Increasing effects of selective 5-HT$_{2C}$ receptor stimulation on evoked momentary urethral closure in female rats and humans

Authors: Izumi Kamo, Hiroshi Nagata, Gale O'Connell, Takuya Kato, Akio Imanishi, Masako Kuno, Satoshi Okanishi, Kyoko Yoshikawa, Yuya Nishiyama

Research (IK, AI, MK, KY), Takeda Development Center Japan (YN), Formerly Japan Development Center (HN, TK), and Formerly Pharmaceutical Research Division (SO), Takeda Pharmaceutical Company Ltd., Kanagawa and Osaka, Japan

Formerly Takeda Development Center Europe Ltd. (GO), London, UK
Running title: Increased momentary urethral closure by 5-HT\textsubscript{2C} agonist

Corresponding author: Izumi Kamo, Ph.D., Research, Takeda Pharmaceutical Company Ltd.,
26-1, Muraoka-Higashi 2-chome, Fujisawa, Kanagawa 251-8555, Japan, Tel +81-80-2518-8419,
Fax +81-466-29-4412. E-mail: izumi.kamo@takeda.com

The number of text page: 46

The number of tables: 3

The number of figures: 7

The number of references: 25

The number of words in Abstract: 250

The number of words in Introduction: 580

The number of words in Discussions: 1457

Abbreviations

5-HT: 5-hydroxytryptamine; 5-HT\textsubscript{2A}: 5-Hydroxytryptamine type 2A; 5-HT\textsubscript{2B}: 5-Hydroxytryptamine type 2B; 5-HT\textsubscript{2C}: 5-Hydroxytryptamine type 2C; i.d.: intraduodenal; i.v.: intravenous; LPP: leak point pressure; MT: motor threshold; P\textsubscript{ves}: intravesical pressure; S-LPP:
sneeze leak point pressure; TMS: transcranial magnetic stimulation

A recommended section: Gastrointestinal, Hepatic, Pulmonary, and Renal
Abstract

Under healthy condition, more than one urethra-closing reflex, including both bladder afferent-independent and -dependent actions function during momentary elevation of intravesical (bladder) pressure to prevent urinary incontinence. In the current study, the effects of a novel selective 5-hydroxytryptamine type 2C (5-HT\textsubscript{2c}) receptor agonist, TAK-233, on evoked momentary urethra-closing functions were investigated in female rats and humans to elucidate 5-HT\textsubscript{2c} receptor functions. In anesthetized female rats, TAK-233 dose-dependently and significantly increased the urethral resistance during sneezing in rats with distended vaginas and bilaterally transected pelvic nerves. The drug also dose-dependently and significantly increased urethral resistance during momentary intravesical pressure elevation by electrical stimulation of abdominal muscles in rats with a transected spinal cord at the T8-9 level and intact pelvic nerves. The increased effects observed during electrical stimulation were abolished by either an intravenously administered selective 5-HT\textsubscript{2c} receptor antagonist, SB 242084, or bilateral transection of the pelvic nerves or somatic nerves innervating the external urethral sphincter and pelvic floor muscles. In the spinal cord-transected and pelvic nerve-intact rats, TAK-233 enlarged the urethra-closing responses induced by both passive and abrupt intravesical pressure elevation measured by a micro-tip transducer located in the middle urethra. Additionally, the effects of TAK-233 on the stimulus threshold of urethral contractile responses induced by transcranial
magnetic stimulation were investigated in healthy female volunteers. The drug dose-dependently and significantly lowered this stimulus threshold, indicating an increased sensitivity of the response. These results demonstrate that 5-HT$_{2C}$ receptor stimulation enhances the evoked momentary urethra-closing functions in both female rats and humans.
Significance Statement

5-hydroxytryptamine (serotonin) type 2C (5-HT$_{2C}$) receptor stimulation by TAK-233 enhanced urethral resistance in rats during an evoked momentary event in which the bladder afferent-independent or -dependent reflex functions via striated muscle-mediated mechanisms. The increases in sensitivity of transcranial magnetic stimulation-evoked urethral contractile responses in healthy female subjects indicates that this mechanism also functions in humans. The evoked momentary conditions activating these reflexes provide a suitable model to demonstrate the exerting effects of 5-HT$_{2C}$ receptor stimulation.
Introduction

Stress urinary incontinence generally occurs as a result of defects in various passive and reflex mechanisms that maintain urethral closure in the presence of elevated abdominal pressure (Yoshimura and Miyazato, 2012). With respect to nerve-mediated mechanisms, at least 2 urethra-closing reflex mechanisms function during the abrupt and momentary elevation of intravesical (bladder) pressure ($P_{ves}$; Kamo et al., 2009, Yoshimura and Miyazato, 2012). One is a bladder afferent-dependent reflex where both abrupt and passive $P_{ves}$ elevation induces urethra-closing responses in spinal cord-transected rats (spinal reflex), and where the responses are totally abolished by the bilateral transection of the pelvic nerves containing bladder afferent nerves (Kamo et al., 2004). A 1-second electrical stimulation of abdominal muscles induces momentary passive $P_{ves}$ elevation in rats, and bilateral transection of the pelvic nerves greatly reduces urethral resistance during electrical stimulation (Kamo and Hashimoto, 2007), demonstrating that the bladder afferent-dependent spinal reflex contributes to the prevention of urinary incontinence during passive $P_{ves}$ elevation. The other reflex is a bladder afferent-independent mechanism and functions during sneezing in spinal cord-intact rats (Kamo et al., 2003, 2006 and 2009). Clear middle urethra-closing responses are observed during sneezing, and those responses are greatly reduced after bilateral transection of the somatic nerves innervating the external urethral sphincter and pelvic floor muscles, whereas bilateral transection
of both pelvic and hypogastric nerves had no effect (Kamo et al., 2003), demonstrating that the reflex is bladder afferent-independent. Bilateral transection of the somatic nerves induces a clear reduction in urethral resistance during sneezing in rats (Kamo et al., 2003), indicating that this reflex contributes to the prevention of urinary incontinence during sneezing. In addition, the urethral reflex is considered to be one component of the pre-programmed sneeze reflexes observed throughout the body, since urethral responses are observed before the start of $P_{\text{ves}}$ elevation during sneezing (Kamo et al., 2003). Since a sneeze is a very brief and sudden event lasting for up to 0.15 s in rats (Kamo et al., 2003), the passive $P_{\text{ves}}$ elevation-induced urethral reflex (bladder afferent-dependent spinal reflex) may occur too late to prevent any urine leakage from the urethral orifice (Kamo et al., 2009). These findings indicate that at least 2 urethra-closing reflexes are essential to prevent urinary leakage during various events that elevate $P_{\text{ves}}$, therefore it is considered important to identify relevant mechanisms that enhance both types of reflexes when developing treatments for stress urinary incontinence.

5-hydroxytryptamine (serotonin; 5-HT) receptors are divided into at least 14 different receptor subtypes that have been classified into seven major families (Hoyer et al., 2002). Stimulation of 5-HT$_{2C}$ receptor increased the urinary storage function of the urethra in a pioneering study in guinea-pigs (Conlon et al., 2012). In rats, receptor stimulation increased the bladder afferent-independent urethra-closing reflex during sneezing (Miyazato et al., 2009; Suzuki et al.,
2018; Ochi et al., 2018). A 5-HT$_{2B/2C}$ agonist, mCPP, and a 5-HT$_{2C}$ agonist, CP-809101, enhanced the urethra-closing responses during sneezing in rats, and these responses were totally abolished by RS-102221 and SB 242084, selective 5-HT$_{2C}$ receptor antagonists, respectively. However, the effect on urethral functions via the bladder afferent-dependent spinal reflex remains unclear.

In the current study, in order to clarify the effects of 5-HT$_{2C}$ receptor stimulation on urethral function, TAK-233, another selective 5-HT$_{2C}$ receptor agonist, was investigated for effects on the urethra-closing reflex under both the bladder afferent-dependent and -independent mechanisms. Furthermore, in order to better understand these effects in humans, the effects of TAK-233 on transcranial magnetic stimulation (TMS)-induced urethra-closing responses were investigated in healthy female subjects.
Materials and Methods

Non-clinical studies

Drugs

TAK-233 (N-Methyl-N-(1-methylethyl)-6,7,8,9-tetrahydropyrazino[2,3-f][1,4]oxazepin-3-amine monohydrochloride; Figure-1, and Example 8 in WO2010/147226A1), and lorcaserin hydrochloride (lorcaserin) were synthesized by Takeda Pharmaceutical Company Ltd. 5-HT, duloxetine hydrochloride (duloxetine), and SB 242084 were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan), Kemprotec (Middlesbrough, UK), and Tocris Bioscience (Bristol, UK), respectively. Drugs were suspended in 0.5% methylcellulose (Shin-Etsu Chemical CO., Ltd., Tokyo, Japan) solution for intraduodenal (i.d.) administration, and dissolved in N,N-dimethylacetamide-polyethyleneglycol 400 (1:1) for intravenous (i.v.) administration. Drug doses are listed as the weight of the free base.

Animals

Adult female rats of Sprague-Dawley strain weighing 203.2 g – 346.0 g (CLEA Japan, Tokyo, Japan) were used. All animal experiments were conducted in accordance with the Guide for the Care and Use of Laboratory Animals as adopted and promulgated by the U.S. National Institutes of Health, and the protocols of the experiments were approved by Takeda’s Experimental Animal
Care and Use Committee.

**In vitro receptor agonist potency**

Recombinant human 5-HT$_{2C}$ receptor-expressing CHO-K1 cells purchased from Euroscreen (Gosselies, Belgium) were cultured in supplemented UltraCHO media, and recombinant human 5-HT$_{2A}$, human 5-HT$_{2B}$ or rat 5-HT$_{2C}$ receptor-expressing CHO dhfr- cells generated by Takeda Pharmaceutical Company Ltd. were cultured in supplemented MEM alpha media.

5-HT receptor agonist potency of 5-HT, TAK-233, and lorcaserin was assessed by performing calcium influx assays using the integrated detection and liquid handling robotics, CellLuxTM (PerkinElmer, Waltham, MA). The recombinant receptor-expressing cells were seeded into black-walled clear-bottom 384-well plates at a density of 10,000 cells per well and treated with 10 mM sodium butyrate and grown for approximately 24 h at 37°C in a 5% CO$_2$ incubator. The culture media were removed, and the cells were loaded with 40 µl/well of Ca$^{2+}$ indicator dye solution, Ca Screening Kit Fluo 4 (DOJINDO laboratories, Kumamoto, Japan) for around 60 min at 37°C in a 5% CO$_2$ incubator. The plates were left for approximately 10 min at room temperature and placed into the CellLuxTM to measure intracellular Ca$^{2+}$ mobilization. Fluorescence was monitored at a 2-second interval for 30 s. After 10 s basal signal recording, 20 µl of 3-fold compound solution was added by CellLuxTM equipped 96-well pipettor. CellLuxTM
repeated the sequence 4 times.

The collected fluorescence counts were calculated by subtracting the background fluorescence in the same well. The change in fluorescence counts by each compound were determined by subtracting the minimum fluorescence count from maximum fluorescence count in the same well. The change in fluorescence counts was normalized to the response observed with 10 μM of 5-HT.

The values of EC$_{50}$ were estimated by a 2-parameter logistic regression analysis. The relative potency was calculated by dividing EC$_{50}$ value of a compound by EC$_{50}$ value of 5-HT. The selectivity for human 5-HT$_{2C}$ receptors was calculated by dividing the relative potency against human 5-HT$_{2A}$ or human 5-HT$_{2B}$ receptors by the relative potency against human 5-HT$_{2C}$ receptor.

*Sneeze leak point pressure (S-LPP)*

The experiment was conducted based on the method described in literature (Kamo et al., 2003). Rats were anesthetized with isoflurane (Abbott Japan, Tokyo, Japan). A 8Fr Foley balloon catheter (TOP Corporation, Tokyo, Japan) was inserted into the vagina, inflated with 3 mL water for 2 hours, and removed. Four days after the vaginal distention, rats were anesthetized with urethane (Wako Pure Chemical Industries) and isoflurane. The bladder and the duodenum were exposed through an abdominal incision. PE-50 and PE-100 polyethylene catheters (Intramedic, Becton
Dickinson and Company, Franklin Lakes, NJ) were inserted into the duodenum and the bladder, respectively, and were secured with ligatures. A 3.5 Fr catheter with a side mounted SPR-524 microtip transducer (Millar Instruments, Houston, TX) connected to an amplifier (ML117, ADInstruments, Castle Hill, NSW, Austria) was inserted into the bladder. The pelvic nerves were cut bilaterally to prevent reflex bladder contractions. The abdominal incision was closed with sutures. S-LPP expressing urethral resistance during sneezing was measured as follows. After the bladder was emptied, 0.3 ml of 0.1% Evans blue (Wako Pure Chemical Industries)-saline solution was injected into the bladder. A fiber of brush was inserted into the nostril to induce sneeze reflexes while recording changes in $P_{\text{ves}}$ at a sampling rate of 1000 Hz while examining if leakage of the Evans blue solution was observed from the urethral orifice. $P_{\text{ves}}$ was recorded using data acquisition software (Chart, ADInstruments) on a computer system equipped with a PowerLab® analog-to-digital converter (ADInstruments). The maximal $P_{\text{ves}}$ during each sneeze event was evaluated and the lowest pressure that induced fluid leakage was defined as S-LPP.

**Leak point pressure (LPP) during electrical stimulation of abdominal muscles**

The experiments were conducted based on the methods described in literature (Kamo and Hashimoto, 2007). Rats were anesthetized by intraperitoneal administration of urethane, and isoflurane inhalation was added during surgery, if necessary. Their spinal cord was transected at
the T8-9 level after laminectomy to prevent reflex bladder contractions. After the abdomen was
opened, nerves innervating the lower urinary tracts and the pelvic floor muscles were transected
as indicated in results. PE-100 and PE-50 catheters were inserted into the bladder and the
duodenum, respectively, and secured. The abdomen was then closed with sutures. The bladder
catheter was connected to a pressure transducer (Life kit; Nihon Kohden, Tokyo, Japan) and an
amplifier (AP-641G; Nihon Kohden). Two sites of abdominal skin were cut, and the stimulus
needle electrodes were inserted in exposed abdominal muscles. After the bladder was emptied, 0.3
mL of 0.1% Evans blue-saline solution was injected into it. While the $P_{ves}$ was digitally recorded
at a sampling rate of 500 Hz using data-acquisition software (Acknowledge; BIOPACK Systems,
Santa Barbara, CA) on a computer system equipped with an analog-to-digital converter
(MP-100A-CE; BIOPACK Systems), the exposed abdominal muscles were stimulated with
rectangular electric pulses (50 Hz, 0.5 ms width pulse trains lasting 1s) by an electrical stimulator
(SEN-3301; Nihon Kohden) and an isolator (SS-202J; Nihon Kohden). The stimulus intensity was
gradually increased from 1 V to 10 V to increase the $P_{ves}$ step by step, and the lowest $P_{ves}$ at which
fluid leakage from the urethral orifice was observed was regarded as LPP indicating urethral
resistance.

Reflex urethra-closing responses
The experiments were conducted based on the method described in literature (Kamo and Hashimoto, 2007). The spinal cord of urethane-anesthetized rats was transected at the T8-9 level after laminectomy. The bladder was exposed, and a PE-100 catheter was inserted into it. The bladder neck was ligated with a suture to prevent fluid leakage from the bladder into the urethra. P\textsubscript{ves} was controlled by connecting a bladder catheter to a saline reservoir and a pressure transducer (MLT0670, AD Instruments) via three-way stopcocks. A SPR-524 microtip transducer catheter was inserted into the middle urethra from the urethral orifice. The microtip transducer catheter and the pressure transducer were connected to an amplifier (ML117, AD Instruments), and urethral responses were digitally recorded at a sampling rate of 1000 Hz using data-acquisition software (Chart, AD Instruments) on a computer system equipped with an analog-to-digital converter (PowerLab 4/52, AD Instruments). P\textsubscript{ves} was abruptly increased by elevating the reservoir and maintaining P\textsubscript{ves} for 30 s at 50 cmH\textsubscript{2}O while the urethral response was recorded. The average values measured with the microtip transducer for 30 s before and during increment of P\textsubscript{ves} were evaluated, and the difference of the average values was defined as a urethra-closing response.

Statistical Analysis

Data are the mean ± SEM. Data are analyzed with Dunnett’s test, Student’s t-test, or Welch’s test, and P values less than 0.05 were considered to be significant.
Clinical study

A randomized, double-blind, single-dose, 4 × 4 crossover Pharmacodynamic study

This study was approved by the Institutional Review Board at the study site and was conducted in accordance with the ethical principles of the Declaration of Helsinki, the International Conference on Harmonization E6 Guidelines for Good Clinical Practice and all applicable local laws and regulations.

Healthy adult Japanese females from 20 to 40 years of age, weighing at least 45 kg, with a body mass index between 18.5 kg/m² and 25.0 kg/m², had signed and dated a written informed consent, and met inclusion criteria were randomized into 4 administration sequences. Main inclusion criteria were as follows.

1. In the opinion of the investigator or subinvestigator, a subject who was capable of understanding and complying with the protocol requirements.

2. A female subject who did not have any conditions that could affect the urethra function (including no vaginal delivery nor urethral surgery, etc.).

A subject received a single dose of either TAK-233 (Takeda Pharmaceutical Company Ltd.) 20 mg, TAK-233 90 mg, duloxetine (Eli Lilly Japan, Kobe, Japan) 40 mg, or placebo (both TAK-233 placebo solution and duloxetine placebo capsule) in Periods 1 through 4 in accordance with the
predefined treatment in each administration sequence. The study consisted of the screening period (Day -28 to Day -2), treatment period (Day -1 to Day 2 in Periods 1 through 4), and follow-up examination (Day 8 in Period 4). TAK-233, duloxetine, and placebo were administered in the fasted state on Day 1 in each period. All subjects who underwent the pharmacodynamic tests on Day 1 received 500 mg of levofloxacin (CRAVIT; Daiichi-Sankyo, Tokyo, Japan) after the test to avoid urinary tract infection. The washout interval between the 2 consecutive periods was at least 7 days.

Based on the method described in the literature (Boy et al., 2006), the pharmacodynamic test identified a motor threshold (MT) for inducing urethral sphincter contraction in response to TMS as a primary endpoint and measured urethral pressure profile at rest as a secondary endpoint. Briefly, after bladder emptying, a 110 mm double cone coil (Magstim, Whitland, UK) that was designed to fit overhead and connected to a Magstim Rapid stimulator (Magstim) was positioned near the vertex for TMS, and a dual microtip pressure transducer catheter (Unisensor AG, Attikon, Switzerland) having its transducers lateral in the three o’clock position was positioned in the bladder and the urethra by inserting it from the urethral orifice. After identifying maximal tolerable TMS within maximal stimulator output and suitable catheter position in the urethra for observing urethral responses, TMSs with decreasing intensities by 5% of the maximal stimulator output were delivered, and the minimum intensity that produced a urethral contractile response
was determined as MT. Urethral pressure profiles at rest were recorded during pulling the catheter at 1 mm/s speed, and the maximal pressure was measured. The pharmacodynamic tests were performed once before pretreatment and 3 times posttreatment on Day 1.

Blood samples were collected, and plasma drug concentrations were measured by a method with Liquid Chromatography with tandem mass spectrometry (LC/M/MS).

Results are expressed as the mean ± SD, and statistical difference among groups were analyzed by analysis of variance (ANOVA) for the primary endpoint.

Safety was also assessed as an endpoint. Treatment-emergent adverse events were coded using the Medical Dictionary for Regulatory Activities (MedDRA) Version 17.1., and vital signs, weight, ECGs, and clinical laboratory test values were also measured.
Results

In vitro receptor agonist potency

5-HT, TAK-233 and lorcaserin showed agonist activities on human and rat 5-HT$_{2C}$ receptors, with relative agonist potencies of TAK-233 and lorcaserin to 5-HT being slightly weaker on rat 5-HT$_{2C}$ receptors compared with those on human 5-HT$_{2C}$ receptors (Table-1). Selectivity of TAK-233 and lorcaserin for human 5-HT$_{2C}$ receptors over human 5-HT$_{2A}$ receptors was 26-fold and 11-fold (TAK-233 and lorcaserin, respectively), and selectivity over 5-HT$_{2B}$ receptors was 282-fold and 70-fold (TAK-233 and lorcaserin, respectively), indicating that selectivity of TAK-233 is slightly higher compared with that of lorcaserin, a selective 5-HT$_{2C}$ receptor agonist marketed for the treatment of obesity.

In a broad screen for receptor binding affinity and effects on various ion channel and enzyme activities, other than the 5-HT$_{2C}$ receptor, TAK-233 showed only slight activities on 5-HT$_{1B}$ and non-selective sigma receptors (50% and 54% radioligand binding inhibition at 10 μM, respectively).

S-LPP

In rats without vaginal distension, 2 out of 4 rats did not show fluid leakage from the urethral orifice during sneezing, whereas in all rats with vaginal distension, fluid leakage was observed.
during sneezing with a S-LPP of 47.7 ± 2.1 cmH₂O.

TAK-233 at 1 and 3 mg/kg dose-dependently and significantly increased the S-LPP (Figure-2). Duloxetine significantly increased the S-LPP at all tested doses, however, the maximal effect of TAK-233 was significantly higher than that of duloxetine (Figure-2).

**LPP during electrical stimulation of abdominal muscles**

All rats with unilateral transection of the nerves to iliococcygeus/pubococcygeus muscles showed fluid leakage from the urethral orifice by electrical stimulation. TAK-233 dose-dependently and significantly increased LPPs with an effect at 3 mg/kg, i.d. of approximately 30 cmH₂O (Figure-3). Duloxetine also dose-dependently and significantly increased the LPP with an effect at 20 mg/kg, i.d. of approximately 10 cmH₂O (Figure-3). In a preliminary experiment, the duloxetine effects at 20 mg/kg, i.d. were maximal among the tested doses up to 40 mg/kg, i.d.

The urethral resistance-increasing effects of TAK-233 at 3 mg/kg, i.d. were dose-dependently inhibited by SB 242084, a specific 5-HT₂C receptor antagonist (Kennett et al, 1997; Figure-4).

In rats with intact nerves innervating the lower urinary tracts and pelvic floor muscles, TAK-233 significantly increased the LPP with the effects totally abolished by bilateral transection of the pelvic nerves (Table-2). While bilateral transection of the hypogastric nerves innervating urethral...
smooth muscles did not affect TAK-233’s effects, bilateral transection of both the pudendal nerves innervating the external urethral sphincter and the coccygeus muscle and the nerves to iliococcygeus/pubococcygeus muscles greatly inhibited these effects (Table-2), indicating a greater contribution of the striated muscle component to the effects of TAK-233.

**Reflex urethra-closing responses in rats**

Abrupt elevation of P$_{ves}$ from 0 cmH$_2$O to 50 cmH$_2$O induced reflex urethra-closing responses (Figure-5; Kamo and Hashimoto., 2007). The responses were measured 2 times before and 30 min after drug administration, respectively, and the ratio of post-value to pre-value was calculated with the means of the 2 trials. TAK-233 at 3 mg/kg, i.d. significantly increased the responses with a post/pre ratio of 2.82 ± 0.29 (Mean ± SEM of 8 rats; P<0.001 v.s. vehicle-treated group by Welch's test), whereas the vehicle had no effect with a post/pre ratio of 1.11 ± 0.13 (Mean ± SEM of 8 rats). In a separate experiment, duloxetine at 20 mg/kg, i.d. also significantly enlarged the responses with a post/pre ratio of 2.00 ± 0.28 (Mean ± SEM of 8 rats; P<0.05 v.s. vehicle-treated group by Welch's test), whereas the vehicle had no effect with a post/pre ratio of 1.07 ± 0.11 (Mean ± SEM of 8 rats).

*A randomized, double-blind, single-dose, 4 × 4 crossover Pharmacodynamic study*
Effects of TAK-233 on urethra-closing functions in healthy female subjects were examined. The mean subject age was 22.4 years (SD, 2.04), the mean weight was 50.91 kg (SD, 3.346), and the mean BMI was 19.57 kg/m\(^2\) (SD, 0.929). Demographic and other baseline characteristics were well balanced across the administration sequences. Twenty-four eligible subjects at 1 study site in Japan received the study drug. Of the eligible subjects, 23 subjects completed the study; 1 subject withdrew due to an adverse event that was considered unrelated to the study drug. Each of the 24 eligible subjects were allocated to a 4-part administration sequence (6 subjects/sequence).

The mean MT for urethral sphincter contraction in response to TMS was significantly decreased by TAK-233 at 20 mg and 90 mg and duloxetine at 40 mg with no change after placebo treatment (Figure-6). The maximum MT change from baseline was -8.1%, -17.2% and -11.8% (TAK-233 20 mg at 0.5 h, TAK-233 90 mg at 3 h, and duloxetine 40 mg at 6 h, respectively), indicating a greater change from baseline following TAK-233 at 90 mg compared with the other treatment groups (Figure-6).

In the TAK-233 90 mg group, the mean change from baseline in the maximum urethral pressure at rest along the entire functional length of the urethra increased at 0.5 h post dose followed by decreases at later time points (Figure-7). In the duloxetine 40 mg group, the mean change from baseline increased at 3 h and 6 h post dose (Figure-7). In the TAK-233 20 mg group and placebo groups, the mean change from baseline appeared unaffected.
TAK-233 was rapidly absorbed after a single oral administration, and the maximal plasma level was observed at 0.5 h and 2 h (20 mg and 90 mg, respectively; Table-3). The maximal plasma level of duloxetine was observed at 6 h following administration (Table-3).

No deaths or serious adverse events and no obvious changes in mean values of clinical laboratory tests were reported during this study.
Discussions

While many events in daily activities cause a rise in abdominal pressure with bladder compression, nerve-mediated active urethral closure contributes to the prevention of urinary incontinence under normal condition (Yoshimura and Miyazato, 2012). Rat studies investigating urethral resistance and reflex urethral responses demonstrate that at least 2 kinds of reflexes function during this rise in $P_{ves}$ (Kamo et al., 2009; Yoshimura and Miyazato, 2012). One is the bladder afferent-dependent spinal reflex induced by passive $P_{ves}$ elevation, and the other is the bladder afferent-independent reflex. The latter reflex functions as a pre-programed reflex at least during sneezing, and 5-HT$_{2C}$ receptor stimulation by mCPP or CP-809101 increases the sneeze-induced urethral response under different conditions (Miyazato et al., 2009; Suzuki et al., 2018). In the current study, another specific 5-HT$_{2C}$ receptor agonist, TAK-233, also increased urethral resistance during sneezing in female rats. These findings obtained across a range of experimental conditions with different drugs demonstrate that 5-HT$_{2C}$ receptor stimulation increases urethral closure during sneezing in rats via an enhancement of the bladder afferent-independent reflex.

The effects of TAK-233 on passive $P_{ves}$ elevation-induced urethra-closing responses were investigated to determine if 5-HT$_{2C}$ receptor stimulation enhances the bladder afferent-dependent
spinal reflex. TAK-233 significantly increased the evoked responses, indicating enhancement of the spinal reflex. TAK-233 also increased the urethral resistance in spinal cord-transected rats during electrical stimulation of abdominal muscles-induced passive $P_{\text{ves}}$ elevation. The increased effects were not observed after bilateral pelvic nerve transection that would have totally abolished the $P_{\text{ves}}$ elevation-induced reflex urethral contraction itself (Kamo et al., 2004). These findings demonstrate that 5-HT$_{2C}$ receptor stimulation also enhances the bladder afferent-dependent spinal reflex to elevate urethral resistance during passive $P_{\text{ves}}$ elevation in rats.

The effects of TAK-233 on the urethral resistance during sneezing in rats were significantly larger than those of duloxetine, a potent and balanced dual 5-HT-norepinephrine reuptake inhibitor (Thor et al., 1995, Mariappan et al., 2007), with effects on the urethral resistance during electrical stimulation clearly larger than those of duloxetine in rats. One of the possible explanations for the larger effects of TAK-233 may be due to duloxetine stimulating both positively effective receptor subtypes such as 5-HT$_{2C}$ receptor, 5-HT$_7$ receptor and $\alpha_1$-adrenoceptor (Miyazato et al., 2009, Suzuki et al, 2018, Kaiho et al, 2007), and negatively effective receptor subtypes such as 5-HT$_{1A}$ receptor and $\alpha_2$-adrenoceptor (Yoshimura and Miyazato, 2012, Miyazato, 2008), that, when taken together, increases urethral function to an overall extent in rats. Whereas TAK-233, on the other hand, may only stimulate the positively effective receptor subtype 5-HT$_{2C}$ receptor with no stimulation of negatively effective receptor
subtypes, thus causing a larger urethral resistance elevation in rats.

A wide range of muscles such as the smooth urethral sphincter muscle, the striated external urethral sphincter muscle and the striated pelvic floor muscles (iliococcygeus, pubococcygeus and coccygeus muscles) contribute to urethra-closing functions in rats, with the striated muscles contributing significantly to the elevation of urethral resistance during sneezing and electrical stimulation of abdominal muscles in rats (Yoshimura and Miyazato, 2012, Kamo et al, 2003, 2004, 2009, Kamo and Hashimoto, 2007). In the current rat study, with electrical stimulation of the abdominal muscles, contribution of the smooth and striated muscles to the urethral resistance-increasing effects of TAK-233 were investigated by bilateral transection of the hypogastric nerves innervating the smooth urethral sphincter muscle (de Groat et al, 1993), the pudendal nerves innervating the external urethral sphincter and the coccygeus muscle (Manzo et al., 2000), and the nerves to iliococcygeus/pubococcygeus muscles (Pacheco et al. 1989, Kamo et al., 2003, 2004), respectively. The increasing effects were greatly suppressed by bilateral transection of both the pudendal nerves and the nerves to iliococcygeus/pubococcygeus muscles. In contrast, bilateral transection of the hypogastric nerves did not change the effect, suggesting a greater contribution of the striated muscles to the effects of TAK-233. In addition, since urethra-closing responses during sneezing are themselves totally abolished by bilateral transection of the nerves innervating the striated muscles (Kamo et al, 2003), it seems reasonable to consider
that the urethral closing-increasing effects of TAK-233 during sneezing are also mediated by elevating striated muscle function. As striated muscle-mediated mechanisms greatly contribute to momentary reflex urethral closures (Kamo et al., 2009), the effects of TAK-233 may be more amenable to detect during momentary urethral closure than during continuous measurement.

It is considered highly likely that the site of action of TAK-233 for urethral closure-enhancing effects is located at Onuf’s nucleus in the sacral spinal cord similar to other 5-HT$_{2C}$ receptor agonists (Yoshimura and Miyazato, 2012). Spinal 5-HT$_{2C}$ receptor stimulation by intrathecal drug injection enhances urethra-closing responses in rats (Miyazato et al., 2009), and the current nerve transection study revealed that striated muscle, not smooth muscle, components greatly contributed to the effects of TAK-233, and the motor neurons innervating the striated external urethral sphincter via the pudendal nerves are located at Onuf’s nucleus (Rajaofetra et al., 1992, Thor, 2003, 2004). Furthermore, the reflex pathway from the Onuf’s nucleus to the urethra is common between the 2 urethra-closing reflexes (Kamo et al., 2009, Yoshimura and Miyazato, 2012) and TAK-233 enhanced the urethra-closing function mediated by both types of reflexes. Therefore, it seems natural to conclude that TAK-233 acts on 5-HT$_{2C}$ receptors on motor neurons in Onuf’s nucleus to enhance both types of reflexes, resulting in the elevation of urethral resistance under momentary $P_{ves}$ elevation.

The effects of 5-HT$_{2C}$ receptor stimulation on urethra-closing functions in healthy female
humans were studied with TAK-233, with duloxetine as a reference drug known to reduce the frequency of stress urinary incontinence episodes in patients (Mariappan et al., 2007). Based on the putative mechanism of TAK-233, effects on the sensitivity of evoked urethral responses were primarily investigated by measuring stimulus threshold for inducing TMS-evoked momentary urethral sphincter contraction. Two published studies demonstrate that duloxetine increases the sensitivity of the evoked urethral responses (Boy et al., 2006; Yono et al., 2015), suggesting this as one potential mechanism for a drug for the treatment of stress urinary incontinence. Our pharmacodynamic study in humans obtained the same finding, with TAK-233 significantly lowering the stimulus threshold even at the lower dose (20 mg), and those effects generally following the time course of its pharmacokinetic profile. These findings indicate that TAK-233 increased the sensitivity of the evoked momentary urethra-closing mechanism and the 5-HT$_{2C}$ receptor-mediated enhancing mechanism functions in humans.

Compared with the effects of TAK-233 on the stimulus threshold, its effect on urethral pressure at rest required higher plasma drug levels. While TAK-233 plasma levels of 125 ng/ml after administration at 90 mg were required for increasing the maximum urethral pressure at rest, a plasma level around 30 ng/ml after administration at 20 mg was enough to increase the sensitivity of the evoked momentary contractions. It seems reasonable to consider that the more the urethra-closing reflex pathway is stimulated, such as during the evoked urethral contractions, the
more the effects of 5-HT$_{2C}$ receptor stimulation become obvious, and that the reflex does not function to any great extent during urethral pressure profile measurement at rest since the bladder was emptied before measurement of the reflex. Recently, another 5-HT$_{2C}$ receptor agonist, ASP2205, demonstrated no positive effects on the opening urethral pressure, both at rest and with squeezing, in healthy female subjects (Klarskov et al., 2019). Healthy subjects may show maximal striated muscle-mediated closure during squeezing even without drug administration, and duloxetine might increase smooth muscle-mediated adrenergic urethral closure mechanism.

As shown in the current human pharmacodynamic study, the time profile of change in evoked urethral closure-enhancing effects of each drug was generally consistent with the time course of the plasma drug levels, indicating that exceeding a plasma level threshold is necessary for the pharmacodynamic effects of respective drugs. Meanwhile, the required duration of an effective treatment may vary depending on each patient with stress urinary incontinence and their situation. For example, a patient may only need a short effective duration while engaging in daily activities for short periods of time, whereas another patient may need a longer effective duration to cover a full working day. The plasma levels of TAK-233 rapidly elevated after oral administration and then sharply declined after reaching its peak level. This pharmacokinetic profile, such as that demonstrated for TAK-233, in combination with slow-release formulation technology, etc., may make it possible to provide various medicines with different effective durations on a flexible basis,
since in almost all cases, rapid achievement of the plasma level threshold is required.

The current studies with TAK-233 in female rats and humans demonstrate that 5-HT$_{2C}$ receptor stimulation enhances striated urethra-closing functions during both evoked and momentary events, and consequently, 5-HT$_{2C}$ receptor agonists may provide effective treatment options for patients with stress urinary incontinence. However, the effects of 5-HT$_{2C}$ receptor stimulation on the frequency of stress urinary incontinence episodes remain unclear, requiring further clinical studies.
Acknowledgement

We would like to thank project chemists, especially Shigekazu Sasaki, Tomokazu Kusumoto, Izumi Nomura, Hironobu Maezaki as TAK-233 inventors, for identifying TAK-233 after their great efforts in medicinal chemistry studies. We would like to thank the investigator and other staff members for dedicated operation for the clinical study.
Authorship Contributions

Participated in research and clinical study design: Izumi, Hiroshi, Gale, Takuya, Akio, Kyoko, Yuya

Conducted experiments and the clinical study: Hiroshi, Takuya, Akio, Masako, Satoshi, Yuya

Performed data analysis: Izumi, Hiroshi, Gale, Takuya, Akio, Masako, Satoshi, Kyoko, Yuya

Wrote or contributed to the writing of the manuscript: Izumi, Akio, Yuya

All authors were full-time employees of Takeda Pharmaceutical Company Ltd or Takeda Development Center Europe Ltd when the studies were conducted, and 5 authors such as Izumi, Akio, Masako, Kyoko and Yuya are currently full-time employees of Takeda Pharmaceutical Company Ltd. Izumi, Akio and Masako own shares in Takeda Pharmaceutical Company Ltd, and Izumi is one of inventors in WO2019131902A1.
References

Boy S, Reitz A, Wirth B, Knapp PA, Braun PM, Haferkamp A, and Schurch B (2006)

Facilitatory neuromodulative effect of duloxetine on pudendal motor neurons controlling
the urethral pressure: a functional urodynamic study in healthy women. *Eur Urol*

50:119-125.

Conlon K, Miner W, McCleary S, and McMurray G (2012) Identification of 5-HT(2C)
mediated mechanisms involved in urethral sphincter reflexes in a guinea-pig model of
urethral function. *BJU international* 110: E113-117.

De Groat WC, Booth AM, and Yoshimura N (1993) Neurophysiology of micturition
and its modification in animal models of human disease, in *The Autonomic Nervous
System Nervous Control of the Urogenital System*, 1st ed (Maggi CA eds) pp 227–290,
Harwood, London.

Hoyer D, Hannon JP, and Martin GR (2002) Molecular, pharmacological and functional
diversity of 5-HT receptors. *Pharmacol Biochem Behav* 71: 533-554.

Kaiho Y, Kamo I, Chancellor MB, Arai Y, de Groat WC, and Yoshimura N (2007) Role of
noradrenergic pathways in sneeze-induced urethral continence reflex in rats. *Am J
Physiol Renal Physiol* 292: F639-646.

Kamo I, Cannon TW, Conway DA, Torimoto K, Chancellor MB, de Groat WC, and
Yoshimura N (2004) The role of bladder-to-urethral reflexes in urinary continence mechanisms in rats. *Am J Physiol Renal Physiol* **287**: F434-441.

Kamo I, and Hashimoto T (2007) Involvement of reflex urethral closure mechanisms in urethral resistance under momentary stress condition induced by electrical stimulation of rat abdomen. *Am J Physiol Renal Physiol* **293**: F920-926.

Kamo I, Kaiho Y, Canon TW, Chancellor MB, de Groat WC, Prantil RL, Vorp DA, and Yoshimura N (2006) Functional analysis of active urethral closure mechanisms under sneeze induced stress condition in a rat model of birth trauma. *J Urol* **176**: 2711-2715.

Kamo I, Kaiho Y, Miyazato M, Torimoto K, and Yoshimura N (2009) Two kinds of urinary continence reflexes during abrupt elevation of intravesical pressure in rats. *Lower urinary tract symptoms* **1**(s1): S40-S43.

Kamo I, Torimoto K, Chancellor MB, de Groat WC, and Yoshimura N (2003) Urethral closure mechanisms under sneeze-induced stress condition in rats: a new animal model for evaluation of stress urinary incontinence. *Am J Physiol Regul Integr Comp Physiol* **285**: R356-365.

Kennett GA, Wood MD, Bright F, Trail B, Riley G, Holland V, Avenell KY, Stean T, Upton N, Bromidge S, Forbes IT, Brown AM, Middlemiss DN, and Blackburn TP (1997) SB 242084, a selective and brain penetrant 5-HT2C receptor antagonist.
Neuropharmacology 36: 609-620.

Klarskov N, Till OV, Sawyer W, Cernus D, and Sawyer W (2019) Effect of a 5-HT\textsubscript{2C} receptor agonist on urethral closure mechanism in healthy women. Neurourol Urodyn 38: 1700-1706.

Manzo J, Vazquez MI, Cruz MR, Hernandez ME, Carrillo P, and Pacheco P (2000) Fertility ratio in male rats: effects after denervation of two pelvic floor muscles. Physiol Behav 68: 611-618.

Mariappan P, Alhasso A, Ballantyne Z, Grant A, and N’Dow J (2007) Duloxetine, a serotonin and noradrenaline reuptake inhibitor (SNRI) for the treatment of stress urinary incontinence: a systematic review. Eur Urol 51: 67-74.

Miyazato M, Kaiho Y, Kamo I, Chancellor MB, Sugaya K, de Groat WC, and Yoshimura N (2008) Effect of duloxetine, a norepinephrine and serotonin reuptake inhibitor, on sneeze-induced urethral continence reflex in rats. Am J Physiol Renal Physiol 295: F264-271.

Miyazato M, Kaiho Y, Kamo I, Kitta T, Chancellor MB, Sugaya K, Arai Y, de Groat WC, and Yoshimura N (2009) Role of spinal serotonergic pathways in sneeze-induced urethral continence reflex in rats. Am J Physiol Renal Physiol 297: F1024-1031.

Ouchi M, Kitta T, Kanno Y, Higuchi M, Togo M, Moriya K, and Shinohara N (2018)
Effect of a 5-HT$_{2c}$ receptor agonist on urethral closure mechanisms in female rats.

*Neurourol Urodyn* **37**: 2382-2388.

Pacheco P, Martinez-Gomez M, Whipple B, Beyer C, and Komisaruk BR (1989) Somato-motor components of the pelvic and pudendal nerves of the female rat. *Brain Res* **490**: 85–94.

Rajaofetra N, Passagia JG, Marlier L, Poulat P, Pellas F, Sandillon F, Verschuere B, Gouy D, Geffard M, and Privat A (1992) Serotonergic, noradrenergic, and peptidergic innervation of Onuf’s nucleus of normal and transected spinal cords of baboons (Papio papio). *J Comp Neurol* **318**: 1-17.

Suzuki T, Shimizu T, Kwon J, Takaoka E, Yoshikawa S, Sumino Y, Kitta T, Miyazato M, Miyake H, and Yoshimura N (2018) Role of the serotonergic system in urethral continence reflexes during sneezing in rats. *Am J Physiol Renal Physiol* **315**: F79-F85.

Thor KB (2003) Serotonin and norepinephrine involvement in efferent pathways to the urethral rhabdosphincter: implications for treating stress urinary incontinence. *Urology* **62**: 3-9.

Thor KB (2004) Targeting serotonin and norepinephrine receptors in stress urinary incontinence. *Int J Gynaecol Obstet* **86**: S38-52.

Thor KB, and Katofiasc MA (1995) Effects of duloxetine, a combined serotonin and
norepinephrine reuptake inhibitor, on central neural control of lower urinary tract function in the chloralose-anesthetized female cat. *J Pharmacol Exp Ther* **274**: 1014-1024.

Yono M, Otani M, Ito K, Inoue Y, Furukawa K, Hori M, Tsuji S, Tanaka T, Sakata Y, and Irie S (2015) Effect of duloxetine on urethral resting pressure and on sphincter contractility in response to coughing and magnetic stimulation in healthy women. *Low Urin Tract Symptoms* **7**: 93-98.

Yoshimura N, and Miyazato M (2012) Neurophysiology and therapeutic receptor targets for stress urinary incontinence. *Int J Urol* **19**: 524-537.
Footnotes

This work was supported by Takeda Pharmaceutical Company Ltd. This work received no external funding.

Data describing in Figures 2 to 6 and “Reflex urethra-closing responses in rats” section in “Results” (body text) have been already disclosed in WO2019131902A1 for patent filing.
Figure Legends

Figure-1. Chemical structure of TAK-233

Figure-2. Effects of TAK-233 and duloxetine on the urethral resistance during sneezing in rats with vaginal distention. In all rats, pelvic nerves were bilaterally transected. Sneeze leak point pressures (S-LPPs) were measured before and 30 min after intraduodenal (i.d.) drug administration. Data are expressed as the mean ± SEM of increase in S-LPP. Number of rats evaluated was 8, 8, 4, 6, 8 and 3 (vehicle, TAK-233 1 and 3 mg/kg, duloxetine 10, 20 and 40 mg/kg, respectively). ** P<0.01, *** P<0.001 compared with the vehicle-treated group by Dunnett’s test. ### P<0.001 compared by Student’s t-test.

Figure-3. Effects of TAK-233 (A) and duloxetine (B) on the urethral resistance during momentary intravesical pressure elevation by electrical stimulation of abdominal muscles in rats. In all rats, the spinal cord was transected at the T8-9 level and the nerve to iliococcygeus/pubococcygeus muscles was unilaterally transected to reduce the urethral resistance. Leak point pressures (LPPs) were measured before and 30 min after intraduodenal (i.d.) drug administration. Data are expressed as the mean ± SEM in 6 or 8 rats for the experiment with TAK-233 (A) or duloxetine (B), respectively. Mean ± SEM
of baseline LPP values were 46.1 ± 6.4 cmH2O, 47.2 ± 5.7 cmH2O, 47.3 ± 5.0 cmH2O, and 47.6 ± 3.1 cmH2O (TAK-233 0, 0.3, 1, and 3 mg/kg, i.d., respectively), and 35.6 ± 4.3 cmH2O, 36.6 ± 3.0 cmH2O, 35.3 ± 3.0 cmH2O, and 36.9 ± 2.8 cmH2O (duloxetine 0, 5, 10, and 20 mg/kg, i.d., respectively). * p≤0.05, ** p≤0.01, *** p≤0.001 compared with the vehicle-treated group by Dunnett’s test.

Figure-4. Blocking effects of intravenously injected SB 242084 on the urethral resistance-increasing effects of TAK-233 during momentary intravesical pressure elevation by electrical stimulation of abdominal muscles in rats. In all rats, the spinal cord was transected at the T8-9 level and the nerve to iliococcygeus/pubococcygeus muscles was unilaterally transected to reduce urethral resistance. Five min after intravenous (i.v.) injection of SB 242084 or vehicle, TAK-233 or vehicle was administered by intraduodenal (i.d.) administration. Leak point pressures (LPPs) were measured before SB 242084 and 30 min after TAK-233 administrations. Data are expressed as the mean ± SEM in 5 rats. Mean ± SEM of baseline LPP values were 46.1 ± 5.2 cmH2O, 45.9 ± 5.4 cmH2O, 46.0 ± 5.1 cmH2O, 44.0 ± 7.4 cmH2O, 45.0 ± 6.7 cmH2O, and 44.3 ± 7.0 cmH2O (TAK-233 0 mg/kg/SB 242084 0 mg/kg, TAK-233 0 mg/kg/SB 242084 0.3 mg/kg, TAK-233 3 mg/kg/SB 242084 0 mg/kg, TAK-233 3 mg/kg/SB 242084 0.03 mg/kg, TAK-233 3 mg/kg/SB 242084 0.1 mg/kg, TAK-233 3 mg/kg/SB 242084 0.3 mg/kg, respectively). * P≤0.05
compared with the vehicles i.v. and i.d. administered group by Dunnett’s test. # P≤0.05 compared with the vehicle i.v. and TAK-233 i.d. administered group by Dunnett’s test.

Figure-5. Typical recordings of the middle urethral responses measured by microtip transducer catheter during increments of intravesical pressure (P<sub>ves</sub>) in rats with the spinal cord transection at T-8-9 level. Responses are shown before (Pre) and 30 min after intraduodenal administration of TAK-233 at 3 mg/kg (Post).

Figure-6. Time profiles of change from baseline in motor threshold (MT) for urethral sphincter contraction in response to transcranial magnetic stimulation in healthy female subjects. Data are expressed as the mean ± SD of % change from the baseline MT values. Mean ± SD of baseline MT values were 68.0 ± 9.97, 66.3 ± 12.27, 70.0 ± 10.22, and 67.8 ± 11.46 (placebo, TAK-233 20 mg, TAK-233 90 mg, Duloxetine 40 mg, respectively). + Changes from baseline was significantly lower than that in the placebo group by analysis of variance (ANOVA).

Figure-7. Time profiles of change from baseline in maximal urethral pressure at rest in the entire functional urethral length in healthy female subjects. Data are expressed as the mean ± SD of change from the baseline values.
### Tables

Table-1. *In vitro* agonist potency of 5-HT, TAK-233, and lorcaserin for human 5-HT\(_{2C}\), 5-HT\(_{2A}\), 5-HT\(_{2B}\) and rat 5-HT\(_{2C}\) receptors in calcium influx assays.

| Receptor     | 5-HT\(_{2C}\) | TAK-233 | Lorcaserin |
|--------------|--------------|---------|------------|
| human 5-HT\(_{2C}\) | EC\(_{50}\) (nM) | 0.12 | 2.6 | 0.81 |
|              | 95% CI       | 0.07 - 0.23 | 1.6 - 4.1 | 0.54 - 1.18 |
|              | Response at 10 μM (%*) | 100.0 | 94.7 | 95.3 |
|              | Relative potency to 5-HT** | 1.0 | 21 | 6.5 |
| human 5-HT\(_{2A}\) | EC\(_{50}\) (nM) | 0.86 | 465 | 61 |
|              | 95% CI       | 0.66 - 0.95 | 377 - 570 | 36 - 104 |
|              | Response at 10 μM (%*) | 100.0 | 93.2 | 83.0 |
|              | Relative potency to 5-HT** | 1.0 | 541 | 71 |
| human 5-HT\(_{2B}\) | EC\(_{50}\) (nM) | 1.5 | 8459 | 666 |
|              | 95% CI       | 1.1 - 1.9 | 7117 - 10249 | 466 - 963 |
|              | Response at 10 μM (%*) | 100.0 | 53.8 | 74.0 |
|              | Relative potency to 5-HT** | 1.0 | 5818 | 458 |

This article has not been copyedited and formatted. The final version may differ from this version.
| rat 5-HT<sub>2C</sub> | EC<sub>50</sub> (nM) | 10 | 633 | 172 |
|---------------------|-------------------|----|-----|-----|
| 95% CI              | 6.5 - 17          | 421 - 958 | 100 - 296 |
| Response at 10 μM (%)* | 100.0            | 78.2 | 82.1 |
| Relative potency to 5-HT** | 1.0             | 61 | 16 |

CI expresses confidence interval.

* Agonistic response at 10 μM was normalized to that observed with 5-HT at 10 μM.

** Relative potency was calculated by dividing EC<sub>50</sub> value of a compound by EC<sub>50</sub> value of 5-HT.
Table 2. Effects of transection of various nerves relating to contractions of the urethra and pelvic floor muscles on the leak point pressure (LPP)-increasing effects of TAK-233 in rats.

| Nerve       | TAK-233 (mg.kg, i.d.) | LPP (cmH₂O) | Increase vs Sham |
|-------------|-----------------------|-------------|------------------|
|             | Pre       | Post       |                  |
| Sham        | 0         | 3          |                  |
|             | 54.1 ± 13.3 | 52.7 ± 13.5 | -1.4 ± 1.9 |
|             | 51.3 ± 3.4  | 76.9 ± 5.2  | 25.6 ± 3.5 ***  |
| Pelvic      | 0         | 3          |                  |
|             | 33.1 ± 6.1 | 34.6 ± 6.9 | 1.5 ± 1.5        |
|             | 30.0 ± 2.5 | 29.8 ± 3.6 | -0.3 ± 1.2      | ### |
| Hypogastric | 0         | 3          |                  |
|             | 40.8 ± 7.0 | 41.5 ± 5.9 | 0.7 ± 1.2        |
|             | 42.9 ± 6.3 | 69.0 ± 7.9 | 26.1 ± 4.2 **   | N.S. |
| Pudendal    | 0         | 3          |                  |
|             | 33.8 ± 4.0 | 32.8 ± 3.5 | -1.0 ± 1.8       |
|             | 30.6 ± 3.1 | 38.9 ± 3.6 | 8.3 ± 2.1 *     | #    |
| Ilio/Pubo   | 0         | 3          |                  |
|             | 35.8 ± 4.9 | 32.7 ± 5.1 | -3.1 ± 1.2       |
|             | 41.9 ± 8.9 | 66.5 ± 11.7| 24.6 ± 6.1 **   | N.S. |
| Pudendal    | 0         | 3          |                  |
|             | 27.0 ± 4.4 | 28.3 ± 4.5 | 1.3 ± 0.7        |
| + Ilio/Pubo | 3         | 36.8 ± 6.9 | 43.0 ± 7.0       | 6.1 ± 2.5 ** |

In all rats, the spinal cord was transected at T8-9 level. The pelvic nerves, the hypogastric...
nerves, the pudendal nerves, the nerves to iliococcygeus/pubococcygeus muscles (Ilio/Pubo), or both the pudendal nerves and Ilio/Pubo were bilaterally transected. Data are expressed as the mean ± SEM in 5 rats.

* P \leq 0.05, ** P \leq 0.01, *** P \leq 0.001 compared with the vehicle-administered group in rats with the same nerves transected by Student’s t-test.

# P \leq 0.05, ## P \leq 0.01, ### P \leq 0.001: LPP increase after TAK-233 administration was compared with that in the sham-operated group by Dunnett’s test.
Table 3. Plasma concentration alteration of TAK-233 and duloxetine in healthy female subjects.

| Drug     | Dose (mg) | Pre       | 0.5 h | 1 h | 2 h | 3 h | 6 h | 8 h | 12 h | 24 h |
|----------|-----------|-----------|-------|-----|-----|-----|-----|-----|------|------|
| TAK-233  | 20        | 0.0       | 29.2  | 25.2| 20.4| 14.3| 3.5 | 1.4 | 0.1  | -1.4 |
|          |           | ±         | ±     | ±   | ±   | ±   | ±   | ±   | ±    | ±    |
|          | 90        | 0.0       | 127.3 | 126.0| 152.8| 122.4| 52.8| 24.7| 5.6  | -    |
|          |           | ±         | ±     | ±   | ±   | ±   | ±   | ±   | ±    | ±    |
|          |           | 0.0       | 79.2  | 59.2| 68.1| 52.4| 31.4| 16.5| 4.5  | -    |
| Duloxetine| 40        | 0.0       | 0.0   | -   | -   | 6.0 | 32.4| 29.2| 21.1 | 9.4  |
|          |           | ±         | ±     | ±   | ±   | ±   | ±   | ±   | ±    | ±    |
|          | 0.0       | 0.0       | 7.3   | 15.7| 13.0| 9.9 | 5.8 | -   | -    | -    |

Data are expressed as the mean ± SD in 23 subjects.

-: not tested
Figure-1

This article has not been copyedited and formatted. The final version may differ from this version.
Figure 2

**Increase in S-LPP (cmH₂O)**

|                | Vehicle | TAK-233 (mg/kg, i.d) | Duloxetine (mg/kg, i.d) |
|----------------|---------|-----------------------|-------------------------|
|                | 1       | 3                     | 10                      |
|                | 20      | 40                    | 20                      |

Significance levels:
- **P < 0.001**
- **P < 0.01**
- **P < 0.05**

This article has not been copyedited and formatted. The final version may differ from this version.

JPET Fast Forward. Published on April 9, 2021 as DOI: 10.1124/jpet.121.000573
Figure-3

(A) Increase in LPP (cmH₂O) for Vehicle and TAK-233 (mg/kg, i.d.).

(B) Increase in LPP (cmH₂O) for Vehicle and Duloxetine (mg/kg, i.d.).
Figure-4

![Graph showing the increase in LPP (cmH₂O) with SB 242084 (mg/kg, i.v.) and TAK-233 (mg/kg, i.d.)]
Figure-5

This article has not been copyedited and formatted. The final version may differ from this version.
Figure-6

% Change in MT from baseline

Time after administration (h)

Placebo (n=23)
TAK-233 20 mg (n=24)
TAK-233 90 mg (n=23)
Duloxetine 40 mg (n=23)
Figure 7

Figure 7

This article has not been copyedited and formatted. The final version may differ from this version.