Genetic Factors in Nonsyndromic Orofacial Clefts

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Introduction

Orofacial clefts (OFCs) are the most common congenital birth defects in humans and immediately recognized at birth. The etiology remains complex and poorly understood and seems to result from multiple genetic and environmental factors along with gene–environment interactions. It can be classified into syndromic (30%) and nonsyndromic (70%) clefts. Nonsyndromic OFCs include clefts without any additional physical or cognitive deficits. Recently, various genetic approaches, such as genome-wide association studies (GWAS), candidate gene association studies, and linkage analysis, have identified multiple genes involved in the etiology of OFCs. This article provides an insight into the multiple genes involved in the etiology of OFCs. Identification of specific genetic causes of clefts helps in a better understanding of the molecular pathogenesis of OFC. In the near future, it helps to provide a more accurate diagnosis, genetic counseling, personalized medicine for better clinical care, and prevention of OFCs.

Epidemiology

The prevalence of OFCs ranges from 1 in 700 to 1,000 newborns worldwide13 and include cleft lip only (CLO), cleft palate only (CPO), and cleft lip and palate (CLP).14 They may occur as unilateral or bilateral, complete, or incomplete, and may involve the lip only, the palate only, or both.15

The prevalence of OFC varies according to geographical location, ethnicity, race, and socioeconomic status.16–19 Asians have the highest prevalence rate (1:500), the intermediate prevalence in Europeans (1:1000), and lowest in Africans (1:2500).20–22 In India, the incidence of clefts is around 1:800 to 1:1000, and three infants are born with some type of cleft every hour.23 These differences appear to persist even after migration, suggesting that they are mediated by genetic, rather than environmental factors.
Factors. Overall, 70% of the OFCs are nonsyndromic (NS) and occur as isolated cases without any additional physical or cognitive deficits. In contrast, 30% of clefts are syndromic and are associated with a few other developmental anomalies.

The frequency of occurrence of OFC differs with regard to gender and side of clefting. Cleft lip is more common in males at a 2:1 male to female ratio, whereas a cleft palate is more common in females. Approximately 90% of OFCs are unilateral with primarily left-sided involvement.

Development of Cleft Lip and Palate

The development of lip and palate begins during the fourth week of gestation where migrating neural crest cells combine with mesodermal cells to establish facial primordia which consist of five different processes (medial nasal, lateral nasal, frontonasal, maxillary, and mandibular processes) derived from the first pharyngeal arch. Once facial prominences are formed, the nasal placodes invaginate to form the medial nasal process (MNP) and lateral nasal process (LNP). The maxillary processes initially grow medially, pushing the LNP toward the upper side. During the sixth and seventh weeks of gestation fusion of maxillary processes with each other and also with lateral and MNPs takes place to form upper lip and primary palate. Defect in fusion or failure in the growth of these processes results in clefts involving the upper lip, alveolus, and/or primary palate.

The secondary palate begins to develop in the seventh week of embryogenesis; the maxillary processes initially outgrow as palatal shelves which move toward each other vertically. After proper growth they settle at a horizontal position above the tongue which entails much extracellular remodeling. The palatal shelves then fuse in the midline both anteriorly and posteriorly like a zipper that forms a midline epithelial seam (MES). The disintegration of MES is required to maintain palatal confluence, which may involve apoptosis, epithelial-mesenchymal transition (EMT), and cell migration. Effective fusion of the secondary palate results in complete separation of the oral and nasal cavities. Cleft palate can result from failure at any of the steps, including palatal shelves elevation, cell migration, or fusion.

In summary, a variety of cellular mechanisms such as cell proliferation, cell migration, cell growth, cell fusion, apoptosis, EMT, and extracellular remodeling are involved in a coordinated manner during the development of lip and palate. Therefore, disruption in the gene/s involved in these processes during lip and palate development may lead to an OFC.

Glimpse into the History of Genetic Etiology of Orofacial Clefts

Although the familiarity of OFC has long been noted, Fogh-Andersen was the first to provide the evidence for genetic factors contributing to the etiology of CLP from family-based studies where it was observed that the siblings of CLP patients had an increased frequency of cleft lip with or without cleft palate. This observation was further confirmed by studies on the familial distribution of congenital clefts of the lip and palate, and Dr. Clark Fraser published a review paper highlighting the conclusions of a workshop on CLP sponsored by the National of Institutes of Health of the United States, and he mentioned the etiology is indeed multifactorial. Later, more evidence in favor of contribution of genetic factors to the etiology of CLP accrued from segregation analysis and twin studies where the monozygotic twins showed high prevalence rate (40%) than the dizygotic twins (4%).

Role of Genetic Factors in the Etiology of Orofacial Clefts

The etiology of OFCs involves genetic factors, environmental influences, and gene–environment interactions, all contributing to its susceptibility. Scientific literature evidence suggests that environmental factors such as maternal tobacco smoking and alcohol consumption, antiepileptic medications, maternal folate deficiency, infections, consanguinity, and geographical location are risk factors for NS cleft lip and palate (NSCLP). Advances in genetics and molecular biology techniques have discovered multiple genes and loci associated with CLP. This article provides an overview of the genes implicated in the etiology of NSCLP. Identification of specific genetic variation contributing to NSCLP has led to an increase in our understanding of the molecular pathogenesis of OFC.

Genes Involved in the Etiology of Nonsyndromic Orofacial Clefts

Special AT-Rich Sequence-Binding Protein

Special AT-rich sequence-binding protein (SATB2) is a DNA binding protein which binds with nuclear matrix attachment regions. It is involved in transcription regulation and chromatin remodeling process. In an animal study, mouse SATB2 is strongly expressed in the developing cleft palate and is similar to the human SATB2 protein. The identification of SATB2 gene responsible for the craniofacial dysmorphologies associated with deletions and translocations at 2q32-q33, only one region of the genome has been significantly associated with the development of isolated cleft palate. Glass syndrome characterized by cleft palate, gum hyperplasia, slight micrognathia, generalized osteoporosis, and mental retardation reported from Thai patient was caused by SATB2 gene mutation. Recently, using salivary miRNAs showed that the SATB2 genes are involved in the development of the cleft palate and lip development. Table 1 provides the list of genes involved in the etiology of NS OFC in humans.

B-Cell Leukemia/Lymphoma 3

B-cell leukemia/lymphoma 3 (BCL3) is a proto-oncogene, acts as a transcriptional co-activator through NF-kappa-B target genes and located on 19q13.2. BCL3 gene has also shown a strong association with NS orofacial clefts (NSOCs). A case-parent trio study showed BCL3 influence risk of CL/P through a parent-of-origin effect with the excess maternal transmission. Several studies in different
Table 1 Genes involved in the etiology of nonsyndromic orofacial clefts in humans

| Gene                        | Gene symbol | Loci          | OMIM       | Evidence | References     |
|-----------------------------|-------------|---------------|------------|----------|----------------|
| Special AT-rich sequence-binding protein | SATB2       | 2q33.1        | 608148     | M        | 43,46          |
| B-cell leukemia/lymphoma 3  | BCL3        | 19q13.32      | 109560     | LD, L    | 47,49          |
| Distal-less homeobox 4      | DLX4        | 17q21.33      | 601911     | M        | 53,55,56       |
| Paired box gene 9           | PAX9        | 14q13.3       | 167416     | L, GWAS  | 6,60,62,64     |
| Netrin 1                    | NTN1        | 17p13.1       | 601614     | GWAS     | 70,71          |
| T-box transcription factor 22| TBX22       | Xq21.1        | 300307     | M, LD    | 75,77,79       |
| Poliovirus receptor like-1  | PVRL1       | 11q23.3       | 600644     | M, LD, GWAS | 81,83,85   |
| Cleft lip and palate associated transmembrane protein 1 | CLPTM1       | 19q13.32     | 604783     | M, L, LD | 89,90          |
| MAF bZIP transcription factor B | MAFB        | 20q12         | 608968     | GWAS     | 98,101         |
| Fibroblast growth factor receptor 1 | FGFR1     | 8p11.23       | 136350     | M, D     | 102,103        |
| Transcription factor AP-2α  | TFAP2A      | 6p24.3        | 107580     | D, L     | 104            |
| S-glutathione transferase T1 | GSTT1       | 22q11.2       | 600436     | LD       | 108,109        |
| Receptor-like tyrosine kinase | RYK         | 3q22.2        | 600524     | M        | 110,111        |
| Gamma-aminobutyric acid receptor, Beta-3 | GABRB3   | 15q12         | 137192     | GWAS     | 112            |
| ATP-binding cassette, subfamily A, member 4 | ABCA4   | 1p22.1        | 601691     | GWAS     | 115,117,118    |

Abbreviations: D, deletions; GWAS, genome-wide association studies; L, linkage; LD, linkage disequilibrium; M, mutations; OMIM, Online Mendelian Inheritance in Man.

populations have implicated the role of BCL3 in development of NSCL/P. It was found that BCL3 contributes to the regulation of cell proliferation, and cell cycle regulation can cause a disturbance in facial formation.49 A study also reported BCL3 contribution in angiogenesis-related genes in the etiology of CLP.50 However, no association of BCL3 gene with NSCLP was found in multigenerational families of Indian population.51

Distal-Less Homeobox 4
Distal-Less Homeobox 4 (DLX4) belongs to DLX gene family containing a homeobox transcription factor which plays an important role in craniofacial development and palatogenesis. It is located on chromosome 17q21.33 and causes orofacial cleft 15 (OFC15) and cleft lip/palate. In an animal study, the DLX genes caused cleft palate showing the importance of these genes in craniofacial morphogenesis.52 Whole-exome sequencing study in a Hispanic mother and son with bilateral CLP confirmed the DLX4 as a potential cause of oral clefts.53–55 Recently, a study showed that none of the distal-less 4 (DLX4) gene SNPs were associated with NSOCs, so it should be interpreted with a caution in the etiology of nonsyndromic orofacial clefts.56

Paired Box Gene 9
Paired Box Gene 9 (PAX9) is a member of the paired box (PAX) family of transcription factors and contains a paired box domain, an octapeptide, and a paired-type homeodomain. It plays a critical role during neural crest and fetal development. PAX9 is located on chromosome 14q13.3, consists of five exons, and associated with the formation of the teeth and palate.57,58 Tooth agenesis and the formation of a cleft palate in PAX9-deficient mice have been reported.59

Schuffenhauer et al, first reported the role of PAX9 in a patient presented with bilateral CLP.60 A linkage analysis of PAX9 gene in two large families and four unrelated families showed that the hypodontia primarily involving molars, suggested that the mutant PAX9 protein acquires functional defects in DNA binding, as well as the loss of function of PAX9 resulting in haploinsufficiency during the morphogenesis of the dentition and the subsequent tooth agenesis.61,62 Several studies identified mutations at PAX9 may increase the risk of NS cleft lip with or without palate.63–66

A combined genome-wide association study (GWAS) used unmixed NS CPO and NS CLO subtypes suggested that PAX9 is a strong genetic factor for NS CPO in the Chinese population. Mutation analysis of PAX9 SNPs rs12885612 and rs12881248 revealed that PAX9 is a promising susceptible gene for NS CLO in Western Han Chinese population.5,6

Netrin 1 (NTN1)
The netrin 1 (OMIM: 601614) located on chromosome 17p13.1 is a family of laminin-related secreted proteins. Its functions include axon guidance and cell migration in the central nervous system, angiogenesis, and semicircular canal formation.

NTN1 is expressed at high levels in cells that will come together to form a fusion plate, a prerequisite for the formation of semicircular canals. In netrin 1 mutant mice, fusion plate formation is severely affected, and it stimulates proliferation of the periotic mesenchymal cells which then push the epithelial cell walls together to form the fusion plate.67,68 It affects the development of the craniofacial region and has been shown to play a vital role in regulating cell migration during embryogenesis, and it is also expressed in the medial edges and oral sides of the palatal shelves.69
A GWAS included 1,409 case-parent trios by several research groups with samples of Asian or European ancestry from Europe, the United States, China, and the Philippines found NTN1 role in the etiology of NSCLP. A case–control study of NTN1, identified SNP (rs9788972) as a risk locus for NSOCs susceptibility in a northern Chinese population.

T-Box Transcription Factor 22
The T-box 22 (TBX22) gene encodes transcription factors involved in the regulation of developmental processes and plays a major role in human palatogenesis. It contains eight coding exons. TBX22 cause X-linked cleft palate and ankyloglossia. It is expressed in the developing palatal shelves and at the base of the tongue prior to elevation to a horizontal position above the tongue.

Genome-wide linkage analysis showed the association of TBX22 role in the development of NSCLP. In addition, mutations in TBX22 were found in individuals with isolated CPO. It plays a significant role in tooth and upper lip development and causes hypodontia and cleft lip. DNA methylation study suggests that cleft palate-susceptible gene Tbx22 is associated with gene expression and might be responsible for the developmental failure of palatal fusion, eventually resulting in the formation of cleft palate.

Poliovirus Receptor-Like 1
Poliovirus receptor-like 1 (PVRL1), also known as NECTIN1 (nectin cell adhesion molecule 1) belongs to the nectin subfamily of immunoglobulin-like adhesion molecules that involve cell–cell adhesion. It plays a vital role in the organization of adhesion junctions and tight junctions in epithelial and endothelial cells. During the developmental process, the palatal shelves and palatal epithelium come in close contact and fuse together. PVRL1 plays a major role in these developments during palatogenesis and genetic variations reported to have a significant relationship with CLP. Diseases associated with PVRL1 include cleft lip/palate-ectodermal dysplasia syndrome (CLPED) and herpes simplex.

In animal experiments, PVRL1 expressed at the medial edge epithelium of the palatal shelves and the skin surface epithelium locations that corresponded to the clinical phenotypes of CLPED and in humans, mutations of the PVRL1 gene resulting CLPED in families from Israel and Brazil. Interestingly, heterozygous mutation of PVRL1 (W185X) associated with NSCLP in northern Venezuela and two novel variants of the PVRL1 gene were identified in Turkish NSCLP patients. In an experimental animal studies, PVRL1 expressed at the medial edge epithelium of the palatal shelves and the skin surface epithelium locations that corresponded to the clinical phenotypes of CLPED and in humans, mutations of the PVRL1 genes caused CLPED in Israel and Brazilian population.

Cleft Lip and Palate-Associated Transmembrane Protein 1
Cleft lip and palate-associated transmembrane protein 1 (CLPTM1) is a multipass transmembrane protein that regulates GABA-A receptors (e.g., GABRA1) and modulates inhibitory synaptic strength. It is located on chromosome 19q13.3 and plays a role in T-cell development. Mutation of CLPTM1 genes suggested that a regulatory element in this gene region get affected and which is responsible for the development of orofacial clefts. However, some studies contradict the role of CLPTM1 in the etiology of NSCL/P as no evidence of an association with oral clefts was found among the SNPs of CLPTM1 selected for testing in Japanese and Irish population.

MAF bZIP Transcription Factor B
The MAF bZIP Transcription Factor B (MAFB) gene encodes a basic leucine zipper (bZIP) transcription factor that plays an important role in the regulation of lineage-specific hematopoiesis. It is located on chromosome 20q11.2, consists of a single exon and spans approximately 3 kb. In a mouse study, MAFB was expressed in the palatal shelves and the medial edge epithelia (or MEE) during palatal fusion. Mutations in the MAFB gene reported causing multicentric carpotarsal osteolysis syndrome and Duane retraction syndrome 3 with or without deafness.

Several GWAS in European and Asian populations identified the role of MAFB in NS CLP. Different systematic review and meta-analyses confirmed that the MAFB gene SNP (rs13041247) is associated with NSCL/P risk in different population; however, this association is not significant in East Asian or Caucasian populations, whereas, the SNPs rs17820943 and rs6072081 of MAFB found to be associated with NSCLP in an East Asian population.

Other Candidate Genes
A variety of genetic approaches have identified several genes located on different chromosomes, contributing to etiology of NSCLP. Advances in genetics and molecular biology techniques have led the way to the discovery of genetic variation involved in NSCLP. Genes such as FGFR1, TFAP2A, GSTT1, receptor-like tyrosine kinase (RYK), and ABCA4 have also been found to be associated with NSCLP.

Riley et al assessed the genes involved in the fibroblast growth factor (FGF) signaling pathway and identified the functional impairment in the FGFR1 gene in NSCLP families and suggested that the FGF signaling pathway may contribute to as much as 3 to 5% of NS cleft lip or palate. Xu et al reported mutations of the FGFR1 gene in Chinese Kallmann syndrome males with cleft lip/palate. TFAP2A located on 6p24 region has been associated with orofacial clefting. Davies et al reported a patient with cleft palate, microretrognathia, frontal bossing, hypertelorism, flat, broad nasal bridge, low set ears, and developmental delay. However, it was found that there was no significant association between TFAP2A and NSCLP in this northern Chinese and Indian population. S-glutathione transferase T1 (GSTT1) play an important role in the detoxification and secretion of smoking byproducts and deficiency of this enzyme may cause a greater risk of NSCLP if the individuals were exposed to smoking byproducts during pregnancy. Hozyasz et al suggested that homozygous deletion of GSTT1 in mother genome might increase the risk of having a child with NSCLP.
Further, mutations in the RYK gene were also found to be associated with orofacial clefting.\textsuperscript{110,111} Significant linkage disequilibrium between Gamma-aminobutyric acid receptor, Beta-3 (GABRB3), and CLP was reported by Scapoli et al and this finding in humans is in agreement with previously reported data obtained with the murine model.\textsuperscript{112} In addition, Baroni et al and Carter et al reported the association of GABRB3 with oral clefts in different populations.\textsuperscript{113,114} ATP-binding cassette, subfamily A, member 4 (ABCA4) encodes an ATP-binding cassette transporter. Several linkages and GWAS showed the role of ABCA4 in NSCLP with stronger evidence among Asian samples. ABCA4 is known to cause the autosomal-recessive retinal degenerative disease Stargardt's disease. A GWAS and a case-parent trio approach by Beaty et al identified ABCA4 gene association with oral clefts in different populations.\textsuperscript{115,116} Several other studies also reported a potential role of ABCA4 in the etiology of CL/P in Brazilian and northern Chinese Han population.\textsuperscript{117,118}

### Conclusion

Genetic studies provide insight into the multiple genes involved in the etiology of OFCs. Identification of specific genetic causes of clefts helps for better understanding of the molecular pathogenesis of OFC. In the near future, it helps to provide a more accurate diagnosis, genetic counseling, personalized medicine for better clinical care and prevention of OFCs.

### Conflict of Interest

None declared.

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