In late December 2018, an outbreak of listeriosis occurred after a group of 32 individuals celebrated in a tavern in Styria, Austria; traditional Austrian food (e.g., meat, meat products and cheese) was served. After the celebration, 11 individuals developed gastrointestinal symptoms, including one case with severe sepsis. Cases had consumed mixed platters with several meat products and pâtés originating from a local production facility (company X). Human, food and environmental samples taken from the tavern and company X were tested for *L. monocytogenes*. Whole genome sequence-based typing detected a novel *L. monocytogenes* strain of serotype IVb, sequence type 4 and CT7652 in 15 samples; 12 human, two food and one environmental sample from company X with an allelic difference of 0 to 1. Active case finding identified two further cases who had not visited the tavern but tested positive for the outbreak strain. In total, 13 cases (seven females and six males; age range: 4–84 years) were identified. Liver pâté produced by company X was identified as the likely source of the outbreak. Control measures were implemented and since the end of December 2018, no more cases were detected.

**Background**

*Listeria monocytogenes* is a Gram-positive bacterium that is typically transmitted to humans through the consumption of contaminated food products. Clinical symptoms of listeriosis vary depending on the immune status of the host, with those immunocompromised at higher risk of presenting severe symptoms [1]. In immunocompetent individuals, infection can be asymptomatic but it more often results in febrile gastroenteritis [2]. Febrile gastroenteritis usually resolves within 2–3 days after the onset of symptoms, while invasive forms of the disease can lead to meningoencephalitis, abortion, sepsis or even death [3,4]; other manifestations such as endophthalmitis have also been associated with infections with *L. monocytogenes* [1,5]. Foods that have been implicated with listeriosis outbreaks are ready-to-eat (RTE) products such as sliced meat, pâté and soft cheese varieties [6]. Since *L. monocytogenes* can persist in the environment for long periods due to its ability to form biofilms and its resistance to disinfectants, this pathogen can be difficult to eradicate from food-processing facilities [7].

In Austria, notification of invasive listeriosis cases is mandatory. The Austrian National Reference Laboratory for Listeria (NRL; Graz Austria) is responsible for performing whole genome sequence (WGS)-based typing of human and non-human (e.g., food, environmental) isolates. In recent years, WGS-based surveillance of *L. monocytogenes* has been successfully used in combination with analysis of epidemiological data in outbreak investigations [8-10].

**Outbreak detection**

On 21 December 2018, the local health authority of Styria (Directorate of Public Health, Graz, Austria) and the NRL confirmed the occurrence of an outbreak of febrile gastroenteritis, including one case of culture-confirmed *L. monocytogenes* bacteraemia, among 32 persons having attended a tavern in the province on 15 December 2018 as part of a celebration. Previously, a pregnant physician who had visited the tavern and was aware that *L. monocytogenes* had been isolated from a blood culture of a tavern guest had informed the local health authority of Styria about a possible listeriosis outbreak. According to her, more than half of the guests started showing symptoms of febrile gastroenteritis and vomiting within 2 days after the tavern visit.
On 29 January 2019, the Austrian Ministry of Health (Vienna, Austria) mandated the Austrian Agency of Health and Food Safety (AGES; Graz, Austria) to investigate the outbreak. The aim of the investigation was to identify the causative agent and the likely source of infection, in order to detect and prevent further cases.

Methods
Outbreak case definition
An outbreak case was defined as an individual who presented with febrile gastroenteritis at least 24 hours after visiting the tavern in Styria, Austria on 15 December 2018 and tested positive for L. monocytogenes by either blood or stool culture.

Specimen collection and trace-back investigations
Of 32 individuals attending the celebration on 15 December 2018, 19 symptomatic individuals provided stool specimens. Of five individuals working in this tavern on this day, three asymptomatic individuals provided stool samples. The local food authority gathered information on involved food suppliers, restricting it to one local meat producer (company X) and three grocery shops (A, B and C).

Between 3 January and 25 January 2019, a total of 73 food and environmental samples were analysed for the presence of L. monocytogenes: (i) 19 environmental (surface) samples and three food samples produced by company X were collected from the tavern, (ii) three samples were collected from food produced at other facilities (shops A to C) (iii) one sample was collected from food produced by the tavern itself, and (iv) 47 food and environmental samples were collected from company X at their production facility. Products from company X were only offered at six locations, including the affected tavern and sold directly from the factory.

Microbiological investigations and WGS-based typing
The detection of L. monocytogenes in human samples was conducted as described elsewhere [11] and colonies were confirmed by Api-Listeria (BioMérieux, Marcy l’Etoile, France) or Maldi Biotyper (Bruker Daltonics, Hamburg, Germany). Food and environmental isolates received at the NRL were cultivated on RAPID’L.mono agar plates (Biorad, Munich, Germany) for species verification and subsequently subcultured overnight on Columbia Broth (BD Difco, Heidelberg, Germany) for extraction of high quality genomic DNA using the HMW MagAttract kit (Qiagen, Hilden, Germany) according to the instructions of the manufacturer for Gram-positive bacteria.

WGS was performed as described previously [12]. For sequencing, an Illumina MiSeq platform (Illumina Inc., San Diego, California, United States) was used. Library preparation was carried out using Nextera XT according to the instructions of the manufacturer (Illumina Inc.). For assembly into draft genomes, raw reads were de novo assembled using SPAdes version 3.11.1 [13]. Classical multilocus sequence typing (MLST) data were extracted from WGS sequence data [14]. A minimum spanning tree (MST) was generated in SeqSphere+ for visualisation of strain relatedness. Assessment of the core-genome multilocus sequence typing (cgMLST) was done as described by Ruppitsch et al. [8]. In parallel, a single nucleotide polymorphism (SNP) analysis was performed for comparison purposes with GenomeGraphR [15]. In addition, we created two task templates that were implemented with Seqsphere+ for detection of the pathogenicity islands LIPI-3 and LIPI-4 using the reference genomes from strain F2365 and strain CLIP 80459, respectively (NC_002973.6, NC_012488.1). Other virulence genes (VGs) were detected among the sequenced genomes by using VirulenceFinder [9].

Active case finding
We conducted active case finding to detect additional human cases (retrospectively and prospectively). A new case definition was created, which included any person testing positive to the outbreak strain by WGS since three months before the tavern gathering. We performed an intensive search and strain comparison using our local L. monocytogenes database, which currently contains nearly 8,000 sequenced isolates.
Ethical statement
Ethical approval to conduct the study was not needed. In Austria, investigation of foodborne outbreaks is a legal obligation.

Results
Microbiological findings
Of 19 stool specimens collected from individuals with febrile gastroenteritis, 10 samples (specimen ID: H03–H10, H12, H14) were positive for L. monocytogenes. Of three stool specimens from asymptomatic staff members of the tavern, only one stool sample from a female in her early 60s (H11) tested positive for L. monocytogenes. In addition, one L. monocytogenes isolate came from a blood culture obtained from a male in his mid-20s who was hospitalised for febrile gastroenteritis 24 hours after the tavern visit (H02).

Of 73 non-human samples, two food samples and one environmental sample tested positive for L. monocytogenes. The two food samples (specimen ID: F01: smoked meat ('Geselchtes') and F02: liver pâté ('Leberstreichwurst') were collected at the tavern and produced by company X; both tested positive for L. monocytogenes. In addition, one L. monocytogenes isolate came from a blood culture obtained from a male in his mid-20s who was hospitalised for febrile gastroenteritis 24 hours after the tavern visit (H02).

In addition to the three non-human isolates (F01–F03), 12 human L. monocytogenes isolates (from specimens: H2–H12, H14) were available for sequencing. cgMLST performed as described by Ruppitsch et al. [8] revealed that all isolates belonged to the same genetic type: they were identified as genoserogroup IVb, clonal complex (CC)4, multilocus sequence type (ST)4 and cg complex type (CT)7652 [8] and displayed 0 to 1 allelic difference (Figure 1). In total, 1,701 loci composing the cgMLST scheme were detected. The SNP analysis revealed a maximum of four SNPs difference between the isolates and we did not find related strains in the database. All isolates yielded the same 72 virulence genes including LIPI-3 and LIPI-4 pathogenicity islands.

International alert and active case finding
Since this outbreak strain had not been detected previously in Austria, an urgent inquiry was launched through the Epidemic Intelligence Information System (EPIS) at the European Centre for Disease Prevention and Control (EPIS-UI-539) on 28 January 2019. Denmark, Finland, France, the Netherlands, Luxembourg, Portugal, Switzerland and the United Kingdom answered the inquiry via the EPIS platform; no cases were linked to this outbreak strain. The closest matches were found at the European Nucleotide Archive repositories showed at least 32 allelic differences when using the cgMLST scheme from Moura et al. [10].

Through active case finding, we identified two more cases of invasive listeriosis. These cases lived in the same two geographical districts ('Bezirk') as the tavern and company X. The first isolate was taken from a blood culture from a case in their early 80s (H1) who had not visited the tavern but developed symptoms.
in November 2018. The case, who later died from the infection, reported having repeatedly consumed liver pâté from company X purchased at a local market. The second isolate originated from an eye chamber aspirate from a case in their mid-50s with endophthalmitis (H13), with onset of clinical symptoms on 23 December 2018; the case had not visited the tavern. The two isolates differed by a maximum of one allele from the other clinical isolates and the food and environmental isolates.

In total, 13 individuals (seven females and six males; age range: 4–84 years) living within area radius of 24 km in Styria, were confirmed as outbreak cases.

A timeline displaying all cases belonging to the outbreak is shown in Figure 2. A geographical representation of the residence for the confirmed \textit{L. monocytogenes} cases, the tavern and the meat producer company X is shown in Figure 3.

**Outbreak control measures**

After the outbreak was confirmed on 21 December 2018, intensive cleaning of the tavern and at company X was performed by professional sanitation companies, under supervision of the local health authority. Only heat-treated products were allowed to be sold until confirmed sanitation (i.e. \textit{L. monocytogenes} was no longer detected after repeated sampling). No more cases were detected by the end of December and the outbreak was declared over.

**Discussion**

The use of epidemiological data and WGS-based typing allowed us to confirm a local outbreak due to an \textit{L. monocytogenes} IVb-CC4-ST4-CT7652 strain, not previously detected in Austria nor anywhere else. Trace-back investigations showed that meat products originating from company X were the most likely source of the outbreak. The 13 confirmed cases lived within a radius of 24 km, further suggesting that the outbreak was caused by a locally sourced ingredient. No more cases were detected after company X implemented control measures, supporting the hypothesis that meat products from company X were the likely source of the outbreak.

In contrast to classical restaurants, a tavern offers a limited selection of traditional food (especially local meat products and cheese) in a buffet-like manner but has a fully functioning kitchen including a fridge and a freezer. In Austria, gatherings of large groups at traditional taverns are common around festivities such as Christmas. It is possible that the time between food preparation and service might have been long increasing the possibility for \textit{L. monocytogenes} to multiply.
Moreover, traditional Austrian meat products e.g. liver pâté and jellied pork do not require heating prior to consumption and have been previously identified as potential risk food for listeriosis due to their growth potential for *L. monocytogenes* [16,17]. In 2009, an outbreak of febrile gastroenteritis was reported that was also associated with a traditional Austrian tavern; this time locally produced jellied liver was identified as the vehicle of infection [11]. Liver pâté (‘Leberstreichwurst’) is a peculiar type of traditional Austrian meat product, made from cured, cooked, comminuted meat with fat as a binder. Its average pH value is 6.19 ± 0.15, with a corresponding water activity value of 0.963 ± 0.003, which provides *L. monocytogenes* with an optimal substrate for its growth [17]. Other types of pork pâté such as ‘pâté de campagne’ were found to be contaminated with *L. monocytogenes* [18]. Although enumeration of *L. monocytogenes* in the pâté seemed low (<10 CFU/gram), no Tween 80 was added to its initial suspension during the enrichment procedure, and that might explain the relatively low number of bacteria found per gram.

The only case without a proven connection to liver pâté of company X was the case with endophthalmitis; only 43 cases of endophthalmitis attributed to an *L. monocytogenes* infection have been reported in the literature so far [5,19-21]. While we were unable to elucidate any connection of this patient to company X, the case confirmed regular consumption of liver pâté so it is possible that the meat product may have been produced at company X.

*L. monocytogenes* serotype IVb strains are commonly associated with outbreaks [22]. Moreover, serotype IVb CC4 is known to be widely distributed in food and it has been described as one of the IVb hypervirulent CCs [23,24]. All 14 human isolates (from the 13 confirmed cases plus the stool isolate from the asymptomatic case), the two food isolates and the environmental isolate carried LIP1-3 and LIP1-4 pathogenicity islands, which have been associated with high invasiveness and severe symptomatology [25]. In contrast, another article reported the hypervirulent CC4 was associated with dairy products, while hypovirulent clones, such CC9 or CC121 were associated with meat products [26].

The main limitation of our study was associated with the closure of the tavern over the Christmas period. Due to the closure, the AGES outbreak investigation did not start until 45 days after the tavern visit. Nevertheless, control measures had been previously put in place preventing new cases to occur.

**Conclusion**

The isolate-based surveillance of *L. monocytogenes* using WGS-based typing and analyses of epidemiological data allowed us to confirm a local outbreak due to a *L. monocytogenes* IVb-CC4-ST4-CT7652 strain not previously detected elsewhere. Epidemiological and trace-back investigations showed the liver pâté produced at company X was the most likely source of infection. The applied control measures were effective in stopping the outbreak. Additional investigations are needed to estimate the risk of infection with *L. monocytogenes* when attending celebrations at taverns serving non-heated products that have been stored at room temperature for extended periods of time.

**Depositing of the sequences**

This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the project number PRJNA528854 and corresponds to the study number SRR8776907. Raw sequence data for each strain were deposited under SRA accession numbers SRR8776907, SRR8776908, SRR8776904, SRR8776910, SRR8776917, SRR8776915, SRR8776918, SRR8776920, SRR8776906, SRR8776905, SRR8776911, SRR8776912, SRR8776913, SRR8776914, SRR8776916, SRR8776919, SRR8776909.

**Acknowledgements**

All co-authors in this manuscript declare that no funding was received from any funding agency in the public, commercial or not-for-profit sectors. AC was supported by a grant from the European Public Health Microbiology Training Programme (EUPHEM), European Centre for Disease Prevention and Control (specific grant agreement number 1 ECD.7550 implementing ECDC/GRANT/2017/003).

**Conflict of interest**

None declared.

**Authors’ contributions**

AP, FA and WR were responsible for the concept and design of the study; AP, MW-N, AC and FA carried out the outbreak investigation; AP, AC, CS and WR conducted the microbiological analyses; AP, MW-N, AC and FA carried out the outbreak; AP, FA and WR were responsible for the concept and design of the study; AP, FA and WR were responsible for the concept and design of the study; AP, MW-N, AC and FA carried out the outbreak investigation; AP, AC, CS and WR conducted the microbiological investigations; all authors contributed to the writing and final version of the manuscript.

**References**

1. Allerberger F, Wagner M. Listeriosis: a resurgent foodborne infection. Clin Microbiol Infect. 2010;16(1):6-23. https://doi.org/10.1111/j.1469-0691.2009.03109.x PMID: 20002687
2. Hernandez-Milian A, Payeras-Cifre A. What is new in listeriosis? BioMed Res Int. 2014;2014:358051. https://doi.org/10.1155/2014/358051 PMID: 24822197
3. Lamont RF, Sobel J, Mazaki-Tovi S, Kusanovic JP, Vaisbich E, Kim SK, et al. Listeriosis in human pregnancy: a systematic review. J Perinat Med. 2011;39(3):227-36. https://doi.org/10.1515/jpm.2011.035 PMID: 21517700
4. Pagliano P, Arslan F, Ascione T. Epidemiology and treatment of the commonest form of listeriosis: meningitis and bacteremia. Infez Med. 2017;25(3):210-6. PMID: 28956537
5. Chersich MF, Takkinnen J, Charlier C, Leclercq A, Adams PE, Godbole G, et al. Diagnosis and treatment of Listeria monocytogenes endophthalmitis: A Systematic Review. Ocul Immunol Inflamm. 2018;26(4):508-17. PMID: 28145786
6. Lopez-Valladares G, Danielsson-Tham ML, Tham W. Implicated food products for listeriosis and changes in serovars of Listeria monocytogenes affecting humans in recent decades. Foodborne Pathog Dis. 2018;15(7):387-97. https://doi.org/10.1089/fpd.2017.2419 PMID: 29958028
7. Leong D, Alvarez-Ordóñez A, Jordan K. Monitoring occurrence and persistence of Listeria monocytogenes in foods and
24. Lee S, Chen Y, Gorski L, Ward TJ, Osborne J, Kathariou S.
23. Maury MM, Tsai Y-H, Charlier C, Touchon M, Chenal-Francisque
22. Todd ECD, Notermans S. Surveillance of listeriosis and
21. Holbach LM, Bialasiewicz AA, Boltze HJ. Necrotizing ring ulcer
20. Gaskell KM, Williams G, Grant K, Lightman S, Godbole G.
19. Bajor A, Luhr A, Brockmann D, Suerbaum S, Framme C,
18. de Valk H, Vaillant V, Jacquet C, Rocourt J, Le Querrec F, Stainer
17. Awaiwanont N, Smulders FJM, Paulsen P. Growth potential of
15. Sanaa M, Pouillot R, Vega FG, Strain E, Van Doren JM.
14. Salcedo C, Arreaza L, Alcalá B, de la Fuente L, Vázquez JA.
13. Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov
12. Lepuschitz S, Mach R, Springer B, Allerberger F, Ruppitsch W.
11. Pichler J, Much P, Kasper S, Fretz R, Auer B, Kathan J, et al.
10. Moura A, Tourdjman M, Leclercq A, Hamelin E, Laurent E,
9. Joensen KG, Scheutz F, Lund O, Hasman H, Kaas RS, Nielsen
8. Ruppitsch W, Pietzka A, Prior K, Bletz S, Fernandez HL, Allerberger F, et al. Defining and Evaluating a Core Genome Multilocus Sequence Typing Scheme for Whole-Genome Sequence-Based Typing of Listeria monocytogenes. J Clin Microbiol. 2015;53(10):4186-97. https://doi.org/10.1128/JCM.01935-15 PMID: 26135865
Joensen KG, Scheutz F, Lund O, Hasman H, Kaas RS, Nielsen EM, et al. Real-time whole-genome sequencing for routine typing, surveillance, and outbreak detection of verotoxigenic Escherichia coli. J Clin Microbiol. 2014;52(5):1501-10. https://doi.org/10.1128/JCM.03617-13 PMID: 24574290

10. Moura A, Tourdjman M, Leclercq A, Hamelin E, Laurent E, Fredriksen N, et al. Real-time whole-genome sequencing for surveillance of Listeria monocytogenes, France. Emerg Infect Dis. 2017;23(9):1462-70. https://doi.org/10.3201/eid2309.170336 PMID: 28643628
11. Pichler J, Much P, Kasper S, Fretz R, Auer B, Kathan J, et al. An outbreak of febrile gastroenteritis associated with jellied pork contaminated with Listeria monocytogenes. Wien Klin Wochenschr. 2009;121(3-4):149-56. https://doi.org/10.1007/s00058-009-1137-3 PMID: 19280142
12. Lepuschitz S, Mach R, Springer B, Allerberger F, Ruppitsch W. Draft Genome Sequence of a Community-Acquired Methicillin-Resistant Staphylococcus aureus USA300 Isolate from a River Sample. Genome Announc. 2017;5(2):e02166-17. https://doi.org/10.1128/genomeA.02166-17 PMID: 29051260
13. Nurb S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, et al. Assembling single-cell genomes and minimetagenomes from chimeric MDA products. J Comput Biol. 2013;20(6):714-37. https://doi.org/10.1089/cmb.2013.0084 PMID: 24093227
14. Salcedo C, Arreaza L, Alcalá B, de la Fuente L, Vázquez JA. Development of a multilocus sequence typing method for analysis of Listeria monocytogenes clones. J Clin Microbiol. 2003;41(2):757-62. https://doi.org/10.1128/JCM.41.2.757-762.2003 PMID: 12574278
15. Sanee M, Pouillot R, Vega FG, Strain E, Van Doren JM. GenomEigraphR: A user-friendly open-source web application for foodborne pathogen whole genome sequencing data integration, analysis, and visualization. PLoS One. 2019;14(2):e0213039. https://doi.org/10.1371/journal.pone.0213039 PMID: 3134816
16. Paulsen P, Awaiwanont N, Smulders FJM. Growth of Listeria monocytogenes in traditional Austrian meat jelly products. Procedia Food Sci. 2015;5:219-22. https://doi.org/10.1016/j.profo.2015.09.027
17. Awaiwanont N, Smulders FJM, Paulsen P. Growth potential of Listeria monocytogenes in traditional Austrian cooked-cured meat products. Food Control. 2015;50:150-6. https://doi.org/10.1016/j.foodcont.2014.08.043
18. de Valk H, Vaillant V, Jacquet C, Rocourt J, Le Querrec F, Stainer F, et al. Two consecutive nationwide outbreaks of Listeriosis in France, October 1999-February 2000. Am J Epidemiol. 2001;154(10):944-50. https://doi.org/10.1093/aje/154.10.944 PMID: 12574278
19. Bajor A, Lahr A, Brockmann D, Suerbaum S, Framme C, Sedlacek L. Listeria monocytogenes endophthalmitis - case report and review of risk factors and treatment outcomes. BMC Infect Dis. 2016;16(1):332. https://doi.org/10.1186/s12879-016-1680-2 PMID: 27424034
20. Gaskell KM, Williams G, Grant K, Lightman S, Godbole G. Listeria Monocytogenes: a rare cause of endophthalmitis, a case report. IDCases. 2017;8:43. https://doi.org/10.1128/idc.00508-17 PMID: 28643628
21. Holbach LM, Bialasiewicz AA, Bötze HJ. Necrotizing ring ulcer of the cornea caused by exogenous Listeria monocytogenes serotype IV b infection. Am J Ophthalmol. 1988;106(1):105-6. https://doi.org/10.1016/S0002-9394(14)76347-3 PMID: 31779749
22. Todd ECD, Notermans S. Surveillance of listeriosis and its causative pathogen, Listeria monocytogenes. Food Control. 2011;22(9):1484-90. https://doi.org/10.1016/j.foodcont.2010.07.021
23. Maury MM, Tsai Y-H, Charlier C, Touchon M, Chenal-Francisque V, Leclercq A, et al. Uncovering Listeria monocytogenes hypervirulence by harnessing its biodiversity. Nat Genet. 2016;48(3):308-13. https://doi.org/10.1038/ng.3501 PMID: 26829754
24. Lee S, Chen Y, Gorski L, Ward TJ, Osborne J, Kathariou S. Listeria monocytogenes source distribution analysis indicates regional heterogeneity and ecological niche preference among serotype 4b clones. MBio. 2018;9(2):e00396-18. https://doi.org/10.1128/mBio.00396-18 PMID: 29662682
25. Vilchis-Rangel RE, Espinoza-Mellado MDR, Salinas-Jaramillo II, Martínez-Peña MD, Rodas-Suárez OR. Association of Listeria monocytogenes LIP1-1 and LIP1-3 marker l1sW with invasiveness. Curr Microbiol. 2019;76(5):637-43. https://doi.org/10.1007/s00284-019-10671-2 PMID: 30888475
26. Maury MM, Bracq-Dieye H, Huang L, Vales G, Lavina M, Thouvenot P, et al. Hypervirulent Listeria monocytogenes clones’ adaption to dairy products. Nat Commun. 2019;10(1):2488. https://doi.org/10.1038/s41467-019-10380-0 PMID: 31177974

License, supplementary text and copyright
This is an open-access article distributed under the terms of the Creative Commons Attribution (CC BY 4.0) Licence. You may share and adapt the material, but must give appropriate credit to the source, provide a link to the licence and indicate if changes were made.

Any supplementary material referenced in the article can be found in the online version.

This article is copyright of the authors or their affiliated institutions, 2019.