Variability of \textit{nm23-H1}/NDPK-A expression in human lymphomas and its relation to tumour aggressiveness

DNT Aryee\textsuperscript{1}, I Simonitsch\textsuperscript{2}, I Mosberger\textsuperscript{2}, K Kos\textsuperscript{1}, G Mann\textsuperscript{1}, E Schlögl\textsuperscript{1}, U Pöttschger\textsuperscript{1}, H Gadner\textsuperscript{1}, T Radaszkiewicz\textsuperscript{2} and H Kovař\textsuperscript{1}

\textsuperscript{1}Children's Cancer Research Institute, St Anna Kinderspital, A-1090 Vienna, Austria; \textsuperscript{2}Institute of Clinical Pathology, University of Vienna, A-1090 Vienna, Austria.

Summary

The \textit{nm23-H1} gene is a putative metastasis-suppressor gene encoding a 17 kDa protein with nucleoside diphosphate kinase activity. Expression of \textit{nm23-H1}/NDPK-A correlates inversely with the metastasising potential of some human tumours and experimental animal cells. No \textit{nm23} expression studies exist for human malignant lymphomas so far. In this study, we examined \textit{nm23-H1} expression by Northern and immunohistochemical analysis in 106 primary lymphoma samples from patients with Hodgkin's disease (HD) (\(n=15\)), high-grade non-Hodgkin's lymphoma (NHL) from different lineages (\(n=71\)) and low-grade NHL (\(n=20\)). Both inter- and intra-subtype variations in \textit{nm23-H1}/NDPK-A expression levels were demonstrated by all disease subtypes. Besides this heterogeneity, a general trend towards highly malignant samples expressing higher \textit{nm23-H1}/NDPK-A levels than the low-grade lymphomas was observed. Both adult and childhood HD and high-grade NHL samples exhibited significantly higher NDPK-A expression than the low-grade NHL found only in adults. High \textit{nm23-H1}/NDPK-A levels in lymphoma samples did not always reflect proliferative activity of tumour cells as monitored by Ki-67 antigen staining. Fifty samples were further investigated for possible mutations in the \textit{nm23-H1} coding sequence by means of reverse transcriptase-polymerase chain reaction (RT–PCR) and single-strand conformation polymorphism (SSCP) analysis. No mutation was found by this screening. Our results suggest a role for \textit{nm23-H1} expression in the disease aggressiveness of lymphomas.

Keywords: \textit{nm23-H1}; non-Hodgkin's lymphoma; Hodgkin's disease; immunohistochemistry

The \textit{nm23} gene seems so far to be the most promising candidate for a gene with metastasis-suppressor function. It was originally identified in differential hybridisation experiments involving murine K-175S melanoma cell line clones of varying metastatic potential (Steeg \textit{et al.}, 1988). A tumour metastasis-suppressor function was implicated by the reduced expression of \textit{nm23} in highly metastatic sublines compared with non-metastatic sublines derived from the same K-175 clone (Steeg \textit{et al.}, 1988; Rosengard \textit{et al.}, 1989; Leone \textit{et al.}, 1991). Two closely related homologues of this gene, namely \textit{nm23-H1} and \textit{nm23-H2}, have been found, both of which map to the chromosomal locus 17q21.3 (Backer \textit{et al.}, 1993). They encode 18 kDa and 17 kDa proteins respectively, which have been demonstrated to have nucleoside diphosphate kinase (NDPK) activity (Biggs \textit{et al.}, 1990; Lacombe \textit{et al.}, 1990; Gilles \textit{et al.}, 1991). The \textit{nm23-H2} gene product was recently shown to be the \textit{c-myc} transcription factor PuF (Postel \textit{et al.}, 1993). The transfection of \textit{nm23} cDNA into low \textit{nm23}-expressing and highly metastatic murine melanoma and human breast cancer cell lines reduced their metastatic potential, independent of growth rate, and further stratified their suppressor role in these tumour cohorts (Leone \textit{et al.}, 1991, 1993a).

Down-regulation of the \textit{nm23-H1} gene expression due to mutation and its allelic deletion at the 17q21.3 chromosomal locus, which might abrogate its suppressor role, has been implicated in metastasis formation of some human tumour types (Bevilacqua \textit{et al.}, 1989; Cohn \textit{et al.}, 1991; Hennessy \textit{et al.}, 1991; Hirayama \textit{et al.}, 1991; Leone \textit{et al.}, 1991; Nakayama \textit{et al.}, 1992). In human breast carcinoma, reduced primary tumour \textit{nm23} expression has been shown to correlate with disease recurrence (Hirayama \textit{et al.}, 1991), significant reductions in survival (Barnes \textit{et al.}, 1991; Hennessy \textit{et al.}, 1991) and the presence of lymph node metastases in one of three reported studies (Bevilacqua \textit{et al.}, 1989; Sastre-Garau \textit{et al.}, 1992; Dawkins \textit{et al.}, 1993). This inverse correlation between low \textit{nm23-H1}/NDPK-A expression and some clinicopathological features has also been found in several other tumours studied, including human hepatocellular carcinoma (Nakayama \textit{et al.}, 1992) and melanoma (Florenes \textit{et al.}, 1992). In other cancer types, however, \textit{nm23-H1}/NDPK-A may be irrelevant to metastatic tumour progression, or it may be altered by means other than reduced expression. In childhood neuroblastoma, for example, \textit{nm23} mutations have been found, and aggressive N-myc-amplified stage III and IV tumours have been reported to express relatively high levels of Nm23-H1/NDPK-A (Leone \textit{et al.}, 1993b; Chang \textit{et al.}, 1994; Hailat \textit{et al.}, 1991). Similarly, higher Nm23-H1/NDPK-A expression levels have been correlated with high grade of malignancy in other neoplastic diseases, such as small-cell lung carcinomas (Engel \textit{et al.}, 1993) and prostate cancers (Igawa \textit{et al.}, 1994).

Malignant lymphomas, Hodgkin's disease (HD) and non-Hodgkin's lymphomas (NHL) constitute a heterogeneous group of disorders. NHL comprise tumours of different histogenesis and variable clinical outcomes. The indices most commonly used in the study of NHL, i.e. histological classification and staging, give an estimate of the average prognosis but do not accurately predict clinical outcome in individual cases. This is reflected within each of the major subgroups of this disease, i.e. low-grade and high-grade lymphomas, in which a great variation in the clinical pattern exists (Horning and Rosenberg, 1984; Gaynor and Ullmann, 1984). Consequently, there is a need for reproducible quantitative methods of tumour description to confer additional prognostic information and so guide the clinician in the selection of the most appropriate therapeutic approach.

The aim of this study was to determine in a large series of primary childhood and adult lymphomas the pattern of \textit{nm23-H1} expression as the basis for future evaluation as a probable prognostic indicator, and whether the \textit{nm23-H1} gene is mutated in some of these tumour subtypes. As it has been shown that low proliferative activity was associated with low-grade NHL while high proliferative activity was associated with high-grade NHL (Kath \textit{et al.}, 1995), we also checked whether there exists a correlation of \textit{nm23-H1}/NDPK-A expression to proliferative activity in the lympho-

Correspondence: DNT Aryee

Received 28 February 1996; revised 24 June 1996; accepted 4 July 1996
Table 1 Relative staining intensities [Mean values (range)] for nm23-H1/NDPK-A and Ki-67 antigen in lymphomas as tested on paraffin sections with MAb 37.6 (recognises only nm23-H1/NDPK-A) and MIB-1 (recognises the Ki-67 antigen)

| Lymphoma type* | n | All cases MAb 37.6 | MIB-1 | Cases aged 0.5–20 years (median 10 years) n | MAb 37.6 | MIB-1 |
|----------------|---|-------------------|-------|--------------------------------------------|---------|-------|
| Low-grade NHL  |   |                   |       |                                            |         |       |
| B-cell CLL     | 7 | 1.4(1–2)          | 1.0(1–1) | 2(1–3)                                    | 1.0(1–1) | 2(1–3) |
| Lymphoplasmacytoid | 4 | 2.5(1–3)         | 1.0(1–1) | 2.0(1–3)                                   | 2(1–3)  | 2(1–3) |
| Mantle cell    | 5 | 6.0(3–3)          | 1.0(1–1) | 2.0(2–3)                                   | 1.0(1–1) | 2(1–3) |
| FCL (diffuse)  | 1 | 2.0(2–3)          | 1.0(1–1) |                                            |         |       |
| FCL (folicular) | 3 | 2.0(2–3)          | 1.0(1–1) |                                            |         |       |
| High-grade NHL |   |                   |       |                                            |         |       |
| Diffuse LBCL   | 10| 3.1(3–4)          | 2.0(1–4) | 2(1–4)                                     | 3.0(3–3) | 3.0(2–4) |
| Precursor B-LL | 18| 2.7(1–4)          | 3.4(2–4) | 14(2–4)                                    | 2.5(1–4) | 4.2(4–4) |
| Burkitt's ALCL | 13| 3.4(3–4)          | 4.0(4–4) | 12(3–4)                                    | 3.3(2–4) | 4.0(4–4) |
| PTCL (LN, NOS) | 4 | 3.0(3–3)          | 2.5(3–4) | 4(3–4)                                     | 3.5(3–3) | 4(3–4)  |
| Precursor T-LL | 11| 3.0(3–3)          | 3.7(3–4) | 9(3–4)                                     | 3.0(3–3) | 3.7(4–3) |
| PTCL (LN, Mf/SS)| 4 | 3.3(3–4)          | 2.8(2–4) |                                            |         |       |
| PTCL (skin, NOS)| 5 | 2.6(2–3)         | 1.4(1–2) |                                            |         |       |
| PTCL (Mf/SS)   | 2 | 3.0(2–4)          | 2.0(1–3) |                                            |         |       |
| Hodgkin’s lymphomas | 2 | 3.0(3–3)          | 1.0(1–1) |                                            |         |       |
| Lymphocyte-rich | 2 | 3.0(3–3)          | 1.0(1–1) |                                            |         |       |
| Mixed cellularity | 4 | 3.5(3–4)          | 3.2(3–4) | 2(3–4)                                     | 3.5(3–4) | 3.0(2–4) |
| Lymphocyte-depleted | 2 | 3.0(3–3)          | 1.0(1–1) | 1(1–3)                                     | 3.0(3–3) | ND     |

*LN, lymph node; NOS, not otherwise specified; Mf/SS, mycosis fungoides/Sezary’s syndrome. n, number of samples. ND, not done.

Materials and methods

**Tissue samples**

Tumour tissue was collected from 106 patients between 1987 and 1993. The diagnosis of HD and NHL was established using standard morphological and immunohistochemical criteria. Low- and high-grade non-Hodgkin’s lymphomas were classified according to the Revised European American Classification of Lymphoid Neoplasms (REAL) classification (Harris et al., 1994). Low-grade NHL comprised small lymphocytic lymphomas, lymphoplasmacytoid, mantle cell lymphoma and follicle centre lymphoma (FCL). High-grade NHL comprised diffuse large B-cell lymphoma (diffuse LBCL), precursor B-lymphoblastic lymphoma (precursor B-LL), Burkitt’s lymphoma, anaaplastic large-cell lymphoma (ALCL) of T and null cell types and peripheral T-cell lymphoma (PTCL). Hodgkin’s disease consisted of lymphocyte-rich (LR), nodular sclerosis (NS), mixed cellularity (MC) and lymphocyte-depleted (LD) subtypes.

Childhood NHL patients were aged 0.5–20 years (median 10 years) while adult NHL patients were aged 23–85 years (median 61 years). Sixty-six of the patients were male and 40 female. Seventy-one patients presented with nodal disease and 35 had extranodal lymphoma. Tissue samples were subdivided for study and routine histopathological examinations. Study samples were snap-frozen in liquid nitrogen and stored at −70°C until processing.

**Immunohistochemical staining**

Formalin-fixed and paraffin-embedded tissue sections (3 μm) from 106 patients were stained for NDPK-A with the anti-nm23-H1/NDPK-A monoclonal antibody MAb 37.6 at a dilution of 1:100 (1 mg ml−1 stock) in 1 × phosphate-buffered saline (PBS). This antibody was generated as has been previously described (Aryee et al., 1995). It does not cross-react with NDPK-B on Western blot and showed the appropriate band at 18 kDa. The immunoreactivity was investigated by a three-step immunoperoxidase procedure (ABC-Elite, Vector, Burlingame, CA, USA) according to the manufacturer’s recommendations. Briefly, sections were deparaffinised and endogenous peroxidase activity was blocked using 0.3% hydrogen peroxide in methanol. After treatment with normal blocking serum, sections were incubated with the NDPK-A specific monoclonal antibody 37.6. Sections of normal lymph nodes and benign unspcific lymphadenitis served as staining controls. The peroxidase reaction used 0.02% 3,3′-diaminobenzidine tetrahydrochloride in the presence of hydrogen peroxide. Sections were counterstained with Mayer’s haemalum.

Staining results for lymphoma specimens were evaluated semiquantitatively by two observers blind to nm23-H1 RNA data, taking into account the percentage of NDPK-A-positive neoplastic cells (which always paralleled staining intensity as determined relative to adjacent non-tumorous lymphocytes or plasma cells), as (0) <5%, (1) 5–20%, (2) 20–50%, (3) 50–90% and (4) >90% positive-staining cells. Representative results of immunohistochemistry are shown in Figure 2 and summarised in Table I. The proliferative activity was evaluated using the Ki-67 specific monoclonal antibody MIB-1 (Dianova Hamburg, dia 505, dilution 1:10).

Statistical analyses were carried out using the non-parametric Kruskal–Wallis analysis of variance. A P-value less than 0.05 was considered to be statistically significant.

**Northern blot analysis**

Total RNA was extracted from primary snap-frozen lymphoma samples using the guanidinium isothiocyanate/phenol method according to Chomczynski et al. (1987). Five micrograms of total RNA was resolved by electrophoresis on 1.2% agarose–formaldehyde gel and transferred onto Hybond N membrane (Amersham, Aylesbury, UK) according to standard protocols. Prehybridisation and hybridisation reactions were performed at 42°C in 50% (w/v) formamide, 5 × standard saline citrate (SSC), 50 mm Tris-HCl, pH 7.5, 5 × Denhardt’s solution, 5% (w/v) sodium dodecyl sulphate (SDS), and 250 μg ml−1 denatured salmon sperm DNA and washed at a final stringency of 0.1 × SSC and 0.1% (w/v) SDS at 65°C. The blot was hybridised with a 32P-labelled cDNA fragment corresponding to a portion of the 3′-untranslated region specific for nm23-H1 (Stahl et al., 1991), and hybridisation was detected by autoradiography. For multiple hybridisations, the bound probe was removed.
by incubating the filter twice for 10 min in 0.1 x SSC and 0.1% SDS at 95–100°C. Quality and the comparable loading of RNA samples were confirmed by including ethidium bromide in the gels and by rehybridisation to β-actin cDNA respectively.

**PCR/SSCP analysis and DNA sequencing**

Three sets of primers were used separately for polymerase chain reaction (PCR) amplification of overlapping fragments from first-strand complementary DNA covering the entire coding sequence of nm23-H1. They were (1) nmS1: 5’-TCTCCGAAACCAGTGGTGC-3’ and nmA1: 5’-GGACGGCTTCTAGGTCAAC-3’ (2) nmS2: 5’-GCTTCCGAAGATCTTCTGAGA-3’ and nmA2: 5’-CCAGTTCCTCAAGGGTAAAC-3’ (3) nmS3: 5’-GCAAGGAGGAGATCGCTTGTG-3’ and nmA3: 5’-CAGATGGTCGGGAGATGTAAC-3’.

Primers were selected on the basis of limiting the fragment sizes to less than 300 bp for SSCP analysis. Polymerase chain reactions were done using 1 μl of the first-round cDNAs with the appropriate primers in the presence of 10 μl reaction volume as recommended by the manufacturer (Perkin Elmer Cetus, Norwalk, CT, USA). The following amplification conditions were used: 30 cycles of 94°C for 30 s, 62°C for 30 s and 72°C for 1 min, preceded by a 94°C, 10 min primary denaturation step and followed by a 72°C, 7 min final extension step. An aliquot of 1 μl of each product was subsequently diluted 1:20 with the loading buffer (95% formamide, 2 μM EDTA, 0.05% bromophenol blue, 0.05% xylene cyanol) and denatured at 90°C for 5 min. A 2 μl aliquot was loaded onto a 6% acrylamide non-denaturing sequencing gel in 89 mm Tris-borate, 2 mm EDTA, pH 8.3 and electrophoresed at 5 μA at 4°C at 35 W. Gels were subsequently dried and bands visualised by autoradiography. DNA sequencing was performed on PCR-amplified nm23-H1 cDNA with the same primers as described above using the DNA cycle sequencing system (BRL, Gaithersburg, MD, USA), according to the manufacturer’s instructions.

**Results**

**nm23-H1 mRNA expression and mutational analysis**

Nm23-H1 RNA steady-state levels in 48 samples were determined in comparison with β-actin and 18S rRNA by Northern blot analysis (Figure 1). Autoradiographs were semiquantitatively evaluated by densitometry. Representative data presented in Figure 1 indicate that, although both inter- and intra-subtype variations in band intensity for the nm23-H1 0.8 kb mRNA levels were observed, there was a trend towards higher expression levels in the high-grade NHL relative to the low-grade NHL. The nm23-H1 mRNA levels in some of the high-grade NHL were similar to that of the highly NDPK-A-expressing MCF-7 breast adenocarcinoma cells (Figure 1). The variation of nm23-H1 gene expression in the various lymphoma subtypes may have resulted from mutations in the coding region of the gene. To examine such a possibility, we screened for nm23-H1 gene mutations in 48 specimens by RT-PCR/SSCP analysis. None of the tumours showed aberrantly migrating bands in SSCP analysis with any of the primer sets used, which spanned the whole coding region. Twenty-four cDNA samples generated by PCR, representing the various lymphoma subtypes studied, were randomly chosen and the coding regions sequenced. None of these specimens harboured a mutation (data not shown).

**Immunohistochemical activity of NDPK-A**

Paraffin-embedded, formalin-fixed sections from 106 resected primary lymphomas, well preserved for immunohistochemical analysis, were studied for tumour cell reactivity with the NDPK-A specific antibody 37.6. Both inter- and intra-subtype variations in NDPK-A expression were exhibited in the immunohistochemical stainings (Figure 2), which corroborated the RNA data. Results of the immunohistochemical analyses are summarised in Table 1. Staining intensity of neoplastic cells was compared with non-neoplastic surrounding lymphoid tissue. Cells usually showed cytoplasmic staining. Additional nuclear staining was observed in high-grade lymphomas, i.e. LBCL and Burkitt’s lymphoma, ALL and in Hodgkin’s cells of mixed cellularity and NS types. In low-grade NHL, staining was restricted to the cytoplasm. Generally, low-grade malignant B-type NHL exhibited only few stained tumour cells; however, mantle cell lymphomas (Figure 2c) exhibited more positive tumour cells than the other low-grade lymphomas (i.e. lymphocytic lymphoma, Figure 2b; lymphoplasmacytoid lymphoma and follicle centre lymphomas). NDPK-A staining intensity was also higher in this group of lymphomas known to have a more aggressive clinical course. Within the rather heterogeneous group of precursor B-lymphoblastic lymphomas, we found low staining intensities in the very immature lymphomas, pre-pre-B and pre-B, whereas more mature lymphoblastic lymphomas presented with considerably more stained tumour cells and a higher staining intensity. Among the other high-grade malignant B-type NHL, almost all tumour cells exhibited strong expression of NDPK-A. The most prominent tumour positivity and staining intensity was observed in Burkitt’s lymphoma specimens (Figure 2d and Table 1).

In T-cell lymphomas, two tumour sites were investigated: nodal lymphomas and the clinically rather non-aggressive cutaneous lymphomas. In cutaneous T-cell lymphomas (CTCL) fewer immunoreactive cells were observed than in lymph node lymphomas with a similar morphology (e.g. pleomorphic CTCL contained usually lower fractions of
reactive cells than the same type of nodal lymphomas). In contrast to high-grade B-cell lymphomas, we did not find striking discrepancies in staining intensity and positivity of cells when looking at T-cell precursor lymphoblastic lymphomas (Table I). Anaplastic large-cell lymphomas (ALCL) behaved like other high-grade lymphomas, usually presenting with high rates of intensely stained tumour cells (Figure 2e). In lymphadenitis, which was used as control, MAb 37.6 reactivity was observed mainly within the cytoplasm of blasts in reactive enlarged germinal centres. (Figure 2a). Centrocytes and mantle cells were usually not stained. Centroblasts and, occasionally, starry-sky macrophages showed the highest reactivity. Within the T zones, single blastic T cells, plasma cells and histiocytes were weakly positive. Small lymphocytes were Mab 37.6 negative.

In Hodgkin’s lymphoma, nodular sclerosing (NS) and mixed cellularity (MC) subtypes showed marked positivity of the Hodgkin and Reed–Sternberg cells. These neoplastic cells had the most intense cytoplasmic staining (Figure 2f) when compared with the staining intensities of the different cellular components in high-grade NHL. Cells from the surrounding infiltrates such as lymphocytes or plasma cells showed weak staining (Figure 2f). In the lymphocyte-rich and, strikingly, lymphocyte-depleted subtypes, NDPK-A staining intensity of the neoplastic cells was lower than in both other subtypes of HD.

Children usually present with high-grade lymphomas of the B and T type or Hodgkin’s disease. They commonly do not present with low-grade lymphomas. Comparing the results between children (<20 years) and adult cases of each subtype, no differences were observed in either staining patterns or intensity (Table I).

**Comparison of the proliferative activity and NDPK-A immunostaining in lymphomas**

MIB-1 is a monoclonal antibody that recognises the cell proliferation marker Ki-67 in paraffin-embedded tissues. To determine whether NDPK-A expression correlates with the proliferative activity, we stained all lymphoma specimens with the MIB-1 monoclonal antibody. Figure 2 shows representative Ki-67 antigen immunoreactivities. There was no consistent correlation between NDPK-A activity and proliferative activity (Table I).

**Correlation between histological subtype and NDPK-A expression**

As exemplified in Figure 2 and summarised in Table I, there was considerable variation in NDPK-A staining intensities among the lymphoma types studied (non-parametric Kruskal–Wallis test; $\chi^2=20.481$, $P=0.0001$). Both high-grade NHL and Hodgkin’s lymphoma samples exhibited significantly higher NDPK-A expression levels than low-grade NHL (Wilcoxon’s test; $P=0.0002$ and $P=0.0001$ respectively). By contrast, there was no significant difference in NDPK-A expression between high-grade NHL and Hodgkin’s lymphoma samples (Wilcoxon’s test; $P=0.0529$).

**Discussion**

The relationship between nm23-H1/NDPK-A and tumour metastatic potential in various human tumours and experi-
mental model systems remains a contentious subject, with conflicting reports from different research groups. Our data demonstrate that nm23-H1/NDPK-A is expressed in varying amounts in all lymphomas irrespective of clinical presentation. In contrast, Engels et al. (1993) and Rosengard et al. (1994) could not detect expression of nm23-H1/NDPK-A in some other tumours, such as breast cancer (Blevilacqua et al., 1989; Barnes et al., 1991; Hennessy et al., 1991; Hirayama et al., 1991), in which high nm23-H1/NDPK-A expression correlates with non-aggressive disease. In this study, a high level of nm23-H1/NDPK-A expression was found in high-grade lymphomas, similar to the situation in Ewing tumours (Aryee et al., 1995), neuroblastomas (Hailat et al., 1991), lung carcinomas (Engel et al., 1993) and prostate cancers (Igawa et al., 1994). These contrasting results might indicate that nm23-H1 can function as a suppressor gene in some types of cancer and can be associated with tumour aggressiveness in others. This is in agreement with the situation in colorectal carcinomas, in which nm23-H1 mRNA expression is correlated with increasing colorectal cancer size and extent of local bowel invasion and therefore is speculated to be associated with local aggressive behaviour (Zeng et al., 1994). In neuroblastomas, in which mutations in the nm23-H1 gene have also been associated with advanced stages of the disease (Chang et al., 1994), no mutations have so far been found in any of the high-grade lymphomas analysed. Within the group of high-grade B-cell lymphomas, usually strong expression of NDPK-A was observed. In the heterogeneous group of precursor B lymphoblastic lymphomas, the mature types showed a relatively higher expression levels than the immature types (pre-pre-B and pre-B). These data might parallel the different clinical behaviour of these tumours as phenotypically mature lymphoblastic B-cell lymphomas require more aggressive and different chemotherapy regimens than the immature lymphomas.

The highest cytoplasmic staining intensity, even compared with the reactivity in high-grade NHL, was observed in HD. NDPK-A antigen was present in Hodgkin and Reed–Sternberg cells, whereas the accompanying plasma cells and lymphocytes showed only faint (if any) staining. In the classical forms of HD (NS, MC and LD), Hodgkin’s and Reed–Sternberg cells express activation markers such as CD30, whereas they usually do not express T- or B-cell markers. The high expression levels of NDPK-A by Hodgkin and Reed–Sternberg cells might be interpreted in different ways because of the still enigmatic origin of the Hodgkin and Reed–Sternberg cell. NDPK-A expression paralleled proliferative activity in nodular scleroses and in the mixed cellularity form of HD but not in the lymphocyte-depleted subtype, which is known to have the worst prognosis of all HD subtypes.

Acknowledgements

This work is dedicated to the memory of Professor Dr T Radaskiewicz. We appreciate the generosity of Drs Michel Veron, Francois Traincard and Ioan Lacu for the monoclonal antibody 37.6. We also thank Dr Patricia Steeg for the pm23-H1 and H2 clones. This work was supported in part by the Fonds zur Förderung der Wissenschaftlichen Forschung, grant no. P9238-MED and by the Österreichische Kinderkrebshilfe.

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