Platelet Distribution Width at First Day of Hospital Admission in Patients with Hemorrhagic Fever with Renal Syndrome Caused by Hantaan Virus May Predict Disease Severity and Critical Patients’ Survival

Xiude Fan,1 Zitong Liu,1,2 Shiqi Fu,1,2 Jiao Sang,1 Huan Deng,1 Fang Li,1 Xiaoge Zhang,1 Na Li,1 Qunying Han,1 and Zhengwen Liu1

1Department of Infectious Diseases, First Affiliated Hospital of Xi’an Jiaotong University, Xi’an, 710061 Shaanxi, China
2Xi’an Medical University, Xi’an, 710021 Shaanxi, China

Correspondence should be addressed to Zhengwen Liu; liuzhengwen113@xjtu.edu.cn

Received 21 March 2018; Accepted 24 May 2018; Published 19 June 2018

Academic Editor: Alexandra Scholze

Copyright © 2018 Xiude Fan et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Thrombocytopenia is one of the main characteristics of hemorrhagic fever with renal syndrome (HFRS). This study aimed to evaluate the associations of platelet distribution width (PDW) with the disease severity and critical patients’ survival of HFRS.

The demographics, clinical data, and white blood cell and platelet parameters including PDW in 260 patients hospitalized for HFRS were analyzed. The results showed that PDW on the first day (PDW1) was positively associated with the disease severity (p = 0.005). Multiple regression analysis showed that in addition to age (odds ratio [OR], 1.091; 95% confidence interval [CI], 1.015–1.172) and occurrence of sepsis (OR, 22.283; 95% CI, 2.985–166.325), PDW1 (OR, 0.782; 95% CI, 0.617–0.992) was a risk factor of the mortality, having an area under the receiver operating characteristic curve of 0.709 (95% CI, 0.572–0.846, p = 0.013) for predicting mortality, with a sensitivity of 70% and a specificity of 67% at a cutoff of 16.5 fL, in patients with critical HFRS. These results suggest the potential of PDW at the first day of hospitalization as a valuable parameter for evaluating the severity of HFRS and a moderate parameter for predicting the prognosis of critical HFRS patients. A prospective study in large patient population is needed to validate these findings.

1. Introduction

Hemorrhagic fever with renal syndrome (HFRS), a zoonotic disease caused by pathogenic hantaviruses, is characterized by altered vascular permeability all over the body and may exhibit severe profile with sudden fever, acute renal failure, shock, and hemorrhage [1]. HFRS is mainly endemic in Asia, Southeast Europe, and North Africa [2–4]. China is the most seriously affected country and accounts for over 90% of the total number of HFRS cases all over the world. In China, two HFRS-causing hantaviruses, Hantaan and Seoul viruses, which induce a severe and a mild form of HFRS, respectively, are the main agents of HFRS diseases and the endemic of these two viruses is geographically quite stable [5]. Due to the promotion of vaccine in China, the prevalence of HFRS has been decreased in recent years [6]. However, there has been no specific antiviral therapy for HFRS up to now and the high mortality of critical cases persists in certain areas [7]. According to the criteria of clinical classification of China, HFRS is classified into four clinical types: mild, medium, severe, and gravis [8]. These criteria of clinical classification depend largely on doctors’ subjective judgment, and the incidence of atypical patients with unusual clinical manifestations has increased. Currently, no reliable prognostic parameters have been available for the patients, especially the critically ill patients. These may hamper the early diagnosis and treatment of HFRS. Therefore, exploring new and early predictors to assess the severity and prognosis of HFRS...
beforehand is important to guide the clinician to initiate effective treatment and improve clinical management.

Platelets (PLTs), fragments of megakaryocyte cytoplasm, play a prothrombotic role in the process of stopping bleeding at the site of the interrupted endothelium [9]. Platelet parameters obtained by automated cell counters consist of PLT count, platelet distribution width (PDW), mean platelet volume (MPV), platelet-large cell rate (P-LCR), and plateletcrit (PCT). Decreased PLT count was shown to precede acute kidney failure in patients with Puumala virus infections [10], and PLT count was an independent predictor of severe HFRS [11] and critically ill patients in intensive care unit (ICU) [12]. In addition, PDW, MPV, P-LCR, and PCT are associated with platelet activation and function and have recently been shown to be useful markers for predicting occurrence and prognosis of certain inflammatory diseases and vascular diseases, such as Crimean-Congo hemorrhagic fever [13], sepsis [14], pulmonary embolism [15], ascitic fluid infection [16], obstructive sleep apnea [17], Crohn’s disease [18], and coronary artery disease [19]. It is well known that platelet parameters are routinely tested on HFRS patients in hospitals almost every day. Nevertheless, there is limited information on their clinical significance in HFRS.

The aim of the current study, therefore, was to investigate whether platelet parameters would be useful in evaluating the severity of HFRS disease and predicting the prognosis of HFRS patients.

2. Materials and Methods

2.1. Study Population. This retrospective study was conducted in patients from the First Affiliated Hospital of Xi’an Jiaotong University. The patients were all residents of Shaanxi Province in China, an endemic area of Hantaan virus infection. Records of 321 HFRS patients (mean age 41.79 ± 16.75 years; 243 men and 78 women) from January 2011 to December 2016 were collected.

The diagnosis of HFRS was confirmed by clinical manifestations in combination with serological evidence of the presence of IgM or IgG antibodies against hantavirus determined by indirect immunofluorescence test. For the present investigation, only adult patients were included and patients who had pregnancy, cancer, hemopath, other kidney diseases, liver diseases, and the use of anticoagulants prior to admission were excluded. Based upon the diagnosis criteria for the prevention and treatment strategy of HFRS published by the Ministry of Health, China, the disease severity in HFRS patients was classified into mild, medium, severe, and grave types. Accordingly, 58 patients were classified as mild type, 75 medium type, 44 severe type, and 83 grave type. The patients with grave type HFRS were divided into nonsurvival and survival groups to analyze factors associated with prognosis.

2.2. Data Management. Personal features and clinical data collected from the patient’s medical chart included the patient’s age, sex, max temperature prior to admission, days from fever onset to hospital admission, blood pressure at the time of the assessment, cigarette and alcohol consumption, hospital stay, blood transfusion, continuous renal replacement therapy (CRRT), comorbidities (hypertension, diabetes mellitus, and coronary heart disease), and HFRS-related complications (hemorrhage, infection, hepatic injury, kidney rupture, sepsis, multiple organ dysfunction syndrome (MODS), and arrhythmia). Hemorrhage refers to patients presenting with bleeding (pulmonary hemorrhage, melena, hematemesis, hematuria, diffuse ecchymosis, and hemotoma) throughout hospitalization. Sepsis was defined according to criteria determined by Vincent et al. [19]. Blood transfusion, CRRT, comorbidity, and HFRS-related complications were reported as a total count.

Blood tests routinely measured by autoanalyzers on the day of hospital admission and on the third day of hospitalization were collected. Variables were as follows: WBC, PLT, PDW, MPV, P-LCR, and PCT. Routine blood test results like PDW values on the day of hospital admission were recorded as PDW1 and those on the third day of admission as PDW3. The study protocol was performed to conform with the Declaration of Helsinki and was approved by the Ethics Committee of the First Affiliated Hospital of Xi’an Jiaotong University. Informed consent was not obtained from the patients as all patient records/information was anonymized and deidentified prior to analysis.

2.3. Statistical Analysis. Statistical analysis was performed using SPSS 20.0 software (SPSS Inc., Chicago, IL, USA) and MedCalc software version 12 (MedCalc Software, Ostend, Belgium). Quantitative variables were expressed as mean and standard deviation or median and interquartile range, where appropriate. The normality of the distribution and the homogeneity of variance were assessed with the Kolmogorov-Smirnoff test and Levene’s test, respectively. Demographic information and blood routine parameters were compared by the independent sample t-test or one-way ANOVA test for normally distributed variables. The nonnormally distributed variables were compared by the nonparametric Kruskal-Wallis test. The distributions of MPV levels in the first day conformed to normal distribution, and one-way ANOVA test was used for statistical analysis. Numerical variables such as age, max temperature, blood pressure, hospital stay, and other laboratory parameters did not coincide with normal distribution; these variables were compared by the nonparametric Kruskal-Wallis test. Chi-square test and Fisher exact test were used for categorical variables. Spearman correlation was performed to indicate the direction of association between parameter and disease severity (positive or negative association). Tolerance and variance inflation factor (VIF) were used to detect multicollinearity among variables. Variables significantly associated with death were tested in a logistic regression model for their potential to predict the corresponding outcome. Variables with statistically significant results in the univariate analyses and without multicollinearity were included in multivariate logistic regression analyses for independent variables. Predictive values of the laboratory parameters for prognosis were tested with receiver operating characteristic curves (ROC) and quantified by calculating the area under the ROC curve (AUC) and the 95% confidence interval (CI). MedCalc
software was used to test the statistical significance of the difference between the AUCs. A p value < 0.05 was considered statistically significant. All p values were from two-sided tests.

3. Results

3.1. Patient Characteristics. According to inclusion and exclusion criteria, records of 260 adult HFRS patients were finally included in this study (Supplementary Figure 1). The characteristics of patients included are shown in Table 1. The 260 patients had a mean age of 44.82 ± 14.99 years, and 74.61% were male (Table 1). Nineteen HFRS patients died, respectively correlated with the severity of disease (all p < 0.001). Among HFRS-related complications, the incidences of hemorrhage, infection, and hepatic injury increased gradually as the severity of the disease increased (Table 1). Kidney rupture, sepsis, and MODS were only observed in severe and gravis patients. Arrhythmia was not identified in mild patients. Furthermore, the severe and gravis patients tended to have longer hospital stays and more frequent blood transfusions and CRRT than the mild and medium patients (p < 0.001, Table 1).

3.2. Complication Related to the Severity of HFRS Patients. Among HFRS-related complications, the incidences of hemorrhage, infection, and hepatic injury increased gradually as the severity of the disease increased (Table 1). Kidney rupture, sepsis, and MODS were only observed in severe and gravis patients. Arrhythmia was not identified in mild patients. Furthermore, the severe and gravis patients tended to have longer hospital stays and more frequent blood transfusions and CRRT than the mild and medium patients (p < 0.001, Table 1).

3.3. Laboratory Parameters Related to the Severity of HFRS Patients. The levels of WBC count, PLT count, PDW, MPV, P-LCR, and PCT on the first day of admission demonstrated significant differences among the four clinical types. Levels of WBC1 and WBC3 increased gradually with the severity of the disease. On the first day of admission, PDW and MPV levels in the severe and gravis patients were higher than those in the mild and medium patients. Levels of PLT and PCT in the first and third day were decreased significantly with the severity of the disease, and these two indices were not significantly different among the four clinical types except the ratio of WBC1 to WBC3 level. WBC1/WBC3 ratios in the medium, severe, and gravis patients were higher than those in the mild patients (Table 2). Spearman correlation analysis further confirmed that WBC1/WBC3 ratios in the medium, severe, and gravis patients were higher than those in the mild patients (Table 2). Spearman correlation analysis further confirmed that WBC1/WBC3 ratios in the medium, severe, and gravis patients were higher than those in the mild patients (Table 2).

Spearman correlation analysis further confirmed that WBC1/WBC3 ratios in the medium, severe, and gravis patients were higher than those in the mild patients (Table 2).
negatively correlated with the severity of HFRS (all \( p < 0.05 \); Supplementary Table 1).

### 3.4. Factors Associated with the Mortality of Patients with Gravis HFRS

Since most of the deaths occurred in the patients with gravis HFRS among the four clinical types (16/19, 84.21%), factors associated with mortality of patients with gravis HFRS were analyzed. Many variables including age, systolic (SBP) and diastolic blood pressure (DBP), and incidences of HFRS-related complications were significantly associated with deaths of HFRS patients (Table 3). Age was significantly higher in the nonsurvivors than that in the survivors. In contrast, lower SBP and DBP was common in those who died compared to those who survived. Among HFRS-related complications, the incidences of secondary infection, sepsis, and MODS in the nonsurvivors were significantly higher than those of the survivors. No significant differences in the frequencies of hemorrhage, hepatic injury, and arrhythmia were identified (Supplementary Table 2). Shorter hospital stays were found in the nonsurvivors compared to survivors due to the death of gravis patients during the disease course.

| Parameters       | Mild (\( n = 58 \)) | Medium (\( n = 75 \)) | Severe (\( n = 44 \)) | Gravis (\( n = 83 \)) | \( p \) value |
|------------------|---------------------|-----------------------|-----------------------|-----------------------|--------------|
| WBC1, \( \times 10^9 \) cells/L | 7.35 (3.97) | 9.2 (7.73) | 10.1 (6.16) | 13.07 (12.18) | <0.001 |
| PLT1, \( \times 10^9 \) cells/L | 100 (96.5) | 61 (68) | 51 (48.50) | 41 (47) | <0.001 |
| PDW1 (fL) | 16.0 (3.50) | 16.6 (6.10) | 17.8 (4.90) | 17.15 (4.45) | 0.003 |
| MPV1 (fL) | 11.88 ± 1.49 | 12.47 ± 1.30 | 12.77 ± 1.15 | 12.46 ± 1.52 | 0.017 |
| P-LCR1 (%) | 39.7 (16.25) | 44.10 (13.40) | 45.9 (6.10) | 43.2 (11.35) | 0.020 |
| PCT1 (%) | 0.12 (0.10) | 0.09 (0.12) | 0.07 (0.07) | 0.06 (0.07) | <0.001 |
| WBC3, \( \times 10^9 \) cells/L | 7.16 (5.22) | 9.49 (4.99) | 8.41 (4.79) | 10.27 (7.76) | <0.001 |
| PLT3, \( \times 10^9 \) cells/L | 115.5 (159.25) | 125 (145.25) | 77 (112) | 60 (67) | <0.001 |
| PDW3 (fL) | 16.2 (2.80) | 16.2 (5.02) | 16.3 (2.35) | 16.8 (3.65) | 0.391 |
| MPV3 (fL) | 12.1 (2.72) | 12.4 (2.05) | 12.3 (1.40) | 12.4 (1.63) | 0.213 |
| P-LCR3 (%) | 41 (20) | 44.2 (15.67) | 42.8 (8) | 42.45 (11.5) | 0.482 |
| PCT3 (%) | 0.17 (0.12) | 0.16 (0.16) | 0.12 (0.13) | 0.09 (0.09) | <0.001 |
| WBC1/WBC3 | 1 (0.6) | 1.06 (0.53) | 1.20 (0.63) | 1.16 (0.69) | 0.005 |
| PLT1/PLT3 | 0.78 (0.43) | 0.64 (0.54) | 0.55 (0.41) | 0.70 (0.68) | 0.082 |
| PDW1/PLW3 | 1 (0.25) | 1.01 (0.26) | 1.14 (0.27) | 1.01 (0.33) | 0.083 |
| MPV1/MPV3 | 1 (0.11) | 1.01 (0.11) | 1.03 (0.13) | 1.02 (0.17) | 0.717 |
| P-LCR1/P-LCR3 | 1 (0.21) | 1.03 (0.24) | 1.05 (0.26) | 1.03 (0.31) | 0.540 |
| PCT1/PCT3 | 0.78 (0.40) | 0.71 (0.53) | 0.64 (0.53) | 0.71 (0.74) | 0.187 |

WBC: white blood cell; PLT: platelet; PDW: platelet distribution width; MPV: mean platelet volume; P-LCR: platelet-large cell rate; PCT: plateletcrit.

| Variables | Univariable logistic regression | Multivariable logistic regression |
|-----------|--------------------------------|---------------------------------|
|           | OR (95% CI)                     | OR (95% CI)                     |
|           | \( p \) value                   | \( p \) value                   |
| Age       | 1.060 (1.009, 1.114)            | 1.091 (1.015, 1.172)            |
|           | 0.019                           | 0.018                           |
| SBP       | 0.981 (0.948, 1.015)            | 0.981 (0.948, 1.015)            |
|           | 0.266                           | 0.264                           |
| DBP       | 0.970 (0.935, 1.006)            | 0.970 (0.935, 1.006)            |
|           | 0.097                           | 0.097                           |
| Secondary | 6.400 (1.349, 30.372)           | 6.400 (1.349, 30.372)           |
| infection | 0.019                           | 0.019                           |
| MODS      | 12.250 (2.981, 50.333)          | 12.250 (2.981, 50.333)          |
|           | 0.001                           | 0.001                           |
| Sepsis    | 14.773 (2.542, 85.859)          | 14.773 (2.542, 85.859)          |
|           | 0.003                           | 0.003                           |
| PLT1      | 0.997 (0.988, 1.005)            | 0.997 (0.988, 1.005)            |
|           | 0.431                           | 0.431                           |
| PLT3      | 0.995 (0.984, 1.005)            | 0.995 (0.984, 1.005)            |
|           | 0.297                           | 0.297                           |
| PDW1      | 0.818 (0.679, 0.985)            | 0.818 (0.679, 0.985)            |
|           | 0.034                           | 0.034                           |
| PDW3      | 1.086 (0.863, 1.368)            | 1.086 (0.863, 1.368)            |
|           | 0.483                           | 0.483                           |
| MPV1      | 0.658 (0.435, 0.995)            | 0.658 (0.435, 0.995)            |
|           | 0.047                           | 0.047                           |
| MPV3      | 0.987 (0.592, 1.647)            | 0.987 (0.592, 1.647)            |
|           | 0.962                           | 0.962                           |
| PDW1/PDW3 | 0.05 (0.003, 0.899)             | 0.05 (0.003, 0.899)             |
|           | 0.042                           | 0.042                           |

OR: odds ratio; CI: confidence interval; SBP: systolic blood pressure; DBP: diastolic blood pressure; MODS: multiple organ dysfunction syndrome; PLT: platelet; PDW: platelet distribution width; MPV: mean platelet volume.
1.091; 95% CI, 1.015

Multiple logistic regression analysis showed that age (OR, nearity (all tolerance values > $p$ nonsurvivor group (% signifi

22.283; 95% CI, 2.985 – and a speci

sepsis on the prognosis of HFRS, ROC analysis was performed. PDW1 was the platelet indi-

mention, and MODS (Table 3).

risk factors for the mortality in HFRS patients after adjust-

Multicollinearity test indicated no significant multicollinearity (all tolerance values > 2.8 and all VIF values < 3.5). Multiple logistic regression analysis showed that age (OR, 1.091; 95% CI, 1.015–1.172; $p = 0.018$), PDW levels at the first day of hospital admission (OR, 0.782; 95% CI, 0.617–0.992; $p = 0.043$), and sepsis secondary to HFRS (OR, 22.283; 95% CI, 2.985–166.325; $p = 0.002$) were independent risk factors for the mortality in HFRS patients after adjust-

To explore the predictive value of age, PDW1 levels, and the incidences of sepsis on the prognosis of HFRS, ROC analysis was performed. PDW1 was the platelet indices showing a high AUC (0.709), with a sensitivity of 70% and a specificity of 67% at a cutoff of 16.5 fl. All pertinent data are shown in Table 4. ROC analysis demonstrated that the specificity, sensitivity, and AUC of the three factors in combination were preferable to any single factor alone in predicting prognosis ($p < 0.05$; Table 4, Figure 1).

4. Discussion

The current study was performed to investigate the predictive role of the routine blood parameters of WBC and PLT in the setting of HFRS. We analyzed the demographic, epidemiological, clinical and laboratory characteristics of 260 patients who were hospitalized due to HFRS. The main findings are that WBC and PLT counts in the first and third day of hospital admission, WBC1/WBC3, PDW1, and PCT3, were corre-

Platelets, small anucleate cell fragments of the mega-
karyocytes, play a key role in regulating haemostasis and protecting vascular integrity [8, 20]. Platelet indices includ-
ing PLT, PDW, MPV, P-LCR, and PCT are standard indi-
cators of platelet function. PDW is an indicator of the heterogeneity in platelet size, which can be a sign of active platelet release. MPV represents the average size of plate-

Figure 1: Receiver operating characteristic curve (ROC) analysis of platelet distribution width at first day of hospital admission (PDW1), age, and sepsis and the combination of PDW1, age, and sepsis to predict prognosis in critical HFRS patients.
tend to require more frequent blood transfusions and CRRT and suffer more fatal complications such as hemorrhage, hepatic injury, kidney rupture, sepsis, and MODS. Furthermore, the nonsurvivors had higher incidences of secondary infection, sepsis, and MODS.

So far, the pathogenesis of HFRS is not fully known. The virus itself, immune response, and interaction between the host immune system and hantaviruses may contribute to disease initiation and development. Vascular endothelial dysfunction is the basic pathological change, which is characterized by a dramatic increase in vascular permeability [25]. Currently, there is no specific therapy for HFRS. Early diagnosis and prognosis is important for the management of the patients. Therefore, early identification of patients with HFRS at risk of developing severe and gravis types would potentially improve the management via prompt initiation of systematic supportive treatment.

Platelet parameters have been suggested as valuable novel biomarkers. Activation of the coagulation system [26, 27], severe infection [13–15, 21, 25], inflammatory diseases [18], trauma [28], cardiovascular diseases [19], and thrombotic diseases [29] could all result in changes in platelet parameters. Previous study showed that PLT counts were negatively correlated with the progression of HFRS, and the AUC values were 0.814 for the severity of HFRS, which indicated the better predictive efficacy of PLT counts [11]. However, the patients in that study [11] were just divided into mild group and severe group, and no analysis of the predictive value of platelet parameters was conducted in HFRS patients in relation to the patients’ survival.

It is generally recognized that when PLTs have been excessively destructed and consumed, bone marrow will produce a mass of immature PLTs which have larger volume than mature ones. At that time, PDW, MPV, and P-LCR will be increased correspondingly, because immature platelets with large volume and mature platelets with small volume synchronously were present in the blood [30]. Because thrombocytopenia is a significant feature of HFRS, the analysis of all the platelet parameters, instead of only PLT count, may provide a more accurate and comprehensive information for the disease severity and patient prognosis.

We analyzed whether WBC count, PLT count, PDW, MPV, P-LCR, or PCT could predict disease severity and mortality in HFRS. Firstly, levels of WBC and platelet parameters were investigated and analyzed in relation to clinical types of HFRS. Consistent with previous studies [11, 31], WBC count on day 1 of hospital admission (WBC1) was found to increase gradually with the exacerbation of the disease severity. Patients with mild type HFRS had higher PLT1 value than those with severe and gravis types. Of the 4 platelet parameters routinely tested, levels of MPV1 and PDW1 in the severe and gravis patients were found to be higher than those in the mild and medium patients. PCT1 was consistent with the variation trend of PLT1. Secondly, this study examined factors associated with patient survival in patients with gravis HFRS. PDW1 was found to be a significant predictor of mortality of patients with gravis type HFRS.

In contrast to the increasing trend of PDW1 with disease severity among the four clinical types of HFRS, PDW1 was decreased in nonsurvivors compared with survivors. This might suggest that the depletive reduction of platelets cannot be compensated by the hemopoietic function due to this critical disease condition. However, the possible mechanism for the decrease of PDW1 in nonsurvivors of patients with gravis type HFRS is not very clear and needs further investigations.

It is not difficult to understand that age and the occurrences of sepsis were also found to be independent risk factors of death in gravis HFRS patients.

Studies on risk scores in patients with other diseases showed that laboratory parameters at day 3 and the ratio of the first-day value to the third-day value had better predictive performance than baseline data [13, 32, 33]. One could hypothesize that the variation of platelet parameters within the first 3 days after hospital admission could be a strong predictor of clinical deterioration and mortality. Therefore, we compared the prediction accuracy of WBC and platelet parameters at day 3 and ratios of WBC and platelet parameters at day 1 to WBC and platelet parameters at day 3, respectively. Although WBC3, PLT3, PCT3, and WBC1/WBC3 were found to be correlated with the severity of HFRS, these parameters were not superior to parameters at day 1 in the prediction accuracy. Therefore, waiting 3 or more days seems to confer no benefit to improving the prediction performance of platelet parameters.

Our study has a few limitations. First, the clinical classification criteria of HFRS contain substantial subjective factors based on symptoms and signs, which may influence the judgment of severity. Second, the study is limited by the relatively small number of critical patients, especially for the binary logistic regression analysis used for identifying risk factors for prognosis. Third, the study is a retrospective design and has no validation analysis in other patient populations. Fourth, this study shows that PDW1 may only moderately predict mortality and thus caution should be exercised when using this marker. Prospective studies with a larger sample of HFRS patients are needed to verify the predictive value of these parameters in the severity and mortality of HFRS patients.

5. Conclusion

The current study showed that WBC, PLT, PDW, and PCT may be used as valuable parameters for the severity of HFRS patients. Furthermore, in gravis HFRS patients, reduced PDW1, age, and the occurrences of sepsis were risk factors for prognosis and the combination of these factors had better performance than any single parameter alone in predicting the mortality of gravis HFRS patients. Platelet distribution width at the first day of hospital admission (PDW1) has the potential as a valuable parameter for the disease severity of HFRS and a moderate parameter for the prognosis of critical HFRS patients in that its alteration is a significant factor associated with the disease severity of HFRS and the survival of gravis HFRS patients. The early determination of PDW and the dynamic and close monitoring of sepsis occurrence may help clinicians to identify high-risk HFRS patients and to stratify risk for optimal management.
Data Availability

The data underlying the findings of this study are all included in this article and its supplementary materials.

Conflicts of Interest

The authors declare that they have no conflict of interest related to this paper or the study it describes.

Supplementary Materials

Supplementary 1. Table 1: correlation of laboratory data and the severity of HFRS.

Supplementary 2. Table 2: demographic and clinical characteristics of survivors and nonsurvivors in patients with gravis HFRS.

Supplementary 3. Table 3: comparison of white blood cell characteristics of survivors and nonsurvivors in patients with gravis HFRS.

Supplementary 4. Figure 1: flowchart of the patients’ selection process in the study.

References

[1] M. Ermonval, F. Baychelier, and N. Tordo, "What do we know about how hantaviruses interact with their different hosts?," *Viruses*, vol. 8, no. 8, 2016.

[2] T. Avsic Zupanc, M. Korva, and A. Markotic, "HFRS and hantaviruses in the Balkans/South-East Europe," *Virus Research*, vol. 187, pp. 27–33, 2014.

[3] H. Jiang, X. Zheng, L. Wang, H. Du, P. Wang, and X. Bai, "Hantavirus infection: a global zoonotic challenge," *Virologica Sinica*, vol. 32, no. 1, pp. 32–43, 2017.

[4] M. H. Miettinen, S. M. Mäkelä, I. O. Ala-Houhala et al., "Ten-year prognosis of Puumala hantavirus-induced acute interstitial nephritis," *Kidney International*, vol. 69, no. 11, pp. 2043–2048, 2006.

[5] M. Liang, D. Li, S.-Y. Xiao, C. Hang, C. A. Rossi, and C. S. Schmaljohn, "Antigenic and molecular characterization of hantavirus isolates from China," *Virus Research*, vol. 31, no. 2, pp. 219–233, 1994.

[6] D. H. Krüger, G. Schönrich, and B. Klempa, "Human pathogenic hantaviruses and prevention of infection," *Human Vaccines*, vol. 7, no. 6, pp. 685–693, 2011.

[7] L. X. Zou, M. J. Chen, and L. Sun, "Haemorrhagic fever with renal syndrome: literature review and distribution analysis in China," *International Journal of Infectious Diseases*, vol. 43, pp. 95–100, 2016.

[8] Z. Liu, Q. Zhao, Q. Han, M. Gao, and N. Zhang, "Serum thrombopoiesis-1 is altered in patients with hemorrhagic fever with renal syndrome," *Journal of Medical Virology*, vol. 80, no. 10, pp. 1799–1803, 2008.

[9] K. Laki, "Our ancient heritage in blood clotting and some of its consequences," *Annals of the New York Academy of Sciences*, vol. 202, no. 1, pp. 297–307, 1972.

[10] S. Skarpheðinsson, H. C. Thiesson, S. A. Shakar, and M. Tepel, "Factors predicting kidney damage in Puumala virus infected patients in Southern Denmark," *European Journal of Clinical Microbiology & Infectious Diseases*, vol. 34, no. 10, pp. 2017–2021, 2015.

[11] Z. Yu, N. Zhou, A. Li et al., "Performance assessment of the SAPS II and SOFA scoring systems in hanta virus hemorrhagic fever with renal syndrome," *International Journal of Infectious Diseases*, vol. 63, pp. 88–94, 2017.

[12] S. Zhang, Y. L. Cui, M. Y. Diao, D. C. Chen, and Z. F. Lin, "Use of platelet indices for determining illness severity and predicting prognosis in critically ill patients," *Chinese Medical Journal*, vol. 128, no. 15, pp. 2012–2018, 2015.

[13] H. Yilmaz, G. Yilmaz, A. Menteşe, U. Kostakoğlu, S. C. Karahan, and İ. Köksal, "Prognostic impact of platelet distribution width in patients with Crimean-Congo hemorrhagic fever," *Journal of Medical Virology*, vol. 88, no. 11, pp. 1862–1866, 2016.

[14] Y. Gao, Y. Li, X. Yu et al., "The impact of various platelet indices as prognostic markers of septic shock," *PloS One*, vol. 9, no. 8, article e103761, 2014.

[15] M. Kostrubiec, A. Labyk, J. Pedowska-Wloszek et al., "Mean platelet volume predicts early death in acute pulmonary embolism," *Heart*, vol. 96, no. 6, pp. 460–465, 2010.

[16] A. Abdel-Razik, W. Eldars, and E. Rizk, "Platelet indices and inflammatory markers as diagnostic predictors for ascitic fluid infection," *European Journal of Gastroenterology & Hepatology*, vol. 26, no. 12, pp. 1342–1347, 2014.

[17] B. M. Sanner, M. Konermann, M. Tepel, J. Groetz, C. Mummenhoff, and W. Zidek, "Platelet function in patients with obstructive sleep apnoea syndrome," *The European Respiratory Journal*, vol. 16, no. 4, pp. 648–652, 2000.

[18] J. Tang, X. Gao, M. Zhi et al., "Plateletcrit: a sensitive biomarker for evaluating disease activity in Crohn’s disease with low hs-CRP," *Journal of Digestive Diseases*, vol. 16, no. 3, pp. 118–124, 2015.

[19] M. H. Seyyed-Mohammadzad, R. Eskandari, Y. Rezaei et al., "Prognostic value of mean platelet volume in patients undergoing elective percutaneous coronary intervention," *Anatolu Kardiyojiji Dergisi/The Anatolian Journal of Cardiology*, vol. 15, no. 1, pp. 25–30, 2015.

[20] K. R. Machlus, J. N. Thon, and J. E. Italiano Jr, "Interpreting the developmental dance of the megakaryocyte: a review of the cellular and molecular processes mediating platelet formation," *British Journal of Haematology*, vol. 165, no. 2, pp. 227–236, 2014.

[21] Z. Fan, J. Pan, Y. Zhang et al., "Mean platelet volume and platelet distribution width as markers in the diagnosis of acute gangrenous appendicitis," *Disease Markers*, vol. 2015, Article ID 542013, 4 pages, 2015.

[22] I. Akpinar, M. R. Sayin, Y. C. Gursoy et al., "Plateletcrit. A platelet marker associated with saphenous vein graft disease," *Herz*, vol. 39, no. 1, pp. 142–148, 2014.

[23] Z. M. Golwala, H. Shah, N. Gupta, V. Sreenivas, and J. M. Puliyel, "Mean platelet volume (MPV), platelet distribution width (PDW), platelet count and plateletcrit (PCT) as predictors of in-hospital paediatric mortality: a case-control study," *African Health Sciences*, vol. 16, no. 2, pp. 356–362, 2016.

[24] H. Jiang, H. Du, L. M. Wang, P. Z. Wang, and X. F. Bai, "Corrigendum: hemorrhagic fever with renal syndrome: pathogenesis and clinical picture," *Frontiers in Cellular and Infection Microbiology*, vol. 6, p. 178, 2016.

[25] I. N. Gavrilovskaya, E. E. Gorbunova, N. A. Mackow, and E. R. Mackow, "Hantaviruses direct endothelial cell permeability by"
sensitizing cells to the vascular permeability factor VEGF, while angiopeptin 1 and sphingosine 1-phosphate inhibit hantavirus-directed permeability,” *Journal of Virology*, vol. 82, no. 12, pp. 5797–5806, 2008.

[26] W. W. Duke, “The relation of blood platelets to hemorrhagic disease,” *JAMA*, vol. 250, no. 9, pp. 1201–1209, 1983.

[27] M. Levi and E. C. Löwenberg, “Thrombocytopenia in critically ill patients,” *Seminars in Thrombosis and Hemostasis*, vol. 34, no. 5, pp. 417–424, 2008.

[28] R. C. Jacoby, J. T. Owings, J. Holmes, F. D. Battistella, R. C. Gosselin, and T. G. Paglieroni, “Platelet activation and function after trauma,” *The Journal of Trauma*, vol. 51, no. 4, pp. 639–647, 2001.

[29] U. Sevuk, R. Altindag, M. V. Bahadir, N. Ay, E. Demirtas, and F. Ayaz, “Value of platelet indices in identifying complete resolution of thrombus in deep venous thrombosis patients,” *Indian Journal of Hematology and Blood Transfusion*, vol. 31, no. 1, pp. 71–76, 2015.

[30] J. L. Vincent, A. M. Habib, C. Verdant, and A. Bruhn, “Sepsis diagnosis and management: work in progress,” *Minerva Anestesiologica*, vol. 72, no. 3, pp. 87–96, 2006.

[31] H. Du, J. Li, H.-T. Yu et al., “Early indicators of severity and construction of a risk model for prognosis based upon laboratory parameters in patients with hemorrhagic fever with renal syndrome,” *Clinical Chemistry and Laboratory Medicine (CCLM)*, vol. 52, no. 11, pp. 1667–1675, 2014.

[32] R. Jalan, M. Pavesi, F. Saliba et al., “The CLIF consortium acute decompensation score (CLIF-C ADs) for prognosis of hospitalised cirrhotic patients without acute-on-chronic liver failure,” *Journal of Hepatology*, vol. 62, no. 4, pp. 831–840, 2015.

[33] R. Jalan, F. Saliba, M. Pavesi et al., “Development and validation of a prognostic score to predict mortality in patients with acute-on-chronic liver failure,” *Journal of Hepatology*, vol. 61, no. 5, pp. 1038–1047, 2014.