Evaluation of Press Mud, Vinasse Powder and Extraction Sludge with Ethanol in a Pyrolysis Process

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Received: 27 August 2019; Accepted: 17 September 2019; Published: 30 October 2019

Abstract: The effluents of the sugar and bio-ethanol industry, mainly vinasse as well as lignocellulosic waste, are produced in high volumes. Therefore, their treatment and valorization would reduce the environmental impact and make this industry more productive and competitive. The purpose of this study was to determine the potential use of press mud (lignocellulosic waste), vinasse powder, and vinasse sludge from an extraction process with ethanol, as raw materials for conventional pyrolysis evaluating the physicochemical characteristics that affect this thermochemical process, such as calorific power, density, ash content, volatile material, moisture and nitrogen, sulfur, carbon and hydrogen content, thermogravimetric profile, and quantification of lignin cellulose and hemicellulose. The batch pyrolysis experiments showed that all three wastes could be converted successfully into more valuable products. The powder vinasse led to the formation of the lowest content of bio-char (42.7%), the highest production of volatiles (61.6 wt.%), and the lowest ash content (20.5 wt.%). Besides, it showed the high heating value of 15.63 MJ/kg. Meanwhile, the extraction sludge presented the highest liquid yield (32%) with the lowest gas formation (18.2 wt.%) and the lowest heating value of 8.57 MJ/kg. Thus, the sludge could be a good feedstock for production of bio-oil and bio-char.

Keywords: lignocellulosic waste; pyrolysis; biowaste; sugar cane

1. Introduction

According to the most recent report of the United States Department of Agriculture (USDA), the world’s sugar production was 178.926 million metric tons for the marketing year 2018/19, with a
stable trend for the past four years [1]. Colombia is within the first 20 countries in the world in terms of sugar production, with 2.4 million tons in the 2018/19 year (Figure 1) [1] and with a well-developed industry that takes advantage of having the world highest productivity, approximately 16.2 tons of sugar per hectare per year [2]. The sugarcane industry is concentrated in the south-western region of the country, specifically in the geographic valley of the Cauca river, with a planted area of 243,232 hectares and 13 industrial sugar mills that generate 286,000 direct and indirect jobs [2].

![Figure 1. Sugar production and consumption worldwide (a) and in Colombia (b) [1].](image)

The productive process of the sugar industry is intensive in the generation of solid and liquid sub-streams. The major solid wastes generated are bagasse and press mud. The bagasse, which is the residue that remains after the sugarcane milling, is produced in a proportion of 0.30–0.34 tons per 1 ton of sugar cane processed [3] and has a high content of cellulose (23%–45%), hemicellulose (12%–28%), and lignin (10%–20%) [3,4]. It is also composed of sugar (5%) and minerals (1%) [3]. Owing to its calorific power, bagasse is mostly used in energy cogeneration in the sugar mills, although a broad range of uses has been proposed for this material; however, most of them have not been commercially exploited [5]. One particular company in Colombia uses 15% of the bagasse to produce paper without mixing with other types of vegetable fibers. The remaining 85% is used in energy cogeneration in the sugar mills [6].

The press mud or filter cake, which is the waste obtained from the clarification and filtration processes of the sugar juice, is produced in a range of 0.01 to 0.07 tons per ton of ground sugarcane [4]. The composition of the press mud is very variable; however, it is characterized by a high content of proteins, sugar, and fiber. In fact, the press mud in Colombia is used in a composting process to produce fertilizers. In recent years, there have been studies reporting different uses for valorization of these solid wastes beyond the agronomic use [5], either as a source of renewable energy (biogas production) [7] or products of greater added value in areas like the pharmaceutical and food industries [3,4].
In addition, the vinasse produced in the distillation towers is the main liquid effluent of the bio-ethanol sub-process from sugarcane [8]. It presents a dark brown color and generates ethanol in a 10 to 1 volumetric ratio (vinasse/ethanol). It is characterized by low pH values, high chemical oxygen demand (COD: 32,000–109,700 mg/L), and biological oxygen demand (BOD₅: 13,414–87,700 mg/L) [9,10]. Vinasse is composed mainly of water; organic solids; and minerals like potassium, calcium, and magnesium [10]. At present, there are different ways for the treatment or disposal of the vinasse such as composting, concentration by evaporation, and fertigation, among others. Concentration by evaporation (concentrating vinasse from 5% in solids to 20%–50%) and composting are the most viable options applied in Colombia. In composting, the organic substrate is degraded, and the water evaporated, resulting in a stable solid product that is rich in nutrients for the soil; however, this process has low added value, and requires a long residence time (more than 40 days) and large land areas—the high volumes of lignocellulosic material (crop residues and press mud) require to be mixed with vinasse in a proportion of 80%–90% solid waste to 10%–20% vinasse to achieve the desired product [11]. Meanwhile, fertigation, which is the direct disposal of vinasse into soils widely used in Brazil, has the purpose of recovering nutrients (mainly the potassium) and reusing the water to meet the nutritional requirements of sugarcane [12]. However, it has raised concerns regarding the ability of the soils to assimilate the high dosages (100–150 m³/ha) that may result in the unbalance of soil nutrients and groundwater contamination [11]. In fact, vinasse management is still an unsolved problem for many sugar mills, and researchers suggest that new studies on green methods need to be developed aiming at the recycling and disposal of sugarcane vinasse [11].

An alternative for the sugarcane industry wastes’ valorization is the pyrolysis process, which can convert the residues into high added-value products. The process consists of the thermal decomposition of materials in the absence of oxygen. The advantages of biomass pyrolysis are the production of both valuable liquid and solid products, as well as a non-condensable gas fraction. The liquid fraction, bio-oil, can be used in combustion engines and gas turbines [13]. Moreover, the liquid pyrolysis fraction can be used for production of various speciality chemicals [14,15]. Bio-char, the solid fraction, has several applications as a fuel (substitute for coke), activated carbon (adsorbent), or fertilizer for soils (hosting microbes, slowly releasing nutrients and water) [14]. The variables of biomass pyrolysis to achieve desired relative amounts of fractions in the pyrolysis process have been intensively studied, including the type of pyrolysis applied (slow, moderate, fast), reactor design, influence of biomass composition, moisture, size, and operating conditions (temperature, heating rate, gas flow, catalyst type, and loading) [15,16].

Previous studies have shown the feasibility of using pyrolysis for sugarcane bagasse, demonstrating that the bio-oil can be potentially valuable as a renewable fuel and can be used as a chemical feedstock [17,18], while the bio-char can be used as a solid fuel, a cheap adsorbent, feedstock for activated carbon production, and in agricultural applications [19–22]. However, studies for press mud [23,24] and vinasse [25,26] pyrolysis are scarce. One of the reasons is because of the additional energy needed to dry both wastes, especially the vinasse, as samples prior to pyrolysis should be dried to a low moisture content to minimize the water content in the resulting bio-oil.

The high organic content in dried vinasse as well as in press mud, and their high availability (only in Colombia, the annual press mud and concentrated vinasse—40% to 55% solids—production can be estimated in 1.4 million tons and 2.7 million tons per year, respectively [2,5,27,28]) suggest an interesting opportunity for these biomass wastes to be used as feedstock in a thermochemical valorization process. Therefore, the objective of this work is to describe a non-catalysed slow pyrolysis study using a batch reactor unit and three biomass waste samples produced in an industrial sugar and bio-ethanol plant. The biomass samples were characterized prior to pyrolysis tests, and the effect of biomass type on the yield and chemical composition of bio-oil and bio-char was investigated.
2. Materials and Methods

2.1. Raw Material

The press mud (Figure 2a) was obtained from the clarification and filtration stage of a commercial sugar mill located at the Cauca River valley (Colombia). The sample had an initial humidity of 62.8%. Preconditioning of the press mud was carried out according to ASTM E 871 standard—drying the sample at 105 °C for one hour. Milling and sieving were carried out to obtain a press mud sample with a particle size smaller than 0.5 mm.

![Figure 2. Biomass waste samples from bioethanol processes. (a) Press mud; (b) powder vinasses; (c) extraction sludge.](image)

Two different solid vinasse samples were studied. The first one was an extraction sludge (Figure 2b) from a protein denaturation process described by Yadav and Chandra [29], in which ethanol and vinasse are mixed in a 500 mL separating funnel in equal volumetric proportions [30]. The mixture was stirred, and the solids were allowed to settle for two hours. Subsequently, the supernatant was filtered with a qualitative filter, the sludge was dried at room temperature for two hours, and the ethanol in the supernatant was separated by distillation.

The second sample was a powdered vinasse (Figure 2c) supplied by Biogreen Corporación S.A.S with a humidity of 5.6% and a maximum particle size of 80 mesh. The sample was obtained by drying the concentrated vinasse (50% in solids) by means of a patented industrial process [31].

Figure 3 shows the sugar and bio-ethanol process block diagram, including the vinasse powder and the vinasse sludge production sections.
2.2. Pyrolysis Tests

All pyrolysis tests were carried out in a batch scale reactor unit. The laboratory-scale pyrolysis unit (Figure 4) consists of a stainless steel reactor with 50 mm internal diameter and 300 mm length, a cooling unit formed by four condensers, and a gas scrubber to retain any other volatile organic compounds that did not condense in the cooling system. While the first two condensers are at room temperature (first condenser to retain any biomass particles that might be driven by gas flow, second condenser to trap the heaviest fraction), the third condenser is cooled with ice, and the fourth condenser is cooled with liquid nitrogen. The reactor is heated inside an electrical furnace fitted with a thermocouple and a temperature controller. For all experiments, 50 g of each biomass sample was loaded directly in the reactor. The loaded reactor was sealed and placed inside the electrical furnace. The reactor was externally heated from room temperature to 450 °C at a heating rate of 7–8 °C/min. When the temperature reached 100 °C, nitrogen flowing at a rate of 50 L/h was injected through the reactor. The pyrolysis tests were performed at 450 °C for 2 h to ensure maximum conversion of biomass waste particles. Afterward, the reactor was cooled down to 100 °C under a nitrogen flow of 50 L/h and then it was allowed to reach room temperature overnight. During pyrolysis and cooling of the reactor,
the liquid fraction was collected in condensers. After cooling, the reactor was removed from the electrical furnace.

![Figure 4. Schematic diagram of the slow pyrolysis unit.](image)

The amount of bio-char and condensed liquid from each condenser was measured, and the liquid products were afterwards recovered from each condenser with ethanol 95% and acetone 98%. The measure of bio-char and condensed liquid allowed the computation of the material balance.

### 2.3. Analysis

The proximate and elementary analyses on biomass samples were carried out in accordance with the following standards: ASTM D 3302 for moisture, ASTM D 3175 for volatile material, ASTM D 5865 for high heating value, ASTM D 3174 for ash content, ASTM D 3172 for fixed carbon content, ASTM D 4239 for sulfur content, ASTM D 5373 for nitrogen and oxygen content, and ASTM D 5865 for low heating value. The cellulose and hemicellulose content was obtained according to the SCAM-CM 71:09 procedure by hydrolysis with H$_2$SO$_4$, followed by the measurement of carbohydrates through high performance liquid chromatography (HPLC) system equipped with refractive index detector and the following columns: Shodex sugar SP0810 or Biorad Aminex HPX-87P column (or equivalent) with ionic form H$^+$/CO$_3$ deashing guard column and Biorad Aminex HPX-87H column (or equivalent) with corresponding guard column. The lignin content was determined by extraction of the solid samples with ethanol according to TAPPI 204 standard and then subjecting the sample free of extractive components, according to the TAPPI 222 standard, to two acid hydrolysis to finally dry, weigh, and measure the lignin using a UV spectrophotometer [32].

The thermogravimetric (TGA) analysis was performed with an STD Q 600 equipment using 10 mg of samples heated in a nitrogen atmosphere up to 1000 °C, at a heating rate of 20° C min$^{-1}$.

For the pyrolysis tests, the moisture content of bio-char was determined by the oven drying method, according to ASTM D 4442-07.

The FT-IR spectra were used to obtain the chemical structure of the waste biomass samples utilized in the pyrolysis tests, as well as their corresponding bio-char. The samples were analysed in a Bruker Vector 22 spectrometer at 4 cm$^{-1}$ resolution using the KBr pellet method at room temperature; 20 scans were single-averaged in the 4000–600 cm$^{-1}$ region. Baseline and smooth corrections of the spectra were performed for each analysis.

The chemical composition of pyrolysis bio-oils was determined using a GC-2010 Shimadzu gas chromatograph connected to GCMS-QP2010 Plus mass spectrometer (MS). A capillary column HP-1
(100% dimethylpolysiloxane) of 25 m with 0.2 mm ID and thickness of 0.33 µm was used for all the analyses. The temperature program started heating up to 40 °C for 5 min; then, the temperature was increased from 40 °C to 300 °C with a ramp of 5 °C min⁻¹. The injection temperature was set at 250 °C. The MS was operated in the electron ionization mode within the 35–650 m/z range. The ion source temperature was set to 220 °C, and the interface temperature to 250 °C.

3. Results and Discussion

3.1. Analysis of Raw Materials

Table 1 shows the proximate and elementary analysis of the press mud, the extraction sludge, and the vinasse powder samples.

| Analysis                        | Method       | Press Mud | Vinasse Powder | Extraction Sludge |
|---------------------------------|--------------|-----------|----------------|-------------------|
| Moisture (% wt)                 | ASTM D 3302  | 2.06      | 5.60           | 7.53              |
| Volatile material (% wt)        | ASTM D 3175  | 40.67     | 61.66          | 47.79             |
| High heating value (MJ/kg)      | ASTM D 5865  | 8.57      | 15.63          | 9.27              |
| Ash (% wt)                      | ASTM D 3174  | 53.83     | 20.56          | 40.13             |
| Fixed Carbon (% wt)             | ASTM D 3172  | 3.45      | 12.18          | 4.55              |
| Sulfur (% wt)                   | ASTM D 4239  | 0.13      | 1.47           | 0.85              |
| Nitrogen (% wt)                 | ASTM D 5373  | 0.81      | 1.84           | 2.73              |
| Carbon (% wt)                   | ASTM D 5373  | 21.50     | 34.99          | 22.97             |
| Hydrogen (% wt)                 | ASTM D 5373  | 3.33      | 4.75           | 3.87              |
| Oxygen (% wt)                   | ASTM D 5373  | 18.34     | 30.78          | 21.91             |
| Low heating value (BTU/lb)      | ASTM D 5865  | 3349      | 6212           | 3545              |

The high heating value (HHV) of the press mud was 8.57 MJ/kg, which is lower than values previously reported in the literature (17.0 MJ/kg [7], 12.5 MJ/kg [24]). This may be because of the widely variable composition of press muds. In previous studies, lower ash contents were reported, likely owing to different technologies used in the industrial filtration stage [33]. Similarly, the extraction sludge also has a relatively low HHV of 9.27 MJ/kg, probably owing to its moisture (7.53%) and rather high ash content (40.13%).

Vinasse powder has the highest HHV, which is similar to reported values for bagasse (18.9 MJ/kg [24]) and other lignocellulosic wastes like rice husk (14.09 MJ/kg) and palm kernel shell (18.96 MJ/kg) [28]. In addition, the vinasse powder has the highest proportion of volatile material, implying higher yields of light bio-oil fractions and gas [28]. As mentioned before, the press mud and the extraction sludge present the highest ash content, which adversely affects the high heating value, while the vinasse powder has an ash content of 20.56%, which is in the range of feedstock material for pyrolysis and gasification processes [24]. This is an indication that powder vinasse has a promising potential as feedstock for pyrolysis.

The results of the fixed carbon, oxygen, and carbon are also higher for the powder vinasse with respect to those of the press mud and those of the extraction sludge, which indicates better yields of the powder vinasse either to form bio-char or solid fuels. The high content of oxygen and carbon suggests the presence of organic groups characteristic of lignocellulosic materials [22]. On the other hand, the sulfur and nitrogen values of both press mud and powder vinasse are low, being consistent with values reported in the literature [34,35].

It can be seen in Table 2 that the lignin content for press mud, powder vinasse, and extraction sludge is high in proportion to the cellulose and hemicellulose, while bagasse is high in cellulose and...
hemicellulose [28]. A higher lignin content relates to the formation of increased bio-char yield during pyrolysis. It is also expected that the press mud and the powder vinasse, in particular, degrade at higher temperatures owing to the high content of lignin, which decomposes in a temperature range of 395–500 °C [7].

| Component          | Press Mud | Powder Vinasse | Extraction Sludge |
|--------------------|-----------|----------------|------------------|
| Lignin (%)         | 17.4      | 23.0           | 12.9             |
| Cellulose (%)      | 12.8      | 12.7           | 7.5              |
| Hemicellulose (%)  | 6.5       | 8.7            | 5.0              |

The thermal behavior of the press mud is shown in Figure 5, which shows the mass loss (TGA) and derivative mass loss (DTA) profiles as a function of temperature, within the temperature range of 25–1000°C. From this figure, the mass loss of press mud can be divided into four phases: the first phase, 25 °C to 150 °C, was attributed to the elimination of the free and bound moisture present in the material [24]. Then, during the second step, a significant loss of the mass (~32%) was observed ranging from 150 °C to 350 °C. In this range, the derivative thermogravimetry (DTG) curve shows a peak at 310 °C. This behavior is the result of the decomposition of hemicellulose and cellulose [26]. Afterward, lignin decomposition occurred in the 350–470 °C range with a mass loss of ~13%. In the last phase, the slope presented small variations corresponding to the degradation of oxygenated compounds that are stable at a high temperature. The residue of the sample is left as char.

The thermogravimetric analysis of vinasse powder is presented in Figure 6. The overall mass loss of vinasse can be divided into five phases. The first phase corresponds to the removal of free moisture, bond moisture, and evaporation of volatile matter in the range of 25–100 °C and 100–200 °C, respectively. In the second phase, there occurs the main volatilization, and a mass loss of ~30% is achieved. Then, a mass loss of ~5%, followed by a weight loss of 30% with a decomposition peak temperature at 475 °C, can be seen in the range of 400–475 °C. Finally, no mass loss was detected up to 1000 °C. Vinasse decomposed at higher temperature than the press mud.

Figure 7 shows the TGA–DTG curves for the extraction sludge. The thermal behavior of the sludge presented seven phases. Four phases were observed between 25 and 150 °C, indicating ethanol
and water evaporation. Two more phases were identified in the range of 150–550 °C with a mass loss of ~45%. Further, above 550 °C, the mass loss was negligible. The DTG curve for the extraction sludge displays several decomposition steps; the peak at 300 °C is more likely owing to the decomposition of hemicellulose [36]. The press mud and the vinasse powder showed an active pyrolysis zone in the range of 200–470 °C, where maximum weight loss occurred, while the passive pyrolysis zone was in the range of 450 °C–650 °C for the press mud sample and in a wider range for vinasse. Regarding the latter, the thermal degradation of the sample was not complete, even at 1000 °C, owing to its complex composition. In contrast, the extraction sludge showed an active pyrolysis zone between 150 and 550 °C with a passive pyrolysis zone in a higher range (550–1000 °C). Similar results have been reported for the thermogravimetric behavior of the dried solid waste of post-methanated distillery effluent [37].

![Figure 6. Thermogravimetric analysis of vinasse powder.](image1)

![Figure 7. Thermogravimetric analysis of extraction sludge.](image2)
3.2. Slow Pyrolysis of Waste Biomass

3.2.1. Influence of Biomass Type on Pyrolysis Product Distribution

Figure 8 presents the difference in pyrolysis products yield when each type of biomass waste—press mud, powder vinasse, and extraction sludge—was used as feedstock for slow pyrolysis. The liquid product was divided into heavy fraction and water (collected in the first two condensers of the pyrolysis unit) and light fraction (collected in the other two condensers). The gas yield was calculated as the mass difference between feedstock, liquid, and char yields.

The results obtained show significant differences between the three waste biomass samples subjected to pyrolysis. It was observed that powder vinasse led to the formation of the lowest content of bio-char (42.7%) and the highest production of volatiles. The results are confirmed by the proximate analysis of the raw biomass samples where powder vinasse showed the highest volatiles content (61.6 wt.%) and the lowest ash content (20.5 wt.%).

Meanwhile, the extraction sludge presented the highest liquid yield (32%) with the lowest gas formation (18.2 wt.%), which is likely owing to the high water content in the extraction sludge (7.5 wt.%). This is consistent with the extraction sludge high heavy fraction yield (6.8 wt.%), low content of volatiles (47.8 wt.%), and high ash content (40.1 wt.%).

The press mud presented the greatest char yield (60.1%) owing to its initial high ash content (53.8%). The elevated char yield of all biomass samples could suggest an incomplete decomposition of the organic phase at 450 °C.

Similar product distributions of biomass wastes through slow pyrolysis were presented by other studies [38–42]. Agar et al. [38] converted wastewater sludge and organic fines from municipal wastewater treatment plants (similar volatiles and ash content as our biomass feedstock) in carbon-rich char through pyrolysis in a laboratory fixed-bed reactor at 600 °C and 700 °C. The product distribution of sewage sludge at 600 °C was gas fraction 19%, liquid 33%, and char 48%, while for solid organic fines, the product yields at the same temperature were gas 33%, liquid 14%, and char 53%. Even at 700 °C, the char produced was 45% for the sewage sludge feedstock, while the gas yield increased to 29%.

Williams and Besler [39] studied the influence of temperature and heating rate on the slow pyrolysis of biomass wood. The pyrolysis experiments were performed for 2 h at temperatures ranging from 300 to 720 °C, and heating rates of 5–80 °C/min. The wood biomass presented a moisture of 7 wt.%, volatiles of 92.2 wt.%, and ash of 0.8 wt.%. At 420 °C and a heating rate of 5 °C/min (similar to our experimental conditions), the compositions were 29.7% for char, 12.4% for oil, 35.9% for the aqueous fraction, and 21.5% for gas. At higher operating temperatures, the char proportion decreased to 23.2%, while the gas increased to 26.8%. Increasing the heating rate promoted the gas yield, but decreased the
oil yield. Al Arni [40] obtained a product distribution of 37.6% char, 26.1% liquid, and 25.1% gas from sugarcane bagasse (84 wt.% volatiles, 5.9 wt.% ash) by conventional pyrolysis at 480 °C, heating rates of 40–50 °C/min, and 1 h residence time.

3.2.2. Influence of Biomass Type on Bio-Char Composition

The bio-char obtained after each pyrolysis test was analyzed in terms of moisture, C, H, N, and S contents, as well as FTIR analysis. The compositions of the bio-char samples are shown in Table 3.

| Feedstock        | Moisture, wt% | C, %  | H, %   | N, %  | S, % |
|------------------|---------------|-------|--------|-------|------|
| Press mud        | 0.034         | 18.12 | 1.823  | 4.379 | 0    |
| Powder vinasse   | 0             | 38.804| 1.639  | 1.16  | 0    |
| Extraction sludge| 0.042         | 29.364| 1.594  | 0.16  | 0    |

After pyrolysis at 450 °C for 2 h, the moisture content in all char samples decreased to almost 0%. The results are also correlated with hydrogen content that decreased in all samples after pyrolysis. The sulfur content was not detected in any bio-char sample, suggesting that organic sulfur components were reduced to light fractions of the increase of gas yield bio-oil or gas. The char produced from powder vinasse was the richest in carbon (favorable as fuel and activated carbon). The increase of carbon content after pyrolysis was also observed by Parthasarathy and Sheeba [41] and Lee et al. [43]; these authors also associated the release of volatiles, such as hydrogen and oxygen by dehydrogenation and deoxygenation, to the relative increase of carbon content. In comparison, the pyrolysis of rubber wood at 550 °C showed a bio-char yield of 28.0% ± 0.35% and a moisture content of 7.5% [44]

FTIR spectra of each waste biomass sample and their corresponding chars are presented in Figure 9. Four regions were detected on FTIR spectra of all biomass feedstock samples: in the range of 3000–3800 cm⁻¹, between 2750 and 3000 cm⁻¹, a region between 1500 and 1800 cm⁻¹, and between 750 and 1250 cm⁻¹. The small peak that appears at 3750–3970 cm⁻¹ can be related to the presence of water in all biomass samples, while the broad peaks with maximum intensity around 3420 cm⁻¹ can be attributed to O–H stretching from aromatic and aliphatic alcohols, in particular, lignin [45]. The small peak from 2920–2930 cm⁻¹ is characteristic of C–H stretching of methyl and methylene groups from cellulose, hemicellulose, and lignin.

The peaks in the region of 1500–1750 cm⁻¹ correspond to C=O stretching of ketones, aldehydes, esters from hemicellulose, and lignin [45,46]. Powder vinasse also shows an intense peak at 1417 cm⁻¹, which is characteristic of C–H deformation of the aromatic ring from lignin. The presence of this peak at higher intensity in the case of the powder vinasse could be the result of the higher content of lignin in this biomass sample. The peaks from 1033–1180 cm⁻¹ can be attributed to C–O stretching from cellulose and hemicellulose.

Significant changes of FTIR spectra appear in the case of all bio-char samples obtained after slow pyrolysis at 450 °C for 2 h. The OH stretching band from 3750–3970 cm⁻¹ decreased drastically in the case of all char samples, disappearing completely in the case of char from extraction sludge (with the lowest content of lignin of 12.7%). The absence of a peak at the 2920–2930 cm⁻¹ range suggests a significant decomposition of the biomass material during pyrolysis [47]. While the bio-char produced from press mud still shows the peaks characteristic to C–O stretching at almost the same intensity as the feedstock, the other two char samples show a significant decrease of the peaks from this region, suggesting the higher degradation of the organic compounds for these two biomass samples.
3.2.3. Influence of Biomass Type on Bio-Oil Composition

The bio-oils produced from each biomass waste pyrolysis were analyzed in terms of chemical composition using the gas chromatography mass spectrometer (GC-MS) technique. For each test, the liquid phase was extracted separately from each condenser of the cooling unit and analyzed with GC-MS; the reason for analyzing separately was to observe the way the organic components condensed in the cooling unit, as well as to obtain a more precise identification of the organic components.

The oil collected from condenser 1 (heavy fraction and water collector) showed a wide variety of organic components in the case of the powder vinasse and the extraction sludge, and fewer components for the press mud (Table 4).
This section may be divided by subheadings. It should provide a concise and precise description of the experimental results, their interpretation, as well as the experimental conclusions that can be drawn.

Table 4. Composition of bio-oil collected on the first condenser (heavy fraction and water) of the pyrolysis unit.

| Press Mud                       | Rel. % | Powder Vinasse          | Rel. % | Extraction Sludge                        | Rel. % |
|--------------------------------|--------|-------------------------|--------|------------------------------------------|--------|
| Propanoic acid. 2-hydroxy-methyl ester | 2.54   | Propanoic acid. 2-hydroxy-methyl ester | 2.72   | Oxalic acid. dicyclobutyl ester         | 8.08   |
| 1-Hydroxy-2-butanol Propane. 2.2-diethoxy | 0.86   | 1-Hydroxy-2-butanol 2-Butanone compounds | 0.72   | Proline                                 | 5.9    |
| 2-Propanone. 1.1-dimethoxy | 0.43   | Pyrrolidine. 2-(hydroxymethyl)-1-cyclopentene compounds | 0.27   | Oxazole. Hexanedioic acid. monoethyl ester | 3.1    |
| 2-Cyclopenten-1-one compounds Furfural | 4.95   | 2-Cyclopenten-1-one compounds Acetamide | 3.74   | 4.5-dihydro-2,4,4-trimethylindolizine    | 0.36   |
| 2-Propanone. 1-(acetoxy) | 3.73   | Oxazole                  | 32.43  | 2-Pentanone. 4-hydroxy-4-methyl-Hexadecanoic acid. ethyl ester | 4.48   |
| 2-Furancarboxaldehyde Phenol compounds | 0.9    | Acetaldehyde. ethylidenehydrazone | 3.87   | Phenol compounds                        | 0.31   |
| Benzofuran. 2.3-dihydro Azidine Alkanes (C12–C18) | 3.33   | Phenol compounds         | 13.36  | Phenol compounds                        | 16.11  |
| Alkenes (C12–C15) | 0.28   | Benzo[1,3]diazin. 2,3-dihydro-Azidine compounds | 1.6    | Butanenitrile. 2-methyl-Azidine compounds | 2.52   |
| Hexadecanoic acid. methyl ester | 1.26   | 1,2-Ethanediol. monoacetate | 18.39  | 18.17                                   |        |
| n-Hexadecanoic acid | 0.27   | Alkenes (C12–C15)       | 0.47   | Alkenes (C12–C15)                       | 2.52   |
| 2-Furanmethanol | 0.15   | 2-Furanmethanol         | 5.37   | Alkenes (C13–C18)                       | 3.47   |
| Pyrazine compounds | 0.12   | Pyrazine compounds      | 1.61   | 2-Furanmethanol                         | 12.6   |
| 1,2-Cyclopentane-1,3-dimethyl Caprolactam | 1.32   | Pyrazine compounds      | 1.94   | Others                                 | 7.52   |
| others | 1.96   | Others                  | 1.96   | Others                                 | 9.04   |
| others | 9.04   | Others                  | 9.04   | Others                                 |        |

Similar compounds were obtained from all three biomass waste samples, although with significant differences in terms of concentration. The powder vinasse and the extraction sludge showed a high concentration of nitrogen compounds, related to the higher N content observed in these two samples. The liquid obtained from the press mud showed a significant amount of 2,2-dimethoxypropane (DMP)—the compound is generally formed from acetone and methanol at room temperature in the presence of an acidic environment [48]. Small amounts of 2-cyclopenten-1-one, phenolic compounds, as well as alkanes (C12–C18) were also observed.

In the case of the powder vinasse, oxazole and aziridine compounds (both with nitrogen in their structure) were in the highest concentration—50.8%, along with phenolic compounds (13.4%) and furanmethanol (5.4%). While the powder vinasse did not show any alkanes or alkenes, it contained a significantly higher concentration of aromatics than the liquid from the press mud. Similar results have been obtained by David et al. [49] from pyrolysis of sugarcane bagasse.

The extraction sludge showed the highest concentration of phenols (16.1%), aziridines and pyrazines (22.1%), and furanmethanol (12.6%), as well as the most elevated content of alkanes and
alkenes (11.6%). In the second condenser of the cooling unit, heavier organic compounds were observed in the composition of the pyrolysis liquids (Table 5).

Table 5. Composition of bio-oil collected from the second condenser of the pyrolysis unit.

| Press Mud                  | Rel.% | Powder Vinasse               | Rel.% | Extraction Sludge            | Rel.% |
|----------------------------|-------|-----------------------------|-------|------------------------------|-------|
| Phenol compounds           | 31.99 | Phenol compounds            | 54.61 | Phenol compounds             | 49.83 |
| Oxazole.4.5-dihydro        | 6.54  | 2-hydroxy                   | 8.24  | 3-Pent-2-one.4-methyl-       | 2.02  |
| 2-Cyclohexene-1-one comp. | 7.81  | 2,6-Butanediol.             | 1.09  | 2-Isopropylpyrazine          | 9.69  |
| Furfural                   | 1.31  | 4-hydroxy-4-methyl-         | 2.46  | 2-Pentanone.4-hydroxy-4-methyl- | 9.58  |
| 2-Propanone.               | 4.17  | 2-Furanmethanol             | 0.5   | 2-N-Cyano-2-methylpyrrolidine | 1.48  |
| 1-(acetyloxy)              |       |                             |       |                              |       |
| Cyclooctane                | 2.19  | 2-Propen-1-amine.N.N-bis(1-methyls) | 1.47 | Toluene                      | 2.66  |
| 2-Furan-2-one              | 1.53  | 1,2-dimethyl-2,3-dihydro    | 0.35  | 1.6-Dioxaacyclododecane-7,12-dione | 1.44 |
| Benzofuran.2,3-dihydro     | 3.09  | 2-Cyclohexene-1-one comp.   | 4.97  | Hexanediolic acid.ethyl.methyl es | 0.46 |
| 1.2-Indole                 | 3.74  | 1.2-Indole                  | 1.88  | Indolizine                   | 6.11  |
| Alkanes (C13–C22)          | 25.09 | Alkanes (C19–C32)          | 4.29  | Alkanes (C16–C25)           | 6.68  |
| Alkenes (C14–C16)          | 3.19  | 2-Amino-5,6-dihydro-4,4,6-trimethyl-4H-1,3-oazaine | 4.12 | Alkenes (C16–C25)           | 2.57  |
| Palmitic acid.             | 1.2   | Indole compounds            | 2.29  | 1H-Indole.3-methyl-          | 1.51  |
| methyl ester               |       |                             |       | Others                       | 2.85  |
| Hexadecanoic acid.         | 0.63  | 4-Piperidinone.             | 1.03  |                             |       |
| propyl ester               |       | 2.2,6,6-tetramethyl-Thymol  | 0.66  |                             |       |
| Others                     | 5.81  | Others                      | 11.82 |                             |       |

For all of the samples, the main components of the liquid collected from the second condenser were phenolic compounds that were produced by the degradation of lignin. Powder vinasse and extraction sludge showed 50% of phenolic compounds, which is a very similar result to those reported in the literature [49–51]. The liquid fraction obtained from the press mud pyrolysis contained the highest proportion of alkanes (25.1%), although powder vinasse presented heavier alkenes, up to C32. A higher content of ketones (aliphatic and cyclic) was formed when the press mud (13.7%) and the extraction sludge (14.7%) were used for pyrolysis.

The liquid extracted from the last two condensers showed a variety of components, including heavy and volatile compounds. A significant amount of phenols was still observed in the liquid collected at the third condenser.

The liquid from the press mud collected in the third condenser (Table 6) also showed a high content of alkanes, phenols, and ketones, similar to the compounds gathered in the previous condenser. For the powder vinasse, the main components observed were ketones (38%); phenols (16.1%); and organic compounds based on nitrogen (17.6%)—azirides, pyrroles, piperidines, and pyrazines. The extraction sludge showed a lower content of ketones (17.5%) and phenols (15.7%), but a higher content of organic nitrogen compounds (29.9%) and alkanes (6.5%). The liquids obtained from the pyrolysis of the press mud and the extraction sludge seemed to be, in general, heavier than that obtained from powder vinasse.
Table 6. Composition of bio-oil collected at the third condenser of the pyrolysis unit.

| Press Mud                          | Rel.% | Powder Vinasse                      | Rel.% | Extraction Sludge           | Rel.% |
|-----------------------------------|-------|-------------------------------------|-------|-----------------------------|-------|
| Propane. 2,2-diethoxy             | 8.26  | Propanoic acid. 2-hydroxy-         | 0.52  | Cyclopentaneone             | 1.20  |
| 2-Cyclopenten-1-one compounds     | 10.49 | 2-hydroxy-5-methyl ester.          |       |                             |       |
| Furfural                          | 5.24  | Penten-2-one compounds Pyrrolidine.| 24.82 | Oxazole                      | 3.64  |
| Propanoic acid. 2-oxo- ethyl ester| 4.13  | 2-(hydroxymethyl)-                 | 0.41  | 3-Penten-2-one. 4-methyl     | 2     |
| 2-Furancarboxaldehyde. 5-methyl-  | 2.37  | Acetaldehyde. ethylidenelydradione  | 3.48  | 5-Hexenonitrile. 2-methyl    | 4.49  |
| Carbamic acid. phenyl ester       | 5.42  | 2-Cyclopenten-1-one compounds      | 2.29  | 2-Pentanone. 4-hydroxy-4-methyl | 13.14 |
| 1,2-Cyclopentanedione. 3-methyl-  | 3.48  | Glycine.                            |       |                             |       |
| Phenol compounds                  | 13.54 | N-(aminoiminomethyl)-N-methyl-     | 1.97  | 2-Furanmethanol              | 5.64  |
| Larixic acid                      | 0.71  | Pentaneone                          | 11.52 | Aziridine compounds         | 4.97  |
| Alkylcarboxaldehyde. 4-methyl-    | 3.83  | 2-(1.1-dimethylethyl)-1,3-dimethyl-| 3.81  | Pyrazine. 2.6-dimethyl-      | 1.20  |
| Alkane (C13–C26)                  | 20.85 | Pyrrole.                            | 1.03  | 2-Cyclopenten-1-one. 3-methyl| 1.16  |
| Alkenes (C14–C26)                 | 5.53  | Phenol compounds                    | 16.07 | 1-Heptene. 2.6-dimethyl-     | 1.59  |
| Hexadecanoic acid. methyl ester   | 2.52  | Pyrazine compounds                  | 1.28  | Phenol compounds             | 15.70 |
| Octadecanoic acid. 2-propenyl ester| 0.44 | Piperazine. 2.5-dimethyl-trans      | 0.37  | Pyrazole. 1.4-dimethyl       | 10.93 |
| Others                            | 13.19 | Caprolactam                         | 1.20  |                             | 2.73  |
|                                   |       | 2-Propan-1-amine. N,N-bis(1-methylethyl) | 3.72  | 2,2,6,6-Tetramethyl-4-piperidone | 4.39 |
|                                   |       | 4-Piperidinone.                     | 11.09 | Alkanes (C13–C26)           | 6.51  |
|                                   |       | 2.2.6.6-tetramethyl-1,3,5,6-piperidone| 1.36  | Alkenes (C13–C18)           | 2.29  |
|                                   |       | Benzofuran.                         |       | Hexadecanoic acid ester      | 0.68  |
|                                   |       | 2.3-dihydro Alkanes (C18–C28)      | 1.49  | Others                       | 11.74 |

In the case of the press mud, it was observed that the pyrolysis liquid from the fourth condenser presented a significant amount of volatiles—acetic acid and toluene (32.9%). The proportion of furfural, benzene, and furan components was also significant (21%), evidencing the decomposition of hemicellulose (Table 7).
Table 7. Composition of the bio-oil collected at the fourth condenser of the pyrolysis unit.

| Press Mud                  | Rel.% | Powder Vinasse                  | Rel.% | Extraction Sludge   | Rel.% |
|----------------------------|-------|---------------------------------|-------|---------------------|-------|
| Acetic acid                | 17.95 | 2,4,4-Trimethyl-2-oxazoline     | 10.52 | 1,3-Butanediol      | 2.92  |
| Tolueno compounds          | 14.97 | Toluene compounds               | 1.27  | Toluene             | 30.86 |
| Furfural                   | 10.96 | Furfuryl alcohol                | 2.65  | 2-Pentanone,        | 5.13  |
|                            |       | Propanoic acid, 2-hydroxy-      |       | 4-hydroxy-4-methyl  |       |
|                            |       | methyl ester                    |       |                     |       |
| Propane, 2,2-dimethoxy-    | 0.46  | 2-Cyclopenten-1-one compounds   | 5.34  | 1-Methyl-1-cyclopent-| 0.44  |
|                            |       | Other Aliphatic ketones         | 0.89  | 3-one               |       |
|                            |       | Other aromatic ketones          | 2.39  |                      |       |
|                            |       | Benzene compounds               | 5.46  |                      |       |
|                            |       | Furan compounds                 | 4.62  |                      |       |
|                            |       | Phenol compounds                | 3.96  |                      |       |
|                            |       | Octadecyl vinyl ether           | 2.26  |                      |       |
|                            |       | p-Methylbenzyl alcohol          | 1.36  |                      |       |
| 3,5-Heptadien-2-ol, 2,6-dimethyl | 1.1  | m-Xylene                        | 0.43  |                      |       |
| Alkanes (C8–C28)           | 14.69 | Xylenol compounds               | 2.87  |                      | 0.57  |
| Alkenes (C7–C22)           | 6.56  | 2-Methyl-2-cyclopentenone       | 3.13  | 2,5-Dimethylpiperazine| 8.56  |
| Other                      | 7.06  | 2-Methylene-4-pentenal          | 1.15  | Alkanes (C9–C27)    | 9.11  |
|                            |       | Pyridine compounds              | 2.57  | Alkenes (C12–C27)   | 4.63  |
|                            |       | Cycloheptanol, 1-methyl-2-methylene- | | Other             | 17.61 |
|                            |       | 4-(Ethylsulfanyl)-2-butanol     | 1.04  |                     |       |
|                            |       | Alkynes (C5–C8)                 | 0.34  |                     |       |
|                            |       | Other                           | 3.71  |                     |       |
|                            |       |                                 | 21.13 |                     |       |

The pyrolysis liquid from the powder vinasse still contained a significant amount of phenolic compounds (18.9%), in particular, phenol, phenol methoxy, phenol methyl, and ethyl. Figure 10 shows the chromatogram of the pyrolysis liquid obtained from powder vinasse with the identification of the peaks characteristic to phenols. This sample contained a higher content of aliphatic and aromatic ketones (19.2%) compared with the other two pyrolysis oils generated from the press mud and the extraction sludge. Moreover, the sample also showed the presence of alkynes and sulfur organic compounds; the powder vinasse presented the highest content of sulfur—1.47%. The extraction sludge showed significant amounts of toluene, xylenes, and benzenes (37.3%), as well as aliphatic and aromatic ketones (6%), alkanes, and alkenes (13.7%).
After individual analysis of pyrolysis liquids samples collected at each condenser, the overall composition of the pyrolysis bio-oil for each biomass waste feedstock was calculated, based on the concentration of each component and the quantity of liquid collected from each condenser. The main components found in each pyrolysis bio-oil are shown in Table 8. As a general observation, the powder vinasse produced a large variety of organic components owing to the thermal decomposition of the lignocellulosic compounds (with high volatiles content).

More than 200 components were identified in the liquid phases collected from each condenser; the separate GC-MS analysis eased the identification of all components. In the case of all three biomass samples, similar compounds were identified, in particular for the powder vinasse and the extraction sludge (high amount of organic components with nitrogen). The press mud produced important amounts of propane alkyloxy (propane, 2,2-diethoxy, 2-propanone, 1,1-dimethoxy, 2-propanone, 1-(acetyloxy)), carboxylic acids, furfural, toluene, and aliphatic and aromatic phenols, as well as alkanes and alkenes. The powder vinasse showed higher produced concentrations of phenolic, aliphatic, and aromatic ketones, as well as organic nitrogen compounds, compared with the bio-oil produced from the press mud. The bio-oil from the powder vinasse showed a high content of aromatics and a low content of hydrocarbons (alkanes, alkenes), which could have a positive impact on the physicochemical properties of the bio-oil.

Conversely, the main component of bio-oil from extraction sludge was toluene (more than 22%) as well as phenolic and organic nitrogen compounds—azirides, piperazines, piridines, and indolizines. The content of hydrocarbons was also high for the bio-oil produced from the extraction sludge.
Table 8. Overall composition of the pyrolysis bio-oil obtained from different biomass feedstock.

| Press Mud               | Mass % | Powder Vinasse           | Mass % | Extraction Sludge         | Mass % |
|-------------------------|--------|--------------------------|--------|---------------------------|--------|
| Carboxylic acid esters  | 3.02   | Carboxylic acid esters   | 1.32   | Carboxylic acid esters    | 2.70   |
| 1-Hydroxy-2-butanone    | 0.20   | 1-Hydroxy-2-butanone     | 0.12   | 5-Hexenenitrile, 2-methyl | 0.21   |
| Propane, propanone,     | 20.61  | 2-Cyclopenten-1-one      | 7.55   | 2-Cyclopenten-1-one       | 1.33   |
| +propanone, alkyloxy-    |        | compounds                |        | compounds                |        |
| 2-Cyclopenten-1-one     | 5.62   | Other Aliphatic ketones  | 6.96   | Other aliphatic ketones   | 0.47   |
| compounds               |        |                          |        |                           |        |
| 2-Pentanone,            | 0.04   | Other aromatic ketones   | 5.82   | Other aromatic ketones    | 0.07   |
| 4-hydroxy-4-methyl-     |        |                          |        |                           |        |
| 1,2-Cyclopentanedione,  | 0.31   | Oxazole compounds        | 13.70  | Oxazoles                  | 0.83   |
| 3-methyl               |        |                          |        |                           |        |
| Other aliphatic ketones | 0.60   | Phenol compounds         | 18.51  | Phenol compounds          | 7.48   |
| Other aromatic ketones  | 1.62   | Benzofuran, 2,3-dihydro- | 0.35   | Ethylenbenzene            | 2.69   |
| Furfural               | 8.16   | Aziridine compounds      | 3.12   | Aziridine compounds       | 4.11   |
| 2-Furan carboxaldehyde  | 0.40   | Furan compounds          | 3.27   | Furan compounds           | 3.59   |
| Phenol compounds        | 5.04   | Pyrazine compounds       | 2.17   | Pyrazine compounds        | 2.64   |
| Benzene compounds       | 4.63   | Furane compounds         | 0.21   | Piperazines               | 6.80   |
| Furan compounds         | 3.14   | Caprolactam              | 0.37   | Piridine compounds        | 0.42   |
| Benzofuran, 2,3-dihydro   | 0.13   | 2-Pentanone, 4-hydroxy-4-methyl- | 0.54   | 2-Pentanone, 4-hydroxy-4-methyl- | 5.18   |
| Benzaldehyde, 4-methyl-  | 0.25   | Amines                   | 0.29   | Indolizine                | 0.44   |
| Acetic acid             | 12.18  | Acetaldehyde, ethylenedihydrazine | 0.78 | Acetaldehyde, ethylenedihydrazine | 0.21   |
| n-Hexadecanoic acid     | 0.03   | Piperidine compounds     | 0.53   | 2,2,6,6-Tetramethyl-4-piperidone | 2.35   |
| Larixic acid            | 0.05   | Toluene compounds        | 1.03   | Toluene                   | 22.69  |
| Oxazole, 4,5-dihydro     | 0.14   | Indole compounds         | 0.02   | Butanenitrile, 2-methyl- | 0.54   |
| Cyclooctane             | 0.05   | Pyrrolidine, 2-(hydroxymethyl)- | 0.21 | Proline                   | 1.26   |
| Toluene compounds       | 10.16  | Furfuryl alcohol         | 2.14   | 2-Picolin                 | 1.49   |
| 3,5-Heptadien-2-ol, 2,6-dimethyl Alkanes (C12-C28) | 0.75 | Picoline compounds      | 1.75   | Xylene compounds          | 2.06   |
| 2-Picolin               | 2.15   |                         |        |                           |        |
| Alkenes (C12-C26)       | 4.94   | m-Xylene                 | 0.35   | Alkanes (C9-C27)          | 8.76   |
| Others                  | 5.78   | Pyridine compounds       | 2.08   | Alkanes (C12-C27)         | 4.26   |
| 2,3-Butanediol,         | 0.07   | 1,3-Butanediol           | 2.15   | Others                    | 15.25  |
| 2-Methylene-4-pentenal  | 0.93   |                           |        |                           |        |
| Cycloheptanol, 1-methyl-2-methylene-4-(Ethylsulfanyl)-2-butanol | 0.84 | Alkenes (C5-C8)          | 3.00   |                       |        |
| Alkynes (C5-C8)         | 3.00   |                           |        |                           |        |
| Alkanes (C19-C32)       | 0.10   |                           |        |                           |        |
| Benzene                 | 0.10   |                           |        |                           |        |
| Others                  | 19.21  |                           |        |                           |        |

4. Conclusions

The vinasse powder presented some of the most favorable physicochemical characteristics to be used as a raw material in a pyrolysis process. Its low ash content leads to a larger high heating value. In addition, the high content of volatile material and lignin favors the production of gaseous and liquid
products during pyrolysis, as this biomass decomposes into additional organic components that lead to high liquid and gas yields.

All three biomass samples have a low concentration of sulfur and nitrogen and relatively high heating values; such characteristics are advantageous for pyrolysis processes. The pyrolysis at 450 °C (heating rate 7–8 °C/min) for 2 h favors the production of char (42.7%–60.1% yield) for all biomass waste feedstock. The press mud showed the highest production of bio-char, but with a low content of carbon, while the powder vinasse produced a bio-char with almost 40% carbon content. The FTIR spectra of all bio-char evidenced the significant decomposition of organic components from all biomass samples; the presence of peaks characteristic to the C–O group points to the capacity of further decomposition of organic compounds at a higher temperature. The results agreed with the TGA–DTG analysis, which showed that none of the biomass samples were completely decomposed at 450 °C.

The pyrolysis of the extraction sludge exhibited the production of the highest amount of bio-oil and the lowest amount of gas, with bio-oil presenting aliphatic compounds (in particular with primary and secondary carbons), aromatics, and carbonyl compounds. The powder vinasse decomposition produced the lowest content of bio-char, along with pyrolysis bio-oil rich in aliphatic compounds, methoxy alcohols, and carbohydrates, as well as aromatic components. This biomass waste also exhibited the highest gas yield (owing to high volatiles content).

The extraction of sludge can be a good feedstock for the production of bio-oil and bio-char. All biomass samples show potential for pyrolysis; for bio-char production (press mud); for valuable chemicals like toluene, carboxylic acids, alkanes, and alkenes (press mud and extraction sludge); and for aliphatic, aromatic phenols, and ketones (powder vinasse).

Author Contributions: Conceptualization, N.M.C., C.E.A.-P., D.D.-V., and A.S.; Methodology, L.E.B.H., N.M.C., D.D.-V., C.E.A.-P., A.S., and E.G.; Software, J.S.D.-O.; Validation, C.E.A.-P., D.D.-V., A.S., and E.G.; Formal Analysis, J.S.D.-O., E.G., B.B., and A.S.; Investigation, J.S.D.-O., C.E.A.-P., D.D.-V., and A.S.; Resources, L.E.B.H., D.D.-V., E.G., and B.B.; Data Curation, J.S.D.-O., C.E.A.-P., and A.S.; Writing—Original Draft Preparation, J.S.D.-O., C.E.A.-P., D.D.-V., L.E.B.H., and A.S.; Writing—Review & Editing, D.E., M.B., E.G., D.L., and A.S.; Visualization, J.S.D.-O. and A.S.; Supervision, C.E.A.-P., D.D.-V., N.M.C., and E.G.; Project Administration, S.G. and M.B.; Funding Acquisition, N.M.C., S.G., M.B., E.G., D.E., and M.S.

Funding: The current work is part of “ERANET-LAC” project under the Horizon 2020 programme. The authors (Devia-Orjuela, Alvarez-Pugliese, Donneys-Victoria, Barba Ho, and Mariaga Cabrales) gratefully acknowledge Colciencia (CT-482-2016 ELAC2015 T03-1018) and Universidad del Valle. The authors (E. Gal and A. Sauciuc) gratefully acknowledge the funding authorities: Romanian National Authority for Scientific Research and Innovation (CCCDI–UEFISCDI, 20/09.2019). Sebastiano Giardinelli and María Briceto gratefully acknowledge SENACYT Panama for financial support, under project ERANET/ELAC2015-T03-1018.

Acknowledgments: The authors (Devia-Orjuela, Alvarez-Pugliese, Donneys-Victoria, Barba Ho, and Mariaga Cabrales) gratefully acknowledge Colciencias, Universidad del Valle, Cenicaña and Leydi Johana Velasco.

Conflicts of Interest: The authors declare no conflict of interest.

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