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CME Review

The infectious complications of atopic dermatitis

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Key Messages

- Factors that contribute to the increased infections in atopic dermatitis (AD) are skin barrier defects, suppression of cutaneous innate immunity by type 2 inflammation, Staphylococcus aureus colonization, and cutaneous dysbiosis.
- Skin infections in AD increase the risk of life-threatening systemic infections.
- The use of antibiotics for AD exacerbation remains controversial, and further studies are needed to define which subsets of these patients can benefit from antibiotics.
- The goals of infection prevention in AD consist of skin barrier improvement, anti-inflammatory therapy, and minimizing the use of antibiotics.

ARTICLE INFO

Article history:
Received for publication June 30, 2020.
Received in revised form July 31, 2020.
Accepted for publication August 4, 2020.

ABSTRACT

Objective: Atopic dermatitis (AD) is a chronic inflammatory skin disease that is complicated by an increased risk for skin and systemic infections. Preventive therapy for AD is based on skin barrier improvement and anti-inflammatory treatments, whereas overt skin and systemic infections require antibiotics or antiviral treatments. This review updates the pathophysiology, diagnosis, management, controversy of antibiotic use, and potential treatments of infectious complications of AD.

Data Sources: Published literature obtained through PubMed database searches and clinical pictures.

Study Selections: Studies relevant to the mechanisms, diagnosis, management, and potential therapy of infectious complications of AD.

Results: Skin barrier defects, type 2 inflammation, Staphylococcus aureus colonization, and cutaneous dysbiosis are the major predisposing factors for the increased infections in AD. Although overt infections require antibiotics, the use of antibiotics in AD exacerbation remains controversial.

Conclusion: Infectious complications are a comorbidity of AD. Although not common, systemic bacterial infections and eczema herpeticum can be life-threatening. Preventive therapy of infections in AD emphasizes skin barrier improvement and anti-inflammatory therapy. The use of antibiotics in AD exacerbation requires further studies.

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Participants will be able to demonstrate increased knowledge of the clinical treatment of allergy/asthma/immunology and how new information can be applied to their own practices.

Learning Objectives
At the conclusion of this activity, participants should be able to:

- Describe the mechanisms that lead to increased infections in atopic dermatitis (AD).
- Discuss the strategies for infection prevention in AD.

Release Date: January 1, 2021
Expiration Date: December 31, 2022
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Physicians involved in providing patient care in the field of allergy/asthma/immunology

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Introduction

Atopic dermatitis (AD) is the most common chronic inflammatory skin disease that affects both children and adults with a prevalence of up to 18% and 7%, respectively. Patients with AD and their caregivers experience decreased quality of life, including disruption in daily activities at school and at work, sleep disturbance, depression, and anxiety.1 In addition to these complications, patients with AD are at increased risk for infections.2 The prevalence of cutaneous and systemic infections in patients with AD is significantly higher than those without AD.3 Infectious complications of AD include skin and soft tissue infections (SSTI), eczema herpeticum (EH), bacteremia, osteomyelitis, septic arthritis, and endocarditis.4 These complications lead to significant financial burden on the health care system.5 In this review, we will summarize the advances in the mechanisms, clinical complications, and management of infections in AD.

What Causes an Increase in Infections in AD?

Skin Barrier Defects

AD is inherently associated with skin barrier defects, as measured by transepidermal water loss.6 Patients with AD have a significantly thinner stratum corneum owing to a lack of terminal keratinocyte differentiation. As a result of skin barrier abnormalities, AD is associated with increased transepidermal water loss, which is greatest in the patients with most severe AD.2 The molecular basis for skin barrier defects is due to a deficiency in proteins and lipids with barrier functions including filaggrin, involucrin, claudins, ceramides, cholesterol, and free fatty acids.7 The filaggrin gene loss-of-function (FLG LoF) was the first evidence for the genetic basis of skin barrier defects in AD.2 FLG LoF leads to decreased skin hydration and renders AD susceptible to environmental insults including allergens and pathogens.2 In healthy skin, filaggrin is broken down into hygroscopic amino acids, including urocanic acid and pyrrolidone carboxylic acid, which maintain the acidic pH of the stratum corneum. The acidic environment in healthy skin decreases the expression of 2 staphyloccocal surface proteins, clumping factor B and fibronectin-binding protein, which bind to host proteins cytokeratin 10 and fibronectin, respectively.2 Defects in filaggrin expression lead to decreased urocanic acid and pyrrolidone carboxylic acid levels and a rise in pH, which favors Staphylococcus aureus proliferation.8 FLG LoF is associated with early-onset AD and is present in approximately 25% to 30% of patients with AD of European and Asian descent.9 A more
Immune Dysregulation

Keratinocytes are skin epithelial cells that contribute to the barrier functions and immune response. In patients with AD, keratinocytes produce an increased amount of thymic stromal lymphopoietin, IL-33, and IL-25, which activate innate lymphoid cells 2 (ILC2) to produce type 2 cytokines, including IL-4, IL-5, and IL-13. IL-4 and IL-13 have been indicated to suppress keratinocyte expression of antimicrobial peptides and skin barrier functions, thus predisposing patients with AD to have increased skin infections. In addition to keratinocytes, endothelial cells, macrophages, mast cells, and basophils are other cellular sources of IL-33. IL-33 is stored preformed in the nuclei of these cells and produced readily to exert its inflammatory effects. It attaches to its receptor (ST2) on ILC2 to activate the production of IL-5 and IL-13. IL-25 acts on both ILC2 and T cells by attaching to its receptor, IL-17RB. In combination with IL-33 and thymic stromal lymphopoietin, it enhances the proliferation and cytokine expression by ILC2. Both IL-33 and IL-25 are highly expressed in AD lesions.

Defects in dendritic cells also contribute to increased infections in AD. Both myeloid and plasmacytoid dendritic cells in patients with AD produced significantly less interferon-α. Toll-like receptor-2 (TLR-2)-sensing of S. aureus by Langerhans cells and inflammatory dendritic epidermal cells has also been found to be impaired in patients with AD. Natural killer cells have recently been found to be deficient in patients with AD. This deficiency may also contribute to increased type 2 inflammation owing to a potential counter-regulatory mechanism between the natural killer cells and type 2 inflammation.

Staphylococcus aureus Colonization

Up to 90% of patients with AD are colonized with S. aureus. This predominance of S. aureus is unique to AD, as compared with healthy individuals and patients with another chronic inflammatory skin disease, psoriasis. The predominance of S. aureus in AD may be attributed to the virulence factors of this bacteria and its ability to evade the cutaneous immunity of patients with AD. S. aureus fibronectin has a special affinity for type 2 inflammation. In addition, S. aureus produces enterotoxins (superantigens) that are known to break down the skin barrier and enhance type 2 inflammation. Superantigens also down-regulate the cutaneous production of interferon-γ and tumor necrosis factor-α, both of which are important mediators of cellular immunity against bacterial and viral infections. Methicillin-resistant S. aureus (MRSA) has been found to produce significantly more superantigens than methicillin-sensitive S. aureus (MSSA). Both superantigens and another staphylococcal toxin, the α toxin, may contribute to keratinocyte apoptosis and barrier defects in AD. Staphylococcal δ toxin may also contribute to AD inflammation by inducing mast cell degranulation.

Dysbiosis of Skin Flora

The maintenance of healthy skin also depends on its commensal microbiome. Normal skin flora is found beyond the surface of the epithelium, which highlights the protective role in immune defense and regulation. The most abundant microbes consist of Cutibacterium acnes (formerly known as Propionibacterium acnes), Corynebacterium, and coagulase-negative Staphylococcus (CoNS). Patients with AD are deficient in commensal bacteria, and this facilitates the virulence of S. aureus in lesional skin. The roles of commensal bacteria are 2-fold as follows: (1) their ability to modulate the host immune system to minimize inflammation and to increase protection against microbial pathogens; (2) their ability to directly outcompete microbial pathogens such as S. aureus. CoNS, S. epidermidis, was found to produce a lipoateic acid that is capable of preventing injury-induced TLR-3-mediated cutaneous inflammation by means of TLR-2 interaction. S. epidermidis also modulates host cytotoxic and regulatory T cells in wound repair and immune tolerance, respectively. In addition to its anti-inflammatory role, S. epidermidis may also up-regulate antimicrobial peptide production by keratinocytes to protect against microbial pathogens. CoNS including S. epidermidis, S. lugdunensis, and S. hominis are capable of producing proteases or antimicrobial factors that either prevent biofilm formation by S. aureus or are bactericidal against it.
Typically presents with oozing serum that has dried up, giving it a honey-crusted appearance surrounded by an erythematous base (Fig 2). Impetigenous lesions may also present with fluid-filled blisters (bullous impetigo), which may be mistaken for EH. Nonpurulent SSTIs include erysipelas and cellulitis. These infections usually start in a focal skin area but may spread rapidly to cover the major parts of the body such as the arms, legs, trunk, or face.23 Focal erythema, swelling, warmth, and tenderness are signs of these infections. These patients may experience fever and bacteremia. Purulent SSTI presents as skin abscesses, which may be fluctuant or nonfluctuant nodules or pustules surrounded by an erythematous swelling. The lesions may be tender and warm. MRSA is a common cause of these lesions. In addition, SSTI in patients with AD may lead to systemic complications, which include bacteremia, osteomyelitis, septic arthritis or bursitis, and, more rarely, endocarditis and staphylococcal scalded skin syndrome, which is mediated by staphylococcal toxins. Persistent fever and specific signs—including an ill-looking appearance, lethargy (bacteremia), focal point tenderness of the bone (osteomyelitis), joint swelling (septic arthritis/bursitis), heart murmur (endocarditis), and widespread desquamation (staphylococcal scalded skin syndrome)—should raise suspicion for these infections. Persistent elevated inflammatory markers such as C-reactive protein or erythrocyte sedimentation rate further increase the index of suspicion for these infections. MSSA and MRSA cause an equal proportion of infectious complications (40% each) in hospitalized children with AD.4 These infection rates are consistent with that of the general pediatric inpatient populations across the United States.24 The second most common cause of SSTI and systemic infections in AD is Streptococcus pyogenes. Str pyogenes may cause infections in patients with AD by itself or in combination with S aureus. These skin infections typically present with pustules or impetigo. The lesions may appear as punched-out erosions with scalloped borders that mimic EH.25 Although SSTI and systemic infections in AD present with overt signs that facilitate diagnosis and antibiotic treatment, the so-called infected eczema associated AD exacerbation is not as clearly defined.26 Patients with severe AD exacerbation tend to have more generalized cutaneous signs and symptoms. These include erythema, swelling, oozing, and tenderness, all of which may also be signs of skin infections. However, Cochrane analysis indicates that antibiotics do not improve the severity of AD in these patients.27 The main concern with the overuse of antibiotics in AD exacerbation is the potential development of bacterial resistance and dysbiosis.28 However, apart from the outcome of AD severity, there may be a subset of patients with severe AD exacerbation who may benefit from antibiotics in terms of infections or the prevention of infectious complications.4,28,29 It has been proposed that these patients may be differentiated by a higher density of S aureus and the amount of tissue damage caused by S aureus—host interaction.29 Children with severe AD exacerbation were found to have elevated C-reactive protein and erythrocyte sedimentation rate, although these levels were significantly less than that of patients with infectious complications.4 There may be a potential use in these inflammatory markers in identifying patients with AD who are at risk for severe infectious complications.

**Viral Infections**

EH is caused by infection with herpes simplex virus (HSV)—1, which is a potentially life-threatening infectious complication in patients with AD. Nearly a third of children who are hospitalized for AD infectious complications were related to EH.2 Younger age and non-White race (African Americans, Asians, and Native Americans) are at increased risk for hospitalization with EH.30 EH can manifest with skin pruritus or pain and presence of vesicles, punched-out erosions (Fig 3), or hemorrhagic crusts that can become more extensive. A local skin infection may progress to disseminated vesicles with skin breakdown. Systemic EH infection may present with fever, malaise, viremia, and complications including keratoconjunctivitis, encephalitis, and septic shock.

Exposure to HSV-1 is common in the general population and is present in 60% of adults and 20% of children.31 Immunologic and genetic elements likely contribute to the vulnerability of a subset of patients with AD, given that EH only affects 3% of patients with AD.31 Patients with AD and EH have been reported to have
Repeat decolonization of patient and all household contacts as follows:

1. Assess level of adherence with above regimen.

2. Education on best personal hygiene practices
   - Mechanisms of *S aureus* transmission (eg, skin-to-skin contact, fomites).
   - Emphasize personal hygiene practices
     - Frequent hand-washing with soap and water or an alcohol-based sanitizer.
     - Avoid reusing or sharing personal hygiene items that contact the skin (eg, towels, loofas, razors, cosmetics, brushes).
     - Avoid contamination of topical medications and moisturizers (use a pump or pour containers).
     - Keep fingernails clean and trimmed; avoid scratching.

3. Environmental hygiene measures
   - Regularly clean high-touch surfaces (eg, counters, door knobs, and appliances) with commercially available disinfectants.
   - Use a barrier between exposed skin and high-touch surfaces touched by multiple people (eg, exercise equipment).
   - Wash hands before and after touching pets.
   - Wash bedding at onset and completion of decolonization regimen.
   - Wash clothing, towels, and washcloths with hot water and detergent before reuse.
   - Use a barrier between exposed skin and high-touch surfaces touched by multiple people (eg, exercise equipment).
   - Dilute bleach baths may be preferable to chlorhexidine solutions in patients with AD because chlorhexidine can cause skin irritation. Repeat exposure can lead to resistance and it is costlier.
   - Chlorhexidine can be applied as a wash or disposable wipe. Care should be taken to avoid contact with the face and the 4% solution should be thoroughly rinsed off with water after application.
   - Can consider changing decolonizing agents.

4. Personal and household decolonization
   - Nasal decolonization with intranasal mupirocin 2% ointment twice daily for 5-10 days.
     - Topical decolonization with either one of the following:
       - Dilute bleach baths for 15 min twice weekly (1 tsp of bleach per gallon of water or ¼ cup of bleach per ¼ standard tub or 13 gallons of water) for 3 mo.
       - Chlorhexidine gluconate 4% solution once daily for 5-14 d.
       - Dilute bleach baths for 15 min twice weekly with chlorhexidine washes daily on days bleach baths not given for 3 mo.

5. If recurrent infections despite decolonization
   - Optimize underlying condition, personal, and environmental hygiene.
   - Assess level of adherence with above regimen.
   - Repeat decolonization of patient and all household contacts as follows:
     - Intranasal mupirocin 2% ointment twice daily for 5 d once or twice a month for 6 mo.
     - Topical decolonization with dilute bleach baths as above twice weekly or chlorhexidine gluconate solution as above for 5 d every 2 wk for 6 mo.

May consider concomitant use of oral antibiotic therapy on a case-by-case basis with rifampin and another oral agent to which the isolate is susceptible to for 5-10 days.

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**Table 1**

| Decolonization strategy |
|--------------------------|
| 1. Optimize underlying condition |
| Daily skin care |
| - Topical corticosteroid or calcineurin inhibitor for eczema areas. |
| - Emollients for unaffected areas. |
| Basic wound care measures for severe eczema lesions (eg, covering open or weeping wounds to prevent the spread and secondary infection). |
| Avoidance of triggers for eczema flares. |
| 2. Education on best personal hygiene practices |
| Mechanisms of *S aureus* transmission (eg, skin-to-skin contact, fomites). |
| Emphasize personal hygiene practices |
| - Frequent hand-washing with soap and water or an alcohol-based sanitizer. |
| - Avoid reusing or sharing personal hygiene items that contact the skin (eg, towels, loofas, razors, cosmetics, brushes). |
| - Avoid contamination of topical medications and moisturizers (use a pump or pour containers). |
| - Keep fingernails clean and trimmed; avoid scratching. |
| 3. Environmental hygiene measures |
| - Regularly clean high-touch surfaces (eg, counters, door knobs, and appliances) with commercially available disinfectants. |
| - Use a barrier between exposed skin and high-touch surfaces touched by multiple people (eg, exercise equipment). |
| - Wash hands before and after touching pets. |
| - Wash bedding at onset and completion of decolonization regimen. |
| - Wash clothing, towels, and washcloths with hot water and detergent before reuse. |
| - Use a barrier between exposed skin and high-touch surfaces touched by multiple people (eg, exercise equipment). |
| - Dilute bleach baths may be preferable to chlorhexidine solutions in patients with AD because chlorhexidine can cause skin irritation. Repeat exposure can lead to resistance and it is costlier. |
| - Chlorhexidine can be applied as a wash or disposable wipe. Care should be taken to avoid contact with the face and the 4% solution should be thoroughly rinsed off with water after application. |
| - Can consider changing decolonizing agents. |

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Molluscum contagiosum (MC) is a poxvirus that belongs to the Molluscipoxvirus subfamily, but it is distinct from vaccinia, variola, and cowpox viruses in the Orthopoxvirinae genus. MC infection in patients with AD may be diffused or along the AD distribution (Fig 5). Skin barrier defects predispose patients with AD to MC, and long-term scratching leads to the spread by autoinoculation. MC infection in AD has been associated with *FLG* LoF.

Eczema vaccinatum (EV) is a life-threatening infection in patients with AD that is caused by a live vaccinia virus in smallpox vaccines. EV is reported to be rare since the discontinuation of routine smallpox vaccination in 1971. In 2002, owing to the concern that smallpox virus may be used as a weapon for bioterrorism, a national program began to vaccinate US military members, select laboratory researchers, and first responders with the smallpox vaccine. Pre-outbreak smallpox vaccine is contraindicated in persons with a history of AD or persons who are in close contact with patients with AD. With careful screening, there have only been a few cases of disseminated EV or EV by autoinoculation since 2002. Most of the affected patients have been either military members or close contacts of military members who had a recent history of smallpox vaccination. Although rare, an acute presentation of vesiculopustular/nodular rash in a patient with AD with a military background, or who had close contact with military
infection, anti-inflammatory treatment should confer protection against infections in patients with AD (Fig 6). Dupilumab, a monoclonal antibody that targets the IL-4α receptor to neutralize the effects of IL-4 and IL-13, was found to decrease S aureus colonization and increase microbial diversity.48 Pooled analysis of dupilumab clinical trials revealed significant improvement in SSTI and EH as compared with placebo.49,50 These observations are consistent with the suppressive effects of IL-4 and IL-13 on skin barrier functions and endogenous antimicrobial peptide expression in AD lesions, predisposing patients with AD to increased infections. Because of the current unprecedented global pandemic caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), there has been some concern about whether systemic anti-inflammatory medications for AD including dupilumab may increase the risk of patients with AD for this viral infection. Case series, mainly from Italy, thus far, have not supported an increased risk of SARS-CoV-2 infection in patients with AD who are treated with dupilumab. A global web-based registry has been set up for clinicians to monitor the risks and outcomes of SARS-CoV-2 in patients with AD receiving systemic agents, including dupilumab (www.covidderm.org).

Attempts to decolonize S aureus is largely experimental. There is insufficient evidence that diluted bleach bath and antibiotics result in sustained decolonization of S aureus in AD.27 Dilute bleach at 0.005% was not suppressive of S aureus growth or toxin production.51 Acetic acid (apple cider vinegar) has been used as an anti-microbial bathing additive for AD,52 although its efficacy in S aureus clearance in AD has not been established. Another study reported that 0.5% acetic acid daily bath for 14 days did not improve skin barrier function or acidity in patients with AD, as compared with plain water baths.53 In contrast, skin irritation was reported in some patients treated with dilute acetic acid. Chlorhexidine bath has been used in the decolonization of MRSA in the general population, but it has not been studied adequately in AD. A potential adverse effect of personnel, along with a recent history of smallpox vaccination, should raise an index of suspicion for EV.

Prevention of Infections in AD

The approach in preventing infections in AD is based on addressing the predisposing factors for infections. Daily skin hydration and moisturization are recommended for patients with AD to maintain skin barrier function.26 Patients with AD should take a daily warm shower or bath, followed by gentle drying and application of a moisturizer or a prescribed topical medication. The choice of moisturizer should be based on the patient’s or parent’s preference and experience. In general, a thick or ointment-based moisturizer (eg, petrolatum) is better than cream in retaining moisture in the skin. Application of petrolatum has been indicated to up-regulate antimicrobial peptides and induce key barrier differentiation markers such as filagrin and involucrin, in patients with AD.27 The use of standard topical anti-inflammatory medications, including topical corticosteroids (TCS) and topical calcineurin inhibitors (TCI), have been reported to improve skin barrier functions based on transepidermal water loss.38-40 Furthermore, TCS and TCI have been reported to decrease S aureus colonization in AD lesions.41-44 Topical anti-inflammatory treatments have been associated with increased microbial diversity in AD lesions.55,46 Although multiple case reports have found an association between EH and the use of anti-inflammatory medications in AD, this was not supported by a recent multicenter study, which reviewed more than 200 cases of EH.47 The authors found that the use of TCS, TCI, systemic corticosteroid, or cyclosporine was not associated with the onset of EH. Uncontrolled AD inflammation is likely the primary risk factor for EH (or bacterial infections), rather than the anti-inflammatory treatment. Therefore, in the absence of an active infection, anti-inflammatory treatment should confer protection
this antimicrobial agent is allergic contact dermatitis. 54 The Infectious Diseases Society of America published guidelines for the management of recurrent SSTI because of MRSA in 2011. 55 Similar principles apply to the management of recurrent SSTI in AD. Based on these guidelines and findings of more recent studies/expert opinion, a suggested approach to the decolonization of S. aureus in patients with AD with recurrent SSTIs is outlined in Table 1. 15–57

Management of Infectious Complications in AD

Approximately 20% of children with AD hospitalized for infectious complications had invasive bacterial infections. 4 For patients with AD who have signs and symptoms of systemic illness, hospitalization and empirical intravenous antibiotics are recommended. The empirical antibiotic regimen should provide coverage against S. aureus because this is the most frequently identified bacterial pathogen in AD. For critically ill patients, coverage for both MRSA and MSSA with vancomycin and an antistaphylococcal beta-lactam is appropriate because vancomycin is inferior to nafcillin or first-generation cephalosporins for the treatment of serious MSSA infections. 56,57 For severe but non–life-threatening infections, vancomycin may be used alone as empirical therapy, pending culture results. Clindamycin can also be considered if there is no concern for an endovascular infection and the local prevalence of clindamycin resistance is less than 15%. 56 Bacteremia because of S. aureus requires the use of a bactericidal intravenous agent initially. For MRSA, vancomycin is the drug of choice. For MSSA, cefazolin and nafcillin are both acceptable first-line agents, though nafcillin can cause venous irritation and phlebitis when administered peripherally. As long as there are no concerns for ongoing bacteremia or an endovascular focus, completion of therapy with an oral agent to which the isolate is susceptible is appropriate in children with S. aureus bacteremia. 58 Duration of therapy should be determined by the clinical response, but typically 7 to 14 days is recommended. Infective endocarditis is a rare complication of AD. 59 Careful auscultation for a heart murmur is recommended.

For patients with AD with uncomplicated, nonpurulent skin infection, a beta-lactam antibiotic that covers both S. aureus and β-hemolytic streptococci (eg, cefazolin or cephalexin) may be sufficient pending clinical response or culture, taking into account local epidemiology and resistance patterns. 60,61 In contrast, for patients with AD with a skin abscess, history of MRSA colonization, close contacts with a history of skin infections, or recent hospitalization, coverage for MSSA should be considered. Clindamycin, doxycycline, trimethoprim-sulfamethoxazole, and linezolid are all acceptable oral options for MRSA skin infections in both children and adults, assuming the isolate is susceptible in-vitro. 55 Of note, the rates of clindamycin resistance have been rising among both MRSA and MSSA nationally, though there is regional variation. 62 Patients with AD with minor, localized skin infections such as impetigo may be treated with topical mupirocin ointment. The duration of therapy typically ranges from 5 to 10 days depending on clinical response. 55,63 Lesional HSV polymerase chain reaction should be sent on suspicion of EH. However, treatment with systemic antiviral should not be withheld pending the results of HSV testing. Coinfection of EH with S. aureus is also common and concurrent treatment with an anti-S. aureus antibiotic should be considered. Table 2 provides the antiviral treatment options for EH and suggested dosing in adults and children. There are no formal guidelines regarding the preferred route of administration of antivirals or indications for hospitalization in patients with EH. For patients with extensive skin involvement, signs of systemic illness, and those less than 1 year of age, parenteral acyclovir should be considered initially. Fever and mild systemic symptoms often accompany mucocutaneous HSV infections, particularly with the initial episode. Once clinical improvement is reported, a transition to an oral agent to complete the course of therapy is appropriate. For mild cases, oral acyclovir can be considered and was associated with faster healing and resolution of pain in a randomized, placebo-controlled trial of 60 adults and adolescents with EH. 64 Valacyclovir, the L-valyl ester prodruk of acyclovir, has a 3 to 5-fold greater bioavailability than oral acyclovir, which can be dosed less frequently, and with plasma concentrations are comparable with parenteral acyclovir. 55 Topical antivirals do not have an appreciable benefit in HSV mucocutaneous disease and do not have a role in the treatment of EH. 58 Patients with recurrent EH may benefit from long-term suppressive therapy, though this has not been studied. Suggested oral suppressive dosages are listed in Table 2. The need for ongoing therapy should be reassessed after 6 to 12 months. The development of resistance to acyclovir is rare in EH, but may be suspected in cases of recalcitrant EH or frequent recurrences of EH despite suppressive therapy and good adherence to long-term therapy. 50,65 Forscarnet is the recommended therapy for acyclovir-resistant HSV infections, because acyclovir-resistant HSV isolates are also resistant to valacyclovir.

The treatment for EC is supportive of the continuation of routine skin care and AD treatments, including TCS. MC is benign, and observation is recommended in most cases. An attempt should be made to minimize scratching that spreads the lesions. This includes daily skin care and topical anti-inflammatory treatments. Sedating fast-acting antihistamines may be helpful in decreasing scratch during sleep. Treatments such as curettage, cryotherapy, salicylic acid, imiquimod, and cantharidin (beetle juice) are associated with either pain, the risk for scarring, or mixed results of efficacy. 66 However, a more recent randomized placebo-controlled trial has indicated efficacy in the use of cantharidin for the treatment of pediatric MC. 67 When evaluating pustule-vesicular rash in patients with AD with a military background or a history of close contact with a military personnel who had a recent vaccination, an index of suspicion for EV should be raised. Suspected cases should be reported to the Centers for Disease Control and Prevention Emergency Operation Center for assistance in diagnosis and management. EV patients with systemic symptoms may require treatment with vaccinia immune globulin.

Potential Therapy in the Pipeline

A number of agents currently in the pipeline that may help in the prevention of infections in AD include anti-inflammatory treatments that target type 2 inflammation. 54 These include monoclonal antibodies that target IL-13, IL-33, thymic stromal lymphopoietin, and OK40. Janus kinase inhibition has also been shown to reduce inflammation and improve skin barrier in AD. Both topical and oral janus kinase inhibitors are in various phases of clinical trials. Topical delgocitinib has been approved for AD in Japan. 55 Pruritus and associated scratching in AD can contribute to significant damage to the skin barrier and new therapeutic options are needed. A long-term trial with nemilizumab (anti–IL-31 receptor A monoclonal antibody) reported improvement in pruritus and AD severity. 54 Other anti-itch treatments under investigation include transient receptor potential melastatin agonists and vanilloid antagonists. 68 Improvement of skin barrier function and cutaneous innate immunity in AD is of interest, because it may prevent external triggers and skin infections. 67 Although the attempt to prevent AD in healthy infants with a daily emollient application has been disappointing 58 whether or not skin barrier functions may be modified in established AD remains to be investigated. Aryl hydrocarbon receptor
agonists, which increase filaggrin expression, were found to improve AD and endogenous antimicrobial production in preliminary studies. Directly targeting *S. aureus* is also an active area of investigation. These treatments include novel products with anti-*S. aureus* activity, synthetic antimicrobial peptides, and *S. aureus* lytic agents. There is currently no approved *S. aureus* vaccine. However, approaches that target *S. aureus* toxins are in development. There is increasing evidence that topicaly applied probiotics may be a viable approach against *S. aureus* in AD. In a small study, a gram-negative bacteria, *Roseomonas mucosa*, was found to improve AD and decrease *S. aureus* burden in adults and children with AD. *S. hominis* strains were found to produce an autoinducing peptide that is capable of inhibiting *S. aureus* accessory gene regulatory quorum sensing system and prevent biofilm formation by *S. aureus*.

**Conclusion**

AD is a complex disease associated with skin barrier defects that result in allergen or pathogen invasion and dysfunctional immune responses, causing a vicious cycle of inflammation. The skin microbiome is altered because of this dysregulation and pathogenic organisms such as *S. aureus* are more likely to colonize the skin. The combination of skin barrier defects, immune dysregulation, and alteration in the skin microbiome result in an increased risk for skin infections.

The prevention of infection in AD should emphasize skin barrier repair and maintenance anti-inflammatory medications without relying on antibiotics. The need for antibiotics in patients with severe AD exacerbations remains controversial. This is because some of the signs and symptoms associated with severe AD exacerbation resemble that of bacterial skin infections. It is possible that there is a threshold at which *S. aureus* levels and the extent of host tissue damage evolve into an infection. Studies are needed to investigate the biomarkers that assist in determining this threshold. Acute-phase response markers such as C-reactive protein and erythrocyte sedimentation rate may be helpful in determining the need for antibiotics in patients with severe AD exacerbation who are suspected of having infections. Future studies should also address whether anti-inflammatory treatments, especially those that specifically target type 2 inflammation, may benefit patients with AD with active infection. This is based on the premise that suppressing type 2 inflammation may lead to the improvement of immunity against microbial pathogens.

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