Mechanism of the Reduction of the Native Intermediate in the Multicopper Oxidases: Insights into Rapid Intramolecular Electron Transfer in Turnover

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Supporting Information

ABSTRACT: The multicopper oxidases (MCOs) are the family of enzymes that catalyze the 4-electron reduction of O₂ to H₂O coupled to the four 1-electron oxidations of substrate. In the catalytic cycle electrons are transferred intramolecularly over ~13 Å from a Type 1 (T1) Cu site that accepts electrons from substrate to a trinuclear Cu cluster (TNC) where O₂ is reduced to H₂O at rapid rates consistent with turnover (560 s⁻¹). The oxygen reduction mechanism for the MCOs is well-characterized, whereas the rereduction is less understood. Our initial study of *Rhus vernicifera* Laccase (Heppner et al. *J. Am. Chem. Soc.* 2013, 135, 12212) experimentally established that the native intermediate (NI), the species formed upon O₂ bond cleavage, is reduced with an IET rate >700 s⁻¹ and is the catalytically relevant fully oxidized form of the enzyme, rather than the resting state. In this report, we present kinetic and spectroscopic results coupled to DFT calculations that evaluate the mechanism of the 3 e⁻/3 H⁺ reduction of NI, where all three catalytically relevant intramolecular electron transfer (IET) steps are rapid and involve three different structural changes. These three rapid IET processes reflect the sophisticated mechanistic control of the TNC to enable rapid turnover. All three IET processes are fast due to the associated protonation of the bridging oxo and hydroxo ligands, generated by O=O cleavage, to form water products that are extruded from the TNC upon full reduction, thereby defining a unifying mechanism for oxygen reduction and rapid IET by the TNC in the catalytic cycle of the MCOs.

INTRODUCTION

The reduction of dioxygen (O₂) to water is performed in nature by the multicopper oxidases (MCOs) in order to carry out a variety of single-electron oxidations of metal ion or organic substrates.¹⁻³ These enzymes have been of particular interest in a number of areas including their relevance to human health (ceruloplasmin),¹⁴ bioremediation (laccases),¹⁵ and application as oxidation catalysts in biofuel cells (laccases and bilirubin oxidases).¹⁶ The MCOs require at least four Cu’s to accomplish their activity: a type 1 (T1)⁴⁵⁻¹⁰ Cu site and a trinuclear Cu (TNC)¹¹⁻¹² site, that in the resting state is composed of a mononuclear type 2 (T2) and antiferromagnetically coupled binuclear type 3 (T3) Cu centers (Figure 1). The T1 site is solvent accessible and receives electrons from substrate. This T1 center then transfers the electrons through the protein via a conserved Cys-His pathway over ~13 Å to the TNC, which is buried in the protein, where O₂ binds and is reduced. The coppers of the TNC in the oxidized resting state are ligated by 8 histidines in conserved His-X-His motifs where three each ligate the T3 coppers and two ligate the T2. The T2 is additionally coordinated by an aquo-derived hydroxide while the T3s are bridged by a μ₂-hydroxide (μ₂-OH) providing a superexchange pathway for the antiferromagnetic coupling of the T3 Cu(II)’s.¹³ Two conserved carboxylate residues are located near the TNC in all MCOs: D94 (numbered from the Fet3p sequence),¹⁴ hydrogen bonded to the T2 hydroxo ligand and near the T3b, functions as an essential negative charge. Also, there is E487 at the bottom of the TNC near the T3s, which is the source of protons for the O₂ reduction at the TNC.¹⁵⁻¹⁸

The mechanism of O₂ reduction by the MCOs has been well-characterized and occurs in two 2-electron steps (Figure 2).² First, the fully reduced (FR) state reacts with O₂ at a rate of 1.7 × 10⁷ M⁻¹ s⁻¹ to form a peroxy intermediate (Figure 2).¹⁹ In this intermediate, O₂ is reduced by 2 electrons and is coordinated by all three Cu’s of the TNC, where the T2 and T3b Cu’s are oxidized, being closest to the negative D94 that lowers their redox potentials.¹⁷ With an electron transferred from the T1 and a proton from E487, the O=O bond is cleaved to give the native intermediate (NI), where all Cu’s are oxidized and the oxygen atoms originating from O₂ are fully reduced and bound as a μ₃-oxo in the center of the cluster and a μ₂-OH bridging the T3s (Figure 2).²⁰ Reoxidation rates of *Rhus vernicifera* Laccase have determined a lower-limit rate for O₂ bond cleavage of >350 s⁻¹ from stopped-flow kinetics (Figure 2) and of *Trametes versicolor* Laccase a lower-limit rate of >25 000 s⁻¹ has been determined from microsecond freeze-hyper quench studies.²¹ These kinetic measurements imply...
rapid IET from the T1 to the TNC in O₂ reduction. The triangular topology of the TNC enables rapid O–O bond cleavage with a low barrier in the second 2-electron step making this overall process an effective 4-electron reduction of O₂. After the O–O bond has been cleaved, NI decays slowly (0.058 s⁻¹ at 23 °C) in the absence of substrate to the resting oxidized form.22 Alternatively, in the presence of substrate, NI is reduced directly to the fully reduced form.23

While much is known about the O₂ reduction mechanism in the MCOs, less is understood concerning the half of the catalytic cycle where substrate is oxidized and the TNC is reduced. In this case, four electrons must reduce the enzyme with three undergoing intramolecular transfer over 13 Å from the T1 to the TNC. Considerable insight into the mechanism of the MCOs has been derived from studies on the Japanese Tree Laccase (Lc) from Rhus vernicifera, which exhibits fast turnover (kcat = 560 s⁻¹).24 For a long time, the mechanism of the reduction of the enzyme in catalysis went undefined since intramolecular electron transfer (IET) rates from the T1 to the TNC in the resting oxidized form were measured to be far slower (kIET = 1.1 s⁻¹)25 than turnover (kcat = 560 s⁻¹).24 Alternatively, it had been proposed that IET rates in the reduction of NI could be fast and therefore catalytically relevant.1,2 We have recently experimentally determined that this is the case and shown that NI, not the fully oxidized resting enzyme, as is studied in crystallography,26,27 is the catalytically relevant fully oxidized form capable of rapid IET (kIET > 700 s⁻¹) consistent with turnover (kcat = 560 s⁻¹).23 This significant rate enhancement (>10³) for IET in NI relative to the resting TNC was determined to be due to the large driving force for proton-coupled electron transfer (PCET) caused by the strong basicity of the μ₃-oxo of NI. Subsequently, fast IET rates (~460 s⁻¹) were reported in turnover conditions of a small laccase from S. coelicolor with single molecule measurements consistent with our findings establishing rapid IET rates in the catalytic cycle.28 However, our understanding to this point only focuses on the first IET step in the reduction of NI; two additional rapid IET steps are required to fully reduce NI and complete the catalytic cycle.

In this report, we extend our initial study on Rhus vernicifera Laccase to now characterize the molecular mechanism of the 3 e⁻/3 H⁺ processes in the reduction of NI in the catalytic cycle of the MCOs. The reduction of NI is examined via kinetic modeling of stopped-flow (SF) absorption data correlated to freeze-quench electron paramagnetic resonance (FQ-EPR) measurements to determine intermediates formed in this process. These data are coupled to density functional theory (DFT) calculations of these electron and proton transfer processes to fully characterize the molecular mechanism of all three rapid IET steps in NI reduction that enable fast turnover in the MCOs.

### RESULTS AND ANALYSIS

**Kinetics.** The reduction of NI was monitored by spectral changes in stopped-flow (SF) absorption. The reaction of fully reduced Lc with equal molar O₂ shows initial rapid (~10⁶ M⁻¹ s⁻¹) formation of NI based on the appearance of the 365 nm band due to the μ₃-oxo of NI. Subsequently, fast IET rates (~460 s⁻¹) were reported in turnover conditions of a small laccase from S. coelicolor with single molecule measurements consistent with our findings establishing rapid IET rates in the catalytic cycle.28 However, our understanding to this point only focuses on the first IET step in the reduction of NI; two additional rapid IET steps are required to fully reduce NI and complete the catalytic cycle.

Results and analysis.
Figure 3. Stopped-flow absorption traces of the reduction of NI. [Lc] = [O2] = 50 μM at pH 7.5 and 4 °C. (A) Scaled absorption traces of the T1 (614 nm, blue) and TNC (365 nm, red) charge transfer bands in the reduction of NI with 0.125 mM H2Q excess. Inset: absorption spectra from 1 to 300 s. (B) Scaled absorption traces (solid light blue) and fits (dashed black) of the 614 nm band with 0.125, 0.250, 0.500, and 1.250 mM [H2Q] excess. (C) Traces (solid light blue) and fits (dashed black) of the 614 nm band with 42.8, 85.5, and 145.0 mM [H2Q] excess. Part A reprinted with permission from ref 23. Copyright 2013 American Chemical Society.

The redox states are denoted with the superscript "ox" or "red" for the T1 and by the oxidation states of the Cu′s of the TNC. The rates governing this process are the [H2Q] dependent T1 reduction rate (kred1) and the [H2Q] independent IET rates (κIET) from the T1 to the TNC. Starting with the formation of NI, this sequence of T1 reduction followed by IET occurs three times to fully reduce the TNC, and the fourth electron reduces the T1 giving the fully reduced state. Since the 614 nm band is proportional to the concentration of the oxidized T1, all species with T1red will contribute to the trace (i.e., intensity) of this band.

The model in Scheme 1 was initially fit to the 614 nm decay kinetics with 50 electron-equivalents (1.25 mM) excess H2Q to ensure pseudo-first-order behavior for the T1 reduction steps. The rates for the first T1 reduction (kred1 = 0.207 s−1) and first IET rate (κIET1 > 700 s−1), where rates slower than this lower limit cause the fit to deviate from the data, were determined from the 614 nm decay in the same experiment and fixed in the fitting of the 614 nm reduction kinetics. The first fit to the data (fit A; Figure 4A) has all four kred rates in Scheme 1 equal to the first T1 reduction rate (kred1 = 0.207 s−1), with the first IET rate set equal to 700 s−1, and the second and third IET rates were modeled as fast (>2 s−1 under these conditions). This fit shows a T1 intensity reduction that is too slow and is inconsistent with the data (dashed black line compared to the data in blue in Figure 4A). In fit B (Figure 4B), the second, third, and fourth T1 reduction rates are increased to approximately twice kred1 with the same IET rates as in fit A. This leads to faster 614 nm decay, but fails to capture the curvature of the trace, in particular at early times (0–5 s in Figure 4B). To properly model the absorbance at early time, fit C (Figure 4C) includes reversibility at the second IET step with KeqIET2 = κIET−1. Including this equilibrium with the same rates as in fit A correctly fits the absorbance decay at early time points, but the overall decay is too slow. In fit D (Figure 4D), the second, third, and fourth T1 reduction rates in fit C are increased as in fit B. This fit properly models the data and represents the minimal model required to describe the time dependence of the 614 nm absorption decrease. This shows that NI is reduced in a mechanism where the three T1 reduction rates are twice as fast as the first, and importantly, the second IET step is reversible implying a low driving force for this process.

This minimal model (Figure 4D) can be expanded to further probe the kinetics of the third IET step, since this step is contained within the kinetics of the 614 nm band and precedes the fourth and final reduction of the T1. First considering the data with 50 electron equivalent excess in Figure 5A, modeling an equilibrium constant for the third IET step shows that it must be irreversible (KeqIET3 > 1). Additionally, fitting IET rates with the 614 nm data at higher H2Q (Figure 5B) affords a lower limit for kIET3 > 500 s−1.

Application of this model to the entire data set (Figure 3B,C), with kIET1 = 700 s−1, KeqIET2 = 1.0, and kIET3 = 500 s−1 and the T1 reduction rates varying proportionally to the concentration of H2Q reveals a global fit to the kinetics of the 614 nm band (Table 1). The second order rates of the T1 reduction steps are obtained from the linear dependence of these rates across the entire data set where kred1 = 150 M−1 s−1 is in accord with the rate observed from loss of the 365 nm band of NI.25 Additionally, this model is consistent with H2Q as the only reductant for all four T1 reduction steps in the reduction of NI. Since the T1 is a single electron acceptor, a semiquinone (SQ) would be produced that could reduce the T1 with an approximately 3 orders of magnitude faster rate than H2Q (k(H2Q) ≈ 105 M−1 s−1; k(SQ) ≈ 107 M−1 s−1) based on its 350 mV lower reduction potential.30,31 However, the rate of disproportionation of SQ is 10−10−10 M−1 s−1.
implying that all SQ formed will disproportionate before reducing the T1. This is consistent with early kinetic studies on the reduction of *Rhus vernicifera* Laccase by H$_2$Q where no significant SQ involvement was observed. The kinetic data at high H$_2$Q concentrations (Figure 3C) contain an additional rate of 1.2 × 10$^6$ M$^{-1}$ s$^{-1}$ for the formation of NI, consistent with a previous estimate in the absence of reductant. This model reveals a number of key findings about the mechanism of the reduction of NI. First, the T1 reduction rate increases for the last three steps. This is consistent with our
earlier spectroscopic findings that the structure and redox state of the TNC affect the structure of the T1 center.24 Second, the curvature at early time points of the 614 nm trace requires that the second IET is reversible with a $K_{\text{eq, IET}} \approx 1$, and therefore, this process occurs with a low driving force. The fits in this model are not sensitive to the actual rates for the forward and reverse processes, but this IET step must be rapid ($k_{\text{IET}} \geq 560 \text{ s}^{-1}$) to be consistent with turnover and the IET rates from catalytic single molecule measurements.24,28 Lastly, the third IET step is irreversible and has a lower limit rate of $k_{\text{IET}} > 500 \text{ s}^{-1}$ consistent with this step having a large driving force. Taken together with the previous findings, where the first IET step also has a large driving force,23 the kinetics of all three rapid IET steps are revealed. Importantly, these IET steps exhibit different kinetic behavior; namely, the second IET has a lower driving force than the first and third. Since these IET steps are all rapid in turnover, there must be a mechanistic difference between at least the second IET step relative to the first and third.

Spectroscopy. Freeze-quench electron paramagnetic resonance (FQ-EPR) data were obtained to correlate to the kinetics and define potential paramagnetic intermediates in NI reduction. NI is trapped by reacting fully reduced Lc with O2 quenched after 1 s.29 The 77 K X-band FQ-EPR spectrum of NI exhibits the typical signal of the oxidized T1 site with $g_{\text{zx}} = 2.30$ and $A_{\text{zx}} = 471 \times 10^{-4} \text{ cm}^{-1}$ (Figure 6A, black), where contributions from the 3 coupled CuH$^+$’s of the TNC in NI are not observed at temperatures $>20 \text{ K.}$29 FQ-EPR samples of reduced NI (Red NI in Figure 6, red) were obtained by reacting fully reduced Lc with O2 but in the presence of 5 electron equivalents of H2O at pH = 7.5. The 77 K X-band EPR spectrum of this reaction quenched after 1 s is composed mostly of the signal of the oxidized T1 but exhibits an additional signal at $g = 2.14$ (Figure 6A, red). The overlay of the X-band FQ-EPR spectra of NI and Red NI show this extra signal has a derivative shape using the signal of the T1 of NI as a reference (Figure 6A, expanded). A similar 77 K X-band FQ-EPR signal was observed by Reinhammar upon reacting Lc reduced with excess electron equivalents of H2O with O2.25 This $g = 2.14$ signal is also observed in FQ-EPR samples at 77 K in Q-band (Figure 6B). An overlay of the Q-band spectra in the $g_{\text{zx}}$ region ($\sim 11500 - 12000 \text{ G}$) of NI and Red NI shows an additional signal intensity contribution in the Red NI FQ-EPR spectrum at $g = 2.05$ (Figure 6B, expanded). No other contribution from this new species is evident in the X-band or Q-band EPR spectra.

From these data, an additional EPR detectable species is present in the reduction of NI exhibiting a derivative signal centered at $g = 2.14$ and potentially another negative feature at $g = 2.05$.

FQ-EPR samples at different reaction times at 4 °C show the time course of the reduction of NI (Figure 6C). The starting point (0 s) is defined by the 77 K X-band EPR spectrum of NI (Figure 6C, black). When NI is reduced, the derivative feature at $g = 2.14$ clearly increases at early times (0 to 1 and 10 s in Figure 6C, expanded black to red to blue). As the reaction proceeds to 35 s (Figure 6C, green), the FQ-EPR intensity of both the T1 and the $g = 2.14$ signals decreases. These data show that as the T1 reduces in NI reduction, the $g = 2.14$ derivative signal first grows in and then decays along with the T1. Attempts to quantify the time dependence of the intermediate EPR signal are difficult due to large errors in spin-quantifying its contribution to the total EPR intensity (see Supporting Information Figure S4). Since this signal is observed at 77 K, this new signal reflects an $S = \frac{1}{2}$ EPR active intermediate, a 1-electron hole (CuIICuI$^+$) form of the TNC generated by the reduction of NI. A geometric perturbation of the T1 can be ruled out since the absorption spectra from stopped-flow show no changes to the T1 region over the course of the reduction of NI (Figure 3A, inset). This signal will be correlated to possible structures obtained from DFT calculations below.

DFT Calculations on Proton-Coupled Electron Transfer to the TNC of NI. Expanding on experimental insights from the kinetic model and FQ-EPR data, DFT calculations were performed to understand, on a molecular level, the nature of the three IET processes in the reduction of NI. The active site model of NI contains the histidines ligated to the Cu’s, the D94 carboxylate near the T2 and T3b Cu’s, which has been shown to provide an essential negative charge,15,17 and a water hydrogen bonded to the hydroxo ligand that bridges the T3 Cu’s, which is observed in the resting crystal structures (Figure 7; PDB: 1GYC).27 To distinguish different reduced and protonated structures, the optimized structures will be denoted with NI$^x$H$^y$ where $x$ and $y$ denote the number of electrons and protons transferred to NI, respectively. The free energies of reduction are computed with respect to a reduced and oxidized T1 model and free energies of protonation from solvent are computed using the solvation free energy of the proton.30 In order to understand the factors that tune rapid IET, the calculated free energies and inner-sphere reorganization energies are correlated with Marcus Theory where rates of
IET are dependent on the thermodynamic driving force (ΔG°), reorganization energy (λ), which contains the energy to reorganize solvent (outer-sphere; λo) and bonds (inner-sphere; λi) upon redox of the donor and acceptor, and the electronic coupling between the donor and acceptor (H DA). These processes can occur in a stepwise manner where electron transfer (ET) occurs prior to proton transfer (PT), denoted “ET/PT” or in the opposite order “PT/ET”. The electron and proton can transfer in a concerted process, denoted “EPT”. The inner-sphere reorganization energies for the concerted EPT processes (λi,EPT) were estimated using a method employed by Hammes-Schiffer and co-workers (see Methods section).40–41

**First IET Step.** The first IET to NI has been shown to be a concerted EPT process, which explains the >10^3 rate enhancement over IET to the resting TNC (Figure 8, Table 2). The first electron into NI reduces the T3a Cu, as seen from spin densities of NIe in Table 2. The T3a Cu of the TNC is furthest from D94 and therefore has a higher redox potential than the T3b and T2 Cu’s. This reduction results in the

Table 2. Selected Distances and Cu Spin Densities of the Structures of the First IET Step

| Distances (Å) | NI | NIe | NIH (T3W) | NIe,H |
|---------------|----|-----|-----------|-------|
| T3a−T3b       | 2.982 | 3.328 | 3.259 | 3.325 | 3.718 |
| T3a−T2        | 3.474 | 3.427 | 3.480 | 3.642 | 4.219 |
| T3b−T2        | 3.463 | 3.288 | 3.397 | 3.934 | 3.719 |
| T3a−O(μ3-oxo) | 1.920 | 1.982 | 1.868 | 2.058 |
| T3b−O(μ3-oxo) | 1.978 | 1.918 | 1.978 | 2.428 | 1.984 |
| T2−O(μ2-oxo)  | 1.881 | 1.914 | 2.018 | 1.934 | 1.867 |
| T3a−O(μ2-OH)  | 1.929 | 3.149 | 1.946 | 2.101 |
| T3b−O(μ2-OH)  | 1.898 | 1.906 | 2.053 | 1.892 | 1.919 |
| T2−OH2        | 2.025 | 2.107 | 2.015 | 1.953 | 1.959 |

Cu Spin Densities

| T3a | 0.66 | -0.03 | 0.57 | 0.68 | 0.03 |
| T3b | 0.62 | 0.58  | 0.64 | 0.69 | 0.61 |
| T2  | -0.56 | -0.51 | 0.61 | -0.62 | -0.60 |

a) O(μ2-OH) in NIH and NIHc. b) O(−OH2) in NIH(T3W).

First IET Step.

Figure 7. Geometry optimized structure of NI determined from spectroscopy. Histidine on the T2 omitted for clarity. Colors: Cu, orange; O, red; N, blue; C, gray; H, white.

Figure 8. DFT calculations of the first IET step. All energies reflect Gibbs free energies (ΔG) in units of kcal/mol. Distances and spin densities for these structures are reported in Table 2. Histidine of the T2 removed for clarity. Colors: Cu, orange; O, red; N, blue; C, gray; H, white. ET/PT, PT/ET, and EPT pathways labeled in red, blue, and black, respectively.

dx.doi.org/10.1021/ja509150j | J. Am. Chem. Soc. 2014, 136, 17788–17801

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Elongation of the T3a bond to the $\mu_2$-OH where the oxidized T3b and T2 are antiferromagnetically coupled ($\text{NIe}$).

Protonation of the $\mu_3$-oxo of $\text{NIe}$ gives $\text{NIe,H}$ where the newly generated $\mu_2$-OH bridge provides a superexchange pathway for antiferromagnetic coupling of the oxidized T2 and T3b Cu’s with a free energy for EPT $\Delta G(e^- + H^+)$ of $-31.8$ kcal/mol and $\lambda_{\text{EPT}} = 1.1$ eV as consistent with previous calculations for this IET step. This large negative free energy is clearly an overestimation that can be attributed to the B3LYP functional, which overestimates the covalency of the T1 site. This can be corrected for by reference to equivalent calculations of the resting state where the calculation gives $-21.3$ kcal/mol while the experimental value is given by the electrochemical potential difference between the T1 and T3 sites in the resting enzyme ($\Delta G = -2.0$ kcal/mol). Using this “resting” calibration, the experimentally calibrated free energy for EPT for the first IET step in NI reduction is $-12.5$ kcal/mol.

If NI is first protonated, this protonation can occur at either the $\mu_3$-oxo or $\mu_2$-OH ligands. Protonation of the $\mu_3$-oxo gives $\text{NIH}$, which is $18.2$ kcal/mol more favorable compared to protonation of the $\mu_2$-OH reflecting a $\Delta pK_a$ of $\sim 13$ between the $\mu_3$-oxo and $\mu_2$-OH ligands quantifying the strong basicity of the $\mu_3$-oxo (Figure 8). Interestingly, the decay of NI due to protonation of the $\mu_3$-oxo is slow.

Table 3. Selected Geometric Parameters and Copper Spin Densities of the Structures of the Second IET Step

| distances (Å)         | $\text{NI}^{2e,H}$ | $\text{NI}^{2e,H}$ | $\text{NI}^{2e,2H}$ (MV) | $\text{NI}^{2e,2H}$ (L) |
|-----------------------|---------------------|---------------------|--------------------------|--------------------------|
| T3a–T3b               | 3.827               | 4.509               | 4.558                    | 4.044                    |
| T3a–T2                | 4.022               | 4.503               | 4.301                    | 4.127                    |
| T3b–T2                | 3.790               | 3.724               | 3.758                    | 4.179                    |
| T3b–O(inner)$^a$      | 1.900               | 2.000               | 1.960                    | 2.081                    |
| T2–O(inner)$^a$       | 2.032               | 1.924               | 1.966                    | 2.254                    |
| T3a–O(outer)$^b$      | 2.090               | 2.197               | 2.217                    | 2.217                    |
| T3b–O(outer)$^b$      | 1.938               | 1.986               | 2.200                    | 1.956                    |
| T2–OH$_2$             | 2.140               | 1.954               | 2.061                    | 2.087                    |

Cu Spin Densities

| Cu Spin Densities | T3a | T3b | T2   |
|-------------------|-----|-----|------|
| $\Delta G$ (e$^- + H^+$) | $-31.8$ kcal/mol | $1.1$ eV |       |

$^a$O(inner) is $\mu_3$-OH in $\text{NI}^{2e,2H}$ (MV) and H$_2$O in $\text{NI}^{2e,2H}$ (L), $^b$O(outer) is $\mu_2$-OH $\text{NI}^{2e,2H}$ (L) and H$_2$O in $\text{NI}^{2e,2H}$ (MV) and $\text{NI}^{2e,2H}$.

Figure 9. DFT calculations of the second IET step. All energies reflect Gibbs free energies ($\Delta G$) in units of kcal/mol. Distances and spin densities for these structures are reported in Table 3. Histidine of the T2 removed for clarity. Colors: Cu, orange; O, red; N, blue; C, gray; H, white. ET/PT, PT/ET, and EPT pathways labeled in red, blue, and black, respectively.

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| T3b–O(outer)$^b$      | 1.938               | 1.986               | 2.200                    | 1.956                    |
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| $\Delta G$ (e$^- + H^+$) | $-31.8$ kcal/mol | $1.1$ eV |       |

$^a$O(inner) is $\mu_3$-OH in $\text{NI}^{2e,2H}$ and $\text{NI}^{2e,2H}$ (MV) and H$_2$O in $\text{NI}^{2e,2H}$ (L), $^b$O(outer) is $\mu_2$-OH $\text{NI}^{2e,2H}$ (L) and H$_2$O in $\text{NI}^{2e,2H}$ (MV) and $\text{NI}^{2e,2H}$. 
ligands are protonated by a carboxylate near the T3 edge of the TNC (E487 in Figure 1), protonation of the less basic μ3-OH would precede protonation of the μ2-oxo leading to slow rate for NI decay. The protonation of NI will be considered below.

These calculations are consistent with the kinetic findings of rapid first IET and show this process has a large driving force for rapid EPT due to the μ2-oxo acting as a strong base (diagonal arrow from NI to NIe,H in Figure 8). Alternatively, the inner-sphere reorganization energy for the EPT in the first IET process is calculated to be 1.1 eV, which is predominantly due to the loss of the T3a-μ1-oxo bond upon protonation.

Second IET Step and Correlation to EPR Active Intermediate. Starting from NIe,H, electron transfer to this intermediate results in reduction of the T2 Cu with a computed ET inner-sphere reorganization energy of 0.4 eV (NIe,2H in Figure 9).

Protonation of this intermediate Protonation of this intermediate (NIe,2H) can occur at the μ2-oxo bridging the T2 and T3b Cu’s (inner μ2-oxo) or at the T3a and T3b Cu’s (outer μ2-oxo). Protonation of the outer μ2-oxo results in a mixed-valent (MV) delocalized intermediate [NIe,2H (MV)] with the electron shared equivalently between the T2 and T3b Cu’s where the bond of the reduced T3a to the μ2-oxo has been cleaved (Cu spin densities in Table 3). Protonation of the inner μ2-oxo yields an intermediate where a water ligand is bound inside the TNC with a localized hole (L) on the oxidized T3b giving NIe,2H (L). Protonation to give NIe,2H (MV) is calculated to be 6.3 kcal/mol more favorable than protonation to form NIe,2H (L).

In the PT/EIT process (blue arrows in Figure 9), protonation of NIe,2H only occurs at the outer μ2-oxo cleaving the reduced T3a μ2-oxo bond to give NIe,2H. Attempts to optimize a protonated inner water ligand (from μ2-oxo) results in proton transfer to the outer μ2-oxo due to a lower pKₐ of the μ2-oxo bridging the oxidized T2 and T3b coppers. Reduction of NIe,2H gives the same NIe,2H (MV) obtained by first reducing NIe,H where the ET inner-sphere reorganization energy for this step is 0.4 eV. This MV structure can be further protonated at the inner μ2-oxo to obtain a structure with an oxidized, localized T3b NIe,2H (L), which is uphill by 7.2 kcal/mol.

From the kinetics analysis above, the second IET step is particularly interesting since it occurs with a low driving force but is required to be rapid to be consistent with turnover. As in the first IET, the computed free energies in this step can be calibrated to the experimental driving force for the reduction of the fully oxidized resting state and compared to the ΔG(e⁻ + H⁺) for EPT.

First, an EPT process (black arrows in Figure 9) for the second IET step is considered. Compared to the ΔG(e⁻ + H⁺) for the first IET step, the computed ΔG(e⁻ + H⁺)s for EPT to give NIe,2H (MV) and (L) intermediates are less favorable by ~10 and ~17 kcal/mol, respectively, consistent with the lower driving force for this step compared to the first from the kinetics. The “resting” calibrated EPT free energy to give NIe,2H (MV) is ΔG(e⁻ + H⁺) = -2.1 kcal/mol and is also consistent with the experimentally determined low driving force. From Marcus Theory, if the first and second IET steps occur with similar, rapid rates (>700 s⁻¹), the low driving force for this second IET step requires it to occur with a reorganization energy at least 0.8 eV lower than the first IET step. However, due to large geometric changes upon EPT (Figure 9), the computed inner-sphere reorganization energies for EPT for the second step are large and comparable to that of the first [1.2 eV for NIe,2H (MV) and 0.9 eV for NIe,2H (L)].

Thus, a concerted process for the second IET step would not be fast or catalytically relevant. The ET/PT mechanism (red arrows in Figure 9) would also be slow as it is calculated to be ~30 kcal/mol less favorable than the first IET. This would require ~2.0 eV lower λ than the first IET. However, the λ is calculated to be only 0.8 eV less than the first.

Alternatively, a PT/EIT process (blue arrows in Figure 9), where the proton transfer to the outer μ2-oxo precedes ET to give the NIe,2H (MV) form, has a driving force for ET ~10 kcal/mol less than the first IET step and an ET inner-sphere reorganization energy of 0.4 eV. This difference in driving force would require a 0.8 eV lower reorganization energy for IET to have the same rate (>700 s⁻¹). This is consistent with the difference in computed reorganization energies for EPT (1.1 eV) in the first step and PT/EIT in the second (0.4 eV).

Therefore, rapid EIT for the second IET step would occur after protonation of the outer μ2-oxo of the NIe,H intermediate.

In order for this IET process to be rapid and stepwise, the proton transfer to the outer μ2-oxo must be fast. Interestingly, the free energy for protonating the outer μ2-oxo of NIe,H is similar to protonating the μ2-oxo in NI, 0.2 and ~2.9 kcal/mol, respectively, implying that these ligands have similar basicities and would rapidly protonate. However, NI decay, which involves this protonation of the μ2-oxo, is slow.

Slow protonation of NI is due to barriers in the PT process where protons originate from E487 near the μ2-oxo bridge of the T3 Cu’s (Figure 1). To determine the differences in the rates of PT to the μ2-oxo of NIe,H relative to the μ2-oxo in NI, PT mechanisms were computed (Figure 10). In these models, a proton was transferred to the TNC from a carboxylic acid modeling E487. PT to the μ2-oxo of NI leads to a transition state (TS; 9.8 kcal/mol) where the T3aCu(II)–μ2-oxo bond is broken due to the incoming proton. Upon cleavage of the T3aCu(II)–μ2-oxo bond, the spin ground state changes from S = 1/2 to 1/2 [NIe,H(T3W); ΔE = 4.0 kcal/mol].

Note that the energy of protonation of the μ2-oxo of NI from solvent is computed to be ΔE = 9.1 kcal/mol (ΔG = 15.3 kcal/mol in Figure 8). The ~5 kcal/mol more favorable protonation by the carboxylic acid reflects greater stability due to the stronger hydrogen bond from the anionic carboxylate compared to a neutral water (Figure 8). PT from T3b–OH₂ to the μ2-oxo leads to another transition state (TS'; 11.0 kcal/mol, modeled in a 2D potential energy surface; see Supporting Information Figure S8) where the T3aCu(II)–μ2-oxo bond is broken due to a steric clash of the water ligand with the T3aCu(II). From here, the proton is transferred to the μ2-oxo (~7.7 kcal/mol), which is in good agreement with the energy for protonation of the μ2-oxo of NI in Figure 8 (ΔE = -8.6 kcal/mol; ΔG = -2.9 kcal/mol). Therefore, protonation of the basic μ2-oxo requires overcoming two barriers due to the cleavage of strong Cu(II) hydroxo and oxo bonds, the highest being ~11.0 kcal/mol, in the range of the experimentally observed activation enthalpy of ~9–14 kcal/mol. Alternatively, PT from the carboxylic acid to the μ2-oxo of NIe,H is calculated to be downhill (~8.6 kcal/mol) and importantly without a barrier (Supporting Information Figure S10) consistent with the relative energy for protonation of the corresponding structure in Figure 9 (ΔE = -10.1 kcal/mol; ΔG = 0.2 kcal/mol), and comparable with the proton affinity for the μ2-oxo (Figure 10). This is due to the weakened T3aCu(II)–μ2-oxo bond in NIe,H, which cleaves prior to PT. This contrasts PT to the μ2-oxo in NI that involves breaking a strong Cu(II)–μ2-oxo bond. Therefore, the μ2-oxo in NIe,H is kinetically accessible and would protonate rapidly whereas the μ2-oxo in NI is just as
basic but slow to protonate due to the required breaking of strong Cu(II)–oxo and hydroxo bonds. This would prevent adventitious protonation of NI before it can be reduced (vide infra).

From the above DFT modeling, there are a number of potential EPR active $S = \frac{1}{2}$ intermediates (Figures 9 and 11). These possible structures for this 1 electron hole intermediate can be correlated to the FQ-EPR data presented in Figure 6 (a derivative feature at $g = 2.14$ and another possible negative contribution at $g = 2.05$).

The $\text{NI}^{2e,H}$ intermediate has a hole localized on the T3b Cu (Figure 11, first structure). In the $g$-tensor coordinate system the ground state is calculated to be $d_{x^2-y^2}$. This predicts an EPR spectrum inconsistent with the experimental signal (Figure 11 bottom center) since this intermediate lacks a derivative feature at low field. The possible $\text{NI}^{2e,2H}$ (L) and $\text{NI}^{2e,3H}$ intermediates
(Figure 11, second and third structures) have localized holes on the T3b Cu also with predominantly $d_{x^2-y^2}$ ground states but with a small amount of $d_{z^2}$ mixing leading to a rhombic splitting of the $g$-values. Therefore, these possible intermediates would exhibit two $g$-values at low field, but only one low field signal is observed experimentally (Figure 11, bottom center). Therefore, these intermediate structures also do not correspond to the FQ-EPR signal.

The fourth potential EPR-active intermediate is $\text{Ni}^{2+\cdot2H} (\text{MV})$, where the hole is delocalized over the T2 and T3b Cu’s, which have $d_{x^2-y^2}$ orbitals with close to equal contributions, but with different orientations (Figure 11 right). The observed $g$-values for a MV system reflect the molecular $g$-tensor that is the vector-coupled product of the local $g$-tensors on each Cu.40 The z-axis of the $d_{x^2-y^2}$ of the T3b is ∼90° rotated about the y-axis relative to the z-axis of the $d_{x^2-y^2}$ of the T2 (Supporting Information Figure S12). This results in a molecular $g$-tensor consisting of two components that are the average of $g_x$ and $g_z$ from the T3b and T2 and a third that is $g_y$. Consistent with this picture, the predicted $g$-values for $\text{Ni}^{2+\cdot2H} (\text{MV})$ are 2.201 and 2.168, accompanied by a low $g$-value at 2.070. The simulated spectrum (Figure 11, right) from this structure exhibits a derivative signal consisting of two similar $g$-values at low field with another at high field. This is very similar to the one-hole intermediate spectrum [obtained by subtraction of the T1 spectrum from the 1s, Figure 6, spectrum shown at the bottom center of Figure 11 (see Supporting Information Figure S3)]. Therefore, the $\text{Ni}^{2+\cdot2H} (\text{MV})$ predicted EPR spectrum is most consistent with the observed EPR signal that correlates with the 1-hole intermediate observed in the kinetics.

In summary, the experimental findings coupled to these calculations indicate that the TNC carries out a second rapid IET but with a low driving force. A stepwise PT/ET process enables a fast IET rate with a low driving force due to the low inner-sphere reorganization for the ET step. An EPT process would be slow since protonation of the bound oxygen ligands of the TNC results in structural rearrangements that lead to large reorganization energies. The $\text{Ni}^{2+\cdot2H}$ intermediate is set up to be rapidly protonated because it is located near the carboxylate proton donor and is strongly basic in this partially reduced site. This second IET step provides structural insight into how the TNC of a partially reduced NI is set up to perform rapid IET with a low driving force.

Third IET Step. Starting from the $\text{Ni}^{2+\cdot2H} (\text{MV})$ species (Figure 12), reduction leads to a fully reduced TNC, where the $\mu_2$-OH shifts from bridging the T2 and T3b Cu’s to bridging the T3 Cu’s with dissociation of the water that was bound to the T3b ($\text{Ni}^{2+\cdot2H}$). The $\mu_2$-OH bridging the T3 Cu’s is accessible to protonation by £E487. Protonation of this $\mu_2$-OH gives the fully reduced TNC, where both oxygen atoms from the 4-electron reduction of O2 are now fully protonated water products. Protonation of $\text{Ni}^{2+\cdot2H} (\text{MV})$ prior to reduction leads to a water bound in the center of the TNC. Compared to the $\mu_3$-oxo of NI or the outer $\mu_2$-OH of $\text{Ni}^{2+\cdot2H}$ evaluated above, this is ∼7-10 kcal/mol less favorable protonation since this water is constrained in the center of the TNC. Reduction of $\text{Ni}^{2+\cdot2H}$ leads to the fully reduced state $\text{Ni}^{3+\cdot2H}$ (FR) where the waters now optimize to outside the TNC.

The kinetic modeling show that the third IET step is rapid and irreversible. The ET/PT pathway (red arrows in Figure 12) for this step lacks sufficient driving force since its uphill from the first EPT step by ∼20 kcal/mol [comparing the ET step (−11.9 kcal/mol) to the $\Delta G(e^- + H^+)$ of the first IET]. PT/ET (red arrows in Figure 12) is also unlikely since the proton affinity of $\text{Ni}^{2+\cdot2H} (\text{MV})$ is ∼7-10 kcal/mol lower than the protonation of the $\mu_3$-oxo of NI and the $\mu_2$-OH of $\text{Ni}^{2+\cdot2H}$, and therefore, this protonation would be slow. However, the free energy for an EPT process [$\Delta G(e^- + H^+) = -10.4\text{ kcal/mol}$ (calibrated to resting)] is comparable with that of the first IET step, indicating a large driving force for this concerted process that would enable rapid IET even with the large calculated $\lambda_{\text{EPT}}$ due to movement of water together with EPT and consistent with experiment.

Although the first and third IET steps have similar driving forces for EPT, they involve different structural changes and therefore have different contributions to the driving force. The

\[
\begin{align*}
\text{Table 4. Selected Distances and Copper Spin Densities of the Structures of the Third IET Step} \\
\hline
\text{Distances (Å)} & \text{Ni}^{2+\cdot2H} & \text{Ni}^{3+\cdot2H} & \text{Ni}^{3+\cdot2H} (\text{FR}) \\
\hline
\text{T3a} & 4.703 & 3.974 & 4.826 \\
\text{T3a} & 4.524 & 4.009 & 4.561 \\
\text{T3b} & 4.492 & 4.193 & 4.344 \\
\text{T3b} & 2.086 & & \\
\text{T3b} & 2.055 & 2.030 & 2.117 \\
\text{T2} & 2.073 & 2.192 & 2.109 \\
\hline
\text{Cu Spin Densities} & & & \\
\text{T3a} & 0.00 & & \\
\text{T3b} & 0.68 & & \\
\text{T2} & 0.00 & & \\
\end{align*}
\]

\(\text{O} = \text{H}_2\text{O in Ni}^{2+\cdot2H}. \quad \text{P} = \mu_2\text{-OH in Ni}^{3+\cdot2H} \) and $\text{H}_2\text{O}$ in $\text{Ni}^{2+\cdot2H}$ and $\text{Ni}^{3+\cdot2H}$.}
major difference is that the oxygen ligand products of the first IET remain bound to the TNC, while after the third IET the oxygen ligands are fully protonated and optimize out of the cluster. Therefore, loss of water from the fully reduced TNC will contribute to the driving force for EPT.

To estimate the energetic contribution associated with water loss, i.e., water extrusion from within the cluster, a fully reduced TNC structure with waters bound inside the TNC was optimized (Figure 12, middle). Forming this intermediate from NI$^{2e,2H}$ (MV) has a free energy of $\Delta G(e^- + H^+) = -14.3$ kcal/mol, which represents the driving force for EPT in the absence of water extrusion. Note that upon EPT the $\mu_2$-OH in NI$^{2e,2H}$ (MV) remains as a water bridge between the T2 and T3b coppers. When the distance of the bridging water to the T2 Cu is elongated from this position, the waters optimize to outside the cluster. The free energy difference to this fully reduced (FR) cluster is downhill by $-15.4$ kcal/mol and is an upper limit on the driving force for water extrusion from the TNC, which is coupled to EPT enabling rapid IET (Figure 12).

The contributions to the driving force are evident from the geometric changes upon water extrusion from the TNC. In the water bound structure (Figure 13, middle), if only the internal waters are optimized (constraining the rest of the cluster), the repulsive interactions of these water ligands with the reduced Cu’s are minimized ($-9.6$ kcal/mol, Figure 13, bottom). Full optimization of the cluster to FR gives shorter Cu–Cu distances reflecting relaxation of the Cu’s to their equilibrium positions ($-6.4$ kcal/mol). The 2.2 Å resolution crystal structure of fully reduced ascorbate oxidase exhibits Cu–Cu distances (T3a–T3b = 5.09 Å, T3a–T2 = 4.03 Å, T3b–T2 = 4.46 Å)$^{26}$ consistent with the FR optimized structure (Figure 13, right) and indicate that the Cu–Cu distances in the water bound cluster (Figure 13 right) are indeed long. Thus, waters bound in the center of the cluster undergo steric repulsions with the reduced Cu’s and cause these Cu’s to distort away from their preferred Cu–Cu distances in the fully reduced cluster. Since the Cu’s are held in position by the protein ligands, these protein constraints oppose the waters inside the fully reduced TNC and cause their favorable extrusion in consort with EPT. This water extrusion serves an additional purpose since all three coppers of the reduced TNC need to be coordinatively unsaturated to go on to react with O$_2$ in catalysis.

DISCUSSION

From a combination of kinetics, EPR spectroscopy, and calculations, the mechanism of the reduction of NI has been evaluated and reveals that the TNC enables three rapid IET processes with three unique structural mechanisms for rapid turnover in catalysis (Figure 14). The first IET step is fast due a large driving force for coupled EPT based on the $\mu_3$-oxo acting as a strong base.$^{23}$ From experimental kinetic data, the second IET step has a low driving force that would result in a slow EPT process due to the sizable reorganization energy resulting from large structural changes upon EPT. Instead, a stepwise PT/ET process is effective. The $\mu_2$-OH bridge of the partially reduced TNC is kinetically accessible and basic enough for rapid protonation. This enables rapid ET with a low driving force because of the low inner-sphere reorganization energy. The third IET step completes the reduction of the TNC again through an EPT process, with a large driving force as reduction and protonation are coupled to water extrusion from the fully reduced TNC due to protein constraints from the protein backbone.

A number of attributes enable rapid IET in the turnover of the MCOs. The most important is that reduction of NI requires both three electrons and three protons. This is enabled by the
triangular arrangement of the three oxidized coppers of the TNC, which stabilize the reduced oxygen ions, bound as µ₃-oxo and µ₂-OH ligands. Therefore, three electrons and three protons are required to fully reduce the TNC and produce water, which are coupled to enable rapid IET.

The delivery of protons to these ligands at the proper step enables the TNC to protonate the bound oxo and hydroxo ligands with the reduction of the Cu’s. This is due in part to the MCOs containing a single entry point for protons (E487) and limited solvent accessibility to prevent adventitious protonation of the basic µ₃-oxo ligand prior to reduction. This is evident from the PT mechanism for NI (Figure 10), where there is a barrier to protonate the basic µ₃-oxo due to the strong Cu(II)–oxo bond in addition to the barrier for protonation of the µ₂-OH. The reduction of the T₃a Cu by the first EPT step (to NT) increases the basicity of the T₃ µ₂-OH bridge, which is directly accessible from E487 and would rapidly protonate, which enables rapid and reversible ET for the second IET step. Proton delivery in the third step is concerted with ET and coupled to water extrusion.

An essential function of the TNC revealed by this mechanism is making product water binding to the fully reduced TNC unfavorable, which contributes to the driving force for water extrusion. Waters bound inside the fully reduced cluster have steric interactions with the Cu’s and distort the reduced Cu’s away from their equilibrium positions enforced by the protein. Therefore, the protein constraints on the Cu’s of the fully reduced TNC lead to favorable water extrusion, and this contributes to the driving force to enable rapid EPT even with the reorganization energy associated with this structural change. Additionally, coupling water extrusion to the last IET step is critically important in enabling the coordination unsaturation required by the fully reduced TNC for O₂ reduction in turnover.

**CONCLUSIONS**

The combination of coupling the transfer of electrons to proton delivery and the protein constrains enabling water extrusion reflects a sophisticated mechanistic control by the TNC of the MCOs that enables the three rapid IETs in turnover. All three of these IET steps are rapid because of the basicity of the O₂-derived ligands that arise from cleavage of the O=O bond. In catalysis, the triangular topology of the TNC enables the fast, concerted 4-electron reduction of O₂ to the water level in the formation of NI. However, only after NI is fully reduced by rapid proton coupled IET processes are the water products of O₂ reduction fully protonated and then extruded from the cluster, enabling reduction of another equivalent of O₂. Therefore, the MCOs enable rapid rereduction of the TNC by coupling the three rapid proton-coupled IET processes with the final synthesis of two H₂O molecules from O₂ (Figure 15). This defines a unifying catalytic mechanism by the TNC for coupling O₂ reduction with rapid proton coupled IETs to enable fast turnover in oxidation catalysis.

**METHODS**

**Experimental Section.** All chemicals were reagent grade and used without further purification. Water was purified to a resistivity of 15–17 MΩ cm⁻¹ using a Barnstead Nanopure deionizing system. *Rhus vernicifera* laccase was isolated from acetone powder (Saito and Co., Osaka, Japan) according to published procedures.⁴³,⁴⁷ Protein concentration was determined using the extinction coefficient of the absorption band at 280 nm (90 000 M⁻¹ cm⁻¹).⁴⁸ Copper content was determined spectrophotometrically using 2,2'-biquinoline.⁴⁹ The concentration of paramagnetic copper was determined from spin quantitation of EPR spectra, using a 1.0 mM CuSO₄·5H₂O solution with 2 mM HCl and 2 M NaClO₄ standard.⁵⁰ Protein samples were buffer-exchanged into 100 mM sodium phosphate buffer (pH = 7.5). Kinetic data with ~2 ms dead time were obtained using an Applied Photophysics SX.20 stopped-flow absorption spectrophotometer equipped with a Hg/xe arc lamp and outfitted with PEEK tubing. The temperature was maintained using a water/ethanol temperature bath (Fisher Scientific Isotemp 3016). Stopped-flow experiments were conducted at 4 °C with a cell path length of 1 cm. All solutions were freshly prepared in an anaerobic glovebox. Protein was deoxygenated under a constant flow of Ar for ~2–3 h and transferred to a glovebox. Both injector ports of the stopped-flow were degassed with ~3.0 mM sodium dithionite for ~20 min and kept anaerobic with nitrogen stream through the system. Concentrations of protein were obtained by measurement of the absorption of the T₁ copper at 614 nm (5600 M⁻¹ cm⁻¹) of an
 aliquant of anaerobic protein immediately preceding a kinetic experiment. All protein samples were loaded on the stopped-flow at concentrations of 100 μM. To ensure maximal enzyme oxidation while only being single turnover upon reaction with O2, dioxygen solutions were prepared by diluting air saturated buffer with degassed buffer ~3 fold to produce solution concentrations of [O2] ~ 100 μM. Kinetics on the native intermediate were conducted by preparing fully reduced protein by adding 4 electron equivalents of excess ascorbate and allowing this to sit for ~30 min to ensure full reduction as also confirmed with color change from blue to colorless. Buffer was added to dilute the reduced enzyme to desired concentration. Native intermediate reduction is measured by addition of excess hydroquinone to the colorless fully reduced protein and dilution to 100 μM concentration followed by reaction with O2. All kinetic traces of the 365 nm band were fit with Origin 6.0 and KaleidaGraph 4.1 software packages and described previously. Kinetic traces of the 614 nm band were fit with the IBM Chemical Kinetics Simulator version 1.0.1.

All X-band spectra were obtained at 77 K in a liquid nitrogen finger dewar with a Bruker EMX spectrometer, ER 051 QF microwave bridge, and ER 4102ST cavity (parameters for recording the X-band EPR: 9.39 GHz frequency, 10 mW power, and 10 G modulation amplitude). Q-band EPR spectra were obtained at 77 K using an ER 051 QF microwave bridge, an ER 5106QT resonator, and an Oxford continuous-flow CF935 cryostat (parameters for recording the Q-band EPR: 34.0 GHz frequency, 0.44 mW power, and 10 G modulation amplitude). X-band freeze-quench samples were made by reacting reduced laccase (prepared anaerobically in a glovebox) and reacting at amplitude). X-band freeze-quench samples were made by reacting ascorbate and allowing this to sit for ~30 min to ensure full reduction and dilution to 100 μM concentration followed by reaction with O2. All kinetic traces of the 614 nm band were fit with the IBM Chemical Kinetics Simulator version 1.0.1.

All X-band spectra were obtained at 77 K in a liquid nitrogen finger dewar with a Bruker EMX spectrometer, ER 051 QF microwave bridge, and ER 4102ST cavity (parameters for recording the X-band EPR: 9.39 GHz frequency, 10 mW power, and 10 G modulation amplitude). Q-band EPR spectra were obtained at 77 K using an ER 051 QF microwave bridge, an ER 5106QT resonator, and an Oxford continuous-flow CF935 cryostat (parameters for recording the Q-band EPR: 34.0 GHz frequency, 0.44 mW power, and 10 G modulation amplitude). X-band freeze-quench samples were made by reacting

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