Constitutive overexpression of the TaNF-YB4 gene in transgenic wheat significantly improves grain yield

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Abstract

Heterotrimeric nuclear factors Y (NF-Ys) are involved in regulation of various vital functions in all eukaryotic organisms. Although a number of NF-Y subunits have been characterized in model plants, only a few have been functionally evaluated in crops. In this work, a number of genes encoding NF-YB and NF-YC subunits were isolated from drought-tolerant wheat (*Triticum aestivum* L. cv. RAC875), and the impact of the overexpression of *TaNF-YB4* in the Australian wheat cultivar Gladius was investigated. *TaNF-YB4* was isolated as a result of two consecutive yeast two-hybrid (Y2H) screens, where ZmNF-YB2a was used as a starting bait. A new NF-YC subunit, designated *TaNF-YC15*, was isolated in the first Y2H screen and used as bait in a second screen, which identified two wheat NF-YB subunits, *TaNF-YB2* and *TaNF-YB4*. Three-dimensional modelling of a *TaNF-YB2/TaNF-YC15* dimer revealed structural determinants that may underlie interaction selectivity. The *TaNF-YB4* gene was placed under the control of the strong constitutive polyubiquitin promoter from maize and introduced into wheat by biolistic bombardment. The growth and yield components of several independent transgenic lines with up-regulated levels of *TaNF-YB4* were evaluated under well-watered conditions (*T*1–*T*3 generations) and under mild drought (*T*2 generation). Analysis of *T*2 plants was performed in large deep containers in conditions close to field trials. Under optimal watering conditions, transgenic wheat plants produced significantly more spikes but other yield components did not change. This resulted in a 20–30% increased grain yield compared with untransformed control plants. Under water-limited conditions transgenic lines maintained parity in yield performance.

Key words: Drought, grain yield, NF-Y genes, plant development, protein–protein interactions, wheat.

Introduction

The *Nuclear Factor Gamma* (*NF-Y*) gene family in plants has recently generated much interest due to involvement of family members in plant performance, development, and stress regulation (Petroni et al., 2012; Laloum et al., 2013). NF-Y transcription factors (TFs), also known as histone- or haem-associated proteins (HAPs) or CCAAT-binding factors (CBFs), are heterotrimeric DNA-binding proteins, structurally conserved in all eukaryotes (Mantovani, 1999; Romier et al., 2003). They bind specifically to the CCAAT box (Sinha et al., 1995; Bi et al., 1997), found in ~25% of eukaryotic promoters (Suzuki...
NF-Y TFs are composed of three different subunits named NF-YA (also known as HAP2 or CBF-B), NF-YB (HAP3 or CBF-A), and NF-YC (HAP5 or CBF-C). The NF-Y subunits B and C contain a histone-fold domain (HFDs) structurally related to histones H2B and H2A, respectively (Baxevanis et al., 1995). These domains mediate formation of a stable, histone-like heterodimer (Romier et al., 2003). The NF-Y subunit A subsequently binds to the B–C heterodimer to form a heterotrimeric complex, which can specifically recognize the CCAAT box (Sinha et al., 1995).

While most eukaryotic genomes contain only one or two genes encoding each NF-Y subunit (Maity and de Crombrugghe, 1998), NF-Y subunits of vascular plants are generally encoded by gene families with at least 10 members (Riechmann et al., 2000; Cagliari et al., 2014). In hexaploid wheat (Triticum aestivum L.), a total of 35 NF-Y genes (10 NF-YA, 11 NF-YB, and 14 NF-YC) have been identified (Stephenson et al., 2007), and some of these have been functionally characterized (Stephenson et al., 2010, 2011). Recently the total number of identified NF-Y genes in wheat reached 80 (Qu et al., 2015). Since the whole-genome sequence of wheat is not yet completed, the exact number of wheat NF-Y genes remains unknown. It was proposed that the opportunity for formation of a large number of heterotrimeric combinations in plants provides substantial plasticity for the involvement of NF-Y complexes in regulation of multiple developmental processes (Edwards et al., 1998).

Subunits of NF-Y TFs were shown to be involved in development and stress responses in different plant species (Edwards et al., 1998; Lotan et al., 1998; Kwong et al., 2003; Lee et al., 2003; Cai et al., 2007; Chen et al., 2007; Nelson et al., 2007; Yazawa and Kamada, 2007; Kumimoto et al., 2008, 2010; Yamamoto et al., 2009; Liu and Howell, 2010; Stephenson et al., 2010, 2011; Wei et al., 2010; Yu et al., 2011; Ni et al., 2013; Qu et al., 2015). A large number of these genes have been evaluated in important cereals such as maize, rice, and wheat (Miyoshi et al., 2003; Nelson et al., 2007; Wei et al., 2010; Ito et al., 2011; Stephenson et al., 2011; Qu et al., 2015). For example, regulation of plant height and yield potential under optimal growth conditions was demonstrated for transgenic rice plants overexpressing the rice gene encoding the HAP3 subunit, DTH8, which appears to play an important role as a suppressor in the signal network of photoperiodic flowering (Wei et al., 2010). Constitutive overexpression of the ZmNF-YB2 gene in transgenic maize significantly improved performance of transgenic plants subjected to mild drought in controlled conditions and during field trials (Nelson et al., 2007). It has recently been demonstrated that the TaNFYA-B1 gene plays an important role in root development and in nitrogen and phosphorus uptake in wheat (Qu et al., 2015). However, given the large number of NF-Y genes in the genome of wheat and the potential for different combinations of these genes to have specific regulatory functions, further characterization of NF-Y genes is warranted.

In this work, a wheat homologue of the maize ZmNF-YB2a subunit was cloned using yeast two-hybrid (Y2H) screens of cDNA libraries prepared from tissues of a drought-tolerant wheat cultivar subjected to drought and heat stress. The effects produced by overexpression of one of the isolated wheat genes in the elite Australian wheat cultivar, Gladius, was evaluated.

**Materials and methods**

**Plant material**

Transgenic plants were generated in a modern Australian wheat variety Gladius. The pedigree of Gladius was as following: RAC-875/Krichauff/3/RAC-875/Krichauff/4/RAC-875/excalibur/Kukri. The variety was directly developed from a doubled haploid and is, consequently, perfectly homozygous and true breeding. This variety performed well in the hot, dry conditions of southern Australia and is regarded as particularly drought tolerant (Fleury et al., 2010).

**Plasmid construction and plant transformation**

A 492 bp fragment of the TaNF-YB4 coding sequence (CDS) was cloned into the pENTR-D-TOPO vector and verified by sequencing using the primers listed in Supplementary Table S1 available at JXB online, and subcloned into the vector pUbi (Eini et al., 2013) by recombination. The resulting construct was designated as pUbi-TaNF-YB4. The pUbi-TaNF-YB4 construct was linearized using the unique Pme1 restriction site and co-transformed with a selection marker cassette (pUbi-Hyg-nos) into the Australian elite wheat cv. Gladius, using a biolistic bombardment method described by Kovalchuk et al. (2009) and Ismagul et al. (2014). Transgene integration was confirmed by PCR using a forward primer derived from the 3′ end of the transgene CDS and a reverse primer from the 5′ end of the nos terminator (Supplementary Table S1), for 17 independent transgenic events. Transgene copy number was estimated by quantitative PCR as described by Fletcher (2014).

**Yeast two-hybrid screen**

The CDS of ZmNF-YB2a (accession no. NP_001105435) was cloned into the vector pGBK T7 (Invitrogen, Victoria, Australia) for Y2H screening. ZmNF-YB2a was used as bait because maize ZmNF-YB2 CDS could not be isolated based on the sequence submitted to public databases by Nelson et al. (2007). The only protein sequence divergence between the two is an insert of seven amino acid residues (GKTIPAN) in the HFD of ZmNF-YB2, which is absent in ZmNF-YB2a and similar NF-YB subunits from wheat and other grasses. ZmNF-YB2a was used as bait to screen WGL and WENDL (Lopato et al., 2006), WHSL (Eini et al., 2013), WGD (developing grain at 0–6 days after pollination collected from the wheat cv. RAC875 subjected to drought at flowering), and WRDL (roots of wheat cv. RAC875 seedlings grown in soil and subjected to drought) cDNA libraries as previously described (Eini et al., 2013). A large number of independent clones containing full-length or partial coding regions of the TaNF-YC15 cDNA were isolated. Because of self-activating properties of the full-length TaNF-YC15, a truncated version of the protein beginning at residue Ala97 was used to re-screen the same cDNA libraries. Clones containing inserts encoding two different full-length NF-YB TFs, designated as TaNF-YB2 (18 independent clones) and TaNF-YB4 (22 independent clones), were isolated.

**Phylogenetic analysis of NF-Y subunits containing histone fold domains**

The amino acid sequences of 53 NF-YB and 32 NF-YC proteins were aligned with Clustal Omega (Sievers et al., 2011) and alignments were further inspected by PROMALS3D (Pe et al., 2008) and the Alignment Annotator (Gille et al., 2014). Unrooted phylogenetic trees, based on crude distance measures, were visualized in TreeView (Page, 1996).
1996). GenBank accession numbers of protein sequences included in the phylogenetic trees can be found in the Supplementary data.

Construction of a three-dimensional (3D) model of the TaNF-YB2/TaNF-YC15 dimer

A 3D model was constructed by using mapping monomeric threading alignments to protein–protein interactions based on oligomeric entries in the Protein Data Bank (PDB) (Guerler et al., 2013). The crystal structure of the NF-YB/NF-YC dimer from Homo sapiens (PDB accession 1N1J, chains A/B, designated as 1N1JA/1N1JB) (Romier et al., 2003) was identified as a suitable quaternary assembly template for structural modelling. The full-length TaNF-YB2 and TaNF-YC15 sequences were analysed by SMART (Letunic et al., 2012), ProDom (Bru et al., 2005), and SBASE (Vlahovic et al., 2002) to determine domain arrangements and the positions of HFDs. The TaNF-YB2 and TaNF-YC15 sequences were aligned with those of 1N1JA and 1N1JB, respectively, using LOMETS (Wu and Zhang, 2007), and the alignment quality was checked by the Alignment Annotator (Gille et al., 2014) and PSI-PRED (Buchan et al., 2013) to confirm that secondary structures remained undisturbed. The aligned sequences were submitted to SPRING (Guerler et al., 2013) and the most suitable model of TaNF-YB2/TaNF-YC15, evaluated by structural criteria, was selected from a library of six models. The model was minimized using AMBER99 force field to achieve optimal stereochemical parameters. A Ramachandran plot of the optimized model indicated that 100% of the residues were in the most favoured, additionally allowed, and generously allowed regions, when excluding glycine and proline residues. The overall G-factor values evaluated by PROCHECK (Laskowski et al., 1993) were 0.55 and 0.11 for 1N1JA/1N1JB and TaNF-YB2/TaNF-YC15, respectively. The Z-score values, deduced from Prosa2003 (Sippl, 1993) and reflecting combined statistical potential energy, were −5.21 and −5.12 for 1N1JA/1N1JB and TaNF-YB2/TaNF-YC15, respectively. The root-mean-square deviation (RMSD) values between 1N1JA/1N1JB (165 residues) and TaNF-YB2/TaNF-YC15 (169 residues) determined with a PyMol ‘super’ algorithm were 1.1 Å for 164 residues in Cα positions. Images of structural models were generated in the PyMol Molecular Graphics System, Version 1.3 Schrödinger, LLC.

Genotyping and transgene expression analyses

Plant DNA was extracted from leaf tissue using a freeze-drying method described by Shavrukov et al. (2010). Individuals were genotyped for the presence of the transgene. Transgene copy number was estimated by efficiency-adjusted real-time quantitative PCR (qRT-PCR), using nos terminator-specific primers (Supplementary Table S1 at JXB online; Kovalchuk et al., 2013). For template loading normalization, primers and a probe complementary to a portion of the single-copy endogenous Parvicolinae-b (Pin-b) gene were used (Li et al., 2004). The oligonucleotide sequences were: forward 5′-ATTTCCGATCTACCTGCCGCC-3′; reverse 5′-TGCTATCTGGCTCAGCTGC-3′; and dual-labelled TaqMan probe 5′-CAL fluor Gold 540-ATGGTGGAAGGG CGGCTGTGA-BHQ1-3′. Total RNA was isolated from leaf tissue using the Direct-zol RNA MiniPrep (Zymo Research Corporation, ACT, Australia). Transgene expression was confirmed using reverse transcription–PCR (RT-PCR) and transgene-specific primers (Supplementary Table S1 at JXB online). For northern analysis of the transgene expression, RNA was electrophoretically separated in a 1.3% agarose gel containing 6% formaldehyde, transferred to a nylon membrane, and hybridized with 32P-labelled DNA probes according to the protocol described by Church and Gilbert (1984).

Gene expression analysis of native NF-Y genes in wheat

qRT-PCR analyses of gene expression in various tissues of non-transgenic wheat were performed on cDNA samples as described by Burton et al. (2008). Four control genes were used to normalize the qRT-PCR data (primers in Supplementary Table S1 at JXB online) as described by Fletcher (2014). Samples included cDNA developmental tissue series and were prepared from different tissues of T. aestivum cv. Chinese Spring (Kovalchuk et al., 2013). A dehydra
tion cDNA series was prepared from detached leaves of 4-week-old seedlings of drought-tolerant wheat cv. RAC875 (Kovalchuk et al., 2013), which were incubated for 0, 0.5, 1, 2, 4, 5, and 7h at room temperature (23 °C) in opened 15 ml plastic tubes, before snap-freezing in liquid nitrogen for RNA extraction. There were three biological replicates in each qPCR experiment, which was repeated four times.

Analysis of transgenic plants

Experiment 1 Three lines estimated to have a single copy of the transgene (L3, L4, and L5) and one line with two copies (L6) were selected for preliminary characterization of plant phenotypes and seed multiplication. Six T1 plants for each line were grown in 15 cm diameter pots, one plant per pot, in coco-peat soil in a greenhouse (24/16 °C, day/night temperature; 16h day) under well-watered conditions. Untransformed wild-type (WT) plants were also grown for the comparison. Plant height, number of tillers and spikes, plant biomass, flowering time, seed number, and seed weight were recorded for each plant. Transgene copy number and transgene expression levels (Supplementary Fig. S1 at JXB online) were also determined, and null segregants were excluded from the analyses.

Experiment 2 Six T2 sub-lines (L3-5, L4-2, L4-4, L5-4, L6-2, and L6-3) derived from four independent T1 lines were selected for a second experiment under a controlled water regime, conducted in two large containers (190 × 68 × 60 cm) filled with a 1:1:1 mix of coco-peat, river sand, and clay soil collected near Adelaide (South Australia). The soil was re-used for two seasons, and 1/5 of coco-peat soil was added before a new sowing to compensate for any loss of nutrients. In general, nutrient supply in this soil was relatively low but comparable with non-fertilized field soil. Plants were grown in rows, with eight plants per row for each line and the WT with three randomized blocks in each container, comprising in total 18–26 biological replicates for each line as well as for the WT and null segregant lines. Experimental plants were flanked by a border row of WT plants on each short side of the container. The experimental design was identical for each container. No significant differences were found in plant growth between the three blocks in preliminary experiments (data not shown), and therefore all replicates for each line, WT, and nulls were used to calculate means and standard errors for the measurements. Containers were watered every second day. The watering was withdrawn in the droughted container when the majority of plants reached tillering; watering was continued in the well-watered container until the end of grain maturation. A soil water potential of −0.3 ± 0.05 MPa was reached in the droughted container a short time before flowering, and plants were kept under slowly increasing drought until ~10 d after the end of flowering. Soil water content was monitored by the system, Magpie-3 (Measuring Engineering Australia), where data for water content in the soil were regularly collected by sensors from each container and from three levels of depth in the soil: 15, 30, and 45 cm. Curve graphs of droughted and well-watered bins were automatically recorded. When water potential in the droughted container reached ~0.5 MPa, watering re-commenced and soil moisture was restored to a level similar to the well-watered container (Supplementary Fig. S2 at JXB online).

Measurements of yield-related plant growth traits and yield components were taken at harvest, as for Experiment 1. Leaf material sampled from all plants was used for genotyping and analysis of gene expression. Five of the six T2, sub-lines (L3-5, L4-2, L4-4, L5-4, and L6-3) were identified as homozygous for the transgene (Supplementary Fig. S1 at JXB online). In the segregating
heterozygous sub-line (L6-2), null segregants were excluded from the analyses, as were progeny of line L3 due to a large number of dead plants (Table 1).

Experiment 3  T1 wheat sub-lines were grown in 22 cm diameter pots, four plants per pot, including one WT and three transgenic plants. Transgenic plants included representatives of two confirmed and one possible homozygous sub-line (L4-4-51, L5-4-52; and L6-3-31). A coco-peat soil amended with five levels (0, 0.7, 1.4, 2.1, and 2.8 gram per litre of soil) of slow-release complete fertilizer (Osmocote Exact Mini, Everris International, Australia) was used in this experiment. The nutrient content of the soil was in the range of medium to high, and comparable with the levels reached in well-fertilized field soil. Three pots of plants (replicates) were grown for each level of fertilizer, except for the unamended treatment, for which there were six pots. Analysis of yield components was performed as described above for Experiments 1 and 2.

Statistical treatment of data
Single-factor analyses of variance (ANOVAs) and Student t-tests (unpaired, two-tails) from Microsoft Excel software were applied for statistical analyses of data from the plant phenotyping experiments. Numbers of biological replicates were 18–26 (Experiment 2) and 3–6 (Experiment 3).

Results
Isolation of wheat genes encoding NF-YB subunits
In order to identify and isolate cDNA clones encoding wheat NF-YB proteins homologous to maize ZmNF-YB2, a two-step Y2H screen was performed. Full-length ZmNF-YB2a protein (accession no. NP_001105435) was used to screen cDNA libraries prepared from different wheat tissues of drought-tolerant wheat cv. RAC875 subjected to drought and heat stresses.

The screen resulted in isolation of several TFs and transcription-related proteins (data not shown). The most abundant of the isolated sequences was from an unreported wheat gene encoding the NF-Y subunit C, designated as TaNF-YC15. To prevent self-activation during the Y2H screen, TaNF-YC15 was truncated at the N-terminal end. Truncation did not affect the HFD, important for interactions with NF-YB proteins. Y2H screens with TaNF-YC15 resulted in isolation of large numbers of independent clones encoding two different NF-YB subunits identical to TaNF-YB2 and TaNF-YB4, which were previously described by Stephenson et al. (2007) (Fig. 1A). Full-length coding regions of isolated wheat NF-YB cDNAs were cloned into the bait vector, and interaction of TaNF-YB2 and TaNF-YB4 with TaNF-YC15 was demonstrated in a reciprocal way (Fig. 1B). In addition, it was found that TaNF-YC15 is the most abundant among several NF-YC subunits isolated in Y2H screens using TaNF-YB2 and TaNF-YB4 as bait proteins (data not shown).

The phylogenetic relationships between NF-YB and NF-YC protein sequences from wheat, maize, rice, and Arabidopsis were revealed in unrooted trees constructed using Clustal Omega (Blackshields et al., 2010) (Fig. 2). The tree for the NF-YB sequences shows that both TaNF-YB2 and TaNF-YB4 belong to two different branches of the same protein clade, and that TaNF-YB2 and ZmNF-YB2 are the products of orthologous genes as they are grouped together. The TaNF-YB4 protein is grouped with a protein from maize, which is designated as ZmNF-YB4, while two rice proteins are grouped within these two branches. In contrast, the Arabidopsis AtNF-YB1 protein has a high level of identity to proteins from both branches containing TaNF-YB2-like and TaNF-YB4-like proteins, and two other proteins, AtNF-YB8 and AtNF-YB10, have much lower levels of identity to TaNF-YB2 (59% and 63%, respectively) or TaNF-YB4 (58% and 61%, respectively), but still are grouped into the same clade (Fig. 2A). The isolated TaNF-YB2 protein has a single amino acid residue difference from that submitted to the GenBank database (accession no. BT009078). This may reflect the fact that these two proteins originate from different wheat cultivars. No differences were found in the protein sequences for TaNF-YB4 isolated from cv. RAC875 in this study and the GenBank entry for TaNF-YB4 (accession no. BT009393).

TaNF-YC15 has a high level of protein sequence identity to TaNF-YC7 (accession no. AED95949). Both proteins belong to the same clade together with orthologous proteins from rice and the more distant TaNF-YC1 (accession no.

Table 1. T, progeny of TaNF-YB4 transgenic lines analysed in large containers

| Growth conditions | Lines | WT | Null | L3-5 | L4-2 | L4-4 | L5-4 | L6-2 | L6-3 |
|-------------------|-------|----|------|------|------|------|------|------|------|
| Trangene copy number | Total no. of plants | 21 | 22 | 10 | 21 | 28 | 28 | 28 | 28 |
| Well-watered | No. of dead plants | 3 | 2 | 10 | 0 | 4 | 3 | 1 | 1 |
| Well-watered | No. of nulls | – | 22 | 0 | 0 | 0 | 0 | 4 | 0 |
| Well-watered | No. of plants with expressed transgene | – | 0 | 10 | 21 | 24 | 25 | 23 | 27 |
| Drought | Total no. of plants | 24 | 23 | 9 | 24 | 24 | 24 | 24 | 24 |
| Drought | No. of dead plants | 2 | 1 | 9 | 2 | 1 | 2 | 2 | 2 |
| Drought | No. of nulls | – | 23 | 0 | 0 | 0 | 0 | 4 | 0 |
| Drought | No. of plants with expressed transgene | – | 0 | 9 | 22 | 23 | 22 | 20 | 28 |

a Progeny of sub-line 3-5 demonstrated a poor growth and high mortality, and therefore it was not shown in Fig. 5 or used in further experiments.
b Homozygous sub-lines.
c Only data for plants with the expressed transgene were used for generation of the graphs in Fig. 6.
CD88515). No proteins from *Arabidopsis* were grouped in the same clade (Fig. 2B).

**TaNF-YB2 and TaNF-YB4 interact with TaNF-YC15**

To explain why only two out of more than a dozen wheat NF-YB factors interacted with TaNF-YC15, the essential segment of the TaNF-YB2 protein was mapped in a Y2H assay using a series of truncated forms of TaNF-YB2 (Fig. 1C). The identified minimal protein segment, which interacted with TaNF-YB2 and TaNF-YB4 interacted with TaNF-YC15, the essential segment indicated that the structural fold has not undergone significant diversification during evolution. The residues involved in dimerization of TaNF-YB2/TaNF-YC15 included nine residues each in chain B (Lys57, Ala59, Asp61, Glu68, Ser75, Lys91, Ile93, Tyr110, and Arg122) and chain C (Asp117, Asp119, Met122, Ser124, Glu126, Gly141, Arg156, Leu158, and Lys160) (Fig. 3C). All nine residues in TaNF-YB2 and eight out of the nine residues in TaNF-YC15 were conserved compared with human NF-YB/NF-YC, except for Lys160 that was substituted with Arg75 in human NF-YC (Fig. 3B, C).

All residues of TaNF-YB2 HFD (Fig. 3) were present in the minimal segment of TaNF-YB2 (residues 50–146; Fig 1C) that were required for interaction with TaNF-YC15 in Y2H assay. A truncated variant of TaNF-YB2 with a C-terminal Glu123 (underlined in the motif QKYR; Fig 3B) lost the ability to bind TaNF-YC15. This suggested a minimal length for TaNF-YB2 protein to ensure correct folding of TaNF-YB2 (Fig. 3B), and allowing Arg122 (that precedes Glu123) to recover its proper protonation state and function during formation of the TaNF-YB2/TaNF-YC15 complex. Gln119 of wheat TaNF-YB2 was located in close proximity to Arg122 in TaNF-YB2 and was also present in TaNF-YB4 (and human NF-YB; Fig. 3B), but was substituted in all other TaNF-YBs by a histidine residue.
Further indirect evidence that wheat TaNF-YC15 acts in concert with either TaNF-YB2 or TaNF-YB4 came from gene expression analyses of plant tissues treated under a variety of conditions. Expression levels of TaNF-YC15 in these wheat tissues correlated well with levels of NF-YB transcripts, particularly with TaNF-YB4 (Fig. 4).
Expression levels of isolated NF-Y transcription factors in different tissues, during slow developing drought and rapid dehydration

Transcript levels of TaNF-YB2, TaNF-YB4, and TaNF-YC15 genes were analysed by qRT-PCR in different tissues of un unstressed wheat plants and in leaves from plants subjected to abscisic acid (ABA) treatment, drought, or rapid dehydration. Expression of both NF-YB factors was detected in all tested tissues of unstressed wheat plants. Higher levels of expression were observed in reproductive tissues and particularly in anthers, where levels of expression of both genes were

Fig. 3. Molecular model of the wheat TaNF-YB2/TaNF-YC15 dimeric substructure and structural bioinformatics of wheat NF-YB and NF-YC transcription factors. (A) A multiple sequence alignment of HFDs (coloured in blue) of 11 TaNF-YB from wheat. The sequence of a human NF-YB (PDB accession 1NIJ-A also designated NF-YB) is also included. Protein sequences were aligned with ProMals3D (Pei et al., 2008). Predicted consensus α-helices (h) are marked. Conservation of residues on a scale of 9–6 is shown at the top of the diagram. Predicted residues involved in binding to HFD of NF-YC are marked in blue boxes. Conserved glutamine or histidine residues at the C-terminal regions are shaded in magenta. TaNF-YB2 and TaNF-YB4 isolated in this work are in bold. (B) A multiple sequence alignment of HFDs (cyan) of 12 TaNF-YCs from wheat. The sequence of a human NF-YC (PDB accession 1NIJ-B; also designated NF-YC) is also included. Predicted residues involved in binding to the HFD of NF-YB are marked in cyan boxes. Other annotations are as specified in (A). (C) Molecular model of the wheat TaNF-YB2/TaNF-YC15 dimeric substructure (sequences highlighted in bold in A and B). Secondary structures are shown in cartoon representations (blue for TaNF-YB, cyan for TaNF-YC). Residues participating in binding (TaNF-YB to TaNF-YC: Lys57, Ala59, Asp61, Glu68, Ser75, Lys91, Ile93, Tyr110, and Arg122; TaNF-YC to TaNF-YB: Asp117, Asp119, Met122, Ser124, Glu126, Gly141, Arg156, Leu158, and Lys160) involved in dimer formation are shown in sticks and atomic colours. Gln119 in close proximity to Arg122 is in magenta. Distances between interacting residues are shown in black dashed lines and vary between 2.7 Å and 3.5 Å.
5- to 6-fold higher than in other tissues (Fig. 4A). Correlation between TaNF-YB2 and TaNF-YB4 levels of expression was observed across most tissue types. The most significant difference between expression levels of NF-YB genes was found in mature endosperm, where the number of TaNF-YB2 transcripts was ~9-fold higher than the number of TaNF-YB4 transcripts (Fig. 4A). Expression levels of TaNF-YC15 in different wheat tissues correlated well with levels of NF-YB transcripts, particularly with TaNF-YB4, with one exception (Fig. 4B). In contrast to NF-YB genes, level of TaNF-YC15 expression in anthers was lower than in bracts and pistil. Overall, the TaNF-YC15 gene was less highly expressed than TaNF-YB2/YB4.

Neither TaNF-YC15 nor TaNF-YB was induced by ABA treatment (data not shown). In a drought cDNA tissue series, the three NF-Y genes showed two periods of induction. The first induction point occurred on the 17th day of drought treatment, when plants had just started to sense drought, and the second induction point happened at the wilting point, 30 d after watering was withheld (Fig. 5A). A control gene known to be drought stress inducible, TaCor39 (Kovalchuk et al., 2013), showed a single increase in expression across the same tissue series, at 30 d after cessation of watering (Fig. 5A). Following moderate re-watering, expression levels of all tested genes decreased, but began to increase again after a further 4 d as the soil profile dried out (day 35).

In contrast to slowly developing drought, the rapid dehydration of detached leaves at room temperature led to different patterns of TaNF-YB2 and TaNF-YB4 expression. While TaNF-YB2 was induced by 3-fold, TaNF-YB4 levels
decreased with dehydration. Interestingly, the patterns of expression of \textit{TaNF-YB4} and \textit{TaNF-YC15} in dehydrating leaves were nearly identical (Fig. 5B). The \textit{TaCor39} gene was used as a dehydration-inducible control gene to confirm the quality of the dehydration cDNA series.

**Evaluation of transgenic wheat plants overexpressing \textit{TaNF-YB4} in well-watered, droughted, and nutrient-depleted conditions**

Transgenic wheat plants (cv. Gladius) were generated with constitutive overexpression of \textit{TaNF-YB4} driven by the maize ubiquitin promoter. In the preliminary evaluation of plant performance (\textit{T}_1 generation), three from four transgenic lines with one or two copies of the transgene produced more grain compared with control plants, without substantial changes in other yield components (data not shown).

In the second experiment, two large containers were used for phenotyping of a large number of transgenic plants (18–28 \textit{T}_2 individuals from each of six \textit{T}_2 sub-lines, L3-5, L4-2, L4-4, L5-4, L6-2, and L6-3; see Table 1). All of these except L6-2 were determined to be homozygous for the transgene. Progeny of the sub-line L3-5 grew more slowly than other plants and showed a high level of mortality (Table 1), possibly due to a very high level of transgene expression (>10-fold higher than in other lines; data not shown), or as a result of effects due to the position of transgene integration. This sub-line was not considered for further analysis.

Under well-watered conditions, the transgenic \textit{T}_2 plants yielded up to 30% more mass of grain per plant (Fig. 6).
Supporting the initial observations, all five analysed transgenic lines showed significant increases in the number of spikes, vegetative biomass, and grain weight per plant compared with control plants (Fig. 6). There was a slight reduction in single-grain weight for four of the lines. Thus, the increased grain yield was attributable to an increased number of spikes per plant (Fig. 6). No yield advantage was observed for transgenics compared with control plants under mild drought.

Preliminary observations of plant performance in nutrient-depleted media indicated that NF-Y factors may impart the greatest yield advantage in poor soils. Thus, a third growth experiment was conducted using TaNF-YB4 homozygous transgenic lines (T3) to investigate this hypothesis. Plants were grown in a coco-peat soil either with no addition or with four different rates of Osmocote, a slow-release complete fertilizer. Increases in grain yield of the transgenic plants compared with control plants were evident across all fertilizer treatments (Fig. 7; Table 2). However, there was also an interaction between fertilizer rate and the yield advantage of transgenic lines. Control plants showed a 25% increase in yield in response to fertilizer addition. In contrast, no positive effect of additional fertilizer on the grain yield of transgenic lines was observed (Fig. 7).

Discussion

ZmNF-YB2, TaNF-YB2, and TaNF-YB4 bind to TaNF-YC15

Plant genomes contain >10 genes for each of three subunits of NF-Y proteins (Riechmann et al., 2000), making it difficult to identify the specific subunits that may come together to form heterotrimeric complexes. However, Y2H screening methods such as employed in this study allow the direct identification of interacting proteins from libraries of expressed genes. ZmNF-YB2a was selected to be used as starting bait in sequential Y2H screens, to find related wheat NF-Y subunits and other proteins. ZmNF-YB2 has been demonstrated to increase grain yield when constitutively overexpressed in maize, particularly under drought (Nelson et al., 2007). Aiming to identify related genes possibly implicated in

![Fig. 6. A comparison of phenotypes and grain yields of transgenic and control (WT and null segregant) wheat plants grown in large containers under well-watered conditions and constantly increasing drought (see Supplementary Fig. S2A at JXB online). From 16 to 28 WT, nulls, and transgenic T2 plants with confirmed transgene expression for each sub-line of three independent lines were used in the experiment (see Table 1). All sub-lines except line 6-2 were confirmed to be homozygous. Differences between transgenic lines and both WT and null plants were tested in the unpaired Student’s t-test (*P<0.05; **P<0.01).](https://academic.oup.com/jxb/article-abstract/66/21/6635/2893305)
drought stress tolerance in wheat, Y2H cDNA libraries prepared from different tissues of the Australian drought-tolerant wheat cultivar RAC875 were screened, including tissues from plants subjected to drought or to combined drought and heat stresses.

In the first round of the Y2H screening, a novel wheat protein, TaNF-YC15, encoded by DNA inserts of most clones isolated with ZmNF-YB2a as bait, was identified. Then a truncated N-terminal version of TaNF-YC15, containing the full-length HFD that is important for protein–protein interactions (Fig. 3B, C), was used to screen for interacting proteins from the same wheat tissue libraries. In these Y2H screens, only two types of NF-YB subunits were found, TaNF-YB2 and TaNF-YB4, full-length cDNAs of which were identified in the majority of isolated independent clones. Both proteins had high levels of sequence identities (83.6% for TaNF-YB2 and 74.9% for TaNF-YB4) to the starting bait ZmNF-YB2a.

It was previously reported that most NF-YB subunits of Arabidopsis can interact in Y2H assays with either the majority of or all NF-YC subunits, suggesting a low level of selectivity between NF-YB and NF-YC (Calvezani et al., 2012; Hackenberg et al., 2012). In this study, however, significant selectivity in recognition of TaNF-YC15 by wheat NF-YB subunits was observed. At least three explanations can be proposed: (i) high enrichment with particular NF-Y cDNAs in the cDNA libraries (e.g. up-regulation of both isolated NF-YB genes by drought; Fig. 5B); (ii) high specificity of recognition between TaNF-YC15 and TaNF-YB2 or TaNF-YB4, as a result of differences in protein sequences that guide protein–protein interactions; or (iii) differences in post-translational modifications of NF-YB subunits. Identification of the same NF-YB genes in several different cDNA libraries and isolation of the same NF-YC subunit in reciprocal screens using TaNF-YB2 and TaNF-YB4 proteins as bait (data not shown) suggested that the specificity of NF-YB/NF-YC complex formation could be guided by a protein sequence rather than by abundance of cDNAs in libraries. It was possible to isolate a number of other TaNF-YB gene family members from the cDNA libraries (data not shown), but these were not identified with TaNF-YC15 in Y2H screens.

Fig. 7. A comparison of phenotypes and grain yields of transgenic and control (WT) wheat plants grown in soil mixes (A), either without added fertilizer (marked as ‘low’), or with the addition of a complete, slow-release fertilizer (marked as ‘high’) (see Table 2). One WT plant and one T3 transgenic plant for each of three homozygous sub-lines with confirmed transgene expression were grown in the same pot. Six pots were used for the soil mix without added fertilizer, and three pots for the fertilized treatments. Differences between transgenic lines and WT plants were tested in the unpaired Student’s t-test (*P<0.05; **P<0.01). Images of spikes from plants grown in separate pots: (B) without addition and (C) with addition of a complete slow-release fertilizer. (This figure is available in colour at JXB online.)
Aiming to find possible determinants of specificity in interactions between various NF-YBs and TaNF-YC15, a TaNF-YB2 segment that may be responsible for the interaction with TaNF-YC15 was mapped (Fig. 1C). A minimal size segment of TaNF-YB2, which preserves a strong ability to bind TaNF-YC15, started at residue 50 (inside the N-terminal end of the HFD) and was finished at residue 146 (downstream of the C-terminal end of the HFD). Further truncations from either side led to disruption of protein–protein interactions, because of either the deletion of key residues responsible for these interaction and/or disruption of HFD folds (Fig. 3). The minimal interacting segment of TaNF-YB2 was compared with similar segments of TaNF-YB4 and other known wheat NF-YB subunits (Fig. 3A). It was found that similarly to Gln90 of human NF-YB, Gln119 of TaNF-YB2 and the respective glutamine of TaNF-YB4 were situated close to Arg93, which was involved in TaNF-YB2/TaNF-YC15 complex formation (Fig. 3C). In all other wheat NF-YB sequences, this glutamine residue was substituted by histidine. Since histidine imidazole side chains play roles in electrostatic interactions of proteins, the histidine to glutamine substitution in TaNF-YB2 and TaNF-YB4 may directly influence the strength of interaction with TaNF-YC15 (Hansen and Kay, 2014). It is suggested that the histidine/glutamine substitution may play a critical role in the specificity of the interaction between TaNF-YC15 and TaNF-YB2 or TaNF-YB4.

Expression of TaNF-YB2, TaNF-YB4, and TaNF-YC15 is regulated in a similar way in most tested tissues and under drought stress, but differs under rapid dehydration

Analysis of expression levels of TaNF-YB2, TaNF-YB4, and TaNF-YC15 revealed that all three genes are expressed in all tested tissues, indicating that their products are in close physical proximity to each other for the formation of protein complexes. However, significant differences in levels of TaNF-YB2 and TaNF-YB4 expression in particular tissues (e.g. mature endosperm) might suggest the possible importance of some specific features of protein sequences for the formation of particular transcriptional complexes, and hence differences in function of TaNF-YB2 and TaNF-YB4. Observations of NF-YB2/4 gene expression in all tested wheat tissues confirm the results of wheat NF-Y expression analysis reported by Stephenson et al. (2007). Furthermore, the presence of transcripts of other NF-YB and NF-YC subunits in the same tissues (Stephenson et al., 2007) and detection of these in the cDNA libraries (data not shown), together with results of the Y2H screens, advocate the fact that interactions between particular NF-YB and NF-YC subunits can be more specific than other similar interactions.

All three isolated NF-Y genes were up-regulated by drought. In contrast to the stress-inducible control gene TaCor39 (Guo et al., 1992; Kovalchuk et al., 2013), which is progressively induced by slowly developing drought, both TaNF-YB and TaNF-YC15 genes were activated at two points, initially when plants began to sense insufficiency of water, and a second time at wilting point, when water deficit became severe (Fig. 5). Maximal levels of TaNF-YB2 activation in the present experiments were considerably lower than the level of activation of the same gene by dehydration reported by Stephenson et al. (2007). This may be explained by different stress conditions used in these experiments, and possibly an insufficient number of data points in the experiment for detecting the peak time of expression. In comparison, TaNF-YB2 transcript levels were increased by 3-fold in detached leaves subjected to rapid dehydration, a similar level of induction to the stress-inducible reference gene TaCor39 (Fig. 5). TaNF-YB4 and TaNF-YC15 showed very similar patterns of expression under dehydration. Both genes were initially down-regulated, then

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**Table 2. Influence of different amounts of the slow-release fertilizer Osmocote Exact Mini on yield components of WT (wheat cv. Gladius) and three homozygous TaNF-YB4 transgenic lines**

| Line                  | Additional Osmocote (gram per litre of soil) |
|-----------------------|---------------------------------------------|
|                       | 0               | 0.7          | 1.4          | 2.1          | 2.8          |
| **Spike number per plant** |                              |               |               |               |               |
| WT                    | 5.7±0.7         | 6.1±0.8      | 6.3±0.7      | 6.5±0.5      | 6.9±0.4      |
| L4-4-51               | 9.2±0.5         | 10.7±0.4     | 10.3±1.2     | 11.0±0.7     | 10.7±0.5     |
| L5-4-52               | 9.5±0.9         | 9.7±1.5      | 9.7±0.4      | 10.3±1.1     | 9.7±1.0      |
| L6-3-31               | 9.0±1.0         | 10.0±1.4     | 11.0±0.9     | 11.0±0.7     | 11.0±1.2     |
| **Seed number per plant** |                              |               |               |               |               |
| WT                    | 162.3±14.6      | 169.0±31.3   | 189.7±20.2   | 206.3±26.0   | 215.7±32.2   |
| L4-4-51               | 301.3±12.3      | 291.3±11.7   | 340.7±28.0   | 341.0±13.7   | 336.3±30.1   |
| L5-4-52               | 339.8±24.0      | 286.7±32.3   | 269.3±30.6   | 298.0±33.4   | 333.3±36.6   |
| L6-3-31               | 290.7±12.3      | 287.7±21.8   | 332.7±29.5   | 324.3±28.8   | 348.0±35.8   |
| **Seed weight per plant (g)** |                              |               |               |               |               |
| WT                    | 4.33±0.34       | 4.29±0.64    | 4.70±0.49    | 5.53±0.65    | 5.38±0.64    |
| L4-4-51               | 8.69±0.31       | 8.28±0.62    | 8.61±0.70    | 8.28±0.21    | 7.73±0.45    |
| L5-4-52               | 8.57±0.48       | 8.02±0.81    | 7.88±0.87    | 7.76±0.35    | 7.65±1.10    |
| L6-3-31               | 7.85±0.38       | 7.82±0.88    | 8.29±0.62    | 8.29±0.88    | 8.22±0.75    |

Data are presented as means (n=3) ±SE with the exception for the first treatment (denoted as 0), where n=6. WT plants (*) were statistically different from each of three lines at least for P<0.05 for each treatment and each trait.
after 2 h of leaf dehydration they partially returned to initial levels of expression and later again decreased by as much as 2.5-fold. These results suggest three interesting conclusions: (i) TaNF-YB2 and TaNF-YB4 may be induced by different components of drought stress, in the case of TaNF-YB2 by dehydration, and therefore may play different roles in drought stress responses; (ii) significant differences in levels of expression of TaNF-YB2 and TaNF-YB4 in mature endosperm may be a result of TaNF-YB2 activation by natural desiccation of grain at this stage of development; or (iii) TaNF-YB4 and TaNF-YC15 are more likely to be constituents of the same NF-Y protein complex than TaNF-YB2 and TaNF-YC15. The activation of NF-Y genes by drought was ABA independent, because no significant changes in expression of all three genes were induced by treatment with 200 μM ABA.

Constitutive overexpression of TaNF-YB4 improves performance of transgenic wheat

While constitutive overexpression of stress-related TFs generally confers improved plant reactions to corresponding stresses, in many cases it does not increase yield (seed weight per plant) (Hsieh et al., 2002; Oh et al., 2005, 2007; Kobayashi et al., 2008a, b; Takumi et al., 2008). In contrast, strong constitutive overexpression of TFs often results in development of undesirable pleiotropic phenotypes, and consequently reduces grain yields (Jaglo-Ottosen et al., 1998; Liu and Zhu, 1998; Kasuga et al., 1999; Hsieh et al., 2002; Ito et al., 2006; Morran et al., 2011; Kovalchuk et al., 2013). Reports of the overexpression of some NF-Y subunits in maize and rice, however, are promising exceptions to this rule (Nelson et al., 2007; Wei et al., 2010). The main purpose of this study was to identify and clone wheat NF-Y gene(s) responsible for yield increase and explore the possibility of improving performance of an elite wheat cultivar by overexpression of such gene(s).

In order to be of practical value, transgenic events need to demonstrate advantages over and above those achieved through conventional breeding. This can best be done through comparison with elite lines bred for the target environments. Evaluation of yield and yield components of transgenic plants would ideally be conducted under field conditions in large plots at multiple locations. This is costly and slow given the complex regulatory environment associated with the evaluation of transgenic plants. Some form of preliminary screening is needed to identify promising genes and transgenic events to proceed to field evaluation. However, assessing yield under controlled growth condition is difficult. In this study, transgenic lines produced in an elite wheat cultivar were generated that showed good yield under the hot, dry conditions experienced in the Mediterranean-type environment of southern Australia, where crops are grown on low nutrient soils with a strong terminal drought stress. Comparisons are commonly made between transgenic and null segregants grown in small pots, but a more challenging test is to show that the transgenics can outperform elite lines under conditions as close to the field as is practical in controlled environments. Although null segregants were included in the experiments, the comparisons are based on a non-transgenic control (WT) grown in large bins, at low nutrient status to best approximate field conditions.

Preliminary phenotyping revealed no negative influence of the constitutively expressed TaNF-YB4 transgene on plant phenotype and yield for three independent transgenic lines showing moderate transgene expression. In contrast, line 3 showed slower growth, lower grain number, and a 10% lower grain yield compared with control plants. Levels of transgene expression in this line were ~10-fold higher than in the other lines, suggesting that limiting transgene expression to more moderate levels may be critical for avoiding deleterious phenotypes.

A replicated experiment was conducted in large containers in which soil moisture could be controlled, to simulate seasonal drought stress for one of the treatments. In this experiment, spike number, biomass, and grain yield of transgenic plants were significantly higher than for WT plants under well-watered conditions. Yield increase was observed for all five analysed T2 sub-lines of three transgenic lines (three independent transformation events), of up to 20–30% (Fig. 6). Yield increase in crop plants as a result of overexpression of NF-Y genes has been reported previously (Nelson et al., 2007; Wei et al., 2010). In the first report, transgenic maize plants with enhanced ZmNF-YB2 expression demonstrated increased tolerance to drought based on improvement of parameters such as chlorophyll content, stomatal conductance, leaf temperature, reduced wilting, and maintenance of photosynthesis. These stress adaptations contributed to a grain yield advantage for transgenic maize lines under water-limited conditions (Nelson et al., 2007). Although TaNF-YB4 is closely related to ZmNF-YB2, under limited water conditions, neither reduced wilting nor increased grain yield were observed in transgenic wheat plants compared with untransformed control plants. This may be explained by differences in protein sequences between ZmNF-YB2/TaNF-YB2 and TaNF-YB4, leading to formation of different complexes with other transcription factors and/or different roles for these proteins. The differences in expression patterns of TaNF-YB2 and TaNF-YB4 under dehydration stress also suggest diverse roles for their products under stress and during grain desiccation. It is planned to investigate the drought stress adaptation of transgenic wheat with constitutive overexpression of TaNF-YB2.

In another study, a rice HAP3H (NF-YB) gene, DTH8 [quantitative trait locus (QTL) for days to heading on chromosome 8], was overexpressed in rice (Wei et al., 2010). It was shown that DTH8 suppresses rice flowering under long-day conditions and plays an important role in the regulation of plant height and yield potential. Although the product of the DTH8 gene belongs to another clade of NF-YB TFs (Fig. 2A), the phenotype described for DTH8 transgenic rice plants to some extent resembles the phenotype of TaNF-YB4 transgenic wheat plants. Both transgenic events led to increased biomass production. TaNF-YB4 transgenic wheat plants in the T0 and T1 generations, similarly to DTH8 rice plants, were slightly taller than control plants, although this difference in height was not so pronounced as in DTH8.
It was reported recently (Qu et al., 2015) that the phenotype of transgenic wheat with constitutive overexpression of the gene encoding the NF-Y subunit A of the trimeric TF, TaNFYA-B1 was similar to that described in this work: transgenic plants produced higher grain yield than control plants because of an increased number of tillers and consequently an increased number of seeds. It was demonstrated that the increased nitrogen and phosphorus uptake may be a result of changed root morphology and up-regulated levels of nitrate and phosphate transporters in roots (Qu et al., 2015). Although the present preliminary results revealed no changes in root size and morphology, it cannot be excluded that the reported TaNFYBA-B1 and currently described TaNF-YB4 interacting partners are parts of the same or a similar trimeric protein complex. It is also suggested that the constitutive up-regulation of each of the subunits may lead to the increased functional efficiency of the whole trimeric NF-Y complex. This hypothesis can explain that similar phenotypes of transgenic wheat plants were observed both in the study with TaNFYBA-B1 (Qu et al., 2015) and in the current study with TaNF-YB4. Further research with the transgenic lines is planned to investigate the possible mechanisms by which TaNF-YB4-overexpressing wheat plants overcome nutrient deficiency in soil.

Supplementary data

Supplementary data are available at JXB online.

Figure S1. Confirmation of transgene (TaNF-YB4) expression in transgenic wheat lines.
Figure S2. Large container systems used for plant growth (soil-water potential and outlook of containers).
Table S1. List of PCR primers used in this study.
Supplementary data. List of GenBank accession numbers of protein sequences used for construction of phylogenetic trees (Fig. 2).

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References

Baxevanis AD, Arents G, Moudrianakis EN, Landsman D. 1995. A variety of DNA-binding and multimeric proteins contain the histone fold motif. Nucleic Acids Research 23, 2685–2691.
Bi W, Wu L, Country F, de Crombrugghe B, Maity SN. 1997. DNA binding specificity of the CCAAT-binding factor CBF/NF-Y. Journal of Biological Chemistry 272, 26562–26572.
Blackshields G, Sievers F, Shi W, Wilm A, Higgins DG. 2010. Sequence embedding for fast construction of guide trees for multiple sequence alignment. Algorithms for Molecular Biology 5, 21.
Bra C, Courcelle E, Carrere S, Beausse Y, Dalmar S, Kahn D. 2005. The ProDom database of protein domain families: more emphasis on 3D. Nucleic Acids Research 33, D212–D215.
Buchan DW, Minneci F, Nugent TC, Bryson K, Jones DT. 2013. Scalable web services for the PSIPRED protein analysis workbench. Nucleic Acids Research 41, W349–W357.

Burton RA, Jobling SA, Harvey AJ, Shirley NJ, Mather DE, Bacic A, Finch GB. 2008. The genetics and transcriptional profiles of the cellulose synthase-like HvCSE5 gene family in barley. Plant Physiology 146, 1821–1833.

Cagliari A, Turchetto-Zolet AC, Korbes AP, Maraschin FD, Margis R, Margis-Pinheiro M. 2014. New insights on the evolution of Leafy cotyledon1 (LEC1) type genes in vascular plants. Genomics 103, 380–387.

Cai X, Balili J, Endo S, Davis E, Liang M, Chen D, DeWald D, Kreps J, Zhu T, Wu Y. 2007. A putative CCAAT-biding transcription factor is a regulator of flowering time in Arabidopsis. Plant Physiology 145, 98–105.

Calvanzani V, Testoni B, Gusmaroli G, Lorenzo M, Gnesutta N, Petroni K, Mantovani R, Tonelli C. 2012. Interactions and CCAAT-binding of Arabidopsis thaliana NF-Y subunits. PLOS One 7, e42902.

Chen NZ, Zhang XQ, Wei PC, Chen QJ, Ren F, Chen J, Wang XC. 2007. ATHAP3b plays a crucial role in the regulation of flowering time in Arabidopsis during osmotic stress. Journal of Biochemistry and Molecular Biology 40, 1083–1089.

Church GM, Gilbert W. 1984. Genomic sequencing. Proceedings of the National Academy of Sciences, USA 81, 1991–1995.

Edwards D, Murray JA, Smith AG. 2010. Using real-time qPCR to determine transgene copy number. Methods in Molecular Biology 1145, 213–238.

Feurly D, Jefferies S, Kuchel H, Langridge P. 2010. Genetic and genomic tools to improve drought tolerance in wheat. Journal of Experimental Botany 61, 3211–3222.

Gille C, Birgit W, Gille A. 2014. Sequence alignment visualization in HTML5 without Java. Bioinformatics 30, 121–122.

Guerler A, Govindarajoo B, Zhang Y. 2013. Mapping monomeric tautomer populations in invisible protein states. Journal of Chemical Information and Modeling 53, 717–725.

Guo W, Ward RW, Thomason MF. 1992. Characterization of a cold-regulated gene related to Arabidopsis cos47. Plant Physiology 100, 915–922.

Hackenberg D, Wu Y, Voigt A, Adams R, Schramm P, Grimm B. 2012. Studies on differential nuclear translocation mechanism and assembly of the three subunits of the Arabidopsis thaliana transcription factor NF-Y. Molecular Plant 5, 876–883.

Hanski AL, Kay LE. 2014. Measurement of histidine pKa values and tauator populations in invisible protein states. Proceedings of the National Academy of Sciences, USA 111, E1705–1712.

Hsieh TH, Lee JT, Yang PT, Chiu LH, Chang YY, Wang YC, Chan MT. 2002. Heterology expression of the Arabidopsis C-repeat/dehydration response element binding factor 1 gene confers elevated tolerance to chilling and oxidative stresses in transgenic tomato. Plant Physiology 129, 1086–1094.

Ismagul A, Iskakova G, Harris JC, Elby S. 2014. Biolistic transformation of wheat with centropheoxine as a synthetic auxin. Methods in Molecular Biology 1145, 191–202.

Ito Y, Katsura K, Maruyama K, Taji T, Kobayashi M, Seki M, Shinozaki K, Yamaguchi-Shinozaki K. 2006. Functional analysis of rice DREB1/CBF-type transcription factors involved in cold-responsive gene expression in transgenic rice. Plant and Cell Physiology 47, 141–153.

Ito Y, Thirumurugan T, Serizawa A, Hiratsu K, Ohme-Takagi M, Kurata N. 2011. Aberrant vegetative and reproductive development by overexpression and lethality by silencing of OsHAP3E in rice. Plant Science 181, 105–110.

Jaglo-Ottosen KR, Gilmour SJ, Zarka DG, Schabenberger O, Thomason MF. 1998. Arabidopsis CBF1 overexpression induces COR genes and enhances freezing tolerance. Science 280, 104–106.

Kasuga M, Liu Q, Miura S, Yamaguchi-Shinozaki K, Shinozaki K. 1999. Improving plant drought, salt, and freezing tolerance by gene transfer of a single stress-inducible transcription factor. Nature Biotechnology 17, 287–291.

Kebrom TH, Chandler PM, Swain SM, King RW, Richards RA, Spielmeyer W. 2012. Inhibition of tiller bud outgrowth in the tin mutant of wheat is associated with precocious internode development. Plant Physiology 160, 308–318.

Kobayashi F, Ishibashi M, Takumi S. 2008a. Transcriptional activation of CCAAT genes and increase in abiotic stress tolerance through expression of a wheat DREB2 homolog in transgenic tobacco. Transgenic Research 17, 755–767.

Kobayashi F, Maeta E, Terashima A, Kawaoka K, Ogihara Y, Takumi S. 2008b. Development of abiotic stress tolerance via bZIP-type transcription factor LIP19 in common wheat. Journal of Experimental Botany 59, 891–905.

Kovalchuk N, Jia W, Eini O, et al. 2013. Optimization of TaDREB3 gene expression in transgenic barley using cold-inducible promoters. Plant Biotechnology Journal 11, 659–670.

Kovalchuk N, Smith J, Pallotta M, et al. 2009. Characterization of the wheat endosperm transfer cell-specific protein TaPR60. Plant Molecular Biology 71, 81–98.

Kumimoto RW, Adam L, Hymus GJ, Repetti PP, Reuber TL, Marion CM, Hempel FD, Ratcliffe OJ. 2008. The nuclear factor Y subunits NF-YB2 and NF-YB3 play additive roles in the promotion of flowering by inductive long-day photoperiods in Arabidopsis. Planta 228, 709–723.

Kumimoto RW, Zhang Y, Siefers N, Holt BFII. 2010. NF-YC3, NF-YC4 and NF-YC9 are required for CONSTANS-mediated, photoperiod-dependent flowering in Arabidopsis thaliana. The Plant Journal 63, 379–391.

Kwong RW, Bui AQ, Lee H, Kwong LW, Fischer RL, Goldberg RB, Harada JJ. 2003. LEAFY COTYLEDON1-LIKE defines a class of regulators essential for embryo development. The Plant Cell 15, 5–18.

Laloum T, De Mita S, Gamas P, Baudin M, Niebel A. 2013. CCAAT-box binding transcription factors in plants: Y so many? Trends in Plant Science 18, 157–166.

Laskowski RA, Moss DS, Thornton JM. 1993. Main-chain bond lengths and bond angles in protein structures. Journal of Molecular Biology 231, 1049–1067.

Lee H, Fischer RL, Goldberg RB, Harada JJ. 2003. Arabidopsis LEAFY COTYLEDON1 represents a functionally specialized subunit of the CCAAT binding transcription factor. Proceedings of the National Academy of Sciences, USA 100, 2152–2156.

Leticnic I, Doerks T, Bork P. 2012. SMART 7: recent updates to the protein domain annotation resource. Nucleic Acids Research 40, D302–D305.

Li Z, Hansen JL, Liu Y, Zemeta RS, Berger PH. 2004. Using real-time PCR to determine transgene copy number in wheat. Plant Molecular Biology Reporter 22, 179–188.

Liu J, Zhu JK. 1998. A calcium sensor homolog required for plant salt tolerance. Science 280, 1943–1945.

Liu JX, Howell SH. 2010. bZIP28 and NF-Y transcription factors are activated by ER stress and assemble into a transcriptional complex to regulate stress response genes in Arabidopsis. The Plant Cell 22, 782–796.

Lopato S, Bazanova N, Morran S, Milligan AS, Shirley N, Langridge P. 2006. Isolation of plant transcription factors using a modified yeast one-hybrid system. Plant Methods 2, 3–17.

Luan T, Ohito M, Yee KM, West MA, Lo R, Kwong RW, Yamagishi K, Fischer RL, Goldberg RB, Harada JJ. 1998. Arabidopsis LEAFY COTYLEDON1 is sufficient to induce embryo development in vegetative cells. Cell 93, 1195–1205.

Maity SN, de Crombrugghe B. 1996. Role of the CCAAT-binding protein CBF/NF-Y in transcription. Trends in Biochemical Sciences 23, 174–178.

Mantovani R. 1999. The molecular biology of the CCAAT-binding factor NF-Y. Gene 239, 15–27.

Marino-Ramirez L, Spouge JL, Kanga GC, Landsman D. 2004. Statistical analysis of over-represented words in human promoter sequences. Nucleic Acids Research 32, 949–958.
Miyoshi K, Ito Y, Serizawa A, Kurata N. 2003. OsHAP3 genes regulate chloroplast biogenesis in rice. The Plant Journal 36, 532–540.

Morran S, Eini O, Pyrovarenko T, Parent B, Singh R, Ismagul A, Elify S, Shirley N, Langridge P, Lopato S. 2011. Improvement of stress tolerance of wheat and barley by modulation of expression of DREB/CFB factors. Plant Biotechnology Journal 9, 230–249.

Nelson DE, Repetti PP, Adams TR, et al. 2007. Plant nuclear factor Y (NF-Y) B subunits confer drought tolerance and lead to improved corn yields on water-limited acres. Proceedings of the National Academy of Sciences, USA 104, 16450–16455.

Ni Z, Hu Z, Jiao Q, Zhang H. 2013. GmNFYA3, a target gene of miR169, is a positive regulator of plant tolerance to drought stress. Plant Molecular Biology 82, 113–129.

Oh SJ, Kwon CW, Choi DW, Song SI, Kim JK. 2007. Expression of barley HvCBF4 enhances tolerance to abiotic stress in transgenic rice. Plant Biotechnology Journal 5, 646–656.

Oh SJ, Song SI, Kim YS, Jang HJ, Kim SY, Kim M, Kim YK, Nahm BH, Kim JK. 2005. Arabidopsis CBF3/DREB1A and ABF3 in transgenic rice increased tolerance to abiotic stresses without stunting growth. Plant Physiology 138, 341–351.

Page RDM. 1996. TreeView: an application to display phylogenetic trees on personal computers. Computer Applications in the Biosciences 12, 357–358.

Pei J, Kim BH, Grishin NV. 2008. PROMALS3D: a tool for multiple protein sequence and structure alignments. Nucleic Acids Research 36, 2295–2300.

Petroni K, Kumimoto RW, Gnesutta N, Calvenzani V, Fornari M, Tonelli C, Holt BFII, Mantovani R. 2012. The promiscuous life of plant NUCLEAR FACTOR Y transcription factors. The Plant Cell 24, 4777–4792.

Qu B, He X, Wang J, et al. 2015. A wheat CCAAT box-binding transcription factor increases the grain yield of wheat with less fertilizer input. Plant Physiology 167, 411–423.

Riechmann JL, Heard J, Martin G, et al. 2000. Arabidopsis transcription factors: genome-wide comparative analysis among eukaryotes. Science 290, 2105–2110.

Romier C, Cocchiarella F, Mantovani R, Moras D. 2003. The NF-YB/NF-YC structure gives insight into DNA binding and transcription regulation by CCAAT factor NF-Y. Journal of Biological Chemistry 278, 1336–1345.

Shavrukov Y, Gupta NK, Miyazaki J, Baho MN, Chalmers KJ, Tester M, Langridge P, Collins NC. 2010. HvNax3—a locus controlling shoot sodium exclusion derived from wild barley (Hordeum vulgare ssp. spontaneum). Functional and Integrative Genomics 10, 277–291.

Sievers F, Wilm A, Dineen D, et al. 2011. Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. Molecular Systems Biology 7, 539.

Sinha S, Maity SN, Lu J, de Crombrugghe B. 1995. Recombinant rat CBF-C, the third subunit of NF-Y/FKF, allows formation of a protein–DNA complex with CBF-A and CBF-B and with yeast HAP2 and HAP3. Proceedings of the National Academy of Sciences, USA 92, 1624–1628.

Sippl MJ. 1993. Recognition of errors in three-dimensional structures of proteins. Proteins 17, 355–362.

Smith TF, Waterman MS. 1981. Identification of common molecular subsequences. Journal of Molecular Biology 147, 195–197.

Stephenson TJ, McIntyre CL, Collet C, Xue GP. 2007. Genome-wide identification and expression analysis of the NF-Y family of transcription factors in Triticum aestivum. Plant Molecular Biology 65, 77–92.

Stephenson TJ, McIntyre CL, Collet C, Xue GP. 2010. TaNF-YC11, one of the light-upregulated NF-YC members in Triticum aestivum, is co-regulated with photosynthesis-related genes. Functional and Integrative Genomics 10, 265–276.

Stephenson TJ, McIntyre CL, Collet C, Xue GP. 2011. TaNF-YB3 is involved in the regulation of photosynthesis genes in Triticum aestivum. Functional and Integrative Genomics 11, 327–340.

Suzuki Y, Tsunoda T, Sese J, et al. 2001. Identification and characterization of the potential promoter regions of 1031 kinds of human genes. Genome Research 11, 677–684.

Takumi S, Shimamura C, Kobayashi F. 2008. Increased freezing tolerance through up-regulation of downstream genes via the wheat CBFB gene in transgenic tobacco. Plant Physiology and Biochemistry 46, 205–211.

Vlahovicek K, Murvai J, Barta E, Pongor S. 2002. The SBASE protein domain library, release 9.0: an online resource for protein domain identification. Nucleic Acids Research 30, 273–275.

Wei X, Xu J, Guo H, Jiang L, Chen S, Yu C, Zhou Z, Hu P, Zhai H, Wan J, 2010. DTH8 suppresses flowering in rice, influencing plant height and yield potential simultaneously. Plant Physiology 153, 1747–1758.

Wu S, Zhang Y. 2007. LOMETS: a local meta-threading-server for protein structure prediction. Nucleic Acids Research 35, 3375–3382.

Yamamoto A, Kagaya Y, Toyoshima R, Kagaya M, Takeda S, Hattori A, 2003. Arabidopsis NF-Y subunits LEC1 and LEC1-LIKE activate transcription by interacting with seed-specific ABRE-binding factors. The Plant Journal 38, 843–856.

Yazawa K, Kamada H. 2007. Identification and characterization of carrot HAP factors that form a complex with the embryo-specific transcription factor C-LEC1. Journal of Experimental Botany 58, 3819–3828.

Yu Y, Li Y, Huang G, Meng Z, Zhang D, Wei J, Yan K, Zheng C, Zhang L. 2011. PwHAP5, a CCAAT-binding transcription factor, interacts with PwFKBP12 and plays a role in pollen tube growth orientation in Picea wilsonii. Journal of Experimental Botany 62, 4805–4817.

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