Response of biotester *Lepidium sativum* to the effects of native herb extracts and phytoinvader *Solidago canadensis*

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**Abstract.** The effects of aqueous extracts from aboveground parts of *Solidago canadensis* L. as well as from dominant species of four main succession stages on the degraded forest soils of the Kemerovo region (*Filipendula ulmaria*, *Chamaenerion angustifolium*, *Cirsium setosum*, *Poa pratensis*) on the germinating capacity and speed of *Lepidium sativum* seeds were investigated. It was showed that *Solidago canadensis* and *Filipendula ulmaria* have the greatest effect on the reduction of germinating capacity of *Lepidium sativum* seeds.

1 **Introduction**

According to a number of scientific reports, *Solidago canadensis* can suppress the development of many herbs on the different stages of ontogenesis [1 – 5] leading to the dysfunction of native phytocenosis and failure of disturbed land restoration caused by *S. canadensis* invasion beyond their native areal (North America) [6 – 10].

To study the effects of *Solidago canadensis* L. on the germinating capacity of other plants’ seeds and to compare the obtained effects with ones of dominant species of four main succession stages on the degraded forest soils of the Kemerovo region (*Filipendula ulmaria*, *Chamaenerion angustifolium*, *Cirsium setosum*, *Poa pratensis*) we selected *Lepidium sativum* L. (cress) characterized by the most balanced differentiating ability in assessing the allelopathic activity of various “donor plants” [11].

2 **Materials and Methods**

Seeds of biotester *Lepidium sativum* L. were couched under exposure to the aqueous extracts from the aboveground parts of five herbs with a different life strategy.

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2.1 Object of study

Testing culture: seeds of *Lepidium sativum* L. (cress). Aqueous extracts were obtained from crushed dry aboveground parts (the competitiveness of herbs decreases in a row during the development of disturbed ecotopes):

1. *Solidago canadensis* L. (Canadian goldenrod) – potentially invasive species;
2. *Filipendula ulmaria* (L.) Maxim. (meadowsweet) – dominant of climax cenosis;
3. *Chamaenerion angustifolium* (L.) Scop. (willow-herb) – dominant of post-fire upland cenosis;
4. *Cirsium arvense* var. *integrifolium* Wimm. & Grab. = *Cirsium setosum* (Willd.) Besser ex M.Bieb. (yellow thistle) – dominant of early successive fallow cenosis;
5. *Poa pratensis* L. (rough-stalked bluegrass) – dominant of meadow cenosis.

2.2 Methods

As a control, distilled water was used. All experiments were performed in triplicate. Generally, 100 seeds were investigated. Germinated seeds were counted daily following the day when the experiment was started. As the samples drying, the new appropriate extracts were added.

Germination capacity and speed of germination were determined at the different stages of the experiment according to the State Standard No. 12038-84 [12]; total germination energy (GE) of seed per day was calculated [13].

2.3 Statistics

Statistical analysis was performed by Microsoft Excel software using the graphical analysis for minute samples characterized by a different distribution.

3 Results and discussion

The maximum germination period for *L. sativum* seeds was 5 days including the day as the experiment was started. The germination capacity and speed (19% on the first day of the experiment and about 70% in total) was discovered in the control group; the extracts obtained from all studying herbs at the first stage of the experiment were associated with significantly reduced germination (Fig. 1). At the same time, the majority of studied extracts lose their effects after third day of the experiment – the viability of seeds of testing culture approach the control values excluding the extracts obtained from *Solidago canadensis* and *Filipendula ulmaria* associated with the suppressed development of testing culture. Moreover, *Filipendula ulmaria* was associated with three-fold decreasing of germination capacity compared to the control, while potentially invasive *Solidago canadensis* – with only one third one. Generally, average coefficients of germination capacity are equal to the dynamics of germ appearance (Fig. 2). The germination rate of *Lepidium sativum* seeds reflecting the germination speed at a certain stage of the experiment was decreased in the majority of the studied extracts excluding *Poa pratensis*, but the overall germination capacity for the most of variants was equal to the control.
The germination rate of *Lepidium sativum* seeds was 5 days in the majority of the studied extracts excluding *S. canadensis* – invasive species. At the same time, the viability of seeds of testing herbs with a different life strategy was associated with three variants: 1. Herb – dominant of early successive fallow c. – *Cirsium setosum* (Willd.) Besser; 2. Herb – dominant of meadow cenosis – *Filipendula ulmaria* (L.) Wimm. & Grab.; 3. Herb – dominant of climax cenosis – *Chamaenerion angustifolium* (Craib.) Schouw & Forss.; 4. Herb – post–fire herb – *Poa pratensis*; 5. Herb – stalked bluegrass – *Poa pratensis*; 6. Herb – *stalked bluegrass* – *Poa pratensis*. Generally, average coefficients of germination were determined at the different stages of the experiment according to the State Standard No. 12038 of the experiment when the experiment was started. As the samples drying, the new appropriate extracts were added. As a control, distilled water (K) was used in all experiments. Aqueous extracts were obtained from the aboveground parts of the plants and their effects associated with their distribution in the majority of the studied extracts excluding *S. canadensis*. The maximum germination period for *S. canadensis* was equal to the dynamics of germ appearance (20% on the first day of the experiment). Moreover, *S. canadensis* culture approach extracts significantly reduce germination capacity compared to the control, while potentially losing their effects after 1 and 3 quartile, a line in the center of the box – median, error bars – minimum and maximum value, point – mean, error bars (thin line) – standard deviation; here and after: K – control (distilled water), S.c. – *Solidago canadensis*, F.u. – Filipendula ulmaria, Ch.a. – Chamaenerion angustifolium, C.s. – Cirsium setosum, P.p. – Poa pratensis.

Germination energy (GE) of seeds including moments as each part of germ appeared. An increased GE reflects the delay in seeds’ germination. In our experiment, the maximum GE values after exposure to *Filipendula ulmaria* and *Solidago canadensis* extracts (3.5 and 3 days, respectively) were significantly different from other samples (about 2 days).
4 Conclusion

It was shown that aqueous extracts from the aboveground parts of *Cirsium setosum* and *Poa pratensis* have no effects on the germination capacity of *Lepidium sativum* seeds compared to the control. *Chamaenerion angustifolium* can suppress the germination in the first days but then an inhibitory effect disappears. The most pronounced effect on the germination capacity of *L. sativum* seeds was showed for *Solidago canadensis* and *Filipendula ulmaria*.

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References

1. G. Chen, Ch. Zhang, L. Ma, Sh. Qiang, J. A. Silander, L. L. Qi, J. Integrat. Agric. **12**(5), 835–845(2013)
2. A. Fenesi, C. I. Vágási, M. Beldean, R. Földesi, L.-P. Kolcsárb, J. T. Shapiro, E. Török, A. Kovács-Hostyánszkie, Basic Appl. Ecol. **16**, 335–346 (2015)
3. A.P. Gusev Rus. J. Biol. Invas. **6**(2), 74–77 (2015)
4. V.M. Shmelev, A.N. Pankrushina, V. Tver SU **3**(55), 130-135 (2019)
5. C. Wang, B. Wu, K. Jiang, J. Zhou, D. Du, Urban. For. Urban. Gree. **38**, 145–156 (2019)
6. B. Tokarska-Guzik, B. Węgrzynek, A. Urbisz, A. Urbisz, T. Nowak, K. Bzdęga, Biodiv. Res. Conserv. **19**, 33–54 (2010)
7. *Black book of the flora of Siberia* (Novosibirsk, “Geo” Publ., 2016), 440
8. Yu.K. Vinogradova, S.R. Mayorov, L.V. Khorun, *Black data book of the flora of Central Russia* (Moscow, Geos, 2010)
9. M. Stefanowicz, M. Stanek, M. L. Majewska, M. Nobis, S. Zubek, Appl. Soil Ecol. **136**, 168–177 (2019)
10. M. Richardson, P. Pyšek, M. Rejmánek, M. G. Barbour, F. D. Panetta, C. J. Divers, Distrib. **6**(2), P. 93–107 (2000)
11. A.F. Buharov, D.N. Baleev, RUDN J. Agro. Animal Ind. **1**, 32-39 (2012)
12. *GOST R 12038-84 Agricultural seeds. Methods for determination of germination.* (1986)
13. *Agricultural dictionary reference* (Moscow, Leningrad, Selkhozgiz, 1934)