Antifilarial activity of *Butea monosperma* L. leaves extracts against *Setaria cervi*

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**Abstract**

**Background:** Usage of herbal drugs in traditional medicine is quite well known but largely empirical. Hence the present study was designed to screen the *in vitro* antifilarial effect of *Butea monosperma* L. plant leaves against *Setaria cervi*.

**Methods:** Antifilarial activity of Methanol/Hexane-ethanol extracts of *Butea monosperma* L. (leaves) and Ciprofloxacin was explored against adult of *Setaria cervi* after incubation for 24 hrs with concentration range of 0.25 to 20 mg/ml for possible antifilarial effect by comparing with suitable control, in terms of motility inhibition assay and MTT reduction assay.

**Results:** *Butea monosperma* L. plant leaves showed significant antifilarial activity against adult as compared to controls whereas activity demonstrated by Ciprofloxacin was comparatively less significant. Inhibitory concentrations (IC50) for the plant extracts with significant antifilarial activity against *Setaria cervi* adult in vitro system have been derived to be 1.25, 3.6 and 7.5 mg/ml Methanol, Hexane-ethanol extracts and Ciprofloxacin respectively.

**Conclusion:** The present research investigation proved to be an additional frame in recording the plant extract's antifilarial activity. Methanol and Hexane-ethanol extracts of *Butea monosperma* L. plant leaves which shown significant antifilarial activity. The present research data highlights the importance of further depth research in this area.

**Keywords:** Filariasis, *in vitro*, Drug discovery, *Setaria cervi*, *Butea monosperma* L.

**Introduction**

Lymphatic filariasis is a vector-borne parasitic disease, caused by nematodes (roundworms) parasitic species *Wuchereria bancrofti, Brugia malayi,* and *Brugia timori.* Lymphatic filariasis causes of disfigurement and disability in endemic areas, leading to significant economic and psychosocial impact. In India, around 45% of its 1–billion plus population lives in known endemic areas and 48 million are infected [1], accounting for 40% of the worldwide filariasis burden [2]. Socioeconomic studies showed that the annual loss caused by this disease is near to a billion U.S. dollars [3]. Diethylcarbamazine (DEC) is the drug of choice to treat patients suffered from filariasis. DEC kills circulating microfilariae and it is less effective against the adult worms. Adult worms may survive for several years in the infected person producing microfilariae and thereby facilitate transmission of the disease through the vector mosquitoes to more individuals. Hence elimination of the parasite by means of microfilaricide alone is extremely difficult [4]. Precisely because of these reasons, it is quite imperative to find out novel antifilarial drugs against the adult filarial worms. The World Health Organization (WHO) has already outlined the nature of traditional medicine including herbal therapeutics [5]. Plants are rich in resource materials for numerous phytomedicine utilized in traditional therapeutics. Plant based active principles can be derived from any part of the plant like bark, leaves, flowers, roots, fruits, seeds, and the others [6]. With the perspective of these encouraging advancements, in the present study plant *Butea monosperma* L. leaves was screened *in vitro* for their possible antifilarial activity against *Setaria cervi* adult filarial parasite.

**Materials and methods**

**Procuring plant material**

Leaves of *Butea monosperma* L. plant leaves (palash) were collected from campus of Barkatullah University, Bhopal in the month of Jan 2013. The botanical identity was confirmed by a botanist Prof. Zia-Ul-Hasan Department of Botany, Safia Science College, Bhopal and reference No. 407/Bot./Safia/15 allotted.

**Extraction**

The leaves of medicinal plant were washed, shade dried and powdered. The powder of *Butea monosperma* L. plant leaves were extracted with Petroleum ether, Hexane-Ethanol and Methanol respectively [7,8]. Different concentrations of extract were prepared by dissolving the extract in Dimethylsulfoxide (DMSO, Merck, Drug use grade) for further study.

**Parasite**

Adult *Setaria cervi* were obtained from the peritoneal cavity of freshly slaughtered cattle. The worms were washed repeatedly with normal saline (0.85%) to free them of any extraneous
In vitro motility inhibition assay
The worms were transferred immediately to DMEM (Dulbecco’s modified eagle’s medium) (Hi-Media, Mumbai, India) with 0.01% Strepto-penicillin (Hi-Media, Mumbai, India) and supplemented with 10% heat-inactivated fetal bovine serum (Hi-Media, Mumbai, India). Dilutions of the methanol/hexane-ethanol extract of *Butea monosperma* L. plant leaves and antibiotic Ciprofloxacin were made in DMSO (Dimethyl sulfoxide) (Merck India, drug use grade) in such a way that 100 µL of which, when distributed to sterile disposable Petri dishes (35-mm diameter and 5-mL capacity) containing 3 ml medium would give the required test concentration. Screening was done at concentrations ranging from 0.25 to 20 mg/mL. A simultaneous control was kept without the test solution but with 100µl DMSO in 3mL of the medium. One male and female worm introduced into each petri-dish. Three replicates each were set up for both test and control. The worms were incubated at 37ºC in 5% CO₂ incubator for 24 hrs. Motility observed after 5-24 hrs respectively. After exposure, the worms were washed twice with fresh medium and transferred to another set of fresh petridish containing fresh medium without the test solution to find out whether any of the immotile worms regained motility in the 2 hrs post treatment period in drug free Medium. If the worms did not revive, the condition was considered as irreversible and the concentration lethal. Each experiment was repeated thrice [9].

MTT Formazan colorimetric assay of crude extract
Effect of crude plant extracts on adult female *Setaria cervi* worms was studied by MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide) (Hi-media, Mumbai, India) - Formazan reduction assay following the method described by Comely et al., [10]. Because of the scarcity of male worms only female worms were used for these tests. The parasites were further incubated for 30 min individually in 0.5mL phosphate buffered saline (pH 7.4) containing 0.25 mgmL⁻¹ MTT. At the end of the incubation, worms were carefully transferred to a microtiter plate containing 400 µL of DMSO (Hi-media, Mumbai, India, Spectroscopic grade) and allowed to be at room temperature for 1 hr, with occasional gentle shaking to extract the colour developed. The absorbance of the resulting formazan solution was then determined at 492 nm in an enzyme-linked immunosorbent assay reader (ELISA plus, Microtiter plate reader) relative to DMSO blank. High values of absorption correlate with high viability of the worms. Adult worms that had previously been heat killed (56ºC for 30 min) and incubated with MTT served as the negative control. Viability of the worms was estimated as percentage inhibition in formazan formation relative to solvent controls and heat killed worms [11] by following the formula:

\[
\% \text{ inhibition reduction (parameter)} = 100 - \left( \frac{T - H}{C - H} \right) \times 100
\]

Where T, C, and H are absorbance values obtained for the formazan produced in treated, control, and heat killed worms respectively.

Statistical analysis
The results were expressed as mean±s.e.m for the triplicate observations made in each observation. For comparison of means of different parameters between the test plant extracts and respective controls, Student’s t test was used. P values of <0.05 were considered as significant.

Results
Preparation of plant extract
The solvent removed from the plant extract under reduced pressure and semisolid extract obtained.

In vitro motility inhibition assay
Crude extract of Methanol, Hexane–ethanol and antibiotic Ciprofloxacin was used for antifilarial screening against adult parasite *Setaria cervi*. Concentrations for Methanol and Hexane–ethanol extract 0.25 to 5.0, 0.50 to 10.0 mg/mL and for Ciprofloxacin concentration 1.0 to 20.0 mg/mL caused complete immobilization of the worms at 5 to 24 hrs exposure respectively at 37ºC, whereas in untreated control, all the worms were active (Table 1). Post exposure incubation in fresh medium (without test solution) for 2 hrs did not revive the worms, confirm their death due to the treatment by drug. The results shown that at high concentrations of drug, the inhibition in motility was faster, while at lower concentrations it was comparatively slow.

MTT-reduction assay
The adulticidal effect of the plant extracts and Ciprofloxacin was confirmed by comparison of the treated worms to un-

| Methanol extract (mgmL⁻¹) | Hexane-ethanol extract (mgmL⁻¹) | Ciprofloxacin (mgmL⁻¹) | Incubation time (end point) in hrs | Worm motility inhibition (Test) | Worm motility inhibition (Control) |
|--------------------------|---------------------------------|------------------------|----------------------------------|-------------------------------|----------------------------------|
| 0.25                     | 0.50                            | 1.00                   | 24.0                             | 100                           | 0                                |
| 0.50                     | 1.00                            | 2.00                   | 20.0                             | 100                           | 0                                |
| 1.00                     | 2.00                            | 5.00                   | 15.0                             | 100                           | 0                                |
| 2.00                     | 5.00                            | 10.0                   | 10.0                             | 100                           | 0                                |
| 5.00                     | 10.00                           | 20.0                   | 5.0                              | 100                           | 0                                |
treated control and heat-killed worms, in terms of MTT-formazan colorimetric assay. MTT is light yellow in solution, when incubated with living parasite, is reduced by live mitochondria to yield dark blue formazan within the cells, the formazan formed is extracted with DMSO and quantitated colorimetrically, during the assay. The very low absorbance value (<0.323) observed for the heat-killed worms was due to the less production of formazan in dead worms. The percentage inhibition (˃50%) was considered significant, was achieved at concentrations 1.0, 2.0, and 5.0 mg/mL for Methanol, for Hexane–ethanol extract 5.0 and 10.0 mg/mL, indicating the significant effect of the plant extract at lower concentrations considered as significant and for Ciprofloxacin it was 10.0 and 20.0 mg/mL (Table 2). Consequently, inhibitory concentration at which 50 per cent of the motility inhibition achieved (IC50), was calculated by plotting the graph of percentage reduction in MTT–assay against different concentrations of herbal drugs/Ciprofloxacin and the obtained values was for Methanol and Hexane–ethanol extract 1.25. 3.6 mg/mL respectively, while for Ciprofloxacin it was 7.5 mg/mL. Both worm motility assay and MTT-reduction assay confirmed the significant macrofilaricidal activity of plant extracts but antibiotic Ciprofloxacin failed to show such similar activity against Setaria cervi.

**Discussion**

In view of the huge socio-economic load of filarial disease, where this disease is much more prevalent, discovery and development of potent antifilarial curative drug candidate is essential. Herbal medicines are quite popular and are time tested for their safety, efficacy and cultural suitability. The plant based drugs are compatible to the human being. WHO has already referred this medicinal system in his TDR mandate [12].

During the present investigation an effort was made by screening of plant Butea monosperma L. leaves extracts for antifilarial activity against Setaria cervi. This plant is a traditionally used medicinal plant in many Ayurvedic drug preparations in India, revealed promising adulticidal activity. In in vitro experiment Methanol extracts shown significant activity than Hexane–ethanol. Another study was carried out for same plant against Brugia malayi human filarial nematode-aqueous extracts of leaves and roots shown significant activity against Brugia malayi filarial nematode [13]. Butea also showed activity against intestinal worms [14]. Other study was also carried out by various workers with other plant extracts against Setaria cervi-Aqueous and alcoholic extracts of the leaves of Mallotus philippensis (Lam.) was reported antifilarial activity [16]. Alcoholic extract of Plumbago indica [4], alcoholic and aqueous extracts extract of Azadirachta indica flowers [17] and Excoecaria agallocha L. leaves extracts [18] shown antifilarial activity. Effect of Asparagus adscendens Roxb [19]. Effect of alcoholic and aqueous extracts of the fruits of the Ficus racemosa Linn., significantly inhibited the spontaneous movements of the whole worm. [20], ethyl acetate extract of Vitex negundo leaves [21], and in another study Methanolic

| Sample          | Treatment            | Test concentration (mgmL-1) | Incubation time(In hrs.) | Absorbance at 492 nm (mean ± s.e.m.) | % reduction relative to solvent control, heat killed & treated worms | IC50 (mgmL-1) |
|-----------------|----------------------|----------------------------|--------------------------|--------------------------------------|---------------------------------------------------------------|--------------|
| Methanol extract | Control              | --                        | 24                       | 1.006 ± 0.003                       | --                                                             |              |
|                 | Heat killed          | --                        | 0.5                      | 0.319 ± 0.004                       | --                                                             |              |
|                 | Plant extract        | 0.25                      | 24.0                     | 0.91 ± 0.009*                       | 14.2                                                           |              |
|                 |                      | 0.50                      | 20.0                     | 0.788 ± 0.008*                      | 31.8                                                           | 1.25         |
|                 |                      | 1.00                      | 15.0                     | 0.622 ± 0.006*                      | 56.3                                                           |              |
|                 |                      | 2.00                      | 10.0                     | 0.522 ± 0.006*                      | 70.5                                                           |              |
|                 |                      | 5.00                      | 5.0                      | 0.336 ± 0.005*                      | 97.6                                                           |              |
| Hexane-ethanol extract | Control            | --                        | 24                       | 1.008 ± 0.002                       | --                                                             |              |
|                 | Heat killed          | --                        | 0.5                      | 0.322 ± 0.004                       | --                                                             |              |
|                 | Plant extract        | 0.50                      | 24.0                     | 0.955 ± 0.005*                      | 7.8                                                            | 3.6          |
|                 |                      | 1.00                      | 20.0                     | 0.87 ± 0.002*                       | 20.7                                                           |              |
|                 |                      | 2.00                      | 15.0                     | 0.745 ± 0.002*                      | 38.4                                                           |              |
|                 |                      | 5.00                      | 10.0                     | 0.56 ± 0.009*                       | 65.4                                                           |              |
|                 |                      | 10.0                      | 5.0                      | 0.345 ± 0.003*                      | 96.7                                                           |              |
| Ciprofloxacin   | Control              | --                        | 24                       | 1.011 ± 0.003                       | --                                                             |              |
|                 | Heat killed          | --                        | 0.5                      | 0.323 ± 0.001                       | --                                                             |              |
|                 | Antibiotics          | 1.0                       | 24.0                     | 0.965 ± 0.002*                      | 6.7                                                            | 7.5          |
|                 |                      | 2.0                       | 20.0                     | 0.886 ± 0.004*                      | 18.2                                                           |              |
|                 |                      | 5.0                       | 15.0                     | 0.688 ± 0.005*                      | 47                                                             |              |
|                 |                      | 10.0                      | 10.0                     | 0.551 ± 0.011*                      | 66.9                                                           |              |
|                 |                      | 20.0                      | 5.0                      | 0.342 ± 0.002*                      | 97.3                                                           |              |

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*P value represents the level of significance P<0.05 when comparing the mean value of absorbance observed for the formazan formed between treated and control worms.
extract of leaves of *Hibiscus mutabilis* exhibited activity against *Setaria cervi* in vitro [22]. Ciprofloxacin was also assessed for antifilarial activity in present research. This antibiotic not showed much more activity as other two extracts used. In similar study Ciprofloxacin was shown antifilarial activity against *Brugia malayi* parasite [23]. Novobiocin antibiotics also found to be active in many experiment against *Brugia malayi* [24,25].

These results revealed that some active molecules present in this plant extract might be responsible for the real antifilarial effect. Therefore, it would be interesting to find out the basis behind the pharmaceutical approach of this potential drug candidate in the light of phytochemical basis of this extract. Usage of herbal drugs in traditional medicine is well known and so now these herbals are largely being explored. In the present investigation methanolic extract found potent antifilarial drug revealed towards the importance of in depth study for design and development of new antifilarial therapeutic drug candidate; which may actually prove better in terms of cost effectiveness and patient fulfilment to combat this disease.

**Competing interest**
The authors declare that they have no competing interests.

**Authors’ contributions**

| Authors’ contributions | MD | KNS | RKP | BM | VS |
|------------------------|----|-----|-----|----|----|
| Research concept and design | -- | -- | -- | -- | ✓ |
| Collection and/or assembly of data | ✓ | -- | ✓ | -- | -- |
| Data analysis and interpretation | -- | ✓ | -- | -- | -- |
| Writing the article | -- | ✓ | -- | ✓ | -- |
| Critical revision of the article | -- | -- | -- | -- | ✓ |
| Final approval of article | -- | -- | -- | -- | ✓ |
| Statistical analysis | -- | ✓ | -- | -- | -- |

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