Capillary Blood Gas in Children Hospitalized Due to Influenza Predicts the Risk of Lower Respiratory Tract Infection

August Wrotek 1,2,* and Teresa Jackowska 1,2,*

1 Department of Pediatrics, Centre of Postgraduate Medical Education, Marymoncka 99/103, 01-813 Warsaw, Poland
2 Department of Pediatrics, Bielanski Hospital, Cegłowska 80, 01-809 Warsaw, Poland
* Correspondence: awrotek@cmkp.edu.pl (A.W.); tjackowska@cmkp.edu.pl (T.J.)

Abstract: Background: Influenza may impair respiratory exchange in the case of lower respiratory tract infections (LRTIs). Capillary blood gas (CBG) reflects arterial blood values but is a less invasive method than arterial blood sampling. We aimed to retrospectively verify the usefulness of CBG in pediatric influenza. Material and methods: CBG parameters (pH, pCO₂, pO₂, SatO₂) in laboratory confirmed influenza cases hospitalized in 2013–2020 were verified in terms of LRTI, chest X-ray (CXR) performance, radiologically confirmed pneumonia (CXR + Pneumonia), prolonged hospitalization, and intensive care transfer. A theoretical CBG-based model for CXR performance was created and the odds ratios were compared to the factual CXR performance. Results: Among 409 children (aged 13 days–17 years 3/12, median 31 months), the usefulness of CBG decreased with the age. The SatO₂ predicted the LRTI with AUC = 0.74 (95%CI: 0.62–0.86), AUC = 0.71 (0.61–0.82), and AUC = 0.602 (0.502–0.702) in children aged <6 months old (mo), 6–23 mo, 24–59 mo, respectively, while pO₂ revealed AUC = 0.73 (0.6–0.85), AUC = 0.67 (0.56–0.78), and AUC = 0.601 (0.501–0.702), respectively. The pCO₂ predicted the LRTI most precisely in children <6 months with AUC = 0.75 (0.63–0.87), yet not in older children. A high negative predictive value for CXR + Pneumonia was seen for SatO₂ < 6 mo (96.7%), SatO₂ 6–23 mo (89.6%), pO₂ < 6 mo (94.3%), pO₂ 6–23 mo (88.9%). The use of a CBG-driven CXR protocol (based on SatO₂ and pO₂) would decrease the odds of an unnecessary CXR in children <2 years old (yo) by 84.15% (74.5–90.14%) and 86.15% (66.46–94.28%), respectively. SatO₂ and pO₂ also predicted a prolonged hospitalization <6 mo AUC = 0.71 (0.59–0.83) and AUC = 0.73 (0.61–0.84), respectively, and in 6–23 mo AUC = 0.66 (0.54–0.78) and AUC = 0.63 (0.52–0.75), respectively. Conclusions: The CBG is useful mainly in children under two years, predicts the risk of LRTI, and can help exclude the risk of CXR + pneumonia. Children under six months of age represent the group that would benefit the most from CBG. A CBG-based protocol for the performance of CXR could significantly decrease the number of unnecessary CXRs.

Keywords: influenza; children; pneumonia; capillary blood gas; acidosis; hypercapnia

1. Introduction

Influenza has a significant impact on the pediatric population due to both the frequency and the severity of the disease. Estimates based on a systematic review of published data report 109.5 million influenza episodes annually only in children under five years of age [1–3]. Influenza undergoes seasonal fluctuations with regards to its frequency, although during the SARS-CoV-2 pandemic, the previously known seasonal patterns were altered in various regions, mainly due to nonpharmaceutical interventions [4,5]. The most important preventive measure, influenza vaccination, is globally available. Nonetheless, vaccine hesitancy and the generally low vaccine uptake impede the fight against the spread of influenza, thus facilitating the virus transmission [6,7]. Children play a pivotal role in the transmission of the virus, as their social activity combines household and community contacts, turning children into active influenza vectors [8,9]. Approximately 9% of the
cases are complicated with acute lower respiratory tract infections (ALRI) attributable to influenza, which each year translate to over 10.1 million cases, and around 870,000 influenza-related ALRI cases require hospitalizations, thus having a huge impact on the consumption of the health care system resources [1]. Roughly, one in 20 hospital admissions due to ALRI is estimated to be caused by the influenza virus, while prospectively collected data show that 3% of all pneumonia cases in infants is associated with influenza [1,10]. Influenza type A has been related to an increased risk of a life-threatening pneumonia in children (odds ratio = 2.55), but influenza B also carries a significant threat to the population, including the risk of a severe lower respiratory tract infection (LRTI) [11,12]. The huge pathogenic potential of influenza results not only from its virulence, but is enhanced with bacterial coinfections or secondary bacterial infections in close proximity after an influenza episode. Assessments in the pediatric group of patients reveal a bacterial codetection in as many as 80% of cases [13–15]. Gas exchange in the course of influenza may be compromised, and potential pathways involved in the regulation of the local immune response and maintenance of gas exchange include the alveolar epithelium (depletion of type I cells might result in serious sequelae), alveolar macrophages, and a number of inter- and intra-cellular crosstalk [16–20]. An altered gas exchange in severe cases may lead to a respiratory failure with the risk of intensive care and a potential fatal outcome [16]. In general estimates, approximately 4% of ALRI deaths in patients younger than five years old might be related to influenza [1]. Influenza may result in ALRI, which possibly affects the gas exchange, and the involvement of the lower respiratory tract is often suspected in hospitalized patients. Thus, a prompt diagnosis of lower respiratory tract involvement is crucial for adequate management, whereas an assessment of the patient’s gas exchange parameters might be helpful in the prediction of ALRI [21].

Arterial blood gas (ABG) sampling is a golden standard in the assessment of the patient’s gas exchange status, but this procedure carries the risk of complications, and as such, is performed almost exclusively in intensive care unit (ICU) settings, not in general pediatric wards [22]. Nonetheless, there is also a promising alternative for ABG, namely, the capillary blood gas (CBG) that measures the gas balance parameters in the peripheral blood taken from the fingertip, earlobe or heel. The CBG allows to indirectly assess the arterial blood parameters without the need for arterial blood sampling at the expense of a slightly lower accuracy, still, this option is much safer and easier to perform [22]. The CBG is also more reliable than venous blood gas (VBG), and as such might be used even in children with an impaired gas exchange [22].

The published data prove a satisfactory correlation between the CBG and the ABG in terms of pH and partial carbon dioxide pressure (pCO₂) and shows a slightly lower but still significant correlation regarding the partial oxygen pressure (pO₂) [22–25]. In addition, the CBG mirrors the ABG values closely, even in the presence of potential confounding factors. The studies cited above have been performed in the intensive care settings, and irrespectively of the patient’s severe condition, reflected by poor perfusion or hypothermia, showed a strong correlation, with the only exception for hypotension, which may distort the results of the pO₂ [22,24].

We sought to verify the usefulness of the CBG in children hospitalized due to a laboratory confirmed influenza (LCI) in predicting the risk of a lower respiratory tract involvement, the need for a chest radiograph, the presence of a radiologically confirmed pneumonia, a prolonged hospital stay, and an ICU admission. Moreover, a theoretical CBG-driven model for CXR qualification was created in order to calculate the odds ratios of an unnecessary CXR performance and missed pneumonia cases.

2. Materials and Methods

This was an observational retrospective study. Children hospitalized at the Department of Pediatrics at the Bielski Hospital, Warsaw, between January 2013 and December 2020 were eligible for the study. An electronic database of the medical charts was searched for the following final diagnoses of influenza, according to the International Classification
of Diseases, 10th Revision (ICD-10): J10 (or J11) with their extensions; while the J10 codes stand for influenza due to an identified influenza virus, and the J11 stand for an influenza due to an unidentified virus. In order to conduct as comprehensive research as possible, patients with J11 codes were also identified and their charts were revised for the existence of a laboratory confirmation of influenza, however, we included only laboratory confirmed cases. The diagnosis was made upon clinical signs/symptoms of influenza alongside a positive rapid influenza diagnostic test (RIDT) and/or a real-time polymerase chain reaction (RT-PCR), performed in a sample from a nasopharyngeal swab.

The exclusion criteria consisted of a previously known (or diagnosed during the hospitalization) immune deficiency, diabetes, a history of or an ongoing proliferative disease, hemodynamically significant heart disease, and cystic fibrosis. Patients discharged on parental request were also excluded, as the effect of the treatment was uncertain.

Only children in whom the CBG was performed were enrolled into the final analysis. The CBG was performed in each case immediately after the hospital admission; a sample of capillary blood was drawn after a local disinfection of a finger: a needle was used for an initial puncture and a CBG tube was promptly contacted with the punctured site. No prior warming of the site of incision had been performed. Each sample was verified against the presence of air bubbles and sent to the local laboratory. The analysis was performed with the Roche Cobas b121 and b221 analyzer, Roche Diagnostics Ltd., Switzerland (in the period between 1 January 2013 to 26 January 2016), and with the RAPIDLab 348EX Blood Gas System by Siemens, Siemens Healthcare Diagnostics, Germany (27 January 2016–31 December 2020). The whole analytic process was conducted in accordance with the manufacturers’ instructions. For the purposes of the final analysis, the following CBG parameters were included: pH, partial carbon dioxide pressure (pCO₂), partial oxygen pressure (pO₂), and oxygen saturation (SatO₂).

The anonymized patients’ information included demographical (age, sex), and laboratory data (C-reactive protein (CRP), procalcitonin, white blood cells count (WBC), absolute neutrophil count (ANC)), as well as clinical data (duration of the signs/symptoms prior to the hospitalization, duration of the fever, the highest fever during the disease; pulse oximetry, breath rate and heart rate at admission).

The patients were assigned to the groups depending on the age: under 6 months of age, 6–23 months of age, 24–59 months of age, and ≥60 months old. The choice of the age groups was based upon the most widely used age-based risk groups of a severe influenza course [3,26].

The major end-point consisted of a diagnosis of a lower respiratory tract infection (LRTI) and was based upon the final diagnoses coded according to the ICD-10: J10.0 (influenza due to an identified influenza virus with pneumonia), or a laboratory confirmed influenza J10.x (influenza due to an identified influenza virus with extensions other than pneumonia), together with the pneumonia code (j12.x-j18.x) and/or bronchitis (J20.x) and/or bronchiolitis (J21.x) and/or not-specified LRTI (J22). The secondary endpoints included the performance of a chest X-ray (CXR), the presence of radiologically confirmed pneumonia, the length of the hospital stay, an ICU admission, and a fatal outcome. A theoretical model which assumed a CBG-driven performance of a CXR was created.

The clinical diagnosis of the LRTI was made upon the clinical signs/symptoms of pneumonia, bronchitis or bronchiolitis. According to the local Polish guidelines on the respiratory tract infection diagnosis and treatment, pneumonia might be diagnosed in the presence of the typical signs and/or symptoms, such as cough, tachypnoea, fever, retraction of the intercostal spaces, a dull percussion note, and abnormalities on the auscultation (crackles or a bronchial murmur), while bronchitis is characterized by cough (productive or unproductive) accompanied by wheezing or rales on auscultation [27]. Bronchiolitis is diagnosed when the first episode of the restriction of bronchioles (expiratory dyspnea, wheezing, rales, and/or hypoxia) is seen in the course of a respiratory tract infection in children under 2 years of age [27].
Radiologically confirmed pneumonia was defined as the presence of clinical signs/symptoms plus a radiological confirmation, irrespective of the character of the CXR findings (interstitial, lobar or mixed); no additional assessment on the correlation with the types of the radiological abnormalities was performed. The chest X-ray performance depended on the individualized decisions made by the team of physicians responsible for the treatment and was not influenced by the study. In the case of a clinical suspicion of pneumonia that was not confirmed with the X-ray, the patient was included into the group of LRTI (if the above conditions were met) but excluded from the analysis of the radiologically confirmed pneumonia. The LOS was considered prolonged if the length of the hospitalization exceeded the median value. The ICU admission was the endpoint itself and no detailed analysis of the type of the respiratory support or length of hospitalization in the ICU was carried out.

The distribution of continuous data was determined with the Kolmogorov–Smirnov test, according to which a mean with a standard deviation (SD) or a median with an interquartile range (IQR) were used to present the normally or abnormally distributed data, respectively. The corresponding parametric (unpaired Student t-test) or non-parametric (Mann–Whitney U test) tests were performed. A ROC (receiver-operating characteristic) curve analysis was performed to calculate the area under the curve (AUC), and the Youden index was used to estimate optimal cut-off values for the CBG parameters in the prediction of the categorical endpoints (LRTI, CXR performance, radiologically confirmed pneumonia, prolonged LOS—for this parameter data was categorized upon the median value). The sensitivity, specificity, positive and negative predictive values (PPV and NPV) were computed for the prediction of a LRTI and for the radiologically confirmed pneumonia. Based upon the negative predictive values for radiologically confirmed pneumonia, the best performing CBG parameters were chosen for the theoretical model of CBG-driven CXR protocol. In such a protocol, the optimal cut-off values were applied to decide on the performance of the CXR. The odds ratios (in comparison to the lack of a CBG-driven protocol) of the unnecessarily performed CXR (i.e., a negative result) and the/omitted radiologically confirmed pneumonia cases with 95% confidence intervals (95%CI) were calculated. The decrease in the odds of an unnecessary CXR (e.g., the one that did not reveal pneumonia) was calculated by subtracting 1 from the odds ratio and its 95% CI. Additionally, a Spearman’s rank test calculated the correlation between the CBG parameters. The p value lower than 0.05 was assumed to be statistically significant. The statistical analysis was performed with Statistica 13.1 software (Statsoft, Tulsa, OK, USA). The study obtained the approval by the Ethics Committee at the Centre of Postgraduate Medical Education in Warsaw, Poland (approval number 141/PB/2020 issued on 9 December 2020). Due to the retrospective character of the analysis, the patient’s and parents’ or tutors’ consent was waived.

3. Results

A total of 495 children were hospitalized due to influenza in the period between 2013 and 2020. One child was discharged upon parental request, and the CBG was performed in 409 cases (82.6%) which finally created the study group (213 boys and 196 girls) (Figure 1). The patients’ age varied from 13 days to 17 years and three months, with a median of 31 months; the patients’ distribution into the age groups was as follows: <6 mo—71 patients (17%), 6–23 mo—97 patients (24%), 24–59 mo—132 patients (32%), ≥60 mo—109 patients (27%).

Children younger than two years old with LRTIs presented with a lower pH (7.42 vs. 7.447, p = 0.025 in <6 mo group and 7.445, p = 0.046 in 6–23 mo), higher pCO₂ (37.57 vs. 32.07, p < 0.01 in <6 mo), lower pO₂ (median 52.4 vs. 60.9, p = 0.002 in <6 mo and median 65.8 vs. 71.2, p < 0.01 in 6–23 mo), and lower SatO₂ (88.2 vs. 93.45 vs. 95.1, p < 0.01 in 6–23 mo) (Table 1). Of note, children above two years showed no statistically significant differences in the CBG parameters, regardless of the lower respiratory tract involvement.
Figure 1. Flow chart of patients in the study.

Table 1. Baseline characteristics of the patient study groups according to the lower respiratory tract involvement; the mean and standard deviation (SD) (marked in blue) or median with the interquartile range are shown according to the data distribution; the standard deviation values are followed by the SD. The corresponding test results (p-values) are shown: Student t-test results are marked with *, while the remaining are the Mann–Whitney U test results. LOS—length of stay, CRP—C-reactive protein, PCT—procalcitonin, WBC—white blood cells, ANC—absolute neutrophil count. Statistically significant results are bolded.
Table 1. Cont.

| Age Group | LRTI | Without LRTI | p      |
|-----------|------|--------------|--------|
| 6–23 mo   | mean/median with SD or IQR | mean/median with SD or IQR |        |
| age [months] | 12   | 9            | 18    | 13   | 9     | 17    | 0.657 |
| duration of signs/syndromes [days] | 4    | 1            | 5     | 1    | 1     | 4     | 0.053 |
| the highest fever [Celsius degrees] | 39.0 | 38.6         | 39.5  | 39.3 | 38.7  | 40.0  | 0.064 |
| LOS [days] | 8    | 6            | 11    | 5    | 4     | 7     |        |
| breath rate [per minute] | 30   | 25           | 40    | 28   | 25    | 30    | 0.121 |
| heart rate [per minute] | 120  | 115          | 140   | 120  | 110   | 130   | 0.345 |
| CRP [mg/L] | 6.33 | 2.72         | 20.72 | 4.28 | 1.20  | 14.15 | 0.071 |
| PCT [ng/mL] | 0.35 | 0.18         | 1.03  | 0.20 | 0.13  | 0.54  | 0.026 |
| WBC [10^3/µL] | 11.95 | 8.55        | 16.77 | 9.05 | 6.71  | 10.86 | 0.011 |
| ANC [10^3/µL] | 4.78 | 2.55         | 8.92  | 4.16 | 2.12  | 6.06  | 0.071 |
| pH         | 7.43 | 0.04 (SD)    | 7.44  | 0.05 (SD) | 0.846 * |
| pCO_2 [mmHg] | 32.75 | 29.15      | 35.05 | 30.40 | 27.00 | 33.50 | 0.069 |
| pO_2 [mmHg] | 65.80 | 59.20       | 70.95 | 71.20 | 65.00 | 76.40 | 0.006 |
| SatO_2     | 93.45 | 91.35       | 94.55 | 95.10 | 93.30 | 96.10 | 0.000 |
| 24–59 mo   | mean/median with SD or IQR | mean/median with SD or IQR |        |
| age [months] | 37   | 29           | 48    | 39   | 31    | 50    | 0.878 |
| duration of signs/syndromes [days] | 4    | 3            | 6     | 3    | 1     | 5     | 0.020 |
| the highest fever [Celsius degrees] | 39.2 | 39.0         | 40.0  | 39.5 | 39.0  | 40.0  | 0.789 |
| LOS [days] | 7    | 5            | 9     | 4    | 3     | 6     | 0.000 |
| breath rate [per minute] | 28   | 24           | 30    | 24   | 22    | 26    | 0.029 |
| heart rate [per minute] | 120  | 100          | 130   | 109  | 100   | 120   | 0.137 |
| CRP [mg/L] | 7.70 | 3.46         | 16.64 | 5.75 | 1.00  | 17.21 | 0.086 |
| PCT [ng/mL] | 0.21 | 0.13         | 0.80  | 0.17 | 0.10  | 0.36  | 0.135 |
| WBC [10^3/µL] | 8.09 | 5.81         | 9.95  | 7.52 | 5.42  | 10.47 | 0.270 |
| ANC [10^3/µL] | 4.32 | 2.31         | 7.26  | 3.95 | 2.19  | 6.69  | 0.389 |
| pH         | 7.43 | 0.04 (SD)    | 7.43  | 0.05 (SD) | 0.974 * |
| pCO_2 [mmHg] | 32.60 | 4.94       | 33.40 | 5.65 | (SD)  | 0.847 * |
| pO_2 [mmHg] | 65.90 | 60.00       | 74.85 | 68.95 | 63.80 | 77.20 | 0.055 |
| SatO_2     | 93.70 | 91.20       | 95.40 | 94.25 | 92.60 | 95.90 | 0.054 |
| >60 mo     | mean/median with SD or IQR | mean/median with SD or IQR |        |
| age [months] | 74   | 64           | 95    | 84   | 70    | 107   | 0.169 |
| duration of signs/syndromes [days] | 5    | 2            | 6     | 3    | 1     | 4     | 0.010 |
| the highest fever [Celsius degrees] | 39.5 | 39.0         | 40.0  | 39.5 | 39.0  | 40.0  | 0.958 |
| LOS [days] | 6    | 4            | 8     | 4    | 3     | 5     | 0.001 |
| breath rate [per minute] | 22   | 20           | 24    | 22   | 20    | 24    | 0.696 |
| heart rate [per minute] | 100  | 90           | 110   | 91   | 85    | 100   | 0.150 |
| CRP [mg/L] | 6.83 | 1.90         | 20.70 | 6.99 | 3.19  | 15.05 | 0.841 |
| PCT [ng/mL] | 0.22 | 0.10         | 0.45  | 0.14 | 0.08  | 0.26  | 0.104 |
| WBC [10^3/µL] | 5.94 | 4.70         | 8.89  | 6.18 | 4.43  | 8.51  | 0.941 |
| ANC [10^3/µL] | 3.56 | 2.09         | 6.89  | 3.52 | 2.28  | 5.42  | 0.961 |
| pH         | 7.44 | 7.41         | 7.44  | 7.42 | 7.41  | 7.45  | 0.658 |
| pCO_2 [mmHg] | 35.30 | 30.90      | 39.90 | 35.80 | 32.20 | 38.60 | 0.773 |
| pO_2 [mmHg] | 67.60 | 59.70       | 74.30 | 69.70 | 63.70 | 75.00 | 0.288 |
| SatO_2     | 94.10 | 89.80       | 95.30 | 94.45 | 92.70 | 95.60 | 0.287 |

The ROC curve analysis revealed that the number of the CBG parameters able to predict the LRTI decreased with the age. While in <6 mo each of the analyzed CBG parameters showed statistically significant AUC, in the 6–23 mo group, the pH, pO_2, and SatO_2 correlated with the LRTI, and in 24–59 mo, only the pO_2 and SatO_2 were associated with the LRTI, and none of the parameters were significant in patients older than five years (Figure 2).

The pCO_2 had the highest area under the curve for the prediction of the LRTI in children <6 mo (AUC = 0.75, 95%CI: 0.63–0.87, p < 0.01), but remained insignificant in older age groups (Table 2). The pH, on the other hand, was relevant both in children aged <6 mo and 6–23 mo, but had a much lower AUC (0.65, 95%CI: 0.52–0.78, p = 0.02, and 0.62, 95%CI: 0.51–0.73, p = 0.33, respectively). The SatO_2 showed high a AUC in children <6 mo,
6–23 mo, and lower in 24–59 mo (AUC = 0.74, 95% CI: 0.62–0.86, p < 0.01, AUC = 0.71, 95% CI: 0.61–0.82, p < 0.01, and AUC = 0.602, 95% CI: 0.502–0.702, p = 0.045, respectively), while a slightly lower AUC was observed in the case of pO$_2$ in the same age groups (AUC = 0.73, 95% CI: 0.6–0.85, p < 0.01, AUC = 0.67, 95% CI: 0.56–0.78, p < 0.01, and AUC = 0.601, 95% CI: 0.501–0.702, p = 0.0475, respectively).

Optimal cut-off values for children <6 mo were calculated and equaled: pH = 7.442 (sensitivity = 73.9%, specificity = 54.2%, PPV = 43.6%, and NPV = 81.3%), pCO$_2$ = 36.1 mmHg (sensitivity = 69.6%, specificity = 72.9%, PPV = 55.2%, and NPV = 83.3%), pO$_2$ = 58 mmHg (sensitivity = 73.9%, specificity = 60.4%, PPV = 47.2%, and NPV = 82.9%), SatO$_2$ = 93% (sensitivity = 91.3%, specificity = 50.0%, PPV = 46.7%, and NPV = 92.3%) (Table 2).

In children 6–23 mo, the following cut-offs were established: pH = 7.445 (sensitivity = 82.5, specificity = 47.4%, PPV = 52.4%, and NPV = 79.4%), pO$_2$ = 68.6 mmHg (sensitivity = 72.5%, specificity = 59.7%, PPV = 55.8%, and NPV = 75.6%), SatO$_2$ = 94% (sensitivity = 72.5%, specificity = 64.9%, PPV = 59.2%, and NPV = 77.1%).

In patients aged 24–59 months, the cut-offs were established at: pO$_2$ = 76.6 mmHg (sensitivity = 91.3%, specificity = 29.1%, PPV = 40.8%, and NPV = 86.2%), SatO$_2$ = 96.2% (sensitivity = 95.7%, specificity = 23.3%, PPV = 40%, and NPV = 90.9%).

For the prediction of the chest X-ray performance (irrespective of its result), the highest AUC in children <6 mo was seen for pO$_2$ (AUC = 0.7, 95% CI: 0.57–0.83, p = 0.022), followed by SatO$_2$ (AUC = 0.68, 95% CI: 0.53–0.82, p = 0.016), and pCO$_2$ (AUC = 0.66, 95% CI: 0.52–0.81, p = 0.027) (Table 3). In children 6–23 mo, the highest AUC was observed for SatO$_2$ (AUC = 0.75, 95% CI: 0.65–0.85, p < 0.01), followed by pO$_2$ (AUC = 0.69, 95% CI: 0.59–0.8, p < 0.01) and pCO$_2$ (AUC = 0.62, 95% CI: 0.51–0.73, p = 0.033). No significant associations were found in older children.

Figure 2. Cont.
Figure 2. The usefulness of capillary blood gas parameters in the prediction of lower respiratory tract infection (LRTI)- the results of the ROC analysis; (A) patients under 6 months of age, (B) patients aged 6–23 months old, (C) patients aged 24–59 months old.
Table 2. The results of the ROC curve analysis for the prediction of a lower respiratory tract infection (LRTI) and radiologically confirmed pneumonia (CXR + pneumonia). The cut-off values were calculated with the Youden index. AUC—area under the curve, 95%CI—95% confidence interval, PPV—positive predictive value, NPV—negative predictive value.

|                  | AUC   | 95%CI   | p     | cut off | Sensitivity 95%CI | Specificity 95%CI | PPV 95%CI | NPV 95%CI |
|------------------|-------|---------|-------|---------|-------------------|-------------------|-----------|-----------|
| **0–6 mo LRTI**  |       |         |       |         |                   |                   |           |           |
| pH               | 0.651 | 0.520   | 0.783 | 0.024   | 7.442             | 73.91%            | 54.17%    | 43.59%    | 81.25%    |
| pCO2             | 0.749 | 0.633   | 0.865 | 0.000   | 36.10             | 69.57%            | 72.92%    | 55.17%    | 83.33%    |
| pO2              | 0.727 | 0.604   | 0.849 | 0.000   | 58.00             | 73.91%            | 60.42%    | 47.22%    | 82.86%    |
| SatO2            | 0.740 | 0.620   | 0.861 | 0.000   | 93.00             | 91.30%            | 50.00%    | 46.67%    | 92.31%    |
| **CXR+ pneumonia** |     |         |       |         |                   |                   |           |           |
| pH               |        | insignificant |
| pCO2             | 0.740 | 0.607   | 0.873 | 0.000   | 36.10             | 73.33%            | 67.86%    | 37.93%    | 90.48%    |
| pO2              | 0.754 | 0.619   | 0.888 | 0.000   | 58.00             | 86.67%            | 58.93%    | 36.11%    | 94.29%    |
| SatO2            | 0.756 | 0.621   | 0.891 | 0.000   | 91.60             | 93.33%            | 51.79%    | 34.15%    | 96.67%    |
| **6–23 mo LRTI** |       |         |       |         |                   |                   |           |           |
| pH               | 0.622 | 0.510   | 0.734 | 0.033   | 7.455             | 82.50%            | 47.37%    | 52.38%    | 79.41%    |
| pCO2             | 0.666 | 0.556   | 0.776 | 0.003   | 68.60             | 72.50%            | 59.65%    | 55.77%    | 75.56%    |
| pO2              | 0.714 | 0.609   | 0.818 | 0.000   | 94.00             | 72.50%            | 64.91%    | 59.18%    | 77.08%    |
| SatO2            | 0.714 | 0.609   | 0.818 | 0.000   | 94.00             | 56.11–85.40%      | 51.13–77.09% | 49.25–68.42% | 66.26–85.21% |
| **CXR+ pneumonia** |     |         |       |         |                   |                   |           |           |
| pH               |        | insignificant |
| pCO2             | 0.640 | 0.527   | 0.754 | 0.016   | 28.00             | 92.86%            | 67.68%    | 37.68%    | 92.86%    |
| pO2              | 0.695 | 0.588   | 0.803 | 0.000   | 68.60             | 76.50–99.12%      | 26.29–50.17% | 32.89–42.73% | 76.77–98.08% |
| SatO2            | 0.755 | 0.655   | 0.855 | 0.000   | 94.00             | 63.11–93.94%      | 45.48–69.76% | 36.39–52.37% | 77.90–94.78% |
Table 2. Cont.

| LRTI      | AUC 95%CI | p   | cut off | Sensitivity 95%CI | Specificity 95%CI | PPV 95%CI | NPV 95%CI |
|-----------|-----------|-----|---------|-------------------|-------------------|-----------|-----------|
| pH        | insign.   |     |         |                   |                   |           |           |
| pCO₂      | insign.   |     |         |                   |                   |           |           |
| pO₂       | 0.601     | 0.501 | 0.702   | 0.048             | 76.60             | 91.30%    | 29.07%    | 40.78%    | 86.21%    |
|           |           |     |         |                   |                   | 79.21–97.58%| 19.78–39.86%| 36.93–44.74%| 69.84–94.40%|
| SatO₂     | 0.602     | 0.502 | 0.702   | 0.045             | 96.20             | 95.65%    | 23.26%    | 40.00%    | 90.91%    |
|           |           |     |         |                   |                   | 85.16–99.47%| 14.82–33.61%| 36.89–43.20%| 70.97–97.61%|
| CXR+ pneumonia | AUC 95%CI | p   | cut off | Sensitivity 95%CI | Specificity 95%CI | PPV 95%CI | NPV 95%CI |
| pH        | insign.   |     |         |                   |                   |           |           |
| pCO₂      | insign.   |     |         |                   |                   |           |           |
| pO₂       | insign.   |     |         |                   |                   |           |           |
| SatO₂     |           |     |         |                   |                   |           |           |

Table 3. The results of the ROC curve analysis for the performance of chest X-ray (CXR) and prolonged length of stay (LOS). The cut-off values are calculated with the Youden index. AUC—area under the curve, 95%CI—95% confidence interval, PPV—positive predictive value, NPV—negative predictive value.

| 0–6 mo | 6–23 mo | 24–59 mo |
|--------|---------|---------|
| CXR    | LOS     | CXR     |
| AUC 95%CI | p   | cut off | AUC 95%CI | p | cut off |
| pH     | insign. |         | pH     | insign. |         |
| pCO₂  | 0.663   | 0.519   | 0.808  | 0.027 | 36.1  | pCO₂  | 0.636   | 0.503   | 0.768  | 0.045  | 36.0  |
| pO₂   | 0.703   | 0.573   | 0.833  | 0.002 | 52.3  | pO₂   | 0.726   | 0.608   | 0.843  | 0.000  | 52.3  |
| SatO₂ | 0.676   | 0.533   | 0.819  | 0.016 | 91.6  | SatO₂ | 0.712   | 0.591   | 0.833  | 0.001  | 88.2  |

| CXR    | LOS     | CXR     |
| AUC 95%CI | p   | cut off | AUC 95%CI | p | cut off |
| pH     | insign. |         | pH     | insign. |         |
| pCO₂  | 0.622   | 0.510   | 0.734  | 0.033 | 28.5  | pCO₂  | insign. |         |
| pO₂   | 0.693   | 0.587   | 0.799  | 0.000 | 68.6  | pO₂   | 0.633   | 0.517   | 0.750  | 0.025  | 68.1  |
| SatO₂ | 0.751   | 0.652   | 0.849  | 0.000 | 94.6  | SatO₂ | 0.659   | 0.544   | 0.775  | 0.007  | 92.9  |

| CXR    | LOS     | CXR     |
| AUC 95%CI | p   | cut off | AUC 95%CI | p | cut off |
| pH     | insign. |         | pH     | insign. |         |
| pCO₂  | insign. |         | pCO₂  | 0.390   | 0.288   | 0.493  | 0.036  |
| pO₂   | insign. |         | pO₂   | insign. |         |
| SatO₂ | insign. |         | SatO₂ | insign. |         |
More significant relationships were observed for a radiologically confirmed pneumonia. SatO2 demonstrated the highest AUC in both <6 mo and 6–23 mo groups (AUC = 0.76, 95%CI: 0.62–0.89, p < 0.01 and AUC = 0.76, 95%CI: 0.66–0.86, p < 0.01, respectively), and was followed by pO2 (AUC = 0.75, 95%CI: 0.62–0.89, p < 0.01 and AUC = 0.7, 95%CI: 0.59–0.8, p < 0.01, respectively). An optimal cut-off for SatO2 was established: SatO2 < 6 mo = 91.6% showed a sensitivity = 93.3%, specificity = 51.8%, PPV = 34.2%, and NPV = 96.7%, while SatO2 6–23 mo = 94% had a sensitivity = 82.1%, specificity = 62.3%, PPV = 46.9%, and NPV = 89.6%; pO2 < 6 mo = 58 mmHg showed a sensitivity = 86.7%, specificity = 58.9%, PPV = 36.1%, and NPV = 94.3%, and pO2 6–23 mo = 68.6 mmHg had a sensitivity = 82.1%, specificity = 58%, PPV = 44.2%, and NPV = 88.9%.

If the calculated cut-off values were to be applied to decide on the CXR performance, the use of SatO2 in children under 2 years old (91.6 mmHg <6 mo and 94 mmHg in 6–23 mo) would decrease the odds of an unnecessary CXR by 84.15% (95%CI: 74.5–90.14%, p < 0.01), while the pO2 use would decrease the odds by 84.58% (95%CI: 75.17–90.42%) (Table 4). The odds of omitting a radiologically confirmed pneumonia were statistically insignificant (with an assumption of no omitted cases when no-CBG protocol was made). Moreover, children with a positive CXR who would not have their CXR performed due to normal SpO2 (six children) or pO2 (seven children) values, 16.7% and 14.3%, respectively (one child in each group), did not obtain antibiotics, meaning that even a positive CXR did not influence their clinical management.
Figure 3. The usefulness of capillary blood gas parameters in the prediction of radiologically-confirmed pneumonia (CXR + Pneumonia)—the results of the ROC analysis; (A) patients under 6 months of age, (B) patients aged 6–23 months old, (C) patients aged 24–59 months old.
Table 4. The odds ratio of a negative CXR; a comparison between the theoretical CBG-driven model and a lack of CBG-driven protocol. The results are shown separately for the age groups and combined for the parameter (SpO₂ or pO₂) as odds ratio and the corresponding odds reduction. OR—odds ratio, 95%CI—95% confidence interval. Note: SatO₂ and pO₂ were selected as tools for the decision-making process due to the highest negative predictive value.

| Negative CXR Performance | Cut-Off   | OR    | 95%CI   | p       | %  | 95%CI   |
|--------------------------|-----------|-------|---------|---------|----|---------|
| SatO₂ < 6 mo             | 91.6%     | 0.1644| 0.0781  | 0.3461  | <0.01| 83.56  | 65.39  | 92.19  |
| SatO₂ 6–23 mo            | 94%       | 0.1486| 0.0793  | 0.2785  | <0.01| 85.14  | 72.15  | 92.07  |
| SatO₂ 0–23 mo            | 0.1585    | 0.0986| 0.2550  |         | <0.01| 84.15  | 74.5   | 90.14  |
| pO₂ < 6 mo               |           |       |         |         |     |         |        |        |
| pO₂ 6–23 mo              |           |       |         |         |     |         |        |        |
| pO₂ 0–23 mo              |           |       |         |         |     |         |        |        |

The AUC for the extended LOS prediction in <6 mo was 0.73 (95%CI: 0.61–0.84, p < 0.01) for pO₂, followed by SatO₂ (AUC = 0.71, 95%CI: 0.59–0.83, p < 0.01) and pCO₂ (AUC = 0.64, 95%CI: 0.5–0.77, p = 0.045), while in 6–23 mo the highest AUC was observed for SatO₂ (AUC = 0.66, 95%CI: 0.54–0.78, p < 0.01) followed by pO₂ (AUC = 0.63, 95%CI: 0.52–0.75, p = 0.025) (Table 3). Inversely, in children 24–59 mo, the increased pCO₂ correlated with a shorter LOS (AUC = 0.39, p = 0.036).

Due to the low number of patients transferred to the ICU (one patient), a further analysis on this endpoint would present a low statistical value and was thus not performed. No fatal cases were reported.

An internal correlation between the CBG parameters was observed between the pH and pCO₂ (rho = –0.78), pO₂ (rho = 0.33), and SatO₂ (rho = 0.57) and between the pCO₂ and pO₂ (rho = –0.49), SatO₂ (rho = –0.66).

4. Discussion

This study shows that the capillary blood gas has a promising value in the assessment of the risk of a lower respiratory tract involvement in young children hospitalized due to influenza and might possibly prompt a new direction in the assessment of pediatric influenza patients. To the best of our knowledge, this is the first study focusing on the use of the CBG in children with influenza. It needs to be underscored that, except for numerical merit, the use of CBG seems to be promising due to practical implications. While capillary blood sampling is a relatively easy-to-perform and repeatable method, arterial blood sampling is far more invasive, technically difficult, and painful [22]. We did not analyze patients’ compliance nor opinion on arterial versus capillary sampling, but we are convinced that when a less invasive method of comparable value is available, it should be the preferred method.

Firstly, the influence of the patients’ age needs to be considered; while in children under six months old each of the analyzed CBG parameters exhibited a significant AUC for the prediction of the lower respiratory tract involvement, in older children the predictive value of the pCO₂, pH, and SatO₂ or pO₂ gradually disappeared with the increasing age (at 6–23, 24–59, and over 60 mo, respectively). It is interesting to note that the upper age limit after which the CBG loses its usefulness coincides with the upper age limit of the risk of a severe influenza course [26]. This might be explained by a lower impact of LRTIs on older children; even if the lower respiratory tract is infected, it does not disturb the gas exchange to such a high degree as observed in younger patients.

In general, the SatO₂ and pO₂ demonstrate the best performance in the prediction of LRTIs in children under 24 mo. Both SatO₂ and pO₂ also showed a statistical significance in children 24–59 mo in the LRTI risk assessment, although its power decreased with age, and became insignificant over the age of five years. In children under 6 mo and 6–23 mo, the
AUC was moderate and differed only slightly between SatO$_2$ and pO$_2$, with a preference for SatO$_2$ (AUC = 0.74 and 0.73 for <6 mo, or 0.71 and 0.67 for 6–23 mo, respectively), although both parameters might be considered for the purposes of a LRTI prediction.

It needs to be emphasized that although some doubts on the true reflection of the arterial parameters have been raised in the case of the SatO$_2$ and pO$_2$, in our group of patients, the capillary SatO$_2$ and pO$_2$ turned out to be independent predictors of LRTIs, not biased by the uncertainties on its true relationship with the arterial blood parameters. The published data show 65–69% of concordance between capillary and arterial blood in the case of pO$_2$ [22–24], yet some authors reported ratios as low as r = 0.358 [25] (Harrison et al. 1997).

The pCO$_2$, on the other hand, seems to reflect arterial values more precisely. However, in our series of patients, its use is restricted to the youngest group [22]. The pCO$_2$ showed the best performance in infants under 6 mo, but it was insignificant in older children. The previous studies on the pCO$_2$ confirmed a strong correlation between the arterial and capillary blood results. Yildildas and colleagues revealed the concordance ratio of 0.988, which is in line with the previous reports showing ratios between r = 0.86, r = 0.9534, and r = 0.955 [22–25]. Interestingly, although the level of agreement between the CBG and the ABG is also high in the case of pH (a slightly lower correlation—between r = 0.823 and r = 0.903) [22,25], the pH (just as pCO$_2$) was relevant to a lower degree than SatO$_2$ or pO$_2$. There are several possible explanations for these alleged discrepancies. The first one is the group selection, as patients included in our study (except for one) did not present a respiratory failure, and obviously a lower respiratory tract involvement is not unequivocally related to a respiratory failure. Promising results of pCO$_2$ and pH use in the prediction of an ICU transfer were observed in children with bronchiolitis [28,29], but not in this group of influenza hospitalizations. In fact, influenza LRTIs may affect oxygen supply to a higher degree than carbon dioxide elimination or acid-base balance. The research by Zhang et al. reported decreased arterial pO$_2$ in 73.5% of patients (adults, only one child was included into the study) hospitalized due to H7N9 influenza, while the pCO$_2$ was increased only in 3% of cases [30]. Secondly, the age-related differences in the respiratory tract pathophysiology might offer another interpretation: limited compensatory capacities in the youngest group of patients may rapidly lead to an increased pCO$_2$, while the older children are not affected so significantly. Bronchiolitis is diagnosed mainly in the first year of life, while influenza patients were seen in each age group. Thirdly, differences between the etiological factors need to be recognized, since the major etiological factor of bronchiolitis is respiratory syncytial virus (RSV), whereas the influenza virus does not result in bronchiolitis so frequently and significant differences in immunological response patterns might be expected [18,31,32].

Another interesting aspect is the high negative predictive value in the prediction of a radiologically confirmed pneumonia, which suggests that a CBG-driven protocol might help decrease the number of needlessly performed chest X-rays, the decrease in the study reached almost 85%. Restraining from the CXR on the sole basis of CBG seems to be somehow controversial, but if the CBG was, for example, one of the steps in the verification of the need for a CXR, then children with influenza could be exposed to radiation less frequently. Certainly, this is only preliminary data, and a lack of a radiological confirmation of pneumonia is not equal to a lack of pneumonia, but the practical approach also shows that, in some cases, a positive CXR does not change the clinical management of the patient.

The relationship between a prolonged hospital stay and the oxygenation parameters in younger children (also an unexpected reverse relationship with pCO$_2$ in those aged 24–59 mo) may suggest that oxygenation is more affected in the course of influenza in children, while the carbon dioxide elimination and acid–base balance remain more stable. The association with a respiratory failure and its consecutive stages, however, needs to be established. The internal correlation between the CBG parameters does not let a replacement of one by another, and attention should be paid to their use in the different age groups.
The study may be limited by several factors, including its retrospective character, resulting in the lack of performance of CBG in each patient (although almost 83% of the patients underwent a CBG), and a single-center setting, which impairs a generalization of the results without a further confirmation. Secondly, the optimal study design would consist of a radiological verification of the lower respiratory tract involvement in each patient, yet ethical concerns would make unnecessary exposure to radiation unacceptable. A change of the CBG analyzer needs to be mentioned as well, although an instrument-related bias is unlikely and no significant aberrations are expected. Furtherly, we did not perform influenza subtype or lineage analysis, considering the limited access to such diagnostic tools, even in hospital settings, although clinical differences related to the specific virus should be recognized. Finally, the group preselection might also play a role, and we cannot exclude the usefulness of the CBG in older children, presumably in patients in a more severe condition. However, our results confirm its value only in younger children.

5. Conclusions

In conclusion, we found the CBG a useful tool, mainly in children under two years of age. Children under six months of age represent the group that would benefit the most from CBG. The CBG is able to predict the risk of a lower respiratory tract involvement and, more importantly, to exclude the risk of a radiologically confirmed pneumonia with an approximated 90% NPV. A CBG-based qualification for a CXR performance could significantly decrease the number of unnecessary radiological chest examinations.

Author Contributions: Conceptualization, A.W. and T.J.; methodology, A.W. and T.J.; validation, A.W. and T.J.; formal analysis, A.W.; investigation, A.W. and T.J.; resources, A.W. and T.J.; data curation, A.W.; writing—original draft preparation, A.W.; writing—review and editing, A.W. and T.J.; visualization, A.W.; supervision, A.W. and T.J.; project administration, A.W. and T.J.; funding acquisition, A.W. and T.J. All authors have read and agreed to the published version of the manuscript.

Funding: The study was supported by CMKP grant 501-1-020-19-22.

Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki and approved by the Ethics Committee at the Centre of Postgraduate Medical Education in Warsaw, Poland (approval number 141/PB/2020 issued on 9 December 2020).

Informed Consent Statement: Due to the retrospective character of the analysis, the patient’s and parents’ or tutors’ consent was waived. No identifying data about specific patients were collected, thus informed consent for paper publication was not necessary.

Data Availability Statement: Data are available on request from the authors.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Wang, X.; Li, Y.; O’Brien, K.L.; Madhi, S.A.; Widdowson, M.-A.; Byass, P.; Omer, S.B.; Abbas, Q.; Ali, A.; Amu, A.; et al. Global burden of respiratory infections associated with seasonal influenza in children under 5 years in 2018: A systematic review and modelling study. *Lancet Glob. Health* 2020, 8, e497–e510. [CrossRef]
2. Maldonado, Y.A.; O’Leary, S.T.; Banerjee, R.; Barnett, E.D.; Campbell, J.D.; Caserta, M.T.; Caserta, M.T.; Gerber, J.S.; Kourtis, A.P.; Lynfield, R.; et al. Recommendations for Prevention and Control of Influenza in Children, 2020–2021. *Pediatrics* 2020, 146, e2020024588. [CrossRef]
3. DISEASES COI. Recommendations for Prevention and Control of Influenza in Children, 2021–2022. *Pediatrics* 2021, 148, e2021053745. [CrossRef] [PubMed]
4. Cui, A.; Xie, Z.; Xu, J.; Hu, K.; Zhu, R.; Li, Z.; Li, Y.; Sun, L.; Xiang, X.; Xu, B.; et al. Comparative analysis of the clinical and epidemiological characteristics of human influenza virus versus human respiratory syncytial virus versus human metapneumovirus infection in nine provinces of China during 2009–2021. *J. Med. Virol.* 2022. [CrossRef]
5. Terliesner, N.; Unterwalder, N.; Edelmann, A.; Corman, V.; Knaust, A.; Rosenfeld, L.; Gratopp, A.; Ring, H.; Martin, L.; von Bernuth, H.; et al. Viral infections in hospitalized children in Germany during the COVID-19 pandemic: Association with non-pharmaceutical interventions. *Front. Pediatr.* 2022, 10, 935483. [CrossRef]
6. Willis, G.; Bloomfield, L.; Berry, M.; Bulsara, C.; Chaney, G.; Cooke, H.; Maticic, J.; Russell, K.; Zic, M.; Mak, D. The impact of a vaccine mandate and the COVID-19 pandemic on influenza vaccination uptake in Western Australian health care students. *Vaccine* 2022, 40, 5651–5656. [CrossRef]

7. Shen, A.K.; Browne, S.; Srivastava, T.; Michel, J.J.; Tan, A.S.L.; Kormides, M.L. Factors Influencing Parental and Individual COVID-19 Vaccine Decision Making in a Pediatric Network. *Vaccines* 2022, 10, 1277. [CrossRef]

8. Grohskopf, L.A.; Alyanak, E.; Ferdinands, J.M.; Broder, K.R.; Blanton, L.H.; Talbot, H.K.; Fry, A.M. Prevention and Control of Seasonal Influenza with Vaccines: Recommendations of the Advisory Committee on Immunization Practices—United States, 2020–2021 Influenza Season. *MMWR Recomm. Rep.* 2020, 70, 1–28. [CrossRef]

9. Shope, T.R.; Walker, B.H.; Aird, L.D.; Southward, L.; McCown, J.S.; Martin, J.M. Pandemic Influenza Preparedness Among Child Care Center Directors in 2008 and 2016. *Pediatrics* 2017, 139, e20163690C. [CrossRef]

10. Gresh, L.; Kuan, G.; Sanchez, N.; Azziz-Baumgartner, E.; Ojeda, S.; Melendez, M.; Lopez, R.; Martin, E.T.; Widdowson, M.-A.; Breesee, J.; et al. Burden of Influenza and Influenza-associated Pneumonia in the First Year of Life in a Prospective Cohort Study in Managua, Nicaragua. *Pediatr. Infect. Dis. J.* 2016, 35, 152–156. [CrossRef]

11. Sharma, L.; Rebaza, A.; Cruz, C.S.D. When “B” becomes “A”: The emerging threat of influenza B virus. *Eur. Respir. J.* 2019, 54, 1901325. [CrossRef] [PubMed]

12. Yu, J.; Qian, S.; Liu, C.; Xiao, Y.; Xu, T.; Wang, Y.; Su, H.; Chen, L.; Yuan, B.; Wang, X.; et al. Viral etiology of life-threatening pediatric pneumonia: A matched case-control study. *Infl. Other Respir. Viruses* 2020, 14, 452–459. [CrossRef]

13. McCullers, J.A. The co-pathogenesis of influenza viruses with bacteria in the lung. *Nat. Rev. Microbiol.* 2014, 12, 252–262. [CrossRef] [PubMed]

14. Kubale, J.; Kuan, G.; Gresh, L.; Ojeda, S.; Schiller, A.; Sanchez, N.; Lopez, R.; Azziz-Baumgartner, E.; Wraith, S.; Harris, E.; et al. Individual-level Association of Influenza Infection With Subsequent Pneumonia: A Case-control and Prospective Cohort Study. *Clin. Infect. Dis.* 2020, 73, e4288–e4295C. [CrossRef] [PubMed]

15. Jung, J.; Seo, E.; Yoo, R.N.; Sung, H.; Lee, J. Clinical significance of viral-bacterial codetection among young children with respiratory tract infections: Findings of RSV, influenza, adenoviral infections. *Medicine (Baltimore)* 2020, 99, e18504. [CrossRef]

16. Sanders, C.J.; Vogel, P.; McClaren, J.L.; Bajracharya, R.; Doherty, P.C.; Thomas, P.G. Compromised respiratory function in lethal influenza infection is characterized by the depletion of type I alveolar epithelial cells beyond threshold levels. *Am. J. Physiol. Cell. Mol. Physiol.* 2013, 304, 1481–1488. [CrossRef]

17. Niethamer, T.K.; Stabler, C.T.; Leach, J.P.; Zepp, J.A.; Morley, M.P.; Babu, A.; Zhou, S.; Morrissey, E.E. Defining the role of pulmonary endothelial cell heterogeneity in the response to acute lung injury. *Elife* 2020, 9, e53072. [CrossRef]

18. LeMessurier, K.S.; Tiwary, M.; Morin, N.P.; Samarasinghe, A.E. Respiratory Barrier as a Safeguard and Regulator of Defense Against Influenza A Virus and Streptococcus pneumoniae. *Front. Immunol.* 2020, 11, 3. [CrossRef]

19. McNally, B.; Ye, F.; Willette, M.; Fláño, E. Local Blockade of Epithelial PDL-1 in the Airways Enhances T Cell Function and Viral Clearance during Influenza Virus Infection. *J. Virol.* 2013, 87, 12916–12924. [CrossRef]

20. Schneider, C.; Nobs, S.P.; Heer, A.K.; Kurrer, M.; Klinke, G.; Van Rooijen, N.; Vogel, J.; Kopf, M. Alveolar Macrophages Are Essential for Protection from Respiratory Failure and Associated Morbidity following Influenza Virus Infection. *PLoS Pathog.* 2014, 10, e1004503. [CrossRef]

21. Peltola, V.; Ziegler, T.; Ruuskanen, O. Influenza A and B Virus Infections in Children. *Clin. Infect. Dis.* 2003, 36, 299–305. [CrossRef] [PubMed]

22. Yildizdas, D.; Yapıcıoğlu, H.; Yilmaz, H.L.; Sertdemir, Y. Correlation of simultaneously obtained capillary, venous, and arterial blood gases of patients in a paediatric intensive care unit. *Arch. Dis. Child.* 2004, 89, 176–180. [CrossRef] [PubMed]

23. Kirubakaran, C.; Gnananayagam, J.E.J.; Sundaravalli, E.K. Comparison of blood gas values in arterial and venous blood. *Indian J. Pediatr.* 2003, 70, 781–785. [CrossRef] [PubMed]

24. Escalante-Kanashiro, R.; Tantaleán-Da-Fieno, J. Capillary blood gases in a pediatric intensive care unit. *Crit. Care Med.* 2000, 28, 224–226. [CrossRef] [PubMed]

25. Harrison, A.M.; Lynch, J.M.; Dean, J.M.; Witte, M.K. Comparison of simultaneously obtained arterial and capillary blood gases in pediatric intensive care unit patients. *Crit. Care Med.* 1997, 25, 1904–1908. [CrossRef] [PubMed]

26. CDC. Available online: https://www.cdc.gov/flu/professionals/antivirals/summary-clinicians.htm (accessed on 30 August 2022).

27. Hryniewicz, W. *Rekomendacje Postępowania w Pozaszpitalnych Zakażeń Układu Oddechowego*; Narodowy Instytut Leków: Warszawa, Poland, 2016.

28. Wrotek, A.; Kobiałka, M.; Jackowska, T. Capillary Blood Gas Predicts Risk of Intensive Care in Children with Bronchiolitis. *Children* 2021, 8, 719. [CrossRef]

29. Vo, A.T.; Liu, D.R.; Schmidt, A.R.; Festekjian, A. Capillary blood gas in infants with bronchiolitis: Can end-tidal capnography replace it? *Am. J. Emerg. Med.* 2021, 45, 144–148. [CrossRef]

30. Zhang, J.; Zhao, Y.; Chen, Y. Laboratory findings in patients with avian-origin influenza A (H7N9) virus infections. *J. Med. Virol.* 2013, 86, 895–898. [CrossRef]
31. Goritzka, M.; Makris, S.; Kausar, F.; Durant, L.; Pereira, C.; Kumagai, Y.; Culley, F.; Mack, M.; Akira, S.; Johansson, C. Alveolar macrophage–derived type I interferons orchestrate innate immunity to RSV through recruitment of antiviral monocytes. *J. Exp. Med.* 2015, 212, 699–714. [CrossRef]

32. Choi, J.; Callaway, Z.; Kim, H.-B.; Fujisawa, T.; Kim, C.-K. The role of TNF-α in eosinophilic inflammation associated with RSV bronchiolitis. *Pediatr. Allergy Immunol.* 2010, 21, 474–479. [CrossRef]