Population genetic structure analysis and identification of backfat thickness loci of Chinese synthetic Yunan pigs

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Yunan is a crossed lean meat pig breed in China. Backfat thickness is the gold standard for carcass quality grading. However, over 14 years after breed registration, the backfat of Yunan thickened and the consistency of backfat thickness decreased. Meanwhile, no genetic study has been ever performed on Yunan population. So, in this study we collected all the 120 nucleus individuals of Yunan and recorded six backfat traits of them, carried out population genetic structure analysis, selection signals analysis and genome-wide association study of Yunan pigs with the help of their founder population Duroc and Chinese native Huainan pigs, to determine the genomic loci on backfat of Yunan. Genetic diversity indexes suggested Yunan pigs had no inbreeding risk while population genetic structure showed they had few molecular pedigrees and were stratified. A total of 71 common selection signals affecting growth and fat deposition were detected by FST and XP-CLR methods. 34 significant loci associated with six backfat traits were detected, among which a 1.40 Mb region on SSC4 (20.03–21.43 Mb) were outstanding as the strong region underlying backfat. This region was common with the results of selection signature analysis, former reported QTLs for backfat and was common for different kinds of backfat traits at different development stage. ENPP2, EXT1 and SLC30A8 genes around were fat deposition related genes and were of Huainan pig's origin, among which Type 2 diabetes related gene SLC30A8 was the most reasonable for being in a 193.21 Kb haplotype block of the 1.40 Mb region. Our results had application value for conservation, mating and breeding improvement of backfat thickness of Yunan pigs and provided evidence for a human function gene might be reproduced in pigs.

KEYWORDS
synthetic pig breed, genetic diversity, population structure, selection signature, GWAS, backfat thickness
Introduction

Huainan is one of the oldest northern China native pig breeds. It is native to south of the upper reaches of Huai River and North of Dabie Mountains in Henan Province. Henan is located in the middle and lower reaches of the Yellow River and is one of the earliest pig domestication areas in China (Zhang et al., 2007). Huainan is an all-black pig breed with large body size, large ears, short mouth, strong and robust limbs, high fertility, strong adaptability and delicious meat quality (Wang et al., 2005). People in and around Huainan pig producing areas have the habit of eating black pigs. Black pig is an essential ingredient of the famous local cuisine there. However, the poor growth rate of Huainan pigs limited its development.

Since 1980s, a large number of foreign commercial pigs with fast growth rate and high lean meat rate have been introduced to China to improve growth performance of Chinese native pig breeds (Xu, 2004). Among them, the American Duroc with golden coat color was widely used because of its high growth rate and high feed conversion rate (Figure 1A). So, in 1996, we started to use Huainan and American Duroc pigs as the base herd to intercross to develop a new high-performance black pig, Yunan pig (Figures 1B,C) (Zhu et al., 2009).

Yunan pig is a black crossbred pig breed obtained by nine generations crossbreeding between Huainan and American Duroc (Figure 2). In 2008, Yunan was registered as a new pig breed in China by National Livestock and Poultry Genetic Resources Committee. Theoretically, it contains 37.50% of Huainan’s lineage and 62.50% of Duroc lineage. Yunan exhibits good growth performances from Duroc with an average daily gain of 648 g during the stage of body weight at 30 kg–90 kg and lean meat rate of 56%, and has good meat quality from Huainan with an excellent meat quality (intramuscular fat content as 4.11%). So, Yunan becomes a popular black pig breed in and around Henan.

Since the year of 2008, the systematic breeding of Yunan pigs have been no longer carried out which resulted in a decline of backfat consistency. Backfat thickness now is still the only gold standard for black pig carcass grading as same as the commercial pigs in China. Therefore, the decline of backfat performance of Yunan brings great economic loss to farmers. Meanwhile, with the emergence of the African swine fever in China in 2018, the population of Yunan pigs reduced seriously.

However, there has never been a genomic study on Yunan pigs. Only a few correlation analyses of several genes and comparative analysis of production traits of Yunan were reported (Li et al., 2016; Wang et al., 2021). Herein, in this study, we recorded six backfat thickness traits of the core group of Yunan population, and used SNP array genotyping data of Yunan, Huainan and Duroc with the following objectives: 1) determine the genetic diversity of Yunan by calculating the observed heterozygosity and the expected heterozygosity; 2) access the inbreeding state by detecting runs of homozygosity (ROH); 3) detect selection signals of Yunan by pairwise FSTS and XP-CLR methods; 4) investigate genomic evidence on the population structure of Yunan by phylogenetic tree, principal component and admixture analysis; 5) identify the genomic region that controlled backfat thickness of Yunan.

Material and methods

Animals sampling, genotyping and phenotyping

Ear tissues were collected from Yunan (n = 120) and Huainan (n = 33) pigs at Sungo Agricultural and Livestock Co. Henan, China. Genomic DNA was extracted using the phenol-chloroform method and genotyped by 50 K SNP (Compass) and 80 K SNP (NEOGEN). Five backfat thickness traits including shoulder backfat (SBF), sixth and seventh ribs backfat (SSRBF), last rib backfat (LRBF), lumbar joint backfat (LBF) and P2 backfat (P2BF) were measured and recorded for Yunan pigs using ultrasonic instrument (EXAGO, France). Average backfat (ABF) was calculated as the average of SBF, SSRBF, LBF and LRBF. Genotype data of American Duroc pigs (n = 40) (Illumina PorcineSNP60 Genotyping Bead Chip) was downloaded from the public Dryad database (http://dx.doi.org/10.5061/dryad.30tk6).

We used PLINK v.1.9 (Purcell et al., 2007) to perform the quality control (QC) of the total 193 individuals genotype data. SNPs without positions on pig reference genome (Sus scrofa 11.1), or with genotyping rate less than 90%, or with minor allele...
frequency (MAF) lower than 1% or on Y chromosome were excluded. Individuals with genotyping rate less than 90% were excluded. Genotypic data of the same SNP in Yunan, Huainan and Duroc populations were extracted for genetic diversity, genetic differentiation and population genetic structure analyses.

Genetic diversity and selection signal analysis

Genetic indicators including the observed heterozygosity (Ho), the expected heterozygosity (He) and MAF were calculated for Yunan, Huainan and Duroc by PLINK v.1.9 to compare the genetic diversity of these populations, command set is `-hardy`. In addition, SNePv1.1 (Barbato et al., 2015) software was used to calculate the effective population size (Ne).

The length and frequency of ROH can reflect the group history. A long ROH indicates recent inbreeding, while a short ROH indicates ancient inbreeding. Genomic homozygous fragments of each individual were detected using runs of homozygosity of PLINK v.1.9. The following parameters were used of define ROHs: 1) the minimum length of ROHs was 500 Kb; 2) a sliding window of 50 SNPs across the genome; 3) each window allowed one heterozygous genotype and five missing SNPs (Xu et al., 2021) to avoid false negatives caused by occasional genotyping errors and missing genotypes. Then the ratio of the total length of the ROH fragment to the total length of autosomal genome was calculated to get the coefficient of inbreeding (Silió et al., 2013) using the following formula:

$$F_{ROH} = \frac{\sum L_{ROH}}{L_{auto}},$$  

where $L_{ROH}$ is the sum of the lengths of the autosomal ROH fragments and $L_{auto}$ is the total size of 18 autosomes of pigs covered by SNPs, which is 2.265 Gb (https://www.ncbi.nlm.nih.gov).

We used SNP data from three breeds, using genetic differentiation coefficient ($F_{ST}$) and cross-population compound likelihood ratio test (XP-CLR) for genome-wide selective detection. The genetic differentiation coefficient ($F_{ST}$) between populations was calculated using Vcftools v.0.1.13 (Sun et al., 2020), The command set was as follows: 1) window size was set to 500 Kb; 2) step length was set to 40 Kb. The $F_{ST}$ values ranging from 0 to 0.05, 0.05 to 0.15, and 0.15 to 0.25 indicate that there are no genetic, moderate, and large differentiations among populations, respectively. XP-CLR was calculated using XP-CLR (Chen et al., 2010) software. XP-CLR models the difference in frequency of multiple alleles between the two populations. Further, we used the parameters ("-w1 0.005 200 2000 -p0 0.95") to calculate the XP-CLR score for each chromosome. The empirical cutoffs for the genomic windows with top 1% $F_{ST}$ and XP-CLR values across the whole genome were considered as selective sweeps.

Population genetic structure analysis

To assess the individual genetic distances between populations to illustrate the relationship between populations, we carried out principal components analysis (PCA) of the SNP dataset using PLINK v.1.9 and selected the first two principal components for visualization by ggplot package in R v.4.1.3 (Team, 2014). To gain more insight into the Yunan pig population, we performed t-distributed stochastic neighbour embedding (T-SNE) analysis on the dataset using the R package "Rtsne" (Krijthe, 2015). The phylogenetic tree was built by neighbor-joining (NJ) method and visualized using R v.4.1.3, the genetic distance matrix was constructed by PLINK v.1.9. To explore the population structure of Yunan, the rapid model of ADMIXTURE v.1.3 (Alexander et al., 2015) software was used to cluster all Yunan samples in a context of some related populations and the results were visualization using the "barplot" package of R v.4.1.3 software.

Genome-wide association analysis of backfat thickness

Genome-wide association analysis for backfat traits of Yunan was performed using a univariate mixed linear model of GEMMA v.0.98.5 (Zhou et al., 2012) as follows.

$$y = W\alpha + x\beta + u + e,$$  

where $y$ is the phenotype vector, $W$ is a design matrix which contains the fixed effects, $x$ is the design matrix which contains the genotypes, $\alpha$ is the fixed effect vector, $\beta$ is the vector of genetic effects, $u$ is the random additive genetic effect vector and $e$ is the vector of residual effects.
TABLE 1 Summary of population genetic diversity indexes.

| Breed     | No | Ne | MAF      | H₀     | He     |
|-----------|----|----|----------|--------|--------|
| Yunan     | 120| 67 | 0.2813   | 0.3756 | 0.3685 |
| Huainan   | 33 | 43 | 0.2337   | 0.3422 | 0.3193 |
| Duroc     | 40 | 77 | 0.2462   | 0.3019 | 0.3277 |

where $y$ is a vector of phenotypic values for the trait; $W$ is a matrix of fixed effects including the first three PCs; $\alpha$ is a vector of corresponding coefficients including the intercept; $x$ is a vector of genotypes for the marker; $\beta$ is an effect size for the marker; $u$ is an n-vector of random effects; and $\epsilon$ is an n-vector of errors.

The results of genome-wide association analysis were visualized using the rMVP (Yin et al., 2021) package in R software. The suggestive significance threshold was $1/N$, where $N$ is the number of SNPs used for the analysis. We used 1 Mb upstream and downstream significant SNPs region as traits related candidate region.

Results

Genotyping

There are 57,466 SNPs sites in the original data. After quality control, SNPs without positions on pig reference genome (Sus scrofa 11.1), or with genotyping rate less than 90%, or with minor allele frequency lower than 1% or on Y chromosome were excluded. Individuals with genotyping rate less than 90% were excluded. Finally, 50,717 SNPs were used for the following analysis.

Genetic diversity analysis

To assess genetic diversity of Yunan, we analyzed MAF, Ho, He and Ne of Yunan, Huainan and Duroc. The results were shown in Table 1. By comparison, the Ne of Huainan was lower than that of Yunan and Duroc. Indexes MAF, Ho and He of Yunan were higher than those of Huainan and Duroc indicating that Yunan was rich in genetic diversity.

ROH analysis results of Yunan, Huainan and Duroc were shown in Table 2. A total of 3,317 homozygous fragments were detected in the three populations. Duroc had the highest average number of ROH per animal (48.8 ± 6.55 with a range of 2–80 Mb) while Huainan had the lowest (7.21 ± 8.14 with a range of 1–105 Mb). Mean length of ROH (MGLROH) was maximum in Yunan (10.01 ± 4.28 Mb) and minimum in Duroc (8.91 ± 1.14 Mb). Duroc revealed the highest ROH based inbreeding ($F_{ROH}$ = 0.1925 ± 0.037). $F_{ROH}$ of Huainan ($F_{ROH} = 0.0324 ± 0.043$) and Yunan ($F_{ROH} = 0.043 ± 0.033$) were lower than Duroc. Yunan did not have severe inbreeding.

ROH fragment size of Yunan mainly concentrated in 5–15 Mb, accounting for 88.98% while that of Duroc and Huainan mainly concentrated in 1–10 Mb, accounting for 75.32% and 71.01%, respectively (Figure 3A). Chromosomes with the most ROH fragments in Yunan, Duroc and Huainan were SSC1 ($n = 170$), SSC1 ($n = 255$) and SSC7 ($n = 22$), and chromosomes with the least ROH fragments were SSC17 ($n = 15$), SSC17 ($n = 53$) and SSC18 ($n = 4$) (Figure 3B). The total average length of ROH of Yunan, Duroc and Huainan were 98.37 Mb, 436.28 Mb and 73.32 Mb (Figure 3C).

Population genetic structure analysis of yunan population

The first and second principal components of Yunan, Huainan and Duroc explained 29.68% and 9.99% of the total variance. Yunan, Huainan, and Duroc were clearly separated (Figure 4A). The first PC clearly separated Yunan, Huainan and duroc. Yunan was located between Huainan and Duroc. We used t-SNE to best classify the populations to perform dimensionality reduction clustering analysis on all the breeds. From Figure 4B, these results indicate that different subpopulations exist in Yunan pig population. From the result of admixture analysis in Figure 4C, When K = 2, Yunan, Huainan and Duroc could be clearly distinguished. Yunan contained 33.78% of Huainan and 66.22% of Duroc blood. When K = 3, a new bloodline in red was appeared in Yunan. When K = 4-5, different degrees of differentiation appeared in Yunan population. To investigate genetic structure of Yunan pigs, we constructed the NJ tree of the 193 individuals, which indicated the existing Yunan pigs could be divided into four main branches, each of which was divided into two or more subbranches (Figure 4D).

Selection signal screening among yunan, Huainan and Duroc

To screen the selection signals among Yunan, Huainan and Duroc, and analyse the possibly origin of the signals, we divided Yunan, Huainan and Duroc into three groups (Yunan-Duroc Vs. Huainan, Yunan-Huainan Vs. Duroc and Yunan-Duroc) and used two methods of $F_{ST}$ and XP-CLR. Top 1% regions of $F_{ST}$ and XP-CLR were considered as salient loci under selected. $F_{ST}$ screening results were shown in Figure 5A (Yunan-Duroc Vs. Huainan), 5C (Yunan-Huainan Vs. Duroc) and 5E (Yunan Vs. Huainan-Duroc). We detected several significant loci on SSC1-12, SSC14-17 and SSCX in Yunan-Duroc Vs. Yunan group, SSC1-16 and SSCX in Yunan-Huainan Vs. Duroc group, and SSC1-10, SSC12-18 and SSCX in Yunan-Duroc Vs. Yunan group. XP-CLR analysis results of the above three groups were shown in Figure 5B (Yunan-Duroc Vs. Huainan), 5D (Yunan-Huainan Vs. Duroc) and 5F (Yunan Vs. Huainan-Duroc).
obtained 71 significant loci on each of the 18 autosomes in each of the three groups.

Then we combined the outputs of FST and XP-CLR analysis. In Yunan-Duroc Vs. Huainan group, we found 37 significant regions that over threshold under selected were overlapped in FST and XP-CLR results. A total of 104 genes were annotated in these regions in the Ensembl (http://ensemble.org) database, 17 out of which were involved in fat production and metabolism, nine out of which were related to growth and development. These 26 genes under selected were thought to be of Duroc origin. Similarly, in Yunan-Huainan Vs. Duroc group, we got 21 overlapping regions containing 103 annotated genes, 14 genes out of which were related to fat regulation while eight genes were related to growth development. These 22 genes under selected were thought to be of Huainan origin. Also, 13 overlapping regions covering 116 genes were found in Yunan Vs. Huainan-Duroc. Only eight genes were involved in fat metabolism and adipose production, three genes were involved in skeletal muscle development. These outstanding 11 genes may have been selected by artificial selection in Yunan population.

To further look into former reported economic characters underlying these genomic regions, we extracted the common regions between these results and PigQTLdb (www.animalgenome.org/cgi-bin/QTLdb/index). QTLs that overlapped with potentially selected regions were QTLs of average daily gain (Fontanesi et al., 2014; Guo et al., 2017), backfat last rib (Gilbert et al., 2007) and days 110 kg (Wang et al., 2015), etc. as shown in Table 3.

**Genome-wide association study on backfat traits in yunan pigs**

To understand genetic background of backfat thickness variation in Yunan, Yunan pigs were divided into gilt (n = 48), 1st parity (n = 48) and 2nd parity (n = 24) according to the age. Descriptive statistics of the six backfat traits (P2BF, SBF, SSRBF, LRBF, LBF and ABF) of this three groups were shown in Table 4. From Table 4, Yunan was much fatter from the ideal body shape. For example, the average P2BF thickness of gilt, 1st parity and 2nd parity in Yunan was 36.90, 35.28 and 35.93 mm. This was much thicker than the commercial sows (usually 16–22 mm). Moreover, the coefficient variation (CV) of the backfat traits was larger, ranged from 12.80% to 34.96%.

With the goal of pinpointing genomic region associated with backfat phenotypes of Yunan, we performed genome-wide association studies of six backfat traits using a linear mixed model. We identified 34 suggestive significant SNPs associated with six backfat traits in total. Thereinto, nine significant SNPs on SSC1-2, SSC4, SSC11-12, SSC14 and SSC16 were detected in 1st parity pigs. 14 significant SNPs on a 1.4 Mb segment of SSC4

### Table 2 Genomic distributions and descriptive statistics of ROH in Yunan, Huainan and Duroc.

| Breeds  | N<sub>ROH</sub> | Range ROH (Mb) | NM<sub>ROH</sub> | MGL<sub>ROH</sub> (Mb) | F<sub>ROH</sub> |
|---------|----------------|----------------|------------------|-------------------------|--------------|
| Yunan   | 1,126          | 3–124          | 9.38 ± 4.55      | 10.01 ± 4.28            | 0.043 ± 0.033|
| Huainan | 238            | 1–105          | 7.21 ± 8.14      | 9.49 ± 6.21             | 0.034 ± 0.043|
| Duroc   | 1953           | 2–80           | 48.82 ± 6.55     | 8.91 ± 1.14             | 0.1925 ± 0.037|

N<sub>ROH</sub>: Total number of ROH per breed; Range ROH: length range of ROH; NM<sub>ROH</sub>: Mean number of ROH in a breed; MGL<sub>ROH</sub>: Breed wise mean genome length covered by ROH in Mb; F<sub>ROH</sub>: Inbreeding coefficient based on ROH.
(20.03–21.43 Mb) were detected in 2nd parity pigs while 11 significant SNPs on SSC1-2, SSC8, SSC13 and SSCX were obtained in gilts. 202 genes were annotated in the 34 genomic regions in the Ensembl database, five, six and three genes of which were involved in fat metabolism or growth in gilt, 1st and 2nd parity pigs (Figure 6).

The genomic regions where these significant loci were located partially overlapped. The most significant associated SNP was the same for P2BF and LRBF on SSC8 (10.43 Mb) in gilts, for P2BF and ABF on SSC16 (21.97 Mb) in 1st parity, for P2BF, LRBF and ABF on SSC4 (20.03–21.43 Mb) and for SSRBF and ABF on SSC4 (106.29 Mb) in 2nd parity pigs. Then we combined these 34 regions with PigQTLdb database (www.animalgenome.org/cgi-bin/QTLdb/index) and found that significant loci on SSC4 (20.03–21.43 Mb) were overlapped with a reported average backfat thickness QTL (20,366,121–21,945,045 bp) in large white pigs (Bink et al., 2000) (Table 5).

In view of the existence of selective signals related to fat deposition and growth, and to investigate whether the genomic regions associated with backfat traits mapped by GWAS might have been selected in the population, we analyzed the regions common to the results of FST, XP-CLR and GWAS. The selected region on SSC4 (21.24–21.83 Mb) in Yunan-Huainan Vs. Duroc group which was thought to be of Huainan origin and the P2BF, LRBF and ABF associated region on SSC4 (20.03–21.43 Mb) in 2nd parity stood out. These two regions were overlapped, and ENPP2 and EXT1 genes (SSC4:19.02–21.06 Mb) in this region were functionally related to fatty acid production (Crespo-Piazuelo et al., 2020). Therefore, the genome region of 20.03–21.83 Mb on SSC4 was considered as an important candidate region for backfat thickness in Yunan pig population.

Finally, we used Haplovie v4.2 (Barrett et al., 2005) software for linkage disequilibrium analysis and constructed haplotype modules for the SSC4 (20.03–21.83 Mb) under the default
parameters. We found four linkage blocks ($r^2 \geq 0.8$), block1 (21, 111, 825–21,196,785 bp), block2 (21, 208, 018–21,218,173 bp), block3 (21, 402, 451–21,421,498 bp) and block4 (21, 516, 432–21,709,640 bp) within the target area (Figure 7). Only one gene SLC30A8 (21, 517, 486–21,555,757 bp) was found in block4. SLC30A8 gene encodes zinc transporter, which transfers zinc from the cytoplasm of pancreatic $\beta$-cells to intracellular vesicles and functions in insulin secretion. Impaired insulin secretion and insulin resistance are the pathogenesis of type 2 diabetes mellitus (Witka et al., 2019).

**Discussion**

The genetic diversity of a population was a key factor in ensuring the survival and evolution of a species. Commercial pigs have been subjected to high-intensity artificial selection for a time, resulting in a decrease in genetic diversity, while local varieties were mainly selected by natural conditions such as environmental factors and geographical factors, thus maintaining a high level of genetic diversity. China’s native pig breeds has a better genetically diverse (SanCristobal et al., 2006; Ai et al., 2013). The genetic diversity of Chinese hybrid pigs was higher than that of Chinese local pig breeds (Yang et al., 2003; Huang et al., 2020) and commercial pig breeds (Wang et al., 2021). There were many statistical methods to evaluate population genetic diversity, such as allele frequency and population heterozygosity. The higher the heterozygosity, the higher the genetic diversity. In this study, the MAF, Ho and He of the hybrid pig breed Yunan were higher than those of its founders Duroc and Huainan, indicating Yunan had a richer genetic diversity. Effective population size ($N_e$) is also an important indicator of diversity and species conservation. If the $N_e$ was below 65, the breed might have a population crisis.
(Simon, 1999). The Ne of Yunan was 67. This suggested that although having experienced the threat of African swine fever, Yunan did not have a group crisis now.

ROH segments contains information about population inbreeding, and its length and frequency can reflect the population history. Compared with pedigree inbreeding number, the calculation of genomic inbreeding number based on ROH was more accurate and can better reflect the real inbreeding number of an individual (Purfield et al., 2012; Zanella et al., 2016; Deniskova et al., 2019; Bhati et al., 2020). A longer ROH usually indicates a closer genetic relationship while a shorter ROH indicates an ancient inbreeding. A larger number of ROHs and fragments means a higher probability of inbreeding (Kirin et al., 2010). Here, the average total length of ROH of Yunan was between Huainan and Duroc. Duroc had the highest average number of ROH per animal while Huainan had the lowest. Mean length of ROH was maximum in Yunan and minimum in Duroc. Duroc had the highest ROH based on inbreeding. 

\[ \text{FRoh} \] of Huainan and Yunan were lower than Duroc. Compared with some other local breed pigs (Mashen and Chun'an) in China (Cai et al., 2021; Dai et al., 2021) and European commercial breeds (Zhan et al., 2020), the average inbreeding coefficient of Yunan was relatively lower, and Yunan black pigs did not show obvious inbreeding.

From few large branches in NJ trees, population stratification in PCA, and having no one main lineage when K = 3 to 5 of

### Table 3: Candidate genes and previous reported QTLs of common selected regions from FST and XP-CLR analysis.

| Group                        | SSC | Position (bp) | Gene             | QTL                                |
|------------------------------|-----|---------------|------------------|-----------------------------------|
| **Yunan-Duroc Vs. Huainan**  | 1   | 44,480,000–4,493,794 | GPRC6A          | Backfat Growth                    |
|                              | 1   | 90,160,000–90,495,794 | SMAD2, ZBTB7C   | Body weight (birth)               |
|                              | 3   | 2,770,524–3,132,524 | SLC26A2         | Body weight (end of test)         |
|                              | 3   | 36,784,268–37,498,268 | SENP6           |                                    |
|                              | 3   | 71,302,268–71,340,268 | ZNF638          |                                    |
|                              | 6   | 54,733,496–55,767,496 | CPT1            |                                    |
|                              | 7   | 14,960,000–15,610,808 | ID4             |                                    |
|                              | 8   | 978,400–106,400    | NSD2            | Backfat at last rib               |
|                              | 12  | 976,400,000–773,400 | TAC1            | Leaf fat weight                   |
|                              | 15  | 109,103,272–109,460,000 | TLX5         |                                    |
|                              | 15  | 3,800,000–4,580,000 | CRTAC1          |                                    |
|                              | 17  | 17,009,180–17,211,180 | MBD5, ACVR2A   |                                    |
| **Yunan-Huainan Vs. Duroc**  | 2   | 56,423,768–56,919,768 | NLRP3          |                                    |
|                              | 2   | 5,807,168–60,123,768 | GATAD2A, SUGP1, NCAN, INSL3, JAK3, CRT1 | Body depth |
|                              | 2   | 68,667,768–68,780,000 | PIN1, OLFM2    | Body width                        |
|                              | 2   | 69,440,001–69,625,768 | DNMT2, CARM1   | Backfat at first rib              |
|                              | 4   | 21,240,001–21,830,268 | SLC30A8        | Average backfat thickness         |
|                              | 4   | 149,794,156–150,244,156 | ANGPTL3      | Body weight                       |
|                              | 4   | 33,334,864–33,480,864 | DOCK3          | Days to 100 kg                    |
| **Yunan Vs. Huainan-Duroc**  | 2   | 44,416,175–44,614,028 | PDE3B          |                                    |
|                              | 6   | 4,808,255–5,832,188 | CDH13          | Body length                       |
|                              | 6   | 29,366,624–29,413,703 | AMFR           |                                    |
|                              | 7   | 28,300,808–29,064,808 | BEND6          | Average daily gain                |
|                              | 12  | 22,829,286–23,387,286 | MED1, PIP4K2B  |                                    |
|                              | 12  | 75,501,272–77,161,272 | MCU, PLAZ2G12B, ANX4A7 | MSS51, MYOZ1 |
|                              | 14  | 112,560,001–113,175,272 | LDB1          |                                    |

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ADMIXTURE implied much more breeding work should be done in Yunan. In addition, different subsets of Yunan may exist according to PCA, which was also indicated by t-SNE analysis. So, even if Yunan pigs had no risk of inbreeding and recession according to the population diversity and inbreeding analysis, but this diversity may partly be due to stratification within groups. Therefore, in the future breeding work of Yunan pigs, we should carry out mating among lineages according to the NJ molecular pedigree to avoid multiple invalid matings, otherwise the consistency of the population would not be improved.

Selection signals can reflect loci and genes that have been strongly selected in a population during long-term domestication. The selective signals we detected in Yunan black pigs were not only related to fat deposition, but also to growth. Both directions had both Duroc origin (Yunan-Duroc VS. Huainan) and Huainan origin (Yunan-Huainan VS. Duroc). For example, GPRC6A (Mukai et al., 2021) and MBDS (Du et al., 2012) for fatness and growth of Duroc while ANGPTL3 (Jiang et al., 2018) and DOCK3 (Reid et al., 2020) of Huainan. These results indicated that both Duroc and Huainan pigs had genetic variants affecting growth and fat deposition. In addition, some genes, such as CDH13 (Göddeke et al., 2018) and RAB23 (Hasan et al., 2020), were found to be involved in fat deposition and growth in the Yunan VS. Huainan-Duroc group. These loci might be derived from the founder effect of the initial breeding group of Yunan or might be the result of the 14 years of breeding process of Yunan.

Genomic loci associated with backfat thickness partially overlapped between different parities and different traits. This suggested the influence of some genes on backfat was a long-term process, there’s no time and space specificity. When it came to the nonoverlapped genes or regions, we could not come to a conclusion of time or space specificity. Because although the population used in this study covers the core population of Yunan black pigs, the number of populations was too small. This reminded us that under the epidemic environment, it was necessary to adopt multi-site conservation for species, especially for local genetic resources.

Three genes related to fat deposition, ENPP2, EXT1 and SLC30A8, were found in the upper and lower 1 Mb of SSC4 (20.03–21.43 Mb) (Du et al., 2009; Hutley et al., 2011; Nishimura et al., 2014). Among the four linkage disequilibrium blocks in the 1.40 Mb interval of SSC4, only one gene, SLC30A8, was present in the fourth block (193.21 Kb). This gene is the star gene of type 2 diabetes. In 2009, there was a study that attempted to find the causal mutation for human type 2 diabetes by analyzing the correlation between the mutation of this gene with fat deposition in pigs (Du et al., 2009) but failed. Further analysis of this gene in Yunan might be needed.

In addition, QTL has been widely used in the study of important economic traits in pigs. The hunt for QTL in pigs has been ongoing for nearly 2 decades, beginning with the first publication of a QTL for fatness on pig chromosome 4 in 1994 (Andersson et al., 1994). In crossbreeding analysis, SNP markers

| Table 4 Descriptive statistical of backfat traits in Yunan pigs. |
|-------------------------|------------------|------------------|------------------|------------------|------------------|
| Number | Trait | Mean (mm) | SD | Min | Max | CV % |
|-------|-------|-----------|----|-----|-----|-----|
| Gilt  | P2BF  | 36.90     | 9.21 | 20.70 | 53.90 | 34.96 |
|       | SBF   | 15.70     | 3.24 | 9.40  | 26.90 | 20.64 |
|       | SSRBF | 40.00     | 9.29 | 23.20 | 58.90 | 23.23 |
|       | LRBF  | 34.64     | 8.37 | 20.10 | 53.90 | 24.16 |
|       | LBF   | 20.58     | 3.24 | 15.00 | 27.60 | 15.74 |
|       | ABF   | 27.70     | 5.04 | 19.58 | 37.93 | 18.19 |
| Paify 1 | P2BF | 35.28     | 7.37 | 20.70 | 50.80 | 20.89 |
|       | SBF   | 18.40     | 2.73 | 13.80 | 26.30 | 14.84 |
|       | SSRBF | 35.77     | 8.74 | 20.70 | 54.50 | 24.43 |
|       | LRBF  | 31.96     | 7.50 | 18.20 | 52.60 | 23.47 |
|       | LBF   | 21.78     | 3.99 | 13.80 | 36.40 | 18.32 |
|       | ABF   | 26.98     | 4.79 | 18.00 | 39.03 | 17.75 |
| Paify 2 | P2BF | 35.93     | 6.74 | 22.60 | 52.00 | 18.76 |
|       | SBF   | 18.36     | 2.35 | 13.80 | 24.40 | 12.80 |
|       | SSRBF | 39.67     | 9.94 | 21.30 | 60.30 | 25.06 |
|       | LRBF  | 32.87     | 6.54 | 21.30 | 50.10 | 19.90 |
|       | LBF   | 21.76     | 2.88 | 17.50 | 30.10 | 13.24 |
|       | ABF   | 28.17     | 5.03 | 19.43 | 40.43 | 17.86 |

P2BF, P2 point backfat, SBF, Shoulder backfat; SSRBF, Six and seven rib backfat; LRBF, Last rib backfat; LBF, Lumbosacral backfat; ABF, Average backfat; SD, Standard Deviation; CV, Coefficient of variance.
FIGURE 6
Manhattan plot of genome-wide association analysis of P2-backfat, shoulder backfat, sixth and seventh ribs backfat, last rib backfat, lumbar joint backfat and average backfat at different parity. The annotated genes represent the relevant candidate genes.
and QTL linkage analysis can be used to identify genomic regions with complementary effects from potential resource groups, so as to increase the degree of genetic complementarity between varieties or lines in a planned way, and thus improve the economic traits of hybrids and the genetic diversity of varieties.

In conclusion, we analyzed genetic structure and mapped genomic regions affecting backfat thickness of Yunan black pigs.

| Parity  | Phenotype | SSC (Mb) | Position of significant SNPs | Most significant SNP Position (bp) | Minor allele frequency | Candidate gene name |
|---------|-----------|---------|-----------------------------|-----------------------------------|------------------------|---------------------|
| Gilt    | P2BF      | 8       | 10.43                       | 10,430,709                        | 6.58E-06               | 0.2396              |
|         | SBF       | 1       | 233.79–235.18               | 234,643,280                       | 8.30E-06               | 0.125               |
|         | 2         | 125.35  | 1                           | 125,353,968                       | 7.77E-06               | 0.375               |
|         | 8         | 83.12   | 1                           | 83,125,662                        | 1.37E-05               | 0.08333             |
|         | 13        | 136.08  | 1                           | 136,088,769                       | 1.40E-05               | 0.2812              |
|         | 23        | 23.89   | 1                           | 23,893,572                        | 1.05E-05               | 0.05208             |
| LRBF    | 8         | 10.43   | 1                           | 10,430,709                        | 5.69E-06               | 0.2396              |
| LBF     | 1         | 63.41   | 1                           | 63,418,505                        | 1.21E-05               | 0.09375             |
| Parity 1| P2BF      | 16      | 21.97                       | 21,976,238                        | 1.61E-05               | 0.4271              |
|         | SBF       | 4       | 83.26                       | 83,269,208                        | 1.21E-05               | 0.4375              |
|         | SSRBF     | 11      | 26.09                       | 26,093,623                        | 1.84E-05               | 0.1354              |
|         | 12        | 17.86   | 1                           | 17,867,041                        | 1.38E-05               | 0.1562              |
|         | 14        | 85.98   | 1                           | 85,986,081                        | 9.06E-06               | 0.0625              |
| LRBF    | 2         | 3.52    | 1                           | 3,528,858                         | 1.27E-05               | 0.1458              |
|         | 2         | 102.23  | 1                           | 102,231,770                       | 1.89E-05               | 0.1562              |
| LBF     | 1         | 213.80  | 1                           | 213,808,326                       | 1.42E-05               | 0.01042             |
| ABF     | 16        | 21.97   | 1                           | 21,976,238                        | 4.61E-06               | 0.4271              |
| Parity 2| P2BF      | 4       | 20.03–21.43                 | 20,038,718                        | 1.44E-05               | 0.4792              |
|         | SSRBF     | 4       | 106.29                      | 106,293,182                       | 4.47E-06               | 0.1250              |
| LRBF    | 4         | 20.03–21.43 | 4                     | 20,038,718                        | 8.73E-06               | 0.4792              |
| ABF     | 4         | 106.29  | 1                           | 106,293,182                       | 2.44E-06               | 0.1250              |
|         | 4         | 20.03–21.43 | 4                     | 20,038,718                        | 1.91E-05               | 0.4792              |

Significance of SNP.

FIGURE 7
Haplview of linkage disequilibrium of SNPs on chromosome four (20.03–21.83 Mb) in Yunan pigs.
Although there was no risk of inbreeding depression, the stratification phenomenon existed in Yunan population. The increasing backfat thickness and decreasing consistency had a particular genetic basis. The 1.40 Mb interval of SSC4 (20.03–21.43 Mb) was a strong candidate region associated with backfat thickness. ENPP2, EXT1, and SLC30A8, particular SLC30A8, were strong candidate genes. Our findings are helpful for the subsequent breeding and conservation, as well as genomic improvement of backfat thickness of Yunan black pigs and suggested the importance of multi-site breeding conservation method.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: https://bigd.big.ac.cn, GVM000386.

Author contributions

RQ contributed to the conception of the study and directed the wrote of the manuscript. PH performed date analysis and wrote the manuscript. MZ and BZ contributed to sample collection and data collection. XnL, XH, KW, XuL and FY contributed to revise the manuscript. All authors have read and agreed to the published version of the manuscript.

References

Ai, H., Huang, L., and Ren, J. (2013). Genetic diversity, linkage disequilibrium and selection signatures in Chinese and Western pigs revealed by genome-wide SNP markers. PLoS One 8 (2), e56001. doi:10.1371/journal.pone.0056001

Alexander, D. H. Shringarpure, S. S. Novembre, J., and Lange, K. L. (2015). Admixture 1.3 software manual.

Andersson, L., Haley, C. S., Ellegren, H., Knott, S. A., JohanssonM., Andersson, K., et al. (1994). Genetic mapping of quantitative trait loci for growth and fatness in pigs. Sci. (New York, N.Y.) 263 (5154), 1771–1774. doi:10.1126/science.8134840

Barbato, M., Orozco-terWengel, P., Tapio, M., and Bruford, M. W. (2015). SNeP: A tool to estimate trends in recent effective population size trajectories using genome-wide SNP data. Front. Genet. 6, 109. doi:10.3389/fgene.2015.00109

Barrett, J. C., Fry, B., Maller, J., and Daly, M. J. (2005). Haploview: Analysis and visualization of LD and haplotype maps. Bioinformatics 21 (2), 263–265. doi:10.1093/bioinformatics/bth457

Bhati, M., Kader, N. K., Crysantho, D., and Pausch, H. (2020). Assessing genomic diversity and signatures of selection in Original Braunvieh cattle using whole-genome sequencing data. BMC Genomics 21 (1), 27. doi:10.1186/s12864-020-6446-y

Bink, M. C., Te Pas, M. F., Harders, F. L., and Janss, L. L. (2000). A transmission/disequilibrium test approach to screen for quantitative trait loci in two selected lines of large white pigs. Genet. Res. 75 (1), 115–121. doi:10.1017/S0016672399004061

Cai, C., Zhang, X., Zhang, W., Yang, Y., Gao, P., Guo, X., et al. (2021). Evaluation of genetic structure in mashen pigs conserved population based on SNP chip. Chin. J. Animal Veterinary Sci. 52 (04), 920–931.

Chen, H., Patterson, N., and Reich, D. (2010). Population differentiation as a test for selective sweeps. Genome Res. 20 (3), 393–402. doi:10.1101/gr.100545.109

Crespo-Pazuelo, D., Grado-Mesas, L., Revilla, M., Castello, A., Noguera, I. L., Fernández, A. I., et al. (2020). Identification of strong candidate genes for backfat and intramuscular fatty acid composition in three crosses based on the Iberian pig. Sci. Rep. 10 (1), 13962. doi:10.1038/s41598-020-70894-2

Dai, L., Zhu, X., Chen, X., Yang, N., Huang, S., and Xu, R. (2021). Analysis of genetic diversity and genetic structure in chun an spotted pigs conserved population.

Dai, L., Zhu, X., Chen, X., Yang, N., Huang, S., and Xu, R. (2021). Analysis of genetic diversity and genetic structure in chun an spotted pigs conserved population.

Deniskova, T., Dotsey, A., Lushihina, E., Shakhin, A., Xunz, E., Medugorac, L., et al. (2019). Population structure and genetic diversity of sheep breeds in the Kyrgyzstan. Front. Genet. 10, 1311. doi:10.3389/fgene.2019.01311

Du, Y., Liu, B., Guo, F., Xu, G., Ding, Y., Liu, Y., et al. (2012). The essential role of Mbd5 in the regulation of somatic growth and glucose homeostasis in mice. PLoS One 7 (10), e47358. doi:10.1371/journal.pone.0047358

Du, Z. Q., Fan, B., Zhao, X., Amouko, R., and Rothschild, M. F. (2009). Association analyses between type 2 diabetes genes and obesity traits in pigs. Obes. (Silver Spring, Md.) 17 (2), 323–329. doi:10.1086/02.2008.557

Fontanesi, L., Schiavo, G., Galimberti, G., Calo, D. G., and Russo, V. (2014). A genomewide association study for average daily gain in Italian Large White pigs. J. Anim. Sci. 92 (4), 1385–1394. doi:10.2527/jas.2013-7059

Gilbert, H., Le Roy, P., Milan, D., and Bidanel, J. P. (2007). Linked and pleiotropic QTLs influencing carcass composition traits detected on porcine chromosome 7. Genet. Res. 89 (2), 65–72. doi:10.1017/S0016672307008701

Goldeke, S., Knebel, B., Fahlbusch, P., Horbelt, T., Poschmann, G., van de Velde, F., et al. (2018). CDH13 abundance interferes with adipocyte differentiation and is a novel biomarker for adipose tissue health. Int. J. Obes. 42 (5), 1039–1056. doi:10.1038/s41366-018-0022-4

Guo, Y., Huang, Y., Hou, L., Ma, J., Chen, C., Ai, H., et al. (2017). Genome-wide detection of genetic markers associated with growth and fatness in four pig populations using four approaches. Genet. Sel. Evol. 49 (1), 21. doi:10.1186/s12711-017-0299-4

Hasan, M. R., Takatalo, M., Ma, H., Rice, R., Mustonen, T., and Rice, D. P. (2020). RAB23 coordinates early oogenesis by repressing FGFR10-ePcK1 and GLI1. eLife 9, e55829. doi:10.7554/eLife.55829

Huang, M., Yang, B., Chen, H., Zhang, H., Wu, Z., Ai, H., et al. (2020). The fine-scale genetic structure and selection signals of Chinese indigenous pigs. Evol. Appl. 13 (2), 485–478. doi:10.1111/eva.12887

Conflict of interest

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A putative role for endogenous FGF-2 in FGF-1 mediated differentiation of human preadipocytes. Mol. Cell. Endocrinol. 339 (1-2), 165–171. doi:10.1016/j.mce.2011.04.012

Jiang, Y., Tang, S., Wang, C., Wang, Y., Qin, Y., Wang, Y., et al. (2018). A genome-wide association study of growth and fattness traits in two pig populations with different genetic backgrounds. J. Anim. Sci. 96 (3), 806–816. doi:10.1093/jas/skx038

Kirin, M., McQuillan, R., Franklin, C. S., Campbell, H., McKeigue, P. M., and Wilson, J. F. (2010). Genomic runs of homozygosity record population history and consanguinity. PLoS One 5 (11), e13996. doi:10.1371/journal.pone.013996

Krejța, J. H. (2015). Rtsne: T-distributed Stochastic neighbor Embed. using a Barnes-Hut Implement.

Li, X., Qiao, R., Li, X., Guo, J., Han, X., Zhang, H., et al. (2016). The genetic characteristics of meat quality and nutritional components of yunan black pig and its hybrid pigs. J. Domest. Animal Sci. 37 (03), 20–26.

Mukai, S., Mizokami, A., Otani, T., Sano, T., Matsuda, M., Chishaki, S., et al. (2021). Adipocyte-specific GPRC6A ablation promotes diet-induced obesity by inhibiting lipolysis. J. Biol. Chem. 296, 100274. doi:10.1016/j.jbc.2021.100274

Nishimura, S., Nagasaki, M., Okudaira, S., Aoki, J., Ohmori, T., Ohkawa, R., et al. (2014). ENPP2 contributes to adipose tissue expansion and insulin resistance in diet-induced obesity. Diabetes 63 (12), 4154–4164. doi:10.2337/db13-1694

Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A., Bender, D., et al. (2007). Plink: A tool set for whole-genome association and population-based linkage analyses. Am. J. Hum. Genet. 81 (3), 556–579. doi:10.1086/519795

Purfield, D. C., Berry, D. P., McParland, S., and Bradley, D. G. (2012). Runs of homozygosity and population history in cattle. BMC Genet. 13, 70. doi:10.1186/1471-2156-13-70

Reid, A. L., Wang, Y., Samani, A., Hightower, R. M., Lopez, M. A., Gilbert, S. R., et al. (2020). DOCK3 is a dosage-sensitive regulator of skeletal muscle and characteristics of meat quality and nutritional components of yunan black pig and its hybrid pigs. J. Domest. Animal Sci. 37 (03), 20–26.

Slåås, L., Rodríguez, M. C., Fernández, A., Barragán, C., Benítez, R., Óvilo, C., et al. (2013). Measuring inbreeding and inbreeding depression on pig growth from pedigree or SNP-derived metrics. Anim. Genet. 44 (4), 381–387. doi:10.1111/j.1365-2052.2013.01885.x

Simon, D. L. (1999). European approaches to conservation of farm animal genetic resources. Anim. Genet. Resour. Inf. 25, 77–97. doi:10.1017/s1014243900005794

Sun, T., Shen, J., Achilli, A., Chen, N., Chen, Q., Dang, R., et al. (2020). Genomic analyses reveal distinct genetic architectures and selective pressures in buffaloes. GigaScience 9 (2), giz166. doi:10.1093/gigascience/giz166

Team, R. C. (2014). R: A language and environment for statistical computing. MSOR Connect. 1.