Genomes of *Alteromonas australica*, a world apart

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**Abstract**

**Background:** *Alteromonas* is a genus of marine bacteria that is very easy to isolate and grow in the laboratory. There are genomes available of the species *Alteromonas macleodii* from different locations around the world and an *Alteromonas* sp. isolated from a sediment in Korea. We have analyzed the genomes of two strains classified by 16S rRNA (>99% similarity) as the recently described species *Alteromonas australica*, and isolated from opposite ends of the world; *A. australica* DE170 was isolated in the South Adriatic (Mediterranean) at 1000 m depth while *A. australica* H17⁴ was isolated from a sea water sample collected in St Kilda Beach, Tasman Sea.

**Results:** Although these two strains belong to a clearly different species from *A. macleodii*, the overall synteny is well preserved and the flexible genomic islands seem to code for equivalent functions and be located at similar positions. Actually the genomes of all the *Alteromonas* species known to date seem to preserve synteny quite well with the only exception of the sediment isolate SN2. Among the specific metabolic features found for the *A. australica* isolates there is the degradation of xylan and production of cellulose as extracellular polymeric substance by DE170 or the potential ethanol/methanol degradation by H17⁴.

**Conclusions:** The genomes of the two *A. australica* isolates are not more different than those of strains of *A. macleodii* isolated from the same sample. Actually the recruitment from metagenomes indicates that all the available genomes are found in most tropical-temperate marine samples analyzed and that they live in consortia of several species and multiple clones within each. Overall the hydrolytic activities of the *Alteromonas* genus as a whole are impressive and fit with its known capabilities to exploit sudden inputs of organic matter in their environment.

**Keywords:** *Alteromonas australica*, Biogeography, *Alteromonas*, Genomic Island, Population genomics, Integron

**Background**

The genus *Alteromonas* contains species of marine bacteria that have been isolated often from the oceans around the world [1,2]. The type species *Alteromonas macleodii* is found worldwide but mostly at temperate or tropical latitudes due to its mesophilic nature (grows between 10 and 45°C) (López-Pérez and Rodriguez-Valera, in press). This microbe was shown to be enriched in the particulate fraction of the water column in off-shore Mediterranean waters [3,4]. In the Mediterranean Sea, bacteria of this species can be found even at high depths due to the relatively warm temperatures of its deep water mass (ca. 13°C) [5]. Related microbes have been found to bloom in mesocosms in the central Pacific gyre when the water was enriched with natural seawater nutrients [6,7]. Other species of *Alteromonas* (e.g., *A. stellipolaris*) have been isolated from polar environments (Antarctic sea water) and are more psychrophilic [8].

Previously the genomes of two *A. macleodii* strains, AltDE and AltDE1 isolated from the South Adriatic at 1000 m depth were analyzed [9]. The average nucleotide identity (ANI) over the core genome between the two strains was 98.5%. From the same sample several other strains were isolated. The characterization by PCR amplification of their 16S rRNA gene classified one of them as belonging (99% identity) to the recently described species *Alteromonas australica* [10]. This bacterium (strain H17⁴) was collected from the first meters below the surface at St. Kilda beach, in Port Phillip Bay (Tasman Sea, Pacific Ocean). When comparing AltDE and AltDE1 [9] in spite of their high similarity, their genomes differ markedly in their content of flexible genomic regions or islands. These can be classified into two main types [11]. One type is typically associated to mobile genetic elements (including lysogenic phages). They tend to be found at the same genomic location and context but they are often

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completely absent from some strains. They have been designated “additive genomic islands” [11] because they typically vary by the presence/absence of gene cassettes in some strains but not in others. They carry lifestyle related genes, that include different metabolic properties (for example the degradation of organic compounds such as urea) or biotechnologically relevant features such as the polysaccharide or the polyketide synthase (PKS) clusters [12]. The second type are the “replacement genomic islands”, in which completely different gene clusters with similar assigned functions are found at the same genomic location. They are typically involved in structural features of the cells exposed to the environment. Recent evidence indicates that they are exchanged by homologous recombination [13]. The gene cluster coding for the genes of the O-chain (sometimes called O-antigen) of the Gram negative lipopolysaccharide being a paradigmatic example.

This study aimed to provide a detailed genomic analysis of the two A. australica strains. We have fully sequenced and assembled these two genomes and compared them with all the available genomes of strains within the genus Alteromonas. The patterns of variation observed between the two A. australica strains are very similar to the ones found before when comparing the isolates from the same location mentioned above [9,11]. Across the genus, the location and functional nature of the genomic islands was preserved, although obviously the genetic content was very different. However, some remarkable examples of conservation were found in the additive genomic islands with the members of A. macleodii.

Results

We have assembled into a single contig the genome of the two isolates, A. australica H17T [10] isolated from the Tasman Sea and A. australica DE170 from the South Adriatic (16000 km away). The general features of the strains and the comparison with other members from the Alteromonas genus sequenced thus far are shown in Table 1. We have included in the comparison all previously assembled genomes of Alteromonas species [14-16] and the genome of the isolate Alteromonas sp. ALT199 from the Pacific Ocean, deposited as a draft genome (although it was assembled into one single contig) at the Joint Genome Institute (JGI, http://www.jgi.doe.gov/) (Table 1). In spite of the different locations and depth of the sampling sites, the two A. australica strains form a highly homogeneous clade with an average nucleotide identity (ANI) of 98.6%. The pairwise genome comparison of the A. australica isolates with all available strain genomes of the Alteromonas genus gave an ANI value of ca. 74% (Table 1). This is consistent with A. australica belonging to a different species within the same genus. The geographic origin of the isolates is presented in Additional file 1: Figure S1. Only 2224 genes were conserved in all the genomes shown in Table 1 (about 50% of the average genome gene content) (see Methods). Regarding the number of strain-specific genes, it was variable, ranging from 1170 for Alteromonas sp. SN2 to 260 for A. australica H17T (Table 1). The alignment of the genomes showed that the synteny was remarkably well preserved in all the strains, except SN2, that showed large rearrangement from rRNA operon 1 to rRNA operon 5 (Figure 1).

To investigate the phylogenetic relationships of both A. australica strains within the Alteromonas genus, whole-genome phylogeny was inferred from a concatenation of shared genes (ca. 1.4 Mb) including all the completely sequenced species (Figure 2). Pseudoalteromonas atlantica T6c, a marine bacterium which belong to the separate family Pseudoalteromonadaceae [17], was used as an outgroup. The A. macleodii isolates are split by the surface (small particle)/deep (larger particle) ecotypes (or subspecies) [11,14,18] while A. australica and Alteromonas sp. SN2 appear clearly separated as expected from different species of the same genus. Alteromonas sp. ALT199 clustered within the same branch of the members of the A. macleodii surface clade, formed by the strains AD45, BS11, 673 and the type strain ATCC 27126T [18].

In order to analyzed the impact of the homologous recombination in the evolution of the core genome of the members of the Alteromonas genomes we used ClonalFrame software v1.2 [19] to estimate two different values, r/m and ρ/θ. The parameter r/m shows the weight of recombination in the diversification of the species relative to mutation, whereas ρ/θ measures the absolute number of events of recombination relative to mutation. The results showed that within the Alteromonas genus the 95% credibility interval of ρ/θ was 0.01-0.02 (mean = 0.01), indicating that recombination happened hundred times less frequently than mutation. However, the impact of recombination had a predicted significant effect in the evolutionary process as showed by the r/m value of, 0.60 (with 95% credibility interval 0.45-0.91), suggesting that recombination introduces several polymorphisms per event. The average size of the fragments affected by recombination was 1102 bp (with a 95% credibility interval of 799 to 1349 bp), similar to those found in other bacteria, such as Bacillus cereus [20] and Chlamydia psittaci [21]. In a previous study carried out with the nine genomes belonging to the A. macleodii deep ecotype [11] we obtained similar results for the comparison of all the A. macleodii members (deep and surface ecotype) with values of r/m = 0.45 and ρ/θ = 0.06. However, when only the deep ecotype members were considered, both parameters were higher, ρ/θ (0.58) and r/m (5.13), suggesting that recombination is an important factor in the evolution within this clade. Using the locally collinear blocks generated from the whole-genome
alignments we have also analyzed the existence of recombination events by building ML trees of randomly selected regions (Additional file 1: Figure S2). The results show frequent variations of the topology suggesting recombination events taking place across the Alteromonas genus, but only in small fragments of less than 3 Kb. However, topology was stable when fragments of more than 5 Kb were selected.

Flexible genomic islands

The alignment of the two A. australica genomes and of other representative species of Alteromonas is shown in Figure 1. As has been shown several times [9,11,22-25] a significant part of the flexible genome is found in genomic islands larger than 15 Kb. As previously [11], we have classified them into two categories, the flexible genomic islands (fGIs) that are found in both isolates and at equivalent position in the genomes (same genome context) but contain different genes, and the unique islands that are found in either one or the other strain (Table 2). fGIs preserve the same location (with regard to the origin of replication) and the same genome context (neighboring genes) even when different species of the same genus are

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**Table 1 General features of genomes**

| Alteromonas strain | A. australica H17 | A. australica DE170 | A. macleodii ATCC 27126 | Alteromonas sp. ALT199 | Alteromonas sp. SN2 |
|--------------------|------------------|---------------------|------------------------|------------------------|-----------------------|
| Size (bp)          | 4308870          | 4447535             | 4480937                | 4607844                | 4635654               |
| GC content (%)     | 44.9             | 44.9                | 44.9                   | 44.6                   | 43.7                  |
| Total ORFs         | 3764             | 4252                | 4346                   | 4444                   | 3955                  |
| Share Genes        | 2289             | 2296                | 2260                   | 2283                   | 2279                  |
| Unique Genes       | 260              | 780                 | 523                    | 542                    | 561                   |
| ANI (%)*           | –                | 98.58               | 74.87                  | 74.02                  | 74.01                 |
| Origen isolated    | Tasman Sea (Australia) | Adriatic Sea | Adriatic Sea | Pacific Ocean (Hawaii) | Pacific Ocean (San Diego) | Yellow Sea (South Korea) |
| Depth of Sample (m)| Surface          | 1000                | 1000                   | Surface                | Surface               |

*ANI, Average nucleotide identity (Konstantinidis and Tiedje, 2005) to A. australica H17 homologous genes.

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**Figure 1 Whole-genome alignment of Alteromonas genomes.** The genomes are arranged by decreasing ANI values. Colored rectangles to the left indicate members of the same species. fGIs identified in the comparison of both A. australica have been highlighted in black, identified by the inferred function on the top panel. Red rectangles indicate the location of the strains-specific genomic island. Blue and green vertical lines along the genomes of A. australica show the rRNA and tRNA locations respectively.
considered (Figure 1). This is remarkable since it shows that the location of these genomic features transcends the species and therefore, likely, very long evolutionary times. As found in *A. macleodii*, in *A. australica* there were two types of fGIs, the additive and the replacement types [11,13].

Additive fGIs
fGI1 is of the additive kind and is a typical example of genomic island in which a conserved tRNA act as an insertion target for gene cassettes [18]. Addition of different cassettes makes the length variable among the strains, going from more than 110 Kb in *A. macleodii* to 200 Kb in *A. australica* (Figure 2).

Table 2 Characteristics of genomic islands found in *A. australica* H17$^T$ and DE170 genomes

| Strain  | No.   | Start      | End        | Size (Kb) | No. genes | GC (%) | Inferred character       |
|---------|-------|------------|------------|-----------|-----------|--------|--------------------------|
| Flexible genomic islands                      |        |            |            |           |           |        |                          |
| H17-DE170 fGI-1     | 310522 | 384692     | 310522     | 74        | 46        | 40.74  | Metal resistance/Hydrogenases |
| H17-DE170 fGI-2     | 634953 | 650653     | 634953     | 16        | 25        | 40.82  | Integron                 |
| H17-DE170 fGI-3     | 1127948| 1152459    | 1127948    | 25        | 25        | 37.52  | Flagellum                |
| H17-DE170 fGI-4     | 1783555| 1790698    | 1783555    | 7         | 6         | 36.22  | Exopolysaccharide         |
| H17-DE170 fGI-5     | 3012274| 3028621    | 3012274    | 16        | 16        | 38.40  | O-chain of LPS            |
| H17-DE170 fGI-6     | 3188385| 3217385    | 3188385    | 29        | 29        | 42.80  | CRISPR cluster           |
| H17-DE170 fGI-7     | 3853337| 3872202    | 3853337    | 19        | 16        | 38.35  | Glycosyltransferases      |
| Strain specific genomic islands                |        |            |            |           |           |        |                          |
| H 17 GI-1          | 79701  | 110606     | 79701      | 31        | 19        | 41.84  | DNA phosphorothioation    |
| H 17 GI-2          | 953402 | 975953     | 953402     | 23        | 17        | 43.29  | Type I RM system          |
| H 17 GI-3          | 1298603| 1317990    | 1298603    | 19        | 28        | 38.46  | ND                       |
| H 17 GI-4          | 3133024| 3162117    | 3133024    | 35        | 25        | 44.53  | Carbohydrates metabolism |
| H 17 GI-5          | 3429813| 3464787    | 3429813    | 35        | 26        | 42.53  | PQQ dehydrogenase         |
| DE170 GI-1         | 1444412| 1484731    | 1444412    | 40        | 54        | 46.07  | Mu Phage                 |
| DE170 GI-2         | 2167586| 2188175    | 2167586    | 20        | 12        | 44.30  | Type I RM system          |
| DE170 GI-3         | 2246523| 2285275    | 2246523    | 36        | 39        | 43.82  | Metal resistance          |
| DE170 GI-4         | 3139215| 3155614    | 3139215    | 16        | 12        | 44.6   | Xylan catalysis           |
| DE170 GI-5         | 3713976| 3741976    | 3713976    | 28        | 26        | 46.17  | Cellulose synthase cluster |

Abbreviations: LPS Lipopolysaccharide, CRISPR clustered regularly interspaced short palindromic repeats, RM restriction modification, ND Not determined, PQQ pyrroloquinoline quinone, MGI Mobilizable genomic island.
DE (AltDE) and A. australica DE170 to a single cassette in A. macleodii ATCC 27126T (Additional file 1: Figure S3). Two different kinds of cassettes were found, one is composed of different clusters of czcABC genes, encoding the components of heavy metal efflux pumps important for resistance to cobalt, zinc and cadmium. The other contains a complete hydrogenase cluster. It is remarkable that this hydrogenase cluster previously described in the strain AltDE [14] from the Adriatic Sea, has been also found with a similarity higher than 99% in the Tasman Sea isolate H17T (Additional file 1: Figure S3). However, it is not present in A. australica strain DE170 that comes from the same water sample than AltDE. This cluster is also present in all the members of the A. macleodii deep clade, with the single exception of the strain A. macleodii 615 isolated from the English Channel. The AltDE cluster has been expressed as an active enzyme in both E. coli [26] and in the cyanobacterium S. elongates [27]. A similar (87% nucleotide identity) hydrogenase cluster was found in a Glaciecola sp. 4H-3-7 + YE-5 (an isolate from marine subseafloor sediments Suruga Bay, Japan) plasmid [28].

fGI2 is an integron already described in A. macleodii [11]. Actually this region was proven to be among the most variable in this species. Specifically, the genomes of A. macleodii DE1 and UM7 which belong to the same deep ecotype clonal frame, differ in only 87 single nucleotide polymorphisms (SNPs) over the core genome and were identical even over the FGIs [11] had different gene cassettes in the integron. (Additional file 1: Figure S4). Integrons have widespread distribution among the genomes of marine bacteria (i.e. Vibrionales, Alteromonadales and Pseudomonadales) [29,30]. In A. australica there is a different version, like in all the other genomes of Alteromonas described thus far. Integron identification is typically based on the presence of two parts. The first contains the elements responsible of the integration: an integrase gene (intI) that catalyzes the excision and integration of gene cassettes, a proximal primary recombination site (attI), multiple target-specific recombination sites (attC), and the promoter Pc that drives the transcription of the captured genes cassette [31]. The second part are the gene cassettes that are variable and responsible of the main differences among the strains. The integrase is the only gene conserved in all the fGI3 versions in Alteromonas. The presence of the five structural motifs of integron integrases (boxes I and II and patches I, II, and III) were well conserved (Additional file 1: Figure S5) identifying clearly this as a bona fide integron. We have built a phylogenetic tree (Additional file 1: Figure S6) with the intI genes from all the Alteromonas and the 104 sequences more similar to the intI gene of A. australica found in GenBank. The tree shows that, leaving aside most of the Vibrio species that form a homogeneous group, all the strains belonging to A. macleodii surface ecotype, A. australica and Alteromonas sp. SN2 were grouped together with the Glacieola species intI while A. macleodii deep ecotype clustered with the corresponding gene of Pseudoalteromonas sp. As could be expected, the tree is markedly different from the phylogenomic tree of Figure 2, reflecting the horizontal transfer of these mobile elements. We have identified the attC site of the Alteromonas integron, a small region between 55 and 87 bp. This conserved sequence is located upstream of each gene cassette. The attC is better conserved than the attI [32] and has been well characterized in several species of Vibrio [33]. We have found also this attC sequence (>96% nucleotide identity) in the chromosome and plasmid of Glacieola sp. 4H-3-7 + YE-5. Unexpectedly, we found attC sites in all the Alteromonas genomes at two other regions of the genome (Figure 3). However, the third region is not present in A. australica H17T and strain DE170 only has two genes at this location. All these large arrays of cassettes found in the Alteromonas chromosome could be designated as a ‘super integron’ [34]. The insertion of new cassettes occurs more often next to the integrase than in the other two regions that do not have the integrase (Additional file 1: Figure S4). Considering the three regions, a total of 913 different genes were identified within all the Alteromonas integrons. Among these, almost 60% of them (572) were classified as hypothetical proteins making it impossible to infer any function. A similar proportion has been found in Vibrio species [30]. The second most abundant group of genes (12%) were assigned to toxin/antitoxin (TA) systems (ParDE, RelBE, HigAB and CcdAB) that have been suggested to be involved in the stabilizing of the integron gene cassettes [32] and 9% corresponded to IS elements normally found in the largest integrons. Most of TA systems in all Alteromonas strains are concentrated in the genome in two of the integron regions (Additional file 1: Figure S4). Although the promoter is normally embedded in the integrase sequence, we know from the data of the recent transcriptional analysis performed of the AltDE and AltDE1 genomes (Kimes et al., submitted; see Additional Methods), that all the three integron regions are expressed (Figure 3). Although the expression pattern is similar in all the conditions (Kimes et al., submitted), the expression level was higher under starvation than under nutrient-rich conditions (Figure 3), suggesting that the integron cassettes might help the bacteria to survive during stress conditions [35]. Although the members of the same phylogenomic clade appear to have more similar cassettes, in the region next to the integrase, the cassettes were totally different for each strain showing a highly dynamic nature. It is remarkable that the two A. australica isolates possess three nearly identical cassettes in the integrase region, in spite of their distant geographical origin and its high variability in all the other strains.
Another fGI that is common to all *Alteromonas* species is fGI6 that, interestingly, in the case of *A. australica* DE170, contains the only CRISPR system described in *A. australica*. This system is used by bacteria and archaea as a defense against foreign DNA, either viral or plasmidic [36]. Twenty two spacers were identified by the CRISPR tool (CRISPRFinder; [37]) in the repeat-spacer array preceded by seven cas genes (cas6, cas3, csy2, csy1, cas2-3 and cas1). Based on the classification made by [38], DE170 CRISPR system belong to the subtype I-F, characterized by the fusion of cas2 and cas3 genes. Using the same method we could not find any CRISPR system in the H17T genome. No match was obtained in the Blast searches of the 22 DE170 spacers against public databases. DE170 CRISPR cluster is flanked by IS elements, suggesting that it could be a mobile element as has been seen before in regions of *Escherichia coli* CRISPR gene clusters [39]. This is only the second CRISPR system found in *Alteromonas*. *A. macleodii* DE contains a CRISPR system of a different kind (subtype I-E) (Figure 4) [14] and none of the spacer sequences is shared between the two systems. The presence of different CRISPR subtypes in members of the same genus has also been described in *E. coli* strain collections [39]. Strain H17T contains at the same location a totally different fGI and no trace of the CRISPR system. However, both *A. australica* isolates share a couple of genes coding for the TA system HipAB (Figure 4). It has been described that CRISPR systems and TA genes are often associated in genomic islands that have been named “defense islands” on the grounds of the anti-phage activities that both systems perform [40].

**Strain specific genomic islands**

In addition, there are genomic islands that are found only in one of the strains. Five unique islands were
found in strain H17\textsuperscript{T} and six in strain DE170 (Table 2). H17\textsuperscript{T} GI5 contains a gene cluster encoding a methanol/ethanol pyrroloquinoline quinone (PQQ)-dependent dehydrogenase and the putative \textit{pqqABCDEF} operon that is used as a cofactor and have also shown to be an important antioxidant agent against oxidative damage [41]. This GI was located next to a single Leu-tRNA and at the end of this GI we found a direct repeat identical to the last 52 nucleotides of the tRNA 3′ end, evidence that the tRNA has acted as an insertion target of the GI in the same way as we found in fGI1. Methanol/ethanol dehydrogenase is a key enzyme used for growth in the presence of methanol/ethanol [42]. However, the growth on a one carbon substrate requires a complex metabolic machinery that has not been detected in \textit{Alteromonas} yet. In fact, phenotypic analyses reveal that none of the two strains are able to grow in minimal medium with 1% of methanol (or ethanol) as sole carbon and energy source.

Some of the GIs appear to be associated to post-segregation killing systems. These systems have been mainly described as defense mechanisms against foreign DNA and involved in the stabilization of mobile elements [43]. GI2 in both strains encoded for a different classic type I restriction modification systems. Along the same lines, in the strain H17\textsuperscript{T} GI1 a complete gene cluster for a DNA backbone S-modification (phosphorothioation) was identified [44]. This system seems to act similarly to the classical restriction systems type I but protects DNA using phosphorothioation instead of methylation. The H17\textsuperscript{T} cluster is composed by seven genes, \textit{dndBCDE} that are required for the DNA S-modification and another set (\textit{dptFGH}) involved in the restriction of the unmodified DNA [45]. \textit{A. australica} DE170 genome contains a mobilizable genomic island (MGI) (GI11). MGIs have been recently described as conjugative mobile elements that utilize the conjugation machinery of Integrative Conjugal Elements (ICE) or plasmids for their transfer. This GI could be considering other kind of “additive islands” [46]. In a recent study, five putative MGIs have been characterized in several strains of \textit{A. macleodii} [11] sharing the same insertion point with the one of DE170.

Most of the heavy metal resistance genes in \textit{Alteromonas} are clustered in fGI1. In addition, strain DE170 have another GI containing a large cluster of genes that code for metal resistance related proteins. This 36 Kb region that was flanked by transposases was nearly identical to a region in the plasmid from \textit{Glaciecola} sp. strain 4H-3-7 + YE-5 (pGLAAG01, 341 Kb). Interestingly, this plasmid was showed to be highly syntenic with the plasmid pAMDE1-300 from \textit{A. macleodii} DE1 isolated from the same location as strain DE170 [9]. However, both plasmids have a large variable region with very different genes. Within the variable part in pGLAAG01 was this metal resistant GI found within the chromosome of strain DE170. In the case of pAMADE1-300 the variable region contains a hybrid NRPS-PKS cluster of 65 Kb [12] that was also found within the chromosome of two other \textit{A. macleodii} strains [11]. These results illustrate
the plasticity of these genomic elements that appear to cross easily the genus barrier.

GI4 and GI5 of DE170 appear to be involved in xylan degradation and cellulose biosynthesis. Xylan, a complex polysaccharide with a β-1,4-linked backbone of xylose, is the second most abundant plant cell wall polysaccharide [47]. The xynTAB and xyliREAB clusters that code for the enzymes involved in the degradation and its conversion to D-xylulose 5-P, a pentose phosphate pathway intermediate, were all found in GI4 of DE170. Phenotypic analysis showed that strain D170 was indeed able to grow using xylan as a unique carbon and energy source while strain H17T was not (see Methods). Also in DE170, the complete operon for bacterial cellulose synthesis (bcs) was found in GI10. This operon contains the genes that code for the two catalytic subunits, BcsA and BcsB, that are a transmembrane complex involved in the synthesis of cellulose from glucose and its and secretion. The other components (BscC and BscZ), are accessory proteins related to the regulation and the assembly of the complex [48]. Bacteria generally produce cellulose as a component of the extracellular matrix involved in biofilm formation and cell adhesion [49].

Lysogenic phages

Both A. australica strains contain a genomic region with high similarity to CP4–57, a putative defective prophage described in Escherichia coli [50]. This prophage participates actively in E. coli biofilm development [51]. Like its E. coli counterpart, this putative prophage was found in both H17T and DE170 genomes inserted at one tRNA (Leu) gene that contains a similar att site to the one described for prophage CP4-57 [51,52]. A similar putative prophage was found at the equivalent location in three highly similar isolates of A. macleodii “deep ecotype” belonging to the same clonal frame and obtained from a deep sample (−3500 m) from the Urania Basin in the Ionian (Mediterranean) [11]. Actually, widely divergent versions of this prophage have been found in most Alteromonas genomes available, including the widely divergent SN2 (Additional file 1: Figure S7). In the A. australica strains the versions of this putative prophage were very divergent in sequence. The H17T version was much more similar to the Urania isolates and the DE170 strain version was more similar to the isolate from the Yellow Sea sediment SN2 (Additional file 1: Figure S7).

In addition, in strain DE170, GI6 (47.5 Kb) (Figure 2 and Table 2) has all the hallmarks of a Mu-like lysogenic phage not present in H17T. DNA attachment sites attL and attR could be identified confirming this region as a prophage [53]. Interestingly, a highly related prophage (Figure 5) was previously described in the strain A. macleodii 673 (related only by a 74.1% ANI with DE170) [18]. This strain comes from the Western English Channel, showing the remarkably wide distribution of this phage. Furthermore, 28 of 30 consecutives nucleotides of the spacer 20 found in AltDE CRISPR system matched at 100% identity a hypothetical protein in the Mu-like prophage found in DE170 indicating that a highly related phage is also found in the Mediterranean. The protosparser region in these phages showed a large accumulation of SNPs in the comparison of both prophage genomes (Figure 5) indicating that it is highly variable. Actually, this region includes one ORF annotated as tail fiber protein, known to be involved in the determination of the host range specificity and often highly diversified [54]. Most of the variation in this gene was concentrated in the last (C-terminal) 200 nucleotides (Additional file 1: Figure S8) suggesting that it could be more important for the specificity. The other gene that showed a lower similarity coded for the tape measure protein that is the determinant of phage tail length, what might also indicate host specificity (in this case adapting the phage to different thickness of the host cell envelope). Both the presence of this spacer in the AltDE CRISPR and the variability found in these regions indicate that the prophage is widespread, often lytic and recognize different host strains.

Recruitment from metagenomes

We have compared the recruitment of the genomes in different published marine metagenomes in order to compare the relative abundance of different species [18,55]. These include both central North Pacific and North Atlantic gyres (Hawaii Ocean Time Series-HOTS and Bermuda Atlantic Time Series-BATS) metagenomes [56,57] and the different collections of the Global Ocean Survey (GOS) [58] among others (see Methods). A restrictive cut-off of 98% of identity in 90% of the length of the environmental read was established to perform BLASTN comparisons against the genome of a representative of each species of Alteromonas. Unfortunately with the single exception of the deepest Mediterranean basin Matapan-Varilov [59] none of the Alteromonas genomes recruit significantly from marine metagenomic datasets (Additional file 1: Figure S9). From the Matapan-Varilov sample the A. australica genomes recruit similarly to A. macleodii DE, indicating a possible large particle habitat [14] also for this species. We also compared the genomes with the Dissolved Organic Matter (DOM), Deep Sea Water (DSW) and Dauphin Island Cubitainer Experiment (DICE) metatranscriptomes [6,7,60]. These three microcosms experiments were designed to analyze the transcriptional response of the bacterial community from ocean surface waters to adding dissolved organic matter (DOM and DICE) and deep seawater samples (DSW). Interestingly, all the species representative genomes (one for each since the method...
cannot distinguish differential recruitment within a single species, see Methods) recruit highly in these metatranscriptomes (Additional file 1: Figure S9).

Discussion

The two genomes described here are the first from a species of *Alteromonas* formally described as different from *A. macleodii*. Actually the genome of strain SN2 also belongs to what certainly is a separate species, although it has not been formally described as such and remains as *Alteromonas* sp. We have also the added advantage that the two available genomes come from isolates obtained in very different locations (actually from nearly exact geographic antipodes) and habitats (surface coastal waters versus bathypelagic and off-shore). The first fact to underline is the remarkable similarity between the two genomes that actually recalls the diversity found for *A. macleodii* concurrent isolates. This supports the global distribution of *Alteromonas* species compounded with a high diversity of concurrent clones at every specific location. We posit that *Alteromonas* populations and actually most, if not all, prokaryotes live in complex consortia of multiple clones that have different gene complements, increasing the activities and environmental ranges in which the population can be successful [61,62].

The core genome of all the strains of *Alteromonas* described to date (ca. 2000 genes) is relatively large (see below) although the ANI found among these microbes (Figure 2) indicates that they are widely divergent lineages as expected from strains from different species within the same genus. The core genome could shrink as new strains are sequenced. Particularly the species that live in cold environments such as *A. stellipolaris* isolated from Antarctic sea water [8], might have quite different gene pools reflecting their different habitats. As a reference, in the comparison of 7 species of the genus *Pseudomonas* which genome has a similar genome size to the genus *Alteromonas*, were found ca.1500 genes making up the core [63]. In the same way, 11 species of the genus *Glaciecola* that belong to the same family had 1257 genes [64]. Actually, from the ANI values *A. macleodii* could be in really composed of at least 3 species, the strains defined as the deep ecotype, the yellow sea isolate and the global surface ecotype with ANIs ranging from 73.7 to 80.6%. With this proviso we would be dealing with the equivalent of 5 different species genomes in this calculation of the *Alteromonas* core genome. The flexible part of the genome thus far would add up to ca. 13000 genes, also a typical value if is compared with the marine bacteria of the related genus *Glaciecola* (17276 gene families) [64] or the more than 17000 of the *Vibrio* pan-genome [65].

A remarkable phenomenon that we have discovered is the continuum at the level of the genes found in the flexible genome across the genus. The fact that flexible genomic islands are found at equivalent genomic context, even when considering different species of the same genus, indicates that they are quite stable genomic features. In addition, we found remarkable examples of conservation in the flexible genome. For example, the presence of a nearly identical hydrogenase cluster shared by *A. macleodii* DE (Mediterranean Sea) and H17T (Tasman Sea), or the presence of nearly identical integron cassettes in the two *A. australica* strains. This was expected from the mobile nature of these genetic elements. However, the absolute lack of geographic patterns was more unexpected [66]. Also, the conservation of these elements is nearly perfect, indicating that they have been exchanged recently, in spite of the large geographic distance. Some fGIs that are not mobile such as the flagellum glycosylation have also been found to be perfectly conserved in strains of distant geographic origins [11]. It seems that the core genome diverges more (at the level of SNPs) than the flexible one, probably because the latter is much more homogenized due to its frequent exchange, either by homologous or illegitimate recombination. In addition, flexible
genes jump over the species barriers much more often than the core genome.

The recruitment, mostly from metatranscriptomes indicates that i) the genus *Alteromonas* is one of the most actively blooming microbes in sea water, ii) they have a worldwide distribution from the deep waters of the Mediterranean to the Pacific and Atlantic oceans and iii) they appear in complex consortia of multiple species. It was already known that *A. macleodii* lives in complex consortia of multiple clonal lineages that appear concurrently at any given sample [9]. A similar situation has been found for other marine microbes such as *Prochlorococcus* [62]. It would seem that the same applies to different species within the same genus. This adds to the plankton paradox [67] including even more diversity levels to the populations within the same communities.

**Conclusions**

The two genomes of *A. australica* described here have a similar pattern of variation than the two isolates of *A. macleodii* isolated from the same location in the deep Mediterranean and previously described. Although the genomes are largely syntenic and with a high ANI over the homologous regions, large flexible genomic islands are markedly different in the two strains. This fGIs are of two types, i) the total replacement fGIs, in which the gene cluster in one strain is completely different in gene content from the other strain but with similar assigned function, and ii) the additive fGIs in which gene cassettes are added or deleted depending on the strains but retain some similar regions. It is remarkable that in spite of the large difference in ANI between *A. australica* and *A. macleodii* the fGIs are located at equivalent positions and with similar assigned functions (at least for the replacement type).

Like in the case of *A. macleodii*, metagenomic recruitments indicate that the different strains coexist at the same locations and have probably a global distribution in temperate and tropical latitudes. Both *A. macleodii* and *A. australica* are bloomers that increase their presence in fertilized mesocosms when an input of nutrients is provided.

**Methods**

**Sample collection and sequencing**

Details of isolation and origin of the *Alteromonas* strains used in this study have been described in Additional file 1: Figure S1. DNA was extracted by phenol-chloroform as described in [68] and checked for quality on a 1% agarose gel. The quantity was measured using Quant-iT® PicoGreen® dsDNA Reagent (Invitrogen). The genomes were sequenced using the IlluminaHiSeq 2000 (100-bp paired-end read) sequencing platform (Macrogen, Korea). The generated reads were trimmed and assembled de novo with VELVET, version 0.7.63 [69] using default parameters except for the k-mer that was 49. Combination of Geneious Pro 5.0.1 (with default parameters) using previously *Alteromonas* assembled genomes as a reference [9,11,18] and oligonucleotides designed from the sequence of the ends of assembled contigs were used in order to obtain one single closed contig.

**Genome annotation and analysis**

The genomes were annotated using the NCBI PGAAP annotation pipeline (http://www.ncbi.nlm.nih.gov/genome/annotation_prok/). The predicted protein sequences were compared using BLASTP to the NCBI nr protein database (e-value $10^{-5}$). ORFs smaller than 100 bp and without significant homology to other proteins were not considered. BioEdit was used to manipulate the sequences [70]. GC content was calculated using the EMBOSS tool geece [71]. For comparative analyses, reciprocal BLASTN and TBLASTXs searches between the genomes were carried out, leading to the identification of regions of similarity, insertions and rearrangements. To allow the interactive visualization of genomic fragment comparisons Artemis v.12 [72], Artemis Comparison Tool ACTv.9 [73] were used to compare the genomes. The ANI between strains was calculated using JSpecies software package v1.2.1 using default parameters [74]. Sequences were aligned using MUSCLE version 3.6 [75] and ClustalW [76] and edited manually as necessary. Nucmer program in the MUMmer3+ package [77] was used to identify the indels and the SNPs between small regions of the genomes. Pan Genome Analysis Pipeline (PGAP) [78] was used to identify all of the orthologous pairs genes between *Alteromonas* genomes. The common dataset of shared genes among the strains was defined as their core genome. The total set of genes within the genomes was defined as the pan genome. The set of genes in each strain not shared with other strains was defined as unique genes.

**Phylogenetic analysis**

To determine the exact phylogenetic relationship of the new isolates within the genus, phylogenetic analysis for all the *Alteromonas* members whose genomes were available were carried out (Additional file 1: Table S1). The tree was rooted using *Pseudoalteromonas atlantica* T6c (NC_008228.1) as an outgroup. The complete genomes were analyzed using TIGRams to identify and concatenated all the conserved proteins. The concatenated proteins were aligned using Kalign [79] and a maximum likelihood tree was made using FastTree [80] using a JTT + CAT model and a gamma approximation.

**Recombination**

The first step to analyze the recombination events among the *Alteromonas* strains was to make a multiple genome
alignment. This was performed by using Mauve multiple alignment software (v2.3.1) [81]. ClonalFrame software v1.2 [20], a Bayesian inference method that reconstructs clonal relationships between the isolates in a sample, was used for estimating mutation and recombination rates. Three independent runs of ClonalFrame were performed each consisting of 100000 Markov chain Monte Carlo iterations. To assess the relative contribution of recombination and mutation, r/m and ρ/θ statistics were used.

Recruits of environmental collections
Genomes recruits were carried out against some available marine metagenomes and metatranscriptomes [BATs (Bermuda Atlantic Time Series) [56], DICE (Dauphin Island Cubitainer Experiment) [60], DOM (Dissolved Organic Matter) [7], DSW (Deep Sea Water) [6], GOS (Global Ocean Survey) [58], HOTS (Hawaii Ocean Time Series) [57,82], Marmara [83], MedDCM (Mediterranean Deep Chlorophyll Maximum) [84] and MVP (Matapan-Vavilov Deep) [59]]. BLASTN was carried out between the genome of a representative member of each Alteromonas species (A. australica DE170; A. macleodii ATCC27126\(^T\); A. macleodii DE; Alteromonas sp. ALT199 and Alteromonas sp. SN2) and the environmental databases. A restrictive cut-off of 98% of identity in 90% of the length of the environmental read was established to guarantee that only similarities at the species level were counted. The number of hits was normalized against the genomes and the database sizes.

Phenotypic analyses
Growth assays in media with xylan as a sole a carbon and energy source were carried out in order to study if the presences of this gene cluster provided extra abilities in strain DE170. A 50 ml culture of both A. australica strains were grown in marine broth at 20°C until they reached to late exponential growth phase (Optical density 600 nm = 1.0). The cultures were washed twice using minimal medium without any carbon sources and then grown in minimal medium agar plates supplemented with 0.5, 1, 5 and 10 g l\(^{-1}\) of xylan from beechwood (Sigma; \(\times4252\)) at 20°C. The same washed cultures were used to analyze the Methanol/Ethanol dehydrogenase activity. To measure growth rate, cultures were grown in minimal medium with 1% ethanol or methanol as a sole carbon source in Erlenmeyer flasks with orbital shaking (200 rpm) and incubated at 20°C. Growth was measured by the optical density at 600 nm.

Accession numbers
The genome sequences have been deposited in GenBank under the following bioproject accession numbers; PRJNA246143 for A. australica H17\(^T\) and PRJNA246140 for A. australica DE170.

Additional file

**Additional file 1: Figure S1.** A. macleodii isolates used in this study and their origin. **Figure S2.** Maximum-likelihood phylogenetic trees of randomly selected collinear blocks of sequence (size indicated under each tree). Members of different species were indicated with different color letters. In a black rectangle the core tree (as in Figure 2) is shown as a reference. A. australica DE170 genome was used to locate the position of the sequences used to perform the trees. **Figure S3.** Alignment of the fGI in some species of Alteromonas. Red arrows under the genomes indicate the 3’ end of the tRNA gene section that is duplicated. The plots above the genomes indicated the number of SNPs in a 500-bp window in comparison with the genome located on the top. The average and the total number of SNPs in the genome are indicated by a red dot line. **Figure S4.** Comparison of the DE1 and UMP integron cluster. The integron integrate is marked with a red rectangle. Gene expression data (expressed as RPMI) is shown mapped to the A. macleodii DE1. **Figure S5.** Alignment of C-terminal integrates Alteromonas sequences. **Figure S6.** Phylogenetic tree of the Alteromonas integron integrates and some reference sequences similar to the intI gene of A. australica found in GenBank. Color code indicates members of the different species of Alteromonas. **Figure S7.** Putative Propophase CP4-57-like found in both A. australica and comparison to the similar propophages found in other Alteromonas. **Figure S8.** Alignment of the tail fiber protein from Mu-like prophages inserted in the A. australica DE170 and A. macleodii 673 genomes. **Figure S9.** Alteromonas species genomes relative recruitment of metagenomic reads at 98% identity and 90% coverage from some marine reference metagenomes. Table S1. Features of the reference genomes.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
FRV conceived the study, and participated in its design and coordination. Assembly and genome analysis was carried out by MJP and AG. EI isolated the strains H17\(^T\) and contributed to the genome analysis. FRV and MLP wrote the manuscript. All authors read and approved the final manuscript.

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