On the relationship between tripartite motif-containing 22 single-nucleotide polymorphisms and COVID-19 infection severity

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Abstract

Background: The tripartite motif containing (TRIM)-22 participates in innate immune responses and exhibits antiviral activities. The present study aimed to assess the relationship between TRIM22 single-nucleotide polymorphisms and clinical parameters with the coronavirus disease 2019 (COVID-19) infection severity.

Methods: TRIM22 polymorphisms (rs7113258, rs7935564, and rs1063303) were genotyped using TaqMan polymerase chain reaction (PCR) assay in 495 dead and 497 improved severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)-positive patients.

Results: In this study, the frequencies of TRIM22 rs1063303 GG, rs7935564 GG, and rs7113258 TT were significantly higher in dead patients than in improved patients, and higher viral load with low PCR Ct value was noticed in dead patients. The multivariate logistic regression analysis revealed that the lower levels of low-density lipoprotein (LDL), cholesterol, PCR Ct value, and lower 25-hydroxyvitamin D, and also higher levels of erythrocyte sedimentation rate (ESR), C-reactive protein (CRP), and TRIM22 rs1063303 GG, rs7113258 TT, and rs3824949 GG genotypes were related to the COVID-19 infection severity.

Conclusion: Our finding proved the probable relationship between the COVID-19 infection severity with the genotypes of TRIM22 SNPs and clinical parameters. More research is required worldwide to show the association between the COVID-19 infection severity and host genetic factors.

Keywords: Tripartite motif-containing 22, Coronavirus disease 2019, Severe acute respiratory syndrome coronavirus 2

Introduction

In December 2019, in Wuhan, China, the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) was the causative agent of an ongoing pandemic of coronavirus disease 2019 (COVID-19), a respiratory disease and exhibiting symptoms similar to pneumonia [1]. In addition to viral genetic variation, social, economic, and host are recognized to have effects on the incidence of COVID-19. Accordingly, the characterization of the host’s genetic factors contributing to this disease will be of great interest in prognosis and treatment [2].

Innate immunity is the first line of defense against viruses and aims to protect a host from the invasion of viruses. A part of this complex network of cells and soluble factors is each cell’s unique ability to initiate a series of intracellular responses to viral infections to limit replication and the further spread of the viruses [3]. The genome of viruses and the early products of viral replication are recognized by many pattern recognition receptors,
resulting in the synthesis of type I interferons (IFNs) and activating the IFN-stimulated genes (ISGs) cascade with antiviral functions. In this regard, some members of the tripartite motif (TRIM) family are antiviral executors [4].

The members of the TRIM protein are divided into 11 families, C-I to C-XI, based on the overall domain structure; however, one group of TRIM proteins remains unclassified due to the absence of the really interesting new gene (RING) domain (e.g., TRIM14 and TRIM20). Many of these proteins have antiviral effects belonging to the C-IV family [5].

TRIM22 is involved in cell restriction of a wide variety of viruses, including human immunodeficiency virus (HIV), hepatitis B virus (HBV), hepatitis C virus (HCV), influenza A virus, encephalomyocarditis virus, and herpes simplex virus, and also TRIM22 single-nucleotide polymorphisms (SNPs) have antiviral effect in several viral infections such as measles, HBV, HCV, and HIV [6, 7].

Many reports have indicated that SNPs might play critical roles in the COVID-19 infection severity, including polymorphisms located around the interferon lambda 3/4 (IFNL3/IFNL4) region, interferon-induced transmembrane protein-3, the transmembrane protease serine 2 (TMPRSS2), and the angiotensin-converting enzyme 2 (ACE2) [8, 9].

However, there are no data on the impact of TRIM22 SNPs on the COVID-19 infection in Iran. Accordingly, the present study aimed at evaluating the effects of three TRIM22 SNPs on the COVID-19 infection recovery and severity.

Materials and methods

Study population

The present study was performed on 992 COVID-19 patients with the same ethnic group at the Pasteur Institute of Iran (PII) from July 2021 to October 2021. In this study, reverse transcriptase real-time polymerase chain reaction (rtReal Time-PCR) from nasopharyngeal or oropharyngeal swab samples was used to detect SARS-CoV-2. There was no underlying disease such as chronic kidney disease, pregnancy, and others.

About 10 ml of blood samples was taken from the patients, and peripheral blood mononuclear cells (PBMCs) were extracted by Ficoll (Ficoll-Paque PLUS, GE Healthcare, USA) density gradient centrifugation and stored at −70 °C. The laboratory parameters including 25-hydroxyvitamin D, C-reactive protein (CRP), white blood cells (WBC), hemoglobin, erythrocyte sedimentation rate (ESR), fasting blood glucose (FBS), low-density lipoprotein (LDL), cholesterol, triglyceride (TG), high-density lipoprotein (HDL), alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), blood urea nitrogen (BUN), serum creatinine, uric acid, triiodothyronine (T3), thyroxine (T4), thyroid-stimulating hormone (TSH), platelets, and real-time PCR Ct values were extracted from the patients’ records.

DNA extraction and TRIM22 SNPs genotyping

According to the manufacturer’s instructions, the genomic DNA of the COVID-19 patients was extracted from PBMCs using the High Pure PCR Template Preparation Kit (Roche Diagnostics Deutschland GmbH, Mannheim, Germany). Three SNPs of TRIM22 (rs1063303, rs7935564, and rs7113258) were genotyped by TaqMan® real-time allelic discrimination method, as previously described [7]. The selection criteria for TRIM22 polymorphisms included SNPs found in a putative regulatory area, minor allelic frequency (MAF) larger than 20% for Asia and Iran, and tagSNP selection based on linkage disequilibrium (LD) > 0.8 [7, 10].

Statistical analysis

All data were analyzed using the statistical software IBM SPSS for Windows version 22.0. (SPSS. Inc., Chicago, IL, USA). The Shapiro–Wilk test was used to evaluate the normality of the ordinal variables. Pearson’s Chi-square and Mann–Whitney U tests were also used to evaluate quantitative and continuous variables, respectively. A multivariate logistic regression analysis was performed using the Hosmer–Lemeshow test to analyze the correlation between COVID19 resistance and some risk factors for susceptibility. A two-tailed P<0.05 was set as the significance level. The area under the receiver-operating characteristic curve (AUC-ROC) analysis was used to assess the effect of TRIM22 SNPs on resistance and susceptibility to COVID19. The correlation between COVID-19 mortality and TRIM22 SNPs and haplotypes analysis were investigated using the SNPStats software inheritance mode analysis. The best-fit model for each SNP was determined using the Akaike information criterion (AIC) and the Bayesian information criterion (BIC). The Chi-square test was used to estimate the Hardy–Weinberg equilibrium (HWE) for genotype distributions [11].

Results

Baseline characteristics of COVID-19 patients

The present study included 992 COVID-19 patients assigned into two groups: dead patients (n = 495) and improved patients (n = 497). The mean ages of dead and improved patients were 62.6 ± 11.9 and 51.1 ± 11.8 years, respectively. The severity of
COVID-19 infection was significantly associated with lower levels of cholesterol \( (P < 0.001) \), TG \( (P = 0.025) \), LDL \( (P < 0.001) \), 25-hydroxyvitamin D \( (P < 0.001) \), and low real-time PCR Ct value \( (P = 0.010) \) and high levels of ESR \( (P < 0.001) \) and CRP \( (P < 0.001) \). Table 1 shows the patient’s laboratory and clinical features.

**Table 1** Comparison laboratory parameters between dead and improved patients infected with COVID-19

| Variables                        | dead patients \( n = 495 \) | Improved patients \( n = 497 \) | \( P \)-value |
|----------------------------------|-----------------------------|---------------------------------|-------------|
| Mean age ± SD                    | 62.6 ± 11.9                 | 51.1 ± 11.8                     | 0.515       |
| Gender (male/female)             | 259/236 (52.3/47.7%)        | 270/227 (54.3/45.7%)            | 0.527       |
| ALT, IU/L (mean ± SD) \( \text{Reference range: 5–40} \) | 32.6 ± 19.5                | 31.2 ± 20.1                     | 0.144       |
| AST, IU/L (mean ± SD) \( \text{Reference range: 5–40} \) | 33.9 ± 18.2                | 30.8 ± 17.4                     | 0.064       |
| ALP, IU/L (mean ± SD) \( \text{Reference range: up to 306} \) | 178.7 ± 81.4               | 163.9 ± 79.4                    | 0.073       |
| Cholesterol, mg/dL (mean ± SD) \( \text{Reference range: 50–200} \) | 116.1 ± 46.4               | 128.5 ± 51.2                    | <0.001*     |
| TG, mg/dL (mean ± SD) \( \text{Reference range: 60–165} \) | 128.7 ± 59.8               | 137.1 ± 61.2                    | 0.025*      |
| LDL, mg/dL (mean ± SD) \( \text{Reference range: up to 150} \) | 62.3 ± 29.7                | 95.4 ± 48.2                     | <0.001*     |
| HDL, mg/dL (mean ± SD) \( \text{Reference range: > 40} \) | 32.1 ± 11.9                | 34.4 ± 12.5                     | 0.813       |
| WBC, 10^9/L (mean ± SD) \( \text{Reference range: 4000–10,000} \) | 7553.0 ± 2680.1            | 7773.5 ± 2865.9                 | 0.245       |
| CRP, mg/L (mean ± SD) \( \text{Reference range: < 10 mg/L Negative} \) | 70.1 ± 21.4                | 51.9 ± 19.2                     | <0.001*     |
| ESR, mm/1st h (mean ± SD) \( \text{Reference range: 0–15} \) | 563.3 ± 15.7               | 45.9 ± 15.1                     | <0.001*     |
| FBS, mg/dL (mean ± SD) \( \text{Reference range: 70–100} \) | 106.8 ± 41.5               | 107.2 ± 42.1                    | 0.806       |
| Platelets \( \times 1000/\text{cumm} \) (mean ± SD) \( \text{Reference range: 140,000–400,000} \) | 185 ± 77                   | 183 ± 68                        | 0.577       |
| T3, ng/dL (mean ± SD) \( \text{Reference range: 2.3–4.2} \) | 3.5 ± 1.6                  | 2.7 ± 1.3                       | 0.066       |
| T4, mcg/dL (mean ± SD) \( \text{Reference range: 5.6–13.7} \) | 9.1 ± 6.8                  | 8.4 ± 6.3                       | 0.128       |
| TSH, mu/L (mean ± SD) \( \text{Reference range: 0.4–4.5} \) | 3.4 ± 2.1                  | 3.1 ± 1.7                       | 0.808       |
| Hemoglobin, g/dL (mean ± SD) \( \text{Reference range: 12–18} \) | 14.2 ± 2.2                 | 12.8 ± 1.5                      | 0.378       |
| BUN, mg/dL (mean ± SD) \( \text{Reference range: 15–45} \) | 39.2 ± 6.2                 | 38.1 ± 7.1                      | 0.527       |
| Creatinine, mg/dL (mean ± SD) \( \text{Reference range: 0.6–1.4} \) | 1.5 ± 0.7                  | 0.9 ± 0.3                       | 0.107       |
| 25-hydroxy vitamin D, ng/mL (mean ± SD) \( \text{Sufficiency: 21–150} \) | 194 ± 9.2                  | 402 ± 12.8                      | <0.001*     |
| Real-time PCR Ct values          | 124.7 ± 6.2                 | 291.9 ± 9.3                     | 0.010*      |
| **TRIM22 rs1063303**             |                             |                                 |             |
| CC genotype                      | 69 (13.9%)                  | 197 (39.6%)                     | <0.001*     |
| GC genotype                      | 258 (52.2%)                 | 224 (45.1%)                     |             |
| GG genotype                      | 168 (33.9%)                 | 76 (15.3%)                      |             |
| **TRIM22 rs7935564**             |                             |                                 |             |
| AA genotype                      | 89 (18.0%)                  | 261 (52.5%)                     | <0.001*     |
| AG genotype                      | 159 (32.1%)                 | 186 (37.4%)                     |             |
| GG genotype                      | 247 (49.9%)                 | 50 (10.1%)                      |             |
| **TRIM22 rs7113258**             |                             |                                 |             |
| AA genotype                      | 109 (22.0%)                 | 294 (59.2%)                     | <0.001*     |
| AT genotype                      | 159 (32.1%)                 | 151 (30.4%)                     |             |
| TT genotype                      | 227 (45.9%)                 | 52 (10.4%)                      |             |

ALT alanine aminotransferase; AST aspartate aminotransferase; ALP alkaline phosphatase; TG triglyceride; LDL low-density lipoprotein; HDL high-density lipoprotein; WBC white blood cells; CRP C-reactive protein; ESR erythrocyte sedimentation rate; FBS fasting blood glucose; T3 triiodothyronine; T4 thyroxine; TSH Thyroid-stimulating hormone; BUN Blood urea nitrogen; Ct cycle threshold; SD standard deviation

*Statistically significant \( (< 0.05) \)
patients with a co-expression of the TRIM22 rs1063303 CC, rs7935564 AA, and rs7113258 AA genotypes (39.0%) had demonstrated a better response to COVID-19 infection compared to patients having other genotypes (32.9%).

SNPStats was used to construct the TRIM22 SNPs inheritance model (codominant, dominant, recessive, and overdominant). The best-fit inheritance model for TRIM22 rs1063303 was codominant with the lowest AIC and BIC. The GG genotype was associated with a higher mortality rate (OR 6.31, 95% CI 4.29–9.28, \( P < 0.0001 \)). Minor allele frequency (G) in improved, dead, and all patients was 0.38, 0.6, and 0.49, respectively (Table 2).

For TRIM22 rs7935564, the best-fitting inheritance model was codominant. A higher risk of death was associated with the GG genotype (OR 14.49, 95% CI 9.83–21.35, \( P < 0.0001 \)). MAF (T-allele) in improved, dead, and all patients was 0.29, 0.66, and 0.47, respectively.

### Table 2

| TRIM22 rs1063303 C > G | Groups | OR (95% CI) | P-value | AIC   | BIC   |
|------------------------|--------|-------------|---------|-------|-------|
|                        | Allele |             |         |       |       |
|                        | C      | 618 (62.3%) | 396 (39.9%) | –     | –     | –     |
|                        | G      | 376 (37.7%) | 594 (60.1%) | –     | –     | –     |
| Codominant             | C/C    | 197 (39.6%) | 69 (13.9%)  | 1.00  | <0.0001* | 1279.0 | 1293.7 |
|                        | C/G    | 224 (45.1%) | 258 (52.1%) | 3.29 (2.37–4.50) | 6.31 (4.29–9.28) |
|                        | G/G    | 76 (15.3%)  | 168 (33.9%) | 6.31 (4.29–9.28) |
| Dominant               | C/C    | 197 (39.6%) | 69 (13.9%)  | 1.00  | <0.0001* | 1293.0 | 1302.8 |
|                        | C/G‑G/G| 300 (60.4%) | 426 (86.1%) | 4.05 (2.97–5.54) |
| Recessive              | C/C–C/G| 421 (84.7%) | 327 (66.1%) | 1.00  | <0.0001* | 1331.8 | 1341.6 |
|                        | G/G    | 76 (15.3%)  | 168 (33.9%) | 2.85 (2.09–3.87) |
| Overdominant           | C/C‑G/G| 273 (54.9%) | 237 (47.9%) | 1.00  | 0.026*  | 1374.3 | 1384.1 |
|                        | C/C    | 224 (45.1%) | 258 (52.1%) | 1.33 (1.03–1.70) |
| Minor allele frequency | G      | 0.38         | 0.6       | –     | –     | –     |

| TRIM22 rs7935564 A > G | Groups | OR (95% CI) | P-value | AIC   | BIC   |
|------------------------|--------|-------------|---------|-------|-------|
|                        | Allele |             |         |       |       |
|                        | A      | 708 (71.4%) | 337 (34.0%) | –     | –     | –     |
|                        | G      | 286 (28.6%) | 653 (66.0%) | –     | –     | –     |
| Codominant             | A/A    | 261 (52.5%) | 89 (18%)  | 1.00  | <0.0001* | 1148.3 | 1163.0 |
|                        | A/G    | 186 (37.4%) | 159 (32.1%) | 2.51 (1.82–3.45) |
|                        | G/G    | 50 (10.1%)  | 247 (49.9%) | 14.49 (9.83–21.35) |
| Dominant               | A/A    | 261 (52.5%) | 89 (18%)  | 1.00  | <0.0001* | 1245.3 | 1255.1 |
|                        | A/G‑G/G| 236 (47.5%) | 406 (82%)  | 5.05 (3.78–6.74) |
| Recessive              | A/A‑A/G| 447 (89.9%) | 248 (50.1%) | 1.00  | <0.0001* | 1178.9 | 1188.7 |
|                        | G/G    | 50 (10.1%)  | 247 (49.9%) | 8.90 (6.33–12.53) |
| Minor allele frequency | G      | 0.29         | 0.69      | –     | –     | –     |

| TRIM22 rs7113258 A > T | Groups | OR (95% CI) | P-value | AIC   | BIC   |
|------------------------|--------|-------------|---------|-------|-------|
|                        | Allele |             |         |       |       |
|                        | A      | 739 (74.5%) | 377 (38.0%) | –     | –     | –     |
|                        | T      | 255 (25.5%) | 613 (62.0%) | –     | –     | –     |
| Codominant             | A/A    | 294 (59.1%) | 109 (22%)  | 1.00  | <0.0001* | 1174.4 | 1189.1 |
|                        | A/T    | 151 (30.4%) | 159 (32.1%) | 2.84 (2.08–3.88) |
|                        | T/T    | 52 (10.5%)  | 227 (45.9%) | 11.77 (8.11–17.10) |
| Dominant               | A/A    | 294 (59.1%) | 109 (22%)  | 1.00  | <0.0001* | 1233.2 | 1243.0 |
|                        | A/T‑T/T| 203 (40.9%) | 386 (78%)  | 5.13 (3.88–6.77) |
| Recessive              | A/A‑A/T| 445 (89.5%) | 268 (54.1%) | 1.00  | <0.0001* | 1216.4 | 1226.2 |
|                        | T/T    | 52 (10.5%)  | 227 (45.9%) | 7.25 (5.17–10.16) |
| Minor allele frequency | T      | 0.26         | 0.62      | –     | –     | –     |

OR odds ratios; CI confidence intervals; AIC Akaike information criterion; BIC Bayesian information criterion
TRIM22 rs7935564 genotypes were compatible with HWE ($P = 0.062$).

The best-fit inheritance model for TRIM22 rs7113258 was codominant with the lowest AIC and BIC. The TT genotype was associated with a higher mortality rate (OR 11.77, 95% CI 8.11–17.10, $P < 0.0001$). MAF (T) in improved, dead, and all patients was 0.26, 0.62, and 0.44, respectively (Table 2). TRIM22 rs7113258 genotypes were compatible with HWE in all patients ($P = 0.221$).

The findings of the haplotype analysis are shown in Table 3. According to our findings, the GGT haplotype was more common in the dead than in the improved patients (OR = 6.83, 95% CI = 4.87–9.59, $P < 0.0001$). Furthermore, the GGA haplotype raised the risk of mortality by OR = 6.34 (95% CI = 3.89–10.34, $P < 0.0001$). In addition, CGT (OR = 4.29, 95% CI = 2.59–7.13, $P < 0.0001$), GAT (OR = 3.02, 95% CI = 1.92–4.87, $P < 0.0001$), and CAT (OR = 2.10, 95% CI = 1.16–3.82, $P = 0.015$) haplotypes caused an increase in the risk of COVID-19 mortality.

Moreover, we used logistic regression for create two different predictor variable with and without the TRIM22 SNPs genotypes. The AUC-ROC values in with and without SNPs were 0.923 and 0.867, respectively, suggesting that host genetic factors are commonly crucial for viral infection resolution (Fig. 1).

Factors associated with COVID-19 infection mortality

We assessed some factors associated with COVID-19 infection mortality using multivariate logistic regression analysis. In dead patients, the COVID-19 infection severity was associated with LDL (OR 0.989, 95% CI 0.981–0.998, $P = 0.021$), cholesterol (OR 1.020, 95% CI 1.003–1.038, $P = 0.024$), 25-hydroxyvitamin D (OR 1.041, 95% CI 1.026–1.056, $P < 0.001$), ESR (OR 0.958, 95% CI 0.947–0.968, $P < 0.001$), CRP (OR 0.980, 95% CI 0.972–0.988, $P < 0.001$), real-time PCR Ct values (OR 1.163, 95% CI 1.047–1.293, $P = 0.005$), TRIM22 rs1063303 GG (OR 0.653, 95% CI 0.508–0.839, $P = 0.001$), TRIM22 rs7935564 GG (OR 0.374, 95% CI 0.295–0.473, $P < 0.001$), and TRIM22 rs7113258 TT (OR 0.446, 95% CI 0.353–0.565, $P < 0.001$) (Table 4).

Table 3  Haplotype analysis of the studied polymorphisms of TRIM22 on COVID-19 mortality

| rs1063303 | rs7935564 | rs7113258 | Improved patients | Dead patients | OR (95% CI) | $P$-value |
|-----------|-----------|-----------|-------------------|---------------|-------------|-----------|
| C         | A         | A         | 0.4054            | 0.1434        | 1.00        | –         |
| G         | G         | T         | 0.0848            | 0.3625        | 6.83 (4.87–9.58) | < 0.0001* |
| G         | A         | A         | 0.1843            | 0.0673        | 0.93 (0.58–1.51) | 0.78      |
| C         | G         | A         | 0.1153            | 0.0711        | 1.35 (0.80–2.29) | 0.26      |
| C         | T         | G         | 0.0491            | 0.1269        | 4.29 (2.59–7.13) | < 0.0001* |
| G         | A         | T         | 0.0707            | 0.0711        | 3.02 (1.92–4.76) | < 0.0001* |
| G         | G         | A         | 0.0386            | 0.0991        | 6.34 (3.89–10.34) | < 0.0001* |
| C         | A         | T         | 0.0520            | 0.0587        | 2.10 (1.16–3.82) | 0.015     |

OR odds ratios; CI confidence intervals

*Statistically significant (< 0.05)

Discussion

The invasion of viral infection into target cells is blocked by several host factors induced by pattern recognition receptors (PRRs) after virus recognition. Primarily, type I IFNs play a role in inducing hundreds of genes with antiviral functions. TRIM22 is one of the genes limiting viral infections. It is constitutively expressed in epithelial cells; however, it is immediately induced by viral infection. In accordance with intracellular innate immunity effector

![ROC Curve for different predictor variable with and without the TRIM22 SNPs genotypes](image-url)
proteins, the function of TRIM22 is broad spectrum and is not specific to a single virus. However, several RNA and DNA viruses are restricted, and many other yet unknown viruses may be sensitive to their antiviral function. Moreover, SNPs in genes involved in the innate immune system can affect the likelihood of a clinical course of viral infection [4, 6, 12]. In other words, this was the first study to investigate whether the TRIM22 SNPs genotypes affect the recovery or severity of COVID-19 infection.

TRIM22 is an interferon-induced protein remarkably inhibiting the various viruses replication, including HIV-1, HCV, and HBV that is located on chromosome 11. Moreover, SNPs on TRIM22 have been associated with several aspects of viral infections such as chronic hepatitis B infection, HIV replication, and specific antibody and cytokine levels after vaccination against measles and rubella [13–18]. Due to TRIM22 constitutive expression in epithelial cells, its potential role in limiting the SARS-CoV-2 infection causing the currently towering COVID-19 pandemic is remarkable. However, additional experiments are recommended to determine whether TRIM22 limits the SARS-CoV-2 replication [19].

In the present study, patients with TRIM22 rs1063303 CC, rs7935564 AA, and rs7113258 AA genotypes had a higher rate of improved COVID-19 infection compared to others.

A significant higher prevalence of TRIM22 rs1063303 C-allele, rs7935564 A-allele, and rs7113258 A-allele carriers was observed in dead patients than in improved patients. The allele frequency of TRIM22 rs1063303, rs7935564, and rs7113258 in this study was 0.49, 0.44, and 0.47, respectively. This was in agreement with our previous study that we indicated the relationship between these SNPs and HCV treatment [7]. The MAF for TRIM22 rs1063303 in European (0.52), African (0.20), Asian (0.12), other Asian (0.43), and Latin American (0.00) was reported in dbSNP the NCBI dbSNP database (https://www.ncbi.nlm.nih.gov/SNP/).

For TRIM22 rs7935564, this value was in European (0.60), African (0.49), Asian (0.18), South Asian (0.52), and Latin American (0.53). Also, for TRIM22 rs7113258, MAF was in European (0.18), African (0.32), Asian (0.36), East Asian (0.41), and Latin American (0.32). Totally, we observed large differences in the frequency of MAF of these SNPs among the various ethnic groups.

GGT and GGA haplotypes had more association with the risk of COVID-19 mortality than other haplotypes. Most patients with COVID-19 who had GGT and GGA haplotypes were affected by the severe form of the disease.

The rs7113258 on TRIM22 is located in 3’ untranslated region (UTR) and could be the binding site for miRNAs. Accordingly, it may have a regulatory effect on the expression of the TRIM22 gene [20]. By binding to levels in the 3’UTR region, miRNAs can function as the negative regulators of gene expression [21]. It is documented that the TRIM22 rs7113258 A allele generates putative target sites for many miRNAs such as hsa-miR-4495, hsa-miR-3148, and hsa-miR-3668; however, the TRIM22 rs7113258 T allele damages these target sites and generates other miRNAs, including hsa-miR-4678 and hsa-miR-3177-5p. Therefore, the differences in miRNAs binding between the TRIM22 rs7113258 genotypes may be associated with the observed correlation [7, 10].

Furthermore, SNPs in TRIM22 are involved in several aspects of viral infections, including the replication of HIV, HBV infection, and HCV infection [7, 10, 17]. In this regard, TRIM22 rs1063303 G>C makes amino acids exchange from arginine to threonine at position 242 of the TRIM22 protein. The TRIM22 rs1063303 GG variant is correlated with the adverse effect of increasing TRIM22 expression and reducing TRIM22’s antiviral activity [22]. In this respect, TRIM22 overexpression was negatively correlated with HCV and HIV-1 viral load [15, 23]. On the other hand, TRIM22 gene silencing increased HIV1 infection in target cells [15]. Moreover, the TRIM22 rs1063303 GG genotype is also correlated with more efficient replication of HIV1 [17]. In this study, the TRIM22 rs1063303 GG genotype was correlated with the COVID-19 infection severity with a high viral load. There is a possibility that the rs1063303 GG genotype may interfere with the regulation of HCV replication and promote a more robust inflammatory response, which could increase the infection severity more efficiently, as reported in HCV infection [10]. Further, the replication of HIV-1 was more effective in PBMCs from patients with TRIM22 rs7935564 G allele than from patients with

### Table 4 Factors associated with dead patients infected with COVID-19

| Factors Baseline predictors | OR (95% CI)      | P-value |
|-----------------------------|-----------------|---------|
| LDL (mg/dL)                 | 0.989 (0.981–0.998) | 0.021*  |
| Cholesterol, mg/dL          | 1.020 (1.003–1.038) | 0.024*  |
| 25-Hydroxyvitamin D, (ng/mL)| 1.041 (1.026–1.056) | <0.001* |
| ESR, (mm/1st h)             | 0.958 (0.947–0.968) | <0.001* |
| CRP, (mg/L)                 | 0.980 (0.972–0.988) | <0.001* |
| Real-time PCR Ct values     | 1.163 (1.047–1.293) | 0.005*  |
| TRIM22 rs1063303 (GG)       | 0.653 (0.508–0.839) | 0.001*  |
| TRIM22 rs7935564 (GG)       | 0.374 (0.295–0.473) | <0.001* |
| TRIM22 rs7113258 (TT)       | 0.446 (0.353–0.565) | <0.001* |

LDL low-density lipoprotein; CRP C-reactive protein; ESR erythrocyte sedimentation rate; Ct cycle threshold; TRIM-22 tripartite motif-22; SD standard deviation

*Statistically significant (<0.05)
The TRIM22 rs7935564 A allele [17]. In the present study, TRIM22 rs7935564 GG was associated with the COVID-19 infection severity.

The TRIM22 rs1063303 GG, rs7935564 GG, and rs7113258 TT genotypes were strongly associated with the severity of COVID-19 infection. Using the ROC curve analysis, the most powerful predictive factor of the COVID-19 infection severity was TRIM22 rs7935564 (GG). To the best of our knowledge, there is no report on the genotypes frequencies of TRIM22 SNPs related to COVID-19 infection. These genotypes of TRIM22 SNPs may be an independent predictor of the COVID-19 infection severity.

In our study, the high levels of CRP and ESR and the low levels of LDL and real-time PCR Ct were significantly associated with the COVID-19 infection severity. These results were in agreement with previous studies, indicating that disease severity was correlated with the lower levels of total LDL and cholesterol [24]. Lowering cellular cholesterol can increase cholesterol uptake from the bloodstream, leading to lower serum HDL and LDL cholesterol levels. This may lead to the upregulation of lipoprotein receptors, especially scavenger receptor class B type 1, thereby enhancing cholesterol uptake into the plasma membrane and SARS-CoV-2 infection rates [25].

T-regulatory lymphocytes (Tregs) are the primary defense against uncontrolled inflammation, and common viral infections. The Treg levels are low in many patients infected with SARS-CoV-2 and may be increased with 25-hydroxyvitamin D supplementation. The low levels of 25-hydroxyvitamin D are associated with an increased inflammatory cytokine and a significantly increased risk of pneumonia and viral upper respiratory tract infections. Moreover, 25-hydroxyvitamin D deficiency is correlated with an increase in thrombotic episodes commonly observed with COVID-19 [26].

In the present study, low Ct of rtRT-PCR was found in dead patients. Some studies have demonstrated that low PCR-Ct levels increase the risk of hospitalization and death in the intensive care unit. A correlation between viral load and the disease severity, as determined by PCR-Ct measurements, was also suggested [9].

One of the limitations of this study was the lack of data on survey correlations between ethnics and these SNPs on TRIM22. Another limitation was the PCR-Ct value; however, this relationship has not yet been fully standardized and quantified.

In conclusion, the present study’s finding revealed a strong correlation between the lower levels of real-time PCR Ct values, 25-hydroxyvitamin D, LDL, cholesterol, and higher levels of ESR and CRP the severity of COVID-19 infection. We also demonstrated that patients with TRIM22 rs1063303 GG, rs7935564 GG, and rs7113258 TT were exposed to more severe COVID-19 infection compared to patients with other genotypes. Further experiments are recommended to confirm the findings.

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Author contributions
NR, SCR, and MM performed the experiments and manuscript preparation; JIMA and MM contributed to clinical samples and data acquisition; FS analyzed data and interpreted data; and AF designed and supervised clinical study, interpreted data, read, and approved manuscript. All authors reviewed the manuscript.

Availability of data and materials
All data generated or analyzed during this study are included in this published article.

Declarations
Ethics approval and consent to participate
The study was approved by the Ethical Committee of PII (IR.PII.REC.1400.042) and was carried out according to the 1975 Declaration of Helsinki and relevant local regulations. Also, written informed consent was directly obtained from all participants.

Consent for publication
Not applicable.

Competing interests
The author(s) declared no potential competing interests with respect to the research, authorship, and/or publication of this article.

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