Fyn Tyrosine Kinase Regulates the Surface Expression of Glycosylphosphatidylinositol-linked Ephrin via the Modulation of Sphingomyelin Metabolism*

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Glycosylphosphatidylinositol-linked ephrin-As play important roles in various biological events, such as neuronal development and immune responses. Because the surface amount of ephrin-As is critical in these events, the trafficking of ephrin-As must be regulated by intracellular machinery. In particular, Src family protein-tyrosine kinases regulate the intracellular trafficking of several membrane molecules and act downstream of ephrin-As; whether they affect the trafficking of ephrin-As, however, has remained unexplored. Here, we report that the activity of Src family protein-tyrosine kinases, particularly Fyn, negatively regulates the cell-surface amount of ephrin-As. The expression of constitutively active Fyn decreases the surface amount of ephrin-As. Conversely, the expression of dominant-negative Fyn or the application of a Src-family inhibitor increases the surface amount of ephrin-A2. The total cellular amount of ephrin-A is inversely correlated with its amount on the surface, suggesting that ephrin-As are more stable in the intracellular compartment. The expression of constitutively active Fyn increases the amount of sphingomyelin clusters on the plasma membrane, whereas inhibiting Fyn decreases it. Moreover, the inhibition of sphingomyelin synthesis greatly increases the surface amount of ephrin-As. Altogether, these results suggest that Fyn regulates the surface amount of ephrin-As by modulating the metabolism of sphingomyelin, which presumably inhibits the trafficking of ephrin-As from endosomes to the plasma membrane. The signaling cascade described here may function as part of the negative feedback loop of ephrin-A function.

Ephrins are the ligands of Eph receptor tyrosine kinases, and these molecules play central roles in many physiological and pathological events, such as neuronal development, adult brain function, angiogenesis, cancer, and immune responses (1, 2). Ephrins are structurally classified into two subclasses: ephrin-As are tethered to the outer leaflet of the plasma membrane via glycosylphosphatidylinositol (GPI)3 linkage, whereas ephrin-Bs are single transmembrane proteins. Ephrin-As and -Bs bind to EphAs and EphBs, respectively, with some exceptions (1, 2). The net concentration of functional ephrins (i.e. those on the plasma membrane) is important, a fact that is best exemplified by the mechanism of retinal axon guidance (3, 4).

The amount of ephrin-A protein is regulated at the transcriptional level (4, 5), but such regulation by itself is insufficient to achieve the tight regulation of ephrin concentration on the plasma membrane. In fact, the surface expression of many membrane receptor and ligand molecules is regulated at the posttranslational level by intracellular signaling cascades. For example, during the oocyte meiotic maturation of nematode Caenorhabditis elegans, the endocytosis of the Eph receptor is controlled both by intracellular adaptor proteins and by G protein signaling (6). The intracellular trafficking of Notch receptor and its ligand molecules are regulated by several proteins (7), ensuring the signaling capacity of the system. On the other hand, the mechanism by which intracellular signaling cascades affect the surface amount of ephrins remains largely unexplored.

Ephrin-A proteins, like many other GPI-linked proteins, are localized to the so-called “lipid-raft” microdomain on the plasma membrane (8), although the definition, size, biochemical nature, biophysical properties, and physiological importance of such microdomains are still matters of debate (9, 10). Whether the amount and localization of such microdomains are spatiotemporally regulated in neuronal cells remains unknown. Conceivably, the surface expression and/or distribution of ephrin-A is influenced, or even governed, by the amount and/or localization of the lipid rafts. Moreover, ephrin-A is not merely a ligand but also functions as a “receptor” of the EphA “ligand” in a phenomenon called “reverse signaling” (1, 2). In

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3 The abbreviations used are: GPI, glycosylphosphatidylinositol; AP, alkaline phosphatase; ApoER2, apolipoprotein E receptor 2; APP, amyloid precursor protein; CA, constitutively active; DN, dominant-negative; FB6, fumonisin B1; HmVNT-Lys, HmVNT-Lys, and monomeric Venus-tagged non-toxic lysenin; HRP, horseradish peroxidase; NPP6, nucleotide pyrophosphatase/phosphodiesterase 6; PP2, 4-amino-5-(4-chlorophenyl)-7-(t-butyl)pyrazolo[3,4-d]pyrimidine; PP3, 4-amino-7-phenylpyrazolo[3,4-d]pyrimidine; RFP, red fluorescent protein; SFK, Src-family tyrosine kinase; SM, sphingomyelin; PBS, phosphate-buffered saline.

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particular, the binding of the ectodomain of EphA to ephrin-A activates Fyn, a member of the Src-family tyrosine kinases (SFKs), in ephrin-A-bearing cells (11) via the p75 neurotrophin receptor (12). Recently, Fyn activity was reported to modulate the intracellular trafficking of TrkB, a receptor for brain-derived neurotrophic factor (13). Fyn induces the translocation of TrkB into the intracellular lipid-raft fraction (13). SFK is also involved in the intracellular trafficking of epidermal growth factor receptors (14–16), Reelin receptor apolipoprotein E receptor 2 (ApoER2) (17), and vascular endothelial growth factor receptor (18). However, whether Fyn activity affects the intracellular trafficking of ephrin-A is still unknown.

In the present study, we analyzed the effect of Fyn activity on the intracellular trafficking of ephrin-A protein. We report that Fyn activity negatively regulates the surface amount of ephrin-A. Moreover, this effect of Fyn is mediated by the metabolism of sphingomyelin (SM), a major component of the lipid raft microdomain and a regulator of intracellular trafficking. These results led us to hypothesize a novel model regarding the regulatory mechanism of ephrin-A and its downstream signaling.

**EXPERIMENTAL PROCEDURES**

**Expression Vectors**—The expression vectors for Myc-tagged ephrin-A2 and ephrin-A5 were described previously (19). The expression vectors for the Fyn mutants (20, 21) were a generous gift from Professor Tadashi Yamamoto (University of Tokyo, Japan). The expression vector for His6- and monomeric Venus-tagged non-toxic lysenin (HmV-NT-Lys) was described previously (22). The expression vector for nucleotide pyrophosphatase/phosphodiesterase 6 (NPP6) was a generous gift from Prof. Junken Aoki (Tohoku University, Japan). The expression vector for red fluorescent protein (RFP, DsRed-Express) was purchased from Clontech-Takara Bio (Mountain View, CA).

**Antibodies and Reagents**—Anti-Myc mouse monoclonal antibody (9E10) and anti-β-actin mouse monoclonal antibody (AC-15) were purchased from Sigma. Anti-caveolin-1 rabbit polyclonal antibody (N-20) and anti-flotilllin-1 rabbit polyclonal antibody (H-104) were purchased from Santa Cruz Biotechnologies (Santa Cruz, CA). Horseradish peroxidase (HRP)-conjugated secondary antibodies and HRP-streptavidin were purchased from Amersham Biosciences. Alexa-Fluor594-conjugated secondary antibody was purchased from Invitrogen. Fumonisin B₁ (FB₁), 4-amino-5-(4-chlorophenyl)-7-(t-buty1)-pyrazolo[3,4-d]pyrimidine (PP3), and 4-amino-7-phenylpyrazolo[3,4-d]pyrimidine (PP3) were purchased from Merck Biosciences (Darmstadt, Germany). N-Hexanoyl-d-sphingosine (C₆-h-ceramide) and sphingomyelin were purchased from Sigma.

**Cell Culture and Transfection**—COS-7 cells were maintained in Dulbecco’s modified Eagle’s medium containing 10% fetal calf serum (Equatech-Bio, Kerrville, TX) and antibiotics. Transfection was performed using Lipofectamine 2000 (Invitrogen), according to the manufacturer’s protocol.

**Quantification of the Surface Amount of Ephrin-A Using EphA3-alanine Phosphatase Fusion Protein**—Culture supernatant containing EphA3-alanine phosphatase (AP) was obtained, and the binding assay was performed as reported previously (23). Briefly, the cells were incubated with EphA3-AP for 60 min, washed five times with Hanks’ balanced salt solution containing 0.5 mg/ml bovine serum albumin, 0.1% NaN₃, and 20 mM HEPES, pH 7.2, treated for 2 min with an aceto-fermalin fixative (60% aceton, 8% formalin, 20 mM HEPES, pH 7.2), washed two times in HEPES-buffered saline (150 mM NaCl, 20 mM HEPES, pH 7.2), and then incubated at 65 °C for 30 min to inactivate the endogenous AP activity. After rinsing twice with AP buffer (5 mM MgCl₂, 100 mM NaCl, 100 mM Tris-HCl, pH 9.5), the cells were stained with 0.17 mg/ml 5-bromo-4-chloro-3-indolyl phosphate and 0.33 mg/ml of nitro blue tetrazolium in AP buffer. The reactions were stopped by changing the solution to phosphate-buffered saline (PBS) containing 20 mM EDTA. Microscopic images were captured using a color-cooled charge-coupled device camera and a VB-6000 image control system (Keyence, Osaka, Japan) equipped with a SZX7 stereoscopic microscope (Olympus, Tokyo, Japan).

**Western Blotting**—Samples were separated by 12% SDS-PAGE and transferred onto a polyvinylidene difluoride membrane. The blotted membranes were blocked with 5% skimmed milk in Tris-buffered saline with 0.05% Tween 20 (TBST) for 1 h and incubated with the primary antibody for 2 h. After washing four times with TBST, the membranes were incubated with HRP-conjugated secondary antibody (1:4000) for 1 h and washed, and the labeled proteins were detected using Immunobilon Western Chemiluminescent HRP Substrate (Millipore, Bedford, MA). Images were captured using a LAS3000 mini system (Fuji, Tokyo, Japan).

**Biotinylation of Cell-surface Proteins**—Biotinylation of the cell-surface proteins was performed as described previously (24) with some modifications. Twenty-four hours after transfection, COS-7 cells were rinsed with ice-cold PBS and incubated with 0.125 mg/ml of sulfo-NHS-biotin (Thermo Scientific, Rockford, IL) for 30 min at 4 °C. After washing twice with PBS containing 100 mM glycine to stop the reaction, the cells were lysed with radioimmune precipitation assay buffer (10 mM phosphate buffer, pH 7.4, 150 mM NaCl, 2 mM EDTA, 1% sodium deoxycholate, 0.1% SDS, 1% Triton X-100). The lysates were recovered and centrifuged at 15,000 × g for 2 min at 4 °C to remove debris. The supernatant was then incubated with streptavidin-conjugated agarose (Sigma) and rotated for 60 min at 4 °C. After centrifugation at 3,300 × g for 1 min, the supernatants were removed and the resins were washed six times with radioimmune precipitation assay buffer and eluted with SDS-PAGE sampling buffer (62.5 mM Tris-HCl, pH 6.8, 2% SDS, 5% β-mercaptoethanol, 10% glycerol, and 0.05% bromophenol blue).

**Detection of SM Clusters in Situ**—HmV-NT-Lys protein was expressed in Escherichia coli BL21-CodonPlus-RP (Stratagene, La Jolla, CA) and purified using nickel-nitritriacetic acid-agarose (Qiagen, Valencia, CA), according to the manufacturer’s protocol. HmV-NT-Lys solutions were diluted in PBS containing 2% bovine serum albumin. To detect the SM clusters in situ, the cells were fixed and incubated with 1–2 μg/ml HmV-NT-Lys for 1.5–3 h and washed with PBS four times. The fluorescent images were captured using a BZ-9000 digital fluorescence microscope (Keyence).
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Immunocytochemistry—The cells were fixed with 4% paraformaldehyde for 10 min or with 4% formaldehyde for 5 min at room temperature. After washing with PBS twice, the cells were permeabilized with PBS containing 0.05% Tween 20 for 30 min and then blocked by incubation with PBS containing 2% bovine serum albumin for 30 min. The cells were then incubated with the primary antibody for 2 h at room temperature, washed four times with PBS, incubated with the secondary antibody for 90 min, and washed four times with PBS and twice with water. To visualize the nucleus, 2 µg/ml Hoechst 33342 (Invitrogen) was added to the secondary antibody solution. Fluorescent images were captured using a BZ-9000 digital fluorescence microscope. In the case of optical sectioning, the samples were examined using an LSM510meta confocal laser-scanning microscope (Carl Zeiss, Tokyo, Japan).

Lipid Extraction and Analysis Using TLC—Prior to the phospholipid analysis, COS-7 cells were preincubated with medium containing 5 µM PP2 or PP3 for 24 h. After washing with PBS twice, the cells were lysed under a hypotonic condition containing 0.1 M acetic acid. The cellular total lipids were extracted twice, the cells were lysed under a hypotonic condition containing 2% bovine serum albumin for 30 min. The cells were then incubated with the primary antibody for 2 h at room temperature, washed four times with PBS, incubated with the secondary antibody for 90 min, and washed four times with PBS and twice with water. To visualize the nucleus, 2 µg/ml Hoechst 33342 (Invitrogen) was added to the secondary antibody solution. Fluorescent images were captured using a BZ-9000 digital fluorescence microscope in the case of optical sectioning, the samples were examined using an LSM510meta confocal laser-scanning microscope (Carl Zeiss, Tokyo, Japan).

Quantification and Analysis of Fluorescent Intensities—For the fluorometric assays, the mean fluorescent intensity was scored using ImageJ analysis software, version 1.41m (rsweb.nih.gov/ij/). Images were captured using a BZ-9000 digital fluorescence microscope (Keyence) with a 20× objective, and individual cells in 10–15 randomly selected microscopic fields were traced. Dividing or dead cells were excluded from the measurements based on the Hoechst staining pattern. To calculate the staining intensity of HmV-NT-Lys, RFP-expressing area in each cell was outlined using an automated program and was defined as the whole cell region. The mean fluorescent intensity of the HmV-NT-Lys image in the corresponding region was calculated by subtracting the background and was expressed as 256 gradations. To calculate the mean fluorescent intensity of surface Myc-ephrin-A2, the optically observed area of each cell was defined as the whole cell region, and the mean fluorescence intensity of the Myc-immunoreactive images was calculated. All measurements from two independent experiments were performed in a blinded manner.

RESULTS

Modulation of Fyn Activity Affects the Surface Amount of Ephrin-A—COS-7 cells were transiently transfected with the expression vector for Myc-ephrin-A2 together with that for dominant negative Fyn (Fyn-DN), that for constitutively active Fyn (Fyn-CA), or pcDNA3.1 (control). Twenty-four hours after transfection, the cells were stained with EphA3-AP to assess the surface amount of Myc-ephrin-A2. When Fyn-DN was co-transfected, the surface amount of Myc-ephrin-A2 increased, compared with that of the control (Fig. 1, B and A, respectively). On the other hand, the co-expression of Fyn-CA dramatically reduced the surface amount of Myc-ephrin-A2 (Fig. 1C). Staining with the control AP gave no signal in the Myc-ephrin-A2-transfected cells (Fig. 1F). These results indicate that Fyn activity negatively regulates the surface amount of Myc-ephrin-A2. To verify the role of Fyn on the surface expression of other types of GPI-linked ephrin-A, Myc-ephrin-A5 was expressed in COS-7 cells either with pcDNA3.1, Fyn-DN, or Fyn-CA. EphA3-AP staining clearly revealed that the surface amount of Myc-ephrin-A5 increased when the cells were co-transfected with Fyn-DN (Fig. 1E) and decreased when the cells were co-transfected with Fyn-CA (Fig. 1F). The cells expressing Myc-ephrin-A5 were not stained by control AP (Fig. 1L). These results indicate that a common mechanism exists for regulating the surface amount of Myc-ephrin-A2 and Myc-ephrin-A5.

We next checked whether the inhibition of endogenous SFK activity was sufficient for increasing the surface amount of Myc-ephrin-As. COS-7 cells expressing either Myc-ephrin-A2 or Myc-ephrin-A5 were incubated with 5 µM PP2, an SFK inhibitor, for 24 h and then stained with EphA3-AP. As a control, the cells were incubated with PP3, an inactive analogue of PP2. The surface amount of Myc-ephrin-A2 in the PP2-treated cells was more abundant than that in the PP3-treated cells (Fig. 1, H and G, respectively). The same tendency was observed in cells expressing Myc-ephrin-A5 (Fig. 1, J and K). Thus, these results indicate that the surface amounts of Myc-ephrin-As are regulated by the endogenous SFK activity.

Fyn Activity Modulates the Surface Amount of Ephrin-A by Altering Its Intracellular Sorting—By which mechanism does SFK regulate the surface amount of ephrin-A? We first speculated that SFK simply regulates the total amount of ephrin-A protein in the cell. To test this hypothesis, COS-7 cells were transfected with the expression vector for Myc-ephrin-A2 together with pcDNA3.1 (control), Fyn-DN, or Fyn-CA, and whole cell lysates were prepared 24 h later. These lysates were then analyzed using SDS-PAGE and Western blotting with anti-Myc antibody (Fig. 2A, top panel, lanes 1–3). Quite surprisingly, the co-expression of Fyn-DN greatly decreased the total amount of Myc-ephrin-A2 (Fig. 2A, top panel, lane 2), whereas the expression of Fyn-CA increased it (Fig. 2A, top panel, lane 3). In other words, the total amount of Myc-ephrin-A2 in the cells expressing Fyn mutants was inversely correlated with the surface amount of Myc-ephrin-A2 (Fig. 1, A–C). The amounts of caveolin-1 and flotillin-1, two proteins that are localized to the “lipid raft,” did not change (Fig. 2A, middle panels, lanes 1–3). The amounts of β-actin were also unaffected (Fig. 2A, bottom panel, lanes 1–3). Likewise, the expression of Fyn-DN and Fyn-CA decreased and slightly increased the total amount of Myc-ephrin-A5, respectively (Fig. 2A, top panel, lanes 5 and 6, respectively), without affecting the amounts of caveolin-1, flotillin-1, or β-actin (Fig. 2A, lanes 4–6).
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We next asked whether the surface amounts of other proteins were affected by modulating Fyn activity. For this purpose, the cells were transfected with expression vectors for Myc-ephrin-A2 and either pcDNA3.1, Fyn-DN, or Fyn-CA, and the cell-surface proteins were labeled with sulfo-NHS-LC-biotin. The biotinylated proteins were precipitated using streptavidin-conjugated agarose, followed by Western blotting analysis. When transfected with Fyn-DN, Myc-ephrin-A2 protein in the biotinylated (cell surface) fraction increased (Fig. 2B, lane 2), whereas that in the corresponding fraction of Fyn-CA co-transfected cells decreased (Fig. 2B, lane 3), confirming the results of EphA3-AP staining (Fig. 1, A–C). On the other hand, the expression of the Fyn mutants hardly affected the surface amounts of most of the other cellular proteins (Fig. 2B, lanes 4–6), as assessed by detection with HRP-streptavidin. These results suggest that the surface amount of ephrin-As is selectively regulated by SFK activity, which is achieved by modulating the intracellular sorting of ephrin-A protein. It is also suggested that ephrin-As in the intracellular compartment are more stable than those on the plasma membrane, because the total amounts of Myc-ephrin-As were more abundant in cells that have them mostly in the intracellular compartment (i.e. Fyn-CA-expressing cells, Fig. 2A, lanes 3 and 6) than in cells with Myc-ephrin-As on the plasma membrane (i.e. Fyn-DN-expressing cells, Fig. 2A, lanes 2 and 5).

The cellular distribution of ephrin-A2 was also examined by immunostaining. Cells expressing Myc-ephrin-A2 alone (Fig. 3A), with Fyn-DN (Fig. 3B), or with Fyn-CA (Fig. 3C) were fixed and stained with anti-Myc antibody without permeabilization to detect the surface Myc-ephrin-A2. Under these conditions, the total fluorescence signal was higher in the Fyn-DN-transfected cells (Fig. 3B) and lower in the Fyn-CA-transfected cells (Fig. 3C) than that in the control cells (Fig. 3A). Then, we next compared the intracellular localization of Myc-ephrin-A2 by immunostaining after membrane permeabilization. As shown in Fig. 3D, Myc-ephrin-A2 protein was distributed mostly along the edge of the cell, the putative cell-surface region, and was also diffusely present in the cytoplasmic region. In cells co-expressing Fyn-DN, the fluorescence signals at the edge of the cell were augmented, whereas those in the intracellular region were diminished (Fig. 3E). In cells co-expressing Fyn-CA, on the other hand, the cell-surface signals were diminished and signals were often found throughout the cytoplasm, distributed in apparently clustered and punctate manner (Fig. 3F, arrowheads), indicating that Myc-ephrin-A2 protein tends to be retained within the cell when Fyn activity is high. These results thus strongly suggest that Fyn activity affects the surface expression of Myc-ephrin-A2 by selectively changing the intracellular trafficking of this protein.

Fyn Regulates the Cellular SM Level and Clustering—We next investigated whether SFK activity can modulate the amount or localization of the lipid-raft microdomain on the plasma membrane, where ephrin-As are thought to localize. For this purpose, COS-7 cells were transfected with either pcDNA3.1, Fyn-DN, or Fyn-CA followed by staining with HmV-NT-Lys, which binds specifically to SM clusters (22, 27). The shape of each transfected cell was simultaneously visual-
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![Diagram of Fyn activity altering the subcellular distribution of ephrin-A2.](image)

**FIGURE 3.** Fyn activity alters the subcellular distribution of ephrin-A2. A–C, COS-7 cells were co-transfected with Myc-ephrin-A2 and with the indicated Fyn mutants, then fixed and immunostained with anti-Myc antibody without membrane permeabilization. Scale bar: in C (for A–C), 200 μm. D–F, intracellular distribution of Myc-ephrin-A2 in pcDNA3.1 (D), Fyn-DN (E), or Fyn-CA (F)-transfected cells. Twenty-four hours after transfection, COS-7 cells were permeabilized and immunostained with anti-Myc antibody. Confocal laser optical sectioning (1 μm thick) was used to visualize the distribution of Myc-ephrin-A2 in the transfected cells. The arrowheads in F indicate the intracellular Myc-ephrin-A2 puncta in Fyn-CA-transfected cells. Scale bar: in F (for D–F), 10 μm.

and stained them with HmV-NT-Lys. Consequently, the amount of SM clusters markedly decreased when the cells were treated with PP2 but not with PP3 (Fig. 4, I and H, respectively). Concomitantly, the total cellular SM amount was much lower in the PP2-treated cells (Fig. 4J, lane 4) than in the PP3-treated cells (Fig. 4J, lane 3). Thus, SFK activity modulates the cellular SM level and clustering.

We next compared the localization of Myc-ephrin-A2 and SM in the Fyn-CA-expressing cells by combining immunocytochemistry and HmV-NT-Lys staining after membrane permeabilization. As shown in Fig. 5A, many clusters or vesicles of Myc-ephrin-A2 were observed when Fyn-CA was co-expressed. Interestingly, few of these clusters or vesicles overlapped with the SM clusters (Fig. 5, C and D) within the cells. Thus, although Myc-ephrin-A2 and SM may be located in the same microdomain on the plasma membrane, they are segregated to within the intracellular compartment of Fyn-CA-expressing cells.

**Cell-surface Amount of Ephrin-A Is Regulated by SM**—We next explored the possibility that a change in the amount of SM might affect the surface amount of ephrin-A2. To alter the cellular sphingolipid levels, the cells were treated with FB1, which inhibits sphingosine N-acyltransferase (28). FB1 reduces sphingolipid biosynthesis without affecting cellular cholesterol levels, and the addition of C₆₆-ceramide to FB1-treated cells restores the cellular sphingolipid levels (29). COS-7 cells expressing Myc-ephrin-A2 were incubated with FB1 in the presence or absence of C₆₆-ceramide and were stained with HmV-NT-Lys (Fig. 6, A–C) or with EphA3-AP (Fig. 6, D–F). Under the control conditions, the COS-7 cells were brightly stained by HmV-NT-Lys (Fig. 6A). When treated with FB1, the amount of SM clusters was greatly reduced in most of the cells (Fig. 6B). This effect was partly restored in the presence of C₆₆-ceramide (Fig. 6C). In the FB1-treated cells, the surface

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**FIGURE 2.** Fyn activity alters the sorting machinery of ephrin-A. A, COS-7 cells were transfected with Myc-ephrin-A2 (lanes 1-3) or -A5 (lanes 4-6) together with pcDNA3.1 (lanes 1 and 4), Fyn-DN (lanes 2 and 5), or Fyn-CA (lanes 3 and 6), respectively. A representative immunoblot of the protein levels of Myc-ephrin-A (top panel), caveolin-1, flotillin-1, and β-actin (bottom panel) in the total cell lysates is shown. The black bar indicates the positions of Myc-ephrin-A2. Transfection with Fyn-DN or Fyn-CA caused a decrease or an increase in Myc-ephrin-A without affecting caveolin-1, flotillin-1, and β-actin. B, COS-7 cells were transfected with the indicated vectors, and their cell-surface proteins were labeled with sulfo-NHS-LC-biotin. The cells were then lysed, and the labeled proteins were precipitated with streptavidin beads, followed by SDS-PAGE and Western blotting using anti-Myc antibody (lanes 1-3). The black bar indicates the positions of Myc-ephrin-A2. Total biotinylated proteins were visualized using streptavidin-HRP (lanes 4-6). The positions of the molecular weight mass (kDa) markers are shown to the left of the panel.
amount of Myc-ephrin-A2 greatly increased (Fig. 6E), compared with that in control cells (Fig. 6D). When Cα-ceramide was present, the increase in the surface amount of Myc-ephrin-A2 was less obvious (Fig. 6F). Therefore, reducing the cellular sphingolipid level was sufficient to increase the surface amount of Myc-ephrin-A2.

We further addressed the possibility that the regulation of sphingolipid metabolism by Fyn activity may alter the surface amount of ephrin-A. COS-7 cells were transfected with the expression vectors for Myc-ephrin-A2 and either pcDNA3.1 (control) or Fyn-CA in the presence of FB1, and Myc-ephrin-A2 protein on the cell surface was labeled with anti-Myc antibody (A; red), Hoechst 33342 (B; green) and HmV-NT-Lys (C; blue). The merged image is shown in D. The intracellular puncta of Myc-ephrin-A2 contained few SM clusters. The white box in C is magnified in D. Scale bar: in A (for A–C), 10 μm.

We further addressed the possibility that the regulation of sphingolipid metabolism by Fyn activity may alter the surface amount of ephrin-A. COS-7 cells were transfected with expression vectors for pcDNA3.1 (A and D), Fyn-DN (B and E), or Fyn-CA (C and F) together with RFP. Twenty-four hours after transfection, the cells were fixed and stained with HmV-NT-Lys (A–C; green). The fluorescence of RFP reveals the cell shape (D–F; red). Scale bar: in F (for A–F), 10 μm. G, summary of the effects of pcDNA3.1, Fyn-DN, and Fyn-CA on the content of the SM clusters in COS-7 cells. The pixel intensity of HmV-NT-Lys fluorescence was measured in each cell, and the average of the mean fluorescence intensity is represented. *, p < 0.05; **, p < 0.01 versus pcDNA3.1. H, I, COS-7 cells were maintained in media containing DMSO alone (A), 40 μg/ml FB1 (B), or 40 μg/ml FB1 and 5 μM of Cα-ceramide (C) for 48 h, followed by incubation with HmV-NT-Lys (green) and Hoechst 33342 (blue). In the presence of FB2, the COS-7 cells showed the reduced binding of HmV-NT-Lys. Scale bar: in I (for H and I), 100 μm. J, TLC analysis of cellular phospholipids levels in DMSO, PP3-, or PP2-treated COS-7 cells (lanes 2, 3, and 4, respectively). The total lipids of the COS-7 cells were extracted and separated using TLC. SM, sphingomyelin standard (lane J). The representative image shows that PP2 treatment decreased the SM levels.
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In this study, we found that Fyn, a member of SFK that acts downstream of the Eph-ephrin reverse signaling pathway (11, 12), regulates the surface amount of ephrin-A by modulating the SM content (or localization) of the cell. This is the first demonstration of the molecular mechanism responsible for the intracellular signaling that regulates the surface amount of ephrin-As at the posttranslational level. The idea that Fyn or SFK affects the localization of a membrane molecule by modulating lipid metabolism is also a novel concept. Recently, Fyn was reported to regulate the surface amounts of amyloid precursor protein (APP) and ApoER2, a receptor for Reelin (17). There are two interesting differences between the effects of Fyn on APP or ApoER2 and those on ephrin-A2. First, the expression of Fyn-CA increases the surface amounts of APP and ApoER2 while it decreases that of ephrin-A. Second, the expression of Fyn-CA significantly increases the total ephrin-A level but has no effect on that of APP or ApoER2 (17). Interestingly, the total amounts of ephrin-A, APP, and ApoER2 are decreased by Fyn-DN (17). APP and ApoER2 contain an intracellular domain that is phosphorylated by Fyn, and they have a number of binding proteins that can modify their intracellular trafficking (17). Ephrin-As, being GPI-linked proteins, are not subject to such regulation. Therefore, although Fyn regulates the intracellular trafficking of ephrin-A, APP, and ApoER2, the underlying mechanisms of these regulations must differ. The trafficking of APP and ApoER2 is regulated by phosphoinositides (17, 24). It would be interesting to test whether it is also regulated by sphingolipids. We also found that co-expression of Fyn mutants did not affect the surface amount of other structurally unrelated GPI-linked proteins NPP6 and placental alkaline phosphatase (data not shown), suggesting that the action of Fyn is selective, if not specific, to ephrin-A.

Generally, GPI-linked proteins mature in the Golgi apparatus, are transported to the plasma membrane by vesicular trafficking, and are probably constantly internalized via a clathrin-independent or -independent mechanism (30–33). Internalized GPI-linked proteins are retained in the endosome and are subsequently subjected to degradation or are “recycled” to the plasma membrane (29, 31–34). Our finding that the surface concentration of SM is negatively correlated with that of ephrin-As raises three possibilities. 1) SM inhibits the maturation or transport of ephrin-A. This possibility is rather unlikely, because we did not observe the accumulation of Myc-ephrin-A2 in the Golgi apparatus in Fyn-CA-expressing cells (Figs. 3F and 5A). 2) SM directly enhances the internalization of ephrin-A. While possible, the observation that intracellular vesicles containing Myc-ephrin-A2 typically do not contain SM clusters (Fig. 5, C and D) suggests that SM and ephrin-A tend to be segregated in Fyn-CA-expressing cells. 3) SM augments the retention/recycling ratio. Currently, we favor this scenario, because the amount of intracellular ephrin-A is increased in Fyn-CA-expressing (i.e. SM-rich) cells (Fig. 2, A and B), and sphingolipid depletion in Chinese hamster ovary cells reportedly leads to the retention of two GPI-linked proteins, folate receptor, and decay-accelerating factor, in the endosome fraction (29). How SM affects intracellular trafficking of ephrin-As remains to be investigated. It was also suggested that ephrin-A in the intracellular compartment is more stable than those on the plasma membrane (Fig. 2). This may be due to selective proteolysis of ephrin-A on the plasma membrane by a disintegrin and metalloprotease family (19). Whether SM directly regulates the proteolysis or degradation of ephrin-As remains to be investigated.

We hoped to confirm that the results obtained in COS-7 cells could be reproduced in neuronal cells, but such studies were hindered by two technical difficulties. First, although HmV-NT-Lys can reportedly stain cultured neurons (35), neither hippocampal neurons nor cortical neurons produced a significant signal when we stained them using HmV-NT-Lys (data not shown). The reason for the lack of staining is unknown, but the high contents of non-SM sphingolipids in neuronal cells may hinder the clustering of SM, which is a prerequisite for HmV-NT-Lys binding (27, 36). Second, the overexpression of ephrin-A and Fyn mutants was not satisfactorily obtained in neuronal cells (data not shown), probably because of their toxicity. Thus, in future studies, the quantification of endogenous ephrin-A in knockout mice for Fyn or some other SFK will be necessary.
How SFK modulates the cellular SM content and/or localization is a daunting question, because our understanding of the regulatory mechanisms underlying the synthesis, transport, and degradation of SM is still quite premature (37–39). There are two isozymes for SM synthase (40), each of which seem to have independent roles (41). To the best of our knowledge, whether the activity of SM synthase is regulated by an intracellular signaling cascade is unknown. However, our result, that an inhibitor of sphingolipid synthesis enabled the recovery of the Fyn-CA-induced decrease in surface ephrin-A2 (Fig. 7), may suggest that activated Fyn can directly up-regulate SM clustering by activating SM synthesis. The regulation of cellular SM amounts by sphingomyelinases is a relatively well studied area (37, 39), but only a few reports have discussed the regulation by phosphorylation. Very recently, Chudakova et al. (42) reported that Lyn, an integrin-associated member of SFKs, suppresses the activities of sphingomyelinases. Thus, the up-regulation of SM by SFK via the inhibition of its hydrolysis may be a universal phenomenon. Finally, we could not exclude the possibility that sphingolipids other than SM play a role in the regulation of ephrin-A trafficking.

The model proposed in the present study is summarized in Fig. 8. Ephrin-A, like other GPI-linked proteins, undergoes continuous cycling between the plasma membrane and the intracellular endosome fraction. When ephrin-A binds to EphA, Fyn is activated in ephrin-A-bearing cells. The activation of SFK up-regulates SM via an unknown mechanism, and SM inhibits the transport of intracellular ephrin-A to the plasma membrane. Therefore, this pathway can be regarded as a negative feedback of Eph-ephrin reverse signaling; that is, when an ephrin-A-bearing cell encounters EphA-bearing cells, the surface amount of ephrin-A decreases. This phenomenon may be important for preventing the hyperactivation of EphA and/or for terminating signaling in both directions. Moreover, cross-talk with other neuronal signaling pathways can be envisaged. For example, SFK, most likely Fyn, is activated by Reelin in neuronal cells (43, 44). Our results presented here predict that Reelin down-regulates ephrin-A by up-regulating SM. Furthermore, amyloid-β-peptide 42, the most likely causative molecule of Alzheimer disease, is known to down-regulate SM (45), which would increase the surface amount of ephrin-A. Because ephrin-A is one of the most important inhibitors of axonal elongation and regeneration (46), this pathway may be involved in the pathogenesis of Alzheimer disease. Thus, we believe that our present findings contribute significantly not only to the elucidation of questions in basic cell biology, but also to the development of novel therapeutics for neurodevelopmental and neurodegenerative diseases.

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