The preventive effect of loganin on oxidative stress-induced cellular damage in human keratinocyte HaCaT cells

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SUMMARY Loganin is a type of iridoid glycosides isolated from Corni fructus and is known to have various pharmacological properties, but studies on its antioxidant activity are still lacking. Therefore, in this study, the preventive effect of loganin on oxidative stress-mediated cellular damage in human keratinocyte HaCaT cells was investigated. Our results show that loganin pretreatment in a non-toxic concentration range significantly improved cell survival in hydrogen peroxide (H₂O₂)-treated HaCaT cells, which was associated with inhibition of cell cycle arrest at the G2/M phase and induction of apoptosis. H₂O₂-induced DNA damage and reactive oxygen species (ROS) generation were also greatly reduced in the presence of loganin. Moreover, H₂O₂ treatment enhanced the cytoplasmic release of cytochrome c, upregulation of the Bax/Bcl-2 ratio and degradation of cleavage of poly (ADP-ribose) polymerase, whereas loganin remarkably suppressed these changes. In addition, loganin obviously attenuated H₂O₂-induced autophagy while inhibiting the increased accumulation of autophagosome proteins, including microtubule-associated protein 1 light chain 3-II and Beclin-1, and p62, an autophagy substrate protein, in H₂O₂-treated cells. In conclusion, our current results suggest that loganin could protect HaCaT keratinocytes from H₂O₂-induced cellular injury by inhibiting mitochondrial dysfunction, autophagy and apoptosis. This finding indicates the applicability of loganin in the prevention and treatment of skin diseases caused by oxidative damage.

Keywords Loganin, ROS, DNA damage, apoptosis, autophagy

1. Introduction

Corni fructus, a fruit of Cornus officinalis Sieb. et Zucc. belonging to the Cornaceae family, has been widely used in East Asia including Korea for the purpose of tonifying the kidneys and liver (1-3). Secondary metabolites, including iridoids, are abundantly present in Corni fructus, and studies on the various pharmacological effects of these substances are increasing (3-5). Among them, loganin is an iridoid glycoside compound found and has been reported to have various beneficial effects, including anti-inflammatory, neuroprotective, inhibiting cartilage degeneration and osteoarthritis, renal protection and improving intestinal function (6-12). These pharmacological activities of loganin are believed to be at least related to its antioxidant effect. For example, loganin protected against hydrogen peroxide (H₂O₂) and amyloid-β-induced neurotoxicity while inhibiting the production of reactive oxygen species (ROS) (13,14), suggesting that its antioxidant effect was closely associated with ROS scavenging activity. In addition, this compound suppressed diabetes mellitus-induced reproductive damage by restoring glutathione level and superoxide dismutase activity, as well as reducing
ROS level (15). Recently, Wen et al. (9) reported that loganin reduced burn-induced intestinal inflammation and oxidative stress, and Cheng et al. (16) found that inhibition of NLRP3 inflammasome activation by the antioxidant activity of loganin contributed to the blockade of Schwann cell pyroptosis. Moreover, Xu et al. (12) have been reported that the antioxidant activity of loganin contributes to neuronal survival by inhibiting autophagy and mitochondrial division. Similar to these results, we also demonstrated that loganin may be a substance capable of preventing inflammatory and oxidative damage in lipopolysaccharide-stimulated macrophages (17).

Although ROS play an important role as a second messenger during normal cellular metabolism, excessive production of ROS results in progressive oxidative damage to organelles [18, 19]. In particular, ROS act as regulators of body homeostasis, including epidermal keratinocyte proliferation, exacerbating skin aging and has been implicated in various skin diseases (20,21). As in other tissues, oxidative stress caused by excessive accumulation of ROS in keratinocytes ultimately induces depolarization of the mitochondrial membrane potential (MMP, ΔΨm), a hallmark of mitochondrial dysfunction. Subsequently, cytochrome c is released from the mitochondria into the cytoplasm, which activates the caspase cascade and ultimately induces apoptosis (22,23). More recently, Liu et al. (24) suggested that catalpol, a type of natural iridoid glucoside, may have therapeutic properties for psoriasis by ameliorating oxidative stress in tumor necrosis factor-α-stimulated keratinocytes. In addition, natural iridoid glucoside derivatives such as geniposide and aucubin have been reported to have protective effects on oxidative stress induced by ultraviolet-B irradiation in human skin fibroblasts (25,26). However, since the beneficial effect of loganin on epidermal keratinocytes has not been clearly elucidated, in this study, the protective potential of loganin against H2O2-induced oxidative stress in human keratinocyte HaCaT cells was investigated.

2. Materials and Methods

2.1. Cell culture and treatment

HaCaT keratinocytes purchased from the American Type Culture Collection (CRL-1458™, Manassas, VA, USA) were cultured in Dulbecco's modified Eagle's medium containing 10% fetal bovine serum and 1% antibiotics at 37°C and 5% CO2. All materials needed for cell culture were obtained from WelGENE Inc. (Gyungsan, Korea). Loganin and H2O2 (Sigma-Aldrich Chemical Co., St. Louis, MO, USA) were dissolved in dimethyl sulfoxide (Thermo Fisher Scientific, Waltham, MA, USA) and distilled water to prepare stock solutions of 100 mM each. The stock solutions were diluted in the medium before utilization.

2.2. Cell viability

A 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay is used to measure cell viability, as previously described (27). Briefly, HaCaT cells were stimulated with the different concentrations of loganin or H2O2 alone for 24 h, or exposed to loganin for 1 h and then treated with H2O2 for 24 h. At the end time, the MTT reaction was run and absorbance was measured with a microplate reader (Beckman Coulter Inc., Brea, CA, USA) at the Core Facility Center for Tissue Regeneration (Dong-eui University, Busan, Korea). Images of cell morphological changes were captured using an inverted microscope (Carl Zeiss, Oberkochen, Germany).

2.3. Cell cycle analysis

For cell cycle analysis of H2O2-treated or untreated cells with and without loganin for 24 h, both adherent and detached cells were washed with phosphate-buffered saline (PBS) and then fixed by ethanol, as previously described (28). After that, cells were exposed with RNAase and propidium iodide (PI) (Thermo Fisher Scientific, Waltham, MA, USA) for 20 min at 4°C. Cell cycle distributions were calculated after appropriate gating of cell populations using flow cytometry (Becton Dickinson, San Jose, CA, USA).

2.4. Apoptosis analysis

To observe whether apoptosis was induced, flow cytometry was performed according to a published method (29). Briefly, cells were stimulated with H2O2 for 24 h with or without loganin, washed with PBS and then fixed with 4% paraformaldehyde solution for 15 min. Subsequently, cells were stained with annexin V-fluorescein isothiocyanate (FITC)/propidium iodide (PI) (Becton Dickinson), and annexin V-positive cells were quantified as cells induced by apoptosis using a flow cytometer.

2.5. Analysis of intracellular ROS

To measure the levels of ROS, the collected cells were stained with 2',7'-dichlorofluorescein diacetate (DCF-DA) dye (Sigma-Aldrich Chemical Co.). In brief, treated cells were collected and preloaded with 10 μM DCF-DA for 20 min at 37°C in the dark. The fluorescence intensity was observed under a fluorescence microscope (Carl Zeiss). A comet assay® kit obtained from Trevigen, Inc.
As indicated in Figure 1A, loganin has been no cytotoxicity up to 12.5 μM of concentration in HaCaT cells. However, cells exposed to 15 μM loganin showed a slight inhibition in cell viability. Therefore, 12.5 μM was selected the maximum concentration of loganin to establish its efficacy, we performed further experiments. The cell viability in HaCaT cells treated with 500 μM H₂O₂ was significantly reduced to approximately 60% compared that in the untreated control cells (Figure 1B). However, pretreatment of loganin significantly inhibited H₂O₂-induced reduction of cell viability in a concentration-dependent manner (Figure 1C). Especially, exposure to NAC as a positive control restored cell viability that could control the level of H₂O₂-stimulated cells. In addition, morphological alterations such as cell shrinkage and cytoplasm vacuolization were observed in H₂O₂-treated cells, but not in the presence of loganin (Figure 1D).

3.2. Loga

To measure MMP, the cells were stained with 5,5′,6,6′-tetrachloro-1,1′,3,3′-tetraethylrhodofluorocyanine iodide (JC-1, Thermo Fisher Scientific) reagent according to the previous methods (35,36), and quantitative analysis was performed using a flow cytometer, and fluorescence images were taken with a fluorescence microscope.

2.10. Autophagic activity detection

To evaluate the occurrence of autophagy, the Cyto-ID® Autophagy Detection Kit (Enzo Life Sciences Inc, Farmingdale, NY, USA) was employed according to the manufacturer’s instructions. In brief, the treated cells were collected, suspended, stained with the Cyto-ID staining solution provided in the kit at RT temperature in the dark, and then autophagy-induced cells were quantitatively analyzed using flow cytometry (36). In parallel with this, DAPI staining was used to counterstain the nuclei, and localized green fluorescence of autophagic cells was detected under a fluorescence microscope (30).

2.11. Statistical analysis

All experiments were independently repeated at least 3 times to determine significance. Results were presented as mean and standard deviation (SD) using SPSS 25.0 (SPSS Inc., Chicago, IL, USA), and differences (p < 0.05) were considered statistically using ANOVA-Tukey’s post-hoc test.

3. Results

3.1. Logain prevented the loss of HaCaT cell viability caused by H₂O₂ treatment

Whole cell lysates for immunoblotting were prepared according to a previous method (33). To isolate the cytoplasmic and mitochondrial fractions, a mitochondrial/cytoplasmic fractionation kit (Active Motif, Inc., Carlsbad, CA, USA) was used. For Western blot analysis, the same amount of protein for each treatment group was separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis and transferred to polyvinylidene difluoride membranes, and then the primary antibodies purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA), Abcam, Inc., Thermo Fisher Scientific and Cell Signaling Technology (Danvers, MA, USA) were probed, as previously described (34). Subsequently, the membranes that had finished reacting with the primary antibodies reacted with the correlated secondary antibodies (Santa Cruz Biotechnology, Inc.). The membranes were then exposed enhanced chemiluminescence solution (Thermo Fisher Scientific) and visualized using a Fusion FX Imaging System (Vilber Lourmat, Torcy, France). Band intensities were quantified and normalized to a loading control by densitometry using ImageJ® software (v1.48, National Institutes of Health, Bethesda, MD, USA).

2.8. Western blotting for protein expression analysis

MMP assay

To isolate the reducing power of 500 μM H₂O₂-induced reduction of cell viability in a concentration-dependent manner (Figure 1C). Especially, exposure to NAC as a positive control restored cell viability that could control the level of H₂O₂-stimulated cells. In addition, morphological alterations such as cell shrinkage and cytoplasm vacuolization were observed in H₂O₂-treated cells, but not in the presence of loganin (Figure 1D).

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3.2. Logain restored cell cycle arrest and apoptotic cell death in H₂O₂-treated HaCaT cells

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As shown in Figure 2A, the population of cells belonging to the G2/M phase in H2O2-treated HaCaT cells was significantly increased compared to the untreated control group, which was attenuated by loganin pretreatment. The proportion of cells with sub-G1 DNA content, which indicates the frequency of apoptosis, also greatly increased after H2O2 treatment compared with control cells, which significantly diminished by pretreatment of loganin (Figure 2A and 2B). In addition, the population of annexin V-positive cells was also significantly increased in H2O2-treated HaCaT cells (Figure 2C and 2D), which was markedly suppressed by loganin pretreatment. The results indicate that, similar to the NAC pretreatment group, loganin substantially attenuated cell cycle arrest and apoptotic cell death following by H2O2, thereby restoring cell viability.

3.3. Loganin inhibited ROS production in H2O2-stimulated HaCaT cells

The results of flow cytometry after DCF-DA staining showed that the level of intracellular ROS production was markedly increased in H2O2-exposed HaCaT cells (Figure 3A and 3B). Consistent with this result, the DCF-fluorescence image in H2O2-treated cells was markedly increased compared to that of control cells (Figure 3C). However, similar to NAC pretreatment, loganin significantly decreased H2O2-induced ROS generation, indicating that the inhibitory effect of loganin on the cytotoxicity of H2O2 in HaCaT cells was related to its antioxidant activity.

3.4. Loganin suppressed DNA damage in H2O2-treated HaCaT cells
Next, we investigated the inhibitory effect of loganin on DNA damage induced by H$_2$O$_2$-treatment. According to the comet assay results, DNA tails caused by damaged DNA fragments were markedly enhanced in H$_2$O$_2$-treated HaCaT cells (Figure 4A). However, these tails were markedly reduced in cells pretreated with loganin and NAC. Immunofluorescence indicated that H$_2$O$_2$ significantly increased the number of p-γH2AX positive-stained nuclei compared to control cells (Figure 4B). In addition, the expression of p-γH2A.X protein was strongly enhanced in H$_2$O$_2$-treated cells (Figure 4C and 4D). However, its expression was attenuated by loganin pretreatment, indicating that the inhibitory effect of loganin against H$_2$O$_2$-induced DNA damage was related to inhibition of ROS generation.

3.5. Loganin alleviated the change of apoptosis regulators expression in H$_2$O$_2$-treated HaCaT cells

As indicated in Figure 5A and 5B, the protein level of Bax was up-regulated in H$_2$O$_2$-treated HaCaT cells, while that of Bcl-2 was down-regulated. In addition, H$_2$O$_2$ enhanced the degradation of poly (ADP-ribose) polymerase (PARP). Furthermore, the level of cytochrome c expression in the mitochondria of cells treated with H$_2$O$_2$ was decreased, but its expression in the cytoplasm was increased (Figure 5C and 5D), which was associated with loss of MMP (Figure 5E and 5F). However, H$_2$O$_2$-induced these alterations were remarkably suppressed with loganin pretreatment, indicating that loganin protected HaCaT cells from apoptosis by blocking mitochondrial damage caused by H$_2$O$_2$.
3.6. Loganin attenuates H$_2$O$_2$-induced autophagy in HaCaT cells

Since the formation of autophagic vacuoles, which is typical of autophagy induction, was increased in H$_2$O$_2$-treated HaCaT cells (Figure 1D), we investigated whether autophagy was induced by H$_2$O$_2$ and whether loganin could inhibit it. The results of flow cytometry using Cyto-ID staining showed that autophagy was definitely induced in the cells treated with H$_2$O$_2$, as shown in Figure 6A and 6B, which was greatly reduced in the presence of loganin, and the same results were observed in the results of fluorescence microscopy (Figure 6C). Subsequently we detected autophagy biomarkers using Western blot analysis to confirm H$_2$O$_2$-induced autophagy, and found that H$_2$O$_2$ increased the accumulation of proteins such as microtubule-associated protein 1 light chain 3 (LC3)-II, beclin-1 and p62 (Figure 6D and 6E). However, their expression was obviously attenuated when loganin and H$_2$O$_2$ were treated together, suggesting that autophagy inhibition was implicated in the protection of HaCaT cells by loganin from oxidative damage caused by H$_2$O$_2$.

4. Discussion

In this study, we examined the efficacy of loganin on oxidative damage in H$_2$O$_2$-stimulated HaCaT keratinocytes. Our finding indicated that loganin significantly inhibited H$_2$O$_2$-induced cellular dysfunctions, including cell cycle arrest at the G2/M phase, DNA damage and apoptotic cell death, which was caused by blocking of ROS accumulation. Furthermore, we showed that the antioxidant potential of loganin was accompanied by inhibition of autophagy H$_2$O$_2$-treated HaCaT cells.

As is well known, H$_2$O$_2$, as oxidative stressor, induce cell cycle arrest at the G2/M phase in most cells, including keratinocytes, causing to DNA damage as well as cell death (37-39). In this study, reduction of cell survival in H$_2$O$_2$-treated HaCaT cells was accompanied by inhibition of cell cycle progression at the G2/M phase, which was in good agreement with the previous findings (38,40). However, these effects were significantly inhibited by loganin pretreatment. We also demonstrated that, in good agreement with previous studies (9,13-17),...
loganin had potent antioxidant activity by significantly inhibiting H\(_2\)O\(_2\)-induced ROS generation. Oxidative stress induces damage to intracellular macromolecules such as nucleic acids, contributing to DNA damage and apoptosis (41,42). The results of the comet assay, a commonly used method to measure DNA strand breaks (43), showed that loganin largely blocked H\(_2\)O\(_2\)-induced comet tail formation. Additionally, in the immunoblotting results for analyzing the phosphorylation level of γH2AX (p-γH2AX), which indicates that the DNA double-strand is broken by oxidative stress (33), enhanced expression of p-γH2AX by H\(_2\)O\(_2\) was effectively suppressed in the presence of loganin. These findings demonstrated that loganin has a remarkable ameliorating effect for H\(_2\)O\(_2\)-triggered DNA damage in HaCaT keratinocytes.

According to the results of previous studies, H\(_2\)O\(_2\)-induced apoptosis in HaCaT cells was strongly correlated with the cytosolic release of apoptogenic factors, including cytochrome c, which is initiated by mitochondrial-mediated intrinsic apoptosis pathway (40,44,45). Cytochrome c released into the cytoplasm following the loss of MMP activates effector caspases such as caspase-3 and -7 through the activation of caspase-9, which induce degradation of matrix proteins including PARP to terminate apoptosis (46,47). In this study, the expression of cytochrome c was up-regulated in the cytoplasm and mitochondria and the level of MMP were maintained at control levels in loganin-pretreated cells, suggesting that mitochondrial integrity in H\(_2\)O\(_2\)-treated HaCaT cells was maintained in the presence of loganin. Subsequently, the increase of Bax/Bcl-2 ratio by H\(_2\)O\(_2\) was also counteracted by loganin pretreatment, which was correlated with the cleavage of PARP. Bcl-2 family members control the release of apoptogenic factors through regulation of mitochondrial membrane permeability (46,47). Therefore, it is presumed that the reduction of the Bax/Bcl-2 expression ratio by loganin...
plays a critical role in attenuating H$_2$O$_2$-induced HaCaT cell apoptosis. Based on this finding, we considered that loganin might be a potential antioxidant that prevents mitochondrial-dependent apoptosis as a scavenger of ROS. Our results agree very well with the results of Kwon et al. (14) on the blocking effect of loganin on the induction of apoptosis by H$_2$O$_2$ in SH-SY5Y neuronal cells.

Cellular damage, including DNA damage and apoptosis due to oxidative stress, is often accompanied by autophagy (48,49). In keratinocytes, it has also been reported that autophagy is involved in DNA damage and apoptosis caused by oxidative stress inducers (50,51), suggesting that ROS are potent and effective triggers of autophagy. Autophagy is a highly conserved self-digestion process for recycling cytoplasmic components such as unwanted protein aggregates and damaged organelles, which are sequestered into newly generated double-membraned structures called autophagosomes (52,53). Autophagosomes fuse sequentially with lysosomes and are eliminated through lysosomal degradation (54,55). Several important proteins are involved in this process, including LC3-II, Beclin-1 and p62. LC-3, an autophagosome membrane protein, controls key steps in the autophagic pathway, such as autophagic membrane growth and lysosome fusing, and the ratio LC3-II/LC3-I is commonly used to reflect autophagosome formation (52,56). Beclin-1 is involved in the initiation of autophagy by mediating the localization of autophagy proteins into the pre-autophagosomal membrane. In addition, Beclin-1 may be importantly involved in the regulation of apoptosis as well as autophagy because pro-autophagy properties may be reduced by Bcl-2 (57,58). p62 is another autophagosome-lysosomal membrane-associated protein that serves as an autophagic substrate (59,60). Therefore, these proteins are important markers of autophagy flux and critical regulators of autophagy regulation. The flow cytometry results of this study showed that the frequency of autophagosomes increased in H$_2$O$_2$-treated HaCaT cells, and the accumulation of LC-3, Beclin-1 and p62 was enhanced after H$_2$O$_2$ treatment, as determined by Western blotting. These observations support that autophagy as well as apoptosis are important mechanisms of H$_2$O$_2$-mediated cytotoxicity in HaCaT cells. And, in the presence of loganin, these autophagy markers were remarkably reversed compared to cells treated with H$_2$O$_2$ alone, showing that H$_2$O$_2$-induced HaCaT cell autophagy was clearly reduced by loganin supplementation. However, since autophagy has dual roles of pro-survival and pro-death depending on circumstances, additional mechanistic studies are required on how the alleviated effect of loganin on H$_2$O$_2$-induced autophagy affects the survival of HaCaT cells.

In the current study, we evaluated the efficacy of loganin on H$_2$O$_2$-mediated oxidative damage in HaCaT keratinocytes. Our results revealed that loganin visibly diminished cell cycle arrest at the G2/M phase, DNA damage, autophagy and apoptosis in H$_2$O$_2$-stimulated HaCaT cells, which was linked to its ability to suppress ROS accumulation. Additionally, the anti-apoptotic effect by loganin was a result of blockade of mitochondrial dysfunction, which correlated with inhibition of cytoplasmic release of cytochrome c due to inhibition of increased Bax/Bcl-2 ratio (Figure 7). This study provides a theoretical basis for the inhibitory mechanism of oxidative stress-mediated cellular damage of loganin and its application as a novel therapeutic agent to counteract oxidative stress-mediated skin diseases.

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