Preparation and Characterization of an Extract of German Cockroach From a Korean Source

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Purpose: The cockroach (CR) is an important cause of respiratory allergic disorders. We prepared a German CR extract in a standardized way and analyzed its allergenic properties. Methods: The extract was prepared from German CR (Blattella germanica) obtained from a Korean colony, and its allergenic activity was compared with that of the commercial Hollister-Stier (HS) extract. The concentrations of Bla g 1 and Bla g 2 were measured, and an in vitro specific IgE binding inhibition assay was performed to assess IgE reactivity. Proteolytic activity was examined by gelatin zymography. Results: Bla g 1 and Bla g 2 were detected at 405 U/mg and 273 ng/mg, respectively, in the Korean extract, and at 187 U/mg and 56 ng/mg, respectively, in the HS extract. The Korean extract showed 94.2% inhibition of IgE reactivity, as compared with the HS extract. A similar pattern of IgE-reactive bands was detected for the two extracts, indicating that their allergenic components are similar. The proteolytic activities of the Korean and HS extracts were found to be similar in gelatin zymography. The endotoxin levels in the Korean and HS extracts were 3,440 EU/mL and 6,580 EU/mL, respectively. Conclusions: The German CR extract was prepared in a standardized way. The extract produced in this study will be useful for the development of allergy diagnostics and immunotherapeutic agents.

Key Words: Allergen; German cockroach; Korea; standardization

INTRODUCTION

Cockroach (CR) is an important source of inhalant allergens, and sensitization to CR is associated with asthma exacerbation.3 Moreover, epidemiologic data show a close relationship between CR sensitization and the prevalence of allergic asthma.2 Standardization of allergen extracts is important for diagnosis and immunotherapy.4 The German CR Blattella germanica is the most commonly found CR in Korean homes.4,5 However, CR extracts have not been standardized. The levels of protein and major allergens (Bla g 1 and Bla g 2) vary in the CR extracts that are commercially available in the USA.6 The concentration of CR extract supplied by the manufacturers is usually expressed in weight to volume (w/v) units. A designation of 1:10 w/v indicates that the solution contains the extractable material from 1 g of raw material added to 10 mL of buffer solution. The biological potencies of commercial German CR extracts have been estimated at 10-8570 bioequivalent allergy units (BAU)/mL in the USA.6,7 Moreover, protease activity is known to play an important role in the pathogenesis of CR allergy.8 CR extracts contain various proteases, which can degrade proteins, including allergens, in the extracts.9 Therefore, it is desirable to produce CR extracts that retain considerable protease activity without any significant degradation of the IgE-reactive components.

In the present study, we used a standardized method to produce extracts of German CRs, which were reared at the Korea National Arthropods of Medical Importance Resource Bank, Yonsei University College of Medicine, Seoul, Korea. The concentrations of the major allergens (Bla g 1 and Bla g 2) in the Korean extracts were compared with those of an extract obtained from a US company, i.e., the Hollister-Stier (HS) extract. The allergenic activities of these extracts were compared in an in vitro inhibition analysis.

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MATERIALS AND METHODS

Allergen extraction
Lyophilized CR (20 g) was pulverized using a mortar and pestle. The powdered CR (10 g) was defatted with ethyl ether (1:5, w/v). The allergen was extracted for 48 hours at 4°C in phosphate-buffered saline (PBS; pH 7.4) that contained 0.2% phenol. The extract was centrifuged at 13,000 ×g for 15 minutes at 4°C, and the supernatant was dialyzed (cut-off, 3,500 Da; Spectrum, Houston, TX, USA) extensively against distilled water. The dialyzed sample was filtered (0.22-μm pore; Millipore, Bedford, MA, USA) and lyophilized once again. The protein concentration was determined by the Bradford assay (Bio-Rad, Hercules, Hercules, CA, USA) after reconstitution in buffers. Thereafter, the extract was aliquoted, lyophilized, and stored at -80°C until use.

Protein analyses
The protein profiles of the CR extracts were examined by SDS-PAGE. Samples (10 μg), which were reconstituted in PBS (pH 7.4) that contained 50% glycerol and 0.03% human serum albumin (HSA), were run on 12.5% gels under reducing conditions. Proteins were visualized by staining with Coomassie Brilliant Blue R250 or transferred onto nitrocellulose membrane (Amersham, Buckinghamshire, UK). The membranes were incubated with 1:4 dilutions of sera (pooled serum from five patients or five healthy controls) after blocking with 3% skim milk in TBST (50 mM Tris [pH 7.5], 0.05% Tween-20). Subsequently, IgE-reactive proteins were probed with alkaline phosphatase-conjugated goat anti-human IgE (1:1,000; Sigma-Aldrich, St. Louis, MO, USA) for 1 hour at room temperature, and then placed in developing buffer (Promega, Madison, WI, USA) overnight at 4°C. The gel was incubated in renaturing buffer (Invitrogen) for 1 hour at room temperature, then placed in developing buffer (Invitrogen) overnight at 4°C. The gel was stained with Coomassie Brilliant Blue R250.

In vitro specific IgE binding inhibition assay
Allergen potencies were compared using the competitive specific IgE binding CAP inhibition system with the HS allergen extract (Korean) Laboratories, Spokane, WA, USA). The Korean extract dissolved in distilled water was used for the inhibition study. The serum samples (diluted 1:4) were pre-incubated with various concentrations of inhibitors (0.001 μg/mL to 20 μg/mL), and anti-human IgE reactivity was measured using the UniCAP system (Phadia, Uppsala, Sweden), according to the manufacturer’s instruction. The percentage of inhibition was calculated as: (1-Ai/Ao) × 1,000, where Ai is the IgE value (kU/mL) with inhibitor and Ao is the IgE value without inhibitor.

Assessments of proteolytic activity
For the analysis of gelatinolytic proteases, samples (200 ng/well), which were reconstituted in PBS (pH 7.4) that contained 50% glycerol and 0.03% HSA, were run on 10% SDS-PAGE gels that contained 0.1% gelatin (Invitrogen). After electrophoresis, the gel was incubated in renaturing buffer (Invitrogen) for 1 hour at room temperature, then placed in developing buffer (Invitrogen) overnight at 4°C. The gel was stained with Coomassie Brilliant Blue R250.

RESULTS

Allergenic activities of the CR extracts

The protein concentration of the Korean extract was very low (410 μg/mL) compared with that of the HS extract (2,300 μg/mL). However, the concentrations of Bla g 1 and Bla g 2 in the Korean extract (405 U/mg and 273 ng/mg, respectively) were higher than those in the HS extract (187 U/mg and 56 ng/mg, respectively) (Table).

The allergenic activities of the CR extracts were examined using an in vitro specific IgE binding inhibition assay. The Korean extract showed 94.2% inhibition of IgE reactivity, as compared with the HS extract (Fig. 2).

The Korean extract had more apparent bands in the SDS-PAGE analysis (Fig. 1A). The presence of a protein band close to the well in the HS extract lane implies some aggregation of the extract. No IgE-reactive bands were detected when sera from healthy controls were used for the immunoblotting (data not shown).

Protease and endotoxin activities of the CR extracts

A broad range of gelatinolytic activity was detected for the proteins in the range of 10-100 kDa in both the Korean and HS extracts (Fig. 1C). In both extracts, the primary proteolytic activ-

Table. Concentration of major allergen and endotoxin in German cockroach extracts

| Allergen extract | Protein (μg/mL) | Bla g 1 (U/mL) | Bla g 2 (ng/mL) | Endotoxin level (EU/mL) | Allergen potency |
|------------------|----------------|---------------|----------------|------------------------|-----------------|
| Korean           | 410            | 166           | 112            | 3,440                  | 94.2% of HS     |
| HS               | 2,300          | 429           | 129            | 6,580                  | 1:10 w/v        |

HS, Hollister-Stier.
The recognition sites on Bla g 2 for the monoclonal antibodies to be quite different (Table). Polymorphisms of mite allergen tracts using an Korea. Further characterization of the allergenicities of the extracts from HS and HS extract. (Fig. 1A). How, the HS extract had less-pronounced bands and some aggregation of proteins around the well. The overall patterns of IgE-reactive proteins in the extracts were similar for the Korean and HS extracts, with the exception of the aggregating proteins around the well. The overall patterns of IgE-reactive proteins in the allergen extracts were found to be non-proportional to the overall allergenicities. It is very interesting to observe strongly IgE-reactive proteins of approximately 60 kDa and 70 kDa in both the Korean and HS extracts (Fig. 1B). The observed similarity of the patterns of IgE-reactive bands in the immunoblots indicates the presence of similar IgE-reactive components in the extracts. Identification of these allergens, which are important in Korea, is necessary for enhanced characterization and standardization of the CR extract.

CR extract is rich in proteases, which can activate the various cell types that are important in allergic airway inflammation. Special care is needed, although not recommended, when combining fungal and CR extracts for immunotherapy, given the stability of the allergens in the mixtures.

High levels of endotoxin were detected in the CR extracts, as compared to house dust mite extracts (1-8,485 EU/mL). The average levels of endotoxin detected in standardized house dust mite extracts were: 4,619 EU/mL (range, 849-8,485 EU/mL) for seven Dermatophagoides farinae extracts; and 11 EU/mL (range, 1-34 EU/mL) for seven D. pteronyssinus extracts. However, up to 33,805 EU/mL of endotoxin was detected in the standardized cat pelt extract. CRs are known to carry various microorganisms, including bacteria. The endotoxin present in the CR extract is believed to be mainly derived from the flora or commensals in the CR gut.

The German CR extract prepared in the present study could be useful for the development of allergy diagnostics and immunotherapies.
Preparation of German Cockroach Extract

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