Characteristics and Clinical Implication of UGT1A1 Heterozygous Mutation in Tumor

Qian LI1, Tao SUN2, Hua ZHANG3, Wei LIU4, Yu XIAO1, Hongqi SUN3, Wencheng YIN1, Yanhong YAO1, Yangchun GU1, Yan'e LIU1, Fumei YI1, Qiqi WANG1, Jinyu YU1, Baoshan CAO1, Li LIANG1

1Department of Medical Oncology and Radiation Sickness; 2Department of General Surgery; 3Research Center of Clinical Epidemiology; 4Department of Pharmacy, Peking University Third Hospital, Beijing 100191, China

Abstract

Background: The literature recommends that reduced dosage of CPT-11 should be applied in patients with UGT1A1 homozygous mutations, but the impact of UGT1A1 heterozygous mutations on the adverse reactions of CPT-11 is still not fully clear.

Methods: A total of 107 patients with UGT1A1 heterozygous mutation or wild-type, who were treated with CPT-11 from January 2018 to September 2021 in Peking University Third Hospital, were retrospectively enrolled. The adverse reaction spectra of patients with UGT1A1*6 and UGT1A1*28 mutations were analyzed. Adverse reactions were evaluated according to National Cancer Institute Common Terminology Criteria for Adverse Events (NCI-CTCAE) 5.0. The efficacy was evaluated according to Response Evaluation Criteria in Solid Tumors (RECIST) 1.1. The genotypes of UGT1A1*6 and UGT1A1*28 were detected by digital fluorescence molecular hybridization.

Results: There were 43 patients with UGT1A1*6 heterozygous mutation, 26 patients with UGT1A1*28 heterozygous mutation, 8 patients with UGT1A1*6 and UGT1A1*28 double heterozygous mutations, 61 patients with heterozygous mutation at any gene locus of UGT1A1*6 and UGT1A1*28. Logistic regression analysis showed that the presence or absence of vomiting ($P=0.013$) and mucositis ($P=0.005$) was significantly correlated with heterozygous mutation of UGT1A1*28, and the severity of vomiting ($P<0.001$) and neutropenia ($P=0.021$) were significantly correlated with heterozygous mutation of UGT1A1*6. In colorectal cancer, UGT1A1*6 was significantly correlated to diarrhea ($P=0.005$), and the other adverse reactions spectrum was similar to that of the whole patient cohort, and efficacy and prognosis were similar between patients with different genotypes and patients treated with reduced CPT-11 dosage or not.

Conclusion: In clinical use, heterozygous mutations of UGT1A1*6 and UGT1A1*28 are related to the risk and severity of vomiting, diarrhea, neutropenia and mucositis in patients with Pan-tumor and colorectal cancer post CPT-11 therapy. In colorectal cancer, UGT1A1*6 is significantly related to diarrhea post CPT-11 use, efficacy and prognosis is not affected by various genotypes or CPT-11 dosage reduction.

Keywords UGT1A1; Heterozygous mutation; Adverse reaction

Introduction

There are more than 50 kinds of polymorphisms in the UGT1A1 gene loci. Among them, the relationship between UGT1A1*28 and UGT1A1*6 gene polymorphisms and CPT-11 chemotherapy-related adverse reactions and efficacy is of particular concern[1]. SN-38 is the active metabolite of CPT-11. Studies have shown that some UGT1A1 genotypes might affect the pharmacokinetics of SN-38 and therefore relate to CPT-11 induced drug toxicity reactions[2].

Previous studies have found that at the same dose of CPT-11, patients with genetic mutations of UGT1A1*28 and UGT1A1*6 might face increased risk of neutropenia and diarrhea, especially in patients with homozygous mutations of UGT1A1*28 and UGT1A1*6, and in that case the dose of CPT-11 needs to be reduced. It has been reported that the gene activity will be reduced up to 70% by homozygous mutation of UGT1A1, and up to 30% by heterozygous mutation of UGT1A1. The dose reduction standard for CPT-11 is about 25%-30% in patients with homozygous mutations of UGT1A1[3,4]. Patients with heterozygous mutations of UGT1A1 were reported to have fewer adverse effects to CPT-11 than patients with homozygous mutations, but the incidence of adverse effects post CPT-11 were still
higher than in wild-type patients\(^6\). Moreover, it was reported that the adverse effects to CPT-11 differed among patients with heterozygous mutations of UGT1A1\(^6-8\), and in patients with heterozygous mutation of UGT1A1 at different gene loci\(^9,10\). Thus, it remains elusive whether dose reduction of CPT-11 should be applied to patients with heterozygous mutation of UGT1A1\(^11,13\), and if dose reduction of CPT-11 would affect the efficacy and prognosis in these patients\(^14\).

Therefore, this study was designed to observe the difference in the adverse reaction spectrum to CPT-11 in panneoplastic and colorectal cancer patients with heterozygous mutation of UGT1A1 and the impact of dose reduction of CPT-11 on efficacy and prognosis in these patients.

Materials and methods

Patient cohort

A total of 107 patients who were treated with CPT-11 in the Peking University Third Hospital from January 2018 to September 2021, and underwent UGT1A1 genotyping screen before CPT-11, were enrolled. The inclusion criteria were: (1) Malignant tumors diagnosed by pathology, including colorectal cancer, lung cancer, gastric cancer and cholangiopancreatic cancer; (2) Eastern Cooperative Oncology Group (ECOG) performance status (PS) of 0-2; (3) The expected survival was more than three months with measurable lesions; (4) Aged between 18 and 80 years old; (5) No severe hepatic and renal insufficiency; (6) UGT1A1*6 or UGT1A1*28 heterozygous mutations or wild-type. The specific chemotherapy protocols were FOLFIRI/xeliri/CPT-11 for colorectal cancer, IP/CPT-11 for lung cancer, CPT-11 single drug±Apatinib for gastric cancer and CPT-11 for cholangiopancreatic cancer. The standard dose of the various protocols was decided according to National Comprehensive Cancer Network (NCCN) and Chinese Society of Clinical Oncology (CSCO) guidelines. The weekly dose intensity of CPT-11 did not exceed 90 mg/m\(^2\). For patients with serious adverse reactions or an initial ECOG score of 2 after the first cycle of chemotherapy, 10%-20% dose reduction of CPT-11 was applied. The exclusion criteria included infection, uncontrolled cardiovascular and cerebrovascular diseases, chronic diarrhea, bone marrow metastasis, lactation and pregnancy, UGT1A1*6/*28 homozygous mutations and double primary carcinoma. Patients were followed up by telephone call or clinical visit until October 2021. The study protocol passed the ethical review of the Peking University Third Hospital.

Efficacy and toxicity assessment

Data including ECOG score, hyperbilirubinemia, smoking, gender, age, tumor type, number of treatment lines, treatment plan, CPT-11 combination or single drug, CPT-11 dose reduction and the amount of dose reduction of CPT-11 were obtained. Related adverse reactions included vomiting, mucositis, neutropenia, diarrhea, anemia, thrombocytopenia and fatigue were analyzed. Adverse reactions were evaluated according to National Cancer Institute Common Terminology Criteria for Adverse Events (NCI-CTCAE) 5.0. The efficacy was evaluated according to the Response Evaluation Criteria in Solid Tumors (RECIST) 1.1. Prognostic indicators include progression-free survival (PFS), which is defined as the time from the beginning of treatment to tumor progression or death. Overall survival (OS) is defined as the time from diagnosis to death due to any cause.

Genotyping of UGT1A1*6 and UGT1A1*28

Whole blood samples were collected using ethylenediamine tetraacetic acid (EDTA) as an anticoagulant. Then, 200 μL of the blood sample was transferred to an Eppendorf (EP) tube, and ammonium chloride solution (Beijing Huaxia Times Gene Technology Development Co., Ltd.) was used for blood sample pretreatment for 5 min. The pretreated specimens were centrifuged at 700 × g for 5 min to obtain precipitated blood cells. Genomic DNA was extracted directly from the above precipitated blood cells using a nucleic acid purification kit (Beijing Huaxia Times Gene Technology Development Co., Ltd.) and vibrated for 1 min. After standing for 15 min, the universal kit for sequencing the reaction of a single gene locus (Beijing Huaxia Times Gene Technology Development Co., Ltd.) was used to detect the UGT1A1 genotype (UGT1A1*6 and UGT1A1*28 gene polymorphisms).

Data statistics

Hardy-Weinberg equilibrium test was used for genetic epidemiology. For continuous (quantitative) data, the Shapiro normality test was used to determine the normality of the sample data. If it conformed to a normal distribution, it was expressed as the Mean±SD. Student’s t tests was used for comparisons between the two groups. If the data did not conform to a normal distribution, they were expressed as the median (25% quantile, 75% quantile), and Wilcoxon test was used for the comparisons between the two groups. Classified (qualitative) data were statistically described by frequency (percentage) and compared by \(\chi^2\) test or Fisher’s exact test. In the multivariate analysis of the association between adverse reactions and genotype mutations, GLM function was used for binary Logistic regression analysis. Each adverse reaction was taken as the dependent variable, and UGT1A1*6 mutation, UGT1A1*28 mutation, number of treatment lines, treatment regimen, smoking, gender, tumor type and hyperbilirubinemia were taken as the independent variables. When the independent variables were categorical
classification variables, the minimum value was taken as the reference. When the independent variables were continuous variables, the continuous variables were directly included in the binary Logistic regression model. Two-tailed $P<0.05$ was considered to be statistically significant.

**Results**

There were 66 cases of colorectal cancer, 16 cases of lung cancer, 15 cases of biliary and pancreatic tumors and 10 cases of gastric cancer in this cohort. There were 43 patients with UGT1A1*6 heterozygous mutation, 26 patients with UGT1A1*28 heterozygous mutation, 8 patients with UGT1A1*6 and UGT1A1*28 double heterozygous mutations, 61 patients with heterozygous mutation at any gene locus of UGT1A1*6 and UGT1A1*28. There were 10 patients with wild-type, 41 with heterozygous mutation and 56 homozygous mutation of SLCO1B1 gene. There were 46 patients receiving the first-line, 42 patients receiving the second-line, and 19 patients receiving the ≥third-line chemotherapy in this cohort. There were 85 patients treated with CPT-11 combined regimen and 22 patients with single CPT-1 regimen (Tab 1).

**Adverse reaction spectrum and baseline characteristics of patients with heterozygous mutation of UGT1A1*6 and UGT1A1*28**

In this study, the heterozygous mutation rates of UGT1A1*6 and UGT1A1*28 were 40.2% (43/107) and 24.3% (26/107), respectively. Results showed that heterozygous mutation of these two genes were significantly correlated with the occurrence and severity of multiple common clinical adverse reactions post CPT-11 treatment, such as vomiting, neutropenia, diarrhea and mucositis (Tab 2). Multivariate regression analysis showed that heterozygous mutation of these two genes were related to vomiting, diarrhea, neutropenia and mucositis post CPT-11 use (Tab 3).

According to the statistical results of the above baseline tables, the UGT1A1*6 mutation is related to hyperbilirubinemia, the amount of CPT-11 dose reduction and whether dose reduction of CPT-11 took place, the degree of vomiting, the severity of diarrhea and neutropenia. These results show that the UGT1A1*6 mutation may be more likely to cause vomiting, diarrhea and neutropenia, which can be used as an important index to guide the CPT-11 dose reduction. At the same time, patients with UGT1A1*6 mutations are also more prone to concurrent hyperbilirubinemia. UGT1A1*28 is related to the severity of vomiting and mucositis.

**Multivariate analysis of common adverse reactions in patients with heterozygous mutation of UGT1A1*6 and UGT1A1*28**

Multifactorial regression analysis showed that after adjusting the cofounders, UGT1A1*28 remained as an independent risk factor for vomiting and mucositis, and UGT1A1*6 was the independent risk factor for the severity of vomiting and degree of neutropenia (Tab 3).

**Analysis of adverse reactions and prognosis in colorectal cancer in patients with heterozygous mutation of UGT1A1*6 and UGT1A1*28**

To eliminate the confounding factors caused by the difference in tumor species, the researchers further analyzed the features of patients with single colorectal cancer disease, which accounted for the highest proportion in this cohort. After adjusting the six variables including treatment line, treatment regimen, smoking, gender, tumor type and hyperbilirubinemia, multivariate Logistic regression analysis showed that UGT1A1*6 was significantly correlated with the presence or absence of vomiting, the severity of vomiting and diarrhea, and UGT1A1*28 was significantly correlated with the severity of vomiting (Tab 4). At the same time, multivariate Cox regression analysis of prognosis and survival of PFS (Tab 5) and OS (Tab 6) in colorectal cancer was carried out. Results showed that genotype, CPT-11 reduction degree or other factors did not affect prognosis and PFS in this patient cohort.

**Discussion**

CPT-11 has been approved as a first-line or second-line drug for colorectal cancer, lung cancer, pancreatic cancer, esophageal cancer, gastric cancer and other tumor types. Common adverse reactions include neutropenia and diarrhea. It can lead to interruption or cessation of the treatment, thereby jeopardizing the patient’s prognosis and quality of life. Most of the metabolites of CPT-11, SN-38, are detoxified by uridine diphosphate-glucuronyl transferase (mainly UGT1A1). In patients with UGT1A1 mutations, the detoxification process slows down, and adverse reactions might increase. Therefore, UGT1A1 has been increasingly used as a reference target for dosage formulations in clinical use[15]. Many previous clinical studies regarding the clinical use of CPT-11 came from European and American patients, but results from Asian patients are scarce. It is known that there is a significant difference in the epidemiological distribution of UGT1A1*6 and UGT1A1*28 between Asians and Caucasians. The UGT1A1*28 polymorphism in the Caucasian population accounts for 30%-45%, and among East Asians, it accounts for only 10%-20%. Among Asians, the UGT1A1*6 polymorphism accounts for 16%-40%[16].
Tab 1 Baseline characteristics of UGT1A1*6 and UGT1A1*28

| Characteristics       | UGT1A1*6 |   | UGT1A1*28 |   |
|-----------------------|----------|---|-----------|---|
|                       | WT (n=64) |   | HM (n=43) |   |
| Gender                | 0.124    |   | 0.129     |   |
| Male                  | 29 (45.31%) |   | 45 (55.56%) |   |
| Female                | 35 (54.69%) |   | 36 (44.44%) |   |
| Smoking history       | 0.587    |   | 0.489     |   |
| Yes                   | 22 (34.38%) |   | 31 (38.27%) |   |
| No                    | 42 (65.62%) |   | 50 (61.73%) |   |
| Hyperbilirubinemia    | <0.001   |   | 0.060     |   |
| Yes                   | 4 (6.25%)  |   | 10 (12.35%) |   |
| No                    | 60 (93.75%) |   | 71 (87.65%) |   |
| SLCO1B1               | 0.742    |   | 0.260     |   |
| WT                    | 5 (7.81%)  |   | 6 (7.41%)  |   |
| HM                    | 24 (37.50%) |   | 34 (41.98%) |   |
| M                     | 35 (54.69%) |   | 41 (50.62%) |   |
| CRC                   | 0.549    |   | 0.986     |   |
| Yes                   | 38 (59.38%) |   | 50 (61.73%) |   |
| No                    | 26 (40.62%) |   | 31 (38.27%) |   |
| Treatment lines       | 0.830    |   | 0.635     |   |
| First-line            | 26 (40.62%) |   | 34 (41.98%) |   |
| Second-line           | 26 (40.62%) |   | 31 (38.27%) |   |
| Third-line            | 12 (18.75%) |   | 16 (19.75%) |   |
| Regimen               | 0.682    |   | 0.191     |   |
| Single                | 14 (21.88%) |   | 19 (23.46%) |   |
| Combined              | 50 (78.12%) |   | 62 (76.54%) |   |
| Dose decrease         | 0.043    |   | 0.619     |   |
| <10%                  | 49 (76.56%) |   | 55 (67.90%) |   |
| >10%                  | 15 (23.44%) |   | 26 (32.10%) |   |
| Dose decrease         | 0.003    |   | 0.321     |   |
| Yes                   | 35 (54.69%) |   | 37 (45.68%) |   |
| No                    | 29 (45.31%) |   | 44 (54.32%) |   |

*WT: wild-type; HM: heterozygous mutation; M: homozygous mutation; CRC: colorectal cancer.

Therefore, more data from Asian patients are needed to better guide the clinical use of CPT-11 in Asian patients.

In this study, patients with heterozygous mutations in UGT1A1 and treated with CPT-11 were included to explore the relationship between genotype and the characteristics of adverse reactions in these patients. We mainly found four points: First, the population in this cohort were all patients with stage III inoperable or stage IV cancer. Results showed that UGT1A1*6 mutations are related to hyperbilirubinemia, how much and whether to reduce the dose, the degree of vomiting, the severity of diarrhea, and the severity of neutropenia, which shows that the patients with UGT1A1*6 heterogeneous mutation may be more likely to cause severe vomiting, diarrhea, and neutropenia, as well as hyperbilirubinemia. Thus, like for UGT1A1*6 homogenous patients, patients with UGT1A1*6 heterogeneous mutation might also undergoing dose reduction of CPT-11. UGT1A1*6 heterogeneous mutation can be used as an important indicator of decision making on dose reduction of CPT-11 and how much should be reduced. UGT1A1*28 heterogeneous mutation is also related to the severity of vomiting and mucositis. Second, multivariate Logistic regression analysis showed that the presence or absence of vomiting and mucositis was more related to the UGT1A1*28 heterogeneous mutation, and severe vomiting and severe neutropenia were more common in patients with heterogeneous mutation of UGT1A1*6. Third, to further clarify whether there is a difference in the adverse reaction
Tab 2  Adverse reaction spectrum of UGT1A1*6 and UGT1A1*28

| Adverse reaction | UGT1A1*6          | P         | UGT1A1*28          | P         |
|------------------|-------------------|-----------|-------------------|-----------|
|                  | WT (n=64) HM (n=43) |           | WT (n=81) HM (n=26) |           |
| Vomit grade      |                   |           |                   |           |
| 0                | 30 (46.88%) 16 (37.21%) | 0.020     | 41 (50.62%) 5 (19.23%) | 0.013     |
| 1-2              | 25 (39.06%) 11 (25.58%) |           | 25 (30.86%) 11 (42.31%) |           |
| 3-4              | 9 (14.06%) 16 (37.21%) |           | 15 (18.52%) 10 (38.46%) |           |
| Mucositis        |                   |           |                   |           |
| Yes              | 46 (71.88%) 27 (46.88%) | 0.322     | 61 (75.31%) 12 (46.15%) | 0.005     |
| No               | 18 (28.12%) 16 (37.21%) |           | 20 (24.69%) 14 (53.85%) |           |
| Diarrhea grade   |                   |           |                   |           |
| 0-2              | 50 (78.12%) 25 (58.14%) | 0.027     | 55 (67.90%) 20 (76.92%) | 0.382     |
| 3-4              | 14 (21.88%) 18 (41.86%) |           | 26 (32.10%) 6 (23.08%) |           |
| Neutropenia grade|                   |           |                   |           |
| 0-2              | 43 (67.19%) 17 (39.53%) | 0.005     | 48 (59.26%) 12 (46.15%) | 0.241     |
| 3-4              | 21 (32.81%) 26 (60.47%) |           | 33 (40.74%) 14 (53.85%) |           |
| Anemia           |                   |           |                   |           |
| Yes              | 53 (82.81%) 37 (86.05%) | 0.654     | 69 (85.19%) 21 (80.77%) | 0.820     |
| No               | 11 (17.19%) 6 (13.95%) |           | 12 (14.81%) 5 (19.23%) |           |
| Thrombocytopenia |                   |           |                   |           |
| Yes              | 54 (84.38%) 34 (79.07%) | 0.481     | 65 (80.25%) 23 (88.46%) | 0.510     |
| No               | 10 (15.62%) 9 (20.93%) |           | 16 (19.75%) 3 (11.54%) |           |
| Fatigue          |                   |           |                   |           |
| Yes              | 53 (82.81%) 37 (86.05%) | 0.654     | 67 (82.72%) 23 (88.46%) | 0.697     |
| No               | 11 (17.19%) 6 (13.95%) |           | 14 (17.28%) 3 (11.54%) |           |

Tab 3  Logistic regression of common adverse reactions

| Adverse events | Gene type | B    | SE   | Z    | P     | OR (95%CI) |
|----------------|-----------|------|------|------|-------|------------|
| *Vomit         | UGT1A1*6  | 0.524| 0.438| 1.198| 0.231 | 1.69 (0.72-3.99) |
|                | UGT1A1*28 | 1.491| 0.568| 2.623| 0.009 | 4.44 (1.46-13.52) |
| *Vomit grade   | UGT1A1*6  | 1.921| 0.573| 3.353| 0.001 | 6.83 (2.22-20.98) |
|                | UGT1A1*28 | 1.081| 0.577| 1.874| 0.061 | 2.95 (0.95-9.14)  |
| *Diarrhea      | UGT1A1*6  | 0.128| 0.453| 0.283| 0.777 | 1.14 (0.47-2.76)  |
|                | UGT1A1*28 | -0.474| 0.499| -0.949| 0.343 | 0.62 (0.23-1.66)  |
| *Mucositis     | UGT1A1*6  | 0.601| 0.447| 1.346| 0.178 | 1.82 (0.76-4.38)  |
|                | UGT1A1*28 | 1.371| 0.484| 2.831| 0.005 | 3.94 (1.52-10.17)|
| *Neutropenia grade | UGT1A1*6 | 1.057| 0.459| 2.303| 0.021 | 2.88 (1.17-7.08)  |
|                | UGT1A1*28 | 0.575| 0.506| 1.136| 0.256 | 1.78 (0.66-4.79)  |

*After adjusting the number of treatment lines, treatment plan, smoking, gender, tumor type and hyperbilirubinemia.

spectrum between colorectal cancer and panneoplastic species, we performed multivariate Logistic regression analysis on 107 panneoplastic patients and 66 patients with simple colorectal cancer after adjusting for multiple variables. We found that the overall adverse reaction spectrum was similar in pantumor species and colorectal cancer, while in colorectal cancer, the UGT1A1*6 mutation is highly correlated with the severity of diarrhea, and in pantumor species, among them, UGT1A1*28 is significantly related to mucositis. Fourth, the efficacy and prognosis of mutant patients were similar from those of wild-type patients in colorectal cancer.

In terms of the types of adverse reactions, there have been numerous reports in the literature about the increased risk of diarrhea and neutropenia in patients with gene mutations. Previous studies have found that diarrhea was significantly
### Tab 4 Logistic regression analysis of adverse reactions in patients with colorectal cancer

| Adverse events                  | Gene type | B   | SE  | Z   | P       | OR (95% CI)     |
|---------------------------------|-----------|-----|-----|-----|---------|-----------------|
| *Vomit                           |           |     |     |     |         |                 |
|                                 | UGT1A1*6  | 1.250 | 0.590 | 2.118 | 0.034 | 3.49 (1.10-11.10) |
|                                 | UGT1A1*28 | 1.029 | 0.680 | 1.513 | 0.130 | 2.80 (0.74-10.62) |
| *Vomit grade                     |           |     |     |     |         |                 |
|                                 | UGT1A1*6  | 2.236 | 0.786 | 2.843 | 0.004 | 9.36 (2.00-43.70) |
|                                 | UGT1A1*28 | 1.481 | 0.761 | 1.945 | 0.052 | 4.40 (0.99-19.55) |
| *Diarrhea grade                  |           |     |     |     |         |                 |
|                                 | UGT1A1*6  | 2.166 | 0.767 | 2.824 | 0.005 | 8.72 (1.94-39.22) |
|                                 | UGT1A1*28 | -0.899 | 0.963 | -0.933 | 0.351 | 0.41 (0.06-2.69) |
| *Neutropenia grade               |           |     |     |     |         |                 |
|                                 | UGT1A1*6  | 1.484 | 0.595 | 2.492 | 0.013 | 4.41 (1.37-14.16) |
|                                 | UGT1A1*28 | 0.825 | 0.668 | 1.235 | 0.217 | 2.28 (0.62-8.44) |

*After adjusting the number of treatment lines, treatment plan, smoking, gender, tumor type and hyperbilirubinemia.

### Tab 5 Results of Cox regression multivariate analysis of progression-free survival

| Independent variable | B   | SE  | Z   | P       | HR (95% CI)     |
|----------------------|-----|-----|-----|---------|-----------------|
| Gender               | 0.132 | 0.433 | 0.304 | 0.761 | 1.14 (0.49-2.67) |
| Hyperbilirubinemia   | 0.137 | 0.719 | 0.190 | 0.849 | 1.15 (0.28-4.69) |
| Complication         | -0.581 | 0.537 | -1.082 | 0.279 | 0.56 (0.20-1.60) |
| Metastasis           | 0.064 | 0.550 | 0.117 | 0.907 | 1.07 (0.36-3.13) |

**Regimen**

- CPT-11: Ref.
- FOLFIRI: -1.238 | 0.689 | -1.796 | 0.073 | 0.29 (0.08-1.12)
- XELIRI: -0.599 | 0.671 | -0.893 | 0.372 | 0.55 (0.15-2.05)

**Age**

- -0.576 | 0.468 | -1.232 | 0.218 | 0.56 (0.22-1.41)
- UGT1A1*6: 0.209 | 0.514 | 0.408 | 0.683 | 1.23 (0.45-3.37)
- UGT1A1*28: 0.724 | 0.521 | 1.391 | 0.164 | 2.06 (0.74-5.72)
- First-line: 1.118 | 0.657 | 1.702 | 0.089 | 3.06 (0.84-11.80)
- ≥Second-line: 1.665 | 1.299 | 1.282 | 0.200 | 5.29 (0.41-67.70)
- Dose decrease: -0.556 | 0.629 | -0.884 | 0.377 | 0.57 (0.17-1.97)

### Tab 6 Multivariate analysis results of Cox regression for overall survival

| Independent variable | B   | SE  | Z   | P       | HR (95% CI)     |
|----------------------|-----|-----|-----|---------|-----------------|
| Gender               | -1.495 | 1.252 | -1.194 | 0.232 | 0.22 (0.02-2.61) |
| Hyperbilirubinemia   | 0.371 | 1.356 | 0.274 | 0.784 | 1.45 (0.10-20.66) |
| Complication         | 2.529 | 2.334 | 1.084 | 0.279 | 12.54 (0.13-1,216.22) |
| Metastasis           | 3.008 | 1.767 | 1.702 | 0.089 | 20.24 (0.63-646.48) |

**Regimen**

- CPT-11: Ref.
- FOLFIRI: -3.384 | 1.645 | -2.057 | 0.040 | 0.03 (0.00-0.85)
- XELIRI: -3.252 | 1.577 | -2.062 | 0.039 | 0.04 (0.00-0.85)

**Age**

- -1.284 | 1.309 | -0.981 | 0.326 | 0.28 (0.02-3.60)
- UGT1A1*6: -0.942 | 1.261 | -0.747 | 0.455 | 0.39 (0.03-4.61)
- UGT1A1*28: 1.058 | 1.201 | 0.881 | 0.378 | 2.88 (0.27-30.29)
- Dose decrease: 0.441 | 1.938 | 0.228 | 0.820 | 1.55 (0.03-69.37)
associated with UGT1A1*28 homozygous mutations\cite{17,18,19}, in the high/medium dose group (>125 mg/m²) of CPT-11, but not in the low-dose (<125 mg/m²) group\cite{20}. A study evaluating the combination of irinotecan and platinum in the treatment of colorectal cancer reported an increased risk of severe diarrhea in patients with UGT1A1*6 alleles\cite{21}. Among white people, risk of neutropenia was high in patients with UGT1A1*28 genotype mutations\cite{22}. In Asians, UGT1A1*6 alleles are reported to be associated with severe hematological toxicity\cite{23}. Studies also demonstrated that in Asian cancer patients, the combination of UGT1A1*6 and UGT1A1*28 polymorphisms was associated with an increased risk of IRI-induced neutropenia\cite{6}. A meta-analysis showed that Asians with lung cancer and UGT1A1*6 genetic polymorphism faced a higher risk of neutropenia and diarrhea after IRI chemotherapy. However, UGT1A1*28 is weakly correlated with diarrhea and neutropenia\cite{24}. However, there are very few reports in the literature about the risk of mucositis and vomiting, and only a few studies are involved\cite{25}. Our research found an increased risk of vomiting and mucositis, which may indicate that in patients with UGT1A1 mutations, we should pay more attention to these two often overlooked adverse reactions related to CPT-11 use in daily clinical practice.

In this study, we also found that UGT1A1*6 is significantly correlated with the increased risk of bilirubin, which is mainly because the metabolism of bilirubin also depends on this gene. This is consistent with the literature report\cite{26}. A number of previous studies have shown that homozygous UGT1A1*28 and homozygous UGT1A1*6 are associated with an increased risk of hyperbilirubinemia\cite{27,28}. It was shown that patients with total bilirubin levels between 1.0 mg/dL and 2.0 mg/dL faced higher risks of grade 3-4 neutropenia. Perhaps in the future, if there is no chance to detect UGT1A1 genotype, clinicians can judge the potential risk of adverse reaction post CPT-11 use through elevating the bilirubin level, patients with elevated bilirubin should be monitored more cautiously during CPT-11 therapy.

Some studies suggest that only high doses of CPT-11 could induce severe adverse effects in patients with UGT1A1 polymorphisms. Especially when the dose is ≥180 mg/m², UGT1A1*28 is related to neutropenia and diarrhea. At normal doses, the alarming role of UGT1A1*6 on neutropenia and diarrhea is greater than UGT1A1*28\cite{29}. This is consistent with our findings. Although high doses CPT-11 would certainly relate to a higher risk of adverse drug effects in patients with UGT1A1 mutations, the achieved benefit might also greater. A study showed that patients with UGT1A1*1/*1 and UGT1A1*1/*28 genotypes could tolerate high doses of CPT-11 and more favorable objective response rate (ORR) was achieved without significant adverse events\cite{30}. Another study found that the dose titration plan of CPT-11 is a satisfactory option for achieving favorable ORR and disease control rate (DCR). Metastatic colorectal cancer patients with the UGT1A1 wild-type genotype can tolerate higher doses of CPT-11 and may obtain more favorable clinical results without significantly increased toxicity. Therefore, under the guidance of UGT1A1 gene detection, use of CPT-11 could be optimized in daily clinical practice.

Most studies have failed to observe the correlation between UGT1A1 genotype and prognosis\cite{31}. However, some studies found that the prognosis is more favorable in patients with mutations. In a study on small cell lung cancer, it was shown that UGT1A1*6 and UGT1A1*28 polymorphisms were associated with increased gastrointestinal toxicity and improved OS, respectively\cite{32,33}. In colorectal cancer, good prognosis was evidenced in patients with the UGT1A1*28 polymorphism\cite{34}. The possible mechanism may be that patients with gene mutations had higher blood drug concentration at the first 1.5 h timepoint\cite{35}, thereby achieving higher efficacy. However, there are some opposite findings in that prognosis is better in wild-type patients. For example, in the third-line treatment of gastric cancer, it was found that patients with wide type UGT1A1 had better OS, DCR and time to progress (TTP) than mutant patients\cite{36}. The possible mechanism may be that higher doses tolerated by these patients might be linked with the better prognosis.

In our study, no difference was found in prognosis between mutant and wild-type patients. The possible reasons might be as follows: (1) A better prognosis in colorectal cancer comes from the exploration of larger doses. In our research, we used conventional standard doses; (2) The types of tumors that suggest a better prognosis in conventional doses are mostly reflected in patients with nonintestinal cancer. This study mainly included patients with intestinal cancer, so there may be differences in tumor types; (3) In our clinical practice, most of the patients with mutations were forced to reduce the dose due to severe adverse reactions, but the prognosis was not affected in the end. This may be because the mutation itself is a protective factor for the prognosis, neutralizing the reduction. The unfavorable factors brought about, so overall no difference in prognosis was found; (4) There is a need for longer-term follow-up and a larger sample size.

Some people think that routine testing of CPT-11 for the gene before use will increase the economic burden of patients. However, from an economic perspective, testing the gene can save costs. Related pharmacoeconomic studies have shown that compared with no genotyping, genetic testing only leads to a marginal increase in quality-adjusted life years, but the cost per patient is reduced by US$651.12 and US$805.22, respectively\cite{37}. One-way sensitivity analysis shows that the model is relatively robust. Therefore, UGT1A1 genotyping saves costs for Chinese patients with colorectal cancer\cite{38}.
Under the guidance of the UGT1A1 test, more bold attempts have been made in the treatment dose and program. Innocenti et al. [39] has reported that the dose of CPT-11 can be individualized according to the differential expression of the UGT1A1*28 genotype. McLeod et al. [39] found that different dose levels of CPT-11 and different combination schemes are closely related to pharmacogenomics predicting adverse reactions. Follow-up scholars have carried out a variety of explorations under the guidance of genes. For example, in the poor prognosis of BRAF mutant metastatic colorectal cancer, the dose of some patients could be increased under the guidance of genes to achieve a better curative effect [40]. Neoadjuvant chemoradiation for rectal cancer also uses genetic testing as a guide to improve pathological complete response (pCR), and the efficacy of high-dose patients is better [41]. In locally advanced gastroesophageal junction tumors, different dose gradient treatments are given according to the genotype, which improves the perioperative downstage rate and R0 resection rate [42]. Under the guidance of UGT1A1 genotype, some researchers explored the efficacy and safety of the regimen of regorafenib combined with increased-dose FOLFIRI for metastatic colorectal cancer patients [43]. In gastrointestinal cancer, giving FOLFIRABRAX to patients who can tolerate it under the guidance of genotype has achieved better efficacy [44]. Therefore, UGT1A1 detection can help us boldly explore more intensive therapy for appropriate patients.

Although a variety of studies have found that UGT1A1 affects adverse reactions, the adverse reaction spectrum and predictive value obtained from each study were different [45]. On the one hand, this may be due to different chemotherapy regimens (dose and cycle) and different tumor types. On the other hand, it may be that UGT1A1 affects the metabolism of CPT-11. The plasma area under curve (AUC) of SN38 is related to many factors, including ABCC, CYP3A, ABCB1 and CES [46-48]. Therefore, the risk of adverse reactions in our patients was largely dependent on and not only dependent on the UGT1A1 gene. The risk and degree of adverse reactions are the result of the joint action of multiple metabolic genes. The clinical judgment of whether to reduce requires a comprehensive judgment based on the actual situation of the patient.

Based on our research results, we have the following tips for clinical practice. In addition to prophylactic leukogenics and antidiarreal therapy, we also need to strengthen antiemetics. Try to give triple or even quadruple antiemetic programs. Patients with UGT1A1*28 mutations also have an increased chance of mucositis. More oral care, vitamin supplementation, etc. should be given to such patients to prevent adverse reactions to ensure that the dose completes the full course of chemotherapy. At the same time, patients with hyperbilirubinemia need to pay special attention. In patients whose UGT1A1 has not been detected clinically, if bilirubin is elevated, the occurrence of adverse reactions can be carefully prevented, and the dose should be reduced if necessary.

This study also has several limitations, such as insufficient sample size, insufficient median follow-up time, and few cases of other tumor types except for colorectal cancer. This study is a retrospective study, and there are many factors of bias. For example, there are differences in the regimen, although studies have suggested that the combination of XELIRI and FOLFIRI has similar safety and effectiveness in different genotypes [49], but there are still differences in other intraspecific research protocols. At the same time, due to the problem of sample size, it has not yet been possible to answer the reduction standard in high-risk groups. It's still not clear to determine the best range of reduction to ensure the best curative effect and minimal adverse reactions.

Conclusion

UGT1A1*6 mutations and hyperbilirubinemia often occur concurrently. UGT1A1*6 heterozygous mutations will affect the amount of dose reduction and whether it is to be reduced. At the same time, it is related to the degree of vomiting, the severity of diarrhea, and the severity of neutropenia. UGT1A1*28 is related to the severity of vomiting and mucositis. In colorectal cancer, the adverse reaction spectrum is similar to pantomoma except UGT1A1*6, which is significantly related to the severity of diarrhea. Genotype, CPT-11 reduction degree or other factors did not affect prognosis and PFS in colorectal cancer patient cohort. Since UGT1A1 is an important metabolic gene, it is not only detected in the peripheral blood of UGT1A1, but now studies are beginning to explore the distribution and role of this metabolic enzyme in organs and tissues [49]. There may be more exploratory studies on UGT1A1 in the future.

Author contributions

Li Q and Li Q conceived and designed the study. Li Q, Sun T, Xiao Y, Liu W, Yin WC, Yao YH, Gu YC, Liu Y, Yi FM, Wang QQ, Yu JY, Cao BS and Liang L performed the experiments. Zhang H and Li Q analyzed the data. Liu W and Sun HQ contributed analysis tools. Cao BS, Zhang H, Sun T, Liu W, Sun HQ and Liang L provided critical inputs on design, analysis, and interpretation of the study. All the authors had access to the data. All authors read and approved the final manuscript as submitted.

Acknowledgments

We would like to acknowledge our instructors for their
valuable advice. We also thank the reviewers for their precious comments on this paper.

Conflict of interests

The authors have no conflicts of interest to declare.

References

1 Takahara N, Nakai Y, Isayama H, et al. Uridine diphosphate glucuronosyl transferase 1 family polyphenylalanine A gene (UGT1A1) polymorphisms are associated with toxicity and efficacy in irinotecan monotherapy for refractory pancreatic cancer. Cancer Chemother Pharmacol, 2013, 71(1): 85-92. doi: 10.1007/s00280-012-1981-0
2 Innocenti F, Kroetz DL, Schultz E, et al. Comprehensive pharmacogenetic analysis of irinotecan neutropenia and pharmacokinetics. J Clin Oncol, 2009, 27(16): 2604-2614. doi: 10.1200/JCO.2008.20.6300
3 Hulshof EC, Deenen MJ, Guchelaar HJ, et al. Pre-therapeutic UGT1A1 genotyping to reduce the risk of irinotecan-induced severe toxicity: Ready for prime time. Eur J Cancer, 2020, 141: 9-20. doi: 10.1016/j.ejca.2020.09.007
4 Irinotecan Hydrochloride - irinotecan hydrochloride injection [package insert], Orlando F, USA; June 5, 2017.
5 Shirasu H, Todaka A, Omae K, et al. Impact of UGT1A1 genetic polymorphism on toxicity in unresectable pancreatic cancer patients undergoing FOLFIRINOX. Cancer Sci, 2019, 110(2): 707-716. doi: 10.1111/cas.13883
6 Han FF, Guo CL, Yu D, et al. Associations between UGT1A1*6 or UGT1A1*6/*28 polymorphisms and irinotecan-induced neutropenia in Asian cancer patients. Cancer Chemother Pharmacol, 2014, 73(4): 779-788. doi: 10.1007/s00280-014-2405-0
7 Chen YJ, Hu F, Li CY, et al. The association of UGT1A1*6 and UGT1A1*28 with irinotecan-induced neutropenia in Asians: a meta-analysis. Biomarker, 2014, 19(1): 56-62. doi: 10.3109/1354750X.2013.867534
8 Cheng L, Li M, Hu J, et al. UGT1A1 polymorphisms are correlated with irinotecan-induced toxicity: a system review and meta-analysis in Asians. Cancer Chemother Pharmacol, 2014, 73(3): 551-560. doi: 10.1007/s00280-014-2382-3
9 Hikino K, Ozeki T, Koido M, et al. Comparison of effects of UGT1A1*6 and UGT1A1*28 on irinotecan-induced adverse reactions in the Japanese population: analysis of the Biobank Japan Project. J Hum Genet, 2019, 64(12): 1195-1202. doi: 10.1038/s10038-019-0677-2
10 Chen X, Liu L, Guo Z, et al. UGT1A1 polymorphisms with irinotecan-induced toxicities and treatments in Asian with lung cancer: a meta-analysis. Cancer Chemother Pharmacol, 2017, 79(Suppl 1): 1109-1117. doi: 10.1007/s00280-017-3306-9
11 Hoskins JM, Goldberg RM, Qu P, et al. UGT1A1*28 genotype and irinotecan-induced neutropenia: dose matters. J Nat Cancer Inst, 2007, 99(17): 1290-1295. doi: 10.1093/jnci/djm115
12 Argevani L, Hughes C, Schuh MJ. Dosage adjustment of irinotecan in patients with UGT1A1 polymorphisms: a review of current literature. Innov Pharm, 2020, 11(3): 5. doi: 10.24926/ip.1113.3203
13 Konaka K, Sakurada T, Saito T, et al. Study on the optimal dose of irinotecan for patients with heterozygous uridine diphosphate-glucuronosyltransferase 1A1 (UGT1A1). Biol Pharm Bull, 2019, 42(11): 1839-1845. doi: 10.1248/bpb.b19-00357
14 Tsai HL, Huang CW, Lin YW, et al. Determination of the UGT1A1 polymorphism as guidance for irinotecan dose escalation in metastatic colorectal cancer treated with first-line bevacizumab and FOLFI Ri (PURE FIST). Eur J Cancer Care, 2020, 138: 19-29. doi: 10.1016/j.ejcc.2020.05.031
15 Yau TO. Precision treatment in colorectal cancer: Now and the future. JGH Open, 2019, 3(5): 361-369. doi: 10.1002/jgh3.12153
16 Fukui T, Mitsufuji H, Kubota M, et al. Prevalence of topoisomerase I genetic mutations and UGT1A1 polymorphisms associated with irinotecan in individuals of Asian descent. Oncol Lett, 2011, 2(5): 923-928. doi: 10.3892/oal.2011.346
17 Marcuello E, Albes A, Menoyo A, et al. UGT1A1 gene variations and irinotecan treatment in patients with metastatic colorectal cancer. Br J Cancer, 2004, 91(4): 678-682. doi: 10.1038/sj.bjc.6602042
18 Ferralideschi R, Minchell LJ, Roberts SA, et al. UGT1A1*28 genotype predicts gastrointestinal toxicity in patients treated with intermediate-dose irinotecan. Pharmacogenomics, 2009, 10(5): 733-739. doi: 10.2217/pgs.09.20
19 Martinez-Balibre E, Abad A, Martinez-Cardas A, et al. UGT1A1 and TYMS genetic variants predict toxicity and response of colorectal cancer patients treated with first-line irinotecan and fluorouracil combination therapy. Br J Cancer, 2010, 103(4): 581-589. doi: 10.1038/sj.bjc.6605776
20 Hu ZY, Yu Q, Pei Q, et al. Dose-dependent association between UGT1A1*28 genotype and irinotecan-induced neutropenia: low doses also increase risk. Clin Cancer Res, 2010, 16(15): 3832-3842. doi: 10.1158/1078-0432.CCR-10-1122
21 Liu X, Cheng D, Kuang Q, et al. Association of UGT1A1*28 polymorphisms with irinotecan-induced toxicities in colorectal cancer: a meta-analysis in Caucasians. Pharmacogenomics J, 2014, 14(2): 120-129. doi: 10.1038/tpj.2013.10
22 Schilsky RL. Personalized medicine in oncology: the future is now. Nat Rev Drug Discov, 2010, 9(5): 363-366. doi: 10.1038/nrd3181
23 Satoh T, Ura T, Yamada Y, et al. Genotype-directed, dose-finding study of irinotecan in cancer patients with UGT1A1*28 and/or UGT1A1*6 polymorphisms. Cancer Sci, 2011, 102(10): 1868-1873. doi: 10.1111/j.1349-7006.2011.02030.x
24 Fujii H, Yamada Y, Watanabe D, et al. Dose adjustment of irinotecan based on UGT1A1 polymorphisms in patients with colorectal cancer. Cancer Chemother Pharmacol, 2019, 83(1): 123-129. doi: 10.1007/s00280-018-3711-8
25 Ma X, Han S, Liu Y, et al. Pharmacogenetic impact of UGT1A1 polymorphisms on pulmonary neuroendocrine tumours treated with metronomic irinotecan-based chemotherapy in Chinese populations. J Pharm Pharmacol, 2020, 72(11): 1528-1535. doi: 10.1111/jphp.13333
26 Chen Z, Su AI, et al. UGT1A1 sequence variants associated with risk of adult hypoalbuminemia: A quantitative analysis. Gene, 2014, 552(1): 32-38. doi: 10.1016/j.gene.2014.09.009
27 Bosma PJ, Chowdhury JR, Bakker C, et al. The genetic basis of the reduced expression of bilirubin UDP-glucuronosyltransferase 1 in Gilbert's syndrome. N Engl J Med, 1995, 333(18): 1171-1175. doi: 10.1056/NEJM199511023331802
28 Nelson RS, Seligson ND, Bottiglieri S, et al. UGT1A1 guided cancer
therapy: review of the evidence and considerations for clinical implementation. Cancers (Basel), 2021, 13(7): 1566. doi: 10.3390/cancers13071566

29 Páez J, Tobea M, Fernández-Plana J, et al. Pharmacogenetic clinical randomised phase II trial to evaluate the efficacy and safety of FOLFIRI with high-dose irinotecan (HD-FOLFIRI) in metastatic colorectal cancer patients according to their UGT1A1 genotype. Br J Cancer, 2019, 120(2): 190-195. doi: 10.1038/s41416-018-0348-7

30 Dias MM, McKinnon RA, Sorich MJ. Impact of the UGT1A1*28 allele on response to irinotecan: a systematic review and meta-analysis. Pharmacogenomics, 2012, 13(8): 889-899. doi: 10.2217/pgs.12.68

31 Yun F, Lulu M, Zhiyu H, et al. Uridine diphosphate glucuronide transferase *A1FN*0128 gene polymorphism and the toxicity of irinotecan in recurrent and refractory small cell lung cancer. J Cancer Res Ther, 2014, 10 Suppl: C195-C200. doi: 10.4103/0973-1482.145871

32 Bandopadhay A, Sharma S, Behera D, et al. UGT1A1 gene polymorphisms in patients with small cell lung cancer treated with irinotecan-platinum doublet chemotherapy and their association with gastrointestinal toxicity and overall survival. Oncologist, 2021, 26(8): 701-713. doi: 10.1002/onco.13757

33 Matsuoka H, Murakami R, Abiko K, et al. UGT1A1 polymorphism has a prognostic effect in patients with stage Ib or II uterine cervical cancer and one or no metastatic pelvic nodes receiving irinotecan chemotherapy: a retrospective study. BMC Cancer, 2020, 20(1): 729. doi: 10.1186/s12885-020-07225-1

34 Toffoli G, Cecchin E, Corona G, et al. The role of UGT1A1*28 polymorphism in the pharmacodynamics and pharmacokinetics of irinotecan in patients with metastatic colorectal cancer. J Clin Oncol, 2006, 24(19): 3061-3068. doi: 10.1200/JCO.2005.55.5400

35 Tian C, Ying H, Zhuang R, et al. Value of plasma SN-38 levels and DPD activity in irinotecan-based individualized chemotherapy for advanced colorectal cancer with heterozygous type UGT1A1*6 or UGT1A1*28. Cancer Manag Res, 2018, 10: 6217-6226. doi: 10.2147/CMAR.S179918

36 Wei X, Cai J, Sun H, et al. Cost-effectiveness analysis of UGT1A1*6/*28 genotyping for preventing FOLFIRI-induced severe neutropenia in Chinese colorectal cancer patients. Pharmacogenomics, 2019, 20(4): 241-249. doi: 10.2217/pgs-2018-0138

37 Sharma MR, Joshi SS, Karrison TG, et al. A UGT1A1 genotype-guided dosing study of modified FOLFIRINOX in previously untreated patients with advanced gastrointestinal malignancies. Cancer, 2019, 125(10): 1629-1636. doi: 10.1002/cncr.31938

38 Innocenti F, Schilsy RL, Ramirez J, et al. Dose-finding and pharmacokinetic study to optimize the dosing of irinotecan according to the UGT1A1 genotype of patients with cancer. J Clin Oncol, 2014, 32(22): 2328-2334. doi: 10.1200/JCO.2014.55.2307

39 McLeod HL, Sargent DJ, Marsh S, et al. Pharmacogenetic predictors of adverse events and response to chemotherapy in metastatic colorectal cancer: results from North American Gastrointestinal Intergroup Trial N9741. J Clin Oncol, 2010, 28(20): 3227-3233. doi: 10.1200/JCO.2009.21.7943

40 Hsieh YC, Chang TK, Su WC, et al. UGT1A1 polymorphism for irinotecan dose escalation in patients with BRAF-mutated metastatic colorectal cancer treated with first-line bevacizumab and FOLFIRI. J Oncol, 2021, 2021: 6868517. doi: 10.1155/2021/6868517

41 Zhu J, Liu A, Sun X, et al. Multicenter, randomized, phase III trial of neoadjuvant chemoradiation with capcitabine and irinotecan guided by UGT1A1 status in patients with locally advanced rectal cancer. J Clin Oncol, 2020, 38(36): 4231-4239. doi: 10.1200/JCO.20.01932

42 Catenacci DVT, Chase L, Lomnicki S, et al. Evaluation of the association of perioperative UGT1A1 genotype-dosed gFOLFIRINOX with margin-negative resection rates and pathologic response grades among patients with locally advanced gastrectomy for adenocarcinoma: a phase 2 clinical trial. JAMA Netw Open, 2020, 3(2): e1921290. doi: 10.1001/jamanetworkopen.2020.7771

43 Ma CJ, Chang TK, Tsai HL, et al. Regorafenib plus FOLFIRI with irinotecan dose escalated according to uridine diphosphate glucuronosyltransferase 1A1 genotyping in previous treated metastatic colorectal cancer patients: study protocol for a randomized controlled trial. Trials, 2019, 20(1): 751. doi: 10.1186/s13063-019-3917-x

44 Joshi SS, Catenacci DVT, Karrison TG, et al. Clinical assessment of 5-fluorouracil/leucovorin, nab-paclitaxel, and irinotecan (FOLIRI/ABRAXIS) in untreated patients with gastrointestinal cancer using UGT1A1 genotype-guided dosing. Clin Cancer Res, 2020, 26(1): 18-24. doi: 10.1158/1078-0432.CCR-19-1483

45 Atasip C, Chansiriwong P, Sirachainan E, et al. Effect of drug metabolizing enzymes and transporters in Thai colorectal cancer patients treated with irinotecan-based chemotherapy. Sci Rep, 2020, 10(1): 13486. doi: 10.1038/s41598-020-70351-0

46 Yokokawa A, Kaneko S, Endo S, et al. Effect of UGT1A1, CYP3A and CES activities on the pharmacokinetics of irinotecan and its metabolites in patients with UGT1A1 gene polymorphisms. Eur J Drug Metab Pharmacokinet, 2021, 46(2): 317-324. doi: 10.1007/s13318-021-00675-3

47 Sara SM, Xandra GG, Garcia M, et al. Clinical utility of ABCB1 genotyping for preventing toxicity in treatment with irinotecan. Pharmacol Res, 2018, 136: 133-139. doi: 10.1016/j.phrs.2018.08.026

48 Iwasa S, Muro K, Morita S, et al. Impact of UGT1A1 genotype on the efficacy and safety of irinotecan-based chemotherapy in metastatic colorectal cancer. Cancer Sci, 2021, 112(11): 4669-4678. doi: 10.1111/cas.15092

49 Mennillo E, Yang X, Weber AA, et al. Intestinal UGT1A1 and protection against Irinotecan-induced toxicity in a novel UGT1A1 tissue-specific humanized mouse model. Drug Metab Dispos, 2021, 50(3): 33-42. doi: 10.1124/dmd.121.006644

(Rceived: 2021-12-17 Revised: 2022-01-20 Accepted: 2022-01-22)