Oocyte-Specific Deletion of Pten in Mice Reveals a Stage-Specific Function of PTEN/PI3K Signaling in Oocytes in Controlling Follicular Activation

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Abstract

Immature ovarian primordial follicles are essential for maintenance of the reproductive lifespan of female mammals. Recently, it was found that overactivation of the phosphatidylinositol 3-kinase (PI3K) signaling in oocytes of primordial follicles by an oocyte-specific deletion of Pten (phosphatase and tensin homolog deleted on chromosome ten), the gene encoding PI3K negative regulator PTEN, results in premature activation of the entire pool of primordial follicles, indicating that activation of the PI3K pathway in oocytes is important for control of follicular activation. To investigate whether PI3K signaling in oocytes of primary and further developed follicles also plays a role at later stages in follicular development and ovulation, we conditionally deleted the Pten gene from oocytes of primary and further developed follicles by using transgenic mice expressing zona pellucida 3 (Zp3) promoter-mediated Cre recombinase. Our results show that Pten was efficiently deleted from oocytes of primary and further developed follicles, as indicated by the elevated phosphorylation of the major PI3K downstream component Akt. However, follicular development was not altered and oocyte maturation was also normal, which led to normal fertility with unaltered litter size in the mutant mice. Our data indicate that properly controlled PTEN/PI3K-Akt signaling in oocytes is essential for control of the development of primordial follicles whereas overactivation of PI3K signaling in oocytes does not appear to affect the development of growing follicles. This suggests that there is a stage-specific function of PTEN/PI3K signaling in mouse oocytes that controls follicular activation.

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Introduction

In women, the 300,000–400,000 ovarian primordial follicles at menarche serve as the source of fertilizable ova for the entire duration of reproductive life [1]. In order to ensure the proper length of reproductive life, the majority of primordial follicles remain in a dormant state, and only limited numbers of them are recruited into the growing follicle pool through follicular activation. Menopause occurs when the pool of primordial follicles has become exhausted [1–3].

Recently, studies from our research group have suggested that the phosphatidylinositol 3-kinase (PI3K) signaling pathway in oocytes plays an important role in oocyte growth during early follicular development [4]. The PI3K pathway is a fundamental signaling pathway for regulation of cell proliferation, survival, migration, and metabolism [5]. PI3Ks are lipid kinases that phosphorylate the 3’-OH group on the inositol ring of inositol phospholipids and thus functions as a major negative regulator of PI3K [5].

We have shown that deletion of the Pten gene from mouse oocytes of primordial follicles using transgenic mice expressing Cre recombinase mediated by the growth differentiation factor 9 (Gdf-9) promoter, results in excessive activation of the entire pool of primordial follicles, which in turn leads to premature depletion of all primordial follicles [6]. However, such mutant mice show normal ovulation and normal litter size in young adulthood before their ovarian follicles are depleted [6].

The current study was designed to investigate the role of PTEN/PI3K–Akt signaling in oocytes of primary and further developed follicles in greater detail. We used a Cre-loxP system to delete the Pten gene from oocytes of primary follicles using transgenic mice carrying zona pellucida 3 (Zp3) promoter-mediated Cre recombinase [7]. We found that elevated PI3K–Akt signaling in oocytes of primary and further developed follicles does not affect the pool of primordial follicles. Moreover, female mice lacking Pten in oocytes of primary follicles showed unaltered follicular...
Results

Generation of mice with deletion of Pten in oocytes of primary and further developed follicles

To inactivate the Pten gene from oocytes of primary follicles, we crossed PtenloxP/loxP mice [6] with transgenic mice carrying Zp3 promoter-mediated Cre recombinase (Zp3-Cre mice) [7], where the Zp3 promoter drives the expression of Cre recombinase in oocytes from the primary follicle stage (Fig. 1). To determine the efficiency of deletion of Pten in oocytes of developing follicles, oocytes were collected by puncturing pregnant mare serum gonadotropin (PMSG)-treated ovaries, followed by western blot analysis. The expression of PTEN protein was found to be completely abolished in PtenloxP/loxP; Zp3-Cre+ oocytes (Fig. 2A).

We also confirmed the deletion of Pten gene from pups of PtenloxP/loxP; Zp3-Cre+ females by PCR. PtenloxP/loxP; Zp3-Cre+ female mice were mated with wild-type male mice and their offspring were genotyped for the deletion of Pten exon 5 in their tail-tip DNA. As shown in Fig. 2B, the pups showed deletion of Pten exon 5 (PtenΔ5) in one allele of their genomic DNA. These results indicate that the Pten gene was successfully deleted from growing oocytes of PtenloxP/loxP; Zp3-Cre+ mice.

Elevated PI3K–Akt signaling in PtenloxP/loxP; Zp3-Cre+ oocytes

We tested whether the PI3K–Akt signaling pathway is enhanced in PtenloxP/loxP; Zp3-Cre+ oocytes. As shown in Fig. 3, phosphorylation of Akt (at Ser473) was found to be elevated in PtenloxP/loxP; Zp3-Cre+ oocytes as compared to control oocytes. The phosphorylation of another important site on Akt, Thr308, which is required for complete activation of Akt, was also elevated upon deletion of Pten in growing PtenloxP/loxP; Zp3-Cre+ oocytes. As an indicator of Akt activity, the phosphorylation of Tsc2 (at Thr1462), which is an Akt substrate, was found to be elevated in PtenloxP/loxP; Zp3-Cre+ oocytes compared to control PtenloxP/loxP oocytes. These results suggest that there is enhanced activation of PI3K–Akt signaling in oocytes of growing follicles upon deletion of Pten in PtenloxP/loxP; Zp3-Cre+ mice.

Normal follicular development of PtenloxP/loxP; Zp3-Cre+ mice

Our previous work has shown that deletion of Pten from oocytes of primordial follicles leads to activation of the entire pool of primordial follicles, which eventually results in follicle depletion and premature ovarian failure (POF) in young adulthood [6]. To determine the function of Pten in oocytes of primary follicles, we first examined ovarian morphology of PtenloxP/loxP; Zp3-Cre+ at postnatal day 13 (PD13), PD23, and at 16 weeks of age in comparison to control PtenloxP/loxP; Zp3-Cre+ ovaries. We found that PtenloxP/loxP; Zp3-Cre+ ovaries at PD13 appeared normal in morphology as compared to control PtenloxP/loxP; GCre+ ovaries, with primordial, primary, secondary, and some early antral follicles (Fig. 4, A and B). In comparison, in ovaries of mice with deletion of Pten in oocytes of primordial follicles (referred to as PtenloxP/loxP; Zp3-Cre+ mice), prematurely activated primordial follicles (transient follicles with enlarged oocytes surrounded by flattened pre-granulosa cells) were observed (Fig. 4C, red arrows), as reported earlier [6]. These results show that deletion of Pten from primary follicles does not affect the primordial follicles in PtenloxP/loxP; Zp3-Cre+ mice.

By PD23, all the primordial follicles were activated in PtenloxP/loxP; GCre+ ovaries (Fig. 4, F and I, arrows). However, PtenloxP/loxP; Zp3-Cre+ ovaries exhibited normal morphology with primordial follicles (Fig. 4H, red arrows) and growing follicles (Fig. 4, E and H), which were comparable to PtenloxP/loxP; GCre+ ovaries (Fig. 4D; Fig. 4G, arrows). The premature activation of primordial follicles in PtenloxP/loxP; GCre+ mice led to complete depletion of follicles by 16 weeks of age (Fig. 4L) [6]. However, in ovaries of PtenloxP/loxP; Zp3-Cre+ mice at 16 weeks of age, the morphology was normal with all types of follicles...
and corpora lutea (CL) (Fig. 4K), which was similar to PtenloxP/loxP ovaries (Fig. 4J). Specifically, healthy CL and preovulatory follicles (Fig. 4K, arrows) were observed in 16-week-old PtenloxP/loxP; Zp3-Cre ovaries. Normal resumption of meiosis in PtenloxP/loxP; Zp3-Cre+ oocytes

It has been suggested that resumption of meiosis in oocytes is regulated by the PI3K–Akt signaling pathway [9–11]. Resumption of meiosis in oocytes involves germinal vesicle breakdown (GVBD), chromatin condensation and segregation, and extrusion of the first polar body [12]. To determine whether the enhanced PI3K–Akt signaling has any affect on resumption of meiosis in oocytes, we examined the rates of GVBD in PtenloxP/loxP; Zp3-Cre+ oocytes. Surprisingly, we found that the rate of GVBD in Pten-deficient oocytes was unaltered as compared to the control oocytes (Fig. 5A). These data indicate that the resumption of meiosis in oocytes was not affected by elevated PI3K–Akt signaling in PtenloxP/loxP; Zp3-Cre+ mice.

Normal ovulation, fertilization, and fertility in PtenloxP/loxP; Zp3-Cre+ mice

To test whether overactivation of PI3K–Akt signaling in oocytes has any effect on ovulation, PtenloxP/loxP; Zp3-Cre+ female mice and control PtenloxP/loxP; Zp3-Cre female mice were mated with wild-type males. Two-cell stage zygotes were retrieved from oviducts at E1.5 and were counted. We found that the numbers of 2-cell embryos in PtenloxP/loxP; Zp3-Cre+ mice were similar to those in control PtenloxP/loxP mice (Fig. 5B), indicating that the rates of ovulation and fertilization are not affected by the loss of Pten in ovulated oocytes. Importantly, during a testing period from 6 to 30 weeks of age, the PtenloxP/loxP; Zp3-Cre+ mice were completely fertile, with normal litter sizes (Fig. 5C).

The above results show that the elevated PI3K–Akt signaling in oocytes caused by loss of Pten does not affect follicular development, ovulation, and fertility in PtenloxP/loxP; Zp3-Cre+ mice.

Discussion

In recent years, the PI3K pathway in primary oocytes has been suggested to have important roles in controlling the activation and early development of ovarian follicles [4,13–15]. Activation of the PI3K pathway in oocytes by somatic cell-produced growth factors, such as Kit ligand (KL, or stem cell factor, SCF), results in activation of Akt, which is thought to mediate oocyte growth during follicular activation and early development [15]. Our previous study provided direct functional evidence that the initial recruitment of primordial follicles is tightly regulated by PI3K signaling in oocytes, as oocyte-specific deletion of Pten at the primordial follicle stage leads to global activation of all primordial follicles [6].

In the present study, to investigate how the PTEN/PI3K–Akt signaling in oocytes affects follicular development and oocyte maturation we deleted Pten in oocytes of primary and further developed follicles by using Zp3 promoter-mediated Cre transgenic mice. As expected, we found that the pool of primordial

Figure 2. Characterization of Pten deletion by western blot and PCR. (A) Oocytes were prepared and lysed for western blot as described in Materials and Methods. PTEN expression was found to be completely absent in PtenloxP/loxP; Zp3-Cre+ oocytes. For each lane, 150 oocytes were used. β-actin was used as internal control. (B) PCR analysis showing the complete deletion of Pten exon 5 (Pten Δ5) in one allele of the genomic DNA of pups from PtenloxP/loxP; Zp3-Cre+ females.

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follicles was unaffected throughout the reproductive life of PtenloxP/loxP; Zp3-Cre+ mice. Moreover, we observed normal follicular development, ovulation, and fertility in PtenloxP/loxP; Zp3-Cre+ mice, indicating that the PI3K signaling in oocytes is essential for follicular activation in a stage-specific fashion.

Foxo3a, which is a transcription factor and also an Akt substrate, has been shown to suppress follicular activation [16]. Oocyte-specific deletion of Foxo3a and Foxo3a-deficient (Foxo3a−/−) mice show similar phenotypes of premature activation of the entire primordial follicle pool [16,17], as we demonstrated in PtenloxP/loxP; GCre+ mice [6]. Interestingly, Foxo3a was found to be specifically required for the prevention of primordial follicles from activation [18]. Thus, these results support the notion that PI3K signaling in oocytes is probably more important for follicular activation in a stage-specific way.

It has been suggested that PI3K–Akt signaling plays an important role in the meiotic maturation of oocytes [9–11,19]. Inhibition of Akt activation in oocytes by a PI3K-specific inhibitor LY294002 results in suppressed GVBD and polar body extrusion in mouse oocytes [19], and inhibition of Akt phosphorylation leads to inactivation of cyclin-dependent kinase 1 (Cdk1), which in turn causes inhibition of meiosis resumption [11]. Furthermore, activation of phosphodiesterase 3A (PDE3A) by Akt-mediated phosphorylation has been suggested to play a role in the control of PDE3A activity in the maturation of mammalian oocytes [9]. The above studies suggest the importance of PI3K–Akt signaling in the oocyte for resumption of meiosis and maturation of mouse oocytes. At the same time, however, results from the current study have shown that oocytes from PtenloxP/loxP; Zp3-Cre+ mice undergo normal maturation, ovulation, and fertilization, and the mutant mice were completely fertile with normal litter sizes, indicating that the loss of Pten in oocytes does not apparently affect the fertility of the mice.

Results from our current work with the two mutant mouse models, the PtenloxP/loxP; GCre+ mice and PtenloxP/loxP; Zp3-Cre+ mice that carry Pten deficiency in oocytes of primordial and primary follicles, respectively, indicate that PTEN/PI3K–Akt signaling in oocytes is critically important for maintenance of the primordial follicle pool. On the other hand, further detailed and preferably in vivo studies are needed to elucidate the roles of PTEN and PI3K signaling in oocytes during oocyte maturation.

Materials and Methods

Mice

The PtenloxP/loxP mice [8] and Zp3-Cre transgenic mice where the Cre recombinase is specifically expressed in oocytes of primary and further developed follicles (Fig. 1) [7], were obtained from the Jackson Laboratory (Bar Harbor, ME). Both strains of mice have a C57BL/6J genomic background. After multiple rounds of crossing, mice with PtenloxP/loxP; Zp3-Cre+ genotype and control female mice with a PtenloxP/loxP genotype were obtained for experiments.

The mice with deletion of Pten in oocytes of primordial follicles (referred to as PtenloxP/loxP; GCre+ mice) were obtained by crossing PtenloxP/loxP mice with transgenic mice carrying Gdf-9 promoter-mediated Cre recombinase, which is specifically expressed in oocytes of primordial and further developed follicles (Fig. 1) [20], as previously described [6].

The mice were housed under controlled environmental conditions with free access to water and food. Illumination was on between 0600 and 1800 h. All experiments were conducted with the approval of the regional ethical committee of Umeå University, Sweden.

Reagents, antibodies, and immunological detection methods

The rabbit polyclonal antibodies to Akt, phospho-Akt (Ser473), tuberin/Tsc2, rabbit monoclonal antibody to phospho-tuberin/Tsc2 (Thr1462), and mouse monoclonal antibody to β-actin were purchased from BD Bioscience (Franklin Lakes, NJ). PMSG and mouse monoclonal antibody to β-actin were purchased from Sigma-Aldrich Sweden AB (Stockholm, Sweden). Western blots were carried out according to the instructions of the suppliers of the different antibodies and were visualized using the ECL Plus Western Blotting Detection System (Amersham Biosciences, Uppsala, Sweden).

Histological analysis

Ovaries were fixed in 4% paraformaldehyde, dehydrated, and embedded in paraffin. The paraffin-embedded ovaries were sectioned at 8-μm thickness and stained with hematoxylin for

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**Figure 3. Enhanced Akt signaling in oocytes of PtenloxP/loxP; Zp3-Cre+ mice.** Oocytes were prepared from ovaries of 3–4 week old mice that were treated with PMSG, as described in Materials and Methods. Signaling studies in PtenloxP/loxP; Zp3-Cre+ oocytes showed elevated levels of p-Akt (Ser473), p-Akt (Thr308), and p-Tsc2 (Thr1462) as compared to PtenloxP/loxP oocytes. Levels of total Akt, Tsc2, and β-actin were used as internal controls. 100–150 oocytes were used for each lane. All experiments were repeated at least three times and representative results are shown.

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morphological observation. Ovarian follicles at different developmental stages were categorized based on the well-accepted standards established by Pedersen and Peters [21]. Growing transient follicles were defined as follicles that have obviously enlarged oocytes but which are still enclosed in flattened pre-granulosa cells. Morphology images were taken with a light microscope (Leica).

Oocyte collection and culture
To stimulate follicular development, immature 23-day-old female mice were injected intraperitoneally with 5 IU of PMSG.

The ovaries were collected 48 h later and were dissected free of fat and connective tissue. Oocytes were collected by puncturing the ovaries with a sharp needle in M2 medium (Sigma), selected using a mouth pipette, and then cultured further in M16 medium (Sigma) at 37°C in an atmosphere of 5% CO2 for 6 h to determine the rate of GVBD.

For western blot, oocytes were lysed immediately after being selected with a mouth pipette, in a buffer containing 50 mM Tris-HCl (pH 8.0), 120 mM NaCl, 20 mM NaF, 20 mM β-glycerophosphate, 1 mM EDTA, 6 mM EGTA (pH 8.0), 1% NP-40, 1 mM DTT, 5 mM benzamidine, 1 mM PMSF, 250 μM sodium...
vandadate, 2 μg/ml aprotinin, 10 μg/ml leupeptin, and 1 μg/ml pepstatin, for 20 min on ice with frequent vortexing. The oocyte extracts were then used for western blot.

Natural ovulation and embryo culture

Adult female mice were housed with wild-type males, and vaginal plugs were checked every morning. Embryonic day 0.5 (E0.5) refers to the day when a vaginal plug was found. The mated females were sacrificed at E1.5, and 2-cell embryos were recovered from their oviducts and cultured further in KSOM medium (Specialty Media). The numbers and stages of development of the embryos were recorded.

Statistical analysis

All experiments were repeated at least 3 times. For comparisons of oocyte numbers, differences between the two groups were calculated with Student’s t-test, and a difference was considered significant if \( P < 0.05 \).

Author Contributions

Conceived and designed the experiments: KJ LL EL KL. Performed the experiments: KJ LL DA. Analyzed the data: KJ LL PR EL KL. Contributed reagents/materials/analysis tools: AI UO EL KL. Wrote the paper: KJ EL KL.

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