Non-invasive measurement and imaging of tissue iron oxide nanoparticle concentrations \textit{in vivo} using proton relaxometry

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Abstract. Magnetic nanoparticles and microparticles can be found in biological tissues for a variety of reasons including pathological deposition of biogenic particles, administration of synthetic particles for scientific or clinical reasons, and the inclusion of biogenic magnetic particles for the sensing of the geomagnetic field. In applied magnetic fields, the magnetisation of tissue protons can be manipulated with radiofrequency radiation such that the macroscopic magnetisation of the protons precesses freely in the plane perpendicular to the applied static field. The presence of magnetic particles within tissue enhances the rate of dephasing of proton precession with higher concentrations of particles resulting in higher dephasing rates. Magnetic resonance imaging instruments can be used to measure and image the rate of decay of spin echo recoverable proton transverse magnetisation (R\(_2\)) within tissues enabling the measurement and imaging of magnetic particle concentrations with the aid of suitable calibration curves. Applications include the non-invasive measurement of liver iron concentrations in iron-overload disorders and measurement and imaging of magnetic particle concentrations used in magnetic hyperthermia therapy. Future applications may include the tracking of magnetically labelled drugs or biomolecules and the measurement of fibrotic liver damage.

1. Introduction

Magnetic iron oxide nanoparticles can occur naturally in biological tissue or can be introduced for scientific or clinical reasons \cite{1}. Although their concentrations and spatial distribution can be deduced in experiments on excised tissue or sacrificed animals, there are many circumstances in which it is desirable to measure and image their concentrations \textit{in vivo}. Magnetic resonance imaging (MRI) is based on the resonance of nuclei (typically protons, which are plentiful in biological tissues) with radiofrequency radiation in the presence of a static magnetic field \cite{2}. Since the rate of precession of tissue protons is proportional to the local magnetic field strength, local fluctuations in field strength induced by the presence of magnetic nanoparticles can enhance the rate at which the transverse magnetisation of tissue protons decays with time \cite{3, 4}. In many situations, magnetic nanoparticle concentrations in tissues are sufficiently high to be the main determinant of the tissue proton transverse relaxation rate (R\(_2\)), with higher concentrations resulting in higher values of tissue R\(_2\). The recent development of techniques for measuring and imaging R\(_2\) in various biological tissues using...
clinical magnetic resonance imagers makes such techniques readily available thus enabling measurement and imaging of magnetic nanoparticle concentrations once suitable calibration curves have been determined.

2. Pathological iron oxide nanoparticles
Iron overload disorders such as thalassaemia and hereditary haemochromatosis affect up to 0.5% of the world’s population. These disorders result in the accumulation of iron in the form of nanoparticles of iron(III) oxyhydroxide in organs of the body such as the liver, heart, spleen, and pancreas. Knowledge of the concentrations of these particles is often essential for appropriate management of iron overload conditions. The currently accepted and widely used method of measurement is chemical analysis of needle biopsy specimens of the liver. However, needle biopsy is an invasive procedure that carries a degree of risk and has associated sampling errors owing to the uneven distribution of iron within tissues.

We have developed a method of measuring and imaging $R_2$ within the liver [5]. An example $R_2$ image of the liver is shown in figure 1. $R_2$ is highly dependent on tissue iron concentration owing to spin-spin interactions between protons of water molecules and the iron oxyhydroxide nanoparticles within the tissue. The high degree of correlation between $R_2$ and tissue iron concentration has enabled the development of a method for non-invasively measuring and imaging liver iron concentrations (figure 2) [6]. The method has a demonstrated dynamic range of measurement of liver iron concentration from 0.3 to 42.7 mg Fe/g dry liver tissue. The large dynamic range and sensitivity and specificity of liver $R_2$ to needle biopsy measurements together with a demonstrated high degree of reproducibility on multiple magnetic resonance imaging units offers readily available non-invasive absolute liver iron concentration measurements to the clinical community.

![Figure 1. $R_2$ image of iron-loaded human liver. High $R_2$ corresponds to high iron concentration. The $R_2$ image of the liver is superimposed over a standard T₂ weighted image. Note that $T_2 = 1/R_2$.](image1)

![Figure 2. Correlation between liver iron concentration (LIC) measured in vivo using proton transverse relaxometry and LIC measured by needle biopsy in 105 human subjects (adapted from [6]).](image2)

Furthermore, there is promise that the ability to image liver iron concentrations may enable the detection of distorted liver architecture such as fibrosis and cirrhosis by virtue of the fact that these distortions result in altered patterns of iron oxyhydroxide nanoparticle deposition within the tissue (figure 3). Fibrosis and cirrhosis involve the proliferation of scar tissue within the liver. The scar tissue tends not to incorporate iron at the same concentrations as the liver cells and hence the presence of the scar tissue dilutes the macroscopic iron concentration in its vicinity [7].
Figure 3. $R_2$ images of liver with corresponding $R_2$ distributions for two chronic hepatitis C patients. (a) patient with zero fibrosis score; (b) patient with cirrhosis (high fibrosis score). Note the distortion of the $R_2$ distribution away from normal for the patient with cirrhosis.

3. Magnetic particles for hyperthermia therapy

Mapping of tissue $R_2$ can also be used to image concentrations of magnetic particles introduced into the body such as those used for magnetic hyperthermia therapy [8, 9]. Magnetic hyperthermia therapy involves the application of alternating magnetic fields to tissues containing magnetic particles that have been targeted to tumour tissue in order to generate heat in the particles. Raised tissue temperatures can help to kill cancer cells and have been shown to increase the effectiveness of radio- and chemo-therapy. A safety concern of the technique is ensuring that the concentration of ferrimagnetic material in healthy parts of the liver is low enough to avoid unwanted heating of healthy liver tissue. Hence imaging of magnetic particle distributions \textit{in vivo} may play a role in assessing the effectiveness of magnetic particle targeting of tumours before application of the ac magnetic field to heat the particles.
Figure 4. In vivo $R_2$-image of rabbit liver embolized with ferrimagnetic iron particles (taken from [8]).

Figure 5. Graph of mean $R_2$ against iron concentration for approximate 1 cm cubes of tissue dissected from livers of rabbits loaded with ferrimagnetic particles (taken from [8]). Vertical bars through each data point indicate the width of the Gaussian distribution of $R_2$ observed for each sample of liver.

Figure 4 shows an $R_2$ image of a rabbit liver embolized with ferrimagnetic particles obtained in vivo. Figure 5 shows the calibration curve relating liver $R_2$ to concentration of the ferrimagnetic particles obtained by dissecting the excised liver into approximately 1 cm sized cubes post mortem and both chemically assaying for iron and measuring $R_2$ for each cube. Studies of heating rates and ferrimagnetic particle concentrations may lead to the $R_2$ images being used to predict local heating rates within the liver.

4. Ferrimagnetic particle detection limits
Since there are many situations where the concentrations of ferrimagnetic particles may be somewhat lower than those encountered in, for example, magnetic hyperthermia therapy, the limit of detection of ferrimagnetic particles by clinical magnetic resonance imaging systems has been investigated [10]. Such situations may include the use of nanoparticle “labels” within the human body. Magnetic nanoparticles functionalised with antigens for biomolecules of interest could be introduced into the body in order to magnetically label the biomolecules. Measurements on agar gels loaded with different concentrations of ferrimagnetic iron oxide nanoparticles indicate that ferrimagnetic iron concentrations as low as 50 $\mu$g/ml can be measured and imaged [10]. Although this detection limit is not sufficiently low to enable measurement and imaging of the reported biogenic magnetite particle concentrations in the human brain [11, 12], the use of MRI microscopy systems with much higher field gradients than those available with clinical systems may aid in the study of these particles in excised human brain samples. MRI microscopy might also play a role in elucidating the distribution of magnetic particles in the geomagnetic sensory organs of animals capable of magnetic navigation.

5. References
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