Unveiling the G4-PAMAM capacity to bind and protect Ang-(1-7) bioactive peptide by molecular dynamics simulations

L. América Chi1· Somayeh Asgharpour2· José Correa-Basurto1· Cindy Rodríguez Bandala1,3· Marlet Martínez-Archundia1

Received: 10 June 2022 / Accepted: 26 July 2022 / Published online: 8 August 2022
© The Author(s), under exclusive licence to Springer Nature Switzerland AG 2022

Abstract
Angiotensin-(1-7) re-balance the Renin-Angiotensin system affected during several pathologies, including the new COVID-19; cardiovascular diseases; and cancer. However, one of the limiting factors for its therapeutic use is its short half-life, which might be overcome with the use of dendrimers as nanoprotectors. In this work, we addressed the following issues: (1) the capacity of our computational protocol to reproduce the experimental structural features of the (hydroxyl/amino)-terminated PAMAM dendrimers as well as the Angiotensin-(1-7) peptide; (2) the coupling of Angiotensin-(1-7) to (hydroxyl/amino)-terminated PAMAM dendrimers in order to gain insight into the structural basis of its molecular binding; (3) the capacity of the dendrimers to protect Angiotensin-(1-7); and (4) the effect of pH changes on the peptide binding and covering. Our Molecular-Dynamics/Metadynamics-based computational protocol well modeled the structural experimental features reported in the literature and our double-docking approach was able to provide reasonable initial structures for stable complexes. At neutral pH, PAMAM dendrimers with both terminal types were able to interact stably with 3 Angiotensin-(1-7) peptides through ASP1, TYR4 and PRO7 key amino acids. In general, they bind on the surface in the case of the hydroxyl-terminated compact dendrimer and in the internal zone in the case of the amino-terminated open dendrimer. At acidic pH, PAMAM dendrimers with both terminal groups are still able to interact with peptides either internalized or in its periphery, however, the number of contacts, the percentage of coverage and the number of hydrogen bonds are lesser than at neutral pH, suggesting a state for peptide release. In summary, amino-terminated PAMAM dendrimer showed slightly better features to bind, load and protect Angiotensin-(1-7) peptides.

Keywords Dendrimer · Nanocarrier · Peptide · Ang-(1-7) · G4-PAMAM · PAMAM-OH · Metadynamics · Molecular dynamics

Introduction
The Renin-Angiotensin system (RAS) regulates the blood pressure and also balances ion and other fluids in the human body. For it to function properly, the RAS rests on two opposite axis (Fig. 1): (1) in the classical arm, inflammatory, profibrotic, vasoconstrictor and oxidative effects are triggered by the binding of bioactive octapeptide Angiotensin II (Ang-II) to Angiotensin Type 1 (AT1) cell membrane receptor; meanwhile, (2) in the protective arm, opposite anti-inflammatory, anti-fibrotic, vasodilator and anti-oxidant effects are triggered by the binding of the bioactive heptapeptide Angiotensin 1-7 (Ang-(1-7)) to Mitochondrial Assembly (MAS) proto-oncogene which is a G-protein-coupled receptor (GPCRs) [1, 2]. There must be a balance between the two main axes, however, under several...
pathological conditions this balance is shifted towards the classical arm leading to catastrophic effects, as in the case of the worsening of COVID-19 disease [3–7].

Recently, Ang-(1-7) has emerged as a novel therapeutic agent who acts as a RAS modulator. It has been proved to be a safe and efficacious (intravenous) IV drug, and since it is not pathogen-specific, it is thought to support a natural healing process [4]. Several clinical and preclinical studies showed that it can protect organs like lung, kidney, and heart [4]. Also, some clinical trials testing the safety and the efficacy of intravenous Ang-(1-7) infusion in COVID-19 patients with severe pneumonia are currently under investigation [8]. However, Ang-(1-7) has faced limited clinical application due to its unfavorable pharmacokinetic profile: short half-life, poor systemic distribution and poor ability to cross physiological barriers as well as a need for controlled dosage [9–12]. To overcome these challenges, several strategies have arisen: Ang-(1-7) key amino acid substitution with a cyclic non-natural amino acid [11], cyclization of Ang-(1-7) by inclusion of a thioether bridge [12, 13], Ang-(1-7) inclusion into β-cyclodextrin molecule [14], liposome-entrapped of angiotensin-(1-7) [15] and an Ang-(1-7) eluting polymer-coated medical device [16]. One of the most promising approaches is the physically entrapment of a drug inside a dendrimer-based nanocarrier using non-covalent interactions [17, 18]. Dendrimers are strong candidates because they offer structural stability, high carrying capacity of molecules, water solubility, modifiable surface functionality, available internal cavities, pH dependence properties and a high capacity to cross cell barriers together with an ease of synthesis or commercial availability [19]. Most of the previous studies have focused on PAMAM, among all dendrimers and hydroxyl-terminated PAMAM dendrimer has been additionally proven to be a non-cytotoxic variant compared with the original amino-terminated PAMAM dendrimer that shows a slight cytotoxic effect [20]. In this regard, it was discovered that, in general, dendrimer cytotoxicity depended on the generation number, the number of terminal groups, and the nature of terminal moieties: anionic, neutral, or cationic [21]. In amino-terminated PAMAM dendrimers, it is thought that cationic dendrimer terminals may disrupt cell membranes that have negatively charged on their surface, the cationic terminal groups extracts negatively charged lipids from the membrane with subsequent lysis and cytotoxicity [22], although low concentrations of these substances can be considered safe, in case of amino-terminated G4-PAMAM dendrimers are safe at doses < 10 mg/kg [23]. To decrease the cytotoxicity of dendrimers several approaches are applied: (1) a protecting molecule could be added to terminal groups, this could be amino acids such as glycine and phenylalanine [23]; (2) a second approach is a surface-charge neutralization, it has been reported that in general cationic dendrimers have shown reduced cytotoxicity after the complexation with another molecule such as indomethacin due to the charges neutralization [21, 24], thus, in this work, we could expect that the amino-terminated PAMAM-G4 toxicity will decrease due to the presence of peptide neutralizing its charges but testing it is outside of our scope; and (3) a third approach to minimize cytotoxicity in dendrimers is the substitution of terminal groups for neutral ones, this was the main reason why we considered hydroxyl-terminated PAMAM (PAMAM-OH) dendrimers, which lacks the cationic surface groups, this dendrimer has been suggested as a safe drug carrier due to non-toxic effects observed during in vitro and in vivo applications. By using the latter strategy, the capacity of PAMAM-OH dendrimer as an Ang(1-7) carrier was previously explored by means of theoretical and experimental studies [25] and main conclusions pointed to: (a) one dendrimer could be able to capture two peptides; (b) Ang-(1-7) could be protected by the dendrimer in a range between 50 and 65 %; (c) Ang-(1-7)/dendrimer forms stable complexes over 60 ns MD which is confirmed by electrophoretic shift assay; and (d) Ang-(1-7)/PAMAM-OH complex had an anti-atrophic effect when administered intraperitoneally in mice. We now extend the strategy to design Ang-(1-7)/PAMAM-(OH/NH) complexes by using Ang-(1-7) NMR structure instead of a molecular model, performing molecular docking taking into account the dendrimer and the peptide flexibility, considering different pH conditions in order to glimpse a possible loading and unloading mechanism and extending sampling of the systems.

In summary, we coupled Ang-(1-7) to the most studied kind of dendrimers. PAMAM dendrimers, in order to evaluate their capacity to improve the Ang-(1-7) pharmacokinetic properties and to gain insight into the host-guest (dendrimer/ Ang-(1-7)) molecular binding. Understanding the basis of the host-guest molecular binding is fundamental when a very specific and controlled drug release is needed. It is important
to note that the beneficial effect of Ang-(1-7) peptide take place only at low concentrations and overdosing might interfere with its receptor-associated functions [12]. Different pH conditions (acidic and neutral) and two different group terminals (amino and hydroxyl) were tested as they are critical factors for modulating drug delivery and cytotoxicity. Rigorous computational approaches, such as molecular dynamics (MD) and metadynamics (MTD) have been used to evaluate the structural and energetic properties of the Ang-(1-7), dendrimer or the new complexes formed. Finally, we propose that the best complexes found can be experimentally tested to validate the properties predicted through In Silico approaches, as a potential therapeutic agent.

Methods

Molecular models The NMR structure of Ang-(1-7) peptide (ASP-ARG-VAL-TYR-ILE-HIS-PRO) in solution was retrieved from the protein database with ID: 2JP8; the topology was created using the pdb2gmx module of GROMACS 5.0.7 [26] and its parameters were downloaded from the GROMOS-compatible 2016H66 force field [27]; protonation states were assigned according to physical-chemical properties predicted with peptide calculator server [28] and verified by using ProPkA software [29] (zero peptide charge for neutral pH and +2 for acidic conditions) as shown in the Figs. S1-S3 of the Supplemental Material.

Two types of PAMAM dendrimers were considered: Generation 4 amino-terminated ethylenediamine (EDA)-cored poly(amidoamine) dendrimer (PAMAM-NH₃) and generation 4 hydroxyl-terminated ethylenediamine (EDA)-cored poly(amidoamine) dendrimer (PAMAM-OH) as depicted in Figs. S4-S5. The initial structures and topologies of the dendrimers were created using the newly developed dendrimer topology builder called pyPolyBuilder [30]. The parameters of the dendrimer and its partial charges were obtained from the GROMOS-compatible 2016H66 force field [27]. Previously, systematic evaluation of the accuracy of this force field was performed in the simulation of PAMAM dendrimers [31]. Characteristics of the dendrimers building-blocks used in this work can be found in Table S.5–S.17 of the Supporting Information from the work of Ramos et. al. [31] and in Table S1 of this work. Two pH values were considered: acidic pH of 3 and neutral pH of 7. To mimic pH conditions, the protonation states for the PAMAM dendrimers were assigned according to the experimental work by Cakara et al. [32]: at neutral pH, the primary amines were considered protonated and at low pH, both primary and tertiary amines were considered protonated (Fig. S6).

MD of isolated molecules in solution Dendrimers and peptide structures were placed separately in a box of SPC water model and then subjected to minimization, equilibration and production of standard MD simulations using GROMACS 5.0.7 [26] software, to obtain initial relaxed structures of both types of molecules. Systems were named as: PNH₃, POH₃, PNH₄ and POH₄, Ang-(1-7)₄ and Ang-(1-7)₄, where letters “n” and “a” correspond to neutral pH and acidic pH, respectively. Abbreviations PNH and POH correspond to amino-terminated PAMAM and hydroxyl-terminated PAMAM dendrimers, respectively.

Clustering Representative structures were selected from each MD using clustering by conformations. In the case of dendrimers, the middle structure of the 3 largest clusters was taken as the representative conformation, meanwhile, for peptide, the middle structures in all of the obtained clusters were used unless the cluster had less than 10 members. The center of a cluster or the middle structure is the structure with the smallest average RMSD from all other structures of the cluster.

Molecular docking A double molecular docking was performed in order to have initial dendrimer/peptide complexes by using Autodock Vina [33] program. We tested all the conformers of the dendrimer and the peptide previously selected. First, a blind rigid docking was performed separately for all combinations of the conformers (each dendrimer conformer is docked to each peptide conformer). Once the peptide conformer found a binding site, a second flexible local docking was performed in this area for each system allowing a better adaptation in the binding site. According to the number of hydrogen bonds (HBs) and Vina affinity, one peptide conformer was selected for each dendrimer site avoiding overlapping. We found that peptides can interact with 3 up to 4 different dendrimer sites for both types of terminal groups. Thus, we decided to choose a configuration with 3 peptides because these peptide conformers fulfill better the above-mentioned criteria, it would be interesting for future works to systematically test the number of peptides attached to each dendrimer. Detailed results are presented in supplemental material (Table S2). The double molecular docking approach was developed using our in-house bash script algorithm [34].

MD for complexes A multi-ligand complex was added to each dendrimer type MD simulation and at each pH value. A total of 4 complexes: [PNH-A]₄, [POH-A]₄, [PNH-A]₄ and [POH-A]₄, where letters correspond to neutral pH (n), acidic pH (a) and amino-terminated PAMAM (PNH), hydroxyl-terminated PAMAM (POH) and ang-(1-7) peptides (A). Each complex was placed separately in a SPC model water box and then the three steps of energy minimization, equilibration and production of standard MD were performed using GROMACS 5.0.7 software [26].

Metadynamics In metadynamics (MTD), an external history-dependent bias potential is constructed in the space of a few selected degrees of freedom namely the collective variables (CVs), the effect of the metadynamics bias potential being
to push the system away from local minima into exploring new regions of the phase space [35]. In the long time limit, the bias potential converges to minus the free energy [36] (or a proportional quantity in the case of well-tempered metadynamics (WT-MTD)) [37] as a function of the CVs. WT-MTD allows an exhaustive sampling of all the possible conformations assumed by the molecules in solution, showing its real flexibility. For Ang-(1-7) metadynamics, we first used the radius of gyration \( R_g \) as CV, however, it was not enough to describe the peptide behaviour in solution and led to a stucked system in low \( R_g \) values as depicted in Fig. S7. In our final set up, we used the \( R_g \) and the total number of donor-acceptor contacts \( (C_{da}) \) as collective variables because they are appropriate descriptors of the behavior of flexible molecules in solution [38]. \( C_{da} \) was calculated using a switching functions inspired by the work of Barducci et al [39],

\[
C_{da} = \sum_i \frac{1 - \left( \frac{d_{ij}}{r_0} \right)^n}{1 - \left( \frac{r_0}{r_0} \right)^m},
\]

where \( r_0 \) was set to 0.25 nm and \( n \) and \( m \) were set to 6 and 12, respectively. The sum runs over all of the pairs \( i, j \) of Hydrogen, Nitrogen and Oxygen polar atoms and \( d_{ij} \) is the distance between each \( i \) and \( j \) atoms. In order to limit the \( R_g \) sampling, an upper and a lower harmonic restraint was applied. The restraining potential is in the form:

\[
U_g(R_g(t)) = \begin{cases} 
\frac{k}{2} (R_g(t) - R_{g,max})^2 & \text{if } R_g(t) > R_{g,max} \\
\frac{k}{2} (R_g(t) - R_{g,min})^2 & \text{if } R_g(t) < R_{g,min} \\
0 & \text{otherwise} 
\end{cases}
\]

where \( k \) has to be large enough to prevent the peptide to get stuck in an extended/compressed conformation, \( R_g(t) \) is the value of the \( R_g \) at time \( t \), \( R_{g,max} \) and \( R_{g,min} \) are the maximum and minimum values of the \( R_g \) that we are interested in.

Computational details

During MD simulations each system was placed in a cubic simulation box such that all dendrimer/peptide/complex atoms were, at least, 1 nm distant from the box edges. The box was then filled with a sufficiently large number of SPC water molecules. The MD simulation box was neutralized by adding the appropriate number of \( \text{Cl}^- \) and \( \text{Na}^+ \) counterions at a physiological concentration (0.15 M). Systems were subsequently energy minimized under periodic boundary conditions. The equilibration of each system was first performed under the NVT canonical ensemble for 1000 ps. Initial velocities were generated according to a Maxwell-Boltzmann distribution corresponding to a temperature of 300 K. Pressure coupling was then switched on and the system was equilibrated under the NPT isothermal-isobaric ensemble for 1000 ps with a reference pressure of 1 bar. Production runs were carried out under NPT ensemble at 298.15 K and 1 bar for 200 ns. The coordinates were written to the output file every 10 ps for the final analysis. Newtonian equations of motion were integrated using the leapfrog scheme with a time step of 2 fs. Periodic boundary conditions were adopted in all cases. All bond lengths were constrained to their reference values employing the LINCS algorithm [40]. Electrostatic interactions were calculated with the PME method [41]. Atomic-based pairlist generation was used as implemented in the Verlet algorithm. When needed V-rescale [42] thermostat at 298 K and Parrinello-Rahman [43] barostat at 1 bar were used. This protocol is based on a systematic study that has been recently carried out, confirming that the present scheme is valid [31]. Details on the molecular systems considered in the present work are presented in Tables 1 and 2.

During clustering by conformation, we used the Gromos clustering algorithm by Daura et al. [44]. During docking, the binding mode that shows the lowest energy was chosen. The grid box during rigid docking was centered on the dendrimer center with a cubic grid box of side 26 Å. The grid box during flexible docking was centered on the site that found in the first docking with a cubic grid box of side 10 Å.

In Ang-(1-7) MTD, we used gaussian width of 0.1 nm, gaussian height of 1.2 kJ/mol, deposition time between gaussians of 500 MD time steps, a biasfactor of 15 and a temperature of 300 K. Convergence was reached in 1 µs according to the behaviour of c(t) (a time-dependent parameter in the definition of the free energy which makes

| Type       | GN     | pH  | Molecule atoms | Total atoms | Cl\(^-\) | Na\(^+\) | Waters | Box side (nm) |
|------------|--------|-----|----------------|-------------|--------|--------|--------|---------------|
| PNH\(^a\)  | 4      | Neutral | 1312           | 138360      | 192    | 128    | 45576  | 11.3          |
| PNH\(^a\)  | 4      | Acid  | 1374           | 125148      | 243    | 117    | 41138  | 11            |
| POH\(^a\)  | 4      | Neutral | 1184           | 118853      | 109    | 109    | 38917  | 10.7          |
| POH\(^a\)  | 4      | Acid  | 1246           | 122457      | 176    | 114    | 40307  | 10.8          |
| Ang-(1-7)\(^a\)  | Neutral | 85    | 5105           | 5           | 5      | 1670   | 3.8    |               |
| Ang-(1-7)\(^a\)  | Acid  | 87    | 5136           | 7           | 5      | 1679   | 3.8    |               |
the free energy asymptotically time-independent) and the diffusion of the collective variable [45] as presented on Fig. S7. Gromacs-2016.3 and Plumed-2.3.3 were used.

The number of contacts between two groups of atoms A and B is defined as:

\[ \sum_{i \in A} \sum_{j \in B} s_{ij} \]

where \( s_{ij} \) is 1 if the contact between atoms \( i \) and \( j \) is formed, zero otherwise. This was used to calculate dendrimer-peptide coordination number.

**Results and Discussion**

**MD of isolated systems in solution**

**PAMAM dendrimers**

The Root-mean-square deviation (RMSD) of atomic positions is a standard measure of the structural changes mainly used for proteins or non-protein small organic/inorganic molecules. Nevertheless, due to the large conformational flexibility of dendrimers in aqueous solution, there is a debate as to whether it is an appropriate molecular descriptor to account for its structural stability. Here, we have employed RMSD analysis to find the most converged part of the simulations, in order to obtain reasonable initial structures for the following steps. As shown in Fig 2a, the PNH\(^a\) RMSD values undergo large changes from the starting structure in the first 50 ns, afterwards the system seems to reach a more stable behavior; meanwhile PNH\(^a\) RMSD values undergo big changes from the starting structure in the first 20 ns, afterwards the system seems to reach a more stable behavior fluctuating around a mean value. Ang-(1-7)\(^a\) and Ang-(1-7)\(^n\) RMSD values show a more stable behavior from the first ns. According to these results, we agreed to use the last 100 ns of the simulations as the most equilibrated data for all the systems. Large changes observed on dendrimer structures are mainly due to the fact that the initial structures were obtained by PyPolyBuilder algorithm which only gives us an initial guess for the dendrimer geometry in vacuum.

The radius of gyration (\( R_g \)) might be a more appropriate molecular descriptor to account for the structural stability of dendrimers, it gives a measure for the compactness of a molecular structure. As shown in Fig. 2b, the PNH\(^a\) and PNH\(^n\) \( R_g \) values show a more stable behavior from the first nanoseconds around a mean value. POH\(^n\) \( R_g \) undergoes a rapid change during the first nanoseconds, afterwards it

| Complex | pH      | No. peptides | Total atoms | Cl\(^-\) | Na\(^+\) | Water molecules |
|---------|---------|--------------|-------------|---------|---------|-----------------|
| [PNH – A]\(^n\) | Neutral | 3            | 57563       | 118     | 54      | 18608           |
| [PNH – A]\(^a\) | Acid    | 3            | 63513       | 191     | 59      | 20541           |
| [POH – A]\(^n\) | Neutral | 3            | 28814       | 27      | 27      | 9107            |
| [POH – A]\(^a\) | Acid    | 3            | 60852       | 124     | 56      | 19720           |

**Table 2** Complexed molecular systems considered in the present work. P in the name corresponds to PAMAM, NH for amino terminated dendrimer, OH for hydroxyl terminated dendrimer, A for Ang-(1-7) followed by the number of peptides in complex with the dendrimer and the last number that identify the two complex conformers for each system.

![Fig. 2](image-url) Structural features of PAMAM-NH\(_2\), PAMAM-OH and Ang-(1-7) systems. a RMSD and b \( R_g \) values as function of time.
steadily deviates slowly from this new conformation, reaching a more steady-state after 100 ns. POH\(^n\) \(R_g\) values show that the dendrimer undergoes a rapid change during the first nanoseconds and afterwards fluctuates around a mean value; finally, Ang-(1-7)\(^n\) and Ang-(1-7)\(^a\) \(R_g\) values show a more stable behavior from the first nanoseconds around a mean value in both cases.

PNH\(^n\) and PNH\(^a\) \(R_g\) mean values in the most equilibrated zone were 1.94 ± 0.05 nm and 2.3 ± 0.04 nm, respectively. On the other hand, POH\(^n\) and PNOH\(^a\) \(R_g\) mean values were 1.46 ± 0.02 nm and 2.16 ± 0.06 nm, respectively. Results show that dendrimers at acidic pH assume a stretched, open configuration similar to the dense-shell model as a consequence of the strong Coulomb repulsion between the charged tertiary/primary amines present in the structure. \(R_g\) values are in very well agreement with experimental and theoretical results available [31, 46–51], as also compared in Table 3, confirming that GROMOS-compatible 2016H66 force field, together with our chosen methodology were capable to represent the structural properties of the dendrimers used in this work. It can be observed that POH\(^n\) and POH\(^a\) has in general a more compact structure (lower \(R_g\)) compared with PNH\(^n\) and PNH\(^a\), respectively (Fig. 2b). At neutral pH, this is partly due to the preference of the group terminals to be surrounded by waters (PNH\(^n\), Fig. 3a) or to interact with internal dendrimer groups (POH\(^n\), Fig. 3b), from the total possible interactions between group terminals and waters/internal dendrimer groups, we found that PNH\(^n\) forms 95% of the interactions with waters and 5% with internal groups, meanwhile, POH\(^n\) forms 85% of interactions with waters and 15% with the internal groups. The \(R_g\) and RMSD fluctuations in the case of PNH\(^n\), PNH\(^a\) and POH\(^n\) throughout the simulation time, compared to that of POH\(^n\), suggests that the formers have a more flexible structure than the latter. This result might be attributed to the continuous movement of the protonated terminal amine groups in PNH\(^n\) and terminal/internal protonated

Table 3  Radius of gyration \(R_g\) of PAMAM dendrimers and comparison with available experimental and theoretical studies in the literature

| System | \(R_g\) [nm] | \(2016H66\) [31] | GAFF [46] | CHARMM [47] | SANS [48] | SAXS [49] |
|--------|-------------|-----------------|------------|-------------|-----------|-----------|
| PNH\(^n\) | 1.94 ± 0.06 | 1.84 ± 0.03     | 2.06 ± 0.02 | 2.10 ± 0.04 | 2.13      | 1.86      |
| This work | 2016H66 [31] |                  |            |             |           |           |
| PNH\(^a\) | 2.3 ± 0.04  | 2.45             |            |             |           |           |
| This work | 2016H66 [31] |                  |            |             |           |           |
| POH\(^n\) | 1.46 ± 0.02 | 1.45 ± 0.01      |            |             |           |           |
| This work | AMBER99SBBL [50] |            | GAFF [51] |             |           |           |
| POH\(^a\) | 2.16 ± 0.06 |                 |            |             |           |           |

Fig. 3 Waters surrounding group terminals and hydrogen bonds formed between terminal groups and waters/dendrimer internal groups. a PAMAM-NH\(_2\) and b PAMAM-OH, at neutral pH
groups in PNH\textsuperscript{a} and POH\textsuperscript{a} to minimize the unfavorable enthalpic penalty due to electrostatic repulsion on charges of the same sign.

To characterize the shape of the dendrimers, an asphericity parameter $\delta$ was estimated. It is important to note that the closer this value is to zero, the more spherical the dendrimer becomes and its calculation is described in detail elsewhere [31]. As shown in Table 4 and Fig. 4, PNH\textsuperscript{n} assume a less spherical structure compared to POH\textsuperscript{n}, meanwhile, PNH\textsuperscript{a} and POH\textsuperscript{a} show an almost spherical behavior. This high asphericity observed in PNH\textsuperscript{n}, might be due to an asymmetry induced by a high degree of charged terminal groups back folding as also reported in the previous works [31], in contrast, the neutral terminal in POH\textsuperscript{n} seems to be able to accommodate in a more compact way due to its less charge-charge terminal repulsion and its preference of interactions with internal dendrimer groups. In the case of PNH\textsuperscript{a} and POH\textsuperscript{a}, an almost spherical shape might be mainly due to the repulsion between charged tertiary amine groups in the interior of the dendrimers.

To characterize how particle density (waters, terminal nitrogens, internal nitrogens and ions) varies as a function of the distance from the dendrimer center, radial distribution functions were measured. As shown in Fig. 6a, a higher level of structuration is evidenced by the pronounced peaks on the distributions on PNH\textsuperscript{n}, PNH\textsuperscript{a} and POH\textsuperscript{n} internal nitrogens compared with those of POH\textsuperscript{a}, this is mainly due to a more dense POH\textsuperscript{n} dendrimer compared with less dense, PNH\textsuperscript{n}, being more evident on PNH\textsuperscript{a} and POH\textsuperscript{a}, as evidenced by the height of the peaks. As it can be seen, the dense-core model [53] is confirmed at neutral pH, this model states that the monomer (internal Nitrogen atoms) density distribution is high close to their centers and decays as it approaches the periphery, as evidenced of the height of the peaks from the center to the periphery at neutral pH. However, at acidic pH, dendrimers assume a less dense inner cavity, stretched, open conformation like to the dense-shell model [53] as a consequence of the strong Coulomb repulsions between the charged units. This is in agreement with theoretical models that predicts that the PAMAM-NH\textsubscript{2} core becomes denser upon decreasing the acidity of the medium. Interestingly, this could led to a different drug interaction behaviour and load capacity, probably allowing better drug encapsulation in the big inner cavities at acidic pH (Fig. 5).

As shown in Fig 6b, PNH\textsuperscript{n} N-ter can be found in the intermediate zone (around 1 nm) as well as in the external

![Dendrimers shape comparison. a PAMAM-NH\textsubscript{2}\textsuperscript{n}, b PAMAM-OH\textsuperscript{n}, c PAMAM-NH\textsubscript{2}\textsuperscript{a} and d PAMAM-OH\textsuperscript{a} at 200 ns of MD simulation. Letters "a" and "n" states for neutral and acidic pH](image)
surface (around 2 nm), compared with the case of PNH\textsuperscript{n} where distribution is near the external surface (around 2.3 nm), and POH\textsuperscript{n} and POH\textsuperscript{a} O-ter which are mainly found on the surface (around 1.5 nm and 2.16 nm). In PNH\textsuperscript{n}, terminal groups exhibit a broader distribution that might indicate a certain degree of backfolding, being the backfolding increased from low to neutral pH, as shown by the intensity of the density peaks.

As shown in Fig 7a, RDF profiles reveal a major presence of buried water molecules within the structure of PNH\textsuperscript{n}, PNH\textsuperscript{a} and POH\textsuperscript{a} compared with POH\textsuperscript{n}, at distances larger than 5 Å from the center of mass of the dendrimer. Hydroxyl terminals when internal tertiary amines are not protonated reduce the penetration of water into the dendrimer structure, which can be interpreted as an enhancement of the hydrophobic character of the dendrimer inner small cavities, ideal for encapsulation of small hydrophobic drugs. In general, dendrimers with both kind of terminal groups at acidic pH show a major presence of buried waters within its structure in order to solvate the charged groups. As shown in Fig 7b-c, RDF profiles reveal that chloride ions penetrate the innermost dendrimer cavities in PNH\textsuperscript{n}, PNH\textsuperscript{a} and POH\textsuperscript{a}, contrary to the case of POH\textsuperscript{n}. Interestingly, a broad peak around 1 nm and a shoulder around 2 nm, where PNH\textsuperscript{n} positive terminals are localized, was found. Sodium counterions are found as far as possible of charged terminals in PNH\textsuperscript{n}, PNH\textsuperscript{a} and POH\textsuperscript{a} and more disperse in POH\textsuperscript{n} around its surface.

**Ang-(1-7) bioactive peptide**

Ang-(1-7)\textsuperscript{n} and Ang-(1-7)\textsuperscript{a}, R\textsubscript{g} mean values in the equilibrated zone were 0.65 ± 0.04 nm and 0.57 ± 0.05 nm, respectively (Fig. 2b). From the structural point of view, the solution structure of Ang-(1-7) was previously determined [54] by means of NMR and circular dichroism experiments performed at acidic pH. There, the authors claimed that Ang-(1-7) structure showed a conformational equilibrium

---

**Table 4** Asphericity \(\delta\) of PAMAM dendrimer and comparison with theoretical studies available. Letters “n” and “a” states for neutral and acidic pH.

| System | This work | 2016H66 [31] | AMBER [52] | GAFF [46] |
|--------|-----------|--------------|-------------|-------------|
| PNH\textsuperscript{n} | 0.04 | 0.08 ± 0.03 | 0.02 | 0.03 |
| PNH\textsuperscript{a} | 0.004 | 0.02 ± 0.01 | 0.01 | 0.01 |
| POH\textsuperscript{n} | 0.01 | 0.01 | 0.006 |
| POH\textsuperscript{a} | 0.006 |  |  

---

**Fig. 5** Dendrimers cavities comparison. 
(a) PAMAM-NH\textsubscript{2}\textsuperscript{n}, 
(b) PAMAM-OH\textsubscript{2}\textsuperscript{n}, 
(c) PAMAM-NH\textsubscript{2}\textsuperscript{a} and 
(d) PAMAM-OH\textsubscript{2}\textsuperscript{a} at 200 ns. Letters “n” and “a” states for neutral and acidic pH.

---

\(\odot\) Springer
between the random coil and β-sheet (or a mixture of bend structures), with a bend stabilized by interactions between residues VAL3 and TYR4, this is in very well agreement with our results in Fig. 8, where we can see that the most frequent secondary motif is bent between VAL3 and TYR4. In our classical MD simulations at acidic pH, we found that 60% of the structures were similar to the NMR solution structure (RMSD < 2 Å when superimposed), meanwhile, at neutral pH, this happened only in 40% of the frames, thus, this conformation was less frequent at neutral pH (Fig. 9).

The conformational sampling of peptides in solution that are fully or partially disordered and flexible is not always guaranteed with conventional MD due to the limitations in the sampling time scale [55]. To get more insights about Ang-(1-7) heptapeptide structure and extend this insight at neutral pH conditions, we performed a MTD simulation at neutral pH employing the Rg and the C_d−a as CVs. The final free energy landscape is shown in Fig. 10a, where it can be shown that the landscape has a broad and unique minimum or basin A, indicating that this peptide exist in a variety of
conformations in solution. The global minima corresponds to a $R_g$ of 0.65 nm and a $C_{d-a}$ of 30, in very well agreement with $R_g$ values from classical MD; however, the basin A samples a wide range of $R_g$ and $C_{d-a}$ values, this is a $R_g$ in the region spanning from 0.6 to 0.7 nm and contacts ranging between 30 and 35. Interestingly, the structures in the basin showed a persistent HB between ASP1 and TYR4 or between ASP1 and VAL3 (Fig. 10b). This HB was also frequently observed on the bigger cluster of the MD of Ang-(1-7)n in solution, this conformation is probably frequent because it allows positive charged ARG2 to be far from positive charged N-terminal, both of them becoming able to interact with polar waters.

The one dimensional FES as a funcion of $R_g$ is shown in Fig. 10c, as can be seen, there are not large free energy barriers (around 1 kT) dividing the metastable states in the main basin A. It is important to note that only 35 % of the structures in basin A were similar to the NMR structure and in agreement with classical MD. This result is coherent with the conformational equilibrium between the random coil and a mixture of bend structures reported experimentally for Ang-(1-7). However, basin A is characterized by a significant degree of flexibility of the N-terminal and C-terminal tails which samples multiple conformations that deviate from the ensemble of the NMR structure as shown in Fig. 11.

Together, the $R_g$, $\delta$ and RDFs results validates our chosen methodology and confirms that GROMOS-compatible 2016H66 force field is capable to model the theoretical and experimental structural features of G4-PAMAM dendrimers with amino or hydroxyl group terminals. In case of Ang-(1-7), structural super impositions of MD and metadynamics structures with NMR structure confirms the experimental results previously reported.

**Clusters and docking**

The middle structure of the most populated clusters for dendrimers and the peptide were used for the molecular docking of each dendrimer conformer with each of the peptide conformers. Our methodology allowed us to find a suitable binding site for the peptide by using a representative peptide conformation taken from the MD, once the peptide finds a reasonable site, the algorithm allows it to better accommodate into this local site, as can be schematized in Fig. 12.
Final complexes resulted from the docking are shown in Fig. 13. As can be seen, PNH<sup>n</sup> allows the interaction of the peptides with its inner shells as well as with its surface. POH<sup>n</sup>, only allow the peptide interactions with its surface due to its more compact structure. We can observe that at acidic pH, dendrimers allow a better encapsulation of 3 peptides. Details on the HBs formed between peptides and dendrimers are presented in Tables 7 and 5.

### Complexes stability

**PAMAM-NH/Ang-(1-7) complex**

To identify global structural changes due to complex formation, the RMSD and the R<sub>g</sub> at bound state were measured and compared with that of the free structures. It can be seen in Fig. 14ac that the binding of peptides stabilizes the
Fig. 12  Double-docking schematic representation. Each of the peptides conformer is allowed to find a site by using a blind rigid docking, afterwards, once the peptide found a reasonable site, a flexible docking allows it to better accommodate into the site.

Fig. 13  MD complexes initial structures obtained from double-docking approach: a) PNH\textsuperscript{n} in complex with 3 Ang-(1-7)\textsuperscript{n} peptides, b) POH\textsuperscript{n} complex with 3 Ang-(1-7)\textsuperscript{n} peptides, c) PNH\textsuperscript{n} in complex with 3 Ang-(1-7)\textsuperscript{n} peptides and b) POH\textsuperscript{n} at in complex with 3 Ang-(1-7)\textsuperscript{n} peptides.
dendrimer structure according to RMSD values. Meanwhile, as can be seen in Fig. 14bd, the radius of gyration of the dendrimer and the peptides are not importantly affected by the binding.

To test complex stability, the distance between the center of mass (COM) of the dendrimer and each peptide was measured. As showed in Figs. 15a, 16a, and 18a, under neutral pH conditions, 2 peptides remained stable bonded during the simulation in the dendrimer internal domain around a distance of 1 nm and a third peptide remained stable bonded at the dendrimer surface around a distance of 3 nm, peptides remain mainly in the same initial sites. The dendrimer kept an open hourglass geometrical shape during the whole simulation time, allowing at least two peptides to be near the core. As shown in Figs. 15d and 22b, under acidic pH conditions, from the 3 peptides initially at around 1 nm in the dendrimer internal domain, one remained very close to the core during most of the simulation time, two went out very quickly and stayed near the superficial zone, even leaving the dendrimer at certain points. From the two peptides in the dendrimer surface, one went to the back of the core at the last 50 ns of the simulation, interacting with the other peptide in the core. It appears that for two peptides, transitions from in to out the dendrimer had low energetic barriers or was an unstable state, in such a way that it is easy to get out, meanwhile, the third peptide appears to be in a stable deep minimum that forces it to stay close to the dendrimer core, but further research is required to confirm this hypothesis, due to the system flexibility, a MTD simulation could give us more insight about this process. Whith these results we can conclude that only one peptide remains stably bonded at the core of the dendrimer at acidic pH during the whole MD simulation.

To test the protection capabilities of each dendrimer, the percent coverage of peptides due to binding, was measured. The free peptide in solution was taken as a reference, representing a molecule that is 100% accessible to the solvent. As presented in Fig. 15b, in the case of the [PNH-A]n, an average of 54% of coverage was found, confirming PNHn drug protection capabilities. As presented in Fig. 15e, in the case of the [PNH-A]a, an average of 20% of coverage was found, a much lower percentage than in the case of neutral pH. This is reasonable considering that 2 peptides are in the superficial zone during most of the simulation time and even leave the dendrimer at certain points. If we consider only the peptide that remains at the core, it still has less coverage percentage compared with the ones at the core at neutral pH, probably because due to the open structure at acidic pH, cavities are also deferentially hydrated. It could mean that the peptide might be a little more accessible to water but due

**Fig. 14** Structural stability changes of peptide or PAMAM-NH upon binding. RMSD of structures in complex compared with same structures free in solvent for a neutral pH and c acidic pH. Rg of structures in complex compared with same structures free in solvent at b neutral pH and d acidic pH
Fig. 15 ([PNH-A]) stability. **a** Distance from dendrimer COM to Ang-(1-7) peptides, **b** peptide coverage according to SASA values and **c** number of hydrogen bonds between dendrimer and peptides. # B-D refers to the different Ang-(1-7) peptides bonded to dendrimers. Grey dashed line represents the $R_g$ of the dendrimer as a reference for dendrimer periphery.

Fig. 16 PAMAM-NH/Ang-(1-7) coordination number along simulation time at **a** neutral pH and **b** acidic pH. Coordination mean the number of atoms from the peptides that are within 0.3 nm of the dendrimer. # B-D refers to the different Ang-(1-7) peptides bonded to dendrimers. Grey dashed line represents the threshold agreed that divides coupled from uncoupled states. A Bezier based smooth curve was applied for the sake of visibility.
to its localization inside the dendrimer, it is not necessarily accessible to peptides.

The peptide protection is especially relevant for bonds susceptible to hydrolytic attack by endopeptidases, as schematized in Fig. 17. Once formed, Ang-(1-7) is rapidly hydrolyzed, especially by ACE [56] and dipeptidyl peptidase 3 (DPP3) [11]. Here, we found a 46% of [PNH-A]° coverage on the region attacked by DPP3 and a 73% of [PNH-A]° coverage on the region attacked by ACE. This result is important because the very short half-life of Ang-(1-7) in the circulation is primarily accounted for peptide metabolism by ACE [57].

The main type of HBs formed between Ang-(1-7) and hydroxyl-terminated PAMAM dendrimer are presented in Fig.s S11 - S12, the number of NHs through simulation time is presented in Fig. 15c and detailed occupancy in Tables 5 and 6. At neutral pH, a number of 5.4 HBs in average are formed between PNH° and Ang-(1-7) peptides, during the whole simulation time. The most populated HBs interactions are between core atoms, internal branches and terminal groups of PNH° and Ang-(1-7) peptides, during half of the simulation time; all the peptide keeps its interaction between its negative charged C-terminal and the main Nitrogen of the positively charged primary amine in the dendrimer terminal groups during at least 40% of the time and also side chain of TYR4 interacts with the amide OH group from the internal dendrimer as an acceptor during half of the simulation time; all the peptide keeps its interaction between its negative charged C-terminal and the main Nitrogen of the positively charged primary amine in the dendrimer terminal groups during almost half of the simulation time; ASP1 negative side chain interacts with the protonated primary amine of the dendrimer terminal groups during at least 40% of the time and also side chain of TYR4 interacts with the amide NH group from the internal dendrimer.

Table 5 Main interactions (HBs) formed between Ang-(1-7) peptide and PNH°. Only occupancies above 35% are shown. SegA corresponds to dendrimer and SegB-SegD to peptides

| Donor            | Acceptor            | Occupancy (%) |
|------------------|---------------------|---------------|
| SegC-TYR4-Side-OH| SegA-INTR1-Main-O   | 53.39         |
| SegA-TYR1-Main-N | SegB-PRO7-Side-O2   | 52.70         |
| SegA-TYR1-Main-N | SegD-PRO7-Side-O1   | 50.30         |
| SegA-TYR1-Main-N | SegC-PRO7-Side-O1   | 49.73         |
| SegA-TYR1-Main-N | SegC-PRO7-Side-O2   | 48.61         |
| SegA-TYR1-Main-N | SegB-PRO7-Side-O1   | 48.60         |

According to Fig. 19a, at neutral pH, TYR4 and ILE5 residues, interacted preferably with internal dendrimer groups rather than with the terminal groups, which was expected due to its hydrophobic/partially hydrophobic nature. On the other hand, ASP1 and PRO7 residues, interacted preferably with terminal dendrimer groups rather than with the internal groups, which was expected due to its negative charged side-chain/C-terminal groups. ARG2, VAL3 and HIS6 interacts similarly with terminal or internal dendrimer groups but in a less frequent way. As expected, arginine residue was more exposed to solvent due to its charged aminoacid side chain. According to Fig. 19b, at acidic pH, ASP1, ARG2, TYR4 and HIS6 keep in contact preferably with internal dendrimer groups. Due to a more open dendrimer cavities, ASP1 and ARG2 are able to enter and interact with internal polar groups compared with the neutral case. Due to its hydrophobic/partially hydrophobic nature TYR4 keeps in contact preferably with internal dendrimer as in the neutral case but less frequently. HIS6 at acidic pH is considered as positively charged, while at neutral pH is neutral, this change modify its preference to be in contact

Table 6 Main interactions (HBs) formed between Ang-(1-7) peptide and PNH°. Only occupancies above 10% are shown. SegA corresponds to dendrimer and SegB-SegD to peptides

| Donor            | Acceptor            | Occupancy (%) |
|------------------|---------------------|---------------|
| SegA-INTR1-Main-N| SegC-ASP1-Side-OD1  | 38.15         |
| SegA-INTR1-Main-N| SegC-ASP1-Side-OD2  | 36.29         |
| SegA-INTR1-Main-N| SegB-ASP1-Side-OD2  | 34.73         |
| SegA-INTR1-Main-N| SegD-ASP1-Side-OD2  | 32.99         |
| SegA-INTR1-Main-N| SegD-ASP1-Side-OD1  | 32.35         |
| SegA-INTR1-Main-N| SegB-ASP1-Side-OD1  | 31.09         |
with internal dendrimer groups instead with positively charged terminal groups. Finally, VAL3, ILE5 and PRO7 hydrophobic residues keep almost no contact with the dendrimer, here C-terminal is neutral compared with the neutral case.

Taking into account the number of peptides stably bonded near the core, the higher percent coverage of peptides from water, the major number of HBs between dendrimer and peptides during the whole simulation time, the frequency in which this HBs occur and the peptide and dendrimer frequency of contacts, it is likely that amino terminated PAMAM dendrimer can act as a better encapsulator near neutral pH and as a release agent under acidic pH. However, further studies are needed in this direction.

**PAMAM-OH/Ang-(1-7) complex**

We measured RMSD as a kind of first indicator of stability upon complex formation. It can be seen in Fig. 20ac that the binding of peptides slightly stabilizes the dendrimer structure at neutral pH but, it does not happen at acidic pH. Besides that, the structure of the peptides are not greatly affected by the binding to the dendrimer. For their part, the $R_g$ of either the dendrimer or the peptide are not importantly affected by the binding. Higher fluctuations on dendrimer RMSD and $R_g$ at acidic pH correspond to the continuous movement of the protonated groups which makes it difficult to be stabilized by ligand binding.
To further evaluate complex stability, the distance between the Center Of Mass (COM) of the dendrimer and each peptide was measured. As shown in Fig. 21a, under neutral pH conditions, all the three Ang-(1-7) peptides remained at a stable distance of 1.6 ± 0.1 nm from the dendrimer COM, this is, mostly on the dendrimer surface taking into account that the $R_g$ is of 1.46 nm, as also illustrated in the timeline evolution of frames in Fig. 23a.

![Fig. 19](image1.png)  
**Fig. 19** a Time evolution of [PNH-A]$^n$ structure and b occupancy percentage of the amino acids in the final 100 ns of MD simulation, determined by considering the contacts per amino acids at 3.0 Å from the dendrimer terminal groups (red bars) and to the dendrimer internal groups (blue bars). Occupancy denotes the average percentage of the three ang-(1-7) peptides that remained stable within the dendrimer.

![Fig. 20](image2.png)  
**Fig. 20** Structural stability changes of peptide or PAMAM-OH upon binding. RMSD of structures in complex compared with same structures free in solvent for a neutral pH and c acidic pH. $R_g$ of structures in complex compared with same structures free in solvent at b neutral pH and d acidic pH.

To further evaluate complex stability, the distance between the Center Of Mass (COM) of the dendrimer and each peptide was measured. As shown in Fig. 21a, under neutral pH conditions, all the three Ang-(1-7) peptides remained at a stable distance of 1.6 ± 0.1 nm from the dendrimer COM, this is, mostly on the dendrimer surface taking into account that the $R_g$ is of 1.46 nm, as also illustrated in the timeline evolution of frames in Fig. 23a.
A closer view into the simulations shows that peptides in \([\text{POH-A}]^n\), explore the nearby area around its initial binding site, which could be due to the symmetry of the dendrimer groups. Moreover, the spherical compacted shape of the dendrimer appears to avoid peptide’s complete internalization. The above results might imply that the energetic barriers between superficial binding sites are low, allowing the peptide to go from one local minimum to another, in contrast, the energy barrier needed to leave the dendrimer appears to be high enough, keeping the peptides interacting with the dendrimer as shown in Fig. 22a, but further investigation is needed to confirm energetic landscape of the process. In the case of \([\text{POH-A}]^n\) in Fig. 21d and Fig. 23b, all the three peptides started at the internal dendrimer region around 1.2 nm, quickly both of them move into the periphery, around the \(R_g\) of 2.3 nm, only transiently re-entering to the dendrimer interior. Conversely, the third peptide stays in the dendrimer interior and only transiently moves to the periphery. The number of contacts between the dendrimer and the peptides are presented in Fig. 22b, were we can observe how the three peptides unbind and bind several times during the MD simulation.

For testing the protection capabilities of each dendrimer, the percent coverage of peptides due to binding, was measured. As presented in Fig. 21b, in the case of \([\text{POH-A}]^n\) protection, the peptides showed a dendrimer coverage of around 44\%. As presented in Fig. 21e, in the case of \([\text{POH-A}]^n\) protection, the peptides showed a dendrimer coverage of around 30\%, lesser coverage than in the case of neutral pH. At neutral pH, where peptides coverage is bigguer, we found a 55\% of \([\text{POH-A}]^n\) coverture on the region attacked by DPP3 and a 57\% of \([\text{POH-A}]^n\) coverture on the region attacked by ACE.

The main type of HBs formed between Ang-(1-7) and amino-terminated PAMAM dendrimer are presented in Figs S13 -S14, the number of hydrogen bonds (NHBs) through simulation time is presented in Fig. 21cf and detailed occupancy in Tables 7 and 8. At neutral pH, a number of 4 HBs in average are formed between POh\(^n\) and Ang-(1-7) peptides, during the whole simulation time. It was found until 144 different HBs between POh\(^n\) and peptides, however they have
Fig. 22 PAMAM-OH/Ang-(1-7) coordination number along simulation time at a neutral pH and b acidic pH. Coordinations mean the number of atoms from the peptides that are within 0.3 nm of the dendrimer. # B-D refers to the different Ang-(1-7) peptides bonded to dendrimers. Grey dashed line represents the threshold agreed that divides coupled from uncoupled states. A Bezier based smooth curve was applied for the sake of visibility.

Fig. 23 Time evolution of PAMAM-OH/Ang-(1-7) complex structures at a neutral pH and b acidic pH.
low % of occupancy (below 36 %), suggesting a low specificity of these bonds. The most populated HBs interactions are between internal branches and terminal groups of POH with PRO7, ASP1 and TYR4 aminoacids in the peptide, no interactions with core are present above 19 % of occupancy. At acidic pH, a number of 3 HBs in average are formed between POH and Ang-(1-7) peptides, during the whole MD simulation time. It was found until 139 different HBs between POH and peptides, however, only ASP1 and TYR4 with internal branches have considerable % of occupancy. The most populated HBs interactions are between internal branches of POH with charged side chain of ASP1 and backbone of TYR4 in the peptide, no interactions with core and terminal groups are present above 19 % of occupancy. At neutral pH, the characteristic ASP1-TYR4/ASP1-VAL3 HB found in Ang-(1-7) metadynamics is still frequently formed (53 % of the simulation time) compared with its formation free in solvent (40 % of the simulation time), this imply that these intra peptide HBs are at least equally broken but this is not necessarily in favor of a peptide/dendrimer interaction (Table 7), it is also because part of the time the residue is solved exposed and interacts with waters.

According to Fig. 24a, at neutral pH, TYR4 interacts preferably with internal dendrimer groups rather than with the terminal groups, which was expected due to its partially hydrophobic aromatic nature which prefers to be buried in a hydrophobic core and its polar hydroxyl (-OH) group also being able to interact with polar groups in the dendrimer. Particularly, the inclusion of the TYR residue is favored in both dendrimer types, as in the case of β-cyclodextrin complexes [14]. In the other hand, ASP1, ARG2, VAL3 and PRO7 residues, stay in contact slightly more with terminal dendrimer groups rather than with the internal groups, in case of negative charged ASP1 and C-terminal PRO7 with polar hydroxil terminal group in the dendrimer, arginine residue was in contact with terminal groups in the dendrimer and exposed to solvent due to its charged aminoacid side

![](image)

**Fig. 24** Occupancy percentage of the amino acids in the final 100 ns of MD simulation at a neutral pH and b acidic pH. Determined by considering the contacts per amino acids at 3.0 Å from the dendrimer terminal groups (red bars) and to the dendrimer internal groups (blue bars). Occupancy denotes the average percentage of the three ang-(1-7) peptides that remained stable within the dendrimer

### Table 7

Main interactions (HBs) found between Ang-(1-7) peptide and POH at neutral pH. Only occupancies above 19% are shown. SegA correspond to dendrimer and SegB-SegD with peptides

| Donor                  | Acceptor             | Occupancy (%) |
|------------------------|----------------------|---------------|
| SegA-INTR1-Main-N      | SegC-PRO7-Cterm-O1   | 35.09         |
| SegA-TER1-Main-O       | SegC-PRO7-Cterm-O2   | 33.50         |
| SegA-INTR1-Main-N      | SegC-PRO7-Cterm-O2   | 22.27         |
| SegA-TER1-Main-N       | SegC-PRO7-Cterm-O2   | 21.82         |
| SegD-TYR4-Side-OH      | SegA-INTR1-Main-O    | 19.92         |
| SegA-TER1-Main-O       | SegD-ASP1-Side-OD2   | 19.74         |
| SegB-TYR4-Side-OH      | SegA-INTR1-Main-O    | 19.31         |
| SegA-TER1-Main-O       | SegD-ASP1-Side-OD1   | 19.30         |

### Table 8

Main interactions (HBs) formed between Ang-(1-7) peptide and POH at acidic pH. Only occupancies above 19% are shown. SegA correspond to dendrimer and SegB-SegD to peptides

| Donor                  | Acceptor             | Occupancy (%) |
|------------------------|----------------------|---------------|
| SegA-INTR1-Main-N      | SegD-ASP1-Side-OD1   | 50.88         |
| SegA-INTR1-Main-N      | SegD-ASP1-Side-OD2   | 49.80         |
| SegA-INTR1-Main-N      | SegB-ASP1-Side-OD1   | 40.68         |
| SegA-INTR1-Main-N      | SegB-ASP1-Side-OD2   | 40.39         |
| SegA-INTR1-Main-N      | SegC-ASP1-Side-OD1   | 39.36         |
| SegA-INTR1-Main-N      | SegC-ASP1-Side-OD2   | 39.32         |
| SegB-TYR4-Main-N       | SegA-INTR1-Main-O    | 19.88         |

At acidic pH, a number of 3 HBs in average are formed between POH and Ang-(1-7) peptides, during the whole MD simulation time. It was found until 139 different HBs between POH and peptides, however, only ASP1 and TYR4 with internal branches have considerable % of occupancy. The most populated HBs interactions are between internal branches of POH with charged side chain of ASP1 and backbone of TYR4 in the peptide, no interactions with core and terminal groups are present above 19 % of occupancy. At neutral pH, the characteristic ASP1-TYR4/ASP1-VAL3 HB found in Ang-(1-7) metadynamics is still frequently formed (53 % of the simulation time) compared with its formation free in solvent (40 % of the simulation time), this imply that these intra peptide HBs are at least equally broken but this is not necessarily in favor of a peptide/dendrimer interaction (Table 7), it is also because part of the time the residue is solved exposed and interacts with waters.
chain. ILE5 and HIS6 keep in contact similarly with terminal or internal dendrimer groups. At acidic pH, ASP1, ARG2, VAL3 and TYR4 interact preferably with internal dendrimer groups. ILE5, HIS6 and PRO7 interact equally with internal and terminal groups but in a less frequent manner.

Taking into account the number of peptides stable bonded at the surface, the higher percent coverage of peptides from water, the slightly major number of HBs between dendrimer and peptides during the whole simulation time, the frequency in which this HBs occur and the peptide and dendrimer frequency of contacts, it is likely that hydroxyl terminated PAMAM dendrimer can act as a Ang-(1-7) carrier at neutral pH and as a release agent under acidic pH. However, further experimental studies are needed in this direction.

So far, it appears that POH is able to interact with 3 peptides but not necessarily encapsulate them, however, this is valid for the case when dendrimer and peptide are prepared at neutral pH separately and afterwards put together. It remains still the question if the encapsulation is reached under different conditions, what happen with the complex when pH becomes neutral? In order to address this question, we designed a new complex (complex 2) by using an open dendrimer structure (from the non-equilibrated region of the dendrimer in solution), two peptides were placed in the internal cavities by our double-docking approach. This complex evolved in an MD during 200 ns, evolution of the peptides is showed in Fig. S7. As can be seen, initially, peptides are close from the core in the internal voids of the dendrimer, eventually the dendrimer becomes more and more compact and finally, the peptides remain interacting at the dendrimer surface. This gives us an idea of what could happen if the encapsulation is previously done, the complex at neutral pH evolve to a situation similar to the presented previously (Fig. 24a), dendrimer prefers a compact structure and peptides interact mostly with external surface. This is the case of reference [25], where dendrimer initial open structure for complex formation appears to be taken from a non-equilibrated structure, eventually dendrimer becomes more and more compact, even releasing one of the peptides, but at the end of the 60 ns MD productions, simulations does not necessarily reach convergence and could show a metastable state where peptides are partially encapsulated. Here, we show that in the last 100 ns of the MD production, most part of the peptides are interacting with external dendrimer parts whether it started with the peptides in or out, with slightly differences. Details are showed in Fig. S8.

Conclusions

Structural parameters such as the radius of gyration, the asphericity and the radial distribution function confirmed that the GROMOS-compatible 2016H66 force field, together with our chosen methodology are capable to model the theoretical and experimental structural features of PAMAM dendrimers evaluated in this work.

The classical MD simulations of PAMAM dendrimers in solution showed that hydroxyl-terminated PAMAM dendrimer at neutral pH has a more compact, less structured, more spherical, less hydrated and less flexible structure than amino-terminated PAMAM dendrimer. These results emphasises how a single change in terminal group could lead to differential behaviour of dendrimer structure and eventually in Ang-(1-7) binding. Also, this structural information is envisioned to prove useful for the encapsulation of other drugs.

The classical MD simulations of Ang-(1-7) in solution yielded to conformations in which a bend secondary structure is stabilized by interactions between residues VAL3 and TYR4, which is in agreement with NMR experimental studies. It was also found that at acidic pH, 60% of the Ang-(1-7) conformations were similar to the NMR solution structure, meanwhile, at neutral pH, this also happened but in a slightly less frequent percentage of conformations.

In the accelerated MD metadynamics simulations, Ang-(1-7) free energy surface showed a unique broad free energy minimum on the radius of gyration and donor-acceptor contact phase space, confirming the Ang-(1-7) flexibility due to its peptide nature. One dimensional free energy surface as a function of radius of gyration showed that free energy barriers between metastable states in the main basin are on the order of 1 kT, thus easily passed.

At neutral pH, PAMAM dendrimers with both terminal types are able to interact with at least 3 peptides in a stable way, with a coverage of up to 70%. However, hydroxyl-terminated PAMAM dendrimer only partially encapsulate the peptides due to its compact structure.

At acidic pH, PAMAM dendrimers with both terminal types are still able to interact with positively charged Ang-(1-7) peptides, either internalized or in its periphery, however, contacts, coverage and HBs are lesser than at neutral pH, suggesting a state for peptide release.

With the results found so far, it appears that amino-terminated PAMAM dendrimer at neutral pH posses slightly better features to load and protect Ang-(1-7) peptides than hydroxyl-terminated PAMAM dendrimer. However, either hydroxyl-terminated PAMAM dendrimer as well as amino-terminated PAMAM dendrimer are able to bind Ang-(1-7) peptides in a differential form, suggesting that hydroxyl-terminated PAMAM dendrimer is able to act as nanocarrier/nanotransporter more than nanoprotector of this peptide.

For the PAMAMNH(OH)/Ang-(1-7) complexes, the therapeutic mechanism could be encapsulate or capture the peptide at near neutral pH and deliver them at acidic pH, where repulsions between protonated primary and/or tertiary amines give rise to a open and hydrated environment.
allowing positively charged peptides to be solvated and released easily.

Finally, we propose that the complexes studied here should be experimentally tested to validate the properties predicted through In Silico approaches, as potential therapeutic agents.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s10822-022-00470-5.

Acknowledgements We acknowledge the use of the computational nodes at the Laboratory for the Design and Development of New Drugs and Biotecnological Innovation, Escuela Superior de Medicina, Instituto Politécnico Nacional.

Author Contributions CRB, JCB and MMA conceived, conceptualized and supervised the project. LAC designed the simulations and developed the code used in the double-docking approach. LAC and SA analyzed the results. SA generated and analyzed the interactions diagrams. LAC performed the simulations and drafted the manuscript. MC, CRB, MMA and SA review and edited the manuscript. All authors read and approved the final manuscript.

Funding LAC was supported by a postdoctoral fellowship from Consejo Nacional de Ciencia y Tecnología (CONACyT).

Data availability Code for this work are available in https://github.com/AmericaChi/Double-docking-approach. Metadynamics simulations files in PLUMED-NEST repository: https://www.plumed-nest.org/eggs/22/029/

Declarations Conflict of interest The authors have no competing interests to declare that are relevant to the content of this article.

References

1. Shylesh CM, Arya VS, Kanthalal SK, Uma Devi P (2020) Renin-angiotensin system modulators in COVID-19 patients with hypertension: friend or foe? Clin Exp Hypertens 44:1–10
2. Bader M, Alenina N, Young D, Santos RA, Touyz RM (2018) The meaning of Mas. Hypertension 72:1072–1075
3. Muslim S, Nasrin N, Alotaibi FO, Prasad G, Singh SK, Alam I, Mustafa G (2020) Treatment options available for COVID-19 and an analysis on possible role of combination of rhACE2, angiotensin (1–7) and angiotensin (1–9) as effective therapeutic measure. SN Comprin Clin Med 2:1761–1766
4. Soto M, DiZerega G, Rodgers KE (2020) Countermeasure and Biotechnological Innovation, Escuela Superior de Medicina, Instituto Politécnico Nacional.
5. Inmanpour H, Rezaee H, Nouri-Vaskeh M (2020) Angiotensin 1–7: a novel strategy in COVID-19 treatment. Adv Pharm Bull 10:488–675
6. Peiró C, Moncada S (2020) Substituting angiotensin-(1–7) to prevent lung damage in SARS-CoV-2 infection? Circulation 141:1665–1666
7. Others et al (2021) Combination of angiotensin (1–7) agonists and convalescent plasma as a new strategy to overcome angiotensin converting enzyme 2 (ACE2) inhibition for the treatment of COVID-19. Front Med 8:278
8. ClinicalTrials.gov. Use of Angiotensin-(1-7) in COVID-19. 2022. https://clinicaltrials.gov/ct2/show/NCT04633772
9. Iusuf D, Henning RH, van Gilst WH, Roks AJ (2008) Angiotensin-(1–7): pharmacological properties and pharmacotherapeutic perspectives. Eur J Pharmacol 585:303–312
10. Petty WJ, Miller AA, McCoy TP, Gallagher PE, Tallant EA, Torti FM (2009) Phase I and pharmacokinetic study of angiotensin-(1–7), an endogenous antiangiogenic hormone. Clin Cancer Res 15:7398–7404
11. Waster A, Devocelle M, Tallant E, Chappell MC, Gallagher PE, Paradisi F (2017) Stabilization of Angiotensin-(1–7) by key substitution with a cyclic non-natural amino acid. Amino Acids 49:1733–1742
12. Durik M, van Veghel R, Kuipers A, Rink R, Haas J, Jinhom Akanbi M, Moll G, Danser A, Roks AJ (2012) The effect of the thioether-bridged, stabilized Angiotensin-(1–7) analogue cyclic ang(1–7) on cardiac remodeling and endothelial function in rats with myocardial infarction. Int J Hypertension 2012:1
13. Kluskens LD, Nelemans SA, Rink R, de Vries L, Meter-Arkema A, Wang Y, Walther T, Kuipers A, Moll GN, Haas M (2009) Angiotensin-(1–7) with thioether bridge: an angiotensin-converting enzyme-resistant, potent angiotensin-(1–7) analog. J Pharmacol Exp Therapeut 328:849–854
14. de Lima GF, Pilo-Veloso D, Heine T, Duarte HA, Santos RA, Sinisterra RD (2007) Study of angiotensin-(1–7) vasoactive peptide and its b-cyclodextrin inclusion complexes: Complete sequence-specific NMR assignments and structural studies. Peptides 28:2199–2210
15. Silva-Barcellos NM, Frézard F, Caligiore S, Santos RA (2001) Long-lasting cardiovascular effects of liposome-entrapped angiotensin-(1–7) at the rostral ventrolateral medulla. Hypertension 38:1266–1271
16. Tijmsa EJ, Djriessen-Levela A, Hendriks M (2007) Angiotensin-(1–7) eluting polymer-coated medical device to reduce restenosis and improve endothelial cell function. US Patent 7,176,261
17. Others et al (2019) In silico search, chemical characterization and immunogenic evaluation of amino-terminated G4-PAMAM-HIV peptide complexes using three-dimensional models of the HIV-1 gp120 protein. Colloids Surf. B 177:77–93
18. Vello M, Rodriguez-Fonseca RA, Correa-Basurto J (2020) Complexation of peptide epitopes with G4-PAMAM dendrimer through ligand diffusion molecular dynamic simulations. J Mol Graph Model 96:107514
19. Others et al (2014) How to study dendrimers and dendriplexes III. Biodistribution, pharmacokinetics and toxicity in vivo. J Control Release 181:40–52
20. Fox LJ, Richardson RM, Briscoe WH (2018) PAMAM dendrimer-cell membrane interactions. Adv Colloid Interface Sci 257:1–18
21. Janaszewska A, Lazniewska J, Trzepiński P, Marcinkowska M, Krajnert-Maculewicz B (2019) Cytotoxicity of dendrimers. Biomolecules 9:330
22. Otto DF, De Villiers MM (2018) Poly (amidoamine) dendrimers as a pharmaceutical excipient. Are we there yet? J Pharm Sci 107:75–83
23. Li X, Naeem A, Xiao S, Hu L, Zhang J, Zheng Q (2022) Safety challenges and application strategies for the use of dendrimers in medicine. Pharmaceutics 14:1292
24. Agashe HB, Dutta T, Garg M, Jain N (2006) Investigations on the toxicological profile of functionalized fifth-generation poly (propylene imine) dendrimer. J Pharm Pharmacol 58:1491–1498
25. Márquez-Miranda V, Abrigo J, Rivera JC, Araya-Duran I, Aravena J, Simon F, Pacheco N, González-Nilo FD, Cabello-Verrugio C (1985) The complex of PAMAM-OH dendrimer with Angiotensin...
(1–7) prevented the disuse-induced skeletal muscle atrophy in mice. Int J Nanomed 2017:12
26. Van Der Spoel D, Lindahl E, Hess B, Groenhof G, Mark AE, Berendsen HJ (2005) GROMACS: fast, flexible, and free. J Comput Chem 26:1701–1718
27. Horta BA, Merz PT, Fuchs PF, Dolenc J, Riniker S, Hunenberger PH (2016) A GROMOS-compatible force field for small organic molecules in the condensed phase: the 2016H66 parameter set. J Chem Theory Comput 12:3825–3850
28. Lear S, Cobb SL (2016) Pep-Calc: com a set of web utilities for the calculation of peptide and peptoid properties and automatic mass spectral peak assignment. J Comput-Aid Mol Des 30:271–277
29. Rostkowski M, Olsson MH, Söndergaard CR, Jensen JE (2011) Graphical analysis of pH-dependent properties of proteins predicted using PROPKA. BMC Struct Biol 11:1–6
30. Ramos MC, Quoika PK, Horta VA, Dias DM, Costa EG, do Amaral JL, Ribeiro LM, Liedl KR, Horta BA (2021) pyPolyBuilder: Automated Preparation of Molecular Topologies and Initial Configurations for Molecular Dynamics Simulations of Arbitrary Supramolecules. J Chem Inform Model 61:1539–1544
31. Ramos MC, Horta VA, Horta BA (2019) Molecular dynamics simulations of PAMAM and PPI dendrimers using the GROMOS-Compatible 2016H66 Force Field. J Chem Inform Model 59:1444–1457
32. Cakara D, Kleimann J, Borkoev M (2003) Microscopic prototation equilibria of poly (amidoamine) dendrimers from macroscopic titrations. Macromolecules 36:4201–4207
33. Trot O, Olson AJ (2010) AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. J Comput Chem 31:455–461
34. Chi LA (2022) Double-docking-approach. https://github.com/AmericaChi/Double-docking-approach
35. Barducci A, Bonomi M, Parrinello M (2011) Metadynamics. Wiley Interdiscipl Rev 1:826–843
36. Laio A, Gervasio FL (2008) Metadynamics: a method to simulate rare events and reconstruct the free energy in biophysics, chemistry and material science. Rep Progr Phys 71:126601
37. Barducci A, Bussi G, Parrinello M (2008) Well-tempered metadynamics: a smoothly converging and tunable free-energy method. Phys Rev Lett 100:020603
38. Pavan GM, Barducci A, Albertazzi L, Parrinello M (2013) Combining metadynamics simulation and experiments to characterize dendrimers in solution. Soft Matter 9:2593–2597
39. Barducci A, Bonomi M, Prakash MK, Parrinello M (2013) Free-energy landscape of protein oligomerization from atomistic simulations. Proc Natl Acad Sci USA 110:E4708–E4713
40. Hess B, Bekker H, Berendsen HJ, Fraaije JG (1997) LINCS: a linear constraint solver for molecular simulations. J Comput Chem 18:1463–1472
41. Darden T, York D, Pedersen L (1993) Particle mesh Ewald: an Nlog(N) method for Ewald sums in large systems. J Chem Phys 98:10089–10092
42. Bussi G, Donadio D, Parrinello M (2007) Canonical sampling through velocity rescaling. J Chem Phys 126:014101
43. Parrinello M, Rahman A (1981) Polymorphic transitions in single crystals: a new molecular dynamics method. J Appl Phys 52:7182–7190
44. Daura X, Gademann K, Jaun B, Seebach D, Van Gunsteren WF, Mark AE (1999) Peptide folding: when simulation meets experiment. Angew Chem Int Ed 38:236–240
45. Tiwary P, Parrinello M (2015) A time-independent free energy estimator for metadynamics. J Phys Chem B 119:736–742
46. Maingi V, Jain V, Bharatam PV, Maiti PK (2012) Dendrimer building toolkit: Model building and characterization of various dendrimer architectures. J Comput Chem 33:1997–2011
47. Caballero J, Poblete H, Navarro C, Alzate-Morales JH (2013) Association of nicotinic acid with a poly (amidoamine) dendrimer studied by molecular dynamics simulations. J Mol Graph Model 39:71–78
48. Porcar L, Liu Y, Verduzco R, Hong K, Butler PD, Magid LJ, Smith GS, Chen W-R (2002) Structural investigation of PAMAM dendrimers in aqueous solutions using small-angle neutron scattering: effect of generation. J Phys Chem B 112:14772–14778
49. Rathgeber S, Monkenbusch M, Kreitischmann M, Urban V, Bruet A (2002) Dynamics of star-burst dendrimers in solution in relation to their structural properties. J Chem Phys 117:4047–4062
50. Badalkhani-Khamseh F, Ehrami-Habibi A, Hadipour NL (2017) Atomistic computer simulations on multi-loaded PAMAM dendrimers: a comparison of amine-and hydroxyl-terminated dendrimers. J Comput-Aid Mol Des 31:1097–1111
51. Garcia Y, Jiménez VA, Alderete JB (2020) Host-guest complexation of curcumin and coumarin 6 with PAMAM-OH: Insight from fluorescence spectroscopy and molecular dynamics simulations. J Lumines 222:117182
52. Tanis I, Karatasos K (2009) Association of a weakly acidic anti-inflammatory drug (ibuprofen) with a poly (amidoamine) dendrimer as studied by molecular dynamics simulations. J Phys Chem B 113:10984–10993
53. Ballauff M, Likos CN (2004) Dendrimers in solution: insight from theory and simulation. Angew Chem Int Ed 43:2998–3020
54. Lula I, Denadai ÂL, Resende JM, de Sousa FB, de Lima GF, Pilo-Veloso D, Heine T, Duarte HA, Santos RA, Sinisterra RD (2007) Study of angiotensin-(1–7) vasoactive peptide and its ß cyclodextrin inclusion complexes: complete sequence-specific NMR assignments and structural studies. Peptides 28:2199–2210
55. Clayton J, Baweja L, Wereszczynski J (2022) Peptide Dynamics and Metadynamics: Leveraging Enhanced Sampling Molecular Dynamics to Robustly Model Long-Timescale Transitions. Methods in Molecular Biology (Clifton, NJ) 2405:151–167
56. Chappell MC, Pirro NT, Sykes A, Ferrario CM (1998) Metabolism of angiotensin-(1–7) by angiotensin-converting enzyme. Hypertension 31:362–367
57. Yamada K, Iyer SN, Chappell MC, Ganten D, Ferrario CM (1998) Metabolism of angiotensin-(1–7). Hypertension 32:496–502
58. Luma I, Denadai ÂL, Resende JM, de Sousa FB, de Lima GF, Pilo-Veloso D, Heine T, Duarte HA, Santos RA, Sinisterra RD (2007) Study of angiotensin-(1–7) vasoactive peptide and its ß-cyclodextrin inclusion complexes: complete sequence-specific NMR assignments and structural studies. Peptides 28:2199–2210
59. Clayton J, Baweja L, Wereszczynski J (2022) Peptide Dynamics and Metadynamics: Leveraging Enhanced Sampling Molecular Dynamics to Robustly Model Long-Timescale Transitions. Methods in Molecular Biology (Clifton, NJ) 2405:151–167
60. Chappell MC, Pirro NT, Sykes A, Ferrario CM (1998) Metabolism of angiotensin-(1–7) by angiotensin-converting enzyme. Hypertension 31:362–367
61. Yamada K, Iyer SN, Chappell MC, Ganten D, Ferrario CM (1998) Metabolism of angiotensin-(1–7). Hypertension 32:496–502
62. Springer Nature or its licensor holds exclusive rights to this article under a publisher agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publisher agreement and applicable law.