Review Article

Russell Bodies and Russell Body Inflammatory Polyp in the Colorectum: A Review of Clinicopathologic Features

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Colorectal mucosa with Russell bodies is a reactive inflammatory lesion composed of mature plasma cells, known as Mott cells, which contain multiple intracytoplasmic eosinophilic globules. To the best of our knowledge, 3 case reports of colorectal Russell body containing lesions have been reported in the English literature (searched from 1980 to date), including just one case report of Colonic Russell body inflammatory polyp. Their importance lies in being aware of this unusual entity, recognizing it as well as the clinical scenario in which this typically arises and differentiating it from its malignant mimics that come in the histologic differential. This review discusses the clinical and endoscopic presentation, histopathologic features, ancillary studies, pathogenesis, differential diagnosis, prognosis, and treatment of this rare lesion.

1. Introduction

Russell bodies are eosinophilic spherical or globular cytoplasmic inclusions that accumulate in the rough endoplasmic reticulum of mature plasma cells. These plasma cells containing Russell bodies are also known as Mott cells [1]. Mott cells were first described in 1890 by William Russell [2]; however, the first case of Russell bodies in the gastrointestinal tract was in the stomach and was not described until 1998 by Tazawa and Tsutsumi [3]. Colorectal lesions containing Russell bodies are extremely rare or are probably under-reported with three case reports published in the English literature in a PubMed search from 1980 to date [4–6]. One of these three cases presented as a Colonic Russell body inflammatory polyp [4]. This review discusses the clinical and endoscopic features, histopathologic features, ancillary studies, pathogenesis, differential diagnosis, prognosis, and treatment of this rare entity.

2. Clinical and Endoscopic Features

Colorectal Russell body lesions are extremely rare. The patient characteristics and clinical and endoscopic features of the 3 cases of colorectal Russell body lesions reported in the literature to date are tabulated in Table 1. Of note, two of these presented as colorectal polyps (one patient with Colonic Russell body inflammatory polyp and the other one as tubulovillous adenoma with high-grade dysplasia with dense lamina propria plasma cell infiltrates containing Russell bodies and Mott cells), while one showed normal colonoscopy [4–6]. The only patient of Colonic Russell body inflammatory polyp (Table 1, case 1) reported in the literature had associated severe diverticulosis in the transverse colon, splenic flexure, descending colon, and sigmoid colon [4]. The patient with normal colonoscopy (Table 1, case 2) showed diffuse involvement by Russell bodies, associated with an immunocompromised state (status post kidney and pancreas transplant).

Though a common occurrence in the hematopathology specimens, Russell body containing lesions are unusual in the gastrointestinal (GI) tract with approximately 40 GI cases reported in the literature to date. In the GI tract, they are most commonly seen in the stomach as Russell body gastritis and have been associated with Helicobacter pylori infection in some studies; however this association
### Table 1: Case reports of Russell body containing Mott cells in the colorectum

| Case no. (Source) | Age, year/ Sex | Location | Clinical Presentation | Endoscopic findings | Histopathologic findings | Ancillary Studies |
|-------------------|---------------|----------|-----------------------|---------------------|--------------------------|-----------------|
| 1 (Coates et al., 2017 [4]) | 62/Male | Sigmoid Colon | Average risk screening colonoscopy, no GI complaints | <5 mm Erythematous polyp; severe diverticulosis | Inflammatory polyp with Russell bodies in the lamina propria. | CD138 and CD79a positive; *polyclonal* |
| 2. (Muthukumarana et al., 2015 [5]) | 44/Female | Colon | Diarrhea, abdominal pain, nausea and vomiting. Status post kidney and pancreas transplant with immunosuppression. | Normal colonic endoscopy; no colonic polyp | Chronic lymphoplasmacytic infiltrate with Mott cells within the lamina propria in colon, terminal ileum, duodenum and stomach | PAS positive; CD138 positive; *polyclonal* |
| 3 (Brink et al., 1999 [6]) | 53/Female | Rectum | Indication not provided. No evidence for malignant myeloma or a gammopathy. | Rectal polyp, not further specified | Tubulovillous adenoma with high-grade epithelial dysplasia and dense plasma cell infiltrates containing Russell bodies and Mott cells | IgG monoclonal for kappa light chain |

* polyclonal nature confirmed by concomitant expression of kappa and lambda light chain.

has not been definitively proven [7–16]. Other cases have been reported as Russell body duodenitis and Russell body esophagitis [7, 17–20]. The Russell body containing lesions in the GI tract span a wide age spectrum ranging from 24 to 88 years of age with a reported male to female ratio of 1.6:1 [5]. The clinical presentation of the upper GI Russell body containing lesions is nonspecific and varies from abdominal pain, nausea, dyspepsia, and diarrhea. Endoscopic features in the upper GI tract are again nonspecific and consist of mucosal erythematous changes, erosion/ulcers, edema, or rarely nodules [5].

### 3. Histopathologic Features

Histologically, Russell body inflammatory polyp typically exhibits features of inflammatory polyp with expansion of lamina propria by Mott cells. The surface epithelium can be intact and show hyperplastic changes or can be partially or totally eroded with the presence of ulcer and granulation tissue (Figures 1 and 2). The lamina propria in the colorectal Russell body lesions (including Russell body inflammatory polyp) shows expansion by inflammatory cells including variable composition of neutrophils, plasma cells, lymphocytes, and eosinophils. However, the most prominent population is of Mott cells, which are plasma cells showing eccentric nuclei and containing numerous intracytoplasmic eosinophilic globules also known as Russell bodies (Figure 1 inset) [3].

The histopathologic features of the 3 cases of colorectal Russell body containing lesions are tabulated in Table 1.

### 4. Ancillary Studies

The Russell bodies are periodic acid-Schiff positive. By immunohistochemistry, the Mott cells in Russell body inflammatory polyps are diffusely positive for plasma cell markers, CD138 (Figure 3) and CD79a, and are negative for pancytokeratins. Typically, the plasma cells are polyclonal as demonstrated by immunohistochemistry and/or in situ hybridization, for Kappa or Lambda light chain expression (Figures 4(a) and 4(b)). However, certain cases of monoclonality in GI tract Russell body containing lesions have been reported, without evidence of a neoplastic lesion [4–9, 20].

The ancillary studies in reported cases of colorectal Russell body containing lesions are tabulated in Table 1.

### 5. Pathogenesis

Russell body containing lesions in the colorectum is considered a rare vigorous form of inflammatory response to
chronic mucosal inflammation, possibly leading to over-stimulation of plasma cells and accumulation of numerous nondegradable immunoglobulins in cisternae of rough endoplasmic reticulum. During episodes of chronic inflammation, Mott cells can present as individual cells or clusters or sheets of cells in the lamina propria or accumulate to form aggregates known as Russell body inflammatory polyp, as seen in the patient of severe diverticulosis presenting with Russell body inflammatory polyp [4, 6]. However, why this response is seen in some subset of patients with diverticulosis or chronic inflammation and not all remains uncertain. It is also possible that these cases may be underreported in the literature.

Overall, majority of cases of Russell body gastroenteritis have been reported in upper GI tract, predominantly in the stomach, and have been associated with Helicobacter pylori infection in multiple studies [7–14]. Although Russell body proliferation is a benign entity, it has rarely been reported in association with gastric malignant epithelial neoplasms such as signet ring cell carcinoma, Epstein-Barr virus-associated gastric carcinoma, and gastric tubular adenocarcinoma [7, 21–23].

Figure 2: Medium power photomicrograph highlighting Russell body inflammatory polyp with surface epithelial erosion and expansion of lamina propria by Mott cells as well as other inflammatory cells (H&E X 200).

Figure 3: CD138 immunohistochemical stain highlights the Mott cells in colorectal Russell body containing lesion (CD138 X 100).

6. Differential Diagnosis

Though considered benign, the importance of Russell body inflammatory polyp or Russell body containing lesion lies in recognizing this unusual entity and differentiating it from its malignant mimics such as a signet ring cell carcinoma, which comes rarely in the differential [24, 25]. Unlike the former, the latter shows nuclear atypia and is positive for cytokeratin and mucicarmine. Russell body containing lesion lacks nuclear atypia, is negative for cytokeratins, and is positive for plasma cell markers. The other differential includes plasma cell neoplasms, such as mucosa associated lymphoid tissue (MALT) lymphoma with plasmacytic differentiation and plasmacytomas, and these typically exhibit cellular atypia and mitotic activity and are monoclonal, unlike most cases of Russell body containing lesions.

7. Prognosis and Treatment

Logically, Russell body inflammatory polyp or lesions can be treated by polypectomy as well as by treatment or reduction of the underlying cause of chronic inflammation. Russell body polyps of the upper GI tract have been seen in association with H. pylori gastritis, and some studies have shown that Russell body polyps occurring in the setting of H. pylori will be eliminated by the eradication of the concurrent infection [8–14].

8. Conclusions

Russell body inflammatory polyp and Russell body lesion with Mott cells are exceedingly rare in the colorectum and have only been described in the literature in the form of three case reports from 1980 to date. These are benign, reactive lesions most likely secondary to overproduction of immunoglobulins and development of Mott cells with Russell bodies as a response to chronic mucosal inflammation. The clinical significance lies in recognizing the benign and reactive nature of this entity, assessing the causative etiology, and differentiating it from its malignant mimics which come in the histologic differential.
Figure 4: Polyclonal plasma cells seen in Russell body inflammatory polyp as demonstrated by Kappa (a) and lambda (b) light chain expression (Kappa, Lambda x200).

Conflicts of Interest
The authors declare that they have no conflicts of interest.

References
[1] S. M. Hsu, P. L. Hsu, P. N. McMillan, and H. Fanger, “Russell bodies. A light and electron microscopic immunoperoxidase study,” American Journal of Clinical Pathology, vol. 77, no. 1, pp. 26–31, 1982.
[2] W. Russell, “An address on a characteristic organism of cancer,” British Medical Journal, vol. 2, pp. 1358–1360, 1890.
[3] K. Tazawa and Y. Tsutsumi, “Localized accumulation of Russell body-containing plasma cells in gastric mucosa with Helicobacter pylori infection: Russell body gastritis,” Pathology International, vol. 48, no. 3, pp. 242–244, 1998.
[4] R. F. Coates, N. Ferrentino, and M. X. Yang, “Russell Body Inflammatory Polyp: A Case Report and Review of Literature,” International Journal of Surgical Pathology, vol. 25, no. 1, pp. 94–96, 2017.
[5] V. Muthukumarana, S. Segura, M. O’Brien, R. Siddiqui, and H. El-Fanek, “Russell Body Gastroenterocolitis” in a Posttransplant Patient,” International Journal of Surgical Pathology, vol. 23, no. 8, pp. 667–672, 2015.
[6] T. Brink, B. Wagner, and J.-O. Gebbers, “Monoclonal plasma and Mott cells in a rectal adenoma [3],” Histopathology, vol. 34, no. 1, pp. 81-82, 1999.
[7] F. Bhajee, K. A. Brown, B. W. Long, and A. S. Brown, “Russell Body Gastroenteritis: An Aberrant Manifestation of Chronic Inflammation in Gastrointestinal Mucosa,” Case Reports in Medicine, vol. 2013, Article ID 797264, 5 pages, 2013.
[8] J. D. Coyne and B. Azadkh, “Russell body gastritis: A case report,” International Journal of Surgical Pathology, vol. 20, no. 1, pp. 69-70, 2012.
[9] P. Karabagli and H. S. Gokturk, “Russell body gastritis: Case report and review of the literature,” Journal of Gastrointestinal and Liver Diseases, vol. 21, no. 1, pp. 97–100, 2012.
[10] S. W. Eum, J. H. Lee, and K. Y. Kim, “A case of Russell body gastritis associated with Helicobacter pylori infection,” Korean Journal of Gastrointestinal Endoscopy, vol. 35, pp. 181–185, 2007.
[11] S. Paik, S.-H. Kim, J.-H. Kim, W. I. Yang, and Y. C. Lee, “Russell body gastritis associated with Helicobacter pylori infection: A case report,” Journal of Clinical Pathology, vol. 59, no. 12, pp. 1316–1319, 2006.
[12] A. Ensari, B. Savas, A. O. Heper, I. Kuzu, and R. Idilman, “An unusual presentation of helicobacter pylori infection: So-called "Russell body gastritis",” Virchows Archiv, vol. 446, no. 4, pp. 463–466, 2005.
[13] S. Pizzolitto, D. Camilot, G. DeMaglio, and G. Falconieri, “Russell body gastritis: Expanding the spectrum of Helicobacter pylori - Related diseases? Pathology - Research and Practice, vol. 203, no. 6, pp. 457–460, 2007.
[14] J. B. Yoon, T. Yeong Lee, J. Sook Lee et al., “Two cases of russell body gastritis treated by Helicobacter pylori eradication,” Clinical Endoscopy, vol. 45, no. 4, pp. 412–416, 2012.
[15] A. del Gobbo, L. Elli, P. Braidotti, F. di Nuovo, S. Bosari, and S. Romagnoli, “Helicobacter pylori-negative russell body gastritis: Case report,” World Journal of Gastroenterology, vol. 17, no. 9, pp. 1234–1236, 2011.
[16] R. Drut and A. B. Olenchuk, “Russell body gastritis in an HIV-positive patient,” International Journal of Surgical Pathology, vol. 14, no. 2, pp. 141-142, 2006.
[17] C. A. Rubio, “Mott cell (Russell bodies) Barrett’s oesophagitis,” In Vivo, vol. 19, no. 6, pp. 1097–1100, 2005.
[18] N. M. Savage, T. Fortson, M. Schubert, S. Chamberlain, J. Lee, and P. Ramalingam, “Isolated Russell body duodenitis,” Digestive Diseases and Sciences, vol. 56, no. 7, pp. 2202–2204, 2011.
[19] A. Paniz Mondolfi, M. Samuel, J. Kikhney et al., “Russell body duodenitis: A histopathological and molecular approach to a rare clinical entity,” Pathology - Research and Practice, vol. 208, no. 7, pp. 415–419, 2012.
restriction,” *Clinics and Research in Hepatology and Gastroenterology*, vol. 38, no. 5, pp. e89–e97, 2014.

[21] E.-M. Wolf, K. Mrak, J. Tschmelitsch, and C. Langner, “Signet ring cell cancer in a patient with Russell body gastritis - a possible diagnostic pitfall,” *Histopathology*, vol. 58, no. 7, pp. 1178–1180, 2011.

[22] J. Choi, H. Eun Lee, S.-J. Byeon, K. H. Nam, M. A. Kim, and W. H. Kim, “Russell body gastritis presented as a colliding lesion with a gastric adenocarcinoma: A case report,” *Basic and Applied Pathology*, vol. 5, no. 2, pp. 54–57, 2012.

[23] A. Shinozaki, T. Ushiku, and M. Fukayama, “Prominent Mott cell proliferation in Epstein-Barr virus-associated gastric carcinoma,” *Human Pathology*, vol. 41, no. 1, pp. 134–138, 2010.

[24] E. S. Yu, Y. I. Kim, C. W. Kim et al., “Russell body: containing plasma cell aggregations mimicking signet ring cell carcinoma of the stomach,” *Korean Journal of Gastrointestinal Endoscopy*, vol. 7, pp. 39–41, 1987.

[25] A. Erbersdobler, S. Petri, and G. Lock, “Russell body gastritis: An unusual, tumor-like lesion of the gastric mucosa,” *Archives of Pathology & Laboratory Medicine*, vol. 128, no. 8, pp. 915–917, 2004.