Acute kidney injury with dark urine: the case of paroxysmal nocturnal hemoglobinuria

Kamel El-Reshaid *, Hasan Sabri **, Shaikha Al-Bader **

* Department of Medicine, Faculty of Medicine, Kuwait University, Kuwait
** Department of Medicine, kidney unit, Al-Amir Hospital, Kuwait

ABSTRACT
In this case report; we describe a patient with severe attack of paroxysmal nocturnal hemoglobinuria (PNH) following Ciprofloxacin therapy. He presented with recurrent abdominal pain, repeated vomiting and dark urine. Physical examination revealed severe pallor and jaundice. Laboratory investigations showed severe intravascular hemolysis with negative Coomb's test, progressive acute kidney injury (AKI), sterile blood cultures and negative serology for autoimmune diseases. Subsequently, he developed pancytopenia. Ham test was positive and flow cytometry, later on, confirmed PNH. He was supported with multiple transfusions of packed blood cells and hemodialysis. Ciprofloxacin was discontinued and his PNH was treated with Solunmedrol 1 g daily for 3 days followed by Prednisone 60 mg/day for 2 months. Two weeks later; his hemolysis abated and his AKI improved. Up to 5 years later, he still has minor PNH clone yet without disease activity. In conclusion; our patient had acute drug-induced hemolytic crisis associated with minor PNH clone. With drug-vigilance; no further relapses were reported and his PNH clone remained stable for 5 years. The case expands the spectrum of PNH phenotypes and its triggering factors.

Keywords: AKI, anemia, autoimmune, flow cytometry, hemolysis, PNH

INTRODUCTION
Paroxysmal nocturnal hemoglobinuria (PNH) is a Coomb's negative intravascular hemolytic anemia characterized by attacks of hemolysis associated with dark urine due to severe hemoglobinuria 1. It is an acquired nonmalignant clonal expansion of one or several hematopoietic stem cells that have acquired somatic mutation leading to loss of complement decay activating factor (DAF). The latter protects progeny of hematopoietic cells from lysis by activated complements during innate immunological responses in infections, allergy and exposure to hapatens 2. It is a rare disorder with a prevalence of 0.5-1.5 per million people in the general population and with an average of 400 cases being diagnosed annually in USA 3. Its multiple phenotypic forms and life-threatening complications mask its definitive diagnosis, alter its management and hence limit patients’ survival 4. In this case report; we describe a patient with this disorder and highlight the available means of diagnosis and management.

THE CASE:
A 45-year-old Egyptian man presented with abdominal pain, vomiting and dark urine for 3 days. Past history was significant for fever and body aches 1 week ago for which Ciprofloxacin was used. On his initial physical examination; he was conscious and oriented X3. He had dyspnea on mild effort and tachycardia yet was normotensive and with normal BP. He had severe pallor and jaundice. Initial and subsequent laboratory investigations are summarized in Table 1. Initially he had mildly decreased hemoglobin and hematocrit yet was normocytic and normochromic. AST and ALT were normal. Total and direct bilirubin, AST and LDH were high. Urine routine showed 4(+) blood yet on microscopy erythrocytes were < 5/HPF. Stool testing was negative for ova, parasites and occult blood. Blood and urine culture were negative for pathogens. Blood film did not show significant abnormality except for fragmented RBCs.
“schistocytes” (Fig. 1). No malaria parasites were seen. Prothrombin and activated thromboplastin time were normal. Serum fibrinogen and procalcitonin levels were normal. Direct and indirect Coombs test were negative. G6PD level and hemoglobin electrophoresis were normal. Serum complements (C3 & C4) and protein electrophoresis was normal. ANA, anti-ds DNA, ANCA, anti-GBM antibodies, hepatitis B surface antigen and anti-HCV antibodies were negative. Abdominal and pelvic ultrasound did not show abnormality and Doppler study did not show venous and arterial renovascular disease. Echocardiogram did not show abnormality. Ham test (sucrose lysis test) was positive and flow cytometry confirmed PNH later. Initially; ciprofloxacin was discontinued. His severe pancytopenia and progressive renal failure were treated with multiple blood transfusions and supportive hemodialysis. Since both did not improve by the 3rd day; Solumedrol was started as 1 g IV daily for 3 days followed by Prednisone 60 mg/day for 2 months. By 2 weeks; his clinical, hematological and biochemical abnormalities had improved and hence dialysis was discontinued. Two months after recovery; 24 hour urine testing for amino acids showed normal values. On follow up, and up to 5 years, his disease remained inactive with stable and minor PNH clone.

Figure 1: Peripheral blood smear showing schistocytes.
Table 1: Initial and follow up laboratory investigations in the case of paroxysmal nocturnal hemoglobinuria (PNH)

| Test/(Normal range)                     | At diagnosis | 3 days later | 2 weeks later | 5 years later |
|-----------------------------------------|-------------|-------------|---------------|--------------|
| **blood counts:**                       |             |             |               |              |
| Peripheral leucocytic count (4-10 X10^9/L) | 3           | 2           | 3             | 6            |
| Hemoglobin (130-160 g/L)                | 75          | 65          | 98            | 120          |
| Peripheral platelets count (150-450 X10^9/L) | 114         | 25          | 135           | 190          |
| Reticulocyte % (0.5-2.5%)               | 8           | 11          | 2             | 0.1          |
| Peripheral blood schistocytes (Absent)  | Excess      | Excess      | Scanty        | Absent       |
| **Biochemistry:**                       |             |             |               |              |
| Haptoglobin (0.5-2.2 g/L)               | 0.06        | 0.04        | 0.4           | 2.1          |
| Serum creatinine (64-104 umol/L)        | 503         | 942         | 218           | 120          |
| Total bilirubin (5-21 umol/L)           | 97          | 120         | 32            | 8            |
| Direct bilirubin (1-5 umol/L)           | 20          | 30          | 8             | 3            |
| Aspartate aminotransferase (AST): (3-35 IU/L) | 478         | 566         | 41            | 26           |
| Lactic dehydrogenase (100-190 IU/L)     | 2895        | 3124        | 631           | 154          |
| **Urine routine:**                      |             |             |               |              |
| Blood (absent)                          | 4(+)        | 4(+)        | 1(+)          | Negative     |
| **Flow cytometry:**                     |             |             |               |              |
| RBC                                      |             |             |               |              |
| Type II (partial CD59 deficiency)        | 0.40%       |             |               | 0.60%        |
| Type III (complete CD59 deficiency)     | 0.80%       |             |               | 1.80%        |
| Total RBC PNH clone size                | 1.20%       |             |               | 2.40%        |
| WBC-Neutrophils (FLAER/CD24 deficiency) | 1.30%       |             |               | 2.60%        |
| WBC-Monocytes (FLAER/CD14 deficiency)   | 1.80%       |             |               | 2.10%        |
| **Diagnosis:**                           | Minor PNH clone | Minor PNH clone |          |              |
DISCUSSION

Our patient presented with progressive renal failure and severe intravascular hemolytic anemia. The increment in serum creatinine > 26 umol/L within 48 hours indicated an AKI event. Moreover, the normal kidney ultrasound, Doppler study, lack of history of hyperperfusion, absence of urinary hematuria and proteinuria excluded pre-renal, post-renal, vascular, glomerular and autoimmune etiologies as well as an underlying chronic disease. Previous Ciprofloxacin exposure had raised an issue of acute interstitial nephritis yet the clinical picture was far from being an isolated event. Though he had evident hemolysis and hemoglobinuria; testing 24 hour urine was done to exclude Alkaptonuria 2 months after clearance of hemoglobinuria. Work up for his severe anemia excluded; inherited membrane defects (G6PD), hemoglobinopathies (thalassemias and sickle cell disease), hereditary spheroctysis and elliptocytosis, falciparum malaria, and autoimmune ones. Moreover, his anemia was (a) an acute event with high reticulocytes, (b) mediated by intravascular hemolysis with high direct bilirubin and LDH levels as well as intravascular with high hemoglobinemia and hemoglobinuria, (c) lacked autoimmune etiology with negative Coombs test, and (d) associated with normal bone marrow biopsy, and (e) associated with pancytopenia and blood “schistocytes” simulating microangiopathic picture. The latter was without malignant hypertension, scleroderma signs, disseminated intravascular coagulation (DIC) with prolonged coagulation tests and low fibrinogen, sepsis with fever and high procalcitonin levels. However, 2 microangiopathic disorders could not be ruled out with certainty viz; sporadic form of hemolytic uremic syndrome and thrombotic thrombocytopenic purpura. However, keeping a higher stage of diagnosis with severe intravascular hemolysis and AKI was safer to avoid side tracts with those 2 diagnoses. With hemoglobinuria; AKI is expected due to proteins liberated from hemolysed cells leading to vasoconstriction and nephrotic acute tubular necrosis.

Regarding intravascular hemolysis; PNH is a rare disorder and can be associated with pancytopenia and may lack a myelodysplastic disease. It can be screened with Ham test yet confirmed with flow cytometry of patient’s serum erythrocytes, neutrophils and monocytes. Definite diagnosis of the etiology of AKI can be established by a kidney biopsy. In our patient, we elected to be conservative since; (a) initially he had severe thrombocytopenia, (b) drug-induced acute interstitial nephritis and hemolytic crisis of PAN usually respond to high-dose Corticosteroids, and (c) Ham test and flow cytometry were positive. PNH is a unique form of immune-mediated hemolytic anemia induced by inadequate suppression of activated complement leading to erythrocytes, leucocytes and platelets lysis. Complement activation is part of innate immunity. Lack of suppression is due to failure to activate complement convertase by a deficient cell-anchor protein or DAF. The latter deficiency is: (a) an acquired one since it develops as a non-malignant expansion of mutated clones of stem cells; (b) the gene mutation is an X-linked with deficiency in glycosyl phosphatidylinositol-anchored proteins (GPI-APs) viz. CD59 on erythrocytes and CD14, 24, 157 on leucocytes. Such clone expansion can develop as an isolated disorder and hence is referred to as the classic form of PNH or in associated with myelodysplastic syndrome (MDS) in type II while type III presents with subclinical PNH yet with evident BM failure. Clinically, the disease may present with an isolated erythrocytic or pancytopenic lytic-attacks according to the extent of the initial stem cell mutation. Moreover, its paroxysmal lytic attacks depends on activation of innate immune system during infections, autoimmune diseases, lymphoproliferative disorders, pregnancy, vaccinations and medications (hapatens) which activates complements. While other cells enhance production of convertase that deactivates the activated complements; the affected clone cells in PNH are deficient from such DAF and hence are lysed during such circumstances. Our patient had PNH without MDS i.e. classic form of disease. Moreover, he had pancytopenic presentation due to complement-activation subsequent to Ciprofloxacin therapy. Fortunately, his severe hemolytic crisis was aborted with drug-discontinuation and short-term high-dose Corticosteroids. Complications were limited to pigment-induced AKI. Most patients report dysphagia, abdominal pain, dyspnea and erectile dysfunction during hemolytic crisis. The latter is attributed to spasm of the smooth muscles due to depletion of nitric acid by hemolytic products. Forty% of patients with PNH are susceptible to venous thrombosis yet limb and pelvic venous thrombosis predispose to recurrent and fatal pulmonary emboli. Moreover, thrombotic disease can involve the portal vein leading to Budd-Chiari syndrome, intestinal gangrene with thrombosis of mesenteric vein and portal hypertension with pulmonary vein disease. Specific therapy of the disease and anticoagulation should be individualized for each case. As in our patient the disease was limited to a single attack and he enjoyed long-lasting remission with precautions regarding drug and vaccination-use. Some patients may need: (a) infrequent Corticosteroid therapy to avoid its long-term side effects, (b) Danozol treatment with caution due to its virilizing effect in women, (c) Eculizumab infusions if side effects, meningococcal vaccination and cost are tolerated, and lastly (d) bone marrow transplantation if donors are available. At present the latter has been proposed as the definitive therapy for the disease and Eculizumab for the acute flares. PNH usually presents in adults yet extremes of age are not spared. Children with severe disease; should have stem-cell bone marrow transplantation to avoid long-term disease management or its thromboembolic complications. The affected females are at high risk for thromboembolic disease (20.8%) and may need life-long anticoagulation if associated with major clone disease and during pregnancy. In conclusion; our patient had acute drug-induced hemolytic crisis associated with minor PNH clone. With drug-vigilance; no further relapses were reported and his PNH clone remained stable for 5 years.

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