Proximate Phytochemical analysis of *Corallocarpus epigaeus* (Arn.) Cl rhizomes and *Vernonia anthelmintica* wild seeds

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**ABSTRACT**

The proximate phytochemical analysis of *Corallocarpus epigaeus* (Arn.) Cl rhizomes and *Vernonia anthelmintica* wild seeds carried out using standard reference methods. The result shown the seeds of *Vernonia anthelmintica* had significant values of ash content, moisture content, total solid content and extractives as compared to *C. epigaeus* rhizomes. The qualitative chemical test indicated that the presence of alkaloid, flavonoid, saponin, amino acids and proteins in *C. epigaeus* rhizomes extract while carbohydrate, flavonoid, saponin, amino acids and proteins were found in the *V. anthelmintica* seeds extracts. Among all the extract, aqueous extract of *V. anthelmintica* seeds contains highest amount of total flavonoid content followed by aqueous extract of *C. epigaeus* rhizomes. The presence of high secondary metabolites in the *C. epigaeus* (Arn.) Cl rhizomes and *V. anthelmintica* seeds wild seeds indicated that if the plant is subjected to further research such as identification and characterization of plant, bioactive compounds with strong biological activities may be isolated and novel compounds may also be identified.

**Keywords:** Ash value, extractive value, total flavonoids content.

**INTRODUCTION**

Because of the presence of phytochemical constituents, the medicinal plants are useful for both healing and curing of human diseases. More people around the world have turned to the use of medicinal plant products in the healthcare system in recent years. The growing need for alternative medicine has resulted in the use of natural plant remedies generating an immense need for knowledge on the properties and uses of medicinal plants; resulting in development of natural product markets and interest in conventional medicinal systems. It is necessary to properly integrate the modern scientific techniques and traditional knowledge. Therefore, the role of medicinal plants in the mainstream health care system in solving healthcare problems is becoming increasingly stressed. For this, it is important to establish standardization of herbal raw materials and herbal formulations using systematic and well-designed methodologies 1-2.

Known in Gujarat as the Jungalisuran, *Corallocarpus epigaeus* (Arn.) Cl. (Cucurbitaceae). It is distributed throughout tropical Africa, region of the Persian Gulf and India. The root has a bitter principle that is allied with bryonin. A phdroyxybenzoyl ester, named epigaeusyl ester, a sesterpene lactone, viz. corallocarpescalider, a pyridine carbayclic ester, designated as corallocarpeonyl ester. It has a bitter and sub-acidic taste and has alternative and laxative properties and is used in syphilis, rheumatism, later dysentery stages. *Vernonia anthelmintica* Willd, commonly known as Vakuchi; is a member of the Asteraceae family. It is an annual herb spread all over India it has been commonly used in the treatment of certain illnesses such as asthma, kidney disorders, inflammatory swelling, leucoderma, worm infection and seizures. The possession of various therapeutic activities was noticed, viz. Anti-inflammatory, anti-arthritic, antidiabetic, antibiotic, antibacterial, anti-cancer and many more 3.

Our research aims at standardizing the drugs as well as analyzing the presence or absence of different phytochemicals in both medicinal plants and justifying their use as a medicinal plant.
MATERIAL AND METHODS

Chemicals and reagents
Methanols, aluminum trichloride, quercetin, toluene, ethyl acetate, formic acid are used for the experiment.

Collection and authentication of plant material
Plant materials, such as rhizomes of C. epigaea (Arn.) Cl and seeds of *V. anethelmintica* willid were collected from rural area of Tirupati, India. The plants were identified and authenticated from Department of Botany, Sri Venkateswara University, Tirupati, India. A voucher specimen (1528 and 1124) of the plant was deposited in the department.

Extraction of plant material
About 50 g of defatted dried plant material of *V. anethelmintica* and *C. epigaea* were exhaustively extracted with methanol and water solvent using maceration method. After extraction, material was subjected to the filtration, filtrate was collected and subject to concentrate on water bath. Finally the percentage yields were calculated on air dried basis (% w/w).

Proximate analysis

**Determination of moisture content and total solid content:**
The percentage of moisture content for both raw plant materials has been calculated because any excess water in medicinal plant materials would promote microbial growth, presence of fungi or insects and cause degradation after hydrolysis. The percentage of moisture content in a previously dried and tared flat weighing container was measured gravimetrically in which 5 g of precisely weighted air-dried material was put in. The sample was dried in an oven at 100°C-105°C until 2 consecutive weighing did not differ by more than 5 mg. Total solid content was determined by the following formula:

\[
\text{Total solids (%) = 100 - Moisture (%)}
\]

**Determination of ash values:**
These are used to determine a crude drug’s consistency and purity, and to establish its identity. That medication contains definite amounts of inorganic radicals, such as phosphates, carbonates and sodium silicates, potassium, magnesium, calcium etc. Quantitative determination in terms of different ash values therefore helps in their standardization as well as in determining foreign inorganic matter that is present as impurity.

**Determination of Total ash:**
Four grams of the powdered material were precisely weighed and put in a crucible of silica which was previously ignited and tared. The material was spread out in an even layer and ignited by gradually raising the heat to a temperature of 500-600 °C until it was white, indicating the carbon absence. The material was refrigerated and measured in a desiccator. The total ash content in the air-dried material was calculated in mg / g.

**Determination of Acid-insoluble ash:**
In addition, 25 ml of hydrochloric acid was applied to the crucible containing complete ash, covered with a watch glass and gently boiled for 5 min. The watch glass was rinsed with 5 ml of hot water and added the liquid to the crucible. The insoluble matter was collected on an ashless filter paper and washed with hot water until neutral to the filtrate. The insoluble material left on the filter paper was transferred to the original crucible, dried on a hot plate, and ignited to weight constant. The residue was allowed to cool for 30 min in a suitable desiccator, then weighed without delay. The amount of acid-insoluble ash in air-dried material was measured in mg / g.

**Determination of extractive values:**
The extractive value gives an idea of the essence of the chemical constituents present in the crude drug; which is useful for determining components extracted from the solvent used for extraction as well as for evaluating a crude drug.

**Determination of water soluble extractives:**
5gm of powdered drug was taken in conical flask, and then 100ml of water was added. After that flasks were closed with the help of the cotton plug; the mixture was shaken after regular interval of time without touching the solution on to the cotton plug. The mixture was kept for 24 hrs (with regular shaking). After the period of 24 hrs the solution was filtered out by using whatman filter paper and collected the filtrate. Empty evaporating dish was taken and weighed the dish and noted down. Then 25ml of 5% solution of each of drug was taken in evaporating dish. Then the evaporating dish was heated until the damp mass was formed. Then cooled the evaporating dish and weighted it. Difference of evaporating dish containing damp mass and empty evaporating dish was taken and directly calculated the water soluble extractive value.

**Determination of alcohol soluble extractives:**
5gm of powdered drug was taken in conical flask, and then 100ml of alcohol (99%) was added. After those flasks were closed with the help of the cotton plug; the mixture was shaken after regular interval of time without touching the solution on to the cotton plug. The mixture was kept for 24 hrs (with regular shaking). After the period of 24 hrs the solution was filtered out by using whatman filter paper and collected the filtrate. Empty evaporating dish was taken and weighed the dish and noted down. Then 25ml of 5% solution of each of drug was taken in evaporating dish. Then the evaporating dish was heated until the damp mass was formed. Then cooled the evaporating dish and weighted it. Difference of evaporating dish containing damp mass and empty evaporating dish was taken and directly calculated the alcohol soluble extractive value.

**Qualitative phytochemical analysis**
Preliminary qualitative phytochemical analysis was carried out to identify the secondary metabolites such as alkaloids, carbohydrates, glycosides, tannins, and phenolic compounds, flavonoids, fixed oils, saponins, proteins and amino acids, and steroids present in the various alcoholic and aqueous extracts of *Corallocarpus epigaea* and *Vernonia anethelmintica*.

**Quantitative estimation of phytochemical constituents**

**Estimation of Total Flavonoids Content (TFC)**
Determination of TFC was based on measurement of the intensity of red color complex formed due to reaction between flavonoids and aluminum trichloride (AlCl₃). Briefly, the AlCl₃-methanolic solution (1 mL, 2% w/v) was mixed to various diluted extracts or standard (1 mL) and allowed to stand for 1 h at 25 ± 2°C before the absorbance was measured at λ_max 420 nm against blank using ultraviolet visible (UV-Vis) spectrophotometer (Shimadzu 1800, Kyoto, Japan). Extract samples were evaluated at a final concentration of 1 mg/mL. The TFC was compared to quercetin equivalent (QE) (mg/100 mg of extract) using
thergression equation $y = 0.040X + 0.009$, $r^2 = 0.999$, where $x$ is the absorbance and $y$ is the QE.

Thin layer chromatography (TLC)
Presence of flavonoid in *V. anthelmintica* and *C. epigaeus* drug were determination by using TLC study using quercetin as standard. The selected extracts were prepared with the respective solvent and made up to 10 ml in different test tubes. Then the extracts were taken in a capillary tube and it was spotted in preparative TLC plates coated with silica gel G. The plates were developed in TLC chamber previously saturated with toluene: ethyl acetate: Formic acid (5:4:1) solvent systems. The different spots developed in solvent system were identified by means of detecting agent and the $R_f$ value are correspondingly calculated.

RESULTS AND DISCUSSION
Aqueous and methanolic extract of *V. anthelmintica* showed 3.8% w/w and 3.4% w/w, percentage yield respectively while aqueous and methanolic extract of *C. epigaeus* showed 4.1% w/w and 2.3% w/w, percentage yield, respectively. Percentage yield of both plants are given in Table 1. The physico-chemical parameters are helpful in judging the purity and quality of the drug. The percentage of active principles in the plant is determined only in the dry condition. Hence the parameters determined for proximate analysis include moisture content, total solid content, ash value and extractive value content of the both drugs drug. Results of each parameter evaluated are given in Table 2. The present study of phytochemical analysis of two different extracts methanol and Aqueous of both plants has suggested the presence of various phyto compounds as shown in Table 3. Total flavonoids content (TFC) present in both of the plant extracts as quercetin equivalent (QE), respectively; have been determined using spectrophotometer. Aqueous extract of *V. anthelmintica* contain more amount of flavonoid content rather than methaolic extract. However in case of *C. epigaeus* methanolic extract does not contains flavonoids. Amount of TFC present is various extracts of both plants are given in Table 4. TLC of the extract proves that both of the plants extracts contain flavonoids (quercetin) along with other compounds (Table 5 and Figure 1).

### Table 1: Yield (%) of *Corallocarpus epigaeus* and *Vernonia anthelmintica* extracts

| Drug          | Methanolic extract | Aqueous extract |
|---------------|--------------------|-----------------|
| *Corallocarpus epigaeus* | 2.3% w/w            | 4.1% w/w        |
| *Vernonia anthelmintica*   | 3.4% w/w            | 3.8% w/w        |

### Table 2: Physiochemical analysis of crude drugs

| Parameter                  | *C. epigaeus* | *V. anthelmintica* |
|----------------------------|---------------|--------------------|
| Moisture content           | 0.47% w/w     | 0.68% w/w          |
| Total solid content        | 99.53% w/w    | 99.32% w/w         |
| Total ash                  | 2.83% w/w     | 3.67% w/w          |
| Acid insoluble ash         | 0.42% w/w     | 1.09% w/w          |
| Alcohol soluble extractive value | 5.2% w/w | 4.7% w/w          |
| Water soluble extractive value | 4.3% w/w | 3.9% w/w           |

### Table 3: Phytochemical screening of extracts of *Corallocarpus epigaeus* and *Vernonia anthelmintica* extracts

| Constituents | *C. epigaeus* | *V. anthelmintica* |
|--------------|---------------|--------------------|
|               | Methanol extract | Aqueous extract | Methanol extract | Aqueous extract |
| Alkaloids    | -              | +                 | -                | -              |
| Glycosides   | -              | -                 | -                | -              |
| Flavonoids   | -              | +                 | +                | +              |
| Saponins     | +              | +                 | +                | +              |
| Phenolics    | -              | -                 | -                | -              |
| Amino Acids  | +              | -                 | +                | +              |
| Carbohydrate | +              | +                 | +                | +              |
| Proteins     | +              | -                 | +                | +              |
| Diterpenes   | -              | -                 | +                | +              |

Where (-) = Negative, (+) = Positive
Table 4: Estimation of TPC and TFC

| Drug               | Methanolic extract | Aqueous Extract |
|--------------------|--------------------|-----------------|
| C. epigaeus        | -                  | 1.127           |
| V. anthelmintica   | 0.780              | 2.082           |

Table 5: Thin layer chromatography of extracts of C. epigaeus and V. anthelmintica

| Extract                        | Day Light | Short U.V. | Long U.V. |
|--------------------------------|-----------|------------|-----------|
| Quercetin                      | 0.51      | 0.51       | 0.51      |
| Aqueous extract of C. epigaeus  | -         | -          | 0.36      |
| Methanol extract of V. anthelmintica | 0.39 | 0.39       | 0.38      |
|                                | 0.51      | 0.48       | 0.51      |
|                                | 0.51      | 0.51       | 0.68      |
|                                |           |            | 0.75      |
| Aqueous extract of V. anthelmintica | 0.51 | 0.79       | 0.79      |
|                                | 0.62      | 0.57       | 0.57      |
|                                | 0.51      | 0.51       | 0.51      |
|                                | 0.49      | 0.49       | 0.49      |

CONCLUSION

From the result of proximate phytocological analysis, concluded that the V. anthelmintica and C. epigaeus plants were found to contain phytocomponents like alkaloids, flavonoids, saponins and tannins which are reported clinically to combat various diseases and disorders in human beings. This study prompted us for the identification, isolation, characterization and investigation novel bioactive compounds.

CONFLICTS OF INTEREST

No conflicts of interest

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