Effect of AgNPs distribution on the cotton fiber to the durability of antibacterial cotton fabrics

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Abstract

Silver nanoparticles (AgNPs) loaded on the cotton fiber can endow the fiber with good antibacterial activity, but the AgNPs on the surface of cotton fiber will leach out with the increase of washing times, which will not only greatly reduce the antibacterial properties of cotton fiber, but also cause pollution to the environment. In this paper, carboxyl groups were introduced into cotton fiber by three methods, namely selective oxidation, butane tetracarboxylic acid (BTCA) grafting and polyacrylic acid (PAA) adsorption. The silver ions (Ag\(^+\)) were adsorbed on the cotton fibers through electrostatic attraction, and after reduction, the AgNPs were loaded onto the cotton fibers, and the AgNPs distribution on the fiber and its effect to the washing resistance of AgNPs were studied. The results showed that the selective oxidation and the BTCA grafting can introduce carboxylic groups into the interior of cotton fibers while the PAA adsorption only imports carboxyl groups on the surface of the fiber. As a consequence, AgNPs were formed inside of the cotton fiber when the selective oxidation and the BTCA grafting were employed, while AgNPs can only be loaded on the surface of the fiber by the PAA adsorption method. AgNPs inside of the fiber have high energetic adherence, resulting in silver-cotton fabric nanocomposites with strong slow-release property and long-term antibacterial ability. The fabric nanocomposites prepared by selective oxidation and BTCA grafting exhibited high AgNPs retention rate and excellent antibacterial activity even after 80 washing cycles.

1. Introduction

Cotton fiber was widely used in textile printing and dyeing industry due to its excellent water absorption and human affinity. However, cotton fibers can easily breed bacteria in a suitable environment, which not only cause color change, mechanical strength decline and odor generation, but also have a significant impact on people's life and health(Hebeish et al. 2011a; Hebeish et al. 2011b). With the concern on personal health, silver nanoparticles (AgNPs) are often loaded onto cotton fibers due to their superb bacteria inhibition and broad-spectrum antibacterial properties(Ahmad 2021; Gao et al. 2021).

There are many methods to introduce AgNPs onto cotton fibers(Bacciarelli-Ulacha et al. 2014; Nam and Condon 2014; Pan et al. 2018; Ravindra et al. 2010), including directly deposition(Cheng et al. 2013), padding(Lee et al. 2003), layer-by-layer disposition(Xu et al. 2016; Xu et al. 2017), in situ synthesis(Lee et al. 2007; Montes-Hernandez et al. 2021), heat evaporation(Smetana et al. 2005), etc. The antibacterial mechanism of AgNPs is considered to be the silver ions (Ag\(^+\)) gradually release from the surface of AgNPs, which can penetrate into the cell wall and kill the surrounding bacteria(Maneerung et al. 2008). The release of Ag\(^+\) means that the loading of AgNPs on fibers is not permanent, which can result in poor durability against washing processing. In addition, the released AgNPs can accumulate in human tissue and organs and cause adverse effects such as cell death and DNA damage(El-Rafie et al. 2014a; van der Zande et al. 2012). Therefore, the effective immobilization of AgNPs onto the cotton fiber and the developing a fiber with a controlled release of Ag\(^+\) are crucial for its long-term antibacterial activity. Great
efforts have been made by scientists to improve the stabilization of AgNPs on cotton fibers. The most studied method is the application of binders. Wang et al. loaded nano-silver particles on the surface of cotton fiber by covering the surface of cotton fiber with polydopamine/polyethyleneimine coating, and the washing resistance and the slow-release property of AgNPs on the fiber were greatly improved (Liu et al. 2020). Zhou et al. grafted CMC and methacrylate onto the cotton fabric to load a coating on the surface of the cotton fabric, then used the coating to fix the AgNPs onto the surface of the cotton fabric, The stability of AgNPs on the surface of cotton fabric was improved (Zhou et al. 2019). Alternatively, crosslinkable polymers or crosslinking agents have been used to strengthen the binding force between AgNPs and cotton fiber. Kang et al. used 3-mercaptopropyltrimethoxysilane (3-MPTMS) as a crosslinking agent to fix AgNPs and quaternary ammonium salts on the surface of the cotton fabric, and the stability of AgNPs on cotton fiber surface was improved (Kang et al. 2016). In addition, AgNPs can penetrate into the multi-scale micropore of cotton fiber and be stabilized inside the fiber. Nam et al. treated cotton fibers with an alkaline solution to generate nanosized microfibrillar channels, which were then introduced with [Ag(NH$_3$)$_2$]$^+$ ions and reduced to AgNPs. The AgNPs was successfully diffused into the interior of the cotton fiber, and the washing resistance of the fiber was greatly improved (Nam et al. 2020). Montes-Hernandez et al. put cotton fabrics in silver nitrate solution, and the silver ions were reduced to AgNPs, FE-SEM results of fiber cross section show AgNPs diffused the interior of fiber (Montes-Hernandez et al. 2021). However, the durability of AgNPs on cotton fibers is still insufficient and the methods to improve stabilization of AgNPs on the fibers still need to be improved.

Take advantage of the multi-scale microporous structure of cotton fiber, the carboxyl groups were introduced into the fiber in this present work to slow down the leaching rate of Ag$^+$ and enhance the stability of silver nanoparticles on the cotton fiber. To demonstrate the effect of AgNPs location on the fiber to the long-term antibacterial ability of fabric, three methods for introducing carboxyl groups onto cotton fibers including selective oxidation, BTCA grafting and PAA adsorption were designed. The influence of different methods to the washing resistance of cotton fibers loading with AgNPs and the release of Ag$^+$ were studied.

2. Experiment

2.1 Materials

Mercerized cotton fabrics (180.6 g/m$^2$) were obtained from Lianfa Textile Co., Ltd (Jiangsu, China). 2,2,6,6-tetramethylpiperidine oxide (TEMPO, 99.69%) and butane tetracarboxylic acid (BTCA, 99%) were bought from Aladdin (Shanghai, China). Polyacrylic acid (Mw 80000 g/moL, solid content 40%) was purchased from Shanghai Yingxin laboratory equipment Co., Ltd (Shanghai, China). Poly(diallyldimethylammonium chloride) (PDADMAC, Mw 100,000 g/mol), peptone, agar, polybrene (94%) were purchased from Beijing Innochem Technology Co., Ltd (Beijing, China). Silver nitrate (AgNO$_3$, 99.9 %), NaBr (99%), NaOH (98%), hypophosphate (98%) and sodium dichloroisocyanurate (NaDCC, 96%) were purchased from Sinopharm Chemical Reagent Co., Ltd (Shanghai, China).
2.2 Introduction of carboxyl groups onto cotton fiber

During the selective oxidation of cotton fiber, 0.047 g of TEMPO and 0.31 g of NaBr were dissolved in 300 mL of water, and 2 g of cotton fabric was then immersed in the above solution. The oxidation process was maintained at room temperature for 4 h, and the pH of the solution was kept at about 10 through the addition of 1 M NaOH during the reaction. Following, the reaction was quenched by adding ethanol. The oxidized cotton fabric (OCF) was washed with water and then air-dried.

During the cotton fiber grafting by BTCA, 2 g of cotton fabric was immersed in 100 mL of water containing 8 g of BTCA and 3 g of hypophosphate, and then padded for a wet pick up of 70%. The immersed cotton fabric was dried at 80°C for 3 min, baked at 170°C for 3 min, cotton fabric grafted with BTCA (BCF) was washed with water at 90°C and sufficiently dried.

During PAA adsorption method, 2 g of cotton fabric was immersed in 100 mL of water containing 4 g of PAA, and then padded for a wet pick up of 70%. The immersed cotton fabric was dried at 80°C for 3 min, then baked at 170°C for 3 min, cotton fabric with PAA adsorbed (PCF) was washed with water at 90°C and sufficiently dried.

2.3 Preparation of AgNPs-loaded cotton

Three kinds of treated cotton fabric samples (PCF, BCF, OCF) (2 g) and a control cotton fabric (CF) sample (2 g) were immersed in aqueous solution (100 mL) containing AgNO₃ (0.17 g) and the pH was adjusted to 7.2 by adding 0.1 M NaOH solution. Then the solution was kept at 70°C with modest stirring for 50 min. Finally, the fabric samples were separately immersed in a solution (100 mL) containing NaBH₄ (0.5 g) for five min. Next, the fabric samples were washed with deionized water. The AgNPs-loaded cotton fabrics (AgNPs-CF, AgNPs-PCF, AgNPs-BCF, AgNPs-OCF) were obtained after drying in the vacuum oven at 50°C for 60 min.

2.4 Determination of carboxylate and charge contents

0.25 g of fabric sample was immersed in aqueous solution (30 mL) containing HCl (0.3 mmol) for 1 h. The sample was washed with water, then immersed in 50 mL NaOH solution (0.05 mmol/L). Then the solution was added with a drop of phenolphthalein indicator solution, then titrated with 0.05 mol/L HCl solution. The titration stopped when the color of the solution changed from red to colorless. The carboxyl content was calculated according to the following Eq. (1):

\[
\text{Carboxyl content} = \frac{C_{\text{NaOH}} \times V_{\text{NaOH}}}{W_s} \text{ (mmol/g)}
\]  

Where \(C_{\text{NaOH}}\) is the concentration of the NaOH solution (mol/L), \(V_{\text{NaOH}}\) is the volume of the NaOH solution consumed (mL) and \(W_s\) is the weight of the sample used for titration after drying (g).

The charge content of cotton fiber was calculated by polyelectrolytes (PE) titration, which was first proposed by Winter et al. (Fras et al. 2005; Fras et al. 2004; Junka et al. 2013; Zhang et al. 2015). Briefly, two cationic polyelectrolytes, poly(diallyldimethylammonium chloride) (PDADMAC, Mw 100,000 g/mol)
and Polybrene (Mw 374 g/mol) were used for the titrations. 0.3 g cotton fiber was immersed in 50 mL cationic polyelectrolytes solution (0.01 meq/L) for 30 min. The fiber was washed with a large amount of water, the washing liquid was collected and then mixed with the reaction liquid and the volume was adjusted to 500 mL. 20 mL solution was titrated with potassium polyvinyl sulfate titration solution (0.0025 M) and toluidine blue indicator solution. Polybrene was used measure the total charge. PDADMAC was used to analyze the charge on the surface of the fiber. The charge content was calculated from the following Eq. \( (2) \). All results are the mean values of three measurements.

\[
\text{Charge content} = \frac{(C_{\text{cat}} \times V_{\text{cat}} - C_{\text{PVSK}} \times V_{\text{PVSK}})}{W_S} \text{ (mmol/g)} \tag{2}
\]

where \( C_{\text{cat}} \) is the charge concentration of cationic polyelectrolytes solution (mol/L), \( V_{\text{cat}} \) is the volume of cationic polyelectrolytes solution (mL), and \( W_S \) is weight of oven-dried samples used for titration (g).

### 2.5 Washing Experiment

To evaluate the washing resistance of AgNPs-cotton, washing experiments were carried out in AATCC standard washing machine (M228, SDL Atlas) according to the AATCC Test Method 61-2006. A 150mL washing solution containing 0.2% WOB standard detergent and 10 stainless steel balls (diameter 6 mm) were added to the stainless steel canister. The cotton fabric was cut into 5 cm \( \times \) 10 cm samples and washed in the washing machine at 40°C \( \pm \) 1°C for 45 min. This process was equivalent to 5 washing cycles, after 20, 40 and 80 washing cycles, the samples were thoroughly rinsed with water and naturally air-dried.

### 2.6 Characterizations

Morphology of the AgNPs on the surface of bers was observed by a scanning electron microscopy (SEM, TM-1000, Hitachi). Cotton yarns were put into a Hashi slicer, and then were cut into fiber sections with a blade. Morphology and distribution of AgNPs on the fiber cross-sections was observed on a field emission scanning electron microscopy (FE-SEM, S-4800, Hitachi).

In order to characterize the washing durability, the fabrics washed 0, 20, 40 and 80 cycles were characterized using an UV-vis-NIR spectrometer (UV-3600 Plus, Shimadzu). Reflection spectra in the wavelength range from 350 nm to 800 nm were recorded. The silver retention rate of cotton fabrics after 0, 40 and 80 washing cycles were measured by an atomic emission spectrometer (Optima 2100DV, PerkinElmer). 150 mg of cotton fabric was put into 30 mL of concentrated nitric acid for 12 h to dissolve the silver on the cotton fabric. The above solution of Ag\(^+\) was diluted to 100 mL with deionized water, in which the concentration of Ag\(^+\) in the solution was measured with ICP-AES. The content of AgNPs was calculated by the following Eq. \( (3) \):

\[
\text{AgNPs loading content} = \frac{C_{\text{Ag}} \times V_{\text{Ag}}}{W_t} \text{ (mg/g)} \tag{3}
\]
Where \( V_{Ag} \) is the volume of the solution obtained after nitrication of the fiber (mL), \( C_{Ag} \) is \( Ag^+ \) concentration of the solution obtained after nitrication of the fiber (g/L) and \( W_t \) is the weight of cotton (g).

In order to characterize the release rate, AgNPs-cotton (300 mg) was placed in a sealed sample bottle containing 20 mL of deionized water and kept at 37°C. The cotton fabric was removed and then put into another sealed sample bottle containing 20 mL of deionized water every 24 h for 5 days. The concentration of \( Ag^+ \) in solution was analyzed by ICP-AES.

In order to characterize the antibacterial properties, the antibacterial activity of AgNPs-cotton was analyzed by shake flask method according to the GB/T 20944.32008 (China). 5 mL of bacterial culture solution (3×10^5 - 4×10^5 CFU/ mL) was placed in a flask containing 70 mL of PBS solution (pH = 7.2) with shaken. 0.75 g sample was cut into 0.5 cm × 0.5 cm pieces and placed in the flask mentioned above. The samples were placed in an incubator at a constant temperature of 24°C and 150 rpm for 18 h. 10 µL of solution was diluted and dropped on agar medium. All the agar medium were incubated at 37°C for 24 h. The antibacterial rate was calculated according to the number of colonies and Eq. (4).

\[
\text{percent reduction (\%) } = \frac{B-A}{B} \times 100% \quad \text{(4)}
\]

where \( B \) is the number of viable bacteria on the control sample, and \( A \) is the number of viable bacteria on the AgNPs-cotton cultured for 18 h.

3. Results And Discussion

In this study, three different methods were used to introduce carboxyl groups on cotton fibers, including selective oxidation, BTCA grafting and PAA adsorption. In selective oxidation, TEMPO-mediated oxidation system is used to oxidize the hydroxyl group on the 6-position of cellulose molecules to carboxyl group(Coseri et al. 2018; Isogai et al. 2011; Shinoda et al. 2012). BTCA grafting introduces carboxyl group into cotton fabric by crosslinking BTCA and cellulose molecules through esterification reaction of carboxyl group and hydroxyl group(Hashem et al. 2020b; Lund and Brelid 2013). TEMPO and BTCA molecules can diffuse into the micropore of cotton fiber because of their small size, therefore, the carboxyl groups can be introduced into the fiber by the above two methods. PAA with \( M_w \) of 80000 has large size, and mainly adsorb on the surface of the fiber through hydrogen bonding and van der Waals force(Li et al. 2020). In the following part, the carboxylate and charge contents of modified cotton fibers by different methods were firstly characterized by titration methods, focusing on the difference of the surface and the inner charges. AgNPs was then in situ formed on the fibers, and the content and distribution of AgNPs on the fibers were investigated. The wash-resistance of AgNPs-loaded fabrics and the antibacterial efficacy of the fabrics after 80 washings were also studied.

3.1 The effect of modification methods to the carboxylate and charge contents of cotton fabrics
The content of carboxyl group changes with the oxidant concentration, BTCA concentration, PAA concentration and reaction conditions (Isogai et al. 2018; Isogai et al. 2011; Montazer et al. 2012a; Montazer et al. 2012b). In this study, the carboxyl group content on OCF kept roughly the same as BCF by adjusting the type of reagents and reaction conditions, and then the influence of different modification methods on the distribution of carboxyl group in fibers was studied. Due to the low adsorption efficiency of PAA on fibers, the content of carboxyl groups introduced into fibers by PAA adsorption method was less than that of OCF and BCF. The specific test results are shown in Table 1, there was almost no carboxyl group and charge in CF, carboxyl groups were introduced into PCF, BCF and OCF respectively. The above results showed that oxidation, BTCA grafting and PAA adsorption had successfully introduced carboxyl groups into cotton fibers, in which the internal charge of OCF accounts for 61% of the total charge. The total charge of BCF was almost the same as the surface charge. Although the carboxyl group content of BCF was the same as OCF, the test results showed that the charge of BCF was much lower than that of OCF. This is because part of the surface of untreated cotton fiber was wrapped by non-cellulose materials such as pectin and protein. Oxidation removed these non-cellulosic materials and exposed the cellulose (Fras et al. 2005). At the same time, the surface of the cotton fiber was destroyed and the surface area became larger (Baron and Coseri 2020). Therefore, the cationic polyelectrolytes can be well combined with the carboxyl group of the fiber, so the PE titrations can more accurately reflect the charge distribution of the oxidized cotton fiber. BTCA in BCF not only acted as a carrier of carboxyl groups to introduce carboxyl groups into the fiber, but also acted as a crosslinking agent to cross-link the molecular chains of cotton fibers together, blocking the voids inside the fibers, which resulted in cationic polyelectrolytes unable to enter the fibers, and non-cellulose materials on the surface of cotton fibers cannot be grafted with carboxyl groups (Hashem et al. 2020a; Ji et al. 2016; Shekarriz et al. 2003). Therefore, the test results showed that the charge of BCF was much lower than that of OCF. The total charge and surface charge of PCF were very small. This is because the molecular weight of polyacrylic acid was too large to enter the fiber and can only be adsorbed on the surface of the cotton fiber. Most of the charge was therefore concentrated on the surface of the cotton fiber.

| Samples | Carboxyl content (mmol/g) | Charge (mmol/g) |
|---------|--------------------------|-----------------|
|         |                          | Total charge    | Surface charge |
| OCF     | 0.34                     | 0.36            | 0.14           |
| BCF     | 0.35                     | 0.09            | 0.08           |
| PCF     | 0.14                     | 0.11            | 0.08           |
| CF      | 0.002                    | 0.002           | 0.001          |
3.2 The content and distribution of AgNPs on fibers

Photographs of AgNPs-loaded cotton and SEM images of their fiber surface are shown in Fig. 1. AgNPs loaded on the cotton fabric changed the color of fabric. The control cotton is white and the fiber surface is very smooth. Compared with the control cotton, the color of AgNPs-CF is only slightly yellow, and the fiber surface has only a small number of nanoparticles; the colors of AgNPs-PCF, AgNPs-BCF and AgNPs-OCF were all obviously changed, in which the color of AgNPs-BCF and AgNPs-OCF became the dark brown. Because the contents of AgNPs on the different fiber surfaces were different, different cotton fabrics showed different colors.

The silver contents of different AgNPs-loading cotton fabrics are shown in Table 2, the silver content of CF and PCF are 1.2 mg/g and 6.1 mg/g. However, the FE-SEM images of the cross-sections of CF and PCF show that the interiors of these two fibers are not loaded with AgNPs (Fig. 2 CF and PCF). This is because the carboxyl groups exist only on the surface of PCF, so AgNPs can only be loaded on the surface of fiber, which can also be proved by the SEM images of the fiber surface (Fig. 1B and C). As shown in Table 2, the silver content of BCF and OCF were 12.8 mg/g and 12.0 mg/g respectively, much higher than that of CF and PCF. The FE-SEM images of the cross-sections of BCF and OCF proved that a large amount of AgNPs was loaded inside the two fibers (Fig. 2 BCF and OCF). The distribution of AgNPs (~ 30 nm) inside BCF and AgNPs (~ 20nm) inside OCF were very dense and uniform. Selective oxidation and BTCA grafting introduced a large number of carboxyl groups into the interior and surface of cotton fibers. The electrostatic attraction made a large amount of Ag⁺ diffuse into the fiber, thus the interior of the fibers was loaded with a large number of AgNPs.

| Samples       | AgNPs-CF | AgNPs-PCF | AgNPs-BCF | AgNPs-OCF |
|---------------|----------|-----------|-----------|-----------|
| AgNPs loading content (mg/g) | 1.2      | 6.1       | 12.8      | 12.0      |

3.3 Slow-release and wash-resistance of AgNPs on fabrics

In order to evaluate the washing resistance of AgNPs-cotton, four kinds of cotton fabrics were washed 0, 20, 40 and 80 cycles respectively according to AATCC Test Method 61-2007. Then the fabrics were characterized by UV-vis-NIR spectrometer. Figure 3 shows the reflectance spectrum of AgNPs-cotton. The load of AgNPs leaded to a steep drop in reflectance between 400 nm and 450 nm, and the reflectance changes with the increase of washing times. Figure 3A shows that the reflectance of the AgNPs-CF increased with the increase of washing cycles. As shown in Fig. 3B, the reflectance of the AgNPs-PCF after 20 cycles has hardly changed, and the reflectance increased slightly after 40 cycles, while the reflectance increased significantly until 80 cycles. AgNPs-BCF and AgNPs-OCF had no significant change in reflectance after 20 and 40 washing cycles, and the reflectance only increased slightly after 80 washing cycles. An increase in reflectivity means a decrease in the loading capacity of silver. Therefore, the above results indicate that PAA adsorption can improve the durability of AgNPs. However, since the
loading capacity of AgNPs is relatively small and distributed on the surface, increasing the number of washing will decrease the loading capacity of silver. The AgNPs of BCF and OCF are not only distributed on the surface of the fiber, but also distributed in the interior of the fiber, and the washable resistance is greatly improved. Only after 80 cycles of washing, the reflectivity is slightly increased.

In order to evaluate the stability of the AgNPs more accurately, ICP-AES was used to measure the AgNPs retention rate of AgNPs-cotton after 40 and 80 washing cycles. As demonstrated in Fig. 4A after the same number of washing, the durability of AgNPs-OCF was the best, followed by AgNPs-BCF, AgNPs-PCF and AgNPs-CF. The above laws reflected the washing durability of AgNPs on different cotton fabrics more accurately.

The results of OCF were compared with that of durable AgNPs-cotton fabrics in other literatures, where the washing procedure was generally the same as this paper. As shown in Fig. 4B, Lee et al. reported that antibacterial cotton fabrics directly impregnated with AgNPs sol, and the silver retention rate was only 7% after 20 washing cycles(Lee et al. 2003). Reed et al. demonstrated that their AgNPs-loaded T-shirts had a silver retention rate of 44% after 20 washing cycles(Reed et al. 2016). Zahran et al. used AgNPs-alginate to endow cotton fabric with antibacterial properties, the silver retention rate on cotton fabrics reached 54% after 20 washing cycles(Zahran et al. 2014). The studies by El-Rafie et al. showed that the addition of adhesives can improve washing durability, the AgNPs retention rate reached 65.8% after 20 washing cycles(El-Rafie et al. 2014b). Although Nam et al.(Nam et al. 2020) and Zhang et al.(Zhang et al. 2013) improved the process to improve the durability of AgNPs on cotton fabrics, silver retention rate reached 67.0% and 81.5% after 20 and 30 washes, respectively, but the retention rate was still lower than that of our samples. The AgNPs loaded on our samples were wrapped inside the fiber, resulting in its strong washing resistance. Although carboxyl groups were introduced into cotton fibers through different methods, but they had different effects on the washing durability of silver in cotton fibers. AgNPs-OCF had the best stability, because the oxidation introduces carboxyl group into the cotton fiber. The complex force of carboxyl group on Ag⁺ a large number of Ag⁺ diffuse into the fiber, thus the fiber was loaded with a large number of AgNPs. The AgNPs in the fiber were not easy to be leach out, and even after several washing cycles, the stability was still very good. AgNPs-PCF had the worst stability, because the dip-rolled PAA can only introduce carboxyl group on the surface of the ber, most of the AgNPs can only be loaded on the surface of the ber, and it is easy to lose with the increase of washing times.

In order to study the slow-release properties of AgNPs-cotton, the release rate of Ag⁺ from antibacterial cotton was measured by ICP-AES. As shown in Fig. 4C, the released Ag⁺ of AgNPs-CF was the highest, which achieved 2.4% after 5 days. The released Ag⁺ of AgNPs-PCF was 1.5%. However, the accumulative release of Ag⁺ from AgNPs-BCF and AgNPs-OCF were significantly lower than 0.9%. Hence, the introduction of carboxyl groups greatly enhanced the sustained release of silver. AgNPs on AgNPs-BCF and AgNPs-OCF were distributed inside the fiber and were not easy to be leached, so they have a strong sustained-release property. AgNPs on AgNPs-PCF was mainly located on the fiber surface, and its slow-
release property was slightly weaker than that of AgNPs of AgNPs-BCF. The prepared AgNPs-BCF and AgNPs-OCF can provide long-term antibacterial property by continuously releasing Ag⁺.

### 3.4 Antibacterial efficacy

The antibacterial activity of AgNPs-cotton was tested after 80 washing cycles by a shake flask method according to GB/T 20944.32008 (China). The bacterial solution after cultivation for 18h was diluted to different multiples, and then 10 µL of the diluted bacterial solution were dropped onto the agar plates. After cultivation for 24 h, the bacterial inhibition rate was calculated. Figure 5 shows the photographs of the number of colonies after culturing for 24 h. Table 3 shows the antibacterial activities of AgNPs-cotton against S. aureus and E. coli, AgNPs-BCF and AgNPs-OCF after 80 washing cycles inactivated all bacteria including S. aureus and E. coli. The percent reductions calculated according to Eq. (2) were both determined to 100% for S. aureus and E. coli. The percent reductions of AgNPs-CF were determined to 98.6% and 94.0% for S. aureus and E. coli. The percent reductions of AgNPs-PCF were determined to 97.9% and 99.0% for S. aureus and E. coli. The results showed that after 80 washing cycles, the antibacterial properties of AgNPs-BCF and AgNPs-OCF did not change significantly, while the antibacterial properties of AgNPs-PCF decreased significantly, and the antibacterial decline of AgNPs-CF was the largest, which also proved that the introduction of carboxyl groups can enhance the stability of AgNPs on cotton fibers, resulting in long-term durable antibacterial cotton fabrics.

**Table 3**

Antibacterial activities of AgNPs-cotton after 80 washing cycles

| Bacteria | Samples   | percent reduction (%) |
|----------|-----------|-----------------------|
|          | AgNPs-CF  | 94.0                  |
| E. coli  | AgNPs-PCF | 97.9                  |
|          | AgNPs-BCF | 100                   |
|          | AgNPs-OCF | 100                   |
|          | AgNPs-CF  | 98.6                  |
| S. aureus| AgNPs-PCF | 99.0                  |
|          | AgNPs-BCF | 100                   |
|          | AgNPs-OCF | 100                   |

**Conclusions**

In this work, carboxyl groups were successfully introduced into the inside of cotton fiber by BTCA grafting and selective oxidation. AgNPs were well-distributed inside the cotton fiber through the electrostatic attraction of carboxyl group and Ag⁺. On the contrary, most of the AgNPs of AgNPs-PCF were
concentrated on the surface of the fiber. Internal synthesis of AgNPs within cotton fiber is an effective and reliable way to improve washing durability and sustained release of Ag\(^+\). After 40 cycles, the silver content of AgNPs-BCF and AgNPs-OCF were 71.2\% and 92.4\%, respectively. However, the AgNPs content of AgNPs-CF and AgNPs-PCF were only 42.9\% and 61.2\%. The accumulative release of Ag\(^+\) from AgNPs-BCF and AgNPs-OCF were significantly lower than 0.9\%, which was much lower than AgNPs-CF and AgNPs-PCF. As expected, after 80 washing cycles, the percent reductions of AgNPs-BCF and AgNPs-OCF were all determined to 100\% against both S. aureus and E. coli. By contrast, the percent reductions of AgNPs-CF and AgNPs-PCF were lower than AgNPs-BCF and AgNPs-OCF.

Declarations

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Compliance with ethical standards

Conflict of interest

The authors declare that they have no conflict of interest.

Ethical approval

There are no studies involving humans or animals in this article.

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Page 14/20
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Figures
Figure 1

Photos of Cotton fiber (Left), SEM images of Cotton fiber (Right). (A) control cotton fiber, (B) AgNPs-CF, (C) AgNPs-PCF, (D) AgNPs-BCF, (E) AgNPs-OCF.
Figure 2

FE-SEM images of a given cross section at two different magnifications.
Figure 3

UV-vis-NIR reflectance spectra of control cotton fabrics and AgNPs-cotton fabrics taken after 0, 20, 40 washing cycles. (A) AgNPs-CF, (B) AgNPs-PCF, (C) AgNPs-BCF, (D) AgNPs-OCF.

Figure 4

(A) Silver retention rate of AgNPs-cotton in this paper, (B) Comparison of washing resistance between AgNPs-OCF and AgNPs-cotton in other literatures, (C) Accumulative release rate of Ag+ from AgNPs-cotton.
Figure 5

Photographs of the test plates of (up) S.aureus and (down) E.coli. (A,E) AgNPs-CF, (B,F) AgNPs-PCF, (C,G) AgNPs-BCF, (D,H) AgNPs-OCF, (I,J) control.

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