Myricetin bioactive effects: moving from preclinical evidence to potential clinical applications

Yasaman Taheri 1,2, Hafiz Ansar Rasul Suleria 3, Natália Martins 4,5, Oksana Sytar 6,7, Ahmet Beyatlı 8, Balâkız Yeskâliyevâ 9, Gulnaz Seitimova 9, Bahare Salehi 10,11, Prabhakar Semwal 12,13, Sakshi Painuli 12,14, Anuj Kumar 15, Elena Azzini 16, Miquel Martorell 17,18, William N. Setzer 19,20, Alfred Maroyi 21, Javad Sharifi-Rad 22

Abstract

Several flavonoids have been recognized as nutraceuticals, and myricetin is a good example. Myricetin is commonly found in plants and their antimicrobial and antioxidant activities is well demonstrated. One of its beneficial biological effects is the neuroprotective activity, showing preclinical activities on Alzheimer, Parkinson, and Huntington diseases, and even in amyotrophic lateral sclerosis. Also, myricetin has revealed other biological activities, among them as antidiabetic, anticancer, immunomodulatory, cardiovascular, analgesic and antihypertensive. However, few clinical trials have been performed using myricetin as nutraceutical. Thus, this review provides new insights on myricetin preclinical pharmacological activities, and role in selected clinical trials.

Keywords: Myricetin, Antimicrobial, Antioxidant, Neuroprotection, Diabetes, Cancer, Immunomodulatory, Cardiovascular disease

Introduction

Polyphenols are a wide group of plant-derived molecules resulting from secondary metabolism, ubiquitously distributed in vegetable kingdom where they display different activities such as protective effect against UV rays, bacteria, virus and fungi infections, modulation of plant hormones, enzyme inhibition and pollinator attraction [1]. In nature, there are a plethora of different polyphenols that can be classified in the following main classes: simple phenolic acids (e.g. gallic, vanillic, syringic, p-hydroxybenzoic), hydroxycinnamic acid derivatives (such as caffeic acid, p-coumaric, ferulic, sinapic), flavonoids, stilbenes and lignans. The largest common class of polyphenols present in human diet is represented by flavonoids [2, 3]. Chemically flavonoids are classified in flavans, flavones, flavonols, and anthocyanidins [4]. Among the flavonols, myricetin, a 3,3′,4′,5,5′,7-hexahydroxyflavone, possess one of the most hydroxylated structures (Fig. 1). The solubility of myricetin in water is poor (16.6 μg/mL) but increases when deprotonated in basic aqueous media and in some organic solvents (dimethylformamide, dimethylacetamide, tetrahydrofuran and acetone) [5]. The chemical stability of myricetin is pH and temperature dependent [6]. Depending on the environment conditions, myricetin can exert, in vitro, both a potent antioxidant and a pro-oxidant effect. Buchter et al. [7] attributed its direct antioxidant action to several structural elements. On the other hand, Chobot and...
Hadacek [8] demonstrated the pro-oxidative properties of myricetin to molecular oxygen reduction to reactive oxygen species (ROS) and iron (III) to iron (II) and also highlighted the ability of myricetin to serve as a substitute for ascorbic acid, albeit less efficiently.

Myricetin is mainly present in the glycoside form (O-glycosides), in vegetables, fruits, nuts, berries, herbs, plants together with beverages, such as tea, wine, fruit and medicinal plants [9–15]. There are numerous factors that can influence myricetin levels in plant foods such as genetic and environmental factors, germination, and ripeness degree, variety, seasonal variation, and storage, processing and cooking. The estimate of total flavonoid intake is difficult to calculate, as appropriate tables of food composition are not yet available. However, reliable data on daily flavonoid intake in a population are needed to develop proper dietary recommendations and even for correct data interpretation from intervention studies. The Flemish Dietetic Association database determined an average daily intake of myricetin of 2.2 ± 2.5 mg Mul-lie et al. [16]. In a Korean adult population, Jun et al. [17] estimated an average intake of 0.8 mg/day representing about 1–2% of flavonol subclass, while a mean intake of myricetin 2 mg/day ranged from 1 to 4 mg/day in adults (18 to 64 years) in the European Union was reported by Vogiatzoglou et al. [18]. The knowledge on habitual flavonoids consumption is also crucial to determine their possible impact on human health. Myricetin exhibited antioxidant properties and free radical-scavenging effects [19]. These activities seem to support a wide range of beneficial outcomes including, anti-platelet aggregation, antihypertensive, immunomodulatory, anti-inflammatory, anti-allergic, analgesic, anticancer actions and so on [6, 20–25]. The main goal of the present review is to provide new insights on myricetin preclinical pharmacological activities, and its role in selected clinical trials.

**Myricetin in plants**

Myricetin glycosides include myricetin-3-O-(4′-acetyl)-α-L-arabinopyranoside, myricetin-3-O-(3′-acetyl)-α-L-arabinopyranoside, myricetin-3-O-β-D-galactopyranoside, myricetin-3-O-α-L-rhamnopyranoside, myricetin-3-O-β-D-xlylopyranoside, myricetin-3-O-α-L-arabinofuranoside, myricetin-3-O-(6″-galloyl)-β-D-galactopyranoside, myricetin-3-O-(3″-O-galloyl)-α-L-rhamnoside, myricetin-3-O-(2″-O-galloyl)-α-L-rhamnoside, and myricetin-3-O-α-L-rhamnoside [27].

The first time myricetin was identified was in plants of the Myricaceae, *Comptonia peregrina* (L.) Coul. and later *Morella cerifera* (L.) Small [28, 29]. The myricetin concentration in the plants such as *Rosa canina* L. (rosa hip), *Urtica dioica* L. (nettle), and *Portulaca oleracea* L. (purslane) found between 3 and 58 mg/kg [13].

Myricetin was isolated from *Polygonum bellardii* All. (Polygonaceae) as yellow needles (50 mg) from aerial parts using MeOH extract [30]. Previously, a prescreening of leaves of 28 polygonaceous plants was estimated that myricetin glycosides were relatively rare consituents [31]. *Trigonella foenum-graecum* L. gemmo-modified extract had the richest content in myricetin (830 mg/kg), followed by *Euphorbia tirucalli* L. (821 mg/kg), rhizomes of *Cyperus rotundus* L. (702 mg/kg) and seed extract of *T. foenum-graecum* (547 mg/kg). *C. rotundus* gemmo-modified extracts contained 104 mg/kg myricetin [10].

The highest level of myricetin content has been identified in the strawberry and spinach [9]. Species of *Anacardium* and *Mangifera* (Anacardiaceae) found to have high levels of hydroxylated compounds like myricetin, gallic acid, proanthocyanidins and flavonols. In *Marantodes pumilum* (Blume) Kuntze (Primulaceae) were identified quercetin, myricetin, kaempferol, catechin and epigallocatechin [32].

The most common sources of myricetin are vegetables, fruits, nuts, berries and tea [33]. Myricetin-rich foods are listed in Table 1 based on the USDA Food Database (compiled data from all fruits and vegetables that contain information on myricetin concentration) [34]. In black fruits the quantities varied between 14 and 142 mg/kg [12]. Myricetin is the most abundant flavonol of black currant, and its quantity varied significantly among black currant cultivars [35]. At the same time, honey is also a source of flavonoids, especially myricetin. The HPLC analyses of honeys from Australian *Eucalyptus* have shown that the flavonoids myricetin, quercetin, tricetin, kaempferol and luteolin exist in all honeys. Myricetin was found in range from 29.2–289.0 μg/100 g honey [36]. In grapes, flavonol glycosides from the following aglycons have been identified: myricetin (3′,4′, 5′-triOH), laricitrin (3′-MeO analog of myricetin) and syringetin (3′,5′-diMeO analog of myricetin), quercetin and kaempferol [37]. The simultaneous presence of these aglycons was detected in different types of red
wine *Vitis vinifera* L. grapes [38], while in white wine, only quercetin, kaempferol and isorhamnetin were detected [37].

**Preclinical pharmacological activities of Myricetin**

Myricetin displays multiple preclinical biological effects [19]. Thus, in the following subsections, the antimicrobial, antioxidant, neuroprotective, anti-diabetic, anticancer, immunomodulatory, cardioprotective, analgesic, anti-hypertensive and wound healing potential of myricetin are briefly discussed and summarized.

**Antimicrobial activities**

Antimicrobial mechanism of flavonoids may involve membrane disruption, inhibition of cell envelope synthesis, inhibition of nucleic acid synthesis, inhibition of bacterial virulence and quorum sensing, which impairs their ability to form biofilms, inhibition of efflux pumps, and inhibition of NADH-cytochrome C reductase activity and ATP synthase [39, 40]. Myricetin inhibited *Escherichia coli* DNA gyrase (IC<sub>50</sub> 1.18 mg/dL) [41], and DnaB helicase (IC<sub>50</sub> 11.3 μM) [42], and cellular DNA and RNA polymerases [43].

Myricetin showed a significant antimicrobial activity against foodborne pathogens in terms of minimum inhibitory concentration (MIC, mg/mL) <15.0, <15.0, <20.0, <10.0 at 24 h and <20.0, <20.0, <15.0, <5.0 at 60 h incubation for *Escherichia coli*, *Salmonella paratyphi*, *Salmonella cholerasuis*, and *Salmonella enteritidis*, respectively [44]. The compound myricetin revealed curli-dependent *E. coli* biofilm formation inhibition (IC<sub>50</sub> = 46.2 μM), curli contributes to the robustness of *E. coli* biofilms [45].

At 100 μM concentration, myricetin exhibited in vitro anti-HIV activity in cell cultures: TZM-bl (> 87%; IC<sub>50</sub> 22.91 μM, 1.76 μM) [46]. Myricetin exhibited the highest anti-HIV reverse transcriptase activity (> 49%, IC<sub>50</sub> 203.65 μM) at the concentration of 100 μM [46].

Yadav et al. [47] demonstrated the anti-tubercular activity of 15 selected flavonoids including myricetin and their structure–activity relationships were evaluated against *Mycobacterium tuberculosis* H37Rv strain radiometrically. Myricetin was found to be active against *M. tuberculosis*, with a MIC of 50 μg/mL, and structure–activity relationships authenticated their anti-tubercular potential due to the presence of hydroxy groups in their structure.

The inhibitory activity of the compounds were evaluated against DNA gyrase from *E. coli* by DNA supercoiling. Mean antibacterial activity in terms of MIC and IC<sub>50</sub> were 142 μg/mL and 1.18 mg/mL respectively. The structure–activity relationship analysis suggests that, the presence of hydroxyl and substitution in the ring A and B position are essential for the best inhibitory effects [41].

The inhibitory effect of myricetin on severe acute respiratory syndrome-coronavirus (SARS-CoV) helicase, nsp13, and hepatitis C virus (HCV) helicase, NS3h was also assessed [48]. Myricetin was found to inhibit SARS-CoV helicase protein by affecting the ATPase activity (IC<sub>50</sub> 2.71 μM), however, it failed to affect the ATPase activity of the HCV NS3 helicase.

DeSouza and Wahidullah [49] reported the antimicrobial activity on *E. coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, *Salmonella typhi*, *Shigella flexneri*, *Staphylococcus aureus*, *Vibrio cholerae* and myricetin showed the best activity against *P. aeruginosa* (MIC 1.5 μg/mL). Gendaram et al. [50] reported the myricetin antibacterial effect against *S. aureus* by the disc diffusion method (300 μg/disc, inhibition zone 9 mm) but reported no antibacterial activities against *P. aeruginosa*, *E. coli*, *Enterococcus faecalis*, or *Micrococcus luteus*. However, at 100 μM concentration, myricetin did not exhibit antimicrobial activity on Gram-positive bacteria but showed inhibitory activity against sortase A (SrtA) from *S. aureus* (92%; IC<sub>50</sub> 4.63 μM) [51]. In vitro antimicrobial activity of six natural phytochemicals including myricetin (alone and with combination) were evaluated against five strains of *P. aeruginosa* by using a time-kill assay. The compound showed the MIC as 500 μg/mL against all five strains of *P. aeruginosa* [52]. Other reports of the compound based on antimicrobial and antiviral studies are presented in Table 2.

**Antioxidant activities**

Plant-based compounds considered as natural antioxidants have attracted a large number of communities of scientist, researchers, industries and traditional healers for their health-promoting characteristics. The antioxidant...
potential of myricetin has been reported by several authors in the last few decades.

Hou et al. [61] studied the antioxidant effect of HS15-Myr micelles and independent myricetin by using FRAP (ferric reducing antioxidant power) and ABTS (2,2′-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid) assays. The ABTS assay displayed an improved value from 22.20 to 41.77% in HS15-Myr micelles and 0 to 6.12% in independent myricetin at two different concentrations and incubation periods. The FRAP assay also presented an improved value from 1.27 to 8.94 mM Fe^{2+}/g in HS15-Myr micelles and 13.63 to 16.33 mM Fe^{2+}/g in independent myricetin at two different concentrations and incubation periods. Myricetin in HS15-Myr micelles exhibited in both assays stronger antioxidant effects when compared to independent myricetin.

Barzegar [62] reported the ROS-protection efficiency of the compound myricetin in a cell-free and cell-based system. A low concentration of compound significantly inhibited intracellular ROS production and also protected cells against toxicity induced by peroxide compounds.

Guitard et al. [63] reported that, myricetin is more efficient than α-tocopherol and synthetic antioxidants on preservation of omega-3 oils. Other studies on antioxidant potential of the compound are presented in Table 3.

Neurobiological activities
Natural flavonoids have exerted positive impacts on body through affecting multiple cell systems and modulating the activity of various pathways to reduce cognitive decline and neuronal dysfunction [79]. Myricetin is one of such flavonoids, and multiple studies have been conducted to assess the neuroprotective effects of this compound and its interaction with brain receptors (Table 4). The main mechanisms are shown in Fig. 2.
Table 3  Antioxidant activities of myricetin

| Assay                        | Model                          | Results                                                                 | Ref. |
|------------------------------|--------------------------------|-------------------------------------------------------------------------|------|
| Density functional theory    | in silico                      | The bond dissociation enthalpy computed and the compound showed ionization potentials 161.4 kcal/mol. | [64] |
| Antioxidant response element (ARE) activation | in vitro                    | Activates Nrf2 antioxidant response element pathways and is involved in myricetin-induced expression profiling in hepatic cells. | [65] |
| Deoxyribose degradation      | in vitro                       | Significant antioxidant activity (complex with iron) in the presence of ascorbic acid. | [8]  |
| DPPH                         | in vitro                       | Myricetin/HP-β-CD inclusion complex formation enhances antioxidant activity of drugs. | [66] |
| DPPH                         | in vitro                       | Significant RSA dose-dependently                                       | [50] |
| DPPH, ABTS                   | in vitro                       | Inhibition activity from 13.3 to 99.8% at doses of 0.03 to 1 mg/ml during 5 to 20 min. | [67] |
| DPPH, FRAP                   | in vitro                       | High RSA in DPPH assay, and intermediate ferric reducing ability in FRAP assay. | [68] |
| DPPH, FRAP, ABTS             | in vitro                       | Mean activity for FRAP (27.2, 26.7) mmol Fe2+/L, DPPH (7.9, 9.3) mmol TEAC/L, and ABTS (9.3, 11.5) mmol TEAC/L. | [69] |
| DPPH, TPTZ, superoxide       | in vitro                       | Myricetin and its derivatives showed IC50 value from 1.82 to 3.27 μg/mL in DPPH assay and 1.86 to 3.83 μg/mL in superoxide assay however, 1.38 to 2.89 μM equivalent to Fe2+/L for TPTZ assay. | [71] |
| H2O2                        | in vitro                       | Increases hydrogen peroxide resistance in Saccharomyces cerevisiae.      | [72] |
| DPPH, ROS                    | in vitro                       | 21–54% scavenging activity in DPPH assay (5–10 μg/mL) and 35–73% intracellular ROS scavenging activity (1–10 μg/mL). Significantly inhibits H2O2-induced cell death and activated antioxidant enzymes. | [73] |
| NO                           | in vitro                       | Mean scavenging activity compared to hydrophilic antioxidants.           | [74] |
| ROS                          | in vitro                       | Inhibits peroxynitrite-mediated DNA damage in primary astrocytes at 5 μM. | [75] |
| ROS                          | in vitro                       | The IC50 value for inhibitory effect on triglyceride and ROS were recorded as > 150 μM and 122.7 μM. | [76] |
| ROS                          | in vitro                       | Inhibits H2O2-induced cell death and increases cell survival (65%).      | [77] |
| DCFH-DA                      | in vivo                        | Inhibits ROS production in normal individuals and in patients with sickle cell anemia. | [78] |

ABTS 2’,2’-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid, ARE antioxidant response element, DCFH-DA dichloro-dihydro-fluorescein diacetate, DPPH 2,2-diphenyl-1-picrylhydrazyl, FRAP ferric reducing antioxidant power, NO nitric oxide, ORAC oxygen radical absorbance capacity, ROS reactive oxygen species, RSA radical scavenging activity, TEAC trolox equivalent antioxidant capacity, TPTZ tri-pyridyl triazine

Antidiabetic activities

Myricetin antidiabetic activity has been reported by several authors in the last few years and limited reports are also available on its anti-obesity activity but in this review, we focused on only its antidiabetic potential. Karunakaran et al. [101] reported the in vitro effect of myricetin on high glucose-induced β-cell apoptosis, possibly via cyclin-dependent kinase 5 (CDK5) inhibition. Data revealed that myricetin (20 μM) significantly protected β-cells reducing apoptosis in INS-1 cells and rat islets that were incubated with glucose at the concentration of 30 mM for 24 and 48 h, respectively. Docking studies predicted myricetin inhibited activation of CDK5.

The effect of myricetin was evaluated in diabetes mellitus-associated kidney injuries and dysfunction in an experimental mouse model with diabetes mellitus induced by 5 consecutive injections of low-dose streptozotocin (STZ) [20]. The data revealed that myricetin (orally twice a day, 100 mg/kg/day, for 6 moths) inhibited the Ixβ3/ΔNF-κB pathway, with this pathway being independent of nuclear factor erythroid 2-related factor (Nrf2) regulation. It was also reported that myricetin activates glucagon-like peptide 1 receptor (GLP-1R) and its long-term oral administration (200 mg/kg, for 40 days) validates its glucoregulatory effects [102].

Insulin’s metabolic action is mediated via the activation of phosphatidylinositol 3-kinase (PI3K) and its downstream effectors, the protein kinase B (PKB/Akt) kinases [103]. In contrast, AMP-activated protein kinase (AMPK) signal pathway is likely to mediate the effect of insulin-independent stimuli for glucose uptake in muscle [104]. In an in vitro study, myricetin enhanced Akt and AMPK protein activity, encouraged glucose uptake and reduced insulin resistance [105]. The mechanisms of myricetin for improving insulin-sensitive tissue might be the amelioration of impaired signaling intermediates downstream of insulin receptors through enhancing the secretion of β-endorphin, which in turn led to the activation of peripheral μ-opioid receptors [106, 107]. Then, myricetin affects insulin receptor phosphorylation, insulin receptor substrate-1 (IRS-1), the p85 regulatory sub-unit of PI3K, Akt and Akt substrate of 160 kD, with subsequent effects on glucose transporter 4 (GLUT4) translocation [108].
Other previous studies on antidiabetic potential of the compound are shown in Table 5.

**Anticancer activities**
Cancer is responsible for second highest cause of death across the globe [124, 125]. It has been reported that number of death due to this devastating disease would expand to over 13 million by 2030 [126, 127]. Laboratory and clinical studies have reported that myricetin from natural sources exerts promising effects against various types of cancer [19, 21]. The dietary compound myricetin also has the potential to inhibit key enzymes involved in cancer initiation and growth.

Myricetin has presented cytotoxic activity in human colon cancer cells. Kim et al. [21] demonstrated that myricetin significantly induces the Bcl2-associated X
protein (BAX)/Bcl2 ratio, and induces apoptosis of HCT-15, in a dose-dependent manner (5 to 100 μM). This study suggested that myricetin can be utilized for the design of therapeutic agents against human colon cancer. Myricetin also acts as a potent inhibitor of human flap endonuclease 1 (hFEN1) protein (IC_{50} 690 nM), based on inhibitory mechanisms, molecular docking, and cancer cell-based assays [128]. The hFEN1 protein is a functional member of the 5′-nuclease superfamily. By chemical nature, hFEN1 is a metal ion-dependent and structure-specific nuclease and also instrumental in DNA replication and repairing processes. Molecular docking studies revealed that ring A of myricetin compound, including 4-keto and 5-OH, was found stretched towards the two divalent metal ions. Both metal ions are critical as they seem to interact with Arg100 and Lys93 amino acids through hydrogen bonds. These interacted residues are well known for their critical interplay in hFEN1’s activity during human colon cancer.

Myricetin has also been shown to protect against ovarian cancer through suppressing ovarian cancer cell angiogenesis [129]. Anti-angiogenic effects of myricetin (5 to 20 μM) assessed through in vitro (HUVEC) and in vivo (CAM) models revealed that this compound significantly inhibits angiogenesis induced by OVCAR-3 cells. In SKOV3 human ovarian cancer cells, myricetin inhibited viability and induced apoptosis (40 μg/mL, time-dependent manner) through endoplasmic reticulum stress and DNA double-strand breaks [130]. Zheng et al. [131] stated that in A2780 and OVCAR3 ovarian cancer cells, the dietary flavonoid myricetin induced significant cytotoxicity (IC_{50} = 25 μM). In a recent study, Tavsan and Kayali [132] reported that myricetin suppressed ovarian cancer cell growth, induced apoptosis, arrested cell cycle and also had the potential to inhibit cell invasion in a significant manner (IC_{50} = 184 μM A2780, 32 μM OVCAR-3, 3.3 μM SKOV3, and > 500 μM OSF). Thus, it can be concluded that myricetin has enough potential to cope with ovarian cancer in a significant manner.

Myricetin has potent anticancer-promoting activity against skin cancer. It was found capable of inhibiting neoplastic cell transformation and mitogen-activated
protein kinase 1 (MEK1) activity (myricetin 1 or 5 μM) [133]. Molecular interaction between myricetin and MEK1 suppressed MEK1 activity leading to downstream signaling to the ERK/p90RSK/AP-1 pathway. In another study, myricetin has been presented as a potent chemoprotective agent against skin cancer [134]. Myricetin can bind directly to central kinases including PI3-K, Akt, JAK1, Raf1, MEK1, MKK4, and Fyn, which regulate multiple cell signaling pathways in cancer cells. Myricetin inhibited 12-O-tetradecanoylphorbol-13-acetate (TPA)- and epidermal growth factor (EGF)-induced cell transformation by 76 and 72%, respectively at 10 μM concentration. Sun et al. [135] recently reported that myricetin has anticancer activity against skin cancer A431 cells viability and proliferation (IC₅₀ = 65 μM).

### Immunomodulatory activities

The preclinical immunomodulatory effects of myricetin have also been increasingly reported. Ghassemi-Rad et al. [140] concluded that myricetin has the potential to inhibit T-lymphocyte activation in a mouse model through bead-immobilized anti-CD3 and anti-CD28 monoclonal antibodies. This study clarified the mechanism of action and reported the suppressive effect of myricetin on T lymphocytes mediated through extracellular H₂O₂ generation. In mouse primary macrophages and RAW264.7 monocytic cell-line, this phenolic compound was found to inhibit the lipopolysaccharide (LPS)-induced interleukin (IL)-12 production in a significant manner through down-regulation of NF-κB binding activity [22]. In isolated rat aortic rings, myricetin induced endothelium-dependent contractile responses at 50 μM.

An earlier in vitro study demonstrated the anti-metastatic effect of myricetin in human lung adenocarcinoma A549 cells [136]. This study revealed that myricetin (5 to 20 μM) suppresses adenocarcinoma A549 cell invasion and migration through inhibition of the ERK pathway in a time-dependent manner. Along with a combination of radiotherapy, myricetin was found responsible to enhance the tumor radio-sensitivity of lung cancer A549 and H1299 cells through significant suppression of cell-surviving fraction and proliferation [137]. Wang et al. [138] found that the combination of myricetin with 5-fluorouracil chemotherapy has the potential to enhance tumor chemosensitivity of esophageal cancer EC9706 cells. Sun et al. [139] investigated the function of myricetin phytochemical against human T24 bladder cancer in a dose- and time-dependent fashion, and stated that myricetin significantly inhibits both T24 cancer cells viability and proliferation (IC₅₀ = 85 μM).

### Table 5

| Compound / Plant species | Model | Results |
|-------------------------|-------|---------|
| Myricetin               | in vivo | Enhanced enzymatic and non-enzymatic antioxidant defense system and showed protective effects against oxidative damage in liver and kidney of streptozotocin-cadmium-induced diabetic model. |
| Myricetin               | in vivo | Inhibitory activity against α-glucosidase (IC₅₀ = 414 μM) in dose dependent manner. |
| Myricetin               | in vivo | Anti-hyperglycemic and renoprotective effects at 1.0 mg/kg. |
| Myricetin               | in vivo | Improved and re-established renal functions and activities of the glutathione peroxidase and xanthine oxidase enzymes in diabetic rat model. |
| Myricetin               | in vivo | Antidiabetic activity against t-BHP-induced oxidative stress. |
| Myricetin               | in vivo | Reduced glycemia in diabetic rats up to 50% after 2 days of treatment at 3 mg/12 h. |
| Myricetin               | in vivo | Stimulated lipogenesis in rat adipocytes and enhanced the stimulatory effect of insulin (EC₅₀ = 65 μM). |
| Myricetin               | in vivo | Inhibited intestinal α-glucosidase (29%) and porcine α-amylase (64%) with IC₅₀ vale of 0.38 mM. |
| Abélmoschus moschatus  Medik. (aerial part) | in vivo | Improved insulin sensitivity in rats. |
| Ampelopsis grossedentata (Hand.-Mazz.) W.T. Wang (leaves) | in vivo | Inhibitory activity against α-glucosidase (IC₅₀ = 319.3 μM). |
| Azadirachta indica A.Juss. (leaves) | in vivo | Enhanced insulin signaling pathway and glucose utilization in skeletal muscle. |
| Hovenia dulcis Thunb. (seeds) | in vitro | Inhibited intestinal α-glucosidase with IC₅₀ = 3 μg/mL and α-amylase with IC₅₀ = 662 μg/mL. |
| Myrtus communis L. (seeds) | in vitro | Significant antidiabetic activity in diabetic models. |
| Syzygium cumini (L.) Skeels (seeds) | in vitro | Inhibitory activity against α-glucosidase (IC₅₀ = 1.7 μg/mL) and α-amylase (IC₅₀ = 7.62 μg/mL). |
| Syzygium malaccense (L.) Merr. & L.M.Perry (leaves) | in vitro | Inhibitory activity against α-glucosidase (IC₅₀ = 15.52 μg/mL) and α-amylase (IC₅₀ = 147.30 μg/mL). |

Previous studies on preclinical antidiabetic potential of myricetin

| Compound / Plant species | Model | Results |
|-------------------------|-------|---------|
| Myricetin               | in vivo | Enhanced enzymatic and non-enzymatic antioxidant defense system and showed protective effects against oxidative damage in liver and kidney of streptozotocin-cadmium-induced diabetic model. |
| Myricetin               | in vivo | Inhibitory activity against α-glucosidase (IC₅₀ = 414 μM) in dose dependent manner. |
| Myricetin               | in vivo | Anti-hyperglycemic and renoprotective effects at 1.0 mg/kg. |
| Myricetin               | in vivo | Improved and re-established renal functions and activities of the glutathione peroxidase and xanthine oxidase enzymes in diabetic rat model. |
| Myricetin               | in vivo | Antidiabetic activity against t-BHP-induced oxidative stress. |
| Myricetin               | in vivo | Reduced glycemia in diabetic rats up to 50% after 2 days of treatment at 3 mg/12 h. |
| Myricetin               | in vivo | Stimulated lipogenesis in rat adipocytes and enhanced the stimulatory effect of insulin (EC₅₀ = 65 μM). |
| Myricetin               | in vivo | Inhibited intestinal α-glucosidase (29%) and porcine α-amylase (64%) with IC₅₀ vale of 0.38 mM. |
| Abélmoschus moschatus  Medik. (aerial part) | in vivo | Improved insulin sensitivity in rats. |
| Ampelopsis grossedentata (Hand.-Mazz.) W.T. Wang (leaves) | in vivo | Inhibitory activity against α-glucosidase (IC₅₀ = 319.3 μM). |
| Azadirachta indica A.Juss. (leaves) | in vivo | Enhanced insulin signaling pathway and glucose utilization in skeletal muscle. |
| Hovenia dulcis Thunb. (seeds) | in vitro | Inhibited intestinal α-glucosidase with IC₅₀ = 3 μg/mL and α-amylase with IC₅₀ = 662 μg/mL. |
| Myrtus communis L. (seeds) | in vitro | Significant antidiabetic activity in diabetic models. |
| Syzygium cumini (L.) Skeels (seeds) | in vitro | Inhibitory activity against α-glucosidase (IC₅₀ = 1.7 μg/mL) and α-amylase (IC₅₀ = 7.62 μg/mL). |
| Syzygium malaccense (L.) Merr. & L.M.Perry (leaves) | in vitro | Inhibitory activity against α-glucosidase (IC₅₀ = 15.52 μg/mL) and α-amylase (IC₅₀ = 147.30 μg/mL). |
dose-dependent manner, myricetin inhibited the secretion of a potent T cell growth factor, namely IL-2 protein from mouse EL-4 T cells, activated with phorbol 12-myristate 13-acetate (PMA) plus ionomycin [142]. In vitro evidence demonstrated that at 5–100 μM, myricetin inhibits CD69 expression and lymphocytes proliferation in a mouse model. Moreover, an in vitro investigation revealed that myricetin significantly affects IL-2 expression. However, further in vitro and in vivo investigations are required to explore myricetin as an immunomodulatory agent.

Cardioprotective activity
Previous studies have demonstrated that myricetin also has beneficial effects on the human vascular system [23]. In human umbilical vein endothelial cells, myricetin (100 μM), revealed vasculoprotective effects through changes at the transcriptional level [143]. Myricetin has been presented as a functional agent towards preventing atherosclerosis through inhibition of CD36 cell surface protein and miRNA expression in a significant manner [144]. In isolated and Langendorff-perfused rat hearts, without affecting contractility and relaxation, myricetin elicited coronary dilation [145]. In Triton-treated hyperlipidemic rats, evidence from an in vivo investigation demonstrated that myricetin exerts lipid-lowering activity and suggests that myricetin can be utilized in the treatment of hyperlipidemia and cardiovascular diseases (CVD) [146].

In Wistar rats, myricetin significantly inhibited the effects of histopathological changes of isoproterenol on heart rate, the levels of different cardiac marker enzymes, including lactate dehydrogenase (LDH), creatine kinase (CK), aspartate aminotransferase (AST), superoxide dismutase (SOD) and catalase (CAT), as well changes in vascular reactivity and electrocardiographic patterns [147].

A mechanism-based study by Scarabelli et al. [148] demonstrated that myricetin exerts strong inhibitory activity against signal transducer and activator of transcription 1 (STAT1) activation, and also protects the heart from ischemia/reperfusion-injury. The available genomic and genetics data from preclinical experiments have shown that myricetin is likely to confer the first line of defense against cardiovascular and other associated diseases.

Analgesic activities
In acetic acid-induced writhing response, formalin-induced paw licking, sedative activity and hot plate test models, myricetin revealed potent analgesic effects, closely related with peripheral analgesia, but not with the opioid system [24]. The compound also produced a significant analgesic effects in a rat model of neuropathic pain, by decreasing spinal nerve ligation-induced mechanical allodynia and thermal hyperalgesia lasting for several hours (0.1–10 mg/kg i.p.) [149].

Antihypertensive activities
The antihypertensive effects of myricetin were evaluated in the deoxycorticosterone acetate (DOCA)-salt-hypertensive rat model. Myricetin reduced systolic blood pressure, vascular reactivity changes and reversed the increased heart rate induced by DOCA. At oral doses of 100 and 300 mg myricetin/kg b.w., the compound displayed antihypertensive properties in the DOCA rat model of hypertension [25]. In another study, the compound lowered the high blood pressure that was induced by fructose doses of 100 and 300 mg/kg p.o. in rats and reversed sugar-triggered metabolic changes [150].

Wound healing
The wound-healing effects of myricetin-3-O-β-rhamnose were investigated on three different types of cells, keratinocytes, fibroblasts, and endothelial cells. The compound exhibited significant wound healing activity at 10 μg/mL [151].

Myricetin in clinical trials
Although the number of clinical studies reporting myricetin health benefits in ailments and disorders is low, the increasing data from preclinical studies have supported its beneficial effects [152, 153].

In a 4-week randomized placebo-controlled clinical trial the effect of 300 mg Blueberin (250 mg Blueberry leaves, Vaccinium arctostaphylos L., and 50 mg myricetin, three times per day) on fasting plasma glucose and some other biochemical parameters has been investigated in 42 female volunteers (46 ± 15 years; body mass index, BMI, 25 ± 3 kg/m²) with diabetes type 2. The Blueberin treatment significantly reduced fasting plasma glucose from 143 ± 5.2 mg/L to 104 ± 5.7 mg/L. In addition to antidiabetic effects, results showed that Blueberin also possessed pharmacologically relevant anti-inflammatory properties, reduced plasma enzyme levels of alanine aminotransferases (ALT), AST, glutamyltransferase (GGT), and reduced serum C-reactive proteins (CRP) [154]. Emulin™ (250 mg of patented blend of chlorogenic acid, myricetin, and quercetin), when regularly consumed, was able not only to lower the acute glycemic impact of foods, but also to chronically decrease blood glucose levels in type 2 diabetic humans (reductions between 1 and 5%) [155]. This study was performed in 40 male and female with fasting glucose range between 126 to 249 mg/mL and a BMI ≥ 30 kg/m².

Data from different studies also indicate the importance of myricetin as a chemopreventive agent, acting on cell proliferation, signaling mechanisms, apoptosis,
angiogenesis, and tumor metastasis [156]. Through the analysis of habitual food consumption of 10,054 participants of Finnish Mobile Clinic Health Examination Survey developed during 1966–1972, Knekt et al. [157] estimated that higher myricetin intakes in men led to lower prostate cancer risk. In a prospective study, Gates et al. [158] analyzed the association between the 5 common dietary flavonoids (myricetin, kaempferol, quercetin, luteolin and apigenin) intake and epithelial ovarian cancer incidence in 66,940 women. No clear association was found between total intake of examined flavonoids and incidence of ovarian cancer (Relative Risk [RR] = 0.75 for the highest versus lowest quintile, 95% confidence interval [CI] = 0.51−1.09; p-trend = 0.02), nor for myricetin intake (RR = 0.72, 95% CI = 0.50–0.94; p-trend = 0.01). However, there was a significant 40 and 34% decrease in ovarian cancer incidence for the highest versus lowest quintile for kaempferol and luteolin intake, respectively [158]. The association between flavonoids and flavonoid-rich foods intake and exocrine pancreatic cancer prevention study cohort were also examined [159]. Of the 27,111 male smokers with 306 pancreatic cancers, the data obtained suggests that a flavonoid-rich diet may decrease pancreatic cancer risk in male smokers not consuming supplemental α-tocopherol and/or β-carotene. Tang et al. [160] showed that high/increased flavonoids (e.g., myricetin) intake is associated with lower lung cancer risk in their studied population (meta-analysis of 8 prospective studies and 4 case-control studies involving 5073 lung cancer cases and 237,981 non-cases).

The intake of 36 g lyophilized grape powder (rich in flavans, anthocyanins, quercetin, myricetin, kaempferol, and resveratrol) also had a great impact in key risk factors for coronary heart disease (lowered levels of triglyceride, low-density lipoproteins, apolipoproteins B and E) in both pre- and post-menopausal women [161]. The study was performed on 24 pre- and 20 post-menopausal women for 4 weeks. However, wide ranges of clinical studies are still needed on the potential activities of myricetin which have been already indicated through in vitro and in vivo experiments.

Conclusions
Myricetin is a flavonoid present in many foods that has shown biological activities in numerous studies and has a potential use as a nutraceutical. Its antimicrobial and antioxidant role is widely studied, and numerous studies have shown neurobiological activities and a potential beneficial impact on AD, PD, HD and ALS. Also, pre-clinical studies have revealed antidiabetic, anticancer, immunomodulatory, anti-cardiovascular, analgesic and antihypertensive activities. These studies investigated the effect of myricetin, pure compound or plant extract rich in this compound. In plant studies, the extracts rich in myricetin always have other flavonoids that have also shown antioxidant activity alone. Nevertheless, new well-designed studies have to be performed to study all of the biological effects described before, as well as pre-clinical studies comparing the effect of myricetin compared to other flavonoids and phytochemicals. In the case of neurological diseases, more in-depth studies have to be designed to show the pre-clinical results.
University of Kyiv, Institute of Biology, Volodymyrska str., 64, Kyiv 01033, Ukraine. 7 Department of Plant Physiology, Slovak University of Agriculture, Nitra, A. Hlinku 2, 94976 Nitra, Slovak Republic. 8 Department of Medicinal and Aromatic Plants, University of Health Sciences, 34668 Istanbul, Turkey. 9 Faculty of Chemistry and Chemical Technology, Al-Fatati Kazakh National University, Almaty, Kazakhstan. 10 Noncommunicable Diseases Research Center, Bam University of Medical Sciences, Bam, Iran. 11 Student Research Committee, School of Medicine, Bam University of Medical Sciences, Bam, Iran. 12 Department of Biotechnology, Graphic Era University, Dehradun, Uttarakhand 248001, India. 13 Uttarakhand State Council for Science and Technology, Vigyan Dham, Dehradun, Uttarakhand 248007, India. 14 Himalayan Environmental Studies and Conservation Organization, Prem Nagar, Dehradun, Uttarakhand 248001, India. 15 Uttarakhand Council for Biotechnology, Silk Park, Prem Nagar, Dehradun, Uttarakhand 248007, India. 16 CREA-Research Centre for Food and Nutrition, Via Ardeatina 546, 00178 Rome, Italy. 17 Department of Nutrition and Dietetics, Faculty of Pharmacy, and Centre for Healthy Living, University of Concepcion, 4070386 Concepcion, Chile. 18 Unidad de Desarrollo Tecnológico, UDT, Universidad de Concepción, 4070386 Concepcion, Chile. 19 Department of Chemistry, University of Alabama in Huntsville, Huntsville, AL 35899, USA. 20 Aromatic Plant Research Center, 230 N 1200 E, Suite 100, Lehi, UT 84043, USA. 21 Department of Botany, University of Fort Hare, Private Bag X1314, Alice 5700, South Africa. 22 Zabol Medicinal Plants Research Center, Zabol University of Medical Sciences, Zabol, Iran.

Received: 21 March 2020 Accepted: 24 July 2020

Published online: 01 August 2020

References

1. Matilla MA. Chapter 10 - metabolic responses of plants upon different plant–pathogen interactions. In: Ahmad P, Ahanger MA, Singh VP, Tripathi DK, Alam P, Alyemeni MN, editors. Plant metabolites and regulation under environmental stress. Cambridge: Academic Press; 2018. p. 195–214.
2. Imran M, Salehi B, Sharif-Rad J, Gondal TA, Salehi B, Saeed F, Imran A, Shahbaz M, et al. Polyphenols: a key emphasis to its anticancer potential. Molecules. 2019;24(12):2277.
3. Sharif-Rad J, Sharif-Rad M, Salehi B, Roointan A, Mnayer D, Sharifi-Rad M. Antiproliferative activity of Pteleopsis suberosa leaf extract and its flavonoid components in human prostate carcinoma cells. Plant Med. 2006;72(7): 604–10.
4. Kang BY, Kim SH, Cho D, Kim TS. Inhibition of interleukin-12 production in mouse macrophages via decreased nuclear factor-κB DNA binding activity by myricetin, a naturally occurring flavonoid. Arch Pharm Res. 2005;28(3): 274–9.
5. Tong Y, Zhou X-M, Wang S-J, Yang Y, Cao Y-L. Analgesic activity of Veronica persica Poir. Cell Mol Biol. 2018;64(8):57–64.
6. De Leo M, Braca A, Sanogo R, Cardile V, DeTommassi N, Russo A. Antiproliferative activity of Peeplosis suberosa leaf extract and its flavonoid components in human prostate carcinoma cells. Plant Med. 2006;72(7): 604–10.
7. Kang NN, Fang ST, Wang JH, Wang ZH, Xia CH. Two new flavonoid glycosides from the halophyte Limonium franchetii. J Nat Prod Res. 2018;14(4):370–5.
8. Lau-Cam CA, Chan HH. Flavonoids from Comptonia peregrina. Phytochemistry. 1973;12:1829.
9. Jones JR, Lerbard MD, Jinwak UL, Absambira JF, Koren J 3rd, Blair L, O’Leary JC, Davey Z, Trotter J, Johnson AG, et al. The diallylheptanoids (±)-AR115-myricanol and two flavones from bayberry (Myrica cerifera) destabilize the microtubule-associated protein tau. J Nat Prod. 2011;74(1):38–44.
10. Abid ElHaker AM, El-Readi MZ, Ahmed AS, Nafady AM, Wink M, Ibrahim ZZ, Polyphebons from aerial parts of Polygonum bellardii and their biological activities. Pharm Biol. 2013;51(8):1026–34.
11. Kawasaki M, Kanomata T, Yoshitama K. Flavonoids in the leaves of twenty-eight polygonaceous plants. Bot Mag. 1986;99(2):259–69.
12. Biechter C, Ackermann D, Havermann S, Honnen S, Chovolou Y, Fritz G, Kampkottler A, Waten M. Myricetin-mediated lifespan extension in Caenorhabditis elegans is modulated by DAF-16. Int J Mol Sci. 2013;14(6):11895–914.
13. Chobot V, Hadaček F. Exploration of pro-oxidant and antioxidant activities of the flavonoid myricetin. Redox Rep. 2011;16(6):242–7.
14. Sultana B, Anwar F. Flavonols (kaempeferol, quercetin, myricetin) contents and antioxidant activity of fruit beers. Food Chem. 2020;305:125437.
15. Nardini M, Garaguso I. Characterization of bioactive compounds and antioxidant activity of fruit beers. Food Chem. 2020;305:125437.
41. Wu T, Zang X, He M, Pan S, Xu X. Structure-activity relationship of flavonoids on their anti-Escherichia coli activity and inhibition of DNA gyrase. J Agric Food Chem. 2013;61(34):8185–90.

42. Giep MA, Blood S, Larson MA, Koepsell SA. Hinrichs SH. Myricitrin Inhibits Escherichia coli DnaB helicase but not primase. Bioorg Med Chem. 2007;15(22):7203–8.

43. Ono K, Nakane H, Fukushima M, Chernmann JC, Barre-Sinoussi F. Differential inhibitory effects of various flavonoids on the activities of reverse transcriptase and cellular DNA and RNA polymerases. Eur J Biochem. 1990;190(3):469–76.

44. Cai L, Wu CD. Compounds from Syzygium aromaticum possessing growth inhibitory activity against oral pathogens. J Nat Prod. 2013;72(2):2706–14.

45. Arita-Morioka K-I, Yamanaka K, Mizunoe Y, Ogura T, Sugimoto S. Novel compounds from Syzygium aromaticum. J Agric Food Chem. 2006;54(12):4949–54.

46. DeSouza L, Wahidullah S. Antibacterial phenolics from the mangrove Lumnitzera racemosa. Curr Med Chem. 2010;17(22):2711–5.

47. Yadav AK, Than J, Prakash O, Khan F, Saikia D, Gupta MM. Screening of flavonoids for the inhibition of HIV-1 activity. J Acupunct Meridian Stud. 2018;11(4):234–40.

48. Yu M-S, Lee J, Lee JM, Kim Y, Chin Y-W, Jeon J-G, Keum Y-S, Jeong Y-J. Identification of myricetin and scutellarein as novel chemical inhibitors of the SARS coronavirus helicase, nS1P. Bioorg Med Chem Lett. 2012;22(12):4049–54.

49. Guitard R, Paul J-F, Aubry J-M. Myricetin, rosmarinic and caffeic acids inhibit microsomal benzphetamine hydroxylation: molecular modeling studies. Bioorg Med Chem. 2007;15(12):3917–23.

50. Alvesalo J, Vuorela H, Tammela P, Leinonen M, Saikku P, Vuorela P. Protective effect of flavonoids against reactive oxygen species and vasodilatory properties of Mediterranean food: the case of Cannonau wine, myrtle berries liqueur and strawberry-tomato honey. Food Chem. 2013;140(4):688–91.

51. Griep MA, Blood S, Larson MA, Koepsell SA, Hinrichs SH. Myricitrin Inhibits Escherichia coli DnaB helicase but not primase. Bioorg Med Chem. 2007;15(22):7203–8.

52. Jayaraman P, Sakharkar MK, Lim CS, Tang TH, Sakharkar KR. Activity and characterization of a novel myricetin-based antibiotic myricetin ether derivative. J Nat Prod. 2006;29(8):1751–6.

53. Chu S-C, Hsieh Y-S, Lin J-Y. Inhibitory effects of flavonoids on Moloney murine leukemia virus in a dual-chamber in vitro model. PLoS One. 2014;9(12):e115323.

54. Cetin-Karaca H, Newman MC. Antimicrobial efficacy of plant phenolic compounds against Salmonella and Escherichia coli. Food Sci Biotechnol. 2015;118–16.

55. Arita-Morioka K-I, Yamanaka K, Mizunoe Y, Ogura T, Sugimoto S. Novel compounds from Myrica rubra Sieb. & Zucc. leaves. PLoS One. 2014(9)(3):e96162.

56. Chen W, Li Y, Li J, Han Q, Ye L, Li A. Myricetin affords protection against ROS and fibril-destabilizing effects for the preservation of omega-3 oils. Food Chem. 2016;213:284–90.

57. Kang KA, Wang ZH, Zhang R, Piao MJ, Jo SH, Kim JS, Kang SS, Lee JS, Park DH, Hyun JW. Myricetin suppresses oxidative stress-induced cell damage via both direct and indirect antioxidant action. Environ Toxicol Pharmacol. 2010;29(1):12–8.

58. Sadasivam K, Kumaresan R. Antioxidant behavior of mearnsetin and rosmarinic acid: antioxidant activity and inhibition of nitric oxide production in RAW 264.7 cells. Food Sci Biotechnol. 2009;18(3):585–9.

59. Vauzour D, Vafeiadou K, Rodriguez-Mateos A, Rendeiro C, Spencer JP. The neuroprotective potential of flavonoids: a multiplicity of effects. Genes Nutr. 2010;5(3):154–66.

60. Tuberoso C, Cig Bo, Dobopik J, Dusim D, Pirisi FM. Antioxidant capacity and vasodilatory properties of Mediterranean food: the case of Cannonau wine, myrtle berries liqueur and strawberry-tomato honey. Food Chem. 2013;140(4):688–91.

61. Naldi M, Fiori J, Pistolozzi M, Andrisano V. Disclosure of a fundamental clue for the elucidation of the myricitrin mechanism of action as amyloid aggregation inhibitor by mass spectrometry. Electrophoresis. 2012;33(22):3380–6.

62. Naldi M, Fiori J, Bartolini M, Andrisano V. Disclosure of a fundamental clue for the elucidation of the myricitrin mechanism of action as amyloid aggregation inhibitor by mass spectrometry. Electrophoresis. 2012;33(22):3380–6.

63. Ono K, Yamada M. Antioxidant compounds have potent anti-fibrillogenic and fibril-destabilizing effects for α-synuclein fibrils in vitro. J Neurochem. 2006;97(1):105–15.
104. Kraegen EW, Bruce C, Hegarty BD, Ye JM, Turner N, Cooney G. AMP-kinase signalling and the regulation of glucose and lipid metabolism. Nature. 2001;414(6865):799–806.

105. Kang S-J, Park J-HY, Choi H-H, Kim J-H. α-glucosidase inhibitory activities of myricetin in animal models of diabetes mellitus. Food Sci Biotechnol. 2015;24(5):1897–900.

106. Karunakaran U, Elumalai S, Moon JS, Jeon J-H, Kim ND, Park K-G, Won KC. Urolithiasis: a potential target for the use of myricetin. Alternat Med. 2012;2012:758097.

107. Khan E, Tawani A, Mishra SK, Verma AK, Upadhyay A, Kumar M, Sandhir R, Mishra A, Kumar A. Myricetin reduces toxic level of CAG repeats RNA in HD patients: a novel therapeutic strategy. Mol Diagn Ther. 2018;22(3):281–90.

108. Liu I-M, Tzeng T-F, Liou S-S, Chiou J-I. Myricetin ameliorates defective post-receptor insulin signaling in the skeletal muscles of fructose-fed rats. Endocrinology. 2012;153(11):5363–9.

109. Li Y, Ding Y. Myricetin: a potent inhibitor of human flap endonuclease 1, which potentially can be used as sensitizing agent against HT-29 human colon cancer cells. J Funct Foods. 2016;22:325–32.

110. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

111. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

112. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

113. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

114. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

115. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

116. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

117. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

118. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

119. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

120. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

121. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

122. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.

123. Lin TL, Mu L, Chen L. Flavonoid myricetin modulates receptor activity through activation of α7-nicotinic acetylcholine receptor. J Nutr Sci Vitaminol. 2015;61(4):295–301.
129. Huang H, Chen AY, Ye X, Li B, Rojas-Asanaka Y, Rankin GO, Chen YC. Myricetin inhibits proliferation of cisplatin-resistant cancer cells through a p53-dependent apoptotic pathway. Int J Oncol. 2015;47(4):1404–9.

130. Xu Y, Xie Q, Wu S, Yi D, Yu Y, Liu S, Li S, Li Z. Myricetin induces apoptosis via endoplasmic reticulum stress and DNA double-strand breaks in human ovarian cancer cells. Mol Med Rep. 2016;13(5):2099–100.

131. Zheng AW, Chen YQ, Zhao LQ, Feng JG. Myricetin induces apoptosis and enhances chemosensitivity in ovarian cancer cells. Oncol Lett. 2017;13(6):4974–8.

132. Tavzan Z, Kayali HA. Flavonoids showed antioxidant effects on the ovarian cancer cells: involvement of reactive oxygen species, apoptosis, cell cycle and invasion. Biomed Pharmacother. 2019;109:000.

133. Lee KW, Kang NJ, Rogozin EA, Kim H-G, Cho YY, Bode AM, Lee HJ, Suri Y-I, Bowden GT, Dong Z. Myricetin is a novel natural inhibitor of neoplastic cell transformation and MEK1. Carcinogenesis. 2007;28(9):1918–27.

134. Kang NJ, Jung SK, Lee KW, Lee HJ. Myricetin is a potent chemopreventive phytochemical in skin carcinogenesis. Ann N Y Acad Sci. 2011;1229(1):124–32.

135. Sun W, Tao Y, Yu D, Zhao T, Wu L, Yu W, Han W. Myricetin exerts potent anticancer effects on human skin tumor cells. Trop J Pharm Res. 2018;17(6):1067–72.

136. Shih Y-W, Wu P-F, Lee Y-C, Shi M-D, Chiang T-A. Myricetin suppresses invasion and migration of human lung adenocarcinoma A549 cells: possible mediation by blocking the ERK signaling pathway. J Agric Food Chem. 2009;57(9):3490–9.

137. Zhang S, Wang L, Liu H, Zhao G, Ming L. Enhancement of recombinant myricetin on the radiosensitivity of lung cancer A549 and H1299 cells. Diagn Pathol. 2014;9(1):68.

138. Wang L, Feng J, Chen X, Gou W, Du Y, Wang Y, Zang W, Zhang S, Zhao G. Myricetin enhance chemosensitivity of S-fluorocouls on esophageal carcinoma in vitro and in vivo. Cancer Cell Int. 2014;14(1):71.

139. Sun F, Zheng XY, Ye J, Wu TT, Wang JL, Chen W. Potential anticancer activity of myricetin in human T24 bladder cancer cells both in vitro and in vivo. Nutr Cancer. 2012;64(4):599–606.

140. Ghassemi-Rad J, Maleki M, Knickel AF, Hoskin DW. Myricetin-induced oxidative stress suppresses murine T lymphocyte activation. Cell Biol Int. 2018;42(8):1069–75.

141. Jiménez R, Andriambeloson E, Duarte J, Andriantsitohaina R, Jiménez J, Pérez-Vázquez F, Zarruolo A, Tamargo J. Involvement of thromboxane A2 in the endothelium-dependent contractions induced by myricetin in rat isolated aorta. Br J Pharmacol. 1999;127(7):1539–44.

142. Cho Y-C, Yoon G, Lee KY, Choi HJ, Kang BY. Inhibition of interleukin-2 activity of myricetin in mouse EL-4 T cells. Arch Pharm Res. 2007;30(9):1075–9.

143. Lee SE, Park YS. Gene expression profiling of human umbilical vein endothelial cells exposed to myricetin. Biochip J. 2013;7(4):335–43.

144. Liu T-W, Wang L, Lo Y-H, Huang F-J, Wu M-J. Fisetin, morin and myricetin attenuate CD36 expression and oxLDL uptake in U937-derived macrophages. Biochim Biophys Acta. 2008;1781(10):601–9.

145. Angeleno T, Pasqua T, Di Majo D, Quintieri A, Ficile E, Armodio N, Tota B, Giannanco M, Cerra M. Distinct signalling mechanisms are involved in the dissimilar myocardial and coronary effects elicited by quercetin and myricetin, two red wine flavonoids. Nutr Metab Cardiovasc Dis. 2011;21(5):662–71.

146. Bhatia G, Khanna A, Sonkar R, Mishra S, Srivastava S, Lakshmi V. Lipid lowering and antioxidant activity of flavonoids in triton treated hypolipidemic rats. Med Chem Res. 2011;20(9):1622–6.

147. Tiwari R, Mohan M, Kasture S, Maxia A, Ballero M. Cardioprotective potential of myricetin in isoproterenol-induced myocardial infarction in Wistar rats. Phytother Res. 2009;23(10):1361–6.

148. Scarabelli TM, Mariotto S, Abdel-Azeim S, Shoji K, Darra E, Stephanou A, Chen-Scarabelli C, Marechal JD, Knight R, Ciampa A. Targeting STAT1 by flavonoids showed anticancer effects on the ovarian cancer cells: involvement of reactive oxygen species, apoptosis, cell cycle and invasion. Biomed Pharmacother. 2019;110:000.

149. Hagenacker T, Hillbrand I, Wissmann A, Büsselberg D, Schäfers M. Anti-allodynic effect of the flavonoid myricetin in a rat model of neuropathic pain: involvement of p38 and protein kinase C mediated modulation of Ca2+ channels. Eur J Pain. 2010;14(10):902–8.

150. Godse S, Mohan M, Kasture V, Kasture S. Effect of myricetin on blood pressure and metabolic alterations in fructose hypertensive rats. Pharm Biol. 2010;48(5):494–8.

151. Moghadam S, Ebrahimi S, Salehi P, Moridi Farimani M, Hamburger M, Jabbarzadeh E. Wound healing potential of chlorogenic acid and Myricetin-3-O-β-Rhamnose isolated from Parrotia persica. Molecules. 2017;22(9):1501.

152. Sun J, Sun J, Zhou X. Protective functions of myricetin in LPS-induced cardiomyocytes H9c2 cells injury by regulation of MALAT1. Eur J Med Res. 2019;24(1):20.

153. Zhu S, Yang C, Zhang L, Wang S, Ma M, Zhao J, Song Z, Wang F, Qu X, Li F, et al. Development of M10, myricetin-3-O-beta-d-lactose sodium salt, a derivative of myricetin as a potent agent of anti-chronic colonic inflammation. Eur J Med Chem. 2019;174(9):15–19.

154. Abidov M, Ramazanov A, Jimenez Del Rio M, Chkhikvishvili I. Effect of Blueberry on fasting glucose, C-reactive protein and plasma aminotransferases, in female volunteers with diabetes type 2: double-blind, placebo controlled clinical study. Georgian Med News. 2006;141:66–72.

155. Ahrens MJ, Thompson DL. Effect of emulin on blood glucose in type 2 diabetics. J Med Food. 2013;16(3):211–5.

156. Devi KP, Rajavel T, Habtemariam S, Nabavi SF, Nabavi SM. Molecular mechanisms underlying anticancer effects of myricetin. Life Sci. 2015;142:19–25.

157. Nektet P, Kumpulainen J, Larviinen R, Rissanen H, Heliovaara M, Reunanen A, Hakulinen T, Aromaa A. Flavonoid intake and risk of chronic diseases. Am J Clin Nutr. 2002;76(3):560–8.

158. Gates MA, Tworoger SS, Hecht JL, De Vivo I, Rosner B, Hankinson SE. A prospective study of dietary flavonoid intake and incidence of epithelial ovarian cancer. Int J Cancer. 2007;121(10):2225–32.

159. Bobe G, Weinstein SJ, Albanes D, Hinvenon T, Ashby J, Taylor PR, Virmanto J, Stolzenberg-Solomon RZ. Flavonoid intake and risk of pancreatic cancer in male smokers (Finland). Cancer Epidemiol Biomark Prev. 2008;17(3):553–62.

160. Tang NP, Zhou B, Wang B, Yu RB, Ma J. Flavonoids intake and risk of lung cancer: a meta-analysis. Jpn J Clin Oncol. 2009;39(6):352–9.

161. Zemm TL, Wood RJ, Greene C, West KL, Liu Y, Aggarwal D, Shachter NS, Fernandez ML. Grape polyphenols exert a cardioprotective effect in pre-and postmenopausal women by lowering plasma lipids and reducing oxidative stress. J Nutr. 2005;135(8):1911–7.

Publisher's Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.
Author/s:
Taheri, Y; Suleria, HAR; Martins, N; Sytar, O; Beyatli, A; Yeskaliyeva, B; Setimova, G; Salehi, B; Semwal, P; Painuli, S; Kumar, A; Azzini, E; Martorell, M; Setzer, WN; Maroyi, A; Sharifi-Rad, J

Title:
Myricetin bioactive effects: moving from preclinical evidence to potential clinical applications

Date:
2020-08-01

Citation:
Taheri, Y., Suleria, H. A. R., Martins, N., Sytar, O., Beyatli, A., Yeskaliyeva, B., Setimova, G., Salehi, B., Semwal, P., Painuli, S., Kumar, A., Azzini, E., Martorell, M., Setzer, W. N., Maroyi, A. & Sharifi-Rad, J. (2020). Myricetin bioactive effects: moving from preclinical evidence to potential clinical applications. BMC COMPLEMENTARY MEDICINE AND THERAPIES, 20 (1), https://doi.org/10.1186/s12906-020-03033-z.

Persistent Link:
http://hdl.handle.net/11343/252542

File Description:
Published version

License:
CC BY