In conclusion, this study yields first evidence from longitudinal data of individual patients for the potential of iron and ferritin as progression marker in PD. A validation of our findings in a larger cohort, more advanced PD patients and a longer follow-up period is warranted.

Acknowledgments: We gratefully appreciate the participation of our patients in this study. We thank our Parkinson’s and study nurses Gudrun Leyerer and Jennifer Heinemann for their excellent assistance. We also thank Peter Lange for providing helpful advice on assay-related issues.

Open Access funding enabled and organized by Projekt DEAL.

Data Availability Statement
The datasets analyzed during the current study are available from the corresponding author on reasonable request.

Fabian Maass, MD,* Bernhard Michalke, PhD,2 Desiree Willkommen, (Ms.),3 Szegi Canaslan, (Ms.),1 Matthias Schmitz, PhD,1,3 Mathias Bähr, MD,1,4 Inga Zerr, MD,1,3 and Paul Lingor, MD5,6
1Department of Neurology, University Medical Center Göttingen, Goettingen, Germany, 2Research Unit Analytical BioGeoChemistry, German Research Center for Environmental Health, Helmholtz Zentrum München, Neuherberg, Germany, 3DZNE, German Center for Neurodegenerative Diseases, Goettingen, Germany, 4Center for Biostructural Imaging of Neurodegeneration (BIN), University of Göttingen Medical Center, Göttingen, Germany, 5Department of Neurology, School of Medicine, Klinikum rechts der Isar, Technical University of Munich, München, Germany, and 6DZNE, German Center for Neurodegenerative Diseases, Munich, Germany

References
1. Ward RJ, Zucca FA, Duyn JH, Crichton RR, Zecca L. The role of iron in brain ageing and neurodegenerative disorders. Lancet Neurol 2014;13(10):1045–1060. https://doi.org/10.1016/S1474-4422(14)70117-6
2. Dexter DT, Wells FR, Lee AJ, et al. Increased nigral iron content and alterations in other metal ions occurring in brain in Parkinson’s disease. J Neurochem 1989;52(6):1830–1836. https://doi.org/10.1111/j.1471-4159.1989.tb07264.x
3. Acosta-Cabronero J, Cardenas-Blanco A, Betts MJ, et al. The whole-brain pattern of magnetic susceptibility perturbations in Parkinson’s disease. Brain 2016;140(2016):aww278. https://doi.org/10.1093/brain/aww278
4. Dexter DT, Carayon A, Javoy-agid F, et al. Alterations in the levels of iron, ferritin and other trace metals in parkinson’s disease and other neurodegenerative diseases affecting the basal ganglia. Brain 1991; 114:1953–1975. https://doi.org/10.1093/brain/114.4.1953
5. Zecca L, Stroppolo A, Gatti A, et al. The role of iron and molecules in the neuronal vulnerability of locus coeruleus and substantia nigra during aging. Proc Natl Acad Sci U S A 2004;101(26):9843–9848. https://doi.org/10.1073/pnas.0403495101
6. Cholanians AB, Phan AV, Ditzel EJ, Camenisch TD, Lau SS, Monks TJ. Arsenic induces accumulation of α-synuclein: implications for synucleinopathies and neurodegeneration. Toxicol Sci 2016;153(2):271–281. https://doi.org/10.1093/toxsci/kfw117
7. Sun H. Association of soil selenium, strontium, and magnesium concentrations with Parkinson’s disease mortality rates in the USA. Environ Geochim Health 2018;40(1):349–357. https://doi.org/10.1007/s10653-017-9915-8
8. Davies KM, Bohic S, Carmona A, et al. Copper pathology in vulnerable brain regions in Parkinson’s disease. Neurobiol Aging 2014;35(4):858–866. https://doi.org/10.1016/j.neurobiolaging.2013.09.034
9. Lucio M, Willkommen D, Schroeter M, Sigurðsson A, Schmitt-Kopplin P, Michalke B. Integrative metabolomic and metalomic analysis in a case-control cohort with Parkinson’s disease. Front Aging Neurosci 2019;11:331. https://doi.org/10.3389/fagi.2019.00331
10. Maass F, Michalke B, Willkommen D, et al. Elemental fingerprint: reassessment of a cerebrospinal fluid biomarker for Parkinson’s disease. Neurobiol Dis 2020;134:104677. https://doi.org/10.1016/j.nbd.2019.104677

Supporting Data
Additional Supporting Information may be found in the online version of this article at the publisher’s web-site.

Lower Circulating Lymphocyte Count Predicts ApoE ε4-Related Cognitive Decline in Parkinson’s Disease

Neuroinflammatory changes in the brain, including infiltration of lymphocytes, particularly T cells, play a critical role in the pathogenesis of Parkinson’s disease (PD).1,2 Interestingly, in the peripheral blood of PD patients, a decrease in circulating lymphocyte counts occurs, mainly due to a decrease in T cells.1,3 Furthermore, it has recently been reported that lower lymphocyte count might be causally related to the subsequent development of PD.4 Inspired by these observations, we aimed at assessing whether low lymphocyte count is associated with the subsequent development of the key milestones in PD’s disease course, specifically cognitive impairment, with a particular attention to the apolipoprotein E (ApoE) ε4 allele, a crucial modifying factor in cognitive impairment.5,6

In this retrospective cohort study, using the Parkinson’s Progression Markers Initiative data, 167 de novo PD patients were enrolled (Fig. S1) and were followed up for 2 years (Tables S1 and S2; Text S1). R scripts made for the analysis are freely available at http://dx.doi.org/10.17632/s8ng9yn8.2 or https://github.com/KazutoTsukita/Mov_Disord_2021.

© 2021 The Authors. Movement Disorders published by Wiley Periodicals LLC on behalf of International Parkinson and Movement Disorder Society
This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.
*Correspondence to: Dr Kazuto Tsukita, Department of Neurology, Graduate School of Medicine, Kyoto University, 54 Shogon-Kawaharacho, Kyoto 606-8507, Japan; E-mail: kazusan@kuhp.kyoto-u.ac.jp

Relevant conflicts of interest/financial disclosures: Nothing to report.
Funding agency: Nothing to report.
Received: 13 August 2021; Accepted: 26 August 2021
Published online 13 October 2021 in Wiley Online Library (wileyonlinelibrary.com). DOI: 10.1002/mds.28799
We primarily used the multivariate linear mixed-effects model adjusted for various covariates (age, sex, levodopa-equivalent dose, disease duration, and baseline severity of smell deficit and rapid-eye-movement sleep behavior). We observed that only in PD patients carrying APOE ε4 allele, baseline lymphocyte count had significant interaction effect on the longitudinal decline in the Montreal Cognitive Assessment (MoCA) total score, such that lower baseline lymphocyte count was associated with accelerated MoCA score decline (carrier, standardized fixed-effects coefficient of the interaction term $\beta_{interaction} = 0.17$ [95% confidence interval, CI: 0.04, 0.30], $P < 0.01$; noncarrier, $\beta_{interaction} = 0.00$ [95% CI: $-0.10$, 0.09], $P < 0.94$). When PD patients, with and without APOE ε4 allele, were dichotomized using the median of baseline lymphocyte count (carrier, $1.72 \times 10^7/\mu L$; noncarrier, $1.74 \times 10^7/\mu L$) (Table S3), the interaction effect was apparent only in PD patients carrying APOE ε4 allele (carrier, $\beta_{interaction} = 0.45$ [95% CI: 0.20, 0.71], $P < 0.001$; noncarrier, $\beta_{interaction} = -0.03$ [95% CI: $-0.22$, 0.15], $P = 0.72$) (Fig. 1A,B). The interaction effects of baseline lymphocyte count on the progression of specific domains of cognitive impairment did not reach statistical significance (Fig. 1C). Sensitivity analyses confirmed the robustness of our result in a range of follow-up periods (Table S4) and even when missing values were imputed (Table S5).

An interesting aspect of the present result is that baseline lymphocyte count was clearly associated with subsequent cognitive decline only in PD patients carrying APOE ε4 allele. Given the importance of APOE ε4 allele and blood–brain barrier (BBB) dysfunction and the role of circulating T cells in PD pathogenesis (Text S2), our result might indicate the cooperative pathological role of BBB dysfunction and circulating lymphocytes in PD. Alternatively, the brain cortex of patients carrying APOE ε4 allele may be particularly vulnerable to lymphocyte infiltration. Admittedly, this study has some limitations (Text S3); however, because many covariates were adjusted for, we believe that our result indicates that biological phenomenon reflected by the decrease in the lymphocyte count might actively exacerbate the pathology driving cognitive dysfunction in synergy with the APOE ε4 allele, thereby providing important clinical and pathophysiological implications.

Acknowledgments: This work was supported by JST [Moonshot R&D] [Grant Number JPMJMS2024]. PPMI—a public–private partnership—is funded by the Michael J. Fox Foundation for Parkinson’s Research funding partners—4D Pharma, AbbVie, Acurex Therapeutics, Allergan, Amathus Therapeutics, ASAP, Avid Radiopharmaceuticals, Biontech, Biogen, BioLegend, Bristol-Myers Squibb, Calico, Celgene, Dacapo Brain Science, Denali, the Edmond J. Safra Foundation, GE Healthcare, Genentech, GlaxoSmithKline, Golub Capital, Handl Therapeutics, Insiro, Jansen Neuroscience, Lilly, Lundbeck, Merck, Mesobole Discovery, Neurocine Biosciences, Pfizer, Piramal, Preval, Roche, Sanofi Genzyme, Servier, Takeda, Teva, UCB, Verily, and Voyager Therapeutics. We thank Dr. Takahiro Kamada for inspiring us to do this study. He died in January 2019, and we wish to dedicate this article in his memory.

Data Availability Statement
Data used in this retrospective cohort study were obtained from the Parkinson’s Progression Markers Initiative (PPMI) database (www.ppmi-info.org/data) on July 28, 2021. For up-to-date information on the study, visit www.ppmi-info.org. R scripts made for the analysis are freely available at http://dx.doi.org/10.17632/7s8m9yn82. or https://github.com/KazutoTsukita/Mov_Distord_2021.

Kazuto Tsukita, MD,1,2,3* Haruhi Sakamaki-Tsukita, MD,1 and Ryosuke Takahashi, MD, PhD1
1Department of Neurology, Graduate School of Medicine, Kyoto University, Kyoto, Japan, 2Advanced Comprehensive Research Organization, Teikyo University, Tokyo, Japan, and 3Division of Sleep Medicine, Kansai Electric Power Medical Research Institute, Osaka, Japan

References
1. Tan E-K, Chao Y-X, West A, Chan I-L, Poewe W, Jankovic J. Parkinson disease and the immune system - associations, mechanisms and therapeutics. Nat Rev Neurol 2020;16:303–318.
Screening of GBA Mutations in Nigerian Patients with Parkinson’s Disease

Heterozygous mutations in the β-glucocerebrosidase (GBA) gene are reported in 5% to 30% of patients with Parkinson’s disease (PD) across White and Asian populations with a relative absence of studies in other populations.1,2 Nigeria is the most populated African country and has more than 5 million people who are aged older than 65 years.3 To date, the only GBA screening reported in Sub-Saharan Africa populations was performed in Black South African patients;4 two novel missense variants (p.F216L and p.G478R) and three previously described (p.K(36)R, p.T36del, and p.Q497*) variants were identified in 30 patients with PD.4 The aim of this study was to assess the frequency of GBA mutations in a series of Nigerian patients with PD and controls by gene sequencing. Blood specimens were collected and participants were clinically characterized by the movement disorder specialists (O.O. and S.A.O.) in the Division of Neurology at Lagos State University Teaching Hospital, Lagos, Nigeria. The patients were diagnosed according to the UK Brain Bank criteria.5 The study protocol was approved by the Institutional Review Board of Lagos State University Teaching Hospital and Mayo Clinic Florida. For 92 patients and 51 controls, the 11 exons of GBA were polymerase chain reaction amplified using previously described primers.6 The identified variants were labeled according to GBA reference sequence (NM_001005742). The pathogenicity of non-synonymous variants was assessed with in silico tools (Combined Annotation Dependent Depletion, PolyPhen-2, Sorting Intolerant From Tolerant, ClinVar); the Genome Aggregation Database (gnomAD) was used to assess the published variant frequencies.

The demographic characteristics of the studied population are in Table 1. In the PD group, there were 10 variants (5 missense, 4 synonymous, and 1 loss of function) and 4 in controls (3 missense and 1 intronic) (Table 1). The most frequently observed variant in both groups was K(36)R (6.0% in cases and 4.0% in controls). Using in silico pathogenicity prediction tools, the potential disease-related variants (p.W184R, n = 1; p.L383PfsX3, n = 2; and p.L444P, n = 3) were observed in six PD cases (6.5%), and no likely pathogenic variants were seen in controls.

A limitation of our study is the relatively low number of samples from the Nigerian population; however, studies in Sub-Saharan Africa can be challenging due to access and availability of healthcare. In addition, although our case control study is the first GBA screening of patients with PD from Nigeria, we only have limited clinical details. Further genetic studies are needed in Sub-Saharan Africa, and there are still many populations in this region in which no genetic analysis has been performed. The recognition of population-specific mutations responsible for PD may also lead to new biomarker discoveries and support clinical genetic advances in Sub-Saharan populations.7 The expansion of current international consortia (eg, Human Heredity and Health in Africa, International Parkinson’s Disease Genomics Consortium Africa, and Genetic Epidemiology of Parkinson’s Disease) and the Global Parkinson’s Genetics Program efforts should help to create large, deeply phenotyped clinical cohorts for future studies.

Supporting Data
Additional Supporting Information may be found in the online version of this article at the publisher’s web-site.