MASCOLINIZATION OF TILAPIA (OREOCHROMIS NILOTICUS) BY IMMERSION METHOD USING METHANOL EXTRACT OF PASAK BUMI ROOTS (EURYCOMA LONGIFOLIA JACK)

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ABSTRACT
The use of synthetic steroids of 17-Methytestosterone (MT) to produce a monosexual population of male Tilapia (Oreochromis niloticus) has been restricted because it can leave harmful residues in fish and the environment. One alternative that can be done is by using natural bioactive derived from plants with androgenic properties, such as Pasak Bumi (Eurycoma longifolia Jack). The aim of this research was to assay the use of methanol extract from Pasak bumi roots with immersion techniques for masculinization of tilapia. The research procedure was carried out with three treatment doses of methanol extract from the Pasak Bumi roots (namely 30, 60, and 90 mg/L), treatment dose amount to 0 mg/L, acted as a negative control and treatment of MT amounted to 50 mg/L acted as a positive control, with three replications on each treatment. Immersion was applied to larvae that was hatched out of egg yolk (seven day-old) for 24 hours and was maintained with a capacity of 50 larvae per aquarium during 60 days. The results showed that the treatment of the methanol extract of Pasak Bumi roots gave male sex ratios ranging from 72.66% - 76.33% higher than the negative control which showed the results of male sex ratios amounted to 49.19% and lower than the positive control of MT which the results amounted to 83.99%. The range of survival rate was from 80.67% - 85.33%, while the range of survival rate in the negative control was 83.33% and in positive control of MT was 82.67%. Thus, the methanol extract of Pasak Bumi roots can be used for masculinization of tilapia by immersion method with the best performance at a dose of 60 mg/L – 90 mg/L.

KEY WORDS
Monosex culture, pasak bumi, sex reversal, testosterone, tilapia fish.

Tilapia (Oreochromis niloticus) is one of the main consumption fish in Indonesia that has beneficial biological properties because it can consume various types of natural food, can grow well in all kinds of waters and has a high tolerance for low water quality. But the tilapia culture is disrupted by gonad which becomes mature too quickly, which can cause unwanted reproduction (wild breeding) which results in slow growth. One method to overcome the wild breeding is monosex culture (in this case, male fish) because the type of male tilapia has a growth rate of almost two times faster than that of female tilapia.

One method used to get male monosex fish is by using androgen hormones. One of the hormones that has commonly been used for maleization is 17a-methytestosterone. However, the use of MT has been limited because it can leave harmful residues in fish and the environment (Pandian & Kirankumar, 2003). The use of the hormone of 17-methyltestosterone in Indonesia has been banned (KKP 2014). One alternative that can be done is by using natural products derived from plants that are androgenic properties, such as Pasak bumi.
Pasak bumi (*Eurycoma longifolia* Jack) is used as an active drug for men or it is commonly referred with the term aphrodisiac. Pasak bumi can increase the levels of testosterone, LH (Luteinizing Hormone), and FSH (Follicle Stimulating Hormone). Pasak bumi contains phytosterol, which has androgenic properties. The most common phytosterol found in plants is β-sitosterol and stigmasterol. β-sitosterol has an estrogenic effect that can increase vitellogenesis in female and male Rainbow Trout fish. Male fish liver cells are able to produce vitellogenin (Tremblay and Van der Kraak, 1998). Parks et al. (2001) reported that phytosterols isolated from paper pulp waste are able to masculinize Mosquitofish females (*Gambusia affinis* holbrooki). Masculinization using Purwaceng’s (*Pimpinella alpina*) extract which is thought to contain stigmasterol is able to increase the population of male Tilapia (Putra, 2011), Betta fish (*Betta splendens*) (Bulkini, 2012 and Cahyani 2014) and Rainbow Fish (*Liatherina werneri*) (Nurkhasanah, 2015).

The sex reversal application is performed to reverse the development of gonad sex in fish, from female to male. This process is carried out when the fish gonad has not been differentiated between male and female at the time of hatching. The gonad tissue of Teleostei fish when differentiated is very unstable. It is sensitive to environmental factors such as exogenous steroid hormones or xenobiotics (Govoroun et al., 2001). Piferrer (2001), the sensitivity of steroid hormones to the development of sex differentiation is very dependent on the phase of gonad development that occurs. In this case, the peak sensitivity occurs after the cell division phase of the gonad tissue or before differentiated gonad tissue. This phenomenon, according to Carman et al. (1998), occurs because the function of the sex chromosome in determining gender is not active yet.

The process of female sex differentiation in fish occurred when the P450 aromatase enzyme is produced during times of sex differentiation. P450 aromatase enzymes work to catalyze changes in androgen hormones to estrogen, so the P450 aromatase enzyme determines the balance between androgen and estrogen hormones. The process of inhibiting P450 aromatase activity at the receptor level during the period of sex differentiation causes changes in the female phenotype into the male phenotype (Pandian, 2013). Inhibition of CYP19 activity can be done by the administration of steroid hormones or with manipulating environment temperature (Bowman et al. 2012).

**MATERIALS AND METHODS OF RESEARCH**

The research was conducted for 4 months in the Fish Domestication and Breeding Laboratory, CV. Griya Aquatica - Fish Farming, Palangka Raya, Central Kalimantan.

This research consisted of three treatments and two control treatments. Each treatment was repeated three times. Each treatment applies different amount of dose of methanol extract of the Pasak Bumi roots by the immersion method. Treatment that was applied to tilapia larvae, namely:

- Treatment without methanol extract of Pasak Bumi roots, as a negative control TH (K-);
- Treatment with 30 mg/L of methanol extract of Pasak Bumi roots (EPB1);
- Treatment with 60 mg/L of methanol extract of Pasak Bumi roots (EPB2);
- Treatment with 90 mg/L of methanol extract of Pasak Bumi roots (EPB3);
- Treatment with 50 mg/L of 17α-Methyltestosterone, as a positive control of MT (K+).

The Pasak bumi (*Eurycoma longifolia* Jack) roots were obtained from Rakumpit Subdistrict, Palangka Raya, Central Kalimantan. The sample was dried and ground into flour. Then some of the Pasak Bumi root flour was wrapped in filter paper. Then it was extracted using the Soxhlet method with methanol solvent. Soxhlet ran at 70°C for 4 hours, so the solvent was evaporated by the condenser and dropped to the sample column. The extraction results were evaporated by using a rotary evaporator at 40°C to get the Pasak bumi extracts. Extracts that have been in the form of pasta were weighed according to the dose of treatment. 1,095 g of Pasak Bumi roots flour produced as much as 54.98 g of methanol extract with 4.93 % percentage of extract mass. Then the Pasak Bumi methanol extract was analyzed by GC-MS (Table 1). The methanol extract of Pasak Bumi and 17α-
methyltestosterone also observed nanostructures with scanning electron microscope (SEM) (Figure 1).

Table 1 – Results of GC-MS analysis of Methanol Extract of Pasak Bumi

| No. | Compounds Name                                                        | Suitability Factors (%) | Total (%) |
|-----|-----------------------------------------------------------------------|-------------------------|-----------|
| 1.  | 3-(2-pentenyl)-1,2,4-cyclopentanetetron                                | 53                      | 5.80      |
| 2.  | Palmitic acid methyl ester                                            | 98                      | 1.63      |
| 3.  | Palmitic acid                                                         | 99                      | 10.39     |
| 4.  | 4-ethoxy-2,5-dimethoxybenzaldehyde                                    | 60                      | 2.23      |
| 5.  | Isophraxidine / 6,8-dimethoxy-7-hydroxy kumarin                        | 96                      | 1.55      |
| 6.  | Methyl ester oleic acid                                               | 99                      | 1.44      |
| 7.  | Linoleic acid                                                         | 96                      | 1.33      |
| 8.  | Oleic acid                                                            | 99                      | 4.78      |
| 9.  | Octadecanoic acid                                                     | 97                      | 1.16      |
| 10. | 4-(5-propyl-2-pyrindyl) -benzonitrile                                  | 56                      | 7.95      |
| 11. | Canthine-6-on/3,4-diazfluoranthem-2(3H)-on                            | 94                      | 4.21      |
| 12. | m-cresol / m-toluol / 3-methylphenol                                   | 64                      | 8.80      |
| 13. | Disookylphthalate                                                     | 91                      | 1.91      |
| 14. | 4-phenyl-8-oxo,4,5,6,7-tetrahydroxyclopenta (b) -1,2,3-triazolo (4,5-e) pyridine | 90                      | 6.98      |
| 15. | 2-methyl-4,4-diphony-2-imidazolinen-5-on                              | 91                      | 3.95      |
| 16. | Methyl ester,1,2-dimethyl-ferocarboxylic acid                         | 49                      | 4.73      |
| 17. | Methyl ester,4,5-dihydroxy-4-(3-methyl-2-butyl benzoic acid            | 46                      | 2.48      |
| 18. | Unknown                                                               | 25                      | 0.93      |
| 19. | 3-[(trimethylsilyl) oxy] - acetate (17.beta) -esta-1,3,5, (10) -trienn-17-ol | 87                      | 1.12      |
| 20. | 5-heptadekatri-9(2),11 (2), 14 (Z)-enilresorcinol                      | 80                      | 1.12      |
| 21. | Piperin                                                               | 99                      | 1.12      |
| 22. | Salisilidhe / beta resorcinaldehyde                                   | 49                      | 1.21      |
| 23. | Metilenetaninhkunion                                                  | 41                      | 1.23      |
| 24. | Unknown                                                               | 10                      | 1.10      |
| 25. | Stigmasta-5,23-dien-3-beta,-ol                                        | 64                      | 5.97      |
| 26. | Unknown                                                               | 15                      | 2.85      |
| 27. | Unknown                                                               | 18                      | 1.53      |
| 28. | Unknown                                                               | 15                      | 1.17      |
| 29. | Spinasteron                                                           | 53                      | 4.62      |
| 30. | Sitosterone / delta.4-sitosterol-3-on                                 | 92                      | 3.22      |
| 31. | Aurantiacid                                                           | 46                      | 1.52      |

Figure 1 – Characterization and Nanostructures Analysis of methanol extract of Pasak Bumi roots and 17a-methyltestosterone with Scanning Electron Microscope (SEM). Magnification of 500x

The containers used in this research were 15 aquarium units with a size of 60 x 40 x 30 cm. The aquarium was filled with water that has been deposited in a reservoir for one week. Water replacement was carried out with a recirculation system.

Tilapia (Oreochromis niloticus) larvae was obtained from the People’s Hatchery Unit (UPR) in Bincau Village, Banjar Regency, South Kalimantan. Retrieval of larvae at 1-day old larvae (egg yolks still exist), for 5-day old, they were kept in the aquarium until egg yolks run out (7-day old), weighing 0.01 - 0.02 g/larvae. Larvae that have run out of egg yolks was collected and soaked for 24 hours in Methanol Extract of Pasak Bumi according to the
dosage of treatment. Subsequently, the larvae was transferred to an aquarium with a density of 50 larvae per aquarium.

Tilapia larvae was fed with shrimp flour with a protein content of 45%, 3-4 times a day with a 5% feeding rate of biomass for 60 days of culture.

Measuring water quality consisted of water temperature, dissolved oxygen, pH and NH₃. Temperature was measured using a thermometer, dissolved oxygen was measured using DO meter, pH was measured with pH meter and NH₃ was measured using a spectrophotometer. The parameters of water temperature, dissolved oxygen and pH were measured every day while NH₃ parameters were measured once a week. The temperature during maintenance ranged from 28 - 30°C, dissolved oxygen ranged from 5.4 to 6.8 mg/L, pH ranged from 6.2 to 7.6 and dissolved ammonia ranged from 0.01 - 0.032 mg/L. Sampling growth and survival rate were carried out every 15 days until the 60 days of trial.

Gonad examination was carried out after the fish was 60 day-old by taking a sample of 30% of the population of assay fish. The fish was dissected and gonad was taken carefully by using tweezers. A portion of the gonad was placed on the object-glass and then was chopped using a scalpel until the portion became smooth. Then the acetocarmine solution was added. Gonad preparation slides were observed under a binocular microscope with 400X magnification. Some of the gonad samples were made into histological preparations by hematoxylin-eosin (HE) staining. Then their structure was examined using the histological method of exploration.

Measurement of testosterone levels was carried out before and after immersion on 0, 15, 30, 45 and 60 days. Testosterone levels were measured using the ELISA (Enzym linked immunosobent assay) method, which is an immunoassay method that uses enzymes as a label.

The variance (ANOVA) of the results was analyzed using the STATISTICA 8 at a 95% confidence level. The data was tabulated with the MS Office Excel 2010. Significantly different treatments were analyzed by using Duncan's technique.

RESULTS AND DISCUSSION

Masculinization of tilapia by immersion method produced the highest percentage of males in the positive control (MT) of 83.99 ± 3.33%. Then in EPB3, it was 76.36 ± 2.79%; in EPB2, it was 75.80 ± 2.78%; in EPB1, it was 72.66 ± 2.84% and in negative control (K-), it was 52.19 ± 0.00% (Figure 2). The ANOVA results showed that immersion with methanol extract of Pasak Bumi roots with different doses showed a very significant effect (p < 0.05) on the percentage (%) of male tilapia. Duncan’s analysis results showed EPB1, EPB2 and EPB3 were not significantly different but were significantly different from MT (K+) and K-.

![Figure 2 – Percentage (%) of Male Tilapia by Immersion Method Using methanol extract of Pasak Bumi roots](image-url)
Figure 2 showed the sex ratio of male tilapia using methanol extract of Pasak Bumi roots was higher than negative control, but lower than positive control (MT). Based on the medicinal properties contained in the Pasak Bumi plant, aphrodisiac played a role in fish masculinization activities. Aphrodisiacs are medicines, food, drinks, and odors that can cause or increase sexual activity (Taufiqqurrachman, 1999). Pasak bumi roots methanol contained phytosterol compounds, namely stigmasterol from the results of GC-MS analysis (Table 1).

Stigmasterol stimulated an increase in androgen hormones in the body. In addition, a phytochemical screening of Pasak Bumi roots extract contains saponins, phytosterols, alkaloids, and oligosaccharides. The steroid saponin compound is the essential ingredients of the sex hormone industry product. Based on the compounds found in Pasak Bumi roots, stigmasterol, and saponin steroid compounds are thought to improve the quality and male sexual behavior after consuming them (Taufiqqurrachman, 1999). The increase in male sex ratio in tilapia is believed to be related to the effect of the stigmasterol compound on the affinity of the androgen receptor so that it can work as an androgen. Tremblay & Van Der Kraak (1998) suggested that stigmasterol compounds can influence the affinity power of androgen receptors.

The highest intersex tilapia found in EPB1 was at 9.99 ± 2.84%, and followed by EPB3 at 9.52 ± 2.79%; positive control (MT) was 7.97 ± 3.33%; EPB2 was 7.83 ± 2.78%, whereas negative control did not find intersex fish (Figure 3). Analysis of variance (ANOVA) showed that the treatment of methanol extract of Pasak Bumi roots with a different dose had a significant effect (P < 0.05) on the intersex of tilapia. Duncan’s analysis showed that EPB1 was not significantly different from EPB2, EPB3, and positive control (MT), but very significantly different from negative control (K-).

![Figure 3 – Intersex Percentage (%) of Tilapia by Immersion Method Using methanol extract of Pasak Bumi roots](image)

Figure 3 showed 30, 60, and 90 mg/L treatments and 50 mg/L positive control (MT) found intersex fish, whereas in the negative control, no intersex fish was found. The intersex value ranged from 7.83 to 9.99%. The highest intersex value occurred at 30 mg/L. It suspected that the content of the stigmasterol compound in the extract of Pasak Bumi given by immersion is still low. This was confirmed by Nakamura et al. (1998) which explained that the administration of steroid hormones with lower doses not able to form males ideally so that it can cause the formation of intersex fish. The intersex phenomenon also occurred in Putra (2011), which showed tilapia with a dose of 20mg/L purwaceng extract experienced an intersex of 13.3% with 8 hours of immersion treatment. Ghosal & Chakraborty (2014) obtained the percentage of tilapia that experienced an intersex of 7.2% in the treatment of Tribulus terrestris by immersion method with a dose of 0.05 g/L at three day-old.
The survival rate of Tilapia on the 60 days showed the highest survival rate in EPB2 of \(85.33 \pm 3.06\)\% and subsequently in EPB3 of \(84.67 \pm 4.16\)\%; negative control of \(83.33 \pm 4.62\)\%; positive control (MT) of \(82.67 \pm 4.16\)\%, and EPB1 of \(80.67 \pm 3.06\)\% (Figure 4). Analysis of Variance (ANOVA) showed that there was no significant difference (\(P>0.05\)) from all treatments on the survival of Tilapia.

![Figure 4](image-url) – Percentage (%) of Survival Rate by Immersion Method Using methanol extract of Pasak Bumi roots

Figure 4 showed the survival rate of tilapia after immersion ranged from 80.67\% - 85.33\%. The results of this research were similar to those conducted by Ghosal & Chakraborty (2014) who did masculization with *Tribulus terrestris* extract, resulting in fish survival from 80.3 to 87.5\%. Mukhherjee et al. (2015) reported the use of *Asparagus racemusus* root extract with immersion method resulted in the highest survival rate of 95.56\%.

Daily specific growth (Figure 5) showed the highest growth rate occurred at EPB2 of \(0.151 \pm 0.005\) \%/day, EPB3 of \(0.150 \pm 0.005\) \%/day, positive control (MT) of \(0.135 \pm 0.004\) \%/day, EPB1 of \(0.134 \pm 0.003\) \%/day and negative control \(0.130 \pm 0.006\) \%/day. Absolute growth (Figure 6) showed the highest weight gain occurred in EPB2 of \(9.08 \pm 0.30\), EPB3 of \(9.02 \pm 0.33\), positive control (MT) of \(8.13 \pm 0.24\), EPB1 of \(8.07 \pm 0.21\) and negative control at \(7.80 \pm 0.39\) g. Analysis of Variance (ANOVA) showed a significant effect (\(P<0.05\)) both on daily specific and absolute growth of Tilapia. Duncan’s analysis showed that EPB2 was not different from EPB3, but both were different from EPB1, positive control (MT), and negative control.

![Figure 5](image-url) – Daily Growth Rate (%/day) of Tilapia by Immersion Method Using methanol extract of Pasak Bumi roots
Figure 6 – Weight (g) of Tilapia by Immersion Method Using methanol extract of Pasak Bumi roots

Treatment of 60 mg/L and 90 mg/L showed better and higher growth performance compared to other treatments. Phelps & Popma (2000) stated that androgen hormones have two physiological actions, which are androgenic, which promotes male character, and are anabolic, androgen hormones stimulate protein biosynthesis in fish bodies.

The profile of testosterone in fish larvae was measured by the ELIZA method. Testosterone levels before immersion were at the range of 0.18 - 0.23 pg/ml. After immersion, testosterone values of EPB1, EPB2, EPB3 and positive control (MT) increased with a range from 0.49 to 0.76 pg/ml, whereas in negative control, there was an increase from 0.218 pg/ml to 0.225 pg/ml. Furthermore, testosterone levels on 15, 30, 45, and 60 days, EPB1, EPB2, EPB3, and positive control (MT) treatments decreased in tilapia. In the negative control, there was a slight increase in the level of testosterone from 15 to 30 days and decreased from 45 to 60 days with a very narrow range of values (Figure 7).

The ELISA method was used in this research to measure testosterone concentrations in fish bodies (Memmat et al., 2015). As shown in Figure 7, testosterone levels decreased sharply until the 15 days after immersion, and after that, it tended to be stable until 60 days.
In negative control treatments, fish testosterone levels were relatively more stable compared to other treatments. A rapid decrease in steroid levels has also been reported by Padian and Kirankumar (2012), who stated that the decrease in steroids occured early at the beginning and gradually stabilized. The level of decrease in hormone levels depended on the species, the purity of the steroid used, the organs detected, and the treatment protocol.

CONCLUSION

The masculinization of Tilapia (Oreochromis niloticus) using methanol extract of Pasak Bumi roots (Eurycoma longifolia Jack) with immersion method resulted in male sex ratio between 72.66-76.33%, survival rate between 80.67-85.33%, weight gain between 8.07-9.08 g. Thereby, methanol extract of Pasak Bumi roots can be used for masculinization of tilapia by immersion method with the best performance at a dose of 60 and 90 mg/L.

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