The Gas6/TAM System and Multiple Sclerosis

Mattia Bellan 1,2,3, Mario Pirisi 1,2,3 and Pier Paolo Sainaghi 2,3,*

1 Department of Translational Medicine, Università del Piemonte Orientale, UPO, via Solaroli 17, 28100 Novara, Italy; bellanmattia@yahoo.it (M.B.); mario.pirisi@med.uniupo.it (M.P.)
2 Immuno-rheumatology Unit, Internal Medicine Division, “Maggiore della Carità” Hospital, 28100 Novara, Italy
3 IRCAD, Interdisciplinary Research Center of Autoimmune Diseases, 28100 Novara, Italy
*
Correspondence: pierpaolo.sainaghi@med.uniupo.it; Tel.: +39-0321-3733966

Academic Editor: Christoph Kleinschnitz
Received: 16 September 2016; Accepted: 26 October 2016; Published: 28 October 2016

Abstract: Growth arrest specific 6 (Gas6) is a multimodular circulating protein, the biological actions of which are mediated by the interaction with three transmembrane tyrosine kinase receptors: Tyro3, Axl, and MerTK, collectively named TAM. Over the last few decades, many progresses have been done in the understanding of the biological activities of this highly pleiotropic system, which plays a role in the regulation of immune response, inflammation, coagulation, cell growth, and clearance of apoptotic bodies. Recent findings have further related Gas6 and TAM receptors to neuroinflammation in general and, specifically, to multiple sclerosis (MS). In this paper, we review the biology of the Gas6/TAM system and the current evidence supporting its potential role in the pathogenesis of MS.

Keywords: Gas6; TAM receptors; Axl; MerTK; Tyro3; multiple sclerosis

1. The Gas6/TAM Receptors System

Growth arrest specific 6 (Gas6) is a gene firstly identified in murine fibroblasts in 1988 [1], expressed during the G0 phase and down-regulated upon induction of growth by serum. The human gene was cloned in 1993 [2] and encodes for a vitamin K-dependent protein which is expressed in different tissues, such as the gut, bone marrow, endothelial cells, and fibroblasts [2–4]. Structurally, Gas6 shares a high homology with protein S (ProS), another vitamin K-dependent circulating protein, which plays an anticoagulant role in vivo. Gas6 and ProS are both characterized by the presence of a C-terminal sex hormone-binding globulin (SHBG)-like structure composed by two globular laminin-G-like domains. The N-terminal region contains 11 γ-carboxyglutamic acid residuals (Gla), a loop region and four epidermal growth factor (EGF)-like domains. The post-translational carboxylation of γ-glutamyl residuals is the vitamin K-dependent process that confers a high affinity for negatively-charged membrane phospholipids, crucial for some Gas6 functions [2,5].

Gas6 and ProS are both ligands of three different tyrosine kinase receptors, collectively named TAM, an acronym for Tyro3, Axl, and MerTK; Axl is characterized by the highest affinity for Gas6 [6–8]. The extracellular region of the receptor consists of an immunoglobulin (Ig) domain, followed by a tandem fibronectin 3 domain; the Ig domain interacts with the SHBG-like structure of the biological ligands. The single transmembrane domain is followed by the intracellular region, which is responsible for the tyrosine kinase activity activated by receptor dimerization [9]. This is coupled to the downstream activation of different pathways, including phosphoinositide 3 kinase (PI3K)/Akt, mitogen-activated protein kinase (MAPK), extracellular signal-regulated kinases (ERK) 1/2, phospholipase C [10–12].
All three TAM receptors can be detected in a circulating, soluble form (respectively named sTyr3, sAxl, and sMer), which is the result of the proteolytic cleavage of the transmembrane receptor by a metalloproteinase (Figure 1) [13,14]. This cleavage results in the inactivation of the receptors; furthermore, these soluble forms act as decoy receptors for the ligands [15].

**Figure 1.** Interaction between growth arrest specific 6 (Gas6) and TAM (Tyro3, Axl and Mer) receptors. The sex hormone-binding globulin (SHBG) domain of Gas6, made of two globular laminin-G like repeats, interacts with the immunoglobulin-like domain of the TAM receptors. The extracellular portion is completed by two fibronectin 3 repeats, while the tyrosine kinase activity is played by the intracellular region of the receptor. ADAM10 and ADAM17 are two metalloproteinases responsible of TAM receptors cleavage. Their proteolytic domain cleaves TAM receptors in close proximity to the transmembrane domain, leading to the formation of soluble TAM (sTAM). sTAM receptors inhibit Gas6 activity by acting both as decoy receptors and reducing the number of ligand sites on the cell membrane (see text for further explanations and references). Ig, immunoglobulin; EGF, epidermal growth factor.

TAM receptors are differentially expressed in human tissues. Tyro3 predominates in mouse and human central nervous systems (CNS) [16,17], but it is also expressed by platelets [18], the heart [19], ovaries and testis [20], breasts [21], osteoclasts [22], and the retina [23]. Axl is widely expressed in many tissues and organs, including the brain [24], liver [25], kidney [26], heart [19], monocytes/macrophages [27], and endothelial [28] and vascular smooth muscle cells [29]. Finally, MerTK is the main mediator of Gas6 activity on immune cells [30,31], but is also expressed by the brain [24], platelets [32], gonads and prostate [33,34], lung [35], retina [23], kidney [36], and heart [19].

The Gas6/TAM system is highly pleiotropic and has many biological functions. Hence, it has been studied in many conditions. Gas6 and TAM regulate cell growth and an overactivation of the system has been associated to several neoplastic conditions and proposed as a novel therapeutic target [37–48]. Furthermore, TAM receptors are involved in haemostasis. It is well known that their ligand ProS is a master regulator of the coagulative cascade, by working as a non-enzymatic cofactor for activated protein C in the breakdown of coagulation factors (F) Va and FVIIIa [49]. Gas6 seems to play a complementary role on platelet function, which is impaired in Gas6 knockout (KO) mice, resulting in a defective thrombus formation [50].
2. Gas6 and TAM Receptors System, a Regulator of Innate Immunity

One of the best defined activities of the Gas6/TAM system, however, is the regulation of innate immunity. MerTK and Axl have been isolated in circulating monocytes and tissue macrophages, but not in granulocytes, T and B lymphocytes [27,51,52]. Studies on sections of spleen, lymph nodes, and thymus in mice confirmed that the mRNAs for Tyro3, Axl, and MerTK are abundant in regions populated by macrophages, but are absent in lymphocyte-rich areas [53]. Interesting lessons come from the TAM receptor KO mouse model; despite a normal phenotype of immune system at birth, within the first year of life these animals develop lympho-splenomegaly and aberrant proliferation of active T and B lymphocytes, with diffuse infiltration of tissues [53]. This constitutive activation of the immune system leads to the development of autoimmune manifestations similar to those of several human autoimmune diseases (rheumatoid arthritis, systemic lupus erythematosus, Sjögren’s syndrome, pemphigus vulgaris) and to high serum autoantibody titres [53,54]. Since lymphocytes do not express any of the TAM receptors, the splenomegaly, lymphadenopathy, and lymphocyte hyperactivation seen in TAM KO mice need to be driven by monocytes and macrophages. In fact, these cells show both an increased expression of major histocompatibility complex (MHC) class II and B7 co-receptors and an enhanced production of pro-inflammatory cytokines, including tumor necrosis factor α (TNF-α) and interleukin (IL) 12 [53].

In vitro experiments have shown that the TAM system is up-regulated when antigen-presenting cells (APCs) become activated. Toll-like receptor (TLR) activation induces the expression of Axl receptors through type I interferon (IFN) signalling, leading to suppressor of cytokine signalling proteins (SOCS) 1 and SOCS3 up-regulation, which have a critical role in switching off the inflammatory response in activated dendritic cells [55–57]. Consistent with these findings is the observation that the Gas6/TAM system exerts an anti-inflammatory role [58]. Gas6 is able to suppress IL-1, IL-6, and TNFα expression through the activation of MerTK-PI3K-Akt pathway in TLR-activated monocytes/macrophages, with the down-regulation of NFκB (nuclear factor kappa light chain enhancer of activated B cells) nuclear translocation [31]. Furthermore, the TAM system is involved in the regulation of type 2 immunity. In a house dust mite (HDM) murine model of allergic airway inflammation, HDM-sensitized wild-type (WT) mice developed classical signs of allergic asthma. Interestingly, HDM-sensitized Tyro3−/− mice displayed a larger increase in leukocytes and eosinophils in bronchoalveolar lavage fluid and lung, an increased infiltration of total and effector memory CD4+ T cells in the mediastinal lymph nodes, a higher percentages of CD4+ T cells producing type-2 cytokines and higher serum immunoglobulin E (IgE). This exacerbated type-2 response correlated with the lung histopathological score [59].

A second key feature of the Gas6/TAM system is the regulation of innate immunity through direct involvement in phagocytosis/efferocytosis. Again, this evidence comes from the TAM receptor single and triple mutant mice. MerTK−/− mice display a delayed clearance of apoptotic thymocytes after dexametason stimulation, and the same occurs with the Axl−/− and Tyro3−/− single and double mutants [60–62]. Gas6 recognizes phosphatidylserine (PtdSer) through its amino-terminal Gla domain [63]; this lipid, normally, is expressed on the inner face of the plasma membrane but, during apoptosis, the inactivation of flippases leads to the exposure of PtdSer on the external cell membrane of apoptotic bodies [64,65]. Consequently, Gas6 bridges this lipid with TAM receptors, driving macrophages to the recognition of apoptotic cells and to their subsequent phagocytosis [54,60]. The clearance of apoptotic bodies and the production of pro-inflammatory cytokines are two tightly linked processes; in vitro, apoptotic cells, but not necrotic cells, are able to inhibit the NFκB-mediated production of pro-inflammatory cytokines by dendritic cells. Notably, MerTK binding of apoptotic bodies is required for mediating this effect. MerTK downstream cascade leads to the activation of the PI3K/Akt pathway, which inhibits IKK (IkB kinase); as a consequence, the release of NFκB from the complex with IkB is blocked, preventing its translocation to the nucleus and the transcription of the genes of pro-inflammatory cytokines, including TNF-α [66].
It is, therefore, not surprising that a dysfunction of this system has been linked to the development of autoimmune diseases, since an impaired clearance of apoptotic bodies and an inappropriate inflammatory response are considered critical for the misdirected immune response observed in these conditions.

3. Gas6/TAM System Regulates Survival and Functions of Neuronal and Glial Cells

In recent years a role for Gas6/TAM receptors has been postulated in the regulation of the nervous system. Gas6 is extensively expressed in the CNS [67], suggesting that interactions between Gas6 and its receptors are likely to have physiologically relevant functions [68]. All three TAM receptors are also expressed in the CNS, as reported since 1991 by Lai and Lemke [69], with Tyro3 being the most represented. The Gas6/TAM system, Tyro3 in particular, is relevant to brain development during embryogenesis. In adult mice, Tyro3 is strongly expressed by cerebral cortex and hippocampal neurons [70]; moreover, it is expressed by the amygdala, cerebellum, olfactory bulbs, and gonadotropin-releasing hormone (GnRH) neurons [71]. On the other hand, Axl and MerTK are expressed at low and constant levels during embryogenesis and adult life in mice, mainly in cerebellar and hippocampal neurons [72]; all three TAM receptors are also expressed by glial cells [73] and by endothelial and vascular smooth muscle cells in the CNS [74–76].

Several experiments have disclosed a role of Gas6 in promoting the survival of different neuronal cell types. In vitro, recombinant Gas6 protects hippocampal rat neurons from apoptosis induced by the deprivation of serum [77]. Moreover, Gas6 protects cortical neurons of mice from apoptosis induced by β amyloid protein and phospholipase A2 (PLA2-IIA), inhibiting chromatin condensation and DNA fragmentation. The fact that the cell cultures of these studies contained few non-neuronal cells indicates that Gas6 has a direct neuroprotective effect, not indirectly through supporting cells [78,79]. The anti-apoptotic action of Gas6 has also been described in gonadotropin-releasing hormone (GnRH) secreting neurons from mice, through the ERK cascade and PI3K [80,81]. The Gas6/TAM functional effect on adult neurons remains to be clarified; Tyro3 has been detected in clusters at dendritic, somatic, and axonal levels but, apparently, not in synaptic connections. In view of its distribution, a role in the regulation and integration of synaptic inputs has been hypothesized; furthermore, Tyro3 might help the axonal pathfinding, being expressed by growth cones [70]. Moreover, a role in cell adhesion and cell migration had been previously suggested for Axl [72], which was identified, together with Tyro3, as a factor involved in GnRH neuronal migration along olfactory nerves from their origin in the olfactory placode to the forebrain [80]. Double KO mice for Tyro3 and Axl are characterized by a defective GnRH neuron number and migration which are responsible for impaired sexual function in female mice [71,82].

With reference to neuroglial cells, microarray analyses revealed that transcripts of tyrosine kinase Axl and MerTK receptors are expressed at high levels in isolated oligodendrocytes in the human fetal spinal cord of the second quarter [83]. The latter study also shows that human oligodendrocyte 2′,3′-cyclic nucleotide 3′-phosphodiesterase+ (CNP+) and myelin basic protein+ (MBP+) obtained from fetal spinal cord grown in the presence of recombinant human Gas6 (rhGas6) are protected from apoptosis, and develop more primary processes and arborization compared to those not treated. The effect is mediated by the Axl receptor and—downstream—by PI3K/Akt activation, and is abolished by the soluble receptor Axl-FC [83]. In a later paper by the same group a protective activity of Gas6 on TNFα-mediated cytotoxicity on human oligodendrocytes was shown, with an increase in the survival rate from 18.7% to 64.3%. This effect was Axl-dependent, being completely abrogated in oligodendrocytes derived from Axl KO mice [84]. Additionally, Gas6 stimulates the growth of human Schwann cells, increasing both the number of cells and the incorporation of tritiated thymidine, and has synergic effects with other mitogens; indeed, human Schwann cells express both Axl and MerTK and their phosphorylation is driven by Gas6 [85]. In the mice model of sciatic nerve injury, Axl is overexpressed after the nerve is damaged, again suggesting a role in the survival and protection against apoptosis [85].

Gas6 is able to regulate the inflammatory activity of glial cells, similarly to what is reported in monocytes and macrophages [31]. In 2008, Grommes et al. [86] showed that the treatment of
cultured murine microglial cells with Gas6 significantly reduces the pro-inflammatory response induced by lipopolysaccharide (LPS) stimulation (IL-1β and inducible nitric oxide synthase, iNOS, are significantly down-regulated by Gas6). The Gas6/TAM system has recently been described to be relevant in physiological functions of microglia, the tissue macrophages of the brain and spinal cord. In fact, MerTK−/− and Axl−/− double-KO mice are characterized by impairment in the clearance of apoptotic bodies, reduced motility of microglial cells, and delayed recruitment to sites of brain injury; moreover, both Gas6 and ProS serve as ligands in this process [87,88].

4. Evidence about the Role of the Gas6/TAM System in Multiple Sclerosis: Lessons from Animal Models and Human Studies

Multiple sclerosis (MS) is an immune-mediated disorder of the CNS. The complex interactions between adaptive and innate immunity determine an inflammatory aggression to the myelin of neuronal fibers. In this context, macrophages and microglial cells are involved in myelin degradation and in oligodendrocyte loss by producing proinflammatory cytokines [89]. Therefore, systems involved in dampening macrophages activation are promising targets for studies addressing MS pathogenesis. Furthermore, in the animal model of myelin oligodendrocyte glycoprotein (MOG)-induced experimental allergic encephalomyelitis (EAE) an increased apoptosis in lymphoid organs, as well as the injection of apoptotic bodies, could worsen the disease course from a relapsing-remitting clinical pattern to a more severe secondary progressive course. However, while the underlying mechanisms have not been fully elucidated, an increase in anti-MOG antibody is observed under these circumstances [90]. Hence, impaired apoptosis also seems of significance in MS pathogenesis as it occurs for other systemic autoimmune diseases, such as systemic lupus erythematosus [91].

As mentioned before, the Gas6/TAM system has been linked to the development of autoimmunity, in the pathogenesis of which an impaired clearance of apoptotic bodies and an inappropriate inflammatory response by macrophages and dendritic cells are considered critical [54]. Additionally, the clearance of cellular and myelin debris after inflammatory demyelination might be an initial and important early step for the recovery of damaged myelin fibers; it turned out that the Gas6/TAM system is also a relevant system for both neuron and glial cell survival, including specialized cells involved in myelination processes of the CNS [83]. Thus, a role of this system in MS pathogenesis seems possible either via control of inflammation, or through the regulation of the myelination process, or both.

Again, the strongest evidence of Gas6/TAM system involvement in MS pathogenesis come from animal models, in particular those in which either Gas6 or TAM receptors are knocked down. Both the cuprizone demyelination and the EAE models have been used (Table 1 and Figure 2).

Cuprizone (bis-cyclohexanone-oxaldihydrazone) induces a toxic demyelination without altering the blood/brain barrier; it determines the loss of oligodendrocytes and microglial/macrophage accumulation in the damaged tissues. This model allows the study of myelin damage and repair, without the confounding factors of the intense inflammation present in mice with EAE [92,93]. In mice undergoing cuprizone challenge, a change in TAM and Gas6 expression occurs, Tyro3 is down-regulated, while Gas6, Axl, and MerTK transcription are enhanced, paralleling microglial activation. Gas6−/− mice, under the same conditions, show a more severe demyelination, a greater reduction in oligodendrocytes number, and an overactivation of microglia [94]. A further study confirmed a possible relevant role of Gas6 in myelin repair processes; in fact, Gas6−/− mice had a delayed remyelination at four weeks after cuprizone discontinuation with respect to WT, along with a reduction of oligodendrocytes. These differences, however, disappeared after 10 weeks; additionally, Gas6 significantly increased remyelination in vitro, in a dose-dependent manner [95]. Others obtained similar results. In fact, injection of rhGas6 into the CNS improved the recovery from damage after cuprizone withdrawal, with a beneficial effect on the clearance of cellular and myelin debris, enhancement of remyelination and of maturation of oligodendrocyte progenitor cells, and an increase in the number of myelinated axons [96]. The effect of Gas6 described above is mediated, at least in part, by the Axl receptor, since the clearance of damaged cells and of myelin debris, which likely impacts
upon remyelination and cell survival, is impaired in Axl\(^{-/-}\) mice. These animals have a delayed clearance of apoptotic oligodendrocytes and of myelin debris with deferred recovery from cuprizone demyelination [97]. Other in vitro and in vivo experiments link MerTK to the phagocytosis of apoptotic debris in the CNS [79,88,98].

Altogether, these data from the cuprizone mice model indicate that the Gas6/TAM receptors’ interaction is important, both during demyelination and remyelination, independently of the effects on inflammation. This system favours myelin repair after damage directly, by enhancing the clearance of cellular and myelin debris and through the support to oligodendrocyte survival and myelin restoration.

Further evidence to support Gas6/TAM involvement in the pathogenesis of MS come from the EAE model. The induction of EAE with MOG administration damages the blood-brain barrier, resulting in the infiltration of T cells and monocytes with a severe inflammation, expression of pro-inflammatory molecules, demyelination, and axonal damage. This model creates an inflammatory demyelination process similar to what happens in MS and allows studying the role of the Gas6/TAM system in neuroinflammation [92].

After EAE induction, Gas6, Axl, and MerTK RNA expression (but not Tyro3 or ProS) are significantly increased in the lumbar spinal cord; the direct intracerebral delivery of Gas6 is protective, with evidence of less demyelination and/or enhanced remyelination relative to controls. When EAE was induced in Gas6\(^{-/-}\) mice, worse clinical scores and delayed recovery from damage were observed, and the inflammation in the spinal cord was more severe, with greater expression of pro-inflammatory molecules and a significantly increased infiltration of macrophages [99]. These data fill the gap of the previously mentioned experiments with the cuprizone model, suggesting that Gas6 is relevant in both limiting inflammatory demyelination and favouring recovery. Consistent with this hypothesis, Axl\(^{-/-}\) mice are characterized by a more severe course of EAE than wild-type (WT) mice. Specifically, these mice develop worse spinal cord lesions with larger infiltrates, more demyelination, and more axonal damage. This is associated with larger amounts of pro-inflammatory cytokines and chemokines, such as TNF\(\alpha\), monocyte chemoattractant protein 1 (MCP1) and CCL5/regulated on activation, normal T cell expressed and secreted (RANTES) in the spinal cord [100]. Of note, Axl\(^{-/-}\) mice had a strikingly impaired clearance of myelin debris by microglia/macrophages [100].

In conclusion, the data from animal models of MS altogether indicate that the Gas6/TAM system is relevant both in dampening inflammatory demyelination and in supporting myelin repair. The Axl receptor seems to be the principal effector of these actions. Thus, the Gas6/Axl interaction may be a promising target of anti-inflammatory, neuroprotective, and promyelinating treatments.

To date, very few studies replicated the above evidences in patients with MS. In 2009 Weinger et al., [101] in an autopsy study on MS patients, reported an up-regulation of sAxl and sMer in homogenates derived from chronic silent and chronic active lesions, respectively. Conversely, the full-length form of these receptors was not upregulated; intra-lesion glial cells are responsible of this expression. In normal tissue homogenates, Gas6 positively correlates with sAxl and sMer; on the contrary, in chronic active and silent lesions, Gas6 correlates inversely with sAxl and sMer. However, it is not known whether the low Gas6 concentration is due to the decoy action of sAxl and sMer leading to ligand removal or, alternatively, Gas6 secretion is reduced and, therefore, receptor cleavage enhances in the attempt to eliminate the excess of membrane-bound receptors and to restore the homeostatic ligand-to-receptor ratio. Axl and MerTK are solubilised by two metalloproteinases also called ADAM10 (a disintegrin and metalloproteinase 10) and ADAM17 (see Figure 1); in established MS lesions, the expression of these enzymes is up-regulated [101]. The shift from a positive correlation in normal tissue to an inverse correlation in established MS lesions could impair Gas6/TAM activity, affecting the functions of the system on immune response and on cell debris clearance, and favouring, in turn, a chronic demyelination environment in spite of ongoing remyelination and repair [101].

Similar observations have been reported in other human autoimmune diseases, such as juvenile systemic lupus erythematosus, where an impairment of the physiological balance between
transmembrane and soluble TAM has been described. This might justify a complex derangement of the system, leading to a deficient phagocytosis and persistent inflammatory response [102].

A further clue of the association between Gas6/TAM and MS in humans has been provided by genome-wide association studies (GWAS). Several single nucleotide polymorphisms (SNPs) within the MerTK gene are associated with susceptibility to MS [103]. This finding was later confirmed in a large cohort of 1140 MS cases and 1140 healthy controls using a candidate gene approach. The authors identified 12 intronic SNPs related to MS susceptibility, in strong linkage disequilibrium with each other [104]. Recently, the same group reported that one specific variant of MerTK gene, so called rs7422195, has discordant association to MS according to HLA (human leukocyte antigen)-DRB1*15:01 status, being protective in DR15 homozygosity and favouring the disease in the absence of DR15. The minor allele of rs7422195 is also associated to an increased gene and protein expression of MerTK in monocytes and CD4+ cells [105]. Whether this subset of cells was formed by T cells or contaminated by monocytes expressing CD4 is not yet known. In any case, a recent study assessed the transcriptomic modification that Th17 CD4+ T cells undergo, derived from mice following induction of EAE. Interestingly, MerTK is among the genes overexpressed upon EAE induction [106].

Very limited evidence comes from clinical studies; our group evaluated both cerebrospinal fluid (CSF) and plasma concentration of Gas6 of MS patients by an enzyme-linked immunosorbent assay (ELISA) validated for human use and tested in other diseases and in CSF [107–109]. Sixty-five consecutive patients with clinically-isolated syndrome (CIS) or MS who underwent a spinal tap to confirm MS diagnosis were evaluated in relation to 45 controls affected by a non-inflammatory neurological disease. All MS patients were sampled during a relapse. The score for each functional system (FS) and the total expanded disability status scale (EDSS) score were calculated at onset, at maximum worsening, and at the first examination after the day of maximum improvement of the relapse. Relapse duration, severity, number of FS involved, and relapse recovery were obtained. We observed that patients with MS do not have substantial alterations of plasma Gas6 concentration with respect to controls, but a CSF/plasma dissociation was observed as being CSF Gas6, significantly higher with respect to other non-inflammatory neurologic diseases. Interestingly, those patients suffering a more severe or longer relapse or with more FS had a lower CSF Gas6 concentration, no different from controls, in comparison to those showing shorter and milder relapses and with fewer FS, who had significantly higher concentrations (nearly 2×). On the other hand, CSF Gas6 concentration did not change according to the completeness of recovery. Finally, neither plasma nor CSF Gas6 were related to the relapse rate or EDSS progression in a follow up cohort [110]. These findings fit with experimental evidences, according to which Gas6 is induced during demyelination in MS murine models and probably acts with a protective role in dampening neuroinflammation and in favouring myelin repair, and are in line with the report from Weinger et al., showing that Gas6 expression was very low in chronic MS lesions [101].

Table 1. Summary of the current in vivo studies supporting a role for the Gas6/TAM system in multiple sclerosis.

| Author          | Year | Main Findings                                                                 |
|-----------------|------|-------------------------------------------------------------------------------|
| Hoehn et al.    | 2008 | The deletion of Axl is associated with a delayed recovery and prolonged axonal damage following cuprizone toxicity |
| Binder et al.   | 2008 | Gas6, Axl and MerTK are upregulated upon cuprizone-induced demyelination; Gas6 knockout (KO) mice have more severe demyelination |
| Weinger et al.  | 2009 | In chronic multiple sclerosis (MS) lesions sAxl and sMer are upregulated and inversely related to cerebrospinal fluid (CSF) Gas6 concentration |
| Tsiperson et al.| 2010 | Gas6 stimulates remyelination following cuprizone toxicity                     |
| Ma et al.       | 2011 | SNPs in MerTK gene confer susceptibility to MS                                  |
| Binder et al.   | 2011 | Gas6 KO mice show a defective remyelination, after cuprizone-induced demyelination, which can be corrected by administering exogenous Gas6 |
Evidence comes from human studies. However, the fact that Gas6 is scarcely expressed and sAxl and Axl have a pro-phagocytic action, mediated by both Axl and MerTK. Fewer, but consistent, inflammatory effect on microglia, and a trophic effect on oligodendrocytes mediated by Axl have been observed. Demyelination, likely as the result of multiple mechanisms: a neurotrophic effect, an anti-inflammatory effect on T cells, and an anti-apoptotic effect on oligodendrocytes.

Table 1. Continued.

| Author                        | Year | Main Findings                                                                 |
|-------------------------------|------|-------------------------------------------------------------------------------|
| ANZgene consortium [103]      | 2011 | Many SNPs of MerTK gene are associated to MS risk in a genome wide association study |
| Weinger et al. [100]          | 2011 | Axl KO murine models of experimental allergic encephalomyelitis (EAE) are characterized by a more severe phenotype than wild type mice |
| Sainaghi et al. [110]         | 2013 | Gas6 CSF concentration is higher in patients with shorter and less severe MS flares |
| Gruber et al. [99]            | 2014 | Intracerebral delivery of Gas6 protects against damage in EAE |
| Hoppmann et al. [106]         | 2015 | CD4+ T cells from EAE mice show an up-regulation of Gas6 and MerTK |
| Binder et al. [105]           | 2016 | SNPs in MerTK can protect or confer risk of MS on the basis of HLA-DRB1*15:01 |

Figure 2. Animal models of MS involving the growth arrest specific 6 (Gas6)/TAM (Tyro3, Axl and Mer) receptors system. On the left are summarized data obtained in the experimental allergic encephalomyelitis (EAE) model, and on the right those coming from the cuprizone challenge model, in Axl−/−, Gas6−/−, or wild-type (WT) mice, respectively. See text for further details. ↑ enhancement, ↓ reduction, ↔ no effect.

The important functions exerted in human biology by TAM receptors and their ligand Gas6, Axl and MerTK receptors system. On the left are those coming from the cuprizone challenge model, in Axl−/−, Gas6−/−, or wild-type (WT) mice, respectively. See text for further details. ↑ enhancement, ↓ reduction, ↔ no effect.
5. Conclusions

The important functions exerted in human biology by TAM receptors and their ligand Gas6, including cell growth regulation, inflammation, and clearance of apoptotic bodies, make this relatively novel system a promising target in different pathological conditions, especially if immune-driven [58]. Specifically, Gas6 and TAM seem to play a protective role against inflammatory demyelination, likely as the result of multiple mechanisms: a neurotrophic effect [24], an anti-inflammatory effect on microglia, and [86] a trophic effect on oligodendrocytes mediated by Axl [83,84], and a pro-phagocytic action, mediated by both Axl and MerTK [61]. Fewer, but consistent, evidence comes from human studies. However, the fact that Gas6 is scarcely expressed and sAxl and sMer decoy receptors are overexpressed in chronic MS lesions [101], and that the higher the CSF Gas6 concentration is, the milder the clinical MS relapse phenotype [110], are strong suggestions that activation of the TAM system in this setting is beneficial. The mechanisms by which Gas6 might exert this protective action remain highly speculative; however, dysregulation of TAM cleavage is a hypothesis worthy of further testing. Indeed, a deeper understanding of the Gas6/RAM system may contribute to elucidation of MS pathogenesis and, possibly, to give us new therapeutic tools.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Schneider, C.; King, R.M.; Philipson, L. Genes specifically expressed at growth arrest of mammalian cells. Cell 1988, 54, 787–793. [CrossRef]
2. Manfioletti, G.; Brancolini, C.; Avanzi, G.; Schneider, C. The protein encoded by a growth arrest-specific gene (Gas6) is a new member of the vitamin K-dependent proteins related to protein S, a negative coregulator in the blood coagulation cascade. Mol. Cell. Biol. 1993, 13, 4976–4985. [CrossRef] [PubMed]
3. Avanzi, G.C.; Gallicchio, M.; Cavalloni, G.; Gammaitoni, L.; Leone, F.; Rosina, A.; Boldorini, R.; Monga, G.; Pegraro, L.; Varnum, B.; et al. Gas6, the ligand of Axl and Rse receptors, is expressed in hematopoietic tissue but lacks mitogenic activity. Exp. Hematol. 1997, 25, 1219–1226. [PubMed]
4. Melaragno, M.G.; Wuthrich, D.A.; Poppa, V.; Gill, D.; Lindner, V.; Berk, B.C.; Corson, M.A. Increased expression of Axl tyrosine kinase after vascular injury and regulation by G protein-coupled receptor agonists in rats. Circ. Res. 1998, 83, 697–704. [CrossRef] [PubMed]
5. Joseph, D.R. Sequence and functional relationships between androgen-binding protein/sex hormone binding globulin and its homologs protein S, Gas6, laminin, and agrin. Steroids 1997, 62, 578–588. [CrossRef]
6. Stitt, T.N.; Conn, G.; Gore, M.; Lai, C.; Bruno, J.; Radziejewski, C.; Mattsson, K.; Fisher, J.; Gies, D.R.; Jones, P.F. The anticoagulation factor protein S and its relative, Gas6, are ligands for the Tyro 3/Axl family of receptor tyrosine kinases. Cell 1995, 80, 661–670. [CrossRef]
7. Varnum, B.C.; Young, C.; Elliott, G.; Garcia, A.; Bartley, T.D.; Fridell, Y.W.; Hunt, R.W.; Trail, G.; Clogston, C.; Toso, R.J. Axl receptor tyrosine kinase stimulated by the vitamin K-dependent protein encoded by growth-arrest-specific gene 6. Nature 1995, 373, 623–626. [CrossRef] [PubMed]
8. Nagata, K.; Ohashi, K.; Nakano, T.; Arita, H.; Zong, C.; Hanafusa, H.; Mizuno, K. Identification of the product of growth arrest-specific gene 6 as a common ligand for Axl, Sky, and Mer receptor tyrosine kinases. J. Biol. Chem. 1996, 271, 30022–30027. [CrossRef] [PubMed]
9. Heiring, C.; Dahlback, B.; Muller, Y.A. Ligand recognition and homophilic interactions in Tyro3: Structural insights into the Axl/Tyro3 receptor tyrosine kinase family. J. Biol. Chem. 2004, 279, 6952–6958. [CrossRef] [PubMed]
10. Braunger, J.; Schleithoff, L.; Schulz, A.S.; Kessler, H.; Lammers, R.; Ulrich, A.; Bartram, C.R.; Janssen, J.W. Intracellular signaling of the Ufo/Axl receptor tyrosine kinase is mediated mainly by a multi-substrate docking-site. Oncogene 1997, 14, 2619–2631. [CrossRef] [PubMed]
11. Goruppi, S.; Russo, E.; Varnum, B.; Schneider, C. Requirement of phosphatidylinositol 3-kinase-dependent pathway and Src for Gas6-Axl mitogenic and survival activities in NIH 3T3 fibroblasts. Mol. Cell. Biol. 1997, 17, 4442–4453. [CrossRef] [PubMed]
12. Keating, A.K.; Kim, G.K.; Jones, A.E.; Donson, A.M.; Ware, K.; Mulcahy, J.M.; Salzberg, D.B.; Foreman, N.K.; Liang, X.; Thorburn, A.; et al. Inhibition of Mer and Axl receptor tyrosine kinases in astrocytoma cells leads to increased apoptosis and improved chemosensitivity. *Mol. Cancer Ther.* 2010, 9, 1298–1307. [CrossRef] [PubMed]

13. O’Brien, J.P.; Fridell, Y.W.; Koski, R.; Varnum, B.; Liu, E.T. The transforming receptor tyrosine kinase, Axl, is post-translationally regulated by proteolytic cleavage. *J. Biol. Chem.* 1995, 270, 551–557. [CrossRef] [PubMed]

14. Thorp, E.; Vaisar, T.; Subramanian, M.; Mautner, L.; Blobel, C.; Tabas, I. Shedding of the Mer tyrosine kinase receptor is mediated by ADAM17 protein through a pathway involving reactive oxygen species, protein kinase Cδ, and p38 mitogen-activated protein kinase (MAPK). *J. Biol. Chem.* 2011, 286, 33335–33344. [CrossRef] [PubMed]

15. Sather, S.; Kenyon, K.D.; Lefkowitz, J.B.; Liang, X.; Varnum, B.C.; Henson, P.M.; Graham, D.K. A soluble form of the Mer receptor tyrosine kinase inhibits macrophage clearance of apoptotic cells and platelet aggregation. *Blood* 2007, 109, 1026–1033. [CrossRef] [PubMed]

16. Prieto, A.L.; Weber, J.L.; Lai, C. Expression of the receptor protein-tyrosine kinases Tyro-3, Axl, and Mer in the developing rat central nervous system. *J. Comp. Neurol.* 2000, 425, 295–314. [CrossRef]

17. Mark, M.R.; Scadden, D.T.; Wang, Z.; Gu, Q.; Goddard, A.; Godowski, P.J. Rse, a novel receptor-type tyrosine kinase with homology to Axl/Ufo, is expressed at high levels in the brain. *J. Biol. Chem.* 1994, 269, 10720–10728. [PubMed]

18. Angelillo-Scherrer, A.; de Frutos, P.; Aparicio, C.; Melis, E.; Savi, P.; Lupu, F.; Arnout, J.; Dewerchin, M.; Snodgrass, H.R.; Huhn, D. Expression of Axl, a transforming receptor tyrosine kinase, in normal and malignant hematopoiesis. *Circ. Res.* 2001, 88, 1423–1432. [CrossRef] [PubMed]

19. Higuchi, Y.; Kubota, T.; Koyanagi, M.; Maeda, T.; Feldman, A.M.; Makino, N. Upregulation of anticoagulant proteins, protein S and tissue factor pathway inhibitor, in the mouse myocardium with cardio-specific TNF-α overexpression. *Am. J. Physiol. Heart Circ. Physiol.* 2012, 302, H2352–H2362. [CrossRef] [PubMed]

20. Matsubara, N.; Takahashi, Y.; Nishina, Y.; Mukouyama, Y.; Yanagisawa, M.; Watanabe, T.; Nakano, T.; Nomura, K.; Arita, H.; Nishimune, Y.; et al. A receptor tyrosine kinase, Sky, and its ligand Gas 6 are expressed in gonads and support primordial germ cell growth or survival in culture. *Dev. Biol.* 1996, 180, 499–510. [CrossRef] [PubMed]

21. Taylor, I.C.; Roy, S.; Yaswen, P.; Stampfer, M.R.; Varmus, H.E. Mouse mammary tumors express elevated levels of RNA encoding the murine homology of SKY, a putative receptor tyrosine kinase. *J. Biol. Chem.* 1995, 270, 6872–6880. [PubMed]

22. Katagiri, M.; Hakeda, Y.; Chikazu, D.; Ogasawara, T.; Takato, T.; Kumeegawa, M.; Nakamura, K.; Kawaguchi, H. Mechanism of stimulation of osteoclastic bone resorption through Gas6/Tyro3, a receptor tyrosine kinase signaling, in mouse osteoclasts. *J. Biol. Chem.* 2001, 276, 7376–7382. [CrossRef] [PubMed]

23. Prasad, D.; Rothlin, C.V.; Burrola, P.; Burstyn-Cohen, T.; Lu, Q.; de Frutos, P.G.; Lemke, G. TAM receptor function in the retinal pigment epithelium. *Mol. Cell. Neurosci.* 2006, 33, 96–108. [CrossRef] [PubMed]

24. Ji, R.; Meng, L.; Jiang, X.; Cvm, N.K.; Ding, J.; Li, Q.; Lu, Q. TAM receptors support neural stem cell survival, proliferation and neuronal differentiation. *PLoS ONE* 2014, 9, e115140. [CrossRef] [PubMed]

25. Lafdil, F.; Chobert, M.N.; Couchie, D.; Brouillet, A.; Zafrani, E.S.; Davier, P.; Lapierce, Y. Induction of Gas6 protein in CCl4-induced rat liver injury and anti-apoptotic effect on hepatic stellate cells. *Hepatology* 2006, 44, 228–239. [CrossRef] [PubMed]

26. Yanagita, M.; Arai, H.; Ishii, K.; Nakano, T.; Ohashi, K.; Mizuno, K.; Varmum, B.; Fukatsu, A.; Doi, T.; Kita, T. Gas6 regulates mesangial cell proliferation through Axl in experimental glomerulonephritis. *Am. J. Pathol.* 2001, 158, 1423–1432. [CrossRef]

27. Neubauer, A.; Fiebelers, A.; Graham, D.K.; O’Brien, J.P.; Schmidt, C.A.; Barckow, P.; Serke, S.; Siegent, W.; Snodgrass, H.R.; Huhn, D. Expression of Axl, a transforming receptor tyrosine kinase, in normal and malignant hematopoiesis. *Blood* 1994, 84, 1931–1941. [PubMed]

28. Hasanbasic, I.; Cuerquis, I.; Varnum, B.; Blostein, M.D. Intracellular signaling pathways involved in Gas6-Axl-mediated survival of endothelial cells. *Am. J. Physiol. Heart Circ. Physiol.* 2004, 287, 1207–1213. [CrossRef] [PubMed]

29. Collett, G.D.; Sage, A.P.; Kirton, J.P.; Alexander, M.Y.; Gilmore, A.P.; Canfield, A.E. Axl/phosphatidylinositol 3-kinase signaling inhibits mineral deposition by vascular smooth muscle cells. *Circ. Res.* 2007, 100, 502–509. [CrossRef] [PubMed]
30. Zizzo, G.; Hilliard, B.A.; Monestier, M.; Cohen, P.L. Efficient clearance of early apoptotic cells by human macrophages requires M2c polarization and MerTK induction. *J. Immunol.* 2012, 189, 3508–3520. [CrossRef] [PubMed]

31. Alciato, F.; Sainaghi, P.P.; Sola, D.; Castello, L.; Avanzi, G.C. TNF-α, IL-6, and IL-1 expression is inhibited by GAS6 in monocytes/macrophages. *J. Leukoc. Biol.* 2010, 87, 869–875. [CrossRef] [PubMed]

32. Chen, C.; Li, Q.; Darrow, A.L.; Wang, Y.; Derian, C.K.; Yang, J.; de Garavilla, L.; Andrade-Gordon, P.; Damiano, B.P. Mer receptor tyrosine kinase signaling participates in platelet function. *Atheroscler. Thromb. Vasc. Biol.* 2004, 24, 1118–1123. [CrossRef] [PubMed]

33. Wang, H.; Chen, Y.; Ge, Y.; Ma, P.; Ma, Q.; Ma, J.; Wang, H.; Xue, S.; Han, D. Immunoexpression of Tyro 3 family receptors—Tyro 3, Axl, and Mer—and their ligand Gas6 in postnatal developing mouse testis. *J. Histochem. Cytochem.* 2005, 53, 1355–1364. [CrossRef] [PubMed]

34. Wu, Y.M.; Robinson, D.R.; Kung, H.J. Signal pathways in up-regulation of chemokines by tyrosine kinase MER/NYK in prostate cancer cells. *Cancer Res.* 2004, 64, 7311–7320. [CrossRef] [PubMed]

35. Xie, S.; Li, Y.; Li, X.; Wang, L.; Yang, N.; Wang, Y.; Wei, H. Mer receptor tyrosine kinase is frequently overexpressed in human non-small cell lung cancer, confirming resistance to erlotinib. *Oncotarget* 2015, 6, 9206–9219. [CrossRef] [PubMed]

36. Shao, W.H.; Zhen, Y.; Rosenbaum, J.; Eisenberg, R.A.; McGaha, T.L.; Birkenbach, M.; Cohen, P.L. A protective role of Mer receptor tyrosine kinase in nephrotoxic serum-induced nephritis. *Clin. Immunol.* 2010, 136, 236–244. [CrossRef] [PubMed]

37. Sainaghi, P.P.; Castello, L.; Bergamasco, L.; Galletti, M.; Bellosta, P.; Avanzi, G.C. Gas6 induces proliferation in prostate carcinoma cell lines expressing the Axl receptor. *J. Cell. Physiol.* 2005, 204, 36–44. [CrossRef] [PubMed]

38. May, C.D.; Garnett, J.; Ma, X.; Landers, S.M.; Ingram, D.R.; Demico, E.G.; Al Sannaa, G.A.; Vu, T.; Han, L.; Zhang, Y.; et al. AXL is a potential therapeutic target in dedifferentiated and pleomorphic liposarcomas. *BMC Cancer* 2015, 15, 901. [CrossRef] [PubMed]

39. Han, J.; Tian, R.; Yong, B.; Luo, C.; Tan, P.; Shen, J.; Peng, T. Gas6/Axl mediates tumor cell apoptosis, migration and invasion and predicts the clinical outcome of osteosarcoma patients. *Biochem. Biophys. Res. Commun.* 2013, 435, 493–500. [CrossRef] [PubMed]

40. Schlegel, J.; Sambade, M.J.; Sather, S.; Moschos, S.J.; Tan, A.C.; Winges, A.; DeRyckere, D.; Carson, C.C.; Trembath, D.G.; Tentler, J.J. MerTK receptor tyrosine kinase is a therapeutic target in melanoma. *J. Clin. Investig.* 2013, 123, 2257–2267. [CrossRef] [PubMed]

41. Lee, H.J.; Jeng, Y.M.; Chen, Y.L.; Chung, L.; Yuan, R.H. Gas6/Axl pathway promotes tumor invasion through the transcriptional activation of Slug in hepatocellular carcinoma. *Carcinogenesis* 2014, 35, 769–775. [CrossRef] [PubMed]

42. Lee, C.H.; Yen, C.Y.; Liu, S.Y.; Chen, C.K.; Chiang, C.F.; Shah, S.G.; Chen, P.H.; Shieh, Y.S. Axl is a prognostic marker in oral squamous cell carcinoma. *Ann. Surg. Oncol.* 2012, 19, S500–S508. [CrossRef] [PubMed]

43. Jiang, T.; Liu, G.; Wang, L.; Liu, H. Elevated Serum Gas6 Is a Novel Prognostic Biomarker in Patients with Oral Squamous Cell Carcinoma. *PLoS ONE* 2015, 10, e0133940. [CrossRef] [PubMed]
48. Gjertdum, C.; Tiron, C.; Hailby, T; Stefansson, I.; Haugen, H.; Sandal, T.; Collett, K.; Li, S.; McCormack, E.; Gjertsd, B.T.; et al. Axol is an essential epithelial-to-mesenchymal transition-induced regulator of breast cancer metastasis and patient survival. Proc. Natl. Acad. Sci. USA 2010, 107, 1124–1129. [CrossRef] [PubMed]

49. Walker, F.J. Regulation of activated protein C by a new protein. A possible function for bovine protein S. J. Biol. Chem. 1980, 255, 5521–5524. [PubMed]

50. Van der Meer, J.H.; van der Poll, T.; van’t Veer, C. TAM receptors, Gas6, and protein S: Roles in inflammation and hemostasis. Blood 2014, 123, 2460–2469. [CrossRef] [PubMed]

51. Graham, D.K.; Dawson, T.L.; Mullaney, D.L.; Snodgrass, H.R.; Earp, H.S.; Matsushima, G.K. Macrophages and dendritic cells use different Axl/MerTK/Tyro3 receptors in clearance of apoptotic cells. J. Immunol. 2007, 178, 5635–5642. [CrossRef] [PubMed]

52. Neubauer, A.; Burchert, A.; Maiwald, C.; Grus, H.J.; Serke, S.; Huhn, D.; Wittig, B.; Liu, E. Recent progress on the role of Axl, a receptor tyrosine kinase, in malignant transformation of myeloid leukemias. Leuk. Lymphoma 1997, 25, 91–96. [CrossRef] [PubMed]

53. Lu, Q.; Lemke, G. Homeostatic regulation of the immune system by receptor tyrosine kinases of the Tyro 3 family. Science 2001, 293, 306–311. [CrossRef] [PubMed]

54. Rothlin, C.V.; Ghosh, S.; Zuniga, E.I.; Oldstone, M.B.; Lemke, G. TAM receptors are pleiotropic inhibitors of the innate immune response. Cell 2007, 131, 1124–1136. [CrossRef] [PubMed]

55. Lu, Q.; Lemke, G. Homeostatic regulation of the immune system by receptor tyrosine kinases of the Tyro 3 family. Curr. Opin. Immunol. 2003, 15, 31–36. [CrossRef]

56. Sharif, M.N.; Sosic, D.; Rothlin, C.V.; Kelly, E.; Lemke, G.; Olson, E.N.; Iwashkiv, L.B. Twist mediates suppression of inflammation by type I IFNs and Axl. J. Exp. Med. 2006, 203, 1891–1901. [CrossRef] [PubMed]

57. Scutera, S.; Froni, T.; Musso, T.; Pierobon, D.; Orinska, Z.; Paus, R.; Builone-Paus, S.; Giovarelli, M. Survival and migration of human dendritic cells are regulated by an IFN-α-inducible Axl/Gas6 pathway. J. Immunol. 2009, 183, 3004–3013. [CrossRef] [PubMed]

58. Rothlin, C.V.; Carrera-Silva, E.A.; Bosurgi, L.; Ghosh, S. TAM receptor signaling in immune homeostasis. Annu. Rev. Immunol. 2015, 33, 355–391. [CrossRef] [PubMed]

59. Chan, P.Y.; Carrera Silva, E.A.; de Kouchkovsky, D.; Joanna, L.D.; Hao, L.; Hu, D.; Huntsman, S.; Eng, C.; Licona-Limón, P.; Weinstein, J.S.; et al. The TAM family receptor tyrosine kinase TYRO3 is a negative regulator of type 2 immunity. Science 2016, 352, 99–103. [CrossRef] [PubMed]

60. Scott, R.S.; McMahon, E.J.; Pop, S.M.; Reap, E.A.; Caricchio, R.; Cohen, P.L.; Earp, H.S.; Matsushima, G.K. Phagocytosis and clearance of apoptotic cells is mediated by MER. Nature 2001, 411, 207–211. [CrossRef] [PubMed]

61. Seitz, H.M.; Camenisch, T.D.; Lemke, G.; Earp, H.S.; Matsushima, G.K. Macrophages and dendritic cells use different Axl/MerTK/Tyro3 receptors in clearance of apoptotic cells. J. Immunol. 2007, 178, 5635–5642. [CrossRef] [PubMed]

62. Cohen, P.L.; Caricchio, R.; Abraham, V.; Camenisch, T.D.; Jennette, J.C.; Roubey, R.A.; Earp, H.S.; Matsushima, G.; Reap, E.A. Delayed apoptotic cell clearance and lupus-like autoimmunity in mice lacking the c-mer membrane tyrosine kinase. J. Exp. Med. 2002, 196, 135–140. [CrossRef] [PubMed]

63. Nakano, T.; Ishimoto, Y.; Kishino, J.; Umeda, M.; Inoue, K.; Nagata, K.; Ohashi, K.; Mizuno, K.; Arita, H. Cell adhesion to phosphatidylserine mediated by a product of growth arrest-specific gene 6. J. Biol. Chem. 1997, 272, 29411–29414. [CrossRef] [PubMed]

64. Daleke, D.L. Regulation of transbilayer plasma membrane phospholipid asymmetry. J. Lipid Res. 2003, 44, 233–242. [CrossRef] [PubMed]

65. Krahling, S.; Callahan, M.K.; Williamson, P.; Schlegel, R.A. Exposure of phosphatidylserine is a general feature in the phagocytosis of apoptotic lymphocytes by macrophages. Cell Death Differ. 1999, 6, 183–189. [CrossRef] [PubMed]

66. Sen, P.; Wallet, M.A.; Yi, Z.; Huang, Y.; Henderson, M.; Mathews, C.E.; Earp, H.S.; Matsushima, G.; Baldwin, A.S., Jr.; Tisch, R.M. Apoptotic cells induce Mer tyrosine kinase-dependent blockade of NF-κB activation in dendritic cells. Blood 2007, 109, 653–660. [CrossRef] [PubMed]

67. Prieto, A.L.; Weber, J.L.; Tracy, S.; Heeb, M.J.; Lai, C. Gas6, a ligand for the receptor protein-tyrosine kinase Tyro-3, is widely expressed in the central nervous system. Brain Res. 1999, 816, 646–661. [CrossRef]
Lai, C.; Lemke, G. An extended family of protein-tyrosine kinase genes differentially expressed in the vertebrate nervous system. *Neuron* 1991, 6, 691–704. [CrossRef]

Prieto, A.L.; O’Dell, S.; Varum, B.; Lai, C. Localization and signaling of the receptor protein tyrosine kinase Tyro3 in cortical and hippocampal neurons. *Neuroscience* 2007, 150, 319–334. [CrossRef] [PubMed]

Pierce, A.; Bliesner, B.; Xu, M.; Nielsen-Preiss, S.; Lemke, G.; Tobet, S.; Wierman, M.E. Axl and Tyro3 modulate female reproduction by influencing gonadotropin-releasing hormone neuron survival and migration. *Mol. Endocrinol.* 2008, 22, 2481–2495. [CrossRef] [PubMed]

Bellosta, P.; Costa, M.; Lin, D.A.; Basilico, C. The receptor tyrosine kinase ARK mediates cell aggregation by homophilic binding. *Mol. Cell. Biol.* 1995, 15, 614–625. [CrossRef] [PubMed]

Cahoy, J.D.; Emery, B.; Kaushal, A.; Foo, L.C.; Zamanian, J.L.; Christopherson, K.S.; Xing, Y.; Lubischer, J.L.; Krieg, P.A.; Krupenko, S.A.; et al. A transcriptome database for astrocytes, neurons, and oligodendrocytes: A new resource for understanding brain development and function. *J. Neurosci.* 2008, 28, 264–278. [CrossRef] [PubMed]

Zhu, D.; Wang, Y.; Singh, I.; Bell, R.D.; Deane, R.; Zhong, Z.; Sagare, A.; Winkler, E.A.; Zlokovic, B.V. Protein S controls hypoxic/ischemic blood-brain barrier disruption through the TAM receptor Tyro3 and sphingosine 1-phosphate receptor. *Blood* 2010, 115, 4963–4972. [CrossRef] [PubMed]

Korshunov, V.A.; Mohan, A.M.; Georger, M.A.; Berk, B.C. Axl, a receptor tyrosine kinase, mediates flow-induced vascular remodeling. *Circ. Res.* 2006, 98, 1446–1452. [CrossRef] [PubMed]

Melaragno, M.G.; Cavet, M.E.; Yan, C.; Tai, L.K.; Jin, Z.G.; Haendeler, J.; Berk, B.C. Gas6 inhibits apoptosis in vascular smooth muscle: Role of Axl kinase and Akt. *J. Mol. Cell. Cardiol.* 2004, 37, 881–887. [CrossRef] [PubMed]

Funakoshi, H.; Yonemasu, T.; Nakano, T.; Matumoto, K.; Nakamura, T. Identification of Gas6, a putative ligand for Sky and Axl receptor tyrosine kinases, as a novel neurotrophic factor for hippocampal neurons. *J. Neurosci. Res.* 2002, 68, 150–160. [CrossRef] [PubMed]

Yagami, T.; Ueda, K.; Asakura, K.; Okamura, N.; Sakaeda, T.; Sakaguchi, G.; Itoh, N.; Hashimoto, Y.; Nakano, T.; Fujimoto, M. Effect of Gas6 on secretory phospholipase A2-2IIA-induced apoptosis in cortical neurons. *Brain Res.* 2003, 985, 142–149. [CrossRef]

Yagami, T.; Ueda, K.; Asakura, K.; Sakaeda, T.; Nakazato, H.; Kuroda, T.; Hata, S.; Sakaguchi, G.; Itoh, N.; Nakano, T.; et al. Gas6 rescues cortical neurons from amyloid β protein-induced apoptosis. *Neuropharmacology* 2002, 43, 1289–1296. [CrossRef]

Allen, M.P.; Zeng, C.; Schneider, K.; Xiong, X.; Meintzer, M.K.; Bellosta, P.; Basilico, C.; Varum, B.; Heidenreich, K.A.; Wierman, M.E. Growth arrest-specific gene 6 (Gas6)/adhesion related kinase (Ark) signaling promotes gonadotropin-releasing hormone neuronal survival via extracellular signal-regulated kinase (ERK) and Akt. *Mol. Endocrinol.* 1999, 13, 191–201. [CrossRef] [PubMed]

Allen, M.P.; Linseman, D.A.; Udo, H.; Xu, M.; Schaack, J.B.; Varum, B.; Kandel, E.R.; Heidenreich, K.A.; Wierman, M.E. Novel mechanism for gonadotropin-releasing hormone neuronal migration involving Gas6/Ark signaling to p38 mitogen-activated protein kinase. *Mol. Cell. Biol.* 2002, 22, 599–613. [CrossRef] [PubMed]

Salian-Mehta, S.; Xu, M.; Wierman, M.E. AXL and MET crosstalk to promote gonadotropin releasing hormone (GnRH) neuronal migration and survival. *Mol. Cell. Endocrinol.* 2013, 374, 92–100. [CrossRef] [PubMed]

Shankar, S.L.; O’Guin, K.; Cammer, M.; McMorris, F.A.; Stitt, T.N.; Basch, R.S.; Varum, B.; Shafit-Zagardo, B. The growth arrest-specific gene product Gas6 promotes the survival of human oligodendrocytes via a phosphatidylinositol 3-kinase-dependent pathway. *J. Neurosci.* 2003, 23, 4208–4218. [PubMed]

Shankar, S.L.; O’Guin, K.; Kim, M.; Varum, B.; Lemke, G.; Brosnan, C.F.; Shafit-Zagardo, B. Gas6/Axl signaling activates the phosphatidylinositol 3-kinase/Akt1 survival pathway to protect oligodendrocytes from tumor necrosis factor α-induced apoptosis. *J. Neurosci.* 2006, 26, 5638–5648. [CrossRef] [PubMed]

Li, R.; Chen, J.; Hammonds, G.; Phillips, H.; Armanini, M.; Wood, P.; Bunge, R.; Godowski, P.J.; Sliwkowski, M.X.; Mather, J.P. Identification of Gas6 as a growth factor for human Schwann cells. *J. Neurosci.* 1996, 16, 2012–2019. [PubMed]

Grommes, C.; Lee, C.Y.; Wilkinson, B.L.; Jiang, Q.; Koenigsknecht-Talboo, J.L.; Varum, B.; Landreth, G.E. Regulation of microglial phagocytosis and inflammatory gene expression by Gas6 acting on the Axl/Mer family of tyrosine kinases. *J. Neuroimmune Pharmacol.* 2008, 3, 130–140. [CrossRef] [PubMed]
87. Fourgeaud, L.; Través, P.G.; Tufail, Y.; Leal-Bailey, H.; Lew, E.D.; Burrola, P.G.; Callaway, P.; Zagórska, A.; Rothlin, C.V.; Nimmerjahn, A.; et al. TAM receptors regulate multiple features of microglial physiology. *Nature* 2016, 532, 240–244. [CrossRef] [PubMed]

88. Tang, Y.; Wu, S.; Liu, Q.; Xie, J.; Zhang, J.; Han, D.; Lu, Q.; Lu, Q. MerTK deficiency affects macrophage directional migration via disruption of cytoskeletal organization. *PLoS ONE* 2015, 10, e0117787. [CrossRef] [PubMed]

89. Dendrou, C.A.; Fugger, L.; Friese, M.A. Immunopathology of multiple sclerosis. *Nat. Rev. Immunol.* 2015, 15, 545–558. [CrossRef] [PubMed]

90. Tsunoda, I.; Libbey, J.E.; Kuang, L.Q.; Terry, E.J.; Fujinami, R.S. Massive apoptosis in lymphoid organs in animal models for primary and secondary progressive multiple sclerosis. *Am. J. Pathol.* 2005, 167, 1631–1646. [CrossRef]

91. Mistry, P.; Kaplan, M.J. Cell death in the pathogenesis of systemic lupus erythematosus and lupus nephritis. *Clin. Immunol.* 2016, in press.

92. Procaccini, C.; de Rosa, V.; Pucino, V.; Formisano, L.; Matarese, G. Animal models of Multiple Sclerosis. *Eur. J. Pharmacol.* 2015, 759, 182–191. [CrossRef] [PubMed]

93. Hiremath, M.M.; Saito, Y.; Knapp, G.W.; Ting, J.P.; Suzuki, K.; Matsushima, G.K. Microglial/macroage accumulation during cuprizone-induced demyelination in C57BL/6 mice. *J. Neuroimmunol.* 1998, 92, 38–49. [CrossRef]

94. Binder, M.D.; Prieto, A.L.; Kemper, D.; Butzkueven, H.; Gresle, M.M.; Cipriani, T.; Jokubaitis, V.G.; Carmeliet, P.; Kilpatrick, T.J. Gas6 deficiency increases oligodendrocyte loss and microglial activation in response to cuprizone-induced demyelination. *J. Neurosci.* 2008, 28, 5195–5206. [CrossRef] [PubMed]

95. Binder, M.D.; Xiao, J.; Kemper, D.; Ma, G.Z.; Murray, S.S.; Kilpatrick, T.J. Gas6 increases myelination by oligodendrocytes and its deficiency delays recovery following cuprizone-induced demyelination. *PLoS ONE* 2011, 6, e17727. [CrossRef] [PubMed]

96. Tsiperson, V.; Li, X.; Schwartz, G.J.; Raine, C.S.; Shafit-Zagardo, B. Gas6 enhances repair following cuprizone-induced demyelination. *PLoS ONE* 2010, 5, e15748. [CrossRef] [PubMed]

97. Hoehn, H.J.; Kress, Y.; Sohn, A.; Brosnan, C.F.; Bourdon, S.; Shafit-Zagardo, B. Axl−/− mice have delayed recovery and prolonged axonal damage following cuprizone toxicity. *Brain Res.* 2008, 1240, 1–11. [CrossRef] [PubMed]

98. Weinger, J.G.; Brosnan, C.F.; Loudig, O.; Goldberg, M.F.; Macian, F.; Arnott, H.A.; Prieto, A.L.; Tsiperson, V.; Shafit-Zagardo, B. Loss of the receptor tyrosine kinase Axl leads to enhanced inflammation in the CNS and delayed removal of myelin debris during experimental autoimmune encephalomyelitis. *J. Neuroinflamm.* 2011, 8, 49. [CrossRef] [PubMed]

99. Weinger, J.G.; Omari, K.M.; Marsden, K.; Raine, C.S.; Shafit-Zagardo, B. Up-regulation of soluble Axl and Mer receptor tyrosine kinases negatively correlates with Gas6 in established multiple sclerosis lesions. *Am. J. Pathol.* 2009, 175, 283–293. [CrossRef] [PubMed]

100. Ballantine, L.; Midgley, A.; Harris, D.; Richards, E.; Burgess, S.; Beresford, M.W. Increased soluble phagocytic receptors sMer, sTyro3 and sAxl and reduced phagocytosis in juvenile-onset systemic lupus erythematosus. *Pediatr. Rheumatol. Online J.* 2015, 13, 10. [CrossRef] [PubMed]

101. Bahlo, M.; Booth, D.R.; Broadley, S.A.; Brown, M.A.; Foote, S.J.; Griffiths, L.R.; Kilpatrick, T.J.; Lechner-Scott, J.; Moscato, P.; Perreau, V.M.; et al. Australia and New Zealand Multiple Sclerosis Genetics Consortium (ANZgene). Genome-wide association study identifies new multiple sclerosis susceptibility loci on chromosomes 12 and 20. *Nat. Genet.* 2009, 41, 824–828. [CrossRef] [PubMed]

102. Ma, G.Z.; Stankovich, J.; Australia and New Zealand Multiple Sclerosis Genetics Consortium (ANZgene); Kilpatrick, T.J.; Binder, M.D.; Field, J. Polymorphisms in the receptor tyrosine kinase MERTK gene are associated with multiple sclerosis susceptibility. *PLoS ONE* 2011, 6, e16964.
105. Binder, M.D.; Fox, A.D.; Merlo, D.; Johnson, L.J.; Giuffrida, L.; Calvert, S.E.; Akkermann, R.; Ma, G.Z.; ANZgene.; Perera, A.A.; et al. Common and Low Frequency Variants in MERTK Are Independently Associated with Multiple Sclerosis Susceptibility with Discordant Association Dependent upon HLA-DRB1*15:01 Status. PLoS Genet. 2016, 12, e1005853. [CrossRef] [PubMed]

106. Hoppmann, N.; Graetz, C.; Paterka, M.; Poisa-Beiro, L.; Larochelle, C.; Hasan, M.; Lill, C.M.; Zipp, F.; Siffrin, V. New candidates for CD4 T cell pathogenicity in experimental neuroinflammation and multiple sclerosis. Brain 2015, 138, 902–917. [CrossRef] [PubMed]

107. Alciato, F.; Sainaghi, P.P.; Castello, L.; Bergamasco, L.; Carnieletto, S.; Avanzi, G.C. Development and validation of an ELISA method for detection of growth arrest specific 6 (GAS6) protein in human plasma. J. Immunoassay Immunochem. 2008, 29, 167–180. [CrossRef] [PubMed]

108. Sainaghi, P.P.; Alciato, F.; Carnieletto, S.; Castello, L.; Bergamasco, L.; Sola, D.; Bongo, A.S.; Inglese, E.; Polosa, R.; Avanzi, G.C. Gas6 evaluation in patients with acute dyspnea due to suspected pulmonary embolism. Respir. Med. 2009, 103, 589–594. [CrossRef] [PubMed]

109. Sainaghi, P.P.; Collimedaglia, L.; Alciato, F.; Leone, M.A.; Puta, E.; Naldi, P.; Castello, L.; Monaco, F.; Avanzi, G.C. Elevation of Gas6 protein concentration in cerebrospinal fluid of patients with chronic inflammatory demyelinating polyneuropathy (CIDP). J. Neurol. Sci. 2008, 269, 138–142. [CrossRef] [PubMed]

110. Sainaghi, P.P.; Collimedaglia, L.; Alciato, F.; Molinari, R.; Sola, D.; Ranza, E.; Naldi, P.; Monaco, F.; Leone, M.; Pirisi, M.; et al. Growth arrest specific gene 6 protein concentration in cerebrospinal fluid correlates with relapse severity in multiple sclerosis. Mediat. Inflamm. 2013, 2013, 406483. [CrossRef] [PubMed]