High stathmin expression is a marker for poor clinical outcome in endometrial cancer: An NRG oncology group/gynecologic oncology group study

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Objective. Gynecologic Oncology Group (GOG) 177 demonstrated that addition of paclitaxel to a backbone of adriamycin/cisplatin improves overall survival (OS) and progression-free survival (PFS) for patients with advanced or recurrent endometrial cancer. Using patient specimens from GOG-177, our objective was to identify potential mechanisms underlying the improved clinical response to taxanes. Stathmin (STMN1) is a recognized poor prognostic marker in endometrial cancer that functions as a microtubule depolymerizing protein, allowing paclitaxel to negate the effects of stathmin overexpression.

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ABSTRACT

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1. Introduction

Endometrial cancer is the most common gynecologic malignancy in women [1]. Early stage disease can be treated surgically, with some cases receiving radiation treatment to decrease local and regional recurrence. However, for recurrent disease or cases diagnosed at an advanced stage, chemotherapy plays a crucial role in management.

GOG-177 was a phase III randomized controlled trial that enrolled patients with advanced (stage III or IV) or recurrent endometrial cancer between two treatment arms: adriamycin (doxorubicin)/cisplatin (AP) vs. adriamycin/cisplatin/paclitaxel with G-CSF (TAP). Results from this trial showed that patients treated with the three-drug regimen TAP had a better overall response rate (57% vs 34%), median progression-free survival (PFS) (8.3 vs. 5.3 months), and median overall survival (OS) (15.3% vs 12.3%) [1]. The TAP regimen, however, was associated with a 27% and 12% incidence of grade 2 and 3 peripheral neuropathy, respectively [2]. Following this trial, the GOG performed a phase III non-inferiority trial (GOG-209), which evaluated the differences in survival and toxicity between the triple regimen (TAP) vs. paclitaxel/carboplatin (TC). The interim analysis showed that high stathmin correlated with poor PFS and OS particularly in patients who received adriamycin/cisplatin only. In contrast, there was no statistically significant association between stathmin expression and OS or PFS in patients treated with paclitaxel/adriamycin/cisplatin.

Conclusions. Our findings demonstrate that high stathmin expression is a poor prognostic marker in endometrial cancer. Paclitaxel may help to negate the impact of stathmin overexpression when treating high risk endometrial cancer cases.

2. Materials and methods

2.1. Human subjects

Formalin-fixed, paraffin-embedded (FFPE) tissue slides were received from patients who were enrolled on the GOG-177 protocol. Out of the 263 eligible patients in the protocol, specimens were available for 86 patients. A total of 69 patients had adequate quantity of available slides to evaluate for IHC. Among the 4 patients alive at last contact, the minimum follow-up was 5 years; the others were followed between 8 and 9 years. All studies were approved by the University of Iowa Institutional Review Board (Approval #200907769: GOG Core Laboratory).

2.2. Immunohistochemistry

IHC was performed on slides from GOG-177. FFPE tumor tissue sections from GOG-177 were evaluated for expression of stathmin (#3352, Cell Signaling, Danvers, MA), via IHC per the manufacturer’s recommendations and as shown by other investigators [5,13]. A western blot analysis using 8 different cell lines and IHC utilizing positive and negative control slides were performed to confirm specificity of the antibody. Staining was independently quantified by three investigators blinded to sample identity using a modified H-scoring system that is calculated by multiplying the percentage of tissue with positive staining (0–100%) by the intensity of staining (0–4) as previously described [14].

2.3. The Cancer Genome Atlas (TCGA)

Using 333 identified endometrial cancer patients from TCGA database (271 endometrioid and 62 serous), we examined the relationship of STMN1 (the gene that codes for the protein stathmin) gene expression with OS and PFS. Of the 333 patients, 102 received chemotherapy (N = 55 paclitaxel-containing chemotherapy; N = 52 no paclitaxel). The remaining 229 patients did not receive chemotherapy. The mean follow-up time was 33.8 months. Dataset extraction of clinicopathologic parameters (age, weight, tumor grade, invasion, histology, stage, residual tumor, lymph node status, total pelvic, and total aortic lymph nodes), risk group stratification, and STMN1 gene expression were performed.

“High risk” patients were defined as those at risk of having extra-uterine disease and most likely needing adjuvant treatment after surgery. Specifically, all patients presenting with stage II, III and IV as defined by 2009 FIGO classification (and sanctioned in 2014) [15], and patients with initial stage I and high-intermediate risk features by GOG 99 criteria [16] were classified as high risk. High-intermediate features of stage I included three risks factors: 2 or 3 tumor grade, presence of lymphovascular invasion, and outer-third myometrial invasion, with the following criteria: 1) at least 70 years of age with only one of the risk factors, 2) at least 50 years of age with any two of the other risk factors, or 3) any age with all three of the other risk factors. “Low risk” patients were the remaining stage I patients, either with no myometrial invasion or low-intermediate risk features by GOG 99 criteria [16]. STMN1 gene expression was divided into quartiles and data analyzed as ≤50% vs. >50% or ≤75% vs. >75%.

2.4. Statistical analysis

Protein expression of stathmin was evaluated by IHC in 69 patients from GOG-177 and dichotomized using modified H-score results of
>50, >75, or >100 as the cut-off. Mean age was compared via a two-sided Wilcoxon rank sum test. Tumor grade, tumor stage (as defined by 1988 FIGO classification [17]), and tumor type were compared via a Chi-squared test. Serous vs. non-serous and endometrioid vs. non-endometrioid types were compared via two-sided Fisher's tests. For TCGA data, multi- and univariate analyses were performed via Cox's proportional hazards ratio. *STMN1* gene expression was dichotomized (High or Low) using either the 50th percentile or 75th percentile of the expression as the cut-off. The associations of *STMN1* expression with PFS and OS were evaluated in patients with endometrial cancer, adjusted for all clinicopathologic parameters of interest, using the Cox proportional hazard ratio, and Kaplan-Meier survival curves were plotted when significant. PFS and OS analysis was also further stratified according to histologic type (serous or endometrioid). Significance was assessed using log rank tests. For all statistical tests, a level of <0.05 was considered statistically significant.

3. Results

3.1. Stathmin overexpression predicts for poorer overall survival and progression-free survival

We first performed a retrospective analysis of FFPE tumors from 69 patients enrolled on GOG-177. Patient characteristics are presented in Table 1. Representative images of stathmin expression by IHC are provided in Fig. 1. Stathmin expression was significantly associated with PFS and OS (Fig. 2A, B). However, stathmin was significantly associated with PFS and OS in arm 1 AP (Fig. 2C, D) but not in arm 2 TAP (Fig. 2E, F).

Table 1  
Demographics and clinicopathological features of patients on GOG177 and association with stathmin expression. Stathmin was dichotomized into low (≤50 H-score) or high (>50 H-score) by IHC (N = 69). Mean age was compared via a two-sided Wilcoxon rank sum test. Tumor grade, tumor stage, and tumor type compared via a Chi-squared test, except for serous vs. non-serous and endometrioid vs. non-endometrioid that were compared via two-sided Fisher's tests. Overall survival and progression free survival were compared with a log rank test.

| Characteristic       | Low stathmin (n = 62) | High stathmin (n = 7) | Association with stathmin (P-value) |
|----------------------|-----------------------|-----------------------|------------------------------------|
| Age at study entry (mean, SD) | 62.45, 10.25          | 60.83, 6.63           | P = 0.5183                          |
| Tumor grade (counts) | 1                     | 14                    | 0                                  |
|                      | 2                     | 21                    | 5                                  |
|                      | 3                     | 25                    | 2                                  |
|                      | N/A                   | 2                     | 0                                  |
| Tumor stage (counts) | 3                     | 6                     | 0                                  |
|                      | 3C                    | 3                     | 0                                  |
|                      | 4                     | 14                    | 1                                  |
|                      | 4B                    | 6                     | 1                                  |
| Recurrent            | 33                    | 5                     |                                    |
| Tumor type (counts)  | Adenocarcinoma        | 1                     | 0                                  |
|                      | Clear cell            | 3                     | 0                                  |
|                      | Endometrioid          | 46                    | 6                                  |
|                      | Mixed epithelial      | 5                     | 0                                  |
|                      | Undifferentiated      | 1                     | 0                                  |
|                      | Serous                | 6                     | 1                                  |
| Tumor type (counts)  | Non-serous            | 56                    | 6                                  |
|                      | Serous                | 6                     | 1                                  |
| Tumor type (counts)  | Endometrioid          | 46                    | 6                                  |
|                      | Non-endometrioid      | 16                    | 1                                  |
| Median PFS (months)  | 8.07                  | 2.79                  | P = 0.0470                         |
| Median OS (months)   | 15.10                 | 10.28                 | P = 0.0096                         |

Fig. 1. Representative images of stathmin staining by IHC. High modified H-score (A) and low modified H-score (B). Images were acquired at 200× magnification.

3.2. Relationship between stathmin expression and histologic subtype

We next examined whether there was a relationship between stathmin expression and histologic type in GOG-177 samples. However, there was no significant association (P = 0.5442) observed when tumors were dichotomized into serous and non-serous types nor when tumors were dichotomized into endometrioid and non-endometrioid types (P = 0.6724, Table 1). Similarly, stathmin expression was not associated with age at study entry, tumor grade, or stage. Nevertheless, we extended this analysis to a larger dataset from TCGA.

3.3. Elevated expression of stathmin portends poor outcomes in TCGA dataset

Using RNA-seq data from all 333 endometrial cancer patients in TCGA dataset for endometrial cancer [18], we confirmed that high stathmin mRNA levels are associated with shorter PFS in both multi- and univariate analysis (Fig. 3A), even after adjusting for tumor grade, myometrial invasion, FIGO stage, lymph node status, and residual tumor after surgery. Specifically, stathmin mRNA expression at >50th percentile significantly correlated with poorer 5-year PFS of 62.5% (P = 0.0485) as compared to cases with low expression (5-year PFS of 82.5%). Stathmin expression at the >75th percentile also was
associated with poorer OS of 75% vs. 89% for patients with stathmin mRNA levels \( \geq \) 75th percentile by univariate analysis (Fig. 3B). In the multivariate analysis, only FIGO stage and myometrial invasion, but not stathmin expression, were independently associated with OS (\( P < 0.01 \)).

We next stratified patients in TCGA dataset by histologic type and found that high stathmin mRNA levels were associated with worse PFS in serous tumors by multivariate analysis (Fig. 4A). Specifically, stathmin expression \( \geq \) 75th percentile in the serous group showed significantly worse 5-year PFS of 22% compared to those with low stathmin expression (5-year PFS of 67%). Stathmin expression was not significantly associated with PFS in the endometrioid group (\( P = 0.0811 \)), yet we did detect a similar trend for stathmin expression \( \geq \) 50th percentile linked to worse PFS, with a 5-year PFS of 70% for those with high expression and 85% for those with low expression (Fig. 4B).

4. Discussion

Stathmin expression at the mRNA and protein levels has been attributed to poor prognosis in a variety of solid tumors. In cancer cells, the overall function of elevated stathmin is to speed cell progression into and out of mitosis [4]. High stathmin has been reported to underlie resistance to paclitaxel by directly antagonizing paclitaxel’s stabilizing effect on the microtubule. Using tumor specimens from patients treated on GOG-177, we confirmed that high expression of stathmin correlated with worse OS and PFS. However, the impact of high stathmin was most significant in patients on arm 1, TA. The addition of paclitaxel in arm 2 (TAP) resulted in a significant overall improvement in PFS and OS, highlighting the important role of paclitaxel in the treatment of patients with advanced endometrial cancer. Importantly, even high stathmin expression did not negate the improved outcomes with the addition of paclitaxel.

Stathmin was first identified based on its overexpression in leukemia cell lines. It is now widely recognized to be overexpressed in nearly all solid tumors examined to date, including hepatocellular, gastric, breast, prostate, cervical, pancreatic, and ovarian cancers. Moreover, stathmin expression is generally considered to be a marker for an aggressive tumor based on the correlation between stathmin levels and OS.
tumor size, metastatic potential, histopathologic grade and overall survival in multiple cancer types [19–29].

Elevated expression of stathmin has also been documented in endometrial tumors [5,16,30–34]. For example, an integrated analysis of genome-wide expression data for primary endometrial carcinoma found that \textit{STMN1} is upregulated at the mRNA level [30]. In addition, qRT-PCR demonstrated 2.5-fold higher \textit{STMN1} expression in tumor vs. normal endometrium [31]. Our analysis of TCGA dataset using a \textit{STMN1} cut-off of 50% suggests that approximately half of all endometrial tumors have elevated stathmin transcripts. In our IHC study of stathmin protein expression (GOG-177 samples), we found that 75% of tumors (53 out of 69) had no detectable stathmin expression by IHC, and only 7 tumors (10%) had an H-score > 50. Similarly, a study of metastatic endometrial cancer in Norway demonstrated that only 18% of primary tumors had high stathmin levels [5]. By contrast, using a cut-off of > 10% cells staining positive for stathmin, others have reported that 60.7% of endometrial tumors express stathmin protein, whereas 36.7% of normal endometrium samples have positive staining [31]. Nevertheless, stathmin expression, while relatively uncommon overall in GOG-177, appears to identify some of the highest risk cases from this study based upon the poor prognosis associated with its presence. The high proportion of tumors with no detectable stathmin protein in the GOG specimens cannot be explained by the difference in scoring systems, since such a large number of tumors were negative by IHC in our study.

Studies in lung cancer suggest that stathmin expression is significantly increased in poorly differentiated lung adenocarcinoma as compared to moderately or well differentiated tumors [35]. Similarly, stathmin expression has been associated with more aggressive phenotypes, including higher expression in late vs. early stage endometrial cancer and in lymph node metastases and increased invasion into the myometrium and lymphovascular space [30,31,34,36]. However, we did not observe a significant correlation between stathmin expression and age, pathological type, or grade in GOG-177 subjects or in TCGA dataset. There was a pronounced association between stathmin expression and serous subtype of endometrial cancer in TCGA dataset, with a large difference in PFS associated with low stathmin in this cohort of serous cases. This observation of higher stathmin in serous cases was not replicated in the GOG-177 dataset, possibly because of the relatively small number of serous and stathmin positive cases.

The significant associations between stathmin mRNA levels and PFS/OS are consistent with several studies in multiple cancer types documenting a link between high stathmin expression and poor response to treatment [5,20–22]. Consistent with a previous report [34], we found a significant inverse relationship between stathmin overexpression and OS as well as PFS in both the GOG-177 specimens as well as TCGA dataset for endometrial cancer. One of the most interesting findings of our study is that the addition of paclitaxel negated the impact of stathmin overexpression on PFS and OS, consistent with a study of ovarian clear cell adenocarcinoma patients that received taxane-containing or taxane-free therapy [37]. However, the precise role of stathmin as a marker for paclitaxel sensitivity or resistance is not clear. For example, a Norwegian study of 38 patients with metastatic...
endometrial cancer treated with paclitaxel-containing chemotherapy reported a significantly lower disease-specific survival in patients with high stathmin [5]. Similarly, stathmin has been associated with resistance to antimicrotubule drugs including paclitaxel in breast cancer [3, 6,7], non-small cell lung cancer [8,9], epithelial ovarian cancer [10], and bladder cancer [11].

At the molecular level, stathmin destabilizes microtubules to promote cell division. Conversely, paclitaxel promotes microtubule polymerization, leading to mitotic arrest and ultimately cell death [12]. We propose that the transition through mitosis in the setting of paclitaxel therapy for cancer is a balance between the microtubule depolymerizing activity of stathmin and the microtubule stabilizing activity of paclitaxel. Our translational analyses herein suggest that paclitaxel negates the deleterious effects of stathmin overexpression.

**Authors’ contributions**

H.D.R and K.K.L. conceptualized the hypotheses and designed the research strategy; P.H., S.G., K.S.T., F.B., N.R., G.F.F., V.F., and M.J.B. accrued patients for the study; H.D.R, E.J.D., Y.Z., J.-M.S., and M.M. performed the experiments; J. M., and J. G.-B. performed the statistical analysis; H.D.R., J. M., K.W.T., and K.K.L. analyzed data; and H.D.R., K.W.T., E.J.D., and K.K.L. wrote the paper. All authors read and approved the final manuscript.

**Conflicts of interest**

Dr. Jeffrey Miecznikowski receives grant funding from NRG Oncology Statistical and Data Management Center. Dr. Kristina Thiel is a co-
founder of Immortagen and holds an equity stake in the company. Dr. Krishnaswami Tewari receives payment for lectures, including service on speakers bureaus, from Roche, AstraZeneca and Merck. He also receives travel/accommodations/meeting expenses unrelated to activities listed from Roche. Dr. Floor Backes serves as consultant for Advaxis on clinical trial steering committee. She also has grants/grants pending from Eisai. Dr. Nilsa Ramirez receives grant funding from GOG Tissue Bank/NGO Oncology Biospecimen Bank-Columbus. Dr. Ramirez also has grants/grants pending through the COG/NGO Oncology Biospecimen Bank support via NCI awarded U24 and U10 grants. Dr. Kimberly Leslie receives grant funding through the NIH – R01CA99908. All other co-authors have no conflicts of interest to declare.

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