Basic mechanisms of fertilization and parthenogenesis in pigs

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Fertilization of the egg, or oocyte, initiates the entire developmental process, but while the mechanism by which the spermatozoa triggers the oocyte to resume meiosis has been studied extensively, conclusions about this process are still elusive. Some workers have suggested that a molecule on the surface of the spermatozoon may interact with a receptor on the plasma membrane of the oocyte, thereby triggering the oocyte to resume meiosis. Other workers have focused on a factor or factors located in the cytoplasm of the spermatozoon that is deposited into the cytoplasm of the oocyte. A hallmark response to fertilization in mammals is an increase in the cytoplasmic concentration of free Ca\(^{2+}\) in the oocyte. Many additional studies have focused on treatments that will induce the oocyte to resume meiosis without being fertilized. The process of resumption of meiosis without a spermatozoon is generally referred to as activation or parthenogenesis. Activation of the oocyte is very important for a number of oocyte- or embryo-related technologies including intracytoplasmic sperm injection (ICSI) and cloning by nuclear transfer. This review will focus on what is known about fertilization and methods to mimic this process, with an emphasis on pigs.

Introduction

Pig oocytes are arrested at metaphase II of meiosis when ovulated. Fertilization induces the resumption of meiosis, or activates the oocyte, resulting in an oocyte with a single pronucleus and two polar bodies (Fig. 1). Although the downstream events of fertilization have been well characterized, there is still debate as to what triggers activation. The two competing theories are: (i) that sperm binding to the plasma membrane triggers oocyte activation; and (ii) that a factor is deposited into the cytoplasm of the oocyte at sperm–oocyte fusion and that this factor triggers oocyte activation. Proponents of either of these theories admit that complete oocyte activation may be triggered by a combination of these events. A complete understanding of the process of fertilization is needed because failure of adequate fertilization would be disastrous for the reproductive process. In addition, many oocyte- or embryo-related technologies rely on adequate activation of the oocyte. These technologies include in vitro fertilization (IVF), intracytoplasmic sperm injection (ICSI) and nuclear transfer. Many reviews have been written on the mechanisms of oocyte activation in other species such as mice and sea urchins (Swann and Lai, 1997; Macháty and Prather, 1998; Macháty et al., 1999a; Parrington, 2001); thus, the focus of the present review will be on pig oocytes.

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Activation by fertilization

As stated above, one of the hallmarks of fertilization is the release of intracellular Ca\(^{2+}\). A series of Ca\(^{2+}\) bursts or oscillations generally follows this first Ca\(^{2+}\) transient and continues until the time the pronuclei form. The amplitude and frequency of the Ca\(^{2+}\) bursts are generally consistent within an oocyte, but do vary between oocytes. Although the Ca\(^{2+}\) transients are the main feature of fertilization, there are many other cellular and subcellular events that participate in this process. Some of the following features are species-specific; however, for context they will all be presented with a later focus on pigs.

The events of activation have been broken down into two classes: early and late events. Early events include: a sodium influx (Dale *et al.*, 1978), hyperpolarization of the membrane (Miyazaki and Igusa, 1982) and Ca\(^{2+}\) entry through voltage-gated Ca\(^{2+}\) channels (Chambers, 1989), followed by an explosive release of Ca\(^{2+}\) starting at the point of sperm–oocyte fusion. These Ca\(^{2+}\) releases may be mediated by inositol triphosphate or ryanodine receptors on the endoplasmic reticulum. The first burst of Ca\(^{2+}\) also causes exocytosis of the cortical granules. It is thought that the contents of the cortical granules modify the zona pellucida such that additional spermatozoa cannot bind to or penetrate the zona pellucida. In mammals, there are a series of intracellular free Ca\(^{2+}\) oscillations after fertilization. The oscillations are coupled with a Ca\(^{2+}\) influx and, in pigs, this influx may be mediated by a *trp* gene homologue (Macháty *et al.*, this supplement). Late events include an increase in intracellular pH, polyadenylation and translation of pre-existing mRNAs, resulting in an increase in protein synthesis as well as a change in the quality of the proteins in the oocyte, a decrease in the activity of maturation-promoting factor (MPF), cyostatic factor, mitogen-activated protein kinase (MAPK), myosin light chain kinase (MLCK) and glutathione concentration. If these early and late events occur, then the oocyte will develop pronuclei and begin the developmental programme.

Parthenogenetic activation

As stated earlier, many manipulations performed on oocytes bypass the normal fertilization process. The two most prominent procedures include ICSI and nuclear transfer. Thus, it is
imperative that we understand the normal responses to fertilization so that we can attempt to mimic them and activate the oocytes parthenogenetically for ICSI and nuclear transfer. The Ca$^{2+}$-dependent and -independent methods of parthenogenetic activation, combination treatments, as well as specific applications to ICSI and nuclear transfer, will be discussed.

**Calcium-dependent activation**

Early studies in mouse oocytes showed that stimulating protein kinase C with phorbol esters induced a series of Ca$^{2+}$ oscillations, and that this was followed by pronuclear formation and, in some cases, limited development. There are numerous ways to induce an intracellular Ca$^{2+}$ transient in pig oocytes, including injection of Ca$^{2+}$ (Macháty et al., 1996), electrically porating the plasma membrane in the presence of extracellular Ca$^{2+}$ (Prather et al., 1989; Sun et al., 1992), treatment with ionophores (Wang et al., 1998a,b, 1999), thimerosal (Macháty et al., 1997a) or ethanol (Didion et al., 1990), stimulation of guanine nucleotide binding proteins (Macháty et al., 1997b) and injection of inositol triphosphate, cyclic-ADP-ribose (Macháty et al., 1997c) or a crude sperm extract (Macháty et al., 2000). Each of these treatments attacks a different pathway or point on a pathway that results in a Ca$^{2+}$ transient.

These observations illustrate why the data on what activates an oocyte at fertilization are so difficult to interpret: many different pathways have the capacity to activate the egg (even nitric oxide has been implicated; Kuo et al., 2000). This is further illustrated by studies in which the mRNA encoding the muscarinic M1 receptor was injected into the cytoplasm of unfertilized oocytes (Macháty et al., 1997b; Kim et al., 1998). After time for translation and insertion into the plasma membrane, the oocytes were treated with acetylcholine. Acetylcholine activated the guanine protein-coupled receptor, which probably activated phospholipase C. Phospholipase C then probably acted on phosphatidyl inositol bisphosphate to yield inositol triphosphate and diacylglycerol. The inositol triphosphate bound to inositol triphosphate receptors located on the endoplasmic reticulum and released Ca$^{2+}$ into the cytoplasm. This Ca$^{2+}$ release caused release of the cortical granules into the extracellular space, a decrease in histone H1 kinase activity, a shift in protein profiles consistent with fertilization and pronuclear formation (Macháty et al., 1997b; Kim et al., 1998). Furthermore, oocytes activated in this manner could develop to the blastocyst stage and these blastocysts had nucleoli and mitochondria that had morphologies consistent with normal blastocyst stage embryos (Kim et al., 1998).

Although a simple electrical pulse in the presence of Ca$^{2+}$ will generally induce an oocyte to begin the developmental process, the pattern of events associated with normal fertilization is not always followed. Normally, histone H1 kinase activity decreases after fertilization and stays low until the first cleavage; however, with electrical activation H1 kinase activity initially decreases, but increases later in the first cell cycle (Leal and Liu, 1998). Although a single electrical pulse provides a single Ca$^{2+}$ transient that is sufficient to induce development to the blastocyst stage, it is thought that the quality (amplitude and frequency) of the Ca$^{2+}$ transient or transients is very important and may determine the degree of development after stimulation (Ozi I, 1990).

As stated earlier, there is an increase in intracellular pH at fertilization in many lower species such as the sea urchin. However, this has not been demonstrated at fertilization in mammals. Recently, Ruddock et al. (2000a) observed an increase in intracellular pH when pig oocytes were treated with 7% (v/v) ethanol or A23187 and there appears to be more than one mechanism regulating the change in pH. Similar observations were made in murine and bovine oocytes (Ruddock et al., 2000b). However, the increase in intracellular pH in pigs is independent of intracellular Ca$^{2+}$ concentration (Ruddock et al., 2001).
Calcium-independent activation

Although intracellular Ca\(^{2+}\) is released at fertilization, this is not a requisite for activation of the oocyte. Indeed, the early studies on treatment with phorbol esters were not consistent with phosphorylation status; cells in metaphase have proteins that are already phosphorylated and need to be dephosphorylated to push the cell into the next interphase (histones, nuclear lamins). Thus, studies that have evaluated treatments that would decrease the phosphorylation status or inhibit phosphorylation would appear to be more relevant. Inhibition of MAPK, MPF and MLCK (Green et al., 1999) has resulted in pronuclear formation of pig oocytes. In contrast, stimulation of protein kinase C (Sun et al., 1997) resulted in cortical granule release, but not pronuclear formation, whereas stimulation of tyrosine kinase activity resulted in both cortical granule release and pronuclear formation (Kim et al., 1999).

In addition to the treatments that inhibit kinase activity, other treatments can result in oocyte activation. One of the more popular techniques is to inhibit protein synthesis. As cyclin B is being made continually and this manufacture is necessary for maintaining high concentrations of MPF, inhibiting the production of cyclin B will cause MPF activity to decrease. This lower MPF activity results in the resumption of meiosis by the oocyte and formation of a pronucleus. This response may be species-specific as Nussbaum and Prather (1995) found that cycloheximide treatment alone did not result in activation, but when used in combination with an electrical pulse, it enhanced the rates of pig oocyte development; in other species, activation can be induced with the cycloheximide treatment only (Macháty and Prather, 1998).

In relation to altering MPF activity, the cyclin-dependent kinase inhibitor butyrolactone I can result in cleavage rates of 40%. However, when combined with an electrical pulse, butyrolactone I can result in 59% of the treated oocytes progressing to the blastocyst stage (Dinnyes et al., 1999).

Combination treatments

As stated above, attacking more than one point on the pathway can result in improved rates of pronuclear formation, as well as improved rates of development. A combination of an electrical pulse and cycloheximide is superior to either treatment alone in promoting pronuclear formation (Nussbaum and Prather, 1995). Similarly, Grocholova et al. (1997) used a combination of A23187 and cycloheximide to improve development of parthenogenetic oocytes. For the thimerosal treatment to result in pronuclear formation the action on the sulphhydryl groups must be reversed or the spindle will be damaged irreversibly. To facilitate that reversal, thimerosal followed by dithiothreitol treatment results in Ca\(^{2+}\) release and 42% of oocytes developing to the compact morula or blastocyst stage (Macháty et al., 1997b). Thimerosal causes Ca\(^{2+}\) oscillations and no development if not reversed with dithiothreitol (Macháty et al., 1999b).

Another treatment that is often used when development of parthenogenotes is desired is cytochalasin B. It is not known whether cytochalasin B affects any of the Ca\(^{2+}\) pathways, but it does depolymerize microfilaments and, thus, prevents polar body emission. Retaining the second polar body should result in a doubling of the number of chromosomes in the developing embryo. Cha et al. (1997) increased the percentage of 2n chromosome spreads by treating the activated oocytes with cytochalasin B. Jolliff and Prather (1997) used electrical activation followed by cytochalasin B and obtained fairly normal in vivo development to day 12. Kure-bayashi et al. (2000) found that development could proceed to day 29 in vivo. The lack of further development is probably a result of genomic imprinting (Reik and Walter, 2001).
Although some of these treatments result in cortical granule release they do not prevent subsequent fertilization (Wang et al., 1997, 1998a,b). Thus, there is much to learn about the zona pellucida and methods of inducing hardening.

In conclusion, the most widely used method of activation for nuclear transfer is electrical activation (Prather et al., 1989). Variations to a single pulse of electricity have been added to improve the rates of development (Liu and Moor, 1997).

**Activation for ICSI or nuclear transfer**

A variety of different activation parameters has been used for ICSI and nuclear transfer. In some cases the injection procedure for ICSI activates the oocyte. Martin (2000) produced an ICSI-derived piglet without any activation stimuli to the in vivo-matured oocytes other than injection of a fresh spermatozoon. In contrast, Kolbe and Holtz (2000) used Ca²⁺ ionophore to activate in vivo-matured ICSI oocytes. Lai et al. (in press) obtained a piglet when electrical activation was applied after injection of the spermatozoon; the oocytes did not develop as well without the electrical application.

Oocytes must be enucleated for nuclear transfer. One of the common chemicals used in the enucleation is bisbenzimide. Unfortunately, exposure to this compound has deleterious effects on the development of pig oocytes to the blastocyst stage (Tao et al., 2000). Thus, compounds used for enucleation might limit development of nuclear transfer and parthenogenetic embryos.

In many cases there is a benefit to fusing the donor cell to the recipient oocyte or microinjecting the nucleus into the oocyte, before the oocyte is actually activated. Miyoshi et al. (2000a) found that a delay of 3 h between fusion and activation improved the rate of blastocyst formation. Tao et al. (1999a) used microinjection to transfer the nuclei and attempted to delay the activation up to 4 h after the microinjection while using a Ca²⁺-free medium. The Ca²⁺-free condition was detrimental to development. The rates of development to the blastocyst stage after nuclear transfer have been as low as 3% (Miyoshi et al., 2000b). When presumptive G0 fibroblast cells were transferred to oocytes and electrically activated, only 7% formed blastocysts (Tao et al., 1999b). Betthauser et al. (2000) reported 4-8% blastocysts resulting from nuclear transfers that were activated by ionophore followed by 6-dimethylaminopurine. Recently, Kühholzer et al. (2001) reported almost 20% blastocysts from electrically activated oocytes. In addition, oocytes from sows resulted in higher rates of development to the blastocyst stage compared with oocytes from gilts. Polejaeva et al. (2000) used natural fertilization to achieve normal activation. Nuclear transfer was generally performed as described above with electrical activation. However, the nuclei were transferred to enucleated fertilized oocytes after electrical activation. Thus, the nuclei were transferred to sperm-activated oocytes, which resulted in a number of live born piglets. Onishi et al. (2000) produced live pigs by electrical activation and microinjection of oocytes, but did not report on development to the blastocyst stage. Park et al. (in press) produced 9-16% blastocysts with two electrical pulses to fuse and activate the oocytes, resulting in five nuclear transfer-derived piglets.

**Conclusion**

In conclusion, there are a number of methods that can be used to activate pig oocytes artificially. These methods often attack different pathways that lead to parthenogenesis (Fig. 2). These include Ca²⁺-dependent and -independent processes. As there are so many pathways in oocytes that can be stimulated to trigger the resumption of meiosis, it might be concluded that
Fig. 2. Summary of pathways that may be activated at fertilization or parthenogenesis. Description of the pathways is included throughout the text. PIP₂: phosphatidyl inositol bisphosphate; DAG: diacylglycerol; cADPr: cyclic ADP ribose; IP₃: inositol trisphosphate; RYR: ryanodine receptor; IP₃R: inositol trisphosphate receptor; PN: pronuclear; PKC: protein kinase C; ER: endoplasmic reticulum; CG: cortical granule; RTK: receptor tyrosine kinase; PLC: phospholipase C; α, β, γ subunits of a G-protein coupled receptor; p: phosphorylation; PKI: protein kinase inhibitor; RGD: arginine-glycine-aspartic acid peptide.

the oocyte is poised to progress to interphase. Upsetting this apparently delicate balance required for maintenance of the oocyte in metaphase can be accomplished via a variety of methods. Generally, if more than one pathway that results in activation is attacked, greater rates of development to the blastocyst stage can be obtained. Further advancements in the understanding of the basic mechanisms of fertilization will aid in designing strategies to artificially activate pig oocytes for ICSI and nuclear transfer.

References

Betthauser I, Forsberg E, Augustein M et al. (2000) Production of cloned pigs from in vitro systems Nature Biotechnology 18 1055–1059

Cha KS, Kim NH, Lee SM, Baik CS, Lee HT and Chung KS (1997) Effect of cytochalasin B and cycloheximide on the activation rate, chromosome constituent and in vitro development of porcine oocytes following parthenogenetic stimulation Reproduction, Fertility and Development 9 441–446

Chambers EL (1989) Fertilization in voltage-clamped sea urchin eggs. In Mechanisms of Egg Activation pp 1–18 Eds R Nuccitelli, GN Cherr and WH Clark. Plenum Press, New York

Dale B, Defelice LJ and Taglietti V (1978) Membrane noise and conductance increase during single spermatozoon–egg interactions Nature 275 217–219

Didion BA, Martin MJ and Markert CL (1990) Parthenogenetic activation of mouse and pig oocytes matured in vitro. Theriogenology 33 1165–1175

Dimnyes A, Hirao Y and Nagai T (1999) Parthenogenetic activation of porcine oocytes by electric pulse and/or butyrolactone I treatment Cloning 1 209–216

Green KM, Kim JH, Wang WH, Day BN and Prather RS (1999) Effect of myosin light chain kinase, protein kinase A, and protein kinase C inhibition of porcine oocyte activation Biology of Reproduction 61 111–119

Grocholova R, Petr J, Rozinik J and Jilek F (1997) The protein phosphatase inhibitor okadaic acid inhibits exit from metaphase II in parthenogenetically activated pig oocytes Journal of Experimental Zoology 277 49–56

Jolliff WJ and Prather RS (1997) Parthenogenic development of in vitro-matured, in vivo-cultured porcine oocytes beyond blastocyst Biology of Reproduction 56 544–548
Basic mechanisms of oocyte activation in pigs

Kim JH, Macháty Z, Cabot RA, Han YM, Do HJ and Prather RS (1998) Development of pig oocytes activated by stimulation of an exogenous G protein-coupled receptor Biology of Reproduction 59 655–660

Kim JH, Do HJ, Wang WH, Macháty Z, Han YM, Day BN and Prather RS (1999) A protein tyrosine phosphatase inhibitor, sodium orthovanadate, causes parthenogenetic activation of pig oocytes via an increase in protein tyrosine kinase activity Biology of Reproduction 61 900–905

Kolbe T and Holtz W (2000) Birth of a piglet derived from an oocyte fertilized by intracytoplasmic sperm injection (ICSI) Animal Reproduction Science 64 97–102

Kühnholzer B, Hawley RJ, Lai L, Kolber-Simonds D and Prather RS (2001) Clonal lines of transgenic fibroblast cells derived from the same fetus result in different development when used for nuclear transfer in pigs Biology of Reproduction 64 1695–1698

Kuo RC, Baxter GT, Thompson SH, Strickers SA, Patton C, Bonaventura J and Epel D (2000) NO is necessary and sufficient for egg activation at fertilization Nature 406 633–636

Kure-bayashi S, Miyake M, Okada K and Kato S (2000) Histone kinase activity in pig oocytes after repetitive pulsatile electrical stimulation of an exogenous G protein-coupled receptor Biology of Reproduction 63 488–492

Lai L, Sun Q, Wu G, Murphy CN, Kühnholzer B, Park KW, Bonk AJ, Day BN and Prather RS Development of porcine embryos and oosperm after ICSI with liposome transfected or non-transfected sperm into vitro-matured porcine oocytes Zygote 8 1105–1119

Lee CL and Liu L (1998) Differential effects of kinase inhibitor and electrical stimulation on activation and histone H1 kinase activity in pig oocytes Animal Reproduction Science 52 51–61

Liu L and Moor RM (1997) Factors affecting electrical activation of porcine oocytes matured in vitro. Animal Reproduction Science 48 67–80

Macháty Z and Prather RS (1998) Strategies for activating nuclear transfer oocytes Reproduction, Fertility and Development 10 599–613

Macháty Z, Funahashi H, Mayes MA, Day BN and Prather RS (1996) Effects of injecting calcium chloride into in vitro-matured porcine oocytes Biology of Reproduction 54 316–322

Macháty Z, Wang WH, Day BN and Prather RS (1997a) Complete activation of porcine oocytes induced by the sulfhydryl reagent, thimerosal Biology of Reproduction 57 1123–1127

Macháty Z, Mayes MA, Kovacs LG, Balatí PA, Kim JH and Prather RS (1997b) Activation of porcine oocytes via an exogenously introduced rat muscarinic M1 receptor Biology of Reproduction 57 85–91

Macháty Z, Funahashi H, Day BN and Prather RS (1997c) Developmental changes in the intracellular calcium release mechanisms in porcine oocytes Biology of Reproduction 56 921–930

Macháty Z, Rickords LF and Prather RS (1999a) Parthenogenetic activation of porcine oocytes after nuclear transfer Cloning 1 101–109

Macháty Z, Wang WH, Day BN and Prather RS (1999b) Calcium release and subsequent development induced by modification of sulfhydryl groups in porcine oocytes Biology of Reproduction 60 1384–1391

Macháty Z, Bonk AJ, Kühnholzer B and Prather RS (2000) Porcine oocyte activation induced by a cytosolic sperm factor Molecular Reproduction and Development 57 290–295

Martin MJ (2000) Development of in vitro-matured porcine oocytes following intracytoplasmic sperm injection Biology of Reproduction 63 109–112

Miyazaki S and Igusa Y (1982) Ca-mediated activation of a K current at fertilization of golden hamster eggs Proceedings National Academy of Sciences USA 79 931–935

Miyoshi K, Sack K and Sato E (2000a) Improvement in development of porcine embryos reconstructed with cells from blastocyst-derived cell lines and enucleated oocytes by optimization of reconstruction methods Cloning 2 175–184

Miyoshi K, Taguchi Y, Sendai Y, Hoshi H and Sato E (2000b) Establishment of a porcine cell line from in vitro-produced blastocysts and transfer of the cells into enucleated oocytes Biology of Reproduction 62 1640–1646

Nussbaum DJ and Prather RS (1995) Differential effects of protein synthesis inhibitors on porcine oocyte activation Molecular Reproduction and Development 41 70–75

Onishi A, lwamoto M, Akita T, Mikawa S, Takeda K, Awata T, Hanada H and Perry AC (2000) Pig cloning by microinjection of fetal fibroblast nuclei Science 289 1188–1190

Ozil IP (1990) The parthenogenetic development of rabbit oocytes after repetitive pulsatile electrical stimulation Development 109 117–127

Park K-W, Cheong H-T, Lai L et al. (2001) Production of nuclear transfer-derived swine that express the enhanced green fluorescent protein Animal Biotechnology (in press)

Parrington J (2001) Does a soluble sperm factor trigger calcium release in the egg at fertilization? Journal of Andrology 22 1–11

Polejaeva IA, Chen SH, Vaught TD et al. (2000) Cloned pigs produced by nuclear transfer from adult somatic cells Nature 407 29–30

Prather RS, Simms MM and First NL (1989) Nuclear transplantation in early pig embryos Biology of Reproduction 41 414–418

Reik W and Walter J (2001) Genomic imprinting: parental influence on the genome Nature Genetics 2 21–32

Ruddock NT, Macháty Z, Milanick M and Prather RS (2000a) Mechanism of intracellular pH increase during parthenogenetic activation of in vitro matured porcine oocytes Biology of Reproduction 63 488–492

Ruddock NT, Macháty Z and Prather RS (2000b) Intracellular pH increases accompany parthenogenetic activation of porcine, bovine and mouse oocytes Reproduction, Fertility and Development 12 201–207

Ruddock NT, Macháty Z, Cabot R and Prather RS (2001) Porcine oocyte activation: the differing roles of calcium and pH Molecular Reproduction and Development 59 227–234

Sun FZ, Hoyland J, Huang X, Mason W and Moor RM
R. S. Prather

(1992) A comparison of intracellular changes in porcine eggs after fertilization and electroactivation Development 115 947–956

Sun QY, Wang WH, Hosoe M, Taniguchi T, Chen DY, Shioya Y (1997) Activation of protein kinase C induces cortical granule exocytosis in a calcium-independent manner, but not resumption of cell cycle in porcine oocytes Development, Growth and Differentiation 39 523–529

Swann K and Lai FA (1997) A novel signaling mechanism for generating Ca^{2+} oscillations at fertilization in mammals Bioessays 19 371–378

Tao T, Macháty Z, Boquest AC, Day BN and Prather RS (1999a) Development of pig embryos reconstructed by microinjection of cultured fetal fibroblast cells into in vitro matured oocytes Animal Reproduction Science 56 133–141

Tao T, Boquest AC, Macháty Z, Petersen AL, Day BN and Prather RS (1999b) Development of pig embryos by nuclear transfer of cultured fibroblast cells Cloning 1 55–62

Tao T, Macháty Z, Abeydeera LR, Day BN and Prather RS (2000) Optimisation of porcine oocyte activation following nuclear transfer Zygote 8 69–77

Wang WH, Sun QY, Hosoe M, Shioya Y and Day BN (1997) Quantified analysis of cortical granule distribution and exocytosis of porcine oocytes during meiotic maturation and activation Biology of Reproduction 56 1376–1382

Wang WH, Abeydeera LR, Prather RS and Day BN (1998a) Functional analysis of activation of porcine oocytes by spermatozoa, calcium ionophore and electric pulse Molecular Reproduction and Development 51 346–353

Wang WH, Macháty Z, Abeydeera LR, Prather RS and Day BN (1998b) Parthenogenetic activation of pig oocytes with calcium ionophore and the block to sperm penetration after activation Biology of Reproduction 58 1357–1366

Wang WH, Macháty Z, Ruddock N, Abeydeera LR, Boquest AC, Prather RS and Day BN (1999) Activation of porcine oocytes with calcium ionophore: effects of extracellular calcium Molecular Reproduction and Development 53 99–107