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A Glass Capillary-based Microsensor for L-Glutamate in in vitro Uses

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1. Introduction

Since the pioneering work by R. Adams (1976), who detected a neurotransmitter catecholamine in mammalian brain by implanting a solid carbon electrode directly in animal brains, a lot of miniaturized in vivo and in vitro sensors for neurotransmitters have been proposed (Hirano & Sugawara, 2006; Sugawara, 2007; Zeyden et al. 2008; O’Neill et al., 1998). The field is slowly, but continually, expanding. Most of researches aim at developing electrochemical sensors with spatial and temporal resolution, which enable us to discern distribution of neurotransmitters within each neuronal subfield and estimate its concentration level and temporal changes in intact brains, acute slices and cultured neurons. In the central neuronal system of mammalian brain, L-glutamate is released from synaptic terminals and plays a vital role in brain development, synaptic plasticity, neurotoxicity, and neuropathological disorders (Reis et al., 2009; Bliss & Collingridge, 1993; Malenka & Nicoll, 1999). L-Glutamate is also involved in neuropathological disorders such as epilepsy, stroke, Parkinson’s disease and Alzheimer’s disease (Nishizawa, 2001; Mattoson et al., 2008). L-Glutamate may activate transmitter receptors located extrasynaptically on neurons and glia at greater distance from the place of exocytosis of synaptic vesicles, though the concentration level of such spillover of L-glutamate from synaptic cleft is not clear yet (Volterra & Meldolesi, 2005). The basal and enhanced level of extracellular L-glutamate plays a key role in neuronal functions, because its level will determine whether L-glutamate has actions or negligible actions on most glutamate receptors (Herman & Jahr, 2007).

Up to date, the in vivo level of L-glutamate in brain has been reported mostly for corpus striatum, while no in vivo data have been reported for the hippocampus. On the other hand, acute slices of hippocampal tissue offer experimental control of the neuronal network environment. In the in vitro case as well, a very limited number of microsensors have been applied to acute brain slices, probably because of technical difficulties in handling thin living slices (thickness 200-400 μm) and the lack of suitable miniaturized sensors.

A glass capillary-based enzyme sensor has been developed in which a three-electrode system is built in the capillary (Nakajima et al., 2003), hence outer reference and auxiliary electrodes are not necessary to be set in brain tissue. The sensor with the tip diameter of approximately 10 μm is promising as a microsensor for monitoring the enhanced extracellular level of L-glutamate release in each neuronal region of acute hippocampal slices.
under chemical and electric stimulation. In this review, we describe the principle, properties and application of a glass capillary-based sensor for \textit{in vitro} monitoring of L-glutamate in hippocampal slices.

2. Preparation and response principle of a glass capillary sensor for L-glutamate

2.1 Preparation of a glutamate oxidase (GluOx)-coated Au electrode

A working Au electrode, which is to be set in a capillary pipette, is prepared in the conventional manner (Oka et al., 2007). The one end of a gold wire (ø 0.30 mm) is coated with 0.3 µl of a detergent solution supplied as refill kit peroxidase (Os-gel-HRP, Bioanalytical systems, USA). The gold is then coated twice with each 0.3 µl of the Os-gel polymer followed by air-drying overnight. The surface of the Os-gel-HRP is coated with 1 µl of ACSF (Mg\(^{2+}\), Ca\(^{2+}\)-free) solution containing 2% bovine serum albumin (BSA), 0.2% glutaraldehyde and 65 U/ml GluOx. The electrode is necessary to be stored at 4°C until use. In this protocol, Nafion coating and ascorbate oxidase immobilization, which are commonly used for eliminating interference from L-ascorbate, are not employed, because the inner solution of a glass capillary sensor contains L-ascorbate oxidase \textit{(vide infra)}.

2.2 Preparation of a capillary sensor

The structure and photo of a glass capillary microsensor is shown in Fig. 1. The capillary sensor is composed of a Borosilicate glass capillary (outer diameter 1.5 mm and inner diameter 0.86 mm) having a tip diameter of approximately 10 µm, prepared by using a three-pull technique with a micropipette puller. The tip diameter can be measured under a microscope. Before use, the glass capillary is filled with a Mg\(^{2+}\), Ca\(^{2+}\)-free ACSF (approximately 3 µl) containing 1x10^3 U/ml ascorbate oxidase (AsOx). A Teflon-coated Pt wire (ø 0.127 mm) with ~2 mm of exposed Pt and a Teflon-coated Ag/AgCl wire (ø 0.127 mm) served as counter and reference electrodes, respectively. The working, counter and reference electrodes are inserted into the capillary pipette. The distance between the tip of the glass pipette and the working electrode is usually ~1.5 mm or shorter, as observed under a microscope. A larger distance leads to a delay in the response to L-glutamate.

![Fig. 1. A photo of a glass capillary sensor and its structure.](www.intechopen.com)
2.3 Electrochemical reaction at a glass capillary sensor

L-Glutamate released in brain slices diffuses into the inner solution of a capillary sensor to reach the top layer (GluOx-BSA layer) of the underlying working electrode. L-Glutamate oxidase (GluOx) catalyzes the oxidation of L-glutamate into α-ketoglutarate, producing electro-active H$_2$O$_2$ (Kusakabe et al., 1983).

\[
\text{L-glutamate} + \text{H}_2\text{O} + \text{O}_2 \rightarrow \alpha \text{keto glutarate} + \text{NH}_4^+ + \text{H}_2\text{O}_2 \quad (1)
\]

The Os-gel-HRP on the working electrode mediates the reduction of H$_2$O$_2$ in the following way (Vreeke et al., 1992).

\[
2\text{Os(II)} + \text{H}_2\text{O}_2 + 2\text{H}^+ \rightarrow 2\text{Os(III)} + 2\text{H}_2\text{O} \quad (2)
\]

\[
\text{Os(III)} + e^- = \text{Os(II)} \quad (3)
\]

The Os(III) produced is reduced at the underlying electrode, giving a reduction current, which is used as a response of the present sensor. The operation potential is 0 V vs. Ag-AgCl.

The capillary electrode has the advantage that the inner solution can contain various enzymes. The interference from ascorbic acid, one of the major components in the brain, is removed by adding ascorbate oxidase to the inner solution. The enzyme catalyzes the oxidation of L-ascorbate to 2-dehydroascorbate (Tokuyama et al., 1965; Nakamura et al., 1968) according to

\[
\text{L-ascorbate} + 1/2 \text{O}_2 \rightarrow \text{dehydroascorbate} + \text{H}_2\text{O} \quad (4)
\]

The pH range (pH 5.6-7.0) for the catalytic action of ascorbate oxidase is very close to that (pH 5.5-10.5) of GluOx (Kusakabe et al., 1983), and hence both enzymes are active at pH 7.0.

2.4 Monitoring L-glutamate with a capillary sensor

Our protocol for monitoring L-glutamate in brain slices is as follows. Prior to its implantation into a hippocampal slice, a glass capillary sensor is operated in air at 0 V vs. Ag-AgCl until a steady current is obtained. Then, the sensor is positioned above the surface of a target neuronal region of a hippocampal slice in interface preparation, followed by lowering into the target region of the slice at a depth of ~100 μm with a manipulator. Appearance of a sharp electric noise indicates the implantation of the sensor into the slice. The sensor is kept in the slice until a steady current is attained. After attainment of a steady current, recording an L-glutamate current is started. Since the volume of the sensor inner solution is maintained, continuous and long-time monitoring of L-glutamate in a brain slice is feasible.

3. Response principles of a capillary sensor in bulk solutions and brain slices

The response profiles of a capillary sensor are categorized into two cases, depending on whether it is used in an aqueous solution just above the target region of a brain slice or it is implanted in the target region of a brain slice (Sugawara, 2007). When a capillary sensor is positioned in a bulk aqueous solution just above a brain slice, capillary action is essentially important for its operation. In an aqueous solution, a small volume of a sample solution
containing L-glutamate is spontaneously sampled into an inner solution by capillary action. On the other hand, such capillary action does not work in brain slices because of viscous nature of extracellular fluid. It is noted that a small fraction of GluOx is leached from the top surface of a GluOx-immobilized Au electrode into a capillary inner solution (Oka et al., 2007) and hence, leached GluOx catalyzes the oxidation of L-glutamate, producing hydrogen peroxide, which is detected at the working electrode.

### 3.1 Capillary action of a pulled glass capillary in an aqueous solution

Figure 2 shows the photos that demonstrate the capillary action of a pulled glass capillary in an aqueous solution. The capillary inner solution contained a visible dye, i.e., methylene blue (MB). One can see that an aqueous solution comes into the capillary with time. When an aqueous solution contained 5% dextrane, the sampling rate was deteriorated significantly due to an increase in viscosity. In another set of experiments, we quantified the capillary action by measuring the weight of a capillary dipped in an aqueous solution (Nakajima et al., 2003). With a tip diameter of 2.5 μm or less, a rise of the solution by capillarity is not observed. On the other hand, in the case that a capillary with a tip diameter of 10 μm is dipped in an aqueous solution, the weight of the solution in the capillary increases linearly with dipping time up to 20 min. The slow rise of the solution is due to the conical tip of the capillary, which decelerates the movement of the solution. Thus, pulled glass capillaries exhibit different magnitudes of capillarity, which depend on tip size and dipping time. The quantitative response of a glass capillary sensor in an aqueous solution is relied on such capillary action.

![Fig. 2](image.png)

Fig. 2. Photos that demonstrate capillarity-based sampling of an aqueous solution. The inner solution of a glass capillary contained methylene blue.

### 3.2 Diffusion of L-glutamate into a capillary inner solution

In contrast to its use in an aqueous solution, the capillary action of a glass capillary does not work in a brain slice because of the viscous nature of extracellular fluid. The volume of an inner solution of a glass capillary is maintained, as shown in Fig. 3, even after its
implantation into a brain slice. Under such circumstance, the response of a capillary-based sensor is based on diffusional entry of L-glutamate into its inner solution.

**Fig. 3.** Fluorometric images of a glass capillary. A pulled glass capillary containing ACSF (Mg²⁺ and Ca²⁺-free) was inserted into a brain slice, loaded in advance with a fluorescence dye BCECF (Oka et al., 2007).

### 3.3 In situ calibration

One of the essential tasks to be considered is how to correlate the sensor response to final L-glutamate concentration. There are two ways for maintaining brain slices alive (Sugawara, 2007), i.e., a brain slice is fully submerged in a bath solution (Fig. 4a) and a slice is kept alive by passing an ACSF underneath the slice (Fig. 4b). Calibrating the implanted glass capillary sensor is also dependent on how slices were maintained. In the submerged case, calibrating sensor responses and stimulation of the slice can be performed by changing the concentration of L-glutamate or a stimulant in the bath solution. For brain slices in interface preparation, and also in submerge preparation, post-in vitro calibration is common for calibrating the responses of an implanted sensor, because the adsorption of extracellular components on the top surface of a sensor deteriorates the sensitivity of the response. In this protocol, an implanted sensor is transferred into an aqueous solution and calibrated with a standard L-glutamate solution. However, the post-in vitro calibration approach is based on the assumption that the sensor exhibits the same degree of deterioration both in a brain slice and an aqueous solution. To improve the uncertainty of the post-in vitro calibration, we suggested a method for calibrating an implanted sensor by injecting a small volume of (5 µl) of a standard L-glutamate solution into the close vicinity of the glutamate sensor through a glass capillary (Oka et al., 2007; Chiba et al., 2010). The sensor exhibits a transient current-time profile rather than a steady one (Fig. 5), due to the active reuptake process and diffusional wash out of L-glutamate. Consequently, an instantaneous current is used for calibration. The calibration has to be done at each neuronal region, because the activity of reuptake process is neuronal region-dependent. It is noted that L-glutamate levels measured with a capillary sensor are dependent on the type of slice preparation (Sugawara, 2007). In the submerged case where a sensor is positioned above the surface of a target neuronal region, an L-glutamate level obtained is the one that diffused out of the slice. Such alignment of a sensor is common for not only capillary sensors but also patch sensors using natural receptors. However, thus-obtained L-
glutamate levels are obviously lower than those in the brain slice (Oka et al., 2009). Consequently, a relative change in the response rather than the very magnitude of the response is a matter of concern for monitoring neuronal events. On the other hand, the implantation of a sensor into a brain slice can measure L-glutamate in the vicinity of neurons, but calibrating the sensor response needs a hard task. The lower detection limit for L-glutamate of GluOx-based sensors has been reported to be sub-μM or better. The detection limit of nM range has also been reported (Tang, et al., 2007; Braeken et al., 2009). However, these values are based on the measurements in an electrolyte solution rather than in brain or brain tissues. The properties of tissue environment, for example viscosity, differ significantly from an electrolyte solution. The diffusion of L-glutamate affected by viscosity may alter the sensitivity of the sensor. Therefore, in situ calibration of the sensor response in brain tissue is important for knowing the detection limit of an implanted sensor.

![Fig. 4. Two types of slice preparation. (a) A brain slice is submerged in a bath solution and (b) a brain slice is placed on a lens paper through which an ACSF flows (Sugawara, 2007).](image)

![Fig. 5. Current-Time profiles for in situ calibration at (a) DG, (b) CA3 and (c) CA1 and corresponding calibration graphs for L-glutamate (Oka et al., 2007, 2009; Chiba et al., 2010).](image)
3.4 Selectivity
A number of interfering species coexist with glutamate in brains. The selectivity issues of GluOx-based sensors for eliminating interference from coexisting ions and molecules have been addressed by several authors. Interference from ascorbate that are present in the brain at concentration much larger than that of L-glutamate has been eliminated by coating Nafion that excludes ascorbate anions electrostatically from the electrode surface (Day et al., 2006; Oldenziel et al., 2006a; Rutherford et al., 2007; Burmeister & Gerhardt, 2001) or by using ascorbate oxidase that mediate the decomposition of ascorbate before it approaches to the surface of an underlying electrode (Oldenziel et al. 2006b; Kulagina et al., 1999; Oka et al. 2007). Conducting polymer-modified electrodes are also effective for eliminating the interferent (Rahman et al., 2005). Another potential interfering compound is L-glutamine. The interference arises probably from the presence of glutaminase as contaminant in isolated GluOx (Yamauchi et al., 1984). The isolated GluOx-based sensor shows a response to glutamine at a few µM level. However, at large glutamine concentration above 300 µM, which corresponds to the concentration of glutamine in brain (Kanamori & Ross, 2004; Lerma et al., 1986), the response to glutamine is saturated. In addition, such interference disappears in the presence of a small amount of L-glutamate (Oldenziel et al., 2006). The response to glutamine is significantly modified by using recombinant GluOx (Hozumi et al., 2011). The recombinant GluOx-based sensor suffers interference from glutamine only at high concentration above 300 µM, and the response to glutamine is very weak (1.83 pA/µM an average between 300 and 500 µM) in comparison with the response to L-glutamate (472 pA/µM). The glutamate sensor based on recombinant GluOx does not exhibit responses to glycine, GABA, serotonin, (each 1.0 mM), and L-aspartic acid (200 µM). Since the typical concentration of L-ascorbic acid in brain is 100-500 µM (Walker et al., 1995; Nedergaard et al., 2002) and the estimated basal concentration of glutamine in brain ranges from 200 to 400 µM (Lerma et al., 1986; Kanomori and Ross, 2004) and that of L-aspartic acid is 0.25 µM or less (Robert et al., 1998), the effect of these compounds on L-glutamate currents appears to be of the minor importance.

4. Monitoring of L-glutamate release in hippocampal slices
Since L-glutamate released from nerve terminals into the synaptic cleft is subject to diffusion and dilution into extracellular space and uptake into neurons and glia by excitatory amino acid transporters, the extracellular level of L-glutamate is essentially important for elucidating neuronal signal transmission processes. Although the in vivo level of L-glutamate in the neuronal subfield of the hippocampus is important (Table 1), no reports have been published yet on hippocampal levels of L-glutamate. A small number of data have been gathered from in vitro study using acute hippocampal slices that consist of freshly isolated brain tissue maintained in a chamber. The data described in this section are mostly taken from our results. The extra-cellular and extra-slice L-glutamate levels in acute mouse hippocampal slices obtained with a capillary sensor are given in Table 2, together with the reported basal levels for hippocampal slices (Oldenziel et al., 2007; MaLamore et al., 2010, Hermann & Jahr, 2007). The reported basal level varies in a wide range from several tens nM to a few µM (Table 1). Therefore, the basal L-glutamate level in brain and brain slices is still a matter of debate.
4.1 Chemical stimulation
The monitoring of enhanced L-glutamate level evoked by physiologically relevant stimuli enables us to discern the role and action of each stimulant as well as the regional distribution of L-glutamate in hippocampal slices. We performed monitoring of L-glutamate release in various neuronal regions of mouse hippocampal slices under stimulation with KCl, tetraethylammonium (TEA) chloride and ischemia.

4.1.1 KCl stimulation
The depolarization evoked by KCl (0.10 M) stimulation enhances extracellular L-glutamate level in hippocampal slices, but the enhanced concentration level at dentate gyrus (DG) (Oka et al., 2007) is much higher than those at cornu ammonis 1 (CA1) (Oka et al., 2009) and cornu ammonis 1 (CA3) (Chiba et al., 2010). The K+ evoked L-glutamate levels in the CA1 and CA3 regions are very low, i.e., approximately 4 μM, owing to reuptake processes. Sodium-dependent excitatory amino acid transporters (EAATs) (Taxt et al., 1984; Rothstein et al., 1994; Furuta et al., 1997) are present differentially within neurons and astroglia (Taxt et al., 1984): EAAC1(EAAT3) is highly enriched in hippocampus with distribution of CA1, CA3 > DG, while EAAT4 is present in trace amount in hippocampus (Furuta et al., 199). The regional transporter distribution suggests that L-glutamate in the CA1 and CA3 regions is more strongly removed from extracellular space than in the DG region and hence, the L-glutamate level is maintained very low.

4.1.2 TEA stimulation
The stimulation by a K+ ion channel blocker tetraethylammonium (TEA) chloride is known to elicit chemically induced synaptic potentiation (clLTP) in CA1 of hippocampal slices (Aniksztejn&Ben-Ari, 1991; Hosokawa et al., 1995). The TEA stimulation activates both NMDA receptor channels (Hanse&Gustafsson, 1994) and voltage-dependent calcium channels (Huang et al., 1993). The activation of the NMDA receptor channels induces a calcium influx, often inducing LTP, which is similar to that evoked by a brief afferent tetanus (electrical stimulation). The regional distribution of extracellular L-glutamate in hippocampal slices under TEA stimulation (Oka et al., 2007; Oka et al., 2009, Chiba et al., 2010) is similar to that obtained by K+ stimulation, though L-glutamate levels at CA1 and CA3 are slightly larger than those obtained by K+ stimulation.

4.1.3 Ischemia
One of the most enigmatic aspects of ischemic injury is the selective vulnerability of the hippocampal CA1 neurons, whereas the neurons in the CA3 and DG regions are relatively spared. The monitoring of L-glutamate with a glass capillary sensor showed that the extracellular level of L-glutamate is in the order of CA1 ≈ CA3 > DG (Nakamura, et al., 2005), which is in accordance with the imaging study in terms of an L-glutamate flux (Hirano et al., 2003; Okumura et al., 2009). The concentration level of L-glutamate is much larger than those observed by other chemical stimulation (Table 2), indicating that the high level of L-glutamate is released into the extracellular space and diffuses out of the slice into the bath. The time course of the L-glutamate flux at region CA1 is biphasic and that at region DG is modestly biphasic. Similar biphasic time course of L-glutamate release has been reported for rat striatum (vulnerable to ischemic injury) by using a dialysis electrode (Asai et al., 1996; Kohno et al., 1998).
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Table 1. Examples of basal extracellular L-glutamate levels in the anesthetized rat brain (in vivo).

| Region          | Concentration (µM) | Method            | Size (µm) | Reference         |
|-----------------|--------------------|-------------------|-----------|-------------------|
| Striatum        | 29.0 ± 9.0         | carbon fiber E    | 10        | Kulagina et al 1999 |
| Brain           | 23.6 ± 5.3         | hydrogel E        | 10        | Oldenziel et al. 2006a |
| Striatum        | 18.2 ± 9.3         | hydrogel E        | 10        | Oldenziel et al. 2006b |
| Striatum        | 7.3 ± 0.9          | self-referencing E| 15        | Rutherford et al. 2007 |
| Dorsal striatum | 2.0 ± 0.5          | conducting polymer E | 25         | Rahman et al. 2005 |
| Frontal cortex  | 1.6 ± 0.3          | self-referencing E| 15        | Day et al. 2007   |
| Left striatum   | 1.5 ± 0.3          | microsensor       | 3.0 (mm)  | Zhang et al. 2004 |
| Striatum        | 1.3                | ceramic-based multi E | 15       | Pomerleau et al. 2003 |
| Right striatum  | 3.0 ± 0.6          | microdialysis     |           | Miele et al. 1996 |
| Striatum        | 2.7 ± 0.9          | microdialysis     |           | Robert et al. 1998 |
| Striatum        | 1.15 ± 0.7         | microdialysis     |           | Lada et al. 1997  |
| Striatum        | 1.97 ± 0.7         | push-pull perfusion |           | Kottergoda et al. 2002 |

Table 2. Regional distribution of extracellular and extra-slice levels of L-glutamate in mouse hippocampal slices.

| Stimulation   | Subfield (concentration, µM) | Method (size, µm) | Reference               |
|---------------|------------------------------|-------------------|-------------------------|
| Extracellular (acute) |                             |                   |                         |
| KCl (0.1M)    | DG (22) > CA3 (4.1) ~ CA1 (4.0) | capillary E (10)  | Nakajima et al. 2003    |
| TEA (25 mM)   | DG (20) > CA3 (7.5) ~ CA1 (6.0) | capillary E (10)  | Chiba et al. 2010       |
| Electric (2 Hz)| CA1 (1.4)                   | capillary E (10)  | Hozumi et al., 2011     |
| Electric (0.052 Hz)| CA1 (32 nM)           |                   |                         |
| Basal         | CA1 (1.7)                   | hydrogel E (10)   | Oldenziel et al., 2007  |
| Basal         | CA1 (~25 nM)                | patch sensor (whole cell) | Hermann and Jahr, 2007 |
| Extracellular (cultured) |                             |                   |                         |
| KCl (0.1M)    | hippocampal slice (0.12)    | microdialysis     | Robert et al. 1998      |
| Extra-slice (acute) |                            |                   |                         |
| Basal         | P19 cell (5.8)              | self-referencing E (2-5) | Maclamore et al. 2010  |
| GABA          | CA1 (1.5) > CA3              | patch sensor (2)  | Shimane et al. 2006     |
| Ischemia      | CA3 (60) ~ CA1 (47) > DG (20) | capillary E (10)  | Nakamura et al. 2005    |
4.2 Electric stimulation

Recording field excitatory postsynaptic potentials (fEPSPs) have been a well-established method for knowing neuronal activities in electrophysiological studies (Bliss et al., 2007). The potentials are produced by a group of cells and reflect in an indirect way the changes in the synaptic and action potentials. Analysis of fEPSPs provides information on the average activity of the neurons in group, including the induction and expression of long-term potentiation (LTP) and long-term depression (LTD). Simultaneous monitoring of L-glutamate release and fEPSPs evoked by physiologically relevant electric stimulation will provide explicit information on the actual amount of L-glutamate released into the synaptic cleft and extracellular space in the vicinity of the stimulation site. However, such a measurement is still a challenging task, because of the requirement of placing multiple electrodes in a confined neuronal region and fairly low level of glutamate will be released by physiologically relevant stimulation.

A glass capillary sensor has the advantage that all electrodes required for amperometric current measurements are built in the capillary interior and hence, the tip size (approximately 10 μm) is small enough to implant it between stimulation and recording electrodes for fEPSP measurements. Fig. 6 shows the setup of simultaneous recordings of a glutamate current with a glass capillary sensor and fEPSPs with stimulation and recording electrodes. The tip of the capillary sensor can be positioned in the middle between stimulation and recording electrodes for fEPSP measurements. The example of simultaneous measurements at CA1 of a hippocampal slice demonstrates that although no significant changes in a glutamate current is detected by application of 0.052 Hz (test stimuli), a transient change in the current is observed by application of 2 Hz stimulation, indicating enhanced release of L-glutamate in CA1 region (Hozumi et al., 2011). The L-glutamate level in region CA1 at 2 Hz stimulation obtained by in situ calibration ranged from 0.8 to 2.2 μM (1.4 μM as an average) from 5 independent measurements. The concentration is slightly smaller than those obtained by KCl stimulation. Although direct monitoring of L-glutamate level at test stimuli (0.052 Hz) is not achieved, we can record an L-glutamate current by changing the intensity of electric stimulation from 1 Hz to 3 Hz. The estimated concentration level at 0.052 Hz is 32 ± 7 nM (n=3), which is very close to the reported one using the whole cell recordings (Herman&Jahr, 2007).

Fig. 6. A photo that shows the setup of simultaneous monitoring of fEPSPs and a glutamate current. The traces of fEPSP and a glutamate current were simultaneously monitored.
5. Conclusions and prospects

The present review demonstrates that a glass capillary-based microsensor is useful for knowing the distribution and level of extracellular L-glutamate in acute hippocampal slices. The L-glutamate levels are markedly dependent on the neuronal regions and types of stimulation. Although discerning the concentration level of L-glutamate in acute brain slices with microsensors is still on the stage of accumulating the local concentration level of L-glutamate, there are increasing efforts for clarifying the sources and places of extracellular L-glutamate release. Since the experimental condition can be controlled easily, microsensors will be promising as a tool for monitoring and estimating the averaged extracellular level of L-glutamate in acute brain slices. On the other hand, recent advances in developing new fluorescent probes have enabled to visualize L-glutamate with spine-sized resolution using a cultured neuron. Such a single synapse study will significantly help the understanding of the molecular events that occur at a synapse. Combining the single synapse data with the extracellular data will significantly advance the understanding of the sources and places of extracellular L-glutamate release. In addition, the sophisticated combination of microsensor studies with an electrophysiological study to correlate L-glutamate level to the neuronal activity will be a promising way for solving debates as to whether LTP is due to presynaptic or postsynaptic changes.

6. References

Adams, R.N. (1976). Probing brain chemistry with electroanalytical techniques. Anal. Chem., 48, 1126A-1138A.
Aniksztejn L. & Ben-Ari, Y. (1991). Novel form of long-term potentiation produced by a K+ channel blocker in the hippocampus. Nature, 349, 67-69.
Asai, S.; Iribe, Y.; Kohno, T. & Ishikawa, K. (1996). Real time monitoring of biphasic glutamate release using dialysis electrode in rat acute brain ischemia. NeuroReport, 7, 1092-1096.
Bliss, T.V.P. & Collingdidge, G.L. (1993). A synaptic model of memory: long-term potentiation in the hippocampus. Nature, 361, 31-39.
Bliss, T.; Collingridge, G. & Morris, R. (2007). Synaptic plasticity in the hippocampus. In The Hippocampus book, Andersen A.; Morris, D.; Amaral, D.; Bliss, T. &O'keefe, J. eds., New York, Oxford university Press., pp343-474.
Braeken, D.; Rand, D.R.; Andrei, A.; Huys, R.; Spira, M.E.; Yitzchaik, S.; Shappir, J.; Borghs, G.; Callewaert, G. & Bartic, C. (2009). Glutamate sensing with enzyme modified floating gate field effect transistors. Biosensors and Bioelectronics, 24, 2384-2389.
Burmeister, J.J. & Gerhardt, G.A. (2001). Self-referencing ceramic-based multisite microelectrodes for the detection and elimination of interferences from the measurements of L-glutamate and other analytes. Anal. Chem., 73, 1037-1042.
Chiba.H.; Deguti,Y.; Kanazawa, E.; Kawai, J.; Nozawa, K.; Shoji, A. & Sugawara, M. (2010). In vitro measurements of extracellular L-glutamate level in region CA3 of mouse hippocampal slices under chemical stimulation. Anal Sci., 26, 1103-1106.
Day, B.K.; Pomerleau, F.; Burmeister, J.J.; Huettl, P. & Gerhardt, G.A. (2006) Microelectrode array studies of basal and potassium-evoked release of L-glutamate in the anesthetized rat brain. J. Neurochem., 96, 1626-1635.
Furuta, A.; Martin, L.J.; Lin, C.-I.; Dykes-Hoberg, M. & Rothstein, J.D. (1997). Cellular and synaptic localization of the neuronal glutamate transporters excitatory amino acid transporter 3 and 4. *Neuroscience*, 81, 1031-1042.

Hanse, E. & Gustafsson, B. (1994). TEA elicits two distinct potentiation of synaptic transmission in the CA1 region of the hippocampal slice. *J. Neurosci.*, 14, 5028-5034.

Herman, M.A. & Jahr, C.E. (2007). Extracellular glutamate concentration in hippocampal slice. *J. Neurosci.*, 27, 9736-9741.

Hirano, A. & Sugawara M. (2006). Receptors and enzymes for medical sensing of L-glutamate. *Mini-review in Medicinal Chemistry*, 6, 1091-1100.

Hirano, A.; Moridera, N.; Akashi, M.; Saito, M.& Sugawara, M. (2003). Imaging of L-glutamate fluxes in mouse brain slices based on an enzyme-based membrane combined with a differential-image analysis. *Anal. Chem.*, 75, 3775-3783.

Hosokawa, T.; Rusakov, D.A.; T. V. P. Bliss, T.V. P. & Fine, A. (1995). Repeated confocal imaging of individual dendritic spines in the living hippocampal slice: evidence for changes in length and orientation associated with chemically induced LTP. *J. Neurosci.*, 15, 5560-5573.

Hozumi, S.; Kana Ikezawa, K.; Atushi Shoji,A.; Hirano-Iwata,A.; Bliss,T.& Sugawara, M. (2011). Simultaneous monitoring of excitatory postsynaptic potentials and extracellular L-glutamate in mouse hippocampal slices. *Biosensors&Bioelectronics*, 26, 2975-2980.

Huang, Y.-Y. & Malenka, R. C. (1993). Examination of TEA-induced synaptic enhancement in area CA1 of the hippocampus: the role of voltage-dependent Ca\(^2+\) channels in the induction of LTP. *Neurosci.*, 13, 568-576.

Kanamori, K. & Ross, B. D. (2004). Quantitative determination of extracellular glutamine concentration in rat brain and its elevation in vivo by system A transport inhibitor, alpha-(methylamino)isobutyrate. *J. Neurochem.*, 90, 203-210.

Kohno, T.; Asai, S.; Iribe, Y.; Hosoi, I.; Shibata, K. & Ishikawa, K. (1998). An improved method for the detection of changes in brain extracellular glutamate levels. *J. Neurosci. Methods*, 81, 199-205.

Kottegoda, S.; Shaik, I. & Shippy, S. A. (2002). Demonstration of low flow push-pull perfusion. *J. Neurosci. Methods*, 121, 93-101.

Kulagina, N.V.; Shankar, L. & Michael, A.C. (1999) Monitoring glutamate and ascorbate in the extracellular space of brain tissue with electrochemical microsensors. *Anal. Chem.*, 71, 5093-5100.

Kusakabe, H.; Midorikawa, Y.; Kuninaka, A.; Fujisima, T. & Yoshino, H.(1983). Purification and properties of a new enzyme, L-glutamate oxidase, from Streptomyces sp. X-119-6 grown on wheat bran. *Agric. Biol. Chem.*, 47, 1323-1328.

Lada, M.W.; Vickroy, T. W. & Kennedy, R.T. (1997). High temporal resolution monitoring of glutamate and aspartate in vivo using microdialysis on-line with capillary electrophoresis with laser-induced fluorescence detection. *Anal. Chem.*, 69, 4560-4565.

Lerma, J.; Herranz, A. S.; Herreras, O.; Abraira, V. & Martin del Rio, R. (1986). In vivo determination of extracellular concentration of amino acids in the rat hippocampus. A method based on brain dialysis and computerized analysis. *Brain research*, 384, 145-155.
Malenka, R. C. & Nicoll, R. A. (1999). Long-term potentiation—a decade of progress?. Science, 285, 1870-1874.

McLamore, E. S.; Mohanty, S.; Shi, J.; Claussen, J.; Jedlicka, S.S.; Rickus, J. L. & Porterfield, D. M. (2010). A self-referencing glutamate biosensor for measuring real time neuronal glutamate flux. J. Neurosci. Method, 189, 14-22.

Mattson M. P. (2008). Glutamate and neurotrophic factors in neuronal plasticity and disease. Ann. N. Y. Acad. Sci., 1144, 97-112.

Miele, M.; Boutelle, M.G. & Filenz, M. (1996). The source of physiologically stimulated glutamate efflux from the striatum of conscious rats. J. Physiol., 497, 745-751.

Nakajima, K.; Yamagiwa, T.; Hirano, A. & Sugawara, M. (2003). A glass capillary microelectrode based on capillarity and its application to the detection of L-glutamate release from mouse brain slices. Anal. Sci., 19, 55-60.

Nakamura, N.; Negishi, K.; Hirano, A. & Sugawara, M. (2005). Real time monitoring of L-glutamate release from mouse brain slices under ischemia with a glass capillary based enzyme electrode. Anal. Bioanal. Chem., 383, 660-667.

Nakamura, T.; Makino, N. & Ogura, Y (1968). Purification and properties of ascorbate oxidase from cucumber. J. Biochem. 64, 189-195.

Nedergaard, M.; Takano, T. & Hansen, J. (2002). Beyond the role of glutamate as a neurotransmitter. Nat. Rev. Neurosci., 3, 748-755

Nishizawa, Y. (2001). Glutamate release and neuronal damage in ischemia. Life Sci., 69, 369-381.

Oldenziel, W.H.; Dijkstra, G.; Cremers, T.I.F.H. & Westrink, B.H.C. (2006). Evaluation of hydrogel-coated glutamate microsensors. Anal. Chem., 78, 3366-3378.

Oldenziel, W.H.; Zeyden M. Dijkstra, G., Chijsen, W.E.J.M.; Karst, H.; Cremers, T.I.F.H. & Westerink, B.H.C. (2007). Monitoring extracellular glutamate in hippocampal slices with a microsensor. J. Neurosci. Methods, 160, 37-44.

Oldenziel, W.H.; Dijkstra, G.; Cremers, T.I.F.H. & Westrink, B.H.C. (2006). In vivo monitoring of extracellular glutamate in the brain with a microsensor. Brain Res., 1118, 34-42.

O’Neill, R. D.; Lowry, J. P. & Mas, M. (1998). Monitoring brain chemistry in vivo: voltammetric techniques, sensors, and behavioral applications. Crit. Rev. Neurobiol., 12, 69-127.

Oka, T.; Tasaki, C.; H. Sezaki, H. & Sugawara, M. (2007). Implantation of a glass capillary electrode in mouse brain slices for monitoring of L-glutamate release. Anal. Bioanal. Chem., 388, 1673-1679.

Oka, T.; Tominaga, Y.; Wakabayashi, Y. Shoji, A. & Sugawara, M. (2009). Comparison of the L-glutamate level in mouse hippocampal slices under tetraethylammonium chloride stimulation as measured with a glass capillary sensor and a patch sensor. Anal. Sci., 25, 353-358.

Okumura, W.; Moridera, N.; Kanazawa, E.; Shoji, A.; Hirano-Iwata, A. & Sugawara, M. (2009). Visualizing L-glutamate fluxes in acute hippocampal slices with glutamate oxidase-immobilized cover slips. Anal. Biochem., 385, 326-333.

Pomerleau, F.; Day, B. K.; Huettl · P.; Burmeister, J. J. & Gerhardt, G. A. (2003). Real time in vivo measures of L-glutamate in the rat central nervous system using ceramic-based multisite microelectrode arrays. Ann. N. Y. Acad. Sci., 1003, 454-457.
Rahman, M.A.; Kwon, N.-H.; Won, M.-S.; Choe, E.S. & Shim, Y.-B. (2005). Functionalized conducting polymer as an enzyme immobilizing substrate: an amperometric glutamate microbiosensor for in vivo measurements. *Anal. Chem.*, 77, 4854-4860.

Reis, H.J.; Guatimosim, C.; Paquet, M.; Santos, M.; Ribeiro, F.M.; Kummer, A.; Schenatto, G.; Vsalgado, J.V.; Vieira, L.B.; Teixeira, A. L. & Palotás, A. (2009). Neuro-transmitters in the central nervous system & their implication in learning and memory processes. *Curr. Med. Chem.*, 16, 796-840.

Robert, F.; Bert, L.; Parrot, S.; Denoroy, L.; Stoppini, L. & Renaud, B. (1998). Coupling on-line brain microdialysis, precolumn derivatization and capillary electrophoresis for routine minute sampling of O-phosphoethanolamine and excitatory amino acids. *J. Chromatog.* A, 817, 195-203.

Rutherford, E.C.; Pomerleau, F.; Huettl, P.; Strömberg, I. & Gerhardt, G.A. (2007). Chronic second-by-second measures of L-glutamate in the central nervous system of freely moving rats. *J. Neurochem.*, 102, 712-722.

Shimane, M.; Miyagawa, K. & Sugawara, M. (2006). Detection of γ-aminobutyric acid-induced glutamate release in acute mouse hippocampal slices with a patch sensor. *Anal. Biochem.*, 353, 83-92.

Sugawara, M. (2007). Methodological Aspects of *in vitro* sensing of L-glutamate in acute brain slices. *The Chemical Records*, 7, 317-325.

Tang, L.; Zhu, Y.; Yang, X. & Li, C. (2007). An enhanced biosensor for glutamate based on self-assembled carbon nanotubes and dendrimer-encapsulated platinum nanobiocomposite-doped polypyrrole film. *Anal. chim. Acta*, 596, 145-150.

Taxt, T. & Storm-Mathisen, J. (1984). Uptake of D-aspartate and L-glutamate in excitatory axon terminals in hippocampus: autoradiographic and biochemical comparison with γ-aminoenbutyric acid and other amino acids in normal rats and in rats with lesions. *Neuroscience*, 11, 79-100.

Tokuyama, K.; Clark, E.E.; Dawson, C.R. (1965) Ascorbate oxidase: a new method of purification. Characterization of the purified enzyme. *Biochemistry*, 4, 1362-1370.

Volterra, A. & Meldolesi, J. (2005). Astrocytes from brain glue to communication elements: the revolution continues. *Nature review Neurosci.*, 6, 626-640.

Vreeke, M.; Maiden, R. & Heller A. (1992). Hydrogen peroxide and β-nicotinamide adenine dinucleotide sensing amperometric electrodes based on electrical connection of horse radish peroxidase redox centers to electrodes through a three-dimensional electron relay polymer network. *Anal. Chem.*, 64, 3084-3090.

Walker, M.C.; Galley, P.T.; Errington, M.L.; Shorvon, S.D. & Jefferys, J. G. R.(1995). Ascorbate and glutamate release in the rat hippocampus after perforant path stimulation: a "dialysis electrode" study. *J. Neurochem.*, 65, 725-731.

Yamauchi · H.; Kusakabe, Y.; Midorikawa, T.; Fujisima, T. & Kuninaka, A. (1984). Enzyme electrode for specific determination of L-glutamate. *Eur. Congr. Biotechnol.*, 1, 705.

Zhang, F.-F.; Wan, Q.; Li, C.-X.; Wang, X.-L.; Zhu, Z.-Q.; Xian, Jin, L.-T. & Yamamoto, K. (2004). Simultaneous assay of glucose, lactate, L-glutamate and hypoxanthine levels in a rat striatum using enzyme electrodes based on neutral red-doped silica nanoparticles. *Anal. Bioanal. Chem.*, 380, 637-642.

Zeyden, M.; Oldenziel, W.H.; Rea, K.; Cremers, T.I. & Westerink, B.H. (2008). Microdialysis of GABA and glutamate: Analysis, interpretation and comparison with microsensors. *Pharmacology, Biochem and Behaviror*, 90, 135-147.
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