The basidiomycetes fungus *Schizophyllum commune* has been used as a model of mushroom development because it takes short time for fruiting body formation on synthetic media, its whole genome sequence is publicly available, and many molecular tools are applicable [1]. In addition, this fungus is easily found on trees where it decomposes recalcitrant lignin which has similar structure appear in some of hard to degrade synthetic dyes [2,3]. Therefore, studying the dye degradation ability on *S. commune* species is a promising area, to be manipulated for using in industrial process such as biodegradation of complex substances or pollutants, processing agricultural products or byproducts of biofuel production in the future.

Recently, we screened 81 *S. commune* strains deposited in the Culture Collection of Mushrooms in Incheon National University for the decolorizing activities for several textile dyes and found one excellent candidate with decolorization for all dyes named as IUM1114 [4]. From this strain, total 64 single basidiospore were isolated and among them, 20 single spores were subjected for testing of mating ability. Within 400 combinations, there were 21 combinations showing the success of mating. The 20 strains were also grown in aerated media using filtered vent cap falcon tubes supplied with different textile dyes, such as: Congo Red, Crystal Violet, Amaranth Red, Brilliant Blue G and Congo Red (Figure 1(A)). The red color of Amaranth Red dye almost disappearing after 10 days of incubation with fungus suggested the decolorization ability of this strain on Amaranth Red. While the blue color of Brilliant Blue G dye just remained in fungus clumps but not in the solution implied the dye absorption ability of this strain (Figure 1(B)). The whole-genome *de novo* sequencing of this *S. commune* IUM1114-SS01 was performed to further understand the dye-decolorizing enzyme systems of this fungus.

Genomic DNA of *S. commune* IUM1114-SS01 was extracted from homogenized mycelia of the 3-day-old subculture on potato dextrose agar at 25 °C using Nucleospin® Plant II kit from Macherey-Nagel company according to manufacturer’s instruction. Genome sequencing was performed using PacBio single-molecule real-time (SMRT) sequencing technology. High-molecular-weight genomic DNA was sheared and selected on a BluePippin system using a cutoff range of >15 kb. The libraries were sequenced on one SMRT cell using a PacBio Sequel platform according to the manufacturer’s instructions (Pacific Biosciences, Menlo Park, CA, USA). For *de novo* assembly, a total of 7.2 billion bases from 753,103 reads were assembled using the SMRT Analysis Hierarchical Genome Assembly Process 3 (HGAP3) pipeline, which resulted in 84 contigs, with the length of the N50 value being...
In addition, a total of 4.9 billion bases from 32.5 million reads were sequenced for error correction through the Illumina HiSeq 4000 platform. The HiSeq reads were applied for accurate genome sequence by using Pilon (v1.21), resulting in 74 contigs with the length of the longest one and the N50 value being 44,38,218 bp and 19,27,765 bp, respectively (Table 1). The total size of the S. commune IUM1114-SS01 genome was 4,10,06,423 bp and the cumulative GC content of the genome was 57.44% (Table 1).

Ab initio gene prediction was carried out using Maker v2.31.10 [5] based on gene model training from S. commune reference strain (H4-8). As a result, 13,386 protein encoding genes were predicted including 214 nuclear tRNAs (Table 1). Genes were functionally characterized using BLAST+ (v2.7.1+) [6] and InterProScan [7]. The carbohydrate-active enzyme (CAZyme) gene content was determined using dbCAN2 v2.0.6 [8, 9] based on three different tools, HMMER v3.2.1, DIAMOND v0.9.30, and Hotpep [10]. The analysis revealed that the S. commune IUM1114-SS01 genome contains 249 glycoside hydrolases (GH), 91 glycosyltransferases (GT), 18 polysaccharide lyases (PL), 66 carbohydrate esterases (CE), 18 carbohydrate-binding modules (CBM), and 89 enzymes with auxiliary activities (AA) which are similar with S. commune H4-8 strain (Table 2) [1].

Table 1. Summary of the genome assembly and annotation of S. commune IUM1114-SS01.

| Metric       | Value          |
|--------------|----------------|
| Total length | 4,10,06,423 bp |
| Contigs      | 74             |
| Largest contig | 44,38,218 bp |
| N50          | 19,27,765 bp   |
| L50          | 8              |
| N75          | 6,81,301 bp    |
| L75          | 18             |
| Proteins     | 13,386         |

Table 2. Comparison of the number of CAZymes of S. commune IUM1114-SS01 with those of other fungi in the order Agaricales.

| Species (Ensembl ID) | AA | CBM | CE | GH  | GT | PL | Total |
|----------------------|----|-----|----|-----|----|----|-------|
| Agaricus bisporus var. burnetttii JB137-S8 (GCA_000300555) | 97 | 16  | 56 | 186 | 71 | 11 | 437   |
| Coprinopsis cinerea okayama#130 (GCA_000182895) | 129 | 28  | 72 | 198 | 90 | 16 | 533   |
| Laccaria bicolor S238N-H82 (GCA_000143565) | 58 | 6   | 35 | 161 | 84 | 8  | 352   |
| Pleurotus ostreatus PC15 (GCA_000697685) | 139 | 34  | 59 | 237 | 80 | 24 | 573   |
| Schizopyllum commune H4-8 (GCA_000143185) | 87 | 17  | 65 | 248 | 92 | 18 | 527   |
| S. commune IUM1114-SS01 | 89 | 18  | 66 | 249 | 91 | 18 | 531   |

*CAZymes: Carbonhydrate-Active Enzymes; AA: Auxiliary Activities family; CBM: Carbohydrate-Binding Module family; CE: Carbohydrate Esterase family; GH: Glycoside Hydrolase family; GT: Glycosyl Transferase family; PL: Polysaccharide Lyase family.
These indicated that this *S. commune* IUM1114-SS01 strain has high potential to be used for biomass conversion. The genomic information of *S. commune* IUM1114-SS01 will provide a better understanding of the genomic network involved in dye degrading metabolic process in the white rot fungi. The draft genome sequence of *S. commune* IUM1114-SS01 has been deposited in GenBank under the accession number JAATO10000000000, BioProject number PRJNA615298, and BioSample number SAMN14450467.

**Disclosure statement**

No potential conflict of interest was reported by the author(s).

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