The first case of Coronavirus disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) was reported in December 2019. This virus belongs to the beta-coronavirus group that contains a single stranded RNA with a nucleoprotein within a capsid. SARS-CoV-2 shares 80% nucleotide identity to SARS-CoV. The virus is disseminated by its binding to the ACE2 receptors on bronchial epithelial cells. The diagnosis of COVID-19 is based on a laboratory-based reverse transcription polymerase chain reaction (RT-PCR) test together with chest computed tomography imaging. To date, no antiviral therapy has been approved, and many aspects of the COVID-19 are unknown. In this review, we will focus on the recent information on genetics and pathogenesis of COVID-19 as well as its clinical presentation and potential treatments.

First draft submitted: 11 May 2020; Accepted for publication: 7 August 2020; Published online: 26 August 2020

Keywords: betacoronavirus • remdisivir • SARS-CoV-2 • spike surface glycoprotein

Coronaviruses (CoVs) are enveloped, positive-sense RNA viruses that are primarily responsible for infections of the upper respiratory and digestive tract [1,2]. CoV originates from the Latin word corona. The peplomers on the surface of the virus create an image reminiscent of a solar corona on electron microscopy. These viruses associated with severe acute respiratory syndrome CoV (SARS-CoV) and Middle East respiratory syndrome CoV (MERS-CoV) [1,2], that occurred as outbreaks in 2002 and 2013, respectively, and both have been associated with severe complications, that include, severe pneumonia and bronchiolitis [1]. CoVs belong to the subfamily Coronavirinae, family Coronaviridae and order Nidovirales, which can be divided into four genera: alphacoronavirus (α-CoVs), betacoronavirus (β-CoVs), gammacoronavirus (γ-CoVs) and deltacoronavirus (δ-CoVs) [3].

There are seven types of CoVs known to cause infections in humans including 229E (α-CoVs), NL63 (α-CoVs), OC43 (β-CoVs), HKU1 (β-CoVs), MERS-CoV (β-CoVs), SARS-CoV (β-CoVs) and recent novel β-CoVs (SARS-CoV-2; Figure 1) [4-6]. The three last viruses can affect respiratory, gastrointestinal, hepatic and nervous systems, and may cause life-threatening respiratory and multiple organ failure and death in some severely affected patients [7,8]. α-CoVs and β-CoVs have been responsible for a heavy disease burden on livestock; for example, an HKU-2 bat origin, swine acute diarrhea syndrome coronavirus emerged in the swine population in 2017 in Guangdong in China [9].

The first case of a coronavirus disease 2019 (COVID-19) infection was reported in December 2019 in Wuhan City, Hubei Province, China and after that it has spread globally to being declared to be a pandemic by the WHO in 2020 [10]. SARS-CoV-2, was first isolated from patients with pneumonia of unknown etiology, and may have originated from a ‘wet market’ where wild animals were being sold in Wuhan. But, Wuhan seafood market probably only served as an amplifying hotspot and not is source of novel virus spreading globally. High levels of SARS-CoV-2 transmission have now been reported in the USA, Italy, Iran, Germany and France. More than 12 million affected cases and 500,000 deaths globally have been reported until 10 July. The estimated mean reproduction number (R0) for COVID-19 without intervention, is around 3.28 which is higher than for SARS-CoV [11].
The SARS-CoV-2 genome & structure

All the CoVs are positive-stranded RNA viruses with a polycistronic genome nearly 29.9 kb in length with 6–11 open reading frames (ORFs), encoding several nonstructural proteins (ORF1a and ORF1b, that are processed into 15 nsp proteins) at the 5′-end plus four structural proteins (spike surface glycoprotein [S], envelope [E], matrix [M] and nucleocapsid [N]) and multiple lineage-specific accessory proteins (i.e., ORF3a, ORF6, ORF7a, ORF7b, ORF8 and ORF10 in SARS-CoV-2) at the 3′-end (Figure 2A) [3,12,13]. The SARS-CoV-2 virus also encodes other poly-proteins, nucleoproteins and membranous proteins, in other words RNA polymerase, helicase and various proteases [13,14].

The initial infection requires viral entry into the host cell. The S-glycoprotein mediates binding of the virus to the human cell surface receptors, followed by fusion of the virus and host cell membranes to assist viral entrance (Figure 3) [15,16]. The S-glycoprotein is a structural protein that accounts for the crown-like shape of the CoVs. The 1200 AA length S-protein (approx. 180 kDa) is a member of class-I viral fusion proteins[17,18]. It is expressed on the surface of the virus as a trimeric protein and also determines the host tropism and pathogenesis [19–21]. The S-glycoprotein can be cleaved by host proteases to an N-terminal S1-ectodomain and C-terminal S2-membrane-anchored protein (Figure 2B) [22]. In several CoVs, the higher amplification of S-protein at the cell membrane can also facilitate membrane fusion of infected cells with adjacent uninfected cells, and this leads to the formation of giant or multinucleated cells (syncytia) and further spread of the virus between cells [23–25]. The CoVs S1-protein includes a receptor-binding domain (RBD), which binds to the host cell via receptors that include the ACE2 receptor [26]. SARS-CoV uses the ACE2 receptor for entry [26] and uses the serine protease, cathepsins and TMPRSS2, for S-protein priming [24,27]. Of 14 AA residues in the S1 subunit of SARS-CoV, eight residues are highly conserved in SARS-CoV-2, suggesting that the ACE2 receptor is also used for cell entry by this virus [28]. Labeling studies have confirmed this [4,29]. The S-glycoprotein of SARS-CoV-2 has a 3D structure within the RBD domain that conserve the van der Waals interactions [4]. The 394 glutamine residue of the SARS-CoV-2’s RBD domain is ligated via the important lysine 31 residue of the human receptor, ACE2 [28].

An E-protein is involved in the assembly and release of virus-like particles [30–32]. M-protein provides the shape of the viral envelope and binding to nucleocapsid [33], whereas, the N-protein binds to the CoV RNA genome, building the nucleocapsid to support replication–transcription complex [34]. Interaction between M and N-proteins stabilize the nucleocapsid and the internal core of virus, and, eventually, enhance the assembly of the virion particles [25,35,36]. M-, E- and S-proteins then enter into the endoplasmic reticulum–Golgi intermediate compartment complex and...
constitute the viral envelope [36]. Accessory proteins appear to promote the adaptation of CoVs to human host cells [37].

Genomic analysis of ten genome of SARS-CoV-2 isolated from nine patients demonstrated 99.98% nucleotide identity [38]. Another report found that 99.8–99.9% sequence similarity in sample of five infected patients [39]. Phylogenetic analysis demonstrates that SARS-CoV-2 shares 50 and 80.0% nucleotide identity to MERS-CoV and SARS-CoV, respectively [8,13,38].

The SARS-CoV-2 constitutes a clade among the sub-genus sarbecovirus [40]. Bioinformatics analysis of the viral genome from one COVID-19 patient shared 89 and 82% sequence similarity with bat SARS-like-CoVZXC21 and human SARS-CoV, respectively [41]. However, the external subunit of Spike RBD of SARS-CoV-2 has only 40% amino acid (AA) identity with other SARS-associated CoVs [41]. The S-protein of SARS-CoV-2 is longer (1282 AA) than for other viruses such as SARS-CoV (1255 AA) and Bat SARS-like CoVs (1246 AA). The S-glycoprotein of SARS-CoV-2 has been found to have three short insertions at the N-terminal end, with four variations in the receptor binding site within the RBD compared with SARS-CoV [42]. Notably, SARS-CoV-2 ORF3b codifies a new short protein. Moreover, its novel ORF8 sequence possibly encode a secreted protein with an α-helical structure.
Figure 3. SARS-CoV-2 life cycle. Spike glycoprotein bind to ACE2 receptor to assist virus entrance. Subsequent to the fusion of viral and plasma membranes, virus RNA undergoes replication and transcription. After the production of SARS-CoV-2 structural proteins, nucleocapsids are assembled in the cytoplasm and, followed by budding into the lumen of the ERGIC. New virions are released via vesicles.

E: Envelope protein; ER: Endoplasmic reticulum; ERGIC: Endoplasmic reticulum–Golgi intermediate compartment; M: Membrane protein; N: Nucleocapsid; S: Spike.

with a β-sheet(s) consisting of six strands [41]. The high levels of genetic identity (96.3%) between the SARS-CoV-2 and Bat-CoV RaTG13 does not indicate the precise variant that may have led to the outbreak in humans, although it has been suggested that the likelihood that the novel CoV has derived from bats is very probable [43]. SARS-CoV-2 and RaTG13 differ with respect to the number of major genomic properties, in which SARS-CoV-2 harbors a polybasic (furin) cleavage site insertion at the connection of the two subunits of the S-protein, S1 and S2 [44].

The N-protein is hidden within phospholipid bilayers and coated via two distinctive forms of S-proteins including the spike glycoprotein trimer which is present in all CoVs, as well as the hemagglutinin-esterase (HE) solely found in certain CoVs. For instance, SARS-CoV-2 does not appear to have the HE gene. The M and E proteins are found inside the S-glycoproteins within the viral envelope [45]. The S, E, M, N and ORF3a genes of SARS-CoV-2 are predicted to be 3822, 228, 669, 1260 and 828 nucleotides in length, respectively. Moreover, SARS-CoV-2 has been predicted to contain an ORF8 gene, of 366 nucleotide size, situated between the M and N corresponding ORF genes [45].

The SARS-CoV sequence reveals serine substituting for glycine in the residue at position 543 of the nsp3 protein in Bat SARS-like and SARS-CoV. This AA substitution could promote local stiffness of the polypeptide chain.
for steric impact and potency of the serine side-chain to constitute H-bonds. Beside, serine is a nucleophile that can establish structural environments, like those at active sites of enzyme. Mutations in the nsp3 protein were reported to affect the replication of SARS-CoV-2 in infected cells [46,47]. It has been reported that the single N501T variant in SARS-CoV-2’s S-protein may enhance binding affinity for the ACE2 cellular receptor [28]. Furthermore, a single N439R mutation in SARS-CoV-2 RBD promotes its ACE2-receptor binding and, thus potentially enhances human-to-human transmission [48,49]. By studying the crystal structure of SARS-CoV-2 RBD binding to the human ACE2 receptor has shown that the ACE2 receptor-binding ridge in SARS-CoV-2 RBD results in a more compact conformation, leading structural alterations at the RBD/ACE2 interface versus the SARS-CoV [48]. Overall, SARS-CoV-2 binding affinity for ACE2 is 10–20-times greater than for other SARS-associated CoVs [50].

A missense mutation at the 614 position of S protein (aspartate to glycine, D614G mutation), in the spike protein of SARS-CoV-2, which has emerged as a predominant clade in Europe (66% sequences) and is spreading worldwide (44% sequences). The D614G mutation promotes viral infectivity and transduction of multiple human cell types and mitigates neutralization sensitivity to individual convalescent sera [51–54].

**SARS-CoV-2 pathogenesis**

Lipids play important roles at different stages in the CoVs life cycle. CoVs recruit intracellular membranes of the host cells to produce new compartments, or double membrane vesicles, which are used for the replication of the virion particle genome [55]. Recently, an important lipid processing enzyme, known as cPLA2 α has been reported to be related to the formation of double membrane vesicle and CoV’s amplification [56]. It has been demonstrated that the enzyme, phospholipase A2 group IID, is involved in anti-inflammation or proresolving lipid mediator regulation which may lead to worse outcomes in a SARS-CoV infection animal model by modulating the immune response [57].

It has been shown that there is a distinct insert that includes basic AAs in the S1/S2 priming loop of SARS-CoV-2, which is not found in SARS-CoV or any SARS-associated CoVs. It may substantially alter the entry pathway of SARS-CoV-2 compared with other viruses of the β-CoVs lineage B [58]. In a recent report it was shown that SARS-CoV-2’s S-protein entry into 293/human ACE2 receptor cells is primarily mediated via endocytosis, and that PIKfyve, a TPC2 and cathepsin L are crucial for virus entry. PIKfyve is the key enzyme in the early endosome involved in the synthesis of PI(3,5)P2 and its main downstream effector, TPC2. The S protein of SARS-CoV-2 could also stimulate syncytia in 293/human ACE2 cells independently of exogenous protease [59].

In a study of 452 SARS-CoV-2 infected patients, it was found that severely affected cases had lower numbers of blood lymphocytes, percentages of monocytes, basophils and eosinophils as well as increased leukocytes numbers and neutrophil-lymphocyte-ratio. In most patients with unfavorable progression of COVID-19, elevated concentrations of infection-associated markers and inflammatory cytokines was observed. The frequency of T cells was significantly lower, and less effective in severely affected subjects. Both T helper (Th) cells and suppressor T cell numbers in patients with COVID-19 were below the reference range. The percentage of naive helper T cells was increased, and memory helper T cells and regulatory T cells reduced in severe conditions [60]. Furthermore, simultaneous to the infection with SARS-CoV-2, CD4+ T lymphocytes are quickly over-activated to switch to the pathogenic Th1 cells producing GM-CSF. The cytokines environment activates inflammatory CD14+CD16+ monocytes, leading to over-expression of IL-6 and enhances the inflammatory response. Regarding the increased infiltrations of inflammatory cells that have been found in lungs of severe SARS-CoV-2 infected patients [61,62], these population of abnormal and noneffective pathogenic Th1 cells and inflammatory granulocytes may go to the pulmonary circulation and by immune stimulation, lead to functional impairment of the lungs and eventually death [63].

Inflammasomes are very large intracellular poly-protein signaling complexes which are constitute in the cytosol as an inflammatory immune reaction to endogenous danger stimuli [64]. NLRP3 responds to wide spectra of pathogen and endogenous signals, and is involved in the molecular pathway of various auto-inflammatory disorders [65]. It has been reported that the SARS-CoV can induce the NLRP3 inflammasome in macrophages through ORF8b. Whereas SARS-CoV infects macrophages or monocytes, sufficient ORF8b may be present to impact on the autophagy-lysosome pathway, and NLRP3 inflammasomes. SARS-CoV replicates efficiently in lung epithelial cells. These cells also amplify NLRP3 and support assembly of NLRP3 inflammasomes. In SARS-CoV patients, the full effect of the ORF-8b on these inflammatory cascades was observed in the lung epithelium. Interestingly, ORF8b may be involved in the ‘cytokine storm’ or ‘cytokine cascade’ and inflammasome induction which happens within intensive SARS-CoV infection [66].
SARS-CoV-2 infection stimulates the immune response in two stages. In the early stages, a particular adaptive immune response is necessary to eradicate the virus and to impede progress to a more severe condition. The protective immune response at this phase requires that the host should have excellent general health and a suitable genetic context which provides antiviral immunity [67]. Although, when the immune response protection is disabling, virus will disseminate and great damage to the affected tissues occurs, particularly in organs with a high levels of ACE2 receptor expression. The injured cells activate innate inflammation within the lungs which is mainly mediated through pro-inflammatory macrophages/monocytes. Lung inflammation is the major reason for the fatal respiratory disease at the severe stage of COVID-19 [68].

In viral infections, host antiviral microRNAs participate in the regulation of immune response to virus and are capable of targeting viral genes and interfere with replication, mRNA expression and protein translation of virion particle gene. Sardar et al. predicted the antiviral host-microRNAs specifically for COVID-19. They reported a list of six microRNAs related to COVID-19 including hsa-let-7a, hsa-miR101, hsa-miR126, hsa-miR23b, hsa-miR378 and hsa-miR98 which has been previously reported to be related to other viral infections, such as HIV [69].

Clinical presentation
Virion particles spread from the respiratory mucosa, by binding to the ACE2 receptors on ciliated bronchial epithelial cells, and after that may engage with other cells [70]. In one report from Wuhan, the average incubation period of 425 SARS-CoV-2 infected patients was 5.2 days, but it this differed between individuals [71,72]. Until now, most patients with COVID-19 have initially presented with mild manifestations in other words dry cough, sore throat and fever which spontaneously resolve. Although, some patients have developed other more severe disease such as organ failure, septic shock, pulmonary edema, dyspnea, myalgia, fatigue and acute respiratory distress syndrome [73]. In contrast to SARS-CoV, patients infected with SARS-CoV-2, development of upper respiratory tract signs and manifestations are less common, suggesting that SARS-CoV-2 may target cells in the lower airway [74]. Among cases with severe dyspnea, more than 50% have required intensive care. Some COVID-19 cases do not present with fever or radiologic abnormalities on admission, which makes initial diagnosis difficult [75].

The main characteristics of COVID-19 on preliminary CT examination including bilateral multi-lobar ground-glass opacities with a peripheral/posterior distribution and patchy consolidation, primarily in the lower lobes and fewer inside the right middle lobe [76]. The main reported laboratory test abnormalities in cases with severe COVID-19 infection include: increased levels of liver enzymes (LDH, ALT and AST), total bilirubin, creatinine, cardiac troponin, D-dimer, prothrombin time, procalcitonin and CRP [77]. The histology of liver specimens of SARS-CoV infected patients have revealed a remarkable liver injury with an increase in mitotic cells, along with eosinophilic bodies as well as balloon-like hepatocytes [78]. Cardiac involvement is another prominent manifestation of COVID-19 and is closely related to a poor outcome [79]. In a recent systematic review, the incidence rate of diarrhea varied from 2 to 50% in COVID-19 patients. It may develop earlier, or following the respiratory symptoms. Findings of several studies showed that viral RNA shedding is detect for a longer time period compared with nasopharyngeal swabs [80].

In an investigation on 1099 COVID-19 patients, of whom 23.7% had severe disease with comorbidities of hypertension, 16.2% diabetes mellitus, 5.8% coronary heart diseases and 2.3% cerebrovascular disease [80]. Another study, of 140 patients with COVID-19, found that 30% and 12% had history of hypertension and diabetes, respectively [81]. Analysis of 487 COVID-19 cases, showed that older age (odds ratio [OR] = 1.06; 95% CI: 1.03–1.1), male gender (OR 3.7; 95% CI: 1.7–7.7) and hypertension as a comorbidity (OR 2.7; 95% CI: 1.3–5.6) are related with more severe disease on admission [82]. Moreover, patients with cancer were more vulnerable to severe events from COVID-19 such as admission to the intensive care unit needing invasive ventilation, or death [83].

It has been reported that the highest viral load in throat swabs occurs at the time of development of symptoms. However, viral shedding was reported to occur before the onset of symptoms, and a major proportion of transmissibility happened before first symptoms in the index case [84]. Furthermore, severe COVID-19 cases tend to have an increased viral load and a long virus-shedding time [85].

Diagnosis
At present, the diagnosis of COVID-19 is largely based on guideline agreement that includes laboratory tests and chest CT imaging technique [75,86]. PCR testing of asymptomatic or mild symptomatic contacts can be used in the
Coronaviruses & health Review

evaluation of peoples who have been in contact with a COVID-19 case [87], and the WHO has not accepted the results of a chest CT without RT-PCR conformation in the diagnosis of COVID-19 [88].

Chest CT is a routine imaging tool for the diagnosis of pneumonia, which is relatively easy and rapid to perform. Chest CT shows typical radiographic characteristics in almost all COVID-19 cases, such as peripheral/posterior distribution and patchy consolidation, and/or interstitial alterations with a peripheral distribution, so may provide benefit for diagnosis of COVID-19 [89].

Respiratory tract samples were collected for the diagnosis and screening of patients with SARS-CoV-2 pneumonia; in the 5–6 days of the initiation of symptoms, patients with COVID-19 have increased viral loads in their upper and lower respiratory tracts [90,91]. For suspected cases, real-time fluorescence (RT-PCR) was performed to detect the positive nucleic acid of SARS-CoV-2 in sputum, throat swabs and secretions of the lower respiratory tract specimens [92]. A nasopharyngeal and/or an oropharyngeal swab are frequently recommended for screening or diagnosis of early infection [10,93]. A single nasopharyngeal swab has become the preferable swab as it is well-tolerated by the patient and safer for the operator.

Serological testing detects presence of IgG, IgM or both. A positive elucidation has been defined as a positive IgM, or convalescent sera with a higher IgG titer > four-times in comparison with the acute phase. SARS-CoV-2 IgG and IgM are detected in whole blood, plasma, serum or specimens. Antibodies increase late in the course of illness; the mean duration of SARS-CoV-2 IgM antibody detection was reported to be 5 days, whereas IgG detection about 2 week following the appearance of symptoms [94]. In contrast to respiratory samples which may disturb from false-negative results because of the sampling factors, the presence of antibodies in blood uniformly is detectable. Specimens are easier to gather versus respiratory samples, such as fewer risks to the operator. The serological assay is very easy, rapid, availability of ELISA platforms, requires no instrumentation and can provide results in just 15 min [95].

Treatment

Based on the recommendation of WHO, COVID-19 management protocols have mostly highlighted infection prevention, patient early detection and monitoring, and best supportive care [96,97]. No specific antiviral treatment is currently recommended for COVID-19 due to lack of evidence. Many treatment regimens have been assessed for COVID-19, some showing promising preliminary results. A total of 2531 trials on COVID-19 have been registered to date in the clinicaltrial.gov (updated 10 July 2020). Several pharmacotherapeutic agents have been used including lopinavir/ritonavir, hydroxychloroquine and IFN-β-1A (Table 1).

Chloroquine

Results from several in vitro and clinical studies demonstrated that chloroquine phosphate, an old agent for the treatment of malaria, had significant efficacy and acceptable safety for treatment of COVID-19 [98,99]. Findings of an open-label nonrandomized clinical trial among 22 infected patients indicated that hydroxychloroquine treatment significantly reduced viral load in COVID-19 cases and its effectiveness is promoted by azithromycin [99]. In a systematic review including six published articles highlighting the potency of chloroquine in attenuation the replication of SARS-CoV-2-associated virus [100]. But several other studies demonstrated no evidence of a strong antiviral function, or clinical benefit of the hydroxychloroquine for the treatment of patients with severe COVID-19 [101,102].

Lopinavir–ritonavir

The combination of lopinavir/ritonavir (LPV–R) is extensively used for treating HIV-infected patients. LPV–R has been suggested for treatment of COVID-19. A total of 199 COVID-19 patients were randomly assigned to receive LPV–R (n = 99) or standard-care (n = 100). Treatment with LPV–R was not different from standard care regarding the time to clinical improvement, mortality rate at 28 days, as well as detection of viral RNA at different time points [103].

Arbidol

Arbidol as a wide-spectrum antiviral compound that can inhibit viral fusion of influenza. In one study, 50 patients with laboratory-confirmed SARS-CoV2 were randomly divided into two arms: 34 cases received LPV–R (400 mg/100 mg, two per day) and 16 patients were administrated arbidol (0.2 g a; three per day). No difference was observed concerning fever duration between the two arms. 2 weeks after the intervention, no viral load was
Table 1. Clinical trials for the treatment of coronavirus disease 2019 (updated at 10 July 2020).

| Intervention | Phase | Status | References |
|--------------|-------|--------|------------|
| Lopinavir/ritonavir | II | Recruiting | NCT04330690 |
| Lopinavir/ritonavir | II | Terminated | NCT04307693 |
| Hydroxychloroquine sulfate | IV | Enrolling by invitation | NCT04350671 |
| Interferon-α 1a | IV | Enrolling by invitation | NCT04350684 |
| Lopinavir/ritonavir | II | Recruiting | NCT04321993 |
| Hydroxychloroquine sulfate | III | Recruiting | NCT04355052 |
| Baricitinib (Janus kinase inhibitor) | II | Recruiting | NCT04329832 |
| Sarilumab (anti-IL-6 receptor) | II | Recruiting | NCT04347915 |
| Hydroxychloroquine | III | Recruiting | NCT04341727 |
| Hydroxychloroquine + camostat mesylate | III | Recruiting | NCT04355052 |
| Hydroxychloroquine + azithromycin | II | Recruiting | NCT04359611 |
| Clevudine | II | Recruiting | NCT043299832 |
| Hydroxychloroquine | III | Recruiting | NCT04341727 |
| Hydroxychloroquine | III | Recruiting | NCT04336328 |
| Azithromycin | III | Not yet recruiting | NCT043438906 |
| Camostat mesilate | III | Recruiting | NCT04336904 |
| Placebo | III | Recruiting | NCT04330638 |
| Hydroxychloroquine | III | Recruiting | NCT04351620 |
| Azithromycin | III | Recruiting | NCT04329611 |
| Camostat mesilate | IV | Not yet recruiting | NCT04338906 |
| Placebo | III | Active, not recruiting | NCT04336904 |
| Usual care | III | Recruiting | NCT04330638 |
| Anakinra | III | Recruiting | NCT04317092 |
| Siltuximab | III | Recruiting | NCT04340232 |
| Tocilizumab | III | Recruiting | NCT04348071 |
| Tocilizumab injection | III | Recruiting | NCT04345445 |
| Baricitinib | II/III | Not yet recruiting | NCT04350455 |
| Ruxolitinib | II/III | Not yet recruiting | NCT04350455 |
| Tocilizumab | III | Not yet recruiting | NCT04350455 |
| Methylprednisolone | III | Not yet recruiting | NCT04350455 |
| Sildenafil citrate tablets | III | Recruiting | NCT0433550 |
| Deferoxamine | I/II | Recruiting | NCT0433550 |
| Naproxen | III | Not yet recruiting | NCT04325633 |
| Standard of care | III | Not yet recruiting | NCT04323332 |
| Traditional Chinese medicine prescription | III | Not yet recruiting | NCT043251871 |
| Conventional medicines + traditional Chinese medicines granules | Not applicable | Recruiting | NCT043251871 |

1 From Clinicaltrial.gov.
MSC: Mesenchymal stem cell; NK: Natural killer.
Table 1. Clinical trials for the treatment of coronavirus disease 2019 (updated at 10 July 2020) (cont.).

| Intervention | Phase | Status | References |
|--------------|-------|--------|------------|
| Dietary supplement: ascorbic acid | Not applicable | Enrolling by invitation | NCT04342728 |
| Dietary supplement: zinc gluconate | | | |
| Dietary supplement: ascorbic acid and zinc gluconate | | | |
| Other: standard of care | | | |
| Losartan | IV | Recruiting | NCT04340557 |
| Captopril or enalapril | III | Not yet recruiting | NCT04345406 |
| chloroquine | | | |
| Hyperimmune plasma | Not applicable | Completed | NCT04321421 |
| Blood donation from convalescent donor | | Recruiting | NCT04351659 |
| Convalescent plasma | II | Recruiting | NCT04343755 |
| Convalescent plasma | II | Withdrawn (due to opening expanded access protocol) | NCT04325672 |
| rhACE2 | Not applicable | Withdrawn (without CDE approval) | NCT04287686 |
| Allogeneic human dental pulp stem cells | I/II | Recruiting | NCT04336254 |
| Placebo | | | |
| Allogeneic adipose-derived mesenchymal stromal cells | I/II | Withdrawn | NCT04341610 |
| Placebo | | | |
| Biological: MSCs | I | Recruiting | NCT04252118 |
| Biological: NK cells, IL15-NK cells, NKG2D CAR-NK cells, ACE2 CAR-NK cells, NKG2D-ACE2 CAR-NK cells | I/II | Recruiting | NCT04324996 |
| Biological: NK cells | I | Recruiting | NCT04280224 |
| Biological: allogeneic NK transfer | I/II | Not yet recruiting | NCT04344548 |

†From Clinicaltrial.gov.
MSC: Mesenchymal stem cell; NK: Natural killer.

found in cases received arbidol, while the viral load was detectable in 44.1% of LPV–R group patients. Moreover, no adverse side effects were reported in either arm [104].

Nelfinavir
Nelfinavir (NFV) is a potent HIV-1 protease inhibitor that received US FDA approval in 1997 for treatment of HIV infection. The antiviral activity of NFV against SARS-CoV-2 was reported in Vero E6 cells [105]. By using an integrative computational drug-discovery method, NFV was introduced as a potential inhibitor of SARS-CoV-2 main protease [106].

Main protease inhibitor
The main protease of CoVs (Mpro) is an important protein necessary for the proteolytic maturation of the virion particle [107]. Therefore, targeting Mpro is considered to have potential as a treatment for COVID-19 through suppression of the polypeptide cleavage virus [108–110]. Concerning the results of molecular docking, natural polyphenols such as hesperidin, rutin, diosmin, apiin and diacetyl-curcumin have been reported to have acceptable efficacy to target SARS-CoV-2 Mpro than NFV [111].

Cytokine antagonists
Cytokine-directed antagonists, in other words adalimumab (TNF-α) and CMAB806 (IL-6) against SARS-CoV-2 have been evaluated in clinical trials. The variety of cytokines such as type-I IFN-I contribute to the ‘cytokine storm’ and pathology of SARS-CoV-2. Therefore, targeting the upstream origin of cytokine generation could be a promising therapeutic approach [112].

Nucleoside analog
Utilizing an in silico model, it has been shown that antipolymerase agents including sofosbuvir, IDX-184, ribavirin (RBV) and remidivir (GS-5734; RDV) can target RNA-dependent RNA polymerase of SARS-CoV-2 [113]. The first severe-infected patient with SARS-CoV-2 in the USA was cured by reception of intravenous RDV [114]. Due to adverse side effects, the appropriate dose of RBV in clinical setting should be given with caution.
Convalescent plasma
In previous experience, for example in pandemic influenza A (H1N1), and avian influenza A (H5N1), passive immunization has been successful for treating of infectious complications [115]. A remarkable reduction in viral load and mortality was observed by using convalescent plasma therapy against severe acute viral respiratory infections, such as those created by CoVs [116]. Patients who have recovered from SARS-CoVs infection often have high titers of neutralizing antibody and may be a precious source of convalescent plasma. The FDA has also approved the administration of plasma from recovered individuals for treatment of severe COVID-19 patients [117].

Monoclonal antibodies
SARS-CoV-specific human monoclonal antibody (mAb) can bind potently with SARS-CoV-2 region. But, some of the most powerful SARS-CoV-particular neutralizing antibodies (i.e., m396) that target the ACE2 binding site of SARS-CoV did not bind to SARS-CoV-2 S-protein, indicating that the disparity in the RBD of SARS-CoV and SARS-CoV-2 has an important effect impact on the cross-reactivity of these mAbs, and so novel mAbs that specifically target SARS-CoV-2 RBD need to be designed [112].

Vaccine
Effective SARS-CoV-2 vaccines are urgently needed in order to reduce infection severity, viral shedding as well as human–human transmission, so assisting the control of the CoV outbreaks. Because S-protein and associated fragments, in other words RBD of SARS- and MERS-CoVs are the main targets for designing vaccines, it is speculated that homologous regions of SARS-CoV-2 can also be applied as prime targets for designing vaccines against this novel CoVs [118]. In addition, other conserved regions of SARS-CoV-2 including two subunits of the S-protein, M-protein as well as N-protein, can be applied as another potential target for design and development of effective vaccines.

Antiviral vaccines can be categorized into two broad groups: DNA- and RNA-based vaccines, in which individuals are injected with genetically engineered plasmid containing the DNA molecule encoding the antigen against which an immune response is eligible, thus the cells machinery creates the antigen, leading to immunological response; and peptide- or protein-based vaccines that include whole-inactivated virus, individual viral proteins or subdomains, and purified or recombinant proteinaceous antigens proteins from the virus, all of which are manufactured in vitro.

The candidate vaccines that have recently entered clinical development include: mRNA-1273, Ad5-nCoV, INO-4800 and LV-SMENP-DC and pathogen-specific aAPC (Table 2). Several platforms have progressed to development with potential for rapid development, including DNA- and RNA-based platforms, followed by those for developing recombinant-subunit vaccines. RNA and DNA vaccines can be made quickly because they do not require culture or fermentation, instead using synthetic processes [119,120].

Even with such promising platforms, SARS-CoV-2 vaccine development faces serious challenges. Although the virus's S glycoprotein is a promising immunogen for protection, optimization of antigen design is crucial to obtaining an optimum host immune system response. Another concern is the possible exacerbation of lung disease, either directly or because of antibody-dependent enhancement due to the type 2 helper T-cell response. Furthermore, as with naturally acquired infection, the optimal duration of immunity is unknown; similarly, whether single-dose vaccines will confer lengthy immunity is doubtful.

Conclusion
In the early phases of the epidemic, early detection assists management of the disease and preventive approaches such as masks, hand hygiene compliances, prevention of public contact, voluntary home quarantine, early diagnosis, contact tracing, intelligence social distance and travel restrictions have been recommended to decrease transmission. Other approaches include limiting events that may facilitate superspreader potential including religious services (marriages and funerals) [121]. Many dimensions of the SARS-CoV-2 and corresponding disease are unknown. For instance, the role of ACE2 receptors in SARS-CoV-2 pathogenesis remains uncertain. Future studies should be concentrate on profound understating of replication, pathogenesis and biological properties applying the relevant biological methods in other words reverse genetics and molecular techniques. Genome wide association studies may provide an opportunity for the identification of potential genetic factors contributed in the development of COVID-19. Although host genetic studies are expensive and complex, more studies are required to determine the role of host genetics (such as variation in HLA genes) in the immune response to CoVs, and the clinical outcome of CoVs-mediated disease. Understanding of the SAR-CoV-2 viral genetics during the time and geography specially
Table 2. Vaccine in clinical trials for the treatment of coronavirus disease 2019 (updated 5 July 2020).

| Intervention | Phase | Status | References       |
|--------------|-------|--------|------------------|
| Pathogen-specific aAPC | I     | Recruiting | NCT04299724     |
| LV-SMENP-DC vaccine and antigen-specific CTLs | I/II | Recruiting | NCT04276896     |
| - GX-19 | I/II | Recruiting | NCT04445389     |
| - Vaccine BCG | III | Not yet recruiting | NCT04362124     |
| - Vaccine (mRNA-1273) | I     | Recruiting | NCT04283461     |
| - Novel coronavirus vaccine (adenovirus Type 5 vector) | I     | Active, not recruiting | NCT04313127     |
| - SCB-2019 | I     | Recruiting | NCT04405908     |
| - Gam-COVID-Vac | I/II | Recruiting | NCT04436471     |
| - Gam-COVID-Vac Lyo | I/II | Recruiting | NCT04437875     |
| - Vaccine (adenovirus type 5 vector) | II    | Active, not recruiting | NCT04341389     |
| - BNT162a1 | I     | Recruiting | NCT04368728     |
| - SARS-CoV-2 rS | I     | Recruiting | NCT04368988     |
| - AV-COVID-19 | I/II | Not yet recruiting | NCT04386252     |
| - Covax-19™ | I     | Not yet recruiting | NCT04428073     |
| - Vaccine ChAdOx1 nCoV-19 | II/III | Not yet recruiting | NCT04400838     |
| - bacTRL-spike vaccine | I     | Not yet recruiting | NCT04334980     |
| - SARS-CoV-2 rS | I     | Recruiting | NCT04368988     |
| - IMM-101 | III   | Not yet recruiting | NCT04442048     |
| - CvnCoV vaccine | I     | Recruiting | NCT04449276     |
| - ChAdOx1 nCoV-19 | I/II | Not yet recruiting | NCT04444674     |
| - VPM1002 | III   | Not yet recruiting | NCT04439045     |
| - INO-4800 | I/II | Not yet recruiting | NCT04447781     |
| - INO-4800 | I     | Recruiting | NCT04336410     |
| - BNT162a1 | I/II | Recruiting | NCT04380701     |
| - VPM1002 | III   | Recruiting | NCT04387409     |

aAPC: Artificial antigen-presenting cell; BCG: Bacille Calmette-Guérin or tuberculosis vaccine; CTL: Cytotoxic T lymphocyte.

regarding to the number and repetition of viral mutations and recombination rates and their association with viral infectivity, transmissibility, severity of disease and clinical manifestation, viral load and disease outcome are important knowledge gaps that navigate our research timetable. Until now, no unique antiviral therapy has been approved; so treatment is mainly based on symptomatic therapy and best supportive care.

The zoonotic link of SARS-CoV-2 infection has not been definitively proven; although, phylogenetic analysis shows that SARS-CoV-2 is very similar to SARS-like bat CoVs. Lessons from other human outbreaks from pathogenic viruses such as SARS-CoV, MERS-CoV and influenza viruses are very informative and valuable. Different wide-spectra antivirals agents previously used for treatment of influenza, SARS- and MERS-CoVs are under assessment for repurposing either monotherapy or in combinations to treat COVID-19 cases. SARS-CoV-2 is a novel human pathogen, and may interact with host antiviral defense via a specific pathway. Altogether, the infection and development of SARS-CoV-2 relies on the interplay between the virus and the patient’s immune response. Investigations of the area of SARS-CoV-2-host interplay provide response to many crucial questions in virus pathogenesis, disease control and prevention. At present, COVID-19 is leading to substantial global concerns. Development of valid, accurate and appropriate serological tests is urgently needed. It will be essential to quickly design and develop effective therapeutic regimen and vaccines to prevent or stop infection of this novel CoVs.

Future perspective

The COVID-19 has caused more infections and deaths compared with either SARS or MERS. According to \( R_0 \) values, it is deemed that SARS-CoV-2 is more infectious than SARS or MERS. As imposition of globalization, CoVs will cause spreads and outbreaks with various mutant strains similarly in the coming years. With promotion of scientific collaboration, which is as a consequence of globalization, we may have more powerful means of combating CoVs, in which we characterize the genome structure and pathogenesis of SARS-CoV-2 infection very well in the near future. A present treatment is mainly supportive, but trials of vaccines and antivirals are in progress. Differences
in the length of the spike as it is longer in SARS-CoV-2 are likely to play a major role in the pathogenesis and treatment of this virus.

Robust coordination and collaboration between researchers, vaccine developers, international regulators, policymakers, financiers, national public health institutes and governments will be required to ensure that potential late-stage vaccine candidates can be produced in adequate amount with high safety and efficacy as well as equitably provided to all affected areas, specially low-resource regions.

Executive summary
The severe acute respiratory syndrome coronavirus 2 genome & structure
• All the coronaviruses are positive-stranded RNA viruses with a polycistronic genome with 6–11 open reading frames, encoding several nonstructural proteins at the 5′-end plus four structural proteins (spike surface glycoprotein [S], envelope [E], matrix [M] and nucleocapsid [N]) and multiple lineage-specific accessory proteins at the 3′-end.
• Phylogenetic analysis demonstrates that severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) shares 50 and 80.0% nucleotide identity to Middle East respiratory syndrome CoV and SARS-CoV, respectively.
• The S-protein of SARS-CoV-2 is longer than for other viruses such as SARS-CoV and Bat SARS-like CoVs.

SARS-CoV-2 pathogenesis
• Virion particles spread from the respiratory mucosa, by binding to the ACE2 receptors on ciliated bronchial epithelial cells, and after that may engage with other cells.
• The S-glycoprotein mediates binding of the virus to the sensitive human cell surface receptors, followed by fusion of the virus and host cell membranes to assist viral entrance.
• SARS-CoV-2 infection stimulates the immune response via two stages. At the incubation and nonalarming stages, a particular adaptive immune response is needed to eradicate the virus and to impede progress to severe condition.
• The injured cells activate innate inflammation within the lungs, which is mainly mediated through pro-inflammatory macrophages/monocytes. Lung inflammation is the major reason for the fatal respiratory disease at the severe stage of coronavirus disease 2019 (COVID-19) infection.
• The main characteristics of COVID-19 on preliminary computed tomography (CT) examination including bilateral multi-lobar ground-glass opacities with a peripheral/posterior distribution and patchy consolidation.

Diagnosis & treatment of COVID-19
• At present, the diagnosis of COVID-19 is largely based on laboratory tests PCR and chest CT imaging technique. Although, no specific antiviral treatment for COVID-19 is currently advised due to lack of evidence.
• Several pharmacotherapeutic agents have been used for treatment of COVID-19 patients consisting lopinavir/ritonavir, hydroxychloroquine and IFN β-1A.
• Effective SARS-CoV-2 vaccines are urgently needed in order to decrease infection severity, viral shedding as well as human–human transmission. The most advanced candidates have recently moved into clinical development, including mRNA-1273, Ad5-nCoV, INO-4800 and LV-SMENP-DC, and pathogen-specific.

Author contributions
All authors contributed to data collection, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Financial & competing interests disclosure
This study was supported by Birjand University of Medical Sciences. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

No writing assistance was utilized in the production of this manuscript.

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