Water Salinity Should Be Reduced for Irrigation to Minimize Its Risk of Increased Soil N₂O Emissions

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Abstract: To reveal the effect of irrigation salinity on soil nitrous oxide (N₂O) emission, pot experiments were designed with three irrigation salinity levels (NaCl and CaCl₂ of 1, 2.5 and 4 g/L equivalence, Ec = 3.6, 8.1 and 12.7 ds/m), either for 0 kg N/ha (N0) or 120 kg N/ha (N120) nitrogen inputs. N₂O emissions from soils irrigated at different salinity levels varied in a similar pattern which was triggered by soil moisture dynamics. Yet, the magnitudes of pulse N₂O fluxes were significantly varied, with the peak flux at 5 g/L irrigation salinity level being much higher than at 2 and 8 g/L. Compared to fresh water irrigated soils, cumulative N₂O fluxes were reduced by 22.7% and 39.6% (N0), 29.1% and 39.2% (N120) for soils irrigated with 2 and 8 g/L saline water, while they were increased by 87.7% (N0) and 58.3% (N120) for soils irrigated with 5 g/L saline water. These results suggested that the effect degree of salinity on consumption and production of N₂O might vary among irrigation salinity ranges. As such, desalinating brackish water to a low salinity level (such as 2 g/L) before it is used for irrigation might be helpful for solving water resources crises and mitigating soil N₂O emissions.

Keywords: brackish water irrigation; N fertilizer; nitrogen oxide; water-filled pore space; electrical conductivity

1. Introduction

Due to a shortage of irrigation water, brackish water (salinity concentration of 2–5 g/L), or marginal saline water, has been adopted as an alternative of fresh water for agricultural irrigation in some areas of the world [1–5]. Prolonged application of brackish or saline water could exacerbate soil salinity, which inevitably affects soil properties, microbial activity, and thereby N transformation [6–10]. Meanwhile, agricultural soil is the main source of nitrous oxide (N₂O), contributing to 59.4% of global anthropogenic source of N₂O. The rise of atmospheric N₂O concentrations is primarily due to unreasonable agricultural management activities [11,12]. Soil moisture affected by irrigation management has been demonstrated to be one of the most important factors associated with soil N₂O emissions [13–15]. Therefore, it is expected that the interaction of soil salt and water will result in a different N₂O emissions from soil [16].

Increasing salt-affected agricultural land has attracted interest to understand the influence of salinity on soil N₂O emissions [17–22], but their conclusions sometimes conflict with each other. For example, Zhang et al. [22] found that N₂O fluxes from a drip-irrigated cotton field without nitrogen (N) fertilizer application increased with irrigation salinity from 0.4 to 8.0 ds/m. Yet, other studies [16,20] showed that salinity reduced N₂O emission significantly, either from soils or industrial waste-water
sludge. Furthermore, some researchers [19,22] found that the relationship between salinity and N$_2$O emissions varied among soil conditions. For instance, at under 65% water hold capacity (WHC), soil N$_2$O fluxes at 11.7 g/L salinity level were greater than at 35.1 g/L, while it showed the opposite behavior at under 40% WHC [19]. Moreover, under 0 kg N/ha, N$_2$O flux at 8.0 ds/m irrigation salinity level was greater than at 0.4 ds/m, but it showed opposite results under 360 kg N/ha [22]. In most of the literature regarding the impact of soil salinity on N$_2$O emissions, few researchers have focused on N$_2$O emissions from soils irrigated with brackish or saline water. The connection between N$_2$O emissions and irrigation salinity is still unclear.

Thus, pot experiments were conducted on a silty clay soil treated with or without N application, under three irrigation salinity levels. The primary objective of this study was to quantify N$_2$O fluxes from soils irrigated with water at different salinity levels, and discuss its responses to variations of soil moisture and salinity.

2. Experiments

2.1. Experimental Design

Experiments were conducted from May to August 2016 on a silty clay soil collected from a plastic greenhouse located at Nanjing Vegetables Scientific Institute in 2012 (31°56′ N, 118°37′ E). A soil sample was air-dried, ground, sieved, homogenized, and finally packed into cylindrical soil columns (I.D. = 30 cm, depth = 50 cm) to a bulk density of 1.25 g/cm$^3$ for 0–5 cm layer and 1.33 g/cm$^3$ for 5–50 cm layer, according to the local soil’s native bulk density. Columns were left undisturbed for over 60 days to avoid the influences of soil disturbance on N$_2$O emission. Detailed soil properties are listed in Table 1.

Table 1. Soil physical and chemical properties (mean ± standard deviation).

| Characteristic         | Value       |
|------------------------|-------------|
| Clay (%)               | 29 ± 1.1    |
| Silt (%)               | 48 ± 1.7    |
| Sand (%)               | 23 ± 0.9    |
| Ec (ds/m)              | 0.4 ± 0.06  |
| pH (H$_2$O)            | 6.2 ± 0.1   |
| Organic matter (OM, g/kg) | 25.2 ± 0.3 |
| Total nitrogen (TN, g/kg) | 1.4 ± 0.02 |
| Total phosphorus (TP, g/kg) | 0.04 ± 0.01 |
| Nitrate nitrogen (NO$_3^-$-N, mg/kg) | 34.2 ± 2.9 |
| Ammonium nitrogen (NH$_4^+$-N, mg/kg) | 8.8 ± 0.7 |

The soil sample was irrigated at three salinity levels: 2, 5 and 8 g/L (namely treatments S2, S5 and S8, with Ec of 3.6, 8.1 and 12.7 ds/m), either for zero (N0) or field N application levels (120 kg N/ha, N120). Saline water was produced by adding NaCl and CaCl$_2$ at a mass ratio of 1:1 to fresh water. Urea (N = 46.2%) was used as the N source. Fresh water (Ec = 0.3 ds/m), either with N0 and N120 application, was set as a control (namely treatments CK and CK + N). All treatments (S2, S5, S8, CK, S2 + N, S5 + N, S8 + N and CK + N) were replicated six times (three for gas sampling and another three for soil sampling). The soil was watered twice, at 22:00 on 19 July and 22:00 on 3 August 2016. For each treatment, 3530 and 3210 mL of fresh or saline water was supplied to replenish surface (0–20 cm) soil moisture to field capacity for the first and second irrigation. Air temperatures after the first irrigation ranged from 33.1 to 38.4 °C, which was on average 8.3% higher than that after the second irrigation (29.9–38.4 °C).
2.2. Gases Sampling and Analyzing

Gas samples from each soil column were collected daily during the observation period using the static chamber (I.D. = 30 cm, depth = 30 cm), which was made of 8 mm polyvinyl chloride (PVC) and equipped with a thermometer and an electric fan at the top for measuring air temperature and mixing air inside (Figure 1). A rubber tube was inserted into the chamber from the top and connected to a three-way stopcock, which was used to draw gas samples using a 10 mL syringe. Four gas samples from each chamber were collected at 10-min intervals between 9:45 h and 10:15 h on each sampling day. When gas samples were collected, air temperatures inside the chambers, as well as soil temperature at 0, 5, 10 and 15 cm layers, were measured using mercury thermometers (accuracy, 0.1 °C).

\[ \text{Figure 1. Sketch of soil box and the chamber.} \]

N$_2$O concentration was analyzed within 48 h after sampling using a gas chromatograph system (Agilent 7890B; Agilent Technologies Inc., Santa Clara, CA, USA) with an electron capture detector (ECD). N$_2$O fluxes were calculated according to the linear increment in N$_2$O concentration within the chamber [11]. Cumulative N$_2$O fluxes were calculated by integrating daily values along the sampling period.

2.3. Soil Moisture and Salinity Measurement

Soil samples (approximately 3–5 g each) from 20 cm depth in 5 cm increment were collected at 10:00 h every 2 days using stainless steel samplers (O.D. = 10 cm). Soil moisture content was determined gravimetrically by drying at 105 °C for 24 h, and water-filled pore space (WFPS) was calculated as the percentage of soil volumetric moisture content relative to the soil porosity. Soil electrical conductivity (EC) was measured (soil:water = 1:5) using an electrical conductivity meter (TES-1381K, Taiwan, China).

2.4. Statistical Analysis

Statistical analysis was performed using SPSS 19.0 (SPSS Inc., Chicago, IL, USA). A Fisher’s least significant difference (LSD) test was performed to determine the difference in soil average or cumulative N$_2$O fluxes (n = 3) among treatments in non-pulse or pulse period. A paired t test was conducted to evaluate the difference among soil depths or treatments soil-related factors (temperature, moisture and salinity) with time series (n = N, the N was the number of data in each time series).
3. Results

3.1. Soil Temperature

No significant differences were found in soil temperature among treatments with fresh or saline water irrigation (Figure 2). For all treatments, soil temperatures at 0, 5, 10, and 15 cm layers mainly fell within the ranges of 27.3–37.9 °C, 23.1–37.1 °C, 21.8–37.6 °C, and 21.4–38.0 °C, respectively. The minimum values were observed 2 days after the second irrigation due to the reduced air temperature and low irrigation water temperature (25.7 °C). Among soil depths, soil temperatures decreased with the increasing of soil layers, but the differences were mostly not significant ($p > 0.1$).

![Figure 2](image_url)

**Figure 2.** Soil temperature at 0, 5, 10 and 15 cm layers after two irrigation events from 20 July to 18 August. Solid arrow and dotted arrow represent irrigation and fertilization event, respectively. S + N and S means the average soil temperature from three saline water irrigated soils (S2, S5 and S8) with N120 and N0 application, respectively.

3.2. Soil Moisture

Soil moisture at 0–15 cm layers varied in the similar pattern and fell within same ranges in both saline water and fresh water irrigated soils, either under N0 or N120 level (Figure 3). Soil moisture varied significantly at early observation period, and then decreased with time from 89.8–93.8%, 90.9–95.9%, 91.1–96.3% and 90.2–94.4% WFPS to 11.7–17.2%, 23.3–32.0%, 32.7–39.4% and 40.1–46.4% WFPS at 0, 5, 10 and 15 cm layers, respectively. Among soil layers, soil WFPS varied more significantly in 0 and 5 cm depths than in 10 and 15 cm depths.
3.3. Soil Salinity

In contrast to soil moisture, soil Ec was low immediately after irrigation and increased gradually along with the decrease of soil moisture (Figure 4). Generally, soil Ec at 5 cm depths was higher than at upper or lower depths, indicating that solute mostly accumulated in soil at a depth of 5 cm. Among soil layers, soil Ec from saline water irrigated soils was significantly higher ($p < 0.05$) than that of fresh water irrigated soils (0.06–0.2 ds/m), either under N0 or N120 level. For S2 + N, S5 + N and S8 + N treatments, soil Ec mainly varied in the ranges of 0.2–2.2 ds/m, 0.3–3.8 ds/m, 0.5–5.4 ds/m, respectively. It increased remarkably with irrigation salinity from 2 to 8 g/L, with averages from S5 + N and S8 + N treatments 59.1% and 139.8%, i.e., significantly higher than from S2 + N treatment ($p < 0.05$). For soils with N0 application (S2, S5 and S8 treatments), soil Ec was slightly lower than that of soils with N120 application, but varied in a similar pattern.

Figure 3. Soil WFPS at 0, 5, 10 and 15 cm depths after two irrigation events from 2 July to 18 August. Solid arrow and dotted arrow represent irrigation and fertilization event, respectively. S + N and S denotes the average WFPS from three saline water irrigated soils (S2, S5 and S8) with N120 and N0 application, respectively.
3.4. **N₂O Emission Fluxes**

Average or pulse N₂O fluxes from saline water irrigated soils were quite different among salinity levels, even though their temporal courses triggered by the soil wetting-drying process were much similar. Under the N120 level, N₂O fluxes from soil irrigated at 5 g/L salinity level were much higher than at 2 and 8 g/L levels after both irrigations. Under the N0 level, a similar phenomenon was found for soil N₂O fluxes after first irrigation, yet it remained almost the same among salinity levels after the second irrigation (Figure 5).

For S2 + N, S5 + N and S8 + N treatments, N₂O fluxes increased rapidly during the first 3 days after first irrigation to 7812.4, 10,275.1, and 5353.2 µg N₂O/(m²h), and then declined to 66.5, 41.9, and 71.7 µg N₂O/(m²h). Irrigation significantly stimulated N₂O emission from soils, with peak N₂O fluxes after second irrigation 32.3–140.9 times greater than observed on 2 August prior to the second irrigation. Compared to CK + N treatment, the peak N₂O flux was enhanced by 8.2% for S5 + N treatment, while it was reduced by 10.2% and 28.8% for S2 + N and S8 + N treatments. As a result, the average N₂O flux from S5 + N treatment increased by 1406.1 µg N₂O/(m² h), and from S2 + N and S8 + N treatments decreased by 702.1 and 944.7 µg N₂O/(m² h), compared to CK + N (Table 2).

For S2, S5 and S8 treatments, pulse N₂O fluxes were observed 3 days after both irrigations, primarily in the ranges of 7590.6–21,245.7 µg N₂O/(m²h) and 2078.2–4424.2 µg N₂O/(m²h). The peak

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**Figure 4.** Soil Ec at 0, 5, 10 and 15 cm depths after two irrigation events from 2 July to 18 August. Solid arrow and dotted arrow represent irrigation and fertilization event, respectively. S2, S5 and S8 represent soils irrigated with water at 2, 5 and 8 g/L salinity level; S2 + N, S5 + N and S8 + N represent soils irrigated with water at 2, 5 and 8 g/L salinity level and nitrogen fertilization (120 kg N/ha) application. CK + N and CK represent soils irrigated with fresh water at N120 and N0 level, respectively.
N₂O fluxes from S2 and S5 soils were 5.7% and 66.5% higher than CK, yet the flux from S8 soil was 40.5% lower. Consequently, the average N₂O fluxes from S2 and S8 treatments were reduced by 22.6% and 39.6%, while from S5 treatment was increased by 87.7% ($p < 0.01$), compared to CK. Across salinity levels, N₂O fluxes were significantly lower in unfertilized soils, decreasing by 39.1% on average ($p < 0.05$) (Table 2).

**Figure 5.** N₂O emissions from soil irrigated at different salinity levels after two irrigation events from 20 July to 18 August. Solid arrow and dotted arrow represent irrigation and fertilization event, respectively. S2, S5, and S8 represent soils irrigated with water at 2, 5, and 8 g/L salinity level; S2 + N, S5 + N and S8 + N represent soils irrigated with water at 2, 5, and 8 g/L salinity level and nitrogen fertilization (120 kg N/ha) application. CK + N and CK represent soils irrigated with fresh water at N120 and N0 level, respectively.

**Table 2.** Average N₂O fluxes from different treatments during pulse and non-pulse periods.

| Treatments (n = 3) | 20 July–26 July | 26 July–2 August | 2 August–9 August | 9 August–18 August | 20 July–18 August |
|--------------------|----------------|------------------|------------------|-------------------|------------------|
| **Pulse Period**   | **Non-Pulse Period** | **Pulse Period** | **Non-Pulse Period** | **The Whole Period** |
|---------------------|---------------------|-----------------|-------------------|-----------------|
| CK+N                | 3353.1 ± 245.9a     | 77.5 ± 11.2a    | 969.8 ± 78.4a     | 54.9 ± 11.2a    | 1043.2 ± 95.8a  |
|                     | (75.0%)             | (1.7%)          | (21.7%)           | (1.6%)           | (100%)          |
| S2+N                | 6476.8 ± 432.1b     | 136.2 ± 21.0ba  | 2938.4 ± 202.6b   | 1008.1 ± 104.3b | 2531.1 ± 286.7b|
|                     | (59.7%)             | (1.2%)          | (27.1%)           | (12.0%)          | (100%)          |
| S5+N                | 21086.6 ± 157.9a    | 64.8 ± 11.8a    | 1134.2 ± 107.0a   | 141.6 ± 20.6a   | 814.3 ± 78.5a   |
|                     | (60.4%)             | (1.9%)          | (32.5%)           | (5.2%)           | (100%)          |
| S8+N                | 3042.3 ± 272.3c     | 110.4 ± 14.0b   | 1863.3 ± 126.2c   | 594.3 ± 32.4c   | 1348.7 ± 91.4a  |
|                     | (52.6%)             | (1.9%)          | (32.3%)           | (13.2%)          | (100%)          |

Different letters in each column indicate significant differences at the 5% level. Values in the bracket represent the contribution to the total N₂O emissions. S2, S5, and S8 represent soils irrigated with water at 2, 5 and 8 g/L salinity level; S2 + N, S5 + N and S8 + N represent soils irrigated with water at 2, 5 and 8 g/L salinity level and nitrogen fertilization (120 kg N/ha) application.
3.5. Cumulative N\textsubscript{2}O Emissions

Over the whole experimental period (30 days), cumulative N\textsubscript{2}O fluxes from saline water irrigated soils under N0 level ranged from 586.3 to 1822.4 mg/m\textsuperscript{2}, with fluxes at 5 g/L salinity level as the highest (Figure 6). The application of exogenous N enhanced N\textsubscript{2}O emissions remarkably; the cumulative N\textsubscript{2}O fluxes from saline water irrigated soils under N120 level were 50.8–80.2% greater than under N0 level. Furthermore, under both N levels, cumulative N\textsubscript{2}O fluxes from 2 and 8 g/L saline water irrigated soils were significantly lower than from fresh water irrigated soils, decreasing on average by 25.9% and 39.4% \((p < 0.05)\). Yet for 5 g/L saline water irrigated soil, it increased by 73.0% on average.

![Figure 6](image)

**Figure 6.** Cumulative N\textsubscript{2}O fluxes from saline water irrigated soils and its reduction rates compared to fresh water irrigated soils. (a) represent cumulative N\textsubscript{2}O fluxes from S2 + N, S5 + N, and S8 + N soils and its reduction rates compared to CK + N soils. (b) represent cumulative N\textsubscript{2}O fluxes from S2, S5, and S8 and its reduction rates compared to CK soils. Different letters on each bar represent significant difference in N\textsubscript{2}O emissions at \(p < 0.05\) level. S2, S5, and S8 represent soils irrigated with water at 2, 5, and 8 g/L salinity level; S2 + N, S5 + N, and S8 + N represent soils irrigated with water at 2, 5, and 8 g/L salinity level and nitrogen fertilization (120 kg N/ha) application. CK + N and CK represent fresh water irrigated soils with or without nitrogen fertilization (120 kg N/ha) application, respectively.

4. Discussion

Both natural rainfall and irrigation can be important regulators of soil N\textsubscript{2}O emissions [23–28], causing large pulses emissions relative to normal conditions [29,30]. The occurrences of short-term pulse N\textsubscript{2}O emissions during drying processes were confirmed from saline water irrigated soils in the current experiment (Figure 5). The peak times, as well as the magnitudes of pulse N\textsubscript{2}O fluxes, were in agreement with the results reported in the aforementioned literature. Moreover, correlation analysis between N\textsubscript{2}O emissions and soil moisture or salinity indicated that N\textsubscript{2}O fluxes from different saline water irrigated soils increased exponentially with the increase of soil WFPS (Table 3). The determined coefficients of the exponential functions between N\textsubscript{2}O flux and soil WFPS in saline water irrigated treatments were higher than fresh water irrigated treatments, with the determined coefficient for the treatment at low irrigation salinity level as the highest. Yet, when it comes to the factor of soil Ec, N\textsubscript{2}O fluxes was found to decrease exponentially with the increase of soil Ec in all treatments (Table 3). The determined coefficients decreased with irrigation salinity from 2 to 8 g/L. Generally, the determined coefficients of N\textsubscript{2}O flux to soil WFPS were much higher than it to soil Ec. These results suggested the dynamic of N\textsubscript{2}O emissions from soils irrigated with saline water were significantly influenced by both soil WFPS and soil Ec.
water irrigated soils were significantly lower than from fresh water irrigated soils (Ec of 0.3 ds/m), and the exogenous N almost did not change the response of N while conditions of soil WFPS and temperature kept almost the same ranges (Table 4). It implied that irrigation salinity was a key factor that affected the magnitudes of pulse N2O fluxes under both N application levels (N0 and N120) were much higher from soils irrigated at 5 g/L salinity level than at 2 and 8 g/L, while conditions of soil WFPS and temperature kept almost the same ranges (Table 4). It implied that irrigation salinity was a key factor that affected the magnitudes of pulse N2O fluxes from saline water irrigated soils, and the exogenous N almost did not change the response of N2O peak fluxes to soil-related factors (soil moisture, Ec or temperatures).

Generally, pulse N2O emissions appeared when soil moisture, salinity and temperature were appropriate (Table 4). In current research, the pulse N2O fluxes under both N application levels (N0 and N120) were much higher from soils irrigated at 5 g/L salinity level than at 2 and 8 g/L, while conditions of soil WFPS and temperature kept almost the same ranges (Table 4). It implied that irrigation salinity was a key factor that affected the magnitudes of pulse N2O fluxes from saline water irrigated soils, and the exogenous N almost did not change the response of N2O peak fluxes to soil-related factors (soil moisture, Ec or temperatures).

**Table 3.** The relationship of N2O emissions to soil moisture or salinity for each treatment.

| Treatments | N2O V.S. WFPS | N2O V.S. Ec |
|------------|---------------|-------------|
|            | Relation      | Determine   | Relation      | Determine   |
|            | Coefficient (R²) | Coefficient (R²) | Coefficient (R²) | Coefficient (R²) |
| CK + N     | y = 13.21x + 0.04x | 0.28 * | y = 2474x + 11.7x | 0.07 ns |
| S2 + N     | y = 12.50x + 0.05x | 0.66 * | y = 1496x + 5.84x | 0.59 * |
| S5 + N     | y = 76.87x + 0.05x | 0.34 * | y = 4100x + 3.43x | 0.56 * |
| S8 + N     | y = 20.41x + 0.05x | 0.47 * | y = 6988x + 1.94x | 0.45 * |
| CK         | y = 87.46x + 0.04x | 0.26 * | y = 2980x + 15.53x | 0.15 ns |
| S2         | y = 10.82x + 0.05x | 0.58 * | y = 4201x + 8.76x | 0.57 * |
| S5         | y = 60.36x + 0.05x | 0.44 * | y = 5636x + 4.26x | 0.55 * |
| S8         | y = 14.75x + 0.05x | 0.57 * | y = 1476x + 2.73x | 0.61 * |

The symbols of * indicate correlation is significant at p < 0.05 level. ns represents no significance. S2 + N, S5 + N and S8 + N represent soils irrigated with water at 2, 5, and 8 g/L level and nitrogen fertilization (120 kg N/ha) application. S2, S5, and S8 represent soils irrigated with water at 2, 5, and 8 g/L salinity level. CK + N and CK represent treatments with fresh water irrigation under N120 and N0 level, respectively.

| Irrigation Events | Treatment | N2O Peak Fluxes (µg N2O/(m²h)) | WFPS (%) | Ec (ds/m) | T (°C) |
|-------------------|-----------|---------------------------------|----------|-----------|--------|
| After first irrigation (20 July–1 August) | S2 + N | 20,509.1 ± 1095.9 | 74.6 ± 6.1 | 0.47 ± 0.08 | 36.8 ± 0.4 |
|                   | S5 + N | 24,795.4 ± 1178.5 | 74.4 ± 5.9 | 0.68 ± 0.10 | 36.4 ± 0.5 |
|                   | S8 + N | 16,312.2 ± 808.4 | 76.3 ± 6.3 | 0.99 ± 0.12 | 36.7 ± 0.3 |
|                   | CK + N | 22,926.2 ± 1252.9 | 72.1 ± 5.6 | 0.10 ± 0.03 | 37.0 ± 0.4 |
| S2                | 13,492.5 ± 618.9 | 70.9 ± 5.7 | 0.41 ± 0.06 | 36.3 ± 0.3 |
| S5                | 21,245.7 ± 1139.9 | 71.5 ± 5.9 | 0.63 ± 0.09 | 36.4 ± 0.4 |
| S8                | 7590.6 ± 323.2  | 72.4 ± 5.3 | 0.87 ± 0.12 | 37.0 ± 0.4 |
| CK                | 12,759.9 ± 631.7 | 68.9 ± 6.3 | 0.07 ± 0.02 | 37.3 ± 0.6 |
| After second irrigation (2 August–18 August) | S2 + N | 3465.7 ± 859.0  | 75.2 ± 6.1 | 0.55 ± 0.08 | 31.0 ± 0.3 |
|                   | S5 + N | 13,619.1 ± 1359.9 | 77.8 ± 5.9 | 0.79 ± 0.11 | 30.9 ± 0.2 |
|                   | S8 + N | 2635.8 ± 251.0  | 78.9 ± 4.3 | 1.22 ± 0.10 | 31.1 ± 0.4 |
|                   | CK + N | 5418.9 ± 275.6  | 73.6 ± 6.0 | 0.16 ± 0.02 | 31.3 ± 0.3 |
| S2                | 2347.0 ± 223.2 | 76.8 ± 5.6 | 0.54 ± 0.08 | 31.1 ± 0.4 |
| S5                | 4424.2 ± 359.9 | 75.7 ± 6.3 | 0.64 ± 0.09 | 31.1 ± 0.3 |
| S8                | 2078.2 ± 275.6 | 78.4 ± 7.1 | 1.14 ± 0.12 | 31.4 ± 0.3 |
| CK                | 3304.2 ± 287.9 | 69.1 ± 6.8 | 0.07 ± 0.03 | 32.1 ± 0.5 |

The values of soil WFPS, Ec, and temperature represent the averages of those values at 0–15 cm layers. S2, S5, and S8 represent soils irrigated with water at 2, 5, and 8 g/L salinity level; S2 + N, S5 + N and S8 + N represent soils irrigated with water at 2, 5, and 8 g/L level and nitrogen fertilization (120 kg N/ha) application. CK + N and CK represent fresh water irrigated soils with or without nitrogen fertilization (120 kg N/ha) application, respectively.

Previous researches had investigated the impact of salinity on N2O emissions, whereas their conclusions were contradictory to each other [17,19,21,22,31] (Table 5). Sometimes it increased remarkably with the increase of salinity level [22], but sometimes reduced [17], or showed no effect [19,21,31]. In present study, N2O fluxes from 2 and 8 g/L (Ec of 3.6 and 12.7 ds/m) saline water irrigated soils were significantly lower than from fresh water irrigated soils (Ec of 0.3 ds/m),
while from 5 g/L (Ec of 8.1 ds/m) saline water irrigated soil, it enhanced remarkably. This phenomenon likely suggested that the effect degree of salinity on consumption and production of N$_2$O might vary among irrigation salinity ranges. Some previous results once indicated that the activity of N$_2$O reductase, which reduces N$_2$O to N$_2$ in nitrogen transformation, was inhibited to a lesser degree under low rather than medium salinity [8,9,21,22,32–34]. That partially accounted for the higher N$_2$O emission at moderate rather than low salinity levels. Furthermore, the activities of ammonia-oxidizing bacteria and ammonia-oxidizing archaea were significantly reduced under high salinity levels, which may result in the inhibition of the first step of the nitrification (ammonia oxidation) process, and thus, possible reduced N$_2$O emission levels [35]. This might be another important reason for decreased N$_2$O emissions under high salinity level. These results also indicated that reducing salinity of brackish water from 5 to 2 g/L could be helpful for N$_2$O mitigation.

With increasing shortage of agricultural water resources, brackish water (salinity concentration of 2–5 g/L) is widely applied for irrigation [2,22,36]. The current research was performed to investigate N$_2$O emissions from soils irrigated with water at different salinity levels. Results indicated that the response of N$_2$O emissions to salinity might vary among irrigation salinity ranges, and irrigation with desalinated brackish water could be a potential measure for N$_2$O mitigation. But unfortunately, the N$_2$O emissions were collected from a soil with low initial salinity (Ec = 0.4 ds/m), and the irrigation salinity level did not set in a finer classification (such as 2.5, 3, 3.5 and 4 g/L). Thereafter, more investigation should be done to reveal the effect of salts on N$_2$O emissions under different initial soil salinity conditions and higher irrigation salinity levels, and to figure out a suitable irrigation salinity level, which is helpful for mitigating N$_2$O emissions while meeting the requirement of crop growth.
Table 5. Summary of results of the effect of salinity on N$_2$O emissions.

| Experiment Type                     | Salt Type                | Salinity Level                                                                 | N Level (kg N/ha) | Cumulative N$_2$O Flux | Conclusion        | Reference |
|-------------------------------------|--------------------------|-------------------------------------------------------------------------------|------------------|-------------------------|-------------------|-----------|
| A pot experiment (30 d)             | Nacl:CaCl$_2$ (1:1)      | 2, 5 and 8 g/L in water (S2, S5 and S8)                                      | 0                | 5.9–18.2 kg/ha          | S5 $>$ CK $>$ S2 $>$ S8 | Current research |
| A incubation experiment (28 d)      | NaCl + NH$_4$Cl          | 0, 5.9, 11.7, 23.4 and 35.1 g/L in soil solution (S0, S5.9, S11.7, S23.4 and S35.1) | /                | 96.3–169.6 ug/(g soil)  | 65% WHC: S5.9 $>$ S11.7 $>$ S35.1 $>$ S0 $>$ S23.4 (ns) | [19]      |
| A incubation experiment             | Na$_2$SO$_4$ + (NH$_4$)$_2$SO$_4$ | /                                                                         | /                | 205.5–322.7 ug/(g soil) | 65% WHC: S0 $>$ S5.9 $>$ S35.1 $>$ S23.4 $>$ S11.7 (ns) |           |
| A incubation experiment             | Ca(NO$_3$)$_2$          | 0, 5.9, 17.6 and 29.3 g/L in soil solution (S0, S5.9, S17.6 and S29.3)        | /                | 122.5–699.8 ug/(g soil) | 70% WHC: S5.9 $>$ S0 $>$ S17.6 $>$ S29.3 (ns) |           |
| A incubation experiment             | Nacl                     | 2.5, 7.5, 10 g/L in water (S2.5, S7.5 and S10)                             | /                | /                       | S10 (5.1) $>$ S7.5 $>$ S2.5 (2.2–2.3) (unit: mg/L) | [21]      |
| A incubation experiment             | Nacl                     | 0, 1, 3 and 6 g/(kg soil) (S0, S1, S3 and S6)                               | /                | /                       | 20% moisture (w/w): S6 (0.5) $<$ S3 (1.2) $<$ S1 (2.8) (unit: ng/(kg h)) | [17]      |
| A bean field experiment (84 d)      | Nacl                     | 0.03 and 0.6 g/L in water (S0.03 and S0.6)                                  | 120              | 0.5 and 0.4 kg/ha       | S0.03 $>$ S0.6 (ns) | [31]      |
| A cotton field experiment (57 d)    | Nacl:CaCl$_2$ (1:1)      | 0.4, 4.6 and 8.1 ds/m in water (C0.4, C4.6 and C8.1)                         | 0                | 0.1–0.2 kg/ha           | C0.4 $<$ C4.6 $<$ C8.1 | [22]      |

ns means the difference in N$_2$O fluxes between treatments was not significant.
5. Conclusions

Pulse N\textsubscript{2}O emissions from soils irrigated with saline water were found to be triggered by soil moisture dynamics, and the magnitude of pulse N\textsubscript{2}O fluxes for 5 g/L treatment were significantly higher than those for 2 and 8 g/L treatments. These results implied that soil wetting-drying processes dominated the temporal process of N\textsubscript{2}O emissions from saline water irrigated soils, while irrigation salinity and N level affected its magnitudes remarkably. Compared to fresh water irrigation, 2 and 8 g/L saline water resulted in lower N\textsubscript{2}O emissions from soils either under N0 or N120 level, while 5 g/L saline water irrigation enhanced it significantly. It can be inferred that the sensitivity of N\textsubscript{2}O production and consumption to irrigation salinity might be different among salinity ranges, and reducing the salinity of brackish water might be a potential measure for solving agricultural water crises and reducing soil N\textsubscript{2}O emissions.

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References

1. Assouline, S.; Moller, M.; Cohen, S.; Ben-Hur, M.; Grava, A.; Narkis, K.; Silber, A. Soil-plant system response to pulsed drip irrigation and salinity: Bell pepper case study. Soil Sci. Soc. Am. J. 2006, 70, 1556–1568. [CrossRef]
2. Baath, G.S.; Shukla, M.K.; Bosland, P.W.; Steiner, R.L.; Walker, S.J. Irrigation water salinity influences at various growth stages of Capsicum annuum. Agric. Water Manag. 2017, 179, 246–253. [CrossRef]
3. Chen, W.P.; Hou, Z.A.; Wu, L.S.; Liang, Y.C.; Wei, C.Z. Effects of salinity and nitrogen on cotton growth in arid environment. Plant Soil 2010, 326, 61–73. [CrossRef]
4. Letey, J.; Feng, G.L. Dynamic versus steady-state approaches to evaluate irrigation management of saline waters. Agric. Water Manag. 2007, 91, 1–10. [CrossRef]
5. Caruso, C.; Maucieri, C.; Berruti, A.; Borin, M.; Barbera, A.C. Responses of Different Panicum miliaceum L. Genotypes to Saline and Water Stress in a Marginal Mediterranean Environment. Agronomy 2018, 8, 8. [CrossRef]
6. Adviento-Borbe, M.A.A.; Doran, J.W.; Drijber, R.A.; Dobermann, A. Soil electrical conductivity and water content affect nitrous oxide and carbon dioxide emissions in intensively managed soils. J. Environ. Qual. 2006, 35, 1999–2010. [CrossRef] [PubMed]
7. Bernhard, A.E.; Donn, T.; Giblin, A.E.; Stahl, D.A. Loss of diversity of ammonia-oxidizing bacteria correlates with increasing salinity in an estuary system. Environ. Microbiol. 2005, 7, 1289–1297. [CrossRef] [PubMed]
8. Kumar, U.; Kumar, V.; Singh, J.P. Effect of different factors on hydrolysis and nitrification of urea in soils. Arch. Agron. Soil Sci. 2007, 53, 173–182. [CrossRef]
9. Lodhi, A.; Arshad, M.; Azam, F.; Sajjad, M.H. Changes in mineral and mineralizable N of soil incubated at varying salinity, moisture and temperature regimes. Pak. J. Bot. 2009, 41, 967–980.
10. Reddy, N.; Crohn, D.M. Effects of soil salinity and carbon availability from organic amendments on nitrous oxide emissions. Geoderma 2014, 235–236, 363–371. [CrossRef]
11. Hou, H.J.; Chen, H.; Cai, H.J.; Yang, F.; Li, D.; Wang, F.T. N\textsubscript{2}O and CO\textsubscript{2} emissions from Lou soils of greenhouse tomato fields under aerated irrigation. Atmos. Environ. 2016, 132, 69–76. [CrossRef]
12. Ussiri, D.; Lal, R. Global Sources of Nitrous Oxide. In Soil Emission of Nitrous Oxide and its Mitigation; Springer: Dordrecht, The Netherland, 2013; pp. 131–175.
13. Lognoul, M.; Theodorakopoulos, N.; Hiel, M.P.; Regaert, D.; Broux, F.; Heinesch, B.; Bodson, B.; Vandenbol, M.; Aubinet, M. Impact of tillage on greenhouse gas emissions by an agricultural crop and dynamics of N\textsubscript{2}O fluxes: Insights from automated closed chamber measurements. *Soil Tillage Res.* 2017, 167, 80–89. [CrossRef]

14. Rafique, R.; Anex, R.; Hennessy, D.; Kiely, G. What are the impacts of grazing and cutting events on the N\textsubscript{2}O dynamics in humid temperate grassland? *Geoderma* 2012, 181–182, 36–44. [CrossRef]

15. Repo, M.E.; Susiluoto, S.; Lind, S.E.; Jokinen, S.; Elsavok, V.; Biai, C.; Vratanen, T.; Martikainen, P.J. Large N\textsubscript{2}O emissions from cryoturbated peat soil in tundra. *Nat. Geosci.* 2009, 2, 189–192. [CrossRef]

16. Maucieri, C.; Zhang, Y.; McDaniel, M.D.; Borin, M.; Adams, M.A. Short-term effects of biochar and salinity on soil greenhouse gas emissions from a semi-arid Australian soil after re-wetting. *Geoderma* 2017, 307, 267–276. [CrossRef]

17. Azam, F.; Muller, C. Effect of sodium chloride on denitrification in glucose amended soil treated with ammonium and nitrate nitrogen. *J. Plant Nutr. Soil Sci.* 2003, 166, 594–600. [CrossRef]

18. Ghosh, U.; Thapa, R.; Desutter, T.; He, Y.B.; Chatterjee, A. Saline-Sodic Soils: Potential Sources of Nitrous Oxide and Carbon Dioxide Emissions? *Pedosphere* 2017, 27, 65–75. [CrossRef]

19. Inubushi, K.; Barahona, M.A.; Yamakawa, K. Effects of salts and moisture content on N\textsubscript{2}O emission and nitrogen dynamics in Yellow soil and Andosol in model experiments. *Biol. Fertil. Soils* 1999, 29, 401–407. [CrossRef]

20. Tsuneda, S.; Mikami, M.; Kimochi, Y.; Hirata, A. Effect of salinity on nitrous oxide emission in the biological nitrogen removal process for industrial wastewater. *J. Hazard. Mater.* 2005, 119, 93–98. [CrossRef] [PubMed]

21. Shang, H.L.; Peng, Y.Z.; Zhang, J.R.; Ye, L.; Zhao, K.F. Effects of salinity concentration on N\textsubscript{2}O production during nitrification (abstract in Chinese). *Environ. Sci.* 2009, 30, 1079–1083.

22. Zhang, W.; Zhou, G.W.; Li, Q.; Liao, N.; Guo, H.J.; Min, W.; Ma, L.J.; Ye, J.; Hou, Z.A. Saline water irrigation stimulate N\textsubscript{2}O emission from a drip-irrigated cotton field. *Acta Agric. Scand. B.* 2016, 66, 141–152. [CrossRef]

23. Kusa, K.; Hu, R.G.; Sawamoto, T.; Hatano, R. Three years of nitrous oxide and nitric oxide emissions from silanic andosols cultivated with maize in Hokkaido, Japan. *Soil Sci. Plant Nutr.* 2006, 52, 103–113. [CrossRef]

24. Kallenbach, C.M.; Rolston, D.E.; Horwath, W.R. Cover cropping affects soil N\textsubscript{2}O and CO\textsubscript{2} emissions differently depending on type of irrigation. *Agric. Ecosyst. Environ.* 2010, 137, 251–260. [CrossRef]

25. Manalil, S.; Riethmuller, G.; Flower, K. Rapid emission of nitrous oxide from fallow over summer following wetting in a Mediterranean-type environment. *Soil Tillage Res.* 2014, 143, 130–136. [CrossRef]

26. Scheer, C.; Wassmann, R.; Butterbach-Bahl, K.; Lamers, J.; Martius, C. The relationship between N\textsubscript{2}O, NO, and N-2 fluxes from fertilized and irrigated dryland soils of the Aral Sea Basin, Uzbekistan. *Plant Soil* 2009, 314, 273–283. [CrossRef]

27. Toma, Y.; Takakai, F.; Darung, U.; Kuramoto, K.; Limin, S.H.; Dohong, S.; Hatano, R. Nitrous oxide emission derived from soil organic matter decomposition from tropical agricultural peat soil in central Kalimantan, Indonesia. *Soil Sci. Plant Nutr.* 2011, 57, 436–451. [CrossRef]

28. Yamamoto, A.; Akiyama, H.; Naokawa, T.; Yagi, K. Effect of lime-nitrogen application on N\textsubscript{2}O emission from an Andosol vegetable field. *Soil Sci. Plant Nutr.* 2012, 58, 245–254. [CrossRef]

29. McDaniel, M.D.; Kaye, J.P.; Kaye, M.W. Do ‘hot moments’ become hotter under climate change? Soil nitrogen dynamics from a climate manipulation experiment in a post-harvest forest. *Biogeochemistry* 2014, 121, 339–354. [CrossRef]

30. Sponseller, R.A. Precipitation pulses and soil CO\textsubscript{2} flux in a Sonoran Desert ecosystem. *Glob. Chang. Biol.* 2007, 13, 426–436. [CrossRef]

31. Kontopoulou, C.K.; Bilalis, D.; Pappa, V.A.; Rees, R.M.; Savvas, D. Effects of organic farming practices and salinity on yield and greenhouse gas emissions from a common bean crop. *Sci. Hortic.* 2015, 183, 48–57. [CrossRef]

32. Irshad, M.; Honna, T.; Yamamoto, S.; Eneji, A.E.; Yamasaki, N. Nitrogen mineralization under saline conditions. *Commun. Soil Sci. Plant Anal.* 2005, 36, 1681–1689. [CrossRef]

33. Rietz, D.D.; Haynes, R.J. Effects of irrigation-induced salinity and sodicity on soil microbial activity. *Soil Biol. Biochem.* 2003, 35, 845–854. [CrossRef]

34. Kim, D.G.; Vargas, R.; Bondlamberty, B.; Turetsky, M.R. Effects of soil rewetting and thawing on soil gas fluxes: A review of current literature and suggestions for future research. *Biogeosciences* 2012, 9, 2459–2483. [CrossRef]
35. Ma, L.J.; Zhang, W.; Zhang, N.; Zhang, X.; Li, F.C.; Zhang, H.M.; Mao, J.S.; Hou, Z.A. Effects of saline water on soil ammonia-oxidizing bacteria and enzyme activity in cotton field under drip irrigation condition. *Xinjiang Agric. Sci.* **2015**, *51*, 2038–2045.

36. Pereira, L.S.; Oweis, T.; Zairi, A. Irrigation management under water scarcity. *Agric. Water Manag.* **2002**, *57*, 175–206. [CrossRef]