Ginsenoside-Rg3 inhibits the proliferation and invasion of hepatoma carcinoma cells via regulating long non-coding RNA HOX antisense intergenic

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ABSTRACT

Ginsenoside Rg3, a natural compound, has been reported to function as an anticancer agent for hepatoma carcinoma, while the mechanisms underlying the anticancer effects are not clear. Therefore, the objective of our study was to explore the impact of RG3 on cell migration and invasion by regulating the lncRNA HOX antisense intergenic (HOTAIR) expression involving PI3K/AKT signaling pathway. qRT-PCR was utilized to measure the mRNA expression of HOTAIR. Furthermore, HOTAIR overexpression plasmids were transfected to SMMC-7721 and SK-Hep-1 cells. Additionally, MTT assay was used to evaluate the proliferation of transfected cells. The protein levels were determined with Western blot. lncRNA HOTAIR was overexpressed in SMMC-7721 and SK-Hep-1 cells. Ginsenoside-Rg3 reduced the level of lncRNA HOTAIR. Overexpressed lncRNA HOTAIR offset ginsenoside-Rg3 inhibited proliferation, migration and invasion of HCC cells. Furthermore, ginsenoside-Rg3 decreased the expression of p-AKT, p-PI3K, matrix metalloproteinase-2 (MMP2) and matrix metalloproteinase-9 (MMP9), which was reversed after the treatment of HOTAIR. LncRNA HOTAIR was overexpressed in SMMC-7721 cells. Ginsenoside-Rg3 could reduce the expression of lncRNA HOTAIR, resulting in the inhibited cell proliferation, migration and invasion. Furthermore, ginsenoside-Rg3 inhibited cell proliferation and invasion ability through the PI3k/AKT pathway. Thus, ginsenoside-Rg3 might be a potential and effective treatment for HCC.

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Introduction

Hepatocellular carcinoma (HCC) is one of the most common cancers around the world, which has high incidence rate and mortality. Nearly 748,300 new cases and 695,900 deaths are reported per year worldwide [1]. Patients with a history of cirrhosis are most likely to develop to HCC [2]. The prevailing therapies for HCC are to rely on surgical treatment or local ablation [3]. However, the lack of specific symptoms in the early stage of HCC is one of the main reasons for the poor curative effect of HCC [4]. More than 60% of patients are diagnosed with advanced cancer with the status of metastasis and an overall 5-year survival rate of less than 16% [5]. Therefore, it is urgent to effectively retard the cell migration and invasion ability in order to prolong the survival rate and cure rate of patients with advanced HCC.

Long non-coding RNA (lncRNA), a single-stranded RNA containing more than 200 nucleotides, which lacks significant open reading frames, are unable to code for proteins [6]. Furthermore, lncRNAs are key regulators of cellular function through epigenetic regulation, miRNA sponging, enzyme cofactors and modulating of proteins [7]. Therefore, lncRNAs are a major class of RNA molecules which exert a powerful effect on various physiological and pathological processes involving in tumor growth, progression, and prognosis. In addition, lncRNAs could provide potential clues for developing novel therapeutic approaches for cancers [8]. Previous studies have found that lncRNA HOX antisense intergenic (HOTAIR) is significantly increased in cancerous tissues in comparison to normal tissues in gastric cancer, urothelial cell carcinoma and renal cell carcinoma [8–10]. Additionally, lncRNA HOTAIR is closely related to cell proliferation and migration ability [10]. In Di et al.’s study, lncRNA HOTAIR epigenetically inhibited the level of miR-122 in hepatocellular carcinoma via DNA methylation, which contributes to the activation of Cyclin G1 and promotes the malignant development of HCC [11]. Furthermore, the study determined that the high level of lncRNA HOTAIR was related to hepatocarcinogenesis and metastasis, and overexpression of lncRNA HOTAIR is also a predictor of tumor recurrence in HCC [12,13].

Ginsenoside Rg3 (C_{42}H_{72}O_{13}; molecular weight 785.01) is an effective component extracted from Ginseng [14]. Ginsenoside Rg3 is the first monomer of traditional Chinese medicine applied to the treatment of various kinds of tumors due to its tumor angiogenesis-inhibiting effect and interaction between endothelial cells and the extracellular matrix [15]. So far, ginsenoside-Rg3 has been regarded as an effective drug for postoperative recovery of most of the malignant tumors, which can significantly alleviate the recurrence and diffusion of tumors [16]. Ginsenoside-Rg3 not only suppresses the migration and invasion but also promotes the apoptosis of colorectal cancer cells through reducing the level of LncRNA CCAT1. Ginsenoside-Rg3 suppresses the migration and invasion of hepatoma cells through regulating ARHGAP9. Phosphoinositide-3-kinase (PI3K)/AKT signaling pathway is closely related to various factors, and its activation and inhibition regulate the important cell activity including cell proliferation, migration, invasion and so forth [17]. Furthermore, the cell migration and invasion-related genes are regulated by PI3K/AKT, such as metalloproteinase 2 (MMP2) and matrix metalloproteinase 9 (MMP9) [18]. However, the specific mechanism of ginsenoside-Rg3 on the lncRNA HOTAIR expression of HCC cells remains unclear.

Therefore, this study is the first to explore the effect and mechanisms of ginsenoside-Rg3 on lncRNA HOTAIR in hepatoma carcinoma cells. We hypothesized that ginsenoside-Rg3 inhibits the lncRNA HOTAIR expression, growth and metastasis of the HCC cells via regulating the PI3k/AKT signaling pathway.

Materials & methods

Cells

SMMC-7721, SK-Hep-1 and HEK293T cells were purchased from ATCC and cultured in RPMI-1640 medium containing 10% fetal bovine serum, 100 U/ml penicillin, and 100 ng/ml streptomycin to the 70–80% confluence for the following experiments at 37°C with 5% CO2.

Transfection

SMMC-7721 and SK-Hep-1 cells were seeded in 24-well plates. Furthermore, there were four groups in the cell experiment: control group,
ginsenoside-Rg3 groups, HOTAIR overexpression group, and ginsenoside-Rg3+ HOTAIR control group. Furthermore, the HOTAIR overexpression and control plasmids were constructed by GenePharma. The cells were transfected with the plasmids with 2.5 µl of Lipofectamine 2000 (Invitrogen, USA) and treated with 8 µg/ml of ginsenoside-Rg3, which was also added to the transfected cells. This experiment was performed in triplicate. After 6-h incubation in RPMI-1640 at 37°C with 5% CO₂.

**qRT-PCR**

qRT-PCR was employed to detect the mRNA expression of IncRNA-HOTAIR. According to the manufacturer’s protocol, RNA was isolated by RNeasy Mini Kit (Qiagen, Germany), and the RNA concentration was determined by spectrophotometer nanodrop 2000. The M-MLV was applied to synthesize cDNA through reverse transcription. The running order is 43°C, 30 min; 97°C, 5 min; and 5°C, 5 min. PrimeScript™ RT-PCR Kit (TaKaRa, Japan) was used for RT-PCR. The running order is 95°C, 5 min; 95°C, 30 s, 40 cycles; 59°C, 30 s; and 72°C, 30 s. Each experiment was performed in triplicate. The abundance of gene expression was determined by 2^−ΔΔCt relative quantification [19]. GAPDH was regarded as an internal control.

**Western blot**

Western blot was applied according to previous study [20]. Cells were lysed by RIPA (Thermo Fisher Scientific, USA). Total protein levels were verified by BCA kit (Thermo Fisher Scientific, USA). 30 µg of proteins was electrophoresed in 15% SDS–PAGE and then transferred to polyvinylidene difluoride membranes (Millipore, Netherlands). The membrane was blocked with 5% skimmed milk for 2 h. Subsequently, the membrane was incubated with the primary antibodies including anti-MMP2 (ab92536, 1:1000, Abcam, USA), -MMP9 (ab38898, 1:1000, Abcam, USA), AKT (ab8805, 1:10,000, Abcam, USA), -PI3K (ab32089, 1:1000, Abcam, USA), -p-AKT (phospho S474, 1:1000, Abcam, USA), and -p-PI3K (phospho Y607, 1:1000, Abcam, USA) for 1 h at indoor temperature. Furthermore, the membrane was incubated with secondary antibody (ab6721, 1:2000, Abcam, USA) conjugated with HRP for 45 min at indoor temperature. Then, ECL Western blotting kit (Santa Cruz Biotechnology, Inc.) was adopted for membrane stain and analyzed with ImageJ software. Each experiment was repeated three times.

**MTT**

We measured cell proliferation by MTT assay as described by Kumar et al. [21]. The transfected SMMC-7721 and SK-Hep-1 cells were placed into 24-well plates and incubated at 37°C with 5% CO₂ for 48 h. Cells were treated with 1, 2, 4, 8, and 16 µg/ml of ginsenoside-Rg3. Subsequently, MTT Cell Viability Assay Kit (Thermo Fisher Scientific, USA) was performed according to the manufacturer’s protocol. Moreover, we detected OD value under 570 nm wavelength using a microplate reader at the 0, 12, 24, 48 and 72 h time points.

Then, the lowest cell viability was in the group treated with 8 µg/ml ginsenoside-Rg3; therefore, 8 µg/ml ginsenoside-Rg3 was selected for the following experiments.

**Scratch assay & transwell assay**

We evaluated the migration and invasion ability of SMMC-7721 and SK-Hep-1 cells through Scratch assay and transwell assay according to Ni et al. [22]. The SMMC-7721 and SK-Hep-1 cells were placed into 96-well plates and a monolayer cell culture was obtained. Then, a scratch across the center of the wells was generated by a new 1-ml pipette tip. Therefore, the width of the scratch was equal to the outer diameter of the tip. Then, the cells were incubated at 37°C with 5% CO₂ for 72 h. The migrated cells were captured with a microscope (Olympus, Tokyo, Japan).

Furthermore, the invasion ability of SMMC-7721 cells and SK-Hep-1 cells was measured by transwell assay. The transwell cell culture inserts (8-mm pore size; Falcon; BD Biosciences) were seeded into the wells of 96-well plates to create the separate upper and lower chambers. Furthermore, the upper side of the membrane was pre-coated with Matrigel (BD Biosciences) and incubated for 1 h at 37°C for gel formation. The membrane was hydrated with FBS 2 h. Then, RPMI-1640 (600 µl) containing 10% FBS and 1 × 10⁵ cells/well was added to the lower and
upper chambers, respectively. Then, we calculated invading cells by counting chamber after 48 h incubation.

**Statistical methods**

All experimental data were presented as the mean ± standard deviation (SD) and analyzed by using GraphPad Prism version 5.01 (GraphPad Software, La Jolla, CA, USA). Moreover, the data comparison was employed by t-test and one-way analysis. \( p < 0.05 \) was taken as being statistically significant difference.

**Results**

The present study aimed to explore the effect of ginsenoside-Rg3 on the tumorigenic behavior of the HCC cells. We hypothesized that ginsenoside-Rg3 downregulated the lncRNA HOTAIR expression and inhibited the proliferation, migration and invasion of the SMMC-7721 and SK-Hep-1 cells via regulating the PI3k/AKT signaling pathway.

**Cell viability under different concentrations of ginsenoside-Rg3**

SMMC-7721 Cells were treated with 1, 2, 4, 8 and 16 μg/ml of ginsenoside-Rg3. As shown in Figure 1, in inhibiting the cell viability of SMMC-7721, 8 μg/ml of ginsenoside-Rg3 was more effective. Therefore, the 8 μg/ml of ginsenoside-Rg3 was selected for the following performance.

**Reduced lncRNA-HOTAIR expression by ginsenoside-Rg3**

Using qRT-PCR, we evaluated the relative expression of lncRNA-HOTAIR. Figure 2 indicates that ginsenoside-Rg3 could significantly suppress the expression of lncRNA-HOTAIR compared to the control group. Therefore, we might conclude that ginsenoside Rg3 could inhibit the lncRNA-HOTAIR expression in SMMC-7721 cells and SK-Hep-1 cells.

**LncRNA-HOTAIR overexpression system**

The lncRNA-HOTAIR-negative control plasmids and lncRNA-HOTAIR overexpression plasmids were added to SMMC-7721 cells and SK-Hep-1 cells. Figure 3 shows that the lncRNA-HOTAIR was dramatically overexpressed, which means that the lncRNA-HOTAIR overexpression plasmids were successfully constructed and can be applied to the following experiments.

![Figure 1. Cell viability under different concentrations of ginsenoside Rg3.](image)

The SMMC-2271 cells were treated with 0, 1, 2, 4, 8 and 16 μg/ml ginsenoside Rg3 and the cell viability was evaluated by MTT assay after incubation.
Inhibited cell proliferation rate by ginsenoside Rg3

The SMMC-7721 cells and SK-Hep-1 cells were treated with IncRNA-HOTAIR-negative control plasmids, ginsenoside Rg3 and IncRNA-HOTAIR overexpression plasmids and cultured RPMI-1640 (600 µl) containing 10% FBS for 48 h at 37°C/5% CO₂. The viability of the cell was measured by MTT assay. Figure 4 demonstrates that the SMMC-7721 cell viability was significantly reduced in ginsenoside Rg3 groups and IncRNA-HOTAIR-negative control groups compared to the control group. Therefore, ginsenoside Rg3 could induce the decreased cell viability, which might inhibit the development of hepatoma carcinoma.

The migration and invasion ability of cell inhibited by ginsenoside Rg3

After incubation with IncRNA-HOTAIR-negative control plasmids, ginsenoside Rg3 and IncRNA-HOTAIR overexpression plasmids for 48 h, the
Figure 3. LncRNA-HOTAIR overexpression system.

The LncRNA-HOTAIR-negative and overexpression plasmids were added to SMMC-7721 cells and incubate with RPMI-1640 (600 µl) containing 10% FBS at 37°C with 5% CO2 for 48 h. The relative expression of LncRNA-HOTAIR was determined by qRT-PCR. Control: non-treated group; pcDNA3.1: cells treated with LncRNA-HOTAIR-negative control plasmids; pcDNA3.1-HOTAIR: cells treated with LncRNA-HOTAIR overexpression plasmids. **p < 0.05 vs. control group.
migration and invasion ability of cells were evaluated with scratch and transwell assay. In Figures 5 and 6, the results indicated that a huge reduction in the migration and invasion ability of cells was found in ginsenoside Rg3 and lncRNA-HOTAIR-negative control groups compared with control and lncRNA-HOTAIR overexpression groups. Therefore, ginsenoside Rg3 might efficiently inhibit the migration and invasion of SMMC-7721 and SK-Hep-1 cells.
Inhibited expression of MMP2, MMP9, p-AKT, and p-PI3K by ginsenoside Rg3

The protein expression was determined by Western blot. There were four groups: control group, ginsenoside Rg3 group, ginsenoside Rg3 plus IncRNA-HOTAIR overexpression group and ginsenoside Rg3 plus IncRNA-HOTAIR-negative control group. In Figure 7, the dramatically inhibited expression of MMP2, MMP9, p-AKT, and p-PI3K was observed in ginsenoside Rg3 groups compared to the control group. Furthermore, the MMP2, MMP9, p-AKT, and p-PI3K were significantly increased in IncRNA-HOTAIR overexpression group treated with ginsenoside Rg3. Hence, ginsenoside Rg3 could induce the reduced expression of MMP2, MMP9, p-AKT, and p-PI3K in SMMC-7721 cells.

Discussion

In our study, we first found that ginsenoside Rg3 downregulates the IncRNA-HOTAIR expression and relieves the carcinogenic behaviors of HCC cells via inhibiting the PI3k/AKT signaling...
pathway. Our results demonstrated that ginsenoside Rg3 was a potent multi-target antitumor agent for the treatment of HCC in conclusion.

The indefinite proliferative ability of HCC cells is the main factor leading to the poor curative effect and recurrence of HCC. Effective inhibition of cell growth and metastasis is the main strategy to alleviate the occurrence and development of HCC [23]. In recent years, a growing number of studies have reported that natural extracts have excellent therapeutic effect on HCC. For example, chrysin can prevent sphere formation in SMMC-7721 cells [24]. Berbamine promoted the apoptosis of SMMC-7721 cell via mitochondrial signaling pathway [25]. Xu et al. [26] also suggested that isovitexin induces apoptosis and suppresses the proliferation of SK-Hep-1 cells. In addition, studies have shown that ginsenosides exhibit significant antitumor effect. Dai et al. [27] found that ginsenoside Rb2 suppresses epithelial–mesenchymal transition development of colorectal cancer cells. Similarly, Li et al. [28] demonstrated that ginsenoside Rh2 can transform tumor-associated macrophages from M2 to M1 subgroup and inhibit the migration ability of lung cancer cells. In this study, we found that ginsenoside Rg3 prevents
the growth and metastasis of the HCC cells, which was also found in the study by Shan et al. [29].

As a prognostic circulating marker and potential therapeutic target in patients with tumor diseases, lncRNA has been studied currently in various cancers [30]. Han et al. [31] confirmed that lncRNA-DNAJC3-AS1 promotes the progression and development of colon cancer. Zhang et al. suggested that lncRNA H19 is considered to be closely related to the extent of malignant degree [32]. These reports indicated that lncRNA may be an effective target for tumor molecular therapy. Previously, it was studied that IncRNA HOTAIR was associated with metastasis, differentiation, and early recurrence. Furthermore, knockdown of HOTAIR IncRNA diminishes cell proliferation and is associated with decreased levels of MMP-9 and vascular endothelial growth factor protein, which are crucial for cell motility and metastasis [33]. According to its asymmetric carbon atom C20, Ginsenoside Rg3 can be divided into the R type and S type. And due to the activation or inhibition of different gene expression, Ginsenoside Rg3 also has diverse pharmacological effects in vivo and in vitro [34]. According to our report, through decreasing the expression of epidermal growth and upregulating the protein expression of pro-apoptotic P53, ginsenoside Rg3 can inhibit cancer cell proliferation and induce apoptosis in HCC [34]. In the present study, we verified that 8 μg/ml ginsenoside Rg3 could induce the downregulation of IncRNA-HOTAIR, leading to the reduced rate of cell proliferation, migration and invasion via PI3K/AKT signaling pathway.

However, due to the limited time and resources, our experimental design has some shortcomings. For instance, ginsenoside Rg3 has various targets in cancers, and lots of targets may result in the same or opposite effects on cancer. In our present study, the group of IncRNA-HOTAIR knockdown lacked. Therefore, we cannot compare the cell proliferation, invasion and migration ability between ginsenoside Rg3 group and ncRNA-HOTAIR knockdown group. Thus, we could not identify the exact effects of ginsenoside Rg3 on SMMC-7721 cells and SK-Hep-1 cells and the associated impacts on HCC.

**Conclusion**

To sum up, we found that ginsenoside Rg3 significantly inhibits the proliferation of the SMMC-7721 and SK-Hep-1 cells. 8 μg/ml ginsenoside Rg3 downregulated the IncRNA-HOTAIR expression and suppressed the growth and metastasis of the SMMC-7721 and SK-Hep-1 cells, while over-expression of IncRNA-HOTAIR reversed the role of ginsenoside Rg3. In addition, this study
demonstrated that ginsenoside Rg3 relieves the carcinogenic behaviors of the SMMC-7721 and SK-Hep-1 cells by inhibiting the PI3k/AKT signaling pathway. Our study laid a theoretical basis for the further promotion and application of ginsenoside Rg3 and provided a novel insight for the research on the therapy of HCC.

**Highlights**

1. Ginsenoside-Rg3 could significantly suppress the expression of lncRNA-HOTAIR
2. Ginsenoside Rg3 could induce the decreased cell viability
3. Inhibition of PI3k/AKT signaling pathway

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**Disclosure of potential conflicts of interest**

The authors declare that they have no competing interests.

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