RESEARCH ARTICLE

Association of GSTP1 Ile105Val polymorphism with the risk of coronary heart disease: An updated meta-analysis

Yadong Song1,2*, Xiaoli Liu1,2*, Cheng Luo3,4, Liangkai Chen3,4, Lin Gong1,2, Hanbin Yu1,2, Bin Wang1,2, Ernan Liu1,2, Huiqiong Xu1,2, Jiansheng Liang1,2

1 Department of Disinfection and Pest Control, Wuhan Centers for Disease Prevention and Control, Wuhan, Hubei, China, 2 Wuhan Healthcare-associated Infection Management Quality Control Center, Wuhan, Hubei, China, 3 Department of Nutrition and Food Hygiene, Hubei Key Laboratory of Food Nutrition and Safety, Tongji Medical College, Hua Zhong University of Science and Technology, Wuhan, People’s Republic of China, 4 Ministry of Education Key Lab of Environment and Health, School of Public Health, Tongji Medical College, Wuhan, People’s Republic of China

These authors contributed equally to this work.
* song465979919@163.com (YS); liuxiaoli20851@126.com (XL)

Abstract

Background
Numerous case-control studies have investigated the association between GSTP1 Ile105-Val polymorphism and CHD risk, but the results from published studies were inconclusive. The present meta-analysis was performed to derive a more precise estimation.

Methods
PubMed, EMBASE, and Web of Science database searches were conducted to retrieve relevant articles.

Results
Ultimately, 5,451 CHD cases and 5,561 controls from 15 studies were included. Pooled analysis did not yield any statistically significant association between GSTP1 Ile105Val polymorphism and CHD risk for the overall population (Val vs. Ile: OR, 1.05; 95% CI, 0.93 to 1.18; Val/Val vs. Ile/Ile: OR, 1.09; 95% CI, 0.83 to 1.42; Val/Ile vs. Ile/Ile: OR, 1.09; 95% CI, 0.93 to 1.28; Val/Val vs. Val/Ile+Ile/Ile: OR, 1.04; 95% CI, 0.83 to 1.30; Val/Val+Val/Ile vs. Ile/Ile: OR, 1.14; 95% CI, 0.97 to 1.33). Subgroup analyses and sensitivity analyses indicated that GSTP1 Ile105Val polymorphism was still not associated with an increased risk of CHD. After excluding studies detected by Galbraith plots as major sources of heterogeneity, these relationships were still not significant.

Conclusions
The overall results did not reveal a major role of the GSTP1 Ile105Val polymorphism in modulating CHD risk. Well-designed studies with large sample sizes are needed to validate our findings and explore the possible gene-gene or gene-environment interactions.
Introduction

Atherosclerosis is a major cause of coronary heart disease (CHD), a major public health problem, and a leading cause of morbidity and mortality in the world [1,2]. It is well known that the complex interaction of environmental and predisposing genetic risk factors plays a crucial role in the underlying pathophysiology of CHD. Multiple traditional risk factors lead to CHD development, including age, a high-fat diet, smoking, alcohol, diabetes mellitus, hyperlipidemia, hypertension, and so on.

Oxidative stress, an imbalance between antioxidant defenses and free radical generation, was implicated as potential pathophysiological mechanisms behind the pathogenesis and progression of CHD [3]. DNA damage, inflammation, smooth muscle cell proliferation, and lipid peroxidation, which are caused by increased production of reactive oxygen species (ROS), can result in atherosclerosis and, hence, CHD [4]. Furthermore, DNA adducts have been detected in patients with severe CHD [5] and atherosclerotic plaques [6]. DNA adducts were considered to be related to atherogenic risk factors including old age, alcohol drinking status, smoking status, oxidative DNA damage, triglycerides, cholesterol, and arterial pressure. Besides, DNA adducts have been thought to be risk factors for reducing the capacity of antioxidants [7].

Glutathione S-transferases (GSTs) are a superfamily of phase II detoxification enzymes that convert various reactive metabolites (endogenous and exogenous products of oxidative stress) to more water-soluble and less harmful forms by conjugating them with glutathione [8]. Therefore, GSTs play vital roles in protecting the cell against oxidative stress. Besides, GSTs can protect DNA from genotoxic damage by inhibiting the formation of DNA adducts [9]. Human GSTs are composed of eight major groups including GSTM (mu), GSTT (theta), GSTP (pi), GSTA (alpha), GSTK (kappa), GSTO (omega), GSTS (sigma) and GSTZ (zeta) [10]. The glutathione S-transferase P1 (GSTP1) gene is 2.8 KB long, which is located on the long arm of chromosome 11 (11q13.3). The GSTP1 Ile105Val polymorphism is a single nucleotide polymorphism (SNP) located in exon 5 which encodes an isoleucine (Ile) to valine (Val) exchange at codon 105. Individuals with GSTP1 protein containing Val but not Ile have significantly reduced enzyme activity and affinity for electrophilic substrates [11]. Thus, decreased detoxification ability of lower enzymatic activity may increase the risk of CHD.

A number of case-control studies [4,7,8,12–23] have investigated the relationship between GSTP1 Ile105Val polymorphism with the risk of CHD, with equivocal conclusions. Recently, one meta-analysis [24] revealed that a significant association exists between GSTP1 null genotype and CHD, and the other one [25] suggested that GSTP1 null genotype could impact individual susceptibility to atherosclerotic cardiovascular diseases. However, numerous relevant publications [4,19,22,23] did not appear in their meta-analyses. Moreover, their meta-analyses did not evaluate the strength of the association between GSTP1 Ile105Val polymorphism and CHD risk in the specific genetic models, dominant model, homozygote model, heterozygote model, recessive model, and an allele comparison, respectively. Therefore, we performed an updated meta-analysis to establish a comprehensive picture of the Ile105Val polymorphism of the GSTP1 and the risk of CHD.

Materials and methods

Identification and eligibility of relevant studies

Basing on the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA), we performed the present meta-analysis. PubMed, EMBASE, and Web of Science database searches were performed before 1 April 2021 using the following terms:
glutathione S-transferase, glutathione S-transferase pi, GST, GSTP1; genetic, polymorphism, variant; and myocardial infarction, MI, coronary heart disease, CHD, coronary artery disease, CAD, ischemic heart disease. The references of eligible articles and relevant reviews were also screened for additional reports. All identified publications should fulfill the following criteria: (a) designed as case-control studies, (b) assessed the association of GSTP1 Ile105Val polymorphism with CHD risk published in English, (c) provided the adequate genotypic information to calculate odds ratios (ORs) and 95% confidence intervals (95% CIs). Editorials, review articles, animal studies, case reports, preliminary results not on GSTP1 gene polymorphism or CHD, and studies without data for estimating OR with 95% CI were excluded.

Data extraction and quality assessment
For each included study, the following data were extracted and entered into the standard form: first author, year of publication, country, ethnicity, source of controls, number of cases, number of controls, G allele (%) in case, G allele (%) in control, Hardy-Weinberg equilibrium (the genotype distribution in controls were consistent with Hardy-Weinberg equilibrium (HWE)), Newcastle-Ottawa Scale (NOS) score, and Adjustment covariance. Because two publications [12,22] only provided the genotype data as “Val/Val+Val/Ile” and “Ile/Ile” without details, we could only estimate the OR for the dominant genetic model. We conducted the quality assessment of eligible publications based on NOS [2,26–28]. Two authors (C.L and L.C) independently performed the database searches, data extraction, and quality assessment of included studies, and any disagreements were discussed and resolved with a third investigator (X.L.L).

Statistical analysis
The pooled ORs and 95% CIs were used to estimate the strength of the association between GSTP1 Ile105Val polymorphism and susceptibility to CHD. We investigated the association using five genetic models, including dominant model (Val/Val+Val/Ile vs. Ile/Ile), homozygote model (Val/Val vs. Ile/Ile), heterozygote model (Val/Ile vs. Ile/Ile), recessive model (Val/Val vs. Val/Ile + Ile/Ile), and allele model (Val vs. Ile). Both the Cochran Q test and $I^2$ test were performed to evaluate the between-study heterogeneity among included studies [29]. Significant heterogeneity was detected when $P<0.10$ for the Q test or $I^2 \geq 50\%$ for $I^2$ statistic, and a random effect model using DerSimonian-Laird method was applied. Otherwise, a fixed effect model using Mantel-Haenszel method was performed if heterogeneity was negligible [10]. The detection of outliers by Galbraith plots was considered as the main source of between-study heterogeneity [30]. Publication bias was assessed using Begg’s funnel plot and Egger’s test, and $P<0.05$ indicated a potential publication bias [31–33]. For the control group in each selected study, the Chi-square goodness-of-fit test was used to evaluate HWE and $P<0.05$ was considered significant. The meta-regression was performed with the ‘metareg’ STATA command to explore the source of between-study heterogeneity. Prespecified sources of heterogeneity included publication year, ethnicity, and control source. Sensitivity analysis, excluding one study at a time, was conducted to evaluate the stability of the results. Moreover, sensitivity analysis by excluding studies without confirmation of HWE was also performed. Stratified analyses were also performed by ethnicity (East Asian, Caucasian); source of controls (population-based, hospital-based). All analyses were performed with STATA (version11.0; Stata Corporation, College Station, TX). All tests presented are 2-tailed with a significance level of 0.05.
Results

Identifying studies and study characteristics

The selection process of articles was presented in S1 Fig, with specification of reasons. 414 publications were found with the search criterion, and fifteen publications finally met the criteria for entering the present analysis. Because two articles [12,22] only reported data on genotypes as “Val/Val + Val/Ile” and “Ile/Ile”, the HWE test could not be conducted in these two studies and we could only estimate the OR for the dominant genetic model. The genotype distribution in controls was not in agreement with the HWE in one study [21]. The main study characteristics are summarized in Table 1 (Table 1). Finally, a total of 15 case-control studies with 5,451 CHD cases and 5,561 controls were included. There were eight studies on subjects of Caucasian and seven studies on subjects of Asia. The controls were divided into population-based population and hospital-based patients. The number of cases ranged from 54 to 2042, and the

| First author | Year | Country | Ethnicity | Control source | Genotyping method | Cases | Controls | G allele (%) | HWE | NOS Score | Adjustment covariates |
|--------------|------|---------|-----------|----------------|------------------|-------|----------|-------------|-----|-----------|----------------------|
| Wilson [4]   | 2000 | UK      | Caucasian | PB             | PCR-RFLP         | 351   | 190      | 34.8        | 36.8| 0.384     |                      |
| Wang [12]    | 2007 | Taiwan  | East Asian| PB             | PCR-RFLP         | 279   | 325      | NA          | NA  | NA        | Age and gender        |
| Cornelis [8] | 2007 | Canada  | Caucasian | PB             | PCR-RFLP         | 2042  | 2042     | 39.3        | 39.4| 0.646     |                      |
| Ramprasath [13] | 2011 | India   | East Asian| PB             | PCR-RFLP         | 290   | 270      | 39.3        | 32.2| 0.093     |                      |
| Singh [14]   | 2011 | India   | East Asian| PB             | PCR-RFLP         | 230   | 300      | 20.9        | 22.8| 0.08      | Age, sex, BMI, smoking, food habit, lipid profile and fasting glucose |
| Nomani [7]   | 2011 | Iran    | Caucasian | HB             | PCR-RFLP         | 209   | 108      | 30.9        | 31.9| 0.371     |                      |
| Kariz [15]   | 2012 | Slovenia | Caucasian | HB             | PCR              | 206   | 257      | 37.9        | 32.9| 0.731     |                      |
| Phulukdaree [16] | 2012 | South Africa | East Asian | PB             | PCR-RFLP         | 102   | 100      | 20.1        | 29.5| 0.413     |                      |
| Yeh [17]     | 2013 | Taiwan  | East Asian | HB             | PCR-RFLP         | 458   | 209      | 15.9        | 16  | 0.176     |                      |
| Kovacs [18]  | 2014 | Hungary | Caucasian | HB             | PCR              | 53    | 78       | 34.9        | 32.7| 0.392     |                      |
| Ding [19]    | 2016 | USA     | Caucasian | HB             | PCR              | 119   | 382      | 27.7        | 32.5| 0.268     | Age, BMI, smoking status (ever/never) and total cholesterol to HDL cholesterol ratio |
| Bhat [20]    | 2017 | India   | East Asian | PB             | PCR-RFLP         | 200   | 200      | 39          | 28.8| 0.056     | Age, gender, BMI, alcohol intake, total cholesterol, hypertension and family history |
| Bhatti [21]  | 2018 | India   | East Asian | PB             | PCR-RFLP         | 560   | 545      | 38.2        | 32.1| 0.0095    |                      |
| Simeunovic [22] | 2019 | Serbia  | Caucasian | PB             | PCR-RFLP         | 107   | 274      | NA          | NA  | NA        | Gender, age, smoking, hypertension, and diabetes |
| Pourkeramati [23] | 2020 | Iran    | Caucasian | PB             | PCR-RFLP         | 244   | 281      | 26.6        | 27.76| 0.277     | Age and sex          |

NA = not available; PB = population based; HB = hospital based; HWE = Hardy-Weinberg Equilibrium; NOS = Newcastle–Ottawa Scale.

https://doi.org/10.1371/journal.pone.0254738.t001
The number of controls ranged from 78 to 2042. The mean distribution frequency of the GSTP1 G allele was 31.28% in cases and the average frequency was 30.41% in controls.

### Quantitative synthesis

We performed a meta-analysis of the GSTP1 Ile105Val polymorphism under specific genetic models (Table 2). We found that GSTP1 Ile105Val polymorphism was not associated with CHD risk for overall populations (Val vs. Ile: OR, 1.05; 95% CI, 0.93 to 1.18, Fig 1; Val/Val vs. Ile/Ile: OR, 1.09; 95% CI, 0.93 to 1.28, Fig 1; Val/Val vs. Val/Ile+Ile/Ile: OR, 1.04; 95% CI, 0.83 to 1.30, Fig 5; Val/Val+Val/Ile vs. Ile/Ile: OR, 1.14; 95% CI, 0.97 to 1.33, Fig 5). By stratifying the analysis by ethnicity, the present meta-analysis revealed that GSTP1 Ile105Val polymorphism was not associated with CHD risk in Caucasian and East Asian (Table 2). In subgroup analysis according to the control source, GSTP1 Ile105Val polymorphism was also not associated with CHD risk among hospital-based controls.

### Table 2. Summary ORs and 95% CIs of the association between GSTP1 Ile105Val polymorphism and CHD risk.

| Contrast model | Studies, n | Odds ratio (95% CI) | Heterogeneity | Model |
|----------------|------------|---------------------|---------------|-------|
| Val vs Ile     | 13         | 1.05 (0.93,1.18)    | 63.3%         | 0.001 Random |
| Val/Val vs Ile/Ile | 13      | 1.09 (0.83,1.32)    | 57.3%         | 0.005 Random |
| Val/Ile vs Ile/Ile | 13      | 1.09 (0.93,1.28)    | 61.7%         | 0.002 Random |
| Val/Val vs Val/Ile+Ile/Ile | 13 | 1.04 (0.83,1.30)    | 43.0%         | 0.05 Random |
| Val/Val+Val/Ile vs Ile/Ile | 15 | 1.14 (0.97,1.33)    | 67.7%         | <0.001 Random |
| Val vs Ile     | 7          | 0.99 (0.92,1.06)    | 0.0%          | 0.499 Fixed |
| Val/Val vs Ile/Ile | 7       | 0.96 (0.82,1.12)    | 0.0%          | 0.551 Fixed |
| Val/Ile vs Ile/Ile | 7       | 1.01 (0.91,1.13)    | 39.7%         | 0.127 Fixed |
| Val/Val vs Val/Ile+Ile/Ile | 7 | 0.96 (0.83,1.11)    | 0.0%          | 0.431 Fixed |
| Val/Val+Val/Ile vs Ile/Ile | 8 | 1.01 (0.92,1.12)    | 22.6%         | 0.25 Fixed |
| Caucasian      |             |                     |               |       |
| Val vs Ile     | 6          | 1.11 (0.89,1.39)    | 74.2%         | 0.002 Random |
| Val/Val vs Ile/Ile | 6       | 1.27 (0.74,2.19)    | 68.8%         | 0.007 Random |
| Val/Ile vs Ile/Ile | 6       | 1.18 (0.89,1.55)    | 68.9%         | 0.007 Random |
| Val/Val vs Val/Ile+Ile/Ile | 6 | 1.17 (0.75,1.82)    | 56.50%        | 0.042 Random |
| Val/Val+Val/Ile vs Ile/Ile | 7 | 1.26 (0.96,1.65)    | 74.80%        | 0.001 Random |
| East Asian     |             |                     |               |       |
| Val vs Ile     | 6          | 1.16 (0.81,1.66)    | 68.8%         | 0.002 Random |
| Val/Val vs Ile/Ile | 6       | 1.09 (0.88,1.34)    | 71.8%         | 0.001 Random |
| Val/Ile vs Ile/Ile | 6       | 1.11 (0.85,1.47)    | 52.0%         | 0.042 Random |
| Val/Val vs Val/Ile+Ile/Ile | 8 | 1.16 (0.95,1.43)    | 76.1%         | <0.001 Random |
| Val/Val+Val/Ile vs Ile/Ile | 10 | 1.16 (0.95,1.43)    | 76.1%         | <0.001 Random |
| PB             |             |                     |               |       |
| Val vs Ile     | 8          | 1.07 (0.91,1.25)    | 74.8%         | <0.001 Random |
| Val/Val vs Ile/Ile | 8       | 1.16 (0.81,1.66)    | 68.8%         | 0.002 Random |
| Val/Ile vs Ile/Ile | 8       | 1.09 (0.88,1.34)    | 71.8%         | 0.001 Random |
| Val/Val vs Val/Ile+Ile/Ile | 8 | 1.11 (0.85,1.47)    | 52.0%         | 0.042 Random |
| Val/Val+Val/Ile vs Ile/Ile | 10 | 1.16 (0.95,1.43)    | 76.1%         | <0.001 Random |
| HB             |             |                     |               |       |
| Val vs Ile     | 5          | 1.02 (0.88,1.18)    | 15.2%         | 0.318 Fixed |
| Val/Val vs Ile/Ile | 5       | 0.92 (0.65,1.32)    | 22.0%         | 0.274 Fixed |
| Val/Ile vs Ile/Ile | 5       | 1.12 (0.92,1.37)    | 37.8%         | 0.169 Fixed |
| Val/Val vs Val/Ile+Ile/Ile | 5 | 0.85 (0.61,1.20)    | 26.8%         | 0.243 Fixed |
| Val/Val+Val/Ile vs Ile/Ile | 5 | 1.08 (0.90,1.31)    | 29.4%         | 0.226 Fixed |

PB = population based; HB = hospital based; 95% CI = 95% Confidence Interval; \( P_H \) = \( P \) value based on Q test for between-study heterogeneity.

https://doi.org/10.1371/journal.pone.0254738.t002
controls and healthy controls (Table 2). Covariates were introduced including the year of publication, ethnicity, and control source for meta-regression analysis. The meta-regression was conducted with the introduction of covariates including the publication year, ethnicity, and control source. However, in any comparison, no covariate was detected as a potential source of between-study heterogeneity.

**Heterogeneity and sensitivity analysis**

Heterogeneity was detected in the overall pooled analysis (Table 2). After excluding studies detected by Galbraith plot as major sources of heterogeneity, no evidence for heterogeneity was detected and the fixed effects summary estimate also indicated that GSTP1 Ile105Val polymorphism was not associated with an increased risk of CHD (Val vs. Ile: OR, 0.98; 95% CI, 0.91 to 1.05; \( I^2 = 27.7\% \), \( P_{H} = 0.256 \), excluding 4 studies [13,16,20,21]; Val/Val vs. Ile/Ile: OR,
Sensitivity analyses were performed by omitting each study at a time to explore the effect of individual study, and the pooled ORs were not noticeably changed, suggesting that the results of present analysis were stable. In the sensitivity analysis, the influence of each study on the pooled OR was examined by repeating the meta-analysis while omitting each study, one at a time. This procedure confirmed the stability of the overall result. After omitting one study [21] departing from HWE and two studies [12,22] lacking the necessary information, the results based on crude ORs remained unchanged.

![Fig 2. Forest plots for the GSTP1 Ile105Val polymorphism and the risk of CHD in overall studies (Val/Val vs. Ile/Ile).](https://doi.org/10.1371/journal.pone.0245738.g002)
Publication bias

Begg’s and Egger’s tests were performed to evaluate publication bias in the overall pooled analysis. Begg’s and Egger’s test indicated no significant evidence of publication bias (Val vs. Ile: Begg’s test, $P = 0.428$; Egger’s test, $P = 0.965$; Val/Val vs. Ile/Ile: Begg’s test, $P = 0.300$; Egger’s test, $P = 0.967$; Val/Ile vs. Ile/Ile: Begg’s test, $P = 0.583$; Egger’s test, $P = 0.934$; Val/Val vs. Val/Ile+Ile/Ile: Begg’s test, $P = 0.360$; Egger’s test, $P = 0.882$; Val/Val+Val/Ile vs. Ile/Ile: Begg’s test, $P = 0.767$; Egger’s test, $P = 0.766$).

Discussion

In the present meta-analysis including 15 studies with a total of 5,451 CHD cases and 5,561 controls, the association between GSTP1 Ile105Val polymorphism and CHD risk was comprehensively assessed, and no positive results were obtained by the overall analysis.
Recently, one previous meta-analysis, conducted by Su et al. [24], included 4,595 cases and 4,390 controls from 11 studies. They proved that GSTP1 null polymorphism was associated with the risk of CHD in the overall population (OR, 1.23; 95% CI, 1.02 to 1.48). Another meta-analysis performed by Li et al. [25] revealed that GSTP1 null genotype could impact individual susceptibility to atherosclerotic cardiovascular diseases. However, our meta-analysis indicated that the GSTP1 Ile105Val polymorphism was not associated with CHD risk in five genetic models. The present study included 5,451 CHD cases and 5,561 controls in 15 studies, which could provide more sufficient statistical power. Compared with previous studies, more than four relevant studies [4,19,22,23] were involved in present meta-analysis but not in theirs. The sample size of two previous studies was relatively small and data was not sufficient for subgroup analysis. The results were not changed after adjustment for heterogeneity by excluding studies spotted by Galbraith plot. When stratifying the analysis by ethnicity, the results of our
study revealed that $\text{GSTP1}$ Ile105Val polymorphism was also not associated with CHD risk in East Asian and Caucasian.

The evaluation of heterogeneity is critical to the interpretation of the results for most meta-analyses [34]. Even modest heterogeneity exists across eligible studies may make meta-analysis miss the true effect. In the present meta-analysis, obvious between-study heterogeneity existed in five genetic models for overall populations. The heterogeneity persisted when stratified analyses were conducted by ethnicity and control source. Furthermore, a Galbraith plot was conducted to explore the source of heterogeneity. After excluding studies with low-quality design, no obvious between-study heterogeneity was observed among the remaining studies. In addition, sensitivity analysis was performed to confirm the robustness of our findings. The between-study heterogeneity may be caused by the inclusion population or study design. For example, Nomani et al. [7], Ramprasath et al. [13] and Cornelis et al. [8] contained a high risk
control group with hypertension, diabetes mellitus, or family history of CHD. The results may be biased when the controls cannot reflect exposure distribution or the genotype of the source population.

It was reported that GSTP1 Ile105Val polymorphism was associated with the altered catalytic and non-catalytic activity of GSTs. The GSTP1*G allele coding for the protein in which amino acid isoleucine (Ile) is substituted with valine (Val) has been shown to reduce enzyme activity and affinity for electrophilic substrates, which may lead to individual susceptibility to CHD. Of the fifteen studies, eight reported no significant association between GSTP1 Ile105-Val polymorphism and the risk of CHD. Among the rest of the studies, Wang et al. [12] observed a 1.8-fold increased CHD risk among subjects with the combination of Val/Val and Val/Ile genotypes of GSTP1 when compared to Ile/Ile genotype. Ramprashat et al. [13] found that GSTP1 Ile105Val polymorphism was associated with a higher risk of CHD. Singh et al. [14] reported that the interactive effect of GSTP1 Val/Val with MI remained significant after adjusting for risk factors. Kariz et al. [15] proved that univariate analysis indicated an association between the GSTP1 Ile105Val polymorphism and MI. Phulukdaree et al. [16] found that a significant association with CHD was observed in GSTP1 A105/A105. Bhat et al. [20] reported a statistically significant association between GSTP1 g.313A>G (A/G, G/G) genotype and CHD was detected. The study by Bhatti et al. [21] indicated the GG genotype of the GSTP1 (313A/G) gene was associated with an approximately two-fold enhanced risk of developing CHD. However, when pooling all studies together, we found no evidence for an association between GSTP1 Ile105Val polymorphism and CHD. The etiological mechanism of CHD is very complicated, in which gene-gene and gene-environment interactions may play important roles. The findings of the study by Phulukdaree et al. [16] supported the association of genotypes GSTM1 0/0 and GSTP1 A105/A105 and smoking with CHD. The study by Simeunovic et al. [22] observed a stronger association in heart failure patients due to CHD, who were carriers of a combined GSTP1(rs1695)/GSTA1“risk-associated” genotype. Singh et al. [14] reported that a significant protective effect of GSTP1 Val genotype against the disease was detected in nonsmokers with AMI. Hence, future studies concerning gene-gene, gene-nutrition, and gene-environment interactions under a systems network biology framework are required [35–39]. N4-acetylcytidine (ac4C) is often known as a conservative, chemically modified nucleoside present on rRNA and tRNA. The abnormal expressions of some gene indications are mediated through mRNA modifications. The recent progress of N4-Acetylcytidine on RNA expression plays a very important part in human diseases [40]. Future studies should be conducted to elucidate the potential biological regulation mechanisms regarding how the genetic variant affect the CHD outcome through N4-Acetylcytidine on RNA expression.

Some limitations of the present meta-analysis should be highlighted. First, our meta-analyses were based on unadjusted estimates. Future studies with potential confounding factors, such as age, ethnicity, sex, lifestyle factors and environmental exposure factors, should be conducted if possible. Second, obvious heterogeneity was detected in this study. We have performed meta-regression, Galbraith plots, sensitivity analysis, and subgroup analysis and the results of the present meta-analysis did not change. Third, the number of including publications and the sample size were relatively small. So, the findings should be interpreted with caution. Four, only one SNP within GSTP1 is not enough to elucidate the role of this gene on the susceptibility to CHD. Genome-wide association studies in the future should be conducted to investigate the association between single nucleotide polymorphisms (SNPs) in the GSTP1 gene and the risk of CHD [41]. Future studies should also be performed to see if the GSTP1 Ile105Val polymorphism or other SNPs in this gene are causally triggering the development of CHD through mediating the expression of this gene in specific tissues, like vascular or heart [42–44]. Finally, gene-gene, gene-nutrition, and gene-environment interactions were not
performed for the lack of sufficient information. Deep learning or machine learning is a hot topic in the classification and prediction of diseases based on biomarkers, and future studies should concentrate on the genetic variants, gene-gene, and gene-environment interactions for the prediction or early diagnosis of CHD [45,46].

Conclusions

In conclusion, our meta-analysis indicated that GSTP1 Ile105Val polymorphism did not appear to confer susceptibility to CHD. Further well-designed studies with detailed personal information are needed to validate the results of the present study.

Supporting information

S1 Checklist. PRISMA-P 2015 checklist. (DOCX)
S1 Fig. PRISMA flowchart describing the included/excluded literature. (TIF)

Acknowledgments

We appreciate the contribution of all the members participating in this study.

Author Contributions

Conceptualization: Yadong Song, Xiaoli Liu.
Data curation: Cheng Luo, Liangkai Chen.
Formal analysis: Yadong Song, Cheng Luo, Liangkai Chen.
Methodology: Yadong Song, Lin Gong.
Software: Cheng Luo, Liangkai Chen.
Supervision: Yadong Song, Xiaoli Liu.
Validation: Yadong Song, Xiaoli Liu.
Visualization: Lin Gong, Hanbin Yu, Bin Wang, Ernan Liu, Huiqiong Xu, Jiansheng Liang.
Writing – original draft: Yadong Song.
Writing – review & editing: Lin Gong, Hanbin Yu, Bin Wang, Ernan Liu, Huiqiong Xu, Jiansheng Liang.

References

1. Fuster V, Badimon L, Badimon JJ, Chesebro JH. The pathogenesis of coronary artery disease and the acute coronary syndromes (2). The New England journal of medicine. 1992; 326(5):310–8. http://dx.doi.org/10.1056/nejm199201303260506. PMID: 1728735.
2. Song Y, Shan Z, Liu X, Chen X, Luo C, Chen L, et al. An updated meta-analysis showed smoking modify the association of GSTM1 null genotype on the risk of coronary heart disease. Bioscience reports. 2021; 41(2). http://dx.doi.org/10.1042/bsr20200490. PMID: 33506866.
3. Harrison D, Griendling KK, Landmesser U, Hornig B, Drexler H. Role of oxidative stress in atherosclerosis. The American journal of cardiology. 2003; 91(3a):7a–11a. http://dx.doi.org/10.1016/s0002-9149(02)03144-2. PMID: 12645638.
4. Wilson MH, Grant PJ, Hardie LJ, Wild CP. Glutathione S-transferase M1 null genotype is associated with a decreased risk of myocardial infarction. FASEB journal: official publication of the Federation of
American Societies for Experimental Biology. 2000; 14(5):791–6. http://dx.doi.org/10.1096/fasebj.14.5.791. PMID: 10744635.

5. Van Schooten FJ, Hirvonen A, Maas LM, De Mol BA, Kleinjans JC, Bell DA, et al. Putative susceptibility markers of coronary artery disease: association between VDR genotype, smoking, and aromatic DNA adduct levels in human right atrial tissue. FASEB journal: official publication of the Federation of American Societies for Experimental Biology. 1998; 12(13):1409–17. http://dx.doi.org/10.1096/fasebj.12.13.1409. PMID: 9761785.

6. Andreassi MG, Botto N, Colombo MG, Biagini A, Clerico A. Genetic instability and atherosclerosis: can somatic mutations account for the development of cardiovascular diseases? Environmental and molecular mutagenesis. 2000; 35(4):265–9. http://dx.doi.org/10.1002/1098-2286(2000)35:4<265::aid-em1>3.0.co;2-m. PMID: 10861945.

7. Nomani H, Mozafari H, Ghobadloo SM, Rahimi Z, Raygani AV, Rahimi MA, et al. The association between GSTT1, M1, and P1 polymorphisms with coronary artery disease in Western Iran. Molecular and cellular biochemistry. 2011; 354(1–2):181–7. http://dx.doi.org/10.1007/s11010-011-0617-2. PMID: 21499713.

8. Cornelis MC, El-Soehmy A, Campos H. GSTT1 genotype modifies the association between cruciferous vegetable intake and the risk of myocardial infarction. The American journal of clinical nutrition. 2007; 86(3):752–8. http://dx.doi.org/10.1093/ajcn/86.3.752. PMID: 17823442.

9. Ryberg D, Skauv D, Hever A, Phillips DH, Harries LW, Wolf CP, et al. Genotypes of glutathione transferase M1 and P1 and their significance for lung DNA adduct levels and cancer risk. Carcinogenesis. 1997; 18(7):1285–9. http://dx.doi.org/10.1093/carcin/18.7.1285. PMID: 9230269.

10. Song Y, Shao Z, Luo C, Kang C, Yang Y, He P, et al. Glutathione S-Transferase T1 (GSTT1) Null Polymorphism, Smoking, and Their Interaction in Coronary Heart Disease: A Comprehensive Meta-Analysis. Heart, lung & circulation. 2017; 26(4):362–70. http://dx.doi.org/10.1016/j.hlc.2016.07.005. PMID: 27686690.

11. Hu X, Ji X, Srivastava SK, Xia H, Awaishi S, Nanduri B, et al. Mechanism of differential catalytic efficiency of two polymorphic forms of human glutathione S-transferase P1-1 in the glutathione conjugation of carcinogenic diol epoxide of chrysene. Archives of biochemistry and biophysics. 1997; 345(1):32–8. http://dx.doi.org/10.1006/abbi.1997.0269. PMID: 9281308.

12. Wang YH, Wu MM, Hong CT, Lien LM, Hsieh YC, Tseng HP, et al. Effects of arsenic exposure and genetic polymorphisms of p53, glutathione S-transferase M1, T1, and P1 on the risk of carotid atherosclerosis in Taiwan. Atherosclerosis. 2007; 192(2):305–12. http://dx.doi.org/10.1016/j.atherosclerosis.2006.07.029. PMID: 16973168.

13. Ramprasath T, Senthil Murugan P, Prabakaran AD, Gomathi P, Rathinavel A, Selvam GS. Potential risk modifications of GSTT1, GSTM1 and GSTP1 (glutathione-S-transferases) variants and their association to CAD in patients with type-2 diabetes. Biochemical and biophysical research communications. 2011; 407(1):49–53. http://dx.doi.org/10.1016/j.bbrc.2011.02.097. PMID: 21352813.

14. Singh N, Sinha N, Kumar S, Pandey CM, Agrawal S. Glutathione S-transferase gene polymorphism as a susceptibility factor for acute myocardial infarction and smoking in the North Indian population. Cardiology. 2011; 118(1):16–21. http://dx.doi.org/10.1159/000324066. PMID: 21389716.

15. Karčić Š, Nikolajević Starčević J, Petrović D. Association of manganese superoxide dismutase and glutathione S-transferases genotypes with myocardial infarction in patients with type 2 diabetes mellitus. Diabetes research and clinical practice. 2012; 98(1):144–50. http://dx.doi.org/10.1016/j.diabres.2012.07.003. PMID: 22853812.

16. Phulukdaree A, Khan S, Moodley D, Chuturgoon AA. GST polymorphisms and early-onset coronary artery disease in young South African Indians. South African medical journal = Suid-Afrikaanse tydskrif vir geneeskunde. 2012; 102(7):627–30. http://dx.doi.org/10.7196/samj.5520. PMID: 22748443.

17. Yeh HL, Kuo LT, Sung FC, Chiang CW, Yeh CC. GSTM1, GSTT1, GSTP1, and GSTA1 genetic variants are not associated with coronary artery disease in Taiwan. Gene. 2013; 523(1):64–9. http://dx.doi.org/10.1016/j.gene.2013.02.052. PMID: 23570881.

18. Kovacs V, Gasz B, Balatonyi B, Jaromi L, Kisfali P, Borsiczky B, et al. Polymorphisms in glutathione S-transferase are risk factors for perioperative acute myocardial infarction after cardiac surgery: a preliminary study. Molecular and cellular biochemistry. 2014; 389(1–2):79–84. http://dx.doi.org/10.1007/s11010-013-1929-7. PMID: 24435850.

19. Ding N, Wang X, Weisskopf MG, Sparrow D, Schwartz J, Hu H, et al. Lead-Related Genetic Loci, Cumulative Lead Exposure and Incident Coronary Heart Disease: The Normative Aging Study. PloS one. 2016; 11(9):e0161472. http://dx.doi.org/10.1371/journal.pone.0161472. PMID: 27584680.

20. Bhat MA, Gandhi G. Glutathione S-transferase P1 gene polymorphisms and susceptibility to coronary artery disease in a subgroup of north Indian population. Journal of genetics. 2017; 96(6):927–32. http://dx.doi.org/10.1007/s12041-017-0863-y. PMID: 29321351.
21. Bhatti JS, Vijayvergiya R, Singh B, Bhatti GK. Genetic susceptibility of glutathione S-transferase genes (GSTM1/T1 and P1) to coronary artery disease in Asian Indians. 2018; 82(6):448–56. http://dx.doi.org/10.1111/ahg.12274. PMID: 3039864.

22. Simeunovic D, Odanovic N, Pjesa-Ercegovac M, Radic T, Radovanovic S, Covic V, et al. Glutathione Transferase P1 Polymorphism Might Be a Risk Determinant in Heart Failure. Annals of human genetics. 2019; 6984845. http://dx.doi.org/10.1111/ahg.12274 PMID: 3127451.

23. Pourkeramati A, Zare Mehrjardi E. Association of GSTP1, GSTT1 and GSTM1 Gene Variants with Coronary Artery Disease in Iranian Population: A Case-Control Study. 2020; 13:249–59. http://dx.doi.org/10.2147/jigm.s252552. PMID: 32547167.

24. Su H, Cao Y, Li J, Zhu Y, Ma X. GST null polymorphisms may affect the risk of coronary artery disease: evidence from a meta-analysis. Thrombosis journal. 2020; 18:20. http://dx.doi.org/10.1186/s12959-020-00234-x. PMID: 32905149.

25. Li Y, Li L, Fan D, Wang Z, Cui Y. Effects of GST null genotypes on individual susceptibility to atherosclerotic cardiovascular diseases: a meta-analysis. Free radical research. 2020; 54(8–9):567–73. http://dx.doi.org/10.1080/10715762.2019.1624743. PMID: 33103508.

26. Xu MQ, Ye Z, Hu FB, He L. Quantitative assessment of the effect of angiotensinogen gene polymorphisms on the risk of coronary heart disease. Circulation. 2007; 116(12):1356–66. http://dx.doi.org/10.1161/circulationaha.107.728857. PMID: 17846284.

27. Xu M, Sham P, Ye Z, Lindpaintner K, He L. A1166C genetic variation of the angiotensin II type I receptor gene and susceptibility to coronary heart disease: collaborative of 53 studies with 20,435 cases and 23,674 controls. Atherosclerosis. 2010; 213(1):191–9. http://dx.doi.org/10.1016/j.atherosclerosis.2010.07.046. PMID: 20732682.

28. Xu M, Lin Z. Genetic influences of dopamine transport gene on alcohol dependence: a pooled analysis of 13 studies with 2483 cases and 1753 controls. Progress in neuro-psychopharmacology & biological psychiatry. 2011; 35(5):1255–60. http://dx.doi.org/10.1016/j.pnpbp.2010.11.001. PMID: 21078357.

29. Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. BMJ (Clinical research ed). 2007; 335(7626):914–6. http://dx.doi.org/10.1136/bmj.39343.408449.80. PMID: 17974687.

30. Ioannidis JP, Patsopoulos NA, Evangelou E. Uncertainty in heterogeneity estimates in meta-analyses. BMJ (Clinical research ed). 2007; 335(7626):914–6. http://dx.doi.org/10.1136/bmj.39343.408449.80. PMID: 17974687.

31. He J, Wang K, Lo K, Zhong Y, Yang A, Fang X, et al. Sex-Specific Association of Circulating Ferritin Level and Risk of Type 2 Diabetes: A Dose-Response Meta-Analysis of Prospective Studies. Translational psychiatry. 2019; 104(10):4539–51. http://dx.doi.org/10.1038/s41398-020-00902-6 PMID: 31074789.

32. Galbraith RF. A note on graphical presentation of estimated odds ratios from several clinical trials. Statistics in medicine. 1998; 7(8):889–94. http://dx.doi.org/10.1002/sim.4780070807. PMID: 3413368.

33. Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for publication bias. Biometrics. 1994; 50(4):1088–101. PMID: 7786990.

34. He J, Wang F, Zhu JH, Chen W, Cui Z, Jia WH. No association between MTR rs1805087 A > G polymorphism and non-Hodgkin lymphoma susceptibility: evidence from 11 486 subjects. Leukemia & lymphoma. 2015; 56(3):763–7. http://dx.doi.org/10.1038/lj.2014.93570. PMID: 24956146.

35. Yan X, Zhao X, Li J, He L, Xu M. Effects of early-life malnutrition on neurodevelopment and neuropsychiatric disorders and the potential mechanims. Progress in neuro-psychopharmacology & biological psychiatry. 2018; 83:64–75. http://dx.doi.org/10.3109/10428194.2014.935370. PMID: 24956146.

36. Li H, Wang X, Lu X, Zhu H, Li S, Duan S, et al. Co-expression network analysis identified hub genes critical to triglyceride and free fatty acid metabolism as key regulators of age-related vascular dysfunction in mice. Aging. 2019; 11(18):7620–38. http://dx.doi.org/10.18632/aging.103150. PMID: 31514170.

37. Zheng S, Zhao T, Yuan S, Yang L, Ding J, Cui L, et al. Immunodeficiency Promotes Adaptive Alterations of Host Gut Microbiome: An Observational Metagenomic Study in Mice. Frontiers in microbiology. 2019; 10:2415. http://dx.doi.org/10.18632/aging.103150. PMID: 31781050.

38. Chen J, Zhao X, Cui L, He G, Wang X, Wang F, et al. Genetic regulatory subnetworks and key regulating genes in rat hippocampus perturbed by prenatal malnutrition: implications for major brain disorders. The Journal of clinical endocrinology and metabolism. 2020; 12(9):8434–58. http://dx.doi.org/10.1210/clinem/dgab454. PMID: 32392183.

39. Wang X, Jiao X, Tian Y, Zhang J, Zhang Y, Li J, et al. Associations between maternal vitamin D status during three trimesters and cord blood 25(OH)D concentrations in newborns: a prospective Shanghai birth cohort study. 2021. http://dx.doi.org/10.1111/pnpbp.2017.12.016 PMID: 33661376.
40. Jin G, Xu M, Zou M, Duan S. The Processing, Gene Regulation, Biological Functions, and Clinical Relevance of N4-Acetylcytidine on RNA: A Systematic Review. Molecular therapy Nucleic acids. 2020; 20:13–24. http://dx.doi.org/10.1016/j.omtn.2020.01.037. PMID: 32171170.

41. Wu Y, Cao H, Baranova A, Huang H, Li S, Cai L, et al. Multi-trait analysis for genome-wide association study of five psychiatric disorders. 2020; 10(1):209. http://dx.doi.org/10.1038/s41398-020-00902-6. PMID: 32606422.

42. Zhang F, Baranova A, Zhou C, Cao H, Chen J, Zhang X, et al. Causal influences of neuroticism on mental health and cardiovascular disease. Human genetics. 2021. http://dx.doi.org/10.1007/s00439-021-02288-x. PMID: 33973063.

43. Zhang F, Rao S, Cao H, Zhang X, Wang Q, Xu Y, et al. Genetic evidence suggests posttraumatic stress disorder as a subtype of major depressive disorder. The Journal of clinical investigation. 2021. http://dx.doi.org/10.1172/jci145942. PMID: 33905376.

44. Wang X, Fang X. Genetic support of a causal relationship between iron status and type 2 diabetes: a Mendelian randomization study. 2021. http://dx.doi.org/10.1210/clinem/dgab454. PMID: 34147035.

45. Liu M, Li F, Yan H, Wang K, Ma Y, Shen L, et al. A multi-model deep convolutional neural network for automatic hippocampus segmentation and classification in Alzheimer’s disease. NeuroImage. 2020; 208:116459. http://dx.doi.org/10.1016/j.neuroimage.2019.116459. PMID: 31837471.

46. Yu H, Pan R, Qi Y, Zheng Z, Li J, Li H, et al. LEPR hypomethylation is significantly associated with gastric cancer in males. Experimental and molecular pathology. 2020; 116:104493. http://dx.doi.org/10.1016/j.yexmp.2020.104493. PMID: 32659237.