higher titers in infants. Infected individuals may shed RSV for up to 3 weeks. It has been observed that this shedding period correlates with the severity of illness but not with the age of the patient. In temperate climates RSV infection may occur throughout the year, but it peaks during winter months with few occurrences in the summer; in tropical areas outbreaks often occur during the rainy season.

22.4.2.5 Influenza Viruses

The influenza viruses belong to three genera of the family Orthomyxoviridae: influenza virus A, influenza virus B, and influenza virus C. These viruses, approximately 80 to 120 nm in size, possess a lipid envelope with a genome consisting of eight segments of single-stranded RNA. Influenza viruses are divided into types A, B, and C. The influenza virus type A is further classified into many subtypes according to host of origin, year, and geographic location of first isolation.

It is not known why several recent influenza pandemics have started in China. It has been suggested that the large pig, duck, and human population in the Canton area may facilitate coinfection of animals with influenza viruses originated from different species, leading to genetic reassortment and to the generation of viruses with novel antigenic and virulence characteristics. There is strong evidence that aquatic birds are the main reservoir of all influenza viruses in other species. For example, the catastrophic influenza pandemic of 1918, in which over 20 million people died, is believed to have been caused by an influenza A virus derived from a bird.

Influenza A virus is a highly contagious agent, causing epidemics of an acute respiratory infection known as influenza, with high mortality in the elderly. About 75% of all influenza deaths occur in individuals over 55 years of age. Influenza A virus infection may be asymptomatic, but more often it may be manifested by a wide variety of clinical conditions, ranging from an annoying flu to a fatal pneumonia. For instance, a subtype of influenza A virus that was responsible for epidemics of severe disease at the beginning of the century caused minor illness in older people during widespread epidemics in the 1980s. The reason for this wide variation is poorly understood, but it has been speculated that age, underlying illnesses, previous exposure to a similar influenza A virus subtype, and virus virulence may be associated with the type of disease presentation. In the United States in recent years more than 90% of influenza-related deaths occurred among persons of 65 years of age or older (Centers for Disease Control, 1996). Mortality due to influenza is significant and varies from season to season. During 9 of 20 influenza seasons in the United States (1972–1992), more than 20,000 people died each season; during four seasons, more than 40,000 deaths were recorded (Centers for Disease Control, 1997).

Influenza A virus can be transmitted most efficiently through aerosols. It has been demonstrated that influenza A virus remains infectious for at least 1 h in aerosols at room temperature (Murphy and Webster, 1996). Clinical manifestations of influenza start rather suddenly, about one to four days after infection. A disabling syndrome, characterized by high fever, together with muscle pain, sore throat, nasal congestion, conjunctivitis, cough, and headache, is usually the norm.

Influenza epidemics occur intermittently. In temperate climates they usually occur from early fall to late spring, but in tropical regions epidemics are observed throughout the year. Influenza epidemics spread rapidly and tend to occur worldwide. Although variable, epidemics caused by influenza A virus are observed every two to four years, whereas influenza B virus epidemics normally occur every three to six years. Immunity to influenza is long lived; however, it is virus subtype specific. Epidemiological studies have shown that individuals previously infected with influenza A subtype H1N1 during the 1957 pandemic were resistant to infection when the same subtype reappeared in the 1977 pandemic, but they were fully susceptible when exposed to other influenza A subtypes.
22.4.2.6 SARS

Severe acute respiratory virus or SARS is a serious respiratory virus that resulted in a worldwide outbreak in 2003 resulting in about 8,000 cases and nearly 800 deaths. It was caused by a coronavirus believed to have originated from bats in Southeast Asia and later transmitted to other animals. It causes a fever with chills and coughing, with 10 to 20% of persons infected developing diarrhea. It is believed to be spread by contact droplets or from fomites contact. The airborne route does not appear to be the major route of spread as close contact appears necessary for effective spread under most conditions. It is present in both the respiratory droplets and feces. It appears capable of surviving for a few days in liquids and for 24 h on fomites (Rabenau et al., 2005).

22.5 FATE AND TRANSPORT OF PATHOGENS IN THE ENVIRONMENT

There are many potential routes for the transmission of excreted enteric pathogens. The ability of an enteric pathogen to be transmitted by any of these routes depends largely on its resistance to environmental factors, which control its survival, and its capacity to be carried by water as it moves through the environment. Some routes can be considered “natural” routes for the transmission of waterborne disease, but others—such as the use of domestic wastewater for groundwater recharge, large-scale aquaculture projects, or land disposal of disposable diapers—are actually new routes created by modern human activities.

Human and animal excreta are sources of pathogens. Humans become infected by pathogens through consumption of contaminated foods, such as shellfish from contaminated waters or crops irrigated with wastewater; from drinking contaminated water; and through exposure to contaminated surface waters as may occur during bathing or at recreational sites. Furthermore, those individuals infected by the preceding processes become sources of infection through their excrement, thereby completing the cycle.

In general, viral and protozoan pathogens survive longer in the environment than enteric bacterial pathogens (Fig. 22.26). How long a pathogen survives in a particular environment depends on a number of complex factors, which are listed in Table 22.13. Of all the factors, temperature is probably the most important. Temperature is a well-defined factor with a consistently predictable effect on enteric pathogen survival in the environment. Usually, the lower the temperature, the longer the survival time. But freezing temperatures generally result in the death of enteric bacteria and protozoan parasites. Viruses, however, can remain infectious for months or years at freezing temperatures. Moisture—or lack thereof—can cause decreased survival, and UV light from the sun is a major factor in the inactivation of indicator bacteria in surface waters; thus, die-off in marine waters can be predicted by amount of

| Excreted load* | Survival (months)* |
|----------------|--------------------|
|                | 1  | 2  | 3  | 4  | 5  | 6  | 7  | 8  | 9  | 10 | 11 | 12 |
| 1. Campylobacter spp. | 10⁷ |     |     |     |     |     |     |     |     |     |     |
| 2. Giardia lamblia | 10⁵ |     |     |     |     |     |     |     |     |     |     |
| 3. Shigella spp. | 10⁷ |     |     |     |     |     |     |     |     |     |     |
| 4. Vibrio cholerae | 10⁷ |     |     |     |     |     |     |     |     |     |     |
| 5. Salmonella spp. | 10⁸ |     |     |     |     |     |     |     |     |     |     |
| 6. Escherichia coli (pathogens) | 10⁸ |     |     |     |     |     |     |     |     |     |     |
| 7. Enteroviruses | 10⁷ |     |     |     |     |     |     |     |     |     |     |
| 8. Hepatitis A virus | 10⁹ |     |     |     |     |     |     |     |     |     |     |
| 9. Ancylostoma duodenale | 10⁹ |     |     |     |     |     |     |     |     |     |     |
| 10. Taenia saginata | 10⁴ |     |     |     |     |     |     |     |     |     |     |
| 11. Ascaris lumbricoides | 10⁴ |     |     |     |     |     |     |     |     |     |     |

*Typical average number of organisms/g feces
*Estimated average life of infective stage at 20–30°C.
(Modified from Feachem et al., 1983).

FIGURE 22.26 Survival times of enteric pathogens in water, wastewater, soil, and on crops.
exposure to daylight. Viruses are much more resistant to inactivation by UV light.

Many laboratory studies have shown that the microflora of natural waters and sewage are antagonistic to the survival of enteric pathogens. It has been shown, for example, that enteric pathogens survive longer in sterile water than in water from lakes, rivers, and oceans. Bacteria in natural waters can feed on indicator bacteria. Suspended matter (clays, organic debris, and the like) and fresh or marine sediments have been shown to prolong their survival time (Fig. 22.27).

### QUESTIONS AND PROBLEMS

1. What are pathogens? What is an enteric pathogen?
2. What is the difference between a waterborne and a water-based pathogen?
3. Which group of enteric pathogens survives longest in the environment and why?
4. What are some of the niches in which *Legionella* can grow to high numbers?
5. Why are *Cryptosporidium* and *Giardia* major causes of waterborne disease in the United States today?
6. What are the names of the environmentally resistant forms of waterborne protozoan parasites?
7. What group of animals are the reservoirs of *Toxoplasma gondii*?
8. What is the difference between a frank pathogen and an opportunistic pathogen? Give examples of each.
9. Which groups of pathogens cannot grow in the environment outside a host? Which ones can?
10. Which virus is the leading cause of childhood gastroenteritis worldwide?
11. Which virus is most commonly associated with waterborne disease outbreaks?
12. Which bacterium is the leading cause of gastroenteritis in the United States?
13. Fomites are important in the spread of what respiratory viruses?
14. Why are respiratory infections more common at certain times of the year?
15. What virus can be transmitted by contact with the eyes?
16. What type of hepatitis has a high mortality in pregnant women?
17. What type of virus causes eye infections in persons swimming in contaminated waters?
18. What are nosocomial infections?
19. What organism is responsible for typhoid? Cholera? Winter diarrhea in infants?
20. Why are only certain strains of E. coli capable of causing disease in humans?
21. What type of pathogenic E. coli is transmitted primarily by cattle feces?

REFERENCES AND RECOMMENDED READINGS

Adam, R. (1991) The biology of Giardia spp. Microbiol. Rev. 55, 706–732.
Allen, M. J., and Geldreich, E. E. (1975) Bacteriological criteria for groundwater quality. Ground Water 13, 45–52.
Atkinson, R. L., Dhurandhar, N. V., Allison, D. B., Bowen, R. L., Israel, B. A., Albu, J. B., and Augustus, A. S. (2005) Human adenovirus-36 is associated with increased body weight and paradoxical reduction of serum lipids. Int. J. Obes. 29, 281–286.
Bagdasar’yan, G. A., and Abieva, R. M. (1971) Survival of enteroviruses and adenoviruses in water. Hyg. Sanit. 36, 333–337.
Balayan, M. S. (1993) Hepatitis E virus infection in Europe: Regional situation regarding laboratory diagnosis and epidemiology. Clin. Diagn. Virol. 1, 1–9.
Baumann, J. (1991) Isolation of Acinetobacter from soil and water. J. Bacteriol. 96, 39–42.
Bifulco, J. M., Shirey, J. J., and Bissonnette, G. K. (1989) Detection of Acinetobacter spp. in rural drinking water supplies. Appl. Environ. Microbiol. 55, 2214–2219.
Bingham, A. K., Jarroll, E., and Meyer, E. (1979) Giardia spp.: Physical factors of excystation in vitro, and excystation vs. eosin exclusion as determinants of viability. Exp. Parasitol. 47, 284–291.
Blaser, M. J., Smith, P. F., Wang, W. L. L., and Hoff, J. C. (1986) Inactivation of Campylobacter by chlorine and monochloramine. Appl. Environ. Microbiol. 51, 307–311.
Bradley, D. W. (1992) Hepatitis E: Epidemiology aetiology and molecular biology. Rev. Med. Virol. 2, 19–28.
Brudastov, A. N., Lemelev, V. R., Kholnukhanedov, S. K., and Krasnos, L. N. (1971) The clinical picture of the migration phase of ascaris in self-infection. Medsko Parazit. 40, 165–168.
Buckland, F. E., and Tyrell, D. A. (1964) Experiments on the spread of colds. I. Laboratory studies on the dispersal of nasal secretion. J. Hyg. 62, 365–377.
Burden, D. J., Whitehead, A., Green, E. A., McFadzean, J. A., and Beer, R. (1976) The treatment of soil infected with human whipworm Trichuris trichiura. J. Hyg. 77, 377–382.
Burns, D., Wallace, R. J., Jr., Schultz, M. E., Zhang, Y., Zabairi, S. Q., Pang, Y., Gilbert, C. L., Brown, B. A., Noel, E. S., and Gordin, F. M. (1991) Nosocomial outbreak of respiratory tract colonization with Mycobacterium fortuitum: Demonstration of the usefulness of pulsed field gel electrophoresis in an epidemiological investigation. Am. Rev. Respir. Dis. 144, 1153–1159.
Carmichael, W. W. (1994) The toxins of cyanobacteria. Sci. Am. 273, 64–72.
Carter, A. M., Pacha, R. E., Clark, G. W., and Williams, E. A. (1987) Seasonal occurrence of Campylobacter spp. in surface waters and their correlation with standard indicator bacteria. Appl. Environ. Microbiol. 53, 523–526.
Centers for Disease Control. (1996) Prevention and control of influenza: Recommendations of the Advisory Committee on Immunization Practices (ACIP). Morbidity and Mortality Weekly Report 45 (RR-5).
Centers for Disease Control. (1997) Influenza Surveillance—United States, 1992–93 and 1993–94. CDC Surveill. Summ. 46 (SS-1).
Centers for Disease Control. (2004) Outbreak of cyclosporiasis associated with snow peas—Pennsylvania. Morbidity and Mortality Weekly Report 53, 876–878.
Chin, D. P., Hopewell, P. C., Yajko, D. M., Vittinghoff, E., Horsburgh, C. R., Jr., Hadley, W. K., Stone, E. N., Nassos, P. S., Ostroff, S. M., Jacobson, M. A., Matkin, C. C., and Reingold, A. L. (1994) Mycobacterium avium complex in the respiratory or gastrointestinal tract and the risk of M. avium bacteremia in patients with human immunodeficiency virus infection. J. Infect. Dis. 169, 289–295.
Clayton, M. (2006) Oscillatoria MC. Instructional Technology Website, Department of Botany, University of Wisconsin, Madison, Wisconsin, http://botit.botany.wisc.edu/images/130/Bacteria/Cyanobacteria/Oscillatoria/Oscillatoria_MC.html.
Colbourne, J. S., Dennis, P. J., Trew, R. M., Beery, C., and Vesey, G. (1988) Legionella and public water supplies. “Proceedings for the International Conference on Water and Wastewater Microbiology,” Newport Beach, CA, February 8–11, 1988, Vol. 1.
Collins, C. H., Grange, J. M., and Yates, M. D. (1984) Mycobacteria in water. J. Bacteriol. 57, 193–211.
Couch, R. B. (1996) Rhinoviruses. In “Virology” (B. N. Fields, D. M. Knipe, P. M. Howley, et al., eds.) Raven Press, New York, pp. 713–734.
Craun, G. F. (1979) Waterborne disease—a status report emphasizing outbreaks in ground-water systems. Ground Water 17, 183–193.
Craun, G. (1986) “Waterborne Diseases in the United States,” CRC Press, Boca Raton, FL, pp. 121–168.
Craun, G. F., Calderon, R. L., and Craun, M. F. (2006) Waterborne disease outbreaks: their causes, problems, and challenges to treatment barriers. “Waterborne Pathogens,” Second Ed. American Water Works Association, Denver, CO.
Cruz, J. R., Bartlett, A. V., Herrmann, J. E., Caceres, P., Blacklow, N. R., and Cano, F. (1992) Astrovirus-associated diarrhea among Guatemalan ambulatory rural children. J. Clin. Microbiol. 30, 1140–1144.
Curry, A., and Canning, E. (1993) Human microsporidiosis. J. Infect. 22, 229–236.
Divizia, M., Gabrieli, R., Donia, D., Macaluso, A., Bosch, A., Guix, S., Sanchez, G., Villena, C., Pinto, R. M., Palombi, L., Buonuomo, E., Cenko, F., Leno, L., Bebci, D., and Bino, S. (2004) Waterborne gastroenteritis outbreak in Albania. Water Sci. Technol. 50, 57–61.
Dubey, J. P. (1996) Strategies to reduce transmission of Toxoplasma gondii to animals and humans. Vet. Parasitol. 64, 65–70.
Dubey, J. P., and Beattey, C. P. (1988) “Toxoplasmosis of Animals and Man,” CRC Press, Boca Raton, FL.
Enriquez, C. E., Hurst, C. J., and Gerba, C. P. (1995b) Survival of the enteric adenoviruses 40 and 41 in tap, sea, and wastewater. Water Res. 29, 2548–2553.
Erlandsen, S. L. (1995) Biotic transmission—giardiasis a zoonosis? In “Giardia: From Molecules to Disease” (R. Thompson, J. Reynolds, and A. Lymbery, eds.), University Press, Cambridge, pp. 83–97.

Erlandsen, S., Sherlock, L., Januszka, M., Schupp, D., Schaefer, F., Jakubowski, W., and Bemrick, W. J. (1988) Cross-species transmission of Giardia spp.: Inoculation of beavers and muskrats with cysts of human, beaver, mouse, and muskrat origin. Appl. Environ. Microbiol. 54, 2777–2785.

Falcao, D. P., Valentini, S. R., and Leite, C. Q. F. (1993) Pathogenic or potentially pathogenic bacteria as contaminants of fresh water from different sources in Araraquara, Brazil. Water Res. 27, 1737–1741.

Falconer, I. R., Runnegar, M. T. C., Buckley, T., Huyn, V. L., and Bradshaw, P. (1989) Using activated charcoal to remove toxicity from drinking water containing cyanobacterial blooms. J. Am. Water Works Assoc. 2, 102–105.

FDA, Anonymous. (1992) Foodborne pathogenic microorganisms and natural toxins. Washington, DC.

Feachem, R. G., Bradley, D. J., Garelick, H., and Mara, D. D. (1983) “Sanitation and Disease Health Aspects of Excreta and Wastewater Management,” World Bank, Washington, DC.

Fields, B. S., Shots, E. B., Jr., Feeley, J. C., Gorman, G. W., and Martin, W. T. (1984) Proliferation of Legionia pneumophila as an intracellular parasite of the ciliated protozoan Tetrahymena pyriformis. Appl. Environ. Microbiol. 47, 467–471.

Frank, A. L., Taber, L. H., Wells, C. R., Wells, J. M., Glezen, W. P., and Paredes, A. (1981) Patterns of shedding of myxoviruses and paramyxoviruses in children. J. Infect. Dis. 144, 433–441.

Freedman, D. O. (1992) Intestinal nematodes. In “Infectious Diseases” (S. L. Gorbach, J. G. Bartlett, and N. Blacklow, eds.), W. B. Saunders, Philadelphia, pp. 2003–2008.

Frenkel, J. K., Ruiz, A., and Chinchilla, M. (1975) Soil survival of Toxoplasma oocysts in KS and Costa Rica. J. Trap. Med. Hyg. 24, 439–443.

Gerba, C. P., and Kayed, D. (2003) A major cause of foodborne illness. J. Food Sci. 68, 1136–1142.

Giai Nobel Y hoc 2005 cho Barry Marshall và Robin Warren về Helicobacter pylori (2005), http://vietsciences.free.fr/nobel/medecine/nobelmedecine2005.htm.

Ginsberg, H. S. (1990) Paramyxoviruses. In “Microbiology” (B. D. Davis, R. Dulbecco, H. N. Eisen, and H. S. Ginsberg, eds.), Fourth Ed., J. B. Lippincott, Philadelphia, pp. 947–959.

Ginsberg, M., Keenan, K., Thompson, M., and Anders, B. (1994) Stool survey of asymptomatic diapered children in day care. Pediatrics 94S, 1026–1027.

Goslee, S., and Wolinsky, E. (1976) Water as a source of potentially pathogenic mycobacteria. Am. Rev. Respir. Dis. 113, 287–292.

Gupta, A., Polyak, C. S., Bishop, R. D., Sobel, J., and Mintz, E. D. (2004) Laboratory-confirmed shigellosis in the United States, 1989–2002: Epidemiologic trends and patterns. Clin. Infect. Dis. 38, 1372–1377.

Gwaltney, J. M. (1997) Rhinoviruses. In “Viral Infections of Humans, Epidemiology and Control” (A. S. Evans, and R. A. Kaslow, eds.), Fourth Ed. Plenum, New York, pp. 815–838.

Hawley, H. B., Morin, D. P., Geraghty, M. E., Tomkow, J., and Phillips, A. (1973) Coxsackievirus B epidemic at a boys’ summer camp. JAMA 226, 33–36.

Hegarty, J. P., Dowd, M. T., and Baker, K. H. (1999) Occurrence of Helicobacter pylori in surface water in the United States. J. Appl. Microbiol. 87, 697–701.

Hendley, J. O., Wenzel, R. P., and Gwaltney, J. M. (1973) Transmission of rhinovirus colds by self-inoculation. N. Engl. J. Med. 288, 1361–1363.

Herwaldt, B. L., Craun, G. F., Stokes, S. L., and Juraneck, D. D. (1992) Outbreaks of waterborne disease in the United States: 1989–90. J. Am. Water Works Assoc. 84, 129–135.

Hibler, C., and Hancock, C. (1990) Waterborne giardiasis. In “Drinking Water Microbiology” (G. McFeters, ed.), Springer-Verlag, New York, pp. 271–293.

Hopkins, D. R. (1992) Homing in on helmiths. Am. J. Top. Med. Hyg. 46, 626.

Horwitz, M. S. (1996) Adenoviruses. In “Fields Virology” (B. N. Fields, D. M. Knipe, and P. M. Howley, et al., Third Ed. Lippincott-Raven, Philadelphia, pp. 2149–2171.

Huang, P., Wever, W., Sosin, D., Griffin, P., Long, E., Murphy, J., Kocka, F., Peters, C., and Kallick, C. (1995) The first reported outbreak of diarrheal illness associated with Cyclospora in the United States. Ann. Intern. Med. 123, 409–414.

Hughes, H. P. A. (1985) How important is toxoplasmosis? Parasitol. Today 1, 41–44.

Hurst, C. J., Benton, W. H., and McClellan, K. A. (1988) Suppression of viral replication by guanidine: a comparison of human adenoviruses and enteroviruses. J. Virol. Methods 22, 1–11.

Isaac-Renton, J., Corderio, C., Sarafis, K., and Shahriar, H. (1993) Characterization of Giardia duodenalis isolates from a waterborne outbreak. J. Infect. Dis. 167, 431–440.

Izaguirre, G. (1992) A copper-tolerant Phomodium species from Lake Mathews, CA, that produces 2-methylisoborneol and geosmin. Water Sci. Technol. 25, 217–223.

Jepsen, A., and Roth, H. (1995) Epizootiology of Cysticercus bovis-resistance of the eggs of Taenia saginata. Report 14. In. Vet. Cong. 22, 43–50.

Jiang, X., Graham, D. Y., Wang, K., and Estes, M. K. (1990) Norwalk virus genome cloning and characterization. Science 250, 1580–1583.

John, D. T. (1982) Primary amebic meningoencephalitis and the biology of Naegleria fowleri. Annu. Rev. Microbiol. 36, 101–103.

Johnson, D. C., Reynolds, K. A., Gerba, C. P., Pepper, I. L., and Rose, J. B. (1995) Detection of Giardia and Cryptosporidium in marine waters. Water Sci. Technol. 31, 439–442.

Johnson, D. C., Enriquez, C. E., Pepper, I. L., Davis, T. L., Gerba, C. P., and Rose, J. B. (1997) Survival of Giardia, Cryptosporidium, poliovirus and Salmonella in marine waters. Water Sci. Technol. 35, 261–268.

Jones, A. R., Bartlett, J., and McCormack, J. G. (1995) Mycobacterium avium complex (MAC) osteomyelitis and septic arthritis in an immunocompetent host. J. Infect. 30, 59–62.

Kapikian, A. Z., Wyatt, R. G., Dolin, R., Thornhill, T. S., Kalica, A. R., and Chanock, R. M. (1972) Visualization by immune electron microscopy of 27nm particle associated with acute infectious non-bacterial gastroenteritis. J. Virol. 10, 1075–1081.

Kappus, K. D., Lundgren, R. G. Jr., Juraneck, D. B., Roberts, J. M., Spencer, H. C. (1994) Intestinal parasitism in the United States: update on a continuing problem. Am. J. Trop. Med. Hyg. 50, 705–713.

Kemp, J. S., Wright, S. E., and Bukhari, Z. (1995) On farm detection of Cryptosporidium parvum in cattle, calves and environmental samples. In “Protozoan Parasites and Water” (W. B. Betts, D. Casemore, C. Fricker, H. Smith, and J. Watkins, eds.), The Royal Society of Chemistry, Cambridge, UK, pp. 154–157.

Klein, P. D., Graham, D. Y., Gaillou, A., Opekun, A. R., and Smith, E. O. (1991) Water source as risk factor for Helicobacter pylori infection in Peruvian children. Lancer 337, 1503–1506.
Knight, P. (1995) Once misidentified human parasite is a cyclosporan. *ASM News* **61**, 520–522.

Kurtz, J. B., and Lee, T. W. (1987) Astroviruses: Human and animal. *In “Novel Diarrhea Viruses”* (G. Bock, and J. Whelan, eds.), Wiley, Chichester, UK, pp. 92–107.

LeChevallier, M. W., Seidler, R. J., and Evans, T. M. (1980) Enumeration and characterization of standard plate count bacteria in chlorinated and raw water supplies. *Appl. Environ. Microbiol.* **40**, 922–930.

Lee, L. A., Shapiro, C. N., Hargrett-Bean, N., and Tauxe, R. V. (1991) Hyperendemic shigellosis in the United States: A review of surveillance data for 1967–1988. *J. Infect. Dis.* **164**, 894–900.

Lee, T. W., and Kurtz, J. B. (1994) Prevalence of human astrovirus serotypes in the Oxford region 1976–92, with evidence for two new serotypes. *Epidemiol. Infect.* **112**, 187–193.

Lipp, E. K., Hug, A., and Colwell, R. R. (2002) Effects of global climate on infectious disease: the cholera model. *Clin. Microbiol. Rev.* **15**, 757–770.

Lucey, D. R., and Maguire, J. H. (1993) Schistosomiasis. *Infect. Dis. North Am.* **7**, 635–653.

MacKenzie, W., Hoxie, N., Proctor, M., Gradus, M., Blair, K., Peterson, D., Kazmierczak, J., Addiss, D., Fox, K., Rose, J., and Davis, J. (1994) A massive outbreak in Milwaukee of Cryptosporidium infection transmitted through the public water supply. *N. Engl. J. Med.* **331**, 161–167.

Mahl, M. C., and Sadler, C. (1975) Virus survival on inanimate surfaces. *Can. J. Microbiol.* **21**, 819–823.

Margolis, H. S., Alter, M. J., and Hadler, S. C. (1997) Viral hepatitis. *In “Viral Infections of Humans, Epidemiology and Control”* (A. S. Evans, and R. A. Kaslow, eds.), Fourth Ed, Plenum, New York, pp. 363–418.

Marrie, T. J. (1994) Community-acquired pneumonia. *Clin. Infect. Dis.* **18**, 501–515.

Mbiti, J. N., Springthorpe, V. S., and Sattar, S. A. (1992) Survival of hepatitis A virus on human hands and its transfer on contact with animate and inanimate surfaces. *J. Clin. Microbiol.* **30**, 757–763.

McHenry, R., Bartlett, M. S., Lehman, G. A., and O’Conner, K. W. (1987) The yield of routine duodenal aspiration for *Giardia lamblia* during oesphagogastroduodenoscopy. *Gastrointest. Endosc.* **33**, 425–426.

McIntosh, K. (1997) Respiratory syncytial virus. In *Viral Infections of Humans* (A.S. Evans and R.A. Kaslow, ed.), Fourth Ed, Plenum, New York, pp. 691–711.

McOist, S., Boid, R., Jones, T., Easterbee, N., Hubbard, A., and Jarrett, O. (1991) *J. Wildlife Dis.* **27**, 693–696.

Meenhorst, P. L., Reingold, A., Groothuis, D. G., Gorman, G. W., Wilkinson, H. W., McKinney, R. M., Feeley, J. C., Brenner, D. J., and Furth, R. V. (1985) Water-related nosocomial pneumonia caused by *Legionella pneumophila* serogroup 1 and 10. *J. Infect. Dis.* **152**, 356–363.

Meisel, et al. (1976).

Meng, Q. S., and Gerba, C. P. (1996) Comparative inactivation of enteric adenovirus, polio virus, and coliphages by ultraviolet irradiation. *Water Res.* **30**, 2665–2668.

Messner, M. J., Chappell, C. L., and Okhuyzen, P. C. (2001) Risk assessment for *Cryptosporidium*: a hierarchical Bayesian analysis of human dose response data. *Water Res.* **35**, 3934–3940.

Miller, W. S., and Artenstein, M. S. (1967) Aerosol stability of three acute disease viruses. *Proc. Soc. Exp. Biol. Med.* **125**, 222–227.

MMWR. (1995) Update, *Vibrio cholerae* O1—Western Hemisphere, 1991–1994, and *V. cholerae* O139—Asia, 1994. *Morbidity and Mortality Weekly Report* **44**(11), 215–219.

Moffat, J. E., and Tompkins, L. S. (1992) A quantitative model of intracellular growth of *Legionella pneumophila* in *Acanthamoeba castellanii*. *Infect. Immun.* **60**, 292–301.

Morens, D. M., and Pallansch, M. A. (1995) Epidemiology. *In “Human Enterovirus Infections”* (A. A. Rothbart, ed.), ASM Press, Washington, DC, pp. 3–24.

Morgan, D. R., Johnson, P. C., DuPont, H. L., Satterwhite, T. K., and Wood, L. V. (1985) Lack of correlation between known virulence properties of *Aeromonas hydrophila* and enteropathogenicity for humans. *Infect. Immun.* **50**, 62–65.

Mullens, A. (1996) I think we have a problem in Victoria: MD’s respond quickly to toxoplasmosis outbreak in BC. *Can. Med. Assoc. J.* **154**, 1721–1724.

Murphy, B. R., and Webster, R. G. (1996) Orthomyxoviruses. *In “Fields Virology”* (B. N. Fields, D. M. Knipe, P. M. Howley, et al., eds.) Third Ed. Lippincott-Raven, Philadelphia, pp. 1397–1445.

Narayan, L. V., and Nunez, W. J., III (1974) Biological control: Isolation and bacterial oxidation of the taste-and-odor compound geosmin. *J. Am. Water Works Assoc.* **66**, 532–536.

Nguyen, A. M., Engstrand, L., Genta, R. M., Graham, D. Y, and el-Zaatari, F. A. (1993) Detection of *Helicobacter pylori* in dental plaque by reverse-transcriptase–polymerase chain reaction. *J. Clin. Microbiol.* **31**, 783–787.

Nuorti, J. P., Niskanen, T., Hallanvuo, S., Kela, E., Ataca, M., Fredrikssoon-Ahornaa, M., Lyytikainen, O., Siitonen, A., Korkeala, H., and Ruutu, P. (2004) A widespread outbreak of *Yersinia pseudotuberculosis* O:3 infection from iceberg lettuce. *J. Infect. Dis.* **189**, 766–774.

Ortega, Y., Sterling, C., Gilman, R., Cama, V., and Diaz, F. (1993) *Cyclospora* species—a new protozoan pathogen of humans. *N. Engl. J. Med.* **328**, 1308–1312.

Payment, P., Gamache, F., and Paquette, G. (1988) Microbiological and virological analysis of water from two water filtration plants and their distribution systems. *Can. J. Microbiol.* **34**, 1304–1309.

Pepper, I. L., Gerba, C. P., and Brusseau, M. L. (2006) Environmental and Pollution Science, 2e. Academic Press, San Diego, CA.

Phillip, R., Brown, M., Bell, R., and Francis, F. (1992) Health risks associated with recreational exposure to blue–green algae (cyanobacteria) when windsurfing and fishing. *Health Hyg.* **13**, 115–119.

Rabenau, H. F., Cinatl, J., Morgenstern, B., Bauer, G., Preiser, W., and Doerr, H. W. (2005) Stability and inactivation of SARS coronavirus. *Med. Microbiol. Immunol.* **194**, 1–6.

Reavis, C. (2005) Rural health alert: *Helicobacter pylori* in well water. *J. Am. Acad. Nurse Pract.* **17**, 283–289.

Rabold, J., Hoge, C., and Shlim, D. (1994) *Cyclospora* outbreak associated with chlorinated drinking water. *Lancet* **344**, 1360–1361.

Rolle-Kampczyk, U. E., Fritz, G. J., Dize, U., Lehmann, I., Richter, M., and Herbarth, O. (2004) Well water—one source of *Helicobacter pylori* colonization. *Int. J. Hyg. Environ. Health* **207**, 363–368.

Rollins, D. M., and Colwell, R. R. (1986) Viable but nonculturable stage of *Campylobacter jejuni* and its role in survival in the natural aquatic environment. *Appl. Environ. Microbiol.* **52**, 531–538.

Samuel, M. C., Vugia, D. I., Shallow, S., Marcus, R., Seglar, S., McGivern, T., Kessenborg, H., Reilly, K., Kennedy, M., Angulo, F., and Tauxe, R. V. (2004) Epidemiology of sporadic *Campylobacter* infection in the United States and declining trend in incidence. *Clin. Infect. Dis.* **38**, S165–S174.

Savioli, L., Renganathan, E., Montresor, A., Davis, A., and Behbehani, K. (1997) Control of schistosomiasis—a global picture. *Parasitol. Today* **13**, 444–448.
