Fentanyl Ameliorates Severe Acute Pancreatitis-Induced Myocardial Injury in Rats by Regulating NF-κB Signaling Pathway

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Background: Acute pancreatitis (AP) is a sudden inflammation of the pancreas. It results in multiple, severe complications, and 15–20% of patients develop severe acute pancreatitis (SAP) with mortality as high as 30%. Consequently, it is imperative to develop an effective therapy for SAP.

Material/Methods: We used 30 adult male Sprague Dawley (SD) rats. Rats were randomly divided into 3 groups – sham, SAP, and fentanyl+SAP – with 10 rats in each group. An automatic biochemical analyzer was used to analyze the concentration of creatine kinase isoenzyme (CK-MB) and lactate dehydrogenase (LDH). Terminal-deoxynucleotidyl transferase-mediated nick-end labeling (TUNEL) assay was applied to assess the cell apoptosis rate. Pathological changes in pancreas/heart were detected with hematoxylin and eosin (HE) staining. Western immunoblot assay was used to analyze protein levels of interleukin (IL)-1β, IL-6, and IκB.

Results: Fentanyl pre-treatment inhibits SAP-induced elevation of CK-MB/LDH concentrations in serum. Compared with the sham group, SAP generates a higher brown/yellow staining rate, which is abated by fentanyl. In the pancreas, SAP generated more serious interstitial edema/hemorrhage and fat necrosis than in the sham group, which are attenuated by fentanyl. Likewise, compared to the sham group, SAP generates swollen/disordered myocardial fibers and congested blood vessels in myocardium, which are ameliorated by fentanyl. In the sham group, there was little IL-1β/IL-6, and fentanyl significantly inhibited SAP-induced up-regulation of IL-1β/IL-6 levels. Compared with the sham group, SAP significantly reduced IκB level, which was rescued by fentanyl.

Conclusions: Fentanyl effectively alleviates SAP-induced pancreas and heart injuries through regulating the nuclear factor-kB (NF-κB) signaling pathway.

MeSH Keywords: Cardiomyopathies • Fentanyl • Pancreatitis, Acute Necrotizing

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Background

AP is a sudden inflammation in the pancreas [1], with severe complications and high mortality despite treatment. Unfortunately, in 15–20% of patients, AP progresses into SAP, with high (30%) mortality [2,3]. Patients with mild SAP can be effectively treated by conservative therapies (fasting and intravenous fluid rehydration), while patients with severe SAP need intensive care. SAP is an abdominal disease frequently encountered by surgeons [4]. Despite mounting achievements in modern medicine and increasing clinical/experimental studies, the specific pathogenesis of SAP remains elusive [5,6]. Surprisingly, nuclear factor-κB (NF-κB), inflammatory cytokines, leukocytic infiltration, and oxidative stress have been shown to be pivotal factors during the process of SAP [7,8]. Inflammation is responsible for the morbidity and mortality of AP [9]. Regrettably, there remains no effective therapy for SAP [10].

SAP causes multiple-organ injuries (e.g., renal failure [11]), and the morbidity of SAP-induced myocardial injury is reported to be 60.5% [12].

Fentanyl (intravenous) has recently been used for pain relief in AP, especially in renal impairment [13], and is increasingly used due to its good safety profile. Consequently, the morbidity of SAP-induced complications has not been substantially reduced to date. It is urgent to seek an effective medicine to successfully treat SAP and SAP-induced myocardial injury. The present study investigated the possible protective role of fentanyl against SAP-induced myocardial injury in rats and to explore the possible molecular mechanism.

Material and Methods

Animals

We randomly separated 30 male Sprague Dawley (SD) rats weighing 200–250 g into 3 different groups — sham, SAP, and fentanyl treatment — with 10 rats in each group. Rats were housed under pathogen-free conditions with controlled temperature (22±2°C), a relative humidity of 30–70%, 12-h light-dark cycle, and provided with water/food ad libitum. Each animal experiment was conducted in accordance with the animal care guidelines of the Chinese National Institutes of Health and the study was approved by the Central Hospital of Wuhan Animal Care and Use Committee.

Model preparation

In brief, anesthetization of rats was performed with 10% chloral hydrate (3 ml/kg) prior to abdomen incision. Upon the location of biliopancreatic duct and opening of duodenal papilla, a syringe needle was placed. Thereafter, the cholangitic porta hepatitis was clipped with a small artery clamp, 5% sodium taurocholate (1 ml/kg) was injected into the pancreatic duct at the speed of 0.1 ml/min. Five min later, the clamp was removed, followed by incision suture and abdomen closing.

At 23–23.5 h after SAP preparation, rats in the fentanyl group received an intravenous administration of fentanyl (30 μg/kg diffused in 0.5 ml saline) and sustained for 30–60 min, while rats in sham and SAP groups were intravenously injected with 0.5 ml saline. For rats in the sham group, an incision was made in the abdomen and pancreatic tissue was marginally rotated several times, then the abdomen was closed. At 24 h after SAP preparation, rats were killed and prepared for further studies.

Specimen collection

Rats were anesthetized by intraperitoneal injection with 10% chloral hydrate, followed by abdomen opening, pancreas/heart removal, and abdominal aorta blood collection under sterile conditions. Heart tissues were used for the investigation of myocardial cell apoptosis and analysis of pathological changes, and pancreas tissues were utilized for determining pathological scores.

Assessment of myocardial injury

We let the obtained arterial blood coagulate naturally at room temperature for about 1 h. Afterwards, serum was collected via centrifuging the coagulated arterial blood at the speed of 1006.2×g at 4°C for 10 min. Concentrations of CK-MB and LDH were detected by use of a Vitros 250 automatic biochemical analyzer.

TUNEL staining

TUNEL staining was carried out using a cell death detection kit in accordance with manufacturer’s instructions to detect the apoptotic rates of myocardial cells in different groups. Nuclei were stained by 3, 3-diaminobenzidine (DAB). Nuclei of healthy cells were stained blue, while those in apoptotic cells presented brown/yellow staining. Ten areas were randomly selected from each section. Double-blind condition was adopted for the counting of TUNEL-positive cells.

HE staining

Pancreatic and heart tissues were fixed in 4% neutral paraformaldehyde (PFA) solution for approximately 24 h, dehydrated through graduated ethanol, embedded with paraffin, and finally sliced into 5-μm-thick sections. In brief, slices were de-waxed in xylene and rehydrated by upgraded ethanol. After rinsing 3 times with distilled water, slices were immersed in...
hematoxylin for approximately 5 min. Before slice differentiation with 1% HCl, a second rinsing was carried out. Thereafter, slices were further stained with eosin for about 4 min. At the end of the experiment, slices were dehydrated and differentiated in alcohol. Thereafter, morphological changes of tissues were studied under light microscopy.

**Pathological score**

According to changes in edema, infiltration of inflammatory cells, hemorrhage, and necrosis in pancreatic tissues, the severity of pathological changes and corresponding scores were recorded: 0, no; 1, mild; 2, moderate; 3, severe; and 4, very severe.

In accordance with pathological changes in infiltration of inflammatory cells and myocardial necrosis in heart tissues, severity of pathological changes and corresponding scores were recorded: 0, no; 1, mild; 2, moderate; 3, severe; 4, very severe; and 5, extremely severe.

**Western immunoblot**

Heart tissues from 3 different groups were homogenized in lysis buffer. In brief, samples were resolved via sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and electro-blotted to Immobilon-P nylon membranes. Filters were first treated with primary antibody at room temperature for about 3 h. Thereafter, filters were treated with the corresponding horseradish peroxidase (HRP)-labeled secondary antibody at room temperature for about 1 h. The immune complexes were visualized with enhanced chemiluminescence.

**Statistical analysis**

All of the experiments were performed in duplicate and repeated at least 3 times. Data were analyzed using two-way analysis of variance (ANOVA) and are expressed as mean ± standard deviation (SD). SPSS (Chicago, IL, USA) was used for statistical analysis. P<0.05 showed a significant difference.
Results

Fentanyl pre-treatment inhibits SAP-induced up-regulation of CK-MB and LDH in fentanyl+SAP group

At 24 hours after modeling, 2 rats in the SAP group and 1 rat in the fentanyl treatment group died, and there was no significant difference. Concentrations of CK-MB and LDH in the serum were detected. Results of automatic biochemical analyzer revealed that the concentration of CK-MB in the sham group was 505 u/ml, in the SAP group it was 1806 u/ml, and in the fentanyl+SAP group it was 1020 u/ml. The concentration of LDH in the sham group was 703 u/l, in SAP group it was 1516 u/ml, and in the fentanyl+SAP group it was 993 u/ml. In comparison with the sham group, SAP notably up-regulated concentrations of CK-MB and LDH, which were significantly repressed by fentanyl pre-treatment.

Fentanyl pre-treatment decreased SAP-induced elevation of myocardial cell apoptosis in the fentanyl+SAP group

TUNEL assay was carried out to identify whether fentanyl affected cell apoptosis of myocardial tissues induced by SAP. In the sham group, the majority of nuclei were stained blue (Figure 1A). Many nuclei were stained brown/yellow in the SAP group (Figure 1B), which was predominantly lowered by fentanyl, but was still higher than in the sham group (Figure 1C).

The apoptotic index was significantly higher in the SAP group than in the sham group (P<0.01), and was alleviated by fentanyl (P<0.05) (Figure 1D).

Fentanyl pre-treatment alleviated SAP-induced pathological features in pancreases in the fentanyl+SAP group

HE staining was conducted to detect the effects of fentanyl on pathological changes pancreases. Pancreases in the sham group exhibited normal morphology (Figure 2A). However, SAP
resulted in serious morphological changes in interstitial edema, interstitial hemorrhage, and fat necrosis (Figure 2B). After treatment with fentanyl, pathological features were notably improved (Figure 2C).

Pathological scores of pancreases are displayed in Figure 2D. Compared with the sham group (1.6), pathological scores in the SAP group were significantly higher (10.3). Moreover, following treatment with fentanyl, pathological scores (5.7) were significantly lower than in the SAP group.

**Fentanyl pre-treatment alleviated SAP-induced pathological features in hearts in the fentanyl+SAP group**

Similarly, HE staining of myocardial tissue in the sham group exhibited normal morphology (Figure 3A), but in the SAP group myocardial fibers were seriously degenerated, swelled, and disordered and the blood vessels were congested (Figure 3B). After fentanyl treatment, pathological features of hearts were improved (Figure 3C).

Pathological scores of hearts are shown in Figure 3D. Pathological scores in the SAP group (2.9) were significantly higher than in the sham group (0.3), and were significantly (1.8) lower in the fentanyl group.

**Fentanyl pre-treatment inhibited SAP-induced IL-1β/IL-6 up-regulation in hearts in the fentanyl+SAP group**

Western blot results showed that in the sham group there was only a low level of IL-1β/IL-6, and fentanyl treatment significantly inhibited SAP-induced up-regulation of IL-1β/IL-6 level (Figure 4A).

We also did band density analysis, and data were in line with that in Western blot experiments (Figure 4B).
Fentanyl pre-treatment rescued SAP-induced reduction of IkB level in rat hearts in the fentanyl+SAP group.

Results demonstrated that, compared with the sham group, SAP significantly reduced IkB level, and fentanyl rescued SAP-induced IkB reduction (Figure 5A).

The corresponding statistical data are presented in Figure 5B.

**Figure 4.** Effect of fentanyl on SAP-induced expression changes of IL-1β and IL-6 in heart (A). Data of band densities were presented (B). **P<0.01 vs. sham group; # P<0.05 vs. SAP group.

**Figure 5.** Effect of fentanyl on SAP-induced expression change of IkB in heart (A). Data of band densities were exhibited (B). **P<0.01 vs. sham group; # P<0.05 vs. SAP group.
ANIMAL STUDY

Discussion

AP is a fatal disease whose pathogenesis remains unclear despite remarkable advances achieved over the past 25 years [14]. Fentanyl was reported to be used for pain relief in AP [13] and is being increasingly used by virtue of its safety profile, especially in renal impairment. The present study aimed to investigate whether fentanyl has a protective role in SAP-induced myocardial injury in rats and to provide a possible molecular mechanism.

CK-MB and LDH in the serum are markers of myocardial injury [15,16]. We first detected changes in CK-MB and LDH concentrations in the present study. Results demonstrated that up-regulation of CK-MB and LDH levels in SAP rats were notably inhibited by fentanyl, which provides direct evidence that fentanyl indeed exerted myocardial protective effects in rats with SAP.

The myocardial protective effects of fentanyl were further verified by TUNEL assay, which displayed that the apoptosis index in the fentanyl group was lower than in the SAP group.

Pathological features in pancreatic and myocardial tissues in the SAP group were remarkably attenuated after treatment with fentanyl, which is consistent with SAP-induced injuries. Consistently, the pathological scores of pancreases and hearts in the fentanyl treatment group were significantly lower than those in the SAP group, indicating that fentanyl can improve SAP-induced pathological changes.

Taken together, our present results verified that fentanyl effectively reduced serum levels of CK-MB and LDH, and inhibited myocardial cell apoptosis, ultimately reducing SAP-generated myocardial injuries. However, the corresponding precise molecular mechanism remains elusive. As the role of fentanyl in pain relief in AP [13] and co-regulators between pain and inflammation [17], we hypothesized the possible mechanism based on previous studies.

AP, together with sepsis, trauma, burns, and surgery, has the potential to generate systemic inflammatory response syndrome (SIRS) [18], in which cytokines act as initiators, enhancers, and damaging agents [19].

Cytokines can activate numerous signaling pathways; for instance, NF-κB can deteriorate inflammation [20,21]. NF-κB is activated at the initial stage of AP and plays a crucial role in inflammatory response during AP [8,22]. Moreover, Satoh et al. reported an obviously elevated activation of NF-κB following the presence of SAP [23]. Free NF-κB is translocated to the nucleus and is bound with the κB site, thus causing massive transcription of plentiful inflammation-correlated genes, including IL-1β and IL-6 [24,25].

Therefore, fentanyl might play a protective role in SAP-induced myocardial injury via inhibiting the activation of the NF-κB signaling pathway, ultimately attenuating the synthesis and release of inflammatory factors. We carried out further studies to investigate whether the aforementioned proposed hypothesis was right.

We first detected the protein levels of IL-1β and IL-6, showing that they were significantly elevated in SAP rats compared to the sham group rats, which were inhibited by fentanyl treatment.

NF-κB can generate transcription of IL-1β/IL-6 and is involved in the process of AP [22–25]. The NF-κB/Rel family contains NF-κB1 (p50), NF-κB2 (p52), Rel A (p65), Rel B, r-Rel, and c-Rel [26–30]. NF-κB is sequestered when bound to IκB (inhibitory element) and stimulated IκB releases NF-κB [29–31]. Furthermore, no detectable changes were found in expression levels of either p50 or p65; IκB levels were reduced in early pancreatitis and elevated with resolving pancreatitis [25]. Thus, we tested IκB levels by Western blot analysis. Results demonstrated that SAP-inhibited activation of IκB was rescued by fentanyl, which is consistent with a previous study [25].

Furthermore, Isik A et al. reported the protective effect of ozone and naringin on intestinal I/R injury [32], and astragaloside IV was recently reported to attenuate AP via reducing the activation of NF-κB in rats [33]. The PI3K/Akt-NF-κB pathway was recently discovered to be activated in AP rats [34]. Our findings are in line with previous studies showing the involvement of the NF-κB pathway in AP, thus providing a possible treatment for SAP via modulating the NF-κB pathway.

Conclusions

Fentanyl had a protective role in SAP-induced myocardial injury in rats via repressing the NF-κB signaling pathway, ultimately reducing the protein level of inflammatory factors.

Disclosure

There are no conflicts of interest to declare.
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