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Humoral and cell-mediated response against SARS-CoV-2 variants elicited by mRNA vaccine BNT162b2 in healthcare workers: a longitudinal observational study

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ABSTRACT
Objectives: To assess the humoral and cell-mediated response to severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) elicited by the mRNA BNT162b2 vaccine in SARS-CoV-2-experienced and -naive subjects against a reference strain and SARS-CoV-2 variants.

Methods: The humoral response (including neutralizing antibodies) and T-cell-mediated response elicited by BNT162b2 vaccine in 145 healthcare workers (both naive and positive for previous SARS-CoV-2 infection) were evaluated. In a subset of subjects, the effect of SARS-CoV-2 variants on antibody level and cell-mediated response was also investigated.

Results: Overall, 125/127 naive subjects (98.4%) developed both neutralizing antibodies and specific T cells after the second dose of vaccine. Moreover, the antibody and T-cell responses were effective against viral variants since SARS-CoV-2 NT Abs were still detectable in 55/68 (80.9%) and 25/29 (86.2%) naive subjects when sera were challenged against β and δ variants, respectively. T-cell response was less affected, with no significant difference in the frequency of responders (p 0.369). Of note, two doses of vaccine were able to elicit sustained neutralizing antibody activity against all the SARS-CoV-2 variants tested in SARS-CoV-2-experienced subjects.

Conclusions: BNT162b2 vaccine elicited a sustained humoral and cell-mediated response in immunocompetent subjects after two-dose administration of the vaccine, and the response seemed to be less affected by SARS-CoV-2 variants, the only exceptions being the β and δ variants. Increased immunogenicity, also against SARS-CoV-2 variant strains, was observed in SARS-CoV-2-experienced subjects. These results suggest that triple exposure to SARS-CoV-2 antigens might be proposed as valuable strategy for vaccination campaigns. Irene Cassaniti, Clin Microbiol Infect 2022;28:301.e1–301.e8 © 2021 European Society of Clinical Microbiology and Infectious Diseases. Published by Elsevier Ltd. All rights reserved.
Introduction

The mRNA BNT162b2 vaccine [1], the first authorized for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection, showed 95% protection against SARS-CoV-2 infection in a phase II/III trial [2]. Another mRNA-based vaccine, mRNA-1273 [3], showed a similar effect. However, data on the kinetics of the immune response elicited by the vaccines are limited to low numbers of analysed subjects, and limited mainly to antibody responses [1,4–8].

The T-cell response elicited by the vaccine may have a crucial role in the long-term protection against SARS-CoV-2 infection and disease. In convalescent subjects, T- and B-cell memory specific for SARS-CoV-2 was found to persist for at least 6–8 months [9–11].

The emergence of new SARS-CoV-2 variants with mutations in the spike (S) protein has raised significant concerns about vaccine efficacy and reinfection risk in previously infected subjects. The new variant 501Y.V1 lineage B.1.1.7 (UK variant or α) includes multiple mutations in both the receptor binding domain (RBD) and the N-terminal domain of the S protein [12], and the 501Y.V3 lineage F.1 (Brazilian, BZ or γ) [13] and the 501Y.V2 lineage B.1.351 (South African, SAF, or β) variants have mutations in the S protein and, especially, in the RBD [14]. More recently, a lineage B.1.617.2 (δ variant) has emerged.

Preliminary data have suggested that convalescent sera and sera from vaccinated individuals efficiently neutralized the α variant, while a reduction in neutralizing (NT) antibody titres has been observed against the β variant [15,16].

In the present study we evaluated humoral and cell-mediated responses elicited by the BNT162b2 vaccine in subjects previously exposed to SARS-CoV-2 and in naive subjects. Moreover, we aimed to define the level of both antibody and cell-mediated responses against the emerging SARS-CoV-2 variants after vaccination.

Methods

We designed an observational, longitudinal, prospective study to evaluate the immune response elicited by the BNT162b2 vaccine against SARS-CoV-2 in 145 healthcare workers (median age 44 years, range 21–69) who received vaccination between 27th December 2020 and 11th February 2021; of these, 127 were SARS-CoV-2-naïve and 18 were SARS-CoV-2-experienced before vaccination. All the subjects were enrolled at Fondazione IRCCS Policlinico San Matteo (Pavia, Italy). The efficacy endpoints were the development of SARS-CoV-2-specific neutralizing antibodies and a T-cell response. Analyses were performed at baseline (before vaccination), at the time of the second vaccine administration (T1), and 21 days after the second vaccine dose (T2).

Antibody response was determined using the chemiluminescent assay Elecsys Anti-SARS-CoV-2 S (Roche Diagnostics Rotkreuz, Switzerland), which provides quantitative measures of antibody (mainly IgG) specific for SARS-CoV-2 RBD. Results are given as units (U)/mL and are considered positive when ≥0.8 U/mL. Moreover, SARS-CoV-2 neutralizing antibodies were quantified using a homemade assay and results higher than 1:10 were considered positive. IgG against RBD of the wild-type (WT) and European (EU, which share the same RBD), α and β strains were determined by ELISA using recombinant proteins.

The SARS-CoV-2-specific T-cell response was quantified by ex-vivo ELISpot assay, and results >10 spot-forming units (SFU)/million peripheral-blood mononuclear cells (PBMCs) were given as positive. Further methodological details are reported in the Supplementary Material.

Results

Antibody response in SARS-CoV-2-naïve and -experienced vaccinated subjects

At T1, all the 18 SARS-CoV-2-experienced subjects showed anti-RBD antibody levels above the upper limit of the quantifiable range of the assay, while 122/127 SARS-CoV-2-naïve subjects (96.1%) developed anti-RBD antibodies although at significantly lower levels than the experienced subjects did. Moreover, levels of anti-RBD antibodies in naïve subjects at T1 were significantly lower (p < 0.009) than the baseline levels of experienced subjects.

At T2, all the SARS-CoV-2-naïve subjects developed a positive anti-RBD response; however, the median anti-RBD level was still significantly higher in SARS-CoV-2-experienced subjects than in naïve ones (p < 0.001) (Fig. 1a).

A similar trend was observed for NT antibodies (Fig. 1b). Serum NT titres were significantly higher in SARS-CoV-2-experienced subjects than in naïve ones at any time point.

SARS-CoV-2-specific T-cell response

All but three SARS-CoV-2-naïve subjects (15/18, 83.3%) showed S-specific T cells at baseline, and levels of S-specific T cells increased at T1 (p < 0.001) and had not increased further at T2 (p 0.510).

Among SARS-CoV-2-naive subjects the percentage of responders was 69.3% at T1 and 98.4% at T2. Levels of S-specific T cells (Fig. 2a) significantly increased from T1 to T2 (p < 0.001); even if the level of S-specific T cells at T2 in experienced subjects was higher than that observed in naive subjects, the difference was not statistically significant (p 0.095). No significant correlation was observed between age and S-specific T-cell response at both T1 (r =0.02, 95%CI −0.21 to 0.17, p 0.8214) and T2 (r =−0.17, 95%CI −0.32 to 0.04, p 0.107). Levels of N-specific T cells did not change with time in either experienced or naive subjects (Fig. 2b). Of note, 30% of SARS-CoV-2-naive subjects showed a detectable SARS-CoV-2-specific T-cell response against S and 18.2% against N at baseline. Data on phenotypical characterization of the T-cell response are shown in the Supplementary Material Figs S1 and S2.

At T1, the overall percentage of full responders (i.e. subjects developing both SARS-CoV2 S-specific T cells and NT antibody) was 77.8% (95%CI 70.1–84.0), corresponding to 100% of experienced and 68.8% of naive subjects (95%CI 60.2–76.3). After T2, the overall percentage of full responders was 98.6% (95%CI 95.0–99.8), corresponding to 100% of experienced and 98.4% of naive subjects (95%CI 94.4–99.7).

Antibody and T-cell response against SARS-CoV-2 variants

Sera collected at T2 from 31 SARS-CoV-2-naïve subjects were tested against RBD of the D614 WT Chinese-derived strain, and the α variant and β variant strains. Median RBD-specific reciprocal antibody titres were 5838 (IQR 2675–16 351) for WT (and EU) strains, 3220 (801–9263) for α and 60.5 (<50–196) for β strains, respectively (Fig. 3a). A reduction of approximately 50% and 99% of RBD-specific antibody titres was observed from WT to α and β strains, respectively (Supplementary Material Fig. S3a,b).

Sera collected at T2 from 61 naïve and 16 experienced subjects were tested against the WT strain by neutralization assay and challenged against EU strain, α, β, and γ variants (Fig. 3b). Additionally, sera at T2 from 29 naïve and 16 SARS-CoV-2-experienced
Subjects were challenged against the δ variant. Among the 61 SARS-CoV-2-naive subjects, an increase in four-fold dilution for SARS-CoV-2 NT antibodies was observed against the EU strain with respect to the WT strain (p < 0.001) (Supplementary Material Fig. S3d), while the level of SARS-CoV-2 NT antibodies was not affected when sera were tested against α and γ strains.

![Graph](image-url)
A five-fold and 20-fold reduction in median titre of SARS-CoV-2 NT antibodies was observed in response to the \( \beta \) strain in comparison to the WT virus and EU strain, respectively (\( p < 0.001 \)) (Supplementary Material Fig. S3g). However, although at lower levels, NT antibodies against the \( \beta \) variant were still detectable in 55/68 subjects (80.9%). Similarly, a decrease of five-fold and 20-fold of median titre of SARS-CoV-2 NT antibodies was observed in response to the \( \delta \) strain.

Fig. 2. Cell-mediated immune response against spike (S) and nucleocapsid (N) peptide pools in 145 vaccinated subjects—127 naive (seronegative) for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection and 18 who had experienced SARS-CoV-2 infection (seropositive)—at baseline (day of first dose administration, T0), at day 21 (day of second dose administration, T1) and at day 42 (21 days after the second dose administration, T2). The dotted horizontal line represents a positive cut-off for SARS-CoV-2 T-cell response.
in comparison to the WT and EU strains (p < 0.001), overall, the response was positive for the δ variant in 25/29 naive subjects (86.2%).

Among the 16 SARS-CoV-2-experienced subjects, a sustained NT level against the all tested variants was observed. The median reduction in SARS-CoV-2 NT against the β variant was four-fold with respect to the WT (p 0.008), and no reduction was observed when sera were challenged against the other variants.

In addition, sera from six vaccinated subjects, five of them naive for SARS-CoV-2 infection, were used to assess the NT level also against the C.36 and B.1.525 variants. No significant difference was observed between SARS-CoV-2 NT titre against WT, α, β and Thai variants (p 0.999). Similar to what was observed for the β variant, a six-fold decrease in NT level was documented against the B.1.525 strain (p 0.081, Supplementary Material Fig. S4). Of note, no correlation was observed between RBD level and NT antibody titre for any variant (Supplementary Material Fig. S5).

In 36 vaccinated subjects (53%), residual PBMCs isolated at T2 were challenged against inactivated virus preparations from the supernatant of Vero E6 cells infected with five SARS-CoV-2 strains (WT, EU, α, β and γ) and the cell-mediated response was analysed by ELISpot assay. The ELISpot assay was less sensitive using WT
**Fig. 4.** (a) Median T-cell response against severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) variants. (b) Numbers of subjects with a positive SARS-CoV-2 T-cell response and a negative SARS-CoV-2 T-cell response.
virus than peptide pools from the homologous S protein. However, even if a slightly lower level of cell-mediated T-cell response was observed against α, β, and γ variants (Fig. 4a), SARS-CoV-2-specific T cells were detected in the large majority of the subjects, with no significant difference in the frequency of responders. In detail, 34/36 (94.4%) were positive for WT SARS-CoV-2 T-cell response and 33/36 (91.7%) were positive for EU SARS-CoV-2 T-cell response; α, β, and γ variant-specific T-cell response was detectable in 30/36 (83.3%), 29/36 (80.5%), and 31/36 (86.1%) vaccinated subjects (Fig. 4b), indicating that the BNT162b2 vaccine was able to elicit T cells able to recognize conserved epitopes from any virus variant tested.

Discussion

In this study, we reported that almost all vaccinated healthy subjects developed a sustained cellular and humoral response after complete vaccination. Moreover, the response level was higher in those subjects with previous exposure to SARS-CoV-2.

SARS-CoV-2-naive individuals developed a humoral response higher than that observed after natural infection after two vaccine doses, but not after one vaccine dose, when about 6% of individuals were still negative for SARS-CoV-2 NT antibodies. On the other hand, the cell-mediated response was detectable in 98.4% naive individuals after two doses of vaccine, whereas one vaccine dose was not sufficient to elicit a detectable SARS-CoV-2 T-cell response in 32.1% of subjects.

Conversely, both humoral and cellular responses were strongly induced after the first dose of the BNT162b2 vaccine in SARS-CoV-2-experienced subjects; therefore, it appears that the prime vaccine dose acts as a boost in these subjects. Moreover, both antibody and T-cell responses did not appear to increase after the second dose in these individuals. According to previous observations [17,18], this evidence strengthens the concept that just one vaccine dose may be sufficient in SARS-CoV-2-experienced subjects [19]. The peptides chosen for T-cell stimulation activated both CD4+ and CD8+ T-cell responses, since they were 15 amino acids long with 10–11 overlap. The peptide pools spanned the entire protein code of the wild-type strain.

Levels of antibody and, especially, cell-mediated response after one vaccine dose was weak or undetectable in SARS-CoV-2-naive subjects. This contrasts with what was observed in a previous report analysing the antibody response in a small number of subjects [17], and suggests that only one dose was not sufficient to elicit complete protection in naive subjects. Furthermore, in these subjects the emergence of viral resistance or escape might be increased if a suboptimal immune response is elicited. Thus far, a single vaccine dose can be unfavourable in SARS-CoV-2-naive subjects.

Only β and δ variants significantly impacted neutralizing antibody levels in vaccinated subjects, while SARS-CoV-2 T-cell response was not significantly affected. NT antibody titres are extremely variable after SARS-CoV-2 infection [20]. Those with low titres could be reinfected, and due to the presence of incomplete protective neutralizing antibodies might develop escape variants [21,22]. The accumulation of mutations in RBD and the N-terminal domain of spike protein might be associated with increased escape from neutralization. Mutation at the 484 site has been related to a significant decrease in neutralization titre against the β variant was observed in plasma from convalescent subjects [25]. Usually, after natural infection or vaccination, polyclonal immune responses arise against multiple antigenic epitopes. Consequently, small numbers of variations in antigen sequence should have little impact on recognition by the immune system, including both NT antibodies and T cells.

Our results show that vaccinated individuals developed an equally effective NT antibody response against the WT, α, and γ variants, while the NT response against the EU variant was increased. On the other hand, sera from vaccinated subjects showed a significant reduction in SARS-CoV-2 NT antibodies when challenged against the β and δ variants. Nevertheless, serum from the majority of vaccinated subjects (almost 80%) maintained a neutralizing activity against these variants that could be effective in preventing the development of severe disease. These results are in contrast to those of another report showing a higher reduction in neutralization titres against pseudoviruses with S protein from α, β, and γ variants, with respect to WT, and no increase in NT against the European variant [26]. Although the binding titre to β RBD was highly reduced with respect to WT (about 100-fold), the NT titre was less affected, showing a five-fold reduction. In addition, we observed that anti-RBD antibody levels, measured in vaccinated subjects with either commercial or in-house assays, have a poor correlation with NT titres. This may suggest that anti-S antibodies binding to sites outside the RBD, which are less involved by the mutations reported, could contribute to virus neutralization. Importantly, our studies rely on the use of wild-type viruses rather than pseudoviruses, which likely could give discrepant results with respect to natural strains [27] because the artificial lentiviral particles cannot resemble the complete biology of actual clinical isolates.

The level of NT antibodies against SARS-CoV-2 variants in SARS-CoV-experienced vaccinated subjects was robust. This preliminary observation might be helpful in the design of vaccination strategies, suggesting that three exposures to SARS-CoV-2 Spike antigen could increase the level of neutralizing antibodies against SARS-CoV-2 variants.

However, and most considerably, our studies on cell-mediated response revealed that SARS-CoV-2 T-cell response is minimally affected by the mutations occurring in SARS-CoV-2 variants, as reported [28], and most subjects tested showed a detectable T-cell response against each virus strain. Unlike Tarke and colleagues, we used inactivated preparations of different SARS-CoV-2 variant isolates in order to recapitulate realistically antigen presenting mechanisms and natural epitope recognition from T cells. The levels of T-cell response and NT antibodies elicited by the vaccine were not correlated; therefore, a low NT titre does not exclude the presence of a protective T-cell response able to control SARS-CoV-2 infection and avoid the development of severe disease.

Potentially cross-reactive T cells originally elicited by the common cold coronaviruses may be boosted by the vaccine and contribute to protection. In our study, about 20–30% of SARS-CoV-2-naive subjects showed a T-cell response specific for S or N proteins. This might be related to the presence of cross-reactive epitopes from common cold coronaviruses that circulate seasonally [11,29–32].

Our study has some limitations. First, the number of SARS-CoV-2-experienced subjects was much lower than that of SARS-CoV-2-naive subjects, and elderly people were not represented. Second, neutralizing and T-cell responses against variants were tested only in a subset of subjects.

In conclusion, the BNT162b2 vaccine appears able to induce robust antibody and T-cell responses in all immunocompetent individuals and paves the way for the clinical evaluation of other mRNA vaccines currently in development [33]. Although the effectiveness of the antibody response may be partially reduced by some mutations naturally arising in virus strains, the T-cell response appears to be marginally affected. A sustained immune response against SARS-CoV-2 variants was observed in vaccinated
SARS-CoV-2–experienced subjects, thus suggesting that a triple exposure to SARS-CoV-2 antigen might represent a valuable strategy for vaccination campaigns.

Transparency declaration

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Author contributions

IC and DL analysed and interpreted the data and drafted the manuscript. IC, FeB and EG collected and managed the data. FeB, EG, KMGA, FZ and GC performed experiments on T-cell response. EP, JCS, AF, PZ and AS performed experiments on antibody response. EP performed virus isolation and production. AP sequenced the viral genome. AR and VZ enrolled the participants. FM and FN collected samples with SARS-CoV-2 variants. LS and LV produced recombinant RBD. DL wrote the protocol. FaB supervised the project and revised the manuscript. All the authors critically reviewed the manuscript.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.cmi.2021.09.016.

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