Value of blood inflammatory markers for predicting intravenous immunoglobulin resistance in Kawasaki disease: A systematic review and meta-analysis

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Background: Previous studies have assessed the diagnostic accuracy of blood inflammatory markers like neutrophil-to-lymphocyte ratio (NLR), platelet-to-lymphocyte ratio (PLR), and C-reactive protein (CRP), and CRP to albumin ratio (CAR) to predict the resistant Kawasaki disease (KD). The aim of the current meta-analysis and systematic review is to compare the prognostic ability of these inflammatory markers to predict the resistance to IVIG in patients with Kawasaki disease.

Methods: A systematic search of online academic databases and search engines such as EMBASE, PubMed Central, MEDLINE, Cochrane library, Google Scholar, and ScienceDirect was conducted for papers that report the diagnostic accuracy of inflammatory markers for resistant KD. The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Results: Twenty-two studies met the inclusion criteria. Pooled sensitivity and specificity of NLR as a predictor of resistant Kawasaki disease was 72% (95% CI: 62%, 80%) and 71% (95% CI: 63%, 78%), with AUC of 0.77 for PLR was 60% (95% CI: 50%, 69%) and 68% (95% CI: 61%, 75%), with area under the curve (AUC) of 0.69. Pooled sensitivity and specificity of CRP was 75% (95% CI: 68%, 81%) and 66% (95% CI: 55%, 76%), respectively, with an AUC value of 0.78. Pooled sensitivity and specificity of combined NLR and PLR was 58% (95% CI: 46%, 69%) and 73% (95% CI: 65%, 79%), respectively, with an AUC value of 0.72.
Conclusion: Our study found that NLR, CRP, PLR, and combined NLR/PLR have a good prognostic value in patients with resistant Kawasaki disease, with moderate to high sensitivity and specificity. More research on the accuracy of these indexes in multiple combinations is needed.

Systematic review registration: [https://www.crd.york.ac.uk/prospero/], identifier [CRD42022322165].

KEYWORDS
C-reactive protein, intravenous immunoglobulin resistance, Kawasaki disease, meta-analysis, neutrophil-lymphocyte ratio

Introduction

Kawasaki disease (KD) is an acute vasculitis that affects coronary arteries in children (1) and is considered the most common cause of acquired cardiac complications in children worldwide.

Intravenous immunoglobulin (IVIG) is the main method of KD treatment that has been known to reduce the risk of coronary artery lesions to 4% from 25% if initiated on time (2). However, 10–20 percent of the patients with Kawasaki disease are IVIG-resistant and have a recurrent or persistent fever even after the initial treatment with IVIG (2). Studies show that IVIG-resistant patients have higher rates of coronary artery lesions development compared to patients that respond to IVIG therapy (3, 4). Combining IVIG with other anti-inflammatory therapeutic agents as the first treatment might lower the incidence of resistance to IVIG and the number of coronary artery lesions (5–8). It is necessary, therefore, to develop ways of early identification of the patients at higher risk of resistant KD, to allow them to benefit from the rapid and aggressive line of management.

Recently, several studies focused on the ability of various blood inflammatory markers like platelet-to-lymphocyte ratio (PLR), neutrophil-to-lymphocyte ratio (NLR), and C-reactive protein (CRP), and CRP to albumin ratio (CAR) to predict the resistant Kawasaki disease (9–15). There is a need for determination of markers for outcome, including IVIG resistance and coronary artery involvement. There is comprehensive review on possible biomarkers for KD and studies comparing the available ten IVIG resistance-predicting scoring system (16, 17). However, these studies show an unsatisfying sensitivity for predicting IVIG resistance. In addition, there are still no reviews that attempted to summarize this evidence systematically. Hence, the main aim of the current systematic review and meta-analysis is to compare the prognostic values of NLR, PLR, CRP, and CAR for predicting the resistance to IVIG in patients with Kawasaki disease.

Methods

Eligibility criteria

Study design

Prospective and retrospective observational studies (cohort, case-control, or cross-sectional analytical) were included. Case reports, series, conference abstracts, and unpublished gray literature were excluded. The study was registered on PROSPERO (No. CRD42022322165).

Study participants

Studies of the Kawasaki disease patients were incorporated irrespective of age and gender of the participants, and comorbidities.

Index test and reference standards

Studies assessing the accuracy of NLR, PLR, CRP, and PAR in predicting the resistant form of Kawasaki disease were included if they satisfied all other inclusion criteria.

Search strategy

An extensive systematic search was done in the databases and search engines such as PubMed Central, EMBASE, MEDLINE, Cochrane library, Google Scholar, and ScienceDirect. The following combination of medical subject headings (MeSH) and free-text headings with the suitable Boolean operators (“AND” and “OR” and “NOT”) in between the pre-defined search terms was used: (“Mucocutaneous Lymph Node Syndrome”[MeSH Terms] OR “Resistant Kawasaki Disease”) OR AND (“immunoglobulins, intravenous”[MeSH Terms] AND “Neutrophil Lymphocyte Ratio”) OR (“Platelet Lymphocyte Ratio”) OR (“CRP Albumin Ratio”) OR (“C-Reactive Protein”) OR (“Neutrophils”[MeSH Terms]) OR “Lymphocytes”[MeSH Terms] OR “Blood Platelets”[MeSH Terms] OR “C-Reactive Protein”[MeSH Terms].
Terms] OR "Albumins"[MeSH Terms]. The following additional filters were also applied during the process of literature search: time point (January 1954–March 2022), and language (English and Chinese). References from the retrieved studies were also searched to identify any additional relevant articles.

**Steps in study selection**

The first step of the study selection process included a screening of the title, keywords, and abstract and was carried out by the two independent reviewers (CL and JW). Full-text studies were then shortlisted independently by two investigators for the second stage of screening based on the eligibility criteria, and studies that met the criteria were selected for further analysis. Disagreements were resolved by discussion. “Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement 2020” was used for reporting this review (18).

**Data extraction procedure**

Data was extracted manually by the two investigators (CL and JW) using a pre-defined semi-structured data collection form that was defined at the stage of the protocol itself. The following information was obtained: name of the authors, study title, publication year, year when the study was done, duration of the study, design, study setting, country/region of the study, sample size, outcome assessment tool, and other details, mean
### TABLE 1 Characteristics of the included studies (N = 22).

| Study no | First author and year | Country | Study design | Sample size | Diagnostic criteria for Kawasaki disease | Indicators for predicting resistant Kawasaki disease | Indicator acquisition time | Cut-off point for indicators | Risk of bias |
|----------|------------------------|---------|--------------|-------------|-------------------------------------------|--------------------------------------------------|----------------------------|-------------------------------|-------------|
| 1        | Chen et al. (25)        | China   | Retrospective | 92          | AHA | NLR | Before IVIG administration | NLR = 2.51 | Higher                      |
| 2        | Cho et al. (23)         | Korea   | Retrospective | 196         | AHA | NLR | Before IVIG administration | NLR = 5 | Higher                      |
| 3        | Faim et al. (38)        | Portugal| Retrospective | 48          | AAP | CRP | Before IVIG administration | CRP = 15.1 mg/dL | Lower                     |
| 4        | Ha et al. (13)          | Korea   | Retrospective | 587         | AHA | NLR | Before and after IVIG administration | NLR = 1.26 (after) NLR = 5.49 (before) | Higher       |
| 5        | Iwashima et al. (24)    | Japan   | Retrospective | 91          | JKDRC | CRP | Before IVIG administration | CRP = 8 mg/dL | Higher                     |
| 6        | Kanai et al. (35)       | Japan   | Retrospective | 520         | JKDRC | NLR and PLR | Before IVIG administration | NLR = 4.11 PLR = 119 | Lower                     |
| 7        | Kawamura et al. (14)    | Japan   | Retrospective | 405         | JKDRC | NLR and PLR | Before and after IVIG administration | NLR = 1.27 and PLR = 201 (after) NLR = 3.83 and PLR = 150 (before) | Higher       |
| 8        | Kim et al. (26)         | Korea   | Prospective   | 129         | JKDRC | CRP | Before IVIG administration | CRP = 6.88 mg/dL | Lower                     |
| 9        | Kim et al. (22)         | Korea   | Retrospective | 5151        | AHA | CRP | Before IVIG administration | CRP = 3.68 mg/dL | Higher                     |
| 10       | Lee et al. (28)         | Korea   | Retrospective | 91          | AHA | CRP | Before and after IVIG administration | CRP = 8.3 mg/dL (before) and 3.4 mg/dL (after) | Higher       |
| 11       | Li et al. (36)          | China   | Retrospective | 957         | AHA | CAR | Before IVIG administration | CAR = 3.15 | Higher                     |
| 12       | Li et al. (32)          | China   | Retrospective | 1257        | AHA | NLR and PLR | Before IVIG administration | NLR = 3.58 PLR = 164 | Higher                     |
| 13       | Liu et al. (33)         | China   | Prospective   | 542         | AHA | NLR and PLR | Before IVIG administration | NLR = 3.37 PLR = 124 | Lower                     |
| 14       | Liu et al. (29)         | China   | Prospective   | 831         | AHA | NLR and PLR | Before IVIG administration | NLR = 3.24 PLR = 134 | Lower                     |
| 15       | Liu et al. (30)         | China   | Prospective   | 550         | AHA | CRP and CAR | Before IVIG administration | CRP = 57.7 mg/L | Lower                     |
| 16       | Lu et al. (27)          | China   | Retrospective | 94          | AHA | CRP | Before IVIG administration | CRP = 45 mg/L | Higher                     |
| 17       | Nandi et al. (20)       | India   | Prospective   | 72          | AHA | CRP | Before IVIG administration | CRP = 40 mg/L | Lower                     |
| 18       | Takeshita et al. (15)   | Japan   | Retrospective | 437         | JKDRC | NLR and PLR | Before IVIG administration | NLR = 3.83 PLR = 150 | Higher                     |
| 19       | Türkmen et al. (34)     | Turkey  | Retrospective | 94          | JKDRC | NLR and CRP | Before IVIG administration | NLR = 1.69 | Higher                     |
| 20       | Wu et al. (21)          | China   | Retrospective | 282         | AAP and AHA | NLR and PLR | Before IVIG administration | NLR = 2.69 PLR = 110.92 | Higher                    |
| 21       | Xie et al. (31)         | China   | Retrospective | 410         | JKDRC | CRP | Before IVIG administration | CRP = 100 mg/L | Higher                     |
| 22       | Yuan et al. (37)        | China   | Retrospective | 404         | AHA | NLR and PLR | Before and after IVIG administration | NLR = 4.36 and PLR = 162 (before) NLR = 1.45 and PLR = 196 (after) | Higher       |

AHA, American Heart Association; AAP, American Academy of Pediatrics; JKDRC, Japanese Kawasaki Disease Research Committee; NLR, Neutrophil Lymphocyte Ratio; PLR, Platelet lymphocyte ratio; CRP, C-reactive protein; CAR, C-reactive protein to albumin ratio; IVIG, Intravenous immunoglobulin.
age, index test, reference standards, true- and false-positive and negative values for each inflammatory marker. The first author (CL) entered the data, and the second author (JW) verified the correctness of the data entry.

Risk of bias assessment

The risk of bias was evaluated independently by two authors (CL and JW) using the quality assessment of diagnostic accuracy studies-2 (QUADAS-2) tool (19). The assessed domains were: patient selection, type and interpretation of index test and reference standards, and flow and timing of outcome assessment. Based on these domains, studies received a high, low, or unclear grade.

Statistical analysis

Statistical analysis was done by STATA "Midas" command package. Pooled indices of predictive accuracy, i.e., sensitivity, specificity, diagnostic odds ratio (DOR), likelihood ratio positive (LRP), and negative (LRN) for the predictive accuracy of inflammatory markers were assessed using bivariate meta-analysis. Graphical representation of the diagnostic accuracy indices was done by the forest plot, LR scattergram (clinical value of inflammatory markers), and Fagan plot (post-test probability of a patient having resistant Kawasaki disease). Summary receive operator characteristic curve (sROC showing area under the curve [AUC]) was done. Between-study variability (heterogeneity) was assessed using the $\chi^2$ test and $I^2$ statistic with graphical representation using a bivariate box plot. Publication bias was statistically assessed using Deek's test and graphically represented by the funnel plot. $P < 0.05$ was statistical significance.

Results

Study selection

The literature search identified 1,115 records. Of them, full texts were obtained from the 109 relevant studies. Three additional articles were identified by screening the references of the retrieved full texts during primary screening. After the final screening against eligibility criteria, 22 papers were included in this study (Figure 1) (13–15, 20–38).

Characteristics of the included studies

Most of the included studies (17 out of 22) were retrospective in nature. Most were conducted in China (10 studies) followed by Korea (5 studies) and Japan (4 studies).

Predictive accuracy of neutrophil-to-lymphocyte ratio

In total, 14 included studies reported on the utility of NLR for predicting resistant Kawasaki disease. Pooled sensitivity and specificity of NLR for predicting resistant KD were 72% (95% CI: 62%, 80%) and 71% (95% CI: 63%, 78%), respectively, with an AUC value of 0.77 (Figures 3A, 4A). The DOR was 6 (95% CI: 4, 10), LRP was 2.5 (95% CI: 2, 3.1), and LRN was 0.40 (95% CI: 0.29, 0.54). LRP and LRN in the right lower quadrant of the LR scattergram (Figure 5A) suggest no confirmation or exclusion value for NLR. As shown in Figure 6A, NLR has a good clinical value for predicting resistant Kawasaki disease (Positive: 37%; Negative: 8%), compared to the pre-test probability (19%). As shown in Figure 7 and by the bivariate box plot (Figure 7A), there was a significant inter-study heterogeneity ($\chi^2$ p < 0.001, $I^2 > 75$). There was no publication bias, as shown by Deck's test and the symmetrical funnel plot (Figure 8A).

Predictive accuracy of platelet-to-lymphocyte ratio

Nine studies reported the use of PLR for predicting resistant Kawasaki disease with pooled sensitivity and specificity of
FIGURE 3
(Continued)
FIGURE 3 (Continued)

Forest plot (A) NLR; (B) PLR; (C) CRP; (D) NLR and PLR combination.
60% (95% CI: 50%, 69%) and 68% (95% CI: 61%, 75%), respectively, and an AUC value of 0.69 (Figures 4B). The DOR was 3 (95% CI: 2.5, 5), LRP was 1.9 (95% CI: 1.5, 2.3), and LRN was 0.59 (95% CI: 0.48, 0.72). Positions of LRP and LRN on the LR scattergram (Figure 5B) indicated no confirmation or exclusion for PLR. PLR has a moderate clinical value in predicting resistant KD (Positive: 25%; Negative: 9%) that differed moderately from the pre-test probability (15%; Figure 6B). There was a significant inter-study variability ($\chi^2 p < 0.001$ and an $I^2 > 75$%). This heterogeneity was confirmed by a bivariate box plot (Figure 7B). We could not estimate the publication bias due to the small number of studies.

**Predictive accuracy of C-reactive protein**

In total, 11 included studies reported on the utility of CRP for predicting resistant KD, with pooled sensitivity and specificity of 75% (95% CI: 68%, 81%) and 66% (95% CI: 55%, 76%), respectively, with an AUC value of 0.78.
Predictive accuracy of the combination of neutrophil-to-lymphocyte ratio and platelet-to-lymphocyte ratio

Six studies reported the ability of NLR-PLR combination to predict the resistant Kawasaki disease with pooled sensitivity and specificity of 58% (95% CI: 46%, 69%) and 73% (95% CI: 65%, 79%), respectively, and an AUC value of 0.72 (Figures 3D, 4D). The DOR was 4 (95% CI: 3, 5), LRP was 2.1 (95% CI: 1.9, 2.4), and LRN was 0.58 (95% CI: 0.48, 0.70). LR scattergram (Figure 5D) showed that the NLR-PLR combination is not suitable for confirmation or exclusion. Figure 6D shows a good clinical utility of NLR and PLR combination for predicting IVIG-resistant KD (Positive: 35%; Negative: 13%) that differed significantly from the pre-test probability (20%). There was a significant inter-study heterogeneity ($\chi^2 p < 0.001; I^2 > 75\%$), shown also by the bivariate box plot (Figure 7D). Publication bias was not assessed due to a small number of studies.

Predictive accuracy of PAR

Only two of the included studies reported on the utility of PAR for predicting resistant Kawasaki disease. It was not possible, therefore, to pool the results and obtain the sensitivity, specificity, and other predictive accuracy indices. However, both
FIGURE 6
Fagan plot (A) NLR, (B) PLR, (C) CRP, (D) NLR and PLR combination.
studies have reported moderate predictive accuracy of sensitivity that ranged from 61 to 70% and specificity of 55–75%.

**Discussion**

Our systematic literature search found 22 studies that reported the use of NLR, PLR, CRP, CAR, and NLR in combination with PLR to predict resistant KD. Out of these markers, CRP showed the highest pooled sensitivity (75%) followed by NLR (72%). However, the pooled specificity was higher for NLR + PLR (73%) followed by NLR alone (71%) for the prediction of resistant KD. CRP and NLR had better overall sensitivity and specificity compared to all other markers. CRP is a protein that is produced by the hepatocytes in response to inflammation. CRP levels rise during the acute phase of the inflammatory process after the IL-6 and TNF-alpha activation (cytokine activation) (39), making it a well-established marker for Kawasaki disease (10, 40, 41). The extent of the increase in the levels of CRP can in turn provide evidence of the possible resistance KD.

Our results, showing the high predictive value of NLR, are in line with the previous review assessing the NLR accuracy for predicting the resistant KD (42). Neutrophils reflect the ongoing inflammatory process, while the lymphocytes act as a marker for immune regulatory response (25). Therefore, NLR can be a marker of balance between the inflammatory process and the immune regulatory response (43). The clinical value of NLR in our study was further confirmed by Fagan’s nomogram, which showed a marked increase in the post-NLR probability vs. pre-NLR probability.

In our review, PLR was identified as a moderately useful marker for predicting the resistant Kawasaki disease. Like the other inflammatory markers, elevated platelet counts are indicative of systemic infection and inflammation, since the increased levels of the proinflammatory cytokines cause the proliferation of megacaryocytes (44, 45). The combination of PLR and NLR showed good specificity but a weaker sensitivity, thus making it not useful as a marker for the identification of resistant KD.

Not enough studies, included in our review, evaluated CAR as a prognostic marker. Therefore, it was not possible to pool the evidence and obtain pooled estimates. However, studies
FIGURE 8

Funnel plot (A) NLR; (B) CRP.
reporting the accuracy of CAR for the identification of resistant KD, reported that its sensitivity and specificity were moderate. CAR, therefore, can be considered a useful marker.

Based on our review findings, all analyzed markers had moderate to a high level of sensitivity and specificity. We could not identify any specific index as the preferred marker for predicting resistant KD. This makes it a very useful marker for the prediction. Further studies, focusing on building a scoring system that combines all the important markers that can be applied to predict resistant Kawasaki disease at the early stages, are needed.

Our review has certain strengths. This meta-analysis involved 22 studies with a large sample size of > 7000 participants. We comprehensively assessed multiple inflammatory markers and identified the level of predictive accuracy for resistant Kawasaki disease. However, our review has certain limitations. Each study included in the review had wide range of biomarkers value considered to put the patient at high risk of IVIG failure. It is unclear which threshold is associated with such risk. Stratification of results based on cut-off was also not possible given the wide range and difference across the studies. Studies included in our review were of lower quality across most of the domains with significant publication bias for most markers, which might affect the credibility of our analysis. Moreover, inter-study heterogeneity that was detected may limit our ability to interpret the pooled results.

In conclusion, our study may contribute to further improvement of the clinical management of resistant Kawasaki disease. Our results suggest that inflammatory markers such as NLR, CRP, and PLR can be utilized as a useful screening and predictive tool. Moreover, early prediction of IVIG resistance in KD patients may lead to early initiation of additional anti-inflammatory therapy and more effective therapeutic management. There is a need for further updated reviews that would compare the predictive performance of the various individual and combined inflammatory markers in KD patients.

Data availability statement

The original contributions presented in this study are included in the article/supplementary material, further inquiries can be directed to the corresponding author.

Author contributions

CL and JW conceived and designed the study, collected the data, and performed the analysis. JW was involved in the writing of the manuscript and is responsible for the integrity of the study. Both authors have read and approved the final manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Modesti AM, Plewa MC. Kawasaki disease. St. Petersburg, FL: StatPearls Publishing (2021).

2. McCrindle BW, Rowley AH, Newburger JW, Burns JC, Bolger AF, Gewitz M, et al. Diagnosis, treatment, and long-term management of Kawasaki disease. A scientific statement for health professionals from the American heart association. Circulation. (2017) 135:e927–99. doi: 10.1161/CIR.0000000000000484

3. Muta H, Ishii M, Furuji J, Nakamura Y, Matusuhi T. Risk factors associated with the need for additional intravenous gamma-globulin therapy for Kawasaki disease. Acta Paediatr. (2006) 95:189–93. doi: 10.1080/08035250500327328

4. Tremoulet AH, Best BM, Song S, Wang S, Corinaldesi E, Eichenfeld JR, et al. Resistance to intravenous immunoglobulin in children with Kawasaki disease. J Pediatr. (2008) 153:117–21. doi: 10.1016/j.jpeds.2007.12.021

5. Tremoulet AH, Jain S, Jagg J, Jimenez-Fernandez S, Pancheri JM, Sun X, et al. Inflammab for intensification of primary therapy for Kawasaki disease: A phase 3 randomised, double-blind, placebo-controlled trial. Lancet. (2014) 383:1731–8. doi: 10.1016/S0140-6736(13)62298-9

6. Kobayashi T, Saji T, Otani T, Takeuchi K, Nakamura T, Arakawa H, et al. Efficacy of immunoglobulin plus prednisolone for prevention of coronary artery abnormalities in severe Kawasaki disease (RAISE study): A randomised, open-label, blinded-endpoints trial. Lancet. (2012) 379:1613–20. doi: 10.1016/S0140-6736(11)61930-2

7. Galeotti C, Kaveri SV, Cimaz R, Koné-Paut I, Barry J. Predisposing factors, pathogenesis and therapeutic intervention of Kawasaki disease. Drug Discov Today. (2016) 21:1850–7. doi: 10.1016/j.drudis.2016.08.004

8. Ogata S, Oghara Y, Honda T, Kon S, Akiyama K, Ishii M. Corticosteroid pulse combination therapy for refractory Kawasaki disease: A randomized trial. Pediatrics. (2012) 129:e17–23. doi: 10.1542/peds.2011-0148

9. Egami K, Muta H, Ishii M, Suda K, Sugahara Y, Iemura M, et al. Prediction of resistance to intravenous immunoglobulin treatment in patients with Kawasaki disease. J Pediatr. (2016) 219:237–40. doi: 10.1016/j.jpeds.2016.03.050

10. Kobayashi T, Inoue Y, Takeuchi K, Okada Y, Tamura K, Tomomasa T, et al. Prediction of intravenous immunoglobulin unresponsiveness in
patients with Kawasaki disease. Circulation. (2006) 113:2606–12. doi: 10.1161/CIRCULATIONAHA.105.592865

11. Sleeper LA, Minich LL, McCrinkle BM, Li JS, Mason W, Colan SD, et al. Evaluation of Kawasaki disease risk-scoring systems for intravenous immunoglobulin resistance. J Pediatr. (2011) 158:831–35.e3. doi: 10.1016/j.jpeds.2010.10.031

12. Fu P, Du Z, Pan Y. Novel predictors of intravenous immunoglobulin resistance in Chinese children with Kawasaki disease. Pediatr Infect Dis J. (2013) 32:e119–23. doi: 10.1097/INF.0b013e31828e87f1

13. Ha K-S, Lee J, Jang GV, Lee J, Lee KC, Snn CS, et al. Value of neutrophil–lymphocyte ratio in predicting outcomes in Kawasaki disease. Am J Cardiol. (2015) 115:301–6. doi: 10.1016/amjcard.2015.04.021

14. Kawamura Y, Takeshita S, Kanai T, Yoshida Y, Nonoyama S. The combined usefulness of the neutrophil-to-lymphocyte and platelet-to-lymphocyte ratios in predicting intravenous immunoglobulin resistance with Kawasaki disease. J Pediatr. (2016) 178:281–284.e1. doi: 10.1016/j.jpeds.2016.07.035

15. Takeshita S, Kanai T, Kawamura Y, Yoshida Y, Nonoyama S. A comparison of the predictive validity of the combination of the neutrophil-to-lymphocyte ratio and platelet-to-lymphocyte ratio and other risk scoring systems for intravenous immunoglobulin (ivig)-resistance in Kawasaki disease. PLoS One. (2017) 12:e0176957. doi: 10.1371/journal.pone.0176957

16. Chaudhary H, Nameirakpam J, Kumrah R, Pandiarajan V, Suri D, Rawat A, et al. Biomarkers for Kawasaki disease: Clinical utility and the challenges ahead. Front Pediatr. (2019) 7:242. doi: 10.3389/fped.2019.00242

17. Yaya Akca U, Arslangolu Aydin E, Ayhan HH, Serin O, Sag E, Demir S, et al. Comparison of IVIG resistance predictive models in Kawasaki disease. Pediatr Res. (2022) 91:621–6.

18. Page MJ, McKenzie JE, Bossoy PM, Bourron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. BMJ. (2021) 372:n71. doi: 10.1136/bmj.n71

19. Whiting PF, Rutjes AWS, Wood MJ, Lewington S, McIsaac G, Collins RA, et al. QUADAS-2: A revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med. (2011) 155:529–36. doi: 10.7326/0003-4819-155-8-201110180-00009

20. Nandi A, Pal P, Basu S. A comparison of serum IL6 and CRP levels with respect to coronary changes and treatment response in Kawasaki disease patients: A prospective study. Rheumatol Int. (2019) 39:1797–801. doi: 10.1007/s00296-019-04357-9

21. Wu S, Long Y, Chen S, Huang Y, Lao Y, Sun Y, et al. A new scoring system for prediction of intravenous immunoglobulin resistance of Kawasaki disease in infants under 1-year old. Front Pediatr. (2019) 7:514. doi: 10.3389/fped.2019.00514

22. Kim MK, Song MS, Kim GB. Factors predicting resistance to intravenous immunoglobulin treatment and coronary artery lesion in patients with Kawasaki disease: Analysis of the Korean nationwide multicenter survey from 2012 to 2014. Korean Circ J. (2015) 45:871–9. doi: 10.7499/j.issn.1008-8830.2015.04.010

23. Cho H-J, Bak SY, Kim SY, Yoo R, Baek H-S, Yang S, et al. High neutrophil–lymphocyte ratio is associated with refractory Kawasaki disease. Pediatr Infect Dis J. (2017) 36:1366–73. doi: 10.1097/INF.0000000000002566

24. Iwashima S, Nameirakpam J, Kumrah R, Pandiarajan V, Suri D, Rawat A, et al. Levels of serum cytokines and acute phase proteins in Kawasaki disease. Circulation. (2016) 178:281–284.e1. doi: 10.1161/circulationaha.105.592865

25. Klinger MHF, Jelkmann W. Role of blood platelets in infection and inflammation. J Interferon Cytokine Res. (2002) 22:913–22. doi: 10.1089/1079990026286623

26. Türkçar S, Yıldız K, Acar C, Dundar HA, Kir M, Unsal E. Risk factors of intravenous immunoglobulin resistance and coronary arterial lesions in Turkish children with Kawasaki disease. Turk J Pediatr. (2020) 62:1–9. doi: 10.24953/turkped.2020.01.001

27. Yuan Y-D, Sun J, Li P, Wei C-L, Yu Y-H. Values of neutrophil–lymphocyte ratio and platelet–lymphocyte ratio in predicting sensitivity to intravenous immunoglobulin in Kawasaki disease. J Chin Med Sci. (2021) 36:1366–73. doi: 10.1007/s00380-021-01819-w

28. Faim D, Henriques C, Brett A, Francisco A, Rodrigues F, Pires A. Kawasaki disease: Predictors of resistance to intravenous immunoglobulin and cardiac complications. Arq Bras Cardiol. (2021) 116:885–91. doi: 10.60660/abc.2019.0758

29. Moon KP, Kim BJ, Lee KI, Oh JH, Han JW, Lee KY, et al. Prediction of nonresponsiveness to medium-dose intravenous immunoglobulin (1 g/kg) treatment: An effective and safe schedule of acute treatment for Kawasaki disease. Korean J Pediatr. (2016) 59:178–82. doi: 10.3345/kjp.2016.59.4.178

30. Sano T, Kurotobi S, Matsuzaki K, Yamamoto T, Maki I, Mikitani T, et al. Prediction of non-responsiveness to standard high-dose gamma-globulin therapy in patients with acute Kawasaki disease before starting initial treatment. Eur J Pediatr. (2020) 179:131–7. doi: 10.1007/s00431-020-02923-z

31. Wu G, Yue P, Ma F, Zhang Y, Zheng X, Li Y. Neutrophil-to-lymphocyte ratio as a biomarker for predicting the intravenous immunoglobulin-resistant Kawasaki disease. Medicine (Baltimore). (2020) 99:e18535. doi: 10.1097/MD.00000000000018535

32. Park JH, Ji J-H, Oh J-Y, Yoon C-H, Suh J-W, Cho Y-S, et al. Prognostic value of neutrophil-to-lymphocyte ratio in patients presenting with ST-elevation myocardial infarction undergoing primary percutaneous coronary intervention. Am J Cardiol. (2015) 116:636–42. doi: 10.1016/j.amjcard.2012.11.012

33. Klinger MHF, Jelkmann W. Role of blood platelets in infection and inflammation. J Interferon Cytokine Res. (2002) 22:913–22. doi: 10.1089/1079990026286623

34. Alexandrakis MG, Passam FH, Moschandrea IA, Christophoridis AV, Pappa CA, Coulocheri SA, et al. Levels of serum cytokines and acute phase proteins in patients with essential and cancer-related thrombocytosis. Am J Clin Oncol. (2003) 26:155–60. doi: 10.1097/00000421-200304000-00007

35. Mackiewicz A, Speroff T, Ganapathi MK, Kushner I. Effects of cytokine combinations on acute phase protein production in two human hepatoma cell lines. J Immunol. (1991) 146:3032–7.