Natural Melanogenesis Stimulator a Potential Tool for the Treatment of Hypopigmentation Disease

Abstract

The synthesis of melanin pigments or melanogenesis has many important physiological functions that include photo protection of the human skin from ultraviolet (UV) irradiation. Melanogenesis is a complex pathway involving melanin synthesis, melanin transport, and melanososome release. Melanin synthesis is stimulated by various effects such as α-melanocyte-stimulating hormone (α-MSH); cyclic AMP (cAMP) elevating agents including palmitic acid, methyl ester, psoralen, thyroquinone, pipерine, berberine and withaferin-A. Loss of the hair shaft melanin is associated with decrease of tyrosinase activity in the bulb of melanocytes. Activators of tyrosinase with stimulatory effects on melanogenesis are beneficial for the treatment of hypopigmentation diseases. This is only report authors found in literature about tyrosinase activation. Melanogenesis stimulator are rarely studied as most industries are more interested on inhibitors of the enzyme in order to reduce the adverse effects of melanin formation in, for example, processed food and human skin. Hence, there is a need to review the successful utilization of tyrosinase stimulators from natural herbs for medicinal applications.

Keywords: Tyrosinase; Herbs; Vitiligo; Melanin; Melanocytes

Introduction

Vitiligo is one of the most well-known conditions of skin depigmentation. It is estimated to occur in up to 2% of the world’s population. A number of people, these patches can prove all over the body. It is an autoimmune disease in which the pigment-inducing cells are injured. There is no therapy for vitiligo, however there are few therapies, including ultraviolet light, cosmetic cover-ups or corticosteroid creams medications [2]. Melanin synthesis is regulated by melanocyte specific enzymes and related transcription factors, tyrosinase, TRP-1, TRP-2 and MITF [1]. Natural herbal extracts have powerful phytochemical properties which are now being exploited world over and there is a sudden surge in Ayurvedic or traditional uses of plant wealth in treatment of diseases like cancers, arthritis, sterility, psoriasis and diabetes. For the treatment of hypopigmentation or vitiligo, Kyoko et al. [4] stated whether Tunisian aromatic plants can induce melanogenesis in aromatic plant extracts; found that melanogenesis in the cultured mouse B16 melanoma cell line was enhanced by Tunisian aromatic plants. The cells cultured with and without Tunisian plant extracts showed no effect on cell growth and shape. This denotes that Tunisian aromatic plants can induce melanogenesis in B16 melanoma cells without causing transformation. Later on Jeon [5] studied the essential oil from lotus flower extract and its effects on melanogenesis in human melanocytes. It was found that the effective compound of lotus flower oil palmitic acid methyl ester induced the expression of tyrosinase, microphthalmia- associated transcription factor M

Abbreviations: α-MSH: α-melanocyte-stimulating hormone; cAMP: Cyclic Adenosine Monophosphate; MITF: Microphthalmia-Associated Transcription Factor; TRP1: Tyrosinase Related Protein1; TRP2: Tyrosinase Related Protein2; CREB: Responsive Element-Binding Protein; mRNA: Messenger Ribonucleic Acid

Effect of herbal extract and their actives on melanogenesis

Herbal extracts most widely used medicinal plants in traditional oriental medicine. Over thousands of years, it has been used to improve the overall condition of skin, as well as to treat a wide variety of diseases. Natural plant extracts have powerful properties which are now being exploited world over and there is a sudden surge in Ayurvedic or traditional uses of plant wealth in treatment of diseases like cancers, arthritis, sterility, psoriasis and diabetes. For the treatment of hypopigmentation or vitiligo, Kyoko et al. [4] stated whether Tunisian aromatic plants can induce melanogenesis in aromatic plant extracts; found that melanogenesis in the cultured mouse B16 melanoma cell line was enhanced by Tunisian aromatic plants. The cells cultured with and without Tunisian plant extracts showed no effect on cell growth and shape. This denotes that Tunisian aromatic plants can induce melanogenesis in B16 melanoma cells without causing transformation. Later on Jeon [5] studied the essential oil from lotus flower extract and its effects on melanogenesis in human melanocytes. It was found that the effective compound of lotus flower oil palmitic acid methyl ester induced the expression of tyrosinase, microphthalmia- associated transcription factor M
addition it induced the tyrosinase activity and altered melb-a cell morphology. Trans well migration assay showed the potential of this herbal candidate to induce direct migration of treated cells. The findings were significant in designing preclinical and clinical studies on the efficacy of C. occidentalis as a stimulant for skin regeneration in vitiligo [10]. Moreira et al. [11] investigated the melanogenic activity of hydro alcoholic extracts from the leaves and flowers of P. venusta on murine B16F10 melanoma cells. Both extracts, leaves increased the melanin content in a dose dependent manner on melanoma cells. Leaves extract promoted enhancement of melanogenesis with maximum effect of (3 μg/mL), and the flower extract increased in (0.1 μg/mL). The cell viability tested concentrations of both extracts no cell death was detected. Actually, either extract was not able to cause any change in the tyrosinase activity. Their findings support the folk medicinal use of P. venusta on the treatment of hypopigmentation diseases, such as vitiligo Table 1.

Later on Ali & Meitei, [12] studied the effects of the root extract of Withania somnifera and its active ingredient Withaferin A on the isolated melanophores of the wall lizard H. flaviviridis were studied in order to establish the mechanism of skin darkening at the cellular level. Significant skin darkening activity of the extract of W. somnifera and Withaferin A was observed on the isolated melanophores of the wall lizard. The melanin dispersal effects leading to the darkening of the skin were antagonized by atropine and hyoscine, and were also found to be highly potentiated by neostigmine. These findings suggested that the extract of W. somnifera, as well as its active principle, mimic the action of acetylcholine in melanin dispersion, thus leading to skin darkening via stimulation of cholinergic receptors of mucaromic nature within the melanophores of the wall lizard. In 2012, Meitei & Ali [13] subsequently studied the effects of fig leaf extract and its bioactive compounds which found to induce skin darkening effect in reptilian melanophores via cholinergic receptor stimulation. They found ethanolic leaf extract of Ficus carica per se can cause melanin stimulatory effects leading to skin darkening. They also found neostigmine an anti cholinesterase agent to potentiate the melanin dispersal effects of both ethanolic leaf extract of F. carica and its active ingredient psoralen. Choudhary et al. [14] investigated effects of extracts of two species of Chlorophytum i.e., Chlorophytum tuberosum & Chlorophytum borivilianum on the isolated scale melanophores of the teleost fish, Channa punctatus. The lyophilized extract of tubers of C. tuberosum had a melanin aggregating effect causing paling of the skin; the action seems to be mediated through alpha adrenergic receptors present dominantly on fish melanophores. The extract of tubers of C. borivilianum had a melanin dispersing effect within the fish melanophores inducing darkening of the skin and the responses seem to be mediated probably through beta adrenergic receptors.

The compound 4’-O-β-d-Glucopyranosyl-quercetin-3-O-β-d-glucopyranosyl-(1→4)-β-d glucopyra-noside from Helminthostachys zeylanica root extract as a melanogenesis acceleration compound and have synthesized using rutin as the starting material. It has been found that isolated compounds were also synthesized to understand the structure-activity relationships in melanin biosynthesis. Melanogenesis activities of the

In vitiligo the active melanocytes in the epidermis are totally missing, whereas melanoblast cells in the outer root sheath of hair follicles are not affected. In an attempt to find potent repigmenting agents for vitiligo therapy, pod extracts of Cassia occidentalis were found to be effective in inducing differentiation and migration of mouse melanoblast cell lines. The induction in melan-a melanoblast cells after 4 days in treatment medium.
glycosides were determined by measuring intracellular melanin content in B16 melanoma cells. Among the synthesized quercetin glycosides, quercetin-3-O-β-D-glucopyranoside, quercetin-3-O-β-D-glucopyranosyl-(1→4)-β-D-glucopyranoside, and 3 showed more potent intracellular melanogenesis acceleration activities than theophylline used as positive control in a dose-dependent manner with no cytotoxic effect [15]. Ali et al. [16] have also determined the ability of berberine, a principal active ingredient present in the roots of the herb *Berberis vulgaris*, to stimulate pigment dispersion in the isolated skin melanophores of the toad *Bufo melanostictus*. It was observed for mean melanophore size index of the isolated skin melanophores of *B. melanostictus* and assayed after treating with various concentrations of berberine. A marked melanin dispersion response leading to skin darkening was observed in the isolated melanophores of toad in response to berberine, which was found to be mediated through beta-2 adrenergic receptors. It was indicated that berberine causes a tremendous, dose-dependent, physiologically significant pigment dispersing in the isolated skin melanophores of *B. melanostictus*.

| Source                        | Bioactive     | In vitro / In vivo | Target site / Mode of action                                    | References |
|-------------------------------|---------------|-------------------|----------------------------------------------------------------|-------------|
| *Agaricus bisporus*           | Tyrosinase    | B16F10 Melanocytes| Phosphorylation cAMP and CREB                                   | Zaidi et al., [18] |
| *Pleurotus ostreatus*         | Tyrosinase    | B16F10 Melanocytes| TRP1, TRP2 and MITF                                             | Zaidi et al., [17] |
| Tunisian aromatic plants      | Plants extracts | B16 melanoma cell line | Tyrosinase TRP1, TRP2                                        | Kyoko et al., [4] |
| *Nelumbo nucifera*            | Palmitic acid | Human melanocytes  | MITF, TRP-2                                                    | Jeon [5]    |
| *Psoralea corylifolia*        | Psoralen      | Fish scale Melanophores | Melanin dispersal responses                                   | Ali et al., [6] |
| *Nigella sativa*              | Thymoquinone  | Wall lizard of melanophores | Melanin dispersal responses                                   | Ali & Meitei, [7] |
| *Piper nigrum*                | Piperine      | Tadpole Melanophores of frog Rana tigerina | Melanin dispersal responses                                   | Sajid & Ali, [8] |
| *Withania somnifera*          | Withaferin-A  | Melanophores of frog | Melanin dispersal effects                                    | Ali & Meitei, [7] |
| *Cassia occidentalis*         | Pod extracts  | Melanoblast cell lines | Reglementation in vitiligo                                    | Babitha et al. [10] |
| *Pyrostegia venusta*          | Murine B16F10 melanoma cells | Hydroalcoholic extracts of leaves and flowers | Increase melanin content and tyrosinase activity | Moreira et al. [11] |
| *Ficus carica*                | Psoralen      | Reptilian melanophores | Melanin dispersal effects                                    | Ali & Meitei [7] |
| *Helminthostachys zeylanica*  | 4'-O-β-D- Glucopyranosyl- quercetin- | B16 melanoma cells | Intracellular melanogenesis                                   | Yamauchi et al. [15] |
| *Berberis vulgaris*           | Berberine     | Melanophores of toad | Melanin dispersion response                                   | Ali et al. [6] |
| *Agaricus bisporus*           | Tyrosinase    | B16F10 Melanocytes | Phosphorylation cAMP and CREB                                   | Zaidi et al. [17] |

α-MSH: α-melanocyte-stimulating hormone.
Camp: cyclic adenosine monophosphate.
CREB: CREB: response element-binding protein.
Recently Zaidi et al. [17] has been investigated the effect of purified mushroom tyrosinase of Agaricus bisporus on B16F10 melanocytes for the melanin production via blocking pigment cell machinery. Using B16F10 melanocytes showed that the stimulation of melanogenesis by purified tyrosinase is due to increased tyrosinase absorption. Cellular tyrosinase activity and melanin content in B16F10 melanocytes were increased by purified tyrosinase in a dose-dependent manner. Western blot analysis revealed that cellular tyrosinase levels were enhanced after treatment with purified tyrosinase for 48 hours. Furthermore, tyrosinase induced phosphorylation of cyclic adenosine monophosphate (cAMP) response element-binding protein (CREB) in a dose-dependent manner. The purified tyrosinase-mediated increase of tyrosinase activity was significantly attenuated by H89, LY294002, Ro-32-0432, and PD98059, cAMP-dependent protein kinase inhibitors. The results indicate that purified tyrosinase can be used as contestant for the treatment of vitiligious skin conditions. Later Zaidi et al. [18] also reported that morphoanatomical effects of purified tyrosinase to determine its skin-darkening potential using B16F10 melanocyte. Phase contrast and immuno fluorescence microscopic analysis of B16F10 melanocytes has been done after treatment with various concentrations of purified tyrosinase along with standard tyrosinase (Sigma) in order to explore the mechanism of action of purified tyrosinase induced skin darkening. The phase contrast microscopic results showed that the number of melanocytes with melanin-loaded dendrites has increased significantly in purified tyrosinase treated cells in a dose dependent manner leading to skin darkening. In addition, immuno fluorescence microscopic analysis revealed purified tyrosinase increase cellular tyrosinase expression in doze dependent manner due to tyrosinase absorption in B16F10 melanocyte. Present findings proved that purified tyrosinase possesses a skin darkening potential and could be used as a safe melanogenic agent for the treatment of hypopigmentation disorders or vitiligo.

Conclusion

Different dermatological disorders, such as vitiligo, albinism and loss of the hair, the tyrosinase stimulators induce cellular melanin biosynthesis, up-regulating CREB phosphorylation and expression of MITF, tyrosinase, TRP-1 and TRP-2, and tyrosinase. These consistent results suggest that melanogenesis stimulators might be useful for treatment of hypopigmentation related disorders. The exploration and characterization of new stimulators of tyrosinase are not only useful for the medicinal purposes, but their potential applications in improving food quality and nutritional value, controlling insect pests etc are also important.

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