Clinical Study

Ceruloplasmin and the Extent of Heart Failure in Ischemic and Nonischemic Cardiomyopathy Patients

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Objective. Ceruloplasmin was elevated in patients with coronary heart disease, but the relationship between ceruloplasmin and heart failure was still unknown. We aimed to evaluate ceruloplasmin in heart failure patients and assess association between ceruloplasmin and the extent of heart failure.

Methods and Results. 202 heart failure patients were divided into ischemic (78 with coronary stenosis) and nonischemic groups (124 without coronary stenosis). 94 subjects without heart failure were included as controls. The extent of heart failure was defined according to NYHA classification. Ceruloplasmin levels in ischemic (𝑃< 0.001) and nonischemic groups (𝑃< 0.001) were higher than those in control group. Ceruloplasmin had a positive linear correlation with C-reactive protein (𝑃< 0.01) and a negative linear correlation with LVEF (𝑃< 0.05). In nonischemic group, CP levels were significantly different among different NYHA subgroups (𝑃< 0.05). The correlation between ceruloplasmin and extent of heart failure was calculated by binary logistic regression. Ceruloplasmin showed an independent association with the extent of heart failure in nonischemic cardiomyopathy patients (𝑃< 0.05). Conclusions. Ceruloplasmin was significantly elevated in patients with ischemic or nonischemic cardiomyopathy and had linear correlation with C-reactive protein and LVEF. In nonischemic cardiomyopathy patients, the ceruloplasmin value was an independent biomarker associated with the extent of heart failure.

1. Introduction

During the progression of cardiomyopathy, heart failure was gradually aggravated and became the major cause of unplanned hospitalization. The mortality of heart failure is still high in the present therapy, and brain natriuretic peptide, which is elevated in acute heart failure, could be false positive in pulmonary disease, like pulmonary embolism or chronic obstructive pulmonary disease.

Metal metabolism, metalloenzymes, and their activities of oxidation and inflammation are involved in progression of heart failure [1]. Many studies have shown that ceruloplasmin (CP) levels were elevated in patients with cardiovascular disorders including arteriosclerosis [2], coronary heart disease [3, 4], and myocardial infarction [5]. In the ischemic heart failure, CP is recognized as an inflammation-related marker protein for its predictive value [6]. Elsherif et al. [7] verified that copper deficiency leads to cardiac dysfunction.

Correale et al. [8] demonstrated that the CP value is a significant marker of acute heart failure in patients with ST segment elevated myocardial infarction. Serum CP might be an independent risk factor for ischemic cardiac disease [9]. However, several studies still suspected the association [10], and the relationship between CP and heart failure was still unclear in either ischemic or nonischemic cardiomyopathy.

Our study aimed to evaluate the predictive value of serum ceruloplasmin in patients with ischemic and nonischemic cardiomyopathies, and assess the association between CP levels and the extent of heart failure, CP levels and other parameters in both ischemic and nonischemic cardiomyopathy patients.

2. Material and Methods

2.1. Patient Recruitment. Between December 2009 and April 2011, a total of 202 patients were recruited consecutively from
2.2. Clinical Data Collection. For each patient, the clinical characteristics were collected, including age, gender, height, weight, history of hypertension, diabetes mellitus, cigarette smoking, and NYHA classification. Medications taken (cardiogenic, diuretic, nitrate, angiotensin-converting enzyme inhibitor/angiotensin receptor blocker, β-blocker, calcium channel blocker, aspirin/clopidogrel, and statin) were also collected. BMI (body mass index) is defined as the individual's weight (kg) divided by square of height (m).

Echocardiography was performed for each subject and parameters were selected for analysis: LVEF (%)—left-ventricular ejection fraction, LVIDd (cm)—left-ventricular internal diameter at end diastole, and LA (cm)—left atrium.

2.3. Blood Sample Analysis. Blood samples were collected on admission, using pyrogen-free tubes containing EDTA for plasma test and sterile tubes containing gel for serum separation (BD, USA) and then immediately centrifuged at 4000 rpm for 10 min at 4°C. All the samples were analyzed in laboratory of Second Affiliated Hospital of Zhejiang University College of Medicine. The results of blood biochemical test including total cholesterol, low-density lipoprotein cholesterol, high-density lipoprotein cholesterol, triglycerides, glucose, creatinine, C-reactive protein, troponin I, creatine kinase-MB, and CP were collected and analyzed.

The levels of serum ceruloplasmin were measured by nephelometry [11] (Beckman Coulter, Immage 800, USA).

2.4. Statistical Analysis. Continuous variables were given as mean ± standard deviation. Categorical variables were defined as percentage. Data were tested for normal distribution using the Kolmogorov-Smirnov test. Student’s t-test and one-way analysis of variance were used for continuous variables and the χ² test for the categorical variables. The correlation between ceruloplasmin and other parameters was evaluated by Pearson correlation. The NYHA II and III subgroups were combined as mild group, NYHA IV was the severe group, and then association between the clinical parameters and the extent of heart failure was assessed by binary logistic regression. All tests of significance were two tailed. Statistical significance was defined as P value less than 0.05.

3. Results

3.1. Patient Enrollment and Group Characterization. 78 heart failure patients with coronary stenosis and 124 heart failure patients without coronary stenosis were included in ischemic and nonischemic groups, respectively. 94 subjects without heart failure were included as controls. Epidemiological characteristics and biochemical parameters of the patients according to the three groups are described in Table 1. When compared with the control group, both the ischemic and nonischemic cardiomyopathy groups were older, had higher levels of creatinine, uric acid, C-reactive protein, homocysteic acid, LVIDd, and LA, and had lower levels of high-density lipoprotein cholesterol and LVEF. Prevalence of hypertension, diabetes and usage of cardiovascular medication were also higher in both ischemic and nonischemic cardiomyopathy groups than in control group. When compared with ischemic cardiomyopathy group, the nonischemic cardiomyopathy had higher levels of uric acid, LVIDd, LA, higher prevalence of diabetes and smoking, and lower levels of triglycerides and LVEF. The nonischemic group still has higher levels of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) than ischemic and control groups.

To assess the relationship between ceruloplasmin and other parameters, the Pearson linear correlation was evaluated in all the subjects in Table 2. The ceruloplasmin had a significant positive linear correlation with ALT, AST, uric acid, C-reactive protein, LVIDd, and LA (P < 0.01). The ceruloplasmin also had a negative linear correlation with LVEF (P < 0.05).

3.2. Ceruloplasmin and the Extent of Heart Failure. Mean CP levels were 250.01 ± 47.54, 313.06 ± 73.60, and 328.80 ± 98.91 mg/L, for patients in the control, ischemic cardiomyopathy, and nonischemic cardiomyopathy groups, respectively. CP levels in ischemic (P < 0.001) and nonischemic cardiomyopathy groups (P < 0.001) were higher than those in the control group (Figure 1), whereas there was no significant difference between the ischemic and nonischemic groups. When CP levels were compared among the different NYHA groups in patients with ischemic cardiomyopathy, there was
no significant difference among three groups (308.69 ± 74.89, 308.68 ± 68.36, and 363.14 ± 84.65, P > 0.05) (Figure 2). In nonischemic cardiomyopathy patients, CP levels were significantly different among NYHA II, NYHA III, and NYHA IV groups (291.81 ± 56.28, 318.54 ± 91.98, and 318.54 ± 91.98, P < 0.05) (Figure 2).

3.3. Ceruloplasmin and the Extent of Heart Failure in Nonischemic Group. According to results previous mentioned, we tried to identify the relation between ceruloplasmin and the extent of heart failure in nonischemic group. All the related parameters were compared, and the results showed that AST, uric acid, CKMB, CRP, and LVEF were significantly different among NYHA II, NYHA III, and NYHA IV groups (Table 3). Because these parameters could not be balanced among groups, we combined the NYHA II and NYHA III patients as mild group, and then binary logistic regression model was applied between mild group and NYHA IV group. The results showed that only CP levels (χ² = 4.489, odds ratio (OR) = 1.010, 95% confidence interval (CI): 1.001–1.019, P = 0.034) and age (P = 0.046) were correlated with the extent of heart failure. The correlation between the CP levels and the extent of heart failure was independent of gender, smoking, alcohol taking, hypertension, diabetes mellitus, AST, uric acid, CKMB, CRP, LVEF, and other parameters (Table 4).

4. Discussion

Ceruloplasmin (CP) is a 132-kDa plasma glycoprotein which binds 95% of the total circulating copper in healthy adults.

Table 1: Baseline characteristics of the study population between groups.

| n (296) | Control (94) | Ischemic cardiomyopathy (78) | Nonischemic cardiomyopathy (124) |
|--------|--------------|------------------------------|---------------------------------|
| Age (year) | 43.67 ± 15.80 | 69.97 ± 11.05* | 66.30 ± 15.23* |
| Male sex [n (%)] | 43 (45.7%) | 40 (51.3%) | 65 (52.4%) |
| Smoking [n (%)] | 19 (20.2%) | 33 (42.3%) | 34 (27.4%) |
| Alcohol [n (%)] | 10 (10.6%) | 16 (20.5%) | 21 (16.9%) |
| Hypertension [n (%)] | 18 (19.1%) | 51 (65.4%) | 65 (52.4%) |
| Diabetes mellitus [n (%)] | 0 (0%) | 21 (26.9%) | 18 (14.5%) |
| BMI (kg/m²) | 22.35 ± 2.77 | 22.32 ± 4.52 | 21.09 ± 5.71 |
| ALT (U/L) | 18.82 ± 10.93 | 20.30 ± 11.80 | 31.33 ± 50.15* |
| AST (U/L) | 21.63 ± 8.20 | 26.17 ± 22.98 | 37.19 ± 56.13* |
| Total cholesterol (mg/dL) | 170.78 ± 37.68 | 158.37 ± 44.85 | 155.53 ± 41.18* |
| LDL-C (mg/dL) | 100.47 ± 28.92 | 93.92 ± 35.86 | 90.84 ± 31.14* |
| Triglycerides (mg/dL) | 114.96 ± 64.85 | 134.37 ± 76.33 | 111.74 ± 59.79* |
| Creatinine (mg/dL) | 0.72 ± 0.19 | 0.94 ± 0.52* | 1.01 ± 0.78* |
| Uric acid (mg/dL) | 5.33 ± 1.47 | 6.22 ± 2.22* | 6.96 ± 2.53* |
| CK (IU/l) | 74.78 ± 32.33 | 100.35 ± 186.27 | 100.14 ± 127.43 |
| CK-MB (IU/l) | 10.60 ± 3.34 | 15.61 ± 27.07 | 13.66 ± 6.91 |
| TNI (ng/mL) | 0.06 ± 0.21 | 0.66 ± 3.75 | 0.28 ± 1.79 |
| CRP (mg/L) | 3.85 ± 7.99 | 14.77 ± 24.84* | 13.80 ± 27.27* |
| Homocysteic acid (μmol/L) | 12.25 ± 5.49 | 16.66 ± 7.15* | 17.12 ± 10.22* |
| LVEF (%) | 63.96 ± 4.85 | 53.29 ± 14.36 | 53.29 ± 14.36 |
| LVIDd (cm) | 4.55 ± 0.42 | 5.19 ± 0.95* | 5.60 ± 1.11 |
| LA (cm) | 3.09 ± 0.38 | 3.93 ± 0.67* | 4.47 ± 1.01* |
| Cardiac tonic [n (%)] | 0 (0%) | 13 (16.7%) | 68 (54.8%) |
| Diuretic [n (%)] | 0 (0%) | 37 (47.4%) | 100 (80.6%) |
| Nitrate [n (%)] | 0 (0%) | 56 (71.8%) | 83 (66.9%) |
| ACEI/ARB [n (%)] | 9 (9.6%) | 72 (92.3%) | 105 (84.7%) |
| Beta blockers [n (%)] | 9 (9.6%) | 56 (71.8%) | 83 (66.9%) |
| CCB [n (%)] | 11 (11.7%) | 29 (37.2%) | 30 (24.2%) |
| Aspirin/clopidogrel [n (%)] | 4 (4.3%) | 74 (94.9%) | 71 (57.3%) |
| Statins [n (%)] | 2 (2.1%) | 72 (92.3%) | 23 (18.5%) |

ACEI: angiotensin-converting enzyme inhibitor; ALT: alanine aminotransferase; ARB: angiotensin receptor blocker; AST: aspartate aminotransferase; BMI: body mass index; CCB: calcium channel blocker; CK: creatinine kinase; CK-MB: creatinine kinase-MB; CRP: C-reactive protein; HDL-C: high-density lipoprotein cholesterol; LA: left atrium; LDL-C: low-density lipoprotein cholesterol; LVEF: left-ventricular ejection fraction; LVIDd: left-ventricular internal diameter at end diastole; TNI: troponin I.

*P < 0.05 versus control group.
†P < 0.05 versus ischemic cardiomyopathy group.
Table 2: Linear correlation between ceruloplasmin and other parameters.

| Parameter          | Pearson correlation | P value |
|--------------------|--------------------|---------|
| Creatinine         | −0.028             | 0.635   |
| ALT                | 0.229              | <0.001* |
| AST                | 0.286              | <0.001* |
| LDL-c              | −0.047             | 0.428   |
| Total cholesterol  | −0.083             | 0.160   |
| Triglycerides      | −0.061             | 0.301   |
| Uric acid          | 0.249              | <0.001* |
| CK                 | 0.046              | 0.435   |
| CK-MB              | 0.037              | 0.553   |
| Troponin I         | 0.006              | 0.926   |
| CRP                | 0.449              | <0.001* |
| Homocysteic acid   | 0.094              | 0.107   |
| LVEF               | −0.151             | 0.032*  |
| LVIDd              | 0.213              | 0.003*  |
| LA                 | 0.215              | 0.003*  |

ALT: alanine aminotransferase; AST: aspartate aminotransferase; CK: creatine kinase; CK-MB: creatine kinase-MB; CRP: C-reactive protein; LA: left atrium; LDL-C: low-density lipoprotein cholesterol; LVEF: left-ventricular ejection fraction; LVIDd: left-ventricular internal diameter at end diastole. *P < 0.05.

**Figure 1:** Ceruloplasmin levels in control, ischemic, and nonischemic cardiomyopathy groups. CP levels in ischemic and nonischemic cardiomyopathy groups were higher than those in the control group (P < 0.001). No difference was found between the ischemic and nonischemic cardiomyopathy groups. *p < 0.001 versus control group.

The liver is the major source of serum CP in adults [13]. The synthesis of serum CP in hepatic cells is increased by proinflammatory agonists [14–16]. The recent research has indicated that increased ceruloplasmin concentrations were associated with an increased risk of myocardial infarction and stroke [17, 18]. To the best of our knowledge, this is the first study on the relationship between CP levels and the extent of heart failure, which indicated that the CP levels were significantly high in patients with ischemic or nonischemic cardiomyopathy. In nonischemic cardiomyopathy patients, the CP value was an independent biomarker associated with the extent of heart failure.

During the progression of heart failure, the inflammatory and oxidative reactions were activated and enhanced as a protective mechanism. So we hypothesized that CP, which was an index of the metal metabolism, still could be the inflammatory and oxidative biomarker in heart failure. The results proved our hypothesis. The CP level was elevated in patients with cardiomyopathy and showed the positive linear correlation with uric acid and C-reactive protein and negative linear correlation with left-ventricular ejection fraction, which meant that CP was increased with other acute-phase biomarkers when heart function got worse. Then we compared the CP levels of different NYHA levels in ischemic and nonischemic cardiomyopathy groups, respectively.

In ischemic heart disease, ischemic injury to cardiac muscle induced the inflammation and redox toxic reactions and the injury ran through every stage of ischemic cardiomyopathy. Copper might be involved in the redox toxic reaction [19]. CP is elevated by proinflammatory agonists to play a role of cytoprotection [20] and should be elevated when heart function was aggravated in our hypothesis. However, when we compared the CP value among different NYHA levels in ischemic cardiomyopathy group, no statistical difference was found. We believed that it was related to the small number of patients included, because only 7 patients were included in NYHA IV subgroup and the mean CP value in NYHA IV patients was much higher than that in other subgroups.

**Figure 2:** Ceruloplasmin levels in NYHA II, NYHA III, and NYHA IV subgroups. There was no significant difference among different NYHA subgroups in ischemic cardiomyopathy patients (P > 0.05). In nonischemic cardiomyopathy patients, the CP levels showed statistical difference among NYHA II, NYHA III, and NYHA IV subgroups (P < 0.05).
### Table 3: Baseline characteristics of patients with nonischemic cardiomyopathy.

|                      | NYHA II (21) | NYHA III (71) | NYHA IV (32) | P value |
|----------------------|--------------|---------------|--------------|---------|
| Age (year)           | 68.52 ± 16.23| 63.69 ± 15.49 | 70.00 ± 13.32| 0.110   |
| Male sex [n (%)]     | 10 (47.6%)   | 42 (59.2%)    | 13 (40.6%)   | 0.195   |
| Smoking [n (%)]      | 5 (23.8%)    | 24 (33.8%)    | 6 (18.8%)    | 0.258   |
| Alcohol [n (%)]      | 4 (19.0%)    | 15 (21.1%)    | 3 (9.4%)     | 0.347   |
| Hypertension [n (%)] | 12 (57.1%)   | 34 (47.9%)    | 19 (59.4%)   | 0.498   |
| Diabetes mellitus [n (%)] | 4 (19.0%) | 7 (9.9%)     | 7 (21.9%)    | 0.225   |
| BMI (kg/m²)          | 21.56 ± 3.11 | 21.29 ± 5.68 | 20.12 ± 7.41 | 0.639   |
| ALT (U/L)            | 17.33 ± 12.50| 27.60 ± 26.02| 48.69 ± 88.41| 0.052   |
| AST (U/L)            | 23.14 ± 6.98 | 31.14 ± 22.77| 59.84 ± 102.72| 0.024*  |
| Total cholesterol (mg/dL) | 165.81 ± 50.27| 154.94 ± 39.12| 151.65 ± 39.59| 0.460   |
| LDL-C (mg/dL)        | 94.52 ± 40.68| 90.74 ± 28.34| 89.23 ± 30.50| 0.832   |
| Triglycerides (mg/dL) | 123.05 ± 77.44| 112.51 ± 60.00| 104.13 ± 44.42| 0.537   |
| Creatinine (mg/dL)   | 0.83 ± 0.25   | 0.95 ± 0.40   | 1.01 ± 0.47  | 0.292   |
| Uric acid (mg/dL)    | 5.92 ± 1.82   | 6.96 ± 2.04   | 7.71 ± 3.55  | 0.044*  |
| CK (IU/L)            | 93.70 ± 84.26| 90.88 ± 135.05| 124.97 ± 133.91| 0.451   |
| CK-MB (IU/L)         | 10.26 ± 3.72  | 13.05 ± 6.50  | 17.17 ± 7.89 | 0.001*  |
| Troponin I (ng/mL)   | 0.02 ± 0.01   | 0.13 ± 0.77   | 0.77 ± 3.25  | 0.213   |
| CRP (mg/L)           | 4.98 ± 4.43   | 9.11 ± 12.07  | 27.43 ± 45.96| 0.002*  |
| Homocysteic acid (µmol/L) | 15.11 ± 5.10 | 17.65 ± 12.10| 17.25 ± 7.89 | 0.609   |
| LVEF (%)             | 61.53 ± 8.54  | 47.14 ± 15.73 | 46.80 ± 31.06| 0.022*  |
| LVIDd (cm)           | 5.09 ± 0.93   | 5.68 ± 1.08   | 5.74 ± 1.24  | 0.101   |
| LA (cm)              | 4.23 ± 0.52   | 4.47 ± 1.11   | 4.63 ± 1.00  | 0.434   |
| Cardiac tonic [n (%)]| 6 (28.6%)     | 40 (56.3%)    | 22 (68.8%)   | 0.017*  |
| Diuretic [n (%)]     | 7 (33.3%)     | 64 (90.1%)    | 30 (93.8%)   | <0.001* |
| Nitrate [n (%)]      | 9 (42.9%)     | 49 (69.0%)    | 25 (78.1%)   | 0.024*  |
| ACEI/ARB [n (%)]     | 15 (71.4%)    | 64 (90.1%)    | 27 (84.4%)   | 0.099   |
| Beta blockers [n (%)]| 12 (57.1%)    | 53 (74.6%)    | 18 (56.2%)   | 0.107   |
| CCB [n (%)]          | 7 (33.3%)     | 12 (16.9%)    | 10 (31.2%)   | 0.140   |
| Aspirin/clopidogrel [n (%)] | 12 (57.1%) | 40 (56.3%) | 18 (56.2%) | 0.998   |
| Statins [n (%)]      | 5 (23.8%)     | 14 (19.7%)    | 4 (12.5%)    | 0.542   |

ACEI: angiotensin-converting enzyme inhibitor; ALT: alanine aminotransferase; ARB: angiotensin receptor blocker; AST: aspartate aminotransferase; BMI: body mass index; CCB: calcium channel blocker; CK: creatinine kinase; CK-MB: creatinine kinase-MB; CRP: C-reactive protein; HDL-C: high-density lipoprotein cholesterol; LA: left atrium; LDL-C: low-density lipoprotein cholesterol; LVEF: left-ventricular ejection fraction; LVIDd: left-ventricular internal diameter at end diastole.

*P < 0.05 among NYHA II, NYHA III, and NYHA IV subgroups.

In nonischemic cardiomyopathy patients, heart failure is always accompanied with tissue hypoxia. The hypoxia was exacerbated with low-pulse oxygen saturation clinically when heart failure was aggravated. CP synthesis was increased as a defense mechanism in response to hypoxia [21]. Our results showed the statistical difference among different NYHA levels in nonischemic group, suggesting that CP’s ferroxidase activity and metal metabolism might play important roles. However, when we analyzed other risk factors and parameters related to the extent of heart failure, AST, uric acid, CKMB, CRP, and LVEF were not well balanced among subgroups, and so the logistic regression was given. The results showed that CP value was well correlated with the extent of heart failure, and this correlation was independent of AST, uric acid, CKMB, LVEF, and even C-reactive protein which is a sensitive marker of inflammation. The correlation was also independent of gender, hypertension, diabetes mellitus, and other risk factors. Age was the only risk factor related to the extent of heart failure.

### 5. Conclusions

We concluded that CP levels were significantly high in patients with ischemic or nonischemic cardiomyopathy and had a positive linear correlation with C-reactive protein and a negative linear correlation with LVEF. In nonischemic cardiomyopathy patients, the CP value was an independent biomarker associated with the extent of heart failure. Further studies should be applied to confirm our data in a larger number of patients and to clarify the mechanism.
Table 4: Relationship between extent of heart failure and clinical and laboratory parameters in patients with nonischemic cardiomyopathy.

| Variables                 | OR   | χ²  | P value | 95% confidence interval |
|---------------------------|------|-----|---------|-------------------------|
| Age                       | 1.072| 3.997| 0.046⁴ | 1.001–1.148             |
| Gender                    | 0.293| 1.426| 0.232   | 0.039–2.198              |
| Smoking                   | 1.499| 0.172| 0.678   | 0.222–10.136             |
| Alcohol                   | 0.993| 0.000| 0.994   | 0.125–7.874              |
| Hypertension              | 2.330| 1.214| 0.270   | 0.518–10.492             |
| Diabetes mellitus         | 1.047| 0.002| 0.967   | 0.121–9.026              |
| Creatinine                | 0.157| 1.029| 0.310   | 0.004–5.609              |
| ALT                       | 1.019| 0.339| 0.560   | 0.957–1.084              |
| AST                       | 0.972| 0.700| 0.403   | 0.910–1.039              |
| LDL-C                     | 1.026| 0.481| 0.488   | 0.954–1.013              |
| Total cholesterol         | 0.981| 0.621| 0.431   | 0.935–1.029              |
| Triglycerides             | 0.998| 0.076| 0.782   | 0.981–1.014              |
| Uric Acid                 | 1.141| 0.530| 0.467   | 0.800–1.629              |
| CK                        | 1.006| 1.962| 0.161   | 0.998–1.013              |
| CKMB                      | 1.069| 0.937| 0.333   | 0.934–1.223              |
| Troponin I                | 1.304| 1.011| 0.315   | 0.778–2.185              |
| CRP                       | 1.012| 0.255| 0.614   | 0.966–1.061              |
| Homocysteic acid          | 1.009| 0.057| 0.811   | 0.937–1.087              |
| LVEF                      | 0.981| 1.479| 0.224   | 0.952–1.012              |
| LVIDd                     | 0.616| 0.502| 0.479   | 0.161–2.354              |
| LA                        | 0.839| 0.097| 0.755   | 0.280–2.519              |
| Ceruloplasmin             | 1.010| 4.489| 0.034⁴ | 1.001–1.019              |

ALT: alanine aminotransferase; AST: aspartate aminotransferase; CK: creatinine kinase; CK-MB: creatinine kinase-MB; CRP: C-reactive protein; LA: left atrium; LDL-C: low-density lipoprotein cholesterol; LVEF: left-ventricular ejection fraction; LVIDd: left-ventricular internal diameter at end diastole. *P < 0.05.

Conflict of Interests

The authors declare that they have no conflict of interests.

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References

[1] C. Leeuwenburgh, J. E. Rasmussen, F. F. Hsu, D. M. Mueller, S. Pennathur, and J. W. Heinecke, “Mass spectrometric quantification of markers for protein oxidation by tyrosyl radical, copper, and hydroxyl radical in low density lipoprotein isolated from human atherosclerotic plaques,” Journal of Biological Chemistry, vol. 272, no. 6, pp. 3520–3526, 1997.

[2] J. B. Bustamante, M. C. Mateo, J. Fernandez, B. de Quiros, and O. O. Manchado, "Zinc, copper and ceruloplasmin in arteriosclerosis," Biomedicine Express, vol. 25, no. 7, pp. 244–245, 1976.

[3] A. Reunanen, P. Knekt, and R. K. Aaran, “Serum ceruloplasmin level and the risk of myocardial infarction and stroke,” American Journal of Epidemiology, vol. 136, no. 9, pp. 1082–1090, 1992.

[4] M. Mänttäri, V. Manninen, J. K. Huttunen et al., “Serum ferritin and ceruloplasmin as coronary risk factors,” European Heart Journal, vol. 15, no. 12, pp. 1599–1603, 1994.

[5] K. Klipstein-Grobusch, D. E. Grobbbee, J. F. Koster et al., “Serum caeruloplasmin as a coronary risk factor in the elderly: the Rotterdam Study,” British Journal of Nutrition, vol. 81, no. 2, pp. 139–144, 1999.

[6] P. M. Ridker, M. Cushman, M. J. Stampfer, R. P. Tracy, and C. H. Hennekens, “Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men,” New England Journal of Medicine, vol. 336, no. 14, pp. 973–979, 1997.

[7] L. Elsherif, R. V. Ortines, J. T. Saari, and Y. J. Kang, “Congestive heart failure in copper-deficient mice,” Experimental Biology and Medicine, vol. 228, no. 7, pp. 811–817, 2003.

[8] M. Correale, N. Daniele Brunetti, L. de Gennaro, and M. di Biase, "Acute phase proteins in atherosclerosis (Acute Coronary Syndrome)," Cardiovascular and Hematological Agents in Medicinal Chemistry, vol. 6, no. 4, pp. 272–277, 2008.

[9] S. J. Adelstein, T. L. Coombs, and B. L. Vallee, “Metalloenzymes and myocardial infarction—I. The relation between serum copper and ceruloplasmin and its catalytic activity,” The New England journal of medicine, vol. 255, no. 3, pp. 105–109, 1956.

[10] A. Enbergs, A. Dorszewski, M. Luft et al., “Failure to confirm ferritin and ceruloplasmin as risk factors for the angiographic extent of coronary arteriosclerosis,” Coronary Artery Disease, vol. 9, no. 2-3, pp. 119–124, 1998.

[11] A. Ziakas, S. Gavrilidis, E. Souliou et al., “Ceruloplasmin is a better predictor of the long-term prognosis compared with fibrinogen, CRP, and IL-6 in patients with severe unstable angina,” Angiology, vol. 60, no. 1, pp. 50–59, 2009.
[12] P. L. Fox, C. Mukhopadhyay, and E. Ehrenwald, “Structure, oxidant activity, and cardiovascular mechanisms of human ceruloplasmin,” *Life Sciences*, vol. 56, no. 21, pp. 1749–1758, 1995.

[13] S. M. Weissman, R. D. Wochner, F. X. Mullins, A. Wynngate, and T. A. Waldmann, “Synthesis of plasma proteins by heptatectomized dogs,” *The American journal of physiology*, vol. 210, no. 1, pp. 128–132, 1966.

[14] G. Ramadori, J. Van Damme, H. Rieder, and K. H. Meyer zum Buschenfelde, “Interleukin 6, the third mediator of acute-phase reaction, modulates hepatic protein synthesis in human and mouse. Comparison with interleukin 1β and tumor necrosis factor-α,” *European Journal of Immunology*, vol. 18, no. 8, pp. 1259–1264, 1988.

[15] B. Mazumder, C. K. Mukhopadhyay, A. Prok, M. K. Cathcart, and P. L. Fox, “Induction of ceruloplasmin synthesis by IFN-γ in human monocytic cells,” *Journal of Immunology*, vol. 159, no. 4, pp. 1938–1944, 1997.

[16] E. J. Lewis, A. D. Sedgwick, and T. H. Hanahoe, “In vivo changes in plasma acute phase protein levels in the rat induced by slow release of IL-1, IL-6 and TNF,” *Mediators of Inflammation*, vol. 1, pp. 39–44, 1992.

[17] G. Engström, P. Lind, B. Hedblad, L. Stavenow, L. Janzon, and F. Lindgärde, “Effects of cholesterol and inflammation-sensitive plasma proteins on incidence of myocardial infarction and stroke in men,” *Circulation*, vol. 105, no. 22, pp. 2632–2637, 2002.

[18] N. Shukla, J. Maher, J. Masters, G. D. Angelini, and J. Y. Jeremy, “Does oxidative stress change ceruloplasmin from a protective to a vasculopathic factor?” *Atherosclerosis*, vol. 187, no. 2, pp. 238–250, 2006.

[19] C. Altamura, R. Squitti, P. Pasqualetti et al., “Ceruloplasmin/Transferrin system is related to clinical status in acute stroke,” *Stroke*, vol. 40, no. 4, pp. 1282–1288, 2009.

[20] S. Shiva, X. Wang, L. A. Ringwood et al., “Ceruloplasmin is a NO oxidase and nitrite synthase that determines endocrine NO homeostasis,” *Nature Chemical Biology*, vol. 2, no. 9, pp. 486–493, 2006.

[21] F. Martin, T. Linden, D. M. Katschinski et al., “Copper-dependent activation of hypoxia-inducible factor (HIF)-1: implications for ceruloplasmin regulation,” *Blood*, vol. 105, no. 12, pp. 4613–4619, 2005.