Microbiology of bile aspirates obtained at ERCP in patients with suspected acute cholangitis

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ABSTRACT
Background The cornerstone of treatment for acute cholangitis is source control with biliary drainage and early antibiotics. The primary aim of this study was to describe the microbiology of bile aspirate pathogens obtained at the time of endoscopic retrograde cholangiopancreatography (ERCP) in patients suspected of having acute cholangitis.

Methods In this single-center retrospective study, patients were included if a bile aspirate was collected at ERCP for suspicion of acute cholangitis, from 1 January 2010 to 31 December 2016.

Results There were 721 ERCP procedures for suspected acute cholangitis with bile culture results, with 662 positive bile cultures (91.8%). Pathogens included: Enterococcus species (spp.) 448 (67.7%); Klebsiella spp. 295 (44.6%); Escherichia coli 269 (40.6%); Pseudomonas spp. 52 (7.9%); and anaerobes 64 (9.7%). Susceptibility of Klebsiella pneumoniae and E.coli isolates to ciprofloxacin was 88% and 64%, respectively. Extended-spectrum beta-lactamases and carbapenem resistance were found in 7.9% and 3.6% of Enterobacteriaceae, respectively. There were 437 concurrent blood cultures, of which 174 were positive (39.8% of cultures drawn). Prior biliary endoscopic sphincterotomy (ES) was evident in 459 ERCP cases (63.7%), and was associated with increased frequency of Klebsiella spp., Pseudomonas aeruginosa, Enterobacter spp., and Enterococcus spp. Prior biliary ES significantly increased the probability of vancomycin-resistant Enterococcus (VRE).

Conclusions The vast majority of bile cultures (91.8%) were positive. The susceptibilities of E.coli and K.pneumoniae to ciprofloxacin are lower than historically noted. A notable portion of cultures contained pathogenic drug-resistant organisms. Prior biliary ES is associated with a higher frequency of certain organisms and higher frequency of VRE.
Introduction

Acute cholangitis is a frequently encountered clinical entity that can have potentially fatal consequences if not detected and treated promptly. There is a high risk of end-organ damage, leading to a significant mortality rate if treatment is not instituted promptly [1], which drops to <5% with timely and appropriate interventions [2]. Treatment for acute cholangitis is incumbent on prompt and adequate hemodynamic resuscitation [3], and initiation of appropriate antimicrobial therapy to limit the local and systemic inflammatory response to sepsis and to contain the spread of infection [3], followed by the achievement of definitive source control by decompressing the biliary tree to relieve biliary obstruction [4]. Biliary decompression is most often achieved by techniques applied at endoscopic retrograde cholangiopancreatography (ERCP). Multiple studies support superior outcomes and improved mortality with ERCP compared with interventional radiology or surgical modalities [5–11]. The selection of appropriate empiric antimicrobial agents in acute cholangitis is an important cornerstone of medical decision-making that is frequently encountered in clinical care.

Our group has previously reported the microbiology of bile cultures aspirated at the time of ERCP in patients with acute cholangitis and cholestasis [12]. In our prior study, the bile aspirates from 160 patients from 1994 to 2000 were analyzed. In the bile aspirates, *Escherichia coli*, *Enterococcus* spp., *Streptococcus* spp., and *Klebsiella pneumoniae* were most commonly isolated. Polymicrobial infections and *Enterococcus* infections were more frequently identified in patients with previously placed plastic biliary stents. Based on susceptibility data, ciprofloxacin and ceftriaxone were effective against Gram-negative bacilli in 96% and 91% of cases, respectively. This has supported the use of these antibiotics for routine empiric antibiotic coverage in patients with suspected acute cholangitis.

With widespread use of antibiotics and the global emergence of multidrug-resistant organisms, efforts are currently underway to identify microbiological characteristics and recognize patterns of drug resistance related to intra-abdominal infections to help in directing prudent antimicrobial choice, early de-escalation to pathogen-directed therapy, and appropriate termination of therapy. This has prompted prior studies and new recommendations from medical societies reinforcing these principles [2, 13–16].

The primary aim of this study was to describe the microbiology patterns of bile aspirates obtained at the time of ERCP in patients suspected of having acute cholangitis. Secondary aims included: (i) the identification of factors that affect the microbiology patterns, such as the presence/absence of a biliary sphincterotomy, presence/absence of a biliary stent; and (ii) a description of the presence of microbiological resistance among the obtained biliary organisms.

Methods

This was a single-center retrospective study. The study was approved by the Institutional Review Board (IRB) of Indiana University School of Medicine (study #1607577831). Patients were included in this study if a bile aspirate was collected at the time of ERCP for clinical suspicion of acute cholangitis, from 1 January 2010 to 31 December 2016. ERCP procedures were carried out at IU Health University Hospital in Indianapolis, Indiana, a large tertiary care teaching hospital. An IRB-approved, prospectively maintained institutional ERCP database and a separate microbiology database were queried to identify patients. The electronic medical record (Cerner, Kansas City, Missouri, USA) and electronic procedure reporting system (ProVation, Minneapolis, Minnesota, USA) were reviewed for the identified patients. Patient demographics, procedure-specific data, and microbial data were collected for each patient and each biliary aspirate.

At our center, bile aspirates are obtained at the time of ERCP at the discretion of the endoscopist, most often in the setting of clinically suspected acute cholangitis. They are routinely drawn for this indication by all endoscopists performing ERCP in our hospital. Very rarely, if the bile is too thick to aspirate, there are concurrent data available that may make bile culture redundant (e.g. a blood culture is already positive for a biliary organism in a patient with pus coming from bile duct); or the endoscopist forgets to obtain the bile specimen, bile may not be aspirated at the time of the ERCP.

For this study, clinical acute cholangitis was defined as the presence of a cholestatic biochemical test profile accompanied by fever, with or without biliary dilatation on imaging, severe abdominal pain, or leukocytosis. Furthermore, patients noted to have purulent bile (milky-white discharge exiting the biliary orifice) at the time of ERCP met the criteria for clinical acute cholangitis. For inpatients undergoing ERCP for suspected acute cholangitis, blood for two sets of aerobic and anaerobic blood culture bottles was routinely drawn within 48 hours of admission (at the clinical discretion of the admitting physician).

Generally, all patients undergoing ERCP for suspected cholangitis have received antibiotics (either as an inpatient, in the emergency room, or pre-procedurally in the ERCP suite). If the patient is an inpatient, their inpatient antibiotic regimen is continued perioperatively (this is usually intravenous cefepime and metronidazole, or intravenous piperacillin–tazobactam). If the patient is an outpatient, peri-procedural antibiotics are given at the discretion of the endoscopist and most often include intravenous cefazolin or ciprofloxacin, depending on the clinical scenario and any medication allergies the patient may have.

The duodenoscope reprocessing techniques during the time period of the study were consistent, with the exception of a more substantial pre-cleaning step and a change to double high level disinfection (DHLD) in May 2015 for all duodenoscopes used at ERCP [17] (Appendix 1s, see online-only Supplementary material). DHLD is one of the four supplementary measures recommended by the US Food and Drug Administration (FDA) for the reprocessing of reusable duodenoscopes [18, 19].

Our techniques for collecting, processing, and reporting biliary aspirates during ERCP have been previously described [12]. Our techniques for collecting, processing, and reporting blood cultures during the period of this study are largely unchanged from our previously reported study [12], except for a few mod-
ifications. When blood cultures were obtained, they were obtained from patients who were admitted as inpatients. Our techniques for collection and processing of bile cultures and blood cultures are described in Appendix 2s.

Statistical analyses
Data analysis was performed using STATA13 (StataCorp., LLC, College Station, Texas, USA), SAS9.4 (SAS Inc., Cary, North Carolina, USA), and Excel (Microsoft Corporation, Redmond, Washington, USA). Descriptive data were reported as median and interquartile range (IQR) for continuous variables. Student’s t-test was used for the comparison of continuous variables. Categorical variables were described using frequency and proportion; the 95 %CI of a proportion obtained using the Clopper–Pearson method was also reported. Chi-squared tests or Fisher’s exact tests were used for comparisons of categorical data.

A crude odds ratio (OR) and its 95 %CI were calculated to compare the relative odds of the occurrence of a particular organism in the bile culture, with relation to prior biliary endoscopic sphincterotomy (ES) and indwelling transpapillary biliary stent. Hochberg’s step-up Bonferroni method was used to adjust for multiple tests performed for multiple organisms in the same encounter (40.5 % [95 %CI35.6 %–45.5 %] vs. 32.6 % [95 % CI19.1 %–48.5 %]). The most common organism isolated on blood culture was E.coli (49/174; 28.2 %).

Of the 721 ERCPs with bile aspirates, the cultures of the bile aspirates were positive in 662 ERCPs (91.8 %), with 403 having been performed on inpatients and 259 on outpatients. Of the negative bile cultures, 19 procedures were performed on inpatients and 40 on outpatients. The characteristics of the patients with suspected acute cholangitis who underwent ERCP with bile aspiration for culture are presented in Table 1. Patients who had a positive bile culture were significantly older (median 65 vs. 57 years) and were significantly more likely to have purulent bile or pus exiting the bile duct at the time of ERCP, as described by the endoscopist (44.7 % vs. 18.6 %).

Among the 721 cases in which a bile aspirate was obtained at the time of ERCP, 437 blood cultures were also obtained within 48 hours of presentation, of which 174 (39.8 %) were positive. Of the cases with positive bile cultures who also had blood drawn for cultures, the corresponding blood cultures grew bacterial isolates in 40.6 % of instances (160/394). The microbiological characteristics of the blood cultures obtained are detailed in Table 2. There was no significant difference in the proportion of patients with positive blood cultures based on whether the corresponding bile culture was positive or negative at the same encounter (40.5 % [95 %CI35.6 %–45.5 %] vs. 32.6 % [95 % CI19.1 %–48.5 %]). The most common organism isolated on blood culture was E.coli (49/174; 28.2 %).

Of the 662 positive bile cultures, 81.6 % were polymicrobial. The following pathogens were identified (as a percentage of total positive bile cultures aspirated; therefore, owing to polymicrobial cultures, percentages add up to >100 %): Enterococcus spp.44 (67.7 %); Klebsiella spp. 295 (44.6 %); E.coli 269 (40.6 %); viridans group streptococci 235 (35.5 %); Candida spp. 189 (28.5 %); Pseudomonas spp. 52 (7.9 %); anaerobes 64 (9.7 %); Clostridium spp. 53 and Bacteroides spp. 11); and Staphylococcus aureus 32 (4.8 %). Select antimicrobial susceptibilities of the Gram-negative organisms isolated from bile cultures are presented in Table 3. Furthermore, SPICE organisms (Serratia spp., Pseudomonas spp., Indole-positive Proteus spp., Citrobac-

### Table 1 Characteristics of patients with suspected acute cholangitis undergoing endoscopic retrograde cholangiopancreatography (ERCP) with bile aspiration for culture.

| Total ERCPs (n=721) | ERCPs with positive bile culture (n=662) | ERCPs with negative bile culture (n=59) | P value |
|---------------------|----------------------------------------|---------------------------------------|---------|
| **Age, median (IQR), years** | 65 (21.0) | 65 (20.0) | 57 (34.5) | <0.001 |
| **Sex, female, n (%)** | 290 (47.1) | 263 (39.7) | 27 (45.8) | 0.36 |
| **Inpatient, n (%)** | 422 (58.5) | 403 (60.9) | 19 (32.2) | <0.001 |
| **Choleclochothlithiasis, n (%)** | 431 (59.8) | 402 (60.7) | 29 (49.2) | 0.41 |
| **Malignant stricture, n (%)** | 232 (32.2) | 226 (34.1) | 6 (10.2) | 0.002 |
| **Benign stricture, n (%)** | 123 (17) | 118 (17.8) | 5 (8.5) | 0.13 |
| **Primary sclerosing cholangitis, n (%)** | 52 (7.2) | 49 (7.4) | 3 (5.1) | 0.79 |
| **Pus exiting bile duct at time of ERCP, n (%)** | 307 (42.5) | 296 (44.7) | 11 (18.6) | 0.006 |

IQR, interquartile range.
Enterobacteriaceae isolates, extended-spectrum beta-lactamase (ESBL)-producing organisms were detected in 7.9% of isolates and carbapenem-resistant Enterobacteriaceae (CRE) in 3.6%. Of S. aureus isolates 50% were methicillin-resistant (MRSA), and vancomycin resistance (VRE) was detected in 14.7% of Enterococcus spp. isolates.

Of the 721 total ERCPs, 459 (63.7%) had a prior biliary ES and 341 (47.3%) had a prior biliary stent. The influence of a prior biliary ES or an indwelling biliary stent on bile cultures are detailed in ▶Table 4. Bile cultures were significantly more likely to be positive when obtained in the presence of a prior biliary ES (96.1% [95% CI 93.9%–97.7%] vs. 84.4% [95% CI 79.4%–88.5%]; OR 4.5). A prior biliary ES significantly increased the probability of VRE in the bile culture (11.5% [95% CI 8.8%–14.8%] vs. 5.0% [95% CI 2.7%–8.3%]; OR 2.5). Certain bacteria were

| Table 2 | Microbiological characteristics of blood cultures for patients with positive or negative bile cultures. |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
|                | Total blood cultures (n = 437) | Blood cultures with positive bile culture (n = 395) | Blood cultures with negative bile culture (n = 43) | P value1 | Adjusted P value2 |
| n | n | % (95% CI) | n | % (95% CI) | n | % (95% CI) | n | (95% CI) |
| Positive blood culture | 174 | 160 | 40.5 (35.6–45.5) | 14 | 32.6 (19.1–48.5) | 0.31 | > 0.99 |
| Escherichia coli | 49 | 43 | 10.9 (8.0–14.4) | 6 | 14.0 (5.3–27.9) | 0.61 | > 0.99 |
| Klebsiella spp. | 36 | 36 | 9.1 (6.5–12.4) | 0 | 0 (0–9.2) | 0.04 | 0.26 |
| Enterococcus spp. | 15 | 14 | 3.5 (2.0–5.9) | 1 | 2.3 (0.1–12.3) | > 0.99 | > 0.99 |
| Enterobacter spp. | 10 | 9 | 2.3 (1.0–4.3) | 1 | 2.3 (0.1–12.3) | > 0.99 | > 0.99 |
| Gram-negative rods, NOS | 25 | 21 | 5.3 (3.3–8.0) | 4 | 9.3 (2.6–22.1) | 0.29 | > 0.99 |
| Other organisms | 39 | 37 | 9.4 (6.7–12.7) | 2 | 4.7 (0.6–15.8) | 0.41 | > 0.99 |

spp., species; NOS, not otherwise specified.
1 P value from chi-squared test or Fisher’s exact test.
2 Adjusted P value from Hochberg’s step-up Bonferroni method.
3 CIs obtained using the Clopper-Pearson method.

| Table 3 | Antimicrobial susceptibilities of isolated Gram-negative bacilli from bile cultures. |
|-----------------|-----------------|-----------------|-----------------|-----------------|
|                | Ampicillin1 | TMP/SMX | Ceftriaxone | Ciprofloxacin |
| Klebsiella pneumoniae | 183/201 (91.0%) | 192/201 (95.5%) | 176/201 (87.6%) |
| Klebsiella oxytoca | 94/94 (100%) | 94/94 (100%) | 94/94 (100%) |
| Escherichia coli | 145/269 (53.9%) | 213/269 (79.2%) | 238/269 (88.5%) | 173/269 (64.3%) |
| Citrobacter freundii complex | 16/17 (94.1%) | 15/17 (88.2%) | 17/17 (100%) |
| Citrobacter freundii | 11/12 (91.7%) | 9/12 (75.0%) | 11/12 (91.7%) |
| Other Citrobacter spp. | 15/16 (93.8%) | 15/16 (93.8%) | 15/16 (93.8%) |
| Pseudomonas aeruginosa | 0/52 (0%) | 0/52 (0%) | 24/52 (46.2%) |
| Enterobacter cloacae | 52/57 (91.2%) | 49/57 (86.0%) | 46/57 (80.7%) |
| Enterobacter cloacae complex | 27/28 (96.4%) | 25/28 (89.3%) | 27/28 (96.4%) |
| Other Enterobacter spp. | 22/23 (95.7%) | 22/23 (95.7%) | 21/23 (91.3%) |
| Aeromonas spp. | 11/13 (84.6%) | 13/13 (100%) | 13/13 (100%) |
| Other Gram-negative rods, NOS | 42/48 (87.5%) | 45/48 (93.8%) | 44/48 (91.7%) |

TMP/SMX, trimethoprim sulfamethoxazole; spp., species; NOS, not otherwise specified.
1 Sensitivities to ampicillin were not routinely reported for Klebsiella spp. and other species because of the high prevalence of resistance.
more common in the presence of a prior biliary ES: Enterococcus spp. (77.1 % [95%CI73.0%–80.9%] vs. 35.9 % [95%CI30.1%–42.0%]; OR6.0); Enterobacter spp. (19.2 % [95%CI15.7%–23.1%] vs. 7.6 % [95%CI4.7%–11.5%]; OR2.9); Pseudomonas aeruginosa (10.0 % [95%CI7.4%–13.1%] vs. 2.3 % [95%CI0.8%–4.9%; OR4.8); and Klebsiella spp. (47.1 % [95%CI42.4%–51.7%] vs. 30.2 % [95%CI24.7%–36.1%]; OR2.1).

Likewise, the presence of a biliary stent significantly increased the probability of VRE (13.5 % [95%CI10.0%–17.6%] vs. 5.3 % [95%CI3.2%–8.0%]; OR2.8). A similar effect regarding the increased presence of certain organisms was seen in the presence of a prior biliary stent: Enterococcus spp. 86.8 % (95%CI82.7%–90.2%) vs. 40.0 % (95%CI35.0%–45.1%; OR9.9; Enterobacter spp. 23.5 % (95%CI19.1%–28.3%) vs. 7.4 % (95%CI5.0%–10.5%; OR3.9); P aeruginosa 12.3 % (95%CI9.0%–16.3%) vs. 2.6 % (95%CI1.3%–4.8%; OR5.2); Klebsiella spp. 53.4 % (95%CI47.9%–58.8%) vs. 29.7 % (95%CI25.2%–34.6%; OR2.7). There was no effect on the incidence of E.coli with regard to the presence of a biliary stent or anaerobes with regard to the presence of biliary ES or a biliary stent.

Table 4  Associations of prior biliary endoscopic sphincterotomy and indwelling transpapillary biliary stent with bile culture.

| Biliary endoscopic sphincterotomy | Biliary stent |
|-----------------------------------|--------------|
| Positive bile culture             |              |
| Yes (n=459) [95%CI]               | Yes (n=341) [95%CI] |
| [93.9%–97.7%]                     | [96.2%–99.4%] |
| No (n=262) [95%CI]                | No (n=380) [95%CI] |
| [79.4%–88.5%]                     | [82.2%–89.4%] |
| Crude odds ratio [95%CI]          | Crude odds ratio [95%CI] |
| 4.5 [2.6–8.1]                     | 9.0 [3.8–21.3] |
| Adjusted Pvalue¹                  | Adjusted Pvalue¹ |
| <0.001                            | <0.001 |

| Multidrug resistant organisms     |              |
|-----------------------------------|--------------|
| ESBL                              |             |
| Yes (n=380) [95%CI]               | No (n=380) [95%CI] |
| [4.6–9.4]                         | [3.0–8.8] |
| 1.3 [0.7–2.5]                     | 1.6 [0.9–2.9] |
| Adjusted Pvalue¹                  |              |
| 0.94                              |              |
| MRSA                              |             |
| Yes (n=31) [95%CI]                | No (n=380) [95%CI] |
| [1.4–4.5]                         | [0.6–3.4] |
| 1.7 [0.6–5.4]                     | 1.9 [0.7–5.2] |
| Adjusted Pvalue¹                  |              |
| 0.94                              |              |
| VRE                               |             |
| Yes (n=53) [95%CI]                | No (n=380) [95%CI] |
| [8.8–14.8]                        | [4.6–10.0] |
| 2.5 [1.3–4.7]                     | 2.8 [1.6–4.9] |
| Adjusted Pvalue¹                  |              |
| 0.002                             |              |
| CRE                               |             |
| Yes (n=22) [95%CI]                | No (n=380) [95%CI] |
| [3.0–7.2]                         | [3.6–8.9] |
| 2.1 [0.9–5.4]                     | 2.9 [1.3–6.7] |
| Adjusted Pvalue¹                  |              |
| 0.09                              |              |

| Organisms                         |              |
|-----------------------------------|--------------|
| Klebsiella spp.                   |             |
| Yes (n=216) [95%CI]               | No (n=380) [95%CI] |
| [42.4–51.7]                       | [82.2%–89.4%] |
| 2.1 [1.5–2.8]                     | 10 [2.6–10.5] |
| Adjusted Pvalue¹                  |              |
| <0.001                            | <0.001 |
| Escherichia coli                  |             |
| Yes (n=174) [95%CI]               | No (n=380) [95%CI] |
| [33.5–42.3]                       | [3.0–14.4] |
| 1.1 [0.8–1.5]                     | 1.4 [1.0–1.9] |
| Adjusted Pvalue¹                  |              |
| 0.17                              |              |
| Citrobacter spp.                  |             |
| Yes (n=31) [95%CI]                | No (n=380) [95%CI] |
| [4.6–9.4]                         | [3.0–8.8] |
| 1.3 [0.7–2.5]                     | 2.6 [1.3–5.1] |
| Adjusted Pvalue¹                  |              |
| 0.94                              |              |
| Pseudomonas aeruginosa            |             |
| Yes (n=46) [95%CI]                | No (n=380) [95%CI] |
| [7.4–13.1]                        | [3.0–14.4] |
| 4.8 [2.0–11.3]                    | 5.2 [2.6–10.5] |
| Adjusted Pvalue¹                  |              |
| <0.001                            | <0.001 |
| Enterobacter spp.                 |             |
| Yes (n=88) [95%CI]                | No (n=380) [95%CI] |
| [15.7–23.1]                       | [5.0–10.5] |
| 2.9 [1.7–4.8]                     | 3.9 [2.4–6.1] |
| Adjusted Pvalue¹                  |              |
| <0.001                            | <0.001 |
| Enterococcus spp.                 |             |
| Yes (n=354) [95%CI]               | No (n=380) [95%CI] |
| [73.0–80.9]                       | [28.2–70.9] |
| 6.0 [4.3–8.4]                     | 9.9 [6.8–14.4] |
| Adjusted Pvalue¹                  |              |
| <0.001                            | <0.001 |
| Aeromonas spp.                    |             |
| Yes (n=10) [95%CI]                | No (n=380) [95%CI] |
| [1.0–4.0]                         | [3.0–8.8] |
| 1.9 [0.5–7.0]                     | 3.8 [1.0–13.9] |
| Adjusted Pvalue¹                  |              |
| 0.17                              |              |
| Other Gram-negative rods, NOS     |             |
| Yes (n=47) [95%CI]                | No (n=380) [95%CI] |
| [7.6–13.4]                        | [3.0–8.8] |
| 2.6 [1.3–5.1]                     | 3.2 [1.8–5.8] |
| Adjusted Pvalue¹                  |              |
| 0.001                             |              |
| Anaerobes                         |             |
| Yes (n=35) [95%CI]                | No (n=380) [95%CI] |
| [5.4–10.4]                        | [6.0–12.3] |
| 0.7 [0.4–1.1]                     | 1.0 [0.6–1.6] |
| Adjusted Pvalue¹                  |              |
| 0.94                              |              |

ESBL, extended spectrum beta-lactamase-producing Enterobacteriaceae; MRSA, methicillin-resistant Staphylococcus aureus; VRE, vancomycin-resistant Enterococcus; CRE, carbapenem-resistant Enterobacteriaceae; spp., species; NOS, not otherwise specified.

¹ Adjusted Pvalue from Hochberg’s step-up Bonferroni method.
Discussion

This single-center retrospective study evaluated the microbiology of biliary aspirates obtained from 721 ERCP procedures in patients with suspected acute cholangitis. Positive bile cultures were found in nearly 92% of these ERCPs. In general, duodenoscope cleaning techniques, culture appropriation techniques, and processing have remained predominantly consistent, with only minor variations [17] from the time of a prior study on this topic published by our institution in 2002 [12].

This study covered a 7-year time period between 2010 and 2016, similar to the previously published bile culture data from our institution for a comparable group of patients covering a 7-year time period between 1994 and 2000. The current study however contains over four times more ERCPs with bile aspiration performed than in the prior study (721 ERCP procedures compared with 180 ERCP procedures), owing to near universal adoption of the practice by all gastroenterologists performing ERCP for this indication at our institution.

The current study confirms and expands upon the prior data that there is an increased incidence in polymicrobial bile cultures, with an increased likelihood of Enterococcus spp. in the setting of an indwelling biliary stent [12]. We also found a significantly higher likelihood of Enterobacter spp. (23.5% vs. 7.4%), P. aeruginosa (12.3% vs. 2.6%), and K. pneumoniae (53.4% vs. 29.7%) in the presence of a biliary stent, which was not appreciated in the prior study. We suspect this association was uncovered in this study because of the much larger sample size and expanded culturing of bile in nearly all patients suspected of having acute cholangitis.

The current study also realized a higher positive bile culture rate in the patients who had not had a prior biliary stent. We found that bile cultures were positive in 86.1% of patients who had previously not had a biliary stent, whereas only 55% of patients in the prior study had a positive bile culture without a prior biliary stent. We suspect this may be related to more patients undergoing ERCP with biliary aspiration who have possibly had other sources of their clinical symptoms, longer duration of antibiotics pre-ERCP, or different antibiotics pre-ERCP, or to there being less resistance to antibiotics in biliary organisms in the prior study.

In the current study, we sought to clarify the influence of a prior biliary ES on the positivity rate and microbiology of bile cultures. We found effects of biliary ES that were similar to that of having a prior indwelling biliary stent. This is to be expected as both biliary ES and an indwelling biliary stent violate the natural sphincter of Oddi barrier between the duodenal lumen and the biliary system [20]. We hypothesize the changes in microbiology with regard to the presence or absence of a biliary ES or a biliary stent is likely due to migration of luminal bacteria into the biliary tree and/or colonization of the existing biliary stent. This is further supported by the increased presence, in these subsets of patients, of organisms that are well-known to produce biofilms and colonize foreign bodies such as urinary catheters and biliary stents (Enterobacter spp., Enterococcus spp., Klebsiella spp., and Pseudomonas spp.) [21–24]. There is a large overlap of patients with a biliary ES and an existing or prior biliary stent. In addition, although the data are not completely available to analyze within this study, patients who receive a biliary ES or biliary stent likely have a higher disease burden from more complex disease, prior hospitalizations, and multiple exposures to antibiotics, which may contribute to higher rates of positive bile culture, antibiotic resistance, and bacterial colonization.

We discovered a notable number of high concern multidrug-resistant organisms (155/662; 23.4%). There was a significant increase in isolation of VRE in the presence of a biliary ES (11.5% vs. 5%; OR2.5 [95%CI1.3–4.7]; adjusted P=0.04) and biliary stents (13.5% vs. 5.3%; OR2.8 [95%CI1.6–4.9]; adjusted P=0.002), but not other multidrug-resistant organisms. This association has been reported previously [25].

A potential benefit of collecting and culturing bile from patients with suspected cholangitis is the ability to tailor antibiotic regimens to the resistance pattern of the biliary aspirates. Furthermore, in patients with recurrent cholangitis from structural disease (e.g., primary sclerosing cholangitis), the presence of recurrent positive bile cultures or highly resistant biliary cultures may prompt liver transplant teams to advocate for exception points for liver transplantation. We also believe that, in the era of known risk of transmission of organisms from reusable duodenoscopes, the knowledge of biliary culture data can inform the decision regarding enhanced endoscope reprocessing and/or future endoscope usage for that patient. In a patient with a known multidrug-resistant organism on bile culture, we would consider reprocessing that endoscope subsequently with an enhanced treatment (ethylene oxide sterilization). Furthermore, we would consider future ERCPs for that patient being done with a single-use duodenoscope.

The recommended choice of empiric antibiotics by medical societies for community acquired biliary infections include a third-generation cephalosporin, a fluoroquinolone, or a penicillin/beta-lactamase inhibitor [2,13,26]. In our earlier study, given the 96% susceptibility of Gram-negative organisms to ciprofloxacin, we continued to consider ciprofloxacin a recommended initial empiric choice for suspected acute cholangitis. In the current study however the susceptibility to ciprofloxacin of the most common Gram-negative organisms has been reduced substantially (E.coli susceptibility 64% and K.pneumoniae susceptibility 88%). This decrease in susceptibility to ciprofloxacin demonstrates an increase in the prevalence of Gram-negative organisms resistant to this commonly used antibiotic in our community. A similar change in resistance pattern has been reported in other geographic areas as well [27–29]. An important strength of this study is that we were able to assess a longitudinal change over time in the antibiotic sensitivity of biliary organisms by comparing with a similar cohort from our prior study [12].

Antibiotic selection for suspected cholangitis is an important decision made routinely in clinical practice. We support a judicious use of empiric antimicrobial agents for biliary infections that should be modified based on the local antibiogram. We no longer use ciprofloxacin as an empiric antibiotic for suspected acute cholangitis. For inpatients suspected of having acute cholangitis, the practice at our institution is to recom-
Gromski Mark A et al. Microbiology of bile aspirates in patients with suspected cholangitis at Digestive Disease Week 2018, Washington DC, USA, on 2 June 2018.

Competing interests

M.A. Gromski serves as a consultant for Boston Scientific. S. Sherman serves as a consultant for Boston Scientific, Cook Medical, and Olympus. G. Lehman serves as a consultant for Cook Endoscopy. J. Easler serves as a consultant for Boston Scientific. The remaining authors declare that they have no conflict of interest.

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