Triglyceride Glucose-Body Mass Index Is a Simple and Clinically Useful Surrogate Marker for Insulin Resistance in Nondiabetic Individuals

Leay-Kiaw Er1, Semon Wu2,3, Hsin-Hua Chou4, Lung-An Hsu5, Ming-Sheng Teng2, Yu-Chen Sun6, Yu-Lin Ko2,4,7*

1 The Division of Endocrinology and Metabolism, Department of Internal Medicine, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei City, Taiwan, 2 Department of Research, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei City, Taiwan, 3 Department of Life Science, Chinese Culture University, Taipei, Taiwan, 4 The Division of Cardiology, Department of Internal Medicine and Cardiovascular Center, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei City, Taiwan, 5 The First Cardiovascular Division, Department of Internal Medicine, Chang Gung Memorial Hospital and Chang Gung University College of Medicine, Taoyuan, Taiwan, 6 Department of Laboratory Medicine, Chang Gung Memorial Hospital, Taipei, Taiwan, 7 School of Medicine, Tzu Chi University, Hualien, Taiwan

* yulinkotw@yahoo.com.tw

Abstract

Background
Insulin resistance (IR) and the consequences of compensatory hyperinsulinemia are pathogenic factors for a set of metabolic abnormalities, which contribute to the development of diabetes mellitus and cardiovascular diseases. We compared traditional lipid levels and ratios and combined them with fasting plasma glucose (FPG) levels or adiposity status for determining their efficiency as independent risk factors for IR.

Methods
We enrolled 511 Taiwanese individuals for the analysis. The clinical usefulness of various parameters—such as traditional lipid levels and ratios; visceral adiposity indicators, visceral adiposity index (VAI), and lipid accumulation product (LAP); the product of triglyceride (TG) and FPG (the TyG index); TyG with adiposity status (TyG-BMI) and TyG-waist circumference index (WC)—was analyzed to identify IR.

Results
For all lipid ratios, the TG/high-density lipoprotein cholesterol (HDL-C) ratio had the highest additional percentage of variation in the homeostasis model assessment of insulin resistance (HOMA-IR; 7.0% in total); for all variables of interest, TyG-BMI and leptin-adiponectin ratio (LAR) were strongly associated with HOMA-IR, with 16.6% and 23.2% of variability.
respectively. A logistic regression analysis revealed similar patterns. A receiver operating characteristic (ROC) curve analysis indicated that TG/HDL-C was a more efficient IR discriminator than other lipid variables or ratios. The area under the ROC curve (AUC) for VAI (0.734) and TyG (0.708) was larger than that for TG/HDL-C (0.707). TyG-BMI and LAR had the largest AUC (0.801 and 0.801, respectively).

Conclusion
TyG-BMI is a simple, powerful, and clinically useful surrogate marker for early identification of IR.

Introduction
Insulin resistance (IR) involves reduced muscle and adipose tissue sensitivities to insulin and reduced ability of the liver to suppress hepatic glucose production and output [1]. IR is considered a major risk factor for type 2 diabetes and cardiovascular (CV) diseases [2]. Because of the clinical importance of IR, the ability to identify individuals with IR before the development of cardiometabolic diseases is of paramount importance. Although the hyperinsulinemic euglycemia clamp remains the gold standard for measuring IR [3], its practical clinical application is limited by the labor intensiveness and cost and by ethical concerns. Therefore, a simple, reliable, and reproducible index for measuring IR is urgently required.

It has been demonstrated that the product of TG and FPG levels (TyG index) presents moderate power as a surrogate marker for estimating HOMA-IR index in healthy subjects [4]. And The TyG index has high sensitivity and specificity compared with the euglycemic hyperinsulinemic clamp test for recognizing IR [5]. The superiority of the TyG index in identifying IR was also reported in many other studies [6,7,8]. With these findings, a highly efficient substitute measure to establish IR can be easily applied in clinical setting.

Previous studies also proposed other new formulas for estimating IR from adiposity. The visceral adiposity index (VAI), a mathematical model used to identify IR, is based on the anthropometric body mass index (BMI), waist circumference (WC), and metabolic (TG and HDL-C) parameters [9,10]. The lipid accumulation product (LAP), another mathematical model calculated from the WC and fasting TG levels, has been demonstrated as an alternative index for lipid accumulation and as a surrogate marker for IR [11,12]. Recently, a cross-sectional study directly compared lipid ratios, VAI, LAP, and TyG for measuring their IR detection efficiency, and the TyG was revealed to be the most efficient marker for early identification of IR [6]. BMI and WC are simple, inexpensive and noninvasive anthropometric parameters and are commonly adopted as useful indicators of obesity and other metabolic risk. As evidence increasingly suggests that obesity exhibits a close relationship with IR and because TyG are well-established as promising surrogate marker of IR, a combination of obesity and TyG can potentially identify IR more strongly than other surrogate markers. Thus, in this investigation, we compare lipid, adipokines, and lipid and adipokine ratios, visceral adiposity indicators, TyG and TyG related parameters (TyG-WC, TyG BMI) for early identification of IR. We analyzed whether a combination of TyG with BMI or WC can be a more powerful, simple and inexpensive clinical surrogate marker for IR.
Subjects and Methods

Study population

This study was approved by the institutional review board of the Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation (IRB number: 98-IRB-021-XD). Participants were recruited during cardiovascular health examinations between October 2003 and September 2005 at Chang Gung Memorial Hospital. All the study individuals provided written informed consent. Medical history- and lifestyle characteristics-related questionnaire responses were collected. Cancer; current renal or liver disease; and a history of myocardial infarction, stroke, or transient ischemic attacks were the exclusion criteria. An initial recruitment revealed 617 individuals. In addition, participants with diabetes mellitus (defined as fasting blood glucose \( \geq 7.0 \text{ mmol/L} \) (7 individuals) or the regular use of medications for diabetes mellitus (11 individuals)), aged \(<18\) years (5 individuals), triglyceride levels \( > 500 \text{ mg/dL} \) (10 individuals) and a history of regular medication for hypertension (66 individuals) or hyperlipidemia (9 individuals) were excluded. Finally, we enrolled 511 people—257 men and 254 women—for the analysis. Blood pressure was measured with patients in the sitting position after a 15-min rest period. Weight was measured on a calibrated beam scale with participants wearing light clothing, and height was measured without shoes. BMI was calculated by dividing weight (in kilograms) by the square of height (in meters). WC was measured using an inelastic tape at the midpoint of the bottom of the rib cage and the top of the iliac crest. Fasting blood samples were obtained from each participant. Current smoker was defined as smoking at least 1 cigarette per day at the time of survey. Alcohol consumption was defined as anyone who consumed alcohol once a day or more. Exercise was defined as regular exercise one or more times per week, at least 20 min for each time. Table 1 summarizes the clinical and biometric features of the study group stratified by sex and IR.

Laboratory examinations and assays

Lipid variables were performed according to the methods described by Teng et al. [13]. Non-HDL-C was calculated by subtracting high density lipoprotein cholesterol (HDL-C) from total cholesterol (TC). FPG was enzymatically determined by using the hexokinase method. Serum insulin levels were measured using an immunoradiometric assay (Bio-source, Nivelles, Belgium). Homeostasis model assessment of insulin resistance (HOMA-IR) was calculated using the following formula: HOMA-IR = fasting insulin (\( \mu \text{U/mL} \)) × FPG (mmol/L)/22.5 [14]. Insulin resistance was recognized when the HOMA-IR index reached the upper quartile. The C-reactive protein (CRP) level was measured by using a sandwich-type enzyme-linked immunosorbent assay (ELISA), which was developed in house. [15]. Serum leptin and adiponectin levels also were measured using an in-house sandwich-type ELISA assay. All in-house kits showed good correlation when compared with commercially available ELISA kits [15,16,17]. Circulating resistin levels were measured using commercially available ELISA kits (R&D Systems, Minneapolis, MN, USA). The within-day precision and day-to-day precision were 2.2% and 2.1% for TC, 3.4% and 4.5% for HDL-C, 2.3% and 3.0% for LDL-C, 2.0% and 2.2% for TG, 1.3% and 2.0% for glucose, 6.0% and 10.0% for insulin, 5.1% and 4.0% for leptin, 3.2% and 8.6% for resistin, 7.7% and 7.0% for adiponectin and 8.2% and 9.5% for CRP, respectively.

Visceral adiposity indicators, TyG related parameters and adipokine ratios were calculated as follows: LAP for men: \( (\text{WC} [\text{cm}] – 65) \times \text{TG} [\text{mmol/L}] \); LAP for women: \( (\text{WC} [\text{cm}] – 58) \times \text{TG} [\text{mmol/L}] \) [18]. VAI for men: \( (\text{WC} [\text{cm}] / 39.68 + (1.88 \times \text{BMI}) \times (\text{TG} [\text{mmol/L}] / 1.03) \times (1.31/\text{HDL-C} [\text{mmol/L}]) \); VAI for women: \( (\text{WC} [\text{cm}] / 36.58 + (1.89 \times \text{BMI}) \times (\text{TG} [\text{mmol/L}] / 0.81) \times (1.52/\text{HDL-C} [\text{mmol/L}]) \) [9]. TyG index: \( \text{Ln} \left[ \text{TG (mg/dL)} \times \text{FPG (mg/dL)/2} \right] \) [4,5].
Table 1. Baseline Characteristics of the Study Subjects.

|                          | Men | Women | $P$  | Non-IR | IR   | $P$  |
|--------------------------|-----|-------|------|--------|------|------|
| Number                   | 257 | 254   |      | 405    | 106  |      |
| Age, year                | 43.0 (38.1–50.0) | 46.0 (40.0–51.2) | 0.002 | 45.0 (39.0–51.0) | 44.7 (39.8–50.0) | 0.974 |
| Body mass index, kg/m²   | 24.8 (22.8–26.4) | 22.8 (20.8–25.5) | < 0.001 | 23.3 (21.3–25.4) | 26.5 (24.5–28.7) | < 0.001 |
| Waist circumference, cm  | 87.0 (82.0–91.0) | 80.0 (74.0–87.3) | < 0.001 | 83.0 (76.0–88.5) | 90.0 (84.8–97.3) | < 0.001 |
| Systolic BP, mmHg        | 112.0 (102.0–122.0) | 110.0 (98.0–122.0) | 0.077 | 108.0 (100.0–122.0) | 116.0 (108.0–130.0) | < 0.001 |
| Diastolic BP, mmHg       | 76.0 (70.0–84.0) | 72.0 (66.0–80.0) | < 0.001 | 72.0 (66.0–82.0) | 80.0 (72.0–86.0) | < 0.001 |
| Mean BP, mmHg            | 88.7 (81.0–95.3) | 84.7 (77.2–94.2) | 0.002 | 85.3 (78.0–93.3) | 92.3 (85.7–98.7) | < 0.001 |
| TC, mg/dL                | 200.0 (176.0–221.0) | 192.0 (170.0–219.5) | 0.085 | 195.0 (172.0–220.0) | 198.0 (178.5–223.0) | 0.319 |
| LDL-C, mg/dL             | 49.0 (42.5–55.0) | 62.0 (52.0–71.0) | < 0.001 | 57.0 (47.0–68.0) | 47.0 (39.0–56.0) | < 0.001 |
| HDL-C, mg/dL             | 120.0 (98.0–140.0) | 109.0 (91.8–134.3) | 0.004 | 114.0 (93.5–137.0) | 119.0 (95.0–139.0) | 0.417 |
| TG, mg/dL                | 131.0 (91.0–193.5) | 87.0 (65.0–129.3) | < 0.001 | 98.0 (68.5–150.0) | 140.0 (101.5–220.5) | < 0.001 |
| Non-HDL-C, mg/dL         | 153.0 (124.0–171.5) | 131.5 (109.0–155.0) | < 0.001 | 137.0 (114.0–163.0) | 153.0 (130.8–175.3) | 0.001 |
| TC/HDL-C                 | 4.13 (3.39–4.89) | 3.12 (2.69–3.78) | < 0.001 | 3.40 (2.83–4.18) | 4.34 (3.48–4.94) | < 0.001 |
| TG/HDL-C                 | 2.78 (1.63–4.32) | 1.43 (0.96–2.32) | < 0.001 | 1.69 (1.11–2.95) | 3.29 (1.75–4.83) | < 0.001 |
| LDL-C/HDL-C              | 2.50 (1.94–3.04) | 1.78 (1.43–2.32) | < 0.001 | 2.00 (1.55–2.60) | 2.53 (1.95–2.98) | < 0.001 |
| Non-HDL-C/HDL-C          | 3.13 (2.39–3.89) | 2.12 (1.69–2.78) | < 0.001 | 2.40 (1.83–3.18) | 3.34 (2.48–3.94) | < 0.001 |
| Current smokers, %       | 34.2 | 3.9   | < 0.001 | 16.8     | 28.3  | 0.007 |
| Alcohol consumption, %   | 5.1  | 0.4   | 0.001  | 2.7      | 2.8   | 0.949 |
| Exercise %               | 31.9 | 31.9  | 0.973  | 32.3     | 30.2  | 0.085 |
| Fasting plasma glucose, mg/dL | 94.0 (89.0–99.0) | 90.0 (85.0–95.0) | < 0.001 | 91.0 (87.0–95.0) | 96.5 (92.0–105.0) | < 0.001 |
| Fasting serum insulin, μU/mL | 8.06 (6.29–10.94) | 7.44 (5.72–10.27) | 0.025 | 7.06 (5.56–8.56) | 13.89 (12.18–18.00) | < 0.001 |
| HOMA-IR index            | 1.89 (1.46–2.54) | 1.69 (1.26–2.35) | 0.002 | 1.58 (1.24–1.98) | 3.33 (2.85–4.30) | < 0.001 |
| Adiponectin, μg/mL       | 4.65 (3.13–7.02) | 8.09 (5.60–11.55) | < 0.001 | 6.81 (4.44–10.05) | 4.11 (2.67–6.28) | < 0.001 |
| Leptin, ng/mL            | 9.17 (5.05–14.13) | 24.40 (15.73–32.41) | < 0.001 | 13.19 (6.88–23.42) | 21.87 (12.77–34.89) | < 0.001 |
| Resistin, ng/mL          | 14.8 (10.2–22.4) | 15.1 (10.7–22.6) | 0.840 | 14.9 (10.4–22.7) | 15.2 (10.0–21.6) | 0.682 |
| LAR                      | 2.07 (0.92–3.67) | 3.03 (1.57–5.29) | < 0.001 | 2.01 (0.98–3.49) | 4.74 (3.27–9.24) | < 0.001 |
| AR index                 | 1.50 (1.31–1.74) | 1.31 (1.03–1.53) | < 0.001 | 1.36 (1.09–1.62) | 1.53 (1.36–1.80) | < 0.001 |
| CRP, mg/L                | 0.64 (0.29–1.31) | 0.50 (0.23–1.16) | 0.050 | 0.47 (0.22–0.98) | 1.19 (0.56–2.31) | < 0.001 |

IR, insulin resistance; BP, blood pressure; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; TC, total cholesterol; TG, triglyceride; HOMA-IR, homeostasis model assessment of insulin resistance; LAR, leptin-adiponectin ratio; AR index, adiponectin-resistin index; CRP, C-reactive protein. Data are presented as median (interquartile range), or percent.

doi:10.1371/journal.pone.0149731.t001

TyG-BMI: TyG index × BMI. TyG-WC: TyG index × WC. LAR: leptin (ng/ml)/adiponectin (mg/ml) [19]. AR index: 1 + log10 (fasting resistin [ng/mL]) – log10 (fasting adiponectin [μg/mL]) [20].

Statistical analyses

All statistical analyses were conducted using SPSS (Version 12.0 for Windows; SPSS, Chicago, IL, USA). Before conducting linear regression, all variables except TyG, TyG-BMI, and TyG-WC were log-transformed (natural logarithm) to adhere to a normality assumption. Multiple linear regression analyses with HOMA-IR as the dependent variable were performed to evaluate associations of these variables with HOMA-IR. The predictive values of each variable were judged by comparing the proportion of the total variation for each index; in other words, the $R^2$ for a base model that excluded each of the indexes subtracted from the $R^2$ for the entire regression model. We divided each index into increasing quartile values and calculated odds
ratios (ORs) and 95% confidence intervals (CIs) for IR, and compared indexes in quartiles 2–4 with those in quartile 1 using logistic regression analysis. To detect individuals with elevated IR, we compared the relative diagnostic strengths of lipid parameters, lipid ratios, visceral adiposity indicators, TyG related parameters and adipokines by plotting receiver operating characteristic (ROC) curves. ROC curves were constructed and the area under the curves (AUCs) was calculated for each evaluated variable using a HOMA-IR index 2.43 as the reference value to define IR. The AUCs for all variables of interest were compared nonparametrically. The significant difference among AUCs was calculated using an online tool [http://vassarstats.net/roc_comp.html] [21]. A two-tailed P value < 0.05 was considered statistically significant.

Results

BMI, WC, systolic, diastolic and mean BP, TG, non-HDL-C, TC/HDL-C, TG/HDL-C, low-density lipoprotein cholesterol (LDL-C)/HDL-C, non-HDL-C/HDL-C, current smokers, FPG, insulin, HOMA-IR, leptin, AR index and CRP were significantly higher, whereas HDL-C, and adiponectin were lower in insulin-resistant individuals than in their noninsulin-resistant counterparts (Table 1). BMI, WC, diastolic, and mean BP, TG, LDL-C, non-HDL-C, TC/HDL-C, TG/HDL-C, LDL-C/HDL-C, non-HDL-C/HDL-C, current smokers, alcohol consumption, FPG, insulin, HOMA-IR, AR index and CRP were significantly higher, whereas age, HDL-C, adiponectin, leptin, and LAR were lower in men than in women (Table 1).

After controlling for potential intermediate variables of lipid traits, studied parameters individually significantly predicted HOMA-IR except TC, LDL-C, and non-HDL-C (Table 2). The models with lipid ratios were consistently superior to those with single variables for predicting HOMA-IR. For all lipid ratios, such as TC/HDL-C, TG/HDL-C, LDL-C/HDL-C, and non-HDL-C/HDL-C, the additional percentage of variation ranged from 0.8% to 7.0% in HOMA-IR, with TG/HDL-C being the strongest predictor. For visceral adiposity indicators and TyG related parameters, TyG-BMI had the strongest association with HOMA-IR, explaining 16.6% of the variability in HOMA-IR, followed by TyG-WC (14.2%), LAP (11.3%), VAI (9.7%) and TyG (8.5%). For adipokines, LAR had the strongest association with HOMA-IR, which explained 23.2% of the variability in HOMA-IR, followed by leptin (22.2%), adiponectin (6.6%), and the AR index (0.7%), whereas resistin was not associated with HOMA-IR.

The direct comparative ORs and 95% CIs for those in the top quartiles of each variable are presented in Table 3. After adjusting for age, sex, smoking status, systolic and diastolic BP, alcohol consumption, exercise and CRP levels, each of the lipid traits or lipid ratios, including TG, HDL-C and TG/HDL-C were strongly associated with IR (P < 0.001), whereas TC, LDL-C, non-HDL-C, and LDL-C/HDL-C were not associated with IR. Among all lipid parameters, in terms of their strengths of association with IR, TG related parameters were more efficient overall than other lipid parameters, with TG and TG/HDL-C had the strongest association (top vs. bottom OR [95% CI]: 3.11 [1.58–6.11] and 2.98 [1.49–5.96], respectively). All visceral adiposity indicators and TyG related parameters were strongly associated with IR (P < 0.001) and TyG-BMI had the highest OR [95% CI] (6.10[2.51–14.80]). For adipokines, leptin, adiponectin, LAR, and the AR index were associated with IR, and LAR had the highest OR [95% CI] (12.99[3.95–42.70]).

To detect each evaluated variables with IR, the ROC curves of the lipid parameters, lipid ratios, visceral adiposity indicators, TyG related parameters and adipokines were plotted, and the AUCs were compared (Table 4). The AUCs derived from the lipid ratios were in general significantly larger than those from individual lipids. For the lipid ratios, the AUC was the largest for TG/HDL-C (0.707). For all variables of interest, TyG-BMI and LAR had the largest AUC (0.801), followed by TyG-WC (0.772), LAP (0.761), VAI (0.743), TyG (0.708), and leptin.
The TyG-BMI AUC in detecting IR was significantly higher than that of TyG, TG/HDL-C, and leptin ($P = 0.004, 0.004$ and $<0.001$, respectively). The AUC of LAR in detecting IR was also larger than that of TyG, TG/HDL-C, and leptin ($P = 0.003, 0.003$ and $<0.001$, respectively).

Discussion

In this study, we compared lipids, adipokines, lipid and adipokine ratios, VAI, LAP, TyG, TyG-BMI, and TyG-WC as variables in identifying IR. Our investigation revealed that TyG-BMI, with the combination of TG, FPG, and adiposity status, was a more efficient surrogate marker for early identification of IR in nondiabetic Taiwanese people. Although LAR has a comparable effect on IR, it is clinically less useful because it is not routinely measured in clinical practice. Thus, our data showed that TyG-BMI is a simple and clinically useful surrogate marker for IR in nondiabetic individuals. In addition, the ROC curve investigation affirmed that TyG-BMI is the most favorable surrogate marker of IR.

### Table 2. Multiple Linear Regression Models for Predicting Homeostatic Model Assessment.

|                          | Additional R² | β     | P     |
|--------------------------|---------------|-------|-------|
| **Lipid measures**       |               |       |       |
| Ln TC                    | 0.001         | -0.199| 0.434 |
| Ln TG                    | 0.059         | 0.476 | < 0.001 |
| Ln LDL-C                 | 0.003         | -0.041| 0.170 |
| Ln non-HDL-C             | 0.002         | 0.034 | 0.330 |
| Ln HDL-C                 | 0.043         | -0.189| < 0.001 |
| **Lipid ratios**         |               |       |       |
| Ln TC/HDL-C              | 0.024         | 0.157 | < 0.001 |
| Ln TG/HDL-C              | 0.070         | 0.087 | < 0.001 |
| Ln LDL-C/HDL-C           | 0.008         | 0.052 | 0.034 |
| Ln non-HDL-C/HDL-C       | 0.026         | 0.094 | < 0.001 |
| **Visceral adiposity indicators** |             |       |       |
| Ln LAP                   | 0.113         | 0.096 | < 0.001 |
| Ln VAI                   | 0.097         | 0.096 | < 0.001 |
| **TyG related parameters** |             |       |       |
| TyG index                | 0.085         | 0.116 | < 0.001 |
| TyG-BMI                  | 0.166         | 0.003 | < 0.001 |
| TyG-WC                   | 0.142         | 0.001 | < 0.001 |
| **Adipokines**           |               |       |       |
| Ln Leptin                | 0.222         | 0.138 | < 0.001 |
| Ln Adiponectin           | 0.066         | -0.092| < 0.001 |
| Ln Resistin              | -0.012        | -0.013| 0.370 |
| Ln LAR                   | 0.232         | 0.100 | < 0.001 |
| AR index                 | 0.007         | 0.077 | 0.001 |

$P$ value was adjusted for age, sex, smoking, systolic blood pressure, diastolic blood pressure, alcohol consumption, exercise and C-reactive protein. TC, total cholesterol; TG, triglycerides; LDL-C, low-density lipoprotein cholesterol; HDL-C, high-density lipoprotein cholesterol; LAP, Lipid accumulation product; VAI, visceral adiposity index; TyG index, the product of triglycerides and fasting glucose; BMI, body mass index; WC, Waist circumference; LAR: leptin-adiponectin ratio; AR index: adiponectin-resistin index.

doi:10.1371/journal.pone.0149731.t002

(0.690). The TyG-BMI AUC in detecting IR was significantly higher than that of TyG, TG/HDL-C, and leptin ($P = 0.004, 0.004$ and $<0.001$, respectively). The AUC of LAR in detecting IR was also larger than that of TyG, TG/HDL-C, and leptin ($P = 0.003, 0.003$ and $<0.001$, respectively).
Previous studies proposed that traditional lipid ratios, particularly the TG/HDL-C ratio based on the proportion between proatheroagenic and antiatherogenic fractions, are more effective than single lipids measures in identifying IR [6,22,23]. Concordant with these studies, our investigation revealed that the magnitude of the association was higher for TG related parameters, especially TG/HDL-C, than for any other lipid measures or ratios. Therefore, TG/HDL-C can be reasonably proposed as a simple and convenient IR surrogate marker. The mechanism through which hypertriglyceridemia and low HDL-C cause IR remains unresolved. One previous study has proposed that hypertriglyceridemia might cause fatty acid accumulation in non-adipose tissues such as the liver, muscle, and heart which resulted in ectopic lipid deposition with lipotoxicity that has been accepted as a mechanism for IR [24].

Adipose tissue has been recognized as a principal endocrine organ. By secreting an increasing number of adipokines, the adipose tissue is involved in the regulation of whole-body energy balance and insulin action [25]. The altered adipokine profile in obesity leads to profound changes in insulin sensitivity and various metabolic derangements. Leptin and adiponectin are the two most easily characterized adipokines that respond reciprocally to increasing adiposity

### Table 3. Adjusted Odds Ratios of Insulin Resistance among Those in the Lowest Quartile and Upper Three Quartiles of Each Variable.

|                        | Odds ratio  |
|------------------------|-------------|
| **Lipid measures**     |             |
| TC                     | 1.17 (0.68–2.02) |
| TG                     | 3.11 (1.58–6.11)* |
| LDL-C                  | 1.04 (0.60–1.79) |
| Non-HDL-C              | 1.59 (0.88–2.86) |
| HDL-C                  | 0.26 (0.15–0.44)* |
| **Lipid ratios**       |             |
| TC/HDL-C               | 2.23 (1.16–4.30) |
| TG/HDL-C               | 2.98 (1.49–5.96)* |
| LDL-C/HDL-C            | 1.83 (0.99–3.38) |
| non-HDL-C/HDL-C        | 2.23 (1.16–4.30) |
| **Visceral adiposity indicators** |         |
| LAP                    | 3.19 (1.56–6.55) |
| VAI                    | 3.20 (1.57–6.55)* |
| **TyG related parameters** |         |
| TyG index              | 4.11 (2.00–8.46)* |
| TyG-BMI                | 6.10 (2.51–14.80)* |
| TyG-WC                 | 3.55 (1.71–7.36)* |
| **Adipokines**         |             |
| Leptin                 | 5.60 (2.54–12.37)* |
| Adiponectin            | 0.32 (0.19–0.54)* |
| Resistin               | 0.73 (0.43–1.24) |
| LAR                    | 12.99 (3.95–42.70)* |
| AR index               | 2.37 (1.31–4.29) |

*P < 0.001

**Abbreviations as in Table 2**

P value was adjusted for age, sex, smoking, systolic blood pressure, diastolic blood pressure, alcohol consumption, exercise and C-reactive protein.

Data are presented as Odds ratio (95% confidence interval).

doi:10.1371/journal.pone.0149731.t003
Accumulating evidence suggests that hyperleptinemia and hypoadiponectinemia are associated with increased adiposity and IR in nondiabetic subjects [19]. The strength of associations between LAR and HOMA-IR is greater than that of leptin or adiponectin alone [19]. The AR index, combined with adiponectin and resistin levels, is a cost-effective and precise diagnostic biomarker for insulin sensitivity and is a predictor of acute coronary syndrome [20]. Our data revealed that the LAR, not the AR index, is an extremely powerful indicator for IR; however, as leptin and adiponectin assays are more expensive than glucose and triglycerides assays, their clinical utility is limited.

Both VAI and LAP are visceral adiposity indicators involving lipid variables and adiposity status for identifying IR. VAI is a sex-specific index in which the BMI and WC measurements are combined with TG and HDL-C to identify IR and cardiometabolic risk [9,10]. LAP, calculated using WC and fasting TG levels, is an index for excessive lipid accumulation and is a favorable surrogate for recognizing IR in non-diabetic people [27] and for identifying diabetes or CV risk in individuals in previous studies [12,18]. Our research showed that VAI and LAP

| Table 4. Areas Under the Receiver Operating Characteristic (ROC) Curves for Each Evaluated Variable in Predicting Homeostatic Model Assessment. |
| --- |
| Lipid measures |
| Ln TC | 0.531***
| Ln TG | 0.690***
| Ln LDL-C | 0.526***
| Ln non-HDL-C | 0.604**
| Ln HDL-C | 0.313***
| Lipid ratios |
| Ln TC/HDL-C | 0.679***
| Ln TG/HDL-C | 0.707*
| Ln LDL-C/HDL-C | 0.645***
| Ln non-HDL-C/HDL-C | 0.679***
| Visceral adiposity indicators |
| Ln LAP | 0.761
| Ln VAI | 0.734
| TyG related parameters |
| TyG index | 0.708*
| TyG-BMI | 0.801
| TyG-WC | 0.772
| Adipokines |
| Ln Leptin | 0.690***
| Ln Adiponectin | 0.293***
| Ln Resistin | 0.487***
| Ln LAR | 0.801
| AR index | 0.645***

*P < 0.005 compare to TyG-BMI,
**P < 0.001 compare to TyG-BMI
* P < 0.005 compare to Ln LAR,
** P < 0.001 compare to Ln LAR
Abbreviations as in Table 2.

doi:10.1371/journal.pone.0149731.t004
are more efficient than TG/HDL-C in recognizing IR because both parameters reflect abdominal obesity, particularly visceral fat, which is strongly correlated with IR.

Previous studies revealed that both TG and FPG levels are well validated for their roles in identifying IR [28,29]. High FPG is an independent risk factor for type 2 diabetes in people with normoglycemic levels [30]. In obese youth with normal FPG levels, insulin sensitivity declines when moving from low to high FPG [31]. With the combination of TG and FPG levels, the TyG index presents moderate power as a surrogate marker for recognizing HOMA-IR index in healthy subjects [4]. The TyG index has high sensitivity and specificity compared with the euglycemic hyperinsulinemic clamp test for recognizing IR [5]. The superiority of the TyG index in identifying IR was also reported in many other studies [6,7,8]. In the current study, TyG is more efficient than TG/HDL-C for identifying IR, supporting that both lipotoxicity and glucotoxicity have crucial roles in IR modulation.

To our knowledge, this is the first study to compare lipid, adipokines, and lipid and adipokine ratios, visceral adiposity indicators and TyG related parameters as surrogate markers of IR. TyG-BMI is a clinically useful surrogate in identification of IR, as it combines TG, FPG, and adiposity, and all the parameters are well validated for their roles in recognizing IR [30]. TyG-BMI positively correlated with HOMA-IR, and the ROC curve analysis suggested that TyG-BMI had the largest AUC (0.801), thus demonstrating its superior performance in recognizing IR than other variables. The availability of a simple and economical tool, such as TyG-BMI, is favorable for identifying high-risk individuals, thus expanding the benefits of screening. The ROC curve analysis showed that both TyG-BMI and LAR were effective in discriminating IR in the study population, revealing a similar capacity to discriminate this correlation. However, TyG-BMI, a simple and economical IR surrogate marker based on a single routine fasting blood test and BMI measurement, could have more favorable clinical values as an early indicator of IR risk.

Although the clinical appeal for the use of a measure of visceral fat is undeniable, results from studies that have attempted to compare different measurements of general and regional adiposity (BMI vs. WC) have not been consistent [32,33,34]. BMI is simple to measure and is commonly adopted as a useful indicator of general obesity and other metabolic abnormalities. Height and weight, used to calculate BMI, can be reliably measured, even though the BMI cannot distinguish body fat from fat-free mass. In our investigation, TyG-BMI performed more efficiently than TyG-WC did in identifying IR, possibly because WC, although easy to obtain, is an inaccurate measure of abdominal obesity [35]. Abdominal obesity includes subcutaneous and visceral adipose tissues. Visceral adipose tissues are crucial in identifying IR. WC alone cannot be used to differentiate between subcutaneous and visceral adipose tissues; therefore, WC cannot fully represent visceral adipose tissues. Imaging techniques, such as MRI and CT, can more accurately measure fat distribution and quantify adiposity; however, they are not readily accessible, economic, or usable in epidemiological studies. Another explanation is that WC is highly correlated with cardiometabolic risks; however, it is subject to inter-observer variability [33]. Unreliable measurement of the exposure variable can attenuate the observed association of exposure with the disease of interest, thereby reducing the power of the variable to detect a true association [36]. Possible measurement errors may arise from inadequate training and measurement difficulties. Inter-observer reliabilities for circumferences observed may range from 86% to 99% for WC [36]. In another study, the intra- and inter-observer variabilities of WC were higher than those of BMI [37]. In addition, WC differs depending on the precise site at which they are measured [38]. Reliable measurements of body fat and fat distribution are vital in epidemiological surveys. Anthropometric measurement error cannot be avoided but can be minimized by closely monitoring all aspects of data collection.
Our study has the following limitations. First, because of its cross-sectional design, the associations are not prospective and causality cannot be inferred. Further longitudinal study is necessary to confirm if the TyG-BMI index may predict future occurrence of IR. Second, because the study sample primarily includes Taiwanese individuals, the results cannot be generalized to other ethnicities; given the variability of TG levels according to ethnicity, further research is required to evaluate TyG-BMI in different populations.

Conclusion
A simple and inexpensive method to measure IR for routine screening, prospective study, risk assessment, and therapeutic monitoring is essential. TyG-BMI comprehensively identifies and measures obesity and metabolic abnormalities in individuals. The simplicity of TyG-BMI calculation and low-cost biochemical measurements warrant further investigation of its role as a surrogate marker for IR to improve the identification of subjects with a high cardiometabolic risk and facilitate the prevention of chronic diseases associated with IR.

Acknowledgments
This study was supported by a grant from the National Science Council, Taiwan (No. NSC101-2314-B-303-023-MY3) and grants from the Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation (No. TCRD-TPE-99-07, TCRD-TPE-NSC-102-01, TCRD-TPE-MOST-103-01 and No. TCRD-TPE-103-R-2) and Tzu Chi University (No. TCRP102001-02Y1 TCRP99001-04Y1) to YL Ko. We greatly appreciate the technical support from the Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation Core Laboratory.

Author Contributions
Conceived and designed the experiments: YLK LKE. Performed the experiments: SW YCS. Analyzed the data: SW MST LKE. Contributed reagents/materials/analysis tools: LAH HHC. Wrote the paper: LKE.

References
1. Matthaei S, Stumvoll M, Kellerer M, Haring HU. Pathophysiology and pharmacological treatment of insulin resistance. Endocr Rev. 2000; 21:585–618. PMID: 11133066
2. Lillioja S, Mott DM, Spraul M, Ferraro R, Foley JE, Ravussin E, et al. Insulin resistance and insulin secretory dysfunction as precursors of non-insulin-dependent diabetes mellitus. Prospective studies of Pima Indians. N Engl J Med. 1993; 329:1988–1992. PMID: 8247074
3. DeFronzo RA, Tobin JD, Andres R. Glucose clamp technique: a method for quantifying insulin secretion and resistance. Am J Physiol. 1979; 237:E214–E223. PMID: 382871
4. Simental-Mendia LE, Rodriguez-Moran M, Guerrero-Romero F. The product of fasting glucose and triglycerides as surrogate for identifying insulin resistance in apparently healthy subjects. Metab Syndr Relat Disord. 2008; 6:299–304. PMID: 19067533
5. Guerrero-Romero F, Simental-Mendia LE, Gonzalez-Ortiz M, Martinez-Abundis E, Ramos-Zavala MG, Hernandez-Gonzalez SO, et al. The product of triglycerides and glucose, a simple measure of insulin sensitivity. Comparison with the euglycemic hyperinsulinemic clamp. J Clin Endocrinol Metab. 2010; 95:3347–3351. PMID: 20494475
6. Du T, Yuan G, Zhang M, Zhou X, Sun X, Yu X. Clinical usefulness of lipid ratios, visceral adiposity indicators, and the triglycerides and glucose index as risk markers of insulin resistance. Cardiovasc Diabetology. 2014; 13:146.146–155. PMID: 25926814
7. Vasques AC, Novaes FS, de Oliveira Mda S, Souza JR, Yamanaka A, Pareja JC, et al. TyG index performs better than HOMA in a Brazilian population: a hyperglycemic clamp validated study. Diabetes Res Clin Pract. 2011; 93:e98–e100. PMID: 21665314 doi: 10.1016/j.diabres.2011.05.030
8. Lee SH, Han K, Yang HK, Kim MK, Yoon KH, Kwon HS, et al. Identifying subgroups of obesity using the product of triglycerides and glucose: the Korea National Health and Nutrition Examination Survey, 2008–2010. Clin Endocrinol (Oxf). 2014; 82:213–20. PMID: 24841432
9. Amato MC, Giordano C, Galia M, Criscimanna A, Vitabile S, Midiri M, et al. Visceral adiposity index: A reliable indicator of visceral fat function associated with cardiometabolic risk. Diabetes Care. 2010; 33:920–922. PMID:20067971 doi: 10.2337/dc09-1825

10. Ciresi A, Amato MC, Pizzolanti G, Giordano Galluzzo C. Visceral adiposity index is associated with insulin sensitivity and adipocytokine levels in newly diagnosed acromegalic patients. J ClinEndocrinol-Metab. 2012; 97:2907–2915. PMID:22679062

11. Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type 2 diabetes. Nature. 2006; 444: 840–846. PMID:17167471

12. Ioachimescu AG, Brennan DM, Hoar BM, Hoogwerf BJ. The lipid accumulation product and all-cause mortality in patients at high cardiovascular risk: a PreCIS Database Study. Obesity. 2010; 18:1836–1844. PMID:20035284 doi: 10.1038/oby.2009.453

13. Teng MS, Hsu LA, Wu S, Sun YC, Juan SH, Ko YL. Association of CDH13 Genotypes/Haplotypes with Circulating Adiponectin Levels, Metabolic Syndrome, and Related Metabolic Phenotypes: The Role of the Suppression Effect. PLoS One. 2015; 10:e0122664. PMID:25875811 doi: 10.1371/journal.pone.0122664

14. Bonora E, Targher G, Albieriche M, Bonadonna RC, Saggiani F, Zenere MB, et al. Homeostasis model assessment closely mirrors the glucose clamp technique in the assessment of insulin sensitivity: studies in subjects with various degrees of glucose tolerance and insulin sensitivity. Diabetes Care. 2000; 23:57–63. PMID:10857969

15. Wu TL, Chen Tsai, Chang PY, Tsao KC, Sun CF, Wu LL, et al. Establishment of an in-house ELISA and the reference range for serum amyloid A (SAA): complementarity between SAA and C-reactive protein as markers of inflammation. ClinChimActa. 2007; 376:72–6. PMID:16916504

16. Hsu LA, Ko YL, Wu S, Teng MS, Chou HH, Chang CJ, et al. Association of soluble intercellular adhesion molecule-1 with insulin resistance and metabolic syndrome in Taiwanese. Metabolism. 2009; 58:983–9. PMID:19394054 doi: 10.1016/j.metabol.2009.02.021

17. Chou HH, Hsu LA, Wu S, Teng MS, Sun YC, Ko YL. Leptin-to-Adiponectin Ratio is Related to Low Grade Inflammation and Insulin Resistance Independent of Obesity in Non-Diabetic Taiwanese: A Cross-Sectional Cohort Study. Acta Cardiol Sin. 2014; 30:204–214

18. Kahn HS. The lipid accumulation product is better than BMI for identifying diabetes. A population-based comparison. Diabetes Care. 2006; 29:151–3. PMID:16373916

19. Finucane FM, Luan J, Wareham NJ, Sharp SJ, O'Rahilly S, Balkau B, et al. Correlation of the leptin:adiponectin ratio with measures of insulin resistance in non-diabetic individuals. Diabetologia. 2009; 52:2345–9. PMID:19756488 doi: 10.1007/s00125-009-1508-3

20. Singh P, Sridhar MG, Rajappa M, Balachander J, Kadhiravan T. Adiponectin-resistin index and its strong association with acute coronary syndrome in South Indian men. Inflamm Res. 2014; 63:961–8. PMID:25217005 doi: 10.1007/s00011-014-0771-z

21. Hanley JA, McNeil BJ. The meaning and use of the area under a Receiver Operating Characteristic (ROC) curve. Radiology. 1982; 143;29–36. PMID:7063747

22. Kannel WB, Vasan RS, Keyes MJ, Sullivan LM, Robins SJ. Usefulness of the triglyceride-high-density lipoprotein versus the cholesterol-high-density lipoprotein ratio for predicting insulin resistance and cardiometabolic risk (from the Framingham Offspring Cohort). Am J Cardiol. 2008; 101:497–501. PMID:18312765 doi: 10.1016/j.amjcard.2007.09.109

23. Kimm H, Lee SW, Lee HS, Shim KW, Cho CY, Yun JE, et al. Associations between lipid measures and metabolic syndrome, insulin resistance and adiponectin.—Usefulness of lipid ratios in Korean men and women. Circ J. 2010; 74:931–937. PMID:20215701

24. Bickerton AS, Roberts R, Fielding BA, Hodson L, Blaak EE, Wagenmakers AJ, et al. Preferential uptake of dietary Fatty acids in adipose tissue and muscle in the postprandial period. Diabetes. 2007; 56:168–76. PMID:17192479

25. Kershaw EE, Flier JS. Adipose tissue as an endocrine organ. J ClinEndocrinol-Metab. 2004; 89:2548–2556. PMID:15181022

26. Badman MK, Flier JS.The adipocyte as an active participant in energy balance and metabolism. Gastroenterology. 2007; 132:2103–2115. PMID:17498506

27. Xia C, Li R, Zhang S, Gong L, Ren W, Wang Z, et al. Lipid accumulation product is a powerful index for recognizing insulin resistance in non-diabetic individuals. Eur J ClinNutr. 2012; 66:1035–8. PMID:22781025

28. Taniguchi A, Fukushima M, Sakai M, Kataoka K, Nagata I, Doi K, et al. The role of the body mass index and triglyceride levels in identifying insulin-sensitive and insulin-resistant variants in Japanese non-insulin-dependent diabetic patients. Metabolism. 2000; 49:1001–5. PMID:10954017
29. Taniguchi A, Nakai Y, Sakai M, Yoshii S, Hamanaka D, Hatae Y, et al. Relationship of regional adiposity to insulin resistance and serum triglyceride levels in nonobese Japanese type 2 diabetic patients. Metabolism. 2002; 51:544–8. PMID: 11979383

30. Tirosh A, Shai I, Bitzur R, Kochba I, Tekes-Manova D, Israeli E, et al. Changes in triglyceride levels over time and risk of type 2 diabetes in young men. Diabetes Care. 2008; 31: 2032–2037. PMID: 18591400 doi: 10.2337/dc08-0825

31. O'Malley G, Santoro N, Northrup V, D'Adamo E, Shaw M, Eldrich S, et al. High normal fasting glucose level in obese youth: a marker for insulin resistance and beta cell dysregulation. Diabetologia. 2010; 53:1199–209. PMID: 20204321 doi: 10.1007/s00125-010-1693-0

32. Wahrenberg H, Hertel K, Leijonhufvud BM, Persson LG, Toft E, Amer P. Use of waist circumference to predict insulin resistance: retrospective study. BMJ. 2005; 330:1363–4. PMID: 15833749

33. Pouliot MC, Després JP, Lemieux S, Moorjani S, Bouchard C, Tremblay A, et al. Waist circumference and abdominal sagittal diameter: best simple anthropometric indexes of abdominal visceral adipose tissue accumulation and related cardiovascular risk in men and women. Am J Cardiol. 1994; 73:460–468. PMID: 8141087

34. Kotlyarevska K, Wolfgram P, Lee JM. Is waist circumference a better predictor of insulin resistance than body mass index in U.S. adolescents? J Adolesc Health. 2011; 49:330–3. PMID: 21856529 doi: 10.1016/j.jadohealth.2010.12.008

35. Kuk JL, Lee S, Heymsfield SB, Ross R. Waist circumference and abdominal adipose tissue distribution: influence of age and sex. Am J Clin Nutr. 2005; 81:1330–1334. PMID: 15941883

36. Ulijaszek SJ, Kerr DA. Anthropometric measurement error and the assessment of nutritional status. Br J Nutr. 1999; 82:165–77. PMID: 10655963

37. Nadas J, Putz Z, Kolev G, Nagy S, Jeremendi G. Intraobserver and interobserver variability of measuring waist circumference. Med Sci Monit. 2008; 14:CR15–8. PMID: 18160939

38. Wang J, Thornton JC, Bari S, Williamson B, Gallagher D, Heymsfield SB, et al. Comparisons of waist circumferences measured at 4 sites. Am J Clin Nutr. 2003; 77:379–84. PMID: 12540397