Distribution and Protective Role of HLAB40 in Iranian Patients with Kawasaki Disease; a Report from Southern Iran

Gholamhossein Ajami1, MD; Khashayar Aflaki2, MD; Soheila Alyasin3, MD; Behrooz Gharesi-fard4, MD; Mohammad Borzouee2, MD; Hamid Amoozgar*5, MD

1Department of Pediatrics Cardiology, 2Department of Pediatrics Intensive Care, 3Department of Pediatric Immunology, 4Department of Immunology, 5Cardiac and Neonatology Research Center, Shiraz University of Medical Sciences, Shiraz, Iran

Abstract

Objective: Kawasaki disease (KD) clinically presents as a systemic vasculitis syndrome with significant cardiovascular involvement. With different incidence among different ethnic groups, the role of certain human leukocyte antigens and their products has been considered as a crucial predisposing factor in the immune responses in this disease.

Methods: We determined the distribution of human leukocyte antigens type B for 90 Iranian patients with Kawasaki disease in order to evaluate a possible association between these antigens and this disease in our area. We used the polymerase chain reaction (PCR) sequence specific primers (PCR-SSP) technique for antigen typing. Distribution of these antigens for 89 healthy Iranians used as control.

Findings: While 7 (3.9%) of our patients were positive for human leukocyte antigen type B 40*, there were 18 (10.1%) subjects from the control group who had this antigen with statistically significant difference between patients and control group (CI= 95%, RR=1.15 and P= 0.02). Data were analyzed by Pearson chi-square test and Fisher’s exact test. SPSS version 15 was used for statistical analysis and a P value less than 0.05 considered statistically significant

Conclusion: The presence of higher frequency of allele type-B40* in the control group may represent a protective role for this antigen with resultant decreased susceptibility to KD in our area.

Key Words: Kawasaki Disease; Gene Protection; Human Leukocyte Antigens; HLA-B40* Allele

Introduction

Kawasaki disease (KD) is an acute febrile illness of childhood which presents as a systemic disease. While many organs are affected, involvement of cardiovascular system during early and late stages of the disease is considered as the leading cause of acquired heart disease in children in developed countries[1,2]. With more recognition of KD, it seems that the reported incidence is also increasing in some developing countries, even more than attacks of acute rheumatic fever in this age group[3-5].

While several decades have elapsed since Dr. Tomisaku Kawasaki described this disease for the first time, the cause is still unknown. Furthermore, epidemiological studies have revealed different incidence among different ethnic groups, and so
the presence of certain human leukocyte antigens has been considered as a possible inherited predisposition for development of the disease[6-13]. It is postulated that exposure of susceptible person to the triggering pathogen may cause release of certain inflammatory mediators which can provoke vasculitis as the main pathological process with resultant clinical manifestations[14]. In this study we determined the distribution of human leukocyte B antigens among 90 Iranian patients with diagnosis of KD according to American Heart Association criteria[15]. We used the PCR-SSP technique as described in literature for antigen typing[16-19].

**Subjects and Methods**

**The study population:** The study population consisted of 90 Iranian patients with final diagnosis of KD at the time of hospital discharge between 1991 and 2008 who were under follow up of pediatric immunology and pediatric cardiology services of Shiraz University of Medical Sciences, Shiraz, Iran. One hundred and nineteen patients were found to fulfill classic criteria of Kawasaki disease according to American Heart Association, among which 90 patients included in our study. Exclusion criteria were all patients presenting less than 4 criteria of Kawasaki. All of the patients were residents of south Iran. Informed consent for participation was obtained from the parents or their guardians. HLA-B distribution of 89 healthy Iranians was used as control[20]. The protocol of the study was approved by Ethical Committee of Shiraz University.

**Method:** Two samples of one milliliter of blood was obtained from each subject and preserved in -20°C for Antigen typing by PCR.

**DNA extraction:** Total DNA was extracted from 200 μL of whole blood by column based method using amplification DNA blood mini Kit (Qiagnene, England).

**Determination of DNA concentration:** DNA concentration was measured by absorbance of the samples at 260 nm and 280 nm versus distilled water as blank. Optical density ratio between 260 nm/280 nm of all samples was above 1.8. The following formula was used to quantify DNA concentration. DNA (μg/ml)=Optical density 260 nm×dilution factor×50.

Finally DNA concentration was adjusted to 50-100 μg/ml with distilled water and stored at -20°C for analysis.

**Antigen-B typing:** Typing was performed by method of PCR-SSP, using BAG HISTO type low resolution kit (BAG Health Care GmbH, Germany). Genomic DNA (50-100 μg/ml) was amplified by PCR-SSP method with 48 pairs of sequence specific primers. Internal positive control was included in each reaction. A total of 48 reactions were done for each sample. This method is based on the fact that primer extension and hence successful PCR relies on an exact match at the 3-end of both primers. So, only if the primers entirely match the target sequence, amplification is obtained (Fig.1). For the PCR procedure the following components were included in 10 μL of reaction mixture: 1 × Polymerase chain reaction buffer (includes dNTPs1, MgC12), Taq DNA polymerase, internal and sequence specific primers, template DNA. Amplification of PCR-SSP was carried out by using Eppendorf Mastercycler.

**Gel electrophoresis:** PCR products (48 for each sample) were subjected to 3% Agarose gel electrophoresis at 100-150 volt for 30-60 minutes. Then after termination of the run, the gels were stained in ethidium bromide solution for 30 minutes, and under ultraviolet illumination (220-310 nm) PCR products were visualized (Fig. 2).

**Documentation and interpretation:** For interpretation of the results, specific tables and
evaluation diagram were used. Only bands with correct size were considered as positive reaction. Internal positive control bands were also observed in all reactions. For further reassurance, score evaluation software (BAG Health Care, Germany) was used for analysis of the results. The control group included 89 healthy Iranians who were residents of Shiraz and selected from a population pool on whom a genetic study was done by Farjadian et al in the area\textsuperscript{[20]}. Data were analyzed by Pearson chi-square test and Fisher’s exact test according to the frequency specific type antigen in the patients. SPSS version 15 was used for statistical analysis and a $P$ value less than 0.05 considered statistically significant.

**Findings**

The frequency of human leukocyte antigen type B of the patients and the control subjects is presented in Table 1. Comparison between frequency of 30 types of B* antigens among patients and control group revealed only 7 (3.9%) patients with positive antigen-B40* type, while 18 (10%) subjects of control group had this antigen with a statistically significant difference (95%CI: 0.13–0.84, RR=1.15 and $P=0.02$). The frequency of the other 29 types of human leukocyte antigen type B had no statistically significant difference between the patients and control group (Table 2).

**Discussion**

While most of the early and late manifestations of KD are controlled with the use of intravenous immunoglobulin and acetylsalicylic acid, the etiology and possible predisposing factors are still undetermined. For development of KD, different etiological factors such as bacteria, viruses, mycoplasma, superantigens or even rug shampoo

**Table 1:** Prevalence of complete Kawasaki criteria in the patients

| Parameter                                             | Frequency |
|-------------------------------------------------------|-----------|
| Sex: male/female                                      | 1:4       |
| Lymph node                                            | 74%       |
| Conjunctivitis                                        | 94%       |
| Oral mucosa involvement                               | 90%       |
| Extremity involvement                                 | 74%       |
| Rash                                                  | 78%       |
| Cardiac involvement including coronary artery aneurism | 26%       |
| Coronary artery aneurism                              | 0.06%     |
have been accused. Rare presentation of KD in infants younger than 4 months may suggest protective role of maternal antibodies. Having higher incidence among those with Asian background, especially 5000-6000 new cases annually in Japan, and the greatest occurrence in twins, suggest the role of a genetic predisposition. This genetic background may predispose the patient to a super-antigen driven response with a selective expression of immune system and then the release of certain mediators of inflammation. The production of various inflammatory cytokines and elevated level of matrix metalloproteinases may mediate the vascular endothelial damages. Accordingly, the presence of certain human leukocyte antigens may increase susceptibility of the patients to KD or may protect them against the disease or its complications. Kato et al demonstrated increased presence of human leukocyte antigen BW22 in patients with KD in comparison with control group ($P<0.0005$). Krensky et al in their report from Boston hospital showed increased prevalence of BW51 antigen among Kawasaki patients ($P<0.002$). While Kaslow et al in their study reported that the combination of different human leukocyte antigens may have a role. Chang et al from China, Hong Kong, did not show any association between presence of human leukocyte antigens, including type B and KD.

Recently Maggioli et al reported the possible role of HLA class III region and susceptibility to KD.

Also with respect to development of coronary artery aneurysms, polymorphism of transmembrane region of human leukocyte antigen

### Table 2: Prevalence and percentage of human leukocyte antigen type B in the patients and controls

| Human Leukocyte Antigen -B | Patients (90) |          | Controls (89) |          |       |
|----------------------------|--------------|----------|---------------|----------|-------|
|                            | n            | %        | n             | %        | $P$   |
| *07                        | 9            | 5        | 4             | 2.2      | 0.2   |
| *08                        | 5            | 2.8      | 11            | 6.2      | 0.1   |
| *13                        | 6            | 3.3      | 2             | 1.1      | 0.3   |
| *14                        | 4            | 2.2      | 4             | 2.2      | 1.0   |
| *15                        | 3            | 1.7      | 3             | 1.7      | 1.0   |
| *18                        | 9            | 5        | 16            | 9        | 0.1   |
| *27                        | 2            | 1.1      | 4             | 2.2      | 0.4   |
| *35                        | 43           | 23.9     | 35            | 19.7     | 0.3   |
| *38                        | 4            | 2.2      | -             | 0        | 0.1   |
| *39                        | 3            | 1.7      | 1             | 0.6      | 0.6   |
| *40                        | 7            | 3.9      | 18            | 10.1     | 0.02  |
| *41                        | 7            | 3.9      | 1             | 0.6      | 0.07  |
| *42                        | -            | 0        | 2             | 1.1      | 0.2   |
| *44                        | 8            | 4.4      | 5             | 2.8      | 0.4   |
| *45                        | 1            | 0.6      | 3             | 1.7      | 0.4   |
| *49                        | 2            | 1.1      | 1             | 0.6      | 1.0   |
| *50                        | 7            | 3.9      | 2             | 1.1      | 0.2   |
| *51                        | 17           | 9.4      | 21            | 11.8     | 0.5   |
| *52                        | 8            | 4.4      | 7             | 3.9      | 1.0   |
| *53                        | 5            | 2.8      | 13            | 7.3      | 0.05  |
| *55                        | 6            | 3.3      | 7             | 3.9      | 0.8   |
| *56                        | 1            | 0.6      | 1             | 0.6      | 1.0   |
| *57                        | 4            | 2.2      | 3             | 1.7      | 1.0   |
| *58                        | 7            | 3.9      | 9             | 5.1      | 0.6   |
| *73                        | 3            | 1.7      | 1             | 0.6      | 0.3   |
| *81                        | -            | 0        | 4             | 2.2      | 0.06  |
| *37                        | 5            | 2.7      | 0             | 0        | 0.06  |
| *54                        | 2            | 1.1      | 0             | 0        | 0.5   |
| *36                        | 1            | 0.6      | 0             | 0        | 1.0   |
| *47                        | 1            | 0.6      | 0             | 0        | 1.0   |
genes and presence of certain alleles with increased susceptibility or a possible protective role for coronary complication in KD has been suggested\[23-25\]. So far studies from around the world have shown varied and inconsistent association between KD and single or summation of alleles (halotype)\[26\]. However, the possibility of a protective role for human leukocyte antigen in KD has not been reported as far as we are aware. For certain infectious diseases Cresio Alves et al have reviewed the association between human leukocyte antigen genes and increased susceptibility or protection against certain infections and their complication\[27\]. In their report, among Chilean patients with Chagas disease the incidence of cardiomyopathy was lower in those patients with antigen type B40*, however a report from Venezuela did not confirm this finding. Also for leprosy a protective role for type-A and type-B summations (halotype) has been suggested. The prevalence of antigen-B40* among our control group was about 10.1%. This antigen also has been the most frequent allele among Mexican, Korean, Chinese from Hainan province and Omani\[9,12,28,29\]. Considering the higher frequency of antigen type-B40* allele among our control subjects, may assume the possibility of a protective role for this antigen in our area. However, more studies are needed to find out the role of B-40* antigen and KD among mentioned ethnic groups with higher frequency of this antigen.

Limitation of study:

It is shown that many genetic characters can be transferred in halotype pattern, i.e. summation of multiple genes. The major limitation of our study is that we checked only the type-B antigens. Certainly further studies are needed to determine the possibility of a protective role for certain single gene or summation of genes for KD in our area.

Conclusion

The presence of higher frequency of allele type-B40* in the control group may represent a protective role for this antigen with resultant decreased susceptibility to KD in our area.

Presence or lack of certain human leukocyte antigens has been assumed for expression of certain genes which can induce production of mediators of inflammatory response in this disease.

Acknowledgment

This work was financially supported by Vice Chancellor for Research of Shiraz University of Medical Sciences (Grant Number 86-3719). We would like to thank the families of Kawasaki patients who kindly contributed to this work. We are also thankful to Mrs. Akbarzadeh for her secretarial assistance.

Authors' Contribution

Ajami Gh: conceived and designed the study and prepared the manuscript .

Kh. Aflaki: acquisition of Data, initially analyzed, interpreted the data and contributed in writing of initial manuscript .

S. Alyasin: performed the literature search and assisted in first draft of manuscript writing .

B. Gharesh-fard: acquisition of Data and HLA analysis.

M. Borzouee: contributed in the revision of the manuscript.

H. Amoozgar: modified the analysis and contributed in the revision of the manuscript.

All authors read and approved the final manuscript.

Conflict of Interest: None

References

1. Takahashi M, Newburger JW. Kawasaki disease. In: Allen HD, Driscoll DJ, Shaddy RE, el al., eds. Moss and Adams’ Heart Disease in Infants, Children, and Adolescents 7th ed. Philadelphia, Lippincott, Williams & Wilkins. 2008; Pp: 1242-56.

2. Satter E, Quinn CY, Kelly AP. Kawasaki disease. E. Medicine, Web Med, 2006; Sept 13.

3. Burns JC. Kawasaki disease update. Indian J Pediatrics 2009; 76(1):71-6.

4. Singh S. Kawasaki disease - An emerging disease in India but still not being commonly diagnosed. Abstract 045. Nineth Internaternal Kawasaki Disease Symposium, Taipei, Taiwan, 2008; Pp: 10-2.

5. Sadeghi IS, Amin R, Ajami GH. Kawasaki syndrome. The Iranian experience. East Mediterr Health J 2001; 7(1-2):16-25.

6. Kato S, Kimura M, Tsuji K, et al. HLA antigens in Kawasaki disease. Pediatrics 1978;61(2):252-5.

7. Krensky AM, Berenberg W, Shaney BA, et al. HLA antigens in mucocutaneous lymph node syndrome in New England. Pediatrics 1981;67(5):741-3.

8. Kaslow RA, Bailowitz A, Lin FYC, et al. Association of
9. Chang CC, Hawkins BR, Kao HK, et al. Human leukocyte antigen in southern Chinese with Kawasaki disease. *Eur J Pediatr* 1992;151(11):866.

10. Terai M, Kohno Y, Namba M, et al. Association of Col 11 cyte antigen wasakiase or cardiac involvement.

11. Barron KS, Silverman ED, Gonzales JC, et al. Major histocompatibility complex class II alleles in Kawasaki syndrome. Lack of consistent correlation with disease or cardiac involvement. *J Rheumatol* 1990;19(11):1790-3.

12. Whang DH, Park MH. Diversity of HLA -Bw alleles and haplotypes in Koreans. *Korean J Lab Med* 2003;23(1):45-51.

13. Oh JH, Han JW, Lee SJ, et al. Polymorphisms of HLA genes in Korean children with Kawasaki disease. *Pediatr Cardiol* 2008;29(2):402-8.

14. Onouchi Y. Molecular genetics of Kawasaki disease. *Pediatr Res* 2009;65(5 Pt 2):46R-54R.

15. Newburger JW, Takahashi M, Gerber MA, et al. Diagnosis, treatment and long-term management of Kawasaki disease: a statement for health professionals from the committee on rheumatic fever, endocarditis and Kawasaki disease, council on cardiovascular disease in the young, American Heart Association. *Circulation* 2004;110(17):2747-71.

16. Cereb N, Maye P, Lee S, et al. Locus specific amplification of human leukocyte antigen class I genes from genomic DNA: locus-specific sequences in the first and third introns of human leukocyte antigen HLA-A, -B and -C alleles. *Tissue Antigens* 1995;45(1):1-11.

17. Guttridge MG, Burr C, Klouda PT. Identification of human leukocyte antigen HLA-B35, B53, B18, B17 and B17 alleles by polymerase chain reaction using sequence specific primers (PCR-SSP). *Tissue Antigens* 1994;44(1):43-6.

18. Bidwell J. Advances in DNA-based HLA typing methods. *Immunol Today* 1994;15(7):303-7.

19. Narouei-Nejad M, Khosravi F, Danesh A, Nikbin B. Polymerase chain reaction-sequence specific primer (PCR-SSP) versus serology in typing of HUMAN LEUKOCYTE ANTIGEN -A, -B and -C in Iranian patients. *Arch Iranian Med* 2003;6(1):23-8.

20. Farjadian S, Kianimanesh N, Abbaszadegan A, et al. HL class I gene polymorphism in Iranian patients with Papillon-Lefevre syndrome. *Iran J Immunol* 2007;4(4):241-5.

21. Burns JC, Shimizu C, Shike H, et al. Family-based association analysis implicates IL-4 in susceptibility to Kawasaki disease. *Genes Immun* 2005;6(5):438-44.

22. Natividad MF, Torres-Villanueva CA, Saloma CP. Superantigen involvement and susceptibility factors in Kawasaki disease: profiles of TCR Vβ2+ T cells and HLA-DRB1, TNF-α and ITPKC genes among Filippo patients. *Int J Mol Epidemiol Genet* 2013;4(1):70-6.

23. Sheu JJ, Lin YJ, Chang J, et al. Association of Col 11 A2 polymorphism with susceptibility to Kawasaki disease and development of coronary artery lesions. *Int J Immunogenet* 2010;37(6):487-92.

24. Huang Y, Lee YJ, Chen MR, et al. Polymorphism of transmembrane region of MICA gene and Kawasaki disease. *Exp Clin Immunogenet* 2000;17(3):130-7.

25. Onouchi Y. Identification of susceptibility genes for Kawasaki disease. *Nihon Rinsho Meneki Gakkai Kaiho* 2010;11(2):73-80.

26. Hata A, Onouchi Y. Susceptibility genes for Kawasaki disease: Toward implementation of personalized medicine. *J human Genetics* 2009;54(2):67-73.

27. Alves C, Souza T, Meyer I, et al. Immunogenetics and infectious diseases; special reference to the major histocompatibility complex. *Braz J Infect Dis* 2009;10(2):122-31.

28. Center for Arab genomic study. The catalogue for transmission genetics in Arab CTGA database, major histocompatibility complex Class I , B. Oman, a division of Sheikh Hamdan award for medical sciences. 2007.

29. Vargas-Alurcon G; Hernandez Pacheco G; Zuniga J, et al. Distribution of HLA alleles in Mexican Amerindian populations. *Immunogenetics* 2003;54(11):754-60.