Ventilator-associated pneumonia caused by multidrug-resistant Gram-negative bacteria: understanding nebulization of aminoglycosides and colistin

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The use of nebulized antibiotics for treating ventilator-associated pneumonia (VAP) caused by multidrug-resistant (MDR) Gram-negative bacteria (GNB) increases worldwide. There is a paradox, however, between the large body of experimental evidence supporting the administration of nebulized rather than intravenous aminoglycosides and colistin to treat inoculation pneumonia caused by GNB [1, 2], and the paucity of clinical studies confirming such a benefit in VAP. Based on the recommendations of the European Society of Clinical Microbiology and Infectious Diseases (ESCMID) [3, 4], the present article examines this apparent contradiction and suggests some directions for further research and clinical practice.

Why and when to administer nebulized aminoglycosides and colistin in VAP

The main reason for nebulizing aminoglycosides and colistin in VAP is to bypass the alveolar–capillary barrier which offers a severe obstacle to lung penetration following intravenous administration. In comparison to intravenous route, nebulized aminoglycosides and colistin can achieve significantly higher lung tissue concentrations necessary for the effective treatment of VAP due to MDR GNB [1, 2]. For colistin, this is achieved with minimal systemic toxicity compared to intravenous administration. Demonstration of high lung tissue deposition following nebulization is difficult in humans as epithelial lining fluid concentrations may be in part falsely elevated because of a heavy contamination of the bronchoscope during bronchoalveolar lavage (Fig. 1e–i) [5]. Evidence of high lung tissue concentrations relies on microdialysis [2] or open lung biopsies [1] that can be performed exclusively in experimental studies.

In healthy sheep, high and homogeneously distributed tobramycin pulmonary interstitial concentrations are observed 30 min after nebulization followed by a bi-compartmental decrease, and contrasting with low concentrations after intravenous administration [2]. In pneumonia, high antibiotic tissue concentrations are also observed but are heterogeneously distributed, and likely influenced by the aeration loss [6, 7]. Peak tissue concentrations remain high in non-aerated lung regions, indicating that aminoglycosides and colistin likely diffuse through bronchiolar mucosa towards adjacent consolidated infected alveoli. Both are concentration-dependent antibiotics with peak interstitial concentrations determining bactericidality. Systemic diffusion of nebulized aminoglycosides is substantial, increasing when the alveolar–capillary membrane is injured by a microorganism [1]. The peak plasma concentration is observed 1 h following nebulisation, with a subsequent bi-compartmental time-dependent decrease. Trough plasma concentrations, which determine the toxicity risk, are similar to those resulting from the intravenous administration when the nebulized dose is equal to the intravenous dose.
plus extrapulmonary deposition (Fig. 1k). In contrast, systemic diffusion of nebulized colistin is weak even in the presence of extensive VAP (Fig. 1j), protecting against nephrotoxicity [1, 7, 8] and offering the possibility of delivering very high doses by nebulization. Measured colistin plasma concentrations result from the hydrolysis of the nebulized prodrug colistin methanesulfonate [9].

According to available PK data, a benefit of nebulization on cure rate and microbiological eradication can be expected with two classes of antibiotics: aminoglycosides and polymyxins, predominantly used in VAP caused by MDR GNB.

**Inhaled substitution rather than adjunctive aminoglycosides or colistin for VAP caused by MDR GNB**

Despite the experimental evidence supporting nebulized antibiotics to treat pneumonia, clinical studies have not shown any mortality benefit when used as adjuvant therapy (nebulized plus intravenous colistin or nebulized aminoglycosides plus intravenous betalactams). However, in VAP caused by MDR GNB, a higher clinical resolution rate was observed with adjuvant therapy [10]. A decrease in the emergence of MDR bacteria was also reported in randomized controlled trials without effect on ventilator-associated pneumonia relapse [11–13].

The ESCMID position paper [4] recommended avoiding the routine use of nebulized antibiotics in VAP, due to a questionable efficacy and the potential for underestimated risks of adverse respiratory events. The panel identified an urgent need for randomized clinical trials of nebulized antibiotic therapy as part of a substitution approach to VAP therapy caused by MDR pathogens. In 2018, the French Society of Anaesthesia and Intensive Care Medicine (FSAICM) and the French Intensive Care Society (FICS) published guidelines regarding hospital-acquired pneumonia (HAP) in the intensive care unit [11], and recommended nebulized colistin and/or aminoglycosides alone in HAP due to MDR GNB susceptible to colistin and/or aminoglycosides, when no other antibiotics can be used.

When considering experimental pharmacokinetic data, the rationale for adjunctive nebulized therapy appears limited [4, 11]. The combination of nebulized and intravenous aminoglycosides is likely to increase the risk of toxicity. When added to intravenous betalactams, nebulized aminoglycosides do not improve therapeutic efficacy in VAP caused by susceptible GNB, likely because double-antimicrobial therapy is not superior to monotherapy. The addition of nebulized to intravenous colistin increases lung tissue concentrations but not plasma concentrations. Thus, it improves efficacy without increasing systemic toxicity. Compared to adjunctive therapy, substitution therapy markedly reduces colistin plasma concentrations and decreases the risk of toxicity as shown in a recent meta-analysis [10]. This is the reason why the ESCMID position paper recommended to perform future randomized control studies comparing substitution therapy (rather than adjunctive therapy), to intravenous administration [4]. The FSAICM and FICS also recommended the use of substitution rather than adjunctive therapy in VAP caused by MDR GNB susceptible to colistin and/or aminoglycosides [11].

**Optimisation of nebulization to maximize antibiotic lung deposition**

Limiting inspiratory flow velocity is required to reduce inertial impaction in the airways and optimize lung deposition [1, 3]. Volume-controlled mode should be preferred to pressure support ventilation [14]. As shown in Fig. 1, it is recommended to select specific ventilator settings during the nebulization, to use specifically designed
Nebulized amikacin and colistin for ventilator-associated pneumonia caused by MDR Gram bacteria

Priority to mesh nebulizers and specifically designed circuits

Mass median aerodynamic diameter → 2 - 5 μ
High lung deposition 20-30% Easy to handle for nurses
Chamber deposition < 5%

Initial dose inserted into the nebulizer’s chamber

Colistimethate 4 m IU diluted in 6 ml x3/24h
Mesh nebulizers → chamber residual volume < 10%
Jet nebulizers → chamber residual volume > 40%

Amikacin 40 mg/kg/24h diluted in 6 ml
Circuits deposit (inspiratory tubing + Y piece + endotracheal tube) around 30%

Nebulization time ≤ 30 min - Specific ventilator settings to limit inspiratory inertial impaction

Volume control ventilation with constant inspiratory flow, no patient’s triggering, no asynchrony between the patient and the ventilator (propofol if necessary)

TV 8 ml/kg, RF 12-15 bpm, I/E 1:2, end-inspiratory pause 20%, PEEP 5-10 cmH₂O

Removal of heat and moisture exchangers, stop of heat humidifier and addition of a filter on the expiratory limb

Return to previous settings after nebulization
Change the expiratory filter

Nebulised dose entering the respiratory system

Tracheobronchial deposition
Contamination of the bronchoscopes during the BAL

ELF is contaminated during the BAL

Lung dose reaching bronchioles and alveoli

Exhaled dose tracheal suctioning

Systemic absorption

Routine dosage of trough plasma levels to detect systemic toxicity

Colistin in presence of lung infection, low systemic diffusion

Amikacin in presence of lung infection, high systemic diffusion

Intravenous AMK → n = 8 pigtets with E. coli inoculation pneumonia
Nebulised AMK → n = 10 pigtets with E. coli inoculation pneumonia

Urinary excretion
tubings, and mesh nebulizers positioned 10–15 cm before the Y piece on the inspiratory limb [3]. Heat and moisture exchanger and heated humidifiers should be removed during the nebulization to avoid hygroscopic growth of the aerosolized particles and a rainout effect in the circuits. Written operating procedures should be implemented to ensure that previous ventilator settings and humidification are resumed at the end of nebulization. The benefit on aerosol delivery far outweighs the additional workload for health care providers.

There is evidence supporting the use of mesh (Fig. 1a and b) rather than jet nebulizers for nebulized antibiotic delivery [1, 3, 11, 15]. In vitro, mesh nebulizers appear superior over jet nebulizers to deliver tobramycin [15]. Aerosol particle size is slightly smaller with jet nebulizers compared to mesh nebulizers, but always remained below five microns, a condition required to reach the distal lung. Lung dose is significantly higher with mesh nebulizers whereas nebulization time and residual volume are significantly reduced. These in vitro benefits were confirmed in animals and in patients treated by salbutamol [16–18]. In conclusion, substitution therapy should be preferred to adjunctive therapy to evidence the therapeutic benefit of nebulized aminoglycosides and colistin in VAP caused by MDR GNB. In addition, mesh nebulizers should be preferred to jet nebulizers to optimize the lung deposition of aerosolized antibiotics.

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Author contributions
JR wrote the first draft of the manuscript. JR and CS‑L contributed towards the critical revision of the manuscript for important intellectual content and confirm the integrity of the work. Each member of the European Investigators Network for Nebulized Antibiotics in Ventilator-associated Pneumonia approved the content of the manuscript and contributed to its revision.

Compliance with ethical standards
Conflicts of interest
JR received grant support from BAYER and served in the advisory board for BAYER and speakers bureau for Norma Helas. OM served as a consultant for SANDOFI. SE declares having received consultancies from Aerogen Ltd, La Diffusion Technique Française and Bayer Healthcare, research support from Aerogen Ltd, Fisher & Paykel healthcare, Hamilton medical, travel reimbursements from Aerogen Ltd and Fisher & Paykel. JD and PFL received an unrestricted grant from Aerogen Ltd for their study [14]. JMQ currently received grant from Sino-European Cooperative Clinical Research Project “Evaluation of clinical efficacy and safety of QBW251 in patients with severe bronchiectasis”. MBB has participated in advisory boards and/or received speaker honoraria from Achaogen, Angelini, Astellas, Bayer, Basilea, Biomerieux, Cidara, Gilead, Menarini, MSD, Nabria, Paratek, Pfizer, Roche, Melinta, Shionogi, Tetraphase, VenatoRx and Vifor and has received study grants from Angelini, Basilea,
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