Research Article

Correlation of Aquaporins and Transmembrane Solute Transporters Revealed by Genome-Wide Analysis in Developing Maize Leaf

Xun Yue,1,2 XiangYu Zhao,2 YuKui Fei,1 and Xiansheng Zhang2

1 College of Information Sciences and Engineering, Shandong Agricultural University, Shandong, Taian 271018, China
2 State Key Laboratory of Crop Biology, College of Life Sciences, Shandong Agricultural University, Shandong, Taian 271018, China

Correspondence should be addressed to Xiansheng Zhang, zhangxs@sdau.edu.cn

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Aquaporins are multifunctional membrane channels that facilitate the transmembrane transport of water and solutes. When transmembrane mineral nutrient transporters exhibit the same expression patterns as aquaporins under diverse temporal and physiological conditions, there is a greater probability that they interact. In this study, genome-wide temporal profiling of transcripts analysis and coexpression network-based approaches are used to examine the significant specificity correlation of aquaporins and transmembrane solute transporters in developing maize leaf. The results indicate that specific maize aquaporins are related to specific transmembrane solute transporters. The analysis demonstrates a systems-level correlation between aquaporins, nutrient transporters, and the homeostasis of mineral nutrients in developing maize leaf. Our results provide a resource for further studies into the physiological function of these aquaporins.

1. Introduction

Water can take different paths on its way through the leaf, in addition to radial water flux, water movement across leaf cell membranes is important for water homeostasis, increasing cell volume, maintaining turgor during expansion, regulating the opening and closure of stomata, and controlling leaf movement [1]. Water movement through cell membranes is facilitated by water channels called aquaporins. Plant aquaporins exhibit multiplicity and diversity, and fall into seven subfamilies loosely based on intracellular locations and sequence similarities: the plasma membrane intrinsic proteins (PIPs), tonoplast intrinsic proteins (TIPs), NOD26-like intrinsic proteins (NIPs), small, basic intrinsic proteins (SIPs), the GlpF-like intrinsic proteins (GIPs), hybrid intrinsic proteins (HIP), and the uncategorized X intrinsic proteins (XIP) [2].

Plant aquaporins are significant not only in plant-water relations, but also in physiological aspects such as nutrient transport and metal/metalloid toxicity [3, 4]. Flexas et al. provided evidence for the in vivo involvement of NtAQPI in mesophyll CO2 conductance, suggesting a significant role for PIPs in CO2 diffusivity [5]. Ludewig and Dynowski have shown that AtTIP1;1 and AtTIP1;2 conduct H2O2 when heterologously expressed in yeast [6]. Azad et al. described TgTIP1;1- and TgTIP1;2-mediated H2O2 conductance by fluorescence assay in Tulipa gesneriana. Recent studies have investigated the selectivity mechanisms of aquaporins, nutrient transporters and homeostasis of mineral nutrients in most plant groups [7]. Hove and Bhave performed a comprehensive analysis of all plant aquaporins, which have been proven to transport ammonia, boron, carbon dioxide, hydrogen peroxide, silicon, and urea [8]. However, their studies focused on aquaporin structures at atomic resolution; there exists little information about the development and growth-dependent expression of aquaporins in leaves and how this relates to nutrient transporters and the homeostasis of mineral nutrients [9].

The use of genomics has resulted in many insights into the fully differentiated state of the maize (Zea mays) leaf [10–12]. Breeze et al. used microarray analysis to obtain a high-resolution time-course profile of gene expression during the development of a single leaf over a 3-week period to senescence [13]. Recently, RNA-Seq has emerged as
2 Comparative and Functional Genomics

Genes under the control of transmembrane transport regulatory mechanism

Coexpression network of aquaporins and transmembrane solutes transports along the leaf developmental gradient

Using genome-wide analysis and coexpression network-based approaches

Detailed temporal and spatial gene expression profiles

High similarity coregulated gene expression patterns of aquaporins and transmembrane solutes transports along the leaf developmental gradient

Figure 1: Schematic of computational pipeline for genome-wide analysis and coexpression network-based approaches.

a powerful new technology for transcriptome analysis [14]. RNA-Seq is particularly advantageous for low-abundance transcripts, for which microarrays are generally insensitive [15]. Matthieu et al. presented a detailed and quantitative analysis of expression in leaf zones, via functional characterization and determination of tissue-specific expression, for a selected group of candidate barley (Hordeum vulgare) aquaporins. These analyses enabled allocation of particular roles during leaf development to specific aquaporin isoforms and proposal of factors that influence their expression [16]. By Illumina sequencing of the maize leaf transcriptome [17], more than 120 million reads were mapped to define gene structure and alternative splicing events, and to quantify transcript abundance, along a leaf developmental gradient. Differential mRNA processing events for most maize genes were detected.

In this study, we use genome-wide analysis and coexpression network-based approaches to examine the wide range of selectivity profiles of aquaporins and associated transmembrane solute transporters in developing maize leaf. We identify specific maize aquaporins related to specific transmembrane solute transporters. Our results provide a resource for further studies into the physiological function of these aquaporins.

2. Materials and Methods

2.1. Materials and Data. Aimed at examining the transcriptional network associated with the development of maize leaf, Pinghua et al. defined gene structure, alternative splicing events, and quantified transcript abundance along a leaf developmental gradient [17]. We used their methods as a template for gene expression analysis. Tissue from leaf three at nine days after planting, three hours into the L period, was collected from four segments: basal (1 cm above the leaf three ligule), transitional (1 cm below the leaf two ligule), maturing (4 cm above the leaf two ligule), and mature (1 cm below the leaf three tip). All data were sequenced and analyzed using an Illumina Genome Analyser 2. RNA-Seq data have been deposited in the NCBI short read archive under accession number SRA012297.

2.2. Method and Computational Pipeline for Genome-Wide Analysis. In this study, our research concentrates on mining the systems-level correlation of aquaporins and associated transmembrane nutrient solute transporters along a leaf developmental gradient. We know that aquaporins and transmembrane mineral nutrient transporters are all under the control of transmembrane transport regulatory mechanisms (GO:0055085). Thus, when transmembrane mineral transporters exhibit the same expression patterns as aquaporins under four segments: basal, transitional, maturing, and mature, there is a greater probability that they interact [18–22]. Using genome-wide analysis and coexpression network-based approaches, systems-level correlation of maize aquaporins, nutrient transporters, and homeostasis of mineral nutrients can be examined. The computational pipeline for genome-wide analysis is shown in Figure 1.

First, we identified characteristic maize aquaporin family and transmembrane transporters under the control of transmembrane transport regulatory mechanisms (GO:0055085).
from the MaizeGDB and Affymetrix GeneChip Maize Genome Array (AFF-900614). Next, detailed temporal and spatial gene expression profiles for aquaporins and nutrient transporters along the developing maize leaf were obtained though RNA-Seq data. Then, high similarity coregulated gene expression patterns were studied and mined by coexpression network-based approaches. Finally, the AmiGO Gene Ontology Database and iHOP (Information Hyperlinked over Proteins) were examined to characterize correlation of ZmTIPs, ZmPIP, and ZmNIPs with associated transmembrane solute transport.

2.3. Rank-Based Network Construction Method. A range of statistical and computational methodologies have been used to extract novel meaning from large biological datasets. Common approaches include both differential expression across conditions and the calculation of correlations between gene expression levels across a large number of samples [18, 23]. The coexpression network-based approach, termed coexpression analysis, considers all samples together and establishes connections between genes based on all available information. Based on the methods from Ruan et al. [24], our computational methodology uses a simple but robust rank-based network construction method. In this experiment, the leaf transcriptome mRNA was isolated from each of the four developmental zones, basal (1 cm above the leaf three ligule), transitional, (1 cm below the leaf two ligule), maturing (4 cm above the leaf two ligule), and mature (1 cm below the leaf three tip). Each gene is associated with a set of expression values, called gene expression profile. We first calculate the Pearson’s correlation coefficient between every pair of genes. We then define a correlation-based similarity score to measure the average coexpression between a gene and other members. For each gene, we rank all other genes by their similarity. We calculate the scores for all the genes. The gene rank of each gene is the position of the score after sorting the list of scores in descending order. We then select these genes to the genes that are most similar to it. For two nodes to be considered as coexpressed, their expression profiles need to satisfy the following (1) Pearson’s correlation coefficient is higher than min_cc (2) One gene is within the max_rank most correlated gene of the other. Cytoscape is used for network visualization and analysis [25]. After we generated the coexpression network, the statistics of the networks (such as, average path lengths, diameters, clustering coefficients, node degree distributions) should be analyzed. But in this study, rank-based network construction method are used mainly to demonstrate a systems-level correlation between aquaporins, nutrient transporters, and the homeostasis of mineral nutrients in developing maize leaf. So, the topological properties of coexpression network will be provided for further studies.

On the other hand, the method has two parameters to be defined in order to run the algorithm: (1) min_cc: threshold on Pearson’s correlation coefficient, (2) max_rank: threshold on the ranks of the correlation coefficient values. In particular, genes in one functional pathway may be strongly mutually coexpressed, while genes in another functional pathway may be only weakly coexpressed. After some preliminary tests with the algorithm, we choose a stringent threshold on Pearson’s correlation coefficient (min_cc = 0.95), it means we attempt to connect the strongly mutually coexpressed genes in one functional pathway, we choose a larger threshold on the ranks of the correlation coefficient values (max_rank = 50), it means we attempt to choose the max subset of the most correlated genes. This rank-based network construction method has successfully elucidated gene function in Arabidopsis thaliana from genome-wide coexpression networks, leading to the identification of genes essential in the life cycle of Arabidopsis [21] and regulation of seed germination [20].

3. Results

3.1. Identification of Candidate Aquaporins and Transmembrane Solute Transporters in Maize. Genome projects have identified 33 aquaporins in maize: 13 PIPs split into 6 PIP1s and 7 PIP2s, 11 TIPs, 6 NIPs, and 3 SIPs. No GIPs, HIPs, or XIPs have been found in maize (see supplemental file 2 in Supplementary Material available online at doi:10.1155/2012/546930). Existing maize aquaporin families were retrieved from the MaizeGDB website. Phylogenetic analysis of maize aquaporins, together with sequences from Arabidopsis and rice, has been performed by Katsuhara et al. [26]. In addition to the aquaporin families, 812 maize transmembrane transport genes (GO:0055085) were identified from the MaizeGDB and Affymetrix GeneChip Maize Genome Array (see supplemental file 1).

In the RNA-Seq dataset, which quantified transcript abundance along a leaf developmental gradient, 24 mRNA maize isoforms were detected, representing three maize aquaporin subfamilies: 11 plasma membrane (PIP), 8 tonoplast (TIP), and 5 NOD26-like (NIP). The others, ZmPIP1;2, ZmPIP2;7, ZmNIP5;1, ZmTIP1;1, ZmTIP4;1, ZmTIP4;3, ZmSIP1;1, ZmSIP1;2, and ZmSIP2;1, showed hardly any or no expression. Surprisingly, ZmPIP1;3 and ZmPIP1;4 were not discernible. Of the maize transmembrane transporters (GO:0055085), only 481 were examined for channel activity by heterologous expression along the leaf developmental gradient (see supplemental file 3).

3.2. Expression Patterns of Maize Aquaporins along a Leaf Developmental Gradient. As shown in Table 1, the expression profiles of aquaporins along a leaf developmental gradient were mostly statistically significant, and expression differed by orders of magnitude.

After examining the dynamic expression patterns of aquaporins in four maize leaf developmental zones, we were interested in studying the specifically expressed genes in each leaf developmental zone. A gene that was expressed at low levels or not detected in the stage was defined as a specifically expressed gene in this stage. Eight genes, ZmTIP4;3 (GRMZM2G146627), ZmTIP4;1 (GRMZM2G103945), ZmTIP1;1 (AAC09245), ZmSIP1;1 (GRMZM2G113470), ZmSIP2;1 (GRMZM2G060922), ZmSIP2;1 (GRMZM2G175038), ZmPIP2;7 (AAK26763, zma:
Table 1: Dynamic expression patterns of aquaporins in four developmental maize leaf zones.

| Aquaporins family | kgeneID | Accession no. | uid | NCBI-GeneID | Expression patterns of maize aquaporins along a leaf developmental gradient (RPKM*) |
|-------------------|---------|---------------|-----|-------------|----------------------------------------------------------------------------------|
| ZmNIP1;1          | GRMZM2G041980 | AAK26750 | 5 | zma:542741 | Basal  542,451  55.78293  1.32962  0.604141  
| ZmNIP2;1          | GRMZM2G028325 | AAK26751 | 5 | zma:542643 | Transitional  0.738541  3.736517  108.7187  151.1207  
| ZmNIP2;2          | GRMZM2G137108 | AAK26752 | 6 | zma:541884 | Maturing  12.83963  65.19496  110.8307  191.0704  
| ZmNIP2;3          | GRMZM2G081239 | AAK26849 | 9 | zma:542497 | Mature  1.390615  0.622895  0.379714  0.028008  
| ZmNIP3;1          | GRMZM2G176209 | AAK26753 | 1 | zma:541885 |  
| ZmPIP1;1          | GRMZM2G174807 | Q41870 | 2 | zma:542434 |  
| ZmPIP1;3          | GRMZM2G392975 | AAK26754 | 4 | zma:541886 |  
| ZmPIP1;4          | GRMZM2G392975 | AAK26754 | 4 | zma:541886 |  
| ZmPIP1;5          | GRMZM2G081843 | AAK26756 | 4 | zma:542014 |  
| ZmPIP1;6          | GRMZM2G136032 | AAK26757 | 9 | zma:541887 |  
| ZmPIP2;1          | GRMZM2G014914 | AAK26758 | 7 | zma:541888 |  
| ZmPIP2;2          | GRMZM2G092125 | AAK26759 | 2 | zma:542644 |  
| ZmPIP2;3          | GRMZM2G081192 | AAK26760 | 4 | zma:541889 |  
| ZmPIP2;4          | GRMZM2G154628 | AAK26761 | 5 | zma:541890 |  
| ZmPIP2;5          | GRMZM2G178693 | AAD28761 | 2 | zma:541891 |  
| ZmPIP2;6          | GRMZM2G047368 | AAK26762 | 7 | zma:541891 |  
| ZmTIP1;2          | GRMZM2G168439 | AAK26767 | 8 | zma:541893 |  
| ZmTIP2;1          | GRMZM2G027098 | AAK26687 | 4 | zma:541894 |  
| ZmTIP2;2          | GRMZM2G056908 | AAK26769 | 5 | zma:541895 |  
| ZmTIP2;3          | GRMZM2G125023 | AAK26770 | 2 | zma:541896 |  
| ZmTIP3;1          | GRMZM2G305446 | AAK26771 | 5 | zma:541896 |  
| ZmTIP3;2          | GRMZM2G103983 | AAK26772 | 1 | zma:541912 |  
| ZmTIP4;2          | GRMZM2G108273 | AAK26773 | 8 | zma:541898 |  
| ZmTIP4;4          | GRMZM2G093090 | AAK26775 | 3 | zma:542647 |  

Basal (1 cm above the leaf three ligule), transitional (1 cm below the leaf two ligule). Maturing (4 cm above the leaf two ligule), mature (1 cm below the leaf three tip).

* Those genes showed hardly any or no expression in leaf tissue.

542645), ZmNIP5-1 (GRMZM2G000471), showed hardly any or no expression in four developmental maize leaf zones. ZmTIP4;4, ZmTIP3;2, ZmNIP1;1, and ZmNIP3;1 were specifically expressed in the mature leaf zone, while ZmNIP2;1 and ZmTIP4;2 were specifically expressed in the leaf basal.

Based on the gene expression profiles of the 24 maize aquaporins along a leaf developmental gradient, a coexpression network-based approach was used to construct coexpression networks. Cytoscape was used to create Figure 2. This approach partitioned the network into two modules, indicating a strong modular structure. We then examined the temporal gene expression profiles of the two modules. As shown in Figure 2, the modules identified by our method contained significant transcripts that showed the most dramatic changes. One modules includes ZmPIP2;1 (GRMZM2G041980), ZmNIP3;1 (GRMZM2G176209), ZmTIP1;6 (GRMZM2G136032), ZmPIP2;4 (GRMZM2G154628), ZmIP1;3 (GRMZM2G081239), ZmTIP3;2 (GRMZM2G103983), ZmPIP2;3 (GRMZM2G081192), ZmPIP1;1 (GRMZM2G174807), ZmPIP2;2 (GRMZM2G092125), ZmNIP1;1 (GRMZM2G041980), ZmTIP2;3 (GRMZM2G125023), ZmTIP1;2 (GRMZM2G168439), ZmTIP3;1 (GRMZM2G305446), ZmTIP4;4 (GRMZM2G093090), 14 genes which showed the highest expression in the basal and transitional zone showed much lower or hardly detectable expression in the maturing and mature zone. While the opposite applied to another modules which are ZmPIP1;5 (GRMZM2G081843), ZmPIP2;5 (GRMZM2G178693), ZmNIP2;3 (GRMZM2G081239), ZmTIP2;1 (GRMZM2G108273), ZmTIP4;2 (GRMZM2G108273), ZmTIP4;2 (GRMZM2G093090), ZmNIP1;3 (GRMZM2G137108), those genes which showed the highest expression in maturing and mature leaf tissue. These results do not seem surprising considering that there are two groups of aquaporins, one positively and the other negatively regulated during different stages of leaf development.

By comparing the expression profiles of ZmNIPs (ZmNIP1;1, ZmNIP2;1, ZmNIP2;2, ZmNIP3;1, ZmNIP3;2, ZmNIP2;3, ZmNIP3;1)
in four maize leaf developmental zones (basal, transitional, maturing, mature), it was evident that the ZmNIP family members are expressed in distinct patterns during leaf development (Table 1). We noticed that ZmNIP2;1, ZmNIP2;2, and ZmNIP2;3 were expressed at low levels at the base of the leaf and gradually increased to reach their highest levels at the leaf tip. In contrast, ZmNIP1;1 and ZmNIP3;1 were expressed at their highest levels at the base of the leaf and decreased significantly to low levels at the leaf tip. The most notable gene, which was upregulated during the whole course of the experiment, was ZmNIP2;2.

Interestingly, with the exception of ZmPIP2;7, which was not expressed, ZmPIPs1 and ZmPIPs2 showed the highest expression in the four leaf base zones. However, these genes showed significant changes in expression levels in the four stages. ZmPIP1;1, ZmPIP1;3, ZmPIP1;4, ZmPIP2;1, ZmPIP2;2, ZmPIP2;3, and ZmPIP2;4 showed their highest expression in the base, with much lower expression in the mature zone. The opposite applied to ZmPIP1;5, ZmPIP1;6, ZmPIP2;5, and ZmPIP2;6, which showed their highest expression in the mature region. Expression in the transitional and maturing zones was at an intermediate level. ZmPIP1;5 showed the most dramatic change in relative expression (a more than 30-fold increase).

Compared with the expression profiles of the ZmNIPs and ZmPIPs, with the exception of ZmTIP1;1, which was not expressed, the ZmTIPS showed distinct patterns during leaf development. We noticed that ZmTIP3;1 showed very low expression or expression near the limit of detection along the leaf developmental gradient. ZmTIP2;3 and ZmTIP4;4 were expressed at low levels in the mature leaf zone, and ZmTIP4;2 showed expression near the limit of detection at the leaf base. ZmTIP1;2, ZmTIP2;1, ZmTIP2;2, and ZmTIP3;2 showed the most dramatic changes in relative expression (more than 10-fold increases).

3.3. ZmTIPS and Associated Transmembrane Solute Transport. While TIPs are predominantly located in the tonoplast, some ZmTIPS are found in specialized organelles such as protein storage vacuoles, lytic vacuoles and small vacuoles, [27]. As shown in Figure 3, we observed significant relationships between ZmTIP genes and neutral solute transporters that
exhibited the same expression patterns in the developing leaf. These included nitrate, hydrogen peroxide (H$_2$O$_2$), sulfate, auxin, sucrose, mercury, drug, sugar, chloride, peptide and oligopeptide transmembrane transporters, and metal ion transporters such as iron, potassium, copper, and cobalt.

**Nitrate.** Chopin et al. showed that the ATNRT2.7 nitrate transporter protein is localized to the vacuolar membrane and plays a specific role in nitrate accumulation in the seed [28]. Our results showed that NRT2.5 (nitrate transporter 2.5, GRMZM2G45124) exhibited the same expression patterns as ZmTIP2;3, ZmTIP3;2, and ZmTIP4;4.

**Hydrogen Peroxide (H$_2$O$_2$).** Evidence for the H$_2$O$_2$ permeability of aquaporins comes from the work of [29] and [7]. Dynowski et al. have shown that the plant TIPs AtTIP1;1 and AtTIP1;2 conduct H$_2$O$_2$ when heterologously expressed in yeast [29]. Azad et al. described TgTIP1;1- and TgTIP1;2-mediated H$_2$O$_2$ conductance by fluorescence assay in *Tulipa gesneriana* [7]. Our results showed that the C2C2(Zn)-GATA transcription factor family (AC202864.3, EntrezGene:100279625) exhibited the same expression patterns as ZmTIP2;1, and the bZIP transcription factor family (GRMZM2G445575, EntrezGene:100192007) exhibited the same expression patterns as ZmTIP2;1. These may be involved in sequence-specific DNA-binding transcription factor activity (GO:0003700) that catalyzes the reaction: methanol + O$_2$ = formaldehyde + H$_2$O$_2$ in developing leaves.

**Sulfate.** Transmembrane sulfate transport is performed by a family of high-affinity sulfate transporters that includes SULTR1;1, SULTR1;3, SULTR1;2, SULTR3;1, SULTR3;2, SULTR3;4, SULTR3;5, SULTR4;1, and SULTR4;2. Kataoka et al. described how SULTR3;5 facilitates the root-to-shoot transport of sulfate through the vasculature [30]. Yoshimoto et al. provided evidence that the Sultr1;3 transporter plays an important role in the loading of sulfate into the sieve tube, initiating the source-to-sink translocation of sulfur nutrients in Arabidopsis [31]. Our results showed that SULTR2;1 (sulfate transporter2;1, GRMZM2G042171) exhibited the same expression patterns as ZmTIP2;2, and SULTR3;4 (sulfate transporter 3;4, GRMZM2G44801) exhibited the same expression patterns as ZmTIP3;2. These genes may be involved in transmembrane sulfate transport in the developing leaf.

**Auxin.** ABCB19 of *A. thaliana* (ATPase multidrug resistance protein) belongs to the multidrug resistance-like (MDR) or B group of the ATP-binding cassette (ABC) transporter superfamily, and mediates polar auxin transport in stems and roots. Lewis et al. suggested that cotyledon expansion during the establishment of photoautotrophic growth depends on ABCB19-mediated auxin import [32]. Our results showed that ABCB19 (GRMZM2G085236) exhibited the same expression patterns as ZmTIP2;3 and ZmTIP3;2, and ABCB1 (GRMZM2G315375, EntrezGene:1003840) exhibited the same expression patterns as ZmTIP2;3. These ABC transporters may be involved in multidrug transport and resistance systems in the developing leaf. We also observed a significant relationship between ZmTIP2;1 and an auxin efflux carrier family protein (GRMZM2G112598).

**Sucrose.** SUT4 (sucrose transporter 4, GRMZM2G307561, EntrezGene:100240688) exhibited the same expression patterns as ZmTIP2;1 and may be involved in sucrose transmembrane transporter activity (GO:0008515) in developing leaves.

**Mercury.** Vacuolar membranes from different species have been commonly characterized by a high, mercury-sensitive osmotic water permeability [33]. TIPs, which function as mercury-sensitive channels and can account for up to 40% of total intrinsic TP protein content [34], supposedly play an important role in this vacuolar function. Our results showed that a heavy-metal-associated domain-containing protein (GRMZM2G096008) exhibited the same expression patterns as ZmTIP2;1 and may be involved in mercury ion transmembrane transporter activity (GO:0015097) in the developing leaf.

We also observed significant relationships between ZmTIP2;1 and a MATE efflux family protein (GRMZM2G170128), and ZmTIP2;3 and another MATE efflux family protein (GRMZM2G423884). These may be involved in transmembrane drug transport (GO:0015238). ATPLT5 (polypeptide transporter 5, GRMZM2G481021) exhibited the same expression patterns as ZmTIP3;2 and may be involved in sugar and substrate-specific transmembrane transporter activity (GO:0022891). GPT2 glucose-6-phosphate transmembrane transporter (GRMZM2G099223, EntrezGene:100281048) exhibited the same expression patterns as ZmTIP2;1 and ZmTIP2;2. Lipoprotein (AC234165.1, EntrezGene:100280698) exhibited the same expression patterns as ZmTIP2;2. Uncharacterized GPI-anchored protein (GRMZM2G041645) exhibited the same expression patterns as ZmTIP2;3 and may be involved in voltage-gated chloride channel activity (GO:0005247) in developing leaves. APP (poly ad-ribose polymerase, GRMZM2G099231) exhibited the same expression patterns as ZmTIP2;3 and may be involved in nucleic acid binding (GO:0003676) that catalyzes the reaction: xanthine + NAD$^+$ + H$_2$O = urate + NADH + H$^+$ in the developing leaf. Peptide transporter PTR2-B, a proton-dependent oligopeptide transport (POT) family protein (GRMZM2G316889, EntrezGene:100381733) exhibited the same expression patterns as ZmTIP2;3 and may be involved in oligopeptide transport (GO:0006857) during leaf development.

**Iron.** Photosynthesis, heme biosynthesis, and Fe-S cluster assembly all take place in the chloroplast, and all require iron. Reduction of iron via a membrane-bound Fe(III) chelate reductase is required before it is transported across...
membranes. Jeong and Cohu reported that the Arabidopsis ferric reductase oxidase (FRO) family, FRO7, localizes to the chloroplast. Their results provided molecular evidence that FRO7 plays a role in chloroplast iron acquisition and is required for efficient photosynthesis in young seedlings and survival under iron-limiting conditions [35]. Wu et al. showed that the six AtFROs encode iron chelate reductases that function in iron homeostasis in Arabidopsis. AtFRO2 displayed the highest iron reduction activity among the AtFROs investigated; further demonstrating that AtFRO2 is a major iron reductase gene in Arabidopsis. AtFRO2 and AtFRO3 were mainly expressed in the roots, AtFRO5 and AtFRO6 in shoots and flowers, AtFRO7 in cotyledons and trichomes, and the transcription of AtFRO8 was specific to leaf veins [36]. Our results showed that ferric reductase-like transmembrane component, FRO7 (GRMZM2G068557, EntrezGene:100281526;) exhibited the same expression patterns as ZmFRO7. Other significant relationships we observed are as follows: a flavin-containing monoxygenase family protein/FMO family protein (GRMZM2G423886, EntrezGene:100272315), a sarcosine oxidase family protein (GRMZM2G428628), and an oxidoreductase family protein (GRMZM2G174773) exhibited the same expression patterns as ZmTIP4;4 and ZmTIP2;3 and may be involved in potassium ion transport (GO:0006813). sks17 (SK5 Similar 17, GRMZM2G043301) exhibited the same expression patterns as ZmTIP2;3 and ZmTIP4;4 and may be involved in copper ion binding (GO:0005507). BAG6 (bcl-2-associated athanogene 6, GRMZM2G063162) exhibited the same expression patterns as ZmTIP2;1 and may be involved in cellular cobalt ion homeostasis (GO:0006877) in the developing leaf.

3.4. ZmPIPs and Associated Transmembrane Solute Transport. PIPs are localized to the plasma membrane and facilitate the movement of water into and out of cells. Flexas et al. provided evidence for the in vivo involvement of NtAQP1 in mesophyll CO2 conductance, suggesting a significant role for PIPs in CO2 diffusivity [5]. Fitzpatrick and Reid established that at least 50% of boron uptake could be facilitated by two barley aquaglyceroporins, HvPIP1;3, and HvPIP1;4 [37]. In fact, most PIP2s show substantial water channel activity, whereas PIP1s either facilitate the movement of neutral solutes or increase water channel activity when coexpressed with PIP2s [3]. We observed significant relationships between ZmPIP genes and the transport of transmembrane solutes, including urate, hydrogen peroxide (H2O2), phosphate, sulfate, sugars, peptides, fatty acids, amino acids, and the metal ions potassium, magnesium, sodium, zinc, and mercury. These results are shown in Figure 3.

Urate. Gaspar et al. showed that ZmPIP1-5b has a low aquaporin activity when expressed in Xenopus oocytes. However, a special feature of ZmPIP1-5, when compared with other plant PIPs, is its capacity to transport urea [38]. Interestingly, our results show that glycine-rich RNA-binding protein 2 (GRMZM2G080603, EntrezGene:542725) exhibited the same expression patterns as ZmPIP1-1 and may be involved in catalysis of the reaction: xanthine + NAD+ + H2O = urate + NADH + H+. These showed their highest expression in the four leaf base zones. Our results also showed that ATP-dependent RNA helicase SUV3 (GRMZM2G078275) and helicase domain-containing protein (GRMZM2G373175) exhibited the same expression patterns as ZmPIP2;4. These may also be involved in catalysis of the reaction: xanthine + NAD+ + H2O = urate + NADH + H+ in developing leaves.

Figure 3: Correlation of maize aquaporins and associated nutrient solute transporters in developing maize leaf. The detailed analysis can be found in Sections 3.3, 3.4, and 3.5.
In addition, **LOS4** (low expression of osmotically responsive genes 4, GRMZM2G000823) exhibited the same expression patterns as ZmPIP1;1 and ZmPIP2;1, and RNA recognition motif (RRM)-containing protein (GRMZM2G071589, EntrezGene:100383130) exhibited the same expression patterns as ZmPIP2;2.

**Hydrogen Peroxide** \( (H_2O_2) \), **TSA**: Zea mays contig14775 mRNA sequence (UniGene:Zm.86314) and **TSA**: Zea mays contig01669 mRNA sequence (UniGene:Zm.42122) exhibited the same expression patterns as ZmPIP2;2. These may be involved in catalysis of the reaction: vanillyl alcohol + \( O_2 \) = vanillin + hydrogen peroxide in the developing leaf.

**Phosphate.** Phosphate transporter (GRMZM2G015401) exhibited the same expression patterns as ZmPIP1;1, M A G 1 (MAIGO 1, GRMZM2G109315, EntrezGene:100192764) exhibited the same expression patterns as ZmPIP2;1, and GRMZM2G152827 (EntrezGene:541617) exhibited the same expression patterns as ZmPIP2;1. These genes may be involved in mitochondrial phosphate transport in developing leaves.

**Sulfate.** In contrast to ZmNIPs, our results show that SULTR3;4 (sulfate transporter 3;4, GRMZM2G444801, EntrezGene:100281787) exhibited the same expression patterns as ZmPIP1;1 and ZmPIP2;1 and may be involved in sulfate transmembrane transport in developing leaves.

**Sugar.** STP (sugar transporter) is a transmembrane carbohydrate transporter. Stadler et al. described the first localization of a guard cell-specific Arabidopsis sugar transporter involved in carbon acquisition in these symplastically-isolated cells [39]. The timing of the transient increase in AtSTP1 expression correlated with guard cell-specific accumulation of sucrose. Sherson et al. investigated the in vivo properties and function of the high-affinity monosaccharide/proton symporter AtSTP1 in Arabidopsis, showing that AtSTP1 is the major monosaccharide transporter in Arabidopsis seedlings and suggesting that active transport by AtSTP1 plays a major role at very high concentrations of exogenous sugar [40]. Our results showed that STP1 (sugar transporter 1, GRMZM2G374812) exhibited the same expression patterns as ZmPIP1;5 and may be involved in transmembrane carbohydrate transport. In addition, Wormit et al. showed that monosaccharide transporter1 (TMT1) is involved in vacuolar monosaccharide transport and plays a major role during stress responses [41]. Our results showed that TMT2 (tonoplast monosaccharide transporter2, GRMZM2G083173, EntrezGene:100285573) exhibited the same expression patterns as ZmPIP1;3/ZmPIP1;4 and may be involved in substrate-specific transmembrane transporter activity (GO:00022891) in developing leaves.

**Peptide.** In contrast to ZmNIPs, our results also showed that OPT4 (oligopeptide transporter 4, GRMZM2G112456) exhibited the same expression patterns as ZmPIP2;4 and may also be involved in transmembrane transport of peptides and oligopeptides. Another gene, ArYSL2 is involved in metal-chelate transport. The investigations of Schaaf et al. support an involvement of ArYSL2 in Fe and Zn homeostasis [42]. Our results showed that YSL2 (yellow stripe like 2, GRMZM2G026391, EntrezGene:100273385) exhibited the same expression patterns as ZmPIP1;6 and may be involved in the transport of peptides and oligopeptides during leaf development.

**Fatty Acid.** Signal recognition particle 54 kDa protein 3/SRP54 (SRP-54C, GRMZM2G038953) exhibited the same expression patterns as ZmPIP2;3 and may enable the directed movement of short-chain fatty acids (less than 10 carbons) into, out of, or within cells. We also observed a significant relationship between the ZmPIP1;1 gene and a transmembrane drug transporter (GRMZM2G079127, EntrezGene:100273132), and ZmPIP1;3/ZmPIP1;4 and an auxin efflux carrier family protein (GRMZM2G050089) and a cationic amino acid transporter (GRMZM2G078292).

**Mercury.** Heavy-metal-associated domain-containing proteins (GRMZM2G155525, EntrezGene:100282516) and (GRMZM2G087101) exhibited the same expression patterns as ZmPIP2;1 and may be involved in mercury ion transmembrane transporter activity (GO:0015097) in the developing leaf.

**Potassium.** Our results showed that the flavin-containing monooxygenase family protein/FMO family protein (GRMZM2G423886, EntrezGene:100272315) exhibited the same expression patterns as ZmPIP2;1, and 3-hydroxybutyryl-CoA dehydrogenase (GRMZM2G106250, EntrezGene:100191282) exhibited the same expression patterns as ZmPIP1;3/ZmPIP1;4. These may be involved in potassium ion transport (GO:0006813) in developing leaves.

**Magnesium.** Magnesium transporter CorA-like family protein (MRS2-2, GRMZM2G159295) exhibited the same expression patterns as ZmPIP1;5 and may be involved in metal ion transmembrane transporter activity (GO:0046873) in the developing leaf.

**Sodium Ion.** NHX2 (sodium hydrogen exchanger 2, GRMZM2G311165) exhibited the same expression patterns as ZmPIP1;1, ZmPIP2;1, ZmTIP2;3, ZmTIP3;1 and ZmTIP4;4 and may be involved in sodium ion transmembrane transporter activity (GO:0015385) during leaf development. Important evidence comes from Yokoi et al. [43], who indicated that AtNHX2 has a major function in vacuolar compartmentalization of Na⁺.

**Zinc Ion.** MPPalpha (mitochondrial processing peptidase alpha subunit, GRMZM2G005036, EntrezGene:100280280) exhibited the same expression patterns as ZmPIP2;1 and may be involved in catalytic/metal ion binding/metalloendopeptidase/zinc ion binding in developing leaves.
3.5. ZmNIPs and Associated Transmembrane Solute Transport. NIPs are located in intracellular membranes. ZmNIPs usually display low water permeability, in fact, these aquaporins are considered plant aquaglyceroporins [27]. We observed significant relationships between ZmNIP genes and the transport of solutes, including glycerol, phosphate, chloride (Cl\(^-\)), drugs, auxin, malic acid, peptides, sugars, mercury, and metal ions such as potassium, copper, zinc, cobalt, and manganese, as shown in Figure 3.

Glycerol. A peroxidase 64 (GRMZM2G160327, EntrezGene: 100281197) and a glycerol-3-phosphate transporter (GRMZM2G078757) exhibited the same expression patterns as ZmNIP1;1. Peroxidase 64 is involved in the regulation of heme binding that is dependent on intracellular accumulation of glycerol (GO:0020037). Our analysis indicate that only ZmNIP1;1 exhibited the same expression patterns as the glycerol-3-phosphate transporter, indicating that ZmNIP1;1 may be involved in transmembrane transport of glycerol.

Phosphorus. Phosphate mobilization into the plant is a complex process requiring numerous transporters for absorption and translocation of this major nutrient. In the A. thaliana genome, nine closely related high-affinity phosphate transporters have been identified but their specific roles remain unclear. Shin et al. showed that eight members of the Arabidopsis Phl1 phosphate transporter family are expressed in roots, with Phl1;1 and Phl1;4 displaying the highest transcript levels [44], Misson et al. demonstrated that the Arabidopsis Phl1;4 high-affinity phosphate transporter was mainly expressed in roots growing on inorganic phosphate limiting medium [45], primarily in the epidermis. In addition, Misson et al. suggested a role for Phl1;4 in phosphate absorption and translocation from the growth medium to the different parts of the plant. Our research showed that phosphate transporters PHT4;2 (GRMZM2G102521) and PHT4;6 (GRMZM2G048363, EntrezGene:100282593) exhibited the same expression patterns as ZmNIP1;1. At the same time, PHT5 (GRMZM2G045473, EntrezGene:100194162) exhibited the same expression patterns as ZmNIP2;1. These genes may be involved in transmembrane inorganic phosphate transport in the developing leaf. Our results also showed that DEG15:endopeptidase (GRMZM2G162699, EntrezGene: 100384043) and GH9A1 (hydrolyzing O-glycosyl compounds 9A1, GRMZM2G003379) exhibited the same expression patterns as ZmNIP3;1 and may be involved in alpha-trehalose-phosphate synthase (UDP-forming) activity (GO:0003825) in developing leaves.

Chloride. Anion transporting proteins of the CLC type are involved in anion homeostasis in a variety of organisms. Seven chloride channel (CLC) members have been identified in the Arabidopsis genome and appear to have different roles in diverse cell organelles. Marmagne et al. have shown that AtCLC-e is targeted to the thylakoid membranes in chloroplasts, in agreement with this subcellular localization [46]. The AtCLC-f protein is localized to Golgi membranes and functionally complements the yeast gef1 mutant, which is disrupted in the single yeast CLC gene that encodes a Golgi-associated protein [47]. CLC-b, a close relative of CLC-a, is localized to the tonoplast, and its expression is strongest in young roots, hypocotyl and cotyledons. Fecth-Bartenbach showed that AtCLC-d is weakly expressed in various tissues, including the root. This suggested that the luminal pH in the trans-Golgi network is adjusted by AtCLC-d-mediated transport of a counter anion such as Cl\(^-\) or NO\(_3^-\) [48]. Our results showed that chloride channel (CLC) members CLC-f (GRMZM2G128969) and plastocyanin-like domain-containing protein (GRMZM2G139193, EntrezGene:100284694) exhibited the same expression patterns as ZmNIP1;1, and chloride channel protein CLC-d (GRMZM2G397836) exhibited the same expression patterns as ZmNIP2;2. These may be involved in chloride transmembrane transport in the developing leaf.

Sulfate. In addition to ZmTIPs and ZmPIPps, our results showed that SULTR1;3 sulfate transmembrane transporter (GRMZM2G159632, EntrezGene:541917) and SULTR3;1 (GRMZM2G158013, EntrezGene:100382058) exhibited the same expression patterns as ZmNIP2;2 and may be involved in sulfate transmembrane transport in developing leaves.

Auxin. Lewis et al. have investigated ABCB19 from A. thaliana, which belongs to the B group of the ATP-binding cassette (ABC) transporter superfamily. ABCB19 mediates polar auxin transport in stems and roots, and cotyledon expansion during the establishment of photoautotrophic growth depends on ABCB19-mediated auxin import [32]. Our results showed that ABCB19:ATPase multidrug resistance protein 11 (GRMZM2G072850) exhibited the same expression patterns as ZmNIP1;1 and may be involved in ABC transporter and ATPase activity coupled to transmembrane movement of substances (GO:0042626), and the G-protein coupled receptor protein signaling pathway (GO:0007186) in the developing leaf. Our results also showed that OsSAUR12:auxin-responsive SAUR gene family member (GRMZM2G154332, EntrezGene:100284645) exhibited the same expression patterns as ZmNIP2;2.

Peptide. In Arabidopsis, AtPTR2, a peptide transport gene, has been suggested to have an important physiological role in plant growth and development [49]. After evaluating the function of this transporter, Song et al. suggested that AtPTR2-B may play a general role in plant nutrition [50]. Our results showed that peptide transporter PTR2-B (GRMZM2G378604, EntrezGene:100281589) and plastocyanin-like domain-containing protein (GRMZM2G139193, EntrezGene:100284694) exhibited the same expression patterns as ZmNIP1;1 and may be involved in oligopeptide transport (GO:0006857) in developing leaves. In addition, our results also showed that OPT7 (oligopeptide transporter 7, GRMZM2G479703, EntrezGene:100383021) exhibited the same expression patterns
as ZmNIP2;1. There is evidence that AtOPTs mediate the uptake of tetra- and pentapeptides [51].

Sugar. A wide range of sugars including hexoses, pentoses, tetrose, a sugar acid, and sugar alcohols, but not disaccharides, induced inward currents in oocytes expressing AtPLT5. Reinders et al. showed that AtPLT5 encodes an ion-coupled uptake transporter [52]. Our results showed that AtPLT5 (polyol transporter 5, GRMZM2G153920, Entrez-Gene:100281055) exhibited the same expression patterns as ZmNIP2;1 and may be involved in sugar and substrate-specific transmembrane transporter activity (GO:0022891). From our results, we also observed significant relationships between the ZmNIP1;1 gene and a transmembrane drug transporter (GRMZM2G069098, Entrez-Gene:100286164), and ZmNIP2;1 and other drug transmembrane transporters (GRMZM2G043075, GRMZM2G115105). These may be involved in transmembrane drug transport (GO:0006855). Malic acid transport protein SLAH3 (GRMZM2G061469, Entrez-Gene:100281575) also had a significant relationship with ZmNIP1;1.

Potassium. Potassium (K+) is a major plant nutrient required for growth and development. K+ uptake in the high-affinity range of concentrations and its components have been widely studied. In A. thaliana, the AtHAK5 transporter and the AtAKT1 channel have been shown to be the main transport proteins involved in this process [53]. Growth analysis shows that AtHAK5 plays a role during severe K+ deprivation. Under K+-deficient conditions, in the presence of Cs+, the high-affinity K+ transporter AtHAK5 and the inward-rectifier K+ channel AtAKT1 have been attributed to K+ uptake in Arabidopsis [54]. Our results showed that HAK5 (high potassium transporter 5, GRMZM2G455817) exhibited the same expression patterns as ZmNIP2;1 and may be involved in potassium ion transmembrane transporter activity (GO:0015079) in the developing leaf. Our results also showed that PDS (phytoene desaturase 3, GRMZM2G088601) exhibited the same expression patterns as ZmNIP2;1 and may be involved in potassium ion transmembrane transporter activity (GO:0015079) in the developing leaf. Our results also showed that PDS (phytoene desaturase 3, GRMZM2G088601) exhibited the same expression patterns as ZmNIP2;1 and may be involved in potassium ion transmembrane transporter activity (GO:0015079) in the developing leaf. Our results also showed that PDS (phytoene desaturase 3, GRMZM2G088601) exhibited the same expression patterns as ZmNIP2;1 and may be involved in potassium ion transmembrane transporter activity (GO:0015079). In the present study, the results show the expression profiles of aquaporins along a leaf developmental gradient were mostly statistically significant, and expression differed by orders of magnitude. ZmTIP4;3, ZmTIP4;1, ZmTIP1;1, ZmSIP1;1, ZmSIP1;2, ZmSIP2;1, ZmSIP2;7, ZmNIP5-1 showed hardly any or no expression in four developmental maize leaf zones. ZmTIP4;4, ZmTIP3;2, ZmNIP1;1, and ZmNIP3;1 were specifically expressed in the mature leaf zone, while ZmNIP2;1 and ZmTIP4;2 were specifically expressed in the leaf basal. Based on the gene expression profiles of the 24 maize aquaporins along a leaf developmental gradient, a coexpression network-based approach was used to construct coexpression networks. The coexpression networks are partitioned into two modules, indicating a strong modular structure. one includes ZmPIP2;1, ZmNIP3;1, ZmPIP1;6, ZmPIP2;4, ZmPIP1;3/ZmPIP1;4, ZmTIP3;2, ZmPIP2;3, ZmPIP1;1, ZmPIP2;2, ZmNIP1;1, ZmTIP2;3, ZmTIP1;2, ZmTIP3;1, ZmTIP4;4, which showed the highest expression in the basal and transitional zone showed much lower or hardly detectable expression in the maturing and mature zone. While the opposite applied to another modules which are ZmPIP1;5, ZmPIP2;5, ZmNIP2;3, ZmNIP2;1, ZmTIP4;2, ZmTIP2;1, ZmNIP2;2, ZmNIP2;6, ZmNIP2;2, those genes showed the highest expression in maturing and mature leaf tissue. These results do not seem surprising considering that there are two groups of aquaporins, one positively and the other negatively regulated during different stages of leaf development.

After examining the dynamic expression patterns of aquaporins in four maize leaf developmental zones, we were interested in studying the correlations of aquaporins and nutrient transporters along a leaf developmental gradient. The results described the correlations of ZmTIPs, ZmPIPs, and other drug transporters involved in cadmium transmembrane transport. ROP9 (rhopas- related protein 9, AC209819.3, EntrezGene:542503) exhibited the same expression patterns as ZmNIP1;1 and may be involved in cadmium transmembrane transporter activity (GO:0006877). This GO category also includes zinc ion transporters and divalent metal ion transporters involved in manganese homeostasis. 4. Discussion

In the present study, the results show the expression profiles of aquaporins along a leaf developmental gradient were mostly statistically significant, and expression differed by orders of magnitude. ZmTIP4;3, ZmTIP4;1, ZmTIP1;1, ZmSIP1;1, ZmSIP1;2, ZmSIP2;1, ZmSIP2;7, ZmNIP5-1 showed hardly any or no expression in four developmental maize leaf zones. ZmTIP4;4, ZmTIP3;2, ZmNIP1;1, and ZmNIP3;1 were specifically expressed in the mature leaf zone, while ZmNIP2;1 and ZmTIP4;2 were specifically expressed in the leaf basal. Based on the gene expression profiles of the 24 maize aquaporins along a leaf developmental gradient, a coexpression network-based approach was used to construct coexpression networks. The coexpression networks are partitioned into two modules, indicating a strong modular structure. one includes ZmPIP2;1, ZmNIP3;1, ZmPIP1;6, ZmPIP2;4, ZmPIP1;3/ZmPIP1;4, ZmTIP3;2, ZmPIP2;3, ZmPIP1;1, ZmPIP2;2, ZmNIP1;1, ZmTIP2;3, ZmTIP1;2, ZmTIP3;1, ZmTIP4;4, which showed the highest expression in the basal and transitional zone showed much lower or hardly detectable expression in the maturing and mature zone. While the opposite applied to another modules which are ZmPIP1;5, ZmPIP2;5, ZmNIP2;3, ZmNIP2;1, ZmTIP4;2, ZmTIP2;1, ZmNIP2;2, ZmNIP2;6, ZmNIP2;2, those genes showed the highest expression in maturing and mature leaf tissue. These results do not seem surprising considering that there are two groups of aquaporins, one positively and the other negatively regulated during different stages of leaf development.
4.1. Specific Maize Aquaporins Are Related to Specific Transmembrane Transporter Families in Developing Leaves. It is proposed that during evolution, plant aquaporins diversified in terms of both the specificity of their expression in plant tissues and their water permeation properties, while maintaining their ability to be induced by the activities of their transmembrane solute transporters. Some aquaporins are more related to specific developmental stages and/or organs. In this study, we confirmed that sulfate transmembrane transporters show specific correlation to certain maize aquaporins, as shown in Figure 4. SULTR1;3 shows a significant relationship with ZmNIP2;1, SULTR3;1 with ZmNIP2;2, SULTR2;1 with ZmTIP2;2, and SULTR3;4 with ZmTIP3;2, ZmPIP1;1 and ZmPIP2;1. This suggests that these maize aquaporins are related to a specific sulfate transmembrane transporter family, SULTR, in the developing leaf.

With chloride channel (CLC) members, our results show that not only does CLC-f exhibit the same expression patterns as ZmNIP1;1 in developing leaves, but also chloride channel protein CLC-d, which is believed to be involved in inorganic phosphate transmembrane transport, displays the same expression patterns as ZmNIP2;2.

Our analysis also reveals that phosphate transporters PHT4;2 and PHT4;6 exhibit the same expression patterns as ZmNIP1;1, and PHT5 the same as ZmNIP2;1, suggesting that these genes may be involved in inorganic phosphate transmembrane transport during leaf development.

4.2. Overlapping Patterns of Expression for Some of Transmembrane Solute Transport Family Members. Overlapping patterns of expression for some of the transmembrane solute transport family members were evident in our analysis. Lavin-containing monooxygenase family protein/FMO family protein not only exhibits the same expression patterns as ZmTIP2;3, but also ZmPIP2;1, suggesting it may be involved in potassium ion transport (GO:0006813) in the developing leaf. ABCB19:ATPase multidrug resistance protein 11 shows the same expression patterns as ZmNIP1;1, ZmTIP2;3, and ZmTIP3;2, suggesting that these aquaporins may be involved in ABC-mediated transport and multidrug resistance systems. Peptide transporter PTR2-B proton-dependent oligopeptide transport (POT) family protein exhibits the same expression patterns as ZmTIP2;3 and ZmTIP3;2, suggesting they may be involved in oligopeptide transport (GO:0006857) during leaf development. ATPLT5 (polyol transporter 5) exhibits the same expression patterns as ZmNIP2;1 and ZmTIP3;2, suggesting they may be involved in the transport of sugars.
Accurate prediction in our approach depends on the conditions associated with each gene. To evaluate the predictive capacity, we examined previously published research work. Important evidence about the reliability of our systems level analysis comes from Kataoka et al. [30]. Sulfate transporters include a family of high-affinity sulfate transporters, with members SULTR1;1, SULTR1;3, SULTR1;2, SULTR3;1, SULTR3;2, SULTR3;4, SULTR3;5, SULTR4;1, and SULTR4;2. Kataoka et al. showed that in Arabidopsis, SULTR3;5 was colocalized with the SULTR2;1 low-affinity sulfate transporter in xylem parenchyma and pericycle cells in roots. The root-to-shoot transport of sulfate is restricted in sultr3;5 mutants, and under conditions of high SULTR2;1 expression in the roots after sulfur limitation, coexpression of SULTR3;5 and SULTR2;1 provides maximal sulfate transport activity. This facilitates retrieval of apoplastic sulfate from the xylem parenchyma cells into the vasculature of Arabidopsis roots and may contribute to the root-to-shoot transport of sulfate. In this study, we found that SULTR3;1 (sulfate transporter 3;1, GRMZM2G158013, EntrezGene:100382058) exhibits the same expression patterns as ZmNIP2;2, and SULTR2;1 (sulfate transporter2;1, GRMZM2G042171) the same as ZmTIP2;2, indicating that these genes are coexpressed and involved in transmembrane sulfate transport in the developing leaf. Other evidence comes from recent investigations on how the structural features of plant aquaporins at atomic resolution influence their substrate selectivity. Our results are in accord with data reported by [6] and [8].

5. Conclusion

The mechanisms controlling the selectivity of aquaporins and nutrient transporters in most plants have been widely studied [6, 8]. However, these studies have focused on aquaporin structures at atomic resolution. Advances in postgenomic technologies and their use by the scientific community are generating increasing quantities of high quality genome-wide transcriptomic data sets. Deposition of these data sets into publicly accessible online databases enables researchers to analyze the collated data and uncover novel information. Therefore, there is a great need for additional analytical approaches to maximize the return on the large collective investment made in data generation. Here, we present a genome-wide analysis and coexpression network-based approach as a powerful novel associative tool for the investigation and prediction of gene function using gene expression data from developing maize leaves. The analysis demonstrates a systems-level correlation between aquaporins, nutrient transporters, and the homeostasis of mineral nutrients in the developing maize leaf. This computational methodology represents a useful alternative approach for the extraction of biological knowledge from existing data. Our results also provide a resource for further studies into the physiological function of these aquaporins.

Authors’ Contribution

X. Yue and X. Zhao contributed equally to this work.

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References

[1] W. Majeran, B. Zybaílov, A. J. Ytterberg, J. Dunsmore, Q. Sun, and K. J. van Wijk, “Consequences of C4 differentiation for chloroplast membrane proteomes in maize mesophyll and bundle sheath cells,” Molecular and Cellular Proteomics, vol. 7, no. 9, pp. 1609–1638, 2008.
[2] J. H. Danielson and U. Johanson, “Unexpected complexity of the Aquaporin gene family in the moss Physcomitrella patens,” BMC Plant Biology, vol. 8, article 45, 2008.
[3] C. Maurel, “Plant aquaporins: novel functions and regulation properties,” FEBS Letters, vol. 581, no. 12, pp. 2227–2236, 2007.
[4] J. M. Ward, P. Maser, and J. I. Schroeder, “Plant ion channels: gene families, physiology, and functional genomics analyses,” Annual Review of Physiology, vol. 71, pp. 59–82, 2009.
[5] J. Flexas, M. Ribas-Carbó, D. T. Hanson et al., “Tobacco aquaporin NtAQP1 is involved in mesophyll conductance to CO2 in vivo,” Plant Journal, vol. 48, no. 3, pp. 427–439, 2006.
[6] U. Ludewig and M. Dynowski, “Plant aquaporin selectivity: where transport assays, computer simulations and physiology meet,” Cellular and Molecular Life Sciences, vol. 66, no. 19, pp. 3161–3175, 2009.
[7] A. K. Azad, N. Yoshikawa, T. Ishikawa, Y. Sawa, and H. Shibata, “Substitution of a single amino acid residue in the aromatic/arginine selectivity filter alters the transport profiles of tonoplast aquaporin homologs,” Biochim Biophys Acta, vol. 1818, no. 1, pp. 1–11, 2011.
[8] R. M. Hove and M. Bhave, “Plant aquaporins with non-aqua functions: deciphering the signature sequences,” Plant Molecular Biology, vol. 75, no. 4-5, pp. 413–430, 2011.
[9] N. V. Obrucheva and I. A. Sin’kevich, “Aquaporins and cell growth,” Russian Journal of Plant Physiology, vol. 57, no. 2, pp. 153–165, 2010.
[10] A. Bräutigam, S. Hoffmann-Benning, and A. P. M. Weber, “Comparative proteomics of chloroplast envelopes from C3 and C4 plants reveals specific adaptations of the plastid envelope to C4 photosynthesis and candidate proteins required for maintaining C4 metabolite fluxes,” Plant Physiology, vol. 148, no. 3, pp. 568–579, 2008.
[11] S. Covshoff, W. Majeran, P. Liu, J. M. Kolkman, K. J. Van Wijk, and T. P. Brutnell, “Deregulation of maize C4 photosynthetic development in a mesophyll cell-defective mutant,” Plant Physiology, vol. 146, no. 4, pp. 1469–1481, 2008.
[12] G. Friso, W. Majeran, M. Huang, Q. Sun, and K. J. van Wijk, “Reconstruction of metabolic pathways, protein expression, and homeostasis machineries across maize bundle sheath and mesophyll chloroplasts: large-scale quantitative proteomics using the first maize genome assembly,” Plant Physiology, vol. 152, no. 3, pp. 1219–1250, 2010.
[13] E. Breeze, E. Harrison, S. McHattie et al., “High-resolution temporal profiling of transcripts during Arabidopsis leaf
senescence reveals a distinct chronology of processes and regulation,” \textit{Plant Cell}, vol. 23, no. 3, pp. 873–894, 2011.

[14] S. Liu, L. Lin, P. Jiang, D. Wang, and Y. Xing, “A comparison of RNA-Seq and high-density exon array for detecting differential gene expression between closely related species,” \textit{Nucleic Acids Research}, vol. 39, no. 2, pp. 578–588, 2011.

[15] Z. Wang, M. Gerstein, and M. Snyder, “RNA-Seq: a revolutionary tool for transcriptomics,” \textit{Nature Reviews Genetics}, vol. 10, no. 1, pp. 57–63, 2009.

[16] B. Matthieu, K. Thorsten, J. M. Anthony, V. Jean-Luc, P. Thomas, and F. Wieland, “Developmental pattern of aquaporin expression in barley (Hordeum vulgare L.) leaves,” \textit{Journal of Experimental Botany}, vol. 62, no. 12, pp. 4127–4142, 2011.

[17] L. Pinghua, P. Lalit, and G. Neeru, “The developmental dynamics of the maize leaf transcriptome,” \textit{Nature Genetics}, vol. 42, no. 12, pp. 1060–1067, 2010.

[18] B. Usadel, T. Obayashi, M. Mutwil et al., “Co-expression tools for plant biology: opportunities for hypothesis generation and caveats,” \textit{Plant, Cell and Environment}, vol. 32, no. 12, pp. 1633–1651, 2009.

[19] I. Lee, B. Ambaru, P. Thakkar, E. M. Marcotte, and S. Y. Rhee, “Rational association of genes with traits using a genome-scale gene network for Arabidopsis thaliana,” \textit{Nature Biotechnology}, vol. 28, no. 2, pp. 149–156, 2010.

[20] G. W. Bassel, H. Lan, E. Glaab et al., “Genome-wide network model capturing seed germination reveals coordinated regulation of plant cellular phase transitions,” \textit{Proceedings of the National Academy of Sciences of the United States of America}, vol. 108, no. 23, pp. 9709–9714, 2011.

[21] M. Mutwil, S. Klic, T. Tohge et al., “PlaNet: combined sequence and expression comparisons across plant networks derived from seven species,” \textit{Plant Cell}, vol. 23, no. 3, pp. 895–910, 2011.

[22] W. B. George, G. Enrico, and M. Julietta, “Functional network construction in arabidopsis using rule-based machine learning on large-scale data sets,” \textit{The Plant Cell}, vol. 23, pp. 3101–3116, 2011.

[23] S. M. Brady and N. J. Provart, “Web-queryable large-scale data sets for hypothesis generation in plant biology,” \textit{Plant Cell}, vol. 21, no. 4, pp. 1034–1051, 2009.

[24] J. Ruan, A. K. Dean, and W. Zhang, “A general co-expression network-based approach to gene expression analysis: comparison and applications,” \textit{BMC Systems Biology}, vol. 4, article 8, 2010.

[25] M. E. Smoot, K. Ono, I. Ruscheinski, P. L. Wang, and T. Ideker, “Cytoscape 2.8: new features for data integration and network visualization,” \textit{Bioinformatics}, vol. 27, no. 3, Article ID btq675, pp. 431–432, 2011.

[26] M. Katsuhara, Y. T. Hanba, K. Shiratake, and M. Maeshima, “Expanding roles of plant aquaporins in plasma membranes and cell organelles,” \textit{Functional Plant Biology}, vol. 35, no. 1, pp. 1–14, 2008.

[27] D. Gomes, A. Agasse, P. Thièbaut, S. Delrot, H. Gerós, and F. Chaumont, “Aquaporins are multifunctional water and solute transporters highly divergent in living organisms,” \textit{Biochimica et Biophysica Acta}, vol. 1788, no. 6, pp. 1213–1228, 2009.

[28] F. Chopin, M. Orsel, M. F. Dorbe et al., “The Arabidopsis ATNRT2.7 nitrate transporter controls nitrate content in seeds,” \textit{Plant Cell}, vol. 19, no. 5, pp. 1590–1602, 2007.

[29] M. Dynowski, G. Schaal, D. Loque, O. Moran, and U. Ludewig, “Plant plasma membrane water channels conduct the signalling molecule H$_2$O$_2$,” \textit{Biochemical Journal}, vol. 414, no. 1, pp. 53–61, 2008.

[30] T. Kataoka, N. Hayashi, T. Yamaya, and H. Takahashi, “Root-to-shoot transport of sulfate in Arabidopsis. Evidence for the role of SULTR3;5 as a component of low-affinity sulfate transport system in the root vasculature,” \textit{Plant physiology}, vol. 136, no. 4, pp. 4198–4204, 2004.

[31] N. Yoshimoto, E. Inoue, K. Saito, T. Yamaya, and H. Takahashi, “Phloem-localizing sulfate transporter, Sult1;3, mediates redistribution of sulfur from source to sink organs in arabidopsis,” \textit{Plant Physiology}, vol. 131, no. 4, pp. 1511–1517, 2003.

[32] D. R. Lewis, G. Wu, K. Ljung, and E. P. Spalding, “Auxin transport into cotyledons and cotyledon growth depend similarly on the ABCB19 Multidrug Resistance-like transporter,” \textit{Plant Journal}, vol. 60, no. 1, pp. 91–101, 2009.

[33] C. Maurel, H. Javot, V. Lavergne et al., “Molecular physiology of aquaporins in plants,” \textit{International Review of Cytology}, vol. 215, pp. 105–148, 2002.

[34] T. Higuchi, S. Suga, T. Tsuchiya et al., “Molecular cloning, water channel activity and tissue specific expression of two isoforms of radish vacuolar aquaporin,” \textit{Plant and Cell Physiology}, vol. 39, no. 9, pp. 905–913, 1998.

[35] J. Jeong, C. Cobo, L. Kerkeb, M. Pilon, E. L. Connelly, and M. L. Guerinot, “Chloroplast Fe(III) chelate reductase activity is essential for seedling viability under iron limiting conditions,” \textit{Proceedings of the National Academy of Sciences of the United States of America}, vol. 105, no. 30, pp. 10619–10624, 2008.

[36] H. Wu, L. Li, J. Du, Y. Yuan, X. Cheng, and H. Q. Ling, “Molecular and biochemical characterization of the Fe(III) chelate reductase gene family in Arabidopsis thaliana,” \textit{Plant and Cell Physiology}, vol. 46, no. 9, pp. 1505–1514, 2005.

[37] K. L. Fitzpatrick and R. J. Reid, “The involvement of aquaglyceroporins in transport of boron in barley roots,” \textit{Plant, Cell and Environment}, vol. 32, no. 10, pp. 1357–1365, 2009.

[38] M. Gaspar, A. Bousser, I. Sissoeff, O. Roche, J. Hoarau, and A. Mahé, “Cloning and characterization of ZmPIP1-5b, an aquaporin transporting water and urea,” \textit{Plant Science}, vol. 165, no. 1, pp. 21–31, 2003.

[39] R. Stadler, M. Büttner, P. Ache et al., “Diurnal and light-regulated expression of AtSTP1 in guard cells of Arabidopsis,” \textit{Plant Physiology}, vol. 133, no. 2, pp. 528–537, 2003.

[40] S. M. Sherson, G. Hemmann, G. Wallace et al., “Monosaccharide/proton symporter AtSTP1 plays a major role in uptake and response of Arabidopsis seeds and seedlings to sugars,” \textit{Plant Journal}, vol. 24, no. 6, pp. 849–857, 2000.

[41] A. Wormit, O. Trentmann, I. Feifer et al., “Molecular identification and physiological characterization of a novel monosaccharide transporter from Arabidopsis involved in vacuolar sugar transport,” \textit{Plant Cell}, vol. 18, no. 12, pp. 3476–3490, 2006.

[42] G. Schaal, A. Schikora, J. Häaberle et al., “A putative function for the Arabidopsis Fe-phytosiderophore transporter homolog AtYSL2 in Fe and Zn homeostasis,” \textit{Plant and Cell Physiology}, vol. 46, no. 5, pp. 762–774, 2005.

[43] S. Yokoi, F. J. Quintero, B. Cubero et al., “Differential expression and function of Arabidopsis thaliana NHX Na+/H+ antiporters in the salt stress response,” \textit{Plant Journal}, vol. 30, no. 5, pp. 529–539, 2002.

[44] H. Shin, H. S. Shin, G. R. Dewbre, and M. J. Harrison, “Phosphate transport in Arabidopsis: Pht1;1 and Pht1;4 play a major role in phosphate acquisition from both low- and high-phosphate environments,” \textit{Plant Journal}, vol. 39, no. 4, pp. 629–642, 2004.
[45] J. Misson, M. C. Thibaud, N. Bechtold, K. Raghothama, and L. Nussaume, “Transcriptional regulation and functional properties of Arabidopsis Ph1;4, a high affinity transporter contributing greatly to phosphate uptake in phosphate deprived plants,” *Plant Molecular Biology*, vol. 55, no. 5, pp. 727–741, 2004.

[46] A. Marmagne, M. Vinauger-Douard, D. Monachello et al., “Two members of the Arabidopsis CLC (chloride channel) family, AtCLCe and AtCLCf, are associated with thylakoid and Golgi membranes, respectively,” *Journal of Experimental Botany*, vol. 58, no. 12, pp. 3385–3393, 2007.

[47] J. Von Der Fecht-Bartenbach, M. Bogner, M. Dynowski, and U. Ludewig, “CLC-b-mediated no3-/H+ exchange across the tonoplast of arabidopsis vacuoles,” *Plant and Cell Physiology*, vol. 51, no. 6, pp. 960–968, 2010.

[48] J. V. D. Fecht-Bartenbach, M. Bogner, M. Krebs, Y. D. Stierhof, K. Schumacher, and U. Ludewig, “Function of the anion transporter AtCLC-d in the trans-Golgi network,” *Plant Journal*, vol. 50, no. 3, pp. 466–474, 2007.

[49] C. S. Chiang, G. Stacey, and Y. F. Tsay, “Mechanisms and functional properties of two peptide transporters, AtPTR2 and fPTR2,” *Journal of Biological Chemistry*, vol. 279, no. 29, pp. 30150–30157, 2004.

[50] W. Song, S. Koh, M. Czako et al., “Antisense expression of the peptide transport gene AtPTR2-B delays flowering and arrests seed development in transgenic Arabidopsis plants,” *Plant Physiology*, vol. 114, no. 3, pp. 927–935, 1997.

[51] O. Cagnac, A. Bourbouloux, D. Chakrabarty, M. Y. Zhang, and S. Delrot, “AtOPT6 transports glutathione derivatives and is induced by primisulfuron,” *Plant Physiology*, vol. 135, no. 3, pp. 1378–1387, 2004.

[52] A. Reinders, J. A. Panshyshyn, and J. M. Ward, “Analysis of transport activity of Arabidopsis sugar alcohol permease homolog AtPLT5,” *Journal of Biological Chemistry*, vol. 280, no. 2, pp. 1594–1602, 2005.

[53] M. Nieves-Cordones, F. Alemán, V. Martínez, and F. Rubio, “The Arabidopsis thaliana HAK5 K+ transporter is required for plant growth and K+ acquisition from low K+ solutions under saline conditions,” *Molecular Plant*, vol. 3, no. 2, pp. 326–333, 2010.

[54] F. Rubio, F. Alemán, M. Nieves-Cordones, and V. Martínez, “Studies on Arabidopsis athak5, atakt1 double mutants disclose the range of concentrations at which AtHAK5, AtAKT1 and unknown systems mediate K+ uptake,” *Physiologia Plantarum*, vol. 139, no. 2, pp. 220–228, 2010.

[55] I. Hwang, H. Sze, and J. F. Harper, “A calcium-dependent protein kinase can inhibit a calmodulin-stimulated Ca2+ pump (ACA2) located in the endoplasmic reticulum of Arabidopsis,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 11, pp. 6224–6229, 2000.