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Citation
Ceyhan-Birsoy, Ozge, Beril Talim, Lindsay C. Swanson, Mert Karakaya, Michelle A. Graff, Alan H. Beggs, and Haluk Topaloglu. 2015. “Whole Exome Sequencing Reveals DYSF, FKTN, and ISPD Mutations in Congenital Muscular Dystrophy Without Brain or Eye Involvement.” Journal of neuromuscular diseases 2 (1): 87-92. doi:10.3233/JND-140038. http://dx.doi.org/10.3233/JND-140038.

Published Version
doi:10.3233/JND-140038

Citable link
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Whole Exome Sequencing Reveals *DYSF*, *FKTN*, and *ISPD* Mutations in Congenital Muscular Dystrophy Without Brain or Eye Involvement

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Abstract

**Background**—Congenital muscular dystrophies (CMDs) are a genetically and clinically heterogeneous group of neuromuscular disorders. Several genes encoding extracellular matrix, nuclear envelope, sarcolemmal proteins and glycosylation enzymes have been implicated in CMDs. The large overlap of clinical presentations due to mutations in different genes poses a challenge for clinicians in determining disease etiology for each patient.

**Objective**—We investigated the use of whole exome sequencing (WES) in identifying the genetic cause of disease in 5 CMD patients from 3 families who presented with highly similar clinical features, including early-onset rapidly progressive weakness without brain or eye abnormalities.

**Methods**—Whole exome sequencing was performed on DNA from affected individuals. Potential functional impacts of mutations were investigated by immunostaining on available muscle biopsies.

**Results**—Pathogenic mutations in 3 different genes, *DYSF*, *FKTN*, and *ISPD* were identified in each family. Mutation in *DYSF* led to absence of dysferlin protein in patient muscle. Mutations in *ISPD* led to impaired ISDP function, as demonstrated by deficiency of α-dystroglycan glycosylation in patient muscle.
Conclusions—This study highlights the benefit of unbiased genomic approaches in molecular diagnosis of neuromuscular disorders with high clinical heterogeneity, such as the phenotypes observed in our patients. Our results suggest that dysferlin deficiency should be in the differential diagnosis of congenital and rapidly progressive muscular dystrophy, and therefore dysferlin antibody should be in the standard immunohistochemistry panel for muscle biopsies in cases with suspected CMD.

Keywords
Muscular dystrophies; DYSF protein human; Dysferlinopathy; FKTN protein human; ISPD protein human; neuromuscular diseases; genetic testing

INTRODUCTION
Congenital muscular dystrophies (CMDs) are a group of genetically heterogeneous neuromuscular disorders that present at birth or early infancy with hypotonia and muscle weakness [1]. The broad phenotypic spectrum and large overlap of clinical presentations resulting from different gene mutations pose a challenge for clinicians in making a diagnosis and determining the underlying genetic cause for each patient. Therefore, an unbiased genetic testing approach may be beneficial to determine disease etiology, guide patient clinical care and aid family counseling.

We report 5 affected children from 3 unrelated families who presented with very similar clinical features, including early-onset generalized weakness, severely elevated creatine kinase (CK) levels, and rapid progression of disease. Using whole exome sequencing (WES), we identified mutations in 3 different genes in the 3 families, including novel mutations, in the dysferlin (DYSF), fukutin (FKTN), and isoprenoid synthase domain-containing protein (ISPD) genes. Our results highlight the benefit of WES as a diagnostic tool for neuromuscular diseases with wide clinical spectrum and high genetic heterogeneity, such as the phenotype observed in our patients.

MATERIALS AND METHODS
Standard protocol approvals and patient consents
This study was approved by the institutional review board of Boston Children’s Hospital. Written informed consent was obtained from all participants.

Molecular studies
Whole exome sequencing of DNA from probands and related individuals was performed by the IDDRC Core Next-Gen Sequencing Facility of Boston Children’s Hospital and Harvard Medical School, in collaboration with Axeq Technologies. Samples were enriched for exomic sequences using the Illumina Exome Enrichment protocol and captured libraries were sequenced using Illumina HiSeq 2000 Sequencers. The reads were mapped to the human genome assembly UCSC hg19 using Burrows-Wheeler Alignment (BWA version 0.5.8, http://bio-bwa.sourceforge.net/). Single nucleotide polymorphisms and small insertions/deletions were called with SAMtools (version 0.1.7, http://
The annotated variants were filtered against variations reported on dbSNP132, the 1000 Genomes project November 2010 edition, and the NHLBI Exome Sequencing Project (http://evs.gs.washington.edu/EVS/) databases. Computational tools SIFT [2] and PolyPhen [3] were used to predict the impact of missense, while MaxEntScan [4] and NNSplice [5] were used to predict the impact of splice variants. All reported mutations were confirmed by Sanger sequencing in the patients and carrier parents.

**Immunostaining on muscle biopsies**

Muscle histology was examined using standard immunohistochemical staining. Frozen muscle sections from Patient C were fixed in ice-cold acetone for 10 minutes, followed by immunostaining with IIH6 antibody.

**RESULTS**

**Case reports**

We tested 3 Turkish families with clinical diagnoses of congenital muscular dystrophy by whole exome sequencing.

Two siblings from a consanguineous family (Family A), 14 and 11 years old, presented at 4 months and 6 months of age, respectively with hypotonia and delayed motor milestones. CK levels of the older sibling (a girl) were 728 and 4909 IU/L at 2 and 6 years of age, respectively. She became ambulant at age 2.5 years but lost the ability to walk at 8 years. She developed hip and ankle contractures gradually starting from age 3. Examination at 11 years revealed muscle power of 4/5 for neck flexors, 3+/5 for deltoids, 3+/5 for triceps, 2/5 for quadriceps, and 2/5 for hamstrings. Patients did not have intellectual disability in neurocognitive evaluation. Clinical presentation of the younger affected sibling (a boy) was relatively milder. Despite early onset of hypotonia by 6 months, he continues to walk, albeit with difficulty at age 11. Currently he cannot rise from the floor.

The proband from Family B presented at 8 months of age with hypotonia and delayed motor milestones. CK levels were 2675 IU/L at 1.5 years and 4025 IU/L at 3 years of age. He became ambulant at age 3.5 years and is currently able to walk slowly at 6 years of age. Cranial MRI was normal. Patient did not have intellectual disability in neurocognitive evaluation.

Both affected siblings from Family C presented at 4 months of age with hypotonia and delayed motor milestones. The older sibling is currently nonambulant at 5 years of age. He can barely bear weight on legs. His CK levels were 4581 and 6640 IU/L at age 1.5 and 3.5 years, respectively. Cranial MRI was normal. Patient did not have intellectual disability in neurocognitive evaluation. His younger sibling who is 13 months old now has a maximum ability to sit without support.

Major clinical and muscle biopsy findings in the 5 patients are summarized in Table 1.
Identification of mutations by WES

We used WES in the affected probands to determine the underlying genetic defects. Samples were sequenced individually. On average, 94% (range 93.3% to 94.3%) of target regions were covered, and 84.4% (range 82.8% to 86.2%) were covered more than 10X. Average median coverage of target regions was 37X (range 30X to 44X across samples). 96.4% of Ref-Seq coding exons were covered. On average, 20,223 (range 19,941 to 20,447) variants were identified in exonic regions. Variants that were absent from or were rare (minor allele frequency <0.1%) in control populations in dbSNP135, 1000 Genomes project (November 2010 edition), and NHLBI Exome Sequencing Project databases, as well as 80 race-matched control chromosomes, were filtered in 82 genes that have been implicated in neuromuscular diseases, including all congenital myopathy, muscular dystrophy, and congenital myasthenia syndrome genes in the literature. We only report variants that were novel or rare in the control populations (<0.1% minor allele frequency that would be consistent with the prevalence of congenital muscular dystrophy) and either 1) were predicted to lead to absent or truncated protein, or 2) have previously been reported in other patients with muscle disease or 3) was in a recessive gene and confirmed to be in trans with a variant meeting one of the above-mentioned criteria. Variants that were rare in the general population but did not meet these criteria were classified as uncertain significance and are not reported.

A homozygous frameshift mutation in the dysferlin gene (DYSF): c.2779delG (p.Ala927LeufsX21) that alters the protein sequence beginning from amino acid 927 and is predicted to lead to a premature termination codon 21 amino acids downstream, was identified in the patients from Family A. This variant was confirmed to be heterozygous in the parents. The same variant has previously been reported in ten individuals with hyper-CKemia who were asymptomatic until the second to third decade of life [6], three individuals with LGMD [7, 8], and two siblings with congenital muscular dystrophy who had normal or mildly elevated CK levels at ages 2 and 3 years [9]. The patients in our study had highly elevated CK levels at 2 years old. Arm muscles were preserved in patients described by Paradas et al. [9], however our patients had significant arm weakness. Interestingly, previously reported individuals carrying this variant were of Jews of the Caucasus, Iranian, and Spanish ethnicity, while the patients in our study were Turkish. Loss of function mutations of the DYSF gene are an established disease mechanism in limb girdle muscular dystrophy (LGMD) and Miyoshi myopathy, although patients with DYSF mutations typically have a later onset and milder disease compared to the patients in our study. Although other patients with earlier onset of muscle weakness have been described, our patients are unique in their presentation with highly elevated CK at an early age and involvement of all proximal muscle groups.

Two compound heterozygous missense variants were identified in the fukutin gene (FKTN): c.[915G>C];[920G>A] (p.[Trp305Cys];[Arg307Gln]) in the proband from Family B. The Arg307Gln variant has previously been reported in 2 patients with LGMD and a patient with CMD in compound heterozygous state with frameshift variants in FKTN and Trp305Cys variant has been reported in one patient with Muscle-Eye-Brain disease-Fukuyama Congenital Muscular Dystrophy phenotype [10, 11], supporting that they are pathogenic.
variants. Variants were confirmed to be in trans by parental studies. Based on these reasons, these variants were determined to be pathogenic in this patient.

In both of the two affected siblings from Family C, we identified two novel compound heterozygous mutations in ISPD: one missense variant, c.458T>C (p.Ile153Thr) and one variant within the splice consensus sequence that was predicted to lead to altered splicing, c.535-3C>G. Mutations were confirmed to be in trans by parental studies.

No other potentially pathogenic variants were identified in neuromuscular disease-related genes in the patients.

**Confirming the impact of DYSF and ISPD mutations by immunostaining on muscle biopsies**

To confirm the predicted impact of the DYSF mutation in Family A, dysferlin immunohistochemistry was performed and it revealed absence of dysferlin protein in the patient muscle (Fig. 1b).

Muscle biopsy was not available from the patient from Family B for α-dystroglycan staining and further studies.

α-Dystroglycan is hypoglycosylated in muscle biopsies from patients with ISPD mutations that cause loss of function of the protein [12, 13]. In order to test whether the ISPD mutations in Family C caused loss of function of the protein, we analyzed the glycosylation status of α-dystroglycan in the muscle biopsy using IIH6 antibody [12] (Fig. 1c). Staining with IIH6 antibody demonstrated lack of α-dystroglycan glycosylation in patient muscle, suggesting deficiency of ISPD function.

**DISCUSSION**

The five patients in our study all presented in the first year of life with rapidly progressing diffuse muscle weakness and highly elevated CK levels at an early age. In Family A, WES revealed a homozygous frameshift mutation in DYSF, a gene that is typically associated with a later onset LGMD phenotype and slower disease progression [9, 14, 15]. Reexamining the muscle biopsy in guidance of the genetic test results demonstrated that the mutation led to absence of dysferlin. The results from this case suggest that dysferlin deficiency should be in the differential diagnosis of congenital muscular dystrophy with significantly high CK at an early age. Thus, we suggest that dysferlin antibody should be used as standard in the immunohistochemistry panel for muscle biopsies in cases with suspected CMD.

We identified compound heterozygous mutations in FKTN and ISPD genes in Families B and C. FKTN and ISPD are typically associated with CMD or LGMD with variable brain and eye involvement [11, 12, 16, 17]. None of the patients in our study had cognitive or ocular abnormalities. Cranial MRI investigations were normal in all index cases. Patients from Family C with ISPD mutations presented with hypotonia and delayed motor milestones at 4 months of age, earlier than previously reported patients with ISPD mutations. α-Dystroglycan deficiencies have a diverse clinical spectrum ranging from quite mild with a rather benign LGMD phenotype to severe weakness with profound eye and brain...
abnormalities. In the vast majority of cases, clinical prediction before genetic analysis may not be accurate. The usefulness of whole exome sequencing for diagnosing various Mendelian diseases have been demonstrated [18]. Our results highlight that unbiased genomic approaches may represent a powerful diagnostic tool for neuromuscular diseases with high clinical, pathological and genetic heterogeneity, such as the phenotype observed in our patients.

Acknowledgments

We would like to thank the patients and families for their enrollment. This work was supported by a Schlumberger Foundation Faculty for the Future grant, National Institutes of Health (USA) grants R01 AR044345 and R01 HD075802, and by the Muscular Dystrophy Association (USA) grant MDA201302. DNA sequencing in the Boston Children’s Hospital IDDRC Molecular Genetics Core was supported by NIH grant P30 HD18655. O.C-B. was supported by the Dubai-Harvard Foundation for Medical Research postdoctoral fellowship.

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Fig. 1.
(a) Pedigrees of families A–C. Full black symbol, affected subjects. (b) Dysferlin immunohistochemistry in Patient A-II:1 muscle biopsy and healthy control muscle. (c) Immunostaining with IIH6 antibody for examining glycosylated α-dystroglycan in Patient C-II:1 and control muscle biopsies demonstrating absence of α-dystroglycan glycosylation in the patient muscle.
Table 1

Clinical and histopathologic features of the patients

| Patient | A-II:1 | A-II:2 | B-II:1 | C-II:1 | C-II:2 |
|---------|--------|--------|--------|--------|--------|
| Age     | 14 yrs | 11 yrs | 6 yrs  | 5 yrs  | 14 mo  |
| Sex     | Female | Male   | Male   | Male   | Male   |
| Clinical diagnosis | CMD | CMD | CMD | CMD | CMD |
| Histopathologic diagnosis | CMD | NA | CMD | CMD | NA |
| Pregnancy | Polyhydramnios | No | No | No | No |
| Fetal movements | Normal | Normal | Normal | Normal | No |
| Birth | Respiration | Normal | Normal | Normal | Normal |
| Hypotonia | No | No | No | No | No |
| Head lag | No | No | No | No | No |
| Contractures | No | No | No | No | No |
| Fractures | No | No | No | No | No |
| High arched palate | Yes | No | NA | NA | NA |
| Motor milestones | Delayed motor milestones | Yes | Yes | Yes | Yes |
| Sat unsupported | 8 mo | 9 mo | 12 mo | 13 mo | 13 mo |
| Walked unsupported | 2.5 yrs | 2.5 yrs | 3.5 yrs | No | No |
| Presentation | Age of onset | 4 mo | 6 mo | 8 mo | 4 mo |
| Presenting symptoms | Hypotonia, delayed motor milestones | Hypotonia, delayed motor milestones | Hypotonia, delayed motor milestones | Hypotonia, delayed motor milestones | Hypotonia, delayed motor milestones |
| Age of diagnosis | 2.5 yrs | 2.5 yrs | 3 yrs | 18 mo | 6 mo |
| Current abilities | Ambulation | Not ambulant since 8 yrs | Walks slowly | Ambulant since 3.5 yrs, walks slowly | Not ambulant | Not ambulant |
| Respiration | Normal | Normal | Normal | Normal | Normal |
| Feeding | Oral | Oral | Oral | Oral | Oral |
| Other health issues | Contractures | Hips and ankles (started to develop at 3 yrs) | Ankles (started to develop at 3 yrs) | No | No |
| Scoliosis | No | No | No | No | No |
| Intelligence | Normal | Normal | Normal | Normal | Normal |
| Brain MRI | Normal | NA | Normal | Normal | NA |
| Cardiac status | Normal | Normal | Normal | Normal | NA |
| Patient | A-II:1 | A-II:2 | B-II:1 | C-II:1 | C-II:2 |
|---------|-------|-------|-------|-------|-------|
| Muscle weakness | Generalized/proximal/distal | Generalized | Generalized | Generalized | Generalized |
| Facial weakness | No | No | No | No | No |
| Ophthalmoplegia | No | No | No | No | No |
| Serum CK levels (IU/L) | | | | | |
| Level 1 | 728 (2 yrs) | 3800 (3 yrs) | 2675 (1.5 yrs) | 4581 (1.5 yrs) | 2016 (9 mo) |
| Level 2 | 4909 (6 yrs) | 3747 (4 yrs) | 4025 (3 yrs) | 6640 (3.5 yrs) | NA |
| Muscle biopsy findings | | | | | |
| Internal nuclei | Yes | No muscle biopsy | NA | Yes | No muscle biopsy |
| Fiber size variation | Yes | Yes | Yes | Yes | Yes |
| Degeneration/regeneration | Yes | NA | Yes | Yes | Yes |
| Endomysial fibrosis | Yes | Yes | Yes | Yes | Yes |
| Fatty infiltration | Yes | Yes | Yes | Yes | Yes |
| IHC (merosin, dystrophin, sarcoglycans (α, β, γ, δ)) | Positive | Positive | Positive | Positive | Positive |
Table 2

Mutations identified in families A–C

| Gene Transcript | Family A                  | Family B                  | Family C                  |
|-----------------|---------------------------|---------------------------|---------------------------|
|                 | **DYSF**                  | **FKTN**                  | **ISPD**                  |
|                 | NM 003494                 | NM 001079802              | NM 001101426              |
| cDNA            | c.2779delG (Hom)          | c.915G>C (Het)            | c.458T>C (Het)            |
| Protein         | p.Ala927LeufsX21          | p.Trp305Cys               | p.Ile153Thr              |
| SIFT            | NA                        | Deleterious               | Deleterious               |
| PolyPhen        | NA                        | Probably damaging         | Probably damaging         |
| MaxEntScan      | Unlikely impact           | Unlikely impact           | Unlikely impact           |
| NNSplice        | Unlikely impact           | Unlikely impact           | Unlikely impact           |

SIFT: Deleterious, PolyPhen: Probably damaging, MaxEntScan: Unlikely impact, NNSplice: Unlikely impact

PolyPhen: Deleterious, MaxEntScan: Likely impact (wt = 10.2, var = 0.7), NNSplice: Likely impact (wt = 0.99, var = 0.5)