Description of a new horned toad of *Megophrys* Kuhl & Van Hasselt, 1822 (Amphibia, Megophryidae) from Zhejiang Province, China

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Abstract

A new species of the Asian horned toad genus *Megophrys* is described from Zhejiang Province, China, based on multiple data. Molecular phylogenetic analyses based on mitochondrial DNA indicated the new species as an independent clade deeply clustered into the *Megophrys* clade. The new species is identified from its congeners by a combination of the following characters: body size small (SVL 28.4–32.4 mm in males); vomerine teeth absent; tongue not notched behind; tympanum distinctly visible, oval; a small horn-like tubercle present at the edge of each upper eyelid; two metacarpal tubercles distinctly visible in hand; toes without webbing; heels overlapped when thighs are positioned at right angles to the body; tibiotarsal articulation reaching the level to middle of eye when leg stretched forward; an internal single subgular vocal sac in male; in breeding male, the nuptial pads present on the dorsal base of the first two fingers.

Keywords

Molecular phylogenetic analyses, morphology, new species, taxonomy, toad

* These authors have contributed equally to this work.

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Introduction

The Asian horned toad *Megophrys* Kuhl & Van Hasselt, 1822 (Anura: Megophryidae Bonaparte, 1850) is widely distributed in eastern and central China, throughout southeastern Asia, and extending to the islands of the Sunda Shelf and the Philippines (Frost 2020). The generic assignment of species in the group has been controversial for decades (e.g., Tian and Hu 1983; Dubois 1987; Rao and Yang 1997; Lathrop 1997; Jiang et al. 2003; Delorme et al. 2006; Fei et al. 2009; Fei and Ye 2016; Chen et al. 2017; Deuti et al. 2017; Mahony et al. 2017; Li et al. 2020). Recent molecular phylogenetic studies proposed this group as a monophyletic group (Chen et al. 2017; Mahony et al. 2017; Li et al. 2018a; Liu et al. 2018; Liu et al. 2020; Wang et al. 2020), which was recognized as a big genus *Megophrys sensu lato* (Mahony et al. 2017; Li et al. 2018b; Liu et al. 2018; Liu et al. 2020; Lyu et al. 2020; Xu et al. 2020; Wang et al. 2020), though some studies still divided the taxa of the group into different genera and/or subgenera (Fei and Ye 2016; Chen et al. 2017; Deuti et al. 2017; Liu et al. 2018). The genus *Megophrys* currently contains 106 species, of which 52 species were described over the last decade (Frost 2020). A number of cryptic species were still indicated in the genus by molecular phylogenetic analyses (e.g., Chen et al. 2017; Liu et al. 2018).

Wuyi Mountain region, located in northern Fujian, southeastern Jiangxi and southern Zhejiang provinces of China, is a biodiversity hotspot. In this region, four *Megophrys* species have been recorded, i.e., *M. boettgeri* (Boulenger, 1899), *M. kuatunensis* Pope, 1929, *M. ombrophila* Messenger & Dahn, 2019, and *M. lishuiensis* Wang, Liu & Jiang, 2017. However, many mountains in this region, especially in southern Zhejiang Province, have been poorly investigated.

During field surveys in Qingyuan County, Zhejiang Province, China, we collected *Megophrys* specimens. Molecular phylogenetic analyses and morphological comparisons supported some of these specimens as an undescribed taxon that we describe herein as a new species.

Materials and methods

Sampling

A total of 15 specimens were sampled in this study: six adult males and one tadpole of the undescribed species and two adult males of *M. boettgeri* from Qingyuan County, Zhejiang Province, China, and one adult male of *M. ombrophila* and six adult males of *M. kuatunensis* from Wuyi Mountain, Fujian Province, China (Table 1; Fig. 1). The developmental stage of tadpole was identified following Gosner (1960). In the field, the toad and tadpole were euthanized using isoflurane, and the specimens were fixed in 75% ethanol. Tissue samples were taken and preserved separately in 95% ethanol prior to fixation. The specimens were deposited in Chengdu Institute of Biology, Chinese Academy of Sciences (*CIB, CAS*).
A new species of *Megophrys*

**Table 1.** Information for samples used in molecular phylogenetic analyses in this study.

| ID | Species | Voucher number | Localities | GenBank accession number |
|----|---------|----------------|------------|-------------------------|
| 1  | *Megophrys baishanzuensis* sp. nov. | CIBQY20200719001 | Baishanzu National Park, Qingyuan, Zhejiang, China | MW001150 MT998291 |
| 2  | *Megophrys baishanzuensis* sp. nov. | CIBQY20200719002 |                  | MW001151 MT998292 |
| 3  | *Megophrys baishanzuensis* sp. nov. | CIBQY20200719003 |                  | MW001152 MT998293 |
| 4  | *Megophrys baishanzuensis* sp. nov. | CIBQY20200719004 |                  | MW001153 MT998294 |
| 5  | *Megophrys baishanzuensis* sp. nov. | CIBQY20200719006 |                  | MW001154 MT998295 |
| 6  | *Megophrys baishanzuensis* sp. nov. | CIBQY20200726001 |                  | MW001155 MT998296 |
| 7  | *Megophrys baishanzuensis* sp. nov. | CIBQY20200726002 |                  | MW001156 MT998297 |
| 8  | *Megophrys kuatsusennis* | CIBWY180828407 | Wuyi Shan, Fujian, China | MW001157 MT998298 |
| 9  | *Megophrys kuatsusennis* | CIBWY180828408 |                  | MW001158 MT998299 |
| 10 | *Megophrys kuatsusennis* | SYS a001579 |                  | KJ560376 – |
| 11 | *Megophrys lini* | SYS a002370 | Suichuan, Jiangxi, China | KJ560412 – |
| 12 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 13 | *Megophrys nanlingensis* | SYS a001959 | Nanling Nature Reserve, Guangdong, China | MK524111 MK524142 |
| 14 | *Megophrys lini* | SYS a002370 | Suichuan, Jiangxi, China | KJ560412 – |
| 15 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 16 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 17 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 18 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 19 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 20 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 21 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 22 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 23 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 24 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 25 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 26 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 27 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 28 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 29 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 30 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 31 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 32 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 33 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 34 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 35 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 36 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 37 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 38 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 39 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 40 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 41 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 42 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 43 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 44 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 45 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 46 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |
| 47 | *Megophrys xiangnanensis* | SYS a002874 | Yangming Shan, Hunan, China | MH406713 MH406165 |

**Notes:**
- COI: Control Region
- 16S: Small Subunit
- MW: GenBank accession number
- MT: GenBank accession number
| ID | Species                  | Voucher number       | Locality                                | GenBank accession number |
|----|--------------------------|----------------------|-----------------------------------------|--------------------------|
| 48 | *Megophrys jingdongensis* | KIZ-LC0805067        | Huanglianshan National Nature Reserve, Yunnan, China | KX811872, KX812131       |
| 49 | *Megophrys fansipanensis* | VNMM 2018.01         | Lao Dai, Sa Pa, Vietnam                 | MH514886                 |
| 50 | *Megophrys huangdiensis*  | VNMM 2018.02         | Lao Dai, Sa Pa, Vietnam                 | MH514889                 |
| 51 | *Megophrys minor*         | KIZ01939             | Qingcheng Shan, Sichuan, China          | KX811896, KX812145       |
| 52 | *Megophrys jiangi*       | CIBKKS20180722006    | Kuankuosui National Reserve, Guizhou, China | MN107743, MN107748       |
| 53 | *Megophrys chibuisiensis* | CIBCS20190518031     | Chishui National Reserve, Guizhou, China | MN954707, MN928958       |
| 54 | *Megophrys brachykolos*  | ROM 16634            | Hong Kong, China                        | KX811897, KX812150       |
| 55 | *Megophrys gerti*        | ITBCZ 1108           | Nuí Chua National Park, Ninh Thuan, Vietnam | KX811917, KX812161       |
| 56 | *Megophrys acuta*        | CIBKKS20180722006    | Phong Dien Nature Reserve, Thua Thien Hue, Vietnam | KX811913, KX812155       |
| 57 | *Megophrys microstoma*   | KIZ048799            | Xiaoqiaogou National Reserve, Yunnan, China | KX811914, KX812156       |
| 58 | *Megophrys pachyproctus* | KIZ010978            | Beibeng, Xizang, China                  | KX811908, KX812153       |
| 59 | *Megophrys nasuta*       | ZMMU ABV-00454       | Bident Mountain, Lam Dong, Vietnam       | KY425379                 |
| 60 | *Megophrys stejnegeri*   | FMNH 237694          | Gunung Kinabalu National Park, Kogopan Trail, Malaysia | KJ831310                 |
| 61 | *Megophrys ligayae*      | ZMMU NAP-05015       | Pasonanca Natural Park, Zamboanga, Philippines | KX811922, KX812052       |
| 62 | *Megophrys kobayashii*   | UNIMAS 8148          | Gunung Kinabalu National Park, Sabah, Malaysia | KJ831313                 |
| 63 | *Megophrys himalayana*   | SDBDU2009.75         | East Siang Dist., Arunachal Pradesh, IN  | KY022309, MH647528       |
| 64 | *Megophrys major*        | SYS a002961          | Nanjiang, Sichuan, China                | MH406728, MH406180       |
| 65 | *Megophrys oreocrypta*   | BNHS 6046            | West Garo Hills Dist., Meghalaya        | KY022306                 |
| 66 | *Megophrys auralensis*   | NCSM 79599           | Aural, Kampong Speu, Cambodia           | KX811807                 |
| 67 | *Megophrys nankiangensis*| CIB ZYC517           | Nanjiang, Sichuan, China                | MH406737                 |
| 68 | *Megophrys dringi*       | UNIMAS 8148          | Nanling National Forest Park, Guangdong, China | KX811790, KX812079       |
| 69 | *Megophrys nannai*       | ZSIA11799            | –                                      | KX894669                 |
| 70 | *Megophrys major*        | SYS a002961          | –                                      | KX894669                 |
| 71 | *Megophrys major*        | BNHS 6046            | –                                      | MH406728, MH406180       |
| 72 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 73 | *Megophrys elongatus*    | BNHS 6046            | –                                      | MH406728, MH406180       |
| 74 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 75 | *Megophrys elongatus*    | BNHS 6046            | –                                      | MH406728, MH406180       |
| 76 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 77 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 78 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 79 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 80 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 81 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 82 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 83 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 84 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 85 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 86 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 87 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 88 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 89 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 90 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 91 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 92 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 93 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 94 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 95 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 96 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 97 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 98 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 99 | *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 100| *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 101| *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
| 102| *Megophrys elongatus*    | UNIMAS 8148          | –                                      | MH406728, MH406180       |
A new species of *Megophrys*

| ID | Species                | Voucher number | Locality                        | GenBank accession number |
|----|------------------------|----------------|---------------------------------|--------------------------|
| 92 | *Megophrys intermedia* | ZFMK 87596     | U Bo, Phong Nha-Ke Bang NP, Vietnam | HQ588950 –              |
| 93 | *Megophrys Montana*     | LSUMZ 81916    | Sukabumi, Java, Indonesia       | KX811927 KX812163        |
| 94 | *Megophrys lanzip*      | MZB: Amp:22233 | –                               | KY679891 –              |
| 95 | *Leptobrachium boriogii*| Tissue ID: YPX37539 | Emei Shan, Sichuan, China | KX811930 KX812164 |
| 96 | *Leptobrachella oshanensis* | KIZ025778 | Emei Shan, Sichuan, China | KX811928 KX812166 |

Figure 1. Sampling localities of *Megophrys baishanzuensis* sp. nov. and its relatives

1. Baishanzu National Park, Qingyuan County, Zhejiang Province, China, inhabited by *Megophrys baishanzuensis* sp. nov. and *M. boettgeri*
2. Wuyi Mountain, Wuyishan City, Fujian Province, China, inhabited by *M. boettgeri*, *M. kuatunensis*, and *M. ombrophila*.

Molecular data and phylogenetic analyses

Six adult males and one tadpole of the undescribed species, two *M. kuatunensis*, one *M. ombrophila*, and two *M. boettgeri* were included in the molecular analyses (Table 1). Total DNA was extracted using a standard phenol-chloroform extraction protocol (Sambrook et al. 1989). Two fragments of the mitochondrial 16S rRNA (16S) and cytochromeoxidase subunit I (COI) genes were amplified. For 16S, the primers P7 (5’-CGCCTGTTTTACCAAAAAACAT-3’) and P8 (5’-CCGGTCTGAACTCAGATCACGT-3’) were used following Simon et al. (1994), and for COI, Chmf4 (5’-TYTGWCAWCCAYAAAGAYATCGG-3’) and Chmr4 (5’-ACYTCTGGRTGRCCRAARAATCA-3’) were used following Che et al. (2012). Gene fragments were amplified under the
following conditions: an initial denaturing step at 95 °C for 4 min; 36 cycles of de-
naturing at 95 °C for 30 s, annealing at 52 °C (for 16S)/47 °C (for COI) for 40 s and
extending at 72 °C for 70 s. Sequencing was conducted using an ABI3730 automated
DNA sequencer in Shanghai DNA BioTechnologies Co., Ltd. (Shanghai, China). New
sequences were deposited in GenBank (for GenBank accession numbers see Table 1).

For molecular analyses, the available sequences for congeners of *Megophrys* were
downloaded from GenBank (Table 1), primarily from previous studies (Chen et al.
2017; Liu et al. 2018). For phylogenetic analyses, corresponding sequences of one
*Leptobrachella oshanensis* (Liu, 1950) and one *Leptobrachium boringii* (Liu, 1945) were
also downloaded (Table 1), and used as outgroups following Mahony et al. (2017).
Sequences were assembled and aligned using the Clustalw module in BioEdit v.7.0.9.0
(Hall 1999) with default settings. Alignments were checked by eye and revised manu-
ally if necessary. For phylogenetic analyses of mitochondrial DNA, the dataset concat-
enated with 16S and COI gene sequences. To avoid under- or over-parameterization
(Lemmon and Moriarty 2004; McGuire et al. 2007), the best partition scheme and the
best evolutionary model for each partition were chosen for the phylogenetic analyses
using PARTITIONFINDER v. 1.1.1 (Robert et al. 2012). In this analysis, 16S gene
and each codon position of COI gene were defined, and Bayesian Inference Criteria
was used. As a result, the analysis suggested that the best partition scheme is 16S gene/
each codon position of COI gene, and selected GTR + G + I model as the best model
for each partition. Phylogenetic analyses were conducted using maximum likelihood
(ML) and Bayesian Inference (BI) methods, implemented in PhyML v. 3.0 (Guindon
et al. 2010) and MrBayes v. 3.12 (Ronquist and Huelsenbeck 2003), respectively. For
the ML tree, branch supports were drawn from 10,000 nonparametric bootstrap rep-
lies. In BI, two runs each with four Markov chains were simultaneously run for 50
million generations with sampling every 1,000 generations. The first 25% trees were
removed as the “burn-in” stage followed by calculations of Bayesian posterior prob-
abilities (BPP) and the 50% majority-rule consensus of the post burn-in trees sampled
at stationarity. Finally, mean genetic distance between *Megophrys* species based on un-
corrected $p$-distance model was estimated respectively on 16S and COI genes using
MEGA v. 6.06 (Tamura et al. 2013).

**Morphological comparisons**

Six adult males and one tadpole of the undescribed species were measured (Table 1
and Suppl. material 1). For comparisons, six adult male specimens of *M. kuatunensis*
were also measured (Supp. material 1). The terminology and methodology followed
Fei et al. (2009). Measurements were taken with a dial caliper to 0.1 mm. Twenty-two
morphometric characters of adult specimens were measured:

- **ED** eye diameter (distance from the anterior corner to the posterior corner of the eye);
- **FIL** first finger length (distance from base to tip of finger I);
- **FIIL** second finger length (distance from base to tip of finger II);
A new species of *Megophrys*

FIIL  third finger length (distance from base to tip of finger III);
FIVL  fourth finger length (distance from base to tip of finger IV);
FL    foot length (distance from tarsus to the tip of fourth toe);
HDL   head length (distance from the tip of the snout to the articulation of jaw);
HDW   maximum head width (greatest width between the left and right articulations of jaw);
HAL   hand length (distance from tip of third digit to proximal edge of inner palmar tubercle);
IND   internasal distance (minimum distance between the inner margins of the external nares);
IOD   interorbital distance (minimum distance between the inner edges of the upper eyelids);
LAL   length of lower arm and hand (distance from the elbow to the distal end of the Finger IV);
LW    lower arm width (maximum width of the lower arm);
SNT   distance between the nasal the posterior edge of the vent;
SVL   snout-vent length (distance from the tip of the snout to the posterior edge of the vent);
SL    snout length (distance from the tip of the snout to the anterior corner of the eye);
TFL   length of foot and tarsus (distance from the tibiotarsal articulation to the distal end of the Toe IV);
THL   thigh length (distance from vent to knee);
TL    tibia length (distance from knee to tarsus);
TW    maximal tibia width;
TYD   maximal tympanum diameter;
UEW   upper eyelid width (greatest width of the upper eyelid margins measured perpendicular to the anterior-posterior axis).

For the single tadpole of the undescribed species, eleven morphometric characters were measured:

BH    maximum body height;
BW    maximum body width;
IOS   interocular distance (minimum distance between eye);
MW    mouth width (distance between two corners of mouth);
SL    snout length (distance from the tip of the snout to the anterior corner of the eye);
SS    snout to spiraculum (distance from spiraculum to the tip of the snout);
SVL   snout-vent length;
TAH   tail height (maximum height between upper and lower edges of tail);
TAL   tail length (distance from base of vent to the tip of tail);
TBW   maximum width of tail base;
TOL   total length (distance from the tip of the snout to the tip of tail).
To reduce the impact of allometry, the correct value from the ratio of each character to SVL was calculated, and then was log-transformed for the following morphometric analyses. Mann-Whitney $U$ tests were conducted to test the significance of differences on morphometric characters between the undescribed species and $M. kuatunensis$. The significance level was set at 0.05. Furthermore, principal component analyses (PCA) were conducted to highlight whether the different species were separated in morphometric space.

The new species was also compared with all other $Megophrys$ species on morphology. Comparative data were obtained for related species as described in literature (Table 2).

Bioacoustics analyses

The advertisement calls of the undescribed species were recorded from the holotype specimen CIBQY20200726001 in the field on 26 July 2020 from Qingyuan County, Zhejiang Province, China. When registering the male in the stream the ambient air temperature was 21.5 °C and there was air humidity of 87%. For comparisons, the advertisement calls of $M. kuatunensis$ from Wuyi Mountain, Fujian Province, China were recorded from the specimens CIBWY18082410, CIBWY18082411 and CIBWY18082412 at an ambient air temperature of 22.0 °C and air humidity of 88% on 24 August 2018. SONY PCM-D50 digital sound recorder was used to record within 20 cm of the calling individual. The sound files in wave format were resampled at 48 kHz with sampling depth 24 bits. The sonograms and waveforms were generated by WaveSurfer software (Sjöander and Beskow 2000) from which all parameters and characters were measured. Ambient temperature was taken by a digital hygrothermograph.

Results

Phylogenetic analyses

Aligned sequence matrix of 16S+COI contains 1104 bp. ML and BI trees of the mitochondrial DNA dataset presented almost consistent topology, and as well, though relationships of many clades were unresolved (Fig. 2). In mitochondrial DNA trees, all samples of the undescribed species were clustered into one clade which was deeply clustered into the $Megophrys$ clade. The species is likely sister to $M. kuatunensis$ (bootstrap supports < 50% and BPP = 0.51) though the relationships between the two species and most other congeners were not resolved (all bootstrap supports < 50% and many BPP < 0.95).

Genetic distances based on 16S and COI genes with uncorrected $p$-distance model between the samples of the undescribed species were all below 0.2%. The genetic distance between the undescribed species and its closest related species $M. kuatunensis$ were 2.1% and 8.1% on 16S and COI respectively, which was higher or at the same level with those among many pairs of sister species, for example, 1.7% and 3.8% on 16S and COI respectively between $M. spinata$ and $M. sangzhiensis$ (Suppl. materials 2 and 3).
Table 2. References for morphological characters for congeners of the genus *Megophrys*.

| Species | Literature obtained |
|---------|---------------------|
| *M. aceras* Boulenger, 1903 | Boulenger 1903 |
| *M. acuta* Wang, Li & Jin, 2014 | Li et al. 2014 |
| *M. annecte* Mahony, Teeling & Biju, 2013 | Mahony et al. 2013 |
| *M. annecte* Wu, Suwannapoom, Poyarkov, Chen, Pawangkhanant, Xu, Jin, Murphy & Che, 2019 | Wu et al. 2019 |
| *M. auralesius* Ohler, Swan & Daltry, 2002 | Ohler et al. 2002 |
| *M. awadh Mahony, Kamei, Teeling, & Biju, 2020* | Mahony et al. 2020 |
| *M. baluensis* (Boulenger, 1899) | Boulenger 1899a |
| *M. baolongensis* Ye, Fei & Xie, 2007 | Ye et al. 2007 |
| *M. binchuanensis* Ye & Fei, 2001 | Fei et al. 2001 |
| *M. brachykelos* Inger & Romer, 1961 | Inger and Romer 1961 |
| *M. carinens* (Boulenger, 1889) | Boulenger 1889 |
| *M. caobangensis* Nguyen, Pham, Nguyen, Luong, & Ziegler, 2020 | Nguyen et al. 2020 |
| *M. cadoperta* Shen, 1994 | Shen. 1994 |
| *M. cheni* (Wang & Liu, 2014) | Wang et al. 2014 |
| *M. chihuansisi* Xu, Li, Liu, Wei & Wang, 2020 | Xu et al. 2020 |
| *M. chuannanensis* (Fei, Ye & Huang, 2001) | Fei et al. 2001 |
| *M. damrei* Mahony, 2011 | Mahony 2011 |
| *M. daweiensis* Rao & Yang, 1997 | Rao and Yang 1997 |
| *M. dongguanensis* Wang & Wang, 2019 | Wang et al. 2019b |
| *M. dringi* Inger, Stuebing & Tan, 1995 | Inger et al. 1995 |
| *M. dzukou* Mahony, Kamei, Teeling & Biju, 2020 | Mahony et al. 2020 |
| *M. edwardinae* Inger, 1989 | Inger 1989 |
| *M. elfina* Poyarkov, Duong, Orlov, Gogoleva, Vassilieva, Nguyen, Nguyen, Che & Mahony, 2017 | Poyarkov et al. 2017 |
| *M. fansipanensis* Tapley, Cutajar, Mahony, Nguyen, Dau, Luong, Le, Nguyen, Nguyen, Portway, Luong & Rowley, 2018 | Tapley et al. 2018 |
| *M. feae* Boulenger, 1887 | Boulenger 1887 |
| *M. feii* Yang, Wang & Wang, 2018 | Yang et al. 2018 |
| *M. flavipunctata* Mahony, Kamei, Teeling & Biju, 2018 | Mahony et al. 2018 |
| *M. gerti* (Ohler, 2003) | Ohler 2003 |
| *M. gigantica* Liu, Hu & Yang, 1960 | Liu et al. 1960 |
| *M. glandulosa* Fei, Ye & Huang, 1990 | Fei et al. 1990 |
| *M. hansi* (Ohler, 2003) | Ohler 2003 |
| *M. himalayana* Mahony, Kamei, Teeling & Biju, 2018 | Mahony et al. 2018 |
| *M. hoanglienensis* Tapley, Cutajar, Mahony, Nguyen, Dau, Luong, Le, Nguyen, Nguyen, Portway, Luong & Rowley, 2018 | Tapley et al. 2018 |
| *M. huangshanensis* Fei & Ye, 2005 | Fei and Ye 2005 |
| *M. hypsiglena* (Wang, Liu, Lyu, Zeng & Wang, 2017) | Wang et al. 2017a |
| *M. intermedia* Smith, 1921 | Smith 1921 |
| *M. jiangi* Liu, Li, Wei, Xu, Cheng, Wang & Wu, 2020 | Liu et al. 2020 |
| *M. jingdongensis* Fei & Ye, 1983 | Fei et al. 1983 |
| *M. jinggangensis* (Wang, 2012) | Wang et al. 2012 |
| *M. julianensis* Wang, Zeng, Lyu & Wang, 2019 | Wang et al. 2019b |
| *M. kalimantanensis* Munir, Hamidy, Matsui, Iskandar, Sidik & Shimada, 2019 | Munir et al. 2019 |
| *M. kobayashii* Malkmus & Matsui, 1997 | Malkmus and Matsui 1997 |
| *M. koni* Mahony, Foley, Biju & Teeling, 2017 | Mahony et al. 2017 |
| *M. kuansanensis* Pope, 1929 | Pope 1929 |
| *M. lancip* Munir, Hamidy, Farajallah & Smith, 2018 | Munir et al. 2018 |
| *M. leishanensis* Li, Xu, Liu, Jiang, Wei & Wang, 2018 | Li et al. 2018 |
| *M. lekaguli* Stuart, Chuyankern, Chan-ard & Inger, 2006 | Stuart et al. 2006 |
| *M. linnii* (Zhang, Li, Xiao, Li, Pan, Wang, Zhang & Zhou, 2017) | Zhang et al. 2017 |
| *M. linyue* Taylor, 1920 | Taylor 1920 |
| Species | Literature obtained |
|---------|---------------------|
| M. lini (Wang & Yang, 2014) | Wang et al. 2014 |
| M. lishuensis (Wang & Yang, 2014) | Wang et al. 2014 |
| M. longipes Bouleenger, 1886 | Bouleenger 1886 |
| M. major Bouleenger, 1908 | Bouleenger 1908 |
| M. mangshanensis Fei & Ye, 1990 | Fei et al. 2012 |
| M. massonensis Bourret, 1937 | Bourret 1937 |
| M. medogensis Fei, Ye & Huang, 1983 | Fei et al. 1983 |
| M. megacephala Mahony, Sengupta, Kamei & Biju, 2011 | Mahony et al. 2011 |
| M. microtoma (Bouleenger, 1903) | Bouleenger 1903 |
| M. minor Stejneger, 1926 | Stejneger 1926 |
| M. mirabilis Lyu, Wang & Zhao | Lyu et al. 2020 |
| M. montana Kuhl & Van Hasselt, 1822 | Kuhl and Van Hasselt 1822 |
| M. monticola (Günther, 1864) | Günther 1864; Mahony et al. 2018 |
| M. mufumontana Wang, Lyu & Wang, 2019 | Wang et al. 2019b |
| M. nankiangensis Liu & Hu, 1966 | Hu and Liu 1966 |
| M. nankunensis Wang, Zeng & Wang, 2019 | Wang et al. 2019b |
| M. nanningensis Lyu, Wang & Li, 1966 | Wang et al. 2019b |
| M. naustia (Schlegel, 1858) | Schlegel 1858 |
| M. nambhuanaeng Mahony, Kamei, Teeling, & Biju, 2020 | Mahony et al. 2020 |
| M. nbea Wang, Li & Zhao, 2014 | Wang et al. 2014 |
| M. ombrophila Messenger & Dahm, 2019 | Messenger et al. 2019 |
| M. omeimontis Liu, 1950 | Liu 1950 |
| M. oorecrpta Mahony, Kamei, Teeling & Biju, 2018 | Mahony et al. 2018 |
| M. oprpedion Mahony, Teeling & Biju, 2013 | Mahony et al. 2013 |
| M. orientalis Li, Lyu, Wang & Wang, 2020 | Li et al. 2020 |
| M. pachyrectus Huang, 1981 | Huang and Fei 1981 |
| M. palpebratipinna Bourret, 1937 | Bourret 1937 |
| M. parallela Inger & Iskandar, 2005 | Inger and Iskandar 2005 |
| M. parva (Bouleenger, 1893) | Bouleenger 1893 |
| M. periata Mahony, Kamei, Teeling & Biju, 2018 | Mahony et al. 2018 |
| M. popei (Zhao, Yang, Chen, Chen & Wang, 2014) | Zhao et al. 2014 |
| M. robusta Bouleenger, 1908 | Bouleenger 1908 |
| M. subrimal Tapley, Cutajar, Mahony, Chung, Dau, Nguyen, Luong & Rowley, 2017 | Tapley et al. 2017 |
| M. sanghiensis Jiang, Ye & Fei, 2008 | Jiang et al. 2008 |
| M. serchhipii (Mathew & Sen, 2007) | Mathew and Sen 2007 |
| M. shappingensis Liu, 1950 | Liu 1950 |
| M. shinumatsina Lyu, Liu & Wang | Lyu et al. 2020 |
| M. shihengensis Tian & Sun, 1995 | Tian and Sun 1995 |
| M. shihuangensis Wang, Deng, Liu, Wu & Liu, 2019 | Wang et al. 2019a |
| M. spinata Liu & Hu, 1973 | Hu et al. 1973 |
| M. stejneri Taylor, 1920 | Taylor 1920 |
| M. synoria (Stuart, Sok & Neang, 2006) | Stuart et al. 2006 |
| M. takensis Mahony, 2011 | Mahony 2011 |
| M. tuberogranulata Shen, Mo & Li, 2010 | Mo et al. 2012 |
| M. vegrandis Mahony, Teeling, Biju, 2013 | Mahony et al. 2013 |
| M. wawuensis Fei, Jiang & Zheng, 2001 | Fei et al. 2012 |
| M. wugongensis Wang, Lyu & Wang, 2019 | Wang et al. 2019b |
| M. wuhuanganensis Ye & Fei, 1995 | Ye and Fei 1995 |
| M. wuhanensis Ye & Fei, 1995 | Ye and Fei 1995 |
| M. xiangduensis Wang, Wu, Peng, Shi, Lu & Wu, 2020 | Wang et al. 2020 |
| M. xianguanensis Lyu, Zeng & Wang | Lyu et al. 2020 |
| M. yuanduensis Lyu, Zeng & Wang | Lyu et al. 2020 |
| M. zhangi Ye & Fei, 1992 | Ye and Fei 1992 |
| M. zunhebotoensis (Mathew & Sen, 2007) | Mathew and Sen 2007 |
In PCA for male group, the total variation of the first two principal components was 47.5%. On the two-dimensional plots of PC1 vs. PC2, the undescribed species was almost separated from *M. kuatunensis* (Fig. 3). The first two principal component axes could separate *M. kuatunensis* from the undescribed species mainly based on limb and head characteristics, namely, HDL, HDW, IND, FIL, FIIL and FL. The results of Mann-Whitney *U* tests indicated that in males, the undescribed species was significantly different from *M. kuatunensis* on UEW and TFL (*p*-values < 0.05; Table 3).
There were two differences in sonograms and waveforms of calls between the undescribed species and *M. kuatunensis* (Fig. 4; Table 4). Firstly, the undescribed species had slower call repetition rate than the latter (0.79 call/s in the former vs. 1.18 call/s in the latter). Secondly, the undescribed species had lower dominant frequency (3.19–3.38 kHz in the former vs. 3.38–3.75 kHz in the latter).

Based on the molecular phylogenetic analyses, morphological comparisons (Supp. material 4), and bioacoustics differences, the specimens from Qiangyuan County, Zhejiang Province, China represent a new species which is described as follows.

**Taxonomic accounts**

*Megophrys baishanzuensis* sp. nov.

http://zoobank.org/563EBE4E-45FF-4956-AB3B-70467B2D338E

Figs 4A, B, E, G, H, 5–8; Tables 1–4, Suppl. materials 1–4

**Holotype.** CIBQY20200726001 (Figs 4A, B, E, G, H, 5), adult male, from Baishanzu National Park, Qingyuan County, Zhejiang Province, China (27.76°N, 119.18°E, ca. 1537 m a.s.l.), collected by Bin Wang on 26 July 2020.
A new species of *Megophrys*

**Paratype.** Five adult males collected from the same place as holotype collected by Bin Wang. CIBQY20200719001-CIBQY20200719004 collected on 19 July 2020 by Bin Wang, and CIBQY20200726002 collected by Zhonghao Luo on 26 July 2020.

**Other material examined.** One tadpole (CIBQY20200719005; Fig. 7) collected by Bin Wang on 19 July 2020.

**Diagnosis.** *Megophrys baishanzuensis* sp. nov. is assigned to the genus *Megophrys* based on molecular phylogenetic analyses and the following generic diagnostic characters: snout shield-like; projecting beyond the lower jaw; canthus rostralis distinct; chest glands small and round, closer to the axilla than to midventral line; femoral glands on rear part of thigh; vertical pupils (Fei et al. 2009).

*Megophrys baishanzuensis* sp. nov. could be distinguished from its congeners by a combination of the following morphological characters: body size small (SVL 28.4–32.4 mm in males); vomerine teeth absent; tongue not notched behind; tympanum distinctly visible, oval; a small horn-like tubercle at the edge of each upper eyelid; two metacarpal tubercles distinctly visible in hand; toes without webbing; heels overlapping when thighs are positioned at right angles to the body; tibiotarsal articulation reaching the level to the middle of eye when leg stretched forward.

**Description of holotype.** (Figs 4A, B, E, G, H, 5). SVL 28.5 mm; head width larger than head length (HDW/HDL ratio ca. 1.3); snout obtusely pointed, protruding well beyond the margin of the lower jaw in ventral view; loreal region vertical and concave; canthus rostralis well-developed; top of head flat in dorsal view; eye large, eye diameter 46.0% of head length; pupils vertical; nostril orientated laterally, closer to snout than eye; tympanum distinct, 55.8% of eye diameter; vomerine ridges present and vomerine teeth absent; margin of tongue smooth, not notched behind.
Forelimbs slender, the length of lower arm and hand 47.0% of SVL; fingers slender, relative finger lengths: I < II < IV < III; tips of digits globular, without lateral fringes; subarticular tubercle distinct at the base of each finger; two metacarpal tubercles, prominent, oval-shaped, the inner one bigger than the outer one.

Hindlimbs slender, tibia length 46.5% times of SVL; heels overlapping when thighs are positioned at right angles to the body, tibiotarsal articulation reaching the middle of eye when leg stretched forward; tibia length longer than thigh length; relative toe lengths I < II < V < III < IV; tips of toes round, slightly dilated; subarticular tubercles absent on each toes; toes without webbing but with narrow lateral fringe; inner metatarsal tubercle oval-shaped; outer metatarsal tubercle absent.

Dorsal skin rough, several large warts scattered on flanks; a small horn-like tubercle at the edge of each upper eyelid; tubercles on the dorsum forming a X-shaped ridge, two dorsolateral parallel ridges on either side of the X-shaped ridges; an inverted triangular brown speckle between two upper eyelids; several tubercles scattered on dorsal, flanks and dorsal surface of thighs and tibias; supratympanic fold distinct.

Numerous granules scattered on ventrum; pectoral and femoral glands distinct; numerous white granules on outer thighs.

Coloration of holotype in life. (Fig. 5). Dorsal brown, several pink tubercles scattered on dorsal, an inverted triangular brown speckle between the eyes; X-shaped ridges...
A new species of *Megophrys*

Figure 6. Photos of the holotype CIBQY20200726001 of *Megophrys baishanzuensis* sp. nov. in life

A dorsal view  B ventral view  C lateral view  D ventral view of hand  E ventral view of foot.

on the dorsum brown, four dark transverse bands on the dorsal surface of the thigh and shank; ventral surface of body white with brown spots; two dark brown dark bars on the flanks, throat brown; white vertical bars on lower and upper lip; ventral surface of anterior limb dark reddish purple, posterior limb orange with numerous white granules; tip of digits pale grey; inner metatarsal tubercle and two metacarpal tubercles pinkish; soles uniform dark reddish purple; pectoral glands white.

**Coloration of holotype in preservation.** (Fig. 4A, B, E, G, H). Color of dorsal surface fades to taupe; the inverted triangular brown speckle between the eyes and brown X-shaped ridges on dorsum are more distinct; ventral surface greyish white; creamy-white substitutes the purple grey on tip of digits; the posterior of ventral surface of body, inner of thigh and upper of tibia fades to creamy-white.

**Variation.** Fig. 6. Measurements and basic statistics of adult specimens are presented in Tables 3 and Supp. material 1. All specimens were similar in morphology but some individuals different from the holotype in color pattern. In CIBQY2020200719001...
Table 3. Morphometric comparisons between the adult specimens of *Megophrys baishanzuensis* sp. nov. and *M. kuatunensis*. Units given in mm. See abbreviations for the morphological characters in Materials and methods section. P-value resulted from Mann-Whitney *U* test. Significant level at 0.05.

| Character | *Megophrys baishanzuensis* sp. nov. | *M. kuatunensis* | Mann-Whitney U value | P-value |
|-----------|------------------------------------|------------------|----------------------|---------|
|           | Male (N = 6)                        |                  |                      |         |
| SVL       | Ranging 28.4–32.4                   | Ranging 28.4–32.4| 13.000               | 0.423   |
|           | Mean ± SD 30.5 ± 1.8                | Mean ± SD 30.5 ± 1.8|                     |         |
| HDL       | 8.0–9.1                            | 8.0–9.1          | 6.000                | 0.055   |
|           | Mean ± SD 8.6 ± 0.4                 | Mean ± SD 8.6 ± 0.4|                     |         |
| HDW       | 9.3–10.5                           | 9.3–10.5         | 8.000                | 0.109   |
|           | Mean ± SD 10.2 ± 0.4                | Mean ± SD 10.2 ± 0.4|                     |         |
| SL        | 3.4–4.1                            | 3.4–4.1          | 16.000               | 0.749   |
|           | Mean ± SD 3.8 ± 0.3                 | Mean ± SD 3.8 ± 0.3|                     |         |
| SNT       | 1.5–2.6                            | 1.5–2.6          | 18.000               | 1.000   |
|           | Mean ± SD 2.0 ± 0.4                 | Mean ± SD 2.0 ± 0.4|                     |         |
| IND       | 3.1–3.7                            | 3.1–3.7          | 16.000               | 0.749   |
|           | Mean ± SD 3.4 ± 0.3                 | Mean ± SD 3.4 ± 0.3|                     |         |
| IOD       | 2.8–3.3                            | 2.8–3.3          | 6.000                | 0.055   |
|           | Mean ± SD 3.0 ± 0.2                 | Mean ± SD 3.0 ± 0.2|                     |         |
| UEW       | 2.3–3.0                            | 2.3–3.0          | 2.000                | 0.010   |
|           | Mean ± SD 2.6 ± 0.2                 | Mean ± SD 2.6 ± 0.2|                     |         |
| ED        | 3.7–4.0                            | 3.7–4.0          | 15.000               | 0.631   |
|           | Mean ± SD 3.8 ± 0.1                 | Mean ± SD 3.8 ± 0.1|                     |         |
| TYD       | 1.5–2.1                            | 1.5–2.1          | 16.000               | 0.749   |
|           | Mean ± SD 1.8 ± 0.2                 | Mean ± SD 1.8 ± 0.2|                     |         |
| LAL       | 13.4–14.6                          | 13.4–14.6        | 9.000                | 0.150   |
|           | Mean ± SD 14.1 ± 0.5                | Mean ± SD 14.1 ± 0.5|                     |         |
| HAL       | 6.6–7.9                            | 6.6–7.9          | 6.000                | 0.055   |
|           | Mean ± SD 7.1 ± 0.5                 | Mean ± SD 7.1 ± 0.5|                     |         |
| LW        | 2.2–2.7                            | 2.2–2.7          | 10.000               | 0.200   |
|           | Mean ± SD 2.4 ± 0.2                 | Mean ± SD 2.4 ± 0.2|                     |         |
| FIL       | 2.2–2.8                            | 2.2–2.8          | 17.000               | 0.873   |
|           | Mean ± SD 2.5 ± 0.2                 | Mean ± SD 2.5 ± 0.2|                     |         |
| FIIL      | 2.4–3.0                            | 2.4–3.0          | 12.000               | 0.200   |
|           | Mean ± SD 2.7 ± 0.2                 | Mean ± SD 2.7 ± 0.2|                     |         |
| FIHII     | 4.3–5.1                            | 4.3–5.1          | 10.000               | 0.200   |
|           | Mean ± SD 4.6 ± 0.3                 | Mean ± SD 4.6 ± 0.3|                     |         |
| FIVL      | 2.6–3.6                            | 2.6–3.6          | 15.000               | 0.631   |
|           | Mean ± SD 3.0 ± 0.4                 | Mean ± SD 3.0 ± 0.4|                     |         |
| THL       | 12.2–13.5                           | 12.2–13.5        | 10.000               | 0.200   |
|           | Mean ± SD 12.9 ± 0.5                | Mean ± SD 12.9 ± 0.5|                     |         |
| TL        | 12.8–14.9                           | 12.8–14.9        | 13.000               | 0.423   |
|           | Mean ± SD 13.9 ± 0.9                | Mean ± SD 13.9 ± 0.9|                     |         |
| TW        | 2.7–4.2                            | 2.7–4.2          | 13.000               | 0.423   |
|           | Mean ± SD 3.3 ± 0.5                 | Mean ± SD 3.3 ± 0.5|                     |         |
| TFL       | 17.8–20.4                           | 17.8–20.4        | 1.000                | 0.006   |
|           | Mean ± SD 19.4 ± 1.0                | Mean ± SD 19.4 ± 1.0|                     |         |
| FL        | 11.2–12.3                           | 11.2–12.3        | 13.000               | 0.423   |
|           | Mean ± SD 11.8 ± 0.4                | Mean ± SD 11.8 ± 0.4|                     |         |

the tubercles on the dorsum forming two > shaped, disconnected ridges (Fig. 6A); in CIBQY2020200719004 the tubercles on the dorsum forming a big and distinct X-shaped speckle (Fig. 6B); in CIBQY2020200719003 ventral surface of body grey with brown spots (Fig. 6C); in CIBQY2020200726002 ventral surface of body and limbs brownish red (Fig. 6D).

**Tadpole description.** Fig. 7. The tadpole CIBQY20200719006 (Fig. 7) was confirmed as *Megophrys baishanzuensis* sp. nov by molecular phylogenetic analyses. Measurements in mm. Stage 31. Body slender, body brownish black and tail pale brown, body height greater than tail height; dorsal fin arising behind the origin of the tail, the highest fin near mid-length, tapering gradually to the narrowly pointed tip; tail approximately 1.9 times as long as snout-vent length; tail height 13.6% of tail length; body width longer than body height (BW/BH 1.2); eyes large, lateral, nostril near eyes; spiracle on the left side of the body and distinct; oral disk terminal, lips expanded and directed upwardly into a umbelliform oral disk; flank of body brownish black with some white spots, tail fins lightly colored, with small white and black spots. TOL 22.7; SVL 8.7; BW 3.0; BH 2.7; SL 2.0; SS 4.0; IOS 1.8; TAL 14.7; TAH 2.2; TBD 1.5; MW 1.3.

**Advertisement call.** Fig. 4. The call description is based on recordings of the holotype CIBQY20200726001 (Fig. 4; Table 4) from a shrub leaf near the streamlet.
A new species of *Megophrys*

Call duration was 151.0–170.0 ms (mean 162.4 ± 5.7). Inter-call interval was 682.0–1869.0 ms (mean 936.8 ± 349.0). Pulse/call was 23.0–30.0 (mean 26.0 ± 2.4); pulse duration was 3.0–6.0 (mean 4.9 ± 6.0) and call repetition rate was 0.79 call/s.

Amplitude modulation within note was apparent, beginning with moderately high energy pulses, increasing to the maximum by approximately quarter, and then decreasing towards the end. The average dominant frequency was 3.36 ± 0.06 (3.19–3.38 kHz).

**Secondary sexual characters.** A single subgular vocal sac present in male. In breeding season, nuptial pads are present on the dorsal base of the first two fingers in males.

**Comparisons.** Supp. material 4. By having small body size, *Megophrys baishanzuensis* sp. nov. differs from *M. ancaea, M. auralensis, M. awuh, M. baluensis, M. baolongensis, M. binlingensis, M. boettgeri, M. caobangensis, M. carinense, M. caudoprocta, M. chishuiensis, M. chuannanensis, M. damrei, M. daweimontis, M. dzukou, M. edwardinae, M..
Table 4. Comparisons of characteristics of advertisement calls of *Megophrys baishanzuensis* sp. nov. and *M. kuatunensis*.

| Call character                | *Megophrys baishanzuensis* sp. nov. | *M. kuatunensis* |
|------------------------------|-------------------------------------|------------------|
|                              | CIBQY20200726001 | CIBWY2018082410 | CIBWY2018082412 | WY2018082411 |
| Number of call groups measured | 11                     | 30               | 30               | 20           |
| Number of notes measured      | 22                     | 30               | 30               | 40           |
| Call duration (ms)            | 151.0–170.0 (162.4 ± 5.7) | 131.0–163.0 (147.2 ± 7.1) | 131.0–163.0 (147.2 ± 7.1) | 130.0–159.0 (120.9 ± 5.9) |
| Call repetition rate (calls/s) | 0.79                  | 1.18             | 1.13             | 1.3          |
| Intercall interval (ms)       | 682.0–1869.0 (936.8 ± 349.0) | 404–1548.0 (687.3 ± 206.8) | 404–1548.0 (687.3 ± 206.8) | 350.0–733.0 (458.4 ± 87.1) |
| Pulses/call                  | 23.0–30.0 (26.0 ± 2.4) | 25.0–36.0 (30.0 ± 2.3) | 25.0–36.0 (30.0 ± 2.3) | 32.0–40.4 (35.7 ± 2.3) |
| Dominant frequency (kHz)      | 3.19–3.38 (3.36 ± 0.06) | 3.38–3.75 (3.46 ± 0.16) | 3.38–3.75 (3.46 ± 0.16) | 3.38–3.38 (3.38±0.01) |
| Pulse duration (ms)          | 3.0–6.0 (4.9 ± 0.6) | 3.0–6.0 (4.4 ± 0.7) | 3.0–6.0 (4.4 ± 0.7) | 3.0–6.0 (4.5 ± 0.6) |

*M. feae*, *M. flavipunctata*, *M. gigantica*, *M. glandulosa*, *M. hansi*, *M. himalayana*, *M. hoanglienis*, *M. huangshanensis*, *M. insularis*, *M. jiangi*, *M. jingdongensis*, *M. jinggangensis*, *M. kalimantanensis*, *M. kobayashii*, *M. lancip*, *M. lekaguli*, *M. liboensis*, *M. ligayae*, *M. lini*, *M. longipes*, *M. major*, *M. mangshanensis*, *M. medogensis*, *M. megacephala*, *M. mirabilis*, *M. montana*, *M. monticola*, *M. nasuta*, *M. obesa*, *M. omeimontis*, *M. orientalis*, *M. pachyprobistes*, *M. palpebralespinosa*, *M. parallela*, *M. parva*, *M. periosa*, *M. platyparietus*, *M. popei*, *M. sangzhienis*, *M. serchhipi*, *M. shangningensis*, *M. spinata*, *M. takensis*, *M. wawuensis*, and *M. xiangnanensis* (maximum SVL < 33.0 mm in the new species vs. minimum SVL > 34.0 mm in the latter).

By vomerine teeth absent, *Megophrys baishanzuensis* sp. nov. differs from *M. ancrae*, *M. baluensis*, *M. carinense*, *M. caudoprocta*, *M. chuannanensis*, *M. damrei*, *M. daoweimontis*, *M. dongguanensis*, *M. dazukou*, *M. fansipanensis*, *M. feae*, *M. flavipunctata*, *M. glandulosa*, *M. himalayana*, *M. hoanglienis*, *M. insularis*, *M. intermedia*, *M. jingdongensis*, *M. jinggangensis*, *M. jiulianensis*, *M. kalimantanensis*, *M. kobayashii*, *M. lancip*, *M. lekaguli*, *M. liboensis*, *M. ligayae*, *M. longipes*, *M. mangshanensis*, *M. maosonensis*, *M. medogensis*, *M. megacephala*, *M. montana*, *M. nankunensis*, *M. nanlingensis*, *M. nasuta*, *M. numbbumaeng*, *M. omeimontis*, *M. oreocrypta*, *M. orientalis*, *M. oropedion*, *M. pachyprobistes*, *M. palpebralespinosa*, *M. parallela*, *M. parva*, *M. periosa*, *M. platyparietus*, *M. popei*, *M. sangzhienis*, *M. serchhipi*, *M. shangningensis*, *M. spinata*, *M. takensis* *M. wawuensis*, and *M. xiangnanensis* (maximum SVL < 33.0 mm in the new species vs. minimum SVL > 34.0 mm in the latter).

By a small horn-like tubercle present at the edge of each upper eyelid, *Megophrys baishanzuensis* sp. nov. differs from *M. aceras*, *M. acuta*, *M. carinense*, *M. caudoprocta*, *M. chuannanensis*, *M. feae*, *M. gerti*, *M. hansi*, *M. intermedia*, *M. intermedia*, *M. jinggangensis*, *M. kalimantanensis*, *M. koui*, *M. lancip*, *M. liboensis*, *M. microstoma*, *M. montana*, *M. nasuta*, *M. orientalis*, *M. palpebralespinosa*, *M. platyparietus*, *M. popei*, *M. shangningensis*, *M. stejnegeri*, and *M. synoria* (vs. having a prominent and elongated tubercle in the latter).
A new species of *Megophrys* A new species of *Megophrys* 91

By tongue not notched behind, *Megophrys baishanzuensis* sp. nov. differs from *M. ancræ*, *M. baolongensis*, *M. binlingensis*, *M. boettgeri*, *M. carinense*, *M. cheni*, *M. chuan-nanensis*, *M. damrei*, *M. dringi*, *M. dzukou*, *M. fansipanensis*, *M. feae*, *M. feii*, *M. flavipunctata*, *M. gerti*, *M. glandulosa*, *M. hoanglienenis*, *M. huangshanensis*, *M. insularis*, *M. jiulianensis*. *M. jingdongensis*, *M. kalimantanensis*, *M. kuatunensis*, *M. liboensis*, *M. mangshanensis*, *M. maasonensis*, *M. medogensis*, *M. minor*, *M. nankiangensis*, *M. nanlin-genis*, *M. numbbumaeng*, *M. omeimontis*, *M. oropedion*, *M. pachyproctus*, *M. parallela*, *M. popei*, *M. robusta*, *M. sanzhiensis*, *M. shapingensis*, *M. shuichengensis*, *M. spinata*, *M. vegrandis*, *M. wawuensis*, *M. zhangi*, and *M. zunhebotoensis* (vs. notched behind in the latter).

By toes with narrow lateral fringes, *Megophrys baishanzuensis* sp. nov. differs from *M. angka*, *M. baolongensis*, *M. brachykolos*, *M. caobangensis*, *M. chishuiensis*, *M. damrei*, *M. daweimontis*, *M. dongguanensis*, *M. fansipanensis*, *M. feae*, *M. himalayana*, *M. hoanglienenis*, *M. huangshanensis*, *M. insularis*, *M. jiangi*, *M. jiulianensis*, *M. kaliman-tanensis*, *M. koui*, *M. leishanensis*, *M. lekaguli*, *M. lishuiensis*, *M. major*, *M. mangshanen-sis*, *M. medogensis*, *M. megacephala*, *M. microstoma*, *M. minor*, *M. nankunensis*, *M. obesa*, *M. ombrophila*, *M. oreocrypta*, *M. oropedion*, *M. pachyproctus*, *M. parva*, *M. periosa*, *M. shunhuangensis*, *M. takensis*, *M. tuberogranulata*, *M. wawuensis*, *M. wugongensis*, *M. wuliangshanensis* and *M. xianjvensis* (vs. lacking in the latter); and differs from *M. bin-chuanensis*, *M. boettgeri*, *M. carinense*, *M. cheni*, *M. chuan-nanensis*, *M. dringi*, *M. feii*, *M. gigantica*, *M. glandulosa*, *M. intermedia*, *M. jingdongensis*, *M. liboensis*, *M. lini*, *M. orientalis*, *M. palpebralespinosa*, *M. platyparietus*, *M. shapingensis*, *M. shuichengensis*, *M. spinata*, and *M. xiangnanensis* (vs. with wide lateral fringes in the latter).

By toes without webbing, *Megophrys baishanzuensis* sp. nov. differs from *M. brach-ykolos*, *M. carinense*, *M. flavipunctata*, *M. jingdongensis*, *M. jinggangensis*, *M. lini,
M. major, M. palpebralespinosa, M. popei, M. shuichengensis, and M. spinata (vs. at least one-fourth webbed in the latter).

By heels overlapping when thighs are positioned at right angles to the body, Megophrys baishanzuensis sp. nov. differs from M. actuta, M. brachykolo, M. dongguanensis, M. huangshanensis, M. kuatunensis, M. nankunensis, M. obesa, M. ombrophila, M. wushanensis, and M. wugongensis (vs. just meeting or not meeting in the latter).

By tibiotarsal articulation reaching to the level to the middle of eye when leg stretched forward, Megophrys baishanzuensis sp. nov. differs from M. daweimontis, M. glandulosa, M. lini, M. major, M. medogensis, M. obesa, M. sangzhiensis, and M. yangmingensis (vs. reaching the anterior corner of the eye or beyond eye or nostril and tip of snout in the latter); differs from M. mufumontana (vs. reaching tympanum in males and to the eye in females in the latter); and differs from M. chishuiensis (vs. reaching the level between tympanum and eye in the latter).

By having an internal single subgular vocal sac in male, Megophrys baishanzuensis sp. nov. differs from M. caudoprocta, M. shapingensis, and M. shuichengensis (vs. vocal sac absent in the latter).

The congeners M. boettgeri, M. lishuiensis, M. ombrophila, and M. xianjuensis all occur in Wuyi Mountains, Fujian Province and/or Zhejiang Province, China, and probably have sympatric distribution with Megophrys baishanzuensis sp. nov. (Fei et al. 2012; Wang et al. 2017b; Messenger et al. 2019; Wang et al. 2020). The new species can be distinguished from these species by a series of morphological characters as follows. The new species differs from M. boettgeri by body size smaller (adult males with 28.4–32.4 mm vs. adult males with 34.5–37.8 mm), and in breeding male nuptial pads present on the dorsal base of the first two fingers (vs. nuptial pad only on the first finger). The new species differs from M. lishuiensis by vomerine ridges present (vs. absent), toes with narrow lateral fringe (vs. without), and tibiotarsal articulation reaching the middle of eye when leg stretched forward (vs. reaching the range from tympanum to eye). The new species differs from M. ombrophila by heels overlapping when thighs are positioned at right angles to the body (vs. not meeting), vomerine ridges present (vs. absent), and toes with narrow lateral fringe (vs. without). The new species differs from M. xianjuensis by tibiotarsal articulation reaching the middle of eye when leg stretched forward (vs. reaching the range from tympanum to eye), and toes with narrow lateral fringe (vs. without).

Megophrys baishanzuensis sp. nov. is phylogenetically closest to M. kuatunensis. Megophrys baishanzuensis sp. nov. could be identified from M. kuatunensis distinctly by tibiotarsal articulation reaching the middle of eye when leg stretched forward (vs. reaching the range from tympanum to eye), heels overlapping when thighs are positioned at right angles to the body (vs. not meeting), tongue not notched behind (vs. notched feebly), the supratympanic fold more expanded in dorsal view and tympanum protruding (vs. concave), and having significantly lower ratios of UEW and TFL to SVL in males (all \(p\)-values < 0.05; Table 3). On call characters, the new species has slower call repetition rate (0.79 call/s in the new species vs. 1.18 call/s in M. kuatunensis), and has lower dominant frequency (3.19–3.38 kHz in the new species vs. 3.38–3.75 kHz in M. kuatunensis).
A new species of *Megophrys*  

**Distribution and habitat.** *Megophrys baishanzuensis* sp. nov. is known from the type locality, Baishanzu National Park, Qingyuan County, Zhejiang Province, China, at elevations between 1400–1600 m. The individuals of the new species were frequently found in the stream surrounded by evergreen broadleaved forests (Fig. 9). *M. boettgeri* was also found in the same stream.  

**Etymology.** The specific name *baishanzuensis* refers to the distribution of this species, Baishanzu National Park, Qingyuan County, Zhejiang Province, China. We propose the common name “Baishanzu horned toad” (English) and Bai Shan Zu Jiao Chan (百山祖角蟾, Chinese).  

**Discussion**  

Although *Megophrys baishanzuensis* sp. nov. superficially resembles *M. kuatunensis*, molecular phylogenetic analyses, detailed morphological comparisons and call data all proposed the distinct differences between them. Moreover, the breeding seasons of them are different. According to our surveys, the breeding season of *M. kuatunensis* is in April to May in Wuyi Mountain, Fujian Province, China. But in this season, we did not find any individual of *Megophrys baishanzuensis* sp. nov. in Qingyuan County,
Zhejiang Province. And, the breeding season of the new species should be later than June because in June, we only listened to the calls of one male in the type locality (< 10 °C), and, in late July, the males of the species started to call when the temperature was just higher than 18 °C (but we did not find any female individual and egg of it). Different call characteristics and breeding ecology most probably promoted separation of the two species.

During our several and extensive surveys, we only found fewer than 15 adult males of *Megophrys baishanzuensis* sp. nov., only in a small stream near the top of the mountain in Baishanzu National Park, Zhejiang Province, China, and even then, we did not find any female, and only found four tadpoles of this species. Obviously, the population of the new species is very endemic and small. Fortunately, this population is in a preserved area in Baishanzu National Park. Of course, we still should make a reinforced plan to preserve this area for this toad species.

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**Supplementary material 1**

**Table S1**

Authors: Bin Wang

Data type: morphological measurements

Explanation note: Measurements of the adult specimens of *Megophrys baishanzuensis* sp. nov. and *M. kuatunensis*. Units given in mm. See abbreviations for the morphological characters in Materials and methods section.

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Link: https://doi.org/10.3897/zookeys.1005.58629.suppl1
Supplementary material 2

Table S2
Authors: Bin Wang
Data type: genetic distance
Explanation note: Uncorrected $p$-distances between the *Megophrys* species on the 16S gene.
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Link: https://doi.org/10.3897/zookeys.1005.58629.suppl2

Supplementary material 3

Table S3
Authors: Bin Wang
Data type: genetic distance
Explanation note: Uncorrected $p$-distances between the *Megophrys* species on the COI gene.
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Supplementary material 4

Table S4
Authors: Bin Wang
Data type: morphological comparisons
Explanation note: Diagnostic characters separating *Megophrys baishanzuensis* sp. nov. from other species of *Megophrys*.
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