Iodine-125 interstitial brachytherapy reduces tumor growth via Warburg effect inhibition in non-small cell lung cancer A549 xenografts

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Abstract. Iodine-125 interstitial brachytherapy (¹²⁵I-IBT) is an alternative and effective treatment option for unresectable non-small cell lung cancer (NSCLC), and the Warburg effect is a determinant of tumor growth. The present study aimed to explore the influence of ¹²⁵I-IBT on tumor growth and the Warburg effect, and the potential mechanisms underlying NSCLC progression. Mice with A549 cell xenografts were evenly divided into a control group without ¹²⁵I-IBT, and three treatment groups receiving ¹²⁵I-IBT with 20, 40 and 60 Gy. Tumor volume (TV), maximum standardized uptake value (SUVmax) determined by ¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) micro-postion emission tomography/computed tomography and mean optical density (MOD) of mammalian target of rapamycin (mTOR), c-Myc, hypoxia inducible factor-1α (HIF-1α) and glucose transporter 1 (GLUT1) staining were compared among groups. Tumor inhibition rate (TIR). ¹⁸F-FDG uptake attenuation rate (FUAR) and expression suppression rate (ESR) were also calculated on day 14 and 28. The results demonstrated that the mean TV in the 60 and 40 Gy groups was smaller compared with the control TVs since days 14 and 16, respectively. The mean SUVmax value of the 60 Gy group at day 14, and all treatment group SUVmax values at day 28 were lower compared with the controls. In addition, the MOD of mTOR and GLUT1 was lower in the 60 Gy group, compared with the other groups, and c-Myc and HIF-1α values were lower in the 40 and 60 Gy groups, compared with the control and 20 Gy group (P<0.05). SUVmax positively correlated to TV (day 14, r=0.711; day 28, r=0.586) and the MOD of c-Myc and GLUT1 (r=0.621 and 0.546, respectively; P<0.01). Furthermore, dose dependent increases were observed for TIR, FUAR and ESR. In conclusion, ¹²⁵I-IBT reduced tumor growth by inhibiting the Warburg effect, which may have resulted from downregulation of mTOR, c-Myc, HIF-1α and GLUT1 expression, particularly c-Myc and GLUT1 in NSCLC A549 xenografts.

Introduction

Lung cancer is the most frequently diagnosed cancer and the leading cause of cancer mortality worldwide. Over 80% of cases are histologically classified as non-small cell lung cancer (NSCLC) (1,2). Surgery is generally the treatment of choice for early stage NSCLC. However, 30-55% of patients develop recurrence even following complete resection, and the mortality rate is high (3). As early disease is typically asymptomatic, the majority of patients are diagnosed at a distant stage, resulting in extremely poor prognosis, even with chemotherapy and/or radiotherapy (2). Recently, computed tomography (CT)-guided iodine-125 interstitial brachytherapy (¹²⁵I-IBT) has been employed to treat NSCLC at the inoperable advanced stage, and in cases with local recurrence or metastasis following surgery or concurrent chemoradiotherapy, alone or in combination with other treatments (3-6). It was demonstrated that ¹²⁵I-IBT significantly improves overall...
survival and local control, by maximizing the radiation dose delivered to the tumor, and minimizing radiation injury of the surrounding normal lung tissue. Certain preclinical studies have further suggested that increased G2/M arrest, increased apoptosis and enhanced bystander effect may serve key roles in the anti-proliferative effects induced by $^{125}$I-IBT in NSCLC (7-9). However, the association between $^{125}$I-IBT and glucose metabolism in NSCLC remains unclear.

The Warburg effect, a hallmark for tumor cells, was discovered by Otto Warburg in 1924. It is well established that in contrast to normal cells, which predominantly rely on mitochondrial oxidative phosphorylation (OXPHOS) to generate the energy required for cellular processes, the majority of tumor cells instead rely on aerobic glycolysis. The Warburg effect is a determinant of tumor cell proliferation (10). The rate of glucose entry into tumor cells is at least 20-30-fold higher than normal cells. This unique phenomenon is the basis for the use of positron emission tomography (PET)/CT with $^{18}$F-fluoro-deoxyglucose ($^{18}$F-FDG), a radioactive glucose analog that is intensely accumulated by tumor cells (11). $^{18}$F-FDG PET/CT is widely used for initial diagnosis, disease staging, recurrence and metastasis detection, as well as for monitoring chemotherapy and/or radiotherapy response in lung cancer (12-16).

In recent years, several studies have investigated the Warburg effect in lung cancer progression, as well as for the development of targeted therapies (17-19). It was suggested that Sad1 and UNC84 domain containing 2 (SUN2) protein may suppress lung cancer cell proliferation and migration by attenuating the Warburg effect, whereas pyruvate dehydrogenase kinase 1 (PDK1), may promote lung cancer cell proliferation and migration, via the inverse mechanism. In addition, dchlo-roacetate, a metabolic agent, may partially revert the Warburg effect in tumor cells, increase X-ray sensitivity and inhibit cell growth via Warburg effect inhibition in NSCLC. Alterations in tumor growth and $^{18}$F-FDG uptake were monitored using micro-PET/CT imaging. In addition, the relationships among these factors, the expression of glycolysis-associated molecules and the radiation dose were analyzed in NSCLC A549 xenografts following $^{125}$I-IBT.

Materials and methods

**Cell culture and animal model.** The A549 cell line was purchased from Shanghai Jianglin Biological Technology Co., Ltd. (Shanghai, China). Cells were cultured in RPMI-1640 (GE Healthcare Life Sciences, Logan, UT, USA) supplemented with 10% fetal bovine serum (FBS; Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA), 100 U/ml penicillin and 0.1 mg/ml streptomycin at 37°C in a 5% CO$_2$ atmosphere. Cells were trypsinized and harvested once 80-90% confluence was reached. Male BALB/c-nu mice (18-20 g; 4-6 weeks old) were purchased from Shanghai SLAC Laboratory Animal Co., Ltd. (Shanghai, China) and allowed to acclimatize for 1 week in a specific pathogen-free room under controlled temperature and humidity prior to experimental initiation. Following this, ~0.2 ml A549 cell suspension (1x10$^7$ cells/1 ml RPMI-1640) was injected subcutaneously into the right armpits of the forelegs. After 4 weeks, mice with a xenograft diameter of ~10 mm were used for study.

**Study design.** A total of 24 prior mice were randomly divided into four groups (n=6 per group). The control group without $^{125}$I seed implantation, and three treatment groups with a prescribed dose of 20, 40 and 60 Gy, via 1-2 $^{125}$I seed implantation with a radioactivity of 0.3-0.6 mCi, respectively. The treatment period was 4 weeks. Tumor growth was monitored by determining the xenograft size every 2 days with a calliper. Before and 14, 28 days after receiving $^{125}$I-IBT, micro-PET/CT imaging was performed to determine the baseline level and post-treatment alterations in $^{18}$F-FDG uptake. At the end of study, the mice were euthanized with dislocation of cervical vertebra after being anesthetized with 3% isoflurane inhalation and all xenografts were histologically analyzed with immunohistochemical (IHC) staining.

**Interstitial brachytherapy.** $^{125}$I seeds (model 6711; diameter, 0.8 mm; length, 4.5 mm; half-life, 59.6 days; half value thickness, 1.7 cm in tissue; main emission, 27.4-31.4 Kev X-ray and 35.5 Kev γ-ray) were provided by Seeds Biological Pharmacy, Ltd. (Tianjin, China). The $^{125}$I seeds were preloaded into 18-gauge needles and subsequently implanted into the xenograft center. The radiation prescribed dose was planned based on the axial micro-CT images using a brachytherapy treatment planning system (Beijing Astro Technology, Beijing, China).

**Tumor volume.** Based on measurements of the short (a) and long (b) tumor diameters, tumor volume (TV) was calculated, according to the formula of TV=1/2 a x b. The TV and tumor inhibition rate (TIR) on day 14 and 28 were defined as $^{18}$TV, $^{18}$TIR$_{14}$ and $^{18}$TIR$_{28}$, respectively. TIR was calculated using the following equation: TIR=(mean of $^{18}$TV$_t$-mean of TV$_j$/mean of TV$_j$ x 100%, where TV$_c$ was the control TV, and TV$_w$ was the treatment group TV.

**Micro-PET/CT imaging.** $^{18}$F-FDG was synthesized by the PET/CT center in our institution. Following an overnight fast, mice were injected with ~0.1 mCi $^{18}$F-FDG via the tail vein. After 1 h, mice were anesthetized with 3% isoflurane inhalation and placed in the prone position in the center of a Siemens Inveon combined micro-PET/CT scanner (Siemens Preclinical Solution USA, Inc., Knoxville, TN, USA). Micro-CT scans were performed with a 80 kV X-ray tube voltage, 500 µA current, 150 ms exposure time and 120 rotation steps. Micro-PET static acquisition was subsequently performed for 10 min and the data were processed using the ordered set expectation maximization algorithm for three dimensional PET reconstruction. Micro-PET/CT images were analyzed with an Inveon Research Workplace 4.1 (Siemens, Erlangen, Germany). The maximum standardized uptake value (SUVmax; g/ml) of the xenograft was measured on day 0, 14 and 28 and referred to as SUVmax$_{0}$, SUVmax$_{14}$, SUVmax$_{28}$, respectively. The $^{18}$F-FDG uptake attenuation rate (FUAR) of the xenograft on day 14 and 28 was defined as FUAR$_{14}$ and FUAR$_{28}$, respectively, using the following equation: FUAR=(mean SUVmax$_t$ value-mean SUVmax$_c$)
value)/mean SUVmax, value x100%, where SUVmax was the control group SUVmax, and the SUVmax was the treatment group SUVmax.

Histological evaluation. All xenografts were fixed in 10% formalin overnight at 4˚C and subsequently embedded in paraffin for histological section preparation (4 μm). The sections were deparaffinized and stained with hematoxylin and eosin for normal histological evaluation. IHC was also performed with antibodies to detect mammalian target of rapamycin (mTOR), c-Myc, hypoxia inducible factor-1α (HIF-1α) and glucose transporter 1 (GLUT1) expression, according to manufacturer's instructions. Primary antibodies (Abcam, Cambridge, UK) were used anti-mTOR ab2732 (diluted 1:200), anti-c-Myc ab32072 (diluted 1:100), anti-HIF-1α ab16066 (diluted 1:400) and anti-GLUT1 ab652 (diluted 1:200). In brief, sections were dewaxed and rehydrated prior to quenching in 3% hydrogen peroxide to block endogenous peroxidase activity for 10 min. Antigen retrieval was performed by immersing the sections in EDTA/Tris solution (pH 9.0) in a microwave oven for 2.15 min. Sections were cooled at room temperature and washed in PBS for 3x10 min. Following this, sections were blocked in 5% bovine serum albumin (BSA) for 30 min and subsequently incubated overnight with primary antibodies at 4˚C and rewarmed for 30 min and washed in PBS for 3x10 min. Then sections were incubated with biotinylated secondary antibodies at 37˚C for 30 min and washed in PBS for 3x10 min, followed by exposure to streptavidin-biotin complex (Ready-to-use SABC-POD kits: SA1021, goat anti-mouse IgG and SA1022, goat anti-rabbit IgG) for 45 min at 37˚C and diaminobenzidine tetrahydrochloride (DAB AR1022; both Boster, Wuhan, China). Staining without primary antibodies was performed in parallel and served as a negative control. Finally, sections were counterstained with hematoxylin, dehydrated in ethanol and xylene and mounted with cover slips using neutral balsam. Each section was assessed in three random microscopic fields (x200). The expression of mTOR, c-Myc, HIF-1α and GLUT1 was quantified by determining the mean optical density (MOD) using Image-Pro Plus software (version 6.0; Media Cybernetics, Inc. Maryland, USA). The expression suppression rate (ESR) of mTOR, c-Myc, HIF-1α and GLUT1 was defined as ESR_{mTOR}, ESR_{c-Myc}, ESR_{HIF-1α} and ESR_{GLUT1}, respectively, using the following equations: ESR=(mean MOD_α-mean MOD_β)/mean MOD_α x100%, where MOD_α was the control group MOD, and MOD_β was the MOD of the treatment groups.

Statistical analysis. All statistical analyses were performed using SPSS 19.0 software (IBM Corp., Armonk, NY, USA). The continuous variables were described as the mean ± standard deviation. Significant differences were evaluated using one-way analysis of variance. If variances were equal, all pairwise comparisons between group means were performed with Fisher’s Least Significant Difference test; if unequal, Dunnett’s T3 test was used. Pearson’s correlation analysis was used to evaluate the relationships among TV, SUVmax and the MOD of mTOR, c-Myc, HIF-1α and GLUT1. P<0.05 was considered to indicate a statistically significant difference.

Results

\(^{125}\text{I}-\text{IBT inhibits A549 xenograft growth}.\) A549 xenograft growth over time following \(^{125}\text{I}-\text{IBT}\) was presented in Fig. 1. Overall, the tumor gradually enlarged over time in all groups, although TV decreased as the radiation dose increased. The mean TV of the 60 Gy group (377.3±227.2 mm³) and 40 Gy group (410.7±174.42 mm³) was significantly reduced compared with controls since day 14 (691.0±258.5 mm³) and 16 (717.5±212.9 mm³), respectively (P<0.05). However, no statistical differences were observed between the 20 Gy group and controls among all treatment groups (P>0.05). In addition, the TIR4 of the 20, 40 and 60 Gy groups was 19.9, 39.8 and 45.4%; and the TIR4 were 18.0, 60.5 and 61.7%, respectively. These data suggested that \(^{125}\text{I}-\text{IBT}\) may have inhibited the growth of A549 xenografts, and inhibition was enhanced as the radiation dose increased.

\(^{125}\text{I}-\text{IBT reduces }^{18}\text{F-FDG uptake in A549 xenografts}.\) \(^{18}\text{F-FDG}\) uptake within A549 xenografts following \(^{125}\text{I}-\text{IBT}\) was presented in Fig. 2. In general, compared with controls, the \(^{18}\text{F-FDG}\) uptake of xenografts gradually reduced over time in \(^{125}\text{I}-\text{IBT}\) treatment groups. Furthermore, the reduction in SUVmax was increased as the radiation dose increased (Fig. 3). The mean of SUVmax\(_α\) of the 60 Gy-group was significantly lower than the control group (0.91±0.13 g/ml vs. 1.08±0.08 g/ml; P<0.05). The mean SUVmax\(_α\) of all treatment groups was significantly lower than the control group (20 Gy, 0.94±0.19 g/ml; 40 Gy, 0.93±0.08 g/ml; 60 Gy, 0.79±0.16 g/ml vs. control group, 1.15±0.22 g/ml; P<0.05), although there were no significant differences among treatment groups on day 14 and 28 (P>0.05). Furthermore, the FUAR\(_α\) of the 20, 40 and 60 Gy groups was 6.5, 8.3 and 15.7%, and the FUAR\(_α\) of the 20, 40 and 60 Gy groups was 18.3, 19.1 and 31.3%, respectively. These data suggested that \(^{125}\text{I}-\text{IBT}\) may have reduced \(^{18}\text{F-FDG}\) uptake within A549 xenografts, and this effect was more marked as the radiation dose increased.
**125**I-IBT reduces tumor growth via Warburg effect inhibition in NSCLC

**125**I-IBT suppresses mTOR, c-Myc, HIF-1α and GLUT1 expression in A549 xenografts. The IHC staining patterns of mTOR, c-Myc, HIF-1α and GLUT1 in A549 xenografts following **125**I-IBT were presented in Fig. 4. In tumor cells, mTOR, c-Myc and HIF-1α were expressed in the nucleus and cytoplasm, whereas GLUT1 was predominantly expressed in the cytomembrane and cytoplasm. Overall, the expression of mTOR, c-Myc, HIF-1α and GLUT1 in xenografts was gradually downregulated as the radiation dose increased in the **125**I-IBT treatment groups, compared with the controls (Fig. 5). The MOD of mTOR staining in the 60 Gy-group (0.0092±0.0062) was markedly decreased compared with the other groups (control, 0.0198±0.0053; 20 Gy, 0.0191±0.0096 and 40 Gy, 0.0194±0.0062; P<0.05). In addition, the MOD of c-Myc staining was significantly lower in the 40 (0.0025±0.0010) and 60 Gy (0.0006±0.0006) treatment groups, compared with the other groups (control, 0.0077±0.0020 and 20 Gy, 0.0055±0.0029; P<0.01). Furthermore, the MOD of HIF-1α staining in the 40 (0.0024±0.0021) and 60 Gy (0.0018±0.0006) treatment groups was significantly than that of the control group (0.0070±0.0058; P<0.05). Finally, the MOD of GLUT1 staining in the 60 Gy (0.0024±0.0025) group

Figure 2. 18F-FDG micro-PET/CT imaging in A549 xenografts following the administration of **125**I-IBT. The 18F-FDG uptake generally decreased over time within treatment group tumors, compared with the control group. 18F-FDG, 18F-fluorodeoxyglucose; micro-PET/CT, micro-positron emission tomography/computed tomography; **125**I-IBT, iodine-125 interstitial brachytherapy.

Figure 3. Comparison of 18F-FDG uptake in A549 xenografts following the administration of **125**I-IBT. The maximum standardized uptake value gradually reduced over time in the treatment groups compared with the control. Reduction was more marked as the radiation dose increased. *P<0.05, **P<0.01 vs. controls. 18F-FDG, 18F-fluorodeoxyglucose; **125**I-IBT, iodine-125 interstitial brachytherapy.
was significantly decreased compared with the other (control, 0.0150±0.0043; 20 Gy, 0.0139±0.0046; 40 Gy, 0.0130±0.0043; P<0.01). However, there were no significant differences between 20 Gy group and the control for mTOR, c-Myc, HIF-1α or GLUT1 expression in A549 xenografts (P>0.05). Additionally, the ESR_{mTOR} in the 20, 40 and 60 Gy groups was 4.0, 2.5 and 53.8%, respectively; the ESR_{c-Myc} was 28.6, 67.5 and 92.2%, respectively; the ESR_{HIF-1α} was 51.4, 65.7 and 74.3%, respectively; and the ESR_{GLUT1} was 7.3, 13.3 and 84.0%, respectively. These data suggested that 125I-IBT may have suppressed the expression of mTOR, c-Myc, HIF-1α and GLUT1 in A549 xenografts, and the suppression effect was increased as the radiation dose increased.

Relationships among tumor growth, 18F-FDG uptake and expression of mTOR, c-Myc, HIF-1α and GLUT1. The correlations among tumor growth, 18F-FDG uptake and expression of mTOR, c-Myc, HIF-1α and GLUT1 were presented in Fig. 6. There was a positive correlation between TV_{28} and SUV_{max}_{28} (r=0.586), respectively (P<0.01). In addition, SUV_{max}_{28} positively correlated to the MOD of c-Myc (r=0.621) and GLUT1 (r=0.546), respectively (P<0.01). However, the MOD of mTOR (r=0.243) and HIF-1α (r=0.029) had no significant correlation with SUV_{max}_{28} (P>0.05). Furthermore, the MOD of mTOR, c-Myc and GLUT1 had a significant correlation with each other (P<0.05; Table I).

**Discussion**

In the present study, it was postulated that 125I-IBT may have suppressed NSCLC tumor growth by inhibiting the Warburg effect. Tumor growth and 18F-FDG uptake was monitored in NSCLC A549 xenografts following 125I-IBT with a radiation dose of 20, 40 or 60 Gy, and the relationships among them were also evaluated. It was demonstrated that both the TV and SUV_{max} of all treatment groups were decreased compared with controls following 125I-IBT, and these reductions were more marked as the time and radiation dose increased, which
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was consistent with results obtained in previous studies of pancreatic carcinoma xenografts (20,21). In addition, the TIR and FUAR increased more or less as the radiation dose increased, and TV positively correlated to SUVmax on day 14 (r=0.682; P<0.01) and day 28 (r=0.546 on day 28, respectively). TV, tumor volume; SUVmax, maximum standardized uptake value; mTOR, mammalian target of rapamycin; HIF-1α, hypoxia inducible factor-1α; GLUT1, glucose transporter 1; 125I-IBT, iodine-125 interstitial brachytherapy.

The Warburg effect is considered to involve multiple intricate mechanisms, including tumor microenvironment support, HIF stabilization, oncogene activation, tumor suppressor gene loss, cancer cell mitochondrial dysfunction, nuclear DNA mutations, epigenetic modifications, microRNA regulation, glutamine metabolism and post-translational modifications (24). HIF-1, a heterodimeric transcription factor consisting of α and β subunits, is known to contribute to the adaptability of tumor cells to hypoxic conditions. It may increase the efficacy of the glycolytic pathway by upregulating the expression of several GLUTs and several other genes linked to aerobic glycolysis (25). For example, GLUT1 is ubiquitously expressed, but overexpressed in tumor cells and therefore increases the uptake of glucose. HIF regulation is predominantly dependent on the HIF-1α subunit, that has a short half-life under normoxic conditions due to proteasomal degradation (24,26,27). The phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT) and the Ras/Raf/mitogen-activated protein kinase kinase (MAPKK)/extracellular regulated protein kinase (ERK) pathways are the major signaling pathways downstream from growth factor-bound receptor tyrosine kinases, which are linked to both growth control and glucose metabolism in lung cancer (16), as well as other malignancies (22,23).

Table I. Associations among expression of mTOR, c-Myc, HIF-1α and GLUT1 in A549 xenograft after 125I-IBT.

| Protein name | mTOR, correlation coefficient (P-value) | c-Myc, correlation coefficient (P-value) | HIF-1α, correlation coefficient (P-value) |
|--------------|----------------------------------------|----------------------------------------|----------------------------------------|
| c-Myc        | 0.500a (0.013)                          | 0.365 (0.079)                          | 0.264 (0.212)                          |
| HIF-1α       | -0.003 (0.988)                          | 0.526a (0.008)                         | 0.614a (0.001)                         |
| GLUT1        |                                       |                                        |                                        |

aP<0.05, bP<0.01. mTOR, mammalian target of rapamycin; HIF-1α, hypoxia inducible factor-1α; GLUT1, glucose transporter 1; 125I-IBT, iodine-125 interstitial brachytherapy.

Figure 6. Relationships among tumor growth, 18F-FDG uptake and expression of mTOR, c-Myc, HIF-1α and GLUT1 in A549 xenografts following 125I-IBT. (A) TV positively correlated to SUVmax on day 14 (r=0.711; P<0.01) and day 28 (r=0.586; P<0.01). TV on day 28 also correlated to SUVmax on day 14 (r=0.682; P<0.01). TV, tumor volume; SUVmax, maximum standardized uptake value; mTOR, mammalian target of rapamycin; HIF-1α, hypoxia inducible factor-1α; GLUT1, glucose transporter 1; 125I-IBT, iodine-125 interstitial brachytherapy.
together with HIF-1 in glycolysis and OXPHOS regulation. Both HIF and c-Myc activate hexokinase 2 (HK2) and PKD1, leading to increased glycolytic rates and conversion of glucose to lactate (27).

In the present study, alterations in mTOR, c-Myc, HIF-1α and GLUT1 expression was analyzed by IHC techniques, and the relationship among them was determined. The findings revealed that the xenograft expression of mTOR, c-Myc, HIF-1α and GLUT1 decreased as the radiation dose increased, with the largest decrease observed in the 60 Gy group, compared with the controls. Furthermore, the ESR2_1α, ESR2_Myc, ESR2_HIF-1α and ESR2_GLU1 was dose-dependently increased, and the MOD of mTOR, c-Myc and GLUT1 were significantly correlated with each other. These findings indicated that the Warburg effect was inhibited by 125I-I-IBT, which may have resulted from mTOR, c-Myc, HIF-1α and GLUT1 downregulation in NSCLC A549 xenografts.

Although the downregulation of the above molecules by 125I-I-IBT has not been explicitly defined in the present study, possible involvement of AKT and ERK-mediated signaling pathways may be suggested in accordance with a previous report demonstrating that 125I-I-IBT may downregulate epidermal growth factor receptor, inhibiting AKT activation in colorectal tumor cells (34). 125I-I-IBT has also been demonstrated to suppress vascular endothelial growth factor-A expression, and consequently inhibit ERK activation in nasopharyngeal tumor cells (35). It is plausible that AKT and ERK inactivation may have reduced the expression of mTOR and c-Myc, resulting in the downregulation of HIF-1α and GLUT1 expression in the present study. In addition, Cron et al (36) reported that 125I seed may significantly increase oxygen partial pressure within 24 h and even on day 3. Furthermore, it may elevate blood perfusion from day 3, in 2-4 mm of the surrounding liver tumor region (36). As a result, the improvement of the tumor hypoxic microenvironment by 125I-I-IBT may have also partially reduced the expression and/or accelerated the degradation of HIF-1α. However, Hu et al (37) demonstrated that 125I-I-IBT increases mitochondrial reactive oxygen species (ROS) production and consequently upregulates the expression of HIF-1α in human colorectal cancer cells (37). Therefore, the downregulation of HIF-1α by 125I-I-IBT may have been a synthetic action of above factors in NSCLC.

In addition, the correlation analysis between glucose metabolism and the expression of mTOR, c-Myc, HIF-1α and GLUT1 in the present study revealed that the SUVmax_a was positively correlated with the MOD of c-Myc (r=0.621) and GLUT1 (r=0.546). This suggested that 18F-FDG uptake may depend upon the overexpression of c-Myc and GLUT1 in xenografts, particularly c-Myc. GLUT1 is the major glucose transporter expressed in NSCLC. Although its overexpression is considered to be linked to 18F-FDG uptake within tumors, it is unlikely that this is the sole or rate-limiting step in this process (38). The role of GLUT1 remains controversial in NSCLC; certain studies have reported a significant correlation (39,40), whereas others have detected no correlation (41,42). This may be attributed to the fact that 18F-FDG uptake involves tumor blood flow and tracer delivery rate, in addition to expression of GLUT1. Once 18F-FDG enters tumor cells, hexokinase and glucose-6-phosphatase (G6Pase) activity determines how much phosphorylated 18F-FDG is trapped in cells (38). c-Myc, as a central regulator of cell growth and proliferation (43), targets genes involved in glucose transport, glycolysis, glutaminolysis and fatty acid synthesis (44). c-Myc activates GLUT1, as well as HK2 (27) and G6Pase (45), leading to increased uptake of 18F-FDG. Downregulation of c-Myc and GLUT1 suppresses the glycolytic pathway and glycolytic enzyme activity, leading to 18F-FDG uptake inhibition (46).

However, it was noted that in the present study, no statistical differences between TV, SUVmax_a and SUVmax_b were observed among the treatment groups, or between the TV and the MOD of mTOR, c-Myc, HIF-1α and GLUT1 in the 20 Gy and control group. This may have been due to the short investigation duration of 28 days, as 125I-I-IBT is typically delivered over >180 days (21). Additionally, 60 Gy is a relatively low radiation dose, considering that the clinically effective dose is typically 100-160 Gy (3-6). Furthermore, there were no significant correlations between HIF-1α expression and the other detected proteins, or between the SUVmax_a and the MOD of mTOR and HIF-1α (P>0.05). This may be attributed in part to the complex, multifactorial regulation of HIF-1α as detailed above. In addition, it suggested that the AKT/mTOR pathway may have less influence on the Warburg effect in NSCLC, compared with the ERK/c-Myc pathway.

Certain limitations of the present study should be considered. First, other key molecules associated with glycolysis were not examined, such as PKM2 and HK2. Second, the expression of glycolysis-associated molecules was only evaluated using IHC rather than with western blotting and/or reverse transcription-quantitative polymerase chain reaction assays for transcriptional and translational expression analyses, respectively. Third, A549 cells harbor the mutant K-ras gene, and the Ras/Raf/ERK signaling pathway is critical for tumor proliferation. Therefore, multiple NSCLC cells, including those with the wild-type K-ras gene, should be used to confirm the results of the present study.

In conclusion, the present study suggested that 125I-I-IBT may have reduced tumor growth via Warburg effect inhibition, and this inhibitory effect increased as the radiation dose increased. Warburg effect inhibition may have resulted from mTOR, c-Myc, HIF-1α and GLUT1 expression downregulation, particularly c-Myc and GLUT1, in NSCLC A549 xenografts. Further research is required to clearly delineate the exact mechanisms underlying Warburg effect inhibition by 125I-I-IBT in NSCLC.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

JZ established animal models, performed animal imaging and drafted the manuscript. YZ was responsible for cell culture and participated in preparation of animal model and imaging. MD, JY and WW participated in the iodine-125 seed implantation and immunohistochemistry. LT conceived and designed the study, performed the statistical analysis and revised the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The animals used in the present study received humane care in compliance with the Guideline to the Care and Use of Experimental Animals established by the Medical Ethical Committee on animal experiments of the Zhejiang University.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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