Lamb Fattening Under Intensive Pasture-Based Systems: A Review

Gonzalo Fernandez-Turren 1,*, José L. Repetto 1, José M. Arroyo 1,2, Analía Pérez-Ruchel 1 and Cecilia Cajarville 1

1 Departamento de Producción Animal, IPAV, Facultad de Veterinaria, Universidad de la República, Ruta 1 km 42,5 km, San José 80100, Uruguay; joselorepetto@gmail.com (J.L.R.); chemaarroyo@gmail.com (J.M.A.); anapevet@gmail.com (A.P.-R.); ccajarville@fvet.edu.uy (C.C.)

2 Programa Nacional de Investigación en Pasturas y Forrajes, Instituto Nacional de Investigación Agropecuaria, La Estanzuela 70002, Colonia, Uruguay

* Correspondence: gonzalofernandezt@gmail.com; Tel.: +598-4030-7023

Received: 27 January 2020; Accepted: 14 February 2020; Published: 27 February 2020

Simple Summary: The use of fresh high-quality pastures in intensive lamb production systems is considered worldwide as economically advantageous, environmentally friendly, and a promoter of animal welfare. Moreover, it generates a desirable meat composition. However, it is known that the availability of pastures in grazing regions is variable throughout the year, and this makes the maintenance of a stable offer of feeds and production difficult. The combination of high-quality pastures and other feedstuffs is very common in dairy cow grazing systems while, for sheep, there is less information available. The objective of the current review is to discuss this topic in light of published information about intake, digestion, rumen environment and health, performance, and carcass quality and composition.

Abstract: The benefits of pasture-based systems on the fatty acid composition of sheep meat appear to be achievable despite variability in the quality of the pastures. Lambs fed high levels of temperate pastures have an excess of N-ammonia derived from protein degradation. Furthermore, animal performance is highly variable depending on the quality of the pasture at the time of grazing, and high animal performance in these systems appears to be linked to the use of high-quality pastures with high availability, and is possibly added to by the inclusion of concentrates that allow increasing energy intake and a better use of the N in the pasture. The combination of high-quality pastures and total mixed ration offers a good alternative to the inclusion of concentrates in the diet, improving the use of N, and avoiding acidosis problems. However, information to determine the effect of a number of nutritional strategies on meat quality, and the minimum level of pasture intake necessary to achieve the benefits of pastoral systems is still lacking.

Keywords: sheep; fresh forage; intake; rumen; digestion; performance; meat quality

1. Introduction

The meat produced on pasture-based systems is recognized as a high-quality product, with positive nutritional contributions to human health [1–3]. In addition, systems that allow animals to graze have a better social perception, as they are associated with natural attributes, environmental care, and animal welfare, which also offer new opportunities for pasture-based systems [4–7]. For fattening lambs, these systems are developed mainly on grasses (Poaceae) and legumes (Fabaceae). They extend across tropical and temperate regions, but the quality of the forage differs between them. Temperate pastures are, in general, considered to be of high quality, due to the higher digestibility and lower fiber
content of temperate compared with tropical species [8]. The most commonly-cultivated pastures include grasses (e.g., *Agrostis* spp., *Festuca* spp., *Lolium* spp., and *Dactylis* spp.) and herbaceous legumes (e.g., *Lotus* spp., *Medicago* spp., and *Trifolium* spp.) [9]. This review will be focused on the use of these type of pastures.

Temperate-cultivated pastures are usually used for intensive grazing or grazing plus supplementation. Compared with sheep, there is abundant information on the use of nutrients for beef cattle and dairy cows in these rearing conditions [10]. Although sheep have been used as an experimental rumen model for feed evaluation and feeding studies, the information on diets specifically designed for sheep is not as abundant as that for other ruminants (dairy and beef cattle), which may be because of the variety of conditions in which sheep are raised. However, the digestive physiology of sheep is quite different from cattle; mainly associated with differences in digesta retention times, rumen digestion, and fermentation characteristics [11–13], so the use of feeding strategies designed for beef or dairy cattle is not recommended for sheep nutrition. For example, Aguerre et al. [13] observed in animals consuming lotus (*Lotus corniculatus*), that increasing supplementation with sorghum grain from 1% to 1.5% of body weight (BW) was effective in increasing feed intake and digestive use in cattle. Nevertheless, the same levels of supplementation in sheep resulted in excessive ruminal fermentation that reduced fiber digestibility and intake. In the same way, van Gastelen et al. [11] highlighted the caution needed before extrapolating results of CH$_4$ mitigation, mainly due to differences between dairy cattle, beef cattle, and sheep.

Grazing sheep occupy an acreage that may be limited [14,15] in order to increase areas of native fauna and flora protection. It is also imperative to reduce greenhouse gas emission intensity (CH$_4$ and N$_2$O released per kg of meat or milk produced), and therefore, it is necessary to increase productivity on pastures worldwide. Low dry matter (DM) intake has been identified as a main constraint of pasture-based systems for high-production animals [16]. In turn, the growth of lambs is conditioned to climatic conditions and excellent grass management is required. Combining pastures with other feedstuffs will allow for the overcoming of restrictions of grazing systems, taking advantage of the benefits of both pasture and confinement.

### 2. The Advantages of Pastures on Meat Characteristics

Recent reviews have been published reporting information on the impact of grazing on the quality of lamb meat [17,18]. Fattening lambs to pasture could be considered as an alternative to produce high quality lamb meat [19], but it is not clear what would be the minimum forage inclusion level at which we could observe improvement the quality of the meat, without limiting the growth, with respect to the lambs reared with concentrates. The demand for healthier and environmentally sustainable meat products has stimulated consumers’ interest in more extensive systems [20]. Lower fat content is, in general, characteristic of the meat of animals fattened on pastures, and is one of the points that has raised more interest. In general, lambs fed on concentrates produce meat with higher fat content than those fed forage-based diets at the same slaughter weight [21–23]. On the other hand, it is necessary to point out that fatty acid composition plays an important role in the definition of meat quality, as it is related to the nutritional value of fats for human consumption—these fatty acids have a broad range of biological actions involving cell membrane integrity, signal transduction, gene expression, and prevention of CVD, metabolic and inflammatory diseases, and cancer [3,24]—as well as with differences in organoleptic attributes, especially taste [24,25], even with undesired sensory characteristics [23].

In particular, forage degradation in the rumen is a complex process that involves multiple microorganisms. In fact, it has been observed that sheep fed only with fresh ryegrass (*Lolium multiflorum*) have greater microbial diversity than those fed with hay and concentrate [26]. The microbiome of pasture-fed ruminants has the ability, through its enzymatic activity, to synthesize the long-chain n-3 fatty acids (FA) (eicosapentaenoic and docosahexaenoic acid) from the α-linolenic acid precursor [27]. In turn, forage-based diets would favor the growth of fibrolytic microorganisms that are primarily
responsible for the intensive hydrogenation activity in the rumen and, consequently, for the production of conjugated linoleic acid (CLA) and vaccenic acid (C18:1 trans 11, precursor of CLA in tissue) that would have benefits for meat quality [28].

Forage species of the pasture may also have effects on the composition of intramuscular fat deposited. In this sense, Fraser et al. [29] compared fatty acid composition of meat derived from lambs fed legumes or grasses, and observed higher concentrations of unsaturated fatty acids when lambs were fed legumes, thus increasing the polyunsaturated:saturated ratio. Meanwhile, De Brito et al. [30] evaluated different herbage species and mixtures in fattening lambs and observed that some mixtures; such as chicory (*Chichorium intybus*) + arrowleaf clover (*Trifolium vesiculosum*), had the ability to promote a better fatty acid profile, while others, such as alfalfa (*Medicago sativa*) + phalaris (*Phalaris aquatica*), led to high Vitamin E concentrations, improving oxidative stability, reducing the conversion of oxymyoglobin to metmyoglobin and, as a consequence, improving the shelf life of meat.

Finally, it is necessary to point out that, frequently in the literature, the effect of the type of diet appears to be confounded by the production system. For example, Popova et al. [31,32], concluded that feeding pastures decreases fat and increases n-3 polyunsaturated content in the carcass. However, these studies compared indoor vs. grazing animals, with all the differences that both systems involve, aside from diets (exercise, comfort, food availability, etc.).

3. Nutrient Intake, Digestion and Rumen Environment of Sheep Consuming Temperate Pastures

In temperate zones, forage species are characterized by higher protein and lower fiber contents compared with tropical ones, and their nutritive value declines less with age, leading to high DM digestibility, which can exceed values of 90% [8]. The chemical composition of temperate pastures and their fermentative characteristics determine the ruminal environment. In turn, it is expected that the inclusion of other feeds in the diets of lambs that consume fresh forage affects the conditions of the rumen environment. Table 1 shows gathered information on the intake, digestibility, and ruminal environment of sheep that consumed diets based on fresh forage.
Table 1. Chemical composition of pasture (g/kg), nutrient intake (g/kg body weight), coefficient of digestibility of dry matter (DMd), rumen environment, microbial nitrogen synthesis (MNS, g/d), and efficiency of microbial nitrogen synthesis (ENMS) in lambs fed fresh forage (FF) or FF plus feedstuffs (concentrate or total mixed ration, TMR).

| Diet                        | FF          | DMCompositionofFreshForage | NutrientIntake | DMIntake | RumenEnvironment | MNS    | ENMS     | Ref |
|-----------------------------|-------------|-----------------------------|----------------|-----------|------------------|--------|----------|-----|
|                             |             | DM                        | N  | NDF          | ADF       | WSC       | DM  | N  | NDF | pH | NH3-N  | VFA  | A:P     |     |
| Ryegrass/white clover       | 100 L       | 240                        | 24.8 | 480          | -          | -        | 121 | 39.0 | 0.927 | 18.7 | OR     | -     | 6.44    | 17.0 | 79.0 | 4.10 | [33]|
| White clover                | 100 L       | 146                        | 44.6 | 264          | -          | -        | 145 | 51.9 | 2.31  | 13.7 | OR     | -     | 6.35    | 28.5 | 96.0 | 3.30 | [33]|
| Alfalfa                     | 100 L       | 212                        | 39.1 | 323          | -          | -        | 123 | 48.1 | 1.88  | 15.5 | OR     | -     | 6.37    | 27.5 | 97.0 | 3.40 | [33]|
| Lotus                       | 100 L       | -                          | -    | -            | -          | -        | NA  | 0.749 | -     | -    | -      | -     | [34]|
| Alfalfa                     | 100 L       | -                          | -    | -            | -          | -        | NA  | 0.763 | -     | -    | -      | -     | [34]|
| Red clover                  | 100 L       | -                          | -    | -            | -          | -        | NA  | 0.745 | -     | -    | -      | -     | [34]|
| Ryegrass                    | 100 L       | -                          | -    | -            | -          | -        | NA  | 0.732 | -     | -    | -      | -     | [34]|
| Alfalfa                     | 100 L       | -                          | -    | -            | -          | -        | NA  | 0.755 | -     | -    | -      | -     | [29]|
| Red clover                  | 100 L       | -                          | -    | -            | -          | -        | NA  | 0.783 | -     | -    | -      | -     | [29]|
| Ryegrass                    | 100 L       | -                          | -    | -            | -          | -        | NA  | 0.745 | -     | -    | -      | -     | [29]|
| Ryegrass                    | 100 L       | 211                        | 24.3 | 482          | 219        | -        | 32.9 | 0.801 | 15.9  | SE   | 0.757  | 6.01  | 14.6   | 85.8 | 2.96 | [35]|
| 75% Ryegrass + 25% white clover | 100 L   | 193                        | 28.5 | 458          | 218        | -        | 33.9 | 0.966 | 15.5  | SE   | 0.765  | 6.04  | 18.3   | 98.1 | 2.93 | [35]|
| 50% Ryegrass + 50% white clover | 100 L  | 175                        | 32.6 | 434          | 218        | -        | 35.9 | 1.17  | 15.6  | SE   | 0.766  | 6.07  | 26.5   | 104  | 3.01 | [35]|
| 25% Ryegrass + 75% white clover | 100 L | 157                        | 36.8 | 410          | 217        | -        | 36.3 | 1.34  | 14.9  | SE   | 0.771  | 6.57  | 26.5   | 106  | 3.02 | [35]|
| White clover                | 100 L       | 139                        | 41.0 | 386          | 216        | -        | 36.9 | 1.51  | 14.2  | SE   | 0.770  | 6.05  | 33.3   | 115  | 3.10 | [35]|
| Ryegrass                    | 100 L       | 168                        | 34.1 | 503          | 240        | -        | 29.3 | 1.03  | 14.7  | OR   | 0.710  | -     | -      | -    | 12.1 | 24.0 | [36]|
| Ryegrass + CM               | 100 L       | 168                        | 34.1 | 503          | 240        | -        | 31.3 | 0.923 | -     | OR   | 0.750  | -     | -      | -    | 13.4 | 23.0 | [36]|
| Ryegrass + CM + CG          | 100 L       | 168                        | 34.1 | 503          | 240        | -        | 33.5 | 1.19  | -     | OR   | 0.720  | -     | -      | -    | 13.3 | 22.0 | [36]|
| Ryegrass + CM + CC          | 100 L       | 168                        | 34.1 | 503          | 240        | -        | 33.2 | 1.19  | -     | OR   | 0.750  | -     | -      | -    | 13.2 | 22.0 | [36]|
| Ryegrass + CG               | 100 L       | 168                        | 34.1 | 503          | 240        | -        | 35.2 | 1.27  | -     | OR   | 0.730  | -     | -      | -    | 12.9 | 21.0 | [36]|
| Alfalfa                     | 100 L       | 296                        | 33.4 | 374          | 211        | 96.8    | 42.8 | 1.46  | 14.9  | OR   | 0.690  | 6.04  | 17.6   | 165  | 1.41 | [37]|
| Alfalfa + TMR75             | 100 L       | 296                        | 33.4 | 374          | 211        | 96.8    | 35.2 | 1.08  | 11.9  | OR   | 0.730  | 6.27  | 23.5   | 161  | 1.60 | [37]|
| Alfalfa + TMR50             | 100 L       | 296                        | 33.4 | 374          | 211        | 96.8    | 36.4 | 1.19  | 11.8  | OR   | 0.700  | 6.20  | 30.5   | 179  | 1.58 | [37]|
| Lotus                       | 100 L       | 214                        | 36.2 | 366          | 223        | -        | 27.2 | 0.985 | 8.97  | OR   | 0.684  | 6.49  | 28.4   | 146  | 3.19 | [38]|
| Alfalfa + TMRa              | 100 L       | 214                        | 36.2 | 366          | 223        | -        | 38.0 | 1.11  | 11.65 | OR   | 0.753  | 6.31  | 21.8   | 154  | 2.46 | [38]|
| Alfalfa + TMri              | 100 L       | 214                        | 36.2 | 366          | 223        | -        | 37.9 | 1.10  | 13.5  | OR   | 0.690  | 6.14  | 16.1   | 152  | 2.01 | [38]|
| Lotus                       | 100 L       | 318                        | 20.2 | 418          | 288        | -        | 40.9 | 0.821 | 17.1  | OR   | 0.680  | 6.80  | 37.4   | 90.4 | 3.50 | 17.6 | 14.1 | [38]|
| Lotus + 5 g/kg sorghum grain | 100 W       | -                          | -    | -            | -          | -        | NA  | 0.752 | -     | -    | -      | -     | [39]|
| Lotus + 10 g/kg sorghum grain | 100 W   | -                          | -    | -            | -          | -        | NA  | 0.752 | -     | -    | -      | -     | [39]|
| Lotus + 15 g/kg sorghum grain | 100 W  | -                          | -    | -            | -          | -        | NA  | 0.752 | -     | -    | -      | -     | [39]|
| Diet Describe | Diet FF  | C  | Chemical Composition of Fresh Forage | Nutrient Intake | DMJ m | DMD | Rumen Environment | MNS | ENMS |
|--------------|---------|----|-------------------------------------|----------------|-------|-----|------------------|-----|------|
| Lotus + clover + ryegrass | 100 * | W | 20.5 | 444 | 285 | - | 17.7 | 0.397 | 7.84 | OR | 0.610 | 6.66 | 23.5 | - | - | 3.19 | 6.54 | [39] |
| Ryegrass diploid (spring) | 100 | W | 144 | 24.3 | 467 | 244 | 253 | 24.5 | 0.594 | 11.4 | OR | - | - | - | - | - | - | [40] |
| Ryegrass high-sugar (spring) | 100 | W | 166 | 22.4 | 418 | 227 | 304 | 27.2 | 0.609 | 11.4 | OR | - | - | - | - | - | - | [40] |
| Ryegrass tetraploid (spring) | 100 | W | 189 | 25.6 | 458 | 246 | 229 | 22.7 | 0.582 | 10.4 | OR | - | - | - | - | - | - | [40] |
| Ryegrass diploid (autumn) | 100 | W | 177 | 33.1 | 631 | 298 | 133 | 22.0 | 0.727 | 13.9 | OR | - | - | - | - | - | - | [40] |
| Ryegrass high-sugar (autumn) | 100 | W | 183 | 35.7 | 580 | 300 | 135 | 23.1 | 0.820 | 13.4 | OR | - | - | - | - | - | - | [40] |
| Ryegrass tetraploid (autumn) | 100 | W | 160 | 38.1 | 611 | 290 | 128 | 20.9 | 0.797 | 12.8 | OR | - | - | - | - | - | - | [40] |
| Italian ryegrass | 100 | L | 243 | 27.5 | 502 | - | 300 | 64.7 | 0.739 | 32.5 | OR | 0.661 | - | - | - | - | - | [41] |
| Barley | 100 | L | 262 | 25.4 | 557 | - | 186 | 64.9 | 0.731 | 36.2 | OR | 0.783 | - | - | - | - | - | [41] |
| Alfalfa | 100 | L | 223 | 34.1 | 356 | 277 | 53.0 | 55.6 | 1.51 | 19.8 | NA | 0.700 | - | - | - | - | - | - | [42] |
| Red clover | 100 | L | 150 | 34.9 | 263 | 198 | 74.0 | 58.2 | 1.74 | 15.3 | NA | 0.730 | - | - | - | - | - | - | [42] |
| White clover | 100 | L | 134 | 39.8 | 223 | 185 | 90.0 | 56.7 | 2.13 | 12.6 | NA | 0.730 | - | - | - | - | - | - | [42] |
| Ryegrass | 100 | L | 187 | 29.8 | 363 | 198 | 166 | 49.5 | 1.30 | 18.0 | NA | 0.750 | - | - | - | - | - | - | [42] |
| Oats + white clover + barley | 70.0 | E | 159 | 18.6 | 554 | 296 | 47.0 | 15.5 | 1.33 | 8.57 | F | 0.670 | - | - | - | - | - | - | [43] |
| Mixed grass | 100 | L | 147 | 23.0 | 499 | 268 | 14.4 | 22.1 | 0.510 | 6.47 | OR | - | 6.47 | 17.8 | 92.5 | 2.34 | 4.71 ** | 11.6 ** | [44] |
| Mixed grass + legumes (forage cut at 7:00 h) | 100 | * | E | 148 | 23.7 | 546 | 279 | 82.0 | 15.5 | 1.85 | 8.44 | F | 0.710 | - | 6.33 | 17.4 | - | - | 8.30 | 20.8 | [43] |
| Mixed grass + legumes + MBP | 70.0 | * | E | 148 | 23.7 | 546 | 279 | 82.0 | 15.5 | 1.52 | 6.67 | F | 0.750 | - | 6.15 | 18.5 | - | - | 8.09 | 16.7 | [43] |
| Mixed grass + legumes + barley + MBP | 70.0 | * | E | 148 | 23.7 | 546 | 279 | 82.0 | 15.5 | 1.64 | 6.31 | F | 0.710 | - | 6.51 | 18.0 | - | - | 7.55 | 16.2 | [43] |
Table 1. Cont.

| Diet       | FF 10 | C 11 | Chemical Composition of Fresh Forage | Nutrient Intake | DMI m DMd | Rumen Environment | MNS | ENMS 21 | Ref |
|------------|-------|------|-------------------------------------|----------------|-----------|-------------------|-----|---------|-----|
|            |       |      | DM 12 | N 13 | NDF 14 | ADF 15 | WSC 16 | DM | N 17 | NDF | pH | NH3-N 18 | VFA 19 | A:P 20 |       |
| Ryegrass   | 100 * | W    | 165   | 31.5 | 423   | 218   | 114   | 13.7 | 0.426 | 5.80 | OR 0.741 | 6.35 | 28.3 | 83.8 | 3.67 | - | - | [46] |
| Ryegrass   | 100 * | W    | 165   | 31.5 | 423   | 218   | 114   | 21.1 | 0.680 | 8.91 | OR 0.753 | 6.18 | 27.1 | 92.2 | 3.38 | - | - | [46] |
| Ryegrass   | 100   | L    | 148   | 29.0 | 464   | 242   | 83.0  | 25.0 | 0.730 | 11.6 | OR 0.646 | 6.71 | - | 74.5 | 3.53 | - | - | [47] |
| Ryegrass   | 100   | L    | 198   | 25.6 | 445   | 231   | 123   | 31.5 | 0.810 | 14.0 | OR 0.750 | 6.71 | - | 58.3 | 2.93 | - | - | [47] |

1 7 g/kg LW daily of cassava meal; 2 cassava meal plus corn gluten meal; 3 cassava meal plus calcium caseinate; 4 corn gluten feed; 5 TMR at a level of 0.75 of the potential intake; 6 TMR at a level 0.50 of the potential intake; 7 TMR with cereal grains; 8 TMR with by-products; 9 molasses-based product; 10 level of FF intake of diets; 11 category of sheep (L: lambs; W: wethers; E: ewes); 12 dry matter; 13 nitrogen; 14 neutral detergent fiber; 15 acid detergent fiber; 16 water soluble carbohydrates; 17 DMI m: methods of intake measure (OR: feed offered less feed refused; NA: n-alkane; SE: sensors; F: Intake was individually fixed); 18 mg/dL; 19 total volatile fatty acids (mM); 20 acetate-propionate ratio; 21 expressed relating to the apparently digestible organic matter ingested; * Restricted forage was offered; ** data available in Thesis Pérez-Ruchel [48].
The studies presented in Table 1 were performed on ryegrass, oats (*Avena sativa*), and barley (*Hordeum vulgare*) as grasses; white (*Trifolium repens*) and red clover (*Trifolium pratense*), alfalfa, and lotus as legumes; and their mixtures. Chicory was also found in mixtures with grasses and legumes. On average, the neutral detergent fiber (NDF) and acid detergent fiber (ADF) values were (440 ± 94 and 244 ± 35 g/kg DM, respectively; mean ± SD). There was a great variability in water-soluble carbohydrate (WSC) concentration (114 ± 70 g/kg DM; mean ± SD) due to the high variation in ryegrass varieties high in WSC content. The relatively low content of WSC is a characteristic of temperate pastures, which is in general considered a limiting factor to the efficiency of N use at rumen level [44,49,50].

The pastures used (Table 1) had average N values of 28.6 ± 7.1 g/kg DM (mean ± SD), and although no data on protein fractionation are reported in these studies, high values of the soluble protein would be expected [51,52], which, in addition to the WSC content, can explain the high N-ammonia concentration in the rumen, as shown in most experimental treatments (20.0 mg/dL ± 8.8; mean ± SD). High selective grazing of sheep may even increase N-ammonia in rumen, as observed by Pérez-Ruchel et al. [53] comparing lambs grazing with lambs fed the same pasture cut and provided in feeders. Rumen fermentation of temperate pastures is usually high [54,55], indicating a good quality of fiber, which explains the high volatile fatty acid (VFA) concentration in the rumen observed in most studies. In sheep consuming temperate pastures as sole feed, the average VFA concentration varied from 58 to 165 Mm, with a net predominance of acetate in most of the studies (Table 1). Only a few papers report microbial protein synthesis in the rumen of lambs or sheep on pastures, and low values seem to be related with restricted feeding regimes, more than to pasture or diet composition.

Some authors proposed that pastures composed of mixtures of grasses and legumes can be a strategy to increase intake, preserving forage quality with maturity in comparison to grass monocultures [56]. Niderkorn et al. [35] studied the supply of different proportions of ryegrass and white clover in cannulated sheep. These authors concluded that a mixture containing 25% to 50% of white clover led to better results with respect to only ryegrass forage. Lambs fed only white clover consumed more, but had high concentrations of N-ammonia, leading to greater N losses. According to Rutter [57], sheep and cattle develop preferences for clover over grass mainly during the morning, but this is reversed during the afternoon. Although the mechanisms involved in these preferences are not clear enough, release of ammonia from the soluble protein fraction of the forage, and subsequent uptake in the blood, in addition to propionate, have been proposed as inductors of satiety in grazing ruminants [58]. Therefore, combining grasses and legumes in the sward could help to maintain intake during a longer period. On the other hand, the inclusion of legumes containing condensed tannins could reduce the rumen degradation of the protein and increase the flow of protein into the duodenum, which in turn can enhance the efficiency of protein digestion by ruminants [55,59–61].

4. Performance of Lambs on Pasture-Based Systems

For finishing lambs at pasture, growth rates of 250–300 g/d from birth to weaning and 150–200 g/d from weaning to finishing could be set as a benchmark according the study published for Orr et al. [62]. However, the performance and nutrient intake from temperate pasture is highly variable. For example, some authors report daily gain of 141 g/d [63], while others report 243 g/d [29], both with lambs grazing on alfalfa. We will focus on factors that allow optimizing the intake of fresh forage in lambs.

In Table 2, growth performance results of lambs grazing on different species of temperate pastures, and pastures supplemented with other feedstuffs, are shown.
Table 2. Dry matter intake (DMI, g/kg body weight (BW)) and average daily gain (ADG, g/d) of sheep fed fresh forage (FF) or FF plus feedstuffs (concentrate or total mixed ration, TMR) with different biomass allowance (t DM/ha) and chemical composition (g/kg BW).

| Diet                      | FF  | Allowance | C  | DM  | N  | NDF | ADF | WSC | Breed         | BW | DMI | DMI m | ADG | Ref |
|---------------------------|-----|-----------|----|-----|----|-----|-----|-----|---------------|----|-----|-------|-----|-----|
| Ryegrass var. AberDart (continuous grazing system) | 100 * | 1.67 | L   | 220 | 32.3 | 537 | 284 | 115 | 20 Brecknock Cheviot | 25.5 | 22.0 | NA    | 47.1 | [64] |
| Ryegrass var. AberDart (rotational grazing system) | 100 | 2.13 | L   | 187 | 33.4 | 482 | 263 | 113 | 20 Brecknock Cheviot | 25.5 | 29.4 | NA    | 98.4 | [64] |
| Ryegrass var. Fennema (continuous grazing system) | 100 * | 1.57 | L   | 216 | 34.1 | 533 | 278 | 100 | 20 Brecknock Cheviot | 25.5 | 27.1 | NA    | 51.5 | [64] |
| Ryegrass var. Fennema (rotational grazing system) | 100 | 2.15 | L   | 188 | 30.1 | 496 | 271 | 100 | 20 Brecknock Cheviot | 25.5 | 30.2 | NA    | 71.7 | [64] |
| Ryegrass high WSC (spring) | 100 |   | L   | 34.6 | 434 |  -  | 196 | 40  | -  | 42.0 | -   | -   | 170   | [65] |
| Ryegrass diploid (spring) | 100 |   | L   | 34.1 | 458 |  -  | 182 | 40  | -  | 42.0 | -   | -   | 158   | [65] |
| Ryegrass tetraploid (spring) | 100 |   | L   | 36.0 | 440 |  -  | 178 | 40  | -  | 42.0 | -   | -   | 164   | [65] |
| Ryegrass high WSC (autumn) | 100 |   | E   | 40.3 | 416 |  -  | 186 | 40  | -  | 30.7 | -   | -   | 179   | [65] |
| Ryegrass diploid (autumn) | 100 |   | L   | 42.7 | 416 |  -  | 162 | 40  | -  | 30.7 | -   | -   | 206   | [65] |
| Ryegrass tetraploid (autumn) | 100 |   | L   | 28.0 | 462 |  -  | 251 | 60  | -  | 28.7 | -   | -   | 133   | [65] |
| Ryegrass diploid (spring) | 100 |   | L   | 28.3 | 497 |  -  | 221 | 60  | -  | 28.7 | -   | -   | 120   | [65] |
| Ryegrass high WSC (spring) | 100 |   | L   | 34.6 | 434 |  -  | 196 | 40  | -  | 42.0 | -   | -   | 170   | [65] |
| Ryegrass diploid (spring) | 100 |   | L   | 34.1 | 458 |  -  | 182 | 40  | -  | 42.0 | -   | -   | 158   | [65] |
| Ryegrass tetraploid (spring) | 100 |   | L   | 36.0 | 440 |  -  | 178 | 40  | -  | 42.0 | -   | -   | 164   | [65] |
| Ryegrass high WSC (autumn) | 100 |   | E   | 40.3 | 416 |  -  | 186 | 40  | -  | 30.7 | -   | -   | 179   | [65] |
| Ryegrass diploid (autumn) | 100 |   | L   | 42.7 | 416 |  -  | 162 | 40  | -  | 30.7 | -   | -   | 206   | [65] |
| Ryegrass tetraploid (autumn) | 100 |   | L   | 28.0 | 462 |  -  | 251 | 60  | -  | 28.7 | -   | -   | 133   | [65] |
| Ryegrass diploid (spring) | 100 |   | L   | 28.3 | 497 |  -  | 221 | 60  | -  | 28.7 | -   | -   | 120   | [65] |
| Ryegrass high WSC (spring) | 100 |   | L   | 34.6 | 434 |  -  | 196 | 40  | -  | 42.0 | -   | -   | 170   | [65] |
| Ryegrass diploid (spring) | 100 |   | L   | 34.1 | 458 |  -  | 182 | 40  | -  | 42.0 | -   | -   | 158   | [65] |
| Ryegrass tetraploid (spring) | 100 |   | L   | 36.0 | 440 |  -  | 178 | 40  | -  | 42.0 | -   | -   | 164   | [65] |
| Ryegrass high WSC (autumn) | 100 |   | E   | 40.3 | 416 |  -  | 186 | 40  | -  | 30.7 | -   | -   | 179   | [65] |
| Ryegrass diploid (autumn) | 100 |   | L   | 42.7 | 416 |  -  | 162 | 40  | -  | 30.7 | -   | -   | 206   | [65] |
| Ryegrass tetraploid (autumn) | 100 |   | L   | 28.0 | 462 |  -  | 251 | 60  | -  | 28.7 | -   | -   | 133   | [65] |
| Ryegrass diploid (spring) | 100 |   | L   | 28.3 | 497 |  -  | 221 | 60  | -  | 28.7 | -   | -   | 120   | [65] |
| Ryegrass high WSC (spring) | 100 |   | L   | 34.6 | 434 |  -  | 196 | 40  | -  | 42.0 | -   | -   | 170   | [65] |
| Ryegrass diploid (spring) | 100 |   | L   | 34.1 | 458 |  -  | 182 | 40  | -  | 42.0 | -   | -   | 158   | [65] |
| Ryegrass tetraploid (spring) | 100 |   | L   | 36.0 | 440 |  -  | 178 | 40  | -  | 42.0 | -   | -   | 164   | [65] |
Table 2. Cont.

| Diet | FF [9] | Allowance | C [10] | Chemical Composition of Forage | Breed | BW [17] | DMI | DMI m [18] | ADG | Ref |
|------|--------|-----------|--------|--------------------------------|-------|--------|------|------------|-----|-----|
|      |        |           |        | DM [11] | N [12] | NDF [13] | ADF [14] | WSC [15] |       |     |
| 6 species of forage 1 | 100 | 1.20 | L | - | 31.4 | 405 | 194 | - | 30 | Suffolk, Texel, others | 33.3 | - | - | 193 [68] |
| 9 species of forage 2 | 100 | 1.20 | L | - | 30.2 | 400 | 190 | - | 30 | Suffolk, Texel, others | 32.1 | - | - | 193 [68] |
| Alfalfa | 100 | - | L | - | 39.8 | - | - | - | 36 | Rambouillet × Suffolk | 30.7 | 42.2 | Cut | 141 [63] |
| Alfalfa + 123 g corn | - | - | L | - | 39.5 | - | - | - | 36 | Rambouillet × Suffolk | 30.7 | 41.6 | Cut | 154 [63] |
| Alfalfa + 247 g corn | - | - | L | - | 39.4 | - | - | - | 36 | Rambouillet × Suffolk | 30.7 | 43.5 | Cut | 169 [63] |
| Alfalfa grazing | 100 | - | L | - | - | - | - | - | 12 | Romane | 21.5 | - | - | 299 [69] |
| Alfalfa + barley | 62.1 | - | L | - | - | - | - | - | 12 | Romane | 21.5 | - | - | 294 [69] |
| Dactylis + alfalfa 3 | 100 | 1.53 | L | 197 | 28.5 | 476 | 308 | - | 10 | Dorset | 23.6 | 75.2 | NDFi | 267 [70] |
| Dactylis + alfalfa 4 | 100 | 1.53 | L | 197 | 28.5 | 476 | 308 | - | 10 | Dorset | 23.6 | 74.0 | NDFi | 295 [70] |
| Orchardgrass | 100 | - | L | 192 | 35.7 | 567 | 305 | - | 8 | - | 25.9 | - | - | 147 [71] |
| Ryegrass | 100 | - | L | 158 | 36.6 | 489 | 286 | - | 8 | - | 25.5 | - | - | 152 [71] |
| Alfalfa | 100 | - | L | 181 | 47.7 | 314 | 234 | - | 8 | - | 25.4 | - | - | 239 [71] |
| Orchardgrass | 100 | - | L | 209 | 37.4 | 559 | 307 | - | 4 | - | 29.2 | - | - | 149 [71] |
| Ryegrass | 100 | - | L | 191 | 33.4 | 473 | 283 | - | 4 | - | 27.7 | - | - | 150 [71] |
| Alfalfa | 100 | - | L | 196 | 46.6 | 293 | 228 | - | 4 | - | 22.9 | - | - | 175 [71] |
| Orchardgrass | 100 | - | L | 237 | 39.0 | 507 | 284 | - | 8 | - | 21.9 | - | - | 112 [71] |
| Ryegrass | 100 | - | L | 213 | 37.9 | 443 | 269 | - | 8 | - | 21.4 | - | - | 85 [71] |
| Alfalfa | 100 | - | L | 230 | 42.9 | 281 | 197 | - | 8 | - | 21.8 | - | - | 256 [71] |
| Alfalfa + TMRa 5 | 40 | - | L | 214 | 36.2 | 366 | 223 | - | 18 | Corriedale × Ile de France | 29.5 | 46.9 | Cut | 336 [72] |
| Alfalfa + TMRf 6 | 41 | - | L | 214 | 36.2 | 366 | 223 | - | 18 | Corriedale × Ile de France | 29.5 | 42.6 | Cut | 305 [72] |
| Lotus | 100 | 5.27 | L | - | 56.0 | 449 | 350 | - | 20 | Romney | 22.8 | 58.2 | C2,O2 | 203 [73] |
| Alfalfa | 100 | 5.24 | L | - | 49.8 | 423 | 311 | - | 20 | Romney | 22.8 | 65.5 | C2,O2 | 185 [73] |
| Ryegrass WSC | 100 | - | L | - | 23.8 | 411 | 222 | 143 | 5 | Bluefaced Leicester | 14.0 | 69.2 | EC | 312 [74] |
| Ryegrass control WSC | 100 | - | L | - | 27.0 | 487 | 255 | 89.0 | 5 | Bluefaced Leicester | 14.0 | 82.9 | EC | 271 [74] |
| Ryegrass WSC | 100 | - | L | - | 32.0 | 473 | 250 | 113 | 5 | Bluefaced Leicester | 14.0 | 120 | EC | 244 [74] |
| Ryegrass control WSC | 100 | - | L | - | 26.6 | 540 | 279 | 75.0 | 5 | Bluefaced Leicester | 14.0 | 94.0 | EC | 194 [74] |
| Ryegrass WSC | 100 | - | L | - | 29.0 | 506 | 267 | 92.0 | 5 | Bluefaced Leicester | 14.0 | 77.9 | EC | 186 [74] |
### Table 2. Cont.

| Diet                          | FF  | Allowance | C  | Chemical Composition of Forage | N  | Breed       | BW  | DMI | DMI_ω | ADG | Ref     |
|-------------------------------|-----|-----------|----|-------------------------------|----|------------|-----|-----|-------|-----|---------|
|                               |     |           |    | DM  | N   | NDF_ω | ADF  | WSC  |       |     |         |
| Ryegrass control WSC          | 100 | -         | L  | -   | 31.0 | 514   | 274  | 10.0 | 5     | Bluefaced Leicester | 14.0 | 82.9 | EC    | 175  | [74]   |
| Trifolium alexandrium +        |     |           |    |     |     |       |      |      |       |                  |       |       |       |      |        |
| concentrate                   | 65.4| -         | L  | -   | -   | -     | -    | -    | 9     | Lohi | 21.0 | 37.1 | Cut   | 130  | [75]   |
| Trifolium alexandrium +        |     |           |    |     |     |       |      |      |       |                  |       |       |       |      |        |
| concentrate                   | 66.0| -         | L  | -   | -   | -     | -    | -    | 9     | Lohi | 21.0 | 37.6 | Cut   | 160  | [75]   |
| Tetraploid ryegrass and white  |     |           |    |     |     |       |      |      |       |                  |       |       |       |      |        |
| clover                        | 66.7| -         | L  | -   | -   | -     | -    | -    | 9     | Lohi | 21.0 | 38.6 | Cut   | 180  | [75]   |
| Herb/clover                   | 100 | 3.45      | L  | 25.3| 281  | -     | -    | 6     | Romney| 33.0 | -    | -    | 247   | [76]   |
| Plantain/pasture              | 100 | 3.79      | L  | 20.6| 399  | -     | -    | 6     | Romney| 33.0 | -    | -    | 107   | [76]   |
| Tetraploid ryegrass and white  | 100 | 3.79      | L  | 31.4| 481  | -     | -    | 6     | Romney| 33.0 | -    | -    | 119   | [76]   |
| clover                        | 100 | 5.55      | L  | 22.6| 537  | -     | -    | 6     | Romney| 33.0 | -    | -    | 119   | [76]   |
| Diploid ryegrass, other grass  | 100 | 5.55      | L  | 24.8| 480  | -     | 121  | 18    | -     | 28.5 | 39.0 | OR    | 105   | [33]   |
| species, and white clover     |     |           |    |     |     |       |      |      |       |                  |       |       |       |      |        |
| Ryegrass/white clover         | 100 | L         | 240| 480 | -    | 121  | 18   | -    | 28.5  | 39.0 | OR    | 105   | [33]   |
| White clover                  | 100 | L         | 146| 264 | 145  | 18   | -    | 28.5  | 51.9  | OR    | 256   | [33]   |
| Alfalfa                       | 100 | L         | 212| 323 | 123  | 18   | -    | 28.5  | 48.1  | OR    | 191   | [33]   |

1 perennial ryegrass, timothy, white clover, red clover, plantain, and chicory; 2 perennial ryegrass, timothy, cocksfoot, white clover, red clover, birdsfoot trefoil, plantain, chicory, and yarrow; 3 cut daily ad libitum; 4 rotational grazing; 5 TMR with cereal grains; 6 TMR with by-products; 7 chicory, plantain, red clover, and white clover; 8 plantain, ryegrass, and white clover; 9 level of FF intake of diets; 10 category of sheep (L: lambs; W: wethers; E: ewes); 11 dry matter; 12 nitrogen; 13 neutral detergent fiber; 14 acid detergent fiber; 15 water soluble carbohydrates; 16 lambs per treatment; 17 body weight (kg); 18 DMI_ω: methods of intake measure (NA: n-alkane; Cr_2O_3: chromic oxide; Cut: disappearance herbage mass; NDFi: indigestible NDF; EC: exclosure cages; OR: feed offered less feed refused); * restrict offered.
The data presented in Table 2 show a high variability in the growth performance of lambs fed only fresh forage, ranging from 47.1 to 336 g/d. In this sense, forage-species grazing was found to significantly affect the liveweight gain of the lambs [34]. This great diversity in productive results, in general, is explained by variations in intake [77], although it is necessary to point out that the procedures to measure intake of pasture differed among studies. It is known that in herbivores, the control of intake is multifactorial [78]. In grazing animals, the total daily herbage intake depends on the biting rate, bite weight, and the time spent grazing [79,80]. The biting rate and bite weight decrease as forage advances in the state of maturity because the animals spend more time selecting the food [81].

Results show that lambs grazing on a lucerne-dominant perennial pasture performed better than did lambs grazing on annual pasture with supplements during the finishing period [82]. In addition, Fraser and Rowarth [67] underlined the importance of feed quality on animal performance when they evaluated ryegrass, white clover, chicory, plantain (*Plantago lanceolata*), or lotus. These variations in grass quality are mainly explained by the differences in the fiber content and protein of the pastures used. In this sense, the intake of NDF is considered the first limiting factor of the intake in forage diets since it is related to the reticulum–rumen distention [83].

Figure 1 presents the relationship between dry matter intake (DMI; g/kg BW) and average daily gain (ADG; g/day) in treatments on lambs fed only fresh forage [33,42,63,64,67,70,73,74].

![Figure 1](image)

*Figure 1*. Relationship between dry matter intake (DMI; g/kg BW) and average daily gain (ADG; g/day) in lambs fed only fresh forage.

Even considering that different methods of intake assessment were used in the studies (see Table 2), and that this fact could add variability to the data set, the consumption of DM in grazing lambs explained almost 40% of the ADG (P < 0.001). In this sense, the quality of the pasture would play a determining role; observed values above the predicting line, corresponded mostly to legumes with a high content of N (g/kg DM) or varieties of ryegrass with high WSC content.

In addition, when the plants advance in the state of maturity, the retention time of the forage in the rumen is increased, limiting voluntary intake, mainly due to the increase of cell walls and lignified tissues, and the decrease of the protein content [84]. Reduction in the concentration of the cell wall of the forage and/or the increase of its digestibility would allow the improvement of the animal’s performance [85] through, above all, an increase in consumption. Moreover, it is also related to other characteristics such as the structure of the plant, its morphology, its density and height, and its allowance or spatial distribution. The differences between grasses and legumes have been reported in the literature through an extensive review by Luscher et al. [56]. The smaller content of structural
components of the cell wall of herbaceous legumes with respect to the grass represents one of the main differences, which is reflected in a greater digestibility of the organic matter, and greater concentrations of net energy and metabolizable protein. Moreover, other authors have reported that forage legumes, such as red clover and alfalfa, offered either fresh or as silage, could increase growth rates in ruminants due to the increased intake of DM, with regard to grasses such as ryegrass [34,86].

In general, voluntary intake of legume forage is 10% to 15% greater than grasses of similar digestibility [56]; this is probably because, in an equivalent phenological state, legumes have a lower resistance to chewing, a faster rate of digestion, and a faster rate of rumen particles, which in turn reduces rumen filling [87]. The inclusion of legumes (red clover and alfalfa) has the potential to improve the quality of the diet of sheep [88], increasing daily gain [29,89], and some legumes, such as lotus, even reduced the total nematode parasite intensities compared to lambs grazing on ryegrass/white clover swards [42,86,90]. Fraser et al. [29] evaluated the performance of lambs fed with clover, alfalfa, or ryegrass. These authors reported daily gains of 243, 305, and 184 g/d in lambs fed alfalfa, red clover, and perennial ryegrass, respectively.

On the other hand, some authors observed that plant species diversity increases feeding motivation [91], such that the mixture of pastures with several species could be considered as positively affecting forage intake and animal performance [68,76]. Niderkorn et al. [35], in the central region of the Alpes (France), studied different mixtures of perennial ryegrass (cv. AberVon) and white clover (cv. Merwi), at different ratios (0, 0.25, 0.50, 0.75, and 1). They observed that the clover/ryegrass ratios of 0.25 and 0.50 optimized consumption and digestion. Moreover, Papadopoulos et al. [88] evaluated the addition of white clover to orchardgrass (Dactylis glomerata) pasture on the performance of grazing lambs. These authors reported lower concentrations of ADF and NDF in a mixture (white clover + orchardgrass) vs. orchardgrass, and body weight gain for lambs grazing on the mixture was 40% greater than for lambs grazing on orchardgrass.

Another constraining factor of intake in grazing is when the moisture concentration of the pasture is too high (above 80%) [92]. Some authors reported that high-moisture forages, compared to dry forages (hay), caused a lower passage rate and lower intake in sheep and had a lower digestibility [93]. However, low moisture can also negatively affect intake of fresh pastures. Kenney et al. [94] evaluated the rate of intake in sheep fed kikuyu grass (Pennisetum clandestinum); the intake rate of fresh forage decreased as DM content increased, but above 40% DM content the intake rate remained relatively constant.

Grazing management is a key point in intensive pasture-based systems, in this sense, the time of access to forage is very important. The effects of this management on pasture intake have been studied in dairy cattle and beef cattle [79,95], while there is less information for sheep. Pérez-Ruchel et al. [39] studied the effect of a time restriction on access to feed in sheep fed only pasture, and reported an increase in the intake rate when the time of access to forage was reduced from 24 to 6 h/d. However, this increase in the intake rate did not compensate for the lower time of access to forage, leading to a decrease in the total intake. In addition, Iason et al. [96] evaluated the effects of food availability (5.5 and 3 cm sward height) on the ability of grazing sheep to compensate for the restriction of daily grazing time. In response to restricted time, the sheep had a higher rate of intake, achieved mainly via larger bites. The behavioral responses to restricted time allowed the sheep to counteract the reduced daily grazing time only for the tallest sward, but in short swards, the time restriction led to a reduction in total daily intake. In another experiment, Luciano et al. [97] compared performance of lambs fed concentrates in stall, grass at pasture for 8 h, or grass at pasture for 4 h in the afternoon. These authors concluded that growing lambs can tolerate a restriction of grazing duration without detrimental effects on performances. In general, the effect of restriction time on sheep grazing consumption would appear to be determined by increasing the bite rate, as long as it has enough availability, height, and is a type of pasture that allows compensation [98].

In grazing, the preference of lambs for some species of pastures may be affected by changes in chemical composition with the season of the year [99]. In addition, the management of the time of day when the animals are in the pasture can act for or against the ability of the animal to
optimize the harvest. Several authors studying this topic [100–103] observed a higher concentration of non-structural carbohydrates (NSC) when forage was harvested in the afternoon. Probably, the greater concentration on NSC increased the palatability of the pasture, and as a consequence, increased daily gain [74]. In addition, NSC may be an alternative to increase the efficiency of N use, as will be seen later. Ciavarella et al. [104] observed that differences in the WSC concentration of ryegrasses had a significant influence on dietary choices of sheep grazing. In a review, Edwards et al. [105] studied opportunities to improve the diet quality, intake, and performance of animals through manipulation of the partial preference commonly shown by grazing animals for different pasture components. This work highlights the preference patterns and the complexity of plant–animal interactions.

Based on the results discussed in this section, to maximize the intake of fresh forage in the fattening of lambs, an important point is to consider a better efficiency in the forage harvest, associated with the grazing duration and moment of the day in which the pasture is supplied. In turn, to maximize the intake of fresh forage, there must be high availability, and mixtures of legumes and grass seem to be the best options.

5. The Addition of Other Feedstuffs to Temperate Forage-Based Diets

5.1. Digestion of Temperate Forages Plus Other Feedstuffs

How the forage is combined with other feedstuffs (concentrate or total mixed ration, TMR) can have a large influence on N metabolism and nutrient utilization and improve efficiency of microbial protein synthesis. There is consensus among authors that the synchrony of carbohydrates (CHO) and nitrogen components is key to increase the synthesis of microbial protein; and that the availability of CHO is decisive in this regard, since in rumen there is no endogenous source for energy to compensate temporary imbalances at the rumen level, as for N through recycling [106]. Given the high level of soluble protein components of the pasture—and the fact that it contains relatively low levels of CHOs with fast fermentation rates (WSC), and high levels with slow ones (cell walls)—one way to achieve synchronization would be the addition to the diet of carbohydrates with rapid fermentation through supplementation with grains or by-products [107]. Theoretically, starch would accompany the rapid release of N, and thus would allow the achievement of higher microbial growth with a positive impact on growth.

Nutrient synchronization has been studied in cattle [108,109] and also in concentrate diets for sheep [110–113]. In grazing lambs, the results have not been consistent. For example, Trevaskis et al. [114] evaluated the effect of nutrient synchronization in a series of experiments—either by the feeding of carbohydrate-based supplements (sucrose and fine-rolled barley grain) to tropical and temperate pasture (kikuyu; *Pennisetum clandestinum*, or ryegrass), or by providing pasture with a higher ratio of carbohydrate/N (kikuyu or ryegrass cut early morning or late afternoon)—on rumen pH, ammonia, and microbial protein synthesis. The results of these studies supported the hypothesis that there are benefits on microbial protein synthesis in synchronizing the availability of rumen-fermentable carbohydrates with N in the rumen, but this is not always associated with significant changes in rumen pH and NH₃-N concentrations. Meanwhile, Tebot et al. [43] evaluated three forage diets with non-fibrous carbohydrate supplementation (100% fresh temperate forage, 70% forage + 30 barley grain, and 70% forage + 15% barley + 15% molasses-based product). The results indicated that supplementation with starch (barley) or sugars (molasses) in sheep grazing did not improve ruminal N-ammonia captured for microbial protein synthesis. In addition, Aguerre et al. [13] evaluated lotus as fresh forage with different levels of sorghum grain supplementation (0, 5, 10, and 15 g/kg BW) in lambs, and also reported that grain did not improve the use of N-ammonia; moreover, microbial protein synthesis decreased as grain supplementation levels increased, probably because there was a net predominance of starch (sorghum grain) in the supplement, and it was supplied separately to forage. Overall, these results could indicate mismatches in the use of CHO and N due to feeding management of pastures (animals are not consuming the pasture and the supplement at the same time) rather than
the quantity of nutrients (CHO and N) ingested throughout the day. Amaral et al. [36] studied the level of starch and the type of protein (high or low degradability) to supplement lambs consuming ryegrass and concluded that the supplement should contain both starch and true protein sources.

Supplementation with cereal grains on pastures of good quality, not only would not appear to be effective in increasing the capture of N, as mentioned, but would also represent an additional disadvantage, linked to the degradation of CHOs and the reduction of ruminal pH as a consequence. This effect is aggravated in sheep, especially due to a greater susceptibility to episodes of ruminal acidosis with respect to cattle [13]. Ruminal pH reduction would affect, among other things, protein degradation, due to a decrease in the activity of proteolytic enzymes when the pH reaches values below 5.5 [115].

Pasture supplementation with TMR (mixture of forage and concentrates) appears as a strategy that can improve the uptake of N-ammonia and improve microbial protein synthesis on a forage-based diet in cattle [116]; but in sheep, this approach has been little explored. Our group evaluated different levels of fresh forage supplementation to a total mixed ration (TMR) (0, 0.50, 0.75, and only fresh forage) in lambs. We observed that the ruminal pH and ammonia concentration increased as the inclusion of forage in the diet [37]. On the other hand, when cereal grains were replaced by corn by-products combined with fresh forage provided for 8 h per day, rumen pH and ammonia decreased, but the microbial protein synthesis did not change [38]. Figure 2 shows the relationship observed in several studies of our group between N intake and rumen NH₃-N in lambs fed forage, forage plus concentrate, or TMR. Ruminal NH₃-N concentration does not seem to be related up to a certain level of N intake.

![Figure 2. Relationship between N intake and NH₃-N in lambs fed fresh forage plus feedstuffs (studies plotted measured the intake by difference offered-refused in feeders).](image)

In addition to the degradation kinetics of CHO sources, there are other factors, such as ruminal pH and the amount of concentrate consumed, that influence the animal and therefore the productive results. Based on the results shown, it seems that such synchrony could not be achieved when good quality pastures are supplemented exclusively with carbohydrates. In addition to the implications on the use of nitrogenous materials from pasture from an environmental point of view, the low capture of N-ammonia in the rumen generally implies a lack in the synthesis of microbial protein and, therefore, a poor use of diet at the digestive level.
5.2. Performance of Lambs Fed Fresh Forage Plus Other Feedstuffs

In pasture-based systems, animal performance in general is limited with respect to the most intensive production systems (feedlot), mainly due to the lower contribution of forage energy to achieve high daily gains. However, it is possible to improve animal performance through different strategies to maximize the consumption of fresh forage, in order to obtain the benefits of pasture consumption over meat quality. In this section, the objective is to update the information on feeding strategies that can achieve high daily gains with high levels of inclusion of fresh pasture.

In confinement systems, daily gains of 350–370 g/d are reported \[117,118\] and over 300 g/d, even with 60% levels of a fibrous by-product (distillers dried grains with solubles) in the diet \[117\]. Jacques et al. \[70\] working with lambs fed with ad libitum concentrate, 60% hay and 40% concentrate, fresh forage cut and offered twice a day, and grazing lambs, reported daily gains of 449, 347, 267, and 295 g/d, respectively. Although forage-fed lambs had lower daily gains and longer termination periods, the use of pasture termination systems would avoid excessively fat carcasses of lambs slaughtered at 47 kg liveweight. In addition, Karnezos et al. \[63\] evaluated the supplementation with corn (123 and 247 g/d) in lambs grazing on alfalfa. ADG in lambs without supplementation reached 141 g/d, and 169 g/d was obtained for lambs supplemented with 247 g/d of corn. In turn, Devincenzi et al. \[69\] evaluated performance in lambs grazing on alfalfa, supplemented with barley, or confined (concentrate + hay), reporting ADG of 294, 299, and 314 g/d respectively.

6. Combining Pastures with TMR Diets for Finishing Lambs

A strategy less studied in sheep is to feed the lambs with a partial mixed ration (PMR) consisting of the combination of a TMR ration and fresh forage of good quality. The use of PMR, alternating daily grazing periods with periods of access to a TMR, seeks to add the positive aspects of pasture-based systems and confinement. Additionally, the use of PMR could overcome the absence of positive results discussed above when grazing grains are supplemented.

Most of the work done with temperate pastures as the only food, and with the aggregate of concentrates or ration completely mixed, have been carried out in dairy cattle \[116,119–121\]. If these effects could be demonstrated in sheep, this strategy could be used to produce high yields of high-quality lamb meat with the inclusion of high levels of fresh forage.

However, the effects of the use of PMR in lambs have not been conclusive so far. Pérez-Ruchel et al. \[37\], upon supplementing, with fresh alfalfa, lambs fed with decreasing levels of a TMR, observed an increase in nutrient consumption. Fernandez-Turren et al. \[38\] evaluated a PMR diet composed of fresh forage (alfalfa), cut and offered for 8 h/d, and TMR (either with cereal grains or fibrous by-products) achieved higher levels of consumption with respect to the use of fresh forage only offered ad libitum throughout the day.

Britos et al. \[122\], studying PMR diets for lambs composed of TMR and fresh alfalfa in a rumen simulation technique (RUSITEC), concluded that these diets could enhance digestibility and rumen conditions with respect to both TMR and forage, and also that the inclusion of a fibrous energy source in the TMR could enhance microbial synthesis. These results, obtained in vitro, coincide with those obtained in vivo in lambs fed PMR diets \[38\]. This better synchronization in the supply of nutrients at the ruminal level in lambs fed PMR could explain the high daily gains (300 g/d) when Urioste et al. \[72\] evaluated the performance with the same diets.

7. Conclusions

As a result of investigating different strategies for fattening lambs under pasture-based systems, this study found that the quality of the pasture plays a fundamental role, especially in allowing greater efficiency in the times of fattening. However, several points are still unclear, such as the level of pasture necessary to achieve the benefits of pasture-feeding on meat characteristics, the level and type of pasture to maximize intake, and the combination to reach both attributes at once. Work is still needed
to deepen strategies that maximize ruminal efficiency and provide data on the digestive use of diets with a high level of forage inclusion in fattening lambs.

The highly productive systems that seek to incorporate high levels of fresh forage would appear to be linked to the use of pastures of high quality and high availability, and possibly added to by the inclusion of other feedstuffs to increase energy consumption and a better use of the N components of the pasture.

**Author Contributions:** Conceptualization, C.C., J.L.R.; data curation, G.F.-T.; writing—original draft preparation, G.F.-T. and C.C.; writing and editing, J.M.A. and J.L.R. visualization, A.P.-R., C.C. All authors have read and agreed to the published version of the manuscript.

**Funding:** We acknowledge financial support by ANII-Fondo Innovagro for J.A. grant (FSA_1_2013_1_12561) and ANII-Sistema Nacional de Becas for G.F grant (POS_NAC_2016_1_130922).

**Conflicts of Interest:** The authors declare no conflict of interest.

**References**

1. Priolo, A.; Micol, D.; Agabriel, J. Effects of grass feeding systems on ruminant meat colour and flavour. A review. *Anim. Res.* 2001, 50, 185–200. [CrossRef]
2. Cabrera, M.C.; Saadoun, A. An overview of the nutritional value of beef and lamb meat from South America. *Meat Sci.* 2014, 98, 435–444. [CrossRef] [PubMed]
3. Chikwanha, O.C.; Vahmani, P.; Muchenje, V.; Dugan, M.E.R.; Mapiye, C. Nutritional enhancement of sheep meat fatty acid profile for human health and wellbeing. *Food Res. Int.* 2018, 104, 25–38. [CrossRef] [PubMed]
4. Hersleth, H.; Naes, T.; Rodbotten, M.; Lind, V.; Monteleone, E. Lamb meat—Importance of origin and grazing system for italian and norwegian consumers. *Meat Sci.* 2012, 90, 899–907. [CrossRef] [PubMed]
5. Baumont, R.; Lewis, E.; Delaby, L.; Prache, S.; Horan, B. Sustainable intensification of grass-based ruminant production. In Proceedings of the 25th General Meeting of the European Grassland Federation, Aberystwyth, Wales, 7–11 September 2014; Volume 19, pp. 521–532.
6. Martin-Collado, D.; Boettcher, P.; Bernuéüs, A. Opinion paper: Livestock agroecosystems provide ecosystem services but not their components and breeds. *Animal* 2019, 13, 2111–2113. [CrossRef]
7. Bernuéüs, A.; Ruiz, R.; Olazíola, A.; Villalba, D.; Casasuís, I. Sustainability of pasture-based livestock farming systems in the European Mediterranean context: Synergies and trade-offs. *Livest. Sci.* 2011, 139, 44–57. [CrossRef]
8. Lee, M.A. A global comparison of the nutritive values of forage plants grown in contrasting environments. *J. Plant Res.* 2018, 131, 641–654. [CrossRef]
9. Capstaff, N.M.; Miller, A.J. Improving the yield and nutritional quality of forage crops. *Front. Plant Sci.* 2018, 9, 1–18. [CrossRef]
10. Wilkinson, J.M.; Lee, M.R.F.; Rivero, M.J.; Chamberlain, A.T. Some challenges and opportunities for grazing dairy cows on temperate pastures. *Grass Forage Sci.* 2019, 75, 1–17. [CrossRef]
11. van Gastelen, S.; Dijkstra, J.; Bannink, A. Are dietary strategies to mitigate enteric methane emission equally effective across dairy cattle, beef cattle, and sheep? *J. Dairy Sci.* 2019, 102, 6109–6130. [CrossRef] [PubMed]
12. Soto-Navarro, S.A.; Lopez, R.; Sankey, C.; Capitan, B.M.; Holland, B.P.; Balstad, L.A.; Kreibiel, C.R. Comparative digestibility by cattle versus sheep: Effect of forage quality. *J. Anim. Sci.* 2014, 92, 1621–1629. [CrossRef] [PubMed]
13. Aguerre, M.; Cajarville, C.; Kozloski, G.V.; Repetto, J.L. Intake and digestive responses by ruminants fed fresh temperate pasture supplemented with increased levels of sorghum grain: A comparison between cattle and sheep. *Anim. Feed Sci. Technol.* 2013, 186, 12–19. [CrossRef]
14. Carrasco, S.; Ripoll, G.; Sanz, A.; Alvarez-Rodriguez, J.; Panea, B.; Revilla, R.; Joy, M. Effect of feeding system on growth and carcass characteristics of Churra Tensina light lambs. *Livest. Sci.* 2009, 121, 56–63. [CrossRef]
15. Murphy, T.A.; Loerch, S.C.; Smith, F.E. Effects of feeding high-concentrate diets at restricted intakes on digestibility and nitrogen metabolism in growing lambs. *J. Anim. Sci.* 1994, 72, 1583–1590. [CrossRef]
16. Kolver, E.S. Nutritional limitations to increased production on pasture-based systems. *Proc. Nutr. Soc.* 2003, 62, 291–300. [CrossRef]
17. De Brito, G.F.; Ponnampalam, E.N.; Hopkins, D.L. The Effect of Extensive Feeding Systems on Growth Rate, Carcass Traits, and Meat Quality of Finishing Lambs. Compr. Rev. Food Sci. Food Saf. 2017, 16, 23–38. [CrossRef]
18. Howes, N.L.; Bekhit, A.E.D.A.; Burritt, D.J.; Campbell, A.W. Opportunities and Implications of Pasture-Based Lamb Fattening to Enhance the Long-Chain Fatty Acid Composition in Meat. Compr. Rev. Food Sci. Food Saf. 2015, 14, 22–36. [CrossRef]
19. Jacques, J.; Chouinard, P.Y.; Gariépy, C.; Cinq-Mars, D. Meat quality, organoleptic characteristics, and fatty acid composition of Dorset lambs fed different forage to concentrate ratios or fresh grass. Can. J. Anim. Sci. 2017, 97, 290–301. [CrossRef]
20. Siró, I.; Kápolna, E.; Kápolna, B.; Lugasi, A. Functional food. Product development, marketing and consumer acceptance-A review. Appetite 2008, 51, 456–467. [CrossRef]
21. Fisher, A.V.; Enser, M.; Richardson, R.I.; Wood, J.D.; Nute, G.R.; Kurt, E.; Sinclair, L.A.; Wilkinson, R.G. Fatty acid composition and eating quality of lamb types derived from four diverse breed × production systems. Meat Sci. 2000, 55, 141–147. [CrossRef]
22. Archiméde, H.; Pellonde, P.; Despois, P.; Etienne, T.; Alexandre, G. Growth performances and carcass traits of Ovin Martinik lambs fed various ratios of tropical forage to concentrate under intensive conditions. Small Rumin. Res. 2008, 75, 162–170. [CrossRef]
23. Resconi, V.C.; Campo, M.M.; i. Furnols, M.F.; Montossi, F.; Sañudo, C. Sensory evaluation of castrated lambs finished on different proportions of pasture and concentrate feeding systems. Meat Sci. 2009, 83, 31–37. [CrossRef] [PubMed]
24. Santos-Silva, J.; Mendes, I.A.; Bessa, R.J.B. The effect of genotype, feeding system and slaughter weight on the quality of light lambs. 1. Growth, carcass composition and meat quality. Livest. Prod. Sci. 2002, 76, 17–25. [CrossRef]
25. Wood, J.D.; Richardson, R.I.; Nute, G.R.; Fisher, A.V.; Campo, M.M.; Kasapidou, E.; Sheard, P.R.; Enser, M. Effects of fatty acids on meat quality: A review. Meat Sci. 2004, 66, 21–32. [CrossRef]
26. Belanche, A.; Kingston-Smith, A.H.; Griffith, G.W.; Newbold, C.J. A multi-kingdom study reveals the plasticity of the rumen microbiota in response to a shift from non-grazing to grazing diets in sheep. Front. Microbiol. 2019, 10, 1–17. [CrossRef] [PubMed]
27. French, P.; Stanton, C.; Lawless, F.; O’Riordan, E.G.; Monahan, F.J.; Caffrey, P.J.; Moloney, A.P. Fatty acid composition, including conjugated linoleic acid, of intramuscular fat from steers offered grazed grass, grass silage, or concentrate-based diets. J. Anim. Sci. 2000, 78, 2849–2855. [CrossRef]
28. Schmid, A.; Collomb, M.; Sieber, R.; Bee, G. Conjugated linoleic acid in meat and meat products: A review. Meat Sci. 2006, 73, 29–41. [CrossRef]
29. Fraser, M.D.; Speijers, M.H.M.; Theobald, V.J.; Fychan, R.; Jones, R. Production performance and meat quality of grazing lambs finished on red clover, lucerne or perennial ryegrass swards. Grass Forage Sci. 2004, 59, 345–356. [CrossRef]
30. De Brito, G.F.; Holman, B.W.B.; McGrath, S.R.; Friend, M.A.; van de Ven, R.; Hopkins, D.L. The effect of forage-types on the fatty acid profile, lipid and protein oxidation, and retail colour stability of muscles from White Dorper lambs. Meat Sci. 2017, 130, 81–90. [CrossRef]
31. Popova, T. Effect of the rearing system on the fatty acid composition and oxidative stability of the M. longissimus lumborum and M. semimembranosus in lambs. Small Rumin. Res. 2007, 71, 150–157. [CrossRef]
32. Popova, T.; Gonzalez-Barron, U.; Cadavez, V. A meta-analysis of the effect of pasture access on the lipid content and fatty acid composition of lamb meat. Food Res. Int. 2015, 77, 476–483. [CrossRef]
33. Burke, J.L.; Waghorn, G.C.; Brookes, I.M.; Kolver, E.S.; Atwood, G.T. An evaluation of sulla (Hedysarum coronarium) with pasture, white clover and lucerne for lambs. In Proceedings of the New Zealand Society of Animal Production, Palmerston North, New Zealand, 24–26 June 2002; Volume 62, pp. 152–156.
34. Speijers, M.H.M.; Fraser, M.D.; Theobald, V.J.; Haresign, W. The effects of grazing forage legumes on the quality of light lambs. 1. Growth, carcass composition and meat quality of finishing lambs. J. Agric. Sci. 2004, 142, 483–493. [CrossRef]
35. Niderkorn, V.; Martin, C.; Le Morvan, A.; Rochette, Y.; Awad, M.; Baumont, R. Associative effects between fresh perennial ryegrass and white clover on dynamics of intake and digestion in sheep. Grass Forage Sci. 2017, 72, 691–699. [CrossRef]
36. Amaral, G.A.; Kozloski, G.V.; Santos, A.B.; Castagnino, D.S. Metabolizable protein and energy supply in lambs fed annual ryegrass (Lolium multiflorum Lam.) supplemented with sources of protein and energy. J. Agric. Sci. 2011, 149, 519–527. [CrossRef]
37. Pérez-Ruchel, A.; Repetto, J.L.; Cajarville, C. Supplementing high-quality fresh forage to growing lambs fed a total mixed ration diet led to higher intake without altering nutrient utilization. Animal 2017, 11, 2175–2183. [CrossRef]
38. Fernández-Turren, G.; Arroyo, J.M.; Fontes, A.; Grignola, S.; Pérez-Ruchel, A.; Repetto, J.L.; Cajarville, C. Dry matter intake and ruminal environment of lambs fed a total mixed ration (amylaceous or fibrous) supplemented with fresh forage. In Proceedings of the 10th International Symposium on the Nutrition of Herbivores, Clermont-Ferrand, France, 2–6 September 2018; Volume 621.
39. Pérez-Ruchel, A.; Repetto, J.L.; Cajarville, C. Suitability of live yeast addition to alleviate the adverse effects due to the restriction of the time of access to feed in sheep fed only pasture. J. Anim. Physiol. Anim. Nutr. 2013, 97, 1043–1050. [CrossRef]
40. Jonker, A.; Cheng, L.; Molano, G.; Sandoval, E. Nitrogen partitioning in sheep offered three perennial ryegrass cultivars at two allowances in spring and autumn. In Proceedings of the New Zealand Society of Animal Production, Dunedin, New Zealand, 28 June 2015; Volume 75, pp. 74–78.
41. Catanese, F.; Distel, R.A.; Arzadun, M. Preferences of lambs offered Italian ryegrass (Lolium multiflorum L.) and barley (Hordeum vulgare L.) herbage as choices. Grass Forage Sci. 2009, 64, 304–309. [CrossRef]
42. Marley, C.L.; Fraser, M.D.; Fychan, R.; Theobald, V.J.; Jones, R. Effect of forage legumes and anthelmintic treatment on the performance, nutritional status and nematode parasites of grazing lambs. Vet. Parasitol. 2005, 131, 267–282. [CrossRef]
43. Tebot, I.; Cajarville, C.; Repetto, J.L.; Cirio, A. Supplementation with non-fibrous carbohydrates reduced fiber digestibility and did not improve microbial protein synthesis in sheep fed fresh forage of two nutritive values. Animal 2012, 6, 617–623. [CrossRef]
44. Cajarville, C.; Pérez, A.; Aguerre, M.; Britos, A.; Repetto, J.L. Effect of the timing of cut on ruminal environment of lambs consuming temperate pastures. In Proceedings of the Joint Annual Meeting Abstracts, American Society of Animal Science and American Dairy Science Association, Minneapolis, MN, USA, 9–13 July 2006; Volume 84, p. 103.
45. Niderkorn, V.; Martin, C.; Bernard, M.; Le Morvan, A.; Rochette, Y.; Baumont, R. Effect of increasing the proportion of chicory in forage-based diets on intake and digestion by sheep. Animal 2019, 13, 718–726. [CrossRef] [PubMed]
46. Sun, X.Z.; Hoskin, S.O.; Zhang, G.G.; Molano, G.; Muetzel, S.; Pinares-Patiño, C.S.; Clark, H.; Pacheco, D. Sheep fed forage chicory (Cichorium intybus) or perennial ryegrass (Lolium perenne) have similar methane emissions. Anim. Feed Sci. Technol. 2012, 172, 217–225. [CrossRef]
47. Sun, X.; Henderson, G.; Cox, F.; Molano, G.; Harrison, S.J.; Luo, D.; Janssen, P.H. Lambs fed fresh winter forage rape (Brassica napus L.) emit less methane than those fed perennial ryegrass (Lolium perenne L.), and possible mechanisms behind the difference. PLoS ONE 2015, 10, 1–16. [CrossRef] [PubMed]
48. Pérez Ruchel, A. pH, amoniacal, ácidos grasos volátiles y producción de proteína microbiana en el rumen de corderos, según el horario de corte de la pastura consumida. Bachelor’s Thesis, Facultad de Veterinaria, Universidad de la República, Montevideo, Uruguay, 2006.
49. Elizalde, J.C.; Santini, F.J.; Pasinato, A.M. The effect of stage of harvest on the process of digestion in cattle fed winter oats indoors. II. Nitrogen digestion and microbial protein synthesis. Anim. Feed Sci. Technol. 1996, 63, 245–255. [CrossRef]
50. Kokko, C.; Soder, K.J.; Brito, A.F.; Hover, R.C.; Berthiaume, R. Effect of time of cutting and maceration on nutrient flow, microbial protein synthesis, and digestibility in dual-flow continuous culture. J. Anim. Sci. 2013, 91, 1765–1774. [CrossRef]
51. Cohen, D.C. Degradability of crude protein from clover herbage in irrigated dairy production systems in northern Victoria. Aust. J. Agric. Res. 2001, 52, 415–425. [CrossRef]
52. Repetto, J.L.; Cajarville, C.; D’Alessandro, J.; Curbelo, A.; Soto, C.; Garin, D. Effect of wilting and ensiling on ruminal degradability of temperate grass and legume mixtures. Anim. Res. 2005, 54, 73–80. [CrossRef]
53. Pérez-Ruchel, A.; Repetto, J.L.; Cajarville, C. Feeding behavior and ruminal environment of lambs fed only pasture indoors or grazing. Veterinaria (Montev.) 2017, 54, 32–38.
54. Berthiaume, R.; Benchaar, C.; Chaves, A.V.; Tremblay, G.F.; Castonguay, Y.; Bertrand, A.; Belanger, G.; Milchaud, R.; Lafreniere, C.; McAllister, T.A.; et al. Effects of nonstructural carbohydrate concentration in alfalfa on fermentation and microbial protein synthesis in continuous culture. J. Dairy Sci. 2010, 93, 693–700. [CrossRef]
55. Niderkorn, V.; Baumont, R.; le Morvan, A.; Macheboeuf, D. Occurrence of associative effects between grasses and legumes in binary mixtures on in vitro rumen fermentation characteristics. J. Anim. Sci. 2011, 89, 1138–1145. [CrossRef]
56. Lüscher, A.; Mueller-Harvey, I.; Soussana, J.F.; Rees, R.M.; Peyraud, J.L. Potential of legume-based grassland-livestock systems in Europe: A review. Grass Forage Sci. 2014, 69, 206–228. [CrossRef] [PubMed]
57. Rutter, S.M. Diet preference for grass and legumes in free-ranging domestic sheep and cattle: Current theory and future application. Appl. Anim. Behav. Sci. 2006, 97, 17–35. [CrossRef]
58. Hill, J.; Chapman, D.F.; Cosgrove, G.P.; Parsons, A.J. Do ruminants alter their preference for pasture species in response to the synchronization of delivery and release of nutrients? Rangel. Ecol. Manag. 2009, 62, 418–427. [CrossRef]
59. Min, B.R.; Barry, T.N.; Attwood, G.T.; McNabb, W.C. The effect of condensed tannins on the nutrition and health of ruminants fed fresh temperate forages: A review. Anim. Feed Sci. Technol. 2003, 106, 3–19. [CrossRef]
60. Min, B.R.; Pinchak, W.E.; Fulford, J.D.; Puchala, R. Wheat pasture bloat dynamics, in vitro ruminal gas production, and potential bloat mitigation with condensed tannins. J. Anim. Sci. 2005, 83, 1322–1331. [CrossRef] [PubMed]
61. Waghrorn, G. Beneficial and detrimental effects of dietary condensed tannins for sustainable sheep and goat production—Progress and challenges. Anim. Feed Sci. Technol. 2008, 147, 116–139. [CrossRef]
62. Orr, R.; Griffith, B.; Rivero, M.; Lee, M. Livestock Performance for Sheep and Cattle Grazing Lowland Permanent Pasture: Benchmarking Potential of Forage-Based Systems. Agronomy 2019, 9, 101. [CrossRef]
63. Karnezos, T.P.; Matches, A.G.; Preston, R.L.; Brown, C.P. Corn supplementation of lambs grazing alfalfa. J. Anim. Sci. 1994, 72, 783–789. [CrossRef]
64. Marley, C.L.; Fraser, M.D.; Fisher, W.J.; Forbes, A.B.; Jones, R.; Moorby, J.M.; MacRae, J.C.; Theodorou, M.K. Effects of continuous or rotational grazing of two perennial ryegrass varieties on the chemical composition of the herbage and the performance of finishing lambs. Grass Forage Sci. 2007, 62, 255–264. [CrossRef]
65. Cosgrove, G.P.; Taylor, P.S.; Jonker, A. Sheep performance on perennial ryegrass differing in concentration of water soluble carbohydrate. J. NZ Grassl. 2015, 77, 123–130.
66. Proctor, L.E.; Craig, H.J.B.; McLean, N.J.; Fennessy, P.F.; Kerslake, J.I.; Behrent, M.J.; Chuah, J.C.L.; Campbell, A.W. The effect of grazing high-sugar ryegrass on lamb performance. In Proceedings of the New Zealand Society of Animal Production, Dunedin, New Zealand, 28 June 2015; Volume 75, pp. 235–238.
67. Fraser, T.; Rowarth, J. Legumes, herbs or grass for lamb performance? In Proceedings of the New Zealand Grassland Association, Oamaru, New Zealand, 21–24 October 1996; Volume 58, pp. 49–52.
68. Grace, C.; Lynch, M.B.; Sheridan, H.; Lott, S.; Fritch, R.; Boland, T.M. Grazing multispecies swards improves ewe and lamb performance. Animal 2019, 13, 1721–1729. [CrossRef] [PubMed]
69. Devincenzi, T.; Prunier, A.; Meteau, K.; Prache, S. How does barley supplementation in lambs grazing alfalfa affect meat sensory quality and authentication? Animal 2019, 13, 427–434. [CrossRef]
70. Jacques, J.; Berthiaume, R.; Cinq-Mars, D. Growth performance and carcass characteristics of Dorset lambs fed different concentrates: Forage ratios or fresh grass. Small Rumin. Res. 2011, 95, 113–119. [CrossRef]
71. McClure, K.E.; Van Keuren, R.W.; Althouse, P.G. Performance and carcass characteristics of weaned lambs either grazed on orchardgrass, ryegrass, or alfalfa or fed all-concentrate diets in drylot. J. Anim. Sci. 1994, 72, 3230–3237. [CrossRef] [PubMed]
72. Urioste, M.J.; Arroyo, J.M.; Pérez-Ruchel, A.; Fariña, V.; Fernandez, G.; Fontes, A.; Martinez, V.; Grignola, S.; Repetto, J.L.; Cajamarca, C. Dieta totalmente mezcladas vs. dietas mixtas difiriendo en la fuente de energía: Desempeño en engorde intensivo de corderos. Rev. Argent. Prod. Anim. 2017, 37, 140.
73. Wang, Y.; Douglas, G.B.; Waghrorn, G.C.; Barry, T.N.; Foote, A.G.; Purchas, R.W. Effect of condensed tannins upon the performance of lambs grazing Lotus corniculatus and lucerne (Medicago sativa). J. Agric. Sci. 1996, 126, 87–98. [CrossRef]
Animals 2020, 10, 382

74. Lee, M.R.F.; Jones, E.L.; Moorby, J.M.; Humphreys, M.O.; Theodorou, M.K.; Scollan, N.D. Production responses from lambs grazed on Lolium perenne selected for an elevated water-soluble carbohydrate concentration. Anim. Res. 2001, 50, 441–449. [CrossRef]

75. Farooq, M.Z.; Abdullah, M.; Ahmad, N.; Sattar, S. Effect of feeding frequency on dry matter intake weight gain feed conversion efficiency and its relation with body measurements in lohi lambs. Pak. J. Agric. Sci. 2017, 54, 689–692.

76. Golding, K.P.; Wilson, E.D.; Kemp, P.D.; Pain, S.J.; Kenyon, P.R.; Morris, S.T.; Hutton, P.G. Mixed herb and legume pasture improves the growth of lambs post-weaning. Anim. Prod. Sci. 2011, 51, 717–723. [CrossRef]

77. Allen, M.S.; Bradford, B.J.; Oba, M. Board-invited review: The hepatic oxidation theory of the control of feed intake and its application to ruminants. J. Anim. Sci. 2009, 87, 3317–3334. [CrossRef]

78. Rhind, S.M.; Archer, Z.A.; Adam, C.L. Seasonality of food intake in ruminants: Recent developments in understanding. Nutr. Res. Rev. 2002, 15, 43–65. [CrossRef] [PubMed]

79. Griffiths, W.M.; Hodgson, J.; Arnold, G.C. The influence of sward canopy structure on foraging decisions by grazing cattle. I. Patch selection. Grass Forage Sci. 2003, 58, 112–124. [CrossRef]

80. Hodgson, J. The control of herbage intake in the grazing ruminant. Proc. Nutr. Soc. 1985, 44, 339–346. [CrossRef] [PubMed]

81. Manteca, X.; Smith, A.J. Effects of poor forage conditions on the behaviour of grazing ruminants. Trop. Anim. Health Prod. 1994, 26, 129–138. [CrossRef]

82. Burnett, V.E.; Seymour, G.R.; Norrg, S.; Jacobs, J.L.; Ponnampalam, E.N. Lamb growth performance and carcass weight from rotationally grazed perennial pasture systems compared with annual pasture systems with supplements. Anim. Prod. Sci. 2012, 52, 248–254. [CrossRef]

83. Forbes, J.M. A personal view of how ruminant animals control their intake and choice of food: Minimal total discomfort. Nutr. Res. Rev. 2007, 20, 132–146. [CrossRef]

84. Arthington, J.D.; Brown, W.F. Estimation of feeding value of four tropical forage species at two stages of maturity. J. Anim. Sci. 2005, 83, 1726–1731. [CrossRef]

85. Jung, H.G.; Allen, M.S. Characteristics of plant cell walls affecting intake and digestibility of forages by ruminants. J. Anim. Sci. 1995, 73, 2774–2790. [CrossRef]

86. Marley, C.L.; Fychan, R.; Fraser, M.D.; Sanderson, R.; Jones, R. Effects of feeding different ensiled forages on the productivity and nutrient-use efficiency of finishing lambs. Grass Forage Sci. 2007, 62, 1–12. [CrossRef]

87. Dewhurst, R.J.; Delaby, L.; Moloney, A.; Boland, T.; Oba, M. Board-invited review: The hepatic oxidation theory of the control of feed intake and its application to ruminants. J. Anim. Sci. 2009, 87, 3317–3334. [CrossRef]

88. Pasha, T.N.; Prigge, E.C.; Russell, R.W.; Bryan, W.B. Influence of moisture content of forage diets on intake and digestion by sheep. J. Anim. Sci. 1994, 72, 2455–2463. [CrossRef]

89. Kenney, P.A.; Black, J.L.; Colebrook, W.F. Factors affecting diet selection by sheep. 3. Dry matter content and particle length of forage. Aust. J. Agric. Res. 1984, 35, 831–838. [CrossRef]

90. Feng, C.; Ding, S.; Zhang, T.; Li, Z.; Wang, D.; Wang, L.; Liu, C.; Sun, J.; Peng, F. High plant diversity stimulates foraging motivation in grazing herbivores. Basic Appl. Ecol. 2016, 17, 43–51. [CrossRef]

91. Cabrera Estrada, J.I.; Delagarde, R.; Faverdin, P.; Peyraud, J.L. Dry matter intake and eating rate of grass- and chicory (Cichorium intybus) and red clover (Trifolium pratense L.) into naturalized pastures in eastern Canada. Anim. Feed Sci. Technol. 2003, 112, 147–155. [CrossRef]

92. Cabrera Estrada, J.I.; Delagarde, R.; Faverdin, P.; Peyraud, J.L. Dry matter intake and eating rate of grass- and chicory (Cichorium intybus) and red clover (Trifolium pratense L.) into naturalized pastures in eastern Canada. Anim. Feed Sci. Technol. 2003, 112, 147–155. [CrossRef]

93. Cabrer, J.I.; Delagarde, R.; Faverdin, P.; Peyraud, J.L. Dry matter intake and eating rate of grass by dairy cows is restricted by internal, but not external water. Anim. Feed Sci. Technol. 2004, 114, 59–74. [CrossRef]

94. Pasha, T.N.; Repetto, J.L.; Hernández, N.; Pérez-Ruchel, A.; Cajamar, C. Restricting the time of access to fresh forage reduces intake and energy balance but does not affect the digestive utilization of nutrients in beef heifers. Anim. Feed Sci. Technol. 2017, 226, 103–112. [CrossRef]
96. Iason, G.R.; Mantecon, A.R.; Sim, D.A.; Gonzalez, J.; Foreman, E.; Bermudez, F.F.; Elston, D.A. Can grazing sheep compensate for a daily foraging time constraint? J. Anim. Ecol. 1999, 68, 87–93. [CrossRef]

97. Luciano, G.; Biondi, L.; Pagano, R.I.; Scerra, M.; Vasta, V.; López-Andrés, P.; Valenti, B.; Lanza, M.; Priolo, A.; Avondo, M. The restriction of grazing duration does not compromise lamb meat colour and oxidative stability. Meat Sci. 2012, 92, 30–35. [CrossRef]

98. Allden, W.G.; McDWhittaker, I.A. The determinants of herbage intake by grazing sheep: The interrelationship of factors influencing herbage intake and availability. Aust. J. Agric. Res. 1970, 21, 755–766. [CrossRef]

99. Pain, S.J.; Corkran, J.R.; Kenyon, P.R.; Morris, S.T.; Kemp, P.D. The influence of season on lambs’ feeding preference for plantain, chicory and red clover. Anim. Prod. Sci. 2015, 55, 1241–1249. [CrossRef]

100. Brito, A.F.; Tremblay, G.F.; Bertrand, A.; Castonguay, Y.; Belanger, G.; Milchaud, R.; Lapierre, H.; Benchaar, C.; Petit, H.V.; Ouellet, D.R.; et al. Alfalfa cut at sundown and harvested as baleage improves milk yield of late-lactation dairy cows. J. Dairy Sci. 2008, 91, 3968–3982. [CrossRef] [PubMed]

101. Pelletier, S.; Tremblay, G.F.; Belanger, G.; Bertrand, A.; Castonguay, Y.; Pageau, D.; Drapeau, R. Forage nonstructural carbohydrates and nutritive value as affected by time of cutting and species. Agron. J. 2010, 102, 1388–1398. [CrossRef]

102. Morin, C.; Belanger, G.; Tremblay, G.F.; Bertrand, A.; Castonguay, Y.; Drapeau, R.; Milchaud, R.; Berthiaume, R.; Allard, G. Short Communication: Diurnal variations of nonstructural carbohydrates and nutritive value in timothy. Can. J. Plant Sci. 2012, 92, 883–887. [CrossRef]

103. Cajarville, C.; Britos, A.; Errandonea, N.; Gutiérrez, L.; Cozzolino, D.; Repetto, J.L. Diurnal changes in water-soluble carbohydrate concentration in lucerne and tall fescue in autumn and the effects on in vitro fermentation. N. Zeal. J. Agric. Res. 2015, 58, 281–291. [CrossRef]

104. Ciavarella, T.A.; Dove, H.; Leury, B.J.; Simpson, R.J. Diet selection by sheep grazing Phalaris aquatica L. pastures of differing water-soluble carbohydrate content. Aust. J. Agric. Res. 2000, 51, 757–764. [CrossRef]

105. Edwards, G.R.; Parsons, A.J.; Bryant, R.H. Manipulating dietary preference to improve animal performance. Aust. J. Exp. Agric. 2008, 48, 773–779. [CrossRef]

106. Hall, M.B.; Huntington, G.B. Nutrient synchrony: Sound in theory, elusive in practice. J. Anim. Sci. 2008, 86, 287–292. [CrossRef]

107. Lee, M.R.F.; Merry, R.J.; Davies, D.R.; Moorby, J.M.; Humphreys, M.O.; Theodorou, M.K.; MacRae, J.C.; Scollan, N.D. Effect of increasing availability of water-soluble carbohydrates on in vitro rumen fermentation. Anim. Feed Sci. Technol. 2003, 104, 59–70. [CrossRef]

108. Qiao, G.H.; Xiao, Z.G.; Li, Y.; Li, G.J.; Zhao, L.C.; Xie, T.M.; Wang, D.W. Effect of diet synchrony on rumen fermentation, production performance, immunity status and endocrine in Chinese Holstein cows. Anim. Prod. Sci. 2018, 59, 664–672. [CrossRef]

109. Valkeners, D.; Théwis, A.; Piron, F.; Beckers, Y. Effect of imbalance between energy and nitrogen supplies on microbial protein synthesis and nitrogen metabolism in growing double-muscled Belgian Blue bulls. J. Anim. Sci. 2004, 82, 1818–1825. [CrossRef] [PubMed]

110. Sinclair, L.A.; Garnsworthy, P.C.; Newbold, J.R.; Butterly, P.J. Effects of synchronizing the rate of dietary energy and nitrogen release in diets with a similar carbohydrate composition on rumen fermentation and microbial protein synthesis in sheep. J. Agric. Sci. 1995, 124, 463–472. [CrossRef]

111. Witt, M.W.; Sinclair, L.A.; Wilkinson, R.G.; Butterly, P.J. The effects of synchronizing the rate of dietary energy and nitrogen supply to the rumen on the production and metabolism of sheep: Food characterization and growth and metabolism of ewe lambs given food ad libitum. Anim. Sci. 1999, 69, 223–235. [CrossRef]

112. Richardson, J.M.; Wilkinson, R.G.; Sinclair, L.A. Synchrony of nutrient supply to the rumen and dietary energy source and their effects on the growth and metabolism of lambs. J. Anim. Sci. 2003, 81, 1332–1347. [CrossRef]

113. Henning, P.H.; Steyn, D.G.; Meissner, H.H. Effect of synchronization of energy and nitrogen supply on ruminal characteristics and microbial growth. J. Anim. Sci. 1993, 71, 2516–2528. [CrossRef]

114. Trevaskis, L.M.; Fulkerson, W.J.; Gooden, J.M. Provision of certain carbohydrate-based supplements to pasture-fed sheep, as well as time of harvesting of the pasture, influences pH, ammonia concentration and microbial protein synthesis in the rumen. Aust. J. Exp. Agric. 2001, 41, 21–27. [CrossRef]

115. Bach, A.; Calsamiglia, S.; Stern, M.D. Nitrogen metabolism in the rumen. J. Dairy Sci. 2005, 88, E9–E21. [CrossRef]
116. Santana, A.; Cajarville, C.; Mendoza, A.; Repetto, J.L. Combination of legume-based herbage and total mixed ration (TMR) maintains intake and nutrient utilization of TMR and improves nitrogen utilization of herbage in heifers. *Animal* 2017, 11, 616–624. [CrossRef]

117. Felix, T.L.; Zerby, H.N.; Moeller, S.J.; Loerch, S.C. Effects of increasing dried distillers grains with solubles on performance, carcass characteristics, and digestibility of feedlot lambs. *J. Anim. Sci.* 2012, 90, 1356–1363. [CrossRef]

118. Jaborek, J.R.; Zerby, H.N.; Moeller, S.J.; Wick, M.P.; Fluharty, F.L.; Garza, H.; Garcia, L.G.; England, E.M. Effect of energy source and level, and animal age and sex on meat characteristics of sheep. *Small Rumin. Res.* 2018, 166, 53–60. [CrossRef]

119. Bargo, F.; Muller, L.D.; Delahoy, J.E.; Cassidy, T.W. Performance of high producing dairy cows with three different feeding systems combining pasture and total mixed rations. *J. Dairy Sci.* 2002, 85, 2948–2963. [CrossRef]

120. Mendoza, A.; Cajarville, C.; Repetto, J.L. Digestive response of dairy cows fed diets combining fresh forage with a total mixed ration. *J. Dairy Sci.* 2016, 99, 8779–8789. [CrossRef] [PubMed]

121. Pastorini, M.; Pomié, N.; Repetto, J.L.; Mendoza, A.; Cajarville, C. Productive performance and digestive response of dairy cows fed different diets combining a total mixed ration and fresh forage. *J. Dairy Sci.* 2019, 102, 4118–4130. [CrossRef] [PubMed]

122. Britos, A.; Dearmas, B.; Repetto, J.L.; Cajarville, C. PSX-2 In vitro evaluation of the inclusion of a fibrous concentrate to partial Total Mixed Ration for lambs. *J. Anim. Sci.* 2018, 96, 414–415. [CrossRef]

© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).