INTRODUCTION

According to the global cancer statistics, breast cancer (BC) is the most commonly diagnosed female cancer worldwide.1,2 It is a heterogeneous disease that can be classified into distinct subgroups: luminal A [estrogen receptor (ER)⁺ and/or progesterone (PR)⁺/human epidermal receptor 2 (HER2)⁻], luminal B (ER⁺ and/or PR⁺/HER2⁺), (ER⁻ and PR⁻/HER2⁺), or triple-negative (ER⁻, PR⁻, and...
HER2) subtypes according to the expression of particular clusters of co-expressed genes.\(^3\) Despite the breast cancer etiology has not been completely elucidated, it is thought to be multifactorial, with implication of both environmental and genetic factors.\(^4\)

The immune system serves as an important natural barrier to cancer development. Innate and adaptive responses are carefully orchestrated through soluble and membrane-bound receptors to eliminate precancerous cells and control neoplastic progression.\(^5\) Avoiding immune destruction has been considered a hallmark of cancer.\(^6,7\) It is therefore of great interest to identify immune genes that influence susceptibility to breast cancer.\(^8\)

One of the best characterized genes that play a crucial role in autoimmunity is the protein-coding autoimmune regulator (AIRE) gene (Gene ID: 326). This gene encodes a transcriptional regulator that plays an essential role to promote the negative selection of autoreactive T cells in the thymus by regulating the expression of a wide array of self-antigens that have the commonality of being tissue-restricted in their expression pattern in the periphery, called tissue-restricted antigens (https://www.genecards.org/cgi-bin/carddisp.pl?gene=AIRE). AIRE protein, in addition, was shown to have E3 ubiquitin ligase activity, which mediates proteasome pathway crucial for developing self-tolerance, cell cycle, DNA repair, and cell trafficking.\(^9\) Interestingly, not only is AIRE highly expressed in medullary thymic epithelial cells (mTECs), but there is an experimental evidence of its expression in peripheral lymphoid organs.\(^10-12\) Gardner et al, have reported the extra-thymic AIRE-expressing cells (eTACs) to be a distinct "bone marrow-derived antigen-presenting CD4\(^{low}\)" population that induce functional inactivation of effectors cells (CD4\(^+\)) T cells mediating tolerance through a mechanism involving deficient co-stimulation.\(^13\) Very recently, AIRE expression was reported in some solid cancers, including BC cells.\(^14-15\) Bianchi et al, for the first time, have detected an unexpected involvement of AIRE in the BC scenario.\(^15\) They reported AIRE expression in BC and an association with prognosis, which provides compelling evidence for the relevance of this unusual transcription factor as a new marker for BC.

Single nucleotide polymorphisms (SNPs) are the most common genetic variations, which influence inter-individual predisposition to breast carcinogenesis and prognosis.\(^16\) SNP association analysis has provided valuable information about the genetic susceptibility of BC.\(^17\) The human AIRE gene is located in the chromosome 21q22.3 region. Polymorphisms in this gene were previously shown to be genetically associated with autoimmune diseases, such as rheumatoid arthritis and autoimmune polyendocrine syndrome type 1 among others.\(^18,19\) The pathogenic variant rs2075876, caused by a point mutation (c.653-387G>A) in intron 5 (chr21:44289270), is located within exons forming the domain in the middle of protein chain, which functions in chromatin-dependent transcriptional regulation, nuclear localization, and oligomer formation.\(^20\)

As no case-control study of AIRE gene variant rs2075876 in sporadic BC has been reported and given the association of AIRE expression with breast cancer prognosis reported by Bianchi et al\(^{15}\) and confirmed by our in silico analysis (Figure 1), the authors were inspired to examine for the first time the possible association between rs2075876 variant and BC risk in a sample of Middle East population.

### 2 | MATERIALS AND METHODS

#### 2.1 | Study participants

In this case-control study, a total of 540 unrelated Egyptian women were enrolled (340 controls and 200 cases with primary BC). The cases were consecutively recruited from the Oncology Diagnostic Unit of Suez Canal University, Ismailia, Egypt. Patients had a histologically confirmed primary BC with relevant stage assigned according to the TNM classification of the Sixth Edition of the American Joint Committee on Cancer (AJCC) Manual for Staging of Cancer\(^21\) and graded by the Nottingham grading system.\(^22\) The participants who have pre-surgical treatment of chemotherapy, radiotherapy, endocrine therapy, or biotherapy were excluded. The unrelated healthy control group was randomly selected from the blood bank donors based on a negative history of any disease interfering with the study and has no family history of breast cancer or any other neoplastic diseases. All participants also had no history of any chronic diseases (eg, heart disease, renal problems, and diabetes mellitus). All relevant clinical and pathological information was revised and completed from the medical files including cancer type, grade and stage at the time of diagnosis, presence of lymph node metastases, and ER, PR, and HER2 expression in breast cancer tissue. The study was conducted in accordance with the guidelines in the Declaration of Helsinki and was approved by the ethics committee of the researchers’ university. The informed consent was obtained from all participants before taking part.

#### 2.2 | Blood sampling

Venous blood samples were collected from all participants under a complete aseptic condition on plain and trisodium ethylene-diamine-tetra-acetic acid (EDTA) (1 mg/mL) tubes. The former tubes were centrifuged at 2500 g for 30 minutes at 4°C, and the serum was divided into aliquots and kept frozen at −80°C until analysis of any required laboratory test that is not available in the patients’ medical records. The EDTA tubes were used for genotyping studies.

#### 2.3 | AIRE mutation analysis in cancer databases

The mutation spectrum of the AIRE gene in cancer was identified from The Cancer Genome Atlas (TCGA, https://cancergenome.nih.gov/), which contains genome maps from >30 cancer types. The role of the AIRE gene in cancer hallmarks was determined using Cancer Hallmarks Analytics Tool (CHAT) (http://chat.lionproject.net/), a web browser for classifying cancer-related text and text mining-associated biological processes from PubMed articles.\(^{23}\) The prognostic value of the AIRE gene in BC patients was explored in Kaplan-Meier
FIGURE 1  Prognostic role of AIRE gene in cancer. A, Alteration frequency of the AIRE gene in 32 pan-cancer studies in The Cancer Genome Atlas (TCGA). B, Type of somatic mutations and copy number variations. C, Enrichment of the AIRE gene in cancer hallmarks. D-G, Kaplan-Meier estimates for overall survival analysis in different molecular intrinsic subtypes.
Plotter (https://kmplot.com/analysis/), which includes gene chip and RNA-seq data from microarray and TCGA data. The following thirty-three datasets were included: GSE11121 (N = 200), GSE12093 (N = 136), GSE12276 (N = 204), GSE1456 (N = 159), GSE16391 (N = 55), GSE6446 (N = 120), GSE16716 (N = 47), GSE17705 (N = 196), GSE17907 (N = 54), GSE1915 (N = 77), GSE18728 (N = 61), GSE19615 (N = 115), GSE20194 (N = 45), GSE20271 (N = 96), GSE2034 (N = 286), GSE20685 (N = 327), GSE20711 (N = 90), GSE21653 (N = 240), GSE2063 (N = 99), GSE26971 (N = 276), GSE2990 (N = 102), GSE31448 (N = 71), GSE31519 (N = 67), GSE32646 (N = 115), GSE3494 (N = 251), GSE37946 (N = 41), GSE41998 (N = 279), GSE42568 (N = 121), GSE45255 (N = 139), GSE4611 (N = 153), GSE5327 (N = 58), GSE6532 (N = 82), and GSE7390 (N = 198). Multivariate analysis was performed with proportionate hazards assumption. Cutoff value for gene expression was set at the median, and the following variables were included in the model MK167, ESR1, and HER2 status. Next, stratified analysis by molecular subtyping was performed.

2.4 Allelic discrimination of rs2075876 G/A variant

Genomic DNA was extracted from the buffy coat using QIAamp DNA Blood Mini kit (Catalog No. 51 104; Qiagen) in accordance with the manufacturer’s instructions. NanoDrop ND-1000 (NanoDrop Technologies, Inc) was used for assessment of DNA concentration and purity. Genotyping of AIRE gene rs2075876 (G/A) variant was identified using StepOne™ Real-Time Polymerase Chain Reaction (Applied Biosystems, USA). PCRs were run in a total volume of 20 µL containing genomic DNA (20 ng) with nuclease-free water up to 9 µL, TaqMan SNP Genotyping Assay Mix (1 µL) (Applied Biosystems, assay ID C_15863141_20), TaqMan Genotyping PCR Master Mix (10-µL).

The thermocycler conditions were followed: denaturation at 95°C for 10 minutes, followed by 40 cycles of 90°C for 15 seconds and 60°C for 1 minutes. Genotyping was conducted blind to the case/control status. Ten percent of the samples were randomly selected and re-genotyped with 100% reproducibility in genotyping recall (Figure 2).

2.5 Statistical analysis

Data were analyzed using SPSS (IBM SPSS Statistics for Windows, version 26.0., IBM Corp.). Chi-square and Fisher’s exact tests were used for qualitative variables. Allele and genotype frequencies were calculated as previously described. The Hardy-Weinberg equilibrium (HWE) was estimated using the Online Encyclopedia for Genetic Epidemiology (OEGE) software (http://www.oeg.org/software/hwe-mr-calc.shtml). Adjusted odds ratios (ORs) with a 95% confidence interval (CI) using logistic regression models were calculated for multiple genetic association models. The study power estimation using G power-3 software (http://www.gpower.hhu.de/) showed that with the specified case-control study design and allowable error rate (alpha error = 0.05), the specified sample size for each group can give 99% study power. A two-tailed P-value of .05 was considered statistically significant.

3 RESULTS

3.1 Baseline characteristics of the study population

The mean age of controls was 46.1 ± 10.8 and that of BC patients was 45.3 ± 12.3 (P = .557). Demographic, clinical, and pathological characteristics are represented in Table 1.
**Table 1** Demographic and clinical characteristics of breast cancer patients

| Characteristics                  | n   | %   |
|----------------------------------|-----|-----|
| **Risk factors**                 |     |     |
| FH of cancer                     |     |     |
| Negative                         | 140 | 70.0|
| Positive                         | 60  | 30.0|
| Degree of relation               |     |     |
| First degree                     | 28  | 14.0|
| Second degree                    | 24  | 12.0|
| Third degree                     | 8   | 4.0 |
| Menarche                         |     |     |
| Late                             | 72  | 36.0|
| Early                            | 128 | 64.0|
| Menopause                        |     |     |
| Early                            | 90  | 45.0|
| Late                             | 10  | 5.0 |
| OCP usage                        |     |     |
| Negative                         | 160 | 80.0|
| Positive                         | 40  | 20.0|
| Parity                           |     |     |
| Multipara                        | 172 | 86.0|
| Nullipara                        | 28  | 14.0|
| Late first gravida               |     |     |
| Negative                         | 192 | 96.0|
| Positive                         | 8   | 4.0 |
| **Clinical presentation**        |     |     |
| Symptoms                         |     |     |
| Mastalgia                        | 62  | 31.0|
| Breast mass                      | 164 | 82.0|
| Skin lesions                     | 32  | 16.0|
| Nipple change                    | 36  | 18.0|
| Axillary pain                    | 14  | 7.0 |
| Axillary mass                    | 36  | 18.0|
| Side                             |     |     |
| Left                             | 74  | 37.0|
| Right                            | 383 | 191.5|
| Site                             |     |     |
| LOQ                              | 18  | 9.0 |
| UIQ                              | 38  | 19.0|
| UOQ                              | 96  | 48.0|
| Retro                            | 48  | 24.0|
| No of masses                     |     |     |
| Single                           | 152 | 76.0|
| Multiple                         | 48  | 24.0|

**Table 1** (Continued)

| Characteristics                 | n   | %   |
|---------------------------------|-----|-----|
| **Pathological analysis**       |     |     |
| HPD                             |     |     |
| Ductal                          | 76  | 38.0|
| Lobular                         | 52  | 26.0|
| Medullary                       | 28  | 14.0|
| Mucinous                        | 20  | 10.0|
| Tubular                         | 12  | 6.0 |
| Metaplastic                     | 12  | 6.0 |
| Grade                           |     |     |
| G2                              | 128 | 64.0|
| G3                              | 72  | 36.0|
| LVI                             |     |     |
| Negative                        | 104 | 52.0|
| Positive                        | 96  | 48.0|
| T stage                         |     |     |
| T2                              | 100 | 50.0|
| T3                              | 44  | 22.0|
| T4B                             | 40  | 20.0|
| T4D                             | 16  | 8.0 |
| N stage                         |     |     |
| N0                              | 56  | 28.0|
| N1                              | 68  | 34.0|
| N2                              | 64  | 32.0|
| N3                              | 12  | 6.0 |
| M stage                         |     |     |
| M0                              | 88  | 44.0|
| M1                              | 84  | 42.0|
| Mx                              | 28  | 14.0|
| Site of metastasis              |     |     |
| Bone                            | 36  | 18.0|
| Liver                           | 12  | 6.0 |
| Lung                            | 30  | 15.0|
| Skin infiltration               |     |     |
| Negative                        | 160 | 80.0|
| Positive                        | 40  | 20.0|
| Receptor status                 |     |     |
| ER/PR                           |     |     |
| Negative                        | 80  | 40.0|
| Positive                        | 120 | 60.0|
| HER2+                           |     |     |
| Negative                        | 164 | 82.0|
| Positive                        | 36  | 18.0|

(Continues)
3.2 | Prognostic value of AIRE from the cancer databases

Screening 10 953 cancer patients in thirty-two TCGA pan-cancer studies revealed an alteration of the AIRE gene in 1.5% of cases (Figure 1A). Somatic mutations included amplification, deep deletions, fusion events, and mutations in various types of cancer (Figure 1B). According to CHAT database, AIRE is enriched in immune destruction, genome instability and mutation, resisting cell death, sustaining proliferative signaling, and tumor-promoting inflammation (Figure 1C). KM survival curves demonstrated an association of AIRE expression level with prognosis in triple-negative basal-like molecular subtype of BC (HR = 2.44, 95% CI = 1.44-4.15, log-rank P-value < .001) (Figure 1G).

3.3 | Genotyping of AIRE polymorphism

AIRE gene variant rs2075876 is located within intron 5 and caused by substitution of G to A (c.653-387G>A). Genotype frequencies of rs2075876 were deviated from HWE in the study population (ie, a marked decrease in the heterozygous frequency of the overall study population was observed; hence, the observed genotype frequency was not in line with the expected one) (Supplementary Table S1). MAF (A allele) accounted for 0.28 in controls. According to 1000 Genome Project (https://www.ncbi.nlm.nih.gov/variation/tools/1000genomes/), this figure was significantly different from that of Africans (MAF = 0.35) and East Asians (MAF = 0.37). On comparing between patients and controls, G/G homozygote was significantly more frequent among patients (94%) compared to controls (66%) (P < .001). In contrast, the other homozygote A/A was less represented among cases, accounting for 6% of cases, while heterozygote genotype G/A was absent in patients (Figure 3).

3.4 | Genetic association model for disease risk assessment

After adjusting confounding variables, individuals with A allele conferred protection against developing BC under allelic model (OR = 0.33, 95% CI = 0.20-0.55), recessive model (OR = 0.25, 95% CI = 0.10-0.60), dominant model (OR = 0.12, 95% CI = 0.05-0.29), and homozygote comparison (OR = 0.20, 95% CI = 0.08-0.50) (Figure 3D). Given the low frequency of A/A in patients (6%) and absence of heterozygote genotype, multiple regression analysis was not feasible. Table 2 shows no significant associations between the homozygous minor allele AIRE variant (A/A) with the clinicopathological features of BC patients.

4 | DISCUSSION

Given the newly emerged implication of several AIRE gene variations in multiple cancers (Figure 1B), and as there is no association analysis of the study AIRE variant with BC risk has been documented, this study aimed to investigate whether AIRE rs2075876 G>A polymorphism is associated with the development of BC in our population. Here, we identified that the study variant is associated with BC susceptibility under several genetic association models.

As a transcriptional and post-transcriptional regulator in medullary thymic epithelial cells and peripheral tissues, AIRE plays a substantial role in “thymocyte education” and “negative selection” by controlling
the peripheral antigen expression in thymus.\textsuperscript{28} Accumulating evidence indicated that AIRE protein is a non-conventional transcriptional factor as it does not bind directly to any DNA motifs; however, it is implied in releasing the stalled RNA polymerase II at the promoter regions of the target genes to induce gene expression.\textsuperscript{29} Our in silico analysis revealed that this transcriptional regulator can significantly interact with several protein partners implied in many physiological/pathological cellular processes with high confidence level (Figure 4).\textsuperscript{30} These proteins, including the AIRE, have been enriched in nucleosome assembly, chromatin silencing, interleukin-7-mediated signaling pathway, telomere organization, regulation of immune system process, response to stress and stimulus,
regulation of transcription and RNA metabolic process, and positive regulation of cell growth among other identified processes (Table S2). KEGG (Kyoto Encyclopedia of Genes and Genomes) enrichment analysis also revealed that AIRE is involved in “transcriptional misregulation in cancer” pathway (hsa05202) (Table S2). Interestingly, all the above-mentioned pathways and processes were identified to play essential roles in BC etiology, progression, and/or outcome, which add evidence to AIRE implication in the BC.

Several AIRE variants were previously shown to be associated with many autoimmune diseases, including rheumatoid

| Characteristics | A/A (n = 12) | G/G (n = 188) | P value |
|-----------------|-------------|--------------|---------|
| Age             | Mean ± SD   | 48.0 ± 15.2  | 45.1 ± 12.3 | .27    |
| FH              | 8 (66.7)    | 172 (70.2)   | .85     |
|                 | 4 (33.3)    | 56 (29.8)    |         |
| Skin infiltration | No         | 10 (83.3)    | 150 (79.8) | .83    |
|                 | Yes         | 2 (16.7)     | 38 (20.2)  |         |
| Metastasis      | M0          | 4 (33.3)     | 84 (44.7)  | .86    |
|                 | M1          | 6 (50)       | 78 (41.5)  |         |
|                 | Mx          | 2 (16.7)     | 26 (13.8)  |         |
| Clinical stage  | IIA         | 2 (16.7)     | 34 (18.1)  | .88    |
|                 | IIB         | 0 (0)        | 28 (14.9)  |         |
|                 | IIIA        | 2 (16.7)     | 22 (11.7)  |         |
|                 | IIIB        | 2 (16.7)     | 26 (13.8)  |         |
|                 | IV          | 6 (50)       | 78 (41.5)  |         |
| NPI             | Good        | 4 (33.3)     | 96 (51.1)  | .67    |
|                 | Poor        | 8 (66.7)     | 92 (48.9)  |         |
| ESMO            | Low risk    | 4 (33.3)     | 72 (38.3)  | .80    |
|                 | High risk   | 8 (66.7)     | 116 (61.7) |         |
| ER/PR           | Negative    | 2 (16.7)     | 78 (41.5)  | .39    |
|                 | Positive    | 10 (83.3)    | 110 (58.5) |         |
| HER2+           | Negative    | 10 (83.3)    | 154 (81.9) | .93    |
|                 | Positive    | 2 (16.7)     | 34 (18.1)  |         |
| Molecular subtype | Luminal A     | 8 (66.7)     | 88 (46.8)  | .67    |
|                 | Luminal B   | 2 (16.7)     | 22 (11.7)  |         |
|                 | HER2+       | 0 (0)        | 12 (6.4)   |         |
|                 | TNBC        | 2 (16.7)     | 66 (35.1)  | .67    |
| IHP             | Good        | 10 (83.3)    | 110 (58.5) | .37    |
|                 | Moderate    | 2 (16.7)     | 66 (35.1)  |         |
|                 | Poor        | 0 (0)        | 12 (6.4)   |         |
| Recurrence      | Negative    | 4 (33.3)     | 92 (48.9)  | .45    |
|                 | Positive    | 8 (66.7)     | 96 (51.1)  |         |
| Survival        | Alive       | 8 (66.7)     | 156 (83)   | .29    |
|                 | Dead        | 4 (33.3)     | 32 (17)    |         |
| DFS             | Mean ± SD   | 12.5 ± 7.8   | 14.3 ± 5.8 | .45    |
| OS              | Mean ± SD   | 16.0 ± 5.4   | 16.5 ± 4.2 | .75    |

Note: Data are presented as mean ± SD or number (percentage). The t test and two-sided chi-square tests were used.

Abbreviations: FH, family history; NPI, Nottingham Prognostic Index; calculated as [0.2 × tumor size in cm] + tumor grade [1-3] + lymph node stage [1-3, according to stages A-C]; ESMO, European Society of Medical Oncology; ER/PR, estrogen and progesterone receptors; HER2+, positive for human epidermal growth factor receptor 2/neu receptor; IHP, Immunohistochemical Prognostic Index estimated based on the three receptor status (HER2, ER, and PR); DFS, disease-free survival; OS, overall survival.

| TABLE 2 | Association between clinicopathological characteristics of BC patients and AIRE rs2075876 G>A variant homozygous genotypes |
arthritis, vitiligo, systemic sclerosis, Addison disease, systemic lupus erythematosus, and alopecia areata. However, a limited number of association studies for rs2075876 G>A polymorphism were identified. Furthermore, no previous study explored the relationship of the specified study variant with the BC.

As rs2075876 G>A is an intronic variant with low pathogenicity prediction score (6.657) based on the online CADD PHRED score prediction tool for single or combined variants, it is difficult to explain the exact biological function of this polymorphism. However, the non-coding variants at the transcription level may disrupt the transcription factor binding sites, the splice sites, and other functional sites as proposed by Chen et al and supported recently by Conteduca et al. According to the GEO (Gene Expression Omnibus) database mining, Terao et al revealed a statistically significant correlation between the risk allele (A) of rs2075876 and AIRE down-regulation. Furthermore, the specified variant may be in linkage disequilibrium with distinct genetic markers, which synergistically influence disease/cancer risk that deserve future functional studies to evaluate the role of this putative variant in disease risk.

It is worth noting that despite the genotype frequencies of the study variant deviated from the HWE, no standard guidelines for rejecting SNPs that depart from HWE have been developed. In practice, all variants for which HWE P-values decrease below a predetermined threshold should be checked manually for genotyping quality. As the current genotyping of this study was based on a high-quality real-time TaqMan allelic discrimination PCR assay with genotype recall rate (100%), and the specified quality control measurements were applied throughout the methodology, the current variant departure from HWE suggested a violation of HWE assumptions in the study groups rather than technical genotyping errors.

Although this study has enough power to detect the association of the specified polymorphism with BC risk, some potential limitations merit careful consideration: first, as the patients and controls recruited from one center may not be representative of the general population. Second, with a deficit in the number of observed heterozygotes as well as the presence of few cases carrying A/A genotype, this could potentially affect HWE congruence. Thus, our findings should be confirmed by large-scale multicenter studies in different populations. Third, the authors have to confirm that despite there was a significant association with BC risk conferred by an individual locus, this might have a small impact on the disease risk as genes act multiplicatively. However, this small impact could be included in more advanced multiplicative polygenic risk scores. Individually, these risk alleles confer a very small increase in BC risk but their joint effect with other lifestyle risk factors could “improve risk-reduction and screening strategies by targeting those most likely to benefit” as concluded by Lakeman et al. Fourth, the underlying mechanisms of the implication of this SNP in BC were not investigated. Further functional analyses are required to explore the molecular mechanisms by which this variant could participate in BC susceptibility.

In conclusion, our study provided evidence that rs2075876 G>A variant of the AIRE gene may contribute to BC susceptibility. It is worth noting that in combination with other genetic and non-genetic data, this variant might be helpful in refining the BC patients profile. The efficiency of population-based preventive programs such as screening mammography could be improved in a near future by targeting women who are at the greatest risk for BC according to their genotyping profile and other environmental risk factors.

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CONFLICT OF INTEREST
The authors declare that they have no conflict of interest.

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SUPPORTING INFORMATION
Additional supporting information may be found online in the Supporting Information section.

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