Effect of Genistein Supplementation on the Progression of Neoplasms and the Level of the Modified Nucleosides in Rats With Mammary Cancer

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Abstract. Background/Aim: The aim of the study was to assess the impact of nano-, micro-, and macro-sized genistein on the growth and development of neoplasms in rats with mammary cancer. Additionally, the effect on the kinetics of changes (9-11-17-20 week of a rat’s life) in the levels of methyl derivatives: 1-methyladenine, 3-methyladenine, 7-methylguanine, 1-methylguanine, 1-methyladenosine, 7-methylguanosine, O-methyl-guanosine and N6-methyl-2’-deoxyguanosine in the urine of rats was analyzed. Materials and Methods: Female Sprague-Dawley rats divided into 4 groups were used in the study. Animals were fed only a control diet or diets supplemented with the nano-, micro- and macro-sized genistein. To induce the mammary adenocarcinoma, rats were treated with 7,12-dimethylbenz[a]anthracene (DMBA). Modified nucleosides were determined by a high-performance liquid chromatography coupled to mass spectrometry method (LC-MS/MS). Results: The supplementation of the diet of animals with genistein resulted in an increase in the excretion of methylated derivatives in the urine of rats. In the animals receiving standard diet, the levels of methyl derivatives increased during the study or remained relatively low. In the case of animals whose diet was supplemented with the various forms of genistein, the levels of methylated derivatives were very high from the beginning. Conclusion: High levels of methyl derivatives can influence carcinogenesis.

Genistein (4’,5,7-trihydroxyisoflavone or 5,7-dihydroxy-3-[4-hydroxyphenyl] chromen-4-one) (C15H10O5) belongs to a multifunctional natural isoflavonoid class of flavonoids with a 15-carbon skeleton. It was isolated for the first time from Genista tinctoria L. in 1899 and named after the genus of this plant (1). Since then, it has been considered the main secondary metabolite of the Trifolium species and in Glycine max L. (synonym Soja hispida) (1). Numerous clinical studies have shown chemopreventive and therapeutic effects of genistein on various types of cancer (2). Genistein has cytostatic and cytotoxic properties on both healthy and cancerous cells. The mechanisms of these actions are associated with inhibition of the activity of tyrosine kinases transmitting cell growth signals and topoisomerase II, a protein that is responsible for DNA stability (3). In this way, genistein induces apoptosis. In addition, it has been shown to cause cell cycle arrest, antiangiogenic, antimitastic, and anti-inflammatory effects (4). These properties are beneficial in the treatment of cancer (4). However, this action can be harmful to healthy cells. By causing DNA strand breaks in genes, it may cause mutations. Based on in vitro and in vivo tests, it has been found that genistein may also accelerate the growth of certain types of cancer, in particular those that are hormone-dependent, and reduce their sensitivity to tamoxifen and letrozole. Genistein is a phytosterol with a structure similar to 17β-estradiol (E2), which is the main estrogen in women. Thus, phytoestrogens may affect the reproductive and endocrine systems leading to the development of some estrogen-related cancers. These types
of cancers include cervical cancer, breast cancer or ovarian cancer (5-8). In addition, studies have shown that high levels of flavonoids in the diet of pregnant women are associated with an increased risk of acute myeloid leukemia (AML) in infants, representing about 15% of the total infant incidence of leukemia in USA (9).

It has been suggested for many years that modified nucleosides may be tumor markers, as their levels are often elevated in patients with oncogenic disease (10-13). It has been shown that they contribute along with genetic mutations to cancer development and progression. It was found that increased gene expression may be caused by increased methylation of N-7 guanine (7-MeG) in methyl-CpG pairs, which may result in a change in protein/DNA interactions and chromatin remodeling (14). Moreover, the majority of lesions created by alkylating agents, such as 3-methyladenine (3-MeA), strongly block replication (15-17). Genistein might modulate the DNA methylation status and expression of cancer-related genes in breast cancer cells (18, 19). However, the precise mechanisms underlying the effect of genistein on breast cancer development, in particular the epigenetic mechanisms, remain unclear. Therefore, further studies are needed to determine the beneficial and harmful effects of genistein, as well as to determine the mechanism of action of genistein, the safe therapeutic doses, and the administration form.

The aim of the present research was to assess the impact of nano-, micro- and macro-sized-genistein on the growth and development of neoplasms in rats with mammary cancer (adenocarcinoma) induced with 7,12-dimethylbenz[a]anthracene. Additionally, we aimed to examine the effect on the kinetics of changes (9-11-17-20 week of a rat’s life) in the levels of the methyl derivatives: 1-methyladenine, 3-methyladenine, 7-methylguanine, 1-methylguanine, 1-methyladenosine, 7-methylguanosine, O-methyl-guanosine, N6-methyl-2’-deoxyguanosine in the urine of rats with mammary cancer (adenocarcinoma) induced with 7,12-dimethylbenz[a]anthracene. The studies of the impact of dietary components on the growth and development of neoplasms and on selected biomarkers can be very important in both cancer prevention and treatment.

Materials and Methods

Preparation of genistein micro- and nanoparticles. PVA (n=approx. 2,000, degree of saponification ca. 80 mol%) and genistein (purity >98.0%) were purchased from Tokyo Chemical Industries (Portland, OR, USA). Ethyl acetate (purity 99.8%) was purchased from Avantor Performance Materials Poland S.A (Gliwice, Poland).

SciInt JY-92 – IIDN 900 Watt (Staufen, Germany) ultrasonic homogenizer with 6 mm titanium tip was used for homogenization. Homogenization parameters were as follows: 15 min homogenization time, 4.5 s pulse, 0.5 s pulse interval, 75% power.

To a strongly stirred (5,488 RCF) 2.5% PVA solution (400 ml, saturated with ethyl acetate), concentrated solution of genistein (1,00 g) in ethyl acetate was added dropwise. Stirring was continued for 5 min after addition. Opaque solution was ho-mogenized for 15 min and slowly stirred (900 rpm) with gentle heating at 40°C. Homogenization was repeated every 24 h until complete removal of ethyl acetate. The mixture was diluted 3x with distilled water and centrifuged (15 min, 4,500 rpm). Resulting nanoparticles were repeatedly washed with distilled water and centrifuged until removal of excess PVA, and dried at room temperature in vacuum for 24 h. The above procedure yielded 1.02 g of nanoparticles. Micronized genistein was prepared by milling in SPEX Sample Prep 6770 Freezer/Mill cryogenic mill (Retsch Polska Verder Polska Sp. z o.o., Katowice, Poland) (coolant: liquid nitrogen, 3 cycles, precool time: 1 min, run time: 2 min, cool time: 1 min, rate: 15 CPS). Macro-size genistein was used as received.

To examine the average size and zeta potential of the particles the dynamic light scattering technique (DLS) was applied. The Zetasizer Nano ZS instrument (Malvern Instruments, Westborough, MA, USA) equipped with a red laser with a wavelength of 633 nm and a scattering angle of 173° at 25°C was utilized for this purpose. Sample was suspended in either distilled water (Zeta potential measurement) or glycerol water solution (size measurement) (Table I, Figures 1 and 2).

Laboratory animals. Female Sprague-Dawley rats (n=32) were obtained from the Animal Laboratory, Department of General and Experimental Pathology, Medical University of Warsaw. The study was approved by the Ethics Committee. All rats were provided Labelled H standard diet (standard diet: Labofeed H, Żurawia 19, 89-240 Kcynia, Poland) and water ad libitum and housed in an environmentally controlled room at 22°C with a 12-h light-dark cycle. The rats were divided into four experimental groups: control animals were fed only standard diet (without supplementation) (received 0.4 ml water), nanosized genistein (0.1 mg/ml; i.e., 0.2 mg/kg bw) (92±41 nm), microsized genistein (0.1 mg/ml; i.e. 0.2 mg/kg bw) (587±83) and macrosized genistein (0.1 mg/ml; i.e. 0.2 mg/kg bw) supplemented groups. The rats were fed extra supplements suspended in water (0.4 ml daily via gavage), from 40 days until 20 weeks of age. The polyphenols dose level was selected based on human average daily consumption (extrapolating on the rats’ body weight). Determining the effect of a selected dose of genistein may allow its application in human cancer prevention or in the improvement of pharmacological treatment.

The rats were treated twice with DMBA (7,12-dimethyl-1,2-benz(a)anthracene; Sigma-Aldrich, St. Louis, MO, USA) in rapeseed oil (via gavage) to induce mammary cancer (adenocarcinoma); the first treatment was given at 60 days of age (80 mg/kg body weight); followed by a repeated dose of 40 mg/kg body weight at 90 days of rat age.

Table I. Dynamic light scattering (DLS) results for genistein particles.

| Parameter          | Size (d.nm)a | Z-average (d.nm) | Zeta potentialb [mV] | Dc |
|--------------------|--------------|------------------|----------------------|----|
| Genistein nanoparticles | 92±41       | 158              | −17.2±5.5            | 1,000 |
| Genistein microparticles | 587±83     | 1467             | −30.2±6.0            | 0,873 |

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aSize±standard deviation; bZeta potential±standard deviation; cdispersity.
The animals were examined by palpation during the study to characterize the time course of tumor development. In order to obtain urine samples, each animal was individually placed in a metabolic cage for 24 h. Urine samples were collected once a week at the 9th, 11th, 17th, 20th week of rodent’s age and stored at –70°C until the test time.

Histopathology. The rats were sacrificed by decapitation at 150 days of age, and tumors were evaluated histopathologically. The tissues were placed in a buffered formalin solution, dehydrated, sealed in paraffin and cut into 4 μm thick sections. Hematoxylin and eosin staining of tissue and cell sections was applied, and the sections were evaluated using a BX43 Olympus research microscope (Olympus Europa SE & Co., Hamburg, Germany).

LC-MS/MS analysis. Reference standards i.e., 1-methyladenine, 3-methyladenine, 7-methylguanine, 1-methylguanine, 7-methylguanosine, 7-methylguanosine, O-methylguanosine, N6-methyl-2’-deoxyguanosine as well as internal standard (tubercidin) were purchased from Sigma Aldrich. Modified nucleosides and nucleobases were determined by validated high performance liquid chromatography coupled to mass spectrometry (LC-MS/MS) method using multiple reaction monitoring (MRM) mode on Agilent 1260 Infinity (Agilent Technologies, Santa Clara, CA, USA) coupled to QTRAP 4000 (AB Sciei, Framingham, MA, USA). MRM transitions, declustering potential (DP), and collision energy (CE) for O-methylguanosine, 1-methyladenosine, 7-methylguanosine, 7-methylguanine, 3-methyladenine, 1-methylguanine and N6-methyl-2’-deoxyadenosine were: (m/z) 298>152 (DP=51 V, CE=17 V), 282>55 (DP=66 V, CE=87 V), 298>166 (DP=71 V, CE=19 V), 166>79 (DP=96 V, CE=43 V), 150>123 (DP=86 V, CE=51 V), 166>135 (DP=81 V, CE=31 V) and 266>150 (DP=61 V, CE=23 V), respectively. Chromatographic separation was achieved using SeQuant® ZIC®-HILIC (50×2.1 mm, 5 μm, Merck KGaA, Darmstadt, Germany) column. The column was maintained at 25°C at the flow rate of 0.5 ml/min. The mobile phases consisted of 20 mM ammonium acetate as the eluent A and acetonitrile with 0.2% formic acid as the eluent B. The gradient (%B) was as follows: 0 min, 95%; 1 min, 95%; 7 min, 50%; 8 min, 50%. The injection volume was 5 μl.

Urine samples (0.1 ml) prior to injection to LC were mixed with tubercidin (0.1 ml, 1 μg/ml) and acetonitrile (0.6 ml), vortexed in high speed (3 min) and centrifuged (5 min at 10,000xg).

The levels of the modified nucleosides and bases in urine were standardized by conversion to the creatinine level. The latter was determined in urine samples with a creatinine test (Hydrex, Warsaw, Poland) based on Jaffe’s reaction.

Statistical analyses. Statistical analyses were performed using the PQStat statistical package, version 1.8.0.324 (BioInforStats, Cracow, Poland). For normally distributed data, Student’s test and ANOVA were used. The results obtained in each group were compared using the analysis of variance and the post-hoc Tukey test. The test probability at the level of p<0.05 was assumed as
Results

Tumor size assessment. Histopathological examination revealed that chemically induced DMBA tumors were breast adenoma. Additionally, the time at which the first tumors appeared, the number and size of tumors and the incidence of cancer in rats were analyzed. The results of tumor induction in 7,12-dimethylbenz[a]anthracene treated groups in relation to supplementation are shown in Tables II and III. In the case of rats that were not supplemented and received a standard diet, the incidence of cancer was 100%, the weight of tumors was in the range 0.1-7.8 g (0.93±1.34) and the number of tumors per rat ranged from 2 to 9 (Table II). Similar results were obtained in the case of rats supplemented with macrosized genistein [incidence 100%, tumors weight 1.27±1.52 g (0.14-6.39 g), number of tumors per rat 1-6]. In contrast, in case of animals supplemented with microgenistein, although the first tumors appeared at week 17, a week later compared to animals receiving only the standard diet, the incidence of tumors was 88%, the number of tumors per rat ranged from 0 to 3; there was a statistically significant stimulation of tumor growth. The mean mass of the tumors in the animals receiving microgenistein was statistically significantly higher compared to tumors of animals receiving standard diet, 1.99±1.75 vs. 0.93±1.34. In the case of animals supplemented with nanogenistein, the first tumors appeared as early as at 14 weeks of animals’ age, 2 weeks earlier than in animals receiving only the standard diet, and 3 weeks earlier than in animals receiving macro and microgenistein. In the case of animals supplemented with nanogenistein, the incidence was 100%, the number of tumors per rat was 2-5 and the weight of tumors at the end of the experiment was 1.59±2.64 g (range=0.06-9.50 g). Histopathological examination showed that all examined tumors had features of breast cancer. In animals receiving only a standard, non-supplemented diet and animals supplemented with macrogenistein, grade II adenoma was found (Figure 3, Table II). Interestingly, in samples obtained from animals supplemented with microgenistein and nanogenistein, the adenoma image indicated grade III malignancy. In the case of animals supplemented with genistein massive (increased) tumor cell proliferation was observed. The tendency of neoplastic cells to proliferate was evidenced by the number of

| Table II. Tumour induction in 7,12-dimethylbenz[a]anthracene treated groups in relation to supplementation. |
|-------------------------------------------------|-------------------------------------------------|---------------------------------|---------------------------------|--|---------------------------------|
| Supplementation                                | The week when first tumor occurred | Number of tumors per animal (week 20) | Tumor incidence (%) (week 20) | Tumor weight (g) (week 20) mean±SD | Tumor grade       | The mean number of mitoses in the field of view area* |
| Standard                                       | 16 (2/8)                             | (2-9)                                          | 100% (8/8)                        | 0.93±1.34 (0.10-7.80)        | Adenocarcinoma 2 grade | 1.79±1.25a,b,c       |
| Macrogenistein                                 | 17 (5/8)                             | (1-6)                                          | 100% (8/8)                        | 1.27±1.52 (0.14-6.39)        | Adenocarcinoma 2 grade | 4.46±2.38a,b,c       |
| Microgenistein                                 | 17 (1/8)                             | (0-3)                                          | 88% (7/8)                         | 1.99±1.75 (0.11-6.11)        | Adenocarcinoma 3 grade | 7.33±1.57b,c        |
| Nanogenistein                                  | 14 (1/8)                             | (2-5)                                          | 100% (8/8)                        | 1.59±2.64 (0.06-9.50)        | Adenocarcinoma 3 grade | 5.82±1.57c          |
| Data are expressed as mean±SD. Values sharing letters (a: standard, b: macro genistein, c: micro genistein, d: nano genistein) indicate statistically significant differences between groups (p<0.01). *Mitoses were counted in slides from randomly selected tumors in 15 fields of view with a 40x objective magnification. SD: Standard deviation. |

| Table III. Tumor incidence (weeks 14-20) (%) (number of animals that developed tumors). |
|-------------------------------------------------|-------------------------------------------------|---------------------------------|---------------------------------|--|---------------------------------|
| Supplementation                                | Week 14                              | Week 15                          | Week 16                          | Week 17                          | Week 18                          | Week 19                          | Week 20                          |
| Standard                                       | 0/8                                  | 0/8                              | 2/8 (25%)                         | 5/8 (63%)                         | 6/8 (75%)                         | 7/8 (88%)                         | 8/8 (100%)                       |
| Macrogenistein                                 | 0/8                                  | 0/8                              | 0/8                              | 5/8 (63%)                         | 6/8 (75%)                         | 8/8 (100%)                         | 8/8 (100%)                       |
| Microgenistein                                 | 0/8                                  | 0/8                              | 0/8                              | 1/8 (13%)                         | 3/8 (38%)                         | 4/8 (50%)                         | 7/8 (88%)                         |
| Nanogenistein                                  | 1/8 (13%)                           | 1/8 (13%)                        | 1/8 (13%)                        | 2/8 (25%)                         | 4/8 (50%)                         | 6/8 (75%)                         | 8/8 (100%)                       |
mitoses in the field of view of the microscope (at 40× lens magnification). The mean number of mitoses in the field of view (at 40× magnification of the microscope objective) in the group of rats fed with the diet without supplementation was 1.79±1.25. In the group of rats fed with the diet supplemented with nanogenistein, microgenistein and macrogenistein the mean number of mitoses was 5.82±1.57, 7.33±1.57, and 4.46±2.38, respectively.

Weight gain of rats. No statistically significant differences were observed in the weight gain of rats during the study period (Table IV, Figure 4).

| Group   | Standard | Macro     | Micro     | Nano     |
|---------|----------|-----------|-----------|----------|
| Arithmetic mean | 99.0000  | 99.1250   | 107.4000  | 96.5000  |
| Standard deviation | 10.9414  | 8.8227    | 8.5370    | 8.7178   |
| Standard error of the mean | 3.8684   | 3.1193    | 3.0183    | 3.0822   |
| ANOVA | F=2.0860 , p=0.1247 |

Figure 3. Hematoxylin-eosin-stained sections of breast tumors. A) rats fed with a standard diet - no supplementation (black arrows show the tubular system of tumor cells); B) rats fed with nano-genistein (red arrow indicates infiltrating tumor cell); C) rats fed with a diet supplemented with micro-genistein (red arrows indicate mitosis and black bands indicate tumor cells); D) rats fed with a diet supplemented with macro-genistein (red arrows show mitosis and black arrows show cancer cells forming vesicles).

Figure 4. Weight gain of rats week 20 (g).
Organs’ weight. No statistically significant differences were observed in rat liver and spleen weights (Table V, Figures 5 and 6).

Kinetic evaluation of methyl derivatives. In this study, the effect of nano-, micro- and macro-sized-genistein on the kinetics (9th, 11th, 17th, 20th week of a rat’s life) of change in the levels of the methyl derivatives: 1-methyladenine, 3-methyladenine, 7-methylguanine, 1-methylguanine 1-methyladenosine, 7-methylguanosine, O-methyl-guanosine, N6-methyl-2'-deoxyguanosine in urine of rats with mammary cancer (adenocarcinoma) induced with 7,12-dimethylbenz[a]anthracene, were analyzed (Figures 7, 8, 9, 10, 11, 12, 13 and 14).

The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the level of O-methylguanosine (ng/mg creatinine) in the urine of rats treated DMBA. Highly significant (p<0.0001) differences between the groups and between measurement dates (p=0.0043) were found. In the case of rats, which received a standard diet, the concentration of O-methylguanosine in the urine increased significantly (p=0.0039) over time (week 9→20) as the cancer develops. However, in the case of animals supplemented with genistein, the levels of methylated derivatives were very high from the beginning and only in the case of microgenistein supplementation it decreased statistically significantly (p=0.007) (Figure 7).

The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of 3-methyladenine (ng/mg creatinine) in the urine of rats treated DMBA. There was no significant (p=0.0873) difference between the groups, whereas highly significant differences (p<0.0001) were found between the measurement dates. Compared to the control group, where the concentration of the biomarker in all weeks was maintained at the same level, a decrease in the concentration of 3-methyladenine was observed in rats supplemented with macrogenistein from the 11th to the 17th week (p=0.04) and at the 20th week (p=0.006). Moreover, a decrease in the concentration of the investigated methyl derivative was observed in the group supplemented with microgenistein between weeks 9 and 20 (p=0.003) and 11 and 20 (p=0.01). A significant decrease in the concentration of 3-

Table V. The results of the weight of rat liver and spleen (g) after decapitation.

|          | Liver          |         | Spleen         |         |
|----------|----------------|---------|----------------|---------|
|          | Standard       | Macro   | Micro          | Nano    |
| Arithmetic mean | 6.6763   | 7.6588  | 7.3413         | 7.2613  |
| Standard deviation | 0.8071   | 0.7510  | 0.6076         | 0.7137  |
| Standard error of the mean | 0.2854   | 0.2655  | 0.2148         | 0.2523  |
| ANOVA    | F=2.5662, p=0.0745 |         |                |         |

ANOVA F=1.1765, p=0.3364
Figure 7. The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of O-methylguanosine (ng/mg creatinine) in the urine of rats treated DMBA. Standard: blue circles, Macro: red squares, Micro: green triangles, Nano: yellow diamonds. The same letter over two averages means no significant difference between them (p>0.05); if in two compared averages the letter is not repeated, then the averages differ significantly (p<0.05). The results obtained in each group were compared using analysis of variance and the post-hoc Tukey test.

Figure 8. The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of 3-methyladenine (ng/mg creatinine) in the urine of rats treated DMBA. Standard: blue circles, Macro: red squares, Micro: green triangles, Nano: yellow diamonds. The same letter over two averages means no significant difference between them (p>0.05); if in two compared averages the letter is not repeated, then the averages differ significantly (p<0.05). The results obtained in each group were compared using analysis of variance and the post-hoc Tukey test.
Figure 9. The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of 1-methylguanine (ng/mg creatinine) in the urine of rats treated DMBA. Standard: blue circles, Macro: red squares, Micro: green triangles, Nano: yellow diamonds. The same letter over two averages means no significant difference between them (p>0.05); if in two compared averages the letter is not repeated, then the averages differ significantly (p<0.05). The results obtained in each group were compared using analysis of variance and the post-hoc Tukey test.

Figure 10. The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of 1-methyladenosine (μg/mg creatinine) in the urine of rats treated DMBA. Standard: blue circles, Macro: red squares, Micro: green triangles, Nano: yellow diamonds. The same letter over two averages means no significant difference between them (p>0.05); if in two compared averages the letter is not repeated, then the averages differ significantly (p<0.05). The results obtained in each group were compared using analysis of variance and the post-hoc Tukey test.
Figure 11. The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of 7-methylguanine (μg/mg creatinine) in the urine of rats treated DMBA. Standard: blue circles, Macro: red squares, Micro: green triangles, Nano: yellow diamonds. The same letter over two averages means no significant difference between them (p>0.05); if in two compared averages the letter is not repeated, then the averages differ significantly (p<0.05). The results obtained in each group were compared using analysis of variance and the post-hoc Tukey test.

Figure 12. The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of 8-hydroxy-2’-deoxyguanosine (ng/mg creatinine) in the urine of rats treated DMBA. Standard: blue circles, Macro: red squares, Micro: green triangles, Nano: yellow diamonds. The same letter over two averages means no significant difference between them (p>0.05); if in two compared averages the letter is not repeated, then the averages differ significantly (p<0.05). The results obtained in each group were compared using analysis of variance and the post-hoc Tukey test.
Figure 13. The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of N6-methyl-2’deoxyadenosine (ng/mg creatinine) in the urine of rats treated DMBA. Standard: blue circles, Macro: red squares, Micro: green triangles, Nano: yellow diamonds. The same letter over two averages means no significant difference between them (p>0.05); if in two compared averages the letter is not repeated, then the averages differ significantly (p<0.05). The results obtained in each group were compared using the analysis of variance and the post-hoc Tukey test.

Figure 14. The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat’s life) in the levels of 7-methylguanosine (ng/mg creatinine) in urine of rats treated DMBA. Standard: blue circles, Macro: red squares, Micro: green triangles, Nano: yellow diamonds. The same letter over two averages means no significant difference between them (p>0.05); if in two compared averages the letter is not repeated, then the averages differ significantly (p<0.05). The results obtained in each group were compared using analysis of variance and the post-hoc Tukey test.
methyladenine was also observed between weeks 11 and 20 (p=0.04) in the nanogenistein-supplemented group (Figure 8).

The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat's life) in the levels of 1-methylguanine (ng/mg creatinine) in the urine of rats treated with DMBA. Highly significant (p<0.0001) differences between the groups were found, including persistently high levels of the methyl derivative in rats that received diets supplemented with macro-, micro- and nano-genistein. The differences between the measurement dates were not significant (p=0.0533). It was found that in the initial period (week 9 and 11) the concentration of urinary 1-methylguanine was significantly lower in the control group compared to groups supplemented with micro- and nano-genistein (p>0.05). Similar results were observed at week 11 in the case of rats supplemented with macro-, micro- and nano-genistein (as compared with rats that received only standard diet) (p>0.05). At week 17, rats supplemented with nanogenistein were characterized by significantly higher levels of the urinary 1-methylguanine than rats without supplementation. Moreover, a statistically significant decrease in the concentration of 1-methylguanine was observed in the group supplemented with microgenistein between week 9 and week 17 (p=0.04) (Figure 9).

The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat's life) in the levels of 1-methyladenosine (μg/mg creatinine) in the urine of rats treated DMBA. Highly significant (p<0.0001) differences between groups and (p=0.0102) between weeks of measurements were found. The rats receiving only standard diet were characterized by lower content of 1-methyladenosine in the urine at week 9 in comparison with the rats supplemented with microgenistein (p<0.05). Statistically significant differences between the groups were observed at week 11; rats fed only with standard diet (without supplementation) had a lower concentration of urinary 1-methyladenosine in comparison with the rats supplemented with macro-, micro- and nano-genistein (p<0.05). At weeks 17 and 20, the results of all groups did not differ significantly (p>0.05). Moreover, a statistically significant decrease in the concentration of 1-methyladenosine was observed in the group of rats supplemented with microgenistein between week 9 and week 17 (p=0.01) (Figure 10).

The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat's life) in the levels of 7-methylguanine (μg/mg creatinine) in the urine of rats treated with DMBA. Highly significant differences between the groups and (p<0.0001) between measurement dates (p<0.0001) were found. At week 9, rats that received only standard diet had a significantly lower concentration of 7-methylguanine in urine compared to rats supplemented with micro- and nano-genistein. A similar trend was also observed at week 11, where the concentration in the “standard” group was significantly lower than those in the macro-, micro-, and nano-genistein supplementation groups. At week 17, rats supplemented with nanogenistein were characterized by significantly higher levels of 7-methylguanine than the rats without any supplementation. However, at week 20 concentration of 7-methylguanine did not differ significantly between the groups (p>0.05). In addition, in rats supplemented with microgenistein, the concentration of 7-methylguanine in the urine was significantly reduced both between week 9 and 17 weeks (p=0.001) and between week 9 and 20 (p=0.040) (Figure 11).

The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat's life) in the levels of 8-hydroxy-2'-deoxyguanosine (ng/mg creatinine) in the urine of rats treated with DMBA. No significant differences (p=0.3497) between groups or between measurement dates were observed (p=0.3228) (Figure 12).

The effect of nano-, micro- and macro-genistein on the kinetics of changes (9th, 11th, 17th, 20th week of rat's life) in the levels of N6-methyl-2’deoxyadenosine (ng/mg creatinine) in the urine of rats treated with DMBA. Highly significant differences between the groups (p<0.01) and between measurement dates (p<0.01) were found. At week 9 the concentration of N6-methyl-2’deoxyadenosine in the urine of rats fed standard diet was significantly lower than in the group of animals supplemented with microgenistein. Also, at weeks 11 and 17, the concentration of N6-methyl-2’deoxyadenosine in the group of animals fed a standard diet were significantly lower than its concentration in the group supplemented with macrogenistein (p<0.01). At week 20, the concentration of N6-methyl-2’deoxyadenosine in the urine of rats did not differ significantly between groups (p>0.05). Moreover, a significant decrease in N6-methyl-2’deoxyadenosine urinary concentration was observed in micro- and nano-genistein-supplemented groups over time. The abovementioned change occurred between week 9 and 11 in the microgenistein group and between week 17 and 20 in the nanogenistein supplemented group (p<0.01, Figure 13).
between week 11 and week 17 in rats that were supplemented with macrogenistein ($p=0.006$). A decrease in the methyl derivative was also observed in the urine of rats supplemented with microgenistein ($p=0.01$). In contrast, in the group of rats subjected to the standard diet, an increase in urinary 7-methylguanosine concentration was observed between week 9 and week 20 ($p=0.01$) (Figure 14).

**Discussion**

Carcinogenesis is accompanied by epigenetic changes. They may be characterized by focal hypermethylation and global hypomethylation. Each of these mechanisms plays an essential role in carcinogenesis. Hypomethylation, which occurs mainly in repetitive regions, is a carcinogenic process; it promotes gene instability, causing erroneous chromosome segregation during cell division (20-22). Hypermethylation may lead to silencing of key tumour suppressors and regulatory regions in the genome, which leads to dysregulation of cell growth and changes in response to cancer therapy (20, 23). In this study, we investigated the effect of the isoflavonoid found in soybean genistein on cancer cells. Soybeans are the most widely used, least expensive, and least caloric way to get large amounts of protein. Soy foods have a lot of isoflavones including genistein, which are estrogen-like. Because their chemical structure is similar to 17-β-estradiol, they bind to estrogen receptors and display higher affinity than for ER-α. Moreover, they can influence the activation of both genomic and nongenomic estrogen signaling pathways. It is important to note that the tumors induced by the carcinogen 7,12-dimethylbenz[a]anthracene are mostly ER/PR+, as in humans (24, 25). The results regarding the effect of genistein are inconclusive, some of them show protective effects and others harmful effects. Nevertheless, the tumorigenic properties of soy isoflavones are well documented in breast cancer cell lines and animal models and are largely associated with their estrogenic properties (26, 27). Estrogen can promote the development and metastasis of breast cancers, therefore, consumption of soy foods or soy isoflavones may be disadvantageous. Researchers from the Memorial Sloan Kettering Cancer Center and Weill Cornell Medical College, have shown that adding a moderate amount of soy to some women’s diets may activate a gene that leads to cancer development (28, 29). Their research confirmed the concerns that soy may affect gene expression associated with breast cancer development. High plasma genistein was associated with a signature characterized by over-expression of FGFR2 and genes that drive the cell cycle and proliferation pathways (28, 29). Our results showed that the supplementation of animals with genistein causes an increase in the excretion of methylated derivatives in the urine of rats. The influence of genistein on the formation of methyl derivatives plays a particularly important role in the early stages of carcinogenesis. In the case of animals receiving only standard diet, the levels of methyl derivatives increased during the course of the study (O-methylguanosine, 1-methyladenosine, 7-methylguanine, 7-methylguanosine) or remained relatively low (3-methyladenine, 1-methylguanine, 7-methylguanine, N6-methyl-2’-deoxyadenosine). However, in the case of animals supplemented with various forms of genistein, the levels of the methylated derivatives were very high from the beginning. It is worth noting that the highest levels of the methyl derivatives were found in the urine of animals supplemented with genistein at 9 and 11 weeks of life, i.e., in the early stage of DMBA-induced neoplastic process. Such high levels of methyl derivatives indicate high cell proliferation. They can be associated with the development of the cancer process. It can be concluded that carcinogenesis was more intensive in rats that were supplemented with genistein. DNA methylation is known to be abnormal in all forms of cancer and carcinogenesis-related changes in DNA methylation can be detected with accuracy in cell-free DNA present in blood, feces, urine and other biological samples. Therefore, cancer-related changes in DNA methylation can be potentially clinically useful for the development of cancer biomarker assays. They would allow initial cancer detection and may be used to monitor treatment efficacy and patients throughout the disease. DNA methylation testing is very promising for the development of simple and specific methods for detecting cancer at an early stage (20). It has been shown that there is homology between the effects of DMBA on rats and estrogens, which can contribute to accelerating tumour development (30). It is also worth mentioning that one study described the occurrence of estrogen receptors after induction of breast tumours using DMBA (31). Considering the histopathological results and the concentration of methyl derivatives, it seems that nanogenistein stimulates cancer cells the most and tumours appear relatively quickly. In the group of animals supplemented with nanogenistein, the first tumours appeared as early as 14 weeks, 2 weeks earlier than in animals receiving only the standard diet and 3 weeks earlier than in animals supplemented with macro- and micro-genistein. The nanoparticles overcome biological barriers readily and enable prolonged blood circulation time, which, in effect, enhances the accumulation of nanoparticles in cancer cells. The nano form changes genistein bioavailability. The nano molecule, which was the smallest form of genistein studied in this study, probably changed the bioavailability. The bioavailability is defined in three basic steps: absorption, penetration into systemic circulation and use by the cells. Reduction of materials to the nano scale can sometimes lead to the development of new structural, phytochemical, electronic, and magnetic properties that are not present in larger sized particles comprised the same material. In comparison with
their large counterpart particles can have novel/different physical properties such as an increased surface area to volume ratio, reactive sites, charge, shape, mobility and thermal properties (32). The form of particles can unambiguously change their properties and therefore, the nanogenistein effects on human physiology. For this reason, nanoparticles are ideally suited for the diagnosis and treatment of various diseases allowing earlier detection of pathological changes and more effective treatment of patients (33). However, considering the structure of genistein and its potential of intensifying the proliferation of cancer cells, the use of nanotechnology in our study had the opposite effect on the severity of cancer. What is interesting, in animals supplemented with microgenistein, although the first tumours appeared at week 17, a week later than in animals receiving only the standard diet, the incidence of tumours was 88%, and the number of tumors per rat ranged from 0 to 3, there was a statistically significant stimulation of tumour growth. The mass of the tumours in the animals receiving microgenistein was significantly higher compared to the mass of the tumours of animals receiving standard diet only, nano- and macro-genistein. Therefore, it is possible that the effect of genistein depends on the size of the particles. Even though we did not expect such results, none of the studied forms of genistein inhibited the development of the cancer process, and on the contrary, they stimulated cancer progression.

In Summary, genistein contained in soy may increase the risk of developing breast cancer. Although, due to estrogenic activity, soy protein products are recommended as a natural alternative to estrogen drugs, many specialists consider that in some cases it may pose a potential risk to health (34). Earlier animal studies have shown that genistein can cause breast cancer (35). In addition, human studies have also provided disturbing results: in the case of a diet with soy protein, rich in this isoflavonoid, an increase in epithelial cells in the breasts, i.e., those that most often become cancer cells, has been observed (36). This article shows that genistein can modify the development of the neoplastic process by changing the methylation status of DNA. Further research is needed to understand the mechanism of action of different sizes of genistein particles on cancer cells and epigenetic alternations.

Conclusion

In this article, genistein has been shown to modify the development of neoplasms, and changes at the epigenetic level, among others, may play an important role in this process. It was observed that supplementation of animals with genistein causes an increase in the excretion of methylated derivatives in the urine of rats. The results of this study and the available literature may indicate stimulation of cancer cells depending on the size of genistein molecules.

Conflicts of Interest

All Authors confirm that there are no conflicts of interest to declare regarding this study.

Authors’ Contributions

All Authors contributed equally to all aspects of this work.

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