Oxysterols Modulate the Acute Effects of Ethanol on Hippocampal N-Methyl-D-Aspartate Receptors, Long-Term Potentiation, and Learning

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ABSTRACT

Ethanol is a noncompetitive inhibitor of N-methyl-D-aspartate receptors (NMDARs) and acutely disrupts hippocampal synaptic plasticity and learning. In the present study, we examined the effects of oxysterol positive allosteric modulators (PAMs) of NMDARs on ethanol-mediated inhibition of NMDARs, block of long-term potentiation (LTP) and long-term depression (LTD) in rat hippocampal slices, and defects in one-trial learning in vivo. We found that 24S-hydroxycholesterol and a synthetic oxysterol analog, SGE-301, overcame effects of ethanol on NMDAR-mediated synaptic responses in the CA1 region but did not alter acute effects of ethanol on LTD; the synthetic oxysterol, however, overcame acute inhibition of LTP. In addition, both oxysterols overcame persistent effects of ethanol on LTP in vitro, and the synthetic analog reversed defects in one-trial inhibitory avoidance learning in vivo. These results indicate that effects of ethanol on both LTP and LTD arise by complex mechanisms beyond NMDAR antagonism and that oxysterol NMDAR PAMS may represent a novel approach for preventing and reversing acute ethanol-mediated changes in cognition.

SIGNIFICANCE STATEMENT

Ethanol acutely inhibits hippocampal NMDARs, LTP, and learning. This study found that certain oxysterols that are NMDAR-positive allosteric modulators can overcome the acute effects of ethanol on NMDARs, LTP, and learning. Oxysterols differ in their effects from agents that inhibit integrated cellular stress responses.

INTRODUCTION

Ethanol can acutely alter memory (Oslin and Cary, 2003; White, 2003; Abrahao et al., 2017) and impair long-term synaptic plasticity involved in memory formation (Izumi et al., 2005b; Zorumski et al., 2014). Ethanol is a noncompetitive inhibitor of N-methyl-D-aspartate receptors (NMDARs) that contribute to synaptic plasticity but is only a partial antagonist at concentrations that block synaptic plasticity. It is thus uncertain whether effects on NMDARs drive ethanol-induced impairments. Some effects of ethanol on synaptic plasticity, particularly inhibition of long-term potentiation (LTP), involve a form of metaplasticity resulting from activation of NMDARs that remain unblocked by ethanol during acute drug exposures (Izumi et al., 2005b; Tokuda et al., 2011). This NMDAR-dependent LTP inhibition involves complex signaling (Zorumski and Izumi, 2012), including activation of the cellular integrated stress response (ISR), a mechanism that regulates neuronal plasticity and memory under pathologic conditions (Izumi and Zorumski, 2020). Ethanol activates ISR (Chen et al., 2008; Magne et al., 2011), and agents that block ISR overcome the effects of ethanol on LTP, long-term depression (LTD), and a form of hippocampal-dependent, one-trial inhibitory avoidance learning (Izumi and Zorumski, 2020). These observations suggest that modulation of stress responses may effectively prevent or reverse detrimental effects of ethanol on cognition, particularly learning defects associated with acute alcohol-induced memory blackouts (White, 2003).

One of the agents that overcomes the acute effects of ethanol on synaptic plasticity and learning is an agonist at liver X receptors (LXRs) (Izumi and Zorumski, 2020). The test of the LXR agonist (GW3965) was prompted by work showing that LXR activation dampens ISR by modulating plasma membrane composition (Rong et al., 2013). Effects of GW3965 suggest that other agents that modulate LXRs may also have beneficial effects on ethanol’s synaptic and cognitive effects.
Certain side-chain–oxidized derivatives of cholesterol (oxysterols) are agonists for LXRs (Russell et al., 2009; Sun et al., 2016b). A prototypic example is 24S-hydroxycholesterol (24S-HC), the major cholesterol metabolite in the brain (Janowski et al., 1999). 24S-HC is an endogenous neuregulator that, in addition to effects on LXR, is a positive allosteric modulator (PAM) of NMDARs that also promotes hippocampal plasticity (Paul et al., 2013; Linsenbardt et al., 2014). Furthermore, synthetic derivatives of 24S-HC (Paul et al., 2013) have shown promise as potential therapeutic agents for treating cognitive dysfunction in certain neuropsychiatric illnesses (Keenig et al., 2019). For these reasons, we were interested in determining whether 24S-HC and a synthetic derivative, SGE-301, would alter acute effects of ethanol on hippocampal function and learning. We chose SGE-301 for these studies because it mimics the effects of 24S-HC on NMDARs and has good brain bioavailability after systemic administration (Paul et al., 2013).

Materials and Methods

Chemicals. SGE-301 was a gift from Sage Therapeutics (Cambridge, MA) (Paul et al., 2013). Other chemicals were purchased from Sigma Chemical Company (St. Louis, MO). For ex vivo slice experiments, SGE-301 or 24S-HC was dissolved in DMSO and administered at concentrations noted in the text. The concentration of DMSO was typically 0.1% and had no effect on hippocampal responses or synaptic plasticity (Izumi and Zorumski, 2020; Popiolek et al., 2018). For in vivo experiments, 5 mg SGE-301 was dissolved in 3.3 ml of 2-hydroxypropyl-ß-cyclodextrin (CDX) (catalog number 332607; Sigma) (1.5 g CDX + 3 ml water) and administered at a final dose of 3 mg/kg i.p. This dose of SGE-301 was selected based on prior behavioral studies we have done with this agent (Paul et al., 2013).

Hippocampal Slice Preparation. The Washington University Institutional Animal Care and Use Committee approved all protocols for animal experiments according to NIH guidelines. Using methods described previously, hippocampal slices were prepared from postnatal day 28–32 Harlan (Indianapolis, IN) Sprague-Dawley male albino rats (Tokuda et al., 2010, 2011). For slice preparation, rats were anesthetized with isoflurane, and dissected hippocampi were pinned at their ventral pole on an agar base in ice-cold artificial cerebrospinal fluid (ACSF) that contained (in millimolars) 124 NaCl, 5 KCl, 2 MgSO4, 2 CaCl2, 1.25 NaH2PO4, 22 NaHCO3, and 10 glucose, bubbled with 95% O2 and 5% CO2 at 4–6°C. The dorsal two-thirds of the hippocampus was cut into 500-μm slices using a rotary tissue slicer and kept in a chamber with gassed ACSF at 30°C for at least 1 hour before use in physiology studies.

Hippocampal Slice Physiology. Slices were transferred one at a time to a submersion-recording chamber at 30°C and perfused with the ACSF described above at 2 ml/min. We did not include inhibitors of GABA2 receptors in our recording solutions to avoid nonphysiologic network effects resulting from disinhibition (Izumi et al., 2005a, 2007; Tokuda et al., 2011; Izumi and Zorumski, 2020). Extracellular recordings were obtained from the apical dendritic layer (stratum radiatum) of area CA1 to monitor field excitatory post synaptic potentials (fEPSPs).

fEPSPs in the stratum radiatum of CA1 were evoked once per minute with 0.1-millisecond constant-current pulses administered to the Schaffer collateral pathway via a bipolar stimulating electrode. Stimulus intensity was set to half-maximal based on a baseline input-output (IO) curve. LTP was induced using a 100 Hz × 1 second high-frequency stimulus (HFS) at the same intensity. LTD was induced with low-frequency stimulation (LFS) consisting of 900 single pulses at 1 Hz. IO curves were repeated 60 minutes after HFS or LFS and were the primary outcome measure of synaptic change by comparison with baseline IO curves.

For experiments examining isolated NMDAR-mediated fEPSPs, dendritic responses were recorded in stratum radiatum at low frequency (1/min) in ACSF containing 0.1 mM Mg2+ and 2.5 mM Ca2+ to promote NMDAR activation and 30 μM 6-cyano-7-nitroquinoxaline-2,3-dione to block AMPARs (Izumi et al., 2005b). fEPSPs under these conditions are completely inhibited by the NMDAR antagonist 2-amino-5-phosphonovalerate (Izumi et al., 2005b, 2006). We quantified NMDAR fEPSPs by the rising slope of the field potential (12–24 points) before and at the end of drug treatment, and effects of drugs were based on comparison of the 50% maximal point on IO curves before and after drug exposure.

Behavioral Studies. To assess in vivo memory formation, P28-32 rats were tested in a one-trial inhibitory avoidance learning task that has been associated with CA1 hippocampal LTP (Whitlock et al., 2006; Tokuda et al., 2010). The testing apparatus has two chambers, one of which is illuminated, whereas the second chamber is kept dark. These chambers have a floor of stainless steel rods (4 mm diameter, spaced 10 mm apart), through which an electrical shock can be delivered in the dark chamber (12 × 20 × 16 cm). The lit compartment (30 × 20 × 16 cm) was illuminated by four 13-W lights, and light intensity in the lit chamber was 100 lux, whereas the dark chamber was <10 lux.

At the time of study, rats were initially placed in the lit chamber and allowed to move freely between the chambers for 10 minutes. Subsequently, animals were administered SGE-301 (3 mg/kg i.p.) or vehicle 2 hours prior to ethanol or saline administration. At 15 minutes prior to being placed back in the apparatus, rats were treated with either saline or ethanol (2 g/kg i.p.). This dose of ethanol results in acute blood and cerebrospinal fluid levels of ethanol near 20 mM 15 minutes after injection (Izumi and Zorumski, 2020). Rats were then placed in the lit compartment and allowed to explore the apparatus for up to 300 seconds. Upon complete entry into the dark chamber, rats were immediately given a foot shock. Animals were then removed from the apparatus and returned to their home cages. On the next day of testing, rats were placed in the lit chamber without drug treatment, and the latency to enter the dark compartment was recorded over a 300-second trial.

Experimental Design and Statistical Analysis. Data were collected and analyzed using PC clamp (Molecular Devices, Union City, CA) and are expressed as means ± S.E.M. At 60 minutes after HFS or LFS, fEPSPs were normalized to baseline recordings (taken as 100%). A two-tailed Student’s t test was used for all comparisons of fEPSPs between groups. When appropriate (Fig. 1), paired t tests were used. Statistical comparisons were based on analysis of IO curves at baseline and 60 minutes after HFS or LFS to determine the degree of change in the maximal rising slope of fEPSPs evoked by stimulus at the 50% point on the IO curves, with P < 0.05 considered significant (Izumi and Zorumski, 2020). For behavioral studies in Fig. 4, data were analyzed by one-way analysis of variance followed by Dunnett’s multiple comparisons test. Numbers reported in the text for statistical analyses are the number (N) of animals studied in a given condition. Statistics were performed using commercial software (SigmaStat (Systat Software, Inc., Richmond City, CA) or GraphPad Prism version 8.3.0 for Windows (www.graphpad.com; GraphPad Software, San Diego, CA)]. Data in figures showing fEPSPs display continuous monitoring of responses at low frequency and thus may differ from numerical results described in the text, which are based on analyses of IO curves.

Results

Previously, we found that 60 mM ethanol acutely and reversibly inhibits isolated NMDAR-mediated fEPSPs in the CA1 region of rat hippocampal slices by about 50%, largely via inhibition of a subtype of NMDARs expressing GluN2B subunits (Izumi et al., 2005b). In initial experiments, we examined whether an oxysterol PAM of NMDARs alters the effects of ethanol on synaptic responses mediated by these
receptors. 24S-HC is an endogenous oxysterol, the major metabolite of cholesterol in brain (Sun et al., 2016a,b), and an NMDAR PAM that enhances CA1 NMDAR fEPSPs at concentrations in the low micromolar range (Paul et al., 2013; Linsenbardt et al., 2014). At a threshold concentration of 1 μM in hippocampal slices, 24S-HC prevented the ability of 60 mM ethanol to inhibit NMDAR fEPSPs when administered prior to and during ethanol exposure (96.2% ± 6.3% of baseline responses vs. 39.5% ± 10.1% of baseline in the presence of ethanol alone, \(P = 0.0004\); \(N = 5\); Fig. 1A). At this concentration, 24S-HC had little direct effect on NMDAR responses (Fig. 1A). 24S-HC also reversed the effects of ethanol on NMDAR fEPSPs when administered after ethanol-induced NMDAR inhibition and in the continued presence of ethanol (95.6% ± 8.5% of baseline after 24S-HC administration vs. 53.9% ± 5.5% with ethanol alone, \(N = 5\), \(P = 0.03137\) by paired \(t\) test vs. ethanol alone; Fig. 1B). Similarly, 1 μM SGE-301, a synthetic mimic of 24S-HC (Paul et al., 2013), prevented the effects of ethanol on NMDA fEPSPs and had no clear effect on these responses on its own at this concentration in slices (104.9% ± 5.0%, \(P = 0.0014\) vs. ethanol alone; Fig. 1C; \(N = 5\) (Sun et al., 2016a)). SGE-301 also overcame the effects of ethanol on NMDA fEPSPs when administered after inhibition by ethanol (55.9% ± 6.5% in the presence of ethanol alone vs. 106.9% ± 4.4% with ethanol + SGE-301, \(N = 5\), \(P < 0.0001\) by paired test; Fig. 1D).

Fig. 1. 24S-HC and a synthetic oxysterol analog prevent effects of ethanol on NMDA EPSPs. (A) The graph shows the time course of change of isolated NMDAR-mediated fEPSPs in the presence of 1 μM 24S-HC (blue bar, denoted as 24S in the graph) followed by 60 mM ethanol (denoted as EtOH in the graph) normalized to baseline (100%). In the absence of 24S-HC, ethanol depressed NMDAR-mediated fEPSPs. (B) Even when administered after 60 mM ethanol, which acutely inhibits NMDAR responses, 24S-HC dampened the block of NMDA fEPSPs. (C) SGE-301 (red bar), a synthetic mimic of 24S-HC, also prevents effects of ethanol on NMDA fEPSPs. (D) Akin to 24S-HC, SGE-301 reverses the effects of ethanol on NMDA fEPSPs when administered after block is established. Traces to the right of the graphs show representative NMDA fEPSPs at the times denoted, with baseline responses shown as dashed lines. Calibration: 1 mV, 5 milliseconds.
baseline 60 minutes after LFS; \( N = 6 \), Fig. 2A). Surprisingly, despite overcoming the effects of ethanol on NMDAR fEPSPs (Fig. 1, A and B), 24S-HC had no effect on the ability of ethanol to block LTD (fEPSP slope: 99.3% ± 3.5% of baseline 60 minutes after LFS, \( N = 5 \) vs. 98.7% ± 3.8% of baseline with ethanol alone, \( N = 6 \); \( P = 0.3796 \); \( P < 0.0001 \) for each vs. control LTD; Fig. 2A). 24S-HC alone does not inhibit LTD (Popiolek et al., 2020). Similarly, SGE-301 failed to overcome the effects of ethanol on LTD (104.2% ± 6.0% of baseline, \( N = 5 \); \( P = 0.0002 \) vs. control LTD; Fig. 2B). Neither 24S-HC (52.3% ± 5.2% of baseline, \( N = 5 \); \( P = 0.8396 \) vs. control LTD) nor SGE-301 (60.6% ± 4.6% of baseline, \( N = 5 \); \( P = 0.3866 \) had any effect on LTD when administered alone.

We previously found that 24S-HC promotes LTP (Paul et al., 2013). Thus, given effects on the block of NMDA EPSPs, we next examined the effects of 24S-HC on acute inhibition of LTP by 60 mM ethanol. We found that 24S-HC failed to alter the ability of ethanol to block LTP induced by 100 Hz \( \times \) 1 second HFS at either 1 or 10 \( \mu M \) (1 \( \mu M \) 24S-HC: 95.5% ± 7.2% of baseline fEPSPs 60 minutes after HFS (white circles); 10 \( \mu M \): 94.3% ± 9.8% of baseline, \( N = 5 \) and \( N = 6 \) vs. ethanol alone: 96.1% ± 3.2%, \( N = 6 \); \( P = 0.9371 \) and 0.8649, respectively; Fig. 2C). In control slices, HFS readily induced LTP (black triangles, 148.4% ± 6.9% of baseline 60 minutes after HFS, \( N = 6 \); \( P = 0.0001 \) vs. ethanol alone (black circles); \( P = 0.0005 \) vs. ethanol + 1 \( \mu M \) 24S-HC and \( P = 0.0011 \) vs. ethanol + 10 \( \mu M \) 24S-HC; Fig. 2C). 24S-HC alone did not alter LTP in naïve slices at either 1 \( \mu M \) (130.0% ± 4.6% of baseline, \( N = 5 \); \( P = 0.3587 \) vs. control LTP; not shown). In contrast to what we observed with 24S-HC, we found that SGE-301 reversed the acute effects of ethanol on LTP (155.6% ± 16.1% of baseline, \( N = 5 \); \( P = 0.0032 \) vs. ethanol alone and \( P = 0.6712 \) vs. control LTP; Fig. 2D). When administered alone, SGE-301 had no effect on LTP induced by a single 100 Hz \( \times \) 1 second HFS (154.7% ± 15.0% of baseline, \( N = 5 \); \( P = 0.6947 \)).

As noted earlier, ethanol’s block of LTP involves a form of negative metaplasticity (Zorumski and Izumi, 2012) dependent upon untimely activation of NMDARs prior to delivery of HFS; this defect in LTP induction persists for over an hour.
after ethanol washout (Izumi et al., 2005b; Tokuda et al., 2011; Zorumski et al., 2014). Thus, we also examined whether 24S-HC altered the persisting effects of ethanol. In contrast to acute LTP inhibition by ethanol, the prolonged effects of ethanol on LTP were eliminated in the presence of 24S-HC (fEPSP slopes: 98.1% ± 6.5% of baseline 60 minutes after HFS following ethanol alone vs. 139.2% ± 7.6% in the presence of 24S-HC, N = 6 and 5, respectively; P = 0.0025 by two-tailed Student’s t test; Fig. 3A). We also found that 24S-HC overcame the persisting effects of ethanol when administered after ethanol exposure (164.7% ± 14.7% of baseline, N = 5; P = 0.0017 vs. ethanol alone; Fig. 3B). Similar to 24S-HC, SGE-301 overcame persistent LTP inhibition by ethanol (153.6% ± 13.4% of baseline, N = 5; P = 0.0034 vs. ethanol; Fig. 3C).

To determine whether the interaction between oxysterols and ethanol on synaptic plasticity predicts behavioral effects of ethanol, we examined whether oxysterols alter cognitive impairment by ethanol in vivo. For these studies, we used a one-trial inhibitory avoidance learning task that has previously been linked to hippocampal LTP (Whitlock et al., 2006; Tokuda et al., 2010) and that we recently showed is altered by acute ethanol administration (Izumi and Zorumski, 2020). When administered at 2 g/kg i.p. 15 minutes prior to training, ethanol markedly reduced learning in the one-trial task when rats were tested 24 hours after training (Fig. 4). Based on our observations with the oxysterols (Figs. 1–3), we examined whether the 24S-HC analog SGE-301 altered these amnesic effects of ethanol. We used SGE-301 for these experiments because of its brain bioavailability after systemic administration (Paul et al., 2013) along with consistent effects on LTP in the presence of ethanol (Figs. 2 and 3). Rats treated with CDX, the vehicle used for in vivo administration of SGE-301, prior to initial exposure to the test chamber learned the task readily and remained in the lit chamber for 260.0 ± 40.0 seconds (N = 5) 24 hours after initial exposure and shock. Animals treated with ethanol plus CDX (vehicle for SGE-301) prior to initial exposure failed to learn the task and entered the dark chamber in 6.0 ± 2.0 seconds (N = 5) upon exposure to the apparatus 24 hours later (P < 0.0003 vs. CDX controls). When administered at 3 mg/kg i.p. 2 hours prior to ethanol, SGE-301 markedly diminished the ethanol-mediated learning defect (latency to enter the dark chamber: 253.0 ± 32.0 seconds, N = 5; P < 0.0003 vs. vehicle + ethanol-treated controls); this duration in the lit chamber did not differ from controls (P = 0.8947; Fig. 4). SGE-301 alone had no effect on learning the task (267.6 ± 32.4 seconds, N = 5, P = 0.8863 vs. CDX-alone controls; Fig. 4).

Discussion

We recently found that the LXR agonist GW3965, an agent that affects sterol metabolism (Wang and Tontonoz, 2018) and inhibits endoplasmic reticulum (ER) stress (Rong et al., 2013), potently and effectively overcomes the acute effects of ethanol.

Fig. 3. 24S-HC and SGE-301 both reverse the persisting effects of ethanol on LTP. (A) The graph shows the block of LTP in slices treated with 60 mM ethanol for 15 minutes and then washed out for 30 minutes prior to delivery of HFS (arrow) (blue squares). 24S-HC (1 μM) overcame the effects of ethanol and allowed robust LTP (gray circles). (B) 24S-HC also overcame the persisting effects of ethanol on LTP when administered after ethanol. (C) A synthetic mimic of 24S-HC, SGE-301 (1 μM), also overcame the persisting effects of ethanol on LTP. Traces show representative fEPSPs as in Fig. 2 with solid traces taken 60 minutes after HFS. In (A), the upper set of traces is with 24S-HC. Calibration: 1 mV, 5 milliseconds.
on hippocampal NMDARs, synaptic plasticity, and one-trial inhibitory avoidance learning (Izumi and Zorumski, 2020). With the exception of effects on the block of NMDARs by ethanol, these actions were mimicked by ISRIB, a selective inhibitor of the ISR (Sidrauski et al., 2013; Sekine et al., 2015; Costa-Mattioli and Walter, 2020; Izumi and Zorumski, 2020). The LXR agonist and ISRIB also overcame prolonged adverse effects of ethanol on LTP that are observed after ethanol washout. Prior studies have demonstrated that neuronal stress resulting from aging and NMDAR activation promotes translocation of CYP46A1, the enzyme responsible for 24S-HC synthesis (Russell et al., 2009), from ER to plasma membrane and 24S-HC synthesis resulting in neuronal survival (Sodero et al., 2011a,b, 2012). Other studies indicate that knockdown of CYP46A1 in the hippocampus activates ER stress responses, apoptotic neuronal damage, and cognitive dysfunction (Burlot et al., 2015; Djelti et al., 2015); related effects on neuronal function are observed in striatum (Boussicault et al., 2015) and cerebellum (Nobrega et al., 2019). These observations prompt us to examine whether the oxysterol 24S-HC shared the effects of GW3965, given that this oxysterol is an endogenous agonist at LXRs (Janowski et al., 1999; Sun et al., 2016). We found that 24S-HC and a synthetic oxysterol analog, SGE-301, shared certain effects of GW3965, overcoming acute block of NMDAR fEPSPs by ethanol and prolonged effects of ethanol on LTP after ethanol washout. SGE-301 alone had no effect on one-trial learning compared with vehicle controls. ****P = 0.0001 by one-way ANOVA, followed by Dunnett’s multiple comparisons test vs. ethanol (F = 17.60).

Our results demonstrate that the oxysterols mimic only certain effects of the LXR agonist and the ISR inhibitor, indicating that the oxysterols clearly differ from both of these agents. Consistent with these observations, prior work indicates that SGE-301 has no clear effects on LXR, unlike 24S-HC (Paul et al., 2013). In contrast, both SGE-301 and 24S-HC are potent and effective NMDAR PAMs (Paul et al., 2013), raising the strong likelihood that effects on NMDARs may primarily be responsible for the common constellation of oxysterol effects we have observed. Based on studies to date, NMDAR enhancement by the oxysterols is not shared by either GW3965 or ISRIB (Izumi and Zorumski, 2020).

In prior studies, we showed that the effects of a high concentration of ethanol on LTP and LTD likely result from different mechanisms. Effects on NMDARs and LTD mimic the effects of selective antagonists of NMDARs that express GluN2B subunits and reverse readily after ethanol washout. Effects on LTP are more complex and persistent and involve a form of metaplasticity after untimely NMDAR activation during the period of ethanol exposure (Izumi et al., 2005b; Zorumski et al., 2014). As a result, complete NMDAR inhibition with 2-amino-5-phosphonovalerate during ethanol exposure prevents persisting effects on LTP (Tokuda et al., 2011). In the present study, we found that the persisting effects of ethanol on LTP were overcome by both oxysterols, but, contrary to expectations, neither oxysterol altered the acute block of LTD, despite overcoming the block of hippocampal NMDARs by ethanol. The latter observations raise the possibility that more complex intracellular and perhaps intercellular signaling contributes to ethanol-mediated block of LTD. As one possibility, ISR inhibition overcomes ethanol-mediated LTD inhibition, despite having no effect on the block of NMDARs (Izumi and Zorumski, 2020). Prior studies indicate that activation of ER stress promotes synthesis of endogenous neurosteroids (pregnenolone) from cholesterol (Barbero-Camps et al., 2014), and we previously found that induction of homosynaptic LTD in the CA1 region involves local NMDAR-dependent synthesis of GABAergic 5α-reduced neurosteroids, downstream products of pregnenolone (Izumi et al., 2013). NMDAR-dependent LTD induction is also accompanied by metaplastic LTD inhibition involving neurosteroids (Izumi et al., 2013), as is LTD inhibition by pharmacological NMDAR activation (Izumi et al., 2007; Tokuda et al., 2011). These observations raise a possible role for ER stress.
and ISR-mediated intracellular signaling in determining effects of a variety of stressors on LTD and LTP (Barbero-Camps et al., 2014; DiPrisco et al., 2014; Costa-Mattioli and Walter, 2020).

We also note that ethanol’s ability to inhibit LTP via an NMDAR-dependent mechanism is shared with other non-competitive NMDAR antagonists that also impair memory, most notably subanesthetic ketamine (Izumi and Zorumski, 2014). Both subanesthetic ketamine and ethanol appear to inhibit GluN2B-expressing NMDARs preferentially in the CA1 region at concentrations that inhibit LTP (Gerhard et al., 2020), and trigger NMDAR activation of unblocked NMDARs. Also, another synthetic oxysterol NMDAR PAM, SGE-201, overcomes persisting adverse effects of subanesthetic ketamine on LTP, and SGE-201 and SGE-301 reverse certain effects of related agents phencyclidine (PCP) and MK-801 on cognition and behavior (Paul et al., 2013). In the latter studies, effects of oxysterols on PCP-induced changes were observed with subchronic PCP dosing and observed after drug washout. Some evidence also suggests that 245-SCB-preferentially enhances tonic GluN2B-expressing NMDARs (Wei et al., 2019, but see Paul et al., 2013).

LXR agonists and ISR inhibitors have therapeutic potential as cognitive enhancers in neuropsychiatric illnesses (Wang et al., 2002; Donkin et al., 2010; Baez-Becerra et al., 2018). LXR agonists also have anti-inflammatory and neuroprotective effects that contribute to their CNS actions (Mouzat et al., 2002; Donkin et al., 2010; Baez-Becerra et al., 2018). With NMDAR enhancement, however, comes potential risks of excitotoxicity, although other studies have found both enhanced (Yamanaka et al., 2010; Camps et al., 2014; DiPrisco et al., 2014; Costa-Mattioli and Walter, 2014) and diminished (Okabe et al., 2013; Ishikawa et al., 2016) neurotoxicity resulting from these agents. It is also intriguing that an oxysterol NMDAR PAM appears to be well tolerated in humans and is now in early testing as a possible cognitive enhancer (Koenig et al., 2019). NMDAR-mediated metaplasticity contributes to defects in LTD and learning in other conditions of neuronal stress; thus, effects observed here could be relevant for other neuropsychiatric illnesses (Zorumski and Izumi, 2012).

It is interesting and surprising that neither oxysterol overcame the acute effects of ethanol on LTD and only SGE-301 overcame acute LTD inhibition; the latter effect may reflect a difference in potency between the oxysterols. SGE-301 also overcame effects on one-trial avoidance learning, a form of learning linked to hippocampal LTP (Whitlock et al., 2006). However, both oxysterols overcame more persistent LTP inhibition after ethanol washout, including reversal of LTD inhibition when the oxysterols were administered after ethanol removal. We did not systematically explore how long after ethanol the oxysterols could be administered, but the combination of ex vivo and in vivo results suggests that persisting changes in synaptic plasticity likely account for at least some of ethanol’s effects on cognition during and after bouts of intoxication. Prior stu(Parada et al., 2011)des have observed persisting cognitive changes, including learning defects after acute ethanol intoxication, resulting in altered cognitive performance during alcohol hangovers (Crews et al., 2004; Silvestre de Ferron et al., 2015; Gunn et al., 2013; West et al., 2018; Contreras et al., 2019) and perhaps contributing to longer-term impairment from ethanol.

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Authorship Contributions

Performed data analysis: Izumi, Zorumski.

Wrote or contributed to the writing of the manuscript: Izumi, Mennerick, Doherty, Zorumski.

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