Micro-Sized Cold Atmospheric Plasma Source for Brain and Breast Cancer Treatment

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ABSTRACT: Micro-sized cold atmospheric plasma (µCAP) has been developed to expand the applications of CAP in cancer therapy. In this paper, µCAP devices with different nozzle lengths were applied to investigate effects on both brain (glioblastoma U87) and breast (MDA-MB-231) cancer cells. Various diagnostic techniques were employed to evaluate the parameters of µCAP devices with different lengths. Parameters included potential distribution, electron density, and optical emission spectroscopy. The generation of short- and long-lived species (such as hydroxyl radical [•OH], superoxide [O2•−], hydrogen peroxide [H2O2], and nitrite [NO2−]) were studied. These data revealed that µCAP treatment with a 20 mm long tube has a stronger effect than that of the 60 mm tube because of the synergetic effects of reactive species and free radicals. Reactive species generated by µCAP enhanced tumor cell death that was dose-dependent and nonspecific to tumor cell type.

KEY WORDS: reactive species, glioblastoma cancer, cancer therapy

I. INTRODUCTION

Cold atmospheric plasma (CAP) has been proposed as a novel therapeutic method for anticancer treatment, which can be applied to living tissues and cells. 1,2 CAP is a partially ionized gas that contains charged particles, reactive oxygen and nitrogen species (ROS and RNS), excited atoms, free radicals, UV photons, electric field, and other substances. 3,4 ROS and RNS, combined or independently, are known to initiate different signaling pathways in cells and to promote oxidative stress. 5,6 Plasma-induced biological effects include damage to lipids, proteins, and DNA, and apoptosis induced through plasma-generated ROS and RNS. 7–10 Moreover, many studies have reported both in vivo and in vitro that plasma is a possible adjunct treatment in oncology. It can kill several cancer cell types, such as glioblastoma, breast cancer, bladder carcinoma, cervical carcinoma, skin carcinoma, pancreatic carcinoma, lung carcinoma, colon carcinoma, gastric carcinoma, melanoma, and hepatocellular carcinoma. 11–27
In plasma medicine, jet plasma, corona discharge, and dielectric barrier discharge (DBD) have been used. These types of plasma can be directly applied to skin cancers; however, they are not suitable for systemic cancer treatment. Some studies investigated the use of micro-sized plasma devices in live animals. However, their devices were only used on xenograft tumors and not systemic cancers. Moreover, delivery of the plasma species is crucial to suppress tumor growth and assess efficiency of micro-sized plasma devices. Hence, this study aims to design micro-sized cold atmospheric plasma devices with different nozzle lengths to enhance delivery of reactive species and evaluate the efficiency of these devices for cancer therapy. Figure 1 shows the potential applications of μCAP for brain and breast tumors in the future.

II. MATERIALS AND METHODS

Figure 2 depicts the schematic of the experimental setup, including high voltage power (see Fig. 2a) and μCAP devices (Fig. 2b). The high voltage power includes DC input, trigger signal and metal-oxide-semiconductor field-effect transistor (MOFSET), and the secondary output. In this work, the DC input was set at 5 V, a square wave signal was obtained from the control unit (see Fig. 2a, upper left), and a high voltage wave was obtained from the square wave signal through the transformer (see Fig. 2a, upper right). The μCAP devices consist of a two-electrode (copper) assembly with a central powered electrode (1 mm in diameter) and a grounded outer electrode wrapped around the outside of a quartz tube (10 mm), as shown in Fig. 2b. The electrodes were connected to the secondary output of the high voltage transformer. The peak-to-peak voltage was approximately 8 kV, and the frequency of the discharge was around 16 kHz (see Fig. 2a, upper right). The secondary output of the high voltage transformer was connected to

**FIG. 1:** Potential applications of μCAP for brain and breast tumors
the first input. At the end of a quartz tube, a capillary tube (stainless steel) with an inner diameter of 275 ± 5 μm and a length of 20 or 60 mm was attached and insulated with epoxy. The feed gas for this study was industrial purity helium, which was injected into the quartz tube with a 0.2 L/min gas flow rate. A longer tube (60 mm) is needed to access deeper tumors in the brain and breast.

In this study, we are assessing the effect of tube length to understand the limitations of depth. For instance, it is believed that a longer tube (60 mm) is needed to access deeper tumors in the brain and breast. UV-visible-NIR, a range of wavelengths from 200 to 850 nm, was investigated on plasma to detect various RNS and ROS (nitrogen [N₂], nitric oxide [–NO], nitrogen cation [N⁺₂], atomic oxygen [O], and hydroxyl radicals [–OH]). The optical probe was placed 1.0 cm in front of the plasma jet nozzle. Data were then collected with an integration time of 100 msec.

A fluorimetric hydrogen peroxide assay kit (Sigma-Aldrich) was used for measuring the amount of H₂O₂, according to the manufacturer’s protocol. Briefly, 50 μL of standard curve, control, and experimental samples were added to 96-well flat-bottom black plates, and then 50 μL of Master Mix was added to each well. The plates were incubated for 20 min at room temperature, protected from light, and fluorescence was measured with a Synergy H1 Hybrid Multi-Mode Microplate Reader at Excitation/Emission (Ex/Em): 540/590 nm.

RNS levels were determined with a Griess Reagent System (Promega Corporation) according to the instructions provided by the manufacturer. Briefly, 50 μL of sample and 50 μL of the provided sulfanilamide solution were added to 96-well flat-bottom plates and incubated for 5 to 10 min at room temperature. Subsequently, 50 μL of the NED solution was added to each well and incubated at room temperature for 5 to 10 minutes. The absorbance was measured at 540 nm with the Synergy H1 Hybrid Multi-Mode Microplate Reader.
XTT sodium salt (2,3-bis(2-methoxy-4-nitro-5-sulfophenyl)-5-[(phenylamino) carbonyl]-inner salt-2H-tetrazolium, monosodium salt) solution, purchased from Cayman Chemical, was prepared by dissolving XTT power in Dulbecco’s Modified Eagle Medium (DMEM, Life Technologies). XTT sodium salt solution (100 μL per well, 500 μM) in a 96-well flat-bottom plate by μCAP for 5, 10, 30, 60, and 120 seconds. The gap between the outlet of the μCAP and the surface of the samples was set at approximately 3 mm. As a control, untreated XTT sodium salt solution in triplicate were transferred to a 96-well flat-bottom plate. As a control, DMEM (100 μL per well) was treated with μCAP for 5, 10, 30, 60, or 120 seconds. The color change of XTT solution was used to indicate the presence of superoxide (O₂⁻). A color change of XTT solution was measured with a Hach DR 6000 UV-VIS spectrophotometer at 470 nm.

A methylene blue (MB) solution was prepared by dissolving MB powder in DMEM. MB solutions (100 μL per well, 0.01g/L) in a 96-well flat-bottom plate were treated by μCAP for 5, 10, 30, 60, or 120 seconds. The gap between the outlet of the μCAP and the surface of the samples was approximately 3 mm. As a control, untreated MB solutions in triplicate were transferred to a 96-well flat-bottom plate. The color change of methylene blue shows the presence of OH radicals via immediate and distinct bleaching of methylene blue dye (qualitatively analysis). The color change of the MB solution was measured at the absorbance at 664 nm by a Synergy H1 Hybrid Multi-Mode Microplate Reader.

Human glioblastoma cancer cells (U87MG, Perkin Elmer) were cultured in DMEM (Life Technologies) supplemented with 10% (v/v) fetal bovine serum (Atlantic Biologicals) and 1% (v/v) penicillin and streptomycin (Life Technologies). Cultures were maintained at 37°C in a humidified incubator containing 5% (v/v) CO₂. The human breast cancer cell line (MDA-MB-231) was cultured in DMEM supplemented with 10% (v/v) fetal bovine serum (Atlantic Biologicals) and 1% (v/v) penicillin and streptomycin. Cultures were maintained at 37°C in a humidified incubator containing 5% (v/v) CO₂.

U87 and MDA-MB-231 cells were plated in 96-well flat-bottom microplates at a density of 3000 cells per well in 100 μL of complete culture medium. Cells were incubated for 24 h to ensure proper cell adherence and stability. On day 2, the cells were treated with He μCAP for 0, 5, 10, 30, 60, or 120 seconds. Cells were further incubated at 37°C for 24 and 48 h. The viability of the glioblastoma and breast cancer cells was measured for each incubation time point with an MTT assay. 100 μL of MTT solution (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) (Sigma-Aldrich) was added to each well, and plates were incubated for 3 h. The MTT solution was discarded, and 100 μL per well of MTT solvent (0.4% (v/v) HCl in anhydrous isopropanol) was added to the wells. The absorbance of the purple solution was recorded at 570 nm with a Synergy H1 Hybrid Multi-Mode Microplate Reader.

III. RESULTS AND DISCUSSION

The reactive species generated by the μCAP device with different tube lengths are detected by optical emission spectroscopy, as shown in Fig. 3. The identification of the
emission line and bands was performed mainly according to reference.\textsuperscript{30} For devices with tubes that were 20 mm and 60 mm long, an $N_2$ second-positive system (315 nm, 337 nm, 357 nm, and 380 nm) representing the photon emission intensity drops from the state $C^3\Pi_u$ to $E^5\Pi_g$ with different upper and lower vibration quantum numbers. There are very weak emission lines in the special range of 250 to 300 nm, which are detected as NO lines. The helium bands were assigned between 500 and 750 nm, as shown in Fig. 3a and 3b. We also observed a high-intensity OH/O$_3$ peak at 309 nm for devices with both 20 mm and 60 mm lengths. Atomic oxygen (O, including the ground state and all the excited states of atomic oxygen) was observed at 777 nm in both devices, which was believed to have a significant effect on cells and therefore a broad biomedical application. Micro-sized plasma is a complicated mix that combines the comprehensive effect of different ions and reactive species. Because its delivery distance is longer, the 60 mm μCAP has fewer electrons and reactive species than the 20 mm μCAP.

The experimental Rayleigh microwave scattering (RMS) system was described previously.\textsuperscript{19} The detection of the scattered signal was accomplished using a homodyne scheme by means of an $I/Q$ mixer, providing in-phase ($I$) and quadrature ($Q$) outputs. For the entire range of scattered signals, the amplifiers and mixer were operated in linear mode. The total amplitude of the scattered microwave signal was determined by: $U = \sqrt{I^2 + Q^2}$. We can calculate the total electron number in the plasma as $N_e = U(\omega^2 + v_w^2)/(2.82 \times 10^{-4} A v_m)$, where $\omega$ is the angular frequency, $v_w$ is the frequency of the electron-neutral collisions, and $A$ is the proportionality coefficient.\textsuperscript{31} The total electron number in the jet from μCAPs with lengths of 20 mm and 60 mm is presented in Fig. 4a and 4b, and the total electron number for one discharge period is $4.60 \times 10^{12}$ and $4.04 \times 10^{12}$, respectively. A very small decrease of electron number has been detected in the 60 mm μCAP compared to the 20 mm μCAP.
XTT solution was used to determine the relative concentration of superoxide ($O_2^-$). Superoxide radical reduced soluble formazans of the tetrazolium dye XTT. Figure 5a and 5b shows the relative superoxide concentration of 20 mm and 60 mm μCAP treatment of DMEM. Relative intensity increases with treatment, which corresponds to the relative concentration of superoxide increasing with treatment. Comparing the 20 mm with 60 mm lengths, the 20 mm μCAP device produced a higher relative concentration of superoxide than the 60 mm device. Methylene blue (MB) was used to assess the relative concentration of hydroxyl radicals (•OH). MB reacts with •OH aqueous solutions, leading to a visible color change. Figure 5c and 5d shows that the relative MB concentration decreases with the treatment time of μCAP, suggesting that more •OH species are generated in DMEM (20 mm > 60 mm). Overall, these findings demonstrate that there is an increase in the relative concentration of $O_2^-$ and •OH as a function of μCAP treatment time.

DMEM treated with the 20 mm and 60 mm μCAP devices induced changes in the concentration of $H_2O_2$ and $NO_2^-$ as a function of the treatment time. These results are shown in Fig. 6, with concentrations produced by the 20 mm and 60 mm He μCAP devices. In Fig. 6a, the $H_2O_2$ concentrations produced by the 20 mm He μCAP device increased with treatment time up to 60 sec; but between 60 and 120 sec, the concentration decreased. The $H_2O_2$ concentration produced by 60 mm He μCAP increased with treatment time (see Fig. 6b). This result means that the $H_2O_2$ concentration reaches saturation earlier in the 20 mm length than with the 60 mm length μCAP device. In Fig. 5, we know that He μCAP produces •OH and $O_2^-$ in DMEM, which are the two most important species in plasma-activated media. In particular, •OH reacting with •OH and $O_2^-$ reacting with $2H^+$ lead to $H_2O_2$ formation. $NO_2^-$ concentrations of both the 20 mm and 60 mm devices increase with treatment time (see Fig. 6c and 6d), and $NO_2^-$ concentrations of the 20 mm device is much higher than that for the 60 mm device. Comparing $NO_2^-$ concentration with the $H_2O_2$ concentration under the same conditions, $NO_2^-$ concentration is much higher than $H_2O_2$ concentration. A possible hypothesis for this result is that DMEM comprises over 30 components such as in-
organic salts, amino acids, and vitamins, and plasma might react with amino acids to form NO₂⁻.

Figure 7 shows the viability of the brain (glioblastoma U87) cancer cells after 24 and 48 h incubation with µCAP treatment during 5, 10, 30, 60, and 120 sec treatment with the 20 mm and 60 mm length µCAP devices, respectively. For the 20 mm length µCAP treatment, the cell viability of brain cancer cells was lower than that for the 60 mm length at each treatment duration (from 5 to 60 sec), and dropped with increasing treatment time. For both 20 mm and 60 mm devices, 120 sec treatment has a similar effect on cell viability of U87 cancer cells. For 48 h incubation under 20 mm µCAP treatment, 60 and 120 sec duration had a similar effect on cell viability. Thus, our overall conclusion is that the 60 mm tube can still produce reactive species in deeper tumors.

Figure 8 shows the viability of the breast (MDA-MB-231) cancer cells after 24 and 48 h incubation with µCAP treatment with the 20 mm and 60 mm length µCAP devices.

**FIG. 5:** Relative O₂⁻ and •OH concentration of 20 mm and 60 mm µCAP-treated DMEM. For relative O₂⁻ concentration: (a) 20 mm and (b) 60 mm. For relative •OH concentration: (c) 20 mm and (d) 60 mm. Student’s t-test was performed, and the statistical significance compared to µCAP 5 s treatment is indicated as *p < 0.05, **p < 0.01, ***p < 0.001 (n = 3).
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During 5, 10, 30, 60, or 120 sec duration. For both 20 mm and 60 mm μCAP treatment, cell viability after 24 and 48 h incubation dropped with increasing treatment time. For 20 mm μCAP treatment, the viability of breast cancer cells was lower than that of the 60 mm length at each treatment duration.

The direct plasma jet irradiation is limited to the skin. It can also be invoked as a supplemental therapy during surgery because it only causes cell death in the upper three to five cell layers. However, the current cannulas from which the plasma emanates are too large for intracranial applications. Thus, we developed micro-sized plasma devices with 20 mm and 60 mm length stainless steel tubes, both of which can effectively kill brain and breast cancer cells. This preliminary study offers significant potential for new treatment applications. Numerous studies reported plasma-induced apoptosis in cancer cells due to various plasma-generated reactive species. Plasma generates multiple ROS and RNS, including hydrogen peroxide (H₂O₂), ozone (O₃),

![Graphs depicting H₂O₂ and NO₂⁻ concentration](image)

**FIG. 6:** H₂O₂ and NO₂⁻ concentration of 20 mm and 60 mm μCAP-treated DMEM. For H₂O₂ concentration: (a) 20 mm and (b) 60 mm. For NO₂⁻ concentration: (c) 20 mm and (d) 60 mm. Student’s t-test was performed, and the statistical significance compared to μCAP 5 sec treatment is indicated as *p < 0.05, **p < 0.01, ***p < 0.001 (n = 3).
hydroxyl radical (•OH), atomic oxygen (O), superoxide (O$_2^-$), nitric oxide (NO) and peroxynitrite anion (ONOO$^-$), singlet delta oxygen (O$_2$($^1\Delta_g$)), and nitrite (NO$_2^-$).\textsuperscript{37,38} All of these are shown in Fig. 3. In these experiments, we have specifically measured relative concentrations of O$_2^-$ and •OH (short-lived species, see Fig. 5) and the concentration of H$_2$O$_2$ and NO$_2^-$ (long-lived species, see Fig. 6). The relative concentration of O$_2^-$ treated by μCAP devices with 20 mm and 60 mm lengths increases with treatment time (see Fig. 5a and 5b). O$_2^-$ can activate mitochondrial-mediated apoptosis by changing the mitochondrial membrane potential. It simultaneously upregulates pro-apoptotic genes and downregulates anti-apoptotic genes for activation of caspases resulting in cell death.\textsuperscript{39} Figure 5c and 5d shows the relative concentration of •OH in DMEM treated by μCAP with 20 mm and 60 mm lengths also increases with treatment time.

**FIG. 7:** Cell viability of U87 after 24 and 48 h incubation with μCAP treatment with 20 mm and 60 mm length during 5, 10, 30, 60, and 120 sec treatments. Cell viability of U87 treated by 20 mm He μCAP at (a) 24 h incubation and (c) 48 h incubation. Cell viability of U87 treated by 60 mm He μCAP at (b) 24 h incubation and (d) 48 h incubation. The ratios of surviving cells for each cell line were normalized relative to controls (DMEM). Student’s t-test was performed, and the statistical significance compared to cells present in DMEM is indicated as *p < 0.05, **p < 0.01, ***p < 0.005 (n = 3).
•OH derived amino acid peroxides can contribute to cell injury because •OH itself and protein (amino acid) peroxides react with DNA, thereby inducing various forms of damage. Compared with cell viability of both cancer lines, the trend of cell death can be partly attributed to the increase of $O_2^-$ and •OH concentrations with treatment time. On the other hand, the 20 mm μCAP device shows higher relative concentrations of $O_2^-$ and •OH, such that the 20 mm μCAP device is more effective in killing both cancer cell lines than the 60 mm μCAP device. Figure 6 shows $H_2O_2$ and $NO_2^-$ concentrations of the DMEM treated with the 20 mm and 60 mm μCAP devices. $H_2O_2$ can induce cell death by apoptosis and necrosis, whereas $NO_2^-$ is known to induce cell death via DNA damage. Thus, the synergism of $H_2O_2$ and $NO_2^-$ might be an important factor in the efficiency of killing cancer cells.

FIG. 8: Cell viability of MDA-MB-231 after 24 and 48 h incubation with μCAP treatment with 20 mm and 60 mm length during 5, 10, 30, 60, and 120 sec treatment. Cell viability of MDA-MB-231 treated by 20 mm He μCAP at (a) 24 h incubation and (c) 48 h incubation. Cell viability of MDA-MB-231 treated by 60 mm He μCAP at (b) 24-h incubation and (d) 48-h incubation. The ratios of surviving cells for each cell line were calculated relative to controls (DMEM). Student’s t-test was performed, and the statistical significance compared to cells present in DMEM is indicated as *p < 0.05, **p < 0.01, ***p < 0.005 (n = 3).
Several methods, such as chemotherapy, surgery, and radiotherapy, are now being used for cancer treatment. The conventional methods have some disadvantages, such as longer treatment times, high cost, and adverse effects. However, plasma treatment may overcome the disadvantages of traditional treatments. Currently, plasma can be directly applied to skin cancers, but they are not useful for more systemic cancer treatment. However, we developed novel μCAP devices with 20 mm and 60 mm lengths, which can be considered as a local treatment tool. Plasma does not have the systemic therapeutic effects seen with chemical therapies. In addition, these devices do not have the limitations normally associated with plasmas. Overall, the previously discussed results indicate that μCAP devices with either a 20 mm or 60 mm length might be useful and should be considered in a clinical medical application.

IV. CONCLUSIONS

In this work, we showed that the newly developed micro-sized cold atmospheric plasma (μCAP) device with 20 mm and 60 mm length stainless steel tubes induce the formation of reactive species and radicals in culture medium. There is an increase in the concentration of O₂⁻, •OH, H₂O₂, and NO₂⁻ as a function of μCAP treatment time, which matches the trend of cell viability of two cancer cells with μCAP treatment time. A synergistic effect of short- and long-lived species present in the plasma treating DMEM is suspected to play a key role in cell death. Both the 20 and 60 mm length devices have significant effects on both U87 and MDA-MB-231 cancer cell viability, allowing access to both superficial and deeper tumors. The results of this study suggest a possibility for clinical applications of this μCAP device on brain and breast tumors. Future work will focus on the use of μCAP devices inside the patient’s body.

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