INTRODUCTION

Myotonic dystrophy type 1 (DM1) is a progressive muscular dystrophy with autosomal dominant inheritance. It has the highest prevalence among adult muscular dystrophies, which is estimated to be 9.13-10.4/100 000. The responsible gene is DMPK (19q13.3), and the mutation is an excessive extension of CTG repeats present in the untranslated region. Depending on the time of onset and...
severity, DM1 is classified into congenital, childhood, adult, and mild form (Table 1). The pathognomonic phenomenon is, known as “anticipation,” that when the affected allele is passed down from generation to generation, the mutation is further elongated as it is inherited; this causes the onset to occur at a younger age and follows the more severe clinical course in offspring than in parents. Based on the gravity of these conditions, preimplantation genetic testing (PGT) has been applied to couples with DM1.

PGT is a highly advanced technique for early prenatal diagnosis of embryo, which requires a series of complex procedures for the accurate diagnosis. As for the PGT for DM1, whole-genome amplification (WGA) is performed on the genomic DNA extracted through an embryo biopsy, and the WGA products are subjected to polymerase chain reaction (PCR) for the amplification of the nucleotide sequence of the CTG repeats region, which is followed by capillary electrophoresis using genetic analyzer with Genescan®. On the basis of the electrophoresis analysis on the sequence, the genetic status of the embryo is determined (direct diagnosis). On the principle of the Japan Society of Obstetrics and Gynecology (JSOG), PGT for monogenic disorder is performed based on the direct diagnosis for the disease responsible gene.

The challenges of PGT for DM1 are as follows:

1. because the gene mutation of DM1 is an excessive extension of CTG repeats, the binding between G and C nucleotides on complementary strands is likely to occur, and the gene amplification efficiency is likely to decrease;
2. in the gene amplification process, amplification errors such as allele dropout (ADO) will occur at a certain level of probability and pose a risk of misdiagnosis;
3. there is a limit to the base length which can be amplified by conventional PCR method, and the reaction fails to occur when the gene mutations cause CTG repeats extensions of 1000 or greater in base length. As a result, the affected alleles are difficult to detect on the capillary electrophoresis, and the analysis of affected embryos can detect only one peak from the unaffected allele;
4. the Southern blotting method, which is conventionally used for the diagnosis of the gene mutation is not capable for the analysis of rare DNAs (in extremely small quantities of a few picograms) extracted from a single cell;
5. in unaffected embryos as well, if the CTG repeats number on the alleles is identical or close to each other, the peaks in the capillary electrophoresis overlap (only one peak can be detected), and consequently, the analysis results is the same as those of embryos with ADO or affected embryos;
6. DM1 is an autosomal dominant inherited disorder, and embryos cannot be diagnosed as “transferable” unless the unaffected alleles are proven to be inherited from both the parents. In other words, the embryo diagnosed as “transferable” in PGT are those which shows two distinguishable peaks in the capillary electrophoresis.

We report the diagnostic outcomes of PGT for DM1 in our center and clarify the clinical problems in Japanese subjects.

## MATERIALS AND METHOD

From among couples in which either spouse was diagnosed with DM1, who opted for PGT and attended the Center for Medical Genetics (Keio University Hospital) for counseling between June 2006 and March 2017, our study was conducted on 32 couples who had received genetic counseling and provided consent to participate in the study. PGT was carried out on the couples who had been approved by the ethics committee of both our institution and JSOG; and 288 embryos retrieved from 17 couples over 43 PGT cycles were subjected for analysis.

Peripheral blood was collected from each couple, cells were lysed by adding an alkaline lyses buffer (50 mmol/L dithiothreitol/200 mmol/L NaOH), followed by 10 minutes incubation at 65°C. Gene amplification was performed on the extracted genomic DNA by conventional PCR method, using primers labeled with fluorescein (FAM). Primer design was as follows: forward primer 5’-(FAM)-GAA-CGG-GGC-TCG-AAG-GGT-CCT-TGT-AGC-3′, reverse primer 5’-CTT-CCC-AGG-CCT-GCA-GTT-TGC-CCA-TC-3’, and PCR conditions were as follows: 94°C for 2 minutes (denaturation), followed by 35 cycles under the condition of 94°C for 30 seconds, 72°C for 1 minute (amplification), and finally one cycle of 72°C for 5 minutes (final extension). For capillary electrophoresis of the amplification products, 3500 Genetic Analyzer (Applied Biosystems®) was used. The capillary length was 50 cm, the electrophoresis and size separation were carried out using a POP-6 polymer, and the results were analyzed using Genescan®. The number of expanded CTG repeats on the affected alleles of the couples and their referrals were analyzed using the Southern blotting method. To ensure the accuracy and safety of the diagnosis, 2 or more difference in CTG repeats number was considered as a criterion for determining whether CTG repeats number can be clearly distinguished on the results of the electrophoretic analysis. Based on the allele combination of the unaffected alleles, the couples were classified into three groups; “full

### TABLE 1 Clinical classification of myotonic dystrophy type 1

| Type          | CTG repeats | Clinical features                               |
|---------------|-------------|-------------------------------------------------|
| (unaffected)  | ≤37         | —                                               |
| Mild          | 50-150      | Weakness and cataract                           |
| Adult         | 100-1000    | Myotonia, muscle weakness, and cataract          |
| Childhood     | 500-2000    | Hypotonia and developmental disorder            |
| Congenital    | 1000-       | Hydramnios, respiratory distress, severe myotonia, and developmental disorder |
informative” who have no indistinguishable allele pattern, "semi informative” who have 1 indistinguishable pattern and “noninformative” who have no distinguishable allele pattern.9

Oocyte retrieval was conducted after controlled ovarian stimulation (COS), which were subjected to micro fertilization, and we conducted on biopsy of 3-5 trophoblast cells from blastocyst. Using the multiple displacement amplification (MDA) method, WGA was carried out on the genomic DNA extracted from each cell; the amplification products were subjected to conventional PCR under the conditions expressed above, and analyses were carried out using the capillary electrophoresis and Genescan®. The results of the electrophoretic analysis were classified according to whether the number of detected peak from the unaffected allele was 2, 1, or 0; and depending on the number, the embryo was diagnosed as “transferable,” “indeterminable,” or “inconclusive,” respectively.

Statistical analysis was carried out using a $\chi^2$ test or Fisher’s exact test. $P < .05$ was considered as the criterion for significance.

3 | RESULTS

The proportion of affected males and females in the 32 couples was as follows: 3 males and 29 females. The affected individuals’ mean age at diagnosis was 31.2(17-38) years; the number of affected individuals who had already developed clinical symptoms such as myotonia was 27; and the number of couples who had family history diagnosed with DM1 accounted for 17. A total of 14 couples had pregnancy and delivery history, and 18 couples had been attending a hospital for counseling before pregnancy.

In the analysis of the CTG repeats number on the unaffected alleles in the 96 chromosomes from the 64 individuals, the minimum number of CTG repeats was 3 and the maximum was 24. The frequency of CTG repeats on the unaffected alleles showed bimodal distribution; the peaks lied in (CTG)4-5 and (CTG)12-13 allele groups and the proportion of the allele groups was 22.9% and 43.7%, respectively (Figure 1). Meanwhile, the number of CTG repeats in the affected alleles ranged from 73 to 900, and the numbers showed no accumulation of specific repeats. The proportion of the “full informative,” “semi informative,” and “noninformative” couples was 46.9% (15/32 couples), 46.9% (15/32 couples), and 6.2% (2/32 couples), respectively. The PGT for the 2 “noninformative” couples had not been approved by our institution’s ethics committee.

In total, 288 embryos were collected from 17 couples during 43 PGT cycles. In the 17 couples, the number of the “full informative” couple was 9 (21 PGT cycles, 114 embryos) and the “semi informative” was 8 (22 PGT cycles, 174 embryos). The “transferable” “indeterminable” and “inconclusive” embryos accounted for 19.1% (55/288 embryos), 52.8% (152/288 embryos), and 28.1% (81/288 embryos), respectively. Analysis about the results of the “full informative” and “semi informative” couples showed that the proportion of the “transferable” embryos accounted for 28.9% (33/114 embryos) and 12.6% (22/174 embryos), and the “indeterminable” embryos were 40.3% (46/114 embryos) and 60.9% (106/174 embryos), respectively. There were significant differences in the results between the couples (Table 2, $\chi^2$ test, $P < .05$). Meanwhile, the “inconclusive” embryos accounted for 30.7% (35/114 embryos) and 26.4% (46/174 embryos) and there was no significant difference ($P < .05$).

**FIGURE 1**  Frequency distribution of the DM1 associated CTG repeats in the 32 couples. In the 96 unaffected alleles of the 32 couples, the allele frequency showed bimodal distribution. (CTG)12-13 alleles was the most common allele group.
The loss of isolated cell during transfer, cell degeneration or cell which incidence is reported as 5%-10% of single cell subjected to (AF) and ADO. AF is a phenomenon of total failure of amplification, to amplification errors of extracted DNA such as amplification failure couples, which values were lower than expected. This might be due tive" couples and 12.6% (22/174 embryos) in the "semi informative" embryos was 28.9% (33/114 embryos) in the "full informative" and "semi informative" couples (Table 3, Fisher's exact test, \( P < .05 \)).

Previous reports on the distribution of "unaffected" CTG repeats (≤37) have shown that, although there is interpopulation variation in the frequencies of CTG repeats, \((\text{CTG})_5\) allele and \((\text{CTG})_{11-14}\) allele group are major in all classes of CTG repeats arrays.\(^{10-12}\) It is also reported that there is a significant difference in the frequency of the major alleles between Japanese and European population, with a smaller frequency of \((\text{CTG})_5\) and greater frequency of \((\text{CTG})_{11-14}\) on Japanese subjects.\(^{10}\) On the other hand, there is no significant difference between Malay and Japan.\(^{11}\) In our study, the CTG repeats number on the unaffected alleles from the 32 couples showed similar distribution to previous reports (Table 3, Fisher’s exact test, \( P < .05 \)).\(^{10}\) Our data may imply the practical problem of PGT for DM1 in Japan, due to the allele combination.

As for the allele combination of the 32 couples, the proportion of "semi informative" and "noninformative" couples accounted for 53.1% (17/32 couples) and this result showed that the unaffected embryos with identical or close number of CTG repeats, diagnosed as "indeterminable" through the direct diagnosis, were easy to arise. Because DM1 follows an autosomal dominant pattern of inheritance, the incidence of unaffected embryos is theoretically 50%, but in practice, the "transferable" embryos accounted for only 19.0% (55/288 embryos), and the "indeterminable" embryos accounted for 52.7% (152/288 embryos), which was the largest percentage. However, considering the limitation of the direct diagnosis that cannot distinguish unaffected embryo from "indeterminable" embryo, the incidence of the "transferable" embryos in the "semi informative" couples decreases to 25%. As a result, the percentage of the "transferable" embryos was 28.9% (33/114 embryos) in the "full informative" couples and 12.6% (22/174 embryos) in the "semi informative" couples, which values were lower than expected. This might be due to amplification errors of extracted DNA such as amplification failure (AF) and ADO. AF is a phenomenon of total failure of amplification, which incidence is reported as 5%-10% of single cell subjected to PCR.\(^{7}\) The loss of isolated cell during transfer, cell degeneration or cell lysis failure are considered as causes of AF.\(^{7}\) On the other hand, ADO is a failure of PCR to amplify one allele of the cell. The incidence of ADO is reported as 5%-15%.\(^{7}\) ADO is explained by technical failures due to broken sequence of the targeted DNA during lysis of single cells, inefficient or no priming of the PCR primers, chromosomal mosaicism, and aneuploidy of embryos due to mitotic nondisjunction or anaphase lagging.\(^{8,13}\) Further, the proportion of 12.6% in the "semi informative" couples was approximately half the expected value, suggesting that they needed to be burdened with a larger number of oocyte retrieval in order to detect "transferable" embryos. In fact, couples from whom the "transferable" embryo could not be collected in one oocyte retrieval trial accounted for 11.1% (1/9 couples) in the "full informative" couples and 62.5% (5/8 couples) in the "semi informative" couples.

In addition, as for the two "noninformative" couples, the direct diagnosis theoretically cannot detect "transferable" embryos. This finding emphasizes the limitation of PGT for DM1. To compensate this limitation, the techniques using linkage analysis and triplet repeat primed PCR (TP-PCR) analysis have been applied to PGT for DM1. The linkage analysis, known as the indirect analysis, uses polymorphic microsatellite markers to detect not the gene mutation but the inherited haplotype. For the linkage analysis, both DNAs of parents-child trios and 2 or more informative markers are imperative because of genetic recombination.\(^{8,14}\) However, the informative markers are limited.

### Table 2: Comparison and \( \chi^2 \) analysis of PGT results from the “full informative” and “semi informative” couples

| Analysis result of PGT | Number of embryo (%) in “full informative” couples | Number of embryo (%) in “semi informative” couples | \( \chi^2 \) test |
|------------------------|-----------------------------------------------|-----------------------------------------------|------------------|
| Transferable           | 33 (28.9%)                                   | 22 (12.6%)                                   | \( P = 7.524e-8 \) |
| Indeterminable         | 46 (40.3%)                                   | 106 (60.9%)                                  | \( P = 1.705e-5 \) |
| Inconclusive           | 35 (30.7%)                                   | 46 (26.4%)                                   | \( P = .4619 \)   |
| Total                  | 114                                          | 174                                          |                  |

Note: There were significant differences in PGT analysis result, the number of "transferable" and "indeterminable" embryos, between the “full informative” and “semi informative” couples (\( P < .05 \)).

### Table 3: Comparison of the frequency of CTG repeats in unaffected alleles of Japanese from 1992 (Davies) and current study

| CTG repeats | Number of alleles from current study (%) | Number of alleles from Davies (%) |
|-------------|------------------------------------------|----------------------------------|
| 5           | 11 (11.4)                                | 21 (19.8)                        |
| 11          | 10 (10.4)                                | 9 (8.4)                          |
| 12          | 19 (19.7)                                | 25 (23.5)                        |
| 13          | 23 (23.9)                                | 29 (27.3)                        |
| 14          | 3 (3.1)                                  | 1 (0.9)                          |
| Others      | 30 (31.2)                                | 21 (19.8)                        |
| Total       | 96                                       | 106                              |

Note: In Davies’ study, 37 unaffected and 32 affected Japanese were subjected.\(^{10}\) Fisher’s exact test was performed to compare the frequency of CTG repeats in unaffected alleles and showed negative correlation (\( P < .05 \)).
in the PGT for DM1\textsuperscript{15-17} and, as mentioned above, PGT for monogenic disorder should be based on the direct diagnosis for the disease responsible gene. Analysis by the indirect diagnosis is supportive but not used as basis of diagnosis. TP-PCR is a PCR technique using multiple primers to detect expanded allele qualitatively. This technique is reported to be efficient in screening affected embryos in PGT for DM1\textsuperscript{9} but it is inevitable ADO will occur at a certain level of probability resulting in the false-negative result, which leads to misdiagnosis. The haplotype analysis using single nucleotide polymorphism (SNP) microarray could have possibility of new diagnostic method, but the accuracy, safety, and limitation should be validated.\textsuperscript{18}

In our study, the frequency of the CTG repeats in the unaffected alleles in 32 Japanese couples showed bimodal distribution, which was consistent of previous reports. The "semi/noninformative" couples accounted for approximately half of the couples, and the allele combination had significant impact on the diagnostic outcomes of PGT for DM1. This might contribute to the differences in terms of PGT-related burden weighing on each couple. Our study also emphasized the limitation of the direct diagnosis based on the PCR in PGT for DM1. For the "noninformative" couples, an alternative diagnostic method, such as haplotype analysis using SNP array, needs to be evaluated the accuracy and usefulness.

ACKNOWLEDGEMENTS
The authors offer their sincere gratitude to Yoko Yasuda and Miyuki Samejima from the Laboratory of reproductive genetics, Department of Obstetrics and Gynecology, Keio University School of Medicine.

DISCLOSURES
Conflict of interest: Hiroshi Senba, Kou Sueoka, Suguru Sato, Nobuhiko Higuchi, Yuki Mizuguchi, Kenji Sato, Mamoru Tanaka declare that they have no conflict of interest. Human rights statements and informed consent: All procedures followed were in accordance with the ethical standards of the responsible committee on human experimentation (institutional and national) and with the Helsinki Declaration of 1964 and its later amendments. Informed consent was obtained from all couples for being included in the study. Animal studies: The protocol and procedure of PGT were approved by the ethics committee both of our institution and the Japan Society of Obstetrics and Gynecology (JSOG) in every couple.

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How to cite this article: Senba H, Sueoka K, Sato S, et al. The impact of parental unaffected allele combination on the diagnostic outcome in the preimplantation genetic testing for myotonic dystrophy type 1 in Japanese ancestry. Reprod Med Biol. 2020;19:265–269. https://doi.org/10.1002/rmb2.12327