ORIGINAL ARTICLE

Phosphoinositide 3-kinase-δ regulates fungus-induced allergic lung inflammation through endoplasmic reticulum stress

Kyung Sun Lee, Jae Seok Jeong, So Ri Kim, Seong Ho Cho, Narasaiyah Kolliputi, Yun Hye Ko, Kyung Bae Lee, Suk Chul Park, Hae Jin Park, Yong Chul Lee

ABSTRACT

Background Sensitisation with Aspergillus fumigatus (Af) is known to be associated with severe allergic lung inflammation, but the mechanism remains to be clarified. Phosphoinositide 3-kinase (PI3K)-δ and endoplasmic reticulum (ER) stress are suggested to be involved in steroid-resistant lung inflammation. We aimed to elucidate the role of PI3K-δ and its relationship with ER stress in fungus-induced allergic lung inflammation.

Methods Using Af-exposed in vivo and in vitro experimental systems, we examined whether PI3K-δ regulates ER stress, thereby contributing to steroid resistance in fungus-induced allergic lung inflammation. Moreover, we checked expression of an ER stress marker in lung tissues isolated from patients with allergic bronchopulmonary aspergillosis.

Results Af-exposed mice showed that ER stress markers, unfolded protein response (UPR)-related proteins, phosphorylated Akt, generation of mitochondrial reactive oxygen species (mtROS), eosinophilic allergic inflammation, and airway hyperresponsiveness (AHR) were increased in the lung. Similarly, glucose-regulated protein 78 was increased in lung tissues of patients with ABPA. A PI3K-δ inhibitor reduced Af-induced increases in ER stress markers, UPR-related proteins, allergic inflammation and AHR in mice. However, dexamethasone failed to reduce Af-induced allergic inflammation, AHR and elevation of ER stress. Administration of an ER stress inhibitor or a mtROS scavenger improved Af-induced allergic inflammation. The PI3K-δ inhibitor reduced Af-induced mtROS generation and the mtROS scavenger ameliorated ER stress. In primary cultured tracheal epithelial cells, Af-induced ER stress was inhibited by blockade of PI3K-δ.

Conclusions These findings suggest that PI3K-δ regulates Af-induced steroid-resistant eosinophilic allergic lung inflammation through ER stress.

INTRODUCTION

Aspergillus fumigatus (Af) is ubiquitous in normal air and the human respiratory system is constantly exposed to airborne spores of Af. Various lung disorders including invasive aspergillosis, aspergillosma, bronchial asthma and allergic bronchopulmonary aspergillosis (ABPA) are commonly associated with Af. Among them, ABPA and bronchial asthma are representative allergic lung inflammatory disorders related to Af. ABPA is one of the complex hypersensitivity reactions to Aspergillus species usually colonised in bronchi and it occurs in patients with asthma. Although many patients with ABPA respond well to treatment with systemic steroids, the condition of some patients is poorly controlled by conventional management. Additionally, in patients with bronchial asthma, sensitisation with Af is known to cause severe phenotypes of the disease. However, the mechanisms by which Af is involved in allergic lung inflammation remain poorly understood.

Class I phosphoinositide 3-kinases (PI3Ks) are involved in the induction of T helper type 2 cell (Th2)-related immune responses. They exist as heterodimeric complexes in which a catalytic p110 subunit (designated as α, β, γ or δ) is associated with a particular regulatory subunit (designated as p85, p55, p50 or p101). These proteins are a family of lipid signalling kinases that generate phosphatidylinositol-3,4,5-trisphosphate (PIP3) and phosphatidylinositol-3-phosphate (PIP2) and phosphorylate effector proteins such as Akt. The p110-α and p110-β isoforms are ubiquitously expressed. However, expression of p110-δ isofrom is restricted to circulating haematogenous cells and endothelial cells. PI3K-δ plays a role in a variety of immune processes involving B-cell and T-cell activation, mast cell degranulation, and trafficking and activation of neutrophils and eosinophils.
In particular, PI3K-δ mediates steroid resistance in chronic respiratory disorders in which neutrophilic inflammation predominates. Therefore, inhibition of this pathway may serve as a critical target for treating steroid-resistant severe asthma and non-severe asthma. Unfortunately, there is a lack of information on whether PI3K-δ contributes to fungus-induced steroid-resistant eosinophilic allergic lung inflammation. The endoplasmic reticulum (ER) serves as a protein-folding factory essential for production of functional proteins and trafficking of secretory and biological membrane proteins in cells. ER stress causes accumulation of unfolded and/or misfolded proteins in ER, interferes with protein synthesis and secretion, induces reactive oxygen species (ROS) generation, and increases inflammation partly via nuclear factor (NF)-κB activation. As a result, ER stress is implicated in the pathogenesis of diverse inflammatory processes. Recently, our group demonstrated that ER stress influences the pathogenesis of bronchial asthma, especially the steroid-resistant neutrophilic asthma model. Although various signalling pathways are involved in ER stress, the relationship between PI3K-δ and ER stress in allergic lung inflammation is not clearly understood. In addition, there are few data about the role of ER stress in fungus-induced allergic lung inflammation.

In this study, we examined whether PI3K-δ regulates ER stress, thereby contributing to steroid resistance in eosinophilic dominant allergic lung inflammation using A. fumigatus–exposed mice and primary cultured tracheal epithelial cells (TECs). In addition, we analysed the expression of a representative ER stress marker, glucose-regulated protein 78 (GRP78), in lung tissues from patients with ABPA to evaluate the association of ER stress with the pathogenesis of A. fumigatus–related lung inflammation in humans.

METHODS

Animals and experimental protocol

Female C57BL/6 mice, 8–10 weeks of age and free of murine specific pathogens, were used. All experimental animals used in this study were under a protocol approved by the Institutional Animal Care and Use Committee of the Chonbuk National University (CBU 2014-00030). For the generation of an A. fumigatus–induced allergic lung inflammation model, mice were sensitised and challenged with A. fumigatus, in which the cellular fungal material was inactivated and lyophilised, as previously described. Bronchoalveolar lavage (BAL) was performed at 24 h after the last challenge with A. fumigatus.

Administration of drugs

A selective p110-δ inhibitor, IC87114 (0.1 or 1 mg/kg body weight/day, Calbiochem, San Diego, California, USA) or vehicle control (0.05% dimethyl sulfoxide, DMSO) diluted with 0.9% NaCl was administered once by intratracheal instillation to each animal 24 h after the last challenge with A. fumigatus. A mitochondrial ROS (mtROS) scavenger, NecroX-5 (30 mg/kg body weight/day, Enzo Life Sciences, Farmingdale, New York, USA) diluted with distilled water was administered twice by intraperitoneal injection to each animal 24 h before and after the last challenge with A. fumigatus. The amount of PI3P produced was quantified by the PI3P competition enzyme immunoassay (Echelon, Inc., Salt Lake City, Utah, USA).

Measurement of PI3K enzyme activity in lung tissue

Total glutathione (GSH) and glutathione disulfide (GSSG) levels were determined using a GSH Assay Kit (Cayman Chemical Co., Ann Arbor, Michigan, USA) according to the manufacturer’s protocol.

Serum total IgE and A. fumigatus–specific IgE

For the A. fumigatus–specific IgE assay, 96-well immunosorbent plates were coated with A. fumigatus antigen (10 μg/mL, Greer Laboratories, Lenoir, North Carolina, USA). Horseradish peroxidase-conjugated goat anti-mouse IgE antibody (Bethyl Laboratories, Montgomery, Texas, USA) was used to detect A. fumigatus–bound IgE. Total serum IgE was measured using a mouse Total IgE ELISA Kit (MD Bioproducts, St Paul, Minnesota, USA) according to the manufacturer’s protocol.

Isolation and primary culture of murine TECs and IC87114 treatment

Murine TECs were isolated under sterile conditions as described previously. To verify the role of PI3K-δ in the regulation of ER stress in A. fumigatus–stimulated primary cultured TECs, cells were treated with IC87114 (10 μmol/L) for 2 h, and then stimulated by A. fumigatus antigen (5 μg/mL) for an additional 12 h.

PI3K-δ specific or scrambled small interfering RNA transfection in primary cultured TECs

Primary cultured murine TECs were transfected with the PI3K-δ specific (PIK3CD) or scrambled small interfering RNA (siRNA) (Dharmacon, Lafayette, Louisiana, USA) by using Lipofectamine RNAiMAX reagent (Invitrogen, Carlsbad, California, USA).

Immunofluorescence staining for GRP78, CCAAT/enhancer-binding protein-homologous protein (CHOP) and mtROS

Immunofluorescence staining for GRP78 and CHOP was performed as previously described. To demonstrate the intensity of mtROS in A. fumigatus–stimulated primary cultured TECs and BAL cells, cells were stained with Mitotracker Red CM-H2ROS (Invitrogen) and analysed using a confocal laser scanning microscope (Carl Zeiss, Jena, Germany).

Western blot analysis

Protein levels were determined by Western blotting as previously described.

Nuclear protein extractions

Extraction of nuclear proteins was performed as previously described.

Histology

Histologic assessment was performed as previously described.
Determination of airway responsiveness to methacholine
Invasive measurement of airway responsiveness was performed as described elsewhere.19

Statistics
We used SPSS statistical software (V18.0, SPSS, Chicago, Illinois, USA). Data are expressed as mean±SEM. Statistical comparisons were performed using one-way ANOVA followed by Scheffe’s test. The Mann–Whitney U test was used to compare the two groups of human subjects. A value of p<0.05 was considered statistically significant.

RESULTS
PI3K-δ regulates ER stress in vivo
To examine whether p110-δ contributes to regulation of ER stress in A-f-exposed mice, we evaluated the effects of IC87114, a selective p110-δ inhibitor, on levels of ER stress markers, GRP78 and CHOP, and unfolded-protein response (UPR)-related markers in lung tissues. Levels of GRP78 and CHOP were substantially increased in A-f-exposed mice compared with those in control mice (figure 1A–D). A-f-induced increases of GRP78 and CHOP were significantly reduced by administration of IC87114.

Moreover, A-f-induced increases in UPR-related markers including p-IRE1α, p-eIF2α, XBP-1 and ATF-4 in A-f-exposed mice were significantly reduced by administration of IC87114 (figure 1E–I).

To demonstrate the involvement of ER stress in A-f-related allergic lung disorder in humans, we performed immunofluorescence and immunohistochemical analyses with lung tissues obtained from patients with ABPA and healthy controls. The results showed that GRP78 was remarkably increased in patients with ABPA compared with that in healthy controls (see online supplementary figure S1A–C).

PI3K-δ regulates ER stress in vitro
Epithelial cells are central in allergic lung inflammation through regulation of T(H)2-related mediators.24 In particular, we have previously shown that PI3K-δ is involved in the pathogenesis of allergic lung disorders through modulating the release of mediators in the lung epithelial cells.25 Therefore, to further assess the above interrelationship between PI3K-δ and ER stress in A-f-induced allergic lung inflammation, primary cultured TECs were used. Levels of GRP78 and CHOP were significantly increased at 12 h after A-f stimulation compared with the levels in control cells (figure 2A–D). Treatment with IC87114 significantly reduced A-f-induced increases of GRP78 and CHOP. Supporting the data, treatment with IC87114 limited A-f-induced increases in immunofluorescence intensities of GRP78 and CHOP in these cells (figure 2E).

To support the link of PI3K-δ to ER stress, PI3K-δ was knocked down in TECs using PI3K-δ-specific siRNA. The level of PI3K-δ mRNA was remarkably increased at 12 h after A-f stimulation compared with control cells, suggesting that A-f regulates PI3K-δ expression at gene level (see online supplementary figure S2). In addition, A-f-induced increase in PI3K-δ mRNA was significantly decreased by transfection of PI3K-δ-specific siRNA. Moreover, A-f-induced increases in levels of GRP78 and CHOP were significantly decreased by PI3K-δ-specific siRNA (figure 2F–I). Similarly, A-f-induced increases in immunofluorescence intensities of GRP78 and CHOP were decreased by PI3K-δ-specific siRNA (figure 2J). However, no significant changes were observed by scrambled siRNA.

IC87114 inhibits PI3K activity
To ensure an involvement of PI3K-δ in A-f-induced allergic lung inflammation, we measured levels of p-Akt and PI3K activity in the lung. The level of p-Akt protein was increased at 72 h after A-f challenge compared with the level in control mice (figure 3A, B). Consistent with these findings, PI3K activity was significantly increased after A-f challenge compared with that in control mice (figure 3C). Intratracheal administration of IC87114 reduced A-f-induced increases of p-Akt and PI3K activity. However, no significant changes in total Akt levels were observed in any of the groups tested.

Effect of IC87114 on A-f-induced allergic lung inflammation
An increase in the numbers of cells, especially eosinophils, in BAL fluids of A-f-exposed mice was significantly reduced by administration of IC87114 (figure 3D).

Airway hyperresponsiveness (AHR) was assessed as a change in respiratory system resistance (Rrs). In A-f-exposed mice, the dose-response curve of Rrs shifted to the left compared with that in control mice (figure 3E). Administration of IC87114 at 1 mg/kg substantially reduced Rrs at 25 and 50 mg/mL of methacholine.

Histological analysis revealed infiltration of various inflammatory cells into bronchioles of A-f-exposed mice (figure 3G) compared with those in control mice (figure 3F). In addition, A-f-exposed mice treated with 0.1 mg/kg (figure 3H) or 1.0 mg/kg (figure 3I) of IC87114 showed marked reduction in infiltration of inflammatory cells.

We determined levels of T(H)2 cytokines including IL-4, IL-5, and IL-13 and IL-17 in the lung. Western blot analyses showed that A-f-induced increase in these cytokines was significantly reduced by administration of IC87114 (figure 3J–Q).

Lastly, we determined whether IC87114 modifies A-f-specific T(H)2 response by analysing IgE levels. Treatment with IC87114 significantly lowered A-f-induced increases in circulating total and A-f-specific IgE level in a dose-dependent manner (figure 3R).

Effects of dexamethasone on pathophysiologic features and ER stress in A-f-induced allergic lung inflammation
Steroid resistance is a common feature in severe allergic inflammation.18 Therefore, we investigated whether dexamethasone improves pathophysiologic features of A-f-induced allergic lung inflammation. Increases in the numbers of cells including eosinophils in BAL fluids from A-f-exposed mice were not significantly changed by treatment with dexamethasone (figure 4A). In addition, treatment with dexamethasone did not improve the increase in Rrs induced by methacholine (figure 4B). In A-f-exposed mice administered dexamethasone, the grade of inflammatory cell infiltration into bronchioles was similar to that of A-f-exposed mice administered the drug vehicle only (figure 4C–E). Moreover, increases in levels of IL-4, IL-5 and IL-13 in the lung were not significantly reduced by dexamethasone (figure 4F–K). These results suggest that the current fungus-induced allergic inflammation model represents a steroid-resistant severe phenotype.

We then examined whether treatment with dexamethasone reduces A-f-induced increases in ER stress markers. Interestingly, treatment with dexamethasone failed to reduce A-f-induced elevation of GRP78 and CHOP in the lung of A-f-exposed mice (figure 4L–O).

Inhibition of ER stress alleviates A-f-induced allergic lung inflammation
To ascertain the association of ER stress with pathophysiologic features of A-f-induced allergic lung inflammation, we evaluated
the therapeutic effects of a potent ER stress inhibitor, 4-PBA. Increased numbers of cells, especially eosinophils, in BAL fluids of Af-exposed mice were reduced by administration of 4-PBA (figure 5A). Treatment with 4-PBA also reduced the Af-induced increase in $R_e$ by methacholine (figure 5B). In addition, Af-exposed mice treated with 4-PBA manifested marked...
reduction of the Af-induced infiltration of inflammatory cells into peribronchiolar regions (figure 5C–E).

4-PBA reduces nuclear translocation of NF-κB p65
To understand the molecular basis upon which inhibition of ER stress improves Af-induced allergic lung inflammation, we measured NF-κB p65 protein, a well known molecule involved in inflammation, in Af-exposed mice. The level of NF-κB p65 in nuclear protein extracts was significantly increased at 72 h after the last Af challenge compared with the control (figure 5F, G). The Af-induced increase in NF-κB p65 was decreased by administration of 4-PBA.

Effect of NF-κB inhibition on pathophysiologic features of Af-induced allergic lung inflammation
To further demonstrate the involvement of NF-κB, we evaluated the effects of a NF-κB inhibitor, BAY 11-7085, on Af-induced allergic lung inflammation.

Af-induced increases in numbers of total cells and eosinophils in BAL fluids were significantly reduced by administration of BAY 11-7085 (figure 5H). Treatment with BAY 11-7085 also attenuated the Rrs produced at 10 and 50 mg/mL of methacholine in Af-exposed mice (figure 5I). In addition, increases in numbers of infiltrated cells around bronchioles after the last Af challenge were markedly reduced by BAY 11-7085.

Figure 2
Phosphoinositide 3-kinase (PI3K)-δ modulates endoplasmic reticulum (ER) stress in vitro. Representative immunoblots of GRP78 (A) and CHOP (C) after stimulation with Aspergillus fumigatus (Af) in primary cultured murine tracheal epithelial cells (TECs) in the presence and absence of IC87114 and densitometric analyses of GRP78 (B) or CHOP (D). Bars represent mean±SEM from five independent experiments. *p<0.05 versus control; #p<0.05 versus cells stimulated with Af alone. (E) Representative confocal laser immunofluorescence photomicrographs of primary cultured TECs showed the localisation of GRP78 and CHOP in controls (no treatment), Af-stimulated cells (Af), Af-stimulated cells administered drug vehicle (dimethyl sulfoxide, DMSO) (Af+DMSO), or Af-stimulated cells administered IC87114 (Af+IC). 4',6-diamidino-2-phenylindole (DAPI) stain was used for nuclear localisation. The right panels show the merger views. Bars indicate 20 μm. Representative immunoblots of GRP78 (F) and CHOP (H) after stimulation with Af in primary cultured murine TECs transfected with either scrambled small interfering RNA (Scram siRNA) or PI3K-δ-specific siRNA and densitometric analyses of GRP78 (G) or CHOP (I). Bars represent mean±SEM from six independent experiments. *p<0.05 versus control; #p<0.05 versus cells stimulated with Af transfected with scrambled siRNA. (J) Representative confocal laser immunofluorescence photomicrographs of primary cultured TECs showed the localisation of CHOP and GRP78 in controls (no treatment), Af-stimulated cells (Af), Af-stimulated cells administered scrambled siRNA (Af+Scram), or Af-stimulated cells administered PI3K-δ-specific siRNA (Af+PI3K-δ). Bars indicate 20 μm.
Figure 3  Inhibition of phosphoinositide 3-kinase (PI3K)-δ ameliorates *Aspergillus fumigatus* (*Af*)-induced allergic lung inflammation. Representative immunoblots of p-Akt and total (t)-Akt in lung tissues (A) from SAL+dimethyl sulfoxide (DMSO), *Af*+DMSO, *Af*+IC 0.1 or *Af*+IC 1.0, and densitometric analyses of p-Akt (B). Bars represent mean±SEM from six mice per group. (C) Enzyme immuncassays of PIP3 in lung tissues. Bars represent mean±SEM from six mice per group. (D) Cellular changes in bronchoalveolar lavage (BAL) fluids. Bars represent mean±SEM from six mice per group. (E) Airway responsiveness assessed by invasive (Rrs) measurements. Bars represent mean±SEM from five mice per group. Representative H&E stained sections of the lung from SAL+DMSO (F), *Af*+DMSO (G), *Af*+IC 0.1 (H) or *Af*+IC 1.0 (I). Bars indicate scale of 50 μm. Representative immunoblots of IL-4 (J), IL-5 (L), IL-13 (N) and IL-17 (P) in lung tissues and densitometric analyses of IL-4 (K), IL-5 (M), IL-13 (O) and IL-17 (Q). Bars represent mean±SEM from six mice per group. *p<0.05 versus SAL+DMSO; **p<0.05 versus *Af*+DMSO.

Lee KS, et al. Thorax 2016;71:52–63. doi:10.1136/thoraxjnl-2015-207096 57}

Thorax: first published as 10.1136/thoraxjnl-2015-207096 on 5 November 2015. Downloaded from http://thorax.bmj.com on April 23, 2024 by guest. Protected by copyright.
Furthermore, Af-induced increases in IL-4, IL-5 and IL-13 protein were substantially reduced by BAY 11-7085 (figure 5M, N). IC87114 restores GSH Oxidative stress is one of the most important features in allergic lung inflammation. Therefore, we measured GSH and GSSG in lung tissues. Af-exposed mice showed significantly lower GSH level than control mice (figure 6A). In contrast, GSSG level was increased after Af challenge (figure 6B). Af-induced decrease in GSH and increase in GSSG were substantially restored by administration of IC87114.

IC87114 reduces Af-induced mtROS generation Mitochondrion has been recognised as a major source of intracellular ROS. We therefore examined the relationship between PI3K-δ and mtROS generation in our experimental systems. Immunofluorescence intensity of mtROS in BAL cells was markedly increased in Af-exposed mice compared with the intensity in control mice (figure 6C). Administration of IC87114 reduced Af-induced increase in immunofluorescence intensity of mtROS. Consistent with in vivo data, Af-induced increase in fluorescence intensity of mtROS in TECs was markedly inhibited by administration of IC87114 (figure 6D). These results indicate that PI3K-δ may induce mtROS generation.

NecroX-5 ameliorates inflammatory cell infiltration and AHR To further investigate the role of mtROS in Af-induced allergic lung inflammation, we evaluated the treatment effects of a potent mtROS scavenger, NecroX-5. Increased numbers of cells, particularly eosinophils, in BAL fluids from Af-exposed mice were markedly reduced by administration of NecroX-5.
In addition, Af-exposed mice treated with NecroX-5 showed marked reduction in infiltration of inflammatory cells into the peribronchiolar region compared with that in control mice (figure 7A). Moreover, administration of NecroX-5 resulted in a significant attenuation in $R_{ns}$ at 50 mg/mL of methacholine (figure 7E).

**NecroX-5 reduces GRP78 and CHOP**

To investigate whether mtROS modulates ER stress, we evaluated the interrelationship between them using NecroX-5. Levels of GRP78 and CHOP were significantly elevated in Af-exposed mice compared with those in control mice (figure 7F–I). Interestingly, administration of NecroX-5 markedly lowered Af-induced increases in these proteins, suggesting that mtROS generation induces ER stress.

**Effect of NecroX-5 on Af-induced mtROS generation in BAL fluids**

Immunofluorescence intensity of mtROS in BAL cells was markedly increased in Af-exposed mice compared with that
control mice (figure 7J). Administration of NecroX-5 substantially reduced Af-induced increase in immunofluorescence intensity of mtROS.

DISCUSSION
In this study, ER stress was increased in the lung of Af-exposed mice. Expression of GRP78 was also elevated in lung tissue from patients with ABPA, a hypersensitivity disorder induced by Af, compared with that in healthy controls. These results suggest that ER stress may be involved in the pathogenesis of Af-related allergic lung disorders. Additionally, we found a novel mechanism of action of PI3K-δ as an upstream regulator of ER stress through mtROS generation in Af-induced allergic lung inflammation. Also, blockade of PI3K-δ remarkably ameliorated Af-induced steroid-resistant allergic lung inflammation. These findings suggest that PI3K-δ plays an essential role in the pathogenesis of fungus-induced severe allergic lung inflammation.

Sensitivity to fungi has been known as one of the crucial determinants complicating clinical course of bronchial asthma. Among various species, Af is a major indoor and outdoor fungi. Allergic sensitisation to Af in patients with bronchial asthma is often associated with reduced lung function and immunologically mediated structural lung damage. Furthermore, the conventional anti-inflammatory agents including corticosteroids may be insufficient for treating severe asthma associated with fungal sensitisation. In this study, treatment with dexamethasone failed to alleviate various pathophysiologic features of Af-induced allergic lung inflammation, including eosinophil-dominant inflammatory cell infiltration into the lung, AHR, and elevation of T H2-dominant pro-inflammatory cytokines. In fact, we revealed that under an identical protocol to this study, the same dose of dexamethasone attenuates the typical features of allergic asthma induced by ovalbumin which is a well established classic asthma model. These results suggest that Af-induced lung inflammation in the current murine experimental model represents a unique endotype for steroid-resistant eosinophil-dominant allergic lung inflammation.

ER stress and UPR activation are known to be closely involved in pathogen immunity by influencing innate and adaptive immune responses and being interconnected with diverse intracellular inflammatory platforms. Previous studies have demonstrated that ER stress is implicated in the pathogenesis of various pulmonary disorders, including bronchial asthma, COPD, idiopathic pulmonary fibrosis and lipopolysaccharide-related acute lung injury. Particularly, we recently reported that ER stress contributes to corticosteroid resistance in a murine model of neutrophil-dominant allergic lung inflammation.
Inflammation. In the current study, sensitisation/challenge with \( Af \) increased expression of ER stress markers and UPR-related proteins in the lung. Expression of ER stress markers was also elevated in \( Af \)-stimulated TECs. Moreover, in lung tissues from patients with ABPA, expression of GRP78 was also increased. In addition, blockade of ER stress improved \( Af \)-induced eosinophilic allergic lung inflammation and AHR. However, \( Af \)-induced increases in the levels of GRP78 and CHOP were not reduced by treatment with dexamethasone. These findings suggest that ER stress is involved in the pathogenesis of \( Af \)-induced allergic lung inflammation.

Activation of transcription factor NF-\( \kappa \)B serves as an important molecular platform interconnecting ER stress and inflammatory process. Recently, the NF-\( \kappa \)B signalling pathway has been reported to be essential in ER stress-related lung inflammation. In the present study, the level of nuclear NF-\( \kappa \)B p65 in the lung was significantly increased after \( Af \) challenge. \( Af \)-induced increase in NF-\( \kappa \)B p65 was decreased by administration of an ER stress inhibitor. Additionally, treatment with a NF-\( \kappa \)B inhibitor substantially reduced increases in pro-inflammatory mediators, inflammatory cell infiltrations specifically eosinophils, and AHR in \( Af \)-exposed mice. Taken together, these findings suggest that NF-\( \kappa \)B is a pivotal player in ER stress-related allergic lung inflammation.

The PI3K pathway mediates a number of critical cellular events, such as cell survival, proliferation and migration. As for \( Af \)-related inflammatory disorders, to date, it remains unclear about the roles of PI3K and its isotypes. In fact, the PI3K pathway has been known to participate not only in host defence against \( Af \) but also in the pathogenesis of \( Af \)-induced inflammatory disorders or aspergillosis. In addition, there is little available information on the PI3K-\( \delta \) subtype in \( Af \)-related pulmonary disorders. Previous studies have reported that PI3K-\( \delta \) is crucially involved in the pathogenesis of a number of pulmonary inflammatory diseases, including bronchial asthma and COPD. In particular, PI3K-\( \delta \) is known to mediate steroid resistance in relation to neutrophil-dominant inflammatory processes in the lung. The basic mechanisms through which PI3K-\( \delta \) modulates neutrophilic inflammation include trafficking and activation of neutrophils, and decrease in histone deacetylase-2...
However, severe allergic lung inflammation is heterogeneous and may arise from diverse pathophysiological processes involving various cell types. Among them, eosinophilic inflammation affects the majority of patients with severe asthma. In our study, Afl challenge in mice increased levels of PIP3 and p-Akt in the lung. Stimulation with Afl also upregulates PI3K-δ mRNA expression in TECs. In Afl-exposed mice, PI3K-δ inhibition with a selective inhibitor significantly lowered Afl-induced increases in PIP3 and p-Akt and reduced infiltration of airway inflammatory cells, especially eosinophils, production of pro-inflammatory mediators, total and Afl-specific IgE, and AHR. Interestingly, PI3K-δ inhibition significantly limited Afl-induced increases in ER stress markers and UPR-related proteins in the lungs. In addition, the PI3K-δ inhibitor significantly limited Afl-induced increases in ER stress markers in TECs. Transfection of PI3K-δ siRNA also significantly limited Afl-induced increases in GRP78 and CHOP in cells. In agreement with these findings, blockade of PI3K-δ using the PI3K-δ inhibitor or PI3K-δ siRNA in cells substantially limited Afl-induced increases in immunofluorescence intensities of GRP78 and CHOP. These observations indicate that PI3K-δ may play an important role in Afl-induced steroid-resistant eosinophilic-dominant allergic lung inflammation. Moreover, PI3K-δ seems to contribute to steroid resistance in the current model through induction of ER stress in the lung.

ROS production, which leads to oxidative stress, is one of critical features in chronic airway disorders. In addition, oxidative stress contributes to diverse pathologic processes through modulation of the PI3K signalling pathway. Emerging evidence has indicated that mitochondrion is an important source of intracellular ROS and that considers interplay between mitochondria and ER in several respiratory disorders has been demonstrated. Moreover, PI3K signalling is closely associated with mitochondrial physiology.

Figure 8 Proposed mechanism for the involvement of phosphoinositide 3-kinase (PI3K)-δ in the regulation of endoplasmic reticulum (ER) stress in Aspergillus fumigatus (Afl)-related steroid-resistant allergic lung inflammation. NF-κB, nuclear factor κB.

In the present study, BAL cells of Afl-exposed mice and Afl-stimulated TECs showed increased production of mtROS. In addition, blockade of PI3K-δ reduced Afl-induced increases in mtROS generation in these cells. PI3K-δ inhibition also blocked Afl-induced decreases in GSH level in the lung of Afl-exposed mice. Moreover, treatment with NecroX-5 effectively attenuated increases in mtROS in BAL cells, reduced increases in GRP78 and CHOP in Afl-exposed mice, and ameliorated pathophysiologic features of Afl-induced allergic airway inflammation. Taken together, PI3K-δ may activate the ER stress pathway at least in part through regulation of mtROS generation in the current model.

In summary, we have shown that PI3K-δ and ER stress are involved in the pathogenesis of Afl-induced allergic lung inflammation and that PI3K-δ may regulate Afl-induced steroid-resistant lung inflammation through ER stress (figure 8). Our results suggest that inhibition of PI3K-δ is efficacious in preclinical models of fungus-induced severe allergic lung disease and may have potential for treating patients with steroid-resistant eosinophil-dominant allergic lung disorders.

Acknowledgements We thank Professor Mie-Jae Im and Peter Lee for critical readings of the manuscript.

Contributors KSL performed experiments and analysed data; JSJ interpreted data and wrote the manuscript; SRK designed research, interpreted data, and edited the manuscript; SCP analysed data and reviewed the manuscript; HJP performed experiments and analysed data; KBL analysed data and reviewed the manuscript; YHK performed experiments and analysed data; KBL designed research, interpreted data, and edited the manuscript.

Funding This work was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT and Future Planning (2014R1A2A1A01002823).

Competing interests None declared.

Ethics approval Institutional Review Board of the Biomedical Research Institute of Chonbuk National University Hospital (IRB file No. 2013-11-007-001).

Provenance and peer review Not commissioned; externally peer reviewed.

Open Access This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.

REFERENCES

1. Krutzen AP, Bush RK, Demain JG, et al. Fungi and allergic lower respiratory tract diseases. J Allergy Clin Immunol 2012;129:280–91.
2. Agarwal R. Allergic bronchopulmonary aspergillosis. Chest 2009;135:805–26.
3. Vlahakis NE, Akssomit TR. Diagnosis and treatment of allergic bronchopulmonary aspergillosis. Mayo Clin Proc 2001;76:930–8.
4. Deming DW, O’Driscoll BR, Hogoaboam CM, et al. The link between fungi and severe asthma: a summary of the evidence. Eur Respir J 2006;27:615–26.
5. Menzies D, Holmes L, McCumesky G, et al. Aspergillus sensitization is associated with airflow limitation and bronchiectasis in severe asthma. Allergy 2011;66:679–85.
6. Kwak YG, Song CH, Yi HK, et al. Involvement of PTEN in airway hyperresponsiveness and inflammation in bronchial asthma. J Clin Invest 2003;111:1083–92.
7. Wyman MP, Pirila L. Structure and function of phosphoinositide 3-kinases. Biochim Biophys Acta 1998;1436:127–50.
8. Vanhaesebroeck B, Ali K, Blanco A, et al. Signalling by PI3K isoforms: insights from gene-targeted mice. Trends Biochem Sci 2005;30:194–204.
9. Okkenhaug K, Blanco A, Farjot G, et al. Impaired B and T cell antigen receptor signaling in p110delta PI 3-kinase mutant mice. Science 2002;297:1031–4.
10. Ali K, Camps M, Pearce WP, et al. Isoform-specific functions of phosphoinositide 3-kinases: p110 delta but not p110 gamma promotes optimal allergic responses in vivo. J Immunol 2008;180:2538–44.
11. Puri KD, Doggett TA, Douangpanya J, et al. Mechanisms and implications of phosphoinositide 3-kinase delta in promoting neutrophil trafficking into inflamed tissue. Blood 2004;103:3448–56.
