Functional Ingredients From *Brassicaceae* Species: Overview and Perspectives

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**Abstract:** *Brassicaceae* vegetables are important crops consumed worldwide due to their unique flavor, and for their broadly recognized functional properties, which are directly related to their phytochemical composition. Isothiocyanates (ITC) are the most characteristic compounds, considered responsible for their pungent taste. Besides ITC, these vegetables are also rich in carotenoids, phenolics, minerals, and vitamins. Consequently, Brassica’s phytochemical profile makes them an ideal natural source for improving the nutritional quality of manufactured foods. In this sense, the inclusion of functional ingredients into food matrices are of growing interest. In the present work, *Brassicaceae* ingredients, functionality, and future perspectives are reviewed.

**Keywords:** *Brassicaceae*; ingredients; phytochemicals; functionality

1. *Brassicaceae* Family: A Rich Mine of Bioactive Phytochemicals

*Brassicaceae* family vegetables have an ample worldwide distribution, which can be found in all continents except Antarctica [1–4]. One of the most striking features of this botanical family is the presence of several kinds of secondary metabolites with a distinctive taste, and also interesting bioactivities. The most deeply studied are the glucosinolates (GSL) and their breakdown products, isothiocyanates and indoles [5–7]. Moreover, these species are also rich and possess unique profiles of phenolic compounds, carotenoids, and other groups of less studied compounds such as phytoalexins, terpenes, phytosteroids, and tocopherols, here reviewed.

1.1. Phenolic Compounds

Phenolic compounds are a large class of plant secondary metabolites characterized by having at least one aromatic ring with one or more hydroxyl groups attached, showing a diversity of structures, ranging from rather simple and low molecular weight structures to complex polymeric compounds. More than 8000 phenolic compounds have been reported on plant kingdom [8]. Phenolic compounds are very important regarding the quality of plant-based foods since they are involved in flavor features (e.g., astringency), and they are responsible for the color of some fruits and vegetables and also because they serve as substrates for enzymatic deterioration [9,10]. Finally, phenolic compounds are considered to contribute to the health benefits associated with dietary consumption of *Brassicaceae* species such as antioxidant capacity, anticarcinogenic power, anti-aggregation activity, activation of...
detoxification enzymes, among others (Brassicaceae bioactive properties are reviewed and discussed in Section 3 of the present study). The most important phenolic compounds present in Brassicaceae family vegetables are the flavonoids and the hydroxyphenolic acids [8]. Among flavonoids, the most important group corresponds to the flavonols. Quercetin, kaempferol, isorhamnetin, and cyanidin are the most representative flavonols in these species, but their qualitative and quantitative profiles vary significantly among species. For example, cauliflower main phenolic compounds are quercetin aglycon and catechins, but in white cabbage, the main compound are kaempferol glucosides and epicatechins [11,12]. The phenolic profile can also vary within the same plant species according to the plant organ being studied; for example, cruciferous sprouts can contain from 2 to 10 times more phenolic compound when compared with roots and inflorescences, which are the most common plant organ consumed [13]. Recently, Fusari et al. (2019) reported for the first time important levels of trans-resveratrol—a p-terostilbene phenolic compound extensively reported as a potent antioxidant molecule—in several members of this botanical family, reaching, for example, 84 µg/g dry weight level in rocket leaves (Eruca sativa L.) [12,14].

Another group of phenolic compounds frequently detected in Brassicaceae vegetables is the hydroxyphenolic acid group, which is characterized by the C6–C3 structure and can be found free or conjugated with sugars or with other acids. The most common in this species are ferulic acid, sinapic acid, caffeic acid, and p-coumaric acid, but as occur with flavonols, this varies significantly according to the plant species considered. For example, sinapic acid is the main hydroxyphenolic acid present in rocket salad but is absent in red cherry and daikon radishes [11]. Another example of this considerable variation among species is ferulic acid, which represents the main hydroxyphenolic acid for red cabbage but is absent in radish and rocket leaves [12]. In Figure 1, the chemical structure of the phenolic compounds usually present in Brassicaceae is shown.

Anthocyanins have also been reported in Brassicaceae vegetables. For example, the red or violet pigmentation of red cabbage, purple cauliflower, or red radishes is caused by anthocyanins. The major type of anthocyanins differs among species. While red radish contains mainly cyanidin and peonidin anthocyanins acylated with aromatic acids [15], red cabbage and broccoli sprouts contain mainly cyaniding glucosides derivatives [16]. Besides, Lo Scalzo et al. (2008) [17] found that the p-coumaryl and feruloyl esterified forms of cyanidin-3-sophoroside-5-glucoside were predominant in cauliflower, while the sinapyl one was mostly present in red cabbage. In another study, Otsuki (2002) [18] found 12 different anthocyanins in radish roots, 6 of them corresponding to the pelargonidin derivatives. In red cabbage cultivars, the predominant anthocyanins resulted in being nonacylated cyanidin-glucosides [19]. Altogether, these results indicate that anthocyanins, as with other phenolic compounds, vary according to the vegetable species, the plant organ, and the cultivars of the single species considered.

1.2. Organosulfur Compounds

Among the organosulfur compounds accumulated by cruciferous vegetables, glucosinolates (GSL)—sulfur-containing glycosides—are the main secondary metabolites found. Their presence is evidenced whenever its tissue is disrupted, and their breakdown products are the principal responsible for the sharp and bitter-tasting flavors of these vegetables. There have been described in plant kingdom more than 120 different GSL, but in Brassicaceae, the amount reaches around 96, of which some are unique of some specific gender or species [20]. While genetic factors determining mainly the type of GSL, environmental factors influence the amount of them. The induction of GSL following abiotic or biotic stresses has frequently been described in order to increase the phytochemicals levels when GSL are hydrolyzed by myrosinase (thioglucoside glucohydrolase, EC 3.2.1.147) upon tissue disruption, numerous breakdown products are formed, including isothiocyanates (ITCs), thiocyanates, nitriles, ascorbigen, indoles, oxazolidine-2-thiones, and epithioalkanes depending upon different factors like pH, temperature, presence of myrosinase-interacting protein, and availability of ferrous ion [21]. Among the different GSL-degradation products formed, the most abundantly formed at
physiological pH are the ITCs, which are also considered the main responsible for cruciferous foods bioactivity. Since ITCs are very unstable, their health benefits depend on numerous variables related to several factors, such as the initial GSL concentration, cooking processes, amount of vegetable intake, and human metabolism.

![Diagram showing various phenolic compounds present in members of Brassicaceae.](image)

**Figure 1.** Phenolic compounds present in members of Brassicaceae [12].

Because the GSL profile varies between different species and the hydrolysis conditions determine the identity and amount of ITC compounds formed, it is possible to find in the literature many ITC profiles for each species, and these profiles do not always coincide with each other. ITC profiles for each cruciferous vegetable have been extensively reviewed [22–25], and the reported information highly variable according to the extraction. It is also important to keep in mind that the majority of GLS will not always give rise to its ITC profile. For example, in radish, it has consistently been reported that the majority GLS is glucoraphasatin; however, the raphasatin levels found when the ITC profile was studied, were negligible or non-quantifiable because of the extraction conditions and
the hydrolysis affecting the results, especially if the medium is polar or aqueous [26]. Furthermore, the ITC profile of each cruciferous species varies according to the cultivar or variety considered and also according to the plant tissue found [27,28]. For example, in rocket leaves, the main ITC generally detected corresponds to sativin [29], but in rocket seeds, the main ITC detected have been erucin and sulforaphane (SFN) [30]. Accordingly, in radish seeds and sprouts, the main ITC detected was sulforaphane according to several reports [13,27,28,31] (Figure 2), but in radish roots, the chief ITC have been raphasatin and sulforaphane [27,32]. In Figure 2, a graphical example of the ITC variation inside a single plant is schematized. The foregoing demonstrates that, in order to inform the ITC profile of a cruciferous vegetable, it is very important to consider, in addition to the species, the cultivation, the plant organ under study, and the extraction and detection techniques in order to make inferences and comparisons among studies.

**Figure 2.** Schematic representation of different isothiocyanate (ITC) profiles in several radish plant organs.

1.3. Carotenoids

Carotenoids are highly pigmented phytochemicals that possess a C40 backbone structure and are classified as symmetrical tetraterpenes. They are produced in many plants and microorganisms, and they can be yellow, orange, or red pigments. Some carotenoids such as β-carotene and α-carotene and β-cryptoxanthin have provitamin A activity since they act as precursors of vitamin A and, therefore, acquire an important function as a human health promoter. These compounds have been extensively studied for their health-enhancing properties and also for their biological functions as attractants to pollinators, as photoprotection pigments, and as light-harvesting pigments. Similarly, to phenolic compounds, the accumulation of carotenoids in cruciferous is regulated by the environment, tissue type, and developmental stage [33,34]. These groups of bioactive compounds have not been deeply characterized for Brassicaceae species, but it has been reported that the predominant ones are β-carotene and luteolin, but variable amounts of zeaxanthin, cryptoxanthin, neoxanthin, and violaxanthin have also been detected [35–37]. Other reports have found that carotenoid-containing cruciferous vegetables include kale (Brassica oleracea L. convar. Acephala var. sabellica), brussels sprouts, broccoli, cauliflower, red cabbage, white cabbage, pakchoi (Brassica rapa subsp. chinensis), and kohlrabi (Brassica oleracea...
var. ganglyoides) [38,39]. Kale is considered the richest cruciferous source of this compound, surpassing cabbage in about 40 times [35]. Among these, kale stands out for its high contents, not only within the cruciferous but also when compared to other vegetables, resulting in one of the main dietary source of carotenoids [35,40,41]. Kale main carotenoids have been proposed to be zeaxanthin and luteolin [40] but important differences were found among several kale cultivars [42]. Carotenoid profile among Brassicaceae species varies greatly; for example, broccoli contains mainly β-carotene and luteolin [43], cabbage contains mainly luteolin, followed by β-carotene, zeaxantin, and α-carotene [44].

1.4. Other Terpenes Present in Brassicaceae Vegetables

Other naturally occurring terpenes compounds that can be found in Brassicaceae vegetables are tocotrienols and tocopherols. According to Podsdeck (2007) [45], the descending order of total tocopherols and tocotrienol content in Brassica vegetables is as follows: broccoli > broccoli sprouts > cabbage. Furthermore, Kurilich et al. (1999) [46] reported that kale was the best source among other Brassicaceae vegetables of α-tocopherol and γ-tocopherol. Beside Brassicaceae vegetables, these compounds have also been reported in oils and cereals [47]. These phytochemicals have been extensively researched due to its anticarcinogenic properties [48,49]. Another bioactivity extensively reported in these lipid-soluble compounds is the antioxidant activity through hydrogen atom transference [50].

Phytosterols are another important terpene subclass. It has been reported to possess anti-inflammatory, anti-neoplastic, anti-pyretic, and immune-modulating activity. Also, it has been reported that phytosterols reduce serum or plasma total cholesterol and low-density lipoprotein (LDL) cholesterol [51]. Among cruciferous vegetables, Brassica napus L., known as rapeseed, is the most abundant natural source of phytosterols, reaching levels of up to 9.79 gr/kg oil [52]. Another rich source of phytosterols among cruciferous vegetables is Brassica Juncea, of which, according to the cultivar analyzed, different compositions and levels of phytosterol can be detected [53].

1.5. Phytoalexins

These groups of compounds were described initially in 1940 and are considered phenolic-related compounds with highly diverse chemical structures and several bioactivities, including anti-cancer properties. They possess low molecular weight and are thought to serve as an important defense mechanism for the plant [54,55]. Brassicaceae members containing phytoalexins present an indolic ring with C3 substitutions with N and S atoms, which confers a unique structure among other vegetables. The proposed biosynthetic pathway for phytoalexins formation includes brassinin formations from GSL; thereafter, other related phytoalexins are produced from brassinin. Klein and Sattely (2017) [55] reported that over 30 compounds arise from oxidative tailoring and rearrangement of Brassinin. Among edible cruciferous, phytoalexins have been reported in Brassica napus [56], Brassica oleracea [57], Brassica Juncea [58], Sinapsis alba [59], Wasabi japonica [60], and in Raphanus sativus [61].

1.6. Alkaloids

Alkaloids are secondary metabolites of plants synthesized from amino acids. These nitrogen compounds have been reported in several Brassicaceae species, including Capsella bursa pastoris, Lepidium cartilacinum, Nasturtium montanum, and Raphanus sativus, among others [62]. Among edible Brassicaceae alkaloid compounds have also been reported in a cabbage cultivar collection [63], in cabbage seeds by Mohammed (2014) [64], in cauliflower leaves [65], in broccoli florets [66], and red cabbage florets [67]. Besides, a screening of tropane alkaloids—a class of alkaloids typically found in Solanaceae vegetables—in 43 different Brassicaceae species reveal that 18 of them presented alkaloid of different structures, and the authors proposed that alkaloid compounds are typical secondary cruciferous metabolites [68].

As described above, the Brassicaceae family is characterized by the presence of GSL and the isothiocyanates that are exclusive of this family. Also, Brassicaceae stands out because they possess
phytochemicals of multiple chemical groups, being these species’ excellent sources of bioactive compounds that have been studied throughout history. Currently, modern analytical techniques allow us to expand the knowledge about new compounds and metabolites. Consequently, it is possible to understand that each species, each Brassica-derived product, is themselves a mixture of multiple components. For this reason, an exhaustive determination of their phytochemical profiles must be made in each case.

2. “Functional” Foods Based on Brassicas: Concepts and Relevance for Development of New Products

2.1. Origin of the “Functional Food” Concept

The origin of the concept of functional food dates to 1980 with the introduction of the concept “FOSHU” (Foods for Specified Human Health) in Japan. This tag system pretended, for first time in the world, to regulate the employment of health claims in the market [69]. From this point, the regulation of functional foods has been in constant evolution in Europe by the EFSA (European Food Safety Authority), in the USA by the FDA (United States Food and Drug Administration), or in Canada by the CFIA (Health Canada’s Food Directorate and the Canadian Food Inspection Agency), among others [70–72]. Nowadays, different unofficial definitions and concepts of functional foods coexist, being dependent of diverse factors: the country of the origin, in the case of a food product; the main characteristics of the manufacturing and the main use of this product; the specific criteria of an author, in the case of an article [73]. In this sense, a global definition can be outlined, based on specific definitions collected in recent literature [74]: A functional food must have a nutritional function that contributes to nutritional benefit on the consumer health, besides have been subjected to a technological process, in order to add a beneficial ingredient or eliminate a harmful one. In addition, it is interesting to mention the concept “nutraceutical” (classified by the European Union like “dietary supplement”) [75], a kind of functional product with a specific format similar to medicines (e.g., pills, tablets), like [76]. However, it would be a priority to clarify the limit between medicine and functional foods or nutraceuticals: it is possible to establish a mandatory common characteristic, and, without discussion, that is the preventive and non-resolving nature of functional foods against different diseases.

In general terms and due to the information exposed, it is feasible to establish the next division of this group of products:

• Functional foods with an added (or enhanced) ingredient that is associated with a health benefit. Example: milk chocolate enriched with kale [77].

• Functional foods without an ingredient (naturally present in the original product) with a health risk associated. Example: reduced fat/cholesterol mayonnaise [78]. Nutraceuticals example: microencapsulation of polyphenols extracted from red chicory and red cabbage [79].

Nowadays, consumers have introduced several modifications in their nutritional habits, due to the growing concern about health. This fact could explain the recent popularization of food products based on functional ingredients, including products based on Brassicas vegetables [80]. However, the introduction of new concepts like bioaccessibility (“digestibility”) and bioavailability has caused a strong controversy due to the questionable functionality of these food products in scientific reality [81]. In fact, the responsible regulations of health claims demand studies in vivo to allow legal commercialization, in addition to official controls in vitro that guarantees the presence of the compounds, related to the health claim of the commercial product and the capacity of the organism to use such compounds.

2.2. Functional Foods Based on Brassica Vegetables

Dietary intake of Brassicas has shown a relevant influence in the control and incidence of diverse diseases like cancer, hypertension, diabetes, chronic inflammation, or oxidative stress, among others [82].
These benefits are associated to specific compounds with active properties over human health, such as polyphenols or GSL, previously described in Section 1.

In this sense, fresh products such as broccoli or cabbage have reached a high production level nowadays, which leads to a high amount of by-products with interesting potential as biocompound sources. This valorization process is especially relevant in popular crops like broccoli, which represents 34% of total cruciferous production in the world, according to the Food and Agriculture Organization Corporate Statistical Database (FAOSTAT, 2017, www.fao.org/faostat/es/). Broccoli floret, the edible part, represents just 15% of the total vegetable, producing 85% (stems and leaves) of valuable by-products [83]. Other Brassica species by-products have been studied as sources of bioactive compounds, among them red radish or kale, among others [84].

Recent work has focused on the increase of the total quantity of bioactive compounds found in Brassica species by elicitation in order to enhance health benefits. In this way, biotic (plant hormones), and abiotic (e.g., LED light, temperature, humidity, irrigation) elicitors, have been applied to improve the quantity of polyphenols and GSL in broccoli sprouts (Brassica oleracea L. var. Italica), red radish sprouts (Raphanus sativus cv. Rambo), or chinese cabbage (Brassica rapa ssp. pekinensis), among others [85,86].

3. Functionality: What Has Been Demonstrated and What Remains to Be Study

The development of functional foods or functional ingredients is possible thanks to the interaction of three actors: (1) consumers demanding healthy foods, (2) the industrial sector motivated to elaborate and labelling their food products with a functional claim, and (3) the scientific sector which is responsible to obtain the knowledge to support those claims. Scientific substantiations of claims are performed by taking into account the totality of the available pertinent scientific data and by weighing up the evidence. To support these claims, scientific evidence on functional assessment procedures is needed, as well as toxicological evaluations and standardize analytical methodologies for functional component quantification [90]. The European Commission Regalement indicates a ranking of tests that can be done to support health statements of certain products. These tests consider, among others, whether a specific effect attributed to the product is representative in a target population, and if the quantity of the food and pattern of consumption required to obtain the claimed effect could reasonably be achieved as part of a balanced diet. In order of decreasing preference, the ‘most reliable’ would be the products that would have demonstrated their benefits in experimental trials in humans. For that reason, among all the available information on the functional properties of Brassicaceae, we consider in the first instance, those that involve the results of epidemiological studies. A bibliographic search was made with the keys “epidemiology” + “Brassicaceae” + “functional property” in the last ten years. It follows that from the spectrum of functional properties attributed to this botanical family, biological properties related to chemoprevention of cancer are the most evaluated (40%), followed very far by properties related to the prevention of cardiovascular diseases (6.6%) and antidiabetics (5.7%). These results allow us, a priori, to warn of the areas in which there is a shortage of epidemiological data.

In this regard, the extent of information supporting the benefits of consuming cruciferous vegetables is plenty and rather proven. However, when analyzing the functionality of Brassicaceae-based products, epidemiological results are scarce. The studies that can be found in this regard focus mainly on product development, and the study of in vitro properties associated with some bioactive compounds present in the final product. Alvarez-Jubete et al. (2013), for example, determined the glucosinolate and isothiocyanate contents as the antioxidant capacity of a Broccoli-based soup [91]. Other authors have evaluated different biological activities of Brassicaceae juices. Broccoli sprout juice has been studied as a potential therapeutic strategy for Alzheimer’s disease [92]. A broccoli–cabbage juice was also efficient
as a lowering LDL-cholesterol agent [93]. The antibacterial properties against food-borne bacteria of *Brassica oleracea* juice have also been addressed [94]. Furthermore, Brussels sprouts juice affects the balance of colorectal cell proliferation and death in an animal model of colorectal neoplasia [95]. Even though these findings help to give birth to the idea that certain products and ingredients have potential functionality, clinical and epidemiological studies that yield more accurate findings regarding a real health-claim are still lacking.

Unlike the above, there is plenty of epidemiological evidence indicating that cruciferous vegetable (CV) consumption has health-promoting effects on the consumers. Although *Brassicaceae* vegetables are associated with numerous biological properties, such as antioxidant, anti-inflammatory, anti-diabetic, neuroprotective, and cholesterol-lowering effects, the cancer-protective effects stand out from the other [93]. It has been proven that CV is protective against a range of cancers, GSL and their breakdown products being considered the biologically active constituents [96].

Meta-analysis are useful tools that allow statistical comparison of results among a large number of research articles that have common variables [97]. In other words, they serve to organize and simplify the findings concerning a specific subject, identifying broad trends and patterns. In this case, updated meta-analysis related to chemopreventive properties associated with CV intake were examined in order to get closer to valid health claims. Data on epidemiological studies were taken from an exhaustive review carried out using the most common and broad search engines available online up to 2010. The databases considered were SCOPUS, SCIENCE.GOV, and SCHOLAR GOOGLE. The current paper limits itself to an overview of epidemiological data on cancers with high incidence and mortality such as lung, colon, prostate, gastric, breast, and ovarian. Based on other reviews, like the one published by Van Poppel et al. (2000) [98], two types of epidemiological studies were considered. On one side, prospective cohort studies, in which diets of large groups of people are recorded, using surveys, and then, these people are controlled for disease occurrence. On the other hand, retrospective case–control studies were also considered. They are based on interviews or questionnaires to estimate dietary patterns in the near or distant past. Data from patients are then compared with data from disease-free controls.

As a result of this search in the most important data bases regarding scientific reports and articles, using as key words the terms “meta-analysis isothiocyanates”, up to 15,000 results appeared in SCHOLAR GOOGLE, but fewer results appeared in SCOPUS and SCIENCE.GOV. It must be point out that not all these results are meta-analysis per se, given that the term “analysis” is also included in the search. Therefore, most results are related to in-vitro or in vivo studies, and not precisely epidemiological studies, like the ones we are interested in. Table 1 shows the most relevant up-dated meta-analysis carried out, considering CV or ITC intake.

| Functional Property Addressed in the Meta-Analysis/Health Claim | Bioactive Compounds/Vegetables/Ingredients to Which the Bioactivity Is Attributed | Reference |
|---------------------------------------------------------------|--------------------------------------------------------------------------------|-----------|
| Chemoprevention of melanoma                                   | Isothiocyanates                                                                | [99]      |
| May reduce ovarian cancer                                     | Cruciferous vegetables                                                         | [100]     |
| Protect against colon cancer                                  | Cruciferous vegetables                                                         | [101]     |
| May decrease risk of renal cancer                             | Cruciferous vegetables                                                         | [102]     |
| Might be inversely associated with pancreatic cancer           | Cruciferous vegetables                                                         | [103]     |
| Chemoprevention activities against bladder cancer             | Isothiocyanates                                                                | [104]     |
| Inversely associated with type 2 diabetes                     | Cruciferous vegetables                                                         | [105]     |
| Inversely associated with lung cancer                         | Cruciferous vegetables                                                         | [106]     |
| Inversely associated with breast cancer                       | Glucosinolates (GSL) and isothiocyanates (ITC)                                | [107]     |
| Decreased risk of renal cell carcinoma                        | Cruciferous vegetables                                                         | [102]     |
| May reduce risk of developing lung cancer in females           | Cruciferous vegetables                                                         | [108]     |
| Decreased risk of developing colorectal and gastric cancer    | Cruciferous vegetables                                                         | [109]     |
| Chemoprevention of breast cancer                              | Sulforaphane (SFN)/Epicathechin Gallate                                        | [110]     |
| Weakly and inversely associated with lung cancer               | Cruciferous vegetables                                                         | [111]     |
Lung cancer derived from tobacco consumption is responsible for approximately 22% of cancer deaths [112]. Regarding this type of cancer, back in 2009, Lam et al. [111] started to systematize the available evidence through a meta-analysis based on epidemiological studies focusing on the potential gene–diet interaction between cruciferous vegetable intake and GSTM1 and GSTT1 (genes involved in detoxification). Back then, the evidence weakly showed an inverse association between lung cancer and CV consumption. Since then, dozens of case–control and cohort studies have been published. Liu et al. (2013) gathered this information to make an emphasis in lung cancer incidence in females, more specifically in Chinese women aged 40 to 70 years, through a prospective cohort study, which added to previous observational evidence up to 2011 [108]. In that occasion, it was observed that CV consumption might reduce the risk of lung cancer in women, especially in non-smoker ones. More recently, Zhang et al. (2018) concluded that CV intake was inversely associated with lung cancer risk. However, they recognized that current evidence is still limited and more short-term clinical Phase II and III trials are needed to elucidate further whether the inverse association reported for CV intake is due to ITC content or other bioactive compounds.

On the other side, cancers related to colorectum, gastric, oesophagus, and oropharyngeal tissues constitute one of the most common causes of death from cancer throughout the world [112]. In regards to this type of cancers, a literature review and meta-analysis carried out by Johnson et al. (2018) [109] proved that higher consumption of CV might reduce the risk of colorectal and gastric cancers by approximately 8% and 19%, respectively. These protective effects in the colorectal tissues are hypothesized to be associated with genetic polymorphism of regulating genes of the glutathione S-transferase enzyme expression; however, evidence is inconclusive on this point. That is why further epidemiological studies with accurate dietary exposure measurements need to be done. A previous study carried out by Tse et al. (2012) also suggested the value of CV intake when considering colon preventive properties. In that meta-analysis, including 33 articles, results showed a statistically significant inverse association between CV intake and colon cancer. Broccoli, particularly, stood out from other species.

Following this line of research, pancreatic cancer is responsible for 4.5% of the total number of deaths in 2018 [112]. In 2015, Li [103] showed that cruciferous plant consumption might be inversely associated with pancreatic cancer risk. Nevertheless, given that the number of studies included in this research is scarce, further evidence needs to be included in future meta-analyses.

On another note, breast cancer is the second most common cancer diagnosed in the entire world. Given this high incidence, joint efforts are being made to prevent the development of this disease, and CV intake has been proven as a chemopreventive strategy against breast cancer cells. Gianfredi et al. (2017) [110] identified sulforaphane (SFN) and epigallocatechin gallate as modulators of epigenetic events in one breast cancer cell line, therefore interfering with tumor growth rate. The following year, Zhang et al. (2018) [107] conducted a case–control study among Chinese women, proving that CV providing GLS and ITC showed a significant statistically inverse association with breast cancer risk. Despite these promising results, future prospective epidemiological studies are needed to positively assess a breast cancer prevention claim concerning CV or CV ingredient consumption. Another woman-affecting disease is ovarian cancer. In regards to this ailment, Hu et al. (2015) [100] showed that CV intake was also effective to prevent this disease.

Cancers affecting the urinary system are also affected by Brassicaceae vegetable consumption. Liu et al. (2013) [102] proved the consumption of these vegetables might decrease the risk of renal cancer, and in 2015, Veeranki and colleagues [104], concluded that ITCs have chemopreventive activities against bladder cancer.

As can be appreciated, there are cumulatively epidemiological studies suggesting that cancer rates are associated with environmental factors, more precisely in this case with diet. Numerous researches have attempted to identify the dietary agents that may inhibit the multistage process of carcinogenesis [113]. In this sense, plenty of evidence regarding naturally occurring compounds that have shown cancer-preventive effects in experimental models is available. Particularly in Brassicaceae
species, as several investigations show, isothiocyanates are the ones responsible for the chemopreventive activities [114]. There are other phytochemicals such as flavonoids present in Brassica species, that have cancer related properties; however, isothiocyanates are by far the ones with the greatest chemopreventive power [114]. Figure 3 shows the mechanism of actions during carcinogenesis process affected by ITCs.

**Figure 3.** Mechanisms of action and signalling pathways implied for inhibiting carcinogenesis by isothiocyanates.

Due to the wide range of GLS breakdown products that can be found in nature, it is not surprising that the number of cancer preventive mechanisms is so diverse. However, it must be noted that there is a dose-dependency in these responses generally; induction of cytoprotective genes and inhibition of CYP activity occurs at low compounds concentrations, whereas activation of cell cycle arrest and apoptosis occurs at higher levels of phytochemicals. Another major problem exists in interpreting experiments when using vegetable extracts because of their variable composition. Hence, uncertainty about attributing biological effects to specific phytochemicals exists. It is also important to consider the bioavailability of GLS breakdown products, which makes the interpretation of in vitro data more complex [115].

In summary, there is abundant scientific evidence that supports functional activities for the vegetables of the Brassicaceae family. However, the number of results related to Brassicaceae derived-products is lower. In agreement with point (2), each Brassicaceae-product (e.g., ingredient, phytotherapeutic, functional food) possesses a unique mixture of bioactive compounds that consequently evidences a spectrum of biological activities because of an additive/synergistic mechanism. From here arises the importance of studying each product particularly.

### 4. Food Products and Ingredients Enriched in Bioactives from Brassicaceae

The functional food market based on Brassicaceae vegetables is relatively recent. However, it contains interesting potential to offer new food products and formats with beneficial effects on health. In this sense, consumers, especially young adults and children, show some adversity in relation to the organoleptic characteristics, like the flavor of many cruciferous such as broccoli or radish [116]. For this reason, diverse new products based on functional ingredients from cruciferous families have been developed to facilitate the inclusion of this group of nutrients to the diet, into different and original formats (e.g., smoothies, soups, breads) [88]. In this way, the manufacturing of these new food
products can include the use of by-products, or side-streams, to also reach a better environmental and socio-economic balance [117]. In addition, diverse nutraceuticals like broccoli pills, tablets, or powders have been commercialized to compensate for the absence of this group of vegetables in the diet [118].

Broccoli, cabbage, and kale are predominant in the search for new functional products based on Brassica vegetables (Table 2), due to their phytochemical composition and the extensive knowledge collected in the scientific literature until this moment. The processing of food to obtain the new functional products can cause diverse changes (advantageous or disadvantageous) into the food matrix. This is crucial for the design of novel food products. In this sense, it has been seen that the addition of broccoli to baked crackers improved the nutritional properties of the final product, whilst the addition of broccoli to a juice results in important losses of sulforaphane (Table 2). On the other hand, the use of byproducts is promising, not only because of the use of non-marketable material but also for showing the advantages of using agro-waste instead of edible florets.
Table 2. Diverse experimental products based on functional ingredients from *Brassicaceae*.

| Functional Food | Nutrients and Bioactive Compounds | Effects of Processing on Bioactive Compounds | References |
|----------------|----------------------------------|--------------------------------------------|------------|
| Juice from Broccoli sprouts (*Brassica oleracea* L. var. *botrytis* subvar. *cyamosa*) | SFN and Glucoraphanin (GRA) | Less amount of SFN present than expected from GRA dosage | [88] |
| Lentil flour fortified bread with addition of kale (*Brassica oleracea* var. *sabellica*) and pea leaves | Carotenoids, Chlorophylls Flavonoid glycosides Hydroxycinnamic acid derivatives | Less quantities of carotenoids and chlorophylls. Formation of derivatives (pheophytins) Losses of flavonoid glycosides and hydroxycinnamic acid derivatives | [119] |
| Muffins enriched with dietary fiber from kimchi byproducts | Dietary fiber | Enhanced antioxidant capacity by adding kimchi fiber Decrease of color, height, and volume of the muffins Increased of hardness due to the weakening of gluten | [120] |
| Milk chocolate enriched with kale (*Brassica oleracea* var. *acephala*) and grapes | Phenolic compounds Dietary fiber Minerals | Some phenolic compounds transferred from kale or grapes to milk chocolate. However, the aniradical activity was not increased Enhanced total amount of fiber and minerals due to the addition of kale powder | [77] |
| Broccoli puree inoculated with lactic acid | Phenolic compounds GLS | Phytochemical total content was enhanced due to the fermentation of the lactic bacteria. The total content of sugar was lower than original broccoli puree | [121] |
| Broccoli puree inoculated with lactic acid | GRA SFN-nitrile | Improved stability of SFN Improved antioxidant capacity Preferential formation of SFN-nitrile (less potential as inducer of phase II detoxification enzymes than SFN) instead of SFN. | [122] |
| Broccoli soup with microalgae addition | Phenolic compounds | Improved antioxidant capacity due to the incorporation of the microalgae rich in bioaccessible phenolic compounds Preferential formation of SFN-nitrile instead of SFN. | [123] |
| Croquets with addition of red and green cabbage aqueous extract | Phenolic compounds | Improved antioxidant activity, better in croquets with green cabbage than croquets with red cabbage Organoleptic analysis indicates acceptability for consumers | [124] |
| “Kimchi” Prepared with Amtap Baechu Cabbage salted in Brine Solution | Gluconasturtin β-carotene, Pyrophyrphorbide A | Increase of the anticancer effect of “kimchi” | [125] |
| Baked crackers with addition of broccoli byproducts | Phenolic compounds Dietary fiber GLS | Improved antioxidant capacity Organoeleptic properties unaffected by elaboration | [126] |
| Puree and juice made with Broccoli by-products (powder) | Epigallocatechin gallate | Improved antioxidant capacity and anti-inflammatory activity by increased Epigallocatechin-gallate in puree Juices is not an optimal carrier of Epigallocatechin-gallate | [127] |
| Sponge cake with substitution of white flour (10% and 20%) by White Cabbage byproduct powder | Total dietary fiber | Increase of dietary fiber Decrease of total quantity of fat and carbohydrates Slight but acceptable decrease of organoleptic properties | [128] |
| Microencapsulation of polyphenols extracted from red chicory and red cabbage (nutraceutical) | Phenolic compounds | Stabilization of pH-dependent light-absorption properties of polyphenols Improvement of the thermal stability of polyphenols, mostly from red cabbage | [79] |
| Microencapsulated SFN from broccoli seed extracts (nutraceutical) | SFN | Powdered complex from Arabic gum and gelatin for encapsulating SFN from broccoli seeds. | [129] |
| Microencapsulated of Broccoli ingredient (nutraceutical) | Phenolic compounds Chlorophylls Carotenoids | Powdered complex obtained by coacervation for the stability of chlorophylls content Odor masking effects to improved acceptability for consumer | [130] |
Natural food matrix contents and bioactive compounds and grants the food the capacity to be adsorbed and metabolized by the organism [131]. However, the addition (or isolation) of some bioactive compound functional food products is not a guaranty of bioaccessibility and bioavailability, and more work is necessary to elucidate solutions against their insufficient presence in new products [132]. On the other hand, the food industry is trying to reduce the food processing to obtain products more similar to fresh-food: the concept called “minimally processed foods”. One example is broccoli hummus, which has shown promising nutritional values [133]. However, it is necessary to clearly establish the difference between fresh products and minimally processed foods to avoid the systematic and total consumption of fruits and vegetables instead of ready-to-eat fruits and salads, among other processed foods in the diet that, in addition, would result in an important impact on the environmental.

5. Food Products and Ingredients from Brassica spp.—Certain Commercialization Aspects

It is true that certain functional foods result in interesting physiological effects (Table 2), but it is necessary to stress that the novel developed “functional food” products cannot replace the nutritive qualities of fresh food [134]. Nevertheless, numerous food products are commercialized as “alternatives” for the consumption of Brassica vegetables, such as vegetable powdered formulas (broccoli https://www.bulkpowders.es/brocoli-en-polvo.html) or kale powder https://saludmediterranea.com/products/kale-col-rizada-en-polvo-salud-viva) in convenient but expensive formats (200 g >10 EUR/unit). We can ask ourselves if these are really necessary products, considering these prices. One kg of fresh broccoli (heads) is between EUR 1–2 in the supermarket, the sample place where these other products are sold in different shelves or areas of the store to highlight these added-value products. Besides the price tag, the effects of industrial processing to obtain the products (e.g., powders) could degrade the phytochemical profile (and the label of these products sometimes is difficult to understand for the consumers) and the composition on the marketed products may be far distant from the natural content of the fresh produce for a given bioactive, and therefore, much less effective.

Taking in consideration the mentioned situation, there is more interest so far in those foods that are minimally processed—once the organoleptic barrier is overpassed for the consumption of these smelly vegetable family—and ready-to-eat salads, sprouts and germinates of different cruciferous species are growing in demand and presence in the stores, even in countries that were not top consumers (such as USA or UK), as in the Mediterranean European countries [135,136], even in snack formats [137]. Therefore, there is a wide range of possibilities to exploit commercially.

When it comes to nutraceuticals or products alike, the situation is quite similar; their potential for specific population groups and pathophysiological situations where they can be used as coadjuvants is clear because of the isolation of the bioactive compounds in pharma-grade products such as pills, powders, capsules, but involving the elimination of the food matrix; in many circumstances, the isolated bioactive is not as bioavailable or metabolically active as in the natural food matrix [138].

The research in this area of functional foods and ingredients for new therapeutic applications keeps going further with the evaluation of functionalities in different chronic diseases; but many more studies will be needed to ascertain the “functionality” of these new products. For this reason, it is important to think about the population target these products are intended for: is the consumption of nutraceuticals for the general population really necessary or advisable? Will his trend lead consumers to avoid or dramatically reduce the consumption of fresh foods? If the current evidence from clinical studies and epidemiological data are still not totally clear or not totally acceptable for many of these new products, which are always much more expensive than fresh food, should we keep the wheel spinning and keep working hard on these products because of their (so far unclear) potential, or should we push for much more work from scientific research in collaboration with dietary and nutrition advice on eating a more sustainable, safe, and rich diet with plenty of “naturally functional” fresh foods (e.g., cruciferous sprouts, fresh foods enriched in bioactives) that would definitively contribute to wellbeing? Many open fronts remain waiting for answers in this global era of plants for food and health.
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Abbreviations
GSL Glucosinolates
ITC Isothiocyanates
CV Cruciferous vegetables
SFN Sulforaphane
GRA Glucoraphanin

References
1. Jiménez-Morales, P.; Sánchez-León, G.; Vargas-Rincón, C. La Producción de Metabolitos Secundarios en la Familia Brassicaceae. Rev. Fac. Ciencias Básicas 2014, 9, 282.
2. Branca, F.; Argento, S.; Alessandro, T. Assessing genetic reserves in Sicily (Italy): The Brassica wild relatives case study. In Agrobiodiversity Conservation: Securing the Diversity of Crop Wild Relatives and Landraces; Maxted, N., Ehsan Dulloo, M., Ford-Lloyd, B.V., Frese, L., Iriondo, J.M., Pinheiro de Carvalho, M.A.A., Eds.; CABI: Wallingford, Oxfordshire, UK, 2012; pp. 52–58.
3. Pinheiro de Carvalho, M.Á. Agrobiodiversity Conservation: Securing the Diversity of Crop Wild Relatives and Landraces; CABI: Wallingford, Oxfordshire, UK, 2012.
4. Branca, F.; Chiarenza, L.; Ragusa, L.; Argento, S. Morphological Characterization of the ECPGR Wild Brassica Species Collection. Acta horticulturae. Acta Hortic. 2013, 1005, 157–164. [CrossRef]
5. Argento, S.; Melilli, M.G.; Branca, F. Enhancing Greenhouse Tomato-Crop Productivity by Using Brassica macrocarpa Guss. Leaves for Controlling Root-Knot Nematodes. Agronomy 2019, 9, 820. [CrossRef]
6. Branca, F.; Lucia, R.; Alessandro, T.; Lo Scalzo, R.; Picchi, V.; Argento, S. The Glucosinolates and Variation of Antioxidant Compounds in Seeds and Sprouts of Broccoli (Brassica oleracea L. var. italica) and Rocket (Eruca sativa L.) in Relation to Temperature and Germinative Stage. Acta Hortic. 2013, 1005, 271–278. [CrossRef]
7. Galletti, S.; Bagatta, M.; Branca, F.; Argento, S.; De Nicola, G.R.; Cianchetta, S.; Iori, R.; Ninfali, P. Isatis canescens is a rich source of glucobrassicin and other health-promoting compounds. J. Sci. Food Agric. 2015, 95, 158–164. [CrossRef]
8. Cartea, M.E.; Francisco, M.; Soengas, P.; Velasco, P. Phenolic Compounds in Brassica Vegetables. Molecules 2011, 16, 251–280. [CrossRef]
9. Cheynier, V. Phenolic compounds: From plants to foods. Phytochem. Rev. 2012, 11, 153–177. [CrossRef]
10. Todaro, A.; Cavallaro, R.; Argento, S.; Branca, F.; Spagna, G. Study and Characterization of Polyphenol Oxidase from Eggplant (Solanum melongena L.). J. Agric. Food Chem. 2011, 59, 11244–11248. [CrossRef]
11. Li, Z.; Lee, H.W.; Liang, X.; Liang, D.; Wang, Q.; Huang, D.; Ong, C.N. Profiling of Phenolic Compounds and Antioxidant Activity of 12 Cruciferous Vegetables. Molecules 2018, 23, 1139. [CrossRef]
12. Fusari, C.; Beretta, H.; Locatelli, D.; Nazareno, M.; Camargo, A. Seasonal isothiocyanates variation and market availability of Brassicaceae species consumed in Mendoza. Rev. Fac. Cient. Agrar. 2019, 51, 403–408.
13. Baenas, N.; Gómez-Jodar, I.; Moreno, D.A.; García-Viguera, C.; Periago, P.M. Broccoli and radish sprouts are safe and rich in bioactive phytochemicals. Postharvest Biol. Technol. 2017, 127, 60–67. [CrossRef]
14. Loedolf, B.; Brooks, J.; Stand, M.; Peters, S.; Kossmann, J. High light bio-fortification stimulates de novo synthesis of resveratrol in Diplotaxis tenuifolia (wild rocket) micro-greens. Funct. Foods Health Dis. 2017, 7, 859–872. [CrossRef]
15. Matera, R.; Gabbanini, S.; De Nicola, G.R.; Iori, R.; Petrillo, G.; Valgimigli, L. Identification and analysis of isothiocyanates and new acylated anthocyanins in the juice of Raphanus sativus cv. Sango sprouts. Food Chem. 2012, 133, 563–572. [CrossRef] [PubMed]

16. Moreno, D.A.; Pérez-Balibrea, S.; Ferreres, F.; Gil-Izquierdo, Á.; García-Viguer, C. Acylated anthocyanins in broccoli sprouts. Food Chem. 2010, 123, 358–363. [CrossRef]

17. Lo Scalzo, R.; Genna, A.; Branca, F.; Chedin, M.; Chassaigne, H. Anthocyanin composition of cauliflower (Brassica oleracea L. var. botrytis) and cabbage (B. oleracea L. var. capitata) and its stability in relation to thermal treatments. Food Chem. 2008, 107, 136–144. [CrossRef]

18. Otsuki, T.; Matsufuji, H.; Takeda, M.; Toyoda, M.; Goda, Y. Acylated anthocyanins from red radish (Raphanus sativus L.). Phytochemistry 2002, 60, 79–87. [CrossRef]

19. Wiczkowski, W.; Szawara-Nowak, D.; Topolska, J. Red cabbage anthocyanins: Profile, isolation, identification, and antioxidant activity. Food Res. Int. 2013, 51, 303–309. [CrossRef]

20. Anjum, N.A.; Gill, S.S.; Ahmad, I.; Pacheco, M.; Duarte, A.C.; Umar, S.; Khan, N.A.; Pereira, M.E. The Plant Family Brassicaceae: An Introduction. In The Plant Family Brassicaceae: Contribution Towards Phytoremediation; Anjum, N.A., Ahmad, I., Pereira, M.E., Duarte, A.C., Umar, S., Khan, N.A., Eds.; Springer: Dordrecht, The Netherlands, 2012; pp. 1–33.

21. Bell, L.; Yahya, H.N.; Oloyede, O.O.; Methven, L.; Wagstaff, C. Changes in rocket salad phytochemicals within the commercial supply chain: Glucosinolates, isothiocyanates, amino acids and bacterial load increase significantly after processing. Food Chem. 2017, 221, 521–534. [CrossRef]

22. Traka, M.; Mithen, R.J.P.R. Glucosinolates, isothiocyanates and human health. Phytochem. Rev. 2009, 8, 269–282. [CrossRef]

23. Fahey, J.W.; Zalcman, A.T.; Talalay, P. The chemical diversity and distribution of glucosinolates and isothiocyanates among plants. Phytochemistry 2001, 56, 5–51. [CrossRef]

24. Cartea, M.E.; Velasco, P.J.P.R. Glucosinolates in Brassica foods: Bioavailability in food and significance for human health. Phytochem. Rev. 2008, 7, 213–229. [CrossRef]

25. Dinkova-Kostova, A.T.; Kostov, R.V. Glucosinolates and isothiocyanates in health and disease. Trends Mol. Med. 2012, 18, 337–347. [CrossRef]

26. Baenas, N.; Piegholdt, S.; Schloesser, A.; Moreno, D.A.; García-Viguera, C.; Rimbach, G.; Wagner, A.E. Metabolic Activity of Radish Sprouts Derived Isothiocyanates in Drosophila melanogaster. Int. J. Mol. Sci. 2016, 17, 251. [CrossRef] [PubMed]

27. Blažević, I.; Mastelić, J. Glucosinolate degradation products and other bound and free volatiles in the leaves and roots of radish (Raphanus sativus L.). Food Chem. 2009, 113, 96–102. [CrossRef]

28. Yi, G.; Lim, S.; Chae, W.B.; Park, J.E.; Park, H.R.; Lee, E.J.; Huh, J.H. Root Glucosinolate Profiles for Screening of Radish (Raphanus sativus L.) Genetic Resources. J. Agric. Food Chem. 2016, 64, 61–70. [CrossRef] [PubMed]

29. Fechner, J.; Kaufmann, M.; Herz, C.; Eisenschmidt, D.; Lamy, E.; Kroh, L.W.; Hanschen, F.S. The major glucosinolate hydrolysis product in rocket (Eruca sativa L.), sativin, is 1,3-thiazepane-2-thione: Elucidation of structure, bioactivity, and stability compared to other rocket isothiocyanates. Food Chem. 2018, 261, 57–65. [CrossRef]

30. Franco, P.; Spinozzi, S.; Pagnotta, E.; Lazzari, L.; Ugolini, L.; Camborata, C.; Roda, A. Development of a liquid chromatography–electrospray ionization–tandem mass spectrometry method for the simultaneous analysis of intact glucosinolates and isothiocyanates in Brassicaceae seeds and functional foods. J. Chromatogr. A 2016, 1428, 154–161. [CrossRef]

31. Kuang, P.; Song, D.; Yuan, Q.; Yi, R.; Lv, X.; Liang, H. Separation and purification of sulforaphene from radish seeds using macroporous resin and preparative high-performance liquid chromatography. Food Chem. 2013, 136, 342–347. [CrossRef]

32. Kim, J.-W.; Kim, M.-B.; Lim, S.-B. Formation and Stabilization of Raphasatin and Sulforaphene from Radish Roots by Endogenous Enzymolysis. Prev. Nutr. Food Sci. 2015, 20, 119–125. [CrossRef]

33. Yu, B.; Lydiate, D.J.; Young, L.W.; Schäfer, U.A.; Hannoufa, A.J.T.R. Enhancing the carotenoid content of Brassica napus seeds by downregulating lycopene epsilon cyclase. Transgenic Res. 2008, 17, 573–585. [CrossRef]

34. Frede, K.; Schreiner, M.; Baldermann, S. Light quality-induced changes of carotenoid composition in pak choi Brassica rapa ssp. chinensis. J. Photochem. Photobiol. B Biol. 2019, 193, 18–30. [CrossRef] [PubMed]
35. Nilsson, J.; Olsson, K.; Engqvist, G.; Ekvall, J.; Olsson, M.; Nyman, M.; Åkesson, B. Variation in the content of glucosinolates, hydroxycinnamic acids, carotenoids, total antioxidant capacity and low-molecular-weight carbohydrates in *Brassica* vegetables. *J. Sci. Food Agric.* 2006, 86, 528–538. [CrossRef]

36. Zhang, B.; Liu, C.; Wang, Y.; Yao, X.; Wang, F.; Wu, J.; King, G.J.; Liu, K. Disruption of a Carotenoid Cleavage Dioxygenase 4 gene converts flower colour from white to yellow in *Brassica* species. *New Phytol.* 2015, 206, 1513–1526. [CrossRef] [PubMed]

37. Guzman, I.; Yousef, G.G.; Brown, A.F. Simultaneous Extraction and Quantitation of Carotenoids, Chlorophylls, and Tocopherols in *Brassica* Vegetables. *J. Agric. Food Chem.* 2012, 60, 7238–7244. [CrossRef] [PubMed]

38. Park, W.T.; Kim, J.K.; Park, S.; Lee, S.-W.; Li, X.; Kim, Y.B.; Uddin, M.R.; Park, N.I.; Kim, S.-J.; Park, S.U. Metabolic Profiling of Glucosinolates, Anthocyanins, Carotenoids, and Other Secondary Metabolites in Kohlrabi (*Brassica oleracea* var. gongylodes). *J. Agric. Food Chem.* 2012, 60, 8111–8116. [CrossRef] [PubMed]

39. Chun, J.-H.; Kim, N.-H.; Seo, M.-S.; Jin, M.; Park, S.U.; Arasu, M.V.; Kim, S.-J.; Al-Dhabi, N.A. Molecular characterization of glucosinolates and carotenoid biosynthetic genes in Chinese cabbage (*Brassica rapa* L. ssp. *pekinesis*). *Saud J. Biol. Sci.* 2018, 25, 71–82. [CrossRef]

40. Walsh, R.P.; Bartlett, H.; Eperjesi, F. Variation in Carotenoid Content of Kale and Other Vegetables: A Review of Pre- and Post-harvest Effects. *J. Agric. Food Chem.* 2015, 63, 9677–9682. [CrossRef]

41. Abdel-Aal, E.-S.M.; Akhtar, H.; Zaheer, K.; Ali, R. Dietary Sources of Lutein and Zeaxanthin Carotenoids and Their Role in Eye Health. *Nutr. Nour.* 2013, 5, 1169–1185. [CrossRef]

42. Mageney, V.; Baldermann, S.; Albach, D.C. Intraspecific Variation in Carotenoids of *Brassica oleracea* var. *sabellica*. *J. Agric. Food Chem.* 2016, 64, 3251–3257. [CrossRef]

43. Fernández-León, M.F.; Fernández-León, A.M.; Lozano, M.; Ayuso, M.C.; González-Gómez, D. Identification, quantification and comparison of the principal bioactive compounds and external quality parameters of two broccoli cultivars. *J. Food Foods Biol.* 2012, 4, 465–473. [CrossRef]

44. Kaulmann, A.; André, C.M.; Schneider, Y.-J.; Hoffmann, L.; Bohn, T. Carotenoid and polyphenol bioaccessibility and cellular uptake from plum and cabbage varieties. *Food Chem.* 2016, 197, 325–332. [CrossRef] [PubMed]

45. Podsędek, A. Natural antioxidants and antioxidant capacity of *Brassica* vegetables: A review. *UWT—Food Sci. Technol.* 2007, 40, 1–11. [CrossRef]

46. Kurilich, A.C.; Tsau, G.J.; Brown, A.; Howard, L.; Klein, B.P.; Jeffery, E.H.; Kushad, M.; Wallig, M.A.; Juvik, J.A. Carotene, Tocopherol, and Ascorbate Contents in Subspecies of *Brassica oleracea*. *J. Agric. Food Chem.* 1999, 47, 1576–1581. [CrossRef] [PubMed]

47. Dan, L.; Yi, Y.; Yongxin, L.; Chengjun, S. Analysis of Tocopherols and Tocotrienols in Pharmaceuticals and Foods: A Critical Review. *Curr. Pharm. Anal.* 2015, 11, 66–78. [CrossRef]

48. Yu, W.; Simmons-Menchaca, M.; Gapor, A.; Sanders, B.G.; Kline, K. Induction of apoptosis in human breast cancer cells by tocopherols and tocotrienols. *Nutr. Cancer* 1999, 33, 26–32. [CrossRef]

49. Mo, H.; Elson, C.E. Apoptosis and Cell-Cycle Arrest in Human and Murine Tumor Cells Are Initiated by Isoprenoids. *J. Nutr. 1999*, 129, 804–813. [CrossRef]

50. Lampi, A.-M.; Kamal-Eldin, A.; Piironen, V. Tocopherols and tocotrienols from oil and cereal grains. *Funct. Foods Biochem. Process. Asp.* 2002, 1–38.

51. Ling, W.H.; Jones, P.J.H. Dietary phytosterols: A review of metabolism, benefits and side effects. *Life Sci.* 1995, 57, 195–206. [CrossRef]

52. Gül, M.; Amar, S. Sterols and the phytosterol content in oilseed rape (*Brassica napus* L.). *J. Cell & Mol. Biol.* 2006, 5, 71–79.

53. Sharma, A.; Rai, P.K.; Prasad, S. GC–MS detection and determination of major volatile compounds in *Brassica juncea* L. leaves and seeds. *Microchem. J.* 2018, 138, 488–493. [CrossRef]

54. Ahuja, I.; Kissen, R.; Bones, A.M. Phytoalexins in defense against pathogens. *Trends Plant Sci.* 2012, 17, 73–90. [CrossRef] [PubMed]

55. Klein, A.P.; Sattely, E.S. Biosynthesis of cabbage phytoalexins from indole glucosinolate. *Proc. Natl. Acad. Sci. USA* 2017, 114, 1910–1915. [CrossRef] [PubMed]

56. Pedras, M.S.C.; Zheng, Q.-A.; Gadagi, R.S.; Rimmer, S.R. Phytoalexins and polar metabolites from the oilseeds canola and rapeseed: Differential metabolic responses to the biotroph Albugo candida and to abiotic stress. *Phytochemistry* 2008, 69, 894–910. [CrossRef] [PubMed]
68. Brock, A.; Herzfeld, T.; Paschke, R.; Koch, M.; Dräger, B. Brassicaceae contain nortropane alkaloids.

57. Pedras, M.S.C.; Adio, A.M.; Suchy, M.; Okinyo, D.P.O.; Zheng, Q.-A.; Jha, M.; Sarwar, M.G. Detection, characterization and identification of crucifer phytoalexins using high-performance liquid chromatography with diode array detection and electrospray ionization mass spectrometry. J. Chromatogr. A 2006, 1133, 172–183. [CrossRef] [PubMed]

58. Pedras, M.S.C.; Ahiahonu, P.W.K. Metabolism and detoxification of phytoalexins and analogs by phytopathogenic fungi. Phytochemistry 2005, 66, 391–411. [CrossRef]

59. Pedras, M.S.C.; Montaut, S.; Zaharia, I.L.; Gai, Y.; Ward, D.E. Transformation of the host-selective toxin destruxin B by wild crucifers: Probing a detoxification pathway. Phytochemistry 2003, 64, 957–963. [CrossRef]

60. Pedras, M.S.C.; Sorensen, J.L.; Okanga, F.I.; Zaharia, I.L. Wasalexins A and B, new phytoalexins from wasabi: Isolation, synthesis, and antifungal activity. Bioorganic Med. Chem. Lett. 1999, 9, 3015–3020. [CrossRef]

59. Pedras, M.S.C.; Montaut, S.; Zaharia, I.L.; Gai, Y.; Ward, D.E. Transformation of the host-selective toxin destruxin B by wild crucifers: Probing a detoxification pathway. Phytochemistry 2003, 64, 957–963. [CrossRef]

61. Monde, K.; Takasugi, M.; Shirata, A. Three sulphur-containing stress metabolites from Japanese radish. Phytochemistry 1995, 39, 581–586. [CrossRef]

62. Nee, M. Plant Alkaloids: A Guide to Their Discovery and Distribution; Raffauf, R.F., Ed.; Brittonia: New York, NY, USA, 1998; Volume 50, p. 35.

63. Guriya, R.; Moon, A.; Talreja, K. Phytochemical profiling and characterization of bioactive compounds from Brassica oleracea. Int. J. Pharmaceut. Phytochem. Res. 2015, 7, 825–831.

64. Khalid, A.; Mohammed, A.D.; Al-Maliki, M. E destruxin B by wild crucifers: Probing a detoxification pathway. Phytochemistry 1999, 50, 55. [CrossRef]

65. Villaño, D.; Giron-Vilapana, A.; García-Viguera, C.; Moreno, D.A. Development of Functional Foods. In In Innovation Strategies in the Food Industry; Galanakis, C.M., Ed.; Academic Press: Cambridge, MA, USA, 2016; Chapter 10; pp. 191–210. [CrossRef]

66. Shah, M.A.; Sarker, M.M.R.; Gousuddin, M. Antidiabetic Potential of Brassica oleracea var. italica seed on glucose level in blood of alloxan-induced diabetes rabbits. World J. Exp. Biosci. 2014, 2, 24–29.

67. Chauhan, E.S.T.; Tiwari, A.; Singh, A. Phytochemical screening of red cabbage (Brassica oleracea) powder and juice—A comparative study. J. Med. Plants Stud. 2016, 4, 196–199.

68. Brock, A.; Herzfeld, T.; Paschke, R.; Koch, M.; Dräger, B. Brassicaceae contain nortropane alkaloids. Phytochemistry 2006, 67, 2050–2057. [CrossRef] [PubMed]

69. Villaño, D.; Giron-Vilapana, A.; García-Viguera, C.; Moreno, D.A. Development of Functional Foods. In In Innovation Strategies in the Food Industry; Galanakis, C.M., Ed.; Academic Press: Cambridge, MA, USA, 2016; Chapter 10; pp. 191–210. [CrossRef]

70. Serafini, M.; Stanzione, A.; Foddai, S. Functional foods: Traditional use and European legislation. Int. J. Food Sci. Nutr. 2012, 63, 7–9. [CrossRef]

71. Bagchi, D. Nutraceutical and Functional Food Regulations in the United States and Around the World. In Nutraceutical and Functional Food Regulations in the United States and Around the World; Bagchi, D., Ed.; Academic Press: San Diego, CA, USA, 2008; pp. ix–xii. [CrossRef]

72. Powers, J.-P.; Farrell, M.; McMullin, C.; Retik, L.; White, J. Regulation of dietary supplements and functional foods in Canada. In Polyphenols: Properties, Recovery and Applications; Alven-Soled, P., Ed.; Elsevier: Amsterdam, The Netherlands, 2018; pp. 327–362.

73. Baenas, N.; Abellán, A.; Rivera, S.; Moreno, D.A.; García-Viguera, C.; Domínguez-Perles, R. Foods and Suplements. In Polyphenols: Properties, Recovery and Applications; Elsevier: Amsterdam, The Netherlands, 2018; pp. 327–362.

74. Bigiardi, B.; Galati, F Innovation trends in the food industry: The case of functional foods. Trends Food Sci. Technol. 2013, 31, 118–129. [CrossRef]

75. Poli, A.; Barbagallo, C.M.; Cicero, A.F.G.; Corsini, A.; Manzato, E.; Trimarco, B.; Bernini, F.; Visioli, F.; Bianchi, A.; Canzone, G.; et al. Nutraceuticals and functional foods for the control of plasma cholesterol levels. An intersociety position paper. Pharmacol. Res. 2018, 134, 51–60. [CrossRef]

76. Fuentes-Alventosa, J.M. Caracterización de Componentes Bioactivos del Espárrago Verde: Obtención de Ingredientes Funcionales a Partir de Los Subproductos Generados Durante su Transformación Industrial; University of Cordoba: Cordoba, Spain, 2010.

77. Carvalho, J.C.S.; Romoff, P.; Lannes, S.C.d.S. Improvement of nutritional and physicochemical proprieties of milk chocolates enriched with kale (Brassica oleracea var. acephala) and grape (Vitis vinifera). Food Sci. Technol. 2018, 38, 551–560. [CrossRef]

78. Ouraji, M.; Alimi, M.; Motamedzadehan, A.; Shokoohi, S. Faba bean protein in reduced fat/cholesterol mayonnaise: Extraction and physico-chemical modification process. J. Food Sci. Technol. 2020. [CrossRef]
97. Herron, J.D.; Nurrenbern, S.C. Chemical Education Research: Improving Chemistry Learning. *J. Chem. Educ.* 1999, 76, 1353. [CrossRef]

98. Van Poppel, G.; Verhagen, D.T.; Verhagen, H.; Goldbohm, R.A. Brassica vegetables and cancer prevention: Epidemiology and mechanisms. *Adv. Exp. Med. Biol.* 2000, 472, 159–168.
99. Mitsiogianni, M.; Koutsidis, G.; Mavroudis, N.; Trafalis, D.T.; Botaitis, S.; Franco, R.; Zoumpourlis, V.; Amery, T.; Galanis, A.; Pappa, A.; et al. The Role of Isothiocyanates as Cancer Chemo-Preventive, Chemotherapeutic and Anti-Melanoma Agents. *Antioxidants* **2019**, *8*, 106. [CrossRef]

100. Hu, J.; Hu, Y.; Hu, Y.; Zheng, S. Intake of cruciferous vegetables is associated with reduced risk of ovarian cancer: A meta-analysis. *Asia Pac. J. Clin. Nutr.* **2015**, *14*, 201–109.

101. Tse, G.; Eslick, G.D. Cruciferous Vegetables and Risk of Colorectal Neoplasms: A Systematic Review and Meta-Analysis. *Nutr. Cancer* **2014**, *66*, 128–139. [CrossRef] [PubMed]

102. Liu, B.; Mao, Q.; Wang, X.; Zhou, F.; Luo, J.; Wang, C.; Lin, Y.; Zheng, X.; Xie, L. Cruciferous Vegetables Consumption and Risk of Renal Cell Carcinoma: A Meta-Analysis. *Nutr. Cancer* **2013**, *65*, 668–676. [CrossRef] [PubMed]

103. Li, L.Y.; Luo, Y.; Lu, M.D.; Xu, X.W.; Lin, H.D.; Zheng, Z.Q. Cruciferous vegetable consumption and the risk of pancreatic cancer: A meta-analysis. *World J. Surg. Oncol.* **2015**, *13*, 44. [CrossRef] [PubMed]

104. Abbaoui, B.; Lucas, C.R.; Riedl, K.M.; Clinton, S.K.; Mortazavi, A. Cruciferous Vegetables, Isothiocyanates, and Bladder Cancer Prevention. *Mol. Nutr. Food Res.* **2018**, *62*, e1800079. [CrossRef]

105. Jia, X.; Zhong, L.; Song, Y.; Hu, Y.; Wang, G.; Sun, S. Consumption of citrus and cruciferous vegetables with incident type 2 diabetes mellitus based on a meta-analysis of prospective study. *Prim. Care Diabetes* **2016**, *10*, 272–280. [CrossRef]

106. Zhang, Z.; Bergan, R.; Shannon, J.; Slatore, C.G.; Bobe, G.; Takata, Y. The Role of Cruciferous Vegetables and Isothiocyanates for Lung Cancer Prevention: Current Status, Challenges, and Future Research Directions. *Mol. Nutr. Food Res.* **2018**, *62*, 1700936. [CrossRef]

107. Zhang, N.-Q.; Ho, S.C.; Mo, X.-F.; Lin, F.-Y.; Huang, W.-Q.; Luo, H.; Huang, J.; Zhang, C.-X. Glucosinolate and isothiocyanate intakes are inversely associated with breast cancer risk: A case–control study in China. *Br. J. Nutr.* **2018**, *119*, 957–964. [CrossRef]

108. Wu, Q.J.; Xie, L.; Zheng, W.; Vogtmann, E.; Li, H.L.; Yang, G.; Ji, B.T.; Gao, Y.T.; Shu, X.O.; Xiang, Y.B. Cruciferous vegetables consumption and the risk of female lung cancer: A prospective study and a meta-analysis. *Ann. Oncol.* **2013**, *24*, 1918–1924. [CrossRef]

109. Johnson, I.T. Cruciferous Vegetables and Risk of Cancers of the Gastrointestinal Tract. *Mol. Nutr. Food Res.* **2018**, *62*, 1701000. [CrossRef]

110. Gianfredi, V.; Vannini, S.; Moretti, M.; Villarini, M.; Bragazzi, N.L.; Izzotti, A.; Nucci, D. Sulforaphane and Epigallocatechin Gallate Restore Estrogen Receptor Expression by Modulating Epigenetic Events in the Breast Cancer Cell Line MDA-MB-231: A Systematic Review and Meta-Analysis. *Lifestyle Genom.* **2017**, *10*, 126–135. [CrossRef]

111. Lam, T.K.; Gallicchio, L.; Shiels, M.; Hammond, E.; Tao, X.G.; Chen, L.; Robinson, K.A.; Caulfield, L.E.; Herman, J.G.; et al. Cruciferous vegetable consumption and lung cancer risk: a systematic review. *Cancer Epidemiol. Biomark. Prev.* **2009**, *18*, 184–195. [CrossRef] [PubMed]

112. International Agency for Research on Cancer (IARC); World Health Organization. *All Cancers Data Sheet*; International Agency for Research on Cancer: Lyon, France, 2018; Volume 876.

113. Murillo, G.; Mehta, R.G. Cruciferous vegetables and cancer prevention. *Nutr. Cancer* **2014**, *61*, 17–28. [CrossRef] [PubMed]

114. Drabinska, N.; Ciska, E.; Szmotowicz, B.; Krupa-Kozak, U. Broccoli by-products improve the nutraceutical potential of gluten-free mini sponge cakes. *Food Chem.* **2018**, *267*, 170–177. [CrossRef]
119. Klopsch, R.; Baldermann, S.; Voss, A.; Rohn, S.; Schreiner, M.; Neugart, S. Narrow-Banded UVB Affects the Stability of Secondary Plant Metabolites in Kale (Brassica oleracea var. sabellica) and Pea (Pisum sativum) Leaves Being Added to Lentil Flour Fortified Bread: A Novel Approach for Producing Functional Foods. *Foods* **2019**, *8*, 427. [CrossRef]

120. Heo, Y.; Kim, M.-J.; Lee, J.-W.; Moon, B. Muffins enriched with dietary fiber from kimchi by-product: Baking properties, physical–chemical properties, and consumer acceptance. *Food Sci. Nutr.* **2019**, *7*, 1778–1785. [CrossRef]

121. Ye, J.-H.; Huang, L.-Y.; Terefe, N.S.; Augustin, M.A. Fermentation-based biotransformation of glucosinolates, phenolics and sugars in retorted broccoli puree by lactic acid bacteria. *Food Chem.* **2019**, *286*, 616–623. [CrossRef] [PubMed]

122. Cai, Y.X.; Wang, J.H.; McAuley, C.; Augustin, M.A.; Terefe, N.S. Fermentation for enhancing the bioconversion of glucoraphanin into sulforaphane and improve the functional attributes of broccoli puree. *J. Funct. Foods* **2019**, *61*, 103461. [CrossRef]

123. Lafarga, T.; Acién-Fernández, F.G.; Castellari, M.; Villaró, S.; Bobo, G.; Aguiló-Aguayo, I. Effect of microalgae incorporation on the physicochemical, nutritional, and sensorial properties of an innovative broccoli soup. *LWT* **2019**, *111*, 167–174. [CrossRef]

124. Ashfaq, F.; Butt, M.S.; Bilal, A.; Tehseen, S.; Suleria, H.A.R. Effect of cabbage or its aqueous extract incorporated croquettes on chemical composition and storage stability in relation to antioxidant potential and sensory profile. *J. Food Process. Preserv.* **2020**, *44*, e14291. [CrossRef]

125. Song, G.-H.; Park, E.-S.; Lee, S.-M.; Park, D.-B.; Park, K.-Y. Beneficial Outcomes of Kimchi Prepared with Amtak Baechu Cabbage and Salting in Brine Solution: Anticancer Effects in Pancreatic and Hepatic Cancer Cells. *J. Environ. Pathol. Toxicol. Oncol.* **2018**, *37*, 151–161. [CrossRef] [PubMed]

126. Lafarga, T.; Gallagher, E.; Bademunt, A.; Bobo, G.; Echeverria, G.; Viñas, I.; Aguiló-Aguayo, I. Physicochemical and nutritional characteristics, bioaccessibility and sensory acceptance of baked crackers containing broccoli co-products. *Int. J. Food Sci. Technol.* **2019**, *54*, 634–640. [CrossRef]

127. Shi, M.; Ying, D.-Y.; Hlaing, M.M.; Ye, J.-H.; Sanguansri, L.; Augustin, M.A. Development of broccoli by-products as carriers for delivering EGCG. *Food Chem.* **2019**, *301*, 125301. [CrossRef]

128. Prokopov, T.; Goranova, Z.; Baeva, M.; Slavov, A.; Galanakis, C.M. Effects of powder from white cabbage outer leaves on sponge cake quality. *Int. Agrophys.* **2015**, *29*, 493–500. [CrossRef]

129. García-Saldaña, J.S.; Campas-Baypoli, O.N.; López-Cervantes, J.; Sánchez-Machado, D.I.; Cantú-Soto, E.U.; Rodríguez-Ramírez, R. Microencapsulation of sulforaphane from broccoli seed extracts by gelatin/gum arabic and gelatin/pectin complexes. *Food Chem.* **2016**, *201*, 94–100. [CrossRef]

130. Sánchez, F.M.; García, F.; Calvo, P.; Bernalte, M.J.; González-Gómez, D. Optimization of broccoli microencapsulation process by complex coacervation using response surface methodology. *Innov. Food Sci. Emerg. Technol.* **2016**, *34*, 243–249. [CrossRef]

131. Soler-Rivas, C.; Marín, F.R.; Santoyo, S.; García-Risco, M.R.; Señoráns, F.J.; Reglero, G. Testing and Enhancing the In Vitro Bioaccessibility and Bioavailability of Rosmarinus officinalis Extracts with a High Level of Antioxidant Abietanes. *J. Agric. Food Chem.* **2010**, *58*, 1144–1152. [CrossRef] [PubMed]

132. Riar, H.; Khatkar, S.; Khatkar, À.; Arora, N.; Mann, S.; Panghal, A.; Kumar, S. The conceptual understanding of nutrigenomics: A futuristic approach for designing health foods. *Nutr. Food Sci.* **2019**. [CrossRef]

133. Klug, T.V.; Martínez-Hernández, G.B.; Collado, E.; Artés, F.; Artés-Hernández, F. Effect of Microwave and High-Pressure Processing on Quality of an Innovative Broccoli Hummus. *Food Bioprocess Technol.* **2018**, *11*, 1464–1477. [CrossRef]

134. Valenzuela, B.A.; Valenzuela, R.; Sanhueza, J.; Morales, I.G. Alimentos funcionales, nutraceúticos y foshu: ¿Vamos hacia un nuevo concepto de alimentación? Revista chilena de nutrición. *Rev. Chil. Nutr.* **2014**, *41*, 198–204. [CrossRef]

135. Cai, Y.X.; Augustin, M.A.; Jegasothy, H.; Wang, J.H.; Terefe, N.S. Mild heat combined with lactic acid fermentation: A novel approach for enhancing sulforaphane yield in broccoli puree. *Food Funct.* **2020**, *11*, 779–786. [CrossRef] [PubMed]

136. Mazzucotelli, C.A.; González-Aguilar, G.A.; Villegas-Ochoa, M.A.; Domínguez-Avila, A.J.; Ansorena, M.R.; Di Scala, K.C. Chemical characterization and functional properties of selected leafy vegetables for innovative mixed salads. *J. Food Biochem.* **2018**, *42*, e12461. [CrossRef]
137. Barakat, H.; Reim, V.; Rohn, S. Stability of saponins from chickpea, soy and faba beans in vegetarian, broccoli-based bars subjected to different cooking techniques. *Food Res. Int.* **2015**, *76*, 142–149. [CrossRef]

138. Atwell, L.L.; Hsu, A.; Wong, C.P.; Stevens, J.F.; Bella, D.; Yu, T.-W.; Pereira, C.B.; Löhr, C.V.; Christensen, J.M.; Dashwood, R.H.; et al. Absorption and chemopreventive targets of sulforaphane in humans following consumption of broccoli sprouts or a myrosinase-treated broccoli sprout extract. *Mol. Nutr. Food Res.* **2015**, *59*, 424–433. [CrossRef]

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