Clinical features of respiratory syncytial virus bronchiolitis in an infant: rapid and fatal brain involvement

Paolo Bottino1†*, Rebecca Miglino1†, Lisa Pastrone1, Anna Maria Barbui1, Giovanni Botta2, Elisa Zanotto1, Francesca Sidoti1, Cristina Costa1 and Rossana Cavallo1

Abstract

Background: Respiratory Syncytial Virus (RSV) infection is a significant cause of bronchiolitis and pneumonia, mostly responsible for hospitalization and infant death worldwide. However, in recent years the importance of extrapulmonary RSV manifestations, especially at neurological level, have become evident. Seizures, lethargy, ataxia and status epilepticus are suggestive of brain involvement, but also in their absence a direct neurological damage RSV-related need to be evaluated.

Case presentation: A 40-day old male infant was admitted to the Emergency Department with severe bronchiolitis and dyspnea. The patient was reported to be coughing for a week with a vomiting episode in the previous two days. The nasopharyngeal swab confirmed the diagnosis of RSV infection and blood gas test showed hypoxemia and respiratory acidosis. For these reasons, the patient was provided with oxygen therapy. A few hours later, after an initial improvement in clinical parameters, a worsening of respiratory dynamics occurred and the patient was prepared for endotracheal intubation, but in the meantime death occurred. During all the observation period in the Emergency Room, no signs of neuropathological damage were evident. Post mortem examination showed lungs congestion with alveolar atelectasis and white matter degradation with severe edema at brain level. Microbiological analysis performed on autopic samples confirmed the presence of RSV genome in tracheobronchial aspirate, meningeal swabs, pericardic and abdominal fluids, lung and brain biopsies.

Conclusions: RSV is usually associated with respiratory diseases, however, as reported by an increasingly number of studies, the systemic dissemination of virus during severe disease can lead to a sudden infant death. The clinical picture herein reported showed a severe bronchiolitis resulting in a fatal and underestimated cerebral involvement due to RSV neurotropic behaviour and underline the need for clinicians to pay more attention to neurological involvement of RSV infection, even in absence of cerebral damage evidence.

Keywords: RSV, Infant, Bronchiolitis, Neurotropism, Sudden infant death, Case report

Background

Respiratory Syncytial Virus (RSV) infection is a significant cause of hospitalization and infant death worldwide [1]. It affects 60–70% of children by the age of 1 year and almost all children by 2 years of age and it is mostly responsible for 45–90% of episodes of bronchiolitis and 15–35% of pneumonia [2, 3]. However, in recent years the importance of extrapulmonary RSV manifestations, such
as neurological, myocardial or endocrine, have become evident [4, 5].

Focusing on neuropathy, the most reported clinical signs and symptoms are seizures (reported in 7% of children and 12.9% of newborn), apnoeas, lethargy, ataxia, status epilepticus, encephalopathies and encephalitis [5–9].

The pathophysiological mechanism of encephalopathy associated with acute bronchiolitis remains undefined, but a direct invasion of the central nervous system by RSV seems to be the major mechanism responsible for cerebral involvement in RSV disease [4, 10]. As reported by Li et al. (2006), RSV can infect pulmonary neurons and interact with the chemokine receptor for CX3CL1 (CX3CR1) expressed in these cells, overrunning the Central Nervous System with resulting neurological abnormalities in patients [11].

Here, we report a case of an infant admitted with RSV-related severe bronchiolitis and a consequent rapid neurological involvement, resulting in a fatal outcome.

**Case presentation**

A 40-day old male infant was admitted to the Emergency Department (ED) with bronchiolitis and dyspnea. He showed no comorbidities during pregnancy, he was born at full-term and normal.

At the time of admission, the patient was reported to be coughing for a week, worsening with a vomiting episode in the previous two days and reduction of feeding. Diuresis was regular. At the same time, he was treated with nasal and aerobic salbutamol administration. Patient's resistance was regular. At the same time, he was treated with nebulized epinephrine and prepared for endotracheal intubation, however he died before this could be done.

The physical examination highlighted persistent dry cough, globus abdomen, marbled skin with thoracic rush, respiratory rate of 52 breaths/minute, heart rate of 160 beats/minute and arterial oxygen saturation (SaO2) of 93%. No sign of neuropathological damage was evident. A nasopharyngeal swab performed with molecular assay Xpert® Xpress Flu/RSV (Cepheid, USA) tested positive for RSV. No Flu A and B were detected. During observation, SaO2 remained 93% until one hour and half from admission in ED when, due to hypoxaemia (SaO2 89%) and respiratory acidosis (pH7.347; pCO2 45.5 mmHg) (Table 1, BG1), the patient was given low-flow oxygen therapy (1 l/minute). After an initial improvement in clinical parameters (respiratory rate: 40 breaths/minute; SaO2 96%), flow oxygen therapy was reduced and stabilized to 0.5l/minute until 12h from admission in ED. Within this time frame, SaO2 was observed to be settled on 96–97%. Subsequently, a sudden worsening of respiratory dynamics happened (respiratory rate: 60 breaths/minute; SaO2 92%) and the therapy was changed to high-flow oxygen administration (8l/minute). The subsequently BG2 performed after oxygen administration showed a correction in the parameters (pH7.447; pCO2 45.5 mmHg) (Table 1, BG2). Nevertheless, three hours after BG2, despite an improvement in the respiratory dynamics (respiratory rate: 48 breaths/minute; SaO2 96%), a re-evaluation of the patient’s parameters was performed showing again hypercapnia and respiratory acidosis (pH7.336; pCO2 59.2 mmHg) (Table 1, BG3).

Moreover, the chest Xrays performed at time of BG3 and two hours apart highlighted how fast was the clinical worsening: the first one (Fig. 1a) showed normo expanded lungs with no opacification, while the second Xray (Fig. 1b) showed diffuse bilateral pulmonary opacification with marked ectasia of the stomach.

Because of the persistent respiratory failure, the patient was treated with nebulized epinephrine and prepared for endotracheal intubation, however he died before this could be done.

Postmortem examination was carried out on the infant’s body.

Lung’s microscopic examination revealed severe vascular congestion with massive bleeding, peribronchiolitis due to lymphocytic infiltrates and sporadic foci of pneumonia with reactive interstitial infiltrates and alveolar atelectasis (Fig. 2).

Also, extensive injury was present in the brain, characterized by white matter degradation in the left hemisphere, basal ganglia, hippocampus, bulb region and pons with edema (Fig. 3a).

A detailed microscopic analysis showed hypertrophic astrocytes, acutely damaged glia and focal necrosis at the bulb level (Fig. 3b), features reported to be associated

| BG | Time from admission (hh:mm) | Temperature | pH | pCO2 | pO2 | BEecf | HCO3 | tCO2 | SvO2 | HCT | HB | %Pcv |
|----|-----------------------------|-------------|----|------|------|--------|------|------|------|-----|----|------|
| 1  | + 01:30                     | 37.0        | 7.347 | 56.6 | 28   | 5      | 30.8 | 32   | 48   | 40  | 13.6 | 5    |
| 2  | + 12:30                     | 37.0        | 7.447 | 45.5 | 39   | 7      | 31.4 | 33   | 75   | 37  | 12.6 | 6    |
| 3  | + 16:00                     | 37.0        | 7.336 | 59.2 | 44   | 6      | 31.7 | 33   | 75   | 31  | 10.5 | 6    |

**Table 1** Results of the laboratory tests

BG Blood Gas, pCO2 partial pressure CO2, pO2 partial pressure O2, BEecf Base Excess in the Extracellular Fluid Compartment, tCO2 total CO2, SvO2 Venous oxygen saturation O2, HCT Hematocrit test, HB Hemoglobin, %Pcv packed-cell volume
with viral infection [12]. Basal ganglia were affected by neuronal necrosis with perineuronal halo and white matter strongly damaged (Fig. 3c, d). The brain's histological examination didn't exclude injuries in the cardiovascular center, probably due to viral infection.

No narcotic or psychotropic drugs were detected in the blood.

Microbiological analysis performed on autopic samples confirmed the viral etiology of brain damage and death. In fact, using molecular assay BioFire® FilmArray® RP2 Panel Plus (BioMérieux, France), RSV was detected in tracheobronchial aspirate, meningeal swabs, pericardic and abdominal fluids, lung and brain biopsies. On the contrary, the samples tested negative for Adenovirus, Coronavirus, Human Metapneumovirus, Human Rhinovirus/Enterovirus, Influenza A and B, Human parainfluenza virus, Bordetella pertussis, Bordetella parapertussis, Chlamydophila pneumoniae and Mycoplasma pneumoniae. Moreover, the microbiological cultures found neither fungal nor specific pathogenic bacteria.

Therefore, post-mortem examination revealed a severe bronchiolitis resulting in a fatal cerebral involvement due to RSV and its neurotropic behaviour.

Discussion and conclusions

Respiratory Syncytial Virus is usually associated with respiratory diseases whose most common symptoms are fever, cough, wheezing and it typically affects children during the first two years of life. However systemic dissemination of RSV during severe disease can lead to cerebral involvement with sudden infant death [13]. The pathogenesis of RSV-related neurological damage is not yet fully understood but it has been hypothesized that RSV may enter the Central Nervous System through the hematogenous/blood-brain barrier route causing the release of several humoral neurotoxic cytokine mediators. Anyways, the direct role of RSV in inducing encephalopathy was supported by the detection of RSV antibodies or viral genome in cerebrospinal fluids (CSF) [5, 14].

Morichi et al. (2011) classified RSV-related encephalopathies into four groups (metabolic error, cytokine storm, excitotoxicity and hypoxic type) and reported that the brain imaging during RSV infection showed massive cerebral edema with subsequent diffuse brain atrophy [15]. Several other studies reported neurological
Fig. 3  Macroscopic and microscopic brain damage. \textbf{b c} 150X magnification, \textbf{d} 125X magnification; Leica DM 2000 Microscope with Leica DFC280 digital camera; the scale bar represents 100 μm.

### Table 2  Clinical features of documented neurological manifestation with RSV confirmed infection at time of admission

| Source, y      | Age / Sex | Clinical picture at time of admission | CNS involvement at time of admission | Diagnosis of RSV infection | Outcome                                      |
|----------------|-----------|---------------------------------------|-------------------------------------|----------------------------|----------------------------------------------|
| Xu L et al., 2018 [14] | 2 years / F | Respiratory and cardiac arrest        | extensive brain edema                | Antigen rapid test        | Death for multiple organ failure            |
| Morichi et al., 2011 [15] | 1 year / M | CHARGE syndrome                       | involuntary movements, impaired consciousness | RT-LAMP                   | Survived with mental retardation            |
|               | 11 months / F | N/A                                   | status epilepticus, impaired consciousness | RT-LAMP                   | Survived with mental retardation            |
|               | 3 years / M   | N/A                                   | generalized tonic-clonic seizure    | RT-LAMP                   | Survived                                     |
|               | 1 year / F    | N/A                                   | generalized tonic-clonic seizure    | RT-LAMP                   | Survived                                     |
|               | 3 years / M   | N/A                                   | Nuchal rigidity, generalized tonic-clonic seizure | RT-LAMP                   | Survived                                     |
|               | 10 days / M   | N/A                                   | RT-LAMP                             | RT-LAMP                   | Survived                                     |
|               | 4 months / M  | cardiopulmonary arrest                | RT-LAMP                             | Survived with mental retardation |
|               | 27 days / M   | respiratory failure                   | RT-LAMP                             | Survived with mental retardation |
|               | 14 days / F   | Apnea                                 | RJ-LAMP                             | N/A                       |
| Otake et al., 2007 [16] | 11 months / M | Fever, cough                          | generalized tonic-clonic seizure    | Antigen rapid test        | N/A                                          |
| Zlateva et al., 2004 [17] | 4 months / M | Fever, cough, tachycardia             | none                                | RT-PCR                    | Survived                                     |
| Present report | 40 days / M  | Dyspnea                               | convulsions                         | RT-PCR                    | Death for multiple organ failure            |

*P/A data not available, RT-LAMP Reverse transcription loop-mediated isothermal amplification, RT-PCR Reverse transcriptase-polymerase chain reaction*
complications of RSV infection, which mainly include central apnea, seizures and encephalopathy [14–17].

According to the clinical pictures described above, the index patient had an RSV infection that dramatically worsened within hours leading to cerebral edema and death. Detection of RSV genome in CSF confirmed neurological involvement.

Also noteworthy, is the finding of the virus in the pericardic fluid that could suggest a myocardial damage as probably contributing cause of death.

Unlike the previously described clinical cases, in which patients reported a confirmed RSV infection together with at least one cerebral event (Table 2), in our report the infant showed no neurological signs and symptoms at the time of admission. For this reason and due to the rapid worsening of the patient's conditions, CSF sample for cytokines and nitrogen oxide determination was not collected.

Respiratory Syncytial Virus usually affect respiratory tract but, although rare, it can also determine a widespread organ involvement, including the brain and heart. Cerebral damage caused by RSV and perhaps concomitant cardiac involvement may led to sudden cardiac arrest in infants with bronchiolitis. Our findings, in conjunction with the above reported cases underline the need for clinicians to pay more attention and awareness to neurological sequelae of RSV infection, even in the absence of evidence of cerebral damage.

Abbreviations
RSV: Respiratory Syncytial Virus; CX3CR1: Chemokine receptor for CX3C; ED: Emergency Department; SaO2: Arterial oxygen saturation; BG: Blood Gas; pCO2: partial pressure CO2; pO2: partial pressure O2; BE: Base Excess in the Extracellular Fluid Compartment; tCO2: total CO2; SvO2: Venous oxygen saturation; HCT: Hematocrit test; HB: Hemoglobin; PCV: Packed-cell volume; CSF: Cerebrospinal fluids; RT-LAMP: Reverse transcription loop-mediated isothermal amplification; RT-PCR: Reverse transcriptase-polymerase chain reaction.

Acknowledgements
Not applicable.

Authors’ contributions
PB, RM and LP collected the data and wrote the manuscript, AB and GB provided microbiological and histological data, EZ and FS participated to interpretation of data, CC and RC participated to interpretation of data, supervised the work and revised the manuscript. All authors have read and agreed to the final version of the manuscript.

Funding
Not applicable.

Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations
Ethics approval and consent to participate
Not applicable.

Consent for publication
Written informed consent for clinical practice and use of clinical details for publication was obtained from relatives of the patient. All sensitive data related to the patient were anonymized.

Competing interests
The authors declare that they have no competing interests.

Author details
1 A.O.U. “Città della Salute e della Scienza di Torino”, S.C. Microbiology and Virology U., Corso Bramante n. 88, 10126 Turin, TO, Italy. 2 A.O.U. “Città della Salute e della Scienza di Torino”, Pathological anatomy and histology U., Turin, Italy.

Received: 24 August 2021   Accepted: 1 December 2021

Published online: 09 December 2021

References
1. Griffiths C, Drews S, Marchant D. Respiratory syncytial virus: infection, detection, and new options for prevention and treatment. Clin Microbiol Rev. 2017;30(1):277–319.
2. Obando-Pacheco P, Justicia-Grande A, Rivero-Calle I, Rodríguez-Teniente C, Sly P, Ramilo O, et al. Respiratory syncytial virus seasonality: a global overview. J Infect Dis. 2018;217(9):1356–64.
3. Eiland L. Respiratory syncytial virus: diagnosis, treatment and prevention. J Pediatric Pharmacol Therapeutics. 2009;14(2):75–85.
4. Antonucci R, Chiappe S, Porcella A, Rosatelli D, Fanos V. Bronchiolitis-associated encephalopathy in critically-ill infants: an underestimated complication? J Matern Fetal Neonatal Med. 2010;23(5):431–6.
5. Eisenhut M. Cerebral involvement in respiratory syncytial virus disease. Brain and Development. 2007;29(7):454.
6. Bohmwald K, Gálvez N, Ríos M, Kalengis A. Neurologic alterations due to respiratory virus infections. Front Cell Neurosci. 2018;12:386.
7. Kho N, Kerrigan J, Tong T, Browne R, Knillians J. Respiratory syncytial virus infection and neurologic abnormalities: retrospective cohort study. J Child Neurol. 2004;19(11):859–64.
8. Sweetman L, Ng Y, Butler I, Bodensteiner J. Neurologic complications associated with respiratory syncytial virus. Pediatr Neurol. 2005;32(5):307–10.
9. Ng Y, Cox C, Atkins J, Butler I. Encephalopathy associated with respiratory syncytial virus bronchiolitis. J Child Neurol. 2001;16(2):105–8.
10. Espinoza J, Bohmwald K, Cespedes P, Gomez R, Riquelme S, Cortes C, et al. Impaired learning resulting from respiratory syncytial virus infection. Proc Natl Acad Sci. 2013;110(22):9112–7.
11. Li X, Fu Z, Alvarez R, Henderson C, Tripp R. Respiratory syncytial virus (RSV) infects neuronal cells and processes that innervate the lung by a process involving RSV G protein. J Virol. 2006;80(1):537–40.
12. Soung A, Klein R. Vinal encephalitis and neurologic diseases: focus on astrocytes. Trends Mol Med. 2018;24(11):950–62.
13. Eisenhut M. Extrapulmonary manifestations of severe respiratory syncytial virus infection—the systematic review. Crit Care. 2006;10(4):R107.
14. Xu L, Gao H, Zeng J, Liu J, Lu C, Guan X, et al. A fatal case associated with respiratory syncytial virus infection in a young child. BMC Infect Dis. 2018;18(1):217.
15. Morichi S, Kawashima H, Ioi H, Yamanaka G, Kashiwagi Y, Hoshika A, et al. Classification of acute encephalopathy in respiratory syncytial virus infection. J Infect Chemother. 2011;17(6):776–81.
16. Otake Y, Yamagata T, Morimoto Y, Imi M, Morii M, Alhara T, et al. Elevated CSF IL-6 in a patient with respiratory syncytial virus encephalopathy. Brain and Development. 2007;29(2):117–20.
17. Zlateva K, Van Ranst M. Detection of subgroup b respiratory syncytial virus in the cerebrospinal fluid of a patient with respiratory syncytial virus pneumonia. Pediatr Infect Dis J. 2004;23(11):1065–6.