Density-dependent effects on the survival and growth of the rodent stomach worm Protospirura muricola in laboratory mice

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Abstract

The spirurid nematode, Protospirura muricola, is of intrinsic interest as a rodent model of gastric nematode infections. Since worm burdens can be very heavy in nature, density dependent processes may constrain parasite growth. Laboratory mice (BKW) were exposed to varying doses of infective larvae of P. muricola in the range 5 to 40 third-stage larvae (L3), in four separate experiments in which progressively higher doses were utilized. All mice were culled 60 days after infection and a total of 518 worms (226 male and 292 female worms) was recovered, measured and weighed. Overall survival was 58.9%, but survival declined significantly with increasing dose by approximately 21% (from 66% at 5 L3 per mouse to 52% at 40 L3 per mouse). The length and weight of worms correlated positively in both sexes. Total worm biomass increased linearly with increasing numbers of worms. However, whilst the length and weight of male worms declined with increasing worm burden (8.4 and 24.6% respectively), female worms were less affected, only length showing a significant reduction with increasing parasite burden (16.0%). Therefore, increasing worm burdens impeded growth of P. muricola, but reduction in length and weight were relatively small in relation to the overall size of this nematode. Increasing worm burdens were associated with loss of host weight and reduction in stomach weight and worm burdens in excess of 20 exerted a measurable cost to the host, which in the field, may be associated with loss of overall host fitness.

Introduction

Density-dependent effects on the growth and reproductive capacity of helminths have been well documented in many different species in naturally infected populations (Anderson & May, 1978; Keymer, 1982) but are perhaps best known from experimental cestode infections. For example the rat tapeworm, Hymenolepis diminuta, shows a marked inverse relationship between the number of parasites harboured and their individual lengths and weights, and this has been recognized for some time (Chandler, 1939; Read, 1951; Kennedy & Behnke, 2001). In contrast, nematodes are generally considered to be more resilient and although their growth in experimental systems can be constrained with increasing parasite burden (Michael & Bundy, 1989; Norozian-Amiri & Behnke, 1994), the effects on length and weight are less marked than in cestode infections. The mechanistic basis of density-dependent effects on helminth size and reproductive effort has been the subject of debate and controversy, and has spawned hypotheses ranging from competition for limited resources, regulatory factors of parasite origin to host immune responses (Andreassen et al., 1999; Bush & Lotz, 2000; Roberts, 2000; Paterson & Viney, 2002).

During field work in the south of the Sinai peninsula of Egypt, we encountered a relatively high prevalence of the spirurid nematode Protospirura muricola in the dominant wild rodent in the region, the spiny mouse Acomys dimidiatus (Behnke et al., 2000). Protospirura muricola is a large stomach nematode reaching over 5 cm in length (females), commonly encountered in the tropics.
in a variety of hosts, including rodents, and often accumulating in hosts with resultant heavy worm burdens (Baylis, 1928; Campos & Vargas, 1978; Behnke et al., 2000). Some naturally infected spiny mice can harbour over 100 worms, with total biomass exceeding 2% of host body weight (Behnke et al., unpublished observations; Behnke et al., 2000) and occupying most of the stomach lumen. On the basis of observations made in the field we suggested that this species may be costly for the host (Behnke et al., 2000; Barnard et al., 2003), but currently virtually nothing is known about the host–parasite relationship of P. muricola.

In this paper, employing a strain of P. muricola recently isolated from A. dimidiatus in the Sinai and under laboratory conditions, we examine the hypothesis that density-dependent constraints on the growth of P. muricola may limit the total biomass that can develop and survive to maturity in mice. We test the predictions that parasite survival, worm length and weight should decline with increasing numbers of worms in the stomach. We also present preliminary observations on the effects of this parasite on host body weight and the weight of the stomach within which the worms reside.

Materials and methods

Animals

All infections were initiated in 6- to 8-week-old male BKW laboratory mice (Mus musculus), purchased from B and K Universal, which have been found to be suitable hosts for the maintenance of this species under laboratory conditions. The mice were fed on Teklad Rat/Mouse/Breeding Diet, supplied by Harlem Laboratory Animal Feed limited, provided with both food and water ad libitum and maintained on a 12 h light: 12 h dark regime.

Maintenance, infection and recovery of Protospirura muricola

The isolate of P. muricola was obtained in 1997 from A. dimidiatus, from the St Katherine’s Protectorate in the Sinai region of Egypt. Eggs from adult female worms were allowed to embryonate and were fed to Tribolium confusum beetles in flour. The第三-stage larvae (L3) developed in the haemocoel of the insect after 4–5 weeks and these were dissected and used to infect mice for passage, by conventional oral gavage. In our experience, this strain of P. muricola develops only in the stomach of its host. At the end of each experiment mice were culled by exposure to carbon dioxide, weighed, and the body cavity opened. The stomach was removed and placed in warm Hanks’ saline in a Petri dish and carefully opened using forceps, to ensure that the worms inside were not damaged. Worms were removed with forceps and the stomach was carefully washed out with Hanks’ saline to free it of any remaining contents. It was then dabbed gently on a paper towel to remove excess liquid and weighed.

Experimental design

Four experiments were carried out in each of which a range of infection doses was employed. In the first experiment we explored doses not exceeding 15 L3, but in subsequent experiments the range was extended gradually to 40 L3. The step-wise exploration with increasing doses was required on ethical grounds, because this species grows to a relatively large size and little is known about its potential pathogenicity to the host (but see Foster & Johnson, 1939). The doses employed and the number of mice surviving to day 60 post-infection, when all mice were culled, are shown in table 1.

Statistical analysis

Throughout, multivariate or univariate GLIM was used, implemented by the software package SPSS for Windows, version 9.0.0. To assess whether survival of worms was affected by the dose of larvae administered, the worm recovery of each mouse was converted to a percentage of the dose administered, the frequency distributions were checked for normality and analysis was by ANOVA with percentage recovery as the dependent variable, the experiment as a factor and the dose of larvae administered as a covariate. Residuals from a one-way ANOVA, with experiment as the factor, on percentage recovery were saved to illustrate the relationship between dose and recovery with between-experiment variation taken into account.

Individual lengths and weights of worms were averaged by sex for each mouse in the study, and the analysis was conducted on these mean values. To ensure that type 1 errors were not likely, a multivariate analysis was first conducted with worm length and weight for both sexes as dependent variables, and in each case using models that incorporated either number of worms (test 1) or total biomass (test 2) as covariates. Experiment was also included as a factor (to control for between-experiment differences) and percentage of female worms as a covariate (to control for sex bias because female worms are longer than males and considerably heavier). For the first of these analyses Wilks’ lambda was 0.554 for number of worms (F4,43 = 8.667, P < 0.001), 0.801 for percentage of female worms (F4,43 = 2.671, P = 0.045) and 0.285 for experiment (F12,114 = 5.769, P < 0.001). For the second, Wilks’ lambda was as 0.633 for total worm biomass (F4,44 = 6.368, P < 0.001) and 0.323 for experiment (F12,117 = 5.19, P < 0.001). Since both indicated a highly significant effect of worm density, in a second step we employed univariate tests on each variable in turn to allow estimation of the gradient of the slope. Pearson’s product moment correlation test

| Experiment no. | 0 | 5 | 10 | 15 | 20 | 25 | 40 |
|---------------|---|---|----|----|----|----|----|
| 1             | 5* | 5 | 5  | 5  | 4  |    |    |
| 2             | 5  | 5 | 5  | 5  | 2  |    |    |
| 3             | 5  | 5 | 5  | 5  |    |    |    |
| 4             | 5  | 5 | 5  | 5  |    |    |    |

* Each group originally comprised 5 mice.
was used to test the relationship between worm weight and length.

**Results**

**Worm recovery and sex ratio**

A total of 518 worms were recovered comprising 226 male and 292 female worms, giving an overall sex ratio of 56.4% females. The sex ratio did not vary between experiments (ANOVA with experiment as factor and number of worms recovered as a covariate; $F_{3,51} = 1.646$, $P = \text{NS}$) nor with increasing worm burden ($F_{1,51} = 0.049$, $P = \text{NS}$). The overall survival of worms was 58.86% of the dose administered.

**Recovery of worms**

Raw data for percentage recovery from all four experiments combined are illustrated in fig. 1a and this indicates that a small reduction in worm survival with increasing dose was detected. The slope of the regression was $\beta = -0.45 \pm 0.2$ and this was significantly different from zero ($t = -2.25$, $P = 0.03$). Figure 1b illustrates this relationship with minor but significant between-experiment differences taken into account (residuals from one-way ANOVA with experiment as the factor).

**Relationship between worm weight and worm length**

Figure 2 shows the relationship between individual worm weight and worm length for each of the sexes. For both sexes this relationship was positive and highly significant (males $r_p = 0.24$, $n = 226$, $P < 0.001$; females $r_p = 0.37$, $n = 292$, $P < 0.001$), although, as is evident from the figure, there was considerable variation in both sexes.

**Relationship between number of worms recovered and total worm biomass**

The relationship between the number of worms recovered from individual mice and the corresponding total biomass of worms is illustrated in fig. 3. This was essentially a linear positive relationship ($r_p = 0.98$, $n = 56$, $P < 0.001$).

**Effect of worm numbers on length of male and female worms**

Figure 4 shows the relationship between the lengths and weights of male and female worms, and the number of worms harboured at autopsy. The results of statistical analyses are summarized in table 2. Male worms were sensitive to the density of worms, their lengths and weights both declining with increasing worm density (by 8.4 and 24.6%). Both parameters were sensitive to the percentage of female worms present, with a negative.
gradient in relation to increasing percentage of female worms. There was also significant variation between experiments. Female worms were less sensitive. Only the length of female worms declined significantly with increasing worm burden (16.0%). There was no significant relationship with the percentage of female worms, and no difference between experiments.

**Effect of increasing worm burden on host body weight**

Mouse weight at autopsy declined with increasing worm burden whether worm recovery was used as a measure of the intensity of infection (fig. 6b) or the total worm biomass (data not shown, ANOVA with experiment as factor and total worm biomass as covariate, main effect of total worm biomass $F_{1,71} = 14.64$, $\beta = -0.008 \pm 0.002$, $P < 0.001$).

**Effect of increasing worm burden on stomach weight**

There was a significant loss of stomach weight with increasing worm recovery (fig. 6a) and with increasing total worm biomass (fig. 6c). Analysis of the stomach weight as a percentage of body weight did not generate significant relationships with number of worms recovered or total worm biomass (fig. 6d), most likely because body weight also declined with increasing worm burden. However, as a final step we fitted ANOVAs with experiment as factor and mouse weight as an additional covariate in order to control for differences in mouse weight. In both cases, the relationship between stomach weight and number of worms recovered and with total biomass was significant. (For statistical analyses see table 2.)

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**Fig. 3.** Relationship between the total worm biomass (wet weight) and the number of worms of *Protospirura muricola* recovered.

**Fig. 4.** Relationships between the length (a and c) and weight (b and d) of male (a and b) and female (c and d) *Protospirura muricola*, and the number of worms recovered on day 60 post-infection. For statistical analysis see table 2.
worm biomass, remained significant ($F_{1,70} = 6.761$, $P = 0.011$ and $F_{1,70} = 6.075$, $P = 0.016$, respectively).

**Discussion**

*Protospirura muricola* is primarily a nematode parasite of rodents living in the tropics, known mostly from records of its occurrence in Africa and South America (Baylis, 1928; Campos & Vargas, 1978). Very little is known about the host–parasite relationship of this species and experimental studies have been concerned largely with elucidating its growth and development in different intermediate and definitive hosts (Quentin, 1969; Campos & Vargas, 1977). *Protospirura muricola* is a long-lived but slowly developing parasite that takes 1.5 to 2 months to mature in its rodent host. Eggs first appear in mouse faeces 62 days after infection and infections can last many months thereafter (over 200 days, Lowrie, Behnke & Barnard unpublished observations). In the present experiments, mice were culled after 60 days to assess density dependent effects on growth within the prepatent

![Table 2. Univariate analysis of the length and weight of male and female worms of *Protospirura muricola* by ANOVA with the number of worms and percentage of female worms as covariates and experiment as a factor (4 levels).](image)

| Parameter                        | Male worms |             | Female worms |             |
|----------------------------------|------------|-------------|--------------|-------------|
|                                  | Length     | Weight      | Length       | Weight      |
| $F_{1,48}$ for no. of worms       | 4.235      | 11.044      | 16.261       | 0.926       |
| Gradient ($\beta$) ± SEM         | $-0.052 \pm 0.025$ | $-0.114 \pm 0.034$ | $-0.135 \pm 0.033$ | $-0.061 \pm 0.063$ |
| $t$                              | -2.058     | -3.323      | -4.033       | -0.962      |
| $P$                              | 0.045      | 0.002       | <0.001       | NS          |
| $F_{1,48}$ for % females          | 28.92      | 10.997      | 0.762        | 0.192       |
| Gradient ($\beta$) ± SEM         | $-0.044 \pm 0.008$ | $-0.037 \pm 0.011$ | 0.010 ± 0.01 | 0.010 ± 0.022 |
| $t$                              | -5.38      | -3.316      | 0.873        | 0.438       |
| $P$                              | <0.001     | 0.002       | NS           | NS          |
| $F_{3,48}$ for experiment        | 8.862      | 7.43        | 2.23         | 1.69        |
| $P$                              | <0.001     | <0.001      | NS           | NS          |

NS, not significant.
The experiments varied in the range of infection levels examined and higher worm burdens were only administered when it had been ascertained that there was no mortality associated with infection at the lower doses. None of the density-dependent effects detected were marked. Parasite survival for 60 days after infection declined by only 14%, indicating that over the range of 5 to 40 worms, total worm burdens were not a major factor in survival. At this stage it is unknown whether some worms failed to establish or whether they were lost during the 60 day infection period. It is conceivable that at higher worm burdens host immune mechanisms become effective or competition between worms for resources restrict numbers, but currently these are unexplored aspects of this host–parasite relationship. Nevertheless, it is noteworthy that in naturally infected hosts worm burdens can exceed 100, so wild *A. dimidiatus* can tolerate such burdens, even though they occupy the vast majority of the stomach lumen and presumably limit the feeding capacity of the host.

Table 3. Univariate analysis of the length and weight of male and female worms of *Protospirura muricola* by ANOVA with total worm biomass as a covariate and experiment as a factor (4 levels).

| Parameter                      | Male worms                  | Female worms                |
|-------------------------------|-----------------------------|-----------------------------|
|                               | Length | Weight | Length | Weight |
| *F*<sub>1,48</sub> for total worm biomass | 4.509  | 10.227 | 11.203 | 0.330  |
| Gradient (β) ± SEM            | −0.003 ± 0.002 | −0.006 ± 0.002 | 0.006 ± 0.002 | 0.002 ± 0.003 |
| *t*                           | −2.124 | −3.198 | −3.347 | 0.574  |
| *P*                           | 0.039  | 0.002  | 0.002  | NS     |
| *F*<sub>3,48</sub> for experiment | 6.145  | 5.635  | 1.851  | 1.896  |
| *P*                           | 0.001  | 0.002  | NS     | NS     |

NS, not significant.

Fig. 6. The effect of *Protospirura muricola* on host stomach weight (a, c and d) and body weight (b). Statistical analysis of host body weight (b) as the dependent variable by ANOVA with experiment as factor and number of worms recovered as covariate, *F*<sub>1,71</sub> = 14.4, β = −0.163 ± 0.043, *P* < 0.001. Analysis of weight of stomach (a) by ANOVA with experiment as factor and number of worms recovered as covariate, gave *F*<sub>1,71</sub> = 11.68, β = −0.003 ± 0.001, *P* = 0.001. Analysis of weight of stomach (c) with experiment as factor and total worm biomass as covariate, gave *F*<sub>1,71</sub> = 10.84, *P* = 0.002.
total worm biomass. Therefore, in contrast to cestode infections, there was no intestinal maximum holding capacity restricting further increases in biomass (Moss, 1971). It is conceivable that such a limit, represented by an expected plateau to the relationship (fig. 3), would have become evident at higher infection doses, but the total biomass in heavily infected hosts was substantial at 1.58% of host body weight, and ethical considerations limited exploration of higher levels.

A decline in the lengths of both males and females was shown with increasing numbers of worms, and of weight in the case of males, but no density-dependent decline in the weight of female worms was observed. Moreover, male worms were sensitive to the percentage of female worms present (after controlling for differences between experiments and variation in worm burden, table 2), both male length and weight declining as the percentage of female worms increased. In contrast, the relative proportion of sexes affected neither female worm lengths nor weights. Male worms are smaller than female worms (fig. 2), and their increased sensitivity to crowding is consistent with both findings. In these experiments, we did not quantify eggs in female worms, but it is possible that the egg content of female worms may have been a complicating issue. Female worms tend to shed eggs quickly when transferred to Hanks’ medium (Lowrie, personal observations) and variation in the duration of incubation prior to fixation in ethanol, as well as between the egg content of individual worms, may have been responsible for generating sufficient noise to cloud any underlying relationship. In any case, all gradients were very shallow (table 2). Extrapolating from the fitted model in table 2, for male worms, those recovered from mice with high worm burdens (32 worms) were 1.56 mm shorter than those in low intensity infections (2 worms), and 3.42 mg lighter (= 24.6% reduction). In the case of females, the reduction in length with increasing worm burden was 4.1 mm (= 16.0%) and weight 1.8 mg (= 6.5%). Thus, from present data, density-dependent constraints on the growth of P. muricola were relatively small.

Observations on captive white-faced monkeys (Cebus capucinus) indicate that P. muricola can also infect primates in which infections can exceed 500 worms. Foster & Johnson (1939) described heavily infected white-faced monkeys in which worms migrated out of the stomach into the body viscera and were responsible for death of the heavily infected animals. In the present system, P. muricola is confined to the stomach of mice but worm burdens are much lower and it could be that movement out of the stomach is initiated when crowding passes a critical threshold. Nevertheless worm burdens in excess of 20 exerted a measurable cost to the host, which in the field, may be associated with loss of overall host fitness.

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References

Anderson, R.M. & May, R.M. (1978) Regulation and stability of host–parasite population interactions I. Regulatory processes. Journal of Animal Ecology 47, 219–247.

Andreassen, J., Bennet-Jenkins, E.M. & Bryant, C. (1999) Immunology and biochemistry of Hymenolepis diminuta. Advances in Parasitology 42, 223–275.

Barnard, C.J., Sayed, E., Barnard, L.E., Behnke, J.M., Abdel Nabi, I., Sherif, N., Shutt, A. & Zalat, S. (2003) Local variation in helminth burdens of Egyptian spiny mice (Acomys cahirinus dimidiatus) from ecologically similar sites: relationships with hormone concentrations and social behaviour. Journal of Helminthology 77, 197–209.

Baylis, H.A. (1928) On a collection of nematodes from Nigerian mammals (chiefly rodents). Parasitology 20, 280–304.

Behnke, J.M., Barnard, C.J., Mason, N., Harris, P.D., Sherif, N.E., Zalat, S. & Gilbert, F.S. (2000) Intestinal helminths of spiny mice (Acomys cahirinus dimidiatus) from St Katherine’s Protectorate in the Sinai, Egypt. Journal of Helminthology 74, 31–43.

Bush, A.O. & Lotz, J.M. (2000) The ecology of “crowding”. Journal of Parasitology 86, 212–213.
Campos, M.Q. & Vargas, M.V. (1977) Biología de Protospirura muricola Gedoelst, 1916 y Mastophorus muris (Gmelin, 1790) (Nematoda: Spiruridae), en Costa Rica, I. Huespedes intermediarios. Revista de Biología Tropical 25, 191–207.

Campos, M.Q. & Vargas, M.V. (1978) Biología de Protospirura muricola Gedoelst, 1916 y Mastophorus muris (Gmelin, 1790) (Nematoda: Spiruridae), en Costa Rica, II. Huespedes definitivos. Revista de Biología Tropical 26, 199–211.

Castro, G.A. (1990) Intestinal pathology. pp. 283–316 in Behnke, J.M. (Ed.) Parasites: immunity and pathology. London, Taylor & Francis.

Chandler, A.C. (1939) The effect of number and age of worms on development of primary and secondary infections with Hymenolepis diminuta in rats and an investigation into the nature of premunition in tapeworm infections. American Journal of Hygiene 29, 105–114.

Foster, A.O. & Johnson, C.M. (1939) A preliminary note on the identity, life cycle and pathogenicity of an important nematode parasite of captive monkeys. American Journal of Tropical Medicine 19, 265–277.

Kennedy, C.R. & Behnke, J.M. (2001) Intraspecific competition in the cestode, Hymenolepis diminuta in rats. pp. 161–165 in Halton, D.W., Behnke, J.M. & Marshall, I. (Eds) Practical exercises in parasitology. Cambridge, Cambridge University Press.

Keymer, A. (1982) Density dependent mechanisms in the regulation of intestinal helminth populations. Parasitology 84, 573–587.

Michael, E. & Bundy, D.A.P. (1989) Density dependence in establishment, growth and worm fecundity in intestinal helminthiasis: the population biology of Trichuris muris (Nematoda) infection in CBA/Ca mice. Parasitology 98, 451–458.

Moss, G.E. (1971) The nature of the immune response of the mouse to the bile duct cestode Hymenolepis microstoma. Parasitology 62, 285–294.

Norozian-Amiri, S.M.B. & Behnke, J.M. (1994) Density dependent regulation of the growth of the hookworms Necator americanus and Ancylostoma ceylanicum. Parasitology 109, 119–128.

Paterson, S. & Viney, M.E. (2002) Host immune responses are necessary for density dependence in nematode infections. Parasitology 125, 283–292.

Quentin, J.C. (1969) Cycle biologique de Protospirura muricola Gedoelst 1916 (Nematoda; Spiruridae). Annales de Parasitologie 44, 485–504.

Read, C.P. (1951) The “crowding” effect in tapeworm infections. Journal of Parasitology 37, 174–178.

Roberts, L.S. (2000) The crowding effect revisited. Journal of Parasitology 86, 209–211.

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