Brain-derived neurotrophic factor is increased in serum levels of patients with symptomatic dermographism

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Abstract
Introduction: Symptomatic dermographism (SD) is the most common form of physical urticaria. However, the role of neuroimmune mechanisms in SD is unclear.
Aim: To investigate circulating levels of brain-derived neurotrophic factor (BDNF) in symptomatic dermographism.
Material and methods: Thirty-two patients suffering from SD and 33 healthy subjects were included in the study. Serum BDNF levels were determined by a human enzyme-linked immunoassay (ELISA) kit. The serum C-reactive protein concentration and eosinophil counts in peripheral blood were evaluated.
Results: The BDNF serum levels were detectable in both SD patients and healthy controls. The BDNF serum levels were significantly higher in patients with SD compared with healthy controls ($p = 0.004$). There was a positive but weak correlation between serum BDNF and CRP levels; it was not statistically significant ($r = 0.211$, $p = 0.255$). There was also a positive but weak correlation between serum BDNF and eosinophil counts; it was not statistically significant ($r = 0.271$, $p = 0.141$).
Conclusions: The present study shows that BDNF is increased in the serum levels of patients with SD, suggesting a role for BDNF in the pathophysiology of this disorder.
Key words: symptomatic dermographism, brain-derived neurotrophic factor, neurotrophin.
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Neurotrophins have been shown to play an important role in the development and maintenance of cutaneous innervation. There is also evidence for non-neurotrophic functions of neurotrophins in the skin. These include the regulation of epidermal proliferation and apoptosis, control of hair follicle development and cycling, and melanogenesis [5, 6].

Aim

The role of neuroimmune mechanisms in SD is not yet clear. Thus we first investigated whether BDNF serum levels in SD are different from control subjects. Secondly, we analysed whether the relationship between serum BDNF, C-reactive protein (CRP), and eosinophil counts.

Material and methods

This study was conducted at the Turgut Ozal University Hospital (Ankara, Turkey). The study was approved by the local ethics committee. All participants gave their written informed consent. Thirty-two patients suffering from SD and 33 healthy subjects were included in the study. Inclusion criteria were: aged 18 years or older; at least a 3-month history of SD; and no history or presence of systemic or allergic diseases. Patients with SD associated with chronic idiopathic urticaria were excluded. In the same time period, age- and sex-matched non-atopic healthy subjects were recruited as controls. The diagnosis of SD was confirmed by stroking the skin of the back with a tongue blade. This procedure elicited within minutes a linear wheal and flare with itching and burning sensations.

Venous blood samples were drawn from the subjects between 08.00 and 10.00 h following a 12-h fasting period, and sera were isolated and stored at −80°C until analysis. Serum BDNF levels were determined by a human enzyme-linked immunoassay (ELISA) kit (catalog number EK0307, Boster Immunoleader, Boster Biological Technology, Ltd., Pleasanton, California, USA) according to the manufacturer’s instructions. Serum BDNF levels were expressed as pg/ml and the minimum detectable level of BDNF is typically 2 pg/ml. The serum CRP concentration was measured by the spectrophotometric method (Roche Hitachi Cobas C501, Tokyo, Japan). An elevated serum CRP level was defined as higher than 5.0 mg/L. Eosinophil counts in peripheral blood were evaluated using a Coulter LH70 counter (Beckman Coulter, Brea, California, USA).

Statistical analysis

Mean ± standard deviation (SD), median and minimum, maximum values are used to describe the quantitative variables. Also, frequency and percentages are given for the nominal data. Normality assumption was checked by Shapiro-Wilk test. The independent sample t-test and the Mann-Whitney U-test are used to compare the groups in terms of age and BDNF, respectively. The distribution of gender within study groups was analyzed by χ² test. Nonparametric Spearman correlation analysis was used to analyze the relationship between BDNF and blood total eosinophil count (TEC). For all analyses the IBM-SPSS version 21.0 was used and the statistical significance was set at p < 0.05.

Results

The mean age for the patient group was 34.7 ±11.03 (22 females, 10 males), and for the control group, the mean age was 33.48 ±10.5 (23 females, 10 males) (Table 1). The BDNF serum levels were detectable in both SD patients and healthy controls. The BDNF serum levels were significantly higher in patients with SD compared with healthy controls (p = 0.004) (Figure 1). The median serum CRP concentration was 1.9 mg/L. There was a positive but weak correlation between serum BDNF and CRP levels but it was not statistically significant (r = 0.211, p = 0.255) (Figure 2). The total eosinophil count was 147 cell/mm³. There was a positive but weak correlation between serum BDNF and eosinophil counts but it was not statistically significant (r = 0.271, p = 0.141) (Figure 3).

![Figure 1. Distribution of brain-derived neurotrophic factor (BDNF) values in patients and controls](image-url)

**Table 1.** Demographic data and serum BDNF levels of patients with symptomatic dermographism and controls

| Variable            | Patients (n = 32) | Controls (n = 33) |
|---------------------|------------------|------------------|
| Gender (female/male)| 22/10            | 23/10            |
| Age                 | 34.7 ±11.03      | 33.48 ±10.5      |
| Serum BDNF [pg/ml]  | 144.37 ±67.7     | 112.19 ±79.9     |
Discussion

Symptomatic dermographism is a distressing condition. The disease is frequently idiopathic though it may have an immunological basis in some patients [1, 3, 4]. Passive transfer of the dermographic response with IgE- or immunoglobulin M (IgM)-containing serum has been reported, but no allergen has been identified [8]. The pathogenesis remains unclear but trauma may release an antigen that interacts with the membrane-bound immunoglobulin E (IgE) of mast cells, thus releasing inflammatory mediators, particularly histamine, into the tissues. This process causes small blood vessels to leak, allowing fluid to accumulate in the skin [3, 4].

Neurotrophins exert immunomodulatory functions not only on nerves but also on immune cells [5, 6]. Bonini et al. were the first to investigate a role of neurotrophins in patients with urticaria. They found increased levels of nerve growth factor (NGF) in urticaria patients compared with healthy controls [9]. Özeker et al. found that NGF and BDNF levels were not increased in urticaria [10]. In another study, it was demonstrated that BDNF levels of serum and skin were increased in patients with chronic spontaneous urticaria compared with non-atopic control subjects with healthy skin [11]. In this study, we found higher serum levels of BDNF in patients with SD compared with non-atopic healthy control subjects. Some studies have suggested that increased levels of NGF and BDNF and a correlation in disease activity are indicators in chronic inflammatory skin diseases including atopic dermatitis [12–14]. Thus neurotrophins seem to play a role (encompassed) in the pathogenesis of several atopic and non-atopic inflammatory skin diseases.

In the literature, there are no detailed or classified data on triggering factors and/or associated diseases in SD. Taşkapan et al. found that psychological factors and a history of stressful life events have been implicated as triggering factors in 30% of patients [15]. Anlíker et al. detected that the psychological status (accidents, fear of life, surgical intervention, chronic pain, and others) were defined in about 90% of patients with SD [16]. In another study, Schoepke et al. found that 44% of patients claimed stress to be the reason for SD [17]. Stress increases mast cell histamine release. Mast cells and their endogenous products are strongly implicated not only in the classical immune processes but also in neuropsychiatric and neuropathologic events [18]. BDNF is involved as a pro-inflammatory factor [5, 6]. Schulte-Herbrüggen et al. demonstrated that BDNF can be produced by human blood monocytes and that this production is up-regulated by inflammatory mediators as tumor necrosis factor α (TNF-α) and interleukin-6 (IL-6) [19]. Also BDNF may be released in response to stroking, rubbing and scratching of the skin of patients with SD in peripheral nerves and may serve as local stress responders that mediate neurogenic inflammation.

In this study we found a correlation between serum BDNF levels and eosinophil count to be positive but not statistically significant. Because eosinophils are a main player in allergic inflammation, neurogenic inflammation may be an important pathway triggering inflammation in the skin. These findings suggest that eosinophil is not a source of increased BDNF in the serum of SD patients and that the pathophysiology of the disease is not characterized by allergic inflammation. Skin biopsy studies should be used to evaluate the roles of eosinophil and BDNF.
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The results presented in this study demonstrate that there was a positive but weak correlation between serum BDNF levels and CRP levels but the correlation was not statistically significant. Neurotrophin’s pivotal role has been described in several inflammatory conditions [9, 20]. Shin et al. showed that plasma BDNF levels correlated positively with inflammatory cytokines in hemodialysis patients, suggesting that plasma BDNF might reflect uremic inflammation in patients undergoing hemodialysis [21]. In our study, we did not observe any significant associations between levels of serum BDNF and CRP.

Conclusions

Our study suggests that SD is associated with alterations of serum BDNF levels. Further studies will be needed to address the functional role of BDNF in this disorder to establish new therapeutic implications.

Conflict of interest

The authors declare no conflict of interest.

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