Research Article

Administration of Myelin Basic Protein Peptides Encapsulated in Mannosylated Liposomes Normalizes Level of Serum TNF-α and IL-2 and Chemoattractants CCL2 and CCL4 in Multiple Sclerosis Patients

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We have previously shown that immunodominant MBP peptides encapsulated in mannosylated liposomes (Xemys) effectively suppressed experimental allergic encephalomyelitis (EAE). Within the frames of the successfully completed phase I clinical trial, we investigated changes in the serum cytokine profile after Xemys administration in MS patients. We observed a statistically significant decrease of MCP-1/CCL2, MIP-1β/CCL4, IL-7, and IL-2 at the time of study completion. In contrast, the serum levels of TNF-α were remarkably elevated. Our data suggest that the administration of Xemys leads to a normalization of cytokine status in MS patients to values commonly reported for healthy subjects. These data are an important contribution for the upcoming Xemys clinical trials.

1. Introduction

Multiple sclerosis (MS) is an autoimmune neurologic disease with unclarified etiology. It is well established that the proliferation of autoreactive T and B cells leads to CNS inflammation and subsequent myelin destruction. CD4+ T cells are believed to be the essential mediators of the inflammatory process in MS. However, the role of B cells, macrophages, and T cells other than CD4+ should not be neglected [1]. Almost all chronic inflammatory diseases and immunopathologies are linked to dysregulated cytokine responses; thus it is evident that cytokine status plays an important role in MS progression [2, 3]. Cytokines and chemokines not only suppress or activate different types of immune cells, but also can facilitate the migration of pathogenic cells through the blood-brain barrier (BBB) [4].

There are a large number of studies describing cytokine status by direct analysis in the blood and cerebrospinal fluid (CSF) [5, 6] and determining mRNA expression in peripheral blood mononuclear cells (PBMCs) of MS patients [7, 8]. Generally, it is believed that the immune response in MS is shifted towards Th1 cytokine production (IL-6, IL-12, IL-2, IFN-γ, and TNF-α). The most notable study of cytokine involvement in MS progression involved the administration of interferons in humans. While IFN-β is fully approved [9, 10] and appears to be one of the most common treatments for MS worldwide the injection of IFN-γ threatens the health and life of MS patients [11].
Along with the physiological assessment, the level of cytokines may be an important diagnostic criterion [12]. During clinical trials, it is a common practice to analyze cytokine levels in different physiological fluids, particularly in terms of the Th1/Th2 ratio; this analysis may shed light on disease pathogenesis, mechanism of drug action and methods of their improvement, and modifications in administration strategy. It was shown that after long-term IFN-β therapy, the serum concentration of IFN-γ decreased [13]. Others have shown that treatment with IFN-β results in increased levels of TNF-α and decreased levels of IL-5. Additionally, the combination of IFN-β administration with atorvastatin therapy increases the levels of proinflammatory cytokine IL-12p70 [14]. A significantly elevated Th2/Th1 ratio was shown to be a hallmark of glatiramer acetate administration [15]. Recently, a similar effect was shown in MS patients treated with monoclonal anti-α4-integrin antibody (natalizumab) [16].

Previously, we reported the efficiency of immunodominant MBP peptides encapsulated in mannosylated liposomes (Xemys) in suppressing experimental allergic encephalomyelitis (EAE) [17, 18]. This therapeutic composition was created in parallel with a variety of techniques to induce tolerance towards MBP [19–24]. Our study demonstrated that the uptake of distinct MBP peptides, which are immunodominant in terms of autoantibody response in MS patients [25], by dendritic cells was enhanced by mannosylation of carrier liposomes. In frames of successfully completed phase I clinical studies, we analyzed the serum cytokine profile in MS patients treated with Xemys. The aim of this study was to characterize changes in cytokine levels and the Th1/Th2 ratio after Xemys administration.

### 2. Materials and Methods

#### 2.1. Patients.

Phase I included patients with relapse–remitting MS (RRMS) or secondary progressive MS (SPMS) according to the McDonald diagnostic criteria in 2005 [26]. A total of 18 MS patients were subcutaneously administered with a total of 2.675 mg of encapsulated MBP peptides. Patients were enrolled to receive weekly subcutaneous injection (s.c.) of Xemys at increasing doses from 50 μg to 900 μg over six weeks. All patients had a follow-up at 6–18 weeks. The study was authorized by the Russian Public Health Ministry #930 [FASEMS-01/01], which was issued on April 28, 2012. All patients provided written informed consent at the time of enrollment.

#### 2.2. Profiling of Serum Cytokines.

A cytokine profile analysis was performed with serum samples (collected at baseline and during all follow-up visits) using a multiplexed fluorescent magnetic bead-based immunoassay (Bio-Rad Laboratories, USA) according to the manufacturer’s instructions. The following 17 different cytokines and chemokines were assessed: IL-1β, IL-2, IL-4, IL-5, IL-6, IL-7, IL-8, IL-10, IL-12 (p70), IL-13, IL-17A, granulocyte colony-stimulating factor (G-CSF), granulocyte macrophage colony-stimulating factor (GM-CSF), IFN-γ, monocyte chemoattractant protein-1 (MCP-1/CCL2), macrophage inflammatory protein (MIP-1β/CCL4), and tumor necrosis factor-alpha (TNF-α). All samples were measured in triplicate. For the baseline, the cytokine and chemokine levels in healthy subjects (median and IQR) from previously published data were used for CCL2, CCL4, IL-6, IL-7, IL-10, IL-12, G-CSF [27], IL-8, IL-17, TNF-α [28], IFN-γ [29], IL-4 [30], IL-13 [31], IL-5 [32], and IL-2 (http://www.quanterix.com/files/assays/IL-2%20Data%20Sheet%20ID%204904%201165.pdf).

#### 2.3. Statistical Analysis.

Data were analyzed by using the Sigma-Plot 12.5 and Statistica 10 software. The difference in the cytokine levels before and after treatment was compared by using Student’s t-test, nonparametric Mann-Whitney U test, and Wilcoxon signed-rank test. If the measured values were under the detection limit, they were considered to be equal to the value of the detection limit. A two-sided p value < 0.05 was considered statistically significant. The difference was considered reliable when confirmed by at least one test.

### 3. Results and Discussion

Phase I Xemys clinical studies involved 16 (80%) patients with RRMS and 4 (20%) patients with SPMS with relapses. Baseline characteristics of MS patients are listed in Table 1. Three (15%) patients had a mild disability according to the expanded disability status scale (EDSS) (3.0) and 17 (85%) patients showed moderate disability (3.5–5.5) at baseline. Nineteen MS patients were subcutaneously administered a total of 2.675 mg encapsulated MBP peptides, which was divided into 6 injections with increasing dose every week (weeks 1–6). One patient received only the initial 50 μg dose of Xemys and was discontinued from the study during the first week of the treatment period. A complete set of serum samples was obtained for 18 patients. To analyze the immunological consequences of Xemys administration, the levels of 17 serum cytokines and chemokines were analyzed...
Table 2: Level of serum cytokines and chemokines (pg/mL) in MS patients at baseline and follow-up period.

| Cytokine   | −2 weeks | IQR       | Median | IQR       | 7 weeks | IQR       | Median | IQR       | 10 weeks | IQR       | Median | IQR       | 18 weeks | IQR       |
|------------|----------|-----------|--------|-----------|---------|-----------|--------|-----------|----------|-----------|--------|-----------|----------|-----------|
| TNF-α      | 5.7      | 4.2–8.7   | 6.1    | 2.6–15.9  | 4.2     | 3.4–9.3   | 9.2    | 6.6–14.5  |          |           |        |           |          |           |
| IFN-γ      | 155      | 107–226   | 155    | 58–332    | 100     | 33–203    | 171    | 91–360    |          |           |        |           |          |           |
| CCL4       | 337      | 311–378   | 293    | 208–429   | 337     | 229–441   | 278    | 216–394   |          |           |        |           |          |           |
| CCL2       | 97       | 78–123    | 90     | 57–142    | 82      | 49.3–111  | 86     | 64–98     |          |           |        |           |          |           |
| G-CSF      | 9.5      | 5.7–23.2  | 10.2   | 7.2–14.6  | 9.0     | 5.6–18.6  | 12     | 8–32      |          |           |        |           |          |           |
| IL-1β      | 2.5      | 2.4–4.0   | 3.6    | 2.9–6.4   | 2.9     | 2.3–3.2   | 3.2    | 3.1–3.7   |          |           |        |           |          |           |
| IL-2       | 1.4      | 0.4–22.4  | 0.4    | 0.4–1.6   | 0.7     | 0.4–10.7  | 0.5    | 0.4–10    |          |           |        |           |          |           |
| IL-4       | 1.6      | 1.2–3.7   | 1.5    | 1.3–2.8   | 1.6     | 1.0–2.2   | 1.6    | 1.3–2.6   |          |           |        |           |          |           |
| IL-5       | 5.6      | 3.6–7.8   | 4.7    | 3.2–9.3   | 4.9     | 3.0–7.2   | 6.4    | 3.4–9.2   |          |           |        |           |          |           |
| IL-6       | 8.6      | 5.6–14.3  | 9.6    | 3.4–14.8  | 8.7     | 5.1–16.6  | 8.5    | 5.7–13.3  |          |           |        |           |          |           |
| IL-7       | 15       | 12–23     | 12.1   | 5.8–17.2  | 12.5    | 9.4–13.6  | 11.5   | 9.2–13.6  |          |           |        |           |          |           |
| IL-8       | 25       | 16–39     | 23     | 17–146    | 19      | 16–35     | 20     | 16–30     |          |           |        |           |          |           |
| IL-10      | 14       | 7–25      | 12.4   | 6.3–15.2  | 9.5     | 6.3–16.8  | 12     | 7–19      |          |           |        |           |          |           |
| IL-12      | 23       | 17–67     | 18     | 13–28     | 17      | 11–60     | 32     | 16–57     |          |           |        |           |          |           |
| IL-13      | 4.4      | 2.7–6.8   | 3.2    | 2.7–7.0   | 3.7     | 2.9–4.8   | 4.9    | 3.6–6.0   |          |           |        |           |          |           |
| IL-17      | 9.3      | 4.6–19.7  | 10.4   | 3.6–18.8  | 11.3    | 4.6–17.1  | 11     | 3.9–14.1  |          |           |        |           |          |           |
| GM-CSF     |          | Under detection limit |          |          |          |           |        |           |          |           |        |           |          |           |

at baseline (week −2) and during the follow-ups (weeks 7, 10, and 18) (Table 2).

As anticipated, in comparison to healthy individuals, MS patients at baseline revealed increased levels of proinflammatory cytokines and chemokines IFN-γ, IL-2, IL-8, IL-17, CCL2, and CCL4 whereas level of anti-inflammatory cytokines IL-4, IL-10, and IL-13 was not significantly dysregulated. These data partially correlate with previously reported data [3] demonstrating increased level of IL-12, IFN-γ, IL-17, and CCL4 in cerebrospinal fluid (CSF) of MS patients. Interestingly, level of CCL2 in opposite was shown to decrease in CSF during MS. In the present study, a statistically significant decrease was observed in the serum level of MCP-1/CCL2, MIP-1β/CCL4, IL-7, and IL-2 at the time of study completion (week 18) (Figure 1). Importantly, the median level of all effector molecules, except IL-7, was outside of the interquartile range that corresponded to the healthy subjects, suggesting that they were significantly upregulated in MS patients. Our data are in accordance with studies reporting that both CCL2 [33] and CCL4 [34, 35] are elevated during the course of MS. These chemokines recruit monocytes, memory T cells, and dendritic cells to the sites of inflammation, across the BBB and within the CNS parenchyma [36]. Cheng et al. reported that treatment with IFN-β results in decreased CCL2 and CCL4 levels in the CNS of C57BL/6 mice inflicted with EAE [37]. The mechanism is complicated for MS because it elevate only in the blood and is decreased in the CSF; this likely involves the action of CCR2-positive migrating cells as they cross the BBB [38].

Previously, we reported that treatment of DA rats with encapsulated MBP peptides resulted in a downregulation of the proinflammatory cytokine IL-2 in the CNS [18]. In line with this, the levels of serum IL-2 were decreased in MS patients after Xemys treatment. It should be mentioned that, for 40% of patients (7/18), IL-2 concentrations were under the detection limit (0.4 pg/mL) at all follow-up time points, while for the rest of the patients, except for 1, elevated IL-2 levels returned to the levels of healthy subjects after Xemys administration.

IL-7 is a cytokine that is important for B and T cell development [39]. This cytokine forms a heterodimer with the hepatocyte growth factor (HGF) and the heterodimer functions as a pre-pro-B cell growth-stimulating factor [40]. In contrast, Lee et al. showed that increased IL-7 in the serum is a hallmark of the Th1-driven form of MS, and the blockade of IL-7 and the IL-7Rα pathway may have a therapeutic potential in MS [41]. Thus, the observed downregulation of IL-7 after Xemys treatment may be therapeutically beneficial.

Our data suggest that the levels of serum TNF-α are elevated after Xemys administration (Figure 1). This statistically significant observation may have important physiological relevance. Interestingly, TNF-α is considered to be a potent mediator of inflammation in MS [42]. It should be noted that, in the CSF, but not in the serum of MS patients, TNF-α levels are significantly higher and correlates with the severity and progression of the disease [43]. However, it was suggested that TNF-α is a potent anti-inflammatory cytokine in autoimmune demyelination [44, 45] and TNF-α-related genes are downregulated during MS progression [8]. Clinical studies with TNF-α-inhibiting agents revealed an increased frequency of MS relapse [46–48]. Additional evidence to show that TNF-α influences MS could be that a shorter TNF-α receptor 1 with two “A” instead of two “G” increases the probability of MS development to 12% by mimicking the effect of TNF-blocking drugs [49, 50].
Serum concentrations of IL-8, IL-6, and IL-10 decreased to the levels of healthy subjects without any statistically significant differences (Figure 1). This observation is promising because previous studies reported that elevated levels of these cytokines were associated with MS progression. PBMC gene expression analysis revealed that IL-8 was significantly overexpressed in patients with MS and other autoimmune diseases [51]. Moreover, the levels of IL-8 are elevated in the CSF of MS patients [52]. Correlative analysis between the cytokine profile and the progression of the disease [29] revealed that the levels and ranges of IL-6 are elevated in MS patients and this increases with disease severity [29]. Our data showed that the median value of IL-6 had not changed because only a few cases showed elevated levels for this cytokine and all levels were restored to normal levels of healthy subjects upon study completion. Recently published data report anti-inflammatory activities of IL-6 protecting mice from organ specific autoimmune disease
by IL-6 classical signalling-dependent IL-1ra induction [53]. Thus IL-6 may be potentially considered as beneficial in combination with Xemics. IL-10 was slightly elevated in MS patients, but there was no correlation with disease severity [29]. The other cytokine levels were unchanged, while the level of GM-CSF was under the detection limit.

The balance between Th1 and Th2 cytokines is very important as they can functionally cross-inhibit each other. MS is considered to be a Th1-mediated disorder; however, number of studies argue with this [54–56]. Th1 cells and their related activation pathways are linked to the production of IFN-γ and, to a lesser extent, with IL-12, IL-2, and TNF-α. Th2 cells are heavily reliant on IL-4, IL-5, and IL-10. Therefore, we defined the Th1/Th2 ratio between IFN-γ, TNF-α, and IL-12 towards IL-4, IL-5, IL-6, and IL-10 (Figure 2). Interestingly, the administration of Xemics shifted the Th1/Th2 ratio towards mostly Th1. The observed shift may be due to the downregulation of the Th2 cytokines IL-6 and IL-10 and an elevated production of TNF-α rather than by a systemic elevation of Th1 cytokines.

4. Conclusions
In terms of serum cytokines, a reduction of inflammation and restriction of monocye cell trafficking are immunological consequences of the administration of encapsulated MBP peptides in MS patients. In the present study, we found that cytokine levels returned to normal levels of healthy subjects after Xemics treatment in MS patients, especially for IL-2, IL-7, CCL2, CCL4, and TNF-α. However, the levels of IFN-γ were unchanged and significantly higher than healthy subjects. The administration of IFN-β decreased IFN-γ secretion and inhibited IFN-γ-related responses in MS patients [57–59]. Therefore, concomitant therapy utilizing
Xemys in combination with an anti-IFN-γ treatment [60] may be more beneficial. This investigation is preliminary and definitive conclusions could not be made because the cohort is limited and heterogeneous, the follow-up period was short, and the placebo group was absent. Nonetheless, the observed changes in serum cytokines and chemokines may be important to consider for the upcoming Xemys phase II clinical trial.

Competing Interests
Dmitry Genkin and Konstantin Zakharov are employees of the sponsoring company Pharmasynthez OJSC (Saint Petersburg, Russia).

Authors’ Contributions
Yakov Lomakin and Alexey Belogurov Jr. contributed equally in this work.

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