Atopic dermatitis (AD), a common chronic inflammatory skin disease, is characterized by inflammatory cell skin infiltration. The JAK-STAT pathway has been shown to play an essential role in the dysregulation of immune responses in AD, including the exaggeration of Th2 cell response, the activation of eosinophils, the maturation of B cells, and the suppression of regulatory T cells (Tregs). In addition, the JAK-STAT pathway, activated by IL-4, also plays a critical role in the pathogenesis of AD by upregulating epidermal chemokines, pro-inflammatory cytokines, and pro-angiogenic factors as well as by downregulating antimicrobial peptides (AMPs) and factors responsible for skin barrier function. In this review, we will highlight the recent advances in our understanding of the JAK-STAT pathway in the pathogenesis of AD.

## Introduction

Atopic dermatitis (AD) is a common skin disease manifested clinically by chronic inflammation and characterized histologically by skin infiltration of inflammatory cells including predominantly lymphocytes, eosinophils, and mast cells. It affects 10–20% of children and 1–3% of adults in developed countries, and the prevalence is increasing. Many of these patients also suffer from asthma and allergic rhinitis along with intense itching and skin infection. Although the pathogenesis and etiology of AD remain to be completely understood, this multifactorial disease likely results from a complex crosstalk between genetic and environmental factors. Exaggerated Th2 response, disruption of the epidermis barrier functions, high levels of serum IgE, and decreased production of antimicrobial peptides (AMPs) are the key findings in AD. Data from human and animal studies indicate that AD is a Th2 dominant inflammatory skin disease at the acute stage followed by Th1 involvement at the chronic stage of the disease. Specifically, this notion of Th2-dominance in AD is validated by a mouse model we have successfully generated through overexpressing IL-4, a key Th2 cytokine, in the basal epidermis using a epidermis-specific keratin 14 promoter. The IL-4 transgenic (Tg) mice spontaneously develop skin lesions, serological abnormalities, and skin infection, which fulfill the clinical and histological diagnostic criteria for human AD. Consistent with an inflammatory disease process, upregulation of chemokines, proinflammatory cytokines, B cell activation molecules, angiogenic factors, and several critical adhesion molecules has been found in these Tg mice.

The JAK-STAT pathway is a classical signal transduction pathway for numerous cytokines and growth factors. The binding of ligands to their receptors leads to JAK activation, which in turn phosphorylates and activates STATs. The activated STATs then translocate to the cell nucleus to regulate their target genes. The JAK family includes JAK1, JAK2, JAK3, and TYK2, and the STAT family includes STAT1, STAT2, STAT3, STAT5A/B, and STAT6. As a negative regulator of the JAK-STAT pathway, the SOCS family consists of cytokine-inducible SH2 domain-containing protein (CIS) and SOCS1–7. SOCSs may act on the activation loop of JAKs, or they may interact with phosphotyrosines in the cytoplasmic domain of cytokine receptors, suppressing the JAK-STAT pathway with a negative feedback mechanism. In this review, we will discuss the roles that the JAK-STAT pathway plays in the pathophysiology of AD.

## JAK-STAT in Th2 Differentiation

Since AD is a Th2-dominant disease, examination of how the JAK-STAT pathway regulates Th2 differentiation would help us to understand the possible roles that JAK-STAT play in AD. It is well established that IL-4 promotes the differentiation of Th2 cells, which in turn produce IL-4, IL-5, IL-10, and IL-13. The study of IL-4 and the IL-4 receptor α-null mice demonstrated clearly that IL-4 is required for a Th2 response and the production
of Th2 cytokines. In addition, studies from the IL-4 Tg mice demonstrated upregulation of the Th2 cytokines. Consistently, the IL-4 Tg mice generated on a Th2-dominant strain, BALB/c mice, develop earlier onset and worse AD-like skin lesions than the IL-4 Tg mice generated on a Th1-dominant strain, C57BL/6 mice, suggesting that the Th2 systemic immune milieu, in addition to cutaneous Th2 immune milieu, plays an essential role in the pathogenesis of AD. Additionally, knocking out Bcl-6, a transcription factor functioning to regulate IL-4 signal transduction, leads to Th2-mediated hyperimmune response in many organs, further supporting the regulatory function of IL-4 on Th2 immunity. Importantly, IL-4 signals through the JAK-STAT6 pathway to regulate Th2-related target genes in lymphocytes, firmly supporting a role of JAK-STAT in Th2 regulation.

In addition to IL-4, other factors are also important for Th2 differentiation. Thymic stromal lymphopoietin (TSLP), known to promote Th2 differentiation, to activate natural killer T cells and basophils, and to affect B cell maturation, has been reported to be associated with AD. While the murine TSLP receptor signals via STAT3, STAT5/Tec, a Src type kinase, the human TSLP receptor activates STAT1, STAT3, STAT5/JAK1, and JAK2.

It has been reported that histamine enhances the secretion of Th2 cytokines such as IL-4, IL-5, IL-10, and IL-13 and inhibits the production of Th1 cytokines. Histamine was shown to stimulate IL-13 production through the JAK-STAT pathway in a murine Th2 cell line. In addition, tyrophostin, a JAK-STAT pathway inhibitor, reversed the effects of histamine on IFNy, IL-5, and IL-10 production. Horr et al. have shown that histamine, by acting through the H4 receptors on T cells, can suppress STAT1 activation and help to drive the Th2 response, leading to the development of AD.

It is generally accepted that TYK2, JAK2, and STAT4, which are IL-12 signaling pathway components, are essential for Th1 cell differentiation, while JAK1, JAK3, and STAT6, which are IL-4 signaling components, are critical for Th2 differentiation. In addition, STAT5A/B, which are involved in the differentiation, IgE class switching, and MHC class II production. While STAT4 null mice fail to generate Th1 cells and conditional STAT3 knockout mice have an exaggerated Th1 response, STAT6 null mice have impaired Th2 cell development and IgE class switching. On the contrary, mice expressing constitutively active STAT6 (STAT6VT) develop an AD-like skin lesion. In addition, several STAT6 polymorphisms are also associated with a predisposition to allergic diseases and high levels of IgE.

JAK3 is expressed in natural killer cells, T cells, and B cells. A JAK3 mutation in severe combined immunodeficiency (SCID) patients leads to the absence of T cells and natural killer cells and the production of dysfunctional B cells. JA3K-null mice also have similar defects.

In humans, a loss-of-function mutation in TYK2 leads to defects in multiple cytokine signaling pathways including type I IFN, IL-6, and IL-12. These patients showed impaired Th1 differentiation and accelerated Th2 differentiation with clinical manifestations including unusual susceptibility to various microorganism infections, AD-like skin lesions, and high serum IgE levels, suggesting a regulatory role of the JAK family in Th2 differentiation.

Another TYK2 null mutation and hypomorphic mutations in STAT3 are associated with the hyper-IgE syndrome (HIES), a primary immunodeficiency characterized by AD-like skin lesions, susceptibility to infections, and high serum IgE levels. In this syndrome, IL-12 and IFN signaling pathways are defective, thus blocking the Th1 differentiation. As a result, the Th2 response is exaggerated, leading to similar clinical findings as AD. Because STAT3 is important for B cell proliferation and differentiation, HIES patients have decreased number of memory B cells as compared with AD patients.

As a negative regulator of the JAK-STAT pathway, suppressor of cytokine signaling 2 (SOCS2) deletion leads to increased susceptibility to AD as compared with wild-type mice. These mice show an exaggerated Th2 response along with significantly elevated serum IgE levels, eosinophilia and low IL-17 levels. SOCS2-knockout CD4+ T cells display constitutively high levels of STAT6 phosphorylation as compared with wild type T cells. In addition, IL-2-induced STAT5 activation, also involved in the early Th2 differentiation, is exaggerated in the SOCS2-knockout CD4+ T cells. On the other hand, IL-12-stimulated STAT4 phosphorylation is unaffected, and IL-6 mediated STAT3 phosphorylation is inhibited. Since it is generally accepted that the development of the Th1, Th2, and Th17 immune responses are mediated by STAT4/STAT1, STAT6/STAT5, and STAT3 respectively, the increased STAT6/STAT5 activation and suppressed STAT3 activation naturally lead to an exaggerated Th2 response at the expense of Th1.

Another example of the JAK family involvement in Th2 immunity is shown in NC/Nga mice that develop AD-like skin inflammation. Nakagawa et al. reported that Pyridone 6 (P6), a pan-JAK inhibitor, delayed the onset and reduced the severity of AD-like skin lesions in NC/Nga mice. P6 suppressed the IFN-γ/STAT1, IL-2/STAT5, and IL-4/STAT6 signaling pathways strongly while it suppressed IL-6/STAT3 pathway only modestly, resulting in reduction of the Th1 and Th2 responses but the enhancement of the Th17 response.

JAK-STAT in Eosinophils

Eosinophil skin infiltration is frequently observed in the skin lesions of AD patients and AD mouse models. Although STAT6 null mice have defects in Th2 differentiation and IgE class switching, crossing these mice with NC/Nga mice cannot prevent the development of AD-like skin lesions. Even though these mice do not produce IgE and Th2 cytokines, the histological features of their skin lesions are similar to that of AD. These authors suggested that a Th2 response is not absolutely necessary for the development of AD-like skin lesions; instead, IFN-γ and eosinophil skin infiltration may play an essential role.
Eosinophils express the IL-31 receptor A (IL-31RA), and IL-31 has the ability to delay the apoptosis of eosinophils and to stimulate the secretion of pro-inflammatory cytokines and chemokines through JAK-STAT. In addition, IL-5, mainly involved in B cell proliferation and differentiation, was shown to induce eosinophilia when upregulated. Conversely IL-5 or IL-5 receptor α chain (IL-5Rα) null mice show defects in B cells and eosinophils. In eosinophils, IL-5 signals through the JAK2-STAT1/STAT5 and MAP kinases pathways to regulate genes involved in cell proliferation, survival and effector functions.

JAK-STAT in Tregs

In the skin, regulatory T cells (Tregs) help to regulate the immune response, and in AD, Treg function is suppressed. Depleting Treg cells significantly increased the severity of the skin inflammation in OVA-sensitized mice. In T cells, IL-4-activated STAT6 upregulates GATA3, the master regulator of Th2 cells. GATA3 in turn suppresses Foxp3, the master regulator in Treg cells. This regulatory pathway could possibly explain why Treg function is suppressed in AD. In addition, STAT5A/B and STAT3 may also play important roles in the regulation of Foxp3. On the other hand, Foxp3 can also suppress GATA3’s ability to regulate Th2 genes. It was reported that the IFN-γ gene transfer stimulates Treg related cytokines and improves AD-like symptoms of NC/Nga mice. This finding, however, seems to contradict the notion that IFN-γ is a promoting factor for AD. On the clinical level, treatment with a low-dose cyclosporine A is effective in reducing skin inflammation with simultaneous increase of Treg population in AD patients, further supporting a role of Treg in AD.

JAK-STAT in Th17 Cells

STAT3, mediating the IL-23 and IL-6 pathways, is critical for the differentiation of Th17 cells. In the early stage of human AD, the number of Th17 cells is increased, and IL-17 has been reported to upregulate adhesion molecules on keratinocytes, enhancing T cell-keratinocyte adhesion and T cell-mediated cytotoxicity. However, in chronic AD, the Th17 pathway is suppressed, which could account for the AMP deficiency in AD.

JAK-STAT in Mast Cells

In the skin lesions of AD patients as well as AD mouse models, the number of mast cells is increased. IL-9 stimulates VEGF production from human mast cells through the activation of STAT3 in an in vitro experiment. Interestingly, IL-9 serum levels in AD patients are not different from those of non-AD controls even though IL-9 and its receptors are upregulated in the lesional skin of the AD patients, suggesting that the upregulation of VEGF in mast cells via IL-9 may be a localized cutaneous phenomenon. The prominent increase of mast cell VEGF production and angiogenesis in an AD animal model also supports the role of JAK-STAT in this immune pathway.

JAK-STAT in Epidermal Keratinocytes

Being a major cell type of the skin and possessing the ability to participate in various immune responses, epidermal keratinocytes are likely an active player in the skin inflammation of AD. We and others have shown that the dysregulation of the keratinocyte genes by IL-4 play an important role in AD. Two types of IL-4 receptors have been identified. The type I receptor consisting of IL-4Rα and IL-2Ryc chains is expressed in hematopoietic cells, while the type II receptor consisting of IL-4Rα and IL-13Rα1 is expressed in other cell types including keratinocytes.

In keratinocytes, both IL-4 AND IL-13 binds to IL-4Rα and IL-13Rα1. IL-13Rα2, which was reported as a dominant negative inhibitor for both IL-4 and IL-13, binds to IL-13 with high affinity, but lacks a significant cytoplasmic domain. We have previously reported that IL-13Rα2 is upregulated by IL-4 in keratinocytes. In AD patients this gene is similarly upregulated. This may be due to a negative feedback mechanism available to keratinocytes to maintain homeostasis.

We have also shown that IL-4 upregulates chemokines CCL8, CCL24, CCL25, CCL26, CCL3L1, CXCL6, and CXCL16, all known for their roles in the pathophysiology of AD. The eotaxin subfamily that consists of CCL11/eotaxin-1, CCL24/eotaxin-2, and CCL26/eotaxin-3 plays an important role in AD. Eotaxins bind to the chemokine cysteine–cysteine receptor 3 (CCR3), which are expressed predominantly on eosinophils, recruiting these cells to the inflammatory site. Skin infiltrating eosinophils release a variety of cytotoxic granule proteins to cause tissue damage. In the regulation of CCL26, though both STAT3 and STAT6 are phosphorylated by IL-4, only STAT6 is functionally activated. Detailed analysis of the promoter of the CCL26 gene has shown that the STAT6 response element consists of two palindromic half sites TTC and GAA spaced by four nucleotides. Others also reported that STAT6, different from other STAT members, prefers the STAT sites with spaced by four nucleotides. In addition to tyrosine phosphorylation, STAT3 activation may also involve serine phosphorylation and lysine acetylation. Knockout mice studies indirectly support our results in that while STAT6 null mice demonstrated defects in eosinophil tissue infiltration, the STAT3 Cre-loxP knockout mice did not show similar defects. Furthermore, IL-4 and IL-13 double knockout mice, when sensitized with ovalbumin, develop a skin lesion characterized by intact CD4+ dermal infiltration, severely diminished eosinophil infiltration, and undetectable OVA-specific IgE levels, suggesting that STAT6 activation is required for eosinophil skin infiltration in AD. In addition to chemokines, IL-4 also upregulates pro-inflammatory factors, such as IL-1α, IL-19, IL-20, IL-25, IL-27, IL-12Rβ2, IL-31RA, and nitric oxide synthase 2 (NOS2). Moreover, we demonstrated that IL-4 signals through the JAK-STAT pathway to regulate IL-19 expression in keratinocytes.

Using PCR array, we have found that that IL-4 upregulates angiogenic or lymphangiogenic-related genes, such as VEGF, ANG-1, ANGL-4, IGF1, Notch4, PFG, and MCP-1. Using...
the IL-4 Tg mice and cell culture, we showed that CD11b+ macrophages attracted to the skin lesion by MCP-1 may account for lymphangiogenesis observed in AD by secreting VEGF-C.71 Interestingly, filaggrin, which is reduced in acute AD skin lesion, is also downregulated by IL-4, IL-10, and IL-13.88 In addition, HBD-2 and human cathelicidin (LL-37) were also found to be reduced in AD skin lesions as compared with psoriasis.89 Howell et al. suggested that low levels of LL-37 suppressed by IL-4 and IL-13 through STAT6 may be the mechanism responsible for increased vaccinia virus replication as occurred in eczema vaccinatum of AD patients.90

Skin barrier function defects are critical for the development of AD.91 In addition to the filaggrin mutations in some AD patients, cytokines are now known to induce the downregulation of several barrier proteins. Loricrin and involucrin are two important factors for skin barrier function. They are downregulated by IL-4 and IL-13 in keratinocytes, and in STAT6 transgenic mice, their expression is significantly decreased as well.92 In addition, filaggrin, which is reduced in acute skin lesion, is also suppressed by IL-4 and IL-13 in keratinocytes.93 Interestingly, while IL-4 markedly reduces ceramide levels in the epidermis by downregulating the expression of serine-palmitoyl transferase-2, acid sphingomyelinase, and β-glucocerebrosidase, Th1 cytokines (GM-CSF/IFN-γ/TNF-α) increase the ceramide levels.94

Taken together, the role that IL-4 plays in the dysregulation of keratinocyte functions in AD involves the upregulation of chemokines, pro-inflammatory factors, and pro-angiogenic factors and the downregulation of AMPs and factors responsible for skin barrier function.

Pruritus in AD is induced by complicated crosstalks among keratinocytes, immune cells, and nerve fibers.95 The data from the Nc/Nga mice studies indicate that although IL-4 is a key mediator of the inflammatory process in AD, IL-31 might be the key causative factor for pruritus.96 In addition, IL-31 transgenic mice developed a Th2 immune response with severe pruritic skin lesions similar to AD.97

IL-31 is upregulated in the skin of AD patients, and it induces the itching sensation either by direct stimulation of the dorsal root ganglion fibers that express IL-31RA or by indirect stimulation of pruritic factors from keratinocytes.98,99 IL-31 receptor activation in keratinocytes induces calcium influx and STAT3 mediated β-endorphin production, which may contribute to the peripheral itching in AD.98

Conclusion

Thus far, data from our laboratory and others seem to indicate an influential role of JAK-STAT in the pathogenesis of AD, with IL-4 being a key mediator (Figs. 1 and 2). Specifically, the JAK-STAT pathway is instrumental for the Th2 cell differentiation. In the immune milieu of AD, the enhancement of Th2 cell proliferation and their release of various cytokines via the JAK-STAT pathway is likely the critical factor for the inflammatory responses in AD. This Th2 immune upregulation could then lead to B cell maturation and plasma cell differentiation, resulting in the hyper-secretion of IgE. IgE binding to skin mast cells causes histamine release, which further exacerbates AD. Similarly, this hyper Th2 immune milieu triggers epidermal cells to release various chemokines, pro-inflammatory cytokines, and angiogenic factors that participate in AD pathophysiology. Moreover, IL-5 released from this hyper Th2 milieu could activate eosinophils that are attracted to the skin by the cotaxin subfamily, further worsening the AD condition. In addition, by way of IL-31, an inducer of pruritus, AD becomes increasingly intensified.

Our understanding of the JAK-STAT pathway and its relationship to the dysregulated immune response and keratinocyte function is still in its infancy. We have many unanswered questions. What are JAKs’ and STATs’ tissue-specific functions? How does the JAK-STAT pathway crosstalk with other pathways in AD? What exact roles do the different JAKs and STATs play in the pathogenesis of AD? Answering these questions will enable us to target more specifically the key components of the complex pathophysiology of AD, thus providing the best treatment for this disease.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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Figure 1. Proposed mechanism of JAK-STAT involvement in atopic dermatitis (AD) development, part I. Skin barrier function defects are critical for the development of AD. In addition to the filaggrin gene mutation defects in some AD patients, IL-4 is also able to downregulate barrier proteins filaggrin, loricin and involucrin through the JAK-STAT pathway making the epidermis more penetrable by allergens and pathogens. Once penetrated through the epidermis, allergens/pathogens are detected by dendritic cells, which become subsequently activated to present these antigens to naïve Th0 cells. The Th0 cell can then differentiate into the Th2 cell through the JAK1,3-STAT6 pathway under the influence of IL-4. In the Th0 cells, the STAT6 pathway can also upregulate GATA3, a master regulator of Th2 cells. GATA3 in turn suppresses Foxp3, the master regulator in Treg cells, thus allowing more T cells to be activated. Black arrows indicate activation pathway. Red lines indicate inhibition pathway.
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Figure 2. Proposed mechanism of JAK-STAT involvement in atopic dermatitis (AD) development, part II. Here, Th2 cells play a significant role. By their abilities to provide IL-4 and IL-5 stimulation via the JAK-STAT pathway, immature B cells could be differentiated into mature B cell and plasma cells would undergo antibody heavy chain switching to IgE class. The subsequent binding of IgE to skin mast cells could lead to release of histamine, which is known to exacerbate AD. Similarly, this hyper Th2 immune milieu, particularly IL-4, could trigger epidermal cells to produce and release various chemokines (such as CCL26), pro-inflammatory cytokines, and angiogenic factors, leading to AD pathophysiology. Moreover, IL-5 released from this hyper Th2 milieu could, through JAK-STAT pathway, activate eosinophils, and attract them to the skin via CCL26, further worsening the AD condition. In addition, by way of IL-31, an inducer of pruritus, AD symptoms could become increasingly intensified. Black arrows indicate activation pathway. Red lines indicate inhibition pathway.
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