Magnetic Resonance Imaging-Guided Adaptive Radiotherapy for Colorectal Liver Metastases

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Simple Summary: Stereotactic ablative body radiotherapy (SABR) is an effective treatment for CRC liver metastases. Advanced radiation techniques are required to ensure safe and efficacious treatment. MR-guided radiotherapy is a new and evolving technology that can overcome many of the challenges of liver SABR. MR imaging before, during and after treatment delivery facilitates direct visualization of tumor target and adjacent normal healthy organs, real time MR imaging facilitates non-invasive tumor tracking and gating, and daily adaptive re-planning permits treatment plans to be adjusted for the anatomy of the day. MR-guided ablative radiation therapy is a promising radiation technology advance that can further facilitate safe tumor dose escalation of colorectal liver metastases.

Abstract: Technological advances have enabled well tolerated and effective radiation treatment for small liver metastases. Stereotactic ablative radiation therapy (SABR) refers to ablative dose delivery (>100 Gy BED) in five fractions or fewer. For larger tumors, the safe delivery of SABR can be challenging due to a more limited volume of healthy normal liver parenchyma and the proximity of the tumor to radiosensitive organs such as the stomach, duodenum, and large intestine. In addition to stereotactic treatment delivery, controlling respiratory motion, the use of image guidance, adaptive planning and increasing the number of radiation fractions are sometimes necessary for the safe delivery of SABR in these situations. Magnetic Resonance (MR) image-guided adaptive radiation therapy (MRgART) is a new and rapidly evolving treatment paradigm. MR imaging before, during and after treatment delivery facilitates direct visualization of both the tumor target and the adjacent normal healthy organs as well as potential intrafraction motion. Real time MR imaging facilitates non-invasive tumor tracking and treatment gating. While daily adaptive re-planning permits treatment plans to be adjusted based on the anatomy of the day. MRgART therapy is a promising radiation technology advance that can overcome many of the challenges of liver SABR and may facilitate the safe tumor dose escalation of colorectal liver metastases.

Keywords: colorectal liver metastasis; stereotactic ablative radiation therapy; proton therapy; MRI guided radiotherapy

1. Introduction

Colorectal cancer commonly metastasizes to the liver and can often be localized there. In the past, liver metastases were thought to be incurable and the only treatment option considered was systemic cytotoxic chemotherapy [1]. The treatment algorithm for colorectal cancer patients with disease confined to the liver has changed dramatically over the prior 20 years, with the advent of highly effective therapies to treat liver metastases [2]. The recognition of the oligometastatic state, defined as limited but not widespread metastatic...
disease, has helped spur further interest in developing effective local therapies to treat metastatic tumors [3].

Surgical metastasectomy series provided the initial data to support the benefit of metastatic directed treatments. Multiple groups have reported prolonged progression-free survival (PFS) and overall survival (OS) following metastasectomy for liver and/or lung metastases in patients with oligometastatic colorectal cancer [4–13]. Re-analysis of the Intergroup 0114 study noted an improvement in OS in rectal cancer patients with isolated metastatic disease treated with surgical resection as compared to no surgical resection (5 year OS 27% vs. 6%, \( p < 0.001 \)) [14]. Review of patients with newly diagnosed metastatic CRC (mCRC) treated at MD Anderson Cancer Center and Mayo Clinic demonstrated improvements in OS, for patients treated between 1998 and 2004, which was attributed primarily to an increase in hepatic resection [15]. Additional large retrospective series have demonstrated 5-year OS rates of nearly 40% in colorectal cancer patients with isolated liver metastasis following resection [4,5,9]. For patients with limited extrahepatic disease, complete surgical resection has also been associated with prolonged survival [13]. Shah et al. reported median disease-free survival and OS durations of 19.8 and 87 months in 39 patients who underwent both lung and liver resections for mCRC [11]. The 5-year OS was significantly longer for patients who underwent both liver and lung resections than those who did not undergo liver resection for pulmonary metastases (74% vs. 42%, \( p = 0.05 \)) [11]. These data, when taken together, demonstrate that for selected patients with oligometastatic CRC surgical metastasectomy can dramatically alter disease progression.

While surgical metastasectomy remains the standard of care, only ~20–30% of patients are surgical candidates [16]. Thus, there is interest in non-surgical treatment schemas, such as interventional radiology (IR) ablation with radiofrequency or microwave techniques or even high-intensity focal ultrasound and cryotherapy [17]. In fact EORTC 40004 demonstrated improved PFS and OS for radiofrequency ablation (RFA), as compared to systemic chemotherapy, in a randomized phase II trial for colorectal cancer patients with surgically unresectable disease limited to the liver [18,19]. Not all patients are good candidates for IR ablation as tumors in close proximity to large blood vessels and large tumors (i.e., >3 cm) have been reported to have worse local control [20,21]. IR ablation is safe, effective, and convenient for appropriate selected patients [20].

Stereotactic ablative body radiotherapy (SABR) is a noninvasive alternative treatment option. The safety of SABR for the treatment of 2 to 4 extracranial metastases has been demonstrated in the cooperative group setting with NRG-BR001, a phase I trial. Preliminary data indicate that SABR to multiple metastatic sites in the liver, lung, abdomen/pelvis, bone, and spine is safe [22]. No dose-limiting toxicity was identified across the evaluable anatomic locations [23,24]. The efficacy of SABR for treating oligometastatic disease was evaluated in SABR-COMET, which was a randomized, phase II tumor agnostic trial in 99 patients with up to 5 metastatic lesions [25]. Of the patients enrolled in the trial, 27% in the control group and 14% in the SABR group had a CRC primary. The trial met its primary endpoint of improved OS, with a median OS of 41 months in the SABR arm versus 28 months in the control arm (HR 0.57, 95% CI 0.30–1.10, \( p = 0.090 \)). While 4.5% of patients in the SABR group had grade 5 treatment-related adverse events, the 5-year OS rate was 17.7% in the control arm as compared to 42.3% in the SABR arm [26]. Given these data, the use of SABR has gained popularity as ~70% of radiation oncologists reported using SABR for the treatment of oligometastatic disease [27].

2. The Evolution of External Beam Radiation Therapy for Liver Tumors

Historically, external beam radiation therapy for the treatment of liver metastases was reserved for patients requiring palliation of painful liver disease. While, whole liver radiation (WLR) can palliate symptoms, WLR is ineffective at eradicating metastatic disease given the risk for radiation induced liver disease (RILD) [21,28–30]. The severity of RILD depends on a variety of factors including radiation dose and fractionation, volume of liver irradiated, and underlying liver dysfunction [31].
To reduce the risk for RILD, the Quantitative Analyses of Normal Tissue Effects in the Clinic (QUANTEC) published recommendations that restricted the volume and dose for whole liver radiation to 30 Gy in 2 Gy fractions or 21 Gy in 3 Gy fractions [32]. As the risk of RILD is correlated with the dose and volume of liver irradiated these constraints limit the risk of RILD to less than 5% [32]. As these constraints precluded doses required to eradicate liver disease many prematurely concluded that radiation therapy had a limited role in the treatment of intrahepatic malignancies and metastases [33].

Advances in radiotherapy technology provided the opportunity to develop techniques to deliver very high radiation doses to partial liver volumes, while simultaneously sparing healthy liver parenchyma [32]. Given these data, QUANTEC updated the practice guidelines for SABR to the liver. Current dose constraints recommend limiting the mean radiation dose to the liver to <15 Gy in 3 fractions and limiting 700 cm$^3$ of functional healthy liver to <15 Gy in 3-5 fractions [32]. These new constraints were developed to limit the risk of RILD. In patients with chronic liver disease or advanced cirrhosis the risk for RILD is higher and more stringent constraints should be employed [34–40]. In addition, it is important to optimize liver function and address any reversible causes of liver decompensation prior to liver SABR.

The Karolinska Institute was the first to report the efficacy and safety of high dose focal liver irradiation using a stereotactic technique in the 1990’s [41]. Additional centers have prospectively reported on the efficacy and safety of liver SABR, Table 1 [42–55]. Our prior review provides more depth on the background and history of the liver SABR [3]. Importantly, there seems to be a correlation between tumor control and radiation dose [42,56,57]. As the treatment of liver metastases is a balance between adequately ablating the tumor while protecting the normal healthy liver, the future will likely entail tailoring radiation dose based on the predicted tumor radiation sensitivity to achieve 90% local control while limiting the risk of RILD to less than 5% or more appropriately as low as possible.
| Study                          | Study Type                  | Patient # | Lesion # | Primary Histology | Dose (Gy)/Fraction Number | BED (a/b = 10) | Toxicity | Local Tumor Control | Overall Survival |
|-------------------------------|----------------------------|-----------|----------|-------------------|---------------------------|----------------|----------|---------------------|-----------------|
| Herfarth et al. [42]          | Phase I/II                 | 37        | 60       | Mixed             | 14-26/1                   | 33–94          |          | 71% @ 1 year        | 72% @ 1 year    |
| Mendez–Romero et al. [43]     | Phase I/II                 | 17        | 39       | Mixed             | 37.5/3                    | 84             | 12% G3 Liver | 100% @ 1 year | 86% @ 2 years |
| Rusthoven et al. [44]         | Phase I/II                 | 47        | 63       | Mixed             | 36–60/3                   | 79–180         | 1.5% G3 (dermatitis) | 95% @ 1 year | 92% @ 2 years |
| Lee et al. [45]               | Dose escalation, phase I   | 68        | 142      | Mixed             | 28-60/6                   | 41–120         | 10% G3+   | 71% @ 1 year        | 47% @ 1.5 years |
| Rule et al. [46]              | Dose escalation, phase I   | 27        | 37       | Mixed             | 30/3–50–60/5              | 60–132         | 4% G3 Liver | 100%, 89%, 56% @ 2 years | 50%, 67%, 56% @ 2 years |
| Comito et al. [47]            | Observational              | 42        | 52       | Colorectal        | 75/3                      | 263            | 60% G2, 0%G3 | 95% @ 1 year        | 85% @ 2 years    |
| Scorsetti et al. [48]         | Phase II                   | 42        | 52       | Colorectal        | 75/3                      | 263            | 25% G2 Liver, 0% G3 | 95% @ 1 year | 91% @ 2 years |
| Goodman et al. [49]           | Dose escalation, phase I   | 26        | 40       | Mixed             | 18–30/1                   | 50–120         | 8% GI bleeding | 77% @ 1 year        | 50% @ 2 years    |
| Meyer et al. [50]             | Dose escalation, phase I   | 14        | 17       | Mixed             | 35–40/1                   | 158–200        | 6% G2     | 100% @ 2.5 years | 78% @ 2 years |
| Hong et al. [51]              | Phase II                   | 89        | 143      | Mixed             | 30–50/5                   | 48–100         | No G3+    | 72% @ 1 year        | 61% @ 3 years    |
| Scorsetti et al. [52]         | Phase II                   | 61        | 76       | Mixed             | 75/3                      | 263            | 2% G3 chest wall pain | 94% @ 1 year | 78% @ 3 years |
| Kang et al. [53]              | Phase I                    | 9         | 14       | Mixed             | 36–60/3                   | 79–180         | No G3+    | NR                  | NR              |
| Dawson et al. [54]            | Dose escalation, phase I, multicenter | 26 | 37 | Mixed | 35–50/10 | 47–75 | 7.7% G3 GI | NR | NR |

# = number, cm = centimeter, BED = biologically effective dose, Gy = Gray, NR = not reported, GI = gastrointestinal, G3 = grade 3, G2 = grade 2, @ = at, * = 60 Gy/12 Gy per fraction, 50 Gy/10 Gy per fraction, and 30 Gy/10 Gy per fraction dose cohorts.
3. Liver SABR Treatment Planning and Delivery Considerations

Liver SABR has the potential for serious side effects if the delivery does not accurately reflect the treatment plan. All patients undergo standard simulation scan in the treatment position with computed tomography (CT). These images are used for treatment planning. As accurate planning requires tumor visualization, the simulation CT scans typically includes intravenous contrast administration as deemed necessary to help facilitate tumor and normal organ delineation. In addition, alternative imaging modalities (positron emission tomography (PET) or magnetic resonance imaging (MRI)) can be utilized to ensure accurate delineation of the tumor target and adjacent organs at risk (OAR) [21,58,59].

To localize the tumor and to protect the radiosensitive adjacent normal tissues such as the stomach, small intestine, and large intestine, respiratory motion must be controlled or accounted for during simulation and treatment delivery. The degree of movement, especially near the liver dome, can be significant [60]. Multiple strategies can be employed to either control or account for tumor motion. One relatively straightforward technique to account for tumor motion is to include a four-dimensional CT (4DCT) with the radiation simulation scan. The 4DCT will allow liver and, more importantly tumor, motion to be accounted for throughout the respiratory cycle. Including the path of tumor movement in the treatment target volume ensures adequate coverage, albeit at the cost of more liver parenchyma being treated given the larger margin around the tumor. Thus, the use of a 4DCT is most appropriate for small tumors that are far away from radiosensitive luminal organs. More advanced motion management techniques deliver treatment during specific phases of respiration. These techniques are known as gating [61,62]. Gating can also use deep inspiratory breath hold (DIBH), where patients are treated while holding near maximal inspiration. For DIBH to be successful patients must be relatively fit and able to hold their breath for 20–40 s intervals. Similar in concept to DIBH is end-expiratory gating. Here, patients are breathing normally, but the delivery of radiation is restricted to the expiration phase of the natural breathing cycle. An alternative strategy is abdominal compression. Abdominal compression treats throughout the respiratory cycle, but the amplitude of the respiratory cycle is restricted, which limits tumor motion to generally less than 5 mm. With abdominal compression a cuff is placed around the abdomen of the patient and insufflated to restrict respiratory motion. As pressures can reach 40–50 mmHg, some patients find abdominal compression to be uncomfortable and anxiety provoking. While the goal of compression is to limit diaphragmatic movement, it also compresses the radiosensitive abdominal organs into the liver. Thus, abdominal compression may not be optimal in all cases and is of particular concern for metastases in the left lobe, which is in close proximity to the stomach, but generally not a concern for metastases in the right lobe [63]. All of these motion management techniques have advantages and disadvantages that need to be considered when determining the most appropriate technique for an individual patient. In addition, most centers and radiation oncology teams only have experience with one or two of these motion management techniques and limited experience with the others, which makes cross comparing them difficult.

For accurate treatment delivery the patient and tumor must be accurately aligned daily for treatment. As the position of the liver in relation to bony anatomy can change by up to 1 cm between fractions, liver SABR presents a particular challenge [64,65]. Since metastatic tumors are difficult to visualize fiducial markers or the shape of the liver can be used to help ensure proper alignment with daily non-contrast cone beam CT (CBCT) scans taken with the patient in the treatment position on the treatment machine [66]. While setting up to liver shape is a significant improvement as compared to aligning relative to bony anatomy, there is still a degree of inter-fractional motion uncertainty especially at the dome of the liver. For tumors near the dome of the liver, it is preferable to add an additional margin cranially and caudally due to the variability of the position of the diaphragm with DIBH if the alignment will be to the liver shape. One way to improve daily tumor localization and decrease inter-fraction motion is to place fiducial markers in and around the tumor. Alternatively, surgical clips can be utilized if already in place and in close proximity to
the tumor [67–69]. Fiducial markers and/or appropriate positioned surgical clips can help ensure target accuracy (~2–3 mm) with real time intrafraction motion assessment [58]. A well utilized strategy for motion management will allow smaller margins for treatment and help ensure the safety and efficacy of SABR for tumors in close proximity of radiosensitive OARs, as well as large tumors where the dose to the healthy liver parenchyma is limiting.

4. Strategies to Overcome Limitations to SABR

SABR to liver metastases can be challenging in patients with large tumors and/or tumors abutting or in close proximity to radiosensitive gastrointestinal organs [70]. When limited by adjacent OARs, radiation oncologists commonly opt to de-intensify treatment to a sub- ablative palliative dose. This strategy while safe, precludes tumor ablation in patients who have a high likelihood of benefit. Proceeding with sub- ablative treatment, fails to meet the goal of oligometastatic directed therapy. An alternative approach is to consider adopting moderate hypofractionation (e.g., expanding to 10, 15, or even 25 fractions) to maintain an ablative treatment [3]. Fractionation results in greater normal tissue tolerance as sublethal DNA damage is repaired between fractions. Moderate hypofractionation maintains an ablative dose, while prioritizing safety (i.e., limits risk of GI bleeding, RILD, and bile duct strictures) [69]. If OAR constraints cannot be met for a tradition hypofractionated SABR plan one must decide to either proceed with an unsafe treatment to maximize tumor control, proceed with a safe treatment that compromises tumor control, or alternatively to adopt moderate hypofractionation to maintain tumor control while ensuring safety. As we have described previously, we strongly favor adopting moderate hypofractionation to ensure safety while maximizing tumor control [3].

In patients with large tumors or underlying liver dysfunction, proton beam radiotherapy should be considered. Protons come to rest and deposit their energy at a prespecified range in the patient, the Bragg peak. As there is minimal exit dose, protons have a dosimetric advantage over photon radiotherapy. Thus, proton SABR for liver metastases can spare more normal liver than photon SABR [71]. In some patients with bi-lobar or large tumors where the liver dose constraints will be impossible to meet with photon based SABR, protons may permit ablative radiation therapy [72]. In fact, patients with primary liver tumors treated with proton, as compared to photon, radiotherapy resulted in less liver decompensation and better survival [73]. This survival benefit was attributed to a significantly lower mean liver dose, which decreased the risk of non-classic RILD in proton patients [73]. Additional prospective clinical trials have demonstrated the efficacy and safety of proton liver SABR [51,53]. Just as with photon based therapy, considerations need to be taken for motion management and image guidance with proton radiotherapy.

5. MR Guided Adaptive Radiation Therapy

MRgART is a promising radiation technology advance that can overcome many of the challenges of traditional liver SABR and may facilitate safe tumor dose escalation of colorectal liver metastases. The US Food and Drug Administration (FDA) has recently approved the Elekta Unity™ (Elekta, Stockholm, Sweden) and the View Ray MRIdian™ (ViewRay, Oakwood Village, OH, USA) MRI-guided radiation treatment systems for clinical use. Unlike traditional image guidance which use on-board CBCT, the coupling of a MRI and treatment LINAC permits image acquisition before, during, and after radiation delivery, with the patient in the treatment position [74].

The addition of on-board MR imaging offers a number of advantages as MR imaging can be done to ensure patient alignment before treatment, as with typical CBCT-guided approaches, albeit without the excessive radiation, and also continuously during treatment delivery. First, MR imaging offers superior soft tissue delineation so that liver metastases can be accurately visualized and thus alleviate the need for multi-phase contrast CT simulation scans (Figure 1A). In fact, many groups are working on MR-based radiation planning and thus avoiding CT based simulations altogether [75,76]. Second, dynamic MR imaging during before, during, and after treatment allows temporospatial tumor
monitoring throughout treatment to ensure accurate targeting and treatment delivery [77].

The combination of superior contrast resolution combined with a fiducial less treatment is a significant advantage over CBCT scans obtained on traditional LINACs for image guided SBRT [3]. Given the superior soft tissue delineation with MRI, liver tumors can be readily identified and patients can thus be treated without fiducials and spared an invasive and now unnecessary procedure. Similar to non-MR guided treatment, motion management must be accounted for. Many of the same techniques discussed above (i.e., DIBH, gating, and abdominal compression) can be used with MR-radiotherapy systems but require special MR compatible equipment. One potential significant advance is MR based tumor tracking and gating. As the tumor is monitored in real time with dynamic MR imaging, treatment can be delivered only when the tumor is within a pre-specified treatment gate [77].

This improved accuracy decreases inter- and intra-fraction motion and permits treatment with tighter margins, thus sparing more healthy viable tissue. Third, MR-guided treatment programs permit adaptive re-planning, daily in real time, to account for inter-fractional tumor and normal organ motion (Figure 1B–D) [78]. Changes to the radiation plan can be made on a daily basis to ensure optimal tumor target and normal tissue sparing. Much of the aforementioned limits of SABR are easily overcome with MR-guided based treatment as MR imaging permits accurate tumor localization thus minimizing inter-fraction motion and alleviating the need for fiducial marker placement, dynamic imaging during treatment delivery provides a non-invasive system to adapt for tumor movement with respiration, and rapid adaptive replanning in real time provides an opportunity to adjust the radiation delivery plan daily depending on changes in internal anatomy.

Initial experiences in using MRgART for primary and metastatic liver tumors have been published [79–82]. Rosenberg et al. reported a multi-institutional retrospective series on 26 patients with liver tumors (45% colorectal liver metastasis) treated with MRgART to a median dose of 50 Gy in 10 Gy fractions [79]. Freedom from local progression was 80% at 21 months (median follow up). Toxicity was low with 7.7% grade 3 gastrointestinal toxicity and no grade 4+ toxicity appreciated. Interestingly, Rosenberg et al. postulate that traditional dose and volume constraints for organs at risk might be overly conservative for MR-based radiotherapy given the significantly improved tumor and normal tissue delineation and temporospatial real time monitoring [79]. Clinical trials are underway to better define normal tissue dose constraints in the MR-guide radiotherapy era [79].

At our institution, MRgART for Liver SABR is performed using Elekta’s Adapt-to-Shape workflow to a prescription of 54–60 Gy in 3–5 fractions [78]. All patients are simulated and treated with abdominal compression to reduce both tumor and OAR motion to be less than 5 mm. Automatic breath-hold and gating or other motion management options are not currently available on the Unity MR-linac system. The GTV volumes on simulation CT are drawn with the help of MR simulation images that includes a 2D T2w turbo spin echo MR (TR/TE = 1250/80 ms, slice thickness = 4 mm, voxel size = 1.3 × 1.6 mm²) and a 3D radial T1w acquisition called 3DVANE radial MR sequence (TR/TE = 4.1/1.68 ms, slice thickness = 4 mm, voxel size = 1.4 × 1.4 mm²) acquired after Eovist contrast injection. During each treatment fraction, a 3D T1w fat saturated MRI (TR/TE = 4.6/2.3 ms, voxel size = 2 × 2 × 2 mm³, FOV = 450 × 450 × 250 mm³) and/or T2 3D navigator triggered (TR/TE = 1900/87 ms, voxel size = 2 × 2 × 2.4 mm³, FOV = 400 × 448 × 250 mm³) sequence is registered to the reference CT/MR by first performing a rigid spine match and then adjusting the fusion to match the GTV. The remaining OARs are propagated using Monaco deformable image registration algorithm and manually edited by the physician and planners. The simulation CT and on-treatment planning MRIs, both represent a motion averaged image of the residual motion with the compression belt. A new adaptive plan is generated in Monaco using fluence optimization. Right before beam-on, another 3D T1w fat saturated MRI is acquired, and the structures and plan-of-the-day are overlaid to assess GTV coverage and evaluate for any potential intrafraction motion that may have occurred during contouring and planning. The contours are adjusted, and a new plan is generated if significant motion occurs. Otherwise, the treatment is delivered using a three orthogonal
plane 2D balanced fast-field echo cine MR monitoring at 5 frames/s (TR/TE = 2.6/1.32 ms, slice thickness = 5 mm, FOV = 400(FH) × 424(RL) mm², voxel size = 3 × 3 mm²) at the centroid of the motion monitoring structure. Our team typically make use of an OAR near the high dose PTV (e.g., stomach or bile ducts) as the monitoring structure. Overall average adapt-to-shape time divided into contouring, planning, physics QA and beam-on time is 21.1 ± 7.2 min, 14.5 ± 5.2 min, 4.4 ± 1.0 min and 14.7 ± 2.2 min, respectively.

Figure 1. MR-guided adaptive radiation therapy. Seventy-two-year-old female with three liver metastases (2 cm lesion in segment 8 and segments 3/4B measuring 1.6 cm) treated using MRg-ART on Elekta Unity MR-linac to a prescription of 60 Gy (12 Gy in 5 fractions). (A) The tumor metastasis was not visible on non-contrast planning CT as the patient was not a candidate for iodinated IV contrast due to poor renal function. Patient was simulated supine in a custom immobilization device and with an abdominal compression belt to minimize respiratory liver motion to <5 mm. (B) Online adaptive plan for a fraction on a T2 3D navigator triggered MRI. (C) T1w 3D fat saturated MRI was used in combination with T2 3D navigator triggered sequence to delineate GTV and OARs during online planning. Daily adaptive planning was performed to account for coverage of multiple lesions (due to varying deformations in the liver) as well as to protect the organs at risk. Bile ducts were used as the monitoring structure on the three orthogonal plane balanced fast field echo cine MRI during radiation delivery. (D) Daily dosimetric coverage for GTV and various relevant OARs. Each fraction dose is displayed in terms total prescription dose.
As MRgART is a new technology, there remain limitations that need to be overcome. Contouring of gross tumor volumes (GTV) and OARs is still one of the most time and resource consuming part of online MRgART planning and require the use of MR-based auto-segmentation algorithms in routine clinical workflow [83]. Due to the lack of robust clinical tools, coverage to OARs are still assessed based on a conservative assumption that the maximum dose lies in the same position from one fraction to another. Clinical tools that can accurately estimate both the spatial and temporal variation of the maximum doses to the OARs will allow further safe delivery of ablative doses to the target by eliminating the need to over-constrain the OARs by adding larger safety margins. Deformable image registration based dose accumulation studies for MRgART are limited and is an area of active development [84]. Current clinical workflows for MRgART allow use of single and limited MR sequences for contouring GTV and OARs during online planning. Alternative workflows on Elekta Unity are emerging that can make use of multiple MR contrast simultaneously [85]. Figure 1 shows the use of T2w 3D navigator triggered MR that was used simultaneously with T1 3D fat saturation MRI for GTV and OAR delineation. Finally, liver lesions by themselves may not be visible on the vendor provided single or multiple orthogonal plane 2D cine monitoring MR scan and may require the use of MRI contrast agent such as Eovist or surrogate structures such as liver or bile ducts for patient position monitoring or gating. Development of real-time volumetric MRI for online planning and tumor tracking will further improve the accuracy of adaptive treatments for GI tumors [86].

Beyond improving clinical outcomes, functional MR imaging before, during, and after treatment may lead to the identification of predictive radiomic markers of response [87]. The MR-guided systems can acquire functional images and assess changes in the tumor and tumor microenvironment during and after treatment and provide an ideal platform to generate reproducible quantitative biomarkers and facilitate collection and validation of large datasets for use in outcome research. Initial pilot studies performed on these systems have shown that such measurements are feasible [88–91]. This will hopefully pave the way for more personalized therapies as our understanding of tumor histology, size, genetics, and radiomics continues to improve. Figure 2 shows the treatment response assessment of one of the liver lesions shown in Figure 1 using apparent diffusion coefficient (ADC) derived from diffusion weighted (DW-) MRI. These images were acquired on the Elekta Unity platform using four b-values (0, 30, 150 and 550 mm²/s). The histogram distribution as a function of treatment fraction shows increasing number of pixels shifting towards high ADC values. The increasing ADC trend over the course of treatment suggest good tumor response. Follow up imaging studies will corroborate that. Studies in various tumor sub-sites have suggested that increase in ADC is a surrogate for good tumor response [92–94]. Multiple groups have just started to acquire such data on their MR-guided systems without adding any additional time for acquisition. Eventual goal of this data will be to correlate during treatment response to long term tumor control.
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Figure 2. Treatment response of an example liver lesion using T2w MRI and diffusion weighted MRI derived apparent diffusion coefficient maps. (A) T2w 3D navigator triggered MRI for during treatment fractions with GTV lesion displayed. (B) Apparent diffusion coefficient maps derived using four b-values (0, 30, 150 and 550 mm²/s). (C) Histogram distribution of ADC values within the GTV. (D) Trend analysis of ADC values using boxplot.

6. Conclusions

SABR is an effective treatment for CRC liver metastases with locoregional control rates in prospective clinical trials in excess of 75%. Advanced radiation techniques are required to ensure safe and efficacious treatment. In patients with large liver metastases or in metastases abutting or in close proximity to radiosensitive gastrointestinal organs SABR can be challenging. Motion management and consideration for moderate hypofractionation treatment schedules are important to ensure efficacy and safety of liver SABR. For patients with exceptionally large tumors, limited liver reserve, or underlying chronic liver disease proton radiotherapy should be considered. MR-guided radiotherapy is a new and evolving technology that can overcome many of the challenges of liver SABR. MR imaging before, during and after treatment delivery facilitates direct visualization of tumor target and adjacent normal healthy organs, real time MR imaging facilitates non-invasive tumor tracking and gating, and daily adaptive re-planning permits treatment plans to be adjusted for the anatomy of the day. MR-guided ablative radiation therapy is a promising radiation technology advance that can further facilitate safe tumor dose escalation of colorectal liver metastases.

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References

1. Aitken, K.L.; Hawkins, M.A. Stereotactic Body Radiotherapy for Liver Metastases. *Clin. Oncol.* 2015, 27, 307–315. [CrossRef] [PubMed]
2. Goodman, K.A.; Kavanagh, B.D. Stereotactic Body Radiotherapy for Liver Metastases. *Semin. Radiat. Oncol.* 2017, 27, 240–246. [CrossRef] [PubMed]
3. Romesser, P.B.; Neal, B.P.; Crane, C.H. External Beam Radiation Therapy for Liver Metastases. *Surg. Oncol. Clin. N. Am.* 2021, 30, 159–173. [CrossRef] [PubMed]
4. Fong, Y.; Fortner, J.; Sun, R.L.; Brennan, M.F.; Blumgart, L.H. Clinical Score for Predicting Recurrence After Hepatic Resection for Metastatic Colorectal Cancer: Analysis of 1001 consecutive cases. *Ann. Surg.* 1999, 230, 309–318. [CrossRef]
5. Morris, E.J.; Forman, D.; Thomas, J.D.; Quirke, P.; Taylor, E.F.; Fairley, L.; Cottier, B.; Poston, G. Surgical management and outcomes of colorectal cancer liver metastases. *Br. J. Surg.* 2010, 97, 1110–1118. [CrossRef]
6. Nordlinger, B.; Guiguet, M.; Vaillant, J.C.; Balladur, P.; Boudjema, K.; Bachellier, P.; Jaek, D. Surgical resection of colorectal carcinoma metastases to the liver: A prognostic scoring system to improve case selection, based on 1568 patients. *Cancer Interdiscip. Int. J. Am. Cancer Soc.* 1996, 77, 1254–1262. [CrossRef]
7. Okumura, T.; Boku, N.; Hishida, T.; Ohde, Y.; Saka, Y.; Yoshiha, K.; Higashiyma, M.; Hyodo, I.; Mori, K.; Kondo, H. Surgical Outcome and Prognostic Stratification for Pulmonary Metastasis from Colorectal Cancer. *Ann. Thorac. Surg.* 2017, 104, 979–987. [CrossRef]
8. Onaitis, M.W.; Petersen, R.P.; Haney, J.C.; Saltz, L.; Park, B.; Flores, R.; Rizk, N.; Bains, M.S.; Dycoco, J.; D’Amico, T.A.; et al. Prognostic Factors for Recurrence After Pulmonary Resection of Colorectal Cancer Metastases. *Ann. Thorac. Surg.* 2009, 87, 1684–1688. [CrossRef]
9. Choti, M.A.; Sitzmann, J.V.; Tiburi, M.F.; Sumetchotimetha, W.; Rangsin, R.; Schulick, R.D.; Lillemoe, K.D.; Yeo, C.J.; Cameron, J.L. Trends in Long-Term Survival Following Liver Resection for Hepatic Colorectal Metastases. *Ann. Surg.* 2002, 235, 759–766. [CrossRef]
10. Lehnert, T.; Knaebel, H.P.; Dück, M.; Bülzebruck, H.; Herfarth, C. Sequential hepatic and pulmonary resections for metastatic colorectal cancer. *Br. J. Surg.* 1999, 86, 241–243. [CrossRef]
11. Shah, S.A.; Haddad, R.; Al-Sukhni, W.; Kim, R.D.; Greig, P.D.; Grant, D.R.; Taylor, B.R.; Langer, B.; Gallinger, S.; Wei, A.C. Surgical Resection of Hepatic and Pulmonary Metastases from Colorectal Carcinoma. *J. Am. Coll. Surg.* 2006, 202, 468–475. [CrossRef]
12. Johnson, B.; Jin, Z.; Truty, M.J.; Smoot, R.L.; Nagorney, D.M.; Kendrick, M.L.; Kipp, B.R.; Grothey, A. Impact of Metastasectomy in the Multimodality Approach forBRAFV600E Metastatic Colorectal Cancer: The Mayo Clinic Experience. *Oncolist 2018*, 23, 128–134. [CrossRef]
13. McCormack, P.M.; Burt, M.E.; Bains, M.S.; Martini, N.; Rusch, V.W.; Ginsberg, R.J. Lung Resection for Colorectal Metastases. 10-year results. *Arch. Surg.* 1992, 127, 1403–1406. [CrossRef]
14. Tepper, J.E.; O’Connell, M.; Hollis, D.; Niedzwiecki, D.; Cooke, E.; Mayer, R.J.; Intergroup, S. Analysis of Surgical Salvage after Failure of Primary Therapy in Rectal Cancer: Results from Intergroup Study 0114. *J. Clin. Oncol.* 2003, 21, 3623–3628. [CrossRef] [PubMed]
15. Kopetz, S.; Chang, G.J.; Overman, M.J.; Eng, C.; Sargent, D.J.; Larson, D.W.; Grothey, A.; Vauthey, J.-N.; Nagorney, D.M.; McWilliams, R.R. Improved Survival in Metastatic Colorectal Cancer Is Associated With Adoption of Hepatic Resection and Improved Chemotherapy. *J. Clin. Oncol.* 2009, 27, 3677–3683. [CrossRef]
16. Hewish, M.; Cunningham, D. First-line treatment of advanced colorectal cancer. *Lancet* 2011, 377, 2060–2062. [CrossRef]
17. Garcea, G.; Lloyd, T.D.; Aylott, C.; Maddern, G.; Berry, D.P. The emerging role of focal liver ablation techniques in the treatment of primary and secondary liver tumours. *Eur. J. Cancer* 2003, 39, 2150–2164. [CrossRef]
18. Ruers, T.; Van Coevorden, F.; Punt, C.J.; Ferie, J.; Borel-Rinke, I.; Ledermann, J.A.; Poston, G.; Bechstein, W.; Lentz, M.A.; Mauer, M.; et al. Local Treatment of Unresectable Colorectal Liver Metastases: Results of a Randomized Phase II Trial. *J. Natl. Cancer Inst.* 2017, 109. [CrossRef]
19. Ruers, T.; Punt, C.; Van Coevorden, F.; Perie, J.P.E.N.; Borel-Rinke, I.; Ledermann, J.A.; Poston, G.; Bechstein, W.; Lentz, M.A.; Mauer, M.; et al. Radiofrequency ablation combined with systemic treatment versus systemic treatment alone in patients with non-resectable colorectal liver metastases: A randomized EORTC Intergroup phase II study (EORTC 40004). *Ann. Oncol.* 2012, 23, 2619–2626. [CrossRef]
20. Wong, S.L.; Mangu, P.B.; Choti, M.A.; Crocenzi, T.S.; Dodd, G.D., 3rd; Dorfman, G.S.; Eng, C.; Fong, Y.; Giusti, A.F.; Lu, D.; et al. American Society of Clinical Oncology 2009 Clinical Evidence Review on Radiofrequency Ablation of Hepatic Metastases From Colorectal Cancer. J. Clin. Oncol. 2010, 28, 493–508. [CrossRef]

21. Hoyer, M.; Swaminath, A.; Bydder, S.; Lock, M.; Romero, A.M.; Kavanagh, B.; Goodman, K.A.; Okunieff, P.; Dawson, L.A. Radiotherapy for Liver Metastases: A Review of Evidence. Int. J. Radiat. Oncol. Biol. Phys. 2012, 82, 1047–1057. [CrossRef] [PubMed]

22. Chmura, S.J.; Winter, K.A.; Salama, J.K.; Robinson, C.G.; Pisansky, T.M.; Borges, V.; Al-Hallaq, H.A.; Matuszak, M.M.; Park, S.S.; Gonzalez, V.J.; et al. Phase I Trial of Stereotoxic Body Radiation Therapy (SBRT) to Multiple Metastatic Sites: A NRG Oncology Study. Int. J. Radiat. Oncol. 2018, 102, 568–569. [CrossRef]

23. Al-Hallaq, H.A.; Chmura, S.; Salama, J.K.; Winter, K.A.; Robinson, C.G.; Pisansky, T.M.; Borges, V.; Lowenstein, J.R.; McNulty, S.; Galvin, J.M.; et al. Rationale of technical requirements for NRG-BR001: The First NCI-sponsored trial of SBRT for the treatment of multiple metastases. Pract. Radiat. Oncol. 2016, 6, e291–e298. [CrossRef] [PubMed]

24. Al-Hallaq, H.A.; Chmura, S.J.; Salama, J.K.; Lowenstein, J.R.; McNulty, S.; Galvin, J.M.; Followill, D.S.; Robinson, C.G.; Pisansky, T.M.; Winter, K.A.; et al. Benchmark Credentialing Results for NRG-BR001: The First National Cancer Institute-Sponsored Trial of Stereotactic Body Radiation Therapy for Multiple Metastases. Int. J. Radiat. Oncol. Biol. Phys. 2017, 97, 155–163. [CrossRef]

25. Palma, D.A.; Olson, R.; Harrow, S.; Gaede, S.; Louie, A.V.; Haasbeek, C.; Mulroy, L.; Lock, M.; Rodrigues, G.B.; Yaremko, B.P.; et al. Stereotoxic ablative radiotherapy versus standard of care palliative treatment in patients with oligometastatic cancers (SABR-COMET): A randomised, phase 2, open-label trial. Lancet 2019, 393, 2051–2058. [CrossRef]

26. Palma, D.A.; Olson, R.; Harrow, S.; Gaede, S.; Louie, A.V.; Haasbeek, C.; Mulroy, L.; Lock, M.; Rodrigues, G.B.; Yaremko, B.P.; et al. Stereotoxic Ablative Radiotherapy for the Comprehensive Treatment of Oligometastatic Cancers: Long-Term Results of the SABR-COMET Phase II Randomized Trial. J. Clin. Oncol. 2020, 38, 2830–2838. [CrossRef]

27. Lewis, S.L.; Porceddu, S.; Nakamura, N.; Palma, D.A.; Lo, S.S.; Hoskin, P.; Moghanaki, D.; Chmura, S.J.; Salama, J.K. Definitive Stereotoxic Body Radiotherapy (SBRT) for Extracranial Oligometastases: An International Survey of >1000 Radiation Oncologists. Am. J. Clin. Oncol. 2017, 40, 418–422. [CrossRef]

28. Bydder, S.; Spry, N.A.; Christie, D.R.; Roos, D.; Burmeister, B.H.; Krawitz, H.; Davis, S.; Joseph, D.J.; Poulsen, M.; Berry, M. A prospective trial of short-fractionation radiotherapy for the palliation of liver metastases. Australas. Radiol. 2003, 47, 284–288. [CrossRef]

29. Borgelt, B.B.; Gelber, R.; Brady, L.W.; Griffin, T.; Hendrickson, F.R. The palliation of hepatic metastases: Results of the radiation therapy oncology group pilot study. Int. J. Radiat. Oncol. Biol. Phys. 1981, 7, 587–591. [CrossRef]

30. Soliman, H.; Ringash, J.; Jiang, H.; Singh, K.; Kim, J.; Dinniwel, R.; Brade, A.; Wong, R.; Brierley, J.; Cummings, B.; et al. Phase II Trial of Palliative Radiotherapy for Hepatocellular Carcinoma and Liver Metastases. J. Clin. Oncol. 2013, 31, 3980–3986. [CrossRef]

31. Toesca, D.A.S.; Braggimov, B.; Koong, A.J.; Xing, L.; Koong, A.C.; Chang, D.T. Strategies for prediction and mitigation of radiation-induced liver toxicity. J. Radiat. Res. 2018, 59, i40–i49. [CrossRef] [PubMed]

32. Pan, C.C.; Kavanagh, B.D.; Dawson, L.A.; Li, X.A.; Das, S.K.; Miften, M.; Haken, R.K.T. Radiation-Associated Liver Injury. Int. J. Radiat. Oncol. Biol. Phys. 2010, 76, S94–S100. [CrossRef] [PubMed]

33. Lawrence, T.S.; Robertson, J.M.; Anscher, M.S.; Jirtle, R.L.; Ensminger, W.D.; Fajardo, L.F. Hepatic toxicity resulting from cancer treatment. Int. J. Radiat. Oncol. 1995, 35, 1237–1248. [CrossRef]

34. Cheng, J.C.; Wu, J.K.; Huang, C.M.; Huang, D.Y.; Cheng, S.H.; Lin, Y-M.; Jian, J.J.; Yang, P-S.; Chuang, V-P.; Huang, A.T. Radiation-induced liver disease after radiotherapy for hepatocellular carcinoma: Clinical manifestation and dosimetric description. Radiother. Oncol. 2002, 63, 41–45. [CrossRef]

35. Cheng, J.C.; Wu, J.K.; Lee, P.C.T.; Liu, H.S.; Jian, J.J.; Lin, Y.M.; Sung, J.L.; Jan, G.J. Biologic susceptibility of hepatocellular carcinoma patients treated with radiotherapy to radiation-induced liver disease. Int. J. Radiat. Oncol. Biol. Phys. 2004, 60, 1502–1509. [CrossRef]

36. Liang, S.-X.; Zhu, X.-D.; Xu, Z.-Y.; Zhu, J.; Zhao, J.-D.; Lu, H.-J.; Yang, Y.-L.; Chen, L.; Wang, A.-Y.; Fu, X.-L.; et al. Radiation-induced liver disease in three-dimensional conformal radiation therapy for primary liver carcinoma: The risk factors and hepatic radiation tolerance. Int. J. Radiat. Oncol. Biol. Phys. 2006, 65, 426–434. [CrossRef]

37. Lawrence, T.S.; Haken, R.K.T.; Kessler, M.L.; Robertson, J.M.; Lyman, J.T.; Lavigne, M.L.; Brown, M.B.; DuRoss, D.J.; Andrews, J.C.; Ensminger, W.D.; et al. The use of 3-D dose volume analysis to predict radiation hepatitis. Int. J. Radiat. Oncol. Biol. Phys. 1992, 23, 781–788. [CrossRef]

38. Dawson, L.A.; McGinn, C.J.; Normolle, D.; Haken, R.K.T.; Walker, S.; Ensminger, W.; Lawrence, T.S. Escalated Focal Liver Radiation and Concurrent Hepatic Artery Fluorodeoxyuridine for Unresectable Intrahepatic Malignancies. J. Clin. Oncol. 2000, 18, 2210–2218. [CrossRef]

39. Dawson, L.A.; Normolle, D.; Balter, J.M.; McGinn, C.J.; Lawrence, T.S.; Haken, R.K.T. Analysis of radiation-induced liver disease using the Lyman NTPC model. Int. J. Radiat. Oncol. Biol. Phys. 2002, 53, 810–821. [CrossRef]

40. Ben-Josef, E.; Normolle, D.; Ensminger, W.D.; Walker, S.; Tatro, D.; Haken, R.K.T.; Knol, J.; Dawson, L.A.; Pan, C.; Lawrence, T.S. Phase II Trial of High-Dose Conformal Radiation Therapy with Concurrent Hepatic Artery Floxuridine for Unresectable Intrahepatic Malignancies. J. Clin. Oncol. 2005, 23, 8739–8747. [CrossRef]

41. Blomgren, H.; Lax, L.; Näslund, I.; Svansström, R. Stereotoxic High Dose Fraction Radiation Therapy of Extracranial Tumors Using an Accelerator: Clinical experience of the first thirty-one patients. Acta Oncol. 1995, 34, 861–870. [CrossRef] [PubMed]
42. Herfarth, K.K.; Debus, J.; Lohr, F.; Bahner, M.L.; Rhein, B.; Fritz, P.; Höss, A.; Schlegel, W.; Wannenmacher, M.F. Stereotactic Single-Dose Radiation Therapy of Liver Tumors: Results of a Phase I/II Trial. J. Clin. Oncol. 2001, 19, 164–170. [CrossRef] [PubMed]
43. Romero, A.M.; Wunderink, W.; Hussain, S.M.; De Pooter, J.A.; Heijmen, B.J.; Nowak, P.C.; Nuyttens, J.J.; Brandwijk, R.P.; Verhoef, C.; Ijzermans, J.N.; et al. Stereotactic body radiation therapy for primary and metastatic liver tumors: A single institution phase i–ii study. Acta Oncol. 2006, 45, 831–837. [CrossRef] [PubMed]
44. Rusthoven, K.E.; Kavanagh, B.D.; Cardenes, H.; Stieber, V.W.; Burri, S.H.; Feigenberg, S.J.; Chidel, M.A.; Pugh, T.J.; Franklin, W.; Kane, M.; et al. Multi-Institutional Phase I/II Trial of Stereotactic Body Radiation Therapy for Liver Metastases. J. Clin. Oncol. 2009, 27, 1572–1578. [CrossRef] [PubMed]
45. Lee, M.T.; Kim, J.J.; Dinniwell, R.; Brierley, J.; Lockwood, G.; Wong, R.; Cummings, B.; Ringash, J.; Tse, R.V.; Knox, J.J.; et al. Phase I Study of Individualized Stereotactic Body Radiotherapy of Liver Metastases. J. Clin. Oncol. 2009, 27, 1585–1591. [CrossRef]
46. Rule, W.; Timmerman, R.; Tong, L.; Abdulrahman, R.; Meyer, J.; Boike, T.; Schwarz, R.E.; Weatherall, P.; Chinsoo Cho, L. Phase I Dose-Escalation Study of Single-Fraction Stereotactic Body Radiotherapy in Patients with Hepatic Metastases. Ann. Surg. Oncol. 2010, 18, 1081–1087. [CrossRef]
47. Comito, T.; Cozzi, L.; Clerici, E.; Campisi, M.C.; Liardo, R.L.; Navarra, P.; Ascolese, A.; Tozzi, A.; Iftode, C.; De Rose, F.; et al. Stereotactic Ablative Radiotherapy (SABR) in inoperable oligometastatic disease from colorectal cancer: A safe and effective approach. BMC Cancer 2014, 14, 619. [CrossRef]
48. Scorsetti, M.; Comito, T.; Tozzi, A.; Navarra, P.; Fogliata, A.; Clerici, E.; Mancosu, P.; Reggiori, G.; Rimassa, L.; Torzilli, G.; et al. Final results of a phase II trial for stereotactic body radiotherapy for patients with inoperable liver metastases from colorectal cancer. J. Cancer Res. Clin. Oncol. 2015, 141, 543–553. [CrossRef]
49. Goodman, K.A.; Wiegner, E.A.; Maturen, K.E.; Zhang, Z.; Mo, Q.; Yang, G.; Gibbs, I.C.; Fisher, G.A.; Koong, A.C. Dose-Escalation Study of Single-Fraction Stereotactic Body Radiotherapy for Liver Malignancies. Int. J. Radiat. Oncol. Biol. Phys. 2010, 78, 486–493. [CrossRef]
50. Meyer, J.J.; Foster, R.D.; Lev-Cohain, N.; Yokoo, T.; Dong, Y.; Schwarz, R.E.; Rule, W.; Tian, J.; Xie, Y.; Hannan, R.; et al. A Phase I Dose-Escalation Trial of Single-Fraction Stereotactic Body Radiotherapy for Liver Metastases. Ann. Surg. Oncol. 2016, 23, 218–224. [CrossRef]
51. Hong, T.S.; Wo, J.Y.; Borger, D.R.; Yeap, B.Y.; McDonnell, E.I.; Willers, H.; Blaszkowsky, L.S.; Kwak, E.L.; Allen, J.N.; Clark, J.W.; et al. Phase II Study of Proton-Based Stereotactic Body Radiation Therapy for Liver Metastases: Importance of Tumor Genotype. J. Natl. Cancer Inst. 2017, 109. [CrossRef]
52. Scorsetti, M.; Comito, T.; Clerici, E.; Franzese, C.; Tozzi, A.; Iftode, C.; Di Brina, L.; Navarra, P.; Mancosu, P.; Reggiori, G.; et al. Phase II trial on SBRT for unresectable liver metastases: Long-term outcome and prognostic factors of survival after 5 years of follow-up. Radiat. Oncol. 2018, 13, 234. [CrossRef]
53. Kang, J.I.; Suffiço, D.C.; Hsueh, C.-T.; Wroe, A.J.; Patyal, B.; Reeves, M.E.; Slater, J.D.; Yang, G.Y. A phase I trial of Proton stereotactic body radiation therapy for liver metastases. J. Gastrointest. Oncol. 2019, 10, 112–117. [CrossRef]
54. Dawson, L.A.; Winter, K.A.; Katz, A.W.; Schell, M.C.; Brierley, J.; Chen, Y.; Kopec, N.; Crane, C.H.; Willett, C.G. NRG Oncology/RTOG 0438: A Phase 1 Trial of Highly Conformal Radiation Therapy for Liver Metastases. Pract. Radiat. Oncol. 2019, 9, e386–e393. [CrossRef]
55. Scheffer, T.E.; Kavanagh, B.D.; Timmerman, R.D.; Cardenes, H.R.; Baron, A.; Gaspar, L.E. A phase I trial of stereotactic body radiation therapy for liver metastases. Int. J. Radiat. Oncol. Biol. Phys. 2005, 62, 1371–1378. [CrossRef] [PubMed]
56. Klement, R.J. Radiobiological parameters of liver and lung metastases derived from tumor control data of 3719 metastases. Radiother. Oncol. 2017, 123, 218–226. [CrossRef] [PubMed]
57. Mahadevan, A.; Blanck, O.; Lanciano, R.; Peddada, A.; Sundaraman, S.; D’Ambrosio, D.; Sharma, S.; Perry, D.; Kolker, J.; Davis, J. Stereotactic Body Radiotherapy (SBRT) for liver metastasis—Clinical outcomes from the international multi-institutional RSSearch® Patient Registry. Radiat. Oncol. 2018, 13, 26. [CrossRef] [PubMed]
58. Wurm, R.E.; Gum, F.; Erbel, S.; Schenger, L.; Scheffler, D.; Agaoglu, D.; Schild, R.; Gebauer, B.; Rogalla, P.; Plotkin, M.; et al. Image guided respiratory gated hypofractionated Stereotactic Body Radiation Therapy (H-SBRT) for liver and lung tumors: Initial experience. Acta Oncol. 2006, 45, 881–889. [CrossRef]
59. Kirilova, A.; Lockwood, G.; Choi, P.; Bana, N.; Haider, M.A.; Brock, K.K.; Eccles, C.; Dawson, L.A. Three-Dimensional Motion of Liver Tumors Using Cine-Magnetic Resonance Imaging. Int. J. Radiat. Oncol. Biol. Phys. 2008, 71, 1189–1195. [CrossRef]
60. Worm, E.S.; Høyer, M.; Fledelius, W.; Poulsen, P.R. Three-dimensional, Time-Resolved, Intrafraction Motion Monitoring Throughout Stereotactic Liver Radiation Therapy on a Conventional Linear Accelerator. Int. J. Radiat. Oncol. Biol. Phys. 2013, 86, 190–197. [CrossRef]
61. Vogel, L.; Sihono, D.S.K.; Weiss, C.; Lohr, F.; Stieler, F.; Wertz, H.; Von Schwietochowski, S.; Simeonova-Chergou, A.; Wenz, F.; Blessing, M.; et al. Intra-breath-hold residual motion of image-guided DIBH liver-SBRT: An estimation by ultrasound-based monitoring correlated with diaphragm position in CBCT. Radiother. Oncol. 2018, 129, 441–448. [CrossRef] [PubMed]
62. Briere, T.M.; Beddar, S.; Balter, P.; Murthy, R.; Gupta, S.; Nelson, C.; Starkschall, G.; Gillin, M.T.; Krishnan, S. Respiratory gating with EPID-based verification: The MDACC experience. Phys. Med. Biol. 2009, 54, 3379–3391. [CrossRef] [PubMed]
63. Heinzerling, J.H.; Anderson, J.F.; Papiez, L.; Boite, T.; Chien, S.; Zhang, G.; Abdulrahman, R.; Timmerman, R. Four-Dimensional Computed Tomography Scan Analysis of Tumor and Organ Motion at Varying Levels of Abdominal Compression During Stereotactic Treatment of Lung and Liver. *Int. J. Radiat. Oncol. Biol. Phys.* 2008, 70, 1571–1578. [CrossRef]

64. Case, R.B.; Moseley, D.J.; Sonke, J.J.; Eccles, C.L.; Dinniwel, R.E.; Kim, J.; Bezjak, A.; Milosevic, M.; Brock, K.K.; Dawson, L.A. Interfraction and Intrafraction Changes in Amplitude of Breathing Motion in Stereotactic Liver Radiotherapy. *Int. J. Radiat. Oncol. Biol. Phys.* 2010, 77, 918–925. [CrossRef] [PubMed]

65. Case, R.B.; Sonke, J.J.; Moseley, D.J.; Kim, J.; Brock, K.K.; Dawson, L.A. Inter- and Intrafraction Variability in Liver Position in Non-Breath-Hold Stereotactic Body Radiotherapy. *Int. J. Radiat. Oncol. Biol. Phys.* 2009, 75, 302–308. [CrossRef]

66. Yang, J.; Cai, J.; Wang, H.; Chang, Z.; Czito, B.G.; Bashir, M.R.; Palta, M.; Yin, F.-F. Is Diaphragm Motion a Good Surrogate for Liver Tumor Motion? *Int. J. Radiat. Oncol. Biol. Phys.* 2014, 90, 952–958. [CrossRef]

67. Wunderink, W.; Romero, A.M.; Seppenwoolde, Y.; De Boer, H.; Levendag, P.; Heijmen, B. Potentials and Limitations of Guiding Liver Stereotactic Body Radiation Therapy Set-Up on Liver-Implanted Fiducial Markers. *Int. J. Radiat. Oncol. Biol. Phys.* 2010, 77, 1573–1583. [CrossRef]

68. Seppenwoolde, Y.; Wunderink, W.; Wunderink-van Veen, S.R.; Storchi, P.; Romero, A.M.; Heijmen, B.J. Treatment precision of image-guided liver SBRT using implanted fiducial markers depends on marker-tumour distance. *Phys. Med. Biol.* 2011, 56, 5445–5468. [CrossRef]

69. Crane, C.H.; Koay, E.J. Solutions that enable ablative radiotherapy for large liver tumors: Fractionated dose painting, simultaneous integrated protection, motion management, and computed tomography image guidance. *Cancer* 2016, 122, 1974–1986. [CrossRef]

70. Bujold, A.; Massey, C.A.; Kim, J.J.; Brierley, J.; Cho, C.; Wong, R.K.; Dinniwel, R.E.; Kassam, Z.; Ringash, J.; Cummings, B.; et al. Sequential Phase I and II Trials of Stereotactic Body Radiotherapy for Locally Advanced Hepatocellular Carcinoma. *J. Clin. Oncol.* 2013, 31, 1631–1639. [CrossRef]

71. Mondlane, G.; Gubanski, M.; Lind, P.A.; Henry, T.; Ureba, A.; Siegbahn, A. Dosimetric Comparison of Plans for Photon- or Proton-Beam Based Radiosurgery of Liver Metastases. *Int. J. Part. Ther.* 2016, 3, 277–284. [CrossRef] [PubMed]

72. Colbert, L.E.; Cloyd, J.M.; Koay, E.J.; Crane, C.H.; Vauthey, J.N. Proton beam radiation as salvage therapy for bilateral colorectal cancer with 4D-MRI based online adaptation and real-time MRI monitoring using a 1.5 Tesla MR-Linac. *Cureus* 2019, 12, e11–e21. [CrossRef] [PubMed]

73. Sanford, N.N.; Pursley, J.; Noe, B.; Yeap, B.Y.; Goyal, L.; Clark, J.W.; Allen, J.N.; Blaszkowsky, L.S.; Ryan, D.P.; Ferrone, C.R.; et al. A Multi-Institutional Experience of MR-Guided Liver Stereotactic Body Radiation Therapy. *Adv. Radiat. Oncol.* 2019, 4, 142–149. [CrossRef]

74. Fast, M.; van de Schoot, A.; van de Lindt, T.; Carbaat, C.; van der Heide, U.; Sonke, J.-J. Tumor Trailing for Liver SBRT on the MR-Linac. *Int. J. Radiat. Oncol. Biol. Phys.* 2019, 103, 468–478. [CrossRef]

75. Feldman, A.M.; Modh, A.; Glide-Hurst, C.; Chetty, I.J.; Movsas, B. Real-time Magnetic Resonance-guided Liver Stereotactic Body Radiation Therapy: An Institutional Report Using a Magnetic Resonance-Linac System. *Cureus* 2019, 11, e5774. [CrossRef]

76. Heinzerling, J.H.; Anderson, J.F.; Papiez, L.; Boike, T.; Woychik, N.; Zlatkis, J.; Schmoll, S.; Abdulrahman, R.; Timmerman, R. Four-Dimensional Computed Tomography Scan Analysis of Tumor and Organ Motion at Varying Levels of Abdominal Compression During Stereotactic Treatment of Lung and Liver. *Int. J. Radiat. Oncol. Biol. Phys.* 2008, 70, 1571–1578. [CrossRef]

77. Sahin, B.; Mustafayev, T.Z.; Gungor, G.; Aydin, G.; Yapici, B.; Atalar, B.; Ozyar, E. First 500 Fractions Delivered with a Magnetic Resonance-guided Radiotherapy System: Initial Experience. *Cureus* 2019, 12, e6457. [CrossRef]

78. Rosenberg, S.A.; Henke, L.E.; Shaverdian, N.; Mittauera, K.; Wojcieszynski, A.P.; Hullett, C.R.; Kamravab, M.; Lamb, J.; Cao, M.; Green, O.L.; et al. A Multi-Institutional Experience of MR-Guided Liver Stereotactic Body Radiation Therapy. *Adv. Radiat. Oncol.* 2019, 4, 142–149. [CrossRef]

79. Hal, W.A.; Straza, M.W.; Chen, X.; Mickevicius, N.; Erickson, B.; Schultz, C.; Awana, M.; Ahunbay, E.; Li, X.A.; Paulson, E.S. Initial clinical experience of Stereotactic Body Radiotherapy (SBRT) for liver metastases, primary liver malignancy, and pancreatic cancer with 4D-MRI based online adaptation and real-time MRI monitoring using a 1.5 Tesla MR-Linac. *PLoS ONE* 2020, 15, e0236570. [CrossRef]

80. Chetty, I.J.; Rosu-Bubulac, M. Deformable Registration for Dose Accumulation. *Semin. Radiat. Oncol.* 2019, 29, 198–208. [CrossRef]

81. Winkel, D.; Bol, G.H.; Kroon, P.S.; Van Asselen, B.; Hackett, S.S.; Werenstein-Honingh, A.M.; Intven, M.P.; Eppinga, W.S.C.; Tijssen, R.H.N.; Kerkmeijer, L.G.W.; et al. Adaptive radiotherapy: The Elekta Unity MR-linac concept. *Clin. Transl. Radiat. Oncol.* 2019, 18, 54–59. [CrossRef]

82. Rosenberg, S.A.; Henke, L.E.; Shaverdian, N.; Mittauera, K.; Wojcieszynski, A.P.; Hullett, C.R.; Kamravab, M.; Lamb, J.; Cao, M.; Green, O.L.; et al. A Multi-Institutional Experience of MR-Guided Liver Stereotactic Body Radiation Therapy. *Adv. Radiat. Oncol.* 2019, 4, 142–149. [CrossRef]

83. Winkel, D.; Bol, G.H.; Kroon, P.S.; Van Asselen, B.; Hackett, S.S.; Werenstein-Honingh, A.M.; Intven, M.P.; Eppinga, W.S.C.; Tijssen, R.H.N.; Kerkmeijer, L.G.W.; et al. Adaptive radiotherapy: The Elekta Unity MR-linac concept. *Clin. Transl. Radiat. Oncol.* 2019, 18, 54–59. [CrossRef]

84. Hal, W.A.; Straza, M.W.; Chen, X.; Mickevicius, N.; Erickson, B.; Schultz, C.; Awana, M.; Ahunbay, E.; Li, X.A.; Paulson, E.S. Initial clinical experience of Stereotactic Body Radiotherapy (SBRT) for liver metastases, primary liver malignancy, and pancreatic cancer with 4D-MRI based online adaptation and real-time MRI monitoring using a 1.5 Tesla MR-Linac. *PLoS ONE* 2020, 15, e0236570. [CrossRef]
86. Bruijnen, T.; Stemkens, B.; Lagendijk, J.J.W.; van den Berg, C.A.T.; Tijssen, R.H.N. Multiresolution radial MRI to reduce IDLE time in pre-beam imaging on an MR-Linac (MR-RIDDLE). *Phys. Med. Biol.* 2019, 64, 055011. [CrossRef] [PubMed]

87. Fiz, F.; Viganò, L.; Gennaro, N.; Costa, G.; La Bella, L.; Boichuk, A.; Cavinato, L.; Sollini, M.; Politi, L.S.; Chiti, A.; et al. Radiomics of Liver Metastases: A Systematic Review. *Cancers* 2020, 12, 2881. [CrossRef] [PubMed]

88. Shaverdian, N.; Yang, Y.; Hu, P.; Hart, S.; Sheng, K.; Lamb, J.; Cao, M.; Agazaryan, N.; Thomas, D.; Steinberg, M.; et al. Feasibility evaluation of diffusion-weighted imaging using an integrated MRI-radiotherapy system for response assessment to neoadjuvant therapy in rectal cancer. *Br. J. Radiol.* 2017, 90, 20160739. [CrossRef] [PubMed]

89. Yang, Y.; Cao, M.; Sheng, K.; Gao, Y.; Chen, A.; Kamrava, M.; Lee, P.; Agazaryan, N.; Lamb, J.; Thomas, D.; et al. Longitudinal diffusion MRI for treatment response assessment: Preliminary experience using an MRI-guided tri-cobalt 60 radiotherapy system. *Med. Phys.* 2016, 43, 1369–1373. [CrossRef] [PubMed]

90. Kooreman, E.S.; van Houdt, P.J.; Nowee, M.E.; van Pelt, V.W.J.; Tijssen, R.H.N.; Paulson, E.S.; Gurney-Champion, O.J.; Wang, J.; Koetsveld, F.; van Buuren, L.D.; et al. Feasibility and accuracy of quantitative imaging on a 1.5 T MR-linear accelerator. *Radiother. Oncol.* 2019, 133, 156–162. [CrossRef] [PubMed]

91. Cusumano, D.; Boldrini, L.; Yadav, P.; Casà, C.; Lee, S.L.; Romano, A.; Piras, A.; Chiloiro, G.; Placidi, L.; Catucci, F.; et al. Delta Radiomics Analysis for Local Control Prediction in Pancreatic Cancer Patients Treated Using Magnetic Resonance Guided Radiotherapy. *Diagnostics* 2021, 11, 72. [CrossRef] [PubMed]

92. Vandecaveye, V.; Dirix, P.; De Keyzer, F.; De Beeck, K.O.; Vander Poorten, V.; Roebben, I.; Nuys, S.; Hermans, R. Predictive value of diffusion-weighted magnetic resonance imaging during chemoradiotherapy for head and neck squamous cell carcinoma. *Eur. Radiol.* 2010, 20, 1703–1714. [CrossRef] [PubMed]

93. Cai, G.; Xu, Y.; Zhu, J.; Gu, W.-L.; Zhang, S.; Ma, X.-J.; Cai, S.-J.; Zhang, Z. Diffusion-weighted magnetic resonance imaging for predicting the response of rectal cancer to neoadjuvant concurrent chemoradiation. *World J. Gastroenterol.* 2013, 19, 5520–5527. [CrossRef] [PubMed]

94. Van Rossum, P.S.; Van Lier, A.L.; Van Vulpen, M.; Reerink, O.; Lagendijk, J.J.; Lin, S.H.; Van Hillegersberg, R.; Ruurda, J.P.; Meijer, G.J.; Lips, I.M. Diffusion-weighted magnetic resonance imaging for the prediction of pathologic response to neoadjuvant chemoradiotherapy in esophageal cancer. *Radiother. Oncol.* 2015, 115, 163–170. [CrossRef] [PubMed]