Osteoarthritis is Prevented in Rats by Verbascoside via Nuclear Factor kappa B (NF-κB) Pathway Downregulation

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Background: Cartilage degeneration during osteoarthritis (OA) most adversely affects the quality of life by hindering the movement. The present study investigated the role of verbascoside in the protection of cartilage degeneration induced by osteoarthritis.

Material/Methods: The enzyme-linked immunosorbent (ELISA) and western blot assays were used for determination of inflammatory cytokine secretion in serum and cartilage tissues, respectively.

Results: Treatment of the OA rats with verbascoside inhibited overproduction of interleukin (IL)-6, tumor necrosis factor (TNF)-α, and IL-1β in serum as well as cartilage tissues. The expression of P2X7R and matrix metalloproteinase (MMP)-13 was much higher in the rats induced with OA. However, administration of verbascoside reversed the OA-induced upregulation of P2X7R and MMP-13 expression in the cartilage tissues. The OA-mediated increase in substance P (SP) and prostaglandin E2 (PGE2) expression was also reduced in the cartilage tissues by the verbascoside treatment. Western blot assay revealed that verbascoside treatment markedly decreased the activation of IκBa and NF-κB p65 in the OA rats.

Conclusions: Thus, verbascoside inhibited inflammatory cytokine secretion in the OA rats by targeting P2X7R expression, production of matrix metalloproteinase, PGE2 and downregulation of NF-κB signaling pathway. Therefore, verbascoside may be used as potent agent for osteoarthritis treatment.

MeSH Keywords: Cytokines • Matrix Metalloproteinase 10 • Osteoarthritis

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Background

Osteoarthritis (OA) is detected most commonly in the people belonging to over 45 years of age group and is characterized by the degradation of cartilage at joints, inflammation, and osteophyte formation [1,2]. Studies for understanding the mechanism of osteoarthritis and development of treatment are being conducted continuously [3–7]. There is degradation of proteoglycan, breakdown of collagen network and finally erosion of the cartilage which is associated with the joint stiffness, pain and loss of movement [5–11]. The proteolysis and decomposition of different components of extra cellular matrix are catalyzed by the overproduction of inflammatory cytokines, interleukins, matrix metalloproteinases (MMPs) [11,12]. MMPs are the endopeptidases which contain zinc and depend on the calcium ions for their activity [6]. Therefore, suppression of MMPs is believed to play a vital role in OA treatment. The interleukin (IL)-1β acts as the main inducer of osteoarthritis by leading to chondrocyte metabolic imbalance through upregulation of catabolic enzymes [13,14]. The overproduction of catabolic enzymes interferes with the normal physiological functioning of the chondrocytes [13–15]. During OA chondrocytes secrete cytokines such as interleukin (IL)-1β and IL-6, prostaglandins (PGs) and tumor necrosis factor (TNF)-α in much higher level compared to normal chondrocytes [16]. The cytokines secreted by the synovial membrane penetrate and erode the cartilage tissues leading to OA development [17]. The prostaglandin E2 (PGE2), a member of PG family also catalyzes erosion of cartilage by promoting MMP production [18]. The tissue inflammation and pain in OA patients is regulated the expression of a neuropeptide known as substance P (SP) [19]. The expression of P2X7R has been found markedly higher in the macrophages and injured spinal cord tissues [20,21]. It has been found that P2X7R is an important target for rheumatoid arthritis treatment [22,23]. Although clinicians are making enormous efforts to develop an effective treatment strategy for OA, but no successful results have been obtained so far. Therefore, present study was designed to investigate the protective role of verbascoside against OA in the rat model. The study found that verbascoside prevented OA induced cartilage tissue damage through inhibition of cytokine production.

Material and Methods

Animals and ethics committee approval

The 60 male Wistar rats weighing 290–395 g (10–13 weeks old) were obtained from the Animal Center, Henan Province, laboratory, Zhengzhou, China. The rats were placed in wire-bottomed cages individually in the animal center under a constant temperature of 25±2°C and 50–60% humidity. The rats were provided access to tap water plus standard pellet diet and exposed to 12-hour light/dark cycles. The Ethics Committee of The Xi’an Hospital, Weiyang, Shaanxi Province, China approved the study. The experiments were conducted in accordance with the guidelines from National Institute of Health, China.

Osteoarthritis induction and treatment strategy

Osteoarthritis was induced in the rats by injecting 5 mg/kg solution of monosodium iodoacetate (MIA) in normal saline after anaesthetization with sodium 5% sorbitol through intra-articular route. The Wistar rats were divided into 6 groups containing 10 rats in each group; OA group, normal control, 5 mg/kg, 10 mg/kg, 15 mg/kg and 20 mg/kg verbascoside treatment groups. Among the 6 groups OA was established in 5 groups including OA group, 5 mg/kg, 10 mg/kg, 15 mg/kg, and 20 mg/kg treatment groups. The treatment groups were administered 5, 10, 15, and 20 mg/kg doses of verbascoside after 2 days of MIA injection by intraperitoneal route alternately for 2 weeks. The rats in the normal control and OA groups were administered equal volumes of normal saline alone.

Determination of cytokine levels in rat serum

The rats were sacrificed on day 28 of MIA injection after sodium sorbitol anaesthetization for determination of cytokine levels in serum. In brief, the blood samples of the rats were collected from the carotid artery and subjected to centrifugation for 25 minutes at 4000×g. The isolated supernatant was stored for measurement of IL-1β, IL-6, and TNF-α content at −80°C. The commercially available enzyme-linked immunosorbent assay (ELISA) kits were used for measurement of IL-1β, IL-6, and TNF-α in the rat serum in accordance with the manufacturer instructions.

Western blot analysis

The articular cartilage of the Wistar rats at the knee joint was excised carefully after sacrificing under sodium sorbitol anesthesia. The excised cartilage was cut into thin sections and then homogenized in the RIPA buffer [sodium chloride (150 Mm), NP-40 (1%), sodium deoxycholate (0.5%), SDS (0.2%), Tris-hydrochloride (50 mM at pH 8.0), EDTA (10 mM) and PMSF (1 mM; Sigma, Chemical, St. Louis, MO, USA) for 45 minutes at 4°C. The lysate was freed of the debris by centrifugation for 10 minutes at 12 000×g at 4°C. The concentration of proteins in the supernatant was measured by BCA protein assay kit (Beyotime Institute of Biotechnology, Nanjing, China) according to manual protocol. The protein samples (20–30 μg) were resolved by electrophoresis on 8–10% sodium dodecyl sulfate-polycrylamide gel (SDS-PAGE) and subsequently electroblotted onto polyvinylidene fluoride (PVDF) membranes. The membrane blocking was achieved by incubation with 3% non-fat milk at 37°C for 2.5 hours. The incubation of membranes with primary
antibodies was carried out for overnight at 4°C. The antibodies used were: anti-IL-6 (ab6672), anti-TNF-α (ab9635), anti-P2X7R (ab48871), anti-MMP-13 (ab39012), anti-SP (ab67006), anti-PGE2 (ab2318), anti-IL-1β (ab9787), anti-p-NF-κBp65 (ab86299), anti-NF-κBp65 (ab16502), anti-IκBα (sc-371), anti-p-IκBα (ab59195) and anti-GAPDH (G9545; all from Abcam, Cambridge, UK). Then incubation was carried out at room temperature with HRP-conjugated secondary antibodies for 2 hours. The enhanced chemiluminescence detection system was employed for visualization of the bands and Image-Pro Plus 6.0 software (Media Cybernetics, Inc., Rockville, MD, USA) was used for densitometry analysis.

Statistical analysis

The data expressed are the mean of ±standard deviation (SD) of triplicate experiments. The statistical analysis of the data was performed by one-way analysis of variance (ANOVA) and Bonferroni tests. The SPSS 13.0 software (SPSS, Inc., Chicago, IL, USA) was used for statistical analysis. Differences were taken significant statistically at P<0.05.

Results

Inhibitory effect of verbascoside on serum cytokine levels in OA-rats

In the serum of OA-rats the levels of inflammatory cytokines after verbascoside treatment were measured using ELISA assays (Figure 1). It was found that OA-induced secretion of IL-6, TNF-α and IL-1β in the serum of rats markedly suppressed than those of normal group. However, the OA-induced levels of IL-6, TNF-α, and IL-1β were reduced by verbascoside treatment significantly (P<0.05). Among the 5, 10, 15, and 20 mg/kg treatment groups, the OA-mediated secretion of IL-6, TNF-α, and IL-1β levels were completely inhibited on treatment with 20 mg/kg doses of verbascoside.

Verbascoside ameliorated OA-induced cytokines in the rat cartilage of knee tissues

The suppression of IL-6, TNF-α, and IL-1β levels by verbascoside in the knee cartilage of OA-rats was determined by western blot assay (Figure 2). The data showed that the levels of
Cytokines were markedly higher in the OA-rats than those of normal control rats. On the other hand, verbascoside treatment of the OA-rats ameliorated the increase of IL-6, TNF-α, and IL-1β levels. Although the reduction of OA-induced cytokines was observed in all the 4 treatment groups, the effect was maximum in 20 mg/kg treatment group.

**Inhibition of P2X7R and MMP-13 expression by verbascoside in OA-rats**

The western blot showed a markedly higher expression of P2X7R and MMP-13 in the cartilage tissues of the OA-rats. Treatment with 5, 10, 15, and 20 mg/kg doses of verbascoside reversed the OA-induced upregulation of P2X7R and MMP-13 expression. Again, the downregulatory effect of verbascoside on OA-mediated P2X7R and MMP-13 expressions was maximum at 20 mg/kg doses.

**Inhibition of SP and PGE2 expression by verbascoside in OA-rats**

It was found that expression of SP and PGE2 was markedly higher in the cartilage tissues of the OA-rats. Treatment with 5, 10, 15, and 20 mg/kg doses of verbascoside inhibited OA-induced SP and PGE2 upregulation.

**Inhibition of NF-κB signaling pathway by verbascoside in OA-rats**

The expression of p-IkBα and p-NF-κB p65 in the OA-rats was much higher in comparison to the normal control rats. Western blot assay showed that verbascoside treatment at 5, 10, 15 and 20 mg/kg doses markedly decreased the expression of p-IkBα and p-NF-κB p65. The OA-induced increase of p-IkBα and p-NF-κB p65 expression was decreased almost completely by verbascoside at 20 mg/kg doses.
Discussion

In the present study preventive role of verbascoside against osteoarthrits was investigated by measuring the changes in cytokines, P2X7R, MMP, PGE2 and NF-κB. The study demonstrated that verbascoside downregulated levels of cytokines, P2X7R, MMP and PGE2 through inhibition of NF-κB signaling pathway in OA-rats. Inflammation of joints is usually followed by the articular cartilage decomposition, subchondral bone degradation and hyperplasia of the synovial tissues in OA patients [24]. The OA induced joint inflammation is accompanied by neuronal activation which leads to acute pain and involves the chondrocytes [16]. During OA, the chondrocytes produce inflammatory cytokines in higher levels in comparison to those of the normal cases [17]. Studies have found overproduction of TNF-α, nitric oxide, and IL-8/6/α in the chondrocytes of OA patients [17]. In consistence with the earlier reports the present study found markedly higher levels of IL-6, TNF-α, and IL-1β in the serum of OA rats. The article cartilage tissues were also found to secrete higher levels of these inflammatory cytokines in the OA rats. However, verbascoside exhibited inhibitory effect on OA induced overproduction of IL-6, TNF-α, and IL-1β. Treatment of the rats with verbascoside reversed the up-regulatory effect of OA on IL-6, TNF-α, and IL-1β. Treatment of the rats with verbascoside reversed the up-regulatory effect of OA on IL-6, TNF-α, and IL-1β inflammatory cytokines. These findings suggested anti-osteoarthritic potential of verbascoside in the rat model. It is reported that production of inflammatory cytokines in the mouse mast cells is promoted by P2X7R activation [25]. The levels of IL6/1β and TNF-α was also markedly increased in the immune cells of rats by P2X7R activation [26]. Moreover, TNF-α blocking or silencing in different animal models has been found to inhibit inflammation [27–29]. The MMP-13 is secreted only in the cartilage of OA patients and is therefore considered to be indicator for the cartilage degradation [30–33]. There is chondrocyte apoptosis and decomposition of cartilage in the patients following MMP activation [32,34]. In the present study expression of P2X7R and MMP-13 in articular cartilage was markedly higher OA rats compared to the control group. However, verbascoside treatment inhibited the OA induced overexpression of P2X7R and MMP-13 in the cartilage tissues. This suggested that verbascoside inhibited inflammatory cytokine production in OA rats by downregulation of P2X7R expression.

The PGE2 catalyzes degradation of the cartilage in OA patients by promoting the generation of matrix metalloproteinase and aggrecanases [18]. In the present study the production of PGE2 was markedly higher in cartilage tissues of OA tissues. Treatment with verbascoside caused a marked reduction in the level of PGE2 in the cartilage tissues of rats with OA. It has been found that overproduction of IL-1β in the chondrocytes leads to activation of NF-κB pathway [35–37]. The NF-κB activation is also triggered by P2X7R which then leads to the development of various inflammatory diseases including osteoarthritis [38]. In the present study lxBα and NF-κB p65 phosphorylation was markedly promoted by OA induction in the rats. Treatment with verbascoside inhibited phosphorylation of lxBα and NF-κB p65 in the OA rat cartilage tissues.

Conclusions

In summary, the present study showed that verbascoside suppressed inflammatory cytokine secretion in the OA rats by targeting the expression of P2X7R, production of MMP, PGE2, and downregulation of NF-κB signaling pathway. Thus, verbascoside may be used as chemotherapeutic candidate for the treatment of osteoarthritis.

Conflict of interest

None.
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