Visualization of Primary Particles in a Tablet Based on Raman Crystal Orientation Mapping

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Abstract

Objective: The morphology of the primary particles in the active pharmaceutical ingredient (API) is one of the most important determinants for formulation function. However, it has not been possible to visualize primary particles in a tablet using various imaging methods, including spectroscopic mappings, because these particles usually exist as aggregated clusters in tablets. We revealed that the Raman spectrum of crystalline particles is determined depending upon the angle between the crystallographic axis and the polarization direction of the excitation laser. In this paper, we report a method to visualize primary particles within the aggregated cluster based on the Raman spectral change on the boundary of primary particles.

Method: Metformin hydrochloride was chosen as a model API for this study. The crystal structure of metformin hydrochloride was solved using X-ray crystal structure analysis. The Raman spectra of metformin hydrochloride crystal along the xyz axes were recorded and resolved into components along the abc axes. Using the abc components, Raman mapping of metformin hydrochloride in tablets was performed to visualize the crystal orientation at each data point.

Results: Metformin hydrochloride crystals recrystallized from water/ethanol formed a primitive monoclinic cell. Datasets of five distinct peak areas from the metformin hydrochloride Raman spectrum were used for analyses. Raman crystal orientation mapping (RCOM) from the tablet cross-section provided an image of primary particles within the aggregation cluster of metformin hydrochloride in the tablet.

Conclusion: Based on the RCOM, we developed a visualization method for primary API particles in tablets. Because the morphology of primary particles is the key factor of formulation function, this method would contribute to better formulation development and quality control.

Keywords: Raman spectroscopy; Raman mapping; Primary particle; Formulation; Metformin

Introduction

Raman chemical imaging has become a powerful tool for understanding solid formulations by visualizing the dispersion of active pharmaceutical ingredients (API) [1-6] or excipients [7]. However, particularly in high dose formulations, the image of API is usually the image of the aggregation cluster. At this stage, it is not possible to resolve the image to the level of the primary particles by Raman imaging. The morphology of primary particles is one of the key factors in determining the formulation functions [8,9]. Development of imaging techniques for the primary particles in the aggregation would greatly contribute to formulation research and quality control.

Using an antiepileptic agent, phenytoin, as a model API, we have revealed that the peak area of the Raman spectrum changes depending upon the angle of the crystallographic axes and polarization direction of the excitation laser [10]. In our report, we showed that the crystal face of the API can be identified according to the combination of Raman spectra where the polarization direction was horizontal and vertical to the morphological long axis of a target face. Applying this to Raman mapping, we suggested that mapping of the angle between the crystallographic axes and the polarization direction could be achieved. Because the axes orientation within a certain primary particle is constant against the polarization direction and this obviously changes on the boundary between particles, the Raman mapping of the axes orientation (Raman crystal orientation mapping (RCOM)) could be used to visualize the primary particle image within the aggregated cluster of API.

In this report, we showed the first attempt at RCOM and the primary particle imaging for a high dose solid formulation of anti-diabetic agent, metformin hydrochloride. We discussed the feasibility of this imaging method for better formulation development and quality control.

Materials and Methods

X-ray single-crystal structural analysis of metformin hydrochloride

Metformin hydrochloride (Tokyo Kasei, Tokyo, Japan) (Figure 1A) was recrystallized from water/ethanol to obtain a crystal for X-ray single-crystal structural analysis. A colorless prism crystal of metformin hydrochloride (Figure 1B) having approximate dimensions of 0.120 × 0.100 × 0.050 mm was mounted in a loop. All measurements were made on a Rigaku Saturn724 diffractometer (Rigaku Corporation, Japan).

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Tokyo, Japan) using a multi-layer mirror monochromated Mo-Kα radiation.

Raman spectra measurement and resolution into abc components

The xyz axes of coordinates were defined so that the xy plane was on the (1 -1 0) face and the x axis was parallel to the morphologically long axis of the crystal (Figure 1C). Raman spectra were recorded on an inVia Raman microscope system (Renishaw plc, Gloucestershire, UK) equipped with a Leica microscope and a 785 nm, 300 mW excitation lasers. 50X objective lens was used and the laser exposure time was 1 s. The datasets of peak areas of five distinct peaks in the Raman spectrum were converted to 0-255 integer. The text images corresponding to the xyz axes of coordinates on the morphology of metformin hydrochloride crystal, (D) the unit cell and the abc axes of metformin hydrochloride crystal. Yellow axes indicate xyz axes in (C).

Figure 1: (A) Chemical structure of metformin hydrochloride. (B) microscopic view of metformin hydrochloride crystal, (C) definition of the xyz axes of coordinates on the morphology of metformin hydrochloride crystal, (D) the unit cell and the abc axes of metformin hydrochloride crystal. Yellow axes indicate xyz axes in (C).

A metformin hydrochloride tablet (METGLUCO® 250 mg, Sumitomo Dainippon Pharma, Osaka, Japan) was fixed on a glass plate with instant glue and milled with a milling machine (Proxxon GmbH, Niersbach, Germany) to expose the cross-section. Raman mapping was performed with Renishaw inVia Raman microscope under a 785 nm, 300 mW excitation laser with 50× objective lens. The laser exposure time was 1s and the mapping area was 250 μm × 250 μm in a spatial resolution of 2 μm (15,676 data points in total). The areas of the five distinct peaks of metformin hydrochloride were calculated for each data point and a matrix B (15876 × 3) was defined. The abc component of each data point, matrix D (15876 × 3), was calculated as follows:

\[ B = D K \]

\[ D = B \cdot K^T \cdot \left( K \cdot K^T \right)^{-1} \]

Image formation from abc component

Components in each column of D were then normalized to 0-255 integer to form a matrix D’. Each column of D’ was transferred to ImageJ software (National Institutes of Health, Bethesda, MD, USA) as a text image and converted to a picture image. The a, b, and c axis component images were colored by red, green, and blue (RGB), respectively, and the three images were merged.

Results and Discussion

Crystal structure of metformin hydrochloride

Structure solution and refinement of the metformin hydrochloride crystal were performed as described in the Materials and Methods. A summary of the crystal data is presented in Table 1. It was found that the metformin hydrochloride crystal formed a primitive monoclinic cell and the widest plane was the (1-1 0) face (Figure 1C). The structural data have been deposited with The Cambridge Crystallographic Data Centre (CCDC: 143396). Being different from the structure of the phenytoin crystal [10], the unit cell of metformin hydrochloride was not a rectangular solid and was settled with a tilt against morphologically xyz axes (Figure 1D). Thus, it was necessary to perform matrix calculations to resolve the abc component as shown in Materials and Method.

Raman crystal orientation mapping (RCOM)

In this study, we developed a novel Raman mapping method, RCOM. The abc component on each data point was resolved and converted to 0-255 integer. The text images corresponding to the a, b, and c axes were transformed to RGB images and merged. Images obtained in this way (RCOM images) represent how much each component contributes to the Raman spectrum on a data point, and this indicates how much each crystallographic axis is parallel to the polarization direction of the excitation laser (that is, the image reveals the orientation of axes). As described in the Introduction, because the axes’ orientation changes on the boundary between primary particles, the RCOM image represents the primary particle image within the aggregated cluster of API. Figure 3B shows the RCOM image of the cross-section of metformin hydrochloride tablet. The difference in color reflects the difference of crystal orientation. For example, the blue region indicates that the c-axis is parallel to the polarization direction, and the green region indicates that the b-axis is parallel, and the boundary of color is the boundary of primary particles. Thus, we can understand the morphology such as the particle size of API within the aggregation in a solid formulation. Compared with the typical Raman imaging [11] using averaged metformin hydrochloride Raman spectrum as an index (Figure 3A), the RCOM image provided more detailed information of API particle in the solid formulation.
Application of RCOM imaging to formulation development and quality control

The RCOM imaging method is applicable to any other API if API is crystalline and the direction of crystallographic axes is clarified using X-ray structural analysis. Whether the solid consists of a pure component or mixture or whether it is before or after tableting is not relevant in recording the ROM image. No special software for calculation is necessary. This imaging method could help formulation development. For example, the distribution of the primary particle size after forming a test tablet could be assessed, which is important information for the dissolving performance of a poorly soluble API. For formulation quality control, it would also provide more useful information, such as the uniformity of the primary particle size or the aggregation states in tablet, than does the typical chemical imaging methods. Assessment of primary particle size distribution of API in tablets based on the RCOM imaging is now under investigation in our laboratory.

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