**Racioethnic Differences in Human Posterior Scleral and Optic Nerve Stump Deformation**

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**Purpose.** The purpose of this study was to quantify the biomechanical response of human posterior ocular tissues from donors of various racioethnic groups to better understand how differences in these properties may play a role in the racioethnic health disparities known to exist in glaucoma.

**Methods.** Sequential digital image correlation (S-DIC) was used to measure the pressure-induced surface deformations of 23 normal human posterior poles from three racioethnic groups: African descent (AD), European descent (ED), and Hispanic ethnicity (HIS). Regional in-plane principal strains were compared across three zones: the optic nerve stump (ONS), the peripapillary (PP) sclera, and non-PP sclera.

**Results.** The PP scleral tensile strains were found to be lower for ED eyes compared with AD and HIS eyes at 15 mm Hg ($P = 0.024$ and 0.039, respectively). The mean compressive strains were significantly higher for AD eyes compared with ED eyes at 15 mm Hg ($P = 0.018$). We also found that the relationship between tensile strain and pressure was significant for those of ED and HIS eyes ($P < 0.001$ and $P = 0.004$, respectively), whereas it was not significant for those of AD ($P = 0.392$).

**Conclusions.** Our results suggest that, assuming glaucomatous nerve loss is caused by mechanical strains in the vicinity of the optic nerve head, the mechanism of increased glaucoma prevalence may be different in those of AD versus HIS. Our ONS strain analysis also suggested that it may be important to account for ONS geometry and material properties in future scleral biomechanical analysis.

Keywords: glaucoma, sclera, strain, race, ethnicity

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**Glaucoma** is a neuronal degenerative disease that damages retinal ganglion cells (RGC), leading to irreversible vision loss. In 2010, glaucoma affected 60.5 million people, a number that is projected to rise to approximately 80 million by 2020. Risk factors for glaucoma include family history of glaucoma, race, age, and increased intraocular pressure (IOP). These factors influence the biomechanical environment of the lamina cribrosa (LC), which is hypothesized to be the primary region of mechanical insult in glaucoma. Changes in tissue properties of the optic nerve head and surrounding sclera are believed to accompany the initiation and progression of glaucoma by either influencing LC deformations or by causing a reduction of ocular blood flow. Open angle glaucoma (OAG) has been shown to be more prevalent in African descent (AD) populations at all ages, but proportionally increases more rapidly in the European descent (ED) population over the same age compared with AD populations. Stiffness of peripapillary (PP) sclera surrounding the optic nerve has been reported to increase with age and in those of AD compared with those of ED. Significant microstructural differences in human posterior scleral tissue are also present between these two racioethnic groups, with those of AD having less equatorially aligned collagen fibers compared with those of ED. These microstructural and material property differences may be responsible for the varying rates of glaucoma occurrence observed between those of AD and ED. In addition, the prevalence of OAG for those of HIS was found to be higher compared with those of ED, but lower compared with those of AD.

From these prior studies, it is important to determine whether there exist any fundamental biomechanical differences in nonglaucomatous eyes in posterior ocular tissues across races and posterior eye regions. Experimental determination of the biomechanical properties of the enucleated optic nerve stump (ONS) and sclera is challenging due to their nonlinear and anisotropic response, as well as their complex geometry. High-resolution deformation protocols of the posterior sclera that use pressure-inflation experiments have
been reported, with three-dimensional (3D) digital image correlation (DIC).\textsuperscript{21–23} 3D ultrasound speckle tracking (UST),\textsuperscript{24–26} and electronic speckle pattern interferometry (ESPI),\textsuperscript{29–33} being among the most commonly used full-field optical techniques. ESPI offers very high scleral deformation sensitivity,\textsuperscript{29,32} whereas 3D DIC has achieved high-resolution measurement of in-plane scleral displacement.\textsuperscript{21,23} 3D UST has been used to acquire volumetric scans quantifying complex internal scleral strains.\textsuperscript{27} Despite their numerous utilization in scleral deformation studies, none of these 3D deformation measuring techniques have been used to quantify human ONS displacement, due to current measurement limitations and the aspherical geometry of posterior ocular tissues.\textsuperscript{27} This complex geometry demands proper contouring and matching of the local shape with the displacement measurement to enable correct strains map calculation.\textsuperscript{21,34,35} For ESPI, these geometry deviations lead to shadows from oblique illumination that results in irregular data analysis.\textsuperscript{36} In standard DIC techniques, such complex nonospherical geometries with sharp angles are not fully captured in the two viewing angles leading to improper correlation and reconstruction. Because these angles cannot be readily changed without influencing in-plane and out-of-plane resolution,\textsuperscript{37} data analysis from these methods is not accurate for these complex geometry regions.

Our research group recently developed an S-DIC method with advanced 3D capabilities to map posterior sclera and ONS deformations.\textsuperscript{37} In this method, we improved the z (in-depth) resolution without loss of x-y (in-plane) sensitivity in DIC measurements. Our approach used one high-resolution camera that recorded videos of a pressure inflation device as it moved through two orthogonal parallax axes using a standard 3D DIC approach to quantify the deformation of posterior sclera and ONS across carefully selected sequential movie frames (up to the frame rate of the camera). The capabilities of the S-DIC technique were assessed in a prior work by the measurements of shape and displacement on a rigid complex geometry (ONS surrogate) with a reconstruction accuracy of 0.17% and 8-μm uncertainty in the out-of-plane direction.\textsuperscript{37} There are few studies in the literature that have studied the biomechanical properties of the ONS.\textsuperscript{13,15,36–44} As for comparisons across racioethnic groups, only variation of laminar depth with age\textsuperscript{42} and LC displacement\textsuperscript{45} in normal eyes across AD and ED groups have recently been reported. The racioethnic variation in IOP-induced deformation of the human ON in nonglaucomatous tissue has not been reported in the literature, in particular for those of HIS. The purpose of this work was to use S-DIC to evaluate the strains of nonglaucomatous scleral shells across three racioethnic groups (AD, ED, and HIS), four regions (temporal [T], nasal [N], superior [S], and inferior [I]), and three zones (PP sclera, non-PP sclera, and ONS). This comparison was done by analyzing data from four IOP states: 5, 15, 30, and 45 mm Hg.

### MATERIALS AND METHODS

#### Posterior Scleral Shells and ONSs

Posterior scleral shells from human donors of AD (57 to 98 years old, n = 7 eyes), of ED (52 to 92 years old, n = 11 eyes), and HIS (56 to 95 years old, n = 5 eyes) were received from the Alabama Eye Bank in Birmingham, AL, USA; the Banner Sun Health Research Institute in Sun City, AZ, USA; the Donor Network of Arizona in Phoenix and Tucson, AZ, USA; the Illinois Eye Bank in Chicago and Bloomington, IL, USA; the Michigan Eye Bank in Ann Arbor, MI, USA; and the San Diego Eye Bank in San Diego, CA, USA. All eyes were classified as nonglaucomatous according to available medical paperwork, next-of-kin questionnaires, and/or later confirmed via optic nerve axon count. Age of all donors for this study was limited to those over 50 years of age. The eyes were kept on ice until they were shipped to our laboratory and were stored in physiologic saline at 4°C. For axon counting, the optic nerves corresponding to the scleral shells were cut and fixed immediately in Poly/LEM (Polyscience, Inc., Warrington, PA, USA) at their respective eye banks, at which point they were sent to our laboratory. Once received, the nerves were transferred to vials containing 2.5% glutaraldehyde, in which they stored for 24 hours, transferred to PBS, and stored at 4°C until they were sent out for processing. For processing, two to three 1-mm-thick slices were cut from the middle of the available axial length of each nerve. In a few cases, a clean slice could not be prepared, and these nerves were left intact. The slices were washed several times with PBS and fixed with 1% osmium tetroxide in PBS for 2 hours on ice. The samples were extensively rinsed with ddH₂O, dehydrated through a grades series of ethanol concentrations, treated with propylene oxide, and infiltrated with Eponate 12 resin overnight (Ted Pella, Redding, CA, USA). The following day, the slices were placed in fresh resin, and polymerized at 60°C overnight. Semi-thin sections were cut from the blocks on an Ultratome E ultramicrotome (Reichert-Jung, Wein, Austria) and stained with 1% toluidine blue in 1% sodium borate. Unfortunately, some of the embedded slides lacked the necessary quality for axon counting and were not used. The Table reports the race, age, sex, and anatomical location of all donor eyes, including whether or not the optic nerve cross section was used for axon counting.

#### Automated S-DIC

All scleral shells were tested using an S-DIC inflation procedure previously developed by our research group.\textsuperscript{57} As part of our research group’s efforts to enhance our previously reported S-DIC imaging approach, significant improvements were made to the system to reduce hands-on imaging time, maintain consistent eye hydration, upgrade vibration control, and streamline postprocessing methods. Overall, the most significant improvement to the S-DIC system was the inclusion of automated motor rotational stages (CRI-Z7, PRM1Z8; Thorlabs, Newton, NJ, USA), controlled by LabView (National Insti-
ments, Inc., Austin, TX, USA) that acted as the primary controller that managed the camera acquisition and rotational stage movements throughout the experiments. The inclusion of the automatic motors allowed for improved vibration control due to the fine incremental rotational motion resolution, as well as eliminated the need to open the humidity chamber doors for manual actuation thus improving humidity consistency. All other S-DIC experimental protocol steps remained identical to our previously published approach, including eye dissection, eye positioning within the clamp system, speckle pattern application, preconditioning, and imaging at the pressure steps of 5, 15, 30, and 45 mm Hg. With decreased image acquisition time, the automated S-DIC method allowed the user interaction with the system to be completely hands-off once the eye was properly installed. The S-DIC postprocessing techniques were also improved with the automated system through acquiring consistent angle sequence images between ocular experiments. This allowed for robust postprocessing scripts, less procedural variations between eyes, and faster generation of ocular strain fields.

To generate strain maps, strains were calculated following the protocol described in our previous study. Briefly, a geometry mesh was generated using measured position data triangulation. Larger elements were remeshed using custom Matlab code so that the element sizes for all regions and zones were similar. The Green–Lagrange strain tensor, \( E \), was computed for each element using the equation

\[
\frac{\partial \mathbf{F}}{\partial \mathbf{F}} = \frac{1}{2} \left( \frac{\partial \mathbf{F}^T \mathbf{F}}{\partial \mathbf{F}} - \mathbf{I} \right),
\]

where \( F \) is the deformation gradient tensor and \( I \) is the identity matrix. \( E \) was used to calculate two in-plane principal strains for each element, \( E_1 \) and \( E_2 \). The strain calculations assumed that the 5-mm Hg base geometry was the undeformed state for each inflation pressure. This resulted in three pairs of elemental principal strains: \( E_1 \) and \( E_2 \) from 5 to 15, 5 to 30, and 5 to 45 mm Hg (simply referred to as pressures 15, 30, and 45 mm Hg for the remainder of this article).

Zonal and Regional Segmentation

To compare strain values from different spatial locations, all geometries were segmented into three zones and four regions. With regards to the zones, each geometry was divided into the ONS, the PP sclera, and the non-PP sclera. Briefly, the geometry was rotated so that the ONS pointed in the positive z-direction. A Hessian matrix was calculated along the geometry and was used to determine the mean curvature for each element. Using a curvature criterion, the saddle points of the ONS ring were determined and used to fit to a dividing plane. All elements above the plane were designated as ONS elements as shown in Figure 1A. The remaining elements within a distance of 2 mm below the plane were designated as PP scleral elements. The remainder of the scleral elements were designated as non-PP scleral elements, as shown in Figure 1B. After zonal segmentation, the ONS elements were imported into Rhinoceros 3D (Robert McNeel & Associates, Seattle, WA, USA), which was used to close the ONS geometry so that the volume of each ONS could be calculated for each sample (mm\(^3\)). In addition, the diameter and length of the ONS were calculated. Due to the nonuniform shape of the ONS, the diameter was calculated for each sample by subtracting the maximum and minimum positions of the ONS in the x- and y-direction. Both of these measurements were averaged and recorded as the diameter (mm). Similarly, the maximum and minimum positions of the ONS were subtracted in the z-direction. This was recorded as the ONS length (mm). It should also be noted that the ONS zone for all eyes encompassed mostly the optic nerve sheath and a small portion of the optic nerve that was exposed during the enucleation process.

For regional definition, the geometry was divided into S, I, T, and N. To do this, the centroid of the scleral opening produced by the dividing plane was set as the origin of the geometry. Furthermore, the geometry was rotated so that the normal of the dividing plane was aligned with the z-axis. The nodes of all the elements were converted from Cartesian to spherical coordinates. The known S direction was set as \( \theta = 0^\circ \) in the meridional direction. All elements within \( 45^\circ \) of the S direction were designated as S elements. The same was done for the remaining regions dividing the whole geometry into four equally spaced regions as shown in Figure 1C. The orientation of the T and N direction was determined by whether the eye was oculus dexterus (OD) or oculus sinister (OS). The overall 12 spatial locations defined by regions and zones are shown in Figure 1D.

Optic Nerve Axon Counting

The embedded optic nerve cross sections were visualized on a Nikon Eclipse 90i microscope using a montaging method with Nikon NIS-Elements software (Nikon, Inc., Millville, NY, USA). The method used individually imaged 60× magnifications with autofocus capabilities and a 15% overlap. A complete montage of an axon cross section is shown in Figure 2. Semi-automated axon counts were executed using image processing techniques in MatLab and methodology adapted from literature. The user was first prompted to identify the bounds of the optic nerve cross section to crop the image. Contrast-limited adaptive histogram equalization was applied to the cropped image to improve user accuracy of axon identification. The cropped portion of the image was divided equally into a sectional grid comprised of 200 sections horizontally by 200 sections vertically. From these 40,000 sections, 10 individual sections of axons were randomly selected and presented one at a time, and the user was prompted to manually click on the axons in each section. These manual counts were averaged across all 10 counted sections, providing an average axon density for the small image size. This was done to create a more accurate representation of the average axon density in these sections with respect to the entire optic nerve area, as axon distribution is somewhat nonuniform by nature. This calculated density was then used to extrapolate across the entire cross section to give an estimated axon count for the whole nerve. Each image was “counted” through the program three times to improve repeatability and accurate averages with SDs of axon counts being calculated for all nerves. Multiple users performed axon counts on all samples, and their counts were averaged to account for interuser variability.

Statistical Analysis

A statistical analysis was performed using scripts written in R (GNU General Public License, R Core Team, Vienna, Austria). Specifically, a parsimonious linear mixed model built to account for repeated measures (lmer) was used. Racioethnicity (AD, ED, and HIS), zone (ONS, PP sclera, and non-PP sclera), region (S, T, I, and N), and pressure (15, 30, and 45 mm Hg) were considered fixed repeated-measure variables. For this model, pressure was considered as a discrete variable to evaluate racial and zonal differences at specific inflation pressures. Each donor was considered an individual subject for the repeated measures portion of the model. Mean E1 and E2 estimates were determined for each spatial location at every inflation pressure. The data were arranged in a factorial form suitable for the statistical model. All data were normalized using a two-step SPSS Statistics (IBM Corporation, Armonk, NY, USA) transformation procedure that used inverse distribution functions. Post hoc mean E1 and E2 estimates were
calculated using an R least squares mean function (lsmeansLT). Pairwise comparisons were calculated using an R differences of least squares means function (difflsmeans). Both of these functions used the R linear mixed model previously mentioned. To account for familywise error, a Bonferroni-type correction was applied on any pairwise test conducted.

For regression analysis, pressure was considered as a continuous variable to determine the relationship of E1 and E2 as a function of pressure for every zone for each racioethnic group. This was done using the linear mixed model in SPSS. Similar to the R model already mentioned, zone and region were considered repeated-measure variables, and each donor considered an individual subject for the model. Additionally, the same regression model was used to determine the relationships of mean E1 and mean E2 at all spatial locations and inflation pressures as a function of ONS volume and optic nerve axon count.

RESULTS
E1 and E2 Strain Maps
The E1 and E2 magnitudes for the 5- to 45-mm Hg inflation of one representative sample from each racioethnic group are shown in Figure 3. The E1 and E2 magnitudes in the ONS region were qualitatively larger than that of the scleral zones.
for all racioethnic groups. E2 magnitudes (compressive) were qualitatively higher than that of E1 magnitudes (tensile) in the same region. Furthermore, more qualitative regional heterogeneity can be observed in the ED and HIS samples compared with the AD sample. Overall, the reported values of E1 and E2 in the scleral zones were consistent with values reported in the literature.

**Statistical Analysis**

**Pressure as Discrete Variable.** A linear mixed model was performed in R using pressure as a discrete variable. The R linear mixed model results showed that all interaction terms higher than a second-order interaction were not significant and were therefore removed from the main model expression. The mean E1 and E2 estimates, which represent the values predicted by the statistical model, were calculated, and pairwise comparisons were performed between racioethnic groups for all three zones at each inflation pressure. The zonal comparisons are shown in Figure 4.

In the ONS (Fig. 4A), the mean E1 values for AD eyes were significantly higher than that of ED eyes \((P = 0.024)\) at 15 mm Hg. Similarly, the mean E2 absolute values were significantly higher for AD eyes compared with that of ED at 15 mm Hg. No significant zonal differences across racioethnic groups were shown at 30 and 45 mm Hg for both mean E1 and E2 values. For the PP scleral zone (Fig. 4B), the mean E1 values for ED eyes were significantly lower than that of AD and HIS eyes \((P = 0.0219\) and 0.0587, respectively) at 15 mm Hg. No significant racioethnic differences were observed at 30 and 45 mm Hg in the PP scleral zone. The mean E2 values were significantly higher for AD eyes compared with ED eyes at 15 mm Hg \((P = 0.049)\). For the non-PP scleral zone (Fig. 4C), there were no significant racioethnic differences for mean E1 or mean E2 values at all inflation pressures.

There were few significant regional differences within racioethnic groups at different inflation pressures. In the PP zone for HIS eyes, the mean E2 absolute values for the T region was lower than that of the N region at 15 mm Hg \((P = 0.048,\) respectively). For the non-PP scleral zone, the ED mean E1 values for I region were higher compared with the S region for 30 mm Hg \((P = 0.048)\). For the mean E2 absolute values, the I region was significantly higher than the T region at 30 and 45 mm Hg for ED eyes \((P = 0.024\) and 0.03, respectively). All other mean E1 and mean E2 regional differences within racioethnic groups were not significant. For zonal differences within racioethnic groups, the mean E1 values and mean E2 absolute values for the ONS zone were significantly higher than that of both the PP and non-PP scleral zone for all racioethnic groups at all inflation pressure \((P < 0.001\) for all). All other mean E1 and mean E2 differences between PP and non-PP scleral regions within racioethnic groups were not significant.

Pairwise comparisons were made between different pressure states for each zone within each racioethnic group. For the AD eyes in the ONS zone, E1 values at 30 mm Hg were significantly lower than that of 45 mm Hg \((P = 0.053)\) and the E2 absolute values at 15 mm Hg were significantly lower than that of 45 mm Hg \((P = 0.012)\). In the non-PP zone, the E1 values for AD eyes at 15 mm Hg were significantly lower than that of 45 mm Hg \((P < 0.001)\). For ED eyes, the ONS mean E1 values at 45 mm Hg were significantly higher than that of 30 and 15 mm Hg \((P = 0.045\) and <0.001, respectively). The ONS mean E2 absolute values at 15 mm Hg were significantly lower than that of 45 mm Hg \((P = 0.026)\). In the PP scleral zone, the ED mean E1 values at 15 mm Hg was significantly lower than that of 30 and 45 mm Hg \((P = 0.033\) and <0.001, respectively). Furthermore, the mean E1 values for ED eyes at 30 mm Hg in the PP scleral zone was significantly lower than that of 45 mm Hg \((P = 0.015)\). As for the non-PP scleral zone, the mean E1 values for ED eyes at 15 mm Hg were significantly lower than that at 45 mm Hg \((P = 0.026)\). As for HIS eyes, in the ONS zone, the mean E2 absolute values at 15 mm Hg were significantly lower than that at 45 mm Hg \((P = 0.025)\). In the PP scleral zone, the mean E1 values at 15 mm Hg were significantly lower than that at 45 mm Hg \((P = 0.028)\). All other comparisons in all zones for all racioethnic groups were not significantly different.
Pressure as Continuous Variable. A mixed linear model was performed in SPSS using pressure as a continuous variable to determine the relationship of mean E1 and E2 values as a function of pressure. This was done for each zone for each racioethnic group. The regression results are shown in Figure 5. In the ONS, only ED eyes had a significant positive relationship between mean E1 values and pressure ($P = 0.003$). AD, ED, and HIS eyes had a significant positive relationship between mean E2 absolute values and pressure ($P = 0.005$, $P < 0.001$, and $P = 0.017$, respectively). In the PP scleral zone, both ED and HIS eyes had a positive relationship between mean E1 values and pressure. Furthermore, the slope of the mean E1 pressure relationship of the ED eyes was significantly higher than that of AD eyes ($P = 0.04$). For mean E2 values, there was no significant relationship with pressure for any of the racioethnic groups. As for the non-PP scleral zone, AD, ED, and HIS eyes had a significant positive relationship between mean E1 values and pressure ($P = 0.003$, $P < 0.001$, and $P = 0.006$, respectively).

Axon Count. The mean axon counts were $9.05 \times 10^5 \pm 1.2 \times 10^5$, $9.65 \times 10^5 \pm 2.1 \times 10^5$, $1.1 \times 10^6 \pm 3.0 \times 10^5$ for the AD ($n = 4$), ED ($n = 10$), and HIS ($n = 2$) donors, respectively, where the ranges indicate SDs. A 2-tailed Student’s $t$-test was performed comparing the AD and ED eyes, which showed that there was no significant difference between both groups in regard to the optic nerve axon count ($P = 0.964$). A mixed linear model was performed in SPSS using axon count as a continuous variable to determine the relationship of mean E1 and E2 values for all eyes as a function of axon count for all three zones at 15, 30, and 45 mm Hg. In the ONS zone, there was a significant negative relationship between mean E1 values and axon count at 45 mm Hg ($P = 0.002$). There was a significant positive relationship between mean E2 absolute values and axon count at 45 mm Hg ($P = 0.006$). In the PP scleral zone at 45 mm Hg, there was no significant relationship between mean E1 values and pressure ($P = 0.964$); however, there was a significant positive relationship between E2 absolute values and axon count ($P = 0.022$). In the non-PP scleral zone at 45 mm Hg, there were no significant
relationships of neither mean E1 values nor mean E2 values with axon count (P = 0.106 and 0.596, respectively). All relationships between mean E1 and E2 and axon counts at 15 and 30 mm Hg at all zones were shown to be not significant. The regression plots for all zones at 45 mm Hg are shown in Figure 6.

**ONS Dimensions.** The mean ONS volumes were 16.2 ± 6.0, 23.6 ± 7.9, and 17.1 ± 11.9 mm³ for the AD, ED, and HIS donors, respectively. The mean ONS diameters were 5.2 ± 0.4, 6.4 ± 0.7, and 6.3 ± 1.7 mm for the AD, ED, and HIS donors, respectively. The mean ONS lengths were 2.9 ± 0.4, 3.2 ± 0.7, and 6.3 ± 0.9 mm for the AD, ED, and HIS donors, respectively. All ranges for all dimensions indicate 95% confidence intervals. Single-factor ANOVAs concluded that there were no significant differences between the different racioethnic groups in regard to the ONS volume (P = 0.330), diameter (P = 0.154), and length (P = 0.632). Comparisons of ONS volume, diameter, and length among racioethnic groups are shown in Figure 7.

In addition, to evaluate the effect of the ONS size on the E1 and E2 values, a mixed linear model was performed in SPSS to determine the relationship of mean E1 and E2 values for all eyes as a function of ONS volume for all three zones at 15, 30, and 45 mm Hg. The results are shown in Figure 8. Overall, it was found that the mean E1 and mean E2 value did not have a significant relationship with the ONS volume at all pressures (P < 0.001 for all). Furthermore, there was also a significant negative relationship between mean E2 absolute values and ONS volume at 30 and 45 mm Hg (P = 0.009 and 0.023, respectively) in the non-PP scleral zone.

**DISCUSSION**

A zonal racioethnic in-plane principal strain comparison was conducted across three inflation pressures for normal human scleral shells using displacement measurements collected via S-DIC. The ONS tensile and compressive strains for the ED eyes were significantly lower than that of AD eyes at 15 mm Hg (Fig. 4A). The PP scleral compressive strains for the AD eyes were significantly larger than that of ED eyes at 15 mm Hg (Fig. 4B). For ED and HIS eyes, tensile strains in the ONS and non-PP scleral zones increased significantly with pressure, whereas AD eyes showed a significant relationship with pressure in the non-PP scleral zone only (Fig. 5). Additionally, only ED and HIS eyes had a significant positive relationship between tensile strain in the PP scleral region, whereas AD PP scleral strains did not have a significant relationship with pressure. All racioethnic groups had significant compressive strain relationships with pressure in the ONS zone (Fig. 5).

Several studies have previously used principal strains as an end point to compare spatial locations in eyes, whereas some used circumferential and meridional strain. The principal strain values reported in our study were found to be within the range of principal strain values shown in other studies. Fazio et al. used ESPI to investigate
the mean maximum principal strain (E1) regional variability in the PP and midperipheral scleral regions (each divided into eight meridional regions) in 10 pairs of normal eyes ages 57 to 90 years old with nonspecified race/ethnic classification. The study by Fazio et al. showed that the I region demonstrated E1 strain values higher than the S and N regions, which was consistent with our regional trend of the ED tensile strains in the non-PP scleral zone. Coudrillier et al. found regional differences in fiber alignment in the PP scleral location for normal eyes (n = 9). In addition, our research group has previously investigated regional microstructural differences for glaucomatous and normal human eyes of ED. In that study, Danford et al. showed that normal ED eyes exhibited lower percent of equatorial fiber organization in the I region compared to all other regions, especially as compared with the N region, which had the highest degree of circumferential fiber alignment. This disparity may explain the higher tensile and compressive strains in the I region of ED samples reported in the current study as a larger degree of noncircumferential fibers may lead to increased fiber realignment into the circumferential direction and subsequently larger in-plane compressive strains. Future microstructural information in AD and HIS eyes could further explain the various regional strain differences observed throughout all race/ethnic groups. These studies are currently ongoing in our laboratory.

With regard to zonal differences within race/ethnic groups, the study by Fazio et al. found that the PP sclera exhibited significantly higher mean maximum values of principal strains (E1) than midperipheral scleral regions for a 5- to 45-mm Hg pressure inflation. This is not consistent with our results that showed tensile differences only between the ONS and the both scleral zones at all pressures. The study by Fazio et al. also reported that PP sclera exhibited significantly higher values of mean E1 strains than midperipheral sclera for both donors of AD and ED. This was also in contrast to our results that indicated that the PP sclera did not differ significantly from the non-PP region for AD and ED eyes. This could be due to our study including non-PP sclera beyond the midperipheral scleral zone. Furthermore, the ONS volume of eyes in our study seemed to have a significant effect on the tensile and compressive strain values in the non-PP scleral zone (Fig. 8). This could further explain why our ED and AD zonal comparison results differed from those of Fazio et al. who did not include the ONS. In addition, the discrepancy between our research groups in zonal differences within race/ethnic groups could be explained by a number of other factors, including scleral thickness variations. For example, a study by Norman et al. showed that strain differences between PP and non-PP strains could be attributed to the thickness increases adjacent to the optic nerve head. This was also confirmed by Coudrillier et al., who measured scleral thickness and reported a different stiffness between the PP and mid-posterior scleral regions for both normal and glaucomatous eyes, which they attributed to the PP sclera having lower degree of fiber alignment and lower mechanical anisotropy compared with midposterior scleral regions.

To the author’s knowledge, Fazio et al. and Grytz et al. are two of the few research groups that have studied race/ethnicity as a factor in normal posterior scleral material properties using ESPI. In a study by Grytz et al., the authors...
report that the in-plane strain, which they describe as the strain tangent to the scleral shell surface, was found to be significantly lower in donors of AD compared with ED in the PP scleral region \((P = 0.015)\) for age groups of 20 to 90 and 23 to 73 years old, respectively, inflated from 5 to 45 mm Hg. The scleral strain results from Grytz et al.\(^{18}\) were not consistent with our results, which showed significantly lower PP scleral tensile strain for ED samples compared with AD samples at 15 mm Hg \((P = 0.024)\). Furthermore, the PP scleral compressive strains for the ED samples were significantly smaller than that of AD at 15 mm Hg \((P = 0.049;\) Fig. 4B). Discrepancies between our results and those of Grytz et al.\(^{18}\) could be attributed to differences in scleral thickness. Grytz et al.\(^{18}\) found no significant difference in scleral thickness between ED and AD eyes. However, racioethnic differences in scleral thickness could have contributed to the discrepancies that we observe between our different groups. Our laboratory is currently analyzing x-ray microcomputed tomography (\(\mu\)CT) images of the same scleral shells used in this study. Our preliminary results suggest that the sclera of ED eyes may be thicker than that of AD eyes, which would explain why the PP scleral strains of ED eyes would be smaller than that of AD eyes in our study. Grytz et al.\(^{18}\) used ultrasound to obtain thickness at 20 points for each eye \((n = 40\) for ED 20 to 90 years old, \(n = 22\) for

**Figure 7.** ONS volume (left), diameter (middle), and length (right) comparisons between racioethnic groups. Error bars denote 95% confidence interval.

**Figure 8.** Regression plots of mean E1 and E2 values as a function of ONS volume for all inflation pressures within each zone.
AD 23 to 73 years old). Our eyes are restricted to donors older than 50 years old, and the sample number in our study is lower than that of Grytz et al.\textsuperscript{18} The differences in thickness measurements between the racioethnic groups could be due to differences in measurement technique, age, the source, and/or the thickness variability. Furthermore, the eyes used by Grytz et al.\textsuperscript{18} had their optic nerves severed flush, which as was previously suggested could significantly alter scleral deformation. Hence, comparing these two results may not be appropriate without accounting for these main differences in method and eye geometry. Grytz et al.\textsuperscript{18} used thickness data in FE simulations to calculate the shear moduli and estimate stiffness. AD eyes were shown to have a higher shear modulus than those of ED.\textsuperscript{18} The study by Fazio et al.\textsuperscript{17} found that age-related stiffening was significantly greater in the PP sclera for donors of AD compared with donors of ED. Both these studies are consistent with our results where the slope of the tensile strain–pressure relationship of the ED eyes was significant, whereas that of AD eyes was not, suggesting that AD eyes deformed less with pressure (stiffer) compared with ED eyes. All axon counts were within ranges that have been considered normal in different studies in the literature.\textsuperscript{53–60} Both axon count and ONS volume were found to have significant effects on tensile and/or compressive strains in the different zones for at least one level of pressure. One possible concern may be that the biomechanical differences found between racioethnic groups could be due to racioethnic differences in ONS volume in our enucleated eyes. However, a statistical comparison on ONS volume (Fig. 7) showed no significant differences between racioethnic groups that would explain differences in tensile and compressive strains. Additionally, it is noteworthy that a significant correlation existed between axon count and strains at 45 mm Hg for normal nonglaucomatous eyes (Fig. 6). The fact that this correlation was found in normal eyes suggests that axon loss may be a normal ongoing process in humans. It remains to be seen if this relationship plays a role in the predisposition of a person to glaucomatous damage. Furthermore, it seems that as proximity to the ONS decreases, the effect of axon count on strain diminishes (from ONS to PP to non-PP). It also seems that a loss of axons is associated with increased deformation in the optic nerve, especially at higher pressures. Further research will be needed to ascertain whether axon loss itself plays a role in exacerbating the biomechanical insult in primary open angle glaucoma.

There are several limitations to the work presented here. Although our study does provide ONS and PP scleral strain information, the reported strain is only surface strain and does not provide information regarding the mechanical environment at the LC, which has been shown to be an important region as it relates to RGC damage. On that note, the authors also wish to recommend that interpretations of the ONS strain results reported here not be construed as “optic nerve deformations” but rather as strains of mostly the sheath surrounding the optic nerve and a small portion of the optic nerve that was exposed during enucleation. It should be noted that all optic nerve stumps most likely were swollen in our experiments compared with in vivo geometry. However, the well-controlled temperature and humidity environment and consistent preconditioning at all inflation pressures did not allow for any additional swelling during SDIC imaging. Unavoidable issues related to tissue edema and a somewhat nonphysiologic unconstrained severed nerve stump should serve as caution to the reader when interpreting these results. That being said, the fact that the presence (and volume) of the ONS volume did influence peripapillary scleral deformation indicate its inclusion may be important in future studies. Our study also focused entirely on assessing normal eyes and did not include glaucomatous eyes of any race or ethnicity. Future studies should be designed to study more thoroughly how glaucoma differentially affects posterior pole deformations in these populations, especially in regions close to the ONS. Furthermore, due to errors in nerve processing, our research group was unable to measure the axon counts for all the eyes used in S-DIC in this study. Therefore, the axon count relationship with strains is restricted to a subset of the eyes analyzed for strain. Last but not least, the authors wish to convey the limited sample sizes used in this study and recommend the reader interpret the presented results and conclusions with this in mind.

Although there is clinical evidence that both people of AD and HIS are more predisposed to developing glaucoma compared with people of ED, ocular biomechanical studies of AD and particularly HIS scleral shells are not common in the literature. Our results showed that those of HIS exhibited PP scleral tensile strains that were significantly different than those of ED at 15 mm Hg. HIS tensile strains in the ONS region did not have a significant relationship with pressure, whereas ED eyes did. However, the behavior of tensile and compressive strains in both PP and non-PP scleral zones did not differ significantly between ED and HIS eyes. Preliminary results from our ongoing research (data not shown) suggest that ED eyes may also have significantly thicker sclera that of HIS eyes. The lack of consistency between HIS and AD strain patterns compared with ED suggests that the mechanism of higher prevalence of OAG in AD eyes could be different than that of HIS. More comprehensive studies are necessary to investigate possible alternative explanations to prevalence of ocular disease in both people of AD and HIS. In addition, it is important to note that these results show increasing pressure above 15 mm Hg did not result in additional strain in the PP for AD eyes. This may be due to the nonlinear stress–strain material behavior presenting a more prominent strain-stiffening effect in that zone for AD eyes, which is considered a higher-risk group. In future work, it would be interesting to explore whether glaucomatous eyes demonstrate this type of behavior and how this may play a role in the incidence or progression of glaucoma. Another prominent result of our study was that the ONS strains were much higher than the scleral strains at every IOP for all racioethnic groups (Fig. 4). Although it should be noted that the strains measured on the ONS in this study should not be interpreted as identical to those occurring in vivo, this result strongly confirms that there is an IOP-dependent mechanical link between the peripapillary sclera and the dural sheath. This link may include bending modes of the sclera/dural sheath and may contribute to the overall biomechanical environment of the optic nerve head. Future research will need to be performed to more directly test the influence of the dural sheath on the optic nerve head mechanical response in OAG.

Our study is the first determine strains for all zones and regions of donors of HIS. Overall, we found that the HIS PP scleral tensile strain relationship with pressure were more similar to those of ED than AD for tensile strains. Our study is also the first to map the strain behavior of the human ONS for any racioethnic group using S-DIC. We were also the first to find a significant relationship between tensile and compressive strains and ONS volume and axon count. This shows that studying the strains of ONS, which serves as the sheath for nerves, may be an important consideration in biomechanical evaluation of the posterior pole. The results in this study indicate that keeping ONSs attached to their scleral shells, while accounting for volume and axon count, may be necessary to conduct a more complete ocular biomechanical analysis. Future studies by our group will combine geometry data collected via µCT with strain data collected from this study.
to develop an inverse finite element model, which will investigate if the strain differences quantified in this study correspond to true mechanical property differences.

Acknowledgments

The authors thank the Alabama Eye Bank, Banner Sun Health Research, Donor Network of Arizona, Illinois Eye Bank, Michigan Eye Bank, and San Diego Eye Bank for help in obtaining the human donor tissue. The authors thank Melanie Brucks and Scott Harrison for statistical and postprocessing contributions in this paper and Thomas Cahir and Forest Danford for contributions in the development of the axon counting method. Finally, the authors thank Marianne Cilluffo at the University of California Los Angeles and Ben Christensen at the University of Utah for help in nerve sample preparation and staining.

Supported by National Eye Institute, National Institutes of Health, Award R01EY020890 (PI: JPVC). Additional support for EAT was also provided by National Institute of Biomedical Imaging and Bioengineering Award EB005392. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Disclosure: E.A. Tamimi, None; J.D. Pyne, None; D.K. Muli, None; K.F. Axman, None; S.J. Howerton, None; M.R. Davis, None; C.A. Girkin, None; J.P. Vande Geest, None

References

1. Weinhre BN, Aung T, Medeiros FA. The pathophysiology and treatment of glaucoma: a review. JAMA. 2014;311:1901–1911.

2. Quigley HA, Broman AT. The number of people with glaucoma worldwide in 2010 and 2020. Br J Ophthalmol. 2006;90:262–267.

3. Downs JC, Roberts MD, Burgoyne CF. The mechanical environment of the optic nerve head in glaucoma. Optometry Vis Sci. 2008;85:425–435.

4. Rudnicka AR, Mt-Isa S, Owen CG, Cook DG, Ashby D. Variations in primary open-angle glaucoma prevalence by age, gender, and race: a Bayesian meta-analysis. Invest Ophthalmol Vis Sci. 2006;47:4254–4261.

5. Anderson DR, Hendrickson AE. Effect of intraocular pressure on rapid axoplasmic transport in monkey optic nerve. Invest Ophthalmol Vis Sci. 1974;13:771–783.

6. Minckler DS, Bunt AH, Johnson GW. Orthograde and retrograde axoplasmic transport during acute ocular hypertension in the monkey. Invest Ophthalmol Vis Sci. 1977;16:426–441.

7. Bellezza AJ, Rintalan CJ, Thompson HW, Downs JC, Hart RT, Burgoyne CF. Deformation of the lamina cribrosa and anterior scleral canal wall in early experimental glaucoma. Invest Ophthalmol Vis Sci. 2005;46:623–637.

8. Burgoyne C. The morphological difference between glaucoma and other optic neuropathies. J Neuro-Optom Vis. 2015;35(suppl 1):S8–S21.

9. Yang H, Thompson H, Roberts MD, Sigal IA, Downs JC, Burgoyne CF. Deformation of the early glaucomatous monkey optic nerve head connective tissue after acute IOP elevation in 3-D histomorphometric reconstructions. Invest Ophthalmol Vis Sci. 2011;52:345–363.

10. Yang H, Downs JC, Sigal IA, Roberts MD, Thompson H, Burgoyne CF. Deformation of the normal monkey optic nerve head connective tissue after acute IOP elevation within 3-D histomorphometric reconstructions. Invest Ophthalmol Vis Sci. 2009;50:5785–5799.

11. Yang H, Ren R, Lockwood H, et al. The connective tissue components of optic nerve head cupping in monkey experimental glaucoma part I: global change. Invest Ophthalmol Vis Sci. 2015;56:7661–7678.

12. Downs JC, Roberts MD, Burgoyne CF. Mechanical environment of the optic nerve head in glaucoma. Optom Vis Sci. 2008;85:425–435.

13. Sigal IA, Flanagan JG, Tertinegg I, Ethier CR. Modeling individual-specific human optic nerve head biomechanics. Part I: IOP-induced deformations and influence of geometry. Biomech Model Mechanobiol. 2009:8:85–98.

14. Burgoyne CF, Downs JC, Bellezza AJ, Suh JK, Hart RT. The optic nerve head as a biomechanical structure: a new paradigm for understanding the role of IOP-related stress and strain in the pathophysiology of glaucomatous optic nerve head damage. Progr Retinal Eye Res. 2005;24:39–73.

15. Sigal IA, Flanagan JG, Tertinegg I, Ethier CR. Modeling individual-specific human optic nerve head biomechanics. Part II: influence of material properties. Biomech Model Mechanobiol. 2009;8:99–109.

16. Bellezza AJ, Hart RT, Burgoyne CF. The optic nerve head as a biomechanical structure: initial finite element modeling. Invest Ophthalmol Vis Sci. 2000;41:2991–3000.

17. Fazio MA, Grytz R, Morris JS, Bruno L, Girkin CA, Downs JC. Human scleral structural stiffness increases more rapidly with age in donors of African descent compared to donors of European descent. Invest Ophthalmol Vis Sci. 2014;55:7189–7198.

18. Grytz R, Fazio MA, Libertau X, et al. Age- and race-related differences in human scleral material properties. Invest Ophthalmol Vis Sci. 2014;55:8163–8172.

19. Yan D, McPheeters S, Johnson G, Utzinger U, Vande Geest JP. Microstructural differences in the human posterior sclera as a function of age and race. Invest Ophthalmol Vis Sci. 2011;52:821–829.

20. Quigley HA, West SK, Rodriguez J, Munoz B, Klein R, Snyder R. The prevalence of glaucoma in a population-based study of Hispanic subjects: Proyecto VER. Arch Ophthalmol. 2001;119:1819–1826.

21. Coudrillier B, Tian J, Alexander S, Myers KM, Quigley HA, Nguyen TD. Biomechanics of the human posterior sclera: age- and glaucoma-related changes measured using inflation testing. Invest Ophthalmol Vis Sci. 2012;53:1714–1728.

22. Boyce BL, Grazier JM, Jones RE, Nguyen TD. Full-field deformation of bovine cornea under constrained inflation conditions. Biomaterials. 2008;29:3896–3904.

23. Coudrillier B, Boote C, Quigley HA, Nguyen TD. Scleral anisotropy and its effects on the mechanical response of the optic nerve head. Biomech Model Mechanobiol. 2013;12:941–963.

24. Cruz Perez B, Pavlatos E, Morris HJ, et al. Mapping 3D strains with ultrasound speckle tracking: method validation and initial results in porcine scleral inflation. Ann Biomed Eng. 2016;44:2302–2312.

25. Cruz Perez B, Tang J, Morris HJ, et al. Biaxial mechanical testing of posterior sclera using high-resolution ultrasound speckle tracking for strain measurements. J Biomech 2014;47:1151–1156.

26. Morris HJ, Tang J, Cruz Perez B, et al. Correlation between biomechanical responses of posterior sclera and IOP elevations during micro intraocular volume change. Invest Ophthalmol Vis Sci. 2013;54:7215–7222.

27. Pavlatos E, Perez BC, Morris HJ, et al. Three-dimensional strains in human posterior sclera using ultrasound speckle tracking. J Biomechanical Eng. 2016;138:021015.

28. Tang J, Liu J. Ultrasonic measurement of scleral cross-sectional strains during elevations of intraocular pressure: method validation and initial results in posterior porcine sclera. J Biomechanical Eng. 2012;134:091007.

29. Fazio MA, Grytz R, Bruno L, et al. Regional variations in mechanical strain in the posterior human sclera. Invest Ophthalmol Vis Sci. 2012;53:5326–5335.
30. Girard MJ, Downs JC, Bottlang M, Burgoyne CF, Suh JK. Peripapillary and posterior scleral mechanics: part II: experimental and inverse finite element characterization. J Biomechanical Eng. 2009;131:051012.

31. Girard MJ, Suh JK, Bottlang M, Burgoyne CF, Downs JC. Scleral biomechanics in the aging monkey eye. Invest Ophtalmol Vis Sci. 2009;50:5226–5237.

32. Fazio MA, Grytz R, Morris JS, et al. Age-related changes in human peripapillary scleral strain. Biomech Model Mechanobiol. 2014;13:551–563.

33. Grytz R, Fazio MA, Girard MJ, et al. Material properties of the posterior human sclera. J Mechanical Behav Biomed Mater. 2014;29:602–617.

34. Wang Y, Thomas D, Zhang P, Yokota H, Yang L. Whole field strain measurement on complex surfaces by digital speckle pattern interferometry. Mater Eval. 2008;66:507–512.

35. Wang Y, Thomas D, Zhang P, Yokota H, Yang L. Whole field strain measurement on complex surfaces by digital speckle pattern interferometry. OPTICE. 2003;42:1257–1266.

36. Ruiz PD. Evaluation of a scale-space filter for speckle noise reduction in electronic speckle pattern interferometry. OPTICE. 1998;37:5287.

37. Pyne JD, Genovese K, Csaletto L, Vande Geest JP. Sequential-digital image correlation for mapping human posterior sclera and optic nerve head deformation. J Biomech Eng. 2014;136:021002.

38. Sigal IA, Flanagan JG, Ethier CR. Factors influencing optic nerve head biomechanics. Invest Ophtalmol Vis Sci. 2005;46:4189–4199.

39. Sigal IA, Flanagan JG, Tertinegg I, Ethier CR. Finite element modeling of optic nerve head biomechanics. Invest Ophtalmol Vis Sci. 2004;45:4378–4387.

40. Sigal IA, Flanagan JG, Tertinegg I, Ethier CR. Reconstruction of human optic nerve heads for finite element modeling. Technol Health Care. 2005;13:313–329.

41. Sigal IA, Flanagan JG, Tertinegg I, Ethier CR. Predicted extension, compression and shearing of optic nerve head tissues. Exp Eye Res. 2007;85:312–322.

42. Rhodes LA, Huisingsh C, Johnstone J, et al. Variation of laminar depth in normal eyes with age and race. Invest Ophtalmol Vis Sci. 2014;55:8123–8133.

43. Fazio MA, Johnstone JK, Smith B, Wang L, Girkin CA. Displacement of the lamina cribrosa in response to acute intraocular pressure elevation in normal individuals of African and European descent. Invest Ophtalmol Vis Sci. 2016;57:3351–3359.

44. Coudrillier B, Pijanka J, Jefferys J, et al. Effects of age and diabetes on scleral stiffness. J Biomech Eng. 2015;137:0710071–07100710.

45. Reynaud J, Cull G, Wang L, et al. Automated quantification of optic nerve axons in primate glaucomatous and normal eyes—method and comparison to semi-automated manual quantification. Invest Ophtalmol Vis Sci. 2012;53:2951–2959.

46. Teixeira LBC, Buhr KA, Bowie O, et al. Quantifying optic nerve axons in a cat glaucoma model by a semi-automated targeted counting method. Molec Vis. 2014;20:376–385.

47. Templeton GF. A two-step approach for transforming continuous variables to normal: implications and recommendations for IS research. Commun Assoc Inform Syst. 2011;28:41–58.

48. Eliaghi A, Flanagan JG, Simmons CA, Ethier CR. Effects of scleral stiffness properties on optic nerve head biomechanics. Ann Biomed Eng. 2010;38:1586–1592.

49. Norman RE, Flanagan JG, Sigal IA, Rausch SM, Tertinegg I, Ethier CR. Finite element modeling of the human sclera: influence on optic nerve head biomechanics and connections with glaucoma. Exp Eye Res. 2011;93:4–12.

50. Coudrillier B, Pijanka J, Jefferys J, et al. Collagen structure and mechanical properties of the human sclera: analysis for the effects of age. J Biomech Eng. 2015;137:e0131396.

51. Coudrillier B, Pijanka JK, Jefferys JL, et al. Glaucoma-related changes in the mechanical properties and collagen microarchitecture of the human sclera. Plos One. 2015;10:2959.

52. Danford FL, Yan D, Dreier RA, Cahir TM, Girkin CA, Vande Geest JP. Differences in the region- and depth-dependent microstructural organization in normal versus glaucomatous human posterior sclera microstructural organization of human posterior sclera. Invest Ophtalmol Vis Sci. 2013;54:7922–7932.

53. Balazsi AG, Rootman J, Drance SM, Schulzer M, Douglas GR. The effect of age on the nerve fiber population of the human optic nerve. Am J Ophthalmol. 1984;97:760–766.

54. Johnson BM, Miao M, Sadun AA. Age-related decline of human optic nerve axon populations. AGE. 1987;10:5–9.

55. Jonas JB, Müller-Bergh JA, Schlötzer-Schrehardt UM, Naumann GO. Histomorphometry of the human optic nerve. Invest Ophtalmol Vis Sci. 1990;31:736–744.

56. Kerrigan-Baumrind LA, Quigley HA, Pease ME, Kerrigan DF, Mitchell RS. Number of ganglion cells in glaucoma eyes compared with threshold visual field tests in the same persons. Invest Ophtalmol Vis Sci. 2000;41:741–748.

57. Kupfer C, Chumbley L, Downner JC. Quantitative histology of optic nerve, optic tract and lateral geniculate nucleus of man. J Anat. 1967;101:393–401.

58. Mikelberg FS, Yidegiligne HM, Schulzer M. Optic nerve axon count and won diameter in patients with ocular hypertension and normal visual fields. Ophtalmology. 1995;102:342–348.

59. Potts AM, Hodges D, Shelman CB, Fritz KJ, Levy NS, Mangnall Y. Morphology of the primate optic nerve. I. Method and total fiber count. Invest Ophtalmol Vis Sci. 1972;11:980–988.

60. Repka MX, Quigley HA. The effect of age on normal human optic nerve fiber number and diameter. Ophtalmology. 1989;96:26–32.