Fluorometric Determination of Deoxyribonucleic Acid in Bacteria with Ethidium Bromide

J. A. DONKERSLOOT, S. A. ROBRISH, AND M. I. KRICEVSKY

Environmental Mechanisms Section, National Institute of Dental Research, National Institutes of Health, Bethesda, Maryland 20014

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A simple, sensitive, and rapid method is presented for the determination of deoxyribonucleic acid (DNA) in both gram-positive and gram-negative bacteria. It is based upon the fluorometric determination of DNA with ethidium bromide after alkaline digestion of the bacteria to hydrolyze the interfering ribonucleic acid. The assay takes less than 2 hr. Its sensitivity is at least 0.2 μg of DNA in a final solution of 4 ml and it uses commonly available filter or double monochromator fluorometers. Judicious choice of light source and filters allows an additional 10-fold increase in sensitivity with a filter fluorometer. Turbidity caused by bacteria or insoluble polysaccharides does not interfere with the fluorescence measurements. There was no significant difference between the results obtained with this method and those obtained with the indole and diphénylamine methods when these assays were applied to Escherichia coli and sucrose- or glucose-grown Streptococcus mutans. The method was also tested by determining the specific growth rate of E. coli. This new procedure should be especially useful for the determination of bacterial DNA in dilute suspensions and for the estimation of bacterial growth or DNA replication where more conventional methods are not applicable or sensitive enough.

The binding of ethidium bromide (EB) to nucleic acids results in a marked increase in its fluorescence which can be used to measure pure deoxyribonucleic acid (DNA) and ribonucleic acid (RNA) either alone or in combination (8). This method has not been extended to the measurement of DNA in microorganisms, although it is potentially about 100-fold more sensitive than the two colorimetric procedures most often used (1, 2) and has equal, or better, selectivity. We have, therefore, developed a convenient and sensitive assay for microbial DNA based on the intercalation of EB with DNA. The method consists of an alkaline digestion of the bacteria to hydrolyze the RNA and eliminate its response. The alkaline digest is then neutralized to effect partial renaturation of the DNA, EB is added, and the fluorescence is measured with a filter or double monochromator fluorometer.

MATERIALS AND METHODS

Maintenance and growth of bacteria. A laboratory strain of Escherichia coli and the SL-1 strain of Streptococcus mutans were maintained by bimonthly transfer into, respectively, Antibiotic Medium 3 (AB-3 medium, Difco) and fluid thioglycolate medium (BBL) supplemented with horse meat infusion and CaCO₃. After overnight growth at 37°C, the cultures were stored at 3°C. For the growth experiment, E. coli was grown in 50 ml of AB-3 medium in a 200-ml Erlenmeyer flask on a reciprocating shaker (120 strokes/min) at 30°C. For the comparative DNA measurements, E. coli was grown overnight into stationary phase on a rotary shaker (100 rev/min) and then stored at 3°C. One-half milliliter of an overnight culture grown under similar conditions was used as inoculum. S. mutans was grown in 30 ml of broth in test tubes (200 × 25 mm). The medium contained per liter: 5 g of Trypticase (BBL), 5 g of yeast extract (BBL), 5 g of K₂HPO₄, 3H₂O, 50 mg of Na₂ CO₃, and 0.5 ml of a salt solution containing (per 100 ml) 800 mg of MgSO₄·7H₂O, 1 mg of FeSO₄·7H₂O, and 19 mg of MnSO₄·4H₂O. The pH was adjusted to 7.2 with HCl before steam sterilization. Afterwards, glucose or sucrose was added to give a final concentration of 0.01 M. The inoculum was grown overnight on the glucose medium, and 0.3 ml was used per tube. The cultures were incubated for 24 hr at 37°C and subsequently stored at 3°C for DNA analysis. The sucrose-grown S. mutans was exposed for 2 min to sonic energy from a Branson sonifier with a microtip and an output of 50 w. Microscopic inspection showed that such treatment releases most of the streptococci from the dextran aggregates.

Chemicals. Calf thymus DNA, EB, and micro-
cocal nuclease (from \textit{Staphylococcus aureus}; EC 3.1.4.7) were dissolved in 0.1 M tris(hydroxymethyl)aminomethane-hydrochloride buffer of pH 8.5 (T8.5 buffer) in 0.1 mg, 0.1 mg, and 3,000-unit amounts (pg. 4 ml of fresh reagent). Yeast RNA was added solved in 0.1 M phosphate buffer (pH 7.0; 0.1 mg/ml of buffer). Nuclease, DNA, and RNA were obtained from Worthington Biochemical Corp.; and EB (B grade) was from Calbiochem.

\textbf{Fluorometric DNA analysis.} A sample from the bacterial suspension containing 0.2 to 10 \(\mu\)g of DNA was filtered over a previously wetted Metrical DM-450 membrane filter (25 mm, 0.45-\(\mu\)m pore size, Gelman Instrument Co.) and washed twice with an equal volume of 0.85\% saline. Each filter was submersed in 2.0 ml of 0.3 \(\times\) KOH in a 12-ml conical centrifuge tube, and the bacteria were resuspended by using a test tube mixer or a platinum needle.

After incubation for 1 hr at 37 C, 0.5 ml of 1.0 \(\times\) HCl was rapidly mixed into the digest, and 2.0 ml of the partially neutralized suspension was added to 2.0 ml of EB (4 \(\mu\)g/ml in T8.5 buffer). The fluorescence was measured (after equilibration to room temperature and resuspension of any precipitate) against a reagent blank and compared with that of DNA standards similarly treated. The Metrical polyvinylchloride filters do not contribute to the fluorescence, whereas certain other polyvinylchloride filters do so in a rather variable manner. Conventional cellulose ester-type filters cannot be used because they degenerate during the alkaline digestion. Fluorescence was routinely measured in round cuvettes (12 \(\times\) 75 mm; Hycel, Inc.) with a Turner 111 fluorometer equipped with a general purpose lamp (General Electric F4T4.BL) and a temperature-stabilized sample holder. Primary filters were Corning 1-60 (nearest to the lamp) and Watten 58 for the selection of 546-nm exciting light, and the secondary filter was a Watten 23A for the passage of emitted light above 570 nm. An Amino fluorocolorimeter was also used with the same light source and filters. For samples containing 0.02 to 0.2 \(\mu\)g of DNA, it is advantageous to use an EB concentration of 1 \(\mu\)g/ml to lower the background fluorescence and to switch to a General Electric F4T5.G green lamp to obtain sufficient response. The Amino-Bowman, Farrand Mk-1, and Perkin-Elmer MPF-3 were used as double monochromator fluorometers. Optimal excitation and emission wavelength settings were determined with a solution containing 10 \(\mu\)g of DNA and 1 \(\mu\)g of EB per ml of T8.5 buffer in a 1-cm square cuvette and using the narrowest slits consistent with adequate response. They were, respectively, 600 nm for excitation and 585 (Farrand, Perkin-Elmer) or 600 (Amino) nm for emission.

\textbf{Colorimetric DNA analysis.} For the diphenylamine method, a sample of the bacterial suspension containing at least 10 \(\mu\)g of DNA was filtered, and the bacteria were washed with saline and released from the filter in 2 ml of 0.5 \(\times\) HClO\(_4\) in a centrifuge tube. After incubation for 30 min at 70 C and cooling of buffer contained in a centrifuge tube, 2 ml of the reagent (1) was added, and the samples were kept for 16 to 20 hr at 30 C. After centrifugation, the absorbance of the supernatant fluid was determined, at 600 nm against a reagent blank with a Gilford 300N spectrophotometer, and compared with that of DNA standards. For the indole method, a modification of the method of Ceriotti (2), the bacteria, containing at least 5 \(\mu\)g of DNA, were collected and washed as described and then suspended in 3 ml of reagent prepared by mixing 60 ml of 0.015\% indole with 5 ml of HCl (38\% w/w) before use. After 2 hr at 70 C and subsequent centrifugation, the absorbance of the supernatant fluid was measured at 490 nm against a reagent blank and compared with that of DNA standards.

\textbf{Enzymatic methods.} To degrade DNA, 150 units of micrococal nuclease were added after measurement of the fluorescence of the DNA-EB complex, the samples were reincubated for 30 min at 37 C, and the fluorescence was measured again after cooling. Preliminary work showed that similar treatment of 10 \(\mu\)g of alkali-denatured DNA reduced the fluorescence to less than 1\%, even in the presence of EB and in the absence of Ca ions. Insoluble polysaccharide was prepared in vitro by using the medium of a 24-hr culture of \textit{S. mutans} grown with glucose as energy source. After filtration through a membrane filter (0.45-\(\mu\)m pore size), the filtrate was aseptically adjusted to 0.05 \(\times\) sucrose and reincubated for 24 hr at 37 C. The particle size of the resultant insoluble polysaccharide was reduced by a 2-min exposure to 50-w sonic energy from a Branson sonifier. To study the effect of turbidity on fluorescence, samples were filtered off, washed, and digested with or without 10 \(\mu\)g of DNA added, and the fluorescence was compared with that of DNA alone. The turbidity of the final solution was also measured with a Gilford 900N spectrophotometer and expressed as apparent absorbance at 550 nm.

\textbf{Other determinations.} The growth of \textit{E. coli} was measured by determining the apparent absorbance of 1-ml samples at 600 nm. The samples were diluted with saline to keep the readings below 0.4 to preserve linearity between the readings and the dry weight (7). Dry weights were determined in triplicate by collecting the cells from a 2-ml sample, on a previously weighed Metrical DM-450 filter, and washing the bacteria with 5 ml of water. The filters were then transferred to aluminum pans and dried for a few hours at 100 C until constant weight was obtained. Control filters were treated similarly to determine the weight loss. The filters were weighed on the 20-mg scale of a Cahn Gram Electrobalance after equilibration over CaCl\(_2\) in a desiccator.

\textbf{RESULTS}

Because EB intercalates with the double-stranded regions of both DNA and RNA, it was necessary to eliminate the RNA binding by a 1-hr hydrolysis in 0.3 \(\times\) KOH at 37 C (10, 13, 18). The data in Fig. 1 confirm that this procedure virtually eliminates the RNA response. The fluorescence of the denatured DNA after this treatment is about 45\% of that of native DNA. This lower value does not only

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result from the alkaline conditions to which the DNA has been subjected, but also from the presence of 0.1 M KCl in the final assay mixture (9). Furthermore, these data suggest that a certain amount of order is restored in the DNA after adjustment of the pH to 8.5. Figure 1 also shows that the response for the DNA after this treatment is linear over a range of at least 0.1 to 10 μg of DNA as measured with a double monochromator fluorometer. Similar results were obtained with the other fluorometers tested. We found that the time period between the partial neutralization with acid and the addition of EB did not critically affect the final fluorescence. In addition, the complex formation between EB and DNA is very rapid compared with the time required for temperature equilibration prior to the fluorescence measurements.

When this method was applied to *S. mutans*, the bacteria were collected and washed on polycrylpropylene membrane filters which withstood the alkaline digestion. After the digestion and partial neutralization, the final solution was slightly turbid because of particulate matter from the bacteria. This turbidity could be removed by membrane filtration or centrifugation, but both methods resulted in considerably lower fluorometer readings, probably as a result of the adherence of the DNA to the streptococcal debris (14).

To confirm that the fluorometric response observed was, indeed, due to streptococcal DNA and not to other fluorescent compounds, micrococcal nuclease was used to degrade the DNA after measurement of the fluorescence. This treatment reduced the fluorescence to less than 5% of the earlier values, even though the conditions used are suboptimal for enzyme activity.

Variations in the length of the alkaline digestion between 0.5 and 2 hr did not affect the final result significantly. A period of 1 hr was, therefore, adopted for this resulted in hydrolysis of at least 100 μg of RNA under the conditions used.

To determine if the turbidity that was observed with the bacterial samples interfered with the measurements by scattering either the exciting (primary) or the emitted (secondary) light, several experiments were performed. The method was first applied to increasing amounts of streptococci, with and without an internal DNA standard added. The results of this reconstruction experiment are shown in Fig. 2. Without added DNA, the relationship between bacteria and response is described by a straight line through the origin over the

![Graph 1](image1.png)

**FIG. 1.** Fluorescence of native and alkali-treated DNA and RNA after reaction with ethidium bromide. The fluorescence was measured against a reagent blank on a Farrand Mk-1 spectrofluorometer and is expressed as the photomultiplier current. The extinction and emission wavelengths were 520 and 585 nm, respectively. Slits with a bandpass of 20 nm were used. To preserve linearity, the amount of EB should be at least equal the amount of nucleic acid in micromicrograms.

![Graph 2](image2.png)

**FIG. 2.** Determination of DNA in *S. mutans* with ethidium bromide. After measurement of the fluorescence (○), 5 μg of DNA was added and the tubes were measured again (□). In the other set of tubes, 10 μg of DNA was added to the streptococci before the digestion (□). Fluorescence was measured on a Turner 111 fluorometer and is expressed in arbitrary units.
range studied. The line obtained with 10 μg of DNA added to the samples is also straight and parallel to the one without the internal standards. This shows that the backscatter of the primary light due to the bacterial debris does not significantly decrease the intensity of the primary light, and hence the fluorescence, over the range studied. Figure 2 also shows that when 5 μg of DNA was added after the measurement of the streptococcal samples, the recovery was 100%. In a second experiment (data not shown), different amounts of insoluble polysaccharide were added to 10 μg of DNA, and both the fluorescence and turbidity were measured after the procedure. We found that an increase in the apparent absorbance at 550 nm from 0 to 0.2 did not significantly alter the fluorescence, which proves that the amount of primary light that reaches the photomultiplier because of scatter does not contribute significantly to the observed fluorescence.

The EB method was developed by using streptococci but can also be used to measure the DNA in a gram-negative organism such as E. coli. Figure 3 shows that the DNA measurements correlate well with the absorbance data. The doubling time derived from either the absorbance or the DNA data was 40 min.

Finally, we compared this new method with the diphenylamine and indole assays for DNA. The data in Table 1 show that there was no significant difference between the three methods when applied to E. coli or S. mutans. The DNA content of the stationary-phase E. coli and glucose-grown S. mutans was 4.3 and 3.0%, respectively. The sucrose-grown S. mutans culture also contained an appreciable amount of insoluble polysaccharide, but the results confirm that this did not interfere significantly with the method. Nuclease treatment of the final solution reduced the fluorescence to less than 5% with pure DNA or glucose-grown S. mutans. But with sucrose-grown S. mutans and with E. coli only a 10 to 20% reduction in fluorescence was observed. In the latter cases, the denatured and partially insoluble DNA is apparently more resistant towards enzymatic degradation.

**DISCUSSION**

The method presented here is based on the binding of EB to double-stranded DNA and, therefore, should be insensitive to interferences encountered with the indole and diphenylamine procedures for DNA, which depend on the deoxyribose moiety. The good agreement observed between the three methods indicates that each one can be used reliably for E. coli and S. mutans and, furthermore, that the pool of low-molecular-weight compounds that might interfere with the colorimetric tests is relatively small in these bacteria.

The RNA response with EB is most conveniently eliminated by an alkaline hydrolysis (13). The length of this digestion was found to be relatively unimportant, but it should be checked whenever the method is applied to a new class of organisms which might be more

**TABLE 1. Comparison of the diphenylamine, indole, and ethidium bromide assays for bacterial DNA**

| Assay method | DNA (μg/ml) |
|--------------|-------------|
|              | *E. coli* | *S. mutans* |
| Diphenylamine | 54.3       | 12.5         | 14.1         |
| Indole       | 50.8       | 12.3         | 15.4         |
| Ethidium bromide | 54.9   | 12.2         | 13.2         |

* E. coli culture had a dry weight of 1.24 mg/ml.
* *Glucose*-grown S. mutans culture had a dry weight of 0.41 mg/ml.
* *Sucrose*-grown S. mutans culture had a dry weight of 0.75 mg/ml and contained insoluble polysaccharides.
resistant. The quantitative response of the alkali-denatured DNA was found to be the same as that of heat-denatured DNA (9). Most of the streptococcal DNA remains insoluble after the alkaline digestion, but the results indicate that this does not interfere with the EB binding or fluorescence. It has been reported that extraction of Streptococcus faecalis, Lactobacillus leichmanni, or Euglena gracilis with 1 N KOH solubilized only 20 to 35% of their total DNA (4, 14). To determine the possible interference of the bacterial debris with the fluorescence measurements, various experiments were performed, but no significant effects were encountered. This agrees with the findings of others as reviewed by Udenfriend (17).

The main advantages of the EB method are its ease and sensitivity. The latter is at least 50 ng per ml of final solution and can be extended further by using a light source with a higher intensity around 520 nm. The sensitivities of the indole and diphenylamine methods in the modifications used here were 1 and 2 μg per ml of final solution, respectively. Another fluorometric assay for DNA is based on the reaction of deoxyribose with 3,5-diaminobenzoic acid (6). Its disadvantage is that lipids have to be removed by extraction, which requires great care, especially with small samples, to prevent the loss of DNA (15). Particular attention also has to be paid to the hot-acid hydrolysis of the DNA to expose the purine-bound deoxyribose and to the purity of the reagent (5, 12). A third fluorometric procedure is based on the degradation of thymine with bromine water to yield acetol, which is subsequently reacted with o-aminobenzaldehyde to give the fluorescent 3-hydroxyquinidine (11). The latter two methods are about equally as sensitive as the EB method but, to our knowledge, have only been tested with animal tissues.

One promising application for the method presented here is to measure growth in situations where solids interfere with the more conventional absorbance or dry-weight assays. Such a case arises when S. mutans is grown with sucrose as energy source (16). Another possible application is in the detection of very low amounts of bacteria. Assuming 10^{-6} μg of dry weight per bacterial cell and 4% DNA, it can be estimated that, with the higher intensity light source, the EB method can be used to measure DNA in a 10-ml sample containing 50,000 cells per ml. The EB method is, therefore, as sensitive for the detection of bacteria as nephelometry with a laser light-scattering photometer (Science Spectrum, Santa Barbara, Calif., 93105), about 10 times more sensitive than direct turbidimetry, but at least an order of magnitude less sensitive than the method using the firefly bioluminescence assay for ATP (3), a method dependent on the efficient extraction of ATP from the cells and special instrumentation.

LITERATURE CITED
1. Burton, K. 1968. Determination of DNA concentration with diphenylamine, p. 163-166. In L. Grossman and K. Moldave (ed.), Methods in enzymology, vol. 12, part B. Academic Press Inc., New York.
2. Ceriotti, G. 1952. A microchemical determination of deoxyribonucleic acid. J. Biol. Chem. 198:297-303.
3. Chappelle, E. W., and G. V. Levin. 1968. Use of the firefly bioluminescent reaction for rapid detection and counting of bacteria. Biochem. Med. 2:41-52.
4. Downing, M., and B. S. Schweigert. 1956. Role of vitamin B12 in nucleic acid metabolism. III. Preparation of C^4 uniformly labeled pyrimidine deoxyribonucleosides. J. Biol. Chem. 229:515-520.
5. Hinegardner, R. T. 1971. An improved fluorometric assay for DNA. Anal. Biochem. 39:197-201.
6. Kissane, J. M., and E. Robins. 1958. The fluorometric measurement of deoxyribonucleic acid in animal tissues with special reference to the central nervous system. J. Biol. Chem. 233:184-188.
7. Koch, A. L. 1970. Turbidity measurements of bacterial cultures in some available commercial instruments. Anal. Biochem. 38:252-259.
8. Le Pecq, J. B., and C. Paoletti. 1966. A new fluorometric method for RNA and DNA determination. Anal. Biochem. 17:100-107.
9. Le Pecq, J. B., and C. Paoletti. 1967. A fluorescent complex between ethidium bromide and nucleic acids. Physical-chemical characterization. J. Mol. Biol. 27:87-106.
10. Munro, H. N., and A. Fleck. 1966. The determination of nucleic acids, p. 113-176. In D. Glick (ed.), Methods of biochemical analysis, vol. 14. Interscience Publishers, Inc., New York.
11. Roberts, D., and M. Friedkin. 1958. The fluorometric determination of thymine in deoxyribonucleic acid and derivatives. J. Biol. Chem. 233:463-467.
12. Robertson, F. W., and K. Tait. 1971. Optimal conditions for the fluorometric determination of DNA. Anal. Biochem. 41:477-481.
13. Schmidt, G., and S. J. Thanhauser. 1945. A method for the determination of deoxyribonucleic acid, ribonucleic acid, and phosphoproteins in animal tissues. J. Biol. Chem. 161:83-89.
14. Sherrat, H. S. A., and A. J. Thomas. 1953. The nucleic acid fractions of a strain of Streptococcus faecalis. J. Gen. Microbiol. 8:217-223.
15. Switzer, B. R., and G. K. Summer. 1971. A modified fluorometric micromethod for DNA. Clin. Chim. Acta 32:203-206.
16. Tanzer, J. M., W. I. Wood, and M. I. Krichevsky. 1969. Linear growth kinetics of plaque-forming streptococci in the presence of sucrose. J. Gen. Microbiol. 58:125-133.
17. Udenfriend, S. 1969. Fluorescence assay in biology and medicine, vol. 2, p. 185-188. Academic Press Inc., New York.
18. Webb, J. M., and H. B. Levy. 1958. New developments in the chemical determination of nucleic acids, p. 1-30. In D. Glick (ed.), Methods of biochemical analysis, vol. 6. Interscience Publishers, Inc., New York.