Multichannel Residual Cues for Fine-Grained Classification in Wireless Capsule Endoscopy

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ABSTRACT Early diagnosis of gastrointestinal pathologies leads to timely medical intervention and prevents disease development. Wireless Capsule Endoscopy (WCE) is used as a non-invasive alternative for gastrointestinal examination. WCE can capture images despite the structural complexity presented by human anatomy and can help in detecting pathologies. However, despite recent progress in fine-grained pathology classification and detection, limited works focus on generalization. We propose a multi-channel encoder-decoder network for learning a generalizable fine-grained pathology classifier. Specifically, we propose to use structural residual cues to explicitly impose the network to learn pathology traces. While residuals are extracted using well-established 2D wavelet decomposition, we also propose to use colour channels to learn discriminative cues in WCE images (like red color in bleeding). With less than 40% data (fewer than 2500 labels) used for training, we demonstrate the effectiveness of our approach in classifying different pathologies on two different WCE datasets (different capsule modalities). With a comprehensive benchmark for WCE abnormality and multi-class classification, we illustrate the generalizability of the proposed approach on both datasets, where our results perform better than the state-of-the-art with much fewer labels in abnormality sensitivity on several of nine different pathologies and establish a new benchmark with specificity > 97% across classes.

INDEX TERMS Pathology classification, wireless capsule endoscopy, fine grained, residual cues.

I. INTRODUCTION

The gastrointestinal tract, which plays a vital role in absorbing essential nutrients, also exhibits susceptibility to various intestinal pathologies such as ulcers, polyps, lesions and inflammation, amongst others. The lack of proper management of inflammatory bowel disease can lead to encumbering economies with substantial costs at different stages of disease management pipeline [2] in addition to reducing the quality of life among patients [3], [4]. Therefore, the early diagnosis of such pathological occurrences as colorectal polyps or obscure intestinal bleeding can lead to timely intervention and possible prevention of further disease development to extreme cases such as inflammatory bowel diseases and cancers [5], [6], [7]. While standard diagnostic approaches like colonoscopy and gastroscopy enable visualization of the upper and lower gastrointestinal tract, anatomical challenges prevent satisfactory examination of the small bowel through these techniques [8]. As a consequence, Wireless Capsule Endoscopy (WCE) has become an increasingly preferred non-invasive alternative, especially for small bowel examinations [9].

Despite progress in WCE imaging [10], a medical practitioner needs to spend reasonable time analysing frames in excess of 60,000 [11], amounting approximately to 45 to 60 minutes [12] of the practitioner’s time for large bowel diagnosis. Despite the substantial time requirement, the diagnosis has susceptibility to human error [13]. Reliable computer-aided classification tools for detecting normal from pathological conditions can be paramount in cutting down individual diagnostic times and associated costs [14]. A number of approaches have been proposed in the last
decade to classify the images/videos using classical techniques as well as deep learning based architectures [15]. While classical approaches are carefully engineered by understanding the structure and appearance of pathology in images [16], deep learning architectures identify and classify pathological appearances by learning to extract task-beneficial features from large datasets. Although classifying a normal image against pathology is of great clinical significance, there are limited works that attempt to do fine-grained classification involving different pathologies within a single framework [8], [17], leading to a problem of generalization in real-time. The dearth of works in this direction can be attributed to limitations pertaining to the private nature of this data and the scarcity of sufficiently varied examples of different cases, often leading to inadequately representative, unbalanced datasets for one or more pathologies. While machine learning-based algorithms enable prediction of normal against pathological conditions with high confidence [18], [19], [20], attempts at discriminating between different pathological conditions presents a considerable generalization challenge, even with large datasets, as also noted in the multi-centric study by Ding et al. [8]. Figure 1 shows few examples of pathological images obtained from WCE from the Kvasir Capsule Dataset [1] illustrating the varying appearances of pathologies in different images and the challenge in classifying them with high confidence, especially with considerations to changing anatomy within the gastrointestinal tract.

We present a new approach for classifying various pathologies from normal images using a multi-channel Convolutional Neural Network (CNN) based cascaded encoder-decoder network with a ResNet [21] backbone. We observe that occurrence of pathology is coupled with tissue transitions in different directions and that these transitions could provide useful cues for pathology identification. In line with this, we propose to extract directional residuals based on 2D-Wavelet decomposition. We illustrate this assertion with a toy example in Figure 2 where an image is decomposed into three directions that capture different directional aspects of the information contained within the original image. Concretely, the original image (top-left) is decomposed in horizontal (top-right), vertical (bottom-left) and diagonal (bottom-right) directions using 2D-Discrete wavelet transform. This directional information for each orientation is then subtracted from the original image to provide the corresponding residuals, which we refer to as structural residual cues. We experimentally verify that in WCE, these directional residuals not only capture the directional transitions resulting from an abnormality but carry additional structural details particular to different abnormalities that allows learning better discrimination between normal and pathological cases (Table. 1) as well as between different pathological cases (Table. 4).

In addition to structural changes, transitions in colour across different spatial regions of an image can provide further clues that medical practitioners often use for deciding on the presence of a pathological condition. For instance, the red colour is pronounced when a blood clot is observed in the tract compared to an ulcer which appears yellowish. Motivated by such observations, we use colour channels in addition to structural residuals to learn the classification model robustly. Thus, our approach works by employing six different channels consisting of both structural residuals and appearance factors in different colour spaces.
Finally, we propose to exploit intermediate representations at different stages of the cascaded Encoder-Decoder network towards obtaining a final decision to fully utilize features of different scales and sensitivity. We do this through two ways: (a) enforcing a symmetrical loss between different intermediate latent representations between the encoder and the decoder, using a standard triplet loss at the batch level. (b) incorporating a pixel-wise reconstruction loss between the ends, i.e., the input and the decoder output. In addition, a standard Negative Log-Likelihood loss is employed to train the network in a supervised manner to classify the normal versus pathology images by taking full advantage of the 6 channel CNN.

We validate our proposed approach on the fine-grained classification of two WCE datasets - Kvasir-Capsule dataset (D1) [1] and the Computer Assisted Diagnosis for Capsule Endoscopy Database (CAD-CAP) from the Giana Endoscopic Vision Challenge (D2) [22], with up to nine different pathology categories. The obtained results surpass previous baselines in per-pathology sensitivity while closing the gap in sensitivity and specificity with other works with significantly higher labels used in training. We demonstrate the applicability of the proposed approach for classifying many different pathologies (using the same network, no additional architectural changes) from normal across two datasets. In addition, we evaluate the strength of features in characterizing challenging categories through a multi-class classification between pathology categories. We present a detailed analysis on the explainability of the network through analysis of channel contributions. The main contributions are:

- We propose a unified multi-channel CNN based cascaded Encoder-Decoder network to classify multiple WCE pathologies with consistently high sensitivity and specificity.
- We demonstrate generalizability in challenging cross-pathology (nine different non-overlapping classes of datasets) and cross-dataset settings (with different capsule modalities).
- Unlike previous works in WCE, our work presents a more comprehensive baseline for evaluating future works, with the inclusion of multiple commonly occurring pathologies in WCE.

In the rest of the paper, we present the rationale and proposed approach in detail in Section II, followed by a summary of the datasets employed for validation. Section IV presents the detailed analysis of the results and Section V provides the discussion on explainability. Further, Section VI provides the results of the ablation studies with respect to the losses and Section VIII discusses the conclusions.

II. METHODOLOGY

The image of a healthy organ typically exhibits different visual properties from the one with diseases/pathology as illustrated in Figure 1. A pathological image $I_p$ can be asserted as a corrupted version of a healthy image $I_h$ where the corruption refers to the specific structural irregularities over and above normal variations, resulting from the pathology. Thus, a pathology image $I_p$, can be described as a some function $f$ of healthy image $I_h$ and corruption $\omega(\cdot)$ i.e, $I_p = f(I_h) + \omega(\cdot)$ (the corruption being a complex interplay of patient related pathological and physiological variables). Assuming that the healthy images are abundant, we formulate pathological characteristics as outliers deviating from normal healthy images with different abnormal appearances. Thus, we propose an approach based on such an assertion to learn the outliers in the presence of pathology as detailed in the upcoming section.

A. MULTICHANNEL INPUT THROUGH STRUCTURAL RESIDUAL CUES AND COLOUR INFORMATION

The pathological and normal images may appear very similar in multiple cases, given the complex nature of these images. However, some superficial deposits or patterns can be observed in images with pathology when captured with WCE, and using only RGB images may not necessarily lead to features robust enough for learning optimal
classifiers often leading to mis-classifications. Therefore, we first seek to obtain crucial discriminative differences. As the pathology characteristics can appear in all possible orientations and shapes, it is vital to extract pathology traces in all such directions. Although such information can be extracted using different spatial approaches, we rely upon the 2D-discrete wavelets transforms (2D-DWT) to capture details in horizontal, vertical, and diagonal directions as illustrated in Figure 2 to obtain the traces of pathology in different orientations. The wavelets from each of the horizontal, vertical, and diagonal directions provide clear transitions between the areas of pathology and non-pathological regions. It has to be, however, noted that such transitions can also appear due to the tissues that are normally present without any pathology. To deal with such a situation, we take the residual differences between the original image and each of the responses from horizontal, vertical, and diagonal directions leading to a set of structural residuals. We first obtain the gray-level image $I_g$ of $I$ and then obtain the residuals along horizontal, vertical, and diagonal directions $I_{h}, I_{v}, I_{d},$ represented as

\[
I_{h} = I_g - \psi^{(h)}I_g
\]

\[
I_{v} = I_g - \psi^{(v)}I_g
\]

\[
I_{d} = I_g - \psi^{(d)}I_g
\]

where $\psi$ represents the 2D-DWT in different directions at level 1.

While all residual images capture structural transitions in different orientations and discard colour information, our approach may suffer from missing colour information. To account for this, we combine the structural residual information with the colour channel information to create a multi-channel input. Thus, the RGB channels of input image $I$ is appended to form a final six channel input resulting in $[I_{h}, I_{v}, I_{d}, I_R, I_G, I_B]$ for learning the classifier.

### B. LEARNING TO CLASSIFY RESIDUAL CUES

As discussed earlier, a pathological image can be considered as a corrupted variant of the healthy image given by $I_p = f(I_h) + \omega()$. Thus, we assert that learning the cues corresponding to $\omega()$ leads to a better classification of the pathological images from healthy images as well as within themselves. We, therefore, propose an Encoder-Decoder architecture to learn the cues more effectively discriminating healthy images from pathology images.

Given the multi-channel input, we extract the features for learning the classifier using a Residual Network (ResNet-18) [21] backbone with 18 layers. Unlike the traditional three-channel input for a deep neural network, the input in our case has six channels, thus we modify the ResNet-18 architecture to process multi-channel inputs for feature extraction. Furthermore, to account for the limited data settings which can result in an overfitted network when learning from scratch, especially in our case for pathological images where the data scarcity cannot be overruled, we use the pre-trained weights learnt on ImageNet [23] for the last three channels (i.e., RGB) and set the weights of other three channels corresponding to residual differences to zero. To make the learning robust and incorporate all intermediate features, we adopt a U-Net [24] like Encoder-Decoder architecture, where we choose four Encoder ($E$) residual blocks followed by a corresponding number of Decoder ($D$) residual blocks. Such an Encoder-Decoder architecture helps us to decode the information back to generate the original six channel input. The feature maps from each layer are upsampled by a nearest-neighbour interpolation through a $2 \times 2$ convolution in each of the successive decoder blocks. We exploit the intermediate symmetrically encoded-decoded representations (i.e., Encoder-1 with Decoder-4) by using it to measure the differences using a standard triplet loss by formulating it as an anomaly problem inspired by a recent work by Feng et al. [25].

Learned feature representations for images of the same class must ideally lie in a compact sphere, while features corresponding to samples from dissimilar classes lie further away in the representation space. In an analogy, the representation of a pathological image must be such that it drifts away (exhibits smaller pairwise similarity) from the center of the cluster of healthy images in the feature space. If the input space is represented by $(X \subseteq \mathbb{R}^d)$ and the output space by $(Z \subseteq \mathbb{R}^p)$, a neural network with $L$ hidden layers and corresponding set of weights $W = \{W^1, \ldots, W^L\}$ learns a mapping between the input space and the output space as $\phi(\cdot; W) : X \rightarrow Z$ such that $Z$ exhibits the desirable properties described where $Z$ is centered on a predetermined point $c$.

Given $n$ healthy samples $(I^h_1, \ldots, I^h_n \subseteq X)$ and $k$ pathological samples $(I^p_1, \ldots, I^p_k \subseteq X)$, the point $c$ in the output space $Z$ should lie close to healthy samples while the pathological samples are encoded such that they lie away from this healthy cluster.

\[
\min_{W} \frac{1}{h} \sum_{i=1}^{h} \left\| \phi(I^h_i; W) - c \right\|^2 \tag{1}
\]

\[
\max_{W} \frac{1}{p} \sum_{i=1}^{k} \left\| \phi(I^p_i; W) - c \right\|^2 \tag{2}
\]

Minimizing the distance for samples from healthy class while maximizing the distance for samples from pathological class from $c$ can lead to better classification. To achieve this, we employ triplet loss $L_t$ to minimize intra-class separability and maximize inter-class separability at the feature level [26]. Using all intermediate feature representations from the last encoder block to the last decoder block, we create triplets of healthy images and pathological images in each batch by randomly choosing healthy samples as anchors and the rest of the healthy samples as positives and pathological samples as negative. Thus, by enforcing a triplet loss that uses 6 input channels (residual and RGB), the loss estimation can be fully used to benefit from learning class separation.
TABLE 1. Comparison of performance between different datasets and approaches for abnormality detection. All pathologies are clubbed as a single class of abnormality and classified against the normal class.

| Approach       | Total no. of images | Sensitivity | Specificity | AUC  | Accuracy | Abnormalities                                      |
|----------------|---------------------|-------------|-------------|------|----------|---------------------------------------------------|
| Ding et. al. [8] | 158323              | 99.9        | 100         | –    | –        | Inflammation, Polypos, bleeding, vascular disease, protruding lesion, diverticulum, parasite, ulcer, lymphatic follicular hyperplasia, lymphangiectasia Vs Normal |
| Iakovidis et. al [27] | 1136                | 92.4        | 85.8        | –    | 89.2     | Vascular anomalies, polypoid anomalies inflammatory anomalies Vs Normal |
| Ours [D1]      | 5489                | 99.3        | 99.6        | 99.4 | 99.42    | Angiectasia, Blood, Erosion, Lymphoid Hyperplasia Vs Normal |
| Ours [D2]      | 1812                | 97.63       | 100         | 98.8 | 99.2     | Inflammatory, Vascular Lesion Vs Normal |

As the pathological cues obtained from the input channels (wavelet residuals and colour channels) can exhibit dissimilar features compared to healthy images, one can deduce that the encoder-decoder network shall result in an image close to the input image for healthy classes in each of the six channels. Thus, the difference between the input and the output image with six channels for the normal image set is expected to ideally result in zero difference if reconstructed pixel-wise. Assuming zero difference for healthy images when reconstructed, we incorporate a pixel-wise loss to boost learning of normal image classes through explicit supervision. Given a six-channel input image \( I \), the final residual representation obtained from the encoder-decoder block results in a low pixel-wise reconstruction loss \( L_r \), which can be represented as:

\[
L_r = \frac{1}{hch} \sum_{i \in h} \sum_{c=1}^{6} ||C_i||_1,
\]

for \( h \) healthy images in a batch of size \( b \), \( ch \) being the total number of input channels (i.e., \( ch = 6 \)) and \( C_i \) is the channel wise difference between the input and output for image \( i \).

The output of the final decoder block using a Tanh activation layer provides an activation map corresponding to the residual cues that are different in pathological images against healthy images. We, therefore, employ it as an attention mechanism to guide the auxiliary classifier to learn the final classification layer for our proposed model. Figure 4 presents a sample illustration of the cues obtained from the proposed approach, and it can be noted that the visual cues learnt by the network localize the regions of pathology, helping the network in better classification.

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\[
L = \lambda_1 L_r + \lambda_2 \sum_{k \in \{E5, D4\}} L^k_t + \lambda_3 L_a,
\]

where \( \lambda s \) are regularization parameters set to \( \lambda_1 = \lambda_2 = \lambda_3 = 1 \).

FIGURE 4. Visual multi-channel attention cues obtained for an example image for Lymphoid Hyperplasia (top) and bleeding (bottom) through proposed network. The left image represents the input image, the top row represents the cues learnt for each input channel and the bottom row represents the corresponding inputs in multi-channel input. *Note-The images are shown in colour-map for better visualization.

III. DATASETS

We employ two databases, Kvasir-Capsule dataset (D1) [1] and the Computer Assisted Diagnosis for Capsule Endoscopy Database (CAD-CAP) from the Giana Endoscopic Vision Challenge (D2) [22] to demonstrate the applicability of the proposed approach. We consider nine different classes with different pathologies such as erythema, blood, erosion, angiectasia, ulcer, Lymphangiectasia, polyp, in addition to 2000 randomly sampled clean images from the normal class. (The classes reduced mucosal view (GI debris) and foreign body (capsule endoscope from previous examination) have been excluded from our experiments since they represent neither pathology nor normal scenario). While the Kvasir-Capsule dataset [1] is heavily imbalanced, Giana Endoscopic Vision Challenge [22] provides balanced samples for a set of two pathological conditions and one normal class. Specifically, D2 provides 607 inflammatory images, 605 vascular lesion images and 600 normal images organized in three classes. Irrespective of the dataset and samples for each pathology, we employ a train-validation-test split of 40:10:50 across all experiments.

IV. EXPERIMENTS AND RESULTS

A. TRAINING DETAILS

All the training and testing was conducted on an Nvidia 2080 Ti GPU enabled computer with Linux operating
TABLE 2. Per-pathology sensitivity against the normal class for each of the nine classes in datasets D1 and D2.

| Abnormality       | Sensitivity [%] | Specificity [%] | AUC [%] | Accuracy [%] |
|-------------------|-----------------|-----------------|---------|--------------|
| Angiectasia       | 97.8            | 99.6            | 98.7    | 99.1         |
| Blood             | 100             | 100             | 100     | 100          |
| Erosion           | 94.9            | 99.3            | 97.1    | 98.5         |
| Erythematous      | 97.4            | 100             | 98.7    | 99.7         |
| Lymphoidous       | 98.9            | 100             | 99.5    | 99.7         |
| Hyperplasia       |                 |                 |         |              |
| Polyp             | 96.8            | 100             | 98.4    | 99.9         |
| Ulcer             | 98.6            | 100             | 99.3    | 99.6         |
| Inflammatory      | 99.6            | 97.3            | 98.4    | 98.5         |
| Vascular Lesion   | 99.3            | 98.3            | 98.8    | 98.8         |

TABLE 3. Pathology-wise performance comparison for pathology sensitivity against normal, in comparison with other recent works with same or similar class of pathology in WCE (metrics below are as reported in original works).

| Approach | Accuracy | Sensitivity | Specificity |
|----------|----------|-------------|-------------|
| Bleeding | Jia et al. (2016) [20] | 99.9 | 99.2 | – |
|          | Pan et al. (2011) [18] | – | 93.1 | 96 |
|          | Aoki et al. (2020) [28] | 99.89 | 96.63 | 99.96 |
|          | Aoki et al. (2021) [29] | – | 100 | – |
|          | Afonso et al. (2021) [30] | – | 98.3 | 98.4 |
| Ours     | 100      | 100         | 100         |

Erosions and Ulcers

| Approach | Accuracy | Sensitivity | Specificity |
|----------|----------|-------------|-------------|
| Fau et al. (Ulcer, 2018) [31] | 95.16 | 96.8 | 94.79 |
| Wang et al. (2019) [32] | 90.1 | 89.71 | 90.48 |
| Aoki et al. (2021) [29] | – | 100 | – |
| Fau et al. (Erosion, 2018) [31] | 95.34 | 93.67 | 95.98 |
| Aoki et al. (2019) [33] | 90.8 | 88.2 | 90.9 |
| Ours (Ulcer) | 99.59 | 98.56 | – |
| Ours (Erosion) | 99.32 | 94.88 | 98.56 |

Polyp (or Protruding Lesions)

| Approach | Accuracy | Sensitivity | Specificity |
|----------|----------|-------------|-------------|
| Yuan et al. (2017) [34] | 98 | – | – |
| Aoki et al. (2021) [29] | – | 99 | – |
| Ours     | 99.91   | 96.78 | 100 |

Angiectasia

| Approach | Accuracy | Sensitivity | Specificity |
|----------|----------|-------------|-------------|
| Noya et al. (2017) [35] | 89.51 | 96.8 | – |
| Leenhardt et al. (2018) [12] | 100 | 96 | – |
| Tsuibo et al. (2019) [36] | – | 98.8 | 98.4 |
| Aoki et al. (2021) [29] | – | 97 | – |
| Ours     | 99.08   | 97.88 | 99.55 |

B. ABNOMALITY DETECTION RESULTS

We first perform abnormality detection as a binary classification problem where all pathologies constitute a single class of abnormality. The other class is of normal images. The results can be seen in Table 1. We compare our approach with Iakovidis et al. [27] and Ding et al. [8] who have also performed similar binary classification in WCE. While a direct comparison across our work and earlier works [27], [8] is not possible due to the absence of a common dataset, our approach significantly closes the gap for abnormality sensitivity as well as achieves high specificity for normal cases. In comparison to Iakovidis et al. [27] with similar scale of data (Table 1. 40% of 1812 and 5489), and Ding et al. [8] with much more data (approx 100 times more images), we perform better than both the earlier approaches under limited training data (40% train and 10% validation), indicating robust representations for filtering suspected abnormality.

C. PER-PATHOLOGY RESULTS

Different pathologies exhibit different levels of complexities, due to varying visual appearances and scales as compared to normal images, for example blood pathology is easier to distinguish due to large changes in overall color in the image whereas small scale inflammation or lesions may be harder. Thus resulting classification models may have varying sensitivities for different pathologies. We test if the model generalization extends to different pathologies through multiple binary classifications against each of the commonly known pathology classes in WCE. Table 2 shows the results. The sensitivity in all cases (except erosion) equals or exceeds 97%. Furthermore, the high specificity of our system (Ubuntu 20.04). The training was conducted with 30 epochs, with a batch size of 32 and a base learning rate of 5e-4. Three milestone epochs were used at epoch number 5, 8 and 12 to decrease the learning rate by 0.3 to converge the learning faster using multi step learning rate scheduler from PyTorch.

TABLE 4. Multiclass pathology classification on Giana Endoscopic Vision Challenge (D2) [22].

| Abnormality       | Sensitivity | Specificity | AUC | Accuracy |
|-------------------|-------------|-------------|-----|----------|
| Valeiro et al. (2019) [11] | – | – | 87 | 93 |
| Vats et al. (2021) [11] | – | – | 56 | 58.7 |
| Ours | 91.38 | 95.52 | 93.52 | 91.35 |
approach on all classes (especially the fine-grained classes like Erosion and Erythema) can be attributed to the residuals being strong indicators of pathological structures, despite the presence of possible confounding factors like debris and bubbles. This can also be observed from Figure 4 which localizes the residuals helping in classifying the pathology better.

Next, Table 3 presents a comparison of pathology-wise sensitivities with other recent works in WCE. As seen, our work surpasses on sensitivity baseline from most other works and shows competitive performance to Aoki et al. [29] who use a dataset of over 66,000 images (approximately 12 times larger than ours).

D. MULTICLASS PATHOLOGY CLASSIFICATION

One crucial aspect for computer-assisted-diagnosis under WCE is when multiple challenging pathologies can be reliably discriminated within a single end-to-end network. To evaluate this, we perform a multi-class pathology classification to evaluate the strength of features in discriminating such categories as the inflammatory and vascular lesion [17] (in contrast to only discriminating pathology from normal). We provide a comparison of our results with other recent works on the same dataset D2 [11], [17] as presented in Table 4. It can be noted that our method surpasses Vats et al. [17] by a large margin and closes the gap with Valerio et al. [11], who also perform the same classification with features transferred from ImageNet [23], but with a larger (DenseNet161) architecture. The superiority of the results can be attributed to residual features and colour channels in a multi-channel ResNet architecture helping learn discriminative features.

V. EXPLAINABILITY ANALYSIS

A commonly employed way of gaining trust in model predictions is to examine its behavior through visual explanations and checking if this agrees with domain knowledge/intuition about the problem. We test the behavior of our classifier in the locality of test samples using ‘Local Interpretable Model-agnostic Explanations’ (LIME) [37]. LIME works by using the classifier to be evaluated as an input and generates post-hoc explanations contributing to a prediction. For images, this amounts to visualizing image components contributing positively to the true class. We visualize these for a random selection of abnormal images from D2 for the multi-class pathology classifier (Table 4) in Fig.5. The image with abnormality show in yellow circle can be seen in the first columns (Image), followed by components that weight positively to pathology class (middle column). The visualizations show that image regions with abnormality (yellow circle) factor in positively to pathology, whereas seemingly normal regions contribute to normalcy. Interestingly also, we see that visually pervasive pathologies (row 4. and 5.) can be explained with sparser components within the image, whereas obscure cases like in row 2. and 3. includes comparatively denser explanatory components.

We also visualized components contributing negatively to abnormality (positively to the normal class), these can be seen in the right column. In line with expectation, most of the right column’s components comprise of non-pathology sub-regions within images.

VI. ABLATION: CHANNELS

In this section, we study the impact of dropping different channels on the model’s performance. Table 5 shows the classification performance by dropping one of the 6 channels. Similarly, Fig. 6 shows the change in the relative contribution of superpixels to the true class as each of the channels are sequentially zeroed out.
We observed that dropping channel 5 produces consistently sparser explanation as channel 5 encodes low level frequency details of input, while channel 0 exhibits higher pathology sensitivity.

TABLE 5. Impact of each channel on model performance. Evaluated for Multi-pathology classification on D2.

| Channel 0 | Channel 1 | Channel 2 | Channel 3 | Channel 4 | Channel 5 | Accuracy | Sensitivity | AUC    |
|-----------|-----------|-----------|-----------|-----------|-----------|----------|-------------|--------|
| x         | ✓         | ✓         | ✓         | ✓         | ✓         | 36.9     | 52.8        | 37.1   |
| ✓         | x         | ✓         | ✓         | ✓         | ✓         | 28.3     | 46.3        | 28.5   |
| ✓         | ✓         | x         | ✓         | ✓         | ✓         | 33.5     | 50          | 33.3   |
| ✓         | ✓         | ✓         | x         | ✓         | ✓         | 72.8     | 72.9        | 79.6   |
| ✓         | ✓         | ✓         | ✓         | x         | ✓         | 40.0     | 39.8        | 54.9   |
| ✓         | ✓         | ✓         | ✓         | ✓         | x         | 36.6     | 36.8        | 52.6   |

VII. ABLATION STUDY: IMPACT OF LOSS FUNCTIONS

We study the impact of the different loss functions in the proposed approach in this section. Table 6 shows performance obtained for each loss function and the combination of losses used in this work. For the sake of simplicity, we present the ablation studies on dataset D2 alone, where the extremes of
the loss function behaviour can be observed. The Negative Log-Likelihood (NLL) loss ($L_{\text{NLL}}$) which operates solely on the embedding of the ultimate layer, is the dominant discriminatory signal for classification between normal and abnormal classes. However, the sensitivity for hard cases is improved even further with the addition of both feature-level (triplet) $L_{\text{t}}$ and pixel-level (Pix-Reg) $L_{\text{p}}$ discrimination. We further observe that in the absence of the dominant discrimination from the supervised categorical classification loss, neither of the two losses or their combination is particularly useful in classification. The combination ($L_{\text{NLL}}$ and $L_{\text{t}}$) tends to almost perfect performance, therefore we interpret the results more strictly in this case, as too good a performance may indicate a lack of generalizability or be caused by overfitting (for e.g., to samples coming from the same patient exhibiting similarities). We suspect the learning to benefit additionally from the regularization effect coming from $L_{\text{p}}$ as seen in the combination ($L_{\text{NLL}}, L_{\text{t}}, L_{\text{p}}$), hence the loss being a weighted average of all three $L_{\text{NLL}}, L_{\text{t}}, L_{\text{p}}$.  

**VIII. CONCLUSION**

We have proposed a multi-channel cascaded encoder-decoder network in this work for learning a generalizable fine-grained pathological classifier. Noting the differences in the colour appearance of pathology in different directions and orientations, we have proposed to extract residual cues by extracting the differences in wavelets across horizontal, vertical, and diagonal directions in addition to using regular colour channels. The proposed encoder-decoder network fully exploits the multi-channel inputs for learning a generalizable pathology classifier. By employing 40% data (fewer than 2500 labels) for training, we have demonstrated the generalizability of the proposed approach on two WCE datasets (two different capsule modalities) and provided a comprehensive benchmark for different pathologies within a single framework. Our results are better than the state-of-the-art in sensitivity for abnormality detection and fine-grained classification on several of nine different pathologies with much fewer labels and establish a new benchmark with a specificity higher than 97% across all pathology classes.

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