Serum Vascular Endothelial Growth Factor and Angiopoietin-2 Are Associated with the Severity of Systemic Inflammation Rather than the Presence of Hemoptysis in Patients with Inflammatory Lung Disease

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Purpose: Vascular endothelial growth factor (VEGF) and angiopoietin-2 (Ang-2) are major mediators of angiogenesis and are induced by tissue inflammation and hypoxia. The purpose of this study was to investigate whether serum VEGF and Ang-2 are associated with the presence of hemoptysis and the extent of systemic inflammation in patients with inflammatory lung diseases. Materials and Methods: We prospectively enrolled 52 patients with inflammatory lung disease between June 2008 and October 2009. Results: The median values of VEGF and Ang-2 were 436 pg/mL and 2383 pg/mL, respectively. There was a significant positive correlation between serum Ang-2 and VEGF levels. VEGF levels were not significantly different according to the presence of hemoptysis. C-reactive protein (CRP) and Ang-2 level were significantly higher in patients without hemoptysis (n=26) than in those with hemoptysis (n=26; p<0.001 and p<0.001, respectively). CRP and arterial oxygen tension (PaO₂) were significantly correlated with both serum VEGF (r=0.032 and p=0.016, respectively) and Ang-2 levels (r=0.001 and p=0.041, respectively), after adjusting for other factors. Age and the absence of hemoptysis were factors correlated with serum Ang-2 levels. Conclusion: Our study suggests that serum VEGF and Ang-2 levels are associated with PaO₂ and the severity of inflammation rather than the presence of hemoptysis in patients with inflammatory lung diseases. Thus, hemoptysis may not be mediated by increased serum levels of VEGF and Ang-2 in patients with inflammatory lung diseases, and further studies are required to determine the mechanisms of hemoptysis.

Key Words: Angiogenesis, angiopoietin-2, hemoptysis, inflammation, vascular endothelial growth factor

INTRODUCTION

Angiogenesis is characterized by the formation of new microvessels from preex-
isting vasculature and is associated with numerous inflammatory conditions, such as atherosclerosis, arthritis, retinopathy, and tumor growth. Previous studies provided evidence that inflammation exists in a mutually dependent association with angiogenesis.

Among the numerous angiogenic factors, vascular endothelial growth factor (VEGF) is the most extensively studied and is significantly related to the severity of inflammatory lung diseases, such as active tuberculosis, chronic bronchitis, pulmonary aspergilloma, and pulmonary disease caused by cystic fibrosis. Overexpression of VEGF is also significantly correlated with lung cancer progression and metastasis. Antiangiogenic agents have received widespread attention as targets for lung cancer therapy. Recent studies reported that VEGF elevation correlates significantly with the presence of hemoptysis in patients with pulmonary aspergilloma. Another major factor involved in angiogenesis, angiopoietin-2 (Ang-2), is also induced by hypoxemia and plays an important role in initiating vessel sprouting in concert with VEGF. In contrast with angiopoietin-1, which stabilizes blood vessels, Ang-2 destabilizes blood vessels, initiating angiogenic changes instead of regression, and promotes the neovascularization of tumor cells.

The purpose of this study was to investigate whether serum VEGF and Ang-2 were associated with the presence of hemoptysis and the level of systemic inflammation in patients with inflammatory lung diseases.

**Materials and Methods**

**Study population**

The study was approved by the Institutional Review Board of Samsung Medical Center (a 2600-bed University affiliated hospital in Seoul, Korea) and was registered with www.clinicaltrials.gov (NCT01171768). Informed written consent was obtained from all participants.

Patients presented to the Samsung Medical Center for the treatment of benign inflammatory lung diseases, including bronchiectasis, aspergilloma, pneumonia, and post-tuberculosis destroyed lung, between June 2008 and December 2009. Diagnosis of post-tuberculosis destroyed lung (inactive tuberculosis) was considered when chest computed tomography (CT) scan showed the findings compatible with fibrotic sequelae of old tuberculosis and all mycobacterial examinations of sputum or bronchial washing fluid were negative. Patients with hemoptysis (hemoptysis group) were enrolled if they had hemoptysis within 2 weeks before study enrollment, and the patients without hemoptysis (no hemoptysis group) were enrolled if they did not have any hemoptysis more than 2 years before study enrollment. All study patients except one were admitted to general ward or emergency department for the management of symptomatic benign lung diseases. We excluded patients with neoplastic conditions, including lung cancer, vasculitis, and lung disease associated with collagen vascular disease. Serum samples for VEGF and Ang-2, clinical data, such as age, gender, smoking status, clinical symptoms, and laboratory data, including peripheral white blood cell (WBC) count, hemoglobin (Hb), prothrombin time (PT), C-reactive protein (CRP), and arterial oxygen tension (PaO2), were obtained at study enrollment.

**Determination of sample size**

The required sample size was calculated based on VEGF levels according to the presence of hemoptysis in patients with aspergilloma. Assuming 437±164 pg/mL of VEGF in the hemoptysis group and 162±102 pg/mL in the no hemoptysis group, a sample size of 42 patients (21 per group) was required to detect a 120 pg/mL difference between the two groups with a power of 90% and α error of 0.05 using a two-tailed test. Anticipating a potential 20% drop-out rate, we planned to include 52 subjects (26 per group).

**Measurement of VEGF and Ang-2**

Serum samples from each individual were obtained at the time of study enrollment. Sera were stored at -80°C. Serum VEGF and Ang-2 concentrations were measured in triplicate for each sample using a commercial enzyme-linked immunosorbent assay (ELISA) kit (R&D Systems, Minneapolis, MN, USA). The detection ranges of ELISA kits for VEGF and Ang-2 were 31.2-2000 pg/mL and 46.9-3000 pg/mL, respectively. Therefore, serum was diluted to measure serum Ang-2 levels.

**Statistical analysis**

Data presented are expressed as either the median and interquartile range (IQR, 25th and 75th percentiles) or the number (percentage) of patients. For univariate analysis or relation with hemoptysis, categorical variables were analyzed using a Pearson χ² test or Fisher’s exact test; continuous variables were analyzed using a Mann-Whitney U test. VEGF and Ang-2 levels did not fit normality assumptions;
The median VEGF levels were 375 pg/mL in bronchiectasis, 472 pg/mL in aspergilloma, 554 pg/mL in post-tuberculosis destroyed lung, and 451 pg/mL in pneumonia. The median Ang-2 levels were 2444 pg/mL in bronchiectasis, 1689 pg/mL in aspergilloma, 3021 pg/mL in post-tuberculosis destroyed lung, and 4344 pg/mL in pneumonia. Serum Ang-2 levels were significantly correlated with serum VEGF levels ($p=0.028$) (Fig. 1). Serum VEGF levels demonstrated a significant positive correlation with WBC and a negative correlation with PaO$_2$ levels (Table 3). Age, gender, smoking status, presence of hemoptysis, and CRP levels showed no significant correlation with VEGF levels.

RESULTS

Baseline clinical and laboratory features

Characteristics of the enrolled patients are listed in Table 1. In this study, 52 study patients underwent extensive evaluations for underlying disease including chest CT scan ($n=46$), bronchoscopy ($n=21$), and sputum bacterial and mycobacterial examinations ($n=44$). There were 25 men and 27 women, with a median age of 58 years (range, 47-66). Of the 52 patients, 14 patients (27%) had a history of smoking, and 22 (42%) had a history of tuberculosis treatment. The most common disease in enrolled patients was bronchiectasis (62%); 14% had an aspergilloma, and 14% had post-tuberculosis destroyed lung. Median VEGF and Ang-2 levels were 436 pg/mL (257-724) and 2383 pg/mL (1807-3209), respectively. In total, 5 patients received oxygen therapy (1-2 L/min) on arterial blood gas analysis.

Comparison of clinical laboratory features depending on the presence of hemoptysis

In patients with hemoptysis ($n=26$), bronchiectasis (54%), aspergilloma (27%), and post-tuberculosis destroyed lung (19%) were observed. In those without hemoptysis ($n=26$), bronchiectasis (69%), post-tuberculosis destroyed lung (8%), pneumonia (19%), and pulmonary tuberculosis (4%) were observed. Patients with hemoptysis had more significant history of tuberculosis treatment compared with those without hemoptysis. However, the median CRP and Ang-2 levels were significantly higher in patients without hemoptysis than in those with hemoptysis (CRP; 0.34 vs. 3.29 mg/dL; $p<0.001$, and Ang-2; 2017 vs. 2946 pg/mL; $p=0.001$). There was no significant difference in age, gender, smoking history, presenting symptoms, or laboratory findings (including WBC, Hb, PT, PaO$_2$, serum VEGF levels) (Table 2).

Correlation between serum VEGF or angiopoietin-2 and other parameters

The median VEGF levels were 375 pg/mL in bronchiectasis, 472 pg/mL in aspergilloma, 554 pg/mL in post-tuberculosis destroyed lung, and 451 pg/mL in pneumonia. The median Ang-2 levels were 2444 pg/mL in bronchiectasis, 1689 pg/mL in aspergilloma, 3021 pg/mL in post-tuberculosis destroyed lung, and 4344 pg/mL in pneumonia.

Serum Ang-2 levels were significantly correlated with serum VEGF levels ($p=0.028$) (Fig. 1). Serum VEGF levels demonstrated a significant positive correlation with WBC and a negative correlation with PaO$_2$ levels (Table 3). Age, gender, smoking status, presence of hemoptysis, and CRP levels showed no significant correlation with VEGF levels.
of hemoptysis were significantly correlated with serum Ang-2 levels (Table 4).

### DISCUSSION

In the present study, serum VEGF levels did not differ according to the presence of hemoptysis. However, we demonstrated that serum VEGF and Ang-2 levels in patients with inflammatory lung diseases correlated significantly with elevations in serum CRP and reduction of PaO\(_2\).

**Table 2. Comparison of Clinical Laboratory Features according to the Presence of Hemoptysis**

|                      | With hemoptysis (n=26) | Without hemoptysis (n=26) | \(p\) value |
|----------------------|------------------------|---------------------------|-------------|
| **Age**              | 56 (47-65)             | 60 (48-66)                | 0.826       |
| **Sex**              |                        |                           | 0.405       |
| Female               | 11 (42%)               | 14 (54%)                  |             |
| Male                 | 15 (58%)               | 12 (46%)                  |             |
| **Smoking status**   |                        |                           | 0.252       |
| Current smoker       | 2 (8%)                 | 0                         |             |
| Ex-smoker            | 7 (27%)                | 5 (19%)                   |             |
| Non-smoker           | 17 (65%)               | 21 (81%)                  |             |
| **Comorbidities**    |                        |                           | 0.005       |
| Hypertension         | 6 (23%)                | 2 (8%)                    | 0.248       |
| Diabetes mellitus    | 1 (4%)                 | 1 (4%)                    | 1.000       |
| Prior therapy for tuberculosis | 15 (58%) | 7 (27%) | 0.025 |
| **Lung diseases**    |                        |                           |             |
| Bronchiectasis       | 14 (54%)               | 18 (69%)                  |             |
| Aspergiloma          | 7 (27%)                | 0                         |             |
| Post-tuberculosis destroyed lung | 5 (19%) | 2 (8%) |             |
| Pneumonia            | 0                      | 5 (19%)                   |             |
| Pulmonary tuberculosis| 0                     | 1 (4%)                    |             |
| **Severity of diseases** |                    |                           |             |
| No. of involved lobes| 2 (1-3)                | 3 (2-4)                   | 0.390       |
| **Presenting Symptoms** |                      |                           |             |
| Cough                | 16 (62%)               | 20 (77%)                  | 0.229       |
| Dyspnea              | 6 (23%)                | 9 (35%)                   | 0.358       |
| Fever                | 2 (8%)                 | 5 (19%)                   | 0.419       |
| **Laboratory findings** |                      |                           |             |
| WBC (×10\(^3\)/μL)  | 7645 (6353-9695)       | 8285 (5973-10638)         | 0.742       |
| Hemoglobin (g/dL)    | 13 (11-14)             | 12.4 (11.8-13.9)          | 0.615       |
| Platelet (×10\(^3\)/μL) | 216 (186-260)         | 250 (195-317)             | 0.359       |
| PT (INR)             | 1.06 (1.01-1.14)       | 1.04 (0.98-1.12)          | 0.359       |
| aPTT (sec)           | 36 (31-38)             | 38 (34-41)                | 0.039       |
| CRP (mg/dL)          | 0.34 (0.13-2.09)       | 3.29 (0.60-7.19)          | 0.001       |
| PaO\(_2\) (mm Hg)   | 83.5 (75.1-94.2)       | 74.7 (64.7-87.6)          | 0.142       |
| VEGF*                | 389 (270-684)          | 449 (224-776)             | 0.934       |
| Ang-2*               | 2017 (1633-2597)       | 2946 (2101-4698)          | <0.001      |

WBC, white blood cell; PT, prothrombin time; aPTT, activated partial thromboplastin time; CRP, C-reactive protein; VEGF, vascular endothelial growth factor; Ang-2, angiopeptin-2; INR, international normalized ratio; PaO\(_2\), arterial oxygen tension.

*The analysis used a mixed model, accounting for the random effect between subjects (logarithmic transformed VEGF and Ang-2 values were used).
Results also demonstrated that VEGF levels were increased in inflammatory lung diseases, such as bronchiectasis, aspergilloma, post-tuberculosis destroyed lung, and pneumonia, when compared with published normal values.20,21 Additionallly, elevated VEGF levels correlated with severity of inflammation. Significant change in the serum VEGF levels...

**Fig. 1.** Correlation between serum VEGF and Ang-2 levels. VEGF, vascular endothelial growth factor; Ang-2, angiopoietin-2.

**Table 3.** Univariate Analysis of Associations between Serum VEGF or Angiopoietin-2 levels and Other Measured Parameters

| Variables                              | Ln [VEGF] Coefficient | SE  | p value | Ln [Ang-2] Coefficient | SE  | p value |
|----------------------------------------|------------------------|-----|---------|------------------------|-----|---------|
| Age                                    | -0.003                 | 0.005 | 0.524   | 0.013                  | 0.003 | <0.001  |
| Sex (female*)                          | 0.114                  | 0.128 | 0.375   | 0.144                  | 0.081 | 0.077   |
| Smoking status† (non-smoker*)          | 0.116                  | 0.144 | 0.423   | 0.092                  | 0.092 | 0.317   |
| Comorbidities                          |                        |      |         |                        |      |         |
| Hypertension (No*)                     | -0.159                 | 0.177 | 0.370   | -0.020                 | 0.113 | 0.862   |
| Diabetes mellitus (No*)                | -0.050                 | 0.333 | 0.880   | 0.260                  | 0.211 | 0.220   |
| Prior therapy for tuberculosis (No*)   | 0.089                  | 0.130 | 0.493   | -0.154                 | 0.082 | 0.061   |
| Presenting symptoms                    |                        |      |         |                        |      |         |
| Cough (No*)                            | 0.247                  | 0.137 | 0.074   | 0.196                  | 0.087 | 0.026   |
| Dyspnea (No*)                          | 0.355                  | 0.139 | 0.011   | 0.286                  | 0.087 | 0.001   |
| Fever (No*)                            | -0.199                 | 0.187 | 0.288   | 0.030                  | 0.120 | 0.801   |
| Hemoptyaxis (No*)                      | 0.011                  | 0.128 | 0.934   | -0.420                 | 0.074 | <0.001  |
| Amount of hemoptyaxis (cc)             | -0.0001                | 0.0003 | 0.672   | 0.0003                 | 0.0002 | 0.121   |
| Laboratory findings                    |                        |      |         |                        |      |         |
| WBC (#/μL)                             | 0.00006                | 0.00002 | 0.005   | 0.000064               | 0.00001 | <0.001  |
| Hemoglobin (g/dL)                      | 0.016                  | 0.017 | 0.358   | -0.004                 | 0.011 | 0.745   |
| PT (INR)                               | 1.085                  | 0.733 | 0.141   | 1.0961                 | 0.462 | 0.019   |
| CRP (mg/dL)                            | 0.022                  | 0.02  | 0.160   | 0.059                  | 0.009 | <0.001  |
| PaO2 (mm Hg)                           | -0.006                 | 0.003 | 0.055   | -0.004                 | 0.002 | 0.027   |

Ln, natural logarithm; WBC, white blood cell; PT, prothrombin time; CRP, C-reactive protein; VEGF, Vascular endothelial growth factor; Ang-2, angiopoietin-2; SE, standard error; PaO2, arterial oxygen tension.

Analysis using a mixed model to account for the random effect between subjects (logarithmic transformed VEGF and Ang-2 values were used).

*Reference category.

†Smoking status: smokers (current smoker or ex-smoker) and non-smoker.

Angiogenesis, the growth of new capillary blood vessels from pre-existing vasculature, is an important contributor to tissue inflammation and abnormal remodeling in inflammatory diseases. Consistent with previous data that levels of VEGF are significantly elevated in inflammatory disorders, including lung inflammatory disease,6,7,18,19 our present results also demonstrated that VEGF levels were increased in inflammatory lung diseases, such as bronchiectasis, aspergilloma, post-tuberculosis destroyed lung, and pneumonia, when compared with published normal values.20,21 Additionally, elevated VEGF levels correlated with severity of inflammation. Significant change in the serum VEGF levels...
with the treatment of inflammatory lung diseases have been reported.4,6 To our best knowledge, however, our present study is the first to demonstrate that serum VEGF levels are positively correlated with serum CRP levels and WBCs. In a previous study on patients undergoing major surgery, there was no significant correlation between increases in serum VEGF levels and serum IL-6 or CRP levels.22 Recently, several clinical studies have shown that circulating levels of Ang-2 are increased in septic shock patients, as well as non-septic patients with, or at risk of, acute lung injury/acute respiratory distress syndrome; this is related to vascular permeability and pulmonary dysfunction.23-26 In these studies, serum Ang-2 levels were positively correlated with serum CRP levels. In the present study, Ang-2 levels were positively correlated with CRP levels and WBCs, both representing markers of inflammation. Compared with the levels of Ang-2 in previous studies,23,25,27 the median level of Ang-2 in septic shock patients was greater than 4000 pg/mL; this is significantly higher than the levels seen in the current study (median Ang-2 level of 2383 pg/mL). However, the levels of Ang-2 in the current study are similar to data measured in cancer patients, including lung, breast, and prostate;15,28 and were greater than the levels of Ang-2 in the healthy volunteers without lung disease (1000-1500 pg/mL).28 Our findings suggest that high Ang-2 levels are associated with the severity of inflammatory lung disease in patients without sepsis, and that the Ang-2 levels in inflammatory diseases are similar to the levels reported in the setting of various cancers.15,28 To our best knowledge, this is the first reported study to evaluate Ang-2 levels in inflammatory lung diseases, including bronchiectasis, post-tuberculosis destroyed lung, and aspergilloma.

During development of inflammatory tissue, hypoxic conditions can promote angiogenesis, leading to induced VEGF and Ang-2 production. Expression of Ang-2 was elevated in cultured endothelial cells exposed to hypoxia, while VEGF mRNA expression was increased.32,33 In the present study, PaO\textsubscript{2} levels were correlated inversely with VEGF and Ang-2 levels; suggesting that angiogenesis mediators may be released in response to low PaO\textsubscript{2} in patients with inflammatory lung diseases. Significant associations between VEGF and Ang-2 levels have been noted in earlier studies in patients with various tumors,28 diabetes mellitus,24 and asthma.15 The present study demonstrated that serum VEGF and Ang-2 levels had a significant positive correlation in inflammatory lung diseases. However, in contrast to a previous study,6 our study did not find a significant association between the presence of hemoptysis and VEGF levels. This lack of an association may be explained by differences in the distribution of disease between the hemoptysis and the no hemoptysis groups. As shown in Table 2, the prevalence of bronchiectasis (54%), aspergilloma (27%), and post-tuberculosis destroyed lung (19%) in the hemoptysis group differed from the no hemoptysis group where bronchiectasis (69%), pneumonia (19%), and post-tuberculosis destroyed lung (8%) were prevalent.

### Table 4. Multivariate Correlations Between Serum VEGF or Angiopoietin-2 Levels and Other Measured Parameters

|                        | Coefficient | SE  | T   | p value |
|------------------------|-------------|-----|-----|---------|
| Ln [VEGF]              |             |     |     |         |
| Age                    | -0.007      | 0.005 | -1.26 | 0.211   |
| Sex (female*)          | -0.142      | 0.177 | -0.80 | 0.424   |
| Smoking status (non-smoker*) | 0.121  | 0.180 | 0.67  | 0.503   |
| The presence of hemoptysis | 0.188  | 0.152 | 1.24  | 0.218   |
| CRP (mg/dL)            | 0.042       | 0.020 | 2.16  | 0.032   |
| PaO\textsubscript{2} (mm Hg) | -0.007  | 0.003 | -2.45 | 0.016   |
| Ln [Ang-2]             |             |     |     |         |
| Age                    | 0.012       | 0.003 | 4.43  | <0.001  |
| Sex (female*)          | 0.127       | 0.090 | 1.42  | 0.157   |
| Smoking status (non-smoker*) | 0.025  | 0.091 | 0.28  | 0.783   |
| The presence of hemoptysis | -0.275  | 0.077 | -3.59 | <0.001  |
| CRP (mg/dL)            | 0.038       | 0.010 | 3.88  | <0.001  |
| PaO\textsubscript{2} (mm Hg) | -0.003  | 0.002 | -2.06 | 0.041   |

Ln, natural logarithm; T, t statistic value; VEGF, vascular endothelial growth factor; Ang-2, angiopoietin-2; CRP, C-reactive protein; SE, standard error; PaO\textsubscript{2}, arterial oxygen tension.

Analysis using a mixed model to account for the random effect between subjects (logarithmic transformed VEGF and Ang-2 values were used).

*Reference category.

†Smoking status: smokers (current smoker or ex-smoker) and non-smoker.
When the VEGF levels were compared between patients with aspergilloma, which were present only in the hemoptysis group, and those with pneumonia, which were present only in the non-hemoptysis group, no significant difference in VEGF levels was observed. Additionally, when comparison was restricted to patients with bronchiectasis from both groups, VEGF levels were not significantly correlated with the presence of hemoptysis. The lack of a significant difference in VEGF levels between the hemoptysis and no hemoptysis groups can be partially explained by the different disease distribution in the two groups. In previous reports of associations between VEGF levels and hemoptysis in 21 pulmonary aspergilloma patients, the mean serum VEGF levels in 6 patients with hemoptysis and 15 patients without hemoptysis were 437 pg/mL and 162 pg/mL, respectively. The reported serum VEGF levels were similar to the aspergilloma patients with hemoptysis in the current study.

The present study has several other limitations. The measurements of VEGF and Ang-2 levels were made with serum, not lung tissue; it is unclear whether serum VEGF or Ang-2 levels reflect the levels in lung tissue. Also, because the present study was conducted with multiple inflammatory lung diseases, the sample size might have influenced the statistical power; this limits the interpretation of the study despite the fact that a formal sample size calculation was performed. Finally, the timing of measurement of serum angiogenesis markers in this study was not uniform in relation to stages of lung injury. Since the level of serum angiogenesis markers could vary in different stages of lung injury, our data should be interpreted conservatively. A larger-scale study in patients with bronchiectasis, aspergilloma, and pneumonia is needed to further clarify the association between hemoptysis and VEGF levels.

In conclusion, serum VEGF and Ang-2 levels were associated with the PaO2 and the severity of inflammation, rather than the presence of hemoptysis in patients with inflammatory lung diseases. Hemoptysis in patients with inflammatory lung diseases may not be mediated by increased serum levels of VEGF and Ang-2. Further studies are required to determine the mechanism(s) of hemoptysis in patients with inflammatory lung disease.

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REFERENCES

1. Folkman J. Angiogenesis in cancer, vascular, rheumatoid and other disease. Nat Med 1995;1:27-31.
2. Jackson JR, Seed MP, Kircher CH, Willoughby DA, Winkler JD. The codependence of angiogenesis and chronic inflammation. FASEB J 1997;11:457-65.
3. Dvorak HF, Brown LF, Detmar M, Dvorak AM. Vascular permeability factor/vascular endothelial growth factor, microvascular hyperpermeability, and angiogenesis. Am J Pathol 1995;146:1029-39.
4. Matsuyma W, Hashiguchi T, Matsumuro K, Iwami F, Hirotsu Y, Kawabata M, et al. Increased serum level of vascular endothelial growth factor in pulmonary tuberculosis. Am J Respir Crit Care Med 2000;162:1120-2.
5. Kanazawa H. Role of vascular endothelial growth factor in the pathogenesis of chronic obstructive pulmonary disease. Med Sci Monit 2007;13:RA189-95.
6. Inoue K, Matsuyma W, Hashiguchi T, Wakimoto J, Hirotsu Y, Kawabata M, et al. Expression of vascular endothelial growth factor in pulmonary aspergilloma. Intern Med 2001;40:1195-9.
7. McColley SA, Stellmach V, Boas SR, Jain M, Crawford SE. Serum vascular endothelial growth factor is elevated in cystic fibrosis and decreases with treatment of acute pulmonary exacerbation. Am J Respir Crit Care Med 2000;161:1877-80.
8. Donovan EA, Kummars S. Targeting VEGF in cancer therapy. Curr Probl Cancer 2006;30:7-32.
9. Mattern J, Koomägi R, Volm M. Association of vascular endothelial growth factor expression with intratumoral microvessel density and tumour cell proliferation in human epidermoid lung carcinoma. Br J Cancer 1996;73:931-4.
10. Nilsson I, Shibuya M, Wennström S. Differential activation of vascular genes by hypoxia in primary endothelial cells. Exp Cell Res 2004;299:476-85.
11. Tsikos S, Koutsilieris M, Papapetropoulos A. Angiopoietins in angiogenesis and beyond. Expert Opin Investig Drugs 2003;12:933-41.
12. Hashizume H, Falcón BL, Kuroda T, Baluk P, Coxon A, Yu D, et al. Complementary actions of inhibitors of angiopoietin-2 and VEGF on tumor angiogenesis and growth. Cancer Res 2010;70:2213-23.
13. Pouysségur J, Dayan F, Mazure NM. Hypoxia signalling in cancer and approaches to enforce tumour regression. Nature 2006;441:437-43.
14. Bousat S, Eddahibi S, Coste A, Fataccioli V, Gouge M, Houssset B, et al. Expression and regulation of vascular endothelial growth factor in human pulmonary epithelial cells. Am J Physiol Lung Cell Mol Physiol 2000;279:L371-8.
15. Park JH, Park KJ, Kim YS, Sheen SS, Lee KS, Lee HN, et al. Serum angiopoietin-2 as a clinical marker for lung cancer. Chest 2007;132:290-6.
16. Jain RK. Molecular regulation of vessel maturation. Nat Med 2003;9:685-93.
17. Hayes RJ, Bennett S. Simple sample size calculation for cluster-randomized trials. Int J Epidemiol 1999;28:319-26.
18. Kraft A, Weinled K, Ochs A, Marth C, Zmija J, Schumacher P, et al. Vascular endothelial growth factor in the sera and effusions of patients with malignant and nonmalignant disease. Cancer 1999;85:178-87.
19. Alatas F, Alatas O, Metintas M, Ozarslan A, Erginel S, Yildirim H. Vascular endothelial growth factor levels in active pulmonary tuberculosis. Chest 2004;125:2156-9.
20. Hyodo I, Doi T, Endo H, Hosokawa Y, Nishikawa Y, Tanimizu M, et al. Clinical significance of plasma vascular endothelial growth factor in gastrointestinal cancer. Eur J Cancer 1998;34:2041-5.
21. Kikuchi K, Kudo M, Kadono T, Yazuwa N, IHN H, Tamaki K. Serum concentrations of vascular endothelial growth factor in colagen diseases. Br J Dermatol 1998;139:1049-51.
22. Futami R, Miyashita M, Nomura T, Makino H, Matsutani T, Sasa jima K, et al. Increased serum vascular endothelial growth factor following major surgical injury. J Nihon Med Sch 2007;74:223-9.
23. Siner JM, Bhandari V, Engle KM, Elias JA, Siegel MD. Elevated serum angiopoietin 2 levels are associated with increased mortality in sepsis. Shock 2009;31:348-53.
24. van der Heijden M, Pickkers P, van Nieuw Amerongen GP, van Hinsbergh VW, Bouw MP, van der Hoeven JG, et al. Circulating angiopoietin-2 levels in the course of septic shock: relation with fluid balance, pulmonary dysfunction and mortality. Intensive Care Med 2009;35:1567-74.
25. van der Heijden M, van Nieuw Amerongen GP, Koolwijk P, van Hinsbergh VW, Groeneveld AB. Angiopoietin-2, permeability oedema, occurrence and severity of ALI/ARDS in septic and non-septic critically ill patients. Thorax 2008;63:903-9.
26. Ricciuto DR, dos Santos CC, Hawkes M, Toldt LJ, Conroy AL, Rajwans N, et al. Angiopoietin-1 and angiopoietin-2 as clinically informative prognostic biomarkers of morbidity and mortality in severe sepsis. Crit Care Med 2011;39:702-10.
27. Orfanos SE, Kotanidou A, Glynos C, Athanasiou C, Tsigkos S, Dimopoulou I, et al. Angiopoietin-2 is increased in severe sepsis: correlation with inflammatory mediators. Crit Care Med 2007;35:199-206.
28. Caine GJ, Blann AD, Stonelake PS, Ryan P, Lip GY. Plasma angiopoietin-1, angiopoietin-2 and Tie-2 in breast and prostate cancer: a comparison with VEGF and Flt-1. Eur J Clin Invest 2003;33:883-90.
29. Pugh CW, Ratcliffe PJ. Regulation of angiogenesis by hypoxia: role of the HIF system. Nat Med 2003;9:677-84.
30. Mandriota SJ, Pyke C, Di Sanza C, Quinodoz P, Pittet B, Pepper MS. Hypoxia-inducible angiopoietin-2 expression is mimicked by iodonium compounds and occurs in the rat brain and skin in response to systemic hypoxia and tissue ischemia. Am J Pathol 2000;156:2077-89.
31. Pichiule P, Chavez JC, LaManna JC. Hypoxic regulation of angiopoietin-2 expression in endothelial cells. J Biol Chem 2004;279:12171-80.
32. Bruick RK, McKnight SL. Building better vasculature. Genes Dev 2001;15:2497-502.
33. Semenza GL. Regulation of hypoxia-induced angiogenesis: a chaperone escorts VEGF to the dance. J Clin Invest 2001;108:39-40.
34. Lim HS, Lip GY, Blann AD. Angiopoietin-1 and angiopoietin-2 in diabetes mellitus: relationship to VEGF, glycaemic control, endothelial damage/dysfunction and atherosclerosis. Atherosclerosis 2005;180:113-8.
35. Kanazawa H, Nomura S, Asai K. Roles of angiopoietin-1 and angiopoietin-2 on airway microvascular permeability in asthmatic patients. Chest 2007;131:1035-41.