CHITOSAN ISOLATION AND ITS APPLICATION TO REDUCE THE CONTENT OF METAL IONS IN WELLBORE WATER

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ABSTRACT

Potential water sources such as white shrimp shell waste (Penaeus merguiensis) can be used as a source of chitosan. Chitosan can be applied as an environmentally friendly adsorbent for water treatment because of its ability to adsorb metal ions. In this study, chitosan was isolated through several stages such as denitrogenization, deproteinization, decolourization and deacetylation. The yield of chitosan obtained from this study was 17.73%. Characterization by infrared spectroscopy (FTIR) showed the absorption at 3355 cm\(^{-1}\) indicating the presence of amine (-NH\(_2\)) and hydroxyl (-OH) groups. The absorption of the carbonyl group (-C=O) at 1642 cm\(^{-1}\) disappeared while the absorption of the free amine group (-NH\(_2\)) at 1590 cm\(^{-1}\) increased indicating the successful deacetylation with a degree of deacetylation (DD) 78%. Application of chitosan in wellbore water did not affect on colour change and decreasing of iron (Fe) content due to low concentration of iron (Fe). However, chitosan can reduce the pH value of water and manganese (Mn) content. The results of ANOVA and DMRT test at 0.05 significance level showed that chitosan with various mass had different effects. The more the mass of chitosan added, the higher the content of manganese (Mn) will decrease.

Keywords: Chitosan; Iron Ion; Manganese Ion; Wellbore Waters

Introduction

Kota Langsa is a city located on the east coast of Aceh and has great potential in the fields of tourism, plantation and marine. Kuala Langsa is one of the areas in Kota Langsa which is bordered by the sea so that can be a producer of various marine resources such as fish, oysters, crabs and clams.

Consumption of shrimp by the community will certainly produce waste in the form of shrimp shells. Shrimp shells waste produced from households can be used as a source of chitosan. Generally, chitosan comes from animals that have outer and hard skin such as crabs, shrimp, insects, oyster shells and other marine animals. Therefore, coastal and marine resources such as white shrimp shells (Penaeus merguiensis) can be used as a potential source of chitosan because they’re composed of chitin (20-25%). There are about 40% of shrimp weight becomes waste which mainly consists of chitin (20-25%), pigments, calcium carbonate (30%), protein and lipid. Chitin itself can be transformed into chitosan which has many advantages. Chitosan can function as a membrane and an adsorbent material so that chitosan can be used as a natural adsorbent for water treatment that is environmentally friendly. During this time, synthetic coagulant material has been applied more in the water treatment process.

Kota Langsa also has problems in the provision of clean water. So far, the community has used water supply from Lengkong (Langsa Baro District, Kota Langsa) and PDAM as a source of clean water. However, the community still has complaints related to PDAM water which is still turbid and yellowish. In addition, some
other residents use wellbore as a source of water for bathing and washing. Water from the wellbore is generally yellowish and turbid so it must be filtered first. The yellowish and turbid water can be caused by the presence of metals such as iron (Fe) and manganese (Mn). The presence of iron (Fe) and manganese (Mn) content in water causes the colour of the water to turn brownish yellow after some time of contact with air.5

In this research, chitosan was isolated from white shrimp shells (Penaeus merguiensis) and then applied as an adsorbent to reduce the content of iron (Fe) and manganese (Mn) in the wellbore water in Paya Bujok Seuleumak, Kota Langsa. Chitosan can be isolated by chemical6 and biological method (using bacteria and enzyme).7,8 Chemical method involves chemical treatment in room temperature1 or high temperature (using conventional heater9 or microwave irradiation).10,11 Previous studies required a longer time1,9 of isolation and produced a low yield9 as well as a low degree of deacetylation of chitosan product.1 In this study, chitosan was isolated by a modification of reported method9 to make isolation process going faster and give higher in yield and degree of deacetylation. This research is expected to overcome problems related to household waste in the form of shrimp shells and water problems with iron (Fe) and manganese (Mn) content in Kota Langsa.

Methods

Isolation and Characterization of Chitosan

White shrimp shell waste (Penaeus merguiensis) was collected from households. Then washed with water and cleaned. The dried shrimp shell waste was crushed with a blender. Then it was weighed as much as 101.16 grams and put into a 1 L beaker. For the deproteinization process, into shrimp shell powder the solution of 5% NaOH has added as much as 750 mL and heated at 75 °C for 1 hour. After that, the shrimp shell powder was washed with water until the pH becomes neutral. The process continued to the demineralization stage by adding 350 mL HCl 10% (v/v) to shrimp shell powder and heated at 70 °C for 1 until no more gas bubbles were generated. The shrimp shell powder then was washed and confirmed to be neutral pH with universal litmus. The decolourization process was carried out by adding 0.4% (v/v) NaOCl as much as 400 mL and soaking for 1 hour. After decolourization was complete, the process continued to the deacetylation stage by adding 400 mL of 50% NaOH (w/v) and heated at 120 °C for 2.5 hours. Chitosan products were washed until neutral and dried. Then it was tested by dissolving the product in 5% acetic acid (CH₃COOH).

Chitosan was characterized by Fourier Transform Infrared (FTIR) spectrophotometer. The yield of each process and the degree of deacetylation (DD) were calculated using the equation (1) and (2) as follow:12,13

\[
\text{Yield} = \frac{\text{mass obtained}}{\text{Initial mass}} \times 100\% \quad (1)
\]

\[
\text{DD} = \left[1 - \left\{ \frac{A_1655}{A_3450} \times \frac{1}{1.33} \right\} \right] \times 100\% \quad (2)
\]

where A 1655 is the absorbance of the carbonyl group (-C=O) at a wavelength of about 1655 cm⁻¹. A 3450 is the absorbance of hydroxyl (-OH) and amine groups (-NH₂) at a wavelength of about 3450 cm⁻¹. Absorbance was calculated from transmittance (%T) value.

Chitosan Application in Wellbore Water

Wellbore water samples were taken from Gampong Paya Bujok Seuleumak, Kota Langsa. Each sample of 100 mL wellbore water were transferred into four 500 mL beakers. Then chitosan was added respectively with a mass of 0 gram, 0.4 gram, 0.8 gram, and 1.2 grams. The mixture was stirred for 25 minutes then left at room temperature for 35 minutes so that the deposition process was complete. After that, supernatant water and precipitate were separated. Supernatant water was taken and each was put into 4 plastic bottles. The pH value of each water was measured by a pH meter. The concentration of iron (Fe) and manganese (Mn) from each water sample were measured using an atomic absorption
spectrophotometer (AAS). Measurements of iron (Fe) and manganese (Mn) from each water sample were carried out 3 times of repetition so that there were 24 measurements. Data of iron (Fe) and manganese (Mn) after treatment with chitosan with several variations of concentration were analyzed using ANOVA at a significance level of 0.05. Duncan's Multiple Range Test (DMRT) was performed to test the real difference.

**Result and Discussion**

**Isolation and Characterization of Chitosan**

One source of chitosan can be obtained from white shrimp shells (Penaeus merguiensis) taken from household waste. Chitosan isolation consists of several stages, namely deproteination, demineralization and deacetylation. Deproteinization process was done by adding 5% NaOH solution which aims to remove protein. The protein contained in the shells of white shrimp (Penaeus merguiensis) will dissolve and undergo hydrolysis in the presence of a strong base solution (NaOH). In the process of hydrolysis, the peptide bonds in proteins will experience a breakdown to produce amino acids. After the deproteination process, the shrimp shell powder was washed to a neutral pH for removing the remaining NaOH solution.

The process continued to demineralization for removing various minerals contained in shrimp shells. In this process, the solution HCl was added to the shrimp shell powder resulting in an acidic reaction with minerals and produces gas bubbles in accordance with the following reactions:

\[
\text{Ca}_3(\text{PO}_4)_2 \text{(s)} + 6 \text{HCl (aq)} \rightarrow 3 \text{CaCl}_2 \text{(aq)} + 2 \text{H}_3\text{PO}_4 \text{(aq)} \quad (1)
\]

\[
\text{CaCO}_3 \text{(s)} + 2 \text{HCl (aq)} \rightarrow \text{CaCl}_2 \text{(aq)} + \text{CO}_2 \text{(g)} + \text{H}_2\text{O} \text{(aq)} \quad (2)
\]

In accordance with the reaction above the gas bubbles produced are carbon dioxide (CO₂). The addition of HCl was stopped when the mineral salts have finished reacting which was indicated by gas bubbles that were no longer produced. The product after this process (demineralization) was chitin. The chitin obtained was 44.45 grams and its colour was brown so it must be continued by the decolourization process using NaOCl solution. The results of decolourization obtained white chitin which was insoluble in either 10% HCl or 5% acetic acid. This chitin was deacetylated into chitosan and obtained 17.94 grams of dry white chitosan powder. The yield of chitosan was 17.73% and dissolved well in a 5% acetic acid (CH₃COOH) solution. This showed that the transformation of chitin to chitosan was successful. The solubility test to determine the formation of chitosan has been done by adding 5% CH₃COOH. Chitosan can dissolve well in 5% acetic acid.

Chitosan was characterized by infrared spectroscopy (FTIR) to determine the value of the degree of deacetylation (DD) and the chemical groups that were characteristic of chitosan. The chitosan structure and FTIR spectrum can be seen in the following Figure.
Based on the above spectrum, there is absorption at the wavenumber 3355 cm\(^{-1}\) with high intensity, which shows the overlapping vibration absorption of the -OH and -NH groups. The peak absorption vibration stretching -\(\text{C} = \text{O}\) appears at wavenumber 2878 cm\(^{-1}\) while the bending vibration of -\(\text{CH}\) appears at wave number 1355 cm\(^{-1}\). The absorption with high intensity at wave number 1001 cm\(^{-1}\) indicated the existence of stretching vibrations of C-O. The peak of the carbonyl group (\(-\text{C} = \text{O}\)) at 1643 cm\(^{-1}\) did not occur because of its small absorption. However, the bending vibration absorption intensity of the free amine group (\(-\text{NH}_2\)) at wavenumber 1590 cm\(^{-1}\) increased which showed a high degree of chitosan deacetylation (DDA).\(^{13}\) This shows that the deacetylation process was successful. When deacetylation occurs, the absorption intensity of the carbonyl group (\(-\text{C} = \text{O}\)) at 1643 cm\(^{-1}\) will decrease while the absorption intensity of the free amine group (\(-\text{NH}_2\)) at wavenumber 1590 cm\(^{-1}\) will increase.\(^{15}\)

The degree of deacetylation (DD) of isolated chitosan was 78%. The value of the degree of deacetylation showed that the transformation of shrimp shells into chitosan was successful with the number of acetyl groups (-\(\text{COCH}_3\)) lost reaching 78%.

**Table 1. Isolation Process of Chitosan**

| Process            | Yield (%) | Colour |
|--------------------|-----------|--------|
| Deproteination     | 76.52     | Brown  |
| Demineralization   | 57.18     | Brown  |
| Decolourization    | 93.88     | White  |
| Deacetylation      | 42.99     | White  |

**Chitosan Application in Wellbore Water**

One function of chitosan is as an adsorbent that can absorb metal ions. Isolated chitosan powder was applied to water taken from the wellbore in Gampong Paya Bujok Seuleumak, Kota Langsa. The results of water treatment with chitosan showed no difference in watercolour before and after treatment. This can be caused by low content of iron (Fe) and other metal ions in the water so that it did not really affect on colour change. Besides the use of chitosan in the form of powder which has a small surface area so that the adsorption of metal ions in water is not optimal. However, the pH value of the water changes after treatment with chitosan as shown in Figure 2.
From Figure 3 it can be seen that the addition of chitosan can reduce the pH of water. The pH value of water will decrease with increasing concentration of chitosan added. This is caused by chitosan acidity and the existence of cationic polyelectrolytes of chitosan. Suptijah also argued that chitosan is polycationic. Measurement of iron (Fe) content in water from treatments with various variations of the mass of chitosan was also carried out. The measurement results can be seen in Table 2.

**Table 2. The Effect of Chitosan on Content of Iron (Fe) Ion**

| Mass of Chitosan (gram) | Iron (ppm) |
|------------------------|------------|
| 0                      | <0,009     |
| 0.4                    | <0,009     |
| 0.8                    | <0,009     |
| 1.2                    | <0,009     |

The concentration of iron (Fe) in water without treatment with chitosan was so low that the addition of chitosan in the water had no effect. This means that the adsorption process was not optimum. In addition, adsorption of iron (Fe) ion depends on pH which iron (Fe) ion adsorption increased at pH 2.8-4.0. When pH>4 the iron (Fe) ion will more be attracted to hydroxide ion (OH-) compared to amine (-NH$_2$) group of chitosan. In this study, adsorption iron (Fe) ion was carried out at pH water 7.50 that the adsorption condition was not optimum. Different results were shown in the measurement of manganese ion content (Mn) in water as in Table 3.

**Table 3. The Effect of Chitosan on Content of Manganese (Mn) Ion**

| Mass of Chitosan (gram) | Mn (ppm) |
|------------------------|----------|
| 0                      | 0,0312$^a$ |
| 0.4                    | 0,0301$^b$ |
| 0.8                    | 0,0156$^c$ |
| 1.2                    | 0,0034$^d$ |

Note: the numbers in the column followed by the same letter indicate no significant difference based on the smallest significant difference test at a significant level of 0.05.

Based on the results of the analysis of variance (ANOVA), it was known that there was an effect of chitosan with variations in mass on decreasing the content of manganese (Mn) in wellbore water. From Table 3 it can be seen that each treatment had a real difference from the other treatments. This means that the higher the mass of chitosan added, the higher the concentration of manganese ions (Mn) in water will decrease. Decreased content of manganese (Mn) in water due to the adsorption of manganese (Mn) ions by adding chitosan powder. Chitosan itself has a structure that contains active sites such as amine groups (-NH$_2$) and hydroxyl groups (-OH) that allow the binding of metal ions such as Mn$^{2+}$ with these groups.

**Conclusion**

Chitosan was successfully isolated from white shrimp shells with a degree of
deacetylation (DD) of 78%. Chitosan had no effect on iron (Fe) content because of its low concentration in wellbore water. Chitosan functions as an adsorbent because it can reduce the concentration of manganese (Mn) ion in wellbore water. The effective mass of chitosan in this study to reduce manganese concentration (Mn) ion in wellbore water was 1.2 grams.

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