The Concentrations of Ambient *Burkholderia Pseudomallei* during Typhoon Season in Endemic Area of Melioidosis in Taiwan

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**Abstract**

Background: Melioidosis is a severe bacterial infection caused by *Burkholderia pseudomallei* with a high case-fatality rate. Epidemiological and animal studies show the possibility of inhalation transmission. However, no *B. pseudomallei* concentrations in ambient air have been researched. Here, we developed a method to quantify ambient *B. pseudomallei* and then measured concentrations of ambient *B. pseudomallei* during the typhoon season and the non-typhoon season to determine the factors influencing ambient *B. pseudomallei* levels.

Methods: We quantified ambient *B. pseudomallei* by using a filter/real-time qPCR method in the Zoynan Region in Kaohsiung, southern Taiwan. Twenty-four hour samples were collected at a sampling rate of 20 L/min every day from June 11 to December 21, 2012 including during the typhoon season (June to September) and reference season (October to December).

Results: We successfully developed a filtration/real-time qPCR method to quantify ambient *B. pseudomallei*. To our knowledge, this is the first report describing concentrations of ambient *B. pseudomallei*. Ambient *B. pseudomallei* were only detected during the typhoon season when compared to the reference season. For the typhoons affecting the Zoynan Region, the positive rates of ambient *B. pseudomallei* were very high at 80% to 100%. During June to December, rainfall was positively correlated with ambient *B. pseudomallei* with a statistical significance. Sediment at a nearby pond significantly influenced the concentration of ambient *B. pseudomallei*. During the typhoon month, the typhoon was positively correlated with ambient *B. pseudomallei* whereas wind speed was reversely correlated with ambient *B. pseudomallei*.

Conclusions: Our data suggest the possibility of transmission of *B. pseudomallei* via inhalation during the typhoon season.

Introduction

Melioidosis, which is endemic in northern Australia and Southeast Asia, is an emerging infection in other Asian regions and in South America [1–6]. Sporadic autochthonous cases have also been reported throughout the world, including Africa, the Caribbean, America, and the Middle East [1,7–8]. Melioidosis is a severe bacterial infection caused by *Burkholderia pseudomallei* with high case-fatality rates of 14% to 40% in Thailand, Australia, Singapore and Taiwan [1,6,9–11]. With increasing worldwide travel of both humans and animals, and the concern of *B. pseudomallei* as an important potential bioweapon, this agent is an emerging global public health problem [1–3,7,10,12].

Risk factors of melioidosis are diabetes, hazardous alcohol use, chronic lung or renal disease, and older age [1,10,13]. Pneumonia is the most common clinical presentation [1,5,10,14–15]. The association between melioidosis and rainfall intensity is well documented from endemic regions [5–6,10,13–14]. A significant linear correlation has been observed between rainfall and melioidosis cases [1,5–6,13,15–16]. The case clusters were also associated with sudden and heavy rainfall related to cyclones and typhoons [1,4,6,10,14–15]. Although outbreaks have been linked to contamination of drinking water [1], it is now believed that percutaneous inoculation is the major mode of acquisition [1,19–21]. Recently,
epidemiological studies support inhalation as the mode of transmission of *B. pseudomallei* 

after heavy monsoonal rains and winds [5,6,12–13,17,21–22]. Animal studies also show that exhaled *B. pseudomallei* aerosols lead to a lower lethal dose, 50% (LD50) and a shorter incubation time compared to intraperitoneal and subcutaneous injection [23–24]. However, no *B. pseudomallei* concentrations in ambient air have been researched. 

There are very limited researches regarding quantifying ambient pathogens. The first study of quantifying airborne pathogens by using filter/real-time qPCR was to determine the concentrations of airborne *M. tuberculosis* in hospitals [25–26]. Then, airborne influenza virus and avian influenza virus was also quantified in poultry markets [27]. Recently, the first report describing the concentration of ambient pathogens implied the possibility of long-range transport of influenza virus because the concentration of ambient influenza A virus was significantly higher during the Asian dust storm days than during the background days [28]. This quantitative method shows promise for quantifying ambient pathogens with high sensitivity and specificity and should provide deeper insight into infectious disease transmissibility and epidemiology, as well as infection control. Therefore, the aim of the current study was to develop a method to quantify ambient *B. pseudomallei* and to determine the factors influencing ambient *B. pseudomallei* levels.

**Materials and Methods**

**Reference bacteria and genomic DNA**

Reference strains of *B. pseudomallei* (vgh19) were kindly provided by the laboratory for Biotechnology, National Kaohsiung Normal University (Kaohsiung, Taiwan); their characteristics have been described in previous studies [29]. We target the *B. pseudomallei* – specific type III secretion system (TTSS) gene cluster encompassing part of open reading frame 2 (orf2) [30], which was recently reported to be the most accurate clinically and has shown sensitivity beyond culture on soil samples for *B. pseudomallei* detection [31]. The target DNA standard solution of *B. pseudomallei* was purchased from Mission Biotech (Taipei, Taiwan).

**DNA isolation and quantification**

We extracted genomic DNA from *B. pseudomallei* with the QIAamp DNA Mini kits (QIAGEN, GmbH, Hilden, Germany) as previously described [26]. The bacterial DNA was stored at −20°C within one month before the analysis.

![Standard curve of known DNA (1.0 × 10⁶ to 1.0 × 10⁷ copies/µl) and threshold cycle (Ct) measured using a real-time qPCR for *B. pseudomallei* with R² of 0.998 and slope of −3.46.](https://doi.org/10.1371/journal.pntd.0002877.g001)
For *B. pseudomallei*, the primers and probe were primarily targeted to TTSS-orf2 with the sensitivity and specificity of 100% for *B. pseudomallei* [30]. The sequence of primers were BpTT4176F (5′-CGTCTCTATACTGTCGAGCAATCG-3′), BpTT4290R (5′-CGTGCACACCGGTCAGTATC-3′), and the probe was fluorogenic probe BpTT4208P (5′-CCGGAATCTGGATCAC-CACCACTTTCC-3′). The PCR assay was performed at a final volume of 25 μl and 45 cycles as previously described [32] and amplification and detection were performed on the 7900HT Fast Real-Time qPCR System (Applied Biosystems, Inc., Foster City, CA).

Ten-fold serial dilutions of the target DNA standard solution was made for the calibration curve. In order to achieve PCR efficiency of 90% to 110%, the slope of the calibration curve must range from −3.6 to −3.1. The R²-value must be above 0.99. The positive and negative controls were analyzed with each run. The negative controls were all uncontaminated. All samples, positive controls and negative controls were analyzed in triplicate.

**Chamber study**

**Comparison of real-time qPCR and culture.** To understand the relationship between culture and the quantitative

![Figure 2](https://www.plosntds.org/article/f10.1371/journal.pntd.0002877.g002)

Figure 2. (a) Teflon and PC filters comparison over 24 hours and 48 hours sampling after spiking the same quantity of *B. pseudomallei* (b) transport temperature comparison of 4°C and 25°C over 24 hours and 48 hours after spiking the same quantity of *B. pseudomallei* on Teflon filters. doi:10.1371/journal.pntd.0002877.g002
analysis of real-time PCR, we activated the bacteria (vgh19) and cultured 100 μl to Ashdown’s agar for two days. We also took 50 μl of the same batch to autoclaved polytetrafluoroethylene (PTFE; Teflon) membrane filters, then extracted DNA and detected the concentrations by real-time qPCR.

Comparison of filters. We spiked the same quantity of B. pseudomallei into 37 mm diameter with 0.1 μm pore size Teflon filter and polycarbonate (PC) filter. We compared the Teflon filter and PC filter in the laboratory after 24 hours and 48 hours sampling at 20 L/min.

Transport temperature. In order to determine the optimal transport temperature for the air samples, we spiked equal volumes of B. pseudomallei on autoclaved Teflon filters and stored at 4°C and 25°C for 24 hours and 48 hours, respectively.

Matrix effect. We collected air samples over 24 hours on Teflon filters at 20 L/min and then spiked with B. pseudomallei. In addition, we also spiked equal volumes on clean Teflon filters. Finally, we extracted DNA of B. pseudomallei to determine the inhibitory effect of air matrix.

Field study

Sampling strategy. From 2005 to 2011, the number of melioidosis cases in Kaohsiung accounted for 70.3% cases in Taiwan and there was a hotspot of both cases and deaths in the Zoynan Region [11,16,22]. We chose the second floor of the elderly activity center with a distance of 11.4 meters to the Lotus Pond in the Zoynan Region in Kaohsiung, southern Taiwan, as our sampling site. Airborne B. pseudomallei was collected on Teflon filters in the disposable plastic cassettes by using a sampling pump operating at 20 L/min with a sampling time of 24 hours. We sampled air every day from June 11 to December 21, 2012 including the typhoon season (June to September) and the reference season (October to December). Before sampling, the filters and support pads were autoclaved, and the plastic cassettes were sterilized with ethylene oxide.

Environmental parameters. During the study period, the air pollutants and climate factors were provided by the Taiwan Environmental Protection Administration and the Taiwan Central Weather Bureau, respectively. The environmental parameters included concentrations of ozone (O₃), sulfur dioxide (SO₂), carbon monoxide (CO), and nitrogen oxide (NOₓ). The climate factors included typhoon information, rainfall, wind speed, and ultraviolet index (UV index; 1 unit equals 25 mW/m²). From these data we calculated the mean air pollutants and climate factors for defined periods before date of ambient B. pseudomallei measurement. Since there is no study regarding the appropriate day-lag of ambient pollutant concentrations and climate factors to consider when studying their relations with ambient B. pseudomallei, we examined the effects of 0-, 1-, 2-, 3-day lags (lag 0, lag 1, lag 2, and lag 3) and 2-days, 3-days, and 4-days sum before the date ambient B. pseudomallei measurement.

Ambient bacteria and fungi. Duplicate total cultivable airborne bacteria and fungi samples were collected using portable microbiological air sampler (MAS-100; MERCK, USA). The tryptic soy agar (TSA, Difco Laboratories, Michigan, USA) and malt extract agar (MEA, Difco Laboratories, Michigan, USA) was used for bacteria and fungi, respectively. Bacteria were incubated at 37°C for 24 hours and fungi were incubated at 25°C for 48 hours. The results were expressed as colony forming unit per cubic meter air (CFU/m³).

Statistical methods

Statistical analyses were performed using SPSS for Windows Chinese Traditional 14.0. The descriptive statistics were used to evaluate the range, mean and standard error of parameters. The Mann-Whitney U test was used to evaluate the difference of ambient B. pseudomallei and environmental factors between different groups. Since the distribution of ambient B. pseudomallei did not fit to normal distribution (significant result was found in Kolmogorov-Smirnov test), we used Spearman correlation to evaluate relationships between B. pseudomallei and meteorological factors. Significance was accepted at p-value<0.05.

Results

Development of filter/real-time qPCR method

Figure 1 shows the calibration curve of B. pseudomallei from 1.0×10⁰ to 1.0×10⁷ copies/μl with R² of 0.998 and slope of −3.46. Because the DNA on each filter was extracted to a final volume of 200 μl, the detection limit of the filter/real-time qPCR method for B. pseudomallei was 6.94 copies/m². If the positive results were found in the 1/100 dilution samples due to the inhibitory effect, then the actual detection limit corrected for this effect was 0.07 copies/m³. Based on our results, 1 copy/μl in the real-time qPCR assay was approximately equal to 3.5 CFU/μl.

Figure 2 (a) shows the B. pseudomallei concentrations on Teflon and PC filters over 24 hours and 48 hours of sampling after spiking the same quantity of B. pseudomallei. The concentrations of B. pseudomallei on Teflon filters of both 24-hour and 48-hour samples were significantly higher than that on PC filters. So, we

| Typhoon | The period of issuing sea alert | Characteristics | B. pseudomallei |
|---------|--------------------------------|----------------|----------------|
|         |                               | Minimum pressure (hPa) | Maximum wind speed (m/s) | Concentration (copies/m³) | Positive rate % |
| TALUM   | 06/19~06/21                   | 985             | 25             | N.D.ª | 0% (0/3) |
| DOKSURI | 06/28~06/29                   | 995             | 23             | N.D.~25.2 | 20% (1/5) |
| SAOLA   | 07/30~08/03                   | 960             | 38             | N.D.~25.2 | 20% (1/5) |
| HAIKUI  | 08/06~08/07                   | 960             | 35             | N.D.~87.2 | 50% (1/2) |
| KAI-TAK | 08/14~08/15                   | 995             | 20             | 142.2~257.5 | 100% (2/2) |
| TEMBIN  | 08/21~08/28                   | 945             | 45             | N.D.~126.1 | 80% (4/5) |
| JELAWAT | 09/27~09/28                   | 910             | 55             | N.D. | 0% (0/2) |

ªN.D. means not detected.

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Figure 3. The path of (a) Talim, (b) Doksur, (c) Saola, (d) Haikui, (e) Kai-Tak, (f) Tembin and (g) Jelawat Typhoon from Taiwan
Central Weather Bureau.

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evaluated the influence of transport temperature on Teflon filters. Figure 2 (b) shows \( B. \) pseudomallei concentrations with the transport temperature at 4°C and 25°C over 24 hours and 48 hours for the spiked samples on Teflon filters. The concentrations of \( B. \) pseudomallei at 4°C for both 24-hour and 48-hour samples were significantly higher than that at 25°C. For inhibitory effect evaluation, our results show that the extracted DNA of \( B. \) pseudomallei at 4°C over 24 hours and 48 hours for the ambient aerosols (9.2 \( \times \) 10^4 copies/m^3) after spiking same quantity of \( B. \) pseudomallei (\( p \)-value = 0.01).

**Ambient \( B. \) pseudomallei and typhoon events**

We measured ambient \( B. \) pseudomallei from June 11 to December 31, with a total of 188 samples (we lost six samples from June 28 to June 30 and August 23 to 26). The positive rate by qPCR (the number of positive samples divided by the number of all samples) is 20.2% with the concentration range from not detected to 4 \( \times \) 10^4 copies/m^3. According to Taiwan Central Weather Bureau, seven typhoons affected Taiwan in 2012 with the definition of “affected periods” as the period of issuing a sea alert to the typhoon [33]. Table 1 shows the information of the seven typhoons as well as ambient \( B. \) pseudomallei and environmental characteristics during the typhoon-affected periods. The positive rate during Kai-Tak and Tembin affected periods was higher than that of other typhoons. The path of Talim, Doksuri, Haikui, Kai-Tak, and Jelawat were far away from Taiwan, whereas Saola and Tembin landed Hualien (East Taiwan), and Pingtung (South Taiwan), respectively (Figure 3). Only Doksuri, Kai-Tak, and Tembin affected the southern part of Taiwan where we sampled ambient \( B. \) pseudomallei.

Table 2 shows the descriptive statistics for ambient \( B. \) pseudomallei and environmental factors during the typhoon season (June to September) and the reference season (October to December) near Lotus Pond in South Taiwan. Table 3 shows the descriptive statistics for ambient \( B. \) pseudomallei and environmental factors in June, July, August and September.

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**Table 2.** Descriptive statistics for ambient \( B. \) pseudomallei and environmental factors during the typhoon season (June to September) and the reference season (October to December) near Lotus Pond in South Taiwan.

|                      | Typhoon season (June–September) | Reference season (October–December) | \( p \)-value |
|----------------------|----------------------------------|-------------------------------------|--------------|
| \( B. \) pseudomallei (%) | Positive rate | 35.9% (38/106) | 0% (0/82) |          |
| Inhibitory effect | 63.2% (24/38) | 0% | | |
| \( B. \) pseudomallei (copies/m^3) | Mean ± SE (Range) | 500.6±3965.4 (N.D.–41376.0) | N.D. (N.D.) | <0.001* |
| Rainfall (mm) | Mean ± SE (Range) | 13.7±31.9 (N.D.–155.0) | 0.7±4.5 (N.D.–39.5) | <0.001* |
| Wind (m/s) | Mean ± SE (Range) | 2.1±0.9 (1.1–7.2) | 1.8±0.5 (1.1–3.6) | 0.029* |
| UV Index^b | Mean ± SE (Range) | 8.5±2.9 (1.0–13.0) | 5.5±1.4 (1.0–8.0) | <0.001* |
| Ambient Fungi (CFU/m^3) | Mean ± SE (Range) | 400.3±781.1 (513.3–2590.4) | 869.0±1399.3 (429.9–457.3) | <0.001* |
| Ambient Bacteria (CFU/m^3) | Mean ± SE (Range) | 241.1±35.4 (68.9–6849.8) | 157.9±9.8 (122.5–7884.0) | 0.357 |

\*N.D. means not detected; \^11 unit equals 25 mW/m^2; \*significant at \( p \)-value < 0.05.

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**Table 3.** Descriptive statistics for ambient \( B. \) pseudomallei and environmental factors in June, July, August and September.

|                      | June | July | August | September |
|----------------------|------|------|--------|-----------|
| \( B. \) pseudomallei (%) | Positive rate | 11.8% (2/17) | 35.5% (11/31) | 58.1% (18/31) | 33.3% (10/30) |
| \( B. \) pseudomallei (copies/m^3) | Range | N.D.–743.0 | N.D.–41376.0 | N.D.–257.5 | N.D.–155.8 |
| Rainfall (mm) | Mean ± SE | 47.9±43.7 | 1656.4±1327.8 | 47.7±11.3 | 30.9±8.8 |
| Wind (m/s) | Mean ± SE | 37.0±11.4 | 1.7±0.6 | 24.4±7.5 | 1.8±1.2 |
| UV Index^b | Range | 1.8–5.0 | 1.2–2.9 | 1.1–7.2 | 1.3–4.8 |

\*N.D. means not detected; \^11 unit equals 25 mW/m^2.

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1 unit equals 25 mW/m^2; 1 unit equals 25 mW/m^2; *significant at \( p \)-value < 0.05.

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difference (p-value < 0.05). However, the concentrations of ambient fungi, O₃, SO₂, CO, and NOx were significantly higher during the reference season than typhoon season. Table 3 shows the descriptive statistics for ambient B. pseudomallei and environmental factors in June, July, August and September, separately. Figure 4 shows the ambient B. pseudomallei concentration profile from June to September.

**Ambient B. pseudomallei and environmental parameters**

During June to December, the concentrations of ambient B. pseudomallei were positively associated with rainfall and UV of lag 0, lag 1, lag 2, 2-day sum, 3-day sum, and 4-day sum (Table 4). However, no significant correlation was observed between ambient B. pseudomallei and environmental parameters during typhoon season. In August, there were four typhoons affecting Taiwan; therefore, we defined August as “typhoon month.” During typhoon month, the concentrations of ambient B. pseudomallei were negatively associated with wind speed at lag 0, 2-day sum, and 3-day sum with marginal p-values of 0.079, 0.093 and 0.080, respectively. When we divide environmental parameters by the mean and median values of samples, we found that ambient B. pseudomallei of rainy days (> median value of 0 mm) was significantly higher than that of no rainfall days with p-value of 0.006 during June to December (Table 5). During typhoon month, ambient B. pseudomallei of high-wind-speed days divided by mean value (2.4 m/s) were significantly lower than that of low-wind-speed days.

**Discussion**

In this study, we developed a filtration/real-time qPCR method to quantify ambient B. pseudomallei with a wide dynamic range over 8 orders of magnitude and a correlation coefficient (r) value of 0.998. The detection limit was 6.94 copy/m³, which is much lower than that in the previous studies for airborne M. tuberculosis [583 copy/m³] and airborne influenza A, B, and A/H5 virus [886, 653 and 1236 copy/m³, respectively] [26–27]. The low detection limit and wide linear range demonstrate that this newly established method is a promising tool for deeper insight into transmissibility and epidemiology of melioidosis, as well as infection control.

In our laboratory evaluation, Teflon filters show better performance when compared to PC filters. Teflon filters were also used for measuring airborne influenza virus in poultry markets and ambient influenza virus [27–28]. Our results showed that samples transported at 4°C were superior to 25°C. Inhibitory effects were also observed. In a previous study, authors found that air samples containing bacteria and fungi may inhibit PCR amplification and dilution of these samples can resolve these problems [34]. Therefore, optimistic protocols for ambient B. pseudomallei quantification in our follow-up field evaluations used Teflon filters to sample ambient B. pseudomallei, transported samples at 4°C to our laboratory within one hour, and analyzed samples simultaneously using 1, 1/10 and 1/100 dilutions.

Melioidosis is a severe bacterial infection with high case-fatality rates of approximately 40% in Thailand, 30% to 14% in Australia, 40% in Singapore, and 22.7% in Taiwan [1,6–9]. It is now believed that percutaneous inoculation is the major mode of acquisition due to exposure history to polluted water and mud, high risk among farmers, and wide isolation of B. pseudomallei from soil, mud, and pooled surface water in endemic areas [1,19]. However, pneumonia was the most common clinical presentation, which accounted for 32.6%, 51%, 45%, 42.1% and 70% of cases in India [5], northern Australia [10], Thailand [1], Malaysia [13], and Taiwan [14], respectively. In addition, epidemiological studies hypothesized that inhalation of B. pseudomallei was the mode of transmission after heavy monsoonal rains and winds [5,6,10,13,17,20–22]. In this study, we successfully quantified ambient B. pseudomallei using filtration/real-time qPCR. To our knowledge, this is the first report describing concentrations of B. pseudomallei in ambient air. Our results provided evidence to support the hypothesis from these epidemiological studies of inhalation transmission.

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B. pseudomallei is widely isolated from soil and water samples [35–38]. In Taiwan, there was a cluster of melioidosis for both cases and deaths in the Zoynan Region [11]. B. pseudomallei was isolated from this region at a rate of 25.9% and 13.5% for soil samples and water samples, respectively [11]. Our results of 20.2% positive air samples was similar to that of water and soil samples.
Table 4. The association between ambient *B. pseudomallei* and environmental parameters of lag 0, lag 1, lag 2, 2-days sum, 3-days sum, and 4-days sum during whole period (June to December), typhoon season (June to September), and typhoon month (August).

|                      | Whole period (June–December) (N = 191) | Typhoon season (June to September) (N = 109) | Typhoon month (August) (N = 31) |
|----------------------|----------------------------------------|--------------------------------------------|-------------------------------|
|                      | r          | p-value  | r          | p-value  | r          | p-value  |
| Rainfall             |            |          |            |          |            |          |
| lag 0                | 0.184      | 0.011*   | −0.041     | 0.671    | −0.050     | 0.788    |
| lag 1                | 0.140      | 0.053    | −0.074     | 0.446    | −0.140     | 0.453    |
| lag 2                | 0.176      | 0.015*   | −0.044     | 0.653    | −0.052     | 0.779    |
| lag 3                | 0.220      | 0.002*   | 0.012      | 0.899    | 0.255      | 0.166    |
| 2-days sum           | 0.181      | 0.012*   | −0.065     | 0.500    | −0.115     | 0.537    |
| 3-days sum           | 0.218      | 0.002*   | −0.038     | 0.694    | −0.135     | 0.468    |
| 4-days sum           | 0.235      | 0.001*   | −0.028     | 0.773    | −0.039     | 0.834    |
| Wind Speed           |            |          |            |          |            |          |
| lag 0                | −0.019     | 0.794    | −0.155     | 0.108    | −0.320     | 0.079    |
| lag 1                | 0.004      | 0.952    | −0.133     | 0.168    | −0.032     | 0.863    |
| lag 2                | 0.102      | 0.161    | 0.014      | 0.888    | −0.066     | 0.722    |
| lag 3                | 0.137      | 0.059    | 0.103      | 0.285    | 0.266      | 0.148    |
| 2-days sum           | −0.014     | 0.848    | −0.176     | 0.067    | −0.307     | 0.093    |
| 3-days sum           | 0.034      | 0.642    | −0.139     | 0.150    | −0.319     | 0.080    |
| 4-days sum           | 0.086      | 0.237    | −0.074     | 0.442    | −0.197     | 0.287    |
| UV                   |            |          |            |          |            |          |
| lag 0                | 0.246      | 0.001*   | 0.021      | 0.832    | 0.045      | 0.809    |
| lag 1                | 0.215      | 0.003*   | −0.027     | 0.777    | 0.126      | 0.500    |
| lag 2                | 0.202      | 0.005*   | −0.035     | 0.721    | 0.159      | 0.392    |
| lag 3                | 0.298      | <0.001*  | 0.139      | 0.150    | 0.020      | 0.913    |
| 2-days sum           | 0.263      | <0.001*  | −0.008     | 0.936    | 0.090      | 0.632    |
| 3-days sum           | 0.282      | <0.001*  | −0.017     | 0.864    | 0.189      | 0.310    |
| 4-days sum           | 0.320      | <0.001*  | 0.050      | 0.608    | 0.150      | 0.420    |

*p-value < 0.05; lag 0, lag 1, lag 2, and lag 3: the effects of 0-, 1-, 2-, 3-day lags before the date ambient *B. pseudomallei* measurement.

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Table 5. Associations between ambient *B. pseudomallei* concentrations and environmental parameters (rainfall and wind speed) divided by the mean and median values during whole period (June to December), typhoon season (June to September), and typhoon month (August).

| Ambient *B. pseudomallei* (copies/m³) | whole period (June–December) | Typhoon season (June–September) | Typhoon month (August) |
|--------------------------------------|-----------------------------|---------------------------------|------------------------|
|                                      | Rainfall                    | Rainfall                         | Rainfall               |
| Divided by the mean value of samples |                              |                                 |                        |
|                                      | N                           | mean    | p-value | N   | mean    | p-value | N   | mean    | p-value |
| ≥ 8.1 mm                             | 26                          | 45.0    | 0.17    | 22  | 49.7    | 0.49    | 10  | 33.3    | 0.85    |
| < 8.1 mm                             | 165                         | 325.3   |          | 87  | 621.2   |          | 21  | 54.5    |          |
| Wind speed                           |                              |         |        | Wind |         |        |     |         |        |
| ≥ 2.0 m/s                            | 65                          | 680.6   | 0.91    | 36  | 153.6   | 0.16    | 9   | 22.5    | <0.001* |
| < 2.0 m/s                            | 126                         | 77.2    |         | 73  | 165.1   |         | 22  | 58.0    |         |
| Divided by the median value of samples |                              |         |        |      |         |        |     |         |        |
|                                      | Rainfall                    | Rainfall |         | Rainfall |         |
|                                      | N                           | mean    | p-value | N   | mean    | p-value | N   | mean    | p-value |
| > 0.0 mm                             | 69                          | 101.5   | 0.006*  | 54  | 120.7   | 0.42    | 14  | 32.7    | 0.60    |
| ≤ 0.0 mm                             | 122                         | 389.9   |         | 55  | 873.7   |         | 17  | 60.0    |         |
| Wind speed                           |                              |         |        | Wind |         |        |     |         |        |
| > 1.8 m/s                            | 94                          | 493.7   | 0.80    | 54  | 836.0   | 0.10    | 15  | 39.7    | 0.36    |
| ≤ 1.8 m/s                            | 97                          | 84.2    |         | 55  | 171.4   |         | 16  | 55.2    |         |

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and implicated that ambient \textit{B. pseudomallei} should be a concern as a significant source of transmission.

In 2012, seven typhoons affected Taiwan whereas only three typhoons (Doksuri, Kai-Tak, and Tembin) affected Zoynan Region. We lost the ambient \textit{B. pseudomallei} samples during Doksuri-affected periods due to communication mistakes with the personnel managing the sampling site. For the other two typhoons affecting the Zoynan Region, the positive rates of ambient \textit{B. pseudomallei} were very high at 100% and 90% for Kai-Tak and Tembin, respectively. In addition, ambient \textit{B. pseudomallei} were only detected during the typhoon season when comparing to the reference season. Our results support the hypothesis that heavy monsoonal rains and winds may cause a shift toward inhalation of \textit{B. pseudomallei} [13] and provide a possible explanation for the observed growth in the number of melioidosis cases in the Taiwan Zoynan Region from the 2005 and 2009 typhoon season [16,20]. Regarding air pollutants, the significantly higher concentrations of ambient fungi, \textit{O}_3, \textit{SO}_2, \textit{CO} and \textit{NO} were observed during the reference season when compared to the typhoon season. Pollutants concentrated in autumn due to high atmospheric pressure may provide the explanation [39].

When investigating ambient \textit{B. pseudomallei} in June, July, August and September separately, we found that although the highest positive rate was observed in August, the highest concentrations were all observed in July with the lowest rainfall among June to September (Table 3, Figure 4). This observation is contrary to the results in the previous studies [5–6,10,13–14]. During our sampling period, we found that the water of the Lotus Pond was pumped out on June 27 to let sediment expose to the air. The authority (Department of Health, Kaohsiung city government) did confirm that water was pumped out on June 27 prior to the typhoon to avoid waterlogging. Since we lost samples from June 28 to June 30, we compared the concentration of June and July to see the association between ambient \textit{B. pseudomallei} and the appearance of sediment. The concentration of ambient \textit{B. pseudomallei} in July was higher than that in June with a marginal \textit{p}-value of 0.052. For evaluating the association between the appearance of sediment and ambient \textit{B. pseudomallei} from June to September, the linear-by-linear association test was used after logarithmic transformation of ambient \textit{B. pseudomallei}. We defined log (ambient \textit{B. pseudomallei} +1) as the dependent variable (Y) and the time period after the appearance of sediment at a nearby pond may be the source of ambient \textit{B. pseudomallei} during the typhoon season.

The association between melioidosis and rainfall intensity is well documented from endemic regions, with 75%, 72%, 80%, 81% and 85% of cases presenting during the wet season in northeast Thailand [6], India [3], Taiwan [14], northern Australia [10] and northern Australia [13], respectively. In regard to the associations between ambient \textit{B. pseudomallei} and environmental parameters, we found that rainfall was positively correlated to ambient \textit{B. pseudomallei} during June to December. This is consistent with the previous observation of significantly linear correlation between melioidosis cases and rainfall [5–6,13,15–16]. However, no significant correlation was observed between ambient \textit{B. pseudomallei} and rainfall during the typhoon season and typhoon month. This may be due to the appearance of sediment strongly affecting the concentration of ambient \textit{B. pseudomallei}. We defined August as “typhoon month” to investigate the association between ambient \textit{B. pseudomallei} and environmental parameters due to four typhoons affecting Taiwan in August, and less effect of sediment appearance in August when compared to July. We defined log (ambient \textit{B. pseudomallei} +1) as the dependent variable (Y). Rainfall, wind speed and typhoon were defined as independent variables. We found that typhoon and wind speed were significantly associated with ambient \textit{B. pseudomallei} with the predicted model as \( Y = 1.350 - 0.272 \) (wind speed) (\( p \)-value 0.039+0.713 (typhoon) (\( p \)-value 0.046) with a \( p \)-value of 0.049 for the whole model. We found that typhoon was positively correlated with ambient \textit{B. pseudomallei}. This supports conclusions from previous studies that the case clusters were associated with sudden and heavy rainfall related to cyclones and typhoons [1,4,6,10,13-18,22]. Wind speed was reversely correlated with ambient \textit{B. pseudomallei}. This may be due to dilution of ambient \textit{B. pseudomallei} by wind.

In conclusion, we successfully developed a filtration/real-time qPCR method to quantify ambient \textit{B. pseudomallei}. To our knowledge, this is the first report describing concentrations of ambient \textit{B. pseudomallei}. Our results provided evidence to support the hypothesis that heavy monsoonal rains and winds may cause a shift toward inhalation of \textit{B. pseudomallei}. Ambient \textit{B. pseudomallei} should be considered as an important source of transmission due to the wind speed during the typhoon season. Ambient \textit{B. pseudomallei} should be taken seriously as a surveillance target in endemic areas of Melioidosis especially during typhoon season.

**Author Contributions**

Conceived and designed the experiments: YLC YCY PSC. Performed the experiments: YLC YCY PSC. Analyzed the data: YLC YCY PSC. Contributed reagents/materials/analysis tools: MSL CKH KDM PYW. Wrote the paper: YLC YCY PSC.

**References**

1. Cheng AC, Currie BJ. (2003) Melioidosis: epidemiology, pathophysiology and management. Clin Microbiol Rev 16(3):416–416.
2. Currie BJ, Dance DA, Cheng AC. (2008) The global distribution of \textit{Burkholderia pseudomallei} and melioidosis: an update. Trans R Soc Trop Med Hyg 102 Suppl 1: S1–4.
3. Dance DA. (2000) Melioidosis as an emerging global problem. Acta Trop 74:113–9.
4. Ko WC, Cheung BM, Tang HJ, Shih HI, Lau YJ, et al. (2007) Melioidosis outbreak after typhoon, southern Taiwan. Emerg Infect Dis Jun;13(6):896–8.
5. Vidyakalakhis K, Lipika S, Vishal S, Damodar S, Chakrapani M. (2012) Emerging clinico-epidemiological trends in melioidosis: analysis of 95 cases from western coastal India. Int J Infect Dis 16(7):e91–7.
6. White NJ. (2003) Melioidosis. Lancet 361:1715–22.
7. Chinnery CJ, Mian M, Htun T, et al. (2003) Melioidosis. Am J Trop Med Hyg 75:947–954.
8. Stewart T, Engelhardler DM, Blaney DD, Tuanloy A, Wangness E, et al. (2011) Epidemiology and investigation of Melioidosis, Southern Arizona. Emerg Infect Dis 17(7):1286–1293.
9. Limamuthuotsakul D, Wonggratanacheewin S, Teerawattanaseek N, Wongguan G, Chaisukka S et al (2010) Increasing incidence of human melioidosis in Northeast Thailand. Am J Trop Med Hyg 2010 Jun;82(6):1113–7.
10. Currie BJ, Ward L, Cheng AC. (2010) The Epidemiology and Clinical Spectrum of Melioidosis: 540 Cases from the 20 Year Darwin Prospective Study. PLoS Negl Trop Dis 2010 Mar;4(1):e900.
11. Dai D, Chen YS, Chen PS, Chen YL. (2012) Case cluster shifting and contaminant source as determinants of melioidosis in Taiwan. Trop Med Int Health 17(3):1093–1093.
12. Wiersinga WJ, van der Poll T, White NJ, Day NP, Peacock SJ. (2006) Melioidosis: insights into the pathogenicity of Burkholderia pseudomallei. Nat Rev Microbiol 4(4):727–722.
13. Currie BJ, Jacups SP. (2003) Intensity of rainfall and severity of melioidosis, Australia. Emerg Infect Dis 9: 1538–1542.
14. Chou DW, Chung KM, Chen CH, Cheung BM. (2007) Bacteremic melioidosis in southern Taiwan: clinical characteristics and outcome. J Formos Med Assoc 106(12):1013–22.
15. Hassan MR, Pani SP, Peng NP, Vorahu K, Vijayalakshmi N, et al. (2010) Incidence, risk factors and clinical epidemiology of melioidosis: a complex socioecological emerging infectious disease in the Alor Setar region of Kedah, Malaysia. BMC Infect Dis 10:302.
16. Su HP, Chan TC, Chang CC. (2011) Typhoon-related leptospirosis and melioidosis, Taiwan. Emerg Infect Dis 17(7):1322–4.
17. Cheng AC, Jacups SP, Gal D, Mayo M, Currie BJ. (2006) Extreme weather events and environmental contamination are associated with case-clusters of melioidosis in the Northern Territory of Australia. Int J Epidemiol 35: 323–329.
18. Parameswaran U, Baird RW, Ward LM, Currie BJ. (2012) Melioidosis at Royal Darwin Hospital in the big 2009–2010 wet season: comparison with the preceding 20 years. Med J Aust 196(5):345–9.
19. Anuradha K, Meena AK, Lakshmi V. (2003) Isolation of Burkholderia pseudomallei from a case of septicaemia – a case report. Indian J Med Microbiol 21(2):129–32.
20. Limmathurotsakul D, Dance DA, Wuthiekanun V, Kaestli M, Mayo M, et al. (2013) Systematic review and consensus guidelines for environmental sampling of Burkholderia pseudomallei. PLoS Negl Trop Dis 7(3):e2105.
21. Lo TJ, Ang LW, James L, Goh KT. (2009) Melioidosis in a tropical city state, Singapore. Emerg Infect Dis 15: 1645–1647.
22. Su HP, Chou CY, Tseng SC, Ferg TL, Chen YL, et al. (2007) Possible Typhoon-related melioidosis epidemic, Taiwan, 2005. Emerg Infect Dis 13(11):1795–7.
23. Tan GY, Liu Y, Sivalingam SP, Sim SH, Wang D, et al. (2008) Burkholderia pseudomallei aerosol infection results in differential inflammatory responses in BALB/c and C57Bl/6 mice. J Med Microbiol 57(Pt 4):508–15.
24. Warawa JM. (2010) Evaluation of surrogate animal models of melioidosis. Front Microbiol 1:141.
25. Chou PS, Li CS. (2005) Quantification of airborne Mycobacterium tuberculosis in health care setting using real-time qPCR coupled to an air-sampling filter method. Aerosol Science and Technology 39 (4): 371–376.
26. Chen P S, Li CS. (2008) Concentration Profiles of Airborne Mycobacterium tuberculosis in a Hospital. Aerosol Science and Technology 42 (5): 194–200.
27. Chen PS, Lin CK, Tsai FT, Yang CY, Lee CH, et al. (2009) Quantification of Airborne Influenza and Avian Influenza Virus in a Wet Poultry Market using a Filter/Real-time qPCR Method. Aerosol Science and Technology 43: 290–297.