INTRODUCTION

Yellow fever (YF) is an acute febrile disease caused by the yellow fever virus (YFV) belonging to the Flaviviridae family and is endemic in mostly African and South American countries [1]. YF is mainly transmitted by mosquitoes of the genera Aedes sp., Haemagogus sp. and Sabethes sp., so it may occur mainly in areas where these mosquitoes are prevalent [2-4]. Most people are asymptomatic or have a mild infection, but a small proportion of patients develop severe symptoms with a mortality of over 50% [5]. YFV belongs to the Flaviviridae family. YFV is composed of 3 structural proteins (capsid: C, membrane: M, and envelope: E) and 7 non-structural proteins (NS1, NS2a, NS2b, NS3, NS4a, NS4b, and NS5) [7]. The Flavivirus non-structural protein 1 (NS1) is a highly preserved glycoprotein dimer approximately 46-55 kDa in size depending on the degree of glycosylation. NS1 can be detected from the cellular surface or in side infected cells, and are secreted effectively from cells [13]. Intracellular NS1 is central to viral replication, secreted and recently bound NS1 is important in inducing immune reactions [3]. NS1 from patients’ serum infected with WNV and DENV in the Flaviviridae family are used as initial markers. However, in contrast to other Flaviviridae, not much is known about the
expression and secretion of YFV.

Laboratory diagnosis can provide a rapid and accurate differential identification of the pathogen and may help in the appropriate management of YF. YFV infection can be detected by several methods such as viral isolation and RNA detection from the blood, and antibody detection from the serum [14,15]. Several methods such as haemaglutination inhibition test, complement fixation test, plaque reduction neutralization test, immunofluorescence assay, IgM-capture enzyme linked immunosorbent assay, IgG ELISA are used for serological diagnosis of viral infections. Although most laboratories use ELISA to detect antibodies that are released after infection, it has an inherent disadvantage that it takes too long to perform. In this study, we developed a rapid diagnostic test kit to diagnose yellow fever by using monoclonal antibodies highly specific to NS1 protein obtained from yellow fever virus itself. And we performed the specificity and sensitivity of the kit among various similar viruses.

MATERIALS AND METHODS

Culture of host cell and virus

African green monkey kidney cell line (Vero, ATCC® CRL-1586, Manassas, Virginia, USA) was used as host cells in this study. YF virus strain 17D was used. Dulbecco’s modified Eagle’s medium (DMEM, Gibco/BRL, Gaithersburg, Maryland, USA) with 2% FBS (Gibco/BRL, 16000-044) and 0.05 mg/ml gentamycin (Gibco/BRL, 15710-064) was used as culture medium. When the Vero cells grew to 70-80% confluency, the culture was washed with PBS (pH 7.4) and then dengue, zika, chikungunya, mayaro and YF viruses were inoculated at a concentration of 0.5 × 10⁴ pfu/ml. After inoculating each virus, the culture was washed with PBS (pH 7.4) and then dengue, zika, chikungunya, mayaro and YF viruses were inoculated at a concentration of 0.5 × 10⁴ pfu/ml. After washing the unbound proteins using the same buffer, YF NS1 bound to the column was eluted with 0.1 M glycine (pH 2.8). The pH level was adjusted by adding 1 M Tris-HCl (pH 7.9) solution to 1/10 th volume of the eluted solution. The eluted YF NS1 was dialyzed with PBS (pH 7.4) and stored at 4°C. The purified YF NS1 was dried by 1 µl dripping method on nitrocellulose membrane. The nitrocellulose dot strips were blocked for 2 hr with PBS containing 5% skim milk, and then the antigen diluted with dilution solution (1% BSA in PBS) was added to each strip and reacted for 30 min. Each dot strip was washed with PBS-Tween for 1 min 5 times, and then reacted for 30 min at room temperature with gold-conjugated protein G diluted 1:1,000 in dilution solution. Finally, it was washed with distilled water and the reaction stopped. In this study, we obtained YF NS1 5 mg/L from the yellow fever culture media, 1 week post infection.

Immunization of mouse with purified YF NS1

Procedures to produce mAb against NS1 of YF were followed the methods described earlier [16,17] with some modifications. Briefly, a 6-week SPF female mouse (Samtaco, Suwon, Korea) was immunized primarily with yellow fever NS1 25 µg in 25 µl PBS mixed with Freund’s Complete adjuvant (Sigma Chem Co., St. Louis, Missouri, USA) of 1:1 volume. Two weeks later, secondary immunization was carried out using incomplete adjuvant (Sigma) by the same method. Two weeks later, antibody titration was identified through indirect ELISA with blood collected from the tail. An absorbance of 1:1,000 diluents of 0.2 or higher seemed successful immunization, and 25 µg of YF NS1 50 µl was injected into the mouse’s feet 5 days before cell fusion to boost the immunity.

Selection of producing hybridoma monoclonal antibody and mouse ascites production

Myeloma cells (SP2/0 cells, ATCC CRL1581) were cultured in T-75 flasks using DMEM containing 20% FBS. The cells were fused with lymph node cells of the immunized mouse under the polyethylene glycol 1500 (PEG 1500, Sigma). Then, after reacting for 5 min at 37°C, the hybridoma cells were cultured in HAT culture medium containing 20% FBS. When the cells multiplied to 60-70% of the well plate at the bottom, the antibody titer was measured by ELISA using a plate coated with YF NS1 protein and the well supernatant in addition to
differentiated with cross-reactions using plates coated with Zika and Dengue viruses. Wells with high antibody titers were moved into 24 well plates and cultured according to limiting dilution so that one cell per well would fit in a 96-well plate, and finally a monoclonal cell line was established.

Murine ascites were harvested after 1 week injecting 0.5 ml of incomplete Freud’s adjuvant in the abdomen of a female adult mouse with at least $5 \times 10^7$ hybridoma cells diluted into 0.5 ml PBS (pH 7.4). The collected ascites was centrifuged at 4°C, 8,000 rpm for 10 min, and then the antibody fractions were collected with 20% to 50% ammonium sulfate precipitation. The precipitate was solved in 20 mM KH$_2$PO$_4$ (pH 7.4) and dialyzed (Spectrumlabs, 132724) with the same buffer. Then, it was loaded into a protein-G column equilibrated with 20 mM KH$_2$PO$_4$ (pH 7.4) to purify antibody by adding 1 M Tris-HCl (pH 7.9) solution. The eluted antibodies were dialyzed with PBS (pH 7.4) and refrigerated. The purified antibodies were quantified at 280 nm, and the protein was confirmed with 12% SDS-PAGE by Coomassie blue staining.

Characterization of monoclonal antibodies

To perform an isotype analysis, an ELISA kit was composed of a certain amount of goat antibody (Sigma) specific for mouse antibody isotypes (IgG1, IgG2a, IgG2b, IgG3, IgM, IgA) to aliquot into immune plates. After 1 hr reaction at 37°C in PBS (pH 7.4), the plate was blocked with 200 μl PBS containing 10% BSA, 5% sucrose, and 0.1% Tween-20 at 37°C for 1 hr, the remaining PBS was removed and dried for 16 hr at room temperature before use. After aliquoting the collected monoclonal culture medium into each well, it was reacted for 1 hr at 37°C, and then washed 3 times with PBS-T (pH 7.4). Peroxidase-conjugated goat anti-mouse IgG (Sigma, A9917) which is specific to mouse IgG is reacted at 37°C for 1 hr, and then substrate containing color fixing agent is used to induce colorization and identify isotypes.

The dissociation constant (Kd) for each monoclonal antibody against antigens was measured by modifying the method of Friquet et al. [18]. After reacting serially diluted antigens and antibodies at a certain concentration for 10 min, dissociation constant was calculated using antigen-coated plates. A dissociation constant value in the range of 10-6 to 10-7 means a low coupling force between antigen-antigen, whereas a value in the range of 10-9 to 10-12 means a high coupling rate.

**RESULTS**

**YF NS1 purification and confirmation of antigenicity**

The analysis of purified YF NS1 with SDS-PAGE showed a band of approximately 45 to 50 kDa (Fig. 1A). To confirm the antigenicity of the purified YF NS1, a dot blot assay was performed. As a result, a distinct positive reaction was observed only in the general sample in which the anti-flavivirus NS1 polyclonal antibodies were spiked, and not in normal serum (Fig. 1B). YF NS1 with confirmed purity and antigenicity was used for immunization.

Development of RDT to detect YF NS1

RDT kit was composed basically of test and control lines which were divided on the nitrocellulose membrane [19]. Membranes were obtained from Advanced Microdevices (Ambala Cantt, Haryana, India) while the absorbant, sample, conjugate, and blood separation pads were obtained from Ahlstrom (Helsinki, Finland). The test line contained antibody at 1.5 mg/ml that worked as a capture site, while the control line contained goat antibody at 2.5 mg/ml. Gold nanoparticle (40 nm in mean diameter) conjugate pads were made by soaking the antibody conjugated gold solution in glass fibers and then drying them, while the sample pads were soaked in cellulose fibers containing 0.5% polyvinyl alcohol and Tween-20 and then drying them. All pads were attached overlapping so that the sample solutions could migrate.

Monkey tissue samples were provided by Bahiafarma (based on the IRB Committee of Bahiafarma 2016-01-02), while negative blood samples were provided by the Catholic University of Korea (based on the IRB committee No. MC18SES10013). Specimens from Bahiafarma and Catholic University were all confirmed by RT-PCR. In brief, we first extracted RNA using QIAamp® Viral RNA Mini Kit (QIAGEN, Hilden, Germany). The reagent and 10 μl of RNA prep sample were then mixed using the RealStar® Yellow Fever Virus RT-PCR Kit 1.0 (Altona, Hamburg, Germany). The final mixture was loaded and run in the RT-PCR machine (CFX96™, Bio-Rad, Hercules, California, USA) after setting the temperature profile. The monkey tissues were crushed with a homogenizer with PBS (pH 7.4) at 1 ml per g tissue, and then used. The samples received were verified 15 min after dropping 100 μl onto the sample pad. The control line was checked in every test, and results without control lines were re-examined. When a band appeared on the test line, it was determined to be positive.

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Immunization and monoclonal cell production

Immunization was performed using YF NS1 with confirmed antigenicity. The fused cells were subjected to a screening process to confirm the secretion of antibodies specifically binding to YF NS1. Screening was performed by indirect ELISA and lateral flow assay (LFA). Using indirect ELISA, 10 candidate clones were selected, in which the antibody titer was high. In addition, screening was carried out using plates coated with Zika NS1 and Dengue NS1 to confirm cross-reaction with other flaviviruses. Among the 10 highly reactive candidate clones, 7 clones were selected, except for those (1A7, 5G8, 5G9) that cross-reacted with Zika or Dengue. In order to select clones that recognize the tertiary structure, LFA was used to confirm that the candidate clones (2B10, 5A6), which did not show cross-reaction in indirect ELISA, showed cross-reaction between Zika and Dengue. Therefore, 3A11, 4H5, 3B7, 6C11, and 3C4, which are specific to yellow fever, and do not cross-react with Dengue and Zika, were selected finally. The cloning procedure was then performed twice to establish a monoclonal cell line (Fig. 2).

Monoclonal antibody production and characterization

The antibody was isolated and purified from the ascites using ammonium sulfate precipitation method and protein-G column affinity chromatography. As a result of SDS-PAGE, bands were observed near 50 kDa and 25 kDa (data not shown).

Fig. 1. Purity and antigenicity of purified yellow fever virus NS1 (nYF-NS1). (A) Purified YF NS1 on SDS-PAGE gel appeared as a single band. (B) Dot blot assay showed strong positive reaction to a normal serum (a) spiked with anti-Flavivirus NS1 polyclonal antibody but not to a normal serum itself (b).

Fig. 2. Screening of YF NS1 monoclonal antibodies against flavivirus NS1 antigens. (A) Screening by indirect ELISA. (B) Screening by western blotting.
shown), that correspond to the heavy chain and light chain in a usual antibody. Isotype and dissociation constants were determined to characterize the purified monoclonal antibodies (3A11, 3B7, 3C4, 4H5, 6C11). All monoclonal antibodies belonged in the IgG class and were confirmed to be isotypes IgG1 and IgG2a. The dissociation constants were determined $10^{-8}$-$10^{-9}$, and the antibodies with the highest affinity were 3A11 and 3B7 (Table 1).

### Table 1. Isotype and dissociation constant of monoclonal antibody

| Hybridoma | Isotype | $K_d$ (M)       |
|-----------|---------|-----------------|
| 3A11      | IgG1    | $7.531 \pm 0.233 \times 10^{-9}$ |
| 3B7       | IgG2a   | $9.114 \pm 0.473 \times 10^{-9}$ |
| 3C4       | IgG1    | $7.781 \pm 0.403 \times 10^{-8}$ |
| 4H5       | IgG1    | $8.121 \pm 0.433 \times 10^{-8}$ |
| 6C11      | IgG2a   | $7.888 \pm 0.307 \times 10^{-8}$ |

Monoclonal Antibody Pair Test and detection limit test

1.5 mg/ml of the 5 candidate clone antibodies (3A11, 3B7, 3C4, 4H5, 6C11) were applied on nitrocellulose membranes and gold nanoparticle conjugate solution was prepared for the pair test. Antibodies for capture and antibodies for gold nanoparticle conjugate were tested in 25 cases. Antibodies 3B7 for capture, and 3A11 for gold nanoparticle conjugate showed the best sensitivity (Fig. 3). These 2 monoclonal antibodies were selected as the best pair to apply in developing kits for the detection of YF NS1.

Detection limit was analyzed by the recombinant protein and the virus culture were diluted by 1/5, which resulted in the positive up to 0.8 ng/ml of recombinant NS1 protein and 1.6 pfu/ml of supernatant from virus culture. (Fig. 4).

### Specificity, sensitivity and cross reaction

Both positive and negative monkey tissues from Bahiafarma, and negative blood samples from the Catholic University of Korea were used for clinical trials. Specificity refers to the value that a person without disease is judged to be normal through a diagnostic test. A total of 100 sera and 50 whole bloods were tested for specificity with the YF NS1 kit. No false positives were observed, the background was clean, whereas control lines were definite. Sensitivity refers to the value that a person with a disease is defined as being positive through a diagnostic test. Positive monkey tissue was used to investigate the sensitivity of the kits produced in this study.

Tests were carried out with virus cultures to determine whether cross-reaction with other epidemic Flaviviruses and Alphaviruses occurred. No cross-reaction was observed with
DISCUSSION

Yellow fever is an acute febrile disease caused by the yellow fever virus belonging to the *Flaviviridae* family, which occurs mainly in the jungle areas of Africa and South America. Although safe and effective vaccines have been in use since 1937, YF remains a major health problem in Africa and South America. Nowadays, the increase in international travelers and global warming have increased the possibility of infectious disease-causing mosquitoes in becoming endemic, so that Korea is not safe from this risk any more.

Since YF has no effective treatment yet, it is imperative to diagnose it early and accurately. Selection of a target protein is most important in making a rapid and accurate diagnosis. Previous studies have shown that the *Flaviviridae* NS1 is a highly conserved protein involved in viral replication that is known to be produced a lot in the early stages of infection [20]. In addition, NS1 is known to be a good candidate for the diagnosis of viral infection since it is secreted out of infected cells at a high concentration that can be detected in the blood, and can be observed up to 9 days after the initial infection [21].

However, not much is known about the expression and secretion of YF NS1 due to the scarcity of related researches, causing us concern about its use as a marker. But as YFV belongs to same *Flaviviridae* family as DENV, we assumed that the similar mechanism as in the previous DENV research would exist and proceeded with the study [22].

Viruses of the *Flaviviridae* family have homology to each other, and NS1 has a high homology of 61% [12] (Table 2). Taking advantage of this homology, anti-DENV polyclonal antibody developed in a previous study was attached to beads, and YF NS1 was purified by affinity chromatography. Subsequent immunization and cell fusion were carried out with this purified YF NS1.

During the screening process for monoclonal antibodies that
Table 2. Homology of NS1 in the Flaviviridae family

| NS1 | DENV 1 | DENV 2 | DENV 3 | DENV 4 | JEV | WNV | ZKV | YFV |
|-----|--------|--------|--------|--------|-----|-----|-----|-----|
| DENV 1 | 100.0 | 73.9 | 79.8 | 69.3 | 51.4 | 51.4 | 53.7 | 41.9 |
| DENV 2 | 73.9 | 100.0 | 74.4 | 73.3 | 53.4 | 56.0 | 54.0 | 43.9 |
| DENV 3 | 79.8 | 74.4 | 100.0 | 73.9 | 51.4 | 53.7 | 55.4 | 41.9 |
| DENV 4 | 69.3 | 73.3 | 73.9 | 100.0 | 52.6 | 53.7 | 54.0 | 42.5 |
| JEV | 51.4 | 53.4 | 51.4 | 52.6 | 100 | 77.0 | 56.8 | 45.9 |
| WNV | 51.4 | 56.0 | 53.7 | 53.7 | 77.0 | 100 | 54.1 | 43.9 |
| ZKV | 53.7 | 54.0 | 55.4 | 54.0 | 56.8 | 54.1 | 100.0 | 46.7 |
| YFV | 41.9 | 43.9 | 41.9 | 42.5 | 45.9 | 43.9 | 46.7 | 100.0 |

*Pairwise sequence comparison of the NS1 proteins from different Flaviviruses.
**Accession no. AUW27546.1 (DENV1); CAE11558.1 (DENV2); YP_001531169.2 (DENV3); NP_740318.1 (DENV4); NP_775667.1 (JEV); YP_009164952.1 (WNV); YP_009430301.1 (ZKV); NP_776002.1 (YFV).

specifically capture YF NS1, this homology of the Flaviviridae family may in turn cause a problem in the actual screening. So screening was conducted with ELISA using not only YFV but also DENV and ZIKV NS1. However, when the native virus is tested in the field, the tertiary structure is recognized and diagnosed, but ELISA has the distinct disadvantage in that this tertiary structure breaks down due to hydrophobic binding when the antigen is coated on the plate, so we judged that screening could not be performed if the epitope of an antibody was a tertiary structure. Therefore, YF NS1 was applied to a membrane and screening was also carried out by lateral flow method.

The 2 results identified 2 candidate clones that did not cross-react in ELISA, but showed cross-reaction in the lateral flow assay. We confirmed that screening could not be performed with only ELISA if dependent solely on the antigenic determinant. Finally, we selected candidate clones that specifically target only YF NS1, without cross-reacting in both ELISA and lateral flow assay.

Finally, clones 3A11 and 3B7 were selected through antibody characterization and pair test, and the kit was prepared, and comparison tests were performed with a commercial pair from Meridian Bioscience. Detection limit was analyzed by comparing the 2 kits using recombinant protein and virus culture medium. The recombinant protein when diluted was detected better with the purchased kit, while the kit developed in this study had much better sensitivity with viral culture medium. In addition, tests with positive monkey tissues provided by Bahiafarm showed that the kit developed in this study showed high sensitivity specificity of 100% (Table 3).

Table 3. Diagnostic performance of in-house RDT compared with RT-PCR

|         | RDT         | Total |
|---------|-------------|-------|
|         | Negative    | Positive |       |
| RT-PCR  | 12          | 0      | 12    |
|         | 0           | 112    | 112   |
| Total   | 12          | 112    | 124   |

Sensitivity = 100%, Specificity = 100%.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.
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