Alteration of Hematologic Parameters in Morphine-Dependent Rats by Long-Term Administration of Orexin Type 1 Receptor Antagonist

Zahra Piri¹, Masoumeh Kouros Arami ², Minoo Shahidi ³ and Somayeh Nazari ⁴

¹Department of Pharmacology and Toxicology, Faculty of Pharmacy, Pharmaceutical Sciences Branch, Islamic Azad University, Tehran, IR Iran
²Department of Neuroscience, School of Advanced Technologies in Medicine, Iran University of Medical Sciences, Tehran, IR Iran
³Department of Hematology, Faculty of Allied Medical Sciences, Iran University of Medical Sciences, Tehran, IR Iran
⁴Department of Physiology, Medical College, Shiraz University of Medical Sciences, Shiraz, IR Iran

*Corresponding author: Department of Neuroscience, School of Advanced Technologies in Medicine, Iran University of Medical Sciences, Tehran, IR Iran. Email: mkourosharami@gmail.com

Received 2019 October 26; Revised 2020 April 14; Accepted 2020 April 15.

Abstract

Background: Orexin peptides that are produced in the hypothalamic nuclei are involved in opioid dependence.

Objectives: In the current study, we aimed to figure out the effect of orexin type 1 receptor (OXR1) antagonist on hematologic factors in morphine-dependent rats.

Patients and Methods: Male Wistar rats were rendered morphine-dependent by subcutaneous injection of morphine sulfate (10 mg/Kg) at an interval of 12 hours twice a day for seven days. In the control and treatment groups, SB-334867 vehicle and SB-334867 were injected during postnatal days 1 to 30 (P1-P30) daily and then before each morphine injection during for days. Data were analyzed using unpaired two-tailed Student t-test and one-way analysis of variance (ANOVA). The defined level of statistical significance was P < 0.05.

Results: Morphine increased white blood cell count (WBC), platelet cell count, and hematocrit. Application of SB-334867 reduced several hematologic factors in morphine-dependent rats, including mean corpuscular hemoglobin concentration (MCHC), WBC, and platelet count compared to morphine-dependent rats.

Conclusions: Inhibition of OXR1 may improve morphine-induced changes in hematologic factors in morphine-dependent rats.

Keywords: SB-334867, Morphine, Dependent Rat, WBC, MCHC, PLT, Hematologic Factors, Leukocyte Count

1. Background

Long-term opioid drug usage results in the development of dependence and tolerance that reduces their beneficial use and creates severe health and social concerns (1). In numerous forms of drug addiction, negative and positive reinforcement include two main elements. Positive strengthening of euphoric consequences leads to leads to pursuing drug, while negative reinforcement of withdrawal signs happens following an interruption of opioid taking (2-4). Opiate dependence is a multifaceted occurrence that engages several areas of the brain (2, 5-8).

The neuropeptides orexin A and orexin B are synthesized, particularly in the hypothalamus, and act on target neurons throughout the central nervous system via two G-protein-coupled receptors, orexin 1 (OXR1) and 2 receptors (OXR2) (9, 10). In the previous studies, it has been revealed that orexergic neurons of the dorsomedial hypothalamus and perifornical area are activated by foot shock stimulus and are implicated in the negative reinforcement of withdrawal symptoms (4, 11). Nucleus raphe magnus that is a thermoregulatory center (12, 13) has high densities of not only opiate receptors but also orexin receptors (14). Orexinergic neurons have projections to the visual cortex that send information to the lateral geniculate nucleus (LGN). Accordingly, LGN and many other thalamic nuclei have orexin receptors (15, 16). Orexin is involved in long-term potentiation that leads to learning and memory (17, 18). Another study described that the injection of orexin in the nucleus tractus solitarius that is a regulator of the cardiovascular system, causes blood pressure increase (19-21). Recently, compelling studies have demonstrated a novel and significant role for the orexin neuronal system in reward processing and addiction. Morphine-conditioned animals had greater Fos-activated orexin neurons than non-conditioned ones (22). Orexin is involved in reward processing and addiction in the nucleus accumbens, ventral tegmental area, and the locus coeruleus that receives glutamatergic afferents mainly from paragiganto cellular nucleus (14, 23-26).
Although several pieces of evidence demonstrated an effect of orexins in arousal and preservation of the waking conditions (27), additional evidence supports a significant and particular role in reward procedures and drug abuse as well (28). Injection of the OXR1 antagonist SB-334867 systemically or into the ventral tegmental area prevented the acquirement of cocaine sensitization (29). Furthermore, orexin is implicated in cue-induced drug craving and motivation to take cocaine when a high effort is essential to acquire the drug, but not in the principal reinforcing properties of cocaine itself (30).

In the previous studies, hematological factors in heroin and opioid-dependent subjects were investigated (31, 32). Nevertheless, in heroin-dependent groups, several alterations occur in immune function and blood lymphocytes (33). Similarly, other studies displayed that opioid intake may exacerbate along with the progress of infectious diseases (31). On the other hand, it was revealed that inhibition of OXR1 reduced the development of morphine tolerance and physical dependence in rats (34). Totally, previous studies concluded that SB-334867 was used for the decrement of morphine dependence and withdrawal behaviors (35).

2. Objectives

Therefore, we aimed to investigate the effect of OXR1 antagonist injection on hematologic factors in morphine-dependent rats.

3. Patients and Methods

3.1. Drugs

Morphine sulfate (Temad, Tehran, Iran) was dissolved in a volume of 10 mg per 1 ml physiological saline then was administered subcutaneously. The selective OXR1 antagonist SB-334867 (Tocris, Bristol, UK) was dissolved in 2% dimethyl sulfoxide (DMSO), 10% 2-hydroxypropyl-β-cyclodextrin in sterile water purchased from Sigma-Aldrich, Germany.

3.2. Animals

Male Wistar rats were accommodated in Plexiglas breeding cages in groups of four for each cage with woodchip bedding and ad libitum access to food and water. Animals were housed in a colony room at ambient temperature and on 12-h light/dark cycles (the light period started at 7 a.m.). Efforts were made to diminish animal distress and reduce the number of animals that were used. All experiments were done in accordance with the ethical guidelines set by the “Ethical Committee of Iran University of Medical Sciences”, which are according to the “NIH Guide for the Care and Use of Laboratory Animals”.

3.3. Induction of Morphine Dependence

To induce morphine dependence, morphine was injected (10 mg/Kg, i.m., subcutaneous [s.c.]) twice a day with an interval of 12 hours for seven days (36).

3.3.1. Assessment of Hematologic Factors

To detect the level of hematological parameters, the subsequent steps were carried out: 2 ml fresh venous blood was collected in test tubes comprising specific EDTA anticoagulant. Afterward, the subsequent tests were conducted on the samples by the usage of Counter Sysmex:

1. Complete blood cell count (CBC) for red and white blood cell
2. Hemoglobin (HGB) level
3. Hematocrit percentage (HCT)
4. Cell indices, including mean corpuscular hemoglobin (MCH), mean corpuscular volume (MCV), and mean corpuscular hemoglobin concentration (MCHC)
5. Differential leukocyte count (neutrophils)
6. Blood platelets (PLT)

3.4. Experimental Groups

Rats were divided into five different experimental groups as the following: Group 1 (sham), rats were naïve without any injection (n = 6); Group 2 (control), rats received SB-334867 vehicle (2% dimethyl sulfoxide (DMSO), 10% 2-hydroxypropyl-β-cyclodextrin in sterile water) P1-30 (37) and then DMSO+ cyclodextrin +Saline (morphine vehicle) P31-P37 (n = 8); Group 3 (M); animals (P31) received morphine administration (10 mg/kg, 1 mL, s.c.) for seven days twice a day (n = 8); Group 4 (SB), animals received SB-334867 (20 mg/kg, Intraperitoneal [i.p.]) (4) daily from P1-P37 (n = 6). Group 5 (SB+M), animals (n = 6) received SB-334867 (20 mg/kg, i.p.) (4) daily from P1-P30 and then before each morphine injection for seven days.

3.5. Data Analysis

Data are expressed as mean ± SEM and were analyzed using unpaired two-tailed Student t-test and one-way analysis of variance (ANOVA) for the comparison of two or more groups, respectively. The defined level of statistical significance was P < 0.05.

4. Results

No significant differences were observed in the WBC, MCHC, HCT, and platelet levels between the naïve group and vehicle-treated rats (Table 1). The comparison between naïve and vehicle-treated rats displayed that the injection process has not any effect on the level of mentioned hematologic factors.
Table 1. Comparison Between Rats Who Received SB-334867 Vehicle Injections with Naïve Ones (Naïve/ SB Vehicle)

| Groups | Hem. Factor | Naïve | SB-334867 Vehicle | Difference |
|--------|-------------|-------|-------------------|------------|
|        | WBC (× 10³/µL) | 4.5 ± 1.15 | 3.73 ± 0.39 | NS        |
|        | HCT (%)     | 35.50 ± 4.20 | 30.75 ± 1.72 | NS        |
|        | MCHC (%)    | 29.85 ± 0.05 | 31.13 ± 0.33 | NS        |
|        | Platelet (× 10³/µL) | 360.7 ± 16.23 | 383.6 ± 5.87 | NS        |

Abbreviations: NS, non-significant

4.1. The Effects of Morphine Addiction and Orexin Antagonist on WBC Counts

The results of our study indicated that the mean number of total WBC in the morphine-dependent rats (5.550 ± 0.289 × 10³ cells/µL) increased compared to the control group (3.733 ± 0.393 × 10³ cells/µL) while SB-334867 could decrease the WBC in morphine-dependent rats (3.067 ± 0.470 × 10³ cells/µL) compared to control rats (P < 0.01, Figure 1).

4.2. The Effects of Morphine Addiction and Orexin Antagonist on Hematocrit Level

The results of the current study indicated that the mean number of HCT in the morphine-addicted rats (36.8 ± 2.78%) increased (P < 0.05) compared to the control group (30.7 ± 3.45%), while SB-334867 failed to change the HCT in morphine-dependent rats (37.8 ± 1.79%) compared to the morphine-addicted rats (Figure 2).

4.3. The Effects of Morphine Addiction and Orexin Antagonist on MCHC Counts

The results of our study indicated that the MCHC in the morphine-addicted rats (34.35 ± 0.45%) increased compared to the control group (31.13 ± 0.33%), while SB-334867 decreased the WBC in morphine-dependent rats (22.98 ± 2.35%) compared to the control rats (P < 0.01, Figure 3).

4.4. The Effects of Morphine Addiction and Orexin Antagonist on Platelet Counts

The platelet count was increased by morphine (592.2 ± 38.970 × 10³ cells/µL) compared to the control group (383.600 ± 5.87 × 10³ cells/µL) (P < 0.01). The injection of SB-334867 in morphine-dependent rats decreased, and so returned platelet counts to the control level (Figure 4).

The lymphocyte numbers in morphine-dependent rats noticeably decreased compared to the control group (2726 ± 99 and 1589 ± 155 cells/µL, P < 0.001, unpaired t-test), while the number of eosinophils in morphine-dependent rats (2726 ± 96 cells/µL) showed no difference in comparison to the control group (2726 ± 95 cells/µL, unpaired t-test). SB-334867 failed to change the number of lymphocytes and eosinophil in the morphine-dependent rats compared to the control group.
Figure 3. The effect of SB-334867 on the mean corpuscular hemoglobin concentration (MCHC) of morphine-dependent rats (M). The graph displays the MCHC in morphine-dependent rats who received SB-334867 (orexin antagonist) (SBc+M) compared to control and morphine-injected rats (M). Data are shown as mean ± SEM (*P < 0.05, **P < 0.01).

Figure 4. Alteration of the platelet count in SB-334867-(orexin antagonist) injected rats and those who received SB-334867 before morphine injection (SBc+M) compared to control and morphine (M) dependent rats. Data are presented as mean ± SEM (**P < 0.01).

were observed in the MCV level between the morphine-addicted (63.6 ± 1.07 fL) and control groups (65.2 ± 2.81 fL) as well as between the SB-treated (63.7 ± 1.67 fL) and the SBc+M (65.8 ± 1.97 fL) rats (Table 2).

The MCH level (22.6 ± 2.08 pg) did not show any difference in the morphine-dependent group in comparison to the control group (21.5 ± 2.49 pg). In addition, SB-treated (19.7 ± 0.61 pg) and SBc+M (20.5 ± 0.45 pg) had not any significant difference (Table 2).

The percentage of the red blood cell distribution width (RDW) in the morphine-dependent group (20.6 ± 0.28) did not reveal any significant difference compared to the control group (23.0 ± 5.42). In addition, there was no significant difference in the RDW level between the SB-treated rats (22.4 ± 2.08) and SBc+M-treated ones (18.4 ± 0.36) (Table 2).

5. Discussion

Along with numerous problems derived from narcotic drugs, the results of the current research unraveled the intense outcomes of morphine on the total and differential counts of peripheral WBC and platelets, as well as HCT and MCHC levels. Furthermore, the current results demonstrated that orexin antagonist was involved in some changes caused by morphine, and hence, it might suppress several depraved effects of morphine on the above-mentioned hematologic factors.

5.1. The Effect of Orexin Antagonist on WBC in Morphine-Dependent Rats

In this study, it has been revealed that morphine addiction shows an intense impact on the mean total WBC count. In previous studies, it has been revealed that morphine, as a component of opium, prompts catecholamine release, which is identified to raise the leukocyte count (38). Rising of the peripheral blood leukocyte count also might be augmented by direct injury of epithelial and endothelial surfaces and/or alterations in cytokine levels (especially interleukin-6 [IL-6]) produced by constituents of opium (39).

It would be mentioned that the hemopoiesis is controlled by a multipart network of cytokines like colony-stimulating factors. It has been established that opium or some of its derivatives may play a role in the secretion of cytokines such as IL-2, IL-4, IL-5, IL-10, IFN-γ, and TGF-β (40), thereby influencing the production of WBC.

It is previously demonstrated that in heroin-dependent people without any malnutrition, blood T cell count was augmented (41, 42). In addition, there are several reports suggesting that the treatment of heroin and morphine would strengthen some parameters of the immune system. In this regard, it was revealed that the creation of several cytokines was augmented a few minutes after morphine injection in mice (32). Moreover, evidence displayed that the monocyte count was greater than the normal level in heroin-addicted persons (43, 44). Morphine is known as the agonist of µ opioids receptors and the central and the active metabolite of heroin. The possible mechanism of the morphine effect might be the regulation of the immune system either directly through mu opiate receptors situated on the immune cells, or indirectly via a central way with the mu receptors in the central nervous system.

In a previous study on the morphine-dependent dogs, it was demonstrated that morphine could not change the
OXR1 and 2 were expressed on human CD34 morphine-dependent rats. It was also revealed that SB-334867 significantly decreased WBC counts in the derivatives on lymphocytes (49, 50). This may be ascribed to the apoptotic aspects of opium or its effects on lymphocytes counts. The mentioned outcomes exemplify that opium addiction has intense adverse effects (47). Moreover, a diminished number of lymphocytes has been reported in addicted dogs (48). The current results indicated that the injection of SB-334867 significantly decreased MCHC in morphine-dependent rats, the results were not statistically significant. Hence, it may propose that the HCT increment was higher than that of HGB, which could decrease the MCHC level indeed.

Table 2. Hematologic Factors in Saline (Control), Morphine (M), and SB-334867 (Orexin Antagonist) Injected Rats (SBc) and Those Who Received SB-334867 Before Morphine Injection (SBc+M)*

| Index                        | SB Vehicle | M        | SBc   | SBc+M   |
|------------------------------|------------|----------|-------|---------|
| RBC (x10^12/L)               | 5.27 ± 0.28| 5.36 ± 0.07| 5.72 ± 0.4 | 6.098 ± 0.40 |
| HGB (g/dL)                   | 10.3 ± 1.07| 11.7 ± 1.24| 11.3 ± 1.18| 12.5 ± 1.96  |
| MCV (fL)                     | 65.2 ± 2.81| 63.6 ± 1.07| 63.7 ± 1.67| 65.8 ± 1.97  |
| MCH (pgm)                    | 21.5 ± 2.49| 22.6 ± 2.08| 19.7 ± 0.61| 20.5 ± 0.45  |
| RDW (%)                      | 23.0 ± 5.42| 20.6 ± 2.08| 22.4 ± 2.08| 18.4 ± 0.36  |

Abbreviations: HGB, hemoglobin; MCH, mean cell hemoglobin; MCV, mean corpuscular volume; RBC, red blood cell; RDW, red cell distribution width.

*Data are shown as mean ± SEM.

5.2. The Effects of Orexin Antagonist on Hematocrit Level and MCHC in Morphine-Dependent Rats

In the current study, morphine did not change MCHC significantly. It is consistent with previous studies, demonstrating that six-month heroin addiction did not make key alterations in erythrocyte parameters, for instance, RBC count and HGB levels (55). Correspondingly, it was revealed that the blood HGB content did not change in morphine-dependent dogs (56). In comparison with the present study, it was revealed that blood HGB was augmented in people who used heroin. Nevertheless, in methadone-addicted people, the amount of HGB reduced to its normal level (31). In another study, it was revealed that HGB concentration decreased in morphine-dependent people, but they found that HCT increased by morphine (31). Thus, the increment of both HGB and HCT in morphine-dependent subjects may lead to MCHC stability by morphine. The current results, however, indicated that the injection of SB-334867 significantly decreased MCHC in morphine-dependent rats. The previous studies demonstrated that stimulation with orexin A and B led to a significant decline of erythroid precursors burst forming unit erythrocyte (BFU-E) and colony-forming unit erythrocyte (CFU-E) (54). Hence, HCT reduction by orexin could be due to the diminishing of these erythroid precursors.

In addition, it was discovered that orexin-A usage enhanced the expression of nuclear factor erythroid-derived 2 related factor 2 and antioxidant response element luciferase activity, resulting in the generation of the cytoprotective enzyme heme oxygenase-1 (HO-1) (57). This could catalyze heme to biliverdin, carbon monoxide, and free iron (58). Consequently, it may be proposed that orexin decreased HGB by HO-1 activating, and as a result, SB-334867 could increase the HGB.

Although in this study, HGB and HCT increased by SB-334867 in morphine-dependent rats, the results were not statistically significant. Hence, it may propose that the HCT increment was higher than that of HGB, which could decrease the MCHC level indeed.
5.3. The Effect of Orexin Antagonist on Platelet Number in Morphine-Dependent Rats

Our results demonstrated that morphine injection could increase platelet counts. Enhanced platelet activity amplifies significantly the risk of arterial thrombotic diseases, including stroke, peripheral ischemia, and myocardial infarction (59).

Our results demonstrated that SB-334867 injection could return platelet counts to the control level in morphine-dependent rats; thus may prevent numerous cardiovascular disorders, which could be due to morphine dependence. Previous studies have been revealed that orexins augmented the excitability and synchronization of rat sympathetic preganglionic neurons. Centrally administered orexins display cardiovascular effects, comprising a rise of blood pressure and heart rate (60). Therefore, totally, it is concluded that SB-334867 can prevent cardiovascular diseases directly by orexin receptor antagonization or indirectly by the prevention of morphine effect on platelet counts.

The current study, however, indicated that SB-334867 did not alter the results of the rest of hematologic factors, including MCV, MCH, RBC, RDW, HGB in morphine-dependent rats. This might be due to the great level of deviation detected in these factors.

5.4. Conclusion

SB-334867 reduced MCHC, WBC, and platelet counts that were increased in morphine-dependent rats. Therefore, totally it may be concluded that blockade of ORXRI can improve morphine-induced changes in hematologic factors of morphine-dependent rats.

Acknowledgments

This work is supported by the Neuroscience Research Center and Department of Basic Sciences in Faculty of Allied Medicine of Iran University of Medical Sciences.

Footnotes

Authors’ Contribution: Zahra Piri: acquisition of data and statistical analysis; Masoumeh Kourosh Arami: study concept and design and drafting the manuscript; Minoo Shahidi: helped in drafting the manuscript and reviewed content; Somayeh Nazari: helped in drafting the manuscript and revision

Conflict of Interests: The authors declare that they have no conflict of interests.

Ethical Approval: There is none to be declared.

Funding/Support: The authors received no financial support for the research.

References

1. Zhu H, Zhou W. Discharge activities of neurons in the nucleus paragigantocellularis during the development of morphine tolerance and dependence: a single unit study in chronically implanted rats. Eur J Pharmacol. 2006;536(1-2):65-72.
2. Koob GF, Maldonado R, Stinus L. Neural substrates of opiate withdrawal. Trends in neurosci. 1992;15(5):186-91.
3. Rezaei Z, Kourosh-Arami M, Azizi H, Semnanian S. Orexin type-1 receptor inhibition in the rat lateral paragigantocellularis nucleus attenuates development of morphine dependence. Neurosci Lett. 2020;734875.
4. Shariﬁ R, Sarhan M, DiLeone RJ. Orexin mediates the expression of precipitated morphine withdrawal and concurrent activation of the nucleus accumbens shell. Biol psychiatry. 2008;64(4):275-83.
5. Abood LG. Mechanisms of tolerance and dependence: An overview. Mechanisms of tolerance and dependence. 1984:4.
6. Kantak KM, Miczek KA. Social, motor, and autonomic signs of morphine withdrawal: differential sensitivities to catecholaminergic drugs in mice. Psychopharmacology. 1988;96(4):468-76.
7. Maldonado R, Stinus L, Gold LH, Koob GF. Role of different brain structures in the expression of the physical morphine withdrawal syndrome. Journal of Pharmacology and Experimental Therapeutics. 1992;261(2):669-77.
8. Redmond Jr DE, Krystal JH. Multiple mechanisms of withdrawal from opioid drugs. Annu Rev Neurosci. 1984;7(1):443-78.
9. Babasafari M, Kourosharami M, Behman J, Farhadi M, Komaki A. Alteration of Phospholipase C Expression in Rat Visual Cortical Neurons by Chronic Blockade of Orexin Receptor 1. Int J Pept Res Ther. 2019. doi: 10.1007/s10998-019-09941-y.
10. Mieda M, Tsuchino N, Sakurai T. Differential roles of orexin receptors in the regulation of sleep/wakefulness. Front Endocrinol. 2013;4:57.
11. Harris GC, Wimmer M, Aston-Jones G. A role for lateral hypothalamic orexin neurons in reward seeking. Nature. 2005;437(7058):556.
12. Arami MK. Nitric oxide in the nucleus raphe magnus modulates cutaneous blood flow in rats during hypothermia. Iran J Basic Med Sci. 2015;18(10):598.
13. Malakouti SM, Kourosh Arami M, Sarari A, Hajjazadeh S, Behzadi G, Shahidi S, et al. Reversible inactivation and excitation of nucleus raphe magnus can modulate tail blood flow of male wistar rats in response to hypothermia. Iran Biomed J. 2008;12(2):237-40.
14. Peyron C, Tighe DK, Van Den Pol AN, De Leece I, Heller HC, Sutcliffe JG, et al. Neurons containing hypocretin (orexin) project to multiple neuronal systems. J Neurosci. 1999;19(21):9996-10015.
15. Govindaiah G, Cox CL. Modulation of thalamic neuron excitability by orexins. Neuropharmacol. 2006;51(3):414-25.
16. Arami MK, Sohya K, Sarari A, Jiang B, Yanagawa Y, Tsumoto T. Reciprocal homosynaptic and heterosynaptic long-term plasticity of corticogenulate projection neurons in layer VI of the mouse visual cortex. J Neurosci. 2013;33(18):7787-98.
17. Akbari I, Motamedif D, Davoodi FG, Norobarakahshia M, Ghahreman E. Orexin-1 receptor mediates long-term potentiation in the dentate gyrus area of freely moving rats. Behav Brain Res. 2011;216(1):375-80.
18. Komaki A, Shahidi S, Sarari A, Hasanein P, Lashgari R, Haghparast A, et al. Effects of neonatal C-fiber depletion on interaction between neocortical short-term and long-term plasticity. Basic and clinical neuroscience. 2015;4(2):336.
19. Arami MK, Sarari A, Behzadi J, Malakouti SM, Amiriz I, Ekbatani RZ. The effect of hyperglycemia on nitric oxidergic neurons in nucleus tractus solitarius and blood pressure regulation in rats with induced diabetes. Iranian Journal of Diabetes and Lipid Disorders. 2005;4(3). E2.
20. Kourosh Arami M, Sarari A, Malakouti SM, Behzadi G, Vahabian M, Amiriz I. The Effect of Nucleus Tractus Solitarius Nitric Oxidergic Neurons on Blood Pressure in Diabetic Rats. Iran Biomed J. 2006;10(1):15-9.
21. Smith PM, Connolly BC, Ferguson AV. Microinjection of orexin into the rat nucleus tractus solitarius causes increases in blood pressure. *Brain Res*. 2002;950(1-2):261-7.

22. Brown RA, Walling SG, Milway JS, Harley CW. Locus ceruleus activation suppresses feedforward interneurons and reduces β-γ electroencephalogram frequencies while it enhances β frequencies in rat dentate gyrus. *J Neurosci*. 2005;25(8):3985-91.

23. Arami MK, Hajizadeh S, Semnanian S. Postnatal development changes in excitatory synaptic activity in the rat locus coeruleus neurons. *Brain Res*. 2016;1648:165-71.

24. Fadel J, Deutch AY. Anatomical substrates of orexin–dopamine interactions: lateral hypothalamic projections to the ventral tegmental area. *Neuroscience*. 2002;111(2):379-87.

25. Georgescu D, Zachariou V, Barrot M, Mieda M, Willie JT, Eisich AJ, et al. Involvement of the lateral hypothalamic peptide orexin in morphine dependence and withdrawal. *The Journal of neuroscience*. 2003;23(8):3106-11.

26. Arami MK, Semnanian S, Javan M, Hajizadeh S, Sarihi A. Postnatal development changes in excitatory synaptic activity in the rat locus coeruleus neurons. *Brain Res*. 2016;1648:165-71.

27. de Lecea L. Hypocretins and the neurobiology of sleep-wake mechanisms. *Prog Brain Res*. 2003;137:365–71.

28. Baimel C, Borgland SL. Hypocretin modulation of drug-induced synaptic plasticity. *Prog Brain Res*. 2012;198:123-31. doi: 10.1016/S0079-6194(12)98900-2. [PubMed: 22813972].

29. Borgland SL, Taha SA, Sarti F, Fields HL, Bonci A. Orexin A in the VTA is necessary for drug-induced locomotion and preference for opioids. *The Journal of neuroscience*. 2012;32(18):6034–47.

30. Mahler SV, Smith RJ, Moorman DE, Sartor GC, Aston-Jones G. Multiple roles for orexin/hypocretin in addiction. *Protein Pept Lett*. 2013;20(4):337-48.

31. Haghpanah T, Afarinesh M, Divsalar K. A review on hematological factors of rat. *Physiol Pharmacol Tissue Pharmacol*. 2012;578:950-9.

32. de Lecea L, Hypocretins and the neurobiology of sleep-wake mechanisms. *Prog Brain Res*. 2003;137:365–71.

33. Govitrapong P, Suttitum T, Kotchabhakdi N, Uneklabh T. Alterations in the levels of interleukin-6, interleukin-1β, tumor necrosis factor α, and interferon γ in heroin and morphine-treated mice. *Int J Addict Health*. 2014;3(1):1-8.

34. Piri Z et al. Orexins induce increased excitability and synchronisation of rat dentate gyrus. *Mol Neurobiol*. 2015;51(4):1777-89.

35. Patil SR, Hartwig JH, Italiano JE. The biogenesis of platelets from megakaryocyte proplatelets. *J Clin Invest*. 2005;115(12):3348-54.

36. Mousavi Y, Azizi H, Mirnajafi-Zadeh J, Javan M, Semnanian S. Blockade of orexin receptor 1 attenuates the development of morphine dependency in rats. *European J Pharmacol*. 2018;843:195-201.

37. Wiskerke J, James MH, Aston-Jones G. The orexin-1 receptor antagonist SB-334867 reduces motivation, but not inhibitory control, in a rat stop signal task. *Brain Res*. 2019;1775:97-105.

38. Nair R, Saini D, Garg S, Dhawan S, Agrawal S. The effect ofSB-334867 on morphine-induced hyperactivity in rats. *J Pharm Pharmacol*. 2007;59(8):1095-1100.

39. McCarty MF. Interleukin-6 as a central mediator of cardiovascular disease. *Curr Opin Pharmacol*. 2002;2(3):15-24. doi: 10.1016/b978-0-444-59489-1.00008-2. [PubMed: 22813972].

40. Ohara T, Itoh T, Takahashi Z, Takahashi M. Immunosuppression by morphine. *Fa yi xue za zhi*. 1981;28(1):1-8.

41. Puri P, Semnanian S, Haghparast A. Blockade of orexin type-1 receptors in locus coeruleus nucleus attenuates the development of morphine dependency in rats. *J Pharmacol Exp Ther*. 2014;350(2):330-8. doi: 10.1124/jpet.113.22813972.

42. Louria DB, Hensle T, Rose J. The major medical complications of cancer patients with opioid addiction. *Am J Med*. 1986;80(3):79-81.

43. Roy S, Ramakrishnan S, Loh HH, Lee NM. Chronic morphine treatment selectively suppresses macrophage colony formation in bone marrow. *Eur J Pharmacol*. 1999;379(3):359-63.

44. Tamba-Berehoiu RADIANA, Popa NC, Popescu S, Popa C. Research regarding the toxic effects of heroin consumption on human homeostacogramm. *Rom Biotech Lett*. 2008;13(3):3605-11.

45. Pierce HI, Plant OH. Studies in chronic morphine poisoning in dogs II. Changes in blood cells and hemoglobin during addiction and withdrawal. *J Pharmacol Exp Ther*. 1982:213(3):359-70.

46. Roy S, Ramakrishnan S, Loh HH, Lee NM. Chronic morphine treatment selectively suppresses macrophage colony formation in bone marrow. *Eur J Pharmacol*. 1999;379(3):359-63.

47. Asadikaram G, Sirati-Sabet M, Asabandeh M, Shahrokhi M, Jafarzadeh A, Khaksari M. Hematological changes in opium addicted diabetic rats. *Int J High Risk Behav Addict*. 2013;2(4):173-81.

48. Kuang YM, Zhu YC, Kuang Y, Sun Y, Hua C, He WY. Changes of the immune cells, cytokines and growth hormone in teenager drug addicts. *Chinese J Cell Mol Immunol*. 2007;23(9):821-3.

49. Liu XS, Zang LQ, Hao ZR, Li ZH, Liu SP, Chen YC, et al. Apoptosis of cultured cortical neurons of rat's brain induced by heroin. *Fa yi xue za zhi*. 2007;27(1):34-7.

50. Ohara T, Itoh T, Takahashi M. Immunosuppression by morphine-induced lymphocyte apoptosis: is it a real issue? *Anesth Analg*. 2005;100(4):1617-22.

51. Kalinkovich A, Spiegel A, Shvitol S, Kollet O, Jordanay N, Piabello W, et al. Blood-forming stem cells are nervous: direct and indirect regulation of immature human CD34+ cells by the nervous system. *Brain Behav Immun*. 2009;23(8):1059-65.

52. Xu L, Fukumura D, Jain RK. Acidic extracellular pH induces vascular endothelial growth factor (VEGF) in human glioblastoma cells via ERK1/2 MAPK signaling pathway. *Mechanism of low pH-induced VEGF*. *J Biol Chem*. 2002;277(1):168-74.

53. Tang J, Chen J, Ramanjanyeya M, Punn A, Conner AC, Randeva HS. The signalling profile of recombinant human orexin-2 receptor. *Cell Signal*. 2008;20(9):1651-61.

54. Haxinen M, Hakanen A, Aalto K, Saarinen J, Sarmozov A, et al. The Neuropeptides Orexin a and B Have An Impact on Functional Properties of Human CD34+ Stem and Progenitor Cells. *Blood*. 2008;111(12):3939-46.

55. Kim J, Li M, Jang J, Na H, Song N, Lee C, et al. 15-Deoxy-D12,14-prostaglandin J2 rescues PC12 cells from H2O2-induced apoptosis through Nrf2-mediated upregulation of heme oxygenase-1: potential roles of Akt and ERK2. *Biochem Pharmacol*. 2008;76(11):1577-89.

56. Poss KD, Tonegawa S. Heme oxygenase-1 is required for mammalian iron utilization. *Proceedings of the National Academy of Sciences*. 1997;94(20):10919-24.

57. Patel SR, Hartwig JH, Italiano JE. The biogenesis of platelets from megakaryocyte proplatelets. *J Clin Invest*. 2005;115(12):3348-54.

58. van den Top M, Nolans M, Lee K, Richardson PJ, Buijs RM, Davies CH, et al. Orexins induce increased excitability and synchronisation of rat sympathetic preganglionic neurones. *J Physiol*. 2003;549(3):809-21.

Int J High Risk Behav Addict. 2020; 9(3):e99081.