From polymers to proteins – novel phases of short compact tubes

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A framework is presented for understanding the common character of proteins. Proteins are linear chain molecules – however, the simple model of a polymer viewed as spheres tethered together does not account for many of the observed characteristics of protein structures. We show here that the notion of a tube of non-zero thickness allows one to bridge the conventional compact polymer phase with a novel phase employed by Nature to house biomolecular structures. We build on the idea that a non-singular continuum description of a tube (or a sheet) of arbitrary thickness entails discarding pairwise interactions and using appropriately chosen many body interactions. We suggest that the structures of folded proteins are selected based on geometrical considerations and are poised at the edge of compaction, thus accounting for their versatility and flexibility. We present an explanation for why helices and sheets are the building blocks of protein structures.
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I. INTRODUCTION

Recent years have witnessed gigantic leaps in the field of molecular biology culminating in the sequencing of the human genome as reported in two historic issues of Science (Volume 291, issue 5507) and Nature (Volume 409, issue 6822) in 2001. Base pairing and the remarkable structure of the DNA molecule (Watson and Crick, 1953) provide a very efficient means of storing and replicating genetic information. The principal role of genes is to serve as a template for the synthesis of m-RNAs that, in turn, are “translated” by ribosomes into the polypeptide chains which then fold into active proteins. These proteins are the work horse molecules of life. They not only carry out a dizzying array of functions but also they interact with each other and play a role in turning the genes on or off. There is little variability in the structure of the information carrying molecule, DNA. On the other hand, there are several thousand geometries which folded proteins can adopt and these structures determine the functionality of the proteins (Branden and Tooze, 1999; Creighton, 1993; Fersht, 1998).

Proteins are the basic constituents of all living cells. Some familiar examples of proteins are hemoglobin, which delivers oxygen to our tissues, actin and myosin which facilitate the contraction of our muscles, insulin which is secreted in the pancreas and signals our body to store excess sugar and antibodies that fight infection. Marvelous machines within the cell known as ribosomes make proteins by stringing together little chemical entities called amino acids into long linear chains. There are twenty types of amino acids which differ only in their side chains. The protein backbone as well as some of the side chains are hydrophobic and shy away from water, while other side chains are polar and yet others have charges associated with them.

Our focus is on small, globular proteins, which, under physiological conditions, fold rapidly and reproducibly (Anfinsen, 1973) in a cooperative fashion into a somewhat compact state in order to expel the water from the core of the folded structure, which predominantly houses
the hydrophobic amino acids. Thus there is an effective attraction between the hydrophobic amino acids arising from their shared tendency to avoid the water.

For proteins, form determines function. The structure of the protein in its folded state (also called its native state structure) controls its functionality (Branden and Tooze, 1999; Creighton, 1993; Fersht, 1998). The rich variety of amino acids allows for many sequences to have the same native state structure. Thus even though our human body may have more than 100,000 proteins, it is believed that the number of distinct folds that they adopt in their native state, is only a few thousand in all (Chothia, 1992). Furthermore, these folds are beautiful (Levitt and Chothia, 1976; Chothia, 1984) – they are not just any compact form but, rather, are made up of building blocks of helices and sheet-like planar structures with tight loops connecting these secondary motifs (see Figure 1).

In 1939, J. D. Bernal (1939) stated the challenge associated with the protein problem: *Any effective picture of protein structure must provide at the same time for the common character of all proteins as exemplified by their many chemical and physical similarities, and for the highly specific nature of each protein type.* Despite many advances in experiments on proteins and the advent of powerful computers, the problem has remained largely unsolved. The key components of the problem are protein folding and design: protein folding entails the prediction of the folded geometry of a protein given its sequence of amino acids while the design problem involves the prediction of the amino acid sequence which would fold into a putative target structure. It is probably not too surprising that progress has been somewhat limited because, until now, there has not been any simple unifying framework for understanding the common character of all proteins. The principal aim of this colloquium is to address this issue. Such a framework must provide an explanation for the relatively small number of protein native structures, for why the building blocks of protein structures are helices and sheets, for the highly cooperative nature of the folding transition of small globular proteins and for the versatility and flexibility of protein structures, which account
for the ability of the proteins to perform a wide range of functions.

II. QUANTUM CHEMISTRY SCORES A MAJOR SUCCESS

Linus Pauling and his collaborators (Pauling, Corey and Branson, 1951; Pauling and Corey, 1951) invoked the chemistry of covalent and hydrogen bonds to show that helices and sheets were periodically repeatable structures for which appropriately placed hydrogen bonds could provide the scaffolding. This stunning prediction was experimentally confirmed in short order. Unfortunately, these observations do not provide a complete explanation of the selection of the protein folds. The difficulty arises because hydrogen bonds can equally easily form between the protein molecule and the water surrounding it. While helices and sheets are nicely stabilized by hydrogen bonds, one may construct other viable structures which do not have helices or sheets as the building blocks but yet have a large number of hydrogen bonds and hence a favorable energy.

A protein is complex because of the many features that one is confronted with. As mentioned before, we need to deal with twenty types of amino acids with their individual properties and, in addition, there is the crucial role played by the solvent. A first principles approach might consist of considering all the numerous atoms making up a protein and the surrounding solvent and carrying out some really heavy computer calculations to simulate the folding process. Very quickly one realizes that, with the somewhat imperfect knowledge of the interactions and the sheer magnitude of the job at hand, this approach is not too likely to yield qualitatively new insights into the protein folding problem. Furthermore, one might worry that, at best, one would be able to mimic Nature but would one obtain an understanding of Nature?
III. A PHYSICS APPROACH LEADS TO A DISCONNECT BETWEEN THE
COMPACT POLYMER PHASE AND THE NOVEL PHASE ADOPTED BY
PROTEIN STRUCTURES

Let us now consider the protein problem afresh from a physics point of view and attempt to identify the key issues. It is of course possible and, one might fear, even likely that many of the details are crucial in understanding the intricate behavior of proteins. In order to make progress, we will take the approach, though, of looking at what we might imagine to be the the most essential features and adding in details as we require them. This will allow us to retain some control over our understanding and we will be able to assess, a posteriori, the relative importance of the features that we may have to incorporate.

The approach is analogous to one commonly used in physics (Chaikin and Lubensky, 1995) of distilling out just the most essential features for understanding emergent phenomena. For example, one can use general geometrical and symmetry arguments to predict the different classes of crystal structures. The existence of these structures does not rely on quantum mechanics or on chemistry. They are a consequence of a deeper and more general mathematical framework. Of course, given a chemical compound like common salt, a careful quantum mechanical study would show that sodium chloride adopts the face centered cubic lattice structure. Also, a clever grocer would use the same crystal structure for the efficient packing of fruits. Thus the structures transcend the specifics of the chemical entities that are housed in them. One might therefore seek to determine what the analogous structures are for protein native states that are determined merely by geometrical considerations. What are the bare essentials that determine the novel phase adopted by biopolymers such as proteins?

Proteins are linear chains and ignoring the details of the amino acid side-chains, all proteins have a backbone. A protein folds because of hydrophobicity or the tendency of certain amino acids to shy away from water. In the folded state, therefore, one would like to have
a conformation which squeezes the water out from certain regions of the protein populated by the hydrophobic amino acids. As stated before, the simplest way of encapsulating such a tendency for compaction is by means of an effective attractive interaction between the backbone atoms promoting a somewhat compact native state.

An early success of the physics approach was the work of Ramachandran, Ramakrishnan and Sasisekharan (Ramachandran and Sasisekharan, 1968), as embodied in the Ramachandran plot. They showed that steric constraints, relating to or involving the arrangement of atoms in space, alone dictated that the backbone conformations of a protein lie predominantly in two regions of the space of the so called torsional angles corresponding to α-helical and β-strand conformations (see Figures 1-3). In other words, the high cost associated with the overlap of two atoms viewed as hard spheres leads to conformations which are consistent with the local structure associated with a helix or a sheet.

We hit a snag in our thought experiment – careful computational studies (Hunt et al., 1994; Yee et al., 1994) have shown that the standard polymer model of chain molecules, viewed as spheres tethered together, when subjected to interactions which promote compactness, have innumerable conformations almost none of which have any secondary motifs. In contrast, proteins have a limited number of folds to choose from for their native state structure and the energy landscape is vastly simpler. In addition, the structures in the polymer phase are not specially sensitive to perturbations and are thus not as flexible and versatile as protein native state structures are in order to accommodate the dizzying array of functions that proteins perform. Indeed, there has been somewhat of a disconnect between the familiar compact polymer phase and the novel phase used by Nature to house biomolecules. To quote from P. J. Flory (1969): *Synthetic analogs of globular proteins are unknown. The capability of adopting a dense globular configuration stabilized by self-interactions and of transforming reversibly to the random coil are peculiar to the chain molecules of globular proteins alone.*
So what new feature should we incorporate next? Are the details of the amino acids important? We expect not because it is known that many sequences fold into the same native state structure (Branden and Tooze, 1999; Creighton, 1993; Fersht, 1998). At a somewhat simpler level, we recall the work of Ramachandran (Ramachandran and Sasisekharan, 1968), who showed that steric interactions or the undesirability of two atoms to sit on top of each other, even when the atoms are treated as effective hard spheres, lead to certain regions of conformational space being excluded for a protein chain (Rose, 1996). The side chains of the amino acids occupy space as well, and thus it seems important to allow for room around the backbone to accommodate these atoms. So we proceed by incorporating a new ingredient – let us treat the protein backbone not as a chain of spheres but as a tube of non-zero thickness analogous to a garden hose. So how does such a tube behave if it has an effective attractive self-interaction that tends to make its conformation somewhat compact? There is some hope on the horizon, because the problem is enriched and we now have two length scales to play with, the thickness of the tube and the range of the attractive interactions.

It is useful to consider what the standard model of a chain represented as tethered spheres is missing. For unconstrained particles, spheres are the simplest objects that one might consider. Of course, symmetry matters a great deal and when these spheres are replaced by asymmetric objects, one gets a host of qualitatively new liquid crystalline phases (Chaikin and Lubensky, 1995). There are two simple ingredients associated with a chain – the particles are tethered to each other (which is well-captured by the standard model of tethered spheres) and associated with each particle of the chain is a special direction representing the local direction associated with the chain as defined by the adjacent particles at that location. This selection of a local direction immediately leads to the requirement that the symmetrical spherical objects comprising the chain must be replaced by anisotropic objects (such as coins) for which one of the three directions is different from the other two. Thus,
if one were to think of a chain as being made up of stacked coins instead of spheres, one naturally arrives at a picture of a tube. Indeed, previous analysis (Banavar et al., 2002 (a); Banavar et al., 2002 (b)) of the native state structures of proteins has shown that a protein backbone may be thought of approximately as a uniform tube of radius 2.7Å. Before we explore the phases associated with a tube subject to compaction, we will have an interlude where we will revisit some issues in polymer physics.

V. STRINGS, SHEETS AND MANY-BODY INTERACTIONS

Strings and chains have been studied over the years in the field of polymer physics. Tubes of non-zero thickness are ubiquitous – garden hoses and spaghetti populate our houses. How does one mathematically describe a tube of non-zero thickness in the continuum limit? Alas, a visit to the library confirms our worst fears, this elementary problem has not been tackled before. A continuum description of a string was put forward by Sam Edwards (Doi and Edwards, 1993) – it captures self-avoidance by means of a singular delta function repulsion between different parts of a string. The delta function describes a situation in which the repulsive interaction is infinitely strong precisely when there is an exact overlap and zero otherwise. This description is therefore valid only for an infinitesimally thin string. An associated complication is that the analysis of a continuum string requires the use of renormalization group theory to regularize the theory by introducing a lower-length scale cut-off combined with a proof that the behavior, at long length scales, is independent of this cut-off length scale. Unfortunately, the renormalization group theory analysis, in this context, is peripheral to the physics being studied.

Recently, with the help of two mathematicians, Oscar Gonzalez and John Maddocks, we were able to write down a singularity-free description of manifolds such as chains or sheets (Banavar et al., 2002(c)). The solution is very simple but not intuitively obvious. In science,
the starting point for describing interacting matter is by means of pair-wise interactions. In order to describe your interactions with your friends, it is a good starting point to consider your pair-wise interactions with each of them – your true interaction will be different from this only because of genuine many body interactions that may be thought of as higher order corrections. With a pair-wise interaction, there is only one length scale that one can construct from a knowledge of where you are and where your friend is. This length scale is your mutual distance. One can define potential energies of interaction between you and your friend which depends on this length scale. Generically, such an interaction may be one in which if you and your friend are separated by a sufficiently long distance, you do not talk to each other and there is no interaction. There is an optimal distance between you and your friend where the interaction is at its happiest. Any closer approach leads to a higher energy with the potential energy becoming infinitely large when you sit on top of each other.

Unfortunately, such an analysis is not very helpful when you and your friends (and your enemies) are formed into a conga line by someone who does not know what your personal relationships are. Let us assume that one is working again with pair-wise interactions and you are told that two people are spatially (not necessarily emotionally) close to each other. With that information alone, you will not be able to tell whether the two people are from different parts of the chain and are close to each other because they like each other or whether the two people are sworn enemies who happen to be close to each other simply because they were constrained to be next to each other in the conga line. In other words, pair-wise interactions merely provide the mutual distance but not the context in which the interacting particles exist.

The basic idea behind the development of a continuum theory of a tube of non-zero thickness is to discard pair-wise interactions and consider appropriately chosen three-body interactions as the basic interacting unit. The requirements for a well-founded theory are that one ought to be able to take a continuum limit on increasing the density of particles, that self-
interactions be properly taken into account and that there be a characteristic microscopic length other than the spacing between neighboring particles along the string.

Let us consider a three-body potential characterizing the interaction between three particles, which lie on the corners of a triangle. Let the sides of the triangle have magnitudes $r_1$, $r_2$ and $r_3$. In order to specify a triangle uniquely, one needs three attributes. The potential of interaction can therefore depend on three independent length scales, which are invariant under translation, rotation and permutation of the three particles. We choose these length scales to be the perimeter, $P$, of the triangle, the ratio of the area, $A$, of the triangle to its perimeter, $P$, and finally $r_1r_2r_3/A$. The first two lengths do not cure the problems alluded to before – they both vanish when the particles approach each other and cannot distinguish between particles from the same region or different regions of the string. The third length scale is proportional to $R$, the radius of a circle drawn through the three particles and has proved to be valuable for the study of knots (Gonzalez and Maddocks, 1999). This length scale neatly solves the contextual problem mentioned above. When two parts of a chain come together, the radius of a circle passing through two of the particles on one side of the chain and one particle from the other side of the chain turns out to be a measure of the distance of approach of the two sides of the chain. On the other hand, when one considers three particles consecutively along the chain, the radius of the circle passing through them is simply the local radius of curvature. Indeed when three such particles form a straight line, the radius goes to infinity and the three particles essentially become non-interacting. The straight line configuration is the best that the particles can do in terms of staying away from each other given that they are constrained to be neighbors along the chain. Our suggestion is to use a generic potential energy function such as the one described previously but with this three-body radius as its argument.

How might one define the thickness of a tube associated with a chain configuration? A simple procedure would be to construct a tube whose axis coincides with the chain and inflate the
tube uniformly until it intersects with itself or has sharp corners. A natural definition of the thickness is then the radius of this largest tube (Katritch et al., 1996). A tube with a large thickness has more space for internal rearrangements of the side chains of the amino acids than a thinner tube. This thickness can also be obtained using the three-body interactions by computing the radius associated with all triplets (contiguous or otherwise) and selecting the smallest among these radii (Gonzalez and Maddocks, 1999). A simple way of describing a tube of non-zero thickness in the continuum limit is to discard pair-wise interactions and to consider triplet interactions. One may choose a simple potential energy which is a sum of 3-body terms whose argument is the 3-body radius and whose form has a hard-core at short distances – any radius (local or non-local) is forbidden from taking on a value less than the thickness of the tube (see Figure 2). Likewise, one may write down a continuum description for the self-avoidance of a sheet of paper (such as the page you are reading) of non-zero thickness by discarding pairwise and three-body interactions and employing appropriately chosen 4-body interactions as the basic interacting unit (Banavar et al., 2002(c)).

The new insight that one obtains with this continuum description is the important role of three-body interactions in characterizing tubes of non-zero thickness. It is important to stress that this elimination of pair-wise potentials and their replacement by effective three-body potentials is necessary only in the continuum limit. Also, the potentials we are discussing are effective potentials obtained on integrating what one hopes are irrelevant finer degrees of freedom. Our formulation not only allows one to carry out a continuum study of thick polymer chains but also it is useful for the study of chains in a given knot class or with a fixed number of knots. Any model employing a pairwise potential allows self-intersections, albeit with an energy penalty, so that the topology of the polymer chain (as measured by the knotting number or linking number) can be changed at will. This, of course, does not happen in real life with closed chains. Thus our non-singular many body potential allows one to formulate an analytic attack on the entropic exponents and weights of polymer configurations with a fixed linking number.
VI. MARGINALLY COMPACT TUBES

We return now to the protein backbone viewed as a tube of non-zero thickness. Consider a uniform tube undergoing compaction to expel the water away from the interior of its structure in the folded state. (We alert the reader to the fact that the tube we are considering is not hollow.) The backbone of all amino acids contains a carbon atom which is called a $C_\alpha$ atom. In a coarse-grained description, this atom may be chosen as the representative of the amino acid. For specificity, let us consider a discrete chain of $C_\alpha$ atoms of the protein backbone. As we have discussed, the notion of a tube thickness is captured by ensuring that none of the three-body radii is smaller than a threshold value equal to the radius of the tube. Let us also postulate that the attractive interactions promoting compaction are pairwise and have a given range. (Because we are considering a discrete situation, it is quite valid to have pairwise interactions.) There is one dimensionless quantity, which we will call $X$, that we will need to specify, which is the ratio of the thickness of the tube to the range of the attractive interactions.

When $X$ is very large compared to 1, the tube is so fat that it is unable to benefit from the attractive interactions. The constraints of the three body interaction dominate (the pairwise interaction plays no role) and one then obtains a swollen phase which consists of all self-avoiding conformations which satisfy the three-body radius constraint associated with the non-zero tube thickness. A vast majority of these conformations are ineffective in expelling the water from the interior of the structure. The non-zero thickness is loosely analogous to restricted space that others are not allowed to trespass. Imagine that your friend sits in the center of his/her room and requests that no one enter the room. The thickness then is proportional to the width of the room. If the range of attractive interactions is very small compared to this size, your ability to benefit from interactions with him/her is compromised.
by the fact that you cannot enter the room and for all practical purposes, it is as though your interactions with him/her were turned off. At the other extreme, for a tube with a very small $X$ compared to 1, one also obtains many, many conformations. This is because, in the room analogy, your interaction with your friend is sufficiently long range so that there is a lot of flexibility in where you position yourself. From a dynamical point of view, the structures obtained when $X \ll 1$ are somewhat inaccessible because the energy landscape is studded with numerous multiple minima. This situation is one in which the pairwise attractive interactions dominate and the three body radii constraints do not matter.

On varying $X$, we therefore expect two regimes, the phase with an effective long range attraction and the swollen phase, both with tremendous degeneracies. There is a twilight zone between these two phases, viewed as day and night, when $X$ is just shy of 1 (Figure 4). (We alert experts in the protein field that this crossover which we characterize colloquially as a twilight zone has no relationship to and should not be confused with the same terminology sometimes used in the studies of sequence similarity.) In this twilight zone, there is a rich interplay of the pairwise attractive interactions and the constraints imposed by the three-body interaction. This is a situation in which you are able to interact with your friend but can only do so by positioning yourself right outside his/her room.

In the twilight zone, a tube is barely able to avail itself of the attractive interactions promoting compaction. In this region of parameter space, the forces promoting compaction just set in and one would expect to obtain marginally compact structures which have the ability to expel the water from the interior. In addition, because the scale of the interaction strength is relatively small, one would expect a low ordering transition temperature with entropic effects not being too important. Furthermore, the physical picture of a tube (recall that a tube can be thought of as many anisotropic coins tethered together) leads to a strongly anisotropic interaction between nearby tube segments – it is better to position them parallel rather than perpendicular to each other. Thus, in the twilight zone, one has
a relatively weak and strongly anisotropic interaction. Because the tube segments have to position themselves next to each other and with the right relative orientation in order to avail themselves of the attractive self-interaction, one would expect a cooperative transition with few intermediates – the tube will need to snap into its correctly folded configuration. Also, because of the loss of flexibility regarding the relative positioning and orientation of nearby tube segments, one would expect a large decrease in the degeneracy.

In proteins, why is this effective value of $X$ tuned to be so close to its transition value of 1? The answer lies in the fact that the atomistic scale interactions are short range due to the screening effects of the water and, at the microscopic scale, the squeezing out of water is facilitated by the outer atoms of nearby side chains coming together. In a coarse-grained level of description, this translates into a value of $X$ which is close to 1, because, on the one hand, the necessity of having some wiggle room for the side chains of the amino acids leads to the tube picture and determines the tube thickness and, on the other hand, the same side-chains are responsible for and control the range of the attractive interactions promoting compaction.

There are several significant advantages in the system being poised in this twilight zone and having a limited number of marginally compact structures as the candidate native state conformations. In the thermodynamic limit of a tube of infinite length, there is a first order transition, on decreasing the tube thickness, between a swollen phase and a compact phase. This phase transition is characterized, nevertheless, by a diverging length scale – the propensity for nearby tube segments to be aligned just right with respect to each other leads to a diverging persistence length, defined as the characteristic length over which memory of the tube orientation is preserved.

Let us briefly review the well studied subject of phase transitions and critical phenomena (Stanley, 1999). Examples of critical points include a magnet at the onset of ordering, a
liquid-vapor system at the critical temperature and pressure, and a binary liquid system which is about to phase separate. The key point is that the fluctuations in a system at its critical point occur at all scales and the system is exquisitely sensitive to tiny perturbations. Even though sharp phase transitions can occur only in infinitely large systems, behavior akin to that at a phase transition is observed for finite size systems as well. Indeed, for a system near a critical point, the largest scale over which fluctuations occur is determined either by how far away one is from the critical point or by the finite size of the system.

A magnet at low temperatures compared to its critical temperature is well magnetized and is not very sensitive to a tiny external field. After all, when the magnetization is large, small perturbations do not lead to major consequences. Similarly, a magnet at very high temperatures is not very sensitive to a tiny external field because the strong thermal fluctuations dominate and the ordering tendencies are rather small. However, at the critical point, where there is about to be an onset of the magnetization, the system is very sensitive to an applied magnetic field and indeed the magnetic susceptibility for an infinite system diverges.

Nature, in her desire to design proteins to serve as smart and versatile machines, has used a system poised near a phase transition to exploit this sensitivity. Indeed, it is well known that proteins utilize conformational flexibility (Jacobs et al., 2001) to achieve optimal catalytic properties (Branden and Tooze, 1999; Creighton, 1993; Fersht, 1998). That protein structures are poised near a phase transition provides the versatility and the flexibility needed for the amazing range of functions that proteins perform.

In this marginally compact state, the number of candidate protein structures is somewhat limited. An energy landscape with relatively few energy minima associated with the protein folds has several consequences and advantages. First, each of these minima will have a correspondingly large basin of attraction. Second, a protein sequence has only a limited menu to choose from in deciding what its native state ought to be. A simple analogy is the
greater ease we have in selecting from a restaurant menu containing a few items than from one with innumerable choices. This selection is further reduced by the requirement that the native state be compatible with general chemical affinities and be free of steric clashes. The quality of match (Banavar and Maritan, 2001) between a sequence and a putative native state structure can be assessed by considering the propensity of the individual amino acids to be in distinct secondary structure elements such as the α-helix or a β-sheet, their likelihood of being buried or exposed and the degree to which the native state structure accommodates the ‘desire’ for certain pairs of amino acids to be in the vicinity of or away from each other. Strikingly, as seen in protein engineering experiments (Fersht, 1998), the ultimate choice of which fold a sequence adopts is dictated by a small number of key amino acids which have distinctly better environments in the native state than in competing folds. The limited number of these special folds underscores the key role played by the native state topology in determining many of the essential aspects of protein folding (Micheletti et al., 1999; Maritan et al., 2000 (a); Maritan et al., 2000(b); Baker, 2000). The powerful forces of evolution (Lesk and Chothia, 1980) operate within the fixed playground of these selected folds yielding better or more versatile sequences. Indeed, multiple protein functionalities can arise within the context of a single fold (Holm and Sander, 1997).

VII. BUILDING BLOCKS OF PROTEIN STRUCTURES

In order to determine the nature of the twilight zone structures (Banavar et al., 2002(a); 2002(b)) that a protein would adopt in its native state and be efficient in terms of expelling water from its interior, we will begin by considering a coarse-grained representation of the protein as a uniform tube of, say, unit radius (recall that 1 unit is approximately 2.7 Å). In the discretized case, one may consider the backbone of the protein with just the $C_\alpha$ atoms. The tube thickness at a given $C_\alpha$ location is obtained by considering all triplets of $C_\alpha$ atoms including the $C_\alpha$ at that location and selecting the smallest among all their radii. Recall
the notion of the private space associated with the thickness, which requires that no three body radius be smaller than 1 unit. In order to promote conformations which are efficient in squeezing the water from the interior of the structure, we could invoke an effective potential which promotes radii close to unity or the tube thickness.

Let us ask now what one obtains for the energetically favored conformations of a short chain made up of discrete particles with three-body potentials whose energy is lowest when the radius is 1 unit. The simplest starting point for obtaining a happy configuration is to choose the local radius of curvature (the radius associated with three contiguous particles) to be 1 unit. Winding the chain around a circle will lead to an overlap of the chain with itself and that is prohibitively expensive. So one would instead choose a helix with a local radius of curvature equal to 1 unit. But how would one select the pitch of the optimal helix? The pitch would be chosen so that the radius characterizing the three body interaction comprising a pair of particles from one turn of the helix and another from the next turn is again equal to unity, thus lowering the energy. This picks out a special pitch to radius ratio of the helix. Strikingly, the corresponding ratio in helices of proteins is within a few percent of this prediction (Maritan et al., 2000(b), Stasiak and Maddocks, 2000). Furthermore, the tube segments corresponding to neighboring turns of the helix are oriented parallel to each other and respect the anisotropy inherent in a tube-like description (see Figure 2). This helical conformation corresponds to the space-filling configuration of a garden hose in which the local radius of curvature equals the tube thickness (any smaller local radius is disallowed) and in which successive turns of the hose lie on top of each other with no intervening space.

How would one deal with a situation when the bulky side chains of amino acids do not allow a segment of a chain to be placed in a tight turn of such a small radius? The local radius of curvature would have to be larger than 2.7Å. In this situation, an alternative way to promote triplets having a radius of 1 unit (or around 2.7Å) is through non-local interactions. One possibility that would be cumbersome from a folding point of view is to have multiple
helices with a larger local radius of curvature winding around each other.

A more versatile way to obtain non-local interactions is by means of a sheet. First, a strand in an extended conformation would form to locally accommodate the larger radius of curvature enforced by the local steric incompatibility. In order to have as many triplets of the desired radius of 1 unit as possible, one would need interactions with a different part of the chain and the problem reduces to determining the optimal placement of two essentially independent parts of the chain. From symmetry considerations, one would expect the most favorable circumstances to occur when two such identical strands from different parts of the chain are in the vicinity of each other and when they both lie essentially in a plane. The three-body interaction encourages planarity by not only allowing for a harmonious fit of the strands but also providing room for the side chains perpendicular to the plane. Pauling and Corey (1951) had shown that two neighboring strands in a protein are replicas of each other or mirror images of each other (Richardson, 1997) (corresponding to parallel and antiparallel sheets respectively) in terms of the backbone atoms. They are located at an optimal distance from each other, which allows the formation of a supporting framework for the assembly of the strands based on hydrogen bonds between atoms in neighboring strands. A sheet is formed by a repetition of the same process (Figure 3). In this case as well, adjoining segments of the tube (neighboring strands) are parallel to each other. Strikingly, one can show analytically (Banavar et al., 2000(b)) that the familiar zig-zag pattern of the strands arise because of the discrete nature of the chain made up of the $C\alpha$ atoms of the backbone.

When one considers longer segments of the proteins, it is not energetically favorable to have just one helix or one sheet because distant regions would not necessarily have triplets characterized by the preferred radius. Thus there is a persistence length associated with a given secondary structure. In order to assemble the tertiary structure, which provides more energetically favored triplets from distinct secondary structure elements, one would need a mechanism for producing tight turns, which would entail having a small local radius
of curvature. This is facilitated by small amino acids like Glycine, which is often found in backward bends. In reality, therefore, a protein is strictly speaking not characterized by a uniform thickness.

The thickness associated with a tightly wound helical geometry would be expected to be slightly less than that associated with a hairpin or a sheet geometry. But are there other structures that might emerge which have many triplets having the optimal radius? A possibility that one might expect is a saddle structure instead of a hairpin. The easiest way to visualize a saddle is to start with a planar hairpin and bend it into a three dimensional object. The distinct advantage of doing this is the ability to create additional contacts at the cost of reducing the thickness somewhat. However, Nature does not seem to adopt this conformation in proteins because of the inability to form hydrogen bonds and provide the necessary scaffolding. Nevertheless, kissing hairpins are found in RNA secondary structures (see Figure 5).

VIII. CONSEQUENCES OF THE TUBE PICTURE

Within the hierarchical picture of folding (Baldwin and Rose, 1999), each short local segment of a sequence may be associated with a propensity or ability to either form very tight turns (as in backward bends), or the regular tight turn associated with a helix, or, indeed, a desirability to be in a strand conformation with a larger local radius of curvature. This local information then has to be put together in a global way in order to provide stability for the strands by forming a hairpin or a sheet structure. The complexity arises because a short segment of the sequence which is able to form a helix may instead choose to form a strand in order to stabilize a nearby segment which can only form a strand. These decisions are of course non-local in character and furthermore one has to ensure that all the turns can be made to assemble the tertiary structure and that all the hydrophobic residues are
shielded from the water in the folded state.

There are an astronomical number of sequences that one can construct, even for modest lengths. Why then are there so few sequences that are protein-like? More generally, for purposes of protein design, what is the selection principle in sequence space? It is likely that, for an overwhelming majority of sequences, different parts of the sequence would attempt to take on conformations corresponding to pieces of secondary structure that simply do not fit together to form one of the folds. This inherent frustration is absent for protein-like sequences and is responsible for selection in sequence space (Bryngelson and Wolynes, 1987). The rich and varied repertoire of amino acids has been used by Nature in evolution to design sequences that are able to fold rapidly and reproducibly to just their native states. There are many sequences that fold into a given structure because once a sequence has selected its native state structure, it is able to tolerate a significant degree of mutability except at certain key locations (Sander and Schneider, 1991; Kamtekar et al., 1993; West et al., 1999). Also, such a design could be carried out in order to create a folding funnel (Bryngelson et al., 1995; Dill and Chan, 1997) with a minimal amount of ruggedness in the energy landscape.

It is interesting to consider the ground state of many long tubes subject to compaction. Packing considerations suggest that the tubes become essentially straight and parallel to each other and are arranged (when viewed end on) in a triangular lattice, analogous to the Abrikosov flux lattice phase in superconductors (Tinkham, 1996). Returning to the case of a single tube, in the very long length limit, a similar phase would be expected with the additional constraint of the bending of the tube segments at the ends. One can show that, for a discrete chain, a planar placement of zig-zag strands is able to accommodate the largest thickness tube that can yet avail of the attraction. However, the thickness for this limiting case is too large to produce the three-dimensional ordering alluded to above. It would be interesting to consider how the ground state structure crosses over from the “flux-lattice” type phase to the familiar planar phase. Indeed, for thick tubes of moderate length, one may
expect to form a large sheet-like structure analogous to the cross-\(\beta\)-scaffold observed as a building block of amyloid fibrils (Dobson, 1999; 2002). Such fibrils have been implicated in a variety of human disorders including Alzheimer’s disease and spongiform encephalopathies such as Creutzfeldt-Jakob disease. The generic fibrillar forms of proteins can be regarded as the intrinsic ‘polymer’ structure of a polypeptide chain (Dobson, 2002) and is a direct confirmation of the tube picture presented here.

**IX. STUDIES OF SHORT TUBES**

We have carried out numerous analytic and computational studies (Banavar et al., 2002(a); 2002(b); 2002(d)) and have quantitatively confirmed the ideas presented here. As an illustration, Figure 5 shows the structures obtained in computer simulations of short tubes in the marginally compact phase (Banavar et al., 2002(b)). Helices and hairpins (sheets) are of course the well-known building blocks of protein structures (see Fig. 5 (A1) and (D1) for two examples from a protein and (A2), (D2) and (D3) for the tube structures in our simulations). It is interesting to note that some of the other marginally compact conformations bear a qualitative resemblance to secondary folds in biopolymers. Helices analogous to Fig. 5 (A3) with an irregular contact map occur, e.g., in the HMG protein NHP6a (Allain et al., 1999) with pdb code 1CG7. Fig. 5 (C1) shows the “kissing hairpins” (Chang and Tinoco, 1997) of RNA (pdb code 1KIS), each of which is a distorted and twisted hairpin structure while Fig. 5 (C2) is the corresponding tube conformation. Fig. 5 (B1) shows a helix of strands found experimentally in Zinc metalloprotease (Baumann et al., 1993) (pdb code: 1KAP), whereas Fig. 5 (B2) is the corresponding marginally compact conformation obtained in our calculations.

Specifically, these studies have shown that a thick short tube in the twilight zone assumes conformations corresponding to helices of the correct pitch to radius ratio and zig-zag hair-
pins and sheets. These building blocks of protein structures are the only ones that are effective in expelling the water from their interior. Furthermore, these structures are poised near a phase transition of a new kind of the corresponding infinite-sized system.

X. SUMMARY AND CONCLUSIONS

We have presented a simple unifying framework for understanding the common character of all proteins. Our analysis is based on just three ingredients – all proteins share a backbone, there are effective forces which promote the folding of a protein and the one and only new idea that a protein can be viewed as a tube. We have shown how one may write down a non-singular continuum description of a tube or a sheet of non-zero thickness. The recipe for doing this has the surprising feature that pairwise interactions potentials need to be discarded and replaced by appropriate many-body potentials.

We have considered a situation in which there is an attractive force, mimicking the hydrophobicity, between different parts of the tube. New physics arises from the interplay between two length scales: the thickness of the tube and the range of attractive interactions. Many of the known polymer phases are found when the tube is very thin compared to other length scales in the problem. However, when the two length scales become comparable, one obtains a novel phase of matter that is used by proteins for its native state structures. This new phase has many properties which explains the character of all small globular proteins, which do not depend on the specific amino acid sequence. These include the ability of the folded structure to expel water efficiently from its interior, the existence of a simple energy landscape with relatively few putative marginally compact native state structures, an explanation for many of the well-known characteristics of globular proteins such as helices and hairpins and sheets being the building blocks of protein structures, the cooperative folding of small proteins, generic formation of fibrils in tube-like polypeptide
chains and the acute sensitivity of protein structures to the right types of perturbations thus accounting for their flexibility and versatility.

Many strategies for attacking the protein folding problem have been put forward which employ a coarse-grained description (Banavar and Maritan, 2001). None of the currently used methods has been successful. Our results suggest that a deficiency of all these methods has been that the context provided by the local tube orientation is neglected while considering the interaction between coarse-grained units (Banavar et al., 2002d). The novel phase discussed here arises from the addition of anisotropy to the well-studied polymer problem just as one obtains rich liquid crystal behavior on studying anisotropic molecules. A mapping of the phase behavior of tubes on varying the nature of interactions, the thickness of the tube, the length of the tube and temperature might yield additional surprises.

It is important to stress that our results are not at odds with or meant as a substitute for the detailed and beautiful work involving the laws of quantum mechanics and biochemistry. The virtue of our approach is that it predicts a novel phase with selected types of structures and the attendant advantages. It is then necessary to complement this information with the principles of quantum chemistry to assess whether a given biomolecule would fit one of these structures. We do not invoke hydrogen bonds as Pauling did in his prediction of protein secondary motifs (Pauling, Corey and Branson, 1951; Pauling and Corey, 1951) and indeed not all the structures in the marginally compact phase are compatible with hydrogen bond placement. What is remarkable, however, is that the lengths of the covalent and hydrogen bonds and the rules of quantum chemistry conspire to provide a perfect fit to the basic structures in this novel phase. One cannot but be amazed at how the evolutionary forces of Nature have shaped the molecules of life ranging from the DNA molecule, which carries the genetic code and is efficiently copied, to proteins, the work horses of life, whose functionality follows from their form, which, in turn, is a novel phase of matter. Protein folds seem to be immutable – they are not subject to Darwinian evolution and are determined
from geometrical considerations, as espoused by Plato (Denton and Marshall, 2001). It is as if evolution acts in the theater of life to shape sequences and functionalities, but does so within the fixed backdrop of these Platonic folds.

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REFERENCES

Allain, F. H. T., M. Yen, J. E. Masse, P. Schultze, T. Dieckmann, R. C. Johnson, and J. Feigon, 1999, “Solution structure of the HMG protein NHP6A and its interaction with DNA reveals the structural determinants for non sequence specific binding,” *Embo J.* **18**, 2563-2579.

Anfinsen, C., 1973, “Principles that govern the folding of protein chains,” *Science* **181**, 223-230.

Baker, D., 2000, “A surprising simplicity to protein folding,” *Nature* **405**, 39-42.

Baldwin, R. L. and G. D. Rose, 1999, “Is protein folding hierarchic? I. Local structure and peptide folding,” *Trends in Biochemical Sciences* **24**, 26-33.

Banavar, J. R. and A. Maritan, 2001, “Computational approach to the protein folding problem,” *Proteins* **42**, 433-435.

Banavar, J. R., A. Maritan, C. Micheletti, and A. Trovato, 2002a, “Geometry and physics of proteins,” *Proteins* **47**, 315-322.

Banavar, J. R., A. Flammini, D. Marenduzzo, A. Maritan, and A. Trovato, 2002b, “Geometry of compact tubes and protein structures,” *ComPlexUs* **1**, XX-YY.

Banavar, J. R., O. Gonzalez, J. H. Maddocks, and A. Maritan, 2002c, “Self-interactions of strands and sheets,” *J. Stat. Phys.* **XX**, YY-ZZ.

Banavar, J. R., A. Maritan and F. Seno, 2002d, “Anisotropic Effective Interactions in a Coarse-Grained Tube Picture of Proteins”, *Proteins** **XX**, YY-ZZ.

Baumann, U., S. Wu, K. M. Flaherty, and D. B. Mckay, 1993, “Three-dimensional structure of the alkaline protease of pseudomonas aeruginosa: a two-domain protein with a calcium binding parallel beta roll motif,” *Embo J.* **12**, 3357-3364.

Bernal, J. D., 1939, “Structure of proteins,” *Nature* **143**, 663-667.

Branden, C. and Tooze, J., 1999, *Introduction to protein structure* 2nd Edition (Garland Pub., New York).

Bryngelson, J. D. and P. G. Wolynes, 1987, “Spin glasses and the statistical-mechanics of protein folding,” *Proc. Nat. Acad. Sci. USA* **84**, 7524-7528.

Bryngelson, J. D., J. N. Onuchic, N. D. Socci, and P. G. Wolynes, 1995, “Funnels, pathways and the energy landscape of protein folding: A synthesis,” *Proteins* **21**, 167-195.

Chaikin, P. M. and T. C. Lubensky, 1995, *Principles of condensed matter physics* (Cambridge University Press, Cambridge).

Chang, K. Y. and I. Tinoco, 1997, “The Structure of an RNA “kissing” hairpin complex of the HIV tar hairpin loop and its complement,” *J. Mol. Biol.* **269**, 52-66.

Chothia, C., 1984, “Principles that determine the structure of proteins,” *Annu. Rev. Biochem.* **53**, 537-572.

Chothia, C., 1992, “One thousand families for the molecular biologist,” *Nature* **357**, 543-544.

Creighton, T. E., 1993, *Proteins, structure and molecular properties* 2nd Edition (W. H. Freeman, New York).

Denton, M. and C. Marshall, 2001, “Laws of form revisited,” *Nature* **410**, 417-417.
Dill, K. A. and H. S. Chan, 1997, “From Levinthal to pathways to funnels,” Nature Structural Biology 4, 10-19.

Dobson, C. M., 1999, “Protein misfolding, evolution and disease,” Trends in Biochem. Sci., 24, 329-332.

Dobson, C. M., 2002, “Protein-misfolding diseases: Getting out of shape,” Nature 418, 729-730.

Doi, M. and S. F. Edwards, 1993, The theory of polymer dynamics (Clarendon Press, New York).

Fersht, A. R., 1998, Structure and mechanism in protein science: A guide to enzyme catalysis and protein folding (W.H. Freeman and Company, New York).

Flory, P. J., 1969, Statistical mechanics of chain molecules p. 301, (Wiley, New York).

Gonzalez, O. and J. H. Maddocks, 1999, “Global curvature, thickness and the ideal shapes of knots,” Proc. Natl. Acad. Sci. USA 96, 4769-4773.

Holm, L. and C. Sander, 1997, “An evolutionary treasure: unification of a broad set of amidohydrolases related to urease,” Proteins 28, 72-82.

Hunt, N. G., L. M. Gregoret, and F. E. Cohen, 1994, “The origins of protein secondary structure,” J. Mol. Biol. 241, 214-225.

Jacobs, D. J., A. J. Rader, L. A. Kuhn, and M. F. Thorpe, 2001, “Protein flexibility predictions using graph theory,” Proteins 44, 150-165.

Kamtekar, S., J. M. Schiffer, H. Y. Xiong, J. M. Babik, M. H. Hecht, 1993, “Protein design by binary patterning of polar and non-polar amino acids,” Science 262, 1680-1685.

Katritch, V., J. Bednar, D. Michoud, R. G. Scharein, J. Dubochet, and A. Stasiak, 1996, “Geometry and physics of knots,” Nature 384, 142-145.

Lesk, A. M. and C. Chothia, 1980, “How different amino acid sequences determine similar protein structures: the structure and evolutionary dynamics of globins,” J. Mol. Biol. 136, 225-270.

Micheletti, C., J. R. Banavar, A. Maritan, and F. Seno, 1999, “Protein structures and optimal folding from a geometrical variational principle,” Phys. Rev. Lett. 82, 3372-3375.

Pauling, L. and R. B. Corey, 1951, “Conformations of polypeptide chains with favored orientations around single bonds: two new pleated sheets,” Proc. Natl. Acad. Sci. USA 37, 729-740.

Pauling, L., R. B. Corey, and H. R. Branson, 1951, “The structure of proteins: two hydrogen-bonded helical conformations of the polypeptide chain,” Proc. Natl. Acad. Sci. USA 37, 205-211.

Ramachandran, G. N. and V. Sasisekharan, 1968, “Conformations of polypeptides and proteins,” Adv. Protein Chem. 23, 283-438.

Richardson, J. S., 1997, “β-sheet topology and the relatedness of proteins,” Nature 268,
495-500.

Rose, G. D., 1996, “No assembly required,” Sciences - New York 36, 26-31.

Sander, C. and R. Schneider, 1991, “Database of homology-derived protein structures and the structural meaning of sequence alignment,” Proteins 9, 56-68.

Stanley, H. E., 1999, “Scaling, universality and renormalization: three pillars of modern critical phenomena,” Rev. Mod. Phys. 71, S358-S366.

Stasiak, A. and J. H. Maddocks, 2000, “Mathematics - Best packing in proteins and DNA,” Nature 406, 251-253.

Tinkham, M., 1996, Introduction to superconductivity (McGraw-Hill, New York).

Yee, D. P., H. S. Chan, T. F. Havel, and K. A. Dill, 1994, “Does compactness induce secondary structure in proteins?,” J. Mol. Biol. 241, 557-573.

Watson, J. D. and F. H. C. Crick, 1953, “A structure for deoxyribose nucleic acid,” Nature 171, 737.

West M. W, W. X. Wang, J. Patterson, et al., 1999, “De novo amyloid proteins from designed combinatorial libraries,” Proc. Nat. Acad. Sci. USA 96, 11211-11216.
FIGURE CAPTIONS

Figure 1:
Native state structure of the B1 domain (Protein data bank code: 1GB1) of Protein G, a small protein produced by several Streptococcal species which binds very tightly to Immunoglobulin. The domain shown has a length of 56 amino acids. The structure contains an efficiently packed hydrophobic core between a 4-stranded β-sheet (shown in blue) and a 4 turn α-helix (shown in red). Strikingly, all protein structures have helices, hairpins and sheets as their building blocks.

Figure 2:
Space filling helix.
The geometry of this helix nicely illustrates the idea behind the 3-body potential. Consider a tube in a compact helical conformation. The smallest value that the local radius of curvature of the helix can adopt equals the tube thickness. Note that if the local radius of curvature were any smaller than the tube thickness, the tube would self-intersect and such configurations are not allowed. Physically, a space-filling helix is obtained when successive turns of the tube lie on top of each other. This translates into the observation that the non-local radius associated with three points, of which two are close together and the third is alongside them in a neighboring turn, is also equal to the tube thickness. Again, a radius smaller than this value would lead to an intersection and is forbidden. The pitch to radius ratio of this helix is within 3% of the corresponding value for α-helices in globular proteins.

Figure 3:
Anti-parallel β-sheet, made of 4 strands, predicted by Linus Pauling. The local radius of curvature of strands is greater than that of helices, but the non-local three-body radius associated with two neighboring Cα atoms in a strand and a nearest Cα atom in an adjoining strand is close to the local radius of curvature associated with a helix.
Figure 4:
Sketch of the maximal number of contacts that a short, compact tube can make as a function of $X$, the dimensionless ratio of the tube thickness to the range of the attractive interaction. When $X$ is large compared to 1, one obtains a swollen phase. At the other extreme, when $X \ll 1$, one finds a highly degenerate compact phase. The twilight zone between these two phases occurs in the vicinity of $X \sim 1$ and is characterized by marginally compact structures. Typical tube conformations in each of the phases are shown in the figure.

Figure 5:
Building blocks of biomolecules and ground state structures associated with the marginally compact phase of a short tube. The top row shows some of the building blocks of biomolecules, while the second row depicts the corresponding structures obtained for a tube in the twilight zone. (A1) is an $\alpha$-helix of a naturally occurring protein, while (A2) and (A3) are the helices obtained in our calculations. (A2) has a regular contact map whereas (A3) is a distorted helix in which the distance between successive atoms along the helical axis is not constant, but has period 2. (B1) is a helix of strands in the alkaline protease of pseudomonas aeruginosa, whereas (B2) shows the corresponding structure obtained in our computer simulations. (C1) shows the “kissing” hairpins of RNA and (C2) the corresponding conformation obtained in our simulations. Finally (D1) and (D2) are two instances of quasi-planar hairpins. The first structure is from the same protein as before (the alkaline protease of pseudomonas aeruginosa) while the second is a typical conformation found in our simulations. The sheet-like structure (D3) is obtained for a longer tube. The tube thickness increases from left to right while the range of interactions is held fixed. For more details, see Banavar et al. (2002b).
