Wrinkle reduction in post-menopausal women consuming a novel oral supplement: a double-blind placebo-controlled randomized study

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Synopsis.
OBJECTIVE: The maintenance of youthful skin appearance is strongly desired by a large proportion of the world’s population. The aim of the present study was therefore to evaluate the effect on skin wrinkling, of a combination of ingredients reported to influence key factors involved in skin ageing, namely inflammation, collagen synthesis and oxidative/UV stress. A supplemented drink was developed containing soy isoflavones, lycopene, vitamin C and vitamin E and given to post-menopausal women with a capsule containing fish oil.

METHOD: We have performed a double-blind randomized controlled human clinical study to assess whether this cocktail of dietary ingredients can significantly improve the appearance of facial wrinkles.

RESULTS: We have shown that this unique combination of micronutrients can significantly reduce the depth of facial wrinkles and that this improvement is associated with increased deposition of new collagen fibres in the dermis.

CONCLUSION: This study demonstrates that consumption of a mixture of soy isoflavones, lycopene, vitamin C, vitamin E and fish oil is able to induce a clinically measurable improvement in the depth of facial wrinkles following long-term use. We have also shown, for the first time with an oral product, that the improvement is associated with increased deposition of new collagen fibres in the dermis.

Introduction
The maintenance of youthful skin appearance is strongly desired by a large proportion of the world’s population. Skin, like all organs of the body, undergoes alterations due to the passage of time and changes are primarily characterized by a loss of elasticity, increased roughness, uneven skin tone and the appearance of wrinkles. The prevalence of pruritus also increases in aged skin, reflecting poor hydration, impaired skin barrier and altered neural function [1, 2].

The underlying biological mechanisms that give rise to these symptoms are numerous, including changes in dermal extracellular matrix (ECM) turnover, increases in inflammatory status and reduction in blood flow. Skin wrinkling in particular arises, in part, from atrophy of the dermal layer of which type I collagen is the major component. This protein is by far the most abundant in human skin, comprising more than 90% of its dry weight and it confers structural integrity on the skin [3]. With increasing age, the amount and integrity of collagen in skin is reduced, a process that is further enhanced in photodamage [4]. This decline is brought about by increased expression of collagen-degrading matrix metalloproteinases (MMPs) with age, coupled with the failure to replace the damaged collagen with newly synthesized protein [5].

The period of most rapid decline in skin quality is around the menopause, with the dramatic decline in circulating oestrogen correlating strongly with loss of dermal collagen and contributing
significantly to pronounced skin atrophy, dryness and sensitivity [6], along with decreased elasticity and wrinkle formation [7]. Furthermore, the positive effects of oestrogen replacement on skin have been demonstrated in clinical studies that have shown that hormone replacement therapy (HRT) greatly improves the skin of menopausal women [8, 9]. Increased collagen content, dermal thickness and elasticity have also been demonstrated [10].

Because many of the age-associated changes in skin occur deep in the dermis, they are often difficult to influence with topical products. Effective delivery of nutrients through the diet to the lower region of the skin, particularly to the collagen-producing dermal fibroblasts, is therefore an attractive option. There has been an increasing amount of evidence in recent years showing the potential effects of diet on skin ageing, with new insights into the relationship between food intake and skin appearance beginning to emerge [11, 12].

There is now convincing evidence from in vitro, animal and human studies that dietary constituents can afford an important degree of endogenous photoprotection and may therefore be presumed to help reduce skin photoaging. Whilst the level of sun protection (measured as the sun protection factor; SPF) with an SPF 2 to 4 is considerably lower than that afforded by topical sun creams with an SPF of 5 to >50, over a lifetime systemic dietary, photoprotection is still expected to contribute very significantly to overall improved skin appearance.

The ability of a nutrient to provide systemic photoprotection requires it to have one or more of the following functions: prevent the absorption of UV light by skin, protect target molecules by acting as an antioxidant scavenger, induce cellular repair systems or suppress cellular responses such as inflammation. Vitamins such as vitamin C and vitamin E [13–16], carotenoids (E.g. lycopene) [17–22] and n-3 polyunsaturated fatty acids [23, 24] have all been shown to provide photoprotective benefits, following oral consumption and are believed to operate through one or more of the above mechanisms. Carotenoids are highly effective scavengers of singlet oxygen [25, 26], whilst n-3 polyunsaturated fatty acids have been shown to influence inflammation in skin through down regulation of prostaglandin E2 (PGE2) and leukotriene B4 (LTB4) [27–29]. Both vitamin E and vitamin C form part of the skin’s natural defence against reactive oxygen species and their interaction is thought to be particularly important in the protection of skin against photo-oxidation [30]. Vitamin C is also an essential cofactor in the biosynthesis of collagen, an important process in the prevention of skin ageing [31].

Given the association of oestrogen levels with deterioration in skin quality at menopause, plant-derived compounds that are able to mimic oestrogen have also received much attention as potential anti-ageing nutrients. These so-called phytoestrogens have been described as a partial alternative to HRT, due to their ability to bind to and activate the oestrogen receptor [32, 33]. Activation of this receptor has also been associated with induction of collagen synthesis by dermal fibroblasts [34]. Phytoestrogens are believed to offer potential therapies for hormonal-dependent conditions, including the menopause, without the unwanted side effects associated with HRT [35]. Indeed, epidemiological studies have indicated a strong correlation between the consumption of soya products in the Far East, containing high levels of phytoestrogens and reduced incidence of hormone-related diseases, including some menopausal symptoms [36–38]. In addition to their hormonal-like effect, dietary phytoestrogens have also been reported to exert their effects via non-hormonal mechanisms [39]. It is therefore hypothesized that long-term consumption of foods containing phytoestrogens may exert anti-ageing effects in post-menopausal women, including alleviation of skin-associated symptoms.

The aim of the present study was therefore to evaluate the effect on skin wrinkling, of a combination of ingredients reported to influence key factors involved in skin ageing; namely inflammation, collagen synthesis and oxidative/UV stress. A supplemented drink was therefore developed containing soy isoflavones, lycopene, vitamin C and vitamin E and given to post-menopausal women with a capsule containing fish oil.

Post-menopausal women are considered a useful cohort to study, with respect to skin ageing, as the quality of skin declines dramatically in women following the menopause. Any intervention aimed at correcting the symptoms of skin ageing are therefore likely to have a greater effect in this target population.

Materials and methods

Randomized controlled study

The study was double-blind and placebo-controlled and conducted by ProDERM Institute for Applied Dermatological Research (Hamburg, Germany) between January and July 2006.

Eligible subjects were healthy, non-smoking, post-menopausal females (skin type I-III as described by the Fitzpatrick skin type scale [40]) who were not receiving HRT, not using dietary supplements and with a BMI between 18 and 33 kg m⁻². Power calculations for the smallest difference between the treatments were performed by bootstrapping data of sample sizes of 101 individuals from a previous double-blind placebo-controlled study with identical primary objectives [41]. These calculations indicated that a sample size of 50 individuals per treatment group would be sufficient to achieve a power of 75%.

Subjects were allocated to treatment groups using a random allocation procedure in SAS (SAS Institute Inc., Cary NC, U.S.A). The randomization was stratified by age, BMI and time at menopause. Study sponsors, investigators and statisticians remained blind to the treatment allocation to avoid bias during data analysis. Subjects were randomized into three groups of equal size in a parallel design. Two arms of the study were given a combination of drinks and capsules containing differing dose levels of the active ingredients. The third arm was given placebo drinks & capsules, containing no active ingredients. Test products were a combination of:

- Total isoflavone (expressed as aglycone) 0 70 (43) 40 (25)
- Lycopene 0 8 3
- Vitamin C 0 250 180
- Vitamin E 0 250 30
- Omega-3 EFA’s (23% EPA, 16% DHA) 0 660 660

Table I  Study treatments and active ingredient doses

| Active ingredient                  | Placebo group | Test group 1 | Test group 2 |
|------------------------------------|---------------|--------------|--------------|
| Dose (quantity per day)            |               | 1            | 1            |
| 100 mL drink                       | 2             | 2            | 2            |

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of a fruit-based drink containing vitamin C, vitamin E, lycopene and isoflavones and capsules containing omega-3 EFA. The respective doses are outlined in Table I.

Subjects were asked to consume one dose of the study treatment per day for 14 weeks. Non-invasive skin measurements were taken at baseline (T1) and week 15 (T15) along with blood samples, following a minimum 6-h fast, to assess plasma nutrient status. In addition, 3-mm full thickness punch biopsies were taken at baseline and at the end of the study to assess skin quality.

The Freiburger Ethik-Kommission International GmbH approved the study and all subjects gave written and informed consent. All the subjects were monitored for the occurrence of serious adverse events up to, and including, 28 days after their involvement with this study.

Study outcomes
The primary endpoint of the study was the measurement of wrinkle depth around the crow’s foot area of the eye. This was assessed by: (i) PRIMOS analysis of Sillo replications taken according to manufacturer’s guidelines and (ii) high-quality photography (Fuji S2 Pro Digital camera). In addition, skin firmness and elasticity (Courage & Khazaka Cutometer® CM825), barrier function and skin hydration (Dermalab® TEWL), skin colour (Konica Minolta Chromameter CR300) and histological biopsy analyses (collagen and elastin) were also measured.

Punch biopsies, at baseline (T1) and following 14 weeks intervention (T15), were obtained and snap-frozen in liquid nitrogen and stored at −70°C prior to analysis. The biopsies were defrosted and fixed in 10% neutral-buffered formalin and embedded in paraffin before being sectioned. Tissue was then stained with H&E and evaluated by a blinded, independent histopathologist for scored, qualitative changes in the quantity and quality of collagen and elastin fibres.

Analysis of vitamin and carotenoid levels in blood samples was performed by the Scottish Trace Element and Micronutrient Reference Laboratory at the Glasgow Royal Infirmary using standard HPLC methods. Triglyceride and isoflavone analyses were carried out at Unilever R&D Colworth using Fatty Acid Methyl Ester-Gas Chromatography (FAME-GC) analysis and automated immunoassay methods, respectively [42, 43].

Study products
Study drinks were prepared on behalf of Unilever at Crown Packaging Ltd, U.K. Drinks (100 mL units) were supplemented with the required level of individual actives, except those used for the placebo group. The capsules were produced by EuroCaps Ltd. Active capsules contained Omega-3 fish oil (Marinol C38; 23% EPA and 16% DHA) and placebo capsules contained sunflower oil (high oleic). To preserve study blinding, all test and placebo products were of similar appearance and were packaged and labelled in an identical way.

Statistical analyses
Study data were analysed using analysis of covariance (ANCOVA) with change from baseline after 14 weeks used as the response variable and the treatment group as the fixed effect of interest. Baseline, age and BMI were included in the model as covariates and results considered significant if \( P < 0.05 \) (95% confidence interval) as calculated using standard methodology for least-squares means (LSMEAN) within SAS.

A per-protocol analysis set with strict outcome exclusions defined prior to the treatment code being broken. A log-transformation was applied to the raw data and subsequently the change from baseline calculated for T15. This was included in a general linear mixed model as the dependent variable with the treatment as the three-level fixed effect of interest. Fixed continuous covariate terms were included for log (baseline) and the stratification variables age, BMI and time since menopause. Estimates were obtained for the least-squares means for the dependent variable for each treatment and also for the differences between test group 1/test group 2 and placebo. P-values for the differences were calculated and adjusted for multiplicity within each model using a Dunnett’s correction. In a post-hoc analysis, the interaction of treatment effect with the baseline wrinkle R3x scores, measured by PRIMOS, was also analysed.

Clinical assessment of biopsies
Differences in the quantity and quality of collagen and elastin in the skin of subjects receiving the placebo and test group 2 products over 14 weeks were assessed visually. Analysis was performed blinded from an independent histopathologist who documented changes in the quantity (i.e. changes in the number of orderly reticular collagen bundles and increases in branch-like elastin fibrils) and quality (such as fibre expansion/regression of collagen and arrangement/orientation of elastin fibrils seen above the capillary plexus). Each panelist was scored for a decrease (−1), maintenance (0) or increase (+1) in the level of collagen and/or elastin over the study period. The significance was assessed using a Fisher’s exact test, to give an overall P-value.

Results
Subject characteristics
One-hundred and sixty six post-menopausal women were recruited onto the study and were subsequently randomized into three equal treatment arms (a study flow chart is outlined in Table II). Baseline demographics for all treatment groups are indicated in Table III. Dropout rates were low, with only three subjects lost to follow-up and four withdrawn due to poor compliance or voluntary withdrawal. A total of 159 subjects completed the study and no serious adverse events were recorded.

Study compliance and uptake of functional ingredients
Compliance, as assessed by urinary isoflavone concentration at fortnightly intervals, was considered to be good throughout the study. Mean urinary isoflavone levels, detected in 24-h urine samples, were significantly higher in the test groups compared to placebo for the duration of the study (\( P < 0.001 \)). In addition, a difference could be observed between the two active treatments at all time points after baseline (\( P < 0.01 \)), with subjects on test group 1 consistently demonstrating higher excretion of both daidzein and genistein (Fig. 1).

The plasma levels of all active ingredients measured are also detailed in Table IV. Data show that all active ingredients were effectively absorbed into the bloodstream and again indicate that compliance was good throughout the study.
Wrinkle replica analysis

At the end of the study, sillsol replicas collected of the crow’s foot area of both sides of the face were analysed using standard Phase-shift Rapid In-vivo Measurement Of Skin (PRIMOS) optical 3D technology [44]. Data were generated for a number of roughness parameters appropriate to the 3D profile characterization of the skin [45, 46] as summarized in a means plot analysis (Fig. 2).

Statistically significant changes were observed over 14 weeks in several of the parameters, indicating that the test groups generally experienced a reduced severity of skin roughness compared with the placebo group over the same period. In particular, the change in R3z value over 14 weeks in both test groups was significantly reduced compared with placebo (Fig. 3). This parameter is considered to be one of the primary indicators of wrinkle depth [47], and the changes observed support a significant reduction in average wrinkle depth in both test groups compared with placebo.

The mean treatment effect on R3z value for both test groups was also examined, and this analysis showed a statistically significant effect ($P < 0.05$) of both groups when compared with placebo (Fig. 4). Furthermore, in a post-hoc analysis, the interaction effect of treatment and baseline R3z was also assessed. As seen in Fig. 5, the gradients of the best-fit lines are distinct and separate to the

Table II Study flow chart

| Assessed for eligibility/ Randomised ($n = 166$). |
|-----------------------------------------------|
| Allocated to Placebo group ($n = 56$). |
| Followed up ($n = 56$). Primary & secondary outcomes measured at weeks 9 & 15. |
| Withdrawn ($n = 1$). |
| Completed trial ($n = 55$). |
| Allocated to Test Group 1. High dose ($n = 55$). |
| Followed up ($n = 55$). Primary & secondary outcomes measured at weeks 9 & 15. |
| Withdrawn ($n = 2$). Lost to follow up ($n = 2$). |
| Completed trial ($n = 51$). |
| Allocated to Test Group 2. Low dose ($n = 55$). |
| Followed up ($n = 55$). Primary & secondary outcomes measured at weeks 9 & 15. |
| Withdrawn ($n = 1$). Lost to follow up ($n = 1$). |
| Completed trial ($n = 53$). |

Table III Mean subject demographics at screening (range)

| Placebo group | Test group 1 | Test group 2 | P-value |
|---------------|-------------|-------------|---------|
| No. of subjects: | 56 | 55 | 55 | N.S |
| Age (years) | 61 (45–71) | 61 (49–71) | 61 (48–70) | N.S |
| Years post-menopausal | 13 (2–19) | 12 (2–34) | 13 (3–30) | N.S |
| BMI | 27 (18–33) | 27 (19–33) | 27 (19–33) | N.S |

Non-invasive skin measures

Wrinkle replica analysis

At the end of the study, sillsol replicas collected of the crow’s foot area of both sides of the face were analysed using standard Phase-shift Rapid In-vivo Measurement Of Skin (PRIMOS) optical 3D technology [44]. Data were generated for a number of roughness parameters appropriate to the 3D profile characterization of the skin [45, 46] as summarized in a means plot analysis (Fig. 2).

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The mean treatment effect on R3z value for both test groups was also examined, and this analysis showed a statistically significant effect ($P < 0.05$) of both groups when compared with placebo (Fig. 4). Furthermore, in a post-hoc analysis, the interaction effect of treatment and baseline R3z was also assessed. As seen in Fig. 5, the gradients of the best-fit lines are distinct and separate to the
greatest extent at higher baseline R3z values, with the relationship being clearly significant for test group 2 (P < 0.01). This interaction highlights that treatment with the active ingredients had a stronger effect on deeper wrinkles.

Firmness & elasticity
Skin elasticity was measured using a Courage & Khazaka Cutometer (probe aperture size 8 mm) at baseline and week 15 on the inner & outer forearm. However, there was no statistical difference from baseline or between placebo and test treatment at any time point in any of the parameters measured (data not shown).

Barrier function & skin hydration
Transepidermal water loss (TEWL) and hydration were also measured at baseline and after 14 weeks of intervention. There was a significant increase from baseline in TEWL with all treatments and at both test sites (Fig. 6a). This was not unexpected as seasonal differences in skin barrier integrity have previously been

Table IV Mean ingredient plasma levels (standard deviation)

| Mean plasma levels | Placebo group | Test group 1 | Test group 2 |
|--------------------|---------------|--------------|--------------|
| Vitamin C (µM)     |               |              |              |
| T1                 | 55.07 (15.66) | 59.50 (16.8) | 56.85 (17.8) |
| T9                 | 52.80 (15.05) | 74.64 (19.54) | 72.37 (13.93) |
| T15                | 55.00 (13.22) | 72.30 (17.06) | 72.46 (14.56) |
| Vitamin E (µM)     |               |              |              |
| T1                 | 34.44 (7.45)  | 34.60 (9)    | 38.50 (10)   |
| T9                 | 34.85 (7.28)  | 50.10 (14)   | 46.90 (13)   |
| T15                | 36.30 (7.58)  | 50.90 (13)   | 47.20 (11)   |
| Lycopene (µg L⁻¹)  |               |              |              |
| T1                 | 114.94 (65.76) | 108.71 (77) | 124.00 (75) |
| T9                 | 120.50 (74.09) | 234.70 (104) | 179.20 (70) |
| T15                | 100.87 (61.00) | 254.40 (103) | 201.4 (75) |
| Plasma EPA (µg mL⁻¹) |           |            |              |
| T1                 | 43.37 (26.11) | 42.91 (28.59) | 44.40 (41.2) |
| T15                | 42.98 (21.38) | 67.32 (26.45) | 71.09 (28.28) |
| Plasma DHA (µg mL⁻¹) |           |            |              |
| T1                 | 80.20 (26.86) | 76.25 (29.37) | 80.36 (23.69) |
| T15                | 82.69 (25.10) | 97.54 (31.18) | 103.56 (33.36) |

*Significant difference from baseline (T1): P < 0.05.
**Significant difference from placebo (within time point): P < 0.05.
†Significant difference from other active treatment (within time point): P < 0.05.

Figure 1 Analysis of Urinary Isoflavones (mean ± standard deviation). (a) ng/ml urinary daidzein (b) ng/ml urinary genistein. Significance tested with students t-test and considered significant if P < 0.05 (95% confidence interval).

Figure 2 Means plot + Standard Error of the Mean (SEM). PRIMOS analysis generates quantitative data for several parameters with relation to the 3D profile of the skin including Ra, Rz, Rq, R3Max, R3z, Rp, W1 and Rc. Placebo (PL), Test group 1 (T1) and Test group 2 (T2).

Figure 3 Mean Replica R3z values, expressed as change from baseline (CFB T15-T1). Values are mean ± standard deviation. Placebo (grey), Test group 1 (white), Test group 2 (black). Significance is displayed vs. placebo using ANOVA (** P < 0.01; *** P < 0.005).

Skin elasticity was measured using a Courage & Khazaka Cutometer (probe aperture size 8 mm) at baseline and week 15 on the inner & outer forearm. However, there was no statistical difference from baseline or between placebo and test treatment at any time point in any of the parameters measured (data not shown).

Barrier function & skin hydration
Transdermal water loss (TEWL) and hydration were also measured at baseline and after 14 weeks of intervention. There was a significant increase from baseline in TEWL with all treatments and at both test sites (Fig. 6a). This was not unexpected as seasonal differences in skin barrier integrity have previously been
However, no significant difference was observed between test and placebo. Similarly, hydration also increased from baseline with all treatments and at both sites measured, but no statistical significance was observed between test and placebo (Fig. 6b).

**Skin colour**

The Minolta Chromameter allows for the objective determination of skin colour using the tristimulus L a* b* system, and increases in
b* values are commonly associated with the accumulation of carotenoids in the skin [49]. Measurements were taken at baseline and after 14 weeks of intervention, and higher b* values were observed in test groups, particularly within areas of the skin with high levels of subcutaneous fat (such as the palm and lower back) (Fig. 7). This data strongly indicates an accumulation of dermal carotenoids in the skin following 14 weeks of intervention [50].

There was no evidence of a significant treatment effect at any site for L and a* values (skin lightness and blood flow, respectively).

Biopsy analysis
Following evaluation from an independent histopathologist, a significant number of individuals in test group 2 demonstrated an increase in collagen quantity and quality in biopsies after 14 weeks compared with those on the placebo treatment when examined by Fisher’s exact test (Fig. 8). Given the high number of individuals recruited into each cell and the time-consuming nature of the histological evaluation, biopsies from individuals on test group 1 treatment were not analysed for changes in collagen levels. No significant differences were observed in elastin quantity and/or quality between control and test group 2.

Digital photography
High-quality photography was taken at the beginning (T0) and end of the study (T15). Examples of subjects are shown in Fig. 9.

Discussion
The purpose of this study was to establish that a unique combination of ingredients given through the diet would have a significant benefit on skin wrinkle formation in a population of post-menopausal Caucasian women. We have shown that a combination of isoflavones, omega-3 fatty acids, lycopene, vitamin C and vitamin E when consumed orally for 14 weeks can significantly reduce the depth of facial wrinkles. We have also demonstrated, for the first time, a significant increase in collagen quantity and quality in biopsies from individuals on test group 2 treatment.
time with an oral product, that the improvement is associated with increased deposition of new collagen fibres.

We have performed a double-blind randomized controlled investigation to assess whether a combination of dietary ingredients, selected for their proposed skin anti-ageing benefits, can significantly improve the appearance of facial wrinkles. The study was conducted following the principles of international good clinical practice standards, with products coded and randomized at source and with subjects, investigators and statistician remaining ‘blind’ to the coding until the study and initial data analysis were complete.

We observed no benefit of the test or placebo products on skin colour, hydration or barrier quality. However, we did demonstrate that individuals consuming the active test products showed significantly improved skin texture and reduced wrinkle depth when compared with the placebo group. On average, there was a 10% reduction in wrinkle depth in the test group when compared with placebo. Interestingly, we also demonstrated a baseline interaction effect with regard to wrinkle depth and response to treatment, that is, the deeper the wrinkle the greater the reduction in depth after 14 weeks. This potentially highlights the mechanistic action of the treatment. The smaller wrinkles as measured by PRIMOS technology are most likely a result of surface changes in the upper epidermal layers, whilst deeper wrinkles are potentially caused by alterations in the deeper dermal layer of skin. Any nutritional intervention therefore, taken orally is likely to have a greater effect on this latter type of wrinkle, so individuals with deeper wrinkles will experience the greatest benefit.

Interest in the role of specific food ingredients on promoting optimal skin condition has grown considerably in the past few years, with new insights into the relationship between food intake and skin health beginning to emerge. There are now comprehensive clinical data demonstrating the photoprotective benefits of dietary carotenoids [51, 52] vitamins E & C [13, 14] and fish oil (containing EPA and DHA) [27, 28]. Furthermore, recent studies have indicated that oral intake of soy isoflavone aglycones appears to improve the aged skin of middle-aged women [53–55].

In addition, we examined the distribution of collagen in skin samples taken at the beginning and end of the study. Collagen is the main structural protein of the skin, and the abrupt onset of visible skin ageing around the menopause is primarily the consequence of a lower production of collagen, the supportive protein network of the skin [56–58].

We found that a significantly higher number of individuals consuming the test product (test group 2) showed increased levels of collagen in skin biopsies after 14 weeks, than those who consumed placebo products. To our knowledge, this is the first published report of increased collagen production in skin as a result of an oral intervention.

A reduction in the level of collagen synthesis and general decline in the quality of existing collagen has been reported to contribute greatly to the increase in wrinkles and loss of firmness associated with ageing skin [59]. To obtain real wrinkle reduction effects therefore, matrix rejuvenation would be needed. The increase in collagen observed provides a potentially underlying reason for the reduction in wrinkle depth demonstrated in the study.

The ability of the ingredients described here to induce skin collagen is further supported by evidence from previously published studies. Cultured human fibroblasts in vitro treated with an isoflavone soy-containing extract demonstrated an increased level of collagen synthesis [60], whilst mice fed dietary soy isoflavones also showed increased deposition of collagen in their skin [61]. In addition, there are numerous reports of omega-3 fatty acids influencing collagen production in ligaments and connective tissue cells [62, 63]. Furthermore, addition of omega-3 fatty acids to mouse 3T3 cells in vitro has also been shown to be associated with enhanced collagen synthesis [64]. More recently, increased levels of collagen were also detected in human skin following topical application of omega-3 fatty acids [65].

Surprisingly, in this investigation, we were unable to detect any significant changes between the quantity and quality of elastin fibres between test and placebo groups. In fact, detailed histological evaluation revealed that the papillary microfibril scaffolds of elastin were already well organized and uniform in distribution and excellent in condition. These histological observations may provide further support to the lack of beneficial effects we observed between test and placebo firmness and elasticity measurements.

Figure 9 Examples of high quality digital photography with a Fuji S2 Pro Digital camera and two Nikon flashes (SB80DX) in three individuals from test group 2.
In conclusion, we believe that the combination of ingredients, rather than the level of each individual active, is the most important aspect for the overall functionality of the mix and that this study demonstrates that consumption of these specific nutritional ingredients is able to induce a clinically measureable improvement in the depth of facial wrinkles following long-term use. This improvement is associated with increased deposition of newly formed collagen in the dermis of panellists consuming the active combination.

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Conflict of interest

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References

1. Garibyan, L., Chio, A.S. and Elmariah, S.B. Advanced aging skin and itch: addressing an unmet need. Dermatol. Ther. 26, 92–103 (2013).
2. Jenkins, G. Molecular mechanisms of skin ageing. Mech. Ageing Dev. 123, 801–810 (2002).
3. Fisher, G.J., Varani, J. and Voorhees, J.J. Looking older: fibroblast collapse and therapeutically implications. Arch. Dermatol. 144, 666–672 (2008).
4. Baroni Edo, R., Biondo-Simões Mde, L., Aurassvald, A. et al. Influence of aging on the quality of the skin of white women: the role of collagen. Acta Cir. Bras. 27, 736–740 (2012).
5. Varani, J., Dame, M.K., Rittie, L. et al. Decreased collagen production in chronologically aged skin. Am. J. Pathol. 168, 1861–1868 (2006).
6. Shah, M.G. and Maibach, H.I. Estrogen and skin aging. In: Aging and Skin: Current and Future Therapeutic Strategies (Rhein, L.D., Fluhr, J.W., eds), pp. 377–394. Allured books, Illinois (2010); Chapter 13.
7. Fuchs, J. and Kern, H. Modulation of UV-light-induced skin inflammation by D-alpha-tocopherol and L-ascorbic acid: a clinical study using solar simulated radiation. Free Radic. Biol. Med. 25, 1006–1012 (1998).
8. Palme-König, B., Palme, M. and Praybilla, B. Protective effect against sunburn of combined systemic ascorbic acid (vitamin C) and d-alpha-tocopherol (vitamin E). J. Am. Acad. Dermatol. 38, 45–48 (1998).
9. La Ruche, G. and Cesarini, J.P. Protective effect of oral selenium plus copper associated with vitamin complex on sunburn cell formation in human skin. Photodermatol. Photoimmunol. Photomed. 8, 212–215 (1991).
10. Werninghaus, K., Meydani, M., Bhawan, J. et al. Evaluation of the photoprotective effect of oral vitamin E supplementation. Arch. Dermatol. 130, 1257–1261 (1994).
11. Smith-Warner, S.A., Elmir, P.J., Tharp, T.M. et al. Increasing vegetable and fruit intake: randomized intervention and monitoring in an at-risk population. Cancer Epidemiol. Biomarkers Prev. 9, 307–317 (2000).
12. Heinrich, U., Wiebusch, M. and Tironni, H. Photoprotection from ingested carotenoids. Nutr. J. 1, 919–925 (2003).
13. Orenjo, I.F., Black, H.S. and Wolf, J.E. Influence of fish oil supplementation on the minimal erythema dose in humans. Arch. Dermatol. Res. 284, 219–221 (1992).
14. Sies, H., Stahl, W. and Sundquist, A.R. Antioxidant functions of vitamins of vitamins E and C and beta-carotene, and other carotenoids. Ann. NY Acad. Sci. 669, 7–20 (1992).
15. Handelman, G. The evolving role of carotenoids in human biochemistry. Nutr. J. 17: 818–822 (2001).
16. Rhodes, L.E., O’Farrell, S., Jackson, M.J. and Friedmann, P.S. Dietary fish-oil supplementation in humans reduces UVB-erythemal sensitivity but increases epidermal lipid peroxidation. J. Invest. Dermatol. 103, 151–154 (1994).
17. Rhodes, L.E., Durham, B.H., Fraser, W.D. and Friedmann, P.S. Dietary fish oil reduces basal and ultraviolet B-generated PG2 levels in skin and increases the threshold to provocation of polymorphic light eruption. J. Invest. Dermatol. 105, 532–535 (1995).
18. Pilington, S.M., Watson, R.E., Nicolau, A. and Rhodes, L.E. Omega-3 polyunsaturated fatty acids: photoprotective macronutrients. Exp. Dermatol. 20, 537–543 (2011).
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30. Wefers, H. and Sies, H. The protection by ascorbic and glutathione against microsomal lipid peroxidation is dependent on vitamin E. Eur. J. Biochem. 174, 353–357 (1988).
31. Pinnel, S.R., Murad, S. and Darr, D. Induction of collagen synthesis by ascorbic acid. A possible mechanism. Arch. Dermatol. 121, 1684–1686 (1987).
32. Albertazzi, P. and Purdie, D. The nature and utility of the phytoestrogens: a review of the evidence (reprinted from Maturitas, 2002; 42: 173-185). Maturitas 61, 214–226 (2008).
33. Cassidy, A. Potential risks and benefits of phytoestrogen-rich diets. Int. J. Vitam. Nutr. Res. 73, 120–126 (2003).
34. Surazynski, A., Jarzabek, K., Haczynski, J. et al. Differential effects of estradiol and raloxifene on collagen biosynthesis in cultured human skin fibroblasts. Int. J. Mol. Med. 12, 803–809 (2003).
35. Setchell, K.D. and Cassidy, A. Dietary isoflavones: biological effects and relevance to human health. J. Nutr. 129, 7585–7678 (1999).
36. North American Menopause Society. The role of soy isoflavones in menopausal health: report of The North American Menopause Society/Wulf H. Utian Translational Science Symposium in Chicago, IL (October 2010). Menopause 18, 732–753 (2011).
37. Welty, F.K., Lee, K.S., Lew, N.S. et al. Global and systematic demonstration for the practical usage of a direct in vivo measurement system to evaluate wrinkles. Int. J. Cosmet. Sci. 29, 421–436 (2007).
38. Pinnel, S.R., Murad, S. and Darr, D. Induc- tion of collagen synthesis by ascorbic acid. Methods Enzymol. 190, 360–371 (1990).
39. Jacob, V., Chen, M., Frankowski, G. et al. Nanosecond pulsed electric field microwaved skin: rapid and non-invasive binding of the non-steroidal anti-inflammatory drug ibuprofen. Skin Res. Technol. 10, 207–214 (2004).
40. Conti, A., Rogers, J., Verdejo, P. et al. Seasonal influences on stratum corneum ceramide 1 fatty acids and the influence of topical essential fatty acids. Int. J. Cosmet. Sci. 18, 1–12 (1996).
41. Alahaf, S., Heinrich, U., Stahl, W. et al. Die- tary carotenoids contribute to normal human skin color and UV photosensitivity. J. Nutr. 132, 399–403 (2002).
42. Stahl, W., Heinrich, U., Jungmann, H. et al. Increased dermal carotenoid levels assessed by non-invasive reflection spectrophotometry correlate with serum levels in women ingesting Betaline. J. Nutr. 128, 903–907 (1998).
43. Talbot, D.C., Ogborne, R.M., Dadd, T. et al. Monoclonal antibody-based time-resolved fluorescence immunoassays for daidzein, genistein, and equol in blood and urine: application to the Inoheut intervention study. Clin. Chem. 53, 748–756 (2007).
44. Leveque, J.L. EEMCO guidance for the assessment of skin topography. J. Eur. Acad. Dermatol. Venereol. 12, 103–114 (1999).
45. Fujimura, T., Haketa, K., Hotta, M. et al. Global and systematic demonstration for the practical usage of a direct in vivo measurement system to evaluate wrinkles. Int. J. Cosmet. Sci. 29, 421–436 (2007).
46. Grove, G.L. and Grove, M.J. Effects of topical retinoids on photoaged skin measured by optical profilometry. Methods Enzymol. 190, 360–371 (1990).
47. Jacobi, V., Chen, M., Frankowski, G. et al. Nanosecond pulsed electric field microwaved skin: rapid and non-invasive binding of the non-steroidal anti-inflammatory drug ibuprofen. Skin Res. Technol. 10, 207–214 (2004).
48. Conti, A., Rogers, J., Verdejo, P. et al. Seasonal influences on stratum corneum ceramide 1 fatty acids and the influence of topical essential fatty acids. Int. J. Cosmet. Sci. 18, 1–12 (1996).
49. Alahaf, S., Heinrich, U., Stahl, W. et al. Die- tary carotenoids contribute to normal human skin color and UV photosensitivity. J. Nutr. 132, 399–403 (2002).
50. Stahl, W., Heinrich, U., Jungmann, H. et al. Increased dermal carotenoid levels assessed by non-invasive reflection spectrophotometry correlate with serum levels in women ingesting Betaline. J. Nutr. 128, 903–907 (1998).
51. Mathews-Roth, M.M., Pathak, M.A., Parrish, J. et al. A clinical trial of the effects of oral beta-carotene on the responses of human skin to solar radiation. J. Invest. Dermatol. 59, 349–353 (1972).
52. Cesarini, J.P., Michel, L., Maurette, J.M. et al. Immediate effects of UV radiation on the skin: modification by an antioxidant complex containing carotenoids. Photodermatol. Photoin- munol. Photomed. 19, 182–189 (2003).
53. Izumi, T., Saito, M., Obata, A. et al. Oral intake of soy isoflavone aglycone improves the aged skin of adult women. J. Nutr. Sci. Vitam. (Tokyo) 53, 57–62 (2007).
54. Skovgaard, G.R., Jensen, A.S. and Sigler, M.L. Effect of a novel dietary supplement on skin aging in post-menopausal women. Eur. J. Clin. Nutr. 60, 1201–1206 (2006).
55. Thom, E. A randomized, double-blind, pla- cebo-controlled study on the clinical efficacy of oral treatment with DermaVite on ageing symptoms of the skin. J. Int. Med. Res. 33, 267–272 (2005).
56. Brincat, M., Kabalan, S., Studd, J.W. et al. A study of the decrease of skin collagen content, skin thickness, and bone mass in the postmenopausal woman. Obstet. Gynecol. 70, 840–845 (1987).
57. Attinato, P., Palomba, S., Sorrentino, C. et al. Effects of postmenopausal hypoestrogenism on skin collagen. Maturitas 31, 239–247 (1999).
58. Adamia, A., Skorupski, P., Recherberger, T. and Jakowicki, J.A. The expression of the gene encoding pro-alpha 1 chain of type I collagen in the skin of premenopausal and postmenopausal women. Eur. J. Obstet. Gynecol. Reprod. Biol. 93, 9–11 (2000).
59. McGrath, J.A., Eady, R.A.J. and Pope, F.M. Anatomy and organization of human skin. In: Rook’s Textbook of Dermatology (Burns, T., Breathnach, S., Cox, N. and Griffiths, C., eds), 7th edn, pp. 3.1–1.84. Blackwell Publishing, Oxford (2004): Chapter 3.
60. Sudek, K.M., Venke, K., Mielke, H. et al. Novel aspects of intrinsic and extrinsic aging of human skin: beneficial effects of soy extract. Photochem. Photobiol. 81, 581–587 (2005).
61. Kim, S.Y., Kim, S.J., Lee, J.Y. et al. Protective effects of dietary soy isoflavones against UV-induced skin-aging in hairless mouse model. J. Am. Coll. Nutr. 2, 157–162 (2004).
62. Jia, Y. and Turek, J.J. Altered NF-kappaB gene expression and collagen formation induced by polyunsaturated fatty acids. J. Nutr. Biochem. 16, 500–506 (2005).
63. Haukensohn, K.D., Watkins, B.A., Schoenlein, I.A. et al. Omega-3 fatty acids enhance liga- ment fibroblast collagen formation in associ- ation with changes in interleukin-6 production. Proc. Soc. Exp. Biol. Med. 223, 88–95 (2000).
64. Jia, Y. and Turek, J.J. Polyenoic fatty acid ratios alter fibroblast collagen production via PGE2 and PGF receptor subtype response. Exp. Biol. Med (Maywood) 229, 676–683 (2004).
65. Kim, I.H., Cho, S., Lee, S. et al. Photoprotective and anti-skin-aging effects of eicosa- pentaenoic acid in human skin in vivo. J. Lipid Res. 47, 921–930 (2006).