Impact of phosphorus and luminosity in the propagation, photochemical reactions and quality of *Lippia alba* (Mill.) N.E.Br. seedlings

Impacto del fósforo y luminosidad en la propagación, reacciones fotoquímicas y calidad de plántulas de *Lippia alba* (Mill.) N.E.Br.

ABSTRACT

*Lippia alba* (Mill.) N.E.Br. (lemongrass) is a spice and medicinal plant species, with fledgling studies related to mineral nutrition and luminosity on foliar and vegetative responses. Thus, the aim of this study was to assess the foliar aspects and production of *L. alba* under two light conditions (full sun and 50% shading) and four levels of phosphorus-P (0, 150, 300, and 450 mg kg$^{-1}$). The *L. alba* seedlings presented changes in physiological indices and photochemical responses based on chlorophyll-$a$ fluorescence. The phosphate fertilization helped mitigate the light stress for the synthesis of chlorophylls. The greatest leaf biomass production occurred with addition of P. The principal components analysis explained 74% of the variability, with leaf number, initial fluorescence and specific leaf area in principal component 1 (PC 1) and bud number, survival and leaf area in PC 2. Two groups formed in the cluster analysis, with lower distances between P300 full sun and P450 shading (2.31). The seedlings cultivated under full sun presented a higher survivability and seedling quality. The production of *L. alba* seedlings should be done under full sun with the addition of 450 mg kg$^{-1}$ of phosphorus.

Additional key words: acclimatization; chlorophyll-$a$ fluorescence; medicinal plant; mineral nutrition.

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*Lippia alba* (Mill.) N.E.Br. (lemongrass, Verbenaceae) is a plant of medicinal interest, and tea made from its leaves has a soothing effect (Hohlenwerger *et al.*, 2017). It can also be used as a natural preservative (Machado *et al.*, 2011). The essential oil from its leaves has shown an antifungal activity against *Aspergillus*, *Fusarium*, *Penicillium* and *Trichoderma* (Glamočlija *et al.*, 2011). There are few studies on cultivation practices that must be established during the production of seedlings propagated with stem cuttings.

The abiotic factors that may influence the vegetative propagation of seedlings include light availability in the cultivation environment, which may affect the morphophysiological responses of plants since it acts directly in the process of photochemical and biochemical reactions of photosynthesis (Leal *et al.*, 2015).

However, light intensity may cause damage to the photosynthetic apparatus (Díez *et al.*, 2017; Taiz *et al.*, 2017) by changing the carboxylation speed of Ribulose 1.5 bisphosphate carboxylase/oxygenase, the assimilation of CO₂ and, consequently, the production of photoassimilates for seedling formation. The plants have foliar functional strategies based on chlorophyll-α fluorescence in photosystem II (PS II), enabling the efficient use of light (Jaimez *et al.*, 2018; Jiménez-Suancha *et al.*, 2015). Therefore, these photochemical parameters may be used in assessing the integrity of photosynthetic machinery under eco-physiological disorders as a result of light stress.

In addition to light, another limiting factor for plants is soil in tropical regions, such as soil in Cerrado, which is highly weathered, has high levels of iron and aluminum oxides, and has high acidity (Souza *et al.*, 2013; Islas-Espinoza *et al.*, 2014), limiting the availability of some nutrients, particularly phosphorus (P). Greater attention is required for the low availability in soil in the tropics because of the greater fixation in soil (Quesada *et al.*, 2011; Gérard *et al.*, 2016).

Generally, P-deficient plants may suffer damage in diffusive and non-diffusive processes of photosynthetic metabolic pathways through a reduction in consumption and regeneration of Rubisco and production of ATP and NADPH (Andrade *et al.*, 2018), which may cause instability in the photochemical process in PS II. Thus, phosphate fertilization is an important agronomic practice since it may assist in photochemical stability (Carstensen *et al.*, 2018) and biomass increases in plants (Kuwahara *et al.*, 2016).

We tested the hypothesis that P may contribute to the production and quality of seedlings, mitigate damage and increase photochemical reactions in PS II under light stress. This study associated the mineral

**INTRODUCTION**

*Lippia alba* (Mill.) N.E.Br. (hierba de limón) es una especie de interés medicinal, con estudios incipientes en relación a la nutrición mineral y a la luminosidad en las respuestas foliares y vegetativas. El objetivo del trabajo fue evaluar los aspectos foliares y la producción de plántulas de *L. alba* bajo dos condiciones de luminosidad (pleno sol y 50% de sombreado) y cuatro niveles de fósforo (0, 150, 300 y 450 mg kg⁻¹). Las plántulas presentaron alteraciones en los índices fisiológicos y respuestas fotoquímicas basadas en la fluorescencia de la clorofila-α. La fertilización fosfatada contribuyó en la mitigación del estrés luminoso en la síntesis de clorofílas. La mayor producción de biomasa discuelas de hojas ocurrió con la adición de P. El análisis de componentes principales explicó el 74% de la variabilidad donde fueron características representativas el número de hojas, la fluorescencia inicial y el área foliar específica dentro del componente principal 1 (PC 1) y el número de brotes, supervivencia y el área foliar en el PC 2. En el análisis de conglomerados, se constató la formación de dos grupos, con distancias más bajas entre 300 pleno sol y 450 sombreado (2.31). Las plántulas cultivadas a pleno sol presentaron mayor capacidad de supervivencia y calidad. La producción de plántulas de *L. alba* debe ser realizada bajo pleno sol con adición de 450 mg kg⁻¹ del fósforo.

**Palabras clave adicionales:** aclimatización; fluorescencia de la clorofila-α; planta medicinal; nutrición mineral.
nutrition and the ecophysiology of the plants in order to assess the photochemical and vegetative response and quality of *L. alba* seedlings produced with stem cuttings under different light availability and phosphate fertilization.

**MATERIAL AND METHODS**

**Collection of plant material and preparation of cuttings**

This species was identified and a voucher was deposited at the Herbarium DDMS of the Federal University of Grande Dourados (UFGD), under number 5226. The collection of plant material was performed from plant matrices in the Horto de Plantas Medicinais (22º11'43.7"S and 54º56'08.5"W, 452 m a.s.l.) of UFGD, in good phytosanitary conditions. The cuttings were removed from the median portion of the branches, standardized as 20 cm in length, mean diameter of 2.45 mm, and a pair of leaves at the apex; 1/3 of the cuttings was buried in the substrate.

**Studied Factors and experiment design**

This experiment was performed from October to December of 2017 in the Faculty Agrarian Science (22º11'43.7"S e 54º56'08.5"W, 452 m a.s.l.) of UFGD in Dourados, Mato Grosso do Sul, Brazil. The climate of the region is classified as Am (Alvares et al., 2013) with an annual average rainfall over 1,500 mm. The factors consisted of two light conditions (full sun and 50% shading) and four levels of phosphorus in the form of simple superphosphate - 18% of P₂O₅ (0, 150, 300 and 450 mg kg⁻¹). The treatments were displayed in a 2×4 factorial scheme, in randomized blocks, with four replicates. The experiment unit consisted of five 500 mL plastic containers, with one cutting each. Shading was simulated using polypropylene black screen with 50% retention of light. The soil was classified as Dystroferric Red Latosol (Santos et al., 2018), clay texture, with the following chemical attributes: pH CaCl₂ = 4.76; P= 0.5 mg dm⁻³; Ca = 1.04 cmol dm⁻³; K = 0.06 cmol dm⁻³; Mg = 0.12 cmol dm⁻³; Al = 1.2 cmol dm⁻³; H+Al = 7.71 cmol dm⁻³; sum of bases = 1.22 cmol dm⁻³; cation exchange capacity = 8.93 cmol dm⁻³ and base saturation = 13.68. No soil correction was performed. Base fertilization was performed in coverage with ammonium sulphate (20% N) and potassium chloride (60% K₂O). The cultivation practices consisted of daily irrigation at 70% of the substrate’s field capacity.

**Chlorophyll-a fluorescence**

At 30 and 60 days after burial (DAB) of cutting, the emission of chlorophyll-a fluorescence was assessed by subjecting the leaves to darkness for 30 min, using leaf clips, between 8:00 and 10:00 am. The initial (F₀) and maximum (Fₘ) chlorophyll-a fluorescence and photochemical efficiency of photosystem II (Fᵥ/Fₘ) were measured under flash of 1,500 µmol m⁻² s⁻¹ with a portable fluorometer (OS-30P; Opti-Sciences Chlorophyll Fluorometer, Hudson, NY). The variables fluorescence (Fᵦ = Fₘ−F₀), efficiency of absorbed energy conversion (Fᵥ/F₀), non-photochemical maximum yield (F₀/Fᵥ) and electron transport rate (ETR) were estimated (Baker, 2008).

**Quantification of photosynthetic pigments**

At 60 DAB, fully expanded leaves were collected, with 1 g macerated in 8 mL of acetone (80%). Afterwards, the samples were centrifuged for 10 min at 3000 rpm, and the absorbance reading was taken at the wavelengths of 470, 645 and 663 nm using a spectrophotometer. The contents of chlorophylls *a*, *b*, total (*a* + *b*) and carotenoids were estimated (Arnon, 1949; Lichtenthaler and Buschmann, 2001).

**Growth indicators**

After 65 DAB, the survival, length of the aerial part (distance from the collet to the inflection of the highest leaf), collar diameter - CD (± 1.0 cm above the substrate level), leaf thickness, and number of buds and leaves were recorded. Seedlings were removed from the containers, washed and assessed for length of the largest root and rooted cuttings (emission of adventitious root of at least 5.0 cm). The leaf area was also assessed (LA) using an area integrator (area meter LI-COR 3100 C; Licor, Lincoln, NE).

**Biomass, physiological and quality indices**

Fresh material from the aerial and root parts were submitted to forced air circulation in an oven at 60±5°C to obtain the dry mass. Using the data for the LA and leaves and total dry biomass (LDM and
TDM, respectively), the physiological indices of leaf area ratio (LAR = LA/LDM), specific leaf area and mass (SLA = LA/TDM and SFM = LDM/TDM, respectively) were calculated (Benincasa, 2003). From the data for total dry biomass, height/diameter ratio (RHD) and aerial part/root ratio (APRR), the Dickson quality index (DQI) was estimated (Dickson et al., 1960).

**Statistical analysis**

The rooting and survival data were transformed into arcsine of $\sqrt{x} + 0.5$ and subjected to normal distribution with the Shapiro-Wilk test for normalization. The data for chlorophyll-a fluorescence were assessed in plots subdivided in time. All data were subjected to analysis of variance (ANOVA), and, when significant according to the F test, the averages were compared with Student’s t test for the luminosity and evaluation periods, along with regression analysis for phosphorus ($P \leq 0.05$), using SISVAR.

Additional multivariate analysis of principal components was carried out with variance and co-variance arrays. A cluster analysis was also performed using the complete linkage method to describe the similarity between the factors, and the grouping was performed with the classical method using Euclidean distances.

**RESULTS**

The greatest seedling survival and collet diameter were observed under full sun (Fig. 1A and B). The greatest height/diameter ratio (RHD) occurred in the shaded seedlings (Fig. 1C). The leaf number (LN) was influenced by the factors independently, with higher amounts under the shaded environment (Fig. 1D), without adjustment to phosphate fertilization (Tab. 1).

The characteristics height, rooting, length of the largest root, stem fresh and dry mass, root fresh mass and total mass were not influenced by the factors ($P > 0.05$), with averages of 22.4 cm, 76%, 12.0 cm, 2.22, 1.04, 0.96 and 6.30 g/plant, respectively. The number of buds, chlorophyll b and fresh and dry mass of the leaves were influenced only by phosphorus, in

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**Figure 1.** Survival (A), collar diameter (B), height/diameter ratio – RHD (C) and leaf number – LN (D) in *L. alba* seedlings produced under light ambience. * significant difference according to the Student’s t test ($P \leq 0.05$).
which the data for LN, NB and chlorophyll b had no adjustment to the mathematical models (Tab. 1) because of the low value of the determination coefficient ($R^2$).

Chlorophyll a and total showed the same trend in the shaded environment, reducing the levels with increasing doses, with maximum levels (17.62 and 28.19 µg cm$^{-2}$) and minimum levels (7.74 and 10.21 µg cm$^{-2}$) without and with (450 mg kg$^{-1}$) P, respectively (Tab. 2). On the other hand, under full sun, chlorophyll a and carotenoids had higher contents (9.96 and 1.84 µg cm$^{-2}$, respectively) with the addition of 450 mg kg$^{-1}$ of P.

Under the shaded environment, the data for $F_v/F_m$ and $F_0/F_v$ had no adjustment to the mathematical models (Tab. 2). Under full sun, the highest ratio was 0.750 and 0.338 electrons quantum$^{-1}$, under 450 mg kg$^{-1}$ of P, respectively. The higher $F_v/F_m$ values were observed at 60 DAB in full sun as a result of the lowest non-photochemical yield ($F_0/F_v$) (Tab. 3). The greatest $F_v$ occurred in the leaves of shaded seedlings (0.162 electron quantum$^{-1}$) (Fig. 2A) and at 30 DAB (0.187 electrons quantum$^{-1}$) (Fig. 2B).

The maximum $F_v$ 0.415 electron quantum$^{-1}$ was found with the addition of 27 mg kg$^{-1}$ of P (Tab. 1). The shaded seedlings had higher $F_v$ and $F_m$ values at 30 DAB, with a reduction of values at 60 DAB in both environments (Tab. 4). The ETR had lower values at 30 DAB (Tab. 4). The greatest ETR occurred in the leaves of seedlings cultivated under full sun at 60 DAB as a result of the higher incident light.

In the shaded environment, the largest LA was 16.43 cm$^2$ under 450 mg kg$^{-1}$ of P. Under full sun, the maximum leaf area (11.85 cm$^2$) occurred in seedlings cultivated with 228.31 mg kg$^{-1}$ of P (Tab. 5). The leaf

| Characteristic | Equation | $R^2$ |
|---------------|----------|------|
| Leaf number (LN) | $\hat{y} = 7.18$ leaves/plant | W/adjust |
| Budding number (BN) | $\hat{y} = 6.34$ budding/plant | W/adjust |
| Chlorophyll b | $\hat{y} = 9.31$ µg cm$^{-2}$ | W/adjust |
| Variable fluorescence | $\hat{y} = 0.40832 + 0.00054*P - 0.00001*P^2$ | 0.98 |
| Leaf fresh mass | $\hat{y} = 0.1564 + 0.0013*P - 0.00002*P^2$ | 0.98 |
| Leaf dry mass | $\hat{y} = 0.14551 + 0.00079*P$ | 0.93 |

* significant difference ($P \leq 0.05$).

| Characteristic | Equation | $R^2$ |
|---------------|----------|------|
| Chlorophyll a ($\mu g$ cm$^{-2}$) | Full sun | Shading |
| $\hat{y} = 5.57350 + 0.00975*P$ | $\hat{y} = 17.64521 - 0.02201*P$ | 0.56 | 0.90 |
| Total chlorophyll ($a+b$) ($\mu g$ cm$^{-2}$) | Full sun | Shading |
| $\hat{y} = 16.73$ W/ adjust | $\hat{y} = 33.33526 - 0.05138*P$ | 0.89 |
| Carotenoids ($\mu g$ cm$^{-2}$) | Full sun | Shading |
| $\hat{y} = 0.58725 + 0.00279*P$ | $\hat{y} = 1.51$ W/ adjust | 0.75 |
| Photochemical efficiency of the photosystem II ($F_v/F_m$) | Full sun | Shading |
| $\hat{y} = 0.71932 + 0.00007*P$ | $\hat{y} = 0.749$ W/ adjust | 0.57 |
| $F_0/F_v$ (electrons quantum$^{-1}$) | Full sun | Shading |
| $\hat{y} = 0.39187 - 0.00013*P$ | $\hat{y} = 0.337$ W/ adjust | 0.50 |
| $F_v/F_0$ (electrons quantum$^{-1}$) | Full sun | Shading |
| $\hat{y} = 2.0480 + 0.00192*P$ | $\hat{y} = 2.958$ W/ adjust | 0.80 |
| $\hat{y} = 3.642$ W/ adjust | $\hat{y} = 3.261$ W/ adjust | 60 DAB |

* significant difference ($P \leq 0.05$).
Table 3. Photochemical efficiency of the photosystem II (Fv/Fm) and maximum non-photochemical yield (F0/Fv) in L. alba seedlings produced with phosphorus under light environment, at 30 and 60 days after burial (DAB).

| Environment   | Fv/Fm 30 DAB | Fm 30 DAB | Fv/Fm 60 DAB | Fm 60 DAB |
|---------------|-------------|----------|-------------|----------|
| Full sun      | 0.699 aB    | 0.611 aB | 0.774 aA    | 0.503 bA |
| Shading       | 0.740 aA    | 0.768 aA | 0.759 aA    | 0.543 bA |
| C.V. (%)      | 7.26        | 8.33     | 11.37       | 10.22    |

Means followed by the same lower case letter in the rows, for day after burial, and upper case in the columns, for light environment, do not differ according to the Student’s t test (P ≤ 0.05).

Figure 2. Initial fluorescence – F0 in leaves of L. alba seedlings produced under phosphorus rates and light environment (a), at 30 and 60 days after burial (b). *significant difference according to the Student’s t test (P ≤ 0.05).

Table 4. Maximum (Fm) and variable (Fv) fluorescence and electron transport rate (ETR) in leaves of L. alba seedlings produced with phosphorus under light environment at 30 and 60 days after burial (DAB).

| Environment   | Fm 30 DAB | Fv 30 DAB | Fm 60 DAB | Fv 60 DAB | ETR 30 DAB | ETR 60 DAB |
|---------------|----------|----------|----------|----------|------------|------------|
| Full sun      | 0.611 aB | 0.433 aB | 0.503 bA | 0.395 aA | 248.38 bA  | 275.29 aA  |
| Shading       | 0.768 aA | 0.572 aA | 0.543 bA | 0.415 bA | 107.94 aB  | 110.78 aB  |
| C.V. (%)      | 8.33     | 10.22    | 8.33     | 10.22    | 6.12       | 6.12       |

Means followed by different lower case letters in a row, for day after burial, and upper case letters in the columns, for light environment, indicate significant differences according to the Student’s t test (P ≤ 0.05).

thickness data were not adjusted, and the sun leaves had higher values. The data for root dry mass and APRR of the seedlings under full sun were not adjusted to the mathematical models, but, under the shading, the higher mean values were 0.462 g/plant and 0.44 with 450 mg kg⁻¹ of P. The fresh and dry biomass of the leaves were influenced only by phosphorus, with higher values (0.177 and 0.0501 g/plant) with the addition of 32.5 and 450 mg kg⁻¹ of P, respectively (Tab. 1).
The greatest LAR (2.027 cm² g⁻¹) and SLM (0.0458 g cm⁻²) occurred in the shaded seedlings (Fig. 3A and B) as a result of the greatest leaf thickness (Tab. 5). The greatest SLA was 48.54 cm² g⁻¹ in the seedlings produced in the shaded environment (Fig. 3C). For the DQI, the greatest value (1.70) was observed in the seedlings cultivated under full sun (Fig. 3D), regardless of phosphorus.

The principal components analysis explained 74% of the variability, in which PC1 and PC2 contributed 41.05% and 32.05%, respectively, of the remaining variance of the characteristics in the L. alba seedlings (Fig. 4A). In the cluster analysis, two groups were formed for the P dose (G1 and G2), with six subgroups (Fig. 4B). The subgroups with lower distances between P dose and light environment (S - shading and PS - full sun) were 300 PS and 450 S (2.31), followed by 450 S and 450 PS (4.61) and 300 S and 0 S (6.65).

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DISCUSSION

The greatest survival was associated with the amount of photoassimilates as a function of greater incident light. Seedlings under excessive light lose more water through transpiration because of an increased temperature, with the greater diameter allowing a greater ability to transport water and photoassimilates to other organs in order to maintain turgidity (Taiz et al., 2017). The greatest RHD occurred in the shaded seedlings, but the values did not indicate that there was etiolation in the seedlings under limited light.

Generally, plants under shading tend to expand the number of leaf limbs, enabling greater light interception (Oliveira et al., 2017) and maximizing photosynthesis. These characteristics are important in the vegetative propagation of the species since the greater the amount of buds, the greater the formation of leaves, contributing to photoassimilate distribution to developing organs.
The reduction of pigments, except carotenoids, in leaves under full sun can be explained by photoinhibition of chlorophylls as a result of excessive light (Fu et al., 2012). However, shaded leaves tend to increase the content of chlorophyll and, thereby, maximize the photosynthetic capacity (Díez et al., 2017). Similar results were observed by Soares et al. (2017) when assessing young Tamarindus indica L. plants under shading, which presented higher concentrations of chlorophylls.

An increase in chlorophyll content is desirable because it is responsible for capturing light photons and transmitting energy to reaction centers (Monteiro et al., 2018), and carotenoids are responsible for chlorophyll photoprotection and reduction of membrane harm (Taiz et al., 2017), maximizing the photosynthetic capacity. Under full sun cultivation, phosphate fertilization can be a potential mitigation strategy for ecophysiological disorders from light stress in the synthesis of chlorophylls.

P, in the form of Pi, is responsible for the control of enzymatic reactions and metabolic regulation in the cytosol and chloroplast (Hawkesford et al., 2012). Higher photochemical indicators, especially the Fv/Fm ratio, with phosphorus result because P favors the speed of ATP synthesis, contributing to the export of protons to the chloroplast stroma and thylakoid lumen (Carstensen et al., 2018), favoring the maintenance of electron mobility in PS II. Increasing Fv/Fm can be a photoprotective mechanism of the photosynthetic apparatus for excessive incident radiation (Blind et al., 2018), i.e. a greater use of electrons produced as a result of higher efficiency in absorbed energy conversion (Fv/Fm), regardless of the shading level during this period (60 DAB) (Tab. 2).

The higher Fv at 30 DAB is desirable in order to mitigate damage in the photosynthetic apparatus (Fig. 2B). Fu et al. (2012) described a higher Fv in Lactuca sativa L. plants cultivated under 100 µmol m-2s-1 (low irradiance) at the initial stage of the experiment, stating that this mechanism can mitigate an increase in reactive oxygen and D1 protein degradation. As for phosphorus in Fv, P is involved in phosphorylation reactions and pyrophosphate release, acting in the activation of catalytic enzymes, forming ATP (Hawkesford et al., 2012, Xing and Wu 2014; Andrade et al., 2018) and favoring photochemical stability because Fv is related to the ability to transfer electrons.

The results for Fm and ETR is important in the photochemical process since larger values favor the flow of chlorophyll molecules between acceptors in photosystems (Farias et al., 2016). The higher photochemical efficiency in PS II effectively contributes to maintaining the integrity of the photosynthetic apparatus and increasing vegetative characteristics.

Plants exposed to low luminosity enhance the expansion of leaves (Gondim et al., 2018) as a strategy for use of light. Increasing APRR and RDM may be associated with the fact that under high irradiance, substrate and leaves tend to lose more water through evapotranspiration and leaf transpiration. Thus, an increase in these characteristics under full sun promotes water absorption for the maintenance of metabolic processes and nutrient transport (Sarto et al., 2018).

The responses of plants to abiotic variants can change according to the species. Oliveira et al. (2017), when assessing the physiological and productive aspects of Origanum vulgare L. plants, found greater root biomass under shaded cultivation. On the other hand, there was greater biomass allocation in Physalis minima L. seedlings under full sun (Silva et al. 2016), similar to L. alba seedlings (Fig. 3).

P plays an important role in plant biomass allocation and is a structural component of nucleic acids, phospholipids and ATP formation, favoring primary metabolism reactions and constituting ~ 0.2% of plant mass (Kuwahara et al., 2016). For seedling production in Dalbergia nigra Benth., it was found that the addition of 500 mg kg-1 of P favored an increase in biomass (Carlos et al., 2018), similar to the L. alba seedlings in this study (Tab. 1).

LAR and SLM show greater biomass allocation through leaf area with greater thickness (Oliveira et al., 2016). These authors found that Melissa officinalis L. plants, at 120 d after transplanting, showed a greater SLM when cultivated under full sun. For LAR, Ribeiro et al. (2018) found higher values in Pogostemon cablin cultivated under shaded conditions. The increase in SLA under this condition indicated the adaptive ability of leaf tissues in optimizing light capture (Guzmán et al., 2016; Liu et al., 2016) because it promotes CO2 assimilation and stomatal control (Gommers et al., 2013). However, leaves with a smaller thickness are less heavy. Similar results were observed in Enterolobium contortisiliquum (Vell.) Morong
(Souza et al., 2017) and *Colocasia esculenta* L. Schott (Gondim et al., 2018). Both authors described higher values in plants under artificial shading.

The DQI demonstrated that this species has survivability and stability capacity when exposed to high irradiance, an important fact since the initial cultivation under this condition reduced the acclimatization period (rustification) of the seedlings, i.e. these will be less susceptible to weather under field conditions, such as excessive sunlight. The use of DQI has been constant when assessing seedling quality since it is an easy implementation analysis, performed by calculating the morphometric stability level, distribution and biomass allocation.

In PC 1, the characteristics that showed positive scores in descending order were RHD (0.271), LN (0.263), \( F_c \) and SFM (0.255), and the characteristics with negative scores were APRR (-0.292), ETR and RDM (-0.288) and SLA (-0.270), which were the most similar. In PC 2, the components that most contributed with positive factorial scores were the number of buds (0.352), survival (0.330), leaf area (0.319) and \( F_v \) (0.289), with no negative absolute scores > 0.30. The cluster analysis consisted of sample classification in order to verify the similarity within the groups and between-group heterogeneity, considering all evaluated characteristics (Araújo et al., 2013). Thus, there was greater heterogeneity between the luminous environments, with G1 comprising the association of P with shading, except for 450 S and G2 under full sun.

**CONCLUSION**

The *Lippia alba* seedlings responded positively to the phosphate fertilization when vegetative propagation was used with stem cuttings. The association of 450 mg of phosphorus with the cultivation under full sun contributed substantially to mitigating ecophysiological disorders in the photosynthetic apparatus as the result of light stress, providing greater photochemical stability, survival and quality in the *Lippia alba* seedlings. No acclimatization process was required.

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