Safety and efficacy of a new spot-on formulation of selamectin plus sarolaner in the treatment and control of naturally occurring flea infestations in cats presented as veterinary patients in Australia

Raj Packianathan¹*, Melissa Pittorino², Andrew Hodge¹, Natalie Bruellke¹ and Kelly Graham³

Abstract

Background: The safety and efficacy of a new spot-on formulation of selamectin plus sarolaner were evaluated for the treatment and control of natural flea infestations on cats in two non-randomised, multi-centre clinical trials conducted in 8 different locations in Queensland, Australia.

Methods: One hundred and four cats from 65 different households were enrolled across the two studies. Demographic characteristics of cats in the two studies were similar. The new spot-on formulation of selamectin and sarolaner was administered topically once a month for 3 consecutive months at a minimum dosage of 6 mg/kg selamectin (dose range 6–12 mg/kg) plus 1 mg/kg sarolaner (dose range 1–2 mg/kg). Cats were dosed on Days 0 (pre-treatment), 30 and 60 and physical examinations and flea counts were conducted on Days 0, 30, 60 and 90. Efficacy assessments were based on the percentage reduction in live flea counts post-treatment compared to Day 0.

Results: In Study A, at enrolment, primary cats had flea counts ranging from 6 to 107 (arithmetic mean 21.0). The selamectin and sarolaner spot-on formulation resulted in arithmetic mean efficacy of 98.0%, 100% and 100% on Days 30, 60 and 90, respectively. In Study B, at enrolment, primary cats had flea counts ranging from 6 to 22 (arithmetic mean 10.0). The selamectin and sarolaner spot-on formulation resulted in arithmetic mean efficacy of 99.7%, 100% and 100% on Days 30, 60 and 90, respectively.

Conclusions: The new spot-on formulation of selamectin plus sarolaner topically administered at monthly intervals at the minimum dosage of 6.0 mg/kg selamectin and 1.0 mg/kg sarolaner was safe and highly effective against natural infestations of fleas under a range of geographical conditions, representative of both tropical and subtropical regions of Australia.

Keywords: Cats, Ctenocephalides felis, Ectoparasites, Efficacy, Field study, Flea, Isoxazoline, Parasiticide, Sarolaner, Selamectin, Revolution® Plus, Topical
Background

Ctenocephalides felis is the dominant flea species affecting cats in Australia [1–3]. Recent studies have confirmed the genetic diversity among the Australian isolates of C. felis with at least three distinct phylogenetic clades identified using mitochondrial DNA markers [4]. Flea infestations cause flea allergy dermatitis, pruritus and anaemia, especially in young cats, but can also transmit a number of zoonotic pathogens such as Rickettsia felis, Bartonella clarridgeiae [5–8] and Bartonella henselae [9]. Flea infestations can occur all year round with a higher incidence during the warmer months. Therefore, flea control throughout the year, including the winter months, is essential to control flea infestation and prevent transmission of pathogens such as R. felis [3, 8]. Optimal flea control requires an integrated approach focusing on controlling fleas on the cats as well as controlling flea reproduction in the environment, killing the fleas before they can lay eggs [1, 3, 10, 11]. Historically, various classes of insecticides with different modes of action have been used for flea control. Some of these products also have an extended spectrum of activity against other ecto- and endoparasites in cats [3] including selamectin, as example of a compound in the macrocyclic lactone class of endectocides, which provides effective control against C. felis [12, 13], biting lice (Felicola subsaurostratus), ear mites (Otodectes cynotis), heartworm (Dirofilaria immitis) and intestinal hookworm (Ancylostoma tubaeforme) and roundworm (Toxocara catti) in cats, but lacks substantial activity against ticks at the labelled dosage [14, 15].

Sarolaner is an oxazoline with a wide spectrum of activity against ectoparasites such as fleas, ticks including the Australian paralysis tick, Ixodes holocyclus [16], and mites including Sarcoptes scabiei var. canis, Demodex canis and Otodectes cynotis in dogs [17, 18]. To broaden the spectrum of activity and provide effective control against both endo- and ectoparasites in cats, a new spot-on formulation was developed with selamectin and sarolaner. This is currently marketed in the USA and Europe as Revolution® Plus or Stronghold® Plus (Zoetis, New Jersey, USA), respectively. The new spot-on formulation has demonstrated efficacy against heartworm [19], fleas [20, 21], various tick species in Europe [22–24] and the USA [25, 26], intestinal hookworms and roundworms [27, 28], and ear mites [29].

Two multi-centric clinical field studies were conducted to evaluate the safety and efficacy of the new spot-on formulation of selamectin plus sarolaner against natural flea infestations in cats presented as veterinary patients under Australian field conditions.

Methods

The studies were conducted in accordance with the World Association for the Advancement of Veterinary Parasitology (WAAVP) guidelines for evaluating the efficacy of parasiticides for the treatment, prevention and control of flea and tick infestation on dogs and cats [30] and principles of Good Clinical Practice (GCP) [31]. The studies were approved by the Community Access Animal Ethics Committee of the Department of Agriculture and Fisheries Queensland.

Study locations

The field studies were conducted in subtropical (Study A: Southern part of Queensland) and tropical (Study B: Northern part of Queensland) regions in Queensland, Australia [32]. The location and number of veterinary clinics where cases were enrolled in the two studies are summarised in Fig. 1.

Animals

Client-owned cats presented at veterinary clinics or in-house visits by the veterinary investigators were used in the studies. Cats were from diverse households and lived both indoors and outdoors. Cats came from both single cat households and households with multiple cats (maximum of 3 cats) and/or dogs. There were no breed or sex restrictions, but cats intended for breeding or that were pregnant or lactating were not eligible for enrolment. For inclusion in the study, at least one cat in the household had to harbor at least 6 live fleas at screening. All cats were at least 8 weeks of age and ≥ 1.25 kg in body weight at enrolment. Cats in the study were not allowed to have been treated with any ectoparasiticide with persistent activity within 30 days or with a short-acting ectoparasiticide within 14 days of the first treatment. When these studies were conducted, there were no long acting ectoparasiticides in the Australian market for use in cats.

Experimental design

Both studies followed the same experimental design; however, studies A and B were analysed and reported separately, as they represented different geographical regions. Each study was conducted as a multi-centric, single group, non-randomised trial. Within each household, the primary cat was the first cat in the household with ≥ 6 fleas counted on Day 0 and up to 2 additional cats were enrolled as supplementary cats. Only the primary cats were included in the efficacy evaluation whereas all cats were included in the safety evaluation.

All cats were confirmed to be in good general health prior to enrolment based on the physical examination performed by a veterinarian. Cats were housed and maintained under their normal home conditions for the
duration of the study. No additional products that had activity against fleas (including systemic, premises, and/or over-the-counter treatments including insecticidal shampoos or collars) were permitted to be used on any animal in the household for the duration of the study. Any concomitant medications used during the study were recorded along with any abnormal health events.

**Treatment administration**

Cats received three consecutive monthly treatments on study days 0, 30 and 60. Day 0 was set as the day the primary cat in each household received the first treatment. For the follow-up treatments on Days 30 and 60, the visits were allowed to deviate by ±3 days of the target date. Treatments were dispensed by the veterinarian. Treatment dispensing was based upon the most recent body weight recorded prior to each treatment. Cats were topically dosed with the recommended label dosage of the spot-on formulation containing 6.0–12.0 mg/kg selamectin and 1.0–2.0 mg/kg sarolaner. After each treatment, cats were not permitted to be bathed for 24 hours or groomed throughout the study.

**Flea counts**

Flea counts on primary cats were conducted prior to treatment on Days 0, 30 and 60 as well as on Day 90 (the post-treatment evaluations could be conducted ±3 days of the target day) by a qualified and/or experienced veterinary staff.

Flea counts were conducted by personnel trained to a standardized methodology. The cats were combed with a fine-toothed flea comb that was uniquely identified for each cat. The combing proceeded in a systematic manner to ensure all areas of the cat were combed. Each cat was
examined for at least 10 min. If any fleas were found, the examination was continued in 1-min increments until no fleas were encountered. Once combing was completed, all fleas were placed back on the cat. Fleas maintaining an upright orientation or moving in a coordinated manner were considered to be alive. Only live flea counts were recorded.

Statistical analysis
Data were summarised and analysed for each of the two studies separately, using SAS version 9.3 (SAS Institute Inc., Cary, NC, USA). The individual animal (primary cat) was the experimental unit for the efficacy analysis. Data were excluded from the efficacy analysis following protocol deviations such as incorrect dosing or where dosing or flea counts were not conducted within ±3 days of the target day (after Day 0). Flea counts were summarised at each time point using arithmetic means and ranges. Percent effectiveness with respect to the baseline was calculated at each time point using the formula 
\[
\left(\frac{C - T}{C}\right) \times 100,
\]
where C is the pre-treatment arithmetic mean and T is the post-treatment arithmetic mean.

Results
Demographics
Enrolment and demographic characteristics are summarised in Table 1.

Study A
A total of 31 primary cats were enrolled across 3 different clinics in the southern part of Queensland. Most of the primary cats were crossbred (n = 30; 97%) and most were neutered (n = 24; 77%), with similar numbers of males (n = 17) and females (n = 14). Out of 31 households, 10 were single cat households and 21 had additional dogs or cats; 11 households had both dogs and cats. All dogs in the households were treated for fleas with ‘Nexgard (afoxolaner) Chewable for dogs’ (Boehringer Ingelheim Pty Ltd, New South Wales, Australia). The ages of the primary cats ranged from 10 weeks to 13 years, with a mean age of 4.7 years and weighed 1.3–7.8 kg. Eighty-four percent (n = 26) of primary cats had short or medium hair length and the majority (n = 22; 71%) had outdoor access, with only 29% (n = 9) living mainly indoors. Of the 31 primary cats, 2 cats did not complete the study due to owner non-compliance.

Study B
A total of 34 primary cats were enrolled across 5 different clinics (including mobile veterinary clinic investigator visiting households) in the northern part of Queensland. Of the 34 primary cats, 2 (6%) were purebred and 32 (94%) were crossbred. Approximately half of the primary cats were neutered (n = 16; 47%), with similar numbers of males (n = 19) and females (n = 15). The ages of the primary cats ranged from 10 weeks to 13 years, with a mean age of 3.2 years and weighed 1.3–6.5 kg. Ninety-four percent (n = 32) of primary cats had short or medium hair length and the majority (n = 21; 62%) had outdoor access, with only 38% (n = 13) living mainly indoors. Out of 34 households, 9 were single cat household only and 25 had additional dogs or cats; 23 households had both dogs and cats. All dogs in the households were treated for fleas with ‘Nexgard (afoxolaner) Chewable for dogs’ (Boehringer Ingelheim Pty Ltd). Of the 34 primary cats, six cats did not complete the study due to owner non-compliance and other reasons.

Safety
In study A and B, a total of 55 and 49 and cats were enrolled for safety evaluation, respectively. A total of nine adverse events in Study A and two in Study B were recorded. Most of the observed adverse events in both studies were sporadic in nature and typical of those commonly seen in the general cat population such as skin infections and gastrointestinal and ocular disorders. One cat (6-year-old, neutered female) in Study A was reported to have vomiting from 2 hours after the second monthly treatment application and vomiting continued for 4 days and the cat was presented at the veterinary clinic for

| Table 1: Demographic characteristics of primary cats enrolled in two clinical field studies in Australia and treated with selamectin (6–12 mg/kg) plus sarolaner (1–2 mg/kg) |
|----------------------------------------|
| Characteristic | Study A (3 clinics, N = 31) | Study B (5 clinics, N = 34) |
| n (%) | n (%) |
| Breed | Purebred | 1 (3) | 2 (6) |
| | Non-purebred | 30 (97) | 32 (94) |
| Living condition | Indoors and outdoors | 17 (55) | 15 (44) |
| | Mostly indoors | 9 (29) | 13 (38) |
| | Mostly outdoors | 5 (16) | 6 (18) |
| Sex | Male | 17 (55) | 19 (56) |
| | Female | 14 (45) | 15 (44) |
| Neutered | Yes | 24 (77) | 16 (47) |
| | No | 7 (23) | 18 (53) |
| Hair type | Long | 5 (16) | 2 (6) |
| | Medium | 6 (19) | 7 (21) |
| | Short | 20 (65) | 25 (74) |
examination 4 days after treatment. Physical examination, blood biochemistry and complete blood counts were performed, and no clinically significant findings were noted. The cat was treated with maropitant injection and the condition resolved without further treatment. Since there was no other attributable cause of vomiting, the vomiting episode was deemed to possibly be related to the treatment. There were no other adverse events assessed as treatment-related in any of the other selamectin and sarolaner spot-on formulation-treated cats.

### Efficacy

#### Study A
At enrolment, primary cats had flea counts ranging from 6 to 107 (arithmetic mean = 21.0). The selamectin and sarolaner spot-on formulation resulted in arithmetic mean efficacy of 98.0%, 100% and 100% on Days 30, 60 and 90, respectively (Table 2).

#### Study B
At enrolment, primary cats had flea counts ranging from 6 to 22 (arithmetic mean = 10.0). The selamectin and sarolaner spot-on formulation resulted in arithmetic mean efficacy of 99.7%, 100% and 100% on Days 30, 60 and 90, respectively (Table 2).

### Discussion

One hundred and four cats from 65 different households were enrolled across the two studies in the subtropical and tropical regions of Australia (Fig. 1). Demographic characteristics of cats in the two studies were generally similar. The new spot-on formulation of selamectin and sarolaner administered topically once a month for 3 consecutive months at a minimum dosage of 6 mg/kg (dose range 6–12 mg/kg) selamectin plus 1 mg/kg (dose range 1–2 mg/kg) sarolaner resulted in excellent treatment and control of naturally occurring flea infestations on client-owned cats. The new spot-on formulation of selamectin and sarolaner was well tolerated, with the observed abnormal clinical signs consistent with conditions most commonly seen in the general cat population and not related to study treatment, except in one cat where vomiting was possibly related to the treatment.

The treatment with selamectin and sarolaner spot-on provided 98.0% (Study A) and 99.7% (Study B) mean efficacy at 30 days after the first treatment and these findings were consistent with previous data reported in overseas field studies in the USA and Europe [20, 22] where following monthly administration on Days 30 and 60, the new spot-on formulation provided 100% efficacy against fleas for 30 days as reported previously [20–22]. Under field conditions, cats are continuously exposed to new flea infestations and therefore the rapid speed of kill is critical in reducing the flea burden on the cats as well as breaking the flea life-cycle and halting their development in the environment [3, 33].

Although identification of fleas in the field was not conducted in these studies, it is well documented that *C. felis* is the most common cause of flea infestations in cats in Australia [2]. Using the mtDNA sequencing of cytochrome c oxidase subunits, different haplotype clades of *C. felis* were identified in different parts of Australia [34]. Although the insecticidal efficacy against different haplotype clades of *C. felis* is unknown, a proportion of enrolled cats in both studies (Fig. 1) came from the regions where the different haplotype clades of *C. felis* have been reported [34].

*Ctenocephalides felis* has been reported to develop resistance to some of the older classes of ectoparasiticides [1], but the lack of or reduced efficacy of flea products reported in the field is generally most likely due to poor owner compliance, high environmental flea burden, poor understanding of flea biology or different susceptibility of fleas [3, 10, 13, 35]. Inclusion of sarolaner in the new spot-on formulation will complement the pulicidal activity of selamectin as well as provide effective control

### Table 2 Flea counts, ranges of counts and arithmetic mean (AM) efficacy at each time point for primary cats treated with selamectin (6–12 mg/kg) plus sarolaner (1–2 mg/kg) in two clinical field studies in Australia

| Day of study | Study A | Study B |
|--------------|---------|---------|
| Day 0 | | |
| No. of animals | 31 | 34 |
| Arithmetic mean count | 21.0 | 10.0 |
| Range of counts | 6–107 | 6–22 |
| Day 30 | | |
| No. of animals | 29 | 29 |
| Arithmetic mean count | 0.4 | 0.0 |
| Range of counts | 0–4 | 0–1 |
| AM efficacy (%) | 98.0 | 99.7 |
| Day 60 | | |
| No. of animals | 27 | 28 |
| Arithmetic mean count | 0.0 | 0.0 |
| Range of counts | 0–0 | 0–0 |
| AM efficacy (%) | 100 | 100 |
| Day 90 | | |
| No. of animals | 21 | 26 |
| Arithmetic mean count | 0.0 | 0.0 |
| Range of counts | 0–0 | 0–0 |
| AM efficacy (%) | 100 | 100 |

* Efficacy calculated based on comparison to arithmetic mean on Day 0
against ticks [22–24]. The new spot-on formulation was also demonstrated to have rapid speed of kill following administration thus killing the fleas before they can lay eggs [20, 21], and was effective in reducing the environmental flea burden [36] thereby reducing the risk of spreading vector-borne diseases caused by R. felis and Bartonella spp. [5–9, 37].

Conclusions
The new spot-on formulation of selamectin plus sarolaner topically administered at monthly intervals at the minimum dosage of 6.0 mg/kg selamectin and 1.0 mg/kg sarolaner, was well tolerated and highly effective against natural infestations of fleas under a range of Australian conditions.

Acknowledgements
The authors are grateful for the dedication of the veterinarians and their clinic staff involved in the study and to owners for their participation.

Authors’ contributions
All authors assisted with the design and conduct of the study and interpretation of the data. Manuscript was written by RP and revised by AH, NB, KG and MP. All authors read and approved the final manuscript.

Funding
This study was funded by Zoetis Australia Research and Manufacturing Pty Ltd, Level 6, 5 Rider Boulevard, Rhodes, NSW 2138, Australia. Eurofins Animal Health, Unit F10, 16 Mars Road, Lane Cove West NSW 2066, Australia was an independent Contract Research Organisation responsible for the conduct and management of the studies.

Availability of data and materials
Relevant datasets generated and/or analysed during these studies are included within the article.

Ethics approval and consent to participate
The studies were approved by the Community Access Animal Ethics Committee of the Department of Agriculture and Fisheries Queensland, Australia on 01 September 2016 prior to the beginning of the study. The animal ethics approval numbers were CA 2016/08/990 (Study A) and CA 2016/08/991 (Study B). This study did not report on any data related to humans. All cats enrolled in the study were client-owned cats presented at veterinary clinics or in-house visits by the veterinary investigators in different parts of Australia. Informed owner consent was obtained prior to enrolment of all study cats.

Consent for publication
Not applicable.

Competing interests
RP, AH, NB and KG are Zoetis employees. MP was a contracted study investigator.

Author details
1 Veterinary Medicine Research and Development, Zoetis Australia Research and Manufacturing Pty Ltd, Level 6, 5 Rider Boulevard, Rhodes, NSW 2138, Australia. 2 Eurofins Animal Health, Unit, F10, 16 Mars Road, Lane Cove West, NSW 2066, Australia. 3 Zoetis, Level 6, 5 Rider Boulevard, Rhodes, NSW 2138, Australia.

Received: 20 November 2019 Accepted: 25 April 2020 Published online: 06 May 2020

References
1. Rust MK, Dryden MW. The biology, ecology, and management of the cat flea. Annu Rev Entomol. 1997;42:451–73.
2. Slapeta J, King J, Mcdonell D, Malik P, Homer D, Hannan P, et al. The cat flea (Ctenocephalides felis felis) is the dominant flea on domestic dogs and cats in Australian veterinary practices. Vet Parasitol. 2011;180:383–8.
3. Burton G, Shipstone M, Burrows M. Veterinary guidelines for the control of fleas in dogs and cats in Australia. Aust Vet Pract. 2003;33:117–24.
4. Ckrcvnic N, Slapeta J. Climate change models predict southerly shift of the cat flea (Ctenocephalides felis) distribution in Australia. Parasites Vectors. 2019;12:137.
5. Hii S-F, Abdad MY, Kopp SR, Stenos J, Rees RL, Traub RJ. Seroprevalence and risk factors for Rickettsia felis exposure in dogs from Southeast Queensland and the Northern Territory, Australia. Parasites Vectors. 2013;6:159.
6. Barns VR, Beatty JA, Wilson BJ, Evans N, Govan R, Baral RM, et al. Prevalence of Bartonella species, Rickettsia felis, haemoplasmas and the Ehrlichia group in the blood of cats and fleas in eastern Australia. Aust Vet J. 2010;88:160–5.
7. Schloeder D, Owen H, Clark P, Stenos J, Fernwick SG. Rickettsia felis in fleas, Western Australia. Emerg Infect Dis. 2006;12:841–3.
8. Teoh YT, Hi SF, Stevenson MA, Graves S, Rees R, Stenos J, et al. Serological evidence of exposure to Rickettsia felis and Rickettsia typhi in Australian veterinarians. Parasites Vectors. 2017;10:129.
9. Chomel BB, Boulouis H-J, Maruyama S, Breitschwerdt EB. Bartonella spp. in pets and effect on human health. Emerg Infect Dis. 2006;12:389–94.
10. Siak M, Burrows M. Flea control in cats: new concepts and the current armoury. J Feline Med Surg. 2013;15:31–40.
11. Dryden M, Payne P, Smith V. Efficacy of selamectin and fipronil-(S)-methoprene spot-on formulations applied to cats against adult cat fleas (Ctenocephalides felis), flea eggs, and adult flea emergence. Vet Ther. 2007;8:255–62.
12. Mctier TL, Evans NA, Martin-Short M, Graton K. Comparison of the activity of selamectin, fipronil, and imidacloprid against flea larvae (Ctenocephalides felis felis) in vitro. Vet Parasitol. 2003;116:45–50.
13. Ritzhaupt LK, Rowan TG, Jones RL, Cracknell VC, Murphy MG, Shankis DJ. Evaluation of the comparative efficacy of selamectin against flea (Ctenocephalides felis felis) infestations on dogs and cats in simulated home environments. Vet Parasitol. 2002;106:165–75.
14. Fisher MA, Shankis DJ. A review of the off-label use of selamectin (Stronghold/Revolution) in dogs and cats. Acta Vet Scand. 2008;50:1–5.
15. Bishop BF, Bruce CI, Evans NA, Goudie AC, Graton KA, Gibson SP, et al. Selamectin: a novel broad-spectrum endectocide for dogs and cats. Vet Parasitol. 2000;91:163–76.
16. Packianathan R, Hodge A, Bruelleke N, Davis K, Maeder S. Comparative speed of kill of sarolaner (Simparica®) and afoxolaner (NexGard®) against induced infestations of Ixodes holocyclus on dogs. Parasites Vectors. 2017;10:98.
17. Mctier TL, Chubb N, Curtis MP, Hedges L, Inskeep GA, Knauer CS, et al. Discovery of sarolaner: a novel, orally administered, broad-spectrum, isoxazoline ectoparasiticide for dogs. Vet Parasitol. 2016;222:3–11.
18. Packianathan R, Colgan S, Hodge A, Davis K, Six RH, Maeder S. Efficacy and safety of sarolaner (Simparica®) in the treatment and control of naturally occurring flea infestations in dogs presented as veterinary patients in Australia. Parasites Vectors. 2017;10:367.
19. Mctier TL, Pullins A, Chapman S, Rugg J, Von Rettenstein M, Mccall JW, et al. The efficacy of a novel topical formulation of selamectin plus sarolaner (Revolution® Plus/Stronghold® Plus) in preventing the development of Dirofilaria immitis in cats. Vet Parasitol. 2019;270:56–62.
20. Becskai C, Cherri JA, Vatta AF, King VL, Lin D, Rugg D. Efficacy and speed of kill of a new spot-on formulation of selamectin plus sarolaner against flea infestations in cats. Vet Parasitol. 2017;238:518–21.
21. Vatta AF, Everett WR, Holzmer SJ, Cherri JA, King VL, Rugg D, et al. Efficacy of a new spot-on formulation of selamectin plus sarolaner for cats against adult Ctenocephalides felis, flea egg production and adult flea emergence. Vet Parasitol. 2017;238:522–6.
23. Becskei C, Lin D, Rugg D, Geurden T. Speed of kill of a new spot-on formulation of selamectin plus sarolaner for cats against induced infestations with *Ixodes ricinus*. Vet Parasitol. 2017;238:S8–11.

24. Geurden T, Botowski S, Wozniakiewicz M, King V, Fournie J, Liebenberg J. Comparative efficacy of a new spot-on combination product containing selamectin and sarolaner (Stronghold®Plus) versus fluralaner (Bravecto®) against induced infestations with *Ixodes ricinus* ticks on cats. Parasites Vectors. 2017;10:319.

25. Vatta AF, Young DR, Everett WR, King VL, Cherni JA, Von Reitzenstein M, et al. Efficacy of a new topical formulation containing selamectin plus sarolaner against three common tick species infesting cats in the United States. Vet Parasitol. 2019;270:S19–25.

26. Vatta AF, Everett WR, Cherni JA, King VL, Rugg D. The speed of kill of a topical combination of selamectin plus sarolaner against induced infestations of *Ixodes scapularis* ticks on cats. Vet Parasitol. 2019;270:S26–30.

27. Vatta AF, Myers MR, Bowman DD, Rugg JJ, Damrah L, Thien C, et al. Efficacy and safety of a new topical formulation of selamectin plus sarolaner in the treatment and control of natural infections of *Ancylostoma tubaeforme* and *Toxocara cati* in cats presented as veterinary patients in the United States. Vet Parasitol. 2019;270:S45–51.

28. Geurden T, Vatta AF, Slootmans N, King VL, Lin D, Mctier T, Rugg D. Efficacy of a new spot-on formulation of selamectin plus sarolaner against *Ancylostoma tubaeforme* and *Toxocara cati* in cats. Vet Parasitol. 2019;270:S31–5.

29. Becskei C, Reinemeyer C, King VL, Lin D, Myers MR, Vatta AF. Efficacy of a new spot-on formulation of selamectin plus sarolaner in the treatment of *Otodectes cynotis* in cats. Vet Parasitol. 2017;238:S27–30.

30. Marchiondo AA, Holdsworth PA, Fournie L, Rugg D, Hellmann K, Snyder DE, et al. World Association for the Advancement of Veterinary Parasitology (W.A.A.V.P) second edition: guidelines for evaluating the efficacy of parasiticides for the treatment, prevention and control of flea and tick infestations on dogs and cats. Vet Parasitol. 2013;194:84–97.

31. VICH GL9. Guideline on good clinical practices. 2001. VICH Topic GL9: https://www.vichsec.org/en/guidelines/pharmaceuticals/pharma-efficacy/good-clinical-practice. Accessed 14 Nov 2019.

32. Teoh YT, Hsi SF, Graves S, Rees R, Stenos J, Traub RJ. The epidemiology of *Rickettsia felis* infecting fleas of companion animals in eastern Australia. Parasites Vectors. 2018;11:138.

33. Dryden M, Payne P, Lowe A, Mailen S, Smith V, Rugg D. Efficacy of a topically applied formulation of metaflumizone on cats against the adult cat flea, flea egg production and hatch, and adult flea emergence. Vet Parasitol. 2007;150:263–7.

34. Lawrence AL, Brown GK, Peters B, Spielman DS, Morin-Adeline V, Slapeta J. High phylogenetic diversity of the cat flea (*Ctenocephalides felis*) at two mitochondrial DNA markers. Med Vet Entomol. 2014;28:330–6.

35. Rust MK. Insecticide resistance in fleas. Insects. 2016;7:1–9.

36. Dryden MW, Canfield MS, Bocon C, Phan L, Niedfeldt E, Kinnon A, et al. In-home assessment of either topical fluralaner or topical selamectin for flea control in naturally infested cats in West Central Florida, USA. Parasites Vectors. 2018;11:422.

37. Chomel B. Tick-borne infections in dogs—an emerging infectious threat. Vet Parasitol. 2011;179:294–301.

**Publisher’s Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.