Mitigating pseudoreplication and bias in resource selection functions with autocorrelation-informed weighting

Jesse M. Alston1,2,3 | Christen H. Fleming4,5 | Roland Kays6,7 | Jarryd P. Streicher8 | Colleen T. Downs8 | Tharmalingam Ramesh8,9 | Björn Reineking10 | Justin M. Calabrese1,2,11

1Center for Advanced Systems Understanding, Görlitz, Germany; 2Helmholtz-Zentrum Dresden Rossendorf (HZDR), Dresden, Germany; 3School of Natural Resources and the Environment, University of Arizona, Tucson, Arizona, USA; 4Smithsonian Conservation Biology Institute, National Zoological Park, Front Royal, Virginia, USA; 5Department of Biology, University of Maryland, College Park, Maryland, USA; 6Department of Forestry and Environmental Resources, North Carolina State University, Raleigh, North Carolina, USA; 7North Carolina Museum of Natural Sciences, Raleigh, North Carolina, USA; 8Centre for Functional Biodiversity, School of Life Sciences, University of KwaZulu-Natal, Pietermaritzburg, South Africa; 9Sálim Ali Centre for Ornithology and Natural History (SACON), Coimbatore, Tamil Nadu, India; 10Université Grenoble Alpes, INRAE, LESSEM, Saint-Martin-d’Hères, France and 11Department of Ecological Modelling, Helmholtz Centre for Environmental Research (UFZ), Leipzig, Germany

Abstract

1. Resource selection functions (RSFs) are among the most commonly used statistical tools in both basic and applied animal ecology. They are typically parameterized using animal tracking data, and advances in animal tracking technology have led to increased levels of autocorrelation between locations in such data sets. Because RSFs assume that data are independent and identically distributed, such autocorrelation can cause misleadingly narrow confidence intervals and biased parameter estimates.

2. Data thinning, generalized estimating equations and step selection functions (SSFs) have been suggested as techniques for mitigating the statistical problems posed by autocorrelation, but these approaches have notable limitations that include statistical inefficiency, unclear or arbitrary targets for adequate levels of statistical independence, constraints in input data and (in the case of SSFs) scale-dependent inference. To remedy these problems, we introduce a method for likelihood weighting of animal locations to mitigate the negative consequences of autocorrelation on RSFs.

3. In this study, we demonstrate that this method weights each observed location in an animal’s movement track according to its level of non-independence, expanding confidence intervals and reducing bias that can arise when there are missing data in the movement track.

4. Ecologists and conservation biologists can use this method to improve the quality of inferences derived from RSFs. We also provide a complete, annotated...
analytical workflow to help new users apply our method to their own animal tracking data using the ctmm R package.

**KEYWORDS**
continuous-time movement models, habitat selection, home range, Ornstein–Uhlenbeck process, space use, spatial point process, stochastic process model, utilization distribution

### 1 | INTRODUCTION

Resource selection functions (RSFs) have been a mainstay of basic and applied ecology for decades (Boyce & McDonald, 1999; Manly et al., 2007), and are commonly used to answer questions such as ‘What landscape features do animals seek or avoid during their movements?’ (e.g. Lamont et al., 2019; Prokopenko et al., 2017; Valls-Fox et al., 2018) and ‘In which areas of the landscape are animals at risk of predation or disease transmission?’ (e.g. Ali et al., 2017; Ng’weno et al., 2019; Rayl et al., 2019). They are typically conceptualized as a Poisson point process and fit in a use–availability framework, whereby environmental covariates at the locations where animals were known to be present (i.e. ‘used’ locations) are compared with covariates at locations taken from an area assumed to be available for selection (i.e. ‘available’ locations; Manly et al., 2007). RSFs are usually parameterized using logistic regression, which allows researchers to easily estimate selection or avoidance of environmental features and to generate maps or data layers for use in downstream analyses (Northrup et al., 2013, 2022).

The relative ease of fitting RSFs has contributed to their popularity in animal ecology. However, the results of RSFs are influenced by a number of methodological challenges (Aarts et al., 2008; Northrup et al., 2013, 2022). One major (and worsening) challenge is the degree of temporal autocorrelation in tracking data, which has increased considerably as advances in tracking devices facilitate the recording of animal locations at ever-smaller intervals (Fieberg et al., 2010). Conventional RSFs assume that data arise from Poisson point processes and are therefore sampled independently, which means that they do not allow for any autocorrelation in the data. Autocorrelated movement data are generally less informative than independent data when estimating coarse-scale parameters, because adjacent data points share information. In other words, an animal’s location at time $t_i$ is a function of both resource selection and its location at time $t_{i-1}$. Estimates of resource selection are therefore overconfident and often biased when autocorrelated data are modelled as independent and identically distributed (IID). This is one form of pseudoreplication, which has long been acknowledged as a problem in ecology (Hurlbert, 1984). Modern animal tracking data are almost always positively autocorrelated (Noonan et al., 2019), which poses a major problem for RSFs.

Such pseudoreplication—and the incorrectly narrow confidence intervals that arise from it—can cause random variation to be incorrectly identified as significant effects. This may (at least in part) explain why a growing number of studies have documented surprising levels of apparent variation in habitat selection within species and even populations (e.g. Leclerc et al., 2018; Newediuk et al., 2022). Individual animals undoubtedly vary somewhat in their habitat preferences because of differences in behavioural traits (Bastille-Rousseau & Wittemyer, 2019; Leclerc et al., 2016; Stuber et al., 2022), population density (Avgar et al., 2020; Matthiopoulos et al., 2015; van Beest et al., 2014) and habitat availability (Aarts et al., 2013; Godvik et al., 2009; Mysterud & Ims, 1998). However, substantial levels of Type I error in selection parameters can easily arise when pseudoreplication from autocorrelated data causes unimportant covariates to be misleadingly found to be significant. Accurate quantification of uncertainty is therefore paramount for accurate inference concerning habitat selection. In addition, RSFs that assume IID data are unable to account for varying levels of autocorrelation in movement data. This might happen when environmental variables inhibit successful estimation of animal locations (e.g. dense vegetation blocking Global Positioning System [GPS] satellite reception), leaving unsampled gaps in an animal movement track (Fleming et al., 2018; Ffair et al., 2010; Lewis et al., 2007). When the gaps in data are correlated with specific covariates of interest, the level of autocorrelation in animal locations is also correlated with those covariates. Despite widespread awareness of these problems, no generally accepted solutions have been described, and researchers often fit IID RSF models to autocorrelated tracking data.

Historically, data thinning has been the most common suggestion for reducing autocorrelation in tracking data used to inform RSFs (e.g. Hooten et al., 2014; Northrup et al., 2013; Swihart & Slade, 1985). While pragmatic and easy to implement, data thinning suffers from a number of limitations as a method of mitigating autocorrelation. First, thinning inherently requires discarding data, which results in the loss of useful information and can lead to imprecise parameter estimates. Second, thinning typically involves using statistical tests for the presence of statistically significant levels of autocorrelation—a target which is both arbitrary and not optimized for the task of estimating distributions (Aarts et al., 2008). Under-thinning results in misleadingly narrow confidence intervals and possibly biased coefficient estimates, while over-thinning results in an unnecessary loss of information and statistical efficiency. Third, highly irregular time-series, such as those obtained from tracking aquatic animals (Breed et al., 2011; Fleming et al., 2018) or sampling designs targeted at identifying behaviours at multiple temporal scales (e.g. Scantlebury et al., 2014; Ullmann et al., 2020), may not be particularly amenable to thinning.


Increasingly, researchers use step selection functions (hereafter SSFs; Avgar et al., 2016, Fortin et al., 2005, Signer et al., 2019; Thurjell et al., 2014) to account for autocorrelation in animal movements during studies of resource selection (e.g. Alston et al., 2020; Dickie et al., 2020; Kohl et al., 2018; Merkle et al., 2016; Prokopenko et al., 2017a). This approach builds upon conventional RSFs by generating available points (or movement steps) using a biologically realistic, movement-informed sampling scheme. However, SSFs still have important limitations. First, because available points in conventional SSFs depend on distributions of step lengths and turning angles on a fixed sampling interval, conventional SSFs cannot handle irregularly sampled data—SSFs can only be parameterized using animal location data that were sampled at a constant sampling interval (but see Brost et al., 2015; Christ et al., 2008; Johnson et al., 2008 for examples of models that are conceptually similar to SSFs, but that replace the correlated random walk movement model underlying conventional SSFs with Gaussian stochastic processes, which can relax the assumption of a constant sampling interval). Second, parameters of SSFs are explicitly scale-dependent—parameters change as the sampling schedule of a movement track changes (Avgar et al., 2016; Fieberg et al., 2021; Signer et al., 2017). In other words, if the same movement track were to be sampled at different intervals, the SSFs parameterized using the different samples would provide different estimates of selection. Third, SSFs do not invariably mitigate the pseudoreplication that arises from treating autocorrelated data as IID—SSFs assume that steps are independent, so when the sampling interval is smaller than the length of time required for steps to be statistically independent of one another, SSFs will also produce misleadingly narrow confidence intervals. Additional measures such as data thinning (Babin et al., 2011; Robb et al., 2022), variance inflation (Nielsen et al., 2002), generalized estimating equations (Craiu et al., 2008; Prima et al., 2017) and modelling interactions between steps can be employed to widen confidence intervals associated with SSF parameters, but these methods lack an objective and easily implementable target for the appropriate width of confidence intervals.

Likelihood weighting offers another potential solution for mitigating the adverse effects of temporal autocorrelation on RSFs. In this framework, the contributions of individual animal locations are weighted according to a joint likelihood function such that the sum of all weights matches an ‘effective sample size’ derived from stochastic process models that describe the autocorrelation structure of an animal movement track. This framework avoids two problems caused by data thinning. First, instead of considering one thinned subsample of data and its set of resulting parameter estimates, or several thinned subsamples of the data and an average over their parameter estimates (which would reduce noise—and potentially bias—arising from thinning), all subsamples of independent data and an average of their log-likelihoods are evaluated during parameter estimation. In other words, autocorrelated data are down-weighted rather than discarded. Second, instead of thinning the data to an ad hoc threshold (e.g. less than 5% autocorrelation), data are weighted using an objective estimate of ‘effective sample size’ that quantifies confidence in a home-range estimate. As an additional benefit, likelihood weights can also account for sampling biases that arise from irregular sampling. Such bias can occur, for example, when covariates are over- or under-sampled, such as when environmental covariates are associated with differential probability of successful triangulation of animal locations (Fleming et al., 2018; Fray et al., 2010; Lewis et al., 2007). This means that irregular sampling of non-independent data (which is incompatible with the IID assumption underlying conventional RSFs) can be explicitly accounted for in an RSF model.

In this study, we introduce a method for autocorrelation-informed likelihood weighting of animal locations to mitigate pseudoreplication and bias in parameter estimates of RSFs. We provide a description of the mathematical principles underlying our method, demonstrate its practical advantages over conventional approaches using simulations and empirical animal tracking data for a water mongoose *Atilax paludinosus*, a caracal *Caracal caracal*, and a serval *Leptailurus serval*, and discuss pathways for continuing to improve upon our method. We also provide an annotated analytical workflow for using the ctmm R package (Calabrese et al., 2016) to apply our method to animal tracking data (Appendix S1).

### 2 | MATERIALS AND METHODS

#### 2.1 | Mathematical concepts and definitions

In a conventional RSF, the log-likelihood of n IID samples of an homogeneous Poisson point process model is given by

\[
\ell(\text{data}|\lambda) = \sum_{i=1}^{n} \log \lambda(x(t_i), y(t_i)) - n \log \int \lambda(X, Y) dX dY, \tag{1}
\]

where \(\lambda(x, y)\) is the intensity function, which contains all model parameters, and where, for simplicity, we have assumed that \(\lambda\) does not change in time. This likelihood function is typically approximated numerically via weighted logistic regression, and \(\lambda\) is usually constructed to be an exponential model (Fieberg et al., 2021; Northrup et al., 2022), but other approaches have also been proposed and used (e.g. Cooper & Millspaugh, 1999; Lele & Keim, 2006; Nielson & Sawyer, 2013).

Within \(\lambda\), we include area terms \((x, y)\) and \(x^2 + y^2\) that produce a bivariate Gaussian distribution of animal locations so that when no resource selection occurs, the weighted likelihood reproduces a Gaussian home range estimate that approximates the maximum likelihood estimate of the autocorrelation model. This Gaussian distribution becomes the null RSF model (i.e. the domain of availability, and the area of movement when there is no habitat selection; Fieberg et al., 2021, Horne et al., 2008). The effective sample size is roughly the number of times an animal has crossed the linear extent of its home range, which dictates how well the area and shape of the Gaussian home range estimate can be resolved from the sampled movement path of an animal (Fleming et al., 2019). When selection parameters are supported, then we obtain a synoptic model of habitat selection (sensu Horne et al., 2008) within a Gaussian area.
Further details of this configuration of the RSF model are found in Appendix S3.

To the conventional IID log-likelihood, we incorporate our RSF log-likelihood weights, which are given by

\[ w(t_i) = w_{AKDE}(t_i) N, \tag{2} \]

where \( w_{AKDE}(t_i) \) denotes weights optimized for non-parametric autocorrelated kernel density estimation at each sampled location (Fleming et al., 2018), which sum to 1. \( N \leq n \) is the effective sample size of the autocorrelated Gaussian area estimate so that our weights, \( w(t_i) \), sum to \( N \). The AKDE weights minimize bias in nonparametric kernel density estimation, where for a sample of \( n \) \( q \)-dimensional locations \( r(t_i) \) at times \( t_i \), a weighted kernel density estimate can be represented as

\[ \hat{p}(r) = \sum_{i=1}^{n} w(t_i) k(r-r(t_i)), \quad \sum_{i=1}^{n} w(t_i) = 1, \tag{3} \]

where \( k \) denotes a Gaussian kernel with covariance \( \sigma_k \) (the bandwidth matrix). Both the bandwidth matrix and the weight vector \( w \) with \( w_i = w(t_i) \) are optimized to minimize the mean integrated square error

\[ \text{MISE}(\sigma_k, w) = \left( \int d^q r |p(r) - \hat{p}(r)|^2 \right). \tag{4} \]

where \( \int d^q r \) denotes the \( q \)-dimensional volume integral and \( (L) \) denotes the expectation value with respect to the distribution of the data, \( r(t_i) \), which may be autocorrelated. Because the true density function \( p(r) \) is unknown and kernel density estimation is non-parametric, the MISE (4) must be approximated, and different approximations correspond to different methods of kernel density estimation—all being asymptotically optimal (Izenman, 1991; Silverman, 1986; Turlach, 1993).

The inclusion of area terms in the RSF model is a form of parametric Gaussian density estimation, which allows weights to serve a similar purpose in RSFs as for kernel density estimation. Weights (2) also provide identical estimates of the mean location in both kernel and Gaussian density estimation, indicating that they provide a bias correction for the Gaussian model. The weights \( w(t_i) \) are used to weight each sampled location so that the final log-likelihood is of the form

\[ \ell(\text{data}| \lambda) = \sum_{i=1}^{n} w(t_i) \log \lambda(x(t_i), y(t_i)) - N \log \int \lambda(X, Y) dX dY. \tag{5} \]

These weights can be used to re-weight data points for any method of approximating inhomogeneous Poisson point processes, but details of implementation will vary for different methods and software platforms. Likelihood (5) can be viewed as an example of a `composite likelihood’ (Varin et al., 2011) that produces an unbiased estimating equation for the mean location, but with the additional property that the total weight is fixed to the effective sample size.

### 2.2 Simulations

To validate our theoretical argument and demonstrate the value of our approach on animal location data, we performed three sets of simulations. For all three sets of simulations, we started by simulating movement paths for animals following an isotropic Ornstein-Uhlenbeck Foraging movement model (Fleming et al., 2014) with a location variance \( \sigma^2 \) of 200,000 m², velocity persistence time-scale \( \tau_0 \) of 1/3 day and home range crossing time-scale \( \tau_0 \) of 1 day. This is equivalent to an animal with a circular home range of 3.76 km², with three correlated bouts of movement each day, and which crosses its home range roughly once per day. Because range crossings occurred daily, the effective sample size in this movement track is equivalent to the number of days the movement track is sampled, and sampling locations from the movement track move more frequently than once per day yields autocorrelated data. We overlaid each movement track on a raster consisting of equal amounts of two habitat types (hereafter, Habitat 1 and Habitat 2) that each take up half of the raster surface. All simulations were performed using the ctmm R package (v0.6.2; Calabrese et al., 2016) in the R statistical software environment (v3.6.2; R Core Team, 2020). Habitat rasters were created using the raster R package (v3.4-10; Hijmans, 2021).

#### 2.2.1 Scenario 1: Absolute vs. effective sample size

We first simulated a scenario that demonstrates that our weighting scheme does not reduce bias simply by inflating confidence intervals such that it becomes impossible to identify habitat selection when it occurs. For this scenario, we simulated habitat selection occurring by arraying habitat in 10 m vertical strips and then simulated habitat-independent movement tracks over this layer. We sampled these tracks while varying (1) the sampling rate (2, 4, 8, 16 and 32 times per day for 90 days) or (2) the sampling duration (12 times per day for 16, 32, 64, 128 and 256 days). We then shifted every other location in Habitat 1 further right by 10 m, which moved those locations into Habitat 2. This alteration changed the fit of the underlying movement model very little, but led to the animals being located in Habitat 2 roughly three times as often as they were located in Habitat 1, emulating habitat selection. Finally, we used weighted RSFs to estimate strength of selection for Habitat 2 compared with Habitat 1, and the confidence intervals around this parameter estimate. In this scenario, parameter estimates should be constant around the expected parameter (ln(3)) in all scenarios, but confidence intervals should only meaningfully contract when the effective sample size (i.e. sampling duration) increases, and not just when the temporal resolution of the data increases.
2.2.2 | Scenario 2: Data thinning

We then simulated a scenario that demonstrates that data thinning creates substantially more variation around RSF parameter estimates than weighting. As in Scenario 1, we simulated habitat selection occurring by arraying habitat in 10 m vertical stripes and then simulated a habitat-independent movement track over this layer. The track was sampled 50 times per day for 64 days. We then shifted every other location in Habitat 1 further right by 10 m, which would be the spatial configuration of habitat affects estimates of resource selection (Northrup et al., 2011). We then created 50 subsampled movement tracks (all possible regular subsamples of this data in which the absolute sample size is roughly equal to the effective sample size, rendering locations independent from one another). We used a weighted RSF to estimate strength of selection for Habitat 2 compared with Habitat 1, and the confidence intervals around this parameter estimate, for the full movement track. We used IID RSFs to estimate strength of selection for Habitat 2 compared with Habitat 1, and the confidence intervals around this parameter estimate, for each subsampled movement track. In this scenario, the parameter estimates for the weighted RSF should be near the expected parameter (ln(3)), but thinning the data will create substantial variation in IID parameter estimates because of loss of information about the movement track.

2.2.3 | Scenario 3: Triangulation failure

We finally simulated a scenario in which bias could arise when certain habitat covariates of interest cause disproportionate amounts of triangulation failure in GPS devices. This is known to happen, for example, in rugged terrain, thick vegetation and underwater (e.g. Breed et al., 2011; O’Neill et al., 2020; Streicher et al., 2021). For this scenario, we simulated no habitat selection occurring (i.e. the animal movement path was simulated independently of the habitat raster), but we censored every other location in Habitat 2, which would be consistent with a habitat-specific triangulation success rate of 50% (an extreme case of habitat-specific triangulation failure). Because the spatial configuration of habitat affects estimates of resource selection (Northrup et al., 2013), we configured our habitat rasters in two different ways to illustrate how weighting responds to habitat configuration: (1) a highly clustered case, in which the two habitats were arrayed in two large contiguous blocks and (2) an unclustered case where the habitats were arrayed in alternating vertical stripes that were 10 m in width (identical to the raster in Scenarios 1 and 2). We sampled these tracks 1, 2, 4, 8 and 16 times per day, for 90 days, on 400 movement tracks in each habitat configuration. We finally used weighted and IID RSFs to estimate strength of selection for Habitat 1 compared with Habitat 2, and the confidence intervals around this parameter estimate.

This scenario demonstrates two useful properties of our weighting scheme. First, by varying the sampling rate, we can demonstrate that an RSF that treats data as IID becomes increasingly confident in its parameter estimates as data become more autocorrelated—even when they are biased—while weighted RSFs maintain stable confidence intervals as locations are sampled more frequently because the effective sample size of the data set is not increasing. In other words, reducing the time interval between sampled animal locations causes the confidence intervals around selection parameters to become narrower in IID RSFs (because increasing autocorrelation in the data causes pseudoreplication) but not in weighted RSFs (because autocorrelation is explicitly modelled and accounted for). Second, by simulating habitat-specific location failure, we demonstrate that this weighting scheme can reduce sampling-induced bias in RSF parameter estimates compared with RSFs that assume data are IID. An IID RSF should detect a pattern of selection of Habitat 1 over Habitat 2 that is a statistical artefact of triangulation failure; a weighted RSF will shift the parameter estimate towards zero and expand the confidence intervals around the parameter estimate, correctly inferring that no habitat selection is occurring when effective sample sizes are large enough to reliably estimate parameters. However, its ability to do this depends on the degree of spatial clustering of habitat covariates—which is known to influence RSF parameter estimates (Northrup et al., 2013; Street et al., 2021). As more consecutive locations occur within a single habitat type, the average weights of locations in the habitat type with frequent triangulation failure become larger, allowing the model to reduce the influence of the habitat-specific sampling intervals on parameter estimates.

2.3 | Empirical examples

To demonstrate the application of our method on real-world animal location data, we performed IID and weighted RSFs on tracking data from a water mongoose (144 locations over 58 days; Appendix S2, Figure S1), a caracal (504 locations over 85 days; Appendix S2, Figure S2), and a serval (3603 locations over 321 days; Appendix S2, Figure S3). All animals were tracked using GPS telemetry; further details on the specifics of animal capture and tracking can be found in Streicher et al. (2021), Ramesh et al. (2016a, 2016b). All three animals are missing a meaningful number of locations because of GPS triangulation failure. For each animal, we estimated the effects of two land cover types (built-up [human settlements, roads, railways, and airfields] and plantation [exotic tree plantations]) on their habitat selection compared with a grouped reference category consisting of the remaining land cover types. For our land cover data, we used a land-use map with 20-m resolution from Ezemvelo KZN Wildlife (EKZN Wildlife & GeoTerraImage, 2018) reclassified into the three categories of interest (i.e. plantation, built-up and reference).

For each animal, we used variogram analysis (Fleming et al., 2014) to ensure animals were range-resident, fit and selected an autocorrelated movement model that best described the animal’s movements using perturbative Hybrid Residual Maximum Likelihood (Fleming et al., 2019) and Akaike’s information criterion corrected for small sample sizes, estimated utilization distributions for each animal (wAKDE for weighted RSFs [Fleming et al., 2018]; conventional KDE for IID RSFs [Worton, 1989]), and fit RSFs using the ctmm R
package (v0.6.2; Calabrese et al., 2016) in the R statistical software environment (v3.6.2; R Core Team, 2020).

For the caracal, we then examined how using IID RSFs on thinned datasets would alter inferences gained from this dataset. To do this, we created 27 subsampled movement tracks (all possible regular subsamples of this data in which the absolute sample size is roughly equal to the effective sample size, rendering locations independent from one another). We used a weighted RSF to estimate strength of selection for built-up areas (accounting for selection for plantation), and the confidence intervals around this parameter estimate, for the full movement track. We used IID RSFs to estimate strength of selection for built-up areas (accounting for plantation), and the confidence intervals around this parameter estimate, for each subsampled movement track.

3 | RESULTS

3.1 | Simulations

3.1.1 | Scenario 1: Absolute vs. effective sample size

In our simulations of habitat selection, we found that weighted RSFs performed as expected, with parameter estimates of weighted RSFs remaining consistently around the expected parameter (ln(3); Figure 1). The width of confidence intervals remained stable as the sampling rate (i.e. autocorrelation) increased (Figure 1a), but contracted as sampling duration (i.e. effective sample size) increased (Figure 1b). Weighted RSFs could detect habitat selection when effective sample sizes were ≥32 (or 32 days) in this example.

3.1.2 | Scenario 2: Data thinning

In our simulations of data thinning, we found that RSFs performed as expected, with the parameter estimates of the weighted RSF very close to the expected parameter (ln(3)) while the IID RSFs parameterized using thinned datasets exhibited substantial variation in parameter estimates (range: [-1.94, -0.44]; Figure 2). The expected parameter fell within 48 out of 50 (96%) of the IID RSF 95% confidence intervals, indicating that our measure of effective sample size is a reasonable measure of independence. One IID RSF failed to identify statistically significant habitat selection.

3.1.3 | Scenario 3: Triangulation failure

In our simulations of triangulation failure, we found that weighted and IID RSFs performed as expected. Across 400 individual movement paths, parameter estimates of weighted RSFs were closer to zero than parameter estimates of IID RSFs (Figure 3; Table 1). Confidence intervals around parameter estimates of weighted RSFs contained zero much more often (Table 1, top half; Figures 3a,c) than confidence intervals around parameter estimates of IID RSFs (Table 1, bottom half; Figures 3b,d). Confidence intervals around parameter estimates of IID RSFs became narrower as the sampling rate (and thus autocorrelation) increased, while the width of confidence intervals around parameter estimates of weighted RSFs remained similar across all sampling rates (Figure 3). Notably, weighted RSFs were better able to successfully identify a lack of habitat selection (i.e. weighted RSF parameter estimates became increasingly closer to zero and a higher proportion of confidence intervals contained zero) as the sampling rate increased, while IID RSFs performed worse as the sampling rate increased (i.e. parameter estimates were largely consistent, but confidence intervals around the [biased] estimates contracted sharply as the sampling rate increased). The benefits of weighted RSFs are also enhanced when habitat is more highly clustered, but likelihood weighting still improves both parameter estimates and confidence intervals when habitat is unclustered (Table 1).

3.2 | Empirical examples

When analysing the water mongoose data (Figure 4a), we found that both IID and weighted RSFs identified statistically significant

![Figure 1](https://besjournals.onlinelibrary.wiley.com/doi/10.1111/2041-210X.14025)

**FIGURE 1** Mean parameter estimates and confidence intervals around those parameter estimates for weighted resource selection functions (RSFs) during simulations of habitat selection. In this case, the animal is selecting habitat such that its movement path contains three times as many locations in Habitat 2 as in Habitat 1. Panel (a) shows the performance of weighted RSFs as the sampling rate (i.e. autocorrelation) increases, while panel (b) shows the performance of weighted RSFs as sampling duration (effective sample size) increases. As expected, point estimates of the selection parameter are constant across all scenarios, while confidence in those parameters increases as sampling duration (i.e. effective sample size), but not sampling rate (i.e. autocorrelation), increases.
avoidance of plantation areas, but the point estimate of avoidance of plantation areas for the weighted RSF lies closer to zero and outside the 95% confidence interval generated by the IID RSF (\(\hat{\beta}_{\text{IID}} = -1.23\) vs. \(\hat{\beta}_w = -1.17\); CIw = [−8.25, −1.45]). Moreover, although point estimates for selection of built-up areas were similar (\(\hat{\beta}_w = 1.68\) vs. \(\hat{\beta}_{\text{IID}} = 1.58\)), confidence intervals were substantially wider for the weighted RSF (CIw = [0.67, 2.69] vs. CI\(\text{IID}\) = [1.39, 1.78]).

When analysing the serval data (Figure 4c), we found that although point estimates for avoidance of plantation areas were similar between weighted and IID RSFs (\(\hat{\beta}_w = -2.04\) vs. \(\hat{\beta}_{\text{IID}} = -1.88\)), avoidance of plantation areas was only statistically significant for the IID RSF.
RSFs (CIw = [-5.65, 1.56] vs. CI_{ID} = [-2.47, -1.30]). Point estimates for avoidance of built-up areas were also similar between weighted and IID RSFs ($\hat{\phi}_w = -0.57$ vs. $\hat{\phi}_{ID} = -0.66$), but this avoidance was again only statistically significant for the IID RSF (CIw = [-1.88, 0.73] vs. CI_{ID} = [-0.90, -0.43]).

When analysing how data thinning would influence inferences of habitat selection by the caracal, we found that IID RSFs parameterized using subsets of the caracal movement track exhibited substantial variation around the parameter estimate provided by the weighted RSF (Figure 5). The parameter identified by the weighted RSF fell within 27 out of 27 (100%) of the IID RSF 95% confidence intervals, but 7 out of 27 (26%) of the IID RSFs did not identify statistically significant habitat selection for this individual.

Our empirical analyses demonstrate that inferences gained from weighted and IID RSFs can diverge substantially. Weighted RSFs produce wider confidence intervals than conventional RSFs and meaningfully different parameter estimates when missing data are common, indicating that the results of our simulations translate to real-world data, even at absolute sample sizes that are relatively small for modern GPS datasets. When data are autocorrelated, thinning data allows IID RSFs to generate more robust confidence intervals, but with more variation in parameter estimates than weighted RSFs.

### TABLE 1

| RSF type | Habitat clustering | Parameter Est. | % non-significant |
|----------|--------------------|----------------|------------------|
| wRSF     | High               | -0.59          | 83.8             |
|          | 2                  | -0.43          | 93.5             |
|          | 4                  | -0.23          | 99.3             |
|          | 8                  | -0.10          | 99.3             |
|          | 16                 | -0.07          | 99.5             |
|          | Low                | -0.60          | 51.5             |
|          | 2                  | -0.55          | 46.3             |
|          | 4                  | -0.50          | 58.0             |
|          | 8                  | -0.49          | 60.8             |
|          | 16                 | -0.49          | 62.5             |
| IID      | High               | -0.68          | 61.0             |
|          | 2                  | -0.68          | 37.5             |
|          | 4                  | -0.71          | 15.8             |
|          | 8                  | -0.71          | 7.3              |
|          | 16                 | -0.74          | 3.8              |
|          | Low                | -0.69          | 24.3             |
|          | 2                  | -0.68          | 1.5              |
|          | 4                  | -0.69          | 0.0              |
|          | 8                  | -0.69          | 0.0              |
|          | 16                 | -0.69          | 0.0              |

**FIGURE 4** Parameter estimates and confidence intervals for independent and identically distributed (IID) (white points) and weighted (black points) resource selection functions during empirical tests on a water mongoose (panel a), a caracal (panel b) and a serval (panel c). Weighting meaningfully alters point estimates of selection for built-up areas for the mongoose, and appropriately expands confidence intervals around parameter estimates, changing inferred habitat selection for all three animals.
become artificially more confident in this bias as the sampling rate of selection of those habitats are negatively biased, and models handle frequent habitat-induced triangulation failure—estimates that do not allow regular sampling intervals. As shown by our animal behaviour at multiple temporal scales, or technological constraints.

Rigorous estimation of confidence intervals on RSF parameters is a challenge using conventional RSF methods. Because consecutive data points in modern animal tracking data are rarely statistically independent, using all the data available in an autocorrelated movement track leads to increasing (and increasingly misleading) confidence in parameter estimates with increasing sampling rates. Data thinning can be used to reduce autocorrelation between successive locations, which can widen confidence intervals that are known to be too narrow. In practice, however, data thinning inherently involves loss of useful information on an animal’s movements, is generally applied in an ad hoc and non-optimized way and can pose other statistical problems (Noonan et al., 2020). Generalized estimating equations can also be used to estimate more robust confidence intervals while using all the data in a movement track, but the ‘cluster’ sizes (akin to effective sample sizes) are often also chosen arbitrarily (Prima et al., 2017), and it is difficult to implement generalized estimating equations jointly with other common techniques to mitigate statistical issues in RSFs (e.g., random effects). The method we propose in this study offers an objective way to estimate effective sample sizes for generating reliable confidence intervals.

In addition to estimating more robust confidence intervals, our method of weighting also offers potential to mitigate bias that can arise from inadequate sampling of landscape features that cause triangulation failure, irregular sampling designs intended to document animal behaviour at multiple temporal scales, or technologies that do not allow regular sampling intervals. As shown by our simulations (Table 1; Figure 3), RSFs that assume IID data cannot handle frequent habitat-induced triangulation failure—estimates of selection of those habitats are negatively biased, and models become artificially more confident in this bias as the sampling rate increases. Our method of weighting reduces this bias, and the quality of estimated parameters improves rather than declines as the sampling rate increases (however, we note that the extent to which inference is improved is dependent upon the spatial arrangement of environmental covariates). Duty cycling, by which researchers periodically suspend or reduce the sampling rate in an attempt to collect high-resolution data over longer total periods of time while conserving battery life, offers another promising use case for our method. For example, researchers may be interested in the foraging behaviour of a crepuscular animal and therefore program a tracking device to collect more locations near dawn and dusk, when an animal is actively foraging. In this case, an IID RSF would result in sampling foraging areas out of proportion to other areas where an animal spends its time, potentially underestimating use of landscape features that play important roles as rest sites, escape cover or thermal refugia. A weighted RSF could down-weight oversampled crepuscular locations more than undersampled locations during the day and night, providing more objective parameter estimates for overall resource selection. Finally, many species are not amenable to wearing tracking devices that acquire fixes at regular intervals. Aquatic or semi-aquatic species may spend long periods of time underwater where they are inaccessible to satellites (Breed et al., 2011; Fleming et al., 2018), and many species remain too small for today’s GPS technology (McMahon et al., 2017; Weller et al., 2016). Weighted RSFs provide a way for researchers to conduct RSFs that are less biased by such technological constraints.

Although applied here to tracking data of individual animals, the statistical principles underlying our method are also applicable at the species and population levels, which would be of great use to researchers. Random effects are often used to account for non-independence of data for individuals within a population. Gillies et al. (2006) recommended using random intercepts to account for unequal sample sizes between individuals in studies of habitat selection, and random slopes to account for differences in habitat selection between individuals. Similarly, Hebblewhite and Merrill (2008) recommended using random intercepts to account for non-independence arising from correlated locations in socially
structured populations (e.g. repeated observations from individuals within the same herd or pack). Incorporating random effects into RSFs and SSFs has since become standard practice (Fieberg et al., 2010; Muff et al., 2020), but remains an imperfect solution for accounting for pseudoreplication. Random effects generate confidence intervals that depend on variation in the point estimates of RSF parameters between individuals, which may or may not reflect true parameter uncertainty. If autocorrelation in individual movement tracks is not properly accounted for, individual sampling variance is underestimated, which inflates population variance estimated using random effects. Checking for this is difficult and rarely (if ever) performed. Population-level likelihood weighting, however, has potential to serve similar purposes. Weighting based on effective sample sizes can account for individuals having been tracked for differing amounts of time, and autocorrelation-informed weighting can account for differing amounts of spatial or temporal autocorrelation between individuals in a study area or study period. Iterated over both individuals and populations, such weight optimization accounts for temporal sampling bias within individual time-series, as it did for the single-individual weighting described in this study, while also accounting for sampling biases among individuals, where certain individuals are better sampled than others.

Because location weights are generated from range distributions using weighted autocorrelated kernel density estimation, which can only be validly calculated on animals that are range-resident, this method cannot be used to conduct RSFs on individuals that are not range-resident (e.g. dispersing or migratory individuals). However, it should be noted that because conventional IID RSFs assume stationary distributions of resource use and availability, conventional RSFs also assume range residency (but this assumption is often ignored). SSFs are therefore more appropriate for studying resource selection by migratory, dispersing or nomadic animals because they do not assume stationary distributions of use and availability. The inferences gained from such studies should be treated with caution, however, because autocorrelation can still lead to pseudoreplication and bias in inferences gained from SSFs of these movement tracks.

Although ecologists and conservation biologists have access to better statistical and computational tools than ever before, new methods are still required to maximize the value of animal tracking data for informing our understanding of the natural world. In this study, we introduced a method of autocorrelation-informed weighting to reduce pseudoreplication and bias when parameterizing RSFs with autocorrelated and irregular animal tracking data, which are pervasive in studies of animal movement. Our method is easily implemented using the ctmR R package (Calabrese et al., 2016), and we provide an annotated workflow in our supplementary materials (Appendix S1) so that other researchers can implement weighted RSFs on their own animal location data. We hope that the analytical technique we have provided here, which is grounded in statistical theory and validated using simulations and empirical data, allows continued progress towards more statistical rigour in studies of resource selection by animals.

**AUTHOR CONTRIBUTIONS**

Christen H. Fleming, Björn Reineking and Justin M. Calabrese conceived the ideas; Christen H. Fleming and Björn Reineking led the mathematical derivation and software implementation; Jesse M. Alston conducted the simulations and empirical analyses; Jarryd P. Streicher, Colleen T. Downs and Tharmalingam Ramesh contributed the empirical data; and Jesse M. Alston and Christen H. Fleming led the writing of the manuscript. All authors contributed critically to drafts of the manuscript and gave final approval for publication.

**ACKNOWLEDGEMENTS**

We thank I. Silva for providing feedback on the weighted RSF tutorial in Appendix S1. J. Calabrese and C. Fleming were supported by NSF IIBR 1915347. R. Kays was supported by NSF ABI 1914928 and NASA Ecological Informatics 80NSSC21K1182. This work was partly funded by the Center for Advanced Systems Understanding (CASUS) which is financed by Germany’s Federal Ministry of Education and Research (BMBF) and by the Saxon Ministry for Science, Culture and Tourism (SMWK) with tax funds on the basis of the budget approved by the Saxon State Parliament.

**CONFLICT OF INTEREST**

We declare no conflict of interest.

**PEER REVIEW**

The peer review history for this article is available at https://publons.com/publon/10.1111/2041-210X.14025.

**DATA AVAILABILITY STATEMENT**

All data and code (Alston et al., 2022) used in analyses for this manuscript are publicly available on the RODARE data repository at http://doi.org/10.14278/rodare.1862.

**ORCID**

Jesse M. Alston  https://orcid.org/0000-0001-5309-7625
Christen H. Fleming  https://orcid.org/0000-0002-9356-6518
Roland Kays  https://orcid.org/0000-0002-2947-6665
Colleen T. Downs  https://orcid.org/0000-0001-8334-1510
Tharmalingam Ramesh  https://orcid.org/0000-0001-6477-4425
Justin M. Calabrese  https://orcid.org/0000-0003-0575-6408

**REFERENCES**

Aarts, G., Fieberg, J., Brasseur, S., & Matthiopoulos, J. (2013). Quantifying the effect of habitat availability on species distributions. Journal of Animal Ecology, 82, 1135–1145.

Aarts, G., Mackenzie, M., McConnell, B., Fedak, M., & Matthiopoulos, J. (2008). Estimating space-use and habitat preference from wildlife telemetry data. Ecography, 31, 140–160.

Ali, A. H., Ford, A. T., Evans, J. S., Mallon, D. P., Hayes, M. M., King, J., Amin, R., & Goheen, J. R. (2017). Resource selection and landscape change reveal mechanisms suppressing population recovery for the world’s most endangered antelope. Journal of Applied Ecology, 54, 1720–1729.

Alston, J. M., Fleming, C. H., Kays, R., Streicher, J. P., Downs, C. T., Ramesh, T., Reineking, B., & Calabrese, J. M. (2022). Data and
Muff, S., Signer, J., & Fieberg, J. (2020). Accounting for individual-specific variation in habitat-selection studies: Efficient estimation of mixed-effects models using Bayesian or frequentist computation. *Journal of Animal Ecology, 89*, 80–92.

Mysterud, A., & Ims, R. A. (1998). Functional responses in habitat use: Availability influences relative use in trade-off situations. *Ecology, 79*, 1435–1441.

Newdick, L., Prokopenko, C. M., & Vander Wal, E. (2022). Individual differences in habitat selection mediate landscape level predictions of a functional response. *Oecologia, 198*, 99–110.

Ng’weno, C. C., Ford, A. T., Kibungei, A. K., & Goheen, J. R. (2019). Interspecific prey neighborhoods shape risk of predation in a savanna ecosystem. *Ecology, 100*, e02698.

Nielsen, S. E., Boyce, M. S., Stenhouse, G. B., & Munro, R. H. M. (2002). Modeling grizzly bear habitats in the Yellowstone ecosystem of Alberta: Taking autocorrelation seriously. *Ursus, 13*, 45–56.

Nielsen, R. M., & Sawyer, H. (2013). Estimating resource selection with count data. *Ecology and Evolution, 3*, 2233–2240.

Noonan, M. J., Fleming, C. H., Tucker, M. A., Kays, R., Harrison, A. L., Crofoot, M. C., Abrahms, B., Alberts, S. C., Ali, A. H., Altmann, J., Antunes, P. C., Attías, N., Belant, J. L., Beyer, D. E., Bidner, L. R., Blaum, N., Boone, R. B., Caillaud, D., Paula, R. C. D., ... Calabrese, J. M. (2020). Effects of body size on estimation of mammalian area requirements. *Conservation Biology, 34*, 1017–1028.

Noonan, M. J., Tucker, M. A., Fleming, C. H., Akre, T. S., Alberts, S. C., Ali, A. H., Altmann, J., Antunes, P. C., Attás, N., Belant, J. L., Beyer, D. E., Bidner, L. R., Blaum, N., Boone, R. B., Caillaud, D., Paula, R. C. D., Dekker, J., Drescher-Lehman, J., Farwig, N., Fichtel, C., Fischer, C., ... Calabrese, J. M. (2019). A comprehensive analysis of autocorrelation and bias in home range estimation. *Ecological Monographs, 89*, e01344.

Northrup, J. M., Hooten, M. B., Anderson, C. R., Jr., & Wittemyer, G. (2013). Practical guidance on characterizing availability in resource selection functions under a use–availability design. *Ecology, 94*, 1456–1463.

Northrup, J. M., Vander Wal, E., Bonar, M., Fieberg, J., Lafarge, M. P., Leclerc, M., Prokopenko, C. M., & Gerber, B. D. (2022). Conceptual and methodological advances in habitat-selection modeling: Guidelines for ecology and evolution. *Ecological Applications, 32*, e02470.

O’Neill, H. M. K., Durant, S. M., & Woodroffe, R. (2020). What wild dogs want: Habitat selection differs across life stages and orders of selection in a wide-ranging carnivore. *BMCL Zoology, 5*, 1.

Prima, M. C., Duchesne, T., & Fortin, D. (2017). Robust inference from conditional logistic regression applied to movement and habitat selection analysis. *PloS ONE, 12*, e0197779.

Prokopenko, C. M., Boyce, M. S., & Avgar, T. (2017a). Characterizing wildlife behavioural responses to roads using integrated step selection analysis. *Journal of Applied Ecology, 54*, 470–479.

Prokopenko, C. M., Boyce, M. S., & Avgar, T. (2017b). Extent-dependent habitat selection in a migratory large herbivore: Road avoidance across scales. *Landscape Ecology, 32*, 313–325.

R Core Team. (2020). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing.

Ramesh, T., Kalle, R., & Downs, C. T. (2016a). Space use in a south African agriculture landscape by the caracal (*Caracal caracal*). *European Journal of Wildlife Research, 63*, 11.

Ramesh, T., Kalle, R., & Downs, C. T. (2016b). Spatiotemporal variation in resource selection of servals: Insights from a landscape under heavy land-use transformation. *Journal of Mammalogy, 97*, 554–567.

Rayl, N. D., Proffitt, K. M., Almberg, E. S., Jones, J. D., Merkle, J. A., Gude, J. A., & Cross, P. C. (2019). Modeling elk-to-livestock transmission risk to predict hotspots of brucellosis spillover. *The Journal of Wildlife Management, 83*, 817–829.

Robb, B. S., Merkle, J. A., Sawyer, H., Beck, J. L., & Kauffman, M. J. (2022). Nowhere to run: Semi-permeable barriers affect pronghorn space use. *Journal of Wildlife Management, 86*, e22212.

Scantlebury, D. M., Mills, M. G. L., Wilson, R. P., Wilson, J. W., Mills, M. E. J., Durant, S. M., Bennett, N. C., Bradford, P., Marks, N. J., & Speakman, J. R. (2014). Flexible energetics of cheetah hunting strategies provide resistance against kleptoparasitism. *Science, 346*, 79–81.

Signer, J., Fieberg, J., & Avgar, T. (2017). Estimating utilization distributions from fitted step-selection functions. *Ecosphere, 8*, e01771.

Signer, J., Fieberg, J., & Avgar, T. (2019). Animal movement tools (amt): R package for managing tracking data and conducting habitat selection analyses. *Ecology and Evolution, 9*, 880–890.

Silverman, B. W. (1986). *Density estimation for statistics and data analysis*. vol. 26 of monographs on statistics and applied probability. *Chapman & Hall*.

Street, G. M., Potts, J. R., Börger, L., Beasley, J. C., Demaraïs, S., Fryxell, J. M., McLoughlin, P. D., Monteith, K. L., Prokopenko, C. M., Ribeiro, M. C., Rodgers, A. R., Strickland, B. K., van Beest, F. M., Bernasconi, D. A., Beumer, L. T., Dharmarajan, G., Dwinnell, S. P., Keiter, D. A., Keuroghlian, A., ... Vander Wal, E. (2021). Solving the sample size problem for resource selection functions. *Methods in Ecology and Evolution, 12*, 2421–2431.

Streicher, J. P., Ramesh, T., & Downs, C. T. (2021). An African urban mesocarnivore: Navigating the urban matrix of Durban, South Africa. *Global Ecology and Conservation, 26*, e01482.

Stuber, E. F., Carlson, B. S., & Jesmer, B. R. (2022). Spatial personalities: A meta-analysis of consistent individual differences in spatial behaviors. *Behavioral Ecology, 33*, 477–486.

Swihart, R. K., & Slade, N. A. (1985). Testing for independence of observations in animal movements. *Ecology, 66*, 1176–1184.

Thurfjell, H., Ciuti, S., & Boyce, M. S. (2014). Applications of step-selection functions in ecology and conservation. *Movement Ecology, 2*, 4.

Turlach, B. A. (1993). Bandwidth selection in kernel density estimation: A review. *Tech. rep., CORE and Institut de Statistique.*

Ullmann, W., Fischer, C., Kramer-Schadt, S., Pirhofer-Walzl, K., Gleimnitz, M., & Blaum, N. (2020). How do agricultural practices affect the movement behaviour of European brown hares (*Lepus europaeus*). *Agriculture Ecosystems & Environment, 292*, 106819.

Valls-Fox, H., De Garine-Wichatitsky, M., Fritz, H., & Chamalé-Jammes, S. (2018). Resource depletion versus landscape complementation: Habitat selection by a multiple central place forager. *Landscape Ecology, 33*, 127–140.

van Beest, F. M., McLoughlin, P. D., Vander Wal, E., & Brook, R. K. (2014). Density-dependent habitat selection and partitioning between two sympatric ungulates. *Oecologia, 175*, 1155–1165.

Varin, C., Reid, N., & Firth, D. (2011). An overview of composite likelihood methods. *Statistica Sinica, 21*, 5–42.

Weller, T. J., Castle, K. T., Liechti, F., Hein, C. D., Schirmacher, M. R., & Cryan, P. M. (2016). First direct evidence of long-distance seasonal movements and hibernation in a migratory bat. *Scientific Reports, 6*, 34585.

Worton, B. J. (1989). Kernel methods for estimating the utilization distribution in home-range studies. *Ecology, 70*, 164–168.

**SUPPORTING INFORMATION**

Additional supporting information can be found online in the Supporting Information section at the end of this article.