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Cytoplasmic Incompatibility Variations in Relation with Wolbachia cid Genes Divergence in Culex pipiens

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ABSTRACT In arthropods, Wolbachia endosymbionts induce conditional sterility, called cytoplasmic incompatibility (CI), resulting from embryonic lethality. CI penetrance (i.e., embryonic death rate) varies depending on host species and Wolbachia strains involved. All Culex pipiens mosquitoes are infected by the endosymbiotic alphaproteobacteria Wolbachia wPip. CI in Culex, characterized as a binary “compatible/incompatible” phenomenon, revealed an unparalleled diversity of patterns linked to the amplification-diversification of cidA and cidB genes. Here, we accurately studied CI penetrance variations in the light of cid genes divergence by generating a C. pipiens compatibility matrix between 11 lines hosting different phylogenetic wPip groups and exhibiting distinct cid gene repertoires. We showed, as expected, that crosses involving wPip from the same group were mostly compatible. In contrast, only 22% of the crosses involving different wPip groups were compatible, while 54% were fully incompatible. For the remaining 24% of the crosses, “intermediate” compatibilities were reported, and a cytological observation of the first zygotic division confirmed the occurrence of “canonical” CI phenotypes in a fraction of the eggs. Backcross experiments demonstrated that intermediate compatibilities were not linked to host genetic background but to the Wolbachia strains involved. This previously unstudied intermediate penetrance CI was more severe and frequent in crosses involving wPip-IV strains exhibiting cid variants markedly divergent from other wPip groups. Our data demonstrate that CI is not always a binary compatible/incompatible phenomenon in C. pipiens but that intermediate compatibilities putatively resulting from partial mismatch due to Cid proteins divergence exist in this species complex.

IMPORTANCE Culex pipiens mosquitoes are infected with wPip. These endosymbionts induce a conditional sterility called CI resulting from embryonic deaths, which constitutes a cornerstone for Wolbachia antivectorial methods. Recent studies revealed that (i) two genes, cidA and cidB, are central in Wolbachia-CI mechanisms, and (ii) compatibility versus incompatibility between mosquito lines depends on the wPip phylogenetic groups at play. Here, we studied CI variations in relation to wPip groups and cid genes divergence. We showed, as expected, that the crosses involving wPip from the same group were compatible. In contrast, 78% of the crosses involving different wPip groups were partially or fully incompatible. In such crosses, we reported defects during the first zygotic division, a hallmark of CI. We showed that CI was more severe and frequent in crosses involving wPip-IV strains exhibiting cid variants, which markedly diverge from those of other wPip groups.

KEYWORDS Culex pipiens, toxin-antitoxin system, Wolbachia, developmental biology, endosymbionts, gene amplification, vectors
In many arthropods, the fertility of two sexual partners undergoes acute reduction due to the presence of the intracellular alphaproteobacteria Wolbachia (1). This conditional sterility, depending on the presence of cytoplasmic factors, is called cytoplasmic incompatibility (CI). CI primarily occurs within crosses between males infected with Wolbachia and uninfected females, thus exhibiting reduced fertility compared to the infected ones. Such “reproductive manipulation” induced by Wolbachia promotes the spread of the infection (2). The loss of fertility for uninfected females, while infected females reproduce well, confers an advantage for Wolbachia transmission, which is the cornerstone of CI evolution. Such loss of fertility does not result from reduced egg production but from a high rate of early embryonic mortality (3, 4). Cytological embryonic observations demonstrated in Culex, Drosophila, and Nasonia that CI induced by Wolbachia is precisely due to defects in paternal chromatin during first zygotic division, suggesting a chromatin modification by some Wolbachia factors (5–9). Such defects during the first embryonic division can be prevented if Wolbachia are present in the eggs. This cytological characterization of the hallmarks of CI has contributed to the formulation of the modification-rescue (mod-resc) model that could putatively be based on toxin-antidote interactions where a toxin (the mod factor) produced by the paternal Wolbachia and introduced in the sperm induces embryonic mortality unless an antidote (the resc factor) is produced by the maternal Wolbachia in the eggs (3).

Recent studies pointed out pairs of adjacent genes called CI factors (cif), within the genomes of CI-inducing Wolbachia, as major molecular actors of CI (10–15). Cif is their general name, while cid or cin are specific names based on their enzymatic domains (deubiquitinase [DUB] for cid and nuclease for cin [16, 17]). The heterologous expression of either cid or cin pairs (each composed of A/B genes) in Drosophila melanogaster males induces early death for a significant number of the embryos when crossed with uninfected females (11, 12, 14). However, the abortive embryo proportion due to CI, also called CI penetrance, varies depending on the cif transgenes. In similar expression conditions, the cid genes induced stronger CI than cin ones (12, 14, 15). Moreover, differences in CI penetrance between the different cid alleles introduced in D. melanogaster have been reported: the cidA/BwPip genomes from Culex pipiens (10–12), induced full CI (i.e., null hatching rate [HR], with HR equal to the proportion of hatched eggs) while cidA/BwMel factors (from the wMel genome) only induced a significant decrease in HR (12, 15). Differences in CI penetrance were also reported between wMel and wPip, harboring different cifA and cifB genes, in the natural context of their native hosts. Indeed, in C. pipiens, all wPip strains induced full CI when infected males were crossed with uninfected females (18, 19) while wMel in Drosophila induced a partial HR reduction (12, 20). In C. pipiens, full CI occurs regardless of male age (21, 22), host genetic background (23, 24), or Wolbachia densities (9, 22). The cumulative presence of both functional cid and cin genes (17, 19, 25, 26) and the massive amplification-diversification of cid genes (9, 19, 27) provided putative genomic bases for this full CI induction. Indeed, unlike Wolbachia strains found in other host species where cid genes are monomorphic, each wPip strain encodes a “repertoire” of cid genes, with up to 6 different variants of cidA and cidB genes in a single Wolbachia genome (19, 27).

The strength of wPip-induced CI represents a force that certainly promoted the initial fixation and the maintenance of wPip in the C. pipiens complex (28). All C. pipiens individuals are currently infected with Wolbachia strains belonging to the monophyletic clade of wPip that is diversified into five groups, wPip-I to wPip-V (29). This diversity of wPip strains is responsible for the unparalleled diversity of CI patterns in the C. pipiens complex described as a binary “compatible/incompatible” phenomenon (30, 31). Indeed, hundreds of crosses between C. pipiens lines from different geographical origins all infected with wPip revealed the following two major outcomes based on their HR (21–23, 29, 31–34): (i) compatible crosses, with 80% ≤ mean HR ≤ 100%; in these cases, the number of unhatched eggs is similar to those of intraline crosses; or (ii) fully incompatible crosses, with null HR except for very few eggs (18, 21, 22, 34). In
the latter situation, incompatibility can be either unidirectional (one cross direction is incompatible, while the reciprocal cross is compatible) or bidirectional (both cross directions are incompatible) (32, 35, 36). Reconstruction of wPip phylogeny revealed that mosquitoes infected with strains from the same group are more likely to be compatible with each other, while the compatibility between host-harboring wPip strains from different groups is mostly unpredictable (31). Moreover, specific variations in cidB repertoires harbored by males correlated with compatibility/incompatibility variations between C. pipiens lines, suggesting that some specific variants may play a strong role in this “yes-or-no” CI (19, 27). However, few cases were also reported with intermediate HR, i.e., 10% ≤ mean HR ≤ 80%, without knowing if those intermediate HR were linked to the Wolbachia strains involved or other factors such as nuclear incompatibilities (30, 37–43). Indeed, at the time of these intermediate HR observations, no diversity between wPip strains was discovered, and it was not possible to decipher the part of nuclear genetic background versus Wolbachia in the observed intermediate HR.

Our recent reconstruction of wPip phylogenetic groups (29, 31) and discovery of cid genes’ amplification and diversification led us to correlate cid and “yes-or-no” CI diversities in C. pipiens (19, 27). In the present study, we accurately monitored CI penetrance variations in the light of cid genes divergence by generating a C. pipiens compatibility matrix involving 11 lines harboring Wolbachia strains belonging to different wPip groups (wPip-I to wPip-IV) and all harboring different cid repertoires (9, 19). This compatibility matrix is composed of estimated HR obtained from (i) 11 intraline crosses (INTRA), (ii) 12 crosses between lines harboring wPip strains from the same group (INTER-INTER), and (iii) 83 crosses between lines harboring wPip from different groups (INTER-INTRA). We showed, as expected, that all INTRA and INTER-INTRA (except two) crosses were fully compatible. Among the INTER-INTER crosses, 54% were totally incompatible, displaying no hatching, and 22% were considered fully compatible, while 24% of the crosses exhibited mean HRs that can be qualified as intermediate. Backcross experiments demonstrated that such intermediate HRs were not linked to host genetic background but to the Wolbachia strains involved. Moreover, we showed that intermediate HR values were particularly low within crosses involving wPip-IV strains that also present marked phylogenetic difference in their cid repertoires from other wPip groups (19). To visualize the developmental defects responsible for intermediate HR, we monitored the embryonic development and found defects during the first zygotic division and subsequent developmental arrest, which are typical hallmarks of “canonical CI” (9, 14). Altogether, our data demonstrate that CI is not always a “yes-or-no” phenomenon in C. pipiens but that subtle CI variations, referred to as “cryptic CI,” putatively resulting from partial mismatch due to Cif protein divergence, exist in this species complex.

RESULTS

HR in fully compatible crosses. Mean HR of the 11 INTRA crosses were comprised between 0.78 and 0.95, showing that an important part of the eggs (up to 22%) failed to develop even in INTRA crosses. Intermediate HR can thus only refer to crosses with mean HR ≤ 78% (Fig. 1; Table S1 in the supplemental material; Data Set S1).

Depriving lines from Wolbachia did not influence INTRA HR. To test for the effect of presence/absence of Wolbachia, two C. pipiens lines were tetracycline treated (SlabTC and IstanbulTC). For these “cured lines”, mean HRs were not significantly different from HRs of the corresponding INTRA crosses with infected lines (Wilcoxon \( W = 356, P = 0.168 \); and \( W = 344, P = 0.119 \) for Slab/SlabTC and Istanbul/IstanbulTC, respectively) (Table S1; Data Set S2).

No influence of host genetic backgrounds on HR. Crosses involving females harboring the same wPip strain in different genetic backgrounds (i.e. from backcrossed lines [Sl(wPip-I-Tunis) and Sl(wPip-IV-Harash)]) did not differ in their HRs when crossed with males from seven different lines (generalized linear models with mixed effects [GLMM]; \( \chi^2 = 2.857 \), degrees of freedom [df] = 1, \( P = 0.091 \)). Crosses involving males harboring the same wPip strain in different genetic backgrounds showed similar
HRs when crossed with females from five different lines (GLMM; $\chi^2 = 0.414, \text{df} = 1, P = 0.520; \chi^2 = 0.0137, \text{df} = 1, P = 0.907$; Table S1). Moreover, reciprocal crosses involving different C. p. species (i.e., Culex quinquefasciatus [Slab] versus C. p. [Istanbul]) without Wolbachia were not significantly different from corresponding intraspecies crosses (Wilcoxon $W = 216, P = 0.764$ and $W = 185, P = 0.327$, respectively; Data Set S3).

**INTER-INTER crosses exhibit significantly reduced HR.** The full distribution of HR per egg raft for all the crosses is presented in Fig. 1A. The mean HR (i.e., calculated on
of INTRA crosses ranged from 0.78 to 0.95; the mean HR of INTER-INTRA crosses (except for two fully incompatible crosses) ranged from 0.75 to 0.93, while the mean HR of INTER-INTRA crosses displayed much more variability, ranging from 0 to 0.96. Fifty-four percent (45/83) of the INTER-INTER crosses were actually fully incompatible, while 46% (38/83) produced numerous larvae (mean HR between 0.48 and 0.96). HR distributions differed significantly among the different cross types (TYPE parameter in the statistical model) that led to larval production, as follows: (i) all the 11 INTRA (330 eggs rafts analyzed for a total of 47,504 eggs), (ii) all the 12 INTER-INTRA (360 egg rafts analyzed for a total of 49,461 eggs), and (iii) 38 out of the 83 INTER-INTER (1,140 egg rafts analyzed for a total of 169,215 eggs [Fig. 1A]). HR from INTER-INTER crosses were significantly lower than others (GLMM; χ² = 8.0371, df = 2, P = 0.018; Fig. 1A). Furthermore, the variance in HR per egg raft was significantly higher in INTER-INTER crosses (Levene’s test, P < 0.001), while it did not differ between INTRA and INTER-INTRA (Levene’s test, P = 0.65; Fig. 1A).

The INTER-INTER crosses category shows a higher occurrence of intermediate HR. Among the 38 INTER-INTER crosses in which eggs hatched (Data Set S4), 20 crosses displayed a mean HR below 78%, referred to as intermediate HR, while only 1 cross out of 12 in the INTER-INTRA showed such intermediate values. INTER-INTER crosses showed significantly more intermediate HR crosses than other types (chi-square test; χ² = 7.346, df = 1, P = 0.006; Table S1).

The lowest HRs were observed in INTER-INTER crosses involving wPip-IV strains. For the 38 INTER-INTER crosses which were not fully incompatible, global models did not reveal any significant effect of the wPip group hosted by either female or male lines (GLMM; χ² = 0.268, df = 3, P = 0.966; χ² = 2.742, df = 3, P = 0.433, respectively) but pointed out a significant interaction effect between the wPip groups involved in the crosses (generalized linear models [GLM]; χ² = 113.764, df = 13, P < 0.001; for detailed statistics, see Text S1). Careful inspection of the HR matrix revealed that 8 INTER-INTER crosses out of 38 showed a mean HR below 60%, here called low HR (Fig. 1B; Table S1). All these eight INTER-INTER crosses with backcrossed line S(wPip-IV-Harash) did not differ from crosses involving Harash lines (GLM; χ² = 0.0137, df = 1, P = 0.907), demonstrating that it was the wPip-IV strain harbored in the cytoplasm and not the host genetic background that explained such a low HR.

Intermediate HR results from cryptic but canonical CI. As low HRs (mean HR under 0.6) were only observed in INTER-INTER crosses involving wPip-IV strains, we (i) studied the first zygotic division resulting from these crosses, and (ii) in an attempt to quantify putative CI defects, compared them with INTER-INTRA and INTRA crosses at 5 h (Table 1 and Table S2). To verify whether intermediate HRs were due to previously described canonical CI cellular mechanisms (5–9), we visualized the first zygotic division with paternal and maternal chromatin labeled in green/yellow and red, respectively. In INTER-INTER crosses with intermediate HR, an important proportion of eggs normally hatched. Such normal embryogenesis, as documented in Fig. 2, is similar to what was observed for all INTRA embryos previously documented (9). After

**TABLE 1** Proportion of embryos that did not reach normal blastoderm stage 5 h postoviposition in one INTRA, one INTER-INTRA, and one INTER-INTER cross

| Cross (male x female) | Cross type | No. of blastoderm-stage embryos | No. of embryos with abnormal development | No. of embryos with no sign of development | Total no. of embryos | % of embryos that did not reach blastoderm stage (5 h postoviposition) |
|-----------------------|------------|-------------------------------|----------------------------------------|------------------------------------------|---------------------|--------------------------------------------------|
| Tunis x Tunis         | INTRA      | 94                            | 0                                      | 5                                        | 99                  | 5                                                 |
| Ichkeul-13 x Harash   | INTER-INTRA| 46                            | 0                                      | 1                                        | 47                  | 2                                                 |
| Ichkeul-13 x S(wPip-I-Tunis) | INTER-INTRA | 36                           | 5                                      | 4                                        | 45                  | 20                                                |
| Ichkeul-13 x Tunis    | INTER-INTRA| 105                           | 20                                     | 20                                       | 145                 | 28                                                |

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fertilization, maternal and paternal pronuclei migrated toward each other and apposed (documented embryos with confocal microscopy images, \( n = 3 \); Fig. 2A). Then, paternal and maternal chromatins condensed and entered into the first zygotic division (\( n = 1 \); Fig. 2B). During the first mitotic division, paternal and maternal chromosomes aligned in separate regions at the metaphase plate (\( n = 2 \), Fig. 2C). Both sets of chromosomes segregated equally during anaphase (\( n = 2 \); Fig. 2D) to produce two diploid nuclei (\( n = 1 \); Fig. 2E). Although our observations of first zygotic division events are not quantitative due to the technical challenge to monitor the different steps of this fast process, observations of embryos’ early development in INTER-INTER crosses with intermediate HR enabled us to document the presence of first zygotic division defects (\( n = 4 \); Fig. 3) that were previously observed in fully incompatible INTER-INTER crosses and absent in INTRA ones (9). As it was the green-labeled chromatin that exhibited such defects, it can be concluded that paternal chromatin is affected (Fig. 3A, A’, B, C, and D).

For only three crosses involving wPip-I and wPip-IV strains (one INTRA between Tunis wPip-I infected individuals; one INTER-INTRA between Ichkeul-13 wPip-IV males and Harash wPip-IV strain females; one INTER-INTER between Harash wPip-IV males and Tunis wPip-I females), we were able to produce enough observable embryos to assess the proportion of embryos with abnormal development 5 h postoviposition as presented in Table 1. At this time, embryos should have reached the syncytial blastoderm stage (~3,200 “normal” nuclei; Fig. 4A and C), while embryos considered “abnormal” only presented few nuclei (less than 50; Fig. 4B). Moreover, atypical mitotic features were observed in these abnormal embryos (Fig. 4D and E). The proportion of abnormal embryos was less than 6% in INTRA and INTER-INTRA crosses while reaching at least 20% in the INTER-INTER cross with intermediate HR (Table 1; chi-square test; \( \chi^2 = 29.998, \text{df} = 3, P < 0.001 \)).
variants from wPip-IV repertoires are divergent from those of other wPip groups. The phylogenetic cidA and cidB networks constructed with wPip strains repertoires showed that wPip strains from the wPip-IV group exhibited markedly divergent cidA and cidB variants. For both cidA and cidB variants, wPip-IV variants clustered remotely from other groups’ variants (Fig. 5; Tables S3 and S4). Two well-separated clusters of wPip-IV cidA variants appeared on the network, while all cidB variants clustered altogether (Fig. 5). For other wPip groups, no clear wPip-group-based clustering was observed (Fig. 5).

**DISCUSSION**

In arthropods in which CI is mainly studied between infected males and uninfected females, including major insect models such as *Drosophila* and *Nasonia*, CI penetrance was proved to depend on *Wolbachia* strains, their densities, host genetic background, age of the males, and environmental factors such as temperature (44–59). On the contrary, in *Culex pipiens* s.l. mosquitoes, these factors did not affect CI penetrance (19, 22–24, 33): full CI (hatching rate [HR], 0) is reported between infected males and uninfected females (cured of *Wolbachia* with antibiotics) whatever their geographical origin, age, or genetic background (9, 18, 23, 33). However, hundreds of crosses between lines infected with *Wolbachia* revealed unparalleled variations in CI patterns in *C. pipiens*. Two main opposite outcomes were observed: either the crosses were compatible (mean HR ≥ 80%) or incompatible, producing almost no larvae (mean HR < 0.01%) (18, 21–23, 31, 32, 34, 40). Early in the study of CI in *Culex*, backcross experiments demonstrated that the host genome does not influence the outcome of a given cross (9, 24, 32, 60). In the present study, we conducted backcross experiments for two of our lines and also performed crosses between cured individuals from different *C. pipiens* species, which again confirmed that host genetic background does not impact compatibility.
Most crosses and backcrosses showed that CI in *C. pipiens* is a binary compatible/incompatible phenotype under the sole control of *Wolbachia*. However, in the numerous articles that presented results of interline *C. pipiens* crosses from different parts of the world, rare cases of intermediate HR were reported (30, 37–43). At the time of these publications, all the *wPip* were considered clonal due to monomorphic genetic markers available (34, 61). Intermediate HRs were thus attributed to putative undiscovered *Wolbachia* variability (including different *wPip* sublines in the same laboratory line) and most probably to putative host “restorer” nuclear factors counteracting *Wolbachia* CI induction (40, 41, 62). In the present paper, we investigated these intermediate HR situations in light of our present knowledge of *wPip* genomes (19, 27, 29, 31). To that extent, we studied 106 crosses between 11 *C. pipiens* isofemale lines infected with different *wPip* strains from different groups (I to IV), each exhibiting different cidA-cidB repertoires (9, 19) (Table S3 in the supplemental material). Different types of crosses were performed, including (i) INTRA crosses between mosquitoes from the same line, (ii) INTER-INTRA crosses between mosquitoes infected with different strains from the same *wPip* group, and (iii) INTER-INTER crosses between mosquitoes infected with different *wPip* groups.

For the 11 INTRA crosses performed in this study, mean HRs were all comprised between 78% and 95%, showing that a significant proportion of eggs never hatched even in fully compatible crosses. Previous cytological observations of *C. pipiens* early development in seven INTRA crosses with or without *Wolbachia* (i.e., after antibiotics

**FIG 4** *Culex pipiens* embryos 5 h postoviposition in INTER-INTER crosses. Green/yellow (acetylated histone H4 labeling) and red (propidium iodide labeling). (A) Global view of a normal *C. pipiens* embryo having reached the expected syncytial stage. (B) Global view of an abnormal *C. pipiens* embryo exhibiting only few (less than 15) nuclei 5 h postoviposition. (C) Normal nuclei in a syncytial embryo. (D and E) Atypical mitotic features observed in abnormal embryos. Confocal stacks were obtained on embryos from several INTER-INTER crosses. Red dots (especially visible at the embryo’s poles in panel B) are propidium iodide-labeled *Wolbachia* in the embryo’s cytoplasm. Scale bar represents 10 μm.
treatment) did not detect any CI typical defects (9). Here, we reported no difference in HR in the same lines with or without Wolbachia, confirming that CI induced by Wolbachia is not responsible for the 5% to 22% of the eggs that did not reach the larval stage. Abortive eggs in INTRA crosses certainly resulted from imperfect fertilization and/or intrinsic mortality during development from eggs to larvae (9, 18, 40). The 12 INTER-INTRA crosses, involving lines from different locations but harboring the same wPip group, exhibited HRs similar to INTRA crosses, except for two cases of unidirectional incompatibility, again demonstrating that the wPip group is a major predictor of compatibility between C. pipiens lines (Fig. 1A; Table S1) (31).

Heterogeneity in compatibility clearly increased in INTER-INTER crosses (Table S1). Among the 83 performed here, we found that 54% of them were fully incompatible, while the other 46% (38/83) were fertile and exhibited HR comprised between 48% and 96%. Global HR statistical analyses, including all fertile crosses (11 INTRA, 12 INTER-INTRA, and 38 INTER-INTER crosses) showed that HR was significantly lower in INTER-INTER crosses and that variance in HRs among egg rafts was significantly higher in INTER-INTER than INTER-INTRA and INTRA crosses (Fig. 1A). Moreover, we found that 53% of the fertile INTER-INTER crosses actually exhibited HRs that were low enough to be characterized as intermediate. We also found that the interaction between the wPip groups infecting the male and female lines significantly influenced HR. Careful inspection of the HR matrix revealed that the crosses with a low HR below 60% (8 crosses out of the 20 with intermediate HR) were only observed in INTER-INTER crosses involving wPip-IV strains (Fig. 1B and Table S1). Atyame et al. (31) had already shown that wPip-IV group-infected C. pipiens lines exhibited markedly different crossing types from lines infected with other wPip groups. Network phylogenetic analyses of all the 34 cidA and 21 cidB different variants characterized in the wPip strains studied here revealed that cid-IV variants (especially cidB) were divergent, gathering in specific clusters, while other wPip groups are mixed altogether. This suggests that Cid proteins that are considered major effectors of CI (15, 17) are divergent in wPip-IV strains compared to other wPip groups (Fig. 5).

To investigate whether intermediate HR resulted from canonical CI, i.e., paternal chromatin defects during first zygotic division, we monitored the first stages of embryonic development in embryos from INTER-INTER crosses. In these crosses, even with low HR, many embryos exhibited normal development into larvae (Fig. 2). However, in a few embryos, we were able to document imperfect paternal chromatin segregation.

![Phylogenetic networks of the cidA and cidB genes](image)

**FIG 5** Phylogenetic networks of the cidA and cidB genes. Networks obtained with 34 cidA (A) and 21 cidB (B) nucleotide variants present in the repertoires of the 11 strains from the four phylogenetic wPip groups studied here. The networks were obtained using the neighbor-net method. Each edge (or set of parallel edges) corresponds to a split in the data set and has a length equal to the weight of the split. Incompatible splits produced by recombination are represented by boxes in the network. wPip-I cid variants are in green, wPip-II cid variants are in yellow, wPip-III cid variants in brown, and wPip-IV cid variants are in pink.
during the first zygotic division (Fig. 3). These embryonic defects, which were never observed in INTRA crosses (9), were similar to those reported in fully incompatible crosses (9). Such defects in the first zygotic division likely produced aneuploid nuclei which might disrupt further development or even arrest the embryogenesis. The proportion of embryos that did not reach blastoderm stage 5 h postoviposition, but presented instead few nuclei only, can be considered a quantitative proxy for the occurrence of CI defects during the first division. We observed a larger amount of abnormal developmental stages, 5 h postoviposition, in INTER-INTER crosses than the INTRA and INTER-INTRA crosses (Table 1). Abnormal embryos, which represented 20% of the embryos in the INTER-INTER crosses studied and 5% in the INTRA one (Table 1), displayed very few (or no) nuclei (Fig. 4B). These observations suggest that embryonic defects during the first division are responsible for the intermediate HR observed in the analyzed INTER-INTER cross (Table 1). The intermediate HR observed in INTER-INTER crosses could be attributed to cryptic CI (in that it has a weak penetrance) but canonical CI (in that it translates into the same cytological defects).

In the light of the toxin-antidote model of CI, penetrance would depend on the interaction between CidA, CidB, and their specific substrates, eventually leading to paternal chromatin defects or its rescue (15, 16, 63). In *C. pipiens*, as all wPip genomes encode a repertoire of several polymorphic variants of CidA and CidB (19, 27), full compatibility could result from multiple interactions between different CidA and CidB variants even in INTRA or INTER-INTRA crosses. In every *C. pipiens* male, several CidB proteins differing in their amino acid sequences might be introduced in the sperm and then in the egg during fertilization where several CidA proteins might also be present. Full compatibilities reported here in some INTER-INTER crosses involving different wPip groups with totally different CidA/CidB repertoires (Fig. 1) suggest (i) that strict specific interactions between cognate variants are not required for full compatibility, and (ii) a potential redundancy in the interaction between CidA/CidB variants. The intermediate HR resulting from cryptic CI in a given INTER-INTER cross can hypothetically result from partial rescue due to imperfect interactions between the CidA and the CidB from the two wPip strains repertoires. Since most of the embryos from intermediate HR crosses developed into living larvae, it certainly means that, in those individuals, CidB toxicity has been efficiently counteracted. On the contrary, in embryos exhibiting CI, CidB toxicity would not have been counteracted properly. This heterogeneity could be explained if embryo rescue depends on one or a few matching CidA variants which might be required in a larger quantity for the rescue to occur. However, it is possible that in certain eggs, the expression of the(se) CidA variant(s) would be too low to counteract the CidB toxicity. This would be especially true for neutralizing CidB proteins encoded by wPip-IV strains that show striking differences in their sequences from other wPip groups (Fig. 5). Less efficacy in the interactions between CidB-IV proteins and CidA from other groups could explain their higher probability to be involved in both (i) full incompatibility as reported in reference 31, and (ii) cryptic CI as reported here.

The interactions between the CidA and CidB repertoires encoded by wPip strains determine the developmental fate of each embryo of a given cross, normal development versus CI. CI penetrance (i.e., the proportion of embryos undergoing CI) in a given cross could then be determined by the diversity of *cidA/cidB* genes of the different wPip genomes hosted by the different *C. pipiens* lines, their expression levels, and the affinity between the resulting proteins.

**MATERIALS AND METHODS**

*Culex pipiens* lines. Eleven isofemale lines were used (Table S5 in the supplemental material). They differed in (i) their geographical origins, (ii) the species they belong to, (iii) the wPip group (I, II, III, or IV), and (iv) their *cid* repertoires. The *Wolbachia* group was checked by performing a pk1 PCR-restriction fragment length polymorphism (RFLP) test (64) on DNA extracted using cetyltrimethylammonium bromide (CTAB) protocol (65). Tetracycline-treated *Wolbachia*-free lines (TC lines), named SlabTC and IstanbulTC, were obtained from Slab- and Istanbul-infected lines as described in reference 33. The absence of *Wolbachia* was checked by PCR on a fragment of the *wsp* gene using the primers designed in
Sequenced variants (accession numbers given in Table S4) were aligned using the Muscle algorithm obtained by PCR cloning followed by Sanger sequencing as previously described in references 9 and 19. Wolbachia strains hosted by the 11 crossed lines were already published except for Brazil that has been performed in the MRI-CRBM platform. Embryos optical observations were performed at the CytoEvol facilities.

Supplemental material is available online only.

DATA SET S1, XLSX file, 0.04 MB.
DATA SET S2, XLSX file, 0.02 MB.
DATA SET S3, XLSX file, 0.04 MB.
DATA SET S4, XLSX file, 0.1 MB.
TEXT S1, DOCX file, 0.01 MB.
TABLE S1, XLSX file, 0.02 MB.
TABLE S2, DOCX file, 0.01 MB.
TABLE S3, DOCX file, 0.02 MB.
TABLE S4, DOCX file, 0.02 MB.
TABLE S5, DOCX file, 0.02 MB.

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