N-n-Butyl Haloperidol Iodide Ameliorates Cardiomyocytes Hypoxia/Reoxygenation Injury by Extracellular Calcium-Dependent and -Independent Mechanisms

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N-n-butyl haloperidol iodide (F₂) has been shown to antagonize myocardial ischemia/reperfusion injury by blocking calcium channels. This study explores the biological functions of ERK pathway in cardiomyocytes hypoxia/reoxygenation injury and clarifies the mechanisms by which F₂ ameliorates cardiomyocytes hypoxia/reoxygenation injury through the extracellular-calcium-dependent and -independent ERK1/2-related pathways. In extracellular-calcium-containing hypoxia/reoxygenation cardiomyocytes, PKCα and ERK1/2 were activated, Egr-1 protein level and cTnI leakage increased, and cell viability decreased. The ERK1/2 inhibitors suppressed extracellular-calcium-containing-hypoxia/reoxygenation-induced Egr-1 overexpression and cardiomyocytes injury. PKCα inhibitor downregulated extracellular-calcium-containing-hypoxia/reoxygenation-induced increase in p-ERK1/2 and Egr-1 expression. F₂ downregulated extracellular-calcium-containing-hypoxia/reoxygenation-induced elevation of p-PKCα, p-ERK1/2, and Egr-1 expression and inhibited cardiomyocytes damage. The ERK1/2 activators antagonized F₂’s effects. In extracellular-calcium-free-hypoxia/reoxygenation cardiomyocytes, ERK1/2 were activated, LDH and cTnI leakage increased, and cell viability decreased. F₂ and ERK1/2 inhibitors antagonized extracellular-calcium-free-hypoxia/reoxygenation-induced Egr-1 overexpression and suppressed cardiomyocytes damage. The ERK1/2 activator antagonized F₂’s above effects. F₂ had no effect on cardiomyocyte cAMP content or PKA and Egr-1 expression. Altogether, ERK activation in extracellular-calcium-containing and extracellular-calcium-free hypoxia/reoxygenation leads to cardiomyocytes damage. F₂ may ameliorate cardiomyocytes hypoxia/reoxygenation injury by regulating the extracellular-calcium-dependent PKCα/ERK1/2/Egr-1 pathway and through the extracellular-calcium-independent ERK1/2 activation independently of the cAMP/PKA pathway or Egr-1 overexpression.

1. Introduction

The phenomenon of exacerbated tissue and organ damage produced by the restoration of blood flow after ischemia is known as ischemia/reperfusion (I/R) injury. Studies have demonstrated that this phenomenon takes place in a variety of tissues and organs such as the brain, heart, liver, lungs, kidneys, gastrointestinal tract, limbs, and skin. Myocardial I/R injury is a pathophysiological phenomenon commonly seen after ischemic heart disease and heart surgery. Reducing and eliminating this damage has become a hot topic in the field.

N-n-Butyl haloperidol iodide (F₂) is a new compound synthesized by our group. A series of previous studies have
shown that F₂ has protective effects on in vivo myocardial I/R injury and in vitro hypoxia/reoxygenation (H/R) injury models [1–4]. Our studies have shown that the F₂ protection is associated with antagonizing intracellular calcium overload through L-type calcium channels and inhibiting early growth response gene-1 (Egr-1) mRNA and protein overexpression [2, 5–7]. Further analysis has shown that F₂ is able to inhibit Egr-1 expression through suppression of the H/R-induced classical calcium-dependent PKCa translocation/activation. However, it can also activate calcium-independent PKCε translocation/activation to protect cardiomyocytes from sustaining H/R injury [8]. In addition, in cardiac microvascular endothelial cells, which do not have L-type calcium channels, F₂ still has a protective effect against H/R injury [6, 9–11]. These studies indicate that F₂ can protect cells from I/R injury through both calcium-dependent and -independent mechanisms. However, it is not clear which signaling pathways are involved.

The extracellular signal-regulated kinase (ERK1/2) pathway, which has attracted extensive attention in recent years, was the first signal transduction pathway of the MAPK family discovered. It is also the most extensively studied of signal transduction pathway [12]. It is not only involved in the regulation of a variety of cellular physiological functions but also plays an important role in the pathogenesis of a variety of diseases. Numerous studies have shown that the ERK1/2 signaling pathway is closely related to myocardial I/R and H/R injury [13]. Upon I/R or H/R stimulation, ERK1/2 is activated and transduced to the nucleus, phosphorylating serine, and threonine residues of transcription factors and leading to the activation and inactivation of gene transcription and subsequent changes in cell functions [12–14]. Moreover, it was reported that both the Ca²⁺-dependent and -independent pathways are necessary for elevating active ERK to a level sufficient to affect gene expression [15]. To explore the role of ERK1/2 in I/R and H/R injury, we first observed the change of ERK1/2 activity in cardiomyocytes after H/R in the presence and absence of extracellular calcium. Based on these results, we further investigated whether F₂ protection of cardiomyocytes from H/R injury might take place through its regulation of the calcium-dependent PKCa/ERK1/2/Egr-1 signaling pathway.

Both cAMP and Ca²⁺ are major second messengers. They not only crosstalk by downstream signal molecule but also transduce intracellular signal independently [16]. The cAMP-dependent PKA is the major downstream molecule in the cAMP signaling pathway. The cAMP/PKA activation has been shown to inhibit ERK1/2 activation in Rat-1 cells, NIH/3T3 cells, HEK-293 cells, and COS-7 cells [17–19]. In PC12 cells and S49 mouse lymphoma cells, cAMP/PKA acts as an upstream signal to activate ERK1/2 and affect cell function [20, 21]. In cardiomyocytes, the cAMP/PKA signaling pathway is also closely related to ERK1/2. After being activated by isoproterenol, β2AR activates the Gs/AC/cAMP/PKA pathways, consequently activates ERK1/2, and causes myocardial apoptosis [22, 23]. These results suggest that the calcium-independent cAMP/PKA/ERK1/2/Egr-1 pathway may be related to H/R-induced myocardial damage. Therefore, in this study, we focused simultaneously on whether the calcium-independent mechanism of F₂ protection is related to its regulation of the cAMP/PKA/ERK1/2/Egr-1 pathway.

2. Materials and Methods

2.1. Culture of Primary Cardiomyocytes. Adult Sprague-Dawley rats (200–250 g) were obtained from Vital River Laboratory Animal Technology Company (Beijing, China). The research protocol was approved by the Medical Animal Care and Welfare Committee of Shantou University Medical College and performed in compliance with the Guide for the Care and Use of Laboratory Animals (NIH Publication, 1996). Primary cardiomyocytes were cultured as described previously with minor modifications [2]. Briefly, neonatal ventricular cardiomyocytes were isolated from 1- to 4-day-old Sprague-Dawley rats with 0.1% trypsin. The dispersed cells were plated in M-199 medium containing 10% fetal bovine serum for 30 min to remove noncardiomyocytes. Then cardiomyocytes, representing 90–95% of total adhering cells, were cultured in the medium with 0.1 mM 5-bromodeoxyuridine for the first 4 days in an incubator with 5% CO₂ at 37 °C. Experiments were performed on day 4 or 5 of the culture.

2.2. Preparation of Reagents and Liquid. F₂ was synthesized in our laboratory. Verapamil was purchased from Shanghai Wellhope Pharmaceuticals (China); ERK inhibitor PD98059 was purchased from Promega (U.S.) and U0126 from Cell Signaling Technology (U.S.); ERK activator EGF was purchased from Pepprotech (U.S.); PKC-α inhibitor Gö6976 was purchased from Plymouth Meeting (U.S.); PKC-ε activator PMA, PKA inhibitor H89, and activator Forskolin were purchased from Sigma (U.S.). Anti-p-PKA, anti-total PKCa, anti-PKA, and cheliuminescence laminol reagents were purchased from Santa Cruz Biotechnology (U.S.); anti-p-ERK1/2, anti-total ERK1/2, and anti-Egr-1 were purchased from Cell Signaling Technology (U.S.); anti-β-actin and horseradish peroxidase-conjugated secondary antibodies were purchased from Wuhan Boster Biotechnology Limited Company (China); all the other chemicals and reagents were purchased from local agencies. Calcium-containing hypoxia solution was composed of the following: 137 mM NaCl, 12 mM KCl, 0.49 mM MgCl₂, 6mM Na₂HPO₄, 0.9 mM of CaCl₂, 4 mM HEPES, and 20 mM Na lactate. Calcium-free hypoxia solution was composed of the following: 137 mM NaCl, 12 mM KCl, 0.49 mM MgCl₂, 6mM Na₂HPO₄, 1 mM EGTA, 4 mM HEPES, and 20 mM Na lactate. The calcium absent reoxygenation solution was normal medium with 2 mM calcium-chelating EGTA added.

2.3. Establishment of Calcium-Containing (Normal Extracellular Calcium) and Calcium-Free (Lacking Extracellular Calcium) H/R Models and Experimental Groups. Cultured cardiomyocytes were randomly grouped (Figure 1). The calcium-containing-H/R model was established as described previously with 2-hour hypoxia instead of 3-hour hypoxia...
Figure 1: Protocols of experimental grouping and reagent administering. (a) Protocol used to investigate role of ERK1/2 and Egr-1 in extracellular-calcium-containing-H/R injury. (b) Protocol used to investigate role of PKC\(\alpha\)/ERK1/2/Egr-1 in extracellular-calcium-containing-H/R injury. (c) Protocol used to investigate role of ERK1/2 and Egr-1 in extracellular-calcium-free-H/R injury. (d) Protocol used to investigate role of cAMP/PKA in extracellular-calcium-free-H/R injury. CaCon, calcium-containing normoxia; CaH/R, calcium-containing H/R; 0CaCon, calcium-free normoxic control; 0CaH/R, calcium-free H/R.

[2]. F\(_2\) (1 × 10\(^{-6}\) mol/L), Ver (2 × 10\(^{-6}\) mol/L), inhibitors (PD98059 (2 × 10\(^{-5}\) mol/L), U0126 (2 × 10\(^{-5}\) mol/L) and Go6976 (1 × 10\(^{-6}\) mol/L)), and activators (EGF (50 ng/mL) and PMA (1 × 10\(^{-7}\) mol/L)) were given in normal medium (for preincubation), hypoxia solution, and/or reoxygenation medium, respectively. The calcium-containing normoxia (CaCon) group was replenished with fresh medium before the experiment and cultured for 3 hours.

The calcium-free-H/R model was established as before only with calcium-free hypoxia solution substituting for calcium-containing hypoxia solution and calcium-free medium for normal medium. F\(_2\), inhibitors (PD98059, U0126, and H89 (1 × 10\(^{-5}\) mol/L)) and activators (EGF and Forskolin (1 × 10\(^{-5}\) mol/L)) were also given as above. The calcium-free normoxic control (0CaCon) group was replenished with calcium-free medium before the experiment and cultured for 3 hours.

2.4. Western Blot Analysis. Total protein extracts were prepared from cultured cells using cell lysis buffer containing a protease inhibitor cocktail (aprotinin, leupeptin, pepstatin A, and PMSF). Western blot analysis was performed as described previously with some modifications [2]. The protein concentration was determined using a bicinchoninic acid assay (Bio-Rad, Hercules, CA, USA). Equal amounts of total protein were subjected to SDS-PAGE (10%) followed by electrophoretic transfer to nitrocellulose membranes. The nonspecific binding sites on the membrane were blocked with Tris buffer containing 5% nonfat dry milk for 1 hour. The membranes were probed with anti-p-PKC\(\alpha\), anti-total PKC\(\alpha\), anti-p-ERK1/2, anti-total ERK1/2, anti-Egr-1, anti-PKA, and anti-\(\beta\)-actin antibodies (1:5000 dilution for anti-\(\beta\)-actin, 1:1000 dilution for other antibodies) at 4°C overnight. Blots were then washed three times for 10 min with 20 mM Tris, 150 mM NaCl, and 0.1% Tween 20 (TBST) and incubated...
with horseradish peroxidase-conjugated secondary antibodies (1:5000) for 1 hour. The detection of immunoreactive bands was performed using Western blotting chemiluminescence luminol reagents. The relative densities of protein bands were quantitated using Gel-pro software of densitometric analysis (Media Cybernetics, USA).

2.5. Measurements of Cardiac Troponin I (cTnI) and Lactate Dehydrogenase (LDH) Levels in Conditioned Medium. The release of cTnI and LDH was detected in conditioned medium after reoxygenation. The levels of cTnI were measured using an ACS 180 Automated Chemiluminescence System (Bayer Corp., U.S.) with a two-site sandwich immunoassay kit (Bayer Corp., U.S.). The levels of LDH in conditioned medium were determined using test kits (Jiancheng Bioengineering Institute, Nanjing, China):

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LDH \text{ activity (U/L)} = \frac{(\text{OD of Assay} - \text{OD of Control})}{(\text{OD of Standard} - \text{OD of Blank})} \times \text{Standard Concentration (2} \mu\text{mol/mL)} \times 1000 \text{ mL.}
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2.6. Assessment of Cardiomyocyte Viability by Cell Counting Kit-8 (CCK-8) Colorimetric Assay. The cardiomyocytes were plated at 5 × 10^4 cells/well in 96 well plates. Then, 4-5 days later, the cells were treated as described previously. 10 μL of CCK-8 solution was added to 100 μL of reoxygenation solution and the cells were incubated for 1 additional hour after reoxygenation. The absorbance was measured by a microplate reader at 450 nm.

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\text{Cell viability} (%) = \frac{(\text{OD of Assay} - \text{OD of Blank})}{(\text{OD of Control} - \text{OD of Blank})} \times 100\%.
\]

2.7. Levels of cAMP in Cultured Cardiomyocytes. The concentration of cAMP in cultured cardiomyocytes was determined by ELISA using a commercially available kit (Enzo Life Sciences International Inc., Switzerland) according to the manufacturer’s instructions. All samples and standards were measured in duplicate. Briefly, cardiomyocytes were treated with 0.1 M HCl for 30 minutes and then harvested. After centrifugation, the supernatant was stored at −30°C for later analysis. Fifty microliters of standards and samples were added to a 96-well plate coated with GxR IgG antibody. Then, cAMP-conjugated alkaline phosphatase and cAMP antibody were added to all wells in sequence. After incubation for 2 hours on a plate shaker at 500 rpm, each well was washed three times with wash buffer and then incubated with substrate solution (p-nitrophenyl phosphate) for 1 hour. The reaction was stopped by addition of the stop solution (trisodium phosphate). The plate was read at 405 nm and the concentration of cAMP was calculated according to the standard curve.

2.8. Statistical Analysis. Data are shown as the mean ± SEM. The significance of differences was determined using one-way ANOVA followed the by Student-Newman-Keuls test. P < 0.05 was considered statistically significant.

3. Results

3.1. \( F_2 \) Inhibited Calcium-Containing-H/R-Induced ERK1/2 Activation and Consequently Reduced Egr-1 Protein Expression and cTnI Leakage and Improved Cell Viability in Myocardial Cells

3.1.1. Effects of \( F_2 \) on Calcium-Containing-H/R-Induced ERK1/2 Activation and Egr-1 Protein Expression. The ratio of p-ERK1/2 density to total ERK1/2 density reflects the degree of ERK activation. The ratio of total ERK density to β-actin density reflects total ERK protein level. The ratio of Egr-1 density to β-actin density reflects Egr-1 protein level. In each experiment, the density ratio in the CaCon group was set as 100% and the density ratio in other groups is here expressed relative to CaCon levels.

As shown in Figure 2, p-ERK1/2 and Egr-1 expression levels were significantly higher in the CaH/R group than in the CaCon group \((P < 0.05)\). p-ERK1/2 and Egr-1 expression levels were significantly lower in the CaH/R group than in the CaCon group \((P < 0.05)\). EGF activated ERK1/2 and Egr-1 expression but had no discernable effect on total ERK1/2 protein expression. EGF activated ERK1/2 under normoxia but did not affect Egr-1 expression. These results suggest that the ERK1/2 signaling pathway mediated calcium-containing-H/R-induced Egr-1 protein upregulation. \( F_2 \) inhibited Egr-1 expression by suppressing the ERK1/2 signaling pathway.

3.1.2. Influence of Inhibition of ERK1/2 Activation on Calcium-Containing-H/R-Induced Leakage of cTnI and Decrease of Cell Viability in Myocardial Cells. cTnI content in cultured cardiomyocyte supernatants was significantly higher and cell viability significantly lower in the CaH/R group than in the CaCon group \((P < 0.05)\). \( F_2 \), Verapamil, and ERK1/2 inhibitors U0126 and PD98059 significantly reduced cTnI content and improved cell viability \((P < 0.05)\). The ERK1/2 activator EGF was found to antagonize \( F_2 \)’s inhibition of cTnI leakage and improvement of cell viability \((P < 0.05)\). Under normoxic conditions, EGF had no effect on cTnI content or cell viability (Table 1).

3.1.3. Regulatory Role of \( F_2 \) on Calcium-Containing-H/R-Induced Abnormal PKCa/ERK1/2/Egr-1 Pathway. The PKCa inhibitor G66976 and activator PMA were used to clarify the effects of \( F_2 \) on the PKCa/ERK1/2/Egr-1 signaling pathway. The ratio of p-PKCa density to total PKCa density was used to determine the degree of PKCa activation, and the ratio of total PKCa density to β-actin density was used to determine
Figure 2: Effects of F₂, Verapamil, and ERK1/2 inhibitors and activator on p-ERK1/2, total ERK1/2, and Egr-1 expression in extracellular-calcium-containing myocardial H/R by western-blot assay. (a) p-ERK1/2 and total ERK1/2; (b) Egr-1 protein. Quantitative densitometric data were expressed as percentages of the level observed in the CaCon group. All values are expressed as mean ± SEM of at least six individual experiments. *P < 0.05 versus CaCon group; †P < 0.05 versus CaH/R group; ‡P < 0.05 versus CaH/R+F₂ group.
was no upstream-downstream correlation between ERK1/2 and Egr-1. F2 had no effect on Egr-1 protein expression in cardiomyocytes under calcium-free H/R injury conditions.

3.2.2. Effects of Inhibition of ERK1/2 Activation on Calcium-Free-H/R-Induced Leakage of LDH and cTnI and Decrease of Cell Viability. LDH and cTnI levels in cultured cardiomyocyte supernatants were significantly higher, and cell viability was significantly lower in the 0CaH/R group than in the 0CaCon group (P < 0.05). F2 and ERK1/2 inhibitors U0126 and PD98059 were found to significantly reduce LDH and cTnI concentration and improve cell viability (P < 0.05). The ERK1/2 activator EGF was found to antagonize F2 inhibition of LDH and cTnI leakage and improvement of cell viability (P < 0.05). Under normoxic conditions, EGF was found to have no effect on LDH or cTnI levels or on cell viability (Table 2).

3.2.3. Role of the cAMP/PKA Pathway in F2 Protection of Cardiomyocytes from Calcium-Free-H/R-Induced Injury. cAMP/PKA is involved in the regulation of myocardial cell function during I/R by acting as an upstream signaling molecule to activate the ERK1/2 signaling pathway [22]. Like Ca2+, cAMP is a transmembrane second messenger. It can be considered a noncalcium second messenger. In this study, we evaluated the effects of F2 on cAMP levels and PKA protein expression and examined the effects of PKA inhibitor H89 and activator Forskolin on LDH leakage in cardiomyocytes during calcium-free H/R. The density ratio of PKA to ß-actin was used to indicate PKA protein expression. The density of PKA in the 0CaCon group was set at 100%, and the density in other groups was calculated relative to these values.

CAMP levels were lower in the 0CaH/R group than in the 0CaCon group, but the difference was not statistically significant (P > 0.05). F2 was found to have no effect on cAMP levels in calcium-free H/R. No significant difference in PKA protein was observed between different groups (P > 0.05). LDH levels in cultured cell supernatants were

### Table 1: Effects of F2, Verapamil, and ERK1/2 inhibitors on cTnI level and cell viability in extracellular-calcium-containing myocardial H/R (n = 9).

| Group            | cTnI (ng/mL) | Survival rate (%) |
|------------------|-------------|-------------------|
| CaCon            | 0.22 ± 0.01 | 100.00            |
| CaH/R            | 0.61 ± 0.03* | 47.51 ± 2.15*     |
| CaH/R + F2       | 0.49 ± 0.02** | 64.23 ± 2.78**    |
| CaH/R + U0126    | 0.50 ± 0.03** | 63.52 ± 2.46**    |
| CaH/R + PD98059  | 0.48 ± 0.02** | 64.20 ± 2.21**    |
| CaH/R + Ver      | 0.47 ± 0.02** | 56.85 ± 2.24**    |
| CaCon + EGF      | 0.22 ± 0.01† | 93.34 ± 3.91†     |
| CaH/R + F2 + EGF | 0.59 ± 0.01† | 48.54 ± 3.56†     |

F2, N-n-butyl haloperidol iodide; cTnI: cardiac troponin I; H/R: hypoxia/reoxygenation. *P < 0.05 versus CaCon group; †P < 0.05 versus CaH/R + F2 group.

### Table 2: Effects of F2, and ERK1/2 inhibitors and activator on LDH level, cTnI level, and cell viability in extracellular-calcium-free myocardial H/R (n = 9).

| Group             | LDH (U/mL) | cTnI (ng/mL) | Survival rate (%) |
|-------------------|------------|-------------|-------------------|
| 0CaCon            | 968.65 ± 12.37 | 0.17 ± 0.01 | 100.00            |
| 0CaH/R            | 1342.35 ± 15.82* | 0.84 ± 0.03* | 50.66 ± 1.90*    |
| 0CaH/R + F2       | 1135.16 ± 22.33** | 0.62 ± 0.04** | 67.38 ± 2.94**   |
| 0CaH/R + U0126    | 1155.04 ± 24.24** | 0.69 ± 0.03* | 62.81 ± 4.58**   |
| 0CaH/R + PD98059  | 1261.39 ± 14.20** | 0.53 ± 0.02** | 57.73 ± 2.71**   |
| 0CaCon + EGF      | 1019.90 ± 17.16† | 0.17 ± 0.01† | 94.01 ± 4.70†    |
| 0CaH/R + F2 + EGF | 1421.43 ± 17.16† | 0.77 ± 0.05† | 53.17 ± 2.48†    |

F2, N-n-butyl haloperidol iodide; LDH: lactate dehydrogenase; cTnI: cardiac troponin I; H/R: hypoxia/reoxygenation. *P < 0.05 versus 0CaCon group; †P < 0.05 versus 0CaH/R + F2 group.
Figure 3: Effects of F₂, Verapamil, and PKCα inhibitor and activator on p-PKCα, total PKCα, p-ERK1/2, total ERK1/2, and Egr-1 expression in extracellular-calcium-containing myocardial H/R by western-blot assay. (a) p-PKCα and total PKCα protein levels; (b) p-ERK1/2 and total ERK1/2 protein levels; (c) Egr-1 protein levels. All values are expressed as mean ± S.E.M. of at least six individual experiments. *P < 0.05 versus CaCon group; †P < 0.05 versus CaH/R group; ‡P < 0.05 versus CaH/R+F₂ group.
4. Discussion

4.1. Effects of ERK1/2 Activation by H/R Stimulation on Cardiomyocyte Damage. The ERK1/2 pathway is an important cell signaling pathway. It can transfer extracellular information into the nuclei and mediate the ultimate cellular reaction. The studies have shown that the ERK1/2 signaling pathway is involved in I/R injury in a variety of tissues and organs, especially in myocardial tissue [13, 24–27]. In this study, we focused on the relationship between myocardial H/R injury and the ERK1/2 signaling pathway. Our results show that H/R stimulation activated ERK1/2 in both the presence and absence of calcium. This was demonstrated by increased levels of p-ERK1/2 and unchanged levels of total ERK. The ERK1/2-specific inhibitors U0126 and PD98059 effectively inhibited calcium-containing- and calcium-free-H/R-induced ERK1/2 activation, leading to reduced cell damage, as demonstrated by reduced levels of LDH and cTnI leakage, and improved cell viability. These indicate that ERK1/2 activation caused cardiomyocyte damage in both calcium-containing- and calcium-free-H/R stimulation.
Although studies have suggested that activation of the ERK1/2 pathway may promote cell survival [28, 29], our results have shown that the activation of ERK1/2 signaling pathway led to cell damage under myocardial H/R conditions. This result has been supported by many studies, such as the study conducted by Kang et al., which demonstrated that activated ERK1/2 induced by H/R might be novel drug target in cardiomyocytes [30], and the studies of Tsoporis et al. and Liu et al., where they found that the activation of ERK1/2-p53 signaling pathway caused cardiomyocyte apoptosis after myocardial infarction or administration of anticancer drug doxorubicin [31, 32]. In this way, ERK1/2 has a significant impact on the pathophysiological status of cells, but its role may be different in various cell types and experimental models. We believe that the ERK1/2 may play different roles during different stages of I/R: the activation of ERK1/2 in a very short period of I/R may initiate the endogenous protective processes, such as ischemic preconditioning, but when accompanied by the extension of I/R processes, ERK1/2 activation may initiate injury signals, leading to cell injury [33, 34].

4.2. Role of Egr-1 Expression Inhibition through the Calcium-Dependent PKCa/ERK1/2/Egr-1 Pathways on F2 Protection against Myocardial H/R Injury. Our preliminary results showed that blocking L-type calcium channels can inhibit calcium influx and reduce intracellular calcium overload, thereby inhibiting calcium-dependent PKCa activation and subsequent abnormal expression of Egr-1. This is one of the important calcium-dependent mechanisms underlying F2 protection from H/R-induced myocardial cell injury. However, it is not clear whether ERK mediates signal transduction between PKCa and Egr-1.

We found in one previous study that H/R could induce PKCa translocation from soluble fraction to particulate fraction in cardiomyocytes [8]. In the present study, we also found that H/R could activate PKCa by increasing its phosphorylation. We also observed that the PKCa inhibitor G66976 inhibited both p-ERK1/2 and Egr-1 protein overexpression, indicating that PKCa activation has an important impact on ERK1/2 activation and Egr-1 overexpression, which means that both ERK1/2 and Egr-1 are downstream molecules of PKCa signal pathway. Using the ERK1/2 inhibitors U0126 and PD98059, we found ERK1/2 to be an upstream signaling molecule of Egr-1. In the present study, we proved that H/R caused abnormal activation of PKCa/ERK1/2/Egr-1 pathway, leading to a series of cellular injuries.

In this study, p-PKCa, p-ERK1/2, and Egr-1 protein expression decreased after F2 treatment, but total PKCa and ERK1/2 protein expression did not change, suggesting that F2 can also inhibit PKCa and ERK1/2 activation in addition to suppressing Egr-1. The PKCa activator PMA can inhibit F2 downregulation of p-PKCa expression and downregulation of p-ERK1/2 and egr-1 protein expression, suggesting that F2 inhibition of ERK1/2 and Egr-1 is dependent on its inhibition of PKCa activation. The ERK1/2 activator EGF can inhibit F2 downregulation of p-ERK1/2 activation and Egr-1 protein expression and F2 protection of cardiomyocytes (including inhibition of cTnI leakage and improvement of cell viability), suggesting that F2 protection of cardiomyocytes is dependent on its inhibition of ERK1/2 activation and subsequent down-regulation of Egr-1 protein expression. In this way, we proved that F2 inhibition of ERK1/2 activation is PKCa-dependent and that F2 inhibition of Egr-1 overexpression is ERK1/2-dependent, suggesting that F2 protection of cardiomyocytes under H/R conditions takes place through its regulation of the abnormal PKCa/ERK1/2/Egr-1 signaling pathway. In addition, in this study, we used Verapamil as a positive control for the calcium antagonist and found that Verapamil, like F2, has an effect on p-PKCa, p-ERK1/2, and Egr-1 protein expression and protects cardiomyocytes from a series of H/R injuries. This suggests that both F2 and Verapamil can regulate the abnormal PKCa/ERK1/2/Egr-1 pathway, which might be initiated through the regulation of calcium. This hypothesis was supported by the fact that PKCa is a calcium-dependent kinase. Under normoxic conditions, Egr-1 expression was low and PMA treatment activated ERK1/2 but did not stimulate Egr-1 protein expression. Similarly, the ERK1/2 activator EGF did not cause Egr-1 protein overexpression or cell damage. These results suggest that the ERK1/2-related cell signaling network is very complicated and the specific intracellular microenvironment at H/R stimulation determines ERK1/2 downstream signaling and its ultimate functions.

4.3. Inhibition of ERK1/2 Activation Is One of the Extracellular Calcium-Independent Mechanisms for F2 Protection against Myocardial H/R Injury. Intracellular calcium overload is an important cause of I/R injury. Calcium antagonists can antagonize intracellular calcium overload and protect cardiomyocytes from I/R injury. However, Hempel et al. found that Nifedipine had no effect on ischemia-induced intracellular calcium increases in endothelial cells but it could prevent ischemia-induced PKC translocation and ameliorate increased ischemia-induced endothelial cell permeability [35]. A study performed by Eickelberg et al. showed that Amlodipine, Diltiazem, and Verapamil could regulate transcription factor NF-IL6 and NF-kB in an intracellular-calcium-independent manner [36]. These results indicate that calcium antagonists can affect cell function in ways other than blockage of calcium channels and affecting intracellular calcium levels. Our preliminary results also showed that F2, a new L-type calcium antagonist, can not only activate calcium-independent PKCa translocation through translocation in rat cardiomyocytes at the H/R stimulation and protect cardiomyocytes but also protect rat coronary endothelial cells, which do not have L-type calcium channels, from H/R injury [6, 8–11]. We therefore have reason to speculate that F2 can protect myocardial cells from H/R injury in an extracellular-calcium-independent manner.

Our results showed that F2 inhibited calcium-free H/R-induced ERK1/2 activation, leading to reduced LDH and cTnI leakage, and improved cell viability. The ERK1/2 activator EGF antagonized F2 inhibition of calcium-free H/R-induced p-ERK1/2 upregulation and inhibited F2 protection of cardiomyocytes. These data indicate that in calcium-free H/R,
F2 can act on ERK1/2 directly or its upstream signal molecule and protect cardiomyocytes from H/R injury. In other words, blocking H/R-induced ERK1/2 pathway activation is an extracellular-calcium-independent mechanism by which F2 protects cardiomyocytes. Under normoxic conditions, EGF was found to activate ERK1/2 in the presence or absence of calcium but it did not cause cardiomyocyte injury. However, ERK1/2 activation can cause cell injury in H/R, indicating that although ERK1/2 is a key mediator, it still needs other factors to cause cell injury.

In the study of ERK1/2-related pathways, our results showed that cAMP levels were lower in the 0CaH/R group than in the 0CaCon group, but this difference was not statistically significant and PKA levels did not change between the groups. In the 0CaH/R group, LDH leakage increased and the PKA activator Forskolin and inhibitor H89 had no effect on calcium-free-H/R-induced LDH leakage, suggesting that calcium-free-H/R-induced cardiomyocyte injury is not mediated by the cAMP/PKA pathway. We also observed that F2 had no effect on cAMP concentration or PKA protein expression in calcium-free-H/R stimulation, indicating that the extracellular-calcium-independent mechanism of F2 protection against H/R injury in cardiomyocytes is related to ERK1/2, but the upstream signaling molecule is not related to cAMP/PKA.

This study and our previous studies showed that the myocardial protective effects of F2 are related to the inhibition of I/R- and H/R-induced Egr-1 mRNA and protein overexpression. But the present study showed that calcium-free H/R did not cause Egr-1 protein upregulation and that F2 had no effect on Egr-1 protein expression in cardiomyocytes in calcium-free H/R. One possible explanation for this is that calcium is involved in the H/R-induced upregulation of Egr-1. We had found that three different types of calcium antagonists, Verapamil, Diltiazem, and Nifedipine, suppressed I/R- and H/R-induced Egr-1 mRNA and protein up-regulation to some extent [7]. Similarly, Lo et al. found that the calcium chelator BAPTA/AM completely inhibited hypoxia-induced Egr-1 overexpression in endothelial cells [37]. We previously observed H/R-induced Egr-1 overexpression and F2 protection and inhibition of Egr-1 in microvascular endothelial cells [9–11]. These cells lack L-type calcium channels. However, these specific cells were in an environment with calcium. We therefore speculate that H/R may cause extracellular calcium influx and ultimately Egr-1 overexpression due to counter-transportation of calcium by the Na+/Ca2+ exchanger (NCX) or membrane integrity destruction. F2 exhibited its regulation of Egr-1 expression and protective effect on microvascular endothelial cells through the inhibition of NCX outward currents and the subsequent reduction in calcium influx [38]. In this study, EGTA chelated all extracellular calcium and the intracellular-extracellular calcium gradient disappeared. All means of extracellular calcium influx were eliminated and so calcium-dependent Egr-1 overexpression and F2’s regulation to it became difficult in calcium-free H/R.

Although we observed abnormal ERK/Egr-1 pathway activity in calcium-containing H/R model, ERK1/2 inhibitors and activator had no effect on Egr-1 protein expression in calcium-free H/R. This indicates that ERK activation is a master switch to trigger myocardial H/R injury whether in extracellular calcium-containing or calcium-free H/R, nevertheless, its downstream signaling is also determined by the specific intracellular microenvironment such as intracellular calcium concentration. Thus, in the present study, we found that Egr-1 expression did not change with ERK activation when the influx of calcium was eliminated in calcium-free H/R. This result has been supported by Josefsen et al., who found that Egr-1 expression was dependent on Ca2+ influx [39]. Of course, other downstream signaling molecules can be involved and worthy of further study.

5. Conclusions
In summary, in cultured cardiomyocytes, both extracellular-calcium-containing and extracellular-calcium-free-H/R were found to activate ERK1/2, leading to cell damage. F2 was found to protect cardiomyocytes against H/R injury by regulating extracellular-calcium-dependent abnormal PKCa/ERK1/2/Egr-1 signaling pathway. F2 was also found to protect cardiomyocytes from H/R injury through extracellular-calcium-independent mechanisms, which may be related to its suppression of H/R-induced ERK1/2 activation but are not related to the cAMP/PKA signaling pathway or to Egr-1 protein expression.

Conflict of Interests
The authors declare that there is no conflict of interests.

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