Karyomorphology, Meiotic Behaviours and Pollen Fertility of Calendula officinalis L. (Calenduleae- Asteraceae)

Pushpa Karna Mallick*

Tribhuvan University, Department of Botany, Tri-Chandra Multiple Campus, Kathmandu, Nepal

Article Information

Received: 05 February 2021
Revised version received: 13 March 2021
Accepted: 18 March 2021
Published: 30 March 2021

Cite this article as:
P.K. Mallick (2021) Int. J. Appl. Sci. Biotechnol. Vol 9(1): 75-79. DOI: 10.3126/ijasbt.v9i1.36011

*Corresponding author
Pushpa Karna Mallick,
Tribhuvan University, Department of Botany, Tri-Chandra Multiple Campus, Kathmandu, Nepal
Email: karnapushpa@Yahoo.com

Peer reviewed under authority of IJASBT
© 2021 International Journal of Applied Sciences and Biotechnology

Open Access

This is an open access article & it is licensed under a Creative Commons Attribution Non-Commercial 4.0 International (https://creativecommons.org/licenses/by-nc/4.0/)

Keywords: Karyomorphology; meiosis; pollen fertility; Asteraceae; Aceto-orcein

Introduction

The taxa Calendula officinalis is the flowering plants of the family Asteraceae and placed in the tribe Calenduleae. The tribe Calenduleae has eight genera and over 110 species, mostly found in South Africa (Judd et al., 2008). Only one species Calendula officinalis found in Nepal so far (Judd et al., 2008). Economically the taxa Calendula officinalis is very important as it grows for ornamental purposes as well as for medicinal purposes. The leaves and flower of this is uses in wounds and burns. Calendula officinalis contained with many chemical constituents such as carotenoids, flavonoids, saponins, sterols, phenolic acids, lipids etc. (Ashlawyen, 2018). Karyomorphological study provides evolutionary characteristics of karyotypes, as well as the cytological mechanisms. Karyomorphological study is a fast and inexpensive approach to classify plant species by identifying the basic cytological parameters of a species, including chromosome number, ploidy level, karyotype asymmetry, and karyotype coefficient (Guerra, 2008). Chromosomes number and karyotype of a species are stable characteristics which can reflect its basic genetic information. Karyomorphological studies was done by different authors in different plant species time to time such
as Yano and Hashino (2007) studied four species of cyperaceae karyomorphologically. Seven species of the

genus Salvia karyomorphologically studied by Martin et al., (2015). Differences in chromosome numbers between

populations are important evidence for determining reproductive isolation (Sun et al., 2020). Meiosis is events

of high evolutionary stability which reduce the chromosome number of chromosomes are one of the important
determinants for the evolutionary study. Pollen fertility is the ability of the pollen to perform its functions of delivering
male gametes to embryo sac. Pollen is a critical stage in the life cycle of the plants as fertile pollen is the crucial for
sexual plant reproduction. Commonly for pollen transport vectors are wind and different insects depending on the
species. In seed formation fertile pollen is very necessary. Studies of pollen fertility are very helpful for recognition of
wide range of variations existing within plants species and differentiating plant species with genera (Noor et al., 2004).

The main objective of this study is to determine chromosome number, karyotype analysis, meiotic behaviors and pollen fertility of the taxa Calendula officinalis L.

Materials and Methods

The plants were collected from Kathmandu, Nepal and transplanted in earthen pots at my home garden. Somatic
chromosomes were prepared from healthy root tips. They were pretreated with aqueous solution of 0.002M 8-
hydroxyquinoline for three hours. The root tip after pretreatment was fixed in mixture of absolute ethanol and

glacial acetic acid (3:1) for 24 hours. The root tip materials were then hydrolyzed and stained in a mixture of 2% aceto-
orcein and 1N HCl (9:1) contained in watch glass and warmed for few seconds and left for 30 minutes to 1 hour.

Squashes were made in 45% acetic acid. Observed under compound microscope. The drawings were made with the
help of 1366 Camera Lucida apparatus. Photomicrographs were taken by using digital camera of 12.1 megapixel using
10 x eye pieces and 100x objective of trinocular compound microscope. For kar

ytotype studies at least three different

stages of meiosis were photographed under the compound
microscope. The meiotic behavior of pollen mother cells was observed under microscope. Pollen fertility of the taxa was estimated on the

basis of stainability test. It was determined by using Muntzing (1941) solution made by one-part aceto-carmine and one-part glyc
erine (1:1). Well inflated, uniformly stained and healthy grains were considered as fertile ones.

Results and Discussion

The plant is a short living, aromatic and erect annual herb, growing up to 80 cm tall with sparsely branched stem. It is
commonly called pot marigold. The stem is angular, glandular and hairy. The leaves are oblong, lanceolate and hairy on both sides, margins entire. The head is yellow comprising a thick capitulum. Flowers are bright orange yellow, 3-toothed, tube hairy. In the wild form they have a single ring of ray florets surrounding the central disc florets. The disc florets are tubular and hermaphrodite, and generally of a more intense orange-yellow color than the ray

florets. Peripheral ray florets tridentate. The flowers may appear throughout year where conditions are suitable. The
fruit is a thorny curved achene. Achenes longer than the involucres, curved boat-shaped dorsally muricate not
beaked, outer longer ventrally crested, beaked (Fig.1).

Somatic chromosome number determined for this taxon is 2n=28. Reproductive chromosome number or haploid
number n=14 determined for this taxon. The somatic chromosome number determined is shown in Fig. 2 and
camera lucida drawing in Fig. 3. Its ideogram is represented in Fig.4. The chromosome measurements are given in Table

1.

Chromosome of metaphase plate shows three types with centromeric at median points, median regions and sub
median regions. The chromosomes length ranged from 0.4 to 2.6 µm with mean length 1.5 µm and absolute length 21.9
µm. Karyotype formula is M16+1(2A+2B) sm8.

Meiotic study shows both regular as well as irregular behavior in this taxon. Normal diakinensis with rod
bivalents were observed. Metaphase exhibit normal behavior. Anaphase with non- synchronous division and non-oriented chromosomes were observed which shows slightly irregular meiosis. Sticky chromosomes with laggar are found in metaphase-I stage. Formation of non oriented chromosomes may be due to early disjunctions of bivalents. Normal telophase was occurred. Diakinesis shown in Figs. 5 and 6. Metaphase-I is shown in Fig. 7-10 are noted. Telophase-I is in Fig. 11. Metaphase- II (Fig. 12-14). Anaphase-II with non- synchronous division (Fig. 15) and Anaphase-II with non-oriented chromosomes (Fig. 16) have been observed. Telophase-II abnormal shown in Fig. 16. Tetrads (Fig. 18) and circular, round, echinate large pollens (Fig. 19) are observed. Pollen fertility determined was 84.4 percent.
Fig. 1. Photograph of living plant.
Fig. 2. Photomicrograph of somatic metaphase plate.
Fig. 3. Camera lucida drawing of the same.
Fig. 4. Ideogram of the above.
Fig. 5-6. Diakinesis.
Fig. 7-9. Metaphase-I.
Fig. 9. Metaphase-I with sticky chromosomes and laggars.
Table 1: Chromosome measurement in *Calendula officinalis* L.

| Chrom. Pairs | Long Arm (µm) | Short Arm (µm) | Total Length (µm) | r-value | Relative Length (µm) | Position of centromere |
|--------------|---------------|----------------|-------------------|---------|----------------------|------------------------|
| I            | 1.3           | 1.3            | 2.6               | 1       | 11.8                 | M                      |
| II           | 1.3           | 0.8            | 2.1               | 1.6     | 9.8                  | m                      |
| III          | 1.3           | 0.8            | 2.1               | 1.6     | 9.8                  | m                      |
| IV           | 0.8           | 0.8            | 1.6               | 1       | 7.8                  | M                      |
| V            | 0.8           | 0.8            | 1.6               | 1       | 7.8                  | M                      |
| VI           | 0.8           | 0.8            | 1.6               | 1       | 7.8                  | M                      |
| VII          | 0.8           | 0.8            | 1.6               | 1       | 7.8                  | M                      |
| VIII         | 0.8           | 0.8            | 1.6               | 1       | 7.8                  | M                      |
| IX           | 0.8           | 0.4            | 1.2               | 2       | 5.9                  | Sm                     |
| X            | 0.8           | 0.4            | 1.2               | 2       | 5.9                  | Sm                     |
| XI           | 0.8           | 0.4            | 1.2               | 2       | 5.9                  | Sm                     |
| XII          | 0.8           | 0.4            | 1.2               | 2       | 5.9                  | Sm                     |
| XIII         | 0.4           | 0.4            | 0.8               | 1       | 3.9                  | M                      |
| XIV          | 0.2           | 0.2            | 0.4               | 1       | 1.9                  | M                      |

Fig. 10 Metaphase-I.  Fig. 11. Telophase-I.  Fig. 12-14 Metaphase-II.  Fig. 15. Anaphase-II with non-synchronized division.  Fig. 16. Anaphase-II with non-oriented chromosomes.  Fig. 17. Telophase-II.  Fig. 18. Tetrad normal.  Fig. 19. Pollen grain.
Karyotype \((M_{1s}+M_{1s}, sm_{s})\) of *Calendula officinalis* shows slightly asymmetrical chromosomes with centromere at median point, median and sub-median regions. Fallahi *et al.* (2020) determined symmetrical karyotypes with metacentric and sub-metacentric chromosomes for these taxa. The somatic chromosomes number in present study was found to be \(2n=28\) but somatic chromosomes, \(2n=32\) found by Fallahi *et al.* in 2020. So, this species found in two different cytotypes. The chromosome lengths ranged from 0.4 to 2.6 \(\mu m\) in this study. The longest and shortest chromosomes ratio indicates its advanceness. Annual habit, sparsely branched stem and large oblong- lanceolate leaves show advanceness of this taxon.

The haploid chromosome number \(n=14\) for the taxa *Calendula officinalis* of the tribe Calenduleae is determined in present investigation. Earlier haploid number \((n=16)\) was reported by Gupta (1969). So, *Calendula officinalis* may be existed with two haploid numbers. Meiosis in this taxon exhibited both regular as well as irregular behaviors. Regular meiosis was observed in this species by Gupta *et al.* (1972). Samatadze *et al.* (2019) determined sixteen bivalent for this species. Meiotic disorders like chaotic disjunction, laggards, bridges, chromosome fragments also revealed by them for this species. Darlington and Wylie (1955) suggested different basic numbers for this species such as \(x=7, 8\) and \(9\). Basic number \(x=8\) was reported for *Calendula officinalis* by Gupta *et al.* (1972). Thus, *Calendula officinalis* could be found in triadic forms.

Pollen fertility for *Calendula officinalis* 95 percent was determined by Noor *et al.* (2004). Pollen fertility determined for this species in present investigation is 84.4 percent. Pollen fertility was affected by development of abnormalities like laggards, non-orientations of chromosomes etc leading to the formation of deficient pollen grains.

**Conclusions**

The longest and shortest chromosomes ratio indicates its advanceness. The variations in chromosome numbers, in meiotic behaviors and karyomorphological structure and high fertility rate shows evolve nature of this species which play a great role in evolution.

**Conflict of Interest**

The author declares that there is no conflict of interest with present publication.

**Acknowledgement**

The Former head, Central Department of Botany, Tribhuvan University, Kathmandu, Nepal, Dr Pramod Kumar Jha highly acknowledged for providing lab facilities.

**References**

Ashwlayan VD, Kumar A and Verma M (2018) Therapeutic Potential of Calendula officinalis. *Pharm Pharmacol Int J*. 6(2):149-155. DOI: 10.15406/ppij.2018.06.00171

Darlington CD and Wylie AP (1955) Chromosome Atlas of Flowering plants. George Allen and Unwin Ltd., Great Britain.

Fallahi M, Mohammadi A and Miri SM (2020) The natural variation in six populations of *Calendula officinalis* L.: A karyotype study. *J Genet Resour* 6(1): 34-40.

Guerra M (2008) Chromosomes numbers in plant cytotomy: concepts and implications. *Cytogenet. Genome Res.*, 120:339-350. DOI: 10.1159/000121083

Gupta PK (1969) Cytological investigations in some Indian Compositae. *Cytologia* 34: 429-438. DOI: 10.1508/cytologia.34.429

Gupta PK, Agarwal DK and Shrivastava AK (1972) Further Cytological investigation in Indian Compositae *Cytologia* 37: 581-593. DOI: 10.1508/cytologia.37.581

Judd Campbell, Kellogg, Stevens, Donoghue (2008) *Plants Systematics: a Phylogenetic Approach*. Third Edition, Sinauer Associates, Sunderland, MA.

Levan A, Fregda K and Sandberg AA (1965) Nomenclature for centromeric position on chromosomes. *Hereditas* 52: 201-220. DOI: 10.1111/j.1601-0836.2014.00195.x

Martin E, Fahim A, Celep F, Ahmet K and Musa D (2015) Karyomorphological studies in seven taxa of the genus Salvia (Lamiaceae) in Turkey. *Caryologia* 68(1), 13-8. DOI: 10.1159/000367114.2014.998127

Muntzing A (1941) Differential response to x-ray treatment of diploid and tetraploid barley. k. Fission. *Sellsk. Forh.* 11: 1-42.

Noor M J, Ahmad M, Asghar R, Kanwal A, Sadaf P (2004) Palynological Studies of Cultivated plants Species at University of Arid agriculture, Rawalpindi, Pakistan, Asian journal of Plant Science 3(4): 76-479. DOI: 10.3923/ajps.2004.476.479

Samatadze T E, Zoshchuk S A, Hazieva F M (2019) Phenotypic and molecular cytogenetic variability in calendula (*Calendula officinalis* L.) cultivars and mutant lines obtained via chemical mutagenesis. *Sci Rep* 9, 9155. DOI: 10.1038/s41598-019-45738-3

Sun W, Wang H, Wu R, Sun H, Li Z (2020) Karyomorphoogy of three endemic plants (Brassicaceae: Euculideae and Arabideae) from the Qinghai-Tibet Plateau and its significance. *Plant Diversity*, 42 (3)135-141. DOI: 10.1016/j.pld.2020.03.002

Yano O and Hashino T (2007) Karyomorphological studies of four species of Japanese Scleria (Cyperaceae). *Cytologia* 72(3): 275-278. DOI: 10.1508/cytologia.72.275