Spontaneous primary hypothyroidism in 7 adult cats

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Background: Naturally occurring hypothyroidism in adult cats is rare, with only 4 cases reported.

Objectives: To describe the historical, clinical, laboratory, and scintigraphic features of adult cats with spontaneous hypothyroidism.

Animals: Seven adult cats referred for suspected hypothyroidism.

Methods: Prospective case series. We collected data on cats’ signalment, clinical signs, results of physical examination, routine laboratory and thyroid hormone testing, and thyroid imaging (thyroid scintigraphy or ultrasound). We subsequently treated cats with levothyroxine and evaluated their response to treatment.

Results: Cats ranged from 3.5 to 11 years, with no apparent breed predilection; 6/7 cats were male. Only 2/7 cats were initially tested because of signs of hypothyroidism (hair-coat changes, lethargy, obesity); others were tested for routine thyroid monitoring or palpable thyroid nodules. Four were azotemic (serum creatinine, 2.2-3.4 mg/dL). Six of the 7 cats had low serum thyroxine (T4) and free T4 (fT4) concentrations, whereas all 7 cats had high thyroid-stimulating hormone (TSH) concentrations. In 6/7 cats, thyroid scintigraphy revealed bilateral goiter with intense radionuclide uptake; imaging showed no visible thyroid tissue in the other. After levothyroxine treatment, serum concentrations of T4 and fT4 increased and TSH fell; high serum creatinine normalized in azotemic cats; and repeat imaging showed reduction in goiter size.

Conclusions and Clinical Importance: Primary hypothyroidism develops in adult cats, with a higher prevalence than previously thought. Most cats appear to develop a goitrous form of hypothyroidism associated with thyroid hyperplasia, whereas thyroid atrophy appears to be less common. With levothyroxine replacement, clinical and laboratory abnormalities improve or resolve.

KEYWORDS
atrophic, feline, goiter, goitrous, thyroid gland, thyroxine, TSH

1 | INTRODUCTION

Spontaneous primary hypothyroidism appears to be an extremely rare clinical disorder in adult cats, with only 4 reported cases over the last 25 years.1–4 Most hypothyroid cats are younger (generally kittens, aged 2–4 months) and suffer from congenital hypothyroidism, which typically results in disproportionate dwarfism (cretinism). Of ~60 cats with congenital hypothyroidism that have been reported,5–9 only 3 were older than 12 months of age at time of diagnosis, but these 3 cats were stunted since kittenhood, consistent with the diagnosis of congenital hypothyroidism.

In dogs and humans, almost all naturally occurring hypothyroidism is attributable to irreversible destruction of the thyroid gland. Histologically, primary hypothyroidism in humans and dogs presents as either lymphocytic thyroiditis or idiopathic thyroid degeneration (idiopathic follicular atrophy).10–12 Of the 4 reported adult cats with naturally occurring primary hypothyroidism, 2 had lymphocytic thyroiditis,1,4 1 had idiopathic atrophy,2 and 1 had a goitrous form of hypothyroidism...
associated with diffuse thyroid follicular hyperplasia of both thyroid lobes. Goitrous hypothyroidism is extremely rare in dogs, with iatrogenic, drug-induced thyroid goiter being the most common cause. However, a subset of adult hypothyroid humans have goiter secondary to iodine deficiency, environmental goitrogens, or ineffective thyroid hormone synthesis caused by biosynthetic defect (dyshormonogenesis).

Given the paucity of data regarding adult-onset feline hypothyroidism, we sought to describe the history, clinical features (including presence or absence of goiter), diagnostic testing, treatment, and long-term outcome of 7 adult cats with spontaneous primary hypothyroidism. For these cats, we used the serum concentrations of T4, free T4 (fT4), and thyroid stimulating hormone (TSH), and results of thyroid scintigraphy to aid in both the diagnosis and long-term monitoring of thyroid hormone replacement treatment.

2 | MATERIALS AND METHODS

2.1 | Case selection and data collection

This prospective case series included 7 adult cats with spontaneous hypothyroidism referred to the Animal Endocrine Clinic for evaluation over a 3.5-year period from March 2014 to September 2017 and then followed until April 2018. Data collected for each cat included the following: age, breed, sex, reason for initial workup by referring veterinarian, clinical signs as reported by owners, dietary history, drugs and supplements administered, known concurrent illnesses, physical examination findings (including the presence or absence of thyroid goiter), routine laboratory findings (eg, complete blood count, serum biochemistry profile, symmetric dimethylarginine [SDMA] concentration), complete thyroid panel (T4, fT4, and TSH concentrations), survey radiographs of the spine and limbs, and results of thyroid imaging studies (eg, thyroid scintigraphy, or ultrasound).

After collection of pretreatment data, all 7 cats were treated with levothyroxine (L-T4), using an initial dosage of 100–150 μg/cat/day. Dose adjustments were titrated (by a 50 μg/cat/day increase or decrease as needed) based on follow-up serum T4 and TSH concentrations, with the aim of restoring serum thyroid values to within reference intervals. Five cats with goitrous hypothyroidism also had follow-up thyroid imaging (4 using scintigraphy and 1 with ultrasonography) to evaluate changes in thyroid function and volume after L-T4 treatment.

Two of the 7 cats had thyroid aspirates collected for cytologic evaluation before any treatment, whereas a 3rd cat with persistent bilateral goiter despite treatment with L-T4 underwent excisional biopsy of the left lobe of the thyroid gland for histopathologic examination.

2.2 | Thyroid imaging

All cats underwent thyroid scintigraphy by injecting 111 MBq of sodium 99mTc-pertechnetate (99mTcO4- IV) and imaging 1 hour later, as previously described. Thyroid activity was quantified by calculation of both the thyroid-to-salivary gland ratio (T/S) and the percent thyroidal uptake of the injected 99mTcO4 (TcTU). The estimated thyroid volume was also calculated from the scintigraphic image, using the equation for a spheroid as previously described. In 1 cat, serial thyroid ultrasound imaging was used to estimate changes in goiter size by measuring the length and width of each lobe and then again calculating thyroid volume using the equation for a spheroid.

2.3 | Assays for serum thyroid hormone and thyrotropin (TSH) Concentrations

Serum total T4, fT4, and TSH concentrations were measured by assays (DRI Thyroxine [T4] assay, Microgenics Corporation, Freemont, California; Free T4 - by Equilibrium Dialysis, Antech Diagnostics, Irvine, California; Immulite Canine TSH, Siemens Healthcare Diagnostics Product, Tarrytown, New York) validated for use in cats, as previous described. The reference intervals for these serum thyroid tests, previously established in clinically normal cats were as follows: total T4 = 0.9-3.9 μg/dL; fT4 = 10-51 pmol/L; and cTSH = <0.03-0.3 ng/mL.

2.4 | Data and statistical analyses

All statistical analyses were performed by proprietary statistical software (GraphPad Prism, version 7.0; GraphPad Software, La Jolla, California). Most data are presented with descriptive and summary statistics (median, range, proportions). For select comparisons, such as reduction of goiter size after thyroid hormone treatment, a paired T test was used to compare the measured thyroid volumes before and after L-T4 replacement. Similarly, before and after comparisons of serum T4, fT4, TSH, and creatinine concentrations in the hypothyroid cats were also analyzed by the paired T test. For all analyses, statistical significance was defined as P < .05.

3 | RESULTS

3.1 | Signalment of hypothyroid cats

The 7 hypothyroid cats ranged in age from 3.5 to 11.0 years (median, 7 years). When age was classified according to the cats’ life stage, 2 were in prime (3–6 years), 4 were in mature (7–10 years), and 1 was in the senior life stage (11–14 years). Breeds included domestic short-hair in 6 cats and American shorthair in 1. Six of the 7 hypothyroid cats were male and 1 was female; all were castrated or spayed.

3.2 | History, clinical presentation, and physical examination findings

Cats were initially tested for thyroid disease by the referring veterinarians for 1 of 3 reasons: (1) routine annual thyroid monitoring (n = 3); (2) palpable goiter (thyroid nodules) detected on physical examination (n = 2); and (3) clinical signs suggestive of hypothyroidism (n = 2). Of the 2 cats with clinical features suggestive of hypothyroidism (ie, hair loss, lethargy, and obesity), owners had first noted clinical signs 2 and 8 months before examination.

The most common clinical signs reported by owners included polyuria and polydipsia, mild changes in hair coat (unkempt appearance, increased shedding, dandruff, or hair thinning), and weight gain...
Two of the owners reported no overt clinical signs for their cats. Diet fed to the 7 cats consisted of a variety of both moist and dry commercial cat foods to 5, moist only to 1, and dry only to 1 cat. None of the cats were being administered any drugs or supplements associated with changes in thyroid function or goiter (eg, sulfonamide antibiotics or iodine/iodide).

On physical examination, symmetric enlargement of both thyroid lobes (goiter) could be palpated in 6 of the 7 cats. The median weight was 5.3 kg (range 3.6–7.3 kg). Three of the 7 cats weighed >6.0 kg and were considered obese (body condition scores, 6/9 to 7/9), whereas the body condition in the remaining 4 cats was considered ideal. All 7 cats had normal muscle condition. Sinus bradycardia (heart rate, 120 bpm) was auscultated in 1 cat. Relatively mild changes in hair coat (unkempt appearance, increased shedding, or hair thinning), were identified in 4 cats (Table 1). None of these cats had overt or complete areas of alopecia.

### 3.3 Routine laboratory findings

The most frequent abnormality on routine laboratory testing was azotemia (defined as serum creatinine concentrations >2.0 mg/dL; Table 1). Serum creatinine concentrations in the 4 azotemic cats ranged from 2.2 to 3.4 mg/dL; concomitant urine specific gravities were less-than-maximally concentrated <1.035 in 2 cats (1.015 and 1.034), and well concentrated in 2 (1.047 and 1.050). Only 1 of the 4 azotemic cats had a slightly high serum SDMA concentration (15 μg/dL); the other 3 had SDMA values (11, 12, and 13 μg/dL) that were in the upper tertile of the reference interval (≤14 μg/dL). In the 3 nonazotemic cats, serum creatinine concentrations were in upper half of the reference interval (1.5, 1.7, and 1.9 mg/dL). Two cats were mildly anemic, as reflected by a low RBC count, hemoglobin concentration, or hematocrit (Table 1).

### 3.4 Serum thyroid hormone and TSH concentrations

Serum T4 and fT4 concentrations were low in 6 of the 7 cats (Table 1; Figure 1A,B). One cat had low-normal concentrations of both T4 and fT4 (within the lower tertile of the reference interval). Serum TSH concentrations were high in all cats (Figure 1C), ranging from 2.1 to 20 ng/mL (median, 10.1 ng/mL).

### TABLE 1

Clinical features, routine laboratory findings, and serum thyroid hormone concentrations in 7 adult cats with naturally occurring primary hypothyroidism

| Historical, owner-reported signs |   |
|--------------------------------|---|
| Polyuria and polydipsia        | 4 |
| Hair coat changes              | 4 |
| Weight gain                    | 4 |
| Lethargy/mental dullness       | 3 |
| Poor appetite                   | 1 |
| Cold intolerance                | 1 |

| Physical examination findings  |   |
|--------------------------------|---|
| Palpable goiter                | 6 |
| Unkempt, dull coat             | 3 |
| Hair thinning/ hypotrichosis   | 3 |
| Overweight/obesity             | 3 |
| Dandruff, flaking dry skin     | 2 |
| Bradycardia (≤150 bpm)         | 1 |

| Routine laboratory findings    |   |
|--------------------------------|---|
| Azotemia (serum creatinine >2.0 mg/dL) | 4 |
| Urine specific gravity <1.035      | 2 |
| Anemia                           | 2 |
| Hypercholesterolemia (>300 mg/dL) | 1 |
| High serum creatine kinase       | 1 |
| High serum SDMA                  | 1 |

| Serum thyroid hormone concentrations |   |
|--------------------------------------|---|
| Low serum T4 (<0.9 μg/dL)            | 6 |
| Low serum fT4 (<10 pmol/L)           | 6 |
| High serum TSH (>0.3 ng/mL)          | 7 |

FIGURE 1 Serum concentrations of (A) total T4, (B) fT4, (C) TSH, and (D) creatinine in 7 hypothyroid cats, measured before and after treatment with levothyroxine. Serum T4 and fT4 concentrations increase, while TSH and creatinine concentrations decrease after administration of thyroid replacement treatment. The shaded area represents the reference interval. To convert T4 concentration from μg/dL to nmol/L, multiply by 12.87. To convert fT4 concentration from pmol/L to ng/dL, divide by 12.87.
3.5 | Survey radiographs of the spine and limbs
Ossification centers of the long bones and vertebrae were evaluated with survey radiographs. All 7 cats had complete closure of the physes (ossification centers) of long bones and vertebrae.

3.6 | Pretreatment thyroid imaging
In 6 of the 7 cats, qualitative thyroid scintigraphy revealed bilateral thyroid lobe enlargement (goiter) and intense radionuclide thyroid uptake (Figures 2B and 3). All 6 of these cats with goitrous hypothyroidism had high values for T/S ratio, percent TcTU, and thyroid volume (Figure 4). In the 7th cat, thyroid imaging showed no visible thyroid tissue, neither in the normal cervical nor an ectopic location (Figure 2C). This cat with atrophic hypothyroidism had a low T/S ratio (0.1), percent TcTU (0.02%), and thyroid volume (0.05 g).

3.7 | Thyroid aspiration cytology and biopsy
Before treatment, 2 cats had thyroid aspirates collected for cytology. In both cats, findings were consistent with thyroid hyperplasia, consisting of variably sized sheets and clusters of epithelial/neuroendocrine-appearing cells ranging from round to polygonal in shape, with small to moderate amounts of light to dark blue cytoplasm. The cells were frequently associated with small amounts of pink globular to fibrillar material that was thought to represent colloid. The cells displayed minimal features of atypia and mitotic figures were not observed.

Histopathologic finding of the thyroid lobe excised from 1 cat with persistent goiter despite L-T4 treatment (with normalization of serum TSH concentrations) for 22 months revealed that thyroid parenchyma was characterized by prominent coalescing nests of small follicles, which contained normal to decreased amounts of colloid (Figure 5). These extensive regions were interposed between areas containing larger thyroid follicles with abundant colloid lined by low cuboidal epithelial cells. The extensive regions of hypercellularity, small thyroid follicles, and diminished colloid were considered consistent with follicular hyperplasia of the thyroid gland.

3.8 | Treatment with levothyroxine and outcome
After treatment with L-T4 (median, 200 μg/cat/day [32.7 μg/kg/day]; range 100–200 μg/cat/day [16.7–55.6 μg/kg/day]) for 3 to 7 months (median time, 150 days), serum concentrations of T4 and fT4 both increased and TSH decreased, generally into the respective reference intervals (Figures 1A-C). In 5 treated cats with goitrous hypothyroidism, repeat thyroid imaging demonstrated decreases in goiter size, as reflected by the reduced thyroid volumes (Figures 3 and 6). The intensity of scintigraphic thyroid uptake also fell, evidenced by the decreases in both T/S ratio and percent TcTU (Figure 4).

After L-T4 treatment, serum creatinine concentrations decreased in all 7 cats (Figure 1D). In the 4 cats that were azotemic before L-T4 treatment, serum creatinine concentrations normalized after L-T4 supplementation (Figure 1D). Of the 2 azotemic cats that had urine specific gravity values that less-than-appropriately concentrated (<1.035) before L-T4 treatment, 1 cat remained isosthenuric, whereas the other concentrated appropriately after treatment. Mild anemia also resolved in the 2 affected cats after L-T4 treatment. By 2–4 months after treatment, owners reported that their cat’s lethargy, hair coat changes, and obesity (if present before treatment) were greatly improved or had resolved. All cats are alive, well, and remain non-azotemic on L-T4 treatment at the time of manuscript preparation, with a median treatment time of 739 days (range, 233–1439 days).

4 | DISCUSSION
Our results provide further information about spontaneous adult-onset primary hypothyroidism in cats. Although the condition is rare, ∼2 cats are diagnosed with spontaneous adult-onset hypothyroidism each year at the primary author’s endocrine referral clinic. We suspect
that this represents only "the tip of the iceberg" of cats affected with this syndrome, because of the mild clinical signs displayed by these cats, and the lack of awareness of this condition by first opinion clinicians. Heightened awareness that adult hypothyroidism can develop in cats, together with increased screening for the disorder, will ultimately determine if this condition is indeed rare or more common than currently thought (Supplemental Data S1).

All of our hypothyroid cats were adults, with 4/7 diagnosed in the "mature" life stage (age 7–10 years), 2 in the younger "prime" life stage (3–6 years) and 1 in the older "senior" life stage (11–14 years). Time of first onset of clinical disease was difficult to determine, but historically, none of our cats displayed any signs longer than 10 months before diagnosis. Almost all of our cats (6/7) were of mixed breeding (DSH), suggesting no specific breed predilection. Six of our 7 cats were castrated males; we have no explanation for this possible sex predilection, especially since human patients and dogs with hypothyroidism have a female, rather than a male, sex predilection. Clearly, more cats need to be diagnosed and studied to clarify sex distribution in feline hypothyroidism.

In contrast to previous reports, most cats in our case series had goitrous hypothyroidism, with only 1 having idiopathic thyroid atrophy. However, this preponderance of goitrous hypothyroidism in our cats could reflect the fact that one-third of cats were referred primarily for evaluation of palpable bilateral thyroid nodules (together with unexpectedly low serum T4 concentrations). Cats without a palpable goiter might not have been as readily referred for thyroid evaluation with scintigraphy at our clinic, or screened for thyroid function on routine evaluation. A larger number of hypothyroid cats need to be diagnosed and evaluated with thyroid imaging or thyroid pathology (or both) to better determine the prevalence of atrophic versus goitrous hypothyroidism in cats.

Most of our cats had only mild clinical features of hypothyroidism and were initially tested for thyroid disease either because of routine thyroid monitoring (eg, annual screening for hyperthyroidism) or a palpable bilateral goiter detected on routine physical examination. Hypothyroidism was initially suspected in only 2 of our cats, both of which had hair thinning, lethargy, and obesity. Even when hair loss or other cutaneous changes were noted, these signs were relatively mild, with no cat having overt or complete areas of alopecia. The most common clinical signs were polydipsia and polyuria, associated with mild-to-moderate azotemia in 4 of our 7 cats (see below). This was followed by weight gain and obesity, signs that are also common in euthyroid, middle-aged adult cats. Because of the lack of overt clinical signs in many cats and the presence of only vague and nonspecific signs in
hypothyroidism can easily be missed and may be misdiagnosed as "normal aging" or concurrent disease (eg, chronic kidney disease [CKD]).

In our 6 cats with goitrous hypothyroidism, the underlying cause of the bilateral thyroid lobe enlargement is not clear. In the 1 cat that had persistent goiter despite long-term L-T₄ treatment, histopathology revealed changes consistent with thyroid hyperplasia, with no evidence of nodular/adenomatous changes, cellular atypia, or capsular invasion. Therefore, these changes are unlike the pathologic findings characteristic of feline adenoma or adenomatous hyperplasia seen in cats with hyperthyroidism, but are similar to those found in the previously reported adult cat with goitrous hypothyroidism. Compared with that cat, the hyperplastic changes in our cat were milder; treating chronically with L-T₄ and normalizing high serum TSH concentrations likely led to partial regression of the thyroid hyperplasia. However, the fact that all 6 cats with goitrous hypothyroidism had bilateral, diffuse thyroid lobe enlargement on physical examination and imaging indicates the presence of intact thyroid tissue, and the massive sizes of these goiters suggest a process that was ongoing for months to years. Acquired causes for goitrous hypothyroidism, such
as iodine deficiency or environmental goitrogens, were considered highly unlikely based on the cats’ history and diet of commercial canned food. In addition, all of our 6 affected cats lived in households with 1 or more other cats, none of which were hypothyroid or had palpable goiter. One goitrous cat lived in the same household with his littermate; evaluation of this unaffected cat revealed completely normal thyroid function, as documented by serum thyroid and TSH testing and qualitative and quantitative thyroid scintigraphy (data not shown).

The hypothesis that best fits our 6 cats with goitrous hypothyroidism is that these cats suffer from a hereditary defect or block in thyroid hormone production (dyshormonogenesis) by an anatomically intact thyroid gland.\(^\text{56}\) This would be similar to the defects reported in kittens with congenital goitrous hypothyroidism, which is most commonly associated with an impaired ability of the thyroid gland to organify iodide (ie, organification or peroxidase defect).\(^\text{7,37–40}\) In congenital, goitrous hypothyroidism, the inability to secrete adequate amounts of T\(_4\) and T\(_3\) leads to the loss of normal negative-feedback inhibition on pituitary thyrotropes, resulting in persistent secretion of excessive amounts of TSH. The unrelenting stimulation of intact thyroid follicular cells by the high circulating concentrations of TSH results in thyroid hyperplasia, enlargement of the intact thyroid, and clinical palpable goiter. In humans, the clinical presentation of patients with dyshormonogenetic goiter depends on the severity of the inborn error in thyroid hormone metabolism. A severe defect will lead to neonatal or congenital hypothyroidism, goiter, mental retardation, and growth abnormalities (cretinism), similar to many of the reported hypothyroid kittens.\(^\text{7,37–40}\) Milder forms of congenital dyshormogenesis can present later in life (ie, adolescence or young adulthood) as goiter and minimal, if any, signs of thyroid dysfunction.\(^\text{18,41–44}\) Such milder or partial defects in thyroid hormone production could explain the pathophysiology in our 6 goitrous cats, in which goiter was one of the major reasons for work-up and few overt clinical signs of thyroid disease were present. If this is the case, our cats were able to compensate for the block in thyroid hormone secretion for many months to years, as evidenced by the cats’ normal rate of growth, body size, and closure of bone physes (ie, none of our adult cats were dwarfed or had open bone plates, both characteristic finding in kittens with congenital hypothyroidism).\(^\text{5–7}\) In addition, none of the 6 cats with goitrous hypothyroidism displayed signs of mental dullness, as reported in younger cats with untreated congenital hypothyroidism.\(^\text{5–7}\) Because severe hypothyroidism in early life leads to lack of bone growth and neurodevelopment (cretinism), moderate to severe intellectual deficits are common, at least in people.\(^\text{45–47}\)

Cats in our study underwent thyroid scintigraphy to confirm the diagnosis and characterize the disease.\(^\text{2–7,48}\) Hypothyroid cats with thyroid atrophy have no detectable thyroid tissue on scintigraphy, and minimal or absent thyroid uptake of the injected radionuclide (\(^{99m}\)TcO\(_4\)^–), as seen in the 1 cat with atrophic hypothyroidism (Figure 2C).\(^\text{2,48}\) In contrast, high circulating TSH concentrations greatly increase \(^{99m}\)TcO\(_4\)^– uptake into the anatomically intact thyroid gland in untreated cats suffering from goitrous hypothyroidism,\(^\text{48,49}\) as evidenced in the 6 cats by the intense (‘hot’) uptake of \(^{99m}\)TcO\(_4\)^– into both thyroid lobes (Figures 2B and 3) and the high calculated values for both the T/S ratio and percent TcTU (Figure 4). The cats with goitrous hypothyroidism also had a marked increase in their measured thyroid volumes (2.5–8 fold higher than that of the upper reference interval).

Our results indicate that measurement of serum TSH concentration is the most sensitive diagnostic test for naturally occurring hypothyroidism in cats, similar to our previous findings in cats with iatrogenic (\(^{131}\)I-induced) hypothyroidism.\(^\text{27,50}\) All 7 hypothyroid cats in the current study had extremely high serum TSH concentrations (7–40 times higher than the upper reference interval). Serum concentrations of T\(_4\) and fT\(_4\) were low in 6/7 cats, diagnostic for overt hypothyroidism. One cat maintained normal serum concentrations of T\(_4\) and fT\(_4\) despite high TSH values, diagnostic for subclinical (mild) hypothyroidism,\(^\text{51–54}\) a pattern similar to that of \(^{131}\)I-treated cats that develop iatrogenic hypothyroidism.\(^\text{27,50}\) In addition, recent prevalence studies of human hypothyroidism reveal that most people (up to 95% in some reports) have subclinical hypothyroidism, with overt hypothyroidism being less common.\(^\text{55–57}\) Additionally, serum TSH concentration is the most specific diagnostic test for hypothyroidism: low serum T\(_4\) and fT\(_4\) concentrations commonly develop in cats with nonthyroidal illness,\(^\text{50,58}\) but high values for TSH have not been reported in these sick, euthyroid cats.\(^\text{50,59}\) However, some human patients with nonthyroidal illness will develop slightly increased serum TSH concentrations,\(^\text{52–64}\) especially during the recovery phase of their illness.\(^\text{60–61}\) In addition, human patients treated with some medications (eg, metoclopramide, amiodarone) can develop slightly increased serum TSH concentrations.\(^\text{52–64}\) Similarly, transiently high serum TSH concentrations can develop in dogs with nonthyroidal disease (with hypoadrenocorticism being the best characterized example).\(^\text{65–68}\) as well as during treatment with certain drugs.\(^\text{53,14,69,70}\) Therefore, it is clear that more studies evaluating serum TSH concentrations in a larger population of cats are needed—especially in cats with nonthyroidal illness during the recovery phase of illness, as well as after treatment with medications known to increase serum TSH in other species.

After levothyroxine supplementation to our hypothyroid cats, high serum TSH concentrations decreased to within the reference interval, as the negative feedback effects of higher circulating T\(_4\) and T\(_3\) concentrations suppressed excessive TSH secretion.\(^\text{5,50}\) In our cats, we used serum TSH concentrations for long-term monitoring, with daily L-T\(_4\) doses titrated to maintain TSH concentrations within its reference range (<0.3 ng/mL) while maintaining serum T\(_4\) and fT\(_4\) concentrations within their respective reference intervals. With that regime, our cats needed a final daily L-T\(_4\) dose ranging from 100 to 200 μg/cat/day (16.7–55.6 μg/kg/day). Our median dose administered per kg (32.7 μg/kg/day) was higher than that generally reported for thyroid hormone replacement in dogs (≤20 μg/kg/day).\(^\text{71–73}\)

Treatment with levothyroxine improved or resolved clinical signs, when present, in all of our hypothyroid cats. Thyroid hormone replacement also led to shrinkage of the bilateral goiter; this might be expected after lowering high serum TSH concentrations to within the reference range, especially if TSH-induced, diffuse thyroid hyperplasia was responsible for the goiter.\(^\text{7}\) As circulating TSH concentrations fell on levothyroxine replacement, scintigraphic evidence of increased thyroid uptake, such as the intensity of \(^{99m}\)TcO\(_4\)^– accumulation and
calculated values for T/S and percent TcTU, also decreased in our goitrous hypothyroid cats (Figure 4). In 1 of our goitrous cats, both the thyroid gland size and uptake values normalized after L-T4 replacement (Figure 3E,F). However, thyroid gland size and radionuclide uptake failed to completely normalize in all of our goitrous cats despite long-term, adequate thyroid hormone replacement, similar to findings reported in younger cats with congenital hypothyroidism, which as well as humans with dysmorphogenic goiter.54

One of the major indications for workup in our cats was the finding of high serum creatinine concentrations, consistent with IRIS Stage 2 to Stage 3 CKD in 4/7 cats. Azotemia is a well-recognized complication of hypothyroidism in humans, developing in over half of patients in some reports. The pathophysiology of hypothyroid-induced azotemia is complex but is related in large part to decreases in cardiac output, renal blood flow, and glomerular filtration rate.84,85 Unless patients have concurrent, primary kidney disease, azotemia is reversible with adequate thyroid hormone replacement treatment, leading to a consistent fall in serum creatinine concentrations.76–85 Cats in our study demonstrated similar decreases in serum creatinine concentrations after treatment, with serum creatinine normalizing (falling back into within the reference interval) in the 4 azotemic cats. Like the hypothyroid cats of this case series, cats with iatrogenic (131I-induced) hypothyroidism commonly become azotemic after treatment, and serum creatinine decreases or normalizes in many of those cats after thyroid hormone replacement.21,50 Overall, the fact that the azotemia can resolve in hypothyroid cats after thyroid hormone replacement implies that these cats likely did not have CKD or that it was not as severe as the high serum creatinine concentration might imply.

Hypothyroidism can also impair urine concentrating ability, an effect that can also be reversed with thyroid hormone treatment.84–87 Cats in our study mostly concentrated their urine (urine specific gravities > 1.035), but 2 of the azotemic cats had values that were less-than-appropriately concentrated. After L-T4 treatment, 1 cat remained isosthenuric, but the other began to concentrate appropriately. This impaired urine concentrating ability likely contributed to the polyuria and polydipsia reported in 4 of our hypothyroid cats, since these signs improved (2 cats) or resolved (2 cats) after thyroid hormone replacement.

This reversible form of azotemia in hypothyroid cats suggests that clinicians should include hypothyroidism as a differential for all adult cats that develop azotemia. This is especially true in relatively young or middle-aged cats that lack other evidence for CKD, such as low urine specific gravity, high serum SDMA concentrations, or small kidney size. These cats should have serum T4 and TSH concentrations measured; finding a low to low-normal serum T4 concentration (ie, in the lower third of the reference interval; generally <1.5 μg/dL) together with a clearly high serum TSH concentration (>1.0 ng/mL) is diagnostic for hypothyroidism and confirms the need for thyroid hormone treatment. Similarly, given the subtlety of clinical signs in cats with spontaneous adult-onset hypothyroidism, clinicians should consider measuring serum TSH concentration as part of routine thyroid monitoring in adult cats, especially in those that have a palpable thyroid nodule(s) but do not show classical clinical features of hyperthyroidism. Finding an undetectable TSH concentration with high-normal to high serum T4 concentrations is consistent with hyperthyroidism and certainly excludes hypothyroidism; a high serum TSH concentration with low-normal T4 concentrations is diagnostic for hypothyroidism and justifies the need for thyroid imaging, L-T4 treatment, or both.

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CONFLICT OF INTEREST DECLARATION

Authors declare no conflict of interest.

OFF-LABEL ANTIMICROBIAL DECLARATION

Authors declare no off-label use of antimicrobials.

INSTITUTIONAL ANIMAL CARE AND USE COMMITTEE (IACUC) OR OTHER APPROVAL DECLARATION

Authors declare no IACUC or other approval was needed.

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Additional supporting information may be found online in the Supporting Information section at the end of the article.

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