Bacteremia among Children in Central Teaching Hospital of Pediatric in Baghdad City

Wafaa Abdulazeez Hadi

ABSTRACT

Introduction: children with Occult bacteremia shows presence of fever, difficulty in breathing, tachycardia, refusal of feeds, malaise or lethargy. In such a potentially life-threatening condition, isolation of the causative pathogen in blood culture is one of the most important bacteriological examinations with important clinical and therapeutic consequences.

Material and methods: This prospective study was carried out in the Central Teaching of Pediatric Hospital in Baghdad. Total 300 febrile patient aged from 1 days to 11 years of age without any localizing clinical features attending central teaching hospital of pediatrics in Baghdad and who were not hospitalized 30 days prior to the initial evaluation were enrolled into the study.

Results: Among 300 patients, 49 were positive for different kinds of bacteria confirming OB and 235 were negative after 18 hrs of incubation. Among different age groups, OB was positive in 9.7% in age group 1day-<29 days, 4.3% in age group 1 month -<11 months, 1.0% in age group 1year-<5 years and age group 5years-10years represent (3/49) respectively and 0.3% in age groups over 11years.

Conclusion: Occult bacteremia is the most prevalent condition among children’s less than one months. Imipenem, class of quinolone antibiotic is found to be sensitive towards bacterial isolates and thus could be effective treatment strategy for management of bacteremia.

Keywords: Bacteremia, Pediatric

INTRODUCTION

Fever is a common presenting symptom in pediatric patients especially in in children < 3 years of age. Among these febrile children, finite number of patients harbors serious bacterial illnesses and clinically indistinguishable from the rest. Such asymptomatic clinical condition with no apparent focus of infection is termed as occult bacteremia (OB). Bacteremia has been reported increasingly in pediatrics age group condition with a high mortality rate that varies between 30 and 70 per cent. OB is defined as the presence of pathogenic bacteria in the blood stream of a febrile child with good clinical state and presents fever without a known infectious process.

Children with OB present with fever, difficulty in breathing, tachycardia, refusal of feeds, malaise or lethargy. Bacteremia also occur in children who have sepsis (ie, clinical evidence other than fever of a systemic response to infection). Children with sepsis generally appear ill, have an increased heart rate or respiratory rate and may have a change in temperature (typically fever, although hypothermia is often seen in very young infants and newborns). Presence of circulating bacteria in bloodstream displays consequences like shock, multiple organ failure, disseminated intravascular coagulation, etc. In such a potentially life-threatening condition, isolation of the causative pathogen in blood culture is one of the most important bacteriological examinations with important clinical and therapeutic consequences. The majority of bacteremia cases are caused by a number of pathogens including Staphylococcus spp, Streptococcus spp, Escherichia coli and Klebsiella pneumonia. OB in children has different implications and different patterns than that in adults. There is a wide variation in the incidence and clinical characteristics of OB caused by different species of bacteria. Identifying the causative species and characterizing the clinical significance in a specific age group in a community is essential for the prevention and treatment of these infections. The successful isolation of causative pathogen from blood is decisive for proper antimicrobial treatment as different organisms have different antimicrobial susceptibilities. Administration of correct drug with antimicrobial susceptibility towards causative pathogen helps in improved prognosis. However, OB in children’s residing in Baghdad region not been investigated. Thus, the present study was focused on estimation of age and gender as a risk factor of OB, identification of the focal bacterial infections and isolation of main bacterial pathogens in children’s with OB aged between 3-11 years. Also, the susceptibility pattern of isolates to the commonly used antimicrobial agents in the treatment of sepsis will be determined.

MATERIAL AND METHODS

Subjects
This prospective study was carried out in the Central Teaching of Pediatric Hospital in Baghdad. Ethical approval was obtained from the faculty research publication committee, department of clinical laboratory science, college of medical and health technology along with official agreement of Ministry of health as well as the director of pediatrics central health hospital, Baghdad. The study was performed in between the July 2012 to October 2012.

Corresponding author: Wafaa Abdulazeez Hadi, Kadhimya Teaching Hospital, Karch Health Directorate, Baghdad, Iraq

How to cite this article: Wafaa Abdulazeez Hadi. Bacteremia among children in central teaching hospital of pediatrics in Baghdad city. International Journal of Contemporary Medical Research 2019;6(1):A1-A6.

DOI: http://dx.doi.org/10.21276/ijcmr.2019.6.1.13
Hadi, et al. Bacteremia among Children in Central Teaching Hospital of Pediatric

300 febrile patient aged from 1 days to 11 years of age without any localizing clinical features attending central teaching hospital of pediatrics in Baghdad and who were not hospitalized 30 days prior to the initial evaluation were enrolled into the study. Those patients with having nephrotic, rheumatologic or hematological diseases and those exposed to corticosteroid treatment for a period greater than five days or chemotherapy or radiotherapy were excluded from the study. For analysis of age and gender as a risk factor of OB, the children were divided into following groups: A-1 day-<29 days; B- 1 month-<11 months; C- 1 year-<5 years; D- 6-10 years and E- over 11 years.

Sample collection

The blood samples were collected from patients before starting antimicrobial treatment in duplicates or triplicates from two different sites at an interval at least 1-hour to check the probable chance of blood isolate contamination. For each blood culture, blood volume ≥0.5 mL for infants ≤1 month of age, ≥1 mL for children between 1 month and 36 months of age and ≥4 mL for children ≥36 months of age were collected.

Microbiology

Blood samples were cultured aseptically on aerobic (brain heart infusion) and anaerobic (thioglycolate broth) by direct inoculation of blood into culture bottles. The bottles were then incubated at 37°C. Bottles were examined daily for one week for signs of turbidity, hemolysis, or other evidence of growth. Positive samples with presence of bacteria in culture bottles were subjected to diagnostic tests by subculturing 0.1 ml of samples into blood agar plate, chocolate agar plate (3% to 10% of carbon dioxide) and MacConkey agar plate for 18 -24 hrs at 37°C for isolation of pathogens. Colonies were identified morphologically by Gram stain. The organisms grown on agar plates were identified by standard laboratory methods including API 20 E system, API Staph, and API 20E (Bio Merieux, France). Further identification of the clinical isolates were done by performing the biochemical tests i.e. indole, hydrogen sulphide (H₂S), citrate utilization, semi-solid mannitol, urease, oxidase, catalase, coagulase and mannitol salt agar. Inoculated blood culture media were discarded as negative if there was no growth after continuous incubation for 24 hours. Organisms were considered ‘contaminants’ if aerobic spores were observed.

Antimicrobial susceptibility test

Isolated microorganisms were further examined using antimicrobial susceptibility method. Antimicrobial susceptibility testing was performed for all blood culture isolates according to the criteria of the National Committee for Clinical Laboratory Standards (NCCLSs) by Mueller–Hinton agar plates by Kirby–Bauer disc diffusion method. Zone diameter was measured and interpreted per the Clinical and Laboratory Standards Institute (CLSI) guidelines. Based on zones sizes mentioned in interpretative chart, the organisms were classified as resistant, intermediate/ moderately susceptible or susceptible.

STATISTICAL ANALYSIS

Data entry and analysis was done using SPSS version computer 19 software. Comparisons were made using Chi-square test with Fisher exact tests. A p-value of <0.05 was considered indicative of a statistically significant difference. Odds ratio and chi-square tests were used to determine presence of association between risk factors and culture results. Logistic regression was used to explain the dependent variable based on the independent variable.

RESULTS

A prospective study was performed on 300 febrile patients with fever aged 1 day to over 11 years admitted to Central Teaching Hospital of Pediatric in Baghdad from July 2012 to October 2012. Blood culture results classified on basis of age and gender Based on blood culture tests, among 300 patients, 16.3% (49/300) were positive for different kinds of bacteria confirming OB and 78% (235/300) were negative after 18
**Table-3:** Distribution of age groups according to the gender

| Sex | Age groups | Total |
|-----|------------|-------|
|     | (1D - <29D) | (1M - <11M) | (1Y -<5Y) | (5Y-10Y) | (11Y+) | Total |
| Male | 119 | 36 | 13 | 6 | 6 | 180 |
|     | 39.7% | 12.0% | 4.3% | 2.0% | 2.0% | 60.0% |
| Femel | 65 | 27 | 15 | 11 | 2 | 120 |
|     | 21.7% | 9.0% | 5.0% | 3.7% | 0.7% | 40.0% |
| Total | 184 | 63 | 28 | 17 | 8 | 300 |
|     | 61.3% | 21.0% | 9.3% | 5.7% | 2.7% | 100.0% |

**MCP>0.05 (Non-significant)**

---

**Table-4:** The relationship between outcome and the first diagnosis

| Result | No growth | Occult bacteremia | Contamination | Total |
|--------|-----------|-------------------|---------------|-------|
|        | RDS | NNG | FIT | Fever | PUO | Jaundice |        |
| No growth | 24 | 8.0% | 9 | 3.0% | 48 | 16.0% | 123 | 41.0% | 20 | 6.7% | 11 | 3.7% | 235 |
| Occult bacteremia | 4 | 1.3% | 0 | 0.0% | 8 | 2.7% | 32 | 10.7% | 4 | 1.3% | 0.3% | 49 |
| Contamination | 2 | 0.7% | 0 | 0.0% | 1 | 0.3% | 13 | 4.3% | 0 | 0.0% | 0.0% | 16 |
| Total | 30 | 10.0% | 9 | 3.0% | 57 | 19.0% | 168 | 56.0% | 24 | 8.0% | 12 | 4.0% | 300 |

**x² = 10.452, P>0.05, (NS)**

*High fever (temperature ≥39ºC) was a predictive factor for occult bacteremia*

RDS=Respiratory disease syndrome, NNJ=Neonatal jaundice, PUO=Pyreximal kunon oregen

**Table-5:** The frequency and percentage of Bacteria isolated from patient with occult bacteremia (N=49)

| Result of growth | Bacteria isolated | Total |
|------------------|-------------------|-------|
|                  | E.coli | Staph. epidermidis | Klebsella | Staph.aureus | Acinetobacter | Enterobacter | Streptococci | Pseudo |      |
| No growth | 235 | 78.3% | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 235 |
| Occult bacteremia | 0 | 0% | 15 | 5.0% | 10 | 3.3% | 9 | 3.0% | 6 | 2.0% | 4 | 1.3% | 3 | 0.3% | 1 | 0.3% | 16 | 16.3% |
| Contamination | 16 | 5.3% | 0 | 0% | 0 | 0% | 0 | 0% | 0 | 0% | 0 | 0% | 0 | 0% | 0 | 0% | 0 | 0% | 16 | 5.3% |
| Total | 251 | 83.7% | 15 | 5.0% | 10 | 3.3% | 9 | 3.0% | 6 | 2.0% | 4 | 1.3% | 3 | 0.3% | 1 | 0.3% | 1 | 0.3% | 300 | 100% |

**MCP<0.01 (Highly significant)**

---

hrs of incubation (Table 1). Among different age groups, OB was positive in 9.7% in age group 1day-<29 days, 4.3% in age group 1 month -<11 months, 1.0% in age group 1year-<5 years and age group 5years-10years represent (3/49) respectively and 0.3% in age groups over 11years (Table 2). Based on the distribution of age groups according to the gender for patient under study found that the male represent high proportion of 60.0% (180/300) while the female represent 40% (120/300) with the male: female ratio 1.5: 1 (Table 3).

2. Focal source of infection and causative species of bacteremia

2.1 The focal bacterial infections

The focal source bacterial infections were identified in all the 300 patients included in the study. Major source of focal infection was found to be fever in 56.0% (168/300) patients with presence of 10.7% (32/49) of isolated bacteria in positive OB cases. 19.0% (57/300) represented FIT cases with 2.7% (8/49) of isolated bacteria in positive OB cases. Other sources of focal infections were respiratory disease syndrome (RDS), 10.0% (30/300) with 1.3%(4/49) the isolated bacteria, pyreximal kunon oregen (PUO) cases, 8.0% (24/ 300) with 1.3%(4/94) the isolated bacteria, jaundice 4.0% (12/300) with 0.3% (1/49) bacteria isolated represent and neonatal jaundice (NNJ) cases 3.0% (9/300) with no bacterial presence (0.0%) (Table 4).

2.2 Causative species of bacteremia

A total of 49 isolates were collected from blood cultures, in which Gram-negative bacteria is higher in proportion representing65.3% (32/49). Most common species
causing bacteremia in these patients were *E. coli* (5.0%), *Staphylococcus epidermidis* (3.3%), *Klebsiella spp.* (3.0%), *Staphylococcus aureus* (2.0%), *Acinetobacterspp.* (1.3%), *Enterobacter spp.* (1.0%) followed by *pseudo monas spp.* and *Streptococcus spp.* (0.3%) respectively (Table 5).

3. Antibiotic susceptibility of bacterial isolates

Antibiotic susceptibility tests were performed for 49 bacterial isolates obtained from positive OB cases. All 14 antibiotics showed different degrees of susceptibility towards the identified bacterial isolates (Table 6). After categorization of identified bacterial isolates based on gram staining, the results of susceptibility test of antibiotics on gram negative bacteria showed that all isolates except *Klebsiellae spp.* were sensitive to imipenem. *Klebsiellae spp.* were found to be 80% sensitive to cefipim.

*E. coli* were sensitive to imipenem (90%) followed by ciprofloxacin (70%), vancomycin, amikacin, azethromycin (60%) respectively, cefitriaxon, oxacillin, amoxicillin (40%) respectively, cefoxitin, picarlicillin, ticarcillin (30%) respectively and cefipim, ampicillin (20%) respectively. *Klebsiella spp.* were sensitive to cefipim (80%), followed by cefitriaxon, imipene, gentamicin, ciprofloxacin, ticarcillin (70%) respectively, vancomycin, amikacin, azethromycin pipracillin, oxacillin (60%) respectively, amoxicillin, cefoxitin (30%) respectively and amoxcillin, ampicillin (20%) respectively.

*Enterobacter spp.* 100% sensitive to gentamicin, imipenem, cefipim, and amikacin respectively, followed by ciprofloxacin, amoxicillin, vancomycin, ticarcillin (70%) respectively and azethromycin, ampicillin, cefoxitin, cefitrixon (30%) respectively. *Acinetobacter spp* were sensitive to imipenem (100%) followed by cefipim, azithromycin, gentamicin (75%) respectively, cefoxitin, cefitrixon vancomycin, amikacin, ciprofloxacin amoxicillin, oxacillin (50%) and ticarcillin (25%). *Pseudomonas spp.* were sensitive

| No | Antibiotic             | E. coli N=15 | Klebsiella Spp N=9 | Acentoebacter spp N=4 | Enterobacter Spp N=3 | Pesudomonas Spp N=1 |
|----|------------------------|--------------|------------------|-----------------------|----------------------|---------------------|
|    |                        | S | R  | S | R  | S | R  | S | R  | S | R  |
| 1  | Cefoxitin              | 30 | 70 | 30 | 70 | 50 | 50 | 30 | 70 | -  | -  |
| 2  | Cefitrixon             | 40 | 60 | 70 | 30 | 50 | 50 | 30 | 70 | 100 | -  |
| 3  | Cefipim                | 20 | 80 | 80 | 20 | 75 | 25 | 100 | -  | -  | -  |
| 4  | Imepnem                | 90 | 10 | 70 | 30 | 100 | -  | -  | 100 | -  | -  |
| 5  | Azetronam              | 60 | 40 | 60 | 40 | 75 | 25 | 50 | 70 | -  | 100 |
| 6  | Amikacin               | 60 | 40 | 60 | 40 | 50 | 50 | 100 | -  | -  | -  |
| 7  | Gentamycin             | 80 | 20 | 70 | 30 | 75 | 25 | 100 | -  | -  | -  |
| 8  | Pipracillin            | 30 | 70 | 60 | 40 | -  | 100 | -  | 100 | -  | -  |
| 9  | Ampcillin              | 20 | 80 | 20 | 80 | -  | 100 | 30 | 70 | -  | 100 |
| 10 | Ciprofluxacin          | 70 | 30 | 70 | 30 | 50 | 50 | 70 | 30 | 100 | -  |
| 11 | Vancomycin             | 60 | 40 | 60 | 40 | 70 | 30 | 50 | 50 | 100 | -  |
| 12 | Oxacillin              | 40 | 60 | 60 | 40 | -  | 100 | 50 | 50 | -  | 100 |
| 13 | Ticarcillin            | 30 | 70 | 70 | 30 | 70 | 30 | 25 | 75 | 100 | -  |
| 14 | Augmutin               | 40 | 60 | 30 | 70 | 70 | 30 | 50 | 50 | 100 | -  |

Mcp < 0.01 (Highly significant), S=sensitive, R= resistant

Table-6: Antibiotic susceptibility patterns of gram negative bacterial isolates

| S.N. | Antibiotic | Staphylococcus epidermidisN=10 | Staphylococcus aureusN=6 | Streptococcus spp N=1 |
|------|------------|--------------------------------|-------------------------|-----------------------|
|      |            | S | R  | S | R  | S | R  |
| 1    | Cefoxitin  | 50 | 50 | 60 | 40 | -  | 100 |
| 2    | Cefitrixon | 60 | 40 | 50 | 50 | 100 | -  |
| 3    | Cefipim    | 90 | 10 | 60 | 40 | 100 | -  |
| 4    | Imepnem    | 80 | 20 | 80 | 20 | 100 | -  |
| 5    | Azetronam  | 40 | 60 | 60 | 40 | 100 | -  |
| 6    | Amikacin   | 80 | 20 | 80 | 20 | -  | 100 |
| 7    | Gentamycin | 30 | 70 | 50 | 50 | 100 | -  |
| 8    | Pipracillin| 50 | 50 | 50 | 50 | 100 | -  |
| 9    | Ampcillin  | 40 | 60 | 60 | 40 | -  | 100 |
| 10   | Ciprofluxacin| 70 | 30 | 60 | 40 | 100 | -  |
| 11   | Vancomycin | 80 | 20 | 80 | 20 | 100 | -  |
| 12   | Oxacillin  | 20 | 80 | 80 | 20 | 100 | -  |
| 13   | Ticarcillin| 60 | 40 | 50 | 50 | 100 | -  |
| 14   | Augmutin   | 70 | 30 | 60 | 40 | 100 | -  |

Mcp < 0.01 (Highly significant), S=sensitive, R= resistance

Table-7: Antibiotic susceptibility tests patterns of gram positive bacterial isolates
to gentamicin, imipenem, cefipime, amikacin, pipracillin, amoxicillin, ceftriaxone (100%) respectively (Table 6).

The patterns of gram-positive bacterial susceptibility to antibiotic is summarized in Table 7. *Staphylococcus epidermidis* were sensitive to ceftriaxone (90%), followed by imipenem, amikacin, vancomycin (80%), ciprofloxacin, amoxicillin (70%), ceftriaxone, ticarcillin (60%), pipracillin, cefoxitin (50%), ampicillin, azithromycin (40%), gentamicin (30%) and oxacillin (20%). *Staphylococcus aureus* were sensitive to imipenem, amikacin, vancomycin (80%), amoxicillin, ciprofloxacin, ampicillin, azithromycin, cefipime, cefoxitin (60%), ceftriaxone, gentamicin, ticarcillin, pipracillin (50%) and oxacillin (40%). *Streptococcus spp.* were sensitive to ceftriaxone, cefipime, imipenem, azithromycin, ciprofloxacin, amoxicillin, vancomycin (100%) respectively and resistant to cefotixin, amikacin, gentamicin, ampicillin, oxacillin, ticarcillin.

**DISCUSSION**

Occult bacteremia (OB) is major cause of morbidity and mortality among children’s in developing countries.4 Thus, identification of susceptible age group, causative organism and treatment of specific bacterial causative organism is needed for definitive treatment and proper management of disease. The goal of the present study was the evaluation of a young child with fever to identify sources of infection and antimicrobial susceptibility pattern.

1. Age and gender as a risk factor of occult bacteremia

Risk factors including age have been evaluated in children with occult bacteremia. In this study, distribution of children’s according to the age group with fever from birth to 11 years were depicted that the rate of cases decrease with increasing age. Infants (1 day–29 days) represented 61.3% while age group over 11 years represented 2.7% of study population. Earlier reports stated the higher incidences of bacteremia in febrile patients aged 0-1 months because of low immunoglobulin-G antibodies response to encapsulated bacteria.5,6 Other study also reported greater preponderance of bacteremia in infants younger than 1 month.10 The ratio of male:female ratio in study population was 1.5:1 slightly higher than study by Al-mousawi M. R, 2016 wherein the ratio between the male:female ratio was 1.35:1.

2. Focal source of infection and causative species of bacteremia

2.1 The focal bacterial infections

Focal signs and symptoms in neonates due to localized infections may be clinically imperceptible, and thus poses difficulty in differentiating generalized and blood stream infections. Often the early signs of neonatal sepsis are non-specific, such as temperature instability, difficulty in breathing, lethargy, poor feeding, and unexplained jaundice.11 Clinical assessment using a combination of symptoms and signs are useful guides to provisional diagnosis of neonatal blood infection.8 In present study, fever was found to be most common source of the focal bacterial infections in 56.0% followed by 19% of FIT cases in the selected study population. Respiratory disease syndrome (RDS), 10.0% (30/300); pyrexial kunon oregen (PUO), 8.0% (24/300); jaundice 4.0% (12/300) and neonatal jaundice (NNJ), 3.0% (9/300) also contributed to focal bacterial infection.

2.2 Causative species of bacteremia

Gram-negative bacteria were higher in proportion representing 65.3% (32/49) of bacterial isolates in accordance with study reported by Jain et al., 2015, Al-mousawi M. R, 2016. The most common gram-ve species isolated were namely *E. coli* 5.0% (15/49), most common gram-ve species isolated from blood cultures6; *Klebsiella spp.* 3.0% (9/49), earlier reported in neonatal sepsis cases in Ethiopia9; *Acinetobacterspp.* 1.3% (4/49); *Enterobacter spp.* 1.0% (3/49) and *pseudomonas spp.* 0.3% (1/49), predominant pathogen reported in case study of children undergoing transplantation.13 On the other hand, gram positive bacterial isolates constituted 34.6% (17/49) of total. *Staphylococcus epidermidis* was the commonest bacteria isolated in 3.3% of the total isolate of gram positive (10/49), *Staphylococcus aureus* 2.0% (6/49) and *Streptococci spp.* 0.3 (1/49) (p-value<0.01). *Staphylococcus epidermidis* and *Staphylococcus aureus* are reported to be present in majority of bacteremia cases because of its highly invasive nature, it’s virulence factors and ability to resist many antibiotics particularly Methicillin Resistant *S. aureus* (MRSA) leading to its widespread dispersal in hospitals and surrounding environment.14

3. Antibiotic susceptibility of bacterial isolates

The susceptibility pattern of gram positive and gram-negative organisms to the most relevant 14 antibiotics were done for 49 bacterial isolates. Patterns of bacterial susceptibility to these antibiotic is summarized in results section. In the present study Imipenem was the most effective drug against the tested gram positive and gram-negative bacteria. Similar finding has been reported by Al-mousawi (2016). Imipenem is a relatively new class of quinolone antibiotics which has recently become very common, particularly in general practice.15

**CONCLUSION**

In conclusion, occult bacteremia is the most prevalent condition among children’s less than one months. Gram-negative bacteria contributes more in bacteremia with presence of bacterial species like *E. coli, Klebsiella spp., Acinetobacter, Enterobacter spp. and pseudomonas spp.* Among gram-positive bacteria *Staphylococcus epidermidis, Staphylococcus aureus* and *Streptococci spp.* are the commonest bacteria causing bacteremia. Fever without a source is found to be a focal source of bacterial infection. Imipenem, class of quinolone antibiotic is found to be sensitive towards bacterial isolates and thus could be effective treatment strategy for management of bacteremia. However, considering the frequencies of resistant bacteria in hospitals, rapid laboratory tests such as Interleukin-8 (IL-8) and/or C reactive protein (CRP) to reduce unnecessary antibiotic therapy is needed.
REFERENCES

1. Jain, S. K., Gour, N., Nadkarni, J., & Bhatia, S. Bacteremia in Febrile Children: Its Correlation with Birth Weight, Feeding Practices, Vaccination and Malnutrition. British Journal of Medicine and Medical Research 2015;5:1131.

2. Stoll, M. L., & Rubin, L. G. Incidence of occult bacteremia among highly febrile young children in the era of the pneumococcal conjugate vaccine: a study from a Children's Hospital Emergency Department and Urgent Care Center. Archives of pediatrics & adolescent medicine 2004;158:671-675.

3. Al-mousawi, M. R. Bacterial profile and antibiogram of bacteremic Children in Karbala city, Iraq. karbala journal of pharmaceutical sciences 2016;11:131-139.

4. Aubais, H. S. Combination of C-Reactive Protein, Erythrosedimentation Rate and White blood cell for the Detection of Gram Negative Bacteremia in Children under 9 Years Old. Medical Journal of Babylon 2009;6:60-67.

5. Al-Husseiny, K. R. Bacteremia and Septicemia in the ChildrenUnder three years at Al-Nasseria Province. Al-Qadisiyah Medical Journal 2008; 4:1-10.

6. Nimri, L. F., Rawashdeh, M., & Meqdam, M. M. Bacteremia in children: etiologic agents, focal sites, and risk factors. Journal of tropical pediatrics 2001;47:356-360.

7. Naher, H. S., Al-Mrzoq, J. M., & Hasson, S. O. Gram-positive bacteremia in febrile children under two years of age in babylon province. Al-qadisiyah medical journal 2012;8:1-9.

8. Gransden, W. R., Eykyn, S. J., & Phillips, I. Septicemia in the newborn and elderly. Journal of Antimicrobial Chemothapy 1994;34:101-119.

9. Wayne, P. A. (2011). Clinical and laboratory standards institute. Performance standards for antimicrobial susceptibility testing.

10. Pantell, R. H., Newman, T. B., Bernzweig, J., Bergman, D. A., Takayama, J. I., Segal, M., & Wasserman, R. C. Management and outcomes of care of fever in early infancy. Jama 2004;291:1203-1212.

11. Shah, B. A., & Padbury, J. F. Neonatal sepsis: an old problem with new insights. Virulence 2014;5:170-178.

12. Ghiorgis, B. Neonatal sepsis in Addis Ababa, Ethiopia: a review of 151 bacteremic neonates. Ethiopian medical journal 1997;35:169-176.

13. Weinstein, M. P., Mirrett, S., Reimer, L. G., Wilson, M. L., Smith-Elekes, S., Chuard, C. R., & Reller, L. B. Controlled evaluation of BacT/Alert standard aerobic and FAN aerobic blood culture bottles for detection of bacteremia and fungemia. Journal of clinical microbiology 1995;33:978-981.

14. Nielsen, M. V., Sarpong, N., Krumkamp, R., Dekker, D., Loag, W., Amemasor, S., & Hagen, R. M. Incidence and characteristics of bacteremia among children in rural Ghana. PloS one 2012;7:44063.

15. Lee, N. Y., Lee, C. C., Huang, W. H., Tsai, K. C., Hsueh, P. R., & Ko, W. C. (2012.). Carabapenem Therapy for Bacteremia due to Extended-spectrum β-lactamase-producing Escherichia coli or Klebsiella pneumoniae: Implications of Ertapenem Susceptibility. Antimicrobial agents and chemotherapy, AAC-06301.