Procyanidins are potent inhibitors of LOX-1: 
a new player in the French Paradox

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Abstract: Lectin-like oxidized LDL receptor-1 (LOX-1) is an endothelial receptor for oxidized LDL (oxLDL) and plays multiple roles in the development of cardiovascular diseases. We screened more than 400 foodstuff extracts for identifying materials that inhibit oxLDL binding to LOX-1. Results showed that 52 extracts inhibited LOX-1 by more than 70% in cell-free assays. Subsequent cell-based assays revealed that a variety of foodstuffs known to be rich in procyanidins such as grape seed extracts and apple polyphenols, potently inhibited oxLDL uptake in Chinese hamster ovary (CHO) cells expressing LOX-1. Indeed, purified procyanidins significantly inhibited oxLDL binding to LOX-1 while other ingredients of apple polyphenols did not. Moreover, chronic administration of oligomeric procyanidins suppressed lipid accumulation in vascular wall in hypertensive rats fed with high fat diet. These results suggest that procyanidins are LOX-1 inhibitors and LOX-1 inhibition might be a possible underlying mechanism of the well-known vascular protective effects of red wine, the French Paradox.

Keywords: LOX-1, cardiovascular diseases, lipid accumulation, procyanidin, French Paradox

Introduction

Atherosclerosis is a leading burden of cardiovascular diseases such as coronary artery diseases and stroke. It is a chronic, multifactorial disease of vascular wall involving lipid accumulation, inflammation, cell death, and thrombosis. The changes in the vascular wall, which precede by far earlier the appearance of coronary artery diseases.1)2) Environmental factors such as diet and cigarette smoking together with such pathological conditions as hyperlipidemia, hypertension and diabetes greatly influence the disease progression over time. Therefore, lifestyle changes such as diet control and smoking cessation are strongly encouraged in patients with high risk of cardiovascular diseases.

Lectin-like oxidized receptor-1 (LOX-1) was originally identified as an endothelial receptor for oxidized LDL (oxLDL), a critical player involved in the initiation of atherosclerosis.3) OxLDL binds to LOX-1 and increases expressions of adhesion molecules and inflammatory cytokines, and decreases nitric oxide release, resulting in endothelial dysfunction.4)–6) LOX-1 also recognizes other ligands such as platelets and C-reactive protein, and mediates platelet adhesion and vascular hyperpermeability, respectively.7),8) LOX-1 is expressed not only in endothelial cells but in macrophages, vascular smooth muscle cells, platelets and cardiomyocytes; and the expression is highly induced under atherogenic settings such as hyperlipidemia, hypertension and diabetes.9)–11) In vivo evidence has also demonstrated that LOX-1 contributes to the initiation and development of a wide range of cardiovascular diseases. For instance, LOX-1 gene-deficient mice display less atherosclerotic lesions on high fat diet and less myocardial injury after ischemia-reperfusion.12),13) Conversely, LOX-1 overexpressing mice

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display increased atheroma-like lesions and impaired endothelium-dependent vasorelaxation on high fat diet.\(^{14,15}\)

Importantly, we have recently demonstrated that high LOX Index, which is calculated by multiplying circulating concentrations of soluble LOX-1 and LOX-1 ligand LDL, associates with an increased risk of coronary heart diseases and stroke in Japanese population.\(^{16}\) These lines of evidence suggest that inhibition of LOX-1 could be a strategy for the prevention and/or treatment of cardiovascular diseases.

Although statin therapy has made a success to reduce the risk of cardiovascular events, multiple lines of evidence have suggested that daily intake of certain foods or beverages could be an effective strategy to prevent the development of cardiovascular diseases. For instance, fish oil and red wine have been long postulated to possess cardioprotective actions.\(^{17,18}\) Moreover, a case-control study involving participants from 52 countries reported an inverse association between the risk of myocardial infarction and intake of prudent diet (high in fruit and vegetables).\(^{19}\) However, the molecular targets and/or the active ingredients of those foods and beverages are largely unknown although a number of health food supplements are available in the market. This study was undertaken to identify materials that inhibit oxLDL binding to LOX-1 from foodstuff extracts.

**Materials and methods**

**Preparation of lipoproteins.** Serum was isolated from healthy volunteers and LDL (density: 1.019–1.063 g/mL) was prepared by sequential ultracentrifugation. Isolated LDL was oxidized with 7.5 µM CuSO\(_4\) for 16 h and labeled with 1,1-dioctadecyl-3,3,3,3-tetramethylindocarbocyanine perchlorate (DiI, Invitrogen, Carlsbad, CA, USA). Cell-associated DiI-oxLDL was determined as fluorescence analysis using the IN Cell analyzer system (GE Healthcare, Fairfield, CT, USA). The binding of DiI-oxLDL to LOX-1-CHO cells were determined similarly to the uptake assay, except that the incubation of DiI-oxLDL was performed at 4°C for 45 min. The samples were subjected to fluorescence microscopic analysis (Axiostar 200M, Zeiss, Oberkochen, Germany) or quantitative fluorescence analysis using the IN Cell analyzer system (GE Healthcare, Fairfield, CT, USA). Cell-associated DiI-oxLDL was determined as a ratio of DiI-oxLDL fluorescence intensity per cell in the presence of foodstuff extracts to that in the absence of extracts. All experiments were conducted in more than triplicates. For the determination of fluorescence quenching by procyanidins, the fluorescence was also measured before washing out unbound DiI-oxLDL (Spectramax, Molecular Devices, Sunnyvale, CA, USA). The final concentration of DMSO was less than 0.1% of total volume.

**Secondary screening in CHO cells expressing LOX-1.** Tetracycline-inducible human LOX-1 (tagged with V5-6×His at C-terminus) expressing CHO-K1 (LOX-1-CHO) cells were maintained as previously described.\(^{8}\) The cells were seeded in 96-well plate at 10^4 cells/well in the presence of doxycycline (1 µg/mL) (Calbiochem, La Jolla, CA, USA) and were incubated in Ham’s F-12 medium containing 10% FBS at 37°C for 24 h. After being washed with the medium without FBS, the cells were treated with foodstuff extracts or an anti-LOX-1 antibody at the final concentration of 10 µg/mL for 1 h. The cells were washed again and incubated with DiI-oxLDL (10 µg/mL) for 2 h. After washing, the cells were fixed with 10% formalin, and were stained with 4’,6-diamidino-2-phenylindole (DAPI) (Sigma, St. Louis, MO, USA). The binding of DiI-oxLDL to LOX-1-CHO cells were determined similarly to the uptake assay, except that the incubation of DiI-oxLDL was performed at 4°C for 45 min. The samples were subjected to fluorescence microscopic analysis (Axiovert 200M, Zeiss, Oberkochen, Germany) or quantitative fluorescence analysis using the IN Cell analyzer system (GE Healthcare, Fairfield, CT, USA). Cell-associated DiI-oxLDL was determined as a ratio of DiI-oxLDL fluorescence intensity per cell in the presence of foodstuff extracts to that in the absence of extracts. All experiments were conducted in more than triplicates. For the determination of fluorescence quenching by procyanidins, the fluorescence was also measured before washing out unbound DiI-oxLDL (Spectramax, Molecular Devices, Sunnyvale, CA, USA). The final concentration of DMSO was less than 0.1% of total volume.

**Animal study.** All protocols were approved by the Institutional Animal Care and Use Committee of the National Cerebral and Cardiovascular Center. Animals were individually housed at 23 ± 2°C with 12 h light-dark cycles (7:00–19:00 light-on). Eight-week-old male stroke-prone spontaneously hypertensive rats (SHR-SP) (CLEA Japan, Tokyo, Japan)
were divided into two groups (n = 8 each) with similar blood pressure, heart rates and body weights (tail cuff and pulse transducer system, BP-98A, Softron, Tokyo, Japan). They were given high fat diet without cholic acid or vitamin E (CLEA Japan, Tokyo, Japan) and physiological saline as drinking water for 2 weeks. Half of the rats received physiological saline containing 0.5% (w/v) oligomeric procyanidins (OPC) with various chain lengths purified from apples. At the end of the study, the rats were euthanized with isoflurane inhalation and blood was collected from vena cava between 11:00–14:00. The rats were perfused systemically with physiological saline for Oil Red O staining. Total cholesterol, high-density lipoprotein (HDL) cholesterol, triglyceride, and thiobarbituric acid reactive substance (TBARS) were determined enzymatically using commercially available kits (Wako Pure Chemical Industries, Osaka, Japan; Zeptometrix corporation, Buffalo, NY, USA). In a separate experiment, rats (n = 4) were given 0.5% OPC for two days and similarly sacrificed between 7:00–8:00 for the determination of plasma concentrations of epicatechin (4/3-8)epicatechin (procyanidin B2) and epicatechin (4/3-8)epicatechin (4/3-8)epicatechin (procyanidin C1) as previously reported.

Oil Red O staining of accumulated lipid in mesenteric arteries. Oil Red O staining was used for the detection of lipid accumulation in mesenteric arteries as previously reported. In brief, the intestine was excised out and the enteric canal was removed. Branches of the mesenteric artery were isolated from adipose tissue and fixed with 10% formalin. The mesenteric arteries were incubated in 60% isopropanol for five minutes followed by staining with 0.18% Oil Red O (Merek KGaA, Darmstadt, Germany). Lipid deposits were manually enumerated under stereomicroscope in the posterior 8 branches of the artery (Stemi 2000, Carl Zeiss, Oberkochen, Germany).

Anti-LOX-1 antibody. A monclonal antibody against human LOX-1 (TS92) was used for blocking ligand binding to LOX-1. Characteristics of the antibody were described in previous reports.

Statistics. One-way ANOVA followed by Dunnett or Student’s t-test was performed for multiple or paired comparison, respectively. P < 0.05 was considered statistically significant and indicated by *.

Results

Screening. 472 products were investigated to identify LOX-1 inhibitors. They included 437 food-stuff extracts and 35 test reagents such as amino acids. Almost all the products were commercially available and many of them have been utilized for healthcare as folk medicine or health food supplements. The assayed products were categorized as shown in Fig. 1.

First we employed a sandwich ELISA using oxLDL as a ligand for LOX-1. Results showed that 52 products inhibited oxLDL binding by more than 70% (data not shown). The products were further examined using a cell-based assay whether they inhibited oxLDL uptake in LOX-1-CHO cells. Control CHO cells did not uptake DiI-labeled oxLDL (DiI-oxLDL) at detectable levels, indicating the specific uptake of DiI-oxLDL by LOX-1 (data not shown). In LOX-1-CHO cells, we found 26 products that inhibited DiI-oxLDL uptake by more than 70% (Table 1). These included a number of foodstuff extracts that are known to contain a large amount of procyanidins such as grape seed extracts, pine bark extracts, peanuts skin and apple polyphenols (highlighted in bold, Table 1). We considered the possibility that procyanidins may inhibit oxLDL binding to LOX-1 and conducted the subsequent studies.

Characterization of apple polyphenols. In the next series of experiments, we characterized the profiles of apple polyphenols as a representative containing a large amount of procyanidins. Apple polyphenols dose-dependently and completely inhibited the DiI-oxLDL (10 µg/mL) binding to LOX-1-CHO cells with an IC50 value of 102 ng/mL (Fig. 2A). As apple polyphenol contains catechin/epicatechin ((epi)catechin), phenolcarboxylic acids, and other ingredients as well as procyanidins (Fig. 2B), we examined which of the ingredients inhibited DiI-oxLDL binding to LOX-1. As shown in Fig. 2C, the fractions containing procyanidins and (epi)catechin significantly inhibited the DiI-oxLDL binding while phenolcarboxylic acids and others did not. These results clearly indicated that procyanidins and/or (epi)catechin were active ingredients for the inhibition of oxLDL binding to LOX-1. Neither apple polyphenol nor procyanidins displayed cytotoxic effects up to 10 µg/mL (data not shown).

In vitro profiles of procyanidins. We next analyzed the activities of procyanidins according to the polymerization levels. (Epi)catechin and oligomeric procyanidins from dimer to heptamer were separated from apple polyphenols as previously reported. Purified procyanidins were used for the analysis. The results showed that procyanidins of
dimer displayed a concentration-dependent inhibition of DiI-oxLDL (10 µg/mL) binding to LOX-1-CHO cells. In contrast, (epi)catechin was without effect up to 1 µg/mL (Fig. 3). Moreover, the efficacy was much more potent with procyanidin of ≥trimer than dimer procyanidin. The IC₅₀ of dimer procyanidin was 330 ng/mL and that of oligomeric procyanidins ranged from 61 ng/mL (trimer) to 27 ng/mL (heptamer). Oligomeric procyanidins with chain length of more than eight also similarly inhibited oxLDL binding (data not shown). These results clearly indicated that oligomeric procyanidins inhibited oxLDL binding to LOX-1.

It has been reported that trimer apple procyanidins comprises six isoforms. In the next study, we prepared major four isomers of apple procyanidins, epicatechin(4-O-8)epicatechin(4-O-8)epicatechin (known as procyanidin C1), epicatechin(4-O-8)epicatechin(4-O-8)catechin, epicatechin(4-O-6)epicatechin(4-O-8)catechin and epicatechin(4-O-6)epicatechin(4-O-8)epicatechin, and examined their activities. As shown in Fig. 4A, the fluorescence of DiI-oxLDL (10 µg/mL) in LOX-1-CHO cells was significantly attenuated by procyanidin C1 (300 ng/mL). The fluorescence represents the specific binding to LOX-1 since it was disrupted by anti-LOX-1 antibody (LOX-1 Ab). These results indicated that procyanidin C1 inhibited oxLDL binding to LOX-1. Quantitative analysis showed that all of the trimer procyanidins similarly inhibited DiI-oxLDL binding to LOX-1-CHO cells with IC₅₀ of 51, 41, 47 and 73 ng/mL, respectively (Fig. 4B). The fluorescence quenching of DiI-oxLDL by procyanidin C1 (300 ng/mL) was less than 25% (data not shown).
shown). The IC50 of procyanidin C1 decreased to 7 ng/mL and 26 ng/mL when DiI-oxLDL was used at the concentrations of 1 and 3 µg/mL, respectively (data not shown).

**In vivo effects of oligomeric procyanidins.**

Finally, we addressed whether procyanidins inhibited lipid accumulation in vascular wall in vivo. Oligomeric procyanidins (OPC), a mixture containing procyanidins with various chain lengths purified from apple, was used for *in vivo* study. SHR-SP rats are well-known disease-model animals for hypertension and stroke, and accumulate lipids in arterial wall in response to high fat diet.22,24 In this study, 8 week old SHR-SP rats were given high fat diet and physiological saline containing 0.5% OPC (OPC rats) for 2 weeks. OPC did not affect the daily food intake and body weight during the experimental period. Systolic and diastolic blood pressure and heart rates were also unaffected (Table 2). At termination, the mesenteric artery isolated from the control rats displayed numerous Oil Red O-positive dots while that from the OPC-treated rats displayed significantly less dots, indicating that OPC suppressed arterial lipid deposition (Fig. 5). Plasma total cholesterol (TC), HDL-cholesterol, triglyceride and TBARS did not differ between the groups (Table 2). The plasma concentrations of procyanidin B2 and C1 in OPC rats were 7.9 ± 1.8 ng/mL and 4.9 ± 1.9 ng/mL, respectively.

**Discussion**

We successfully identified several foodstuff extracts that inhibit LOX-1 using ELISA and DiI-oxLDL uptake assay. The results obtained with the cell assays suggested that procyanidins might be one of the components that inhibit oxLDL uptake since nearly half of the potent hit extracts are known to contain a large amount of procyanidins. Indeed, purified procyanidins inhibited oxLDL binding in LOX-1-CHO cells. Moreover, OPC suppressed lipid accumulation in vascular wall of SHR-SP rats in which an anti-LOX-1 antibody was also effective.22 To date, several natural products with cardioprotective effects

| Extract/Product Name       | Inhibition (%) |
|----------------------------|----------------|
| Black Soybean Hull         | 99             |
| Propolis                   | 98             |
| Oligomeric Proanthocyanidins from Grape Seed*1 | 98 |
| Grape Seed*2               | 98             |
| Peanut Seed Coat*3         | 98             |
| Apple Condensed Tannin     | 97             |
| Grape Seed*4               | 97             |
| Peanut Seed Coat*5         | 97             |
| French Maritime Pine Bark  | 96             |
| Proanthocyanidins          | 95             |
| New Zealand Pine Bark      | 95             |
| Grape Seed*6               | 91             |
| Apple Polyphenol           | 88             |
| Pomegranate Skin           | 86             |
| Roselle Tea                | 86             |
| Mangosteen Fruit Skin      | 84             |
| Wasabi Leaf                | 83             |
| Gingko Leaf                | 80             |
| Bilberry*7                 | 80             |
| Bilberry*8                 | 78             |
| Guava Leaf                 | 76             |
| Vitis Coignetiae           | 76             |
| Quercus Salicina           | 76             |
| Pine Bark                  | 75             |
| Hop Polyphenol             | 73             |
| Cat’s Claw                 | 72             |

*1Kikkoman, *2Sun Globe Food, *3Kishimoto Sanyo, *4Hoechst Marion Roussel, *5Tokiwa Phytochemical, *6Chika Industries, *7Indena Japan, *8Tokiwa Phytochemical.
Fig. 4. DiI-oxLDL (10 µg/mL) binding in LOX-1-CHO cells in the presence of various isomers of trimer procyanidins. Fluorescent images of DiI-oxLDL (red) binding to LOX-1-CHO cells in the absence or presence of procyanidin C1 (300 ng/mL) or in the presence of anti-LOX-1 antibody (A). Nuclei were stained by DAPI (blue). DiI-oxLDL binding in LOX-1-CHO cells in the presence of various concentrations of trimer procyanidin isomers (B). Chemical structures of the trimer procyanidin isomers used in the experiments are shown. Difference in the hydroxyl groups indicated by red jagged lines causes stereoisomers of catechin and epicatechin.
have been identified to reduce the biological actions of oxLDL and/or the expression of LOX-1 such as flavonoids from seabuckthorn, bergamot oil and mulberry leaf aqueous fractions. However, their molecular targets are not known. To our knowledge, this is the first report demonstrating a direct antagonistic action of natural products to LOX-1.

LOX-1-inhibiting properties were almost identical among procyanidins ≥ trimer and the dimer also potently inhibited LOX-1. Moreover, four different isomers of trimer procyanidins almost equally inhibited oxLDL binding to LOX-1. These results implicate that intake of procyanidin-rich foods potentially inhibits LOX-1, regardless of food source since the polymerization levels of procyanidins significantly differ among foods. In support to this, OPC, a mixture of various chain lengths of procyanidins, inhibited vascular lipid accumulation in SHR-SP rats. We have previously demonstrated that an anti-LOX-1 antibody inhibited vascular lipid accumulation in this animal model, suggesting that procyanidins function as a LOX-1 inhibitor in vivo as well. A wide range of in vivo effects of procyanidin-

Table 2. Body weight, cardiovascular parameters, food intake and plasma parameters of SHR-SP rats before and after the administration of procyanidins (OPC).

| Parameters                  | Control Before | Control After | OPC Before | OPC After |
|-----------------------------|----------------|----------------|------------|-----------|
| Body Weight (g)             | 198 ± 5        | 247 ± 4        | 199 ± 5    | 237 ± 4   |
| Systolic Blood Pressure (mmHg) | 173 ± 7       | 174 ± 4       | 191 ± 6    | 197 ± 3   |
| Diastolic Blood Pressure (mmHg) | 119 ± 6       | 121 ± 3       | 137 ± 3    | 142 ± 4   |
| Heart Rate (/min)           | 351 ± 7        | 364 ± 8       | 348 ± 5    | 364 ± 7   |
| Food Intake (g/day)         | 18.0 ± 0.4     | 17.1 ± 0.3    | 18.0 ± 0.4 | 17.1 ± 0.3 |
| Plasma Triglyceride (mg/dL) | 404.8 ± 45.3   | 378.2 ± 46.1  | 404.8 ± 45.3 | 378.2 ± 46.1 |
| Total Cholesterol (mg/dL)   | 157.6 ± 8.4    | 152.3 ± 6.4   | 157.6 ± 8.4 | 152.3 ± 6.4 |
| HDL-cholesterol (mg/dL)     | 36.1 ± 1.6     | 35.6 ± 0.9    | 36.1 ± 1.6 | 35.6 ± 0.9 |
| TBARS (nmol/mL)             | 0.89 ± 0.05    | 0.84 ± 0.04   | 0.89 ± 0.05 | 0.84 ± 0.04 |

Fig. 5. Arterial lipid deposition in SHR-SP rats on high fat diet. Oil Red-O image (A) and the number of lipid deposition (B) in control and OPC rats. * represents statistical significance from control.
rich extracts such as anti-oxidative, anti-hypertensive and anti-hyperlipidemic actions have been reported.\textsuperscript{29–32} These actions potentially reduce vascular lipid accumulation, since in this animal model vascular lipid accumulation is increased by high fat feeding and decreased by the treatment with anti-oxidant or hypotensive agents\textsuperscript{22} (unpublished observation). It may be less likely, however, since OPC did not affect cardiovascular parameters, plasma lipids, or TBARS, a marker of oxidative state, in the present study. Different experimental conditions such as dosage and experimental period may have affected the readouts. Shoji \textit{et al.} have demonstrated that procyanidin B2 and C1 are orally available.\textsuperscript{20} In agreement to this, we were able to detect procyanidin B2 and C1 in the plasma of rats treated with orally administered OPC. However, the plasma levels of procyanidins were small and close to the detection limits. These results may argue against that OPC suppressed vascular lipid accumulation by inhibiting LOX-1 on vascular wall. Yet, the plasma level may be sufficient to inhibit LOX-1 since plasma oxLDL level in SHR-SP rats is about 200 ng/mL\textsuperscript{22} and procyanidin C1 halved the oxLDL binding in LOX-1-CHO cells even at 7 ng/mL\textsuperscript{22} when 1 µg/mL oxLDL was used. Alternatively, plasma procyanidin levels may have reached higher when rats actively intake OPC during light-off period. It is also possible that OPC may inhibit the function of LOX-1 expressed on the apical membrane of intestinal epithelium, in which LOX-1 mediates the transcytosis of bile salt-dependent lipase.\textsuperscript{33} Since intestinal lumen is exposed to a significant amount of OPC during the absorption process, OPC may have inhibited intestinal LOX-1 and thereby modulated lipid metabolism leading to less lipid accumulation. Further studies are definitely called for to reach positive evidences to determine whether OPC inhibits LOX-1 \textit{in vivo} or other unknown factors might be involved.

Out of more than 400 foodstuff extracts derived from various sources, more than half of those displaying potent LOX-1 inhibition are known to contain a large amount of procyanidin. In addition, not well-known compared to apples or grape seeds, procyanidins are also contained in other extracts listed in Table 1 such as pomegranate skin,\textsuperscript{34} mangosteen fruit skin,\textsuperscript{35} bilberry,\textsuperscript{36} Quercus salicina\textsuperscript{37} and cat’s claw.\textsuperscript{38} These results implicate that LOX-1 inhibition may underlie the health benefits of these extracts. Of particular interest, procyanidins as well as resveratrol are considered to be one of the bioactive ingredients in red wine for the cardioprotective effects, known as “French Paradox.”\textsuperscript{39–41} The present findings implicate that LOX-1 is a molecular target of procyanidins as is sirtuin 1 for resveratrol. Furthermore, Corder \textit{et al.} identified procyanidins from red wine as the molecule that suppressed endothelin-1 release from vascular endothelial cells.\textsuperscript{42,43} They also found that the concentrations of procyanidins were high in wines from certain geographic areas, which correlated with longevity in such regions.\textsuperscript{42,43} As LOX-1 activation leads to endothelin-1 release from endothelial cells,\textsuperscript{44} these results may further support the hypothesis that LOX-1 inhibition is an underlying mechanism of the cardiovascular protective effects of red wine.

In conclusion, we demonstrate that LOX-1-inhibiting property is rich in foodstuffs containing a large amount of procyanidins and that procyanidins are potent inhibitors of LOX-1. These results implicate the role of LOX-1 in the French Paradox. Moreover, the LOX-1 ELISA and CHO cell assays employed in this study are likely to make powerful methods to identify LOX-1 inhibitors not only from foodstuffs but also from chemical compounds for future drug discovery.

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