Molecular analysis of the BRCA1 and BRCA2 genes in 32 breast and/or ovarian cancer Spanish families

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Summary It is estimated that about 5–10% of breast cancer cases may be due to inherited predisposition. Until now, two main susceptibility genes have been identified: BRCA1 and BRCA2. The first linkage and mutational studies suggested that mutations in these two genes would account for the majority of high-risk breast cancer families, but recent studies show how the proportion of families due to BRCA1 or BRCA2 mutations strongly depends on the population and the types of family analyzed. It is now clear that, in the context of families with a modest cancer profile, which are the most commonly found in the clinical practice, the percentage of mutations found is much lower than that suggested by the first studies. In the present study, we analyze a group of 32 Spanish families, which contained at least three cases of female breast cancer (at least one of them diagnosed before the age of 50 years), for the presence of mutations in the BRCA genes. The total proportion of mutations was low (25%), although the percentage of mutations in the BRCA1 and BRCA2 genes was higher, considering the breast and ovarian cancer families and the male breast cancer families respectively. Our results are in agreement with the idea that a great proportion of moderate-risk cancer families could be due to low penetrance susceptibility genes distinct from BRCA1 or BRCA2. The study was to establish the proportion of cases attributable to mutations in the BRCA1 or BRCA2 genes in our population and to contribute to the general knowledge of the proportion of mutations found in families with a modest cancer profile, which comprise the majority of the families seeking genetic advice. Although the percentage of mutations was low (25%), striking differences were observed when families were classified attending to the presence of ovarian cancer or male breast cancer. These results are useful to establish a guideline in the development of mutation screening for the BRCA genes.

Keywords: BRCA1; BRCA2; mutation; ovarian cancer; male breast cancer

PATIENTS AND METHODS

Breast/ovarian cancer families

Thirty-two breast and/or ovarian cancer families were selected from women who underwent surgery in Fundación Jiménez Díaz (Madrid) in the last 7 years. These families can be considered representative of the Spanish population as a whole, as they are ascertained in every area of the country. The selection criteria were as follows:

1. Families with three or more cases of women affected with breast cancer and at least one of them diagnosed before the age of 50 (27 cases).
2. Families with three or more cases of women affected with breast cancer and at least one case of male breast cancer diagnosed at any age (five cases).

Mutation detection

We performed a molecular analysis of the complete coding sequence and exon–intron boundaries of the BRCA1 and BRCA2 genes.
genes. For this study we used the PTT (protein truncation test), SSCP (single-strand conformation polymorphism) and direct sequencing methods.

**PTT**

We used this method for the analysis of exon 11 of the **BRCA1** gene and exons 10 and 11 of the **BRCA2** gene. Exons were amplified by polymerase chain reaction (PCR) in overlapping fragments of 1000–2000 bp using primers previously described (Hogervorst et al, 1995; BIC website), which contained an T7 promoter, a eukariotic translation initiation sequence and gene-specific sequences. The PCR conditions were the following: 250 ng of genomic DNA; 10·PCR buffer (Boehringer Mannheim), a mix of 200 mM of dATP, dTTP, dGTP and dCTP; 40 pmol each primer and 2.5 U of Taq polymerase (Boehringer Mannheim). Approximately 1 μg of the PCR product was ‘in vitro’ transcribed and translated using the TnT R T7 Quick Coupled Transcription/Translation System from Promega. The proteins were labelled with biotinilated lysine (Promega) and electrophoresed in a 12.5% polyacrylamide-sodium dodecyl sulphate (SDS) gel. The detection was performed using the ‘Transcend™ Chemiluminescent Non-Radioactive Translation’ from Promega.

**SSCP**

SSCP was used for the analysis of exons 2–10 and 12–24 of the **BRCA1** gene, and 2–10 and 12–27 of the **BRCA2** gene using primers previously described (Simard et al, 1994; Stratton et al, personal communication). Fragments of 200–300 bp were amplified under standard PCR conditions containing 100 ng genomic DNA; 10 × PCR buffer (Boehringer Mannheim), a mix of 200 μM of dATP, dTTP, dGTP and dCTP; 40 pmol each primer and 2.5 U of Taq polymerase (Boehringer Mannheim). Approximately 1 μg of the PCR product was ‘in vitro’ transcribed and translated using the TriT<sup>R</sup> T7 Quick Coupled Transcription/Translation System from Promega. The proteins were labelled with biotinilated lysine (Promega) and electrophoresed in a 12.5% polyacrylamide-sodium dodecyl sulphate (SDS) gel. The detection was performed using the ‘Transcend™ Chemiluminescent Non-Radioactive Translation’ from Promega.

**RESULTS**

**BRCA1 and BRCA2 mutations**

Only three of the 22 site-specific female breast cancer families were found to harbour mutations, one in the **BRCA1** gene and two in the **BRCA2** gene (Table 1). The **BRCA1** mutation identified was a splice-site mutation previously described. The two families with a **BRCA2** mutation showed the same alteration, 936delAAAC in exon 11, which is considered one of the recurrent mutations in the **BRCA2** gene (Neuhausen et al, 1998).

Two of five families with breast and ovarian cancer and no males affected showed mutations in the **BRCA1** gene. One was a frameshift mutation in exon 3, 39delTG, not previously described. The second alteration was an amino acid substitution in exon 18 that was classified as a missense mutation, as it was found in two affected sisters from the same family, but it was not detected in 200 control chromosomes. This alteration would be expected to be significant, since it results in the substitution of a small hydrophobic amino acid by a hydrophilic-charged one, and it is localized in the COOH terminal region of the gene which is...

| Table 1a | BRCA1 mutations |
| --- | --- |
| Family | Female breast cancer age | Ovarian cancer | BRCA1 mutation Exon Codon Effect |
| M36 | 8 | 42.0 | – | 5272–1–G/A Intron 18 – Splicing |
| M72 | 4 (2B)<sup>a</sup> | 38.6 | 3 | 5236GGA/GAA 18 1706 G1706E |
| M73 | 2 | 43.0 | 4 | 236delAG 3 39 Ter39 |

<sup>a</sup>B = bilateral breast cancer.

| Table 1b | BRCA2 mutations |
| --- | --- |
| Family | Female breast cancer age | Male breast cancer | Other cancers<sup>b</sup> | BRCA2 mutation Exon Codon Effect |
| M21 | 4(2B) | 44.5 | – Pan, Hep, Lung, Int | DelAAAC 11 936 Ter956 |
| M22 | 4(3B) | 44.7 | 3 Ov, Leu, Ut, Osteo, St | Del5 23 3009 Ter3015 |
| M24<sup>a</sup> | 1 | 49.0 | 1 Ov (3), NHL, St | DelAA 14 2370 Ter2390 |
| M45 | 11 | 47.7 | 1 | DelAA 25 3104 Ter3109 |
| M59 | 4 | 38.0 | – Prostate | delAAAC 11 936 Ter956 |

<sup>a</sup>Five paternal relatives affected by breast cancer at unknown ages and four relatives affected by other cancers. <sup>b</sup>Pan = pancreatic cancer, Hep = hepatic cancer, Int = intestine cancer, Leu = leukaemia, Ut = uterine cancer, Osteo = osteosarcoma, NHL = non-Hodgkin’s lymphoma, St = stomach cancer, Ov = ovarian cancer.
highly conserved and supposed to be functionally important (Abel et al, 1995).

Three of five families with male breast cancer (60%), revealed frameshift mutations in the \textit{BRCA2} gene (Table 1). All the families with mutations also contained cases of ovarian cancer and in two of them, there were several members affected by other types of cancer.

Surprisingly, in the case of the \textit{BRCA1} gene we did not detect any mutation in exon 11, which, to date, is supposed to accumulate a great proportion of the mutations identified in most of the studies. Our results are in agreement with those derived from other studies in Spanish population (Baiget et al, personal communication; Caldés et al, personal communication). In the case of \textit{BRCA2}, the mutations identified were distributed between exon 11 and exon 27. We found unknown significance variants in the \textit{BRCA2} gene in two families (Table 2): one of them (family 44), contained three cases of female breast cancer and four additional members of the family affected by thyroid, stomach, intestine and liver cancer, the second case (family 68) contained three cases of female breast cancer. We did not have enough data to accept or rule out the implication of these variants in the pathology of the disease in none of the cases. We also found a great number of polymorphisms in both genes (Table 2).

### Table 2a Polymorphisms found in the \textit{BRCA1} gene

| Variant* | Exon | Codon | Amino acid change | Frequency* |
|----------|------|-------|-------------------|------------|
| IVS8-57delT | Intron 8 | – | – | 16/54 |
| 4447T/C | 13 | 1436 | Ser-Ser | 72/180 |
| 4956A/G | 16 | 1613 | Ser-Gly | 6/50 |
| 5272+66G/A | Intron 18 | – | – | 17/54 |

*The number indicates the nucleotide in which the alteration occurs. All the variants have been previously described. *The frequency is obtained by the number of chromosomes which showed the variant/total number of chromosomes analysed in our breast cancer population.

### Table 2b Polymorphisms found in the \textit{BRCA2} gene

| Variant | Exon | Codon | Amino acid change | Frequency | Frequency in control population |
|---------|------|-------|-------------------|-----------|---------------------------------|
| CAG/CAA | 2 | – | – | 3/26 | – |
| 909+56C/T | Intron 8 | – | – | 2/26 | – |
| AAT/CAT | 10 | 289 | Asn-His | 1/32 | 4/112 |
| TCA/TCG | 10 | 470 | Ser-Ser | 1/24 | – |
| AAA/AAGa | 11 | 1132 | Lys-Lys | 2/22 | 19/80 |
| AAA/GAAa | 11 | 1286 | Lys-Glu | 9/26 | 3/116 |
| AGC/AGT | 11 | 1528 | Ser-Ser | 1/26 | – |
| ACG/ATG | 11 | 1915 | Thr-Met | 1/26 | – |
| TCA/TCG | 14 | 2414 | Ser-Ser | 3/24 | – |
| AAT/AGTa | 4 | 108 | Asn/Ser | 1/26 | 1/200 |
| ACC/GCCb | 27 | 3349 | Thr/Ala | 1/26 | 1/200 |

*Polymorphisms not previously described. aUnknown significant variants.

### Table 3 Proportion of mutations in the \textit{BRCA1} and \textit{BRCA2} genes

| Group of families | No. of families | \textit{BRCA1} mutations | \textit{BRCA2} mutations | Total proportion of mutations |
|------------------|----------------|------------------------|------------------------|-----------------------------|
| Breast site-specific cancer families | | | | |
| 3–5 cases | 18 | – | 11% (2) | 11% (2) |
| > 5 cases | 4 | 25% (1) | – | 25% (1) |
| Total | 22 | 4.5% (1) | 9% (2) | 13.5% (3) |
| Breast and ovarian cancer families | 5 | 40% (2) | – | 40% (2) |
| Male breast cancer families | 5 | – | 60% (3) | 60% (3) |
| All families | 32 | 9.3% (3) | 15.6% (5) | 25% (8) |

Classification of families

Although the proportion of mutations was low considering all the families (25%), the percentages were strongly modified when we analysed different groups of families. We categorized the families, according to the number of breast cancer cases, the presence of ovarian cancer and the presence of male breast cancer (Table 3).
The lowest proportion of mutations was found in the site-specific female breast cancer families (13.5%). In the breast and ovarian cancer families the percentage of mutations was 40% and all of them were found in the BRCA1 gene. In the case of male breast cancer families, we found 60% of mutations and all of them were found in the BRCA2 gene.

DISCUSSION

Although there are some BRCA1 and BRCA2 mutational studies in the Spanish population, they are focused on women with sporadic or early onset breast cancer (Garcia-Patino et al, 1998) or they are limited to the analysis of possible recurrent mutations (Osorio et al, 1998; Díez et al, 1999). The present study is the first one in which the whole coding region of the BRCA1 and BRCA2 genes are analysed in a group of selected breast and/or ovarian cancer Spanish families. The proportion of mutations in the BRCA1 or BRCA2 genes found in our 32 families was low (25%). These results are in agreement with several recent studies that show how the percentage of mutations found in the BRCA genes is much lower than the predicted from the linkage studies. Most of the first linkage and mutational analysis of familial breast cancer suggested that mutations in the BRCA1 and BRCA2 genes together would account for most of the high-risk breast and/or ovarian cancer families (Easton et al, 1993). However, these studies were focused on rare families with an extremely large number of affected women. This and other studies evidence how the majority of families attending genetic counselling show a modest cancer profile and that it is important to establish the real implication of the BRCA genes in this group (Ford et al, 1998; Malone et al, 1998).

The lowest percentage of mutations, 13.5%, was found in the breast site-specific cancer families. Recent studies suggest that in this types of family, BRCA1 mutations would account for only 10–20% of the cases and probably the same would be for BRCA2 (Blackwood & Weber, 1998). These studies suggest that most of the families with five or fewer cases of women affected with breast cancer and without any case of ovarian or male breast cancer would be attributable to mutations in other low-penetration susceptibility genes distinct from BRCA1 and BRCA2 (Schurert et al, 1997; Serova et al, 1997; Ford et al, 1998; Malone et al, 1998); the majority of our families contained 3–5 cases of breast cancer, which explains the low proportion of mutations found.

In the group of breast and ovarian cancer families, the percentage of mutations was 40% and all of them were found in the BRCA1 gene. This proportion is identical to that found in the last studies, in which up to 45% of breast and ovarian cancer families turned out to be attributable to mutations in the BRCA1 gene (Couch et al, 1997; Serova et al, 1997; Shattuck-Eidens et al, 1997). We did not find any relationship between the number of breast cancer cases in the family or the existence of bilateral breast cancer, and the presence of mutation, but this was probably due to the small number of families analysed.

Sixty per cent of the families with at least one case of male breast cancer harbour a mutation in the BRCA2 gene. This percentage is high and similar to the found in other studies (Ford et al, 1998), and shows once again the relationship which exists between male breast cancer and mutations in the BRCA2 gene.

We observed a number of tumours other than breast and ovary, in the BRCA2 families. This is in agreement with previous studies that suggest that mutations in BRCA2 confer an increased risk for different types of cancer. A recent work from the Breast Cancer Linkage Consortium shows that pancreatic and prostate cancers are the most frequent in BRCA2 families. In our case, we found two families with one case each of prostate and pancreatic cancer, but all the rest of the tumours observed were of different types.

It is important to note that up to 30% of the mutations can be localized outside the coding region of the genes or consist of great deletions or rearrangements not detectable by conventional techniques. On the other hand, we can find mutations that cause an amino acid substitution whose effect in the protein function is unknown. These variants cannot be considered as mutations but it is not possible to rule out their role in the development of the disease. We found unknown significance in variants in two of our families. Although all these factors can contribute to the low number of mutations detected, they are not supposed to strongly affect the percentages.

In summary, our study confirms once again that the proportion of mutations in the BRCA1 and BRCA2 genes in breast cancer families is low, making the genetic testing and counselling extremely difficult. However, the presence of at least one case of ovarian cancer or male breast cancer is a strong predictor for the presence of mutations in the BRCA1 and BRCA2 genes respectively, which makes these kinds of family specially eligible for the mutational studies.

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