ABSTRACT

Worldwide, tomato leaf miner *Tuta absoluta* (Lepidoptera: Gelechiidae) is one of the most devastating invasive pests of tomato crops. It is one of the most important biotic constraints for tomato production which may lead to the production loss up to 100% if it cannot be controlled. *T. absoluta* comprises four developmental stages: egg, larvae, pupa, and adult with a rapid growth rate. Among these larvae is the most devastating stage which affects the fruit, leaves, and stem. Recently, *T. absoluta* is turned into a key pest of tomato in Bangladesh. This pest was first detected by IPM lab, Horticulture Research Centre, Bangladesh Agricultural Research Institute,
1. INTRODUCTION

Globally, tomato (Solanum lycopersicum L.) is one of the most important economically growing vegetable crops in terms of human consumption due to its valuable nutritional components such as niacin, riboflavin, thiamine, beta-carotene, lycopene, iron, magnesium, phosphorus, potassium, and sodium [1,2]. Tomato belongs to the family Solanaceae which are diploid with 12 pairs of chromosomes (2n = 24) [3]. This crop originated from the South American Andes and the first used as a food in Mexico and extended worldwide. After potato, tomato is the second-largest vegetable crop grown all over the world. Worldwide 4,762,129 ha cultivable land was devoted to tomato cultivation in 2018 and the total production was about 182,258,016 metric tons [4]. In Bangladesh, Tomato has been growing as the second horticultural crop after potato which is cultivated in two seasons annually. For tomato cultivation in both winter and summer season, 68,366 acres cultivable land (8.59% of total cultivable land) was dedicated and the total production was about 3,88,725 metric tons in the year of 2016-2017 [5].

Recently, the production of tomato crops has been considerably affected by several insect pests and diseases [6,7]. Numerous insect pests feed on tomatoes [8] such as thrips, whitefly, Africa fruit bollworm, tomato fruit worm, leaf miner, leafhopper, aphid, mites, and mealy bug [9-11]. Among these, an invasive insect pest, tomato leaf miner, Tuta absoluta Meyrick (Lepidoptera: Gelechiidae) has become the most destructive pest for tomato production in different parts of the world [12-14]. In Bangladesh, Tomato leaf miner, Tuta absoluta is newly emerged as an invasive pest of tomato and causing an explicative decrease in tomato production. T. absoluta can cause high production losses of tomato (i.e. up to 100%) in open field and greenhouse cultivation if left uncontrolled [14-17]. Thus, this review paper aims at establishing the problems package and management strategies towards an available, eco-friendly, cost-effective, and sustainable management of the pest. Many insect pests can be controlled by the integration of different control measures that are not completely feasible when treated alone and T. absoluta is one of them. Therefore, it would be very important to integrate all applicable control methods such as biological and cultural methods, mass trapping, and appropriate use of registered insecticides to control those pests effectively [18,19].

2. GENERAL BACKGROUND OF T. absoluta

2.1 Origin and Distribution of T. absoluta

Tomato leaf miner T. absoluta, commonly known as the South American tomato leaf miner and the pest was originated from central Chile [10,20,21]. T. absoluta was first identified in 1917 and as a pest of tomato in the 1960s in Peru [22]. However, T. absoluta was first recorded in Spain in 2006 [23,24]. Since the origin, T. absoluta has been widespread in South America, Europe then Asia and Africa [10,25,26]. Currently T. absoluta has been widespread almost all over the world on different solanaceous crops [27-31], but it has been introduced as an exotic invasive pest for tomato production in the world-leading tomato producer countries such as Egypt, India, Iran, Italy, Spain and Turkey [30,32]. In recent years, tomato leaf miner, T. absoluta has been officially reported in some South Asian countries like India in 2014 [33] and Nepal in 2016 [15]. In Bangladesh, tomato leaf miner, T. absoluta has first detected its infestation in a tomato field in the village of Chaklarhat (26019‘N, 88043‘E) under the Tunirhat union of Panchagarh Sadar Upazila, Panchagarh district of Bangladesh in May 2016 [34]. Since T. absoluta is one of the newly emerged insect pests of tomato in Bangladesh, the pest has caused significant economic losses in the tomato fields at Jessore, Comilla and Panchagarh Districts [34]. Extensive damage is imposed by this pest as a result of its...
short life cycle, high reproduction capacity, a wide range of host plants and endophytic feeding habits [10,35,36].

2.2 Biology and Life Cycle

*T. absoluta* is a micro lepidopteron moth with rapid reproduction capability [28,37-40]. This pest may complete about 10–12 generations per year under suitable environmental conditions. The life cycle of *T. absoluta* consists of 4 developmental stages: egg, larva, pupa and adult which takes about 24-28 days to complete it, depending on temperatures [36,41]. The developmental periods (Egg to adult emergence) are 76.3, 39.8 and 23.8 days at 14°C, 19.7°C and 27.1°C, respectively [42]. Adults are nocturnal moth these active at night and hide between leaves in the day time. The adult (Fig. 1a) is consists of black spots [28] and grey to silvery scales [43] and black spots with silver or brown speckle on wings. They have also one pair of filiform (bead-like structure) antennae [44]. Adult lifespan ranges between 6–7 days for males and 10–15 days for females [45]. Females lay about 200 eggs during their lifespan [34]. Usually, the pest deposits eggs on the underside of leaves, stems and occasionally on fruits (Fig. 1b) [46]. Egg colour varies between creamy-white to bright yellow [10,47] with 0.2 mm diameter and 0.4 mm length [48]. Eggs hatch within 6-7 days into larvae between temperature range 25-30°C [29]. The most destructive stage of *T. absoluta* is the larval stage that can damage tomato plants during all the growing stages [10]. After hatching, young larvae penetrate tomato fruits (Fig. 1c) [46] and feeding damage is caused when the larvae enter into the leaf and feed on the mesophyll tissue of leaves, as a result, developed irregular mines on the leaf surface and reduce the photosynthetic capacity of the plant [14,49].

These mines are blotchy (Fig. 2a), which are differing from the wavy and narrow mines caused by *Liriomyza* spp. (Fig. 2b) [34]. Larvae pass through four instars and they will be completed within 20 days under favorable environmental conditions and they damage seriously on buds, fruits, leaves and stem [29,45,50]. Pupation can complete its lifespan within 21 days and mostly occur in the soil and it occasionally may take place in the calyx or on leaves [48]. The pupae are green in colour during emergence and changing it to brown near-adult bearing [29].

Fig. 1. *T. absoluta* (a) adult [14]; (b) Egg [46]; (c) Larva on tomato fruit [46]

Fig. 2. Infestation of tomato leaf caused by (a) *Tuta absoluta*; (b) *Liriomyza* spp. [34,47]
2.3 Hosts

Tomato leaf miner *T. absoluta* is a harmful oligophagous insect attacking several plant species with a high preference towards the species of the Solanaceae family. It has a strong preference for tomato (*Solanum lycopersicum* L.) which is considered as the main host [51-53]. Despite the clear favouritism of this insect on tomato plants it also attacks some other cultivated species of Solanaceae including: potato (*Solanum tuberosum*), eggplant (*Solanum melongena*), tobacco (*Nicotiana tabacum* L.), pepinodulce (*Solanum muricatum*), and peppers (*Capsicum annuum* L.) [10,45]. Besides, this pest also attacks some wild species of Solanaceae such as *Solanum bonariense* L., *Solanum sisymbriifolium* L., *Lycopersicon hirsutum*, *S. elaeagnifolium*, *S. hirtum*, *S. lyratum*, *S. nigrum*, *S. puberulum*, *Physalis angulata*, and *Nicotiana glauca*, etc [28,45]. Occasionally, it is also capable to infest on different plant species, like bean (*Phaseolus vulgaris* L.), Jerusalem cherry (*Solanum pseudocapsicum* L.), Peruvian groundberrry (*Physalis peruviana*), green beans (*P. vulgaris*), Dutch eggplant (*Solanum aculeatissimum* Jacq.), *Malva* sp., and corallillo (*Lycium chilense*) [40,45]. Recently, *T. absoluta* is capable of attacking some weeds species as an alternative host like; *Datura ferox*, *Datura stramonium*, *Lycium chilense*, *Nicotiana glauca*, and *Solanum nigrum* [38,39,50,54].

2.4 Management Strategies of *Tuta absoluta*

Considering feeding habits, rapid reproduction capacity, and resistance to the pesticide of *T. absoluta* in the affected territories, it is challenging to control. In Bangladesh, there has not been any reliable, cost effective and sustainable control measures that can be used in the management of the pest by the resource poor farmers. However, there exist several efficient and sustainable control methods that can help to reduce the infestation of *T. absoluta* [47,55]. Therefore, several researchers across the world have been reported using different management strategies for effective control of *T. absoluta*. Methods such as Physical methods, Cultural methods, Biological methods, Chemical methods, the use of pheromone traps and mass trapping, cultivation of *T. absoluta* resistant tomato varieties [47], and Integrated pest management (IPM) have been recommended [28,36,53]. However, a combination of two or more of the above control measures could be much more effective than any single practice for controlling *T. absoluta* [10].

2.4.1 Physical control methods

The aim of the Physical control methods is to prevent or decrease pest attack in a crop. several physical methods act either mechanically or by attacking pests’ sustainability or behavior [56]. Tomato leaf miner and other flying insects such as whitefly, thrips, and bollworm can be physically eliminated from the tomato field as well as greenhouse by practicing diverse techniques [28,40,20]. There are several Physical control methods such as insect suction traps (i.e. yellow sticky traps, Black Sticky Traps, water trap, Tuta Trap Tray, Tray Trap Technology (TTT), and so on), Trap cropping, Field burning, and Elimination of breeding grounds. These methods used to reduce the populations of *T. absoluta*. Insect exclusion screens can also suppress insect density without the mortality of insects. Moreover, insect behavior also can be changed by using colour and chemicals [56,47].

2.4.1.1 Screening the greenhouse

This is one of the most effective physical control measures against *T. absoluta* inside the greenhouse tomato cultivation [10]. The main source of pest invasion is the infested seedling. So, the use of insect-free seedlings is very essential for obtaining a leaf miner free plant. According to PPD [41], to reduce the infestation of tomato seedling, protection of nursery bed by 1.6 mm mesh size net house is very effective. The greenhouse must be sealed properly and screened at vents in the roof and sides. To restrict the entrance of the insects, nets with a minimum density of 9 x 6 threads/cm² should be used [40]. The well-regulated use of double-entry doors can minimize the entry of insects inside the greenhouse [28]. Special consideration should be made for ventilation when the net house or greenhouse will be used. Moreover, the collection and destruction of infested plant parts should be performed regularly [36]. Human movements should be avoided from infested to non-infested greenhouses and cultivators should make secure that live adult moths are not present on their persons before entering greenhouses may also be effective. Soil infested with pupa may also be carried from one place to another.
**2.4.2 Cultural control methods**

Cultural control methods are the oldest practices used to control pests’ infestations. These techniques are normally used to create unfavorable environments for the pests to avoid high-risk of infestations or alleviate the pest density. Cultural control methods are aimed to prevent infestations from developing of pest population because only cultural practices cannot suppress the outbreak of the pests. Moreover, in IPM programs, cultural practices play an important role to control an insect [57]. There are several cultural control methods that aid to reduce the density of the pests. There are some cultural practices such as Good agricultural practices, Management of plant material, and Destruction of crop residues showed the effective result to reduce the population of *T. absoluta* [10].

**2.4.2.1 Good agricultural practices**

*Tuta absoluta* can be controlled by good agricultural practices including ploughing, cultural practices, crop rotation with non-Solanaceous crops (preferably Cruciferous crops), sufficient irrigation, and optimum use of fertilizers, destruction of pests infected plant parts, leaves, and fruits [28,58]. Intercropping tomato with a gallant soldier (*Galinsoga parviflora* L., Asteraceae) and coriander (*Coriandrum sativum* L., Parsley) shows a positive effect on decreasing pest density and enhancing natural enemies [45,59]. Keeping a proper planting distance between rows is also helpful, not having crowding of plants, as is the removal of weeds and keeping the area clean from fascicle residues. It is also encouraged to destruct the wild solanaceous host plants surrounding the growing area because the pest can re-infest the growing crop which is hosted by these wild solanaceous species at all developmental stages of *T. absoluta*. This pest can also be eradicated in greenhouses with the help of some important cultural control practices like selective removal and destruction of infested plant materials and crop rotation. Moreover, the wild host plants should also be removed to keep the crop growing area free from further growth of a strong population of *T. absoluta* that might cause significant damage [10].

**2.4.2.2 Management of plant material**

The use of pest-free transplants is an effective control measure against *T. absoluta* [28]. At the beginning stage of pest infestation, it is important to eradicate any of the symptomatic leaves and stems infested by larvae or pupae and place them in plastic bags to destroy. Weeds (i.e. *Lycium chilense*, *Nicotiana glauca*, and *Solanum nigrum*) should be removed from the crop growing area that may be host to the pest. Infested plant material (from weeding or pruning) should not leave on the ground so that later it will not allow growing a potential population, as the larvae will quickly leave them and colonize on a new plant [10,28].

**2.4.2.3 Destruction of crop residues**

After harvesting, the crop residues should be destroyed as soon as possible. Crop residues covered with plastic for a minimum three weeks reportedly decrease the number of *T. absoluta* adults up to 94% during the fall. Crop residues can also be destroyed by burning or grinding combined with insecticide sprays [60]. Soil solarisation is also effective to reduce the number of pupae that remain in the soil in warm climates [10,60].

**2.4.3 Biological control methods**

**2.4.3.1 Predators**

Several researchers have found greater effectiveness of predators as natural enemies to reduce the population of *T. absoluta* in both open-field tomato farms and greenhouses. It also plays an important role in the IPM program for its plenty and outstanding benefits as it is self-prolific, safe, effective therewith environment-friendly, economical, and sustainable [10,18,36], [12] and [61] defined the mirid *Nesidiocoris tenuis* as a universal and effective predator against *T. absoluta*. This species can prey up to 100 eggs of *T. absoluta* per day [53,62]. It was found in Tunisian [63], Cyprus, Egypt, France, Jordan, Iran [64], Israel, Italy, Morocco, Spain, and Turkey [12]. According to [65], *Macrolophus pygmaeus*, *Nesidiocoris tenuis* (Reuter), and *Orius spp.* are efficient predators, among which, *M. pygmaeus* and *N. tenuis* can control 75 to 97% leaves and 56 to 100% fruits infestations caused by *T. absoluta*, respectively [53,66]; by killing the eggs and larvae [53,67]. In 2012-13 and 2013-14, an experiment was conducted in Turkey to ascertain the efficiency of individual and collective use of predacious insects, *N. tenuis* and the egg parasitoid, *Trichogramma evanescens* as biological control agents against *T. absoluta*. This experiment revealed that the capability to control *T. absoluta* by collective use
of *N. tenuis* and *T. evanescens* was about 95% which showed a higher damage reduction percentage than the single use of *N. tenuis*. However, using the *N. tenuis* only was effective to control the *T. absoluta* about 90% in tomato fruit [68]. In the Mediterranean region using *Nesidio corridentis*, showed highly promising results and effectiveness of predator use when combined with other methods in controlling *T. absoluta* [69,70].

### 2.4.3.2 Parasitoids

Parasitoids are one of the most potent biological control agents that can be used to control the population growth of *T. absoluta*. The most significant *T. absoluta* egg parasitoids have been originated from the family Trichogrammatidae, Encyrtidae, and Eupelmidae [20,22,36]. The egg parasitoid *T. achaeae* was found most effective against *T. absoluta* [71] while compared the efficacy of *Trichogramma todeabactrae*, *Trichogramma pretiosum* [72], and *Trichogramma urquiioli*. The parasitism rates of *T. absoluta* eggs are 57.05% and 63.92% for *Trichogramma cacoeae* and *Trichogramma bourrarae* which reduce the yield losses up to 78.89% and 87.62%, respectively in greenhouse tomato production [73]. In different geographical locations, various parasitoids showed their efficacy such as *Trichogramma pretiosum* was an effective parasitoid in Sub-Saharan Africa [22] and *Stenomesius* sp. was a potential parasitoid in the arid region of Algeria [65]. A research was carried out in Tunisia on recently adapted parasitoid for organic tomato production and found two ectoparasitoids that are *Bracon* sp. and *Necremnusartynes* (Hymenoptera: Eulophidae), attacking and developing on *T. absoluta*, where the eulophid wasp was the most abundant with average parasitism rates of 25.5% [74].

#### 2.4.3.3 Entomo-pathogenic microbes

The uses of microorganisms, like fungi, bacteria, and nematodes as biopesticides for the control of *T. absoluta* are effective [75]. Microbes invade the pest by their pathogenic effect which leads to the death of the pests [14]. In Europe and America, *Bacillus subtilis* (bacteria) and *Metarhizium anisopliae* (fungus) observed as the successful formulation to reduce the population of *T. absoluta* on tomato where *M. anisopliae* and *Beauveria bassiana* have also been observed as an effective formulation [22]. The fungi *Aspergillus flavus* and *Fusarium* sp. caused 100% larval mortality of *T. absoluta* within 6 days of treatment in a laboratory study of Algeria [76]. A study performed under open field conditions showed that *B. bassiana* and *M. anisopliae* at 2.5 × 10^9 conidial/ml gave similar mortality of *T. absoluta* as that of *Chlorantraniliprole* (Fig. 3). Similarly, maximum mortality ensued at 2.5 × 10^9 conidial/ml of *B. bassiana* (84.04%) followed by *M. anisopliae* (76.31%) after 10 days of the treatment under glasshouse condition [75].

![Fig. 3. Mortality of T. absoluta adult using Entomo-pathogenic microbes [75]](image-url)

Huda et al.; JAERI, 21(3): 1-16, 2020; Article no.JAERI.55950
Likewise, a study was conducted to determine the effectiveness of three different entomopathogenic nematodes (EPN) against *T. absoluta*, namely *Heterorhabditis bacteriophora*, *Steinernema carpocapsae*, and *Steinernema feltiae*, where *S. feltiae* was the most effective nematode species under laboratory conditions in Turkey during 2013-2014. The results exposed that these EPN had a good potentiality to control the *T. absoluta* larvae [36,77]. Similarly, the effectiveness of entomopathogenic nematodes *Steinernema carpocapsae*, *Steinernema feltiae*, and *Heterorhabditis bacteriophora* also observed against *T. absoluta* at its late larval instars [10,54,78]. Another entomopathogenic bacterium *Bacillus thuringiensis*, that has also been used as very effective bio-insecticide for control of tomato pests [10,54,78,79]. In IPM based programs, biological control has been used extensively to control the pest in tomato crops. Bio-insecticides like *B. thuringiensis* do not create any environmental hazard as they are environmentally safe and eco-friendly.

### 2.4.3.4 Bio-pesticides (Plant extracts)

Plant-based pesticides are better management strategies than synthetic pesticides because plant-based pesticides are eco-friendly, biodegradable, easily obtainable, economical, and sustainable [36]. Active metabolites exist in plants act as a toxicant for the control of pests which may be contact or systemic [36]. Currently, several plant species have been used to control different agricultural pests by using crude extracts of seeds, leaves, bark, bulbs, and fruits globally [80]. Ethanol and hexane extracts of *Acmella oleracea* showed the highest mortality of 88 to 100% after six hours of application [81]. Neem extracts at various concentrations caused 24.5% egg and 86.7 to 100% larval death [82]. Petroleum ether extract of *Jatropha* caused 18 to 25% egg and 87 to 100% larval mortality within 4 days of treatment and 33 to 46.7% egg and 23.5 to 48.5% larval death within 24 hours of application at several concentrations [36,83,84].

An experiment was carried out in Egypt where Neem oil (Azadirachtin) showed the highest mortality (92%) on *T. absoluta* larva where the combined use of garlic + neem + green miracle and garlic + basil showed 43% and 40% larval mortality, respectively inside the mines [85]. In Tunisia, an experiment was conducted on botanical extracts of Neem oil + Azadiractin named “Biocide” and biological insecticide “Thuricide” based on *B. thuringiensis* var. *kurstaki* (Btk) along with mass trapping significantly reduced the damage of *T. absoluta* [86]. Similarly, in Ethiopia it was reported that *Azadirachta indica*, *Nicotiana spp.*, *A. sativum*, and *Cymbopogon citratus* have shown significant effects on the larvae of *T. absoluta* at 10% concentration after a week of application [75]. Crude extracts of *A. sativum*, *Cymbopogon citrates*, and *A. indica* seed showed 95%, 97%, and 98% larval death, successively after 7 days of application [87]. Abdel-Baky and Al-Sooqer [88] observed that jojoba seed extracts achieved 75% larval death at second instar while applied at 100% concentration. [89] reported that Castor bean extracts and thyme extracts achieved 58% and 95% larval death, respectively. Moreover, extracts from some other plants such as basil, chinaberry, eucalyptus, garlic, geranium, and onion have also been showed insecticidal action into different efficacies to achieve larval death of *T. absoluta* [53,85,90].

### 2.4.4 Chemical control methods

*T. absoluta* can be effectively controlled by the chemical control methods as the main control measure of this pest. However, these control measures using chemical insecticides have significant side effects on beneficial arthropods [27,91,92], negative impacts on both human health and on the environment [93], and quit development of insecticide resistance [28,94]. Moreover, the effectiveness of chemical insecticides to control tomato leaf miner has been poor due to mine-feeding behaviour of its larvae or improper spraying method [51]. To avoid these, synthetic pesticides should not use at the beginning of the cropping season and obviously should avoid when symptoms of *T. absoluta* are not present. By using chemical insecticides with a minimal dose, low infestations by *T. absoluta* should be controlled. Insecticides should be selected carefully in the early stage of the crop growth. Several active ingredients should use alternately and not combined all at once so that this will not allow the pest to develop resistance in a shorter lifetime.

Several chemicals have been used in different parts of the world to minimize the infestation of *T. absoluta*. Synthetic pesticides like Organophosphates and Pyrethroids have been used in the 1970s and abamectin, Chlorfenpyr, Tebufonzide, and Spinosad have been introduced in the 1990s to control *T. absoluta* in Egypt [95]. In Africa, chemicals like Abamectin, Emamectin benzoate, Organophosphates, Pyrethroids, and Spinosad have been used against *T. absoluta* [22]. In Nepal, Plant...
Protection Directorate has recommended to spray, Chlorantraniliprole (Trade name: Coragen, Alcora), Novaluron (Trade Name: Remon, Pedestral, Remo 10) and Spinosad (Trade name: Tracer) at 1 ml per liter of water, 3 ml per 10 liter of water and 1 ml per 3 liter of water, respectively against T. absoluta [41]. In Central-East Tunisian a research was conducted where insecticide indoxacarb (50 cc/hl) controlled T. absoluta larvae (with more than 95% efficacy nine days after insecticides application) successfully compared to diafenthion (125 cc/hl) and triflumuron (50 cc/hl) in greenhouse conditions [46].

On the other hand, in Northeastern Tunisian tomato greenhouses, insecticides flubendiamid (30 g/ml) and cyromazine (30 g/ml) attained 77% and 96% larval mortality of T. absoluta after three weeks of treatment, while the efficacy of azadirachtin is not more than 40% [96]. Besides, a good performance was observed by applying active ingredients indoxacarb, pyrethroids and spinosad, against T. absoluta larvae under laboratory and field conditions in Tunisia [46].

2.4.5 Sex pheromone-based control strategies

The sex pheromone trap is an authentic technique to investigate the existence of T. absoluta [10,20,28,40,63] and also helps to determine the critical time for pesticide applications leading to optimum use of registered insecticides [43]. Sex pheromones are chemicals released by an organism in the form of fluid which attracts an individual of the same species of the opposite sex that cause sexual excitement. They act as natural sexual attractants [10,40]. T. absoluta sexpheromone bait has been recognized [38] as (3E,8Z,11Z) 3,8,11-tetradecatrienyl acetate as a major and (3E,8Z) tetradedien-1-yl acetate as a minor element using for mating disruption [10,40]. Most of the female sex pheromones have been detected in Lepidopterans which consist of a mixture of two or more components that attract male from long-distances and also express courtship behaviour [40]. Sex pheromone-based control of T. absoluta is an eco-friendly management technique, therefore this technique is more recommended to be used along with other control methods. Sex pheromone trap can be used for pest detection, population monitoring, mass annihilation, and mating disruption [40,63,97]. In South America, Europe, Asia, and North Africa, the sex pheromone trap has been successfully applied in controlling tomato leaf miner on both greenhouses and open fields. In Egypt, sex-pheromone trap showed a significant result when integrated with other pesticides for controlling the infestation of tomato by T. absoluta [98].

2.4.6 Development of Tuta absoluta Resistance tomato varieties

Selecting tomato varieties that resistant to insect pests are one of the most important management practices that help to protect the tomato plants from infestation by these pests. Recently, breeding programs for the development of insect resistance tomato varieties are gaining more attention in many parts of the world for cost-effective pest control methods [57,99,100]. Generally, wild relatives of genus Solanum contain insect registrant genes. The presence of glandular trichome and allelochemicals are linked with insect-resistant [101]. Trichomes secrete insecticidal compounds that are effective against larvae of T. absoluta [102]. Recently, several countries are focusing their research on the development of insect resistance tomato varieties by incorporating alleles responsible for resistance to insect existing in wild species into popular cultivated varieties to produce the alleles linked with resistance [103-106].

Three types of allelochemicals (acyl-sugars, zingiberene and 2-tridecanone) which was linked with resistant to T. absoluta [107,108]. These allelochemicals function as a toxicant, antinutritional, herbal and medicinal for pest and disease resistant factors [109]. Glandular trichome density and allelochemicals have been linked with conferring resistance to T. absoluta and other insect pests [104,108]. Generally greater resistant showed by those genotypes which have a higher density of glandular trichome [106]. Several researchers have identified these allelochemicals in some wild species of Solanum. Acyl sugars (AA) has detected in S. pennelli [107,109,110] and S. galapagense [112], Zingiberene (ZGB) has detected in S. habrochaites var. hirsutum [113], and 2-tridecanone (2-TD) has detected in S. habrochaites var. glabratum [107,114,115]. The presence of these allelochemicals, the oviposition rate, plant damage severity, injuries to the leaves, and the percentage of affected leaves were reduced significantly [105]. High concentrations of 2-tridecanone, Zingiberene, and Acyl sugars directly associated with the reduction of T. absoluta oviposition rate in tomato varieties [103-105].
Globally total 17 species have been recognized as wild relatives of the genus Solanum; S. cheesmaniae, S. galapagense, S. chilense, S. chmielewskii, S. lycopersicum, S. habrochaites, S. neoickii, S. pennelli, S. arcanum, S. corneliomulleri, S. huylasense, S. peruvianum, S. pimpinellifoillum, S. juglandifolium, S. lycopersicoideas, S. ochranthum, and S. sitiens [116,117]. Among these, six wild species of Solanum, S. Arcanum, S. chilense, S. corneliomulleri, S. lycopersicum, and S. pennellii which are resistance to T. absoluta [118]. These genotypes showed significant larval death and continuing larval and pupal lifespan which led to reduce adult emergence of T. absoluta. On the other hand, S. pennelli and S. habrochaites showed outstanding resistant against T. absoluta [119]. Vitta et al. [120] also detected a wild species S. habrochaites as highly resistant to T. absoluta. Moreover, it will be very helpful to conduct further breeding programs for the development of T. absoluta resistant tomato varieties by utilizing glandular trichome and allelochemicals existing in different wild species of genus Solanum.

2.4.7 Integrated pest management (IPM)

The IPM is an efficient and eco-friendly sensitive approach for the control of pests that depends on combinations of all available pest control methods [28,53,121]. To control T. absoluta successfully the use of Integrated Pest Management (IPM) strategy might be very effective [37]. The integration of physical, cultural (crop rotation, changing planting/harvest dates to avoid pest's infestation, elimination and burning of infected plant parts) [53], biological agents such as predators, parasitoids, entomopathogenic microbes [48,122], biopesticides [10,18], and cultivation of T. absoluta resistant tomato varieties [47] can be develop an effective IPM [61]. The combination of these pest control methods with each other with appropriate use of registered insecticides [54,121] that are less hazardous to the environment could be effective for controlling T. absoluta without hampered the agro-ecosystems.

3. CONCLUSION AND RECOMMENDATIONS

Tomato is one of the most important and popular vegetable crops in Bangladesh which is grown on both open fields and greenhouses. However, recently, the average yield of tomato has been decreasing due to several factors including diseases and pests. T. absoluta has recently been introduced as a devastating insect pest in Bangladesh which has become a major economic pest causing yield losses of up to 100% in tomato production. Therefore, the havoc caused by this pest demands urgent action while the pest’s status has been increasing all over the world as it spreads into new belts over the last decade. Currently, Tuta Trap Tray and Tray Trap Technology (TTT) control measures have been shown more effective against T. absoluta as a single control method compare to other control methods. Moreover, the continuous application of chemical pesticides to control T. absoluta has led to the development of resistance against pesticides, disturbing the ecological and biological world. Therefore, to control this pest effectively, it is important to integrate all available control methods include physical, cultural, biological methods, and optimum use of registered pesticides. Besides these, there is need for multidisciplinary endeavour involving research scientists to detect genetic mechanisms and strategies that will be helpful to pause further colonization of T. absoluta in Bangladesh. Moreover, policymakers should establish appropriate policies; agricultural extension workers should communicate with farmers to provide successful agricultural solutions; to train up extension workers and farmers using accurate measures to control T. absoluta; and farmers to apply innovations for sustainable tomato production in this country. Moreover, the proper utilization of conventional breeding and molecular approaches to develop T. absoluta resistant tomato varieties could be the best sustainable control strategy. Finally, there is need for more collaborative efforts among the research scientists from public and private sectors as well as tomato growers to control T. absoluta in Bangladesh.

ACKNOWLEDGEMENTS

Authors would like to express gratitude to the Department of Entomology, Haje Mohammad Danesh Science and Technology University, Dinajpur, Bangladesh for their collaboration toward achieve this work. Special thanks to Dr. Md. Adnan Al Bachchu and Dr. Md. Mominur Rahman for their inspirations to do this review work.

COMPETING INTERESTS

Authors have declared that no competing interests exist.
REFERENCES

1. Kavanaugh CJ, Trumbo PR, Ellwood KC. The US food and drug administration’s evidence-based review for qualified health claims: tomatoes, lycopene, and cancer. Journal of the National Cancer Institute. 2007;99(14):1074-1085.

2. Rouphael Y, Schwarz D, Krumbein A, Colla G. Impact of grafting on product quality of fruit vegetables. Scientia Horticulturae. 2010;127:172-179.

3. Jenkins JA. The origin of cultivated tomato. Economic Botany. 1948;2(4):379-392.

4. FAO. Food and Agriculture Organization of the United Nations. Faostat. Production/yield quantities of Tomatoes in World + (Total); 2018. (Accessed 6 February 2020) Available:http://www.fao.org/faostat/en/#data/QC/visualize.

5. BBS. Yearbook of Agricultural statistics of Bangladesh. Bangladesh Bureau of Statistics, Statistics and Informatics Division (SID), Ministry of Planning, Government of the People’s Republic of Bangladesh, Dhaka. 2017:301-302.

6. Materu CL, Shao EA, Losujaki E, Chidege M, Mwambela N. Farmer’s perception knowledge and practices on management of Tuta absoluta Meyerick (Lepidoptera Gelechiidae) in Tomato growing areas in Tanzania. International Journal of Research in Agriculture and Forestry. 2016;3(2):1-5.

7. Chidege M, Al-zaidi S, Hassan N, Julie A, Kaaya M, Mrogoro S. First record of tomato leaf miner Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) in Tanzania. Agriculture & Food Security. 2016;5:17.

8. Assaf LH, Hassan FR, Ismael HR, Saeed SA. Population density of tomato leaf miner Tuta absoluta Meyrick (Lepidoptera: Gelechiidae) under plastic houses conditions (b). IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS). 2013; 5(4):7-10.

9. Omond J. Effects of insecticide treated nets in the management of tomato pests and their impact on natural enemies and yield in Nairobi and Murang’a counties (Master’s thesis). The School of Pure and Applied Sciences of Kenyatta University, Kenya; 2015.

10. Illakwahhi DT, Srivastava BBL. Control and management of tomato Leafminer-Tuta Absoluta (Meyrick) (Lepidoptera, Gelechiidae). A review. IOSR Journal of Applied Chemistry (IOSR-JAC). 2017; 10(6):14-22.

11. Shiberu T, Getu E. Evaluation of biopesticides on integrated management of tomato Leafminer, Tuta absoluta (Meyrick) (Gelechiidae: Lepidoptera) on Tomato Crops in Western Shewa of Central Ethiopia. Journal of Entomology, Ornithology & Herpetology. 2018;6(4):206. DOI:10.4172/2161-0983.1000206

12. Zappala L, Biondi A, Alma A, Al-Jboory J, Arno J, BayramA.et al. Natural enemies of the South American moth, Tuta absoluta, in Europe, North Africa and Middle East, and their potential use in pest control strategies. Journal of Pest Science. 2013;86(4):635-647.

13. Tonnang HEZ, Mohamed SA, Khamis F, Ekies S. Identification and risk assessment for worldwide invasion and spread of Tuta absoluta with a focus on Sub-Saharan Africa: Implications for Phytosanitary measures and management. Plos one. 2015;10(9):e0138319. DOI:10.1371/journal.pone.0135283

14. Biondi A, Guedes R, Wan F. Ecology, worldwide spread, and management of the invasive South American tomato pinworm, Tuta absoluta: Past, present, and future. Annual Review of Entomology. 2018;63: 239-258.

15. Bajracharya AS, Bhat B, Mainali RP, Bista S, Shashank PR, Meshram NM. The first record of South American tomato leaf miner, Tuta absoluta (Meyrick, 1917) (Lepidoptera: Gelechiidae) in Nepal. Journal of Entomology and Zoology Studies. 2016; 4(4):1359-1363.

16. Uulu TE, Ulusoy MR, Caliskan AF. First record of tomato leafminer Tuta absoluta Meyrick (Lepidoptera: Gelechiidae) in Kyrgyzstan. Bulletin OEPP/EPPO Bulletin. 2017;47(2):285-287.

17. Saidov N, Srinivasan R, Mavlyanova R, Qurbanov Z. First report of invasive South American tomato leaf miner Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) in Tajikistan. Florida Entomologist. 2018; 101(1):147-149.

18. Muniappan R. Tuta absoluta: The tomato leafminer. Paper presented in the international workshop in Nepal. Asian Vegetable and Mango Integrated Pest Management Innovation Lab (AVMIPM-IL), IDE Nepal, 2016.
19. Zekeya N, Chacha M, Ndakidemi PA, Materu C, Chidege M, Mbega ER. Tomato leaf miner (Tuta absoluta Meyrick 1917): A threat to tomato production in Africa. Journal of Agriculture and Ecology Research International. 2017b;10(1):1-10.

20. Ghoneim K. Parasitic insects and mites as potential biocontrol agents for a devastative pest of tomato, Tuta absoluta Meyrick (Lepidoptera: Gelechiidae) in the world: A review. International Journal of Advanced Research. 2014;2(8):81-115.

21. Guillemaud T, Blin A, Le Goffl, Desneux N, Reyes M, Tabone E, et al. The tomato borer, Tuta absoluta, invading the Mediterranean Basin, originates from a single introduction from Central Chile. Scientific Report. 2015;5:8371. DOI: 10.1038/srep08371.

22. Zekeya N, Ndakidemi PA, Chacha M, Mbega E. Tomato leaf miner, Tuta absoluta (Meyrick, 1917), an emerging agricultural pest in Sub-Saharan Africa: Current and prospective management strategies. African Journal of Agricultural Research. 2017a; 12(6):389-396.

23. Desneux N, Luna MG, Guillemaud T, Urbaneja A. The invasive South American tomato pinworm, Tuta Absoluta, continues to spread in Afro-Eurasia and beyond: The new threat to tomato world production. Journal of Pest Science. 2011;84(4):403-408.

24. Ogor E, Unlu L, Karaca M. Chenopodium album L.: A new host plant of Tuta absoluta povolny (Lepidoptera: Gelechiidae). Turkish Bulletin of Entomology. 2014;4(1):61-65.

25. Mansour R, Brevault T, Chailleux A, Cherif A, Grissa-Lebdik, Haddik, et al. Occurrence, biology, natural enemies and management of Tuta absoluta in Africa. Entomologia Generalis. 2018;38(2):83-112.

26. Han P, Bayram Y, Shaitel-Harpaz L, Sohrabi F, Saji A, Esenali UT, et al. Tuta absoluta continues to disperse in Asia: damage, ongoing management and future challenges. Journal of Pest Science. 2019;92(4):1317-1327.

27. Ontijo GC, Picanço MC, Pereira EJG, Martins JC, Chediak M, Guedes RNC. Spatial and temporal variation in the control failure likelihood of the tomato leaf miner, Tuta absoluta. Annals of Applied Biology. 2013;162:50-59.

28. Retta AN, Berhe DH. Tomato leaf miner–Tuta absoluta (Meyrick), a devastating pest of tomatoes in the highlands of Northern Ethiopia: A call for attention and action. Research Journal of Agricultural and Environmental Management. 2015;4(6):264-269.

29. CABI. Invasive species compendium: Tomato leaf miner (Tuta absoluta); 2016. Available: http://www.cabi.org/isc/datasheet/49260.

30. Campos MR, Biondi A, Adiga A, Guedes RNC, Desneux N. From the Western Palaearctic region to beyond: Tuta absoluta 10 years after invading Europe. Journal of Pest Science. 2017;90(3):787-796.

31. Ndor DC. Incidence of Tomato leaf miner (Tuta absoluta Meyrick) damage on Tomato fields in Pankshin and Kanke Local Government Areas of Plateau State. Agricultural Science Research Journal. 2018;8(1):15-19.

32. EPPO. EPPO Global database (available online). Paris, France; 2018. Available: https://gd.eppo.int/taxon/GNORA B.

33. Sankarganesh E, Firake DM, Sharma B, Verma VK, Behere GT. Invasion of the South American Tomato Pinworm, Tuta absoluta, in northeastern India: a new challenge and biosecurity concerns. Entomologia Generalis. 2017;36(4):335-345.

34. Hossain MS, Mian MY, Muniappan R. First Record of Tuta absoluta (Lepidoptera: Gelechiidae) from Bangladesh. Journal of agricultural and urban entomology. 2016;32(1):101-105.

35. Birgucu AK, Bayindir A, Celikpence Y, Karacal. Growth inhibitory effects of bio- and synthetic insecticides on Tuta absoluta (Meyrick, 1917) (Lepidoptera: Gelechiidae). Turkish Journal of Entomology. 2014;38(4):389-400.

36. Joshi A, Thapa RB, Kalauni D. Integrated management of South American tomato leaf miner [Tuta absoluta (meyrick)]: a review. Journal of the Plant Protection Society. 2018;5:70-86.

37. Miranda MMM, Picanco MC, Zanuncio JC, Bacci L, da Silva EM. Impact of integrated pest management on the population of leafminers, fruit borers, and natural enemies in tomato. Ciencia Rural. 2005;35(1):204-208.
38. EPPO. Tuta absoluta Found on Phaseolus vulgaris in Sicilia (IT). EPPO Reporting Service. 2009;8(154):3.

39. Harizanov V, Stoeva A, Mohamedova M. Tomato Leaf Miner, Tuta absoluta (Povolny) (Lepidoptera: gelechiidae)—First Record in Bulgaria. Agricultural Science and Technology. 2009;1(3):95-98.

40. Megido RC, Haibrugge E, Verheggen FJ. Pheromone-based management strategies to control the tomato leafminer, Tuta absoluta (Lepidoptera: Gelechiidae). A review. Biotechnology, Agronomy, Society and Environment. 2013;17(3):475-482.

41. PPD. Tomato leaf miner; Tuta absoluta (Meyrick). Retrieved from Government of Nepal, Plant Protection Directorate; 2017. Available:http://ppdnepal.gov.np.

42. Barrientos ZR, Apablanza HJ, Norero SA, Estay PP. Threshold temperature and thermal constant for development of the South American tomato moth, Tuta absoluta (Lepidoptera, Gelechiidae). Ciencia e Investigacion Agraria. 1998;25(3):133-137.

43. Berxolli A, Shahini S. Population dynamic of tomato leaf miner, Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae). Albanian Journal of Agricultural Sciences (Special Edition). 2017:85-89.

44. Ballal CR, Gupta A, Mohan M, Lalitha Y, Verghese A. The new invasive pest Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) in India and its natural enemies alongwith evaluation of Trichogrammatids for its biological control. Current Science. 2016;110(11):2155-2159.

45. Desneux N, Wajnberg E, Wyckhuys KAG, Burgio G, Arpaia S, Narváez-Vasquez CA. et al. Biological Invasion of European Tomato Crops by Tuta Absoluta: Ecology, Geographic Expansion and Prospects for Biological Control. Journal of Pest Science. 2010;83(3):197-215.

46. Braham M, Hajji L. Management of Tuta absoluta (Lepidoptera, Gelechiidae) with insecticides on tomatoes. Insecticides—Pest Engineering. 2012:333-354.

47. Sanda NB, Sunusi M, Hamisu HS, Wudil BS, Sule H, Abdullahi AM. Biological invasion of tomato leaf miner, Tuta absoluta (Meyrick) in Nigeria: Problems and management strategies optimization: A review. Asian Journal of Agricultural and Horticultural Research. 2018;1(4):1-14.

48. Urbanuja A, Desneux N, Gabarra R, Amo J, Gonzalez-Cabrera J, Mafra Neto A.et al. Biology, ecology and management of the South American tomato pinworm, Tuta absoluta. In: Pena JE, editor. Potential invasive pests of agricultural crops. CABI, Wallingford, UK; 2013:98-125.

49. Rwomushana I, Beale T, Chipabika G, Day R, Gonzalez-Moreno P, Lamontagne-Godwin J. et al. Evidence Note. Tomato leafminer (Tuta absoluta): impacts and coping strategies for Africa. CABI Working Paper 12. 2019:1-56. Available:https://dx.doi.org/10.1079/CABICOMM62-8100.

50. Borisade OA, Kolawole AO, Adebo GM, Uwaidem YI. The tomato leafminer (Tuta absoluta) (Lepidoptera: Gelechiidae) attack in Nigeria: Effect of climate change on over-sighted pest or agro-biotorerism? Journal of Agricultural Extension and Rural Development. 2017;9(8):163-171.

51. Cocco A, Deliperi S, Delrio G. Control of Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) in greenhouse tomato crops using the mating disruption technique. Journal of Applied Entomology. 2012;137:16-28.

52. Shehata I, Ebada I, Ismaill A, Fouda M, Salama SH. On the population dynamics of the tomato leaf miner Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) in Egypt. Ecologia Balkanica. 2016;8(2):65-75.

53. Aynalem B. Tomato leafminer [(Tuta absoluta Meyrick) (Lepidoptera: Gelechiidae)] and its current ecofriendly management strategies: A review. Journal of Agricultural Biotechnology and Sustainable Development. 2018;10(2):11-24. DOI:10.5897/JABSD2018.0306

54. Gozel C, KasapI. Efficacy of entomopathogenic nematodes against the Tomato leafminer, Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) in tomato field. Turkish Journal of Entomology. 2015;39(3):229-237.

55. Oke A, Awe SA. XXV International Congress of Entomology. Entomological Society of America. Orlando, Florida, USA. 2018;25-30.

56. Vincent C, Weintraub P, Hallman G. Physical Control of Insect Pests. In: Encyclopedia of Insects. 2nd ed. Elsevier B.V; 2009:794-798. Available:https://doi.org/10.1016/B978-0-12-374144-8.00209-5
57. All J. Cultural Control of Insect Pests. In: Encyclopedia of Entomology. Springer, Dordrecht; 2005. Available:https://doi.org/10.1007/0-306-48380-7_1108

58. Markovi D. Crop diversification affects biological pest control Intercropping influence pest control. Agroznajance. 2013; 14(2):449-459.

59. Mohamed ESI, Mahmoud MEE, Elhaj MAM, Mohamed SA, Ekesi S. Host plant records of tomato leaf miner Tuta absoluta (Meyrick) in Sudan. Bulletin OEPP/EPPO Bulletin. 2015;45(1):108-111.

60. Balzan M, Moonen A. Management strategies for the control of Tuta absoluta (Lepidoptera: Gelechiidae) damage in open field cultivations of processing tomato in Tuscany (Italy). Bulletin OEPP/EPPO Bulletin. 2012;42(2):217-225.

61. Giorgini M, Guerrieri E, Cascone P,Gontijo C, Rodriguez D. et al. Managing the Neotropical tomato Pest Tuta absoluta (Meyrick) in the Mediterranean Basin. Neotropical Entomology. 2019;48(1): 1-17.

62. Arno J, Sorribas R, Prat M, Matas M, Pozo C, Rodriguez D. et al. Tuta absoluta, a new pest in IPM tomatoes in the Northeast of Spain. IOBC/WPRS Bulletin. 2009;49:203-208.

63. Refki E, Sadok BM, Ali BB. Study of the biotic potential of indigenous predator Nesidiocoris tenuis on Tuta absoluta pest of geothermal culture in the south of Tunisia. Journal of Entomology and Zoology Studies. 2016;4(6):692-695.

64. Sohrabi F, Hosseini R. Nesidiocoris tenuis (Reuter) (Heteroptera: Miridae), a predatory species of the tomato leafminer, Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) in Iran. Journal of Plant Protection Research. 2015;55(3):222-223.

65. Guenauoi Y, Dehlizb A. Natural enemies of Tuta absoluta (Lepidoptera: Gelechiidae) in Oued Righ Region, an Arid Area of Algeria. Academic Journal of Entomology. 2015; 8(2):72-79.

66. Molla O, Monton H, Vanaclocha P, Belitia F, Urbanseja A. Predation by the mirids Nesidiocoris tenuis and Macrolopus pygmaeus on the tomato borer Tuta absoluta. IOBC/WPRS Bulletin. 2009;49: 209-214.

67. Gabarra R, Arno J. Results of the biological control experiences of tomato moth in greenhouse and open air cultivation in Catalonia. Phytoma Espana. 2010;217:66-68.Spanish.

68. Kececi M, Oztop A. Possibilities for biological control of Tuta absoluta (Meyrick, 1917) (Lepidoptera: Gelechiidae) in the western Mediterranean Region of Turkey. Turkish Journal of Entomology. 2017; 41(2):219-230.

69. Sanchez JA, La-spina M, Lacasa A. Numerical response of Nesidiocoris tenuis (Homoptera: Miridae) preying on Tuta absoluta (Lepidoptera: Gelechiidae) in tomato crops. European Journal of Entomology. 2014;111(3):387-395.

70. Chhetri LB. Tomato Leaf miner (Tuta absoluta) an emerging agricultural pest: Control and management strategies: A Review. World Scientific News. 2018;114: 30-43.

71. Cabello T, Gamez M, Varga Z, Garay J, Carreño R, Gallego JR. et al. Selection of Trichogramma spp. (Hymenoptera: Trichogrammatidae) for the biological control of Tuta absoluta (Lepidoptera: Gelechiidae) in greenhouses by an entomo-ecological simulation model. Integrated Control in Protected Crops, Mediterranean Climate. IOBC/WPRS Bulletin. 2012;80: 171-176.

72. Lewis WJ, Vet LEM, Tumlimson JH, Van Lenteren JC, Papaj DR. Variations in parasitoid foraging behavior: Essential element of a sound biological control theory. Environmental Entomology. 1990;19(5): 1183-1193.

73. Zouba A, Chermiti B, Chraiet R, Mahjoubi K. Effect of two indigenous Trichogramma species on the infestation level by tomato miner, Tuta absoluta, in tomato greenhouses in the south-west of Tunisia. Tunisian Journal of Plant Protection. 2013; 8(2):87-106.

74. Chermiti B, Abbes K. Newly adapted parasitoids of the tomato leaf miner Tuta absoluta (Lepidoptera: Gelechiidae) in Tunisia: An ally in organic tomato crops. In: Kheder MB, Neuhoff D, editors. Book of abstracts crop protection management in Mediterranean Organic Agriculture. ISOFAR /MOAN /CTAB Symposium, Sousse (Tunisia); 2013.

75. Shiberu T, Getu E. Effects of crude extracts of medicinal plants in the management of Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) under laboratory and glasshouse conditions in
76. Lakhdari W, Dehliz A, Acheuk F, Milk R, Hammi H, Matallah S et al. Biocontrol test against the leaf miner of tomato *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) by using entomopathogenic fungi in the Algerian Sahara. Journal Algérien des Régions Arides (JARA). 2016; 13(1):85-89.

77. Turkoz S, Kaskavalci G. Determination of the efficacy of some entomopathogenic nematodes against *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) under laboratory conditions. Turkish Journal of Entomology. 2016; 40(2):175-183.

78. Mahmoud MF. Biology and Use of Entomopathogenic Nematodes in Insect Pests Biocontrol, A Generic View. Cercetari Agronomice in Moldova. 2016; 4(168):85-105.

79. Youssef NA, Hassan GM. Bioinsecticide activity of *Bacillus thuringiensis* isolates on tomato borer, *Tuta absoluta* (Meyrick) and their molecular identification. African Journal of Biotechnology. 2013; 12(23):3699-3709.

80. Isman MB, Seffrin R. Natural Insecticides from the Annonaceae: A Unique Example for Developing Biopesticides. In: Singh D, editor. Advances in Plant Biopesticides. Springer, New Delhi. 2014; 21-33.

81. Moreno SC, Carvalho GA, Piccano MC, Morais EG, Pereira RM. Bioactivity of compounds from *Acmella oleracea* against *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) and selectivity to two non-target species. Pest Management Science. 2011; 68(3):386-393.

82. Kona NEM, Taha AK, Mahmoud MEE. Effects of botanical extracts of Neem (*Azadirachta indica*) and Jatropha (*Jatropha curcus*) on Eggs and Larvae of Tomato Leaf Miner, *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae). Persian Gulf Crop Protection. 2014; 3(3):41-46.

83. Yankova V, Valchev N, Markova D. Effectiveness of phytopesticide neem azal T/S® against tomato leaf miner (*Tuta absoluta* Meyrick) in greenhouse tomato. Bulgarian Journal of Agricultural Science. 2014; 20(5):1116-1118.

84. Gebremariam G. *Tuta absoluta*: A global looming challenge in tomato production, Review Paper. Journal of Biology, Agriculture and Healthcare. 2015; 5(14):57-62.

85. Abd El-Ghany NM, Abdel-Razek AS, Ebadah IMA, Mahmoud YA. Evaluation of some microbial agents, natural and chemical compounds for controlling tomato leaf miner, *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae). Journal of Plant Protection Research. 2016; 56(4):372-379.

86. Harbi A, Abbes K, Karboul H, Yousfi S, Chermiti B. Can biological and botanical insecticides combined with mass trapping control the tomato leaf miner *Tuta absoluta* in tomato crops in Tunisia? In: Kheder MB, Neuhoff D, editors. Book of abstracts crop protection management in Mediterranean Organic Agriculture. ISOFA /MOAN /CTAB Symposium, Sousse (Tunisia). 2013: 54.

87. Shiberu T, Getu E. Estimate of yield losses due to *Tuta absoluta* Meyrick (Lepidoptera: Gelechiidae) on tomato crops under glasshouse and field conditions in Western Shewa of Central Ethiopia. International Journal of Fauna and Biological Studies. 2017a; 4(5):104-108.

88. Abdel-Baky FN, Al-Sooqer AA. Controlling the 2nd instars larvae of *Tuta absoluta* Meyrick (Lepidoptera: Gelechiidae) by simmondsin extracted from Jojoba seeds in KSA. Journal of Entomology. 2017; 14(2): 73-80.

89. Nilahyane A, Bouharroud R, Hormatallah A, Taadaouit NA. Larvicidal effect of plant extracts on *Tuta absoluta* (Lepidoptera: Gelechiidae). Integrated Control in Protected Crops, Mediterranean Climate. IOBC-WPRS Bulletin. 2012; 80:305-310.

90. Ghanim NM, Ghanis SBA. Controlling *Tuta absoluta* (Lepidoptera: Gelechiidae) and *Aphis gossypii* (Homoptera: Aphididae) by aqueous plant extracts. Life Science Journal. 2014; 11(3):299-307.

91. Han P, Niu CY, Lei CL, Cui JJ, Desneux N. Use of an innovative T-tube maze assay and the proboscis extension response assay to assess sublethal effects of GM products and pesticides on learning capacity of the honey bee *Apis mellifera* L. Ecotoxicology. 2010; 19(8):1612-1619.

92. Perez-Agulgar DA, Soares MA, Passos LC, Martinez AM, Pineda S, Carvalho GA. Lethal and sublethal effects of insecticides on *Engyptatus varians* (Heteroptera: Miridae), a predator of *Tuta absoluta*.
involved in its resistance to *T. typicum* and possible compounds RNC. Variability of *Lycopersicon hirsutum* Ecole CC, Picanco M, Jham GN, Guedes RN. *The Plant Journal.* 2012;70(1):51

93. Abdel-Raheem A, Ismail A, Abdel-Rahman S, Abdel-Rhman E, Naglaa F. Efficacy of three entomopathogenic fungi on tomato leaf miner, *Tuta absoluta* in tomato crop in Egypt. *Swift Journal of Agricultural Research.* 2015;1(2):15-21.

94. Roditakis E, Vasakis E, Garcia-Vida IL, Martínez-Aguirre MDR, Rison JL, Haxaire-Lutun MO. et al. A four-year survey on insecticide resistance and likelihood of chemical control failure for tomato leaf miner *Tuta absoluta* in the European/Asian region. *Journal of Pest Science.* 2018;91(1):421-435.

95. Abdelgaleil SA, El-bakary AS, Shawir MS, Ramadan GR. Efficacy of various insecticides against tomato leaf miner, *Tuta absoluta*, in Egypt. *Applied Biological Research.* 2015;17(3):297-301.

96. Cherif A, Harbaoui K, Zappalà L, Grissa-Lebdi K. Efficacy of mass trapping and insecticides to control *Tuta absoluta* in Tunisia. *Journal of Plant Diseases and Protection.* 2018;125(1):51-61.

97. Abbès K, Chermiti B. Comparison of two Marks of Sex Pheromone Dispensers Commercialized in Tunisia for their Efficiency to Monitor and to Control by Mass-Trapping *Tuta absoluta* under Greenhouses, *Tunisian Journal of Plant Protection.* 2011;6(2):133-148.

98. El-aassar MR, Soliman MHA, Elaal AAA. Efficiency of sex pheromone traps and some bio and chemical insecticides against tomato borer larvae, *Tuta absoluta* (Meyrick) and estimate the damages of leaves and fruit tomato plant. *Annals of Agricultural Science.* 2015;60(1):153-156.

99. Proffit M, Birgersson G, Bengtsson M, Reis R, Witzgall P, Lima E. Attraction and oviposition of *Tuta absoluta* females in response to tomato leaf volatiles. *Journal of Chemical Ecology.* 2011;37(6):565-574.

100. Portakaldali M, Oztemiz S, Kutuk H. A new host plant for *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae) in Turkey. *Journal of the Entomological Research Society.* 2013;15(3):21-24.

101. Tissier A. Glandular trichomes: what comes after expressed sequence tags? *The Plant Journal.* 2012;70(1):51-68.

102. Ecole CC, Picanco M, Jham GN, Guedes RN. Variability of *Lycopersicon hirsutum* f. typicum and possible compounds involved in its resistance to *Tuta absoluta.* *Agricultural and Forest Entomology.* 2001;1(4):249-254.

103. Goncalves Neto AC, Silva VF, Maluf WR, Maciel GM, Nizio DAC, Gomes LA et al. Resistance to the South American tomato pinworm in tomato plants with high foliar acylsugar contents. *Horticultura Brasileira.* 2010;28(2):203-208. Portuguese.

104. Maluf WR, Maciel GM, Gomes LA et al. Broad-spectrum arthropod resistance in hybrids between high- and low-acylsugar tomato lines. *Crop Science.* 2010;50(2):439-450.

105. Silva GA, Picanco MC, Bacci L, Crespo AL, Rosado JF, Guedes RN. Control Failure Likelihood and Spatial Dependence of Insecticide Resistance in the Tomato Pinworm, *Tuta absoluta.* *Pest Management Science.* 2011;67(8):913-920.

106. Oliveira CMd, Andrade Júnior VCd, Maluf WR, Neiva IP, Maciel GM. Resistance of tomato strains to the moth *Tuta Absoluta* impaired by allelochemicals and trichome density. *Ciencia e Agrotecnologia.* 2012;36(1):45-52.

107. Maluf WR, Barbosa LV, Santa-CeciliaL VC. 2-Tridecanone-mediated mechanisms of resistance to the South American tomato pinworm *Scrobipalpuloides absoluta* (Meyrick, 1917) (Lepidoptera: Gelechiidae) in *Lycopersicon* spp. *Euphytica.* 1997;93(2):189-194.

108. Resende JTVd, Maluf WR, Faria MV, Pfann AZ, Nascimento IRd. Acylsugars in tomato leaflets confer resistance to the South American tomato pinworm, *Tuta absoluta* Meyr. *Scientia Agricola.* 2006;63(1):20-25. DOI:10.1590/S0103-901306000100004

109. Zeist AR, Silva AAda, Resende JTV de, Maluf WR, Gabriel A, Zanin DS. et al. Tomato Breeding for Insect-Pest resistance. In: Nyaku ST, Danquah A, editors. Recent Advances in Tomato Breeding and Production. IntechOpen, London, United Kingdom. 2018:27-46.

110. Goffreda JC, Mutschler MA. Inheritance of potato aphid resistance in hybrids between *Lycopersicon esculentum* and *pennellii.* *Theoretical and Applied Genetics.* 1989;78(2):210-216.

111. Dias DM, Resende JTVd, Marodin JC, Matos R, Lustosa IF, Resende NCV. Acyl sugars and whitefly (Bemisia tabaci) resistance in segregating populations of...
tomato genotypes. Genetics and Molecular Research. 2016;16(1):1-11.

112. Silva AA, Andrade MC, Carvalho RC, Neiva IP, Santos DC, Maluf WR. Resistance to Helicoverpa armigera in tomato genotypes obtained from the crossing of Solanum lycopersicum x Solanum galapagense. Pesquisa Agropecuária Brasileira. 2016;51(7):801-808. Portuguese.

113. Carter CD, Sacalis JN, Gianfagna TJ. Zingiberene and resistance to Colorado potato beetle in Lycopersicon hirsutum f. hirsutum. Journal of Agricultural and Food Chemistry. 1989;37(1):206-210.

114. Williams WG, Kennedy GG, Yamamoto RT, Thacker JD, Bordner J. Tridecanone—a naturally occurring insecticide from the wild tomato Lycopersicon hirsutum f. glabratum. Science. 1980;207(4433):888-889.

115. Goncalves-Gervasio RCR, Ciociola AI, Santa-Cecilia LVC, Maluf WR. Biological aspects of Tuta Absoluta (Meyrick, 1917) (Lepidoptera: Gelechiidae) in two contrasting tomato genotypes regarding 2-tridecanone content in leaflets. Ciencia e Agrotecnologia. 1999;23(2):247-251. Portuguese.

116. Peralta IE, Knapp S, Spooner DM. New Species of Wild Tomatoes (Solanum Section Lycopersicon: Solanaceae) from Northern Peru. Systematic Botany. 2005; 30(2):424-434.

117. Peralta IE, Spooner DM, Knapp S. Taxonomy of wild tomatoes and their relatives (solanum sect. lycopersicoides, sect. juglandifolia, sect. lycopersicon; solanaceae). American Society of Plant Taxonomists, United States. 2008;84:1-186.

118. Sridhar V, Sadashiva AT, Rao VK, Swathi P, Gadad HS. Trichome and biochemical basis of resistance against Tuta absoluta in tomato genotypes. Plant Genetic Resources. 2019;17(3):301-305.

119. Bitew MK. Significant role of wild genotypes of tomato trichomes for Tuta Absoluta Resistance. Journal of Plant Breeding and Genetics. 2018;2(1):1-12.

120. Vitta N, Estay P, Chorbadjian RA. Characterization of resistance expression in genotypes of Solanum Section Lycopersicon against Tuta absoluta (Lepidoptera: Gelechiidae), Ciencia e Investigación Agraria. 2016;43(3):366-373.

121. Taha AM, Afsah AFE, Fargalla FH. Evaluation of the effect of integrated control of tomato leafminer Tuta absoluta with sex pheromone and insecticides. Nature and Science. 2013;11(7):26-29.

122. Chailleux A, Bearez P, Pizzo IJ, Amiens-Desneux E, Ramirez-Romero R, Desneux N. Potential for combined use of parasitoids and generalist predators for biological control of the key invasive tomato pests Tuta absoluta. Journal of Pest Science. 2013;86(3):533-541.

© 2020 Huda et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
http://www.sdiarticle4.com/review-history/55950