Past and Contemporaneous Otolith Fingerprints Reveal Potential Anthropogenic Interferences and Allows Refinement of the Population Structure of *Isopisthus parvipinnis* in the South Brazil Bight

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Abstract: Otolith geochemical signatures were important tools used to investigate the population of commercially exploited fish species. Historical and contemporary otolith samples of *Isopisthus parvipinnis*, Bigtooth corvina, an economically and ecologically important Brazilian fish species, collected in five subareas [São Paulo: North—NSP, Center—CSP and South—SSP; Paraná (PR) and Santa Catarina (SC)] of the shallow waters off the coast of the South Brazil Bight were used in this study. Univariate and multivariate statistical analyses showed spatial differences in otolith chemical composition over the years, suggesting that long-term temporal variability in oceanographic conditions, anthropogenic influence, and climate change on this coastal ecosystem influenced the geochemical signatures. Moreover, these results also confirm that *I. parvipinnis* is not a single and homogeneous fish stock in this geographic area, supporting the existence of a metapopulation structure scenario and corroborating previous studies that used alternative, complementary phenotypic tags.

Simple Summary: Otolith geochemical signatures were important tools used to investigate the population of commercially exploited fish species. Historical and contemporary otolith samples of *Isopisthus parvipinnis*, Bigtooth corvina, an economically and ecologically important Brazilian fish species, collected in five subareas [São Paulo: North—NSP, Center—CSP and South—SSP; Paraná (PR) and Santa Catarina (SC)] of the shallow waters off the coast of the South Brazil Bight were used in this study. Univariate and multivariate statistical analyses showed spatial differences in otolith chemical composition over the years, suggesting that long-term temporal variability in oceanographic conditions, anthropogenic influence, and climate change on this coastal ecosystem influenced the geochemical signatures. Moreover, these results also confirm that *I. parvipinnis* is not a single and homogeneous fish stock in this geographic area, supporting the existence of a metapopulation structure scenario and corroborating previous studies that used alternative, complementary phenotypic tags.
and (4) SC. This spatial differentiation on the geochemical signatures probably reflects the effects of long-term temporal variability in oceanographic conditions, anthropogenic influence, and climate change on this coastal ecosystem. The data also corroborate and refines the population structure scenario of *I. parvipinnis* recently described using complementary phenotypic tags.

**Keywords:** Sciaenidae; otolith fingerprints; population units; temporal shifts

### 1. Introduction

A proper understanding of the fish population structure and dynamics is essential for the rational management of a fishery [1]. Since population units may respond differently to exploitation, they must be managed separately to optimize their maximum sustainable yield [2]. Accurate knowledge of the population structure can have significant impacts on the rational and sustained management of fish stock, including the necessary adjustments in response to fishing pressure and environmental changes, which play a key role in the species’ persistence [3]. Several natural tags, such as otolith’s shape and its chemical composition, body meristic and morphometric characters, and the presence and prevalence of parasites, among others, have been commonly used in fisheries biology, providing evidence for stock discreteness [4–6]. Otolith microchemistry, for instance, can be pointed to as a successful approach not only to infer about fish population structure but also to help us to solve questions such as natal origin, migration patterns, habitat use, and connectivity, namely, where environmental heterogeneity exists [7–9]. Indeed, coastal systems embrace an inherent variability of abiotic factors that are under the influence of human action and climate change [10–12]. The Brazilian coast, in particular, encompasses a great variety of estuarine and oceanic ecosystems that can provide different chemical signatures for fish otoliths. Therefore, otoliths were successfully used to assess the estuarine dependency and habitat use of *Micropogonias furnieri* Whitemouth croaker (Sciaenidae) [13], *Centropomus parallelus* Fat snook (Centropomidae) [14], and *Cathorops spixii* Madamango sea catfish (Arriidae) [15] in the South Brazil Bight (SBB: 23° S to 29° S of latitude); connectivity and population structure of *Stegastes fuscus* Brazilian damsel (Pomacentridae) [7] and *Abudeflaf saxatilis* Sergeant-major (Pomacentridae) [8] from coastal systems of São Paulo, Paraná and Santa Catarina (25° S to 26.5° S); stock structure and life-time movements of *Chaetodipterus faber* Atlantic spadefish (Ephippidae) from Espírito Santo to Santa Catarina states (20° S to 28° S) [9,16]; and fish stocks and nursery areas of *Genidens genidens* Guri sea catfish (Arriidae) along the southeastern and southern coast of Brazil (21.6° S to 25.5° S) [17].

Drums and croakers are fishes belonging to the Sciaenidae family (order: Acanthuro- forms) with over 250 species worldwide [18], from which more than 20 can be found in the estuarine and coastal waters of the western Atlantic Ocean [19,20]. They are also one of the most important demersal fisheries resources in the shallow waters of the South Brazil Bight (SSB) [21–23]. The Bigtooth corvina, *Isopisthus parvipinnis*, is widely distributed along the western Atlantic Ocean, from Costa Rica to Brazil, being caught as bycatch of the small-scale artisanal fisheries, mainly in the Brazilian Southeast-South region [24]. It represents 6% of the artisanal discards in south and southeastern Brazil [25], and depending on its size, it can be used as bait or sold in a mixture of different fish species or individually [26]. *I. parvipinnis* is a seasonal species usually found on the inner continental shelf at 23° S [27]. *I. parvipinnis* is mainly a piscivorous fish, and its feeding regime varies with season and ontogeny [28,29]. During spring and summer, individuals sporadically move to the most open areas of the Guaratuba Bay, southern Brazil, and feed on fish and crustaceans. During this period, the spawning occurs either in the mangrove or in the open sea next to the state of Paraná. However, in autumn and winter, an important part of the population of *I. parvipinnis* enters into channels and pools, and its diet changes to become entirely based on fish [30]. The reproductive period of the species in the northern Santa Catarina (southern Brazil) occurs mainly in spring and summer, which coincides with the season closure of the...
shrimp fishery [31]. Fish population recruitment appears to occur between summer and autumn to winter in northeast Brazil [32]. The population structure of *I. parvipinnis* was recently studied in southeast–south Brazil through the study of the otolith shape [33] and body geometric morphometrics [34], revealing a complex metapopulational scenario with discrete population units.

The aim of this work was twofold. Firstly, to investigate the long-term stability of the population structure of *I. parvipinnis* in the SBB using trace elements recorded in otoliths collected in a temporal window of 43 years apart (1975 vs. 2018/2019); and, secondly, to evaluate for the first time, the spatial-temporal variations in the chemical composition of otoliths, as environmental time-lag recorders, taking into consideration the long-term effects of human activities and climate change on the marine ecosystems.

2. Materials and Methods

2.1. Fish Sampling and Otolith Preparation

Two-hundred and ninety-seven individuals [all juveniles, since their total length (TL: 66–139 mm) was less than the length at first maturity (159 mm) according to [32]] were caught from commercial and research vessels in the shallow waters of the SBB (from 23° S to 29° S) in five sampling areas: Northern (NSP), Center (CSP), and Southern (SSP) of São Paulo, Paraná (PR) and Santa Catarina (SC) (Figure 1 and Table 1). The individuals were collected (i) between September and November 1975 (*n* = 149) by the Oceanographic Institute of the University of São Paulo during the Nectonic Fauna research project (FAUNEC) and thereafter archived at the ColBIO (Coleção Biológica Edmundo Ferraz Nonato IO-USP) (for more details see [35]); and (ii) between September 2018 and May 2019 (*n* = 148) by local fishermen, preserved on ice and thereafter processed at the laboratory. For both sampling periods, an otter trawl was used.

![Figure 1. Sampling locations of *Isopisthus parvipinnis* juvenile individuals collected in 1975 (squares) and 2018/2019 (stars) in the South Brazil Bight from 23° S to 26.9° S, between São Paulo and Santa Catarina states. Legend: North of São Paulo (NSP), Center of São Paulo (CSP), South of São Paulo (SSP), Paraná (PR), Santa Catarina (SC).](image-url)
Table 1. States, locations, and their respective code, period of capture, sample size (n), total length (TL, mm) and otolith mass (OM, mg) of *Isopisthus parvipinnis* used in this study. For TL and OM, mean and standard deviation are presented. Legend: North of São Paulo (NSP), Center of São Paulo (CSP), South of São Paulo (SSP), Paraná (PR), Santa Catarina (SC).

| States          | Location | Code | Period       | n  | TL (mm)   | OM (mg) | Period       | n  | TL (mm)   | OM (mg) |
|-----------------|----------|------|--------------|----|-----------|---------|--------------|----|-----------|---------|
| São Paulo       | Ubatuba  | NSP  | Sep. and Nov.| 30 | 109 ± 12  | 26.39 ± 6.64 | Nov.         | 30 | 110 ± 11  | 30.74 ± 7.38 |
|                 | Peruíbe  | CSP  | Sep. and Nov.| 30 | 108 ± 14  | 27.89 ± 8.14 | Nov.         | 30 | 134 ± 5   | 46.29 ± 4.03 |
|                 | Cananéia | SSP  | Sep. and Nov.| 30 | 109 ± 14  | 28.97 ± 8.67 | Oct.         | 30 | 113 ± 8   | 32.95 ± 4.95 |
| Paraná          | Paranaguá| PR   | Sep. Nov.    | 30 | 109 ± 14  | 30.06 ± 9.22 | Sep.         | 30 | 112 ± 11  | 32.83 ± 7.30 |
| Santa Catarina  | Itajaí   | SC   | Sep. and Nov.| 29 | 111 ± 14  | 29.45 ± 8.77 | May          | 28 | 76 ± 10   | 11.74 ± 4.40 |

The individuals selected for otolith elemental analysis were restricted, as much as possible, to a narrow size range for each year, with the exception of SC to 2018/2019 (Table 1), to minimize phenotypic variations resulting from ontogenetic processes [36]. The fish were identified through meristic and morphometrics characters to the species level [37]. The total length (TL, 0.1 mm) and body weight (W, 0.01 g) were measured for each individual. Sagittal otoliths were carefully extracted, cleaned from organic tissues, washed with distilled water, dried and stored in plastic tubes.

The right otoliths were cleaned in an ultrasonic bath using ultrapure water (H$_2$O Milli-Q-Water: >18.2 MΩ.cm at 25 °C) for five minutes, followed by immersion in 3% (v/v) ultrapure hydrogen peroxide (H$_2$O$_2$: Honeywell Fluka, TraceSELECT ™, ≥30.0%) for 15 min to remove biological residues, and thereafter superficially decontaminated in 1% (v/v) nitric acid (HNO$_3$: Honeywell Fluka, TraceSELECT ™, ≥69.0%) for 10 s [38]. Thereafter, the otoliths were rinsed by triple immersion in ultrapure water (H$_2$O: Milli-Q-Water > 18.2 MΩ.cm at 25 °C) for five minutes, dried in a laminar flow cabinet hood, and stored in decontaminated plastic tubes [39].

2.2. Otolith Elemental Analysis

Otoliths were weighed on an analytical balance (OM, 0.0001 g), dissolved for 15 min in 0.1 mL of ultrapure nitric acid (HNO$_3$: Honeywell Fluka, TraceSELECT™, ≥69.0%), and diluted with ultrapure water (H$_2$O: Milli-Q-Water > 18.2 MΩ.cm at 25 °C) to a final volume of 5.0 mL (2% of HNO$_3$ v/v and 0.2% of TDS m/v) [9].

Multi-elemental analyses of trace elements (µg/L in liquid sample) were performed by Solution-Based Inductively Coupled Plasma Mass Spectrometry (SB-ICP-MS) using an iCAP™ Q instrument (Thermo Fisher Scientific, Bremen, Germany) equipped with a concentric glass nebulizer, a Peltier-cooled baffled cyclonic spray chamber, a standard quartz torch and a two-cone interface design (sample and skimmer nickel cones). High-purity (99.9997%) argon (Gasin II, Leça da Palmeira, Portugal) was used as the nebulizer and plasma gas. The equipment control and data acquisition were performed on Qtegra software (Thermo Fisher Scientific). Indium(115In), Scandium (45Sc), Terbium (159Tb), and Yttrium (89Y) were monitored as internal standards to minimize the effect of plasma fluctuations or different nebulizer aspiration rates among the samples [6]. The limits of detection (LOD) were calculated as the concentration corresponding to three times the standard deviation of 10 sample blanks. However, minor elements such as calcium, sodium, and strontium, because of their high concentrations in the aragonite matrix (mg/L in the liquid sample), which may precipitate in the nebulizer or overload the plasma, were determined by a Flame Atomic Absorption Spectrometry (FAAS) instrument (Perkin Elmer, Überlingen, Germany). Moreover, with low-mass otoliths, such as the otoliths of *I. parvipinnis*, and when we are already working with a small liquid sample volume (5 mL), further dilution of the samples to evaluate all the elements (minor and trace) in the ICP-MS, would necessarily lead to the loss of some trace informative elements [40]. In any case,
analytical control was performed in both techniques. In order to avoid possible sequence
effects, all samples (ICP-MS and FAAS) were analyzed randomly.

A preliminary analysis detected 20 trace elements, but the concentration of 8 of
them (75As, 111Cd, 52Cr, 98Mo, 121Sb, 82Se, 205Tl, and 66Zn) was consistently below the
LOD, and therefore they are excluded from analyses. Twelve elements were above the
LOD: 137Ba (0.032µg.L\(^{-1}\)), 43Ca (2.384µg.L\(^{-1}\)), 59Co (0.001µg.L\(^{-1}\)), 65Cu (0.020µg.L\(^{-1}\)),
7Li (0.002µg.L\(^{-1}\)), 26Mg (0.268µg.L\(^{-1}\)), 55Mn (0.010µg.L\(^{-1}\)), 23Na (0.138µg.L\(^{-1}\)), 60Ni
(0.016µg.L\(^{-1}\)), 208Pb (0.003µg.L\(^{-1}\)), 85Rb (0.005µg.L\(^{-1}\)), and 88Sr (0.006µg.L\(^{-1}\)). These
elements, commonly used for fish population assessment purposes [9,36,41], were consid-
ered useful biogeochemical tags for \(I.\ parvipinnis\). NIES CRM 22 (a fish otolith certified
reference material from the National Institute for Environmental Studies, Japan) was used
for accuracy control, with recovery values between 83% and 90%. The precision of replicate
analyses (relative standard deviation) of individual elements was, in general, below 5%.
Concentrations of trace elements, originally in µg element.L\(^{-1}\) solution, were transformed
to µg element.g\(^{-1}\) otolith, and finally to µg element.g\(^{-1}\) calcium [42].

2.3. Statistical Analysis

Prior to statistical analyses, the data were checked for normality (Shapiro–Wilk’s test),
homoscedasticity (Levene’s test), and the presence of outliers (Grubbs’ test). The relation-
ship between elemental concentration and fish size (expressed as otolith mass and used
as a covariate) was tested with Analysis of Covariance (ANCOVA). For all the element:Ca
ratios that showed a negative (Ba:Ca, Li:Ca, Mg:Ca, Mn:Ca, Na:Ca, Pb:Ca, Sr:Ca) or positive
(Co:Ca, Cu:Ca, Ni:Ca) relationship with OM (ANCOVA, \(p<0.05\)), the individual data
were weight-detrended by the subtraction of the common within-group linear slope [36].
Differences in single elemental fingerprints among locations and years were explored by a
Two-Way Analysis of Variance (Two-Way ANOVA), respectively, followed by a Tukey’s
post-hoc test if significant \((p<0.05)\). Differences in multi-elemental fingerprints among
locations and years were tested using an overall and pairwise Permutational Multivariate
Analysis of Variance (PERMANOVA) based on the Euclidean distance measure using
9999 random permutations. A Canonical Analysis of Principal Coordinates (CAP) based
on Euclidian distances was performed to visualize regional differences in each year. Vari-
ables that most contributed to each axis, based on the Pearson correlation \((r>0.50)\), were
displayed in CAP two-dimensional plots. The reclassification accuracy of the discriminant
functions for each location was evaluated through the percentage of correctly re-classified
individuals to the origin using a leave-one-out cross-validation [33].

All of the statistical analyses were performed using Past—Version 4.03 and PRIMER
7 + PERMANOVA software, with a statistical level of significance \((\alpha)\) of 0.05.

3. Results

Six element:Ca ratios (Ba:Ca, Cu:Ca, Li:Ca, Mg:Ca, Mn:Ca, and Sr:Ca) presented
significant differences among locations, between years, and for the interaction between
them ANOVA, \(p<0.05\), Table 2); but Pb:Ca only resulted in significant differences among
locations and years (Two-Way ANOVA, \(p<0.05\), Table 2). The other four element: ratios
(Co:Ca, Na:Ca, Ni:Ca and Rb:Ca) did not present any differences between years (Two-Way
ANOVA, \(p>0.05\), Table 2), but differences among locations and locations x years were
observed (Two-Way ANOVA, \(p<0.05\), Table 2).

Li:Ca and Sr:Ca showed significant differences between years in SSP, PR, and SC;
but Ba:Ca and Mg:Ca recorded temporal differences for all locations, except SC and NSP,
respectively (Tukey’s post-hoc test, \(p>0.05\); Figure 2).
Table 2: Two-Way ANOVA comparisons among the five sampling locations regarding the chemical composition of *Isopisthus parvipinnis* otoliths collected in 1975 and 2018/2019. Statistically significant differences (\(p < 0.05\)) were marked in bold.

|          | Ba:Ca                  | Co:Ca                  |
|----------|------------------------|------------------------|
|          | DF | SS   | MS  | F    | p   | DF | SS   | MS  | F    | p   |
| Year     | 1  | 29,477 | 29,477 | 174.20 | <0.000 | 1  | 0.0205 | 0.0205 | 0.3514 | 0.5538 |
| Location | 4  | 17,719 | 4429.7 | 26.180 | <0.000 | 4  | 29.862 | 7.4654 | 128.20 | <0.000  |
| Interaction | 4  | 14,975 | 3743.7 | 22.130 | <0.000 | 4  | 2.8357 | 0.7089 | 12.180 | <0.000  |
| Within   | 287 | 48,560 | 169.20 | 287 | 16.706 | 0.0582 |
| Total    | 296 | 110,568 | 49.435 | |

|          | Cu:Ca                  | Li:Ca                  |
|----------|------------------------|------------------------|
|          | DF | SS   | MS  | F    | p   | DF | SS   | MS  | F    | p   |
| Year     | 1  | 6.8509 | 6.8509 | 45.080 | <0.000 | 1  | 0.2463 | 0.246 | 31.740 | <0.000  |
| Location | 4  | 4.2744 | 1.0686 | 7.0320 | <0.000 | 4  | 0.3217 | 0.080 | 10.370 | <0.000  |
| Interaction | 4  | 24.288 | 6.0720 | 39.960 | <0.000 | 4  | 0.1510 | 0.038 | 4.8650 | <0.000  |
| Within   | 271 | 41.184 | 0.1520 | 286 | 2.2191 | 0.008 |
| Total    | 280 | 77.142 | 2.9420 | |

|          | Mg:Ca                  | Mn:Ca                  |
|----------|------------------------|------------------------|
|          | DF | SS   | MS  | F    | p   | DF | SS   | MS  | F    | p   |
| Year     | 1  | 9181.7 | 9181.7 | 284.800 | <0.000 | 1 | 1851.8 | 1851.8 | 101.40 | <0.000  |
| Location | 4  | 9145.9 | 2286.5 | 70.910 | <0.000 | 4 | 2494.5 | 623.62 | 34.160 | <0.000  |
| Interaction | 4  | 5683.1 | 1420.8 | 44.060 | <0.000 | 4 | 1708.1 | 427.03 | 23.390 | <0.000  |
| Within   | 287 | 9253.7 | 32.243 | 287 | 5239.6 | 18.256 |
| Total    | 296 | 33,363 | 11,323 | |

|          | Na:Ca                  | Ni:Ca                  |
|----------|------------------------|------------------------|
|          | DF | SS   | MS  | F    | p   | DF | SS   | MS  | F    | p   |
| Year     | 1  | 2.58 × 10^5 | 2.58 × 10^5 | 1.4630 | 0.2275 | 1 | 7.3276 | 7.3276 | 0.6257 | 0.4296 |
| Location | 4  | 6.40 × 10^6 | 1.60 × 10^6 | 9.0870 | <0.000 | 4 | 3935.9 | 983.97 | 84.020 | <0.000  |
| Interaction | 4  | 6.60 × 10^6 | 1.65 × 10^6 | 9.3770 | <0.000 | 4 | 241.86 | 60.465 | 5.1630 | <0.000  |
| Within   | 287 | 5.05 × 10^7 | 1.76 × 10^7 | 287 | 3361.3 | 11.712 |
| Total    | 296 | 6.38 × 10^7 | 7548.7 | |

|          | Pb:Ca                  | Rb:Ca                  |
|----------|------------------------|------------------------|
|          | DF | SS   | MS  | F    | p   | DF | SS   | MS  | F    | p   |
| Year     | 1  | 3.3757 | 3.3757 | 788.8 | <0.000 | 1 | 0.0010 | 0.0010 | 2.4540 | 0.1183 |
| Location | 4  | 0.4424 | 0.1106 | 25.84 | <0.000 | 4 | 0.0298 | 0.0074 | 17.910 | <0.000  |
| Interaction | 4  | 0.0370 | 0.0093 | 2.163 | 0.0732 | 4 | 0.0060 | 0.0015 | 3.5890 | <0.000  |
| Within   | 287 | 1.2283 | 0.0043 | 287 | 0.1192 | 0.0004 |
| Total    | 296 | 5.0685 | 0.1560 | |

|          | Sr:Ca                  |
|----------|------------------------|
|          | DF | SS   | MS  | F    | p   |
| Year     | 1  | 7.18 × 10^6 | 7.18 × 10^6 | 53.850 | <0.000 |
| Location | 4  | 1.97 × 10^7 | 4.92 × 10^6 | 36.890 | <0.000  |
| Interaction | 4  | 3.37 × 10^6 | 8.42 × 10^5 | 6.3160 | <0.000  |
| Within   | 287 | 3.83 × 10^7 | 1.33 × 10^5 | |
| Total    | 296 | 6.86 × 10^7 | | |
Regarding the year 1975, six element:Ca ratios distinguished all locations from SC. The former location presented the highest levels of Mg:Ca and Pb:Ca, and the lowest levels of Co:Ca, Cu:Ca, Na:Ca, and Ni:Ca. Moreover, Li:Ca was significantly higher in NSP than in other locations. Ba:Ca and Sr:Ca presented the same patterns, with higher values in SSP and PR. A slight increase in Rb:Ca southwards was also observed. However, Mn:Ca did not show any pattern (Tukey’s post-hoc test, \( p > 0.05 \); Figure 2). For 2018/2019, Ba:Ca, Mn:Ca, and Rb:Ca presented values significantly higher in SC compared to other locations, while Co:Ca and Ni:Ca presented the lowest values in the same region; Li:Ca was significantly higher in NSP, PR, and SC; Mg:Ca was able to differentiate among locations, with a latitudinal increase in concentration from NSP to SC; Cu:Ca showed values significantly lower for NSP/SSP/PR than for CSP and SC; and Na:Ca presented significant differences between NSP, CSP and SSP/PR/SC; Pb:Ca was only detected in CSP and SC, with much lower values compared to 1975; and Sr:Ca presented the same pattern observed in 1975, with higher values in SSP and PR (Tukey’s post-hoc test, \( p < 0.05 \); Figure 2).

Regarding the multi-elemental fingerprints, the PERMANOVA analysis showed significant differences between years and among sampling locations and also detected a significant interaction between both factors (overall PERMANOVA, \( p < 0.05 \); Table 3). Pairwise analysis showed significant differences among all locations for both years, except between SSP and PR, also for both years’ comparisons (pairwise PERMANOVA, \( p > 0.05 \), Table 3).

Regarding the historical samples, CAP showed some overlap between samples from SP and PR but a clear distinction between SC samples (Figure 3, 1975). Vector overlays indicated that group separation was primarily driven by Ni:Ca (\( r = -0.94 \)), Co:Ca (\( r = -0.93 \)) and Cu:Ca (\( r = -0.54 \)) on CAP Axis 1, and by Sr:Ca (\( r = -0.91 \)) and Ba:Ca (\( r = -0.83 \)) on CAP Axis 2. The leave-one-out cross-validation presented a low overall reclassification
success of 58%, highlighting the SC samples that were all fully reallocated to the original location (100%) and the NSP samples that reached 70% of correct reclassification, contrary to CSP and SSP (Table 4).

Table 3. Overall and pairwise PERMANOVA comparisons among the five sampling locations regarding the chemical composition of *Isopisthus parvipinnis* otoliths collected in 1975 and 2018/2019. Significant statistical differences (*p* < 0.05) were marked in bold. Legend: North of São Paulo (NSP), Center of São Paulo (CSP), South of São Paulo (SSP), Paraná (PR), Santa Catarina (SC).

| Source          | SS         | DF | MS        | F          | p     | NSP       | CSP       | SSP       | PR     | SC     | % Correct |
|-----------------|------------|----|-----------|------------|-------|-----------|-----------|-----------|--------|--------|-----------|
| **Overall PERMANOVA** |            |    |           |            |       |           |           |           |        |        | 2018/2019 |
| Year            | 9.99 × 10⁶ | 1  | 9.99 × 10⁶| 30.100     | 0.0001 | NSP 0.0152| CSP 0.0001| SSP 0.0002| PR 0.0028| SC 0.0001|           |
| Location        | 2.74 × 10⁷ | 4  | 6.86 × 10⁶| 20.676     | 0.0001 | CSP 0.0001| SSP 0.0002| PR 0.9031 | SC 0.0001|        |           |
| Interaction     | 8.23 × 10⁶ | 4  | 2.06 × 10⁶| 6.1994     | 0.0001 | SSP 0.0002| PR 0.9367 | SC 0.0001|        |        | 1975      |
| Residual        | 9.54 × 10⁷ | 287| 3.32 × 10⁵| 9.54 × 10⁶ | 0.0001 | PR 0.0001 | SC 0.0005 |        |        |        | 2018/2019 |
| Total           | 1.40 × 10⁸ | 296|           | 1.40 × 10⁸ |       |           |           |           |        |        |           |

Table 4. Jackknifed cross-validation reclassification matrices obtained from otolith’s elemental composition of *Isopisthus parvipinnis* from all sampling locations for the years 1975 and 2018/2019. Legend: North of São Paulo (NSP), Center of São Paulo (CSP), South of São Paulo (SSP), Paraná (PR), Santa Catarina (SC).

| Original Location | NSP | CSP | Predicted Location | SSP | PR | SC | % Correct |
|-------------------|-----|-----|--------------------|-----|----|----|----------|
| NSP               | 21  | 7   | 0                  | 2   | 0  | 0  | 70       |
| CSP               | 12  | 9   | 5                  | 4   | 0  | 0  | 30       |
| SSP               | 0   | 7   | 13                 | 10  | 0  | 0  | 43       |
| PR                | 3   | 1   | 11                 | 15  | 0  | 0  | 50       |
| SC                | 0   | 0   | 0                  | 0   | 29 | 0  | 58       |
| **Total**         |     |     |                    |     |    |    | 100      |
Table 4. Cont.

| Original Location | 2018/2019 | Predicted Location | % Correct |
|-------------------|-----------|--------------------|-----------|
|                   | NSP       | CSP    | SSP    | PR    | SC     |         |
| NSP               | 29        | 0      | 0      | 1     | 0      | 97       |
| CSP               | 4         | 26     | 0      | 0     | 0      | 87       |
| SSP               | 0         | 2      | 18     | 10    | 0      | 60       |
| PR                | 1         | 2      | 10     | 17    | 0      | 57       |
| SC                | 0         | 0      | 0      | 0     | 28     | 100      |
| **Total**         | **80**    |        |        |       |        |          |

The overall reclassification success increased to 80% in 2018/2019, with reclassification percentages increased for all sampling locations, mainly in NSP (97%) and CSP (87%, Table 4). Again, samples from SC were all fully reclassified (100%), while SSP and PR samples were reallocated mainly between them. CAP results were similar again, with the three locations from São Paulo being well distinguished but with an overlap between SSP and PR samples (Figure 3, 2018/2019). Vector overlays indicated that group separation was primarily driven by Mg:Ca ($r = -0.64$), Mn:Ca ($r = -0.71$), Rb:Ca (Pearson, $r = -0.66$), Cu:Ca (Pearson, $r = -0.68$), Ba:Ca (Pearson, $r = -0.82$), Ni:Ca (Pearson, $r = 0.56$), Co:Ca (Pearson, $r = 0.59$), and Sr:Ca (Pearson, $r = 0.35$) on CAP Axis 1, and by Mg:Ca (Pearson, $r = 0.62$), Cu:Ca (Pearson, $r = -0.37$), Sr:Ca (Pearson, $r = 0.37$) on CAP Axis 2.3.1.

4. Discussion

The otolith elemental composition of *I. parvipinnis* recorded significant regional and temporal variations. All single element: Ca ratios showed significant regional differences within years, but not necessarily for all locations. Regarding the multivariate analysis, the correct reallocation of individuals to the original locations was smaller in 1975 (overall 58%) compared to 2018/2019 (overall 80%), where an increment for the reclassification rates was observed for all locations. Moreover, the SC samples reached a full reclassification success of 100% in both periods. The coastal zone of SC, the southernmost region of the study area, is under the influence of important oceanographic processes: wind-driven South Atlantic Central Water intrusions towards the coast take place in a large portion of the shelf, nearly 28.5° S [43]; and the cold (14–17°C) and less salty (33.0–34.0) water transported by the Brazil Coastal Current, consisted of waters from the Argentina continental shelf, near the Rio de la Plata mouth, and the Brazil-Malvinas Confluence [43–45]. As a consequence, SC waters are nutrient-enriched, being classified as mesotrophic, reaching eutrophic conditions during some periods of the year [46,47]. These oceanographic characteristics are very different from the other areas, such as NSP, in which the waters are oligotrophic, which could be reflected in the element:Ca ratios signatures and consequently in the 100% reclassification success for SC.

Taking into consideration the elemental signatures recorded in 1975, two population units were recorded: one including SP and PR states but already suggesting an initial segregation between NSP/CSP and SSP/PR, and SC, a fully isolated population. Similar results were already observed in the past (in the 1970s and 1980s) for other sciaenid species in the SSB inferred from the body and/or otolith shape (e.g., *Cynoscion jamaicensis*, [48,49]; *Macrodon ancylodon*, [50]; *Micropogonias furnieri*, [51]; *Nebris microps*, [52]). However, at present (2018/2019), and 43 years later, four main population units, NSP, CSP, SSP/PR, and SC, were clearly depicted regarding *I. parvipinnis* population structure, as recently suggested by previous studies using alternative approaches [33,34].

The study area, with about 700 km of coastline, covers a wide variety of aquatic ecosystems under the influence of different oceanographic features and anthropogenic activities [53]. NSP and SC are exposed to seasonal intrusions toward the coast by the cold oceanic South Atlantic Central Water (SACW), a water mass transported by the Brazil Current along the Brazilian continental slope [54,55]; CSP is under coastal events,
and minor rivers influence; and SSP and PR are under the influence of great estuarine ecosystems that have changed over time as a result of intense human activities [56,57]. These environmental factors can lead to regional geochemical differences or similarities between fish populations [6,9,58]. Moreover, fish physiological dynamics and experiential life-history traits could result in population-specific differences in otolith chemical composition, even in chemically homogeneous conditions [59,60]. Although some general hydrographic processes remain unchanged over time, such as the SACW intrusions, the biotic integrity of SBB’s coastal ecosystems is changing [61–63]. Therefore, it would be challenging to pinpoint all the causes behind the regional (i.e., spatial) and temporal otolith chemistry variability recorded in the hereby study. Additionally, the incorporation of minor and trace elements in fish otoliths depends on many factors, including their concentration and bioavailability in water, physiological processes, individual somatic growth, and the affinity of the aragonite matrix otolith for the different elements [60,64].

All element:Ca ratios presented significant regional differences in 2018/2019, limited to Ba:Ca, Co:Ca, Cu:Ca, Ni:Ca, and Sr:Ca in 1975. Barium (Ba) and strontium (Sr) are considered the elements that best represent the surrounding water composition since they may pass through calcium channels, directly replacing them in the calcium carbonate matrix, which results in higher abundances in otoliths [60]. These elements usually present a direct relationship with salinity, negative for Ba:Ca and positive for Sr:Ca, which could explain higher values of Ba:Ca in SSP and PR, but not the Sr:Ca patterns observed for both periods, which seems to be also influenced by ontogenetic processes, such as growth and spawning [65–67]. Likewise, sodium (Na) is also essential for cellular processes (e.g., N, K—ATPase), with high physiological response in its regulation, but the influence of environmental factors in the otolith absorption remains somewhat unclear [36,68,69], as well as its profile in this study area. Lithium (Li) incorporation into biogenic calcium carbonates involves little biological control, also replacing the Ca in the aragonite/calcite matrices directly [7,70]; thus, Li:Ca is tied to environmental influences such as salinity, temperature, upwelling conditions, or primary productivity, rather than to otolith specificities [59,71–73]. So, a possible explanation for higher Li:Ca values in NSP and SC in 1975 and 2018/2019, respectively, could be related to the new primary production accompanying upwelling events in these locations. The presence of cobalt (Co) and nickel (Ni), as well as zinc (Zn) and lead (Pb), is primarily through their physiological roles as biomolecule co-factors rather than as a result of the environment [74]; regardless, Co:Ca and Ni:Ca, were significantly lower in both periods for SC. Magnesium (Mg) is unrelated to either temperature or salinity in marine fish otoliths [75], but it is negatively correlated with metabolic processes and growth rate [59,72]; therefore, it is expected that changes in metabolic rates caused by environmental fluctuations (e.g., temperature and food resources variations) could cause alterations in elemental assimilation rates [72]. Studies about manganese (Mn) are still contradictory. It is unclear if it reflects changes in the physicochemical environment experienced during life as a substitute for calcium [74] or if it reflects physiological events, such as maternal transfer since it is a co-factor of the protein FAM20C found in the primordium [76]. Mg and Mn presented similar patterns between years and among areas, with the exception of NSP, which is probably related to environmental conditions and/or physiological events. Rubidium (Rb) usually presents a negative relationship with salinity that was not observed in the I. parvipinnis data, possibly due to interactions between Rb, salinity, and physiological processes (e.g., ionic balance or osmoregulatory pathways), interchanging it with K+, that may render Rb more or less available for incorporation into the otolith [71,77,78]. Copper (Cu) is involved in the activity of many essential enzymes and is required in oocyte formation in vertebrates [65]; nevertheless, peaks of Cu and Pb coinciding with the most industrialized locations and under intense port activity (such as CSP and SC in the present study) could be indicative of environmental contamination [79,80].

Unlike other chemical elements, such as Cu, Zn, Ni, Co, and Cr, which are essential in trace amounts but toxic in higher doses, Pb has no known nutritive function in fish, but it is toxic at low levels interfering with essential nutrients of similar characteristics as Ca and
Zn [80,81]. However, lead is a trace metal less frequently investigated due to its analytical challenges for quantification [60]. Higher Pb concentrations in 1975 could be a result of the intensified occupation of the coast of the SBB, beginning in the 1950s. Since then, numerous human activities have occurred in the study area, such as sandy extraction from beaches and dunes, modification of river channels, canal construction, dredging, construction of sea walls, and contamination from different sources such as port activity, industries, and domestic sewage discharge [12,82–85].

The low levels of Pb observed in 2018/2019 compared to 1975 in *I. parvipinnis* otoliths could be the result of the environmental policies that resulted in a worldwide decrease in Pb levels since the early 1980’s due to reduced consumption of leaded petrol [86]. Those temporal variations in Pb concentration were also registered in sediment cores: a reduction in SSP and PR from the 1970s to 2000–2010s [87,88] in contrast to an increase in CSP [89]. The literature also shows the anthropogenic impacts of oil exposure, industrial, gasoline, sewage, and agriculture input not only in Pb concentration in otoliths but also in Ba, Mg, Mn, Na, Sr, and Zn [90–94]. Anthropogenic climate changes, mainly after the 1950s, are also affecting marine biodiversity, ecosystems, fisheries, and ecosystem services [95–97]. The literature has already reported evidence of an increase in sea surface temperature in the South Atlantic Ocean mainly after the 1980s [12,98] and its consequences in the ichthyofauna of a transitional zone between the tropical and subtropical regions in Southeastern Brazil [12]. Ocean acidification, warming, and deoxygenation resulting from climate change could influence the ecophysiology of marine organisms and the abundance, distribution, and composition of fish communities [10,99,100]. Once several physiological processes (e.g., ion transport, homeostasis, osmoregulation, growth, or gonad development) are often distressed by those environmental changes, otolith growth and chemistry may also respond to anthropogenic climate changes [59,60,101], explaining the observed long-term changes in the geochemical signatures of *I. parvipinnis*.

Alternative research methods can provide different scenarios about the population structure of marine fishes. A holistic approach is recommended to deal with species and/or geographic areas whose characteristics are not fully understood. An improvement in the reclassification rates has been observed when using otolith elemental signatures compared to otolith shape for several species [9,41,102]. Regarding *I. parvipinnis*, those rates slightly increased from whole-body morphometry (79% in 2018/2019; [34]) to otoliths shape (42% in 1975, and 81% in 2018/2019; [33]), and, finally, to otolith chemical composition (58% in 1975, and 80% in 2018/2019; present study). Moreover, compared with the previous data from otolith shape [33], the present study allowed the detection of significant differences between SC and the other locations in both periods, clearly detached it as an isolated population unit for both periods, and increased the regional cross-validation reclassification success.

5. Conclusions

The multi-elemental otolith approach showed clear regional differences between *I. parvipinnis* individuals indicating the presence of distinct population connectivity of the SC population with the populations located northwards and the decrease in the degree of adult populations mixing along the SBB over time were also recorded, recommending regional fishery management for the species. The present study also recorded a noticeable anthropogenic interference in fish otoliths fingerprints over a large temporal scale without excluding the potential effects of climate change.

Author Contributions: Conceptualization, N.T.H. and A.T.C.; methodology, N.T.H., E.P., A.A., R.S. and A.T.C.; formal analysis, N.T.H. and A.T.C.; investigation, N.T.H. and A.T.C.; resources, J.F.D. and A.T.C.; writing—original draft preparation, N.T.H. and A.T.C.; writing—review and editing, N.T.H., J.F.D. and A.T.C.; supervision, A.T.C.; funding acquisition, A.T.C. All authors have read and agreed to the published version of the manuscript.
**Funding:** This research is part of a PhD scholarship funded by CNPq (163116/2015-6) and PDSE—CAPES (88881.189862/2018-01). This study was supported by national funds through FCT—Foundation for Science and Technology within the scope of UIDB/04423/2020 and UIDP/04423/2020.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** NTH thanks to Lucy Satiko Hashimoto Soares and Edmund F. Nonato Biological Collection (IO-USP) for historical data; Sylvio da Conceição, José Roberto Marques, Leandro Martins (IO-USP), Marcelo Soeth (CEM—UFPR), Cláudio Henrique B. Silva, and Eliana Diniz (president from Fishermen Colony Z5—Peruíbe) for sampling support.

**Conflicts of Interest:** The authors declare no conflict of interest.

**References**

1. Begg, G.; Brown, R. Stock identification of Haddock Melanogrammus aeglefinus on Georges Bank based on otolith shape analysis. *Trans. Am. Fish. Soc.* 2000, 129, 935–945. [CrossRef]

2. Galli, O.; Norbis, W. Morphometric and meristic spatial differences and mixed groups of the whitemouth croaker (Micropogonias furnieri (Demearest, 1823)) during the spawning season: Implications for management. *J. Appl. Ichthyol.* 2013, 29, 782–788. [CrossRef]

3. Kerr, L.; Hintzen, N.; Cadrin, S.; Clausen, L.; Worsoedickey-Collas, M.; Goethel, D.; Hatfield, E.; Kritzer, J.; Nash, R. Lessons learned from practical approaches to reconcile mismatches between biological population structure and stock units of marine fish. *ICES J. Mar. Sci.* 2017, 74, 1708–1722. [CrossRef]

4. Poulin, R.; Kamiya, T. Parasites as biological tags of fish stocks: A meta-analysis of their discriminatory power. *Parasitology 2013*, 142, 145–155. [CrossRef] [PubMed]

5. Moreira, C.; Froufe, E.; Vaz-Pires, P.; Triay-Portella, R.; Correia, A.T. Landmark-based geometric morphometrics analysis of body shape variation among populations of the blue jack mackerel, Trachurus picturatus, from the North-East Atlantic. *J. Sea Res.* 2020, 163, 101926. [CrossRef]

6. Moura, A.; Muniz, A.A.; Mullins, R.; Wilson, J.M.; Vieira, R.P.; Almeida, A.A.; Pinto, E.; Brummer, G.J.A.; Gaever, P.V.; Gon-çalves, J.M.S.; et al. Population structure and dynamics of the Atlantic mackerel (*Scomber scombrus*) in the North-Atlantic inferred from otolith chemical and shape signatures. *Fish. Res.* 2020, 230, 105621. [CrossRef]

7. Daros, F.A.; Spach, H.L.; Sial, A.N.; Correia, A.T. Otolith fingerprints of the coral reef fish Stegastes fuscus in southeast Brazil: A useful tool for population and connectivity studies. *Reg. Stud. Mar. Sci.* 2016, 3, 262–272. [CrossRef]

8. Adelir-Alves, J.; Daros, F.; Spach, H.L.; Soeth, M.; Correia, A.T. Otoliths as a tool to study reef fish population structure from coastal islands of South Brazil. *Mar. Biol. Res.* 2018, 14, 973–988. [CrossRef]

9. Soeth, M.; Spach, H.L.; Daros, F.; Adelir-Alves, J.; de Almeida, A.C.O.; Correia, A.T. Stock structure of Atlantic spadefish Chaetodipterus faber from Southwest Atlantic Ocean inferred from otolith elemental and shape signatures. *Fish. Res.* 2019, 211, 81–90. [CrossRef]

10. Cheung, W.W.L.; Sarmiento, J.L.; Dunne, J.P.; Frölicher, T.L.; Lam, VW.Y.; Deng Palomares, M.L.; Watson, R.; Pauly, D. Shrinking water mass associations of settlement-stage rockfishes. *Mar. Ecol. Prog. Ser.* 2013, 430, 251–258. [CrossRef]

11. Wheeler, S.; Russell, A.; Fehrenbacher, J.; Morgan, S. Evaluating chemical signatures in a coastal upwelling region to reconstruct water mass associations of settlement-stage rockfishes. *Mar. Ecol. Prog. Ser.* 2016, 550, 191–206. [CrossRef]

12. Araújo, F.G.; Teixeira, T.P.; Guedes, A.P.P.; de Azevedo, M.C.C.; Pessanha, A.L.M. Shifts in the abundance and distribution of shallow water fish fauna on the southeastern Brazilian coast: A response to climate change. *Hydrobiologia* 2018, 814, 205–218. [CrossRef]

13. Albuquerque, C.Q.; Miekeley, N.; Muller, J.H.; Walther, B.D.; Jaureguizar, A.J. Estuarine dependency in a marine fish evaluated with otolith chemistry. *Mar. Biol.* 2012, 159, 2229–2239. [CrossRef]

14. Daros, F.A.; Spach, H.L.; Correia, A.T. Habitat residency and movement patterns of *Centropomus parallelus* juveniles in a subtropical estuarine complex. *J. Fish Biol.* 2016, 88, 1796–1810. [CrossRef]

15. Carvalho, B.M.; Pupo, D.V.; Volpedo, A.V.; Pisonero, J.; Méndez, A.; Avigliano, E. Spatial environmental variability of nat-ural markers and habitat use of *Cathorops spixii* in a neotropical estuary from otolith chemistry. *J. Mar. Biol. Assoc. UK* 2020, 100, 783–793. [CrossRef]

16. Soeth, M.; Spach, H.L.; Daros, F.A.; Jorge Pisonero, J.; Correia, A.T. Use of otolith elemental signatures to unravel lifetime movements patterns of Atlantic spadefish, Chaetodipterus faber, in the Southwest Atlantic Ocean. *J. Sea Res.* 2020, 158, 101873. [CrossRef]

17. Maciel, T.R.; Avigliano, E.; de Carvalho, B.M.; Miller, N.; Vianna, M. Population structure and habitat connectivity of *Genidens genidens* (Siluriformes) in tropical and subtropical coasts from Southwestern Atlantic. *Estuar. Coast. Shelf Sci.* 2020, 242, 106839. [CrossRef]
18. Menezes, N. Checklist dos peixes marinhos do Estado de São Paulo, Brasil. *Bol. Inst. Oceanogr.* 1980, 37, 95–105. [CrossRef]

19. Santos, M.N.; Rocha, G.R.A.; Freire, K.M.F. Diet composition for three sciaenids caught off northeastern Brazil. *Rev. Biol. Mar. Oceanogr.* 2016, 51, 493–504. [CrossRef]

20. Chaves, P.; Rickli, A.; Bouchereau, J. Spatio-temporal evaluation of the population structure of the bigtooth corvina *Isopisthus parvipinnis* (Teleostei: Sciaenidae) in the continental shelf of the Southeastern Brazilian Bight. In *Biota Neotrop*, 2011; 10, 69–86. [CrossRef]

21. Haimovich, M.; Ignacio, J.M. Pesca da corvina no sul do Brasil. In *Análise das Principais Pescarias Comerciais da Região Sudeste-Sul do Brasil: Dinâmica Populacional das Espécies em Exploração, Série Documentos Revizoe-Score Sul*; Rossi-Wongtschowski, C.L.B., Cergole, M.C., Avila-da-Silva, A.O., Eds.; IOUSP: São Paulo, Brazil, 2005, pp. 101–107.

22. Souza, U.P.; Costa, R.C.; Martins, I.A.; Fransozo, A. Associações entre as biomassas de peixes Sciaenidae (Teleostei: Perci-formes) e de camarões Penaeoida (Decapoda: Dendrobranchiata) no litoral norte do Estado de São Paulo. *Biota Neotrop.* 2008, 8, 83–92. [CrossRef]

23. Souza, U.P.; Soares, L.S.H. Alimentação de Isopisthus parvipinnis (Teleostei: Sciaenidae) na Baía de Santos, São Paulo. *Bol. Inst. Oceanogr.* 1989, 37, 95–105. [CrossRef]

24. Aguiar Soccorro, O. 2020. *Isopisthus parvipinnis*. The IUCN Red List of Threatened Species 2020: e.T47147702A82679809. Available online: https://www.iucnredlist.org/species/47147702/82679809 (accessed on 3 June 2022).

25. Freire, K.M.F.; Pauly, D. Fisheries catch reconstructions for Brazil’s mainland and oceanic islands. *Fish. Cent. Res. Rep.* 2015, 23, 47.

26. Graça Lopes, R.; Tomé, V.;Freire, K.M.F.; Rossi-Wongtschowski, C.L.B., Paes, E. Padrões espaciais e temporais da comunidade de peixes demersais no litoral norte do Estado de São Paulo—Ubatuba, Brasil. *Rev. Inst. Pesca.* 2002, 28, 173–188.

27. Rossi-Wongtschowski, C.L.B.; Paes, E. Padrões espaciais e temporais da comunidade de peixes demersais no litoral norte do Estado de São Paulo—Ubatuba, Brasil. *Inst. Oceanogr.* 1993, 10, 169–188.

28. Soares, L.S.H. Alimentação de *Isopisthus parvipinnis* (Teleostei: Sciaenidae) na Baía de Santos, São Paulo. *Bol. Inst. Oceanogr.* 1989, 37, 95–105. [CrossRef]

29. Santos, M.N.; Rocha, G.R.A.; Freire, K.M.F. Diet composition for three sciaenids caught off northeastern Brazil. *Rev. Biol. Mar. Oceanogr.* 2016, 51, 493–504. [CrossRef]

30. Chaves, P.; Rickli, A.; Bouchereau, J. Spatio-temporal evaluation of the population structure of the bigtooth corvina *Isopisthus parvipinnis* (Teleostei: Sciaenidae) in the continental shelf of the Southeastern Brazilian Bight. In *Biota Neotrop*, 2011; 10, 69–86. [CrossRef]

31. Souza, U.P.; Soares, L.S.H. Alimentação de Isopisthus parvipinnis (Teleostei: Sciaenidae) na Baía de Santos, São Paulo. *Bol. Inst. Oceanogr.* 1989, 37, 95–105. [CrossRef]

32. Romero, R.; Moraes, L.; Santos, M.; Rocha, G.; Cetra, M. Biology of Isopisthus parvipinnis: An abundant sciaenid species cap-tured bycatch during sea-bob shrimp fishery in Brazil. *Neotrop. Ichthyol.* 2006, 8, 67–74. [CrossRef]

33. Hoff, N.T.; Dias, J.F.; Zani-Teixeira, M.D.L.; Correia, A.T. Spatio-temporal evaluation of the population structure of the bigtooth corvina *Isopisthus parvipinnis* from the Southwest Atlantic Ocean using otolith shape signatures. *J. Appl. Ichthyol.* 2020, 36, 439–450. [CrossRef]

34. Hoff, N.T.; Dias, J.F.; Zani-Teixeira, M.L.; Soeth, M.; Correia, A.T. Population structure of the bigtooth corvina *Isopisthus parvipinnis* from the Southwest Atlantic Ocean as determined by whole body morphology. *Reg. Stud. Mar. Sci.* 2020, 39, 101379. [CrossRef]

35. Lamas, R.A.; Soares, L.S.H. Isopisthus parvipinnis (Cuvier, 1830) at the continental shelf of the Southeastern Brazilian Bight. In *Growth of Fisheries Resources from the Southwestern Atlantic*, 1st ed.; Vaz-dos-Santos, A.M., Rossi-Wongtschowski, C.L.B., Eds.; IOUSP: São Paulo, Brazil, 2019; pp. 133–135.

36. Campana, S.; Chouinard, G.; Hanson, J.; Frechet, A.; Brattey, J. Otolith elemental fingerprints as biological tracers of fish stocks. *Fish. Res.* 2000, 46, 343–357. [CrossRef]

37. Menezes, N.; Figueiredo, J. *Manual de Peixes Marinhos do Sudeste do Brasil. IV. Actinopterygii* (3); Museu de Zoolo gia da Universidade de São Paulo: São Paulo, Brasil, 1980.

38. Stocker, J.R.; Zdanowicz, V.S.; Secor, D.H. Chemistry of tuna otoliths: Assessment of base composition and postmortem handling effects. *Mar. Biol.* 2001, 139, 35–43.

39. Patterson, H.M.; Thorrold, S.R.; Shenker, J. Analysis of otolith chemistry in Nassau grouper (Epinephelus striatus) from the Bahamas and Belize using solution-based ICP-MS. *Coral Reefs* 1999, 18, 171–178. [CrossRef]

40. Correa, A.; Moura, A.; Triay-Portella, R.; Santos, P.; Pinto, E.; Almeida, A.; Sial, A.; Muniz, A. Population structure of the chub mackerel *Scomber colias* captured bycatch during sea-bob shrimp fishery in Brazil. *Bol. Inst. Oceanogr.* 2002, 28, 173–188.

41. Moreira, C.; Froufe, E.; Sial, A.; Caeiro, A.; Vaz-Pires, P.; Correia, A. Population structure of the blue jack mackerel (Trachurus picturatus) in the NE Atlantic inferred from otolith microchemistry. *Fish. Res.* 2018, 197, 113–122. [CrossRef]

42. Higgins, R.; Isidro, E.; Menezes, G.; Correia, A. Otolith elemental signatures indicate population separation in deep-sea rockfish, Helicolenus dactylopterus and Pontinus kuhlii, from the Azores. *J. Sea Res.* 2013, 83, 202–208. [CrossRef]

43. Castro, B.M.; Lorenzzetti, J.A.; Miranda, L.B. Estrutura termohalina e circulação na região entre o Cabo de São Tomé (RI) e o Chuí (RS). In *O Ambiente Oceanográfico da Plataforma Continental e do Talude na Região Sudeste-Sul do Brasil*; Rossi-Wongtschowski, C.L.B., Madureira, L.S.P., Eds.; Edusp: São Paulo, Brazil, 2006; pp. 11–120.

44. Campos, E.J.D.; Lorenzzetti, J.A.; Stevenson, M.R.; Stech, J.L.; Souza, R.B. Penetration of waters from the Brazil-Malvinas Confluence region along the South American Continental Shelf up to 23° S. *An. Acad. Bras. Cienc.* 1996, 68, 49–58.
45. Campos, E.J.D.; Lentini, C.A.D.; Miller, J.L.; Piola, A.R. Interannual variability of the sea surface temperature in the South Brazil Bight. *Geophys. Res. Lett.* 1999, 26, 2061–2064. [CrossRef]
46. Souza, R.B.; Robinson, I.S. Lagrangian and satellite observations of the Brazilian Coastal Current. *Cont. Shelf Res.* 2004, 24, 241–262. [CrossRef]
47. Simonassi, J.C.; Henemann, M.C.; Talgatti, D.; Marques, A.N., Jr. Nutrient variations and coastal water quality of Santa Ca-tarina Island, Brazil. *Biotemas* 2010, 23, 211–223.
48. Spach, H.; Yamaguti, N. Variação geográfica de Cynoscion jamaicensis (Pisces: Sciaenidae) entre as latitudes 20°18´S (Vitória, ES)—32°-10´S (Barra do Rio Grande, RS). II—Caracteres morfométricos. *Rev. Nuterica* 1989, 4, 77–104.
49. Sturrock, A.M.; Hunter, E.; Milton, J.A.; EIMF; Johnson, R.C.; Waring, C.P.; Trueman, C.N. Quantifying physiological influences on otolith microchemistry. *Methods Ecol. Evol.* 2015, 6, 806–816. [CrossRef]
50. Souza, R.B.; Robinson, I.S. Lagrangian and satellite observations of the Brazilian Coastal Current.
51. Figueira, R.C.L.; Tessler, M.G.; Mahiques, M.M.; Cunha, I.I.L. Distribution of 137Cs, 238Pu and 239+240Pu in sediments of the Rio Grande estuarine system, southern Brazil. *Braz. J. Oceanogr.* 2005, 53, 169–181. [CrossRef]
52. Castro, B.M. Summer/winter stratification variability in the central part of the South Brazil Bight. *Biol. Annu. Rev.* 2008, 297–330. [CrossRef]
53. De Mahiques, M.M.; Burone, L.; Figueira, R.; Lavenère-Wanderley, A.A.D.O.; Capellari, B.; Rogacheski, C.E.; Barroso, C.P.; Dos Santos, L.A.S.; Cordero, L.M.; Cussioli, M.C. Anthropogenic influences in a lagoon environment: A multiproxys approach at the valo grande mouth, Cananeia-Iguape system (SE Brazil). *Braz. J. Oceanogr.* 2009, 57, 325–337. [CrossRef]
54. Garcia, M.R.; Cattani, A.P.; Lana, P.D.C.; Figueira, R.C.L.; Martins, C.C. Petroleum biomarkers as tracers of low-level chronic oil contamination of coastal environments: A systematic approach in a subtropical mangrove. *Environ. Pollut.* 2019, 249, 1060–1070. [CrossRef]
55. Vazzoler, A. Diversidade fisiológica e morfológica de Micropogon furnieri (Desmarest, 1822) ao sul de Cabo Frio, Brasil. *Bol. Inst. Oceanogr.* 1971, 20, 1–70. [CrossRef]
56. De Mahiques, M.M.; Burone, L.; Figueira, R.; Lavenère-Wanderley, A.A.D.O.; Capellari, B.; Rogacheski, C.E.; Barroso, C.P.; Dos Santos, L.A.S.; Cordero, L.M.; Cussioli, M.C. Anthropogenic influences in a lagoon environment: A multiproxys approach at the valo grande mouth, Cananeia-Iguape system (SE Brazil). *Braz. J. Oceanogr.* 2009, 57, 325–337. [CrossRef]
57. Elsdon, T.; Wells, B.; Campmana, S.; Gillanders, B.; Jones, C.; Limburg, K.; Secor, D.; Thorrold, S.; Walther, B. Otolith chemis-try to describe movements and life-history parameters of fishes: Hypotheses, assumptions, limitations and inferences. *Oceanogr. Mar. Biol. Annu. Rev.* 2008, 46, 297–330.
58. Elsdon, T.; Wells, B.; Campmana, S.; Gillanders, B.; Jones, C.; Limburg, K.; Secor, D.; Thorrold, S.; Walther, B. Otolith chemis-try to describe movements and life-history parameters of fishes: Hypotheses, assumptions, limitations and inferences. *Oceanogr. Mar. Biol. Annu. Rev.* 2008, 46, 297–330. [CrossRef]
59. Mazzini, P.L.F.; Barth, J.A. A comparison of mechanisms generating vertical transport in the Brazilian coastal upwelling regions. *J. Geophys. Res. Oceans* 2013, 118, 5977–5993. [CrossRef]
60. Filho, A.M.P.; Cergole, M.C. Diferenciação geográfica de Nebris microps (Cuvier, 1830), na costa sudeste do Brasil. *Bolm. Inst. Oceanogr.* 1988, 36, 37–45. [CrossRef]
61. Figueira, R.C.L.; Tessler, M.G.; Mahiques, M.M.; Cunha, I.I.L. Distribution of 137Cs, 238Pu and 239+240Pu in sediments of the Rio Grande estuarine system, southern Brazil. *Braz. J. Oceanogr.* 2005, 53, 169–181. [CrossRef]
62. De Mahiques, M.M.; Burone, L.; Figueira, R.; Lavenère-Wanderley, A.A.D.O.; Capellari, B.; Rogacheski, C.E.; Barroso, C.P.; Dos Santos, L.A.S.; Cordero, L.M.; Cussioli, M.C. Anthropogenic influences in a lagoon environment: A multiproxys approach at the valo grande mouth, Cananeia-Iguape system (SE Brazil). *Braz. J. Oceanogr.* 2009, 57, 325–337. [CrossRef]
63. Elsdon, T.; Wells, B.; Campmana, S.; Gillanders, B.; Jones, C.; Limburg, K.; Secor, D.; Thorrold, S.; Walther, B. Otolith chemis-try to describe movements and life-history parameters of fishes: Hypotheses, assumptions, limitations and inferences. *Oceanogr. Mar. Biol. Annu. Rev.* 2008, 46, 297–330.
64. Souza, R.B.; Robinson, I.S. Lagrangian and satellite observations of the Brazilian Coastal Current.
65. De Mahiques, M.M.; Hanebuth, T.; Martins, C.D.C.; Montoya-Montes, I.; Alcântara-Carrió, J.; Figueira, R.; Bicego, M. Mud depocentres on the continental shelf: A neglected sink for anthropogenic contaminants from the coastal zone. *Environ. Earth Sci.* 2016, 75, 44. [CrossRef]
66. Barnes, T.C.; Gillanders, B.M. Combined effects of extrinsic and intrinsic factors on otolith chemistry: Implications for environ-mental reconstructions. *Can. J. Fish. Aquat. Sci.* 2013, 70, 1159–1166. [CrossRef]
67. Hüssy, K.; Limburg, K.E.; de Pontual, H.; Thomas, O.R.B.; Cook, P.K.; Heimbrand, Y.; Blass, M.; Sturrock, A.M. Trace Element Patterns in Otoliths: The Role of Biomineralization. *Rev. Fish. Sci. Aquac.* 2020, 29, 445–477. [CrossRef]
68. Geffen, A.J.; Pearce, N.J.G.; Perkins, W.T. Metal concentrations in fish otoliths in relation to body composition after labora-tory exposure to mercury and lead. *Mar. Ecol. Progr. Ser.* 1998, 165, 235–245. [CrossRef]
69. Barnes, T.C.; Gillanders, B.M. Combined effects of extrinsic and intrinsic factors on otolith chemistry: Implications for environ-mental reconstructions. *Can. J. Fish. Aquat. Sci.* 2013, 70, 1159–1166. [CrossRef]
70. Souza, R.B.; Robinson, I.S. Lagrangian and satellite observations of the Brazilian Coastal Current.
71. Barnes, T.C.; Gillanders, B.M. Combined effects of extrinsic and intrinsic factors on otolith chemistry: Implications for environ-mental reconstructions. *Can. J. Fish. Aquat. Sci.* 2013, 70, 1159–1166. [CrossRef]
72. Grammer, G.L.; Morrongiello, J.R.; Izzo, C.; Hawthorne, P.J.; Middleton, J.F.; Gillanders, B.M. Coupling biogeochemical tracers with fish growth reveals physiological and environmental controls on otolith chemistry. *Ecol. Monogr.* 2017, 87, 487–507. [CrossRef]

73. Pan, X.; Ye, Z.; Xu, B.; Jiang, T.; Yang, J.; Tian, Y. Population connectivity in a highly migratory fish, Japanese Spanish mackerel (*Scomberomorus niphonius*), along the Chinese coast, implications from otolith chemistry. *Fish. Res.* 2020, 231, 105690. [CrossRef]

74. Thomas, O.R.B.; Ganio, K.; Roberts, B.R.; Swearer, S.E. Trace element-protein interactions in endolymph from the inner ear of fish: Implications for environmental reconstructions using fish otolith chemistry. *Metallokins 2017*, 9, 239–249. [CrossRef]

75. Sturrock, A.; Trueman, C.; Darnaude, A.; Hunter, E. Can otolith elemental chemistry retrospectively track migrations in fully marine fishes? *J. Fish Biol.* 2012, 81, 766–795. [CrossRef]

76. Thomas, O.R.B.; Swearer, S.; Kapp, E.A.; Peng, P.; Tonkin-Hill, G.Q.; Papenfuss, A.; Roberts, A.; Bernard, P.; Roberts, B.R. The inner ear proteome of teleost fish. *FEBS J.* 2019, 286, 66–81. [CrossRef]

77. Evans, D.H. Teleost fish osmoregulation: What have we learned since August Krogh, Homer Smith, and Ancel Keys. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 2008, 295, R704–R713. [CrossRef] [PubMed]

78. Evans, D.H. Teleost fish osmoregulation: What have we learned since August Krogh, Homer Smith, and Ancel Keys. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 2008, 295, R704–R713. [CrossRef] [PubMed]

79. Friedrich, L.A.; Halden, N.M. Determining Exposure History of Northern Pike and Walleye to Tailings Effluence Using Trace Metal Uptake in Otoliths. *Environ. Sci. Technol.* 2010, 44, 1551–1558. [CrossRef] [PubMed]

80. Mayer, E.M. Lead. *Fish Physiol.* 2011, 31, 185–236.

81. Wood, C.M. An introduction to metals in fish physiology and toxicology: Basic principles. *Fish Physiol.* 2011, 31, 1–51. [CrossRef]

82. Angulo, R.; Soares, C.; Marone, E.; Souza, M.; Odreski, L.; Noernberg, M. Paraná. In *Erosão e Progradação do Litoral Brasileiro*; Muehe, D., Ed.; Ministério do Meio Ambiente (MMA): Governo Federal, Brasília, 2006; pp. 347–400. [CrossRef]

83. Klein, A.; Menezes, J.; diehl, F.; Abreu, J.; Polete, M.; Sperb, R.; Sperb, R.; Horn, N. Santa Catarina. In *Erosão e Progradação do Litoral Brasileiro*; Muehe, D., Ed.; Ministério do Meio Ambiente (MMA): Brasília, Brazil, 2006; pp. 401–436. [CrossRef]

84. Tessler, M.; Goya, S.; Yoshikawa, P.; Hurtado, S. São Paulo. In *Erosão e Progradação do Litoral Brasileiro*; Muehe, D., Ed.; Ministério do Meio Ambiente (MMA): Brasília, Brazil, 2006; pp. 297–346. [CrossRef]

85. Tessler, M.; Goya, S.; Yoshikawa, P.; Hurtado, S. São Paulo. In *Erosão e Progradação do Litoral Brasileiro*; Muehe, D., Ed.; Ministério do Meio Ambiente (MMA): Brasília, Brazil, 2006; pp. 297–346. [CrossRef]

86. Moura, A.M.M. Trajetória da política ambiental federal no Brasil. In *Governança Ambiental no Brasil: Instituições, Atores e Políticas Públicas*; Moura, A.M.M., Ed.; Ipea: Brasília, Brazil, 2016; pp. 13–44. [CrossRef]

87. Mahiques, M.M.; Figueira, R.C.L.; Salaroli, A.B.; Alves, D.P.V.; Gonçalves, C. 150 years of anthropogenic metal input in a Biosphere Reserve: The case study of the Cananeia Iguape coastal system, Southeastern Brazil. *Environ. Earth Sci.* 2013, 68, 1073–1087. [CrossRef]

88. Conrad, S.R.; Sanders, C.J. Influence of anthropogenic activities on trace metal accumulation in Brazilian mangrove sedi-ments. *Rev. Virtual Quim.* 2017, 9, 2017–2031. [CrossRef]

89. Jesus, M.S.D.S.D.; Frontalini, F.; Bouchet, V.M.; Yamashita, C.; Sartoretto, J.R.; Figueire, R.C.; Sousa, S.H.D.M.E. Reconstruction of the palaeo-ecological quality status in an impacted estuary using benthic foraminifera: The Santos Estuary (São Paulo state, SE Brazil). *Mar. Environ. Res.* 2020, 162, 105121. [CrossRef]

90. Spencer, K.; Shafer, D.J.; Gauldie, R.W.; DeCarlo, E.H. Stable lead isotope ratios from distinct anthropogenic sources in fish otoliths: A potential nursery stock marker. *Comp. Biochem. Physiol. Part A Mol. Integr. Physiol.* 2000, 127, 273–284. [CrossRef] [PubMed]

91. Geffen, A.I.; Jarvis, K.; Thorpe, J.P.; Leah, R.T.; Nash, R.D.M. Spatial differences in the trace element concentrations of Irish Sea plaice Pleuronectes platessa and whiting Merlangius merlangus otoliths. *J. Sea Res.* 2003, 50, 245–254. [CrossRef] [PubMed]

92. Morales-Nin, B.; Geffen, A.; Cardona, F.; Kruber, C.; Saborido-Rey, F. The effect of Prestige oil ingestion on the growth and chemical composition of turbot otoliths. *Mar. Pollut. Bull.* 2007, 54, 1732–1741. [CrossRef] [PubMed]

93. López-Duarte, P.C.; Fodrie, F.J.; Jensen, O.P.; Whitehead, A.; Galvez, F.; Dubansky, B.; Able, K.W. Is exposure to Ma-condo Oil reflected in the otolith chemistry of marsh-resident fish? *PLoS ONE* 2016, 11, e0162699. [CrossRef] [PubMed]

94. Vrdoljak, D.; Matic-Skoko, S.; Peharda, M.; Uvanovic, H.; Markulin, K.; Mertz-Kraus, R. Otolith fingerprints reveals poten-tial pollution exposure of newly settled juvenile Sparus aurata. *Mar. Pollut. Bull.* 2020, 160, 111695. [CrossRef]

95. Barnett, T.P.; Pierce, D.W.; Schnur, R. Detection of Anthropogenic Climate Change in the World’s Oceans. *Science* 2001, 292, 270–274. [CrossRef]

96. Pörtner, H.-O. Ecosystem effects of ocean acidification in times of ocean warming: A physiologist’s view. *Mar. Ecol. Prog. Ser.* 2008, 373, 203–217. [CrossRef]

97. IPCC. Summary for Policymakers. In *Climate Change and Land: An IPCC Special Report on Climate Change, Desertification, Land Degradation, Sustainable Land Man-agement, Food Security, and Greenhouse Gas Fluxes in Terrestrial Ecosystems*; Shukla, P.R., Skea, J., Calvo Buendia, E., Masson-Delmotte, V., Pörtner, H.-O., Roberts, D.C., Zhai, P., Slade, R., Connors, S., van Diemen, R., et al., Eds.; IPCC: Geneva, Switzerland, 2020.

98. Lumpkin, R.; Garzoli, S. Interannual to decadal changes in the western South Atlantic’s surface circulation. *J. Geophys. Res. Earth Surf.* 2011, 116, C01014. [CrossRef]
99. Koenigstein, S.; Mark, F.; Gößling-Reisemann, S.; Reuter, H.; Poertner, H.-O. Modelling climate change impacts on marine fish populations: Process-based integration of ocean warming, acidification and other environmental drivers. *Fish Fish.* 2016, 17, 972–1004. [CrossRef]

100. Kuczynski, L.; Chevalier, M.; Laffaille, P.; Legrand, M.; Grenouillet, G. Indirect effect of temperature on fish population abundances through phenological changes. *PLoS ONE* 2017, 12, e0175735. [CrossRef]

101. Heuer, R.M.; Grosell, M. Physiological impacts of elevated carbon dioxide and ocean acidification on fish. *Am. J. Physiol. Integr. Comp. Physiol.* 2014, 307, R1061–R1084. [CrossRef]

102. Moreira, C.; Froufe, E.; Vaz-Pires, P.; Correia, A. Otolith shape analysis as a tool to infer the population structure of the blue jack mackerel, *Trachurus picturatus*, in the NE Atlantic. *Fish. Res.* 2019, 209, 40–48. [CrossRef]