Prognostic impact of pretreatment T790M mutation on outcomes for patients with resected, EGFR-mutated, non-small cell lung cancer

Yoshiya Matsumoto1, Tomoya Kawaguchi1, Masaru Watanabe2, Shun-ichi Isa3, Masahiko Ando4, Akihiro Tamiya5, Akihito Kubo6, Chiyoe Kitagawa7, Naoki Yoshimoto5 and Yasuhiro Koh2,9*

Abstract
Background: Many previous studies have demonstrated that minor-frequency pretreatment T790M mutation (preT790M) could be detected by ultrasensitive methods in a considerable number of treatment-naïve, epidermal growth factor receptor (EGFR)-mutated, non-small cell lung cancer (NSCLC) cases. However, the impact of preT790M in resected cases on prognosis remains unclear.

Methods: We previously reported that preT790M could be detected in 298 (79.9%) of 373 surgically resected, EGFR-mutated NSCLC patients. Therefore, we investigated the impact of preT790M on recurrence-free survival (RFS) and overall survival (OS) in this cohort by multivariate analysis. All patients were enrolled from July 2012 to December 2013, with follow-up until November 30, 2017.

Results: The median follow-up time was 48.6 months. Using a cutoff value of the median preT790M allele frequency, the high-preT790M group (n = 151) had significantly shorter RFS (hazard ratio [HR] = 1.51, 95% confidence interval [CI]: 1.01–2.25, P = 0.045) and a tendency for a shorter OS (HR = 1.87, 95% CI: 0.99–3.55, P = 0.055) than the low-preT790M group (n = 222). On multivariate analysis, higher preT790M was independently associated with shorter RFS (high vs low, HR = 1.56, 95% CI: 1.03–2.36, P = 0.035), irrespective of advanced stage, older age, and male sex, and was also associated with shorter OS (high vs low, HR = 2.16, 95% CI: 1.11–4.20, P = 0.024) irrespective of advanced stage, older age, EGFR mutation subtype, and history of adjuvant chemotherapy.

Conclusions: Minor-frequency, especially high-abundance of, preT790M was an independent factor associated with a poor prognosis in patients with surgically resected, EGFR-mutated NSCLC.

Keywords: Non-small cell lung cancer, EGFR mutation, Pretreatment T790M, Resection, Recurrence-free survival

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Background
Over the past decade, the treatment paradigm for advanced non-small cell lung cancer (NSCLC) has evolved dramatically due to the detection of “druggable” gene alterations and the development of molecular-targeted therapies [1]. Mutations in the epidermal growth factor receptor (EGFR) gene, such as exon 19 deletions (19del) and Leu858Arg point mutations in exon

*Correspondence: ykoh@wakayama-med.ac.jp
9 Center for Biomedical Sciences, CIMS, Wakayama Medical University, 811-1 Kimiidera, Wakayama-shi, Wakayama 641-8509, Japan
Full list of author information is available at the end of the article
21 (L858R), are among the most common driver oncogenes in NSCLC. EGFR tyrosine kinase inhibitors (TKIs) are the recommended first-line treatment for advanced NSCLC patients with EGFR-activating mutations and improve survival significantly in those patients. However, most patients received EGFR-TKI therapies eventually develop resistance. The most common mechanism of acquired resistance to first-generation or second-generation EGFR-TKIs, such as gefitinib, erlotinib, and afatinib, is secondary EGFR T790M mutation, which is observed in 50–60% of acquired resistance cases [2, 3]. Osimertinib is a third-generation EGFR-TKI that selectively inhibits both EGFR-activating and T790M mutations. Based on the remarkable results of the AURA and FLAURA trials, osimertinib has been approved for the treatment of advanced NSCLC patients with EGFR mutations as first-line treatment and EGFR T790M-positive patients who had disease progression after prior-line EGFR-TKI treatment [4, 5].

The origin of EGFR-TKI resistance due to T790M mutation is not yet well understood. One of the hypotheses was that a T790M clone as a minor de novo clone preexists in treatment-naive tumors, and the pretreatment/de novo T790M (preT790M) clone is selected and enriched by exposure to EGFR-TKIs [6, 7]. Actually, earlier studies indicated the presence of minor-frequency T790M in pretreatment tumor samples in a small cohort, but detection rates varied from 2 to 79%, depending on the mutation detection methods and their sensitivities [7–13]. Therefore, we previously conducted a study aimed to accurately detect preT790M and clarify the prevalence of preT790M in a larger cohort that was an EGFR-mutant subset within the Japan Molecular Epidemiology (JME) study cohort, and we reported that preT790M was detected in 298 (79.9%) of 373 EGFR-mutated NSCLC patients using the ultra-sensitive droplet digital polymerase chain reaction (ddPCR), the analytical sensitivity of which was approximately 0.001% [14]. Recent studies using ddPCR also reported that the detection rates of minor-frequency preT790M were 7.9–70.6% [15–19].

In a recent report, the phase 3 randomized ADAURA trial, which investigated the efficacy and safety of osimertinib as adjuvant treatment compared with placebo after adjuvant chemotherapy in patients with completely resected stage IIB to IIIA EGFR-mutated NSCLC, showed significant improvement in disease-free survival (DFS) in the adjuvant osimertinib arm [20]. Based on the result, osimertinib as adjuvant therapy for NSCLC patients with EGFR mutations has been approved in the United States, China, and the European Union. However, there remain issues with adjuvant EGFR-TKI therapy, such as unclarity of overall survival (OS) benefit, the optimal administration period, cost, and adverse events. Therefore, development of biomarkers that identify high-risk populations for postoperative recurrence are needed to avoid unnecessary treatment. In advanced stage settings, previous researches demonstrated that preT790M was related to poor efficacy or shorter progression-free survival with the early-generation EGFR-TKI treatment [8–10, 12, 13, 16, 18, 19, 21, 22]. Moreover, several studies showed that a higher mutant-allele frequency (MAF) of preT790M might have greater impact on the efficacy of EGFR-TKIs than the presence of preT790M [12, 13, 18, 19]. These data suggest the negative predictive value of preT790M abundance for the efficacy of early-generation EGFR-TKIs in patients with advanced stage EGFR-mutated NSCLC. Therefore, preT790M could be a potential biomarker candidate to identify patients who may benefit from adjuvant osimertinib treatment. However, the clinical significance or prognostic implications of minor-frequency preT790M in patients with early-stage EGFR-mutated NSCLC who had undergone surgical resection have not yet been determined.

The JME study is a prospective, multicenter, molecular epidemiology study collecting samples from 876 surgically resected NSCLC cases and examining the somatic mutations to tackle associations between driver oncogenes and smoking and other environmental factors. Molecular profiling of that cohort as the primary endpoint of the JME study has been previously reported [23]. The secondary endpoints of the study were recurrence-free survival (RFS) and OS analyses (UMIN 000008177). Thus, the follow-up data and clinical outcomes were collected prospectively with the intent to investigate the impact of somatic mutations on RFS and OS for resected NSCLC. In this report, the follow-up data and clinical outcomes, focused on the EGFR mutant cohort of the JME study, are presented, and the impact of preT790M in patients with surgically resected EGFR-mutated NSCLC on RFS and OS is elucidated.

Methods

Patients

Eligible subjects from the JME study were pathologically diagnosed NSCLC patients with clinical stage I to IIIB disease (TNM classification version 7) who had undergone surgery for therapeutic purposes. Full details of the study design have been published previously [23]. All patients were enrolled from July 2012 to December 2013. Somatic mutations were analyzed by multiplex-targeted deep sequencing, and mutations in EGFR were also confirmed by PCR methods by an independent clinical laboratory (SRL, Tokyo, Japan). As a result, 373 samples with an EGFR-activating mutation based on the Cycleave PCR method were analyzed by ddPCR in the current JME study design have been published previously [23]. All patients were enrolled from July 2012 to December 2013. Somatic mutations were analyzed by multiplex-targeted deep sequencing, and mutations in EGFR were also confirmed by PCR methods by an independent clinical laboratory (SRL, Tokyo, Japan). As a result, 373 samples with an EGFR-activating mutation based on the Cycleave PCR method were analyzed by ddPCR in the current JME study.
substudy. This study was approved by the Institutional Review Board of the National Hospital Organization of Japan. All patients provided written, informed consent before surgery. The study was conducted in accordance with the Declaration of Helsinki.

Detection of \textit{EGFR} T790M mutation by droplet digital PCR
The ddPCR was carried out with a RainDrop Digital PCR system (Bio-Rad, Hercules, CA, USA), and the details of this procedure have been published previously [14]. Briefly, the duplex assay is based on the concurrent amplification of wildtype and specific mutant sequences in picoliter-sized compartmentalized liquid droplets and measurement of the terminal fluorescence signal from each droplet by flow cytometric techniques.

Statistical analysis
Clinical data, including age, sex, smoking history, pathological stage, history of adjuvant chemotherapy, \textit{EGFR} mutation subtype, and study findings including preT790M were used for the analysis of the current JME substudy. Fisher’s exact test was carried out for comparison of categorical data. The Kaplan–Meier method was used to estimate the survival curves for RFS and OS. Log-rank tests were used to compare the survival curves among the patients by preT790M status. RFS was defined as the period from the date of operation to the date of confirmed recurrence from any cause. Patients that were alive on the date of the last follow-up were censored at the time. All \(P\)-values were according to a two-sided hypothesis, and a \(P\)-value < 0.05 was considered significant. A Cox proportional hazards model was used to evaluate the impact of preT790M on RFS and OS. Statistical analysis was performed using IBM SPSS software (version 25).

Results

Patient characteristics
In the current JME substudy, 373 \textit{EGFR}-mutated NSCLC patients were obtained, and all samples could be analyzed for preT790M by ultra-sensitive ddPCR. As reported previously, preT790M was detected in 298 (79.9%) of 373 \textit{EGFR}-mutated NSCLC patients using ddPCR, of which the analytical sensitivity was approximately 0.001%. The preT790M-MAF ranged from 0.009 to 26.9% (median MAF 0.044%), and most tumors had preT790M-MAF < 0.1% [14].

All 373 patients’ clinical and prognostic data were collected prospectively. The data cutoff date for the JME study was November 30, 2017, and the median follow-up time in this study was 48.6 months. The characteristics of the patients are shown in Table 1. In the current analysis, the median age was 69 years (range 30–92 years), 182 patients (48.8%) were 70 years old or older, 276 (74.0%) were female, 361 (96.8%) were diagnosed with adenocarcinoma, and 93 (24.9%) had a smoking history. The number of patients by pathological stage was 219 patients (56.7%) in stage IA, 71 (19.0%) in stage IB, 43 (11.5%) in stage II, and 40 (10.7%) in stage III-IV. A total of 120 patients (32.2%) received the adjuvant chemotherapy, and of these, 2 patients with stage II and 5 patients with stage III-IV received gefitinib as adjuvant chemotherapy. Regarding \textit{EGFR} mutation status, 155 patients (41.6%) had 19del, 199 (53.4%) had L858R, and 19 (5.1%) had uncommon mutations.

When tumor samples were classified as having low or high levels of preT790M, using MAF of 0.045% as the cutoff based on the median MAF of 0.044%, that is, low-preT790M was defined as consisting of preT790M-MAF < 0.045% plus preT790M-negative, and high-preT790M was defined as preT790M-MAF ≥ 0.045%, the low group had 222 patients (59.5%), and the high group had 151 patients (40.5%). According to preT790M status, there were no significant differences in preT790M status by age, sex, tumor stage, \textit{EGFR} mutation status, or history of adjuvant chemotherapy. However, there were significantly more never smokers among patients with high-preT790M than among those with low-preT790M (Table 1).

Impact of pretreatment T790M on RFS

First, RFS was analyzed according to preT790M status. RFS tended to be shorter in patients with preT790M than in those without preT790M (hazard ratio [HR] = 1.76, 95% confidence interval [CI]: 0.98–3.15, \(P = 0.056\)) (Supplementary Fig. S1A), whereas RFS was significantly shorter in patients with high-preT790M than among those with low-preT790M (< 0.00001). Age (≥ 70 years), pathological stage (III-IV > IB-II > IA), and adjuvant chemotherapy (yes) were factors related to shorter RFS. On the other hand, sex, smoking history, and \textit{EGFR} mutation status did not affect RFS (Table 2). During this observational period, 96 RFS events were occurred.

Multivariate analysis demonstrated that higher preT790M-MAF (high vs low, HR = 1.56, 95% CI: 1.03–2.36, \(P = 0.035\)), male sex (male vs female, HR = 1.93, 95% CI: 1.06–3.52, \(P = 0.032\)), and advanced pathological stage (IB-II vs IA, HR = 6.30, 95% CI: 3.46–11.47, \(P < 0.00001\); III-IV vs IA, HR = 25.46, 95% CI: 13.11–49.44, \(P < 0.00001\)) were significantly associated with shorter RFS (Table 2). Older age tended to be related to shorter RFS (≥ 70 vs < 70 years, HR = 1.55, 95% CI: 0.99–2.43, \(P = 0.056\)). According to pathological stage, RFS was shorter in patients with high-preT790M than in those
with low-preT790M in stage IB-IV, but there was no significant difference in RFS regardless of preT790M status in stage IA (Fig. 2A-B and Supplementary Fig. S2).

**Impact of pretreatment T790M on OS**
Second, OS was analyzed according to preT790M status. There was no significant difference in OS between patients with and without preT790M (HR = 1.32, 95% CI: 0.53–3.27, \( P = 0.55 \)) (Supplementary Fig. S1B), but OS tended to be shorter in patients with high-preT790M than in those with low-preT790M (HR = 1.87, 95% CI: 0.99–3.55, \( P = 0.055 \)) (Fig. 1B). On univariate analysis, age (\( \geq 70 \) years), uncommon EGFR mutation, and pathological stage (III-IV > IB-II > IA) were factors related to shorter OS. On the other hand, sex, smoking history, and adjuvant chemotherapy did not affect OS (Table 3). During this observational period, 38 OS events were occurred.

Multivariate analysis demonstrated that higher preT790M-MAF (high vs low, HR = 2.16, 95% CI: 3.3–3.27, \( P = 0.0051 \)) was associated with shorter OS in patients with preT790M (Table 3).

### Table 1 Baseline characteristics of patients according to pretreatment T790M status

| Characteristics | Number of patients, n (%) | \( P \) |
|-----------------|--------------------------|-------|
| **All (n = 373)** |                          |       |
| Pretreatment T790M |                        |       |
| High (n = 151, 40.5%) |                  |       |
| Low (n = 222, 59.5%) |                   |       |
| **Age** |                         | 0.67  |
| Median (range) | 69 (30–92) | 69 (37–88) | 69 (30–92) |
| < 70 | 191 (51.2) | 75 (49.7) | 116 (52.3) |
| \( \geq 70 \) | 182 (48.8) | 76 (50.3) | 106 (47.7) |
| **Gender** |                         | 0.47  |
| Male | 97 (26.0) | 36 (23.8) | 61 (27.5) |
| Female | 276 (74.0) | 115 (76.2) | 161 (72.5) |
| **Smoking** |                       | 0.0050 |
| Never smoker | 280 (75.1) | 125 (82.8) | 155 (69.8) |
| Smoker | 93 (24.9) | 26 (17.2) | 67 (30.2) |
| **Histology** |                       | 0.24  |
| Adenocarcinoma | 361 (96.8) | 144 (95.4) | 217 (97.7) |
| Other | 12 (3.2) | 7 (4.6) | 5 (2.3) |
| **Pathological Stage (7th)** |                   | 0.42  |
| IA | 219 (58.7) | 84 (55.6) | 135 (60.8) |
| IB-II | 114 (30.6) | 52 (34.4) | 62 (27.9) |
| III-IV | 40 (10.7) | 15 (9.9) | 25 (11.3) |
| **EGFR Mt Status** |                     | 0.92  |
| Exon21 L858R | 199 (53.4) | 82 (54.3) | 117 (52.7) |
| Exon19 deletion | 155 (41.6) | 62 (41.1) | 93 (41.9) |
| Uncommon | 19 (5.1) | 7 (4.6) | 12 (5.4) |
| **Adjuvant Chemotherapy** |                   | 0.37  |
| No | 253 (67.8) | 98 (64.9) | 155 (70.2) |
| Yes | 120 (32.2) | 53 (35.1) | 67 |
| UFT | 73 | 32 | 41 |
| Platinum doublet | 34 | 18 | 16 |
| EGFR-TKI | 7 | 1 | 6 |
| Other | 6 | 2 | 4 |
| **TP53 Mt Status** |                     | 0.077 |
| Wild-type | 304 (81.5) | 130 (86.1) | 174 |
| Mutant | 69 (18.5) | 21 (13.9) | 48 (21.6) |
| Co-existing except TP53 Mt |                   | 0.52  |
| No | 329 (88.2) | 131 (86.8) | 198 (90.8) |
| Yes | 44 (11.8) | 20 (13.2) | 24 |

EGFR Epidermal growth factor receptor, Mt Mutation, UFT Tegafur-uracil, TKI Tyrosine kinase inhibitor, TP53 Tumor protein P53


1.11–4.20, \( P = 0.024 \)), uncommon \( EGFR \) mutation (vs L858R, \( HR = 3.33, 95\% \) CI: 1.13–9.83, \( P = 0.029 \)), and advanced pathological stage (IB-II vs IA, \( HR = 4.30, 95\% \) CI: 1.77–10.40, \( P = 0.001 \); III-IV vs IA, \( HR = 12.10, 95\% \) CI: 4.83–30.29, \( P < 0.00001 \)) were significantly associated with shorter OS (Table 3). Older age (\( \geq 70 \) vs <70 years, \( HR = 1.79, 95\% \) CI: 0.87–3.69, \( P = 0.12 \)) and 19del (vs L858R, \( HR = 1.80, 95\% \) CI: 0.88–3.67, \( P = 0.11 \)) tended to have shorter OS, and adjuvant chemotherapy tended to be related to longer OS (HR = 0.47, 95% CI: 0.22–1.01, \( P = 0.053 \)). According to pathological stage, OS was shorter in patients with high-preT790M than in those with low-preT790M in stage IB-IV, but there was no significant difference in OS regardless of preT790M status in stage IA (Fig. 2C-D and Supplementary Fig. S3).

During this observational period, 92 postoperative recurrence events were recorded. According to the preT790M status, 44 (29.1%) and 48 (21.6%) recurrences...
were observed in the high- and low-preT790M groups, respectively. Post-recurrence treatments are shown in Table 4.

Discussion

Before the current study, many studies reported the predictive impact of minor-frequency preT790M on the efficacy of initial EGFR-TKI treatment or on the prognosis, mainly in advanced stage settings. However, there were few reports regarding the clinical significance of preT790M in patients with resected NSCLC. In this prospective exploratory analysis, minor-frequency preT790M in the resected EGFR-mutated NSCLC samples was shown to potentially affect RFS and OS.

In previous analyses of prognostic factors for resected EGFR-mutated NSCLC, older age, male sex, advanced stage, and smoker were shown to be independent factors associated with a poor prognosis (RFS or OS) in many studies, although the covariates evaluated differed depending on the studies [24–34]. In the current study, multivariate analysis, which considered these previous reports, demonstrated that male sex and advanced pathological stage were correlated with shorter RFS, and older age tended to be associated with shorter RFS. Pathological stage was also correlated with shorter OS, and in fact, pathological stage was the most important factor affecting RFS and OS. There were many reports showing that EGFR 19del was associated with worse RFS or OS than L858R [26, 28, 33], but some studies reported that 19del had better RFS or OS than L858R [30, 35], and others reported that EGFR mutation subtype has no prognostic impact [25]. Therefore, whether the EGFR mutation subtype has an impact on prognosis remained controversial. On multivariate analysis in the current study, there was no significant difference in RFS by EGFR mutation subtype, but 19del tended to have

Fig. 2 Kaplan-Meier curves for recurrence-free survival (A, B) and overall survival (C, D) according to pretreatment T790M status (high vs low). (A) and (C) for the population with pathological stage IA NSCLC. (B) and (D) for the population with pathological stage IB-IV NSCLC. Plus signs denote censoring.
worse OS, and uncommon mutations had significantly worse OS than L858R.

Let us consider the impact of preT790M as a prognostic factor for survival. In the current study using ultra-sensitive ddPCR, the overall detection rate of preT790M was 79.9%, and the T790M-MAF ranged from 0.009 to 26.9% (median MAF 0.044%). Several previous studies indicated that higher MAF of preT790M might have a greater impact on the efficacy of EGFR-TKIs than the presence of preT790M [12, 13, 18, 19]. Therefore, when tumor samples were classified into two groups based on the abundance of T790M-MAF, multivariate analysis demonstrated that high-preT790M was the independent factor related to a poor prognosis (RFS), irrespective of patient background, including pathological stage, age, and sex. A previous retrospective study by Tatematsu et al., which analyzed the incidence of minor-frequency preT790M using competitive allele-specific PCR in 153 surgically resected EGFR-mutated lung adenocarcinoma tissues, the incidence of preT790M was 29.4%, and T790M-MAF ranged from 0.13 to 2.65% (median MAF 0.20%) [36]. However, in their study, no significant

### Table 3 Prognostic factors for overall survival (OS): univariate and multivariate analyses

| Factor                        | univariate         | multivariate       |
|-------------------------------|--------------------|--------------------|
|                               | HR (95% CI) | P     | HR (95% CI) | P     |
| Pretreatment T790M (ref = Low) |         |       |         |       |
| High                          | 1.87 (0.99–3.55) | .055 | 2.16 (1.11–4.20) | .024 |
| Age (ref = < 70)               |         |       |         |       |
| ≥ 70                          | 2.13 (1.09–4.17) | .028 | 1.79 (0.87–3.69) | .12  |
| Gender (ref = Female)         |         |       |         |       |
| Male                          | 1.12 (0.56–2.26) | .75  | 1.17 (0.40–3.44) | .77  |
| Smoking (ref = Never smoker)  |         |       |         |       |
| Smoker                        | 1.34 (0.68–2.66) | .40  | 1.25 (0.43–3.61) | .69  |
| EGFR Mt (ref = Exon21 L858R)  |         |       |         |       |
| Exon19 deletion               | 1.56 (0.79–3.10) | .20  | 1.80 (0.88–3.67) | .11  |
| Uncommon                      | 3.54 (1.28–9.77) | .015 | 3.33 (1.13–9.83) | .029 |
| Pathological Stage (7th) (ref = IA) |         |       |         |       |
| IB-II                         | 3.51 (1.55–7.95) | <.01 | 4.30 (1.77–10.40) | .001 |
| III-IV                        | 9.68 (4.14–22.67) | <.0001 | 12.10 (4.83–30.29) | <.0001 |
| Adjuvant Chemotherapy (ref = No) |         |       |         |       |
| Yes                           | 1.07 (0.55–2.09) | .85  | 0.47 (0.22–1.01) | .053 |

### Table 4 Postoperative recurrence events and post-recurrence treatment

| Number of patients, n (%) | Pretreatment T790M | P     |
|---------------------------|--------------------|-------|
| All (n = 373)             | High (n = 151)     | Low (n = 222) |
| Recurrence events         | 92 (24.7)          | 48 (21.6) | 0.11 |
| Locoregional              | 45 (12.1)          | 24 (10.8) | 0.42 |
| Distant                   | 58 (15.5)          | 30 (13.5) | 0.19 |
| Post-recurrence treatment |                    |       |
| Local therapy             |                    |       |
| Radiation therapy         | 24                 | 13    | 11  |
| Surgery                   | 4                  | 1     | 3   |
| Systemic therapy          |                    |       |
| Chemotherapy              | 9                  | 5     | 4   |
| EGFR-TKI                  | 53                 | 25    | 28  |
| Best Supportive Care      | 5                  | 1     | 4   |
impact of preT790M on RFS was shown. A previous analysis demonstrated that the impact of T790M shifts according to the cutoff level of T790M-MAF [13]. Therefore, differences in analytical sensitivity, the detection rate of preT790M, sample size, and population grouping might result in the differences in the impact of preT790M between their study and the present one. Furthermore, disease stage was the most crucial factor affecting prognosis [24–29, 31–34]. Therefore, it might be important to consider pathological stage in the analysis of clinical significance of minor-frequency preT790M, although the association of preT790M status with patient characteristics was not reported in their study. In fact, the present study showed that higher-preT790M affected RFS in stage IB or more advanced disease, but it seemed unlikely in stage IA. On the other hand, Gao et al. analyzed clinical outcomes of coexisting T790M in a surgically resected, EGFR-mutated NSCLC cohort using the Amplification Refractory Mutation System, of which the analytical sensitivity was known to be generally 1% [37]. Their study also demonstrated that RFS of patients with coexisting EGFR T790M was significantly shorter than that of those without T790M mutations, and according to the stage, this tendency was observed not only in stage IB-III A, but also in stage IA. Greater T790M-MAF, which could be detected by routine clinical genotyping tests, might affect RFS even in stage IA, although ultra-low-level preT790M was thought not to have an impact on RFS in stage IA.

The above-mentioned study by Tatematsu et al. also did not show a significant effect of preT790M on OS in surgically resected EGFR-mutated NSCLC [36]. However, in the current study, multivariate analysis demonstrated that preT790M was the independent factor related to a poor prognosis in patients with resected EGFR-mutated NSCLC, irrespective of patient background including pathological stage, age, EGFR mutation subtype, and history of adjuvant chemotherapy. According to pathological stage, high-preT790M showed no prognostic impact in stage I A, but OS in high-preT790M tended to be shorter in stage IB or more advanced settings, as well as in the RFS analysis. To the best of our knowledge, the present research is the first to show that preT790M has a significant impact on OS in resected EGFR-mutated NSCLC in a larger cohort.

In metastatic stage settings, the appearance of T790M mutation after resistance to initial EGFR-TKI treatment (acquired T790M) has been reported to be associated with a good prognosis in the patients with EGFR-mutated NSCLC [22, 38, 39]. A basic research study found that the acquisition of T790M was associated with a slowing of tumor growth, which might underlie the good prognosis of EGFR-mutated NSCLC with acquired T790M [40]. On the other hand, positivity or high-abundance of preT790M has been demonstrated to be associated with poor efficacy of initial EGFR-TKI treatment or a poor prognosis [8–10, 12, 13, 16, 18, 19, 21, 22]. In the same way, the present study demonstrated that high-abundance of preT790M was correlated with poor RFS and OS in patients with early-stage NSCLC who had undergone surgical resection. These results suggest that clinical features are likely to be different between preT790M and acquired T790M [22]. However, the reason why patients with EGFR-mutated NSCLC harboring preT790M appear to have a poor prognosis has not yet been elucidated, even though the tumor harbors a low-level amount of T790M clones and has undergone surgical resection. The previous research showed that β-Catenin, which is involved in the pathogenesis and progression of malignant tumors, especially cancer stem cells, was upregulated and activated in EGFR-sensitizing mutant cells, and more in EGFR-mutant cells bearing T790M than in wild-type EGFR cells, and suggested that a cooperative association between β-catenin and EGFR-sensitizing mutations or with T790M plays a significant role in lung tumorigenesis [41]. Several studies also demonstrated that inhibition of β-catenin suppressed EGFR-activating and T790M mutated lung tumor growth or increased the anticancer effects of EGFR-TKIs [41, 42]. These data suggested that β-Catenin mediated stem-cell like properties of cancer cells harboring dual EGFR-activating mutation and T790M mutation may contribute to the activation of cell growth, proliferation, and the progression of disease; therefore, such research findings could explain the worse RFS in patients with NSCLC harboring concomitant preT790M. Further investigations and validation are needed.

Postoperative adjuvant chemotherapy is recommended for patients with completely resected stage II-III A and a subset of stage I NSCLC according to the results from large, randomized trials and meta-analyses that have demonstrated a significant OS benefit [43, 44]. However, whether driver mutation-positive patients with resected stage NSCLC also benefit from adjuvant chemotherapy had not been accurately clarified. In particular, for patients with resected NSCLC harboring EGFR mutations, given the role of EGFR-TKIs in advanced EGFR-mutant NSCLC, many clinical trials have been conducted to investigate the efficacy of EGFR-TKIs in the adjuvant setting [45–50]. Most trials demonstrated that adjuvant treatment using first-generation EGFR-TKIs can decrease the risk of recurrence and prolong DFS compared to placebo or chemotherapy, but these DFS advantages did not always translate to OS [51, 52]. A meta-analysis that evaluated the role of EGFR-TKIs as an adjuvant therapy for patients with completely resected EGFR-mutated
NSCLC demonstrated that, compared to mono chemotherapy, early-generation EGFR-TKI monotherapy had a superior DFS benefit, but did not show a significant OS benefit, whereas treatment with EGFR-TKIs plus chemotherapy was associated with significantly longer DFS and OS compared to mono chemotherapy [53]. Therefore, these data suggested that it was necessary for the prolongation of DFS and OS in patients with resected EGFR-mutated NSCLC not only to add EGFR-TKIs as adjuvant treatment, but also to perform standard adjuvant chemotherapy as much as possible. On multivariate analysis in the current study, it was observed that OS in the patients who received adjuvant chemotherapy tended to be better than in those who did not receive adjuvant chemotherapy.

The ADAURA trial demonstrated significant improvement of DFS in the adjuvant osimertinib arm (HR for disease recurrence or death, 0.17 [99% CI, 0.11–0.26] in patients with stage II to IIIA disease, and 0.20 [99% CI, 0.14–0.30] in patients with stage IB to IIIA disease) [20]. In that trial, administration of standard postoperative adjuvant chemotherapy was allowed, but not mandatory, although the DFS benefit from osimertinib was documented regardless of whether patients underwent adjuvant chemotherapy. Based on the present study findings, high-preT790M was an independent factor related to a poor prognosis for both RFS and OS in stage IB or more advanced stages. In advanced settings, the AZENT study (NCT02841579) showed promising results for first-line osimertinib for patients with EGFR-mutated NSCLC with a coexisting low allelic fraction of T790M, in which the objective response rate was 77.3% and the median PFS was 23.1 months [54]. Moreover, the WJOG13119I study demonstrated that the time to treatment failure (TTF) of the micro-T790M-positive group treated by the first-generation EGFR-TKIs was shorter than the negative group, although the TTF of the micro-T790M-positive group treated by osimertinib was longer than that of the negative group [55]. Based on these reports, osimertinib may be expected to be effective for EGFR-mutated NSCLC with a low-frequency preT790M. Although the basic correlation between poor prognosis and minor-frequency preT790M in resected EGFR-mutated NSCLC has not yet been elucidated, osimertinib treatment may have had an impact on DFS for the population harboring potential preT790M in the ADAURA trial. Therefore, the addition of adjuvant osimertinib to standard adjuvant chemotherapy might also be expected to have a greater impact on improving OS. The NeoAD-AURA (NCT04351555) trial, investigating the efficacy and safety of neoadjuvant osimertinib in patients with EGFR-mutated resectable NSCLC, is ongoing. If translational research assessing minor-frequency T790M before osimertinib and after surgery could be conducted, the clinical significance of treatment of preT790M might be elucidated.

The limitations of the current study include the relatively short observation period and the low number of recurrence and death events, which results in a lack of statistical power, although the current study was the largest prospective trial, and the prognostic information was collected exactly. The number of patients in Stage I, especially stage IA, who have good prognosis because of the progress of diagnostic techniques and developments in improved surgical techniques was considerably large, which resulted in decreased incidences of recurrence and death. Therefore, further validation in a larger cohort might be needed for analysis of the prognosis for stage IB or more advanced settings. In stage IA cases, tumors mainly composed of ground glass opacity (GGO) components are often identified. The proportion of GGO tumors may have the potential to be related to why no difference between the high-preT790M group and the low-preT790M group was observed in RFS and OS in stage IA patients. However, information about the proportion of GGO tumor included in stage IA cases in the current study is not available.

There were several other limitations in the present study. In several previous analyses of prognostic factors for resected NSCLC, various covariates regarding postoperative pathological findings, such as lymphatic infiltration, vascular infiltration, and pleural invasion, were shown to be associated with a poor RFS or OS [25, 26]. However, the information regarding them was not collected in the JME study, and it remains unclear whether the factors of postoperative pathological findings affected prognosis in the present study. There was no provision regarding the CT scan interval in the JME study; it was left to the discretion of the attending physician, and, therefore, it may have affected RFS. Although the tumor specimens were dissected by pathologists, and those with as high a tumor content and as low a necrotic component as possible were chosen for analysis, the effect of normal cells could not be eliminated completely, resulting in a possible effect on accurate calculation of the preT790M-MAF.

Conclusions
The current prospective, multicenter, observational study showed that a higher mutant-allele frequency of pretreatment T790M in patients with surgically resected EGFR-mutated NSCLC was associated with poorer RFS, independent of male sex, advanced pathological stage, and older age, and was also associated with worse OS, independent of EGFR mutation genotype, advanced pathological stage, older age, and no adjuvant chemotherapy.
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Authors’ contributions

TK, MW, and YK provided the concept, TK acquired fund, and TK, SI, MA, AT, AK, CK, NY, and YK acquired study participants, samples or data. MW and YK designed and performed the experiments. YM analyzed the data. YM drafted the manuscript, and YK proofread and revised the manuscript. All authors read and approved the final manuscript.

Availability of data and materials

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

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Additional file 1.

References

1. Howlader N, Forjaz G, Mooradian MJ, Meza R, Kong CY, Cronin KA, et al. The effect of advances in lung-Cancer treatment on population mortality. N Engl J Med. 2020;383(7):640–9.
2. Yu HA, Arcila ME, Rekhtman N, Sima CS, Zakowski MF, Pao W, et al. Analysis of tumor specimens at the time of acquired resistance to EGFR-TKI therapy in 155 patients with EGFR-mutant lung cancers. Clin Cancer Res. 2013;19(8):2240–7.
3. Campo M, Gerber D, Gainor JF, Heist RS, Temel JS, Shaw AT, et al. Acquired resistance to first-line Afatinib and the challenges of prearranged progression biopsies. J Thorac Oncol. 2016;11(1):2022–6.
4. Mok TS, Wu YL, Ahn MJ, Garassino MC, Kim HR, Ramalingam SS, et al. Osimertinib or platinum-Pemetrexed in EGFR T790M-positive lung Cancer. N Engl J Med. 2017;376(7):629–40.
5. Soria JC, Ohe Y, Vansteenkiste J, Reungwetwattana T, Chesskuklyong B, Lee KH, et al. Osimertinib in untreated EGFR-mutated advanced non-small-cell lung Cancer. N Engl J Med. 2018;378(2):113–25.
6. Godin-Heymann N, Bryant J, Rivera MN, Ullrich L, Bell DW, Riese DJ, 2nd, et al. Oncogenic activity of epidermal growth factor receptor kinase mutant alleles is enhanced by the T790M drug resistance mutation. Cancer Research 2007;67(15):7319–26.
7. Inukai M, Toyooka S, Ito S, Asano H, Ichihara S, Sod J, et al. Presence of epidermal growth factor receptor gene T790M mutation as a minor clone in non-small cell lung cancer. Cancer Res. 2006;66(16):7854–8.
8. Mahesswaran S, Sequist LV, Nagrath S, Ullrich L, Brannigan B, Collura CV, et al. Detection of mutations in EGFR in circulating lung cancer cells. N Engl J Med. 2008;359(4):366–77.
9. Rosell R, Molina MA, Costa C, Simonetti S, Gimenez-Capitan A, Bertran-Alamillo J, et al. Pretreatment EGFR T790M mutation and BRCa1 mRNA expression in erlotinib-treated advanced non-small-cell lung cancer patients with EGFR mutations. Clin Cancer Res. 2011;17(5):1160–8.
10. Su KY, Chen HY, Li KC, Kuo ML, Yang JC, Chan WK, et al. Pretreatment epidermal growth factor receptor (EGFR) T790M mutation predicts shorter EGFR tyrosine kinase inhibitor response duration in patients with non-small-cell lung cancer. J Clin Oncol. 2012;30(4):433–40.
11. Fujita Y, Suda K, Kimura H, Matsumoto K, Arao T, Naga T, et al. Highly sensitive detection of EGFR T790M mutation using colony hybridization predicts favorable prognosis of patients with lung cancer harboring activating EGFR mutation. J Thorac Oncol. 2012;7(11):1640–4.
12. Costa C, Molina MA, Drozdowski A, Gimenez-Capitan A, Bertran-Alamillo J, Karachallou N, et al. The impact of EGFR T790M mutations and BIM mRNA expression on outcome in patients with EGFR-mutant NSCLC treated with erlotinib or chemotherapy in the randomized phase III EURTAC trial. Clin Cancer Res. 2014;20(7):2001–10.
13. Lee Y, Lee GK, Lee YS, Zhang W, Hwang JA, Nam BH, et al. Clinical outcome according to the level of preexisting epidermal growth factor receptor T790M mutation in patients with lung cancer harboring sensitive epidermal growth factor receptor mutations. Cancer. 2014;120(14):2090–8.
14. Watanabe M, Kawaguchi T, Isa S, Ando M, Tamiya A, Kudo A, et al. Ultra-sensitive detection of the pretreatment EGFR T790M mutation in non-small cell lung Cancer patients with an EGFR-activating mutation using droplet digital PCR. Clin Cancer Res. 2015;21(15):3552–60.
15. Vendrell JA, Mazieres J, Senal R, Rouquette J, Quantin X, Pujol JL, et al. Ultra-sensitive (EGFR T790M) detection as an independent prognostic marker for lung Cancer patients harboring EGFR (delE19) mutations and treated with first-generation TKIs. Clin Cancer Res. 2019;25(14):4280–9.
16. Dong Y, Zhou Z, Wang J, Ma L, Liu Z, Wang Y, et al. Origin of the T790M mutation and its impact on the clinical outcomes of patients with lung adenocarcinoma receiving EGFR-TKIs. Pathol Res Pract. 2019;215(5):946–51.
17. Lee SH, Kim EY, Kim A, Chang YS. Clinical implication and usefulness of de novo EGFR T790M mutation in lung adenocarcinoma with EGFR-tyrosine kinase inhibitor sensitizing mutation. Cancer Biol Ther. 2020;21(8):741–8.
18. Matsumoto Y, Sawa K, Fukushima M, Oyanagi Y, Yoshimoto N, Suzumura T, et al. Predictive impact of low-frequency pretreatment T790M mutation in patients with EGFR-mutated non-small cell lung cancer treated with EGFR tyrosine kinase inhibitors. Lung Cancer. 2020;13983–8.
19. Beau-Faller M, Pencrach E, Leduc C, Blons H, Merlio JP, Bringuer PE, et al. Independent prognostic value of ultra-sensitive quantification of tumor pretreatment T790M subclones in EGFR-mutated non-small cell lung cancer.

Abbreviations

NSCLC: Non-small cell lung cancer; EGFR: Epidermal growth factor receptor; 19del: Exon 19 deletions; L858R: Leu858Arg point mutations in exon 21, T790M: Tyrosine kinase inhibitor, preT790M: Pretreatment T790M; ddPCR: Droplet digital polymerase chain reaction; DFS: Disease-free survival; MAF: Mutant-allele frequency; RFS: Recurrence-free survival; OS: Overall survival; HR: Hazard ratio; CI: Confidence interval.

Supplementary Information

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Ethical approval and consent to participate

This study was approved by the Institutional Review Board of the National Hospital Organization of Japan (UMIN00008177). All patients provided written, informed consent before surgery. All methods in this study were performed in accordance with the relevant guidelines and regulations.

Consent for publication

Not applicable.

Competing interests

All authors declare no potential conflicts of interest related to the submitted work.

Author details

1 Department of Respiratory Medicine, Graduate School of Medicine, Osaka Metropolitan University, Osaka, Japan. 2 Internal Medicine III, Wakayama Medical University, Wakayama, Japan. 3 Clinical Research Center, National Hospital Organization Kinki-chuo Chest Medical Center, Sakai, Japan. 4 Advanced Medicine and Clinical Research, Naqoya University Hospital, Naqoya, Japan. 5 Internal Medicine, National Hospital Organization Kinki-chuo Chest Medical Center, Sakai, Japan. 6 Division of Respiratory Medicine and Allergology, Department of Internal Medicine, Aichi Medical University School of Medicine, Nagakute, Japan. 7 Medical Oncology and Respiratory Medicine, Nagoya Medical Center, Nagoya, Japan. 8 Respiratory Medicine, Ishikiriseiki Hospital, Higashiosaka, Japan. 9 Center for Biomedical Sciences, CIEMS, Wakayama Medical University, 811-1 Kimidera, Wakayama-shi, Wakayama 641-8509, Japan.

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Matsuo N, Azuma K, Sakai K, Katoh S, Wakama K, Westphal M, et al. Localization of EGFR exon 19 deletion and EGFR-TKI treatment duration with frequency of T790M mutation in EGFR-mutant lung cancer patients. Sci Rep. 2016;6:36458.

Chmielecki J, Foo J, Oxnard GR, Hutchinson K, Ohashi K, Somwar R, et al. Optimization of dosing for EGFR-mutant non-small-cell lung cancer with evolutionary cancer modeling. Sci Transl Med. 2011;3(98):90oa59.

Nakayama S, Sng N, Carretero J, Welner R, Hayashi Y, Yamamoto M, et al. beta-catenin contributes to lung tumor development induced by EGFR mutations. Cancer Res. 2014;74(20):5891–902.

Togashi Y, Hayashi H, Terasima M, de Velasco MA, Sakai K, Fujita Y, et al. Inhibition of beta-catenin enhances the anticancer effect of irreversible EGFR-TKI in EGFR-mutated non-small-cell lung cancer with a T790M mutation. J Thorac Oncol. 2015;10(1):93–101.

Pignon JP, Tribodet H, Scagliotti GV, Douillard JY, Shepherd FA, Stephens RJ, et al. Lung adjuvant cisplatin evaluation: a pooled analysis by the LACE collaborative group. J Clin Oncol. 2008;26(21):3552–9.

Hamada C, Tanaka F, Ohba M, Fujimura S, Kodama K, Imaizumi M, et al. Meta-analysis of postoperative adjuvant chemotherapy with tegafur-uracil in non-small-cell lung cancer. J Clin Oncol. 2015;33(22):2499–5006.

Goss GD, O'Callaghan C, Lorimer R, Tiao MS, Masters GA, Jett J, et al. Gefitinib versus placebo in completely resected non-small-cell lung cancer: results of the NOC CTC BR19 study. J Clin Oncol. 2013;31(27):3320–6.

Kelly K, Altorki NK, Eberhardt WE, O'Brien ME, Spiegel DR, Cino L, et al. Adjuvant Erlotinib versus placebo in patients with stage II-III A non-small-cell lung cancer (RADIANT): a randomized, double-blind, Phase III Trial. J Clin Oncol. 2015;33(34):4007–14.

Pennell NA, Neal JW, Chaft JE, Azzoli CG, Janne PA, Govindan R, et al. Select: a phase II trial of adjuvant Erlotinib in patients with resected epidermal growth factor receptor-mutant non-small-cell lung cancer. J Clin Oncol. 2019;37(2):97–104.

Zhong WZ, Wang Q, Mao WM, Xu ST, Wu L, Shen Y, et al. Gefitinib versus vinorelbine plus cisplatin as adjuvant treatment for stage II-IIIA (N1-N2) EGFR-mutant NSCLC (ADJUVANT/CTONG1104): a randomised, open-label, phase 3 study. Lancet Oncol. 2018;19(1):139–48.

Li N, Ou W, Ye X, Sun HB, Zhang L, Fang Q, et al. Pemetrexed-carboplatin adjuvant chemotherapy with or without gefitinib in resected stage IIIA-N2 non-small-cell lung cancer harbouring EGFR mutations: a randomized, phase II study. Ann Surg Oncol. 2014;21(8):2091–6.

Yue D, Xu S, Wang Q, Li X, Shen Y, Zhao H, et al. Erlotinib versus vinorelbine plus cisplatin as adjuvant therapy in Chinese patients with stage IIA EGFR mutation-positive non-small-cell lung cancer (EVAN): a randomised, open-label, phase 2 trial. Lancet Respir Med. 2016;8(11):985–93.

Huang Q, Li J, Sun Y, Wang R, Cheng X, Chen H. Efficacy of EGFR tyrosine kinase inhibitors in the adjuvant treatment for operable non-small-cell lung Cancer by a Meta-analysis. Chest. 2016;149(6):1384–92.

Raphael J, Vincent M, Boldt G, Shah PS, Rodrigues G, Blanchette P Adjunctive epidermal growth factor receptor tyrosine kinase inhibitors (TKIs) in resected non-small-cell lung Cancer (NSCLC): a systematic review and Meta-analysis. Ann J Clin Oncol. 2019;42(5):440–5.

Tang W, Li X, Xie X, Sun X, Liu J, Zhang J, et al. EGFR inhibitors as adjuvant therapy for resected non-small-cell lung cancer harboring EGFR mutations. Lung Cancer. 2019;136:6–14.

Majem M, Sullivan I, Viteri S, Lopez-Vivanco G, Cobo M, Sanchez JM, et al. The significance of micro-EGFR T790M mutation on EGFR-TKI efficacy in patients with NSCLC: the WJOG13119L study. J Clin Oncol. 2022;40(16_suppl):e21177.