Supplementary Information

Supplementary Table

| Compound    | Suggested Mechanisms                  | Assay                        | Reference |
|-------------|---------------------------------------|------------------------------|-----------|
| Resveratrol | Sirtuin activator, PDE inhibitor, antioxidant | Worm PolyQ neuroprotection | (1)       |
|             |                                       | Mouse amyloid plaque reduction | (2)       |
| Mithramycin | DNA binding agent                      | Worm PolyQ neuroprotection   | (3)       |
|             |                                       | Mouse HD survival            | (4)       |
| Clioquinol  | Metal chelator and CLK-1 inhibitor     | Worm clk-1 partial mimic     | (5)       |
|             |                                       | Mouse Alzheimer cognition    | (6)       |
| Valproic acid| Anticonvulsant (HDAC inhibitor)        | Worm lifespan increase       | (7)       |
|             |                                       | Fly PolyQ neuroprotection    | (8)       |
| Mianserin   | Antidepressant (5-HT2 blocker)         | Worm lifespan increase       | (9)       |
| C2–8        | unknown                               | Fly PolyQ neuroprotection    | (10)      |
|             |                                       | Mouse HD survival            | (11)      |
| Rapamycin   | mTOR inhibitor                         | Cell synuclein/PolyQ         | (12)      |
|             |                                       | Mouse AD cognition           | (13)      |

Table S1: Compounds used in *dnj-14* lifespan screen.
**Figure legends**

**FIG. S1: dnj-14 gene structure, mutations and homology to human dnajc5.**

(A) Sequence alignment of the predicted products of the human DNAJC5 and *C. elegans* dnj-14a genes (CSPα and DNJ-14 proteins, respectively).

(B) Exon structure of the *dnj-14* gene and location of the *ok237* and *tm3223* alleles.

(C) Confirmation of the *ok327* deletion. Genomic DNA from wild type N2 and *dnj-14(ok237)* worms was amplified using primers flanking the deletion (left panel) or with one flanking primer and a primer within exon 2 of *dnj-14* (right panel).

(D) Characterisation of the *tm3223* insertion/deletion. Genomic DNA from wild type N2 and *dnj-14(tm3223)* worms was amplified using primers flanking the mutation site.

**FIG. S2: Quantification of neurodegeneration in *dnj-14(ok237)* mutants.**

Wild type N2 and *dnj-14(ok237)* worms were synchronised and grown on NGM plates containing vehicle control (ethanol) or 100 μM resveratrol for at least 9 days. Animals were then immobilised for GFP imaging and scored for head neuron abnormalities (‘neuron loss’) based on loss of neuronal cell bodies, a reduction in the number of visible neurites, or the presence of contorted neuronal processes in the head of the worms; and the presence of large fluorescent punctae in the dorsal nerve cord (‘punctae’). The number of worms analysed was 45 for N2, 35 for *dnj-14* and 20 for *dnj-14* Resveratrol.

**FIG. S3: Visualisation of neurodegeneration in *dnj-14(ok237)* mutants.**

Wild type N2 and *dnj-14(ok237)* worms were synchronised and grown on NGM plates for at least 9 days. Animals were then immobilised for GFP imaging. Typical images are shown for various animals. These illustrate the loss of neuronal cell bodies, reduction in the number of
visible neurites, or the presence of contorted neuronal processes in the head of the worms that was frequently seen in *dnj-14* mutants. In contrast, age-matched wild type N2 animals generally exhibited obvious neuronal cell bodies and had clearly labelled multiple neurites that extended straight to the end of the worm’s head without twisting.

**FIG. S4: Mechanosensation is unaffected in aged *dnj-14* mutants.**

Wild type N2, *dnj-14(tm3223)* and *dnj-14(ok237)* worms were synchronised and grown on NGM plates until 6 days of age. After transfer to unseeded plates, mechanosensation was assessed by gently touching an eyelash to the side of the worm’s head at a perpendicular angle. Each worm was assayed ten times and the number of times the worm either stopped or reversed its direction of movement was recorded. No significant differences between wild type and *dnj-14* mutants were seen (n=15 animals per strain).

**FIG. S5: Pharyngeal pumping is unaffected in aged *dnj-14* mutants.**

Wild type N2, *dnj-14(tm3223)* and *dnj-14(ok237)* worms were synchronised and grown on NGM plates until 6 days of age. After transfer to plates freshly seeded with OP50 bacteria, the number of contraction/relaxation cycles of the pharynx per thirty seconds was for each worm was recorded. No significant differences between wild type and *dnj-14* mutants were seen (n=15 animals per strain).

**FIG. S6: Measurement of cAMP levels in worms treated with resveratrol and rolipram.**

Wild type N2 worms were age-synchronised and grown on 60-mm NGM plates seeded with JB1669 adenylyl cyclase deficient bacteria. Approximately fifteen such plates of day 2 worms were washed in M9 buffer and then treated in M9 buffer containing vehicle control (ethanol), 100 µM resveratrol or 100 µM rolipram for 120 mins. The worms were then lysed,
centrifuged to pellet any debris, and the supernatant used immediately for assay. Endogenous cyclic AMP levels were measured by ELISA and normalised to total protein concentration. Data shown are mean ± SEM (n = 4 biological replicates). No significant differences between treatments were seen.

**MOVIE S1: Superficially normal locomotion in *dnj-14(ok237)* mutants**

The movement of worms on NGM agar plates was recorded. Wild type N2 worms are shown first, followed by *dnj-14(ok237)* mutants.
Supplementary References

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Fig S1 Kashyap et al
Fig S2 Kashyap et al
Fig S3 Kashyap et al
Fig S4 Kashyap et al
Pharyngeal pumps per 30 s

- Wild type
- tm3223
- ok237
Fig S6 Kashyap et al