Climate Stressors on Growth, Yield, and Functional Biochemistry of two Brassica Species, Kale and Mustard

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Abstract: Due to climate change, the attainment of global food security is facing serious challenges in meeting the growing food demand. Abiotic stresses are the foremost limiting factors for agricultural productivity. However, not much information is available on the effect of multiple abiotic stresses on the morphological and biochemical aspects of kale and mustard. Therefore, an experiment was designed to study the effects of UV-B radiation, CO₂ concentration, and high temperature on the growth, yield, and biochemistry of two Brassica species, namely B. oleracea L. var. acephala Winterbor F1 (hybrid kale) and B. juncea var. Green wave O.G. (mustard greens), which were grown under optimal nutrients and soil moisture conditions in soil–plant–atmosphere–research (SPAR) units. Two levels of UV-B radiation (0 and 10 kJ m⁻² d⁻¹), two concentrations of CO₂ (420 and 720 ppm), and two different temperature treatments (25/17 °C and 35/27 °C) were imposed 12 days after sowing (DAS). Several morphological and biochemical parameters were measured at harvest (40 DAS) in both species. All the traits declined considerably under individual and multi-stress conditions in both species except under elevated CO₂ levels, which had a positive impact. Marketable fresh weight decreased by 64% and 58% in kale and mustard plants, respectively, growing under UV-B treatment. A slight increase in the chlorophyll content was observed in both species under the UV-B treatment alone and in combination with high temperature and elevated CO₂. Understanding the impacts of high temperature, CO₂, and UV-B radiation treatments on leafy vegetables, such as kale and mustard, can help to improve existing varieties to enhance resilience towards environmental stresses while simultaneously improving yield, morphology, and biochemistry in plants.

Keywords: temperature stress; elevated CO₂; Ultraviolet (UV)-B; Brassica oleracea; Brassica juncea; chlorophyll; carotenoids

1. Introduction

In the coming decades, a significant rise in agricultural productivity will be required to meet the food requirements of ~800 million undernourished people, which has been growing at an alarming pace, along with shrinking arable land [1–3]. In addition, adverse climate change has further exacerbated this, with increased abiotic stress conditions that detrimentally affect crop productivity and global ecosystem diversity [1,4–6]. Current temperatures are approximately 1 °C above pre-industrial levels, and a rise of 0.5 °C in global temperatures would increase the associated risks of high-temperature stress [7]. Furthermore, according to the fourth US climate assessment, a rise of 1.5 °C to 4.5 °C in global temperature has been projected in the next century due to an alarming increase in the levels of atmospheric CO₂ and other greenhouse gases. The global atmospheric CO₂ concentration is currently 417 ppm, as reported in March 2021 by the Mauna Loa
observatory, Hawaii, while it was only 270 ppm during the pre-industrial era and is projected by climate models to reach 540 to 970 ppm by the year 2100 due to anthropogenic activities, reducing carbon sinks, and natural global cycles [8,9]. A substantial number of abiotic stress-related studies have been conducted during the past decade, but most of the experiments have focused on plants’ responses to individual stress treatment. The studies have overlooked the complex stress response generated in plants against combined or sequential abiotic stresses and the interaction of stresses with each other [10,11]. Therefore, it is imperative to understand the mechanisms associated with crops’ response to various abiotic stresses to manage future food production.

Kale (Brassica oleracea L. var. acephala.) and mustard (Brassica juncea L.) are leafy green brassica vegetables that have increased in production over the past ten years in response to demand by North American consumers. Both vegetables are considered highly nutritious leafy green vegetables even though their nutrient profile has not been well characterized to date [12]. Kale and mustard are rich sources of dietary fiber and low molecular weight nondigestible carbohydrates (LMWC), as well as vitamins A, K, and C; they also contain the essential minerals potassium (K), calcium (Ca), and magnesium (Mg). Despite the nutritional benefits, along with significant quantities of carotenoids and folates, none of these nutritional quality traits have been well characterized to date [13]. A recent study revealed that kale grown in the Southern US has the potential to provide significant quantities of several essential minerals and adequate quantities of LMWC, also known as “prebiotic carbohydrates”, with moderate to low levels of protein and energy [12].

Due to the suitability of kale for southern fall and winter growing conditions, kale has become a significant Brassica vegetable crop in the Southern US. Indeed, the Southern US has emerged as a leading kale production region over the last five years, responsible for more than two-thirds of the US crop [14]. Kale production in southern areas of the US has been increasing to meet American consumer demand, which has been growing at near-doubling rates in recent years [15]. Although kale production has been increasing, Brassica vegetables are generally widely under-consumed by Americans; this has been acknowledged in the current Dietary Guidelines Advisory Committee, which calls for Americans to increase their consumption of fruits and vegetables [15]. Both kale and mustard are hardy, cool season crop that tolerates summer heat but grows best in the fall and winter in the southern regions of the US. The suitable growth temperature is 15–22 °C [16,17]. However, kale and mustard are sensitive to high temperatures [18]. Thus, elevated temperatures and high UV-B levels can damage growth and developmental processes. Since the abiotic stresses are interlinked, the combination of their effects on the morphological, physiological, and cellular processes results in various changes in plant growth, productivity, and yield. Understanding plants’ mechanisms in response to multiple abiotic stresses are essential in devising management and breeding decisions in the near future.

Deryng et al. [19] contributed significantly to the current understanding of climate change impacts on crops under high temperatures and elevated CO₂ concentration. Heat stress leads to poor germination and plant establishment, reduced photosynthesis, leaf senescence, decreased pollen viability, and fewer grains with smaller grain sizes [20,21]. It has been revealed in previous studies that a significant direct effect of increased CO₂ on plant growth and yield could be seen that can compensate for a potentially hotter climate. Although increased CO₂ concentrations have been reported to significantly increase yield in C₃ plants [22–24], not many direct effects have been recorded on kale and mustard plants. The projected higher doses of incoming UV-B radiation can stimulate a variety of responses in higher plants [25,26]. Some harmful effects of UV-B radiation on plants include DNA damage, dilation, and disintegration of cellular membranes, photooxidation of leaf pigments and phytohormones, and inhibition of photosynthesis [27–29]. Additionally, UV-B radiation leads to changes in leaf thickness, anatomy, and canopy morphology, eventually affecting photosynthesis in plants [30]. Therefore, to enhance the production of green leafy vegetables with proper management and breeding strategies in the coming years, it is crucial to understand the effects of UV-B radiation individually and its interaction with
other stresses. While some data exist on the crop’s growth, development, and productivity in response to individual CO$_2$, temperature, or UV-B stresses, very little data is available on the effects of the interaction of these multiple factors on the growth and development of kale and mustard.

Field studies for understanding the interactive effect of abiotic stresses on Brassica spp. are tedious, discrepant, and seasonally limited. Therefore, simple, rapid, and reliable techniques are required to understand the response of these crops to various environmental stresses. The present study’s results can help to quantify Brassica species’ response to these abiotic stresses and improve existing varieties for enhanced resilience while improving the plants’ yield, morphology, and biochemistry. Compared to other controlled environment facilities, the soil–plant–atmosphere–research (SPAR) systems have the advantage of precisely controlling air temperature, CO$_2$ concentration, UV-B dosage, and air humidity under natural solar radiation compared to other controlled environment facilities [31].

Since only limited information is available, the present study was designed to evaluate the interactive effects of elevated CO$_2$, high temperature, and UV-B stress on kale and mustard plants’ morphology, physiology, and phytochemistry.

2. Materials and Methods

2.1. Experimental Conditions and Plant Material

This study included two Brassica species, namely B. oleracea var. acephala Winterbor F1 (hybrid kale) and B. juncea var. Green Wave OG (mustard greens). The experiment was conducted in a controlled environment facility (SPAR units) at the Environmental Plant Physiology Laboratory, Mississippi State University (33°28′ N, 88°47′ W), Mississippi State, MS, July–August 2019. The specifications and operation of SPAR units have been discussed in detail in Reddy et al. [32].

Seeds of the two Brassica sp. were sown in 30.5 by 15.2 cm (height by diameter) polyvinyl chloride pots filled with a soil medium consisting of 3:1 sand/topsoil (v/v). Before the start of treatments, the seedlings were thinned down to one plant per pot. The plants were watered and fertilized with a full-strength Hoagland nutrient solution [33] based on daily evapotranspiration (Table 1). Pots were arranged in 10 rows with three pots per row in each SPAR chamber with alternating rows of kale and mustard plants. The experiment consisted of 2 factors (8 levels of treatments × 2 species) with 15 replications. The pots were randomly arranged within each SPAR unit to avoid positional effects. In this study, 240 plants (2 species × 8 treatments × 15 replications) were used to estimate the impact of multiple stresses on the two Brassica species.

Table 1. The set treatments and results measured for day, night, and average temperatures, chamber [CO$_2$], daytime and nighttime vapor pressure deficit (VPD), and evapotranspiration (ET) during the experimental period of each treatment on kale and mustard.

| Treatments          | Measured Temperature (°C) | CO$_2$ (µmol mol$^{-1}$) | VPD (kPa) | Mean ET (L H$_2$O d$^{-1}$) |
|---------------------|---------------------------|---------------------------|-----------|----------------------------|
|                     | DAY                       | NIGHT                     | DAY       | NIGHT                     | DAY/NIGHT                     |
| Control             | 24.90 ± 0.08              | 17.42 ± 0.03              | 21.56 ± 0.04 | 433 ± 1.92              | 1.35 ± 0.02              | 0.94 ± 0.01              | 11.55 ± 0.84             |
| +CO$_2$             | 25.13 ± 0.06              | 17.54 ± 0.02              | 21.73 ± 0.04 | 721.22 ± 1.27            | 1.33 ± 0.02              | 0.97 ± 0.01              | 11.71 ± 1.15             |
| +T                  | 31.31 ± 0.66              | 23.73 ± 0.65              | 27.93 ± 0.64 | 434.31 ± 1.30            | 2.30 ± 0.11              | 1.68 ± 0.08              | 12.56 ± 1.13             |
| +UV-B               | 24.84 ± 0.10              | 17.31 ± 0.03              | 21.48 ± 0.05 | 439.69 ± 1.72            | 1.37 ± 0.01              | 0.97 ± 0.01              | 8.48 ± 0.57              |
| +T + CO$_2$         | 31.69 ± 0.70              | 24.04 ± 0.69              | 28.28 ± 0.68 | 720.04 ± 2.49            | 2.75 ± 0.11              | 1.91 ± 0.08              | 13.79 ± 1.40             |
| +UV-B + CO$_2$      | 24.90 ± 0.09              | 17.35 ± 0.03              | 21.53 ± 0.05 | 715.98 ± 2.17            | 1.32 ± 0.02              | 0.90 ± 0.01              | 8.71 ± 0.01              |
| +UV-B + T           | 31.32 ± 0.67              | 23.75 ± 0.66              | 27.94 ± 0.65 | 435.33 ± 1.27            | 2.74 ± 0.11              | 1.90 ± 0.09              | 11.68 ± 0.96             |
| +UV-B + CO$_2$ + T  | 31.34 ± 0.67              | 23.75 ± 0.66              | 27.96 ± 0.65 | 729.37 ± 1.46            | 2.89 ± 0.14              | 1.99 ± 0.10              | 11.34 ± 1.21             |

During the experiment, the incoming daily solar radiation measured with a pyranometer (Model 4–8; The Eppley Laboratory Inc., Newport, RI, USA) outside the SPAR units ranged from 11.3 to 31.3 MJ m$^{-2}$ d$^{-1}$ with an average value of 25.10 ± 0.82 MJ m$^{-2}$ d$^{-1}$. 
2.2. Treatments

The treatments included combinations of two \([\text{CO}_2]\) concentrations, namely 400 and 720 \(\mu\text{mol mol}^{-1}\) (+CO\(_2\)), two different temperatures, namely 25/17 °C and 35/27 °C (+T) (day/night), and two daily biologically effective UV-B radiation intensities, namely 0 and 10 kJ m\(^{-2}\) d\(^{-1}\) (+UV-B).

The control treatment was 400 \(\mu\text{mol mol}^{-1}\) [CO\(_2\)], at 25/17 °C (day/night) temperature, and 0 kJ m\(^{-2}\) d\(^{-1}\) UV-B treatment. All SPAR chambers were maintained at control conditions until 12 days after sowing (DAS). Subsequently, each chamber was set at one of the following eight treatments until the final harvest (40 DAS): (1) a control treatment with optimum temperature, ambient CO\(_2\) levels, and no UV-B; (2) optimum temperature with elevated CO\(_2\) levels and no UV-B (+CO\(_2\)); (3) elevated temperature with ambient CO\(_2\) levels and no UV-B (+T); (4) optimum temperature and ambient CO\(_2\) levels with 10 kJ UV-B (+UV-B); (5) elevated temperature and CO\(_2\) levels with no UV-B (+T + CO\(_2\)); (6) optimum temperature with elevated CO\(_2\) levels and 10 kJ UV-B (+CO\(_2\) + UV-B); (7) elevated temperature with 10 kJ UV-B at ambient CO\(_2\) levels (+T + UV-B); (8) elevated temperature and elevated CO\(_2\) levels with 10 kJ UV-B (+UV-B + CO\(_2\) + T). The set and measured environmental variables in this study using eight different SPAR units are provided in Table 1.

2.3. Measurements

2.3.1. Morphological Measurements

At 40 DAS, kale and mustard plants from each SPAR unit were hand-harvested to obtain their phenotype and growth data on the effects of multiple abiotic stresses. Plant height (PH, cm) was measured, leaf number (LN) was counted, and then total leaf area (LA, cm\(^2\) plant\(^{-1}\)) was determined using an LI-3100 leaf area meter (LI-COR, Lincoln, NE, USA). Plant components, such as aboveground dry weights and root weights (g plant\(^{-1}\)), were determined by drying the samples at 80 °C until a constant weight was reached.

2.3.2. Physiological Measurements

Leaf chlorophyll content, epidermal flavonoids index, epidermal anthocyanin, and nitrogen balance index (the ratio of chlorophyll content/flavonoids) were measured on the uppermost, fully expanded leaf, second from the top, across all treatments using a handheld Dualex Scientific instrument (Force A DX16641, Paris, France) at 35 DAS.

2.3.3. Epicuticular Wax Content Determination

The extraction and quantitative analysis of leaf epicuticular waxes were carried out as per the method of Ébercon et al. [34] with minor modifications. Ten leaf discs constituting an area of 35.36 cm\(^2\) from the third or fourth leaf from the stem apex were cut from both species from five plants in each treatment. Leaf discs were stirred in 15 mL of chloroform (Sigma-Aldrich, Inc., St. Louis, MO, USA) in a test tube for 20 s to remove leaf waxes. The wax extract was evaporated on a water bath maintained at 80 °C, and then cooled to room temperature; 5 mL of dichromate reagent was added and heated on a water bath held at 80 °C for 30 min. The samples were removed from the water bath and cooled, followed by the addition of 12 mL of de-ionized water. The samples were then allowed to stand for 15 min. The intensity of the color was measured at 590 nm using a Bio-Rad UV/VIS spectrophotometer (Bio-Rad Laboratories, Hercules, CA, USA). The wax content was expressed on a leaf area basis (µg cm\(^{-2}\)) using a standard curve developed from the wax obtained from the same species.

2.3.4. Carotenoid Analysis

Carotenoid pigments were extracted and analyzed from freeze-dried leaf tissues, according to Kopsell et al. [35,36], with a few changes, as described by Barickman et al. [37].
2.4. Data Analysis

2.4.1. Combined Stress Response Index (CSRI)

Based on the summation of relative individual stress responses at each treatment and similar to the cumulative response index quoted in other UV-B studies [38], the combined stress response index (CSRI) was calculated to evaluate the interactive effects of eight treatments (+ CO$_2$, + T, + UV-B, + CO$_2$ + T, + CO$_2$ + UV-B, + UV-B + T, and + CO$_2$ + T + UV-B) in comparison to control treatment. The CSRI was calculated as the value of a parameter under control (c) subtracted from the value of the parameter under treatment (t), and then by dividing from the value of a parameter under control (c) as follows:

$$\text{CSRI} = \frac{(\text{PH}_t - \text{PH}_c)}{(\text{PH}_c)} + \frac{(\text{LN}_t - \text{LN}_c)}{(\text{LN}_c)} + \frac{(\text{LA}_t - \text{LA}_c)}{(\text{LA}_c)} + \frac{(\text{MFW}_t - \text{MFW}_c)}{(\text{MFW}_c)} + \frac{(\text{ADW}_t - \text{ADW}_c)}{(\text{ADW}_c)} + \frac{(\text{RDW}_t - \text{RDW}_c)}{(\text{RDW}_c)} + \frac{(\text{TDW}_t - \text{TDW}_c)}{(\text{TDW}_c)} + \frac{(\text{RS}_t - \text{RS}_c)}{(\text{RS}_c)} + \frac{(\text{Neo}_t - \text{Neo}_c)}{(\text{Neo}_c)} + \frac{(\text{Viol}_t - \text{Viol}_c)}{(\text{Viol}_c)} + \frac{(\text{Zea}_t - \text{Zea}_c)}{(\text{Zea}_c)} + \frac{(\text{Lut}_t - \text{Lut}_c)}{(\text{Lut}_c)} + \frac{(\text{Bcar}_t - \text{Bcar}_c)}{(\text{Bcar}_c)} + \frac{(\text{TXan}_t - \text{TXan}_c)}{(\text{TXan}_c)} + \frac{(\text{ZAV}_t - \text{ZAV}_c)}{(\text{ZAV}_c)} + \frac{(\text{Chl}_t - \text{Chl}_c)}{(\text{Chl}_c)} + \frac{(\text{Flav}_t - \text{Flav}_c)}{(\text{Flav}_c)} + \frac{(\text{Anth}_t - \text{Anth}_c)}{(\text{Anth}_c)} + \frac{(\text{NBI}_t - \text{NBI}_c)}{(\text{NBI}_c)} + \frac{(\text{Wax}_t - \text{Wax}_c)}{(\text{Wax}_c)}$$

Here, CSRI is the combined stress response index, PH is the plant height, LN is the leaf number, LA is the leaf area of the plant, MFW is the marketable fresh weight, ADW is the aboveground dry weight, RDW is the root dry weight, TDW is the total dry weight, RS is the root-to-shoot ratio, Neo is the neoxanthin concentration, Viol is the violaxanthin concentration, Zea is the zeaxanthin concentration, Lut is the lutein concentration, Bcar is the β-carotene concentration, TXan is the total xanthophyll concentration, ZA/ZAV is the xanthophyll cycle ratio, Chl is the chlorophyll concentration, Flav is the flavonoid index, Anth is the anthocyanin index, NBI is the nitrogen balance index, Wax is the wax content, under t (treatment) and c (control).

2.4.2. Statistical Analysis

The experimental layout was a split plot with a complete randomized block design, considering multi-stress treatments as the whole plot and species as the subplot. The one-way ANOVA of the general linear model, PROC GLM, was performed to test the effects of treatments, species, and their interactions on the measured traits using SAS 9.2 (SAS Institute, Cary, NC, USA). Fisher’s protected least significant difference tests at $p = 0.05$ were employed to test the differences among treatments for measured parameters. The standard errors of the mean were calculated and presented in the figures as error bars. Graphs were generated using Sigma Plot 13.0 (Systat Software, San Jose, CA, USA).

3. Results

3.1. Aboveground Morphology Parameters

Here, UV-B and mostly elevated temperatures reduced vegetative growth in both crops. However, CO$_2$ masked most of the other stresses’ adverse effects. Treatment and species interaction significantly affected the plant height, number of leaves, leaf area, and marketable fresh weight parameters (Table 2; Figure 1). Plants grown under +UV-B + T treatment were significantly shorter in both Brassica sp. compared to the control plants.

Elevated CO$_2$ (+CO$_2$) slightly increased plant height (7%) in kale and mustard compared to the control treatment. The highest reduction in plant height (47%) was observed in kale plants growing under +UV-B treatment and +T + CO$_2$ (30.7%) for mustard. Mustard had taller plants and fewer reductions among the two species (Table 3).
Table 2. The analysis of variance across the treatments of CO2 concentration, temperature, UV-B radiation, and two crops (kale and mustard), and their interactions on kale and mustard root and shoot growth and developmental traits, plant height (PH), mainstem leaves (LN), whole plant leaf area (LA), marketable fresh weight (MFW), aboveground dry weight (ADW), root dry weight (RDW), total plant dry weight (TDW), root/shoot ratio (RS), neoxanthin concentration (Neo), violaxanthin concentration (Viol), zeaxanthin concentration (Zea), lutein concentration (Lut), β-carotene concentration (Bcar), total xanthophyll concentration (TXan), xanthophyll cycle ratio (ZA/ZAV), chlorophyll concentration (Chl), flavonoid index (Flav), anthocyanin index (Anth), nitrogen balance index (NBI), and wax content (Wax).

| Source of Variance | PH (cm) | LN (plant⁻¹) | LA (cm² plant⁻¹) | MFW (g plant⁻¹) |
|--------------------|---------|--------------|------------------|-----------------|
| Crop               | Control | +CO₂ | +T | +UV-B | +T + CO₂ | +UV-B + CO₂ | +UV-B + T | +UV-B + T |
| Kale               | 53.8    | 57.7 (+7%) | 37.7 (−29.8%) | 28.3 (−47.3%) | 35.7 (−33.6%) | 37.1 (−31%) | 31.1 (−42%) | 35.1 (−34.7%) |
| Mustard            | 52.4    | 56.1 (+7%) | 38.7 (−26%) | 40.3 (−23%) | 36.3 (−30.7%) | 44 (−16%) | 39.6 (−24.5%) | 42.4 (−19%) |
| Kale               | 14.2    | 14.7 (+3%) | 13 (−8.5%) | 14.3 (0%) | 13.8 (−3%) | 15.3 (+7.8%) | 13.4 (−5.4%) | 14 (−1.5%) |
| Mustard            | 32      | 43 (+34%) | 36.2 (+13%) | 20.6 (−35.6%) | 50.1 (−50.6%) | 28.6 (−10.7%) | 30.7 (−4%) | 36.7 (+14.8%) |
| Kale               | 1805.7  | 2558.7 (+39%) | 1387.3 (−24.8%) | 570.1 (−68.4%) | 1843.6 (−2%) | 1136.9 (−37%) | 882.8 (−51%) | 1153.1 (−36%) |
| Mustard            | 4613.7  | 6220.2 (+34.8%) | 3724.6 (−19%) | 1934.7 (−58%) | 4993.5 (−8%) | 2857.1 (−38%) | 2637.4 (−42.8%) | 3830.9 (−17%) |
| Kale               | 241.3   | 359.2 (+48.8%) | 151 (−37.4%) | 85.9 (−64.3%) | 188.8 (−21.7%) | 176.8 (−26.7%) | 105.3 (−56.3%) | 151.7 (37%) |
| Mustard            | 569.2   | 755 (+32.6%) | 353 (−40%) | 256 (−55%) | 437.8 (−23%) | 384.6 (−32.4%) | 266.2 (−53%) | 404.2 (−30%) |
Table 3. Cont.

| Traits | Crop | Treatments |
|--------|------|------------|
|        | Kale | Mustard    |
|        |       |            |
| RS     | 0.27 | 0.35 (+29.6%) | 0.32 (+18.5%) | 0.46 (+70%) | 0.28 (+3.7%) | 0.51 (+88.8%) | 0.31 (+14.8%) | 0.37 (+37%) |
|        | 0.27 | 0.35 (+29.6%) | 0.32 (+18.5%) | 0.36 (+33.3%) | 0.23 (+14.8%) | 0.38 (+40.7%) | 0.33 (+22.2%) | 0.28 (+3.7%) |
| ADW (g plant−1) | Kale | 21.7 (+72.1%) | 17 (+21.7%) | 8.9 (+59%) | 22.4 (+3.2%) | 19.4 (+10.8%) | 10.9 (+49.4%) | 17.3 (+20%) |
|        | Mustard | 38.6 (+50%) | 33.2 (+14%) | 21.7 (+34.6%) | 40.4 (+4.7%) | 33.4 (+13.3%) | 26.3 (+31.8%) | 38 (+1.5%) |
| RDW (g plant−1) | Kale | 6 (+95%) | 5.5 (+8.5%) | 4.3 (+28%) | 6.4 (+6.5%) | 9.6 (+66.5%) | 3.3 (+45.3%) | 6.3 (+5.6%) |
|        | Mustard | 10 (+39%) | 10.4 (+3.9%) | 7.6 (+24%) | 8.9 (+10.3%) | 12.6 (+25.6%) | 8.7 (+12.8%) | 10.4 (+4.4%) |
| TDW (g plant−1) | Kale | 27.7 (+78.8%) | 22 (−19%) | 13.2 (+52.3%) | 28.8 (+4%) | 29 (+4.6%) | 14.2 (+48.5%) | 23.7 (+14.5%) |
|        | Mustard | 48.6 (+47.8%) | 43.6 (+10%) | 29.3 (+39.6%) | 49.4 (+1.6%) | 45.9 (+5.3%) | 35 (+28%) | 48.5 (0%) |

The two *Brassica* species exhibited different responses for the number of leaves produced, among other treatments. A maximum reduction of 35% in the mustard leaves was observed under the +UV-B treatment, whereas the leaf number in kale remained unaffected (Table 3). Elevated CO$_2$ treatment (+CO$_2$) significantly increased the leaf number in both species. The highest number of leaves in mustard was found under +T + CO$_2$ treatment and +UV-B + CO$_2$ treatment in kale plants. Leaf number under the combination of all three stresses (+UV-B + CO$_2$ + T) decreased by 1.5% in kale, whereas an increase of +14.8% was observed in mustard.

Even though +UV-B + CO$_2$ treatment did not alleviate the negative effect of UV-B on leaf area, +CO$_2$ treatment alone or in combination with high-temperature treatment (+T + CO$_2$) increased leaf area by 39% and 2% in kale and 34.8% and 8% in mustard, respectively, in comparison to the control (Table 3, Figure 2A). Maximum reduction in leaf area was observed under UV-B treatment, with 68.4% (kale) and 58% (mustard) compared to the control condition (Figure 3). Maximum values for fresh weight were recorded under +CO$_2$ in kale and mustard compared to their control showed a 49% and 32.6% increase.

3.2. Dry Weight Components

All the dry weight components displayed insignificant differences under the treatments–crop interaction (Table 2). Aboveground dry weight, root dry weight, and the total dry weight (Figure 2B) increased under +CO$_2$ treatment, whereas they decreased to the lowest under +UV-B treatment in both kale and mustard (Table 3). Under +CO$_2$ treatment, 78% and 48% more dry matter was produced in kale and mustard, respectively, compared to their control counterparts. High temperature, either alone (+T) or in combination with UV-B (+UV-B + T), also showed a considerable reduction in dry weight components (Table 3). Root/shoot ratio increased under all the treatments, but a maximum increase was observed under +UV-B treatment in both kale and mustard (Figure 2C, Table 3).
unaffected (Table 3). Elevated CO\textsubscript{2} treatment (+CO\textsubscript{2}) significantly increased the leaf and mustard. Bars indicate standard errors of the mean.

**Figure 2.** Impact of CO\textsubscript{2} concentration (control, 400 \textmu mol mol\textsuperscript{-1} and + CO\textsubscript{2}, 720 \textmu mol mol\textsuperscript{-1}), elevated temperatures (25/17 °C and 35/27 °C (day/night)), and UV-B radiation (control, 0 and + UV-B, 10 kJ m\textsuperscript{-2} d\textsuperscript{-1}), and their interactions on (A) leaf area, (B) total dry weight, and (C) root/shoot ratio for kale and mustard. Bars indicate standard errors of the mean.

**Figure 3.** Impact of CO\textsubscript{2} concentration (control, 400 \textmu mol mol\textsuperscript{-1} and + CO\textsubscript{2}, 720 \textmu mol mol\textsuperscript{-1}), elevated temperatures (25/17 °C and 35/27 °C (day/night)), and UV-B radiation (control, 0 and + UV-B, 10 kJ m\textsuperscript{-2} d\textsuperscript{-1}), and their interactions on marketable fresh weight for kale and mustard. Bars indicate standard errors of the mean.
3.3. Physiological Parameters

The different treatments and species affected the chlorophyll content, flavonoid index, anthocyanin index, and nitrogen balance index (Table 4). The interaction of treatment and crop for all four parameters was significant (Table 2). Chlorophyll content and flavonoid index increased, whereas anthocyanin index decreased under all treatments in both species. The chlorophyll content showed a maximum increase of 43.4% under +UV-B treatment in kale and 93.3% under +UV-B + T treatment in mustard (Figure 4, Table 4). The average flavonoid index ranged from 0.72–1.40, showing the highest increase under the +UV-B + CO₂ + T treatment and a minimum increase under +T alone in kale and mustard.

Table 4. Mean values and percent change for chlorophyll concentration (Chl), flavonoid index (Flav), anthocyanin index (Anth), nitrogen balance index (NBI), and wax content (Wax) under CO₂ concentration (control, 400 µmol mol⁻¹ and + CO₂, 720 µmol mol⁻¹), elevated temperatures (25/17 °C and 35/27 °C (day/night)), and UV-B radiation (control, 0 kJ m⁻² d⁻¹ and + UV-B, 10 kJ m⁻² d⁻¹), and their interactions for kale and mustard at 35 DAS.

| Traits        | Crop       | Control | +CO₂ | +T | +UV-B | +T + CO₂ | +UV-B + CO₂ | +UV-B + T | +UV-B + CO₂ + T |
|---------------|------------|---------|------|----|-------|----------|-------------|----------|------------------|
| Chlorophyll   | Kale       | 30.4    | 37   | 31.5| 43.6  | 33.8      | 42 (+38%)   | 38.8     | 38.8 (+27.6%)    |
| conc. (µg/cm²)| Mustard    | 18.77   | 20.6 | 24  | 27.1  | 24.7      | 29.3        | 36.3     | 29.8 (+58.7%)    |
| Flavonoid     | Kale       | 0.72    | 0.81 | 0.74| 1.21  | 0.81      | 1.26        | 1.25     | 1.32 (+83.3%)    |
| index         | Mustard    | 0.82    | 0.85 | 0.83| 1 (+30%)| 0.84      | 1.12        | 1.19     | 1.40 (+70.7%)    |
| Anthocyanin   | Kale       | 0.09    | 0.09 | 0.08| 0.06  | 0.09      | 0.06        | 0.07     | 0.06 (+33.3%)    |
| index         | Mustard    | 0.14    | 0.13 | 0.12| 0.12  | 0.12      | 0.11        | 0.09     | 0.10 (+28.5%)    |
| NBI           | Kale       | 42.8    | 46.4 | 46.2| 37.7  | 34.4      | 31.5        | 30 (-30%) |                  |
|               | Mustard    | 23.4    | 24.6 | 31.4| 28.5  | 36.5      | 27.4        | 31.8     | 21.9 (-6.4%)     |
| Waxes         | Kale       | 122.5   | 115.6| 133 | 92.5  | 140.8     | 87.9        | 66.3     | 70.1 (-42.7%)    |
| (µg/cm²)      | Mustard    | 17.4    | 13   | 14.7| 13.4  | 18.18     | 11          | 17.5     | 21 (+20.6%)      |

The minimum adverse effect on the anthocyanin index was observed under +T treatment for kale and +CO₂ treatment for mustard, with an average decrease of 11% and 7%, respectively, compared to the control. There was no change observed in the anthocyanin index under +CO₂ treatment alone and in combination with high temperature (+T + CO₂) in kale. Additionally, UV-B treatment alone (+UV-B) and in combination with CO₂ and T (+UV-B + CO₂, +UV-B + T, +UV-B + CO₂ + T) showed significant reductions in the anthocyanin index (Table 4).

Nitrogen balance index (NBI) increased under all treatments in mustard except under +UV-B + CO₂ + T, where a 6% decrease was observed compared to the control. A maximum decrease of 30% was observed in kale under +UV-B + CO₂ + T, whereas an increase in NBI was exhibited under +CO₂, +T, and their combination (Table 4).
3.4. Epicuticular Wax Content

Wax production was significantly increased under +T + CO₂ treatment for kale and +UV-B + CO₂ treatment for mustard; the increase was 15% and 20.6%, respectively, compared to their control treatment (Table 4). The average wax content ranged from 66 to 140.8 µg cm⁻² in kale and 21 to 11 µg cm⁻² in mustard leaves. The lowest epicuticular wax was recorded under +UV-B + CO₂ treatment in kale and under UV-B + CO₂ treatment in mustard (Table 4). Kale plants grown under +UV-B + T + CO₂ treatment and mustard plants grown under +UV-B treatment produced less wax on the leaves, and the reduction was 42.7% and 30%, respectively, compared to the plants growing under control conditions. Kale plants had the highest wax content under all the treatments.

3.5. Total Carotenoid Concentration

None of the pigments displayed significant differences under the treatments–crop interaction (Table 2); however, a substantial reduction in ZA/ZV in kale (38.3%) was observed under +UV-B + CO₂ treatment and in mustard (19%) under +UV-B + T treatment compared to their respective control. Total xanthophyll content increased highest in kale under +UV-B + CO₂ treatment, whereas a decrease of 10% was observed in mustard under the same treatment. An increase of 19% was recorded in total xanthophylls in mustard plants at elevated CO₂ concentrations. Furthermore, β-carotene was atypically much lower (20%) under elevated CO₂ in mustard than in control plants (Table 5). Neoxanthin, violaxanthin, zeaxanthin, lutein, and β-carotene increased significantly in kale plants at high CO₂ concentrations (Table 5). Under +UV-B treatment, both Brassica species increased the concentration of neoxanthin, violaxanthin, and lutein.
Table 5. Mean values and percent change for neoxanthin concentration (Neo), violaxanthin concentration (Viol), zeaxanthin concentration (Zea), lutein concentration (Lut), β-carotene concentration (Bcar), total xanthophyll concentration (TXan), and xanthophyll cycle ratio (ZA/ZAV), measured under CO₂ concentration (control, 400 μmol mol⁻¹ and + CO₂, 720 μmol mol⁻¹), elevated temperatures (25/17 °C and 35/27 °C (day/night)), and UV-B radiation (control, 0 kJ m⁻² d⁻¹ and + UV-B, 10 kJ m⁻² d⁻¹), and their interactions for kale and mustard at 40 DAS.

| Traits            | Crop       | Treatments                  | Control          | +CO₂       | +T         | +UV-B       | +T + CO₂    | +UV-B + CO₂ | +UV-B + T   | +UV-B + CO₂ + T |
|-------------------|------------|-----------------------------|------------------|------------|------------|-------------|-------------|-------------|-------------|----------------|
|                   |            |                             | (µg/g dry mass)   | (µg/g dry mass) | (µg/g dry mass) | (µg/g dry mass) | (µg/g dry mass) | (µg/g dry mass) | (µg/g dry mass) | (µg/g dry mass) |
| Neo               | Kale       | 290                         | 355 (+22.4%)     | 276.8 (−4.5%) | 407.7 (+40.5%) | 298.5 (+3%)  | 372.1 (+28.3%) | 371.5 (+28.1%) | 397 (+36.8%) |
|                   | Mustard    | 273.3                       | 256.3 (−6.2%)    | 246.3 (−9.8%) | 370.4 (+35.5%) | 269 (−1.6%)  | 361.2 (+32%)  | 378.5 (+38.4%) | 319.3 (+16.8%) |
| Viol              | Kale       | 163.7                       | 195 (+19%)       | 167.5 (−2.3%) | 314.4 (+92%)  | 150 (−8.3%)  | 337.8 (+106%) | 276.2 (+68.8%) | 242 (+47.8%)  |
|                   | Mustard    | 323.9                       | 217.3 (−40%)     | 248.8 (+1%)  | 327.7 (+35.7%) | 208 (−15%)   | 274.8 (+20.7%) | 391.2 (+18.4%) | 264 (-14.8%) |
| Anth              | Kale       | 37.6                        | 39.3 (+4.5%)     | 23.8 (−36.7%) | 46.6 (+23.9%) | 34.6 (−8%)   | 41.6 (+10.6%) | 43.6 (+16%)   | 31.2 (-17%)   |
|                   | Mustard    | 64.1                        | 48.2 (−24.8%)    | 44.6 (−50.4%) | 67.7 (+5.6%)  | 38.3 (−40%)  | 51.8 (−19%)   | 45.4 (+29%)   | 37.7 (-41%)   |
| Zea               | Kale       | 209.5                       | 219.2 (+4.6%)    | 132.9 (−36.5%) | 149.4 (+28.6%) | 139.2 (−33.5%) | 157.4 (−24.8%) | 140.8 (−32.7%) | 147.5 (-29.5%) |
|                   | Mustard    | 129.6                       | 126.2 (−2.6%)    | 99.1 (−23.5%) | 148.7 (+14.7%) | 146.8 (+13.2%) | 138.5 (+6.8%) | 123.9 (-4.3%) | 139.2 (+7.4%) |
| Lut               | Kale       | 898.3                       | 967.3 (+7.6%)    | 825 (−8%)    | 1093.6 (+21.7%) | 679.2 (−24.3%) | 1105.5 (+18.7%) | 1066.9 (+5.5%) | 948.1 (-5.5%) |
|                   | Mustard    | 746.7                       | 635.7 (−14.8%)   | 676.4 (−9.4%) | 766.7 (+2.6%) | 654.8 (−12.3%) | 742.3 (+20.4%) | 899.6 (+6.9%) | 695 (-5.6%)   |
| Bcar              | Kale       | 548.4                       | 565.7 (+19.7%)   | 580.5 (+5.8%) | 477.6 (−13%)  | 439.8 (+19.8%) | 710.7 (+29.5%) | 766.6 (+16%)  | 636.8 (+16%)  |
|                   | Mustard    | 649                         | 521.1 (−19.7%)   | 588.2 (−9.3%) | 620.9 (−4.3%) | 476.6 (−26.9%) | 517.7 (+20%)  | 745.2 (+14.8%) | 568.3 (-12.4%) |
| Total Xanth       | Kale       | 410.8                       | 453.5 (+10.3%)   | 324.1 (−21%) | 510.3 (−21.2%) | 323.7 (−21.2%) | 536.7 (+30.6%) | 460.6 (+12%)  | 420.7 (+2.4%)  |
|                   | Mustard    | 517.6                       | 391.8 (−24.3%)   | 392.5 (−24.1%) | 544 (+5.2%)   | 393.3 (−24%)  | 465 (−10%)    | 560.5 (+8.2%)  | 440.9 (-14.8%) |
| ZA/ZAV            | Kale       | 0.60                        | 0.57 (−5%)       | 0.48 (−20%)  | 0.38 (−36.6%) | 0.53 (−11.6%) | 0.37 (−38.3%) | 0.40 (−33.3%) | 0.44 (-26.6%) |
|                   | Mustard    | 0.37                        | 0.44 (−18.9%)    | 0.36 (−2.7%) | 0.41 (−10.8%) | 0.47 +27%  | 0.41 (−18.9%) | 0.30 (−9.8%)  | 0.40 (+8.1%)  |

3.6. Combined Stress Response Index (CSRI)

The CSRI values ranged from −4 to 6.4 in kale and −0.6 to 9.3 in mustard. The lowest CSRI value for both crops was observed under the +T treatment. The highest CSRI value for kale was observed under +UV-B + CO₂ treatment, whereas for mustard it was observed under +T + CO₂ treatment. The CSRI values under all the treatments except +T treatment were positive in mustard. However, CSRI values under +T treatment and its combination with high UV-B levels (+UV-B + T) were negative for kale (Figure 5).
These factors can lead to lower absorption of sunlight and affect photosynthetic activity, been reported in Capsicum annuum [39,40] and Brassica napus [41]. Reduced height in plants exposed to UV-B radiation implied that the specific photomorphogenic response of plants could be related to a UV-B photoreceptor by UV-B radiation [42]. Moreover, to some extent, low photosynthetically active radiation (PAR, 400–700 nm) might have also affected plant height. Colett et al. [43] reported that increased PAR reduces the impacts of UV-B radiation on plant height [43]. In another study conducted by Conner and Zangori [44] with two other Brassica species (B. rapa and B. nigra), reduced plant height was observed in plants exposed to high UV-B radiation [44].

Heat stress decreases stem growth by reducing cell size through the loss of cell water content, resulting in reduced plant height [45–47]. Following our results, many crops have reported reduced plant height at higher temperatures, including recent reports in Brassica juncea [48] and Oryza sativa [49]. Heat stress mainly affects the plant meristems, promotes leaf senescence and abscission, and reduces photosynthesis, ultimately reducing plant growth [50,51]. The number of leaves was decreased in Brassica napus under high UV-B levels [52] and at high temperatures [53]. Another study, under controlled conditions, showed a decrease in leaf number in different quinoa (Chenopodium quinoa Willd.) varieties under elevated UV-B radiation levels [54,55], further confirming our results.

Raghuvanshi and Sharma [56] suggested that decreased concentration of photosynthetic pigments was associated with a decline of leaf area in Phaseolus vulgaris under UV-B treatment, which further resulted in reduced growth, stem length, and root dry weight. These factors can lead to lower absorption of sunlight and affect photosynthetic activity, leading to a decrease in photosynthesis and indirectly affecting plant growth. The UV-B radiations mainly reduce cell division and expansion, reducing leaf area [56–58]. In contrast, Nedunchezhian and Kulandaivelu [59] observed in cowpea that slightly elevated UV-B radiation increases leaf area i.
The decrease in marketable fresh weight under UV-B treatment in the present study is supported by Cechin et al. [60], who reported that the decline in fresh weight production in crops is predominately due to UV-B exposure. Plants exposed to UV-B might allocate more energy for other physiological activities, especially defense mechanisms, rather than biomass production, resulting in a UV-B-mediated synthesis of protective pigments, such as carotenoids, anthocyanins, or phenolic acids [61,62]. The increased CO$_2$ through increased activity of rubisco enzyme and reduced photorespiration increases leaf photosynthesis inside the leaf, leading to increased fresh weight and dry weight [63].

Results observed in *Arabidopsis thaliana* leaves [64], soybean [65], cotton [25], maize [29, 66], *Phaseolus vulgaris* [67], sweet potato [68], and basil [69] further corroborated our results, such as a reduction in total leaf area and fresh and dry weights, number of leaves, and height of plants (Table 3) under higher UV-B levels. In contrast to our results, Sakalauskaite et al. [70] reported an increase in plant height, leaf area, and dry weight under elevated UV-B in *Ocimum basilicum*. Reduction in leaf area in rice plants under high temperature alone and combination with elevated CO$_2$ concentration has recently been reported by Wang et al. [71]. High temperature and UV-B interaction decreased leaf area in *Brassica napus* [41]. A decrease in fresh and dry weight was reported in *B. campesteris* under high-temperature conditions [72].

Our study’s dry weight reduction can be explained as a reaction to stress caused by UV-B radiation in plant development and metabolism [73]. In *Beta vulgaris*, a decrease of 10–12% in dry weight was reported under high UV-B levels [74]. On the contrary, increased dry weight was observed under high UV-B levels in broad bean and wheat plants [75]. Similarly, studies on broad bean and wheat, *Prunella vulgaris* plants, when exposed to 15-day UV-B radiations in a growth chamber, showed an increase in whole plant dry weight [76]. This suggests that the UV-B effect is species/cultivar specific, and sometimes it benefits the growth and development of some crops [77]. Indeed, UV-B exposure led to a significant decrease in root weight which could have caused an increased shoot/root ratio. Similar results under high UV-B levels were observed in *Manihot esculentum* [78] and under +CO$_2$ levels in *Raphanus sativus* and *Daucus carota* [79].

Reduced plant weight under high temperatures can also be related to decreased photosynthesis, increased transpiration [80], and, in turn, reduced water use efficiency (WUE) [81]. A decrease in dry weight has been recently reported in three *Brassica* sp. [82], namely *Brassica oleracea* [83], *Raphanus sativus* [84], and *Chenopodium quinoa* [85], which further validates our study. High temperature and UV-B interaction decreased leaf weight in *Brassica napus* [41]. Mustard plants, compared to kale, showed higher dry weight under all the treatments. The concentration of CO$_2$ lessened the effects of high temperature and UV-B, resulting in lesser reductions. Interaction of high temperatures and elevated CO$_2$ increased plant height, the number of leaves, and leaf area in *Fragaria x ananassa* [86], *Capsicum annum* [87], and *Solanum lycopersicum* [88]. In contrast to our results, Wang et al. [71] reported reduced whole plant dry weight in rice under high temperatures and elevated CO$_2$ [71].

An increase in chlorophyll content under high temperatures has also been recently reported in other vegetable crops, such as tomatoes [89] and basil [90]. A recent study reported increased flavonoid content in kale under high UV-B levels [91], confirming our findings. Due to the strong antioxidant activity that flavonoids possess, higher total flavonoid content in the leaf implies a higher nutritional value in leaves [92]. Our results are consistent with the results of a study conducted by Olsson et al. [93], which reported a 70–150% increase in the overall flavonoid content of *B. napus* when subjected to high UV-B levels. Analogous to our results, an increase in anthocyanin content was reported in *Ocimum basilicum* under elevated UV-B levels by Sakalauskaite et al. [70]. The nitrogen balance index (NBI) is one of the critical indicators for crop growth. The NBI indicates C/N allocation changes due to N deficiency [94–96]. An increase in NBI was also observed in basil [90] and canola [97] under high temperatures.
A higher amount of wax for those leaves that were developed under elevated CO\(_2\) with UV-B radiation might have reduced the amount of incident UV-B radiation penetrating the plant tissue. The reduction in UV-B penetration most likely caused minor damage to the plants, which continued their relatively normal developmental process without a considerable loss in final yield. Qaderi and Reid [39] reported similar results under \(+\text{UV-B} + \text{CO}_2\) treatment in \(\text{Brassica napus}\). In contrast to our findings, Martel et al. [97] observed higher epicuticular wax in leaves of \(\text{Brassica napus}\) under high temperatures, and Steinmüller and Tevini [98] reported that enhanced UV-B radiation increased wax content by 23\% in barley and 28\% in the bean. In a study by Gonzalez et al. [99], six pea genotypes differing in their surface waxiness showed increased wax content under 6.5 kJ m\(^{-2}\) per day UV-B radiation. Higher wax content on the leaves of plants exposed to UV-B radiation indicates the importance of this chemical in plant defense mechanisms against environmental stresses. Both kale and mustard produced less wax content under high UV-B levels, pointing toward their weaker chemical defense against abiotic stresses.

The combined stress response index used in this study integrates the morphological and physiological responses, which reflect the overall sensitivity of kale and mustard to multiple stress conditions. The lowest CSRI value for both crops was observed under \(+T\) treatment, suggesting higher harmful effects of high-temperature treatment on all the parameters. The highest value for kale observed under \(+\text{UV-B} + \text{CO}_2\) treatment and for mustard under \(+T + \text{CO}_2\) treatment indicate the positive impacts of elevated CO\(_2\) concentrations.

5. Conclusions

Most current studies on plant stress response have mainly focused on the effect of individual stresses. However, combined, and sequential stress responses must be thoroughly studied to gain a meaningful understanding. The interaction of temperature stress, elevated CO\(_2\), and UV-B levels significantly impacted kale and mustard plants’ morphological and physiological processes. High temperature and UV-B conditions had considerable detrimental effects on most of the parameters while, under elevated CO\(_2\) concentration, a positive increase in all morphological and physiological traits was observed. This study recommends that varying the temperature and UV-B radiation levels in kale and mustard plants would significantly affect the growth and developmental rates and biochemistry compared to increasing the CO\(_2\) concentrations, which mitigates the constraining effects of temperature and UV-B stress. Farmers and researchers should, thus, attach much more importance to optimizing environmental conditions to enhance vegetable production [100,101]. Vegetables, such as kale and mustard, have been widely recommended in people’s daily diets as they provide various healthy compounds, such as antioxidants, vitamins, minerals, and dietary fiber [102]. Therefore, more research needs to be focused on the effect of multiple stresses on the nutritional quality of leafy vegetables.

**Author Contributions:** Conceptualization, K.R.R. and T.C.B.; methodology, K.R.R., C.H.W. and T.C.B.; software, A.S. and C.H.W.; validation, K.R.R. and T.C.B.; formal analysis, K.R.R. and T.C.B.; investigation, K.R.R., C.H.W. and T.C.B.; resources, K.R.R. and T.C.B.; data curation, A.S., K.R.R. and T.C.B.; writing—original draft preparation, A.S.; writing—review and editing, A.S., K.R.R., C.H.W., T.C.B., S.B., D.C. and W.G.; visualization, A.S. and K.R.R.; supervision, K.R.R. and D.C.; project administration, K.R.R.; funding acquisition, K.R.R., T.C.B. and D.C. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research is partially funded by the National Institute of Food and Agriculture, NIFA 2019-34263-30552 and MIS 043050, and the USDA-NIFA Hatch Project’s work under accession number 149210.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.
Acknowledgments: This research was partly supported by the US Department of Agriculture (USDA), Agricultural Research Service, under agreement number 58-6066-6-045 and National Institute for Food and Agriculture, NIFA 2016-34263-25763 and MIS 043040. The findings and conclusions in this publication are those of the authors and should not be construed to represent any official USDA or US Government determination or policy. Mention of trade names or commercial products in this publication provides specific information and does not imply recommendation or endorsement by the USDA. USDA is an equal opportunity provider and employer. We thank David Brand for his technical assistance and graduate students at the Environmental Plant Physiology Laboratory for helping during data collection.

Conflicts of Interest: The authors declare no conflict of interest.

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