A Multiscale View of the Mechanisms Underlying Ketamine’s Antidepressant Effects: An Update on Neuronal Calcium Signaling

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INTRODUCTION

Major depressive disorder (MDD) is the leading cause of disability worldwide. Despite considerable research, biological mechanisms underlying MDD pathophysiology remain unclear, with significant unmet needs for treatment. Typical antidepressants, including selective serotonin reuptake inhibitors (SSRIs) and serotonin and noradrenaline reuptake inhibitors, increase
monoamine concentration in the synaptic cleft, resulting in antidepressant effects (Berton and Nestler, 2006). However, although increased monoamine concentration in the synapse occurs relatively quickly as an acute pharmacological action, recovery from depression takes several weeks to months in clinical practice (Krishnan and Nestler, 2008). Electroconvulsive therapy (ECT) is also an effective treatment for drug-resistant depression, although achieving clinically meaningful or sustained remission with ECT required at least 1 month (Yamasaki et al., 2020). Such substantial time lags are a major concern since patients with depression are at high risk for suicide. Thus, there is an urgent need to develop antidepressants with rapid onset and sustained effectiveness.

Ketamine, a non-competitive glutamate N-methyl-D-aspartate receptor (NMDAR) antagonist, has gained considerable interest in the neuropsychiatric field. A single administration of ketamine elicits rapid and sustained antidepressant effects for 1–2 weeks in both humans and animals (Berman et al., 2000; Zarate et al., 2006; Li et al., 2010; Autry et al., 2011). This discovery offered new insight into the investigation of a whole new class of agents beyond the monoamine system to treat depression (Chaki, 2017). *Esketamine*, an enantiomer of (R,S)-ketamine, has been approved by the U.S. Food and Drug Administration (USFDA) for treating patients with treatment-resistant depression. Thus, research on pathophysiology and drug discovery for MDD has transitioned from the monoaminergic to the glutamatergic system. Recently, the importance of multiscale neuroscience to study cross-scale interactions at genetic, molecular, cellular, and macroscale levels of brain circuitry, connectivity, and behavior has been emphasized to establish a comprehensive understanding of neuropsychiatric disease (Van Den Heuvel et al., 2019). This mini-review aims to update the current knowledge regarding ketamine effect on the brain, focusing on the glutamatergic signaling pathway from a multiscale perspective at the behavioral, cellular, molecular, and epigenetic levels.

**THE GLUTAMATERGIC SYSTEM IN NEUROPLASTICITY, INTRACELLULAR SIGNALING, AND GENE EXPRESSION**

Glutamate is the major excitatory neurotransmitter in the brain, and increasing evidence indicates that dysfunction in glutamatergic signaling contributes to MDD pathophysiology (Popoli et al., 2011; Duman and Aghajanian, 2012; Thompson et al., 2015; Duman et al., 2019; Xia et al., 2021). The glutamatergic system is modulated by both ionotropic [NMDARs, α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPARs), and kainate receptors] and metabotropic glutamate receptors (mGluRs). NMDARs are found throughout the central nervous system and contribute to synaptic calcium (Ca\(^{2+}\)) influx, which is required for activity-dependent synaptic plasticity (Koester and Sakmann, 1998; Reid et al., 2001; Ngo-Anh et al., 2005; Bloodgood and Sabatini, 2007; Carter et al., 2007). NMDAR function is tightly linked to AMPAR, which gates sodium and mediates fast excitatory transmission. Increased AMPAR density in the postsynaptic membrane causes NMDAR-dependent long-term potentiation (LTP) (Huganir and Nicoll, 2013). AMPARs can also have several direct effects on synaptic transmission (i.e., LTP) and intracellular signals without the proper functioning of NMDARs. This NMDAR-independent and AMPAR-dependent intracellular signaling pathway is also hypothesized to underlie ketamine’s antidepressant actions (Zanos et al., 2016; Duman et al., 2019; Wei et al., 2021).

Ca\(^{2+}\) influx into the postsynaptic neuron stimulates a signaling-cascade, such as calcium/calmodulin-dependent kinases [CAMKs; e.g., calcium/calmodulin-dependent kinase II (CaMKIIs), eukaryotic elongation factor 2 (eEF2) kinase]. Brain-derived neurotrophic factor (BDNF) and its receptor, neurotrophic receptor tyrosine kinase 2 (TrkB), also plays a key role in synaptic plasticity (Minichiello, 2009). TrkB activation stimulates phospholipase Cγ1 (PLCγ1), which results in CaMK activation (Minichiello, 2009). Calcium-signaling activation further sends its signal toward downstream epigenetic and transcription modulators, such as MEF2, MeCP2, and HDAC5. These pathways modulate gene expression that affects dendritic growth, synaptic development, and neuronal plasticity (Greer and Greenberg, 2008; Graff and Tsai, 2013; Takemoto-Kimura et al., 2017; Uchida and Shumyatsky, 2018a,b; Figure 1). Taken together, calcium-signaling stimulation through NMDARs and/or AMPARs activates multiple downstream nucleocytoplasmic pathways; it induces activity-dependent epigenetic genetic expression, contributing to depression and antidepressant action.

Chronic stress initiates and exacerbates several psychiatric illnesses. Indeed, adverse stressful environments are associated with the pathophysiology of major psychiatric disorders, including mood and anxiety disorders (McEwen, 2007; Krishnan and Nestler, 2008; Duman and Aghajanian, 2012). There are several evidences demonstrating alterations in the expression and/or function of glutamatergic signaling and its downstream molecules (e.g., NMDARs, AMPARs, CaMKIIs, MEF2, MeCP2, and HDAC5), which is associated with plasticity and behaviors induced by chronic stress, traditional antidepressant drugs, and/or ketamine (Table 1). Moreover, molecular dysregulation associated with glutamatergic system is visible in postmortem brain tissues of patients with MDD (Table 1). Thus, such clinical and preclinical evidences suggest that calcium-signaling is a downstream target of the glutamatergic system in MDD pathophysiology and antidepressant effects.

**MECHANISMS OF KETAMINE’S ANTIDEPRESSANT EFFECTS: A MULTISCALE VIEW**

Less than one-third of patients with MDD achieve remission using traditional antidepressant pharmacotherapy (Trivedi et al., 2006). Treatment resistance occurs in up to 30% of patients with MDD (Fava, 2003). However, a single subanesthetic dose of ketamine produces a therapeutic response within a few hours that lasts for several days in patients with depression.

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Preclinical data indicate that ketamine’s enantiomer (et al., 1995; Chan et al., 2013; Giorgetti et al., 2015). Recent prepulse-inhibition deficits, cognitive deficits, and USFDA for treating depressive symptoms in adults with MDD respectively (Zarate et al., 2006). Additionally, Ketamine reduces suicidal ideation of treatment-resistant patients with MDD, respectively (Zarate et al., 2006). Intravenous infusion of ketamine results in clinical response and remission in 70 and 30% (Berman et al., 2000; Zarate et al., 2006). Ketamine is known to increase the phosphorylation of CaMKII, MeCP2, and promotes nuclear export and increases activity-dependent transcription. MEF2 recruits HDAC5 and removes transcriptional regulator, binds to methylated CpG sites on the genomic region downstream targets, including MeCP2, MEF2, and HDAC5. MeCP2, a transcriptional regulator, binds to methylated CpG sites on the genomic region and interacts with other transcription repressors, including HDACs. CaMKII phosphorylates MeCP2, promotes its nuclear export, and increases activity-dependent transcription. MEF2 recruits HDAC5 and removes activating acetyl groups from histones, which results in a silenced or repressed state of transcription. CaMKII phosphorylates HDAC5, which promotes nuclear export and increases activity-dependent transcription. Ketamine is known to increase the phosphorylation of CaMKII, MeCP2, and HDAC5 (see detail in the main text). Thus, ketamine-mediated enhancement of intracellular Ca2+ signaling is linked to epigenetic regulation of transcription, which leads to long-term synaptic plasticity and, consequently, prolonged antidepressant-like effects.

(Berman et al., 2000; Zarate et al., 2006). Intravenous infusion of ketamine results in clinical response and remission in 70 and 30% of treatment-resistant patients with MDD, respectively (Zarate et al., 2006). Additionally, Ketamine reduces suicidal ideation (Krystal et al., 2013). In 2020, esketamine was approved by the USFDA for treating depressive symptoms in adults with MDD having acute suicidal ideation or behavior.

Ketamine elicits robust unwanted side effects, including precipulse-inhibition deficits, cognitive deficits, and schizophrenia-like psychotic symptoms in humans (Lahti et al., 1995; Chan et al., 2013; Giorgetti et al., 2015). Recent preclinical data indicate that ketamine’s enantiomer (R)-ketamine (Hashimoto, 2019; Wei et al., 2021) and its metabolites (2R, 6R)-hydroxynorketamine (HNK) (Zanos et al., 2016) exert antidepressant effects with fewer adverse effects than do ketamine or (S)-ketamine. Since potential mechanisms underlying the rapid antidepressant actions of ketamine and its metabolites have been reviewed elsewhere (Fukumoto et al., 2017; Yang et al., 2018; Duman et al., 2019; Krystal et al., 2019; Sial et al., 2020; Highland et al., 2021; Shinohara et al., 2021; Wei et al., 2021; Xia et al., 2021), we review the recent progress in deciphering mechanisms underlying ketamine’s sustained antidepressant effects, with a particular focus on the role of calcium signaling from a multiscale perspective.

**Behavioral Effects of Ketamine**

Several animal studies have demonstrated antidepressant-like responses to ketamine. A single intraperitoneal injection of ketamine or its metabolites produces rapid (30 min–1 h) and long-lasting (24 h–7 days) antidepressant effects (Autry et al., 2011; Koike et al., 2011; Zhou et al., 2014; Sun et al., 2016; Zanos et al., 2016; Yang et al., 2018; Kim et al., 2021). Moreover, such ketamine antidepressant effects have been observed in not only naïve, non-stressed animals but also in animals subjected to adverse stressful life events. Animals exposed to chronic stress show despair-like behavior, anhedonia, anxiety, and/or social avoidance, whereas a single injection of ketamine or its metabolites rapidly reverses these deleterious effects and exerts long-term effects (Li et al., 2011; Zanos et al., 2016; Duman et al., 2019; Wei et al., 2021).

**Neurobiological Effects of Ketamine**

Neuroimaging studies have shown structural and functional alterations in the hippocampus and dorsomedial prefrontal cortex (dmPFC) of patients with MDD (Price and Drevets, 2010; Macqueen and Frodl, 2011). Human functional magnetic resonance imaging (fMRI) studies have demonstrated that a single dose of ketamine ameliorates reductions in functional connectivity in the prefrontal cortex (PFC), which is associated with the alleviation of depressive symptoms (Abdallah et al., 2017). Interestingly, a recent MRI study in animals demonstrated short- and long-term effects of ketamine on distinct brain circuitries (Gass et al., 2019). Interestingly, a recent fMRI study in animals demonstrated short- and long-term effects of ketamine on distinct brain circuitries. Gass et al. (2019) found in an animal model of depression that ketamine causes a rapid response in the amygdala, anterodorsal hippocampus, and ventral pallidum, which are related to cognitive, sensory, emotional, and reward functions. However, 48 h after administration, ketamine showed a long-term normalization of the habenula, midline thalamus, and hippocampal connectivity. They mediate cognitive flexibility for processing contextual information, distinguish contextual cues in safe versus threatening situations, and modulate fear and emotional responses in non-threatening environments (Gass et al., 2019).

There is increasing evidence suggesting altered neuronal and structural plasticity in animal models of depression as well as in patients with MDD (Duman and Aghajanian, 2012; Kang et al., 2012; Abe-Higuchi et al., 2016; Higuchi et al., 2016; Nie et al., 2018; Uchida et al., 2018; Sakai et al., 2021). Ketamine rapidly increases the number and function of spine synapses. Furthermore, Li et al. found that ketamine increases the number and function of spine synapses in the medial PFC (mPFC)
TABLE 1 | Example evidence indicates alterations in behavior, glutamatergic signaling, and its downstream pathways regarding depression, chronic stress, and antidepressants: translational and multiscale views.

### Behaviors

| Findings | References |
|----------|------------|
| Ketamine’s effects on a stress-induced animal model of depression | CUMS-induced increase of immobility in TST were reversed 0.5 and 72 h after ketamine treatment in rats | Sun et al., 2016 |
| | CUS-induced reduction in sucrose preference in SPT was reversed by ketamine 24 h after injection in rats | Li et al., 2011 |
| | CSDDS-induced reduction of social interaction was reversed 24 h after (2R, 6R)-HNK treatment in mice | Moda-Sava et al., 2019 |
| | Ketamine’s effects on pharmacological model of depression | Chronic CORT effects on immobility in TST, open-arm exploration in an elevated plus maze and sucrose preference were reversed 24h after ketamine treatment in mice | Moda-Sava et al., 2019 |
| Neuroplasticity | | |
| MDD patients | Postmortem brain of MDD patients showed a lower number of synapses in dlPFC | Kang et al., 2012 |
| | Meta-analysis of structural imaging studies demonstrated that MDD patients have smaller hippocampus volumes | Macqueen and Frodl, 2011 |
| Stress-induced animal model of depression | Meta-analysis of imaging showed the structural and functional decline in dmpFC of MDD patients | Price and Drevets, 2010 |
| | CUS decreases the number and function of spine synapses in the mPFC | Li et al., 2011 |
| | Reduced spine density in the hippocampus and mPFC of mice susceptible to CUMS and CSDDS | Abe-Higuchi et al., 2016; Higuchi et al., 2016; Nie et al., 2018; Sakai et al., 2021 |
| Ketamine’s effect | Repeated stress impairs glutamatergic transmission in PFC pyramidal neurons | Yuen et al., 2012 |
| | (S)-ketamine normalized habenula, midline thalamus, and hippocampal connectivity at 48 h in fMRI imaging of stressed rats | Gass et al., 2019 |
| | Ketamine blocks NMDAR spontaneous activity | Autry et al., 2011 |
| | Ketamine treatment restores lost spines by chronic CORT exposure and promote generating functional synapses in mice | Moda-Sava et al., 2019 |
| | Ketamine treatment increases the number and function of spine synapse in rat mPFC | Li et al., 2010 |
| | (2R,6R)-HNK increased fEPSC slope in SC-CA1 of rats | Zanos et al., 2016 |
| | (2S,6S)-HNK increased fEPSC slope in SC-CA1 of rats | Chen et al., 2012 |

### Molecular pathway/Intracellular signaling

| Molecules | Findings | References |
|-----------|----------|------------|
| NMDARs | A postmortem prefrontal cortex showed increased levels of NR1 in MDD | Rodríguez-Munoz et al., 2017 |
| | Reduced GluN2A in prefrontal cortex of MDD | Beneyto and Meador-Woodruff, 2008 |
| | MK801, a NMDAR antagonist, injection reduced immobility in FST | Autry et al., 2011 |
| | CUS-induced reduction in sucrose preference in SPT was reversed by a selective NR2B antagonist, Ro 25-6981, 24 h after injection in rats | Li et al., 2011 |
| | Ketamine treatment increases NR1 expression levels in mouse PFC | Liu et al., 2011 |
| | Ketamine and a high dose of (2R, 6R)-HNK influences NMDAR-mediated eEF2 phosphorylation | Autry et al., 2011; Suzuki et al., 2017 |
| | (2R, 6R)-HNK do not block NMDAR function | Lumsden et al., 2019 |
| AMPARs | MD and stress model | Beneyto et al., 2007 |
| | Postmortem cortical tissue from MDD patients showed decreased GluA1 levels | Sakai et al., 2021 |
| | Reduced GluA1 level in the hippocampus of stress-susceptible mice | AMPAR potentiator drives stress resilience, whereas GluA1 inhibition leads to stress susceptibility | Ketamine | Beurel et al., 2016 |
| | Ketamine increased the level of GluA1 subunit in the mouse hippocampus | Zanos et al., 2016 |
| | (2R, 6R)-HNK increased synaptic GluA1 and GluA2 protein expression in the mouse hippocampus | Dvivedi et al., 2003 |
| | BDNF and Tbr2 | Youssef et al., 2018 |
| BDNF/Tbr2 | MD and stress model | Beneyto et al., 2007 |
| | Postmortem brain tissues from the hippocampus and prefrontal cortex in suicide subjects showed reduced expression of BDNF and TrkB | Dvivedi et al., 2003 |
| | BDNF levels were lower in the anterior cingulate of postmortem brains of subjects with early life adversity and/or died by suicide | Youssef et al., 2018 |
and rapidly reverses synaptic abnormalities caused by chronic stress exposure (Li et al., 2010). Although this evidence suggests an association between ketamine-induced spinogenesis and antidepressant-like behavior, the causal relationship is unclear. However, a recent report by Moda-Sava et al. has addressed this issue. They used a photoactivatable proof to selectively reverse ketamine effects on spine formation in the PFC. They found that newly formed spines are necessary for and play a specific role in the sustained antidepressant-like behavior induced by ketamine treatment (Moda-Sava et al., 2019).

Ketamine-Induced Synaptic Plasticity

Brain-derived neurotrophic factor and its receptor TrkB play key roles in synaptic plasticity, stress, and depression (Duman and Monteggia, 2006; Minichiello, 2009; Castren and Monteggia, 2021). A recent report discovered that several antidepressants, including fluoxetine, imipramine, and ketamine, directly bind to TrkB, facilitating BDNF action and plasticity (Casarotto et al., 2021). In addition, increased BDNF-TrkB signaling in rodent frontocortical/hippocampal circuits has been observed following acute treatment with ketamine (Li et al., 2010; Autry et al., 2011). Clinical evidence suggests that repeated ketamine administration allows cumulative and sustained antidepressant effects and that it is more effective than a single injection in patients with MDD (Aan Het Rot et al., 2010; Murrough et al., 2013; Phillips et al., 2019). The threshold and sensitivity of the persistent increase and decrease of synaptic strength are subject to activity-dependent regulation. This type of plasticity, called “metaplasticity,” is important for stabilizing synaptic strength and preventing LTP saturation and long-term depression, leading to homeostatic alternations of synaptic activation (Bienenstock et al., 1982; Turrigiano et al., 1998;
Kavalali and Monteggia, 2020). Notably, a preclinical study suggested that ketamine administration elicits metaplastic effects on LTP modulation and potentially other processes for long term. Kim et al. (2021) reported that, by using slice recordings of the Schaffer collateral-CA1 pathway in the hippocampus, ketamine induces AMPAR-mediated synaptic potentiation. Interestingly, this effect was more than two-fold higher in brain slices of mice that had received ketamine 7 days earlier, suggesting a priming effect of ketamine treatment such that subsequent ketamine augments synaptic potentiation. Further experiments to understand the mechanisms of this metaplasticity will provide critical insight into mechanisms underlying ketamine’s potent and prolonged antidepressant effects.

**Ketamine-Induced Ca^{2+} Signaling Cascades**

N-methyl-D-aspartate receptors activate eEF2 via CaMKs (eEF2 kinases) and depress BDNF levels (Scheetz et al., 2000). Ketamine-induced suppression of postsynaptic NMDARs deactivates eEF2 kinase, leading to reduced eEF2 phosphorylation and increased translation of BDNF in the hippocampus (Autry et al., 2011; Suzuki and Monteggia, 2020). This signaling pathway then potentiates synaptic AMPAR responses through the insertion of GluA1/2 subunits (Autry et al., 2011). In contrast, ketamine’s metabolite (2R, 6R)-HNK has NMDAR inhibition-independent antidepressant actions (Zanos et al., 2016; Lumsden et al., 2019), whereas other reports have shown that NMDAR inhibition at a high dose of (2R, 6R)-HNK triggers intracellular signaling via eEF2 (Suzuki et al., 2017).

A transient burst of glutamate via NMDAR blockade on GABAergic interneurons by ketamine activates postsynaptic AMPARs in excitatory neurons. This activation induces depolarization and activation of NMDARs that trigger Ca^{2+} influx, releasing BDNF (Krystal et al., 2019). Local release of BDNF is thought to activate TrkB on the postsynaptic membrane, stimulating the ERK and PI3K-Akt signaling pathways and mammalian target of rapamycin complex 1 (mTORC1) phosphorylation to promote synapse formation by stimulating synaptic proteins, such as GluA1 and PSD-95, which are required for synaptic plasticity (Cavalleri et al., 2018). Recently, mTORC1 effectors 4E-BP2 and 4-EB2 in excitatory or inhibitory neurons underlie behavioral and neurobiological responses to ketamine (Aguilar-Valles et al., 2021). Ketamine-induced activation of TrkB increases GSK-3β phosphorylation via the ERK signaling pathway, decreasing PSD-95 phosphorylation and internalizing the AMPA GluA1 subunit, which upregulates signaling through the GluA1 to promote synapse formation (Liu et al., 2013; Beurel et al., 2016). Ketamine-dependent changes in dendritic arborization and soma size are abolished by AMPAR antagonists or mTOR complex/signaling inhibitors (Cavalleri et al., 2018). Intracellular molecular signaling cascades stimulated by the glutamatergic pathway may be associated with ketamine-induced structural and synaptic plasticity and its antidepressant effects.

As mentioned earlier, CaMKIIs are major downstream target for the glutamatergic pathway and might be involved in stress and depression. TrkB activation stimulates phospholipase Cγ1 (PLCγ1) and also results in the activation of CaMKs (Minichiello, 2009). Activated CaMKIIs further stimulate MeCP2 phosphorylation (Zhou et al., 2006), allowing the transcription of downstream target genes. A recent study showed that MeCP2 phosphorylation at S421 (p-MeCP2) is essential for the expression of metaplasticity and the sustained, but not acute, antidepressant effects of ketamine (Kim et al., 2021). Hippocampal BDNF protein levels were shown to increase rapidly 30 min after ketamine administration but returned to baseline 3 days after injection. In contrast, hippocampal p-MeCP2 levels increased 3 and 7 days, but not 30 min, after ketamine injection. CaMKIIβ were elevated at 3 days after ketamine injection but returned to baseline at 7 days. These findings indicate that CaMKIIβ plays a role in the intermediary process between BDNF activation and MeCP2 phosphorylation required for the sustained antidepressant effects of ketamine. This hypothesis is also supported, at least in part, by a recent finding that hippocampal CaMKIIβ is downregulated in chronic stress-susceptible mice and that short-term (within 4 days) CaMKIIβ activation ameliorates depression-like behaviors (Sakai et al., 2021).

**Epigenetic Regulation of Gene Transcription by Ketamine**

The interplay between genetic and environmental factors underlies depression pathophysiology, and epigenetic mechanisms might contribute to these interactions (Nestler et al., 2016; Uchida et al., 2018; Kawatake-Kuno et al., 2021). Although accumulating evidence demonstrated altered epigenetic functioning in animal models of depression and postpartum MDD-patient brains, few studies have used ketamine-induced transcriptome and epigenome analyses to characterize ketamine’s antidepressant effects. Genome-wide transcriptome and epigenome mapping offer a template for several strategies to identify novel drug targets in unbiased ways to develop more effective treatments for MDD (Bagot et al., 2017). Here we summarize how ketamine-induced activation of Ca^{2+} signal influences epigenetic regulation of gene transcription.

MeCP2, MEF2, and HDAC5 functions are regulated by Ca^{2+} signaling and are associated with stress and depression (Table 1). As mentioned above, p-MeCP2 is necessary for sustained antidepressant response to ketamine (Kim et al., 2021). MeCP2 is a methylated cytosine reader that impacts chromatin organization with any change in DNA methylation. A previous report showed that chronic stress differentially modulates MeCP2 activity in stress-resistant and -susceptible mice and subsequent epigenetic gene transcription (Uchida et al., 2011). Thus, ketamine-induced enhancement of p-MeCP2 may be associated with the formation of chromatin-remodeling complexes on target genes and, thus, transcription regulation. HDAC5 is a histone deacetylase, and its phosphorylation by CaMKs is associated with transcription repression (Mckinsey et al., 2000). Hippocampal HDAC5 is associated with behavioral response to chronic stress and traditional antidepressants.
(e.g., imipramine and SSRIs) (Tsankova et al., 2006; Higuchi et al., 2016). A recent study suggested that ketamine rapidly induces HDAC5 phosphorylation and nuclear export through CaMKII-dependent pathways, which leads to enhanced ME2F transcription that regulates neuronal structural and functional plasticity (Choi et al., 2015). Correspondingly, HDAC5 knockdown occludes the actions of ketamine. Moreover, MeCP2 is considered as a master regulator of metaplasticity (Chen et al., 2012). Ca^{2+}-signal-mediated modulation of MeCP2, HDAC5, and ME2F functions may be involved in the sustained antidepressant response of ketamine through epigenetic transcription.

CONCLUSION

This mini-review highlights that the glutamatergic pathway is associated with behavioral, neuroplastic, neurobiological, molecular, and epigenetic effects of ketamine, focusing on CA^{2+} signaling wherein its dysfunction is involved in depression pathophysiology according to both clinical and animal studies. Such (reverse) translational implications for bridging the research gap between human depression and animal models will provide a better understanding of how ketamine affects and modulates depression pathophysiology and ultimately contribute to the clinical application of ketamine or the development of related compounds for wide range of psychiatric disorders. Glutamatergic transmission and monoaminergic systems induce rapid biological changes that induce fast antidepressant effects. In contrast, ketamine’s sustained antidepressant actions are likely mediated by intracellular CA^{2+} signaling cascades that affect neurobiological processes, including dendritic spine formation, epigenetic modifications, and long-term synaptic plasticity, and consequently, maintain physiological functioning.

In this mini-review, we particularly focused on the hippocampus and prefrontal cortex, key brain regions associated with MDD pathophysiology and ketamine’s antidepressant effect. However, other brain regions were suggested to also be involved in these processes, such as the lateral habenula. Emerging evidence from preclinical and clinical studies identified an important role of the lateral habenula in depression and ketamine’s antidepressant effect through a glutamatergic pathway (Li et al., 2013; Cui et al., 2018a,b, 2019; Yang et al., 2018; Hu, 2019, Hu et al., 2020). In addition, dynamic molecular changes were observed in the nucleus accumbens of animal models of depression and ketamine-treated animals (Bagot et al., 2017). Thus, future studies are warranted to clarify how ketamine impacts neuronal circuit activity and identify underlying molecular and epigenetic mechanisms.

In summary, ketamine has great potential in the development of groundbreaking neuropsychiatric therapies. Our current understanding of depression pathophysiology and ketamine’s action suggests that diverse drug actions converge around CA^{2+}-signaling-mediated neural plasticity. However, ketamine plays diverse roles in the glutamatergic pathway and other neurotransmitter systems, neurogenesis, inflammation, and even body-brain crosstalk. Furthermore, several studies have suggested the distinct roles of ketamine enantiomers ([S]-ketamine and [R]-ketamine) and their metabolites ([2R,6R]-HNK and [2S,6S]-HNK) in plasticity and behavior (Zanos et al., 2016; Yamaguchi et al., 2018; Hashimoto, 2019; Lumsden et al., 2019; Yokoyama et al., 2020; Highland et al., 2021; Wei et al., 2021). Thus, mechanisms underlying ketamine’s actions remain controversial. Moreover, ketamine effects at the mesoscale of neural architecture and macroscale of neural connectivity, cognition, and behavior are poorly understood. Further investigations at both the multiscale and multisystem levels are necessary to comprehensively understand mechanisms underlying ketamine’s antidepressant effects and develop novel drugs for treating MDD.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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