**Supplementary Figure Legends**

**Figure S1.** *In Vitro* translation analysis of wild-type (WT) and c.354G>A variant *ADAMTS13* transcripts. Representative autoradiogram of WT and p.P118P (c.354G>A) variant *in vitro* translation products. *ADAMTS13* protein is indicated by an arrow. Significantly higher translation of c.354G>A variant relative to WT transcript was observed. *In vitro* translation products of *ADAMTS13* variants not included in current study are indicated.

**Figure S2.** Cumulative sum of (a) log (Ribosome Protected Fragment) *ADAMTS13* values. Cumulative sum of the normalized log (Ribosome Protected Fragments) of WT and c.354G>A (p.P118P) transcripts is shown. Data series for each construct represents two biological replicates of three technical replicates each.

**Figure S3.** Average normalized reads by codon number for *ADAMTS13* (WT and c.354G>A (p.P118P)), *ACTB* and *GAPDH*. Panels A and B respectively show ribosome profiles for *ADAMTS13* variants along the entire gene and focused around codon 118. Largely similar profiles were observed in both the plots. Panels C and D show ribosome profiles for control genes *ACTB* and *GAPDH* respectively. Data series for each construct represents two biological replicates of three technical replicates each.

**Figure S4.** Circular dichroism (CD) analysis of wild-type (WT) and p.P118P variants. CD spectra of WT and p.P118P variant proteins from three independent runs were displayed.
**Supplementary Figure Legends (continued)**

**Figure S5.** Glycosylation of W-387 in ADAMTS13. The MS/MS spectrum of m/z 519.2+ showing the b and y'' ions used to assign the sequence within wild-type ADAMTS13 as WSSWGPR. The quasimolecular ion is 162 Da, a Hexose unit, higher in mass than the theoretical peptide mass, and the position of substitution of the Hexose is on the first Tryptophan, W387, as proven by the interpretation of the fragment ions observed, shown in schematic with suggested mechanisms. There is no evidence of W-390 being Hexosylated. The principal fragment ions observed correspond to the novel formation of a 2-Ethynyl-Indole (Acetylenic substituent) on the side chain of W-387 resulting from loss of 138 Da due to water loss and partial cleavage of the Hexose ring, in preference to the normal β-elimination (162 Da) seen in O-linked glycosylation chemistry.

**Figure S6.** Glycosylation of S-1170 in ADAMTS13. The partial Mass Spectrum (m/z 550-1200) at the corresponding LC-MS elution time where the MS/MS of a signal at 1065.52+ gave b and y'' signals attributable to the sequence GLLFSPAPQPR in ADAMTS13. The spectrum shows an ion at 1065.52+ and a corresponding quasimolecular ion for the peptide component of 1182.6 (591.82+) which calculates for a NeuAc₂HexHexNAc unit attached to S1170 of the peptide sequence assigned. From a knowledge of mammalian biosynthetic pathways this likely corresponds to a DiSialyl Core-1 structure, and the sequential losses of individual sugar units observed (and annotated in the Figure) suggests the basic structure illustrated.

**Figure S7.** Rosetta Interface Score vs RMSD for the modelled metalloprotease docked on to the DTCS (Crystal structure of Disintegrin (D), TSR Repeat-1 (T), Cysteine-rich (C) and Spacer (S) domains of ADAMTS13) run. Each point represents a structure created by the ROSIE server with the top scoring structures shown in bold. The vertical axis gives the interface score (total score of the protein complex minus the total score of each partner in isolation, as described in the ROSIE documentation), while the horizontal axis gives the root-mean-square deviation (RMSD), a measure of distance from the starting position.
Figure S1

ADAMTS13 variants not included in the current study.
Figure S3

A: ADAMTS13

B: ADAMTS13 Around P118P

C: ACTB

D: GAPDH
Figure S4
Figure S6

GLLFSPAPQPR $[\text{M+H}]^+$

GLLFSPAPQPR $[\text{M+2H}]^{2+}$

HexNAc
Hex
NeuAc
Hex
NeuAc
NeuAc

GLYCOPEPTIDE $[\text{M+2H}]^{2+}$

GLLFSPAPQPR $[\text{M+H}]^+$
Figure S7