Research Article

Evaluation of the Effect of Four Bioactive Compounds in Combination with Chemical Product against Two Spider Mites Tetranychus urticae and Eutetranychus orientalis (Acari: Tetranychidae)

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Currently, pests control using chemical acaricides constitutes worries for ecologists and health care people as these chemical products create damage to the ecosystem as well as the development of spider mites resistance. Such concerns request deep and rapid feedback by looking for new alternative and eco-friendly methods. In recent years, a new field is evolving in the use of essential oils in pest management practices. Essential oils have been considered as potential pest management agents, because they demonstrate to have a broad range of bioactivity, possess contact, and fumigant toxicity. In addition, the major advantages of many plant-based acaricides lie in their low toxicity to agroecosystems. Botanical acaricides composed of essential oils may prove to be a good choice for the more persistent synthetic acaricides. In this study, the acaricidal effect of four plant-derived essential oils against adults of the two important crop pests, Tetranychus urticae (Koch) 1836 and Eutetranychus orientalis (Klein) 1936 are studied. The fumigant toxicity revealed that all the essential oils tested Mentha pulegium L., Lavandula stoechas L., Rosmarinus officinalis L., and Origanum compactum Benth (Lamiaceae family) displayed an acaricidal effect. At the highest dose (625 μl/ml), mortalities recorded were found between 91 and 98% and 92 and 99% at 24 and 48 h, respectively, for T. urticae, and between 90 and 98% and 94 and 99% at 24 and 48 h, respectively, for E. orientalis. The M. pulegium L. essential oil represents the highest activity against E. orientalis and T. urticae. For the binary combination between the EOs (essential oils) and the acaricide based on the active ingredient acequinocyl, the results showed that the mixture of O. compactum EO (essential oil) + acequinocyl exhibited an important acaricidal effect on T. urticae and E. orientalis with 99% at 24 h and 100% at 48 h of mortality, followed by M. pulegium L. EO + acequinocyl with 92% at 24 h and 95% at 48 h for T. urticae as well as 99% at 24 h and 100% at 48 h for E. orientalis of mortality. Whereas, the mixture of L. stoechas EO + acequinocyl presented the lowest activity against T. urticae and
1. Introduction

Spider mites including *Tetranychus urticae* (Koch) 1836 and *Eutetranychus orientalis* (Klein) 1936 are a major pest of citrus in many countries including Morocco [1, 2]. These two pests cause considerable losses to citrus crops [3, 4]. In recent years, these pests have become of great economic importance [5, 6], by sucking the cellular juice of the leaves causing yellowing, necrosis, and leaf fall which weakens the plant and reduces the photosynthetic capacity [7]. Their control is often provided by intensive and preventive chemical control [8, 9]. In Morocco, the Acequinocyl, Abamectin, Malathion, Spirodiclofen, Bifenazate, Fenpyroximate, Fenzaquzin, Tebufenpyrad, Propargite, Milbemectine, Bifenthrene, Dicofol, Clofentezine, Maltdoxetine, and Pyridaben are the most acaricides employed to control spider mites including *T. urticae* and *E. orientalis* under greenhouses and field [1, 10]. Despite advantages such as easy use, the decrease of pests numbers, and their immediate action when compared with natural extracts from plants [11]. This approach has considerable limitations; the examination of the spectrum activity of the functional components used throughout the globe demonstrates that 72% of insecticides, 28% of fungicides, and 46% of acaricides are globally toxic toward arthropods and public health [12, 13].

Acequinocyl is a naphthoquinone compound discovered in the 1970s by DuPont [14]. It is a proacaricide that breaks down into the active metabolite, a deacetylated product. A mechanistic study showed that the deacetylated metabolite of acequinocyl inhibits respiration in mitochondria at the deacetylated metabolite site (Q0) of complex III of the electron transfer chain [15, 16].

The control by *Mentha pulegium*, *Lavandula stoechas*, *Rosmarinus officinalis*, and *Origanum compactum* (Lamiaceae family) can be used as alternative approaches for control. Essential oils include a plentiful interest for researchers who always try through their scientific studies to discover the great potential of these substances as anti-inflammatory, antioxidant, antimicrobial, and AC agents [17, 18]. Plants from the Lamiaceae family including *M. pulegium*, *L. stoechas*, *R. officinalis*, and *O. compactum* are well known for their acaricidal dynamic biological activities and many reports have demonstrated the existence of different compounds like terpenes, iridoids, flavonoids, and phenolic constituents the species of this family [19, 20]. Moreover, it had been conveyed that the two Lamiaceae plants, *M. pulegium* and *R. officinalis* have multiple significant acaricidal activities against spider mites [21, 22]. *M. pulegium* possesses several acaricidal properties; this aromatic plant has demonstrated antioxidant and anticholinesterase, anthelmintic, antimicrobial, insecticidal, and acaricidal effects [23–25]. Similarly, *R. officinalis* is well known for its beneficial effect as a medicinal mechanism and it also showed an acaricidal potential against pests [26]. The objective of this work was to evaluate the acaricidal effect of four essential oils derived from *M. pulegium*, *L. stoechas*, *R. officinalis*, and *O. compactum*, alone and in combination with a chemical acaricide based on acequinocyl, against two spider mites, *T. urticae* and *E. orientalis*.

2. Materials and Methods

2.1. Plant Material and Extraction of Essential Oils. Samples of *Lavandula stoechas*, *Mentha pulegium*, and *Origanum compactum* (Lamiaceae family) were collected between April and June (the flowering stage) in the mountainous area that is located in the rural community of Timezgana (Northeastern Morocco, 34°34′48″ North, 4°43′48″ West), at an altitude of approximately 800 m.

Samples of *R. officinalis* come from the experimental garden of the Faculty of Science and Technology of Fez. The collected plants were identified at the National Agency for Medicinal and Aromatic Plants in Taounate, Morocco.

These products were chosen on the basis of their availability and their effectiveness as a natural acaricide.

2.2. Chromatographic Analysis. Chemical analysis of all essential oils was realized using gas chromatography coupled with mass spectrometry (GC/MS) and coupled with flame ionization detection (GC-FID). The GC/MS analysis was utilized for identification, while GC-FID analysis was used for the quantification of components.

2.2.1. Gas Chromatography (GC-FID) Analysis. Gas chromatography analyses for all the samples were realized using a Hewlett-Packard (HP 6890) gas chromatograph equipped with an HP-5 capillary column (30 m × 0.25 mm, film thickness of 0.25 μm), FID a detector, and an injector fixed at 275°C. The oven temperature was at 50°C for 5 min and then increased to 250°C at 4°C/min. The nitrogen (1.8 ml/min) was utilized as carrier gas. The samples were diluted to 1/50 in methanol and the injected volume was 1 μl using a split mode ratio: 1/50, flow: 72.1 ml/min. The relative proportions of the EOs components were reported as percentages determined using peak-area normalization. The retention indices (RI) on HP-5 MS column, were identified using homologous series of (C8–C28) alkanes.

2.2.2. GC/MS Analysis. Chemical analysis was carried out using a Hewlett-Packard gas chromatograph (HP 6890) coupled with a mass spectrometer (HP 5973). The column
utilized was HP-5MS (cross-linked 5% PHME siloxane; 30 m × 0.25 mm, film thickness 0.25 µm). The column temperature was set at 50°C and increased to 250°C at a rate of 2°C/min. The carrier gas is helium at 1.5 ml/min, using a split mode ratio: 1/74.7, flow rate: 112 ml/min. MS identities of components were confirmed using the NIST 98 spectral library. The ion source temperature of 230°C with ionization voltages of 70 eV and a scan mass range of 35–450 m/z. The identification of the components was also confirmed by comparing the elution order of the compounds with the retention indices reported in the literature.

2.3. Spider Mites. The adult spider mites of T. urticae and E. orientalis (Acari: Tetranychidae) were collected from a farmer in Fez city 34°02′36″N, 05°00′12″W planted by (Citrus aurantium L., 1753) not previously treated with chemical products (Acaricides). The collected leaves were transferred directly into referenced polyethylene bags to the laboratory for inspection. T. urticae and E. orientalis spider mites on each leaf were determined and counted on both leaf surfaces using a binocular microscope before and after the addition of the tested products.

2.4. Toxicity Test. The acaricidal tests were conducted according to the methodology of Sertkaya et al. (2010) with slight modifications. Firstly, various doses (39.06, 78.125, 156.25, 312.5, and 625 µL/L) of each EO and the chemical acaricide (Kanemite, 15%) were prepared. Thereafter, 20 adults of each Tetranychidae species (T. urticae and E. orientalis) survived in citrus leaves were placed in glass petri dishes (90 mm × 20 mm). Disks of filter papers that were previously attached to the interior surface of the petri dish cover were impregnated with 10 µl of each concentration. DMSO (0.01%) was used as a negative control. Five replications of treatments and controls were carried out. The toxicity was evaluated after 24 and 48 h of exposure, and the mortality was calculated using the formula of Abbott (1).

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\text{%Mortality Corrected} = \left( \frac{\text{%Mortality Observed} - \text{%Mortality Control}}{\text{%Mortality Control}} \right) \times 100. \tag{1}
\]

2.5. Statistical Treatment. Mortality percentages were analyzed by the SPSS software (IBM SPSS Statistics 25.0). The obtained results were also examined with OriginPro 2021 software to determine the significant effect between mean values using the Tukey test (a probability of \( p \leq 0.05 \) is regarded as statistically significant). The different principal component analysis was carried out by using the JMP Pro 14. The LD\(_{50}\) values were estimated using a probit regression analysis (with SPSS software) according to Finney’s mathematical methods [27].

3. Results

3.1. Chemical Composition of the Four Tested EOs. As presented in Table 1, M. pulegium EO was dominated by \( r \)-pulegone (74.03%), carvone (5.45%), and dihydrocarvone (3.66%). L. stoechas EO was characterized by camphor (43.97%), fenchone (30.39%), camphene (4.09%), borneol (2.92%), and \( \alpha \)-pinene (2.84%); whereas 1,8 cineole (29.31%), camphor (24.66%), and \( \alpha \)-pinene (12.76%) were the main components in R. officinalis. EO. Thymol (30.05%), carvacrol (25.33%), and \( \gamma \)-Terpinene (17.23%) presented the main volatile components in O. compactum EO.

3.2. Toxicity of EOs on T. urticae and E. orientalis. Figures 1 and 2 illustrate the significant differences of the acaricidal effects of the four EOs and the chemical acaricide on T. urticae (Figure 1) and E. orientalis (Figure 2) according to mortality time (Figure 2(a)) and different concentration (Figure 2(b)). A significant difference (\( p < 0.05 \)) was noted between M. pulegium, R. officinalis, L. stoechas, and O. compactum (\( p = 0.008 \)) as well as between the four EOs and acaricide (\( p = 0.009 \)) in terms of mortality (%). In addition, a significant difference was observed between concentration (C1, C2, C3, and C4) of EOs and acaricide (\( p = 0.004 \)) in terms of mortality (%). However, no significant difference (\( p > 0.05 \)) was detected between EOs and acaricide (\( p = 0.08 \)) in terms of concentration C5, except R. officinalis EO for T. urticae. No significant difference was found between the mortality time of the four EOs and acaricide (\( p = 0.18 \)) in terms of T. urticae and E. orientalis mortality. It could be said that the mortality varies according to the oil and the acaricide tested and the treated mite species, generally; it was concentration-dependent. Indeed, M. pulegium and O. compactum EOs were the most effective against the two mites at all concentrations used, yet all EOs caused more than 90% of mortalities at the highest dose (C5: 625 µL/L). These results showed the important acaricidal potential of the different EOs. The chemical acaricide, Kanemite with the acequinocyl as the active molecule also displayed toxicity that varied in function of doses.

The lethal doses (LD\(_{50}\)) values obtained for each essential oil and the chemical acaricide are presented in Table 2. It was found that M. pulegium EO, O. compactum EO, and the chemical acaricide displayed the lowest LD\(_{50}\) values on adults of T. urticae after 24 h (65.78, 87.92, and 36.25 µL/L, respectively) and after 48 h of treatment (56.1, 79.72, and 32.09 µL/L, respectively). For E. orientalis, the lowest LD\(_{50}\) values were obtained for
Table 1: The major chemical constituents of the four EOs.

| Compound          | RI \(^a\) | RI \(^b\) | M. pulegium | L. stoechas | R. officinalis | O. compactum |
|-------------------|-----------|-----------|-------------|-------------|---------------|--------------|
| α-pinene          | 939       | 932       | 0.42        | 2.84        | 12.76         | 2.24         |
| Camphene          | 953       | 946       | —           | 4.09        | 2.47          | Tr           |
| α-Terpinene       | 1002      | 1007      | —           | —           | Tr            | 3.45         |
| p-Cymene          | 1026      | 1022      | —           | 0.32        | 0.09          | 11.22        |
| 1,8-cineole       | 1033      | 1033      | —           | 1.42        | 31.15         | Tr           |
| γ-Terpinene       | 1062      | 1024      | —           | 0.34        | —             | 17.23        |
| Fenchone          | 1088      | 1087      | —           | 30.39       | —             | Tr           |
| Camphor           | 1143      | 1146      | —           | 43.97       | 26.96         | 0.14         |
| Bornceol          | 1165      | 1166      | —           | 2.92        | 5.46          | 0.42         |
| Dihydrocarvone    | 1194      | 1191      | 3.66        | —           | —             | —            |
| R (+)-pulegone     | 1238      | 1233      | 74.03       | —           | Tr            | —            |
| Carvone           | 1242      | 1239      | 5.45        | 0.12        | —             | Tr           |
| Thymol            | 1290      | 1293      | —           | —           | —             | 30.05        |
| Carvacrol         | 1298      | 1291      | —           | —           | —             | 25.33        |
| Total %           |           |           | 83.56       | 86.41       | 78.89         | 90.08        |

Figure 1: Mortality rate (%) of *Tetranychus urticae* adults (a) according to mortality time. T1: 24 h; T2: 48 h, (b) according to various concentrations. C1: 39.06 µl/ml, C2: 78.125 µl/ml, C3: 156.25 µl/ml, C4: 312.5 µl/ml, C5: 625 µl/ml. Different letters indicate significant differences according to the Tukey test.

3.3. Toxicity of the EOs and the Acaricide on *T. urticae* and *E. orientalis*. As shown in Figure 3, the four binary mixtures exhibited an important acaricidal effect on both *T. urticae* and *E. orientalis* after 24 and 48 h of treatment. A significant difference \(p < 0.05\) was indicated in the mortality percentage of *T. urticae* between the *M. pulegium* EO + chemical acaricide (acequinocyl), *L. stoechas* EO + chemical acaricide (acequinocyl), and *O. compactum* EO + chemical acaricide (acequinocyl). No significant difference \(p > 0.05\) was identified between *M. pulegium* EO + chemical acaricide (acequinocyl), *R. officinalis* EO + chemical acaricide (acequinocyl), and *O. compactum* EO + chemical acaricide (acequinocyl) \(p = 1.00\), whereas a significant difference \(p < 0.05\) was found between *L. stoechas* EO + chemical acaricide (acequinocyl), and *R. officinalis* EO + chemical acaricide (acequinocyl) \(p = 0.014\).
acaricide (acequinocyl) and the three mixtures in terms of the mortality percentage of *E. orientalis* ($p = 0.544$). There was no significant in the mortality time of the four EOs and acaricide in terms of *T. urticae* and *E. orientalis* ($p = 0.115$). The binary combination (*M. pulegium* EO + chemical acaricide) and (*O. compactum* EO + chemical acaricide) exerted a high acaricidal effect against *T. urticae* with $92 \pm 2.74$–$99 \pm 2.23\%$ at 24 h and $95 \pm 3.54$–$100 \pm 0.00\%$ of mortalities, respectively. However, the mixtures (*M. pulegium* EO + chemical acaricide, *R. officinalis* EO + chemical acaricide, and *O. compactum* EO + chemical acaricide) were the most effective on spider mites adults of *E. orientalis* with $99 \pm 2.24\%$ at 24 h and $100 \pm 0.00\%$ at 48 h of mortality. These results are very relevant and proved the acaricidal potential of EOs if they compared with the individual one.

### 3.4. Principal Component Analysis

#### 3.4.1. The Effect of the Main Compounds of the Four Tested EOs and Their Acaricidal Activity

The first principal component analysis (PCA) allowed identifying the existence of correlations (positive and negative) between the major compounds of the four essential oils and the acaricidal activity against *E. orientalis* and *T. urticae* in 24 and 48 h. The correlations of the different parameters studied are shown in a correlation matrix (Table 1).

The loading plot (Figure 4(a)) and the correlation matrix (Table 3) revealed some correlation between the variables studied. A strong positive correlation ($>0.7$) exists between (a) α-pinene and LD$_{50}$ of *T. urticae*, (b) 1,8-cineole and LD$_{50}$ of *T. urticae*, and (c) Borneol and LD$_{50}$ of *T. urticae*. A medium positive correlation (0.6–0.5) exists between fenchone and LD$_{50}$ of *E. orientalis*. A strong negative correlation ($>-0.7$) exists between (a) dihydrocarvone and LD$_{50}$E. orientalis, (b) r(+)-pulegone and LD$_{50}$E. orientalis, and (c) carvone and LD$_{50}$ of *E. orientalis*. That means that when the amount of 1,8-cineole, α-pinene, and borneol compounds increased with increased LD$_{50}$ of *T. urticae*, and when the LD$_{50}$ of *E. orientalis* was low, the amount of dihydrocarvone, r(+)-pulegone, and carvone were important. The biplot (Figure 4(b)) showed that essential oils of *L. stoechas* and *R. officinalis* are dominated by the highest amount of 1,8-cineole, α-pinene, borneol, and fenchone compounds and they require a high concentration to eliminate *E. orientalis*. However, *O. compactum* essential oil is presented with a high amount of thymol and α-terpinene compounds. The *M. pulegium* was characterized by having the highest amount of r(+)-pulegone and carvone compounds and they require lowest concentration to eliminate *E. orientalis*.

Based on the result from Figures 4(a) and 4(b), it can be concluded that *M. pulegium* essential oil exhibited the best acaricide activity of the four essential oils. In fact, this can be explained by a high amount of r(+)-pulegone and carvone compounds in *M. pulegium* essential oil (74.03 and 5.45%, respectively) compared with *L. stoechas*, *R. officinalis*, and *O. compactum* (the absence of these compounds), but the contributions of others compounds should also be noted. The effect of synergy could also be the origin of this activity.
Table 2: LD50 values calculated for *Tetranychus urticae* and *Eutetranychus orientalis*.

| Spider mites species | Sample         | LD50 (µL/L air, 24 h, 95% CI) | Slope (24 h) | Intercept (24 h) | df  | X2  | LD50 (µL/L air, 48 h, 95% CI) | df  | X2  | Slope (48 h) | Intercept (48 h) |
|----------------------|----------------|-------------------------------|--------------|-----------------|-----|-----|--------------------------|-----|-----|--------------|-----------------|
|                      | *Tetranychidae urticae* |                    |               |                 |     |     |                           |     |     |              |                 |
|                      | *M. pulegium*    | 65.78 (44.63–10110.12)       | 1.69 ± 0.17  | 1.93 ± 0.38     | 4   | 4.21| 56.1 (33.8–69.5)          | 4   | 4.68| 1.86 ± 0.18  | 1.75 ± 0.41     |
|                      | *L. stoechas*    | 100.03 (85.88–120.12)        | 1.80 ± 0.24  | 1.41 ± 0.55     | 4   | 2.64| 85.35 (65.15–115.12)      | 4   | 2.89| 1.87 ± 0.27  | 1.38 ± 0.60     |
|                      | *R. officinalis* | 171.87 (163.24–184.96)       | 1.59 ± 0.41  | 1.44 ± 0.91     | 4   | 1.95| 141.51 (120.39–162.10)    | 4   | 2.28| 1.67 ± 0.48  | 1.40 ± 0.07     |
|                      | *O. compactum*   | 87.92 (68.15–99.65)          | 1.29 ± 0.27  | 2.50 ± 0.56     | 4   | 3.92| 79.72 (58.9–95.42)        | 4   | 4.10| 1.79 ± 0.33  | 1.27 ± 0.75     |
|                      | Chemical acaricide | 36.25 (27.77–52.48)         | 1.42 ± 0.37  | 2.79 ± 0.83     | 4   | 4.62| 32.09 (19.81–61.10)       | 4   | 4.9 | 1.72 ± 0.35  | 2.21 ± 0.78     |
|                      | *Eutetranychus orientalis* |            |               |                 |     |     |                           |     |     |              |                 |
|                      | *M. pulegium*    | 59.10 (40.5–78.9)            | 1.86 ± 0.15  | 1.71 ± 0.35     | 4   | 3.99| 47.37 (39.80–63.65)       | 4   | 4.51| 1.91 ± 0.19  | 1.80 ± 0.42     |
|                      | *L. stoechas*    | 120.87 (101.19–146.62)       | 1.79 ± 0.14  | 1.26 ± 0.31     | 4   | 1.98| 100.35 (85.98–121.72)     | 4   | 2.15| 1.75 ± 0.15  | 1.50 ± 0.33     |
|                      | *R. officinalis* | 88.2 (66.6–112.24)           | 1.76 ± 0.28  | 1.57 ± 0.63     | 4   | 3.10| 82.44 (70.06–109.2)       | 4   | 3.84| 1.89 ± 0.24  | 1.43 ± 0.54     |
|                      | *O. compactum*   | 112.77 (89.5–135.7)          | 2.36 ± 0.24  | 0.16 ± 0.54     | 4   | 2.92| 96.85 (81.45–140.16)      | 4   | 3.8 | 2.38 ± 0.27  | 0.28 ± 0.61     |
|                      | Chemical acaricide | 34.07 (25.10–50.23)         | 1.49 ± 0.29  | 2.72 ± 0.62     | 4   | 4.49| 29.15 (17.43–62.10)       | 4   | 4.95| 1.47 ± 0.27  | 2.84 ± 0.57     |
3.4.2. The Effect of the Concentrations of the Four Tested EOs and Their Acaricidal Activity. The second PCA permeated to identify the similar acaricidal activity effect of essential oil and concentration, the first principal component analysis was carried out. The individuals are represented by four essential oil samples (M. pulegium, L. stoechas, O. compactum, and R. officinalis) in five different concentrations (C1, C2, C3, C4, and C5). The variables are represented by acaricidal activity against E. orientalis and T. urticae in 24 and 48 h.

The biplot (Figure 5(a)) demonstrated that the distribution of individuals (concentration) in accordance with...
|          | α-pinene | Camphene | α-Terpinene | p-Cymene | 1,8-Cineole | γ-Terpinene | Fenchone | Camphor | Borneol | Dihydrocarvone | R(+)pulegone | Carvone | Thymol | Carvacrol | LD₅₀ E. orientalis | LD₅₀ T. urticae |
|----------|----------|----------|-------------|----------|------------|-------------|----------|----------|---------|---------------|-------------|---------|--------|-----------|---------------------|----------------|
| α-pinene | 1        | 0.39     | -0.28       | -0.28    | 0.99       | -0.28       | -0.21    | 0.4      | 0.92    | -0.5          | -0.5        | -0.51   | -0.28  | -0.28     | 0.02                | 0.99          |
| Camphene | 0.39     | 1        | -0.55       | -0.53    | 0.32       | -0.33       | 0.81     | 1        | 0.72    | -0.55         | -0.55       | -0.53  | -0.55  | -0.55     | 0.52                | 0.49          |
| α-Terpinene | -0.28  | -0.55    | 1           | 1        | -0.35      | 1           | -0.33    | -0.54   | -0.47   | -0.33         | -0.33       | -0.34  | 1      | 1         | 0.42                | -0.27         |
| p-Cymene | -0.28    | -0.53    | 1           | 1        | -0.36      | 1           | -0.31    | -0.53   | -0.46   | -0.35         | -0.35       | -0.36  | 1      | 1         | 0.44                | -0.27         |
| 1,8-Cineole | 0.99   | 0.32     | -0.35       | -0.36    | 1          | -0.36      | -0.29    | 0.33     | 0.88     | -0.35         | -0.35       | -0.36  | -0.35 | -0.35     | -0.14               | 0.96          |
| γ-Terpinene | -0.28  | -0.53    | 1           | 1        | -0.36      | 1           | -0.32    | -0.53   | -0.47   | -0.34         | -0.34       | -0.35  | 1      | 1         | 0.44                | -0.27         |
| Fenchone | -0.21    | 0.81     | -0.33       | -0.31    | -0.29      | -0.32      | 1        | 0.81     | 0.19     | -0.33         | -0.33       | -0.31  | -0.33 | -0.33     | 0.61                | -0.09         |
| Camphor | 0.4      | 1        | -0.54       | -0.53    | 0.33       | -0.53       | 0.81     | 1        | 0.73     | -0.55         | -0.55       | -0.53  | -0.54 | -0.54     | 0.52                | 0.5           |
| Borneol | 0.92     | 0.72     | -0.47       | -0.46    | 0.88       | -0.47       | 0.19     | 0.73     | 1        | -0.58         | -0.58       | -0.58  | -0.47 | -0.47     | 0.21                | 0.95          |
| Dihydrocarvone | -0.5   | -0.55    | -0.33       | -0.35    | -0.34      | -0.33      | -0.55   | -0.58   | 1        | 1             | 1           | -0.33  | -0.33 | -0.33     | -0.87               | -0.59         |
| R(+)pulegone | -0.5   | -0.55    | -0.33       | -0.35    | -0.34      | -0.33      | -0.55   | -0.58   | 1        | 1             | 1           | -0.33  | -0.33 | -0.33     | -0.87               | -0.59         |
| Carvone | -0.51    | -0.53    | -0.34       | -0.36    | -0.36      | -0.35      | -0.31   | -0.53   | -0.58   | 1             | 1           | 1      | -0.34 | -0.34     | -0.86               | -0.6          |
| Thymol | -0.28    | -0.55    | 1           | 1        | -0.35      | 1           | -0.33   | -0.54   | -0.47   | -0.33         | -0.33       | -0.34  | 1      | 1         | 0.42                | -0.27         |
| Carvacrol | -0.28   | -0.55    | 1           | 1        | -0.35      | 1           | -0.33   | -0.54   | -0.47   | -0.33         | -0.33       | -0.34  | 1      | 1         | 0.42                | -0.27         |
| LD₅₀ E. orientalis | 0.02  | 0.52     | 0.42        | 0.44     | 0.14       | 0.44       | 0.61     | 0.52     | 0.21     | -0.87         | -0.87       | -0.86  | 0.42  | 0.42      | 0.14                | 0.14          |
| LD₅₀ T. urticae | 0.99  | 0.49     | -0.27       | -0.27    | 0.96       | -0.27      | -0.09   | 0.5      | 0.95     | -0.59         | -0.59       | -0.6   | -0.27 | -0.27     | 0.14                | 1             |
PC1 and PC2 shows that individuals were separated into three main groups. These results indicated that the concentration has an effect on the acaricidal activity, as previously shown by ANOVA. The first group consisted of C1 (39.06 μl/ml) and C2 (78.125 μl/ml), or the low concentration. The second group is composed of C4 (312.5 μl/ml) and C5 (625 μl/ml), or the high concentration. The third groups were C3 (156.25 μl/ml), or the medium concentration. This graph indicated that when the concentration was high, the acaricidal activity against *E. orientalis* and *T. urticae* was elevated, and when the concentration was low, the acaricidal activity against *E. orientalis* and *T. urticae* was decreased.

The biplot (Figure 5(b)) showed that the acaricidal activity against *E. orientalis* and *T. urticae* improved when the *M. pulegium* essential oil was used. In contrast, the *L. stoechas* and *O. compactum* essential oils revealed a medium acaricidal activity against *T. urticae*. In addition, the *R. officinalis* showed a medium acaricidal activity against *E. orientalis*.

### 3.4.3. The Effect of the Four Mixtures Between Acaricide and Tested EOs and Their Acaricidal Activity.

The third PCA was enabled to determine the similar mixtures that have the same acaricidal activity between the four mixtures studied (*M. pulegium* EO + acaricide, *R. officinalis* EO + acaricide, *O. compactum* EO + acaricide, and *L. stoechas* EO + acaricide).

The biplot (Figure 6) illustrated that the individuals (mixtures) were divided into three main groups. The first group was the mixture of *M. pulegium* EO + acaricide and *R. officinalis* EO + acaricide. The second group was the mixture of *O. compactum* EO + acaricide. The third group was *L. stoechas* EO + acaricide. The mixture of (a) *M. pulegium* EO + acaricide and (b) *R. officinalis* EO + acaricide showed the same acaricidal activity, a medium effect against the *T. urticae* and a low effect against *E. orientalis*. This graph also indicated that the acaricidal activity has been improved with the mixture of *O. compactum* EO + acaricide (synergetic effect), while the mixture of *L. stoechas* EO + acaricide presented a low acaricidal activity against the *E. orientalis* and *T. urticae* (antagonistic effect).

### 3.5. The Hierarchical Cluster Analysis.

The hierarchical cluster analysis (HCA) allowed us to better classify the studied samples (the four essential oils) according to the
acaricidal activity against the *E. orientalis* and *T. urticae* (Figure 7). As a confirmation of the third PCA, the individuals were divided into three main clusters: cluster 1 (*M. pulegium* EO + acaricide and *R. officinalis* EO + acaricide) characterized by a high acaricidal activity against *T. urticae*, while it was characterized by a low acaricidal activity against the *E. orientalis*. However, cluster 2 (*O. compactum* EO + acaricide) was characterized by a high acaricidal activity against the *E. orientalis* and *T. urticae*. Cluster 3 (*L. stoechas* EO + acaricide) was featured by a low acaricidal activity against the *E. orientalis* and *T. urticae*. Both the PCA and HCA concluded that the acaricidal activity against the *E. orientalis* and *T. urticae* were significantly influenced by adding the acaricide to essential oils. A synergetic effect with *O. compactum* EO + acaricide and an antagonistic effect with *L. stoechas* EO + acaricide.

4. Discussion

In commercial orchards, the protection against pests is currently assured by intensive and preventive chemical products. Despite advantages such as rapid action in reducing the number of pests and their ease of use compared with natural plant extracts [20], this strategy has many limitations; the examination of the action spectrum of the active components used throughout the world reveals that 46% of acaricides are globally toxic toward auxiliary arthropods and public health [1, 28]. The extracts of plants are remarkably rich in toxins and inhibitors and can be the source of many insecticidal and acaricidal substances exploitable in the control of pests [29, 30]. On the contrary, the problem of volatility and the high cost causes several limitations of their usage [31]. The essential oils lose their force against environmental conditions. Nanoformulation is a method that may preserve the pesticide performance of plant essential oils [31, 32].

Phytochemical products are environmentally friendly, target-specificity, decreased the number of applications, higher acceptability, suitability for rural areas, low cost, biodegradable, easy preparation, and universally accepted. Botanicals are used as an alternative to synthetic acaricides and have been projected as a tool in the future for spider mites control agents, which are shown to function as highly polyphagous herbivore and primary agricultural pests in the world that provokes hard damage to crops.

In this study, *M. pulegium*, *L. stoechas*, *R. officinalis*, and *O. compactum* (Lamiaceae family) EOs showed an acaricidal potential toward adults of the two crop pests, *T. urticae* and *E. orientalis*. Indeed, the mortality varied according to the EO tested, the concentrations used, and the mite species.

*R. officinalis* was often effective against agricultural crop pests including tetranychid mites [22, 33]. The essential oil of *R. officinalis* and mixtures of its main constituents gave good results in the control of *T. urticae* Koch, on bean and tomato plants [34].

Thymol, a hydrophobic compound, constitutes the major monoterpene in *O. compactum*; it can penetrate the compartment membrane, which diminishes its impermeability, this mechanism stimulates the entry of other components into the cytoplasm [35, 36]. There is also evidence in
the literature that thymol and eugenol have a neurotoxic effect on arthropods, with different mechanisms of action. Thymol can bind to GABA receptors located in the membrane of post-synaptic neurons and disrupt the functioning of synapses [37] under laboratory conditions. A study shows that the essential oils of O. compactum is often toxic against Rhyzopertha dominica and Sitophilus oryzae Linnaeus, 1763 [38]. This was confirmed in our study against T. urticae and E. orientalis. O. compactum significantly inhibited oviposition of T. urticae; it was also toxic to females of this pest. Inhibition of fecundity exceeding 80% [39]. The essential oil of M. pulegium was very toxic against the strawberry spider mite Tetranychus urticae [19]. The same results in another study were observed on T. urticae, a serious pest of many agricultural crops. The phytotoxicity of M. pulegium was significant against this pest after 24 h [40, 41].

The statistical analysis (first PCA) permitted to conclude that the acaricidal activity depends on the major compounds of the essential oils tested. A strong correlation was identified between the LD$_{50}$ dihydrcarvone and carvone. Thus, when the quantity of dihydrocarvone and carvone increased, the acaricidal activity increased. Several studies reported that the principal compound present in the essential oils of thyme and oregano increases (thujone 65.78% and carvone 39.55%) the acaricidal activities against spider mite Tetranychus cinnabarinus Bois are more important [42]. In another study, 28 monoterpenes including monoterpenic hydrocarbons and oxygenated monoterpenes (camphor, 3-carene, carvone, 1,8-cineole, citronellal, β-citronellene, dihydrocarvone, and α-terpineol) of essential oils obtained from different plant species were tested against adults of Sitophilus zeamais Motschulsky under laboratory conditions. The results show that most of the monoterpenes with high concentrations have significantly insecticidal and acaricidal effects on the tested pests [43, 44].

The second PCA showed that M. pulegium was found to be more effective, followed by O. compactum EO and L. stoechas. EOs acquired from multiple plants have already been tested for their potential fumigant toward pests species (Choi et al., 2006) [42]. As an example, [45] examined the fumigant toxicity of L. stoechas EOs against three spider mites pests attacking stored products (Lasioderma serricorne Fabricius 1792, R. dominica Fabricius 1792, and Tribolium castaneum Herbst, 1797), the findings of this analysis showed that this EO had strong acaricidal properties with the high rates of mortality were obtained after 24 and 48 h of fumigation. L. stoechas was also tested for its acaricidal action against Tetranychus cinnabarinus Koch, 1836, one of the most important pests, the registered LC$_{50}$ and LC$_{90}$ values were in the range of 2.92 and 13.0 µg/ml, respectively [42]. Author [35] also recorded an effective acaricidal effect of L. stoechas against Orgyia trigotephras Boisduval, 1828 (Lepidoptera, Lymantriidae).

The third PCA and HCA illustrated that the activity of O. compactum EO has been ameliorated when it is mixed with the acaricide (synergetic effect). The R. officinalis and M. pulegium remain have the same activity when they were mixed with the acaricide. However, the activity of L. stoechas reduced when it was mixed with the acaricide (antagonistic effect). Several studies confirm that the mixture of several products often triggers an antagonistic effect or a synergistic effect and all depends on the obtained combination [39, 46, 47].

Moreover, the fumigant toxicity for EOs mixtures with the chemical acaricides revealed encouraging results where the mortality reached more than 95% for both spider mites. The search for synergistic mixtures aims to reduce the concentrations needed by increasing the biological activity against the target organism. This reduction of concentration may also result in decreased toxicological and environmental risks, and lower production cost [48, 49].

5. Conclusions

This scientific investigation highlighted the acaricidal effect of four essential oils against two important spider mites involved in agriculture damage. The obtained results proved the acaricidal potential of the tested oils and showed the efficacy of the binary mixtures with an acaricidal product reaching the mortality to 100%. These plant-based natural products can be applied in large field applications using low concentrations (based on the LD$_{50}$ values) and minimizing the production cost; they since could be involved in pest management programs as a new tool that could solve pest problems reducing risks to humans and the environment. Their inclusion in an integrated pest management program can contribute greatly to the achievement of the acaricide reduction objectives set by several countries and organizations. In this context, responsible agricultural organizations should actively support the development and implementation of essential integrated pest protection programs. Further studies are needed to test the toxicity of these natural products.

Data Availability

All related data are included within the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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References

[1] A. Assouguem, M. Kara, H. Mechchate et al., “Current situation of Tetranychus urticae (Acari: Tetranychidae) in northern Africa: the sustainable control methods and priorities for future research,” Sustainability, vol. 14, no. 4, p. 2395, 2022.

[2] M. Afzal, M. I. Ullah, M. H. Bashir, S. N. Mukhtar, M. Arshad, and N. Altarf, “Diversity and abundance of mite species in citrus orchards of sargodha, Pakistan,” Punjab University Journal of Zoology, vol. 36, no. 1, pp. 37–46, 2021.
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[3] F. Ferragut, D. Navia, and R. Ochoa, “New mite invasions in citrus in the early years of the 21st century,” Experimental & Applied Acarology, vol. 59, no. 1-2, pp. 145–164, 2013.

[4] A. Mazih, “Status of citrus IPM in the southern mediterranean basin Morocco, North Africa,” Acta Horticulturae, vol. 1065, pp. 1097–1103, 2015.

[5] M. C. Smaili, A. Boutaleb-Joutei, and A. Blenzar, “Beneficial insect community of Moroccan citrus groves: assessment of their potential to enhance biocontrol services,” Egypt. J. Biol. Pest Control, vol. 47, no. 1, 2020.

[6] A. Assouguem, A. Farah, R. Ullah et al., “Evaluation of the Varietal Impact of Two citrus Species on Fluctuations of Tetranychus urticae (Acari: Tetranychidae) and Beneficial Phytoseid Mites,” Sustainability, vol. 14, no. 5, p. 3088, 2022.

[7] A. Assouguem, M. Kara, H. Mechchate et al., “Evaluation of the impact of different management methods on Tetranychus urticae (Acari: Tetranychidae) and their predators in citrus orchards,” Plants, vol. 11, no. 5, p. 623, 2022.

[8] M. E. Sato, M. Z. da Silva, A. Raga, and M. F. d. Souza Filho, “de Abamectin resistance in Tetranychus urticae Koch (Acari: Tetranychidae): selection, cross-resistance and stability of resistance,” Neotropical Entomology, vol. 34, no. 6, pp. 991–998, 2005.

[9] W. Xue, S. Snoeck, C. Njiru, E. Inak, W. Dermauw, and W. Xue et al., “Geographical distribution and molecular insights into abamectin and milbemectin cross-resistance in European field populations of Tetranychus urticae,” Pest Management Science, vol. 76, no. 8, pp. 2569–2581, 2020.

[10] Smaili, “Mc Current pest status and the integrated,” 2021, https://scholar.google.com/scholar?hl=en&sa_d=0&2C58s=Smaili+MC%282017%29+Current+pest+status+and+the+integrated+pest+management+strategy+in+the+2 citrus+groves+in+Morocco.+In%3A+Abstracts+of+the+2C+disea.

[11] Z. Wang, T. Cang, S. Wu et al., “Screening for suitable chemical acaricides against two-spotted spider mites, Tetranychus urticae, on greenhouse strawberries in China,” Ecotoxicology and Environmental Safety, vol. 163, pp. 63–68, 2018.

[12] N. Icli and D. Tahmas Kahyao˘glu, “Investigation of pesticide residues in fresh sultani grapes and antioxidant properties of fresh/sun-dried/oven-dried grapes,” Turkish Journal of Agriculture and Forestry, vol. 44, no. 4, pp. 350–360, 2020.

[13] A. Takafuji, A. Ozawa, H. Nemoto, and T. Gotoh, “Spider mites of Japan: their biology and control,” Experimental & Applied Acarology, vol. 24, no. 5-6, pp. 319–335, 2000.

[14] D. Lynn, R. U. S. A. Data, and P. E. J. Robinson, “Miticidal and aphicidal method utilizing 2-higher alkyl-3-hydroxy-1,4-naphthoqui none carboxylc acd esters,” 1977, https://patents.google.com/patent/US4134517.

[15] S. Y. Salman and E. Sarıtas ¸, “Acequinocyl resistance in Tetranychus urticae Koch (Acari: Tetranychidae): inheritance, synergists, cross-resistance and biochemical resistance mechanisms,” International Journal of Acarology, vol. 40, pp. 428–435, 2014.

[16] Y. Koura, S. Kinoshita, K. Takasuka et al., “Respiratory inhibition of acaricide AKD-2032 and its deacetyl metabolite,” Journal of Pesticide Science, vol. 23, no. 1, pp. 18–21, 1998.

[17] J. S. Raut and S. M. Karuppayil, “A status review on the medicinal properties of essential oils,” Industrial Crops and Products, vol. 62, pp. 250–264, 2014.

[18] J. M. S. Faria and A. M. Rodrigues, “Essential oils as potential biopesticides in the control of the genus Meloidogyne: a review,” Proceedings, vol. 68, 2021.

[19] N. Zandi-Sohani and L. Ramezani, “Evaluation of five essential oils as botanical acaricides against the strawberry spider mite Tetranychus turkestani Ugurov and Nikolskii,” International Biodeterioration & Biodegradation, vol. 98, pp. 101–106, 2015.

[20] S. Atita, K. L. Grissa, G. Lognay, E. Bitume, T. Hance, and A. C. Mailleux, “A review of the major biological approaches to control the worldwide pest Tetranychus urticae (Acari: Tetranychidae) with special reference to natural pesticides: biological approaches to control Tetranychus urticae,” Journal of Pest Science, vol. 86, no. 3, pp. 361–386, 2013.

[21] M. A. Aziz, M. Adnan, A. H. Khan, A. A. Shahat, M. S. Al-Said, and R. Ullah, “Traditional uses of medicinal plants practiced by the indigenous communities at Mohmand Agency, FATA, Pakistan,” Journal of Ethnobiology and Ethnomedicine, vol. 14, no. 1, pp. 2–16, 2018.

[22] R. Ullah, A. S. Alqahtani, O. M. Noman, A. M. Alqahtani, S. Ibenmoussa, and M. Bourhia, “A review on ethno-medicinal plants used in traditional medicine in the Kingdom of Saudi Arabia,” Saudi Journal of Biological Sciences, vol. 27, no. 10, pp. 2706–2718, 2020.

[23] M. Mohammadosseini, A. Venditti, and B. Mahdavi, “Characterization of essential oils and volatiles from the aerial parts of Mentha pulegium L. (Lamiaceae) using microwave-assisted hydrodistillation (MAHD) and headspace solid phase microextraction (HS-SPME) in combination with GC-MS,” Natural Product Research, 2021.

[24] H. Hassanpour, R. A. Khavari-Nejad, V. Niknam, K. Razavi, and F. Najafi, “Effect of penconazole and drought stress on the essential oil composition and gene expression of Mentha pulegium L. (Lamiaceae) at flowering stage,” Acta Physiologica Plantarum, vol. 365, no. 5, pp. 1167–1175, 2014.

[25] M. Hanana, B. Mansour, M. Algrab et al., “Potential use of essential oils from four Tunisian species of Lamiaceae: biological alternative for fungal and weed control,” Natural Product Research, vol. 11, pp. 258–269, 2017.

[26] M. Bourhia, F. E. Laari, H. Aourik et al., “Antioxidant and antiproliferative activities of bioactive compounds contained in Rosmarinus officinalis used in the Mediterranean diet,” Evidence-based Complementary and Alternative Medicine, vol. 2019, Article ID 7623830, 7 pages, 2019.

[27] A. Assouguem, M. Kara, I. Mansouri et al., “Evaluation of the effectiveness of spirotetramat on the diaspine scale parlatoria pergandii in citrus orchards,” Agronomy, vol. 11, no. 8, pp. 1562–1611, 2021.

[28] C. Vicente, A. Boutaleb, and P. Lebrun, “Quelles stratégies de lutte contre les acariens ravageurs du pommier au Maroc,” Parasitica, vol. 59, pp. 25–41, 2003.

[29] M. B. Isman, A. J. Wan, and C. M. Passreiter, “Insecticidal activity of essential oils to the tobacco cutworm, Spodoptera litura,” Fitoterapia, vol. 72, no. 1, pp. 65–68, 2001.

[30] S. A. M. Abdelgalel, M. E. I. Badawy, N. F. Mahmoud, and A. E. S. M. Marei, “Acaricidal activity, biochemical effects and molecular docking of some monoterpens against two-spotted spider mite (Tetranychus urticae Koch),” Pesticide Biochemistry and Physiology, vol. 156, pp. 105–115, 2019.

[31] M. Negahban, S. Moharramipour, M. Zandi, and S. A. Hashemi, “Efficiency of nanoencapsulated essential oil of Artemisia sieberi Besser on nutritional indices of Plutella xylostella,” 2022, https://www.cabdirect.org/cabdirect/abstract/2014337342.

[32] A. Taban, M. J. Saharkhiz, and M. Khorram, “Formulation and assessment of nano-encapsulated bioherbicides based on
biopolymers and essential oil,” *Industrial Crops and Products*, vol. 149, Article ID 112348, 2020.

[33] H. Jahanian, N. Kahkeshani, A. Sanei-Dehkordi, M. B. Isman, M. Saeedi, and M. Khanavi, “secticide Rosmarinus officinalis as a Natural Inc: A Review,” *International Journal of Pest Management*, 2022.

[34] S. Miresmaili, R. Bradbury, and M. B. Isman, “Comparative toxicity of Rosmarinus officinalis L. essential oil and blends of its major constituents against Tetranychus urticae Koch (Acar: Tetranychidae) on two different host plants,” *Pest Management Science*, vol. 62, no. 4, pp. 366–371, 2006.

[35] H. Akkari, O. Ezzine, S. Dhahri et al., “Chemical composition, insecticidal and in vitro anthelmintic activities of Ruta chalepensis (Rutaceae) essential oil,” *Industrial Crops and Products*, vol. 74, pp. 745–751, 2015.

[36] M. Ahmad, S. Benjakul, T. Prodpran, and T. W. Agustini, “Physico-mechanical and antimicrobial properties of gelatin film from the skin of unicorn leatherjacket incorporated with essential oils,” *Food Hydrocolloids*, vol. 28, no. 1, pp. 189–199, 2012.

[37] C. M. Priestley, E. M. Williamson, K. A. Wafford, and D. B. Sattelle, “Thymol, a constituent of thyme essential oil, is a positive allosteric modulator of human GABAA receptors and a homo-oligomeric GABA receptor from *Drosophila melanogaster*,” *British Journal of Pharmacology*, vol. 140, no. 8, pp. 1363–1372, 2003.

[38] S. Bounoua-Fraoucene, A. Kellouche, and J. F. Debras, “Toxicity of four essential oils against two insect pests of stored grains, *Rhyzopertha Dominica* (Coleoptera: bostrochidae) and *Sitophilus oryzae* (Coleoptera: Curculionidae),” *African Entomology*, vol. 27, no. 2, pp. 344–359, 2019.

[39] A. Ramzi, A. El Ouali Lalami, Y. Ez zoubi et al., “Insecticidal effect of wild-grown Mentha pulegium and *Rosmarinus officinalis* essential oils and their main monoterpenes against *Culex pipiens* (Diptera: Culicidae),” *Plants*, vol. 11, no. 9, p. 1193, 2022.

[40] F. Mozaffari, H. Abbasipour, A. S. Garjan, A. R. Saboori, and M. Mahmoudvand, “Various effects of ethanolic extract of Mentha pulegium on the two-spotted spider mite, Tetranychus urticae (Tetranychidae),” *Archives of Phytopathology and Plant Protection*, vol. 45, pp. 1347–1355, 2012.

[41] F. Mozaffari, H. Abbasipour, A. S. Garjan, A. Saboori, and M. Mahmoudvand, “Toxicity and Oviposition Deterrence and Repellency of Mentha pulegium (Lamiacaeae) Essential Oils against Tetranychus urticae Koch (Teteranychidae),” *Journal of essential oil-bearing plants*, vol. 16, pp. 575–581, 2013.

[42] E. Sertkaya, K. Kaya, and S. Soylu, “Acaricidal activities of the essential oils from several medicinal plants against the carmine spider mite (*Tetranychus cinnabarinus* Boisd.) (Acar: Tetranychidae),” *Industrial Crops and Products*, vol. 31, no. 1, pp. 107–112, 2010.

[43] E. Yildirim, B. Emsen, and S. Kordali, “Insecticidal effects of monoterpenes on *Sitophilus zeamais* Motschulsky (Coleo- pertera: Curculionidae),” *Journal of Applied Botany and Food Quality*, vol. 86, pp. 198–204, 2013.

[44] X. Yang, H. Han, B. Li, D. Zhang, Z. Zhang, and Y. Xie, “Fumigant toxicity and physiological effects of spearmint (*Mentha spicata*, Lamiaceae) essential oil and its major constituents against Reticulitermes dabieshanensis,” *Industrial Crops and Products*, vol. 171, Article ID 113894, 2021.

[45] A. Ebadollahi, M. Safaralizadeh, and A. Pourmirza, “Fumigant toxicity of lavandula stoechas L. Oil against three insect pests attacking stored products,” *Journal of Plant Protection Research*, vol. 50, no. 1, pp. 56–60, 2010.

[46] S. Shezryna, N. Anisah, I. Saleh, and R. A. Syamsa, “Acaricidal activity of the essential oils from *citrus hystrix* (Rutaceae) and *cymbopogon citratus* (poaceae) on the cattle tick *rhipicephalus* (boophilus) microplus larvae (acari: ixodidae),” *Tropical Biomedicine*, vol. 37, no. 2, pp. 433–442, 2020.

[47] N. K. Singh, H. Singh, N. Mehta, and S. S. Rath, “In vitro assessment of synergistic combinations of essential oils against *Rhipicephalus* (Boophilus) microplus (Acari: Ixodi- dae),” *Experimental Parasitology*, vol. 201, pp. 42–48, 2019.

[48] O. Belhoussaine, C. El Kourchi, H. Harhar et al., “Chemical composition, antioxidant, insecticidal activity, and comparative analysis of essential oils of leaves and fruits of *schinus molle* and *schinus terebinthifolius*,” *Evidence-based Complementary and Alternative Medicine*, vol. 2022, Article ID 428890, 12 pages, 2022.

[49] F. T. Maestre, J. L. Quero, N. J. Gotelli et al. “Plant species richness and ecosystem multifunctionality in global dry- lands,” *Science*, vol. 335, pp. 214–218, 2012.