Evaluation of the Anticonvulsant and Anxiolytic Potentials of Methyl Jasmonate in Mice

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Abstract
Methyl jasmonate (MJ) is one of the most well-studied plant stress hormones belonging to the jasmonate family. Previous studies have shown that MJ potentiated pentobarbitone sleeping time and enhanced GABA-mediated inhibitory neurotransmission, suggesting potential benefits in disorders associated with hyperactivity of the brain. This study was carried out to evaluate whether MJ has anticonvulsant and anxiolytic properties in mice. The anticonvulsant effect was assessed based on the prevention of tonic-clonic seizures induced by chemoconvulsant agents in mice. The anxiolytic property was evaluated utilizing the elevated plus maze (EPM) and light/dark transition paradigms. The effect of MJ on spontaneous locomotor activity (SMA) was also assessed. Mice received intraperitoneal (i.p.) injections of MJ 30 min before the tests were carried out and diazepam (2 mg/kg, i.p.) was used as the reference drug. MJ (50–400 mg/kg) did not protect the mice against tonic-clonic convulsions induced by picrotoxin (10 mg/kg, i.p.) or strychnine (3 mg/kg, i.p.). However, MJ (100, 200, and 400 mg/kg) offered 20, 60, and 100% protection against pentylenetetrazole (100 mg/kg, i.p.)-induced convulsions. In a similar manner to diazepam (2 mg/kg), MJ (400 mg/kg) produced a marked sedative effect as shown by decreases in the number of lines crossed and the duration of ambulation in the open field test. In contrast to diazepam (2 mg/kg), MJ (5–50 mg/kg) did not show anxiolytic effects in the EPM and light/dark transition paradigms. These findings suggest that methyl jasmonate at high doses possessed anticonvulsant properties in the pentylenetetrazole animal model of epilepsy, but did not produce anxiolytic activity in mice.
Keywords
Methyl jasmonate • Picrotoxin • Pentylenetetrazole • Strychnine • Convulsions • Anxiolytic

Introduction

Convulsive seizures and anxiety are central nervous system disorders associated with an imbalance between excitatory and inhibitory mechanisms of the brain and impose untold suffering on patients [1–3]. Convulsion manifests itself in forms of recurrent seizures, whereas anxiety is related to sustained anxious feelings in response to a perceived life threatening situation [1, 2, 4]. Anxiety shares common neurochemical pathways with convulsions such as reduced GABAergic and increased glutaminergic neurotransmissions [2, 4]. Although large proportions of people suffer from these ailments worldwide, the development of new pharmacological interventions has lagged behind in comparison with many other therapeutic counterparts [4]. The standard anticonvulsant and anxiolytic drugs have a limited spectrum of activities with adverse effects that have limited their clinical usefulness and compromised patients’ compliance [2, 4]. Thus, there is need to search for newer agents with better clinical profiles for the treatment of convulsions and relief of anxiety symptoms.

Methyl jasmonate (MJ) is a cyclopentanoic compound secreted by plants in response to external stressors [5]. MJ was first isolated from the essential oil of Jasminum grandiflorum, but is now obtained through chemical synthesis [6]. Previous investigations have shown that MJ has anticancer, antinociceptive, antiaggressive, and antidepressant properties [5, 7, 8]. In addition, Hossain et al. [9] reported that MJ potentiated pentobarbitone sleeping time and enhanced GABA-mediated inhibitory neurotransmission, suggesting the potential benefits in disorders associated with hyperexcitability of the brain such as convulsions and anxiety states. Thus, this present study was carried out to evaluate whether MJ exhibits anticonvulsant and anxiolytic activities in mice.

Experimental

Laboratory Animals

Albino Swiss mice of either sex (18–22 g) were obtained from the Animal Centre, University of Ibadan, Nigeria and were kept in plastic cages at room temperature. They were fed with balanced rodent pellet diet and water ad libitum and were acclimatized for one week before using them for experiments. The procedures used in the study were in compliance with the ethical guidelines of the University of Ibadan and National Institutes of Health guide for the Care and Use of Laboratory Animals for experimental investigations.

Drugs and Treatment

The following drugs were used: picrotoxin (Sigma-Aldrich, St. Louis, USA), pentylenetetrazole, PTZ (Sigma-Aldrich, St. Louis, USA), strychnine (Sigma, USA), Diazepam, DPZ (Sigma, USA), and methyl jasmonate, MJ (Sigma-Aldrich Chemie GmbH, Steinheim, Germany). Picrotoxin, pentylenetetrazole, and strychnine were dissolved in distilled water. One hundred milligrams of MJ were dissolved in 1 ml of ethanol (95%) to give 100 mg/ml stock solution. This stock solution was further diluted with distilled water to obtain the
different concentrations used in the study. The final concentration of ethanol in each solution used for the study did not exceed 1%.

**Effect of Methyl Jasmonate on Picrotoxin-Induced Convulsions**

The effect of MJ on picrotoxin-induced convulsions was evaluated according to the method described by Loscher [10]. Mice (n = 6) were treated with intraperitoneal (i.p.) injections of MJ (50–400 mg/kg), DPZ (2 mg/kg), or a vehicle (10 ml/kg of 1.0% ethanol) 30 min prior to the induction of convulsions with i.p. injections of picrotoxin (10 mg/kg). Thereafter, the animals were placed individually in a transparent chamber and the latency to tonic-clonic seizures as well as the mortality rate was assessed for the period of 30 min after the administration of picrotoxin [10].

**Effect of Methyl Jasmonate on Convulsions Induced by Pentylentetrazole**

The PTZ animal model of epilepsy was further used to evaluate the anticonvulsant property of MJ following the procedure previously described by [11]. Mice were treated with i.p. injections of MJ (50–400 mg/kg), DPZ (2 mg/kg), or a vehicle (10 ml/kg of 1.0% ethanol) 30 min before the induction of seizures with i.p. injections of PTZ (100 mg/kg). The animals were then observed for the appearance of tonic-clonic seizures and death as earlier described [11].

**Effect of Methyl Jasmonate on Convulsions Induced by Strychnine**

The effect of MJ on seizures induced by strychnine was also evaluated in this study as described above. Mice were treated with i.p. injections of MJ (50–400 mg/kg), DPZ (2 mg/kg), or a vehicle (10 ml/kg of 1.0% ethanol) 30 min prior to the induction of convulsions with strychnine (3 mg/kg, i.p.) and then monitored for the appearance of tonic-clonic seizures and death.

**Performance of Methyl Jasmonate in the Elevated Plus-Maze Test**

The EPM animal model of anxiety was used to evaluate the anxiolytic property of MJ according to the procedure previously described [12]. Mice were randomly distributed into six treatment groups (n = 5). The first four groups received MJ (5, 10, 20, and 50 mg/kg), whereas the fifth group was treated with DPZ (2 mg/kg). The last group received a vehicle (10 ml/kg of 1.0% ethanol), which served as a negative control. Thirty minutes later, each mouse was placed at the centre of the maze, facing an open arm and allowed to explore it for 5 min. The number of entries and time spent (s) in each arm of the EPM were recorded for the period of 5 min.

**Performance of Methyl Jasmonate in the Light/Dark Transition Test**

The light/dark transition test was further employed to assess the anxiolytic property of MJ according to the method of [13]. Mice (five per group) were treated with MJ (5, 10, 20, and 50 mg/kg, i.p.), DPZ (2 mg/kg, i.p.), or a vehicle (10 ml/kg of 1.0% of ethanol, i.p.). Thirty minutes after treatment, each mouse was placed in the illuminated compartment of the box. Thereafter, the number of entries and time spent in the light and dark compartments of the box were measured for the 5 min session [14].
Effect of Methyl Jasmonate on Spontaneous Locomotor Activity in the Open Field Test

The open field test as a measure of spontaneous locomotor activity was utilized to evaluate the sedative property of MJ following the procedure described by Bruhwiler et al. [14]. Mice (five per group) received i.p. injections of MJ (5–400 mg/kg), DPZ (2 mg/kg, i.p.), or a vehicle (10 ml/kg of 1.0 % of ethanol) and were placed individually at the centre of the box 30 min later. Thereafter, the number of line crosses and duration of ambulation were measured for the 5 min period.

Statistical Analysis

The data were expressed as the mean ± S.E.M. The data were analyzed with Graph Pad Prism software version 4.03. Statistical analysis of the data was done by One-way ANOVA, followed by Newman-Keuls multiple comparison test. P-values less than 0.05 were considered statistically significant.

Results

Effect of Methyl Jasmonate on Picrotoxin-Induced Convulsions

Mice treated with the vehicle exhibited tonic-clonic convulsions with a latency of 5.56±0.13 seconds after the i.p. injection of picrotoxin (10 mg/kg, i.p.) and all of the animals died after seizures. One-way ANOVA revealed that there were significant differences between the treatment groups: latency to convulsions [F(5, 24) = 19.16, P = 0.0097] and latency to death [F(5, 24) = 20.73, P <0.0001]. Post-hoc analysis by the Newman-Keuls multiple comparison test showed that MJ (200-400 mg/kg) as well as DPZ (2 m/kg) delayed the latency to the tonic-clonic components of picrotoxin-induced seizures in mice (Table 1). In addition, MJ (200–400 mg/kg) as well as DPZ only delayed the latency to death, as all of the animals eventually died (Table 1).

Table 1. Effect of methyl jasmonate on picrotoxin-induced convulsions

| Treatment Group | Dose (mg/kg) | Latency to Convulsion (min) | Latency to Death (min) | Convulsion (%) / Death (%) |
|-----------------|--------------|-----------------------------|------------------------|---------------------------|
| Control         | –            | 5.56±0.13                   | 10.6±0.30              | 100 / 100                 |
| MJ              | 50           | 6.04±0.55                   | 11.2±0.26              | 100 / 100                 |
| MJ              | 100          | 6.11±0.41                   | 11.3±0.31              | 100 / 100                 |
| MJ              | 200          | 6.26±0.29                   | 11.4±0.51              | 100 / 100                 |
| MJ              | 400          | 9.56±0.64*                  | 19.6±0.55*             | 100 / 100                 |
| DZP             | 2            | 11.20±0.85*                 | 17.8±0.56              | 100 / 100                 |

Values represent the Mean ± S.E.M for 5 animals per group.
*P<0.05 compared to control (ANOVA followed by the Newman-Keuls multiple comparison Test. Latency to convulsion [F (5, 24) = 19.16, P = 0.0097] and latency to death [F(5, 24) = 20.73, P<0.0001].

Effect of Methyl Jasmonate on Pentylentetrazole-Induced Convulsions

The effects of MJ (50–400 mg/kg, i.p) on convulsions induced by PTZ (100 mg/kg, i.p) in mice are shown in Table 2. One-way ANOVA showed that there were significant
differences between the treatment groups: latency to convulsions \( [F_{(5, 24)} = 31.03, P<0.0001] \) and latency to death \( [F_{(5, 24)} = 180.1, P<0.0001] \). Post-hoc test analysis by the Newman-Keuls multiple comparison test showed that MJ (100–400 mg/kg) increased the latency to the tonic-clonic components of PTZ-induced seizures in mice (Table 2). At these doses, MJ reduced the mortality rate, whereas the death rate was 100% in the control group (Table 2). The reference drug, diazepam (2 mg/kg) offered 100% protection against convulsions, as none of the animals exhibited tonic-clonic seizures induced by PTZ in mice (Table 2).

**Tab. 2.** Effect of methyl jasmonate on pentylenetetrazole-induced convulsions

| Treatment | Dose (mg/kg) | Latency to Convulsion (min) | Latency to Death (min) | Convulsion (%) / Death (%) |
|-----------|-------------|-----------------------------|------------------------|---------------------------|
| Control   | –           | 0.89±0.14                   | 1.3±0.04               | 100 / 100                 |
| MJ        | 50          | 1.06±0.13                   | 1.6±0.04               | 100 / 100                 |
| MJ        | 100         | 1.36±0.17*                  | 2.7±0.08*              | 80 / 80                   |
| MJ        | 200         | 1.75±0.19*                  | 3.2±0.07*              | 60 / 40                   |
| MJ        | 400         | –                           | –                      | 0 / 0                     |
| DZP       | 2           | –                           | –                      | 0 / 0                     |

Values represent the Mean ± S.E.M for 5 animals per group. *P<0.05 compared to control (ANOVA followed by the Newman-Keuls multiple comparison test). Latency to Convulsion \( [F_{(5, 24)} = 31.03, P<0.0001] \) and latency to death \( [F_{(5, 24)} = 180.1, P<0.0001] \).

**Effect of Methyl Jasmonate on Strychnine-Induced Convulsions**

One-way ANOVA showed that there were no significant differences between the treatment groups: latency to convulsions \( [F_{(5, 24)} = 3.921, P = 0.0097] \) and latency to death \( [F_{(5, 24)} = 130.9, P<0.0001] \). Post-hoc analysis by the Newman-Keuls multiple comparison test revealed that MJ (50–400 mg/kg) did not significantly \( (p > 0.05) \) prolong the latency to convulsions produced by strychnine in mice (Table 3). In addition, it did not offer any protection against convulsions, as all of the animals experienced tonic-clonic seizures. As shown in Table 3, a similar pattern of effects were observed in the group given DPZ (2 mg/kg, i.p.).

**Tab. 3.** Effect of methyl jasmonate on strychnine-induced convulsions in mice

| Treatment Group | Dose (mg/kg) | Latency to Convulsion (min) | Latency to Death (min) | Convulsion (%) / Death (%) |
|----------------|-------------|-----------------------------|------------------------|---------------------------|
| Control        | –           | 2.36±0.29                   | 2.4±0.02               | 100 / 100                 |
| MJ             | 50          | 2.38±0.17                   | 2.5±0.02               | 100 / 100                 |
| MJ             | 100         | 2.41±0.26                   | 2.6±0.02               | 100 / 100                 |
| MJ             | 200         | 2.44±0.28                   | 2.6±0.04               | 100 / 100                 |
| MJ             | 400         | 3.57±0.58                   | 3.7 ±0.39*             | 100 / 100                 |
| DZP            | 2           | 3.18±0.19                   | 3.2±0.06               | 100 / 100                 |

Values represent the Mean ± S.E.M for 5 animals per group. *P<0.05 compared to control (ANOVA followed by the Newman-Keuls multiple comparison test). Latency to convulsion \( [F_{(5, 24)} = 3.921, P = 0.0097] \) and latency to death \( [F_{(5, 24)} = 130.9, P<0.0001] \).
**Performance of Methyl Jasmonate in Elevated Plus-Maze Paradigm**

The effect of MJ on the explorative behaviors of mice in the EPM test, as assessed by the number of arm entries and time spent in the arms, are shown in figures 1 and 2. One-way ANOVA revealed that there were significant differences between the treatment groups: the number of open arm entries \( F(5, 24) = 10.17, P<0.0001 \) and number of closed arm entries \( F(5, 24) = 26.36, P<0.0001 \); time spent in the open arm \( F(5, 24) = 295.4, P<0.0001 \) and time spent in the closed arm \( F(5, 24) = 122.6, P<0.0001 \). Post-hoc analysis by the Newman-Keuls multiple comparison test revealed that MJ (5, 10, 20, 50 mg/kg, i.p.) did not significantly \((p > 0.05)\) increase the number of open arm entries and duration of time spent in the open arm as compared with the vehicle, which suggests absence of anxiolytic property. In contrast, DPZ (2 mg/kg, i.p.) showed anxiolytic effects as it significantly \((p < 0.05)\) modified all of the components of the EPM paradigm (Fig. 1 and 2).

![Fig. 1. Effect of methyl jasmonate on time spent in closed and open arms in the elevated plus maze. Values represent the Mean ± S.E.M for 5 animals per group. *P<0.05 compared to control (ANOVA followed by the Newman-Keuls multiple comparison test). Time spent in the closed arm \( F(5, 24) = 122.6, P < 0.0001 \); Time spent in the open arm \( F(5, 24) = 295.4, P<0.0001 \).](image)

**Performance of Methyl Jasmonate in the Light/Dark Transition Test**

The effect of MJ on the performance of mice in the light/dark transition test, as measured by the amount of time spent in the light and dark compartments, are shown in Figure 3. One-way ANOVA showed that there were significant differences between the treatment groups: the time spent in the light compartment \( F(5, 24) = 231.2, P < 0.0001 \) and time spent in the dark compartment \( F(5, 24) = 133.8, P<0.0001 \). Post-hoc analysis by the Newman-Keuls multiple comparison test showed that MJ (5, 10, 20, 50 mg/kg) did not significantly \((p > 0.05)\) increase the amount of time spent in the light compartment when compared with the vehicle, which further suggests absence of anxiolytic property in mice (Fig. 3).
However, DPZ (2 mg/kg) significantly ($p < 0.05$) increased the duration of time spent in the light compartment in comparison with the vehicle, indicating anxiolytic effects (Fig. 3).

**Fig. 2.** Effect of methyl jasmonate on the frequency of entries into the closed and open arms in the elevated plus maze. Values represent the Mean ± S.E.M for 5 animals per group. *$P<0.05$ compared to control (ANOVA followed by the Newman-Keuls multiple comparison test). Number of opened arm entries [$F(5, 24) = 10.17, P<0.0001$] and number of closed arm entries [$F(5, 24) = 26.36, P < 0.0001$].

**Fig. 3.** Effect of methyl jasmonate on the time spent in the dark and light compartments in the light and dark box. Values represent the Mean ± S.E.M for 5 animals per group. *$P<0.05$ compared to control (ANOVA followed by the Newman-Keuls multiple comparison test). Time spent in the light compartment [$F(5, 24) = 231.2, P<0.0001$] and time spent in the dark compartment [$F(5, 24) = 133.8, P<0.0001$].
Sedative Effect of Methyl Jasmonate

The effect of MJ on SMA as assessed by the number of line crosses and duration of ambulation in the open field test are presented in Table 4. One-way ANOVA revealed that there were significant differences between the treatment groups: the number of line of crosses [F(9, 40) = 46.59, P<0.0001] and duration of ambulation [F(9, 40) = 43.63, P<0.0001]. Post-hoc analysis by the Newmann-Keuls multiple comparison test revealed that MJ (400 mg/kg) produced marked sedation as shown by the reduced number of line crosses and duration of ambulation in mice (Table 4). However, at lower doses, it did not significantly change the SMA when compared with the control value (Table 4).

Tab. 4. Effect of methyl jasmonate on spontaneous motor activity in mice

| Treatment Group | Dose (mg/kg) | Number of line crossed | Duration of ambulation (s) |
|-----------------|--------------|------------------------|---------------------------|
| Control         | –            | 70.60±4.23             | 80.40±5.20                |
| MJ 5            | 71.40±3.87   |                        | 81.80±5.09                |
| MJ 10           | 73.80±4.12   |                        | 83.40±4.77                |
| MJ 20           | 75.20±2.75   |                        | 85.00±3.87                |
| MJ 50           | 77.80±4.08   |                        | 86.60±6.40                |
| MJ 200          | 65.60±3.59   |                        | 66.60±4.90*               |
| MJ 400          | 2.20±0.37*   |                        | 2.60±0.25*                |
| DZP             | 2            | 33.20±2.42*            | 35.00±2.72*               |

Values represent the Mean ± S.E.M for 5 animals per group. *P<0.05 compared to control (ANOVA followed by the Newman-Keuls multiple comparison test).

Discussion

The results of this study showed that MJ given intraperitoneally offered significant protection against tonic-clonic convulsions induced by PTZ in mice. At the dose of 400 mg/kg, it blocks the action of PTZ, as none of the animals exhibited tonic-clonic convulsions. However, MJ could only delay the onset of seizures, as all of the animals experienced convulsions induced by picrotoxin. In a similar manner to diazepam, the reference drug, MJ did not modify the different components of strychnine-induced seizures. In the open field test, MJ at high doses suppressed spontaneous locomotor activity as shown by decreases in the duration of ambulation and the number of lines crossed. However, in contrast to diazepam, MJ did not exhibit anxiolytic activity at doses used in this study in the EPM and light/dark animal paradigms of anxiety.

Picrotoxin, pentylenetetrazole, and strychnine are widely used to induce convulsions in experimental animals [1, 4, 15]. However, the PTZ animal model has been shown to more closely reflect clonic convulsive seizures seen in humans and is known to be sensitive to the blocking action of anticonvulsant drugs [1, 4]. Pentylenetetrazole induces convulsions by antagonizing gamma aminobutyric acid (GABA) in a competitive manner [15]. The antagonism of the postsynaptic GABA receptor leads to excessive neuronal excitation that terminates in convulsions and death in laboratory animals [1, 15, 16]. Picrotoxin is a non-
competitive antagonist of the GABA receptor chloride ion complex, whereas strychnine causes convulsions through the competitive antagonism of glycine at glycinergic receptors in the spinal cord [16–19]. The major indicator of the anticonvulsant property of a novel compound is based on the prevention of tonic-clonic convulsions especially in the PTZ animal model of epilepsy [1, 4]. Thus, the inhibitory effect shown by MJ against PTZ-induced seizure episodes suggests anticonvulsant activity. In addition, MJ significantly delayed the onset of seizures produced by both PTZ and picrotoxin, which further suggest a beneficial role in retarding the spread of seizures in epileptic brains [20]. Conversely, the failures of MJ and diazepam to modify the convulsant action of strychnine clearly show a lack of effect on the glycinergic system. It is worthy to note that DPZ mediates its anticonvulsant activity through the enhancement of GABA A-mediated inhibitory neurotransmission [1, 10, 21]. This may perhaps explain its efficacy in convulsions induced by PTZ [22].

Generally, drugs which enhanced GABA currents are known to exhibit anticonvulsant activity against PTZ, but might be weak or ineffective against picrotoxin-induced convulsions [4, 21, 23]. The apparent insensitivity of picrotoxin to MJ may be related to the site of its action at the chloride ion gate, which is not readily accessible to most anticonvulsant drugs [18, 23]. Thus, the inability to gain access to the closed chloride ion gate in order to reopen it might explain the failure of MJ to prevent seizures due to picrotoxin. It is our opinion that the enhancement of GABA currents previously reported for MJ [9] might perhaps account for its ability to modify some of the components of convulsions induced by PTZ and picrotoxin in mice. However, further studies using experimental models such as kindling, are necessary to determine if MJ is able to modify the epileptogenesis process.

The EPM and light/dark transition tests are animal models commonly used for elucidating the neurobiological mechanisms of anxiety and for detecting drugs with anxiolytic properties [24]. The behavioral alteration seen in EPM is based on the natural aversion of rodents for open environments. Thus, mice taken from the home cage naturally show a pattern of behavior characterized by open arm avoidance with consistent preference for closed arms [4, 24]. This behavioral trait is suppressed by anxiolytics and enhanced by anxiogenics [4, 24]. Thus, the measures of anxiety in the EPM paradigm are based on the alterations in the number of open arm entries and the total time spent in the open arm [24, 25]. In this model, MJ at the tested doses did not significantly alter the number of open arm entries and time spent in the open arm, suggesting absence of anxiolytic property. Similarly, the natural tendency for rodents is to avoid the light compartment and show preference for the dark region of the light/dark apparatus [26]. MJ also failed to significantly modify the behavioral performance of the animals in the light/dark transition test, which further suggests absence of anxiolytic property. However, gross sedating effect was observed at high doses of MJ in the open field test. Traditionally, the open field test has served as the animal paradigm for assessing pharmacological agents with CNS stimulants or depressant activity [14, 27]. In this test, MJ suppressed SMA at high doses, which suggests sedative or central depressant effect.

**Conclusion**

This study suggests that methyl jasmonate possessed anticonvulsant properties in the PTZ animal model of epilepsy, but did not demonstrate anxiolytic activity in mice.
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Authors’ Statements

Competing Interests
The authors declare no conflict of interest.

Animal Rights
The institutional and international ethical guidelines for the Care and Use of Laboratory Animals were followed. See the experimental parts for details.

References
[1] Rogawski MA, Porter RJ. Antiepileptic drugs: pharmacological mechanisms and clinical efficacy with consideration of promising developmental stage compounds. Pharmacological Rev 1990; 42: 224–247. http://www.ncbi.nlm.nih.gov/pubmed/2217531
[2] McNamara JO. Emerging insight into the genesis of epilepsy. Nature. 1999; 399: 15–22. http://dx.doi.org/10.1038/399a015
[3] Millan MJ. The neurobiology and control of anxious state. Prog Neurobiol 2003; 70: 84–244. http://dx.doi.org/10.1016/S0301-0082(03)00087-X
[4] White HS. Clinical significance of animal seizure models and mechanism of action studies of potential antiepileptic drugs. Epilepsia. 1997; 38: S9–S17. http://dx.doi.org/10.1111/j.1528-1157.1997.tb04523.x
[5] Fingrut O, Flescher E. Plant stress hormones suppress the proliferation and induce apoptosis in human cancer cells. Leukemia. 2002; 16: 608–616. http://dx.doi.org/10.1038/sj.leu.2402419
[6] Demole E, Lederer E, Mercier D. Isolement et determination de la structure du jasmonate de methyl, constituant odoerant caracterisique de l'essence de jassmin. Helv Chim Acta. 1962; 45: 675–685. http://dx.doi.org/10.1002/hlca.19620450233
[7] Umukoro S, Eduviere AT, Aladeokin AC. Anti-aggressive activity of methyl jasmonate and the probable mechanism of its action in mice. Pharmacol Biochem Behav. 2012; 101: 271–277. http://dx.doi.org/10.1016/j.pbb.2011.12.015
[8] Umukoro S, Olugbemide AS. Antinociceptive effects of methyl jasmonate in experimental animals. J Nat Med. 2011; 65: 466–470. http://dx.doi.org/10.1007/s11418-011-0520-3
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[9] Hossain SJ, Aoshima H, Corda H, Kiso Y. Fragrances in oolong tea that enhance the response of GABAA receptors. Biosci Biochnol Biochem 2004; 68: 1242–1248. http://dx.doi.org/10.1271/bbb.68.1842

[10] Löschler W, Hönack D, Fassbender CP, Nolting B. The role of technical, biological and pharmacological factors in the laboratory evaluation of anticonvulsant drugs-pentylentetrazole seizure models. Epilepsy Res. 1991; 8: 171–189. http://dx.doi.org/10.1016/0920-1211(91)90062-K

[11] Loscher W. Comparative assay of anticonvulsant and toxic potencies of sixteen GABAmimetic drugs. Neuropharmacology. 1982; 21: 803–810. http://www.ncbi.nlm.nih.gov/pubmed/7121752

[12] Pellow S, Chopin P, File SE, Briely B. Validation of open-closed arm entries in elevated plus maze as a measure of anxiety in the rat. J Neurosci Meth. 1985; 14: 149–167. http://dx.doi.org/10.1016/0165-0270(85)90031-7

[13] Pellmar TC, Wilson WA. Synaptic mechanism of pentylenetetrazole: Selectivity for chloride conductance. Science. 1977; 197: 912–914. http://dx.doi.org/10.1126/science.887932

[14] Crawley JN. Neuropharmacologic specificity of a simple animal model for the behavioral actions of benzodiazepines. Pharmacol Biochem Behav. 1981; 15: 695–699. http://dx.doi.org/10.1016/0091-3057(81)90007-1

[15] Bruhwylre J, Chleide E, Houbeau G, Mercier M. Stimulant effect of the beta-carboline FG 7142 in the open-field test. Eur J Pharmacol. 1991; 200: 183–185. http://dx.doi.org/10.1016/0014-2999(91)90685-J

[16] Akaike W, Hattori K, Ocmura Y, Carpenter DO. Bicuculline and picrotoxin block gamma-amniobutyric acid-gated chloride ion conductance by different mechanisms. Experiential. 1985; 41: 70–71. http://dx.doi.org/10.1007/BF02005880

[17] Taylor P, Insel PA. Molecular basis of drug action. In: Pratti WB, Taylor P; eds. Principles of drug action. United Kingdom: Churchill, Livingstone, 1990; 34–139.

[18] Yoon KW, Covey DF, Rothman SM. Multiple mechanisms of picrotoxin block of GABA-induced currents in rat hippocampal neurons. J Physiol. 1993; 464: 423–439. http://www.ncbi.nlm.nih.gov/pubmed/8229811

[19] Sayin U, Cengiz S, Altug T. Vigabatin as an anticonvulsant against pentylentetrazole seizures. Pharmacol Res. 1993; 28: 325–333. http://dx.doi.org/10.1006/phrs.1993.1134

[20] Kendall DA, Fox DA, Enna SJ. Effects of gamma vinyl GABA on bicuculline induced seizures. Neuropharmacol. 1981; 20: 351–355. http://dx.doi.org/10.1016/0028-3908(81)90008-3
[21] Olsen RW. The gamma-aminobutyric acid/benzodiazepine/barbiturate receptor-chloride ion channel complex of mammalian brain. In: Edelman M, Gall WE, Corvan WM; eds. Synaptic function. New York: John Wiley & Sons, 1987, 257–71.

[22] De Sarro A, Cechetti V, Fravolini V, Naccari F, Tabarini O, De Sarro G. Effects of novel 6-desfluroquinolones and classic quinolones on pentylenetetrazole-induced seizures in mice. Antimicrob Agents Chemother. 1999; 43: 1729–1736. http://www.ncbi.nlm.nih.gov/pubmed/10390231

[23] Zetler G. Central depressant effects of caerulein and cholecystokinin ocatapeptide (CCK-8) differ from those of diazepam and haloperidol. Neuropharmacol. 1981; 20: 277–283. http://www.ncbi.nlm.nih.gov/pubmed/6267504

[24] Bourin M, Hascoet M. The mouse light/dark box test. Eur J Pharmacol. 2003; 463: 55–65. http://dx.doi.org/10.1016/S0014-2999(03)01274-3

[25] Crawley J, Goodwin FK. Preliminary report of a simple animal behavior model for the anxiolytic effects of benzodiazepines. Pharmacol Biochem Behav. 1980; 13: 167–170. http://dx.doi.org/10.1016/0091-3057(80)90067-2

[26] Bourin M, Petit-Demouliere B, Donnchadha BN, Hascoet M. Animal models of anxiety in mice. Fundament Clin Pharmacol. 2007; 21: 567–571. http://dx.doi.org/10.1111/j.1472-8206.2007.00526.x

[27] Walsh RN, Cummins RA. The open field test: a critical review. Psychol Bull. 1976; 83: 482–504. http://www.ncbi.nlm.nih.gov/pubmed/17582919