Identification of immune-related proteins of *Dreissena polymorpha* hemocytes and plasma involved in host-microbe interactions by differential proteomics

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Biological responses of zebra mussel *Dreissena polymorpha* are investigated to assess the impact of contaminants on aquatic organisms and ecosystems. In addition to concentrate chemical contaminants in their tissues, zebra mussels accumulate several microorganisms such as viruses, protozoa and bacteria. In order to understand the molecular mechanisms involved in the defence against microorganisms this study aims at identifying immune proteins from *D. polymorpha* hemolymph involved in defence against protozoa and viruses. For this purpose, hemolymph were exposed *ex vivo* to *Cryptosporidium parvum* and RNA poly I:C. Differential proteomics on both hemocytes and plasma revealed immune proteins modulated under exposures. Different patterns of response were observed after *C. parvum* and RNA poly I:C exposures. The number of modulated proteins per hemolymphatic compartments suggest that *C. parvum* is managed in cells while RNA poly I:C is managed in plasma after 4 h exposure. BLAST annotation and GO terms enrichment analysis revealed further characteristics of immune mechanisms. Results showed that many proteins involved in the recognition and destruction of microorganisms were modulated in both exposure conditions, while proteins related to phagocytosis and apoptosis were exclusively modulated by *C. parvum*. This differential proteomic analysis highlights in zebra mussels modulated proteins involved in the response to microorganisms, which reflect a broad range of immune mechanisms such as recognition, internalization and destruction of microorganisms. This study paves the way for the identification of new markers of immune processes that can be used to assess the impact of both chemical and biological contaminations on the health status of aquatic organisms.

The Zebra mussel, *Dreissena polymorpha*, is a relevant sentinel species used in aquatic ecotoxicology, due to its ability to accumulate and concentrate the chemical and biological contaminants1. While most ecotoxicological studies focus on the impact of chemical contamination on aquatic organisms, few studies assess the impact of the biological contamination. However, recent studies have shown the ability of zebra mussels to bioaccumulate and concentrate several microorganisms. Gu and Mitchell2 considered the bivalve *D. polymorpha* as a reservoir of opportunistic pathogenic microorganisms to aquatic organisms and humans. Bioaccumulation of several protozoa pathogenic to humans in the zebra mussel tissues have been evidenced by numerous studies3. Furthermore, Mezzanotte et al.4 have demonstrated the ability of *D. polymorpha* to remove enteric viruses and the bacteria *Escherichia coli* found in municipal treated effluent. While literature proved the accumulation of several microorganisms in the tissues of zebra mussel, their interactions with the physiology of *D. polymorpha* remain to be clarified.

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Probably more than other biological processes, the innate immune defence of bivalves is directly involved in the interaction with microbes. Indeed, the immune system acts as the first line of defence against microorganisms, involving physical barriers, phagocytic cells and a variety of immune effectors. Innate immune system process can be summarized in three main steps: (i) the recognition of molecular motifs associated with microorganisms (MAMPs, Microbe-associated molecular patterns) or endogenous molecules secreted by damaged tissues (DAMPs, Damage-associated molecular patterns) by soluble compounds and cellular receptors, (ii) the activation of different signalling pathways, (iii) the production of molecular effectors involved in host defence and cellular defence responses. In bivalves, the immune defence is mainly undertaken by their circulating fluid named hemolymph, which includes circulating immune cells, the hemocytes, and an acellular fraction, the plasma. A cross-talk between hemocytes and plasma appears crucial for an effective immune response. In addition to their hemolymph, which includes circulating immune cells, the hemocytes, and an acellular fraction, the plasma. Juhel observed a decrease in total hemocytes. For of freshwater bivalves and microorganisms are rarely investigated despite their interest in ecotoxicological studies. Indeed, few studies investigated the biological interactions with non-pathogenic microbes which could be found in water courses. Furthermore, in contrast with marine bivalves, interactions between the immune system of freshwater bivalves and microorganisms are rarely investigated in ecotoxicological studies. For D. polymorpha, information regarding modulation of the immune system by biological contamination is relatively scarce. However, the limited literatures reported that microorganisms that interact with D. polymorpha modulate cellular mediated immune responses of hemocytes. Juhel et al. observed a decrease in total hemocytes count, a modulation of phagocytic rate of hemocytes and an increase of the concentration of lysozymes in mussels fed with a microcystin-producing cyanobacterium. Recently, the interaction between three human protozoan parasites and D. polymorpha immunity was investigated at the cellular level. Authors revealed the ability of protozoa to induce cytotoxic effects and modulate phagocytosis of D. polymorpha hemocytes. Moreover, involvement of apoptosis during host-parasite interactions have been evidenced in hemocytes of zebra mussels. While the interactions between the immune system of D. polymorpha and microorganisms are investigated at the cellular level, the immune molecular mechanisms involved in host-microbe interactions are still not documented. Characterization of these interactions at the molecular level would expand our knowledge on microbe-bivalve interactions and more specifically on the innate immune system of D. polymorpha.

Next generation shotgun proteomics appears to be a useful methodology to discover and prioritize new biomarkers of contamination, while targeted proteomics allows the validation of these biomarkers for diagnostic purpose. Recently, a proteogenomic analysis was performed on the hemocytes and plasma of D. polymorpha, resulting in the identification of more than 3,000 proteins. In order to get further insights into the immune proteome of D. polymorpha, many proteins potentially involved in the recognition, internalization and destruction of microorganisms were pinpointed in both hemocytes and plasma fractions. Considering the high number of immune proteins observed in plasma, the need to consider both intracellular and extracellular fractions to investigate the immune defence occurring in the hemolymph compartments was pointed out. Nevertheless, this proteogenomic analysis was performed on zebra mussels that were physiologically acclimated to laboratory conditions and thus only gave a cartography of potential immune-related proteins without exploring their interactions with microorganisms.

The present study aims to identify proteins involved in the interaction between microorganisms and the immune system of D. polymorpha. For this purpose, ex vivo challenges were performed to decipher the immune system response of D. polymorpha upon protozoan and virus challenges. D. polymorpha hemolymph was exposed either to a protozoan, Cryptosporidium parvum, or a synthetic double stranded RNA (dsRNA) known to induce a non-specific antiviral immune responses in bivalves. A comparative proteomic analysis was then performed on both hemocytes and plasma fractions to reveal immune-related proteins modulated by C. parvum and RNA polyinosinic-polyricydlyc acid (poly I:C) exposures.

Results and Discussion

Nano LC-MS/MS strategy for the discovery of modulated proteins in hemocytes and plasma of D. polymorpha. A differential proteomic analysis was conducted on hemocytes and plasma from the hemolymph challenged with immune modulators in order to identified proteins involved in the immune defence of D. polymorpha. Our strategy was to start from a unique pool of hemolymph to perform the three exposure conditions (Fig. 1). In this way, biological variability is minimized as much as possible and changes in protein abundance are more likely the results of exposures to C. parvum and RNA poly I:C. In total, 30 runs of LC-MS/MS analysis were performed, resulting in the recording of 1,667,920 MS/MS spectra from the 15 hemocyte samples and 1,543,356 MS/MS spectra from the 15 plasma samples. MS/MS spectra were assigned using the protein database “Dreissene_cascade” which contains 54,097 putative polypeptide sequences that have been prioritized with a first round of search with experimental MS/MS datasets as described in Leprêtre et al. From the whole dataset comprising 3,211,276 MS/MS spectra, a total of 1,198,994 MS/MS spectra (36%) were interpreted into peptide sequences, leading to the discovery of 33,409 unique peptides and detection of 3,784 proteins from all samples (hemocytes and plasma) (Fig. 1). Results of comparative proteomics reflect the proteome changes of D. polymorpha after four hours of exposure. A higher number of proteins were identified with at least 2 peptides in hemocytes (2,740) compared to plasma (2,076). From them, 1,032 proteins were common between plasma and
hemocytes, 1,708 proteins were exclusive to hemocytes and 1,044 to plasma (Fig. 1). Regarding BLAST annotation, the sequence similarity search against NCBInr and Swissprot Databases resulted in the functional annotation of 3,656 proteins, letting about 3% of proteins without BLAST annotation. Once again, proteogenomic analysis has shown its efficiency in the quick identification of proteins from non-model species without genome sequence and annotation since more than 3,500 proteins could be identified in the hemolymph of *D. polymorpha*.

As described in Leprêtre *et al.*15, a greater number of proteins were observed in hemocytes than in plasma, as expected as the dynamic ranges of protein abundance in cells and plasma are quite different in all animals. Furthermore, several proteins were found in both compartments, suggesting that they are secreted by hemocytes in the extracellular space. The enrichment of some proteins in the latter compartment indicates their longer half-life in this compartment or their origin that could differ.

The patternLab’s T-Fold module was used to pinpoint significantly modulated proteins in hemocytes or plasma from the hemolymph samples exposed to *C. parvum* or RNA poly I:C in comparison to the control condition. As five replicates per condition have been performed and analyzed with a robust methodology, the differential proteomics results are relevant. A total of 643 proteins were significantly modulated in the hemolymph exposed to *C. parvum* and 328 proteins were also significantly modulated in the hemolymph exposed to RNA poly I:C (Fig. 2). The abundance of several proteins was found modulated in both hemocytes and plasma exposed to biological stressors. Different patterns of modulation were observed between the hemolymph challenged with *C. parvum* and RNA poly I:C (Fig. 2). While in *C. parvum* exposure, a greater number of proteins were modulated in terms of abundance in hemocytes compared to plasma, the opposite was observed in the hemolymph exposed to RNA poly I:C, with a higher number of modulated proteins observed in plasma compared to hemocytes. Such results may suggest, that after four hours of exposure, *C. parvum* is managed in hemocytes while RNA poly I:C seems to be managed in the extracellular space. Several studies identified significant changes in the proteome of mollusks challenged with a protozoan. As our results, a higher number of proteins were modulated in hemocytes compared to plasma in the octopus, *Octopus vulgaris*, infected with the protozoan parasite *Aggregata octopiana* and the Manila clam, *Ruditapes philippinarum*, challenged with *Perkinsus olseni* using a 2D-gel electrophoresis methodology18,19. The internalization of *C. parvum* by hemocytes have been reported in the freshwater benthic clam *Corbicula fluminea* during *in vitro* and *in vivo* exposures20. Using flow cytometry, labeled *C. parvum* were

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**Figure 1.** Experimental design. Schematic workflow of the protocol followed for the identification and quantification of proteins in the hemolymph of *Dreissena polymorpha*. 
endocytosed by zebra mussel hemocytes exposed 4 h in ex vivo condition. In this case, the management of protozoa by immune cells involves a dynamic modulation of intracellular proteins to internalize and eliminate protozoa. For viral infection, molecular responses were till now mainly investigated in immune cells using transcriptomic analysis. However, the immune responses against viruses occurring in extracellular fraction is gaining attention. For example, a comparative proteomic analysis was performed in the plasma of the zebra fish to understand spring viremia of carp virus pathogenesis. Infections of zebra fish resulted in the modulation of more than 150 proteins and provided the identification of multiple viral-infection biomarkers. In invertebrates, only Tao et al. performed a proteomic analysis on the plasma of shrimp infected with the white spot syndrome virus (WSSV). Among the 486 proteins identified in the plasma, 59 proteins were modulated upon WSSV infection. Both studies pointed out the importance of protein responses occurring in the extracellular fluid during viral infections. Furthermore multifunctional plasma proteins from marine mollusks have been identified to have broad-spectrum antiviral activity against mammalian viruses.

The Fig. 2 shows overlapped and specific proteins modulated in hemocytes and plasma exposed to C. parvum and RNA poly I:C. Only two proteins, a peroxiredoxin and a cysteine protease, were modulated in all conditions. In hemocytes, 418 proteins were exclusively modulated in C. parvum exposure and 55 proteins were exclusively modulated in the RNA poly I:C condition. In addition, 72 proteins were modulated in both treatments, which represent about 50% of the proteins modulated in hemocytes challenge with RNA poly I:C. In plasma, 106 proteins were modulated exclusively in the RNA poly I:C treatment against 38 proteins exclusively modulated in the C. parvum challenge. More than 50% of plasmatic proteins modulated by C. parvum were also modulated in the plasma of RNA poly I:C exposure. Overall, these results support that common and distinct biological processes are taking place between the hemolymph exposed to C. parvum and RNA poly I:C. While bivalves lack the mechanisms conferring adaptive immunity, recent research demonstrates that they possess a certain level of immune specificity depending on the structural composition and biological properties of microorganisms. In our study, C. parvum was used in its oocyst form and RNA poly I:C is a synthetic dsRNA which mimic virus. Their difference in term of molecular structure probably requires an appropriate immune response for their elimination in D. polymorpha hemolymph, which could explain the different pattern of modulation observed.

Identification of immune-related proteins involved in host-microbial interactions. A Gene set enrichment analysis (GSEA) was performed on significantly modulated proteins in the hemolymph exposed to C. parvum and RNA poly I:C with the aim to reveal biological processes modulated under these two exposure conditions (Table 1). Results showed that under both exposure conditions, several immune-related biological processes were identified, in particular those related to the cytoskeletal reorganization during endocytosis. In addition, BLAST annotation of significantly modulated proteins provided the identification of proteins involved in the defence against C. parvum and RNA poly I:C. Although these proteins can take part in multiple biological processes, they were categorized into one of the following immune functional groups: 1) pattern recognition receptors (PRRs), 2) immune signaling, 3) internalization of microorganisms, 4) immune effectors, 5) apoptosis or 6) other processes. While many of them were modulated in both exposure conditions (Table 2), some were modulated exclusively under C. parvum (Table 3) or RNA poly I:C challenges (Table 4).

Immune proteins modulated in both exposures. The recognition of non-self molecules is an essential step to trigger an appropriate and an effective immune response. Immune responses are initiated by the recognition of MAMs or DAMPs by PRRS. Signal transduction pathways induced by PRRs result in the activation of gene expression and synthesis of a broad range of molecules involved in immune defence. Exposure of the hemolymph
to *C. parvum* and RNA poly I:C resulted in the modulation of several potential PRRs (Table 2). Among them, several lectins were up-regulated in both exposures. Lectins are made of carbohydrate recognition domains which allow the recognition of non-self-carbohydrate domains located at the surface of microorganisms such as glycoproteins of the viral envelope or glycans of parasites. The cation-independent mannose-6-phosphate receptor (M6PR) is a P-type lectin that was positively regulated in the plasma exposed to contaminants with a fold change (FC) higher than 1.6. In vertebrates, M6PRs are believed to play a critical role in the biogenesis of lysosomes by targeting hydrolytic enzymes to lysosomes. Induction of M6PRs also plays an important role during viral infection, by facilitating the entry of viruses into cells. In invertebrates, few studies have investigated the role of M6PRs in the immunity. Recently, Zhang et al. reported an induction of M6PRs genes in shrimp challenged with a broad range of bacteria species. Authors demonstrated that M6PR functioned as a pattern recognition receptor by binding to peptidoglycan (PGN), lipopolysaccharide (LPS), and lipoteichoic acid (LTA) in addition to their implication in antimicrobial peptides production. While involvement of M6PR in response to protozoa and viruses is still not investigated in bivalves, the up-regulation of this protein observed in plasma exposed to *C. parvum* and RNA poly I:C might suggest its involvement in *D. polymorpha* immunity. Interestingly, a natterin-like protein was down-regulated (FC: −1.7) in hemocytes exposed to *C. parvum* and up-regulated in the plasma exposed to RNA poly I:C (FC: 1.3). Natterin-like protein is one of the most abundant protein of *D. polymorpha* hemolymph and is composed of a jacalin-lectin domain, which may confer an affinity with carbohydrate motifs found in microorganisms. Natterins were first identified as the major toxin of the venomous fish *Thalassophryne nattereri* and were described as an important defence proteins against predators. The implication of natterins in the immune defence of bivalves is still not investigated and its involvement in the recognition of microorganisms is not clearly demonstrated. However, a recent study showed that proteins structurally close to natterins serve as a multipotent pattern recognition receptor in the oyster immune defence. The presence of jacalin-lectin and its modulation pattern during exposures to *C. parvum* and dsRNA may indicate its implication in the *D. polymorpha* immune defence against microbes. Galactose binding lectins (galectins) and a Beta-1,3-glucan binding protein (BGP) were also modulated in the hemolymph exposed to protozoan and dsRNA. As reviewed in Vasta et al., galectins are non-self-recognition receptors involved in both innate and adaptive immune systems. In aquatic mollusks, the role of galectins in the defence against microorganisms is well documented. It appears that galectins work as PRRs and interact with bacteria, protozoa and viruses. In innate immunity, BGP functions as a PRR that recognizes glucans of bacteria cell walls. To our knowledge, only Roux et al. have reported an overexpression of Lipopolysaccharide and B-1,3-Glucan Binding Protein (LBGP) gene in shrimp infected by virus. The authors hypothesized that in addition to their PRR functions, LBGP may play a critical role in viral pathogenesis. Oocyst wall of *Cryptosporidium* is composed of carbohydrates, proteins and lipids but lacks β-1,3-glucan polysaccharides. However, numerous

| Biological process (BPs) | Count | Modulated protein (%) | p-value |
|--------------------------|-------|-----------------------|---------|
| vesicle-mediated transport | 84 | 16.4 | 1.8E-02 |
| cytoskeleton organization | 70 | 13.6 | 1.8E-05 |
| locomotion | 67 | 13.1 | 4.1E-02 |
| small GTPase mediated signal transduction | 26 | 5.1 | 2.6E-02 |
| actin polymerization or depolymerization | 23 | 4.5 | 8.7E-04 |
| Ras protein signal transduction | 17 | 3.3 | 3.4E-02 |
| regulation of apoptotic signaling pathway | 17 | 3.3 | 1.8E-02 |
| vesicle organization | 16 | 3.1 | 2.5E-02 |
| establishment or maintenance of cell polarity | 16 | 3.1 | 1.5E-02 |
| Arp2/3 complex-mediated actin nucleation | 8 | 1.6 | 5.1E-02 |
| apoptotic cell clearance | 8 | 1.6 | 5.1E-02 |
| lamellipodium assembly | 7 | 1.4 | 3.6E-02 |
| response to stress | 68 | 28.0 | 2.2E-03 |
| response to chemical | 62 | 25.5 | 1.2E-02 |
| locomotion | 42 | 17.3 | 6.2E-04 |
| biological adhesion | 41 | 16.9 | 1.1E-03 |
| immune system process | 40 | 16.5 | 3.3E-02 |
| cell adhesion | 40 | 16.5 | 1.7E-03 |
| cytoskeleton organization | 38 | 15.6 | 1.3E-04 |
| neurogenesis | 31 | 12.8 | 1.6E-02 |
| negative regulation of hydrolase activity | 12 | 4.9 | 3.1E-02 |
| humoral immune response | 10 | 4.1 | 2.3E-02 |
| oxidoreduction coenzyme metabolic process | 10 | 4.1 | 9.9E-03 |
| complement activation | 8 | 3.3 | 5.0E-02 |

Table 1. Immune-related biological processes enriched in the hemolymph challenged with *C. parvum* or RNA poly I:C. Count: number of modulated proteins related to BPs; Modulated protein: percentage of modulated proteins related to BPs; p-value: modified Fisher’s exact test p-value.
| BLAST annotation (NCBInr) | Immune proteins | E-value | Conserved domain | C. parvum | RNA poly I:C |
|--------------------------|-----------------|---------|------------------|-----------|-------------|
| **Pattern recognition receptors (PRRs)** | | | | | |
| natterin-like protein | 2.5E-09 | jacalin-like lectin domain, C-terminal toxin domain | ~1.7 | ~ 1.3 | 1.3 |
| Cation-independent mannos-6-phosphate receptor (M6PR) | 0.0E + 00 | M6PR repeat domains | ~ 1.6 | ~ 2 | |
| Galactose binding lectin | 5.7E-19 | Galactose binding lectin domain | ~1.5 | ~1.6 | ~1.9 |
| Beta-1,3-glucan-binding protein | 5.2E-99 | cytotytic factor, Glycosyl hydroxylase domain, beta-1,3-glucan recognition protein | ~ 1.3 | ~ 1.3 | |
| **Immune signaling** | | | | | |
| Macrophage migration inhibitory factor (MIF) | 1.2E-40 | MIF domain | 2.5 | ~ 2.3 | ~ |
| **Microorganism Internalization** | | | | | |
| myosin-2 essential light chain-like | 2.9E-58 | EF-hand, calcium binding motif, Ca2+ binding protein | 1.7 | ~ 1.4 | ~ |
| integrin beta-like protein | 2.8E-140 | Integrin beta chain VWA domain, Integrin plexin domain | ~1.4 | ~ 1.8 | ~ |
| integrin alpha-9-like isoform X2 | 1.9E-54 | integrin alpha domain | ~ 1.4 | ~ 1.7 | ~ |
| **Immune effectors** | | | | | |
| Natterin-like protein | 2.5E-09 | jacalin-like lectin domain, C-terminal toxin domain | ~1.7 | ~1.3 | 1.3 |
| Cation-independent mannos-6-phosphate receptor (M6PR) | 0.0E + 00 | M6PR repeat domains | ~ 1.6 | ~ 2 | |
| Galactose binding lectin | 5.7E-19 | Galactose binding lectin domain | ~1.5 | ~1.6 | ~1.9 |
| Beta-1,3-glucan-binding protein | 5.2E-99 | cytotytic factor, Glycosyl hydroxylase domain, beta-1,3-glucan recognition protein | ~ 1.3 | ~ 1.3 | |
| **Antimicrobial peptides** | | | | | |
| Histone H2A.V | 1.3E-54 | Core histone domain, C-terminus of histone H2A | 6 | ~ 4.6 | ~ |
| Proteases, protease inhibitors and hydrolases | | | | | |
| cysteine proteinase | 9.3E-145 | Cathepsin propeptide inhibitor domain, Peptidase C1A subfamily | 1.4 | 4.4 | 2.5 |
| serine protease-like protein | 7.4E-19 | Trypsin-like serine protease domain | 1.6 | 2.9 | 3.2 |
| Metalloproteinase-like protein | 2.9E-212 | Zinc-dependent metalloprotease domain | ~1.9 | ~ 1.6 | ~ |
| cystatin B | 1.5E-18 | Cystatin domain | 1.5 | 1.4 | 1.4 |
| Saccharase-isomaltase | 0.0E + 00 | Glycoside hydrolases domains | ~ 1.4 | ~ 2.2 | ~ |
| **Complement system** | | | | | |
| complement C1q-like protein | 2.3E-03 | C1q domain | ~ 1.7 | ~ 2.3 | |
| complement C1q-like protein | 2.2E-13 | C1q domain | ~ 1.4 | ~ 1.5 | ~ |
| complement C1q-like protein | 2.3E-03 | C1q domain | ~ 1.6 | ~ 1.7 | ~ |
| complement factor B-like | 2.8E-22 | Von Willebrand factor domain; Complement control protein (CCP) | ~ 1.8 | ~ 1.6 | ~ |
| complement component C2 | 8.9E-16 | Trypsin-like serine protease, Von Willebrand factor type A (VWA) domain | ~ 1.7 | ~ 2.2 | ~ |
| complement component C3 | 4.0E-24 | Alpha-2-macroglobulin domain | ~ 1.7 | ~ 1.5 | ~ |
| complement component C3 | 1.5E-08 | Alpha-2-macroglobulin domain | ~ 2.9 | ~ 3.6 | ~ |
| **Other immune-related process** | | | | | |
| 26S protease regulatory subunit | 7.3E-217 | 26S proteasome regulatory subunit | 1.5 | 1.7 | ~ |
| Cytochrome c oxidase | 1.2E-15 | Cytochrome c oxidase subunit Va | 4 | 2.7 | |
| Ubiquitin-conjugating enzyme E2 | 3.3E-72 | Catalytic domain of Ubiquitin-conjugating enzyme E2 | 2.5 | 1.9 | ~ |
| Laccase-like protein | 3.2E-43 | ~ | ~ 1.5 | ~ 2.4 | ~ |

Table 2. Immune related proteins significantly modulated in the hemolymph of *D. polymorpha* under both C. parvum and RNA poly I:C challenge. The BLAST annotation and expect value are derived from the sequence similarity search against NCBInr database. H = hemocytes and P = plasma and the fold change of significantly modulated proteins (p-value < 0.05) between treated and control sample is indicated.

> Immune related proteins significantly modulated in the hemolymph of *D. polymorpha* under both C. parvum and RNA poly I:C challenge. The BLAST annotation and expect value are derived from the sequence similarity search against NCBInr database. H = hemocytes and P = plasma and the fold change of significantly modulated proteins (p-value < 0.05) between treated and control sample is indicated.

oocyst walls of human pathogens contain β-1,3-glucan which may have affinity with BGP proteins. In this way, BPG from *D. polymorpha* may play a role in the general anti-protozoal response. Proteins containing C1q domains were also highly modulated in plasma exposed to *C. parvum* and RNA poly I:C (Table 2). C1q proteins are highly diversified proteins in bivalves and work as PRRs that recognize a broad range of microorganisms and may trigger the activation of the complement pathways. Immune signal transduction is an important process supervising an appropriate response against microorganisms. The macrophage migration inhibitory factor (MIF) was found two times more abundant in the plasma of *D. polymorpha* exposed to both *C. parvum* and RNA poly I:C in comparison with the non-exposed hemolymph (Table 2). MIF is a pro-inflammatory cytokine that plays a major role in the regulation of innate immunity against bacteria, protozoa and viruses. Several studies reported the involvement of such proteins in the immune defence of bivalves. MIF cytokines have been found modulated under bacterial challenge in several bivalves such as the scallop *Chlamys farreri*, the clam *Ruditapes philippinarum*, the mussel *Mytilus galloprovincialis* and the pearl oyster *Pinctada fucata* and the pearl oyster *Pinctada fucata*. These studies concluded that MIFs are involved in proinflammatory responses and promote hemocytes migration to the wound healing site. To our knowledge, it is the first report of a modulation of MIF in the hemolymph of bivalves exposed to a protozoan and RNA poly I:C. These results suggest that both *C. parvum* and RNA poly I:C may induce pro-inflammatory responses in *D. polymorpha* hemolymph.
The GSEA revealed that locomotion, and cytoskeleton organization BPs were enriched in the hemolymph exposed to protozoa and dsRNA (Table 1). Such result may indicate an activation of hemocytes to counteract *C. parvum* and RNA poly I:C. Regarding the modulated proteins, integrins and myosin-2 were significantly upregulated in hemocytes and plasma in both exposure conditions (Table 2). Integrins are transmembrane receptors involved in adhesion and internalization of microorganisms44. Their role in the immune defence of invertebrates have been evidenced in numerous studies19,45–47. These authors showed that integrins are involved in the immune defence against bacteria, viruses and protozoa, by promoting cytoskeleton remodeling, locomotion, cell-adhesion, phagocytosis and participate to immune signaling of hemocytes. Myosin is a protein that converts ATP molecules to mechanical energy, generating physical force and movement48. While myosin proteins are famous for their involvement in muscle contraction, they are also involved in a variety of movements of non-muscle cells. In hemocytes, myosin-2 is the major motor of cytoskeletal contraction for generating cell movement49. In particular, myosin-2 is responsible for many types of cell movements, generating forces behind cell motility and phagocytosis49,50. Overall, the enriched BPs and the modulation of both integrins and myosin-2 in *D. polymorpha* hemolymph exposed to *C. parvum* and RNA poly I:C suggest an interaction between hemocytes and microorganisms. Recruitment and migration of hemocytes to the site of infection is one of the key steps to neutralize invaders51. Cell motility also contribute to the phagocytosis of invading microbes and encapsulation of larger or refractive invaders. In *D. polymorpha*, interaction between hemocytes and protozoa have already been evidenced by Le Guernic et al.51. The authors observed hemocyte aggregates around the protozoa *Toxoplasma gondii* and *Giardia duodenalis*, suggesting encapsulation processes to neutralize them. Moreover, Palos-Ladeiro et al.52 have evidenced a phagocytosis of *C. parvum* oocysts by zebra hemocytes during *ex vivo* challenges.

Antimicrobial peptides (AMP), hydrolases, proteases and protease inhibitors are known to play a crucial role in the elimination of microorganisms by animals. Some proteins considered as AMP were induced in both *C. parvum* and RNA poly I:C exposures (Table 2). Among them, histone H2A was upregulated in hemocytes with a fold change higher than 4.5. The role of histones in the immune responses of aquatic invertebrates was reviewed by Nikapitiya53. Histones H4, H2A and H2B of aquatic invertebrates were up-regulated in response to a variety of environmental stressors, including microbes such as viruses and protozoa. These proteins exert antimicrobial activity against bacteria, fungi, viruses, protozoa and recent studies showed that histones are involved in DNA extracellular trap mechanisms that neutralize invaders53,54. In addition to histones, several proteolytic and hydrolytic proteins were modulated in both exposure conditions. In the plasma from both exposure conditions, a papain-like cysteine protease was the most up-regulated protease with a fold change greater than 4.4. A trypsin-like serine protease was also up-regulated with a fold change greater than 2.9 while a metalloproteumase-like protein was down-regulated. Proteases, protease inhibitors and hydrolytic enzymes are considered as powerful weapons against microbes in bivalve immunity55. Indeed, the concerted action of proteases and inhibitors is involved in several immune processes by cleaving regulator subunits of endogenous proteins, leading to their biological activation and/or exogenous proteins produced by microbes and parasites, leading to their inactivation and/or destruction56. Among them, cysteine proteases and their inhibitors such as cystatins play a central role in the Host-Pathogen Cross Talk56. A sucrase-isomaltase, was also up-regulated in the plasma challenged with *C. parvum* (FC: 1.4) and RNA poly I:C (FC: 2.2). This digestive-enzyme possess a glycoside hydrolase domain, which may contribute to the destruction of protozoa and viruses by hydrolysis of their glycosidic bonds in *D. polymorpha* plasma.

While biological process related to the complement system were exclusively enriched in the hemolymph exposed to RNA poly I:C (Table 1), many proteins related to the complement system were modulated in both *C. parvum* and RNA poly I:C challenges (Table 2). Among them, complement C2/factor B-like and several C3 complement components were significantly up-regulated or down-regulated in the plasma of *D. polymorpha* exposed to *C. parvum* and RNA poly I:C. The complement system is a sophisticated proteolytic system which acts as a critical first-line defence of innate immunity by promoting the recognition, phagocytosis, elimination of microorganisms56. While the existence of the complement system in bivalves remains contradicted, an increasing number of studies described the presence of a proto-complement system in the hemolymph of bivalves, including mussels, oysters and razor clam57–59. Several complement-related proteins (i.e. C3 component, complement factor-B) of bivalves have been shown to interact with several microorganisms such as bacteria, viruses and protozoa50,60–62. The C3 component plays a pivotal role in the complement system and was considered as the “Swiss Army Knife” of cell defence by Ricklin et al.56. Complement factor-B is a proteolytic enzyme which may act as a regulator of bivalve complement system by cleaving C3 components60. In *D. polymorpha* hemolymph, C3 complement components are the major proteins of plasma60 and both exposure to *C. parvum* and RNA poly I:C resulted in the modulation of the abundances of complement-related proteins. Taking together, these results suggest a strong involvement of the complement system in the immune defence occurring in the hemolymph of *D. polymorpha*.

**Immune proteins modulated exclusively in *C. parvum* exposure.** The biological interactions between protozoa and zebra mussel have only been studied at the cellular level. Studies have shown that protozoa bioaccumulated by the mussel interact with the immune system of *D. polymorpha* by inducing changes in apoptosis and/or phagocytosis capacity of hemocytes5–11. The comparative analysis conducted in hemocytes and plasma exposed to *C. parvum* led to the identification of numerous immune-related proteins involved exclusively modulated in *C. parvum* exposure (Table 3). Among the PRRs modulated exclusively in the plasma exposed to *C. parvum*, a Fibrinogen-related protein (FReD) was observed twice more abundant compared to control condition, and gaelctin-like protein was up-regulated in hemocytes (FC: 1.6). In invertebrates, the primary role of FReDs is defence61. Molluscan FReDs are highly diversified molecules involved in pathogen recognition63. Numerous studies have reported up-regulation of FReD genes in the hemolymph of bivalve challenged with bacteria and protozoa63–68. Authors agreed that FReDs are able to recognize and bind carbohydrates in pathogens and activate immune responses.
such as the complement system. In the hemolymph of *D. polymorpha*, the up regulation of a FReD in plasma exposed to *C. parvum* may indicate that FReDs contribute to the recognition and destruction of protozoa.

**Table 3.** Immune-related proteins significantly modulated exclusively in the hemolymph of *D. polymorpha* challenged with *C. parvum*. The BLAST annotation and expect value are derived from the sequence similarity search against NCBI nr database. H = hemocytes and P = plasma and the fold change of significantly modulated proteins (p-value < 0.05) between treated and control sample is indicated.
to *C. parvum*, no modulation was observed for the RNA poly I:C challenge. STAT proteins are certainly involved in the immune response of *D. polymorpha* but further analysis is needed to clarify their role in the interaction with microorganisms.

Vesicle transport and Ras protein signal transduction BPs were enriched in the hemolymph exposed to *C. parvum* (Table 1). In addition, several Ras-related proteins of the Rab-family were modulated in hemocytes (Table 3). In both vertebrates and invertebrates, Rab-like proteins are considered as important regulators of intracellular transport and fusion of intracellular structures. Vesicle trafficking is crucial for several immune processes. For example, the release of immune proteins in plasma and the endocytosis of microorganisms necessarily involves a significant vesicular traffic and fusion. Biological processes related to the internalization of microorganisms were also enriched in the hemolymph exposed to *C. parvum*, including actin polymerization or depolymerization, ARP2/3 complex-mediated actin nucleation and lamellipodium assembly BPs (Table 1). In addition, numerous proteins related to the actin-related protein 2/3 complex (Arp 2/3) were modulated exclusively in the hemocytes challenged with *C. parvum* (Table 3). Among them actin-related protein 2/3 complex subunit (FC: 1.9), ras-related C3 botulinum toxin substrate (FC: 1.6), Cell division control protein 42 (FC: 1.4) were upregulated while the neural Wiskott-Aldrich syndrome protein was down regulated (FC: 1.9). The Arp 2/3 complex is a membrane complex that is considered to play a central role in the assembly of actin filaments during the migration and phagocytosis processes of macrophages. In human cells, implication of the Arp 2/3 complex during *C. parvum* infection has been underlined by Elliott et al. The authors showed that *C. parvum* induced actin polymerization at sites of infection by triggering nucleation machinery of the Arp 2/3 complex. Focusing on hemocytes, Lau et al. observed an up-regulation of Arp 2/3-related genes correlated with an increase of cell motility when hemocytes of the oyster *Crassostrea virginica* were *in vitro* challenged with the intracellular parasite *Perkinus marinus*. In shrimp exposed to the bacteria *Vibrio anguillarum*, Xu et al. observed a modulation of genes involved in the regulation of Arp 2/3 complex when bacteria were phagocytosed by hemocytes. Studies have observed, by flow cytometry analyses, a phagocytosis of *C. parvum* oocysts by zebra mussel hemocytes after 4 hours of *ex vivo* exposure. As evidenced by both BP enrichment analysis and BLAST annotation of modulated proteins, our results may indicate that *C. parvum* are phagocytosed by *D. polymorpha* via the Arp 2/3 complex.

Proteins devoted to the destruction of *C. parvum* were mainly modulated in the hemocytes (Table 2). Numerous cathepsins were upregulated in hemocytes exposed to *C. parvum*. Of them three different types of cathepsins were up-regulated in hemocytes with a fold change higher than 1.7 and a cathepsin D was up-regulated in plasma. Cathepsins have been subject to several investigation and are linked to the immune defence in bivalves. Cathepsins are lysosomal cysteine proteases that regulate the immune and cell death process and contribute to the degradation of phagocytosed microorganisms. The high number of modulated cathepsins in *D. polymorpha* hemocytes strongly suggest that the lysosome-mediated antimicrobial defence is engaged against *C. parvum*. Histone H2B (FC: 1.4) and Ferritin (FC: 1.5) were also up-regulated exclusively in the hemocytes exposed to *C. parvum*. To survive and replicate in hosts, many protozoan invaders secure host iron. Ferritins are iron-sequestering proteins that regulate iron homeostasis in cells and tissues, which are described as important immune regulators that limit the spreading of microbes such as bacteria. Numerous studies highlighted the immune function of ferritins in bivalve. In scallop and freshwater mussel, ferritin genes were up-regulated in response to bacteria correlated with an inhibition of bacteria growth. Furthermore, in response to quahog parasite unknown (QPX), ferritin genes were highly expressed in mantle and gills tissues of the clam *Mercenaria mercenaria*. Iron is probably an important element for the proliferation of *C. parvum*. The up-regulation of the abundance of a ferritin in the hemocytes exposed to *C. parvum* may suggest that this protein prevents the growth of *C. parvum* in zebra mussel hemolymph.

The modulation of apoptosis in the hemolymph exposed to *C. parvum* were evidenced by both BPs enrichment analysis and the BLAST annotation of modulated proteins. Indeed, the regulation of apoptotic signaling pathway and apoptotic cell clearance BPs were significantly enriched in the hemolymph challenged with *C. parvum*. Key proteins involved in the regulation and induction of the intrinsic apoptotic pathway were modulated in hemocytes. Among them, the apoptosis regulator BAX, caspases, cytochrome C and voltage-dependent anion channel were up-regulated in hemocytes challenged with *C. parvum* while another caspase was down regulated. Apoptosis is fundamental biological process involved in immune system homeostasis by preventing inflammatory damage and limiting the spread of pathogens. While apoptosis is described as a mechanism that removes parasite cells, it has been shown induction of inhibition of programmed cell death by protozoa can be linked to parasite survival strategies. In bivalve hemocytes, apoptosis is a common response to protozoa. In particular, the involvement of apoptosis in interactions between zebra mussel hemocytes and their intracellular parasite has been evidenced by Minguez et al.

Finally, other proteins exclusively modulated in the hemolymph exposed to *C. parvum* were related to oxidative metabolism. Of them, a dual oxidase (Duox) was down-regulated in hemocytes. Duox proteins are well-defined sources of reactive oxygen species (ROS) production. ROS production in bivalve hemocytes is regarded a conserved and widespread immune defence mechanisms in response to microorganisms, since cytotoxic ROS interact with a wide variety of organic and inorganic molecules, altering their functions. Induction of Duox genes have been associated to successful defence response against microorganisms in several bivalves, probably by promoting the production of ROS that degrade internalized microorganisms. In this context, the modulation of a Duox protein in hemocytes exposed to *C. parvum* may indicate that ROS-dependent killing processes are involved in defence against *C. parvum*. While ROS have a biocidal effect on invading microbes, ROS production can also injure the cells of the host by causing oxidative stress. In this case, the excess ROS should be removed by antioxidant enzymes to avoid damage on host cells. Noteworthy, two antioxidant enzymes were upregulated in *D. polymorpha* hemocytes exposed to *C. parvum*. More specifically, a manganese superoxide dismutase (SOD) (FC: 2.0) and Cu/Zn SOD (FC: 1.5) were up regulated in hemocytes. SOD are anti-oxidant enzymes involved in antioxidative defence system, by protecting organisms from excess of ROS and maintaining...
the redox balance of immune system. In the oyster *C. gigas*, SODs considered as the major protein group in plasma was able to bind to various microorganisms. In addition to its antioxidative activity, extracellular SODs enhanced cellular immune response in oysters exposed to *Vibrio splendidus* by acting as a PRR and by contributing to the immune priming of oysters.

**Immune proteins modulated exclusively in RNA poly I:C exposure.** Molecular mechanisms involved in anti-viral responses of bivalves remained till now the “blackbox” to solve since tractable systems to induce viral infection have only recently developed. Recently, the double-stranded RNA poly I:C emerged as a powerful inducer of antiviral response in bivalve species. In this study, the hemolymph of *D. polymorpha* was challenged with RNA poly I:C to simulate a viral infection and the proteins involved in the antiviral defence of zebra mussels were identified. Our results have shown that the most enriched BPs related to immunity were “response to stress” and “response to chemical”, both representing more than 27% of the modulated proteins in the hemolymph challenged with RNA poly I:C (Table 1). The GSEA also revealed that immune system processes were modulated in the hemolymph exposed to RNA poly I:C since many BPs related to immunity were enriched, including the immune system process and humoral immune response BPs. Recent results suggest that antiviral defence of bivalves are executed through an interferon pathway like in vertebrates. Key proteins involved in the interferon pathway were identified in the hemolymph of *D. polymorpha*. In our study, no proteins related to the interferon pathway were detected as modulated in response to RNA poly I:C. However, many proteins that have already shown interactions with viruses in non-bivalve organisms were modulated exclusively in RNA poly I:C exposure, suggesting a potential role in the anti-viral responses of *D. polymorpha*. In contrast with the hemolymph exposed to *C. parvum*, immune proteins exclusively modulated in the hemolymph challenged with RNA poly I:C were mainly modulated in the plasma fraction (Table 4). Like in other marine bivalves, this result suggests that plasma proteins may play a central role in *D. polymorpha* antiviral response.

Of the proteins acting as PRRs, a Peptidoglycan-recognition protein (PGPR) containing a lysozyme domain and a galectin protein were exclusively modulated in the plasma exposed to RNA poly I:C (Table 4). PRGPs have multiple immune-related functions. PRGPs were first considered as a PRRs that recognize peptidoglycans from bacteria cell walls. In addition to their role of recognition of microbes, a lysozyme domain was observed in the PRGP of oyster hemocytes which may confer a lytic function. In insects PRPGs interact with several immune signaling pathways. Involvement of PRPG in the immune defence against bacterial invasions was evidenced in several bivalve species such as the manila clam *Ruditapes philippinarum*, the bay scallop and the pacific oyster. Few studies have investigated the role of PRPGs in anti-viral defence. In the grass carp and human cells, PRPGs genes were strongly expressed during infection with dsRNA poly I:C. In the silkworms *Bombyx mori*, PRPGs are believed to exert distinct functions in host responses to bacteria and viruses. Authors suggest that silkworms PRPG act as PR in response to bacteria while PRPGs regulate immune signaling during viral infection. Recently, Wang et al. observed an up-regulation of PRPGs in the oyster *C. gigas* challenged with RNA poly I:C and considered PRPGs as a PRRs involved in virus recognition.

A granulin protein was up-regulated in plasma exposed to RNA poly I:C (FC: 1.7). In vertebrates, granulins are considered as important modulators of cell growth and are believed to be involved in the immune signalling. Furthermore, granulins were able to bind synthetic DNA molecules and were considered as a central actor in the signaling of viral invasion. Especially, granulins were described as soluble co-factor of the toll-like receptor signaling pathway in macrophages. To our knowledge, the role of granulins in the immune system of invertebrates has only been studied in a context of snail infection by trematodes. In these studies, granulins are described as a cytokine that promotes hemocyte activation and proliferation, making the snail resistant to its parasite. The up-regulation of the extracellular granulin-like protein in the hemolymph of *D. polymorpha* challenged with RNA poly I:C suggest that granulins participate to the immune signalling against dsRNA.

Proteins involved in nucleotide metabolism were also modulated in response to RNA poly I:C. An RNA helicase protein containing a DEAD box helicase domain was down-regulated in hemocytes while an ectonucleotide pyrophosphatase/phosphodiesterase (ENPP) protein containing a DNA/RNA nuclease domain was up-regulated in plasma with a fold change greater than 4.5. The role of RNA helicases in anti-viral responses has been subject to extensive investigations and contribute as both sensors and effectors of antiviral immunity. RNA helicases have been shown to be up regulated in the hemocyte of shrimp infected by the WSSV. In bivalves, retinoic acid-inducible gene-1-like receptors (RIG-like receptors) which contain a DEAD box RNA helicase are recognized as fundamental receptors of viral infections. The RNA helicase modulated in the hemolymph of *D. polymorpha* exposed to RNA poly I:C is probably a protein that functions similarly to RIG-like proteins found in bivalves. ENPPs hydrolyze phosphates from nucleotides and their derivatives, and the extracellular form have already been linked to viral infections. For example, ENPP was one of the most significantly up-regulated secretory proteins associated with *Hepatitis B* virus replication in human cells. The ENPP modulated in *D. polymorpha* contained a DNA/RNA non-specific nuclease domain whose function is to cleave double-stranded and single-stranded nucleic acids. In insects, *in vivo* assay revealed that dsRNA was rapidly degraded in the plasma by proteins that contain a DNA/RNA nuclease domain. The conserved domains of both RNA helicase and ENPP and their function in nucleic acid biology suggest their involvement in *D. polymorpha* response to dsRNA.

The abundance of a Thaumatin-like protein (TLP) was significantly down-regulated in the plasma of exposed to RNA poly I:C (FC: −1.5) (Table 4). In plants, TLPs are believed to exert antifungal activity, xylanase inhibitor activity and glucan binding and glucanase activities. In bivalves, information on TLP functions are relatively scarce. To our knowledge, only Niu et al. investigated the role of TLPs in the oyster *C. gigas*. Authors observed a higher expression level of TLPs genes in immune-related organs such as hemocytes and hepatopancreas. Moreover, they observed an up-regulation of the expression of TLP genes in oysters challenged with mannan, *Pichia pastoris* yeast and RNA poly I:C. They concluded that TLPs play a vital role in defending against fungal infection. In our study, TLPs were modulated in the plasma of *D. polymorpha* exposed to RNA poly I:C suggest...
between hemocytes and biological stressors. This strategy has proven its efficiency for identifying proteins that

**Ex vivo**

(p-value = hemocytes and P NCBInr database. H plasma and the fold change of significantly modulated proteins

RNA poly I:C. The BLAST annotation and expect value are derived from the sequence similarity search against

studies have shown that the immune system of bivalves is sensitive to a broad range of chemical contaminants

the immune system of organisms, thus posing a risk to the health of individuals and populations. Numerous

microorganisms. Investigations have shown that many molecules released in the environment are likely to alter

system contributes to ensuring the integrity of the host organism by eliminating foreign constituents such as

D. polymorpha in order to understand the interaction between contaminations and biological responses of

a large panel of immune processes (Fig. 3). In both exposure conditions, several proteins involved in cell motility

hemolymph exposed to C. parvum and RNA poly I:C provided the identification of numerous proteins reflecting

defence were observed in the hemolymph exposed to RNA poly I:C. All these proteins constitute potential immu-

resulted in the identification of key proteins involved in immune signaling, phagocytosis and apoptosis process

C. parvum and destruction of microorganisms were modulated. In challenge, the comparative proteomic analysis

their involvement in zebra mussel immunity. A mucin-like protein was also significantly modulated in the plasma

exposed to RNA poly I:C with a fold change greater than 2. Lieleg et al. considered that mucin biopolymers exert a broad-spectrum antiviral activity. Thus, it is not excluded that this mucin-like protein act as anti-viral agents of the D. polymorpha hemolymph.

Other immune proteins exclusively modulated by RNA poly I:C were related to proteolytic and hydrolytic enzymes. Among them, the protease inhibitor alpha-2 macroglobulin was modulated with a fold change greater than 1.5 in plasma. Serine-protease inhibitor (Serpin) and lysosomal alpha-mannosidase were down-regulated in

immunomarkers for ecotoxicological studies. The zebra mussel, Dreissena polymorpha is a sentinel species used in aquatic ecotoxicology in part because this filter bivalve accumulates and concentrates a large number of biological and chemical contaminants. Current studies seek to understand the interaction between contaminations and biological responses of D. polymorpha in order to assess the impacts of contaminants on living organisms and ecosystems. Among biological processes, the immune system contributes to ensuring the integrity of the host organism by eliminating foreign constituents such as microorganisms. Investigations have shown that many molecules released in the environment are likely to alter the immune system of organisms, thus posing a risk to the health of individuals and populations. Numerous studies have shown that the immune system of bivalves is sensitive to a broad range of chemical contaminants which affect their immunocompetence. The comparative proteomic analysis performed on D. polymorpha hemolymph exposed to C. parvum and RNA poly I:C provided the identification of numerous proteins reflecting a large panel of immune processes (Fig. 3). In both exposure conditions, several proteins involved in cell motility and destruction of microorganisms were modulated. In C. parvum challenge, the comparative proteomic analysis resulted in the identification of key proteins involved in immune signaling, phagocytosis and apoptosis process as well as proteins of the lysosome-mediated antimicrobial defence. Moreover proteins that may act in anti-viral defence were observed in the hemolymph exposed to RNA poly I:C. All these proteins constitute potential immunomarkers for evaluating the impact of contaminans on zebra mussel immunity.

In this study, the hemolymph was challenged during 4 h to C. parvum and RNA poly I:C in ex vivo conditions. Ex vivo exposures were initially designed to simplify the biological system and promote a strong interaction between hemocytes and biological stressors. This strategy has proven its efficiency for identifying proteins that
may play a key role in the management of *C. parvum* and RNA poly I:C. However, such exposures do not take into account the integrity of the body's biology and the dynamic of microorganisms within mussel tissues. We recommend that future exposure should be performed with longer *in vivo* condition in order to be as close as possible to biological reality. *In vivo* assays are more appropriate for studying the immune response of organisms because changes in immune responses may be related to hormonal, nutritional, physical and biological alterations such as changes in the microbiota\(^{127-129}\). Furthermore, our results only give a picture of the early immune responses occurring after 4h of exposure. Experimental challenges with different kinetics of contamination would provide a better understanding of the involvement of immune proteins in the contaminant management. Finally, the present differential proteomics performed with the hemolymph of *D. polymorpha* has led to the identification of key proteins reflecting a variety of immune mechanisms, including recognition, internalization and destruction of microorganisms. In addition to providing information on host-microbial interaction in the zebra mussel, this study paves the way for the identification of new immunomarkers that reflect the broad immune processes of *D. polymorpha*. The modulation of these proteins and the analysis of their genetic regulation should be further validated by other methodologies, such as targeted proteomics and RNAseq\(^{14,130}\). Once approved, these immunomarkers will provide a better understanding of the impact of biological contaminations on the immune system of aquatic organisms. Finally, work remains for a better understanding of the immune response of *D. polymorpha*.

**Conclusion**

Overall, this study helped to understand the interaction between the immune system of *D. polymorpha* and microorganisms. The comparative proteomic analysis performed on hemocytes and plasma exposed to *C. parvum* and RNA poly I:C has provided a large set of information on the immune processes involved in anti-protozoal and anti-viral defence of *D. polymorpha*. The GSEA and the BLAST annotation performed on modulated proteins have shown that common immune responses were induced by *C. parvum* and RNA poly I:C. Indeed, several proteins involved in the recognition and destruction were modulated in both exposure conditions. Among them, proteins related to the complement system were up and down regulated, suggesting a significant implication of the complement system in *D. polymorpha* immune defence. Other immune-related proteins were exclusively modulated in *C. parvum* or RNA poly I:C exposures, which suggest a certain immune specificity depending on the nature of encountered microbes. Multiple clues showed that *C. parvum* was managed by hemocytes of *D. polymorpha* hemolymph. Firstly, a higher number of proteins were modulated in hemocytes compared to plasma. Then, key proteins involved in phagocytosis and apoptosis process as well as proteins of the lysosome-mediated antimicrobial defence were modulated in hemocytes exposed to *C. parvum*. In contrast, a higher number of modulated proteins was observed in plasma for the RNA poly I:C challenge, which suggests that at four hours of exposure dsRNA is managed in the extracellular medium of hemolymph. Among the proteins modulated

![Figure 3. Summary of immune proteins from *D. polymorpha* hemolymph modulated during interaction with microorganisms. Proteins written in black were modulated by both *C. parvum* and RNA poly I:C, the green ones were exclusively modulated in *C. parvum* exposure while the red ones were exclusively modulated in RNA poly I:C exposure.](image-url)
Sequencing Grade Trypsin (Roche) in presence of 0.01% ProteaseMAX surfactant (Promega) for 1 h at 50 °C as each polyacrylamide band was first alkylated with iodoacetamide, reduced with DTT and then proteolyzed with ultra-pure water, each polyacrylamide band corresponding to each whole proteome was excised. Proteins from formed at 200 V in MES buffer (Invitrogen). After Coomassie Blue Safe staining (Invitrogen) and destaining with 0.38 mM EDTA, 0.17 mM SERV A Blue G250, 0.131 mM phenol red, buffered at pH 8.5 and supplemented with (Invitrogen) consisting of 80 mM Tris / HCl, 106 mM Tris base, 1.5% lithium dodecyl sulfate, 7.5% glycerol, et al.10 and (iii) the antiviral challenge where the hemolymph was

**Material and Methods**

**Sampling and maintenance of animals.** Sampling and maintenance of animals were performed as described in Leprêtre et al.13. Zebra mussels between 15- and 25-mm long were collected at the Der lake (Grand Est, France) in May 2017. Mussels were maintained for at least two weeks at the laboratory in 20 L tanks filled with spring water Cristaline Aurèle (Jandun, France), at constant temperature (14 °C) and with a controlled aeration. Mussels were fed twice a week with a mixture of two species of microalgae, Chlorella pyrenoidosa and Scenedesmus obliquus (two million of microalgae per day and per mussel). Tank water was removed weekly.

**Ex vivo challenge experiment.** After two weeks of acclimation in the laboratory, 150 μL of hemolymph per mussel was withdrawn from the posterior adductor muscle by gentle aspiration with a 30 g × 0.3 mL syringe. The quality of each sample was checked under a microscope for confirming the exclusive presence of hemocytes. Samples were pooled to obtain 3 ml hemolymph. As described in Fig. 1, from this pool (3 mL), 1 mL of hemolymph was distributed in order to perform three exposure conditions: (i) the control condition with hemolymph free, (ii) the anti/protozoal challenge where the hemolymph was exposed to Cryptosporidium parvum oocysts purified in phosphate-buffered saline (pH 7.4) (5:1 protozoa:hemocyte; INRA, Val de Loire research center, France) as referred in Le Guernic et al.19 and (iii) the antiviral challenge where the hemolymph was exposed to 100 μg/ml of RNA poly I:C (Sigma-Aldrich). For all conditions, samples were incubated at 16 °C during 4 hours and the experiment was repeated 5 times in order to obtain 5 replicates per exposure conditions.

**Protein extraction and trypsin digestion.** Sample preparation for mass spectrometry analysis was performed as described in Leprêtre et al.13. Briefly, pools of hemolymph were centrifuged at 800 × g for 15 min at 4 °C to separate hemocytes from plasma. 500 μL of Tri-reagent (Sigma-Aldrich) were added to hemocyte pellets and stored at −20 °C until the protein extraction, 3 volumes of ice-cold acetone were immediately added to the plasma and the resulting samples were stored overnight at −20 °C for protein precipitation.

For hemocytes, cells were lysed by passing them ten times through a syringe equipped with a 25 G needle and then centrifuged at 12,000 × g during 2 min for an initial clean up. RNAs were removed from the resulting supernatant by adding 100 μL of chloroform, incubating for 15 min, centrifugation at 12,000 × g at 4 °C for 15 min, and removing the colorless aqueous phase. DNA was discarded after adding 150 μL of absolute ethanol for 3 min followed by centrifugation at 2,000 × g at 4 °C for 5 min. Then, proteins from the resulting supernatant were precipitated by adding 3 volumes of ice-cold acetone to the supernatant. Samples were collected at room temperature for 10 min and then centrifuged during 10 min at 12,000 × g and 4 °C. The remaining protein pellet was then washed three times during 10 min in 1 mL of 0.3 M guanidine hydrochloride in 95% ethanol (Vol:Vol) followed by centrifugation at 8,000 × g and 4 °C for 5 min and removal of the supernatant. A final wash was performed with 1 mL of 95% ethanol before protein pellets were dried at room temperature and re-suspended in 80 μL of resuspension buffer.

Proteins precipitated from plasma were centrifuged at 15,000 × g for 10 min at 4 °C. The protein pellets were washed twice with 1 mL of 80% cold acetone followed by centrifugation at 15,000 × g for 10 min at 4 °C, and removal of the supernatant. Then, the pellets were dried at room temperature and re-suspended in 80 μL of re-suspension buffer consisting of 6 M urea; 2 M thiourea; 4% 3-[[(3-cholamidopropyl)dimethyl-ammonio] -1-propanesulfonate (CHAPS); 65 mM dithiothreitol (DTT), 0.1% Triton X100 (Vol:Vol), pH 7.8.

The concentration of total proteins in each sample was measured by Bradford against a bovine serum albumin (BSA) calibration curve. Biological replicates were diluted in re-suspension buffer to obtain the same protein concentration of 2.25 μg/mL. A volume of 20 μL of each sample was diluted with 10 μL of LDS 3X buffer (Invitrogen) consisting of 80 mM Tris / HCl, 106 mM Tris base, 1.5% lithium dodecyl sulfate, 7.5% glycerol, 0.38 mM EDTA, 0.17 mM SERVA Blue G250, 0.131 mM phenol red, buffered at pH 8.5 and supplemented with 2.5% beta-mercaptoethanol. Then, samples were boiled for 5 min at 95 °C, centrifuged briefly, and loaded into a 4–12% gradient 10-well NuPAGE gel (Invitrogen). A 5 min electrophoretic migration of the proteins was performed at 200 V in MES buffer (Invitrogen). After Coomassie Blue Safe staining (Invitrogen) and destaining with ultra-pure water, each polyacrylamide band corresponding to each whole proteome was excised. Proteins from each polyacrylamide band was first alkylated with iodoacetamide, reduced with DTT and then proteolyzed with Sequencing Grade Trypsin (Roche) (in presence of 0.01% ProteaseMAX surfactant (Promega) for 1 h at 50 °C as previously described13. The proteolysis was then stopped with the addition of TFA 5% to reach a final concentration of 0.5%.

**NanoLC-MS/MS analysis.** The resulting peptide mixtures were analyzed in a data-dependent mode with a Q Exactive HF tandem mass spectrometer (ThermoFischer Scientific) coupled on line to an Ultimate 3000 chromatography system (ThermoFisher), essentially as previously described14. A volume of 5 μL of each sample was injected, first desalted with a reverse-phase capillary precolumn C18 PepMap 100 (LC-Packing) and then resolved with a nanoscale 500 mm C18 PepMap 100 (LC-Packing) column using a 90 min gradient from 2% to 20% in 70 min and from 20% to 36% in 20 min of acetonitrile, 0.1% formic acid and then 80% for 1 min. Mass spectra of peptide ions with +2+ and +3+ charge states were acquired in the full scan range 350–1800 m/z, with a resolution set at 60,000, an AGC target at 3 × 10⁶. MS/MS mass spectra were acquired following a Top20 method for selecting peptides for fragmentation with an AGC target set at 10⁶, a loop count of 60 msec, an isolation
window of 1.6 m/z, a resolution at 15,000, an exclusion time of 10 sec, and a scan carried out in the mass range from 200 to 2,000 m/z.

**Interpretation of MS/MS spectra.** MS/MS spectra were assigned by using Mascot search engine against the polypeptide database "Dreissene_cascade", a database derived from proteogenomic investigations which comprises 54,097 specific polypeptide sequences, totaling 20,419,358 residues. Peptide assignments were validated with the following parameters: full-trypsin specificity, a maximum of two missed cleavages, a mass tolerance of 5 ppm on the parent ion and 0.02 Da on the MS/MS analysis, and a p-value for peptide validation at 0.05. Modifications taken into account were carboxamidomethylated cysteine (+57.0215) as static modification and oxidized methionine (+15.9949) and deamidation of asparagine and glutamine (+0.9848) as dynamic modifications. A protein was validated when at least two unique peptide sequences were detected in at least one of the 30 samples used in this study. The false positive rate for protein identification was estimated, through a search with the reverse decoy MASCOT option, as less than 1% using the same parameters. For each condition and for each replicate, the numbers of peptide-to-spectrum assignments observed for the different detected peptides of the same polypeptide were summed (spectral counts). Identified proteins which share the same attributed peptides, transcript annotation and BLAST annotation were used for parsimony and creation of protein families for which the longest polypeptide isof orm sequence was selected to be the representative.

**Differential proteomics.** The abundance of each protein in hemocytes and plasma from the hemolymph exposed to *C. parvum* or RNA poly I:C were compared to the abundance of proteins from the unexposed hemolymph, using the T-fold module of the Patternlab program for proteomics version 4.0.0.59. The comparative analysis of the expression of proteins in plasma and hemocytes was based on the spectral counts obtained with the five replicates of each exposure condition for each protein. The difference was analyzed using a L-stringency set at 0.6, a minimum of four measurements out of five replicates, and normalization of the dataset upon the total signal. From this analysis, proteins were classified into 4 groups: the first ones correspond to identifications that satisfied both the automatic fold-change threshold (FC > 1.3) and statistical criterion (p-value < 0.05), the second ones did not meet the fold and p-value criteria, the third ones were filtered out by the L-stringency which needs further experimentation to verify if they are indeed differentially abundant and the fourth ones are not considered as they satisfied the fold criterion but not the p-value. The protein classified in the first category were considered as potential protein signatures of the treatment.

**Bioinformatics analysis.** BLAST annotation of identified proteins was performed using the BLASTp module of DIAMOND (version 0.8.22). Sequence similarity search with an E-value threshold set at 1E-03 was carried out without taxonomic restriction against two databases: (i) the National Center for Biotechnology Information (NCBI) database of non-redundant protein sequences (nr) (131,025,392 sequences released 05-09-2017) and (ii) the Swissprot database (465,089 sequences released, 30-09-2016). Then a search of the conserved domain (CD) of proteins with the Batch CD-Search tool of NCBI server was performed to support BLAST annotations.

An enrichment analysis of biological processes (BPs) in the hemolymph exposed to *C. parvum* and RNA poly I:C was performed with the Database for Annotation, Visualization and Integrated Discovery (DAVID) tools for GO annotation functional enrichment analysis of proteins. Using Uniprot accessions, a Fisher’ exact test was performed on significantly modulated proteins in hemolymph exposed to *C. parvum* or RNA poly I:C to reveal enriched BPs. The entire set of proteins identified in both hemocytes and plasma samples from all samples was used as background to perform the enrichment analysis. Only enriched BPs related to immune process with a σ risk lower than 5% are considered in this study.

**Mass spectrometry and proteomics data.** The mass spectrometry and proteomics data were deposited to the ProteomeXchange Consortium via the PRIDE137 partner repository with the dataset identifier PXD017747 and 10.6019/PXD017747.

Received: 16 January 2020; Accepted: 27 March 2020; Published online: 10 April 2020

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Acknowledgements
The authors are grateful to Dr. Damien RIOULT and Isabelle BONNARD for their contribution and numerous advices to this study. This PhD work was funded by the University of Reims Champagne-Ardenne (project PROMETHEE). Financial support was provided by the French National Research Agency (ANR) project MOBIDIC (Mollusc bivalves as indicators of contamination of water bodies by protozoan parasites, Project ANR-15-CE34-0005).

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Competing interests
The authors declare no competing interests.

Additional information
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