Far UV-C radiation: An emerging tool for pandemic control

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ABSTRACT

Far UV-C, informally defined as electromagnetic radiation with wavelengths between 200 and 230 nm, has characteristics that are well-suited to control of airborne pathogens. Specifically, Far UV-C has been shown to be highly effective for inactivation of airborne pathogens; yet this same radiation has minimal potential to cause damage to human skin and eye tissues. Critically, unlike UV-B, Far UV-C radiation does not substantially penetrate the dead cell layer of skin (stratum corneum) and does not reach germinative cells in the basal layer. Similarly, Far UV-C radiation does not substantially penetrate through corneal epithelium of the eye, thereby preventing exposure of germinative cells within the eye. The most common source of Far UV-C radiation is the krypton chloride excimer (KrCl) lamp, which has a primary emission centered at 222 nm. Ozone production from KrCl lamps is modest, such that control of indoor ozone from these systems can be accomplished easily using conventional ventilation systems. This set of characteristics offers the potential for Far UV-C devices to be used in occupied spaces, thereby allowing for improved effectiveness for inactivation of airborne pathogens, including those that are responsible for COVID-19.

KEYWORDS Air disinfection; Coronavirus; disinfection; Far UV-C; ozone; safety

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Introduction

Beginning as early as the 1930s, UV-C radiation (100–280 nm) has been applied for disinfection of air and surfaces (Wells & Fair, 1935; Reed, 2010). Since that time, UV-C-based disinfection systems have been demonstrated to be effective for reduction of disease transmission, even for highly contagious viral diseases, such as measles, influenza, and the common cold.

Conventional UV-C applications have been based on low-pressure mercury lamps, which are characterized by a primary emission line at 253.7 nm. UV-C radiation at this wavelength causes inactivation of pathogens via damage to nucleic acids (DNA and RNA). However, this same radiation is known to cause damage to mammalian skin and eyes. Consequently, conventional UV-C systems for disinfection of air and surfaces must be designed and operated in a manner that minimizes exposure of human tissues.

Recent research has demonstrated that Far UV-C radiation (200–230 nm) has minimal potential to damage mammalian skin and eye tissues. Yet, Far UV-C radiation tends to be at least as effective as conventional UV-C radiation for inactivation of most microbial and viral pathogens, including those that are responsible for COVID-19 and other diseases that are transmitted via aerosols. And while Far UV-C sources will promote photochemical generation of ozone from molecular oxygen, the rates of ozone formation from these sources are modest, such that ozone concentrations in the vicinity of Far UV-C sources can be effectively controlled through conventional ventilation.

This combination of characteristics indicates that Far UV-C radiation may provide opportunities for new modes of application of UV-based disinfection systems. Specifically, it appears that it may be possible to implement Far UV-C radiation in systems that allow greater exposure of human tissues to the output from these UV devices than with conventional UV-C systems. This has the potential to fundamentally change how UV-C radiation can be applied for disinfection of air and surfaces.

Much of the knowledge base around Far UV-C radiation, its sources, and its application has emerged over the last 3–5 years. The goal of this article is to provide a comprehensive review of the current state-of-knowledge related to Far UV-C sources, their application for disinfection of air and surfaces, and the safety concerns related to Far UV-C radiation. Research needs related to Far UV-C sources and their applications are also presented.

UV radiation and far UV-C

UV radiation is defined as a band of electromagnetic radiation with wavelengths ranging from 100 to 400 nm (see Fig. 1). The UV spectrum has been divided into three major photobiological bands: UV-A (315–400 nm), UV-B (280–315 nm), and UV-C (100–280 nm); vacuum-UV (VUV, 100–200 nm) is defined as a sub-band of UV-C. For purposes of this review, we further sub-divide UV-C into the informally defined categories of conventional germicidal UV irradiation (UVGI, 250–280 nm) and Far UV-C (200–230 nm) radiation.¹

Sources of far UV-C radiation

UV-C disinfection processes depend on generation of radiation from artificial sources. Conventional UVGI systems are based on low-pressure mercury lamps, with primary and secondary emission lines at 253.7 and 184.9 nm, respectively; the latter emission line is normally eliminated by use of an envelope around the lamp that absorbs short wavelength radiation that is responsible for ozone generation. For Far UV-C radiation, the most common sources are barrier discharge excimer lamps (Sosnin et al., 2006) first developed in the 1980s (Tarasenko & Sosnin, 2012). In these devices, an electrical potential is applied across a sealed chamber containing a rare
gas-halogen mixture, thereby exciting the gas into a plasma state. The resulting excited dimer (i.e., ‘excimer’) then undergoes an electronic transition to emit quasi-monochromatic radiation. The plasma may contain several excimers, leading to primary and secondary (weaker) emission peaks. To a limited degree, the contributions from different electronic transitions can be enhanced or suppressed by adjusting gas pressure and composition to ‘tune’ lamp output (Shuaibov et al., 2004).

In the context of Far UV-C radiation, most recent activity has focused on krypton-chloride excimer (KrCl*) lamps. Emission spectra vary somewhat depending on the lamp design, but KrCl* lamps typically exhibit a primary peak at \(\sim 222\) nm with full-width-half-maximum \(\sim 4\) nm; in addition there are also secondary emissions extending to 200 nm and below, as well as at wavelengths longer than the primary peak from other KrCl* and Cl2* transitions (Sosnin et al., 2006),
extending to ~300 nm and above, with only trace emission at longer wavelengths, including in the UV-B range (Fig. 2). These secondary emissions can represent up to ~15% of the total power output of a typical unfiltered KrCl* lamp, as illustrated by their unfiltered output spectra (see Fig. 2).

The primary peak from a KrCl* lamp centered at 222 nm is well-suited to the applications that are discussed herein. Radiation at longer wavelengths can be problematic in terms of human eye and skin exposure risks, while shorter wavelengths can be problematic due to potential ozone generation. Consequently, most practical applications of KrCl* lamps include optical filters to reduce contributions of radiation from the secondary peaks (also illustrated in Fig. 2). A summary and comparison of basic characteristics of low-pressure mercury lamps and krypton chloride excimer lamps are presented in Table SI-1.

Other sources of Far UV-C radiation have been developed, including metallic vapor lamps (Zelikoff et al., 1952), optically-filtered medium-pressure mercury lamps (Beck et al., 2015), solid state emitters (e.g., Far UV-C LEDs), and lasers. To date, applications of these alternative Far UV-C sources have been limited to small-scale applications, often in research.

**Inactivation by UV-C/far UV-C radiation**

**Mechanisms of inactivation**

Wavelength and total UV exposure (commonly referred to as UV dose or fluence) are the two key parameters that control UV disinfection (Harm, 1980; Jagger, 1967). The germicidal efficacy of UV-C varies with wavelength and each microbe or virus being targeted has a unique response described by its germicidal action spectrum. Though germicidal action spectra among microbial and viral species are unique (Beck et al., 2015), a local peak in the 250–270 nm region is common, as this is where UV-C most effectively causes genomic damage resulting in pathogen inactivation. This fact, and the widespread availability of mercury vapor-based lamps (emitting predominantly at 254 nm), led to a focus on conventional mercury vapor lamps for UVGI systems.

Wavelengths in the Far UV-C region (200–230 nm) are highly efficient for pathogen inactivation due to the absorption of radiation by both nucleic acids and proteins (see Fig. SI-1); however, less is known about disinfection efficacy in the Far UV-C because sources operating at this region have only recently emerged as commercially viable. Far UV-C absorbance by nucleic acids and proteins can photochemically damage both biomolecule types, providing dual pathways to microorganism and virus inactivation.

While shorter wavelength Far UV-C photons can readily inactivate target pathogens, they can also be more highly absorbed by environmental (water, aerosols, and air) media, resulting in reduced photon flux and lower inactivation. For example, airborne viruses are often embedded within aerosol droplets, which themselves can contain proteins at relatively high concentrations. The presence of any absorbing compounds (e.g., proteins) can inhibit the ability of UV-C radiation, including radiation in the Far UV-C range, to penetrate aerosols. In quantitative terms, aerosol diameter and composition will control delivery of photons to aerosolized target pathogens (Barancheshme et al., 2021). The effects of absorbance by proteins and other constituents in aerosols are expected to be minimal due to the short optical pathlength, but improvement in understanding the effects of this behavior more quantitatively is an area for future research.

**UV-C inactivation data**

UV-C irradiation is effective for inactivation of microbial and viral pathogens in air, water, and on surfaces. Coronaviruses specifically are rapidly inactivated by UV-C radiation, as illustrated in Fig. 3. Only data from studies conducted employing accepted guidelines or best practice and
methods for measuring UV dose and for assessing viral inactivation were included in this graphical representation (Bolton & Linden, 2003). In aqueous (liquid) suspensions of viruses, high levels of inactivation are achieved with irradiation from a conventional UV254 lamps, which was also true for sources that emit at other UV-C wavelengths, including Far UV-C.

At 254 nm approximately 1 log10 reduction of coronaviruses is achieved for each 2 mJ/cm² delivered UV-C fluence (dose). For comparison, other human pathogenic viruses, such as poliovirus and rotavirus require about 4–5 times that amount (i.e., 8–10 mJ/cm²) for each log10 unit reduction (Masjoudi et al., 2021).

UV222 irradiation is at least as effective as UV254 irradiation for inactivation of viruses, with approximately 1 log10 reduction of coronaviruses achieved for each 1 mJ/cm² of delivered UV-C.

Figure 3. (Top) UV254 inactivation of coronaviruses, in aqueous suspension (Bianco et al., 2020; Ma, Gundy, et al., 2021; Ma, Linden, et al., 2021; Storm et al., 2020). Genomic model predictions of coronavirus responses are indicated by dashed lines (Pendyala et al., 2020; Rockey et al., 2020; Sagripanti & Lytle, 2020). (Bottom) UV222 inactivation of coronaviruses in aqueous suspension (Jung et al., 2021; Ma, Linden, et al., 2021; Robinson et al., 2021).
fluence or less. In other words, irradiation at 222 nm provides roughly twice the rate of inactivation as observed at 254 nm.

Genome size and make-up are key factors governing the response of viruses to UV-C radiation (Lytle & Sagripanti, 2005; Pendyala et al., 2020; Rockey et al., 2020; Sagripanti & Lytle, 2020). In general, larger genomes provide more targets for photochemical damage; therefore, viruses and other pathogens with relatively large genomes tend to be inactivated relatively quickly. The type of genome (either single-stranded [ss], double-stranded [ds], RNA, or DNA) also influences the response of viruses to UV-C exposure. Coronaviruses have the largest known genomes among ss-RNA viruses; this is likely an important contributing factor to the facile inactivation of coronaviruses by UV-C exposure. Viruses with ds DNA genomes are often relatively resistant to UV-C exposure because the complementary nucleic acid strand can facilitate repair within its host (Boszko & Rainbow, 1999; Rodriguez et al., 2014; Shin et al., 2009). The presence of a viral envelope can influence the susceptibility of viruses to exposure to some physical agents, such as heat or shear forces. However, the presence of a viral envelope may not have any influence on the response of viruses to UV-C exposure. A summary of the genetic properties of the viruses and bacteriophage reported herein is presented in Table 1.

Inactivation models have recently been developed based on genomic properties of viruses for UV254 irradiation (Pendyala et al., 2020; Rockey et al., 2020; Sagripanti & Lytle, 2020). These models yield predictions of viral inactivation at 254 nm that are similar to reported measurements; moreover, the predictions of these independently-developed models are consistent with each other. Currently, no models have been developed to characterize UV dose-response behavior for 222 nm UV irradiation. The inclusion of terms to address protein damage in Far UV-C inactivation (Beck et al., 2018) will complicate predictions that are based solely on genomic characteristics. Enhanced inactivation of viruses at Far UV-C wavelengths has been shown to be due to protein damage (Beck et al., 2018), as represented by studies on respiratory adenoviruses illustrated in Fig. SI-2 (Beck et al., 2014; Linden et al., 2007).

The kinetics of coronavirus inactivation by UV irradiation on surfaces or in aerosols appear to be at least as fast as those for the same virus in aqueous suspension, with greater than 1 log10 reduction achieved for each 2 mJ/cm² delivered UV fluence (see Fig. SI-3). More generally, inactivation of viruses suspended in water or other aqueous media appears to provide an equivalent or slightly more conservative estimate of the intrinsic inactivation response that can be achieved on surfaces or in aerosols, based on these and other available UV inactivation data.

While these data suggest that UV inactivation of viruses on surfaces or in aerosols is expected to be as fast or faster than in aqueous suspension, there are other complicating factors.

### Table 1. Summary of genetic properties of human viruses and bacteriophage described herein.

| Virus                  | Genome Type | Enveloped or non-enveloped | Genome size (kilobases or kilobase pairs) | NCBI accession |
|------------------------|-------------|----------------------------|------------------------------------------|----------------|
| SARS-CoV-2             | ssRNA       | Enveloped                  | 29.8 kb                                  | MN908947       |
| SARS-CoV-1             | ssRNA       | Enveloped                  | 29.7 kb                                  | NC_004718      |
| MERS                   | ssRNA       | Enveloped                  | 30.1 kb                                  | NC_019843.3    |
| MHV                    | ssRNA       | Enveloped                  | 31.3 kb                                  | AY700211       |
| HuCoV-229E             | ssRNA       | Enveloped                  | 27.3 kb                                  | NC_002645      |
| HuCoV-OC43             | ssRNA       | Enveloped                  | 30.7 kb                                  | NC_006213      |
| Influenza A (H1N1)     | segmented ssRNA | Enveloped                  | 13.5 kb                                  | NC_002016-002023 |
| Adenovirus (Type 2)    | dsDNA       | Non-enveloped              | 35.9 kb                                  | AC_000007      |
| Φ6 Bacteriophage       | segmented dsRNA | Enveloped                  | 13.4 kb                                  | NC_000174-000176 |
| T1 Bacteriophage       | dsDNA       | Non-enveloped              | 48.8 kb                                  | NC_005833      |
| T1UV Bacteriophage     | dsDNA       | Non-enveloped (Assumed similar to T1) | n/a                                      |
| Qi Bacteriophage       | ssRNA       | Non-enveloped              | 4.2 kb                                   | NC_001890      |
| M52 Bacteriophage      | ssRNA       | Non-enveloped              | 3.6 kb                                   | NC_001417      |

Sequencing information can be found in [https://www.ncbi.nlm.nih.gov/](https://www.ncbi.nlm.nih.gov/), using NCBI accession number.
Inactivation of pathogens (including viruses) in aerosols and on surfaces by UV-C irradiation is also influenced by the optical properties of the surrounding media, including absorbance, reflectance, and refraction. As a result, quantifying UV-C dose delivery to (viral) pathogens in aerosols and on surfaces can be challenging. Current research is expanding on these initial findings to include other surface types and conditions.

Validation and modeling

UV-C disinfection experimental protocols for pathogen inactivation in aqueous suspensions are well standardized to deliver a controllable, quantifiable, single-valued dose to a viral population via a (pseudo) collimated beam (Blatchley III, 1997; Bolton et al., 2015; Bolton & Linden, 2003). For airborne and surface-associated viruses, no such standard currently exists, although guidelines and laboratory devices have been proposed and developed (Welch et al., 2018).

Practical applications of UV-C radiation for disinfection will be influenced by both the intrinsic kinetics of inactivation and the distribution of UV doses delivered to the pathogen population. Well-defined methods have been developed for quantification of the dose distribution delivered by UV photoreactors that are used for disinfection of water. These methods, which now represent the industry standards for design and analysis of UV disinfection systems, generally involve integrated applications of computational fluid dynamics, fluence rate field models, and a model to describe the intrinsic kinetics of inactivation (Ahmed et al., 2018, 2019; Chiu et al., 1999; Ducoste, Linden, et al., 2005; Ducoste, Liu, et al., 2005; Imoberdorf et al., 2008; Lyn et al., 1999; Naunovic et al., 2008; Sozzi & Taghipour, 2006). Analogous methods have been applied for evaluation of UV-C systems that are used for disinfection of air (Buchan et al., 2020). Similar numerical models may be developed to simulate dynamic behavior of UV-C disinfection systems used to disinfect surfaces. Like all numerical models, validation based on physical measurements is required. Research in this area is likely to continue to evolve as interest and demand for UV-based disinfection systems grows.

Beyond these numerical models, it is also important that empirical (experiment-based) methods be developed and applied for testing and validation of UV-C systems that are used to disinfect air and surfaces. It is likely that these methods will be derived from the analogous methods that have been standardized for validation of UV disinfection systems that are applied in water treatment in the USA (Emerick & Tchobanoglous, 2012; EPA, 2006) and in western Europe (German & Standardization I. F., 2020a, 2020b; ÖNORM, 2003, 2020a, 2020b). These standardized validation protocols are based on the use of surrogate challenge microbes or viruses to quantify disinfection efficacy. Given the history and success of these applications, which are based on measurements of inactivation of nonpathogenic challenge agents, it is likely that testing and validation protocols for UV disinfection systems for air and water will adopt similar strategies.

Surrogates and indicators of UV inactivation of respiratory viruses

A critical need exists to develop standardized protocols for validation of disinfection efficacy among UV systems designed for disinfection of air and surfaces. These protocols will almost certainly be based on surrogates for airborne pathogens that do not require high levels of biosafety. A good surrogate will be nonpathogenic to humans, easy to culture and assay, no more sensitive to UV exposure than the target pathogen at the wavelength(s) emitted by the UV source of interest, and not exhibit post-irradiation repair. Ideally, this viral surrogate will also be from a similar pathogen class and share genetic and structural similarities, such as nucleic acid make-up and the presence/absence of an envelope. UV inactivation of select viral surrogates can represent UV inactivation of coronaviruses at either 254 or 222 nm, as illustrated in Fig. 4.
T1UV phage, T1 phage, and Qβ phage all provide conservative representations of the rate of inactivation of coronaviruses at 254 nm. For 222 nm irradiation, T1UV, T1, and Φ6 phage also provide conservative representations of the rate of inactivation of coronaviruses. MHV, a murine coronavirus with high genetic similarity to SARS-CoV-2, shows similar inactivation to it for both 254 and 222 nm irradiation. As noted in Table 1, T1 and T1UV phage are dsDNA viruses and are non-enveloped. Qβ phage is a ssRNA virus, also non-enveloped. Φ6 phage is an enveloped dsRNA virus. Data presented in Fig. 4 demonstrate that Φ6 is only about 10% as sensitive to UV_{254} as the coronaviruses, whereas at 222 nm, Φ6 represents a slightly conservative surrogate for the coronaviruses. As such, it appears that Φ6 phage may represent a relevant surrogate for UV_{222} systems but not for UV_{254} systems. While the T1, T1UV, Qβ, and Φ6 phage are all easy to
propagate and assay using bacterial hosts, use of MHV requires mammalian cell culture infectivity assays which require more complex laboratory facilities and skilled virologists.

Conservatism is desirable in the selection of a viral surrogate, but overly conservative surrogate selections can provide misleading information about the performance of UV disinfection systems and may disqualify some technologies that can provide effective protection against airborne pathogens. This is not as simple a case as providing an extra degree of caution in the definition of standards for UV disinfection equipment. A minimum standard that is set too high will limit or negate the implementation of demonstrably effective disinfection apparatus without providing a viable alternative in their place; under such circumstances, a ‘good enough’ system that meets an established standard may not be implemented and result in an absence of intervention where an efficacious system could have been deployed. Therefore, it is generally desirable to select a surrogate that demonstrates UV dose–response behavior that is as close as possible to that of the target pathogen. Examples of phage that will meet this objective include T1UV at 254 nm and T1 at 222 nm.

For some applications, it may be desirable to quantify the performance of a UV disinfection system in a public setting using ambient airborne bacteria or viruses. This would involve the use of indicator viruses or bacteria routinely found in the public setting of interest. Desirable characteristics among indicators that may be used for this purpose include presence at sufficient concentration to allow quantification of disinfection efficacy, availability of a simple yet selective assay for viability or infectivity, lack of pathogenicity, and similarity in UV dose–response behavior to that of the target pathogen. These characteristics are similar to those of surrogates, described above. However, an important distinction is that tests with an indicator would involve ambient microbes or viruses in a public setting, whereas surrogates would be added to the air space in a controlled laboratory facility for purposes of testing.

The vast majority of UV dose–response data sets have been generated using aqueous suspensions of the target microbe or virus. However, there is evidence to indicate that air-exposed microbes or viruses may respond differently than those in aqueous suspension; more specifically, there is evidence to indicate that air-exposed microbes and viruses are inactivated faster than in aqueous suspension (Fletcher, 2004; McDevitt et al., 2007; Peccia et al., 2001; Xu et al., 2005; McDevitt et al., 2012; Lim & Blatchley, 2012). Figure SI-4 illustrates the UV222 dose–response behavior of three aerosolized viruses and an aerosolized bacterium.

The data presented in Fig. SI-4 also indicate that Staphylococcus aureus may represent an appropriate, conservative surrogate for coronaviruses. In general, bacteria are easier to culture and assay than viruses as they do not require a host cell line or cell culture facilities to evaluate viability. However, many bacteria have access to repair pathways that do not exist for viruses, which could complicate the use of bacteria as a viral surrogate. Given the broad differences in the types and suitability of acceptable viruses and an aerosolized bacterium.

The data presented in Fig. SI-4 also indicate that Staphylococcus aureus may represent an appropriate, conservative surrogate for coronaviruses. In general, bacteria are easier to culture and assay than viruses as they do not require a host cell line or cell culture facilities to evaluate viability. However, many bacteria have access to repair pathways that do not exist for viruses, which could complicate the use of bacteria as a viral surrogate. Given the broad differences in the types and suitability of acceptable surrogates, a tailored approach to future validation protocols for these technologies should be taken across the range of possible Far UV-C applications.

In a recent study, measurements were made in a room-scale bioaerosol chamber of aerosolized airborne pathogen reduction by filtered 222 nm Far UV-C (Eadie et al., 2021). Airborne pathogen reduction, using S. aureus as a surrogate, was measured in a 4.2 × 3.4 × 2.3 m room containing a continuous source of aerosolized airborne pathogens, typical ventilation (3 air changes per hour [ACH]), and ceiling-mounted Far UV-C lamps. The experiments were performed at Far UV-C exposure conditions consistent with the current 222 nm threshold limit values (TLVs) for UV-C exposure and at exposure conditions consistent with proposed new TLVs (discussed below). Using lamps operated within the current TLV, a > 92% reduction in live airborne pathogens was achieved after 15 min – equivalent to an additional 35 effective ACH (eACH) (see Fig. 5). Using exposure conditions consistent with the proposed new TLV (Sliney & Stuck, 2021) a 99.9% reduction in viable airborne pathogens was achieved after 5 min.
Safety aspects of Far UV-C radiation exposure of human eyes and skin

The recent interest in applying Far UV-C to germicidal applications has stimulated investigations of health and safety of Far UV-C. Collectively, the studies demonstrate that exposures of both the eye and skin at wavelengths less than approximately 230 nm are substantially safer than longer UV-C wavelengths (Barnard et al., 2020; Buonanno et al., 2017, 2013, 2016, 2021; Cadet, 2020; Fukui et al., 2020; Hanamura et al., 2020; Hickerson et al., 2021; Kaidzu et al., 2019; Woods et al., 2015; Yamano et al., 2020). The experimental data are expected from the biophysical knowledge that Far UV-C radiation has much shorter penetration depths than longer wavelength UV-C radiation (Finlayson et al., 2021). UV-C radiation at wavelengths less than approximately 230 nm is strongly absorbed by all proteins (particularly through the peptide bond) as well as other biomolecules (Buonanno et al., 2013; Goldfarb & Saidel, 1951; Setlow, 1966) thus penetration into living tissue is very shallow, as shown in Fig. 6. These experimental findings have led to recommendations to revise Far UV-C guidelines to limit human exposure that can be applied to whole-room exposure in occupied settings (see below) (ACGIH, 2021; Sliney, 2013; Sliney & Stuck, 2021).

Skin

The greatest safety concern associated with germicidal UV-C applications has long been the potential for promotion of skin cancer (CIE, 2010; ISO, 2016; Sliney, 2013). This concern often originates without recognizing the action spectrum of photocarcinogenesis, which peaks with UV-B wavelengths (Fig. 6) (Forbes et al., 2020).

Epidemiological studies of solar-induced skin cancer clearly show the causal factor to be primarily attributable to UV-B radiation (Moan et al., 2015). Because so few people have been exposed to UV-C there have been no human epidemiological studies solely on UV-C exposure. Even so, it is clear that the outermost layer of skin (i.e., the human stratum corneum) strongly attenuates UV-C radiation such that very little UV-C radiation reaches the germinative, basal layer of the epidermis, where the DNA would have to be altered to potentially initiate skin cancer (CIE, 2010; Barnard et al., 2020; Cadet, 2020; Forbes et al., 2020). Since Far UV-C radiation is so
strongly absorbed in the stratum corneum, very little of this radiation can even reach the superficial layers of the epidermis, and essentially none can penetrate to the deeper basal layer, negating Far UV-C as a cancer risk (see Fig. 6).

All but one of the ten recently published experimental, peer-reviewed safety studies – both in vitro and in vivo – of the effects of Far UV-C radiation on human skin and hairless mouse skin showed no measurable negative impacts (Barnard et al., 2020; Buonanno et al., 2017, 2016, 2021; Cadet, 2020; Fukui et al., 2020; Hanamura et al., 2020; Hickerson et al., 2021; Yamano et al., 2020) despite including much larger doses than would be permitted in a human exposure scenario. The one published study (Woods et al., 2015) that did show evidence of skin
erythema used an unfiltered Far UV-C excimer lamp, i.e., with a biologically significant component of UV radiation at wavelengths >230 nm. This 2015 study was replicated by Fukui et al. (2020) using a spectrally-filtered 222 nm lamp (i.e., without the longer wavelengths) at similar or higher doses, showing no detectable erythema.

The *in-vivo* human skin erythema results support the laboratory findings by Buonanno et al. using an *ex-vivo* human skin model (EpiDerm) to assess photocarcinogenic effects. They found no significant increased occurrences of potentially-mutagenic cyclobutane-pyrimidine-dimer (CPD) DNA lesions after exposure to 125 mJ/cm² of unfiltered Far-UVC radiation. Only a slight increase of CPDs was detected at 500 mJ/cm². Further evidence of a lack of photocarcinogenicity was shown by studies using hairless mice by Yamano et al. (2020) using two genotypes that were highly susceptible to photocarcinogenesis; they showed only a faint detection of CPDs only in the uppermost cells of the epidermis following exposure to a Far UV-C dose of 500 mJ/cm². These findings further support *in-vivo* human skin studies (Hickerson et al., 2021).

It is important to note that the superficial, outer-most cells of the epidermis can no longer divide and are about to become the dead cells to form the outermost skin layer – the stratum corneum. All of these studies show an absence of, or non-consequential skin damage at very high exposures, above those typically required for disinfection thus supporting the use of germicidal Far UV-C exposure in occupied spaces.

**Eye**

Only a few studies of the effects of exposures to Far UV-C wavelengths on eye tissue exist (Oksala & Lehtinen, 1960; Pitts, 1974; Kaidzu et al., 2019; Kaidzu et al., 2021a, 2021b). Kaidzu et al. recently determined threshold photokeratitis in a rodent eye model at 24 hr post exposure employing slit-lamp bio-microscopy, surface mapping, and staining. They determined photokeratitis thresholds at four UV-C wavelengths (207, 222, 235, and 254 nm) and at one UV-B wavelength (311 nm). Thresholds at 207 and 222 nm were above 5000 and 15,000 mJ/cm² respectively, which are well above current safety guidelines. Kaidzu et al. (2019) and Kaidzu et al. (2021a, 2021b) also conducted a histological study of the cornea by measuring CPDs to detect DNA damage, which were analyzed for each wavelength and as a function of exposure dose. The depth of CPD biomarkers seen at 207 and 222 nm were limited to the upper superficial cells of the corneal epithelium, which are sloughed off in a day during a normal corneal epithelial life cycle. The germinative cells of the cornea are located in the limbus bordering the cornea and conjunctiva and are protected – as in the skin – from Far UV-C radiation by several cell layers (see Fig. 6). By contrast, at a longer UV-B wavelength, specifically 311 nm, CPDs were observed in all layers of the cornea including the corneal endothelium.

The corneal injury thresholds reported by Kaidzu et al. (2019) and Kaidzu et al. (2021a, 2021b) for Far UV-C radiation are significantly higher than those reported by Pitts (1974). The lower thresholds reported by Pitts can be explained by their use of wide-bandwidth, arc-monomonochromator exposures at 220, 240, and 250 nm, since the wide bandwidths (10 nm full-width, half-maximum, FWHM) led to an apparent action spectrum that was flattened – similar to an error in the 310-nm region (Chaney & Sliney, 2005). Since the Pitts data were applied during the development of the current UV exposure limits (Sliney et al., 1971; Sliney & Stuck, 2021) the lower UV-C limits now appear to be quite overly conservative, as explained further below.

**Exposure limit guidelines**

The current human exposure guidelines do not differ around the world. The most widely recognized limits are those of the American Conference of Governmental Industrial Hygienists (ACGIH) and the International Commission on Non-Ionizing Radiation Protection (ICNIRP).
These had not changed for decades (ACGIH, 2021; ICNIRP, 2004), until recent guideline values were accepted (see discussion below). Both employed the same spectral weighting function, $S(\lambda)$, based upon threshold data for corneal and skin injury thresholds available in the 1970s. The UV-B data from monochromatic line spectra were given the highest importance (Sliney & Stuck, 2021). Figure SI-6 graphically summarizes these guideline values of $S(\lambda)$. Figure 7 illustrates the corresponding 8-h exposure limits.

The ACGIH issued a ‘Notice of Intended Change’ of UV-C TLVs in 2021. They proposed for the first time different limits for the eye and skin at wavelengths less than 300 nm as well as an increase in ocular limits at wavelengths below 250 nm (ACGIH, 2021). These revised guideline values were recently accepted by ACGIH (2022). The updated $S(\lambda)$ values for the skin and eye exposure are shown in Fig. SI-6 along with previous values. The corresponding exposure limits from the updated and previous values for $S(\lambda)$ are given in Fig. 7. The daily (based on an 8-hr exposure period), time-weighted exposure limits at 222 nm were increased from 23 mJ/cm² (for both skin and eyes) to 161 mJ/cm² for the eye and 479 mJ/cm² for the skin.

**Risks for sensitive individuals for exposure to far UV-C radiation**

The TLVs are guidelines to apply to nearly all individuals and are intended to be sufficiently conservative for the wide range of normal skin and eye sensitivities, including for individuals who are considered to have sensitive or photosensitive skin, or who have ‘dry eye’. Photosensitivity resulting from medications or other photosensitizers requires penetration well into the epidermis where photosensitizers would exist. Far UV-C exposure of the skin is considered much safer than longer UV-C wavelengths because of such shallow penetration depths; almost all incident photons are absorbed in the stratum corneum. The thickness of this layer does not vary significantly among individuals who may or may not possess ‘sensitive skin’ (Richters et al., 2017). This observation would argue against any individual differences because of skin diseases or by age, sex, or skin pigmentation (Lock-Andersen et al., 1997; Tagami, 2015). Correspondingly, the tear-layer thickness of the muco-aqueous tear layer or the lipid tear layer in healthy vs. even severe dry-eye patients show little difference (Segev et al., 2020). Therefore, the weight of evidence suggests that there are no significant subgroups of individuals who, by virtue of age, sex, race, health

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**Figure 7.** Comparison of 8-hr Threshold Limit Values (TLVs) for UV-C radiation, based on relative spectral weighting values shown in Fig. SI-6. Data points indicate values as listed in the Guidance (ACGIH, 2022).
conditions, or genetics, are systematically more sensitive than average to the effects of Far UV-C radiation.

**Ozone generation by far UV-C lamps**

Another safety-related concern related to Far UV-C-based applications is the potential to generate ozone. Ozone can be generated and decomposed by photochemical reactions; however, it should be noted that ozone may also be formed from corona discharges, which are associated with some Far UV-C sources. Moreover, the accumulation of ozone in ambient air will be governed not only by reactions that lead to formation and decay of ozone, but also air circulation and mixing in the vicinity of the Far UV-C source and room air exchange.

**Reactions responsible for ozone formation and decay from far UV-C sources**

Photochemical formation and destruction of ozone in the gas phase have been investigated extensively, largely because of the roles played by ozone in tropospheric and stratospheric chemistry, as well as with the use of vacuum UV (VUV) sources as ozone generators. Ozone \( O_3 \) is generated by short wavelength UV radiation \( (\lambda \approx 242 \text{ nm}) \) or in electrical discharges (corona) when molecular oxygen \( O_2 \) is present. Ozone can also be generated photochemically by a combination of ozone precursors (e.g., nitrogen oxides and hydrocarbons) at longer wavelengths UV \( (290 \text{ nm} \leq \lambda \leq 400 \text{ nm}) \), but this mechanism of ozone formation is unlikely to contribute significantly to ozone generation by Far UV-C sources.

Much of our current understanding of the potential for ozone formation from Far UV-C sources is based on studies of ozone formation and destruction in the stratosphere, which are initiated by photochemical reactions. The basic reactions that are responsible for photochemically-initiated formation and decomposition of ozone were first proposed by Sydney Chapman in 1930. The ‘Chapman Cycle’ comprises the following set of reactions (Chapman, 1930; Wayne, 1985). Note that ‘M’ refers to a non-reacting (chaperone) molecule:

\[
O_2 + h\nu \rightarrow O + O \quad (1) \\
O + O_2 + M \rightarrow O_3 + M \quad (2) \\
O_3 + h\nu \rightarrow O + O_2 \quad (3) \\
O + O_3 \rightarrow O_2 + O_2 \quad (4) \\
O + O + M \rightarrow O_2 + M \quad (5)
\]

Reactions 1 and 3 in this sequence are both photochemical. The rates of these reactions are governed by the rate at which UV photons in the relevant wavelength range are absorbed by the reactant molecules. In the case of reaction 1, the reactant is molecular oxygen \( O_2 \), while for reaction 3 the reactant is ozone \( O_3 \).

Net generation of ozone will depend on balance between the rates of ozone formation and decay; both processes are initiated photochemically. Ozone is a much stronger absorber of UV-C radiation than molecular oxygen on a per-molecule basis; however, the concentration of \( O_2 \) (in air) is many orders of magnitude larger than the concentration of \( O_3 \). Consequently, the rate of reaction 1 is generally faster than the rate of reaction 3 with Far UV-C or VUV sources. For example, for a system operating with an air mixture at a total pressure of 1.0 atm with a temperature of 20°C and \( O_3 \) present near the regulatory limit for \( O_3 \) (0.1 ppmv), with the ambient air being exposed to radiation at 222 nm (the peak wavelength from a KrCl* lamp), reaction 1 will proceed roughly 4.3 times faster than reaction 3. In a room where (essentially) no ozone is present (i.e., when a Far UV-C lamp is turned on), the ratio of the rates of ozone formation and
decay will be much larger. Therefore, the potential exists for generation of O$_3$ from Far UV-C sources.

Net production of O$_3$ by a Far UV-C source will depend on the fundamental chemistry described above, but also the conditions of application. Relevant conditions will include the output power and output spectrum of the Far UV-C source, as well as air circulation and mixing in the vicinity of the lamp.

Additionally, it should be noted that excimer lamps, depending on power and design, are driven by voltages in the $2\text{–}10\text{kV}$ range, which are applied to the outside of the lamp. At these high voltages, there can be corona discharges on the exterior surface of the lamp. Since the discharge will happen in air, ozone generation by corona discharge can be an important contributor to ozone formation from a Far UV-C lamp and can be significantly higher than ozone generation by photochemical processes.

**Ozone exposure limits**

Ozone has been shown to affect the respiratory system as well as the cardiovascular and central nervous systems. Exposure limits for gas-phase ozone have been established by many governmental and non-governmental organizations that have responsibilities to ensure public health. Table SI-2 provides a summary of exposure limits for O$_3$ that have been established by several of these organizations. In general, these limits call for gas-phase O$_3$ concentration to be maintained less than 0.05–0.1 ppmv.

**Ozone generation measurements**

Two leading manufacturers of KrCl$^*$ lamps provided typical, measured ozone generation rates for their lamps. The lamps were operated at $<12\text{ W}$ electrical power. The manufacturers reported significantly different generation rates of 70 µg/hr (7.7 µg O$_3$/W·h) and 12 µg/hr (1.1 µg O$_3$/ W·hr). One manufacturer also reported data for a 70 W lamp of 12 mg/hr (170 µg O$_3$/W·hr).

It must be noted that the coaxial design of that lamp is quite different from the previously-mentioned low power lamps. The substantially higher ozone generation rate of this lamp is attributable to the contributions of corona discharges on the outside of the lamp, which led to much higher ozone generation than by just photochemical processes. It should also be noted that experiments show that ozone generation can be reduced by three times or more when optical filters are applied that absorb photons at wavelengths less than 200 nm.

It is also worth noting that the methods used to measure ozone generation by these lamps were different between the two manufacturers. The development of an industry standard for this measurement is needed to clarify the potential for ozone production in applications of Far UV-C sources.

**Ozone generation in applications**

Control of ozone exposure to humans and the environment will require that ozone generation rates from these lamps be quantified using a standardized method. Since the lamps will always be used in fixtures, it is the responsibility of the fixture manufacturer to provide data to customers about ozone emission from the fixture.

For rough estimates of the room ozone concentration the following formula can be used:

$$\text{Ozone concentration} = \frac{\text{Ozone generation rate} \times \text{time}}{\text{Room volume}}$$

(7)
Using one of the above-mentioned lamps (<12 W) as an example, the maximum ozone accumulation within a room of 1000 cubic feet (28.3 m³ – standard room size for UL867) over 24 hr will be 0.027 ppmv; such an installation would very likely fall below the UL867 upper limit (Underwriters Laboratories, 2000) of 0.050 ppmv, particularly when ‘real world’ conditions of air mixing and in/outflow are also considered. In practical applications of these devices, air circulation from a conventional HVAC system can be used to maintain the ambient ozone concentration well below the limits that are presented in Table SI-2. By extension, this indicates that practical applications of Far UV-C sources should be developed by considering air circulation and mixing that is imposed on indoor air space by the existing HVAC system.

Given the number of variables in lamp design, emission spectrum, lamp power, lamp duty cycle, room volume, air circulation, and air exchange rates, it is not possible to say whether a given Far UV-C installation presents an ozone risk without assessing its specifics. Ozone generation by current Far UV-C lamps must be discussed by fixture manufacturers and considered by those installing or managing these fixtures.

Conclusions

Conventional germicidal UV-C radiation has been demonstrated to be effective for inactivation of airborne pathogens. Far UV-C radiation appears to be at least as effective as conventional UV-C radiation for microbial and viral inactivation, and available evidence gives a strong indication that this portion of the spectrum can be effective at controlling airborne disease transmission.

Implementation of UV-C systems for disinfection of air must be conducted in a manner that balances the need to inactivate airborne pathogens with the risks of UV-C exposure to humans. In the case of conventional UV-C systems, based on low-pressure mercury lamps as the source of radiation, this balance requires that humans be shielded from exposure to the output from these lamps. This is commonly achieved by any of three approaches: removal of human presence from the UV radiation sources (including presence-detection interlocks), isolation of the UV sources from occupied spaces (including in-duct designs and louvered housings), and controlling exposure to within accepted safe levels (based on radiometric measurements and occupation residence times). The same approaches to control equally apply to air disinfection systems based on Far UV-C sources.

In contrast with conventional germicidal UV-C, the potential for damage to human eye and skin tissues is greatly diminished with Far UV-C radiation, largely because of the limited extent of penetration of Far UV-C radiation into these tissues. As a result, the design and implementation of currently available Far UV-C sources (e.g., KrCl* lamps) may allow greater exposure of humans to this radiation than is acceptable for sources of conventional germicidal UV-C radiation. Along with the demonstrated efficacy in microbial inactivation, this gives a reasonable operating window where safe and effective solutions can be developed.

Current Far UV-C sources present two potential pathways for ozone generation through photolysis and electrostatic ionization of molecular oxygen. However, their potential for ozone generation is modest, indicating that control of ambient ozone concentration in rooms where Far UV-C is implemented can be accomplished with conventional HVAC systems and well-established approaches to ventilation.

Taken together, these attributes suggest that air disinfection systems based on Far UV-C radiation have the potential to function as effective measures for reducing the likelihood of airborne disease transmission, including highly transmissible diseases such as influenza, the measles, the common cold, and COVID-19. As such, these devices appear to have characteristics that will allow them to play important roles in controlling the current pandemic, as well as future pandemics and epidemics involving airborne pathogens.
Note

1. The wavelength ranges listed above conform to the definitions provided by the International Commission on Illumination (CIE) (see http://cie.co.at/eilvterm/17-21-008). The International Union of Pure and Applied Chemistry (IUPAC) presents a slightly different set of wavelength ranges for these categories of UV radiation (see https://goldbook.iupac.org/terms/view/UT07492). Specifically, the IUPAC defines UV-A and UV-B in the same manner as CIE, but defines UV-C as radiation with wavelengths of 200–280 nm, while VUV is defined as radiation with wavelengths of 100–200 nm.

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Disclosure of interest statement

No potential conflict of interest was reported by the authors.

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