HIV-1 DNA decay is faster in children who initiate ART shortly after birth than later

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Abstract

Introduction: There is limited data in children on whether persistence of HIV-1 infected cells is affected by age at initiating antiretroviral therapy (ART), its duration or any subsequent ART interruption. We therefore investigated the effects of both age of ART initiation and duration of ART interruption on HIV-1 DNA decay in children.

Methods: We investigated HIV-1 DNA decay in three groups of children on ART: Group-1 (n = 7) started uninterrupted ART within eight days of life; Group-2 (n = 8) started uninterrupted ART at a median of five months of age; and Group-3 (n = 23) started ART at a median age of 1.8 months for either 40 or 96 weeks, then interrupted ART (median of seven months), and restarted ART based on CD4 count and clinical criteria. Total HIV-1 DNA was assayed using a sensitive HIV-1 subtype C-adapted quantitative PCR for integrase. The duration of ART was square root transformed to fit the observed slowing of HIV-1 DNA decay rate. For each group, point estimates for decay rates were determined after six months of continuous suppressive ART in groups 1 and 2 or six months after restarting ART in Group-3. Groups-2 and 3 were combined using a mixed effect regression model to investigate covariates of HIV-1 DNA decay rate.

Results and Discussion: At six months of continuous suppressive ART, the HIV-1 DNA t½ (95% CI) was shorter in Group-1 (n = 7): 2.7 months (2.1 to 3.8), than 9.2 months (7.4 to 12.1) in Group-2 (n = 8); and 9.6 months (7.6 to 12.6) in Group-3 (n = 23) (p < 0.01). In multivariable analyses, HIV-1 DNA before treatment (p < 0.001) and the change in HIV-1 DNA during interruption (p < 0.01) were independent predictors of slower HIV-1 DNA decay.

Conclusions: These data suggest that ART initiation within the first week of life can reduce the persistence of long-lived infected cells. Delaying ART is associated with slower decay of infected cells.

Keywords: HIV-1 DNA kinetics; early treatment; early infant HIV-1 diagnosis; HIV-1 persistence; paediatrics; Africa

1 INTRODUCTION

ART initiation during early stages of HIV-1 infection, Fiebig I-IV, in adults [1] and in the first six months of life in children [2] is associated with lower levels of persistent HIV-1 infected cells. Some individuals, such as the Mississippi baby [3], who started ART at 30 hours of life, and adults who started ART between 10 and 12 days after infection [4], had delayed viral rebound after stopping ART, which is most likely due to a smaller pool of replication-competent proviruses that could stochastically reactivate [5]. Early therapy is also associated with an increased probability of post-treatment control in adults [6,7], and in two children [8,9]. A cross-sectional study of South African children revealed lower HIV-1 total DNA levels in those starting ART before two months-of-age compared to later [10]. There are limited data in children, however, on the influence of age at ART initiation and respective duration of initial ART and ART interruption on the levels of HIV-1 infected cells or their longitudinal decay. In a recent study of adults with an ART interruption for a median of 57 days, restarted when plasma HIV-1 RNA was >1000 copies/mL, for more than two weeks, there was no difference in intact HIV-1 sequences before and after interruption [11].

The Children with Early Antiretroviral Therapy (CHER) trial randomized children to early, time-limited ART or delayed continuous ART. Children were enrolled into the CHER study between August 2005 and December 2007 [12] when children were first diagnosed between four and six weeks of age, representing a mixture of perinatal and intra-uterine HIV-1 infection. As the risk-benefit of early versus delayed ART in perinatally infected children had not been known, children from the CHER trial were randomized to elective early time-limited ART: either 40 (ART-40W) or 96 weeks (ART-96W) of initial treatment, or continuous therapy, deferred until clinical progression or CD4 decline meeting concurrent ART start criteria (ART-Def) [13]. Children, whose ART was interrupted, reinitiated ART based on the same CD4 or clinical criteria,
resulting in variable periods off ART. All interrupted participants except one, from another site, met reinitiation criteria [8]. Despite a period of ART interruption, the CHER study showed that early elective ART had clinical and immunological benefit over delayed uninterrupted ART after a median follow-up period of 4.8 years on study [13]. Subsequently, ART programmes introduced infant HIV-1 PCR testing followed by immediate ART initiation to avert rapid disease progression and early mortality [12,14,15]. The impact of this very early treatment strategy on the long-term persistence of HIV-1-infected cells is unknown. Accordingly, the first aim of the current study was to compare the HIV-1 decay rate in three Groups of children: Group-1 who started continuous ART as soon as possible after birth [16], to two groups from the CHER trial: Group-2 started continuous ART at a median of five months of age; Group-3 initiated ART at a median of 1.8 months of age for either 40 or 96 weeks, and then interrupted ART for median of seven months, restarting ART based on CD4+ T cell count criteria. The second aim was to describe factors associated with the rate of HIV-1 DNA decay in the CHER participants.

2 | METHODS

2.1 | Participant selection

Parents or legal guardians provided written informed consent for all participants. The study was approved by the Stellenbosch University Health Research Ethics Committee: M14/07/029.

Children included in the CHER trial primary endpoint analysis which was time-to-failure of first-line ART (immunological/clinical/virological) or death had a CD4 percentage ≥25% at baseline. A small group with a baseline CD4% of <25% at screening, referred to as part B, commenced early ART, had samples stored and were retained in follow-up. Although some of them initially interrupted ART, the data safety monitoring board subsequently recommended that all Part B remain on continuous ART. Some children, who were randomized to time limited therapy (ART-96W), were not interrupted as they had already met a trial endpoint. Although plasma HIV-1 RNA was not measured during interruption, plasma samples were stored and some were available for retrospective plasma HIV-1 RNA testing. To compare data from interrupted and never-interrupted participants, HIV-1 DNA decay was assessed only during “continued” phase ART, defined as therapy initiation in never-interrupted patients or reinitiation of a protocol defined treatment interruption. All participants in the present study had at least one sample before continuous ART, that is, before treatment in never-interrupted children and before reinitiation in those interrupted. The following time points were included to study HIV-1 DNA kinetics: a baseline (pre-ART sample in never-interrupted children, or a sample during protocol-defined ART interruption, prior to reinitiating ART). Once back on ART, samples were collected closest to the following time points: six, twelve and eighteen months, during the trial. Thereafter samples were collected between two to three years and/or four to five years later. To assess HIV-1 decay kinetics in the absence of viraemia, participants were censored after the first of two viraemic episodes (plasma HIV-1 RNA >200 copies/mL) or excluded if less than two samples (including baseline) were collected before viraemia. Based on these criteria, 31 participants were included. Of these, eight individuals were never interrupted (Group-2), six because they were already in ART-Def and started a few months later and two randomized to ART-96W who had already met a trial endpoint. Twenty-three participants were interrupted based on randomization (Group-3).

HIV-1 DNA decay from children in the CHER study was compared to that of seven infants who initiated ART within eight days of life (Group-1). These children were diagnosed through a public sector birth diagnosis and early treatment initiation programme. Inclusion in this study was based on virological suppression defined as continuous downward trend in plasma HIV-1 RNA and no HIV-1 RNA >100 copies/mL at the first measurement after six months on ART [17].

2.2 | Sample processing and laboratory assays

Samples were processed and stored according to the HIV/AIDS Network Coordination (HANC) peripheral blood mononuclear cell (PBMC) processing standard operating procedure (https://www.hanc.info/labs/labresources/procedures/Pages/pbmcsop.aspx).

HIV-1 total DNA was extracted and measured by a sensitive quantitative PCR adapted for HIV-1 subtype C, targeting a conserved region in HIV-1 integrase (limit of detection: 3 copies/reaction), as previously described [16,18,19].

Plasma HIV-1 RNA monitoring was initially performed with the Roche Amplicor HIV Monitor assay version 1.0 (lower limit of detection (LOD) of 400 copies/mL), then with the Roche Amplicor HIV Monitor assay ultrasensitive protocol (LOD of 50 copies/mL). After completion of the CHER study in August 2011, the Abbott Diagnostics Realtime HIV-1 assay was used (LOD of 150 copies/mL for 200 µL input or 40 copies/mL for 1.0 mL input). The Roche CAP/CTM v2.0 assay (LOD of 100 copies/mL for 200 µL input or 20 copies/mL for 1 mL input) was used to assess plasma HIV-1 RNA in the seven infants initiated shortly after birth.

2.3 | Statistics and modelling

Statistical analyses were performed and graphics were generated using R 3.4.3 [20]. Because the rate of log_{10} HIV-1 DNA decline decreased with the time on ART, a square root transformation of time on ART improved the fit of the decay model as determined by conditional R^2 [21]. This model allowed exploring factors affecting decay over the full observed period, rather in separate phases. Viraemia-copy-years as cumulative measure of viraemia, during the period of interruption, was calculated as the product of the level of viraemia (log_{10} HIV-1 RNA copies/mL) multiplied by the duration of viraemia.

Separate mixed effect models with time treated as fixed effect and participant as random effect (random intercepts only) were used to describe decay of HIV-1 DNA in each of the three groups. As the decay rate decreased the longer participants were treated, the first derivative of the decay curve was used for estimates of HIV-1 DNA decay rate at six months on continued ART to compare HIV-1 DNA decay between groups. Wilcoxon Rank Sum Tests were used to
assess differences between groups: Linear models (log$_{10}$ HIV-1 DNA change against the square root of days treated) were fitted for each individual in each group; endpoint HIV-1 DNA was compared between Groups-2 and 3.

To assess which factors were independent predictors of HIV-1 DNA decay among the 31 CHER participants from the point of continued treatment, the following variables with a priori evidence of possible association with HIV-1 DNA decay, as fixed effects, were included in a full model: baseline HIV-1 DNA, pre-therapy HIV-1 plasma RNA and time-interrupted; participant was included as random effect (independent random intercepts and slopes with linear time effects).

### 3 | RESULTS

The baseline and clinical characteristics of the patients included are shown in Table 1.

Three groups of children were compared: Group-1 comprised seven infants who started continued ART within eight days of birth; median 5 (IQR: 0 to 7) days, Group-2 comprised eight children who started continuous ART at a median of 5.1 (IQR: 3.6 to 8.4) months, and Group-3 comprised 23 children who started ART at a median of 1.8 (IQR: 1.7 to 2.1) months but with ART interruption for a median of 7 (IQR: 4.6 to 9.3) months and who reinitiated at a median age of 20.2 (IQR: 16.0 to 30.9) months.

#### 3.1 | Endpoint HIV-1 DNA

In Group-1, HIV-1 DNA declined to <10 copies/million cells in six of seven (85.7%) infants at a median of 6.9 months on ART [17]. In Group-2, five of the eight children (62.5%) had samples collected, after at least four years of suppressive ART median 8.6 years). Here HIV-1 DNA became undetectable (<3 copies/million cells) in two children, declined to 20.0 copies/million cells in 2, and 39.8 copies/million cells in another child (the remaining three children were censored due to episodes of viraemia before four years on ART). In Group-3, 18 of the 23 (78.3%) had samples collected after at least four years (after a median of 7.8 years after resuming continuous ART), HIV-1 DNA became undetectable in 4, and declined to median of 3.1 (range 7.9 to 39.8) copies/million cells in another 14 (the remaining five were censored before four years on ART due to episodes of viraemia). Despite ART interruption in Group-3 there was no difference in endpoint HIV-1 DNA at a similar period on treatment between Groups-2 and 3 ($p = 0.5$) and no difference in total CD4 count median (IQR) at study end. Group-2: 1032 (814 to 1199); Group-3: 1088 (847 to 1294) cells/µl ($p = 0.7$).

#### 3.2 | Comparison of HIV-1 DNA decay rates

The conditional $R^2$ (95% CI) values for the model HIV-1 DNA decay estimates were 0.82 (0.65 to 0.93) for Group-1 (early start), 0.85 (0.67 to 0.94) for Group-2 (later start) and 0.79 (0.68 to 0.86) for Group-3 (interrupted), indicating an overall good model fit. The $t_\frac{1}{2}$ of HIV-1 DNA in Group-1 at six months of continuous treatment was 2.7 (95% CI: 2.1 to 3.8) months, being significantly shorter than both the eight children from Group-2 at 9.2 months (95% CI: 7.4 to 12.1, $p < 0.01$), and the 23 children from Group-3 (9.6 months; 95% CI: 7.6 to 12.6 months; $p < 0.01$) (Figure 1). Decay rates in the latter two groups were similar ($p = 0.81$).

#### 3.3 | Associations with HIV-1 DNA decay rates

Univariate analyses of the 31 CHER children, using a mixed effect model of decay, showed that only pre-treatment total HIV-1 DNA ($p < 0.001$) was a significant predictor of slower decay. Neither age at ART start, nadir CD4 count or nadir CD4%, baseline plasma HIV-1 RNA, time off ART, viraemia-

| Table 1. Participant characteristics |
|-------------------------------------|
| Participant characteristic          | Group-1: Early continued ART (n = 7) | Group-2: Suppressed viraemia and continued ART (n = 8) | Group-3: Suppressed viraemia and interrupted (n = 23) |
| Treatment regimen                   | ABC, 3TC, LPV/r$^a$               | AZT, 3TC, LPV/r$^b$                        | AZT, 3TC, LPV/r |
| CHER Study arms                     | NA                               | ART-Def (n = 6)                            | ART-40W (n = 15) |
| Age ART first initiated (days); median (IQR) | 5 (1.5 to 6.5)                   | 156.5 (110.3 to 256.8)                     | 55 (50.5 to 64.5) |
| Pre-treatment log 10 HIV-1 RNA load; median (IQR) | 3.1 (2.7 to 3.3)               | 5.6 (5.3 to 5.9)                           | 5.3 (4.0 to 5.8) |
| Baseline ART HIV-1 DNA copies/million cells; median (IQR) | 158 (40 to 398)               | 1107 (468 to 2999)                         | 832 (363 to 1371) |
| CD4% nadir$^c$; median (IQR)       | 40 (38.7 to 54.5)                | 16.9 (14.0 to 19.1)                        | 218 (15.9 to 25.2) |
| Absolute CD4 count nadir cells/microlitre$^d$; median (IQR) | 1955 (1193 to 2064)            | 505 (440.5 to 759.5)                       | 871 (577.5 to 1081.5) |
| Time interrupted (days); median (IQR) | -                              | -                                         | 214 (141 to 284) |
| Age at last sample (years); median (IQR) | 1.0 (0.6 to 1.0)               | 8.8 (1.3 to 9.7)                          | 9.2 (7.8 to 10.6) |
| CD4% (IQR) at last sample           | 34 (28.5 to 37.5)                | 34 (31 to 42)                             | 37 (35 to 39) |
| Absolute CD4 (IQR) (per microlitre) at last sample | 1920 (1440 to 2618)          | 1032 (814 to 1199)                        | 1088 (847 to 1294) |

$^a$Original regimen was AZT, 3TC, NVP, with NVP replaced by LPV/r at 42 weeks of age and AZT replaced by ABC at 3 months of age; $^b$group-2 included 2 ART-96W participants who were not interrupted as both had signs suggestive of HIV encephalopathy; $^c$part B of CHER had screening CD4 <25%; $^d$before continued phase of treatment.
copy-years during interruption, nor original CHER study arm, were significant predictors of the rate of decay.

In multivariable mixed effects models (Table 2), slower decay was associated with HIV-1 DNA level at baseline \( (p < 0.0001) \), faster decay with a higher baseline plasma HIV-1 RNA load \( (p = 0.033) \) and belonging to ART-96W or ART-40W (versus ART-Def) of the CHER study. Time-interrupted \( (p = 0.314) \), was not an independent significant predictor of HIV-1 DNA decay.

4 | DISCUSSION

The decay of HIV-1 DNA was much faster in children diagnosed at birth who started ART at a median of 5 days of life (Group-1) than those starting several weeks later, Groups-2 and 3, with no difference between the latter groups. In Groups-2 and 3 it is uncertain whether infection occurred in utero or intra-partum, which could have influenced decay rate, as in utero infection had time to become more established. In Groups-2 and 3, the strongest predictors of a slower decay rate was a higher pre-treatment HIV-1 DNA level. The median age of ART initiation was later in Group-2 (uninterrupted) than in Group-3 (interrupted), who reinitiated ART based on CD4 and clinical criteria. Therefore, those with more rapid deterioration during interruption were initiated faster. These factors could have counteracted any seeding of infected cells during interruption or effect of earlier ART.

Very early therapy may prevent the seeding of long-lived reservoirs such as CD4 clones that harbour replication
DNA decay than in the current study [28,29].

inhibitors (INSTI) which might have resulted in faster HIV-1 cells. No child in this study received integrase strand transfer reduce the seeding and persistence of long-surviving reservoir of HIV-1 infected cells is more labile and when ART may this suggests that there is a very early window when the pool

of very early treated children that involve intensive monitoring dates for functional cure studies. There is a need for studies

to therapy initiation. In addition, we had access to longitudinal samples of only 31 individuals with suppressed viraemia from the CHER study, a larger sample size may have enabled us to detect a smaller effect of treatment interruption on HIV-1 DNA decay. Moreover, although HIV-1 DNA levels decayed to undetectable levels in a number of children, this does not indicate the absence of persisting replication-competent reservoirs. Children in CHER were reinitiated based on CD4 and clinical criteria. One such study found no difference in the rate of decay in studies in children that describe longitudinal HIV-1 DNA data. There are few studies in children that describe longitudinal HIV-1 DNA data. One such study found no difference in the rate of decay in children starting ART <3 months or later [24]. Taken together, this suggests that there is a very early window when the pool of HIV-1 infected cells is more labile and when ART may reduce the seeding and persistence of long-surviving reservoir cells. No child in this study received integrase strand transfer inhibitors (INSTI) which might have resulted in faster HIV-1 DNA decay than in the current study [28,29].

The limitations of this study included the following: We had longitudinal samples from only seven children treated shortly after birth, none of whom had PBMC samples collected prior to therapy initiation. In addition, we had access to longitudinal samples of only 31 individuals with suppressed viraemia from the CHER study, a larger sample size may have enabled us to detect a smaller effect of treatment interruption on HIV-1 DNA decay. Moreover, although HIV-1 DNA levels decayed to undetectable levels in a number of children, this does not indicate the absence of persisting replication-competent reservoirs. Children in CHER were reinitiated based on CD4 and clinical criteria. The current norm for studies that involve therapy interruption is to perform regular plasma HIV-1 RNA, immunologic and clinical safety monitoring to decide when to reinitiate therapy, referred to as intensively monitored antiretroviral therapy pause (IMAP). Nevertheless, there was no statistical difference in the decay rates and endpoint HIV-1 DNA between interrupted and uninterrupted children in the participants from CHER, but any affect could have been obscured by a later start in participants who did not interrupt ART.

5 | CONCLUSIONS

Birth diagnosis followed by early treatment initiation before eight days of life was associated with a significantly faster decay of HIV-1-infected cells compared to those tested around four to six weeks of life and then receiving early time-limited ART or delayed continuous ART. Early elective treatment initiation and a sufficiently long duration of initial ART may limit the seeding of long-surviving HIV-1 infected cells, and impact health and quality of life outcomes. Due to their small pools of HIV-1 infected cells and largely intact immune responses, early treated individuals may be valuable candidates for functional cure studies. There is a need for studies of very early treated children that involve intensive monitoring during periods of interruption to investigate safety and the impact of interruption on the HIV-1 reservoir.

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COMPETING INTERESTS

GvZ has received an honorarium for a scientific presentation from VIVl health not related to the current work. JW M is a paid consultant to Gilead Sciences and Merck and owns share options in Co-Crystal Pharma, Inc.

AUTHORS’ CONTRIBUTIONS

GvZ drafted the manuscript and performed analyses with advice from CL. KAV performed the HIV-1 DNA assays and summarized the laboratory results. SI and SN were responsible for the laboratory study database and processing of samples. AWT recorded clinical data and recruited patients. BL and MFC were responsible for clinical oversight. JW M and MFC provided scientific guidance. All authors reviewed and accepted the final manuscript.

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