Hypoxia-Related Radiomics and Immunotherapy Response: A Multicohort Study of Non-Small Cell Lung Cancer

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Abstract

Background: Immunotherapy yields survival benefit for some advanced stage non-small cell lung cancer (NSCLC) patients. Because highly predictive biomarkers of immunotherapy response are an unmet clinical need, we used pretreatment radiomics and clinical data to train and validate a parsimonious model associated with survival outcomes among NSCLC patients treated with immunotherapy. Methods: Three cohorts of NSCLC patients treated with immunotherapy were analyzed: training (n = 180), validation 1 (n = 90), and validation 2 (n = 62). The most informative clinical and radiomic features were subjected to decision tree analysis, which stratified patients into risk groups of low, moderate, high, and very high risk of death after initiation of immunotherapy. All statistical tests were 2-sided. Results: The very high-risk group was associated with extremely poor overall survival (OS) in validation cohorts 1 (hazard ratio [HR] = 5.35, 95% confidence interval [CI] = 2.14 to 13.36; 1-year OS = 11.1%, 95% CI = 1.9% to 29.8%; 3-year OS = 0%) and 2 (HR = 13.81, 95% CI = 47.6%, 95% CI = 18.2% to 72.4%; 3-year OS = 0%) when compared with the low-risk group (HR = 1.00) in validation cohorts 1 (1-year OS = 85.0%, 95% CI = 60.4% to 94.9%: 3-year OS = 38.9%, 95% CI = 17.1% to 60.3%) and 2 (1-year OS = 80.2%, 95% CI = 40.3% to 94.8%; 3-year OS = 40.1%, 95% CI = 1.3% to 83.5%). The most informative radiomic feature, gray-level co-occurrence matrix (GLCM) inverse difference, was positively associated with hypoxia-related carbonic anhydrase 9 using gene-expression profiling and immunohistochemistry. Conclusion: Utilizing standard-of-care imaging and clinical data, we identified and validated a novel parsimonious model associated with survival outcomes among NSCLC patients treated with immunotherapy. Based on this model, clinicians can identify patients who are unlikely to respond to immunotherapy.

Checkpoint blockade immunotherapy demonstrates durable and long-term survival benefit in 20%-50% patients with advanced stage non-small cell lung cancer (NSCLC) (1-6). Because patterns of immunotherapy response (7) are complex, including rapid disease progression (8), hyperprogression (9), and acquired resistance (10), there is an urgent need to identify highly predictive biomarkers that can stratify patients into distinct risk groups of survival and progression. Tumor programmed cell death ligand-1 (PD-L1) expression by immunohistochemistry (IHC) is a standard-of-care biomarker; however, recent clinical trials demonstrated statistically significant survival benefit for patients irrespective of tumor PD-L1 expression (6,11). Although tumor mutational burden (TMB) (12) has also shown to be a predictor of immunotherapy response (13-15), tumor specimens have to be sufficient in both quantity and quality to assess TMB (14), and laboratory methods to calculate TMB can be timely and expensive. As such, complimentary biomarkers that are predictive, noninvasive, and measured in a timely fashion using standard-of-care modalities would have direct translational implications.

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Quantitative image-based features, or radiomics, reflect the underlying pathophysiology and tumor heterogeneity (16) and have advantages over tissue-based biomarkers as they can be rapidly extracted using standard-of-care medical images and can capture data from the entire region of interest (eg, tumor) rather than a small portion of the tumor that is biopsied and assayed. Therefore, in this study, we used pretreatment clinical data and radiomic features extracted from pretreatment computed tomography (CT) scans to develop a risk model that is associated with survival outcomes among NSCLC patients treated with immunotherapy. The biological underpinnings of the radiomics signature were assessed by gene expression and IHC analyses.

Methods

Immunotherapy-Treated Lung Cancer Patients

This study was approved by the University of South Florida institutional review board, and a waiver of informed consent was obtained. Three cohorts of NSCLC patients treated with immunotherapy were analyzed: training (Moffitt Cancer Center [MCC1], n = 180), validation 1 (MCC2, n = 90), and validation 2 (Veterans Health Administration [VA], n = 62). Patients in MCC1 and MCC2 cohorts were treated at the H. Lee Moffitt Cancer Center and Research Institute, Tampa, Florida, and patients in the VA cohort were treated at James A. Haley Veterans’ Hospital, Tampa, Florida. All patients were treated with PD-1 or PD-L1 therapy or combinations of PD-L1 or PD-1 with cytotoxic T-lymphocyte-associated protein 4 doublet therapy. Inclusion criteria included pretreatment CT or positron emission tomography-CT imaging acquired less than 90 days prior to initiation of immunotherapy and having at least 1 measurable lung lesion defined by Response Evaluation Criteria in Solid Tumors (RECIST) criteria. Additional details about the lung cancer patients and patient data are provided in the Supplementary Methods (available online).

Radiogenomics Dataset

A previously described (17) patient dataset of 103 surgically resected adenocarcinomas (Lung1) who had CT radiomics and tumor gene-expression profiling was used to identify biological underpinnings of the most informative radiomic feature; additional details are provided in the Supplementary Methods (available online).

Immunohistochemistry

To validate the radiogenomics data, IHC of CAIX was performed on 16 NSCLC patient samples that had a presurgical CT. The IHC staining was quantified by automated evaluation of positive staining percentage defined by the Aperio ImageScope (http://www.leicabiosystems.com) and by H-scoring metric (18) quantified by a board-certified pathologist (JS) who was masked to radiomics and automated IHC scoring data.

Prognostic Datasets

The most informative radiomics was analyzed in 4 independent datasets of NSCLC patients not treated with immunotherapy. Overall survival (OS) was the dependent variable. The first dataset (Lung1) was the radiogenomics dataset described above. The second dataset (Lung2) included 234 patients diagnosed with incidental lung cancer from the National Lung Screening Trial (16,19). The third dataset (Lung3) included 62 patients who had presurgical CTs and underwent resection at MCC (20,21). The fourth dataset (Lung4) (20,21) included 47 patients who had presurgical CT scans and underwent surgical resection at the Maastricht Radiation Oncology Clinic (MAASTRO).

Radiomic Feature Extraction

Figure 1 is an overview of our radiomics pipeline. Pretreatment thoracic CT scans were retrieved from the picture archiving and communication system and loaded into the HealthMyne software (http://www.healthmyne.com) for segmentation. Using the HealthMyne software, 2 radiologists (YT and JQ) with more than 10 years of clinical experience initialized the 3D segmentation algorithm on the largest lung lesion. The radiologists either confirmed or edited the segmentations as necessary. The tumor masks were imported into in-house radiomic feature extraction toolboxes created in MATLAB 2015b (The Mathworks Inc, Natick, MA) and C++ (https://isocpp.org). A total of 213 radiomic features were extracted from the intratumoral (122 features) and the peritumoral (3 mm outside of tumor boundary) regions (91 features) using standardized algorithms defined by the Image Biomarker Standardization Initiative (22). Stable and reproducible radiomic features were identified using previously described methods (23); details are included in the Supplementary Methods (available online).

Statistical Analysis

Statistical analyses were performed using Stata/MP 14.2 (StataCorp LP, College Station, TX) and R Project for Statistical Computing version 3.4.3 (http://www.r-project.org/). All statistical tests were 2-sided, and a P value of less than .05 was considered statistically significant. Differences between cohorts for clinical covariates were tested using Fisher exact test for categorical variables and Mann-Whitney U test and Student’s t test for continuous variables. Survival analyses were performed using Cox regression, Kaplan-Meier survival estimates, and log-rank test. OS and progression-free survival (PFS) were the 2 dependent variables. Index date was the date of initiation of immunotherapy. For OS, an event was defined as death. For PFS, an event was defined as death or either clinical- or RECIST-based progression of cancer, and the data were right censored at 36 months.

Rigorous model building approaches were employed to reduce the number of variables and identify the most informative clinical covariates and radiomic features associated with OS in the training cohort (MCC1). The most informative clinical covariates and radiomic features that were identified in the training cohort were then tested in 2 validation cohorts (MCC2 and VA). For the clinical covariates, univariable Cox regression identified variables statistically significantly associated with OS. To generate a parsimonious clinical model, statistically significant clinical covariates from the univariable analyses were included in a stepwise backward elimination model using a threshold of P less than .01 for inclusion. For the radiomic features, we removed radiomic features correlated with tumor volume using a Pearson correlation coefficient of 0.80 or higher; among intra-correlated features using an absolute Pearson correlation coefficient of 0.80 or higher, the feature with the lowest P value in univariable analysis was retained, and the other(s) was
removed. Among the remaining features, univariable Cox regression identified those statistically significantly associated with OS. Among the remaining features, a parsimonious model was identified using stepwise backward elimination using a threshold of $P$ less than .01 for inclusion. Bonferroni-Holm method was used for multiple testing corrections.

The variables from the parsimonious clinical and radiomics models were used as inputs into a survival Classification and Regression Tree (CART) (24), which yielded risk groups associated with OS in MCC1 and then was validated in MCC2 and VA. Time-dependent area under the curve (AUC) values were calculated for 6, 12, 24, and 36 months.

Radiomics Quality Score (RQS)

RQS was calculated using an established metric that assesses the quality of a radiomics study (25). The RQS contains 16 items with a maximum score of 36.

Results

Patient Demographics

Type of checkpoint inhibitor, Eastern Cooperative Oncology Group (ECOG) performance status, number of previous lines of therapy, serum albumin, lymphocyte counts, and neutrophils to lymphocytes ratio were statistically significantly different between MCC1 and MCC2 (Table 1). Median age, sex, smoking status, stage, type of checkpoint inhibitor, ECOG performance status, lymphocyte counts, and neutrophils to lymphocytes ratio were statistically significantly different between MCC1 and VA.

The OS and PFS rates for all cohorts are presented in Supplementary Table 1 (available online), and the Kaplan-Meier survival curves are presented in Supplementary Figure 1 (available online). MCC1 and MCC2 had statistically significantly different OS (36-month OS = 32.6% vs 19.2%; $P < .001$) and PFS (36-month PFS = 20.8% vs 9.5%; $P = .003$). PFS was not available for the VA cohort.

Covariate Reduction

Among the 16 covariates (Table 1) considered for the clinical model, 5 covariates (serum albumin, number of metastatic sites, previous lines of therapy, white blood cell, neutrophil count) were statistically significant in univariable analysis in MCC1 (Supplementary Table 2, available online). The parsimonious clinical model included 2 clinical covariates that were statistically significantly associated with OS: serum albumin (hazard
Among the 213 intratumoral and peritumoral radiomic features, 67 were found to be stable and reproducible (Figure 2, A). Of 67 features, 8 were correlated with tumor volume and removed. Ten of the remaining features were statistically significantly associated with OS in univariable analysis (Supplementary Table 3, available online), and 2 features

### Table 1. Patient characteristics of the training and 2 validation cohorts

| Characteristics | Training MCC1 cohort (n = 180) | Validation MCC2 cohort (n = 90) | p<sup>b,c</sup> | Validation VA cohort (n = 62) | p<sup>b,d</sup> |
|----------------|-------------------------------|---------------------------------|----------------|-----------------------------|----------------|
| Age at initiation of treatment, No. (%) | | | | | |
| Dichotomized | | | | | |
| <65 y | 68 (37.8) | 37 (41.1) | .60 | 15 (24.2) | .06 |
| ≥65 y | 112 (62.2) | 53 (58.9) | | 47 (75.8) | |
| Median (95% CI), y | 67 (65 to 68) | 67 (64 to 69) | .78 | 68 (67 to 71) | .03 |
| Sex, No. (%) | | | | | |
| Female | 95 (52.8) | 43 (47.8) | | 3 (4.8) | |
| Male | 85 (47.2) | 47 (52.2) | | 59 (95.2) | <.001 |
| Smoking status, No. (%)<sup>d</sup> | | | | | |
| Never smoker | 30 (16.7) | 16 (17.8) | | 2 (3.2) | |
| Ever smoker | 146 (81.1) | 74 (82.2) | | 60 (96.8) | .004 |
| Unknown/Missing | 4 (2.2) | 0 (0) | | 0 (0) | |
| Stage, No. (%) | | | | | |
| III | 6 (3.3) | 4 (4.4) | | 13 (21.0) | |
| IV | 174 (96.7) | 86 (95.6) | | 49 (79.0) | <.001 |
| Histology, No. (%) | | | | | |
| Adenocarcinoma/others | 137 (76.1) | 71 (78.9) | | 43 (69.3) | |
| Squamous cell carcinoma | 43 (23.9) | 19 (21.1) | | 19 (30.7) | .14 |
| Checkpoint inhibitors, No. (%) | | | | | |
| Anti–PD-L1 | 48 (26.6) | 18 (20.0) | | 8 (12.9) | |
| Anti–PD-1 | 57 (31.7) | 69 (76.7) | | 54 (87.1) | |
| Doublet | 75 (41.7) | 3 (3.3) | <.001 | 0 (0) | <.001 |
| ECOG performance status, No. (%)<sup>e</sup> | | | | | |
| 0 | 39 (21.7) | 10 (11.1) | | 12 (19.4) | |
| 1 | 141 (78.3) | 67 (74.4) | | 39 (62.9) | |
| 2 | 0 (0) | 13 (14.4) | <.001 | 11 (17.7) | <.001 |
| Previous lines of therapy on current diagnosis, No. (%) | | | | | |
| None | 70 (43.9) | 21 (23.3) | | N/A | |
| 1 | 48 (26.7) | 47 (52.2) | | N/A | |
| ≥2 | 62 (34.4) | 22 (24.4) | <.001 | N/A | |
| Number of metastatic sites, No. (%) | | | | | |
| 1 | 82 (46.6) | 51 (56.7) | | 25 (40.3) | |
| ≥2 | 98 (54.4) | 39 (43.3) | .09 | 37 (59.7) | .55 |
| EGFR mutational status, No. (%)<sup>f</sup> | | | | | |
| Not detected | 107 (59.4) | 37 (41.1) | | N/A | |
| Detected | 25 (13.9) | 5 (5.6) | .36 | N/A | |
| Missing/Inconclusive | 48 (26.7) | 48 (53.3) | | N/A | |
| KRAS mutational status, No. (%)<sup>f</sup> | | | | | |
| Not detected | 61 (33.9) | 20 (22.2) | | N/A | |
| Detected | 29 (16.1) | 12 (13.3) | .66 | N/A | |
| Missing/Inconclusive | 90 (50.0) | 58 (64.4) | | N/A | |
| Hematology, median (95% CI) | | | | | |
| Serum albumin, g/dL | 4.0 (2.8 to 4.9) | 3.8 (2.3 to 4.7) | <.001 | 3.9 (2.9 to 4.6) | .09 |
| Lymphocytes, 1x10<sup>9</sup>/L | 1.3 (0.3 to 3.6) | 1.0 (0.3 to 3.4) | <.001 | 1.0 (0.2 to 3.2) | .01 |
| WBC, 1x10<sup>9</sup>/L | 7.1 (3.2 to 61.5) | 7.7 (1.4 to 45.1) | .25 | 7.5 (1.8 to 19.4) | .38 |
| Neutrophils, 1x10<sup>9</sup>/L | 4.8 (1.6 to 31.7) | 5.3 (0.4 to 40.2) | .13 | 5.6 (1.1 to 15.1) | .33 |
| Ratio of neutrophils: Lymphocytes | 3.7 (1.1 to 30.4) | 5.2 (0.5 to 53.1) | <.001 | 5.3 (0.8 to 34.3) | .004 |

<sup>a</sup>The majority of the MCC1 (95.3%) and the MCC2 cohorts (86.7%) were self-reported White race and majority of MCC1 cohort (97.0%) and the MCC2 cohort (88.9%) were self-reported non-Hispanic. For the VA cohort, racial and ethnicity data were not available. CI = confidence interval; ECOG = Eastern Cooperative Oncology Group; EGFR = estimated glomerular filtration rate; MCC = Moffitt Cancer Center; PD-L1 = programmed cell death ligand-1; VA = Veterans Health Administration; WBC = white blood cell; N/A = not available (i.e., these covariates were not available in the VA validation cohort).

<sup>b</sup>P values for continuous variables were calculated using Mann-Whitney test and Fisher exact test for categorical variables. All P values are 2-sided. P values were not calculated for N/A cells and an em dash was denoted.

<sup>c</sup>P values were calculated comparing MCC1 and MCC2 cohorts.

<sup>d</sup>P values were calculated comparing MCC1 and VA cohorts.

<sup>e</sup>P values for smoking status, EGFR mutational status, and KRAS mutational status were calculated excluding missing or inconclusive data.
remained after removing correlated features (Figure 2, B). Among the 2 remaining features (gray level co-occurrence matrix [GLCM] inverse difference and peritumoral quartile coefficient of dispersion), stepwise backward elimination identified a parsimonious model with 1 feature: GLCM inverse difference (HR $= 1.41$, 95% CI $= 1.19$ to 1.67).

**CART Analysis**

Using the 2 covariates from the parsimonious clinical model and 1 radiomic feature from the parsimonious radiomic model as inputs, CART analysis identified a novel tree structure (Figure 2, C) and grouped patients into 6 risk groups associated with OS (Supplementary Figure 2, available online). Of the original 6 groups, groups 2 and 3 were combined and groups 4 and 5 were combined as the hazard ratios for groups 2 and 3 and groups 4 and 5 were not statistically significantly different, resulting in 4 risk groups: low risk, moderate risk, high risk, and very high risk (Figure 3, A). The risk groups from the MCC1 cohort were also analyzed in both validation cohorts (MCC2 and VA; Figure 3, B and C). Similar findings were observed when the risk groups were analyzed for PFS for MCC1 and MCC2 (Figure 3, D and E). PFS data were not available for VA.

Time-dependent AUCs demonstrated that the final model (Supplementary Figure 3, A, available online) consistently had
higher AUCs compared with the clinical only model (Supplementary Figure 3, B, available online).

Multivariable Analyses

Multivariable Cox regression analyses demonstrate that the main effects for the risk groups were consistent when adjusting for clinical covariates (Table 1) that were statistically significant between MCC1 and MCC2. The multivariable models revealed the very high-risk group was associated with OS in validation cohort 1 (1-year OS = 40.1%, 95% CI = 1.3% to 83.5%; Table 2; Supplementary Table 4, available online). The results were similar for the CART risk groups (Supplementary Table 4, available online) for clinical covariates that were statistically significant between MCC1 and VA cohorts. Progressive-free survival (PFS) is presented for (A) MCC1 cohort, (B) MCC2 cohort, and (C) VA cohort. Progressive-free survival (PFS) is presented for (D) MCC1 cohort and (E) MCC2 cohort. PFS was not available for the VA cohort. Risk groups 2 and 3 were combined, and risk groups 4 and 5 were combined (as shown in Supplementary Figure 2, A, available online) to create the moderate-risk and the high-risk groups, respectively. The log-rank test was used to calculate two-sided P values. MCC = Moffitt Cancer Center; VA = Veterans Health Administration.

As noted above, 8 features were found to be correlated with tumor volume. Three (longest diameter, minor axis length, and least axis length) of the 8 features were statistically significantly associated with OS. However, the hazard ratios for these 3 features were not statistically significant when they were included in a multivariable model containing the most informative clinical covariates and radiomic features.

Other Model Building Approaches

Other model approaches were explored and yielded identical results (data not shown). First, all radiomic and clinical covariates that were statistically significant in univariable analyses (Supplementary Table 6, available online) were subjected to CART analysis. Second, we substituted least absolute shrinkage and selection operator for the stepwise backward approach, in combination with the CART approach. These extensive experiments demonstrated that our model is robust and reproducible by different modeling approaches.

Radiogenomics Analyses

The most informative radiomic feature was compared with every gene probe set using 2 different approaches: 2-group analysis and correlation analysis. For the 2-group analyses, GLCM inverse difference was dichotomized at the cut-point (0.43) determined by CART. Correlation and 2-group analyses identified 123 statistically significant probe sets representing 91 unique genes that were associated with the GLCM inverse difference radiomic feature (Supplementary Table 7, available online). Pathway analysis found no statistically significant enrichment after
Table 2. Univariable and multivariable Cox regression analysis for overall survival and progression-free survival

| Characteristics          | MCC1 cohort (n = 180) | MCC2 cohort (n = 90) |
|--------------------------|-----------------------|----------------------|
|                          | Univariable modela   | Multivariable modelb | Univariable modela | Multivariable modelb |
|                          | HR (95% CI)           | HR (95% CI)          | HR (95% CI)        | HR (95% CI)          |
| Overall survival         |                       |                      |                   |                      |
| Risk groupd              |                       |                      |                   |                      |
| Low risk                 | 1.00 (Referent)       | 1.00 (Referent)      | 1.00 (Referent)   | 1.00 (Referent)      |
| Moderate risk            | 3.79 (1.13 to 12.68)* | 3.08 (0.89 to 10.66) | 3.56 (1.02 to 12.48)* | 1.70 (0.75 to 3.87)* |
| High risk                | 8.02 (2.47 to 26.09)* | 7.87 (2.38 to 25.97)* | 6.98 (2.10 to 23.18)* | 2.73 (1.33 to 5.69)* |
| Very high risk           | 19.32 (5.80 to 64.32)*| 17.33 (5.11 to 58.72)* | 17.24 (5.09 to 58.36)* | 10.52 (4.58 to 24.17)* |
| ECOG                     | —                     | 1.22 (0.70 to 2.11)  | 1.20 (0.69 to 2.07) | —                     |
| Pr. treatment            | —                     | —                    | —                 | —                     |
| Lymphocytes              | —                     | 1.04 (0.74 to 1.46)  | 0.98 (0.88 to 1.09) | —                     |
| WBC                      | —                     | —                    | 1.10 (0.89 to 1.34) | —                     |
| Neutrophils              | —                     | —                    | —                 | —                     |
| NLR                      | —                     | 1.01 (0.97 to 1.06)  | 0.98 (0.92 to 1.05) | —                     |
| Progression-free survival|                       |                      |                   |                      |
| Risk groupd              |                       |                      |                   |                      |
| Low risk                 | 1.00 (Referent)       | 1.00 (Referent)      | 1.00 (Referent)   | 1.00 (Referent)      |
| Moderate risk            | 2.02 (0.89 to 4.64)   | 2.03 (0.88 to 4.76)  | 2.36 (1.00 to 5.58)* | 2.96 (1.43 to 6.14)* |
| High risk                | 5.15 (2.33 to 11.36)* | 5.55 (2.46 to 12.49)*| 4.89 (2.15 to 11.14)* | 2.58 (1.29 to 5.16)* |
| Very high risk           | 9.62 (4.12 to 22.44)* | 9.03 (3.77 to 21.63)*| 8.79 (3.66 to 21.11)* | 7.13 (3.31 to 15.35)* |
| ECOG                     | —                     | 1.09 (0.68 to 1.74)  | 1.05 (0.66 to 1.68) | —                     |
| Pr. treatment            | —                     | —                    | 1.32 (1.04 to 1.67)*| —                     |
| Lymphocytes              | —                     | 0.83 (0.63 to 1.09)  | —                 | 0.88 (0.59 to 1.33)  |
| WBC                      | —                     | —                    | 1.00 (0.90 to 1.11) | —                     |
| Neutrophils              | —                     | —                    | 1.04 (0.86 to 1.26) | —                     |
| NLR                      | —                     | 1.04 (0.99 to 1.09)  | 1.01 (0.95 to 1.07) | 1.05 (1.02 to 1.08)* |

aUnivariable model
bMultivariable model
cUnivariable model

dThe main effects for each risk group with the low-risk group as the referent category (ie, HR = 1.00). Hazard ratios were not calculated for the cells with the em dash.
CI = confidence interval; ECOG = Eastern Cooperative Oncology Group; HR = hazard ratio; MCC = Moffitt Cancer Center; NLR = neutrophils to lymphocytes ratio; PFS = progression-free survival; Pr. treatment = previous lines of treatments at current diagnosis; WBC = white blood cell.

The models included the clinical covariates that were found to be statistically significantly different between the MCC1 and MCC2 cohorts (Table 1). The low-risk group was the referent category.

These models included the clinical covariates that were found to be statistically significantly different between the Classification and Regression Tree risk groups in Supplemental Table 5 (available online).

Low-risk group refers to patients who have low gray level co-occurrence matrix (GLCM) inverse difference (<0.43) and lower number of metastatic sites (n = 1). The moderate-risk group refers to patients who have low GLCM inverse difference (<0.43) and higher number of metastatic sites (n = 1). The high-risk group refers to either patients who have higher GLCM inverse difference (>0.43), higher serum albumin (>3.9), and higher number of metastatic sites (n = 1). The very high-risk group refers to patients who have higher GLCM inverse difference (>0.43), lower serum albumin (<3.9), and lower number of metastatic sites (n = 1). The very high-risk group refers to patients who have higher GLCM inverse difference (>0.43), lower serum albumin (<3.9), and lower number of metastatic sites (n = 1). Hazard ratios are statistically significant.

The effects for each risk group with the low-risk group as the referent category (ie, HR = 1.00). Hazard ratios were not calculated for the cells with the em dash.

The models included the clinical covariates that were found to be statistically significantly different between the Classification and Regression Tree risk groups in Supplemental Table 5 (available online).

GLCM inverse difference was statistically significantly associated with OS in 3 independent NSCLC cohorts (Lung1, Lung2, and Lung3 in Figure 5) using the identified CART cut-point (0.43). Although this a priori cut-point was not statistically significant in Lung4, GLCM inverse difference as a continuous covariate was statistically significantly associated with OS in univariable analysis (HR = 2.74, 95% CI = 1.04 to 7.24). Hazard ratios were not calculated for the cells with the em dash.

This study yields a radiomic quality score of 22 out of a possible maximum of 36 (Supplementary Table 8, available online).

**Immunohistochemistry Analyses**

The automated pathology scoring was correlated with the pathologist-scored H-score (r = 0.629; P = 0.009). CAIX IHC by automated pathology scoring was statistically significantly different by GLCM inverse difference (Mann-Whitney U test P = 0.008; Figure 4, E), and CAIX IHC by automated pathology scoring was modestly correlated with GLCM inverse difference (r = 0.514; P = 0.04; Figure 4, F).
This radiomics pipeline to identify a parsimonious clinical-radiomic risk model that stratified into 4 risk groups based on risk of patient death (i.e., overall survival). The very high-risk group was associated with extremely poor OS and PFS in the training cohort and validated in internal and external cohorts (Figure 3, A-E). These results suggest very high-risk patients should either avoid immunotherapy altogether or utilize upfront combination treatments that may yield an improved response. The 4 risk groups identified were derived from a decision tree structure based on 1 radiomic feature (GLCM inverse difference) and 2 clinical covariates (baseline number of metastatic sites and serum albumin). The most informative radiomic feature, GLCM inverse difference, was positively associated with CAIX, which is related to tumor hypoxia.

We classify GLCM inverse difference as an “avatar” radiomic feature as it is correlated with 7 other radiomic features (Figure 2, B). Higher GLCM inverse difference, which is associated with poor survival, was found in dense and uniform lesions (Supplementary Figure 4, available online). Radiogenomics and IHC analyses revealed that the avatar feature was positively associated with CAIX expression, which is an important pH regulatory enzyme upregulated in hypoxic tumors leading to an acidic tumor microenvironment (26). Hypoxia is associated with angiogenesis, tumor growth, invasiveness, metastases, therapy resistance by inducing cell quiescence and an immunosuppressive phenotype (27), and poor survival in cancer patients including NSCLC (28–31). Because our results demonstrate that GLCM inverse difference is associated with tumor hypoxia, this biomarker should have wider reaching implications beyond immunotherapy. Thus, it is not surprising that GLCM inverse difference was associated with OS in the prognostic datasets suggesting this avatar feature could have broad clinical relevance beyond immunotherapy. Importantly, GLCM inverse difference was not associated with routinely obtained clinical covariates (other than ECOG; $P = .04$) implicating an independent effect (Supplementary Table 9, available online).

The most informative clinical covariates demonstrate the utility of standard-of-care information to predict patient outcome.
outcomes. A higher number of metastatic sites increase patient disease burden and may lead to mixed immunotherapy response, where some lesions are regressing and others are progressing, which ultimately leads to patient-level progressive disease. The other clinical covariate, serum albumin, has been shown to be associated with survival in NSCLC patients (32,33) and is used in existing cancer prognostic scores [ie, Royal Marsden Hospital prognostic score (34) and MD Anderson risk score (35)]. Low serum albumin is an indicator of malnutrition, inflammation, and hepatic dysfunction. The cut-point of 3.9 identified by CART was within the normal limits of serum albumin (lower limit = 3.5), which may suggest a potentially novel threshold for survival outcomes in immunotherapy.

Emerging evidence demonstrates the utility of radiomics as a noninvasive approach to predict lung cancer treatment response of tyrosine kinase inhibitors (36,37), platinum-based chemotherapy (36), neo-adjuvant chemoradiation (38,39), stereotactic body radiation therapy (40,41), and immunotherapy (8,42,43). We previously (8) demonstrated that pretreatment radiomic models predict rapid disease progression phenotypes, including hyperprogression (AUCs ranging 0.804-0.865) among 228 NSCLC patients treated with immunotherapy. Sun et al. (42) developed a radiomic signature that predicts CD8 cell infiltration and used the signature to predict immunotherapy outcomes among different cancers (AUC = 0.67); however, NSCLCs were only a small subset (22%) of the dataset. Utilizing 123 NSCLC patients treated with immunotherapy, Trebeschi et al. (43) developed a machine learning model that discriminates progressive disease from stable disease and responsive disease (AUC = 0.83). This current study represents one of the largest radiomics study of NSCLC patients treated with immunotherapy.

We do acknowledge some limitations of this study. TMB information was not available for any of the patients, and PD-L1 IHC was only available for 8 patients in MCC1 and 29 patients in MCC2. Thus, we are unable to verify the performance of TMB or PD-L1 expression related to outcomes. However, recent studies have shown that patients respond to immunotherapy regardless of PD-L1 expression (6,11), so inclusion of PD-L1 status may add little or no improvement to our models. Although MCC1 and MCC2 were curated from the same cancer center, the OS of the 2 cohorts were statistically significantly different (Supplementary Figure 1, available online). These differences in OS may be attributed to the MCC1 cohort comprised of clinical trial patients who typically have higher performance status, and the majority of patients in the MCC2 cohort were treated with standard-of-care immunotherapy. However, the hazard ratios were not appreciably modified in multivariable analysis adjusting for covariates that differed between the 2 cohorts. Lastly, clinical covariates were not available to validate the clinical-radiomic model for the prognostic datasets that were
not treated with immunotherapy. Despite these modest limitations, this study yields one of the highest radiomic quality scores of 22 (Supplementary Table 8, available online), which is a stringent metric that quantifies the quality of analysis and the potential clinical relevance of a radiomics study (25).

In conclusion, using standard-of-care imaging and clinical covariates, we identified and validated a novel parsimonious radiomic feature was found to be positively associated with survival outcomes among NSCLC patients treated with immunotherapy. The most informative radiomic feature was found to be positively associated with CAIX, an important enzyme upregulated in hypoxic and acidic tumors, which is related to treatment resistance. The potential clinical application of this work is that baseline radiomics and clinical covariates can identify patients who are unlikely to respond to immunotherapy and eliminate unnecessary treatments. Confirmation is required on the biological tumor-hypoxia underpinnings of GLCM inverse difference.

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**Data Availability**

The data and analysis code will be available upon reasonable request from the corresponding author (MBS). The MATLAB scripts to create peritumoral masks derived from intratumoral masks are publicly available (https://github.com/TunaliIlke/peritumoral_regions/). The radiomic features were extracted using algorithms from the IBSI (22). The algorithms were written in C++ and MATLAB v2015a, and these codes are available upon request from the corresponding author (MBS).

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