Chromosome numbers in antlions (Myrmeleontidae) and owlflies (Ascalaphidae) (Insecta, Neuroptera)

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Abstract

A short review of main cytogenetic features of insects belonging to the sister neuropteran families Myrmeleontidae (antlions) and Ascalaphidae (owlflies) is presented, with a particular focus on their chromosome numbers and sex chromosome systems. Diploid male chromosome numbers are listed for 37 species, 21 genera from 9 subfamilies of the antlions as well as for seven species and five genera of the owlfly subfamily Ascalaphinae. The list includes data on five species whose karyotypes were studied in the present work. It is shown here that antlions and owlflies share a simple sex chromosome system XY/XX; a similar range of chromosome numbers, 2n = 14–26 and 2n = 18–22 respectively; and a peculiar distant pairing of sex chromosomes in male meiosis. Usually the karyotype is particularly stable within a genus but there are some exceptions in both families (in the genera Palpares and Libelloides respectively). The Myrmeleontidae and Ascalaphidae differ in their modal chromosome numbers. Most antlions exhibit 2n = 14 and 16, and Palparinae are the only subfamily characterized by higher numbers, 2n = 22, 24, and 26. The higher numbers, 2n = 20 and 22, are also found in owlflies. Since the Palparinae represent a basal phylogenetic lineage of the Myrmeleontidae, it is hypothesized that higher chromosome numbers are ancestral for antlions and were inherited from the common ancestor of Myrmeleontidae + Ascalaphidae. They were preserved in the Palparinae (Mymrmeleontidae), but changed via chromosomal fusions toward lower numbers in other subfamilies.

Keywords

Male chromosome numbers, sex chromosomes, distant pairing of sex chromosomes, lacewings, Myrmeleontoidea

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Introduction

Within the holometabolous (= Endopterygota) insect order Neuroptera (lacewings) including a total of 17 or 18 currently recognized families (Aspöck et al. 2012), the Myrmeleontidae (antlions) comprise the most species-rich and most widespread family, with over 1500 valid extant species in 191 genera (Stange 2004). The closely related Ascalaphidae (owlflies) are a moderately speciose neuropteran family encompassing approximately 400 valid extant species assigned to about 65 genera, with wide distributional range in tropical and temperate areas of the world (Sekimoto and Yoshizawa 2007).

The Myrmeleontidae and Ascalaphidae belong to the superfamily Myrmeleontoidea (suborder Myrmeleontiformia), together with another four extant families, Nemopteridae, Crocidae, Psychopsidae, and Nymphidae. Despite the controversial hypotheses on the interfamilial phylogenetic relationships within this group, different phylogenetic analyses based on morphological and genetic data provide almost universal support for the monophyly of Myrmeleontoidea and the sister relationship between Myrmeleontidae and Ascalaphidae (Stange 1994, Aspöck 2002, Haring and Aspöck 2004, Winterton et al. 2010, Aspöck et al. 2012). However, molecular analyses are not always concordant with the monophyly of these families (Winterton et al. 2010).

Within Myrmeleontidae, the higher-level classification is controversial (reviewed in Mansell 1999), with several authors proposing various taxonomic divisions at the subfamily, tribe and subtribe levels (e.g. Banks 1899, 1927, New 1985a, b, c, Stange 1994, 2004, Krivokhatsky 2011). In his recent monography on the world fauna of Myrmeleontidae, Stange (2004) recognized three subfamilies, Stilbopteryginae, Palparinae, and Myrmeleontinae, with 14 tribes and 191 genera. Myrmeleontidae were further classified by Krivokhatsky (2011) who subdivided the family into 12 subfamilies (Palparinae, Pseudimarinae, Stilbopteryginae, Dimarinae, Echthromyrmicininae, Dendroleontinae, Nemoleontinae, Glenurinae, Myrmecaelurinae, Acanthaclisinae, Brachynemurinae, and Myrmeleontinae), with 23 tribes.

The Ascalaphidae are poorly-understood and taxonomically weakly-elaborated family. It was extensively revised only by van der Weele (1908) and now it comprises at least three subfamilies, Schizophthalminae (now Ascalaphinae), Holophthalminae (now Haplogleniinae), and Albardiinae, with a total of 15 tribes. Two-thirds of the species are placed in the first subfamily, and the remaining species (approximately 90) are placed in the second one, whereas the third subfamily contains only one species (van der Weele 1908, Sekimoto and Yoshizawa 2007). To date, no wide-ranging modern phylogenetic analyses of higher ascalaphid relationships have been published (Fischer et al. 2006).

Mansell (1999: p. 3) pointed out that the antlions, “apart from their obvious biological significance, are ideal subjects for the study of insect behavior, physiology, biogeography and evolution, and consequently a group urgently warrants study and con-
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Although chromosomal investigations have a long history in systematics and evolutionary biology (White 1973, King 1993), and a large body of data has been accumulated for insects (e.g., butterflies: Lukhtanov 2014; beetles: Angus et al. 2013, Blackmon and Demuth 2014, 2015; true bugs: Papeschi and Bressa 2006, Kuznetsova et al. 2011; aphids: Gavrilov-Zimin et al. 2015; coccids: Gavrilov 2007; cicadas: Kuznetsova and Aguin-Pombo 2015; grasshoppers: Warchałowska-Śliwa et al. 2005, parasitic wasps: Gokhman 2009), both antlions and owlflies were largely ignored in this respect. Our present knowledge of their karyotypes is scarce and fragmentary, being completely confined to the number of chromosomes and, additionally, to the meiotic behavior of the sex chromosomes that is of a very peculiar type in many neuropteran groups (Naville and de Beaumont 1932, 1933, Hughes-Schrader 1969, 1975a, b, 1979, Nokkala 1986) including the Myrmeleontidae (Naville and de Beaumont 1932, 1933, Hughes-Schrader 1983). In the Myrmeleontidae and Ascalaphidae, chromosomal studies were initiated in the 1930s with the pioneering works of Oguma and Asana (1932), Naville and de Beaumont (1932, 1933, 1936), Ikeda and Kichijo (1935), Asana and Kichijo (1936), and Katayama (1939). Since that time only scarce chromosome studies were performed on the Myrmeleontidae (Hirai 1955a, b, Highes-Schrader 1983, Klok and Chown 1993) while no further work on the Ascalaphidae appeared except for the re-investigation of Ascalohybris subjacens (Walker, 1853) karyotype (Hirai 1955a, b: as Hybris Lefèbvre, 1842) earlier studied by Katayama (1939: as Hybris).

Thus, cytogenetic studies on the families Myrmeleontidae and Ascalaphidae virtually ceased a few decades ago. The latest checklist of chromosome numbers in antlions published by Klok and Chown (1993) suffers from many shortcomings including imperfect references, erroneous identifications, outdated species names and synonymy. In order to fill this gap, an updated and comprehensive checklist of chromosome numbers of antlions and owlflys is provided here by integrating the published data together with our latest unpublished results.

**Material and methods**

**Insects**

Four antlion species (only males), namely Palpares libelluloides, Distoleon tetragrammicus, Macronemurus bilineatus, Myrmecaelurus trigrammus, and male owlfly Buropsis hamatus, were used in the present study. The specimens were collected from May to October 2013 in the Republic of Dagestan (North-East Caucasus, Russia). The material was collected by G. Khabiev. Collection sites, sampling dates, and the numbers of studied males are given in Table 1. In the field, adult individuals were fixed in a solution of 96% alcohol and glacial acetic acid (3:1) and then stored at 4 °C until required.
Table 1. Material used.

| Taxon                      | Sampling locality and date of collection | No. of studied males |
|----------------------------|------------------------------------------|----------------------|
| Myrmeleontidae             |                                          |                      |
| Palparinae                 |                                          |                      |
| *Palpares libelluloides*   | Russia, Dagestan, near Makhachkala 43°00'00"N, 47°13'33"E; V.2013 | 2                    |
| Nemoleontinae              |                                          |                      |
| *Distoleon tetragrammicus* | Russia, Dagestan, near Makhachkala 43°00'29"N, 47°14'51"E VII.2013 | 1                    |
| *Macronemurus bilineatus*  | Russia, Dagestan, near Makhachkala 42°59'58"N, 47°13'30"E; VI.2013 | 7                    |
| Myrmecaelurinae            |                                          |                      |
| *Myrmecaelurus trigrammus* | Russia, Dagestan, near Makhachkala 43°01'26"N, 47°15'12"E; 42°57'19"N, 47°28'51"E; 42°58'07.2"N, 47°20'03"E; VI-VII.2013 | 23                   |
| Ascalaphidae               |                                          |                      |
| *Bubopsis hamatus*         | Dagestan, Gumbetovsky district, near Chirkata village; 42°47°53"N, 46°41'14"E; VII.2013 | 2                    |

Chromosome preparation

Air-dried preparations were made by macerating testicular follicles in a drop of 45% acetic acid on a glass microscope slide and squashing under a cover slip. The preparations were frozen using dry ice, the cover slips were removed with a razor blade, and the preparations were dehydrated in fresh fixative (3:1) for 20 min and air dried. Slides were first examined under a phase-contrast microscope to check for the availability of meiotic divisions and quality of chromosome spreads. Counts were based on samples of one to 23 individuals. The preparations and remains of the specimens are stored at the Department of Karyosystematics, Zoological Institute, RAS.

Chromosome staining

Meiotic chromosomes were stained using the Feulgen-Giemsa method developed by Grozeva and Nokkala (1996).

Microscopy and imaging

Chromosome preparations were analyzed under a Leica DM 4000B microscope with a 100x objective. Images were taken with a Leica DFC 345 FX camera using Leica Application Suite 3.7 software with an Image Overlay module.
Results

Only meiotic divisions in adult males were available for analysis during the present study. In five examined species belonging to the families Myrmeleontidae (four species) and Ascalaphidae (one species) (Table 1), as many as three different chromosome numbers were found. Males of *P. libelluloides* showed 12 autosomal bivalents and univalent X and Y chromosomes at spermatocyte metaphases I (MI) suggesting the diploid karyotype formula of this species is $2n = 26(24A + XY)$. Unfortunately, our method proved to be inappropriate for effective and reliable detection of the centromere positions in chromosomes and, hence, analysis of their morphology. Nonetheless, most autosomes were suggested to be one-armed, with at least one clear exception of a large pair of bi-armed submetacentric chromosomes (Fig. 1). Males of *Macronemurus bilineatus* and *Myrmecaelurus trigrammus* showed 7 autosomal bivalents and univalent X and Y chromosomes at spermatocyte MI suggesting the diploid karyotype formula is $2n = 16(14A + XY)$. Males of *D. tetragrammicus* and *B. hamatus* showed 8 autosomal bivalents and univalent X and Y chromosomes at spermatocyte MI suggesting the diploid karyotype formula is $2n = 18(16A + XY)$. In the four low-numbered species, the chromosomes seemed to be essentially bi-armed (Figs 2–5).

A peculiar feature of all the species was that at metaphase I, the univalent X and Y chromosomes were disposed on the opposite sides of the division spindle whereas autosomal bivalents showed a typical metaphase location on the equator of the nucleus (Figs 1–5). In each species, the behavior of sex chromosomes was traced in the meiotic nuclei throughout all stages and these data will be presented elsewhere.

The new findings and references to previous reports of chromosome numbers in Myrmeleontidae and Ascalaphidae are given in Table 2. The subfamilial and tribal classification of the Myrmeleontidae used in this paper follows Krivokhatsky (2011) and that of the Ascalaphidae follows van der Weele (1908).

Discussion

Chromosome numbers

In the Myrmeleontidae, with the original data presented here, karyotype data have been made available for 37 species and 21 genera in 9 out of 12 subfamilies accepted by Krivokhatsky (2011). Having regard to 1500 valid species and 191 valid genera in this family (Stange 2004), the proportion of the studied species and genera is approximately 2.5% and 11% respectively. The karyotypes (chromosome numbers and sex chromosome systems) are presently known for the subfamilies Palparinae (3 species/2 genera), Pseudimarinae (1/1), Dendroleontinae (2/2), Nemoleontinae (6/4), Glenurinae (2/2), Myrmeleontinae (11/3), Brachynemurinae (7/3), Myrmecaelurinae (2/1), and Acanthaclisinae (3/3). The family demonstrates a relatively high diversity of karyotypes, with diploid chromosome numbers (2n) of 37 studied species ranging
Table 2. Data on karyotypes in the Myrmeleontidae and the Ascalaphidae (Neuroptera: Myrmeleontoidea).

| No | Taxon                        | 2n (karyotype formula) | Sampling locality       | Reference                          |
|----|------------------------------|------------------------|-------------------------|------------------------------------|
| 1  | *Indopalpares pardus* (Rambur, 1842) | 24(22+XY)              | East India: Ahmedabad     | Oguma and Asana 1932 (as *Palpares* sp.) |
| 2  | *Palpares libelluloides* (Linnaeus, 1764) | 26(24+XY) 26(24+XY) | Switzerland: Geneve, France: Banyuls-sur-Mer | Naville and De Beaumont 1936 Present data |
| 3  | *Palpares sobrinus* Péringuey, 1911 | 22(20+XY)              | South Africa: Transvaal   | Klok and Chown 1993                |
| 4  | *Palparidius concinnus* Péringuey, 1910 | 18(16+XY)              | South Africa: Transvaal   | Klok and Chown 1993                |
| 5  | *Epacanthaclisis moiwana* (Okamoto, 1906) | 16(14+XX)              | Japan                    | Hirai 1955a, b                     |
| 6  | *Dendroleon jezoensis* Okamoto, 1910 | 16(14+XY)              | Japan                    | Hirai 1955 a, b                    |
| 7  | *Distoleon tetrogrammicus* (Fabricius, 1798) | 18(16+XY)              | Russia: Dagestan          | Present data                       |
| 8  | *Neuroleon* sp. ²             | 16(14+XY)              | Western India: Bombay     | Asana and Kichijo 1936             |
| 9  | *Macronemurus appendiculatus* (Latreille, 1807) | 16(14+XY)              | France: Banyuls-sur-Mer   | Naville and De Beaumont 1933       |
| 10 | *Macronemurus bilineatus* Brauer, 1868 | 16(14+XY)              | Russia: Dagestan          | Present data                       |
| 11 | *Macronemurus* sp.            | 16(14+XY)              | Western India: Bombay     | Asana and Kichijo 1936 (as *Macronemurus* sp.? |
| 12 | *Creoleon lugdunensis* (Villers, 1789) | 18(16+XY)              | France: Banyuls-sur-Mer   | Naville and De Beaumont 1936 (as *Creagris plumbea* Navás, 1928) |
| 13 | *Euptilon arizonensis* (Banks, 1935) | 16(14+XY)              | USA                      | Hughes-Schrader 1983 (as *Pammoleon arizonensis* Banks, 1935) |
| 14 | *Paraglenurus japonicus* (MacLachlan, 1867) | 16(14+XY)              | Japan                    | Hirai 1955a, b (as *Glenuroides japonicus* MacLachlan, 1867) |
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| No | Taxon                        | 2n (karyotype formula) | Sampling locality                      | Reference                                      |
|----|------------------------------|------------------------|----------------------------------------|-----------------------------------------------|
|    | Subfamily Myrmeleontinae Latreille, 1802 |                        |                                        |                                               |
|    | Tribe Myrmeleontini Latreille, 1802 |                        |                                        |                                               |
| 15 | Baliga micans (McLachlan, 1875) | 14(12+XY)              | Japan                                  | Hirai 1955a, b (as Hagenomyia micans McLachlan, 1875) |
| 16 | Baliga sagax (Walker, 1853)    | 14(12+XY)              | Western India: Bombay [Mumbai]         | Asana and Kichijo 1936 (as Myrmeleon sp., probably M. sagax Walker, 1853) |
| 17 | Euroleon nostras (Fourcroy, 1785) | 14(12+XY)              | Switzerland, Geneva                    | Naville and De Beaumont 1932, 1933 (as Myrmeleon europaeus McLachlan, 1873) |
| 18 | Myrmeleon akestris Banks, 1911 | 14(12+XY)              | South Africa: Transvaal                | Klok and Chown 1993                           |
| 19 | Myrmeleon californicus Banks, 1943 | 14(12+XY)              | USA                                    | Hughes-Schrader 1983                          |
| 20 | Myrmeleon exitialis Walker, 1853 | 14(12+XY)              | USA                                    | Hughes-Schrader 1983                          |
| 21 | Myrmeleon formicarius Linnaeus, 1767 | 14(12+XY)              | Western India: Bombay [Mumbai]         | Ikeda and Kichijo 1935, Hirai 1955a, b         |
| 22 | Myrmeleon hyalinus Olivier, 1811 | 14(12+XY)              | France: Corse                          | Naville and De Beaumont 1936 (as Morter hyalinus Olivier, 1811) |
| 23 | Myrmeleon immaculatus DeGeer, 1773 | 14(12+XY)              | USA                                    | Hughes-Schrader 1983                          |
| 24 | Myrmeleon mexicanus Banks, 1903 | 14(12+XY)              | USA                                    | Hughes-Schrader 1983                          |
| 25 | Myrmeleon obscurus Rambur, 1842 | 14(12+XY)              | South Africa: Transvaal                | Klok and Chown 1993                           |
|    | Subfamily Brachynemurinae Banks, 1927 |                        |                                        |                                               |
|    | Tribe Brachynemurini Banks, 1927 |                        |                                        |                                               |
| 26 | Brachynemurus hubbardi Currie, 1898 | 14(12+XY)              | USA                                    | Hughes-Schrader 1983                          |
| 27 | Brachynemurus mexicanus Banks, 1895 | 14(12+XY)              | USA                                    | Hughes-Schrader 1983                          |
| 28 | Clathroneuria coquillettii (Currie, 1898) | 14(12+XY)              | USA                                    | Hughes-Schrader 1983 (as Brachynemurus coquillettii Currie, 1898) |
| 29 | Clathroneuria schwarzi (Currie, 1903) | 14(12+XY)              | USA                                    | Hughes-Schrader, 1983 (as Brachynemurus schwarzi Currie, 1903) |
| 30 | Scotoeon dissimilis (Banks, 1903) | 16(14+XY)              | USA                                    | Hughes-Schrader 1983 (as Brachynemurus dissimilis Banks, 1903) |
| 31 | Scotoeon niger (Currie, 1898)    | 16(14+XY)              | USA                                    | Hughes-Schrader 1983 (as Brachynemurus niger Currie, 1898) |
| 32 | Scotoeon nigrilabris (Hagen, 1888) | 16(14+XY)              | USA                                    | Hughes-Schrader 1983 (as Brachynemurus nigrilabris Hagen, 1888) |
| No | Taxon | 2n (karyotype formula) | Sampling locality | Reference |
|----|-------|------------------------|-------------------|-----------|
| 33 | Myrmecaelurus sp. ¹ | 14(12+XY) | Western India: Bombay [Mumbai] | Asana and Kichijo 1936 (as *Myrmecaelurus sp.*, probably *M. acerbus* (Walker, 1853)) |
| 34 | Myrmecaelurus trigrammus (Pallas, 1771) | 16(14+XY) | Russia: Dagestan | Present data |
| 35 | Synclisis japonica (McLachlan, 1875) | 14(12+XY) | Western India: Bombay [Mumbai] | Japan Ikeda and Kichijo 1935, Hirai 1955a, b (as *Acanthaclisis japonica* Hagen, 1866) |
| 36 | Centroclisis brachygaster (Rambur, 1842) | 14(12+XY) | South Africa: Transvaal | Klok and Chown 1993 |
| 37 | Vella fallax (Rambur, 1842) | 14(12+XY) | USA | Hughes-Schrader 1983 |

**Subfamily Acanthaclisinae Navás, 1912**

| No | Taxon | 2n (karyotype formula) | Sampling locality | Reference |
|----|-------|------------------------|-------------------|-----------|
| 38 | Ascalohybris subjacens (Walker, 1853) | 22(20+XY) | Japan | Japan Katayama 1939 (as *Hybris subjacens* (Walker, 1853)), Hirai 1955a, b (as *Hybris subjacens*) |
| 39 | Glyptobasis dentifera (Westwood, 1847) | 22(20+XY) | Western India: Bombay [Mumbai] | Asana and Kichijo 1936 |

**Tribe Hybrisini Lefèbvre, 1842**

| No | Taxon | 2n (karyotype formula) | Sampling locality | Reference |
|----|-------|------------------------|-------------------|-----------|
| 40 | Libelloides corsicus Rambur, 1842 | 20 | France: Corse | Naville and De Beaumont 1936 (as *Ascalaphus ictericus corsicus* Rambur, 1842) |
| 41 | Libelloides coccajus (Denis & Schiffermüller, 1775) | 22(20+XY) | Switzerland: Geneva, Valais | Naville and De Beaumont 1933, 1936 (as *Ascalaphus libellaloides* Schäffer, 1763) |

**Tribe Encyposini McLachlan, 1871**

| No | Taxon | 2n (karyotype formula) | Sampling locality | Reference |
|----|-------|------------------------|-------------------|-----------|
| 42 | Libelloides longicornis (Linnaeus, 1764) | 22(20+XY) | Switzerland: Valais | Naville and De Beaumont 1936 (as *Ascalaphus longicornis* (Linnaeus, 1764)) |

**Tribe Ascalaphini Rambur, 1842**

| No | Taxon | 2n (karyotype formula) | Sampling locality | Reference |
|----|-------|------------------------|-------------------|-----------|
| 43 | Bubopsis hamatus (Klug in Ehrenberg, 1834) | 18(16+XY) | Russia: Dagestan | Present data |
| 44 | Ogcogaster segmentator (Westwood, 1847) | 22(20+XY) | Western India: Bombay [Mumbai] | Asana and Kichijo 1936 |

¹ Later described as *Palpares pardus asanai* Kuwayama, 1933 (Oguma and Asana 1932, Kuwayama 1933)
² Presence of these taxa in Bombay [Mumbai] is doubtful
³ Wrong identifications: all records of *Creoleon plumbeus* from West Europe actually belong to *C. lugdunensis* (Hölzel 1976, Krivokhatsky 2011)
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Figures 1–5. Meiotic (MI) karyotypes of antlions (1–4) and owlfies (5). 1 *Palpares libelluloides*, $n = 12AA+XY$ (2$n = 26$, XY) 2 *Distoleon tetragrammicus*, $n = 8AA+XY$ (2$n = 18$, XY) 3 *Myrmecaelurus trigrammus*, $n = 7AA+XY$ (2$n = 16$, XY) 4 *Macronemurus bilineatus*, $n = 7AA+XY$ (2$n = 16$, XY), 5 *Bubopsis hamatus*, $n = 8AA+XY$ (2$n = 18$, XY). Arrows point to X and Y sex chromosomes. Scale bars = 10 µm
from 14 to 26 including four intermediate counts, i.e. 16, 18, 22 and 24. The highest numbers, 26, 24 and 22, occur only in the subfamily Palparinae, in three species of the genera *Palpares* Rambur, 1842 and *Indopalpares* Insom & Carfi, 1988. Other numbers, 2n = 14, 16 and 18, are encountered in the remaining subfamilies. In the Pseudimariinae, the only studied species, *Palparidius concinnus*, exhibits the next highest number found in antlions, i.e. 2n = 18. In the Nemoleontinae, with the exception of *Distoleon tetragrammicus* and *Creoleon lugdunensis* displaying 2n = 18, all studied species, including three *Macronemurus* Costa, 1855 species, have karyotypes with 2n = 16. The latter value is also found in all studied Dendroleontinae and Glenurinae. The subfamilies Brachynemurinae, Myrmecaelurinae and Acanthaclisinae include species both with 2n = 16 and 2n = 14, whereas Myrmeleontinae show 2n = 14 in all the studied species. It is noteworthy that, with the exception of *Palpares* (but see below), all these genera do not show interspecific variation in the chromosome number. This is especially remarkable for those genera where several species have been studied, e.g. *Macronemurus* (Nemoleontinae) and *Scotoleon* Banks, 1913 (Brachynemurinae). In each of these genera, three studied species share 2n = 16. Moreover, in *Myrmeleon* Linnaeus, 1767 (Myrmeleontinae) all eight studied species have 2n = 14. It is noteworthy that closely related genera, *Baliga* Navás, 1912 and *Euroleon* Esben-Petersen, 1918 in the Myrmeleontini, show the same karyotype with 2n = 14. It is unclear at present whether the chromosome number varies within the genus *Palpares*. The highest chromosome number, 2n = 26, is found in *P. libelluloides*, the type species of the genus. *Palpares pardus asanai* Kuwayama, 1933 with 2n = 24 (Oguma and Asana 1932) is treated here as a member of *Indopalpares*. Additionally, there is a possibility that *Palpares sobrinus* with 2n = 22 (Klok and Chown 1993) represents in fact *Pseudopalpares sparsus* (McLachlan, 1867). Although few members of the Palparinae are studied at present, karyotypic differences between the genera of this subfamily probably occur.

Compared to the Myrmeleontidae, karyotypes of the Ascalaphidae are less studied. The chromosome numbers are currently known in only seven owlfly species from the genera *Ascalohybris* Sziraki, 1998, *Ocgogaster* Westwood, 1847, *Libelloides* Schaeffer, 1766, *Bubopsis* McLachlan, 1898, and *Glyptobasis* McLachlan, 1871, all presently classified within the subfamily Ascalaphinae. The species studied show relatively high chromosome numbers, i.e. 2n = 18 in *Bubopsis hamatus*, 20 in *Libelloides corsicus*, and 22 in all the remaining species, including two other studied members of the genus *Libelloides*.

Although Myrmeleontidae and Ascalaphidae show a similar range of chromosome numbers (2n = 14 - 26 in the former and 18 - 22 in the latter), these families differ in the modal numbers. Of 37 species studied in the Myrmeleontidae, 19 species display 2n = 14, and 12 species have 2n = 16. On the other hand, five of seven species studied in the Ascalaphidae display 2n = 22. Other chromosome numbers occur only occasionally within the families except for high numbers characteristic of the antlion subfamily Palparinae.

In different eukaryotic organisms, evolutionary changes in the chromosome number happen via polyploidy, aneuploidy or fusion/fission events. In animals polyploidy is known to be rare, whereas chromosomal fusions and fissions are common. As stated
above, most Myrmeleontidae possess lower chromosome numbers, \(2n = 14\) and \(2n = 16\), which are encountered in all subfamilies, with the only exception of the Palparinae. The latter is the only subfamily characterized by higher numbers, \(2n = 26, 24,\) and \(22\), and the higher number, \(2n = 18\), is also found in the only studied species of the related subfamily Pseudimarininae. The higher numbers, \(2n = 22, 20\) and \(18\), are also characteristic of the sister family Ascalaphidae. Since Palparinae represent a basal phylogenetic lineage of the Myrmeleontidae (Krivokhatsky 2011), it is hypothesized that higher chromosome numbers are ancestral for antlions. Most likely, the higher chromosome numbers were inherited from the common ancestor of Myrmeleontidae + Ascalaphidae. It was preserved in the subfamily Palparinae (Myrmeleontidae) but changed via chromosomal fusions toward lower numbers, \(2n = 18, 16\) and \(14\), in other subfamilies.

Knowledge of the chromosome morphology in the low-numbered and high-numbered chromosome complements would help in understanding the karyotype evolution in the Myrmeleontidae and Ascalaphidae and testing the above hypothesis. Unfortunately, despite several efforts to identify chromosomal morphology within particular karyotypes (e.g. Asana and Kichijo 1936, Hughes-Schrader 1983, present study), this important question remains unresolved. Special staining methods, e.g. C-banding, are therefore needed to identify the centromeric position in the chromosomes and thus their morphology. However, these techniques have never been used in neuropteran cytogenetics, and therefore this is the most serious objective in the chromosome research of antlions and owflies.

**Sex chromosome system**

All Myrmeleontidae and Ascalaphidae species, including those studied here, exhibit a simple sex chromosome system XY/XX, which is characteristic of the whole order Neuroptera (White 1973, Blackman 1995). Both antlions and owflies demonstrate a very peculiar behavior of sex chromosomes in males (Naville and de Beaumont 1933, 1936, Asana and Kichijo 1936, Hughes-Schrader 1983, Klok and Chown 1993, present paper). In spermatocyte meiosis of those insects, sex chromosomes take up positions at opposite halves of the meiotic spindle at metaphase I before segregating into the daughter spermatocytes. It means that the X and Y chromosomes get segregated to opposite poles of the spindle long before the autosomal half-bivalents disjoin at anaphase I and move to the poles. The same pattern, the so-called “distance pairing” of sex chromosomes first discovered by Naville and de Beaumont (1933) in antlions, is known to be characteristic of the related neuropteran families Chrysopidae, Mantispidae, Sisyridae, Osmylidae, and Hemerobiidae (Naville and de Beaumont 1936, Hughes-Schrader 1969, 1975b, 1980, Nokkala 1986) and probably of the order Neuroptera in general. The biological role of this unusual behavior of sex chromosomes is unclear. In any case, this mechanism observed in brown lacewings (Hemerobiidae) showed no significance for the regular segregation of the sex chromosomes in meiosis (Nokkala 1986).
The order Neuroptera belongs to the superorder Neuropterida, which comprises another two orders, namely, Raphidioptera with two extant families, Raphidiidae and Inocelliidae, and Megaloptera with two extant families, Corydalidae and Sialidae (Aspöck and Aspöck 2007). Interestingly, the Neuroptera share the “distance pairing” of sex chromosomes with Raphidioptera (Naville and de Beaumont 1936, Hughes-Schrader 1975a) but not with Megaloptera. In the latter group, all hitherto studied species, which belong to the single family Corydalidae, show another very specific “parachute-like” sex bivalent in spermatocyte meiosis (Hughes-Schrader 1980, Takeuchi et al. 2002). In this case, the X and Y chromosomes form a pseudo-bivalent that is situated together with the autosomes on the equator of the spindle and segregates synchronously with them at the first meiotic anaphase. This unique meiotic sex chromosome configuration called X_y_p (Smith 1950) is the well-known characteristic feature of the related order Coleoptera, and is encountered in almost all coleopteran families. Therefore X_y_p is considered ancestral for beetles (Smith 1950), at least for the suborder Polyphaga (Petitpierre 1987).

The variety and distribution of sex chromosome systems in different orders of the class Insecta have been comprehensively reviewed by Blackman (1995). The X(0) system was shown to predominate in the lower orders and is considered as ancestral condition for several major groups and for Insecta as a whole. The XY systems when occur are all derived from an X(0) one. The sex chromosome systems seem to provide useful phylogenetic evidence. Within Holometabola orders, besides simple X(0) and XY, there are some peculiar systems, e.g., those involving female heterogamety (XY/XX or ZW/ZZ) shared by Lepidoptera and Trichoptera, haplodiploid sex determination characteristic of Hymenoptera, and some others. Of these, distance pairing of the X and Y chromosomes in spermatocyte meiosis and the parachute X_y_p system are hypothesized to be synapomorphies respectively of the clade Neuroptera + Raphidioptera and of the clade Megaloptera + Coleoptera (Blackman 1995, Takeuchi et al. 2002).

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