A Smart and Responsive Crystalline Porous Organic Cage Membrane with Switchable Pore Apertures for Graded Molecular Sieving

Citation for published version:
He, A, Jiang, Z, Wu, Y, Hussain, H, Rawle, J, Briggs, ME, Little, MA, Livingston, AG & Cooper, AI 2022, 'A Smart and Responsive Crystalline Porous Organic Cage Membrane with Switchable Pore Apertures for Graded Molecular Sieving', Nature Materials, vol. 21, no. 4, pp. 463-470. https://doi.org/10.1038/s41563-021-01168-z

Digital Object Identifier (DOI):
10.1038/s41563-021-01168-z

Link:
Link to publication record in Heriot-Watt Research Portal

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Nature Materials

Publisher Rights Statement:
© The Author(s) 2021.

General rights
Copyright for the publications made accessible via Heriot-Watt Research Portal is retained by the author(s) and/or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
Heriot-Watt University has made every reasonable effort to ensure that the content in Heriot-Watt Research Portal complies with UK legislation. If you believe that the public display of this file breaches copyright please contact open.access@hw.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
A smart and responsive crystalline porous organic cage membrane with switchable pore apertures for graded molecular sieving

Ai He1,6, Zhiwei Jiang2,3,6, Yue Wu1, Hadeel Hussain4, Jonathan Rawle4, Michael E. Briggs1, Marc A. Little1, Andrew G. Livingston2,3 and Andrew I. Cooper1,5,✉

Membranes with high selectivity offer an attractive route to molecular separations, where technologies such as distillation and chromatography are energy intensive. However, it remains challenging to fine tune the structure and porosity in membranes, particularly to separate molecules of similar size. Here, we report a process for producing composite membranes that comprise crystalline porous organic cage films fabricated by interfacial synthesis on a polyacrylonitrile support. These membranes exhibit ultrafast solvent permeance and high rejection of organic dyes with molecular weights over 600 g mol⁻¹. The crystalline cage film is dynamic, and its pore aperture can be switched in methanol to generate larger pores that provide increased methanol permeance and higher molecular weight cut-offs (1,400 g mol⁻¹). By varying the water/methanol ratio, the film can be switched between two phases that have different selectivities, such that a single, ‘smart’ crystalline membrane can perform graded molecular sieving. We exemplify this by separating three organic dyes in a single-stage, single-membrane process.

Porous organic cages (POCs)1,2 are discrete molecules with intrinsic cavities that can create porosity in molecular crystals3, amorphous solids1 and porous liquids4. The adsorption properties of POCs can sometimes be predicted in silico from knowledge of their molecular structures in isolation1,6. However, the adsorption properties of POC materials are also affected by their solid-state packing2,7. For example, extrinsic pores in POC crystals can selectively adsorb guests, including rare gases1. Indeed, inefficient packing of POCs can generate solids with considerably more porosity than would be expected from the cage cavities alone1–7. This combination of intrinsic and extrinsic porosity determines the functionality of POC-based materials in selective adsorption processes.

Most separation studies involving POCs have used molecular crystals3,5, which can exhibit slow adsorption kinetics. Also, many POC crystals rely on selective adsorption governed by thermodynamics, rather than kinetics, which limits their practical use in size- and shape-selective membrane filters. Given their solution processability, however, there is scope to develop crystalline POC-based membranes that operate by selectively removing guests that are either too large or that have the wrong shape to diffuse through the POC pore structure.

There is growing interest in membrane technologies that perform industrial and environmentally relevant separations where two or more solutes are separated one from each other, as in distillation or chromatography, as opposed to separations where a whole set of solutes is concentrated, such as in evaporation or seawater reverse osmosis8–13. A major advantage of membranes is that they can perform separations in the liquid phase, which is often more practically useful than vapour phase separations.

Membranes for liquid separations are typically produced using phase inversion, which can be followed by coating14 or interfacial polymerization15. This produces amorphous polymer networks with a modest degree of pore tunability. There is a strong demand to develop membranes with more tunable and modular pore structures. Various porous solids, including zeolites16, POCs1,2, organic polymers1, metal–organic frameworks18, covalent organic frameworks (COFs)19 and hydrogen-bonded organic frameworks20 have been explored. Banerjee et al. reported COF films with 1.4 to 2.6 nm pores that showed good performance in dye rejection21. Dichtel et al. reported COF films with 3.4 nm pores and tunable thicknesses over the range of 100 μm to 2.5 nm that rejected Rhodamine WT from water22. The same team also reduced the effective pore size of their COF membrane to 3.3 and 3.2 nm using reticular chemistry23. In addition to COFs19,23, metal–organic frameworks and their composites have been used to produce membranes24,26. However, it remains challenging to produce continuous nanofiltration membranes with extended porous frameworks that perform exclusively as size-based molecular sieves rather than selective adsorbents27. POCs are solution processable and their solid-state structures are defined by non-covalent intermolecular interactions, which can be switched using chemical stimuli to alter their bulk porosity28,29. As such, POCs are intriguing but relatively unexplored candidates for new types of membrane materials30–37.

Many practically important molecular separations involve ternary systems or more complex mixtures—for example, separating multiple hydrocarbon fractions from light crude oil by distillation, pervaporation or organic solvent reverse osmosis38,39; purification of fatty acids40,41, such as the practical recovery of omega-3 polyunsaturated fatty acids from fish oil by nanofiltration42; or sieving out by-products from reactions, for example in the liquid-phase peptide synthesis of pharmaceuticals43. To achieve equivalent...
separations for complex mixtures using membranes, cascades of membranes with graded molecular weight cut-offs (MWCOs) have been developed12, using phase inversion (polymeric membranes)13 or sol–gel processing (ceramic membranes)14 by manipulating the recipe for dope solution or fabrication conditions to produce multiple membranes with a variety of pore sizes. This places membranes at a disadvantage for ternary and higher separations—by contrast, a single distillation or chromatography column can produce multiple fractions with differing compositions. Separating more than binary solute systems using a membrane cascade requires multiple pumped recycle streams and complex fluid controls15. While solvent gradients are used in chromatography to modulate solid–liquid interactions, to the best of our knowledge, there are as yet no reports of membranes that respond to solvent gradients by changing their solute selectivity.

Here, we report the fabrication of close-packed and defect-free films of a shape-persistent imine POC, CC3, which grow at the liquid–liquid interface between water and dichloromethane (Fig. 1a). These films comprise highly crystalline domains of CC3 in its most thermodynamically stable polymorph, CC3α (Fig. 1b). By coating the CC3α film on polyacrylonitrile (PAN), we produce a continuous membrane (CC3α–PAN) that has excellent permeance for both polar and non-polar solvents, including water (43.0 l m⁻² h⁻¹ bar⁻¹) and toluene (55.9 l m⁻² h⁻¹ bar⁻¹). Furthermore, we found that it is possible to rapidly and reversibly switch the membrane pore aperture using common solvents. Exposure of the non-covalent crystal packing of CC3 in methanol (MeOH) induces a rapid phase transition from CC3α to a different crystalline phase, CC3γ′, which is less densely packed. This systematically increases the effective pore aperture of the resulting membrane, CC3γ′–PAN (Fig. 1c). This switching property of the CC3γ′–PAN membrane allows the permeation of larger organic dyes that can be rejected in water while the large pore apertures are turned ‘off’ in the CC3α–PAN membrane. This switchable porosity is reversible, and surprisingly, it does not compromise the continuity of the membrane. This allowed us to separate three organic dyes with different sizes via graded sieving using a single membrane.

Fig. 1 | Synthesis of a crystalline CC3 film and its crystal structures. a, Scheme showing the interfacial synthesis method used to fabricate crystalline CC3 films, which were subsequently adhered to a PAN support. These crystalline cage films can cycle between two different forms, CC3α–PAN and CC3γ′–PAN, by cycling the solvent between water and MeOH. CH2Cl2, dichloromethane. b, CC3α structure with its 3D pore network shown in yellow. c, The CC3γ′ structure, formed by soaking in MeOH, has additional extrinsic solvent-filled channels, shown here in orange, that open up additional porosity in the membrane in response to the MeOH solvent.

Fabrication of crystalline CC3 films
Continuous films with highly crystalline domains of CC3 were produced using a combined interfacial condensation reaction and crystallization process at a water–dichloromethane interface (Fig. 1a). This interfacial process allows the two-component reaction of CC3, which is synthesized via a [4+6] cycloimination reaction using 1,3,5-triformylbenzene (TFB) and (1R,2R)-1,2-diaminocyclohexane (CHDA), while simultaneously directing the formation of CC3 films at the interface (Methods). Continuous and free-standing CC3 films were transferred from the liquid–liquid interface onto various substrates (for example, glass, steel mesh, carbon tape and silicon wafers; Supplementary Fig. 1) for further analysis of the crystallinity and surface morphology. Before performing permeance and dye rejection studies, the CC3 film was coated onto a PAN support by filtration to form the composite membrane (Fig. 2a). The resulting membrane, referred to hereafter as CC3–PAN, was free of macroscopic defects up to at least 7.4 cm in diameter using this preparation process, with no evidence of delamination after cutting the membrane into smaller pieces (Supplementary Fig. 2). The CC3 film was characterized by Fourier transform infrared spectroscopy (Supplementary Fig. 3), Raman spectroscopy, nuclear magnetic resonance (NMR) spectroscopy (Supplementary Fig. 4), scanning electron microscopy (SEM), focused ion beam SEM (FIB-SEM), X-ray diffraction and atomic force microscopy (AFM). For spectroscopic measurements, a crystalline CC3α sample was used as a reference1. CC3α has a three-dimensional (3D) diamondoid pore structure and is the thermodynamically most stable polymorph CC3 (ref. 1). A Raman map was performed on an 80 × 80 μm² CC3 film deposited on glass (Fig. 2e and Supplementary Fig. 5), which indicated that the CC3 film comprised crystalline domains with the same solid-state structure as the CC3α polymorph (Fig. 2i). SEM images showed a continuous, apparently defect-free film in the CC3–PAN composite (Fig. 2b and Supplementary Fig. 6a) with a thickness of ~80 nm measured on a free-standing film (Fig. 2d), which contained embedded, octahedral CC3 crystals (Supplementary Fig. 7). Cross-sectional SEM images were obtained after step-by-step FIB trenching and polishing of both CC3–PAN (Supplementary Fig. 8)
and a CC3 film coated on a silicon wafer (Supplementary Fig. 9). The FIB-SEM images showed a clear boundary between the CC3 film layered on top of the supports. After transferring the as-synthesized CC3 film onto a silicon wafer, we performed AFM measurements to investigate the film thickness further. AFM again confirmed that the CC3 film was continuous with a constant thickness of ~80 nm (Fig. 2c and Supplementary Figs. 6b, 8 and 9).

A key advantage of interfacial synthesis is that it can create continuous films of the product\(^{15,21}\). Here, we also modified the reaction conditions to optimize the thickness, continuity and crystallinity of the CC3 film (Fig. 2g,h). This allowed us to create CC3 films from the interfacial reaction that were four times thinner than the CC3 film created by spin coating\(^{40}\). To further confirm the crystalline structure of the film, we performed a series of powder X-ray diffraction (PXRD) and grazing incidence X-ray diffraction (GIXRD) measurements on CC3-PAN (Methods). These diffraction measurements revealed that the CC3 film was crystalline and had the same structure as CC3 \(\alpha\) (Supplementary Figs. 10 and 11).

To further investigate the crystallization process of CC3 films at the solvent interface, we varied the reaction time from 4 to 96 hours.
The properties of CC3α-PAN-24 h-0.8%, referred to hereafter as CC3α-PAN.

Membrane performance of CC3α-PAN
To determine the permeance and dye rejection performance of CC3α-PAN, we performed filtration experiments in dead-end cells using solvents and dyes with different sizes and chemical functionalities (Supplementary Table 1 and Supplementary Figs. 22 and 23).

With a water contact angle of 94° (Supplementary Fig. 24), the CC3α-PAN membrane was stable in a range of polar and non-polar solvents (Supplementary Fig. 25), proving that these solvents do not dissolve CC3. This led to ultrafast solvent permeances (Fig. 3a; Supplementary Fig. 26 for blank PAN data). We attribute this to the 3D interconnected porosity through the CC3α crystals in the film. By comparison, an amorphous CC3 membrane prepared by spin coating (Supplementary Section 1.2) exhibited a 30-fold lower solvent permeance under the same testing conditions (Supplementary Fig. 27), although it should also be noted that the amorphous CC3 membrane was four times thicker[20]. Apparently, the crystalline CC3α-PAN provides sufficient robustness to support the interconnected channels under high applied pressures. To further confirm the importance of crystallinity, CC3α membranes with different crystallinity levels were fabricated at each of the four reaction stages simply by controlling the reaction time. A partially crystalline membrane (CC3α-PAN-8 h-0.8%) at Stage 2 exhibited a water permeance of 3.01 m⁻² h⁻¹ bar⁻¹; that is, an order of magnitude lower than the fully crystalline Stage 3 membrane (49.51 m⁻² h⁻¹ bar⁻¹ for...
**Fig. 4 | X-ray diffraction characterization and switchable separation performance of CC3-PAN membranes.**

**a,** GIXRD of CC3α-PAN in air and water, and CC3γ'-PAN in MeOH. Experimental PXRD patterns of CC3α and CC3γ' powders are included as references. **b,** MWCO curve for CC3α-PAN in water and CC3γ'-PAN in MeOH containing 20 ppm dye solutes. The MWCO was determined by interpolating from the plot of rejection against the molecular weight of the dyes and corresponds to the molecular weight for which rejection reaches 90%. All error bars depict the s.d. of the data points obtained from at least three independent membranes. **c,** Reversible dye rejection of BB and solvent permeance of the CC3-PAN membrane observed upon switching the feedstock solvent between water and MeOH. All error bars denote the s.d. for measurements from at least three independent membranes. **d,** Photographs of CC3-PAN filtration dead-end cell captured from Supplementary Video 1 during the different cycles; BB is rejected in water by CC3α-PAN while CC3γ'-PAN does not reject BB in MeOH. **e,** In situ GIXRD patterns showing the reversible phase transition between CC3α-PAN and CC3γ'-PAN, by cycling between water and MeOH. **f,** Acetone permeance versus MWCO of general solutes in acetone for nanofiltration membranes reported in the literature and CC3α-PAN. MOF, metal–organic framework; MC, macrocycle; NPs, nanoparticles; GO, graphene oxide; PTMSp, poly(1-(trimethylsilyl)-1-propyne); PA, polyamide; PEI, polyethyleneimine; PANi, polyaniline; PI, polyimide; PE, polyethylene; PEEK, poly(ether ether ketone); PEO, poly(ethylene oxide); PIM, polymers of intrinsic microporosity; PIP, pipierazine; PAR, polyacrylate (Supplementary Table 9 for full details).

CC3α-PAN-48 h-0.8%) prepared with prolonged reaction times (Fig. 3b). CC3α-PAN-8 h-0.8% and CC3α-PAN-48 h-0.8% exhibited the same MWCO, as determined by filtering a range of dyes through the membranes (Fig. 3c). By comparison, amorphous oligomeric membranes produced in Stage 1 (CC3α-PAN-4 h-0.8%) and cracked, highly crystalline membranes produced in Stage 4.
(CC3α-PAN-96 h-0.8%) exhibited unexpectedly higher water permeances but failed to achieve comparable separation performances, indicating that they contained physical defects.

Two limitations of membranes produced from other crystalline porous materials, such as COFs, are poor stability at high pressures\(^{21}\) and the interference of adsorption processes\(^{27}\). Here, the CC3α-PAN membrane was tested under a range of applied pressures, up to a maximum of 35 bar. The water flux increased linearly with increasing applied pressure (Fig. 3d) without affecting the MWCO (Fig. 3e). Longer duration studies demonstrated the mechanical robustness of CC3α-PAN and showed consistent dye rejection (99.7% for rose bengal) and water permeance (~43 l m\(^{-2}\) h\(^{-1}\) bar\(^{-1}\)) over 20 hours (Supplementary Fig. 28). The applied pressure of 35 bar is an order of magnitude higher than that used for liquid filtration through COF membranes\(^{21}\), which suggests that these CC3α-PAN membranes might be more competitive for separations that require higher pressures.

To confirm that dye adsorption did not contribute to the selectivity performance of CC3α-PAN, mass balance calculations were used to measure the dye concentration in the retentate. After permeating 48 ml of Congo red from the 100 ml feed, the absorption intensity of Congo red in the retentate increased from 1.24 to 2.53, while its absorption intensity in the permeate was only 0.02. In combination, these values are consistent with ~100% dye rejection (Fig. 3f and Supplementary Figs. 29 and 30). These measurements agree with the colourless membrane surface observed after the dye filtration (Supplementary Fig. 2). In addition, soaking powdered crystals of CC3α (100 mg) in the dye solution (100 ml) did not lead to adsorption in the crystals after seven days (Supplementary Fig. 31). These results all indicate that the dyes were rejected by the membrane.
Switchable pore aperture for graded sieving

Previous studies showed that certain POCs can be switched between multiple polymorphs to modify their porosity\(^{28,29}\). The solid-state structure of CC3 has been directed into different polymorphs by crystallization from specific solvents\(^{48}\), but until now, the solid-state transformation of CC3 crystals was not explored, to the best of our knowledge.

We found that both air-dried and water-solvated films exhibited the same diffraction patterns as the reference peaks of CC3\(\alpha\)-powders measured by PXRD (Fig. 4a). A series of GIXRD patterns were then recorded after submerging the membrane in various organic solvents (Supplementary Figs. 32 and 33). The crystalline CC3\(\alpha\)-film transformed into a new structure when submerged in MeOH (Fig. 4a). By indexing the GIXRD pattern, we confirmed this was a MeOH-solvated CC3 phase (CC3\(\gamma\); Supplementary Fig. 34) that was isolated previously by crystallizing CC3 from dichloromethane and MeOH (ref. \(^{48}\)). The CC3\(\gamma\) structure is very different from its thermodynamically most stable polymorph, CC3\(\alpha\) (ref. \(^{48}\)), where the cage packs in a window-to-window arrangement to generate a diamondoid pore network (yellow channels in Fig. 1b). By contrast, the CC3 molecules in the CC3\(\gamma\) phase are packed less densely, thus providing large extrinsic pores between hexagonally arranged CC3 molecules (orange channels in Fig. 1c).

To investigate the structural transformation between CC3\(\alpha\)-PAN and CC3\(\gamma\)-PAN, we performed a series of in situ GIXRD measurements while dosing the membrane surface with solvent vapour and after coating the membrane surface in a thin solvent layer (Methods). CC3\(\gamma\)-PAN formed by immersion in MeOH transformed back into CC3\(\alpha\)-PAN after being immersed in water (Fig. 4c), with evidence of both phases found when the membrane was immersed in a mixture of water and MeOH (Supplementary Figs. 35 and 36). High-resolution PXRD also confirmed that CC3\(\gamma\) cleanly transforms into CC3\(\alpha\) after thermally desolvating a powdered sample of CC3\(\gamma\) suspended in MeOH in a capillary (Supplementary Fig. 37).

We next used MeOH rather than water to dissolve the dyes and filtered these solutions through the CC3-PAN membrane under the same conditions. Interestingly, the MWCO shifted from 600 g mol\(^{-1}\) in water to 1,400 g mol\(^{-1}\) in MeOH for the same membrane (Fig. 4b and Supplementary Figs. 38 and 39). By contrast, a commercial Synder NDX nanofiltration membrane with a comparable MWCO (500–700 g mol\(^{-1}\)) exhibited similar rejection behaviour in both water and MeOH (Supplementary Figs. 40 and 41). We attribute this dramatic change in MWCO to the phase transformation to CC3\(\gamma\)-PAN in MeOH. We further investigated how crystallinity influences the switchable pore aperture by measuring dye rejection of CC3-PAN membranes with lower crystallinity (fabricated using lower concentrations or shorter reaction times; Supplementary Figs. 42–45). CC3-PAN-4h-0.8% rejected 78.2% of brilliant blue (BB) dye from water compared to 52.7% from MeOH, while the less crystalline CC3-PAN-4h-0.2% had a less distinct BB rejection performance (68.6% from water versus 52.8% from MeOH). Hence, the high crystallinity in the CC3 membrane is essential for regulating its separation performance after switching its pore aperture using a solvent stimulus.

We next performed molecular separations while cycling between CC3\(\alpha\)-PAN and CC3\(\gamma\)-PAN using a single membrane and water and MeOH feedstocks containing the BB dye. We found that both water and MeOH permeances remained high after cycling between CC3\(\alpha\)-PAN and CC3\(\gamma\)-PAN (Fig. 4c, Supplementary Tables 5–7 and Supplementary Video 1). More importantly, the rejection of BB switches between ~100% in water to ~0% in MeOH in each cycle (Fig. 4c,d); that is, the membrane can be switched ‘on’ and ‘off’ using a solvent. The reversible transition between CC3\(\alpha\)-PAN and CC3\(\gamma\)-PAN appears to be complete within the one minute it takes to switch the feedstock (Supplementary Video 1) and creates alternative diffusion pathways through the membrane structure. From in situ GIXRD measurements on solvated CC3 films, while performing two consecutive cycles, we found that the composite membrane transformed cleanly between CC3\(\alpha\)-PAN and CC3\(\gamma\)-PAN when the solvent was switched between water and MeOH and back again (Fig. 4e). We attributed this switching phenomenon solely to the phase transition of CC3 films, rather than swelling of the membranes. To validate this, the CC3-PAN membrane was soaked in acetonitrile, and the nanofiltration tests were repeated. CC3-PAN exhibits comparable MWCOs in acetonitrile than those observed in water (Supplementary Figs. 41 and 46) because the same phase, CC3\(\alpha\)-PAN, is present in these solvents (Supplementary Figs. 32 and 33). Remarkably, the acetonitrile permeance of CC3\(\alpha\)-PAN reached 1771 lm\(^{-2}\)h\(^{-1}\)bar\(^{-1}\) with a MWCO of ~600 g mol\(^{-1}\), which is well above the upper bound performance for nanofiltration membranes reported in the literature (Fig. 4f, Supplementary Fig. 47 and Supplementary Table 9).

A series of water and MeOH feedstocks containing the dye BB were used to determine the dynamic transformation between CC3\(\alpha\)-PAN and CC3\(\gamma\)-PAN (Fig. 5a). Understanding this dynamic transformation allowed us to manipulate the pore aperture in a single CC3-PAN membrane by simply adjusting the water concentration in a water–MeOH mixture, without any activation processes\(^{31}\) or the use of multiple membranes\(^{21}\). To demonstrate this, we performed a graded sieving experiment to separate molecules from a ternary mixture using a single membrane. Initially, a water feedstock containing three dyes, 4-nitrophenol (NP; yellow, 139 g mol\(^{-1}\)), BB (blue, 826 g mol\(^{-1}\)) and direct red 80 (DR; red, 1,373 g mol\(^{-1}\)) was filtered through the CC3-PAN membrane (Fig. 5c). Since CC3-PAN adopts its CC3\(\alpha\)-PAN structure in water, the narrower pore aperture allowed only the smallest molecule, NP, to diffuse through the membrane, while the larger molecules, BB and DR, were rejected. Excess water was used for flushing residual NP from the retentate, and this process was repeated until the NP concentration in the permeate was below 1%. Subsequently, 90 vol% of MeOH was added into the water retentate to generate a feedstock that transformed the membrane structure to CC3\(\gamma\)-PAN with the larger pore aperture. BB could then diffuse through the membrane alone, while DR was retained in the cell (Fig. 5b,d). Finally, excess MeOH was used to flush any residual BB from the cell to leave only DR in the retentate, where it could be collected in pure form (Supplementary Section 1.4).

Conclusion and outlook

Continuous, defect-free POC membranes can achieve high permeances for a range of organic solvents—in some cases exceeding upper performance bounds—while also showing excellent separation performances. These highly ordered crystalline POC membranes exhibit a switchable phase transition between two crystalline forms, CC3\(\alpha\)-PAN and CC3\(\gamma\)-PAN. This allows graded sieving to separate a mixture of three organic dyes using a single, smart membrane and creates a membrane-based parallel to the widespread and highly effective use of solvent gradients in chromatography\(^{52}\). POC membranes with switchable pore apertures could also lead to new applications in triggered drug delivery\(^{53}\), biosensors\(^{31}\) or fermentation/fractionation processes\(^{55}\).

While the current synthesis process makes it challenging to scale and implement these POC membranes in commercial processes, it is conceivable that a more scalable production method might be developed by exploiting the solution processability of these molecular cages. Future efforts will focus on using computational methods, such as crystal structure prediction, to design POC crystals with specific properties that can be designed from first principles.

Online content

Any methods, additional references, Nature Research reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of
25. Wang, Z., Zhang, S., Chen, Y., Zhang, Z. & Ma, S. Covalent organic framework films. ACS Chem. Sci. 4, 1663–1676 (2018).
26. Jelts, K. E. & Cooper, A. I. Molecular simulations to understand and to design porous organic molecules. Curr. Opin. Solid State Mater. Sci. 17, 19–30 (2013).
27. Little, M. A. & Cooper, A. I. The chemistry of porous organic molecular materials. Adv. Funct. Mater. 30, 1909842 (2020).
28. Chen, L. et al. Separation of rare gases and chiral molecules by selective binding in porous organic cages. Nat. Mater. 13, 954–960 (2014).
29. Nunes, S. P. & Peinemann, K.-V. Membrane Technology in the Chemical Industry (John Wiley & Sons, 2006).
30. Kim, D., Salazar, O. R. & Nunes, S. P. Membrane manufacture for peptide separation. Grimm. J. 41, 5115–5119 (2014).
31. Marchetti, P., Jimenez Solomon, M. F., Szekely, G. & Livingston, A. G. Molecular separation with organic solvent nanofiltration: a critical review. Chem. Rev. 114, 10735–10806 (2014).
32. Sholl, D. S. & Lively, R. P. Seven chemical separations to change the world. Nat. Chem. 7, 114–117 (2015).
33. Jiang, Z., Karam, S. & Livingston, A. G. Water transport through ultrathin polyamide nanofilms used for reverse osmosis. Adv. Mater. 30, 1709793 (2018).
34. Cundy, C. S. & Cox, P. A. The hydrothermal synthesis of zeolites: precursors, intermediates and reaction mechanism. Micropor. Mesopor. Mat. 82, 1–78 (2005).
35. Thomas, A. Functional materials: from hard to soft porous frameworks. Angew. Chem. Int. Ed. 49, 8328–8344 (2010).
36. Furukawa, H., Cordova, K. E., O’Keeffe, M. & Yaghi, O. M. The chemistry and applications of metal-organic frameworks. Science 341, 1230444 (2014).
37. Cöte, A. P. et al. Porous, crystalline, covalent organic frameworks. Science 316, 1166–1170 (2007).
38. Lin, R.-B. et al. Multifunctional porous hydrogen-bonded organic framework materials. Chem. Soc. Rev. 48, 1362–1389 (2019).
39. Dey, K. et al. Selective molecular separation by interfacially crystallized covalent organic framework thin films. J. Am. Chem. Soc. 139, 13083–13091 (2017).
40. Marchesini, M. et al. Lewis-acid-catalyzed interfacial polymerization of covalent organic framework films. Chem. 4, 308–317 (2018).
41. Corcos, A. R. et al. Reducing the pore size of covalent organic frameworks in thin-film composite membranes enhances solute rejection. ACS Mater. Lett. 1, 440–446 (2019).
42. Zhang, C., Wu, B.-H., Ma, M.-Q., Wang, Z. & Xu, Z.-K. Ultrathin metal-covalent-organic framework membranes towards ultimate separation. Chem. Soc. Rev. 48, 3811–3841 (2019).
43. Wang, Z., Zhang, S., Chen, Y., Zhang, Z. & Ma, S. Covalent organic frameworks for separation applications. Chem. Soc. Rev. 49, 708–735 (2020).
44. Venna, S. R. & Carreau, M. A. Metal organic framework membranes for carbon dioxide separation. J. Membr. Sci. 561, 165–178 (2018).
45. Fenton, I. L., Burke, D. W., Qian, D., Oliveira da Cruz, M. & Dichtel, W. R. Polycrystalline covalent organic framework films act as adsorbents, not membranes. J. Am. Chem. Soc. 143, 1466–1473 (2021).
46. Jones, J. T. A. et al. On–off porosity switching in a molecular organic solid. Angew. Chem. Int. Ed. 50, 749–753 (2011).
47. Bera, S. et al. Porosity switching in polymorphic porous organic cages with exceptional chemical stability. Angew. Chem. Int. Ed. 58, 4243–4247 (2019).
48. Song, Q. et al. Porous organic cage thin films and molecular-sieving membranes. Adv. Mater. 28, 2629–2637 (2016).
49. Brutschi, M., Schneider, M. W., Mastalerz, M. & Waldvogel, S. R. Porous organic cage compounds as highly potent affinity materials for sensing by quartz crystal microbalances. Adv. Mater. 24, 6049–6052 (2012).
50. Bushell, A. F. et al. Nanoporous organic polymer/cage composite membranes. Angew. Chem. Int. Ed. 52, 1253–1256 (2013).

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.
© The Author(s) 2021
Methods

Interfacial synthesis of crystalline CC3 films. An aqueous solution of CHDA (0.26 g, 2.24 mmol, 0.8 wt%) in water (32 ml) was carefully layered on top of a dichloromethane solution (30 ml) that contained TFB (0.241 g, 1.48 mmol, 0.8 wt%) and was stored at room temperature. A hexagonal film grew at the dichloromethane–water interface and was then isolated as a free-standing film that could be layered directly onto different substrates, including glass, stainless steel, mesh, carbon tape and silicon wafers. To perform liquid permeation studies, the CC3 film was transferred onto a PAN support to form composite CC3-PAN membranes, which were then soaked in pure water for 1 day (Supplementary Figs. 48 and 49; Supplementary Section 1.2 for full experimental details and Supplementary Fig. 50 for the reaction set-up). Fabrication of PAN supports via phase inversion is presented in Supplementary Section 1.2.

X-ray diffraction. GIXRD measurements were performed using the 107 beamline at Diamond Light Source in the United Kingdom (wavelength, \(\lambda = 0.689\) Å), using a vertical (2 + 2)-type diffractometer equipped with a Pilatus 100 K area detector\(^*\). Membrane samples were cut into 1 x 2 cm\(^2\) pieces and stuck on glass supports, which were then mounted on a hexapod (PI-Micos) to allow independent alignment with six degrees of freedom during the data collection (Supplementary Fig. 51a). The measurements were conducted by moving the detector while maintaining a fixed sample position. The grazing incidence angle is set at 2°. Data collection was performed at room temperature using in-plane (over the 2\(00\) range 3–40°, 0.5° step size and out-of-plane (over the 2\(00\) range 2–40°, 0.25° step size) measurement geometries, and GIXRD scans were processed in DAWN 2 (ref. \(^*\)). GIXRD patterns were refined by Pawley refinement through TOPAS Academic\(^*\). High-resolution synchrotron PXRD data were collected using the I11 beamline at Diamond Light Source (\(\lambda = 0.827\) Å). The full PXRD details are presented in the Supplementary Information.

For the in situ GIXRD measurements performed on solvated samples, pieces of Mylar film were used to cover the membrane surface with a thin layer of solvent (water, MeOH, acetone and acetonitrile) during the GIXRD scans (Supplementary Fig. 51b). To investigate the reversible transformation between CC3-PAN and CC3-P, a membrane sample was removed from water without drying and covered with 1.0 ml of a MeOH solvent layer before recording the GIXRD data (Supplementary Fig. 35). To more closely mimic the reversible membrane separation experiment where the feedstock was cycled between water and MeOH, a CC3-PAN sample was removed from water without drying, soaked in 100 ml MeOH for 1 minute and covered with a thin layer of MeOH (1.0 ml) before the GIXRD measurement. The same process was repeated with the identical CC3-PAN sample using water or MeOH (Fig. 6c). For the in situ measurements performed using solvent vapours, nitrogen gas was bubbled through a 21 bottle that contained the organic solvent at a flow rate of 10 ml min\(^{-1}\). The 'wet gas' generated during this process was then continually flowed over the membrane sample during the full measurement and Mylar film was used to seal the sample environment.

Separation measurements. Solvent permeance and dye rejection measurements were performed using a Sterilehtick HP4750 dead-end membrane filtration system (Supplementary Fig. 22). We also used a commercial bench-scale 50 ml transparent Merck Millipore Amicon dead-end stirred cell that was connected to an 800 ml Merck Millipore Amicon R8000 reservoir, to visualize the filtration process (Supplementary Fig. 23). During these measurements, the feedstocks were kept under a 10 bar nitrogen pressure (3 bar for Merck Millipore Amicon cells) at room temperature, and the feedstock was continually stirred using a stirring bar rotating at 400 rpm. The Hansen solubility parameter (\(\delta\)) and the physical properties of the organic solvents (Supplementary Table 2) were used to investigate the relationships between pure solvent permeances and the combined solvent properties. Flux \(J (\text{ml} \cdot \text{h}^{-1} \cdot \text{cm}^{-2})\) was calculated according to the following equation:

\[
J = \frac{V}{A} \times (\frac{\Delta V}{\Delta t}) \tag{1}
\]

where \(\Delta V\) is the volume of permeate collected in litres in a given amount of time, \(A\) is the membrane surface area in square metres and \(\Delta t\) is the time in hours between the start and end of the measurement. Solvent permeance \(P (\text{ml} \cdot \text{h}^{-1} \cdot \text{cm}^{-2} \cdot \text{bar}^{-1})\) was calculated according to the following equation:

\[
P = \frac{V}{A} \times (\frac{\Delta V}{\Delta t} \times \rho) \tag{2}
\]

where \(\Delta V\) is the volume of permeate collected in litres in a given amount of time, \(A\) is the membrane surface area in square metres, \(\Delta t\) is the time in hours between the start and end of the measurement and \(\rho\) is the transmembrane pressure. To calculate solvent permeance, typical feedstock pressure of 0.21 bar for pure solvent or dye feedstock (20 ppm dye concentration) was added to the feedstock tank. The cell was then pressurized to 10 bar under nitrogen. The solvent permeate was then calculated based on the amount of time it took –0.1 l of pure solvent or dye feedstock to flow through the membrane (Supplementary Tables 3 and 4 for full details). The retentate was collected after each measurement. Error bars (s.d.) were calculated by the STDDEV function using data obtained from at least three independent membranes.

For the dye rejection measurements, a series of dye feedstock solutions in different solvents (water, MeOH, acetone and acetonitrile) were prepared with a dye concentration of 20 ppm using the following dyes: reactive red 120 (1,470 g mol\(^{-1}\)), DR (1,373 g mol\(^{-1}\)), rose benzal (1,018 g mol\(^{-1}\)), BB (826 g mol\(^{-1}\)), Congo red (697 g mol\(^{-1}\)), protoporphyrin IX disodium (607 g mol\(^{-1}\)), acid fuchsin (585 g mol\(^{-1}\)), sunset yellow (452 g mol\(^{-1}\)), methyl orange (327 g mol\(^{-1}\)), neutral red (289 g mol\(^{-1}\)) and NP (139 g mol\(^{-1}\); Supplementary Table 1 for full details).

Ultrafiltration–visible spectrophotometry was used to measure the dye concentration in the permeate to calculate dye rejection performance. Dye rejection, \(R\) (%), of the membranes was calculated as follows:

\[
R = \left(1 - \frac{C_r}{C_p}\right) \times 100 \tag{3}
\]

where \(C_r\) and \(C_p\) represent the dye concentrations in the permeate (\(C_r\)) and feed (\(C_p\)). Dye concentrations in the permeate and feed were determined using a Cary 5000 ultrascan/visible/near-infrared spectrophotometer with the wavelengths specified in Supplementary Table 1. The MWCO was determined by interpolating from the plot of rejection against the molecular weight of the dyes and corresponds to the molecular weight for which rejection is 90%. During these measurements, the volume and the concentration of the permeate and the retentate were measured, and the mass balance of the feed solution could be calculated as follows:

\[
C_t \times V_t = C_p \times V_p + C_r \times V_r \tag{4}
\]

where \(C_t\), \(C_r\) and \(C_p\) are the dye concentrations in parts per million (grams per litre) of the feed, permeate and retentate, respectively; \(V_t\), \(V_r\) and \(V_p\) represent the volume of the feed, permeate and retentate in litres, respectively. Typically, 0.2 l of the feed solution was added into the cell, then 0.1 l permeate was collected and 0.1 l retentate was left in the cell. Reversible filtration tests, membrane absorption tests, long-term operation information, membrane stability tests, water and MeOH feedstock mixture separation experiments and graded sieving experiments for the ternary system are presented in Supplementary Section 1.4. The set-up of a commercial bench-scale dead-end stirred filtration unit with transparent cells (a 50 ml transparent Merck Millipore Amicon dead-end stirred cell connected to an 800 ml Merck Millipore Amicon R8000 reservoir) is shown in Supplementary Fig. 23. Reversible filtration measurement data in water and MeOH are shown in Supplementary Tables 5–7, and dye rejection measurement data in a water and MeOH mixture are shown in Supplementary Table 8.

Data availability

Source data are provided with this paper. In addition, source data are deposited in the University of Liverpool Research Data Catalogue (https://doi.org/10.17638/ datacat.liverpool.ac.uk/1512). Further details can be obtained from the authors upon request.

References

56. Nicklin, C., Arnold, T., Rawle, J. & Warne, A. Diamond beamline I07: a beamline for surface and interfacial diffraction. J. Synchrotron Radiat. 23, 1245–1253 (2016).
57. Basham, M. et al. Data analysis workbench (DAWN). J. Synchrotron Radiat. 22, 853–858 (2015).
58. Coelho, A. TOPAS Academic v5 (Coelho Software, 2012).

Acknowledgements

A.I.C., M.A.L., M.E.B., Y.W. and A.H. acknowledge the Engineering and Physical Sciences Research Council (EP/N004884/1). A.I.C. and A.H. acknowledge the Leverhulme Trust via the Leverhulme Research Centre for Functional Materials Design for funding. A.H. acknowledges the China Scholarship Council for a studentship and the Royal Society of Chemistry for a Researcher Mobility Grant (M19–2442). Z.J. and A.G.L. acknowledge the Engineering and Physical Sciences Research Council for funding (EP/1801847/1). A.I.C., M.A.L., A.H., Y.W., H.H. and J.R acknowledge Diamond Light Source for access to beamlines I07 (SI24359) and I11 (C123666) that contributed to the results presented here. We thank M. Liu for advice about the cage synthesis conditions, H. Chen for assistance during the I07 measurements, P. Cui and H. Gao for assistance during the I11 measurements, Y. Li for fitting the logical curve (Fig. 5a), R. Clowes for instrument support and setting up the filtration cell, H. Yang and T. Mitra for assistance collecting SEM images and K. Arnold for collecting the FIB-SEM images.

Author contributions

A.I.C. and A.G.L. conceived the project. A.H. and M.A.L. conceived the graded molecular sieving part of the study and performed the associated measurements. A.H. and Z.J. prepared the membranes. A.H. led the characterization and membrane performance tests. Z.J. synthesized the PAN membrane support and conducted the AFM measurements. M.E.B. helped with the cage synthesis. H.H. and J.R. designed and set up the in situ GIXRD system and helped with these measurements. A.H., M.A.L. and
Y.W. contributed to the GIXRD measurements and processed the X-ray data. A.G.L. provided facility support and insight into advanced membrane science. Z.J. designed the interfacial synthesis method for the membrane. M.A.L. cosupervised the project with A.I.C. and A.G.L. and wrote the manuscript with A.H. and Z.J., with contributions from all coauthors. All of the authors participated in the discussion of data and commented on the manuscript.

**Competing interests**
The authors declare no competing interests.

**Additional information**

**Supplementary information** The online version contains supplementary material available at https://doi.org/10.1038/s41563-021-01168-z.

**Correspondence and requests for materials** should be addressed to Andrew G. Livingston or Andrew I. Cooper.

**Peer review information** Nature Materials thanks Michael Mastalerz and the other anonymous, reviewer(s) for their contribution to the peer review of this work.

**Reprints and permissions information** is available at www.nature.com/reprints.