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Antibody response after a booster dose of BNT162B2mRNA and inactivated COVID-19 vaccine

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\textbf{A R T I C L E   I N F O}

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\textbf{A B S T R A C T}

\textbf{Introduction:} The necessity of a booster dose is a matter that has not been as yet illuminated, although it is noted that neutralizing antibody titers decrease over time. We aimed therefore to evaluate antibody titers and seroconversion rates after a booster mRNA vaccine and a booster inactivated vaccine.

\textbf{Methods:} A total of 322 participants were divided into three main groups, with two subgroups each, based on their vaccinations and previous infection history. The levels of anti-SARS-CoV-2 IgG were analyzed with the Elecsys® Anti-SARS-CoV-2 S assay.

\textbf{Results:} The antibody titers showed a linear and significant increase from one vaccine group to the other, displaying progressive changes from group 2IV to group 3IV, and then to group 2IV/mRNA. All of the seronegative participants were in the 2IV(−) subgroup; 93.3% of the participants whose antibody titers were above the upper limit were in the 2IV/mRNA group. Doctors were much more inclined to have a booster dose and mRNA vaccines than nurses. The status of being a doctor increases the rate of having a booster dose 7.8 times; likewise, each annual increase in age increases the rate 1.05 times.

\textbf{Conclusion:} Anti-SARS-CoV-2 IgG levels decrease over time. The antibody response rate to only two doses of the inactivated vaccine was meager, so a booster dose is necessary to maintain the effectiveness of inactivated vaccines. The third dose of the vaccine, especially that of the mRNA vaccine, which was found to be much more superior to the inactivated vaccine, should be strongly recommended.

\section{1. Introduction}

Many vaccine and drug development studies were initiated after the Coronavirus 2019 (COVID-19) pandemic caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) broke out in December 2019. While an effective antiviral agent has not been developed yet, inactivated vaccines, messenger RNA (mRNA) vaccines, and adenovirus vaccines were developed not long after the identification of SARS-CoV-2, and emergency use approvals were obtained for these vaccines [1–3]. Vaccination campaigns were launched rapidly and widely around the world. Shortly after the inactivated vaccine (CoronaVac) was given emergency use approval by the Republic of Turkey Ministry of Health, vaccinations were first given to health care workers (HCW), beginning in January 2021. The HCWs received two doses of the CoronaVac vaccine with a recommended dosing interval of 28 days between the first and second doses. In addition to the inactivated vaccine, the BNT162b2mRNA vaccine (Pfizer and BioNTech) began to be implemented in Turkey at the beginning of April 2021. The Turkish Ministry of Health recommended administering the third booster dose with an optional inactivated or mRNA vaccine as of the beginning of July 2021 for healthcare workers (HCWs) and the elderly.

CoronaVac is an inactivated and aluminum-adjuvant vaccine developed by Sinovac Life Science Company, Beijing, China [4]. The BNT162b2mRNA vaccine produced by Pfizer and BioNTech was the first vaccine approved for emergency use by the United States Food and Drug Administration [5].

In this study, we aimed to evaluate antibody titers and seroconversion rates after a booster mRNA vaccine and a booster inactivated vaccine, administered after two doses of inactivated vaccines, by examining the anti-SARS-CoV-2 IgG response in HCWs with and without a previ-
ous history of SARS-CoV-2 infection, and to determine the inclination toward vaccination of nurses and doctors.

2. Materials and methods

2.1. Study design and population

The study group consists of 322 participants (232 (72%) women, 90 (28%) men, 116 (36%) doctors, 206 (64%) nurses) who worked at the Children’s Hospital of Ankara City Hospital. The mean age was 31.4 ± 8.01 years old, and the median age was 28 years old. Our study includes three groups: those who had only two doses of inactivated vaccine [2IV group, 83 participants (25.8%)], those who had two doses of inactivated vaccine and a booster dose of inactivated vaccine [3IV group, 23 participants (7.1%)], and those who had two doses of inactivated vaccine and a booster dose of mRNA vaccine [2IV/mRNA group, 216 participants (67.1%)]. Among the participants, 93 (28.9%) had a previous history of COVID-19 infection (diagnosed by nasopharyngeal swab polymerase chain reaction (PCR) result), 229 (71.1%) did not. Participants in each group who had a previous history of COVID-19 infection were indicated as (+), and those who did not were indicated as (-). The study did not include those with immunodeficiency, those receiving immunosuppressive therapy, and those with a history of multiple COVID-19 infections. In order to determine the anti-SARS-CoV-2 immunoglobulin G (IgG) antibody levels, blood samples from 322 HCWs who were volunteers were collected. The duration between the last vaccination and sampling was 6 to 8 months for those who had only two doses of inactivated vaccine, and 2 to 4 months for those who had inactivated or mRNA booster doses.

2.2. Detection of anti-SARS-CoV-2 antibodies

The samples were analyzed at the Ankara Microbiology Reference Laboratories of the Public Health General Directorate of Turkey. Analyses were performed in a macroELISA device (Roche Cobas 8000-e801/analyser) with an electrochemiluminescent (ECLIA) technique using a commercial kit (Elecsys Anti-SARS-CoV-2 S, Roche). The Elecsys Anti-SARS-CoV-2 S assay utilizes a recombinant protein representing the RBD of the virus’s S antigen in a double antigen sandwich test format which favors the quantitative determination of high-affinity antibodies to SARS-CoV-2. The tests were run and evaluated according to the manufacturer’s instructions. Values below the measurement limit resulted in < 0.40 U/mL. Values above the measuring range resulted in >250 U/mL, and these samples were reanalysed at 1:50 automatic dilution. Values above the measurement limit resulted in >12,500 U/mL. A cut-off index (COI) < 0.80 U/mL was considered negative for anti-SARS-CoV-2 antibodies, while a COI ≥ 0.80 U/mL was considered positive. In order to correlate the units (U) of the Elecsys® Anti-SARS-CoV-2 S assay with the “binding antibody units” (BAU) of the first WHO International Standard for anti-SARS-CoV-2 immunoglobulin, a 0.972 coefficient was used in line with the manufacturer’s recommendations [6]. A mathematical transposition of Roche specific U to BAU of the WHO International Standard followed the equation: Roche $U = 0.972 \times$ BAU.

2.3. Statistical analysis

The Statistical Package for the Social Sciences (SPSS) version 23.0 (IBM Corp., Armonk, New York, USA) was used to analyze the data. Descriptive statistics (including frequencies and means) for all variables were calculated. The results were expressed as mean ± standard deviation, median, range (minimum-maximum), and number (%). The difference between groups in categorical variables was analyzed using the chi-square test. The Kolmogorov Smirnov test was used to examine whether the numerical variables showed normal distribution. The Mann-Whitney U test investigated differences between the two independent groups in terms of binary variables. The Kruskal-Wallis test was used for comparisons of more than one independent group. The level of statistical significance was established as p < 0.05. A logistic regression model was created to reveal the factors affecting having a booster dose and to estimate the odds ratio (OR) and confidence interval (CI).

The data that support the findings of this study are available on request from the corresponding author. The data are not publicly available due to privacy or ethical restrictions.

This study was conducted in conformity with the principles of the Declaration of Helsinki and approved by the Republic of Turkey Ministry of Health, the Ethics Committee of Ankara City Hospital Ethics Committee, and the Institutional Review Board of the Children’s Hospital of Ankara City Hospital.

3. Results

Descriptive features of the participants were demonstrated in Table 1. While 239 (74.2%) of the participants had a booster dose with an inactivated or mRNA vaccine, 83 (25.8%) did not. No relationship was found between gender or previous history of COVID-19 infection and having a booster dose (p > 0.05 each). A significant relationship was found between occupation and having a booster dose; the rate of having a booster dose was 93% for doctors and 63% for nurses (p < 0.001) (Table 2). When vaccine preferences were examined according to professions, 90.5% of the doctors preferred an mRNA booster, 2.6% preferred an inactive vaccine booster, while among nurses, 53.9% preferred an mRNA booster, and 9.7% preferred an inactive vaccine booster (p < 0.001). Additionally, there was a significant relationship between age and having a booster dose; the older the age of the HCWs, the higher was the rate of having a booster dose (p = 0.002). In the present study, the multivariate logistic regression analysis was used to examine associations, and it was revealed that status as a doctor increases the rate of having a booster dose 7.8 times (OR = 7.81, 95% CI 3.59–16.95, p < 0.001), likewise, each annual increase in age increases the rate 1.05 times (OR = 1.05, 95% CI 1.009–1.085, p = 0.014) (Table 2).

No relationship was found between age, gender, or previous history of COVID-19 infection and antibody levels (p > 0.05 each). When the antibody levels of the HCWs were examined, 3.41% (n = 11) of the samples were seronegative (< 20 BAU), and 36.95% (n = 119) were above the upper limit of 12,860 BAU that the test could detect. All of the seronegative participants were in the 2IV- subgroup; 93.3% of the participants whose antibody titers were above the upper limit were in the 2IV/mRNA group.

When the antibody levels of the participants who had and did not have a booster dose were compared, it was seen that the antibody levels of those who had a booster dose were much higher than the others (p < 0.001) (Table 2). Anti-SARS-CoV-2 IgG levels based on vaccines showed

| Variables                      | n = 322 |
|--------------------------------|---------|
| Age (year) (mean±SD)           | 31.4 ± 8.01 (median: 28) |
| Gender                         |         |
| Female                         | 232 (72%) |
| Male                           | 90 (28%)  |
| Occupation                     |         |
| Pediatricist                    | 116 (36%) |
| Pediatric nurse                | 206 (64%) |
| Previous history of COVID-19 infection | 93 (28.9%) |
| Yes                            | 229 (71.1%) |
| No                             |         |
| Vaccine preferences            |         |
| 2IV                            | 83 (25.8%) |
| 3IV                            | 23 (7.1%)  |
| 2IV/mRNA                       | 216 (67.1%) |
Table 2
Descriptive features of the participants based on vaccination preferences.

| Variables                     | Booster (-) (n = 83) | Booster (+) (n = 239) | p value  | Adjusted OR (95% CI) for having a booster dose |
|-------------------------------|----------------------|-----------------------|----------|-----------------------------------------------|
| Age (year)                    | 29 ± 6.88 (median: 26) | 32.22 ± 8.22 (median: 29) | p = 0.002 | 1.05 (1.009–1.085) (p = 0.014) |
| Gender                        |                       |                       |          |                                               |
| Female                        | 62 (25.3%)            | 69 (28.9%)            | p > 0.05 | 0.91 (0.48–1.65) (p = 0.72) |
| Male                          | 21 (74.7%)            | 170 (71.1%)           |          |                                               |
| Occupation                    |                       |                       |          |                                               |
| Pediatrician                  | 8 (9.6%)              | 108 (45.2%)           | p < 0.001 | 7.81 (3.59–16.95) (p < 0.001) |
| Pediatric nurse               | 75 (90.4%)            | 131 (54.8%)           |          |                                               |
| Previous history of COVID-19 infection |                   |                       |          |                                               |
| Yes                           | 29 (34.9%)            | 64 (26.8%)            | p > 0.05 | 0.69 (0.39–1.23) (p = 0.21) |
| No                            | 54 (65.1%)            | 175 (73.2%)           |          |                                               |
| Anti-SARS-CoV-2 IgG (BAU)     | 2000.43 ± 4050.51     | 1085.02 ± 4293.32     | p < 0.001 | 0.71 (0.43–1.17) (p = 0.19) |
| (mean±SD,median, IQR)         | (57.61–777.77)        | (715.34–12,860)       |          |                                               |

OR: Odds ratio, C.I.: Confidence interval.

Table 3
Anti-SARS-CoV-2 IgG levels based on vaccines, doses and COVID-19 infection history.

| Groups                  | Number (%) | Median (IQR) | Anti-SARS-CoV-2 IgG levels (BAU) | p     | Seronegativity rate,% (n) | Rate of participants with a maximum titer,% (n) |
|-------------------------|------------|--------------|----------------------------------|-------|---------------------------|-----------------------------------------------|
| 2IV(-)                  | 54 (16.8%) | 88.27 (25.64–247.17) | p < 0.001 | 20.37% (n = 11) | 7.4%, (n = 4) |
| 2IV(+)                  | 29 (9.0%)  | 626.54 (238.68–4782.92) | 0%     | 10.34%, (n = 3) |                                   |
| 3IV(-)                  | 13 (4.0%)  | 1361.11 (441.36–5128.09) | p > 0.05 | 0% | 0% |
| 3IV(+)                  | 10 (3.1%)  | 1409.47 (969.39–4060.70) | 0%     | 10%, (n = 1) |                                   |
| 2IV/mRNA(-)             | 162 (50.3%) | 12,860 (9437.76–12,860) | p > 0.05 | 0% | 53.9%, (n = 86) |
| 2IV/mRNA(+)             | 54 (16.8%) | 11,871.91 (5912.55–12,860) | 0%     | 46.3%, (n = 25) |                                   |

statistically significant differences between groups. Antibody titers were the highest in the 2IV/mRNA group (p < 0.001). The antibody levels of the 3IV group were lower than the 2IV/mRNA group but higher than the 2IV group (p < 0.001). The lowest antibody levels were observed in the 2IV group.

The antibody levels of the subgroups were compared with each other based on their past COVID-19 infection history. Measured anti-SARS-CoV-2 IgG levels for the 2IV(-) and 2IV(+) subgroups showed statistically significant differences, it was higher in the 2IV+ subgroup (p < 0.001). But the 3IV(-) and 3IV(+) subgroups and the 2IV/mRNA(-) and 2IV/mRNA(+) subgroups did not showed difference (p > 0.05 each) (Table 3 and Fig. 1). Although there was no significant difference in antibody levels between the 2IV/mRNA(-) and 2IV/mRNA(+) subgroups, the antibody levels of each of these two subgroups were significantly much higher than all other subgroups (p < 0.001 each). While the antibody levels of both the 3IV(+) and 3IV(-) subgroups were significantly higher than the 2IV(-) subgroup (p < 0.001 each), surprisingly, no difference was found between the 2IV(+) subgroup and the 3IV(-) subgroup (p > 0.05), or likewise between the 2IV(+) and the 3IV(+) subgroups (p > 0.05).

4. Discussion

In this study, the seroconversion rates and anti-SARS-CoV-2 IgG antibody levels of HCWs who had only two doses of inactivated vaccine, two doses of inactivated vaccine with a booster dose of inactivated vaccine, and two doses of inactivated vaccine with a booster dose of mRNA vaccine were evaluated. To the best of the authors’ knowledge, this is one of the first studies to show the difference between the antibody levels induced by a single inactivated vaccine booster dose and a single mRNA vaccine booster dose, each after two doses of inactivated COVID-19 vaccine.

In several studies conducted in the prevaccination period, it was found that nurses were less inclined to get vaccinated against COVID-19 than physicians, meaning that vaccine reluctance was higher in nurses [7–10]. In addition to the low vaccine acceptance rate noted among nurses for COVID-19 vaccines, a low vaccine acceptance rate for seasonal influenza vaccine was also displayed by this group as compared to physicians in the literature [11,12]. In terms of our study, nurses were less likely to accept a third booster dose against COVID-19 than physicians, which is consistent with previous vaccine hesitancy findings. Furthermore, nurses showed a remarkable reluctance compared to doctors with regard to receiving mRNA vaccines. Having more confidence in vaccines, having a higher degree of medical training, a higher level of risk perception, and fear of COVID-19 may contribute to an increased uptake of vaccines among doctors [13].

In previous studies, the antibody response induced by COVID-19 infection or vaccination was found to be sufficient [3,14–16]. In later studies, however, it was observed that antibody titers decreased in patient follow-ups [17,18]. In a study with an extended period of time,
it was shown that antibody levels increased at a peak after 30 days of BNT162b2 vaccination and, subsequently, reduced until they reached a plateau at 60 days after the second dose, with similar values observed after 90 days [19]. It was also shown that mRNA vaccines elicit rapid immune responses in seropositive individuals with post-vaccine antibody titres that are comparable to or exceed titres in naïve individuals vaccinated with two doses [19,20]. In recent publications, therefore, a booster dose has been recommended for individuals who have been vaccinated with two doses of inactivated vaccine and for special groups such as individuals with a kidney transplant or solid organ transplant who had received the mRNA vaccine but have low seroconversion rates [18,21–23]. The antibody response in our study was found at the lowest titer in the group that received two doses of inactivated vaccine, which supports this recommendation. Similarly, all seronegative participants in our study were in the two-doses inactivated vaccine group. No seronegative participants were found in the mRNA or inactivated booster groups.

Kesin et al. found in their study that an inactivated vaccine booster dose after two doses of inactivated vaccine yielded 1.7 times an increase in median values of anti-spike antibody titers; BNT162b2 administration as the third vaccine dose boosted anti-spike antibody median titers by a factor of 46.6 [24]. In a study of the Ministry of Health of Chile led by Araos et al., 4.7 million participants initially immunized with CoronaVac were analyzed, of whom nearly two million had a booster dose. In the case of CoronaVac, effectiveness increased from 56% to 80.2%, while Pfizer-BioNTech rose from 56% to 90%, and AstraZeneca climbed from 56% to 93%. Regarding the prevention of hospitalization, the effectiveness of the CoronaVac booster vaccine increased from 84% to 88%, Pfizer-BioNTech from 84% to 87%, and AstraZeneca from 84% to 96.3% [25]. Similar to these studies, in our study, it was found that the mRNA booster dose induced the highest antibody level. In addition, most of the participants (93.3%) with the highest antibody levels were in the mRNA booster group. Interestingly, it was also found in the current study that there was no difference between the antibody levels of patients with a previous history of COVID-19 infection who had not had a booster dose, and the antibody levels of patients with or without a previous history of COVID-19 infection who had received a third inactivated vaccine.

The present study had several limitations. First, this was a single-center study. Secondly, the prevaccination levels of SARS-CoV-2 antibodies in the participants were unknown. Moreover, since it was a cross-sectional study, the time between vaccination and sampling could not be standardized. Additionally, the antibody level that predicts protection is unknown.

In conclusion, Anti-SARS-CoV-2 IgG levels decrease over time. The antibody response rate to only two doses of the inactivated vaccine was meager, so the results suggest that a booster dose is necessary to maintain the effectiveness of inactivated vaccines. The third dose of the vaccine, especially that of the mRNA vaccine, which was found to be much more superior to the inactivated vaccine, should be strongly recommended. We consider that doing so will ensure the continuity of herd immunity until multicenter and prospective clinical efficacy studies are conducted.

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Conflicts of interest

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CRediT authorship contribution statement

**Metin Yigit:** Conceptualization, Methodology, Formal analysis, Writing – original draft, Writing – review & editing, Visualization, Supervision. **Aslinur Ozkaya-Parlakay:** Conceptualization, Methodology, Supervision. **Yasemin Cosgun:** Conceptualization, Methodology, Resources, Data curation. **Yunus Emre Ince:** Writing – original draft, Investigation, Visualization. **Furkan Kalayci:** Investigation, Resources, Data curation. **Naci Yılmaz:** Investigation, Resources, Data curation.

**Emrah Senel:** Conceptualization, Writing – review & editing, Supervision.

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