Research Article

Occurrence of Granulomas in Bovines: An Abattoir-Based Study

Sambath Uma, Madhavan Gopalakrishnan Nair, and Khub Chandra Varshney

Department of Veterinary Pathology, Rajiv Gandhi College of Veterinary and Animal Sciences, Kurumbapet, Puducherry 605 009, India

Correspondence should be addressed to Khub Chandra Varshney, headpath@yahoo.co.in

Received 30 November 2011; Accepted 25 December 2011

1. Introduction

Granulomas are tissue reactions typically elicited by persistent, indigestible pathogens, pathogen products, or inert foreign material. Granulomas are pathologically defined by the presence of macrophage-derived epithelioid cells and giant cells, whose organization can be modified by the cells of acquired immunity. The structural similarity in granulomas of diverse etiology suggests that a common set of pathophysiological signals regulates granulomatous inflammation. Observations that granulomas are of differing morphology and caused by a wide variety of irritants have led to numerous attempts to classify granulomatous inflammation in man, either to help in the diagnosis of granulomatous disease or to further the understanding of the granulomatous process [1]. However, in animals, there are limited studies on naturally occurring granulomas and very few attempts have been made to record and classify naturally occurring granulomas in large animals [2, 3]. The objective of the present study was to record the occurrence of granulomas in various tissues and organs in slaughtered cattle and buffaloes.

2. Materials and Methods

2.1. Period of Study. The collection of the samples for the study was carried out during the period from June 2006 to April 2007.

2.2. Collection and Processing of Samples. Organs/tissue samples with gross features of granuloma (ranging from necrotic spots, hard nodules, and growths to abscesses) were obtained from various abattoirs in India located at Puducherry, Chennai, Bangalore, Namakkal, and Kanpur. A detailed gross examination of the lesion with respect to the size, shape, color, and consistency was carried out. Representative samples of the tissues were fixed in 10% neutral buffered formalin for routine paraffin embedding and microtomy. From the fixed and processed tissues, 4-5 µm thick sections were stained with haematoxylin and eosin [4]. After histological assessment, adjacent sections of tissue samples with granulomas were stained using special staining techniques to ascertain the possible etiology. These included Giemsa’s staining for general identification.
of bacteria, fungi, or parasites, Ziehl-Neelsen’s/Auramine O staining for mycobacterium and Periodic Acid-Schiff’s/Gridley’s/Grocott’s stain for fungal elements as per methods described by Luna [4].

2.3. Classification of Granulomas. All the granulomas were classified based on (a) main cellular component as: (i) epithelioid/macrophage rich granuloma, (ii) supplicative/pyogranuloma, or (iii) eosinophilic granuloma and (b) etiological basis as (i) infectious granulomas, (ii) noninfectious/foreign body granulomas, or (iii) granulomas of unknown etiology.

3. Results

In the present study, out of 1600 cattle and buffalo carcasses examined at different locations, 336 tissue samples (21%) were suspected for granulomas. The gross features were characterized by focal grayish-white necrotic spots, large caseated masses, hard nodules as well as growths and abscesses. Based on histopathological assessment, 102 tissue samples (6.38%) were ascertained to be granulomas (Table 1).

The maximum number of granulomas was recorded in the liver, and they accounted for 35.56% (42/102). Pulmonary granulomas were recorded in 29.41% cases (22 lungs and 8 associated lymph nodes). Granulomas were recorded in 6 kidneys (5.88%) and in three associated renal lymph nodes. In 14 cases, subcutaneous granulomas were observed as hard, white, encapsulated nodules in the intercostal spaces and in the brisket region. In four cases, growths found in the body cavities were found to be granulomas. These growths were (i) small nodules in the peritoneum, (ii) a case of multiple nodular lesions in the thoracic cavity attached to the pleura, (iii) a growth attached to the rib, and (iv) a mass attached to the peritoneal wall. Solitary cases of granuloma were recorded in the tongue, muscle, and urinary bladder.

Histologically, based on the main cellular components involved, the granulomas in various tissues and organs were classified into epithelioid, eosinophilic, and supplicative granulomas. It was observed that majority of the granulomas were epithelioid in nature (53%). Eosinophilic granulomas accounted for 37% and supplicative granulomas for 10% of the granulomas.

Special staining techniques were employed to ascertain the possible etiology of the granulomas. Based on the histopathological findings and etiological identification, the granulomas were classified and are shown in Table 2.

Out of 102 granulomas recorded in various organs, the etiology of 75 granulomas could be identified. It was observed that 70 granulomas (68.63%) were of infectious nature; 5 cases (4.90%) were noninfectious and in 27 cases (26.47%) no definite cause could be attributed. The infectious causes identified were parasitic (37 cases, 36.28%), bacterial (32 cases, 31.37%) and one case of a fungal granuloma (0.98%). Non-infectious causes identified were lipid granulomas (2 cases, 1.96%), neoplasm-associated granulomas (2 cases, 1.96%), and a granuloma associated with renal calculi (0.31%).

The distribution and classification of granulomas recorded in various organs and tissues of slaughtered cattle and buffaloes are represented in Table 3.

4. Discussion

The results of the present study are in accordance with the findings of previous reports [2, 3]. Granulomas were histopathologically ascertained in 102 tissue samples (6.38%). Majority of the granulomas were found in the liver (42.11%), followed by lungs (22.05%), lymph nodes (10.78%), kidneys (5.88%), and 14 cases (13.73%) were in the form of subcutaneous nodules/growth. Solitary cases were found in the tongue, muscle, and urinary bladder, whereas 4 granulomas (3.92%) were found in body cavities. In a survey of naturally occurring bovine granulomas carried out at Tirupati (Andhra Pradesh, South India), Yadgirkar [2] recorded granulomas in 315/24,225 (1.3%) carcasses of cattle and buffaloes. Among the 315 granulomas recorded by Yadgirkar [2] 121 (38.41%) were found in the liver, 91 (28.88%) in lungs, 74 (23.49%) in lymph nodes, 28 (8.8%) in intestines, and one (0.31%) in the testis.

The difference in the overall percentage occurrence of granulomas reported in the present study carried out at various locations in South India and North India and that reported by Yadgirkar [2] is attributed to the variation in the total number of carcasses examined in these two studies and also due to the different geographical locations in which these studies were carried out. Further, due to lack of reliable trace back animal identification system in the regions where the study was carried out, it was difficult to precisely

Table 1: Granulomas recorded in various organs/tissues of cattle and buffaloes.

| Organs/tissues          | Suspected | Confirmed |
|-------------------------|-----------|-----------|
| Liver                   | 100       | 42        |
| Lung and Pleura         | 82        | 22        |
| Kidney                  | 17        | 6         |
| Spleen                  | 65        | 11        |
| Lymph node              | 40        | 14        |
| Subcutaneous nodule/mass| 10        | 4         |
| Growth in body cavities | 3         | 1         |
| Intestine               | 2         | N.D.      |
| Uterus                  | 2         | N.D.      |
| Bile duct               | 1         | N.D.      |
| Skin                    | 1         | N.D.      |
| Mammary gland           | 1         | N.D.      |
| Testis                  | 1         | N.D.      |
| Tongue                  | 1         | 1         |
| Urinary bladder         | 1         | 1         |
| Oesophagus              | 1         | N.D.      |
| Total                   | 336       | 102       |

N.D.: not detected.

The maximum number of granulomas was recorded in the liver, and they accounted for 35.56% (42/102). Pulmonary granulomas were recorded in 29.41% cases (22 lungs and 8 associated lymph nodes). Granulomas were recorded in 6 kidneys (5.88%) and in three associated renal lymph nodes. In 14 cases, subcutaneous granulomas were observed as hard, white, encapsulated nodules in the intercostal spaces and in the brisket region. In four cases, growths found in the body cavities were found to be granulomas. These growths were (i) small nodules in the peritoneum, (ii) a case of multiple nodular lesions in the thoracic cavity attached to the pleura, (iii) a growth attached to the rib, and (iv) a mass attached to the peritoneal wall. Solitary cases of granuloma were recorded in the tongue, muscle, and urinary bladder.

Histologically, based on the main cellular components involved, the granulomas in various tissues and organs were classified into epithelioid, eosinophilic, and supplicative granulomas. It was observed that majority of the granulomas were epithelioid in nature (53%). Eosinophilic granulomas accounted for 37% and supplicative granulomas for 10% of the granulomas.

Special staining techniques were employed to ascertain the possible etiology of the granulomas. Based on the histopathological findings and etiological identification, the granulomas were classified and are shown in Table 2.

Out of 102 granulomas recorded in various organs, the etiology of 75 granulomas could be identified. It was observed that 70 granulomas (68.63%) were of infectious nature; 5 cases (4.90%) were noninfectious and in 27 cases (26.47%) no definite cause could be attributed. The infectious causes identified were parasitic (37 cases, 36.28%), bacterial (32 cases, 31.37%) and one case of a fungal granuloma (0.98%). Non-infectious causes identified were lipid granulomas (2 cases, 1.96%), neoplasm-associated granulomas (2 cases, 1.96%), and a granuloma associated with renal calculi (0.31%).

The distribution and classification of granulomas recorded in various organs and tissues of slaughtered cattle and buffaloes are represented in Table 3.

4. Discussion

The results of the present study are in accordance with the findings of previous reports [2, 3]. Granulomas were histopathologically ascertained in 102 tissue samples (6.38%). Majority of the granulomas were found in the liver (42.11%), followed by lungs (22.05%), lymph nodes (10.78%), kidneys (5.88%), and 14 cases (13.73%) were in the form of subcutaneous nodules/growths. Solitary cases were found in the tongue, muscle, and urinary bladder, whereas 4 granulomas (3.92%) were found in body cavities. In a survey of naturally occurring bovine granulomas carried out at Tirupati (Andhra Pradesh, South India), Yadgirkar [2] recorded granulomas in 315/24,225 (1.3%) carcasses of cattle and buffaloes. Among the 315 granulomas recorded by Yadgirkar [2] 121 (38.41%) were found in the liver, 91 (28.88%) in lungs, 74 (23.49%) in lymph nodes, 28 (8.8%) in intestines, and one (0.31%) in the testis.

The difference in the overall percentage occurrence of granulomas reported in the present study carried out at various locations in South India and North India and that reported by Yadgirkar [2] is attributed to the variation in the total number of carcasses examined in these two studies and also due to the different geographical locations in which these studies were carried out. Further, due to lack of reliable trace back animal identification system in the regions where the study was carried out, it was difficult to precisely
trace the origin and the husbandry practices of the animals slaughtered in these locations.

Out of 102 granulomas, the etiology of 75 granulomas could be identified. It was observed that 70 granulomas (68.63%) were of infectious nature; 5 cases (4.90%) were non-infectious, and in 27 cases (26.47%) no definite cause could be attributed. The infectious causes identified were parasites (37 cases, 36.28%), bacterial (32 cases, 31.37%) and one case of a fungal granuloma (0.98%). Non-infectious causes identified were lipid granulomas (2 cases, 1.96%), neoplasm-associated granulomas (2 cases, 1.96%), and a granuloma associated with renal calculi (0.31%). The 315 granulomas recorded by Yadgirkar [2] in cattle and buffaloes were categorized as parasitic (149, 47.30%), bacterial (62, 11.68%), fungal (1, 0.31%), and nonspecific (103, 32.69%).

Etiopathological classification of the granulomas indicated that majority of the granulomas were epithelioid in nature (53%). Eosinophilic granulomas accounted for 37% and suppurative granulomas for 10% of the granulomas.

Hepatic granulomas accounted for 41.18% of all the granulomas, and amongst these 42 granulomas, 26 (61.90%) were eosinophilic, 15 (35.71%) were epithelioid, and one (2.38%) was suppurative. Eosinophilic hepatic granulomas were either parasite induced or were reactions noticed along the migratory tracts of parasites. In a slaughter house-based study of hepatic pathology in bovines carried out at Puducherry, India, Nair et al. [3] recorded hepatic eosinophilic granulomas in 18% of the carcasses examined and suggested these granulomas to be of parasitic origin. In abattoir surveys carried out in Tanzania, Makundi et al. [5] reported the presence of *Schistosoma bovis* in 34% cattle livers. The prevalence and intensities of infection observed in the slaughtered cattle in that study were not related to age, sex and grade of the animals. Agrawal and Banerjee [6] reported that variations in the geographical areas have an important impact on the occurrence and pathogenesis of diseases of parasitic origin.

Out of the 15 epithelioid hepatic granulomas recorded, 12 were due to hydatid cysts and the other three were of unknown etiology. Gram-negative bacteria were detected in a solitary case of suppurative granuloma in the liver. According to Skoric et al. [7], Gram-negative bacteria that
cause granuloma in liver include *Brucella* sp., *Francisella* sp., *Pseudomonas* sp., *Salmonella* sp., *Yersinia* sp., and *Actinobacillus* sp.

The liver is the prime repository for granulomatous inflammation. In humans, about 10% of the routine liver biopsy specimens demonstrate granulomas, and most systemic granulomatous diseases are found to involve the liver, at least to some extent [8, 9]. This may be due to the increased population of mononuclear phagocytes, including Kupffer cells and other macrophages in the liver. These cells serve as sentinels screening the blood for such stimuli as microorganisms, circulating antigens and immune complexes, inflammatory mediators, and partially degraded foreign substance and, then, through incompletely understood mechanisms assemble into granulomas [10].

Pulmonary granulomas were recorded in 22 cases (21.58%). Majority of these granulomas (15 cases, 14.7%) were Ziehl-Neelsen acid-fast-positive tuberculous epithelioid granulomas. Pleural involvement was seen in two cases. In addition to the lungs, most of the tuberculous lesions were confined to the bronchial and mediastinal lymph nodes (8 cases, 7.84%). Balani [11] reported an incidence of 10.97% (34/310) of granulomatous lesions in the lungs of slaughtered cattle. Yadgirkar [2] reported an incidence of 28.8% (91/315) of granulomatous lesions in the lungs of slaughtered cattle and buffaloes, and 0.97% of these were tuberculous. Choudary [12] recorded tuberculous pneumonia in 2% (5/250) slaughtered cattle. It is clear from the present study that hepatic granulomas are not be ascertained. However, in the present case, the source of the lipid could not be ascertained.

In the present study, granulomas were observed in six kidneys (6/15, 35.29%) and 3 associated lymph nodes. Features of epithelioid granuloma were noticed in three kidneys. These included a case with tuberculosis, another case associated with renal calculi as well as in one case of unknown etiology. Suppurative renal granulomas were recorded in three cases. In one case, Gram-negative bacteria were identified and the other two were associated with lymphomas. According to Skoric et al. [7] Gram-negative organisms which have been implicated in granulomas are *Corynebacterium renale*, *Brucella* sp., *Francisella* sp., *Pseudomonas* sp., *Salmonella* sp., *Yersinia* sp., and *Actinobacillus* sp.

In the present study, 13.73% (14/102) granulomas were recorded in the subcutaneous tissues. In 11 subcutaneous eosinophilic granulomas, *Onchocerca gibsoni* (a medium sized filarioid which usually inhabit subcutaneous tissues, ligaments and aponeuroses of large mammals mostly found in south-east Asia) could be demonstrated. The gross and histopathology of eosinophilic granulomas evoked by *O. gibsoni* recorded in this study are in accordance with those reported by other workers (Shastri [14] and Balachandran et al. [15]). Solitary cases of epithelioid lipogranuloma (characterized by multiple necrotic foci, multinucleated giant cells and cholesterol clefts), epithelioid acid-fast-positive tuberculous granuloma, and supplicative granuloma were also recorded in the subcutaneous tissues.

Granulomas were also recorded from the body cavities. A peritoneal nodule showed an eosinophilic granulomatous reaction suspected to be parasite induced. The gross and histopathological features of a large growth attached to the peritoneal wall had features of actinomycosis. Cattle have a relatively high capacity to wall off septic foci within the peritoneal cavity. These infections are usually secondary to perforating injuries caused by bite wounds or foreign bodies [16]. In the present study, supplicative granuloma was seen in the thigh muscle of a cow. Suppurative myositis in muscle may sometimes be haematogenous in origin, but more often they result from inoculation or from an extension of a supplicative focus in adjacent structures such as joints, tendon sheaths, or lymph nodes [16].

The granulomatous inflammatory response is ubiquitous in pathology, being a manifestation of many infective, toxic, allergic, autoimmune, and neoplastic diseases and also conditions of unknown aetiology. Granulomas induced by members of *Mycobacterium tuberculosis* complex (MTC) group, *Schistosomes* sp., *Echinococcus* sp. and even *Capillaria* sp. may pose a potential zoonotic risk. Some of these lesions such as those caused by *Mycobacterium tuberculosis* complex (MTC) group, *Brucella* sp. may occur at abnormal locations and would probably be overlooked by the consulting veterinarian. In the case of tuberculosis, studies have indicated that confirmatory diagnosis of tuberculous granuloma were made only after histopathological evaluation, bacteriological culture, or polymerase chain reaction analysis of tissues collected from affected animals. Considering these aspects, all granulomas in ruminants should be thoroughly evaluated to determine the underlying etiological agent in order to formulate a control and eradication strategy. In this respect, granuloma submission programs have played a major role in eradication of *Mycobacterium bovis* in some developed countries [17].

Another important aspect of granuloma is the economic loss incurred by farmers in animals bearing granulomas. It is clear from the present study that hepatic granulomas account for the majority of the cases. It is likely that affected animals may not show any pathognomonic signs and the condition usually remains clinically undiagnosed. However, due to the hepatic involvement the productivity of the animal is likely to be reduced. Since, in all the cases of granulomas, the affected organs are to be condemned, this is also of much concern because of its potential to cause economic losses. Hence, continued research into granulomatous inflammation is essential, not only for its theoretical value, but also for its potential clinical, zoonotic, and economic implications.
Acknowledgment

The authors thank the Dean, RAGACOVAS, Puducherry for providing facilities to conduct the study.

References

[1] G. T. Williams and W. J. Williams, “Granulomatous inflammation: a review,” Journal of Clinical Pathology, vol. 36, no. 7, pp. 723–733, 1983.
[2] G. Yadgirkar, A study of granulomas in bovines, Ph.D. thesis, Andhra Pradesh Agricultural University, Hyderabad, India, 1983.
[3] M. G. Nair, R. Kumar, A. W. Lakkawar, and K. C. Varshney, “A slaughter house and necropsy based study of lesions in bovines,” Indian Veterinary Journal, vol. 83, no. 5, pp. 490–493, 2006.
[4] L. G. Luna, Ed., Manual of Histologic Staining Methods of the Armed Forces Institute of Pathology, McGraw Hill, New York, NY, USA, 1968.
[5] A. E. Makundi, A. A. Kassuku, R. M. Maselle, and M. E. Boa, “Distribution, prevalence and intensity of Schistosoma bovis infection in cattle in Iringa district, Tanzania,” Veterinary Parasitology, vol. 75, no. 1, pp. 59–69, 1998.
[6] M. C. Agrawal and P. S. Banerjee, “Problems confronting helminthic diseases of domestic animals in India,” Journal of Parasitic Diseases, vol. 31, no. 1, pp. 3–13, 2007.
[7] M. Skoric, E. J. Shitaye, R. Halouzka et al., “Tuberculous and tuberculoid lesions in free living small terrestrial mammals and the risk of infection to humans and animals: a review,” Veterinarini Medicina, vol. 52, no. 4, pp. 144–161, 2007.
[8] J. C. Guckian and J. E. Perry, “Granulomatous hepatitis: an analysis of 63 cases and review of the literature,” Annals of Internal Medicine, vol. 65, no. 5, pp. 1081–1100, 1966.
[9] G. Klatskin, “Hepatic granuloma: problems in interpretation,” Mount Sinai Journal of Medicine, vol. 44, no. 6, pp. 798–812, 1977.
[10] P. T. Harrington, J. J. Gutiérrez, C. H. Ramirez-Ronda, R. Quiones-Soto, R. H. Bermúdez, and J. Chaffey, “Granulomatous hepatitis,” Reviews of Infectious Diseases, vol. 4, no. 3, pp. 638–655, 1982.
[11] D. K. Balani, Studies on the pathology of buffalo lungs showing macroscopic lesions collected from slaughter houses, M.V.Sc. thesis, Agra University, Agra, India, 1967.
[12] C. Choudary, Incidence and histopathology of lung lesions in bovines, M.V.Sc thesis, OUAT, Bhubaneswar, India, 1970.
[13] K. Sato, H. Takahashi, H. Amano, T. Uekusa, T. Dambra, and S. Kira, “Diffuse progressive pulmonary interstitial and intra-alevolar cholestrol granulomas in childhood,” European Respiratory Journal, vol. 9, no. 11, pp. 2419–2422, 1996.
[14] U. V. Shastri, “Preliminary note on filarial infections in cattle (Bos indicus) and buffaloes (Bubalus bubalis),” Indian Journal of Animal Health, vol. 12, pp. 193–195, 1973.
[15] C. Balachandran, N. Pazhanivelu, S. Variamuthu, M. Raman, C. Sriekumar, and B. Muralimanohar, “Pathological changes in concurrent Onchocerca gibsoni and Onchocerca armillata infestation in cow,” Indian Journal of Veterinary Pathology, vol. 29, no. 2, pp. 131–132, 2005.
[16] K. V. F. Jubb, P. C. Kennedy, and N. Palmer, Eds., Pathology of Domestic Animals, vol. 2, Academic Press, San Diego, Calif, USA, 4th edition, 1993.
[17] B. Radunz, “Surveillance and risk management during the latter stages of eradication: experiences from Australia,” Veterinary Microbiology, vol. 112, no. 2–4, pp. 283–290, 2006.