Exposure to polychlorinated biphenyl (PCB) congeners measured shortly after giving birth and subsequent risk of maternal breast cancer before age 50

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Abstract Discrete windows of susceptibility to toxicants have been identified for the breast, including in utero, puberty, pregnancy, and postpartum. We tested the hypothesis that polychlorinated biphenyls (PCBs) measured during the early postpartum predict increased risk of maternal breast cancer diagnosed before age 50. We analyzed archived early postpartum serum samples collected from 1959 to 1967, an average of 17 years before diagnosis (mean diagnosis age 43 years) for 16 PCB congeners in a nested case–control study in the Child Health and Development Studies cohort (N = 112 cases matched to controls on birth year). We used conditional logistic regression to adjust for lipids, race, year, lactation, and body mass. We observed strong breast cancer associations with three congeners. PCB 167 was associated with a lower risk (odds ratio (OR), 75th vs. 25th percentile = 0.2, 95 % confidence interval (95 % CI) 0.1, 0.8) as was PCB 187 (OR, 75th vs. 25th percentile = 0.4, 95 % CI 0.1, 1.1). In contrast, PCB 203 was associated with a sixfold increased risk (OR, 75th vs. 25th percentile = 6.3, 95 % CI 1.9, 21.7). The net association of PCB exposure, estimated by a post-hoc score, was nearly a threefold increase in risk (OR, 75th vs. 25th percentile = 2.8, 95 % CI 1.1, 7.1) among women with a higher proportion of PCB 203 in relation to the sum of PCBs 167 and 187. Postpartum PCB exposure likely also represents pregnancy exposure, and may predict increased risk for early breast cancer depending on the mixture that represents internal dose. It remains unclear whether individual differences in exposure, response to exposure, or both explain risk patterns observed.

Keywords PCBs · Polychlorinated biphenyls · Breast cancer · Pregnancy · Postpartum · Prospective · Human

Abbreviations
OR Odds ratio
PCB(s) Polychlorinated biphenyl(s)
95 % CI 95 percent confidence interval

Introduction

Discrete windows of susceptibility to toxicants have been identified for the breast, including in utero, during puberty, and during pregnancy and postpartum [1–4]. Human studies of breast cancer have not been able to assess the effects of measured exposure to environmental chemicals during windows of susceptibility for the breast [2, 5]. The existing literature on polychlorinated biphenyls (PCBs) and breast cancer typically has focused primarily on postmenopausal breast cancer [6–8]. In addition, exposure in prior studies was largely measured in middle age or at time of diagnosis, was variable on how PCBs were classified and
whether individual congeners were considered, and was based on samples obtained after PCB use was regulated [6–8]. The literature on the pesticide DDT and breast cancer is subject to similar limitations [9].

Findings on the relation of PCBs measured in middle age to breast cancer are complex and have been reviewed previously in detail (see Table 3, pp. 2872–2876 in Brody et al. [8]). Briefly, some studies reported positive associations with individual congeners, others reported no associations for total PCBs or any individual congeners, and others reported negative associations with some tumor types. The most consistent, positive findings were reported where interaction between exposure and CYP1A1 gene polymorphisms could be considered. Three studies found higher PCB exposures with postmenopausal breast cancer in combination with the CYP1A1 polymorphism, M2 type [10–12] and another reported this association with premenopausal breast cancer [13].

The present prospective study adds a unique perspective by testing the hypothesis that exposure to PCBs, measured during the early postpartum, is associated with increased risk of early breast cancer. Because of the long half-life of PCBs [14], early postpartum levels also reflect exposure during pregnancy [15] and possibly during childhood and adolescence. To our knowledge, this is the first study to report on breast cancer associations in relation to measured PCBs levels during critical periods of vulnerability for the breast.

Materials and methods

Subjects

Subjects were participants in the Child Health and Development Studies (CHDS), residents of the Oakland, California area and members of the Kaiser Permanente Health Plan who sought obstetric care between 1959 and 1967 [16]. Subjects voluntarily participated in the CHDS, giving an oral informed consent for an in-person interview, collection of blood specimens at several points in pregnancy and the early postpartum, and permission for medical record access. This study was reviewed and approved by The Institutional Review Board of the Public Health Institute and we have complied with all federal guidelines governing use of human participants.

Breast cancer cases were identified by linkage to the California Cancer Registry, and the California Vital Status Records [17]. All names for each CHDS subject are submitted for cancer linkages using fixed (i.e. birth date, sex, race, and name) and changeable (i.e. address and patient record number) identifiers. A rigorous protocol is used to verify cases, comparing fixed versus changeable identifiers by manual review. The California Cancer Registry is reported to be >99 % complete after a lag time of about 2 years [18].

Cases were defined as women with incident invasive or non-invasive breast cancer diagnosed before age 50, or deaths due to breast cancer before age 50, obtained from linkage conducted in early 1998. There were 133 cases who met study criteria. All members of the CHDS cohort are additionally linked to the California Department of Motor Vehicles (DMV) files on a regular basis to determine residence history allowing us to assess their control status and to update any name changes. All names registered with the DMV are used in establishing a match. Simultaneous linkage of multiple family members enhances matching. The regular DMV matching provides a history of location for each subject which is used to determine the population at risk for cancer, corresponding with geographic surveillance by California’s cancer registries. Subjects who cannot be located are considered lost to follow-up at the date of their last definitive classification as a California resident. One control, matched exactly on birth year, was selected at random for each case from those who were under cancer surveillance and known to be free of breast cancer at the age of diagnosis for the matching case. The median time to diagnosis for cases was 17 years. The mean age at diagnosis was 43 years.

Serum assays

Exposure to PCBs was measured by assays of serum samples drawn during the early postpartum period, within 1–3 days of delivery. Postpartum samples were used to conserve valuable archived serum samples drawn in each trimester for future studies where timed samples are essential. PCBs have a long half-life and prior work has established stability of organochlorine levels assayed across all trimesters of pregnancy and the early postpartum [15]. For this reason, postpartum levels of PCBs may accurately rank women on their pregnancy exposure, covering two potential vulnerable periods of susceptibility of the breast to toxicants.

PCBs were assayed in the laboratory of Dr. Mary Wolff [19] using modifications of methods developed previously by Brock et al. [20]. Briefly, a polar extraction of serum lipids is followed by a column chromatographic clean-up and enrichment step, with analysis by gas chromatography with electron capture detection. Limits of detection were approximately 0.07 ng/mL for individual compounds based on three times the standard deviation of the levels found in the lowest quality control plasma pool [21]. When the serum pool and blanks were considered together [22], the limit of detection was 0.01–0.1 ng/mL; the instrumental limit of detection based on a peak-to-noise ratio of
3 was 0.01–0.03 ng/mL for tetra- through hepta-chlorobiphenyls, using 1–1.5 mL plasma. As described previously [23], we used all observed positive values of PCBs in analyses, even those reported to be below the limit of detection.

We were able to assay archived serum samples for the PCB congeners which are shown in Table 1. These congeners include those most commonly reported in prior studies, comprised the congeners with highest concentrations, or grouped by potential biological activity as previously recommended [24]. We randomly assigned the order of samples within and across batches and analyzed case–control pairs in the same batches to minimize differences due to laboratory drift. The laboratory was blind as to case or control status of the samples. Intra-batch coefficients of variation ranged from 5 % for PCB 180 to 18 % for PCB 101. Total cholesterol and total triglycerides were measured enzymatically on the Hitachi 911 analyzer (Roche Diagnostics, Indianapolis, IN) in a lab certified by the Centers for Disease Control and Prevention and the National Heart Lung and Blood Institute Lipid Standardization Program.

### Statistical analysis

This report is based on 112 case–control pairs, matched on year of birth, after excluding 2 pairs with insufficient serum for lipid assays, 2 pairs with missing data on body mass index, and 17 pairs where one or more member of the pair was missing information on PCB 167, which was not quantified in one of the assay batches. In a sensitivity analysis, we imputed missing PCB 167 by matching on case status and date of blood draw and assigning a PCB value for the missing individual based on a random pick of all matches. Results were similar (associations were of a similar magnitude with overlapping 95 % confidence intervals and all associations remained statistically significant, defined as $p < 0.05$) whether or not the individuals with imputed values were included. We present findings only for individuals with non-missing data on all study

### Table 1 Distribution of PCBs in breast cancer controls and cases

| Classification [24] | PCB | Controls ($N = 117$) | Cases ($N = 123$) | Difference within matched pairs (case–control, $N = 112$) | Percent $> \text{LOD}$ per cent |
|---------------------|-----|----------------------|-------------------|----------------------------------------------------------|-------------------------------|
|                     |     | 25th 50th 75th       | 25th 50th 75th    | Mean (mmol/l)                                            | p value                       |
| Estrogenic          |     |                      |                   |                                                          |                               |
| 101                 |     | 0.37 0.74 1.07       | 0.34 0.55 0.92    | −0.06                                                    | 0.19                          |
| 187                 |     | 0.38 0.48 0.66       | 0.38 0.48 0.63    | −0.07                                                    | 0.30                          |
| 201                 |     | 0.09 0.23 0.33       | 0.14 0.23 0.30    | −0.01                                                    | 0.71                          |
| Anti-estrogenic     |     |                      |                   |                                                          |                               |
| A. Non-ortho, mono-ortho, dioxin-like | 66 | 0.89 1.34 1.78       | 0.96 1.47 1.95    | 0.10                                                     | 0.17                          |
|                     | 74  | 0.62 0.89 1.23       | 0.62 0.86 1.27    | 0.01                                                     | 0.54                          |
|                     | 105 | 0.25 0.40 0.64       | 0.25 0.43 0.61    | −0.01                                                    | 0.92                          |
|                     | 118 | 1.19 1.62 2.02       | 1.19 1.53 2.11    | −0.03                                                    | 0.94                          |
|                     | 156 | 0.11 0.28 0.42       | 0.11 0.28 0.39    | −0.03                                                    | 0.68                          |
|                     | 167 | 0.08 0.19 0.30       | 0.08 0.14 0.25    | −0.03                                                    | 0.12                          |
| B. Di-ortho, limited dioxin activity | 138 | 1.58 2.05 2.55       | 1.50 2.02 2.77    | −0.09                                                    | 0.56                          |
|                     | 170 | 0.30 0.46 0.66       | 0.35 0.48 0.63    | −0.01                                                    | 0.74                          |
| Phenobarbital, CYP1A, and CYP2B inducers | 99 | 0.28 0.58 0.80       | 0.25 0.52 0.86    | −0.04                                                    | 0.30                          |
|                     | 153 | 1.88 2.38 3.08       | 1.86 2.43 3.08    | −0.08                                                    | 0.40                          |
|                     | 180 | 0.89 1.21 1.54       | 0.94 1.21 1.57    | 0.01                                                     | 0.63                          |
|                     | 183 | 0.13 0.23 0.35       | 0.10 0.23 0.33    | −0.03                                                    | 0.32                          |
|                     | 203 | 0.21 0.30 0.42       | 0.23 0.35 0.47    | 0.02                                                     | 0.20                          |
|                     | 203/167 + 187 | 0.34 0.43 0.52       | 0.38 0.50 0.63    | 0.09                                                     | NA                            |

Variable number of subjects in each group is due to missing data on one or more of the PCB congeners shown. The PCB score ($\text{PCB 203/(PCB 187 + PCB 167)}$) was based on the best fitting model as described in the text, and is provided to describe the distribution of the mixture of significant PCB predictors. $p$ values are not presented for the PCB score because the score was created after analysis, as described in text. $p$ values shown are for Wilcoxon Signed Rank Test of differences (case–control) within age-matched case–control pairs.

LOD limit of detection
variables, as this is a conservative choice based on observed values.

We performed data analysis using age-matched conditional logistic regression as in our prior study of DDT and breast cancer in this same population [25]. We began with a full model, entering all PCBs classified by Wolff et al. [24], as potentially relevant to human health: Group 1 consisted of congeners detected in the CHDS serum samples that were considered to be potentially estrogenic and persistent (PCB 101, 187, 201). Group 2 consisted of congeners detected in the CHDS serum samples that were considered to be potentially antiestrogenic, immunotoxic, dioxin-like: Group 2A which are non-ortho or mono-ortho in their structure (PCB 66, 74, 105, 118, 156, 167) and Group 2B which are di-ortho and have more limited dioxin-like activity (PCB 138 and 170). Group 3 consisted of phenobarbital, CYP1A and CYP2B inducers (PCB 99, 153, 180, 183, 203). Congeners that had individual \( p > 0.20 \) were tested for removal as a group, based on a likelihood ratio test (\( p < 0.15 \) as the criterion). At the next step, we used a more stringent criterion, eliminating remaining PCB terms with individual significance probabilities >0.05. In addition to using the likelihood ratio test to test the hypothesis that the coefficients for these terms were each 0 using the criterion, \( p < 0.15 \), we also examined the sign and size of coefficients of remaining predictors before and after elimination to rule out major confounding by the eliminated predictors. Our goal was to identify the minimal number of PCBs that predicted risk. Once the best PCB model was identified, we examined whether further adjustment for blood lipids (total cholesterol, total triglycerides), parity, year of blood draw, body mass index (lower tertile, upper tertile vs. middle tertile as the reference category) and breast feeding following the current pregnancy altered PCB associations with breast cancer.

To describe the net effect of PCB exposure on breast cancer, we constructed a post-hoc score that consisted of the ratio of the sum of PCB congener(s) associated with higher risk of breast cancer to the sum PCB congeners associated with lower risk as in a previous report on health effects of PCB exposure in this population [26]. We then examined the variation in the post-hoc score and described its association with breast cancer in this population.

### Results

Table 1 shows the distribution of PCB congeners and a post-hoc PCB score based on the final model shown in Table 2. In this study population, PCB congener concentrations and the distribution of the PCB score were highly variable. The comparisons between cases and controls in Table 1 show no remarkable, statistically significant associations between single PCB congeners and breast cancer risk, in the absence of control for other congeners. This observation suggests that confounding would play a role in any associations observed in multivariate models. There were no associations between the sum of total PCBs or with PCB groups (groups shown in Table 1) and risk of breast cancer (data not shown). Table 2 shows results of multivariate conditional logistic regression models for individual PCB congeners. PCB 203 is the only congener with a consistent and statistically significant positive coefficient, indicating that it was associated with increased risk of breast cancer (Table 2, both models). In contrast, PCB 167 and PCB 187 were inversely associated with risk of breast cancer (Table 2, both models). To check for dose response, we also estimated associations for these PCBs by quartile shown in Table 3. Results were largely consistent with a monotonic trend for each congener.

### Table 2: Associations of individual PCB congeners with breast cancer diagnosed before 50 years of age

| Classification | PCB Coefficient | Model with all PCBs Coefficient | Final model Coefficient |
|----------------|-----------------|---------------------------------|-------------------------|
| Estrogenic     | 101 0.39 0.41   | 187 −4.18 0.03                  | −2.07 0.02 4.36 0.001   |
| Anti-estrogenic|                 |                                 |                         |
| A. Non-ortho, mono-ortho, dioxin-like | 66 0.11 0.61 | 74 0.36 0.53 | 105 1.37 0.19 | 118 −0.82 0.07 | 156 −0.42 0.61 | 167 −2.50 0.17 | 138 0.10 0.87 | 170 −1.45 0.34 |
| B. Di-ortho, limited dioxin activity | 99 −0.45 0.56 | 153 0.10 0.91 | 180 1.26 0.28 | 183 1.15 0.48 | 203 6.17 0.01 | 4.36 0.001 |

In the column labeled, “Model with all PCBs”, congeners were entered into a single model as the first step in model selection. Congeners PCB 203, 187, 167, 118, and 105 were initially retained based on the criterion of \( p < 0.20 \). PCB 118 and 105 were then removed either one at a time or together. They were deleted from the final model by applying the likelihood ratio test where \( \chi^2 = 3.81, 2df, p = 0.15 \) for the test to retain both PCB 105 and PCB 118 in the model. Both were tested for removal simultaneously because deleting either PCB 118 or PCB 105 alone greatly affected the size of coefficient for the other. There were \( N = 112 \) age-matched case–control pairs for all models shown and all models tested. Coefficients are reported per 1 mmol/l.
Table 3  Associations of PCB 167, 187, and 203 with breast cancer diagnosed before 50 years of age

| PCB     | Quartile | Odds ratio (95 % CI) | p value for linear trend |
|---------|----------|----------------------|--------------------------|
| 167     | Q1       | 1.00                 | <0.04                    |
|         | Q2       | 1.09 (0.48, 2.47)    |                          |
|         | Q3       | 0.70 (0.27, 1.78)    |                          |
|         | Q4       | 0.24** (0.07, 0.79)  |                          |
| 187     | Q1       | 1.00                 | <0.02                    |
|         | Q2       | 0.94 (0.41, 2.17)    |                          |
|         | Q3       | 0.92 (0.36, 2.38)    |                          |
|         | Q4       | 0.35* (0.11, 1.14)   |                          |
| 203     | Q1       | 1.00                 | <0.001                   |
|         | Q2       | 1.21 (0.46, 3.18)    |                          |
|         | Q3       | 2.89** (0.98, 8.55)  |                          |
|         | Q4       | 6.34† (1.85, 21.73)  |                          |

Each of the three PCBs is coded as quartiles, based on the distribution in controls. Quartile 1 is the reference category and quartiles 2, 3, 4 are entered as dummy variables for each PCB shown. Associations shown are based on a single model where quartile terms for all PCBs are entered. P value for trend is estimated from a linear model where all three PCBs are entered as continuous variables. N = 112 case–control pairs

CI confidence interval, p significance probability

* p ≤ 0.10
** p ≤ 0.05
† p ≤ 0.01

Table 4  Estimated net effects of PCB exposure on risk of breast cancer before 50 years of age

| Level of adjustment | Quartile of PCB scorea | Odds ratio (95 % CI) |
|---------------------|------------------------|----------------------|
| Unadjusted          | Q1                     | 1.00                 |
|         | Q2                     | 1.26 (0.53, 3.00)    |
|         | Q3                     | 1.52 (0.64, 3.62)    |
|         | Q4                     | 3.01 (1.34, 6.78)    |
| Cholesterol and triglycerides | Q1       | 1.00                 |
|         | Q2                     | 1.23 (0.53, 3.07)    |
|         | Q3                     | 1.53 (0.64, 3.68)    |
|         | Q4                     | 3.09 (1.34, 7.16)    |
| Cholesterol, triglycerides, race, parity, lactation, body mass index, and year of blood sampling | Q1       | 1.00                 |
|         | Q2                     | 1.36 (0.53, 3.52)    |
|         | Q3                     | 1.78 (0.70, 4.55)    |
|         | Q4                     | 2.81 (1.11, 7.09)    |

CI confidence interval

a A post-hoc PCB score was defined to describe the net effect of PCB exposure in this study sample (described in text). PCB 203 was associated with increased risk, while PCBs 167 and 187 were associated with decreased risk (see final model, Table 2). Therefore, the PCB score was defined as the ratio of PCB 203 relative to the sum of PCBs 167 and 187: PCB 203/(PCB 167 + PCB 187). N = 112 age-matched case–control pairs for all models shown

Based on the results in Tables 2 and 3, a PCB score was constructed as the ratio of PCB 203 (positively related to risk) to the sum of PCBs 167 and 187 (each negatively related to risk). The PCB score was highly variable in the cohort, ranging from 8 % PCB 203 (compared to the sum of PCBs 187 and 167) to 131 % PCB 203. Overall for cases and controls combined, the median of the 4th quartile (0.67) was 2.3 times higher than the median of the 1st quartile (0.30). Among controls, the median of the 4th quartile of the PCB score (0.61) was 2.5 times higher than the median of the 1st quartile of the PCB score (0.25). Among cases, the median of the 4th quartile of the PCB score (0.71) was 2.2 times higher than the median of the 1st quartile of the PCB score (0.32).

Table 4 shows the net effect of exposure based on the proportion of PCB 203 compared to the sum of PCBs 167 and 187. Women in the top 25 % of the PCB score had nearly three times the risk of breast cancer as women in the bottom 25 % of the PCB score. Adjustment for lipids or other breast cancer risk factors had little effect on this result (Table 4), nor did adjustment for the denominator of the PCB score or adjustment for \( p.p'-DDT, o.p'-DDT, \) and \( p.p'-DDE \) (data not shown).

Figure 1 shows the actual distribution of the within-pair differences for the PCB score for cases versus controls in this study sample. In the majority of case–control pairs (62 %), the PCB score was higher among the woman who subsequently developed breast cancer. Pairs where the case had a higher PCB score also showed greater differences on the PCB score than pairs where the control had a higher score (seen in Fig. 1; compare the right side of the \( Y \)-axis (center axis) which shows pairs where cases within the pair had a higher PCB score to the left side of the \( Y \)-axis which shows pairs where controls had a higher PCB score). Figure 1 is consistent with the modeling results shown in Tables 2, 3 and 4.

Discussion

The net effect of PCB exposure in this study population was nearly a threefold increase in breast cancer risk among women who had a higher proportion of PCB 203 in relation to the sum of PCB 167 and PCB 187 (75th percentile vs. 25th percentile). These results are novel, but not inconsistent with the prior literature on PCBs and breast cancer.

Three comprehensive reviews concluded previously that human studies of PCBs and breast cancer, which measured exposure in midlife, had variable findings [6–8]. Most prior studies reported primarily on total PCBs, which are largely determined by the PCBs found in highest concentration in humans (PCB 153, PCB 138, PCB 118, and PCB 180). We too found no associations for total PCBs, high
endocrine disruption, including early life, when the breast might be more susceptible to cancer in young women, during windows of susceptibility during the three lower concentration congeners that predicted prior studies reported on the independent contribution of groupings previously proposed by Wolff et al. [24]. No PCB score \((N = 112\) age-matched case control pairs). Each point represents one case–control pair. The points on the right side of the y-axis (center axis) are positive values that represent pairs where the woman who developed breast cancer had a higher PCB score postpartum than her matched control. In a majority of pairs (62 \%), the woman who subsequently developed breast cancer had a higher PCB score. The differential for the PCB score was also greater for pairs where the case had a higher score than her matched control (compare points on the right of the y-axis to points on the left of the y-axis)

concentration PCBs or sums of PCBs in functional groupings previously proposed by Wolff et al. [24]. No prior studies reported on the independent contribution of the three lower concentration congeners that predicted breast cancer in this study.

It is notable that prior studies did not measure exposure in young women, during windows of susceptibility during early life, when the breast might be more susceptible to endocrine disruption, including in utero, puberty, pregnancy, or the postpartum [1–3]. Our ability to directly measure exposure during the early postpartum is a particular strength of this study. A recent meta-analysis of pregnancy-associated breast cancer outcomes found that women diagnosed with breast cancer in pregnancy, and particularly women diagnosed postpartum, had poorer survival [4], providing further evidence supporting the hypothesis that pregnancy and the postpartum period are vulnerable periods for the breast.

As PCBs are highly persistent, it is likely that postpartum levels also reflect pregnancy levels, as suggested by one longitudinal study that reported high correspondence between early postpartum levels and levels across all three trimesters of pregnancy [15]. As women were young at the time of blood collection, it is also possible that the early postpartum levels of PCBs reflect exposure even prior to pregnancy, possibly during puberty, as well. This might explain the strength of the association observed for PCB 203, a higher chlorinated compound, as compounds with this structure tend to have longer half-lives [14].

Other strengths of this study include prospective assessment of exposure an average of 17 years before diagnosis, simultaneous consideration of individual PCB congener effects, and the opportunity to observe a population during active exposure because blood samples were obtained before PCBs were restricted.

Our focus on breast cancer at a young age is an additional strength. Molecular studies strongly suggest that premenopausal breast cancer may not share the same features or risk factors as breast cancer diagnosed in middle age and older [27, 28]. Our findings could lead to better understanding about etiology, prevention, and treatment of early breast cancer, if the mechanisms for the associations we observed can be validated and investigated by experimental toxicology and molecular studies.

Our choice to estimate the net effect of exposure to observed PCB mixtures found in our study participants is an additional strength. While our approach will likely be improved upon as the methods for analyzing mixed exposures advances; here, we applied an empirical approach to describe more than individual congener associations. It is of interest that the protective associations for PCBs 187 and 167 did not overcome the stronger, deleterious association observed for PCB 203. There are several speculative explanations for this finding, the first being that PCB 203 is a particularly strong risk factor, as evidenced by its point estimate. Alternatively, as the higher chlorinated PCBs are eliminated more slowly, it is possible that postpartum levels of PCB 203 more accurately reflect exposure even earlier in life, including accumulations in utero, childhood and during puberty, periods of susceptibility for the breast in addition to pregnancy and postpartum.

Limitations of our study include the possibility of unmeasured confounding by other exposures. In particular, we were unable to measure dioxin exposure or activity, raising the possibility that this or other unmeasured confounders could have masked or accounted for the associations we report here. However, as we observed dose response, unmeasured confounding would have to follow the same pattern, making this alternative explanation of our findings less likely. It is also possible that host factors that influence the metabolism or selective excretion of various PCB congeners underlie the associations we observed. We used early postpartum samples to save valuable timed serum samples during pregnancy for other studies in the cohort. However, prior studies conducted in serum samples of the same age found good correspondence of these persistent organochlorines across all trimesters and the early postpartum [15], suggesting that postpartum PCB levels...
may also reflect pregnancy exposures. Still our study cannot establish the age or developmental period when PCB exposure was acquired, other than establishing that exposure preceded the mean age of blood collection (age 26 years). Storage of serum samples is unlikely to have biased study results, as all samples were similarly stored. Randomization of samples within and between batches and inclusion of controls and cases in the same batches minimized inter- and intra-batch laboratory error.

Interpretation is limited by a lack of understanding about the potential mechanism for PCB associations observed. The direct association between PCB 203 and early breast cancer was sizable and significant. However, the mode of action for PCB 203, classified as a phenobarbital (PB) inducer [24], is unknown. A PubMed search for “PCB 203” returned no citations, compared to 589 citations for “PCB 153,” for example. The Agency for Toxic Substances and Disease Registry report on the toxic effects of PCBs makes no specific mention of PCB 203 effects [29].

PCB 187 has been classified as potentially estrogenic [24], but it is unclear how postpartum or pregnancy exposure might be associated with a lower risk of breast cancer. This study provides little information regarding the validity of the classification of PCB 187 as estrogenic. In our previous report on the relation of prenatal PCB exposure to time to pregnancy in daughters in this same cohort [26], we found that a longer time to pregnancy in daughters was associated with prenatal exposure to PCB 187. The significant associations observed for breast cancer in mothers and time to pregnancy in daughters following prenatal exposure to PCB 187 deserves additional study, perhaps in experimental models or in vitro systems. It could be of interest to characterize PCB 187 effects in the case of low versus high endogenous levels of estrogen and during pregnancy and postpartum in particular.

We were unable to investigate gene/PCB interactions in this study. The absence of associations for some of the PCBs investigated might be explained by failure to identify susceptible sub-populations of women. Previously, associations were more consistently observed in sub-populations characterized by variant alleles for enzymes that metabolize PCBs (Cytochrome P4501A1 variants) [10–13]. We were also unable to characterize the receptor status of the breast tumors in our study. We cannot account for lactation in subsequent pregnancies. However, breast feeding following the current pregnancy did not predict breast cancer in this sample and was not a confounder of PCB associations. There was no correlation between breast feeding following the observed pregnancy and PCB 203, 187, or 167. These findings may be explained by the low frequency of long-term breast feeding in our cohort. Rates of lactation in this sample were low (34%) and among those who did breast feed, most (60%) breast fed for <4 months [25]. We suggest that it is unlikely that lactation in subsequent pregnancies explains our results, as lactation behavior is highly correlated among pregnancies [30], but this remains a possibility.

The variable distribution of PCB congeners observed in study subjects is to be expected as the distribution of congeners depends on source of exposure which is influenced by the chemical structure of each congener. Exposure also depends on the fate of the congener in the ecosystem which ultimately forms the source of human exposure and on the individual response to the exposure. All these factors contribute differently to the measured serum level [31]. In a previous report on in utero PCB exposure and daughter’s time to pregnancy, we also found considerable variability in the mixture of PCB congeners in CHDS mothers [26]. In the US, total PCB levels in adipose tissue declined steadily after 1972 when restrictions were implemented. A decline in PCBs was also observed in archived blood samples in Norway during the same period [32] and in human adipose tissue in the United States [33]. However, secular trends in the mix of congeners are unknown [33]. As the fate of individual congeners in the environment depends on their structure, environmental topography, and climate, and because individual characteristics may determine routes of exposure, and metabolic fate, it is unlikely that trends for individual congeners are the same over time, or within individuals, or across geographic areas [14]. There is little human data on this topic, but results of repeated blood sampling in a Danish cohort provides limited support for the concept that congener proportions vary over time: over a 5-year period (1976–1978 vs. 1981–1983) median concentration of total PCBs declined 11%, but median concentration of PCB 118 declined 34%, PCB 180 declined 4%, and PCB 153 declined 9% (adapted from Hoyer, et al., Table 1, p. 179) [34]. If congener mixture is the underlying risk factor, then we might expect different results for epidemiological studies, depending on place, time, age, and other individual characteristics that might alter external and internal dose to these compounds.

Given the variability of congener mixtures observed in our cohort, we speculate that an underlying host factor related to metabolism of these compounds might contribute to the PCB associations with breast cancer that we have observed in this study. These data do not allow us to determine whether PCB exposure would be necessary to trigger an effect, or whether some host factor might be sufficient to increase breast cancer risk. Our analysis does indicate that PCB associations observed in this study are independent of DDT associations previously observed in this cohort [25]. Mechanistic studies in experimental models, or in vitro are likely to be very important to explaining the associations we report here, and for
explaining the human breast cancer associations previously reported for CYP1A1 polymorphisms in relation to PCB exposure in middle-aged women [10–13].

In summary, in this study, the mixture of PCB congeners predicted the estimated effect of PCB exposure on risk of breast cancer. Overall, women in this study showed a variable distribution for the three PCB congeners that predicted breast cancer. Women with a high proportion of PCB 203 (top 25 % of the study population) relative to PCBs 167 and 187 had a nearly threefold increase in subsequent risk of breast cancer, compared to women with a lower proportion of PCB 203 (bottom 25 % of the study population). The relation of PCB exposure to breast cancer might be clarified by additional laboratory, experimental and human population studies that account for timing of exposure in relation to windows of susceptibility for the breast and for concomitant host factors. It is likely to be particularly important to study congener mixtures and individual response to multiple exposures. It remains unclear whether individual differences in exposure, response to exposure, or both explain risk patterns observed.

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Conflict of interest The authors declare that they have no conflict of interest.

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