Oral administration of L-arginine in patients with angina or following myocardial infarction may be protective by increasing plasma superoxide dismutase and total thiols with reduction in serum cholesterol and xanthine oxidase

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Abbreviations: SOD, super oxide dismutase; SH, sulfhydryl; XO, xanthine oxidase; ROS, reactive oxygen species; SL, sarcolemmal; SR, sarcoplasmic reticulum; GPx, glutathione peroxidase; GR, glutathione reductase; EDTA, ethylene diamine tetra acetate acid; NBT, nitroblue tetrazolium; TCA, tri chloro acetic acid; DTC, dinitrophenylhydrazine, thiourea, copper sulphate; DTNB, 2,2-dithio bis nitrobenzoic acid; MDA, malondialdehyde; TBA, thiobarbituric acid; SDS, sodium dodecyl sulphate; TEP, 1,1,3,3-tetraethoxy propane; DNPH, 2,4-dinitrophenyl hydrazine; DHA, dehydro ascorbic acid; ANOVA, analysis of variance; MI, myocardial infarction; AA, acute angina; NO, nitric oxide

Key words: L-arginine, oxidative stress, myocardial injury, nitric oxide, myocardial infarction, acute angina

Introduction

Myocardial ischemia can result from a variety of pathological and physiological disorders. Whatever the cause of ischemia, the consequence is always decreased oxygen availability to the myocardium. Insufficient oxygen supply fails to support oxidative phosphorylation. Within few minutes of ischemia, to meet the energy demand, glycolytic pathway is greatly stimulated with resultant accumulation of lactic acid resulting in acidosis. Anaerobic production of ATP in this situation is insufficient to meet the energy demand of the tissues. With continuing ischemia the tissue levels of ATP fall and this initiates a series of events, especially oxidative stress conditions, which are deleterious to the endothelial cells. Oxidative stress, associated with increased formation of reactive oxygen species (ROS), modifies phospholipids and proteins leading to lipid per-oxidation and oxidation of thiol groups. These changes are considered to alter the membrane permeability and configuration.1,2 Oxidative stress may result in cellular defects including a depression in the sarcolemmal (SL) Ca²⁺ pump, ATPase and Na⁺-K⁺ ATPase activities. These changes lead to decreased Ca²⁺ efflux and increased Ca²⁺ influx and thus inhibit Ca²⁺ sequestration from the cytoplasm in the cardiomyocytes.3,4 The oxidative stress induced changes in sarcoplasmic reticulum (SR) Ca²⁺ and SL Na⁺-K⁺ pumps are not limited only to cardiomyocytes but have also been observed in the coronary artery smooth muscle cells. The depression in Ca²⁺ regulatory mechanism by ROS ultimately results in intracellular Ca²⁺ overload and cell death. On the other hand, an increase in Ca²⁺ during ischemia induces conversion of xanthine dehydrogenase to xanthine oxidase, by selective proteolysis, that generates more of super oxide radicals. The super
oxide radicals, though less toxic by itself, triggers the formation of other reactive oxygen species. These include OH•, H2O2 and HOCl. The hydroxyl radicals, in particular, interact with lipids, proteins and nucleic acids resulting in the loss of membrane integrity, structural and functional changes in enzymes, proteins and genetic material. These deleterious effects result in the loss of heart contractile function and severe myocardial cell damage. To counter the damaging effects of ROS, body has evolved certain defense mechanisms to deactivate/destroy them before they can cause damage. The oxidative damage will occur only in situations where the defense mechanisms are deficient, made less active or production of ROS exceeds the capabilities of defense mechanism or a combination of all these. The first line of defense in the body are enzymatic free radical scavengers such as superoxide dismutase (SOD), catalase, glutathione peroxidase (GPx) and glutathione reductase (GR). SOD is considered to be the first line of defense against oxidative insult, as this enzyme efficiently converts oxygen derived free radical to hydrogen peroxide. Hydrogen peroxide is more toxic than oxygen derived free radicals and is capable of producing most toxic OH• radical. It has, therefore, to be removed very efficiently. Catalase is the highly reactive enzyme in most of the tissues that converts toxic H2O2 to water. Another enzyme that removes H2O2 is glutathione peroxidase. This enzyme, almost equally efficient as catalase, removes H2O2 at the expense of oxidation of glutathione. Oxidized glutathione is reduced back by glutathione peroxidase. In fact catalase and glutathione peroxidase cooperate in removing H2O2 in many tissues. For example, mammalian red blood cells (RBCs) have both these enzymes. The normal low rate of production of H2O2 by hemoglobin and SOD seems mainly dealt with glutathione peroxidase. However, catalase also contributes to removal of H2O2 if its concentration is raised. All these enzymes convert ROS to less toxic or non toxic products. The second line of defense to ROS is provided by anti-oxidant compounds present in the body such as thiols, or taken from outside such as α-tocopherol, β-carotenoids, flavonoids and L-arginine. We have earlier shown that administration of vitamin C and vitamin E to the patients of ischemic myocardial syndromes lower lipid per-oxidation, increase the activities of free radical scavenging enzymes, which are decreased in these patients, and lower the activity of pro-oxidant enzyme which are elevated in the patients. Oxidized low density lipo-proteins are involved in the progression of atherosclerotic lesions which can be prevented by L-arginine administration. Moreover there are evidences that this amino acid preserves arterial vasodilation even in the presence of oxidative stress. A fine balance between ROS and various anti-oxidant mechanisms is crucial for avoiding myocardial injury. In the present study we have tried to evaluate the effect of L-arginine administration on certain pro-oxidant and anti-oxidant parameters in myocardial ischemic syndromes like acute myocardial infarction and acute angina.

Results

Myocardial ischemic syndromes are almost invariably associated with alterations with oxidant and anti-oxidant parameters. Levels of SOD, an important enzymatic free radical scavenger and XO, the major source of free radical generation have been selected in the present study. MDA levels has been used as marker of free radical mediated damage along with carbonyl content. Levels of some non-enzymatic anti-oxidants namely total thiols and ascorbic acid and levels of serum cholesterol have been monitored before and after administration of L-arginine.

The results are expressed as Mean ± SD. One way analysis of variance (ANOVA) followed by Newman-Keul’s multiple comparison test is applied to assess the significance of the data. A p < 0.05 is considered to be statistically significant. Personal data of the subjects included in the study are reported in Table 1. Age group, blood pressure and serum cholesterol levels of the subjects are reported in Table 2. Biochemical parameters investigated are presented in Tables 3 and 4. All the data are also represented graphically in Figures 1–7.

SOD. The specific activity of SOD in plasma of MI patients receiving L-arginine treatment along with regular anti-hypertensive therapy (Group 2) showed significant increase (p < 0.05) when compared to those receiving only anti-hypertensive therapy
Table 3 Effect of L-arginine (Arg) administration on the activities of SOD and XO and MDA levels in patients of acute myocardial infarction (MI) and acute angina (AA)

| Groups   | SOD  | XO   | MDA  |
|----------|------|------|------|
|          | unit/mg protein | unit/mg protein | (nmole/ml) |
| Group 1  | 1.91 ± 0.79 | 0.94 ± 0.17 | 4.61 ± 1.27 |
| Group 2  | 2.32 ± 0.68 | 0.68 ± 0.16 | 3.65 ± 1.24 |
| Group 3  | 2.80 ± 0.71 | 0.82 ± 0.15 | 1.91 ± 0.75 |
| Group 4  | 3.05 ± 0.77 | 0.65 ± 0.14 | 1.40 ± 0.48 |

SOD activity in patients was estimated by the method of Misra and Fridovich. Activity of XO was estimated by the method of Fried and Fried. Levels of MDA was determined by Ohkawa et al. Values are presented as mean ± SD. Statistical significance of the groups was determined by one way ANOVA followed by Newman-Keul’s multiple comparison test. p values: Superoxide dismutase (SOD), Group 1 vs. Group 2 p < 0.05; Group 3 vs. Group 4 p > 0.05. Xanthine oxidase (XO), Group 1 vs. Group 2 p < 0.005; Group 3 vs. Group 4 p < 0.05. Malondialdehyde (MDA), Group 1 vs. Group 2 p < 0.05; Group 3 vs. Group 4 p < 0.05.

Table 4 Effect of L-arginine (Arg) administration on the levels of total thiols, ascorbic acid and carbonyl content in patients of acute myocardial infarction (MI) and acute angina (AA)

| Groups   | Total thiols | Ascorbate level | Carbonyl protein |
|----------|--------------|-----------------|-----------------|
|          | (nmole/ml)   | (mg/dl)         | (μmole/ml)      |
| Group 1  | 0.22 ± 0.067 | 0.26 ± 0.88     | 54.28 ± 22.92   |
| Group 2  | 0.31 ± 0.068 | 0.37 ± 0.16     | 38.38 ± 17.48   |
| Group 3  | 0.32 ± 0.041 | 0.39 ± 0.10     | 21.27 ± 11.57   |
| Group 4  | 0.39 ± 0.072 | 0.49 ± 0.15     | 16.82 ± 7.11    |

Levels of total thiols in patients were estimated by the method of Omaye et al. Ascorbic acid level was determined by the method of Omura et al. Carbonyl content was determined by the method of Leveni et al. Values are presented as mean ± SD. Statistical significance of the groups was determined by one way ANOVA followed by Newman-Keul’s multiple comparison test. p values: Total thiols: Group 1 vs. Group 2 p < 0.005; Group 3 vs. Group 4 p < 0.05. Ascorbic acid: Group 1 vs. Group 2 p < 0.05; Group 3 vs. Group 4 p < 0.05. Carbonyl content: Group 1 vs. Group 2 p < 0.05; Group 3 vs. Group 4 p > 0.05.

Discussion

As reported in the results section, most of the oxidant-antioxidant parameters studied, showed a tendency to improve upon administration of L-arginine. Most of the xanthine oxidizing activity present in healthy animal tissues occurs as a dehydrogenase enzyme that oxidizes hypoxanthine and/or xanthine to uric acid at the expense of NAD. During ischemia, insufficient oxygen supply fails to support oxidative phosphorylation, leading to lowering of tissue ATP levels and acidosis. ATP hydrolysis eventually leads to accumulation of substrates of xanthine oxidase.18 As mentioned in the introduction section, levels of Ca^2+ are also increased during ischemia. This increase in Ca^2+ stimulates certain proteases that selectively act on xanthine dehydrogenase to convert it to xanthine oxidase.19 Xanthine oxidase mediated production of OFRs plays major role in causing tissue damage observed in ischemia/reperfusion injury as evidenced by the fact that incorporation of specific inhibitors of xanthine oxidase, such as allopurinol and anti-oxidants, are capable to protect such damage in experimental models of myocardial injury.20-22 The series of events initiated by ischemia, lead to enhanced production of free radicals which is further increased upon reperfusion of ischemic myocardium.10,11 Oxy free radicals are removed by superoxide dismutase in healthy organisms, but during ischemia, the lowered activity of SOD caused by inhibition of the enzyme by excess H2O2,
production fails to cope with excessive production of free radicals and H$_2$O$_2$. This excess H$_2$O$_2$, besides inhibiting SOD, can cause degradation of heme rings of hemoglobin releasing iron which is capable of OH production via Fenton reaction and lipid peroxidation. Lipid per-oxidation and free radical mediated damage during ischemia is indicated by increased MDA levels in patients suffering from myocardial infarction and acute angina in the present study. Total thiols can either chemically or enzymatically reduce dehydroascorbic acid (DHA) to ascorbate. The lower level of total thiols in patients may be responsible for the decreased conversion of DHA back to ascorbate. Ascorbic acid, an effective antioxidant is capable of completely protecting lipids against oxidative damage induced by peroxyl radicals. It has chain breaking properties, reacts directly with superoxide ions, hydroxyl radicals and singlet oxygen species and acts to generate tocopherol.27 During ischemia the lowered levels of total thiols may be responsible for decreased conversion of DHA to ascorbate. This may be one of the reasons that we observed decreased levels of ascorbic acid in the patients. Decrease in ascorbate level fails to cope with per-oxidative damage of lipids and modification of phospholipids, thereby resulting in increased lipid per-oxidation. Decreased levels of total thiols also indicate the effect of oxidative damage of protein molecules due to excessive free radical generation during ischemia. Major parts of thiols are derived from proteins. Oxidative damage modifies proteins leading to oxidation of thiol groups.28 The modifications in lipids and proteins caused due to decreased levels of ascorbic acid and thiols in patients alter membrane permeability and configuration in addition to producing functional modification of various cellular proteins during ischemia. Excessive free radical generation during ischemia causes the inactivation of enzymes and introduction of carbonyl groups into amino acid side chains of proteins. Increased carbonyl derivatives within proteins and enzyme inactivation suggest mixed—function oxidative modification.29

During ischemia, the myocardial tissue is exposed to an exacerbated injury caused by un-scavenged superoxide radicals, other free radicals, increased WBC, vasoconstriction and platelet adhesion.30-32 Higher oxidant stress and diminished antioxidant status along with higher MDA levels constitutes the key factors in the progression of ischemic injury.

L-arginine serves as a precursor of nitric oxide (NO) synthesis. NO can reduce oxidative stress by inhibiting XO, scavenge superoxide radicals and terminate free radical chain reaction within the lipid membranes, thereby reducing inflammatory mediators.33,34 Decreased ROS formation attenuates inhibition of SOD which is increased by L-arginine supplementation. In the present study, it was found that the level of ascorbic acid and total thiols also increased after L-arginine supplementation. Our present finding is in accordance with.35 The improved levels of ascorbic acid and total thiols cope with lipid and protein per-oxidation thus decreasing the damage of membrane and cellular proteins. By efficiently reducing or eliminating these toxic metabolites L-arginine via NO retards the damaging effects of ischemic injury.36

L-arginine controls the modification of proteins due to excess carbonylation and that is why in our study, L-arginine was found to significantly decrease the plasma carbonyl contents in the patients.
Effect of L-arginine in certain ischemic myocardial disorders

The observed lowering of serum cholesterol upon L-arginine administration, has been ascribed to be mediated by NO derived from this amino acid. The adhesiveness of mononuclear cells is increased in hyper-cholesterolemic conditions. The increase in adhesiveness and cholesterol levels lead to atherosclerosis and myocardial infarction and can be taken care of by L-arginine supplementation to these patients. Possible pathway presenting the action of L-arginine on the parameters examined in the present study is provided in Figure 8.

L-arginine supplementation, along with standard therapy in the patients under present study, can maintain oxidant-antioxidant homeostasis and control the levels of lipid, cholesterol, ascorbic acid and protein oxidation in patients of myocardial ischemic syndromes.

Material and Methods

The study was cleared by the departmental ethical committee. Informed consent was obtained from individuals enrolled in the study. The study included 60 patients of the age group between 40–75 suffering from myocardial infarction and acute angina.

The patients suffering from non segment elevation myocardial infarction before reperfusion were included in MI group of the study. These patients were receiving regular antihypertensive medication.

Patients with chest pain at rest and/or on effort, positive exercise tests (chest pain associated with ST-segment depression of >0.1 mV) normal coronary arteriograms and no spasm of large epicardial arteries provoked by acetylcholine were included in...
acute angina group. All these patients exhibited evidence suggestive of myocardial ischemia. None of these patients had any ST-segment abnormalities in their resting ECG’s. The patients were receiving anti anginal medication.

Patients with previous cardiovascular or other organic diseases and those with severe trauma or surgery in preceding eight weeks were excluded from the study.

The patients were grouped in the following way:

- Group (1): included 35 MI patients receiving regular therapy without L-arginine administration.
- Group (2): included 20 MI patients assigned to regular therapy along with supplementation of oral dose of L-arginine (3 g/day) for 15 days.
- Group (3): included 25 patients of acute angina receiving regular therapy without L-arginine supplementation.
- Group (4): comprised of 20 patients of acute angina with conventional therapy along with supplementation of L-arginine (3 g/day) for 15 days.

Regular treatments for both the groups (MI and acute angina) included antihypertensive drugs like nitroglycerine, metaprolol, aspirin and clopivas.

All the chemicals employed in the study were AnalaR grade of Qualigens, biochemicals employed were procured from Sigma Chemical Co., US.

Preparation of sample. Venous blood (4.0 ml) was drawn aseptically from the subjects under study and transferred in polypropylene tubes containing 0.5 ml 3.8% (w/v) sodium citrate, pH 7.2. The tubes were gently rotated to mix the contents and centrifuged at 2,000 xg for 20 min at 4°C. The supernatant plasma was used for study.

Estimation of SOD. SOD was assayed in plasma by the method of Misra and Fridovich. Three ml reaction mixture consisted of 1.5 ml of 100 mM carbonate buffer, pH 10.3; 0.01 ml of 30 mM EDTA (ethylene diamine tetra acet acid), suitable aliquot of enzyme preparation and water to make up the volume to 2.94 ml. The reaction was started by addition of 0.06 ml of 15 mM epinephrine. Change in absorbance was recorded at 480 nm for one min at 15 sec interval. Control consisting of all the ingredients, except enzyme preparation, was run simultaneously.

One unit of enzyme activity has been defined to cause 50% inhibition of auto-oxidation of epinephrine present in the assay system by 1 ml enzyme preparation.

Estimation of XO. XO (Xanthine oxidase) was assayed in plasma by the method of Fried and Fried. The reaction mixture consisted of 0.9 ml of 0.1 M phosphate buffer pH 7.8; 0.75 ml 10 mM EDTA; 0.15 ml 0.2 mg/ml phenazine methosulphate; 0.45 ml 4 mg/ml nitroblue tetrazolium (NBT) salt; 0.5 ml 1 mM xanthine and water to make up the volume to 3.5 ml. The reaction was started at 37°C by addition of 0.5 ml of 1 mM Xanthine for 1 min. Extinction coefficient of the reduced NBT at 540 nm is 7.2 cm/μmole.

One unit of enzyme activity has been defined as amount of enzyme that converts 1 μmole of xanthine to uric acid in one minute at specified conditions of assay.

Estimation of ascorbic acid. Ascorbate level was estimated in the plasma by the method of Omaye, Turnbull and Sauberlich. In the assay, 1.0 ml of plasma was precipitated with 1.0 ml of 10% (w/v) TCA (trichloro acetic acid). To 0.5 ml of the supernatant 0.2 ml of DTC reagent (3.0 g dinitrophenylhydrazine, 0.4 g thiourea and 0.05 g CuSO4·5H2O in 100 ml 9 N H2SO4) was added and the mixture was incubated at 37°C for 3 h. 1.5 ml of 65% H2SO4 was then added to the reaction mixture and kept at room temperature for 30 min. Absorbance was measured at 520 nm. L-ascorbic acid in 5% TCA was used as reference.

Estimation of total thiols. Total thiols (T-SH) were estimated by the method of Hu using Ellman’s reagent. In final volume of 4.0 ml, was added 0.2 ml plasma and 0.6 ml of 20 mM tris-HCl buffer pH 8.2, followed by addition of 0.04 ml of 10 mM 5,5′-dithionitrobenzoic acid (DTNB) in absolute methanol and 3.16 ml of absolute methanol. The tubes were capped and color was developed for 15 min at room temperature. The tubes were then centrifuged at 3,000 xg for 20 min. Supernatant was collected and absorbance measured at 412 nm.

Estimation of MDA. Levels of malondialdehyde (MDA) were measured as an index of oxidative stress by the method of Ohkawa, Ohishi and Yagi, using thiobarbituric acid (TBA) reagent. To 0.2 ml plasma, in a Folin tube, was added 0.2 ml, 8.1% (w/v), sodium dodecyl sulphate (SDS); 1.5 ml, 20% (v/v) acetic acid and 1.1 ml of 0.8% (w/v) thio-barbituric acid to make up the volume to 3 ml. The tubes were heated in a water bath at 95°C for 1 h and cooled immediately under running tap water. To each tube, 1.0 ml water and 5.0 ml solution of butanol and pyridine (15:1 v/v) were added, vortexed and centrifuged at 800 xg for 20 min. The upper layer was aspirated out and color intensity measured at 532 nm. 1,1,3,3-tetra ethoxy propane (TEP) was used as the reference.

Estimation of carbonyl content of plasma protein. Protein oxidation was determined by protein carbonyl content in plasma by a modified method of Levi et al. From 0.5 ml of plasma, proteins were precipitated using 0.25 ml of 10% TCA. 0.75 ml of 10 mM DNPH (2,4-dinitrophenylhydrazine) in 2 M HCl was added to the precipitate and incubated at room temp for 30 min. The reaction mixture was re-centrifuged using the same amount of TCA as before. The precipitate, thus obtained, was washed twice with 1 ml ethanol/ethyl acetate (1:1 v/v) mixture. The precipitate was dissolved in 0.75 ml protein dissolving solution (2 g SDS and 50 mg EDTA in 100 ml 80 mM phosphate buffer, pH 8.0), incubated for 10 min at room temp. The color intensity was measured at 370 nm against 2 M HCl. Carbonyl content was calculated using the molar extinction coefficient 22,000 M-1cm-1.

Estimation of serum cholesterol. Serum cholesterol was determined by the method of Sperry and Webb. To 0.4 ml serum, 0.96 ml water was added to make up the volume to 1.0 ml. To this 4.0 ml of color reagent (0.1 ml of 10.0 g FeCl3·6H2O per 100 ml saline and 1.1 ml of 36 N H2SO4) and 6.0 ml acetic acid was added. The mixture was allowed to stand for 20 min. The color intensity was measured at 570 nm.

Protein estimation. Protein estimation has been carried out by the method of Lowry et al. using Folin phenol reagent. Bovine serum albumin was used as standard.
Specific activity of the enzymes has been defined as unit/mg protein.

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