Corresponding Author Name: Zhiqun Shang
Manuscript Number: CDDIS-22-2076

Reporting Summary

Springer Nature wishes to improve the reproducibility of the work that we publish. This checklist is used to ensure good reporting standards and to improve the reproducibility. Please respond completely to all questions relevant to your manuscript. For more information, please read the journal’s Guide to Authors.

☐ Check here to confirm that the following information is available in the Material & Methods section:

- The exact sample size (n) for each experimental group/condition, given as a number, not a range
- A description of the sample collection allowing the reader to understand whether the samples represent technical or biological replicates (including how many animals, litters, culture, etc.)
- A statement of how many times the experiment shown was replicated in the laboratory
- Definitions of statistical methods and measures: For small sample sizes (n<5) descriptive statistics are not appropriate, instead plot individual data points
  - Very common tests, such as t-test, simple χ² tests, Wilcoxon and Mann-Whitney tests, can be unambiguously identified by name only, but more complex techniques should be described in the methods section
  - Are tests one-sided or two-sided?
  - Are there adjustments for multiple comparisons?
  - Statistical test results, e.g., P values
  - Definition of ‘center values’ as median or mean;
  - Definition of error bars as s.d. or s.e.m. or c.i.

Please ensure that the answers to the following questions are reported in the manuscript itself. We encourage you to include a specific subsection in the methods section for statistics, reagents and animal models. Below, provide the page number or section and paragraph number.

Statistics and general methods

1. How was the sample size chosen to ensure adequate power to detect a pre-specified effect size? (Give section/paragraph or page #)

   For animal studies, include a statement about sample size estimate even if no statistical methods were used.

2. Describe inclusion/exclusion criteria if samples or animals were excluded from the analysis. Were the criteria pre-established? (Give section/paragraph or page #)

   For animal studies, include a statement about the analysis even if no statistical methods were used.

3. If a method of randomization was used to determine how samples/animals were allocated to experimental groups and processed, describe it. (Give section/paragraph or page #)

   For animal studies, include a statement about randomization even if no randomization was used.

| Reported in section/paragraph or page # |
|----------------------------------------|
| See "Statistical analysis” section in the Material and Method of the manuscript. No statistical methods were used to predetermine the sample size. |
| No statistical methods were used to predetermine the sample size. See "Statistical analysis” section in the Material and Method of the manuscript. |
| No data were excluded. See "Statistical analysis" section in the Material and Method of the manuscript. |
| Nude mice were randomly allocated to experimental groups. See "Xenograft assay" section in the Material and Method of the manuscript. |
| Animals are randomized based on the body weight. |
4. If the investigator was blinded to the group allocation during the experiment and/or when assessing the outcome, state the extent of blinding. (Give section/paragraph or page #)

For animal studies, include a statement about blinding even if no blinding was done.

5. For every figure, are statistical tests justified as appropriate?

Do the data meet the assumptions of the tests (e.g., normal distribution)?

Is there an estimate of variation within each group of data?

Is the variance similar between the groups that are being statistically compared? (Give section/paragraph or page #)

**Reagents**

6. Report the source of antibodies (vendor and catalog number)

7. Identify the source of cell lines and report if they were recently authenticated (e.g., by STR profiling) and tested for mycoplasma contamination

**Animal Models**

8. Report species, strain, sex and age of animals

9. For experiments involving live vertebrates, include a statement of compliance with ethical regulations and identify the committee(s) approving the experiments.

10. We recommend consulting the ARRIVE guidelines (PloS Biol. 8(6), e1000412, 2010) to ensure that other relevant aspects of animal studies are adequately reported.

---

The investigators were not blinded to group allocation during experiments. Conclusions were made based on quantitative parameters and statistical significance of the data, and thus on experimental observations, independent of blinding.

The investigators were not blinded to group allocation during experiments.

The statistical test used in each figure is indicated in the figure legends.

Given that sample size is less than 30, we assume that the data did not follow Gaussian distribution and therefore systematically apply a non-parametric analysis.

No estimation of variation within each group was computed.

It does not require estimation of the variance.

Yes, it is done in the Material and Method section.

Yes, the source of the cell lines are mentioned in the Materials and Methods section of the manuscript. We test for mycoplasma contamination every six months at the laboratory.

TRAMP (Transgenic Adenocarcinoma of Mouse Prostate) [C57BL/6] mice were obtained from the Jackson

See "Xenograft assay" section in the Material and Method of the manuscript.
**Human subjects**

11. Identify the committee(s) approving the study protocol.

12. Include a statement confirming that informed consent was obtained from all subjects.

13. For publication of patient photos, include a statement confirming that consent to publish was obtained.

14. Report the clinical trial registration number (at ClinicalTrials.gov or equivalent).

15. For phase II and III randomized controlled trials, please refer to the CONSORT statement and submit the CONSORT checklist with your submission.

16. For tumor marker prognostic studies, we recommend that you follow the REMARK reporting guidelines.

**Data deposition**

17. Provide accession codes for deposited data. Data deposition in a public repository is mandatory for:
   a. Protein, DNA and RNA sequences
   b. Macromolecular structures
   c. Crystallographic data for small molecules
   d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available in the Guide to Authors. We encourage the provision of other source data in supplementary information or in unstructured repositories such as Figshare and Dryad. We encourage publication of Data Descriptors (see Scientific Data) to maximize data reuse.

18. If computer code was used to generate results that are central to the paper’s conclusions, include a statement in the Methods section under “Code availability” to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

---

**Reported in section/paragraph or page #**

|   |   |
|---|---|
| Tissue microarrays (TMA) for prostate cancer (PCa) specimens were obtained from the second hospital of Tianjin Medical University, acquiring the due consent from the patients and Yes, See "Human prostate cancer specimens" section in the Material and Method of the manuscript. |
| n/a | n/a |
| The gene expression data for DHT-treated LNCaP cells generated in this study has been submitted to the NCBI Gene Expression Omnibus under the accession number GSE163539. The gene expression data and binding data in this study have been deposited into CNGB Sequence Archive (CNSA) of China National GeneBank DataBase (CNGBdb) with accession number CNP0001560 and CNP0001628. |

See "RNA-seq and pathway analysis" section in the Material and Method of the manuscript.