Abstract

There are many restrictions on topical medications for the oral cavity. Various factors affect the topical application of drugs in the oral cavity, an open and complex environment. The complex physical and chemical environment of the oral cavity, such as saliva and food, will influence the effect of free drugs. Therefore, drug delivery systems have served as supporting structures or as carriers loading active ingredients, such as antimicrobial agents and growth factors (GFs), to promote antibacterial properties, tissue regeneration, and engineering for drug diffusion. These drug delivery systems are considered in the prevention and treatment of dental caries, periodontal disease, periapical disease, the delivery of anesthetic drugs, etc. These carrier materials are designed in different ways for clinical application, including nanoparticles, hydrogels, nanofibers, films, and scaffolds. This review aimed to summarize the advantages and disadvantages of different carrier materials. We discuss synthesis methods and their application scope to provide new perspectives for the development and preparation of more favorable and effective local oral drug delivery systems.

Keywords: Delivery systems. Dental caries. Hydrogels. Nanoparticles. Periodontal disease.
Introduction

The oral cavity is a complex environment which communicates with the external environment, the upper respiratory tract, and the digestive system. A wide variety of microorganisms is present in the oral cavity, including bacteria and fungi, such as Streptococcus mutans (S. mutans), Lactobacillus spp., Porphyromonas gingivalis (P. gingivalis), and Candida albicans. These microorganisms colonize different parts of the oral cavity, including tooth surfaces and the periodontium, and form biofilms which invade oral cavity tissues. Periodontitis is a consequence of alterations in the ecology of resident microbial communities. The intimate interaction of bacteria with the host leads to an inflammatory reaction.

Dental caries is a consequence of dietary, sugar-driven biofilm accumulation and localized acidification. Due to frequent sugar consumption, the development of a predominantly acidic environment will favor the growth of aciduric bacteria in the biofilm. Aciduric bacteria include Streptococcus spp., Lactobacillus spp., etc. As a result, the dynamic balance between commensals and opportunistic pathogens is disrupted, causing deleterious microbial community shifts and disrupting tooth-enamel mineral homeostasis.

According to the global burden of disease (GBD) study, permanent tooth caries is among the ten diseases with the highest incidence for years lived with disability. Periodontal diseases comprise a wide range of inflammatory conditions affecting tooth-supporting structures (the gingiva, bone, and periodontal ligaments), which starts with the localized inflammation of the gingiva, initiated by a microbial biofilm that forms on the teeth and gingiva. Periodontal diseases lead to teeth loss and contribute to systemic inflammation, which is highly prevalent worldwide. Thus, periodontal diseases represent a significant public health problem. Regarding the topical application of drugs to the oral cavity, the influence of the oral environment on the drugs should be considered. Its complex physical and chemical environment and the complexity of bacterial biofilm affect drug application via the oral cavity. For example, salivary flow rates in the mouth might affect the efficacy of topical anesthesia to some extent, whereas side effects and drug resistance are inevitable in systemic administration. Salivary clearance might dilute and weaken the active ingredients in the oral environment. Therefore, carrier systems had to be designed to consistently release active ingredients so their estimated concentrations can be effective. Recently, emerging advanced biomaterials, including hydrogels, films, nanofibers, and particles hold great potential as cell/drug carriers for local drug delivery and biomimetic scaffolds. Biofilms decrease the effects of drugs on microorganisms, and antibacterial substances are easily metabolized. Moreover, there is no perfect way of delivering drugs to sites such as the periodontal pocket and the periapical area. Drug delivery systems have many advantages, such as increasing drug solubility, prolonging drug action time, improving drug targeting, and reducing cytotoxicity. Moreover, delivery systems play a role in inhibiting bacteria by releasing active ingredients, such as chlorine, and can serve as engineering scaffolds to promote tissue regeneration. For example, patients receiving ibuprofen-incorporated, chitosan-based microspheres show better anti-inflammatory properties than the orally administered ibuprofen group. Moreover, bone morphogenetic protein 2 (BMP-2) loaded into calcium silicate scaffolds showed continuous release and better stimulation for the induction of mesenchymal stem cells. In the field of anesthesia, experiments have also proved that appropriate hybrid nanofilms containing the eutectic mixture of 5% lidocaine-prilocaine (LDC-PLC) show a better anesthetic effect, with higher permeability and no cytotoxicity.

In the reviewing process, "oral/oral cavity/mouth/mouth cavity" and "drug delivery systems/carrier materials/drug targeting" were used as research items in the Pubmed, MEDLINE, and Web of Science databases. Relevant articles with an impact factor greater than five and published after 2015 were included. Moreover, we added some articles about the progress of experiments based on previous classic reviews. All drug delivery systems were classified as nanoparticles, hydrogels, nanofibers, and films according to their material properties. The carrier materials used as oral drug delivery systems in recent years are summarized in Table 1, and their different forms are described in Figure 1. This review discusses drug delivery systems in the oral cavity, which are prepared by various methods to adapt to complex situations. For example, they can be used as support structures to promote regeneration in defective tissues or as drug carriers to release active ingredients which...
Table 1 - Carriers materials used as oral drug delivery systems in recent years

| Drug delivery system | Materials | Active ingredients | Biological activity | Diseases | Experiments | References |
|----------------------|-----------|--------------------|---------------------|----------|-------------|------------|
| **Inorganic nanoparticles** | Amine-functionalized expanded pore mesoporous silica (aMSN) | Polyacrylic acid-stabilized amorphous calcium phosphate (pHA-ACP) | Promote re-mineralization | Enamel white spot lesions | In vitro | Hua et al., 2020 |
| | Silver-decorated mesoporous silica nanoparticles (Ag-MSNs) | Chlorhexidine (CHX), Silver nanoparticles | Antibacterial effects | Periodontal diseases | In vitro; In vivo | Lu et al., 2018 |
| | Mesoporous calcium silicate nanoparticles (MCSNs) | Gentamicin; Fibroblast growth factor-2 (FGF-2) | Antibacterial effects; Promote tissue regeneration | | In vitro | Huang et al., 2017 |
| | Bone Morphogenetic Protein 2 (BMP-2) | Poly (L-lactide-co-glycolide acid) (PLGA) | Promote re-mineralization | Immature permanent tooth with necrotic pulp | In vitro | Huang et al., 2018 |
| **Nanoparticles** | Amine-functionalized expanded pore mesoporous silica (aMSN) | Polyacrylic acid-stabilized amorphous calcium phosphate (pHA-ACP) | Promote re-mineralization | Enamel white spot lesions | In vitro | Hua et al., 2020 |
| | Silver-decorated mesoporous silica nanoparticles (Ag-MSNs) | Chlorhexidine (CHX), Silver nanoparticles | Antibacterial effects | Periodontal diseases | In vitro; In vivo | Lu et al., 2018 |
| | Mesoporous calcium silicate nanoparticles (MCSNs) | Gentamicin; Fibroblast growth factor-2 (FGF-2) | Antibacterial effects; Promote tissue regeneration | | In vitro | Huang et al., 2017 |
| | Bone Morphogenetic Protein 2 (BMP-2) | Poly (L-lactide-co-glycolide acid) (PLGA) | Promote re-mineralization | Immature permanent tooth with necrotic pulp | In vitro | Huang et al., 2018 |

Nanoparticles have widely served as drug delivery systems in the oral cavity. Moreover, to support the future development of carrier materials, we summarized their composition and application.
inorganic carriers, such as mesoporous silica and mesoporous calcium silicate and organic carriers, such as poly (d,l-lactide-co-glycolide acid) (PLGA) and chitosan. Nanoparticle size lies mostly between 1-100 nm, which results in good diffusion properties and loading performances. Inorganic nanoparticles are mainly composed of inorganic materials which are crystallized and amorphous solids at ambient temperature. Their reactivity is different in various solution environments, which significantly influence their toxicity. Organic nanoparticles mainly consist of organic substances such as lipids, proteins, etc. Depending on their composition, organic nanoparticles can be degraded and digested in different parts in vivo. Organic nanoparticles are generally less toxic than inorganic nanoparticles, but they might cause toxicity under particular circumstances. In recent years, inorganic nanoparticles have received attention due to their unique materials, which are more inert, stable, and easy to functionalize.

Inorganic nanoparticles

In general, mesoporous materials are used as inorganic nanoparticle carrier systems. Mesoporous materials include mesoporous silica nanoparticles (MSNs) and mesoporous calcium silicate nanoparticles (MCSNs). MSNs have been modified by amination to make them more suitable for use in the oral environment. In recent studies, the amine-functionalized expanded pore mesoporous silica (aMSN) was synthesized to load polyacrylic acid–stabilized amorphous calcium phosphate (PAA-ACP). PAA-ACP loaded aMSN has the potential to treat white enamel spot lesions and serve as filler in resin adhesives. Regarding their antimicrobial properties, incorporated silver ions into MSNs, synthesizing silver-decorated mesoporous silica nanoparticles (Ag-MSNs) and using them to load chlorhexidine (CHX). They showed redox/pH-responsive release properties due to CHX and silver ions which could inhibit S. mutans biofilm growth. Ag-MSNs\textsuperscript{6}CHX were more effective than an equivalent amount of free CHX in limiting S. mutans biofilm formation since they induced bacterial cell death, particularly in the long term. Though S. mutans was only used as a model to evaluate the effect of Ag-MSNs\textsuperscript{6}CHX, the material may be considered for treating diseases caused by bacterial biofilms, such as periodontal diseases.

A practical antibacterial component, silver ions were also loaded into mesoporous calcium silicate to prepare nano-silver-incorporated mesoporous calcium-silicate nanoparticles (Ag-MCSNs), resulting in lower cytotoxicity. The nanoparticles slowly released...
silver ions which could inhibit Enterococcus faecalis (E. faecalis) growth and colonization.\textsuperscript{34} CHX can be successfully loaded onto MCSNs by the mixing-coupling method. These nanoparticles released CHX, showing low cytotoxicity and excellent anti-E. faecalis properties. Moreover, they can also release Ca\textsuperscript{2+}/SiO\textsubscript{2−} ions, showing an \textit{in vitro} mineralization property. These nanoparticles could be developed into bone-defect-filling material or intracanal medications in dentistry.\textsuperscript{35}

Huang, et al.\textsuperscript{36} (2017) loaded gentamicin and fibroblast growth factor 2 (FGF-2) with mesoporous calcium silicate nanoparticles, which could sustain the release of FGF-2 and gentamicin. These nanoparticles showed bone/cementum tissue regeneration and inhibited bacterial viability, suggesting their use in endodontics as a valuable, biocompatible dental pulp tissue regenerative material capable of odontogenesis. MCSNs can also be loaded with BMP-2 and prepared into a three-dimensional scaffold by 3D printing, inducing the differentiation of dental pulp cells and odontoblasts. The BMP-2-loaded mesoporous calcium silicate 3D scaffold could serve as an intracanal scaffold for both hemostatic clot retention and odontogenesis in reparative endodontic therapy.\textsuperscript{21} Nano-metal particles, such as Ag, Cu, copper oxide (CuO), and titanium oxide (TiO2), can serve as antibacterial agents in the oral cavity.\textsuperscript{37,38} In recent years, the toxic effects of metals and metal oxides have attracted significant attention.\textsuperscript{39,40} Thus, the safety of metal nanoparticles should be further investigated.\textsuperscript{41}

Although many articles have highlighted the role of antimicrobials, there are still many disadvantages in using antibacterial drugs, including drug resistance, dysbiosis, etc. Therefore, active agents are needed to inhibit bacterial biofilms without affecting the ecological balance of flora, such as probiotics and prebiotics, thus reestablishing ecological balance or the biodiversity of oral microbiota.

**Organic nanoparticles**

In recent years, nanoparticle carriers have commonly been prepared by the self-assembly of organic molecules, such as PLGA, chitosan, and poly(lactic acid).\textsuperscript{42-45} PLGA is a conventionally used component in preparing organic nanocarriers. In previous studies,\textsuperscript{40,43} PLGA-nanoparticles were prepared to encapsulate CHX. These nanoparticles could be delivered to the demineralized dentin matrix and the resin-dentin interface by dentinal tubules, in which they might play a role not only in inhibiting bacteria but also in using matrix metalloproteinase (MMP).\textsuperscript{46} PLGA is used to load grapeseed extract (GSE), which could improve the biodegradation resistance of demineralized dentin. Biodegradable polymer nanoparticles can deliver GSE via demineralized dentinal tubules, reducing the biodegradability of the dentin collagen matrix.\textsuperscript{47} PLGA is also used to load lovastatin and control its release, which can induce dentin differentiation at appropriate concentrations. Thus, these nanoparticles could serve as an adjunctive treatment in indirect pulp capping procedures.\textsuperscript{48} PLGA is used to deliver metronidazole or N-phenacylthiazolium bromide to modulate periodontitis progression.\textsuperscript{49} PLGA-chitosan nanoparticles are also loaded with simvastatin and doxycycline to promote the repair of infected periodontal sites and non-infected osseous defects.\textsuperscript{50} Compared with gel carriers, a PLGA particle carrier prolonged the release of active components, indicating its functional suitability for clinical applications.\textsuperscript{51}

Liposomes are often used as drug carriers to encapsulate active ingredients.\textsuperscript{52} They were used to develop curcumin-loaded solid lipid nanoparticles (CurSLN) to treat oral mucosal infections, showing increased antibacterial activity.\textsuperscript{53} Furthermore, liposomes have served as carriers to prepare pH-responsive quaternary ammonium chitosan-liposome nanoparticles by first loading doxycycline with liposomes, then coating the latter with N, N, N-Trimethyl chitosan (TMC). When pH is low, the free amino group in the quaternary ammonium chitosan on the surface of nanoparticles will be protonated. The positive charge on the TMC surface could be increased to destabilize the nanoparticles, thereby triggering DOX release. These nanoparticles could be closely linked to biofilms and effectively destroy their membrane structure and disrupt bacterial biofilms \textit{in vivo}.\textsuperscript{54}

Chitosan is usually used as a carrier to load active ingredients. Chitosan nanoparticles loaded with toothpaste actives such as sodium fluoride (NaF) and cetylpyridinium chloride (CPC) are prepared by emulsion dispersion or ionic gelation. These nanoparticles continuously released their active ingredients, which makes them highly promising as dental delivery systems in the protection against caries.\textsuperscript{55,56} After functionalization with
glutathione, chitosan showed a stronger ability to bind to chlorhexidine, resulting in a better release of chlorhexidine within 48 h.\textsuperscript{57} Moreover, polycaprolactone has been used to encase chlorhexidine to form Poly (ε-caprolactone) (PCL)-coated CHX nanocapsules. In this process, the nanocapsules could reach the demineralized dentin region via micron-scale dentin tubules, slowly releasing chlorhexidine, thereby playing an antibacterial role.\textsuperscript{58}

Furthermore, chitosan has been used as a carrier to load dexamethasone (DEX), and alginate solution, containing transforming growth factor-β1 (TGF-β1), to wrap the resulting nanoparticles. In turn, the core-shell nanosystem (TD-NS) could release TGF-β1 and DEX so materials might treat periapical inflammation.\textsuperscript{59} Zhu, et al.\textsuperscript{60} (2018) used the phosphorylated poly (amide-amine) dendrimer (PAMMA) as a carrier to load apigenin, a type of antibiotic insoluble in water. The material can both play an antibacterial role and promote remineralization. Polylactic acid (PLA) and PLGA have been used to load cells and BMP-2, respectively, promoting odontogenic differentiation and the formation of dentoid tissue via human stem cells of apical papilla (SCAP). The data showed great promise in promoting dentin tissue regeneration.\textsuperscript{61}

Moreover, continuous release systems for CHX were developed based on chitosan (CS) and montmorillonite (MMT). The CHX-hybrid nanosystem containing positively-charged chitosan showed good mucoadhesion properties which enabled the drug delivery system to stay longer at the absorption site and, therefore, contributed to a beneficial effect on drug bioavailability.\textsuperscript{62} A pH-responsive and redox-sensitive polymer-based AmB-delivery carrier system was developed based on O-Carboxymethyl chitosan (OCMCH). This system was functionalized by conjugation with the antifungal peptide histatin 5, which targeted ligands and synergistic antifungal molecules against \textit{Candida albicans}.\textsuperscript{63} The cellulose acetate phthalate nanoparticle is used to load chlorhexidine, enabling much greater infiltration into the subgingival tissue.\textsuperscript{64} Diblock copolymers underwent ultrasound treatment to form nanoparticle carriers by self-assembly. The recent nanoparticle carrier (NPC) DDS is flexible enough to co-load farnesol and myricetin. Therefore, nanoparticles could reduce \textit{S. mutans} biofilm organization and acid production.\textsuperscript{65} Some inorganic nanoparticles are also used in oral mucosal diseases, such as hydroxyapatite nanoparticles and hyperbranched core-multishell nanocarriers.\textsuperscript{66,67}

### Hydrogels

Hydrogels composed a natural or synthetic 3D network by physical or chemical strategies and could absorb a large amount of water (up to 1000-fold compared to the dry weight). Hydrogels consist of long polymer chains filled with water molecules between them.\textsuperscript{68,69,70} Natural hydrogels are biocompatible and can degrade into nontoxic by-products which can interact with biological macromolecules. However, their limitations lie in their weak mechanical strength and immunogenicity.\textsuperscript{71} By contrast, synthesized hydrogels can be better regulated and have better stability, but they might degrade into toxic by-products.\textsuperscript{72} Hydrogels can control drug release due to changes (swelling, dissolution or degradation) in the gel structure in response to internal or external stimuli.\textsuperscript{73} With appropriate release mechanisms, active drugs can be maintained locally at high concentrations for a longer period. Currently, the clinical applications of hydrogels for drug delivery include ophthalmology, cardiovascular diseases, and cancer, having attracted the attention of oral medicine in recent years.\textsuperscript{74,75,76,77} Compared to nanopore carriers, hydrogel carriers can maintain or trigger drug delivery and enable multi-drug delivery. At the same time, hydrogels avoid intravenous injection and have better local application prospects, making them interesting platforms for drug delivery in the oral cavity.\textsuperscript{78}

### Chitosan hydrogels

In recent years, chitosan/β-glycerophosphate (CS/β-GP) thermosensitive hydrogels have served as drug delivery systems to load bone morphogenetic protein-7 (BMP-7) and ornidazole (ORN), continuously releasing them to treat periodontal diseases.\textsuperscript{79} Moreover, chitosan/β-glycerophosphate (CS/β-GP) thermosensitive hydrogels were loaded with aspirin to control inflammation and erythropoietin. Hydrogels can promote periodontal regeneration.\textsuperscript{80} They are also used to load naringin, a naturally derived polymethoxylated flavonoid compound with anti-inflammatory properties that inhibit the inflammatory state of periodontitis.\textsuperscript{81} Regarding other types of thermosensitive hydrogels,
A thermosensitive micellar hydrogel was prepared from amphiphilic copolymer poly (ε-caprolactone-co-1,4,8-trioxaspiro[4.6]undecanone)poly(ethylene glycol)-poly(ε-caprolactone-co-1,4,8-trioxaspiro[4.6]undecanone) (PECT) nanoparticles. Ibuprofen and basic fibroblast growth factor (bFGF) were encapsulated in these thermosensitive micellar hydrogels. Thus, these materials can promote anti-inflammatory activity and soft tissue healing.

Other hydrogels

There are other components of hydrogels. To promote tissue regeneration, biocompatible hydrogelator Nap-Phe-Phe-Tyr-OH (NapFFY) hydrogels were loaded with stromal cell-derived factor-1 (SDF-1) and bone morphogenetic proteins (BMPs) to promote and accelerate periodontal bone regeneration. Pan, et al. (2019) used PLGA–PEG–PLGA triblock copolymers as a carrier to load human periodontal ligament stem cells, which had been modified to overexpress platelet-derived growth factor-BB. These cells are capable of mediating enhanced alveolar bone regeneration in vivo. High-stiffness transglutaminase crosslinked gelatins (TG-gels) were developed, incorporated with IL-4 and stromal cell-derived factor (SDF)-1α, and used for periodontal tissue regeneration. The presence of IL-4 might promote M2 polarization of macrophages (Mψs) and osteogenesis in bone marrow-derived stromal cells (BMSCs). In the field of dental pulp regeneration, hyaluronic acid (HA) hydrogels were reinforced with cellulose nanocrystals (CNCs) and then, enriched with platelet lysate (PL) and GFs. The PL-loaded hydrogels showed preferential supportive properties for encapsulated human dental pulp cells (hDPCs) in in vitro culture conditions. These hydrogels could serve as scaffold for GF delivery and cell recruitment, with great potential in future developments for regenerative dentistry. Moreover, antibiotics and nitric oxide (NO), releasing biomimetic nanomatrix gel, were synthesized by the self-assembly of peptide amphiphiles, which could promote tooth revascularization with the maturation of root canals. Hydrogels can also be used to treat fungal infections and deliver anesthetics. Hydroxypropyl methylcellulose (HPMC) was used to load Histatin-5 (Hst-5), an antimicrobial peptide, which could treat oral candidiasis. A nanostructured lipid-biopolymer hydrogel was developed to continuously deliver lidocaine-prilocaine for trans-buccal preanesthesia, which showed stability (for 6 months in critical conditions) and suitable mechanical properties for oral administration. Natural products can also be used to make thermosensitive hydrogels for oral applications.

Nanofibers

Nanofibers are commonly produced from ceramic materials, metallic compounds, and synthetic polymers by electrospinning, phase separation, self-assembly or laser spinning. Nanofibers are defined as fibers approximately 100 nm in diameter, which can carry diverse antimicrobial molecules, thereby showing biocompatibility and a porous structure. High surface area and porosity are the main advantages of nanofibers. Drug release can be regulated by changing the porosity of nanofibers. Porous nanofibers can promote cell adhesion and shorten the drug release cycle, which is conducive to wound healing. In the oral cavity, nanofibers have structural characteristics similar to dental pulp and periodontal tissues when compared with the most prevalent nanoparticles. Moreover, nanofibers have a larger contact surface which helps distant delivery. Their advantage of easy modulation of drug release profiles depends on the properties of polymer/polymeric blends/other materials used. To treat Actinomyces naeslundii, P. gingivalis or Enterococcus faecalis infections, polydioxanone nanofibers were used to load antibiotics, such as metronidazole (MET), ciprofloxacin (CIP), and minocycline (MINO). The use of nanofibers suggested a significant potential in the eradication/elimination of bacterial biofilms. Moreover, a type of polydioxanone nanofiber was developed to load vascular endothelial growth factor (VEGF), which might be a promising scaffold for additional optimization in endodontic regenerative procedures.

Moreover, polyvinyl alcohol, polylactic acid, and some natural products are also used to prepare nanofiber carriers. Thiolated chitosan (CS-SH) blended with polyvinyl alcohol (PVA) was used as the carrier to load α-mangostin, an antibacterial substance, and prepared into nanofibers. These nontoxic nanofibers could rapidly adhere to the buccal mucosa in the oral cavity, in which α-mangostin was released to inhibit oral bacterial flora and promisingly prevent the formation of dental caries. Resorbable PLA fibers based on electrospinning were developed, containing
the antibiotic metronidazole. These fibers could slowly release metronidazole for the topical treatment of periodontitis. These fibers could slowly release metronidazole for the topical treatment of periodontitis. Cellulose nanofibers (CNF) were modified with κ-carrageenan oligosaccharides (CO) to deliver drugs, load surfactin, and manufacture surfactin-loaded CO-CN. The surface-loaded CO-CN showed antioxidant activity and inhibited the growth of *S. mutans* and *P. gingivalis* by preventing biofilm formation and reducing metabolic activity, which promoted oxidative stress in a concentration-dependent manner. However, most polymers currently used in nanofibers are expensive and not suitable for patients who have financial difficulties. At the same time, the stability of nanofibers and hosts’ immune responses should be further evaluated.

### Films

In general, oral films serve as carriers of antibacterial or antimycotic agents for gradual and direct release at the target area in the oral cavity, which might be suitable for long-term local effects and improve therapy effectiveness. Mucoadhesive oral films (MOF) were prepared from carmellose by incorporating a nanotechnologically modified clay mineral intercalated with antiseptic drugs (chlorhexidine diacetate and digluconate) which could inhibit the growth of *Staphylococcus* spp. and *Candida* spp. Gelatin films, as carriers to load econazole nitrate (ECN), were used as an imidazole antifungal agent to treat skin infections and mucosal candidiasis. The plasma treatment completely dissolved the gelatin within 48 hours in a simulated saliva solution, which suggested a potential application for gelatin films as the buccal delivery of econazole to treat oral candidiasis. Different hybrid nanofilms, composed of biopolymer matrices (chitosan, pectin, and chitosanpectin), were prepared and blended with nanostructured lipid carriers (NLC) and loaded in a eutectic mixture of 5% lidocaine-prilocaine (LDC–PLC). These nanohybrid films prolonged LDC-PLC release profile for more than 8 h in vitro. LDC-PLC shows higher drug permeation values across porcine oral mucosa and longer-lasting anesthesia without compromising its safety profile, indicating its significant potential to deliver local anesthetics.

### Other Drug-delivery Systems

In dental tissue engineering, growth factors and scaffolds are widely used to provide a 3D-scaffold structure with a highly porous, interconnected network that enables the transport of cellular nutrients. Polycaprolactone (PCL)/polyactic-co-glycolic acid was combined with amorphous polycaprolactone (PCL) to synthesize scaffolds which were immobilized with adenosviruses onto the scaffold surface to locally deliver gene vectors encoding platelet-derived growth factor-BB or bone morphogenetic protein-7. Controlled scaffold microtopography, combined with localized growth factor gene delivery, improved the regeneration of periodontal bone–periodontal ligament (PDL) interfaces. Chitosan/alginate/PLGA hybrid scaffolds were prepared for control and the sequential delivery of competence factors, such as insulin-like growth factor (IGF-1), progression factor, and bone morphogenetic factor-6 (BMP-6), which induced proliferation and osteoblastic differentiation of cementoblasts for periodontal tissue regeneration.

### Conclusion

Oral drug delivery systems have been designed in different ways to adapt to the physical and chemical environment of oral cavities for more suitable clinical applications. Carrier materials must have good biocompatibility and stable physical and chemical properties. The drugs loaded onto carriers are primarily antibacterial agents and bioactive substances that promote tissue growth. Temporary treatment with antibiotics is helpful in cases with severe symptoms of oral disease. Antibiotics can be used temporarily to suppress the dominant microbiota when severe dysbiosis occurs.

Indeed, we need to be alert to the abuse of antibiotics, which may lead to the dysbiosis of oral microbiota. Therefore, researchers need to pay attention to this issue when preparing carrier materials. Other drugs that preserve ecological balance may be considered for delivery in the future, such as prebiotic substrates. Drug delivery systems can protect active substances in the complex oral environment and control their release to maintain an effective concentration. In general, hydrogels, scaffolds, and nanofibers can be used in tissue regeneration to treat periodontal...
and jaw defects because of their advantageous cell growth support. Carrier materials with good mucosal adhesion, such as films and hydrogels, can be used for mucosal drug delivery. Nanoparticles are often used to load active substances due to their strong dispersion, targeting bacteria or inflammatory cells. They have been widely used as drug delivery systems in the oral cavity to treat oral infectious diseases, such as caries and periodontal diseases. Therefore, effective carrier materials should continuously and steadily release effective drugs and target pathogenic bacteria, especially oral biofilms, to reduce cariogenic conditions without affecting the homeostasis of the oral flora and adapting to the oral environment.

Targeted and specific delivery systems may be the future development direction. For example, using the targeted delivery systems to adjust biofilm pH may prevent caries.

Researchers mainly use organic compounds to prepare carrier materials, but their biocompatibility and degraded byproducts need further verification. However, at present, drug delivery still faces challenges to its design and synthesis, and we lack a clear conclusion about the durability of its effect on resistant bacteria, hindering its clinical delivery. At the same time, there are few in vivo studies and animal experiments, and the biocompatibility of different drug delivery systems should be investigated further. The complexity of the oral environment and the lack of detailed studies on release rates and stability of specific drugs in drug delivery systems have become obstacles to their clinical application.

Conflicts of interest

The authors declare no conflict of interest

Authors’ contributions

Zhang, Yue: Conceptualization (Equal); Data curation (Equal); Formal analysis (Equal); Validation (Equal); Writing – original draft (Equal); Writing – review & editing (Equal). Jiang, Ruining: Conceptualization (Equal); Data curation (Equal); Formal analysis (Equal); Validation (Equal); Writing – original draft (Equal); Writing – review & editing (Equal). Lei, Lei: Conceptualization (Equal); Data curation (Equal); Formal analysis (Equal); Validation (Equal); Writing – original draft (Equal); Writing – review & editing (Equal). Yang, Yingming: Conceptualization (Equal); Data curation (Equal); Formal analysis (Equal); Validation (Equal); Writing – review & editing (Equal). Hu, Tao: Conceptualization (Supporting); Formal analysis (Supporting); Funding acquisition (Lead); Writing – review & editing (Equal).

Acknowledgments

This work was supported by West China Hospital of Stomatology, Research and Develop Program, (RD-03-202002); Sichuan University new Century Higher Education Teaching Reform Project (the ninth phase) research project NO. SCU9361

References

1- Kamrani P, Sadiq NM. Anatomy, head and neck, oral cavity (mouth) [Internet]. In: StatPearls; 2021 [cited January 24, 2022]. Treasure Island (FL): StatPearls Publishing; 2022. Available from: https://www.ncbi.nlm.nih.gov/books/NBK545271/
2- Kitamoto S, Nagao-Kitamoto H, Hein R, Schmidt TM, Kamada N. The bacterial connection between the oral cavity and the gut diseases. J Dent Res. 2020;99(9):1021-9. doi: 10.1177/002203452094633.
3- Rathee M, Sapra A. Dental caries [Internet]. In: StatPearls. In: StatPearls; 2021 [cited January 24, 222]. Treasure Island (FL): StatPearls Publishing; 2022. Available from: https://www.ncbi.nlm.nih.gov/books/NBK551699/
4- Taylor M, Raja A. Oral candidiasis [Internet]. In: StatPearls; 2021 [cited January 24, 222]. Treasure Island (FL): StatPearls Publishing; 2022. Available from: https://www.ncbi.nlm.nih.gov/books/NBK545282/
5- Dashper SG, Mitchell HL, KA LC, Carpenter L, Gussy MG, Calache H, et al. Temporal development of the oral microbiome and prediction of early childhood caries. Sci Rep. 2019;9(1):19732. doi: 10.1038/s41598-019-56233-0.
6- Abusleme L, Dupuy AK, Dutzan N, Silva N, Burleson JA, Straussbaugh LD, et al. The subgingival microbiome in health and periodontitis and its relationship with community biomass and inflammation. Isme J. 2013;7(5):1016-25. doi: 10.1038/ismej.2012.174
7- Philip N, Suneja B, Walsh LJ. Ecological approaches to dental caries prevention: paradigm shift or shibboleth? Caries Res. 2018;52(1-2):153-65. doi:10.1159/000484985
8- Rosier BT, Marsh PD, Mira A. Resilience of the oral microbiota in health: mechanisms that prevent dysbiosis. J Dent Res. 2018;97(4):371-80. doi:1177/0022034517742139
9- Manji F, Dahlen G, Fejerskov O. Caries and periodontitis: contesting the conventional wisdom on their aetiology. Caries Res. 2018;52(6):548-64. doi:11059/000488948
10- Lamont Rj, Koo H, Hajsinghollis G. The oral microbiota: dynamic communities and host interactions. Nat Rev Microbiol. 2018;16(12):745-59. doi:10.1038/s41579-018-0089-x
11- Vos T, Abajobir AA, Abbafati C, Abbas KM, Abate KH, Abd-Allah F, et al. Global, regional, and national incidence, prevalence, and years lived with disability for 328 diseases and injuries for 195 countries, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. Lancet. 2017;390(10100):1211-59. doi:10.1016/S0140-6736(17)32154-2
12- Kinane DF, Stathopoulou PG, Papapanou PN. Periodontal diseases. Nat Rev Dis Primers. 2017;3:17038. doi: 10.1038/nrdp.2017.38

J Appl Oral Sci. 9/12 2022;30:e20210349
activity and mechanism of nanosilver- and nanozinc-incorporated

29- Chen L, Zhou X, He C. Mesoporous silica nanoparticles for tissue-

30- Leng D, Li Y, Zhu J, Liang R, Zhang C, Zhou Y, et al. The antibiofilm
treatments. ACS Nano. 2019;13(5):4869-75. doi: 10.1021/

31- Hua F, Yan J, Zhao S, Yang H, He H. In vitro remineralization of
eosilver- and nanosilver-incorporated mesoporous calcium-silicate nanoparticles. Int J Nanomedicine. 2020;15:3921-36. doi: 10.2147/ijn.S244686

32- Luo XJ, Yang HY, Niu LN, Mao J, Huang C, Pashley DH, et al. Translation of a solution-based biominerilization concept into a carrier-based delivery system via the use of expanded-pore mesoporous silica. Acta Biomater. 2016;31:378-87. doi: 10.1016/j.actbio.2015.11.062

33- Lu MM, Ge Y, Qiu J, Shao D, Zhang Y, Bai J, et al. Redox/ph dual-controlled release of chlorhexidine and silver ions from biodegradable mesoporous silica nanoparticles against oral biofilms. Int J Nanomedicine. 2018;13:7697-709. doi: 10.2147/ijn.S5181168

34- Fan W, Wu D, Tay FR, Ma T, Wu Y, Fan B. Effects of adsorbed and templated nanosilver in mesoporous calcium-silicate nanoparticles on inhibition of bacteria colonization of dentin. Int J Nanomedicine. 2014;9:5217-30. doi: 10.2147/ijn.S573144

35- Fan W, Li Y, Sun Q, Ma T, Fan B. Calcium-silicate mesoporous nanoparticles loaded with chlorhexidine for both anti- Enterococcus faecalis and mineralization properties. J Nanobiotechnology. 2016;14(1):72. doi: 10.1186/s12951-016-0224-7

36- Huang CY, Huang TH, Kao CT, Wu YH, Chen WC, Shie MY. Mesoporous calcium silicate nanoparticles with drug delivery and odontogenesis properties. J Endod. 2017;43(1):69-76. doi: 10.1016/j.

37- Allaker RP. The use of nanoparticles to control oral biofilm formation. J Dent Res. 2010;89(11):1175-86. doi: 10.1177/0022034510777794

38- Allaker RP, Memarzadeh K. Nanoparticles and the control of oral infections. Int J Antimicrob Agents. 2014;43(2):95-104. doi: 10.1016/j.

39- Dizaj SM, Lotfipour F, Barzegar-Jalali M, Zarrintan MH, Adlikia K. Antimicrobial activity of the metals and metal oxide nanoparticles. Mater Sci Eng C Mater Biol Appl. 2014;44:278-84. doi: 10.1016/j.msec.2014.08.031

40- Mathur P, Jha S, Ramteke S, Jain NK. Pharmaceutical aspects of silver nanoparticles. Artif Cells Nanomed Biotechnol. 2018;46(sup1):115-26. doi: 10.1080/21691401.2017.1414825

41- Tang H, Xu M, Zhou X, Zhang Y, Zhao L, Ye G, et al. Acute toxicity and biodistribution of different sized copper nano-particles in rats after oral administration. Mater Sci Eng C Mater Biol Appl. 2018;93:649-63. doi: 10.1016/j.msec.2018.08.032

42- Ding D, Zhu Q. Recent advances of PLGA micro/nanoparticles for the delivery of biomacromolecular therapeutics. Mater Sci Eng C Mater Biol Appl. 2018;92:1041-60. doi: 10.1016/j.msec.2017.12.036

43- Kumar G, Shafigh N, Malhotra S. Drug-loaded PLGA nanoparticles for oral administration: fundamental issues and challenges ahead. Crit Rev Ther Drug Carrier Syst. 2012;29(2):149-82. doi: 10.1615/
critrevtherdrugcarrierssyst.v29.i2.20

44- Mir M, Ahmed N, Rehman AU. Recent applications of PLGA based nanostructures in drug delivery. Colloids Surf B Biointerfaces. 2017;159:217-31. doi: 10.1016/j.colsurfb.2017.07.038

45- Rizae BR, Younes NH, Rasool K, Nasrallah G. Synthesis, bioapplications, and toxicity evaluation of chitosan-based nanoparticles. Int J Mol Sci. 2019;20(22). doi: 10.3390/ijms2025776

46- Priyadarshini BM, Mitali K, Lu TB, Handral HK, Dubey N, Fawzy AS. PLGA nanoparticles as chlorhexidine-delivery carrier to resin-dentin adhesive interface. Dent Mater. 2017;33(7):830-46. doi: 10.1016/j.
dental.2017.04.015

47- Fawzy AS, Priyadarshini BM, Selvan ST, Lu TB, Neo J. Pantothenoyl-cyanoalanine-loaded nanoparticles enhance dentin degradation resistance. J Dent Res. 2017;96(7):780-9. doi: 10.1177/0022034517691757

48- Lin HP, Tu HP, Hsieh YP, Lee BS. Controlled release of lovastatin from poly(lactic-co-glycolic acid) nanoparticles for direct pulp capping in rat teeth. Int J Nanomedicine. 2017;12:5473-85. doi: 10.2147/ijn.S138410
56- Chang PC, Tai WC, Luo HT, Lai CH, Lin HH, Lin ZJ, et al. Core-Shell poly-(D,L-Lactide-co-Glycolide)-chitosan Nanospheres with simvastatin-doxycycline for periodontal and osseous repair. Int J Biol Macromol. 2020;158:627-35. doi: 10.1016/j.ijbiomac.2020.04.183
57- Spencer PP, Shah SR, Henslee AM, Watson BM, Kiniard LA, Kretlow JD, et al. Evaluation of antibiotic releasing porous poly(methylmethacrylate) space maintainers in an infected composite tissue defect model. Acta Biomater. 2013;9(11):8832-9. doi: 10.1016/j.actbio.2013.07.018
58- Priyadarshini BM, Selvan ST, Lu TB, Xie H, Neo J, Fawzy FC, Tofoli GR, et al. Recent advances and perspectives in topical drug delivery for oral cancer therapy. Molecules. 2019;24(10):24193521.
59- Chronopoulou L, Nocca G, Castagnola M, Paludetti G, Ortaggi GM, Sciubba F, et al. Chitosan based nanoparticles functionalized in targeted drug delivery. Molecules. 2019;24(3). doi: 10.3390/molecules24030603
60-ASE. Chlorhexidine nanocapsule drug delivery approach to the toothpaste actives and adhesion on tooth analogs. 2007;106(6):4248-56. https://doi.org/10.1002/app.27078
61- Nguyen S, Escudero C, Sediqi N, Smisdat G, Hiorth M. Fluoride loaded polymeric nanoparticles for dental delivery. Eur J Pharm Sci. 2017;104(11):3913-24. doi: 10.1016/j.ejps.2017.04.004
62- Chronopoulou L, Nocca G, Castagnola M, Paludetti G, Ortaggi GM, Sciubba F, et al. Chitosan based nanoparticles functionalized with peptidomimetic derivatives for oral drug delivery. N Biotechnol. 2016;33(1):23-31. doi: 10.1016/j.nbt.2015.07.005
63- Priyadarshini BM, Selvan ST, Lu TB, Xie H, Neo J, Fawzy AS. Chlorhexidine nanocapsule drug delivery approach to the resin-dentin interface. J Dent Res. 2016;95(9):1065-72. doi: 10.1177/002203451665135
64- Shrestha S, Kishen A. Temporal-controlled bioactive molecules releasing core-shell nano-system for tissue engineering strategies in endodontics. Nanomedicine. 2019;18:11-20. doi: 10.1016/j.nano.2019.02.013
65- Zhu B, Li X, Xu X, Liu J, Ding C, Zhao C, et al. One-step phosphorylated poly(amide-amine) dendrimer loaded with apigenin for simultaneous remineralization and antibacterial of dentine. Colloids Surf B Biointerfaces. 2018;172:760-8. doi: 10.1016/j.colsurfb.2018.09.036
66- Wang W, Dang M, Zhang Z, Hu J, Eyster TW, Ni L, et al. Dentin regeneration by stem cells of apical papilla on injectable nanofibrous microspheres and stimulated by controlled BMP-2 release. Acta Biomater. 2016;36:63-72. doi: 10.1016/j.actbio.2016.03.015
67- Omainty R, Onida B, Pérez P, Longhi M, Barresi A, Granero G. Targeted chitosan-based bioinamocomposites for controlled oral mucosal delivery of chlorhexidine. Int J Pharm. 2016;509(1-2):408-18. doi: 10.1016/j.ijpharm.2016.06.011
68- Park SC, Kim YM, Lee JK, Kim NH, Kim EJ, Heo H, et al. Targeting and synergistic action of an antianginal peptide in an antibiotic drug delivery system. J Control Release. 2017;256:46-55. doi: 10.1016/j.jconrel.2017.04.023
69- Vidal-Romero G, Zambrano-Zaragoza ML, Martinez-Acevedo L, Leyva-Gómez G, Mendoza-Elvira SE, Quintanar-Guerrero D. Design and evaluation of pH-dependent nanosystems based on cellulose acetate phthalate, nanoparticles loaded with chlorhexidine for periodontal treatment. Pharmaceutics. 2019;11(11). doi: 10.3390/pharmaceutics1110604
Drug delivery systems for oral disease applications

82- Chen W, Zhi M, Feng Z, Gao P, Yuan Y, Zhang C, et al. Sustained co-delivery of ibuprofen and basic fibroblast growth factor by thermosensitive nanoparticle hydrogel as early local treatment of peri-implantitis. Int J Nanomedicine. 2019;14:1347-58. doi: 10.2147/ijn.S190781

83- Tan J, Zhang M, Hai Z, Wu C, Lin J, Kuang W, et al. Sustained release of two bioactive factors from supramolecular hydrogel promotes periodontal bone regeneration. ACS Nano. 2019;13(5):5616-22. doi: 10.1021/acsnano.9b00788

84- Pan J, Deng J, Luo Y, Yu L, Zhang W, Han X, et al. Thermosensitive hydrogel delivery of human periodontal stem cells overexpressing platelet-derived growth factor-bb enhances alveolar bone defect repair. Stem Cells Dev. 2019;28(24):1620-31. doi: 10.1089/scd.2019.0184

85- He XT, Li X, Xia Y, Yin Y, Wu RX, Sun HH, et al. Building capacity for macrophage modulation and stem cell delivery in high-stiffness hydrogels for complex periodontal regeneration: experimental studies in vitro and in rats. Acta Biomater. 2019;88:162-80. doi: 10.1016/j.actbio.2019.02.004

86- Silva CR, Babo PS, Gulino M, Costa L, Oliveira JM, Silva-Correia J, et al. Injectable and tunable hyaluronic acid hydrogels releasing chemoattractant and angiogenic growth factors for endodontic regeneration. Acta Biomater. 2018;77:155-71. doi: 10.1016/j.actbio.2018.07.035

87- Moon CY, Nam OH, Kim M, Lee HS, Kaushik SN, Cruz Walma DA, et al. Effects of the nitric oxide releasing biomimetic nanomatix gel on pulp-dentin regeneration: Pilot study. PLOS One. 2018;13(10):e0205534. doi: 10.1371/journal.pone.0205534

88- Kong EF, Tsui C, Boyce H, Ibrahim A, Hoag SW, Karlsson AJ, et al. Development and in vivo evaluation of a novel histatin-5 bioadhesive hydrogel formulation for oral candidiasis. Antimicrob Agents Chemother. 2016;60(2):881-9. doi: 10.1128/aac.02624-15.

89- Ribeiro LN, Franz-Montan M, Breitkreitz MC, Rodrigues da Silva GH, Castro SR, Guilherme VA, et al. Nanohybrid hydrogels designed for transbuccal anesthesia. Int J Nanomedicine. 2018;13:6453-63. doi: 10.2147/ijn.S180080

90- H RR, Dhameca D, Jagwani S, Patil D, Hegde S, Potdar R, et al. Formulation of thermoreversible gel of cranberry juice concentrate: Evaluation, biocompatibility studies and its antimicrobial activity against periodontal pathogens. Mater Sci Eng C Mater Biol Appl. 2017;75:1506-14. doi: 10.1016/j.msec.2017.03.054

91- Sousa MG, Maximiano MR, Costa RA, Rezende TM, Franco OL. Nanofibers as drug-delivery systems for infection control in dentistry. Expert Opin Drug Deliv. 2020;17(7):919-30. doi: 10.1080/0302252.2019.1642912

92- Chen X, Li H, Lu W, Guo Y. Antibacterial porous coaxial drug-carrying nanofibers for sustained drug-releasing applications. Nanomaterials. 2019;9(1):371. doi: 10.2147/ijn.S238476

93- Dolci LS, Liguori A, Panzavolta S, Miserocchi A, Passerini N, Gherardi D, et al. Non-equilibrium atmospheric pressure plasma as innovative method to crosslink and enhance mucoadhesion of econazole-loaded niosomes as oromuco-adhesive films: A randomized clinical trial of a therapeutic drug delivery platform for the treatment of oral recurrent aphthous ulcers. Sci Rep. 2018;8(18):18056. doi: 10.1038/s41598-018-37157-7

94- Ibrahim YH, Regdon G Jr, Kristó K, Kelemen A, Adam ME, Hamedelniel EI, et al. Design and characterization of chitosan/citrate films as carrier for oral macromolecule delivery. Eur J Pharm Sci. 2020;146:105270. doi: 10.1016/j.ejps.2020.105270

95- He XT, Hölösvy S, Štembírek J, Pazdziora E, Landová H, Doležel P, et al. Carmellose mucoadhesive oral films containing vermiculite/chlorhexidine nanocomposites as innovative biomaterials for treatment of oral infections. Biomed Res Int. 2015;2015:580146. doi: 10.1155/2015/580146

96- Sari EF, Tsui C, Boyce H, Ibrahim A, Hoag SW, Karlsson AJ, et al. Therapeutic drug delivery platform for the treatment of oral recurrent aphthous ulcers. Sci Rep. 2018;8(18):18056. doi: 10.1038/s41598-018-37157-7

97- Spolnik KJ, Nör JE, et al. A novel patient-specific three-dimensional drug delivery construct for regenerative endodontics. J Biomed Mater Res B Appl Biomater. 2019;107(5):1576-86. doi: 10.1002/jbmb.34250

98- Bottino MC, Albuquerque MT, Azabi A, Münchow EA, Spolnik KJ, Nör JE, et al. A novel patient-specific three-dimensional drug delivery construct for regenerative endodontics. J Biomed Mater Res B Appl Biomater. 2019;107(5):1576-86. doi: 10.1002/jbmb.34250

99- Bottino MC, Kamocki K, Yassen GH, Platt JA, Vail MM, Ehrich Y, et al. Bioactive nanofibrous scaffolds for regenerative endodontics. J Dent Res. 2013;92(11):963-9. doi: 10.1177/0022034513505770

100- Albuquerque MT, Ryan SJ, Münchow EA, Kamocka MM, Gregory RL, Valera MC, et al. Antimicrobial effects of novel triple antibiotic paste-mimic scaffolds on actinomyces naeslundii biofilm. J Endod. 2015;41(8):1337-43. doi: 10.1016/j.joen.2015.03.005