Mild Hypoxic-Ischemic Injury in the Neonatal Rat Brain: Longitudinal Evaluation of White Matter Using Diffusion Tensor MR Imaging

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BACKGROUND AND PURPOSE: Selective white matter (WM) damage is a known sequela of mild hypoxic-ischemic (HI) injury in the neonatal rat model. The aim of this study was to evaluate longitudinally mild HI-induced WM damage (represented by the external capsule [EC]) by diffusion tensor MR imaging (DTI) and to correlate the findings with histology.

MATERIALS AND METHODS: Seven-day-old Sprague-Dawley rats (n = 19) underwent unilateral ligation of the left common carotid artery followed by hypoxia for 50 minutes to create mild HI injury. DTI was performed longitudinally at 5 time points from day 1 to day 90 postinjury (n = 19, 16, 13, 11, 9, respectively), and fractional anisotropy (FA), trace, radial diffusivity (\(\lambda_r\)), and axial diffusivity (\(\lambda_a\)) of the injury and control contralateral ECs were quantified. Rats were randomly sacrificed (n = 15, in total), and the corresponding ECs were stained with hematoxylin-eosin, Luxol fast blue (LFB), and neurofilament (NF) to evaluate morphologic changes, amount of myelin, and axonal count at every time point. A paired t test was applied to evaluate statistical differences between both ECs, and the Pearson correlation test was used to evaluate the relationships between DTI indices and histologic evaluations. In addition, longitudinal changes in DTI indices and histologic evaluations were analyzed by a linear mixed model and an analysis of variance test, respectively.

RESULTS: We demonstrated significantly decreased FA, increased \(\lambda_r\), and similar \(\lambda_a\) in the injury compared with the control EC, which was persistent through all time points. Histologic evaluation by LFB and NF staining showed reduced myelin stain intensity in the injury EC and similar axonal counts in both ECs. Longitudinally, there was an increase in FA, a decrease in \(\lambda_r\) and trace, and stability in \(\lambda_a\) in both ECs. Also, there was progressive reduction in the differences in FA, trace, and \(\lambda_r\) between the injury and control EC, especially between day 1 and day 7 postinjury and in tandem with changes in myelin stain. FA was significantly correlated with myelin stain (\(r = 0.681, P < .01\)) and axonal count (\(r = 0.673, P < .01\)), whereas \(\lambda_r\) was significantly correlated with myelin stain only (\(r = -0.528, P < .01\)), and \(\lambda_a\) with axonal count only (\(r = 0.372, P = .043\)).

CONCLUSIONS: Diffusion indices can reflect dysmyelination in mild HI injury, continual myelination of both injury and control ECs with growth, and the partial recovery of myelin postinjury. We propose that diffusion indices may be used as biomarkers to monitor noninvasively the longitudinal changes of mild HI-induced WM damage.
young adult period, and correlated the diffusion indices at every time point with histology. We aimed to evaluate the evolution of HI-induced WM damage, represented by an external capsule (EC), the main region of insult in this mild HI model,1,4 in particular to determine if there were continual maturation and a component of recovery of WM damage and if this may be monitored by DTI indices.

Materials and Methods

Animal Model Preparation

According to the local government legislation, the experiment was approved by the University Animal Ethics Committee. Seven-day-old Sprague-Dawley rats (n = 19) with a mean body weight of 18 ± 2 g were obtained from the Laboratory Animal Unit of The University of Hong Kong. Briefly, the 7-day-old rats underwent unilateral ligation of the left common carotid artery via a midline neck incision after anesthesia with 0.2 mL of inhalational isoflurane in an airtight box for 2 minutes. After that, they were returned to their mother for nursing for 2 hours until they regained normal movement. These rats were subsequently placed in a hypoxic chamber of 8% O2/92% N2 main-tained at 37°C for 50 minutes to create mild HI injury.18

MR Imaging Data Acquisition

The rats underwent MR imaging at post-HI injury day 1 (n = 19), day 7 (n = 16), day 14 (n = 13), day 30 (n = 11), and day 90 (n = 9) postsurgery. DTI and T2-weighted MR imaging were performed by using a 7T nuclear MR scanner with a maximum gradient of 360 mT/m (Version 70/16; PharmaScan, Bruker Biospin, Rheinstetten, Germany) with a microimaging mouse brain coil (for day 1 and day 7) or a rat brain coil (day 14, day 30, and day 90). During scanning, rats were prostrate on a custom-made holder with strapping to minimize head motion while respiration was monitored. Under inhaled isoflu-rane anesthesia (3% for induction and 1.5% for maintenance), scout images were first acquired in 3 planes with a T2-weighted sequence to position the subsequent DTI images along standard anatomic orienta-tions. Coronal T2-weighted images were obtained to confirm the lack of cystic lesions from 2 mm anterior to the corpus callosum to the end of the cerebrum with the following parameters: TR = 11,189 ms, TE = 20 ms, FOV = 2.5 cm², acquisition matrix = 128 × 128, section thickness = 1.0 mm. DTI images were acquired with a respiration-gated spin-echo 4-shot echo-planar imaging sequence and an encoding scheme of 35 gradient directions, which were uniformly distrib-uted on the unit sphere. We used the following imaging parameters: TR = 3000 ms, TE = 32 ms, Δ = 20 ms, δ = 4 ms, FOV = 3.2 cm² (for day 1 and day 7), 4.0 cm² (for day 14, day 30, and day 90), thickness = 0.5 mm (for day 1 and day 7), 0.7 mm (for day 14, day 30, and day 90), acquisition matrix = 128 × 128 (zero-filled to 256 × 256), image resolution = 250 × 250 μm² (for day 1 and day 7), and 313 × 313 μm² (for day 14, day 30, and day 90), acquisition time = 8 minutes, b-value = 0 and 1000 s/mm².

All diffusion-weighted images were first coregistered by using AIR, Version 5.2.5 (http://bishopw.loni.ucla.edu/AIR5/index.html) to compensate for the eddy current–induced displacements that were dependent on the diffusion-gradient directions. DTI indices, including FA, Trace, and eigenvalues (λ1, λ2, and λ3), were obtained by using DTIStudio, Version 2.30 (Johns Hopkins University, Balti-more, Md). Then, FA, Trace, λ1, and λ2 maps were created for quanti-tative analysis by the following equations:

1) $FA = \sqrt[3]{\frac{1}{2} \left( \frac{(λ_1 - λ_2)^2 + (λ_2 - λ_3)^2 + (λ_3 - λ_1)^2}{λ_1^2 + λ_2^2 + λ_3^2} \right)}$

2) $Trace = λ_1 + λ_2 + λ_3$

3) $λ_i = λ_1$

4) $λ_1 = 0.5 \times (λ_2 + λ_3)$

Image Analysis

After the generation of FA, Trace, λ1, and λ2 maps, a region of interest was manually drawn over the both sides of the EC on an FA map at day 30 post-HI injury (circled area). Then, the regions of interest are placed on the identical sites on the trace, λ1, and λ2, maps.

Histopathology Evaluation

Rats were randomly selected for histologic evaluation of WM damage (day 1, n = 3; day 7, n = 3; day 14, n = 2; day 30, n = 2; day 90, n = 5). Brain specimens were processed by using standard histologic protocols. Briefly, rat brains were perfusion-fixed through the left cardiac ventricle with phosphate-buffered saline (PBS) followed by 4% para-formaldehyde (PFA) in PBS. The specimens were fixed in 4% PFA in PBS (pH 7.4) at 4°C overnight. The brains were cut into a 30 μm-thick coronal sections between the locations corresponding to the most posterior and anterior MR imaging sections. Then the brain sections were stored at −70°C until immunohistochemical analysis.

Hematoxylin-eosin (HE) stain was used to evaluate morphologic characteristics of WM. Luxol fast blue (LFB) staining was performed to evaluate the amount of myelin in WM. Tissue sections were pro-cessed as free-floating and were incubated in the monoclonal anti-body to panaxonal neurofilament marker (NF) (SMI-312, 1:1000) for

Fig 1. Regions of interest are manually drawn over the both sides of the EC on an FA map at day 30 post-HI injury (circled area). Then, the regions of interest are placed on the identical sites on the trace, λ1, and λ2 maps.
immunohistochemistry staining of axons. Appropriate secondary antibodies were used at a dilution of 1:200 in 0.1 mol/L PBS and incubated in the secondary antibody goat antimouse immunoglobulin G fluorescein isothiocyanate.

Histologic specimens were analyzed within the region of EC corresponding to the quantitative MR imaging measurements. All sections were examined by using a light microscope (Axioplan 2 imaging system; Carl Zeiss, Goettingen, Germany) under 10×-400× magnification. Histologic images were acquired at the same exposure level by digital photomicrography (SPOT Advanced; Diagnostic Instruments, Farmington Hills, Mich) and quantitatively analyzed by using ImageJ software. To quantitatively evaluate the LFB-stained sections in both symmetric ECs at 200× histologic digital images by using the automated software ImageJ. Optical attenuation is a measurement of the degree of staining intensity based on a gray-scale and calibrated to a standardized optical attenuation value. With a higher LFB staining intensity, the optical attenuation value is higher. For NF stain, the axonal count was automatically calculated by ImageJ at the symmetric EC at 200× histologic digital images.23

Statistical Analysis
All results were expressed as mean ± SD. Ratios of injury/control DTI indices (e.g., FA/FAC) and injury/control quantitative evaluations of LFB staining intensity and axon count (LFB/LFBc, NF/NFc) were calculated for statistical analysis. A paired t test was used to detect statistical differences in the DTI indices and histologic evaluations between the injury and control ECs. After correction for multiple comparisons (5 times the repeated measurement of DTI indices), P < .01 was regarded as a significant difference in the comparison of injury/control DTI indices. Because DTI indices are repeated-measures data, longitudinal changes of DTI indices were analyzed by using a linear mixed model, followed by a least significant difference post hoc pair-wise comparison test. The ratios and absolute values of DTI indices were the dependent variables, whereas the independent variables were subjects (a random factor) and time points (a categoric variable). Because the histologic results are cross-sectional data, the changes of absolute and ratios of injury/control histologic evaluations between different time points were evaluated by the 1-way analysis of variance (ANOVA) test, followed by the Tukey test. The Pearson correlation test was used to evaluate correlations between DTI indices and histologic staining intensity. To evaluate the intraobserver reliability of region-of-interest measurement of DTI indices, we randomly selected 4 rats in every time point (total of 20 rats) for remeasurement. To determine the consistency of DTI indices between different FOVs, we repeated measurements of DTI indices in 3 randomly selected rats, which were scanned by using the 2 different FOVs (3.2 cm² and 4.0 cm²). Intraobserver reliability and consistency of different FOVs were assessed by calculating the 1-way random intraclass correlation coefficients (ICC). All statistical analyses were performed by using the Statistical Package for the Social Sciences for Windows (Version 15, SPSS Inc, Chicago, Ill). A P value of <.05 was considered to indicate statistical significance.

Results
General Results of DTI Scanning
Of all DTI sections, 98.1% (n = 1334/1360) were satisfactory in image quality and were included in the region-of-interest analysis. Twenty-six sections belonging to 5 rats were excluded due to motion artifacts, as determined visually. None of the rats were excluded from image analysis. Intraobserver reliability analysis of manual region-of-interest drawings showed good agreement (ICC = 0.94, 0.96, 0.96, and 0.91 for FA, trace, λ1, and λ2). An excellent consistency was also obtained in the measurement of DTI indices by different FOVs (ICC = 0.991, P < .001).

Comparison of DTI Indices between Injury and Control ECs
DTI indices of FA, trace, λ1, and λ2 in injury and control ECs at every time point are shown in the Table 1.

FA

| Time Points | Injury EC  | Control EC  | Ratio  | P    |
|-------------|-----------|-------------|--------|------|
| D1          | 0.240 ± 0.051a | 0.262 ± 0.057b | 0.922 ± 0.091a | <.01 |
| D7          | 0.307 ± 0.068a | 0.321 ± 0.053a | 0.962 ± 0.132a | <.01 |
| D14         | 0.326 ± 0.040a | 0.337 ± 0.061a | 0.967 ± 0.056a | <.01 |
| D30         | 0.377 ± 0.043a | 0.382 ± 0.054a | 0.958 ± 0.114a | <.01 |
| D90         | 0.416 ± 0.032a | 0.430 ± 0.040a | 0.970 ± 0.098a | <.01 |

Trace (μm²/ms)

| Time Points | Injury EC  | Control EC  | Ratio  | P    |
|-------------|-----------|-------------|--------|------|
| D1          | 2.908 ± 0.204a | 2.683 ± 0.351a | 1.059 ± 0.121a | <.01 |
| D7          | 2.772 ± 0.019a | 2.723 ± 0.016b | 1.001 ± 0.054b | .97 |
| D14         | 2.162 ± 0.154a | 2.145 ± 0.130b | 1.008 ± 0.033b | .08 |
| D30         | 2.284 ± 0.277a | 2.260 ± 0.125b | 1.012 ± 0.053b | .13 |
| D90         | 2.272 ± 0.317a | 2.247 ± 0.303b | 1.011 ± 0.042b | .09 |

λ1 (μm²/ms)

| Time Points | Injury EC  | Control EC  | Ratio  | P    |
|-------------|-----------|-------------|--------|------|
| D1          | 1.107 ± 0.108 | 1.125 ± 0.107 | 0.987 ± 0.083 | .10 |
| D7          | 1.089 ± 0.109 | 1.091 ± 0.099 | 1.000 ± 0.060 | .86 |
| D14         | 1.082 ± 0.079 | 1.080 ± 0.079 | 1.003 ± 0.039 | .73 |
| D30         | 1.075 ± 0.084 | 1.073 ± 0.098 | 1.008 ± 0.113 | .93 |
| D90         | 1.087 ± 0.159 | 1.081 ± 0.157 | 1.009 ± 0.089 | .56 |

λ2 (μm²/ms)

| Time Points | Injury EC  | Control EC  | Ratio  | P    |
|-------------|-----------|-------------|--------|------|
| D1          | 0.835 ± 0.083a | 0.769 ± 0.086a | 1.092 ± 0.079a | <.01 |
| D7          | 0.645 ± 0.060a | 0.616 ± 0.048a | 1.050 ± 0.100a | <.01 |
| D14         | 0.621 ± 0.061a | 0.594 ± 0.052a | 1.048 ± 0.093a | <.01 |
| D30         | 0.617 ± 0.045a | 0.591 ± 0.043a | 1.046 ± 0.073a | <.01 |
| D90         | 0.583 ± 0.076a | 0.570 ± 0.078a | 1.042 ± 0.062a | <.01 |

Note.—FA indicates fractional anisotropy; EC, extracapsular; λ1, axial diffusivity; λ2, radial diffusivity; Ratio, injury/control diffusion tensor imaging indices of EC D1-D90, day 1-day 90 post HI; HI, hypoxic-ischemic.

* A paired t test was used to evaluate the statistical significance between injury and control ECs. A linear mixed model, followed by a post hoc pair-wise comparison test, was applied to evaluate significant differences among longitudinal time points. Superscript a,b,c,d,e reflect significant differences between time points. Values in the same column without a common superscript indicate a significant difference less than 0.05 between 2 time points. Values are shown as mean ± SD.

Table 1: FA, trace, λ1, and λ2 in injury and control ECs from D1 to D90 in a mild-HI neonatal rat model

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Longitudinal Changes of DTI Indices
Longitudinal changes of DTI indices from day 1 to day 90 are shown in the Table 1.
There was a progressive increase in FA from day 1 to day 90, a decrease in trace from day 1 to day 14, and a decrease in \( \lambda_{\perp} \) from day 1 to day 90 in injury and control ECs.

**FA.** There were statistically significant differences in FA among the time points in both injury and control ECs (\( P < .01 \) for both). A subsequent post hoc pair-wise comparison test showed a significant increase in FA among the consecutive time points except between day 7 and day 14 on both sides of the EC (all \( P < .01 \), except between day 7 and day 14).

**Trace.** Statistically significant differences were found in trace among the time points on both sides of the EC (\( P < .01 \) for both). A post hoc pair-wise comparison test showed a significantly decreased trace between day 1 and all the subsequent time points (all \( P < .01 \), but there was no significant difference among the time points day 7, day 14, day 30, and day 90.

**\( \lambda_{\perp} \).** Statistically significant differences were found in \( \lambda_{\perp} \) among the time points on both sides of the EC (\( P < .01 \) for both). A post hoc pair-wise comparison test showed significantly decreased \( \lambda_{\perp} \) between day 1 and all subsequent time points (all \( P < .01 \), but there were no significant differences among the time points day 7, day 14, day 30, and day 90.

**\( \lambda_{\parallel} \).** There was no significant difference found in \( \lambda_{\parallel} \) among all time points on both sides of the EC.

Evaluation of the ratios of the diffusion indices showed statistically significant differences in \( \text{FA}/\text{FA}_{C} \), \( \text{trace}/\text{trace}_{C} \), and \( \lambda_{\perp}/\lambda_{\perp,C} \) among the time points (\( P = .006 \), \( P < .001 \), and \( P = .002 \), respectively) but not in \( \lambda_{\parallel}/\lambda_{\parallel,C} \) (\( P = .328 \)). A subsequent post hoc pair-wise comparison test showed significantly increased \( \text{FA}/\text{FA}_{C} \), decreased \( \text{trace}/\text{trace}_{C} \), and decreased \( \lambda_{\perp}/\lambda_{\perp,C} \) between day 1 and all the subsequent time points (all \( P < .05 \)), but there was no significant difference among the time points day 7, day 14, day 30, and day 90.

**Histopathological Evaluation**

**HE Stain.** With HE staining, on day 1 post-HI injury, all 3 rats were found to have mild vacuolation and a thinner EC in the injury EC compared with the control EC (Fig 2A, -B). Mild vacuolation change was observed in 1 rat in the injury EC on day 14 post-HI injury, but the other 2 rats had symmetric hemispheres. No necrosis or infarct regions were observed in the injury EC at any time points.

**LFB Stain.** LFB stain was observed on both sides of the EC, indicating the presence of myelin. Weaker LFB staining intensity was observed in the injury EC compared with the control EC in all rats from day 1 to day 90 post-HI injury (Fig 2C, -D). Quantitative analysis showed significantly decreased optical
attenuation of LFB staining intensity in the injury EC compared with control EC in all time points (Table 2A). The ANOVA test showed significant differences in LFB staining intensity among different time points (P < .001 for both ECs). A subsequent post hoc Tukey test showed that the progressive increase in LFB staining intensity was statistically significant between all consecutive time points from day 1 to day 90 in both injury and control ECs, except between day 7 and day 14 (P > .05). The increase in LFB staining intensity was statistically significant less than 0.05 between 2 time points. Values are shown as mean ± SD.

Table 2: Histologic evaluations of mild HI-induced WM damage in injury and control ECs from D1 to D90 post-HI

|                | Injury EC | Control EC | Ratio | P     |
|----------------|-----------|------------|-------|-------|
| LFB staining intensity (optical density) |           |            |       |       |
| D1             | 0.055 ± 0.006* | 0.081 ± 0.008* | 0.684 ± 0.086* | <.01  |
| D7             | 0.098 ± 0.008* | 0.129 ± 0.012* | 0.768 ± 0.093* | <.01  |
| D14            | 0.113 ± 0.014* | 0.143 ± 0.015* | 0.801 ± 0.169* | <.01  |
| D30            | 0.143 ± 0.015* | 0.172 ± 0.012* | 0.818 ± 0.083* | <.01  |
| D90            | 0.177 ± 0.021* | 0.214 ± 0.026* | 0.824 ± 0.092* | <.01  |
| Axon count     |            |            |       |       |
| D1             | 201 ± 29a  | 234 ± 32a  | 0.859 ± 0.129 | .074  |
| D7             | 463 ± 48a  | 503 ± 85a  | 0.920 ± 0.084 | .254  |
| D14            | 679 ± 19   | 697 ± 32   | 0.974 ± 0.025 | .087  |
| D30            | 701 ± 132  | 716 ± 89   | 0.979 ± 0.084 | .724  |
| D90            | 1059 ± 476b| 1240 ± 402b| 0.854 ± 0.025 | .335  |

Note:—WM indicates white matter; LFB, Luxol fast blue.
* A paired t test was used to evaluate the statistical significance between injury and control ECs. One-way analysis of variance followed by a Tukey test was used to evaluate the statistical significance among longitudinal time points. Superscript a,b,c,d,e reflect significant differences between time points. Values in the same column without a common superscript indicate a significant difference less than 0.05 between 2 time points. Values are shown as mean ± SD.

Mild HI insult produces predominant injury in the WM in rats with a relative sparing of the GM.1,4,18 In a similar rodent model, Qiao et al1 found that there were atrophy and vacuolar changes in the WM, whereas the GM appeared normal. In another study,4 most examined ECs (8 of 14 brains) showed spongiform and rarefied tracts. However, most of the cortex regions (11 of 14 brains) appeared normal. The other 3 brains had mild neuronal vacuolar changes in the cortex. Selective injury to the WM in mild hypoxia-ischemia is due to the high susceptibility of oligodendrocytes to HI injury.22,23 It was found that immature oligodendrocyte progenitors were dying as early as 3 hours after HI insult,23 and at 24–48 hours post-HI injury, there was dysfunction of maturing oligodendrocytes, which resulted in ≤50% decrease in myelin basic protein.24 Thus, dysmyelination is a major pathologic process in mild HI-induced WM damage, and this is reflected by decreased LFB staining intensity.4,8,25

Correlations between DTI Indices and Histologic Evaluations

FA was significantly correlated with both LFB staining intensity (r = 0.681, P < .01) and axonal count (r = 0.673, P < .01). λ₁ was significantly correlated with LFB staining intensity only (r = −0.528, P < .01), and λ₁ was significantly correlated with axonal count only (r = 0.372, P = .043). No significant correlations were demonstrated between trace and histologic findings.

Discussion

In this longitudinal study, we demonstrated that changes in DTI indices could reflect the pathologic changes of reduced myelination in the injury EC by a reduction in FA, increase in λ₁, and a similar λ₁/λ₀ and this was persistent from day 1 to day 90. Moreover, the longitudinal changes in DTI indices with an increase in FA and a decrease in λ₁ and trace in both the injury and control EC, in parallel with histologic evidence of increased myelin, are in keeping with the pattern of normal development and continual maturation of WM. Finally, the gradual reduction in the differences in FA, trace, and λ₁ between injury and control ECs in tandem with LFB staining intensity suggests a partial recovery process in the injury EC, and this was most evident between day 1 and day 7. Indeed, LFB staining intensity was found to correlate significantly with FA and λ₁.

Dynamic change in trace reflects the mean diffusivities in different directions. Dynamic change in trace reflects the water distribution between extra- and intracellular space. Initial increase in trace post-HI injury as found in our study is commonly explained as water influx from vessels to brain tissue,27 namely vasogenic edema. In the subsequent time points, similar trace was found between both sides of the EC. This is likely due to absorption of vasogenic edema and an additional component of restoration of myelin.
Studies have shown that directional diffusivities could provide more specific information about the pathologic changes in WM such as myelin damage or axonal degeneration compared with FA and trace. Because $\lambda_1$ represents water diffusion perpendicular to the myelin sheaths, myelination causes $\lambda_1$ to reduce, and conversely, loss of myelin causes $\lambda_1$ to increase. Significantly increased $\lambda_1$ has been demonstrated in a shiverer mouse model with dysmyelination. Dynamic changes of FA and trace may be explained by concomitant maturation-induced changes in tissue microstructure, such as reduction in water content, greater cohesiveness of fiber tracts or fiber organization, maturation of axons, and myelination. These processes modify water diffusion during brain development and influence diffusivities. It has been proposed that during maturation, increase of longitudinally oriented neurofibrils and the elevation of fast axonal transport increase water diffusion along axons, which increases $\lambda_J$. However, axonal pruning reduces intermingling axon branches and shortens the length of axons, which decreases $\lambda_J$. Therefore, longitudinal changes of $\lambda_J$ may not be significant. Myelination is another important maturation process that influences water diffusion in the WM. It is suggested that increased FA and reduced $\lambda_1$ occur in parallel with and therefore reflect myelin concentration during the maturation process.

We found similar longitudinal changes of DTI indices in both the injury and control sides of the EC corresponding to the changes in both injury and control WM, and the partial recovery of myelin in post-HI injury WM. 

Conclusions

In conclusion, our results support the use of DTI indices as biomarkers to monitor the longitudinal changes of mild HI-induced WM damage noninvasively. DTI indices are able to reflect dysmyelination, the process of continual myelination of both injury and control WM, and the partial recovery of myelin in post-HI injury WM.

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