Bioprospective Screening of Ricinodendron Heudelotii Seeds

Introduction

The plant Ricinodendron heudelotii commonly called Njangsa is an oily seed tree that has long been used as a medicinal plant. Its uses include: an antidote against poison, because the extract of the bark are said to contain lupeol, to cure various diseases such as cough, malaria, yellow fever, stomach pain, rheumatism. Other characteristics of the plant are its aphrodisiac and anti-inflammatory properties. It has also been used for treatment of infertility and menstrual pain.

According to World Health Organization [1], a medicinal plant is any plant which contains substances that can be used for the therapeutic purpose or substances which are precursors for the synthesis. A plant becomes a medicinal plant only when its biological activities has been ethno-botanically reported and scientifically established [2,3].

In contrast to synthetic conventional pharmaceuticals, many medicinal plants exert their beneficial effects through the additive or synergistic action of several chemical compounds acting at single or multiple target sites associated with a physiological process [4]. Hence the need to screen the seed of Ricinodendron heudelotii for the presence and concentration of the secondary metabolites (phytochemicals), as the seed is majorly used in preparing various dishes.

Phytochemicals are chemical compound that occur naturally in plant, provide the plant with it various characteristics such as colour, flavor, odor and other organoleptic properties. These secondary metabolites are medicinal and can also be obtained from various anatomical structures of plants [2,5] reported that most of the phytochemicals are produced through biosynthesis in the metabolic pathway [6,7] also reported that the various phytochemicals have the following potentials:

a. Carotenoids: May inhibit cancer cell growth, act as an antioxidant and improve immune response.
b. Flavonoids (anthocyanins): May inhibit inflammation and tumour growth, also aid immunity and boost production of detoxifying enzymes in the body.
c. Inositol (phytic acid): May retard cell growth and work as antioxidants.
d. Isoflavones: May inhibit tumour growth, limit production of cancer related hormones and generally work as antioxidants.
e. Isothiocyanates: may induce detoxification of carcinogens, block tumour growth and work as antioxidants.
f. Polyphenols: may prevent cancer formation, inflammation and work as antioxidants.
g. Terpenes: may protect cells from becoming cancerous, slow cancer cell growth, strengthen immune function, limit production of cancer related hormones, fight viruses, and work as antioxidants.
h. Indoles and glucosinolates: may induce detoxification of carcinogens, limit production of cancer related hormones, block carcinogens and prevent tumour growth.

This study therefore screens for the presence (qualitative) and concentration (quantitative) phytochemical constituents of the extract of the dried Ricinodendron heudelotii seed, aimed at enhancing the therapeutic value and its use in pharmaceutical industries.
Materials and Methods

Collection and identification of the plant sample

The dried seeds of Ricinodendron heudelotii were collected from a local food spice market in Port Harcourt, Rivers state, Nigeria. The sample was identified by the department of Plant Science and Biotechnology, University of Port Harcourt, Nigeria. The seed was ground into fine powder using a manual grinding machine in the Department of Chemistry laboratory, Rivers state university of science and technology, Nigeria.

Quantitative (concentration) screening

1g of the sample powder was weighed and transferred into a test tube. 15ml ethanol and 10ml of 50% m/v potassium hydroxide was added. The test tube was allowed to heat in a water bath at 60°C for 60mins. After the reaction time, the reaction product contained in the test tube was transferred to a separatory funnel. The tube was thoroughly washed with 20ml ethanol, 10ml cold water, 10ml hot water and 3ml hexane, which was all transferred to the funnel. The solution was dried with anhydrous sodium sulfate and the solvent was evaporated. The sample was solubilized in 1000µl of pyridine of which 200µl was transferred to a vial for analysis.

Quantification by gas chromatography flame ionization detector (GC-FID)

The analysis for phytochemical was performed on a BUCK M910 Gas chromatography equipped with a flame ionization detector. A RESTEK 15 meter MXF-1 column (15m×250um×0.15um) was used. The injector temperature was 280°C with splitless injection of 2µl of samples and a linear velocity of 3°C ms⁻¹, helium 5.0pa.S was the carrier gas with a flow rate of 40mlmin⁻¹. The oven operated initially at 200°C, it was heated at 330°C at a rate of 30°C min⁻¹ and was kept at this temperature for 5mins. The detector operated at a temperature of 320°C.

Phytochemicals were determined by the ratio between the area and mass of internal standard and the area of the identified phytochemicals. The concentration of the different phytochemicals is expressed in ug/g.

Qualitative (Presence) Screening

Method of extraction

Chemical tests were carried out on the aqueous extract of the powdered sample of the Ricinodendron heudelotii using standard procedures to identify the constituents [8].

Test for tannins

0.5g of the dried powder sample was boiled in 20ml of water in a test tube and filtered. 3 drops 0.1% ferric chloride was added and observed for brownish green or blue black coloration.

Test for saponins

1g of powdered sample was separately boiled with 10ml of distilled water in a beaker for 10minutes. The mixture was filtered while hot and allowed to cool. The following tests were then carried out:

Frothing test: 2.5ml of filtrate was diluted to 10ml with distilled water and shaken vigorously for 2 minutes; Frothing indicated the presence of saponin in the filtrate.

Emulsion test: 2 drops of olive oil was added to the solution obtained from diluting 2.5ml filtrate to 10ml with distilled water (above), shaken vigorously for a few minutes Formation of a fairly stable emulsion indicated the presence of saponins.

Test for phlobatannins

5ml aqueous extract of sample was boiled with 1% aqueous hydrochloric acid. Deposition of red precipitate was taken as evidence for phlobatannins.

Test for terpenoids

5ml of aqueous extract of sample seed was added in 2ml of chloroform. 3ml of concentrated H2SO4 was then added to form a layer. A reddish brown precipitate coloration at the interface formed indicated the presence of terpenoids.

Test for flavonoids

1g of the powder sample was heated with 10ml of distilled water for 5minutes and filtered while hot. Few drops of 20% sodium hydroxide solution were added to 1ml of the cooled filtrate. A change to yellow colour which on addition of acid changed to colorless solution depicted the presence of flavonoids.

Test for cardiac glycosides

5ml of aqueous extract was treated with 2ml of glacial acetic acid containing one drop of ferric chloride solution. This was underplayed with 1ml of concentrated sulphuric acid. A brown ring at the interface indicated the deoxy sugar characteristics of cardenolides.

Test for combined anthraquinones

1g of powdered sample was boiled with 2ml of 10% hydrochloric acid for 5mins. The mixture was filtered while hot and filtrate was allowed to cool. The cooled filtrate was partitioned against equal volume of chloroform and the chloroform layer was transferred into a clean dry test tube using a clean pipette. Equal volume of 10% ammonia solution was added into the chloroform
layer, shaken and allowed to separate. The separated aqueous layer was observed for any colour change. Light rose pink colour showed the presence of combined anthraquinone.

**Test for free anthraquinones**

5ml of chloroform was added to 0.5g of the powdered sample. The resulting mixture was shaken for 5mins after which it was filtered. The filtrate was then shaken with equal volume of 10% ammonia solution. The presence of a bright pink colour in the aqueous layer indicated the presence of free anthraquinones.

**Test for carotenoids**

1g of powdered sample was extracted with 10ml of chloroform in a test tube, filtered and 85% sulphuric acid was added. A blue colour at the interface showed the presence of carotenoids.

**Test for reducing compounds**

To 1g of sample powder, 10ml distilled water was added. The mixture was allowed to boil for 5minutes, filtered while hot and the cooled filtrate made alkaline to litmus paper with 20% sodium hydroxide solution. The resulting solution was boiled with an equal volume of Benedict qualitative solution on a water bath. The formation of a brick red precipitate depicted the presence of reducing compound.

**Test for alkaloids**

1g of powdered sample was boiled with 2ml distilled water and 10ml (10%) hydrochloric acid on water bath and filtered. The pH of the filtrate was adjusted with ammonia to 10. The following reagents were added separately to about 0.5ml of the filtrate in a different test tube and observed:

a. 10% tannic solution - 1ml tannic solution, formation of precipitate indicates presence of alkaloid.

b. Mayer’s reagent (Potassium mercuric iodide solution). 1ml Mayer’s reagent. Turbidity indicates the presence of alkaloid.

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**Figure 2:** Quantitative phytochemical analysis of ethanolic seed extract of Ricinodendron heudelotii showing the concentration of the various phytochemical component.

**Figure 3:** Percentage concentration of sub-class of Alkaloids of ethanolic seed extract of Ricinodendron heudelotii.
Table 1: Quantitative phytochemical analysis of ricinodendron heudelotii seed.

| Component | Sub-class | Concentration µg/g |
|-----------|-----------|--------------------|
| Phenol    |           | 2.2357             |
| Tannin    |           | 39.9463            |
| Oxalate   |           | 1.1581             |
| Phytate   |           | 0.2946             |
| Alkaloid  | Lumarine  | 7.9079             |
|           | Ribalidine| 5.608              |
|           | Spartein  | 0.0002             |
| Saponin   | Sapogenin | 14.2632            |
| Flavonoid | Catechin  | 11.5693            |
|           | Epicatechin| 1.9145            |
|           | Rutin     | 5.9573             |
|           | Kaempferol| 3.1418             |
|           | Naringenin| 18.7939            |
|           | Anthocyanin| 2.4412            |
Table 2: Qualitative phytochemical analysis of ricinodendron heudelotii seeds.

| Component               | Result |
|-------------------------|--------|
| Tannin                  | +ve    |
| Phlobatannins           | -ve    |
| Flavonoid               | +ve    |
| Alkaloid                | +ve    |
| Cardiac glycoside       | +ve    |
| Terpenoid               | +ve    |
| Saponin frothing        | +ve    |
| Saponin emulsifying     | +ve    |
| Combined anthraquinone  | +ve    |
| Free Anthraquinone      | -ve    |
| Carotenoid              | +ve    |
| Reducing compound       | -ve    |

Discussion

The results obtained from the qualitative and quantitative phytochemical analysis reveals that tannin was present with the highest concentration [9]. Reported that tannins have a natural defense mechanism against microbial infection, exert other physiological effects such as reduce blood pressure, decrease serum lipid level, produce liver necrosis, accelerate blood clotting and modulate immune responses. Carotenoids were also present in the seed. Carotenoids have been reported to neutralize free radicals that cause cell damage [2]. The seed of the plant Ricinodendron heudelotii was found to contain phenolic compounds. Plants containing phenolic compounds possess antioxidant and anti-inflammatory properties.

Alkaloids and flavonoids were seen in appreciable concentration in the seed extract. Their content in plant materials have severally been reported to be a major anti-oxidant, anti-inflammatory and analgesic active principle. Alkaloid has widely been used medicinally as antiarrhythmic, anticholinergic, stimulant, antalgic, antimalarial and antithypertensive [10].

The concentration of Saponin was appreciable (14.2632) [11] reported saponins to be included in the formulations used in the treatment of wounds, cough, asthma, hay fever and generally as antimicrobial and antiviral application. Saponins have also been used as adjuvants in vaccines to stimulate the TH immune response and production of cytotoxic T-Lymphocytes against exogenous antigens [11].

The low concentration of oxalate may be related to the fact that low amount of oxalate is needed by the body as too much of it may pose a risk of kidney stone [12]. Also Phytate has been reported to have antioxidative properties and anti cancerogenic activities [13]. The result of the analysis indicated the absence of Phlobatannins, free anthraquinone and reducing compounds. The presence of these phytochemicals in the plant Ricinodendron heudelotii suggests its potential therapeutically and its use in pharmaceutical industries [14-19].

Conclusion

The plant Ricinodendron heudelotii has been used traditionally for the treatment of various diseases and sickness. The phytochemical screening of the seed of this plant has revealed the phytochemical constituent of the plant, hence suggesting its curative potency for various diseases and as such, its potential use therapeutically and in the pharmaceutical industries.

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