Puccinia sambuci Infection of American Elderberry Plants

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Abstract. Elderberry rust (Puccinia sambuci Schwein.) Arthur (=P. bolleyana) (Arthur, 1921) disease is frequently found in commercial American elderberry (Sambucus nigra L. subsp. canadensis L.) plantings when an alternate host, Carex sp., is present. To evaluate potential infection periods of P. sambuci on elderberry plants, micrometeorological conditions were monitored. Rust symptoms were observed on elderberry on 5 Apr. 2016, and conditions favorable for possible infection were 9 to 18 °C, 23 hours of continuous leaf wetness, and ≥85% relative humidity. Studies were also conducted to ascertain whether P. sambuci with varying pustule numbers affects fruiting, berry puree quality, or vegetative growth. Fruit yield was reduced by 31% when potted 'Bob Gordon' elderberry averaged six rust pustules per plant compared with noninfected plants. In another experiment, field-grown ‘Wyldewood’ plants averaging 137 rust pustules/cane at harvest had 47% less fruit weight on canes than uninfected canes. Titratable acidity of fruit puree from plants was lower when plants had either 690 rust pustules/plant or 137/ pustules/cane, but soluble solids and pH of puree were unaffected by P. sambuci infection. The effect of rust infection on vegetative growth of elderberry plants also varied with pustule numbers. With a low infection level (six pustules per plant), P. sambuci did not induce premature leaf loss on ‘Bob Gordon’ plants or adversely affect shoot dry weight at the end of the growing season. When P. sambuci infection on ‘Wyldewood’ plants was more severe (137 pustules/cane), greater leaf loss occurred on infected canes than on uninfected canes. At very high infection levels (690 pustules/plant), 'Bob Gordon' plant dry weight was reduced. Because of the potential for fruit yield loss on elderberry plants, control of P. sambuci at relatively low infection levels on this plant may be warranted. Strategies that eliminate or suppress the alternate host would likely reduce the P. sambuci inoculum and limit the potential for elderberry plant infection.

Elderberry [Sambucus nigra L. ssp. canadensis (L.) Boll]] is a high-value crop that is grown and processed into products for niche markets (Charlebois et al., 2010; Mohebalian et al., 2012). Elderberry plants are fruit-bearing, multi-stemmed shrubs with compound leaves native to eastern and central North America. Inflorescences are indeterminate compound umbels, where the outer fruit is the first to mature (Zomlefer, 1994). With the increasing production of elderberry, wild germplasm with vigorous vegetative growth and a high number of umbels with large fruit size has been selected to enhance yields. ‘Wyldewood’, a cultivar that matures during the late season and has high-quality fruit, was selected near Brush Hill, OK, in 1998 (Byers et al., 2010). ‘Bob Gordon’, a productive midseason-ripening cultivar with pendulous umbels, was selected in Osceola, MO, in 1999 (Byers and Thomas, 2011).

Because of continuous elderberry production in a monoculture system, pests have become prevalent. Elderberry rust (Puccinia sambuci Schwein.) Arthur (P. bolleyana) is a common disease found on American elderberry that causes foliar and shoot distortion (Arthur, 1962; Kellerman, 1904; Warmund, 2017). Puccinia sambuci is a heteroecious fungus that requires two hosts, sedge (Carex spp.) and American elderberry, to complete its lifecycle (Mims, 1981; Saccardo, 1891). Of the five spore stages of P. sambuci, pycniospores and aeciospores develop on elderberry, whereas urediniospores, teliospores, and basidiospores develop on Carex spp. (Mims, 1981). At least 13 species of sedge have been reported as an alternate host for P. sambuci (Afshan and Khalid, 2009; Arthur, 1962).

Pycnia are the first signs observed on elderberry leaflets and petioles during early spring and appear as small yellow pustules on adaxial and abaxial surfaces of leaflets and stems. Pycnia are flase-shaped and contain receptive hyphae and pycniospores (Littlefield and Heath, 1979; Petersen, 1974). When pycniospores contact and adhere to receptive hyphae of a compatible mating type, they undergo plasmodagyn, resulting in the formation of dikaryotic mycelium (Mims, 1981). After dikaryotization, mycelia grow intercellularly on the adaxial leaf surface of elderberry plant tissue while producing intracellular haustoria to gain nutrients and develop aecia containing aeciospores (Petersen, 1974). Puc- cinia sambuci aecia are often observed on elderberry in May, with large yellow-orange pustules that cause deformed leaves, stems, and petioles. Aecia produce chains of aeciospores that are wind-blown to the alternate host, a Carex species (Mims, 1981).

After germination and subsequent infection of sedge leaf tissue in the summer, uredinia form on the adaxial surface of leaflets and produce urediniospores that can re-infect sedge plants (Bolley, 1889). During the late summer, uredinia develop into telia that produce two-cell, thick-wall teliospores that can withstand low winter temperatures (Arthur, 1962). During favorable environmental conditions in March or April, each cell of the teliospore germinates and produces a basidium, the site of meiosis, resulting in four haploid basidiospores that are wind-blown to elderberry plants (Petersen, 1974). Basidiospore germination occurs on elderberry tissue and a germ tube is produced, which penetrates the host directly through the cuticle and epidermis, with subsequent development of monokaryon hyphae that form pycnia (Agrios, 2005).

Although the disease cycle of P. sambuci has been described, the epidemiology of elderberry rust and the consequences of infection on host productivity have not been investigated. Therefore, studies were conducted to: 1) determine the effect of P. sambuci infection on fruiting and vegetative growth of elderberry plants; 2) compare soluble solids, pH, and titratable acidity of berry puree from infected and uninfected plants; and 3) characterize environmental conditions associated with potential rust infection of elderberry.

Materials and Methods

Inoculation of elderberry plants in 2014. Twenty-five sedge (Carex frankii Kunth) plants producing P. sambuci teliospores were obtained from a commercial elderberry planting near Hartsburg, MO, on 15 Sept. 2013. Plants were placed in 8.5-L polyethylene containers (A.M. Leonard, Piqua, OH) with native soil (Haymond silt loam) and transported to the University of Missouri Horticulture and Agroforestry Research Center (HARC) near New Franklin, MO. On 26 Nov. 2013, potted plants were maintained in an isolated nursery area until they were covered with a polyethylene foam blanket (Hummer International, St. Louis, MO) and plastic sheeting for winter protection. On 20 Apr. 2014, sedge plants were uncovered.

One-year-old ‘Bob Gordon’ plants were obtained from a commercial nursery (Botany Shop, Joplin, MO) on 10 Mar. 2014 and were transplanted to 8.5-L polyethylene containers using Pro-Mix BX (Premier Tech Horticulture, Quakertown, PA). Plants were pruned to a height of 30 cm above the medium surface, leaving three canes per plant. Dormant oil (Danoil; Drexel Chemical Company, Memphis, TN) was applied to elderberry plants at 7.5 mL·L−1 to control overwintered eriophyid mites (Phyllocoptes wisconsinensis Kiefer). On 14 Mar. 2014, 50 g 15N–9P–12K controlled-release fertilizer (Osmoste; Scotts 880 HORTSCIENCE VOL. 54(5) MAY 2019
Company, Marysville, OH) was applied to the medium surface of potted elderberry plants. After 21 d in the nursery, eight elderberry plants were treated with propiconazole (PropiMax EC; Dow AgroSciences, Indianapolis, IN) fungicide at 11.4 mL L⁻¹. On 20 Apr., a humid environment was established by filling each 189-L polyethylene bag (HDX; Homer TLC, Wilmington, DE) with 10 L of water; then, the bag was sealed. Two days later, eight elderberry plants not treated with propiconazole were misted with sterile water and inoculated with P. sambuci. A 1-x-1-cm piece of infected sedge foliage with approximately four spores was used to mechanically transfer fungal spores to the entire elderberry leaf surface by direct contact. A different leaf section of sedge was used to inoculate each of the six leaves per plant. Each inoculated plant was then placed on top of an inverted 7.6-L pot within a sealed bag for 48 h of incubation at 18 °C in the dark. Eight fungicide-treated elderberry plants were also misted with sterile water, sealed in bags, and incubated to serve as controls. After incubation, plants were removed from bags and arranged in a completely randomized design in the nursery area at HARC. Plants were irrigated with an overhead sprinkler system as needed to prevent moisture stress. The number of rust pustules on each leaflet was recorded on 17 June 2014. At peak ripeness (all berries were dark purple) in August, umbels were removed and the harvest dates were recorded. Berry number and fresh weights of destemmed berries were recorded before fruit was sealed in polyethylene bags and stored at –22 °C. On 1 Oct. 2014, leaves and petioles were harvested and oven-dried for 24 h at 65 °C for dry weight measurements. On 13 Nov. 2014, canes were pruned to 30 cm above the medium surface for dry weight measurements and covered on 28 Nov. 2014 for winter protection (as previously described).

After 60 d in cold storage, fruit was thawed for compositional analyses. Berries from each elderberry plant were pooled and randomly sampled for analyses. To prepare each berry puree sample, 50 g of fruit was placed in a blender cup (Waring, Stamford, CT) with 50 mL of double-distilled water and processed for 30 s. A 0.3-mL aliquot of puree was used to determine soluble solids concentrations with a digital refractometer (Atago USA, Bellevue, WA), and a 10-mL aliquot was used to measure pH (HI222; Hanna Instruments, Woonsocket, RI). To measure titratable acidity, another 10-mL aliquot was diluted with 48 mL of degassed deionized water and titrated (G20 Compact Titrator; Mettler-Toledo, Columbus, OH) to 8.2 pH with 0.1 N of sodium hydroxide. Titratable acidity, expressed as citric acid, was then calculated.

On 1 Apr. 2015, elderberry plants were uncovered, fertilized as described, and maintained in the nursery without further P. sambuci inoculation. Fruit yield was recorded during Aug. 2015 (the second sea-
of pustules per leaf on each elderberry cane were recorded on 31 May.

Fruit harvest, berry number, and berry weight were recorded from 20 July to 23 Aug. 2016. Leaves on each cane were counted on 26 Aug., and the number of those lost since 18 May was calculated. Canes were pruned to 10 cm on 13 Oct., and the dry weight of canes with their foliage was determined. Fruit compositional analyses from berry puree were performed as described. When the harvested fruit was fewer than 50 g per cane, the 1:1 berry weight/double-distilled water volume was used to prepare the puree. A t-test using pooled data from rust-infected canes and uninfected canes was performed for statistical analyses of fruit yield per cane, mean berry weight, puree characteristics, leaf loss, and cane weight data.

**Results**

*Elderberry plants inoculated in 2014.* 'Bob Gordon' elderberry inoculated with *P. sambuci* averaged 690 pustules/plant on 17 June 2014, whereas no rust symptoms were observed on noninoculated plants. Infected plants had lower fruit yield, lower mean berry weight, and lower shoot dry weight than that of noninoculated controls (Table 1). Soluble solids of fruit puree were unaffected by *P. sambuci* infection. Additionally, the pH levels of puree from inoculated (4.7 pH) and noninoculated (4.8 pH) plants were similar (*P* = 0.21). However, fruit puree processed from *P. sambuci*-inoculated plants had lower titratable acidity than that from noninoculated plants. During the following growing season, plants that had been inoculated with *P. sambuci* in 2014 still had reduced yield and plant dry weight compared with those of noninoculated plants, even though rust pustules were not observed in 2015.

**Potential infection periods of *P. sambuci* in 2016.** Between 11 Mar., when 'Bob Gordon' elderberry plants were uncovered, and 17 May 2016, there were 30 potential infection periods, as defined by Beraha et al. (1960), with ≥3 h of continuous leaf wetness, ≥85% relative humidity, and mean maximum hourly temperatures ranging from 9 to 18 °C (Table 2). The minimum drying time of elderberry canes in 2016. The first harvest date for umbels from *P. sambuci*-inoculated plants was 137 on 31 May 2016. The first harvest date for umbels on rust-infected plants occurred 5 d earlier (6 Aug.) than that for control canes (*P* = 0.02). The last flower harvest date (14 Aug.) for infected canes was also 3 d earlier than that for control canes (*P* = 0.04). The fruit yield was reduced by rust infection, but the mean berry weights were similar among all canes (Table 5). Soluble solids and pH (4.4 to 4.5) of berry purees were also similar among canes. However, the titratable acidity of puree from rust-infected canes was lower than that of puree from asymptomatic canes. By 26 Aug., rust-infected canes lost more leaves but had similar cane dry weights on 13 Oct. than the controls.

**Discussion**

Foliar symptoms of *P. sambuci* rust (i.e., pustules) were first observed on 'Bob Gordon' elderberry plants as early as 5 Apr. Many rust pustules were observed later (29 May) on foliage and stems of 'Wyldewood' elderberry of the commercial planting, with the initial infection occurring earlier. In some cases, stem distortion occurred when numerous rust pustules were present. The numbers of pustules on leaflets and stems for each cane or plant were recorded during these studies, but the severity of stem distortion was not evaluated. Although rust pustules were also observed on a few closed flowers and pedicels before bloom through harvest on 'Wyldewood' elderberry of the field planting, canes with infected floral tissue were not included in this study. Rust infection on floral tissue, which has not been previously reported, contributed to mortality and fruit loss. *P. sambuci* infection occurs when germinated basidiospores from sedge plants penetrate susceptible elderberry tissue under favorable environmental conditions, including high humidity or adequate moisture on the plant tissue, for a long enough time over a suitable range of temperatures. During the initial rust infection observed on 5 Apr. 2016, there were five potential infection periods before the first observation of disease symptoms and signs (Table 2). Temperatures during these periods ranged from 10.9 to 13.2 °C, and leaf wetness durations ranged from 3 to 18 h. Of these potential infection periods, conditions occurring from 12 to 13 Mar. may have resulted in puree development because 5.6 mm of precipitation occurred with 18 h of leaf wetness and the mean hourly temperature was 13.2 °C. The number of plants infected and the number of observed pustules (both pycnia and aecia) increased throughout April and May 2016. From 6 to 27 Apr., there were 14 potential infection periods; five of these occurred during precipitation events. Conditions from 18 to 19 Apr. may have resulted in significant infection since 58.7 mm of precipitation occurred with 16 h of leaf wetness and the mean hourly temperature was 16.2 °C. From 29 Apr. to 19 May, 50 more

| Treatment         | 2014         | 2015         |
|-------------------|--------------|--------------|
|                    | Fruit yield/plant (g) | Mean berry wt (mg) | Total soluble solids ('Brix) | Titratable acidity (g/100 mL) | Shoot dry wt (g) | Fruit yield/plant (g) | Plant dry wt (g) |
| Inoculated        | 138 b        | 113 b        | 10.2 a         | 0.37 b                  | 40 b          | 183 b                | 240 b            |
| Noninoculated     | 180 a        | 146 a        | 9.9 a          | 0.63 a                  | 50 a          | 235 a                | 275 a            |

*Table 1. Fruit characteristics, berry puree composition, and vegetative dry weights of ‘Bob Gordon’ elderberry plants inoculated with *Puccinia sambuci* or noninoculated plants in 2014.*

In 2014, inoculated plants averaged 690 rust pustules/plant. In 2015, none of the elderberry plants were re-infected with *P. sambuci*. Values represent the means of eight replications of each treatment. Means followed by different letters in columns are significantly different (*P* ≤ 0.05). Means were separated by a Satterthwaite test when variances were unequal or by a pooled test when variances were equal.
Table 2. Continuous elderberry leaf wetness periods with corresponding mean maximum hourly temperatures and precipitation recorded at the Horticulture and Agroforestry Research Center near New Franklin, MO, during Spring 2016.

| Date           | Continuous leaf wetness (no. hours) | Mean hourly temp during wetness period (°C) | Precipitation during leaf wetness period (mm) |
|----------------|-------------------------------------|-------------------------------------------|---------------------------------------------|
| 12 Mar.        | 3                                   | 12.5                                      | 3                                           |
| 12–13 Mar.     | 18                                  | 13.2                                      | 5.6                                         |
| 13–14 Mar.     | 13                                  | 10.9                                      | 0                                           |
| 14–15 Mar.     | 13                                  | 11.4                                      | 0                                           |
| 31 Mar.        | 7.5                                 | 13.1                                      | 5.1                                         |
| 6 Apr.         | 3                                   | 13.3                                      | 0.5                                         |
| 10–11 Apr.     | 13                                  | 11.9                                      | 17.5                                        |
| 16 Apr.        | 5                                   | 10.8                                      | 0                                           |
| 16 Apr.        | 10                                  | 10.8                                      | 0                                           |
| 17 Apr.        | 11                                  | 12.3                                      | 0                                           |
| 18–19 Apr.     | 16                                  | 16.2                                      | 58.7                                        |
| 19–20 Apr.     | 11                                  | 15.0                                      | 35.1                                        |
| 20–21 Apr.     | 13                                  | 9.7                                       | 0                                           |
| 22 Apr.        | 9                                   | 8.9                                       | 0                                           |
| 23 Apr.        | 11                                  | 11.4                                      | 0                                           |
| 25–26 Apr.     | 10                                  | 16.2                                      | 0                                           |
| 26 Apr.        | 3                                   | 15.4                                      | 0                                           |
| 27 Apr.        | 3                                   | 15.8                                      | 1.9                                         |
| 27 Apr.        | 4                                   | 12.4                                      | 0                                           |
| 29–30 Apr.     | 15                                  | 14.2                                      | 34.5                                        |
| 1 May          | 8.5                                 | 12.0                                      | 0                                           |
| 6–7 May        | 9.5                                 | 12.2                                      | 0                                           |
| 7 May          | 3                                   | 17.9                                      | 0                                           |
| 8 May          | 7                                   | 14.3                                      | 0                                           |
| 8–9 May        | 16                                  | 17.1                                      | 0                                           |
| 9–10 May       | 11                                  | 16.0                                      | 6.9                                         |
| 10–11 May      | 14                                  | 16.1                                      | 1.0                                         |
| 12 May         | 5                                   | 14.4                                      | 0                                           |
| 15–16 May      | 10                                  | 11.2                                      | 17.3                                        |
| 16–17 May      | 22                                  | 10.4                                      | 81.8                                        |

*On 5 Apr., five pustules were first observed on elderberry plants. By 29 Apr., a total of 304 pustules were observed on 59 plants, and 350 total pustules on 63 plants were observed on 19 May.

Table 3. Mean number of Puccinia sambuci-infected leaves, leaflets, and rust pustules on ‘Bob Gordon’ elderberry plants at three distances from Carex frankii plants on 29 Apr. 2016.6

| Distance from sedge (cm) | No. of pustules/plant | No. of infected leaves/plant | No. of infected leaflets/plant |
|-------------------------|-----------------------|------------------------------|--------------------------------|
| 23                      | 7.6 a                 | 3.3 a                        | 5.8 a                          |
| 59                      | 1.7 b                 | 1.7 b                        | 2.5 b                          |
| 95                      | 2.8 ab                | 2.8 ab                       | 3.4 ab                         |

*Values represent 24 replications of each spacing from sedge. Means followed by different letters in columns are significantly different (P ≤ 0.05).

Table 4. Fruit characteristics, berry puree composition, and shoot dry weight of ‘Bob Gordon’ elderberry plants with or without Puccinia sambuci rust infection in 2016.6

| Treatment | Fruit yield/plant (g) | Berry wt (mg) | Total soluble solids (°Brix) | Titratable acidity (g/100 mL) | Shoot dry wt (g) |
|-----------|-----------------------|---------------|-----------------------------|-----------------------------|------------------|
| Rust-infected | 84 b                  | 116 b         | 10.6 a                     | 0.68 a                      | 49 a             |
| Uninfected | 121 a                 | 127 a         | 11.2 a                     | 0.48 a                      | 31 b             |

*Pooled data from rust-infected elderberry plants (n = 42) and noninfected plants (n = 12) were subjected to a t test. Means followed by different letters in columns are significantly different (P ≤ 0.05). Means were separated by a Satterthwaite test when variances were unequal or by a pooled test when variances were equal. Rust-infected plants averaged six pustules per plant on 19 May 2016.

Table 5. Fruit characteristics, berry puree composition, leaf loss, and cane dry weight of ‘Wyldewood’ elderberry canes with or without Puccinia sambuci rust infection in 2016.6

| Treatment | Fruit yield/cane (g) | Berry wt (mg) | Total soluble solids (°Brix) | Titratable acidity (g/100 mL) | Leaf loss (no.) | Cane dry wt (g) |
|-----------|---------------------|---------------|-----------------------------|-----------------------------|----------------|----------------|
| Rust-infected | 47 b                | 43 a          | 10.9 a                     | 0.68 b                      | 19 a           | 58 a          |
| Uninfected | 88 a                | 41 a          | 11.7 a                     | 0.78 a                      | 10 b           | 86 a          |

*Pooled data from rust-infected elderberry plants (n = 33) and noninfected plants (n = 11) were subjected to a t test. Means followed by different letters in columns are significantly different (P ≤ 0.05). Means were separated by a Satterthwaite test when variances were unequal or by a pooled test when variances were equal. Rust-infected plants averaged 137 pustules/cane.
statistical differences among rust-infected and control canes may not have been detected during this experiment. However, vegetative growth (evaluated by dry weight) was reduced when ‘Bob Gordon’ elderberry plants averaged 690 pustules/plant at the end of the first growing season. Plant dry weight of these elderberry plants was also adversely affected during the successive growing season without P. sambuci reinfection (Table 1).

All levels of P. sambuci infection in these experiments resulted in loss of fruit yield. For young potted ‘Bob Gordon’ plants averaging 6 and 690 pustules/plant, there were 23% and 31% reductions in fruit yield, respectively (Tables 1 and 4). Three-year-old ‘Wyldewood’ plants, with an average of 137 pustules/cane, had 47% less fruit weight than controls at harvest (Table 5). The greater yield loss from rust-infected ‘Wyldewood’ plants may be attributed to cultivar differences, plant age, or the unrestricted root growth in the field as compared with younger container-grown plants. Using a typical planting density (2688 plants/ha), berry yields reported herein, and a selling price of $2.27/kg of fruit (M. Warming, personal communication), the estimated loss of income due to rust infection is $225 to $256/ha for ‘Bob Gordon’ and $1250/ha for ‘Wyldewood’ (assuming five canes per plant with similar berry weight). These estimated losses are likely conservative, as fruit yields for field-grown ‘Bob Gordon’ plants in Oregon have exceeded 5 kg/plant (Finn et al., 2008).

Soluble solids contents of berry puree from elderberry plants were not statistically different in any of the experiments. Due to the naturally low soluble solids content of elderberry fruit, sweeteners are added during processing to enhance the palatability of value-added products (Warming et al., 2016). P. sambuci infection also did not affect the pH, but the titratable acidity of the berry puree was lower than that of control samples when fruit was harvested from elderberry plants averaging 690 rust pustules/plant or 137 pustules/cane. The lower titratable acidity of puree resulting from P. sambuci infection would alter the sugar–acid balance and may adversely affect the flavor of processed elderberry juice or wine products compared with juice products from uninfected fruit.

In conclusion, P. sambuci infection of elderberry plants occurred during early spring when sedge plants were in close proximity. Conditions favorable for possible infection were 9 to 18 °C, ≥3 h continuous leaf wetness, and ≥85% relative humidity. Low infection rates of P. sambuci (six pustules per plant) did not adversely affect vegetative growth, even on young elderberry plants, whereas higher infection levels (137/pustules/cane) resulted in premature leaf loss. In contrast, low infection levels of rust resulted in fruit yield loss. Because of the potential for fruit yield loss on elderberry plants at relatively low infection levels, control of P. sambuci with fungicides may be warranted. Additionally, strategies that eliminate the alternate host or suppress its spread by underground rhizomes may be useful for reducing the leaf area on which rust inoculum resides, thereby limiting the potential for elderberry plant infection.

### Literature Cited

Afshan, N.S. and A.N. Khalid. 2009. New records of Puccinia and Pucciniastrium from Pakistan. Mycotaxon 108:137-146.

Agrios, G.N. 2005. Plant pathology. 5th ed. Elsevier Academic Press, Burlington, MA.

Arthur, J.C. 1921. Memoranda and index of cultures of Uredineae. 1899-1917. Mycologia 13: 230-262.

Arthur, J.C. 1962. Manual of the rusts in the United States and Canada. Hafner Publishing, New York.

Beraha, L., M.B. Linn, and H.W. Anderson. 1960. Development of the asparagus rust pathogen in relation to temperature and moisture. Plant Dis. Rptr. 44:82–86.

Bolley, H.L. 1889. The heteroecismal Puccinia. Amer. Monthly Microsc. J. 8:169–180.

Byers, P.L. and A.L. Thomas. 2011. ‘Bob Gordon’ elderberry. J. Amer. Pomol. Soc. 65:52–55.

Byers, P.L., A.L. Thomas, and M. Millican. 2010. ‘Wyldewood’ elderberry. HortScience 45:312–313.

Charlebois, D., P.L. Byers, C.E. Finn, and A.L. Thomas. 2010. Elderberry: Botany, horticulture, potential. Hort. Rev. 37:213–280.

Finn, C.E., A.L. Thomas, P.L. Byers, and S. Serçe. 2008. Evaluation of American (Sambucus canadensis) and European (S. nigra) elderberry genotypes grown in diverse environments and implications for cultivar development. HortScience 43:1385–1391.

Kellerman, W.A. 1904. Uredineous infection experiments in 1904. J. Mycol. 11:26–33.

Littlefield, L.J. and M.C. Heath. 1979. Ultrastructure of rust fungi. Academic Press, New York.

Mims, C.W. 1981. SEM of ascospore formation in Puccinia sambuci. Scan. Electron Microsc. 3:299–303.

Mohebalian, P.M., M.M. Cernusca, and F.X. Aguilar. 2012. Discovering niche markets for elderberry juice in the United States. HortTechnology 22:556-566.

Mohlenbrock, R.H. 1999. The illustrated flora of Illinois. Sedges: Carex. Southern Illinois Univ. Press, Carbondale, IL.

Petersen, R. 1974. The rust fungus life cycle. Bot. Rev. 40:453–513.

Saccardo, P.A. 1891. Sylloge fungorum omnium hucusque cognitorum. R. Friedlander and Sohn, Berlin.

Warming, M., M. Kwasniewski, J. Elmore, A. Thomas, and K. Adhikari. 2016. Sensory attributes of juice from North American-grown elderberry cultivars. HortScience 51:1561–1565.

Warming, M.R. 2012. Elderberry rust. Univ. Missouri Extension, 20 Oct. 2018. <https://extension2.missouri.edu/ipm1036>.

Zomlefer, W.B. 1994. Guide to flowering plant families. Univ. North Carolina Press, Chapel Hill, NC.