Introduction

Parasitic diseases have worldwide distribution with increased prevalence in areas of poor sanitation. In nonendemic areas, cases also occur because of an increase in international travel and host immunosuppression caused by HIV infection or immunosuppressive therapy. Despite less common than bacterial or viral infection, parasitic infection of central nervous system (CNS) can be fatal if undetected. Especially, parasitic infection of pituitary gland is very rare and unexpected. It can be difficult to make a correct diagnosis which can lead to delay in treatment and be associated with the risk of life-threatening conditions. Although many reviews have been published about parasitic infection of the brain, parasitic infection in sella turcica area has received little attention. In this review, we bring parasitic infection of pituitary gland to light, emphasizing on clinical features, significance and diagnosis of this entity.

Methods

A literature search of case reports or case series of patients with parasitic infection of pituitary was performed using the electronic database of MEDLINE, PubMed, and Scopus. The search was updated to June 2016. The keywords used in the literature were the combinations of name of organism or parasitic diseases such as toxoplasma/toxoplasmosis, cysticercosis/neurocysticercosis and words associated with pituitary gland such as “sellar”, “intrasellar”, “hypophysis”, “hypophyseal”, and “pituitary”. Full articles written in English and relevant to the topic were included in this review. We included the original publication which allows data abstraction on clinical, laboratory and neuroimaging findings whether intrasellar parasitic infection occurred as the only presentation of the disease or as the part of a more disseminated infection.
Results

We assessed information of 23 full-text papers (24 case reports and 3 case series). The total 34 patients were analyzed and information was included in this review [1-24].

Demographic data

For infection in 28 adults, patient age ranged in age from 18 to 81 years (median 42 years); 13 (46%) of patients were male. For infection of children, 6 cases of congenital toxoplasmosis were included; patient age ranged from at birth to 1 year (median 2 months); 4 (67%) of patients were male. The patients were reported from the USA, Mexico, Germany, Turkey, China, Colombia, India, Thailand, Korea, Belgium, Switzerland, Greece and Saudi Arabia.

Parasitology

Out of 28 adult patients and 6 children, twenty patients were infected with Taenia solium presenting with intrasellar neurocysticercosis. Ten of patients were infected with Toxoplasma gondii (6) cases involved with congenital toxoplasmosis and 3 cases associated with pituitary adenoma and the other case presenting as complication from HIV infection). Two of patients had cerebral malaria. The others were one case of pituitary amebiasis and one case of pituitary hydatid disease. Life cycles, route of transmission, and frequent clinical presentation of each parasitic organism are presented in Table 1.

Clinical presentation

In adult group, headache was the most common presenting symptom (64%) and the most common symptom (86%). Second most common symptom was ophthalmological disturbance (61%) defined by decrease visual acuity, visual field defect (bitemporal hemianopia and superior hemianopsia) and bilateral exophthalmos. Endocrine abnormalities were noted in 19 patients (68%) including 7 with panhypopituitarism (with co-existing diabetes insipidus), 7 with hyperprolactinemia, 2 with isolated diabetes insipidus, 3 with partial hypopituitarism (isolate hypothyroidism, isolate adrenal insufficiency with diabetes insipidus and one not classified). Other symptoms included seizure, fever, nausea and focal neurological symptoms. Fever was present in 2 patients (7%) who both had malarial infection.

From 6 pediatric cases of congenital toxoplasmosis, sepsis-like symptoms and seizure are common (50%). Other symptoms included microcephaly, hydrocephalus and polyuria. Endocrine abnormalities included 2 cases with isolated diabetes insipidus, four with panhypopituitarism (2 cases with co-existing diabetes insipidus and one with precocious puberty during follow-up visits). The most common endocrine disturbance was diabetes insipidus (67%).

Diagnosis

The majority of the patients have a definitive diagnosis of infection from identifying parasitic organism from pathology except all cases of congenital toxoplasmosis, malaria and 4 cases of cysticercosis which diagnosis was established on the basis of neuroimaging findings and immune diagnosis tests. Duration from onset of symptoms to diagnosis varied from each parasitic disease. Acute to subacute onset included amebiasis, malaria (1 week) and toxoplasmosis in HIV person (3 weeks). Chronic onset included cysticercosis (4 months), hydatid cyst (7 months) and toxoplasmosis with pituitary adenoma (3.5 years).

| Parasite            | Geographical Distribution | Definitive host | Intermediate host | Transmission                                             | Disease                    | Clinical manifestation                                |
|---------------------|---------------------------|----------------|-------------------|----------------------------------------------------------|----------------------------|------------------------------------------------------|
| Taenia solium       | Worldwide                  | Humans         | Swine             | Ingestion of cysticercus cellulose                       | Taeniasis                  | Gastrointestinal symptoms                             |
|                     |                           |                |                   | Ingestion of eggs                                      | Cysticercosis              | Seizures, headache, focal neurologic deficits, altered cognition, and psychiatric disease |
| Toxoplasma gondii   | Worldwide                  | Cat            | Humans, animals   | Ingestion of tachyzoites or oocysts                     | Primary infection in the immunocompetent person | Asymptomatic, lymphadenopathy syndrome               |
|                     |                           |                |                   | Placenta                                               | Congenital toxoplasmosis   | Hydrocephaly, microcephaly, mental retardation, seizures, visual defects |
|                     |                           |                |                   | Reactivation from latent infection                     | Disease in the immunocompromised person | Encephalitis, brain abscess, chorioretinitis and pneumonitis |
| Plasmodium spp.     | Subtropics and tropics     | Humans         | Mosquito          | Ingestion of Anopheles bite                             | Malaria                    | Recurrent fever, shaking chills, sweats, headache, myalgia, vomiting, splenomegaly |
| Echinococcosis spp. | Sheep-raising and hunting areas | Dogs, cats | Humans, sheep | Ingestion of eggs | Hydatid disease | Abdominal or chest pain, biliary obstruction, cholangitis, cough, chest pain, dyspnea and hemoptysis, severe allergic reaction, |
| Entamoeba histolytica | Worldwide, especially tropics | Humans | None, survive encysted in environment | Ingestion of viable cysts | Intestinal amebiasis | Asymptomatic, amoebic dysentery, colitis |

Citation: Assavapongpaiboon B, Salehian B (2017) Intrasellar Parasitic Infection: Finding the Culprit - How to Get the Right Diagnosis. Int J Clin Endocrinol Metab 3(1): 008-015.
Discussion

Neurocysticercosis

*Taenia solium* is the most common parasite which involves the sellar region causing neurocysticercosis. Over 50 million people around the world are estimated to have cysticercosis infection, although estimates are probably low since many infections are subclinical. Central and South America, sub-Saharan Africa, India, and Asia are endemic areas for *T. solium* [25]. Neurocysticercosis is divided into parenchymal and extraparenchymal forms. Extraparenchymal forms include intraventricular, subarachnoid space of brain and spinal and intraocular. However, sellar involvement is rare. Definite diagnosis can be difficult without the presence of parasite (see the absolute criteria below). Therefore, diagnostic criteria using clinical presentation, radiographic studies, serologic tests, and exposure history was delineated by a consensus conference [26].

The characteristic imaging of neurocysticercosis includes cystic lesions with or without eccentric scolex which can be associated with perilesional edema and enhancing lesion depending on the stages. One or more nodular calcifications can be seen (Figure 2). Parenchymal cysterci are usually round and 10–20 mm in diameter. In contrast, cystic lesions in the subarachnoid space are larger and various in size as there is no limiting parenchymal tissue in these sites [27]. The form of subarachnoid neurocysticercosis has distinguishing characteristic of grape–liked clusters of cysts in the cisterns called “racemosecysticercus”. This is a proliferating form of larval tapeworm containing multiple interconnected cysts of different sizes without scolecis [28]. Racemosecysticercosis occurs primarily in parts of the nervous system where the parasite is not closely enclosed by host tissues, such as the cisterns at the base of the brain including suprasellar cistern which may extend into the sella turcica [2,8]. Diagnosis is more challenging when the sellar cisterceri are the sole manifestation of the disease and no densities within the cysts that indicated the presence of scolecis. In our review, nearly 50% of lesions confined to the sellar region which should be differentiated from other causes of cystic lesions such as craniopharyngioma, cystic pituitary adenoma, arachnoid cyst, Rathke’s cleft cyst, epidermoid cyst and hydatid cyst. Extracellular finding such as subarachnoid cistercicercosis involving the basal CSF cisterns (associated or not with hydrocephalus) and the brain parenchymal cysts may facilitate diagnosis. Specific antibodies to cysterci were included in criteria. An immunoblot assay using lentil lectin purified glycoproteins is 98% sensitive and 100% specific [29]. However, patients with single intracranial lesions or with calcifications may be seronegative [30]. With this assay, it is 10% more sensitive using serum than for CSF specimens. On the other hand, enzyme–linked immunosorbent assays for CSF, not serum, are reliable and interpretation should be done with caution due to high false–positive and false–negative rates [31].

Toxoplasmosis

Interestingly, toxoplasmosis has been incidentally found in 3 patients diagnosed with prolactinoma presenting with amenorrhea and MRI suggestive of pituitary adenomas [13,16]. Due to the recurrent of pituitary adenoma after surgery and inadequate response to dopamine agonist therapy; surgical removal of tumor was needed. The presence of toxoplasma brady cyst was revealed in all cases, 2 of them had prolactin producing adenoma and one with nonfunctioning adenoma. It seems this is a coincidental toxoplasmosis of pituitary and pituitary adenoma, which was detected at the time of the surgery; the prevalence of pseudocyst of toxoplasmosis in normal pituitary or microadenoma is unknown.

Approximately 25 to 30% of the world’s human population is infected by *Toxoplasma gondii* [32]. Europe, Central America, Brazil, and Central Africa are the areas which the highest rates of infection with *T. gondii* have been reported [33]. *T. gondii* infection presents a wide range of clinical manifestation. In congenital form, majority of cases are asymptomatic. However, acute manifestations may occur as nonspecific septic–like features with fever, anemia, jaundice, seizure and hydrocephalus. Occasionally, few infants have late presentation with neurologic symptoms such as sensory–motor...
neural deafness, microcephaly and developmental delay [34].

Differential diagnosis includes other intrauterine infections that have similar manifestation in the newborn such as rubella, cytomegalovirus, syphilis and congenital lymphocytic choriomeningitis virus syndrome and Zika virus [35,36].

Computed Tomography (CT) without contrast is preferred because it is fast, less expensive and can be done without sedation. Intracranial calcifications scattered all over the brain, hydrocephalus and cortical atrophy are characteristic findings. Usually, serologic testing has become the routine method of diagnosis. For accurate diagnosis, testing of blood sample from both the infant and the mother is required. Results from mother typically have positive Toxoplasma IgG and IgM antibody. For diagnosis of newborn, presence of Toxoplasmic IgM is diagnostic [37]. When IgM titers are negative or unequivocal, IgA and IgE ELISA should be performed [38,39].

Serial serologic testing during the first year of life is required for diagnosis when the initial results are equivocal. T. gondii polymerase chain reaction (PCR) on amniotic fluid and infant CSF are also useful for diagnosis in the case with characteristic clinical findings (chorioretinitis, intracranial calcification and/or hydrocephalus) [40]. In this review, diabetes insipidus is the most common endocrine (67%). Therefore, polyuria in the background of congenital toxoplasmosis raises concern of pituitary involvement.

Reactivation of toxoplasmosis commonly seen in HIV-infected person and other immunocompromised conditions is another manifestation presenting with encephalopathy. For HIV-infected patients, a presumptive toxoplasmosis can be made if the patient has a CD4 count <100 cells/μL, has not been receiving proper prophylaxis to prevent toxoplasmosis and has all of the following: 1) compatible clinical syndrome: fever, headache, focal neurologic deficits, seizures, confusion and mental status change 2) positive T. gondii IgG antibody 3) Brain imaging (preferably magnetic resonance imaging) that demonstrates a typical radiographic appearance (e.g., single or multiple ring-enhancing lesions <2 cm) with the basal ganglia and cortico-medullary junction most commonly involved. Toxoplasma encephalitis can be diagnosed with a 90% probability with the presence of these criteria [41,42].

CSF analysis for T. gondii PCR should also be performed to establish the diagnosis, when the patient is safe to undergo lumbar puncture, due to high specificity (96 to 100%) [43,44]. Moreover, CSF analysis can suggest alternative diagnoses such as primary CNS lymphoma (Epstein–Barr virus positive), progressive multifocal leukoencephalopathy (JC virus positive), mycobacterial infection (culture for mycobacterial positive), cryptococcosis (cryptococcal antigen positive) and bacterial abscess [45]. Also, a therapeutic trial of anti-Toxoplasma medications (pyrimethamine plus sulfadiazine or clindamycin) is frequently used to assess the diagnosis. If patients fail to respond or have lymphoma, clinical signs and symptoms worsen by day 7. These patients require brain biopsy by a stereotactic CT-guided method with or without a change in therapy.

Malaria

In general, up to 21% of severe Plasmodium falciparum infections are complicated with polyuria. However, about 66% of polyuria cases has central diabetes insipidus (DI) after laboratory investigation [46]. Other 2 case reports showing cerebral malaria with documented central DI [20,21]. Pathogenesis of DI in cerebral malaria could have been a result of obstruction of neurohypophyseal microvasculature by the malarial parasites. With treatment of the parasitemia, the microvascular obstruction gradually resolves and hence the requirement of desmopressin is temporary. In addition, severe falciparum malaria is associated with secondary adrenal insufficiency. Blunted ACTH response to CRH stimulation was found in patients with severe malaria. This may reflect impairment of corticotroph function due to parasitic erythrocyte sequestration in hypothalamic–pituitary portal system or somatostatin-like peptide produced by parasite [47].

Malaria prevails throughout most of the tropical regions of the world, with P. falciparum causing the largest burden of disease predominating in Africa, New Guinea, and Hispaniola (Haiti and the Dominican Republic) [48]. One of the pictures of severe P. falciparum infection is cerebral malaria. Malaria should be suspected in the setting of fever and history of exposure to endemic area (residence in or travel to an area where malaria is endemic) [49]. Clinical tools for parasite-based diagnosis include microscopy (visualization of parasites in stained blood smears) and rapid diagnostic tests (RDTs; which detect antigen or antibody). Detection of parasites on Giemsa-stained blood smears by light microscopy is the standard tool for diagnosis of malaria [50]. However, microscopy cannot reliably detect very low parasitemia (<5 to 10 parasites/μL) or cases where the majority of the parasite biomass is sequestered (e.g., in the case of placental sequestration during pregnancy) [51]. RDTs should be used if microscopy is not readily available due to their accuracy and ease of use. RDTs usually detect Plasmodium antigens including histidine-rich protein 2 (HRP2), Plasmodium lactate dehydrogenase (pLDH), and aldolase. The approach to RDT selection for Plasmodium species depends.
on the epidemiology of infection and goals for control in the region where the test is used [52]. Molecular techniques for detection of genetic material are limited to research settings.

**Hydatid disease**

Cystic echinococcosis is a disease caused by tapeworm *Echinococcus granulosus* infection. These infections are prevalent in most areas where livestock is raised in association with dogs. These parasites are found on all continents which areas of high prevalence include China, central Asia, the Middle East, the Mediterranean region, eastern Africa, and parts of South America [53]. Echinococcal cysts enlarge slowly and patient is usually asymptomatic. Their expanding size and space-occupying effect in an involved organ elicit symptoms. The liver and the lungs are the most common sites of these cysts. Common symptoms are right upper quadrant pain, nausea, and vomiting in liver involvement or cough, chest pain, dyspnea, and hemoptysis in lung involvement [54]. Cerebral cysts are approximately present in 2–3% of all cases. Headache is the most common presenting symptom of cerebral hydatid cysts (70–75%), followed by weakness in the extremities. Other symptoms are epilepsy, mental changes, skull deformities and, more rarely, dyskinetic phenomena [55]. Complication includes cyst rupture presenting with fever and acute hypersensitivity reactions, including anaphylaxis.

For untreated and uncomplicated hydrated brain lesion, MRI brain usually reveals well-defined large, non-enhancing cystic lesion with no perilesional edema [56]. The cyst may be single or multiple and may or may not show septation. Calcifications are rare and occur in less than 1% [57]. However, the pathognomonic finding, daughter cysts within the larger cyst is rare [58]. Infection can complicate the cysts which may turn to have a multilocular appearance with septations with perilesional edema. Infected cysts resemble gliomatous tumors, but gliomas usually have thick and nodular enhancing walls [59]. Serology is both useful for diagnosis and for follow-up after treatment [60,61]. Antibody detection is more sensitive than antigen detection for diagnosis of *E. granulosus*. The methods most frequently employed for initial screening tests are ELISA, most sensitive and specific of the available assays, and indirect hemagglutination (IHA). These tests use crude antigens such as hydatid fluid or protoscoleces extracts. Confirmatory tests using specific antigens can then be performed, such as immunoelectrophoresis and immunoblotting [62].

**Amoebic disease**

Amoebic disease is a result of *Entamoeba histolytica* infection. The disease is 7 to 10 times more common among adult men, generally seen in migrants from and long-term travelers to endemic areas including India, Africa, Mexico, and parts of Central and South America [63,64]. Worldwide, approximately 50 million people develop colitis or extraintestinal disease, with over 100,000 deaths annually [65]. Extraintestinal manifestations include amebic liver abscess and other more rare manifestations such as liver, pulmonary, cardiac, or brain involvement. Cerebral amebiasis results from with hematogenous spread of infection. Abrupt onset of symptoms such as meningeval signs, facial nerve palsy, motor paralysis, and seizure and rapid progression to death if untreated is characteristic [66]. There is no pathognomonic finding on CT scan. Irregular foci without a capsule or surrounding enhancement, however, are suggestive [67]. Serology is helpful for diagnosis in amoebiasis involving the brain especially cases

| Parasite | Disease | Suggested diagnostic method | Parasitological method | Other serologic methods and molecular diagnosis | Medication for treatment |
|----------|---------|-----------------------------|------------------------|-----------------------------------------------|--------------------------|
| **Taeniasis ovis** [69,70] | Neurocysticercosis | Serum EITB, CSF ELISA | Recognize cysticercus cellulosae in CNS | Complement fixation, IHA | Active lesions*: albendazole, praziquantel plus corticosteroid Inactive lesion**: No treatment |
| **Toxoplasma gondii** [71-73] | Congenital toxoplasmosis | Maternal and infant serum ELISA or ISAGA | Recognize trophozoite or cyst in tissue biopsy (brain, eye) and CSF, animal inoculation | The Sabin-Feldman dye test, IFA, IIF, avidity testing and PCR | Pyrimethamine plus sulfadiazine or atovaquone or clarithromycin or dapsone, clindamycin, trimethoprim-sulfamethoxazole |
| **Plasmodium falciparum** [49,74] | Malaria | Microscopic examination ± rapid diagnosis tests (immunochromatography) | Microscopic examination of thick and thin blood smears | ELISA, IFA, IHA and PCR | Quinine, quinidine, mefloquine,atovaquone-proguanil, doxycycline, artemisinins , fansidar, pyronaridine |
| **Echinococcosis spp.** [75,76] | Hydatid disease | Serum ELISA | Recognize hydatid cyst in liver and other organs | IHA, immunoelectrophoresis, immunoblotting, EIA | Albendazole or mebendazole |
| **Entamoeba Histolytica** [77,78] | Amoebiasis brain abscess | Serum ELISA | Recognize trophozoite or cyst in feces, or liver | EIA, IHA, latex agglutination, immunoelectrophoresis, CIE and PCR | Metronidazole or tinidazole plus luminal agent (idoquinoxalin, paromomycin, diloxanidefuroate) |

EITB: enzyme-linked immunotransferblot assay, ELISA: enzyme-linked immunosorbent assay, IHA: indirect hemagglutination assay, ISAGA: immunosorbent agglutination assay, IIF: indirect immunofluorescence, IFA: Immunofluorescent antibody test, EIA: enzyme immunoassay, CIE: counterimmunoelectrophoresis, CNS: central nervous system, CSF: cerebrospinal fluid

*Active lesion: vesicular stage, colloidal vesicular stage and granular nodular stage  
**Inactive lesion: nodular calcified stage

Citation: Assavapongpaiboon B, Salehian B (2017) Intrasellar Parasitic Infection: Finding the Culprit - How to Get the Right Diagnosis. Int J Clin Endocrinol Metab 3(1): 008-015.
with liver abscess. Enzyme–linked immunosorbent assays or agar gel diffusion assays positive result with appropriate clinical syndrome suggests active disease. Sensitivity of enzyme immunoassays (EIA) for \textit{E. histolytica} is approximately 95% in patients with extraintestinal amebiasis, 70% in patients with active intestinal infections, and 10% in patients with asymptomatic intestinal \textit{E. histolytica} infections [68]. Interpretation of indirect hemaggulination test should be aware because it may remain positive for as long as 10 years. In addition, PCR of brain abscess aspirate or CSF is adjunctive for diagnosis [66]. The definitive diagnosis of amebic colitis is made by the demonstration of \textit{E.histolytica} trophozoites ingesting red blood cell. However, most cases have not been microscopically observed in cerebral amebiasis. At least three fresh stool specimens should be collected because trophozoites are killed rapidly by water, drying, or barium. For circumstances in which amebic brain abscess is suspected based on epidemiology, clinical manifestations, and radiographic findings, treatment (metronidazole) should be started immediately. Surgical intervention for decompression and tissue biopsy may be required.

**Conclusion**

History of risk for exposure and clinical manifestations of headache, visual disturbance, endocrine disturbance and eosinophilia are initial clues that lead to diagnosis of parasitic pituitary infection. Further investigation should include visual evaluation, hormonal evaluation for hypopituitarism and neuroimaging. Attention should be given to correcting endocrine deficiencies in particular adrenal insufficiency and diabetes insipidus which is more prevalent in cases of severe malaria. Although identification of the causative parasite is difficult, immunodiagnostics and molecular diagnosis can be applied to achieve the correct diagnosis. Neuroimaging plays an important role to differentiate diagnosis, while serological and molecular tests can confirm the diagnosis. Treatment of these parasitic diseases is complex, involving surgery, and different modalities based on each parasitic organism. In addition, drug resistance is reported worldwide. In Table 2, we summarized the parasitological and serological methods for the diagnosis of associated parasitic infection and commonly used drugs for treatment of these infections in non tropical area help of tropical medicine and infectious–disease specialist is crucial for correct diagnosis and management.

**References**

1. Zhang X, Huang G, Ji T, Liu W, Gao Y, et al. (2014) Sellar cysticercosis and septum pellucidum cyst: a case report and review of the literature. Southeast Asian J Trop Med Public Health 45: 584-587. Link: https://goo.gl/HEvQ9t

2. Dutta D, Kumar M, Ghosh S, Mukhopadhyay S, Chowdhury S, et al. (2013) Pituitary hormone deficiency due to racemose neurocysticercosis. Lancet Diabetes Endocrinol 1: e13. Link: https://goo.gl/023mU

3. Ramirez Mdel P, Restrepo JE, Syro LV, Fabio Rotondo, Francisco Londoño J, et al. (2012) Neurocysticercosis, meningioma, and silent corticotroph pituitary adenoma in a 61-year-old woman. Case Rep Pathol 2012: 340840. Link: https://goo.gl/0Ypso5

4. Keleidis T, Tsiodras S (2011) Hypopituitarism caused by neurocysticercosis. Am J Med Sci 341: 414-416. Link: https://goo.gl/HIGB9h

5. Cheong JH, Kim JM, Kim CH (2010) Neurocysticercosis involving the pituitary stalk: case report and literature review. J Korean Neurosurg Soc 48: 91-93. Link: https://goo.gl/i4hHfk

6. Ruangkanchanasetri P, Bangchua T, Sithinamsuwan P, Yupin Benjasuratwong, Warun Chuankerkul, et al. (2006) Hypothalamic neurocysticercosis presenting with polyuria: a first report of an unusual manifestation. Nephrol Dial Transplant 21: 2308-2310. Link: https://goo.gl/Hu2RLV

7. Arraida-Mendica N, Celis-Lopez MA, Higuera-Calleja J, Corona-Vazquez T (2003) Imaging features of sellar cysticercosis. AJNR Am J Neuroradiol 24(4): 1386-1389. Link: https://goo.gl/TwFmNC

8. (1993) Case reports of the Massachusetts General Hospital. Weekly clinicopathological exercises. Case 8-1993, A 62-year-old Cape Verdean woman with blured vision, diplopia, a suprasellar mass, and lymphocytic meningitis. N Engl J Med 326: 566-573. Link: https://goo.gl/4PMN5D

9. Boecher-Schwarz HG, Hey O, Higer HP, Perneczky A (1991) Intrasellar cysticercosis mimicking a pituitary adenoma. Br J Neurosurg 5: 405-407. Link: https://goo.gl/SGLjMB

10. Del Brutto OH, Guevara J, Sotelo J (1988) Intrasellar cysticercosis. J Neurosurg 69: 58-60. Link: https://goo.gl/5UekYG

11. Rafael H, Gomez-Llata S (1988) Intrasellar cysticercosis. Case report. J Neurosurg 63: 975-976. Link: https://goo.gl/CTV7NK

12. Prosser PR, Wilson CB, Forsham PH (1978) Intrasellar cysticercosis presenting as a pituitary tumor: successful transsphenoidal cystectomy with preservation of pituitary function. Am J Trop Med Hyg 27: 976-978. Link: https://goo.gl/1tVH921

13. Berkmann S, Fischer I, Sonderregger B, Fischli S, Fandino J, et al. (2015) Sellar Toxoplasmosis and Nonfunctioning Pituitary Adenoma. World Neurosurg 84(1491-1494). Link: https://goo.gl/udKnn

14. Mohamed S, Osman A, Jurayyan NA AI, Nemri A AI, Salih MA, et al. (2014) Congenital toxoplasmosis presenting as central diabetes insipidus in an infant: a case report. BMC Res Notes 7: 184. Link: https://goo.gl/EjRjRr

15. Siahonidou T, Tsoumas D, Kanaka-Gantenbein C, Mandyla H (2006) Neuroendocrine abnormalities in a neonate with congenital toxoplasmosis. J Pediatr Endocrinol Metab 19: 1363-1366. Link: https://goo.gl/ni20Ac

16. Zhang X, Li Q, Hu P, Cheng H, Huang G (2002) Two case reports of pituitary stalk : case report and literature review. J Korean Neurosurg Soc 48: 91-93. Link: https://goo.gl/i4hHfk

17. Oygur N, Yilmaz G, Ozkaynak C, Guven AG (1998) Central diabetes insipidus in a patient with malaria tropica. J Endocrinol Invest 21: 2308-2310. Link: https://goo.gl/9dmJ8

18. Massa G, Vanderschueren-Lodeweyckx M, Van Vliet G, Craen M, de Zeger F, et al. (1989) Hypothalamo-pituitary dysfunction in congenital toxoplasmosis. Eur J Pediatr 148: 742-744. Link: https://goo.gl/jbaZK

19. Milligan SA, Katz MS, Craven PC, Strandberg DA, Russell JL, et al. (1984) Toxoplasmosis and Nonfunctioning Pituitary Adenoma. World Neurosurg 84(1491-1494). Link: https://goo.gl/udKnn

20. Premji R, Roorpaarenewisinh N, Cohen J, Sen S (2016) Cerebral Malaria: An Unusual Cause of Central Diabetes Insipidus. Case Rep Endocrinol 2016:2047410. Link: https://goo.gl/9dMj8

21. Schubert S, Achenbach H, Engelmann L, Borte G, Stumvoll M, et al. (2006) Central diabetes insipidus in a patient with malaria tropica. J Endocrinol Invest 29: 265-266. Link: https://goo.gl/gkG9CU

22. Becker GL, Jr, Keph S, Lance KP, Kaufman L (1980) Amebic abscess of the brain. Neurosurgery 6: 192-194. Link: https://goo.gl/5LLzZ0
60. McManus DP, Zhang W, Li J, Bartley PB (2003) Echinococcosis. Lancet 362: 1295-1304. Link: https://goo.gl/mczl6t

61. Riganò R, Profumo E, Ioppolo, S, Notargiacomo, E, Ortona, et al. (1995) Immunological markers indicating the effectiveness of pharmacological treatment in human hydatid disease. Clin Exp Immunol 102: 281-285. Link: https://goo.gl/wYALSk

62. Elena Ortona, Rachele Riganò, Brigitta Buttari, Federica Delunardoa, Salvatore Ioppolo, et al. (2003) An update on immunodiagnosis of cystic echinococcosis. Acta tropica 85: 165-171. Link: https://goo.gl/W9ihax

63. Acuna-Soto R, Maguire JH, Wirth DF (2000) Gender distribution in asymptomatic and invasive amebiasis. Am J Gastroenterol 95: 1277-1283. Link: https://goo.gl/bsB8NF

64. Aucott JN, Ravdin JT (1993) Amebiasis and “nonpathogenic” intestinal protozoa. Infect Dis Clin North Am 7: 467-485. Link: https://goo.gl/0ZK6s2

65. Petri WA, Haque R (2013) Entamoeba histolytica brain abscess. Handb Clin Neurol 114: 147-152. Link: https://goo.gl/v1p5ZI

66. Maldonado-Barrera CA, Campos-Esparza Mdel R, Muñoz-Fernández L, Victoria-Hernández JA, Campos-Rodríguez R, et al. (2012) Clinical case of cerebral amebiasis caused by E. histolytica. Parasitol Res 110: 1291-1296. Link: https://goo.gl/ffs78e

67. Wilson M, Schantz PM, Nutman T (2006) Molecular and immunological approaches to the diagnosis of parasitic infection. In: Detrick B, Hamilton RG, Folds JD, eds. Manual of Molecular and Clinical Laboratory Immunology. Washington, DC: American Society for Microbiology 557–568.

68. Fogang YF, Savadogo AA, Camara M, Toffa DH, Basse A, et al. (2015) Managing neurocysticercosis: challenges and solutions. Int J Gen Med 8: 333-344. Link: https://goo.gl/hcz0UG

69. Garcia HH, Nash TE, Del Brutto OH (2014) Clinical symptoms, diagnosis, and treatment of neurocysticercosis. Lancet Neurol 13: 1202-1215. Link: https://goo.gl/MpTaK

70. Liu Q, Wang ZD, Huang SY, Zhu XQ (2015) Diagnosis of toxoplasmosis and typing of Toxoplasma gondii. Parasit Vectors 8:292. Link: https://goo.gl/4PeUTP

71. Kieffer F, Wallon M (2013) Congenital toxoplasmosis. Handbook of clinical neurology112: 1099-1101. Link: https://goo.gl/YB62QH

72. Dard C, Fricker-Hidalgo H, Brenier-Pichart MP, Pelloux H (2016) Relevance of and New Developments in Serology for Toxoplasmosis. Trends Parasitol 32: 492-506. Link: https://goo.gl/BwZtpq

73. Sahu PK, Satpathi S, Behera PK, Mishra SK, Mohanty S, Wassmer SC (2015) Pathogenesis of cerebral malaria: new diagnostic tools, biomarkers, and therapeutic approaches. Front Cell Infect Microbiol 5: 75. Link: https://goo.gl/GELyi

74. Agudelo Higuita Nl, Brunetti E, McCloskey C (2016) Cystic Echinococcosis. J Clin Microbiol 54: 518-523. Link: https://goo.gl/pSf96S

75. Manzano-Roman R, Sanchez-Ovejero C, Hernandez-Gonzalez A, Casulli A, Siles-Lucas M (2015) Serological Diagnosis and Follow-Up of Human Cystic Echinococcosis: A New Hope for the Future? Biomed Res Int 2015: 428205. Link: https://goo.gl/rpgpN6

76. Choudhuri G, Rangan M (2012) Amebic infection in humans. Indian J Gastroenterol 31: 153-162. Link: https://goo.gl/aeGRvm

77. Wilkins PP (2013) Immunodiagnosis of CNS parasitic infections. Handb Clin Neurol 114: 23-36. Link: https://goo.gl/hwUbsP

Copyright: © 2017 Assavapongpaiboon B, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Assavapongpaiboon B, Salehian B (2017) Intrasellar Parasitic Infection: Finding the Culprit - How to Get the Right Diagnosis. Int J Clin Endocrinol Metab 3(1): 008-015.