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A phase III, open-label, randomised multicentre study to evaluate the immunogenicity and safety of a booster dose of two different reduced antigen diphtheria-tetanus-acellular pertussis-polio vaccines, when co-administered with measles-mumps-rubella vaccine in 3 and 4-year-old healthy children in the UK

Robin Marlow a, Sherine Kuriyakose b, Narcisa Mesaros c, Htay Htay Han d, Richard Tomlinson e, Saul N. Faust f, Matthew D. Snape g, Andrew J. Pollard g, Adam Finn a

a Bristol Children’s Vaccine Centre, University of Bristol, Upper Maudlin Street, Bristol BS2 8AE, UK
b GSK, 5 Embassy Links, Cunningham Road, Bangalore 560052, India
c GSK, 20 Avenue Fleming, Wavre 1300, Belgium
d GSK, 2301 Renaissance Blvd, King of Prussia, PA 19406, United States
e Royal Devon & Exeter NHS Trust, Exeter EX2 5DW, UK
f Southampton NIHR Wellcome Trust Clinical Research Facility and Faculty of Medicine, University of Southampton and University Hospital Southampton NHS Foundation Trust, Southampton SO16 6YD, UK
g Oxford Vaccine Group, Department of Paediatrics, University of Oxford and the NIHR Oxford Biomedical Research Centre, Headington, Oxford OX3 9DU, UK

Aim: To evaluate the immunogenicity and safety of a reduced antigen diphtheria-tetanus-acellular pertussis-inactivated poliovirus (dTap-IPV B) vaccine (Boostrix-IPV, GSK) as a pre-school booster in 3–4 year old children as compared to dTap-IPV R (Repevax, Sanofi Pasteur), when co-administered with mumps-measles-rubella vaccine (MMRV).

Methods: This phase III, open label, randomised study was conducted in the UK between April 2011 and April 2012. Children due their pre-school dTap-IPV booster vaccination were randomised 2:1 to receive one of two different dTap-IPV vaccines (dTap-IPV B or dTap-IPV R) with blood sample for immunogenicity assessment just prior and one month after vaccination. Immune responses to diphtheria, tetanus and polio antigens were compared between the study vaccines (inferential comparison). In the absence of an accepted pertussis correlate of protection, the immunogenicity of dTap-IPV B vaccine against pertussis was compared with historical pertussis efficacy data (inferential comparison). Safety and reactogenicity of both study vaccines were evaluated.

Results: 387 children were randomised and 385 vaccinated: 255 in the dTap-IPV B group and 130 in the dTap-IPV R group. Prior to vaccination, 76.8% of children had anti-diphtheria and 65.5% had anti-tetanus titres above the protection threshold; for pertussis, the pre-vaccination seropositivity rate ranged between 18.1 and 70.6%. Both vaccines were immunogenic with 99.2–100% of children achieving titres above the pre-specified seroprotection/seropositivity thresholds. One serious adverse event not considered as causally related to the study vaccination by the study investigator was reported in the dTap-IPV B group.
1. Introduction

The timing of primary and booster doses of diphtheria (d), tetanus (T), acellular pertussis (ap) and inactivated poliovirus (IPV) varies widely across Europe [1]. Most countries give two or three doses in the first six months (m) followed by one at 12–18 m (termed 2 + 1/3 + 1 schedules) with a further booster before starting school. However the UK schedule just has three infant doses with no DTaP-IPV booster at 12–18 m; at this visit children already receive four injections (Haemophilus influenzae type B, pneumococcal conjugate, mumps-measles-rubella [MMR] and meningococcal B). By three to five years of age the protection gained from primary vaccinations in infancy is starting to wane [2] so the pre-school booster is given between three and a half and four years. At this age low dose diphtheria vaccines have been shown to induce adequate immune responses but with the advantage of lower rates of local side effects [3]. dTap-IPV (dTape-IPV R; Boostrix-IPV, GSK) is already used as a pre-school booster in many countries around the world but is licensed only from the age of four. The aim of this study was to generate evidence to support the use of dTap-IPV R in three to four year olds so it could potentially be used as a pre-school booster vaccine at this age. Thus we aimed to demonstrate that the immunogenicity of dTap-IPV R is not inferior to that of dTap-IPV B (Repevax, Sanofi-Pasteur), the vaccine in routine use at the time in the UK and which is approved for use in persons from three years of age upwards. In the absence of accepted correlate of protection for pertussis, the immunogenicity of dTap-IPV R was evaluated by comparison with historical pertussis efficacy data [4,5].

2. Methods

2.1. Study design and setting

We conducted an open-label, randomised, multicentre trial in five paediatric research centres in the UK (Bristol, Exeter, Oxford, Southampton and Taunton) and seven general practices. Ethical approval was obtained from the South West 2 Research Ethics Committee (NHS REC Ref: 10/H0206/43). The trial was registered with the European Clinical Trials Database (2009-012202-39) and ClinicalTrials.gov (NCT01245049).

2.2. Participants

Eligible participants were healthy children between three and less than five years of age who had had their previous vaccines on time as per national immunization program in the UK (three diphtheria, tetanus, pertussis and polio doses primary schedule completed before six months of age and first MMR vaccine before two years of age) but had not already received their routine preschool dTap-IPV booster. Children were excluded from participating if they had a known allergy to the vaccine components, known immunodeficiency, chronic use of steroids or were concurrently in another clinical trial. Full exclusion criteria are listed in online supplement (Supplementary Table 1). Families were recruited either using postal mailings through local Child Health Databases or from their general practices. Vaccination was postponed for any intercurrent febrile illness with axillary temperature ≥37.5 °C or other moderate to severe acute illness.

2.3. Study procedures

After initial contact and eligibility checking, the study comprised two visits. At the first, written informed consent was obtained from the parent/legal guardian. Children were then randomised using GSK's central Internet Randomisation system (SBIR) (using a block size of six and a minimisation procedure accounting for centre) and allocated to receive either dTap-IPV B (lots AC39B034B, AC39B026A, AC39B032A) or dTap-IPV R (lots DEXTA397AZ, DEXTA419AZ) in a 2:1 ratio as their pre-school dTap-IPV booster with both groups also receiving a dose of MMR booster (Priorix, GSK, Lots AMJRB892AZ, AMJRC160AZ, AD01B679C, AD01B801A, AD01B733B). After randomisation, the study was open label with both investigator and child’s parents aware of their allocation. A blood sample (2.5 mls) was drawn before vaccination. Vaccines were given intramuscularly dTap-IPV into the left deltoid, MMR into right deltoid, using 25 mm 23 G needles, respectively. Solicited local and general symptoms occurring within four days following vaccination were recorded in diary cards as were other (unsolicited) adverse events (AEs) occurring within 30 days of vaccination. Information on serious adverse events (SAEs) occurring at any time-point during the study was also collected. The second and final study visit was 30 days (range: 21–48 days) after the first visit and comprised a second blood sample and collection of diary cards.

2.4. Laboratory assays

All assays were performed at the laboratories of GSK Biologicals (Rixensart, Belgium) with laboratory staff blinded to the participant group. Antibodies against diphtheria toxoid (antidiphtheria), tetanus toxoid (anti-tetanus) and pertussis components (pertussis toxoid [PT], filamentous haemagglutinin [FHA]...
and pertactin [PRN]) were measured by enzyme-linked immunosorbent assays (ELISA) developed in-house. Specific IgG antibodies to measles, mumps and rubella were measured using commercially available ELISAs following the manufacturers’ instructions (Dade Behring, Germany). For both anti-diphtheria and tetanus thresholds for correlates of clinical protection were defined as 0.1 international units per millilitre (IU/ml), three times the lower quantification limit of the assays as previously demonstrated [6,7]. With no established serological correlates of protection for pertussis antigens [8–10], measles [11], mumps [12] or rubella, the thresholds for serological responses (positive: greater than or equal to) were arbitrarily defined as the lower limits of the assays: 5 ELU/ml (all pertussis antigens), 150 mIU/ml, 231 U/ml and 4 IU/ml, respectively. Seroconversion was defined as seropositivity in the relevant assay in participants seronegative before vaccination. Antibody titres against poliovirus types 1, 2 and 3 were determined by the manufacturer’s in-house virus micro-neutralisation test standardized according to WHO guidance [13] and were expressed as the reciprocal of the dilution resulting in 50% inhibition. Polio antibody titres greater than or equal to 1:8 dilution were considered seropositive and protective [14].

2.5. Statistics

2.5.1. Analysis of immunogenicity

The protocol pre-defined standard non-inferiority criteria [15]. For diphtheria and tetanus responses – the upper 95% confidence interval (CI) limit of the (DTaP-IPV B group minus DTaP-IPV R group) difference between the percentage of participants in the two groups with post-vaccine antibody concentrations above the predefined protective threshold was to be less than 10%; for poliovirus types 1, 2 and 3 – the upper 95% CI limit of the ratio of geometric mean titres (GMTs) (DTaP-IPV B group divided by DTaP-IPV R group) was to be less than or equal to two and for pertussis antigens – the upper 95% CI limit of the ratio of geometric mean concentrations (GMCs) (historic DTaP data [4,5] divided by DTaP-IPV group in this study) was to be less than or equal to 1.5.

Secondary objectives were to describe the immunogenicity of both study vaccines in terms of seroprotection/seropositivity rates and GMCs/GMTs for all antigens, prior to and one month after booster vaccination; the percentage of participants with booster response to the pertussis and polio antigens; to describe the immune responses to the MMR vaccine in terms of seroconversion rates against mumps, measles and rubella, one month after booster vaccination; and to assess the safety and reactogenicity of the study vaccines in terms of solicited symptoms, unsolicited AEs and SAEs.

The primary analysis was based on the cohort of participants who completed the study according to protocol (ATP) for analysis of immunogenicity. If, in any vaccine group, the percentage of vaccinated participants with serological results excluded from this ATP cohort was 5% or more, a second analysis of the total vaccinated cohort was to be performed to complement the ATP analysis.

Given the absence of serologic correlates of protection against pertussis, an immuno-bridging approach was used to assess immune responses to pertussis antigens, by extrapolating the efficacy of a vaccine against pertussis as demonstrated in infants to an older age group, as previously described [16].

2.5.2. Safety

Safety and reactogenicity of the study vaccines were assessed in terms of solicited symptoms (local and general), unsolicited symptoms and SAEs. The intensity of solicited symptoms was graded as mild, moderate or severe (defined in Supplementary Table 2) and assessment of causality by vaccination was assessed by investigators. Additionally, p values for the difference in proportion of participants reporting the solicited symptom, and solicited and unsolicited symptoms combined, were computed post hoc using the continuity adjusted chi-square method. However, the results should be interpreted with caution since there was no adjustment for multiplicity.

2.5.3. Sample size

It was calculated that 230 participants in the DTaP-IPV group and 115 in the DTaP-IPV group (a total of 345) would provide at least 90% overall power to reach conclusions on all the co-primary objectives simultaneously (with Bonferroni adjustment). Assuming a dropout rate of 10%, a total of 384 participants (256 participants in DTaP-IPV group and 128 in DTaP-IPV group) was chosen to ensure that a sufficient number of evaluable participants was available for inclusion in the per protocol cohort for analysis of immunogenicity. Statistical analysis was done by GSK with independent statistical verification carried out by KJC statistics (www.kjctstatistics.com).

3. Results

The study ran from April 2011 to April 2012. 387 children were enrolled and 385 were vaccinated: 255 in DTaP-IPV arm, 130 in DTaP-IPV group (Fig. 1). There were no significant differences in baseline characteristics in the ATP immunogenicity cohort: in both groups mean age at vaccination was 3.1 years with an equal proportion of genders and similar heritage (Table 2). The ATP immunogenicity cohort comprised 76.5%/73.8% (DTaP-IPV/dTap-IPV, respectively) of the total vaccinated cohort mainly due to missing serological data through difficulties in obtaining blood samples. As per protocol, an additional total vaccinated cohort immunogenicity analysis was performed but found no significant difference in primary or secondary outcomes – results are provided as an online appendix (Supplementary Tables 3–5).

3.1. Immunogenicity results

For the study’s primary inferential analyses; at 30 days post booster vaccination the non-inferiority criteria for the upper bounds of the 95% CI of differences between comparator vaccines

### Table 1
Comparison of vaccine components.

| Vaccine | Diphtheria toxoid (IU) | Tetanus toxoid (IU) | Pertussis antigens (µg) | Polio antigens (D-antigen units) | Adjuvant |
|---------|------------------------|---------------------|-------------------------|---------------------------------|---------|
|         | PT | FHA | PRN | Fimbriae 2/3 | 1 | 2 | 3 |                                   |         |
| DTaP    | 30 | 40  | 25  | 25  | 8  | –  | 40 | 8  | 32 | Aluminium phosphate: 0.33 mg Al³⁺ |
| DTaP−IPV | ≥2 | 20  | 2.5 | 5   | 3  | 5  | –  | 40 | 8  | 32 | Aluminium hydroxide and phosphate: 0.5 mg Al³⁺ |

IU, international units; PT, pertussis toxoid; FHA, filamentous haemaggglutinin; PRN, pertactin; DTaP−IPV, reduced diphtheria-tetanus-acellular pertussis-polio vaccine (Boostrix); DTaP−IPV, reduced diphtheria-tetanus-acellular pertussis-polio vaccine (Repevax); DTaP, diphtheria-tetanus-acellular pertussis vaccine (Infanrix).
were met for all component antigens (i.e., seroprotection levels for diphtheria and tetanus, and GMCs ratio for polio and pertussis antigens) (Table 3).

For the secondary analyses the pre booster titres (Table 4) showed that prior to boosting for all antigens a high proportion of children in both groups had antibody levels below the defined seroprotection/seropositivity thresholds; with 16.7–34.5% for diphtheria and tetanus and 30.4–40.6% for polio. For pertussis 29.4–81.9% of participants had IgG titres below the pre-determined optimal serological thresholds. For all antigens, 30 days post dTap-IPV booster vaccination the proportions of seroprotected/seropositive children had risen to >99% indicating adequate serological response. Comparing antibody GMC/GMT between the two study vaccines at one month post dTap-IPV vaccination (Table 4), marginally higher point estimates for diphtheria, tetanus, Polio3 and PRN after dTap-IPV R vaccination, and marginally higher point estimates for the Polio1, Polio2, PT and FHA after dTap-IPV B vaccination were observed.

Prior to boosting, evidence of antibodies persistence for the MMR vaccine (Table 4) was observed for at least 89.7% of children meeting the immunological criteria for measles, mumps or rubella in each group. One month after the MMR booster dose all children had antibody titres above the defined immunological criteria with no significant differences in GMTs between groups.

3.2. Safety results

In the four day post vaccination period, 216/255 (84.7%; dTap-IPV B) and 108/130 (83.1%; dTap-IPV R) (p = 0.79) of participants reported at least one solicited symptom or unsolicited AE. For both vaccines redness and pain were the most commonly reported. Severe (grade 3) symptoms were reported by a maximum of 11.0% (redness in the dTap-IPV B group) and 18.4% (redness in the dTap-IPV R group) (p = 0.08) of participants (Table 5).

During the 31-day (Days 0–30) post vaccination unsolicited reporting period, 88/255 (34.5%; dTap-IPV B) and 36/130 (27.7%; dTap-IPV R) (p = 0.22) of participants reported at least one AE of which 4.7% and 4.6% (p = 1.0) respectively were considered to be severe.

Diarrhoea and vomiting, reported for 9/255 (3.5%) participants were the most frequently reported unsolicited AE in the dTap-IPV B group while rash, reported for 6/130 (4.6%) participants was the most frequently reported unsolicited AE in the dTap-IPV R group.

One SAE was reported during the whole clinical study duration; pneumonia requiring hospitalisation reported for one participant in the dTap-IPV B arm. This was felt not to be causally related to vaccination by the investigator. No study participants withdrew from the study due to an AE or SAE.
According to protocol pre-post vaccination serology results as proportion above serological threshold and pre-post vaccination geometric mean concentrations/titres (descriptive analyses).

Table 3

| Vaccine antigen | Control data | dTap-IPV group | Control group minus dTap-IPV group % (95% CI) | Criteria for non-inferiority |
|-----------------|--------------|----------------|---------------------------------------------|----------------------------|
|                  | (N) | (N) | |
| Diphtheria       | 100 | (90) | 99.4 | (177) | 0.56% (–3.55; 3.14) | UL ≤ 10%
| Tetanus          | 100 | (90) | 98.3 | (176) | 1.70% (–2.43; 4.90) |
| Polio 1          | 1983.1 | (63) | 2175.6 | (131) | 0.91% (0.65; 1.28) | UL ≤ 2
| Polio 2          | 2168.7 | (61) | 2796.9 | (100) | 0.78% (0.54; 1.12) |
| Polio 3          | 4522.5 | (68) | 3468.8 | (126) | 1.30% (0.93; 1.84) |
| GFTs             |      |   |   |   |   |
| PT (IU/ml)       | 45.7 | (2884) | 69.8 | (203) | 0.65% (0.59; 0.72) | UL ≤ 1.5
| FHA              | 83.6 | (685) | 362.1 | (204) | 0.23% (0.20; 0.27) |
| PRN              | 112.3 | (631) | 148.5 | (204) | 0.76% (0.64; 0.89) |

The group difference in booster response to the diphtheria and tetanus antigens and the adjusted GMT ratios between groups for the poliovirus types 1, 2 and 3 antigens, one month post-booster vaccinations are based on the according-to-protocol cohorts. The GMC ratios between groups for the anti-PT, anti-FHA and anti-PRN antigens one month post-booster vaccination is based on the total vaccinated cohort. Control data represent the data from the dTap-IPV group for diphtheria, tetanus and polio types 1, 2 and 3 antigens, and from the DTaP-APV039 study for the pertussis antigens (PT, FHA and PRN). Bold values represent that the statistical criterion for non-inferiority was met. CI, confidence interval; GMC, geometric mean concentration; GMT, geometric mean titre adjusted for baseline titre; EL.U/ml, Enzyme-Linked Immunosorbent Assay units per millilitre; ED50, median effective dose; mIU/ml, milli-international units per millilitre; MMR, mumps-measles-rubella; N, number of participants with available results; PT, pertussis toxoid; FHA, filamentous haemagglutinin; PRN, pertactin; dTap-IPV, reduced diphtheria-tetanus-acellular pertussis-polio vaccine (Boostrix-IPV); dTap-IPV, reduced diphtheria-tetanus-acellular pertussis-polio vaccine (Repevax); DTaP, diphtheria-tetanus-acellular pertussis vaccine (Infanrix).

Table 4

According to protocol pre-post vaccination serology results as proportion above serological threshold and pre-post vaccination geometric mean concentrations/titres (descriptive analyses).

| Vaccine antigen | dTap-IPV group | dTap-IPV group | |
|-----------------|----------------|----------------|-----------------|
|                  | Pre (95% CI)  | Post (95% CI) | Pre (95% CI)  | Post (95% CI) |
| Diphtheria       | (>0.1 IU/ml) | 76.8 (69.9–82.8) | 100 (98.1–100) | 83.3 (74.0–90.4) |
| (GMC IU/ml)      | 0.228 (0.194–0.267) | 8.113 (7.259–9.068) |
| Tetanus          | (>0.1 IU/ml) | 65.5 (58.0–72.5) | 100 (98.1–100) | 70.0 (59.4–79.2) |
| (GMC IU/ml)      | 0.209 (0.173–0.253) | 6.787 (5.961–7.727) |
| Pertussis PT     | (>5 EL.U/ml) | 18.1 (12.7–24.7) | 100 (98.1–100) | 20.0 (12.3–29.8) |
| (GMC EL.U/ml)    | 3.4 (3.0–3.9) | 70.1 (62.2–79.0) |
| FHA              | (>5 EL.U/ml) | 64.4 (56.8–71.5) | 100 (98.1–100) | 70.6 (59.7–70.8) |
| (GMC EL.U/ml)    | 12.9 (10.0–16.6) | 358.3 (312.5–410.8) |
| PRN              | (>5 EL.U/ml) | 34.9 (27.8–42.4) | 99.5 (79.0–100) | 36.0 (32.4–40.5) |
| (GMC EL.U/ml)    | 4.3 (3.8–5.0) | 75.9 (67.3–84.6) |
| Polio 1          | (>8 ED50)   | 59.4 (51.3–67.1) | 99.4 (96.5–100) | 63.4 (51.9–74.3) |
| (GMC ED50)       | 12.8 (10.6–15.5) | 1783.1 (1612.4–2630.1) |
| Polio 2          | (>8 ED50)   | 68.6 (60.7–75.7) | 99.2 (95.6–100) | 66.9 (58.2–79.5) |
| (GMC ED50)       | 15.5 (12.8–18.8) | 1783.1 (1612.4–2630.1) |
| Polio 3          | (>8 ED50)   | 64.1 (56.0–71.6) | 99.4 (96.5–100) | 59.5 (47.9–70.4) |
| (GMC ED50)       | 15.4 (12.7–18.8) | 1783.1 (1612.4–2630.1) |
| Measles          | (<150 mIU/ml) | 97.5 (93.7–99.3) | 100 (97.3–100) | 97.4 (90.8–99.7) |
| (GMC mIU/ml)     | 2644.0 (2261.3–3091.6) | 3817.7 (3422.3–4258.8) |
| Mumps            | (>231 U/ml) | 89.7 (83.9–94.0) | 100 (97.3–100) | 90.8 (81.9–96.2) |
| (GMC U/ml)       | 1015.3 (906.8–1232.3) | 6801.9 (6155.0–7516.8) |
| Rubella          | (<4 IU/ml)  | 100 (97.7–100) | 100 (97.3–100) | 100 (95.3–100) |
| (GMC IU/ml)      | 66.5 (59.1–74.8) | 134.3 (120.7–149.4) |

CI, confidence interval; GMC, geometric mean concentration; GMT, geometric mean titre; ED50, median effective dose; mIU/ml, milli-international units per millilitre; MMR, mumps-measles-rubella; N, number of participants with available results; PT, pertussis toxoid; FHA, filamentous haemagglutinin; PRN, pertactin; U/ml, units per millilitre; dTap-IPV, reduced diphtheria-tetanus-acellular pertussis-polio vaccine (Boostrix-IPV); dTap-IPVs, reduced diphtheria-tetanus-acellular pertussis-polio vaccine (Repevax); pre, pre-booster vaccination blood sampling; post, post-booster vaccination blood sampling.

4. Discussion

This study compares the immunogenicity and AEs of two alternative dTap-IPV vaccines for use as a pre-school booster in the UK vaccine schedule. According to the primary outcomes, the study vaccine response was non-inferior to its comparator at one month post-vaccination – both vaccines boosting serological responses above accepted thresholds. There were no significant differences in solicited or unsolicited local reactions between the two vaccines, with mild redness and pain reported in 49.8–58.4% of cases.

In this study which followed the UK infant schedule in use at that time, we found that by the age of three and a half years, almost a third of children had antibody levels that had waned below desired levels for at least one vaccine component (Table 4) – reinforcing the force for a pre-school booster. This is in line with a previous study showing serological evidence of protection from UK primary course DTaP had substantially waned by the age of three and a half years [17].

For diphtheria, although post-booster all children had antibody levels above threshold, GMCs were marginally higher for
dTap-IPV<sub>R</sub> than dTap-IPV<sub>B</sub> but still 80-fold above the level considered protective. Due to the long half-life of these antibodies, this degree of boosting above the protective threshold has been shown to remain clinically effective for ten years, based on a mathematical model [18].

With no agreed immunological correlate of protection against pertussis, it was not appropriate to simply compare the antibody response between the two study vaccines because dTap-IPV<sub>R</sub> contains two additional pertussis antigens. Accordingly, our predefined primary analysis was to compare the immune responses to the booster dose of dTap-IPV<sub>R</sub> given in this study against historical immunogenicity data from the original study that demonstrated clinical effectiveness for this combination of antigens. This primary analysis showed non-inferiority. In a secondary analysis we compared the GMCs for the antigens common to both vaccines. All titres were significantly boosted but, as might be expected, with minor difference between the vaccines. GMCs for PT and FHA were higher for dTap-IPV<sub>B</sub> than dTap-IPV<sub>R</sub>, while the reverse was true for PRN. Clearly both vaccines used as boosters can provide a serological response. However even with adequate pertussis vaccine responses, data from the US have shown rapid waning of clinical protection (27% per year after the fifth dose of pertussis vaccine) [19] leading the US to introduce an adolescent booster dose for pertussis [20]. Further investigations would need to be done to determine if the additional pertussis antigen contained in dTap-IPV<sub>R</sub> resulted in any significant difference in the rate of waning between the vaccines.

The limitation of this study is that whilst randomised, because of the difference in the visual aspects of the study vaccines, the study was designed to be open label. This meant that for AE recording the parents were not blinded to the study arm – despite this there were no significant differences between AE outcomes. Since the study the UK infant vaccine schedule has changed with the introduction of the rotavirus and meningococcal B immunisation programmes. In response to rising rates of neonatal pertussis, in 2012 the UK introduced maternal dTap vaccination at 28–32 weeks gestation. This has been highly successful with significant reduction in the number of cases [21]. However there is some evidence that increased maternal titres may adversely affect early infant vaccine responses [22,23], increasing the importance of an effective pre-school booster.

In conclusion, we found dTap-IPV<sub>B</sub> to be non-inferior to dTap-IPV<sub>R</sub> with all children making an equivalent immune response after boosting indicative of protection being afforded at the same level as demonstrated now in the population as monitored by national surveillance systems. There were minor differences between vaccines for individual antigens but these are unlikely to be of any clinical significance.

**Trademark statement**

*Boostrix-IPV, Infanrix and Priorix* are trademarks of the GSK group of companies. *Repevax* is a trade mark of Sanofi-Pasteur.

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**Declaration of interest**

R. Marlow declares receiving conference travel expenses from GSK group of companies in 2014.

A. Finn received grants paid to his institution from Novartis (Novartis Vaccines Division was subsequently acquired by the GSK group of companies), GSK group of companies and Sanofi Pasteur MSD for studies outside the submitted work. Owing to his membership on the Joint Committee on Vaccination and Immunisation for the United Kingdom Department of Health, Adam Finn no longer gives lectures or undertakes advisory work for industry, either paid or unpaid.

M. Snape reports grants from GSK group of companies, Novartis, Johnson and Johnson, MedImmune where he acted as investigator for clinical vaccine studies; speaker fees from Novartis; and consultancy fees from MedImmune.

A.J. Pollard reports previous grants from Pfizer and Okairos in the past 36 months. His department received unrestricted educational grants from Pfizer/GSK/Astra Zeneca in July 2016 and Gilead/MSD/GSK/Astra Zeneca in June 2017 for a course on Infec- tion and Immunity in Children. A.J. Pollard is chair of the UK Department of Health’s (DH) Joint Committee on Vaccination and Immunization (JCVI) and the scientific advisory group on vaccines for the European Medicines Agency and is a member of the WHO’s...
SAGE; the views presented in this manuscript do not necessarily represent the views of DHJCV, EMA or WHO.

S.N. Faust reports grants from GSK group of companies; S.N. Faust has been advisory board member to vaccine and antimicrobial manufacturers (GSK, AstraZeneca, Pfizer, Novartis, Sanofi, Cubist Pharmaceuticals, Actelion Pharmaceuticals, Astellas, Merck).

M. Snape, A. Pollard, S.N. Faust and R. Tomlinson declare that all honoraria and fees were paid to the employing institution and no personal fee or honoraria of any kind were received at any time.

S. Kuriyakose and N. Mesaros declare they are employed by the GSK group of companies; N. Mesaros holds shares in the GSK group of companies. H.H. Han was a GSK employee at the time the study was conducted; she is now employee of Takeda Pharmaceuticals.

R. Tomlinson declares receipt of recruitment fees from GSK; all honoraria were paid to the employing institution (Royal Devon & Exeter NHS Trust) and no personal fee or honoraria of any kind were received at any time.

Authors’ contributions

R. Marlow, A. Finn, A.J. Pollard, S.N. Faust, M. Snape, S. Kuriyakose were involved in the conception and design of the study; R. Marlow, A. Finn, A.J. Pollard, M. Snape, S.N. Faust, R. Tomlinson, S. Kuriyakose, N. Mesaros and H.H. Han were involved in data acquisition, analysis or interpretation. R. Marlow produced the first draft of the manuscript, all authors revised the work critically, approved the final version to be published and take full accountability for all aspects of the work.

Appendix A. Supplementary material

Supplementary data associated with this article can be found, in the online version, at https://doi.org/10.1016/j.vaccine.2018.03.021.

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