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Spatial patterns of coral survivorship: Impacts of adult proximity versus other drivers of localized mortality

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Abstract

Species-specific enemies may promote prey coexistence through negative distance- and density-dependent survival of juveniles near conspecific adults. We tested this mechanism by transplanting juvenile-sized fragments of the brooding corals *Pocillopora damicornis* and *Seriatopora hystrix* 3, 12, 24 and 182 cm up- and down-current of conspecific adults and monitoring their survival and condition over time. We also characterized the spatial distribution of *P. damicornis* and *S. hystrix* within replicate plots on three Fijian reef flats and measured the distribution of small colonies within 2 m of larger colonies of each species. Juvenile-sized transplants exhibited no differences in survivorship as a function of distance from adult *P. damicornis* or *S. hystrix*. Additionally, both *P. damicornis* and *S. hystrix* were aggregated rather than overdispersed on natural reefs. However, a pattern of juveniles being aggregated near adults while larger (and probably older) colonies were not suggests that greater mortality near large adults could occur over longer periods of time or that size-dependent mortality was occurring.

While we found minimal evidence of greater mortality of small colonies near adult conspecifics in our transplant experiments, we did document hot-spots of species-specific corallivory. We detected spatially localized and temporally persistent predation on *P. damicornis* by the territorial triggerfish *Balistapus undulatus*. This patchy predation did not occur for *S. hystrix*.

This variable selective regime in an otherwise more uniform environment could be one mechanism maintaining diversity of corals on Indo-Pacific reefs.
Introduction

The processes maintaining high numbers of species in tropical rainforests and coral reefs have long been investigated (Connell 1978). One suggested mechanism for maintaining diversity is the Janzen-Connell hypothesis (Janzen 1970; Connell 1971), which proposes that species-specific enemies clustered near adults increase the local mortality of conspecific juveniles and prevent any single species from monopolizing resources. It has generally been applied to long-lived, stationary, terrestrial organisms such as trees (Zhu et al. 2013). Although there are examples of species-specific distance- or density-dependent mortality affecting community species richness (e.g., Packer and Clay 2000; Petermann et al. 2008; Bagchi et al. 2014), a meta-analysis found no general, net effect of distance from parent on offspring mortality across a variety of plant types, habitats, or life stages (Hyatt et al. 2003). Thus, some tree species may experience Janzen-Connell effects (Johnson et al. 2012) but the generality of the pattern has been difficult to document (Hyatt et al. 2003). In part, this may be because numerous other processes (habitat heterogeneity, spatial patterns of competitors, etc.) could obscure Janzen-Connell effects. This makes experimental tests difficult in field settings, especially when spatial scales over which they may be relevant are unclear.

Research addressing Janzen-Connell effects on coral reefs is rare (Marhaver et al. 2013). Explanations for maintenance of coral diversity more often invoke disturbance regimes, abiotic gradients (e.g., light, sedimentation), and competition hierarchies (Lang 1973; Connell 1978; Buss and Jackson 1979; Porter et al. 1981). One reason for the paucity of tests in marine systems may be that the hypothesis assumes that dispersal decreases monotonically with distance from parents and that the average dispersal distance is greater than the average distance from the parent at which predation occurs but that they are on the same order of magnitude (Nathan and
Casagrandi 2004), neither of which necessarily applies to marine species with pelagic larvae.

Coral larvae are competent to settle within hours of release to months after release (Richmond 1987; Miller and Mundy 2003; Nozawa and Harrison 2008) and may disperse up to hundreds of kilometers (Jones et al. 2009; Torda et al. 2013). Therefore, unlike seeds of many tree species, coral larvae need not be distributed as “seed shadows” with juveniles clustered near parents.

Nevertheless, distance- or density-dependent mortality of juveniles could affect coral species employing either of the sexual reproduction methods that corals use: broadcast spawning, in which eggs and sperm are released into the water column and fertilized eggs develop outside corals, or brooding, in which sperm are released into the water column and fertilize eggs retained inside adult coral colonies. Larvae produced by both methods select their settlement sites and can be attracted to the chemical cues of conspecifics (Dixson, Abrego, and Hay 2014); this could lead to larvae settling near conspecific adults or in aggregations (e.g., Dunstan and Johnson 1998). Additionally, brooding corals may cast larval shadows akin to terrestrial seed shadows because larvae from brooding corals frequently settle quickly and close to their parents (Carlon and Olson 1993; Tioho, Tokeshi, and Nojima 2001; Vermeij 2005; Vermeij and Sandin 2008; Torda et al. 2013).

Selective mortality of juveniles near conspecific adults is often assumed to be due to specialist enemies that accumulate near adults over their lifetimes. While this may be the case for terrestrial plants where many herbivores and pests are specialists (Bernays 1989), it is unclear to what extent this applies to corals, of which there are relatively few identified species-specific consumers that might be expected to accumulate near adults of specific prey species (Cornell and Karlson 2000; Rotjan and Lewis 2008; but see Neudecker 1979 and Jayewardeen et al. 2009 for examples of coral-specific predation).
However, there is growing evidence from both terrestrial (Packer and Clay 2000; Bagchi et al. 2014; Fricke, Tewksbury, and Rogers 2014) and marine systems (Marhaver et al. 2013) that microbial pathogens may accumulate near adults and suppress the survivorship of conspecific recruits or juveniles. In the most direct test of the Janzen-Connell hypothesis in corals, Marhaver et al. (2013) used a series of lab and field investigations in the Caribbean to attribute higher mortality of *Orbicella* (formerly *Montastraea*) *faveolata* recruits placed near adult conspecifics to adult-associated microbial enemies. They found a complex relationship between distance from adult colonies, current direction, and recruit mortality. In less direct tests, Vermeij (2005) and Vermeij and Sandin (2008) observed that survival of coral recruits decreased with increasing cover of conspecifics; they hypothesized that this was due to species-specific microorganisms rather than to saturation of a limiting resource.

No study has explicitly tested the Janzen-Connell hypothesis in brooding corals or in fragments typical of juvenile-sized corals. These conditions may generate different results from previously tested conditions because brooding corals may be subject to different distance-dependent mortality patterns compared to broadcast spawning corals and because small fragments may differ from newly settled larvae. For instance, distance-dependent mortality is known to affect some seedlings more than seeds in terrestrial systems (Hyatt et al. 2003). We experimentally evaluated distance-dependent mortality of juvenile-sized corals in the field and correlatively surveyed multiple reefs for patterns of spatial distribution suggestive of Janzen-Connell effects. We focused on two brooding coral species (*Seriatopora hystrix* and *Pocillopora damicornis*) whose planulae recruit over short distances, the latter of which is known to be a preferred prey for some coral consumers (Neudecker 1979).
Methods

Study site characteristics

This study was conducted on reef flats within no-take marine protected areas (MPAs) adjacent to Votua, Vatu-o-lailai, and Namada villages along the Coral Coast of Viti Levu, Fiji. These reserves are scattered along 11 km of fringing reef and are separated by ~3-8 km. The reserves have high coral cover (38-56%), low macroalgal cover (1-3%), and a high biomass and diversity of herbivorous fishes (Rasher, Hoey, and Hay 2013; Bonaldo and Hay 2014). The reef flats range from ~1-3 m deep at high tide, exposed to ~1.5 m deep at low tide, extend ~500-600 m from shore to the reef crest, and are typical of exposed reef flats occurring throughout Fiji. Except during low tides in calm weather, waves push water over the reef front, and water then flows laterally across the reef flats to discharge through channels bisecting the flats. This creates a relatively predictable current direction at most locations.

Survival experiments

To test whether juvenile corals experienced distance-dependent mortality near adult conspecifics, we created ~5 mm tall fragments of P. damicornis and S. hystrix, selected suitable adult focal colonies (defined below), attached conspecific fragments 3, 12, 24 and 182 cm up- and down-current from each focal adult, and monitored fragment survival. General direction of current (east to west) was determined by in-water observations over ~7 years of working at this site. We conducted this experiment in Votua village’s MPA, which supports a diverse assemblage of corals covering about 50% of hard substrates (Rasher, Hoey, and Hay 2013). Water flow across this reef flat is often negligible around low tide, reducing the potential for constantly dissipating microbial enemies away from focal colonies.
We used pliers to clip 16 fragments of 30-40 polyps each from the tips of each of 24 large *P. damicornis* and 24 large *S. hystrix* colonies in the Votua village MPA. We employed fragments from older colonies as proxies for ~6 month old juveniles (Sato 1985) because, despite these species reproducing monthly in some locations (Fan et al. 2002; Kuanui et al. 2008), neither species planulated at our site during the months of this study (August through October 2013). The fragments from each of four source colonies for a species were collected in six rounds over two days. Each round was taken to shore and four fragments (one from each source colony) were epoxied (Emerkit epoxy) onto the unglazed side of 16 2.54 x 2.54 cm tiles. Thus, each tile had fragments from four different colonies and sets of 16 tiles had fragments from the same four colonies of the same species. After epoxying, tiles were held in a tub of seawater for ~1 h, allowing the epoxy to harden. Tiles were then cable-tied onto metal racks at ~1 m deep in the MPA and allowed to acclimate for two weeks before deployment in the experiment. Survivorship during acclimation was 100%, producing 384 fragments on 96 tiles for each coral species.

Within the MPA, 10 adult *P. damicornis* and 10 adult *S. hystrix* colonies served as focal colonies. Focal colonies: i) were >10 cm at their smallest diameter (10 to 35 cm for *P. damicornis* and 10 to 75 cm for *S. hystrix*), ii) had no conspecific colonies within 4 m (so as not to confound effects of the focal colony with effects of nearby conspecifics), iii) were 5-40 cm deep at low tide, and iv) had space for 190 cm PVC pipes to be placed roughly east and west (the predominant current direction was west) without disturbing other corals. Focal colonies were photographed from above and their size determined using ImageJ (Rasband 1997). Twenty mm diameter by 190 cm long PVC pipes served as platforms to which we attached the tiles. Pipes were anchored to the reef by driving steel rebar through pre-drilled holes.
and cementing the rebar to the pipe. Notches 2.54 cm long allowed us to cable-tie tiles onto the pipes at distances of 3, 12, 24 and 182 cm from focal colonies (Fig 1a, 1b). This approach secured all pipes and tiles throughout the experiment. These distances and this scale were chosen to match a previous experiment in the Caribbean that had detected distance-dependent mortality of newly settled recruits for a broadcast spawning coral (Marhaver et al. 2013).

Tiles were randomly assigned to positions on pipes. Thus, fragments at each distance and around each conspecific focal colony were random with respect to source colony. Unassigned tiles were kept on the rack as spares (64 fragments on 16 tiles for each coral species).

Every 1-2 d after deployment, we examined all fragments, recording survivorship, consumption, overgrowth by algae, bleaching, or other changes in status. The *P. damicornis* fragments were observed for 59 days. The *S. hystrix* fragments were deployed one month later and observed for 29 days.

On some *P. damicornis* tiles, three or four of the fragments disappeared within a 24 h period between checks on their condition, appearing to have been bitten off. To determine the agents of this localized mortality, we replaced tiles whose four fragments had been eaten with spare tiles holding four healthy fragments around three of the focal colonies that had experienced localized mortality and videotaped the tiles (GoPro II HD) from about 1 m away during the following high tides. Cameras were retrieved at the next low tide and the videos watched.

We evaluated overall survival patterns and mortality specifically due to bleaching or predation using mixed-effects Cox proportional hazards survival models (coxme package, Therneau 2012) in R (R Core Team 2013). Distance and direction from focal colony were fixed effects (4 levels and 2 levels, respectively) and focal colony and tile nested within focal colony were random effects because fragments were blocked by tile and focal colony. The size of the
focal colony and the depth of the tiles were included as covariates. Finally, we compared the relative levels of predation and bleaching in *P. damicornis* and *S. hystririx* using a chi-square test.

### Distribution surveys

We characterized the spatial distribution of *P. damicornis* and *S. hystririx* in the reef flat MPAs of Namada, Vatu-o-lailai, and Votua villages at two scales (August through October 2013). For the larger-scale survey, we mapped each colony within 8 x 8 m plots (N=5, 5, and 10 for Namada, Vatu-o-lailai, and Votua, respectively). Each plot was divided into 256 0.5 x 0.5 m cells and each coral >1 cm across mapped into a cell. The location of each survey plot was determined by randomly choosing a point on shore, swimming 100, 200, or 300 kicks directly away from shore at that point, and surveying the closest bommie (coral patch) large enough to fill more than three quarters of an 8 x 8 m plot. In four of 10 surveys at Votua and in all five surveys at Vatu-o-lailai and Namada, we also measured the largest diameter of each *P. damicornis* colony. We did not measure *S. hystririx* colony size because the frequent discontinuity of colonies made accurate estimation of colony area too error-prone. To avoid confounding biotically-driven spatial distribution with patterns caused by patchiness of suitable substrate, we also recorded which cells were comprised primarily of unsuitable habitats such as sand-scoured pools or channels and bommie tops covered in rubble.

We analyzed these data using the neighborhood density function O(r) in the point pattern analysis program Programita (Wiegand and Moloney 2004). This analysis identifies distances at which individuals are aggregated, randomly spaced, or overdispersed compared to a specified null model. Unlike the more frequently used Ripley’s K(r) statistic, each distance category is not affected by those inside it; expected aggregation at each distance is compared to the observed
value independently of nearer distances. Each concentric ring centered on an individual coral is
separately placed on the aggregated-overdispersed continuum and displays the spatial pattern
within a different distance category. Ring width was 0.5 m extending up to 4 m. The null model
for this analysis was complete spatial randomness (CSR). Because the variance in substrate types
violated CSR’s assumption of uniform likelihood of coral placement, we conducted the below
analyses once using the entirety of all 8 x 8 m plots and a second time excluding cells of
unsuitable habitat (which should better meet CSR’s assumption of uniform likelihood).

To determine whether the observed spatial pattern was random, significantly aggregated,
or overdispersed, Programita simulated placement of each plot’s colonies 999 times using CSR,
calculated O(r) for each simulation, then combined replicate O(r)’s from each reef and from all
three reefs. This generated a distribution of simulated O(r)’s from which we established the
significance of the observed spatial patterns. The distance(s) at which significant aggregation or
overdispersion occurred were determined by the distances at which the observed pattern fell
above or below the 95% simulation envelopes, respectively. This analysis does not parse
aggregating and overdispersing processes; it shows the net resulting pattern.

In addition to analyzing all *P. damicornis* and *S. hystrix* colonies, we analyzed *P.
damicornis* <5 cm, ≥5 cm, ≥10 cm, and ≥15 cm in diameter to see if spatial patterns changed
with colony size. The <5 cm and ≥5 cm categories were mutually exclusive but because there
were not enough colonies between 5 and 10 cm and between 10 and 15 cm to analyze as
mutually exclusive groups, larger size categories were subsets of smaller ones.

The 8 x 8 m quadrat surveys could not resolve spatial patterns below the cell size of 0.5 x
0.5 m, meaning that patterns occurring at less than 0.25\(^2\) m could be undocumented. To
determine the spatial distribution of *P. damicornis* and *S. hystrix* at smaller scales, we conducted
211 2 m radius circular surveys around focal *P. damicornis* and *S. hystrix* colonies that i) were the
212 largest colony of that species within 4 m (to reduce the effects of conspecifics), and ii) occurred
213 where >75% of the substrate within 2 m was suitable habitat for *P. damicornis* and *S. hystrix*,
214 again to equalize the likelihood of colonies occurring everywhere in the survey.
215
216 The distance to each surrounding (radial) *P. damicornis* and *S. hystrix* colony was the
217 average of the distance to that colony’s near and far sides (N=45 focal colonies for *P. damicornis*
218 around *P. damicornis*, 10 for *S. hystrix* around *P. damicornis*, and 24 each for *P. damicornis* and
219 *S. hystrix* around *S. hystrix*). We analyzed radial colony counts in 10 cm concentric rings using a
220 generalized linear mixed effects model with Poisson errors and the canonical log link function in
221 R (*lme4* package, Bates et al. 2013). Distance was a fixed effect and focal colony was a random
222 effect, with the $\log_{10}$ of the ring sizes as an offset to control for unequal area sampled at each
223 distance (i.e. ring area increased with distance from the focal colony). We repeated this analysis
224 with just the closest 0.5 m and 1 m of the circles in case radial colonies beyond those distances
225 were masking short-range effects of the focal colonies.
226
227 We also analyzed the *P. damicornis* data from the 8 x 8 m plots in the same manner as we
228 did the circular surveys. To convert the plot data, a function was written in R to identify every
229 surveyed *P. damicornis* colony $\geq$2 m from all edges of its plot and equal to or larger than a
230 specified diameter (either 15 or 20 cm) as a focal colony (N=38 and 19 focal colonies,
231 respectively). In order to have an appreciable sample size, we did not restrict focal colonies to
232 those that were the largest within 4 m. The script then calculated the distances to all *P.
233 damicornis* colonies less than the specified focal colony diameter within 2 m and placed them
234 into 10 cm concentric rings as above. We analyzed the resulting data using the same procedures
235 as described for the circular surveys.
Results

Survival experiments

In our field experiment, neither distance nor direction from focal colony significantly affected survival of *P. damicornis* or *S. hystrix* fragments (Fig. 2a & b, respectively). We observed two main categories of mortality: bleaching preceding death in place (potentially due to microbes [e.g., Ben-Haim, Zicherman-Keren, and Rosenberg 2003]) and partial or complete disappearance, putatively due to predation (akin to Lenihan et al. 2011). Bleaching (47 and 46 fragments out of 320 for *P. damicornis* and *S. hystrix*, respectively) of neither species was affected by distance or direction (Fig. 2c & d). Distance and direction did not affect the number of *P. damicornis* fragments that partially or fully disappeared (putative predation), and direction did not affect this for *S. hystrix* but distance was significant (z=2.23, p=0.03) (Fig. 2e & f), with disappearance increasing with distance from the focal colony. In contrast, 0% of the 64 extra fragments of each species remaining on the coral rack where we originally acclimated the corals bleached or disappeared despite being on the same reef at the same time (Cox proportional hazards survival analysis, likelihood ratio for *P. damicornis*=16.5, likelihood ratio for *S. hystrix*=24.7, p<0.0001 for both species). Fragments on the coral rack were ~1 m above the benthos and may have experienced more flow or fewer benthic-associated biotic or physical stressors compared to the fragments on PVC pipes, which were 5-15 cm above the benthos.

The rapid disappearance of *P. damicornis* fragments around some focal colonies suggested spatially localized predation. Therefore, we further divided deaths due to putative predation between isolated predation incidents (disappearance of one or two fragments on a tile in 24 h) and localized predation episodes (disappearance of three or four fragments from a tile in
We used this classification scheme because three or four fragments tended to disappear from multiple tiles within the same 24 h period at certain replicates, whereas the disappearance of one or two fragments was not often temporally coincident across tiles within a replicate. We distinguished between these two types of putative predation because their causes were potentially different and therefore either one could have been distance-dependent or masked distance-dependence in the other. Six of 10 *P. damicornis* replicates (23 out of 160 tiles) experienced localized predation on at least one of their eight tiles; three of those experienced localized predation on five or more tiles within 24 h. Two of 10 *S. hystrix* replicates experienced localized predation (on one tile each). We further investigated localized predation only for *P. damicornis* because localized predation on *S. hystrix* was infrequent.

When tiles that had experienced localized predation around three focal colonies were replaced with spare tiles holding healthy fragments, all three sets of replacement tiles again experienced localized predation and their collective survival was significantly lower than that of the replicates that did not experience localized predation in the initial run (mixed effect Cox proportional hazards, $z=3.5$, $p<0.0005$). Videos of these tiles showed the territorial triggerfish *Balistapus undulatus* consuming multiple fragments from multiple tiles around two of the three focal colonies. *Balistapus undulatus* feeding resulted in fragments irregularly broken at or above the top of the epoxy, as was seen for most localized predation episodes in the initial outplanting.

We next examined whether localized predation was distance-dependent and whether it masked distance-dependent mortality in replicates that did not experience localized predation. Distance and direction did not significantly affect mortality in replicates that did not experience localized predation (Fig 3a). Considering only replicates that experienced localized predation...
(both original and replacement tiles), neither distance nor direction significantly affected mortality from all causes (Fig. 3b) or just from localized predation (Fig. 3c).

*Pocillopora damicornis* fragments were significantly more likely to die of putative predation as opposed to bleach and die in place than were *S. hystrix* fragments (chi-square test, $\chi^2=17.2$, df=1, p<0.0001). More than three times as many *P. damicornis* fragments died from putative predation as bleached prior to death (169 vs. 47 out of 320, respectively), while numbers of *S. hystrix* fragments that died from putative predation versus bleaching did not differ significantly (58 vs. 46 out of 320, respectively). Excluding replicates with localized predation, *P. damicornis* and *S. hystrix* appeared equally susceptible to isolated predation and bleaching ($\chi^2=0.022$, df=1, p=0.88).

**Distribution surveys**

We analyzed patterns of distribution using both entire 8 x 8 m plots and after excluding habitat deemed unsuitable for *P. damicornis* or *S. hystrix* (e.g., sand-scoured channels and pools, bommie tops covered in rubble). The analyses using only suitable habitat were quantitatively similar to those using the entire plots but were more conservative. Neighborhood density graphs using only suitable habitat are included here. Neighborhood density analysis indicated that both *P. damicornis* and *S. hystrix* were significantly aggregated at up to 1 m when all size classes were considered and surveys from all villages were pooled (Fig. 4a & b, respectively). When analyzed by site, the distance below which colonies were aggregated ranged from < 1 m in Votua and Vatu-o-lailai to nearly 3 m in Namada (see appendix). At no distance on any reef were colonies significantly overdispersed.
Identical analyses with *P. damicornis* separated into size categories (Fig. 4c-f) indicated that the largest colonies (≥15 cm) were not aggregated at any scale, but all smaller size classes were strongly aggregated at scales of up to 1 m. Thus, smaller colonies appeared to drive the aggregation at up to ~1 m when we analyzed all sizes together. However, the limited sample size for large colonies (n=187) may have constrained our ability to detect spatial patterns for large colonies.

To resolve the spatial distribution of *P. damicornis* and *S. hystrix* more finely, we conducted separate circular surveys (radius = 2 m) around focal colonies that met specific criteria. Across all 2 m, there was a significant negative relationship between distance from focal *P. damicornis* colonies and radial *P. damicornis* count (corrected for area surveyed at each distance and henceforth called density), focal *P. damicornis* and radial *S. hystrix* density, and focal *S. hystrix* and radial *P. damicornis* density (GLM: $z=-4.4, p<0.0001$; $z=-3.9, p<0.0005$; $z=-3.6, p<0.0005$, respectively) (Fig. 5a & b). The relationships within the first 0.5 m or 1 m for these focal-radial combinations were not significant (see Table 1 for all values not provided in text).

Across all 2 m, there was no significant relationship between distance from focal *S. hystrix* colony and radial *S. hystrix* density (GLM, $z=-1.9, p=0.06$) (Fig. 5b). However, there was a significant positive relationship between distance and density within the first 0.5 m (GLM, $z=-12.99, p<0.05$) but not within the first 1 m.

When we converted the 8 x 8 m surveys into data analogous to the circular surveys and considered any *P. damicornis* colony ≥15 cm across as a focal colony and any smaller individual as a radial colony, there was a significant negative relationship between distance and radial *P. damicornis* density (GLM, $z=-3.6, p<0.0005$) across all 2 m but not across the first 0.5 m or 1 m.
However, when the cutoff for focal colonies was 20 cm, there was no relationship between distance and *P. damicornis* colony count at 0.5 m, 1 m, or 2 m (Fig. 5c; Table 2).

**Discussion**

Using small portions of adult *P. damicornis* and *S. hystrix* colonies to represent ~6 month old juveniles, we tested for distance-dependent survivorship as a function of proximity to adult conspecifics. Survival experiments with *P. damicornis* and *S. hystrix* fragments did not show distance-dependent mortality around conspecific adults (Fig. 2a & b). The lack of distance-dependent mortality in this study is consistent with a meta-analysis of distance-dependent mortality studies of the seeds and seedlings of terrestrial plants (Hyatt et al. 2003), in which distance from parents did not affect overall survival. However, when separated by life stage, that meta-analysis found that seedling survival increased with distance from parents while seed survival was not affected, suggesting that the strength of distance-dependent mortality may be a function of age. Our experiment using small coral fragments to represent juveniles attempted to conduct a similar test with corals. Our procedures would not have detected distance-dependent mortality of larvae occurring just after settlement. We would have preferred to conduct a reciprocal transplant experiment of fragments from both corals at differing distances to both conspecific and heterospecific adults but we were unable to gain permission to use that many coral colonies. Thus, we could document spatial patterns of survivorship relative to conspecific adults but not relative to heterospecific adults.

Spatial analyses of the distribution of conspecific colonies might uncover patterns that our short-term experiment could not detect. Observed spatial patterns represent the balance of
multiple, potentially opposing processes, such as greater recruit density near brooding parents (similar to terrestrial seed shadows) versus detrimental effects of adult-associated enemies or intraspecific competition on aggregated, nearby recruits. Rather than overdispersion, we found significant clumping within 1 m of conspecifics for both *P. damicornis* and *S. hystrix* (Fig. 4a & b). The 8 x 8 m surveys and the 2 m radius surveys both supported this pattern; there was a significant negative relationship between *P. damicornis* radial colony density and distance from focal *P. damicornis* and a nearly significant negative relationship (with a much more limited sample size) between *S. hystrix* radial colony density and distance from focal *S. hystrix* (Fig. 5a & b). We also observed a significant negative relationship between *S. hystrix* density and distance from *P. damicornis* and *P. damicornis* density and distance from *S. hystrix* (Fig. 5a & b), suggesting that the cause of declining density with distance need not be species-specific.

Since *P. damicornis* and *S. hystrix* are confamilial, it is possible that they aggregate because a location that is physiologically beneficial for one might also be beneficial for the other. We did detect one pattern consistent with the Janzen-Connell hypothesis: small colonies of *P. damicornis* were aggregated at scales of up to 1 m, while colonies $\geq 15$ cm in diameter were not aggregated at any scale (Fig. 4). This selective loss of small colonies near adults is consistent with the Janzen-Connell hypothesis, but is also consistent with self thinning from intraspecific competition without mortality due to enemies aggregated near adults (Zhu et al. 2013).

There are a few potential causes for the observed clumping of conspecifics that could counteract Janzen-Connell effects (Carlon and Olson 1993). Aggregated settlement near maternal adults may occur for *P. damicornis* and *S. hystrix* because brooded planulae can settle quickly after release (Richmond 1987; Isomura and Nishihira 2001; Underwood et al. 2007; Torda et al. 2013), and even if planulae disperse meters or kilometers, they may still aggregate
near conspecific adults (Babcock 1988; Tioho, Tokeshi, and Nojima 2001; Doropoulos et al. 2015). Moreover, pocilloporid recruitment is inherently spatially heterogeneous (Dunstan and Johnson 1998) and occurs in hotspots that may be partially determined by water flow, density of adult confamilials (Eagle 2006), and substrate suitability (Harriott 1983; Lee, Walford and Goh 2009). Thus, multiple ecologically important processes and interactions can generate aggregation of juveniles, and some of these could overwhelm Janzen-Connell effects and make them seem unimportant in the field (at least in the short term), even if they were occurring.

The only other direct test of the Janzen-Connell hypothesis in corals was conducted on planulae and recruits of broadcasting *Orbicella* (formerly *Montastraea*) *faveolata* in the Caribbean (Marhaver et al. 2013). In that study distance-dependent mortality appeared to be microbially mediated, with effects differing upstream and downstream of focal *O. faveolata*. The design of that study and ours differs in several ways.

First, Marhaver et al. (2013) used planulae and recruits a few days old in their distance-dependent survival experiments, whereas we used fragments taken from mature colonies. There are potential differences between fragments from adult corals and recruits. For example, the physiology, skeletal structure, and microbiomes of fragments from adults may differ from those of recruits and similarly sized juveniles (Vandermeulen and Watabe 1973; Harriott 1983; Le Tissier 1988; Christiansen et al. 2009). The planulae of *P. damicornis* have higher lipid percentages in their tissue than do adult colonies (Figueiredo et al. 2012), which could affect palatability and microbial defense. Most recruit mortality in the study by Marhaver et al. (2013) appeared to be microbe-related as opposed to predator-generated. In contrast, predators generated considerable mortality of our juvenile sized transplants. Liberally assuming that every bleaching death in our study was due to microbes, only about one quarter of *P. damicornis* and
half of *S. hystrix* fragments could have died directly because of microbes; thus, about 50-75% of the mortality we observed appeared to be due to consumption by fish. One way in which using colony fragments may not have been so different from using recruits was that both age categories may have similar zooxanthellae endosymbionts, since *P. damicornis* vertically transfers zooxanthellae to its young (LaJeunesse et al. 2004).

Second, Marhaver et al. (2013) studied the broadcast spawning species *O. faveolata*, while we studied two brooding species whose planulae may be more likely to settle near their parents, and whose larvae may receive critical components of their microbiome via vertical transmission from adults. Data on the make-up and function of juvenile coral microbiomes are limited but at present there is some evidence that larvae from brooding species may be more consistently endowed with parental components of the microbiome than are the larvae of broadcast spawners (Littman, Willis, and Bourne 2009; Apprill et al. 2012; Sharp, Distel and Paul 2012; Lema, Willis, and Bourne 2014). In some acroporid corals, juveniles do not develop microbiomes typical of adult colonies until greater than 9 months of age (Littman, Willis, and Bourne 2009) but a core component of the microbiome appears in all the early stages, despite additions of other microbial species from the environment later in development (Lema, Willis, and Bourne 2014). However, in brooding species such as *Porites* and *Pocillopora*, critical microbes are transmitted from adults to larvae, or very quickly acquired from the environment, and even very young juvenile stages resemble adults in their composition of key microbes comprising the symbiotic microbiome (Apprill et al. 2012; Sharp, Distel, and Paul 2012). We do not know these relationships for the species we investigated but if their microbiomes take months to develop and are important defenses against microbial enemies, then our use of small adult portions may not mimic juvenile susceptibility to adult-associated pathogens. In contrast, if the
critical components of the microbiome are present in even the earliest stages, then our adult fragments should be more representative. We would have preferred to use recently recruited larvae but neither *P. damicornis* nor *S. hystrix* planulated at our study site during our experiment.

Finally, the focal adult colonies of *Orbicella* investigated by Marhaver et al. (2013) form larger, longer-lived colonies than the colonies of *Pocillopora* and *Seriatopora* that we investigated. It is possible that larger, longer-lived colonies accumulate more species-specific enemies over their lifetimes; if so, this could more strongly suppress juvenile survivorship near these longer-lived adults.

Although we did not detect distance-dependent mortality, we did document spatially heterogeneous corallivory on *P. damicornis*. This may promote species coexistence by producing a mosaic of favorable and unfavorable patches for *P. damicornis* across the reef (Levin and Paine 1974; Holt 1984). Corallivore activity can structure coral distribution on reefs in both the Pacific and Caribbean (Neudecker 1979; Littler, Taylor, and Littler 1989) and parrotfish and butterflyfish density can impact coral recruit and juvenile mortality, respectively (Penin et al. 2010). Localized predation by the triggerfish *Balistapus undulatus* on small *P. damicornis* could have a similar effect here. *Balistapus undulatus* is a generalist with territories of 100-200 m² (McClanahan 2000) and eats the tips of branching corals, including *P. damicornis* (Hiatt and Strasburg 1960; Neudecker 1979). This triggerfish species’ territoriality may delineate certain patches on reefs in which some species (e.g., *P. damicornis*) have high mortality while other species (e.g., *S. hystrix*) are not directly affected, akin to what is seen with seaweed in territories of the steephead parrotfish on the Great Barrier Reef (Welsh and Bellwood 2012) or *Pocillopora* and *Pavona* in the interaction between damselfish territories and roving corallivores in the Eastern Pacific (Wellington 1982). Additional experiments are necessary to determine how
patchy corallivory contributes to the coexistence of *P. damicornis*, *S. hystrix*, and corals in general.

Overall we found little evidence for distance-dependent mortality relative to focal conspecific adults and for the pattern of over-dispersion that distance-dependent mortality would be expected to produce. Instead, both *P. damicornis* and *S. hystrix* aggregated at the scale of 1 m or less, with a tendency for small colonies to be clumped around larger ones. These findings suggest that local dispersal shadows or areas of physiological benefit near prospering adult conspecifics equal or exceed Janzen-Connell effects for the brooding corals we studied on these Fijian reef flats. Our experiments using small coral fragments did not detect distance-dependent mortality by species-specific enemies; we did, however, observe spatially heterogeneous corallivory on *P. damicornis*, which could facilitate species coexistence by delineating reef patches that are more or less favorable to different corals.

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Figure 1- Experimental set up of coral fragments epoxied to tiles on PVC pipes around focal adult colonies. a) PVC pipes radiating outwards from a focal *Seriatopora hystrix* colony. b) Fragments epoxied onto ceramic tiles around an *S. hystrix* colony.
Figure 2- Left column is *Pocillopora damicornis* and right column is *Seriatopora hystrix*.

Statistical values are from mixed-effects Cox proportional hazards survival analyses.

n=80 fragments at each distance across 10 focal colonies and pooled between both directions. a & b) Survivorship through time for *Pocillopora damicornis* and *Seriatopora hystrix* fragments. c & d) Cumulative number of fragments that bleached over time. e & f) Cumulative number of fragments that partially or fully disappeared over time (putative predation).
Figure 3- a) Survival of *Pocillopora damicornis* fragments in replicates (4 focal colonies, 32 fragments at each distance) that did not experience localized predation. b) Survival of *P. damicornis* fragments in the six focal colony replicates that did experience localized predation and in the replacement replicates. Deaths are from all causes. c) Fraction of *P. damicornis* fragments not killed by localized predation episodes in original replicates that experienced localized predation and in the replacement replicates. Direction not shown. Analyses as in Fig. 2.
Figure 4- Neighborhood density analysis of *Pocillopora damicornis* and *Seriatopora hystrix* in 8 x 8 m quadrats with replicates from all three reefs combined. Black lines are observed patterns; grey lines are the 95% simulation envelopes from 999 simulations. Where black lines are above the upper grey line colonies are significantly aggregated, where they are between the grey lines colonies are randomly spaced, and where they are below the lower grey line colonies are significantly overdispersed. These analyses used only areas of suitable substrate (see text for definition).
Manuscript to be reviewed
Figure 5- Density (± SE) of *Pocillopora damicornis* and *Seriatopora hystrix* at 10 cm intervals from focal a) *P. damicornis* and b) *S. hystrix* colonies. The linear regressions shown are to indicate the slope of the relationship found in the generalized linear mixed effects models but do not represent the models’ outputs. Radial colony count significantly declined with distance from focal colony over 2 m for three of the four focal-radial combinations (focal *P. damicornis*-radial *P. damicornis*—z=-4.4, p<0.001; focal *P. damicornis*-radial *S. hystrix*—z=-3.9, p<0.001; focal *S. hystrix*-radial *P. damicornis*—z=-3.6, p<0.001; focal *S. hystrix*-radial *S. hystrix*—z=-1.9, p=0.06). c) Density (mean ± SE) of *Pocillopora damicornis* within 2 m of focal *P. damicornis* based on the 8 x 8 m surveys. Focal colonies are *P. damicornis* that are ≥15 cm across or ≥20 cm across. Radial colonies are any colonies below that size. Radial colony count significantly declined with distance over 2 m when colonies ≥15 cm were considered focal (focal colonies ≥15 cm—z=-3.6, p<0.0005; focal colonies ≥20 cm—z=-1.09, p<0.27).
Table 1- Relationship between count of radial *Pocillopora damicornis* and *Seriatopora hystrix* colonies and distance from focal *P. damicornis* and *S. hystrix* colonies using generalized linear mixed effects models. “Maximum distance” is the distance up to which radial colonies were considered.

| Focal species-radial species | Maximum distance | Slope | z value | p-value |
|------------------------------|-----------------|-------|---------|---------|
| *P. damicornis-P. damicornis* | 0.50 m          | -0.0098 | -1.0    | 0.32    |
|                              | 1.0 m          | 0.00055 | 0.22   | 0.83    |
|                              | 2.0 m          | -0.0032 | -4.4    | <0.0001 |
| *P. damicornis-S. hystrix*   | 0.50 m          | 0.016  | 0.59    | 0.56    |
|                              | 1.0 m          | 0.0012 | 0.19    | 0.85    |
|                              | 2.0 m          | -0.0057 | -3.9 | <0.0005 |
| *S. hystrix-P. damicornis*   | 0.50 m          | 0.0099 | 0.57    | 0.57    |
|                              | 1.0 m          | 0.0042 | 1.1     | 0.28    |
|                              | 2.0 m          | -0.0036 | -3.6 | <0.0005 |
| *S. hystrix-S. hystrix*      | 0.50 m          | 0.042  | 2.3     | <0.05   |
|                              | 1.0 m          | -0.00065 | -0.18  | 0.86    |
|                              | 2.0 m          | -0.0017 | -1.9  | 0.06    |
Table 2- Relationship between count of radial *Pocillopora damicornis* colonies and distance from focal *Pocillopora damicornis* colonies using the data from the 8 x 8 m surveys. “Threshold size for focal colony” is the size above which surveyed colonies were designated “focal” and below which colonies were designated “radial.” “Maximum distance” is the distance up to which radial colonies were considered.

| Threshold size for focal colony | Maximum distance | Slope  | z value | p-value |
|---------------------------------|------------------|--------|---------|---------|
| 15 cm                           | 0.50 m           | -0.0064|-0.46    | 0.65    |
|                                 | 1.0 m            | -0.0063|-1.5     | 0.14    |
|                                 | 2.0 m            | -0.0034|-3.6     | <0.0005 |
| 20 cm                           | 0.50 m           | 0.047   | 1.2     | 0.23    |
|                                 | 1.0 m            | 0.00032 | 0.045   | 0.96    |
|                                 | 2.0 m            | -0.0016 |-1.09    | 0.27    |
1

*Seriatopora hystrix* fragments on tiles on PVC array

Figure 1- Experimental set up of coral fragments epoxied to tiles on PVC pipes around focal adult colonies. a) PVC pipes radiating outwards from a focal *Seriatopora hystrix* colony. b) Fragments epoxied onto ceramic tiles around an *S. hystrix* colony.
2

Survival and predation of *Pocillopora damicornis* and *Seriatopora hystrix*

Figure 2- Left column is *Pocillopora damicornis* and right column is *Seriatopora hystrix*. Statistical values are from mixed-effects Cox proportional hazards survival analyses. n=80 fragments at each distance across 10 focal colonies and pooled between both directions. a & b) Survivorship through time for *Pocillopora damicornis* and *Seriatopora hystrix* fragments. c & d) Cumulative number of fragments that bleached over time. e & f) Cumulative number of fragments that partially or fully disappeared over time (putative predation).
3

Pocillopora damicornis mortality relating to localized predation episodes

Figure 3- a) Survival of Pocillopora damicornis fragments in replicates (4 focal colonies, 32 fragments at each distance) that did not experience localized predation. b) Survival of P. damicornis fragments in the six focal colony replicates that did experience localized predation and in the replacement replicates. Deaths are from all causes. c) Fraction of P. damicornis fragments not killed by localized predation episodes in original replicates that experienced localized predation and in the replacement replicates. Direction not shown. Analyses as in Fig. 2.
Neighborhood density analysis for *Pocillopora damicornis* and *Seriatopora hystrix*

Figure 4- Neighborhood density analysis of *Pocillopora damicornis* and *Seriatopora hystrix* in 8 x 8 m quadrats with replicates from all three reefs combined. Black lines are observed patterns; grey lines are the 95% simulation envelopes from 999 simulations. Where black lines are above the upper grey line colonies are significantly aggregated, where they are between the grey lines colonies are randomly spaced, and where they are below the lower grey line colonies are significantly overdispersed. These analyses used only areas of suitable substrate (see text for definition).
Density of *Pocillopora damicornis* and *Seriatopora hystrix* at 10 cm intervals around focal colonies

Figure 5- Density (+/- SE) of *Pocillopora damicornis* and *Seriatopora hystrix* at 10 cm intervals from focal a) *P. damicornis* and b) *S. hystrix* colonies. The linear regressions shown are to indicate the slope of the relationship found in the generalized linear mixed effects models but do not represent the models’ outputs. Radial colony count significantly declined with distance from focal colony over 2 m for three of the four focal-radial combinations (focal *P. damicornis*-radial *P. damicornis*— $z=-4.4$, $p<0.001$; focal *P. damicornis*-radial *S. hystrix*— $z=-3.9$, $p<0.001$; focal *S. hystrix*-radial *P. damicornis*— $z=-3.6$, $p<0.001$; focal *S. hystrix*-radial *S. hystrix*— $z=-1.9$, $p=0.06$). c) Density (mean +/- SE) of *Pocillopora damicornis* within 2 m of focal *P. damicornis* based on the 8 x 8 m surveys. Focal colonies are *P. damicornis* that are $\geq 15$ cm across or $\geq 20$ cm across. Radial colonies are any colonies below that size. Radial colony count significantly declined with distance over 2 m when colonies $\geq 15$ cm were considered focal (focal colonies $\geq 15$ cm— $z=-3.6$, $p<0.0005$; focal colonies $\geq 20$ cm— $z=-1.09$, $p<0.27$).
**Table 1** (on next page)

Relationship between radial colony count and distance from focal colony

Table 1- Relationship between count of radial *Pocillopora damicornis* and *Seriatopora hystrix* colonies and distance from focal *P. damicornis* and *S. hystrix* colonies using generalized linear mixed effects models. “Maximum distance” is the distance up to which radial colonies were considered.
| Focal species-radial species | Maximum distance | Slope  | z value | p-value |
|------------------------------|------------------|--------|---------|---------|
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|                              | 1.0 m            | 0.00055| 0.22    | 0.83    |
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|                              | 1.0 m            | 0.0012 | 0.19    | 0.85    |
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| *S. hystrix*-*P. damicornis*  | 0.50 m           | 0.0099 | 0.57    | 0.57    |
|                              | 1.0 m            | 0.0042 | 1.1     | 0.28    |
|                              | 2.0 m            | -0.0036| -3.6    | <0.0005 |
| *S. hystrix*-*S. hystrix*     | 0.50 m           | 0.042  | 2.3     | <0.05   |
|                              | 1.0 m            | -0.00065| -0.18  | 0.86    |
|                              | 2.0 m            | -0.0017| -1.9    | 0.06    |
### Table 2

8x8 survey results converted to radial survey results

Table 2- Relationship between count of radial *Pocillopora damicornis* colonies and distance from focal *Pocillopora damicornis* colonies using the data from the 8 x 8 m surveys.

“Threshold size for focal colony” is the size above which surveyed colonies were designated “focal” and below which colonies were designated “radial.” “Maximum distance” is the distance up to which radial colonies were considered.
| Threshold size for focal colony | Maximum distance | Slope  | z value | p-value |
|--------------------------------|------------------|--------|---------|---------|
| 15 cm                          |                  |        |         |         |
|                                | 0.50 m           | -0.0064| -0.46   | 0.65    |
|                                | 1.0 m            | -0.0063| -1.5    | 0.14    |
|                                | 2.0 m            | -0.0034| -3.6    | <0.0005 |
| 20 cm                          |                  |        |         |         |
|                                | 0.50 m           | 0.047  | 1.2     | 0.23    |
|                                | 1.0 m            | 0.00032| 0.045   | 0.96    |
|                                | 2.0 m            | -0.0016| -1.09   | 0.27    |