Immune responses in mice vaccinated with a DNA vaccine expressing serine protease-like protein from the new-born larval stage of *Trichinella spiralis*

JING XU1†, XUE BAI1†, LI BO WANG1, HAI NING SHI2, JOKE W. B. VAN DER GIESSEN3, PASCAL BOIREAU1,4, MING YUAN LIU1,5*, and XIAO LEI LIU1*

1 Key Laboratory for Zoonoses Research, Ministry of Education, Institute of Zoonoses, Jilin University, Changchun, People’s Republic of China
2 Mucosal Immunology Laboratory, Pediatric Gastroenterology Unit, Massachusetts General Hospital East, Massachusetts, USA
3 Centre for Infectious Disease Control, National Institute for Public Health and the Environment, Amsterdam, The Netherlands
4 Laboratory for Animal Health, ANSES, INRA, ENVA, Université Paris Est, Maisons Alfort, France
5 Jiangsu Co-innovation Center for Prevention and Control of Important Animal Infectious Diseases and Zoonoses, Yangzhou, Jiangsu, People’s Republic of China

(Received 22 August 2016; revised 12 November 2016; accepted 16 November 2016; first published online 10 January 2017)

SUMMARY

*Trichinella spiralis* is a parasitic helminth that can infect almost all mammals, including humans. *Trichinella spiralis* infection elicits a typical type 2 immune responses, while suppresses type 1 immune responses, which is in favour of their parasitism. DNA vaccines have been shown to be capable of eliciting balanced CD4+ and CD8+ T cell responses as well as humoral immune responses in small-animal models, which will be advantage to induce protective immune response against helminth infection. In this study, serine protease (Ts-NBLsp) was encoded by a cDNA fragment of new-born *T. spiralis* larvae, and was inserted after CMV promoter to construct a DNA vaccine [pcDNA3·1 (+)-Ts-NBLsp]. Ts-NBLsp expression was demonstrated by immunofluorescence. Sera samples were obtained from vaccinated mice, and they showed strong anti-Ts-NBLsp-specific IgG response. Mice immunized with the pcDNA3·1 (+)-Ts-NBLsp DNA vaccine showed a 77.93% reduction in muscle larvae (ML) following challenge with *T. spiralis* ML. Our results demonstrate that the vaccination with pcDNA3·1 (+)-Ts-NBLsp plasmid promoted the balance of type 1 and 2 immune responses and produced a significant protection against *T. spiralis* infection in mice.

Key words: *Trichinella spiralis*, DNA vaccine, protective immunity, new-born larvae.

INTRODUCTION

*Trichinella* spp. are intestinal nematode parasites that can cause trichinellosis in humans and animals (Dupouy-Camet, 2000). More than 100 species of mammals, birds and reptiles can be infected by *T. spiralis* (Pozio and Zarlenga, 2013); domestic pigs, horses, dogs and cats are known as the most important hosts of *T. spiralis*. Infection occurs when humans consume raw or undercooked meat of different animal origins containing *T. spiralis* muscle larvae (ML). Trichinellosis is a public health hazard, and it also an economic problem in animal production and food safety (Dorny et al. 2009). It is difficult to control this zoonosis due to its wide distribution of domestic and wild animal reservoirs (Wang and Cui, 2001; Wang et al. 2006; Cui et al. 2011; Murrell and Pozio, 2011). So far, useful and stable anti-*Trichinella* vaccines that can be used in animal husbandry have not been developed yet. Therefore, it is necessary to develop a vaccine to prevent *Trichinella* infection in domestic animals and humans.

*Trichinella spiralis* is a nematode parasite that spends its larval and adult life stages in the same host. It has three major antigenic stages, ML, adult worms (AD) and new-born larvae (NBL). During natural course of *T. spiralis* infection, AD get pregnant and NBL are released in the intestines of the host, then all the AD evacuate from the host through intestines between 10 and 15 days post-infection (dpi). The NBL arrive at striated muscle through lymphatic vessels and blood circulation, and then develop into encapsulated ML in 20 days. Then ML start long-term parasitizing in striated muscle (Pozio, 1989; Gottstein et al. 2009). NBL is a key stage for the growth of *T. spiralis*, without the protection of capsule, larvae will be exposed to muscle immune system. Therefore, it is necessary to think highly of antigens from the NBL stage.

*Parasitology* (2017), 144, 712–719. © Cambridge University Press 2017. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.

doi:10.1017/S0031182016002493

https://doi.org/10.1017/S0031182016002493 Published online by Cambridge University Press
A highly antigenic NBL stage-specific serine protease gene, Ts-NBLsp, was obtained via a subtractive cDNA library of T. spiralis NBL (Liu et al. 2007). Ts-NBLsp showed encouraging potential in the early detection of Trichinella infection, but the function of it in vivo is unclear. Multiple serine proteases have been identified at different stages of T. spiralis, evidences have shown that serine proteinases are abundant in excretory/secretory products or crude extract proteins from T. spiralis. Most of these serine proteases are involved in parasite survival and establishment of infection (Todorova, 2000; Todorova and Stoyanov, 2000; Bien et al. 2012; Wang et al. 2014). Furthermore, serine proteases are involved in reproduction and evasion of the host immune system (Dzik, 2006; Balasubramanian et al. 2010). It has been reported that the antibody response against serine proteases from parasite can inhibit the protease activity and possibly contribute to inhibit invasion of the parasite in a sensitized host (Ros-Moreno et al. 2000; Todorova and Stoyanov, 2000).

Therefore, in this study the plasmid of pcDNA3-1 (+)-Ts-NBLsp was constructed, purified and used as an experimental DNA vaccine to immunize Kunming mice, and to evaluate the immune response and protective effects in a murine model of T. spiralis infection.

MATERIALS AND METHODS

Parasites and animals

Trichinella spiralis (ISS534) used in this study was maintained in Wistar rats in our laboratory by serial passage infections. Muscle larvae were recovered from mice 35 dpi with artificial digestion solution (1% pepsin/HCl) (Li et al. 2010). Female Kunming mice aged 6–8 weeks were obtained from Norman Bethune University of Medical Science (NBUMS), China.

Amplify the cDNA of Ts-NBLsp

The complete sequence of Ts-NBLsp gene has been submitted to Genbank (GenBank AY491941-1). The full-length cDNA of Ts-NBLsp was amplified by PCR from a plasmid of cDNA library of T. spiralis NBL with the following primers: 5'-CCGTGTTGAGATGCGTGTTGCC-3' (forward) and 5'-CGCGTCGAGTTACTTAGAAAACTGATA-3' (reverse). The KpnI and XhoI restriction sites are underlined. The PCR cycles consisted of an initial denaturation step at 94 °C for 5 min, 30 cycles of denaturation at 94 °C for 1 min and annealing at 53 °C for 1 min with an extension at 72 °C for 1 min, and a final extension at this same temperature for another 10 min.

Recombinant protein rTs-NBLsp expression, purification and identified

The DNA fragment of Ts-NBLsp, described above, was cloned into pET28a expression vector using the T7 promoter. The recombinant plasmid pET28a/Ts-NBLsp was determined by DNA sequencing and transformed in Escherichia coli BL21 (DE3) chemically competent cells. The BL21 cells containing pET28a/Ts-NBLsp grown in 1 L LB and were shaken vigorously of 120 rpm at 37 °C up to an optical density of 0.6, calculated at 600 nm. Then, the cells were induced with isopropyl-b-D-thiogalactopyranoside (IPTG) to a final concentration of 1 mM with vigorous shaking of 120 rpm at 37 °C for 5 h. The cells were centrifuged at 7000 g for 15 min and the pellet was resuspended with binding buffer (20 mM Tris–HCl, pH: 7.9, 0.5 M NaCl, 8 M urea, 1 mM PMSF). Purification of the recombinant protein was performed as previously described (Feng et al. 2013). Briefly, the inclusion body was purified by Ni-affinity chromatography and refolded by drop-wise dilution. Then sodium dodecyl sulphate–polyacrylamide gel electrophoresis (SDS–PAGE) and Western blotting were used to identify the protein. Briefly, the protein was separated by 10% SDS–PAGE and electrophoretically transferred onto a nitrocellulose membrane (Bio-Rad, Hercules, CA, USA). The membrane was blocked with 5% skimmed milk in PBS containing 0.05% Tween-20 (PBST) overnight at 4 °C and incubated with mouse anti-Ts-NBLsp mAbs (1:500, this monoclonal antibody to Ts-NBLsp was produced by Laboratory for Animal Health, Maisons Alfort, France) for 1 h at 37 °C. HRP (horseradish peroxidase)-conjugated goat antimouse IgG (Beijing Dingguo Changsheng Biotechnology CO.LTD) was used as the secondary antibody. Finally, the protein bands were developed using ECL (enhanced chemiluminescence) reagents (Applygen Technologies Inc., Beijing, China). The images were photographed in a Chemi Doc image scanner from Bio Rad.

Plasmid DNA vaccine constructs

The DNA fragment of Ts-NBLsp, described above, was cloned into the eukaryotic expression vector pcDNA3-1 (+) using the cytomegalovirus promoter. The sequence accuracy of recombinant plasmid was authenticated by double digestion and DNA sequencing. The recombinant plasmid was then purified from transformed E. coli DH5α cells by Endofree Plasmid Midiprep Kit (BioMIGA, San Diego, USA) following the manufacturer’s instructions, dissolved in sterile endotoxin-free TE buffer and stored at −20 °C. The recombinant plasmid was named pcDNA3-1(+)-Ts-NBLsp.
Immunization and challenge

Six-week-old female Kunming mice (20 per experimental group) were immunized (by bilateral intramuscular (IM) injection into the quadriceps) twice with 2 weeks interval. In pcDNA3·1(+)·Ts-NBLsp-vaccinated group, each mouse was injected with 60 µg plasmid dissolved in 100 µL sterile PBS at each immunization. As negative control, one group of mice were injected with 60 µg empty pcDNA3·1(+) vector, and another group of mice injected with 100 µL sterile PBS constituted the blank control. Two weeks after the last immunization, mice of each group were challenged with 250 T. spiralis ML. Blood was collected from the tail vein before and weekly after vaccination until the challenge infection, and sera samples were separated and stored at −20 °C. Pre-immune serum samples were used as negative controls.

Detection of target gene expression in immunized mice by immunofluorescence test

To determine the expression of recombinant Ts-NBLsp in vivo, quadriceps femoris of three mice in each group were obtained at 48 h post the first immunization. These tissues were fixed in 4% paraformaldehyde (v/v) in PBS, embedded in paraffin, and cut into 3 µm paraffin sections, and then immunofluorescence test (IFT) was performed as described previously. Briefly, the sections were incubated with 0.1% Triton X-100 in PBS at 4 °C for 1 h, blocked for non-specific protein binding by incubation in 10% goat serum diluted in PBS at room temperature for 1 h, and incubated with mouse anti-Ts-NBLsp mAbs (1:200 dilution) at 37 °C for 1 h. After washing, the sections were incubated with a 1:100 dilution of FITC-conjugated goat anti-mouse IgG (Santa Cruz, USA) at 37 °C for 1 h, and then stained with Hoechst (Beyotime Biotechnology, Beijing, China) for 5 min at room temperature. The sections incubated with serum from an unimmunized mouse at the same dilution served as a negative control. The sections were examined and photographed under fluorescent microscope (Olympus, Japan).

Determination of antibodies

Anti-Ts-NBLsp antibodies (total IgG, IgG1 and IgG2a) in serum of immunized mice were checked by enzyme-linked immunosorbent assay (ELISA). Briefly, microtitre plates (Corning, Acton, MA) were coated with 100 µL of the recombinant protein (rTs-NBLsp) (50 µg mL⁻¹ in coating buffer, overnight at 4 °C). The plates were washed with PBST and blocked with 5% non-fat dry milk (Sigma, St. Louis, MO) in PBST at 37 °C for 2 h, followed by incubation at 37 °C for 2 h with 100 µL of the serum samples with a 1:50 dilution in PBST with 5% non-fat dry milk. After three washes, the plates were incubated at 37 °C for 2 h with 100 µL well⁻¹ of horseradish peroxidase-conjugated goat anti-mouse IgG antibody (Beijing Dingguo Changsheng Biotechnology Co. Ltd.) at a 1:4000 dilution, or goat anti-mouse IgG1 or goat anti-mouse IgG2a (Abcam, Cambridge, UK) at a 1:500 dilution, which was used for determination of total IgG antibody levels and isotype analysis, respectively. ELISA was developed by chromogen 3,3′,5,5′-tetramethylbenzidine (TMB, Tiangen Biotech Co., Beijing, China), the reaction was terminated by 2 N H2SO4 solution at 15 min after substrate addition, and the OD 450 (Optical density at 450 nm) value was measured.

Cytokine assays

To analysis the cellular immune responses of mice immunized with DNA construct, the concentrations of interleukin 4 (IL-4), IL-10 and interferon (IFN)-γ in serum sample collected before and weekly after vaccination were measured by ELISA according to the manufacturer’s instructions (eBioscience, San Diego, California). The concentration of cytokines was determined by comparison with the standard curves constructed with known amounts of the respective mouse recombinant cytokines. Results were expressed in picograms per millilitre (pg mL⁻¹).

Flow cytometry analysis of T lymphocytes

One week after the final immunization, blood was collected from five mice in each group and poured
slowly into anticoagulation tubes. Staining of the cells for flow cytometry analysis was performed by direct staining. Briefly, the cells were stained with optimal concentrations of APC-conjugated Hamster anti-mouse CD3e (0·2 mg mL⁻¹, clone 145–2C11, BD Biosciences, San Jose, CA, USA), FITC-conjugated Rat anti-mouse CD4 (0·5 mg mL⁻¹, clone GK 1·5, BD Biosciences, San Jose, CA, USA), and PE-conjugated Rat anti-mouse CD8a (0·2 mg mL⁻¹, clone 53–6·7, BD Biosciences, San Jose, CA, USA), and incubated at room temperature for 30 min. Red blood cells were lysed by the addition of a triple volume of lysis solution (Solarbio, Beijing, China) on ice for 15 min. Cells were then washed twice in PBS, suspended in 300 µL PBS, and immediately analysed with a BD FACSCalibur™ flow cytometer (BD Biosciences, Heidelberg, Germany).

**Evaluation of immune protection**

Twelve mice of each group were sacrificed 42 days after challenge, the carcass weight and the ML of each mouse were examined. The protective immunity was calculated as the worm reduction rate of recovered ML per gram from the vaccinated group compared with the blank control group.

**Statistical analysis**

All statistical analyses were performed by SPSS 16·0 Data Editor. Data were expressed as the mean ± standard deviation (S.D.), and the differences of the data between all the groups were evaluated by one-way ANOVA analysis. The difference between groups were regarded as statistically significant if $P < 0·05$.

**RESULTS**

Identification plasmids of pcDNA3·1(+)-Ts-NBLsp and pET28a/Ts-NBLsp

The constructed plasmids were subjected to digestion with KpnI and XhoI. Electrophoretic separation of the digestion products showed that the construction of the recombinant plasmids was successful (data not shown). DNA sequence analysis indicated that the amplified fragment of Ts-NBLsp gene consisting of 1209 bp was correctly cloned into prokaryotic expression vector pet28a and eukaryotic expression vector pcDNA3·1(+).
Recombinant protein rTs-NBLsp was identified by Western blot

Western blot results demonstrated that the expression of recombinant protein rTs-NBLsp could be induced after addition of IPTG. The mouse anti-Ts-NBLsp monoclonal antibody recognized the recombinant protein, which is around 44-46 kDa in recombinant plasmid pET28a/Ts-NBLsp transformed E. coli cells, no protein band was observed in the pET28a transformed E. coli cells. (Fig. 1).

Ts-NBLsp was identified in vaccinated mice by immunofluorescence

In immunofluorescence analysis, after incubated with mouse anti-Ts-NBLsp mAbs, the sections from mouse vaccinated with plasmid of pcDNA3·1 (+)-Ts-NBLsp at 48 h showed specific green fluorescence. However, the negative control, which were vaccinated with pcDNA3·1(+) or PBS, did not show any fluorescence emission (Fig. 2). These results demonstrate that the Ts-NBLsp protein was expressed in quadriceps femoris of the mice vaccinated with pcDNA3·1(+) -Ts-NBLsp, where it retained its antigenic reactivity.

Humoral immune responses induced by vaccination

As shown in (Fig. 3), specific anti-Ts-NBLsp IgG has been induced in pcDNA3·1(+)-Ts-NBLsp-vaccinated group. The analysis of the IgG isotypes showed that in the pcDNA3·1(+) -Ts-NBLsp-vaccinated group, the IgG2a-predominated IgG antibody elevation suggesting a Th1-predominated Th1/Th2 mixed immune response had been induced.

Evaluation of the phenotypic change of the T lymphocytes

One week after the final immunization, when compared with the control group [PBS and pcDNA3·1 (+)], the CD4+ T lymphocyte count was significantly reduced (P<0.01), while the CD8+ T lymphocyte count was significantly increased (P<0.01). The CD4+/CD8+ ratio (55:36:27-66 = 2.00) was decreased in the pcDNA3·1(+) -Ts-NBLsp-vaccinated group compared with PBS-vaccinated group (68:45:19-41 = 3.53) (Fig. 4).

Evaluation of cytokine production

As shown in Fig. 5, compared with the control group (PBS), in the pcDNA3·1(+) -Ts-NBLsp-vaccinated group and pcDNA3·1(+) -vaccinated group, the levels of cytokine of Th1-type (IFN-γ) were all significantly increased (P<0.05); however, the levels of Th2-type cytokines (IL-10 and IL-4) were significantly increased (P<0.05) only in pcDNA3·1(+) -Ts-NBLsp-vaccinated group.

Assessment of protective effects of DNA vaccine

The pcDNA3·1(+) -Ts-NBLsp-immunized group exhibited a 77.93% reduction in the ML burden 42 days after challenge relative to the blank control group (PBS) (Fig. 6). Compared with the PBS-vaccinated control mice, the pcDNA3·1(+) -immunized group exhibited 30.34% reduction in ML burden.

DISCUSSION

DNA vaccines can express antigens in proper conformation with post-translational modifications and intracellular trafficking (Ulmer et al. 1993; Xiang et al. 1994; Boudinot et al. 1998). In the present study, we constructed the plasmid of pcDNA3·1(+) -Ts-NBLsp and immunized mice by IM needle...
injection to mimic a NBL stage-specific serine protease expressed during natural infection process. The expression of recombinant Ts-NBLsp in vivo was confirmed by immunofluorescence test, the recombinant Ts-NBLsp was distributed on the inner surface of the membrane of the muscle cells in quadriceps femoris of pcDNA3·1(+)-Ts-NBLsp-vaccinated mice at 48 h post first immunization.

The phenotypic change of T lymphocytes in peripheral blood was evaluated one week after the final immunization, and compared to the control group (PBS). The CD8+ T lymphocyte count significantly increased \((P<0.01)\) in both pcDNA3·1(+)-Ts-NBLsp-vaccinated mice and pcDNA3·1(+)-vaccinated mice, and no significant difference was observed between these two groups. Furthermore, the IFN-γ levels in serum were significantly increased \((P<0.05)\) in both groups. It indicated that in the present study the DNA vaccine has triggered the non-specific immune response. In fact, it has been shown that immune response induced by DNA vaccine is not dependent solely on the expression of antigen, and the DNA molecule itself can act as an adjuvant to enhance the immune response in mammals (Klinman et al. 1997; Krieg et al. 1998).

The result of the present study shows that the pcDNA3·1(+)-vaccinated group exhibited a 30.34% reduction in the ML burden 42 days after challenge compared to the control group. It is suggesting that non-specific immune responses might be useful for the protection against the ML of \(T. spiralis\).

There are two major effector arms of the adaptive immune system, antibodies and T cells, and both are important for resistance to primary infections. The pcDNA3·1(+)-Ts-NBLsp-vaccinated group exhibited a 77.93% reduction in the ML burden. The antibodies and T cells response have been analysed. Compared to the control groups, serum IgG were significantly increased after boost immune in this group, and the IgG isotypes, both IgG1 and IgG2a, were increased, with IgG2a taking the dominant place. Because the IgG isotypes are controlled
by cytokines secreted by the CD4+ T cells, the IgG2 isotype is considered to be associated with a Th1 immune response, whereas the IgG1 isotype is associated with a Th2 response. It is suggested that Th1 predominated Th1/Th2 mixed immune response has been induced in the pcDNA3·1(+)-Ts-NBLsp-vaccinated group.

For the cellular immunologic response, IL-4 is one of the key cytokines in induction of Th2 responses, and IL-10 is an important effector of Th2 responses. The increased expression of these two cytokines suggests that the specific CD4+ Th2 response has been induced. Furthermore, the expression of IFN-γ was significantly increased (P < 0·05) in the pcDNA3·1(+)·Ts-NBLsp-vaccinated group. IFN-γ is a key effector of CD4+ Type 1 and CD8+ T cell, it can activate macrophages and dendritic cells, stimulate increased expression of MHC–peptide complexes, and defend against the intracellular infection. Trichinella sp. are intracellular parasites of muscle cells, and the NBL are without the protection of a capsule. It has been reported that IFN-γ is crucially involved in protection against NBL (Helmby and Grencis, 2003).

Trichinella spiralis can evoke a stage specific protective host immune response through their cuticular and ES antigens of each stage (Wang, 1997). During T. spiralis intestinal infection, CD4+ Th2 cells are critical in host protective immune and inflammatory responses (Ha et al., 1983; Khan et al., 2001). However, at muscular phase of T. spiralis infection, Th1 mediated the destructive granulomatous response, while Th2 are largely not destructive to ML (Li and Ko, 2001; Beiting et al. 2007). Furthermore, Th2-type response will inhibit the Th1-type response. So, the pcDNA3·1(+)·Ts-NBLsp plasmid may have promoted Th1 and Th2 immune balance by inducing Th1 predominated Th1/Th2 mixed immune response.

In conclusion, Ts-NBLsp gene induced an effectively specific immune response in vaccinated mice. We presume that Ts-NBLsp may be an important gene for the survival of T. spiralis; further studies are required to elucidate the roles of this gene. Additionally, for the research on vaccine against
trichinellosis, the DNA vaccine successfully promoted the balance of Th1 and Th2 response, and induced the activity of CD8+ T cells. We presume that these two factors may play an important role in producing a significant protection against *T. spiralis* infection in mice.

**CONFLICT OF INTEREST STATEMENT**

None of the authors has any financial or personal relationship with other people or organizations that could inappropriately influence or bias the paper entitled ‘Immune responses in mice vaccinated with a DNA vaccine expressing serine protease-like protein from the NBL stage of *Trichinella spiralis*’.

**ACKNOWLEDGEMENTS**

We thank Xuejin Su and Xinrui Wang for their technical assistance, our thanks are extended to express our gratitude to all the people who made this work.

**FINANCIAL SUPPORT**

This study was supported by the National Natural Science Foundation of China (grant numbers NSFC31520103916 and NSFC 31402185) and Guangdong Innovative and Entrepreneurial Research Team Program (no. 2014ZT05S123).

**REFERENCES**

Balasubramanian, N., Toubarro, D. and Simoes, N. (2010). Biochemical study and in *vivo* insect immune suppression by a trypsin-like secreted protease from the nematode *Steinernema carpocapsae*. *Parasite Immunology* 32, 165–175.

Beiting, D. P., Gagliardo, L. F., Hesse, M., Bliss, S. K., Meskill, D. and Appleton, J. A. (2007). Coordinated control of immunity to muscle stage *Trichinella spiralis* by IL-10, regulatory T cells, and TGF-beta. *Journal of Immunology* 178, 1039–1047.

Bien, J., Nareaho, A., Varmanen, P., Gozdzik, K., Moskwa, B., Boudinot, P., Blanco, M., de Kinkelin, P. and Benmansour, A. (1998). Proteome digestion in the inspection of meat for *Trichinella spiralis* and Trichinella britovi muscle larvae by two-dimensional difference gel electrophoresis. *Proteome Sciences* 10, 10.

Boudinot, P., Blanco, M., de Kinkelin, P. and Benmansour, A. (1998). Combined immunization with the glycoprotein gene of viral hemorrhagic septicaemia virus and infectious hematopoietic necrosis virus induces double-specific protective immunity and nonspecific response in rainbow trout. *Virology* 249, 297–306.

Cui, J., Wang, Z. Q. and Xu, B. L. (2011). The epidemiology of human trichinellosis in China during 2004–2009. *Acta Tropica* 118, 1–5.

Dorny, P., Praet, N., Deckers, N. and Gabriel, S. (2009). Emerging food-borne parasites. *Veterinary Parasitology* 163, 196–206.

Dupouy-Camet, J. (2000). *Trichinella*: a worldwide zoonosis. *Veterinary Parasitology* 93, 191–200.

Dzik, J. M. (2006). Molecules released by helminth parasites involved in host colonization. *Acta Biochimica Polonica* 53, 33–64.

Feng, S., Wu, X., Wang, X., Bai, X., Shi, H., Tang, B., Liu, X., Song, Y., Boireau, P., Wang, F., Zhao, Y. and Liu, M. (2013). Vaccination of mice with an antigenic serine protease-like protein elicits a protective immune response against *Trichinella spiralis* infection. *Journal of Parasitology* 99, 426–432.

Gottstein, B., Pozio, E. and Nockler, K. (2009). Epidemiology, diagnosis, treatment, and control of trichinellosis. *Clinical Microbiology Reviews* 22, 127–145; Table of Contents.

Ha, T. Y., Reed, N. D. and Crowle, P. K. (1983). Delayed expulsion of adult *Trichinella spiralis* by mast cell-deficient W/Wv mice. *Infection and Immunity* 41, 445–447.

Helmy, H. and Grecius, R. K. (2003). Contrasting roles for IL-10 in protective immunity to different life cycle stages of intestinal nematode parasites. *European Journal of Immunology* 33, 2382–2390.

Khan, W. I., Vaillance, B. A., Biennerhassett, P. A., Deng, Y., Verdu, E. F., Mattheai, K. I. and Collins, S. M. (2001). Critical role for signal transducer and activator of transcription factor 6 in mediating intestinal muscle hypercontractility and worm expulsion in *Trichinella spiralis*-infected mice. *Infection and Immunity* 69, 838–844.

Klinman, D. M., Yamshchikov, G. and Ishigatsubo, Y. (1997). Contribution of CpG motifs to the immunogenicity of DNA vaccines. *Virology* 150, 3635–3639.

Krieg, A. M., Yi, A. K., Schorr, J. and Davis, H. L. (1998). The role of CpG dinucleotides in DNA vaccines. * Trends in Microbiology* 6, 23–27.

Li, C. K. and Ko, R. C. (2001). Inflammatory response during the muscle phase of *Trichinella spiralis* and *T. pseudospiralis* infections. *Parasitology Research* 87, 708–714.

Li, F., Cui, J., Wang, Z. Q. and Jiang, P. (2010). Sensitivity and optimization of artificial digestion in the inspection of meat for *Trichinella spiralis*. *Foodborne Pathogens and Disease* 7, 879–885.

Liu, M. Y., Wang, X. L., Fu, B. Q., Li, C. Y., Wu, X. P., Le Bhun, D., Chen, Q. J. and Boireau, P. (2007). Identification of stage-specifically expressed genes of *Trichinella spiralis* by suppression subtractive hybridization. *Parasitology* 134, 1443–1455.

Murrell, K. D. and Pozio, E. (2011). Worldwide occurrence and impact of human trichinellosis, 1906–2009. *Emerging Infectious Diseases* 17, 2194–2202.

Pozio, E. (1989). [Present knowledge of the taxonomy, distribution and biology of genera of *Trichinella* (Nematoda, Trichinelldae)]. *Annales Dell’istituto Superiore di Sanita* 25, 615–623.

Pozio, E. and Zarlanga, D. S. (2013). New pieces of the *Trichinella* puzzle. *International Journal for Parasitology* 43, 983–997.

Ros-Moreno, R. M., Vazquez-Lopez, C., Gimenez-Pardo, C., de Armas-Serra, C. and Rodriguez-Cabeiro, F. (2000). A study of proteins throughout the life cycle of *Trichinella spiralis*. *Folia Parasitologica* (Praga) 47, 49–54.

Todorova, V. K. (2000). Proteolytic enzymes secreted by larval stage of the parasitic nematode *Trichinella spiralis*. *Folia Parasitologica* (Praga) 47, 141–145.

Todorova, V. K. and Stoyanov, D. I. (2000). Partial characterization of serine proteinases secreted by adult *Trichinella spiralis*. *Parasitology Research* 86, 684–687.

Ulmer, J. B., Donnelly, J. J., Parker, S. E., Rhodes, G. H., Felgner, P. L., Dwarki, V. J., Gromkowski, S. H., Deck, R. R., DeWitt, C. M., Friedman, A., Hawe, L. A., Leander, K. R., Martinez, D., Perry, H. C., Shiver, J. W., Montgomery, D. L. and Liu, M. A. (1993). Heterologous protection against influenza by injection of DNA encoding a viral protein. *Science* 259, 1745–1749.

Wang, C. H. (1997). Study of biological properties of *Trichinella spiralis* newborn larva and the antiparasitic mucosal immunity of the host. *Frontiers in Bioscience* 2, d317–d330.

Wang, Z. Q. and Cui, J. (2001). The epidemiology of human trichinellosis in China during 1964–1999. *Parasite* 8, 863–866.

Wang, Z. Q., Cui, J. and Xu, B. L. (2006). The epidemiology of human trichinellosis in China during 2000–2003. *Acta Tropica* 97, 247–251.

Wang, L., Cui, J., Hu, D. D., Liu, R. D. and Wang, Z. Q. (2014). Identification of early diagnostic antigens from major excretory–secretory proteins of *Trichinella spiralis* muscle larvae using immunoproteomics. *Parasites & Vectors* 7, 40.

Xiang, Z. Q., Spitalnik, S., Tran, M., Wunner, W. H., Cheng, J. and Erdi, H. C. (1994). Vaccination with a plasmid vector carrying the rabies virus glycoprotein gene induces protective immunity against rabies virus. *Virology* 199, 132–140.