Y-short tandem repeat haplotype and paternal lineage of the Ezhava population of Kerala, south India

**Aim** To analyze the haplotype of the Ezhava population of Kerala, south India, using 8 short tandem repeat (STR) loci on the Y chromosome and trace the paternal genetic lineage of the population.

**Methods** Whole blood samples (n = 104) were collected from unrelated healthy men of the Ezhava population over a period of one year from October 2009. Genomic DNA was extracted by salting out method. All samples were genotyped for the 8 Y-STR loci by the AmpFiSTR Y-filer PCR Amplification Kit. The haplotype and allele frequencies were determined by direct counting and analyzed using Arlequin 3.1 software, and molecular variance was calculated with the Y-chromosome haplotype reference database online analysis tool, www.yhrd.org.

**Results** Among the 104 examined haplotypes, we found 98 unique ones. The average gene diversity was 0.669, with the highest diversity of 0.9462 observed for the bi-allelic Y-STR marker DYS 385. The allele frequency among DYS loci varied between 0.0096 and 0.75. Out of the 104 haplotypes, 10 were identical to the Jat Sikh population of Punjab, which is the greatest number among the Indian populations, and 4 to the Turkish population, which is the greatest number among the European populations. According to the allele frequency of Y-STR, the Ezhavas were genetically more similar to the Europeans (60%) than to the East Asians (40%).

**Conclusion** The vast majority of haplotypes were observed only once, reflecting the enormous genetic heterogeneity of the Ezhavas. Based on the genotype, the Ezhavas showed more resemblance to Jat Sikh population of Punjab and the Turkish populations than to the East Asians, hence indicating a paternal lineage of European origin.
Due to the geographical position of the Indian Peninsula between Africa, the Pacific, and west and east Eurasia, different populations have moved through its territory. This is why ethnic Indian population shows enormous cultural, linguistic, and genetic diversity. Indian tribal and caste populations derive heritage largely from the Pleistocene southern and western Asians, receiving limited gene flow from external regions since the Holocene (1). Also, Indian castes have been found to be more closely related to the Central Asians than to the Indian tribal groups (2).

The long seacoast of Kerala on the southern-most part of India has provided a gateway to India for many Asian, European, and Sri Lankan missionaries and traders. Non-tribal communities of Kerala, as shown by a human leukocyte antigen (HLA) analysis, were influenced by Dravidian, Indo-European, and East Asian gene pools (3). The Ezhava population of Kerala, according to the allele frequency distribution, had features of European, Central Asians, and East Asian gene pools. Mitochondrial DNA studies also validated the presence of two distinct, eastern and western Eurasian-specific lineage groups in India, suggesting that there were at least two separate migration events to India (4).

Due to the unique biology of the Y-chromosome, its genetic markers have been used in many forensic and evolutionary studies to determine patrilineal relationships within and between population groups. It has been suggested that, due to different distribution of region-specific allele frequencies, Y-short tandem repeats (STR) can be used to compare closely related populations (5,6). Previous genetic studies on the Ezhavas of south India failed to achieve a consensus on their paternal origin. In view of these diverse opinions based on HLA polymorphism and mitochondrial DNA analysis, this study aims to collect conclusive genetic data for a better understanding of the paternal origin of the Ezhavas. We present the haplotype analysis of the 8 Y-STR loci included in the European minimal haplotype set in 104 men from the Ezhava population to explore their genetic relationships with the European and East Asian populations. This is the first report on the Y-STR profile in Kerala population.

MATERIALS AND METHODS

Study samples

Whole blood samples (n = 104) were collected from unrelated healthy male individuals of the Ezhava population of Kerala over a period of one year from October 2009. The familial histories of the participants were recorded to exclude related individuals before sample collection. Blood samples were collected using standard procedures in ethylene diamine tetra acetic acid (EDTA)-coated tubes. The individuals gave their written informed consent and the ethical approval was received from the Ethics Committee of the institution.

DNA analysis

DNA was extracted from EDTA blood samples with the salting out method (7). All samples were genotyped for the 8 Y-STR loci (DYS19, DYS385, DYS389I, DYS389II, DYS390, DYS391, DYS392, and DYS393) by the AmpFiSTR Y-filer PCR Amplification Kit (Applied Biosystems, Foster City, CA, USA). The Y-STR data of Turkish, German, and North Indian Jat Sikh and other Indian populations were obtained from previous studies (8-10) and the Y-STR haplotype reference database (www.yhrd.org).

STATISTICAL ANALYSIS

Allelic and haplotype frequencies were estimated by direct counting. Gene diversities were calculated using Arlequin 3.1 software (11), according to the formula G.D = N (1-Σpi2) / N-1, where N is the population size and Pi is the allele frequency of the i-th haplotype. Furthermore, we used YHRD (www.yhrd.org) in order to determine the similarity of the Y-STR markers with the other publicly available population data. The Ezhava population was compared with other Indian populations and with selected world populations in order to investigate the pattern of paternal contributions. A total of 1890 samples were analyzed: German population sample with 685 haplotypes, Turkish population sample with 160 haplotypes, Ezhava population (our sample) with 104 haplotypes, Haryana Jat population sample with 91 haplotypes, Punjab Jat Sikh population with 108 haplotypes, Andhra Pradesh, India (Brahmin) with 109 haplotypes, Himachal Pradesh, India (Saraswat Brahmin) with 61 haplotypes, Jammu, India (Saraswat Brahmin) with 61 haplotypes, Jharkhand, India (Munda) with 68 haplotypes, Jharkhand, India (Sakaldwipi Brahmin) with 65 haplotypes, Madhya Pradesh, India (Kanyakubja Brahmin) with 78 haplotypes, Punjab, India (Balmiki) with 62 haplotypes, Tamil Nadu, India (Iyengar) with 67 haplotypes, and Tamil Nadu, India (Kuruman) with 67 haplotypes. Analysis of molecular variance (AMOVA) was used to establish the total variance among and within groups. AMOVA was done using fixation index analysis and with 10,000 permutations. From the AMOVA calculation, we obtained pairwise differ-
| ID | DYS | DYS | DYS | DYS | DYS | DYS | DYS | DYS | DYS | DYS | DYS |
|----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| E 01 | 15 | 13,16 | 12 | 27 | 24 | 10 | 11 | 12 |
| E 02 | 14 | 14,18 | 13 | 30 | 25 | 10 | 11 | 14 |
| E 04 | 15 | 16,17 | 14 | 31 | 22 | 10 | 11 | 12 |
| E 05 | 15 | 13,16 | 13 | 29 | 22 | 10 | 14 | 11 |
| E 06 | 15 | 14,19 | 13 | 28 | 23 | 10 | 10 | 14 |
| E 09 | 13 | 13,17 | 13 | 32 | 26 | 11 | 13 | 13 |
| E 11 | 16 | 11,14 | 13 | 30 | 25 | 11 | 11 | 13 |
| E 12 | 15 | 15,17 | 14 | 30 | 22 | 10 | 11 | 12 |
| E 13 | 15 | 15,17 | 14 | 30 | 22 | 10 | 11 | 12 |
| E 15 | 13 | 12,17 | 13 | 30 | 26 | 10 | 13 | 13 |
| E 16 | 15 | 11,19 | 13 | 31 | 22 | 9 | 10 | 14 |
| E 17 | 15 | 15,16 | 13 | 29 | 22 | 10 | 11 | 12 |
| E 18 | 14 | 13,15 | 12 | 28 | 22 | 11 | 14 | 11 |
| E 19 | 15 | 15,17 | 14 | 30 | 22 | 10 | 11 | 12 |
| E 20 | 14 | 14,17 | 14 | 31 | 24 | 10 | 11 | 14 |
| E 21 | 15 | 15,17 | 13 | 29 | 22 | 10 | 11 | 12 |
| E 23 | 15 | 14,17 | 13 | 31 | 24 | 10 | 11 | 14 |
| E 24 | 14 | 15,19 | 13 | 29 | 23 | 10 | 11 | 12 |
| E 25 | 15 | 12,14 | 14 | 32 | 25 | 10 | 11 | 13 |
| E 26 | 15 | 12,14 | 14 | 32 | 25 | 10 | 11 | 13 |
| E 27 | 17 | 13,19 | 14 | 31 | 23 | 10 | 10 | 14 |
| E 28 | 14 | 11,14 | 14 | 31 | 25 | 10 | 11 | 13 |
| E 29 | 17 | 14,15 | 13 | 29 | 21 | 10 | 11 | 13 |
| E 32 | 17 | 11,14 | 13 | 30 | 25 | 10 | 11 | 12 |
| E 33 | 15 | 15,16 | 13 | 29 | 22 | 10 | 11 | 12 |
| E 36 | 15 | 11,13 | 14 | 32 | 25 | 10 | 11 | 13 |
| E 37 | 16 | 11,12 | 13 | 30 | 24 | 11 | 11 | 13 |
| E 39 | 16 | 11,14 | 13 | 31 | 25 | 10 | 11 | 13 |
| E 41 | 14 | 13,17 | 12 | 28 | 22 | 11 | 14 | 11 |
| E 42 | 15 | 11,14 | 13 | 30 | 25 | 10 | 11 | 13 |
| E 43 | 14 | 11,19 | 13 | 29 | 23 | 10 | 10 | 14 |
| E 44 | 15 | 12,14 | 14 | 32 | 25 | 10 | 11 | 13 |
| E 46 | 15 | 15,17 | 14 | 30 | 22 | 10 | 11 | 12 |
| E 47 | 14 | 14,18 | 12 | 28 | 22 | 10 | 14 | 11 |
| E 49 | 17 | 11,14 | 14 | 30 | 25 | 10 | 11 | 13 |
| E 50 | 14 | 13,18 | 12 | 28 | 22 | 11 | 14 | 11 |
| E 51 | 14 | 13,17 | 14 | 31 | 22 | 10 | 14 | 12 |
| E 52 | 15 | 11,14 | 13 | 30 | 25 | 10 | 11 | 13 |
| E 54 | 15 | 13,16 | 12 | 29 | 23 | 10 | 14 | 14 |
| E 55 | 15 | 12,14 | 14 | 32 | 25 | 10 | 11 | 11 |
| E 56 | 14 | 11,14 | 14 | 31 | 25 | 10 | 11 | 13 |
| E 57 | 16 | 11,12 | 13 | 30 | 24 | 11 | 11 | 13 |
| E 59 | 15 | 13,16 | 12 | 27 | 24 | 10 | 11 | 12 |
| E 61 | 16 | 11,12 | 13 | 30 | 24 | 11 | 11 | 13 |
| E 62 | 14 | 14,18 | 12 | 28 | 22 | 10 | 14 | 11 |
| E 66 | 15 | 11,14 | 13 | 29 | 25 | 10 | 11 | 13 |
| E 67 | 14 | 14,17 | 14 | 32 | 25 | 10 | 11 | 14 |
| E 68 | 15 | 12,15 | 12 | 29 | 24 | 11 | 11 | 13 |
| E 69 | 15 | 14,19 | 13 | 30 | 22 | 10 | 11 | 12 |
| E 70 | 14 | 15,19 | 13 | 32 | 23 | 10 | 11 | 12 |
| E 71 | 16 | 11,15 | 13 | 31 | 26 | 10 | 11 | 13 |
| E 72 | 15 | 15,16 | 13 | 29 | 22 | 10 | 11 | 12 |

*E, R, X – Ezhava samples.

www.cmj.hr
ence and genetic distance between populations (multidi-

dimensional scaling analysis).

RESULTS

The haplotypes of 104 unrelated men from the Ezhava population of Kerala were investigated for 8 markers included in the European minimal haplotype set (Table 1). The majority of haplotypes were unique (98/104). While a larger number of individuals showed monoallelic condition at the DYS19, DYS389I, DYS389II, and DYS393 loci, a few samples (4/104) were unique, showing biallelic condition or duplication at the loci. A total of 12.5% (13/104) of the individuals showed monoallelic condition at the normally biallelic DYS385 locus. The variant allele DYS 385*17.1 was observed in 3 samples (Table 1).

Allele frequencies at DYS loci varied between 0.75 to 0.0096 (Table 2). Gene diversity ranged from 0.3875 to 0.9462 (average, 0.6671) and the most polymorphic single locus marker was DYS389II (Table 2).

The haplotypes of the Ezhavas were compared with the haplotypes of other Indian populations (Table 3). The pairwise difference analysis results were 0.0528 for Punjab Jat, 0.0691 for the Turkish population, 0.1024 for the Haryana.

### TABLE 2. Allele frequencies and gene diversity values at 8 DYS short tandem repeat loci of the Ezhava population of Kerala

| DYS Allele | 389 I | 390 | 389 II | 19 | 393 | 391 | 392 | Genotype | 385* |
|-----------|------|-----|--------|----|-----|-----|-----|---------|-----|
| 9         |      |     |        |    |     |     |     |         |     |
| 10        | 0.0192 |     | 7.18   | 0.0096 |
| 11        | 0.75  | 0.0865 | 10.14 | 0.0096 |
| 12        | 0.1154 | 0.2308 | 0.6827 | 10.18 | 0.0096 |
| 13        | 0.3462 |     | 0.0096 | 11.12 | 0.0481 |
| 14        | 0.0192 | 0.4038 | 0.0673 | 11.13 | 0.0288 |
| 15        | 0.3077 | 0.1346 | 0.1442 | 11.14 | 0.1731 |
| 16        | 0.4904 |     |         | 11.15 | 0.0288 |
| 17        | 0.1442 |     |         | 11.19 | 0.0288 |
| 18        | 0.0481 |     |         | 12.14 | 0.0385 |
| 19        |      |     |        |    |     |     |     |         |     |
| 20        | 0.0096 |     |         | 12.15 | 0.0096 |
| 21        | 0.3365 |     |         | 12.17 | 0.0096 |
| 22        | 0.1346 |     |         | 13.15 | 0.0096 |
| 23        | 0.2019 |     |         | 13.16 | 0.0577 |
| 24        | 0.2692 |     |         | 13.17 | 0.0481 |
| 25        | 0.1346 |     |         | 13.17 | 0.0096 |
| 26        | 0.0481 |     |         | 13.17 | 0.0096 |
| 27        | 0.0385 |     |         | 13.18 | 0.0096 |
| 28        | 0.1154 |     |         | 13.19 | 0.0192 |
| 29        | 0.3077 |     |         | 14.15 | 0.0096 |
| 30        | 0.2596 |     |         | 14.17 | 0.0192 |
| 31        | 0.1538 |     |         | 14.18 | 0.0673 |
| 32        | 0.125  |     |         | 14.19 | 0.0192 |

| Genotype | 15,16 | 0.0288 |
|-----------|-------|-------|
| 15        | 0.0769 |
| 16        | 0.0192 |
| 17        | 0.0385 |
| 18        | 0.0096 |

| Gene diversity value (GDV) | 0.6063 | 0.7602 | 0.7908 | 0.6475 | 0.6923 | 0.3875 | 0.5059 | 0.9462 |

*The allele frequencies for DYS385 were calculated for the combination of two alleles.
†Gene diversity value.
na Jat, and 0.1785 for Germans. These values show that Ezha population is distant from other Indian populations and close to the Punjab Jat and Turkish population, as well as the Jammu population (Saraswat Brahmin), with a pairwise distance value of 0.0561, and the Jharkhand population (Sakaldwipi Brahmin), with a value of 0.0617 (Figure 1).

Similarities in the Y-STR data were found between the Ezhas, north Indian, and Turkish population (Table 4). Out of the 104 haplotypes, 10 haplotypes were identical to Jat Sikhs, 7 to the German population, and 4 to the Turkish population. One particular haplotype was found to be common to all 4 populations, with a very high incidence (8/108) in the Jat Sikh population of north India.

Allelic distribution of the Y-STR markers of the Ezhas was compared with the European and East Asian populations (Figure 2). The histogram showed that 60% of the markers had similar allele frequencies to the European population, while only 40% showed similarity to the East Asian populations. The Y-STR alleles showed marked similarities to European populations when compared using European minimal haplotype.

**DISCUSSION**

This study found that the Ezhas showed more genotypic resemblance to the Jat Sikh population of Punjab and the Turkish populations than to East Asians, hence indicating a paternal lineage of European origin.

![Figure 1.](image)

**TABLE 3.** Analysis of molecular variance pair-wise distances based on the fixation index (Fst) values between the Kerala Ezha population and other Indian populations

| Population samples* | Kerala Ezha |
|----------------------|-------------|
| Jammu (Saraswat Brahmin) | 0.0573 |
| Jharkhand (Sakaldwipi Brahmin) | 0.0653 |
| Tamil Nadu (Iyengar) | 0.0671 |
| Himachal Pradesh (Saraswat Brahmin) | 0.0693 |
| Punjab (Balmiki) | 0.0713 |
| Tamil Nadu (Kuruman) | 0.0734 |
| Andhra Pradesh (Brahmin) | 0.0824 |
| Madhya Pradesh (Kanyakubja Brahmin) | 0.0840 |
| Jharkhand (Munda) | 0.1321 |

*Names in parenthesis refer to the YHRD designations (available from [http://www.yhrd.org](http://www.yhrd.org)).

**TABLE 4.** Comparison of haplotypes of Y chromosomal short tandem repeats in populations from Germany (G), north India (NI), Turkey (T), and Ezha population (KE) of South India

| Y-STR marker | G | NI | T | KE |
|--------------|---|----|---|----|
| DYS 19 | 15,16 | 13 | 29 | 22 | 10 | 11 | 12 | 1 | 3 |
| DYS 385 | 14,19 | 13 | 28 | 23 | 10 | 10 | 14 | 1 | 1 |
| DYS 389 I | 16 | 11,14 | 13 | 30 | 25 | 11 | 11 | 13 | 1 | 8 | 2 | 2 |
| DYS 389 II | 15 | 15,17 | 14 | 30 | 22 | 10 | 11 | 12 | 1 | 1 |
| DYS 390 | 14,18 | 12 | 28 | 22 | 10 | 14 | 11 | 1 | 2 |
| DYS 391 | 15 | 11,14 | 13 | 29 | 25 | 10 | 11 | 13 | 1 | 2 |
| DYS 392 | 15 | 15,17 | 14 | 30 | 22 | 10 | 11 | 12 | 1 | 1 |
| DYS 393 | 15 | 11,14 | 13 | 30 | 25 | 10 | 11 | 13 | 2 | 2 |

**Multi Dimensional Scaling plot showing the relationship between Kerala Ezha population with other Indian and European populations.**
To our knowledge, there are no published data about the genetic structure of Y-chromosome in Kerala population, although data on various other ethnic groups have been reported in the last decade (12-19). Y-STR haplotypes are useful for investigating and reconstructing the phylogeny of the more recently diverged paternal lineages (5,6), as well as for forensic/paternity testing (20-24). Based on HLA polymorphism analysis, Thomas et al (3) reported that the Ezhava population had features of European populations such as Belgian, German, and Scottish and that non-tribal communities of Kerala showed influences of Dravidian, Indo-European, and East Asian gene pools. Our study using Y-STR markers also confirmed their findings that the Ezhavas derived their paternal lineage from European and East Asian gene pools. Our study using Y-STR markers also confirmed their findings that the Ezhavas derived their paternal lineage from European and East Asian populations. There are also similar reports that placed Indians closer to European populations than to either East Asians or Africans in the genetic distance trees (25).

Duplications, though rare in the Ezhava population, have been observed in monoallelic markers like DYS19, DYS389I, DYS389II, and DYS393 loci in other populations (8,20,24). Duplications at DYS19 locus have been reported by previous studies in the Turkish populations (8). Except one report (26) of the variant allele DYS385 17.1 in Croatian population, the Ezhava population seem to be the only other population with variant DYS 385 17.1 allele.

The present study established the European paternal lineage of the Ezhavas, as well as contributed to the creation of a database of 8 Y-STR markers in the population. A limitation of the study is the non-availability of comparative Y-STR-based data for other Kerala communities. Also, the study was limited to a particular geographic area with the predominant Ezhava community. The sample size was limited to 104 participants as it was a pioneering study to establish the sample size for a study with a larger sample size and wider geographic coverage. Future studies are planned to create a database for 17 Y-STR markers in all the communities in Kerala population and to enhance the sample number and geographic area of the study.

We thank Prof K. Sasikumar for the conception of this study and the critical evaluation of the historical aspects of the manuscript. We thank all the DNA donors and to Rani. S. Jayamohan and student volunteer Sreejit Vallatham, Sreetha Sreenadhavan, Sruthi P Vijayaraghavan, Sumi Thomas, Surya Deva Ranjan, Swapna Chandran, Ashwarya B Chandran, Aruna G S, Akhila G Sathyan, Ambili Babu, and Nihaz Nizar of the Department of Biotechnology and Biochemical Engineering, SBCE, who gave us their assistance in sample collection.

Funding: Received from the Sree Buddha Educational Trust.
Ethical approval: Received from the Ethics Committee of the Sree Buddha College of Engineering.
Declaration of authorship: PSN contributed to designing the work, implementing, and manuscript preparation. AG contributed to sample collection and lab work. CJ contributed to sample collection and lab work.
Competing interests: All authors have completed the Unified Competing Interest form at www.icmje.org/coi_disclosure.pdf (available on request from the corresponding author) and declare: no support from any organization for the submitted work; no financial relationships with any organizations that might have an interest in the submitted work in the previous 3 years; no other relationships or activities that could appear to have influenced the submitted work.

References
1. Kivisild T, Rootsi S, Metspalu M, Mastana S, Kalmda K, Parik J, et al. The genetic heritage of the earliest settlers persists both in Indian tribal and caste populations. Am J Hum Genet. 2003;72:313-32. Medline:12536373 doi:10.1086/346068
2. Cordaux R, Auinger R, Bentley G, Nasidze I, Sirajuddin SM, Stoneking M. Independent origins of Indian caste and tribal paternal lineages. Curr Biol. 2004;14:231-5. Medline:14761656
3. Thomas R, Nair SB, Banerjee M. A crypto-Dravidian origin for the nontribal communities of South India based on human leukocyte antigen class I diversity. Tissue Antigens. 2006;68:225-34. Medline:16948643 doi:10.1111/j.1399-0039.2006.00652.x
4. Kivisild T, Kalmda K, Metspalu M, Parik J, Papisa S, Villes R. The place of the Indian mtDNA variants in the global network of maternal lineages and the peopling of the Old world. Genomic Diversity. 1999:135-52.
5. Roewer L, Kayser M, Dietjies R, Nagy M, Bakker E, Krawczak M, et al. Analysis of molecular variance (AMOVA) of Y-chromosome-specific microsatellites in two closely related human populations. Hum Mol Genet. 1996;5:1029-33. Medline:8817342 doi:10.1093/hmg/5.7.1029
6. Fabrico R. Santos, Chris Tyler-Smith. Reading the human Y chromosome: the emerging DNA markers and human genetic history. Brazilian Journal of Genetics. 1996;19:665-70. doi:10.1590/S0100-84519600000025

7. Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. Nucleic Acids Res. 1988;16:1215. Medline:3344216 doi:10.1093/nar/16.3.1215

8. Henke J, Henke L, Chattopadhyay P, Kayser M, Dülmer M, Cleef S, et al. Application of Y-chromosomal STR haplotypes to forensic genetics. Croat Med J. 2001;42:292-7. Medline:11387642

9. Yadav B, Raina R, Dogre TD. Haplotype diversity of 17 Y chromosomal STRs in Saraswat Brahmin Community. Forensic Sci Int Genet. 2011;5:e63-70.

10. Ghosh T, Kalpana D, Mukerjee S, Sharma AK, Nath S, et al. Genetic Diversity of 17 Y Short tandem repeats in Indian populations. Forensic Sci Int Genet. 2011 Jan 27. [Epub ahead of print] Medline:21277727 doi:10.1016/j.fsigen.2010.12.007

11. Excoffier L, Laval G, Schneider S. Arlequin (version 3.0): an integrated software for population genetics data analysis. Evol Bioinform Online. 2005;1:47-50. Medline:19325852

12. Asamurah H, Fujimori S, Ota M, Oki T, Fukushima H. Evaluation of mini-Y-STR multiplex PCR systems for extended 16 Y-STR loci. Int J Legal Med. 2008;122:43-9. Medline:17899155 doi:10.1007/s00414-007-0193-3

13. Butler JM, Schoske R, Vallone PM, Kline MC, Redd AJ, Hammer MF. A novel multiplex for simultaneous amplification of 20 Y chromosome STR markers. Forensic Sci Int. 2002;129:10-24. Medline:12230993 doi:10.1016/S0349-3706(02)00195-0

14. Dai HL, Wang XD, Li YB, Wu J, Zhang J, Zhang HJ, et al. Characterization and haplotype analysis of 10 novel Y-STR loci in Chinese Han population. Forensic Sci Int. 2004;145:47-55. Medline:15374594

15. Frank WE, Ralph HC, Tahir MA. Y chromosome STR haplotypes and allele frequencies in a southern Indian male population. J Forensic Sci. 2008;53:248-51. Medline:18279271 doi:10.1111/j.1556-4029.2007.00633.x

16. Iida R, Tsubota E, Sawazaki K, Masuyama M, Matsuji T, Yasuda T, et al. Characterization and haplotype analysis of the polymorphic Y-STRs DYS443, DYS444 and DYS445 in a Japanese population. Int J Legal Med. 2002;116:191-4. Medline:12111327 doi:10.1007/s00414-001-0266-7

17. Kayser M, Brauer S, Willuweit S, Schädlich H, Batzer MA, Zawacki J, et al. Online Y-chromosomal short tandem repeat haplotype reference database (YHRD) for U.S. populations. J Forensic Sci. 2002;47:513-9. Medline:12051329

18. Redd AJ, Agellon AB, Kearney VA, Contreras VA, Karafet T, Park H, et al. Forensic value of 14 novel STRs on the human Y chromosome. Forensic Sci Int. 2002;130:97-111. Medline:12477629 doi:10.1016/S0379-0738(02)000347-X

19. Schoske R, Vallone PM, Kline MC, Redd AJM. High-throughput Y-STR typing of U.S. populations with 27 regions of the Y chromosome using two multiplex PCR assays. Forensic Sci Int. 2004;139:107-21. Medline:15040904 doi:10.1016/j.fsigen.2003.10.007

20. Kayser M, Caglia A, Corach D, Fretwell N, Gehrig C, Graziosi G, et al. Evaluation of Y-chromosomal STRs: a multicenter study. Int J Legal Med. 1997;110:125-33, 41-9. Medline:9228563

21. Edwards A, Hammond HA, Jin L, Caskey CT, Chakraborty R. Genetic variation at five trimeric and tetrameric tandem repeat loci in four human population groups. Genomics. 1992;12:241-53. Medline:1704333 doi:10.1016/0888-7543(92)90371-X

22. Hammond HA, Jin L, Zhong Y, Caskey CT, Chakraborty R. Evaluation of 13 short tandem repeat loci for use in personal identification applications. Am J Hum Genet. 1994;55:175-89. Medline:7912887

23. de Knijff P, Kayser M, Caglia A, Corach D, Fretwell N, Gehrig C, et al. Chromosome Y microsatellites: population genetic and evolutionary aspects. Int J Legal Med. 1997;110:134-49. Medline:9228564 doi:10.1007/s004140050052

24. Kayser M, Brauer S, Weiss G, Schiefenhovel W, Underhill PA, Stoneking M. Independent histories of human Y chromosomes from Melanesia and Australia. Am J Hum Genet. 2001;68:173-90. Medline:11115381 doi:10.1086/316949

25. Cavalli-Sforza LL, Menozzi P, Piazza A. The history and geography of human genes. Princeton (NJ): Princeton University Press; 1994.

26. Grskovic B, Mrsic G, Polasek O, Vrdaljak A, Merkas S, Andelinkovic S. Population data for 17 short tandem repeat loci on Y chromosome in northern Croatia. Mol Biol Rep. 2011;38:2203-9. Medline:20859689 doi:10.1007/s11033-010-0349-y