Dietary Niacin Intake Predicts the Decrease of Liver Fat Content During a Lifestyle Intervention

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Niacin inhibits fatty acid flux from adipose tissue to liver, reduces hepatic triglyceride synthesis and increases hepatic lipid oxidation. Thus, niacin may have a role in the regulation of liver fat content in humans. We tested if dietary intake of niacin predicts change of liver fat content during a lifestyle intervention. To this end, we estimated the composition of diet from diaries of 202 healthy subjects at risk of type 2 diabetes undergoing lifestyle intervention comprising physical activity and diet counselling. Total-, subcutaneous- and visceral adipose tissue mass were measured by magnetic resonance (MR) tomography and liver fat content by 1H-MR spectroscopy at baseline and after 9 months of follow-up. Among fat compartments, liver fat content showed the largest decrease (−32%, p < 0.0001). High baseline niacin intake predicted a larger decrease of liver fat (p = 0.004). Subjects in the highest quartile of niacin intake at baseline also had the largest decrease of liver fat (1st: −10%; 2nd: −27%; 3rd: −35%; 4th: −37%). Among 58 subjects with nonalcoholic fatty liver disease (NAFLD) at baseline, NAFLD resolved in 23 subjects during the lifestyle intervention. For one standard deviation increase in niacin intake, the odds ratio for resolution of NAFLD was 1.77 (95% CI, 1.00–3.43). High dietary niacin intake may have a favorable effect on the reduction of liver fat during lifestyle intervention.

Nonalcoholic fatty liver disease (NAFLD) is the most common chronic liver disease in developed countries, affecting more than one-quarter of adults, and its prevalence is continuing to increase in parallel with the epidemic of obesity1. NAFLD is not only a potential precursor for the development of advanced liver diseases but also represents a strong risk factor for the development of insulin resistance, type 2 diabetes and cardiovascular events2–6. Thus, the prevention and treatment of fatty liver has high priority, not only on a clinical, but also on a population level.

While pharmacological treatment of NAFLD is under intensive investigation and some studies showed promising results7, lifestyle intervention with increase in physical activity and modification of the diet so far represents the most effective and safe approach to prevent and treat NAFLD8,9. A plausible mechanism responsible for the beneficial effect of lifestyle intervention on the change in liver fat content is the loss of fat mass, particularly visceral fat, resulting in a reduced portal free fatty acid supply to the liver and favorable changes in secretion of adipocytokines. However, because the decrease in total body- or visceral fat mass is not necessarily accompanied by a decrease in liver fat content, other mechanisms may also be involved in this process9. In support, we found that a higher cardiorespiratory fitness at baseline predicted a larger decrease in liver fat content during a lifestyle intervention, independently of total adiposity, body fat distribution and exercise intensity8.

Because considerable weight loss and maintenance of reduced body weight in the long term is difficult to achieve, and a high cardiorespiratory fitness is largely genetically determined10, it seems highly important to investigate the potential impact of nutrition factors on liver fat content.

In this respect, dietary niacin intake (nicotinic acid, vitamin B3, or its corresponding amid nicotinamide) could be a potential modulator of hepatic steatosis. Niacin inhibits lipolysis in adipose tissue, decreases the...
activity of diacylglycerol acyltransferase-2 (DGAT2) in hepatocytes and possibly increases hepatic lipid oxidation. Therefore, we investigated whether dietary niacin intake predicted the change in liver fat content during a lifestyle intervention in subjects who are at increased risk for type 2 diabetes.

Methods

Subjects. The data analyzed in this report derive from the Tübingen Lifestyle Intervention Program (TULIP). This is a non-randomized lifestyle intervention study that was designed in 2002 and which has been performed between 2003 and 2009. The study has been registered with the German Research Foundation as 'KFO114' (http://gepris.dfg.de/gepris/projekt/5396893?language=en). The study recruited individuals from the Tübingen region in Southern Germany who fulfilled at least one of the following criteria: a family history of type 2 diabetes, a BMI $\geq 27$ kg/m$^2$, a previous diagnosis of impaired glucose tolerance and/or of gestational diabetes. They were considered healthy according to a physical examination and routine laboratory tests. The participants had no history of liver disease and did not consume more than 2 alcoholic drinks per day. A total of 202 subjects met the aforementioned requirements and complete data about the dietary intake and measurements of body fat distribution and liver fat content using magnetic resonance techniques at baseline and at follow-up were available. Informed written consent was obtained from all participants. The medical ethics committee of the University of Tübingen had approved the protocol which was conform to the Declaration of Helsinki.

Lifestyle intervention. After the baseline measurements, individuals underwent 30 minutes one-to-one dietary counselling with trained dietitians. The whole lifestyle intervention had a duration of two years divided into two phases. The first phase lasted for 6 months and comprised monthly counselling visits. The second phase lasted for 18 months with a total of 6 counselling visits (one every 3 months in average). During the duration of the next 9 months, every participant had up to ten individual counselling sessions. At each visit, the participants presented a 3-day food diary and discussed the results with the dieticians. Applying established lifestyle intervention strategies, counselling was aimed to reduce body weight by $\geq 5\%$, to reduce the intake of calories from fat to $< 30\%$ and particularly the intake of saturated fat to $\leq 10\%$ of energy consumed and to increase the intake of fibres to at least 15 g/1000 kcal. Participants were advised to avoid consuming alcohol within the frame of recommendations aiming at reducing body weight and to increase fruit and vegetables consumption within the frame of recommendations aiming at increasing fiber intake. No dietary advice about salt and coffee intake was not given. Other specific dietary recommendations were not provided. Diet composition was evaluated with a validated computer program using 2 representative days of a 3-day diary (DGE-PC 3.0, Deutsche Gesellschaft für Ernährung, Bonn, Germany). Niacin intake was assessed by the computer program based upon the niacin content of each food component, and provided in mg/d. Mean daily niacin intake from diet protocols before and during the lifestyle intervention were used as variables in our models. Individuals were asked to perform at least 3 hours of moderate sports per week. Aerobic endurance exercise (e.g. walking, swimming) with an only moderate increase in the heart rate was encouraged. All subjects completed a standardised self-administered validated questionnaire to measure physical activity and a habitual physical activity score was calculated. Participants were seen by the staff on a regular basis to ensure that these recommendations were accomplished.

Total body fat mass, body fat distribution and liver fat content. Measurements of total body-, subcutaneous- and visceral fat mass were performed by an axial T1-weighted fast spin echo technique with a 1.5 T whole-body imager (Magnetom Sonata, Siemens Medical Solutions). Liver fat content was measured by localized proton magnetic resonance (1HMR) spectroscopy. NAFLD was defined as liver fat content $> 5.56\%$.

Peak aerobic capacity ($V_{O_2peak}$). The individuals underwent a continuous, incremental exercise test to volitional exhaustion using a cycle ergometer. The cycle ergometer test was performed on an electromagnetically braked cycle ergometer (Ergometrics 800 S; Ergoline, Bitz, Germany). Oxygen consumption was measured using a spiroergometer (MedGraphics System Breese Ex 3.02 A; MedGraphics). $V_{O_2peak}$ is expressed as VO$_2$ (ml/min) per kg lean body mass (ml-min$^{-1}$-kg$^{-1}$).

Oral glucose tolerance test and laboratory measurements. All individuals underwent a 75 g oral glucose tolerance test (OGTT) and venous blood samples were obtained at 0, 30, 60, 90 and 120 minutes for determination of plasma glucose and insulin. Blood glucose was determined using a bedside glucose analyzer (glucose-oxidase method; YSI, Yellow Springs Instruments, Yellow Springs, CO). For measurements of insulin, blood was placed on ice after drawing, immediately transferred to the lab and subsequently analyzed. Plasma triglycerides, total cholesterol, and HDL and LDL cholesterol were measured using the ADVIA 1650 clinical chemical analyzer, and insulin was analyzed using the ADVIA Centaur immunoassay system according to the instructions of the manufacturers. Both analyzers were from Siemens Medical Solutions (Fernwald, Germany). We measured the serum free fatty acid (FFA) concentration with an enzymatic method (WAKO Chemicals, Neuss, Germany). Insulin sensitivity was calculated from glucose and insulin values during the OGTT, as proposed by Matsuda and deFronzo. The adipose tissue insulin resistance index was determined as previously described. The single nucleotide polymorphism (SNP) rs1944438 C $\rightarrow$ T in DGAT2 was genotyped by direct sequencing and using the TaqMan® assay (Applied Biosystems) as previously described.

Statistical analyses. Unless otherwise stated, data are given as mean $\pm$ SE (standard error). Differences between baseline and follow-up data were tested using the Wilcoxon signed-rank test. Change of liver fat content was assessed as fold change (liver fat$_{post-intervention}$/liver fat$_{pre-intervention}$). Relationships were tested with forward stepwise regression (using the Bayesian Information Criterion to decide on the inclusion of a variable) and ordinary multivariable regression models. For the inclusion in the regression models, parameters with skewed distributions were logarithmically transformed to approximate a normal distribution. Effect sizes were provided as
The largest effect of the lifestyle intervention was found for liver fat content (32%, Table 1). From protein, increased. The intake of niacin also tended to increase, however, the change of niacin intake was from fat decreased. On the other hand, the intake of fibers and of calories from carbohydrates, and particularly reference intakes (16 g/d and 14 g/d, respectively22). Niacin intake correlated positively with total energy intake, Nevertheless, a large intra-individual variability was observed for this change.

β larger decrease of liver fat content (β of niacin intake with change of liver fat content remained statistically significant (β = −0.21, p = 0.008) also after additional adjustment for sex, age, total body fat mass at baseline and at follow-up and total daily energy intake at baseline (Table 2, model 1). Niacin intake did not significantly associate with liver fat reduction in the above model (p = 0.26).

Next, we determined whether the change of liver fat content depended on the intake of other nutrients or the amount of physical activity. For this, we ran a forward stepwise regression analysis investigating the impact of total energy intake, intake of calories from carbohydrates, fat and protein, intake of fibers, niacin and physical activity at baseline, to predict the change of liver fat content. In this model, the intake of niacin strongly predicted the change of liver fat content and only the intake of calories from carbohydrates, was entered as another considerably weaker predictor into the stepwise regression model (Table 3). In addition, in a multivariable model, the

| Parameters                              | Before intervention | After intervention | Change (%) | p    |
|-----------------------------------------|---------------------|--------------------|------------|------|
| Gender (Males/Females)                  | 82/120              |                    |            |      |
| Age (years)                             | 45 (38;54)          |                    |            |      |
| Duration of intervention (days)         | 253 (219;289)       |                    |            |      |
| BMI (kg·m⁻²)                            | 29.0 (26.0;32.1)    | 27.9 (25.3;31.7)   | −4%        | <0.0001 |
| Waist circumference (cm)                | 97.0 (87.8;104.0)   | 92.3 (84.0;102.1)  | −5%        | <0.0001 |
| Total body fat mass (kg)                | 24.9 (17.9;32.2)    | 22.1 (16.1;29.2)   | −12%       | <0.0001 |
| Visceral fat mass (kg)                  | 2.6 (1.5;4.1)       | 2.0 (1.1;3.6)      | −21%       | <0.0001 |
| Subc. fat mass (kg)                     | 21.9 (15.4;28.8)    | 19.9 (13.8;26.5)   | −9%        | <0.0001 |
| Liver fat content (%)                   | 3.0 (1.4;6.9)       | 2.0 (1.0;5.0)      | −32%       | <0.0001 |
| Fasting glucose (mM)                    | 5.2 (4.8;5.6)       | 5.1 (4.8;5.5)      | −2%        | 0.003  |
| 2h glucose (mM)                         | 6.6 (5.8;8.0)       | 6.4 (5.6;7.7)      | −4%        | 0.03   |
| Insulin sensitivity (AU)                | 11.8 (7.1;17.4)     | 13.3 (8.9;19.3)    | 13%        | <0.0001 |
| Total cholesterol (mg/dl)               | 190.5 (164.8;215.0) | 186.0 (160.0;208.5) | −2%        | 0.046  |
| LDL cholesterol (mg/dl)                 | 119.0 (98.0;139.0)  | 115.0 (95.0;137.0) | −3%        | 0.003  |
| HDL cholesterol (mg/dl)                 | 50.0 (43.0;60.0)    | 52.0 (43.0;60.5)   | 4%         | 0.58   |
| Triglycerides (mg/dl)                   | 99.0 (76.0;137.5)   | 94.0 (69.0;131.5)  | −5%        | 0.02   |
| Fasting free fatty acids (μmol/l)       | 654.0 (503.0;800.0) | 602.5 (467.8;726.3) | −8%        | 0.0002 |
| 2h free fatty acids (μmol/l)            | 610.0 (43.0;93.0)   | 52.5 (34.0;76.0)   | −14%       | <0.0001 |
| Adipo-IR, (μmol/l/µM)                   | 31819 (20955;54918) | 24608 (16020;41159) | −23%       | <0.0001 |
| Physical activity (arb. u)              | 8.25 (7.50;9.00)    | 8.50 (8.09;9.25)   | 3%         | <0.0001 |
| VO₂peak (ml·min⁻¹·kg⁻¹)                | 24.5 (20.2;28.5)    | 25.3 (21.0;30.6)   | 3%         | 0.0002 |
| Total energy intake (kCal/day)          | 1925 (1628;2310)    | 1765 (1563;2082)   | −8%        | <0.0001 |
| Calories from carb. (%)                 | 47.2 (42.89;52.02)  | 48.77 (44.81;51.77)| 3%         | 0.046  |
| Calories from fat (%)                   | 32.48 (28.22;37.11) | 30.96 (27.90;34.67)| −5%        | 0.002  |
| Calories from protein (%)               | 15.38 (13.80;17.48)| 16.15 (14.86;17.94)| 5%         | 0.0001 |
| Fibers (g/d)                            | 22.25 (17.88;26.82) | 25.14 (21.47;30.15)| 13%        | <0.0001 |
| Niacin intake (mg/d)                    | 12.3 (9.8;15.9)     | 13.1 (10.7;16.2)   | 6%         | 0.06   |

Table 1. Subject characteristics before and after 9 months of the lifestyle intervention. Data are given as medians (interquartile range). P values were calculated with the Wilcoxon signed rank test. Adipo-IR, adipose tissue insulin resistance index; BMI, body mass index; MRT, magnetic resonance tomography; MRS, magnetic resonance (MR) spectroscopy; VO₂peak, Peak oxygen uptake.

standardized beta (β) or as non-standardized estimates. The relationship of the SNP rs1944438 C > T in DGAT2 with the change in liver fat content was studied using a dominant model. Logistic regression was applied to determine the odds ratio for achieving a resolution of NAFLD. A two-sided p-value ≤ 0.05 was considered statistically significant. The statistical software package JMP 10 (SAS Institute Inc, Cary, NC, USA) was used.

Results

The lifestyle intervention resulted in a mean weight reduction of 2.6 kg (p < 0.0001). Total body-, subcutaneous- and visceral fat mass and glycemia also decreased, while insulin sensitivity, the HPA score, representing an estimate of the average daily physical activity, and VO₂peak increased. Total energy intake and the intake of calories from fat decreased. On the other hand, the intake of fibers and of calories from carbohydrates, and particularly from protein, increased. The intake of niacin also tended to increase, however, the change of niacin intake was relatively small. The largest effect of the lifestyle intervention was found for liver fat content (−32%, Table 1). Nevertheless, a large intra-individual variability was observed for this change.

In the study, 32% of male subjects and 31% of female subjects had baseline niacin intakes fulfilling dietary

Next, we determined whether the change of liver fat content depended on the intake of other nutrients or the amount of physical activity. For this, we ran a forward stepwise regression analysis investigating the impact of total energy intake, intake of calories from carbohydrates, fat and protein, intake of fibers, niacin and physical activity at baseline, to predict the change of liver fat content. In this model, the intake of niacin strongly predicted the change of liver fat content and only the intake of calories from carbohydrates, was entered as another considerably weaker predictor into the stepwise regression model (Table 3). In addition, in a multivariable model, the

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change of liver fat content, after adjustment for total daily energy intake, intake of calories from carbohydrates and physical activity, all at baseline and at follow-up, the niacin intake at baseline was predictive of the change of liver fat content (Table 4).

Finally, using our initial model, we tested whether niacin intake predicted the change of liver fat content also after adjustment for the established predictor of liver fat content, aerobic fitness. Niacin intake at baseline remained significantly associated with the change of liver fat content, adjusted for baseline liver fat content, sex, age, total body fat mass at baseline and at follow-up, and total daily energy intake and VO2peak at baseline (n = 190, β = −0.20, p = 0.02).
Niacin intake. To better depict the relationships of niacin intake with changes of total body fat mass, body fat distribution and liver fat content, we divided the subjects into quartiles according to niacin intake at baseline. A significant association of niacin intake quartiles was observed with the change of liver fat content, while total body-, subcutaneous- and visceral fat mass did not associate with the niacin intake (Fig. 1).

From 202 subjects studied, 58 persons had NAFLD at baseline, with a median liver fat content of 12.6% (IQR 7.9–17.6). NAFLD resolved in 23 subjects during the lifestyle intervention. For one standard deviation increase in niacin intake, the odds ratio for resolution of NAFLD was 1.77 (95% CI, 1.00–3.43). To depict these relationships in the 58 subjects with NAFLD, they were then stratified by the median niacin intake. While liver fat content at baseline was similar in both groups, more subjects experienced a resolution of NAFLD in the group having a higher than in the group having a lower niacin intake (Fig. 2). In the subset of participants with NAFLD at baseline, there was a significant inverse association of niacin intake with fold-change liver fat content (using the same model covariates as given in Table 2, model 1, $\beta = -0.76 \pm -0.32$, $p = 0.02$ for log-transformed baseline niacin intake). Of note, the absolute effect size of niacin intake increased in the subgroup with fatty liver, indicating a higher reduction of liver fat content for a given amount of niacin intake in these subjects, compared to the whole dataset.

We observed an interaction of baseline niacin intake with BMI change during lifestyle intervention on the reduction of liver fat ($p = 0.04$, see Supplementary Fig. 1A,B), suggesting that the baseline-niacin dependent reduction of liver fat content was higher in participants loosing more weight.

Because we previously found that carriers of the minor T allele of the SNP rs1944438 in DGAT2 had less decrease in liver fat content during our lifestyle intervention compared to homozygous carriers of the major C allele, and niacin is known to inhibit DGAT2 activity in hepatocytes, we investigated whether this SNP interacts with niacin intake on the change of liver fat content. In fact, the interaction term niacin*SNP was significant ($p = 0.035$) (Table 2, model 2) and niacin more strongly predicted a decrease of liver fat content in homozygous carriers of the major C allele ($n = 68; \beta = -0.90, p = 0.0008$) than in carriers of the T allele ($n = 125; \beta = -0.22, p = 0.24$).

As niacin intake may also influence insulin sensitivity, we also tested whether niacin intake has an independent effect on the change of insulin sensitivity during the lifestyle intervention. Niacin intake at baseline was not associated with the change of insulin sensitivity after adjustment for baseline insulin sensitivity, sex, age, energy intake and total body fat mass at baseline and at follow-up ($\beta = 0.14, p = 0.13$). Also, no relationship of niacin intake with the change of the adipose tissue insulin resistance index, adjusted for the baseline adipose tissue insulin resistance index, sex, age, energy intake and total body fat mass at baseline and at follow-up was observed ($\beta = -0.002, p = 0.99$).
Discussion
Our data indicate that higher dietary niacin intake predicts a larger decrease of liver fat content during a lifestyle intervention. These data suggest that an elevated dietary niacin intake may facilitate the effect of lifestyle intervention on the treatment of hepatic steatosis in humans.

The initial hypothesis about a relationship of niacin intake with liver fat content was based on data about the intricate role of niacin in lipid metabolism. While niacin supplementation increases HDL-cholesterol, it reduces triglyceride levels, also lowering LDL-cholesterol and lipoprotein(a). The full mechanism of niacin action still remains elusive. Based upon available data, the reduction of hepatic triglyceride content that was seen in our subjects with high niacin intake could potentially be attributed to three possible mechanisms. First, niacin inhibits lipolysis in peripheral adipose tissue through its action on the specific nicotinic acid receptor hydroxy-carboxylic acid (HCA) receptor 2 (HCA2)12, thereby, reducing the flux of free fatty acids to the liver. However, the niacin-mediated suppression of lipolysis is subject to a rebound phenomenon23. This consecutive elevation of plasma FFAs most probably cannot be abrogated by the use of an extended-release niacin formulation23. Second, in vitro niacin directly and non-competitively inhibits DGAT2, the hepatic DGAT isoenzyme responsible for the committing step of triglyceride synthesis24. Third, niacin intake is associated with changes in the microbiome which plays an important role in human metabolism25.

The effect of niacin on liver fat content has so far been examined by two randomized controlled trials using extended release niacin preparations. In the first study comprising 9 participants on niacin and 9 on placebo, no niacin-associated change of liver fat content was observed23. In the second study involving 39 participants, niacin was shown to reduce liver fat content by a mean of 47.2% during the study duration of 23 weeks 24. However, a reduction of insulin sensitivity was observed in both studies.

In our study, high niacin intake during lifestyle intervention was associated with a larger reduction of liver fat content without affecting insulin stasis in humans.

What may explain the different findings between these studies in respect to insulin sensitivity? The studies by Fabbrini et al.23 and Hu et al.24 used pharmacologic niacin doses titrated to 2000 mg/day and 2000 mg/week, respectively. In contrast, dietary niacin intake amounted to up to 2 orders of magnitude less in our study (median, 12.3 mg/day, IQR 9.8, 15.9). Because a rebound elevation of FFAs is thought to be among the possible causes of niacin-induced decrease in insulin sensitivity when using higher doses26, we speculate that these effects were absent in our study. In support of this hypothesis, niacin intake was not associated with the change of adipose tissue insulin resistance index, which is an estimate of adipose tissue lipolysis. Thus, it can be speculated that the uptake of niacin by nutrients in lower doses less likely result in an FFA rebound. In context with recent data from Fangmann et al., we could also hypothesize that diet-bound niacin is predominantly released in specific intestinal segments such as the terminal ileum25. Niacin effect in the distal gut could directly or via the microbiome indirectly be responsible for reduction of liver fat.

Furthermore, we pinpointed a gene-nutrient interaction. A higher niacin intake was predictive of a larger decrease in liver fat content predominantly in homozygous carriers of the major C allele of rs1944438 in DGAT2.

Figure 2. Relationships between baseline niacin intake and resolution of NAFLD. In subjects with non-alcoholic fatty liver disease (NAFLD) before lifestyle intervention, liver fat content before (black bars) and after (grey bars) lifestyle intervention is depicted across groups with lower (below median) and higher (above median) niacin intake. The dashed line represents the percent liver fat content above that NAFLD is present (5.56%). In more participants NAFLD resolved during the lifestyle intervention in the higher niacin intake group than in the lower niacin intake group (p = 0.03, pie charts with grey slices representing the proportion of subjects experiencing a resolution of NAFLD.)
Interestingly, Hu et al. found that the effect of niacin treatment on the change of liver fat content also depended on a SNP in DGAT2. These data mechanistically support in vitro results on a direct effect of niacin to inhibit DGAT2 activity in hepatocytes. In addition, the fact that niacin intake predicted the decrease of liver fat content independently of the SNP rs1944438 in DGAT2 indicates that niacin may regulate liver fat content also by other mechanisms.

Our study has limitations. This work is retrospective analysis of a cohort study performed several years ago. Thus, dietary recommendations that may derive from the time when the study was conducted (2003–2009) may not necessarily be applicable to the setting today. Also, subjects were not asked to increase their niacin intake in a randomized controlled setting and, therefore, could only perform a retrospective analysis. However, as many longer lasting nutritional intervention strategies are prone to false reporting and poor compliance of the participants, we did not introduce such a bias that may largely affect the results. The results should be interpreted in the context of the cohort’s elevated type 2 diabetes and cardiovascular risk. We also did not account for intestinal niacin production in our work. It has been long known that intestinal bacteria can synthesize niacin. Despite careful adjustment of our models for possible confounders, we cannot exclude that a high dietary niacin intake may reflect a yet unknown beneficial behavioral or nutritional aspect that resulted in a larger decrease in liver fat content in these subjects. The fact that the niacin intake was associated with the change of liver fat content independently of the changes of body fat content, energy intake and physical activity, makes this hypothesis less probable. The interaction of niacin intake with rs1944438 in DGAT2 affected liver fat content. Since DGAT2 is the probable mediator of niacin’s effects on triglyceride synthesis, this strengthens the notion that the observed association was a result of differences in niacin intake. While DGAT2 constitutes the most probable link between niacin and hepatic steatosis via regulation of hepatic triglyceride synthesis, other potential mechanisms include the expression of ATP synthase beta chain in the cell surface of hepatocytes and the transcription of ATP-binding cassette transporter A1 (ABCA1) which promotes apoAI lipiddation. However, we have both precise measurements of liver fat content and well documented data about food diaries, enabling analyses in the present setting. Furthermore, the fact that the relationship of dietary niacin intake with the change of liver fat content was not only present in all 202 subjects, but also in the subgroup of 58 subjects with NAFLD at baseline, supports that the chance of the results being subject to a type I error is relatively small.

In conclusion, our study demonstrates that higher niacin intake favorably modulates the effect of a lifestyle intervention on liver fat content. This suggests a beneficial effect of higher niacin consumption in reaching the therapeutic goals of a lifestyle intervention to improve hepatic steatosis. Intervention studies testing the influence of niacin-fortified foods on outcomes of a lifestyle intervention to treat NAFLD are now needed to validate this concept.

Data Availability
The dataset analysed during the current study is not publicly available due to local data protection rules, but it is available from the corresponding author on reasonable request.

References
1. Younossi, Z. M. et al. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* **64**, 73–84 (2016).
2. Roden, M. Mechanisms of Disease: hepatic steatosis in type 2 diabetes—pathogenesis and clinical relevance. *Nature Clinical Practice Endocrinology & Metabolism* **2**, 335–348 (2006).
3. Gastaldelli, A. et al. Relationship Between Hepatic/Visceral Fat and Hepatic Insulin Resistance in Nondiabetic and Type 2 Diabetic Subjects. *Gastroenterology* **133**, 496–506 (2007).
4. Byrne, C. D. & Targher, G. NAFLD: a multisystem disease. *J. Hepatol.* **62**, S47–64 (2015).
5. Stefan, N., Fritsche, A., Schick, F. & Häring, H.-U. Phenotypes of prediabetes and stratification of cardiometabolic risk. *Lancet Diabetes Endocrinol* **4**, 789–798 (2016).
6. Bril, F. & Cusi, K. Management of Nonalcoholic Fatty Liver Disease in Patients With Type 2 Diabetes: A Call to Action. *Diabetes Care* **40**, 419–430 (2017).
7. Barb, D., Portillo-Sanchez, P. & Cusi, K. Pharmacological management of nonalcoholic fatty liver disease. *Metab. Clin. Exp.* **65**, 1183–1195 (2016).
8. Kantartzis, K. et al. High cardiorespiratory fitness is an independent predictor of the reduction in liver fat during a lifestyle intervention in non-alcoholic fatty liver disease. *Gut* **58**, 1281–1288 (2009).
9. Marchesini, G., Petta, S. & Dalle Grave, R. Diet, weight loss, and liver health in nonalcoholic fatty liver disease: Pathophysiology, evidence, and practice. *Hepatology* **63**, 2032–2043 (2016).
10. Rankinen, T. & Bouchard, C. Invited Commentary: Physical Activity, Mortality, and Genetics. *Am. J. Epidemiol.* **166**, 260–262 (2007).
11. Lukasova, M., Hanson, J., Tunaru, S. & Offermans, S. Nicotinic acid (niacin): new lipid-independent mechanisms of action and therapeutic potentials. *Trends in Pharmacological Sciences* **32**, 700–707 (2011).
12. Ganji, S. H. et al. Niacin noncompetitively inhibits DGAT2 but not DGAT1 activity in HepG2 cells. *J. Lipid Res.* **45**, 1835–1845 (2004).
13. Lehmann, R. et al. Circulating lysophosphatidylcholines are markers of a metabolically benign nonalcoholic fatty liver. *Diabetes Care* **36**, 2331–2338 (2013).
14. Stefan, N. et al. A high-risk phenotype associates with reduced improvement in glycaemia during a lifestyle intervention in prediabetes. *Diabetologia* **1–8**, https://doi.org/10.1007/s00125-015-3760-z (2015).
15. Stefan, N., Häring, H.-U. & Schulze, M. B. Metabolically healthy obesity: the low-hanging fruit in obesity treatment? *Lancet Diabetes Endocrinol* **6**, 249–258 (2018).
16. Tuomilehto, J. et al. Prevention of type 2 diabetes mellitus by changes in lifestyle among subjects with impaired glucose tolerance. *N. Engl. J. Med.* **344**, 1343–1350 (2001).
17. Baecke, J. A., Buurema, J. & Fritsche, J. E. A Short Questionnaire for the Measurement of Habitual Physical Activity in Epidemiological Studies. *Am J Clin Nutr* **36**, 936–942 (1982).
18. Stefan, N. et al. Inhibition of 11β-HSD1 with ROS93151 for non-alcoholic fatty liver disease: a multicentre, randomised, double-blind, placebo-controlled trial. *The Lancet Diabetes & Endocrinology* **2**, 406–416 (2014).
19. Szczepaniak, L. S. et al. Magnetic resonance spectroscopy to measure hepatic triglyceride content: prevalence of hepatic steatosis in the general population. *Am. J. Physiol. Endocrinol. Metab.* 288, E462–468 (2005).

20. Matsuda, M. & DeFronzo, R. A. Insulin sensitivity indices obtained from oral glucose tolerance testing: comparison with the euglycemic insulin clamp. *Diabetes Care* 22, 1462–1470 (1999).

21. Lomonaco, R. et al. Effect of adipose tissue insulin resistance on metabolic parameters and liver histology in obese patients with nonalcoholic fatty liver disease. *Hepatology* 55, 1389–1397 (2012).

22. Institute of Medicine (US) Standing Committee on the Scientific Evaluation of Dietary Reference Intakes and its Panel on Folate, Other B Vitamins, and Choline and Subcommittee on Upper Reference Levels of Nutrients. *Dietary reference intakes for thiamin, riboflavin, niacin, vitamin B6, folate, vitamin B12, pantothenic acid, biotin, and choline.* (National Academies Press (US) 1998).

23. Fabbrini, E. et al. Effect of Fenofibrate and Niacin on Intrahepatic Triglyceride Content, Very Low-Density Lipoprotein Kinetics, and Insulin Action in Obese Subjects with Nonalcoholic Fatty Liver Disease. *Journal of Clinical Endocrinology & Metabolism* 95, 2727–2735 (2010).

24. Hu, M. et al. Liver fat reduction with niacin is influenced by DGAT-2 polymorphisms in hypertriglyceridemic patients. *J. Lipid Res.* 53, 802–809 (2012).

25. Fangman, D. et al. Targeted Microbiome Intervention by Microencapsulated Delayed-Release Niacin Beneficially Affects Insulin Sensitivity in Humans. *Diabetes Care* 41, 398–405 (2018).

26. Oh, Y. T. et al. Continuous 24-h nicotinic acid infusion in rats causes FFA rebound and insulin resistance by altering gene expression and basal lipolysis in adipose tissue. *Am. J. Physiol. Endocrinol. Metab.* 300, E1012–1021 (2011).

27. Burkholder, P. R. & McVeigh, I. Synthesis of Vitamins by Intestinal Bacteria. *Proc. Natl. Acad. Sci. USA* 28, 285–289 (1942).

28. Nagayama, K. & Fujita, A. The synthesis of vitamins by intestinal bacteria in man and the effect of cellulose. *V. Synthesis of niacin.* *J. Vitaminol (Kyoto)* 3, 24–29 (1957).

29. McLaren, J., Ngo, D. T. & Olivera, B. M. Pyridine nucleotide metabolism in Escherichia coli. *I. Biosynthesis from alternative precursors in vivo. J. Biol. Chem.* 248, 5144–5149 (1973).

30. Magniásdóttir, S., Ravcheev, D., de Crécy-Lagard, V. & Thiele, I. Systematic genome assessment of B-vitamin biosynthesis suggests co-operation among gut microbes. *Front Genet* 6 (2015).

31. Zhang, L.-H., Kamanna, V. S., Zhang, M. C. & Kashyap, M. L. Niacin inhibits surface expression of ATP synthase beta chain in HepG2 cells: implications for raising HDL. *J. Lipid Res.* 49, 1195–1201 (2008).

32. Kamanna, V. S., Ganji, S. H. & Kashyap, M. L. Recent advances in niacin and lipid metabolism. *Curr. Opin. Lipidol.* 24, 239–245 (2013).

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**Author Contributions**

All authors provided substantial contributions to conception and design, acquisition of data or analysis and interpretation of data, drafting the article or revising it critically for important intellectual content and final approval of the version to be published. Here are the most important contributions of each author: K.L., N.S. and R.W. researched data and wrote the manuscript. C.W., K.K., J.M., F.S., M.G. and S.K. researched data and edited the manuscript. A.F. and H.U.H. designed the study, contributed to the discussion and reviewed the manuscript.

**Additional Information**

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