Brain MRI Volumetry Analysis in an Indonesian Family of SCA 3 Patients: A Case-Based Study

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Introduction: Spinocerebellar ataxia type-3 (SCA3) is an adult-onset autosomal dominant neurodegenerative disease. It is caused by expanding of CAG repeat in ATXN3 gene that later on would affect brain structures. This brain changes could be evaluated using brain MRI volumetric. However, findings across published brain volumetric studies have been inconsistent. Here, we report MRI brain volumetric analysis in a family of SCA 3 patients, which included pre-symptomatic and symptomatic patients.

Methodology: The study included affected and unaffected members from a large six-generation family of SCA 3, genetically confirmed using PolyQ/CAG repeat expansion analysis, Sanger sequencing, and PCR. Clinical evaluation was performed using Scale for the Assessment and Rating of Ataxia (SARA). Subjects’ brains were scanned using 3.0-T MRI with a 3D T1 BRAVO sequence. Evaluations were performed by 2 independent neuroradiologists. An automated volumetric analysis was performed using FreeSurfer and CERES (for the cerebellum).

Result: We evaluated 7 subjects from this SCA3 family, including 3 subjects with SCA3 and 4 unaffected subjects. The volumetric evaluation revealed smaller brain volumes ($p < 0.05$) in the corpus callosum, cerebellar volume of lobules I-II, lobule IV, lobule VIIb and lobule IX; and in cerebellar gray matter volume of lobule IV, and VIIIA; in the pathologic/expanded CAG repeat group (SCA3).

Conclusion: Brain MRI volumetry of SCA3 subjects showed smaller brain volumes in multiple brain regions including the corpus callosum and gray matter volumes of several cerebellar lobules.

Keywords: SCA3, volumetry, brain, MRI, Indonesia
INTRODUCTION

Spinocerebellar ataxia type-3 (SCA3) is an autosomal dominant neurodegenerative disease characterized by progressive ataxia that usually starts in early to mid-adulthood decade resulting in early wheelchair dependence and immobilization. Although it is rare in the general population with a prevalence of 1–2/100,000 it is the most common subtype of spinocerebellar ataxias (SCA) worldwide (1, 2). SCA3 patients typically present with bilateral cerebellar syndrome, characterized by gait and limb ataxia, although non-ataxia symptoms are also quite common. The non-ataxia features of SCA3 are varied and include extrapyramidal manifestations such as dystonia, parkinsonism, tremor and spasticity (3, 4).

SCA3 is caused by CAG trinucleotide repeats which exceed 46 repeats in the ATXN3 gene located at the N-terminus of chromosome 14 (1, 2, 5–8). To date, there are no curative treatment or treatments that could halt the progression of SCA3. Current treatment is still aimed at symptomatic management using pharmacological therapies; while non-pharmacological management such as various forms of rehabilitation therapies are utilized to improve balance, mobility and speech. Promising disease-modifying therapies such anti-ATXN3 aggregation compounds, ATXN3 mRNA silencing therapies, autophagy stimulation, stem cell and gene therapies are still in the pipeline and or in Phase 2 clinical trials, and are still not available (1, 2, 6, 9).

Current evaluation of patients with SCA 3 include a detailed clinical assessment, coupled with the measurement of ataxia severity using validated instruments such as the Scale for the Assessment and Rating of Ataxia (SARA) (4, 10). Genetic analysis using Sanger sequencing methods for PolyQ/CAG repeat expansion is required for confirmatory diagnosis, and the detection of the number of CAG repeats. Magnetic resonance imaging (MRI) is part of the clinical workup, although there are no specific distinguishing or diagnostic features unique for SCA3, and typically shows cerebellar atrophy (7, 11–14). An MRI with 3.0-T magnetic field, which is already available in some of the developing countries, is perhaps necessary for a reliable spatial and contrast resolution for volumetric analysis (15, 16).

Recent publications have reported variable findings in the volumetry of various anatomical brain structures in patients with SCA3 (17–24). One recent study reported that patients with SCA 3 had decreased cerebellar and cerebral volumes, and also the basal ganglia, gray matter and the thalamus (18). Given the lack of reports on MRI brain volumetry in SCA3 from the South East Asia region, especially from Indonesia, we therefore conducted this study to evaluate the MRI brain volumetry in SCA 3 patients from a large Indonesian family and compared the findings to unaffected members. We also included volumetric analysis of other brain regions than previously reported.

METHODS

This was a cross-sectional, case-based study involving a large family with SCA 3 within the Garut Regency in Indonesia.

Ethics Statement

This study was reviewed and approved by the Research Ethics Committee of Universitas Padjadjaran no 032107017/2021. Written informed consent in accordance with the ethics committee and Declaration of Helsinki were obtained from all subjects.

Patient Recruitment

Subjects of this study were from a family with progressive hereditary ataxia who were genetically confirmed with ATXN3 gene mutation. This family has 208 members; of these, 34 members had already been clinically evaluated, and 16 of these members had been genetically screened for SCA repeats. Of these 16 subjects, seven consented to take part in this volumetric study (Figures 1A,C).

Clinical Assessment

Clinical assessment was measured using the Scale for Assessment and Rating of Ataxia (SARA). SARA consists of eight items, including tests of gait, stance, sitting, speech, finger-chase test, finger-nose test, fast alternating movements, and heel-shin test (4). According to criteria by Velazquez-Perez et al., an asymptomatic subject is a subject with SARA score of 0 and without any symptoms; those with SARA score of 0–2 and with ataxia symptoms are categorized as prodromal; while those with SARA score of 3 and above are included into ataxic group (10).

Genetic Evaluation

Genetic evaluation was done using blood sample with polyQ/CAG repeat expansion analysis according to Sanger sequencing. According to criteria by du Montcel et al., a person has SCA3 if the CAG repeat length is expanded ≥ 47; otherwise, the individual is considered an unaffected subject (8).

Brain MRI Protocols

Subjects' brain was scanned using 24 channel head coil in a 3.0-Tesla MRI scanner (Signa Pioneer; GE Healthcare, Chicago, United States). Scanning was done in the sagittal plane using isotropic 3D T1 BRAVO sequence (TR/TE 9 ms/3.5 ms, NEX 1, slice thickness 1 x 1 mm). This sequence is able to differentiate brain structure best suited for automatic identification by FreeSurfer and CERES (explained in following section). All brain MRI imaging and analysis were performed by (FH, RH) and two neuroradiology consultants (RH, TSK), who independently reviewed the images.

Image Capture

Captured MRI images were retrieved from DICOM images, using RadiANT ver 2020.2 (Medixant, Poznand, Polanad) (25) (Figure 2). Briefly, DICOM data were mounted to the software, then sagittal plane T1 BRAVO image sequences were loaded. Reconstruction method using multiplanar reformation

Abbreviations: ATXN3, ataxin3 (could refer to gene or protein); CAG, trinucleotide repeat of CAG; CERES, Cerebellum Segmentation; MMSE, Mini-Mental State Examination; MoCa-Ina, Montreal Cognitive Assessment, Indonesian version; MRI, magnetic resonance imaging; PolyQ, polyglutamine; SARA, Scale for the Assessment and Rating of Ataxia; SCA, spinocerebellar ataxia.
Volumetric Calculation

Brain MRI images were analyzed automatically (except for cerebellum) using FreeSurfer ver 7.1.1, software from Martinos Center of Biomedical Imaging, and Mass General Hospital / Harvard University (Boston, MA), https://surfer.nmr.mgh.harvard.edu/. Briefly, after installation of Freesurfer computer (Mac/Linux OS), designated T1 3D BRAVO images were loaded into the software. First, -reconall command was executed to get skull stripping, and brain mask feature applied to the image for volumetric evaluation. Volumetric evaluation was computed automatically which required ∼8–12 h per subject. The results were marked brain region, and inspected by two independent neuroradiology consultants. If the brain marking was not satisfactory, a tweak would be done, and automatic reevaluation would be repeated from the beginning. And if after re-evaluation, brain marking was still not satisfactory, a second brain MRI scan was performed (26, 27).

For automatic cerebellar evaluation, CERES (Cerebellum Segmentation) ver 1.0 was used (developed by Universidad Politecnica de Valencia and Université de Bordeaux [Informatica biomedica-IBIME, Valencia, Spanyol], https://www.volbrain.upv.es/) (28). Briefly, DICOM image were converted to NIFTI format, then loaded to VolBrain server. CERES pipeline was selected, and evaluation gray matter volume, white matter volume, and cortical thickness of each cerebellar lobule was done automatically.

Volumetric Evaluation

Volumetric data calculated by FreeSurfer and CERES were processed and compared with the percentage intracranial volume that was provided by the volumetric analysis. Brain volumetry of the anatomical structures in this study was calculated as ratio of the individual subject’s intracranial volume (ICV). The volume ratio is used since there is no standard or normal range for the volume of the various brain structures yet. In addition, the shape of the head and the volume of the human brain varies. Therefore, an objective standard is needed in the volumetric measurement of the brain. This can be achieved by measuring the ratio of volume relative to ICV. Since we use this ICV as the denominator in the volume calculation, so we name it percentage to ICV (%ICV), to be more precise.

Heat Map

In order to make the assessment easier, a heat-map color grading pattern was employed (Tables 1A,B). Briefly, the cells in the table are colored by conditional formatting using color scale feature, with green for the largest number gradating to yellow and red for the smallest number.

Statistical Analysis

Since the sample size is small, the non-parametric Mann-Whitney U test was used to compare two groups in terms of quantitative variables. Data were analyzed using SPSS ver 25.0 for Windows (IBM Corp. IBM SPSS Statistics for Windows, Version 25.0 Armonk, NY: IBM Corp). Mann-Whitney U test
was employed for non-parametric. \( P < 0.05 \) is considered statistically significance.

**RESULT**

Our subjects came from a large six-generation family with 208 members with a history of progressive cerebellar syndrome, genetically confirmed as SCA3. Clinically affected family members were one person from the first generation; two members from the second generation, 10 members from the third generation, 54 people in the fourth generation, 89 people in the fifth generation and 11 people, thus far, in the sixth generation. The relevant pedigree is shown in (Figures 1A,B).

Out of those 208 members, only seven were willing to take part in this volumetric study (Figure 1A). Subjects 1–3 are SCA3 patients and siblings with Subject 4. They are descendants from a father (IV.40), paternal grandfather (III.8), great grandmother (II.1) and great-great grandmother (I.1), who had ataxia complaints. Subjects 5–7 are unaffected. Subject 5 had parents and grandparents with no ataxia complaints. Subject 6 is a descendant of a father (IV.5), paternal grandfather (III.1), great grandmother (II.1) and great-great grandmother (I.1), who had progressive ataxia complaints. The parents of Subject 7 had no ataxia complaint, despite a great grandmother (II.1) and great-grandmother (I.1), who had ataxia complaints.

Genetic evaluation showed that subjects 1–3 were confirmed as SCA3 subjects (Figure 1C) as they had heterogeneous alleles, with one of the alleles having 68, 71 and 73 expanded CAG repeats, respectively. The other allele had 8 repeats in all three subjects. Subject 4 had an intermediate number of repeats with 23 and 30 CAG repeats. Subject 5 had medium number of CAG repeats (17 and 23), while Subjects 6 and 7 are included as short CAG repeats (13).

From a clinical point of view, according to criteria by Velazquez-Perez et al. (10), Subjects 1 and 2 are considered to have genetically confirmed SCA 3, while Subject 3 is considered prodromal. All other subjects were considered as unaffected. The clinical details of SCA 3 subjects and controls are shown in (Figure 1C).

The ages of subjects 1, 2 and 3 were 35, 34 and 18 years old, respectively and the onset of ataxia symptoms were at 22, 13 and 15 years of age, with disease duration of 13, 21, and 3 years, respectively. The clinical features of all three subjects are summarized in (Figure 1C). Subject 1, a female had gait ataxia and balance difficulties and muscle cramps, with mild cognitive impairment (MoCA-Ina: 22; MMSE: 24) and a SARA score of 3 (gait: 1; standing: 1; and heel-shin coordination: 1). Subject 2, a male, is considered ataxic, complaining of walking and postural problems with severe talking and mild/moderate coordination problems, with mild cognitive impairment (MoCA-Ina: 23) and a SARA score of 14 (gait: 4; standing: 2; talk: 3; finger chase: 1; nose-finger: 1; hand alternating movement: 1; and heel-shin: 2). Subject 3, a male, is considered prodromal and, despite a normal motor condition, has had cognitive complaints from 15 years old (MoCA-Ina 22; MMSE: 21), and a SARA score of 0, plus numbness on his skin. All other

### TABLE 1A | Brain MRI Volumetry

| Group | Subject | Total Gray Matter Volume | Subcortical Gray Matter Volume | Cortical Volume | Diencephalon Corpus Callosum | Cerebrum and Brainstem | Thalamus Basal Ganglia | Ventricle | Brainstem Hippo-campus Amygdala nucleus Accumbens Diencephalon Corpus Callosum |
|-------|---------|--------------------------|-------------------------------|---------------|-----------------------------|-----------------------|-----------------------|-----------|---------------------------------|-----------------------------|
| SCA3  | 1       | 1.05%                    | 1.26%                         | 1.42%         | 1.51%                       | 0.61%                 | 0.20%                 | 0.09%     | 0.54%                           | 0.19%                       |
|       | 2       | 0.90%                    | 1.26%                         | 2.33%         | 1.26%                       | 0.55%                 | 0.20%                 | 0.07%     | 0.57%                           | 0.64%                       |
|       | 3       | 1.12%                    | 1.58%                         | 1.03%         | 1.46%                       | 0.56%                 | 0.23%                 | 0.07%     | 0.57%                           | 0.54%                       |
| Normal | 4       | 1.09%                    | 1.63%                         | 1.63%         | 1.48%                       | 0.66%                 | 0.20%                 | 0.07%     | 0.57%                           | 0.56%                       |
|       | 5       | 0.99%                    | 1.58%                         | 1.58%         | 1.49%                       | 0.50%                 | 0.23%                 | 0.07%     | 0.57%                           | 0.56%                       |
|       | 6       | 0.99%                    | 1.49%                         | 1.33%         | 1.49%                       | 0.50%                 | 0.22%                 | 0.07%     | 0.57%                           | 0.56%                       |
|       | 7       | 1.07%                    | 1.26%                         | 1.33%         | 1.50%                       | 0.51%                 | 0.23%                 | 0.07%     | 0.57%                           | 0.56%                       |
| p-value |       | 0.571                    | 0.200                         | 0.029*        | 0.114                       | 0.114                 | 0.314                 | 0.114     | 0.114                           | 0.114                       |

\* \( p < 0.05 \).
### TABLE 1B | Cerebellum.

| Group  | Subject No | Lobule I-II | Lobule III | Lobule IV | Lobule V | Lobule VI | Lobule Crus I | Lobule Crus II | Lobule VIIIB | Lobule VIIIA | Lobule VIIIB | Lobule IX | Lobule X | Total |
|--------|------------|-------------|------------|-----------|----------|-----------|---------------|---------------|--------------|--------------|--------------|-----------|----------|-------|
| SCA3   | 1          | 0.01%       | 0.08%      | 0.32%     | 0.59%    | 1.30%     | 1.72%         | 1.39%         | 0.71%        | 0.80%        | 0.52%        | 0.48%     | 0.08%    | 8.99% |
|        | 2          | 0.01%       | 0.10%      | 0.26%     | 0.50%    | 1.11%     | 1.55%         | 1.13%         | 0.59%        | 0.79%        | 0.51%        | 0.44%     | 0.06%    | 7.98% |
|        | 3          | 0.01%       | 0.08%      | 0.33%     | 0.56%    | 1.16%     | 1.62%         | 1.07%         | 0.57%        | 0.90%        | 0.55%        | 0.45%     | 0.07%    | 8.42% |
| Normal | 4          | 0.01%       | 0.09%      | 0.34%     | 0.58%    | 1.27%     | 1.80%         | 1.11%         | 0.58%        | 0.86%        | 0.62%        | 0.66%     | 0.09%    | 9.73% |
|        | 5          | 0.01%       | 0.08%      | 0.35%     | 0.58%    | 1.15%     | 1.45%         | 1.18%         | 0.62%        | 0.82%        | 0.56%        | 0.48%     | 0.07%    | 8.50% |
|        | 6          | 0.01%       | 0.10%      | 0.35%     | 0.59%    | 1.39%     | 2.03%         | 1.14%         | 0.66%        | 0.82%        | 0.68%        | 0.64%     | 0.07%    | 9.72% |

**p-value**

0.029* 0.2 0.029* 0.314 0.2 0.429 0.314 0.029* 0.029* 0.114 0.057

### Gray Matter Volume (%ICV)

| Group  | Subject No | Lobule I-II | Lobule III | Lobule IV | Lobule V | Lobule VI | Lobule Crus I | Lobule Crus II | Lobule VIIIB | Lobule VIIIA | Lobule VIIIB | Lobule IX | Lobule X | Total |
|--------|------------|-------------|------------|-----------|----------|-----------|---------------|---------------|--------------|--------------|--------------|-----------|----------|-------|
| SCA3   | 1          | 0.00%       | 0.06%      | 0.29%     | 0.54%    | 1.18%     | 1.50%         | 1.21%         | 0.63%        | 0.72%        | 0.45%        | 0.41%     | 0.07%    | 7.06% |
|        | 2          | 0.00%       | 0.08%      | 0.24%     | 0.44%    | 1.01%     | 1.36%         | 0.98%         | 0.53%        | 0.70%        | 0.45%        | 0.36%     | 0.05%    | 6.21% |
|        | 3          | 0.00%       | 0.07%      | 0.28%     | 0.46%    | 1.10%     | 1.58%         | 0.90%         | 0.63%        | 0.72%        | 0.48%        | 0.40%     | 0.07%    | 6.70% |
| Normal | 4          | 0.00%       | 0.09%      | 0.35%     | 0.57%    | 1.33%     | 1.83%         | 1.14%         | 0.60%        | 0.89%        | 0.62%        | 0.63%     | 0.09%    | 8.15% |
|        | 5          | 0.00%       | 0.06%      | 0.32%     | 0.51%    | 1.05%     | 1.29%         | 1.05%         | 0.56%        | 0.73%        | 0.49%        | 0.41%     | 0.06%    | 6.53% |
|        | 6          | 0.01%       | 0.08%      | 0.30%     | 0.53%    | 1.26%     | 1.78%         | 0.98%         | 0.58%        | 0.72%        | 0.58%        | 0.53%     | 0.07%    | 7.44% |

**p-value**

0.314 0.571 0.029* 0.2 0.314 0.429 0.429 0.2 0.029* 0.057 0.2 0.571 0.314

### Cortical Thickness (mm)

| Group  | Subject No | Lobule I-II | Lobule III | Lobule IV | Lobule V | Lobule VI | Lobule Crus I | Lobule Crus II | Lobule VIIIB | Lobule VIIIA | Lobule VIIIB | Lobule IX | Lobule X | Total |
|--------|------------|-------------|------------|-----------|----------|-----------|---------------|---------------|--------------|--------------|--------------|-----------|----------|-------|
| SCA3   | 1          | 2.03        | 3.48       | 5.12      | 5.12     | 5.12      | 4.97          | 4.97          | 5.00         | 4.96         | 5.04         | 4.64      | 3.63     | 4.97  |
|        | 2          | 3.56        | 3.96       | 5.21      | 5.11     | 5.12      | 5.11          | 5.02          | 4.94         | 4.87         | 5.04         | 4.43      | 3.47     | 4.98  |
|        | 3          | 1.95        | 3.66       | 5.18      | 5.14     | 5.14      | 5.25          | 5.04          | 5.18         | 5.03         | 5.01         | 4.74      | 3.23     | 5.04  |
| Normal | 4          | 2.38        | 3.70       | 5.22      | 5.15     | 5.21      | 5.31          | 5.23          | 5.12         | 5.00         | 5.18         | 4.94      | 3.71     | 5.14  |
|        | 5          | 2.28        | 3.55       | 5.13      | 5.01     | 5.15      | 5.02          | 4.84          | 5.03         | 4.87         | 5.04         | 4.52      | 3.22     | 4.94  |
|        | 6          | 1.16        | 3.18       | 5.22      | 4.91     | 5.19      | 5.17          | 5.18          | 5.04         | 4.98         | 4.96         | 4.54      | 3.51     | 5.07  |
|        | 7          | 2.52        | 3.30       | 5.07      | 5.17     | 5.17      | 5.10          | 4.83          | 4.96         | 4.93         | 4.97         | 4.54      | 3.63     | 4.97  |

**p-value**

0.571 0.2 0.371 0.571 0.257 0.2 0.543 0.429 0.486 0.457 0.314 0.371 0.457

Volume is presented as a percentage of intracranial volume; cortical thickness is presented in mm.

* A heatmap color scale is used. *p < 0.05.
**TABLE 1C** | Cerebellar atrophy SCA3 subjects.

| Subject No. | 1     | 2     | 3     | Function          |
|-------------|-------|-------|-------|------------------|
| Lobules I-II| v     | v     | v     | Motor            |
| Lobule III  | v     | -     | v     | Motor            |
| Lobule IV   | v     | v     | v     | Motor            |
| Lobule V    | -     | v     | o     | Motor            |
| Lobule VI   | -     | v     | o     | Motor/cognitive  |
| Crus I      | -     | o     | o     | Cognitive        |
| Crus II     | -     | o     | v     | Cognitive        |
| Lobule VIIB | -     | o     | v     | Cognitive        |
| Lobule VIII | v     | v     | -     | Motor            |
| Lobule VIII | v     | v     | v     | Motor            |
| Lobule IX   | v     | v     | v     | Cognitive        |
| Lobule X    | -     | v     | v     | Cognitive        |

$v =$ smaller volume than in normal subjects.  
$o =$ smaller volume than all but 1 normal subject.  
$- =$ approximately the same volume as in normal subjects.

Subjects (4–7) had <47 CAG repeats, so they were considered unaffected subjects.

Qualitatively, we compared MRI brain images from SCA3 subjects (Subjects 1–3) and an unaffected subject (Subject 7) (Figure 2). Dilation of the lateral ventricle (red arrow) and fourth ventricle (green arrow) was observed in Subjects 1 and 2. We also observed smaller brainstem (yellow arrow) and cerebellar (orange arrowhead) volumes. In contrast, the quantitative MRI brain image of Subject 3 is presumably equal to that of an unaffected subject. This might be due to the duration of disease and young age.

Detailed brain volumetric results of subjects grouped by CAG repeats and clinical data can be found in Tables 1A,B. Statistical analysis showed a statistical significance difference between SCA3 and unaffected subjects in volume of corpus callosum, and cerebellum (lobules I-II, lobule IV, lobule VII and lobule IX) and also in volume in gray matter of cerebellum (lobule IV, and VIII) but not in other measurements.

**DISCUSSION**

SCA3 is a neurological disease which begins as early as the 2nd decade, characterized by progressive cerebellar syndrome and cerebellar atrophy. Brain atrophy occurs as a result of a toxic gain-of-function, mitochondrial bioenergetic problems, ion channel disturbance, RNA toxicity, repeat-associated non-AUG (RAN) translation, transcription problems, DNA damage, protein transport disturbance and altered ubiquitination processes in the brain. This leads to axonal damage and neuronal and glial death (2, 8). Cortical thinning in SCA3 patients has been associated with the death of Purkinje cells as the sole processing neuron in the cerebellum and damage to cortical glia (Bergmann's glia, stellate cells and basket cells), which contributes to progressive incoordination and ataxia (29, 30).

In this study, we were able to compare brain volumetric findings among members from the same family, who were clinically affected and had genetic confirmation of SCA 3 with unaffected subjects who did not carry SCA 3 genetic mutation. Expectedly, the cerebellar volume of the SCA3 subjects was lower than that of the unaffected subjects. There were however variations in the lobules affected among the three subjects affected by SCA 3. Motor and cognitive function are currently known to be controlled not only by the cerebrum but also by the cerebellum. According to the double motor and triple non-motor concept of cerebellar function, the cerebellar lobules are divided into two motor areas: (1) lobules I-VI and (2) lobule VIII, while the three non-motor areas are (1) lobule VI/crus I, (2) crus II/lobule VII, and (3) lobules IX/X (31–33).

In Subject 1, with an SCA3 duration of 13 years, there were lower brain volumes in almost all motor lobules with partial atrophy in the cognition lobules. Atrophy in the motor
### TABLE 2 | Findings in this study compared to other studies.

| Brain region | This study | Wan et al. | Klaes et al. | Arruda et al. | de Rezende et al. | Guo et al. | Faber et al. | Hernandez-Castillo et al. |
|--------------|------------|------------|--------------|----------------|-------------------|-----------|-------------|--------------------------|
| **Study**    | Indonesian population | Review of 12 studies | Review of 4 studies | Brazilian population | Brazilian population | Chinese population | Brazil, China, France, Germany, Netherland, Spain, UK, and US | Mexican Population |
| **Number of subjects** | 3 SCA3 patients, 4 normal subjects | 252 SCA3 patients, 250 normal subjects | 246 SCA3 patients, 246 normal subjects | 19 SCA3 patients, 19 normal subjects | 49 SCA3 patients, 49 normal subjects | 47 SCA3 patients, 49 normal subjects | 210 SCA3 patients, 63 healthy subject | 17 SCA3 patients, 17 healthy subject |
| **Journal/year** | - | FNeu: 2020 | Am J Neurorad: 2016 | Cerebellum: 2020 | Eur J Neu: 2014 | Neurology: 2020 | Mov Dis: 2021 | Cerebellum Ataxias: 2017 |
| **Brain region** | **Cerebellum – Gray matter (lobule V, VIIIA) ↓** * | Cerebellum – hemispheric ↓ (100%) | Cerebellum – cortex ↓ | Cerebellum – cortex ↓ | Cerebellum – cortex ↓ | Cerebellum – volume ↓ | Crun II, vermis IX, lobule I-IV – gray ↓ |
| **Cerebellum (lobules I-II, IV, VIIIB, IX) ↓** * | Cerebellum – vermis ↓ (75%) | Cerebellum – white matter ↓ | Cerebellum – white matter ↓ |
| **Cerebellum – cortical thickness – ns** | Brainstem – ns | Brainstem ↓ (75%) | Brainstem ↓ (100%) | Midbrain ↓ | Brainstem ↓ |
| **Brainstem – ns** | Brainstem ↓ (50%) | Midbrain ↓ | Brainstem ↓ | Pons ↓ |
| **Ventricle – ns** | Ventricle ↓ | Cerebrum – cortex ↓ | Cerebrum – cortex ↓ | Left lingual gyrus ↓ |
| **Cerebrum – white matter – ns** | Cerebrum ↓ (67%) | Thalamus – ns | Thalamus – ns | Thalamus ↓ |
| **Thalamus – ns** | Thalamus ↓ (25%) | Basal ganglia – ns | Pallidum – ns | Caudate nucleus, putamen, pallidum ↓ |
| **Basal ganglia – ns** | Basal ganglia ↓ (25%) | Amygdala – ns | Amygdala ↓ (42%) | Thalamus ↓ |
| **Amygdala – ns** | Limbic regions ↓ | Nucleus accumbens – ns | Diencephalon ↓ | Caudate nucleus, pallidum ↓ |
| **Corpus callosum ↓** | Subcortical gray matter – ns | Subcortical & total gray matter |

↓: Volume decrease; *: p < 0.05; ns, not significant; (%), percentage of patients showing the indicated change.
regions was in accordance with the ataxia complaints; however, there were some discrepancies in the lobules that command cognition and the subject's cognitive condition. In Subject 2, the volumetric findings corresponded to the motor and cognitive conditions. In Subject 3, the cognitive findings were in accordance with the volumetric findings, although the motoric findings were not in accordance with the volumetric findings (Table 1C).

Based on recent publications, there have been different findings on volumetric differences in various brain anatomical structure of patients with SCA3 (17–24) (Table 2). Researchers from many regions reported their findings; however, there were no report found on ASEAN populations. Wan et al. reviewed 12 publications with a total of 252 subjects and found atrophy in the cerebellum, brain stem, cerebrum, limbic system, basal ganglia and thalamus (19). Klaes et al. reviewed 4 volumetric studies with 246 subjects and found brain atrophy in the cerebellum and brainstem (20). Arruda et al. studied 19 SCA3 subjects in a Brazilian population and showed atrophy in the brainstem and cerebellum (18). Rezende et al. studied 43 SCA3 subjects in a Brazilian population and found vast atrophy in cortical and subcortical regions, including the cerebellum, thalamus, putamen, globus pallidum, diencephalon and brainstem (21). Guo et al. studied 47 SCA3 subjects in a Chinese population and showed progressive atrophy in cerebellar gray matter and several cerebrum regions (22). Faber et al. gathered 210 SCA3 subjects from 8 countries (Brazil, China, France, Germany, Netherland, Spain, UK, and US) and found atrophy in cerebellum, brainstem, thalamus, basal ganglia with dilation of ventricle (23). Hernandez-Castillo et al. studied 17 SCA3 patients from Mexican population and found atrophy in cerebellum, pons and left lingual gyrus (24). Overall there is an agreement that there is decreased cerebellar and cerebral volumes with various other affected regions and with varying degree. Hence, we evaluated MRI brain volumetry with more region than existing publication in SCA3 patients, moreover there are no study examining brain MRI of ASEAN and Indonesian origin. A more thorough evaluation with tractography and an analysis with all the related motor and cognitive brain regions is necessary to clarify the discrepancies observed.

Our patients had onset of disease in the second and third decades, which is relatively early compared to the average published mean age of onset which is in the fifth decade but still fell within the age range of second to sixth decade. Subject 3 was the youngest, with the shortest duration of disease, and therefore had unclear amount of brain atrophy, as age and duration of disease affect the degree of brain atrophy (8, 34–36).

**LIMITATIONS OF STUDY**

Being case-based and a pilot study, our study is limited by a small sample size, which was affected further by the COVID-19 pandemic during the study period. The COVID-19 pandemic hindered the ability of family members to participate and be examined, due to transportation limitations and isolation protocols from the regional government, in the efforts to prevent the spread of COVID-19 pandemic. A study with larger sample size would be necessary for a more generalizable conclusion.

Despite the limitations, we believe our study findings could contribute to the existing literature on brain volumetry and clinical details of SCA 3 patients, in the South East Asia region, given the paucity of data in this region. To our knowledge, this is the first volumetric evaluation of SCA3 patients conducted in Indonesia. Additionally, as we had also included more brain regions in our evaluation as compared to previous studies, we measured the thalamus, basal ganglia, ventricles, brainstem, amygdala, nucleus accumbens, diencephalon, corpus callosum, white matter, and subcortical gray matter, in addition to the measurement of cerebellar volume, cerebellar gray matter volume and cerebellar cortical thickness. Statistically significant differences were found in volume of corpus callosum, and cerebellum (lobules I-II, lobule IV, lobule VIIA and lobule IX) and also in volume in gray matter of cerebellum (lobule IV, and VIIA) but not in other measurements.

In conclusion, our findings confirm previous findings that various brain regions are atrophied in SCA 3, and perhaps may involve other parts of the brain not previously reported. However, our findings remain preliminary and need to be verified further in larger studies. Brain MRI volumetric analysis could thus be an additional tool for non-invasive evaluation of patients with degenerative cerebellar ataxia in addition to the currently utilized clinical scales (20, 37).

**DATA AVAILABILITY STATEMENT**

The original contributions presented in the study are included in the article material, further inquiries can be directed to the corresponding authors.

**ETHICS STATEMENT**

The studies involving human participants were reviewed and approved by Universitas Padjadjaran, Research Ethic Committee. The patients/participants provided their written informed consent to participate in this study.

**AUTHOR CONTRIBUTIONS**

SS: study design, conceptualization, data collection, data analysis, manuscript revision, and funding. FH: study design, conceptualization, data collection, data analysis, manuscript drafting, and manuscript revision. TK: study design, data collection, and data analysis. RH: study design, data collection, data analysis, and manuscript drafting. YS: study design, conceptualization, data analysis, manuscript drafting, manuscript revision, and funding. SD, PO, and ND: data analysis
and manuscript revision. IP: study design, data collection, and manuscript revision. UK: study design, data analysis, and manuscript revision. NM: conceptualization and manuscript revision. TA: study design, conceptualization, manuscript revision, and funding. All authors contributed to the article and approved the submitted version.

REFERENCES

1. Buijsen RaM, Toonen LJA, Gardiner SL, Van Roon-Mom WMC. Genetics, mechanisms, and therapeutic progress in polyglutamine spinocerebellar ataxias. Neurotherapeutics. (2019) 16:263–86. doi: 10.1007/s13311-018-00696-y
2. Klockgether T, Mariotti C, Paulson HL. Spinocerebellar ataxia. Nat Rev Dis Primers. (2019) 5:24. doi: 10.1038/s41572-019-0074-3
3. Jacobi H, Rakowicz M, Rola R, Fancellu R, Mariotti C, Charles P, et al. Inventory of non-ataxia signs (inas): validation of a new clinical assessment instrument. Cerebellum. (2013) 12:418–28. doi: 10.1007/s12311-012-0421-3
4. Schmitz-Hubsch T, Du Montcel ST, Baliko L, Hercianio J, Boesch S, Depondt C, et al. Scale for the assessment and rating of ataxia: development of a new clinical scale. Neurology. (2006) 66:1717–20. doi: 10.1212/01.wnl.0000219042.60538.92
5. Kazachkova N, Lima M. Towards a therapeutic intervention in polyglutamine ataxias: from models to clinical trials. Front Clin Res-CNS Neurolog Disord. (2019) 13:36. doi: 10.3389/fncrn.2019.00036
6. Matos CA, De Almeida LP, Nobrega C. Machado-joseph disease/spinocerebellar ataxia type 3 (machado-joseph disease). Acta Neuropathol. (2010) 120:449–60. doi: 10.1007/s00401-010-0717-7
7. Seidel K, Siswanto S, Brunt ER, Den Dunnen W, Korf HW, Rub U. Brain pathology of spinocerebellar ataxias. Acta Neuropathol. (2012) 124:1–21. doi: 10.1007/s00401-012-1000-x
8. Mccord MR, Bigio EH, Kam KL, Fernandez-Ruiz J, Bruni O. Redefining cerebellar genes. J Neuropathol Exp Neurol. (2020) 79:641–46. doi: 10.1093/jnen/nla01033
9. Tada M, Nishizawa M, Onodera O. Axonal inclusions in spinocerebellar ataxia type 3. Acta Neuropathol. (2010) 120:449–60. doi: 10.1007/s00401-010-0717-7
10. Chow N, Hwang KS, Hurtz S, Green AE, Sømme JH, Thompson PM, et al. Comparing 3t and 1.5t Mri for mapping hippocampal atrophy in the Alzheimer's disease neuroimaging initiative. J Neurosci. (2021) 41:13311–121. doi: 10.1177/0022305320983549
11. Ladd ME, Bachert P, Meyerspeer M, Moser E, Nagel AM, Norris DG, et al. Pros and cons of ultra-high-field mri/mrs for human application. Prog Nucl Magn Reson Spectrosc. (2018) 109:1–50. doi: 10.1016/j.pnmrs.2018.06.001
12. Piccinin CC, Rezende TJR, De Paiva JLR, Moyes PC, Martinez ARM, Cendes F, et al. A 5-year longitudinal clinical and magnetic resonance imaging study in spinocerebellar ataxia type 3. Mov Disord. (2020) 35:1679–84. doi: 10.1002/mds.28113
13. Arruda WO, Meira AT, Ono SE, De Carvalho Neto A, Betting L, Raskin S, et al. Volumetric Mri changes in spinocerebellar ataxia (Sca3 and Sca10) patients. Cerebellum. (2020) 19:536–43. doi: 10.1007/s12311-020-01137-3
14. Wan N, Chen Z, Wan L, Tang B, Jiang H. Mr imaging of Sca3/Mjd. Front Neurosci. (2020) 14:749. doi: 10.3389/fnins.2020.00749
15. Klaes A, Reckziegel E, Franca MC Jr, Rezende TJ, Vedolin LM, Jardim LB, et al. Mr imaging in spinocerebellar ataxias: a systematic review AJNR. Am J Neuroradiol. (2016) 37:1405–12. doi: 10.3174/ajnr.A4760
16. Rezende TJ, L’abreu A, Guimarães RP, Lopes TM, Lopes-Cendes I, Cendes F, et al. Cerebral cortex involvement in machado-joseph disease. Eur J Neurolog. (2015) 22:277–83. e23–4. doi: 10.1111/ejn.12559
17. Guo J, Chen H, Biswal BB, Guo X, Zhang H, Dai L, et al. Gray matter atrophy patterns within the cerebellum-neostriatum-cortical network in Sca3. Neurology. (2020) 95:e3064–12. doi: 10.1212/WNL.0000000000010986
18. Faber J, Schaprian T, Berkan K, Reetz K, Franca MC Jr, De Rezende TJ, et al. Regional brain and spinal cord volume loss in spinocerebellar ataxia type 3. Mov Disord. (2021) 36:2273–81. doi: 10.1002/mds.28610
19. Hernandez-Castillo CR, Diaz R, Campos-Romo A, Fernandez-Ruiz J. Neural correlates of ataxia severity in spinocerebellar ataxia type 3/machado-joseph disease. Cerebellum Ataxias. (2017) 4:7. doi: 10.1186/s40673-017-0065-7
20. Medixent. Radiant Dicom Viewer. 2020.2 ed 2020. Available online at: https://www.radiantviewer.com
21. Reuter M, Schmansky NJ, Rosas HD, Fischl B. Within-subject template estimation for unbiased longitudinal image analysis. Neuroimage. (2012) 61:1402–18. doi: 10.1016/j.neuroimage.2012.02.084
22. Imaging MCBF. Freesurfer. 711 ed. Harvard Medical School, Massachusetts General Hospital (2021). Available online at: https://surfer.nmr.mgh.harvard.edu
23. Manjohn IV, Coupe P, Volbrin: an online MRI brain volumetry system. Front Neuroinform. (2016) 10:30. doi: 10.3389/fninf.2016.00030
24. Scherzed W, Brunt ER, Heinsen H, De Vos RA, Seidel K, Bürk K, et al. Pathoanatomy of cerebellar degeneration in spinocerebellar ataxia type 2 (Sca2) and type 3 (Sca3). Cerebellum. (2012) 11:749–60. doi: 10.1007/s12311-011-0340-8
25. Robinson KJ, Watchon M, Laird AS. Aberrant cerebellar circuitry in the spinocerebellar ataxias. Front Neurosci. (2020) 14:707. doi: 10.3389/fnins.2020.00707
26. Guell X, Schmahmann J. Cerebellar functional anatomy: a didactic summary based on human fMRI evidence. The Cerebellum. (2020) 19:1–5. doi: 10.1007/s12311-019-01083-9
27. Guell X, Gabrieli JDE, Schmahmann JD. Triple representation of language, working memory, social and emotion processing in the cerebellum: convergent evidence from task and seed-based resting-state fMRI analyses in a single large cohort. Neuroimage. (2018) 172:437–49. doi: 10.1016/j.neuroimage.2018.01.082
28. Buckner RL, Krienen FM, Castellanos A, Diaz JC, Yeo BT. The organization of the human cerebellum estimated by intrinsic functional connectivity. J Neurophysiol. (2011) 106:2322–45. doi: 10.1152/jn.00339.2011
29. Paulson HL. The spinocerebellar ataxias. J Neuropsychiat Disord. (2009) 29:227–37. doi: 10.1097/WN00b013e3181b416de
30. McLaughlin HS, Moore LR, Paulson HL. Pathogenesis of Sca3 and Implications for other polyglutamine diseases. Neurobiol Dis. (2020) 134:104635. doi: 10.1016/j.nbd.2019.104635
31. Guell X, Schmahmann J. Implications for other polyglutamine diseases. Neurobiol Dis. (2020) 134:104635. doi: 10.1016/j.nbd.2019.104635
32. Paulson HL, Shakkottai VG, Clark HB, Orr HT. Polyglutamine spinocerebellar ataxias - from genes to potential treatments. Nat Rev Neurosci. (2017) 18:613–26. doi: 10.1038/nrn.2017.92

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