Yield Stability Analysis of Maize (*Zea mays* L.) Hybrids Using Parametric and AMMI Methods

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The present study investigated the stability and adaptability of maize (*Zea mays* L.) hybrids. In this study, 12 maize hybrids were planted and examined considering the grain yield. The experiment was arranged in a randomized complete block design (RCBD) with three replications in four research stations in Iran during two crop years. The combined analysis of variance showed that genotype-environment interactions were significant at one percent probability level. The grain yield can stabilize, and hybrids with specific adaptability are recommended to each environment. Hybrids with specific adaptability can be recommended to all types of the environment. Means comparison yield of the genotypes identified DC370 as a high-yield genotype. Regarding AMMI analysis, genotype × environment interactions (GEIs) and two first components were found significant. The SC647 genotype was identified as the most stable genotype. Regarding the stability parameters, SC647 and KSC705 genotypes were selected as the most stable genotypes. From AMMI1 and AMMI2 graphs, the SC647 genotype was identified as the most stable genotype compared with other hybrids.

1. Introduction

Maize is one of the various crucial annual cereal crops, accounting for more than 188 Mha of croplands worldwide and contributing nearly 50% (1.171 Mt) of global food production [1]. The production and trade of maize (*Zea mays* L.) targeted at animal husbandry. However, maize has been a leading and integral staple food for humans. Considering the importance of maize and its cultivation potential worldwide and in Iran, this crop’s yield sustainability was studied in previous research. Genotype × environment interaction (GEI) is responsible for genotype sustainability characteristics [2–4]. One of the usual exacting effects in plant breeding development is to dissect the genotype × environment (GE) interaction correctly. In most trials, the $G \times E$ interaction was witnessed and then modeled statistically and elucidated. Genotype × environment interaction adjusts the reasonable grain yield of genotypes in diverse environments and makes it hard to select the better ones [5]. Multilocation and multiyear trials can help identify superior and sustainable genotypes [6]. It can also help to identify different environments through the differences between genotypes with minimal replicates. Environments with better yield can distinguish superior genotypes in target environments [7]. Identifying high-yielding and sustainable genotypes in
diverse environments has been a constant challenge for plant breeders worldwide. Features of sustainable genotypes made possible by successive genotype × environment interactions (GEI). Since the environment very much influences genotypes, locations, and years, performance related to the stability of genotypes is a prerequisite and must carry out in different environments over different years in a given location [8]. Plant breeders are evaluating genotypes in multienvironment trials (MET), including in favorable and unfavorable environmental conditions on the genotype × environment interaction (GEI). MET variance analysis provides reasonable estimates of the critical effects of genotype (G) and environment (E) and genotype × environment interaction (GEI) in which the effects of G and GEI mainly lead to cultivar evaluation. The value of observed genetic variance varies from one environment to another due to interaction effect, and it tends to improve in superior environments higher than unfavourable environments [9]. The genotype-environment interaction (GEI) for quantitative traits such as grain yield can limit the selection of superior genotypes to develop modified cultivars [10]. To calculate GEI, breeders evaluate genotypes in various environments to identify high-yielding and stable genotypes. Genotypes with no GEI are considered stable genotypes [11]. Yield stability is one of the most desirable traits of a genotype that allows it to identify as a cultivar. The construction of a large-scale facility is required to study genotypes, referring to as specific adaptability. To achieve maximum production, it is essential to develop hybrid varieties that best fit given target environments and have specific adaptability. Additive main effects and multiplicative interaction analysis (AMMI) allow for a large set of technical interpretations, and they were used more commonly to evaluate the genotype-environment interactions. The additive main effects and multiplicative interaction (AMMI) model addresses the limitations of ANOVA and PCA. The AMMI model effectively explains the GEI patterns. The AMMI method is used for three primary purposes for stability. The first is that the model involves variance analysis and principal component analysis. Second, AMMI patterns. The AMMI method is used for three primary purposes for stability. The first is that the model involves variance analysis and principal component analysis. Second, AMMI patterns. The AMMI method is used for three primary purposes for stability. The first is that the model involves variance analysis and principal component analysis. Second, AMMI patterns. The AMMI method is used for three primary purposes for stability. The first is that the model involves variance analysis and principal component analysis. Second, AMMI patterns. (The AMMI method is used for three primary purposes for stability. The first is that the model involves variance analysis and principal component analysis. Second, AMMI patterns. The AMMI method is used for three primary purposes for stability. The first is that the model involves variance analysis and principal component analysis. Second, AMMI patterns.)
environments are linked to environmental impacts. The interaction sum of squares is separated into two sections: one section expresses the heterogeneity of linear regression coefficients \((b_i)\), and the second section expresses the pooled differences from unique regression lines \(S_d^2\).

Eberhart and Russell [22] suggested an evaluation of hybrid responses to environmental variations applying a linear regression coefficient \(b_i\) plus the variance of the regression differences \(S_d^2\):

\[
b_i = 1 + \frac{\sum (X_{ij} - \bar{X}_i - \bar{X}_j + \bar{X}) (\bar{X}_j - \bar{X})}{\sum (\bar{X}_j - \bar{X})^2}
\]

\[
S_d^2 = \frac{1}{E-2} \left[ \sum (X_{ij} - \bar{X}_i - \bar{X}_j + \bar{X}) - (b_i - 1)^2 \sum (\bar{X}_j - \bar{X})^2 \right]
\]

\[
(5)
\]

\(X_{ij}\) is the grain yield of genotype \(i\) in environment \(j\); \(\bar{X}_i\) is the average yield of hybrid \(i\) and \(\bar{X}_j\) is the average performance of the environment \(j\); \(\bar{X}\) is the grand mean, and \(E\) is the number of environments.

The environmental variance is one of the significant stability measures of the static stability concept and is calculated for each genotype across test environments. A genotype with minimum variance under different environments was considered to be stable. This measure was calculated as follows:

\[
S_{xi}^2 = \frac{\sum (X_{ij} - \bar{X}_i)^2}{(E-1)}
\]

where \(X_{ij}\) is the grain yield of genotype \(i\) in environment \(j\); \(\bar{X}_i\) is the mean yield of genotype \(i\), and \(E\) is the number of environments.

The AMMI stability value was calculated as previously described by Purchase et al [15] and used by Abate [23] and Adjeeng-Danquah et al. [24]:

\[
ASV = \sqrt{\frac{IPCA_{1 SQ}}{PCA_{2 SQ}} (PCA_{1 score})^2 + (PCA_{2 score})^2}
\]

where \(IPCA_{1 SQ}/PCA_{2 SQ}\) is the weight obtained from separating the IPCA1 sum of squares by the IPCA2 sum of squares. The greater the total amount of IPCA, the higher the adaptability of a particular variety for a particular environment. Conversely, lower ASV scores indicate more excellent stability in different environments.

The performance stability index calculated applying the sum of the ranking based on performance and ranking based on the AMMI stability amount.

\[
YSI = RASV + RY.
\]

RASV is the genotypes’ rank based on the AMMI stability value, and \(RY\) is the rank of the genotypes based on yield across environments [12, 25].

### 3. Results

A combine analysis of variance showed that environmental effect, genotype effect, and genotype-environment interaction were significant at one percent probability level. Significance of the effect of environments indicated that environments differed in yield of genotypes, and the significance of GEI showed that the yield of genotypes varied in different environments (Table 3).

Considering Duncan’s mean comparison in 1% probability level on these data and yield of genotypes, KSC707 ranked the highest and KSC704 and SC301 ranked the lowest (Table 2).

Kandus et al. [26] stated that the best model is when only the first two principal components are significant, and the other components showed little variation. The first and second principal components are representing the genotypes and environments, respectively, as can be seen in Table 4.

The environments with the first significant interactions are suitable for identifying and screening genotypes. In this study, the E2 environment was selected as the most suitable environment for screening the genotypes. SC647 genotype with a relatively high yield showed the lowest first principal component of interaction, so it is highly stable. It can be used as a desirable genotype for planting in different regions and years (Table 4). According to the environmental variance method, a genotype with the lowest environmental variance has been selected as the most desirable hybrid; SC604 and KSC703 genotypes had less environmental variance than the rest of the genotypes and hence were considered the most desirable genotypes in this method. SC302 and KSC400 genotypes having the highest environmental variance were identified as the most unstable hybrids. According to the environmental coefficient of variation (CVi), a stable genotype has the least value of this parameter. KSC707 and SC604 genotypes had the highest values of mean rank of parameters, and hence, they were identified as the most stable hybrids.

### Table 1: Annual rainfall mean, codes, and geographical parameters for different environments.

| Location code | Location | Longitude | Latitude | Elevation AMSL (m) | Average rainfall (mm) |
|---------------|----------|-----------|----------|------------------|-----------------------|
| Year 1        | Year 2   |           |          |                  |                       |
| E1            | E2       | Karaj     | 51.00    | 35.48            | 1321                  | 295                   |
| E3            | E4       | Birjand   | 59.19    | 32.53            | 1420                  | 171                   |
| E5            | E6       | Shiraz    | 52.32    | 29.36            | 1670                  | 337.8                 |
| E7            | E8       | Arak      | 49.4     | 34.00            | 1708                  | 341.7                 |
in this method. Also, KSC400 and SC302 genotypes having the highest environmental variation coefficient were identified as the most unstable hybrids. These hybrid had a smaller percentage in genotype and environment interaction, according to Wricke’s ecovalence and Shukla’s stability variance parameters. The Based on the results of these two methods, KSC705, KSC706, and KSC704 genotypes were identified as the most stable hybrids. Also, KSC703, KSC707, and DC370 genotypes were identified as the most unstable genotypes because of the high level of these two parameters. The high value of regression ($b_i > 1$) indicates that the variety is more responsive for the input-rich environment, while the low value of regression ($b_i < 1$) is an indication that the variety may be adopted in a low environment. Thus, the KSC400 genotype is more responsive for the input-rich environment, and the KSC703 genotype is adopted in a low environment. Considering Eberhart and Russell’s regression coefficient ($s^2_{di}$), any genotype with the least value was more stable. Consequently, KSC704 and SC647 genotypes having the lowest mean yield parameter were recognized as the most stable hybrids. SC302 and SC301 genotypes having the highest mean yield parameter were identified as the most unstable genotypes. Based on the detection coefficient method, a stable genotype benefits from this highest statistic value, according to which KSC705, KSC706, KSC707, and SC647 hybrids with the highest parameters were identified as the most stable genotypes. The KSC707 genotype was identified as one of the most unstable hybrids due to its high statistic level. Although the methods mentioned above have similarities in identifying the stable genotypes, some differences indicate that the above parameters’ results are not wholly consistent. In the environmental variance method, KSC703 and SC604 hybrids were selected as the most stable genotypes; in the coefficient of variation, SC604 and KSC707 hybrids; in the genotypic variance-based method, KSC705, KSC706, and KSC704 hybrids; in Eberhart and Russell’s regression coefficient method, KSC704 and SC647; and in detection coefficient method; KSC706, KSC704, and SC647 hybrids were selected as the most stable genotypes, which reveal the differences in the hybrid selection. KSC400 and SC302 genotypes were selected as the most unstable genotypes in identifying the most unstable hybrids based on environmental coefficient of variation; based on variance methods, KSC703, DC370, and KSC707 genotypes; based on Eberhart and Russell’s regression coefficient method, SC302 and SC301 genotypes; and based on detection coefficient, KSC707 were selected as the most unstable hybrids. The univariate analysis was performed on the data of four regions in the second crop year. A genotype with the least environmental variation was selected as the most desirable hybrid. The SC647 genotype showed less environmental variation than the rest of the genotypes and hence was recognized as the most desirable genotype of this method. KSC703 and KSC705 genotypes identified as the most unstable hybrids because of having the highest environmental variance. According to the environmental coefficient of variation (CVi), a stable genotype showed the lowest value of this parameter, and accordingly, SC647 and KSC707 genotypes had the lowest value of this parameter and hence were identified as the most stable hybrids of this method. Also, KSC705, KSC706, and KSC704 genotypes were identified as the most unstable hybrids because of having the highest environmental variation coefficient. According to GEI and environmental variance methods, including Wricke’s ecovalence and Shukla’s stability variance parameters, the lower these two statistics, the higher the stability of the hybrids. Based on the results of these two methods, SC647, SC302, and SC301 genotypes were identified as the most stable hybrids because of having the lowest values in both statistics. Also, KSC706 and KSC704 genotypes were identified as the most unstable genotypes because of having the highest values in both statistics. According to the $b_i$ parameter, DC370, SC604, KSC707, and KSC705 genotypes are more responsive for input-rich environments, and the KSC706 genotype adopted in a low environment. Regarding Eberhart and Russell regression coefficient, any genotype with the least value of this parameter is more stable. Consequently, SC302 and SC647 genotypes with the least value of this parameter identified as the most stable hybrids. In addition, KSC703, KSC704, and KSC706 were identified as the most unstable genotypes because of their high value of this parameter. Based on the detection coefficient method, a stable genotype showed the highest

| Genotype no. | Genotype | Means |
|------------|----------|-------|
| G1 | KSC 703 | 6.76 ab |
| G2 | KSC 260 | 5.99 bc |
| G3 | KSC 705 | 6.02 bc |
| G4 | KSC 400 | 6.05 bc |
| G5 | KSC 706 | 6.3 abc |
| G6 | KSC 704 | 5.55 c |
| G7 | KSC 707 | 6.9 a |
| G8 | SC 307 | 6.3 abc |
| G9 | SC 647 | 5.78 c |
| G10 | SC 302 | 5.72 c |
| G11 | SC 604 | 6.08 bc |
| G12 | SC 301 | 5.51 c |
value, and so KSC705, SC647, and SC301 hybrids were identified as the most stable genotypes. KSC400, KSC706, and DC370 genotypes were identified as the most unstable hybrids due to their high value. The univariate analysis was performed on the data of four regions in two crop years. The genotypes with the lowest value were selected as the most desirable hybrids considering the environmental variance, based on which KSC707, DC370, SC647, and SC604 genotypes showed less environmental variance than the rest of the genotypes, and hence they were identified as the most desirable genotypes of this method. The KSC705 genotype was identified as the most unstable hybrid since it showed the highest environmental variance. According to the environmental coefficient of variation (CVi), a stable genotype valued the least, and based on which the KSC707 genotype showed the lowest value of the parameter and hence was identified as the most stable hybrid of this method. Also, KSC705, KSC400, KSC706, and KSC704 genotypes were identified as the most unstable hybrids with the highest environmental coefficient of variation. According to GEI and environmental variance methods, including Wricke's ecovalence and Shukla's stability variance parameters, the lower these statistics in the genotypes, the higher the stability of the hybrids; and based on the obtained results, KSC260, KSC705, and SC647 genotypes were identified as the most stable hybrids due to their lowest values in both statistics. Also, KSC703, KSC706, KSC707, and DC370 genotypes were identified as the most unstable genotypes due to their highest levels of these two statistics. According to the b parameter, KSC705 is more responsive for an input-rich environment, and the DC370 genotype was adopted in a low environment. According to Eberhart and Russell regression coefficient, any genotype with the least value of this parameter is more stable. Consequently, KSC260, KSC707, and SC647 genotypes had the least value of this parameter, and they were identified as the most stable hybrids. KSC706 was identified as the most unstable genotype because it showed the highest levels of this parameter. The genotype was identified as stable when it showed the highest value of this parameter considering the detection coefficient (Table 5).

4. Discussion

The important goals of plant breeding are to select hybrids with wide adaptation in a series of condition that usually has good and stable performance. The most appropriate methods include identifying desirable cultivars with high productivity genetic potential and testing wide adaptability to most conditions by multi-condition experiments in target environments [27]. The present study determined the stability and adaptability of maize hybrids. Due to the significant combined analysis, the grain yield stability could be examined and hybrids with specific adaptability to each environment and hybrids with general adaptability to all environments can be measured. The significance of genotype × environment interaction indicated that environments could classify GEI [28, 29]. The SC647 genotype with the lowest AMMI stability value (0.001) was identified as the most stable genotype among 12 genotypes. The highest ASV related to the SC302 genotype (0.072). The sum of the yield and stability rankings (YSI) ranked KSC707 as the genotype that combined high yield with stability. The KSC706 genotype though high yielding was unstable because of its low rank according to the YSI. The two genotypes KSC400 and DC370 can be considered high yielding and stable (Table 4). Therefore, genotypes that have been considered stable genotypes based on ASV and IPCA parameters tend to have a lower average yield [30]. Using stability parameters on data obtained from four regions in the first crop year, KSC260, KSC705, SC647, and SC301 hybrids showed the highest value of the parameters. Also, KSC703, KSC707, and DC370 genotypes identified as the most unstable hybrids due to their high statistic levels. Based on the analysis of the four tested regions in the first crop year, the second crop year, and the mean of two crop years, and considering all the calculated parameters, SC647 and KSC705 genotypes identified as the most stable and KSC707 and KSC706 genotypes as the most unstable genotypes in case of grain yield. AMMI can have several models: AMMI0, which estimates the main additive effect of genotypes and environments and does not include any major axis (IPCA); AMMI1, which combines AMNI0 genotype additive effects with environmental interactions estimated and combines from the first major axis (IPCA 1); and AMMI2 and others up to the full model with all IPCA axes. [31]. According to the AMMI1 graph, which explained 73.4% of the total squares of the data variance, the environmental share was 38.36%, the genotype share was 12.01%, and the first component share was 23%. Accordingly, a genotype close to the mean axis and on the positive axis is more stable. KSC400, KSC705, and SC647 genotypes were more stable in comparison with other genotypes. KSC706, KSC703, and KSC707 genotypes are the most unstable, low-yielding genotypes, and the DC370 genotype, the most stable, high-yielding genotype. Environmental stability is essential for demonstrating the reliability of genotype ordering in a given environment about the rating for the environments in question [32]. According to the AMMI2 graph, 68.1% of the data variance explained 46.4% of the first component and 21.7% to the second component. Considering this graph, SC604 and SC302 genotypes in E7 and E1 environments; DC370 genotype in E4 environment; KSC400, KSC707, and KSC706 genotypes in E5 environment; and KSC260 and SC301 genotypes in E3 environment benefited from a desirable yield. Regarding the AMMI2 graph, the SC647 genotype was more stable than other genotypes, but unlike the AMMI1 graph with more stable KSC705 and KSC400 genotypes, these genotypes identified as unstable (Figures 1 and 2).
Table 3: Combined analysis of variance for maize hybrid in four environments and two years.

| Source of variation | DF | SS    | MS    |
|---------------------|----|-------|-------|
| Local               | 3  | 110.1 | 36.7* |
| Year                | 1  | 35.2  | 35.2* |
| Year × Local        | 3  | 13.02 | 4.34* |
| Error 1             | 16 | 46.4  | 2.9   |
| Genotype            | 11 | 49.5  | 4.5   |
| Genotype × Local    | 33 | 84.94 | 2.5*  |
| Genotype × Year     | 11 | 32.7  | 2.9** |
| Genotype × Year × Local | 33 | 87.18 | 2.6*  |
| Error 2             | 176| 293.92| 1.67  |

Symbols *, **, and ns denote significance at 5%, significance at 1%, and not significant, respectively.

Table 4: Ranking of 12 hybrids of maize and 8 environments based on the AMMI stability value (ASV).

| Genotypes | Mean (t/ha) | Rank (A) | IPCA 1 | IPCA 2 | ASV | ASV rank (B) | YSI (A + B) | YSI rank |
|-----------|-------------|----------|--------|--------|-----|--------------|-------------|----------|
| G1        | 6.76        | 2        | 0.025  | −0.001 | 0.056| 8            | 10          | 4        |
| G2        | 5.99        | 8        | −0.025 | 0.005  | 0.055| 7            | 15          | 9        |
| G3        | 6.02        | 7        | 0.0004 | 0.044  | 0.044| 5            | 12          | 7        |
| G4        | 6.05        | 6        | 0.005  | −0.019 | 0.022| 2            | 8           | 3        |
| G5        | 6.31        | 3        | 0.029  | −0.011 | 0.066| 11           | 14          | 8        |
| G6        | 5.55        | 11       | 0.026  | −0.002 | 0.058| 9            | 20          | 10       |
| G7        | 6.9         | 1        | 0.019  | −0.010 | 0.043| 4            | 5           | 1        |
| G8        | 6.3         | 4        | −0.003 | −0.022 | 0.023| 3            | 7           | 2        |
| G9        | 5.78        | 9        | 0.001  | −0.003 | 0.005| 1            | 10          | 5        |
| G10       | 5.72        | 10       | 0.032  | −0.0007| 0.072| 12           | 22          | 11       |
| G11       | 6.08        | 5        | −0.021 | −0.006 | 0.047| 6            | 11          | 6        |
| G12       | 5.51        | 12       | −0.027 | 0.001  | 0.060| 10           | 22          | 12       |

Table 5: Mean gain yield and 8 estimates of parametric stability measures for maize yield across 4 environments in 2 years.

| Genotype | Mean | $S^2X_i$ | CV   | $W^2$ | $SH$ | $b_i$ | $S_{ma}^2$ | $R_i^2$ |
|----------|------|----------|------|-------|------|-------|------------|---------|
| G1       | 6.33 | 0.34     | 9.33 | 8.32  | 3.23 | −0.06 | 0.51       | 0.02    |
| G2       | 5.96 | 1.90     | 23.16| 3.79  | 1.57 | 0.64  | 1.49       | 0.47    |
| G3       | 5.97 | 1.54     | 20.81| 1.24  | 0.28 | 0.76  | 0.44       | 0.8     |
| G4       | 7.01 | 3.81     | 27.48| 2.18  | 0.67 | 1.21  | 0.93       | 0.83    |
| G5       | 6.97 | 1.32     | 16.53| 1.02  | 0.19 | 0.72  | 0.27       | 0.86    |
| G6       | 6.18 | 1.08     | 16.79| 1.09  | 0.22 | 0.66  | 0.18       | 0.88    |
| G7       | 6.95 | 1.42     | 9.31 | 8.04  | 3.11 | −0.02 | 0.62       | 0.003   |
| G8       | 6.27 | 1.07     | 16.51| 7.8   | 3.02 | 0.14  | 1.54       | 0.04    |
| G9       | 6.20 | 0.08     | 14.48| 1.43  | 0.36 | 0.57  | 0.14       | 0.88    |
| G10      | 6.32 | 2.85     | 26.7 | 5.86  | 2.21 | 0.7   | 2.65       | 0.37    |
| G11      | 6.51 | 0.45     | 10.37| 6.1   | 2.54 | 0.13  | 0.68       | 0.08    |
| G12      | 6.38 | 1.75     | 20.76| 2.99  | 1.24 | 0.67  | 2.63       | 0.55    |
Table 5: Continued.

| Genotype | Mean | $S^2X_i$ | CV  | $W^2$ | SH  | $b_i$ | $S^2_{ei}$ | $R^2$ |
|----------|------|----------|-----|-------|-----|-------|------------|-------|
| G1       | 6.84 | 3.05     | 25.5| 6.34  | 2.36| 0.83  | 3.12       | 0.31  |
| G2       | 5.74 | 1.23     | 19.3| 2.65  | 1.1 | 0.62  | 1.04       | 0.43  |
| G3       | 6.25 | 3.66     | 30.6| 2.76  | 0.87| 1.49  | 0.86       | 0.54  |
| G4       | 5.53 | 1.13     | 19.29| 7.33  | 2.77| 0.02  | 1.7        | 0.007 |
| G5       | 5.85 | 2.98     | 29.5| 13.59 | 5.38| 0.06  | 4.47       | 0.001 |
| G6       | 5.21 | 1.41     | 0.03| 3.12  | 0.23| 0.33  | 8.65       | 28.19 |
| G7       | 6.76 | 0.63     | 0.11| 0.67  | 0.2 | 1.37  | 3.96       | 10.54 |
| G8       | 6.14 | –0.13    | 0.003| 1.21  | 0.04| 2.3   | 6.19       | 14.66 |
| G9       | 5.58 | 0.66     | 0.87| 0.03  | 0.35| 0.27  | 1.80       | 7.88  |
| G10      | 4.90 | 0.35     | 0.46| 0.35  | 0.38| 0.67  | 2.27       | 13.55 |
| G11      | 5.35 | 0.38     | 0.34| 1.75  | 0.54| 1.32  | 3.18       | 20.22 |
| G12      | 4.59 | 1.05     | 0.77| 1.34  | 0.71| 0.39  | 0.95       | 20.61 |

Year 2

| Genotype | Mean | $S^2X_i$ | CV  | $W^2$ | SH  | $b_i$ | $S^2_{ei}$ | $R^2$ |
|----------|------|----------|-----|-------|-----|-------|------------|-------|
| G1       | 6.58 | 1.03     | 15.44| 4.97  | 1.9 | 0.31  | 1.29       | 0.16  |
| G2       | 5.85 | 0.62     | 13.53| 1.12  | 0.46| 0.57  | 0.1        | 0.89  |
| G3       | 6.11 | 2.06     | 23.52| 0.67  | 0.11| 1.04  | 0.33       | 0.89  |
| G4       | 6.31 | 1.29     | 17.99| 2.29  | 0.79| 0.65  | 0.84       | 0.56  |
| G5       | 6.41 | 1.29     | 17.74| 4.46  | 1.69| 0.44  | 1.44       | 0.25  |
| G6       | 6.7  | 1.01     | 17.65| 3.02  | 1.09| 0.5   | 0.88       | 0.41  |
| G7       | 6.86 | 0.24     | 7.18 | 4.42  | 1.67| 0.13  | 0.31       | 0.13  |
| G8       | 6.21 | 0.46     | 10.97| 5.7   | 2.22| 0.07  | 0.68       | 0.01  |
| G9       | 5.89 | 0.44     | 11.33| 1.48  | 0.45| 0.48  | 0.07       | 0.89  |
| G10      | 5.61 | 0.85     | 16.43| 2.68  | 0.95| 0.48  | 0.67       | 0.47  |
| G11      | 5.93 | 0.41     | 10.82| 3.07  | 1.28| 0.31  | 0.61       | 0.41  |
| G12      | 5.48 | 0.83     | 16.6 | 1.17  | 0.49| 0.62  | 1.24       | 0.8   |

Average of two years

| Genotype | Mean | $S^2X_i$ | CV  | $W^2$ | SH  | $b_i$ | $S^2_{ei}$ | $R^2$ |
|----------|------|----------|-----|-------|-----|-------|------------|-------|
| G1       | 6.07 | 0.35     | 15.44| 4.97  | 1.9 | 0.31  | 1.29       | 0.16  |
| G2       | 5.85 | 0.62     | 13.53| 1.12  | 0.46| 0.57  | 0.1        | 0.89  |
| G3       | 6.11 | 2.06     | 23.52| 0.67  | 0.11| 1.04  | 0.33       | 0.89  |
| G4       | 6.31 | 1.29     | 17.99| 2.29  | 0.79| 0.65  | 0.84       | 0.56  |
| G5       | 6.41 | 1.29     | 17.74| 4.46  | 1.69| 0.44  | 1.44       | 0.25  |
| G6       | 6.7  | 1.01     | 17.65| 3.02  | 1.09| 0.5   | 0.88       | 0.41  |
| G7       | 6.86 | 0.24     | 7.18 | 4.42  | 1.67| 0.13  | 0.31       | 0.13  |
| G8       | 6.21 | 0.46     | 10.97| 5.7   | 2.22| 0.07  | 0.68       | 0.01  |
| G9       | 5.89 | 0.44     | 11.33| 1.48  | 0.45| 0.48  | 0.07       | 0.89  |
| G10      | 5.61 | 0.85     | 16.43| 2.68  | 0.95| 0.48  | 0.67       | 0.47  |
| G11      | 5.93 | 0.41     | 10.82| 3.07  | 1.28| 0.31  | 0.61       | 0.41  |
| G12      | 5.48 | 0.83     | 16.6 | 1.17  | 0.49| 0.62  | 1.24       | 0.8   |

G1: KSC703; G2: KSC260; G3: KSC705; G4: KSC400; G5: KSC706; G6: KSC704; G7: KSC707; G8: DC370; G9: SC647; G10: SC302; G11: SC604; G12: SC301.

Figure 1: AMMI 1 biplot for main effects vs. IPCA1 in 12 hybrids of Zea mays L. from eight environments. G1: KSC703; G2: KSC260; G3: KSC705; G4: KSC400; G5: KSC706; G6: KSC704; G7: KSC707; G8: DC370; G9: SC647; G10: SC302; G11: SC604; G12: SC301. E1: Karaj Year 1; E2: Karaj Year 2; E3: Birjand Year 1; E4: Birjand Year 2; E5: Shiraz Year 1; E6: Shiraz Year 2; E7: Arak Year 1; E8: Arak Year 2.
5. Conclusion

Regarding the AMMI model, the results of the analysis of variance indicated significant genotype × environment interaction. E2 environment selected as the most suitable environment for screening the genotypes. Based on the results of the data analysis of the four tested regions in the first crop year, the second crop year, and the mean of two crop years, and considering all the calculated parameters, SC647 and KSC705 genotypes were identified as the most stable and KSC707 and KSC706 genotypes as the most unstable genotypes in case of grain yield.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare no conflicts of interest.

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