Feeding wheat dried distillers’ grains with solubles increases conjugated linoleic acid and unsaturated lipids in ovine milk without adversely affecting milk yield

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Abstract

The aim of this research communication was to examine the effect of dietary supplementation with wheat-based dried distillers’ grains with solubles (DDGS), a by-product of bioethanol production, on yield, composition, and fatty acid (FA) profile of ewe milk. Forty-five purebred mid-lactating Chios ewes (average milk yield 2.23 kg/d in 96 ± 5 d in lactation) were offered three iso-nitrogenous and iso-energetic diets (15 animals per diet) for a 10 d adaptation period followed by a 5-week recording and sampling period. The diets contained 0, 6, and 12% DDGS on DM basis for the DG0, DG6, and DG12 treatment, respectively, as a replacement of concentrate mix, whilst concentrate-to-forage ratio remained at 60:40 in all treatments. Individual milk yield, milk composition, and FA profile were recorded weekly and analyzed using a complete randomized design with repeated measurements. No significant differences were observed among groups concerning dry matter intake (overall mean of 2.59 kg/d), milk yield or 6% fat-corrected milk and milk protein percentage or protein yield. Milk fat percentage was decreased in the DG12 (4.76%) compared to DG0 (5.69%) without, however, significantly affecting the daily output of milk fat. The concentration of all major saturated FA between C4:0 to C16:0 was reduced, whereas long-chain (>16 carbons), mono-unsaturated and poly-unsaturated FAs were increased in the milk of DDGS groups. Among individual FA, increments of oleic acid and C18:1 trans-monoenes like C18:1 trans-10 and C18:1 trans-11 were demonstrated in DG12 group, whereas linoleic and conjugated linoleic acid (CLA cis-9, trans-11) were elevated in both DDGS groups compared to control. Changes in FA profile resulted in a decline in the atherogenic index of milk by 20% and 35% in DG6 and DG12 treatments, respectively, compared with control. In conclusion, feeding DDGS to dairy ewes increased the levels of unsaturated FA that are potentially beneficial for human health without adversely affecting milk, protein or fat yield.

The rapid growth of bioethanol production from grains such as corn, wheat and sugarcane has generated significant quantities of by-products that can be used as alternative feedstuffs in dairy animals (Schingoethe et al., 2009). There are two different bioethanol production processes, wet milling and dry grind, with the latter predominating (Schingoethe et al., 2009). During the dry grinding process, extractions may exist in different forms including dried distillers’ grain (DDG) and dried distillers grain with solubles (DDGS). DDGS represents an interesting supplement for ruminants due to its high protein, fat, and energy content (30%, 11.2% and 9.4 MJ NE/kg in DM, respectively, Schingoethe et al., 2009). High concentration of DDGS in the diets may cause milk fat depression when it coincides with reduced forage NDF content as shown in studies in cows with increasing concentration of DDGS in diet (Cyriac, 2005; Leonardi et al., 2005). In most studies, this reduction in fat percentage was not accompanied by a reduction in milk or fat yield although a concomitant reduction in milk yield with 25% DDGS inclusion level has been reported (Testroet et al., 2015). Nevertheless, the majority of studies in dairy cattle investigating DDGS from different grains with inclusion rates up to around 20% of DM intake showed that milk production and fat yield were increased or not adversely affected, when diets contained adequate levels of forage fiber (Schingoethe et al., 2009; Chibisa et al., 2012; Gaillard et al., 2017a, 2017b). Furthermore, the use of DDGS in cows enriched milk lipids with fatty acids (FA) potentially beneficial for human health; the content of saturated fatty acid (SFA) was decreased and the levels of long-chain FA, mono-unsaturated FA (MUFA) and poly-unsaturated FA (PUFA) including conjugated linoleic acid (CLA cis-9, trans-11) were increased (Leonardi et al., 2005; Anderson et al., 2006; Sasikala-Appukuttan et al., 2008; Abdellqader et al., 2009; Kurokawa et al., 2013; Testroet et al., 2015; Gaillard et al., 2017a, 2017b).
With regards to small ruminants, only two studies have investigated the use of DDGS in lactating animals (Cais-Sokolińska et al., 2015; Alshdaifat and Obeidat, 2019). In the latter the inclusion of 20 and 30% DDGS in the diet (DM basis) of dairy Awassi ewes increased milk yield without affecting the percentages of milk fat or protein, while the FA content of milk fat was not investigated. The former study (Cais-Sokolińska et al., 2015) examined only the chemical composition of sheep and goat milk and their products, and not milk yield. They reported that the inclusion of maize DDGS at the level of 3.5% (on DM basis) did not affect milk fat and protein percentages, but did affect FA content showing increased concentrations of total PUFA and trans – MUFA for both ovine and caprine milk, and reduced SFA, as well as increased MUFA and CLA content in caprine but not in ovine milk.

The present study was designed to investigate the effects of feeding DDGS on milk production and content, as well as on milk FA composition of dairy Chios ewes during a five-week sampling period in mid lactation. The hypothesis was that inclusion of DDGS at rates of 6% and 12% in dietary DM with wheat DDGS would positively affect yield, composition or FA profile of milk, suggesting therefore its use as an alternative protein-rich supplement in sheep rations.

### Materials and methods

Forty-five multiparous purebred, mid-lactating Chios ewes (average ± SD: 96 ± 5 d in lactation) were randomly distributed to 9 pens of 5 animals and allocated at random to 3 experimental feeding treatments (3 pens or 15 animals per treatment) resulting in overall means (± SEM) of 2.23 ± 0.01 kg for daily milk yield and 59.3 ± 0.4 kg for live weight. Feeding treatments contained 0, 6, and 12% on DM of wheat DDGS replacing corn grain and soybean meal (DG0, DG6 and DG12 treatments, respectively online Supplementary Table S1).

As shown in this table, animals received also barley hay of good quality as forage, obtaining a concentrate to forage ratio of 60:40 for all treatments. Feed ingredients and chemical composition are shown in Tables S1 and S2, respectively, in the online supplementary material. The three feeding regimes were isoenergetic and iso-nitrogenous and were offered for a 10 d adaptation period followed by a 5-week recording and sampling period. All animals were machine milked twice daily, and milk yields were recorded electronically. Milk samples for the determination of milk composition and lipid profile were collected weekly from each ewe for five consecutive weeks. Measurements for total fat, protein, and solids non-fat were performed with the use of Lactostar 3510 (Funke Gerber, Berlin, Germany) according to Tzamaloukas et al. (2015). Lipids from milk and feeds were extracted and methylated as described by Symeou et al. (2019) and (2020), and Tsiafoulis et al. (2014), respectively. Milk atherogenic (AI) and desaturation index (DI) were determined and fat-corrected milk yield at 6% of fat content was estimated. Further details of all methodologies and calculations are given in the online Supplementary Materials and Methods.

### Statistical analysis

Performance, milk content and milk FA composition data were analyzed using repeated measures analysis for a Completely Randomized Design, using SAS PROC MIXED (SAS version 9.4. SAS Institute Inc., Cary, NC). The model included the fixed effects of diet (D), time (T), and their interaction (T×D) and the random effect of ewes and pen. Statistical significance declared at $P < 0.05$. $P$-values between $>0.05$ and $<0.10$ were interpreted as trends toward significance. Data referring to DMI, chemical and FA composition of feeds were analyzed using one-way ANOVA with 3 replications, respectively.

### Results and discussion

Table 1 shows the DM intake, 6% fat-corrected milk, the daily yield of milk, fat, and protein, as well as the composition of milk. DDGS inclusion in the ewe diets did not affect intake, the

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**Table 1.** Milk production and chemical composition of milk from ewes fed diets contained 0 (control, DG0), 6 (DG6) or 12 (DG12) g of wheat – based dried distillers' grains with solubles per 100 g DM

| Item | Treatment | $SEM$ | $D$ | $T$ | $D \times T$ |
|------|-----------|-------|-----|-----|-------------|
| DMI, kg/d | DG0 | 2.61 | 2.59 | 2.55 | 0.02 | NS | – | – |
| Yield, kg/d | Milk | 1.63 | 1.62 | 1.75 | 0.06 | NS | *** | NS |
| | 6% FCM | 1.55 | 1.56 | 1.53 | 0.06 | NS | *** | NS |
| | Fat | 0.098 | 0.097 | 0.088 | 0.003 | NS | *** | NS |
| | Protein | 0.081 | 0.080 | 0.084 | 0.002 | NS | *** | ↑ |
| Milk composition, % | Fat | 5.69a | 5.60a | 4.76b | 0.18 | ** | *** | NS |
| | Protein | 4.80 | 4.73 | 4.70 | 0.03 | NS | ** | * |
| | Lactose | 4.94 | 4.83 | 4.85 | 0.03 | NS | NS | NS |
| | SNF | 10.88 | 10.64 | 10.72 | 0.07 | ↑ | ↑ | *** |

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$a,b,c$Means within a row not sharing a common superscript differ due to the different diet examined ($P < 0.05$); $*P < 0.01$; **$P < 0.001$; NS, Non-significant; $P < 0.1$: tendency; $6\%$ FCM, milk yield ($\times 0.453 + 0.0912 \times T$); $^\dagger$SNF, Solids non-fat.
Table 2. Fatty acid composition (expressed as a percentage of total fatty acid methyl esters) of milk from ewes fed diets contained 0 (control, DG0), 6 (DG6) or 12 (DG12) g of wheat–based dried distillers’ grains with solubles per 100 g DM

| Item       | Treatment | P-value¹ | SEM³ | D   | T   | D × T |
|------------|-----------|----------|------|-----|-----|-------|
|            | DG0       | DG6      | DG12 |     |     |       |
| C4:0       | 2.73a     | 2.79a    | 2.67b | 0.05 | *   | †     |
| C6:0       | 2.69a     | 2.65a    | 2.33b | 0.08 | *** | NS    |
| C7:0       | 0.08      | 0.07     | 0.08  | 0.009 | NS  | †     |
| C8:0       | 3.11a     | 3.01a    | 2.55b | 0.08 | **  | ***   |
| C10:0      | 7.95a     | 7.23b    | 6.17c | 0.18 | *** | NS    |
| C10:1 cis-9| 0.46a    | 0.40b    | 0.34c | 0.01 | *** | NS    |
| C12:0      | 5.84a     | 5.04b    | 4.46c | 0.13 | *** | NS    |
| C12:1 cis-9| 0.10a    | 0.08b    | 0.07b | 0.005 | NS  | **    |
| C13:0      | 0.33      | 0.25     | 0.30  | 0.02 | †   | NS    |
| C14:0      | 12.28a    | 11.35b   | 10.36c | 0.17 | *** | NS    |
| C14:0 iso  | 0.10      | 0.10     | 0.11  | 0.01 | NS  | NS    |
| C14:1, cis-9| 0.30a   | 0.21b    | 0.20b | 0.01 | NS  | **    |
| C15:0      | 1.72a     | 1.46b    | 1.75c | 0.06 | *** | ***   |
| C15:0 iso  | 0.25a     | 0.25ab   | 0.23b | 0.008 | NS  | NS    |
| C15:0 antiso| 0.75a  | 0.67b    | 0.69b | 0.02 | NS  | NS    |
| C16:0      | 22.26a    | 21.86a   | 20.61b | 0.34 | *   | NS    |
| C16:0 iso  | 0.30      | 0.31     | 0.31  | 0.01 | NS  | NS    |
| C16:1 cis-9| 2.17     | 2.02     | 2.00  | 0.09 | NS  | NS    |
| C16:1 trans-9| 0.60 | 0.57     | 0.62  | 0.02 | NS  | NS    |
| C17:0      | 0.74ab    | 0.73b    | 0.80a | 0.02 | *   | NS    |
| C17:0 iso  | 0.46      | 0.45     | 0.44  | 0.02 | NS  | NS    |
| C17:0 antiso| 0.71 | 0.67     | 0.70  | 0.04 | NS  | NS    |
| C17:1 cis-9| 0.29b    | 0.29b    | 0.34a | 0.01 | *   | NS    |
| C18:0      | 5.22      | 6.20     | 5.39  | 0.19 | †   | NS    |
| C18:1 trans-9| 0.35 | 0.39     | 0.48  | 0.06 | †   | NS    |
| C18:1 trans-10| 0.46b | 0.53b    | 0.77a | 0.05 | **  | *     |
| C18:1 trans-11| 3.10b | 3.26b    | 4.99a | 0.32 | **  | NS    |
| C18:1 trans-16| 0.10 | 0.09     | 0.12  | 0.02 | NS  | NS    |
| C18:1 cis-9| 14.03ab  | 15.24b   | 15.43a | 0.27 | *   | NS    |
| Other cis-C18:1²| 1.07 | 1.18     | 1.32  | 0.06 | NS  | ***   |
| C18:2 trans-9, cis-13| 0.19 | 0.23     | 0.26  | 0.007 | NS  | NS    |
| C18:2 trans-8, cis-13| 0.08b | 0.11a    | 0.12a | 0.006 | *   | NS    |
| C18:2 trans-11, cis-15| 0.13b | 0.18a    | 0.19a | 0.02 | *   | NS    |
| C18:2 trans-9, cis-12| 0.04 | 0.04     | 0.06  | 0.01 | NS  | NS    |
| C18:2n-6| 3.65c     | 4.15b    | 5.21a | 0.14 | *** | NS    |
| C18:3n-6| 0.10      | 0.08     | 0.09  | 0.005 | †   | **    |
| C18:3n-3| 0.33      | 0.33     | 0.41  | 0.01 | NS  | ***   |
| CLA cis-9, trans-11| 0.55b | 0.67a    | 0.72a | 0.03 | *   | ***   |
| CLA trans-9, cis-11| 0.08b | 0.08b    | 0.15a | 0.01 | **  | NS    |
| CLA trans-10, cis-12| 0.05 | 0.05     | 0.06  | 0.004 | NS  | **    |
| C20:4n-6| 0.23      | 0.23     | 0.26  | 0.01 | NS  | ***   |
yield of milk, or the fat-corrected milk. There is only one study (Alshdaifat and Obeidat, 2019) testing corn DDGS in dairy sheep production. This reported increased milk yields with DDGS inclusion in the diets (daily milk production of 0.95, 1.17 and 1.19 kg/d for inclusion rates of 0, 20 and 30% DM, respectively). Small increases (not always significant) have also been reported in some studies with lactating cows (Janicek et al., 2008; Chibisa et al., 2012; Benchaar et al., 2013) and it has been attributed to either the slightly increased energy density, due to higher fat content in DDGS diets, or to greater DM intake and, therefore, higher energy acquisition by the animals offered DDGS. Nevertheless, most cow studies with isoleucine and iso-nitrogenous experimental diets, and similar intakes, as those applied in the present experiment, reported no effect on milk yield, when the applied inclusion level of DDGS was up to 30% DM (Abdelqader et al., 2009; Gaillard et al., 2017a, 2017b; Testroet et al., 2018).

DDGS inclusion did not affect milk protein levels in the present study (Table 1), a fact that was also reported in a previous related study in sheep (Alshdaifat and Obeidat, 2019). Nonetheless, the results in previous studies with cows on the effect of DDGS on milk protein content are controversial, reporting either an increase (Testroet et al., 2015), or a decrease (Kleinschmit et al., 2006; Benchaar et al., 2013; Kurokawa et al., 2013), or no significant effect (Anderson et al., 2006; Janicek et al., 2008; Sasikala-Appukuttan et al., 2008; Chibisa et al., 2012). Reviewing the studies in cows, Kalscheur et al. (2012) concluded that milk protein content is rarely reduced by feeding DDGS, unless protein is limited in the diet or DDGS levels are higher than 30% DM of the diet reflecting the high rumen undegradable protein of the diet, which may result in a reduction of provided lysine for milk protein synthesis in the mammary gland. However, none of these prerequisites took place in our study where adequate feed protein levels and lower DDGS inclusion rates were used.

Milk fat percentage in the present study was reduced with the inclusion rate of 12%, but not with 6% of DDGS in the diet (DM basis) compared to control, without however affecting the fat yield (Table 1). Reduction of milk fat was not observed in a previous study in sheep with either 20 or 30% DM inclusion rates of corn DDGS (Alshdaifat and Obeidat, 2019). Nevertheless, it is well established in lactating cows that the addition of high levels of DDGS in diets could result in a decrease of milk fat percentages when NDF content and forage participation in the diets are low, a fact that has also been observed in practice (Schingoethe et al., 2009). A meta-analysis by Kalscheur et al. (2012) demonstrated that milk fat depression in cows fed with DDGS occurred only when diets contained less than 22% forage NDF and 50% total forage. This finding led these authors to suggest that the key of maintaining milk fat percentage is to provide not only adequate NDF and specific percentage of forages in the diet but also sufficient amounts of effective fiber (e.g. hay as opposed to corn silage). Although our study was designed to provide adequate dietary NDF content (34 and 35% DM in DG6 and DG12, respectively) and the barley straw component was at 40% of the diet DM (usual practice in Cyprus), it is not known if these practices were adequate to prevent milk fat reduction in sheep with the 12% DDGS inclusion in the diet.

All individual FA and their aggregated groups determined in milk of DG0, DG6, and DG12 ewes are presented in Table 2. The DDGS inclusion had a marked effect on milk FA composition by reducing SFA and increasing unsaturated FAs in both DG6 and DG12 treatments. The content of saturated FAs with less than 16 carbons were linearly reduced with increased DDGS proportion in the ewe diets, while the only saturated FA that came close to increase was C18:0 and this difference was not significant. These results are in line with those of a previous study in ewes (Cais-Sokolinski et al., 2015) and with many studies conducted in cows (Leonardi et al., 2005; Anderson et al., 2006; Sasikala-Appukuttan et al., 2008; Abdelqader et al., 2009; Hippen et al., 2010; Kurokawa et al., 2013; Gaillard et al., 2017a, 2017b) showing diminished de novo FA percentages and increased levels of long-chain FA (LCFA) as a result of DDGS dietary inclusion. The reduction of SFA in the present study was accompanied by a linear increase in total PUFA and MUFA with the increasing concentrations of DDGS in the

### Table 2. (Continued.)

| Item                  | Treatment | P-value | D | T | D × T |
|-----------------------|-----------|---------|---|---|-------|
| SCFA                  |           |         |   |   |       |
| DG0                   | 8.61a     | 8.52a   | 7.63b | 0.16 | ***   |
| DG6                   | 54.43a    | 50.69b  | 47.23c | 0.53 | ***   |
| DG12                  | 34.97b    | 39.01b  | 43.05c | 0.62 | ***   |
| MUFAs                 |           |         |   |   |       |
| DG0                   | 68.70b    | 66.62b  | 61.29b | 0.71 | ***   |
| DG6                   | 23.69b    | 25.41b  | 28.12b | 0.41 | ***   |
| DG12                  | 5.66a     | 6.35b   | 7.80a  | 0.21 | ***   |
| AI                    |           |         |   |   |       |
| DG0                   | 2.82a     | 2.27a   | 1.82c  | 0.06 | ***   |
| DG6                   | 2.84      | 2.41    | 2.50  | 0.11 | NS    |
| DG12                  |           |         |   |   |       |

### Notes:

a Means within a row not sharing a common superscript differ due to the different diet examined (P < 0.05), b Probability of significant effects due to diet (D), time (T), and their interaction (D × T); cMeans within a row not sharing a common superscript differ due to the different diet examined (P < 0.05). **P < 0.01; ***P < 0.001, NS, Non-significant; † Probability of significant effects due to diet (D), time (T), and their interaction (D × T); ‡ Means within a row not sharing a common superscript differ due to the different diet examined (P < 0.05).
sheep diets. Individual FA with a special interest for human health such as oleic, vaccenic, linoleic, and conjugated linoleic acid (CLA cis-9, trans-11), were linearly elevated with the addition of 6% and 12% DDGS in diet DM (Table 2). Similar effects have been reported in studies that examined diets from 5 up to 20% DM inclusion levels of DDGS in cows (Kalscheur et al., 2012; Kurokawa et al., 2013; Gaillard et al., 2017a, 2017b) and also observed in ewes with low DDGS inclusion (3.5% in diet DM, Cais-Sokolinski et al., 2015).

The reduction of FA with carbon chain less than 16 is due to either a higher secretion of LCFA from the blood and/or a lower de novo synthesis of FAs in the mammary gland (Chilliard et al., 2007; Dorea and Armentano, 2017). DDGS diets contained higher levels of ether extract than the control diet, indicating that ewes fed with DDGS consumed greater amounts of LCFA, including PUFA such as linoleic acid. Furthermore, de novo FA secretion can be inhibited by bioactive PUFA, produced during incomplete ruminal biohydrogenation of dietary LCFA, particularly CLA trans-10, cis-12 and trans monoenes, such as C18:1 trans-10 (Dorea and Armentano, 2017) with the latter found elevated in the milk of ewes fed DDGS in our study (Table 2). With regards to the predominant CLA isomer, cis-9, trans-11, (rumenic acid), it is well known that the majority of rumenic acid in the milk fat is synthesized endogenously, in the mammary gland through the action of mammary Δ9-desaturase with the substrate being vaccenic acid, while a small amount of this specific CLA originates from ruminal biohydrogenation of unsaturated FA by rumen bacteria (Dorea and Armentano, 2017). Thus, the elevated proportions of rumenic acid observed in the milk of DDGS groups are either due to the higher concentrations of vaccenic acid observed in those groups (Table 2) or due to the incomplete biohydrogenation of longer FA that took place in the rumen of ewes fed with the DDGS diets.

In conclusion, the observed changes in the FA profile as a consequence of feeding dried distillers’ grains with solubles resulted in an enrichment with beneficial FA and a reduction of atherogenic index of sheep milk by 20% (6% by DM inclusion) and 35% (12% inclusion), suggesting an improvement of health-related attributes of milk and possibly related dairy products. These results are useful for exploiting a cost-effective bioethanol byproduct to improve the FA profile of ewe milk and possibly other related dairy products without adverse effects on milk production.

Supplementary material. The supplementary material for this article can be found at https://doi.org/10.1017/S0022029921000443

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References

Abdelqader MM, Hippen AR, Kalscheur KF, Schingoethe DJ and Garcia AD (2009) Isolipidic additions of fat from corn germ, corn distillers grains, or corn oil in dairy cow diets. Journal of Dairy Science 92, 5523–5533.

Alshdaifat SN and Obeidat BS (2019) The impact of feeding corn dried distillers grains with solubles on milk yield and composition in lactating Awassi ewes and digestibility and N partitioning in Awassi ewe lambs. Italian Journal of Animal Science 18, 522–529.

Anderson JL, Schingoethe DJ, Kalscheur KF and Hippen AR (2006) Evaluation of dried and wet distillers grains included at two concentrations in the diets of lactating dairy cows. Journal of Dairy Science 89, 3133–3142.

Benchaz C, Hassanat F, Gervais R, Chouinard PY, Julien C, Petit H V and Massé DI (2013) Effects of increasing amounts of corn dried distillers grains with solubles in dairy cow diets on methane production, ruminal fermentation, digestion, N balance, and milk production. Journal of Dairy Science 96, 2413–2427.

Cais-Sokolinski D, Wójtowski J, Pikul J, Danków R, Majcher M, Teichert J and Bagnicka E (2015) Formation of volatile compounds in kefir made of goat and sheep milk with high polyunsaturated fatty acid content. Journal of Dairy Science 98, 6692–6705.

Chibisa GE, Christensen DA and Mutsamugwa T (2012) Effects of replacing canola meal as the major protein source with wheat dried distillers grains with solubles on ruminal function, microbial protein synthesis, omasal flow, and milk production in cows. Journal of Dairy Science 95, 824–841.

Chilliard Y, Glasser F, Ferlay A, Bernard L, Rouel J and Doreau M (2007) Diet, rumen biohydrogenation and nutritional quality of cow and goat milk fat. European Journal of Lipid Science and Technology 109, 828–855.

Cyrick J (2005) Effects of Replacing Forage Fiber with non-Forage Fiber in Lactating Dairy Cow Diets (Electronic Theses and Dissertations), Dorea JRR and Armentano LE (2017) Effects of common dietary fatty acids on milk yield and concentrations of fat and fatty acids in dairy cattle. Animal Production Science 57, 2224–2236.

Gaillard C, Sørensen MT, Vestergaard M, Weisbjerg MR, Basar A, Larsen MK, Martinussen H, Kidmose U and Sehested J (2017a) Effect of substituting soybean meal and canola cake with grain-based dried distillers grains with solubles as a protein source on feed intake, milk production, and milk quality in dairy cows. Journal of Dairy Science 100, 7988–7998.

Gaillard C, Sørensen MT, Vestergaard M, Weisbjerg MR, Basar A, Larsen MK, Martinussen H, Kidmose U and Sehested J (2017b) Effect of substituting soybean meal and canola cake with dried distillers grains with solubles at 2 dietary crude protein levels on feed intake, milk production, and milk quality in dairy cows. Journal of Dairy Science 100, 8928–8938.

Hippen AR, Schingoethe DJ, Kalscheur KF, Linke PL, Rennich DR, Abdelqader MM and Yoon I (2010) Saccharomyces cerevisiae fermentation product in dairy cow diets containing dried distillers grains plus solubles. Journal of Dairy Science 93, 2661–2669.

Janicek BN, Kononoff PJ, Gehman AM and Doane PH (2008) The effect of feeding dried distillers grains plus solubles on milk production and excretion of urinary purine derivatives. Journal of Dairy Science 91, 3544–3553.

Kalscheur K, Garcia A, Schingoethe D and Diaz-Royon F (2012) Feeding biofuel co-products to dairy cattle. In “Biofuel co-products as livestock feed - Opportunities and challenges”, edited by Harinder P.S. Makkar. Rome, FAO.

Kleinschmit DH, Schingoethe DJ, Kalscheur KF and Hippen AR (2006) Evaluation of various sources of corn dried distillers grains plus solubles for lactating dairy cattle. Journal of Dairy Science 89, 4784–4794.

Kurokawa Y, Shibata H, Tateno S, Kanda S, Takaura K, Ishida S and Itabashi H (2013) Rumen fermentation, milk production and conjugated linoleic acid in the milk of cows fed high fiber diets added with dried distillers grains with solubles. Animal Science Journal 84, 106–112.

Leonardi C, Bertics S and Armentano LE (2005) Effect of increasing oil from distillers grains or corn oil on lactation performance. Journal of Dairy Science 88, 2820–2827.

Sasikala-Appukuttan AK, Schingoethe DJ, Hippen AR, Kalscheur KF, Karges K and Gibson MI (2008) The feeding value of corn distillers grains for lactating dairy cows. Journal of Dairy Science 91, 279–287.

Schingoethe DJ, Kalscheur KF, Hippen AR and Garcia AD (2009) Invited review: the use of distillers products in dairy cattle diets. Journal of Dairy Science 92, 5802–5813.

Symoeu S, Tsaiopolis CG, Gerothanassi IP, Miliadou D and Tzamaloukas O (2009) Nuclear magnetic resonance screening of changes in fatty acid and cholesterol content of ovine milk induced by ensiled olive cake inclusion in Chios sheep diets. Small Ruminant Research 117, 111–116.

Symoeu S, Tzamaloukas O, Banos G and Miliadou D (2020) ACAAA2 and FASN polymorphisms affect the fatty acid profile of Chios sheep milk. Journal of Dairy Research 87, 23–26.

Testroet ED, Li G, Beitz DC and Clark S (2015) Feeding dried distillers grains with solubles affects composition but not oxidative stability of milk. Journal of Dairy Science 98, 2908–2919.
Testroet ED, Beitz DC, O’Neil MR, Mueller AL, Ramirez-Ramirez HA and Clark S (2018) Feeding reduced-fat dried distillers grains with solubles to lactating Holstein dairy cows does not alter milk composition or cause late blowing in cheese. Journal of Dairy Science 101, 5838–5850.

Tsiafoulis CG, Skarlas T, Tzamaloukas O, Miltiadou D and Gerothanassis IP (2014) Direct nuclear magnetic resonance identification and quantification of geometric isomers of conjugated linoleic acid in milk lipid fraction without derivatization steps: overcoming sensitivity and resolution barriers. Analytica Chimica Acta 821, 62–71.

Tzamaloukas O, Orford M, Miltiadou D and Papachristoforou C (2015) Partial suckling of lambs reduced the linoleic and conjugated linoleic acid contents of marketable milk in Chios ewes. Journal of Dairy Science 98, 1739–1749.