Controlling the adsorption of osteopontin for mediating cell behaviour by using self-assembled monolayers with varying surface chemistry†

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Osteopontin (OPN) is an important protein for mediating cell behaviour on biomaterials. However, the interactions between the chemical groups on the biomaterial surface and OPN still need to be further clarified, which has restricted the application of OPN in biomaterial functionalization. In the present study, we developed different self-assembled monolayers (SAMs) with specific chemical groups, including SAMs-OH, SAMs-OEG, SAMs-COOH, SAMs-NH2, and SAMs-PO3H2, to study the behavior of OPN on these SAMs. The results showed that SAMs-NH2 could strongly adsorb OPN, and the amount of protein was highest on this material. Meanwhile, the lowest amount of OPN was present on SAMs-OEG. Interestingly, the unit-mass trend of bound OPN monoclonal antibodies (mAbs) on the SAMs was opposite to the OPN adsorption trend: lowest on SAMs-NH2 but highest on SAMs-OEG. In vitro cell assay results showed that mouse bone marrow mesenchymal stem cells (mBMSCs) on SAMs-COOH, SAMs-NH2, and SAMs-PO3H2 with pre-adsorbed OPN showed promoted behaviour, in terms of spreading, viability, and the expression levels of αv and β3 genes, compared with the other two SAMs, demonstrating the higher bioactivity of the adsorbed OPN. We believe that our findings will have great potential for developing OPN-activated biomaterials.

Among these strategies, surface chemistry can greatly influence protein adsorption and subsequent cellular behaviors. To obtain general information, self-assembled monolayers (SAMs) on model substrates with single or mixed chemical groups have been developed. By this method, researchers have explored the surface-related interactions between cells and proteins on different surfaces, e.g., in studies involving fibronectin, vitronectin, laminin, and collagen, and demonstrated that various chemical groups on SAMs can direct the fates of cells. These universal conclusions have greatly improved the performances of biomaterials.

However, it is still meaningful to further explore the relationships between surface chemical groups and other significant proteins. Among these proteins, osteopontin (OPN) is a member of the small integrin-binding ligand N-linked glycoprotein (SIBLING) family, and it is closely involved in the biological functions of several tissue types. For example, OPN present in bone tissue can activate alkaline phosphatase to improve bone remodeling. Therefore, it is worth clarifying OPN adsorption behavior and subsequent cell adhesion for the development of biomaterials.

In this study, we evaluated OPN adsorption on SAMs with different chemical groups, i.e., -OH, -OEG, -COOH, -NH2, and -PO3H2 (Fig. 1a). Subsequently, we explored the correlation between OPN adsorption and the behavior of mouse bone marrow mesenchymal stem cells (mBMSCs). These findings will be of great significance for the rational design and development of OPN-activated biomaterials.

1. Introduction

The properties of biomaterials are mainly determined by the proteins adhering to their surfaces. These proteins will regulate the cellular behaviors, e.g., the cell adhesion and subsequent cell migration, proliferation, and differentiation. Therefore, regulating the adhesion of proteins on the surface has become a significant strategy when developing novel biomaterials. Recently, researchers have regulated the adhesion of proteins via controlling the topology and composition of the surface, or even through pre-immobilizing expected proteins or peptides. Through these strategies, the performances of biomaterials can be improved significantly.
2. Experimental

2.1 Preparation of SAMs

After being cleaned via nitrogen plasma treatment for 2 min, freshly cleaned gold slides (4 mm × 4 mm) were immediately immersed in either 11-mercapto-1-undecanol (HS(CH$_2$)$_{11}$OH, Dojindo, Japan), (11-mercaptoundecyl)hexa(ethylene glycol) (HS(CH$_2$)$_{11}$(OCH$_2$CH$_2$)$_6$OH, Dojindo, Japan), 10-carboxy-1-decanethiol (HS(CH$_2$)$_{10}$COOH, Dojindo, Japan), 11-amino-1-undecanethiol hydrochloride (HS(CH$_2$)$_{11}$NH$_2$HCl, Dojindo, Japan), or 11-mercaptoundecylphosphoric acid (HS(CH$_2$)$_{11}$-OPO$_3$H$_2$, Sigma, USA) at 1 mmol L$^{-1}$ and left overnight. Then, the substrates were rinsed alternately with ethanol and deionized water and dried under a stream of nitrogen gas. These samples are referred to as SAMs-OH, SAMs-OEG, SAMs-COOH, SAMs-NH$_2$, and SAMs-PO$_3$H$_2$.

2.2 Surface characterization

Static water contact angle measurements were performed using a contact angle meter (OCA15, Data Physics, Germany) ($n = 3$). The zeta potentials of surfaces were determined with an electrokinetic analyzer for solid surface analysis (SurPass, Anton Paar Austria) at a given pH value via pumping 10$^{-3}$ mM KCl electrolyte ($n = 4$).

2.3 Protein adsorption

Recom mouse osteopontin (OPN) (R&D Systems, USA) was radiolabeled with $^{125}$I, as described in another reference,$^{23}$ and then purified via size exclusion chromatography using Sephadex$^\text{TM}$ G-25 medium (GE Healthcare, Bio-Sciences AB, Sweden). SAM samples were immersed in protein solution (10 $\mu$g mL$^{-1}$ in PBS) and incubated at 37 $^\circ$C for 1 h. After rinsing with PBS, the radioactivity levels of samples were measured using a gamma radio-immunoassay counter (GC-300, USTC Chuangxin Co., Ltd, Zonkia Branch, China). Then the samples with adsorbed OPN were incubated in complete medium with 10% fetal bovine serum (FBS, Gibco, USA) at 37 $^\circ$C for 1 h, rinsed, and measured again as previously described. The amount of adsorbed OPN on the surface (4 mm × 4 mm) was calculated from the radioactivity using the following equation ($n = 3$):

\[
\text{Protein (ng cm}^{-2}\text{)} = \frac{\text{count (cpm)}}{[A_{\text{solution (cpm ng}^{-1})} \times S (\text{cm}^2)]}
\]

where count represents the radioactivity of the sample, and $A_{\text{solution}}$ and $S$ are the radioactivity of the protein solution and the surface area, respectively.

The exposure levels of cell-binding domains in adsorbed OPN were characterized via enzyme-linked immunosorbent assay (ELISA) studies with OPN monoclonal antibodies (OPN mAbs) (R&D Systems, USA) according to the manufacturer instructions ($n = 3$).

Besides OPN, we further characterized the adsorption of bovine serum albumin (BSA) (Sigma, USA) on the SAMs via surface plasmon resonance (SPR, Plexera$^\text{®}$ Bioscience LLC, USA) studies ($n = 5$). Briefly, SAMs were prepared on the SPR sensor (25 mm × 75 mm × 1 mm) surface via immersing the sensor in 12.5 mL of reaction solution (1 mM) for 24 h at room temperature, followed by washed by ethanol and deionized water. Then, each surface was treated with PBS, 50 $\mu$g mL$^{-1}$ BSA, and PBS, successively, to remove weakly adsorbed proteins. An auto-injection system was used to control the sample flow at...
a rate of 2 μL s⁻¹. The SPR sensorgram is expressed in refractive units (RU), where 1000 RU corresponds to a protein density of 1 ng mm⁻².

2.4 Cell adhesion and gene expression on SAMs

Mouse bone marrow mesenchymal stem cells (mBMSCs) (CRL-12424, ATCC, USA) from early passages (≤6) were collected for cell assays. The SAM substrates (10 mm × 10 mm) were immersed in 10 μg mL⁻¹ OPN solution and incubated at 37 °C for 1 h. After rinsing with PBS, cells (5 × 10⁴ cells per sample) were seeded on SAM samples with pre-adsorbed OPN and cultured in a serum-free medium. After 12 h of culturing, the cell cytoskeletal filamentous actin and nuclei were stained with Phalloidin FITC probe (AAT Bioquest® Inc., USA) for 20 min and DAPI (Beyotime, China) for 5 min, respectively. The cell morphologies were observed via confocal laser scanning microscopy (CLSM, Leica TCS SP5, Germany). Additionally, after 12 h of culturing, the viabilities of the adhered cells on SAMs was evaluated via Cell Counting Kit-8 (CCK-8, Dojindo, Japan) assays, immersing the samples into 300 μL of CCK-8 working solution for 1 h of incubation. The absorbance at 450 nm was measured with a microplate reader (Thermo3001, USA).

The gene expression levels of αv, β3, and β1 integrin in cells were tested via qRT-PCR assays. Briefly, after 12 h of culturing (5 × 10⁴ cells per sample), the total RNA in the cells was isolated using HiPure Total RNA Kits (Magentec, China) and reverse-transcribed into cDNA using a PrimeScript® RT Reagent Kit with a gDNA Eraser (TaKaRa Biotechnology, Japan) according to the manufacturer protocol. RT-PCR reactions were conducted using an SYBR green system (Invitrogen, USA) as follows: heating at 95 °C for 10 min and then 40 cycles at 95 °C for 15 s and 60 °C for 1 min. The adopted primers (TaKaRa Biotechnology, Japan) are as follows – β-actin: forward: TGCTCTCAGAACCTGCAAG, reverse: GCTGGAAGGTGACAGTGGA; αv integrin: forward: TGAACCTGACCGCAGATACAG, reverse: TAGCGGCGGAGTCTGTCTCAC-TAC; β3 integrin: forward: TCAATGGCCACCTGGCTCAA, reverse: CCTTGGCTCGATACAAAGCTCA.

2.5 Statistical analysis

Statistical comparisons were carried out via analysis of variance (ANOVA) followed by post-hoc tests. A p value < 0.05 was considered to be statistically significant.

3. Results and discussion

The chemical groups strongly affected the wettability and zeta potential of the SAMs. The contact-angle results showed that all the SAMs were hydrophilic (Fig. 1b). Specifically, SAMs-OH had the lowest contact angle of 22.6°, while SAMs-OEG and SAMs-COOH had contact angles of 37.7° and 37.3°, respectively. Meanwhile, the contact angles of SAMs-NH₂ and SAMs-PO₃H₂ increased to 67.5° and 72.8°, respectively. Additionally, the zeta potential results at pH 7.4 (Fig. 1c) showed that positively charged SAMs-OEG and SAMs-NH₂ had the highest zeta potentials of −21.49 mV and −22.09 mV and negatively charged SAMs-COOH had the lowest zeta potential of −83.63 mV. The zeta potentials of SAMs-OH and SAMs-PO₃H₂ were −59.95 mV and −55.82 mV, respectively. Some of the results had a similar trend to what was seen in our previous study, demonstrating that SAMs exhibit distinct properties.

OPN adsorbed on the SAMs with significantly different adsorption behaviours. To quantify the amounts of OPN on the surfaces, we labelled the protein via radiolabeling, and the calculated iodination yield of 125I-OPN was as high as 93.9% (Fig. 2a). The 125I-OPN count results (Fig. 2b) showed that SAMs-NH₂ could strongly adsorb OPN; the adsorbed amount was 89.01 ± 13.62 ng cm⁻², which was 4.36-, 26.24-, 12.31- and 10.47-times the values of SAMs-OH, SAMs-OEG, SAMs-COOH and SAMs-PO₃H₂, respectively. This is likely to be caused by electrostatic forces between negatively charged OPN and the positively charged surface. We also demonstrated that with an increase in pH from 5 to 9, the adsorbed amount of OPN increased on SAMs-NH₂, further verifying the presence of electrostatic forces, as an increase in the pH value will increase the negative charge of OPN (Fig. S1 and S2†). Notably, the adsorbed amounts of OPN on SAM-OEG, SAM-OH, SAM-COOH, and SAM-PO₃H₂ showed a negligible relationship to the pH value, demonstrating that electrostatic interactions were not dominant on these surfaces. Protein adsorption and interactions are governed by many factors, such as solution pH, ionic strength, the chemical nature of the salt, surface hydrophobicity, the surface charge distribution, and the protein phase behavior. Meanwhile, the amount of OPN on neutral SAMs-OH was 20.44 ± 4.73 ng cm⁻², which was higher than the amounts on negatively charged SAMs-COOH and SAMs-PO₃H₂ (7.23 ± 1.02 ng cm⁻² and 8.50 ± 1.98 ng cm⁻², respectively). The amount of OPN was lowest on SAMs-OEG (3.39 ± 0.63 ng cm⁻²) due to the non-fouling effects of OEG molecules. Additionally, the retained amounts of adsorbed 125I-OPN on SAMs after treatment with serum-containing medium were also measured (Fig. 2b). The results showed that serum proteins reduced the amounts of adsorbed OPN, demonstrating that there was competitive adsorption between serum proteins and pre-adsorbed OPN. However, treatment did not change the final adsorption trend, implying that adsorption between OPN and the SAMs was relatively stable.

Besides the adsorption amount, the chemical groups further affected the conformation of the proteins on SAMs. The ELISA results from bound OPN mAbs (Fig. 2c) showed that the exposure levels of cell-binding domains in adsorbed OPN were consistent with the adsorbed amounts of OPN. The amount of bound OPN mAbs on SAMs-NH₂ was highest, and it was 1.49-, 2.38-, 1.48-, and 1.32-times the amounts on SAMs-OH, SAMs-OEG, SAMs-COOH, and SAMs-PO₃H₂, respectively. Additionally, although there was more OPN on SAMs-OH than on SAMs-COOH and SAMs-PO₃H₂ (Fig. 2b), these three SAMs had comparable levels of bound OPN mAbs. Interestingly, the unit-mass trend of bound OPN mAbs on the SAMs was opposite to the OPN adsorption trend. Specifically, the unit mass of bound OPN mAbs on SAMs-NH₂ was lowest (Fig. 2d). One of the possible reasons was electrostatic forces between the surface and the protein, which could limit the flexibility of the adsorbed...
protein and decrease the exposure of bound OPN mAbs. The unit mass of bound OPN mAb on SAMs-OEG was highest, as the weak interactions resulted in the high flexibility of the protein on the surface. Steric factors are another possible reason. The OPN molecules on SAMs would interact with other OPN molecules, and the higher density could significantly decrease the exposure of molecules. Additionally, a difference in OPN attachment points on the SAMs could also impact the unit mass of bound OPN mAbs.

We further characterized the adsorption behaviour of BSA on the SAMs (Fig. 3), as serum albumin is one of the most abundant plasma proteins. The SPR results showed that BSA had similar adsorption behaviour to OPN, and there was much more BSA on SAMs-NH₂ than the other four SAMs. Additionally, BSA has less negative charge, is less phosphorylated, and has more hydrophobic pockets; the amount of BSA absorbed on SAMs-NH₂ was higher than the amounts of OPN adsorbed on SAMs-PO₃H₂, SAMs-OH, SAMs-OEG, and SAMs-COOH. As expected, the minimum amount of adsorption was observed on SAMs-OEG.

As previously reported, cell adhesion is important for transmitting signals, and it can regulate subsequent cell behaviour. Therefore, we employed in vitro cell assays to verify the bioactivity of OPN for cell adhesion. The morphologies of mBMSCs on SAMs with pre-adsorbed OPN (Fig. 4a) showed that the cells spread with obvious pseudopodia on SAMs-COOH, SAMs-NH₂, and SAMs-PO₃H₂ had higher viability than those on SAMs-OH (2.15-, 2.17-, and 2.21-fold higher, respectively) and SAMs-OEG (3.54-, 3.57-, and 3.64-fold higher, respectively). As reported, a high level of bound OPN mAbs indicates more binding domains for cells. However, although the level of bound OPN mAbs on SAMs-NH₂ was highest, its cell viability was similar to those of SAMs-COOH and SAMs-PO₃H₂. This might be because bound OPN mAbs have a critical value for cell adhesion. Additionally, although the level of bound OPN mAbs on SAMs-OH was similar to those on SAMs-COOH and SAMs-PO₃H₂, its cell viability was much lower. This may be caused by the...
superhydrophilicity of the surface, which would impact the adhesion of cells due to the existence of a water membrane.\textsuperscript{35}

Additionally, higher expression levels of \( \alpha_v \) and \( \beta_3 \) by cells on SAMs-COOH, SAMs-NH\(_2\), and SAMs-PO\(_3\)H\(_2\) were observed. Specifically, the \( \alpha_v \) gene expression levels on these three surfaces were 1.49, 1.28, 1.37-times higher, respectively, than on SAMs-OH, and 6.18, 5.33, 5.71-times higher, respectively, than on SAMs-OEG. The \( \beta_3 \) gene expression levels on these three surfaces were 1.61, 1.35, and 1.36-times higher, respectively, than on SAMs-OH, and 13.29, 11.08, and 11.21-times higher, respectively, than on SAMs-OEG. As has been reported, primary interactions between cells and underlying proteins occur via integrins.\textsuperscript{36} The upregulation of integrins was observed when their preferred ligands were available.\textsuperscript{37} OPN has RGD adhesion motifs that are particularly likely to bind with integrins like \( \alpha_v \beta_3 \), \( \alpha_v \beta_5 \), and so on,\textsuperscript{38} and it has been suggested that the expression of integrin subunit \( \alpha_v \) and \( \beta_3 \) binding adhesion protein ligands can influence cell adhesion and subsequent activity.\textsuperscript{39} Therefore, the different expression levels of \( \alpha_v \) and \( \beta_3 \) by the cells on SAMs demonstrated that different chemical groups influenced cell adhesion through integrin-OPN binding.

Above all, we demonstrated that different chemical groups on SAMs affect the adsorption and conformation of OPN, further leading to different cell behaviours.

4. Conclusions

We demonstrated that the chemical groups on SAMs strongly affected the behavior toward OPN and the performance of OPN. The amount of adsorbed OPN was highest on SAMs-NH\(_2\) (89.01 \( \pm \) 13.62 ng cm\(^{-2}\)) and lowest on SAMs-OEG (3.39 \( \pm \) 0.63 ng cm\(^{-2}\)). Interestingly, the unit-mass trend of bound OPN mAbs on SAMs was opposite to the OPN adsorption trend, which was because stronger interactions resulted in lower flexibility being shown by the adsorbed proteins. Meanwhile, cells on SAMs-COOH, SAMs-NH\(_2\), and SAMs-PO\(_3\)H\(_2\) with pre-adsorbed OPN showed larger spreading, better viabilities, and higher expression levels of \( \alpha_v/\beta_3 \) genes than those on SAMs-OH and SAMs-OEG, demonstrating that OPN on these three SAMs exhibited higher bioactivity. Considering the significant effects of OPN on the biological functions of several tissue types, we believe that our research will provide great guidance for developing OPN-activated biomaterials.

Conflicts of interest

There are no conflicts of interest to declare.

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