ABSTRACT

Recent studies have reported the use of IgY antibody in the prevention or treatment of diseases in animals. IgY can be obtained in large amounts from the yolk of chicken eggs through a low-cost process. This study evaluated the effect of different routes of inoculation on antibody production and spleen morphological characteristics of laying hens (White Leghorn) inoculated with sheep red blood cells. The analysis of the results showed that the intramuscular route is the most efficient for total antibody production in the primary immune response, while the intravenous route is the most efficient in producing IgY antibodies in the secondary immune response. No histological changes were observed in the spleen of laying hens. This study could be useful for developing protocols of antigen inoculation in laying hens for IgY antibody production.

INTRODUCTION

The main class of immunoglobulins that occurs in chickens is IgY (Warr et al., 1995). It is found in high concentrations in the egg yolk and its structural and functional features are similar to those of mammalian IgG (Dias da Silva & Tambourgi, 2011; Taylor et al., 2009). For instance, the Fc region of IgY is responsible for its biological functions, such as complement fixation and opsonization (Warr et al., 1995). Compared with antibody production in mammals, IgY from the yolk of chicken eggs has several advantages, including the possibility of large scale production without bleeding or sacrificing the chicken and the lack of reactivity with the complement system or immunoglobulin receptors of mammals (Dias da Silva & Tambourgi, 2010; Xiao & Gao, 2010). Furthermore, several studies have investigated the usefulness of IgY antibodies in the prevention or treatment of diseases in animals (Vega et al., 2011; Xu et al., 2011).

The route of inoculation is a critical point in immunization and for antibody production in mammals and birds (Fyfe et al., 1991; Kreukniet et al., 1992; Boa-Amponsem et al., 2006). Moreover, the route of inoculation influences the class of antibodies produced (Hu et al., 2007). Thus, identifying the optimal route of antigen inoculation in laying hens could result in new technologies based on IgY antibodies for the diagnosis, prevention and treatment of disease in humans and animals.

This study investigated the effect of the inoculation route on antibody production and spleen histological characteristics of laying hens inoculated with sheep red blood cells.

MATERIAL AND METHODS

Sixteen 45-week-old laying hens (White Leghorn) were randomly
assigned to four groups (n=4/group) and maintained in individual cages at ambient temperature, receiving water and feed (Purina, Brazil) ad libitum. Control (c) group chickens were inoculated with PBS pH 7.2 (250 µl), while each bird of the intravenous (iv), intramuscular (im) or subcutaneous (sc) groups was inoculated with 2.5% sheep red blood cells (SRBC) in PBS pH 7.2 (250 µl). SRBC inoculation was performed in the brachial vein (iv group), the breast muscle (im group), or the right wing (sc group).

All chickens were inoculated on days 1 and 28, and sera samples were obtained before the first inoculation and on days 7 and 35. The birds were sacrificed on day 35 and their spleens were collected and processed, as previously described by Beçak & Paulete (1970). Antibody titers against SRBC were determined as described in Boa-Amponsem et al., 2001. Morphometric analysis of the spleen was performed by image analysis (MOTICAN, Motic Company, Xiamen, China) using an optical microscope at 100X (germinal centers) and 400X (areas of white and red pulp) magnifications. The germinal centers were counted on five slices per bird. Ten images per slice from each chicken were analyzed to determine the areas of white and red pulp. The experimental procedures were approved by the Ethics Committee for Animal Research of the Londrina State University (Universidade Estadual de Londrina).

Normality distribution was assessed by the Shapiro-Wilk’s test; the significance level was set at p<0.05 for non-normalized data. Intergroup differences were analyzed by means of ANOVA and the post hoc Tukey’s test (parametric data) or Kruskal-Wallì’s and post hoc Dunn’s test (nonparametric data), with differences considered significant when p<0.05.

RESULTS

Regarding the primary response (day 7), the im and sc groups presented increased total antibody production; however, neither the IgM nor IgY titers increased significantly, regardless of the route analyzed.

Considering the secondary immune response (day 35), analysis verified a significant increase in total antibody titers in the iv, im and sc groups compared with the controls. Furthermore, the total antibody titers of the iv group were increased compared to the sc, but not the im group. Regarding the IgY antibody titers, increased levels in the iv group were observed in relation to the remaining groups. The levels of IgM antibodies showed a significant difference only between the control and im groups (Figure 1).

Histological analysis of the spleen following the 2nd inoculation of SRBC showed no differences in the areas of red and white pulp (Figure 2). No significant differences occurred in the number of germinal centers (Figure 2).

DISCUSSION

The capacity for antibody production can be influenced by several factors, including the inoculation route (Mozdzanowska et al., 2007; Kreukniet et al., 1992; Muir et al., 1998). In the present study, for the
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Regarding the secondary immune response, the iv route resulted in higher total antibody titers than the sc route. The difference in the total antibody levels was mainly due to increased levels of IgY. However, chickens from the im group also presented increased total antibody titers, and this difference was caused by IgM antibody titers, rather than increases in IgY antibody levels, as expected for secondary immune response (Freitas et al., 2011). The predominance of IgM in the secondary immune response was also observed in chickens selected for low antibody production (Boa-Amponsem et al., 2001). These results suggest that the effectiveness of the routes studied is different in primary and secondary responses and that the iv route seems to provide better performance in the second immunization.

Since the iv route delivers antigens to the spleen lymphoid tissues, the primary site of antibody production after immunization by this route (White et al., 1975), analysis of morphological changes in the spleen of the inoculated birds was performed. No significant changes were detected in the spleens of the birds inoculated with 2.5% SRBC, independently of the route used, probably because the concentration of SRBC was not sufficient to stimulate significant changes in the spleen. Other secondary lymphoid tissues could also contribute to IgY production.

In summary, this study demonstrated that the inoculation route influences both the antibody levels and the class of antibodies produced by laying hens. These results could be meaningful for establishing protocols for the inoculation of laying hens to produce antibodies.

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