A biliverdin-binding cyanobacteriochrome from the chlorophyll d–bearing cyanobacterium Acaryochloris marina

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Cyanobacteriochromes (CBCRs) are linear tetrapyrrole-binding photoreceptor proteins in cyanobacteria that absorb visible and near-ultraviolet light. CBCRs are divided into two types based on the type of chromophore they contain: phycocyanobilin (PCB) or phycoviolobilin (PVB). PCB-binding CBCRs reversibly photoconvert at relatively long wavelengths, i.e., the blue-to-red region, whereas PVB-binding CBCRs reversibly photoconvert at shorter wavelengths, i.e., the near-ultraviolet to green region. Notably, prior to this report, CBCRs containing biliverdin (BV), which absorbs at longer wavelengths than do PCB and PVB, have not been found. Herein, we report that the typical red/green CBCR AM1_1557 from the chlorophyll d–bearing cyanobacterium Acaryochloris marina can bind BV almost comparable to PCB. This BV-bound holoprotein reversibly photoconverts between a far red light–absorbing form (Pfr, λmax = 697 nm) and an orange light–absorbing form (Po, λmax = 622 nm). At room temperature, Pfr fluoresces with a maximum at 730 nm. These spectral features are red-shifted by 48–77 nm compared with those of the PCB-bound domain. Because the absorbance of chlorophyll d is red-shifted compared with that of chlorophyll a, the BV-bound AM1_1557 may be a physiologically relevant feature of A. marina and is potentially useful as an optogenetic switch and/or fluorescence imager.

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absorbing metastable state (Pg; C15-E PCB)\textsuperscript{8,20–24}, whereas the green/red types photoconvert between a green light–absorbing thermostable state (Pg; C15-Z PCB) and a red light–absorbing metastable state (Pr; C15-E PCB).

We recently determined the crystal structure of the red/green CBCR AnPixJg2 Pr and found that its chromophore and tertiary structure are quite similar to those of phytochrome Pr, although the detailed chromophore-protein interaction is distinctive\textsuperscript{25,26}. Conversely, the green light–absorbing mechanism of AnPixJg2 must be quite different from that of the far-red light–absorbing mechanism of phytochromes. Although hydration and ring D distortion may occur during photoconversion, their color-tuning mechanisms remain unclear\textsuperscript{25,27}. For green/red CBCRs, color tuning depends on PCB protonation in Pr and deprotonation in Pg, irrespective of the C15-Z/E configuration\textsuperscript{19}.

CBCRs have been found only in cyanobacteria. Although cyanobacteria form a monophyletic clade, they have extensively diversified morphology, habitats, and photosynthetic properties. Among them, \textit{Acaryochloris marina} is unique with respect to its photosynthetic pigments\textsuperscript{28,29}. As its photosynthetic pigment, \textit{A. marina} contains chlorophyll (Chl) \textit{d}—which has an absorbance maximum at $\sim$710 nm—instead of Chl \textit{a}—which has an absorbance maximum at $\sim$675 nm. Photochemical reaction center pigments for photosystem II and photosystem I, P680 and P700, in more typical cyanobacteria...
bacteria are substituted by P713 and P740, respectively, in A. marina. A. marina produces Chl d-binding photosynthetic antenna complexes, such that A. marina uses red-shifted light sources for photosynthesis, which normal cyanobacteria and algae do not. Given these observations, we hypothesized that A. marina might also possess CBCR photoreceptors in its genome that absorb light of longer wavelengths than do typical cyanobacteria.

Optogenetics and bio-imaging are now powerful techniques for regulating and monitoring cellular activities, and so various light-absorbing proteins are applied to these techniques. Particularly, proteins absorbing long-wavelength light are needed for penetration to animal tissues. In this context, it is meaningful to discover or artificially synthesize CBCRs that absorb long-wavelength light. Here, we focused on biliverdin (BV, Fig. 1B) chromophore that absorbs longer wavelength light than PCB. BV has been reported to covalently ligate to bacterial phytochromes (Bphs) but not to known CBCRs. The non-photoconvertible long wavelength form of CBCR is suitable for the stable fluorescent probe. We selected red/green CBCR to look for BV-binding, because its Pr form is a thermostable form.

We report herein that a recombinant red/green CBCR from A. marina MBIC11017 effectively binds BV to form a photoconvertible complex that absorbs and fluoresces at longer wavelengths than does the PCB-binding complex, suggesting that it may be relevant to far-red light-responsive feature of A. marina and suitable as an optogenetic switch or fluorescent imaging tool.

Results

Sequence Characteristics of AM1_1557g2. AM1_1557 is a typical bacterial two-component signal-transduction protein of 883 amino acid residues, two GAF domains, one His kinase (HK) domain, and one response regulator (RR) domain (Fig. 1C). The second GAF domain (AM1_1557g2, residues 220–364) is a red/green CBCR according to our sequence alignment (Fig. S1) and cluster analysis (Fig. 1D). Its sequence has 50% residue identity with that of AnPixJg2 and contains residues that are highly conserved in red/green CBCRs, i.e., Trp272, Asp274, and Tyr335 (Fig. S1).

Photoconversion of AM1_1557g2-PCB and –BV. AM1_1557g2 expressed in a PCB- or BV-producing E. coli was purified to near homogeneity (Fig. 1E, CBB). PCB and BV covalently bound AM1_1557g2 judging from the Zn-dependent fluorescence assay (Fig. 1E, +Zn). AM1_1557g2-PCB reversibly photoconverts between a red light–absorbing form (Pr; absorbance maximum, 649 nm) and a green light–absorbing form (Pγ; absorbance maximum, 545 nm; Fig. 2A). This photoconversion is quite similar to that of AnPixJg2, a typical red/green CBCR. The blue–pink color change of a solution of AM1_1557g2-PCB is clearly seen (Fig. 2G).

Conversely, AM1_1557g2-BV reversibly photoconverts between a far-red light–absorbing form (Pfr; absorbance maximum, 697 nm) and an orange light–absorbing form (Po; absorbance maximum, 622 nm; Fig. 2B). The green–blue color change of a solution of AM1_1557g2-BV is also clearly seen (Fig. 2H). The absorbance maxima of AM1_1557g2-BV Pr and Po are 48- and 77-nm red-shifted compared with those of AM1_1557g2-PCB Pr and Pg. The (Pr – Po) difference spectrum has maxima at 699 and 378 nm, and a minimum at 606 nm, whereas the (Pr – Pg) difference spectrum has maxima at 649 and 351 nm, and a minimum at 540 nm (Fig. 2C). Because the maximum absorbances of free BV and PCB in solution are at 670 and 610 nm, respectively, the difference in the absorbance maxima of the two AM1_1557g2 forms roughly corresponds to the difference in the absorbances of the free chromophores. Isosbestic points are present in spectra recorded during the photoconversion processes of AM1_1557g2-PCB (584 and 449 nm) and AM1_1557g2-BV (652 and 480 nm), indicating no noticeable heterogeneity or intermediates (Fig. 3). Further, photoconversion could be repeated many times without appreciable deterioration of the spectra.

Chromophore Species, Their Configurations and Dark Reversion Kinetics. To conclusively identify the chromophore species and their configurations, spectra were obtained for acid-denatured AM1_1557g2-PCB and –BV. Absorption maxima of denatured AM1_1557g2-PCB Pr and Pg were observed at ~664 and ~594 nm, respectively (Fig. 2D, Fig. S2). Absorption maxima of denatured AM1_1557g2-BV Pr and Po were observed at ~700 and ~620 nm, respectively (Fig. 2E, Fig. S2). These absorbance maxima of AM1_1557g2-BV Pr and Po forms are about 50–60 nm red-shifted compared to those of AM1_1557g2-PCB Pr and Pg forms. Irradiation of denatured Pg and Po with white light resulted in red shift of the absorption spectra (Fig. S3). Further, the spectral difference between denatured AM1_1557g2-BV Pr and Po forms is identical to those of denatured PaBph-BV (bacterial phytochrome that covalently binds BV from Pseudomonas aeruginosa) Pr and Pr forms (Fig. S4), indicating that AM1_1557g2 Pr is the thermostable state containing 15Z-BV, whereas its metastable state is Po containing 15E-BV. In terms of their chromophore configurations, AM1_1557g2-BV Pr and Po correspond to AM1_1557g2-PCB Pr and Pg, respectively. We measured dark reversion kinetics of the thermostable states at room temperature. Unexpectedly, AM1_1557g2-BV showed very slow dark reversion with half-life of 93 hours, whereas AM1_1557g2-PCB showed quick dark reversion with half-life of 1 hour (Fig. 4).

Covalent Attachment of BV to AM1_1557g2 via Cys304. Recently, we solved the crystal structure of AnPixJg2 Pr in which Cys321 within its GAF domain is ligated to PCB C31 (Fig. S5). Our Zn-blot study indicates that AM1_1557g2 is covalently bound to BV (Fig. 1E). AnPixJg2 Cys321 corresponds to AM1_1557g2 Cys304 according to our sequence alignment (Fig. S1). To show that AM1_1557g2 Cys304 is covalently bound to the chromophore, we prepared the mutant C304A with an Ala substituted for Cys304. The Zn blot of C304A indicated that BV was not covalent bound to C304A (Fig. 5A). In addition, C304A did not absorb visible light (Fig. 5B). These results strongly indicate that AM1_1557g2 covalently binds BV via Cys304. This is the first report showing that the conserved CBCR GAF Cys can covalently ligate BV.

Fluorescence Spectroscopy. Room temperature fluorescence spectra of the thermostable states, AM1_1557g2-PCB Pr and -BV Pfr, were measured to evaluate their potential as fluorescence imagers. AM1_1557g2-BV Pr fluoresces with a maximum at 730 nm, whereas AM1_1557g2-PCB Pr fluoresces with a maximum at 676 nm (Fig. 6). The fluorescence maximum of AM1_1557g2-BV Pfr is red-shifted by 54 nm compared with that of AM1_1557g2-PCB, a red shift quite similar to that found for their absorption spectra maxima. Fluorescence quantum yields of the Pr and Pfr form were 1.7% and 0.3%.

The fluorescence of AM1_1557g2-PCB, AM1_1557g2-BV, and free PCB and BV were directly observed under a fluorescence stereomicroscope. AM1_1557g2-PCB Pr intensely fluoresced, whereas free PCB did not (Fig. 7A and B). In addition, we detected a change in fluorescent intensity from Pr during the photoconversion of Pr and Pg (Fig. 7C and Movie S1). The solution of AM1_1557g2-PCB was constantly red-light irradiated, resulting in photoconversion of Pr to Pg concomitant with a fluorescence decrease to almost the background level. Upon green light irradiation Pg photoconverted to Pr, and the fluorescence largely increased.

Similarly, the fluorescence of AM1_1557g2-BV Pfr did not, but that of free BV, was clearly observed (Fig. 7D and E). In addition, we detected a change in fluorescent intensity from Pfr during the Pfr to Po photoconversion (Fig. 7F and Movie S2). The solution of AM1_1557g2-BV was constantly irradiated with far-red light (FRL, 710/75 nm), resulting in photoconversion of Pfr to Po con-
comitant with a decrease in fluorescence. When irradiated with orange light, Po photoconverted to Pfr, and the fluorescence largely increased. The Pfr fluorescence was not completely abolished by far-red irradiation possibly due to incomplete photoconversion. Thus, irradiation by red-shifted far-red light (FRL-2, 720/40 nm) further decreased its fluorescence (Fig. S6 and Movie S3). In both cases, photoconversion was almost complete within 1 min and repetitive photoconversion did not affect the cyclic increase and decrease of fluorescence intensity (Fig. 7C and F).

Discussion

In this study, we prepared the BV-binding CBCR GAF domain, AM1_1557g2-BV, which is the first time a CBCR GAF domain has been shown to bind BV. AM1_1557g2 efficiently binds BV via the canonical GAF Cys304 and reversibly photoconverts in its BV form between Pfr with an absorbance maximum at 697 nm and Po with an absorbance maximum at 622 nm. Furthermore, at room temperature, Pfr fluoresces with a maximum at 730 nm. These experiments suggest that AM1_1557 binds BV in vivo, which would make it highly useful as an optogenetic switch and/or fluorescent imaging tool.

A. marina is unusual cyanobacterium, in that it possesses Chl d, instead of Chl a, as its main photosynthetic pigment. Chl d absorbs light at ~710 nm, whereas Chl a absorbs light at ~675 nm, suggesting that for effective photosynthesis A. marina may need to absorb light of longer wavelengths than do cyanobacteria that possess only Chl a. Because the maximum absorbance of BV is red-shifted by ~60 nm compared with that of PCB, we hypothesized that A. marina may also possess a CBCR(s) that can bind BV instead of PCB to sense light of longer wavelengths. We therefore focused on the red/green CBCR, selected AM1_1557 from A. marina because its GAF domain is highly similar to the representative AnPixJg2, and obtained the recombinant BV-binding GAF domain, AM1_1557g2.

Here, we estimated binding efficiency of BV to AM1_1557g2 in comparison with that of PCB based on fluorescence intensities on SDS-PAGE gel (Fig. 1E), native protein absorptions (Fig. 2A and B) and denatured protein absorptions (Fig. 2D and E) that are standardized by free PCB and BV data (Fig. S7). By these three different
calculations, the binding efficiency of BV was estimated at approximately 55% (52–57%) in comparison with PCB. Because the structure of BV differs from that of PCB at C18 (an ethylidene vs. a vinyl moiety, respectively) and at ring A, which covalently binds to the apoprotein (Fig. 1A and B), it is somewhat surprising that the two chromophores bind AM1_1557g2 similarly. Specifically, the difference in the ring A substituents has been assumed to be critical for chromophore selectivity. Crystal structures of Bph-BV40, and Cph1-41 and CBCR-PCBs25 clearly show different orientations of the canonical Cys-ring A covalent bond, which may reflect different positional reactivities of the double bond caused by the ethylidene and vinyl groups. The N-terminal conserved Cys of Bph covalently ligates C30, which in the free state of BV forms a double bond with C31. Conversely, PCB has a double bond between C31 and C3 and covalently ligates the conserved Cys within the GAF domain of Cph1 and CBCRs via C31. Because there have been no reports that a Cys within a CBCR GAF domain covalently ligates BV, it is difficult to predict whether the covalent bonding site of the ring A to Cys304 is C31 or C30. However, based on the AnPixJg2 structure, Cys304 is more likely to be physically near C30 of BV rather than C31, suggesting that C30 is involved in the covalent bond (Fig. S5). On the other hand, red/green CBCR, AnPixJg2, did not bind BV. For those GAF domains, a different orientation of the ring D may control chromophore binding, i.e., the BV C18 ethylidene of ring D may sterically interfere with BV binding, whereas such steric hindrance in AM1_1557g2 may be absent because the local environment surrounding ring D is different. In this context, residues unique to AM1_1557g2 surrounding ring D would be possible determinants for chromophore selectivity. Although we could not detect obvious differences in residues that directly interact with ring D among AM1_1557g2 and the other red/green CBCRs, notably, AnPixJg2 Asn354 that is within van der Waals distance of Tyr352 (Tyr335 in AM1_1557g2) (Fig. S5), which interacts with the ring D carbonyl, is replaced with a Leu in AM1_1557g2 (Leu337, Fig. S1). This replacement may affect the position of Tyr335, resulting in a different arrangement of ring D within its binding pocket. To examine role of Leu337 in the chromophore selectivity, we replaced Leu337 with an Asn to form AM1_1557g2_L337N. As a result, binding efficiency of L337N to BV was approximately half of that of the wild type protein, whereas binding efficiencies of L337N and wild type proteins to PCB were almost same (Fig. 2 and Fig. S8). This result indicates that Leu337 is a major factor for potential to bind BV. The L337N protein still retains a potential to ligate BV. Additional residues may also be involved in chromophore selectivity.

This study clearly demonstrated that AM1_1557g2 Cys304 can covalently ligate BV as well as PCB, suggesting that BV may also bind AM1_1557 in vivo. If such a protein exists in vivo, it would sense far-red light, which is more efficiently absorbed by Chl d than by Chl a. Notably, the effect of far-red light on the physiology of A.

Figure 3 | Presence of isosbestic points in the spectra of AM1_1557g2-PCB and -BV obtained during photoconversion. (A) Absorption spectra AM1_1557g2-PCB acquired during its (Pr – Pg) photoconversion. (B) Absorption spectra of AM1_1557g2-BV acquired during its (Pfr – Po) photoconversion. Spectral measurements after irradiation with light intensity of 100 μmol m⁻²s⁻¹ for approximately 3-to-30 s were performed and representative spectra are shown.

Figure 4 | Dark reversion kinetics of Pr and Pfr forms of AM1_1557g2-PCB and AM1_1557g2-BV, respectively, at room temperature. Absorbances at 649 nm and 696 nm were monitored for AM1_1557g2-PCB and AM1_1557g2-BV, respectively.
A. marina has been studied. Kiss et al. reported that expression of *psbE2* and *psbD3*, extra copy genes for photosystem II unique to *A. marina*, is induced under far-red light (720 nm). *A. marina* MBIC11017 accumulates phycobilisome under orange light irradiation (625 nm), but downregulates phycobiliprotein expression under far-red light irradiation (720 nm). These far-red light–inducible and far-red/orange light–reversible photo-acclimation processes are compatible with the far-red/orange photoconversion property of AM1_1557g2-BV, suggesting that AM1_1557g2-BV may be involved in such photo-acclimation process(es).

Figure 5 | Replacement of Cys304 with Ala in AM1_1557g2. (A) CBB-stained C304A after SDS-PAGE (CBB) and in-gel Zn-dependent fluorescence assay (+Zn) showing C304A does not bind BV. (B) Absorption spectra of AM1_1557g2-BV Pfr (broken line) and C304A (solid line).

Figure 6 | Fluorescence spectra of AM1_1557g2-PCB Pr (broken line) and AM1_1557g2-BV Pfr (solid line). Excitation maxima: 590 and 660 nm, respectively.
Figure 7 | Fluorescent microscopy. (A) Transmission and (B) fluorescence images of solutions of (a) free PCB and (b) AM1_1557g2-PCB. PCB was obtained from Frontier Scientific. As a control, 50 μM PCB in Buffer A was used. (C) Change in fluorescent intensity of AM1_1557g2-PCB during (Pr – Pg) photoconversion. Fluorescence from Pr was detected when AM1_1557g2-PCB was irradiated with red light (RL). Red-light irradiation was continuous throughout the experiment. Photoconversion was induced by intermittent green light (GL) irradiation. The fluorescence intensities of AM1_1557g2-PCB and free PCB are plotted against time. (D) Transmission and (E) fluorescence images of solutions of (a) free BV and (b) AM1_1557g2-BV. BV was obtained from Frontier Scientific. As a control, 50 μM BV in Buffer A was used. (F) Change in the fluorescent intensity of AM1_1557g2-BV during (Pfr – Po) photoconversion. Fluorescence from Pfr was detected when AM1_1557g2-BV was irradiated with far-red light (FRL). Far-red light irradiation was continuous throughout the experiment. Photoconversion was induced by intermittent orange-light (OL) irradiation. The fluorescence intensities of AM1_1557g2-BV and free BV are plotted against time.
alizing deep tissues in mammals using GFPs and rhodopsins are problematic because the absorbance of hemoglobin and skin melanin interferes with the analyses. Instead, proteins that can be used for these purposes should have absorption maxima within the far-red to the near-infrared spectral region (650–900 nm), as light absorbance within this region by mammalian tissues is negligible. Consequently, red/far-red light sensitive photochromes have been found for these purposes.

Among various photochromes, BV-binding Bphs have been studied for their potential as optogenetic and bio-imaging tools because BV is present in mammalian cells and absorbs light of the longest wavelengths found for linear tetrapyrrole molecules. BV-binding Bphs, however, have drawbacks as their chromophore-binding unit consisting of the three domains is large and as they have a tendency to polymerize. Conversely, CBCRs have the advantage of having a compact chromophore-binding unit composed of only a GAF domain (25 kDa) and do not polymerize. Further, BV-binding CBCR GAF domain discovered in this study provides large advantages for application in animal deep tissues. Because the light signals acquired by CBCR GAF domains are transferred to their enzymatic domains, e.g., the His kinase domain of AM1_1557, optogenetic switches can be created by fusing an input AM1_1557 GAF domain and an enzymatic domain. Detection of clear isoosbestic points in the spectrum of AM1_1557g2-BV during photoconversion and its ability to repetitively and reversibly photoconvert without appreciable deterioration of its spectral ensures its reliable performance as a photoconvertible switch.

AM1_1557g2-BV Pfr emits fluorescence with a maximum at 730 nm, which is a wavelength comparable with or slightly longer than those of Bph-derived IRFP (near-infrared fluorescent probe) and IFP (infrared fluorescent protein). Further, the repetitive photoconversion do not affect its fluorescent properties. These characteristics may be advantageous for super-resolution imaging. AM1_1557g2-BV as a fluorescent probe may also be useful for plant-cell studies. Plants possess a large quantity of Chl a that absorbs red light and emit light at ~680 nm, which would largely interfere with a red light–absorbing optogenetic switch or bio-imaging probe. Conversely, the far-red–absorbing property of AM1_1557g2-BV would be immune to the spectral properties of Chl. The quantum yield of Pfr form of AM1_1557g2-BV is not so high (0.3%), but is comparable to those of the native photochromes. In the case of photochromes, random and site-directed mutagenesis succeeded in elevating the quantum yields to ~5–10% and so we would expect similar improvement of AM1_1557g2-BV by introducing replacements of amino acid residues. The crystal structure of AnPixg2 Pr has been solved, which enables further development and improvement of BV-binding CBCRs. So, based on this structural information, we are now performing further analyses such as mutagenesis for stable and bright fluorescence probes and chimeric protein construction for useful light switches.

Methods

In silico Characterization of AM1_1557. The domain composition of AM1_1557 was determined using SMART (http://smart.embl-heidelberg.de).

Alignment and phylogenetic clustering of CBCR and photochrome GAF domain sequences were performed by CLUSTAL X.

Plasmid Construction. The nucleotide sequence of AM1_1557g2 was cloned into pET28a (Novagen) using the In-Fusion HD Cloning kit (Takara). The DNA fragment corresponding to AM1_1557g2 was PCR amplified using the synthetic primers: 5'-GGATCCCATATGTCGCTATCTC-3' (forward primer) and 5'-CTCGAATTCCGAGATCTCTAGGCTGTTATTTTGCTCT-3' (reverse primer), genomic DNA from A. marina MB11017, and PrimeSTAR Max DNA polymerase. pET28a was PCR amplified using the synthetic primers 5'-CATATGTCGCTGCAGCCCCATGATGTTCT-3' (forward primer) and 5'-GGATCCCAATCGGAGATCTCTAGGCTGTTATTTTGCTCT-3' (reverse primer), pET28a, and PrimeSTAR Max DNA polymerase. A plasmid expressing AM1_1557g2 (pET28a_AM1_1557g2) was then constructed with the Takara in-fusion system reagents.

Expression and Purification of His-tagged AM1_1557g2, C304A and L337N. E. coli C41 (Novagen) carrying pET27o or pET27i was used for expression of AM1_1557g2, C304A and L337N. Each culture was incubated at 37°C for 2.5 h in 1 L of Luria–Bertani medium, 20 mg ml−1 kanamycin, and 20 mg ml−1 chloramphenicol, pH 7.2. Each Bph (dual-Cys cyanobacteriochrome GAF domain found in cyanobacterium Synechococcus sp. PCC 6803, which has an unusual red/blue reversible photoconversion cycle). Biochemistry 53, 5051–5059, doi:10.1021/bi3013565 (2014).

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**Acknowledgments**

We thank Dr. Nathan C. Rockwell for helpful discussion and kind reading of the manuscript. This work was supported by Precursory Research for Embryonic Science and Technology, Japan Science and Technology Agency, 4-1-8 Honcho Kawaguchi, Saitama 332-0012 (to R.N.) and Grants-in-Aid for Young Scientists (to R.N.).

**Author contributions**

R.N., M.S. and M.I. designed the research. R.N., N.N.W. and K.F. prepared plasmids for expression of AM1_1557g2. R.N. purified AM1_1557g2 proteins and performed spectroscopic analyses. G.E. performed purification and spectroscopic analysis of PaBphP. R.N., T.N., Y.A. and S.I. detected fluorescence from AM1_1557g2. R.N., M.S. and M.I. analyzed the data and wrote the manuscript.

**Additional information**

Supplementary information accompanies this paper at http://www.nature.com/scientificreports

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Narikawa, R. *et al*. A biliverdin-binding cyanobacteriochrome from the chlorophyll α-bearing cyanobacterium *Acaryochloris marina*. *Sci. Rep.* 5, 7950; DOI:10.1038/srep07950 (2015).