Safety and Immunogenicity of a Second Dose of an Investigational Maternal Trivalent Group B Streptococcus Vaccine in Nonpregnant Women 4–6 Years After a First Dose: Results From a Phase 2 Trial

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Background. Maternal immunization against group B streptococcus (GBS) could protect infants from invasive GBS disease. Additional doses in subsequent pregnancies may be needed. We evaluated the safety and immunogenicity of a second dose of an investigational trivalent CRM197-glycoconjugate GBS vaccine (targeting serotypes Ia/II/III), administered to nonpregnant women 4–6 years postdose 1.

Methods. Healthy women either previously vaccinated with 1 dose of trivalent GBS vaccine 4–6 years before enrollment (n = 53) or never GBS vaccinated (n = 27) received a single trivalent GBS vaccine injection. Adverse events (AEs) were recorded. Serotype-specific (Ia/II/III) anti-GBS antibodies were measured by multiplex immunoassay prevaccination and 30/60 days postvaccination.

Results. AEs were reported with similar rates after a first or second dose; none were serious. Of previously GBS-vaccinated women, 92%–98% had anti-GBS concentrations that exceeded an arbitrary threshold (8 μg/mL) for each serotype 60 days postdose 2 vs 36%–56% postdose 1 in previously non–GBS-vaccinated women. Of previously GBS-vaccinated women with undetectable baseline (predose 1) anti-GBS levels, 90%–98% reached this threshold postdose 2. For each serotype, anti-GBS geometric mean concentrations (GMCs) 30/60 days postdose 2 in previously GBS-vaccinated women were ≥200-fold higher than baseline GMCs. Among women with undetectable baseline anti-GBS levels, postdose 2 GMCs in previously GBS-vaccinated women exceeded postdose 1 GMCs in previously non–GBS-vaccinated women (≥7-fold).

Conclusions. A second trivalent GBS vaccine dose administered 4–6 years postdose 1 was immunogenic with a favorable safety profile. Women with undetectable preexisting anti-GBS concentrations may benefit from a sufficiently spaced second vaccine dose.

Clinical Trials Registration. NCT02690181

Keywords. group B streptococcus; maternal immunization; second dose; safety; immunogenicity.
contains CPS from GBS serotypes Ia, Ib, and III, which together cause >85% of infant invasive GBS disease [2], conjugated to the CRM197 carrier protein (nontoxic mutant of diphtheria toxin) was well tolerated and immunogenic in nonpregnant and pregnant women [20–23]. Studies have also shown that anti-CPS antibodies induced by maternal immunization with this vaccine were transferred transplacentally to infants and persisted through a minimum of 3 months of age [20, 21, 23, 24].

The World Health Organization has articulated a preference for a single-dose maternal regimen, while acknowledging that a 2-dose regimen may need to be considered and that additional doses in subsequent pregnancies should be investigated [13]. The current study (an extension of a previous trivalent GBS vaccine trial in nonpregnant women [22]) was conducted to evaluate the safety and immunogenicity of a second dose of the trivalent GBS vaccine administered 4–6 years after the initial dose.

METHODS

Study Design, Participants, and Vaccines

In a previous study (NCT01150123; parent study) aimed at selecting trivalent GBS vaccine formulations and schedules for potential use in pregnant and nonpregnant women, nonpregnant women were randomized to receive 1 or 2 injections (1 month apart) of 2 vaccine antigen dosages (5 or 20 µg), either nonadjuvanted, aluminum hydroxide (alum)-adjuvanted or adjuvanted with MF59 (half- or full-dosage; Figure 1) [22, 25]. The study included a placebo group as the comparator.

The current phase 2 extension study was a nonrandomized, controlled, open-label study with 6 parallel groups. Healthy, nonpregnant women aged 22–46 years who had received a single injection of any of the 5-µg formulations (nonadjuvanted, alum-adjuvanted, half- or full-dosage MF59-adjuvanted) or placebo 4–6 years earlier in the parent study were enrolled. To ensure an adequate number of controls, women who were not part of the parent study and never received a GBS vaccine (naïve group) were also enrolled (Figure 1). Inclusion and exclusion criteria are detailed in the Supplementary Methods.

Women in each of the 6 groups received a single injection of nonadjuvanted trivalent GBS vaccine containing CRM197-conjugated GBS CPS of serotypes Ia, Ib, and III (5 µg each). This constituted a second dose for 4 groups (all previously GBS-vaccinated in the parent study: prior GBS groups) and a first dose for the placebo group from the parent study (prior placebo group) and the newly enrolled naïve group (Figure 1). The vaccine was administered intramuscularly in the deltoid muscle of the nondominant arm.

The study was conducted at the Center for Vaccinology, Ghent University Hospital, Belgium, between March 2016 and November 2016 according to the principles of Good Clinical Practice, the Declaration of Helsinki, and applicable regulations. The Ghent University Hospital Commission for Medical Ethics approved the protocol and informed consent form. Each participant provided written informed consent before enrollment.

Reactogenicity and Safety Assessment

Women received diary cards to record solicited adverse events (AEs) that occurred on days 1–7. Unsolicited AEs that occurred on days 1–31 were also recorded. On days 15, 121, and 181
(study end), participants received safety follow-up calls. The intensity of AEs was graded as mild, moderate, or severe. Serious AEs (SAEs), medically attended AEs, and AEs that led to withdrawal were recorded throughout the study (days 1–181). The investigators assessed the relationship of all unsolicited AEs and SAEs to vaccination.

Immunogenicity Assessment

Blood samples were drawn prevaccination (day 1) and 30 and 60 days postvaccination (days 31 and 61; Figure 1). Sera were stored frozen (less than or equal to −18 °C) until analysis. Anti-GBS CPS immunoglobulin G antibodies for serotypes Ia, Ib, and III were measured by ELISA using a multiplex immunoassay, instead of the per protocol planned enzyme-linked immunosorbent assay (ELISA) because of unavailability of the ELISA. The multiplex immunoassay has been described elsewhere and is summarized in the Supplementary Materials. The assay's lower limits of quantitation (LLQs) were 0.233 µg/mL (Ia), 0.155 µg/mL (Ib), and 0.293 µg/mL (III). Serum samples for the baseline prevaccination time point of the parent study, previously analyzed by ELISA, were retested with the multiplex immunoassay to allow comparison with the concentrations obtained for the samples from the extension study.

Statistical Analyses

The sample size was determined by the number of women from the parent study who agreed to participate in the extension study, plus 20 women who did not participate in the parent study.

The safety analysis was performed on the safety set, that is, all women vaccinated in the extension study and who provided postvaccination safety data. The primary safety endpoint was to assess the percentages of women who reported AEs over the time periods described above.

The immunogenicity analysis was performed on the per protocol immunogenicity set (PPS), that is, all women who had correctly received the study vaccine in the parent study (prior GBS and prior placebo groups) and received the study vaccine dose, complied with protocol-defined procedures, and had immunogenicity data available for at least 1 time point in the extension study. The primary immunogenicity endpoint was to assess the percentage of women who reached a range of arbitrary, prespecified serotype-specific anti-GBS concentration thresholds (0.5–8 µg/mL) 60 days postvaccination, reflecting the average time between third-trimester maternal vaccination and delivery. Secondary immunogenicity endpoints included the evaluation of percentages of women who reached these thresholds 30 days post-GBS vaccination and 30/60 days post-GBS vaccination by baseline serotype-specific anti-GBS concentrations (<LLQ). Baseline referred to the prevaccination time point in the parent study for the prior GBS and prior placebo groups and to the prevaccination time point (day 1) in the extension study for the naïve group (Figure 1). We computed percentages and 2-sided 95% Clopper-Pearson confidence intervals (CIs) for each group and different pooled groups (nonadjuvanted plus alum-adjuvanted, half- plus full-dose MF59-adjuvanted, any prior GBS, and prior placebo plus naïve; Figure 1). Differences in percentages between the pooled prior GBS and pooled no prior GBS groups were calculated with 2-sided 95% CIs using the Miettinen and Nurminen method. Reverse cumulative distribution curves of anti-GBS concentrations were generated.

We also assessed serotype-specific geometric mean concentrations (GMCs) and within-subject geometric mean ratios (GMRs) 30 and 60 days postvaccination in all women and by baseline serotype-specific anti-GBS concentration (<LLQ or ≥LLQ). GMRs were determined relative to parent study baseline levels and relative to day 1 levels in the extension study. For GMC calculations, antibody concentrations <LLQ were given an arbitrary value of half the LLQ. Adjusted GMCs with 2-sided 95% CIs were calculated from log_{10}-transformed antibody concentrations with an analysis of covariance model with vaccine group as the qualitative factor and log_{10}-transformed baseline concentration as the covariate. Baseline and day 1 GMCs were calculated using an analysis of variance (ANOVA) model. GMCs with 2-sided 95% CIs were calculated from the log_{10}-transformed within-subject ratios of antibody concentrations (postvaccination/prevaccination) using an ANOVA model with vaccine group as the qualitative factor.

The protocol included an assessment of antipneumococcal antibody levels to evaluate if the CRM197 carrier–lowered preexisting antipneumococcal titers. This analysis was canceled because no such interference was observed for other CRM197-conjugated vaccines [27, 28].

RESULTS

Study Participants

A total of 80 women were enrolled in the extension study: 53 who had received 1 dose of the trivalent GBS vaccine 4–6 years earlier in the parent study (prior GBS groups) and 27 not previously GBS-vaccinated (no prior GBS groups), 6 of whom had received placebo in the parent study (prior placebo group) and 21 newly enrolled (naïve group). All enrolled women received 1 dose of trivalent GBS vaccine and completed the study (Figure 2). The PPS comprised 79 participants (Figure 2). Baseline characteristics and time between parent and extension study vaccinations were similar across groups (Table 1).

Reactogenicity and Safety

Injection site pain was the most common solicited local AE (>50% of women across groups). The most frequent systemic AEs were fatigue and headache (Table 2). Most solicited AEs were mild or moderate. No fever was reported. Across groups,
29%–67% of women reported unsolicited AEs within 31 days postvaccination. Four women experienced unsolicited AEs considered as at least possibly related to vaccination: 2 in the prior GBS groups (upper respiratory tract infection, hot flush) and 2 in the no prior GBS groups (infection site erythema, nasal congestion; Table 2). These AEs were mild and resolved without

Table 1. Demographic Characteristics at Enrollment: All Enrolled Set

| Parent Study Vaccine | Prior GBS | Prior Placebo | Naïve |
|----------------------|-----------|-------------|-------|
| Extension Study Vaccine | GBS No Adj | GBS Alum | GBS MF59 Full | GBS MF59 Half | Placebo | Not Applicable |
| GBS 5 μg No Adj N = 14 | GBS 5 μg Alum N = 14 | GBS 5 μg MF59 Full N = 10 | GBS 5 μg MF59 Half N = 15 | Placebo N = 6 | GBS 5 μg No Adj N = 21 |
| N completed | N = 14 | N = 14 | N = 10 | N = 15 | N = 6 | N = 21 |

Mean age ± SD, y: 32.4 ± 6.9, 31.6 ± 6.1, 29.7 ± 6.2, 28.9 ± 4.7, 28.3 ± 6.4, 29.2 ± 7.0
Caucasian ethnicity, n: 14, 14, 10, 15, 6, 21
Mean weight ± SD, kg: 73.1 ± 12.5, 63.4 ± 6.9, 67.6 ± 12.3, 62.9 ± 8.2, 62.5 ± 7.9, 62.8 ± 6.4
Mean height ± SD, cm: 168.9 ± 5.4, 166.1 ± 4.9, 170.5 ± 5.1, 164.4 ± 7.1, 163.3 ± 5.0, 169.0 ± 6.0
Mean body mass index ± SD, kg/m²: 25.7 ± 4.6, 23.0 ± 2.6, 23.2 ± 3.4, 23.3 ± 2.9, 23.5 ± 4.0, 22.0 ± 2.2
Mean time between parent and extension study vaccination ± SD, y: 5.8 ± 0.06, 5.8 ± 0.05, 5.5 ± 0.05, 5.5 ± 0.06, 5.7 ± 0.13, NA

Abbreviations: alum, aluminum hydroxide-adjuvanted formulation; GBS, group B streptococcus; MF59 full, full-dosage MF59-adjuvanted formulation; MF59 half, half-dosage MF59-adjuvanted formulation; N, number of enrolled women in each group; n, number of women in the specified category; no adj, nonadjuvanted formulation; SD, standard deviation.
sequelae. Throughout the study and across groups, 0%–47% of women reported unsolicited AEs that needed medical attention; none were deemed related to vaccination (Table 2). No SAEs were reported during the study, and no women withdrew due to AEs.

**Immunogenicity**

All women in the PPS who had been vaccinated with trivalent GBS vaccine 4–6 years earlier had serotype-specific anti-GBS concentrations that exceeded an arbitrary threshold of 1 µg/mL for serotypes Ia, Ib, and III 60 days after receiving their second GBS vaccine dose in the extension study (Table 3, Supplementary Table 1). In addition, 92%–98% of women in the pooled prior GBS group reached an arbitrary threshold of 8 µg/mL 60 days postdose 2 across serotypes. This was substantially higher than percentages that achieved that threshold 60 days postdose 1 among previously non-GBS-vaccinated women (36%–56%; Table 3). Similar trends were noted across the other thresholds and for the 30 days postvaccination time point, with higher percentages reaching these thresholds after a second rather than after a first dose (Supplementary Table 1). The GBS vaccine formulation used in the parent study did not appear to have an observable impact on these percentages (Supplementary Table 1). Reverse cumulative distribution curves confirm these results (Figure 3, Supplementary Figure 1).

For each serotype, anti-GBS GMCs 30/60 days postdose 2 (prior GBS groups) were substantially higher than baseline GMCs (approximately 200- to 600-fold) and GMCs 30/60 days postdose 1 in the no prior GBS group (approximately 5- to 30-fold; Figure 4, Supplementary Table 2). No significant differences in postdose 2 antibody GMCs were observed between the prior GBS groups based on the vaccine formulation received.

### Table 2. Number (%) of Participants Reporting Adverse Events Postvaccination in the Extension Study: Safety Set

| Parent Study Vaccine | Extension Study Vaccine | Prior GBS | Prior Placebo | Naive |
|----------------------|-------------------------|-----------|--------------|-------|
| GBS No Adj           | GBS No Adj              | N = 14    |              |       |
| GBS Alum             | GBS No Adj              | N = 14    |              |       |
| GBS MF59 Full        | GBS No Adj              | N = 10    |              |       |
| GBS MF59 Half        | GBS No Adj              | N = 15    |              |       |
| Placebo              | GBS No Adj              | N = 5     |              |       |
| Not Applicable       | GBS No Adj              | N = 21    |              |       |

*Includes possibly related and probably related AEs.*
as the first dose in the parent study (Supplementary Table 2). Significant increases (approximately 20- to 70-fold) in antibody GMCs compared to baseline were also observed for each serotype 30/60 days postdose 1 in the no prior GBS groups (Figure 4, Supplementary Table 2).

In previous studies, women with undetectable prevaccination concentrations mounted a less robust response to GBS vaccination [20–23, 29]. We therefore assessed the anti-GBS response based on whether women had detectable or undetectable antibody concentrations at baseline (ie, above or below the serotype-specific LLQs before the first GBS vaccine dose). Most women across groups included in our analysis had baseline antibody concentrations <LLQs (49/77 [64%] for Ia, 59/77 [77%] for Ib, and 62/77 [81%] for III). Also, 90%–98% of women with undetectable baseline concentrations in the pooled prior GBS group reached the 8-µg/mL threshold 60 days postdose 1 (Table 3), with similar percentages for the other thresholds and 30 days postdose 2 (Supplementary Table 1). Previously GBS-vaccinated women with undetectable baseline antibody concentrations showed higher antibody GMCs 30/60 days postdose 2 vs baseline and vs GMCs 30/60 days postdose 1 in previously non–GBS-vaccinated women (approximately 7- to 70-fold; Figure 4, Supplementary Table 3). Among the small subset of women with detectable baseline antibody concentrations, serotype-specific anti-GBS GMCs were high 30/60 days postdose 1 in the no prior GBS groups and postdose 2 in the prior GBS groups, without consistent differences between postdose 1 and postdose 2 responses (Supplementary Table 3).

DISCUSSION
We are the first to evaluate the safety and immunogenicity of a second dose of the investigational trivalent GBS vaccine given at an interval after the first dose close to the average interpregnancy interval in some populations [30, 31]. Our study did not reveal any tolerability or safety concerns of a second dose given 4–6 years postfirst dose. The second dose elicited a robust immune response, particularly in women with undetectable antibody levels prefirst dose. In these women, a second vaccine dose induced more robust responses than a first dose.

While an inverse relation between maternal serotype-specific anti-GBS levels and the risk of invasive GBS disease in young infants has been demonstrated [6, 14–19], there is no established serological correlate of protection against invasive GBS disease in young infants. The absence of a standardized anti-GBS immunoassay complicates the establishment of a correlate of

### Table 3. Percentages of Women With Serotype-specific Anti-group B Streptococcus Antibody Concentrations ≥1 µg/mL and ≥8 µg/mL in the Pooled Groups and Group Differences 60 Days Postvaccination, per Protocol Immunogenicity Set

| GBS serotype | No Prior GBS | Prior GBS | Abbreviation | N | N | N | N | Group Difference
|--------------|-------------|-----------|--------------|---|---|---|---|------------------|
| Ia           |             |           |              |   |   |   |   | Prior GBS % (95% CI)
| All          | N = 27      | N = 24    | N = 51       | N = 25 |   |   |   |   |   |
| % ≥1 µg/mL   | 100 (87.2, 100) | 100 (85.8, 100) | 100 (93.0, 100) | 92 (74.0, 99.0) | 8 (0.5, 25.1) |
| % ≥8 µg/mL   | 96 (78.9, 99.9) | 98 (86.9, 100) | 56 (34.9, 76.6) | 42 (23.8, 61.4) |   |   |   |   |   |
| <LLQ         | N = 17      | N = 16    | N = 33       | N = 15 |   |   |   |   |   |
| % ≥1 µg/mL   | 100 (80.5, 100) | 100 (79.4, 100) | 100 (89.4, 100) | 93 (68.1, 99.8) | 7 (−4.5, 30.1) |
| % ≥8 µg/mL   | 94 (69.8, 99.8) | 97 (84.2, 99.9) | 47 (21.3, 73.4) | 50 (25.4, 73.0) |   |   |   |   |   |
| Ib           |             |           |              |   |   |   |   | Prior GBS % (95% CI)
| All          | N = 27      | N = 25    | N = 52       | N = 25 |   |   |   |   |   |
| % ≥1 µg/mL   | 100 (87.2, 100) | 100 (86.3, 100) | 100 (93.2, 100) | 64 (42.5, 82.0) | 36 (20.2, 55.6) |
| % ≥8 µg/mL   | 89 (70.8, 97.6) | 96 (79.6, 99.9) | 92 (81.5, 97.9) | 36 (18.0, 57.5) | 56 (34.8, 73.5) |
| <LLQ         | N = 19      | N = 22    | N = 41       | N = 18 |   |   |   |   |   |
| % ≥1 µg/mL   | 100 (82.4, 100) | 100 (84.6, 100) | 100 (91.4, 100) | 50 (26.0, 74.0) | 50 (28.9, 71.1) |
| % ≥8 µg/mL   | 84 (60.4, 96.6) | 95 (77.2, 99.9) | 90 (76.9, 97.3) | 17 (3.6, 41.4) | 74 (48.6, 87.3) |
| III          |             |           |              |   |   |   |   | Prior GBS % (95% CI)
| All          | N = 26      | N = 25    | N = 51       | N = 25 |   |   |   |   |   |
| % ≥1 µg/mL   | 100 (86.8, 100) | 100 (86.3, 100) | 100 (93.0, 100) | 68 (46.5, 85.1) | 32 (17.1, 51.7) |
| % ≥8 µg/mL   | 98 (80.4, 99.9) | 100 (86.3, 100) | 98 (89.6, 100) | 48 (27.8, 68.7) | 50 (30.7, 68.4) |
| <LLQ         | N = 22      | N = 20    | N = 42       | N = 19 |   |   |   |   |   |
| % ≥1 µg/mL   | 100 (84.6, 100) | 100 (83.2, 100) | 100 (91.6, 100) | 58 (33.5, 79.7) | 42 (23.0, 63.9) |
| % ≥8 µg/mL   | 95 (77.2, 99.9) | 100 (83.2, 100) | 98 (87.4, 99.9) | 37 (16.3, 61.6) | 61 (37.7, 79.1) |

Abbreviations: CI, confidence interval; GBS, group B streptococcus; LLQ, lower limit of quantitation; N, number of women with available results in each group.

See Figure 1 for group names. Analyses on all women regardless of their baseline LLQ status (“all”) and on women with baseline serotype-specific anti-GBS antibody concentrations below the LLQs (“<LLQ”). Baseline refers to the prevaccination time point in the parent study for the prior GBS and prior placebo groups and to the prevaccination time point in the current extension study (day 1) for the naïve group.
We therefore assessed several arbitrary thresholds between 0.5 and 8 µg/mL (close to the range of proposed seroprotection thresholds for other assays [1–10 µg/mL] [11, 14, 15, 17–19, 32–34]). Nearly all women reached the 8-µg/mL threshold after a second dose regardless of their baseline antibody levels, while a notably smaller proportion (≤50% 60 days postvaccination) reached this threshold after a single dose among women with undetectable baseline antibody levels.

Few studies have assessed the response to 2 GBS vaccine doses [11, 22, 23, 29]. The parent study compared groups receiving...
Figure 4. Serotype-specific geometric mean anti-GBS antibody concentrations at different time points for the pooled groups (per protocol immunogenicity set). See Figure 1 for group names. Analyses on all women regardless of their baseline lower limit of quantitation (LLQ) status (“all,” left side) and on women with baseline serotype-specific anti-GBS antibody concentrations below the LLQs (“<LLQ,” right side). Baseline refers to the prevaccination time point in the parent study for the prior GBS and prior placebo groups and to the prevaccination time point in the extension study (day 1) for the naïve group. Antibody concentrations <LLQ were given an arbitrary value of half the LLQ. Day 1 refers to the prevaccination time point in the extension study for all groups. Confidence intervals are depicted as error bars. Abbreviations: GBS, group B streptococcus; GMC, geometric mean concentration; LLQ, lower limit of quantitation; N, maximum number of women with available results across time points.
One or 2 doses (administered 1 month apart) of various trivalent GBS vaccine formulations and found no clear differences across groups in serotype-specific antibody GMCs postvaccination [22]. A second dose did not improve the immune response in women with undetectable baseline antibody levels, although subgroup sizes were small, making it difficult to draw definite conclusions [22]. A study in South Africa compared immune responses 1 month postfirst and postsecond dose in women receiving a 2-dose trivalent GBS vaccine regimen with doses spaced 1 month apart. No increases in antibody GMCs were observed after a second dose. No analysis by baseline LLQ status was performed [23]. A beneficial effect of a second dose given 21 months after first vaccination was demonstrated in a phase 1 study with a monovalent GBS serotype III CPS tetanus toxoid conjugate vaccine (GBS III-TT) [29]. While postdose 2 serotype III-specific antibody GMCs were similar to postdose 1 GMCs in the overall study population, in the small subset of participants (n = 8) with undetectable prevaccination antibody concentrations, the serotype III-specific GMC postdose 2 was 3-fold higher than postdose 1 [29]. This is consistent with what was observed in our study, where the subset of participants with undetectable baseline antibody levels was greater (64%–81% vs 22% in the GBS III-TT study). These results indicate that women with undetectable prevaccination anti-GBS concentrations, previously shown to mount a less robust response to women with undetectable prevaccination anti-GBS concentrations, 22% in the GBS III-TT study). These results indicate that women with undetectable prevaccination anti-GBS concentrations, previously shown to mount a less robust response to
GBS vaccination [20–23, 29], may benefit from receiving a sufficiently spaced second GBS vaccine dose.

In the parent study, no additional benefit from adding alum or MF59 adjuvant could be shown [22]. Likewise, we observed no major effect of the vaccine formulation used in the parent study on the immune response after a second dose given 4–6 years later. The small sample size warrants caution when interpreting these results.

As GBS vaccination is intended for use during pregnancy, a favorable reactogenicity and safety profile is essential [13]. In our study, severe solicited AEs were rare and no fever or SAEs were reported. Four women experienced mild, nonserious unsolicited AEs considered as possibly or probably vaccination-related. While the number of participants per group was too small for conclusive group comparisons, no obvious differences were observed between the rates of solicited or unsolicited AEs after a second or first trivalent GBS vaccine dose. Likewise, reactogenicity after a second dose in the extension study was similar to that of 1 or 2 doses of the 5-µg nonadjuvanted formulation in the parent study [22].

Aside from the small sample size and descriptive nature of the analyses, our study has other limitations. We only assessed a 4- to 6-year interval between doses, which is longer than the interpregnancy interval in many settings [29, 30, 34], and we did not establish the minimum interval required for the second vaccine dose to mount a robust immune response. No information was collected on whether women became GBS-colonized/serotype-specific anti-GBS levels and protection [6, 14–19], a second dose could improve the immune response in the parent study [22]. Likewise, we observed a 4- to 6-year interval between doses, which is longer than the interpregnancy interval in many settings [29, 30, 34], and we did not establish the minimum interval required for the second vaccine dose to mount a robust immune response. No information was collected on whether women became GBS-colonized/serotype-specific anti-GBS levels and protection [6, 14–19], a second dose could improve the immune response in infants whose mothers were seronegative before initial vaccination.

In summary, our results show that a second dose of the investigational trivalent GBS vaccine administered 4–6 years after a first dose had a favorable safety profile and was immunogenic, regardless of baseline antibody concentrations. A 2-dose schedule with doses spaced far enough apart may be beneficial for women with very low preexisting antibody concentrations.

**Supplementary Data**

Supplementary materials are available at Clinical Infectious Diseases online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

**Notes**

*Author contributions.* G. L. R., Z. B., I. M., A. K., S. C., and K. S. designed the study. G. L. R., Z. B., C. M., A. A., F. D. B., G. B., I. M., and A. K. acquired the data. G. L. R., Z. B., L. G., G. B., I. M., A. K., B. C., and O. H. analyzed the data. G. L. R., C. M., A. A., F. D. B., A. K., and B. C. contributed to the conduct of the study. All authors reviewed and revised the manuscript and approved the final manuscript as submitted.

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*Potential conflicts of interest.* Z. B., L. G., G. B., I. M., A. K., B. C., and O. H. are employees of the GSK group of companies. I. M., A. K., B. C., and O. H. hold shares in the GSK group of companies. K. S. was an employee of Novartis Vaccines Division and the GSK group of companies. S. C. was an employee of Novartis Vaccines Division and the GSK group of companies and is currently an employee of Novartis and holds shares in the GSK group of companies and Novartis. I. M. is listed as inventor on patents owned by the GSK group of companies. G. L. R. and A. A. report no potential conflicts of interest.

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*Supplementary materials* are available at Clinical Infectious Diseases online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

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References
1. Edmond KM, Kort술oidaki C, Scott S, et al. Group B streptococcal disease in infants aged younger than 3 months: systematic review and meta-analysis. Lancet 2012; 379:547–56.
2. Madrid L, Seale AC, Kohli-Lynch M, et al; Infant GBS Disease Investigator Group. Infant group B streptococcal disease incidence and serotypes worldwide: systematic review and meta-analyses. Clin Infect Dis 2017; 65:160–72.
3. Seale AC, Bianchi Jassir F, Russell NJ, et al. Estimates of the burden of group B streptococcal disease worldwide for pregnant women, stillbirths, and children. Clin Infect Dis 2017; 65:200–19.
4. Seale AC, Blencowe H, Bianchi-Jassir F, et al. Stillbirth with group B streptococcal disease worldwide: systematic review and meta-analyses. Clin Infect Dis 2017; 65:125–32.
5. Bianchi-Jassir F, Seale AC, Kohli-Lynch M, et al. Preterm birth associated with group B streptococcal maternal colonization worldwide: systematic review and meta-analyses. Clin Infect Dis 2017; 65:100–11.
6. Kobayashi M, Vekemans J, Baker CJ, Ratner AJ, Le Doare K, Schrag SJ. Group B streptococcal vaccine development: present status and future considerations, with emphasis on perspectives for low and middle income countries. F1000Res 2016; 5:2355.
7. Russell NJ, Seale AC, O’Driscoll M, et al; GBS Maternal Colonization Investigator Group. Maternal colonization with group B streptococcus and serotype distribution worldwide: systematic review and meta-analyses. Clin Infect Dis 2017; 65:100–11.
8. Le Doare K, O’Driscoll M, Turner K, et al; GBS Intrapartum Antibiotic Investigator Group. Intrapartum antibiotic chemoprophylaxis policies for the prevention of group B streptococcal disease worldwide: systematic review. Clin Infect Dis 2017; 65:143–51.
9. Schrag SJ, Verani JR. Intrapartum antibiotic prophylaxis for the prevention of perinatal group B streptococcal disease: experience in the United States and implications for a potential group B streptococcus vaccine. Vaccine 2013; 31(Suppl 4):D20–6.
10. World Health Organization. WHO recommendations for prevention and treatment of maternal peripartum infections. Available at: https://www.who.int/reproductivehealth/publications/maternal_perinatal_health/peripartum-infections-guidelines/en/. Accessed 19 November 2018.
11. Dzanibe S, Madhi SA. Systematic review of the clinical development of group B streptococcal serotype-specific capsular polysaccharide-based vaccines. Expert Rev Vaccines 2018; 17:635–51.
12. Heath PT. Status of vaccine research and development of vaccines for GBS. Vaccine 2016; 34:2876–9.
13. World Health Organization. WHO preferred product characteristics for group B streptococcal vaccines. Available at: http://apps.who.int/iris/bitstream/handle/10665/258703/WHO-IVB-17.09-eng.pdf?sequence=1. Accessed 19 November 2018.
14. Baker CJ, Carey VJ, Renc MA, et al. Maternal antibody at delivery protects neonates from early-onset group B streptococcal disease. J Infect Dis 2014; 209:781–8.
15. Baker CJ, Edwards MS, Kasper DL. Role of antibody to native type III polysaccharide of group B streptococcus in infant infection. Pediatrics 1981; 68:544–9.
16. Baker CJ, Kasper DL. Correlation of maternal antibody deficiency with susceptibility to neonatal group B streptococcal infection. N Engl J Med 1976; 294:753–6.
17. Dangor Z, Kwarata G, Izu A, Lala SG, Madhi SA. Review on the association of group B streptococcus capsular antibody and protection against invasive disease in infants. Expert Rev Vaccines 2015; 14:135–49.
18. Lin FY, Philips JB 3rd, Azimi PH, et al. Level of maternal antibody required to protect neonates against early-onset disease caused by group B streptococcus type Ia: a multicenter, seroepidemiology study. J Infect Dis 2001; 184:1022–8.
19. Lin FY, Weisman LE, Azimi PH, et al. Level of maternal IgG anti-group B streptococcus type III antibody correlated with protection of neonates against early-onset disease caused by this pathogen. J Infect Dis 2004; 190:928–34.
20. Donders GG, Halperin SA, Devlieger R, et al. Maternal immunization with an investigational trivalent group B streptococcal vaccine: a randomized controlled trial. Obstet Gynecol 2016; 127:213–21.
21. Heyderman RS, Madhi SA, French N, et al. Group B streptococcus vaccination in pregnant women with or without HIV in Africa: a non-randomised phase 2, open-label, multicentre trial. Lancet Infect Dis 2016; 16:536–55.
22. Leroux-Roels G, Maes C, Willekens J, et al. A randomized, observer-blind phase Ib study to identify formulations and vaccine schedules of a trivalent Group B streptococcal vaccine for use in non-pregnant and pregnant women. Vaccine 2016; 34:1786–91.
23. Madhi SA, Cutland CL, Jose L, et al. Safety and immunogenicity of an investigational maternal trivalent group B streptococcal vaccine in healthy women and their infants: a randomised phase 1b/2 trial. Lancet Infect Dis 2016; 16:923–34.
24. Madhi SA, Koen A, Cutland CL, et al. Antibody kinetics and response to routine vaccinations in infants born to women who received an investigational trivalent group B streptococcal polysaccharide CRM197-conjugate vaccine during pregnancy. Clin Infect Dis 2017; 65:1897–904.
25. Schultz V, D’Agosto V, Wack A, Novicki D, Zorn J, Hennig R. Safety of MF59 adjuvant. Vaccine 2008; 26:3209–22.
26. Buñi G, Galletti B, Stella M, et al. Novel multiplex immunoassays for Quantification of IgG against group B Streptococcus capsular polysaccharides in human sera. mSphere 2019; 4:e00273-19.
27. McVernon J, MacLennan J, Clutterbuck E, Buttery J, Moxon ER. Effect of infant immunisation with meningococcus serogroup C-CRM(197) conjugate vaccine on diphtheria immunity and reactogenicity in pre-school aged children. Vaccine 2003; 21:2573–9.
28. Bröker M, Berti E, Schneider J, Vojtek I. Polysaccharide conjugate vaccine protein carriers as a “neglected valency” — Potential and limitations. Vaccine 2017; 35:3286–94.
29. Paolletti LC, Renc MA, Kasper DL, Molrine D, Ambrosino D, Baker CJ. Effects of alum adjuvant or a booster dose on immunogenicity during clinical trials of group B streptococcal type III conjugate vaccines. Infect Immun 2001; 69:6969–703.
30. Gemmill A, Lindberg LD. Short interpregnancy intervals in the United States. Obstet Gynecol 2013; 122:64–71.
31. Kreyenfeld M, Geisler E, Castro-Martin T, et al. Social policies, separation, and second birth spacing in Western Europe. Demogr Res 2017; 37:1245–74.
32. Dangor Z, Lala SG, Kwarata G, Madhi SA. Group B streptococcus: developing a correlate of protection for a vaccine against neonatal infections. Curr Opin Infect Dis 2016; 29:262–7.
33. Dangor Z, Kwarata G, Izu A, et al. Correlates of protection of serotype-specific capsular antibody and invasive group B streptococcal disease in South African infants. Vaccine 2015; 33:6793–9.
34. Fabbrini M, Rigat F, Tuscano G, et al. Functional activity of maternal and cord antibodies elicited by an investigational group B streptococcal trivalent glycoconjugate vaccine in pregnant women. J Infect Dis 2018; 76:449–56.
35. Migini LE, Carroló G, Betran AP, et al. Interpregnancy interval and perinatal outcomes across Latin America from 1990 to 2009: a large multi-country study. BJOG 2016; 123:730–7.