The donor of carbon monoxide (CORM-2) affects the level of serum immunoglobulins and the state of the bone marrow during the immune response in mice

S. P. Beschasnyi, O. M. Hasiuk

Kherson State University, Ukraine

A – research concept and design; B – collection and/or assembly of data; C – data analysis and interpretation; D – writing the article; E – critical revision of the article; F – final approval of the article

Toxic carbon monoxide in small concentrations has pro-apoptotic, anti-allergic, vasodilator effects, and stimulates angiogenesis. The problem with the therapeutic use of low doses of carbon monoxide is that it is difficult to dose. To control the amount and gradual release of carbon monoxide, non-toxic preparation is used – CO donor based on carbonyl compound of ruthenium (CORM-2).

The aim — is to identify the effect of CORM-2 on the level of immunoglobulins in the blood serum and bone marrow of mice under conditions of inducing an immune response.

Materials and methods. 3 groups of 15 white laboratory mice each were formed. Induction of the immune response was due to the intraperitoneal administration of xenogenic red blood cells. The first experimental group on the first day of immunization received CORM-2 (20 mg/kg), group No. 2 – on the 5th day after immunization (period of the productive phase). The control group consisted of immunized animals that did not receive CORM-2. The amount of IgA, IgM, and IgG in blood serum was determined by ELISA on the 2nd and 6th day after immunization. At the end of the experiment, bone marrow cell populations were counted.

Results. After the injection of CORM-2 during the induction phase of the immune response, it inhibits the production of immunoglobulins. In comparison with the control, the level of IgA and IgG is reduced, but the amount of IgM remains unchanged. In the bone marrow, the number of monocytes, erythroblasts, and normoblasts, as well as lymphoblasts and plasma cells, increased. At the same time, the number of myeloblasts, myelocytes, basophilic normoblasts, and megakaryocytes decreased. The use of CORM-2 during the productive phase caused a decrease in the level of IgM and IgG with a simultaneous increase in the level of IgA. The number of neutrophils, eosinophils, monocytes, polychromophilic and oxyphilic normoblasts, lymphocytes, and plasma cells in the bone marrow increased. However, the number of myeloblasts, promyelocytes, myelocytes, metamyelocytes, basophilic normoblasts, and megakaryocytes decreased.

Conclusions. The impact of the CORM-2 on the inductive phase of the immune response leads to inhibition of the production of immunoglobulins. The injection of CORM-2 during the productive phase of the immune response decreased the level of IgM and IgG, but at the same time, an increase in the level of IgA was observed. After the injection of CORM-2, in the bone marrow, the number of monocytes, lymphocytes, and plasma cells increased. The results indicate that CORM-2 is able to modulate the immune response.

Key words: gas transmitter, carbon monoxide, CORM-2, humoral immune response.
риментальная группа в первый день иммунизации получила CORM-2 (20 мг/кг), группа № 2 – на 5 день после иммунизации (период продуктивной фазы). Контрольная группа – иммунизированные мыши, которые не получали CORM-2. В группе № 1 костный мозг получили CORM-2 в индукционную fazу иммунной ответа. В группе № 2 CORM-2 был введен в период продуктивной фазы иммунной реакции. В конце эксперимента подсчитывали популяции клеток костного мозга.

Результаты. CORM-2 в индукционную fazу иммунной ответа стимулирует продукцию иммуноглобулинов. Уменьшение количества IgM и IgG с одновременным повышением уровня IgA указывает на то, что CORM-2 способен стимулировать продукцию иммуноглобулинов в костном мозге. В конце эксперимента подсчитывали популяции клеток костного мозга.

Выводы. Введение CORM-2 в период продуктивной фазы иммунной реакции стимулирует продукцию иммуноглобулинов. Уменьшение количества IgM и IgG с одновременным повышением уровня IgA указывает на то, что CORM-2 способен стимулировать продукцию иммуноглобулинов в костном мозге.

Ключевые слова: газотрансмиттер, монооксид углерода, CORM-2, иммунный ответ.

Актуальные вопросы фармацевтической и медицинской науки и практики. 2020. Т. 13, № 3(34). С. 415–420

Among pharmacological drugs, there is a lot that has immunomodulatory properties [1]. These drugs can be divided into 2 groups: immunostimulants and immunosuppressants. The latter is widely used for the treatment of autoimmune diseases after an organ transplant procedure. Some antibiotics also belong to immunosuppressants, cytostatics, and hormonal drugs [1]. All of them cause adverse reactions, which increase with increasing dose. That is why search for drugs that can gently suppress the activity of the immune system and not cause catastrophic changes in other organs and systems of the body remains relevant. One of these applicants is a representative of the gas transmitter group – carbon monoxide (CO) [2,3].

CO is known as poison gas, however, in 1968, Tenhunen et al. reported that this gas is formed in the body during the breakdown of heme. After that, the period of studying the physiological effects of CO began [4,5]. Today, it is known that CO in a picomolar amount is able to suppress cell apoptosis [6], stimulates Ca2+ dependent K-channels [7], blocks T-cell proliferation [8], and is able to affect mitochondrial activity [9].
The only problem with using CO was that the dosage of this gas is difficult to carry out, poisoning is possible. In this regard, further studies of the therapeutic effects of CO are inhibited. This problem was solved by creating compounds—donors of CO [10]. After the introduction of such a compound into the body, a slow release of CO occurs. The results of the studies were performed by an overdose of isoflurane. Femur bones were harvested from mice, and the influence of CORM-2 on the level of immunoglobulins under conditions of antigenic stimulation was determined. To determine the amount of IgA, IgM, and IgG immunoglobulins, an enzyme-linked immunosorbent assay kit (“Gra num,” Ukraine) was used. The results of the study were performed using an ELISA analyzer Immunohem-21100, HTI (USA) on the 2nd and 6th day after immunization.

At the end of the experiment (6th day), euthanasia was performed by an overdose of isoflurane. Femur bones were isolated and bone marrow imprints were obtained. The obtained imprints were stained with Romanovsky–Giemsa paint and bone marrow cell populations were counted using a microscope.

All procedures with laboratory animals were carried out in compliance with the Council of Europe directives of the EU in 2010/63/EU and were approved by the bioethical commission of Kherson State University. Statistical processing was performed using the Mann–Whitney and Wilcoxon criteria. In this case, the result was considered reliable at P ≤ 0.05. The degree of correlation of indicators was calculated using the Pearson correlation coefficient.

**Results**

Comparison of serum immunoglobulin levels under conditions of induction of an immune response showed differences. In the control group, the level of IgA increased by 81.0 ± 4.0 %, IgM – by 35.3 ± 1.8 %, and IgG – by 47.0 ± 1.9 % (Fig. 1).

The result of the level of immunoglobulins in the serum of animals that were injected with CORM-2 on the first day of the immune response turned out to be interesting. In this group, the content of IgA increased by 100 ± 5 %, IgM remained at the same level, IgG increased only by 6.8 ± 0.3 % (Fig. 2).

But, after comparing this group with the control group, it was found that the level of IgA on the next day (after CORM-2 injection) was reduced by 22.0 ± 1.1 % (1.09 g/L versus 1.41 g/L of the control group). On the 6th day, it was reduced by 15.0 ± 0.75 % (2.18 g/L versus 2.56 g/L of the control group).

The IgM level (on the 2nd day) after comparison with the control group was reduced by 12.0 ± 0.6 % (1.49 g/L versus 1.7 g/L of the control) on the 6th day, it was reduced by 35.0 ± 1.75 % (1.49 g/L versus 2.3 g/L of the control group).

Similar results were obtained after comparing the IgG values. Its level was reduced on the 2nd day by 10.5 ± 0.5 % (9.32 g/L versus 10.42 g/L of the control). On the 6th day, the level was also reduced, but already by 35.0 ± 1.75 % (9.95 g/L versus 15.32 g/L of the control group).

The level of immunoglobulins in the serum of mice that were injected with CORM-2 in the productive period of the immune response was radically different from the other two (Fig. 3). The amount of IgA in serum increased by 67.0 ± 3.3 %, while IgM decreased by 48.7 ± 2.4 %, IgG decreased by 17.0 ± 0.8 %.

A study of bone marrow changes showed that the injection of CORM-2 influenced the proliferation of individual cell types after stimulation of the immune response (Fig. 4).

Compared with the control, in the group of animals to which CORM-2 was injected on the first day of the induction of the immune response (group No. 1), the number of myeloblasts decreased by 34.0 ± 1.7 %, myelocytes by 42.5 ± 1.7 %, basophilic normoblasts by 73.0 ± 3.6 %, megakaryocytes – by 76.0 ± 3.8 %. At the same time, there was an increase in the content of the following bone marrow elements: megakaryocytes by 76.0 ± 3.8 %, basophilic normoblasts by 73.0 ± 3.6 %, myelocytes by 42.5 ± 1.7 %, myeloblasts by 34.0 ± 1.7 %.
marrow cells: monocytes by 56.0 ± 2.8 %, erythroblasts by 57.0 ± 2.8 %, polychrome normoblasts by 171.0 ± 8.5 %, lymphocytes/lymphoblasts by 56.0 ± 2.8 %, plasma cells by 70.0 ± 3.5 %. Against this background, the content of promyelocytes, metamyelocytes, oxyphilic normoblasts, eosinophils, and basophils did not change.

In the bone marrow of mice that were injected with CORM-2 during the productive period of the immune response (group No. 2), a decrease in myeloblast levels by 25.0 ± 1.0 %, promyelocytes by 52.0 ± 2.6 %, myelocytes by 51.0 ± 2.4 %, and metamyelocytes by 58.0 ± 3.5 %, basophilic normoblasts by 83.0 ± 4.2 %, megakaryocytes – by 99.0 ± 4.9 % was observed. Moreover, the content of the following cells was increased: neutrophils by 42.0 ± 2.1 %, eosinophils by 178.0 ± 8.9 %, monocytes by 22.0 ± 1.1 %, polychromophilic normoblasts by 146.0 ± 7.3 %, oxyphilic normoblasts by 65.0 ± 3.2 %, lymphocytes/lymphoblasts by 73.0 ± 3.6 %, plasma cells by 35.0 ± 1.7 %. At the same time, the level of basophils and erythroblasts did not change (compared with the control group).

A correlation analysis revealed a direct, close relationship between immunoglobulin counts and a decrease in the population level of certain bone marrow cells. In group No. 1, a close correlation was observed between the level of serum immunoglobulins and the content of polychromic and oxyphilic normoblasts, lymphocytes/lymphoblasts, plasma cells, megakaryocytes.

In group No. 2, the level of immunoglobulins correlates with the levels of myeloblasts, basophilic normoblasts, polychrome, and oxyphilic normoblasts, lymphocytes, and plasma cells.

**Discussion**

The results indicate that intraperitoneal injection of CORM-2 directly affects the course of the immune response. CORM-2 inhibits the production of immunoglobulins during the induction phase of the immune response. In comparison with the control, the level of IgA and IgG was reduced, but the amount of IgM did not change. It is known that an increase in serum IgM is characteristic of the primary immune response stage. This indicates that CORM-2 is a promising anti-inflammatory agent. This property of CORM-2 indicates the possibility of its use in autoimmune diseases or to prevent the development of chronic inflammation.

The injection of CORM-2 during the productive phase of the immune response decreased the level of IgM and IgG, however, an increase in the level of IgA was observed at the same time (it is known that IgA is “responsible” for the so-called local immunity). This property of CORM-2 is promising to use in cases of immunodeficiency, reactions after transfusion measures, or medications that cause selective IgA deficiency.

Injection of CORM-2 during the inductive period of the immune response stimulated (or led to a delay in the bone marrow) separation of monocytes, erythroblasts and normoblasts, as well as lymphoblasts, and plasma cells. This was observed against a background of a decrease in the number of myeloblasts, myelocytes, basophilic normoblasts, and megakaryocytes. Injection of CORM-2 during the productive period also showed a decrease in the number of myeloblasts, promyelocytes, myelocytes, and plasma cells.
The donor of carbon monoxide (CORM-2) affects the level of serum immunoglobulins and the state of the bone marrow during the immune response...

normoblasts, and megakaryocytes. At the same time, the level of neutrophils, eosinophils, monocytes, polychromophilic and oxyphilic normoblasts, lymphocytes and plasma cells increased. Thus, it was found that CORM-2 stimulates an increase in the number of monocytes, lymphocytes, and plasma cells, regardless of the stage of the immune response.

After the action of CO, which is released from CORM-2, inducible heme oxygenase (HO-1) is activated. It should be noted that inhalation of CO does not increase the activity of HO-1 [14]. Its activity is enhanced only by CORM-2, which once again confirms that CORMs are suitable for these purposes.

Thus, the release of CO leads to the activation of the HMOX1 gene. Activation of this gene increases HO-1 activity. HO-1 is known to be a metabolite to fight inflammation. The anti-inflammatory ability of HO-1 is realized by increasing the expression of interleukin 10 (IL-10) and an antagonist of the pro-inflammatory interleukin 1 receptor (IL-1RA) [9].

After we stimulated the immune response and introduced anti-inflammatory CORM-2, HO-1, likely, affects the cells of the immune system, bone marrow stem cells. This mechanism may explain the findings of the study.

Conclusions

1. The injection of CORM-2 during the induction phase of the immune response enhances the production of IgA and IgG. The amount of IgM remained at the same level. An increase in the number of monocytes, erythroblasts, and normoblasts, as well as lymphoblasts and plasma cells, was observed in the bone marrow. At the same time, there was a decrease in the number of myeloblasts, myelocytes, basophilic normoblasts, and megakaryocytes.

2. During the productive phase of the immune response, CORM-2 lowered the level of IgM and IgG with a simultaneous increase in IgA level. The number of neutrophils, eosinophils, monocytes, polychromophilic and oxyphilic normoblasts, lymphocytes, and plasma cells in the bone marrow increased. However, there was a decrease in the number of myeloblasts, promyelocytes, myelocytes, metamyelocytes, basophilic normoblasts, and megakaryocytes.

Prospects for further research. In the future, a study of the properties of CORM-2 regarding the processes of tissue regeneration and healing will be conducted in vivo.

Funding

The research was performed within the research work of Kherson State University “Effects of certain vasoactive substances on central and peripheral lymphatic organs of white mice” state registration number 0117U001764.

Conflicts of interest: authors have no conflict of interest to declare.

Information about authors:
Beschasnyi S. P., PhD, Associate Professor of the Department of Human Biology and Immunology, Kherson State University, Ukraine.
ORCID ID: 0000-0002-7423-4112

Hasiuk O. M., PhD, Associate Professor, Head of the Department of Human Biology and Immunology, Kherson State University, Ukraine.
ORCID ID: 0000-0003-1055-2848

Відомості про авторів:
Бесчасний С. П., канд. біол. наук, доцент каф. біології людини та імунології, Херсонський державний університет, Україна.
Гасюк О. М., канд. біол. наук, доцент, зав. каф. біології людини та імунології, Херсонський державний університет, Україна.
Сведения об авторах:
Бесчасный С. П., канд. биол. наук, доцент каф. биологии человека и иммунологии, Херсонский государственный университет, Украина.
Гасюк Е. Н., канд. биол. наук, доцент, зав. каф. биологии человека и иммунологии, Херсонский государственный университет, Украина.

References

[1] Gladka, I. V., & Shkuropat, A. V. (2016). Efektyvnist khimichnykh ta biolohichnykh metodiv preventsi rozvytku bakterioziv plodiv Capsicum anuum [Effectiveness of chemical and biological methods of prevention of bacteriosis Capsicum anuum]. Pryrodonezyi almanakh. Seria: Biolohichni nauky, (23), 13-19. [in Ukrainian]. http://na.kspu.edu/index.php/na/article/view/462

[2] Fayad-Kobeissi, S., Ratovonantenaina, J., Dabiré, H., Wilson, J. L., Rodríguez, A. M., Berdeaux, A., Dubois-Randé, J., Mann, B., Motterlini, R., & Foresti, R. (2018). Vascular and angiogenic activities of CORM-401, an oxidant-sensitive CO-releasing molecule. Biochemical Pharmacology, 102, 64-77. https://doi.org/10.1016/j.bcp.2015.12.014

[3] Ji, X., Damera, K., Zheng, Y., Yu, B., Otterbein, L. E., & Wang, B. (2016). Toward Carbon Monoxide-Based Therapeutics: Critical Drug Delivery and Developability Issues. Journal of pharmaceutical sciences, 105(2), 406-416. https://doi.org/10.1016/j.xphs.2015.10.018

[4] Kolupaev, Yu. E., Karpets, Yu. V., Beschasnyi, S. P., & Dmitriev, A. P. (2019). Gasotransmitters and their role in adaptive reactions of plant cells. Cytology and Genetics. 53, 392-406. https://doi.org/10.3103/S0095452719050098

[5] Rose, J. J., Wang, L., Xu, Q., McTiernan, C. F., Shiva, S., Tejero, J., & Gladwin, M. T. (2017). Carbon Monoxide Poisoning: Pathogenesis, Management, and Future Directions of Therapy. American journal of respiratory and critical care medicine, 195(5), 596-606. https://doi.org/10.1164/ajrccm.201606-1275CC

[6] Olas, B. (2014). Carbon monoxide is not always a poison gas for human organism: Physiological and pharmacological features of CO. Chemico-biological interactions, 222, 37-43. https://doi.org/10.1016/j.cbi.2014.08.005

[7] Beschasnyi, S., & Hasiuk, O. (2020). CO-Releasing Molecule (CORM-2) in the Regulation of Ca2+-Dependent K+-Permeability of Erythrocyte. Ukrainian Journal of Medicine, Biology and Sport, 5(2), 166-171. https://doi.org/10.26693/jmsb05.02.166

[8] Park, J., Joe, Y., Ryter, S. W., Suh, Y. J., & Chung, H. T. (2019). Similarities and Distinctions in the Effects of Metformin and Carbon Monoxide in Immunometabolism. Molecules and cells, 42(4), 292-300. https://doi.org/10.14348/molcells.2019.0016

[9] Motterlini, R., & Foresti, R. (2017). Biological signaling by carbon monoxide and carbon monoxide-releasing molecules. American journal of physiology. Cell physiology, 312(3), C302-C313. https://doi.org/10.1152/ajpcell.00380.2016

[10] Ryter, S. W., & Choi, A. M. (2016). Targeting heme oxygenase-1 and carbon monoxide for therapeutic modulation of inflammation. Translational research : the journal of laboratory and clinical medicine, 167(1), 7-34. https://doi.org/10.1016/j.trsl.2015.06.011

[11] Adach, W., & Olas, B. (2017). The role of CORM-2 as a modulator of oxidative stress and hemostatic parameters of human plasma in vitro. PLoS one, 12(8), e0184787. https://doi.org/10.1371/journal.pone.0184787

[12] Magerowski, M., Magierowska, K., Hubalewska-Mazgaj, M., Siłłowski, Z., Ginter, G., Pąjdó, R., Chmura, A., Kwiecien, S., & Brzozowski, T. (2017). Carbon monoxide released from its pharmacological donor, tricarbonyldichlororuthenium (II) dimer, accelerates the healing of pre-existing gastric ulcers. British journal of pharmacology, 174(20), 3654-3668. https://doi.org/10.1111/bph.13968

[13] Tashineva, L. A., Starikova, E. G., Novickij, V. V., & Rjazancova, N. V. (2012). Vnutrikletochnye misheni proapoptoticheskogo vliyanija gazovyh transmitterov [Intracellular targets of proapoptotic influence of gaseous transmitters]. Annals of the Russian academy of medical sciences, 67(10), 77-81. [in Russian]. https://doi.org/10.15690/vramn.v67i10.420

[14] Starikova, Ye. G. (2012). Antiproliferativnyi potencial monooksida uglidera [Antiproliferative potential of carbon monoxide]. Bulletin of Siberian Medicine, 11(4), 68-71. [in Russian]. https://doi.org/10.26693/1862-9833-2012-4-68-71

[15] Mahan V. L. (2020). Cardiac function dependence on carbon monoxide. Medical gas research, 10(1), 37-46. https://doi.org/10.4103/2045-9912.279982