**Mass Death of Predatory Carp, *Chanodichthys erythropterus*, Induced by Plerocercoid Larvae of *Ligula intestinalis* (Cestoda: Diphyllobothriidae)**

Woon-Mok Sohn¹*, Byong-Kuk Na¹, Soo Gun Jung², Koo Hwan Kim³

¹Department of Parasitology and Tropical Medicine, and Institute of Health Sciences, Gyeongsang National University School of Medicine, Jinju 52727, Korea; ²Korea Federation for Environmental Movements in Daegu, Daegu 41259, Korea, ³Nakdong River Integrated Operations Center, Korea Water Resources Corporation, Busan 49300, Korea

**Abstract:** We describe here the mass death of predatory carp, *Chanodichthys erythropterus*, in Korea induced by plerocercoid larvae of *Ligula intestinalis* as a result of host manipulation. The carcasses of fish with ligulid larvae were first found in the river-edge areas of Chilgok-bo in Nakdong-gang (River), Korea at early February 2016. This ecological phenomena also occurred in the adjacent areas of 3 dams of Nakdong-gang, i.e., Gangjeong-bo, Dalseong-bo, and Haecheon-Changnyeong-bo. Total 1,173 fish carcasses were collected from the 4 regions. To examine the cause of death, we captured 10 wondering carp in the river-edge areas of Haecheon-Changnyeong-bo with a landing net. They were 24.0-28.5 cm in length and 147-257 g in weight, and had 2-11 plerocercoid larvae in the abdominal cavity. Their digestive organs were slender and empty, and reproductive organs were not observed at all. The plerocercoid larvae occupied almost all spaces of the abdominal cavity under the air bladders. The proportion of larvae per fish was 14.6-32.1% of body weight. The larvae were ivory-white, 21.5-63.0 cm long, and 6.0-13.8 g in weight. We suggest that the preference for the river-edge in infected fish during winter is a modified behavioral response by host manipulation of the tapeworm larvae. The life cycle of this tapeworm seems to be successfully continued as the infected fish can be easily eaten by avian definitive hosts.

**Key words:** *Ligula intestinalis*, plerocercoid, host manipulation, predatory carp, *Chanodichthys erythropterus*

---

*Ligula intestinalis* (Cestoda: Diphyllobothriidae) is one of the tapeworm species inhabiting the intestine of fish-eating birds. This tapeworm is widely distributed in the world, and has a complicate 3-host life cycle like *Diphyllobothrium* and *Spirometra* spp. [1]. Unembryonated eggs discharged in the feces of avian hosts develop and hatch out in the water as the coracidium. The ciliated coracidia freely swimming in the water are eaten by the first intermediate host, i.e., crustacean copepods, and then develop into procercooids in the abdominal cavity of copepods within 2-3 weeks. Copepods with procercooids are eaten by the second intermediate hosts, zooplankton-eating fish. Infected procercooids develop into plerocercoids in the abdominal cavity of the fish hosts. The plerocercoid larvae stay more than 10 months in the fish host, whereas in the definitive host, fish-eating birds, the larvae quickly mature into adults having gravid proglottids which stay no longer than 5-6 days [1,2].

There have been many studies on the infection status of ligulid larvae in the second intermediate fish hosts [2-8]. However in the Republic of Korea (Korea), only 1 study was performed. Moreover, it was a brief manuscript submitted to the 38th Korea Science Exhibition by Ryu and Lee [9]. After that, a local news paper reported that more than 80% of pale chubs, *Zacco platypus*, in Hoengseong-dam were highly infected with these tapeworm larvae in December 2003. On the other hand, it has been known that the plerocercoid larvae of *L. intestinalis* induce endocrine disruption, in particular, parasitic castration [10-14], growth suppression, morphological modification [15,16], and mortality increase in the fish [17,18] as well as modified behavioral responses [4,19-22]. Recently, we observed a mass death of predatory carp, *Chanodichthys erythropterus*, infected with the plerocercoid larvae of *L. intestinalis* in the 4 sites of Nakdong-gang (‘gang’ means river). Thus, in the present study, we intended to describe this exceptional ecologi-
The predatory carp, *Chanodichthys erythropterus*, found in the river-edge areas of Chilgok-bo in Nakdong-gang (River), Korea. (A) A slowly swimming carp with an extruding larva. (B) An exhausted one just before dying with an escaping larva. (C) Four carcasses of fish with escaping holes of larvae. All scale bars are 10 cm.

**Fig. 1.** The predatory carp, *Chanodichthys erythropterus*, found in the river-edge areas of Chilgok-bo in Nakdong-gang (River), Korea. (A) A slowly swimming carp with an extruding larva. (B) An exhausted one just before dying with an escaping larva. (C) Four carcasses of fish with escaping holes of larvae. All scale bars are 10 cm.

**Table 1.** Fatal status* of predatory carp, *Chanodichthys erythropterus*, infected with the plerocercoid larvae of *L. intestinalis* in the 4 sites of Nakdong-gang (River)

| Duration of fish collection (2016) | CG-bo | GJ-bo | DS-bo | HC-CN-bo | Total |
|-----------------------------------|-------|-------|-------|----------|-------|
| 7-11 Feb.                         | 154*  | 0     | 0     | 0        | 154   |
| 12-16 Feb.                        | 39†   | 0     | 0     | 0        | 39    |
| 17-21 Feb.                        | 252   | 0     | 0     | 0        | 252   |
| 22-26 Feb.                        | 175   | 17    | 51    | 146      | 399   |
| 27 Feb.-2 Mar.                    | 113   | 2     | 32    | 62       | 209   |
| 3-7 Mar.                          | 25    | 0     | 24    | 4        | 53    |
| 8-12 Mar.                         | 56    | 0     | 16    | 0        | 72    |
| 13-17 Mar.                        | 3     | 0     | 2     | 0        | 5     |
| Total                             | 817   | 19    | 125   | 212      | 1,173 |

*Data from the Nakdong River Integrated Operations Center, Korea Water Resources Corporation, Korea.
†CG, Chilgok; GJ, Gangjeong; DS, Dalseong; HC-CN, Hapcheon, Changnyeong.
Personal data (92 and 20 fish) included.

Then, this ecological phenomenon also occurred in the adjacent areas of 3 dams of Nakdong-gang, i.e., Gangjeong-bo, Dalseong-bo, and Hapcheon-Changnyeong-bo, as well as Chilgok-bo. A total of 1,173 fish carcasses were collected from the 4 regions during about 40 days. The fatal status of predatory carp infected with the *L. intestinalis* plerocercoids was designated in Table 1.
Fig. 2. The predatory carp captured in the river-edge areas of Hapcheon-Changnyeong-bo with a landing net. (A) A complete one with the bulging abdomen before necropsy. (B) Two fish opened their abdominal cavity to examine the cause of death. Plerocercoid larvae (white mass) of *L. intestinalis* are occupying almost all space of the abdominal cavity under the air bladders. The digestive organs of fish are slender and empty without intestinal contents (arrow marks).

Fig. 3. Plerocercoid larvae of *L. intestinalis* recovered in the abdominal cavity of fish. (A) All larvae recovered from a heavily infected fish. (B) Two larvae showing their gross morphology with the blunting anterior end (scolex part).
To examine the cause of death, we captured 10 wondering carp in the river-edge area of Hapcheon-Changnyeong-bo with a landing net. They were 24.0-28.5 (av. 25.6) cm in length and 147-257 (av. 178) g in weight, and had 2-11 (5.3) plerocercoid larvae in the abdominal cavity. Their digestive organs were slender and empty without intestinal contents, and reproductive organs were not observed at all, while the plerocercoid larvae occupied almost all spaces of the abdominal cavity under the air bladders (Fig. 2). The proportion in weight of larvae per fish was 14.6-32.1% (av. 23.0%) of the total body weight. The larvae were ivory-white, long slender with the blunt anterior end (scolex part), 21.5-63.0 (av. 38.1) cm in length and 6.0-13.8 (av. 8.6) g in weight (Fig. 3).

Host manipulation by parasites is known as one of the strategies for parasite survival. Various examples have been studied with aspects of host-parasite relationships [23]. As an interesting example, mice infected with Toxoplasma gondii are easily eaten by the definitive host, the cat, because they become lack of phobia response against the predator [24]. In case of L. intestinalis, it has been known that the plerocercoid larvae induce endocrine disruption that can lead to parasitic castration [10-14], growth suppression, morphological modification [15,16], mortality increase in the host fish [17,18], and modified behavioral responses [4,19-22]. Therefore, the preference for the river-edge in L. intestinalis-infected fish during winter season is suggested to be a modified behavioral response similar to host manipulation by ligulid larvae. The infected carp slowly swimming in the river-edge area can be easily eaten by avian definitive hosts (migratory birds) during their visiting season, winter, and the life cycle of this tapeworm will be successfully continued.

During the life cycle of this tapeworm, the plerocercoid stage is most conspicuous in aspects of host-parasite relationships. The long-lasting inhabitation and development of L. intestinalis in the abdominal cavity will give considerable effects on the fish host, i.e., physical damage, morphological modifications, growth retardation, hematological changes, immune response disturbance, endocrine disruption, and behavioral changes [4,19-27]. Among these effects, the physiological function related to reproduction was frequently studied using the fish-ligulid parasite model. The decrease of pituitary gonadotropins, sex steroids, and brain aromatase expressions, as well as the inhibition of gonad maturation and development were mainly revealed as the results of these studies [10-14,27]. In the present study, although we did not approach functionally, we were able to know the infertility of infected carp view-
ing from absence of their reproductive organs at necropsy of the fish, probably because of degeneration. On the other hand, it has been suggested that parasites can obviously influence host population in a density-dependent manner, if they reduce host survival and/or fecundity, and the abundance of potential hosts and the temporal dynamics of their occurrence strongly affect local host specificity [28]. Whether the mass death of predatory carp infected with plerocercoid larvae of L. intestinalis affects the host population or not? It is one of the problems to be solved in the near future from ecological aspects of fish and parasites.

As the second intermediate host of this tapeworm, cyprinid fish are typically reported from a variety of location worldwide. The roach, Rutilus rutilus, is the most well known fish host in European countries, and it has been most frequently studied as a suitable fish model. In African continent, Barbus species have been broadly examined to know the infection status of plerocercoid larvae of L. intestinalis [4,7]. In Asian countries, including China and Korea, the cultrinid fish (subfamily Cultrinae), in particular, Hemiculter bleekeri, was reported as the fish intermediate host of L. intestinalis [9]. By the present study, another species of cultrinid fish, C. erythropterus, has been confirmed as a suitable second intermediate host of this tapeworm in Korea. Accordingly, until now, total 4 species of freshwater fish, i.e., Zacco platypus, Carassius auratus, H. bleekeri, and C. erythropterus, are listed as the second intermediate hosts of L. intestinalis in Korea.

The carcasses of predatory carp found in the river-edge areas had the escape holes of larvae just behind the thoracic fin. The plerocercoid larvae probably escaped through these holes when fish hosts were dead. At necropsy of fish, all visceral organs, including the digestive and reproductive ones of fish, were severely atrophied, and the plerocercoid larvae occupied almost all spaces of the abdominal cavity under air bladders. The parasite weight reached about 23.0% of the total body weight of the fish. The proportion of larval weight per fish was somewhat higher than those of the previous studies. Loot et al. [2] reported that average 6 plerocercoid larvae were infected in 3-year-old roach, R. rutilus, from a gravel pit in River Garonne in France, and their weight was 17.5% weight of fish infected. Britton et al. [4] reported about 20% weight of ligulid larvae in Barbus lineomaculatus from Lake Baringo in Kenya.

The ligulid larval infections occurred more frequently in fish from lakes and dams rather than rivers. In case of Korea, they were detected in fish from 5 lakes in Chungcheongnam-do.
and Hoengeeong-dam in Gangwon-do [9], and 4 dams in Nakdong-gang in this study. Most of the studies in other countries were performed with fish from lakes and dams except a few [4-8]. What kinds of ecological factors act in the life cycle of this tapeworm? The preference of stagnated water environment for the life cycle of this tapeworm should be investigated in the near future.

In conclusion, the plerocercoid larvae of *L. intestinalis* recently caused mass fatality of the fish host, *C. erythropterus*, in Korea. The host-parasite relationships between the fish and the tapeworm seem to be evolutionally well maintained, and the larvae were grown well in the abdominal cavity of fish without negative host responses. Ecological factors and the reason why the mass death of predatory carp occurred exceptionally in the dams of Nakdong-gang should be clarified in the near future.

In addition, *L. intestinalis* is an avian parasite but not a human parasite, although Joyeux and Noyer [29] described it from the vomitus of a Romanian person, and thus some textbooks and websites treat this as a human tapeworm [30]. *L. intestinalis* should have strong host specificity for fish and birds during a long evolutional process.

**ACKNOWLEDGMENT**

We thank Jung-A Kim and Hee-Ju Kim, Department of Parasitology and Tropical Medicine, Gyeongsang National University School of Medicine, Jinju, Korea, for their help in fish examinations.

**CONFLICT OF INTEREST**

The authors declare that they have no conflict of interest related to this article.

**REFERENCES**

1. Dubinina MN. Tapeworms (Cestoda, Ligulidae) of the Fauna of the USSR. New Delhi, India. Amerind Publishing Co. 1980.

2. Loot G, Francisco P, Santoul FG, Lek S, Guegan JF. The three hosts of the *Ligula intestinalis* (Cestoda) life cycle in Lavernos-Lacasse gravel pit, France. Arch Hydrobil 2001; 152: 511-525.

3. Innal D, Keskin N, Erkakan F. Distribution of *Ligula intestinalis* (L.) in Turkey. Turkish J Fish Aqu Sci 2007; 7: 19-22.

4. Britton JR, Jackson MC, Harper DM. *Ligula intestinalis* (Cestoda: Diphyllobothriidae) in Kenya: a field investigation into host specificity and behavioural alterations. Parasitology 2009; 136: 1367-1373.

5. Kurupinar E, Oztürk MO. A study on the helminth fauna linked to seasonal changes and size of the fish host, *Leuciscus cephalus* L., from Lake Dam Orenler, Afyonkarahisar. Türkiye Parazitol Derg 2009; 33: 248-253 (in Turkish).

6. Rusuwa B, Ngochera M, Maruyama A. *Ligula intestinalis* (Cestoda: Pseudophyllidea) infection of *Enguagnycoris sardella* (Pisces: Cyprinidae) in Lake Malawi. Malawi J Sci Tech 2014; 10: 8-14.

7. Tizie E, Baye D, Mohamed A. Prevalence of *Ligula intestinalis* larvae in *Barbus* fish genera at Lake Tana, Ethiopia. World J Fish Marine Sci 2014; 6: 408-416.

8. Arslan MÖ, Yılmaz M, Taçö GT. Infections of *Ligula intestinalis* on freshwater fish in Kars plateau of north-eastern Anatolia, Turkey. Türkiye Parazitol Derg 2015; 39: 218-221.

9. Ryu SS, Lee HJ. Study on the *Ligula intestinalis* larvae parasitized in freshwater fish. The 38th Korea Science Exhibition, Biological (Animal) Section, 1992, pp 1-14 (in Korean).

10. Geraudie P, Boulange-Lecomte C, Gerbron M, Hinfray N, Brion F, Minier C. Endocrine effects of the tapeworm *Ligula intestinalis* in its teleost host, the roach (*Rutilus rutilus*). Parasitology 2010; 137: 697-704.

11. Trubiroha A, Wuertz S, Frank SN, Sures B, Kloas W. Expression of gonadotropin subunits in roach (*Rutilus rutilus*, Cyprinidae) infected with plerocercoids of the tapeworm *Ligula intestinalis* (Cestoda). Int J Parasitol 2009; 39: 1465-1473.

12. Trubiroha A, Kroupova H, Wuertz S, Frank SN, Sures B, Kloas W. Naturally-induced endocrine disruption by the parasite *Ligula intestinalis* (Cestoda) in roach (*Rutilus rutilus*). Gen Comp Endocrinol 2010; 166: 234-240.

13. Trubiroha A, Kroupova H, Frank SN, Sures B, Kloas W. Inhibition of gametogenesis by the cestode *Ligula intestinalis* in roach (*Rutilus rutilus*) is attenuated under laboratory conditions. Parasitology 2011; 138: 648-659.

14. Boulange-Lecomte C, Geraudie P, Forget-Leray J, Gerbron M, Minier C. *Ligula intestinalis* infection is associated with alterations of both brain and gonad aromatase expression in roach (*Rutilus rutilus*). J Helminthol 2011; 85: 339-344.

15. Hadou-Sanoun G, Arab A, Lek-Ang S, Lek S. Impact of *Ligula intestinalis* (L. 1758) (Cestode), on the growth of *Barbus setiwmensis* (Cyprinidae) in a lake system in Algeria. C R Biol 2012; 335: 300-309 (in French).

16. Kroupova H, Trubiroha A, Wuertz S, Frank SN, Sures B, Kloas W. Nutritional status and gene expression along the somatotropic axis in roach (*Rutilus rutilus*) infected with the tapeworm *Ligula intestinalis*. Gen Comp Endocrinol 2012; 177: 270-277.

17. Loot G, Lek S, Dejean D, Guegan JF. Parasite-induced mortality in three host populations of the roach *Rutilus rutilus* (L.) by the tapeworm *Ligula intestinalis* (L.). Ann Limnol 2001; 37: 151-159.

18. Loot G, Poulin R, Lek S, Guegan JF. The differential effects of *Ligula intestinalis* (L.) plerocercoids on host growth in three natural populations of roach, *Rutilus rutilus* (L.). Ecol Freshw Fish 2002; 11: 168-177.

19. Barber I, Hoare D, Krause J. Effects of parasites on fish behav-
20. Brown SP, Loot G, Grenfell BT, Guégan JF. Host manipulation by *Ligula intestinalis*: accident or adaptation? Parasitology 2001; 123: 519-529.
21. Brown SP, Loot G, Teriokhin A, Brunel A, Brunel C, Guégan JF. Host manipulation by *Ligula intestinalis*: a cause or consequence of parasite aggregation? Int J Parasitol 2002; 32: 817-824.
22. Loot G, Aulagnier S, Lek S, Thomas F, Guégan JF. Experimental demonstration of a behavioural modification in a cyprinid fish, *Rutilus rutilus* (L.), induced by a parasite, *Ligula intestinalis* (L.). Can J Zool 2002; 80: 738-744.
23. Chai JY, Hong ST, Choi MH, Shin EH, Bae YM, Hong SJ, Sohn WM, Yu JR, Kho WG, Seo M, Park YK, Han ET. Seo and Lee’s Clinical Parasitology. Seoul, Korea. Seoul National University Press, 2011, pp 37-39 (in Korean).
24. Vyas A, Kim SK, Giacomini N, Boothroyd JC, Sapolsky RM. Behavioural changes induced by *Toxoplasma* infection of rodents are highly specific to aversion of cat odors. Proc Natl Acad Sci USA 2007; 104: 6442-6447.
25. Hayatbakhsh MR, Khara H, Movahed R, Sayadborani M, Rohi JD, Ahmadnezhad M, Rahbar M, Rad AS. Haematological characteristics associated with parasitism in bream, *Abramis brama orientalis*. J Parasit Dis 2014; 38: 383-388.
26. Halimi M, Colagar AH, Youssefi MR. Immune response in spirulins (*Alburnoides bipunctatus*, Bloch 1782) infested by *Ligula intestinalis* parasite. Vet Italiana 2013; 49: 243-246.
27. Carter V, Pierce R, Dufour S, Arme C, Hoole D. The tapeworm *Ligula intestinalis* (Cestoda: Pseudophyllidea) inhibits LH expression and puberty in its teleost host, *Rutilus rutilus*. Reproduction 2005; 130: 939-945.
28. Loot G, Park YS, Lek S, Brosse S. Encounter rate between local populations shapes host selection in complex parasite life cycle. Biol J Linn Soc 2006; 89: 99-106.
29. Joyeux CE, Noyer R. Presence d’une larve de ligule dans des vomissements Marseille Med 1931; 68: 235-236.
30. Beaver PC, Jung RC, Cupp EW. Clinical Parasitology. 9th ed. Philadelphia, USA. Lea & Febiger. 1984, p 499.