Incidence and antibiotic sensitivity pattern of New Delhi metallo-β-lactamase-1 (NDM-1) producers among carbapenem resistant enterobacteriaceae in a tertiary care teaching hospital, Bareilly: A cross sectional study

Anju Saxena¹, Sujata Singh², Rahul Kumar Goyal³, Jaspreet Kaur⁴, Sumit Saxena⁵*

¹,⁵Assistant Professor, ²Professor & HOD, ³Professor, ⁴Research Associate, ¹,²,⁴Dept. of Pharmacology, ⁵Dept. of Community Medicine,

*Corresponding Author:
Email: drsumitsaxena22@gmail.com

Abstract

Objectives: Antibiotic resistance in microorganisms has become a critical health issue these days and has evolved to become a worldwide health threat. Carbapenem-resistant Enterbactriaceae (CRE) is one example which really is a nightmare bacteria, resistant to most, and in some cases all, antibiotics. The commonest cause is elaboration of various types of carbapenemases amongst which the recent detection of New Delhi metallo beta-lactamase-1 (NDM-1), a superbug, further compounded the problem. So the present study was undertaken to study the pattern of carbapenem resistance and role of carbapenamase (blaNDM-1) gene towards it.

Materials and Methods: The study was a cross sectional study conducted in the department of Pharmacology in collaboration with Microbiology and Biochemistry, SRMS IMS, Bareilly. All clinical isolates, their sensitivity pattern and MBL detection of carbapenem resistant ESBLs were obtained from Microbiology records. blaNDM-1 gene was detected by PCR.

Results: A total of 312 ESBL producers were isolated. Out of which 42 were found to be carbapenem resistant. Amongst these 19.05% (8) were found to be MBL positive and 5 (62.5%) MBL positive isolates were NDM positive. ESBL producers were sensitive to very few antibiotics and NDM producers were sensitive only to polypeptide antibiotics.

Conclusion: ESBL is a great problem for gram negative organisms and now the frequency of carbapenem resistance is high which has limited antibiotics available for the treatment. Furthermore carbapenemase (blaNDM-1) gene has also shown a substantial role. This high incidence of antibacterial resistance demands development of newer antibiotics.

Keywords: Carbapenem resistance, ESBL, blaNDM-1, MBL.

Introduction

Resistance to β-lactams is a long recognised problem in Gram-negative bacteria¹ and with the introduction of new classes of β-lactams, novel β-lactamases have emerged.¹,² Carabapenem resistance has become a growing problem over the last decade with the emergence of readily transferable plasmid mediated carbapenem-hydrolysing β-lactamases and has posed a threat to the antimicrobial world.³,⁴ According to a WHO report the “…world is heading towards a post-antibiotic era in which many common infections will no longer have a cure and once again kill unabated”.⁵ Carbapenemases are the agents of last resort against many multi drug resistant, gram negative bacteria. Pseudomonas and Acinetobacter baumannii have significant carbapenem resistance. These enzymes confer resistance to the other beta-lactam agents as well, including extended spectrum cephalosporins. Metallo beta lactamases (MBLs) are one such type of carbapenemase, that are characterized by the ability to hydrolyze carbapenemabs. A novel MBL named NDM-1 (New Delhi metallo-β-lactamase) was identified from Klebsiella pneumoniae (strain 05-506) and Escherichia coli isolates recovered from a Swedish patient transferred from India.⁶ Since then, there have been an increasing number of infections in patients from India, Pakistan and the United Kingdom.⁷ Various studies conducted in the past few years have shown its dissemination across the gram negative organisms and included E.coli, Klebsiella pneumonia, Citrobacter.⁸ Effective surveillance systems have been put in place in some countries to track the emergence and spread of resistance to anti-infective. Such surveillance has been able to bring about changes in national policies and practices. With this background, the present study was undertaken to study the variability in antimicrobial resistance pattern in carbapenem resistant organisms amongst indoor and outdoor patients and genotypic identification of carbapenemase (blaNDM-1) gene.

Materials and Methods

Study design: Cross sectional study.

Place of study: Department of Pharmacology in collaboration with Department of Microbiology and Department of Biochemistry, Shri Ram Murti Smarak Institute of Medical Sciences, Bhojipura, Bareilly.

Study period: April 2013 to March 2014.

Sample: Clinical isolates of various gram positive and gram negative organisms, from sputum, endotracheal tip, tracheal aspirate, urine, pus swab, pus aspirate, bronchial wash, catheter tip, blood, pleural fluid, peritoneal fluid, pericardial fluid, ascitic fluid, sample from shunt tube, corneal swab, intracervical swab, wound tissue, CSF and drainage tip from the inpatients and outpatients of Obstetrics and Gynaecology, Surgery, Medicine, Orthopaedics, Ophthalmology,
The study was conducted in compliance with the protocol and the Institutional Ethics Research Committee (IERC).

Antimicrobial susceptibility test was done by Kirby-Bauer sensitivity testing method. Carbapenem resistance was analysed amongst various ESBL positive isolates and was grouped into IPD and OPD isolates. The sensitivity of ESBL producing carbapenem resistant organisms was also evaluated and then this was compared between the two groups. Metallo β-lactamase (MBL) was tested using double disk diffusion test (DDDT) in carbapenem resistant ESBLs. blaNDM-1 gene was detected by PCR for which the positive control was obtained from department of Microbiology, Institute of Medical Sciences, Banaras Hindu University, Varanasi. The primers for this gene were obtained from HiMedia Laboratories Pvt Limited, Mumbai. Samples of carbapenem resistant ESBLs were processed further for PCR. Further we analysed positive MBL and blaNDM-1 gene for their role towards carbapenem resistance and their ultimate contribution towards antibacterial resistance and the available treatment options. The data was analysed using IBM SPSS version 20. Chi-square test was applied wherever applicable to check the significant difference among the different groups. p value of ≤ 0.05 was considered to be significant.

Results

Present cross sectional study includes a total of 2008 samples from various clinical departments. Of these 2008 samples, 655 (32.62%) specimen gave significant growth of bacteria while rest were either non-pathogenic or sterile (Fig. 1).

Of the total number of positive isolates 576 (87.94%) were from IPD and 79 (12.06%) specimen were from OPD. Majority of the specimen were infected by gram negative bacteria, E.coli (25.3%) (Fig. 2). Out of total positive samples, 47.63% (312) isolates were ESBL producers (Fig. 3) and out of these, 42 ESBL producing isolates were found to be carbapenem resistant and maximum incidence was observed with Acinetobacter followed by Pseudomonas and none of the Proteus isolates were carbapenem resistant. Carbapenem resistance was significantly more in IPD isolates as compared to OPD isolates (p<0.05) (Table 1). On further analysing the sensitivity pattern of carbapenem resistant isolates, it was found that E.coli was sensitive in decreasing order to Cl & PB followed by Cot & C while Klebsiella & Pseudomonas were sensitive only to Cl & PB. (Table 2a & 2b). Out of 42 carbapenem resistant ESBL positive isolates, 8 were MBL positive & when genotypic identification (Fig. 4-6) was done for carbapenemase gene it was found that 5 of MBL positive isolates were NDM positive (Table 3).

The data also reveals that there is a positive correlation between carbapenem resistant ESBL & MBL and also between MBL & NDM (p<0.05).

| Organism       | IPD      | OPD      | P-value |
|----------------|----------|----------|---------|
| Acinetobacter  | 12       | 0        | N.A.    |
| Citrobacter    | 7        | 0        | N.A.    |
| Proteus        | 0        | 0        | N.A.    |
| Klebsiella     | 8        | 0        | >0.05   |
| Pseudomonas    | 9        | 0        | >0.05   |
| E.coli         | 4        | 2        | 10.53   | >0.05   |

Fig. 1: Total number of samples

Fig. 2: Distribution of organisms in IPD and OPD groups

Fig. 3: Phenotypically positive ESBL

Table 1: Distribution of Carbapenem Resistant ESBL Organisms
Table 2a: Antibiotic Sensitivity Pattern of Carbapenem Resistant ESBL Positive Enterobacteriaceae

|          | IPD (n=4) | OPD (n=2) | p-value | Lebsiella | IPD (n=8) | OPD (n=0) | p-value | IPD (n=7) | OPD (n=0) | p-value |
|----------|-----------|-----------|---------|-----------|-----------|-----------|---------|-----------|-----------|---------|
| Cip      | 0         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| Le       | 0         | 0         | NA      | 1         | 0         | 0         | NA      | 1         | 0         | NA      |
| Of       | 0         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| AK       | 0         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| Gen      | 0         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| Tb       | 0         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| Tet      | 0         | 1         | >0.05   | 1         | 0         | NA       | 1       | 0         | NA       |         |
| C        | 1         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| Cot      | 1         | 1         | >0.05   | 0         | 0         | NA       | 0       | 0         | NA       |         |
| CFS      | 0         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| AMC      | 0         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| Pit      | 0         | 0         | NA      | 0         | 0         | 0         | NA      | 0         | 0         | NA      |
| Nit      | -         | -         | -       | 0         | 0         | NA       | 0       | 0         | NA       |         |
| Cl       | 4         | 2         | >0.05   | 7         | 0         | NA       | 6       | 0         | NA       |         |
| PB       | 4         | 2         | >0.05   | 7         | 0         | NA       | 6       | 0         | NA       |         |

Table 2b: Antibiotic Sensitivity Pattern of Carbapenem Resistant ESBL Positive Enterobacteriaceae

|          | IPD (n=12) | OPD (n=0) | p-value | IPD (n=9) | OPD (n=0) | p-value |
|----------|------------|-----------|---------|-----------|-----------|---------|
| Cip      | 0          | 0         | NA      | 0         | 0         | NA      |
| Le       | 0          | 0         | NA      | 0         | 0         | NA      |
| Of       | 0          | 0         | NA      | 0         | 0         | NA      |
| AK       | 0          | 0         | NA      | 1         | 0         | NA      |
| Gen      | 0          | 0         | NA      | 1         | 0         | NA      |
| Tb       | 0          | 0         | NA      | 1         | 0         | NA      |
| Tet      | 0          | 0         | NA      | -         | -         | -       |
| C        | 0          | 0         | NA      | -         | -         | -       |
| Cot      | 0          | 0         | NA      | -         | -         | -       |
| CFS      | 1          | 0         | NA      | 0         | 0         | NA      |
| AMC      | 0          | 0         | NA      | 0         | 0         | NA      |
| Pit      | 0          | 0         | NA      | 1         | 0         | NA      |
| Nit      | 0          | 0         | NA      | -         | -         | -       |
| Cl       | 12         | 0         | NA      | 9         | 0         | NA      |
| PB       | 12         | 0         | NA      | 9         | 0         | NA      |

Fig. 4: Positive control of NDM-1
Fig. 5: Four NDM-1 positive strains
In India, the prevalence of ESBLs has been reported since the 1990s. The phenotypic data generated in the current study indicates a high prevalence (47.63%) of ESBL producers. This number is less than that previously reported by Dalela et al in 2012 and Narayanswamy and Mallika in 2011. Out of the total ESBL producers, maximum frequency was observed with Escherichia coli (79.52%) in the current study which is in accordance to a study conducted in Chennai which showed ESBL production among 75.5% Escherichia coli isolates while 49.32% Escherichia coli were ESBL producer in a study conducted in Mumbai by Aruna et al 2012.

The reports presented by different authors clearly indicate that the prevalence of ESBL producing organisms among clinical isolates vary greatly geographically and rapidly changing over time. Antibiotic options in the treatment of ESBL producing organisms are extremely limited. Carbapenems are the treatment of choice for serious infections due to ESBL-producing organisms and are often used as “last line agents” or “antibiotics of last resort”.

Our study also analysed that 42 ESBL positive organisms are carbapenem resistant leaving only polypeptide antibiotics and a trickle of drugs for their treatment. Our results were found to be similar to study conducted in 2013 in Mumbai which showed 12.26% Enterobacteriaceae to be carbapenem resistant. Other studies conducted in various parts of India showed carbapenem resistance in Enterobacteriaceae to be in the range of 7.87% to 51%. This variability could be due to difference in study design, study population, geographical distribution and different drug pressure in community.

When the sensitivity pattern of these carbapenem resistant ESBL positive isolates was analysed it was noted that 87.5% Klebsiella was sensitive only to polypeptide antibiotics and no drugs were available for the rest 12.5% making them eternal and thereby spreading resistance to other organisms by genetic transfer. In the same way, carbapenem resistant ESBL producing Acinetobacter and Pseudomonas were found to be sensitive only to polypeptide antibiotics suggesting need for development of innovative and improved drugs and their vigilant use.

As carbapenems have long been considered the antibiotic class of last resort in the treatment of infections caused by multidrug-resistant organisms, the dissemination of carbapenem resistance among pathogenic bacteria has been declared a “global sentinel event”.

The current condition suggests judicious and rational use of the antibiotics so that the resistance rate is minimised. Many countries have also given Standard treatment guidelines (STG) to direct choice of antibiotics.

Carbapenem resistance due to acquired carbapenemases has emerged and spread worldwide since the early 2000s and the number of bacteria that produce metallo-β-lactamase (MBL) is on the rise. In the present study, 19.05% carbapenem resistant ESBL positive isolates were MBL positive.

In the present study we also observed that out of 8 MBL positive strains, 62.5% isolates harboured blaNDM-1 gene. Similar to this study, Seema et al., reported 54 (84.38%) NDM – 1 producing Enterobacteriaceae. Out of this, 33.33% were found in E.coli and 37.5% were found in Klebsiella which was comparable to a study conducted by Deshpande et al which reported the incidence of NDM – 1 in 9 E.coli isolates among 24 carbapenem resistant Enterobacteriaceae (37.5%) in a tertiary care centre in Mumbai. Another northeastern study by Bora et al, 2013 showed 5.2% blaNDM-1 gene in E.coli and a study carried out in the intensive care unit (ICU) and wards of Sir Ganga Ram Hospital, Delhi showed 8.1% overall prevalence of NDM – 1 in E.coli isolates. Plasmid transfer is one of the major factors in disseminating antibacterial drug-resistant bacteria. This difference in the percentage of presence of blaNDM-1 gene could be because of the different geographical areas. These reports indicate an alarming risk of rapid dissemination of NDM-1. Identification of a significant number of
NDM – 1 producer in E.coli is an additional source of concern as it suggests that the resistance is being disseminated in the environment as well as in the hospital. E.coli is the cause of many community acquired infections, such as diarrhea and urinary tract infections. Overcrowding coupled with poor hygiene, difficulty in obtaining potable water, poor sanitation, sale of non – prescribed antibiotics (self-medication) and weak hospital antibiotic policies may be contributing factors for selecting and facilitating the spread of NDM – 1 producers. The possible spread of NDM – 1 producer reinforces the urgent need to develop novel anti – gram – negative molecules and the implementation of a worldwide network of sentinel laboratories for antibiotic resistance surveillance. The need of the hour is to preserve antibiotics and trials should be encouraged to study the impact of carbapenem restriction for treatment of ESBLs on the incidence of MBL – producing gram negative pathogens.

Careful surveillance of antibiotic use and resistance patterns could aid in the development of antimicrobial guidelines to improve appropriate use and delay the spread of resistance and thus surveillance studies should be done periodically and in different geographical areas and the results should be made available to all the treating physicians in order to guide proper use of antibiotics.

Conclusion
Antibiotic resistance is a serious public health threat that is on the rise — bacteria that are resistant to our best antibiotics continue to emerge. ESBL and now blαNDM-1 (carbapenemase) gene has also found its place in the world of antibiotic resistance. This high incidence of antibacterial resistance demands development of newer antibiotics.

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Limitations of the study
This study was conducted in a limited area and thus may not represent the whole population.

Conflicts of Interest
There are no conflicts of interest.

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