Antiulcer activity of ethanolic leaf extract of *Capparis zeylanica* against chemically induced ulcers

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**Abstract**

**Background:** Peptic ulcer is the term which refers to acid peptic injury of the digestive tract, and it results in mucosal break reaching the submucosa. Leaves of *Capparis zeylanica* are used as counterirritant, rubefacient, as a cataplasm in piles, boils and swellings. The objective of the present study was to evaluate the antiulcer activity of *C. zeylanica* ethanolic extract against chemically induced ulcers. The leaves were extracted with ethanol (50%) as solvent using hot perforation method. The extract was evaluated against acute and chronic ulcer models. Further, extract was evaluated for gastric autopsy of animals infected with *Helicobacter pylori* bacteria. The genes of rats were evaluated by gel electrophoresis method. Morphology of stomach was also studied after treatment with plant extract.

**Results:** Results exhibited that the area of ulcer was significantly reduced in both acute [naproxen-induced ulcer model (3.62 mm²), histamine-induced ulcer model (3.2 mm²) and ethanol-induced ulcer model (106.4 mm²)] and chronic [chronic naproxen-induced ulcer model (2.14 mm²), chronic histamine-induced ulcer model (0.16 mm²)]. The animals of naproxen-induced ulcer infected with *H. pylori* showed 91.48% reduction of ulcer area on 9th week after treatment with *C. zeylanica* extract (360 mg/kg). The rapid urease test and DNA observation revealed that no infection was present from 4th week after treatment with *C. zeylanica* extract (480 mg/kg). Morphological studies showed less conspicuous petechial marks and hemorrhages in stomach tissues after treatment with test drugs. Histopathological study revealed that *C. zeylanica* extract reduced stomach damages and eradicated *H. pylori* infections.

**Conclusion:** It can be concluded from the study that *C. zeylanica* possess antiulcer and anti-helicobacter activities.

**Keywords:** *Helicobacter pylori*, Naproxen, Ethanol, Histamine, Ulcer index, Ulcer area

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**Background**

Peptic ulcer is one of the most prevalent diseases around the world affecting four million people each year. Peptic ulcer is the term which refers to acid peptic injury of the digestive tract, and it results in mucosal break reaching the submucosa [1]. The disease involves an imbalance between offensive and defensive factors such as pepsin, acid and *Helicobacter pylori*; and bicarbonates, prostaglandins, mucin, nitric oxide and growth factors, respectively [2]. It has been also found that there is a chronic remitting course of peptic ulcer disease with imperfect correlation between symptoms and the presence of an ulcer [3]. *Helicobacter pylori* infection is a very common cause of primary peptic ulcers. It is associated with 70% of gastric ulcers and 95% of duodenal ulcers [4]. Other risk factors responsible to produce peptic ulcer disease are alcohol consumption, cocaine, tobacco and amphetamine use, chronic administration of nonsteroidal anti-inflammatory drugs (NSAIDs), fasting, Zollinger–Ellison syndrome and cancer treatment with angiogenesis inhibitors [5, 6].

Peptic ulcer treatment involves using a number of chemically produced drugs with aim to reduce the rate of stomach acid secretion, protection of the mucosa that...
line the stomach and upper portion of the small intestine or to eliminate *H. pylori* infestation [7]. The existing drugs cause several adverse effects; conversely, indigenous herbal drugs are devoid of side effects which might better treat peptic ulcers [8]. Medicinal plants possess numerous active phytoconstituents that are responsible for several biological activities [9–18]. The herbal drugs are less toxic than synthetic drugs; however, the toxicity evaluation is required to determine the safety profile of herbal drug [19–23]. In this concern, the drugs of natural origin can be used for the management of gastric ulcers as a better alternative to synthetic drugs [24].

*Capparis zeylanica* Linn is a rigid branched shrub, most widely distributed in India, Bangladesh, Sri Lanka and Malaysia. It is commonly known as Indian caper belonging to family Capparidaceae [25, 26]. Ethnopharmacologically, all parts of plant have been used as a folk medicine in Ayurvedic preparations [27]. Phytochemical investigation revealed that the plant possesses numerous active constituents that give different biological activities such as antioxidant, immunostimulant, antitumor, antidiabetic, antiscerosis and antibacterial activities [28]. It has been traditionally used as stomachic, antihydrotic, analgesic, sedative and in cholera, neuralgia, hemiplegia and rheumatism. Leaves of *C. zeylanica* are used as counterirritant, rubefacient, as a cataplasm in piles, boils and swellings [29]. Based on the tradition use of *C. zeylanica* in ulcer healing treatments, the protocol of the present study was designed to evaluate its effect against peptic ulcer. The different inducing agents were selected to determine the efficacy of plant extract against variety of toxic compounds in both acute and chronic conditions. In addition, the antimicrobial activity of the extract was observed in the prevention of ulcer infection.

Methods

Drugs, chemical and reagents

Naproxen sodium (98.0–102.0%), ranitidine (98.0%) and omeprazole (98.0%) were obtained as gift samples from Symed Labs, Hyderabad. Histamine hydrochloride was obtained from Sigma-Aldrich, St Louis, USA. Ethanol absolute was obtained from J. B. Chemicals Mumbai.

Extraction of *C. zeylanica* leaves

Fresh leaves of *C. zeylanica* were collected from the forests of Allahabad in the month of April 2019. It was identified and authenticated by Dr. Sunil Singh of the respective department. The samples of leaves were deposited in the herbarium of the institute with voucher no.1243. The leaves were dried, ground and treated with petroleum ether to remove fatty substances. The marc was extracted with ethanol (50%) as solvent using hot perforation method. Vacuum distillation was performed to reduce the volume of extract to 1/10, and remaining solvent was evaporated by boiling on a water bath. The final extracted product was dried in a lyophilized to get it in a powdered form. The yield of the product was 10%, w/w. The powdered extract was packed in an airtight container and used for further studies [30].

Experimental animals

Swiss albino mice (25–30 gm) and male Wistar rats (200–250 gm) were obtained from Central animal house of the United Institute of Pharmacy, Allahabad. The animals had free access to feed pellets (Amrut Laboratory Animal Feed, manufactured by Nav Maharashtra Chakan oil mills Ltd., Purchased from Pranav Agro Industries Ltd., Sangli, Maharashtra) and water ad libitum. All the experiments were approved by the Institutional Animal Ethics Committee (IAEC) of United Institute of Pharmacy, Allahabad (Approval No-UIP/IAEC/Dec/2016), constituted under Committee for the Purpose of Control and Supervision of Experiments on Animals (Reg. No-145/PO/E/11/CPCSEA Dated 29/7/2015).

Acute toxicity study

Both male and female swiss albino mice (18–22 g) were individually identified and allowed to acclimatize to the laboratory conditions for 7 days prior to study. The animals were administered different doses of test drugs, i.e., 50 mg/kg, 100 mg/kg, 300 mg/kg, 500 mg/kg, 1000 mg/kg and 2000 mg/kg at the starting of experiment, and observed for 14 days. The observation parameters were change in color of furs and eyes, change in behavior, pain and lethargy and change in feeding habits. Acute oral toxicity study was performed according to OECD guidelines 423 [31]. The LD 50 value was calculated using software (Environmental Protection Agency, USA).

Acute ulcer study

*Naproxen-induced ulcer model*

This experiment was performed according to the method of Satoh et al. [32]. Briefly, male Wistar rats of 200–230 g were selected and weighed and marked for identification. All animals were fasted for 24 h. Prophylactic treatment of *C. zeylanica* extract (30, 60 and 120 mg/kg p.o.) was given to three test groups, respectively. Distilled water (1 ml) and omeprazole (30 mg/kg p.o.) were administered to control and standard groups, respectively. Naproxen 30 mg/kg was administered p.o. after 1 h of *C. zeylanica* extract pretreatment. Animals were killed after 6 h of naproxen treatment. Stomach of all treated animals was isolated and opened along greater curvature to expose inner surface. Inner surface was washed thoroughly with normal saline and scanned for analysis. The ulcer index was calculated using the following formula:
Ulcer index (UI) = Total area of ulcer (mm²) /Total area of stomach (mm²)

Histamine-induced ulcer model
All animals were fasted for 24 h before treatment. *Caps paris zeylanica* extract (30, 60 and 120 mg/kg p.o.) was given to three test groups, respectively, as prophylactic dose. In this experimental model, ulcer was induced by oral administration of histamine (300 mg/kg) after 1 h of *C. zeylanica* extract pretreatment. Animals were protected from histamine toxicity by intraperitoneal injection of promethazine hydrochloride (5 mg) 15 min prior to and 15 min after induction of ulcer. Ranitidine (100 mg/kg p.o.) was used as a standard drug. Animals were killed after 4 h of histamine administration followed by dissecting stomach to determine ulcer index [33].

Ethanol-induced ulcer model
All animals were fasted for 24 h before treatment. *Caps paris zeylanica* extract (30, 60 and 120 mg/kg p.o.) was given to three test groups, respectively, as prophylactic dose. Ulcer was induced by administering ethanol (8 ml/kg) p.o. after 1 h of *C. zeylanica* extract pretreatment. Sucralfate (200 mg/kg p.o.) was administered to standard group. The animals were killed after 1 h of ulcer induction and killed by cervical dislocation. The stomach of all animals was dissected out and observed under microscope [34].

Chronic ulcer study
Chronic naproxen-induced ulcer model
In this animals model, healthy male Wistar rats, weighing 200–230 g, were selected. Naproxen (30 mg/kg, p.o.) was administered for consecutive 3 days. Therapeutic treatment was initiated with distilled water (1 ml), omeprazole (30 mg/kg p.o.) and *C. zeylanica* extract (120 mg/kg p.o.) to control, standard and test group, respectively, and treatment was continued daily for next 8 weeks. One animal from each group was killed every week for subsequent 8 weeks. Stomach was isolated and opened along with greater curvature to expose inner surface. Stomach was isolated and opened along greater curvature to expose inner surface. Inner surface was washed thoroughly with normal saline and scanned for analysis. The animals were divided into six groups each consisting 10 animals. Naproxen (30 mg/kg, p.o.) was administered for consecutive 3 days. Brucella broth solution of viable *H. pylori* (10⁸ CFU) was administered (1 ml/animal) after 24 h and continuing for 1 week. Therapeutic treatment was initiated after 24 h with distilled water (1 ml/animal), clarithromycin (30 mg/kg, p.o.) and *C. zeylanica* extract (120 O.D, 240 O.D., 360 O.D. and 480 O.D. mg/kg, p.o.) to control, standard and test groups, respectively, and treatment was continued daily for next 8 weeks. It is to be noted that *C. zeylanica* extract 120 mg/kg O.D. was continued, initially for 4 weeks, and the regimen was changed to *C. zeylanica* extract 120 mg/kg B.D. for the remaining 4 weeks. One animal from each group was killed every week for subsequent 8 weeks.

Gastric autopsy of animals infected with *H. pylori* bacteria
One animal was killed after a week, and *H. pylori* infection was confirmed by rapid urease test (RUT) and molecular biology techniques (DNA isolation and gene amplification by PCR). Stomach of the killed animal was isolated and opened along with greater curvature to expose inner surface. Inner surface was washed thoroughly with normal saline and scanned for analysis. Pylorus of isolated stomach was dissected, a portion of it was used for RUT, and remaining portion was used for PCR techniques [35, 36].

Analysis of genes by gel electrophoresis method
DNA was extracted and analyzed according to the method of Tiwari et al. [37]. The biopsies of stomach tissues were collected in Eppendorf tubes, which was previously filled with sterile phosphate-buffered saline (500 μl) and then vortexed for 2 min. Further, tubes were boiled in a water bath for 15 min, then cooled in ice and centrifuged for 1 min at 13,000×g. The supernatant collected after centrifugation was transferred to a tube, followed by amplification for 1 μl of this template. PCR amplification was carried out using DNA (10 ng), Taq polymerase (1 U) and oligonucleotide primers (10 pmol) for the selected genes (16 s rRNA and hrgA). The standard PCR buffer contains deoxynucleotide triphosphate (0.25 mmol) and MgCl₂ (2–3 mmol). The initially denaturation was performed for 5 min at 95 °C, followed by 35 cycles of denaturation at for 30 min at 94 °C. The annealing was
performed for 1 min at 52 °C, and extension was done for 1 min at 72 °C, followed by final extension at 72 °C for 7 min. The positive control was the DNA of H. pylori. The gel electrophoresis was performed for PCR products in agarose gel (20 g) with ethidium bromide (0.3%) containing 10% Tris–borate–EDTA buffer. UV transilluminator was used for gel visualization. The digital Bio-Rad system (Bio-Rad, India) was used for getting gel images.

**Morphology of stomach after treatment with plant extract**
The stomach of all animals was observed for morphological changes after treatment with plant extract. The observation was based on any glandular change, erythema and ulcer with petechial and conspicuous hemorrhages [38].

**Histopathology**
Stomach was removed, kept in 10% formalin for 12 h and then washed and processed using isopropyl alcohol, xylene and paraffin embedded for light microscopic study (Nikon E200). Paraffin-embedded tissue section (5 µm thickness) was prepared and stained after deparaffinization using hematoxylin and eosin stain to verify morphological assessment and presence of H. pylori. Mucosal inflammation, congestion of blood vessels and presence of H. pylori were recorded for each specimen. Histopathological changes were scored on a scale of none (−), mild (+), moderate (++) and severe (+++).

**Statistical analysis**
Data are represented as mean ± SEM (n = 6). Statistical analysis was performed using one-way analysis of variance (ANOVA) via Bonferroni’s test in a Graph Pad prism software version 8.0. The data are statistically different at *p* < 0.5, **p** < 0.1 and ***p*** < 0.01 in comparison with control group.

**Result**
**Acute toxicity study**
*Capparis zeylanica* extract was found safe up to 2000 mg/kg. No change in fur color, lacrimation, behavior or lethargy was seen in experimental animals during 14 days of observation.

**Acute study**
**Naproxen-induced ulcers**
Result showed that administration of naproxen produced ulcers in all treated animals and the mean ulcer area was 5.505 ± 0.1584 mm² and ulcer index (UI) 0.78 ± 0.03 was noted in control group that indicated the ulcerogenic effect of naproxen. Treatment with *C. zeylanica* extract showed significant (*p* < 0.01) reduction in the ulcer area (60 and 120 mg/kg) dose dependently. Pretreatment of animals with *C. zeylanica* extract (30 mg/kg) did not reduce the ulcer area. However, higher dose of *C. zeylanica* extract, i.e., 120 mg/kg, was effective in protecting against ulcerogenic action of naproxen in comparison with control group. Treatment with *C. zeylanica* extract (120 mg/kg) reduced area of ulcers (3.62 ± 0.15 mm²) and ulcer index (UI) (0.32 ± 0.01) significantly, which was near to standard drug omeprazole as shown in Table 1. Administration of omeprazole reduced area of ulcer (2.33 ± 0.10 mm²) and ulcer index (UI) (0.38 ± 0.03) in ulcerogenic condition.

**Histamine-induced ulcers**
Administration of histamine (300 mg/kg/i.p.) produced ulcers in all treated animals and the mean ulcer area was 10.66 ± 0.13 mm² and ulcer index (UI) 1.48 ± 0.01 was found in control group animals, indicating the ulcerogenic effect of histamine. Treatment with *C. zeylanica* extract showed significant reduction in the ulcer area (3.2 ± 0.15 mm²) and ulcer index (UI) (0.43 ± 0.02) dose dependently in comparison with control group as shown in Table 2. Treatment with standard drug ranitidine (100 mg/kg p.o.) reduced ulcer area (2.7 ± 0.14 mm²) and ulcer index (UI) (0.38 ± 0.03) in all the animals of the group.

**Ethanol-induced ulcers**
Administration of ethanol (8 ml/kg p.o.) produced ulcers in the treated animals and the mean ulcer area was 174.4 ± 5.814 mm² and ulcer index (UI) 22.36 ± 0.82 was found in control group. Results exhibited the ulcer area was significantly (*p* < 0.05 and *p* < 0.01) reduced in the animals pretreated with *C. zeylanica* extract (60 and 120 mg/kg), respectively. *Capparis zeylanica* extract (120 mg/kg)

| Parameters                | Control | Omeprazole 30 mg/kg | Extract 30 mg/kg | Extract 60 mg/kg | Extract 120 mg/kg |
|---------------------------|---------|---------------------|------------------|------------------|------------------|
| Area of ulcer (mm²)       | 5.50 ± 0.16 | 2.33 ± 0.11 | 5.27 ± 0.14 | 4.55 ± 0.14 | 3.62 ± 0.05 |
| Ulcer index (UI)          | 0.78 ± 0.03 | 0.32 ± 0.01 | 0.82 ± 0.02 | 0.64 ± 0.02 | 0.51 ± 0.01 |
kg p.o.) reduced area of ulcers (106.4 ± 3.956 mm²) and ulcer index (UI) (13.54 ± 0.48) near to normal in comparison with control as shown in Table 3. The results thus indicated that the higher dose of C. zeylanica extract, i.e., 120 mg/kg was effective in protecting against ulcerogenic effect of ethanol. However, the standard drug sucralfate (200 mg/kg p.o.) reduced ulcer area (68.12 ± 2.187 mm²) and ulcer index (UI) (8.89 ± 0.29), which was most significant among all groups. Thus, C. zeylanica extract was found effective in preventing ethanol-induced ulcers dose dependently.

**Chronic (therapeutic) study**

**Naproxen-induced ulcers**

Administration of naproxen (30 mg/kg p.o.) for three consecutive days produced ulcer in all treated animals, which was confirmed by killing one animal on 4th day from each group, and then, one animal was killed per week during the course of the experiment. Results indicated that C. zeylanica extract treatment healed naproxen-induced ulcers during the study period. However, a time-dependent reduction in the area of ulcer was observed on the completion of 8 weeks. Treatment resulted in minimum ulcer area (2.14 mm²) in comparison with control group (6.0 mm²) at corresponding time period. The results thus indicated that treatment of more than 8 weeks is required to exhibit optimum ulcer healing activity of C. zeylanica extract. Omeprazole (30 mg/kg/day p.o.) was effective in completely healing histamine-induced ulcers within 7 week treatment. Thus, C. zeylanica extract (120 mg/kg/day, p.o.) showed ulcer healing activity in histamine-induced ulcers (Table 5).

**Histamine-induced ulcers**

Chronic administration of histamine (300 mg/kg p.o.) resulted in ulcers formation during 8 week of the experiment. Results exhibited that C. zeylanica extract treatment healed histamine-induced ulcers during the study period. However, a time-dependent reduction in the area of ulcer was observed on the completion of 8 weeks. Treatment resulted in minimum ulcer area (0.16 mm²) in comparison with control group (12.92 mm²). The result thus indicated that treatment of more than 8 weeks is required to exhibit ulcer healing activity of C. zeylanica extract. The standard drug ranitidine (100 mg/kg/day, p.o.) was effective in completely healing histamine-induced ulcers within 7 week treatment. Thus, C. zeylanica extract (120 mg/kg/day, p.o.) showed ulcer healing activity in histamine-induced ulcers (Table 5).

**Naproxen-induced ulcers infected with H. Pylori**

Administration of naproxen (30 mg/kg, p.o.) for three consecutive days produced ulcers, and infection was induced through H. pylori inoculum (10⁸ C.F.U./ml) (1 ml/day for 7 days) in all animals. It resulted in no death of animals during the study period. The killed animal in the control group showed rather uniform area of ulceration up to 6 weeks, but after seventh week, there was gradual decline in ulcer area indicating natural healing process. Further, evidence of natural

| Table 2 | Effect of C. zeylanica extract on area of ulcer and ulcer index in histamine-induced acute ulcers |
|---------|-----------------------------------------------|
| Parameters | Control | Ranitidine 100 mg/kg | Extract 30 mg/kg | Extract 60 mg/kg | Extract 120 mg/kg |
| Area of ulcer (mm²) | 10.66 ± 0.08 | 2.71 ± 0.1 | 8.17 ± 0.29 | 5.19 ± 0.11 | 3.2 ± 0.11 |
| Ulcer index (UI) | 1.48 ± 0.01 | 0.38 ± 0.03 | 1.15 ± 0.06 | 0.72 ± 0.03 | 0.43 ± 0.02 |

| Table 3 | Effect of C. zeylanica extract on area of ulcer and ulcer index in ethanol-induced acute ulcers |
|---------|-----------------------------------------------|
| Parameters | Control | Sucralfate 200 mg/kg | Extract 30 mg/kg | Extract 60 mg/kg | Extract 120 mg/kg |
| Area of ulcer (mm²) | 174.4 ± 5.81 | 68.12 ± 2.19 | 152.7 ± 3.10 | 122.2 ± 2.58 | 106.4 ± 3.96 |
| Ulcer index (UI) | 22.36 ± 0.82 | 8.87 ± 0.29 | 19.36 ± 0.37 | 15.58 ± 0.31 | 13.54 ± 0.48 |

| Table 4 | Effect of omeprazole and C. zeylanica extract on area of ulcer in naproxen-induced therapeutic ulcers |
|---------|-----------------------------------------------|
| Week | Area of ulcer (mm²) | Control | Omeprazole (30 mg/kg) | Extract (120 mg/kg) |
| 1 | 8.61 | 8.69 | 8.22 |
| 2 | 8.16 | 5.31 | 8.02 |
| 3 | 7.54 | 3.22 | 7.01 |
| 4 | 6.99 | 0.98 | 6.87 |
| 5 | 6.87 | 0.1 | 5.77 |
| 6 | 6.58 | Nil | 3.94 |
| 7 | 6.31 | Nil | 3.2 |
| 8 | 6.00 | Nil | 2.14 |
healing was seen in the animal killed on the 9th week. Observance of 57.58% less area of ulcers was compared to that present in animal killed after 1 week. In case of clarithromycin 97.34%, C. zeylanica extract (120 mg/kg/day B.D. p.o.) 74% and C. zeylanica extract (240 and 360 mg/kg/day p.o.) 84.77% and 91.48% less ulcer area were seen, respectively, in comparison with respective first week ulcer area (Table 6). Clarithromycin (30 mg/kg p.o.) treatment showed a time-dependent reduction in the area of ulceration with maximum reduction in the area of ulceration after 9 weeks of treatment. The presence of this small area of ulcer (0.39 mm²) indicated that clarithromycin reduced the area of ulcers significantly. Similarly, C. zeylanica extract treatment showed a dose-dependent and time-dependent reduction in the ulcer area during the treatment period. Capparis zeylanica extract in the highest dose, i.e., 480 mg/kg p.o. O.D., reduced ulcer area up to 98.60% in comparison with control group.

Gastric autopsy of animal’s stomach after treatment with plant extract

The infection was determined by immersing gastric autopsy specimen, or tissue from the pylorus region into the rapid urease test (RUT) solution, which turned yellow to pink, indicated presence of H. pylori. The same protocol was followed for all the groups of animals treated with clarithromycin (30 mg/kg p.o.) and various doses of C. zeylanica extract. Clarithromycin (30 mg/kg p.o.) showed complete eradication of H. pylori after 3 weeks of treatment, and no infection was observed on 9th week. Capparis zeylanica extract at 120 mg/kg p.o. B.D. and 240 mg/kg p.o. O.D. was failed to eradicate H. pylori up to 9th week of treatment. In the group treated with C. zeylanica extract 360 mg/kg p.o. O.D., the H. pylori infection was found up to 6th week after which the infection was completely ameliorated (Table 7). In case of C. zeylanica extract 480 mg/kg p.o. O.D., the H. pylori infection was found up to 6th week after which the infection was completely ameliorated (Table 7). In case of C. zeylanica extract 480 mg/kg p.o. O.D., the H. pylori infection was found up to 6th week after which the infection was completely ameliorated (Table 7). In case of C. zeylanica extract 480 mg/kg p.o. O.D., the H. pylori infection was found up to 6th week after which the infection was completely ameliorated (Table 7).

Analysis of genes by gel electrophoresis

The infection was further confirmed by isolating DNA from gastric autopsy sample and amplification of two non-mutant genes (16s rRNA and hrgA) using polymerase chain reaction (Table 7).

### Table 5

**Effect of ranitidine and C. zeylanica extract on area of ulcer in histamine-induced acute ulcers**

| Week | Area of ulcer (mm²) |
|------|-------------------|
|      | Control | Ranitidine (100 mg/kg) | Extract (120 mg/kg) |
| 1    | 12.92   | 12.39   | 12.66   |
| 2    | 12.95   | 9.18    | 11.03   |
| 3    | 11.88   | 8.15    | 8.02    |
| 4    | 10.53   | 5.33    | 7.57    |
| 5    | 10.12   | 2.07    | 6.62    |
| 6    | 9.01    | 0.92    | 4.09    |
| 7    | 8.91    | Nil     | 2.06    |
| 8    | 8.79    | Nil     | 0.16    |

### Table 6

**Effect of C. zeylanica extract on area of ulcer naproxen-induced H. pylori-infected therapeutic ulcers**

| Week | Area of ulcer (mm²) |
|------|-------------------|
|      | Control | Clarithromycin 30 mg/kg | Extract (120 mg/kg) | Extract (240 mg/kg) | Extract (360 mg/kg) | Extract (480 mg/kg) |
| 1    | 9.97   | 10.65   | 11.03   | 10.31   | 10.92   | 10.11   |
| 2    | 9.08   | 8.67    | 10.58   | 9.63    | 9.01    | 8.23    |
| 3    | 8.89   | 7.11    | 8.78    | 8.51    | 8.87    | 6.92    |
| 4    | 9.03   | 5.93    | 8.83    | 8.00    | 6.52    | 6.66    |
| 5    | 8.66   | 5.47    | 6.98    | 6.67    | 5.23    | 5.23    |
| 6    | 8.97   | 5.34    | 5.29    | 6.00    | 4.01    | 3.92    |
| 7    | 7.62   | 3.31    | 4.01    | 4.33    | 3.98    | 1.66    |
| 8    | 5.82   | 1.15    | 3.23    | 2.98    | 2.85    | 0.51    |
| 9    | 4.23   | 0.39    | 2.87    | 1.57    | 0.93    | 0.5     |
Table 7 Effect of *C. zeylanica* extract on infection status in naproxen-induced *H. pylori*-infected therapeutic ulcers

| Week | RUT | DNA |
|------|-----|-----|
|      | Control | Clarithromycin 30 mg/kg | Extract (120 mg/kg) | Extract (240 mg/kg) | Extract (360 mg/kg) | Extract (480 mg/kg) | Control | Clarithromycin 30 mg/kg | Extract (120 mg/kg) | Extract (240 mg/kg) | Extract (360 mg/kg) | Extract (480 mg/kg) |
| 1    | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     |
| 2    | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     |
| 3    | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     | +     |
| 4    | +     | −     | +     | +     | +     | −     | +     | −     | +     | +     | +     | +     |
| 5    | +     | −     | +     | +     | +     | −     | +     | −     | +     | +     | +     | +     |
| 6    | +     | −     | +     | +     | +     | −     | +     | −     | +     | +     | +     | +     |
| 7    | +     | −     | +     | +     | −     | −     | +     | −     | +     | +     | −     | −     |
| 8    | +     | −     | +     | +     | −     | −     | +     | −     | +     | +     | −     | −     |
| 9    | +     | −     | +     | +     | −     | −     | +     | −     | +     | +     | −     | −     |

*RUT* Rapid urease test
Identification of hrgA gene by gel documentation

Figure 1 shows the UV transilluminated gel image with DNA ladder on both sides from 100 base pair onward (bp). The image shows the PCR amplified product using mastermix preparation and thermal cycle for hrgA gene. It corresponds to 594 bp when compared with ladder and confirmed the DNA isolated was that of *H. pylori*. The image was captured by gel documentation. The DNA was isolated and amplified gene throughout the period of 9 weeks in control group. However, it was interesting to find the absence of gene at 594 bp in the clarithromycin-treated group from the 4th week onward up to the 9th week. Similarly, higher doses of *C. zeylanica* extract (360 and 480 mg/kg p.o. O.D.) exhibited the absence of gene at 594 bp from 7th week (*C. zeylanica* extract 360 mg/kg p.o. O.D.) and 4th week onward (*C. zeylanica* extract 480 mg/kg p.o. O.D.). The result thus indicated that in both clarithromycin 30 mg/kg and *C. zeylanica* extract 480 mg/kg groups complete eradication of *H. pylori* occurred after 4 weeks of treatment.

Identification of 16srRNA gene by gel documentation

Figure 2 shows the UV transilluminated gel image with DNA ladder on both sides from 100 base pair onward (bp). The image shows the PCR amplified product using mastermix preparation and thermal cycle for 16 s rRNA gene. This amplified product corresponds to 534 bp when compared with ladder and confirmed the DNA isolated was that of *H. pylori*. The image was captured by gel documentation. The DNA was isolated and gene amplified throughout the period of 9 weeks in control group. However, it was interesting to find the absence of gene at 534 bp in the clarithromycin-treated group from the 4th week onward up to the 9th week. Likewise, higher doses of *C. zeylanica* extract (360 and 480 mg/kg p.o. O.D.) exhibited absence of gene at 594 bp from 7th week (*C. zeylanica* extract 360 mg/kg p.o. O.D.) and 4th week onward (*C. zeylanica* extract 480 mg/kg p.o. O.D.). The result thus indicated that in both clarithromycin 30 mg/kg p.o. O.D. and *C. zeylanica* extract 480 mg/kg p.o. O.D. groups complete eradication of *H. pylori* occurred after 4 weeks of treatment.
Effect of *C. zeylanica* extract on morphology of animal's stomach

Effect of *C. zeylanica* extract on morphology of different ulcer-inducing experimental models is shown in Fig. 3. In naproxen-induced ulcer model, the control group exhibited glandular region with petechial hemorrhages (black spots) indicated presence of ulcers (Fig. 3 NC). However, omeprazole showed less conspicuous petechial spots compared to that of control group (Fig. 3 NS). *Capparis zeylanica* extract showed less conspicuous petechial marks than control group, but erythema was present (Fig. 3 NT). In histamine-induced ulcer model, control showed glandular region with conspicuous hemorrhages along the margin (Fig. 3 HC); however, treatment with ranitidine showed less conspicuous hemorrhages were observed compared to toxic group, but erythema was present (Fig. 3 HS). On the other hand, *C. zeylanica* extract exhibited less conspicuous hemorrhages with erythema present (Fig. 3 HT). In ethanol-induced ulcer model, control group showed prominent erythema and conspicuous hemorrhages and complete disruption of the gastric mucosa, which was reduced to less conspicuous erosion of gastric mucosa in standard (sucralfate) and *C. zeylanica* extract-treated animals, but erythema was present in comparison with toxic control group.

![Fig. 3](image-url) Morphological changes due to administration of *C. zeylanica* extract in different experimental models of ulcers. N-C Naproxen-treated control group, N-S naproxen-treated standard group, N-T naproxen-treated test group, H-C histamine-treated control group, H-S histamine-treated standard group, H-T histamine-treated test group, E-C ethanol-treated control group, E-S ethanol-treated standard group, E-T ethanol-treated test group
Histopathological determination of *C. zeylanica* extract on gastric ulcers produced by different chemicals

Light microscopic examination of stomach of various groups of animals exhibited presence of inflammation, congestion and epithelial damage in the control group revealing successful induction of ulcers. Figure 4 shows

![Histopathological study of C. zeylanica extract-treated ulcers-induced animals models. N-C Naproxen-treated control group, N-S naproxen-treated standard group, N-T naproxen-treated test group, H-C histamine-treated control group, H-S histamine-treated standard group, H-T histamine-treated test group, E-C ethanol-treated control group, E-S ethanol-treated standard group, E-T ethanol-treated test group, NHy-C naproxen-induced *H. pylori* control group, NHy-S naproxen-induced *H. pylori* standard group, NHy-T naproxen-induced *H. pylori* test group.](image-url)
effect of *C. zeylanica* extract against naproxen-induced ulcers. The lower doses of *C. zeylanica* extract (30 and 60 mg/kg) exhibited presence of ulceration, inflammation and congestion in histology of animals. However, the highest dose of *C. zeylanica* extract was significantly restored stomach damages. The standard drug omeprazole was effective in healing ulcer at 30 mg/kg p.o. dose. Histamine also caused severe damage to animals stomach of all groups as shown in Fig. 4. Histopathological study revealed that *C. zeylanica* extract (120 mg/kg p.o.) reduced stomach damages and exhibited mild inflammation congestion and epithelial damage to animals stomach. Similar, results was obtained with standard drug ranitidine. Histopathology of ethanol-induced ulcer is shown in Fig. 4. Treatment with *C. zeylanica* extract (120 mg/kg p.o.) and standard drug sucralfate healed ulcer produced by ethanol. Mild inflammation congestion and epithelial damage were observed after treatment.

Infection with *H. pylori* leading successful induction of ulcers up to 9th week in all groups. In clarithromycin (30 mg/kg p.o.)-treated group, mild inflammation, congestion and presence of *H. pylori* were observed up to 5th, 6th and 3rd week, respectively. However, *C. zeylanica* extract (120 mg/kg/day p.o. B.D. and 240 mg/kg/day p.o. O.D.) group exhibited mild inflammation, congestion and presence of *H. pylori* were observed up to 3rd, 5th and 9th week, respectively, which was reduced to 3rd, 5th and 4th week, respectively, in the highest dose to *C. zeylanica* extract (480 mg/kg/day p.o. O.D.) as shown in Fig. 4.

**Discussion**

In the present investigation, the various groups of animals were treated prophylactically with the *C. zeylanica* extract to determine its ulcer protective potential. Naproxen was used as the ulcerogenic tool to produce acute gastric lesions, and it is due to non-selective inhibition of cyclooxygenase I and II, leading to reduced PGE2 synthesis and decreased mucus secretion [39]. Results showed that *C. zeylanica* extract has mild ulcer protective activity against naproxen-induced ulcer at a dose of 120 mg/kg p.o. Our results are in accordance with the observations of Halter et al. [40].

Histamine was used as the ulcerogenic tool to produce acute gastric lesions when administered at a dose of 300 mg/kg i.p. [32]. Its ulcerogenic potential was exhibited due to the hyper secretion of histamine from the parietal cells of the gastric mucosa, particularly in the glandular portion. A large area of redness devoid of any perforations was observed in the glandular potion of stomach which is characteristic of histamine-induced ulcers [41]. It is evident that the *C. zeylanica* extract has mild ulcerprotective effect against histamine-induced ulcers at doses of 60 and 120 mg/kg p.o., which elucidated that the *C. zeylanica* extract may act by blocking H2 receptors. Our findings are in accordance with those of Warzecha et al. [41]. Our results were in agreement with the study of Narra et al. evaluated antiulcer activity of *Cucumis Sativus* fruit extract. It was significantly reduced ulcer index in animals [42].

Ethanol produces acute experimental gastric lesions due to generation of free radicals primarily superoxide anions, hydroxyl radicals, lipid peroxidases localized inflammatory changes [43]. In the present investigation, gastric ulcer was produced using 8 ml/kg p.o. dose of ethanol as the ulcerogenic tool [34]. The various groups of animals were treated prophylactically with the drugs to determine the ulcer healing potential of the *C. zeylanica* extract. However, results showed that the *C. zeylanica* extract was unable to protect the gastric mucosal damage at all the three doses. *Capparis zeylanica* extract was very weak in amelioration of ethanol-induced gastric lesions when compared with sucralfate, which confirmed that the drug does not have any cement-like or ulcer painting activity [44]. The results of Srivasta et al. indicated that fruit extract of *Cucumis melo* Var. Monordica (Roxb.) was useful in the ethanol-induced ulcer model in rats. It was significantly reduced acid secretion and thus reduced ulcer index [45]. Table 8 shows the comparative study of antiulcer activity of our finding with previous publications [46–48].

In the present investigation, naproxen-induced and *H. pylori*-infected ulcer model was used to evaluate the ulcer healing and antimicrobial potential of the *C. zeylanica* extract [35]. When *H. pylori* is administered by oral gavage, this panmictic, spiral, gram negative organism resides in the disrupted mucosa for a long time exhibiting its ulcerogenic and oncopathogenic effects. Hence, this model is best suited to evaluate the antiulcer and anti-*H. pylori* effect of any drug [49]. The oncopathogenic effects of *H. pylori* are due to overexpression of COX2 mRNA leading to altered cellular

| S. no. | Ulcer                | Our findings (% efficacy) | Published works (% efficacy) | References          |
|-------|----------------------|---------------------------|-----------------------------|---------------------|
| 1     | Naproxen-induced ulcer | 35                        | 47                          | [46]                |
| 2     | Histamine-induced ulcer | 70                        | 80                          | [47]                |
| 3     | Ethanol-induced ulcer  | 60                        | 85                          | [48]                |
kinetics programmed cell death tumor angiogenesis and increased in the amount of nitrotyrosine leading to a precancerous which later culminates to produce metaplastic lesions [50]. The infection status was determined using rapid urease test and molecular confirmation by DNA extraction and amplification of 16sra RNA and hrg Agene [36]. The animals treated with C. zeylanica extract 480 mg/kg/day p.o. O.D. showed rapid healing and amelioration of infection within 4 weeks of treatment. This signifies the weak antimicrobial activity of C. zeylanica extract. The absence of any mortality showed that the drug was safe at high dose 480 mg/kg/day p.o. O.D. over a period of 9 weeks.

Conclusion
The present study suggests that the C. zeylanica extract possesses potent antulcer activity against chemicals-induced ulcer with significant antimicrobial activity. The extract showed effectiveness in both acute and chronic ulcers by reducing area of ulcers. Capparis zeylanica extract significantly restored the morphology of ulcerated stomach to normal one. Thus, ethanolic extract of C. zeylanica leaves may be considered as a potential therapeutic candidate in gastric ulcers infected with H. pylori. It can be developed into suitable formulation after clinical trials.

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All authors have read and approved the manuscript. AT contributed to manuscript preparation and proofreading. SKS was involved in laboratory work and performance of experimental task. AM contributed to statistics and analysis of data. All authors read and approved the final manuscript.

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Declarations
Ethics approval and consent to participate
Institutional Animal Ethical Committee approved the above protocol with approval no. CPCSEA/44/2006-07. Plant authentication: Plant leaves were authenticated by the Department of Pharmacognosy, United Institute of Pharmacy, Allahabad, and samples were deposited in the herbarium of the institute with voucher no.1243.

Consent for publication
Not applicable.

Competing interests
There is no competing of interest.

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