Association of SARS-CoV-2 clades with clinical, inflammatory and virologic outcomes: An observational study

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\textbf{ABSTRACT}

\textbf{Background:} Host determinants of severe coronavirus disease 2019 include advanced age, comorbidities and male sex. Virologic factors may also be important in determining clinical outcome and transmission rates, but limited patient-level data is available.

\textbf{Methods:} We conducted an observational cohort study at seven public hospitals in Singapore. Clinical and laboratory data were collected and compared between individuals infected with different SARS-CoV-2 clades. Firth’s logistic regression was used to examine the association between SARS-CoV-2 clade and development of hypoxia, and quasi-Poisson regression to compare transmission rates. Plasma samples were tested for immune mediator levels and the kinetics of viral replication in cell culture were compared.

\textbf{Findings:} 319 patients with PCR-confirmed SARS-CoV-2 infection had clinical and virologic data available for analysis. 29 (9%) were infected with clade S, 90 (28%) with clade L/V, 96 (30%) with clade G (containing D614G variant), and 104 (33%) with other clades ‘O’ were assigned to lineage B.6. After adjusting for age and other covariates, infections with clade S (adjusted odds ratio (aOR) 0.030 (95% confidence intervals (CI): 0.0002–0.29)) or clade O (B.6) (aOR 0.26 (95% CI 0.064–0.93)) were associated with lower odds of developing hypoxia compared with clade L/V. Patients infected with clade L/V had more pronounced systemic inflammation with higher concentrations of pro-inflammatory cytokines, chemokines and growth factors. No significant difference in the severity of clade G infections was observed (aOR 0.95 (95% CI: 0.35–2.52)). Though viral loads were significantly higher, there was no evidence of increased transmissibility of clade G, and replicative fitness in cell culture was similar for all clades.

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The severity of coronavirus disease 2019 (COVID-19) ranges from asymptomatic infection to pneumonia, critical illness and death [1–3]. Major determinants of disease severity include host age, pre-existing comorbidities and sex, but virologic factors may also be important [4,5]. The characterisation of a severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) variant with a 382-nucleotide deletion in open reading frame 8 (ORF8) and its association with milder clinical illness is illustrative [6]. However, currently clinical comparison of the transmissibility and virulence of common SARS-CoV-2 variants is lacking. The recent emergence of several variants of concerns (VOC) in South Africa and the UK, and suspected implications for the effectiveness of vaccines and monoclonal antibody therapeutics highlights the importance of this type of analysis [7].

Several classification schemes have been introduced to describe the diversity of SARS-CoV-2 viruses [8]. The global initiative on sharing all influenza data (GISAID) nomenclature describes four major SARS-CoV-2 clades in the early outbreak: clade L, clade V (variant of the ORF3a coding protein NS3-G251), clade G (variant of the spike protein S-D614G with further subclades GR, GH and GV collectively called clade G here), and clade S (variant ORF8-L845S). S and L are the earliest known strains co-occurring in December 2019/January 2020 with Clade L being characteristic for the Wuhan outbreak. G and V clades are descendants of L, though V and L remain genetically closely related. “O” for “Other clades” is assigned for viruses which do not meet these clade definitions, in which case the more detailed lineage information (e.g. B.6) can be used. Similar to other classification schemes this nomenclature describes genetic differences, but does not attempt to reflect antigenic or other phenotypic properties of the virus. A literature search was done through Medline up to October 23, 2020 with the following keywords: Coronavirus disease 2019; COVID-19; SARS-CoV-2; clade; lineage; D614G. Viruses containing the genetic variant D614G (clade G) have become the most common reported worldwide since March 2020, and in vitro and animal models suggest this virus may be more infectious. Country-level data has also correlated the prevalence of clade G with mortality, however, little patient level data was available, and no study compared the outcomes of all four major clades.

### 1. Introduction

The severity of coronavirus disease 2019 (COVID-19) ranges from asymptomatic infection to pneumonia, critical illness and death [1–3]. Major determinants of disease severity include host age, pre-existing comorbidities and sex, but virologic factors may also be important [4,5]. The characterisation of a severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) variant with a 382-nucleotide deletion in open reading frame 8 (ORF8) and its association with milder clinical illness is illustrative [6]. However, currently clinical comparison of the transmissibility and virulence of common SARS-CoV-2 variants is lacking. The recent emergence of several variants of concerns (VOC) in South Africa and the UK, and suspected implications for the effectiveness of vaccines and monoclonal antibody therapeutics highlights the importance of this type of analysis [7].

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The various SARS-CoV-2 nomenclatures are generally in agreement in their categorisation of genetic diversity but importantly, they do not attempt to reflect antigenic or other phenotypic properties of the virus. Viruses containing the genetic variant G614 (G-clade) have become the most common reported worldwide since March 2020, leading to the hypothesis that this variant is more infectious [11]. However, the current dominance of G614 viruses may reflect a founder effect, while country-level data correlating the prevalence of G614 with mortality requires a more detailed study [12–14]. Whether other clades are associated with phenotypic differences is unknown.

Singapore experienced a wide diversity of SARS-CoV-2 clade infections over the first three months of the pandemic, with cases imported from China, Europe, the United States and other major travel routes [15]. We aim to compare clinical outcomes and transmissibility following infection with different SARS-CoV-2 clades using patient-level data generated through comprehensive contact tracing, case finding and isolation measures in Singapore since the beginning of the local outbreak.

### 2. Methods

#### 2.1. Study design and participants

Details of the first 10,000 COVID-19 cases in Singapore (from January 22 to April 22, 2020), by date of notification were extracted from the Ministry of Health database and matched with SARS-CoV-2
sequences shared on GISAID via laboratory accession numbers (see supplementary file for acknowledgment table) [16]. Detailed clinical information was obtained from all patients who were participants of a multi-centre prospective observational cohort study conducted at seven major public hospitals in Singapore (Appendix p1). Over the first three months of the COVID-19 pandemic this study attempted to recruit all individuals admitted with virologically-confirmed SARS-CoV-2 infection. Serial blood samples were collected from consenting participants during hospitalisation and post-discharge. Clinical data was obtained from a review of medical records. In addition, we obtained clinical data from a retrospective study of all patients with COVID-19 who were admitted to the National Centre for Infectious Diseases, the main infectious diseases management centre in Singapore.

2.2. Epidemiological investigation and testing

All COVID-19 cases were interviewed by the Singapore Ministry of Health to determine activity histories from 14 days preceding symptom onset until isolation in hospital, including recent travel history and possible contact with confirmed cases. Contact tracing was also initiated to identify contacts who had significant interactions with each case. Active case finding through testing of symptomatic contacts was performed to detect additional COVID-19 cases amongst these contacts and amongst travellers quarantined on arrival in Singapore. In addition, COVID-19 testing was mandated for persons meeting suspect case definitions, including persons who presented with pneumonia and influenza-like illnesses [17].

2.3. Estimating transmissibility by clade

The number of secondary cases that resulted from each case was derived from the epidemiological findings of all 10,000 cases. It was calculated through repeated probabilistic sampling (1000 repetitions) based on the assumption that in a cluster with 3 or more cases, cases with symptoms onset after an index case could have acquired the infection from the index case or from other secondary case(s). We determine the possible parent infectors for an offspring infectee after factoring for (a) the serial intervals between the infectee and all other possible infectors and (b) the overall serial interval distribution that we have observed amongst cases in Singapore [15]. Only symptomatic cases were considered.

For estimating transmissibility clades for cases that were not genotyped were imputed from those of epidemiologically linked cases that were genotyped. Clusters with more than one clade implicated were excluded. The number of secondary cases amongst cases with assigned viral clade were analysed for associations between viral clade and disease transmissibility. Due to significantly different transmission patterns, cases in the dormitory setting where there are large outbreaks were excluded in the analysis of secondary transmission.

2.4. Clinical management

Over the course of this study, all patients with COVID-19 were isolated with airborne transmission precautions regardless of disease severity. Supplemental oxygen was administered for pulse oximeter saturations ≤ 94%. Patients with moderate to severe hypoxia (defined as requiring fraction of inspired oxygen [FiO2] ≥ 40%) were transferred to the intensive care unit (ICU) for high-flow oxygen via nasal cannula and invasive mechanical ventilation as required. De-isolation was contingent on two consecutive nasopharyngeal swabs > 24 h apart that were negative for SARS-CoV-2 by polymerase chain reaction (PCR).

2.5. Correlation with clinical outcomes

Primary endpoint was the proportion of patients who developed severe COVID-19, defined as hypoxia requiring supplemental oxygen. This endpoint was selected due to the low incidence of ICU admission or death in Singapore. Secondary outcomes were the levels of immune mediators in plasma samples and the SARS-CoV-2 viral load as defined by the PCR cycle threshold (Ct) value from the first available nasopharyngeal swab. Other clinical outcomes are exploratory.

2.6. Viral RNA sequencing and clade determination

SARS-CoV-2 PCR was performed with a range of commercially available assays used at the participating hospitals. Sequencing of SARS-CoV-2 was performed in one of four laboratories in Singapore together with GISAID submission and case matching. Pangolin COVID-19 Lineage Assigner and CoVsurver were used to assign lineage and clade to each sequence, respectively [10,18]. For analysis, L and V clades were merged given their close genetic relatedness, similarly G, GH and GR were merged (Appendix p2; GV had not been described at this point in the pandemic). Sequences labelled as ‘O’ (other clades) were analysed based on assigned Pangolin lineage.

2.7. SARS-CoV-2 culture

Nasopharyngeal swabs were used to inoculate Vero-E6 cells for virus isolation in an Animal Biological Safety Level 3 (ABSL3) laboratory. Cells were cultured at 37 °C until cytopathic effect (CPE) was observed. Positive isolation was confirmed by the observation of CPE and virus-specific PCR. Following virus isolation, virus stocks were prepared in Vero-E6 cells.

For virus growth kinetics, Vero-E6 cells were seeded into 12-well plates at a density of 2 × 10^5 cells per well. Cells were infected with the respective viruses at a multiplicity of infection (MOI) of 0.01. Cell-associated virus was harvested at 24, 48, 72 and 96 h post-infection and titrated by limiting dilution on Vero-E6 cells and are expressed as the 50% tissue culture infectious dose (TCID50) per millilitre.

2.8. Multiplex microbead-based immunoassay

Levels of specific immune mediators in the first plasma samples collected during hospitalisation were quantified by multiplex microbead-based immunoassays. Plasma samples were treated with 1% Triton™ X-100 for virus inactivation [19]. Immune mediator levels were measured with the Lumiplex™ assay using the Cytokine/Chemokine/Growth Factor 45-plex Human ProcartaPlex™ Panel 1 (ThermoFisher Scientific, Waltham, MA) (Appendix p3). Patient samples with concentrations out of measurement range were assigned the value of logarithmic transformation of Limit of Quantification (LOQ). Data analysis was conducted with Bio-Plex Manager™ 6.1.1 software (Bio-Rad, Hercules, CA).

2.9. Data processing and statistical analysis

Kruskal-Wallis test and Conover’s post-hoc pairwise multiple comparison test (for continuous variables) and chi-square test (for categorical variables) were used to compare characteristics and outcomes between groups. Firth’s logistic regression was used to examine the association between the SARS-CoV-2 clade and development of hypoxia due to the presence of separation with zero cell frequency of hypoxia for a particular clade. The following covariates were selected for inclusion in the multivariable logistic regression model: age group, sex, comorbidities, diagnosis through active case finding, month of COVID-19 diagnosis and clade/lineage. An multivariable ordinal regression model was also developed using the same
covariates and outcomes ordered as: 0 = mild (no pneumonia), 1 = pneumonia, no hypoxia, 2 = hypoxia requiring supplemental oxygen, 3 = critical illness and/or death. A quasi-Poisson regression was used to examine the association between SARS-CoV-2 clades and transmission rate, using the number of secondary cases resulting from a case as the dependent variable and the days between disease onset to isolation as an offset to the linear predictor. The latter is used as a surrogate for the period of transmissibility, the true period of which is generally longer due to pre-symptomatic transmission. Subsequent adjustment was performed for this model, for age (represented by three bands: < 45 years (referent), 45–64 years and ≥ 65 years) and sex (male as referent).

One-way analysis of variance (ANOVA) followed by post-hoc t-test with Bonferroni correction was performed on the logarithmically transformed concentrations in order to discern the differences in immune mediator levels amongst patients with different SARS-CoV-2 clade infections. Heatmap and plots were generated using GraphPad Prism version 8 (San Diego, CA). In the heat map presentation, the concentrations of immune mediators were scaled between 0 and 1 for visualisation.

Data processing and analysis were done in the R statistical language (version 3.6.1) and Stata version 15 (StataCorp, College Station, TX). All statistical tests were two-sided, and p-values < 0.05 considered statistically significant. Adjustment for multiple testing was not performed.

2.10. Ethics statement

The epidemiological investigation was conducted under the Infectious Diseases Act (Singapore). Study protocols were approved by ethics committees of the National Healthcare Group and SingHealth. Written informed consent was obtained from participants for clinical data and biological sample collection as part of the PROTECT study (2012/00917; 2018/3045). A waiver of informed consent for retrospective data collection only was granted for individuals admitted to the National Centre of Infectious Diseases (2020/01122). Healthy donor samples were collected under study numbers 2017/2806 and NUS IRB 04-140.

2.11. Role of the funding source

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3. Results

3.1. Descriptive epidemiology

The first confirmed COVID-19 case in Singapore was an imported infection notified on January 22, 2020, and the first locally transmitted cases were reported on February 3, 2020 (Fig. 1). In response to the outbreak travel restrictions were progressively introduced from February 1, 2020 and a heightened disease alert level was announced on February 7, 2020. A surge in imported cases occurred in March 2020 when residents returned from overseas, with entry-restrictions and 14-day quarantine for arrivals imposed on affected countries. The first foreign worker dormitory cluster was announced at the end of March 2020. This spread to involve numerous foreign worker dormitories, resulting in a steep increase in cases that peaked at more than 1000 cases per day by late April 2020. The increase in cases triggered social distancing restrictions on March 26, 2020, with broad restrictions imposed from April 7, 2020 similar to the lockdowns in other countries.

Of the first 10,000 COVID-19 cases reported in Singapore, 8132 (81.3%) were male foreign workers residing in dormitories. Overall, the proportion of elderly cases (age 65 and above) was 2.4% (240/10,000); decreasing from 11.0% (98/865) in January to March, to 1.5% (134/9033) in April 2020. 0.2% (23/10,000) died with COVID-19 listed as a direct cause. 540 of the first 10,000 cases had SARS-CoV-2
genotyping information available through whole genome sequencing. The four major clades, S, L, V and G were all present in Singapore in January 2020 with repeated importation and local clusters of infection until April 2020. The large outbreak amongst foreign workers was almost exclusively due to O (B.6) [20].

3.2. Cohort study

320/540 (59%) patients with genotyped virus had detailed clinical data available for analysis (Appendix p1). 29 (9%) were clade S, of which 18 (62%) were known to be infected with the attenuated A382 variant (Table 1). 67 (21%) were infected with clade L, 23 (7%) with clade V, and 96 (30%) with clade G (of which 33 were GH and 41 GR). 104 (33%) infections with O label (other clades) were assigned to lineage B.6, while one infection with lineage B.4 was excluded from further analysis.

Overall, the cohort for analysis of 319 individuals included 47 (15%) with severe infection (hypoxia with requirement for supplemental oxygen) and 23 (7%) with critical illness (Table 1). 89% of the cohort was aged less than 65 years, and 82% had a Charlson’s comorbidity index of 0. 47% of the cohort was diagnosed through active case finding, a proportion that did not significantly change from January to April (January/February, 26/49 (53%); March 82/189 (43%); April 41/82 (50%), p = 0.95, Chi square for trend). There was no significant change in the crude proportion of individuals with severe infection from January to April (January/February 11/49 (22%); March 25/189 (13%); April 11/82 (13%), p = 0.25, Chi square for trend).

Significant baseline differences between the groups infected with different clades were observed – including in age, ethnicity and comorbidities. After adjusting for co-variates in the multivariable logistic regression model, compared with L/V there was no significant difference in the severity of G-clade infection, while infections with clade S (adjusted odds ratio (aOR) 0.030 (95% confidence intervals (CI): 0.0002–0.29)) or clade O (B.6) (aOR 0.26 (95% CI 0.064–0.93)) were associated with lower odds of developing hypoxia requiring supplemental oxygen compared with infection (Table 2). There were 30 dormitory residents amongst the 104 individuals infected with O (B.6) in the cohort, none of whom had severe infection (dormitory: 0/30 (0%), non-dormitory: 12/74 (16%)). After excluding dormitories residents from the analysis, infections with O (B.6) remained significantly less severe compared with L/V (aOR 0.29 (95% CI 0.071–1.00), p = 0.049, Appendix p4).

| Table 1. Characteristics of 319 genotyped COVID-19 cases. |
|-----------------------------------------------------------|
| Clade | All | S | L/V | G | O (B.6) | p-value |
|-------|-----|---|-----|---|----------|---------|
| N*    | 319 | 29| 90  | 96| 104      | <0.0001 |
| Month of diagnosis                                   |
| January/February/                                  |
| 49 (15%) 23 (80%) 24 (27%) 2 (2%) 0 (0%) 0.0001 |
| March     |
| 188 (59%) 5 (17%) 65 (72%) 89 (93%) 29 (28%) 0.0005 |
| April     |
| 82 (26%) 1 (3%) 1 (1%) 5 (5%) 75 (72%) 0.0005 |
| Symptom onset to admission (days)                    |
| 4 (2–7) 4 (2–5) 3 (2–7) 3 (2–5) 4 (3–8) 0.031 |
| Diagnosis through active case finding†               |
| 149 (47%) 15 (52%) 49 (54%) 34 (35%) 51 (49%) 0.055 |

Demographics

| Age (years)               |
|---------------------------|
| 42 (30–55.5)             |
| 39 (31–53)               |
| 48 (32–62)               |
| 32 (25–46)               |
| 43 (34–57)               |
| < 0.0001                 |
| 185 (58%)                |
| 18 (62%)                 |
| 190 (59%)                |
| 39 (34%)                 |
| 69 (72%)                 |
| 59 (57%)                 |
| 0.0005                   |
| 45–64                    |
| 34 (11%)                 |
| 37 (41%)                 |
| 25 (26%)                 |
| 28 (27%)                 |
| 0.0001                   |
| ≥ 65                     |
| 14 (6%)                  |
| 12 (6%)                  |
| 20 (10%)                 |
| 16 (8%)                  |
| 8 (4%)                   |
| 0.0001                   |
| Male sex                 |
| Chinese                  |
| 19 (6%)                  |
| 12 (6%)                  |
| 5 (3%)                   |
| 8 (4%)                   |
| < 0.001                  |
| Malay                    |
| 19 (6%)                  |
| 12 (6%)                  |
| 6 (3%)                   |
| 8 (4%)                   |
| 0.0001                   |
| South Asian†             |
| 19 (6%)                  |
| 12 (6%)                  |
| 6 (3%)                   |
| 8 (4%)                   |
| 0.0001                   |
| Other                    |
| 19 (6%)                  |
| 12 (6%)                  |
| 6 (3%)                   |
| 8 (4%)                   |
| 0.0001                   |
| Charlson score (median, IQR) |
| 0 (0–0)                  |
| 0 (0–0)                  |
| 0 (0–0)                  |
| 0 (0–0)                  |
| 0.060                    |
| > ≥ 1                    |
| Diabetes                 |
| 19 (6%)                  |
| 11 (6%)                  |
| 7 (4%)                   |
| 9 (5%)                   |
| 0.009                    |
| Hypertension             |
| 22 (7%)                  |
| 11 (6%)                  |
| 7 (4%)                   |
| 9 (5%)                   |
| 0.015                    |
| Co-morbid conditions†    |
| 19 (6%)                  |
| 11 (6%)                  |
| 7 (4%)                   |
| 9 (5%)                   |
| 0.003                    |
| Laboratory values        |
| LDH                       |
| 369 (321–418)            |
| 405 (339–532)            |
| 370 (312–428)            |
| 383 (332–457)            |
| 0.032                    |
| CRP                       |
| 5 (3.7–12.7)             |
| 7 (6.2–12.7)             |
| 5 (2.5–12.5)             |
| 4 (2.7–12.6)             |
| 0.038                    |
| Lymphocyte count          |
| 1.3 (0.9–1.8)            |
| 1.1 (0.9–1.7)            |
| 1.0 (0.9–1.6)            |
| 1.1 (0.9–1.4)            |
| <0.0001                  |
| Neutrophil count          |
| 3 (1.5–3.5)              |
| 3 (1.5–3.5)              |
| 2 (1.0–3.2)              |
| 3 (1.2–3.6)              |
| 0.052                    |
| Cq Value                  |
| 25.7 (22.0–30.4)         |
| 21.9 (19.1–26.4)         |
| 25.8 (23.7–30.7)         | <0.0001 |

Outcomes

| Pneumonia                  |
| 122 (38%)                 |
| 10 (34%)                  |
| 45 (50%)                  |
| 33 (34%)                  |
| 34 (33%)                  |
| 0.060                    |
| Supplemental oxygen        |
| 47 (15%)                  |
| 10 (0%)                   |
| 24 (27%)                  |
| 11 (13%)                  |
| 12 (12%)                  |
| 0.0007                   |
| ICU/Death                 |
| 23 (7%)                   |
| 10 (0%)                   |
| 14 (16%)                  |
| 0 (0%)                    |
| 9 (9%)                    |
| 0.0002                   |

Data is presented as median (inter-quartile range) or n (%) unless indicated.
* One case (lineage B.4) excluded.
† 23 Jan to 29 Feb 2020.
‡ Including contact tracing, quarantine orders and surveillance.
§ India, Bangladesh, Sri Lanka.
∥ Categorical variable defined as Charlson Comorbidity Index ≥1 or hypertension. See Appendix p4 for breakdown of comorbid conditions by clade.
Table 2
Odds ratios of candidate predictors for (a) hypoxia requiring supplemental oxygen from Firth’s logistic regression analysis, and (b) clinical outcome ordered from mild infection to critical illness and/or death from multivariable ordinal logistic regression;

| Age Group (Years) | Univariable LR model | Multivariable LR model | Proportional odds model |
|-------------------|-----------------------|-------------------------|-------------------------|
|                   | OR (95% CI) | p-value | aOR (95% CI) | p-value | aOR (95% CI) | p-value |
| <45               | 1.00 [reference] | - | 1.00 [reference] | - | 1.00 [reference] | - |
| 45–64             | 6.25 (2.83–15.1) | <0.0001 | 4.55 (1.93–11.57) | 0.0004 | 4.25 (2.44–7.50) | <0.0001 |
| ≥65               | 18.6 (7.36–50.6) | <0.0001 | 7.80 (2.45–26.6) | 0.0005 | 9.82 (3.95–25.00) | <0.0001 |
| Female sex        | 0.84 (0.43–1.57) | 0.58 | 0.70 (0.33–1.47) | 0.35 | 0.85 (0.50–1.44) | 0.55 |
| Co-morbid conditions1 | 7.06 (3.69–13.8) | <0.0001 | 4.57 (1.94–11.1) | 0.00053 | 3.48 (1.84–6.59) | 0.00013 |
| Clade             |                      |               |                      |               |                      |               |
|                   | S                    | 0.046 (0.004–0.35) | 0.00048 | 0.030 (0.0002–0.29) | 0.00059 | 1.00 (reference) | - |
|                   | G                    | 0.37 (0.16–0.77) | 0.0082 | 0.95 (0.35–2.52) | 0.91 | 0.30 (0.10–0.81) | 0.021 |
|                   | O (8.6)              | 0.37 (0.17–0.76) | 0.0072 | 0.26 (0.064–0.93) | 0.037 | 0.30 (0.13–0.92) | 0.037 |
| Diagnosis through active case finding |                      |               |                      |               |                      |               |
|                   | 0.68 (0.36–1.26) | 0.22 | 0.89 (0.42–1.90) | 0.77 | 0.83 (0.49–1.40) | 0.48 |
| Month of diagnosis |                      |               |                      |               |                      |               |
|                   | January/February     | 1.00 [reference] | - | 1.00 [reference] | - | 1.00 [reference] | - |
|                   | March                 | 0.66 (0.29–1.63) | 0.36 | 0.21 (0.061–0.70) | 0.012 | 0.21 (0.088–0.52) | 0.00070 |
|                   | April                 | 0.63 (0.24–1.73) | 0.36 | 0.42 (0.07–2.43) | 0.32 | 0.34 (0.10–1.40) | 0.086 |

3.3. Comparison of early immune response

Acute plasma samples at the first collection timepoint upon hospital admission were available from 99 patients (Fig. 2a). No significant difference in the sample collection time point was observed between the different clades (Appendix S6). Collectively, higher concentrations of chemokines MCP-1, RANTES, SDF-1α, and IP-10, pro-inflammatory IFN-γ, IL-6 and IL-1β and the anti-inflammatory IL-1RA were detected in patients infected with clades L and V compared with patients infected with either clade S or G (Fig. 2b) at a median of 5 days post-infection onset (interquartile range 3–8). In addition, these patients had higher levels of growth factors associated with lung injury and regeneration, including SCF, VEGF-A, PIGF-1 and PDGF-BB. Notably, patients infected with clade G SARS-CoV-2 had lower concentrations of growth factors including VEGF-A, HGF, PDGF-BB and PIGF-1, pro-inflammatory IL-1β, IFN-γ and chemokines such as RANTES, SDF-1α and IP-10, at the early acute phase of infection (Fig. 2B).

Fig. 2. Plasma immune mediator levels of COVID-19 patients infected with different SARS-CoV-2 clades. Concentrations of 45 immune mediators were quantified using a 45-plex microbead-based immunoassay. (a) Heatmap of immune mediator levels in plasma samples of patients infected with different SARS-CoV-2 clades (clade S, n = 21; clade O (B.6), n = 10; clade G, n = 22; clades L and V, n = 46) during first collection timepoint upon hospital admission (median 5 days from symptom onset). Each colour represents the relative concentration of a particular analyte. Blue and red indicates low and high concentration, respectively. (b) Profiles of significant immune mediators of COVID-19 patients infected with different SARS-CoV-2 clades at the first collection timepoint upon hospital admission are illustrated as scatter plots. Immune mediator levels in plasma fraction samples from first collection timepoint during hospital admission (median 5 days from symptom onset) were compared amongst the patients infected with different SARS-CoV-2 clades. One-way ANOVA followed by post-hoc t-test with Bonferroni correction was performed on the logarithmically transformed concentration (*p < 0.05; **p < 0.01; ***p < 0.0001). Immune mediator levels for healthy controls (n = 23) are indicated by the black dotted line. Patient samples with concentration out of measurement range are presented as the value of logarithm transformation of Limit of Quantification (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.).
3.4. Transmissibility

In addition to the 540 cases with SARS-CoV-2 genotyping information available, a further 5626 cases had virus genotype inferred by epidemiologically linkages. The majority (5460/6166, 88.6%) were dormitory residents and were excluded from the analysis. Amongst 706 cases included in the analysis (Appendix p7–8), 546 (76.7%) were non-dormitory cases with genotyped and inferred clades. The majority (5460/6166, 88.6%) were dormitory residents and were excluded from the analysis. Amongst those who were exposed to the community after symptom onset. In addition to the 540 cases with SARS-CoV-2 genotyping information available, a further 5626 cases had virus genotype inferred by epidemiologically linkages. The majority (5460/6166, 88.6%) were dormitory residents and were excluded from the analysis. Amongst those who were exposed to the community after symptom onset.

There was no significant difference between the viral loads of Clade L/V, S and O (B.6), whereas Clade G was associated with significantly higher viral loads compared with the other three clades (Conover post hoc pairwise comparison, p < 0.05). Day of sample collection was not significantly different (Kruskal–Wallis, p = 0.64). The kinetics of viral replication in cell culture were similar between the different clades as measured up to 96 h after inoculation (Fig. 3).

3.5. Viral culture

There was no significant difference between the viral loads of Clade L/V, S and O (B.6), whereas Clade G was associated with significantly higher viral loads compared with the other three clades (Conover post hoc pairwise comparison, p < 0.05). Day of sample collection was not significantly different (Kruskal–Wallis, p = 0.64). The kinetics of viral replication in cell culture were similar between the different clades as measured up to 96 h after inoculation (Fig. 3).

Table 3.
Relative risk of onward transmission (i.e. any secondary transmission) by clade amongst non-dormitory cases with genotyped and inferred clades (n = 587).

| Clade | Crude RR (95%CI) | p-value | Adjusted RR (95%CI) | p-value |
|-------|----------------|---------|---------------------|---------|
| L/V   | 1.00 (reference) | -       | 1.00 (reference)    | -       |
| S     | 1.28 (0.94–1.73) | 0.12    | 1.28 (0.94–1.75)   | 0.12    |
| G     | 0.35 (0.23–0.56) | < 0.001 | 0.36 (0.23–0.58)   | < 0.001 |
| O (B.6) | 0.78 (0.61–1) | 0.048   | 0.80 (0.62–1.02)   | 0.07    |

Note: Modified Poisson regression with binary outcome of transmission occurrence as regressand, amongst those who were exposed to the community after symptom onset. CI: confidence interval; RR: relative risk.

4. Discussion

In this large cohort study we found no evidence for attenuated or increased severity of clade G when comparing infections with the L/V clades. However, clade G was associated with significantly higher viral loads (as measured by PCR Ct value). Clade S was associated with less severe infection, likely reflecting infection by the ORF7/8 deletion variant that we previously reported to be associated with attenuated severity [6]. O (B.6) was associated with attenuated severity of infection for reasons that are unclear and merits further investigation.

G-clade viruses emerged in late January 2020 and became the most common clade worldwide (based on samples submitted to GISAID) in March 2020 [21]. To date there is little patient level data comparing outcomes from G-clade viruses with other variants. A study of 999 patients in Sheffield, UK, reported that infection with G614 was not associated with an increased risk of hospital admission, however, it was associated with higher viral load [22]. A study of 113 infections in Hong Kong reported no significant increase in the severity of infection [23]. We found that clade G is not associated with different disease severity compared with L/V.

The significantly lower PCR Ct value with G-clade infection observed in our cohort supports other data that this virus may be more infectious [24]. In vitro experiments indicate that the G614 variant opens the structure of the spike protein, facilitating binding to the ACE-2 receptor and that this virus is more infectious in tissue culture [21,25]. While previous studies have used a pseudovirus, we observed no difference in viral kinetics between G and other clades. In a hamster model, the SARS-CoV-2 virus with the G614 virus
produced higher infectious titres in nasal washes and trachea, but not lungs, than the D614 virus [26].

At the same time, we observed significantly lower transmission from individuals infected with G-clade viruses compared with other clades. This is likely to be due to significantly earlier isolation of this group, and may reflect either improvements in community diagnosis or behavioural differences in then affected population. For all clades the reproduction number, R, was significantly below 1, and overall 0.34 reflecting the success of Singapore’s COVID-19 public health measures at reducing transmission in the community and indicating that these interventions are effective regardless of clade [15]. While an underestimate because only symptomatic infections are considered, it is still low compared to other R values estimated from symptomatic infections [27]. Higher SARS-CoV-2 viral load has been correlated with COVID-19 mortality, suggesting host immune response to G-clade infections may differ and be less pathogenic – either as a direct result of D614G or other genetic variation associated with this clade [28,29].

We found patients infected with clade L/V had more pronounced systemic inflammation with higher concentrations of pro-inflammatory cytokines, chemokines and growth factors that are strongly associated with severe COVID-19 during the early acute phase of infection [30,31]. Excessive production of pro-inflammatory cytokines in SARS-CoV-2 infection damages endothelial cells resulting in capillary leakage and vascular injury [32]. The more robust production of pro-inflammatory IL-1β, IL-6, MCP-1, RANTES and MIP-1β at the early phase of infection observed in patients infected with clade L/V could promote lung damage, which may in turn result in severe clinical outcomes, as shown by the higher proportion of patients in this group requiring oxygen. Intriguingly, those with clade G infections had a less overactive inflammatory response [22]. In addition, patients with clade S infections had lower levels of inflammation compared with those infected with L/V, and this may reflect the known association between the ORF8 deletion variant and inflammatory responses [6].

This observational study has a number of limitations. We only included data up to April 2020, a period which saw the greatest diversity of SARS-CoV-2 clades in Singapore. Viral adaptation to infection in humans may have occurred since then which affects COVID-19 severity. We also merged several closely genetically related clades (L/V and G/GH/GR) to improve study power which prevents us from analysing the effects of these individual clades. SARS-CoV-2 testing during the study period was largely symptom driven and it is possible that asymptomatic or pauci-symptomatic infection rates vary with different clades. As the pandemic progressed beyond March 2020 the cohort study was unable to enrol all individuals with COVID-19 due to the large numbers. Recruitment was biased towards more severe infections, suggesting that for G and O (B.6) clades in particular severity is likely to have been overestimated. Clinical management evolved over the first few months of the pandemic which may have improved outcomes. However, it remained largely supportive: remdesivir was yet to be licensed, corticosteroids were avoided, while convalescent plasma and monoclonal antibodies were not available [33,34]. SARS-CoV-2 PCR was performed on different commercial platforms, with varied gene targets, limiting the comparability of Ct values. Also, clade distribution in the population is associated with certain subpopulations and clusters. As such, associations of severity with clades may reflect underlying human behaviour or characteristics in the subpopulations, rather than the true virulence or infectiousness of the virus. These differences are difficult to measure and challenging to control for in an observational study.

SARS-CoV-2 genotypes imputed through epidemiological linkage data risks miscategorization. However, the community disease incidence in Singapore was low (e.g. 0.7 per 100,000 persons per day in April 2020) in a setting of aggressive contact tracing. As a result it was unlikely that infection within a cluster arose from more than one source of infection providing high confidence in the relationships between cases. Furthermore, clusters with more than one genotyped clade were excluded. While there may be uncertainties in determining the exact direction of transmission within a cluster, a probabilistic approach to transmission was undertaken. A limitation is that only symptomatic cases were considered in assessing transmissibility, as this allows greater confidence in the direction of transmission. The results may not necessarily reflect the transmissibility of asymptomatic disease unless the clade distribution of asymptomatic cases is similar to that of symptomatic disease.

In conclusion, as a newly emerged human pathogen, SARS-CoV-2 is likely to be under strong selection pressure to adapt to infection in humans. The recent emergence of new variants which may also be able to evade host immunity is concerning, though claims of increased transmissibility or changes in virulence require careful evaluation [35]. In our large cohort study, with detailed epidemiological, clinical and immunological data we find patient-level evidence that infections by G-clade viruses are associated with higher viral titres, without necessarily affecting disease severity or increasing transmissibility. The association of O (B.6) infections with attenuated severity and stronger immune response as compared to early L/V vs later G-clades is intriguing but requires further study.

Declaration of Competing Interest

BY has received honoraria from Sanofi and Roche, outside the submitted work. All other authors no conflicts of interest declared.

Contributors

BY, WW, SWF, TMM, DA, RL, YSL, LR, LW, LN, SMS, DL, VL conceived and designed the study. BY, WW, TMM, DA, SK, SP, LC, PP, SYT, LS, DL collected clinical samples and data. BY, WW, SWF, TMM, DA, YHC, RP, CH, LWA, AK, BL performed the experiments and analyzed the data. BY, WW, SWF, TMM, DA wrote the first draft of the manuscript. All authors contributed to data interpretation, critically reviewed the manuscript, and approved the final manuscript for submission.

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Data sharing

Qualified researchers may request access to anonymized individual patient level data by sending a request to the corresponding author. Data will be shared after approval of a proposal, with a signed data access agreement.

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.ebiom.2021.103319.
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