Exosomes as therapeutic vehicles in liver diseases

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Abstract: The diagnosis and treatment of various liver diseases have progressed greatly over the years, but clinical outcomes are still not satisfying. New research on the mechanisms and application thereof may effectuate positive changes. Exosomes are membrane-derived nanovesicles ranging in size from 40 to 160 nm and are released by a diversity of cells. They contain a variety of cargo, including lipids, proteins, coding RNAs, and noncoding RNAs. Recent studies have recognized exosomes as intercellular communication agents, which play important roles in physiological or biological processes in acute or chronic liver disorders by horizontal transferring of genetic bioinformation from donor cells to neighboring or distal target cells. In the hope that exosomes can potentially be used as vehicles for clinical intervention, this review aims to focus on the roles of exosomes and their cargo in the field of various liver disorders, including virus-related liver diseases, alcoholic liver diseases (ALD), nonalcoholic fatty liver diseases (NAFLD), and liver cancer. In addition, many studies have indicated that mesenchymal stem cell (MSC)-derived exosomes or engineered MSC-derived exosomes can also exert hepatoprotection, antioxidation, or enhance drug sensitivity on corresponding liver diseases with the advantage of low immunogenicity and high biocompatibility. Overall, exosomes are expected to serve as an important therapeutic tool for various liver diseases. However, there are still many problems that need to be resolved by further research and a greater body of evidence before exosomes are ready for clinical application.

Keywords: Exosomes; miRNA; mesenchymal stem cell (MSC); liver disease

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Introduction

Exosomes were first described as a means to eliminate cellular waste by Pan et al. in 1985 (1). While studying the maturation process of erythrocytes, they found that exosomes served to remove unnecessary components from cells. The terms “exosome” and “extracellular vesicle (EV)” were once used interchangeably in many publications. However, it is now widely accepted that an exosome is a membrane vesicle structure ranging from 40 to 160 nm in size that originates from multivesicular bodies (MVBs) through endosomal pathways, which consist of endosome formation and exocytosis (2). The formation of MVBs starts from the early sorting of endosomes (ESEs) which are generated from the invagination of plasma membrane and the endocytosis of biomolecules (3). After periods of maturation, the late endosomal membrane buds inward

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to form the intraluminal vesicles (ILVs), which leads to the formation of MVBs (4). Some of MBVs can be fused with plasma membrane, and subsequently release ILVs into the extracellular space, and these can be defined as exosomes (2,5-7). Other MBVs can be fused with lysosomes or autophagosomes for self-degradation (3) (Figure 1, A). Recent research (8,9) has recognized that exosomes serve as intercellular communication agents which contain various biomolecules, such as lipids, proteins, amino acids, coding RNAs, and noncoding RNAs. In this capacity, exosomes play critical roles in physiological or biological processes in acute or chronic liver disorders by the horizontal transfer of genetic bioinformation from donor cells to neighboring or distal target cells. Other studies (4) have shown that exosomes can be secreted by the majority of cell types. Depending on the cell types released, the quantity, contents, and biological characteristics of exosomes can promote or inhibit the physiological or pathological progression of cells. For instance, normal hepatocytes were found to release a small number of exosomes to regulate liver repair and regeneration (10). Meanwhile, stressed hepatocytes were found to increase exosome release and the expression of cellular mRNA, which modulates the transcriptional process of adjacent hepatocytes and hepatic stellate cells (HSCs) (11). Exosomes released by nonparenchymal cells, such as sinusoidal endothelial cells, Kupffer cells, and HSCs, also participate in the regulation of liver function physiologically or pathologically. Moreover, liver tumor cells exert great influence on regulating the growth, angiogenesis, proliferation, and metastasis of neoplasm by releasing exosomes (12). Such exosome-modulated responses can be disease promoting or suppressing (3).

Accumulating evidence (13,14) has clarified the functions of exosomes in the hepatic pathological state: due to their low immunogenicity and good biocompatibility, exosomes may transfer biomolecules to the target cells without RNA degradation and loss of biological information. Thus, there has been increasing emphasis on the use of exosomes to control the progression of liver diseases. There are three main ways in which exosomes can be manipulated to facilitate liver regeneration, regulate inflammation and fibrosis, or inhibit tumor growth and metastasis: (I) direct regulation of the release of exosomes from particular cell types (15,16); (II) interference with exosome cargo (17); and (III) delivery of drugs into exosomes (18). Indeed, the study of exosomes has become a very active field. Ongoing experiments may discover more about exosome functions, which may in turn help to enhance the diagnosis and treatment of a variety of diseases (3).

**Hepatitis B**

Hepatitis B virus (HBV) infection has become a major global health problem (19). According to the World Health Organization (WHO) Global Hepatitis Report 2017, 257 million people were estimated to be chronically infected with HBV worldwide. People who are chronically infected with HBV may develop cirrhosis or hepatocellular carcinoma (HCC), a potentially fatal disease. Hence, it is crucial to understand the pathogenesis of HBV in order to better intervene in liver diseases. After HBV reaches the liver and infects the hepatocytes through the bloodstream, its envelope fuses with the hepatocyte membrane, and the HBV genome, a partially double-stranded, relaxed circular DNA (rcDNA), is transported to the nucleus of host hepatocytes and converted into covalently closed circular DNA (cccDNA) which serves as the template to synthesize the pregenomic mRNA after multiple steps (20-22). HBV DNA is then created by reverse transcription and may be integrated into the chromosome. Based on these characteristics, exploring the roles of exosomes in HBV replication, transmission, and immune responses is particularly important. HBV X protein (HBx), a multifunctional viral regulator, figures prominently in HBV duplication and viral carcinogenesis by interacting with host factors (19). It can surpass the host exosomal biogenesis mechanism by enhancing the activity of neutral sphingomyelinase 2 and interacting with foreign biomarkers such as neutral sphingomyelinase 2, CD9, and CD81 (23). Moreover, a growing body of evidence suggests that exosomes that contain the HBV genome can be released from infected hepatocytes to adjacent normal cells, leading to the spread of virus (24,25). Yang et al. (26) highlighted that carboxyfluorescein diacetate succinimidyl ester (CFSE)-labeled HLCZ01 cells incubated with HBV-positive exosomes labeled with 4-chlorobenzenesulfnate salt (DiD) were shown to be positive for hepatitis B surface antigen (HBsAg) and core antigen (HBcAg) after 2 days of exposure. We can thus infer that the spread of the virus can be blocked by regulated exosomes. Furthermore, Wang et al. (27) confirmed that the exosome-mediated clustered regularly interspaced short palindromic repeats (CRISPR)-CRISPR-associated protein 9 (Cas9) system could cut the intercellular transmission function of the HBV genome (Figure 1, A). Additionally, exosomes released after virus infection can also modulate immune responses to control
Figure 1 Role of exosomes in liver diseases. (A) The biogenesis of exosomes. Exosomes are generated from the formation of ESE, LSE, and MVBs in sequence. In cytoplasm, ILVs are either degraded by lysosomes or released to the extracellular environment as exosomes. (B) Exosomes exert antiviral effects during viral hepatitis. Exosomes could induce viral genome degradation and activate immune cells. (C) Exosomes have functions in antifibrosis in alcoholic liver disease and nonalcoholic fatty liver diseases. (D) Role of exosomes in HCC. Exosomes participate in the inhibition of tumor growth, metastasis, cell division, and the activation of CTL. (E) Multiple roles of MSC-derived exosomes. MSC-derived exosomes involved in injured hepatocyte repair, HSC inactivation, anti-inflammation, and antitumor and chemosensitivity promotion. ESE, early sorting endosome; LSE, late sorting endosome; MVB, multivesicular bodies; ILV, intraluminal vesicle; HCC, hepatocellular carcinoma; MSC, mesenchymal stem cell; HSC, hepatic stellate cell; ECM, extracellular matrix; NK cell, natural killer cell; CTL, cytotoxic T cell; ER, endoplasmic reticulum; Golgi, Golgi apparatus.
the virus (28). Kouwaki et al. (29) discovered that exosomes containing viral genes derived from HBV-infected hepatocytes in vitro could induce the expression of natural killer group 2, member D (NKG2D) ligand in macrophages through myeloid differentiation factor 88 (MyD88), TIR domain-containing adaptor molecule-1 (TICAM-1), and mitochondrial antiviral signaling (MAVS)-dependent pathways (Figure 1, B). The depletion of exosomes may significantly reduce the expression of NKG2D ligand, so as to postpone disease progression.

**Hepatitis C**

Hepatitis C virus (HCV), a small, enveloped RNA virus, can progress to chronic hepatitis, cirrhosis, or even HCC under long-term exposure (30). It is reported that approximately 170 million people suffering from this virus (31). Therefore, it is urgent to explore new treatments for HCV. Cosset et al. (32) reported that exosomes derived from HCV-infected hepatocytes (HCV-exo) had the ability to transmit pathogenic substances to human hepatoma HuH7.5.1 cells through resisting neutralizing antibodies. However, in an experiment of HCV-replicating HuH7.5 cells treated by the MVB inhibitor, U18666A, the quantity of the viral genomes released to the supernatant showed a significant decrease (31). HCV-exo can also transfer viral nucleic acid from HCV-infected cells to adjacent immune cells, stimulate the expansion of myeloid-derived suppressor cells (MDSCs), subsequently promote the differentiation of T follicular regulatory cells (TFR), and inhibit the functions of T follicular helper cells (TFH), all of which constitute a newly discovered mechanism of immune dysfunction in the process of chronic viral infection (33). Meanwhile, it was reported that the promotion of TFR differentiation and repression of TFH function were associated with the inhibition of miRNA (miR)-124 expression in MDSCs stimulated by HCV RNA-containing exosomes (33). Reintroduction of miR-124 into peripheral blood mononuclear cells (PBMCs) may reduce the induction of MDSCs by viral nucleic acid-containing exosomes (33).

Activation and differentiation of HSCs into myofibroblasts compose a key mechanism for liver fibrosis. Exosomes released by the HBx-containing cells can stimulate proliferation signals in HSCs (23). miR-192 can be delivered to HSCs through exosomes secreted from HCV-replicating hepatocytes, and upregulate fibroblast markers by upregulating transforming growth factor β1 (TGF-β1) in HSCs (34). Devhare et al. also demonstrated that miR-19a in HCV-exo was internalized to activate HSC through regulating the suppressor of cytokine signaling (SOCS)-signal transducers and activators of transcription 3 (STAT3) axis in vitro (35). Thus, it can be seen that after infection with the hepatitis virus, exosomes and their cargo play a critical role in information communication between hepatocytes and nonparenchymal cells, determining the fate of the virus and the evolution of diseases. Researchers have speculated that exosomes can serve as a new target of viral hepatitis or further fibrosis intervention through the inhibition of their release and the modification of their cargo. Grünvogel et al. (36) demonstrated in vitro that exosomes could transfer the intermediate products of HCV replication, and thus blocking exosomal release might inhibit the replication of HCV through activating toll-like receptor 3 (TLR-3). Moreover, exosomes containing HBV-miR-3 derived from HBV-infected hepatocytes may restrain macrophages from expressing SOCS5 and facilitate M1 polarization by activating the Janus kinase (JAK)/STAT pathway. HBV-miR-3 is capable of enhancing the expression of epidermal growth factor receptor (EGFR) via suppressing SOCS5-mediated ubiquitination, and then stimulating IL-6 secretion to inhibit HBV replication (37). Taken together, the above findings point to the potential value of exosomes in interfering or reversing disease progression after viral infection. However, numerous animal experiments and preclinical studies are still needed before this potential can be confirmed and realized.

**Alcoholic liver disease (ALD)**

ALD, which manifests as a wide range of diseases, including alcoholic hepatitis (AH) and cirrhosis, accounts for about 5% of the global disease burden and 6% of total global deaths annually. AH is a syndrome characterized by inflammatory cell infiltration and hepatocyte damage which can stimulate HSCs to secrete excessive extracellular matrix (ECM), such as collagen, and facilitate liver cirrhosis (38) after long-term excessive drinking. With the development of exosome isolation and detection technology, scientists have started to pay attention to their roles in the pathological progression of ALD. A recent study showed that exosome quantity in the serum of healthy people increased after alcohol abuse or long-term drinking (39), with statistically significant correlation with alanine aminotransferase (ALT) levels (40). Ethanol treatment has been found to increase the release of hepatocyte exosomes, which were further preferentially localized in hepatocytes or HSCs, and stimulated the expression of the mRNA involved in exosome biogenesis.
through ceramide or endosomal sorting complexes required for transport (ESCRT) pathways (11). Hence, interference of RNA involved in ESCRT or ceramide pathways in ethanol-treated hepatocytes may reduce exosome production. Meanwhile, in vivo, it has been shown that the exosome inhibitor, GW4869, may suppress intercellular transport of RNA from hepatocytes to target cells (11).

MicroRNAs are the most studied small RNAs in exosomes from mice models and patients with AH. It was demonstrated that miR-30a, miR-30b, miR-122, miR-130a, miR-192, miR-744, and miR-1246 were upregulated in serum exosomes from chronic alcohol-fed mice (40). Babuta et al. (41) further discovered that the inhibition of autophagy and impaired autophagosome and lysosome function were correlated with increased exosome production through the alcohol-related increase of miR-155 in ALD mice models and human livers with ALD. Thus, it may be possible to reduce alcohol-induced liver damage by decreasing the production of exosomes via miR-155 regulation. Transferring exosome miR-122 may induce the sensitization of lipopolysaccharide (LPS) and inhibit the heme oxygenase 1 (HO-1) pathway which can suppress the cell injury induced by cytokines and reactive oxygen species. Hence, these effects can be prevented by the exosome-mediated transmission of miR-122 inhibitor (39). Primary HSCs exhibit profibrotic markers, pri-miR-17-92, and connective tissue growth factor (CCN2) (42), and reduce the expression of miR-19b (43) when exposed to alcohol. During HSC activation, the overexpression of exosome miR-19b may change the responsiveness and epigenetic regulatory factors of TGF-β, further suppressing collagen production (43) (Figure 1, C). Exosomal transfer of miR-214 into the recipient HSCs may also revert the HSC phenotype by directly inhibiting the transcription of CCN2 (42). Understanding these phenomena may be critical to the future development of exosomes as ALD therapeutics.

Other types of cargo in exosomes can also influence the pathological progress of ALD. It was reported that endoplasmic reticulum (ER) stress and oxidative stress may increase the amount of cytochrome P450-2E1 (CYP2E1) in EVs which may promote cell death by stimulating the apoptosis signaling pathway (44). Similarly, alcohol was found to significantly induce CYP2E1 levels of plasma exosomes in an overdrinking mouse model, and these CYP2E1-enriched exosomes worsened alcohol-induced hepatotoxicity and monocyte toxicity, which might be reduced by selective CYP2E1 enzyme activity inhibitors (45). In summary, exosomes have the capacity to regulate inflammatory and fibrotic pathways by delivering cargo to target cells in the occurrence and progression of ALD. The above experiments (44,45) provide clues for future clinical treatment, but determining the exact method by which exosomes and their cargo can be used to intervene in the pathogenesis of ALD requires more experimental research.

**Nonalcoholic fatty liver diseases (NAFLD)**

NAFLD is a complex disease, ranging from simple steatosis, nonalcoholic steatohepatitis (NASH), cirrhosis, and even to HCC. Its emergence is attributed to systemic inflammation, insulin resistance, and hepatocyte apoptosis. NAFLD is becoming one of the most common chronic liver disorders, with a worldwide prevalence of 25.2% (46) and an Asian prevalence of 29.6% (47). Furthermore, the fibrosis progression proportion and HCC incidence were found to be 40.76% and 0.44 per 1,000 person-years, respectively (46), while the risk ratios of liver-specific and overall mortality of NAFLD were reported to be 1.94 and 1.05, respectively (46).

Lipotoxicity plays a crucial role in the pathogenesis of NASH through macrophage-associated inflammatory responses, activation of proapoptotic signaling, and angiogenesis (48,49). It may induce lysosomal dysfunction in hepatocytes, then further increase exosome release, causing M1 polarization and macrophage-induced inflammation in an miR-122-5p-dependent (50) or miR-192-5p-dependent manner (51). Moreover, palmitic acid (PA) treated (52) or cholesterol-induced hepatocytes (50) were found to display a significant increase in exosome production, showing distinctive miRNA expression patterns. Thus, reducing lipid deposition and inhibiting the release of exosomes may ameliorate the liver inflammation caused by lipotoxicity. Meanwhile, hepatocytes stimulated by lipids were also demonstrated to release EVs containing TNF-related apoptosis-inducing ligand (TRAIL) (53) and C-X-C motif ligand 10 (CXCL10) (54), which induced macrophage chemotaxis and inflammation phenotype activation. These two effects might be blocked by CXCL10-neutralizing antisera, mixed lineage kinase 3 (MLK3) inhibitor (54), or rho-associated, coiled-coil-containing protein kinase 1 (ROCK1) inhibitor, fasudil (53).

NASH is characterized by neutrophil infiltration around lipotoxic hepatocytes, which is thought to result in the liver inflammation and injury (55). However, recent research (55) has indicated that inflammation and fibrosis could be ameliorated by the communication between neutrophils and hepatocytes through low-density lipoprotein receptor (LDLR)-dependent miR-223-enriched EV transfer.
Furthermore, Watanabe et al. (56) confirmed that small extracellular vesicles (sEVs) derived from adipose tissue-derived MSCs (AD-MSCs) could attenuate inflammation and inhibit liver fibrosis in a rapid NASH fibrosis model (Figure 1, C). Consequently, exosomes and their cargo are significant to the pathogenesis of NAFLD, but may also be attractive therapeutic tools for treating liver diseases. In the future, making full use of exosomes may provide scientists with novel strategies for the treatment of nonalcoholic liver disease.

### Hepatocellular carcinoma

HCC, the most common type of liver cancer, is a major global public health issue of concern. Chronic HBV and HCV infections are generally considered to be risk factors for HCC, and account for 56% and 20% of cases, respectively (57,58). Only a small portion of HCC patients who have underlying basic chronic liver diseases and cirrhosis may be rescued by resection or liver transplantation (59) due to insufficient donors, financial considerations, and other factors. The survival of advanced HCC still remains poor because of insensitivity or drug resistance to chemotherapy (60). Consequently, it is necessary to explore a new treatment for HCC. Exosomes are extremely useful for the horizontal delivery of multiple RNAs and protein molecules to adjacent or distant cells via paracrine and autocrine forms, and are implicated in the mechanisms of the occurrence and progression of HCC tumor, including angiogenesis, epithelial-mesenchymal transition (EMT), immune escape, and chemotherapy drug resistance (61,62). He et al. (61) conducted proteomic analysis and RNA sequencing of the cargo in HCC-derived and immortalized hepatocyte-derived exosomes. The results showed exosomes carried many protumorigenic proteins and RNAs, such as caveolins, RRAS, CLND3e, and S100A4. Exosome-enriched pathogenic genes derived from hepatoma cells could significantly enhance the migration and invasion of normal hepatocytes by triggering mitogen-activated protein kinase (MAPK) and phosphatidylinositol 3 kinase (AKT) signaling pathways, as well as increasing secretion of active matrix metalloproteinases-2 (MMP-2) and matrix metalloproteinases-9 (MMP-9) (61). Exosomes may exert hepatoma inhibitory effects by the overexpression of a key regulator of exosome biogenesis—vacuolar protein sorting 4 homolog A (Vps4A) to inactivate PI3K/AKT pathway and utilize exosomes to modulate the secretion and uptake of miRNAs (63). Hepatoma cell-secreted exosomal miR-210 (64) and miR-155 (65) may promote tumor angiogenesis of endothelial cells and elevate the proliferation of HCC cells. Conversely, repression or knockdown of these two miRNAs may have the opposite effect, inhibiting angiogenic activity in HCC (64,65). Furthermore, knockdown of phosphatase and tensin homolog deleted on chromosome 10 (PTEN) may attenuate the proliferation of HCC cells treated with the exosomal miR-155 (66). Earlier studies (7) showed that exosomes could transfer RNA between cells, acting as a good adjuster to diseases (Table 1).

### Table 1 The roles of exosomal small RNA in HCC therapy

| Small RNA | Function | Mechanism | Reference |
|-----------|----------|-----------|-----------|
| miR-335-5p | Inhibit proliferation and invasion, increase apoptosis | Shuttle between hepatoma cells and HSCs, downregulate mRNA targets for miR-335 | (67) |
| circ-0051443 | Promote apoptosis | Arrest the cell cycle in the G0/G1 phase | (68) |
| miR-490 | Suppress metastasis | Mast cells stimulated by HCV-E2 secrete exosomes to block the ERK1/2 pathway | (6) |
| miR-142, miR-223 | Hamper proliferation | Affect posttranscriptional regulation of proteins | (69) |
| miR-26a | Repress proliferation and migration | Target HepG2 cells through scavenger receptor class B 1-Apo-a1 complex | (14) |
| miR-320a | Repress proliferation and metastasis | Bind to PBX3 and inhibit the activation of the MAPK pathway | (70) |
| miR-24, miR-223, miR-31 | Inhibit proliferation and promote apoptosis | Downregulate target proteins involved in cell proliferation | (13) |

HCC, hepatocellular carcinoma; HSC, hepatic stellate cell; HCV-E2, hepatitis C virus E2 envelope glycoprotein; ERK1/2, extracellular signal-regulated kinase 1/2; PBX3, pre-B-cell leukemia homeobox 3; MAPK, mitogen-activated protein kinase.
Wang et al. (67) demonstrated in vivo and in vitro that miR-335-5p could be delivered by exosomes to hepatoma cells, so as to inhibit their proliferation and invasion and increase apoptosis (Figure 1, D). Meanwhile, Chen et al. (68) showed that when exosomal circular RNA (circRNA)-0051443 was transmitted to hepatoma cells, the cell cycle ceased.

Moreover, immunocyte-derived exosomes can exert a great influence on antitumor immune responses (6). For example, mast cells stimulated by HCV-E2 envelope glycoprotein (HCV-E2) are capable of secreting numerous exosomes rich in miR-490, which can be transferred into hepatoma cells and suppress their metastasis through blocking the extracellular signal-regulated kinase 1/2 (ERK1/2) pathway (6). MiR-142 and miR-223 expressed in macrophages can be transferred by exosomes to affect posttranscriptional regulation of proteins in HCC cells which hampers the proliferation of these cancer cells (69). Dendritic cells (DCs) play a key role in both innate and adaptive immune responses, and their maturation and activation can be disturbed by the HCC-induced tumor microenvironment (71). Tumor-derived exosomes can activate DCs, stimulate the proliferation of immature T cells, and induce T cells to differentiate into antigen-specific cytotoxic T lymphocytes (CTLs), thus increasing antineoplastic efficacy (72,73) (Figure 1, D). Rao et al. reported that the tumor immune microenvironment in HCC mice was significantly improved upon application of HCC-derived exosome-pulsed DCs. The number of T cells and the level of interferon gamma (IFN-γ) increased, while IL-10 and TGF-β decreased, which eventually resulted in tumor growth inhibition and a strong immune response (74). They also found that the exosomes released by alpha-fetoprotein (AFP)-expressing DCs had antitumor properties similar to those of HCC-derived exosome-pulsed DCs, thus providing a cell-free vehicle for tumor immunotherapy (75).

Accumulating evidence has shown that cancer cells can release exosomes to promote carcinogenesis and the resistance or insensitivity to multiple chemotherapeutic drugs, and thus a greater research focus has been placed on exosome-based drug delivery to inhibit cancer development through interference techniques (76). For instance, some studies have found that the exposure of HCC cells to sorafenib increased long intergenic noncoding RNA-VLDLR (linc-VLDLR) and long noncoding RNA ROR (lncRNA ROR) expression in cells and EVs. RNA interference-mediated knockdown of these two lncRNAs increased chemotherapy-induced cytotoxicity and apoptosis, leading to enhanced chemosensitivity on HCC (77,78). Some preclinical findings have suggested that exosome-mediated drug delivery has good prospects in cancer treatment. For example, engineered exosomes packed with miR-26a were shown to selectively target HepG2 cells through scavenger receptor class B 1-Apo-a1 complex, so as to repress cell proliferation and migration (14). Furthermore, when miR-320a-enriched exosomes were injected into rats via the caudal vein, it was observed that the proliferation and metastasis of hepatoma cells were effectively inhibited via binding to the downstream target pre-B-cell leukemia homeobox 3 (PBX3) (70), further inhibiting the activation of the MAPK pathway (70). MiR-24, miR-223, and miR-31 delivered by exosomes can also restrain the growth and invasion of HCC and increase apoptosis, exerting potential antitumor activity in vivo (13).

These preclinical experiments have clarified the functions of exosomes and their cargo in the inhibition of HCC progression. Due to the advantages of low immunogenicity and toxicity, and the natural release or uptake by tumor cells (76,79), exosomes are expected to become an attractive therapeutic strategy for the treatment of HCC in the future.

**MSC-derived exosomes**

MSCs are multipotent stromal cells possessing various biological functions, such as self-renewal, multilineage differentiation, and anti-inflammation (80,81). Recently, use of MSC-derived exosomes functioning as potent therapeutic vehicles has become a promising strategy for various diseases (80,81) (Table 2). Jiang et al. found that human umbilical cord MSC (UC-MSC)-derived exosomes might alleviate acute liver injury and fibrosis induced by carbon tetrachloride (CCl4) in mouse models via antioxidant potentials (82). MiR-455-3p-containing exosomes released by UC-MSCs stimulated with IL-6 could suppress macrophage activation and reduce cytokine production by targeting PI3K signaling in a chemical liver injury animal model, and consequently ameliorate liver histology and retrieve function (83) (Figure 1, E). Furthermore, Rong et al. applied other exosomes derived from human bone marrow MSCs (BM-MSCs) in the treatment of liver fibrosis induced by CCl4. The results showed fibrosis amelioration and HSC activation was inhibited via Wnt/β-catenin pathway (85) (Figure 1, E). BM-MSC-derived exosomes may also attenuate the hepatic inflammatory response and reduce the release of inflammatory cytokines from macrophages in autoimmune hepatitis, which may be associated with the expression levels of miR-223-3p and STAT3 in macrophages (89).
Acute liver failure (ALF) is a fatal illness with high mortality, which can only be controlled by liver transplantation and artificial liver therapy (90). However, various constraints including ischemia/reperfusion (I/R) injury have prompted researchers to explore alternative treatments. MSC exosomes have shown the potential to prolong or save lives. Studies have demonstrated that TNF-alpha pretreatment of UC-MSC-derived exosomes (T-Exo) and adipose tissue-derived MSC exosomes (AMSC-exo) decreased serum AL T, aspartate transaminase (AST), and proinflammatory cytokine level, inhibited activation of nucleotide-binding and oligomerization domain-like receptor 3 (NLRP3) in macrophages, and reduced pathological liver damage caused by ALF (84,91).

Furthermore, anti-inflammatory-related miR-299-3p packaged into exosomes is upregulated by TNF-alpha-stimulated MSCs, which exerts a therapeutic effect (84); meanwhile, miR-17 is elevated in AMSC-exo through suppressing thioredoxin-interacting protein/nucleotide-binding and oligomerization domain-like receptor 3 (TXNIP/NLRP3) signaling pathway (91). In addition to having anti-inflammation effects, BM-MSC exosomes applied in ALF may also significantly decrease the levels of cleaved caspase-3 and Bax, upregulate the expression of Bcl-2, which can attenuate hepatocyte apoptosis and promote autophagy (86).

I/R injury, including inflammation, necrosis, and apoptosis, is the main problem in liver transplantation (92). MiR-20a secreted by MSC exosomes can bind to the 3’ untranslated region (3’ UTR) of Fas and Beclin-1 to regulate the gene expression involving apoptosis and autophagy (93). Furthermore, BM-MSC-differentiated hepatocyte-like cell exosomes (MSC-Heps-exo) may alleviate hepatic I/R injury effectively and reduce hepatocyte apoptosis and the levels of liver enzyme in vivo and in vitro (92). Lai et al. (87) found BM-MSC-exo could also inhibit Th17 cells and induce regulatory T cells (Tregs) to reduce injury and ameliorate the survival of chronic graft-versus-host disease (cGVHD). In addition, studies (94) have demonstrated that fetal liver MSC-derived exosomes suppress proliferation, activation, and cytotoxicity of NK cells via TGF-β/Smad2/3 signaling in allogeneic reactions, and that applying anti-TGF-β antibody may restore NK cell function. Therefore, using MSC-derived exosomes to ameliorate I/R injury shows considerable promise.

Currently, the application of MSCs exosomes in liver cancer has become a key goal of clinical research, and a large number of related studies have been conducted. Jiang et al. (82) administered UC-MSC-derived exosomes into CCl₄-induced liver tumor in vivo. They found these exosomes could provide more antioxidant effects than bifendate treatment and exerted hepatoprotective effects, subsequently restraining the growth of tumors (82). Exosomes from adipose-derived mesenchymal stem cells (AD-MSCs) could promote NK cells to exert antitumor roles on rat HCC, thereby facilitating low-grade tumor differentiation and inhibiting tumor growth (95) (Figure 1, E). Accumulating evidence has indicated that miR-122 has the property of promoting the chemosensitivity of HCC cells. Hence, Lou et al. chose AD-MSC exosomes as biological

| Origin    | Disease                                      | Function                                                                 | Reference |
|-----------|----------------------------------------------|--------------------------------------------------------------------------|-----------|
| UC-MSCs   | Acute liver injury and fibrosis              | Antioxidant potentials                                                   | (82)      |
| UC-MSCs   | Chemical liver injury                        | Suppress macrophage activation and reduce cytokine production           | (83)      |
| UC-MSCs   | ALF                                           | Inhibit NLRP3 activation in macrophage and decrease proinflammatory cytokines level | (84)      |
| BM-MSCs   | Liver fibrosis induced by CCl₄               | Inhibit HSC activation via Wnt/β-Catenin pathway                        | (85)      |
| BM-MSCs   | ALF                                           | Decrease the levels of cleaved caspase 3 and Bax, upregulate the expression of Bcl-2 | (86)      |
| BM-MSCs   | I/R injury                                    | Inhibit Th17 cells and induce Treg cells                               | (87)      |
| AD-MSCs   | HCC                                           | Improve the sensitivity of chemotherapeutic drugs                       | (88)      |

MSC, mesenchymal stem cell; UC-MSC, umbilical cord mesenchymal stem cell; BM-MSC, bone marrow mesenchymal stem cell; AD-MSC, adipose-derived mesenchymal stem cell; ALF, acute liver failure; NLRP3, nucleotide-binding and oligomerization domain-like receptor 3; CCl₄, carbon tetrachloride; I/R injury, ischemia/reperfusion injury; HCC, hepatocellular carcinoma; HSC, hepatic stellate cell.
vehicles for miR-122 delivery. The results showed that miR-122-transfected AMSCs successfully mediated miR-122 transmission between AMSCs and HCC cells, thereby inducing tumor cells to be sensitive to chemotherapeutic drugs (88). Meanwhile, injecting MSC-derived exosomes containing miR-122 into tumor could effectively enhance the antitumor efficacy of sorafenib (88) (Figure 1, E). Collectively, MSC-exos are being increasingly considered as attractive candidates to control disease progression and improve liver function.

Conclusions

With more evidence being generated from preclinical and animal experiments, researchers have begun to draw the blueprint for the clinical application of exosomes and their cargo in the treatment of liver diseases. Although the above results are encouraging, there are still many problems that remain to be solved. The establishment of standard methods to isolate and identify the types and sources of valuable exosomes is the first step. The quality and sufficient quantity of these exosomes need to be ensured in order to optimize the therapeutic functions of exosomes during treatment. Secondly, the mechanisms by which exosomes are released by cells and recognized and fused by target cells need to be fully understood. Thirdly, comprehensive examination of the genes and protein functions carried by exosomes are needed, as some molecules demonstrate therapeutic effects, while others do not, or may even be deleterious. Once the properties of these molecules are grasped, valuable traits may be exploited, while others can be discarded. Research on exosomes have opened a door to discovering the mechanisms underlying the occurrence and development of various liver diseases, and providing a new drug vehicle with low immunogenicity and high biocompatibility. There is still a long way to go before clinical application can be actualized, and more experimental evidence is needed to support future clinical research.

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