Albiflorin Attenuates Mood Disorders Under Neuropathic Pain State By Suppressing The Hippocampal NLRP3 Inflammasome Activation During Chronic Constriction Injury

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Significance Statement

Neuropathic pain (NP) is a severe condition with several comorbidities, such as mood disorders, affecting many individuals worldwide. This study examines the effect of the Al biflorin (AF) on the NLRP3 inflammasome, which is involved in the neuroinflammatory pathogenesis of NP, anxiety, and depression. We found that the NLRP3 inflammasome contributed to the NP and comorbid anxiety and depression-like behaviors induced by chronic constriction injury of the sciatic nerve, and that AF may relieve these symptoms via inhibition of the NLRP3 inflammasome activity. Moreover, AF enhanced the translocation of Nrf2 into the nucleus and suppressed NF-kB activity in the hippocampus. We believe that our study makes a significant contribution to the literature because it strongly suggests that AF directly regulates the NLRP3 inflammasome, proposing AF as a potential therapeutic agent for NP and related mood disorders.
Abstract

**Background:** Neuropathic pain (NP) is a multifaceted and ubiquitous disease across the globe. Mood disorders, such as anxiety and depression, are frequently observed in patients suffering from NP. Both NP and comorbid mood disorders seriously impacts the quality of life. Accumulated evidence shows that activation of the NOD-like receptor protein 3 (NLRP3) inflammasome is involved in the neuroinflammatory pathogenesis of NP, anxiety, and depression. However, the role of the NLRP3 inflammasome in the pathological process of anxiety and depression under the NP state has not been fully described. Albiflorin (AF), a monoterpenic glycoside, may be a potential regulator of the NLRP3 inflammasome, but it is not clear whether AF relates to NLRP3 inflammasome activation.

**Methods:** We used a systematic pharmacological method to confirm whether the activation of the NLRP3 inflammasome in the hippocampus was involved in the development of NP associated with mood disorders, and whether AF could be an effective treatment for these symptoms.

**Results:** The NLRP3 inflammasome contributed to the NP and comorbid anxiety and depression-like behaviors induced by chronic constriction injury of the sciatic nerve, and that AF may relieve these symptoms via inhibition of the NLRP3 inflammasome activity. Moreover, AF enhanced the translocation of the nuclear factor erythroid 2-related factor 2 (Nrf2) into the nucleus and suppressed NF-κB activity in the hippocampus.

**Conclusions:** AF, as a potential therapeutic agent, might greatly improve the overall symptoms of NP.

**Keywords:** albiflorin, neuropathic pain, anxiety and depression, NLRP3 inflammasome
Introduction

Neuropathic pain is defined as pain caused by a lesion or disease of the somatosensory system and is a multifaceted and ubiquitous disease across the globe (Jensen et al., 2011). Mood disorders, such as anxiety and depression, are frequently observed in patients who are suffering from chronic pain (Davis et al., 2011; Radat et al., 2013). These mental comorbidities increase neuropathic hypersensitivity (Yalcin et al., 2014; Doan et al., 2015; Humo et al., 2019). Thus, both neuropathic pain and comorbid mood disorders constitute a severe burden for patients, which can seriously impact the quality of life (Yalcin et al., 2014; Barthas et al., 2015; Humo et al., 2019). Current drugs for neuropathic pain exhibit both poor efficacy and tolerability due to the lack of knowledge regarding the mechanisms of pathological pain (Cornelius et al., 2013; Peng et al., 2017; Inoue and Tsuda, 2018; Calvo et al., 2019; Meyer et al., 2019). Therefore, it is urgent to further clarify the mechanisms of neuropathic pain and identify more effective and non-addictive pharmacological therapies.

Accumulated studies have verified that neuroinflammation is involved in the pathogenesis of various neurogenic diseases and the overproduction of inflammatory cytokines are essential molecules responsible for neuroinflammation (Capuron and Miller, 2011; Fleshner et al., 2017). Among various inflammatory cytokines, a wide array of evidence supports the vital role of interleukin-1β in the pathogenesis of multiple neuroinflammatory symptoms (Koo and Duman, 2008; Heneka et al., 2013; Clausen et al., 2016; Chen et al., 2018; Gordon et al., 2018; Li et al., 2019b). Nevertheless, IL-1β is only active after proteolysis by cysteine proteases (Afonina et al., 2015). Inflammasomes as cytosolic multiprotein complexes serve as platforms for the activation of cysteinyll aspartate specific proteinase (caspase)-1, which is the main mediator promoting the processing and
secretion of IL-1β (Swanson et al., 2019). Nucleotide binding and oligomerization domain-like receptor family pyrin domain-containing 3 (NLRP3) is the most widely implicated regulator of caspase-1 activation. The NLRP3 inflammasome consists of NLRP3, apoptosis-associated speck-like protein (ASC), and caspase-1. When activated by exogenous irritants or endogenous danger signals, the NLRP3 inflammasome generates and activates caspase-1, which subsequently cleaves pro-IL-1β to its mature form, IL-1β (Swanson et al., 2019). Various studies have reported that the NLRP3 inflammasome is involved in the pathogenesis of neuropathic pain (Jia et al., 2017; Pan et al., 2018; Tonkin et al., 2018; Xu et al., 2019a), or anxiety and depression symptoms (Xu et al., 2016; Alcocer-Gómez and Cordero, 2017; Carney and Freedland, 2017; Yue et al., 2017; Wang et al., 2018; Feng et al., 2019; Zhang et al., 2019). Importantly, inhibition of the NLRP3 inflammasome could independently relieve each of these symptoms. However, to our knowledge, NLRP3 inflammasome has not been reported to be involved in the pathological process of anxiety and depression under neuropathic pain state.

In addition to NLRP3, Kelch-like ECH associated protein 1 (Keap1) / nuclear factor erythroid 2-related factor 2 (Nrf2) signaling pathway is also involved in neuropathic pain and depression in preclinical and clinical studies (Braidy et al., 2018; Li et al., 2018; Bansal et al., 2019; Chu et al., 2019; Dang et al., 2019; Swanson et al., 2019; Zborowski et al., 2020; Zhou et al., 2020a, 2020b). Keap1 suppresses Nrf2 activity under physiological conditions, whereas Nrf2 is liberated from Keap1-mediated suppression on exposure to stresses. When Keap1-Nrf2 signaling pathway was activated, Keap1 is chemically modified and releases Nrf2, which results in Nrf2 translocating into the nucleus. Nuclear Nrf2-driven gene transcription regulates the levels of antioxidant genes limiting ROS levels. Therefore, the significant
increase of Nrf2 in nucleus (Nuclear Nrf2) is a good marker indicating the activation of Keap1-Nrf2 pathway (Sies et al., 2017; Yamamoto et al., 2018). Moreover, Keap1-Nrf2 signaling pathway has been reported to regulate the levels of antioxidant genes limiting the reactive oxygen species (ROS) levels and attenuate the activation of NF-κB thus inhibiting NLRP3 activation (Li et al. 2008; Liu et al. 2017; Chu et al., 2019; Dang et al., 2019), which implies that regulating the Keap1-NRF2 / NLRP3 pathway might be a potential therapy against neuropathic pain and depression.

*Paeoniae Alba* Radix, the dried roots of *P. lactiflora* Pallas or *P. veitchii* Lynch, possesses various medicinal properties including activating blood and alleviating pain (Ma et al., 2015). It has been widely used in traditional Chinese prescriptions to alleviate depression-like symptom for hundreds of years (Mao et al., 2012; Li et al., 2013; Zhu et al., 2015). Albiflorin (Figure 1) is a major constituent among the glucosides contained in *P. alba* Radix. Albiflorin has been reported to exert powerful anti-inflammatory effects and antidepressant activity in clinical reports and preclinical studies (Song et al., 2015; Wang et al., 2016; Han et al., 2018; Zhao et al., 2018; Cai et al., 2019; Wang et al., 2019; Zhang and Wei, 2020). Our previous study (Zhou et al., 2016) has demonstrated that albiflorin displays a significant analgesic effect on peripheral nerve injury-induced neuropathic pain rats. We also found that albiflorin efficiently reduced the elevated spinal IL-1β levels in neuropathic pain rats (Zhou et al., 2016). Previous studies have reported that albiflorin exerted antioxidant effects via reducing ROS levels (Suh et al., 2013; Xu et al., 2019b). Moreover, some Chinese herbal formulas containing albiflorin have been reported to ameliorate oxidative stress via the activation of the Keap1-Nrf2 signaling pathways to limit ROS (Ma et al., 2015; Yang et al., 2015; Hong et al., 2017). According to the above evidence, we hypothesized that albiflorin...
might reduce mood disorders accompanied with neuropathic pain through inhibition of the NLRP3 inflammasome, which is related to the regulation of Keap1-Nrf2 pathway. It should be noted that data regarding the biological activities of albiflorin are scarce, and no report has examined the effect of albiflorin on the NLRP3 inflammasome.

In the present study, we confirmed that activation of the NLRP3 inflammasome was involved in the pathogenesis of comorbid mood disorders (anxiety and depression) associated with neuropathic pain. Blocking the activation of the inflammasome constituted an ideal approach to improve anxiety and depression-like behaviors under the neuropathic pain state. Moreover, the present study showed a novel role for albiflorin, which exerted an anti-anxiety and antidepressant effects highly associated with the inhibition of hippocampal inflammasome activation. Finally, we found that albiflorin inhibits the activation of NLRP3 inflammasome involved in enhancing nuclear factor erythroid 2-related factor 2 (Nrf2) translocation into the nucleus.
Methods

Drugs

MCC950 (a selective NLRP3 inhibitor) was obtained from MedChemExpress (Monmouth Junction, NJ, USA). Albiflorin was purchased from Chengdu Refmedic (Chengdu, China). The purity of albiflorin, as determined by high-performance liquid chromatography (Figure 1), was greater than 97%.

Animals

Adult male Sprague–Dawley (SD) rats (180–200 g) were purchased from Beijing Vital River Laboratory Animal Technology Co., Ltd (Beijing, China). The rats were housed under a scheduled 12-h light/dark cycle at a constant temperature of 23 °C ± 1 °C. This study was carried out in accordance with the principles of the Basel Declaration and recommendations of Guide for the Care and Use of Laboratory Animals (Ministry of Science and Technology of China, 2006). The protocol was approved by the Chengde Medical University Animal Care Committee.

Chronic constriction injury (CCI) of the sciatic nerve

The SD rats were anesthetized by intraperitoneal (i.p.) injection of chloral hydrate (300 mg/kg) and intramuscular (i.m.) injection of ketorolac tromethamine (1 mg/kg). The operation was performed as described by Bennett and Xie (Bennett and Xie, 1988). Briefly, a small incision was made in the right thigh and the sciatic nerve was exposed. Then, four ligatures (4.0 chromic catgut) were tied loosely at the mid-thigh level of the sciatic nerve. Sham surgery comprised of exposure of the right sciatic nerve without ligation.

Drug administration
Albiflorin and MCC950 were dissolved in sterile normal saline solution before use. The optimal administration doses of albiflorin (50 mg/kg) and MCC950 (10 mg/kg) were selected based on previous studies (Zhou et al., 2016; Khan et al., 2018; Chen et al., 2019b) and the results of preliminary experiments. MCC950 and albiflorin or vehicle were administered by i.p. injection daily for 15 days, starting on day 15 after CCI.

**Behavioral measurements**

Before initiating the behavioral tests, rats were acclimated to experimental room conditions for at least 30 min. All tests were carried out according to the schedules depicted in Figure 2A and Figure 4A. Each of the individual behavioral tests was performed by the same observers blind to the grouping scheme.

**Mechanical allodynia**

Mechanical allodynia was measured using an electronic Von Frey Aesthesiometer (BIO-EVF4, Bioseb, Vitrolles, France). Briefly, the rats were placed on an elevated mesh platform in acrylic cages to acclimate for 15 min before the test. The elastic spring-type tip was fitted on the hand-held force transducer and stimulated the plantar surface of the right hind paw. The tests were repeated three times, with an interval of 5 min between each test, and the mechanical threshold was defined as the force (in g) initiating a withdrawal response averaged from the three measurements. Then the paw withdrawal threshold (PWT) was captured as the mechanical allodynia threshold.

**Open field test (OFT)**

Rats were placed in the center of an non-transparent arena (100 cm × 100 cm × 40 cm) individually; then, total distance traveled and duration of central zone visits were recorded for 5 min and analyzed using a behavior analysis system (SMART 3.0, Panlab, Barcelona,
Spain). The method was modified from that used in a previous study (Bambico et al., 2007; Hu et al., 2009).

**Elevated plus maze test (EPM)**

The test was conducted in a cross shaped elevated maze consisting of two closed arm (50 cm × 10 cm × 40 cm) and two open arms (50 cm × 10 cm). Rats were singly placed in the middle of the maze, facing one of the open arms; then, their behavior was recorded for 5 min. The method is modified from that described in a previous study (Pellow et al., 1985; Bambico et al., 2010). The open arm entries and the duration of visiting the open arms were detected and analyzed using the SMART 3.0 software.

**Forced swim test (FST)**

The test was conducted in a glass cylinder (diameter, 22 cm; height, 50 cm) containing 30 cm of water at 23 °C ± 1 °C. The test was examined using the SMART 3.0 software according to a previously described protocol (Porsolt et al., 1978; Bambico et al., 2007). First, the rats were placed in the water for a 15-min training exposure and 24 h later, they were re-exposed for a 5-min test session, during which the duration of immobility was analyzed.

**Immunofluorescence Labeling**

On day 29 after CCI surgery, rats were deeply anesthetized by injections of chloral hydrate (350 mg/kg, i.p.) and ketorolac tromethamine (1 mg/kg, i.m.) after finishing the behavioral tests. The hippocampus was collected, post-fixed, embedded in paraffin, and cut into 5-μm thick sections. The sections were deparaffinized and incubated in citrate repair buffer (pH 6.0); sections were incubated in 3% BSA (Bovine Serum Albumin) for 30 min at 25 °C to block non-specific proteins. The hippocampal sections were incubated with a mixture of mouse anti-neuronal nuclear antigen (NeuN) (1:200, Millipore, Billerica, MA, USA), and rabbit anti-
NLRP3 antibody (1:50, Novus, Littleton, CO, USA) or rabbit anti-caspase-1 (1:500, servicebio, Wuhan, China) overnight at 4 °C. After three washes in PBS (pH 7.4), sections were incubated with Cy3-conjugated goat anti-rabbit (1:300, Servicebio) or 488-conjugated goat anti-mouse (1:400, Servicebio) secondary antibodies. Next, the sections were washed in PBS for 15 min, mounted on slides, and covered with coverslips. For control, primary antibodies were omitted in the process. Sections were imaged with a fluorescence microscope (BX53, Olympus, Tokyo, Japan). Images were taken using a digital camera system (DP74, Olympus, Tokyo, Japan). Analysis was performed by the same observer who was blinded to the grouping scheme.

**Western blot analysis**

On day 29 after CCI, the hippocampus was collected, frozen in liquid nitrogen, and stored at −80 °C until further processing. The total protein was supplemented with protease inhibitors and extracted using radioimmunoprecipitation assay (RIPA) lysis buffer (Beyotime Biotechnology, Shanghai, China). The nuclear protein fraction was extracted using the Nuclear Extraction Reagents Kit (Beyotime Biotechnology). Proteins were separated using 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and the resolved proteins were transferred onto nitrocellulose membranes (Millipore, Billerica, MA, USA). Membranes were incubated in 5% nonfat dry milk for 2 h at 25 °C and overnight at 4 °C with primary antibodies. Primary antibodies used in this study were: a rabbit anti-NLRP3 (1:500, Abcam, Cambridge, MA, USA); a mouse anti-caspase-1 p20 (1:500, Santa Cruz, CA, USA); a rabbit anti-Nrf2 (1:1000, Abcam); a rabbit anti-Kelch-like ECH associated protein 1 (Keap1) (1:1000, Abcam); a rabbit anti-β-actin (1:1000, Cell Signaling Technology, Danvers, MA, USA); a rabbit anti-histone H3 (1:2000, Cell Signaling Technology). The blots were then
incubated with goat anti-rabbit IgG H&L (HRP) (1:10000, Abcam) or goat anti-mouse H&L (HRP) (1:1000, Beyotime, Shanghai, China) for 1 h at 25 °C. The membranes were visualized using an enhanced chemiluminescence Western blot kit (eECL, Beijing ComWin Biotech Co., Ltd., Beijing, China). The band analysis was performed using the ImageJ software (National Institutes of Health, Bethesda, MD, USA).

**Caspase-1 activity assay**

On day 29 after CCI surgery, after finishing the final behavior test, the hippocampus was removed and homogenized in cold lysis buffer. The supernatant was collected after centrifugation at 20 000 ×g for 15 min. The activity of caspase-1 in the hippocampus was detected using a caspase-1 activity assay kit (Beyotime Biotechnology, Shanghai, China) according to the manufacturer’s instruction. Absorbance measurements were performed using a microplate reader (1510, ThermoFisher, Vantaa, Finland) at 405 nm.

**Enzyme-Linked Immunosorbent Assay (ELISA)**

The frozen hippocampus was homogenized in cold phosphate-buffered solution (PBS). Next, the supernatant was collected after centrifugation at 20 000 g for 10 min. The concentration of IL-1β was measured using an ELISA kit (RLB00, R&D Systems, Minneapolis, MN, USA), according to the manufacturer’s instruction.

**Detection of reactive oxygen species (ROS) generation in the hippocampus tissue**

After finishing the final behavioral test, the hippocampus was collected and frozen in liquid nitrogen using Tissue-Tek O.C.T. Compound (Sakura Finetek, VWR, Radnor, PA, USA). The hippocampus tissue was cut into 8-μm thick sections and rewarmed at 25 °C for 30 min and then incubated in ROS dye solution (1:200, Dihydroethidium, Servicebio) at 37 °C for 30 min.
The sections were placed in PBS (pH 7.4) and washed three times, with shaking, for 5 min. After washing in PBS (pH 7.4), the sections were incubated with DAPI (Servicebio) at 25 °C for 10 min, followed by a 15-min wash in PBS, mounting on slides, and covered with coverslips. The hippocampal slices were imaged using a fluorescence microscope (BX53, Olympus, Tokyo, Japan). Images were captured with a digital camera (DP74, Olympus, Tokyo, Japan). The fluorescence intensity was examined to determine the ROS levels. Data are presented as fold-changes of the fluorescence signal.

**NF-κB activity assay**

The nuclear protein of hippocampus was extracted using a nuclear extraction kit (Cayman Chemical, Ann Arbor, MI, USA), and the NF-κB activity of the hippocampus tissue was measured using a NF-κB transcription factor assay kit (Cayman Chemical) according to the manufacturer’s instruction.

**Molecular docking between albiflorin and Keap1**

The protein-ligand docking studies were performed using Discovery Studio Client software 4.0 (Nebraska, Omaha, USA) to calculate the possible interaction of albiflorin with KEAP1. Chemical structure of albiflorin (CID - 51346141) was available from PubChem database (https://pubchem.ncbi.nlm.nih.gov/). The crystal structures of Keap1 was available in the RCSB Protein Data Bank (http://www.rcsb.org/, PDB ID: 6TYP), and the protein structure is further processed by the Prepare Protein protocol of Macromolecules function module. CDOCKER protocol was used to analyze the binding affinity between albiflorin and the protein.

**Statistical analysis**
Data were analyzed using the SPSS software (17.0, IBM, Chicago, IL, USA). The behavioral data from different time point were analyzed using Two-way ANOVA analysis (timecourses with repeated measures). Statistical comparisons between the two groups were performed using the Student’s t-test. One-way analysis of variance (ANOVA) was used to compare the results of behavioral and biochemical data from multiple groups followed by the least-significant difference (LSD) post-hoc test or Dunnett’s T3 test, where appropriate. A P value < 0.05 was considered statistically significant.

**Results**

**Chronic constriction injury surgery induced neuropathic pain and anxiety and depression-like behaviors in model rats**

Mechanical allodynia (Figure 2B) was significantly induced by CCI from day 7 to 28 after surgery (two-way ANOVA, main effect group: $F_{1,18} = 318.21, P < 0.001$; main effect time: $F_{4,72} = 21.096, P < 0.001$; group × time interaction: $F_{4,72} = 16.762, P < 0.001$; Figure 2B). The open field test (OFT) was used to assess locomotor activity and anxiety-like behavior in model rats. The time CCI rats spent in the center zone of the open field (OF) was significantly reduced on days 14 and 28 after CCI surgery compared to that of the sham group (two-way ANOVA, main effect group: $F_{1,18} = 13.949, P = 0.002$; main effect time: $F_{1,18} = 3.055, P = 0.098$; group × time interaction: $F_{1,18} = 0.486, P = 0.495$; Figure 2E). In addition, the total distance traveled by the CCI rats in the OF was not significantly different to that of the sham group on day 15, but significantly decreased on day 28 after CCI surgery (two-way ANOVA, main effect group: $F_{1,18} = 10.12, P = 0.005$; main effect time: $F_{1,18} = 2.92, P = 0.105$; group × time interaction: $F_{1,18} = 2.176, P = 0.157$; Figure 2C, D). This result suggests that the nerve damage did not affect the locomotor activity of rats on day 14, while the decreased total distance on day 28 after surgery was a manifestation of a depression-like behavior.
elevated plus maze test (EPM) test was used to assess anxiety-like behavior. The time CCI rats spent in the open arms of the EPM was significantly decreased on days 14 and 28 after surgery compared to that of the sham group (two-way ANOVA, main effect group: $F_{1,18} = 14.442, P = 0.001$; main effect time: $F_{1,18} = 1.256, P = 0.277$; group × time interaction: $F_{1,18} = 0.837, P = 0.372$; Figure 2F, G). The forced swim test (FST) was used to assess depression-like behavior. FST results showed that the duration of immobility of CCI rats was significantly extended on day 29 after surgery compared to sham group (two-way ANOVA, main effect group: $F_{1,18} = 13.458, P = 0.002$; main effect time: $F_{1,18} = 9.31, P = 0.007$; group × time interaction: $F_{1,18} = 8.27, P = 0.01$; Figure 2H). Altogether, these results indicate that CCI surgery not only induces neuropathic pain but also anxiety and depression-like behaviors in model rats.

CCI surgery induced activation of the hippocampal NLRP3 inflammasome

To test whether the NLRP3 inflammasome was activated in mood disorders associated with neuropathic pain, we examined activation of NLRP3 inflammasome in the hippocampus. NLRP3 (but not NLRP1, Supplementary Figure S1) and caspase-1 expression were significantly increased in hippocampal neurons in CCI rats compared to sham group (Figure 3 A–C). Moreover, CCI surgery significantly enhanced caspase-1 activity and increased IL-1β levels in the hippocampus on day 29 after surgery (Figure 3 D, E). To further confirm the cellular localization of the NLRP3 inflammasome after CCI surgery, we used double immunofluorescence for NLRP3 and the neuronal marker NeuN, and caspase-1 and NeuN. NLRP3 and caspase-1 were mainly expressed in the CA1 layer and dentate gyrus (DG) regions of the hippocampus and predominantly colocalized with NeuN (Figure 3F–I). These
results indicate that hippocampal NLRP3 inflammasomes are activated in the state of neuropathic pain associated with anxiety/depression-like behaviors.

**MCC950 and Albiflorin relieved CCI-induced pain, anxiety and depression-like behavior and suppressed hippocampal NLRP3 inflammasome activation**

To investigate the role of the NLRP3 inflammasome in CCI-induced mood disorders, MCC950, a selective NLRP3 inflammasome inhibitor, was administered by i.p. injection to CCI rats (Figure 4A). The behavioral tests were conducted after MCC950 treatment following the experimental scheme shown in Figure 4A. As illustrated in Figure 4B, administration of MCC950 relieved CCI-induced mechanical allodynia during days 21 to 28 after surgery (two-way ANOVA, main effect group: F\(_{5,48} = 164.649, P < 0.001\); main effect time: F\(_{4,192} = 41.702, P < 0.001\); group × time interaction: F\(_{20,192} = 10.755, P < 0.001\)). Meanwhile, MCC950 significantly reduced the anxiety and depression-like behavior induced by CCI surgery on days 28 and 29 after CCI (Figure 4D–F). In addition, MCC950 significantly reduced the expression of NLRP3 and caspase-1 in the hippocampus on day 29 after CCI (Figure 4G–I). Furthermore, MCC950 treatment decreased the CCI-elevated activity of caspase-1 and the content of IL-1β in the hippocampus (Figure 4J, K). Broadly analogous to that of MCC950, administration of albiflorin significantly relieved CCI-induced pain, anxiety and depression-like behavior and suppressed hippocampal NLRP3 inflammasome activation (Figure 4B–K). Moreover, both the administration of albiflorin and MCC950 had no significant effects on the behavior nor hippocampal NLRP3 inflammasome activation of sham-operated rats (Supplementary Figure S2).

Altogether, these results demonstrated that NLRP3 inflammasome activation was involved in the pathological process of neuropathic pain associated with anxiety and
depression-like behavior. In addition, the effect of albiflorin in attenuating the mood disorder under neuropathic pain state was related to the inhibition of the NLRP3 inflammasome.

Albiflorin promotes Nrf2 translocation into the nucleus and limits the elevated level of ROS and activity of NF-κB in the hippocampus

To investigate the role of albiflorin in the inhibition of the NLRP3 inflammasome, we examined the effect of albiflorin on the Keap1-Nrf2 signaling pathway. As illustrated in (Figure 5A, B), albiflorin treatment significantly limited the increased levels of ROS induced by CCI in the hippocampus. In addition, albiflorin administration clearly enhanced the translocation of Nrf2 into the nucleus but did not affect the total expression levels of Nrf2 and Keap1. These results suggest that albiflorin activates Keap1-Nrf2 signaling pathway thus attenuating the ROS levels after CCI. In addition, albiflorin significantly limited the activity of hippocampal NF-κB (Figure 5G). Furthermore, albiflorin had no significant effect on the sham group (Figure 5A–G).

Molecular docking of albiflorin to Keap1

To further investigate the possible mechanism of albiflorin in the regulation of the Keap1-Nrf2 signaling pathway, we used molecular docking to examine the binding modes of albiflorin with Keap1. As shown in Figure 5H, Albiflorin is predicted to dock with KEAP1 Kelch domain, a recognition module for Nrf2 (CDOCKER_INTERACTION_ENERGY: -50.142 kcal / mol). Figure 5I, J show the predicted binding sites of albiflorin to Keap1. These data suggest that albiflorin displays non-covalent binds to Keap1.

Discussion
The NLRP3 inflammasome is the most widely implicated regulator of caspase-1 activation. Excessive NLRP3 inflammasome activity leads to systemic "sterile" inflammation in both periphery and brain (Youn et al., 2013). The hippocampus is an important brain region in stress response and adjustment for emotion and memory (Liu et al., 2013; Pan et al., 2014). Some animal studies reported that harmful chemicals (including lipopolysaccharides [LPS]) or chronic stress can lead to NLRP3 inflammasome activation in the hippocampus. Meanwhile, animal models displayed anxiety and depression-like behavior and memory loss symptoms. Suppressing the activation of the NLRP3 inflammasome exerted neuroprotective effects and improved the aforementioned neurological symptoms (Lu et al., 2014; Zhang et al., 2014; Xu et al., 2016; Liu et al., 2019; Fu et al., 2020; Li et al., 2020). In addition, several recent studies have shown that the NLPR3 inflammasome is involved in the pathogenesis of neuropathic pain, and inhibition of NLPR3 inflammasome can effectively alleviate nociceptive hypersensitivity (Jia et al., 2017; Zhang et al., 2017; Grace et al., 2018; Khan et al., 2018; Pan et al., 2018; Chen et al., 2019a, 2019b). Although many studies have been conducted on NLRP3 inflammasome's involvement in the pathological process of neuropathic pain, anxiety, and depression, so far there are no studies on NLPR3 inflammasome's involvement in neuropathic pain associated with anxiety and depression-like behaviors. A recent study indicated that the hippocampal NLRP1 inflammasome pathway plays a pivotal role in the development of anxiety and depressive behaviors after chronic neuropathic pain (Li et al., 2019a). To our surprise, our present data showed that CCI significantly induced elevated NLPR3, instead of NLRP1 (Supplementary Figure S1), expression in hippocampal neurons, contributing to anxiety and depression-like behaviors. To further confirm that the activation of NLRP3 inflammasome contributes to these mood disorders, we treated CCI rats with a selective NLRP3 inhibitor. We found that suppressing
the activation of the NLRP3 inflammasome not only significantly decreased the hyperalgesia but also attenuated the anxiety and depression-like behaviors in CCI rats. These results indicate that the activation of NLRP3 inflammasome is involved in the pathological process of anxiety and depression-like behaviors associated with neuropathic pain. Moreover, NLPR3 inflammasome might be a potential therapeutic target for neuropathic pain-associated anxious and depressive disorders.

Albiflorin and paeoniflorin are the main monoterpene glycosides among the glucosides contained in *P. alba* Radix. The anti-inflammatory and antidepressant effects of albiflorin and paeoniflorin have been well documented in existing reports (Wang et al., 2019; Zhang and Wei, 2020). Albiflorin displays more powerful antidepressant activity than its isomer paeoniflorin (Wang et al., 2016). However, there has been no study on albiflorin's role in the regulation of NLRP3 inflammasome. Our results show that albiflorin effectively alleviates neuropathic pain induced by CCI surgery, which is consistent with results of a previous study (Zhou et al., 2016). Importantly, albiflorin significantly inhibited the activation of NLRP3 inflammasome in the hippocampus and relieved the anxiety-depressive behavior of model rats. These results suggest that albiflorin produces these effects by inhibiting the activation of NLPR3, similar to the action of the selective NLRP3 inhibitor MCC950.

It is well documented that ROS are pivotal upstream events implicated in the activation of NLRP3 (Dostert et al., 2008; Zhou et al., 2011; Groß et al., 2016). ROS are continuously released by mitochondria as byproducts of oxidative phosphorylation. Under physiological conditions, cellular antioxidants reduce ROS, maintaining the redox level balance. During the inflammatory response, excessive production of ROS causes oxidative stress, which usually leads to NLRP3 activation (Dostert et al., 2008; Groß et al., 2016). In addition to ROS, the
Keap1-Nrf2 pathway participates in the regulation of the inflammasome. Under physiological conditions, Nrf2 is sequestered in the cytosol and maintained at a low level through a Keap1-Nrf2 protein to protein interaction. Under oxidative stress conditions, Keap1 is chemically modified and releases Nrf2, which results in Nrf2 translocating into the nucleus. Nrf2-driven gene transcription regulates the levels of antioxidant genes limiting ROS levels, thus, inhibiting NLRP3 activation (Liu et al., 2017; Chu et al., 2019; Dang et al., 2019). Moreover, Nrf2 attenuates the activation of NF-κB; this leads to suppression of the inflammasome activity by downregulating the expression of the inflammasome components NLRP3, caspase-1, and pro-IL-1β (Li et al., 2008; Chu et al., 2019; Dang et al., 2019). In addition to its role in NLRP3 inflammasome, the Kea1-Nrf2 pathway is also involved in depression in preclinical and clinical studies (Bansal et al., 2019; Chu et al., 2019; Dang et al., 2019; Swanson et al., 2019; Zborowski et al., 2020), which implies that interrupting the Keap1-NRF2 protein-protein interaction might be a potential therapy against depression. Furthermore, a recent animal study indicated that the Nrf2-NLRP3 signaling pathway, which is responsible for adjusting inflammation levels, plays a vital role in the depression induced by environmental factors (Chu et al., 2019). Albiflorin has shown strong antioxidant activity in in vitro and in vivo studies (Suh et al., 2013; Ho et al., 2015; Hong et al., 2017; Xu et al., 2019b). However, to our knowledge, no previous reports have shown that albiflorin activates the Keap1-Nrf2 pathway to regulate the NLRP3 inflammasome. In the present study, albiflorin treatment significantly promoted the Nrf2 translocation to the nucleus and reduced the levels of hippocampal ROS and caspase-1 activity. To our surprise, albiflorin had no significant effect on Keap1 or total Nrf2 expression. However, molecular docking revealed that albiflorin displayed affinity for Keap1. These data suggest that albiflorin may disrupt Keap1-Nrf2 interaction through a non-covalent binding, which deserves further
investigation. In addition, consistent with previous studies (Zhou et al., 2016; Cai et al., 2019), our present results indicate that albiflorin also markedly reduces the elevated NF-κB activity induced by CCI surgery. Moreover, the current results indicate that the regulation of antioxidant levels may be one of the mechanisms through which albiflorin inhibits the activation of the NLRP3 inflammasome.

In conclusion, our study confirms that activation of the NLRP3 inflammasome is involved in CCI-induced anxiety and depression. The administration at analgesic dose of albiflorin simultaneously exerted an antidepressant and anxiolytic-like activity. Our present data also demonstrated that inhibiting the activation of the NLRP3 inflammasome in the hippocampus is highly associated with the therapeutic effects of albiflorin. Albiflorin represents a potential therapeutic agent in the treatment of neuropathic pain conditions, which might greatly improve the overall symptoms of neuropathic pain with relevant clinical benefit.

Supplementary Materials
Supplementary data are available at International Journal of Neuropsychopharmacology (IJNPPY) online.

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Statement of Interest

None.

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**Figure Legends**
Figure 1. Chemical structure and HPLC chromatogram of albiflorin

Figure 2. Chronic constriction injury (CCI) surgery induced neuropathic pain and anxiety and depression-like behaviors in model rats. The scheme of this animal experiment is presented in panel A. (B) Paw withdrawal threshold (PWT) was measured using an electronic Von Frey Aesthesiometer. (C) Representative tracks showed the horizontal movement traces in the open field test (OFT) of Sham and CCI rats. (D) Total distance traveled (cm) in the OFT by sham and CCI rats. (E) Time (s) spent in the central zone. (F) Representative tracks showed the horizontal movement traces in the elevated plus maze test (EPM) of Sham and CCI rats. (G) Time (s) spent in the open arms. (H) Time (s) of immobility in the forced swim test (FST).

Data are expressed as the mean ± SE for n = 10 rats/group. *P < 0.05, **P < 0.01 and ***P < 0.001 compared to the sham group.

Figure 3. Chronic constriction injury (CCI) surgery induced activation of the hippocampal NLRP3 inflammasome. (A) Representative Western blot bands of NLRP3 and caspase-1 (p20). Quantitative analysis of Western blot expression of NLRP3 (B) and caspase-1 (C) in sham-operated and CCI-operated rats. The fold change in the protein band density was normalized to that of β-actin, and the results are presented as fold change from the sham group (n = 3 rats/group). Activity of caspase-1 (D) and levels of interleukin-1β (E) in the hippocampus (n = 5 rats/group). Data are expressed as the mean ± SD; *P < 0.05, **P < 0.01, and ***P < 0.001 compared to the sham group. Double-immunofluorescence representative photomicrographs displaying the cellular localization of NLRP3 (red) and NeuN (green) (F, G)
and caspase-1 (red) and NeuN (green) (H, I) on day 29 after CCI. Photomicrographs are of the hippocampus of rats with CCI.

**Figure 4.** MCC950 and Albiflorin (AF) relieve chronic constriction injury (CCI) - induced pain, anxiety and depression-like behavior and suppress hippocampal NLRP3 inflammasome activation. (A) The scheme of this animal experiment. (B) Paw withdrawal threshold (PWT) was detected 1 day before chronic constriction injury (CCI) and on days 7, 14, 21, and 28 after CCI surgery. (C) Total distance traveled in the open field test (OFT) of sham and CCI rats. (D) Time spent in the central area of the OF. (E) Time spent in the open arms of the elevated plus maze (EPM). (F) Time of immobility in the forced swim test (FST). Data are expressed as the mean ± SE for n = 8 - 12 rats/group; * P < 0.05, ** P < 0.01, and *** P < 0.001 compared to the sham group; # P < 0.05, ## P < 0.01 compared to the CCI group. (G) Representative Western blot bands of NLRP3 and caspase-1 (p20). Quantitative analysis of Western blot expression of NLRP3 (H) and caspase-1 (I) in sham-operated and CCI-operated rats. The fold change in the protein band density was normalized to that of β-actin. The activity of caspase-1 (J) and levels of interleukin (IL)-1β (K) in the hippocampus. Data are expressed as the mean ± SD of n = 4 rats/group; ** P < 0.01, *** P < 0.001 compared to the sham group; # P < 0.05, ## P < 0.01 and ### P < 0.001 compared to the CCI group.

**Figure 5.** Albiflorin (AF) promotes Nrf2 translocation into the nucleus and limits the elevated level of reactive oxygen species (ROS) and activity of NF-κB in the hippocampus. (A) Representative photographs of hippocampal sections of ROS. (B) Quantification of ROS.
immunoreactivity in the CA1 layer and dentate gyrus (DG) regions of the hippocampus. (C) - (F) Western blot analysis showing the expression of total and nuclear Nrf2 and Keap1 in the hippocampus on day 29 after CCI. The fold changes in the density of total Nrf2 and Keap1 are normalized to that of β-actin, and that of nuclear Nrf2 is normalized to that of histone H3. (G) NF-κB activity. The results are presented as fold changes compared to those of the sham group. Data are expressed as the mean ± SD for n = 4 rats/group; **P < 0.01, ***P < 0.001 compared to the sham group; # P < 0.01, ## P < 0.01 compared to the CCI group. (H) - (J) Molecular docking of AF to Keap1. (H) KEAP1 Kelch domain in complex with AF (CDOCKER_INTERACTION_ENERGY: -50.142 kcal/mol). (I) Magnified portion of 3D interaction map between AF and 6TYP binding site. (J) 2D diagram of docking structure of AF with 6TYP.
Figure 1

Albiflorin 97.6%

8.100 min
Figure 2

A

B

C

D

E

F

G

H

-1 0 7 14 15 21 28 29

PWT PWT PWT PWT

CCl surgery

Collecting hippocampus

Postoperative days (d)

15 day after CCl

28 day after CCl

Sham CC

OFT

Distance travelled (cm)

14 28

Postoperative days (day)

Time in center zone (s)

14 28

Postoperative days (day)

Sham EPM CCI

15 day after CCI

28 day after CCI

Time in open arms (s)

14 28

Postoperative days (day)

FST

Immobility duration (s)

15 29

Postoperative days (day)

Sham CCI
Figure 3

A

NLRP3
caspase-1 (p20)
β-actin

Sham    CCI

B

NLRP3 / β-actin
relative to Sham

Sham    CCI

C

caspase-1 / β-actin
relative to Sham

Sham    CCI

D

Caspase-1 activity
relative to Sham

Sham    CCI

E

IL-1β ng/total protein

Sham    CCI

F

CA1    NLRP3    NeuN

Sham    CCI

G

DG    NLRP3    NeuN

Sham    CCI

H

CA1    caspase-1    NeuN

Sham    CCI

I

DG    caspase-1    NeuN

Sham    CCI
Figure 5

A) Sham+Vehicle  |  Sham+AF  |  CCI+Vehicle  |  CCI+AF  
CA1                      
DG                      

B) ROS immunoreactivity relative to Sham

C) Keap1

D) Keap1/β-actin relative to Sham

E) Total Nrf2/β-actin relative to Sham

F) Nuclear Nrf2/Histone relative to Sham

G) NF-κB activity relative to Sham

H) Molecular model of the protein

I) Molecular model of the protein

J) Molecular docking results
