Noncontiguous finished genome sequence and description of Bacillus testis strain SIT10 sp. nov.

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Abstract

Bacillus testis strain SIT10 (= CSUR P1492 = DSMZ 101190) is the new type strain collected from stool from a 2-year-old boy from Senegal during a culturomics study. This Gram-positive bacterium is a facultative anaerobic rod and a member of the Bacillaceae family. We describe here the features of this bacterium, together with the complete genome sequence and annotation. The 3 987 349 bp long genome (one chromosome but no plasmid) with 42.8% GC content contains 4005 protein-coding and 171 sRNA genes, including 19 5S rRNA gene, 15 16S rRNA genes and ten 23S rRNA genes. © 2016 The Authors. Published by Elsevier Ltd on behalf of European Society of Clinical Microbiology and Infectious Diseases. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

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Introduction

Culturomics was developed in 2012 in order to extend knowledge of the human gut repertoire [1]. A polyphasic approach that combines proteomic by matrix-assisted desorption ionization–time of flight mass spectrometry (MALDI-TOF MS) analysis, genomic data and phenotypic characterization is widely used to describe new bacterial species [2].

Bacillus testis strain SIT10 was isolated from a 2-year-old boy in Senegal as part of a culturomics study aiming to isolate all bacterial species present in the human gut [1]. The genus Bacillus currently comprises 268 species and seven subspecies, although a few of these have been assigned to other genera commonly found in the environment and as laboratory contaminants; however, few species have been linked to infections in humans (http://www.bacterio.net/) [3]. Two Bacillus species are considered medically significant: B. anthracis, which causes anthrax, and B. cereus, which causes a foodborne illness [4]. This genus is one of the largest and most ubiquitous, and it has gained notoriety with taxonomists for its extreme phenotypic diversity and heterogeneity. Bacilli are ubiquitous bacteria that exploit a wide variety of organic and inorganic substrates as nutrient sources [3].

Herein we present a summary of the classification and set of features for Bacillus testis sp. nov. strain SIT10 (= CSUR P1492 = DSMZ 101190) together with the description of the complete genomic sequencing and annotation. These characteristics support the creation of this Bacillus testis species.

Organism Information

Classification and features

A stool sample was collected from a 2-year-old boy living in Senegal. The study was approved by the ethics committee of
the Institut Fédératif de Recherche IFR48, Faculty of Medicine, Marseille, France, under agreement 09-002.

The faecal specimen was preserved at −80°C after collection. Strain SIT10 was isolated on Columbia agar supplemented with 5% sheep’s blood (bioMérieux, Marcy-l’Étoile, France) in aerobic and anaerobic condition using GasPak EZ Anaerobe Container System Sachets (Becton Dickinson (BD), San Diego, CA, USA) at 37°C. Strain SIT10 exhibited a 97.6% 16S rRNA sequence identity with Bacillus massiliogorillae strain G2 (NZ_CAVL000000000.2), the phylogenetically closest bacterial species withstanding in nomenclature (Fig. 1). Its 16S rRNA sequence was deposited in GenBank under accession number LN827531. This value was lower than the 98.7% 16S rRNA gene sequence threshold recommended by Stackebrandt and Ebers [5] to delineate a new species without carrying out DNA-DNA hybridization. The spectrum from SIT10 was added to our MALDI-TOF MS database.

Growth conditions and identification
Growth at different temperatures (25, 37, 45°C) was tested. Growth of the strain was tested in 5% sheep’s blood–enriched Columbia agar (bioMérieux) under anaerobic using GENbag anaer systems (bioMérieux). Growth was achieved only both aerobically and anaerobically. Gram staining and electron microscopy were performed with a TechnaiG2 Cryo (FEI Company, Limel-Brevannes, France) at an operating voltage of 200 keV (Fig. 2). Cells were grown on 5% sheep’s blood agar for 24 hours. A bacterial suspension was prefixed in 5% (v/v) glutaraldehyde in phosphate buffer (Thermo Fisher Scientific Life Sciences, Waltham, MA, USA) for at least 1 hour at room temperature, washed in the same buffer and stained with 1% (w/v) ammonium molybdate 1%. Catalase activity was determined by an ID Color catalase test kit (bioMérieux), and oxidase activity was assayed by applying the cells to moistened discs impregnated with dimethyl-p-phenylenediamine (bio-Mérieux). Biochemical tests were performed with the commercially available API ZYM and API 50 CH strips and were used to characterize the biochemical properties of the strain according to the manufacturer’s instructions. The antibiotic susceptibility was tested using SirScan Discs antibiotics (i2a, Montpellier, France).

Extended Features Descriptions
MALDI-TOF MS [6] protein analysis was carried out as previously described. The SIT10 spectra were imported into the MALDI BioTyper software (version 3.0; Bruker Daltonics, Leipzig, Germany) and analysed by standard pattern matching (with default parameter settings) against 7765 spectra of bacteria, including 231 spectra from Bacillus genus. The method of identification included the m/z from 3000 to 15 000 Da. A maximum of 100 peaks were compared with spectra in the database for every spectrum. The resulting score enabled the identification (or not) of tested species: a score of ≥2 with a validated published species enabled identification at the species level.
level, a score of ≥ 1.7 but <2 enabled identification at the genus level, and a score of <1.7 did not enable any identification. No significant MALDI-TOF MS score was obtained for strain SIT10 against the Bruker database, suggesting that our isolate was not a member of a known species. We added the spectrum from strain SIT10 to our database.

**Genome sequencing and assembly**

Genomic DNA of *Bacillus testis* strain SIT10 was sequenced on the MiSeq Technology (Illumina, San Diego, CA, USA) with the mate pair strategy. The assembly was performed using Soap De Novo with 179% coverage, and 1 910 663 paired reads were filtered according to the read qualities. Its leads to scaffolds and large contigs (>1500 bp) generated a genome size of 3.99 Mb.

**Genome annotation and comparison**

Genome was annotated by Rapid Annotation using Subsystem Technology (RAST) bioserver [7]. The predicted bacterial protein sequences were searched against the GenBank database and the Clusters of Orthologous Groups (COGs) databases using BLASTP (E value 1e-03, coverage 0.7 and identity percentage 30%). The tRNAscanSE tool was used to find tRNA genes [8], whereas ribosomal RNAs were found by using RNAmmer and BLASTn against the GenBank database [9]. The resistome was analysed with ARG-ANNOT (Antibiotic Resistance Gene-ANNOTation) database [10]. The exhaustive search for bacteriocin was performed using the database available in our laboratories (Bacteriocins of the URMITE database BUR; http://drissifatima.wix.com/bacteriocins). Protein sequences from this database allowed putative bacteriocins from human gut microbiota to be identified using BLASTp methodology [11]. Analysis of the presence of polyketide synthase and nonribosomal peptide synthase was performed by discriminating the gene with large size using a database realized in our laboratory, and predicted proteins were compared against the nonredundant (nr) GenBank database using BLASTp.

Phylogenetic relationships with closely related species were determined by MEGA6. The evolutionary history was inferred by using the maximum likelihood method based on the JTT matrix-based model. We compared the genome sequence of *Bacillus testis* strain SIT10 with those of *Bacillus massiliogorillae* strain G2b (NZ_CAVL00000000.2), *Bacillus cereus* ATCC 14579 (NC_004722.1) and *Bacillus psychrosaccharolyticus* ATCC 23296 (NZ_AJTN00000000.2).

**Results**

**Phenotypic properties**

*Bacillus testis* strain SIT10 grew in anaerobic and aerobic conditions on 5% sheep’s blood–Columbia agar at 37°C. Colonies were 0.4 to 0.5 mm in diameter on Columbia agar, and they appeared smooth and grey in color at 37 °C. No growth was observed at 45°C. Gram staining showed Gram-positive bacilli (Fig. 2). A motility test was positive. Strain SIT10 exhibits positive catalase and negative oxidase activity. Acid production was observed for the following carbohydrates with API 50 CH strip (bioMérieux): L-arabinose, D-ribose, D-xylose, D-glucose, D-fructose, D-mannose, N-acetylglucosamine, amygdalin, aesculin, aesculin, arbutin, aesculin, salicin, cellobiose and maltose (Table 1). By using API ZYM, positive reactions were observed for phosphatase alkaline, esterase, leucine aminopeptidase, valine

### Table 1. Differential characteristics of *Bacillus testis* strain SIT10, *B. massiliogorillae*, *B. cereus* ATCC 14579 and *B. psychrosaccharolyticus* ATCC 23296

| Test                  | *B. testis* SIT10 | *B. massiliogorillae* G2 | *B. cereus* ATCC 14579 | *B. psychrosaccharolyticus* ATCC 23296 |
|-----------------------|-------------------|--------------------------|------------------------|----------------------------------------|
| Glycerol              | +/−               | −                        | ND                     | +                                      |
| L-Arabinose           | +                 | +                        | ND                     | +                                      |
| D-Ribose              | +                 | +                        | ND                     | +                                      |
| D-Xylose              | +                 | +                        | ND                     | +                                      |
| D-Galactose           | −                 | −                        | +                      | ND                                     |
| D-Glucose             | +                 | +                        | ND                     | +                                      |
| D-Fructose            | +                 | +                        | ND                     | +                                      |
| D-Mannose             | +                 | +                        | ND                     | +                                      |
| Inositol              | −                 | −                        | −                      | ND                                     |
| D-Mannitol            | −                 | −                        | −                      | −                                      |
| Methyl β-D-glucopyranoside | −     | ND                       | ND                     | ND                                     |
| N-acetylglucosamine  | +                 | +                        | ND                     | +                                      |
| Amygdalin             | +                 | +                        | ND                     | +                                      |
| Aesculin              | +                 | +                        | ND                     | +                                      |
| Salicin               | +                 | +                        | ND                     | +                                      |
| Cellobiose            | +                 | +                        | ND                     | +                                      |
| Lactose               | +                 | +                        | ND                     | +                                      |
| Melibiose             | −                 | −                        | −                      | −                                      |
| Raffinose             | −                 | −                        | −                      | −                                      |
| Gentiose              | +/−               | −                        | ND                     | +                                      |
| Turanose              | −                 | −                        | −                      | −                                      |
| D-Lyxose              | −                 | −                        | −                      | −                                      |

* +, positive result; −, negative result; NA, data not available.*

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TABLE 2. Genome features of Bacillus testis strain SIT10

| Attribute                  | Value       |
|----------------------------|-------------|
| Size (bp)                  | 3,987,349   |
| G+C content (bp)           | 42.8%       |
| mRNAs gene                 | 171         |
| 5S rRNA                    | 19          |
| 16S rRNA                   | 15          |
| 23S rRNA                   | 10          |
| Protein coding gene        | 4005        |
| Genes with unknown function| 264         |
| Genes assigned to COGs     | 2673        |
| CRISPRs                    | 0           |
| Genes associated to PKS    | 0           |
| Genes associated to toxin  | 6           |
| Genes associated to resistome| 3         |

COGs, Clusters of Orthologous Groups database; CRISPR, clustered regularly interspaced short palindromic repeat; G+C, guanine cytosine; NRPS, nonribosomal peptide synthase; PKS, polyketide synthase; rRNA, ribosomal RNA.

Genome properties
The genome size of Bacillus testis strain SIT10 is 3,987,349 bp with a 42.8% G+C content and is assembled into nine scaffolds (28 large contigs). A total of 4005 protein-coding genes are annotated; 171 were mRNAs (19 genes were 5S rRNA, 15 were 16S rRNA, ten were 23S rRNA and 124 were tRNA). A total of 2673 genes were assigned as putative function, while 264 were assigned as unknown function (by COGs or by nr BLAST) (Table 2). The distribution of genes into COGs functional categories is presented in Table 3. The properties and comparisons of the genome are summarized in Table 4. It contains two intact phases 68.2 and 20.5 kb in size with 39% and 35.90% GC content, respectively.

Resistome
The resistome of Bacillus testis strain SIT10 includes (bla) AmpS β-lactamase encoding gene and the major facilitator superfamily (MFS).

Specific features
Analysis of the genome revealed the absence of nonribosomal polyketide synthesis but the presence of two bacteriocin peptides that showed 65% similarity with Clostridium sulfidigenes (Fig. 3) and a pseudouridine synthase that showed 81% similarity with Bacillus massiliogorilae. The genome of B. testis contains the T2SS (type 2 secretion system) operon and approximately 36 genes distributed in five clusters encoding flagellar system disseminated in the genome.

TABLE 3. Number of genes associated with 25 general COGs functional categories

| Code | Value total | Description                                      |
|------|-------------|--------------------------------------------------|
| J    | 168         | Translation                                      |
| A    | 0           | RNA processing and modification                  |
| K    | 216         | Transcription                                    |
| L    | 181         | Replication, recombination and repair            |
| B    | 1           | Chromatin structure and dynamics                 |
| D    | 32          | Cell cycle control, mitosis and meiosis          |
| Y    | 0           | Nuclear structure                                |
| V    | 48          | Defense mechanisms                               |
| T    | 105         | Signal transduction mechanisms                   |
| M    | 103         | Cell wall/membrane biogenesis                    |
| N    | 48          | Cell motility                                    |
| Z    | 0           | Cytoskeleton                                     |
| W    | 0           | Extracellular structures                         |
| U    | 42          | Intracellular trafficking and secretion          |
| O    | 97          | Posttranslational modification, protein turnover, chaperones |
| C    | 159         | Energy production and conversion                 |
| G    | 173         | Carbohydrate transport and metabolism            |
| E    | 304         | Amino acid transport and metabolism              |
| F    | 86          | Nucleotide transport and metabolism              |
| H    | 89          | Coenzyme transport and metabolism                |
| I    | 124         | Lipid transport and metabolism                   |
| P    | 203         | Inorganic ion transport and metabolism           |
| Q    | 71          | Secondary metabolites biosynthesis, transport and catabolism |
| R    | 423         | General function prediction only                 |
| S    | 63          | Function unknown                                 |
|      | 1256        | Not in COGs                                     |

COGs, Clusters of Orthologous Groups database.

Genome comparison
Here we compared the genome of Bacillus testis strain SIT10 with those of Bacillus massiliogorilae G2, Bacillus cereus ATCC 14579 and Bacillus psychrosaccharolyticus ATCC 23296. The draft genome of Bacillus testis strain SIT10 is smaller in size than those of B. massiliogorilae, B. psychrosaccharolyticus and B. cereus (3.9 vs. 5.45 and 4.59 Mb, respectively). The G+C content of Bacillus testis is larger than those of B. massiliogorilae, B. psychrosaccharolyticus and B. cereus (42.7% vs. 34.9%, 38.8% and 35.51%).

Conclusions
On the basis of phenotypic, phylogenetic and genomic analyses, we formally propose the creation of Bacillus testis strain SIT10 sp. nov. The strain was isolated from the stool sample of a 2-year-old boy from Senegal. Several other previously undescribed bacterial species were also cultivated from...
different faecal samples through diversification of culture conditions [1].

**Description of Bacillus testis strain SIT10 sp. nov**

*Bacillus testis* strain SIT10 (= CSUR P1492 = DSMZ 101190) is the type strain of the genus *Bacillus*. It was isolated from a 2-year-old boy living in Senegal as part of a culturomics study aiming to isolate all bacterial species present in the human gut. The main scope of the culturomics study is to cultivate all the species within human faeces. *Bacillus testis* is a motile Gram-positive bacilli that exhibits positive catalase and negative oxidase activities. Colonies were 0.4 to 0.5 mm in diameter. It is a facultative anaerobic bacterium. Using API ZYM and API 50CH, positive reactions were found for phosphatase alkaline, esterase, leucine aminopeptidase, valine aminopeptidase, cystine aminopeptidase, chymotrypsin, phosphatase acid, α-galactosidase, β-galactosidase, α-glucosidase, β-glucosidase L-arabinose, D-ribose, D-xylene, D-glucose, D-fructose, D-mannose, N-acetylglucosamine, amygdalin, arbutin, aesculin, salicin, cellobiose and maltose. Antimicrobial susceptibility testing demonstrated that the cells were resistant to trimethoprim–sulfamethoxazole, cephalosporins (cefazidine, ceftriaxone) and ticarcillin–clavulanic acid. The analysis of genome revealed the absence of nonribosomal polyketide synthesis but the presence of two bacteriocins.

**Genome sequence accession number**

The genome of *Bacillus testis* strain SIT10 has been submitted to the EBI database under bioproject ID PRJEB9400 with GenBank accession number CVQX0000000.1 and 16S RNA accession number LN827531.

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**Conflict of Interest**

None declared.

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