Conformational Plasticity in Broadly Neutralizing 
HIV-1 Antibodies Triggers Polyreactivity

Highlights
- Most HIV-1 bNAbs are polyreactive and often cross-react with self-antigens
- Polyreactivity of bNAbs involves hydrophobic interactions
- Mutations enhancing HIV-1 bNAbs’ capacities also induce de novo polyreactivity
- Enhanced neutralization and polyreactivity co-emerge by antibody conformational flexibility

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In Brief
HIV-1 bNAbs are frequently polyreactive and bind to self-antigens. Prigent et al. show that specific mutations in bNAbs that enhance their neutralizing capacity create an intrinsic structural flexibility of the antibody paratope. This promotes the conformational adaptation that facilitates binding to HIV-1 variants and polyreactivity to topologically distinct non-HIV-1 molecules.
Conformational Plasticity in Broadly Neutralizing HIV-1 Antibodies Triggers Polyreactivity

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SUMMARY

Human high-affinity antibodies to pathogens often recognize unrelated ligands. The molecular origin and the role of this polyreactivity are largely unknown. Here, we report that HIV-1 broadly neutralizing antibodies (bNAbs) are frequently polyreactive, cross-reacting with non-HIV-1 molecules, including self-antigens. Mutating bNAb genes to increase HIV-1 binding and neutralization also results in de novo polyreactivity. Unliganded paratopes of polyreactive bNAbs with improved HIV-1 neutralization exhibit a conformational flexibility, which contributes to enhanced affinity of bNAbs to various HIV-1 envelope glycoproteins and non-HIV antigens. Binding adaptation of polyreactive bNAbs to the divergent ligands mainly involves hydrophobic interactions. Plasticity of bNAbs’ paratopes may, therefore, facilitate accommodating divergent viral variants, but it simultaneously triggers promiscuous binding to non-HIV-1 antigens. Thus, a certain level of polyreactivity can be a mark of adaptable antibodies displaying optimal pathogens’ recognition.

INTRODUCTION

The humoral arm of the immune system relies on the remarkable variety of immunoglobulin molecules (or antibodies) expressed as receptors on B cells or secreted in body fluids, which ensure the recognition of a theoretically infinite number of foreign antigens. This diversity results mainly from the random rearrangement of germline V(D)J genes coding for the immunoglobulin heavy- and light-chain variable domains (Tonegawa, 1983), which associate to form the antigen-binding site or paratope. The primary antibody repertoire is further broadened by the conformational malleability of germline-encoded paratopes that permits low-affinity-binding adaptation to structurally distinct molecules (Kaur and Salunke, 2015). Adaptive immune responses naturally occurring upon infection or induced by vaccination generate high-affinity antibodies to culprit antigens and memory B cell subsets to respond to further antigenic challenges. Upon antigen encounter in secondary lymphoid organs, special microenvironments called germinal centers (GCs) form and support the clonal expansion, maturation, and affinity-based selection of B cells (Victora and Nussenzweig, 2012). In GCs, B cells undergo somatic hypermutation (SHM) of immunoglobulin (Ig) genes that potentially increases antibody affinity to the target antigen (Victora and Nussenzweig, 2012). Concomitantly, antibody paratopes of matured B cells tend to lose the flexibility characterizing polyspecific unmutated B cell precursors and rigidify, allowing optimal contacts with cognate antigens (Jimenez et al., 2003; Kaur and Salunke, 2015; Wedemayer et al., 1997; Yin et al., 2003; Zimmermann et al., 2006). Class-switched memory B cells shaped during GC reactions to express high-affinity Ig can acquire poly- and self-reactivity, which are normally counterselected by tolerance mechanisms during B cell ontogeny (Goodnow et al., 2005; Koelsch et al., 2007; Prigent et al., 2016; Tiller et al., 2007; Wardemann and Nussenzweig, 2007). De facto, serological antibody polyreactivity is commonly associated with several human infections (Mouquet and Nussenzweig, 2012), and high-affinity human antibodies to diverse pathogens, including Plasmodium falciparum (Muellenbeck et al., 2013), influenza virus (Andrews et al., 2015), and HIV-1 (Liu et al., 2015; Mouquet and Nussenzweig, 2012; Mouquet et al., 2010), are also frequently polyreactive.
Figure 1. Poly- and Self-Reactivity of HIV-1 bNAbs

(A) Heatmap showing the antibody binding to the selected antigens as measured by ELISA (blue) and surface plasmon resonance (SPR) (red) in Figure S1. Color intensity is proportional to the reactivity level with darker colors indicating high binding while light colors show moderate binding (white, no binding). KLH, keyhole limpet hemocyanin; SS and DS, single- and double-stranded DNA; INS, insulin; LPS, lipopolysaccharide; TB, thyroglobulin; Lyso., lysozyme; p62-E1, Chikungunya virus (CHIKV) Env protein; IgG, human immunoglobulin G.

(B) Linear regression plot comparing the binding values obtained by ELISA and SPR for KLH alone (gray dots) and for all tested antigens (black dots). Polyreactive bNAbs are depicted in red.

(C) Schematic diagram of HIV-1 envelope glycoprotein showing the epitopes of polyreactive bNAbs.

(D) Antibody binding to HEp-2 cells assayed by immunofluorescence assay (IFA) with all bNAbs at 100 μg/mL.

(E) Titration of HEp-2 IFA reactivity as shown in (D).

(F) Graphs show ELISA binding curves of HIV-1 bNAbs against HEp-2 lysate antigens.

(G) Heatmap showing the IFA reactivity of HIV-1 bNAbs to antigens expressed by mouse kidney, liver, and stomach (Stom.).

(H) ELISA graphs show the binding of PGT128, m2, and m7 to purified Influenza hemagglutinin (HA, left) and Chikungunya virus (CHIKV) p62-E1 (right) glycoproteins.
Neutralizing antibodies to HIV-1 target the surface envelope glycoprotein (gp160) (Levy, 1998), and, among them, particular antibodies are able to neutralize most HIV-1 strains (Mouquet, 2014). Several unusual but essential Ig features characterize these HIV-1 broadly neutralizing antibodies (bNAbs), e.g., high load of SHM, nucleotide insertions or deletions (indels), long and tyrosine-rich CDR\_3, some of which are associated with antibody polyreactivity (Haynes et al., 2005; Liao et al., 2011, 2013). Polyreactivity may confer them with a selective advantage for viral binding (Mouquet et al., 2010) and neutralization (Alam et al., 2009; Mouquet et al., 2012b; Scherer et al., 2010). Immune tolerance control may, however, limit or block the development of bNAbs with exacerbated cross-reactivity to self-antigens (Chen et al., 2013; Kelsoe and Haynes, 2017; Schroeder et al., 2017; Vercoczy et al., 2010, 2011; Zhang et al., 2016). Importantly, bNAbs possess prophylactic and therapeutic capacities for preventing and treating HIV-1 infection (Mouquet, 2014), and they are thought to be mandatory mediators of potentially successful HIV-1 vaccines (Mouquet, 2015). Therefore, elucidating the molecular mechanisms linking the acquisition of bNAb activity with poly-/auto-reactivity is essential to better appreciate whether polyreactivity contributes to HIV-1 neutralization or is simply a bystander effect of affinity maturation. Notably, knowing whether the conformational flexibility typical of germline B cell precursors is also responsible for the dual recognition of HIV-1 Env and non-HIV-1 antigens by highly matured bNAbs is key in the comprehension of this phenomenon.

Here we show that most HIV-1 bNAbs are polyreactive. Certain bNAbs have Ig gene mutations enhancing HIV-1 cross-binding and -neutralization capacities but concomitantly creating de novo polyreactivity. Using binding affinity, thermodynamic, and molecular dynamics analyses, we report that the co-emergence of enhanced neutralizing capacities and polyreactivity is due to an intrinsic conformational flexibility of the antigen-binding sites of bNAbs, which allows a better accommodation of divergent HIV-1 Env variants.

RESULTS

High Prevalence of Polyreactive HIV-1 bNAbs Reacting with Self-Antigens

Polyreactivity is a common feature of HIV-1 antibodies (Mouquet et al., 2010), and it was also reported for some HIV-1 bNAbs (Haynes et al., 2009; Mouquet et al., 2012b; Scherer et al., 2010). Immune tolerance control may, however, limit or block the development of bNAbs with exacerbated cross-reactivity to self-antigens (Chen et al., 2013; Kelsoe and Haynes, 2017; Schroeder et al., 2017; Vercoczy et al., 2010, 2011; Zhang et al., 2016). Importantly, bNAbs possess prophylactic and therapeutic capacities for preventing and treating HIV-1 infection (Mouquet, 2014), and they are thought to be mandatory mediators of potentially successful HIV-1 vaccines (Mouquet, 2015). Therefore, elucidating the molecular mechanisms linking the acquisition of bNAb activity with poly-/auto-reactivity is essential to better appreciate whether polyreactivity contributes to HIV-1 neutralization or is simply a bystander effect of affinity maturation. Notably, knowing whether the conformational flexibility typical of germline B cell precursors is also responsible for the dual recognition of HIV-1 Env and non-HIV-1 antigens by highly matured bNAbs is key in the comprehension of this phenomenon.

Here we show that most HIV-1 bNAbs are polyreactive. Certain bNAbs have Ig gene mutations enhancing HIV-1 cross-binding and -neutralization capacities but concomitantly creating de novo polyreactivity. Using binding affinity, thermodynamic, and molecular dynamics analyses, we report that the co-emergence of enhanced neutralizing capacities and polyreactivity is due to an intrinsic conformational flexibility of the antigen-binding sites of bNAbs, which allows a better accommodation of divergent HIV-1 Env variants.
Figure 2. Poly- and Self-Reactivity of bNAb Variants

(A) Ribbon diagrams showing the crystal structures of NIH45-46 (PDB: 3U7W), VRC01 (PDB: 3NGB), and m66 variable domain (PDB: 4NRY). Amino acid residues distinguishing bNAb variants are depicted as yellow sticks. The complementary determining regions (CDRs) and framework regions (FRWs) in which these mutations are located are indicated on each structure.

(B) Polyreactive binding of rationally designed NIH45-46 antibody variants and controls (20 μg/mL) was assayed by immunoblot analyses on HEp-2 and E. coli protein extracts. Representative infrared scans from two independent experiments are shown.

(C) Table shows all combinations of three amino acid substitutions (IgHW47V, IgHG54W, and IgLS28Y) in NIH45-46 mutant antibodies (left). Heatmap compares the ELISA binding of the selected antibodies to various antigens as measured in Figure S2 (right). KLH, keyhole limpet hemocyanin; DS, double-stranded DNA; INS, insulin; LPS, lipopolysaccharide; TG, thyroglobulin.

(D) Immunoblotting on HEp-2 protein extract comparing the polyreactive binding of novel NIH45-46 mutant (mt1 to mt4) and control antibodies. Representative infrared scans from two independent experiments are shown.

(E) HEp-2 reactivity of NIH45-46 mt1-mt4 antibodies was evaluated by IFA. Representative IFA images from two independent experiments are shown.

(F) Same as for (B) but for VRC01/3 and mutant antibodies.

(G) Same as for (D) but for VRC01/3 and mutant antibodies.

(H) Heatmap comparing the ELISA binding of the selected antibodies to various antigens as measured in Figure S2.

(I) HEp-2 reactivity of m66 and m66.6 (100 μg/mL) was evaluated by IFA (top). Heatmap shows titration of HEp-2 IFA reactivity for m66 antibody variants (bottom).

(J) Binding of selected IgG bNAbs and corresponding Fabs to KLH was evaluated by SPR analysis as shown in Figure S2I. The y axis shows the response units (RUs) obtained at a given time (s, seconds) indicated on the x axis.

See also Figure S2.
Figure 3. HIV-1 Binding and Neutralization by bNAb Variants

(A) ELISA graphs comparing the binding of selected bNAbs to trimeric SS1196 and ZM96 gp140 glycoproteins. Error bars indicate the SEM of duplicate or triplicate OD405 nm values from 1 to 2 experiments.

(legend continued on next page)
escape mutant strains (Diskin et al., 2013) (Figure 2A). Strikingly, we observed a gradual increase of poly- and self-reactivity levels following the accumulation of substitutions in NIH45-46 (m7 > m2 > NIH45-46W > NIH45-46) (Figures 1 and S1). Immunoblot analyses on total HEp-2 and E. coli proteins also revealed increasing numbers of immune-reactive antigens (m7/m2 > NIH45-46W > NIH45-46) (Figures 2B and S2A), suggesting a simultaneous augmentation of polyreactivity strength and spectrum.

To identify the contribution of single versus combined substitutions (IgH^{W47V}, IgH^{S54W}, and IgL^{S28Y}) in enhanced polyreactivity, we generated a series of NIH45-46 variants (M1 to M4) comprising all combinations of unique and double mutations (Figure 2C). Neither IgL^{S28Y} and IgH^{W47V} alone nor combined promoted high polyreactive binding (Figures 2C–2E and S2D). Although 45-46 M1, which has both IgH mutations (W47V and G54W), had a slight increase of poly- and self-reactivity compared to NIH45-46W, the magnitude of binding was much lower than for m2 (IgH^{G54W} and IgL^{S28Y}) (Figures 2C–2E). Thus, the IgH^{G54W} mutation promoted the appearance of polyreactivity, but its association with IgL^{S28Y} in m2 amplified the binding to non-HIV-1 ligands, suggesting a synergetic effect of these two mutations for the induction of polyspecificity. NIH45-46 shares 85% and 52% IgH sequence identity (96% and 61% for IgL) with its clonal variants VRC01 and VRC03, respectively. Interestingly, compared to NIH45-46, VRC01 and VRC03 naturally possess IgL^{S28} and IgH^{W47} residues, respectively (Figure 2A). To test whether the IgH^{W54}/IgL^{S28} combination could confer polyreactivity to VRC01 and VRC03, we generated two new mutants, VRC01^{G54W} (VRC01^{W}) and VRC03^{S28Y} (VRC03^{*}). Interestingly, VRC01^{W}, but not VRC03^{*}, exhibited de novo poly- and auto-reactive binding compared to its wild-type counterpart (Figures 2F, 2G, S2E, and S2F). Thus, as previously shown for VRC07 (Rudicell et al., 2014), the IgH^{W47} mutation can potentiating for polyreactivity in NIH45-46 clonal variants, but only when IgH and IgL genes are not too divergent.

Anti-MPER m66 and m66.6 are clonally related antibody variants. Both antibodies differ in terms of neutralization potency and breadth but also poly- and self-reactivity; m66.6, but not m66, is a broad neutralizer and highly polyspecific (Zhu et al., 2011). We thus evaluated the polyreactive pattern of m66, m66.6, and three mutant antibodies with amino acid substitutions in the m66 CDR1 loop; m66^{H} (m66 IgL^{S28H}, m66^{46K} (m66 IgL^{S28H-G30K}), and m66^{SKK} (m66 IgL^{S28H-G30K-S31K}) (Figure 2A), which displayed intermediate neutralization activity (Ofek et al., 2014). We observed a gradual increase of poly-/self-reactivity according to the number of amino acid substitutions (m66.6 > m66^{SKK} > m66^{HK} > m66^{H} > m66) (Figures 2H, 2I, S2G, and S2H). SPR-binding experiments confirmed that both IgG and Fabs of polyreactive antibody variants have an increased apparent affinity to exogenous non-HIV ligands when compared to non-polyreactive bNAbs (Figures 2J and S2J).

In sum, polyreactivity qualitatively and quantitatively varies among members of bNAbs’ B cell lineages. This variation is due to a handful of IgH and IgL residues located outside the antigen-binding loops, which trigger polyreactivity when engrafted into non-polyreactive clonal variants.

**Polyreactivity Co-emerges with Enhanced Antibody Cross-Reactivity and -Neutralization of HIV-1**

Rationally designed NIH45-46 antibodies and m66 variants are known to possess superior Env-binding affinity and neutralization potency and breadth compared to their parental bNAbs (Diskin et al., 2011, 2013). To further evaluate the extent of enhanced HIV-1 Env cross-reactivity, we measured the binding of NIH45-46, VRC01/3, and m66 IgG variants to five different trimeric gp140 glycoproteins by ELISA. We observed a gradual increase of the apparent affinity to HIV-1 Env across antibody variants (Figures 3A, 3B, and S3A), which followed the magnitude of polyreactivity (Figure 2). For instance, augmented binding for NIH45-46 engineered variants (m7/m2 > NIH45-46W > NIH45-46) and VRC01^{W}, but not VRC03^{*}, was evidenced for 4 of 5 gp140 strains. Comparably, polyreactive m66 antibody variants (m66.6 > m66^{HK} > m66^{HK} and m66^{H} > m66) showed increased reactivity against 3 of the 5 Env proteins (Figures 3A, 3B, S3A, and S3B). SPR analyses performed on YU-2 and 2M96 trimer-immobilized chips confirmed the higher affinity to HIV-1 Env of most polyreactive IgG antibodies (Figures 3C and S3C). Interestingly, mutated variants of NIH45-46, VRC01, and m66, but not VRC03, also displayed enhanced binding to HIV-1 Env exposed at the surface of infected cells (Figures 3D, 3E, and S3D).

Augmented neutralization potency and breadth of NIH45-46^{W} and m2 and m7 were linked to an increased affinity and slower off-rates (Diskin et al., 2011, 2013). We asked whether VRC01^{W} exhibits enhanced neutralization properties against a cross-clade panel of 12 HIV-1 isolates, when compared to VRC01. VRC01^{W} was more potent (geometric mean IC_{50} =
0.40 versus 3.69 μg/mL for VRC01) and broader, with 100% of the panel’s viruses neutralized, while only 75% were sensitive to VRC01 (Figures 3F and S3E). On the other hand, the presence of the IgL S28Y mutation in VRC03 did not improve neutralization (geometric mean IC₅₀ = 7.87 ± 4.0 versus 8.65 ± 5.6 μg/mL for VRC03 and VRC03Y, respectively) (Figures 3F and S3E). Finally, combining already published and new in vitro neutralization data, we confirmed that m66.6 and m66HK have comparable neutralization activities, which largely surpassed the one of the m66 antibody (Figure 3G) (Ofek et al., 2014; Zhu et al., 2011).

Overall, our data show that the most polyreactive variants of various bNAbs also display broad Env cross-reactivity and enhanced neutralization breadth and potency.
Polyreactive bNAb Variants Display Conformational Adaptation upon HIV-1 Env Binding

To gain insights into the mechanisms of polyreactivity-associated HIV-1 Env cross-reactivity, we performed kinetic and thermodynamic analyses of monovalent interactions by SPR using bNAb Fab fragments. Real-time binding data revealed that NIH45-46, VRC01, and m66 variants interact with high affinity with both YU-2 and ZM96 HIV-1 gp140 trimers. In contrast, Fab fragments of m66 and its variants bound only to YU-2 gp140 (Figure 4A). Evaluation of the kinetic rate constants (k_a and k_d) and the equilibrium dissociation constant (K_D) confirmed that mutations that augmented polyreactivity also increased the affinity of bNAb Fab to YU-2 and ZM96 gp140 proteins (Figure 4B). For all bNAb variants, higher binding affinities were due to synergistic changes in the kinetic rate constants. Thus, compared to their wild-type counterparts, NIH45-46, VRC01, and m66 variants displayed both increased association to gp140 ligands and decreased dissociation (Figures 4A and 4B). We then performed kinetic analyses on the binding of bNAb Fab to the non-cognate antigen KLH. Improved affinity of polyreactive variants against KLH was mostly due to a faster association for m2, m66, and m66.6 and to a slower dissociation for NIH45-46_W (Figures 4B and S2I).

To examine more precisely the molecular mechanisms responsible for the dual recognition of HIV-1 and non-HIV antigens by prototypic bNAb and their polyreactive variants, we next performed thermodynamic analyses by measuring binding kinetics as a function of the temperature. We focused our analyses on the association phase, as it strongly depends on conformational changes in the interacting partners. For all bNAb Fab fragments binding to YU-2 trimers, an increase in temperature correlated with a concomitant rise in the association rates (Figures 4C and S4). Mutations amplifying bNAb polyreactivity had no influence on the temperature sensitivity of the association rate constant, as deduced by almost identical slopes of the Arrhenius plots (Figure 4C). However, antibodies exhibited a distinct behavior when binding to ZM96 gp140. The association rates of NIH45-46 and VRC01 Fab to ZM96 had a pronounced sensitivity to temperature, which is typical of interactions made by pre-formed binding sites (lock-and-key binding model; Manivel et al., 2000). Interestingly, polyreactive mutants of NIH45-46 and VRC01 lost sensitivity to elevated temperatures (Figure 4C).

We next derived activation energies and thermodynamic parameters that characterize the protein-protein interactions from Arrhenius plots. We observed identical qualitative changes of the activation enthalpy (ΔH^‡) and the activation entropy (ΔS^‡) during the association of wild-type and mutant Fabs to YU-2 gp140 (Figure 4D). ΔH^‡ and TΔS^‡ were respectively positive and negative, indicating that those changes had unfavorable contributions to the overall binding energy of all bNAb. Activation thermodynamic changes showed distinct trends when considering the antibody association to ZM96 trimers; NIH45-46 and VRC01 presented contrasting qualitative changes of TΔS^‡ as compared to their mutant counterparts. The interactions of parental antibodies with ZM96 were characterized by positive TΔS^‡, but polyreactive mutant bNAb displayed negative TΔS^‡ values (Figure 4D). Also, although changes in ΔH^‡ remained positive, absolute values for polyreactive mutants decreased about 3-fold compared to parental bNAb (Figure 4D). The detected high favorable values of TΔS^‡ (>40 kJ/mol) indicated that binding of NIH45-46 and VRC01 to ZM-96 Env protein was accompanied by an increase in the disorder of the system, most probably mediated by a disruption of ordered solvent shells surrounding interacting proteins. These changes in the association energetics are consistent with the interactions of rigid antibody-binding sites. In contrast, negative TΔS^‡ values observed for polyreactive bNAb suggest that, upon complex formation, the net disorder in the system decreases. This may be explained by a reduction in the intrinsic conformational freedom in the proteins, and it suggests the presence of conformational adaptations upon formation of the intermolecular complexes.

Hydrophobic Interactions Enhance Affinity of Polyreactive bNAb to HIV-1 Env

Changes in ΔH^‡ (the heat absorbed or released by the system) depend on the nature and the quantity of non-covalent contacts between interacting molecular surfaces. To provide a better explanation for the marked decrease of ΔH^‡ characterizing the binding of polyreactive bNAb to ZM96 HIV-1 Env trimers, we analyzed the non-covalent forces involved in the formation of the complexes. First, we compared the hydrophobicity of Fab fragments of bNAb and their variants by using a fluorescent probe. Mutations in bNAb, associated with both an increase in HIV-1 Env cross-reactivity and polyreactivity toward various non-HIV-1 antigens, did not change the overall hydrophobicity of the Fab (Figure 5A). We then analyzed the antibody binding to various HIV-1 Env and non-HIV polyreactive antigens as a function of the ionic strength. The interactions of IgG bNAb with gp120/gp140 glycoproteins and polyreactive ligands were sensitive to changes in the ionic environment, as evidenced by the decreased ELISA reactivity with increased salt concentrations (Figure 5B). This is consistent with the implication of polar bonds in the complexes’ formation. Of note, polyreactive variants were not sensitive to ionic strength or much less than their parental counterparts (Figure 5B). This suggests that hydrophobic interactions were the main driving force involved in the complexes’ formation. Thus, introducing mutations that allow for enhanced HIV-1 cross-binding and neutralization as well as de novo polyreactivity qualitatively changed the type of non-covalent forces from polar to non-polar. This may provide an explanation for the significant drop in ΔH^‡ values.

Collectively, these results suggest that mutations in bNAb augmenting cross-Env-binding affinity and neutralizing activity and polyreactivity are associated with enhanced structural dynamics of the antibody paratope. These mutations also change the type of non-covalent forces driving the formation of antigen-bNAb complexes.

Polyreactive HIV-1 bNAb with Enhanced Neutralization Display Paratope Conformational Flexibility

To better understand the conformational adaptation of certain bNAb upon binding to HIV-1 Env and non-HIV antigens, we explored changes in structural dynamics of parental and mutated antibody paratopes using molecular dynamic (MD) simulations. MD analyses performed in parallel for NIH45-46, NIH45-46_W, and m2 showed that introduction of the single
IgH_{54W} mutation decreased the total energy of the antibody structure (Figures 6A and S5A). Adding the second mutation in the light chain, IgL_{S28Y}, led to a drastic drop in the total energy of the molecular system (Figures 6A and S5A). Both mutations affected the molecular flexibility of the respective chains. An increase in the root-mean-square (RMS) values reflecting the dynamics of the IgH was noticed for NIH45-46W compared to the parental bNAb (2.40 versus 1.90 Å in NIH45-46), while the additional mutation in m2 increased the IgL RMS values from 1.44 to 1.66 Å (Figures 6B, 6C, and S5B). The overall RMS values considering IgH and IgL chains of NIH45-46 antibody variants were stable in the range of 2.2 Å (Figure 6B). The trajectory of the VRC01 W, which has the IgH_{G54W} mutation in association with a naturally present Tyrosine residue in position 28 of the IgL, closely resembled that observed for the analogous m2 antibody, with low total energies (E_{tot}/C_{24}/C_{0} \approx 9.5 \times 10^{4} \text{ kcal/mol}) and RMS values around 1.49 Å (Figures 6A–6C and S5). Here the IgH_{G54W} exchange resulted in an energetic drop (Figures 6A and S5A). This indicates that the major changes in the molecular dynamics were mainly due to the interplay between antibody variable domains, each with the mutated site. A similar tendency, albeit not as pronounced as for NIH45-46/VRC01, was observed for m66-derived antibodies. Both m66.6 and m66_{HKK} mutant exhibited higher total energies compared to m66 (~9 \times 10^{4} versus –10.5 \times 10^{4} \text{ kcal/mol}) (Figures 6A and S5A). This indicates that the major changes in the molecular dynamics were mainly due to the interplay between antibody variable domains, each with the mutated site. A similar trend, albeit not as pronounced as for NIH45-46/VRC01, was observed for m66-derived antibodies. Both m66.6 and m66_{HKK} mutant exhibited higher total energies compared to m66 (~9 \times 10^{4} versus –10.5 \times 10^{4} \text{ kcal/mol}) (Figures 6A and S5A). Moreover, the flexibility of the whole antibody reflected in the total RMS showed higher and similar values, respectively, for m66_{HKK} and m66.6 as compared to m66 (1.85 and 1.80 Å versus 1.64 Å, respectively) (Figures 6B, 6C, and S5). However, the m66.6 IgL exhibited a considerably lower flexibility (Figure S5; Table S1) than m66 and m66_{HKK}, suggesting that the additional mutations in m66.6 could counterbalance the flexibility effect induced by H_{28K30K31} mutations. Structural features further support this as the H_{28K30K31} mutations in the CDR_{H1} loop were located close to the CDR_{H2} loop harboring G_{50L52N53} residues, which also distinguished both antibody variants (Figure S6). In addition, the higher flexibility of m66.6 within the m66 family can be attributed to motional contributions of the common IgH.

We conclude that specific mutations create an intrinsic structural flexibility of the antigen-binding site of certain bNAbs, which promotes the conformational adaptation facilitating binding to HIV-1 Env variants and polyreactivity.

**DISCUSSION**

HIV-1 bNAbs are likely key effectors of protective HIV-1 vaccines. However, their induction by immunization remains difficult as they are naturally elicited from complex co-evolution host-pathogen processes, which are not yet fully resolved (Victora and Mouquet, 2018). Immune tolerance control represents one of the putative roadblocks preventing the selection of bNAb-expressing B cell clones when they express highly poly-/auto-reactive B cell receptors (BCRs) (Kelsoe and Haynes, 2017; Verkoczy et al., 2017).

In this study, we investigated the polyreactive properties of natural and artificially engineered HIV-1 bNAbs. Combining various immunoassays, we show that almost 60% of the tested HIV-1 bNAbs exhibited low to high polyreactivity. This is in agreement with a former estimation of polyreactive HIV-1 bNAbs (Liu et al., 2015; Verkoczy and Diaz, 2014). We further revealed a previously unappreciated polyreactive binding for PGT121, PGT128, NIH45-46W, m2, and m7. The magnitude of polyreactivity differed among antibodies from low to high binders, with only a few being highly cross-reacting with...
Autoantigens (e.g., 4E10, m2 and m7, and m66.6). Abnormally high polyreactivity was commonly found in rationally engineered and phage display-derived bNAbs, which could be explained by the absence of natural counterselection by in vivo tolerance mechanisms. Single mutations in more potent engineered variants of VRC07 and 10E8 have been previously shown to create de novo poly- and cross-reactivity (Kwon et al., 2018; Rudicell et al., 2014). This may limit the use of these types of antibodies as immunotherapeutics, due to their short half-life as evidenced in vivo (Rudicell et al., 2014; Shingai et al., 2013, 2014). However, strategies aiming at eliminating (e.g., by N-glycans engraftment) and selecting for a lack of binding promiscuity (by screening methods) gave rise to enhanced mutants with no or tolerable polyreactivity (Chuang et al., 2015; Jardine et al., 2016; Kwon et al., 2018), which could be more safely evaluated in clinical trials. In infected humans, B cells expressing poly- and self-reactive bNAbs could be proscribed by diverse tolerance mechanisms (Kelsoe and Haynes, 2017; Verkoczy et al., 2017). Thus, even if not pathogenic, self-reactivity of bNAbs could interfere with their development upon infection or vaccination (Kelsoe and Haynes, 2017; Verkoczy et al., 2017). Whether most HIV-1 bNAbs, which display no or only moderate promiscuous binding, are also subjected to immune tolerance control remains to be explored (Mouquet, 2014). Conversely, relaxed or defective tolerance may certainly facilitate bNAbs’ induction (Bonsignori et al., 2016; Schroeder et al., 2017). This led to envisioning various strategies to overcome tolerance control over bNAbs’ induction upon vaccination (Kelsoe and Haynes, 2017; Verkoczy et al., 2017). Strikingly, polyreactivity can naturally arise in parallel with enhanced HIV-1 binding and neutralization during affinity maturation, but the most potent antibody variants are not always polyspecific (Bonsignori et al., 2016; Liao et al., 2011, 2013). Hence, affinity-enhancing mutations do not always lead to polyreactivity, and, even so, compensatory mutations could possibly suppress polyreactivity and/or stabilize antibody structure (in case of a deleterious mutation), allowing affinity and breadth gain without counterselection. We showed nonetheless here that specific Ig mutations confer simultaneously enhanced HIV-1 binding affinity and neutralization and polyreactivity for bNAbs variants to the CD4bs and MPER epitopes. Similarly, hydrophobic residues of 2F5 and 4E10 CDR H3 loops interacting with membrane lipids, and likely involved in their polyspecificity, are required for neutralization (Alam et al., 2007, 2009; Scherer et al., 2010). Our data also show that particular mutations can supply bNAbs variants with a coincident gain of HIV-1 binding affinity, neutralization capacities, and promiscuous reactivity to numerous non-HIV-1 antigens. Thus, polyreactivity seems to be intimately linked to the functional properties of some HIV-1 bNAbs, as previously demonstrated with HIV-1 antibodies capable of hetero-agglutination (Mouquet et al., 2010). It will be of interest to determine whether the links between polyreactivity and neutralization that we describe here also occur with bNAbs targeting epitopes other than the CD4bs and MPER.

The capacity of germline-encoded B cell precursors to accommodate topologically dissimilar antigens with low affinity has been linked to the conformational flexibility of their antibody paratope (Adhikary et al., 2012; Khan and Salunke, 2014; Manivel et al., 2000, 2002; Thielges et al., 2008; Wedemayer et al., 1997). Conformational plasticity can also be found in affinity-matured antibodies, including antiviral bNAbs. One such antibody, H5.3, a potent H5N1 influenza neutralizer, undergoes conformational change upon binding to the H5 subunit, which also implies peripheral interactions to polymorphic residues (Winarski et al., 2015). Our thermodynamics and MD analyses suggest that intrinsic conformational flexibility enabled higher affinity variants of HIV-1 bNAbs to bind divergent HIV-1 Env and non-HIV antigens. Promiscuous binding to disparate
Our observations are reminiscent of those described for murine monoclonal antibodies to hen white-egg lysozyme (HEL). In these studies, the SHM-derived increased affinity of anti-HEL antibodies was associated with decreased thermostability and increased conformational flexibility of the binding sites, which formed plastic hydrophobic bonds with mimetic antigens (Acchione et al., 2009; Mohan et al., 2003, 2009; Sinha et al., 2002). Thus, we hypothesize that the paratope plasticity of HIV-1 bNAbs provides sufficient flexibility to accommodate subtle epitopic variations in divergent HIV-1 Env, as previously suggested for anti-HIV-1 gp41 peptide antibodies (Bhowmick and Salunke, 2013). For instance, conformational flexibility may help m66.6 accommodating different MPER conformers. During B cell ontogeny, increased affinity and flexibility of the antigen-binding site can be accompanied by a gain of auto-reactivity, as shown for HIV-1 bNAb 4E10 (Finton et al., 2014). Acquisition of poly- and auto-reactivity occurs during the affinity maturation toward neutralization breadth of both gp41- and gp120-specific B cells (Liao et al., 2011, 2013). This phenomenon could originate from the selection of clones with more plastic antigen-binding sites. On the other hand, in agreement with the paradigm of paratope rigidification upon affinity maturation (Jimenez et al., 2003; Manivel et al., 2000; Schmidt et al., 2013; Wedemayer et al., 1997; Zimmermann et al., 2006), the accumulation of SHM in HIV-1 bNAbs can optimize paratope complementarity with Env and restrict structural motility (Davenport et al., 2016). Therefore, affinity maturation of B cells likely selects for high affinity antibodies with either lock-and-key preconfigured binding sites or intrinsic flexibility by modeling paratope, as proposed earlier (Mohan et al., 2003). In summary, enhanced binding and neutralization abilities of HIV-1 bNAbs are often linked to the recognition of non-HIV-1 antigens. This promiscuous reactivity can be input into the binding sites’ flexibility, triggered by SHMs, which aid in accommodating not only divergent HIV-1 Env glycoproteins but also topologically distinct non-HIV-1 molecules via plastic hydrophobic structural interfaces. Thus, affinity maturation of HIV-1-reactive B cells toward neutralizing breadth may be tightly regulated between the selection of clones with adaptable antibody-binding sites for an expanded HIV-1 recognition and the counterselection by immune tolerance of highly polyreactive clones reacting with self-antigens. It is tempting to speculate that this phenomenon may occur with other pathogens with diversifying antigens, making antibodies’ natural development or elicitation by vaccines particularly difficult.

**Recombinant HIV-1 Proteins and Antibodies**

Recombinant human IgG1 antibodies, Fab molecules, and gp140 proteins were produced by co-transfection of FreeStyle 293-F cells (Fisher Scientific) using the polyethyleneimine (PEI)-precipitation method and purified as previously described (Lorin and Mouquet, 2015; Mouquet et al., 2012a). See the Supplemental Experimental Procedures for further details.

**Poly- and Auto-reactivity Assays**

Polyreactivity, HEP-2 and viral envelope-specific ELISAs, and HEP-2 IFAs were performed as previously described (Prigent et al., 2016). SPR-binding experiments were performed as previously described (Mouquet et al., 2010). Polyreactivity was also evaluated by infrared immunoblotting on HEP-2 cell and E. coli protein extracts. See the Supplemental Experimental Procedures for further details.

**Viral Binding and Neutralization Assays**

Antibody binding to viruses on infected cells was assayed by flow cytometry as recently described (Bruel et al., 2018). Neutralization of cell-free HIV-1 was measured using TZM-bl cells as previously described (Li et al., 2005; Lorin et al., 2017). To assay for CHIKV neutralization, luciferase-based and plaque neutralization assays were used, as described in the Supplemental Experimental Procedures.

**SPR Affinity and Thermodynamic Analyses**

The apparent affinity of HIV-1 bNAbs (IgG and Fabs) to KLH, YU-2, and ZM-96 gp140 trimers, CMS chips (Biacore) were measured as previously described (Mouquet et al., 2010). Thermodynamic parameters were determined using Eyring’s analyses as previously described (Hadzhieva et al., 2017). See the Supplemental Experimental Procedures for further details.

**Hydrophobicity Analyses of Fabs**

Hydrophobicity of Fabs was assessed using the 8-Anilinonaphthalene-1-sulfonic acid (ANS) fluorescence probe (Sigma-Aldrich, St. Louis, MO). Fabs (at 2 µM in PBS) was incubated for 15 min at room temperature in the absence or presence of 25 µM ANS. Fluorescence intensity of ANS was measured at 500 nm after excitation at 380 nm, with excitation and emission slits set at 9 and 20 nm, respectively. The fluorescence measurements were performed using a Tecan Infinite M200 Pro multimode plate reader (Tecan, Mannedorf, Switzerland). For evaluation of the specific signal of protein-bound ANS, the fluorescence intensities of ANS in the presence of Fabs were subtracted from the fluorescence of ANS in PBS only.

**Molecular Dynamic Simulations**

PDB coordinates were obtained from the Research Collaboratory for Structural Bioinformatics (RCSB). In all of the selected X-ray structures (PDB: 3U7W, 3NGB, 4NRY, and 4NRZ) (Diskin et al., 2011; Ofek et al., 2014; Wu et al., 2013), missing coordinates of flexible loop areas and of mutated residues were introduced using the program Modeler version (v.9.19) (Sali and Blundell, 1993). In each case, the structure with the lowest van der Waals violations was selected as the starting structure from a set of 10 conformers generated. Molecular dynamic simulations of the antibodies were then performed as described in the Supplemental Experimental Procedures.

**SUPPLEMENTAL INFORMATION**

Supplemental Information includes Supplemental Experimental Procedures, six figures, and one table and can be found with this article online at https://doi.org/10.1016/j.cell.2018.04.101.

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AUTHOR CONTRIBUTIONS

H.M. conceived the study. O.S., J.D.D., and H.M. designed experiments. J.P., A.J., C.P., C.E., J.D., A.K., V.L., D.V., T.C., T.B., M.S.S., O.O., J.D.D., and H.M. performed and analyzed experiments. O.S., O.O., J.D.D., and H.M. wrote the manuscript with contributions from all the authors.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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