Review

Evolution, revolution and heresy in the genetics of infectious disease susceptibility

Adrian V. S. Hill*

Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford OX3 7BN, UK

Infectious pathogens have long been recognized as potentially powerful agents impacting on the evolution of human genetic diversity. Analysis of large-scale case–control studies provides one of the most direct means of identifying human genetic variants that currently impact on susceptibility to particular infectious diseases. For over 50 years candidate gene studies have been used to identify loci for many major causes of human infectious mortality, including malaria, tuberculosis, human immunodeficiency virus/acquired immunodeficiency syndrome, bacterial pneumonia and hepatitis. But with the advent of genome-wide approaches, many new loci have been identified in diverse populations. Genome-wide linkage studies identified a few loci, but genome-wide association studies are proving more successful, and both exome and whole-genome sequencing now offer a revolutionary increase in power. Opinions differ on the extent to which the genetic component to common disease susceptibility is encoded by multiple high frequency or rare variants, and the heretical view that most infectious diseases might even be monogenic has been advocated recently. Review of findings to date suggests that the genetic architecture of infectious disease susceptibility may be importantly different from that of non-infectious diseases, and it is suggested that natural selection may be the driving force underlying this difference.

Keywords: susceptibility; genomics; pathogen-driven selection; evolution; diversity; exome

1. WHY STUDY THE GENETICS OF INFECTIOUS DISEASE SUSCEPTIBILITY?

Studies of genetic susceptibility to infectious diseases in humans have a long history, starting with blood groups and other phenotypic markers and a striking success in the early 1950s with the identification of the sickle haemoglobin variant as a major resistance factor for malaria [1]. Today, a great variety of approaches are used from twin studies to whole-genome sequencing. These efforts have several goals. Foremost is a better understanding of disease pathogenesis and resistance in the expectation that this will lead, in time, to improved interventions such as better drugs or vaccines to prevent or attenuate the great global burden of infectious disease morbidity and mortality. With over 10 million deaths annually from infectious diseases and the threat of new epidemics and pandemics, this is a very high priority. There are clear examples where human genetic research has supported the development of new drugs and vaccines. A recently introduced class of human immunodeficiency virus (HIV) drugs known as entry inhibitors were developed in part based on the knowledge that a specific deletion in the gene for the CCR5 co-receptor greatly reduced the risk of HIV infection and the rate of disease progression after infection [2]. The leading vaccine candidate for Plasmodium vivax malaria [3], the Duffy binding protein, was identified and developed supported by evidence that genetic absence of its red blood cell entry receptor provides almost complete protection from this type of malaria [4]. Awareness of the prevalence of a common immunodeficiency of mannose-binding lectin, conferring increased risk of bacterial disease, has encouraged the development of replacement therapy [5].

A second application that is gaining increasing attention is the potential to stratify populations for risk of infectious disease based on genetic profiling. This has not been a priority until now as most preventive interventions such as childhood vaccines have been aimed at universal coverage. However, as more potentially useful vaccines are licensed and the costs of new vaccines escalate, targeted use is becoming a consideration. When a genetic profile costs less than a vaccine and the profile has many other applications in predicting disease risk, it may well be cost-effective to target newer vaccines to those who will benefit most from them. The recent awareness that low-frequency large-effect variants may make a large contribution to inter-individual genetic variation in susceptibility to many diseases [6] should increase interest in defining, early in life, the constellation of potentially deleterious variants that comprise an individual’s inheritance.

But the third, and one of the most interesting aspects of this field, is that most relevant to the theme of this issue. The evolutionary significance of genetic variation in susceptibility to infection has long fascinated the public as well as physicians and infection specialists. Questions such as the importance

*adrian.hill@well.ox.ac.uk

One contribution of 14 to a Discussion Meeting Issue ‘Immunity, infection, migration and human evolution’.
of infectious diseases in generating and maintaining the great diversity that we can now readily define in our genomes have long been debated. I will discuss some aspects of evolutionary interest towards the end of this review, after first providing an overview of approaches and recent progress in this field. The focus will be on information that has been provided by large well-designed case–control studies, which have provided the most compelling evidence of the relevance of specific genetic variants to infectious disease susceptibility.

2. INFECTIOUS DISEASE SUSCEPTIBILITY IS GENETICALLY CONTROLLED

There are some well-studied examples of familial clustering of severe infectious disease syndromes, and these very rare monogenic disorders have been reviewed elsewhere [7]. A more challenging question is the extent to which common major infectious diseases are affected by host genetics. Here, the standard genetic measure used for complex traits, lambda-s, a measure of the increase in risk to siblings of an affected case compared with an unrelated individual, is confounded by the tendency of people to live with their relatives, so that dissecting the effect of environment from shared genes become very difficult. A better approach is to compare the concordance of disease in fraternal and identical twin pairs, where a greater concordance in the latter provides a measure of heritability. Such studies have been undertaken for several infectious diseases, usually many years ago, and a relatively consistent picture of significant heritability for chronic infectious diseases emerges. The evidence is less clear for acute infections such as measles, where exposure and infection rates were very high when these early twin studies were performed [8]. But in tuberculosis [9], leprosy [10], Helicobacter pylori infection [11], chronic hepatitis B infection [12] as well as in the phenotype of immune responses to vaccination [13,14], there is evidence of greater concordance in monozygotic compared with dizygotic twin pairs.

3. APPROACHES TO GENE IDENTIFICATION

One of the most interesting aspects of this field is the wide variety of approaches that can be, and have been, taken towards identification of relevant susceptibility genes for infectious diseases. Some have advocated the use of animal models of infectious diseases as a route to finding human infectious diseases susceptibility loci [15]. It is clear that rodent strains can differ strikingly in susceptibility to infections and genes can be mapped and identified in controlled challenge studies. The difficulty has been in extrapolating these findings to humans who will often not show comparable polymorphism in the relevant loci or may even lack a homologous gene. A related approach is to define the susceptibility profile of artificially generated knockout mice. Such efforts continue and are clearly of value in identifying pathways of potential relevance to human infectious disease susceptibility, but in terms of actual definition of known human susceptibility loci, very few of these have emerged from animal model analysis. Recently, a tuberculosus susceptibility gene was suggested by an analysis of zebrafish genetics [16], but the homologous gene was not associated with human tuberculosis susceptibility in a very large study of over 3700 clinical cases [17].

Prior to the availability of genome-wide association technologies, linkage mapping was attempted to map major genes in common infectious diseases. This required the laborious collection of families with multiple cases, usually affected sibling pairs, and the use of microsatellite markers to define chromosomal segments segregating with disease. It was possible to identify a small number of regions linked to leprosy [18–20], tuberculosis [21,22] and chronic hepatitis [23] in these family studies and some positional candidate genes were associated with disease. But it has been difficult to define the mapped loci in large-scale association studies and to date none appears to have been independently confirmed. These searches for genetic loci of very large effect that might be detectable in such small-scale linkage studies were encouraged by a form of pedigree analysis known as complex segregation analysis, which was claimed to identify the existence (but not the location) of major loci for many infectious diseases [24,25]. However, the clearest result from these genome-wide linkage studies is that such prevalent major genes must be very rare in infectious diseases, in contrast to some autoimmune diseases where major loci, especially in the human leucocyte antigen (HLA) complex, could be detected by affected sibling pair studies. In summary, the central problem with this approach is that most infectious disease susceptibility genes have too small effect sizes and/or too low a frequency to allow detection in linkage studies of even hundreds of families.

After this foray into family studies, the field returned to association studies of infectious disease susceptibility comparing the frequency of variants in affected cases with those of healthy controls. Ever since the successful identification of sickle haemoglobin as a resistance locus with those of healthy controls. Ever since the successful identification of sickle haemoglobin as a resistance locus in the 1950s [1], and extensive blood group association studies in the 1960s [26], such case–control studies have been the most widely used and most successful approach to identifying convincing infectious disease susceptibility genes in humans.

However, the very simplicity of the approach and the reducing cost of genotyping have generated their own problems. A proliferation of studies of modest size, and with apparently statistically significant results, led to a greater focus of thresholds for claiming statistical significance and to a preference, and later a requirement, for replication studies to ‘confirm’ an association. More recently, with the advent of whole-genome association arrays and very recent exome and whole-genome sequencing studies, criteria for ‘statistical significance’ are being assessed again. We remain in a transitional stage where some journals will accept reports with significant levels as modest as $p < 0.05$, and others will require replication studies and a ‘genome-wide significance’ level of $p < 5 \times 10^{-7}$ or less. This dramatic difference in publication thresholds complicates review of the field as well as interpretation of results, as does the long-standing [27] tendency for positive results to be published more readily than negative ones.

*Phil. Trans. R. Soc. B* (2012)
4. CANDIDATE GENE STUDIES

I shall start with some examples of infectious disease associations that appear convincing and are unusual, in that the effect size is very large. All of these are effectively candidate gene studies, in that the investigators chose to assess these particular genes based on some biological information rather than a genome-wide scan. I am aware of six associations that are major effects [28], arbitrarily defined as an odds ratio exceeding 3, where the variant allele is polymorphic in some human populations. These six (table 1) are all supported by case–control analysis of allele and genotype frequencies but, interestingly, three were discovered in a surprising manner. The first reports of the sickle haemoglobin [1] and Duffy blood group associations [4] with malaria resistance included analysis in deliberately infected volunteers, as did the identification of the importance of the non-secretor status for blood group antigens, determined by the fucosyltransferase (FUT2) gene, in resistance to infection by norovirus [31], a frequent viral cause of diarrhoeal disease. The ability to identify these associations in small microbial challenge studies reflects both the large effect sizes of these variants and excellent biological insights suggesting the relevant candidate gene.

In contrast, other candidate gene associations (table 2) that appear compelling typically have smaller effect sizes, with odds ratios of 0.5–2, but most of these are still larger effects than are generally found by genome-wide analyses of non-infectious diseases (with the sole exception of HLA associations with autoimmunity where occasional exceptionally strong associations are seen).

It is impossible to review in any detail here the plethora of reports of candidate genes that have now been associated with various infectious diseases, and there have been several recent reviews [28,42] and an entire monograph on this subject [43]. Some of the more compelling associations are listed in table 2, with references. Instead, I will discuss some selected candidates of immunological relevance that may impact on the risk of disease from several infectious pathogens. Associations with susceptibility to several infectious diseases may of course increase the evolutionary importance of specific genetic variants.

5. INNATE IMMUNE SIGNALLING PATHWAYS

An explosion of research in innate immunity over the last decade has led to considerable clarification of the role of
several classes of innate receptors in triggering inflammation and acquired immunity [44]. The toll-like receptor (TLR) signalling pathway has been the best characterized leading to a particular emphasis on polymorphisms in this pathway by genetic studies. A striking example is the prevalence of a variant of one of the four TLR signalling adaptors, named MAL (encoded by the gene TIRAP) in many populations [38]. This appears to be a functional knockout allele, and heterozygotes have been reported to have an approximately twofold reduction in risk of several infectious diseases, including pneumococcal disease, tuberculosis and malaria [38]. The homozygous genotype is rare in many populations but in Europe, where this genotype is less uncommon, homozygotes may be at a slightly increased risk of bacterial disease [38]. If so, this surprising genotypic pattern could contribute to the evolutionary maintenance of this polymorphism over long periods of time.

Another example of an immunological gene that impacts on multiple infectious diseases was reported recently. Cytokine-inducible SRC homology 2 domain protein (CISH) was the first member of the regulatory suppressor of cytokine signalling (or SOCS) family to be described, and CISH is the gene most consistently upregulated by interleukin (IL)-2 stimulation in humans, and is a key negative regulator of cytokines, especially IL-2. Single-nucleotide polymorphisms (SNPs) flanking the promoter of this gene were associated with risk of bacteremia, tuberculosis and severe malaria in diverse populations and at least one of these SNPs was shown to be functionally relevant [45].

A coding change at position 602 in the TLR1 protein, a key innate receptor component for many bacteria, was shown to impact on the ability of this receptor to reach the cell surface. The variant is found at low frequencies in Africa, but is the more frequent allele in many Caucasian populations. This variant has recently been associated with a substantial reduction in risk of leprosy in Turkey and India, with the cumulative data reaching impressive statistical significance (p < 10^{-7}) [39,46]. Intriguingly, the same variant was associated with a reduced risk of tuberculosis in an exon resequencing study of TLR genes in African Americans [47]. So this may provide another example of a variant impacting on several diseases, and in this case, there is suggestive evidence that the protective variant may have increased in frequency by providing protection against mycobacterial disease.

Finally, one of the best studied genes impacting on a wide variety of autoimmune diseases is the lymphoid tyrosine phosphatase (Lyp) gene, PTPN22. An R620W change is common in Europeans and increases the risk of type I diabetes, rheumatoid arthritis, systemic lupus erythematosus and several other autoimmune diseases [48]. Interestingly, the same variant was associated with an increased risk of invasive pneumococcal disease and empyema (a form of suppurative lung disease) [49] and very recently with leprosy in India [50]. Although it might be expected that alleles associated with an increased risk of autoimmunity might protect against infectious diseases, in this case the observed association is in the opposite direction.

Many other infectious disease associations with genetically variable components of the innate and acquired immune systems are appearing. Several, such as the relevance of variants in genes encoding inhibitors for NFkB signalling [51–53], have been studied only in a small number of infections (in this case pneumococcal disease), and more extensive surveys of infectious diseases are justified. The challenge continues to be accessing large well characterized clinical series with suitable controls to allow adequately powered studies.

In summary, just as HLA genes may impact on risk of several infectious diseases as well as autoimmunity, there are now several examples of non-HLA loci that also appear relevant to multiple infections as well as examples of rare monogenic variants that impact on the risk of several infectious diseases [54]. The consequences of this for evolutionary change in allele frequencies over time will undoubtedly be complex. Not only will selection pressures fluctuate with the prevalence and strain of infectious pathogens (see below), but the presence of different pathogens might have either opposing or additive impacts on the change in allele frequencies of particular genetic variants.

6. GENOME-WIDE ASSOCIATION STUDIES

The introduction of genome-wide association studies (GWAS) has revolutionized the field of complex disease genetics. The availability of millions of SNPs mapped across the human genome and of microarrays that allow cost-effective genotyping of millions of SNPs in thousands of individuals provided, for the first time, the opportunity to test fairly comprehensively for genetic markers that would tag causative variants [55]. In many diseases, the yield of this approach has been spectacular, with hundreds of loci now identified, for example, autoimmune diseases. This is providing new insights into the molecular pathogenesis of these diseases as a route to designing and developing new treatments. There is also potential value in devising diagnostic arrays that would allow some prediction of risk of certain diseases.

Despite these spectacular successes, there are limitations to this approach. Even in the diseases where many dozens of loci have been identified and replicated, the proportion of the estimated genetic variance that can be explained by all the identified genetic associations is modest, often less that 20 per cent of the total genetic variances suggested by twin studies [56]. This is because the individual associations are generally of modest magnitude, with odds ratios typically of 1.1 to 1.5. Furthermore, there is little evidence of epistatic interactions between loci that could account for some of the missing genetic component. Searches for structural variants that might account for the missing component have met with little success. The leading current hypothesis to explain the missing component is that rare variants, perhaps with larger effect sizes, are cumulatively responsible [56], but because of their low frequencies these are very poorly assessed with available GWAS microarrays.

For infectious diseases, the picture is yet more complicated. Fewer diseases have been studied by GWAS, in part because the approach requires very large sample sizes to be worthwhile. This results from the very large number of SNPs assayed, leading to a major challenge in sorting true signals from the noise or false positives that inevitably turn up when assaying about a
Although this region is gene-poor, a flanking analysis with genome-wide significance on chromosome 18 from The Gambia and Ghana identified a significant signal [57]. A parallel GWAS study of malaria in Africa, a combined analysis of tuberculosis disease have been undertaken with some notable successes. In Africa, a further challenge is found in studies of African populations, which were among the first to be studied by GWAS for infectious diseases, as part of the Wellcome Trust Case–Control Consortium [57,58]. The shorter GWAS Ghana million variants. The general approach is to require replication of positive signals and to use a challenging threshold level of ‘genome-wide’ statistical significance of about \( p = 5 \times 10^{-7} \). This requirement for typically thousands of cases and suitable matched controls has limited GWAS studies of infectious diseases. Despite the high prevalence of many infectious diseases in developing countries, resource limitations still make it harder to collect 2000 tuberculosis cases in West Africa than 20 000 diabetes cases in western Europe. A further challenge is found in studies of African populations, which were among the first to be studied by GWAS for infectious diseases, as part of the Wellcome Trust Case–Control Consortium [57,58]. The shorter extent of linkage disequilibrium in Africans compared to non-Africans leads to a requirement for larger numbers of SNPs to attain the same coverage of the genome as in Caucasian and Asian populations. Greater diversity in populations in Africa and complexities in haplotype structure may also complicate fine mapping of causative SNPs (figure 1) [58].

Nonetheless, several GWAS studies of infectious diseases have been undertaken with some notable successes. In Africa, a combined analysis of tuberculosis cases from The Gambia and Ghana identified a signal with genome-wide significance on chromosome 18 [57]. Although this region is gene-poor, a flanking gene that encodes the transcription factor GATA6 is an excellent positional candidate for tuberculosis. Another signal was found on chromosome 2 in the P4LBD3B cell polarity gene [57], which interestingly also emerged from a GWAS of HIV disease progression [59] (although the associated regions of the gene in the two studies are different). Further statistical analysis of the tuberculosis GWAS datasets using an imputation technique to estimate association with less well-covered genomic regions has identified a further highly significant signal [60]. A parallel GWAS study of malaria in Gambia identified the haemoglobin AS variant with clear statistical significance but highlighted the challenges of using limited microarray coverage to map even such strong hits in Africans [58]. A very striking positive association was reported for hepatitis C where one of the interferon lambda genes, IL-28B, showed a striking association with viral clearance and with response to treatment [40].

But probably the most marked GWAS success in infectious diseases has been a study of leprosy reported by a Chinese group [32]. By studying about 4000 cases and 7000 controls, five new genes showed strongly significant associations (\( p < 10^{-10} \)) in addition to the expected HLA class II region association. Three of these six genes could be replicated in an independent study of Indian patients with leprosy [61]. Most interestingly, the susceptibility genes identified in Chinese leprosy show a striking overlap with genes previously associated with Crohn’s disease [32]. Although this does not necessarily implicate mycobacteria in the aetiology of Crohn’s disease, a theory for which there is some support [62], this does point to the same pathways, related to NOD2 signalling, being relevant in these two granulomatous diseases.

In contrast, large GWAS efforts in HIV/AIDS have been less successful [63]. Although the associations in the HLA class I regions with rate of progression to AIDS were reconfirmed and well mapped, no convincing new associations were identified in other chromosomal regions [33,64]. However, these GWASs of disease progression have been limited to non-African populations and studies in the continent most affected by the epidemic and with the greatest human genetic diversity should still be worthwhile. Similarly, a somewhat smaller GWAS in meningococcal disease confirmed an association with the complement factor \( H \) (\( CFH \)) gene, previously identified in a candidate gene study of this disease [37]. And in malaria a first GWAS in Gambians failed to identify new loci with genome-wide significance [58], although large-scale GWASs in other African populations are continuing as part of the extensive MalariaGEN consortium [65].

Figure 1. Association plot of the main associated locus identified in combined analysis of genome-wide association studies of tuberculosis in The Gambia and Ghana. The y-axis show the negative log of the \( p \)-value for the association test. The peak of association is in a gene-poor region, but the positions of flanking genes are shown. Adapted from Thye et al. [57].

### 7. GENETIC ARCHITECTURE OF INFECTIOUS DISEASES

The limited success, at least to date, of GWAS approaches in infectious diseases raises the question of whether this is
Table 3. Theories of the genetic architecture of human infectious disease susceptibility. Theory 1 suggests that most relevant genetic variation is encoded by relatively common variants that cumulatively account for most of the genetic variance. Theory 2, in complete contrast, is the extreme view that most relevant genetic variation is encoded by very rare new mutations that have almost complete penetrance, as in primary immunodeficiency diseases. The more recent theory 3 suggests a predominant role for many individually rare variants with incomplete penetrance, which are generally not new mutations, but cumulatively account for most of the genetic component to infectious disease susceptibility.

| Theory   | Description                                                                 | Examples                                                                 |
|----------|-----------------------------------------------------------------------------|--------------------------------------------------------------------------|
| Theory 1 | **Common variants** higher frequency variants                                | *Identifiable by genome-wide association scans*                           |
|          | *Not fully penetrant*                                                       |                                                                          |
| Theory 2 | **Very rare monogenic variants** highly penetrant mutations                 | *Mainly novel mutations*                                                 |
|          | *Not fully penetrant*                                                       |                                                                          |
| Theory 3 | **Multiple rare diverse variants** not fully penetrant                       | *Mainly standing genetic variation*                                       |
|          |                                                                             |                                                                          |

The most efficient means of identifying the genetic factors underlying variable susceptibility. Indeed, for many non-infectious diseases the field is moving towards next-generation sequencing approaches to try to identify the genes accounting for the heritability not identified through GWAS. This discussion raises the key question of the genetic architecture of variable susceptibility to disease (table 3). There are at least three competing views. GWAS analysis was based on the hypothesis that most of the genetic component to common diseases can be explained by common variants; this is sometimes dubbed the common disease–common variant hypothesis [66]. Because GWAS relies on linkage disequilibrium between common genotyped markers and relatively common causative variants, it has inadequate power to detect significant associations with rare causative variants. The second hypothesis listed in table 3 is that rare monogenic variants with high penetrance account for the bulk, or all, of the genetic component to common disease [67]. Although this may sound implausible, given the lack of Mendelian inheritance of most common disease, it has been seriously proposed for at least paediatric infectious diseases [68]. This idea emerged from the primary immunodeficiency disease field where it has been fairly straightforward to identify some causal mutations by sequencing of candidate genes in rare pedigrees with familial severe susceptibility or rare novel mutations. Emboldened by success with these rare phenotypes, one group has suggested that the same monogenic major gene genetic architecture underlies common infectious disease phenotypes [68,69].

A newer and more plausible third hypothesis (table 3) is advocated here for infectious diseases. This is the concept that much of the genetic component to common infectious disease may be accounted for by the cumulative effects of many rare mutations with limited penetrance [66]. While, in this view, rare mutations would account for much of the genetic component, a consensus view would allow for hypothesis 1 to also be partly true, with common variants accounting for part of the genetic component: of course, there is good evidence that this is the case for many diseases (as reviewed earlier). Finally, one should allow that rare familial cases of severe susceptibility, usually in young children, can indeed be Mendelian as has been documented in selected rare cases [67]. However, the lack of evident Mendelian transmission in most cases of infectious disease susceptibility and the evolutionary consideration that severe deleterious mutations should be rapidly eliminated from populations indicates that the contribution of highly penetrant deleterious mutations to the great majority of infectious disease cases must be very limited.

There is a practical relevance to the understanding of which genetic architecture is most correct. If common variants are of over-riding importance, an extension of the GWAS approach, with microarrays of improved coverage and analysis of larger sample sizes, might be the best way forward. But if rare variants account for most susceptibility, sequencing approaches will be required. Fortunately, next-generation sequencing costs continue to fall rapidly and promise to allow hypotheses 2 and 3 to be assessed in the foreseeable future.

8. TOWARDS EXOMICS

Sequencing costs remain a major limitation today and, even though whole genome sequencing of over three trillion base pairs is feasible, the costs of full genomic sequencing of thousands of samples in just one infectious disease are still prohibitive. Fortunately, sequencing of just the exome, composed of all the exons in the genome, and some limited exon flanking sequence, might provide a cost-effective alternative [70–72]. This is feasible using a variety of capture matrices. The exome comprises just one per cent of the human genome, and costs per sample are falling rapidly from thousands to hundreds of pounds per sample. Although costs will still be high initially, a limited number of samples can be sequenced for the full exome, to search for suggestive associations, and these can be reassessed in a replication study by more limited sequencing of genes of interest on larger sample numbers. As well as analysing individual mutations, the mutational load in a gene, comprising the full set of coding changes identified, can be compared between cases and controls.

Initial data on exon sequencing in infectious diseases are encouraging. Smirnova et al. [73] focused on the TLR4 gene in meningococcal disease and identified a substantial excess of rare coding changes in cases compared with those in controls, with a remarkably large suggested effect size. Ma et al. [47] studied five TLR genes and again found an excess of rare (and some more frequent) coding changes in tuberculosis cases compared with controls.

An argument against focusing on exons is that when one reviews the variants that have been associated with non-infectious disease susceptibility using GWAS analyses, there is strong evidence that the majority of changes identified are regulatory or at least non-coding changes [55]. If this is also true of infectious diseases, then exome sequencing could miss most of the relevant...
changes. However, I suggest that matters may be different for infectious diseases, based on review of the available data on known associations. Table 2 lists the majority of the most convincing associations with infectious diseases where the causative genetic changes are both polymorphic in human populations and reasonably well defined. It is striking that in sixteen out of seventeen cases, the association could have been identified by exome sequencing. Either the mutation is in an exon or, as in the case of the IL-28B–hepatitis C association, a change in an exon tracks very well a flanking (possibly regulatory) change. This excess of exonic changes contrasts strikingly with GWAS-identified changes in say autoimmune diseases. A second difference is also evident. In most GWAS studies of non-infectious diseases, the minor allele is associated with an increased risk of disease whereas, as indicated in table 2, there are more examples of protective than susceptible minor alleles in infectious disease associations.

There are many possible reasons for these striking differences, including some differences in ascertainment methods, but if these differences are real a likely contributor is the action of natural selection by infectious pathogens. In well-studied examples, such as malarial selection of haemoglobin S, it is clear that the protective minor allele has reached polymorphic diversity in humans over long periods of evolutionary time, as is well illustrated by consideration of the maintenance of the extraordinary diversity of HLA genes in the human major histocompatibility complex. Although heterozygote advantage [75] and frequency-dependent selection [76,77] are the most frequently invoked modes of selection capable of maintaining polymorphic diversity, fluctuating selection pressures [76,78] (varying either spatially or temporally, or both) are also capable of maintaining polymorphism. Indeed, epidemiological records provide powerful evidence that fluctuating selection by lethal infectious pathogens is commonplace in recent human history and may always have been so, at least since the increase in population densities following the introduction of agriculture.

Such fluctuation in selection pressures will result not just from the transient occurrence of particular epidemics but from temporal and spatial changes in the antigenic composition of pathogens, as different strains are likely to interact differentially with particular host resistance genotypes. The impact on signatures of selection [79], as currently deduced from genomic sequence data, could be profound in that short periods of selection and then a reversal of the selection pressure would tend to minimize any detectable signals in the genome over time. As discussed earlier, several host genes affect resistance to multiple infectious pathogens and this too could tend to weaken signals if different pathogens interact differentially with various host genotypes. The overall effect of these complexities is that many statistical searches for genomic signatures of selection may underestimate the evolutionary impact of infectious pathogens, because of the complexity of selection involved, in contrast to simple directional selection pressures provided by some other factors such as climate or dietary change.

A major interest of the Discussion Meeting that led to this publication has been the use of human genetic variants as markers of migration routes and population differentiation in recent human evolution. Although some theoretical considerations would point to a preference for non-selected markers for this purpose, in practice, variants that have clearly been selected by infectious pathogens have proved very useful for tracing migration and population affinities. These include examples from the highly polymorphic HLA genes [80], and also specific haemoglobinopathies that were selected to high frequency by malaria but subsequently could be used to trace population migrations. For example, a specific alpha globin gene deletion in the southwest Pacific helped early genetic studies of the colonization of Oceania [81].

9. NATURAL SELECTION AND INFECTIOUS DISEASE SUSCEPTIBILITY GENES

Case–control studies have provided the strongest evidence of ongoing selection by infectious pathogens on genetic polymorphism in humans. Less direct evidence of selection is provided by a variety of other statistical approaches to the analysis of genomic data, some of which are described in other chapters of this issue. A recent development is the use of a composite of multiple signals that can pinpoint variants likely to have undergone selection [74]. These regions and SNPs can then be prioritized for analysis in case–control studies of particular infectious diseases, illustrating the potential complementarity of these approaches.

However, some caveats in each approach should be noted. Some signals of selection extant in our genomes will have been generated by pathogens or strains of pathogens that have died out or been eradicated, preventing case–control studies of their effects, although in vitro studies or animal models might still allow some analysis and inferences. Conversely, the temporally varying natural history of epidemics and ever-varying genetic diversity of pathogens may tend to minimize signals of selection in our genomes, particularly when compared with the more continuous effects of other selective agents such as sunlight and climate. In this context, it is worth recalling that at least three types of selection have been demonstrated to be capable of maintaining genetic diversity in humans over long periods of evolutionary time, as is well illustrated by consideration of the maintenance of the extraordinary diversity of HLA genes in the human major histocompatibility complex. Although heterozygote advantage [75] and frequency-dependent selection [76,77] are the most frequently invoked modes of selection capable of maintaining polymorphic diversity, fluctuating selection pressures [76,78] (varying either spatially or temporally, or both) are also capable of maintaining polymorphism. Indeed, epidemiological records provide powerful evidence that fluctuating selection by lethal infectious pathogens is commonplace in recent human history and may always have been so, at least since the increase in population densities following the introduction of agriculture.

Such fluctuation in selection pressures will result not just from the transient occurrence of particular epidemics but from temporal and spatial changes in the antigenic composition of pathogens, as different strains are likely to interact differentially with particular host resistance genotypes. The impact on signatures of selection [79], as currently deduced from genomic sequence data, could be profound in that short periods of selection and then a reversal of the selection pressure would tend to minimize any detectable signals in the genome over time. As discussed earlier, several host genes affect resistance to multiple infectious pathogens and this too could tend to weaken signals if different pathogens interact differentially with various host genotypes. The overall effect of these complexities is that many statistical searches for genomic signatures of selection may underestimate the evolutionary impact of infectious pathogens, because of the complexity of selection involved, in contrast to simple directional selection pressures provided by some other factors such as climate or dietary change.
An overview of the infectious diseases that have received most attention by human geneticists and those that have produced the clearest successes provides a striking outlier. HIV, tuberculosis and malaria have probably had most investment and attention, in accordance with their global public health burden, but the star performer in terms of results must be leprosy. This was the first infectious disease shown to be HLA-associated [82], the most successful disease for mapping loci by linkage analysis [18–20] and, to date at least, the most successful example of a GWAS [32]. In the past, this may have reflected easier sample collection for leprosy but this is no longer the case. A more likely explanation may relate to the extreme lack of genetic polymorphism in the causative bacterium, Mycobacterium leprae, which has now been well documented by whole genome sequencing [83]. If this explanation is correct, then the implications may be profound. Many of the difficulties in mapping and identifying strong genetic factors for human infectious diseases may relate to the ‘pooling’ of genetically diverse strains of a single pathogen in studies of a single infectious disease. Hence, separating out cases according to strain or genotype of the pathogen may increase power to detect strain-specific host effects, and already there is some preliminary evidence that pathogen strain may influence infectious disease associations [84]. Now that full genome sequences of viral, bacterial and even some protozoan pathogens can be determined routinely, such an approach is becoming more feasible. Allowing for strain diversity in host genetic studies will undoubtedly increase complexity and the required sample sizes but may be particularly rewarding for studies of immunogenetic polymorphisms.

The study of the genetics of infectious disease susceptibility has undergone revolutionary change over the last decade, driven by spectacular advances in human genomics, and the rate of progress shows no sign of abating. Fascinating insights have been uncovered into the long battle between pathogens and their human host and the consequences for our genetic heritage. Although our power to understand and control major infectious pathogens is greater than ever, the true scale of the challenge posed by our need to maintain a long-term accommodation with the microbial world is only beginning to be appreciated.

I am grateful to a succession of excellent graduate students, post-doctoral scientists and fellows in my human genetics research group whose data and ideas formed the basis for this review, especially more recently Anna Rautanen, Fred Vannberg, Stephen Chapman, Tara Mills, Tom Parks, Magda Ellis, Sunny Wong, Sophie Roeynten, Chiea Khor, Branwen Hennig, Angela Framboth, Lyna Zhang, Graham Cooke, Emily Lyons, Kerrie Tosh, Ruby Siddiqui, Simon Hellier, Jordene Fitness, Christoph Aucan and Andrew Walley; to outstanding collaborators in four continents; and to our many funders including the Wellcome Trust, the European Commission, the UK MRC and the UK NIHR.

REFERENCES

1 Allison, A. C. 1954 Protection afforded by sickle-cell trait against subtertian malarial infection. Br. Med. J. 1, 290–294. (doi:10.1136/bmj.1.4857.290)

2 Huang, Y. et al. 1996 The role of a mutant CCR5 allele in HIV-1 transmission and disease progression. Nat. Med. 2, 1240–1243. (doi:10.1038/nm1196-1240)

3 Arevalo-Herrera, M. et al. 2005 Immunogenicity and protective efficacy of recombinant vaccine based on the receptor-binding domain of the Plasmodium vivax Duffy binding protein in Aotus monkeys. Am. J. Trop. Med. Hyg. 73, 25–31.

4 Miller, L. H., Mason, S. J., Clyde, D. F. & McGinniss, M. H. 1976 The resistance factor to Plasmodium vivax in blacks. The Duffy-blood-group genotype, FyFy. N. Engl. J. Med. 295, 302–304. (doi:10.1056/NEJM197608052950602)

5 Petersen, K. A., Matthiasen, F., Agger, T., Kongslev, L., Thiel, S., Cornelissen, K. & Axelsen, M. 2006 Phase I safety, tolerability, and pharmacokinetic study of recombinant human mann-binding lectin. J. Clin. Immunol. 26, 465–475. (doi:10.1007/s10875-006-9037-2)

6 McClellan, J. & King, M. C. 2010 Genetic heterogeneity in human disease. Cell 141, 210–217. (doi:10.1016/j.cell.2010.03.032)

7 Bousfiha, A. et al. 2010 Primary immunodeficiencies of protective immunity to primary infections. Clin. Immunol. 135, 204–209. (doi:10.1016/j.clim.2010.02.001)

8 Gedda, L. et al. 1984 Hereditary and infectious diseases: a twin study. Acta Genet. Med. Gemellol (Roma) 33, 497–500.

9 Comstock, G. W. 1978 Tuberculosis in twins: a re-analysis of the Prophit survey. Am. Rev. Respir. Dis. 117, 621–624.

10 Chakravarti, M. R. & Vogel, F. 1973 A twin study on leprosy. Stuttgart, Germany: Thieme.

11 Malaty, H. M., Engstrand, L., Pedersen, N. L. & Graham, D. Y. 1994 Helicobacter pylori infection: genetic and environmental influences. A study of twins. Ann. Intern. Med. 120, 982–986.

12 Lin, T. M. et al. 1989 Hepatitis B virus markers in Chinese twins. Anticancer Res. 9, 737–741.

13 Newport, M. J., Goetghueber, T., Weiss, H. A., The MRC Gambia Twin Study Group, Whittle, H., Siegrist, C.-A. & Marchant, A. 2004 Genetic regulation of immune responses to vaccines in early life. Genes Immun. 5, 122–129. (doi:10.1038/sj.geni.6364051)

14 Tan, P. L., Jacobson, R. M., Poland, G. A., Jacobsen, S. J. & Pankratz, V. S. 2001 Twin studies of immunogenetics–determining the genetic contribution to vaccine failure. Vaccine 19, 2434–2439. (doi:10.1016/S0264-410X(00)00468-0)

15 Fortin, A., Abel, L., Casanova, J. L. & Gros, P. 2007 Host genetics of mycobacterial diseases in mice and men: forward genetic studies of BCG-osis and tuberculosis. Annu. Rev. Genomics Hum. Genet. 8, 163–192. (doi:10.1146/annurev.genom.8.080706.092315)

16 Tobin, D. M. et al. 2010 The lta4h locus modulates susceptibility to mycobacterial infection in zebrafish and humans. Cell 140, 717–730. (doi:10.1016/j.cell.2010.02.013)

17 Curtis, J. et al. 2011 Association analysis of the LTA4H gene polymorphisms and pulmonary tuberculosis in 9115 subjects. Tuberculosis (Edinb.) 91, 22–25. (doi:10.1016/j.tube.2010.11.001)

18 Siddiqui, M. R. et al. 2001 A major susceptibility locus for leprosy in India maps to chromosome 10p13. Nat. Genet. 27, 439–441. (doi:10.1038/ng6958)

19 Mira, M. T. et al. 2004 Susceptibility to leprosy is associated with PARK2 and PAGCR. Nature 427, 636–640. (doi:10.1038/nature02326)

20 Tosh, K., Meisner, S., Siddiqui, M. R., Balakrishnan, K., Ghei, S., Golding, M., Sengupta, U., Pitchappan, R. M. & Hill, A. V. K. 2002 A region of chromosome 20 is linked to leprosy susceptibility in a South Indian
population. J. Infect. Dis. 186, 1190–1193. (doi:10.1086/343806)

21 Bellamy, R. et al. 2000 Genetic susceptibility to tuberculosis in Africans: a genome-wide scan. Proc. Natl Acad. Sci. USA 97, 8005–8009. (doi:10.1073/pnas.140228097)

22 Cooke, G. S. et al. 2008 Mapping of a novel susceptibility locus suggests a role for MCM3 and CTSZ in human tuberculosis. Am. J. Respir. Crit. Care Med. 178, 203–207. (doi:10.1164/rccm.200710-1554OC)

23 Frodsham, A. J. et al. 2006 Class II cytokine receptor gene cluster is a major locus for hepatitis B persistence. Proc. Natl Acad. Sci. USA 103, 9148–9153. (doi:10.1073/pnas.0602800103)

24 Abel, L. & Demenais, F. 1988 Detection of major genes for susceptibility to leprosy and its subtypes in a Caribbean island: Desirade island. Am. J. Hum. Genet. 42, 256–266.

25 Abel, L., Cot, M., Mulder, L., Carneval, P. & Feingold, J. 1992 Segregation analysis detects a major gene controlling blood infection levels in human malaria. Am. J. Hum. Genet. 50, 1308–1317.

26 Vogel, F. 1970 Controversy in human genetics. ABO blood groups and disease. Am. J. Hum. Genet. 22, 404–475.

27 Mourant, A. E. 1973 Associations between hereditary disease, bacteremia, malaria and tuberculosis. Bull. World Health Organ. 49, 93–101.

28 Hill, A. V. 2006 Aspects of genetic susceptibility to human infectious diseases. Annu. Rev. Genet. 40, 469–486. (doi:10.1146/annurev.genet.40.110405.090546)

29 Palmer, M. S., Dryden, A. J., Hughes, J. T. & Collinge, J. 1991 Homozygous prion protein genotype predisposes to sporadic Creutzfeldt–Jakob disease. Nature 352, 340–342. (doi:10.1038/352340a0)

30 Genton, B., Al-Yaman, F., Mgone, C. S., Alexander, N., Panui, M. M., Alpers, M. P. & Mokela, D. 1995 Ovalocytosis and cerebral malaria. Nature 378, 564–565. (doi:10.1038/378564a0)

31 Lindesmith, L., Moe, C., Marionneau, S., Ruvoen, N., Jiang, X., Lindblad, L., Stewart, P., LePendu, J. & Baric, R. 2003 Human susceptibility and resistance to Norwalk virus infection. Nat. Med. 9, 548–553. (doi:10.1038/nm860)

32 Zhang, F. R. et al. 2009 Genomewide association study of leprosy. N. Engl. J. Med. 361, 2092–2098. (doi:10.1056/NEJMoa0905606)

33 Pereyra, F. et al. 2010 The major determinant genotypes of HIV-1 control affect HLA class I and II peptide presentation. Science 330, 1551–1557. (doi:10.1126/science.1195271)

34 Mbarek, H. et al. 2011 A genome-wide association study of chronic hepatitis B identified novel risk loci in a Japanese population. Hum. Mol. Genet. 20, 3884–3892. (doi:10.1093/hmg/ddr301)

35 Fry, A. E. et al. 2008 Common variation in the ABO glycosyltransferase is associated with susceptibility to severe Plasmodium falciparum malaria. Hum. Mol. Genet. 17, 567–576. (doi:10.1093/hmg/ddn331)

36 Ruwende, C. et al. 1995 Natural selection of hemi- and heterozygotes for G6PD deficiency in Africa by resistance to severe malaria. Nature 376, 246–249. (doi:10.1038/376246a0)

37 Davila, S. et al. 2010 Genome-wide association study identifies variants in the CFHR region associated with host susceptibility to meningococcal disease. Nat. Genet. 42, 772–776. (doi:10.1038/ng.640)

38 Khor, C. C. et al. 2007 A Mal functional variant is associated with protection against invasive pneumococcal disease, bacteremia, malaria and tuberculosis. Nat. Genet. 39, 523–528. (doi:10.1038/ng1976)

39 Wong, S. H. et al. 2010 Leprosy and the adaptation of human toll-like receptor 1. PLoS Pathog. 6, e1000979. (doi:10.1371/journal.ppat.1000979)

40 Thomas, D. L. et al. 2009 Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. Nature 461, U752–U798. (doi:10.1038/nature08463)

41 Roy, S. et al. 2002 MBL gene type and risk of invasive pneumococcal disease: a case–control study. Lancet 359, 1569–1573. (doi:10.1016/S0140-6736(02)08581-6)

42 Vannberg, F. O., Chapman, S. J. & Hill, A. V. 2011 Human genetic susceptibility to intracellular pathogens. Immunol. Rev. 240, 105–116. (doi:10.1111/j.1600-065X.2010.00996.x)

43 Kaslow, R. A., McNicholl, J. & Hill, A. V. 2008 Genetic susceptibility to infectious diseases. Oxford, UK: Oxford University Press.

44 O’Neill, L. A. & Bowie, A. G. 2010 Sensing and signaling in antiviral innate immunity. Curr. Biol. 20, R328–R333. (doi:10.1016/j.cub.2010.01.044)

45 Khor, C. C. et al. 2010 CISH and susceptibility to infectious diseases. N. Engl. J. Med. 362, 2092–2101. (doi:10.1056/NEJMoa0905606)

46 Johnson, C. M. et al. 2007 A common polymorphism impairs cell surface trafficking and functional responses of TLR1 but protects against leprosy. J. Infect. Dis. 178, 7520–7524.

47 Ma, X., Liu, Y., Gowen, B. B., Graviss, E. A., Clark, A. G. & Musser, J. M. 2007 Full-exon resequencing reveals toll-like receptor variants contribute to human susceptibility to tuberculosis disease. PLoS ONE 2, e1318. (doi:10.1371/journal.pone.0001318)

48 Lee, Y. H., Rho, Y. H., Choi, S. J., Ji, J. D., Song, G. G., Nath, S. K. & Harley, J. B. 2006 The PTENP22 C1858T functional polymorphism and autoimmune diseases: a meta-analysis. Rheumatology (Oxford) 46, 49–56. (doi:10.1093/rheumatology/kel170)

49 Chapman, S. J. et al. 2006 PTPN22 and invasive bacterial disease. Nat. Genet. 38, 499–500. (doi:10.1038/ng0506-499)

50 Rani, R., Singh, A., Israni, N., Sharma, P. & Kar, H. K. 2009 The role of polymorphic protein tyrosine phosphatase non-receptor type 22 in leprosy. J. Invest. Dermatol. 129, 2726–2728. (doi:10.1038/jid.2009.140)

51 Chapman, S. J. et al. 2007 IkapapaB genetic polymorphisms and invasive pneumococcal disease. Am. J. Respir. Crit. Care Med. 176, 181–187. (doi:10.1164/rccm.200702-169OC)

52 Chapman, S. J. et al. 2010 NFKBIZ polymorphisms and susceptibility to pneumococcal disease in European and African populations. Genes Immun. 11, 319–325. (doi:10.1038/gene.2009.76)

53 Chapman, S. J. et al. 2010 Common NFKBIL2 polymorphisms and susceptibility to pneumococcal disease: a genetic association study. Crit. Care 14, R227. (doi:10.1186/cc9377)

54 Liu, L. et al. 2011 Gain-of-function human STAT1 mutations impair IL-17 immunity and underlie chronic mucocutaneous candidiasis. J. Exp. Med. 208, 1635–1648. (doi:10.1084/jem.20110958)

55 Wellcome Trust Case–Control Consortium. 2007 Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature 447, 661–678. (doi:10.1038/nature05911)

56 Manolio, T. A. et al. 2009 Finding the missing heritability of complex diseases. Nature 467, 747–753. (doi:10.1038/nature09494)

57 Thye, T. et al. 2010 Genome-wide association analyses identifies a susceptibility locus for tuberculosis on chromosome 18q11.2. Nat. Genet. 42, 739–741. (doi:10.1038/ng.639)
58 Jallow, M. et al. 2009 Genome-wide and fine-resolution association analysis of malaria in West Africa. Nat. Genet. 41, 657–665. (doi:10.1038/ng.388)

59 Troyer, J. L. et al. 2011 Genome-wide association study implicates PARQ3B-based AIDS restriction. J. Infect. Dis. 203, 1491–1502. (doi:10.1093/infdis/jir046)

60 Thye, T. et al. In press. Common variants at 11p13 are associated with susceptibility to tuberculosis. Nat. Genet.

61 Wong, S. H., Hill, A. V. & Vannberg, F. O. 2010 Genomewide association study of leprosy. N. Engl. J. Med. 362, 1446–1447; author reply 1447–1448. (doi:10.1056/nejmc1001451)

62 Behr, M. A. & Schurr, E. 2006 Mycobacteria in Crohn's disease: a persistent hypothesis. Inflammm. Bowel Dis. 12, 1000–1004. (doi:10.1097/mib.0000228183.70197.dd)

63 Fellay, J., Shianna, K. V., Telenti, A. & Goldstein, D. B. 2009 Common genetic variation and the control of HIV-1 in humans. PLoS Genet. 5, e1000791. (doi:10.1371/journal.pgen.1000791)

64 Fellay, J. et al. 2010 Host genetics and HIV-1: the final phase? PLoS Pathog. 6, e1001033. (doi:10.1371/journal.ppat.1001033)

65 Malaria Genomic Epidemiology Network. 2008 A global network for investigating the genomic epidemiology of malaria. Nature 456, 732–737. (doi:10.1038/nature07632)

66 Gorlov, I. P., Gorlova, O. Y., Frazier, M. L., Spitz, M. R. & Amos, C. I. 2011 Evolutionary evidence of the effect of rare variants on disease etiology. Clin. Genet. 79, 199–206. (doi:10.1111/j.1399-0004.2010.01535.x)

67 Casanova, J. L. & Abel, L. 2007 Primary immunodeficiencies: a field in its infancy. Science 317, 617–619. (doi:10.1126/science.1142963)

68 Alcais, A., Quintana-Murci, L., Thaler, D. S., Schurr, E., Abel, L. & Casanova, J.-L. 2010 Life-threatening infectious diseases of childhood: single-gene inborn errors of immunity? Ann. NY Acad. Sci. 1214, 18–33. (doi:1111/j.1749-6632.2010.05834.x)

69 Casanova, J. L. & Abel, L. 2005 Inborn errors of immunity to infection: the rule rather than the exception. J. Exp. Med. 202, 197–201. (doi:10.1084/jem.20050854)

70 Choi, M. et al. 2009 Genetic diagnosis by whole exome capture and massively parallel DNA sequencing. Proc. Natl Acad. Sci. USA 106, 19 096–19 101. (doi:10.1073/pnas.0910672106)

71 Kim, D. W., Nam, S. H., Kim, R. N., Choi, S. H. & Park, H. S. 2010 Whole human exome capture for high-throughput sequencing. Genome 53, 568–574. (doi:10.1139/G10-025)

72 Teer, J. K. & Mullikin, J. C. 2010 Exome sequencing: the sweet spot before whole genomes. Hum. Mol. Genet. 19, R145–151. (doi:10.1093/hmg/ddq333)

73 Smirnova, I., Mann, N., Dols, A., Derksen, H. H., Hibberd, M. L., Levin, M. & Beutler, B. 2003 Assay of locus-specific genetic load implicates rare Toll-like receptor 4 mutations in meningococcal susceptibility. Proc. Natl Acad. Sci. USA 100, 6075–6080. (doi:10.1073/pnas.1031605100)

74 Grossman, S. R. et al. 2010 A composite of multiple signals distinguishes causal variants in regions of positive selection. Science 327, 883–886. (doi:10.1126/science.1183863)

75 Fincham, J. R. 1972 Heterozygous advantage as a likely general basis for enzyme polymorphisms. Heredity 28, 387–391. (doi:10.1038/hdy.1972.49)

76 Hedrick, P. W. 1972 Maintenance of genetic variation with a frequency-dependent selection model as compared to the overdominant model. Genetics 72, 771–775.

77 Bodmer, W. F. 1975 Evolution of HL-A and other major histocompatibility systems. Genetics 79(Suppl.), 293–304.

78 Hill, A. V. S. 1991 HLA associations with malaria in Africa: some implications for MHC evolution. In Molecular evolution of the major histocompatibility complex (eds J. Klein & D. Klein), pp. 403–420. Berlin, Germany: Springer.

79 Sabeti, P. C. et al. 2007 Genome-wide detection and characterization of positive selection in human populations. Nature 449, 913–918. (doi:10.1038/nature06250)

80 Sanchez-Mazas, A. et al. 2011 Immunoge genetics as a tool in anthropological studies. Immunology 133, 143–164. (doi:10.1111/j.1365-2567.2011.03438.x)

81 Hill, A. V. S. et al. 1985 Melanesians and Polynesians share a unique alpha-thalassemia mutation. Am. J. Hum. Genet. 37, 571–80.

82 de Vries, R. R., Fat, R. F., Nijenhuis, L. E. & van Rood, J. J. 1976 HLA-linked genetic control of host response to Mycobacterium leprae. Lancet 2, 1328–1330. (doi:10.1016/S0140-6736(76)91975-9)

83 Monot, M. et al. 2009 Comparative genomic and phylogeographic analysis of Mycobacterium leprae. Nat. Genet. 41, 1282–1289. (doi:10.1038/ng.477)

84 Caws, M. et al. 2008 The influence of host and bacterial genotype on the development of disseminated disease with Mycobacterium tuberculosis. PLoS Pathog. 4, e1000034. (doi:10.1371/journal.ppat.1000034)