Immunotherapy in triple negative breast cancer: beyond checkpoint inhibitors

Yara Abdou, Atta Goudarzi, Jia Xin Yu, Samik Upadhyaya, Benjamin Vincent and Lisa A. Carey

The development of immunotherapy agents has revolutionized the field of oncology. The only FDA-approved immunotherapeutic approach in breast cancer consists of immune checkpoint inhibitors, yet several novel immune-modulatory strategies are being actively studied and appear promising. Innovative immunotherapeutic strategies are urgently needed in triple negative breast cancer (TNBC), a subtype of breast cancer known for its poor prognosis and its resistance to conventional treatments. TNBC is more primed to respond to immunotherapy given the presence of more tumor infiltrating lymphocytes, higher PD-L1 expression, and higher tumor mutation burden relative to the other breast cancer subtypes, and therefore, immuno-oncology represents a key area of promise for TNBC research. The aim of this review is to highlight current data and ongoing efforts to establish the safety and efficacy of immunotherapeutic approaches beyond checkpoint inhibitors in TNBC.

INTRODUCTION

Triple negative breast cancer (TNBC) accounts for ~15–20% of incident breast cancers and is characterized by negativity for estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2). TNBC encompasses more than one molecular subtype, with the majority of tumors found to be of the basal-like RNA expression phenotype. However, other molecular subtypes can be present; including the HER2-Enriched group, Luminal A, Luminal B, Claudin-low, and a few normal-like molecular subtypes. More recent studies have further refined TNBC subtyping by identifying targetable molecules highly expressed in each TNBC subtype for effective immune-based treatment strategies. Despite significant biological heterogeneity within this disease, and recent advances in neoadjuvant and adjuvant therapy, overall, TNBC has been associated with a higher risk for recurrence and disease progression, and poorer outcomes. Once metastatic, this breast cancer subtype has an estimated median overall survival (OS) of 16 months and a median progression-free survival (PFS) of 5.6 months with standard chemotherapy in the first-line setting. There is an unmet clinical need to develop more efficacious and less toxic therapies for patients with TNBC.

The development of immunotherapy agents has revolutionized the field of oncology with durable responses and improvements in OS. Different tumor types including melanoma, renal cell carcinoma, lung cancer, and bladder cancer have greatly benefited from immune checkpoint inhibitors (ICIs), however, in breast cancer, early trials with ICI as monotherapy achieved limited objective responses. Immunotherapy for treatment of breast cancer has not been prioritized, largely because breast cancer has been considered poorly immunogenic making it less likely to respond to immunotherapies. Nevertheless, there is increasing evidence to suggest the presence of variable immunogenic activity in different breast cancer subtypes, with TNBC likely exhibiting the strongest immunogenicity. TNBC has been shown to have a higher proportion of tumor infiltrating lymphocytes (TILs) compared to other subtypes, relatively high tumor mutational burden and PD-L1 expression, and survival associations with degree of T cell and B cell infiltration, making immunotherapy a promising option against this disease. This has encouraged the development of more immunotherapy drugs to treat TNBC patients.

The first FDA accelerated approval of an ICI for the treatment of breast cancer came in March 2019 when the anti-PD-L1 antibody atezolizumab was approved in combination with nab-paclitaxel as a first-line treatment for patients with PD-L1-positive, metastatic TNBC based on the IMpassion130 trial. Continued approval of this combination was contingent upon results of the IMpassion131 trial evaluating first-line atezolizumab and paclitaxel in TNBC, however, updated results in 2021 indicated that the trial failed to meet the primary end point of PFS superiority in the frontline treatment of patients with PD-L1 positivity and there was no difference in survival advantage in the PD-L1-positive nor the intention to treat population. Based on this data, atezolizumab-chemotherapy combination has been withdrawn as an indication for treatment of TNBC. Alternatively, KEYNOTE-355 continues to demonstrate a clinically meaningful improvement in PFS with pembrolizumab, an anti-PD-1 antibody, in combination with chemotherapy, vs. chemotherapy alone among patients with PDL-1 positive, metastatic TNBC with CPS ≥ 10. Based on these results, on November 2020, the FDA granted accelerated approval to pembrolizumab in combination with chemotherapy for the treatment of patients with metastatic TNBC whose tumors express PD-L1 (CPS ≥ 10). Similarly, in July 2021, pembrolizumab was approved for high-risk, early-stage, TNBC in combination with chemotherapy as neoadjuvant treatment and continued as adjuvant treatment, based on results from the KEYNOTE-522 trial showing substantial benefit in terms of event-free survival and distant recurrence-free survival, regardless of PD-L1 status. There are several additional ongoing clinical trials evaluating the role of other types of immunotherapy combinations in TNBC. Ongoing efforts have revolved around modulating the tumor microenvironment (TME) to increase breast cancer immunogenicity and the therapeutic efficacy of immunotherapeutic agents.

1Lineberger Comprehensive Cancer Center, University of North Carolina, Chapel Hill, NC 27599, USA. 2Department of Medicine, University at Buffalo, Buffalo, NY 14203, USA. 3Parker Institute for Cancer Immunotherapy, San Francisco, CA 94129, USA. 4Cancer Research Institute, New York, NY 10006, USA. ✉email: Yara_Abdou@med.unc.edu
The objective of this review is to discuss emerging immunotherapy agents in TNBC patients, highlighting therapies beyond ICIs.

LANDSCAPE OF TNBC IMMUNOTHERAPIES

Using their immuno-oncology (IO) database, the Cancer Research Institute (CRI) explored the number and type of IO agents being developed for use in breast cancer, specifically TNBC (Fig. 1a). The data pull completed in March 2022 showed 778 total agents actively being developed for use in breast cancer and TNBC, at various developmental stages. ICIs, which have had recent approvals in TNBC, lag behind cancer vaccines, adoptive cell therapies and “Other Immunomodulators”; these include immunomodulators to natural killer cells, B cells, and other immune cells. Newer modalities such as T cell engagers, which are bispecific antibodies that simultaneously bind with the T-cell and the tumor cell, are mostly in preclinical or early phase, while cytokine-based therapies, have progressed to later phases of development given its approved use in other cancers such as melanoma, particularly peginterferon alfa-2b and interleukin-2. Due to the antigen-specific nature of some of these IO agents, CRI further explored the most
common targets of these IO agents (Fig. 1b). While HER2 is the most common target identified in breast cancer, other major targets include, adenosine receptor targets, nonspecific tumor-associated antigens (TAA), immune checkpoints (PD-1, PD-L1), toll-like receptors (TLRs), and colony-stimulating factor-1 (CSF1/R).

Clinical trial data was pulled from clinicaltrials.gov as of March 2022 to select for TNBC trials utilizing at least one IO agent, regardless of recruitment status. Despite the development of many categories of IO agents, ICIs dominate in TNBC, representing 295 of 354 (83%) of TNBC IO trials (Fig. 2a). Most of the IO studies are still in early phases, with ~40 trials currently in phase 3. Many of these studies have yet to disclose efficacy. We further divided the trials based on initiation date to estimate how many trials could possibly have efficacy data in the near term (Fig. 2b). The majority of trials were initiated in the past 6 years, peaking in 2020, with a noted decline in the last 2 years, presumably due to the COVID-19 pandemic. As these trials complete, we expect more immunotherapy options for TNBC patients to emerge beyond checkpoint inhibitors.

**VACCINE THERAPY**

Therapeutic vaccines typically target known breast tumor antigens to enhance tumor-specific immune responses through active immunization. Some newer methods generate vaccines based on the mutanome of a patient’s particular tumor. Vaccines can variably generate cytotoxic CD8+ T-cell (CTLs) and NK responses, as well as affect tumor growth directly by altering the TME through chemokines. Different modalities of therapeutic vaccines exist utilizing either peptides, carbohydrates, DNA or RNA (usually recombinant), whole cells, or dendritic cells (DC). Peptides and carbohydrates present limited epitopes, whereas genetic vaccines can generate the entirety of epitopes associated with an antigen. DC vaccines utilize the antigen-presenting and T-cell stimulating properties of DC, as well as modulating cytokines and chemokines to control inflammation and lymphocyte migration to help produce long-lasting anti-tumor effects.

**Peptide and carbohydrate-based vaccines**

Monovalent peptide and carbohydrate-based vaccines generate an immune response to a single antigen. Sialyl-Tn (STn) is a
carbohydrate antigen associated with poor prognosis in multiple cancers including breast cancer\textsuperscript{31}. In a phase III study, patients receiving endocrine therapy plus a vaccine consisting of STn conjugated to the carrier protein keyhole-limpet hemocyanin (KLH) showed increased OS in 180 metastatic breast cancer patients (all subtypes) compared to immunization with just the KLH protein (37 vs. 31 months). The analysis did not take into consideration the molecular subtypes, and the number of TNBC patients included in the study was not disclosed, therefore there is unknown applicability to TNBC\textsuperscript{32}.

AE37 is the iI-Key hybrid of the Major Histocompatibility Complex (MHC) class II peptide that is capable of stimulating CD4\textsuperscript{+} helper T cells. The final analysis of the randomized phase II trial of AE37 + GM-CSF vaccine verses GM-CSF alone for the prevention of breast cancer recurrence in node-positive or high-risk, node negative patients, showed no significant differences in the 5-year disease-free survival (DFS) between treatment arms. However, the subgroup analysis showed that the TNBC cohort (n = 50) had an \textasciitilde 35% reduction in the relative risk of recurrence\textsuperscript{33,34}. Further studies are needed to evaluate the clinical benefit of this vaccine in TNBC.

Folate receptor alpha (FRA) is another target for a therapeutic vaccine in patients with breast cancer. A phase 1 study of FRA peptide vaccine in breast and ovarian cancer patients was well tolerated and elicited augmented immunity in more than 90% of patients examined, with responses that persisted at least 12 months\textsuperscript{35}. Two phase II trials are investigating cyclophosphamide combined with FRA peptide vaccine in large cohorts of women with TNBC (NCT03012100, NCT02593227).

Vaccines utilizing the membrane-bound carbohydrate antigens NeuGCGM3 and Muc1, have shown trends toward better clinical outcome and prolonged periods of no evidence of disease (NED) respectively, in advanced-stage breast cancer\textsuperscript{36,38}. Uncontrolled studies of vaccines targeting telomerase (hTERT) and survivin, an anti-apoptosis protein, have produced periods of stable disease and immunological response in studies of advanced-stage breast cancers of undisclosed subtypes\textsuperscript{19,40}. Other peptide-based vaccines including P10s (a peptide that mimics a carbohydrate antigen) and personalized peptide vaccine (PPV) (where vaccine antigens are selected based on pre-existing immunoglobulin G (IgG) responses) have shown enhanced immunogenicity and possible clinical benefits in early clinical studies, especially in TNBC subgroups\textsuperscript{41,42}.

### Table 1. Clinical trials for breast cancer treatment using vaccine therapy in TNBC.

| Vaccine                                                                 | Patients                  | Phase | NCT              | Status          |
|------------------------------------------------------------------------|---------------------------|-------|------------------|----------------|
| Peptide—AE37 with Pembrolizumab                                        | Stage IV TNBC             | II    | NCT04024800      | Active, not recruiting |
| Peptide—P10s-PADRE with or without standard chemotherapy                | Stage II–III TNBC         | II    | NCT02938442      | Recruiting      |
| Neo-antigen peptide—long peptide with nab-paclitaxel and Durvalumab    | Stage IV TNBC             | II    | NCT03606967      | Recruiting      |
| Multi-peptide—PVX-410 with Durvalumab                                  | Stage II–III TNBC         | Ib    | NCT02826434      | Active, not recruiting |
| Multi-peptide—PVX-410 with or without Pembrolizumab                    | Stage IV or inoperable HLA A2+ TNBC | Ib    | NCT03362060      | Active, not recruiting |
| Multi-peptide—Galinpepimut (WT1)                                       | Select advanced cancers including TNBC | I/I   | NCT03761914      | Active, not recruiting |
| Multi-peptide—Folate Receptor Alpha with GM-CSF following cyclophosphamide | Stage Ib–IV TNBC          | II    | NCT03012100      | Recruiting      |
| DNA—adenovirus CEA, MUC1, and brachyury Peptide—RAS, CEA, and brachyury; with various chemotherapeutics and targeted inhibitors | Progressive TNBC post standard therapy | Ib/II | NCT03387085      | Active, not recruiting |
| DNA—vaccinia p53                                                        | Solid tumors that failed prior therapy (including TNBC) | I     | NCT02432963      | Active, not recruiting |
| Neo-antigen DNA—with or without Durvalumab                             | Stage II/III TNBC         | I     | NCT03199040      | Active, not recruiting |
| Neo-antigen RNA liposomes                                               | All stage TNBC            | I     | NCT02316457      | Active, not recruiting |
| DC—Autologous DC’s pulsed with autologous neo-antigen peptides          | All stage TNBC            | I     | NCT04105582      | Completed as of June 2022 |
| DC targeted against Her2/Her3 with pembrolizumab and a cytokine modulation regimen | Stage IV TNBC or HER2+ breast cancer | II | NCT04348747 | Recruiting |

**Polyvalent peptide vaccines**

Polyvalent peptide vaccines target multiple antigens in a single vaccine. Takahashi and colleagues reported 9 clinical responses among 79 breast cancer patients using peptides selected from a pool of 31 peptides targeting various antigens based on HLA typing and in vivo antibody response, including 1 TNBC with complete response and a second TNBC with partial response\textsuperscript{42}. Additionally, 9 of 10 TNBC patients in this study showed augmented antibody responses, and 7 of 14 patients showed augmented cytotoxic cellular response\textsuperscript{42}. Berinstein and colleagues used a 7-peptide vaccine targeting 7 different tumor antigens in a cohort of 23 adenocarcinomas that included three metastatic breast cancers and showed SD lasting 8 months in one patient and CD8\textsuperscript{+} T-cell responses in all three patients\textsuperscript{43}. Studies using PVX-410, a four-peptide vaccine targeting three antigens, and Galinpepimut, a four-peptide vaccine targeting WT1, have shown clinical effects in studies of smoldering myeloma and mesothelioma, respectively\textsuperscript{44,45}. Clinical trials are currently underway for TNBC patients utilizing PVX-410, and Galinpepimut vaccines (Table 1).

**Neoantigen vaccines**

Neoantigen vaccines use peptides that are unique to particular mutations in the patient’s tumor and not present in normal cells, therefore theoretically avoid host self-tolerance\textsuperscript{27,46}. These
vaccines were shown to elicit robust anti-tumor immune responses through activation of tumor antigen-specific CD8+ and CD4+ T cells. A randomized phase II study of nab-paclitaxel, durvalumab and neoadjuvant vaccine vs. nab-paclitaxel and durvalumab alone in metastatic TNBC is currently recruiting (NCT03606967). Another phase I clinical trial of a neoadjuvant vaccine with or without durvalumab to treat stage II–III TNBC patients who have residual disease after neoadjuvant therapy is also recruiting (NCT03199040).

Genetic vaccines

Genetic vaccines utilize recombinant DNA of an antigen in a vector of isolated plasmid, virus, bacterial or yeast cell. The antigens are then expressed by host cells (in the case of isolated plasmid), or are expressed by the viral, bacterial or yeast cells in the vaccines. These methods are advantageous because these vectors utilize complete target complementary DNA (cDNA) sequences and therefore can generate multiple antigenic epitopes per target. Furthermore, some vectors themselves are immunogenic, which potentiates the targeted immune response, and the vectors can further be transfected with cDNA of T-cell stimulating proteins to enhance that effect. The PANVAC vaccine is a poxvirus transfected with cDNA of CEA and MUC-1, as well as the T-cell stimulating proteins B7.1, ICAM-1 and LFA-3. This vaccine has produced clinical responses in 3 patients, including 1 TNBC. Four vaccines—PANVAC; a related CEA vaccine with the same stimulating proteins, a yeast-based vaccine targeting brachyury; a transcription factor associated with tumor epithelial-mesenchymal transition, and viral vaccines containing the NY-ESO-1 antigen, have all produced SD in varying proportions of carcinomas, although these studies contained small numbers of breast cancer patients and did not have control groups. A study of INVAC-1, an isolated plasmid vaccine containing cDNA of hTERT, in a population of 26 adenocarcinomas, including 5 patients with stage III-IV TNBC, showed two instances of PR and 15 cases of SD of up to 10 months. Additionally, a vaccine of P53-transfected vaccinia cells along with pembrolizumab produced regression of cutaneous metastasis in a woman with stage IV TNBC. Clinical trials of genetic vaccines containing CEA1, MUC1, and Brachyury, and P53 are currently underway for TNBC patients (Table 1).

Dendritic cell vaccines

DC vaccines use autologous patient-derived DC as a vaccine vector; either loaded/transfected with tumor antigens or fused with autologous or allogeneic preparations. A study of ten patients with stage III TNBC investigated the use of neoadjuvant DC loaded with WT1 and cyclin B1 antigens, and a third using DC vaccine targeting HER2/HER3 antigens with pembrolizumab in breast cancer patients with brain metastasis (Table 1).

ADAPTIVE CELL THERAPIES

T cells play an important role in cell-mediated immunity. Chimeric antigen receptor T-cell (CAR-T) therapy and T-cell receptor (TCR)-engineered T-cell therapy are two types of adaptive cell therapies (ACTs) that can genetically modify the patient’s natural T cells ex vivo and inject them back into the patient’s body to make them tumor-specific and enhance their ability to destroy tumor cells. The mechanisms by which they recognize antigens are quite different. CAR T-cells are engineered to recognize only surface antigens through its antigen-binding site of antibody fragments fused to the T-cell. A major advantage of CAR-T cells is relatively high antigen-binding affinity, typically in the nanomolar range compared to micromolar TCR binding affinities. On the other hand, TCRs use an alpha-beta chain heterodimer to recognize intracellular antigens that are expressed on the cell surface by MHC. Therefore, TCRs may have an advantage over CAR-T in solid tumors, given they are able to target a wider range of antigens.

Chimeric antigen receptor-modified T (CAR-T) cell therapy

Current CAR T-cell therapies are approved for the treatment of certain patients with non-Hodgkin lymphomas and leukemias. Despite the exciting clinical efficacy seen in hematologic malignancies, several challenges still exist for the use of CAR T-cell therapies in solid tumors. Many studies are focusing on overcoming these challenges and improving the efficacy of this novel approach to immunotherapy in solid tumors. A key challenge for CAR-T therapy to overcome in breast cancer is improving CAR-T cell infiltration into tumors, which may be overcome by using potent stimulation of antigen-presenting cells to make chemotactic cytokines combined with administration of CAR-T cells.

A cell-surface molecule, c-Met, was found to be highly expressed in ~50% of breast tumors, and is associated with basal-like TNBC, supporting the production of a CAR-T cell specific for c-Met. Tchou and colleagues published results from a phase 0 trial (NCT01873620) evaluating the safety and feasibility of intratumoral injections of RNA c-Met-CAR-T cells in patients with c-Met-expressing metastatic breast cancer. Four out of six patients had TNBC. Results showed that the c-Met-CAR-T-cell injections were well tolerated and elicited an inflammatory response intratumorally. A phase 1 trial (NCT03060356) to evaluate the feasibility, safety and efficacy of intravenously administered mRNA c-Met-CAR-T cells in patients with metastatic breast cancer is underway.

Mesothelin expression was found to be highly expressed in TNBC and is associated with poor prognosis. This prompted the production of mesothelin-specific CAR-T cells which have shown to have in vitro anti-tumor cytotoxicity against primary breast tumor cells. Preliminary results from the phase I/Ii study (NCT02414269) evaluating the safety and efficacy of mesothelin-targeted CAR-T cells in patients with advanced solid tumors showed evidence of CAR-T cell anti-tumor activity and no major toxicities. A phase I clinical trial (NCT02792114) to evaluate the safety and tolerability of mesothelin-targeted CAR-T cells in patients with pretreated metastatic mesothelin-expressing breast cancer is currently recruiting.

Mucin 1 (MUC1) is a heterodimeric protein that is highly expressed in over 90% of TNBC and is associated with poor prognosis. Tumor MUC1 specific CAR-T cells were shown to have potent anti-tumor cytotoxicity both in vitro and in vivo. A Phase I study of anti-MUC1 CAR-T cells for patients with advanced
MUC1 positive breast cancer is currently recruiting (NCT04020575). Receptor tyrosine kinase-like orphan receptor 1 (ROR1) is expressed on tumor cells of primary breast cancer and high expression of ROR1 has been associated with poor prognosis81. ROR1 CAR-T cells were shown to confer a potent anti-tumor effect against TNBC cell lines in vitro90. A phase I study of ROR1 CAR-T cells in patients with advanced ROR1+ malignancies including TNBC is currently ongoing (NCT02706392). Preliminary results from 6 patients (4 with TNBC) showed no dose-limiting toxicities with some evidence of disease control81. NKR-2 malignancies including TNBC is currently ongoing (NCT02706392).

**ONCOLYTIC VIRUS THERAPY**

Oncolytic viruses have shown promising therapeutic efficacy in preclinical breast cancer models. Currently, different types of oncolytic viruses are being developed to target early and metastatic breast cancer. TVEC is a genetically modified type 1 herpes simplex virus (HSV-1) to preferentially replicate in tumor cells99. An early-phase trial including nine patients with TNBC found that adding the oncolytic virus talimogene laherparevoc (T-VEC) to standard chemotherapy in the neoadjuvant setting was well tolerated and showed promising efficacy90. Although the sample size was small, results showed increases in cytopathic T cell infiltration in most of the resected tumor specimens in addition to a reduction in regulatory T cells90. Results of the phase II trial (NCT02779855), with forty patients enrolled, confirmed that the addition of TVEC to neoadjuvant chemotherapy was safe with increased rates of pathological complete response91. Another phase II trial evaluated the efficacy of intratumoral T-VEC as monotherapy for inoperable locoregional recurrence of breast cancer with or without distant recurrence (NCT02658812). The study showed that intratumoral T-VEC as monotherapy did not have optimal outcomes due to uncontrolled disease progression, and administration of concurrent systemic therapy may be warranted92. Furthermore, an early phase, multi-institutional study to evaluate the safety of intratumoral T-VEC injections in combination with atezolizumab in TNBC patients with liver metastasis (NCT03256344) observed no dose-limiting toxicities, and reported one patient that had a partial response93. A phase II trial of in situ oncolytic virus therapy consisting of adenovirus-mediated expression of herpes simplex virus thymidine kinase plus ganciclovir and stereotactic body radiation therapy followed by pembrolizumab was tested in twenty-eight patients (eighteen with PD-L1 negative tumors) with locally advanced or metastatic TNBC (NCT03004183). Results showed that the combination was well-tolerated, with promising efficacy in heavily pretreated metastatic TNBC patients. One patient had complete response, and has remained disease-free without any systemic therapy for 39 months despite early discontinuation of pembrolizumab due to Grade 3 pneumonitis84.

LTX-315, a novel oncolytic peptide, has shown promising results when administered as monotherapy or in combination with pembrolizumab in TNBC patients with transfusional accessible tumors (NCT01986426)95. In a TNBC in vivo model, LTX-315 combined with doxorubicin induced immune-mediated changes in the TME and demonstrated promising therapeutic potential96.

Pelareorep, a serotype 3 reovirus, was evaluated in patients with metastatic breast cancer, including TNBC. The final analysis of the randomized phase II study showed that pelareorep was well tolerated and the combination arm had a significantly longer OS97. The AWARE-1 trial (NCT04102618) is currently enrolling early-stage breast cancer patients to 5 different cohorts with pelareorep. Six TNBC patients will be treated with combination of pelareorep and atezolizumab. Preliminary data demonstrated enhanced inflammatory markers in the tumor after the combination treatment98.

Adenovirus is the most studied oncolytic virus platform in breast cancer research. A phase I trial using adenovirus, ICOVIR-7, enrolled three patients with advanced breast cancer. While the drug was rendered to be safe, the breast cancer patients did not meet their efficacy endpoints99. On the other hand, an oncolytic adenovirus coding for GMCSF (Ad5/3-D24-GMCSF) was shown to induce anti-tumoral immunity and efficacy in patients with advanced breast tumors, including TNBC100.

Several other oncolytic viruses, including Maraba, Measles, Polio, Coxackie, Vaccinia, Newcastle disease, have been tested in breast cancer in vivo and ex vivo models, paving the way for future safety studies in humans101. Ongoing clinical trials are currently investigating the efficacy of different oncolytic viruses in solid tumors and breast cancer (Table 2).
administration of a recombinant adenovirus encoding IL-12 (AdIL-12) resulted in significant tumor regression in breast cancer animal models. The Phase 1 pilot study of IL-12 monotherapy in treatment refractory, metastatic TNBC patients showed evidence of treatment-related increase in CD8+ TIL density and enhanced antigen presentation after intratumoral administration of IL-12. The Phase 2 KEYNOTE-890 trial (NCT03567720) evaluated the efficacy of intratumoral tachykinin teleplasmin, a plasmid encoding IL-12, followed by electroporation and pembrolizumab in metastatic TNBC patients. Evidence of enhanced tumor immunogenicity was observed in addition to 28.6% objective responses, regardless of PDL-1 status. NKTR-214, also known as bempegaldesleukin, is an engineered cytokine that specifically stimulates the IL-2 receptor. NKTR-214 has been tested in a phase 1 trial (NCT02869295) targeting metastatic solid tumor, including TNBC. Based on a favorable safety profile and evidence of a substantial increase in CD8+ T and NK cells within the TME after NKTR-214 treatment, a phase 1/II clinical trial combining NKTR-214 and nivolumab was initiated (NCT02983045). 38 patients with metastatic solid tumors were enrolled, preliminary results showed early evidence of clinical activity with an ORR of 13.2% and no dose-limiting toxicities.

**Novel immunotherapy targeting the TME**

Several clinical trials are investigating novel therapeutic approaches to overcome the immunosuppressive effects of the breast cancer TME. Some of these approaches include expanding effector T-cells, NK cells and other immunostimulatory cells while suppressing regulatory T cells, tumor-associated M2 macrophages and myeloid derived suppressor cells (MDSCs). The goal is to modify the TME by increasing anti-tumor immune responses, suppressing pro-tumor immune responses to produce high immunogenicity and ultimately, a more favorable response to cancer immunotherapy.

SD-101; an intratumoral toll-like receptor 9 (TLR9) agonist, is a novel immune priming strategy that was shown to modify the TME by increasing local production of type 1 Interferon, resulting in cytotoxic T-cell infiltration and an anti-tumor response. The combination of neoadjuvant SD-101 and pembrolizumab in addition to weekly paclitaxel followed by doxorubicin and cyclophosphamide was investigated in a treatment arm of the I-SPY 2 trial (NCT01042379). Results showed a non-statistically significant increase in estimated pCR rates in seventy-five patients with high-risk, HER2-negative stage I/II breast cancer (~30 patients with TNBC). Another novel and interesting method for TME remodeling and enhancing therapy outcomes is through targeting the chemokine system. Chemokines are small signaling proteins that direct the migration and trafficking of immune cells within the TME. Chemokines play a critical role in shaping the immune cell composition and mediating the balance between anti-tumor and pro-tumor responses. CCL5 (ligand for CCR5) and CXCL9, CXCL10, CXCL11 (ligands for CXCR3) are the main chemokines attracting CTLs, type-1 helper (Th1) and NK cells producing an inflammatory response in the TME. On the other hand, production of CCL2, CCL22, and CXCL12 promotes intratumoral infiltration of suppressive immune cells, such as CXCR4+ MDSCs, M2 macrophages, and CCR4+CXCR4+ regulatory T-cells (Tregs). A single arm study (NCT03599453) is investigating how well chemokine modulation therapy works when given prior to check point inhibitor (pembrolizumab) in patients with metastatic TNBC. The study uses a combination of celecoxib (COX-2 inhibitor), recombinant interferon alfa-2b (IFN-alpha), and rintatolimod (a selective toll-like receptors 3 (TLR-3) agonist) given systemically on 3 consecutive days, 1 week apart for a total of 2 weeks prior to initiating pembrolizumab every 3 weeks. Theodoraki and colleagues have shown that the combination of TLR-3 agonist with COX-2 blockers (and/or with IFN-alpha), allowed selective enhancement of type-1 immunity, promoting CTLs migration, while suppressing Treg/MDSC attraction.

**FUTURE DIRECTIONS**

Immune checkpoint inhibition added to chemotherapy improves survival outcomes in TNBC patients. Several other novel immunotherapeutic approaches show promise in this patient cohort. While the IO field continues to grow, a deeper understanding of breast cancer and its microenvironment is still needed to overcome the apparent low immunogenicity in this disease and to optimize immune-therapeutic approaches to their full potential.
At least three conditions are required to ensure optimal immunotherapy responses: (1) generation of tumor antigen-specific T cells, (2) influx of T cells into the tumor, and (3) reversal of immunosuppression mechanisms operating in the tumor immune microenvironment. Though tumor immunotherapy has classically focused on the T cell arm of the adaptive immune system, other approaches, such as tumor antigen-specific B cells, may be important as well to immunotherapy response in breast cancer. Thus, it is likely that combination immunotherapy strategies will be needed going forward. With an increasing number of clinical trials and available immunotherapeutic agents, we anticipate that these promising strategies will improve clinical outcomes in TNBC patients, while decreasing our dependence on cytotoxic therapies.

Received: 30 January 2022; Accepted: 13 October 2022; Published online: 09 November 2022

REFERENCES
1. Perou, C. M. Molecular stratification of triple-negative breast cancers. Oncologist 16, 61–70 (2011).
2. Burstein, M. D. et al. Comprehensive genomic analysis identifies novel subtypes and targets of triple-negative breast cancer. Clin. Cancer Res. 21, 1688–1698 (2015).
3. Rugga, H. S. et al. KEYNOTE-355: final results from a randomized, double-blind phase III study of first-line pembrolizumab + chemotherapy vs placebo + chemotherapy for metastatic triple-negative breast cancer. ESMO Congress 2021. LB A16. (2021).
4. Sharma, P. & Allison, J. P. Immune checkpoint targeting in cancer therapy: toward combination strategies with curative potential. Cell 161, 205–214 (2015).
5. Vinay, D. S. & Kwon, B. S. Harnessing immune checkpoints for cancer therapy. Immunotherapy 10, 1265–1284 (2018).
6. Planes-Laine, G. et al. PD-1/PD-L1 targeting in breast cancer: the first clinical evidences are emerging. A literature review. Cancers 11, 1033 (2019).
7. De La Cruz, L. M. & Czerniecki, B. J. Immunotherapy for breast cancer is controlled, randomised phase III trial of atezolizumab + chemotherapy vs. placebo + chemotherapy for early-stage TNBC. Ann. Oncol. 32, 1198–1200 (2021).
8. Emens, L. A. Breast cancer immunotherapy: facts and hopes. Clin. Cancer Res. 24, 511–520 (2018).
9. Sisirak, V. et al. Impaired IFN-γ-mediated survival in breast cancer: effect of vaccination on tumor immunosurveillance. Proc. Natl Acad. Sci. USA 105, 3014 (2018).
10. Liu, Z., Li, M., Jiang, Z. & Wang, X. A comprehensive immunologic portrait of triple-negative breast cancer. Transl. Oncol. 11, 311–329 (2018).
11. Miller, L. D. et al. Immunogenic subtypes of breast cancer delineated by gene classifiers of immune responsiveness. Cancer Immunol. Res. 4, 600 (2016).
12. Huang, J., Liu, L., Li, L., Gao, G. & Zhou, Y. Targeting tumor-infiltrating lymphocytes in triple negative breast cancer: the future of immune targeting. Clin. Med. Insights Oncol. 10, 31–39 (2016).
13. Stanton, S. E., Adams, S. & Disis, M. L. Variation in the incidence and magnitude of tumor-infiltrating lymphocytes in breast cancer subtypes: a systematic review. JAMA Oncol. 2, 1354–1360 (2016).
14. Thomas, A. et al. Tumor mutational burden is a determinant of immune-mediated survival in breast cancer. Oncoimmunology 7, e1490854 (2018).
15. Wimmerly, H. et al. PD-L1 expression correlates with tumor-infiltrating lymphocytes and response to neoadjuvant chemotherapy in breast cancer. Cancer Immunol. Res. 3, 326 (2015).
16. Iglesias, M. D. et al. Prognostic B-cell signatures using miRNA-seq in patients with subtype-specific breast and ovarian cancer. Clin. Cancer Res. 20, 3818–3829 (2014).
17. Emens, L. A. et al. LBA16 IMpassion130: final OS analysis from the pivotal phase III study of atezolizumab + nab-paclitaxel vs placebo + nab-paclitaxel in previously untreated locally advanced or metastatic triple-negative breast cancer. Ann. Oncol. 31, S1148 (2020).
18. Miles, D. et al. Primary results from IMpassion131, a double-blind, placebo-controlled, randomised phase III trial of first-line paclitaxel with or without atezolizumab for unresectable locally advanced/metastatic triple-negative breast cancer. Ann. Oncol. 32, 994–1004 (2021).
19. Cortes, J. et al. Pembrolizumab plus chemotherapy versus placebo plus chemotherapy for previously untreated locally recurrent inoperable or metastatic triple-negative breast cancer (KEYNOTE-355): a randomised, placebo-controlled, double-blind, phase 3 clinical trial. Lancet Oncol. 19, 1817–1828 (2020).
20. Emens, L. A. et al. KEYNOTE-522: phase III study of neoadjuvant pembrolizumab + chemotherapy vs. placebo + chemotherapy, followed by adjuvant pembrolizumab vs. placebo for early-stage TNBC. Ann. Oncol. 32, 1198–1200 (2021).
21. Thomas, A. et al. Current landscape of immunotherapy in breast cancer: a review. JAMA Oncol. https://doi.org/10.1001/jamaoncol.2018.7147 (2019).
22. Upadhyaya, S., Hubbard-Lucey, V. M. & Yu, J. X. Immuno-oncology drug development: new horizons in the era of COVID-19. Nat. Rev. Drug Discov. 19, 751–752 (2020).
23. Zhang, X., Sharma, P. K., Peter Goedegebuure, S. & Gillanders, W. E. Personalized cancer vaccines: targeting the cancer mutanome. Vaccine 35, 1094–1100 (2017).
24. Burke, E. E., Kodumudi, K., Ramamoorthy, G. & Czerniecki, B. J. Vaccine therapies for breast cancer. Surg. Oncol. Clin. N. Am. 28, 353–367 (2019).
25. Williams, A. D. et al. Immunotherapy for breast cancer: current and future strategies. Curr. Surg. Rep. 5, 516–527 (2019).
26. Perez, C. R. & De Palma, M. Engineering dendritic cell vaccines to improve cancer immunotherapy. Nat. Commun. 10, 3408 (2019).
27. Munkley, J. The Role of Sialyl-Tn in Cancer. Int. J. Mol. Sci. 17, 275 (2016).
28. Ibrahim, N. K. et al. Survival advantage in patients with metastatic breast cancer receiving endocrine therapy plus Sialyl Tn-KLH vaccine: post hoc analysis of a large randomized trial. J. Cancer 10, 377–584 (2013).
29. Mittendorf, E. et al. Primary analysis of a prospective, randomized, single-blinded phase II trial evaluating the HER2 peptide AE37 vaccine in breast cancer patients to prevent recurrence. Ann. Oncol. 27, 1241–1248 (2016).
30. Mittendorf, E. et al. Final report of the phase I/II clinical trial of the E75 (nelipimatum-S) vaccine with booster inoculations to prevent disease recurrence in high-risk breast cancer patients. Ann. Oncol. 25, 1735–1742 (2014).
31. Kalli, K. R. et al. Folate receptor alpha peptide vaccine generates immunity in breast and ovarian cancer patients. Clin. Cancer Res. 24, 3014 (2018).
32. Carr, A. et al. Immunotherapy of advanced breast cancer with a heterophilic ganglioside (NeuGcGM3) cancer vaccine. J. Clin. Oncol. 21, 1015–1021 (2003).
33. Mulems, V. et al. Immunogenicity and safety of a NeuGcGM3 based cancer vaccine: results from a controlled study in metastatic breast cancer patients. Hum. Vaccin. 6, 736–744 (2010).
34. Gilewski, T. et al. Immunization of metastatic breast cancer patients with a fully synthetic globo H conjugate: a phase I trial. Proc. Natl Acad. Sci. USA 98, 3270–3275 (2001).
35. Domchek, S. M. et al. Telomerase-specific T-cell immunity in breast cancer: effect of vaccination on tumor immunosurveillance. Cancer Res. 67, 10546–10555 (2007).
36. Tsuchida, T. et al. Clinical and immunological evaluation of anti-apoptosis protein, survivin-derived peptide vaccine in phase I clinical study for patients with advanced or recurrent breast cancer. J. Transl. Med. 6, 24 (2008).
37. Hutchins, L. F. et al. Targeting tumor-associated carbohydrate antigens: a phase I study of a carbohydrate mimetic-peptide vaccine in stage IV breast cancer subjects. Oncotarget 8, 99161–99178 (2017).
38. Takahashi, R. et al. Feasibility study of personalized peptide vaccination for metastatic recurrent triple-negative breast cancer patients. Breast Cancer Res. 16, R70 (2014).
39. Berinstein, N. L. et al. First-in-man application of a novel therapeutic cancer vaccine formulation with the capacity to induce multi-functional T cell responses in ovarian, breast and prostate cancer patients. J. Transl. Med. 10, 156 (2012).
40. Nooka, A. K. et al. Assessment of safety and immunogenicity of PVX-410 vaccine with or without lenalidomide in patients with smoldering multiple myeloma: a nonrandomized clinical trial. JAMA Oncol. 4, 1562 (2018).
41. Zauderer, M. G. et al. A randomized phase II trial of adjuvant galapimutem-5, WT-1 analogue peptide vaccine, after multi-modality therapy for patients with malignant pleural mesothelioma. Clin. Cancer Res. 23, 7483–7489 (2017).
42. Zhang, X. et al. Breast cancer neoantigens can induce CD8(+) T-cell responses and antitumor immunity. Cancer Immunol. Res. 5, 516–523 (2017).
43. Alsopch, E. et al. MHC-II neoantigens shape tumour immunity and response to immunotherapy. Nature 574, 696–701 (2019).
52. Mohebtash, M. et al. A pilot study of MUC-1/CEA/TRICOM poxviral-based vac-
53. Heery, C. R. et al. Phase I trial of a yeast-based therapeutic cancer vaccine (GI-
59. Joyce, O. et al. Safety and initial clinical ef–
65. Chick, R. C. et al. Subgroup analysis of nelipepimut-S plus GM-CSF combined
68. Garber, K. Driving T-cell immunotherapy to solid tumors.
69. Fucá, G., Reppel, L., Landoni, E., Savoldo, B. & Dotti, G. Enhancing chimeric
73. Li, Y. R. et al. Mesothelin expression is associated with poor outcomes in breast
cancer. Breast Cancer Res. Treat. 147, 675–684 (2014).

74. Tchou, J. et al. Mesothelin, a novel immunotherapy target for triple negative breast
cancer. Breast Cancer Res. Treat. 133, 799–804 (2012).
75. Adusumilli, P. S. et al. A phase I clinical trial of malignant pleural disease treated
with regionally delivered autologous mesothelin-targeted CAR-T cells. 2019 AACR Annual Meeting. Abstract CT068. (2019).
76. Kufe, D. W. MUC1-C oncoprotein as a target in breast cancer: activation of
tumor-infiltrating T cells. Cancer Res. 70, 5598–5599 (2010).
77. Jing, X., Liang, H., Hao, C., Yang, X. & Cui, X. Overexpression of MUC1 predicts
poor prognosis in patients with breast cancer. Oncol. Rep. 41, 801–810 (2019).
78. Zhou, R. et al. CAR T cells targeting the tumor MUC1 glycoprotein reduce triple-
negative breast cancer growth. Front. Immunol. 10, 1149–1149 (2019).
79. Zhang, S. et al. ROR1 is over-expressed in human breast cancer and associated
with enhanced tumor-cell growth. PLOS ONE 7, e31127 (2012).
80. Wallstabe, L. et al. ROR1-CAR T cells are effective against lung and breast cancer
in advanced microphysiological 3D tumor models. JCI Insight 4 https://doi.org/
10.1172/jci.insight.126345 (2019).
81. Specht, J. M. et al. Abstract CT131: a phase I study of adoptive immunotherapy
for advanced ROR1+ malignancies with defined subsets of autologous T cells
expressing a ROR1-specific chimeric antigen receptor (ROR1-CAR). Breast Cancer
Res. 78, CT131 (2018).
82. Ademuyiwa, F. O. et al. NY-ESO-1 cancer testis antigen demonstrates high
immunogenicity in triple negative breast cancer. PLoS ONE 7, e38783 (2012).
83. Ayyoub, M., Sciarlata, C. M., Hamai, A., Pignon, P. & Valmori, D. Expression
of MAGE-A3/6 in primary breast cancer is associated with hormone receptor
negative status, high histologic grade, and poor survival. J. Immunother. 37,
73–76 (2014).
84. Karn, T. et al. Melanoma antigen family A identified by the bimodality index
defines a subset of triple negative breast cancers as candidates for immune
response augmentation. Eur. J. Cancer 48, 12–23 (2012).
85. Gaglia, P. et al. Prognostic value of CEA and ferritin assay in breast cancer:
a multivariate analysis. Eur. J. Cancer Clin. Oncol. 4, 1151–1155 (1998).
86. Paret, C. et al. Cx0rf61 is a target for T cell based immunotherapy of triple-
negative breast cancer. Oncotarget 6, 25356–25367 (2015).
87. Marcinkowski, B. et al. Cancer targeting by TCR gene-engineered T cells directed
against Kita-Kyushu lung cancer antigen-1. J. Immunother. 7, 229 (2019).
88. Zacharaks, N. et al. Immune recognition of somatic mutations leading to
complete durable regression in metastatic breast cancer. Nat. Med. 24, 724–730
(2018).
89. Conry, R. M., Westbrook, B., McKeever, S. & Norwood, T. G. Talimogene laherpar-
epvex: first in class oncolytic virotherapy. Hum. Vaccin. Immunother. 14, 839–846
(2018).
90. Soliman, H. et al. A phase I trial of talimogene laherpevec in combination
with neoadjuvant chemotherapy for the treatment of nonmetastatic triple-
negative breast cancer. Cancer Res. 77, 1012–1018 (2021).
91. Soliman, H. H. et al. A phase 2 trial of talimogene laherpevec (TVEC) in
combination with neoadjuvant chemotherapy for the treatment of nonmeta-
static triple-negative breast cancer. J. Clin. Oncol. 39, 578–578 (2021).
92. Kai, M. et al. A phase II study of talimogene laherpevec for patients with
inoperable locoregional recurrence of breast cancer. Sci. Rep. 11, 22242 (2021).
93. Haskett, S. et al. Breast P3-09-09: Preliminary safety data of intrahepatic
talimogene laherpevec and intravenous atezolizumab in patients with triple
negative breast cancer. Cancer Res. 80, P3–09–19 https://doi.org/10.1158/1538-
7445.SABCS19-P3-09-19 (2020).
94. Sun, K. et al. A phase II trial of stereotactic radiation therapy and in situ oncolytic
virus therapy in metastatic triple-negative breast cancer (mTNBC) patients fol-
lowed by pembrolizumab (STOMP). J. Clin. Oncol. 39, 1079–1079 (2021).
95. Spicer, J. F. et al. LTX-315, an oncolytic peptide, to convert immunogenically
‘cold’ tumors to ‘hot’ in patients with advanced or metastatic tumors: results
from an ongoing phase I study. J. Clin. Oncol. 35, 3085–3085 (2017).
96. Camilio, K. A. et al. Combining the oncolytic peptide LTX-315 with doxorubicin
therapy in metastatic triple-negative breast cancer (mTNBC) patients fol-
lowed by pembrolizumab (STOMP). J. Clin. Oncol. 39, 1079–1079 (2021).
97. Bernstein, V. et al. A randomized phase II study of weekly paclitaxel with or
without palbociclib in patients with metastatic breast cancer: final analysis of
Canadian Cancer Trials Group IND213. Breast Cancer Res. Treat. 167, 485–493
(2018).
98. Manso, L. et al. Abstract PS12-08: a window-of-opportunity study with atezoli-
99. Nokisalmi, P. et al. Oncolytic adenovirus ICOVIR-7 in patients with advanced and
metastatic triple-negative breast cancer. Cancer Res. Treat. 51, 218–218 (2016).
100. Bronte, S. et al. Oncolytic virotherapy for treatment of breast cancer,
including triple-negative breast cancer. Oncoimmunology 5, e1078057–e1078057 (2015).

Published in partnership with the Breast Cancer Research Foundation
npj Breast Cancer (2022) 121
101. SM, O. B. & Mathis, J. M. Oncolytic virotherapy for breast cancer treatment. Curr. Gene Ther. 18, 192–205 (2018).
102. Berraondo, P. et al. Cytokines in clinical cancer immunotherapy. Br. J. Cancer 120, 6–15 (2019).
103. Siegel, J. P. & Puri, R. K. Interleukin-2 toxicity. J. Clin. Oncol. 9, 694–704 (1991).
104. Lasek, W., Zagórzon, R. & Jakobsiak, M. Interleukin 12: still a promising candidate for tumor immunotherapy. Cancer Immunol. Immunother. 63, 419–435 (2014).
105. Divino, C. M. et al. Anti-tumor immunity induced by interleukin-12 gene therapy in a metastatic model of breast cancer is mediated by natural killer cells. Breast Cancer Res. Treat. 60, 129–134 (2000).
106. Telli, M. L. et al. Intratumoral plasmid IL12 expands CD8+ T cells and induces a CXCR3 gene signature in triple-negative breast tumors that sensitizes patients to anti-PD-1 therapy. Clin. Cancer Res. 27, 2481–2493 (2021).
107. Ribas, A. et al. SD-101 in combination with pembrolizumab in advanced or metastatic triple-negative breast cancer (mTNBC) (KEYNOTE-890/2545). Cancer Res. 80, P3-09-04-P03-09-04 (2020).
108. Bernatchez, C. et al. Effect of a novel IL-2 cytokine immune agonist (NKTR-214) on proliferating CD8+ T cells and PD-1 expression on immune cells in the tumor microenvironment in patients with prior checkpoint therapy. J. Clin. Oncol. 35, 2545–2545 (2017).
109. Diab, A. et al. Bempegaldesleukin (NKTR-214) plus Nivolumab in patients with advanced solid tumors: phase 1 dose-escalation study of safety, efficacy, and immune activation (PIVOT-02). Cancer Discov. 10, 1158–1173 (2020).
110. Gatti-Mays, M. E. et al. If we build it they will come: targeting the immune microenvironment in patients with inoperable locally advanced or metastatic triple-negative breast cancer (mTNBC) (KEYNOTE-890/2545). Cancer Res. 80, P3-09-04-P03-09-04 (2020).
111. Gatti-Mays, M. E. et al. Abstract P3-09-04: phase 2, open-label study of intratumoral tavokinogene telseplasmid (tavo) plus electroporation in combination with intravenous pembrolizumab therapy in patients with inoperable locally advanced or metastatic triple-negative breast cancer (mTNBC). J. Clin. Oncol. 37, 2493 (2019).
112. Ribas, A. et al. SD-101 in combination with pembrolizumab in advanced melanoma: results of a phase Ib multicenter study. Cancer Discov. 8, 1250 (2018).
113. Vlegelm, A. E. & Richmond, A. Chemokines modulate immune surveillance in tumorigenesis, metastasis, and response to immunotherapy. Front. Immunol. 10, https://doi.org/10.3389/fimmu.2019.00333 (2019).
114. Francis, L. et al. Modulation of chemokines in the tumor microenvironment enhances oncolytic virotherapy for colorectal cancer. Oncotarget 7, 22174–22185 (2016).
115. Muthuswamy, R. et al. NF-kB hyperactivation in tumor tissues allows tumor-selective reprogramming of the chemokine microenvironment to enhance the recruitment of cytolytic T effector cells. Cancer Res. 72, 3735–3743 (2012).
116. Obermajer, N. et al. Promoting the accumulation of tumor-specific T cells in tumor tissues by dendritic cell vaccines and chemokine-modulating agents. Nat. Protoc. 13, 335–357 (2018).
117. Theodoraki, M. N. et al. Helicase-driven activation of Nfkb-COX2 pathway mediates the immunosuppressive component of dsRNA-driven inflammation in the human tumor microenvironment. Cancer Res. 78, 4292–4302 (2018).

AUTHOR CONTRIBUTIONS
The study was designed, directed, and coordinated by Y.A., A.G., B.V. and L.A.C. J.X.Y. and S.U. explored the landscape of IO agents being developed for use in breast cancer, specifically TNBC, facilitated the data pulls and developed Figs. 1 and 2. Y.A. and A.G. wrote the manuscript, which was reviewed, revised, and improved by all authors. All authors read and approved the final manuscript.

COMPETING INTERESTS
The authors declare no competing interests.

ADDITIONAL INFORMATION
Correspondence and requests for materials should be addressed to Yara Abdou.

Reprints and permission information is available at http://www.nature.com/reprints

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

© The Author(s) 2022

npj Breast Cancer (2022) 121
Published in partnership with the Breast Cancer Research Foundation