Sphingosine-1-phosphate (S1P) receptors: Promising drug targets for treating bone-related diseases

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Abstract
Sphingosine-1-phosphate (S1P) is a natural bioactive lipid molecule and a common first or second messenger in the cardiovascular and immune systems. By binding with its receptors, S1P can serve as mediator of signalling during cell migration, differentiation, proliferation and apoptosis. Although the predominant role of S1P in bone regeneration has been noted in many studies, this role is not as well-known as its roles in the cardiovascular and immune systems. In this review, we summarize previous research on the role of S1P receptors (S1PRs) in osteoblasts and osteoclasts. In addition, S1P is regarded as a bridge between bone resorption and formation, which brings hope to patients with bone-related diseases. Finally, we discuss S1P and its receptors as therapeutic targets for treating osteoporosis, inflammatory osteolysis and bone metastasis based on the biological effects of S1P in osteoclastic/osteoblastic cells, immune cells and tumour cells.

KEYWORDS
oncology-related bone metastasis, inflammatory osteolysis, osteoporosis, S1P receptors

1 | INTRODUCTION

Bone is regarded as a highly dynamic tissue that constantly undergoes cycles of bone resorption and bone formation. The whole cycle is divided into three phases: (a) the interaction of osteoclast precursors (OCPs) and osteoblasts (OBs) on the bone surface, including OCP recruitment, RANKL-RANK binding and adhesion to the bone surface; (b) the differentiation of osteoblast precursors (OBPs) is facilitated by various cytokines from mature osteoclasts; and (c) the apoptosis of osteoclasts (OCs), as well as mineralization of the bone matrix. During these OCPs and OBPs migrate to the bone surface, and various coupling factors, such as receptor activator of nuclear factor-κ B ligand (RANKL), have a dramatic influence on the bone regeneration cycle. Currently, S1P has been recognized to participate in this remodelling cycle, suggesting its significance in bone pathology.

Sphingosine-1-phosphate (S1P) is a natural bioactive lipid molecule and a common first or second messenger in the cardiovascular and immune systems. At present, research on the role of S1P in the...
regulation of cell migration, differentiation, proliferation and apoptosis is superficial.\textsuperscript{2} As a functional molecule that regulates numerous processes in the human body, S1P is increased in blood vessels and reduced in other tissues, thus creating an S1P gradient between blood and interstitial fluid. Moreover, this gradient determines the direction of cell migration, indicating that S1P is another chemokine that participates in chemotaxis in addition to C-X-C motif chemokine ligand 12 (CXCL12)/stromal cell-derived factor 1 (SDF-1). One of the causes of osteoporosis is chemokine-mediated recruitment of osteoclasts into bone resorption sites, and inflammatory osteolysis, as well as bone metastasis, is due to the migration of inflammatory cells or tumour cells through chemokine gradients. In this context, studies on the S1P gradient are of great clinical value.

S1P is derived from ceramide, which is transformed into sphingosine primarily in vascular endothelial cells or circulating red blood cells/platelets. In addition, S1P is phosphorylated by sphingosine kinase 1/2 (SPHK1/2) and transported into blood vessels in different manners. While several transporters of S1P have previously been identified, spinster homologue 2 (SPNS2), a member of a large family of non-ATP-dependent organic ion transporters, has recently attracted much attention as an S1P transporter. Moreover, S1P is also exported from mast cells independently of their degranulation in a manner that is mediated by ATP-dependent ABC transporters. Interestingly, the remaining S1P in different cells participates in several cellular behaviours via many other intracellular signalling pathways.\textsuperscript{4,5} The difference between SPHK1 and SPHK2 is that SPHK1 mediates cytoplasmic S1P secretion, while SPHK2 phosphorylates S1P in cell nuclei to regulate histone acetylation.\textsuperscript{6} For the purpose of travelling through the blood, S1P must bind to high-density lipoprotein (HDL) or albumin. Furthermore, over-production of S1P can be inhibited to maintain a stable level. One mechanism is that intracellular S1P is degraded by sphingosine-1-phosphate lyase (SPL) or dephosphorylated by sphingosine-1-phosphate phosphohydrolase 1/2 (SPP1/2) after production, suggesting that S1P secretion is strictly controlled.\textsuperscript{5} As these enzymes are inactive in haemocytes, the S1P concentration in blood is much higher than that in other tissues, such as bone matrix. Another mechanism involves the lipid phosphohydrolase (LPP) family on the plasma membrane, which dephosphorylates extracellular S1P, resulting in reduced local extracellular S1P concentrations and increased sphingosine.\textsuperscript{7} The accumulated sphingosine takes part in other signal pathways or is converted back into S1P. Under normal circumstances, the generation and degradation of S1P maintain a dynamic balance. Disruption of this balance leads to dysfunction and diseases of many organs. In summary, S1P-related enzymes may be future targets for treating bone disorders.

At present, five different S1P receptors (S1PR1-5) have been identified and are encoded by Edg1, 5, 3, 6 and 8.\textsuperscript{8,9} These receptors are G protein–coupled receptors and activate downstream signals after binding with S1P.\textsuperscript{10} Each receptor has a unique function and downstream signalling pathway based on its specific structure. S1PR1 is widely expressed in almost all kinds of cells and primarily couples to G\textsubscript{12/13} proteins. S1PR2 and S1PR3 are less widely distributed and are coupled to G\textsubscript{q}, G\textsubscript{i}, and G\textsubscript{i}q. The expression of S1PR4 and S1PR5 is even more restricted and only detected in specific tissues. As the research on S1PRs is insufficient, especially in skeletal systems, there is an urgent need for more studies to investigate this field. Hence, this review will focus on how S1PR1-3 influences bone metabolism and how to use these receptors as novel drug targets for bone-related diseases. Because a multitude of S1P agonists or antagonists are recognized and used in the laboratory to determine which S1PR is involved in certain functions, the development of our understanding of S1P has largely broadened.\textsuperscript{10} However, what is much more meaningful is that these activators or inhibitors can be used as drugs to treat bone diseases after commercial processing and extensive clinical trials. As osteolytic diseases such as osteoporosis and inflammatory osteolysis are partly caused by excessive numbers of osteoclasts, searching for a way to reverse excessive osteolysis without affecting bone turnover has incomparable superiority. However, raising this theoretical new therapeutic target to a practical level requires a large amount of effort. In this review, we will also focus on strategies for future research directions and drug targets.

\section{The Function of S1P on Osteoclasts and Osteoblasts}

As a dynamic metabolic system, the skeletal system undergoes degradation and renewal. Healthy people stably maintain this cycle, while patients with bone disorders suffer from disequilibrium of bone formation and resorption, leading to increased or decreased bone mass. As S1P has been discovered to play an essential role in bone metabolism, targeting S1PRs on osteoclastic and osteoblastic cells is a novel direction of scientific research. More importantly, S1P cooperates with RANKL to participate as a coupling factor in osteoclast-osteoblast crosstalk. Thus, the clinical value of S1P involves the whole bone regeneration cycle.

\subsection{The Effect of S1P on Osteoclasts}

Osteoclasts are derived from the monocyte/macrophage lineage and fuse into multinuclear cells after stimulation with RANKL and macrophage colony-stimulating factor (M-CSF). These cells are responsible for bone matrix demineralization by secreting cathepsin K (CTSK) and HCl.\textsuperscript{11} Previous studies have discovered that S1P is responsible for the migration and differentiation of osteoclastic cells. Some chemokines determine the direction of OCP migration, which play important role in bone resorption. S1PR1 and S1PR2, which were originally thought to be correlated with cell migration in lymphocytes, are more highly expressed than other S1PRs, indicating the impact of S1P on OCP migration.\textsuperscript{12} Based on a large number of experimental studies, S1P impacts OCP mobilization and recruitment. When the extracellular S1P concentration is low, S1P binds with S1PR1 to promote cell chemoattraction.\textsuperscript{13} Treatment with pertussis toxin, a Gi protein blocker, in a mouse
model suggested that Gi and Rac were involved in S1PR1-mediated chemotraction. In contrast to this conclusion, a high level of extracellular S1P concentration results in S1PR2-dominant regulation of OCP chemorepulsion. Many studies have shown that the G_{12/13}/Rho signalling axis is downstream of S1PR2. Because OCP chemotraction is enhanced when there are defects in S1PR2, a negative effect of S1PR2 signalling on the function of S1PR1 was determined. Interestingly, S1PR2^{-/-} mice showed a significant attenuation in osteoblasts in vivo but not in vitro. This difference indicates that S1PR2 alone may not be sufficient to recruit osteoclast precursors. Ideas about the mechanisms of OCP chemotaxis suggest that S1PR1 is activated by S1P and rapidly internalizes in a high S1P environment and is transported back to the cell membrane in a low S1P environment. Therefore, S1PR2 is dominant when OCPs are circulating in the blood and is inhibited when OCPs are in the bone marrow. In addition, the binding of S1P to S1PR2 leads to reduced S1PR2 but increased S1PR1 expression, which forms a negative feedback loop. Moreover, osteoclast precursors, as well as bone marrow-derived macrophages/monocytes (BMMs), are stored in bone marrow and are mobilized to the blood when the expression of S1PR1 reaches a level that can be activated by low S1P concentrations.

Based on previous studies, RANKL and M-CSF play important roles in osteoclast differentiation. In addition, S1P participates in OCP differentiation in an indirect manner. Usually, S1P impacts this process by regulating RANKL expression or its downstream signalling pathway. The existence of SPHK1, which is responsible for S1P production, sharply reduces osteoclastogenesis in BMM cultures by blocking p38 MAPK, c-Fos and NFATc1 and augmenting ERK. Intriguingly, coculture of BMMs with osteoblasts had the opposite effect and increased RANKL expression. The difference between these two culture systems subtly revealed that intracellular S1P in OCPs suppresses OCP maturation and boosts RANKL expression by osteoblasts after secretion from the cells. Moreover, enhanced RANKL expression induces a dramatic increase in BMM differentiation. Previous studies found two different mechanisms for bone resorption: (a) chemorepulsion of osteoclast precursors through the S1P gradient and (b) an increase in RANKL due to osteoclast-secrated S1P. The S1P gradient between the blood and bone matrix, as well as S1PR2 on OCPs, is more important for bone resorption, suggesting that the increased number of osteoclasts plays a more superior role than enhanced osteoclast maturity. However, this does not mean that the differential effect of RANKL is unimportant. In contrast, monocytes are unable to fuse with mature osteoclasts in the absence of RANKL. After fusing into multinuclear cells, osteoclasts must adhere to the bone surface to secrete H^{+}, cathepsin K and matrix metalloproteinases (MMPs). The markers of osteoclast formation and activation, including NFATc1, CTSK, acid phosphatase 5 (ACP5), osteoclast-associated receptor (OSCAR), dendritic cell–specific transmembrane protein (DC-STAMP) and osteoclastic cell–specific transmembrane protein (OC-STAMP), sharply increase after S1PR2 stimulation.

### 2.2 The effect of S1P on osteoblasts

Osteoblasts, unlike osteoclasts, are derived from mesenchymal stem cells (MSCs). These cells generally remain on the bone surface and develop into osteocytes in the bone matrix. Collagen and other substances are secreted by osteoblasts for bone mineralization and formation, which is a key step in bone remodelling. Because osteoblasts are responsible for bone mineralization, their recruitment, differentiation and proliferation are fundamental for bone formation.

Similar to OCPs, S1P regulates OBP mobilization and recruitment through S1PR1/S1PR2 downstream signals. S1PR1 stimulates the JAK/STAT signalling axis, while S1PR2 activates the FAK/PI3K/AKT signalling axis, both of which promote MSC migration. S1PR2 mediates chemorepulsion of OBPs, while OBP chemotraction is modulated by S1PR1. Further studies also showed that the JAK/STAT and FAK/PI3K/AKT signalling axes were independent and had no crosstalk.

In addition to osteoblast location, S1P is also crucial for OBP differentiation. S1PR1-3 is largely expressed in osteoblastic cells. In contrast to the decrease in S1PR1 and S1PR2 expression, S1PR3 expression increases sharply during OBP differentiation. Moreover, when S1PR3 is knocked out, bone matrix mineralization is not impacted by S1P, which means that S1PR3 regulates osteoblastogenesis. Other studies also found that S1PR1 and S1PR2 participate in osteoblastogenesis despite their decreased expression during differentiation. Bone morphogenetic protein 2 (BMP2), a crucial protein for OBP differentiation, is the target for S1PR1/2 downstream signalling pathways. The phosphorylation of Smad1/5/8, as well as ERK1/2, and the expression of Runx2 increase dramatically after activation of S1PR1 and, to a lesser extent, S1PR2. However, recent studies discovered that the PI3K/AKT/GSK-3β/κ-catenin and RhoA/ROCK/Smad1/5/8/Runx2/ALP signalling axes were non-BMP2-dependent pathways for osteoblastogenesis that stimulated through S1PR1 and S1PR2, respectively. RhoA was also responsible for Smad 6/7 phosphorylation, which inhibits Smad1/5/8 activation and represents negative feedback regulation. In contrast, S1P enhances S1PR1 and S1PR2 expression, which is positive feedback regulation. Overall, S1PR1-3 promotes OBP differentiation via a regulatory signalling network. In this network, S1PR3 directly regulates osteoblastogenesis, while the differential effect of S1PR1/2 is less powerful than that of S1PR3.

Since mature osteoblasts still undergo mitosis, osteoblast proliferation and survival are crucial for osteogenesis. Various studies have found that S1P facilitates osteoblast proliferation and survival. Initially, only p42/44 MAPK and G_{i} were identified as downstream signalling pathways of S1P in the proliferation process. Later, studies discovered that PKCs are increased by S1P, and protein kinase C α (PKCα) was considered a downstream signal of S1PR1. Some researchers even held the idea that PKC cooperates with MAPK to exert its proliferation effect. However, a group of scientists found that intracellular [Ca^{2+}] but not PKC was required for S1P-induced p42/44 MAPK activation, indicating that PKC exerts a proliferative effect without activating MAPK. Calcium is found mainly in bone,
and intracellular \([\text{Ca}^{2+}]\) plays an essential role in bone metabolism and metastasis.\textsuperscript{23–25} Therefore, the involvement of the calcium signalling pathway in S1P-induced osteoblast proliferation significantly improves our understanding of the function of \([\text{Ca}^{2+}]\) in bone homeostasis. Furthermore, pertussis toxin (a Gi inhibitor), LY294002 or wortmannin (PI3K inhibitors) and calphostin C (a PKC inhibitor) treatment were used on osteoblasts to determine which downstream signal of S1PR impacts osteoblast apoptosis. The results revealed, as expected, that Gi proteins and PI3K are the downstream signals of S1P for anti-apoptotic effects in osteoblasts.\textsuperscript{36} Research on the function of S1P receptors in osteoblast proliferation and survival is still far from sufficient, and further studies will be of great clinical value for bone-related diseases. Despite limited evidence, mechanical loading was found to stimulate S1P production in osteocytes via up-regulating SPHK1 and down-regulating SPP, SPL and SPNS2. The mechanism occurred by the osteocyte network translating the force they have experienced into a biological response, such as bone regeneration.\textsuperscript{37}

## 2.3 The effect of S1P on the crosstalk between osteoblasts and osteoclasts

As previously stated, S1P is crucial for the differentiation, proliferation and migration of both osteoclasts and osteoblasts. For cell migration of osteoclasts and osteoblasts, the same S1PR (S1PR1/2) may exert the same effect. Therefore, by selectively activating or blocking specific S1PRs on specific cells or by impacting S1PRs on different cells to different degrees we can selectively regulate the migration of OCPs or OBPs individually. In addition, it has been gradually accepted that osteoclasts and osteoblasts are not completely antagonistic cells. In contrast, they are tightly coupled and interact with each other to maintain homeostasis and bone turnover (Figure 1). Currently, S1P is generally regarded as another coupling factor that mediates bone regeneration. More specifically, S1P depends on stimulating certain S1PRs or facilitating the downstream signalling of other coupling factors, such as RANKL, to perform osteoclast-osteoblast crosstalk. Since S1P is mainly secreted by osteoclasts in the bone matrix, osteoclasts may play an important role in regulating osteoclast migration, differentiation and proliferation (Figure 1). Moreover, S1P enhances the expression of RANKL in osteoclasts, which promotes osteoclast differentiation and controls osteolyis (Figure 1). Thus, S1P, as a coupling factor between osteoclasts and osteoblasts, stimulates the mutual promotion and balance of osteolysis and osteogenesis. A recent study found that overexpression of intracellular S1P in BMMs attenuated osteoclastogenesis, while it stimulated RANKL expression to promote osteoclast differentiation after secretion into the bone matrix.\textsuperscript{36} Therefore, inhibiting the release of S1P from osteoclastic cells while increasing S1P production in osteoclastic/osteoblastic cells may greatly disrupt osteoclastogenesis without affecting bone turnover. Although some studies have already been carried out in this field, there is a strong need to perform further studies (Table 1).

## 3 The effect of S1P on osteoporosis

Osteoporosis is currently the most common osteolytic disease among elderly individuals and is typically characterized by a decrease in bone mass/density and bone strength. Osteoporosis is prone to complications such as long bone fracture. Osteoporosis occurs for many reasons, but it is essentially a disorder of bone formation and resorption.\textsuperscript{41} Because of the widespread prevalence of osteoporosis, studies searching for potential therapeutic targets are relatively old.

The traditional drug for osteoporosis is bisphosphonate, which is also used in the treatment of Paget’s disease, multiple myeloma and bone metastasis.\textsuperscript{42} It is a synthetic analogue of pyrophosphate combined with hydroxyapatite, and its anti-osteolytic effect is due to inhibition of bone resorption as well as promotion of osteoclast apoptosis, which may lead to severe side effects such as gastrointestinal, renal and ocular toxicities and osteonecrosis of the jaw (ONJ) after long-term treatment.\textsuperscript{43} At present, bisphosphonate is still the primary drug for osteoporosis, which means that exploring a novel treatment to reduce the side effects still has clinical value. Based on this purpose, we found a new drug called denosumab, a RANKL monoclonal antibody, that inhibits osteoclast differentiation with a much less negative effect than that of bisphosphonate. However, long-term use of denosumab leads to low bone turnover, which means that patients with severe bone regeneration problems should avoid using it.\textsuperscript{44} Currently, a humanized monoclonal antibody called romosozumab promotes the Wnt signalling pathway in osteoblastic cells, which increases bone formation and mineral density.\textsuperscript{45} Although it has significant advantages over other osteoporosis drugs, it still leads to a much larger number of osteoblasts than osteoclasts and will weaken the coupling between these two cell lineages.\textsuperscript{46} Compared with the effects of these drugs, treatments that work by modulating S1P can influence both osteoclasts and osteoblasts at the same time. Osteoclasts will not be over-inhibited when treating osteoporosis. Therefore, S1P, acting as a crucial factor to enhance bone regeneration, can be used to treat osteoporosis by restoring the dynamic balance between osteolysis and osteogenesis.

By modulating S1PRs on osteoclasts or osteoblasts, their effects against osteoporosis were discovered. For example, S1PR1 agonists or S1PR2 antagonists decrease osteoclasts in the bone matrix, while S1PR3-specific agonists are responsible for osteoblast differentiation. These agonists or antagonists serve as effective and valuable drugs for osteoporosis. However, the most promising S1P-related therapeutic targets for osteoporosis at present are the enzymes involved in S1P metabolism. Weske et al provided a potential anabolic therapy for bone loss by targeting S1P lyase.\textsuperscript{47} In addition, calcitonin (CT) was proven to promote Spns2 (the gene determining the expression of SPNS2) transcription, which enhanced the release of S1P from osteoclasts.\textsuperscript{48} Similarly, the formation of S1P can be modulated by treatments targeting SPHK. Using SKi (10 μmol/L), a SPHK inhibitor, in hOB assays further confirmed this hypothesis.\textsuperscript{49} Because these enzymes have limited distribution and their activation can be easily controlled, controlling the S1P concentration by regulating
enzyme activation has tissue specificity, and they have great advantages over directly regulating S1PRs.

Moreover, S1P also regulates the function of hormones associated with osteoporosis. Calcitonin (CT) was found to be involved in bone loss in a non-canonical manner, which abolished the secretion of S1P from osteoclastic cells to inhibit osteoblast differentiation.48 Oestrogen acts as an important mediator in regulating bone matrix metabolism, which also participates in the activation of an intracellular network composed of many cytoplasmic and nuclear mediators. Additionally, some oestrogen effects can also be mediated by sphingolipids. Furthermore, oestrogen activates S1P receptors (S1PRs) and induces growth factor receptor transactivation.30,50 17β-oes-tradiol (E2), one of the three major types of endogenous oestrogens, exerts a direct osteogenic effect or regulates osteogenesis via the S1PR1/SPHK/S1P signalling axis (the concentration of E2 used was 10 nmol/L).30 Another hormone that is highly correlated with S1P is glucocorticoid. In addition to their anti-inflammatory and immunosuppressive functions, glucocorticoids have also been widely used in clinical treatment and in rescuing emergency patients in the medical community. However, the negative effect on osteoblast survival is a major concern when considering glucocorticoid use. A recent experiment using K6PC-5, an activator of SPHK1, in MC3T3-E1 osteoblastic cells successfully reversed Dex-induced apoptosis, indicating that S1P greatly alleviates the side effects of glucocorticoids on bone.38 Overall, excessive secretion or the use of these osteolytic hormones leads to decreased osteoblasts or increased osteoclasts, and regulating S1P is a novel idea to treat osteoporosis. However, the links between oestrogen signalling and activation of sphingosine kinase axis in bone cells are unclear and further trials are needed to determine whether the treatment can be used clinically. It has also been shown that sphingosine kinase and S1P receptors participate in oestrogen-mediated EGF receptor transactivation. Moreover, some
functional proteins cooperate with S1P to mediate bone metabolism, including epidermal growth factor (EGF) and bone morphogenetic protein 6 (BMP6). In previous studies, EGF was found to trigger osteoblast proliferation by increasing intracellular S1P concentrations, while OC-secreted BMP6, together with S1P, modulated osteoblastogenesis and mineralization. In addition, a current study using CTSK 

Intracellular 

PKCa 

Survival 

IL-6 synthesis 

OC Activation 

TABLE 1 Biological functions of S1P in skeleton system and its downstream signal axis

| Cell | Effect | Signal pathway | References |
|------|--------|----------------|------------|
| OCP  | Migration | S1PR1/Gi/Rac/chemoattraction | 12,13 |
|      |         | S1PR2/Gi2/13/Rho/chemorepulsion | 13,14 |
|      | Differentiation | S1P/COX2 and mPGES1/PGE2/RANKL/RANK | 16 |
| OBP  | Migration | S1PR1/JAK/STAT/chemoattraction | 22 |
|      |         | S1PR2/FAK/PI3K/AKT/chemorepulsion | 22 |
|      | Differentiation | S1PR1/PI3K/AKT/GSK-3β/β-catenin | 25,26 |
|      |         | S1PR2/RhoA/ROCK/Smad1/5/8/Runx2 and ALP | 25,26 |
|      |         | SPHK1/S1P1/S1PR3/Runx2 and ALP | 39 |
| OB   | Proliferation | Gi/p42/44 MAPK | 31 |
|      |         | Intracellular [Ca^{2+}]/p42/44 MAPK | 33-35 |
|      |         | PKCa | 7 |
|      | Survival | Gi/PI3K | 38 |
|      | IL-6 synthesis | p42/p44 MAPK | 40 |
| OC   | Activation | S1PR2/RANKL/NFATc1, CTSK, ACP5, OSCAR, DC-STAMP and OC-STAMP secretion | 20 |

by binding to Toll-like receptor 4 (TLR4). Activated macrophages, called M1 macrophages, release a variety of pro-inflammatory cytokines, such as tumour necrosis factor-α (TNF-α), interleukin-1β (IL-1β) and interleukin-6 (IL-6), and enhance the expression of inducible NO synthase (iNOS), C-C motif chemokine receptor 7 (CCR7) and CD86. TNF-α and IL-1β directly promote osteoclastogenesis, while IL-6 only stimulates osteolysis by stimulating RANKL expression on osteoblasts (Figure 2). More importantly, M1 macrophages differentiate into osteoclasts after binding to RANKL, indicating that IL-6 also promotes osteoclast differentiation from M1 macrophages by increasing RANKL. Recently, a group of scientists discovered that M1 macrophages also stimulate RANKL expression via the SPHK1/S1PR1/RANKL signalling axis, forming a positive feedback loop. Surprisingly, other experiments showed that S1P increased COX-2, iNOS, prostaglandin E2 (PGE2), IL-1β and TNF-α in several cell lineages, including murine peritoneal macrophages. These findings indicate that S1P may also play an essential role in LPS-mediated inflammatory osteolysis. Many macrophage lineages express S1PR1-3, and their pro-/anti-inflammatory functions have not been completely discovered. Heo et al examined the expression of pro-inflammatory cytokines induced by LPS. Not only COX-2 and iNOS but also IL-1β, IL-6 and TNF-α were inhibited after treatment with the S1PR3 inhibitor TY52156, demonstrating the pro-inflammatory effect of S1PR3 (Figure 2). Moreover, in a previous study by Keul et al, S1PR3 

When G- bacteria infect bone tissue, their cell wall component LPS is released to the bone matrix, which activates macrophages

4 | THE EFFECT OF S1P ON INFLAMMATORY OSTEOLYSIS

Inflammatory osteolysis is a common but severe disease, causing millions of people around the world to suffer from bone density loss. It is the result of bone infection and is characterized by overactivated osteoclasts and an imbalanced bone remodelling cycle. Currently, surgical treatment is the only efficient method to address inflammatory osteolysis, but it may cause serious prognostic difficulties and mobility problems for patients. Therefore, finding a non-surgical treatment has great clinical value. Based on previous studies, there is a connection between S1P and inflammatory reactions, indicating that S1P may be a future target for inflammatory osteolysis.

When G- bacteria infect bone tissue, their cell wall component LPS is released to the bone matrix, which activates macrophages to migrate towards the infection site. Activated macrophages, called M1 macrophages, release a variety of pro-inflammatory cytokines, such as tumour necrosis factor-α (TNF-α), interleukin-1β (IL-1β) and interleukin-6 (IL-6), and enhance the expression of inducible NO synthase (iNOS), C-C motif chemokine receptor 7 (CCR7) and CD86. TNF-α and IL-1β directly promote osteoclastogenesis, while IL-6 only stimulates osteolysis by stimulating RANKL expression on osteoblasts (Figure 2). More importantly, M1 macrophages differentiate into osteoclasts after binding to RANKL, indicating that IL-6 also promotes osteoclast differentiation from M1 macrophages by increasing RANKL. Recently, a group of scientists discovered that M1 macrophages also stimulate RANKL expression via the SPHK1/S1PR1/RANKL signalling axis, forming a positive feedback loop. Surprisingly, other experiments showed that S1P increased COX-2, iNOS, prostaglandin E2 (PGE2), IL-1β and TNF-α in several cell lineages, including murine peritoneal macrophages. These findings indicate that S1P may also play an essential role in LPS-mediated inflammatory osteolysis. Many macrophage lineages express S1PR1-3, and their pro-/anti-inflammatory functions have not been completely discovered. Heo et al examined the expression of pro-inflammatory cytokines induced by LPS. Not only COX-2 and iNOS but also IL-1β, IL-6 and TNF-α were inhibited after treatment with the S1PR3 inhibitor TY52156, demonstrating the pro-inflammatory effect of S1PR3 (Figure 2). Moreover, in a previous study by Keul et al, S1PR3-/- macrophages did not migrate towards S1P, suggesting that S1PR3 participates in macrophage chemotraction. Thus, S1PR3 signalling is responsible for both macrophage mobilization and M1 macrophage polarization (pro-inflammatory effect). A recent study discovered that S1PR2/3 stimulates M1 polarization via the G_{i/o}/PI3K/JNK signalling axis in liver inflammation. However, whether this signalling pathway is involved in inflammatory osteolysis is still unclear. Another study carried out
by Hughes et al identified that S1PR1 on macrophages exerts an anti-inflammatory effect.66 Macrophages that produce IL-4 and IL-13 as anti-inflammatory cytokines are M2 macrophages. Further study using VPC44116, an S1PR1-specific antagonist, showed decreased arginase I (Arg I) activity (a marker of M2 macrophages) and increased iNOS activity (a marker of M1 macrophages), indicating that S1PR1 signalling is responsible for M2 macrophage polarization.66 Intriguingly, in ovariectomized (OVX) mice, M2 macrophages differentiated into osteoclasts, and an increased M1/M2 ratio was also discovered, suggesting a connection between oestrogen deficiency, osteolysis and M1 macrophage polarization.67 As the direction of macrophage polarization is determined by the expression of certain S1PRs or environmental S1P concentrations, it is possible to transform M1 macrophages into M2 macrophages in the bone matrix. If this idea can be used in clinical treatment, we would be able to reverse excess osteoclasts or even control the progression of inflammatory osteolysis. In addition, M1 and M2 macrophages exert completely opposite effects, and so identifying how S1PR1 and S1PR3 affect macrophages will help in regulating the M1/M2 ratio. We also hypothesize that the same S1PRs (S1PR1 or S1PR3) may exert different functions in different tissues during inflammatory reactions, and so the function of S1PR1 and S1PR3 may not strictly follow the conclusions above. For example, a recent study found that S1PR1 signalling up-regulates IL-6 expression in primary mouse macrophages by activating JAK2.68 In addition to S1PR1 and S1PR3, S1PR2 also induces its anti-inflammatory effect by inhibiting macrophage recruitment.69 More recently, using an anti-IL-6 receptor antibody efficiently down-regulated S1PR2 in a collagen-induced arthritis (CIA) model, causing severe osteolytic disease, which further suggests the anti-inflammatory effect of S1PR2.70 Thus, S1PR1-3 on

FIGURE 2 As one of inflammatory mediators, S1P serves as a mediator for inflammatory osteolysis. Excessive osteoclast proliferation and destruction of bone homeostasis are the main characteristics of inflammatory osteolysis, and macrophages are found to participate in this process. Macrophages can be recruited into inflamed sites of bone marrow, and S1P acts as a promoter for the polarization of M0-Mφs into M1-Mφs via S1PR3 signal pathway. Besides, the release of TNF-α, IL-1β and IL-6 from M1-Mφs can interact with S1P to participate in the regulation of bone turnover. TNF-α and IL-1β exert direct promotional effect on osteoclast differentiation, while IL-6 elevates RANKL expression to boost osteoclastogenesis. In addition, the polarization of M2-Mφs can be stimulated by S1PR1 signal pathway, which exerts osteogenic effect by secreting TGF-β.

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Macrophage-produced cytokines not only directly or indirectly impact the formation and activation of osteoclastic cells but also act as inflammatory mediators or chemokines to recruit lymphocytes to execute adaptive immunity reactions. Surprisingly, the S1P gradient between the blood/lymph and other organs is also responsible for lymphocyte trafficking from the thymus to inflamed sites and is mediated by S1PR1 signalling.\(^2\)\(^,\)\(^7\)\(^1\)\(^-\)\(^3\) If the inflamed site is bone, recruited lymphocytes directly eliminate pathogenic bacteria and control the progression of inflammatory osteolysis. In addition, lymphocytes are currently thought to be the only cell lineage to express S1PR4. A previous study showed that S1PR4 was associated with neutrophil migration during inflammation.\(^7\)\(^1\)

Therefore, S1PR4 may be a new drug target for inflammatory osteolysis. In addition, inflammation-induced lymphangiogenesis is widely accepted and is beneficial for lymphocytes and tumour cells. Recently, S1PR1 on HLECs was found to stimulate TNF-\(\alpha\) and IL-1\(\beta\) secretion via the NF-\(\kappa\)B signalling pathway, and TNF-\(\alpha\) and IL-1\(\beta\) are essential for HLEC proliferation, migration and lymphangiogenesis.\(^7\)\(^4\)

Although inflammation is a hot spot in basic medical research, there are still relatively few studies on the skeleton system. Based on the results shown in other tissues, lymphocyte recruitment and cytokine secretion may be potential therapeutic targets for inflammatory osteolysis.

Inhibiting inflammatory osteolysis through the intervention of S1P and lymphocyte recruitment has great advantages compared with those of formal surgical methods (mainly by removal of a large segment of infected bone tissue). Thus, there is a strong need to investigate the mechanisms by which S1P affects inflammation and lymphocyte recruitment.

### 5 | THE EFFECT OF S1P ON CANCER-RELATED BONE METASTASIS

Cancer is regarded as the top killer among patients worldwide and is characterized by uncontrolled growth, infiltration and metastasis. Tumour metastasis is the leading cause of cancer death, and bone is the third most common site for tumour metastasis after the lung and liver.\(^7\)\(^5\) According to previous studies, breast cancer (BCa) and lung cancer (LCa) usually metastasize to bone, causing severe pain and osteolysis. In addition, prostate cancer (PCa)-related bone metastasis always results in bone fracture.

Recently, various studies have found a link between S1P and bone metastasis, providing us with a novel drug target for bone metastasis. At the early stage of metastasis, S1P participates in the metabolism and migration of primary tumour cells in the tumour microenvironment by cooperating with tumour-associated macrophages (TAMs). TAMs act to eliminate tumour cells and are recruited to the tumour microenvironment via S1P-mediated ‘find me signals’.\(^7\)\(^6\) S1P in the tumour microenvironment is secreted and released by apoptotic tumour cells due to SPHK1 activation.\(^7\)\(^6\) Recruited monocytes undergo macrophage polarization (differentiation into M2 macrophages) after stimulation with S1P (Figure 3).\(^7\)\(^7\)\(^,\)\(^7\)\(^8\) Among the cytokines formed by M2 macrophages are IL-4 and IL-10, which promote tumour evasion and chemotherapy resistance (Figure 3).\(^7\)\(^9\) Moreover, M2 macrophages in the inflammatory tumour microenvironment also release PGE2 to modulate angiogenesis, further promoting tumour progression and metastasis.\(^8\)\(^0\)

Because S1PR3 downstream signalling promotes M2 macrophage polarization, using an S1PR3 antagonist in the tumour microenvironment may block the formation of M2 macrophages and fundamentally reduce the possibility of bone metastasis. Furthermore, transforming M2 macrophages into M1 macrophages may be a future strategy for controlling tumour spread. However, there are relatively few studies about how to transform M2 macrophages into M1 macrophages, and the changes caused by M1 macrophage-secreted cytokines cannot be estimated. Therefore, the effect of M1 macrophage-secreted cytokines on the tumour microenvironment is of great academic and clinical value. In addition, primary tumours are difficult to cure because of immune tolerance, which makes it difficult to achieve the expected therapeutic effect by immunological methods. Therefore, targeting M2 macrophages to control bone metastasis will be possible only if we find a way to reduce immune tolerance. In the last few years, various studies have focused on the role of T\(_{\text{reg}}\) cells in immune tolerance. Tumour-specific T\(_{\text{reg}}\) cells were thought to egress from bone marrow to tumour tissue in breast cancer patients by the stimulation of S1PR1, according to Rathinasamy et al. This group also found that S1PR1 expression in T\(_{\text{reg}}\) cells was induced after binding with antigen.\(^8\)\(^1\) Thus, S1P is correlated with T\(_{\text{reg}}\) cell-mediated immune tolerance. However, a brain-specific mechanism by which tumours escape immunosurveillance though the loss of S1PR1 on T cells was discovered.\(^2\)\(^7\) Thus, the exact mechanism by which S1P induces immune tolerance is not obvious, and it is unwise to blindly suppress S1PR1 on T cells or change the environmental S1P concentration.

Migration of tumour cells through blood and lymph vessels is a key step in tumour metastasis, and S1P seems to participate in this process via various mechanisms. On the one hand, the function of cytotoxic T cells is attenuated after binding to circulating S1P, promoting immune tolerance and preventing the elimination of tumour cells.\(^8\)\(^2\) On the other hand, S1P secreted by lymphatic or vascular endothelial cells inhibits breast cancer metastasis-suppressor 1 (BRMS1) after binding to S1PR2 on tumour cells.\(^8\)\(^3\) Furthermore, lymphangiogenesis and angiogenesis are both essential elements in tumour metastasis, and it is the main reason why tumour metastasis has a close interaction with inflammation. As mentioned above, stimulating S1PR1 on HLECs promotes lymphangiogenesis via the secretion of TNF-\(\alpha\) and IL-1\(\beta\), which are known pro-inflammatory cytokines. In addition, a previous study investigated another mechanism of S1P-induced lymphangiogenesis, in which
S1P was produced by tumour cells. As these results were not determined in the cancer lineages that usually metastasize to bone, we are not sure whether these mechanisms are involved in bone metastasis. Thus, determining the mechanisms in PCa/BCa cell lineages is a future strategy for treating bone metastasis. Finally, OB/OC-secreted S1P recruits certain tumour cells by enhancing the connection between tumour cells and the osteoclastic/osteoblastic niche, which is called bone colonization (Figure 3). An early experiment using several PCa cell lines co-cultured with osteoblastic cell lines (mainly MC3T3 cells) showed increased proliferation and resistance to standard therapy. This effect was due to OB-secreted S1P binding to S1PR1 on PCa cells and forming a positive feedback loop by promoting S1PR1 expression. Because PCa is involved osteogenic bone metastasis, osteoblastic niches may act as S1P reservoirs for tumour cell proliferation. Although experimental records have not shown whether osteoclastic niches act as S1P reservoirs for bone metastasis in BCa/LCa, we make this assumption based on the consensus that osteoclasts are better at secreting S1P than osteoblasts and that BCa/LCa are associated with osteolytic bone metastasis. More importantly, regulating S1P-related enzymes may also serve our purpose. However, we discovered that targeting a single receptor–ligand pro-metastatic axis to treat tumour metastasis cannot effectively inhibit the spread of cancer in the body. Thus, there is a strong need for more research to find other therapeutic targets, and a therapy that combines different mechanisms of bone metastasis is also a future direction for clinical treatment.

6 | THE CLINICAL DEVELOPMENT OF DRUGS TARGETING S1P RECEPTORS IN TREATING BONE-RELATED DISEASES

Due to the predominant role of S1P receptors in bone-related diseases have been widely noted, it is necessary to develop and test drugs that target S1P receptors in patients with bone-related
diseases. Among the several kinds of different drugs targeting one or more S1P receptors, fingolimod, ponesimod, siponimod, ozanimod and other related drugs are participated in clinical application and further development. Fingolimod (FTY720) acts as an agonist of the S1P receptor (S1PR). It could induce internalization and subsequent degradation of the receptor and consequently render lymphocytes to physiological S1P stimulation upon binding to the S1PR expressed on lymphocytes.\(^5\) In addition to its immunology modulatory effects in MS, fingolimod may have a beneficial effect on bone mass loss in female MS patients.\(^5\) Furthermore, fingolimod has been reported that it dramatically prevented bone loss in vivo via inhibiting RANKL-induced osteoclastogenesis in periodontitis model.\(^5\) Additionally, fingolimod was shown that it could also attenuate cancer-induced spontaneous pain.\(^6\) Moreover, ponesimod is a selective S1PR1 regulator to participate in the dose-dependent sequestration of lymphocytes in lymphoid organs.\(^8\) It has been noted that ponesimod, a selective S1PR1R regulator, is in great demand.\(^5\) In addition to the role of S1P signal pathways, is in great demand.\(^5\) In addition to the role of S1P signaling in osteoclastic/osteoblastic cells, as well as related facet roles and therapeutic potential.\(^4\) Sphingosine 1-phosphate signaling in bone remodeling: multi-faceted roles and therapeutic potential. Expert Opin Ther Targets. 2017;21:725-737.

7 CONCLUSION

Although scientific research on the relationship between S1P and bone biology is much more restricted than that of the cardiovascular and immune systems, S1P still has tremendous clinical value in bone pathology and will become a promising target for the treatment of many bone-related disorders, such as osteoporosis, inflammatory osteolysis and bone metastasis.\(^9\) A better understanding of S1P in osteoclastic/osteoblastic cells, as well as related signal pathways, is in great demand.\(^5\) In addition to the role of S1P as a first or second messenger that regulates cell migration, differentiation, proliferation and apoptosis, we focused on osteoclast-osteoblast crosstalk coupled with S1P.\(^2\) Interestingly, FTY720 acts as an inhibitor of the S1P receptor and was tested in clinical experiments in 2017.\(^9\) In addition, S1P acts as a bridge linking macrophages with inflammatory osteolysis and bone metastasis, which provides us with a novel drug target. Thus, research on S1P and its receptors is of great academic and clinical value. However, we have little evidence to support our hypotheses, and more studies are needed to determine whether S1P exerts the same effects in bone as in other tissues.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

Shiwu Dong and Yueqi Chen conceived the idea. Lincheng Zhang, Wenhui Hu and Yueqi Chen wrote and edited the manuscript. Lincheng Zhang, Yutong Dong and Yiran Wang drew the figures and organized the table. Shiwu Dong and Yueqi Chen approved the final version of the manuscript. All authors critically revised the manuscript.

DATA AVAILABILITY STATEMENT

The data used to support the findings of this study are available from the corresponding author upon request.

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REFERENCES

1. Matsuo K, Irie N. Osteoclast-osteoblast communication. Arch Biochem Biophys. 2008;473:201-209.
2. Rosen H, Goetzl EJ. Sphingosine 1-phosphate and its receptors: an autocrine and paracrine network. Nat Rev Immunol. 2005;5:560-570.
3. Sato K, Okajima F. Role of sphingosine 1-phosphate receptors. 2002;1582:112-120.
4. Takuwa Y. Subtype-specific differential regulation of Rho family G proteins and cell migration by the Edg family sphingosine-1-phosphate receptors. Biochem Biophys Acta. 2002;1582:112-120.
5. Fukushima S, Simmons S, Kawamura S, et al. The sphingosine-1-phosphate transporter Sgsp2 expressed on endothelial cells regulates lymphocyte trafficking in mice. J Clin Invest. 2012;122:1416-1426.
6. Hait NC, Allegood J, Maceyka M, et al. Regulation of histone acetylation in the nucleus by sphingosine-1-phosphate. Science. 2009;325:1254-1257.
7. Meshcheryakova A, Mechtcheriakova D, Pietschmann P. Sphingosine 1-phosphate signaling in bone remodeling: multi-faceted roles and therapeutic potential. Expert Opin Ther Targets. 2017;21:725-737.
8. Zhou J, Saba JD. Identification of the first mammalian sphingosine phosphate lyase gene and its functional expression in yeast. Biochem Biophys Res Commun. 1998;242:502-507.
9. van Veldhoven PP, Mannaerts GP. Sphingosine-phosphate lyase. Adv Lipid Res. 1993;26:69-98.
10. Sartawi Z, Schipani E, Ryan KB, Waebler C. Sphingosine 1-phosphate (S1P) signalling: role in bone biology and potential therapeutic target for bone repair. Pharmacol Res. 2017;125:232-245.
11. Chen Y, Dou CE, Yi J, et al. Inhibitory effect of vanillin on RANKL-induced osteoclast formation and function through activating mitochondrial-dependent apoptosis signaling pathway. Life Sci. 2018;208:305-314.

12. Ishii M, Egen JG, Klauschen F, et al. Sphingosine-1-phosphate mediates osteoclast precursors and regulates bone homeostasis. Nature. 2009;458:524-528.

13. Ishii M, Kikuta J. Sphingosine-1-phosphate signaling controlling osteoclasts and bone homeostasis. Biochim Biophys Acta. 2013;1831:223-227.

14. Ishii M, Kikuta J, Shimazu Y, Meier-Schellersheim M, Germain RN. Chemorepulsion by blood S1P regulates osteoclast precursor mobilization and bone remodeling in vivo. J Exp Med. 2010;207:2793-2798.

15. Kono M, Belyantseva IA, Skoura A, et al. Deafness and stria vascularis defects in S1P2 receptor-null mice. J Biol Chem. 2007;282:10690-10696.

16. Ryu J, Kim HJ, Chang E-J, Huang H, Banno Y, Kim H-H. Sphingosine 1-phosphate as a regulator of osteoclast differentiation and osteoclast-osteoblast coupling. EMBO J. 2006;25:5840-5851.

17. Kikuta J, Kawamura S, Okiji F, et al. Sphingosine-1-phosphate-mediated osteoclast precursor monocyte migration is a critical point of control in antigen/resorptive action of active vitamin D. Proc Natl Acad Sci U S A. 2013;110:7007-7013.

18. Matsuzaki E, Hiratsuka S, Hamachi T, et al. Sphingosine-1-phosphate promotes the nuclear translocation of beta-catenin and thereby induces osteoprotegerin gene expression in osteoblast-like cell lines. Bone. 2013;55:315-324.

19. Kim B-J, Shin K-O, Kim H, et al. The effect of sphingosine-1-phosphate on bone metabolism in humans depends on its plasma/bone marrow gradient. J Endocrinol Invest. 2016;39:297-303.

20. Hsu L-C, Reddy SV, Yilmaz Ö, Yu H. Sphingosine-1-phosphate receptor 2 controls podosome components induced by RANKL affecting osteoclastogenesis and bone resorption. Cells. 2019;8(1):17.

21. Udagawa N, Takahashi N, Akatsu T, et al. Origin of osteoclasts: mature monocytes and macrophages are capable of differentiating into osteoclasts under a suitable microenvironment prepared by bone marrow-derived stromal cells. Proc Natl Acad Sci U S A. 1990;87:7260-7264.

22. Quint P, Ruan M, Pederson L, et al. Sphingosine 1-phosphate (S1P) receptors 1 and 2 coordinately induce mesenchymal cell migration through S1P activation of complementary kinase pathways. J Bio Chem. 2013;288:5398-5406.

23. Okamoto H, Takawa N, Yokomizo T, et al. Inhibitory regulation of Rac activation, membrane ruffling, and cell migration by the G protein-coupled sphingosine-1-phosphate receptor EDG5 but not EDG1 or EDG3. Mol Cell Biol. 2000;20:9247-9261.

24. Lepley D, Paik JH, Hla T, et al. The G protein-coupled receptor S1P2 regulates Rho/Rho kinase pathway to inhibit tumor cell migration. Cancer Res. 2005;65:3788-3795.

25. Sato C, Iwatsuki T, Kito S, Tsunemi S, Sano H. Sphingosine 1-phosphate receptor activation enhances BMP-2-induced osteoblast differentiation. Biochem Biophys Res Commun. 2012;423:200-205.

26. Higashi K, Matsuzaki E, Hashimoto Y, et al. Sphingosine-1-phosphate/S1PR2-mediated signaling triggers Smad1/5/8 phosphorylation and thereby induces Runx2 expression in osteoblasts. Bone. 2016;93:1-11.

27. Vella G, Bergers G. Where have all the T cells gone? Immunity. 2018;49:592-594.

28. Grey A, Xu X, Hill B, et al. Osteoblast cells express phospholipid receptors and phospholipases and proliferate in response to sphingosine-1-phosphate. Calcif Tissue Int. 2004;74:542-550.

29. Grey A, Chen Q, Callon K, et al. The phospholipids sphingosine-1-phosphate and lysophosphatidic acid prevent apoptosis in osteoblastic cells via a signaling pathway involving G(i) proteins and phosphatidylinositol-3 kinase. Endocrinology. 2002;143:4755-4763.

30. Sukovecha OA. Expansion of sphingosine kinase and sphingosine-1-phosphate receptor function in normal and cancer cells: from membrane restructuring to mediation of estrogen signaling and stem cell programming. Int J Mol Sci. 2018;19:E420.

31. Lampa JD, Kamer A, Margarone J, Dziak R. Sphingosine-1-phosphate effects on PKC isoform expression in human osteoblastic cells. Prostaglandins Leukot Essent Fatty Acids. 2001;65:139-146.

32. Hutami IR, Izawa T, Mino-Oka A, et al. Fas/S1P1 crosstalk via NF-kappaB activation in osteoclasts controls subchondral bone remodeling in murine TMJ arthritis. Biochem Biophys Res Commun. 2017;490:1274-1281.

33. Hwang SY, Putney JW Jr. Calcium signaling in osteoclasts. Biochim Biophys Acta. 2011;1813:979-983.

34. Wang H, Tian L, Liu J, et al. The osteogenic niche is a calcium reservoir of bone micrometastases and confers unexpected therapeutic vulnerability. Cancer Cell. 2018;34(5):823-839.e7.

35. Duncan RL, Akanbi KA, Farach-Carson MC. Calcium signals and calcium channels in osteoblastic cells. Semin Nephrol. 1998;18:178-190.

36. Wu X, Pan G, McKenna MA, Zayzafoon M, Xiong W-C, McDonald JM. RANKL regulates Fas expression and Fas-mediated apoptosis in osteoclasts. J Bone Miner Res. 2005;20:107-116.

37. Dobrosak C, Gooi JH. Increased sphingosine-1-phosphate production in response to osteocyte mechanotransduction. Bone Rep. 2017;7:114-120.

38. Ji F, Mao LL, Liu Y, et al. K6PC-S, a novel sphingosine kinase 1 (SphK1) activator, alleviates dexamethasone-induced damages to osteoblasts through activating SphK1-Akt signaling. Biochim Biophys Res Commun. 2015;458:568-575.

39. Brizuela L, Martin C, Jeannot P, et al. Osteoblast-derived sphingosine-1-phosphate to induce proliferation and confer resistance to therapeutics to bone metastasis-derived prostate cancer cells. Mol Oncol. 2014;8:1181-1195.

40. Kozawa O, Tokuda H, Matsuno H, Uematsu T. Activation of mitogen-activated protein kinase is involved in sphingosine-1-phosphate-stimulated interleukin-6 synthesis in osteoblasts. FEBS Lett. 1997;418:149-151.

41. Coughlan T, Dockery F. Osteoporosis and fracture risk in older people. Clin Med. 2014;14:187-191.

42. Jung J, Park JS, Righesso L, et al. Effects of an oral bisphosphonate and three intravenous bisphosphonates on several cell types in vitro. Clin Oral Investig. 2018;22:2527-2534.

43. Diel U, Bergner R, Grotz KA. Adverse effects of bisphosphonates: current issues. Supportive Oncol. 2007;5:475-482.

44. Bone HG, Bolognese MA, Yuen CK, et al. Effects of denosumab treatment and discontinuation on bone mineral density and bone turnover markers in postmenopausal women with low bone mass. J Endocrinol Metabol. 2011;96:972-980.

45. Bandeira L, Lewiecki EM, Bliezkijan JP. Rosomozumab for the treatment of osteoporosis. Expert Opin Biol Ther. 2017;17:255-263.

46. Lemaire V, Cox DR. Dynamics of bone cell interactions and differential responses to PTH and antibody-based therapies. Bull Math Biol. 2019;81(9):3575-3622.

47. Weske S, Vaidya M, Reese A, et al. Targeting sphingosine-1-phosphate lyase as an anabolic therapy for bone loss. Nat Med. 2018;24:667-678.

48. Keller J, Catala-Lehnen P, Huebner AK, et al. Calcitonin controls bone formation by inhibiting the release of sphingosine 1-phosphate from osteoclasts. Nat Commun. 2014;5:5215.

49. Tantikanlayaporn D, Tourkova IL, Larrouette Q, et al. Sphingosine-1-phosphate modulates the effect of estrogen in human osteoblasts. JBMR Plus. 2018;2:217-226.
63. Heo JY, Im DS. Pro-Inflammatory role of S1P3 in macrophages.

62. Hammad SM, Crellin HG, Wu BX, Melton J, Anelli V, Obeid LM.

67. Dou CE, Ding N, Zhao C, et al. Estrogen deficiency-mediated M2

65. Yang J, Yang L, Tian L, et al. Sphingosine 1-phosphate (S1P)/S1P

64. Keul P, Lucke S, von Wnuck Lipinski K, et al. Sphingosine-1-phosphate

59. Lee H, Liao JJ, Graeler M, et al. Lysophospholipid regulation of mono

53. Pederson L, Ruan M, Westendorf JJ, Khosla S, Oursler MJ.

58. Huang R, Wang X, Zhou Y, Xiao Y. RANKL-induced M1 macro

55. Hutami IR, Tanaka E, Izawa T. Crosstalk between Fas and S1P1 sig

52. Carpio LC, Shiau H, Dziak R. Changes in sphingolipid levels induced

50. Sukocheva O, Wadham C, Holmes A, et al. Estrogen transactivates

44. RANKL-induced M1 macrophages with IL-10, IL-4 and TGF-β signals to

43. Heo JY, Im DS. Pro-Inflammatory role of S1P3 in macrophages.

42. Pederson L, Ruan M, Westendorf JJ, Khosla S, Oursler MJ.

41. Huang R, Wang X, Zhou Y, Xiao Y. RANKL-induced M1 macro

40. Lee H, Liao JJ, Graeler M, et al. Lysophospholipid regulation of mono

39. Hutami IR, Tanaka E, Izawa T. Crosstalk between Fas and S1P1 sig

38. Heo JY, Im DS. Pro-Inflammatory role of S1P3 in macrophages.

37. Xiao L, Zhou Y, Zhu L, et al. SPHK1/S1PR1-RANKL Axis Regulates the Interactions Between Macrophages and BMSCs in Inflammatory Bone Loss. J Bone Miner Res. 2018;33:1090-1104.

36. Huang R, Wang X, Zhou Y, Xiao Y. RANKL-induced M1 macro

35. Lee H, Liao JJ, Graeler M, et al. Lysophospholipid regulation of mono

34. Hutami IR, Tanaka E, Izawa T. Crosstalk between Fas and S1P1 sig

33. Xiao L, Zhou Y, Zhu L, et al. SPHK1-S1PR1-RANKL Axis Regulates the Interactions Between Macrophages and BMSCs in Inflammatory Bone Loss. J Bone Miner Res. 2018;33:1090-1104.

32. van der Weyden L, Arends MJ, Campbell AD, et al. Genome-wide in

31. Huang R, Wang X, Zhou Y, Xiao Y. RANKL-induced M1 macro

30. Lee H, Liao JJ, Graeler M, et al. Lysophospholipid regulation of mono

29. Hutami IR, Tanaka E, Izawa T. Crosstalk between Fas and S1P1 sig

28. Heo JY, Im DS. Pro-Inflammatory role of S1P3 in macrophages.

27. Keul P, Lucke S, von Wnuck Lipinski K, et al. Sphingosine-1-phosphate receptor 3 promotes recruitment of monocyte/macrophages in in

26. Hammad SM, Crellin HG, Wu BX, Melton J, Anelli V, Obeid LM. Dual and distinct roles for sphingosine kinase 1 and sphingosine 1

25. Yang J, Yang L, Tian L, et al. Sphingosine-1-phosphate (S1P)/S1P

24. Keul P, Lucke S, von Wnuck Lipinski K, et al. Sphingosine-1-phosphate receptor 3 promotes recruitment of monocyte/macrophages in in

23. Hammad SM, Crellin HG, Wu BX, Melton J, Anelli V, Obeid LM. Dual and distinct roles for sphingosine kinase 1 and sphingosine 1

22. Yang J, Yang L, Tian L, et al. Sphingosine-1-phosphate (S1P)/S1P

21. Keul P, Lucke S, von Wnuck Lipinski K, et al. Sphingosine-1-phosphate receptor 3 promotes recruitment of monocyte/macrophages in in

20. Hammad SM, Crellin HG, Wu BX, Melton J, Anelli V, Obeid LM. Dual and distinct roles for sphingosine kinase 1 and sphingosine 1

19. Yang J, Yang L, Tian L, et al. Sphingosine-1-phosphate (S1P)/S1P

18. Keul P, Lucke S, von Wnuck Lipinski K, et al. Sphingosine-1-phosphate receptor 3 promotes recruitment of monocyte/macrophages in in

17. Hammad SM, Crellin HG, Wu BX, Melton J, Anelli V, Obeid LM. Dual and distinct roles for sphingosine kinase 1 and sphingosine 1
90. El Jamal A, Bougault C, Mebarek S, Magne D, Cuvillier O, Brizuela L. The role of sphingosine 1-phosphate metabolism in bone and joint pathologies and ectopic calcification. *Bone*. 2020;130:115087.

91. Xiao L, Zhou Y, Friis T, Beagley K, Xiao Y. S1P–S1PR1 signaling: the “Sphinx” in osteoimmunology. *Front Immunol*. 2019;10:1409.

92. Ng ML, Yarla NS, Menschikowski M, Sukocheva OA. Regulatory role of sphingosine kinase and sphingosine-1-phosphate receptor signaling in progenitor/stem cells. *World J Stem Cells*. 2018;10(9):119-133.

93. Beider K, Rosenberg E, Bitner H, et al. The sphingosine-1-phosphate modulator FTY720 targets multiple myeloma via the CXCR4/CXCL12 pathway. *Clin Cancer Res*. 2017;23:1733-1747.

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