Therapeutic potential of active components of saffron in post-surgical adhesion band formation

Article (Published Version)

Arjmand, Mohammad-Hassan, Hashemzehi, Milad, Soleimani, Atena, Asgharzadeh, Fereshteh, Avan, Amir, Mehraban, Saeedeh, Fakhraei, Maryam, Ferns, Gordon A, Ryzhikov, Mikhail, Gharib, Masoumeh, Salari, Roshanak, Hoseinian, Sayyed Hadi Sayyed, Parizadeh, Mohammad Reza, Khazaee, Majid and Hassanian, Seyed Mahdi (2021) Therapeutic potential of active components of saffron in post-surgical adhesion band formation. Journal of Traditional and Complementary Medicine, 11 (4). pp. 328-335. ISSN 2225-4110

This version is available from Sussex Research Online: http://sro.sussex.ac.uk/id/eprint/96738/

This document is made available in accordance with publisher policies and may differ from the published version or from the version of record. If you wish to cite this item you are advised to consult the publisher’s version. Please see the URL above for details on accessing the published version.

Copyright and reuse:
Sussex Research Online is a digital repository of the research output of the University.

Copyright and all moral rights to the version of the paper presented here belong to the individual author(s) and/or other copyright owners. To the extent reasonable and practicable, the material made available in SRO has been checked for eligibility before being made available.

Copies of full text items generally can be reproduced, displayed or performed and given to third parties in any format or medium for personal research or study, educational, or not-for-profit purposes without prior permission or charge, provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way.
Original Article

Therapeutic potential of active components of saffron in post-surgical adhesion band formation

Mohammad-Hassan Arjmand a,1, Milad Hashemzehi b,1, Atena Soleimani c,1, Fereshteh Asgharzadeh d,1, Amir Avan e,1, Saeedeh Mehraban g, Maryam Fakhraei d, Gordon A. Ferns h, Mikhail Ryzhikov i, Masoumeh Gharibi j, Roshanak Salari k, Sayyed Hadi Sayyed Hoseinian l, Mohammad Reza Parizadeh c, Majid Khazaee d, e, **, Seyed Mahdi Hassanian c, e, *

a Medical Plants Research Center, Basic Health Sciences Institute, Shahrekord University of Medical Sciences, Shahrekord, Iran
b Transahur University of Medical Sciences, Transahur, Iran
c Department of Clinical Biochemistry, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran
d Department of Physiology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran
e Metabolic Syndrome Research Center, Mashhad University of Medical Sciences, Mashhad, Iran
f Student Research Committee and Department of Medical Genetics, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

Purpose: The aim of this study was to investigate the therapeutic potential of the pharmacological active component of saffron in attenuating the formation of post-operative adhesion bands using different administration methods in a murine model.

Material method: saffron extract (100 mg/kg), Crocin (100 mg/kg), and Crocetin (100 mg/kg) were administered intraperitoneally and by gavage in various groups of male Wistar rat post-surgery. Also three groups were first treated intra-peritoneally by saffron extract, Crocin, and Crocetin (100 mg/kg) for 10 days and then had surgery. At the end of the experiments, animals sacrificed for biological assessment.

Result: A hydro-alcoholic extract of saffron and crocin but not crocetin potently reduced the adhesion band frequency in treatment and pre-treatment groups in the mice given intra-peritoneal (i.p) injections. Following the saffron or crocin administration, histological evaluation and quantitative analysis represented less inflammatory cell infiltration and less collagen composition, compared to control group. Moreover, the oxidative stress was significantly reduced in treatment groups.

Abbreviations: APC, activated protein C; DSS, dextran sodium sulfate; HE, Hematoxylin & Eosin; IP, intera-peritoneal; MDA, malondialdehyde; PSAB, post-surgical adhesion band; PDGF, platelet-derived growth factor; TGF-β, transforming growth factor-beta; α-SMA, α-smooth muscle actin; SOD, superoxide dismutase; TAA, thioacetamide.

* ** Corresponding author. Department of Medical Physiology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran.
E-mail addresses: khazaeeim@mums.ac.ir [M. Khazaee], hasanianmehrm@mums.ac.ir [S.M. Hassanian].
1 Made equal contribution to this study. This study was supported by grants awarded by the National Institute for Medical Research Development (Grant No.977089).

https://doi.org/10.1016/j.jtcme.2021.01.002

Please cite this article as: M.-H. Arjmand, M. Hashemzehi, A. Soleimani et al., Therapeutic potential of active components of saffron in post-surgical adhesion band formation, Journal of Traditional and Complementary Medicine, https://doi.org/10.1016/j.jtcme.2021.01.002
1. Introduction

Intraperitoneal adhesion formation is a serious worldwide complication of surgery developing between abdominal wall and intra-abdominal surfaces following peritoneal irritation. This condition is highly prevalent (up to 90%) in patients undergoing open gynecological pelvic or open abdominal surgery.1,2 Intestinal obstruction,3 female infertility,4 chronic abdominal and pelvic pain5 are the most important consequences of post-operative adhesion bands. Placement of some solid barriers such as Sephradi film® is a standard strategy for adhesion bands prevention which needs to accurately recognize the damage sites.6 Predicting where these fibrotic bands may develop is a barrier usage limitation encouraging the researchers to find an efficient therapeutic method.7 Therefore, identifying the exact mechanisms involved in adhesion band formation would help to progress the treatment process.

It is generally accepted that pathological adhesions generation is the end-result of improper healing process in peritoneal cavity.8 In post-operative physiological responses, immune cell–released inflammatory cytokines, as well as platelet-derived growth factor (PDGF), and fibroblast-induced transforming growth factor-beta (TGF-β) are increased in peritoneum.9,10 It has been shown that inflammation and fibrosis play key roles in the pathogenesis of adhesion band formation.11–13 We previously investigated the effects of anti-fibrotic or anti-inflammatory compounds, EW-719714 and activated protein C (APC)15 on post-surgical adhesions, representing the hopeful results in reduction of these fibrotic bands. Although there are extensive studies on post-surgical adhesion band formation, our knowledge of pathogenic mechanism of this surgery-associated complication is still limited.

The plant-based therapies to improve pathologic conditions have been used for centuries and counted as potential treatments due to low complications and minimal toxicities. The huge parts of FDA-approved components are contained the therapeutics agents derived from natural compounds.16,17 The natural products continue to play an important role in drug discovery programs for different diseases. Using different technical approaches,18,19 studies showed the effectiveness of these components for various disorders such as carcinomas,20–26 diabetes,27 various inflammatory and fibrotic conditions.28–32 One of these natural products was isolated from Crocus sativus, a flowering plant in the Iridaceae family and is commonly known as saffron.33

Saffron, the dried stigmas of Crocus sativus, is cultivated in Iran and other countries such as Spain and Turkey34 and has a wide range of activities including oxytocic, anti-carcinogenic,35,36 and etc. There are several active pharmacological components in saffron including crocin and crocetin that are responsible for the biological effects of saffron.37,38 Saffron contains essential mineral and various important vitamins. Generally, due to anti-aging and anti-oxidant function of saffron, its daily consumption is very high in many parts of the world. Also, the ability of saffron, crocin or crocetin to reducing adverse effects of chemotherapeutic components showed their drug modulator activities.39 Animal studies showed low or non-toxicity of saffron and its extract.40 A temporary immunomodulatory activity with no renal, hepatic, hematological, or any side effects were reported in sub-chronic use of 100 mg/day saffron in a randomized controlled trial study.41 Studies revealed that these compounds elicit anti-inflammatory and anti-fibrotic responses by decreasing the inflammatory mediators in different animal models42–44 which is consistent with our previous work, showing anti-inflammatory effects of crocin in mice model.45 In line with this, saffron potently decreased the level of Malondialdehyde (MDA) and inflammatory cytokines in bleomycin-induced pulmonary fibrosis.46 Similarly, crocin significantly reduced the over-expression of fibrotic molecules such as collagen 1a, α-smooth muscle actin (α-SMA), and TGF-β in hepatic fibrosis condition.47

In this study, we investigated the protective effects of saffron, crocin, or crocetin in preventing post-surgical adhesion band formation in animal models. Our results showed that saffron or crocin (i.p.) could suppress the formation of adhesion bands through anti-inflammatory, anti-oxidant and anti-fibrosis function in rat model, suggesting the therapeutic potential of this non-toxic compounds against post-surgery-associated complications.

2. Material and methods

2.1. Animal

Animal experiments were conducted in according to the guideline for Care and Use of Laboratory Animals from Mashhad University of Medical Sciences (MUMS). Animals were purchased from pasture institute (Tehran, Iran) and were subjected to controlled conditions of temperature (20 ± 2 °C) and humidity (50%). The animals were housed in a standard cage in a 12 h light-dark cycle room with free access to both food and water. This study was approved by ethical committee of Mashhad University of Medical Sciences, ethical approval number: 960917.

2.2. Reagents

Pure crocin and crocetin were purchased from BuAli Pharmacological Research Center (MUMS, Mashhad, Iran). Hydro-alcoholic extract of Saffron was isolated by Maceration method as described.48,49 Reagents need for hematoxylin and eosin, malonyl dialdehyde (MDA), total thiol, and catalase were purchased from Sigma Co (Saint Louis, MO).

2.3. Surgical abrasion model

Surgical method for adhesion band formation was performed as described.50,51 Briefly, the animals were anesthetized by intraperitoneal injection of ketamine.52 After shaving and preparation of the site of surgery with 1% antiseptic povilone-iodine solution, the abdominal cavity was opened with 1.5 cm incision. The anterior cecal surface was gently rubbed until partial petechial hemorrhages were generated and the interior surface of abdomen was damaged using a medical electric scalpel. The cecum placed in front of
damaged surface of abdomen and the muscular layer was sutured. Intraperitoneal adhesion formation occurs with this procedure probability above 90% between damaged surfaces. The evaluation of post-surgical adhesion bands was performed according to the previously explained adhesions grade criteria by Nair et al. in a blinded fashion, assessing the incidence and frequency of fibrotic bands. The detailed of Nair scoring system is shown in Table 1.

2.4. Experimental design

Rats were randomly divided into the following groups of 6 rats in each group: 1) Control group (animals with surgical abrasion received saline as vehicle intraperitoneally (i.p)); 2) animals with surgical abrasion treated intraperitoneally with 100 mg/kg/day saffron extract; 3) animals with surgical abrasion treated intraperitoneally with 100 mg/kg/day crocin; 4) animals with surgical abrasion treated intraperitoneally with 100 mg/kg/day crocetin.

To investigate whether type of administration, i. p or gavage could affect protective effects of these compounds, we also add the following groups: 5) animals with surgical abrasion treated by oral gavage with 100 mg/kg/day saffron extract; 6) animals with surgical abrasion treated by oral gavage with 100 mg/kg/day crocin; 7) animals with surgical abrasion treated by oral gavage with 100 mg/kg/day crocetin.

Next, we were interested in evaluating the preventive potential of these compounds in post-surgical adhesion band formation. In this case, before the surgery, we consider more groups: 8) animals were first treated intra-peritoneally with 100 mg/kg/day saffron extract for 10 days and then had surgery; 9) animals were first treated intra-peritoneally with 100 mg/kg/day crocin for 10 days and then had surgery; 10) animals were first treated intra-peritoneally with 100 mg/kg/day crocetin for 10 days and then had surgery.

All animals weighted before surgical abrasion and during the experiment. At the end of the experiments, animals sacrificed for biological assessment. The excised tissue samples were quickly frozen in liquid nitrogen or fixed in 10% formalin solution.

2.5. Histological analysis

Tissue sections were fixed in formalin 10% for 24–72 h. After processing and paraffin embedding, staining was done with either Hematoxylin & Eosin (HE) or Masson’s Trichrome for assessing inflammation and collagen deposition, respectively. Quantifying the tissue staining presented histological grading of inflammation and fibrosis scores. The slides were seen under light microscope (magnification ×400).  

2.6. Preparation tissue homogenates

Adhesion bands-linked cecum was collected for making tissue homogenate. The tissue sample (100 mg tissue sample for each animal) was weighted and homogenized with PBS as the homogenization medium. The supernatant of the tissue homogenate was used for the assay of stress oxidative markers.

2.7. Measurement of stress oxidative markers

The oxidative stress was evaluated in each subject by measuring the concentration levels of malondialdehyde (MDA) and thiol, as well as catalase activity in the tissue homogenates. All procedures were conducted according to the kit protocol. Quantitative variables were described as mean ± SEM. One-way ANOVA test was used for comparison between different groups. The collected data were imported into GraphPad Prism for statistical analysis. The statistical significance was set to 5%.

3. Results

3.1. Saffron and crocin significantly reduced adhesion scores

We investigated the therapeutic effect of hydro-alcoholic extracted of saffron, crocin, and crocetin in different administration in adhesion rat model. No animal death occurred during the experiment, and the body weight did not significantly differ after second surgery. There was no infection or bleeding in post-operative time. Schematic representation of the study protocol is shown in Fig. 1A. According to clinical observations, pre-treatment with crocin is the treatment of choice, offering more recovery and reducing the adhesion formation probability (Fig. 1B). We quantitatively analyzed the incidence of adhesion band in different groups, using Nair54 scoring scheme (Fig. 1C). Compared to the rat control group, pre-treatment or treatment (i.p) with saffron or crocin significantly improved the frequency of post-operative intra-abdominal adhesion bands (P value < 0.05). Moreover, decrease in adhesion bands frequency (Nair grade) was not significant in oral administration groups. No reduction of adhesion bands were found in the case of crocetin treatment (Fig. 1B and C), compared to control cases. We did not continue experiments on crocetin due to adverse results.

3.2. Saffron extract and crocin decreased inflammation in post-surgical adhesion bands

Since inflammatory responses are increased during adhesion bands formation, we evaluated the inhibitory effects of saffron and its pharmacological active components on inflammation in adhesion bands tissues, using Hematoxylin & eosin (HE) staining. Histological analysis and quantitative evaluation showed a significant reduction of inflammation in treatment as well as pre-treatment groups (Fig. 2A–C). As expected, oral treatment with hydro-alcoholic extract of Saffron showed no significant differences, compared to control tissues (Fig. 2C). The detailed of inflammation scoring system is shown in Table 2.

Table 1

| Adhesion score system for macroscopic evaluation (Nair’s et al.) | Adhesion grade |
|---|---|
| 0 to 4 | Complete absence of adhesion |
| 1 | Single band of adhesion, between viscera or from viscera to abdominal wall |
| 2 | Two bands, either or from viscera to abdominal wall |
| 3 | More than two bands, between viscera or viscera to abdominal wall |
| 4 | Viscera directly adherent to abdominal wall, irrespective of number and extent of adhesive bands |
3.3. Saffron and crocin decreased inflammatory responses via reducing oxidative stress markers in post-surgical adhesion band

To further determine the anti-inflammatory mechanisms of Crocin and Saffron in adhesion bands, we measured concentration of oxidant marker, Malon deladehyde (MDA), as well as antioxidants agents including total Thiol concentration and Catalase activity in adhesive tissue homogenates. Our findings showed significant reduction of MDA concentration in all treatment group, compared to control group (Fig. 3A). Consistently, the level of total Thiol (Fig. 3B) and catalase activity (Fig. 3C) were higher in treated groups than control tissues. These results supported the hypothesis that saffron and crocin elicited their protective functions at least partially by attenuating oxidative stress reactions in post-operative adhesion rat model.

3.4. Inhibitory effects of saffron and crocin on tissue fibrosis in adhesion rat model

Next, we investigated the regulatory effects of saffron and crocin on fibrosis as key factors in the pathogenesis of adhesion band formation. Masson’s trichrome staining accompanied with quantitative analysis demonstrated that crocin or hydro-alcoholic extracted of Saffron could significantly decrease the areas of fibrosis and collagen deposition in adhesive tissues (Fig. 4A–C). The therapeutic effect was more potent in crocin administration group. The fibroblast activity quantified according to the scoring system presented in Table 2. Similar to previous results, oral treatment of Saffron elicited no protective effect in adhesion tissues (Fig. 4B) (P value < 0.05).

4. Discussion

The present study investigated the protective effects of hydro-alcoholic extract of saffron, crocin and crocetin on intra-abdominal adhesion models, using different administration methods. Macroscopic results demonstrated a significant reduction of adhesion bands frequency in intraperitoneal administration of saffron or crocin as well as the pre-treatment group. However, neither crocetin, nor oral administration of saffron could prevent post-surgical adhesion band formation in rat model. We also showed that decrease in the oxidative stress responses as well as attenuating fibrosis and collagen depositions are some of the mechanisms by which saffron and crocin (i.p) exert their protective responses in adhesion model, supporting the therapeutic potential of these low-toxic compounds against post-surgical adhesion band formation.

Damage to peritoneum leading to the deposition of peritoneal fibrin and local inflammation are the key steps of adhesiogenesis. Studies have shown that using fibrinolytic compounds prevent the progress of adhesion formation via stimulation of the intraperitoneal fibrinolytic system. Moreover, it has been found that adhesion formation is accompanied by lower tissue oxygenation and free oxygen radical generation. Ezberci et al. showed that decrease in oxidant agent including MDA whereas increase in catalase activity attenuated adhesion score in Bacterial peritonitis rat model. Consistent with these results, our previous finding revealed that decrease in oxidative stress markers, as well as collagen deposition and infiltration of inflammatory cells to injured site attenuated the severity of adhesions in animal model. Also, we showed that human APC significantly reduced the formation of adhesion bands which is correlated with lower concentration of pro-inflammatory cytokines and higher tissues plasminogen activator (tPA) in vivo.

Furthermore, there are several studies supporting the anti-oxidative, anti-inflammatory, and anti-fibrotic properties of saffron and crocin in different models. For instance, Hemshekhar et al. showed that crocin enhanced anti-oxidant status by
increasing catalase, and superoxide dismutase (SOD) activities as well as attenuating serum level of inflammatory factors in arteritis rat model. Moreover, Samarghandian et al. reported that crocin improved aged rat kidney functions by reducing oxidative stress and inflammatory responses in rat. Consistently, Hashemi et al. indicated the effects of saffron carotenoids, crocin and crocetin on oxidative stress in breast tumor. They showed that crocin and crocetin increase the catalase and superoxide dismutase activities in BALB/c mice, after 28 days of treatment. Since crocin treatment has been shown antioxidant activities under various conditions, Nasimian et al. showed that crocin elevated the apoptotic death of human breast cancer cell lines, partially via ROS-activated FOXO3a axis. Moreover, clinical trial studies evaluated the effect of saffron aqueous extract, crocin and crocetin in coronary artery disease (CAD). It has been shown that crocin and crocetin treatment resulted in a significant reduction in lectin-like oxidized LDL receptor 1 (LOX1), nuclear factor kappa-B (NF-κB) and Serum ox-LDL. Also, the levels of monocyte chemoattractant protein 1 (MCP-1) was reduced in all treatment groups in CAD patients. Similarly, we recently showed that crocin significantly inhibited oxidative stress and histopathological scores, representing a reduction of inflammation and fibrosis in dextran sodium sulfate (DSS)-induced colitis model. Another study showed that 100 mg/kg of saffron reduced MDA, Myeloperoxidase, and tumor necrosis factor-alpha (TNF-α) in pulmonary fibrosis.

Consistent with the fibrinolysis effect of crocin in post-surgical adhesion model, it has been shown that saffron could significantly prevent the thickness of alveolar septa and collagen deposition in bleomycin-induced pulmonary fibrosis. Chhimwal et al. showed that crocin decreased the hepatic fibrosis via expression of peroxisome proliferator-activated receptor γ (PPAR-γ), modulating the inflammatory and fibrogenic pathways. These results are consistent with another study showing the anti-fibrotic and anti-inflammatory properties of crocin in thioacetamide (TAA)-induced liver fibrosis.

In conclusion, this study suggested that saffron and crocin have potential therapeutic value in preventing intra-abdominal adhesion band formation. The mechanism underlying abdominal adhesions has not yet been completely understood. Further animal and clinical studies are required to clarify this issue and to assess the exact mechanism of action for Saffron and its pharmacologically active component, crocin, for preventing adhesion bands formation.

Table 2
Infiltration of inflammatory cells and fibrosis scoring according to Swolin.

| Grade | Inflammatory cell infiltrate | Fibrosis |
|-------|-----------------------------|----------|
| 0     | Absent or normal in number  | None     |
| 1     | Slight increase             | Slight   |
| 2     | Moderate infiltration       | Moderate |
| 3     | Dense                       | Dense    |

Fig. 2. The inhibitory effects of Saffron and Crocin on inflammation in adhesive tissues. (A, C) Intraperitoneal injection of Crocin (100 mg/kg/day) or Saffron (100 mg/kg/day) reduced inflammatory cell infiltration (black arrows) in (A) treatment and (C) pre-treatment groups. (B) Despite crocin, oral administration of Saffron showed no significant protective responses in adhesion rat model. ***p < 0.001.
Fig. 3. Oxidative stress is attenuated following Saffron- or Crocin-treatment in adhesive tissues. (A–C) The tissue concentration of (A) MDA, (B) total Thiol, and (C) the activity of Catalase were compared between different groups. *p < 0.05; **p < 0.01; ***p < 0.001.

Fig. 4. Saffron and Crocin suppressed fibrosis and Collagenesis in adhesive tissues. (A–C) Masson’s trichrome staining showed a significant reduction of deposition of Collagen (asterix) in Crocin- or Saffron-treated rat in different groups (B) The decrease in collagen thickening was not significant in the case of oral administration of Saffron, compared to control. ***p < 0.001.

Data availability statement

Research Data are not shared.

Declaration of competing interest

The authors have no conflicts of interest.
References

1. Menzies D, Ellis H. Intestinal obstruction from adhesions–how big is the problem? Am J Surg. 1990;160:72–75.
2. Okabayashi K, Ashrafian H, Zacharakis E, et al. Adhesions after abdominal surgery: a systematic review of the incidence, distribution and severity. Surg Today. 2014;44(3):405–420.
3. Menzies D. Peritoneal adhesions. Incidence, cause, and prevention. Surg Ann. 1992;24:27.
4. Kavic SM, Kavic SM. Adhesions and adhesiolysis: the role of laparoscopy. J Soc Laparoscop Surg. 2007;2(2):99.
5. Sulaiman H, Gabella G, Davis C, et al. Growth of nerve fibers into murine peritoneal adhesions. J Pathol. 2000;192(3):396–403.
6. Cai X, Hu S, Yu B, et al. Transglutaminase-catalyzed preparation of crosslinked carboxymethyl chitosan/carboxymethyl cellulose/collagen composite membrane for post surgical peritoneal adhesion prevention. Carbohydr Polym. 2018;201:201–210.
7. diZerega Gs, Campeau JD. Peritoneal repair and post-surgical adhesion formation. Hum Reprod Update. 2001;7(6):547–555.
8. Surgeons PG. ColorectalSurgerySurgical. Pathogenesis, consequences, and control of peritoneal adhesions in gynecologic surgery: a committee opinion. Fertil Steril. 2013;99(6):1550–1555.
9. Saed GM, Zhang W, Diamond MP. Molecular characterization of fibroblasts isolated from human peritoneum and adhesions. Fertil Steril. 2001;75(4):763–768.
10. Saed GM, Diamond MP. Modulation of the expression of tissue plasminogen activator and its inhibitor by hypoxia in human peritoneal and adhesion fibroblasts. Fertil Steril. 2007;87(1):164–171.
11. Rout UK, Saed GM, Diamond MP. Transforming growth factor-β1 modulates expression of adhesion and cytoskeletal proteins in human peritoneal fibroblasts. Fertil Steril. 2001;75(5):154–161.
12. Chegini N. TGFB system: the principal profibrotic mediator of peritoneal adhesion formation. Paper presented at: Seminars in reproductive medicine 2008.
13. White JC, Jiang ZL, Diamond MP, Saed GM. Macrophages induce the adhesion phenotype in normal peritoneal fibroblasts. Fertil Steril. 2011;96(3):758–763, e753.
14. Soleimani A, Agharzadeh F, Rahmani F, et al. Novel oral transforming growth factor-β1 signaling inhibitor potently inhibits postsurgical adhesion band formation. J Cell Physiol.2000.
15. Dinavard P, Hassannia SM, Weiler H, Rezaee AR. Intrapерitoneal administration of activated protein C prevents postsurgical adhesion formation. Blood. 2015;125(8):1339–1348.
16. Baig B, Hilla-Alnaqbi A, Amin A. Cancer and biotechnology: a match-up that should never slowdown. In: Biotechnology and Prevention of Anti-cancer Compounds. Springer. 2017;73–97.
17. Al-Dabbagh B, Elhayy IA, Al Hrout A, et al. Antioxidant and anticancer activities of Trigonella foenum-graecum, Cassia acutifolia and Rhazya stricta. BMC Compl Alternative Med. 2018;18(1):240.
18. Hanna EM, Zaki N, Amin A. Detecting protein complexes in protein interaction networks modeled as gene expression biclusters. PloS One. 2015;10(12), e0144163.
19. Mahmood-Ghoneim D, Amin A, Corr P. MRI-based texture analysis: a potential technique to assess protectors against induced-liver fibrosis. J Cell Biochem. 2009;108(1):203–212.
20. Kamal H, Jafar S, Mudgil P, Murali C, Amin A, Maqsood S. Inhibitory properties of Melissa officinalis against rat peritoneal adhesions in rats. Fertil Steril. 2001;75(4):763–768.
21. Al-Dabbagh B, Chaiboonchoe A, Khraiwesh B, et al. Safranal induces DNA double-strand breakage and ER-stress-mediated cell death in hepatocellular carcinoma cells. J Cell Biochem. 2018;118(5):1311–1319.
22. Vahedi M, Govil S, Kumar S, Shrivastava D, Karimi R, S Bisen P. Therapeutic applications of Crocus sativus L. (saffron): a review. Nat Prod J. 2016;6(3):161–171.
23. Aghaee Z, Jafari SM, Dehnad D. Effect of different drying methods on the physicochemical properties and bioactive components of saffron powder. Plant Foods Hum Nutr. 2019;74(2):171–178.
24. Riaziabakh S, Ghazavi A. Immunomodulatory effects of saffron: a randomized double-blind placebo-controlled clinical trial. Phytotherapy research. PTR. 2011;25(2):1801–1805.
25. Nam KN, Park Y-J, Jung H-J, et al. Anti-inflammatory effects of crocin and crocetin in rat brain microglial cells. Eur J Pharmacol. 2010;648(1–3):110–116.
26. Yarjani ZM, Pourmotabbed A, Pourmotabbed T, Najafi H. Crocin has anti-inflammatory and protective effects in ischemia-reperfusion induced renal injuries. Iran J Basic Med Sci. 2017;20(7):753.
27. Hosseinzadeh H, Younesi HM. Anticancer and anti-inflammatory effects of Crocus sativus L. stigma and petal extracts in mice. BMC Pharmacol. 2002;2(1):7.
28. Rezaei N, Avan A, Pashazhad M, et al. Crocin as a novel therapeutic agent against colitis. Drug Chem Toxicol. 2019;1–8.
29. Bahnoue M, Fatiemka H, Pourkhahli K, et al. Saffron protection against bleomycin-induced pulmonary fibrosis in rats. Iranian J Toxicol. 2017;11(6):73–83.
30. Algardamy MM. Antiinflammatory effects of crocin on thioacetamide-induced liver fibrosis in mice. Saudi J Biol Sci. 2018;25(4):747–754.
31. Makhlov H, Saksouk M, Habib J, Chahine R. Determination of antioxidant activity of saffron taken from the flower of Crocus sativus grown in Lebanon. Afr J Biotechnol. 2011;10(4):554–558.
32. Ghaedroost B, Vafaei AA, Rashidy-Pour A, et al. Protective effects of saffron extract and its active constituent crocin against oxidative stress and spatial learning and memory deficits induced by chronic stress in rats. Eur J Pharmacol. 2015;772(1–3):222–229.
33. Hemadeh O, Chilukuri S, Bonet V, Hussein S, Chaudry IH. Prevention of peritoneal adhesions by administration of sodium carboxymethyl cellulose and oral vitamin E. Surgery. 1993;114(5):907–910.
34. Wei G, Chen X, Wang G, et al. Inhibition of cyclooxygenase-2 prevents intra-abdominal adhesions by decreasing activity of peritoneal fibroblasts. Drug Des Dev Ther. 2015;9:3083.
35. Green CJ, Knight J, Piquette-Simon S. Ketamine alone and combined with diazepam or xylazine in laboratory animals: a 10 year experience. Lab Anim. 1981;15(2):163–170.
36. Hemadeh O, Chilukuri S, Bonet V, Hussein S, Chaudry IH. Prevention of peritoneal adhesions by administration of sodium carboxymethyl cellulose and oral vitamin E. Surgery. 1993;114(5):907–910.
37. Nair SK, Bhat IK, Aurora AL. Role of proteolytic enzyme in the prevention of postoperative intraperitoneal adhesions. Arch Surg. 1974;108(6):849–853.
38. Bostan HB, Mehriz S, Hosseinzadeh H. Toxicology effects of saffron and its constituents: a review. Iranian J Basic Med Sci. 2017;20(2):110.
39. Shirali S, Zahra Batiahe S, Nakjavan M. Effect of crocin on the insulin resistance and lipid profile of streptozotocin-induced diabetic rats. Physiol Res. 2017;66(7):1042–1049.
40. Batiahe S, Houshyar R, Mirdi H, Sadeghizadeh M. Anticancer effects of crocein in both human adenocarcinoma gastric cancer cells and rat model of gastric cancer. Biochem Cell Biol. 2013;91(6):397–403.
41. Zhao C, Bai W, Chen Q, et al. Anticancer effect of crocin against burn-induced intestinal injury. J Surg Res. 2015;198(1):99–107.
42. Sravya T, Rao GV, Kumar MG, Sagir YV, Sivarajanan Y, Sudheerkanth K. Evaluation of biosafe alternatives as xylene substitutes in hepatotoxicity and eosin staining procedure: a comparative pilot study. J Oral Maxillofac Pathol. JOMPP.
M.-H. Arjmand, M. Hashemzehi, A. Soleimani et al.

2018;22(1):148.

O’Connor WN, Valle S. A combination Verhoefts elastic and Masson’s trichrome stain for routine histology. Stain Technol. 1982;57(4):207–210.

Swolin K. Experimented studien zur prophylaxe von intraabdominalen verwachsungen: versuche an der Ratte mit einer Emulsion aus Lipid und Prednisolon. Acta Obstet Gynecol Scand. 1966;45(4):473–498.

Giusto G, Vercelli C, Hussich S, et al. A pericin-honey hydrogel prevents post-operative intraperitoneal adhesions in a rat model. BMC Vet Res. 2017;13(1):55, 55.

Roghani M, Baluchnejadmojarad T, Roghani DF. Involvement of Oxidative Stress Attenuates Vascular Permeability in Rats with Streptozotocin-Induced Diabetes: The Sesame Lignan Sesamin. J Cell Physiol. 2017;232(10):6785–6798.

Hashemzehi M, Behnam-Rassouli R, Hassanian SM, et al. Phytosomal-curcumin antagonizes cell growth and migration, induced by thrombin through AMP-kinase in breast cancer. J Cell Biochem. 2018;119(7):5996–6007.

Asgharzadeh F, Bargi R, Beheshti F, Hosseini M, Farzadnia M, Khazaei M. Thymoquinone restores liver fibrosis and improves oxidative stress status in a lipopolysaccharide-induced inflammation model in rats. Avicenna J. Phytomed. 2017;7(6):502.

Beck G, van Kalsbeek W, van Huijgevoort K. Phytosome-curcumin inhibits tumor growth in colitis-associated colorectal cancer. J Nutr Biochem. 2018;50:1–11.

Hashemi SA, Karami M, Bathaie SZ. Cytosolic and mitochondrial superoxide dismutase activity in breast cancer: in vitro, in vivo and in silico studies. Int J Biol Macromol. 2020;158:845–853.

Srivastava R, Desai S, Agarwal R, et al. Crocin: a protective natural antioxidant against pulmonary fibrosis induced by bleomycin. Pharmacol Rep. 2020;1–10.

Chhimwal J, Sharma S, Kulurkar P, Patel V. Crocin attenuates CCH4-induced liver fibrosis via PPAR-y mediated modulation of inflammation and fibrogenesis in rats. Hum Exp Toxicol. 2020;39(12):1639–1649.