Maize Germplasm Characterization Using Principal Component and Cluster Analysis

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ABSTRACT

In Ethiopian Biodiversity Institute Gene bank, huge collections of maize germplasm are not yet characterized for the magnitude of genetic variability from each other. Though knowing the contribution of individual characters is essential to focus on particular characters in cultivar development. Hence, this experiment was conducted on 92 maize accessions which were not yet characterized and 2 local checks to estimate the magnitude of genetic diversity among the genotypes and to identify the major agro-morphological characters contributing for the observed variations. The experiment was arranged in Augmented Design in seven blocks at Arsi-Negele in 2016 main cropping season. The characters used for analysis were days to flowering, plant height, ear height, ear per plant, days to maturity, ear length, kernel rows per ear, thousand grain weight and yield per plot. The 94 genotypes were grouped into four clusters where cluster I, II, III and IV comprised 30, 21, 23 and 20 genotypes respectively. Early matured and short genotypes were grouped in cluster IV, late matured in cluster II and high yielding and tall genotypes in cluster I. The principal component analysis indicated that the first principal component (PC1) had an eigenvalue of 4.4 and reflects 48.85% of the total variation, this represents the equivalent of two individual variables and the two variables that weighted higher than the other variables are plant height and ear length. The second principal component (PC2) was recorded eigenvalue of 1.63 and maintaining 18.11% of the total variation and related to diversity among genotype due to ear per plant (EPP). Moreover, principal components 3 to 9 were showed more than one eigenvalue, thus they represent equivalent of one individual variable each accounted for 0.98%, 0.78%, 0.68%, 0.35%, 0.15%, 0.03% and 0% respectively toward the variation observed among genotypes. The result ensures the existence of high genetic divergence among the studied maize genotypes.
INTRODUCTION

Maize (Zea mays L.) is one of the popular crops grown in the world, ranking second to wheat and followed by rice [1]. It occupies an important position in the world economy as food, feed, and industrial grain crop. It is a staple food for several million people in the developing world where they derive their protein and calorie requirements from it. Maize is among the leading cereal crops selected to achieve food self-sufficiency in Ethiopia [2]. Although, improved cultivars have been largely included in the national extension package, the national average yield of maize is only 3.45 tons/ha [3], which is far below the world average of 5.5 tons/ha. In any crop, germplasm resource not only serves as a valuable source of useful genes but also provides a wide genetic variability. Bringing improvement over existing crop varieties is a continuous process in plant breeding. To achieve this objective, the breeder has to identify diverse parents having superior genetic variability for combining desirable characters. Hence, knowledge of sound genetic diversity is crucial for undertaking any recombination breeding program. Multivariate statistical techniques used to analyze multiple measurements on each individual and used in the analysis of genetic diversity. Among the multivariate techniques, principal component analysis (PCA) and cluster analysis had been shown to be very useful in selecting genotypes for breeding program that meet the objective of a plant breeder [4]. PCA may be used to reveal patterns and eliminate redundancy in data sets [5], as morphological and physiological variations routinely occur in crop species. Cluster analysis is commonly used to study genetic diversity and for forming core subset for grouping accessions with similar characteristic into homogenous category. Cluster analysis is frequently used to classify maize accessions and can be used by breeders and geneticists to identify subsets of accessions which have potential utility for specific breeding or genetic purposes [6]. Therefore, the objective of this study was aimed to estimate the magnitude of genetic diversity among the maize genotypes and to identify the major agromorphological characters contributing for the observed variations.

MATERIALS AND METHODS

The study was conducted during the year 2016 at the experimental field of Arsi-Negele, Oromia Regional State, Ethiopia. It is located at 7°21’N 38°42’E and at an elevation of 1940 m.a.s.l. It has a chromic and pellic vertisols with PH value of 5-7. The annual rainfall of the location is measured 915 mm with 27 ± 0.30°C mean daily temperature. 92 maize accessions obtained from Ethiopian Biodiversity Institute and two local checks named as check 1 and 2 were grown at farm site. The ninety two maize accessions without replication along with two replicated checks were arranged in augmented design. Individual plot size measured 9m × 1.5m with 4 rows planted at a spacing of 75 × 30cm. Recommended doses of fertilizers were applied. The other management operations were done timely and properly to raise the crop uniformly. Twenty randomly selected plants were used for recording observations on days to flowering, plant height, ear height, ear per plant, days to maturity, ear length and kernel rows per ear, thousand grain weight, and yield per plot. The data collected for all quantitative characters were subjected to analysis of basic statistics, correlation, cluster and principal component analysis using the software statistical package for the social sciences (SPSS) 16.0 package [7].

RESULTS

In the present study, genetic diversity was analyzed among 94 maize genotypes (Table 6) on the basis of 9 agromorphological characters. The results of descriptive analysis (Table 1), ear height (EH) was showed the highest
variation (35.52%) followed by number of ear per plant (30.39%). Conversely, the lowest variation was recorded from kernel rows per ear (6.01%) and days to maturity (9.61%).

Table 1. Basic statistics for various characters of maize genotypes.

| Characters                | Mean  | Minimum | Maximum | Range  | SD    | CV (%) |
|---------------------------|-------|---------|---------|--------|-------|--------|
| Days to flowering         | 107.00| 59.00   | 134.00  | 75.00  | 13.75 | 12.85  |
| Plant height (m)          | 2.23  | 1.10    | 3.04    | 1.94   | 0.41  | 18.36  |
| Ear height (m)            | 1.05  | 0.26    | 1.95    | 1.69   | 0.37  | 35.52  |
| Ear per plant             | 1.95  | 0.00    | 3.10    | 3.10   | 0.59  | 30.39  |
| Days to maturity          | 143.00| 95.00   | 170.00  | 75.00  | 13.75 | 9.61   |
| Ear length (cm)           | 15.13 | 10.70   | 19.00   | 8.30   | 1.80  | 11.90  |
| Kernel rows per ear       | 12.28 | 9.80    | 14.40   | 4.60   | 0.74  | 6.01   |
| Thousand grain weight (g) | 338.37| 196.00  | 504.00  | 308.00 | 54.47 | 16.10  |
| Yield (kg/plot)           | 5.78  | 3.37    | 7.69    | 4.32   | 0.95  | 16.45  |

SD: Standard deviation, CV: Coefficient of variation

Table 2. Phenotypic correlation coefficients for different traits on maize genotypes.

Simple correlation coefficients confirmed that yield per plot was recorded highly significant positive correlations among plant height, ear length, and thousand grain weight and maintained positive significant correlation among days to flower, ear height, days to maturity and kernel rows per ear. Likewise, ear per plant was recorded insignificant negative correlations with days to flowering, plant height and ear height (Table 2). Similarly, found highly significant positive correlation of grain yield with cob diameter and thousand kernels weight and significant positive correlation with plant height.

Table 2. Phenotypic correlation coefficients for different traits on maize genotypes.

| DF  | PH     | EH     | EPP    | DM     | EL     | KRPE   | TGW    | YPP    |
|-----|--------|--------|--------|--------|--------|--------|--------|--------|
| DF  | 1.00   | 0.59218** | 0.59323** | -0.092 | 1.00000** | 0.43338** | 0.18989 | 0.22982* | 0.25984* |
| PH  | 1.00   | 0.96212** | 0.909  | -0.009 | 0.59218** | 0.68246** | 0.37618* | 0.47818** | 0.41423** |
| EH  | 1.00   | -0.092 | 0.59323** | 0.62418** | 0.31869* | 0.41280** | 0.37949* |
| EPP | 1.00   | -0.09127 | 0.23183* | 0.13729 | 0.17099 | 0.25984* |
| DM  | 1.00   | -0.092 | 0.43338** | 0.18989 | 0.22982* | 0.25984* |
| EL  | 1.00   | -0.09127 | 0.42315** | 0.57258** | 0.45637** |
| KRPE| 1.00   | 0.59218** | 0.59323** | -0.092 | 1.00000** | 0.43338** | 0.18989 | 0.22982* | 0.25984* |

*Significant at P = 0.05, **Significant at P = 0.01. DF= Days to flowering, PH= Plant height, EH= Ear height, EPP= Ear per plant, DM=Days to maturity, EL= Ear length, KRPE= Kernel rows per ear, TGW= Thousand grain weight, and YPP= Yield per plot.
Principal component analysis

The nine components which had eigenvalues equal to or greater than one were retained as meaningful interpretation (Table 3). The principal component analysis indicated that the first principal component (PC1) had an eigenvalue of 4.4 and reflects 48.85% of the total variation (Table 3), this represents the equivalent of three individual variables and the three variables that weighted higher than the other variables are plant height (PH), days to maturity and ear length (EL). The second principal component (PC2) was recorded eigenvalue of 1.63 and maintaining 18.11% of the total variation and related to diversity among genotype due to ear per plant (EPP). Moreover, principal components 3 to 9 were showed more than one eigenvalues, thus they represent equivalent of one individual variable each accounted for 0.98%, 0.78%, 0.68%, 0.35%, 0.15%, 0.03% and 0% respectively towards the total variation (Table 4).

| Character | PC1   | PC2   | PC3   | PC4   | PC5   | PC6   | PC7   | PC8   | PC9   |
|-----------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| DF        | 0.36  | -0.43 | 0.07  | 0.29  | 0.30  | 0.00  | -0.04 | 0.00  | 0.71  |
| PH        | 0.43  | -0.09 | 0.06  | -0.27 | -0.38 | 0.22  | -0.06 | -0.73 | 0.00  |
| EH        | 0.41  | -0.13 | 0.07  | -0.25 | -0.43 | 0.33  | 0.07  | 0.67  | 0.00  |
| EPP       | 0.03  | 0.40  | 0.76  | 0.42  | -0.03 | 0.28  | 0.06  | -0.02 | 0.00  |
| DM        | 0.36  | -0.43 | 0.07  | 0.29  | 0.30  | 0.00  | -0.04 | 0.00  | -0.71 |
| EL        | 0.38  | 0.18  | 0.21  | -0.05 | -0.17 | -0.85 | 0.17  | 0.05  | 0.00  |
| KRPE      | 0.24  | 0.27  | 0.18  | -0.62 | 0.66  | 0.11  | 0.00  | 0.03  | 0.00  |
| TGW       | 0.32  | 0.44  | -0.31 | 0.23  | 0.00  | 0.01  | -0.73 | 0.09  | 0.00  |
| YPP       | 0.30  | 0.38  | -0.48 | 0.27  | 0.10  | 0.18  | 0.65  | -0.06 | 0.00  |

Table 4. The principal component of traits used for cluster analysis.

The PC3 showed high weights in ear per plant (EPP) and probably reflecting yield. The fifth principal component (PC5) kernel rows per ear (KRPE) had the largest weight, thus reflecting yield. The seventh principal component (PC7) was showed high value on yield per plot (YPP). Eighth principal component (PC8) had weighted high value of ear height (EH); this is probably reflecting the plant structure. Moreover, the ninth principal component (PC9) was recorded highest value on days to flowering (DF), thus reflecting flower development. In this study the principal component analysis was categorized the total variance into nine (9) principal components and contributing maximum towards the total diversity. Similarly, [9,10] reported important contribution of the first pcs in the total variability while studying various traits. Principal component analysis (PCA) is usually used in plant sciences for the reduction of variables and grouping of genotypes. Several authors suggested first principal component (PC) scores as input variables for the clustering process (Figure 1)[11].
Cluster analysis
Clustering pattern of maize accessions under this experiment reveals that the maize germplasm showed considerable genetic diversity among them by occupying four different clusters (Table 5). These maize germplasm were grouped based on mainly day to flowering, plant height, ear height, ear per plant, days to maturity, ear length, kernel rows per ear, thousand grain weights and yield as variables. Ninety four maize genotypes were grouped into 4 clusters based on various agro-morphological characters. Cluster I to IV were comprised 30, 21, 23 and 20 maize genotypes respectively (Table 6). Thus, Cluster I (Table 5) was maintained maximum plant height (2.42 m), ear
height (1.21 m), ear per plant (2.25), ear length (16.21 cm), kernel rows per ear (12.61) and yield (6.58 kg/plot). Cluster II was showed late days to flowering (120.9 days) and maturing nature (156.9 days). Cluster III also maintained higher yield (5.88 kg/plot). Moreover cluster IV was showed relatively early maturing characters (133.5 days) but had minimum values of plant height (1.84 m), ear height (0.74 m), ear length (13.34 cm), kernel row per ear (11.82), and low yield (4.5 kg/plot). Similarly, hierarchical cluster analysis has been suggested for classifying entries of germplasm collections based on degree of similarity and dissimilarity [12]. A combination of cluster and principal component analysis has been used to classify maize (Zea mays L.) Accessions [13].

The tree diagram comprising 4 main cluster groups and each of them further subdivided into sub-clusters (Figure 2). The information regarding association among various traits is an important part for the initiation of any breeding programme and gives good chance for the selection of superior genotypes having desirable traits simultaneously [14].

### Table 5.
Cluster means values for different agro- morphological characters of 94 genotypes.

| Cluster name | No. Of Genotypes | Name of accessions in each cluster |
|--------------|------------------|-----------------------------------|
| I            | 30               | ACC-9994, ACC-18108, ACC-15459   |
|              |                  | ACC-16226, ACC-18112, ACC-15460  |
|              |                  | ACC-16233, ACC-18121, ACC-241584 |
|              |                  | ACC-16570, ACC-16571, ACC-24297  |
|              |                  | ACC-1671, ACC-18106, ACC-15325   |
|              |                  | ACC-987, ACC-18127, ACC-237657   |
| II           | 21               | ACC-18112, ACC-18112, ACC-15460  |
|              |                  | ACC-18121, ACC-18121, ACC-241584 |
|              |                  | ACC-16570, ACC-16571, ACC-24297  |
|              |                  | ACC-1671, ACC-18106, ACC-15325   |
|              |                  | ACC-987, ACC-18127, ACC-237657   |
| III          | 23               | ACC-9993, ACC-18102, ACC-18112,  |
|              |                  | ACC-16023, ACC-16024, ACC-16582  |
|              |                  | ACC-98098, ACC-98098, ACC-237597 |
| IV           | 20               | ACC-18108, ACC-18112, ACC-18121  |
|              |                  | ACC-16570, ACC-16571, ACC-24297  |
|              |                  | ACC-1671, ACC-18106, ACC-15325   |
|              |                  | ACC-987, ACC-18127, ACC-237657   |

### Table 6.
Clustering pattern of the 94 accessions based on agro-morphological characters.
CONCLUSION

Different methods are available to assessing genetic variability among and within crop species. In the present study, principal component and cluster analysis techniques were employed to examine the amount of genetic
variability present in a set of 94 maize accessions. Thus, it can be inferred from the present investigation that high level of genetic variability was present in agronomic and morphological traits like days to flowering, plant height, ear height, ear per plant, days to maturity, ear length, kernel rows per ear, thousand grain weight and yield per plot in the tested germplasm. Therefore, promising maize genotypes with more genetic divergences were identified. The identification of high level of genetic variability during the current study could be indicated for germplasm characterization, conservation and further improvement in maize breeding program.

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