Assessment of Organophosphate Pesticides Exposure in Men with Idiopathic Abnormal Semen Analysis: A Cross-Sectional Pilot Study

Induja Manikandan, M.D.¹, Sushmita Bora, M.Sc.¹, Prashant Shankarrao Adole, M.D.®, Chitra Thyagaraju, M.D.², Rajesh Nachiappa Ganesh, M.D., D.N.B.³

1. Department of Biochemistry, Jawaharlal Institute of Postgraduate Medical Education and Research, Pondicherry, India
2. Department of Obstetrics and Gynecology, Jawaharlal Institute of Postgraduate Medical Education and Research, Pondicherry, India
3. Department of Pathology, Jawaharlal Institute of Postgraduate Medical Education and Research, Pondicherry, India

Abstract

Background: Because of the widespread use of organophosphate (OP) pesticides in agriculture, they are major environmental contaminants in developing countries. OP pesticides decrease sperm concentration and affect its quality, viability, and motility. Studies have demonstrated the association between abnormal semen analysis and OP pesticides exposure among the high-risk population. As there is limited data on the percentage of OP pesticides exposure, the study aimed to determine the OP pesticides exposure in Southern Indian men with idiopathic abnormal semen analysis and find the possible source of their OP pesticides exposure.

Materials and Methods: In this cross-sectional pilot study, fifty men with idiopathic abnormal semen analysis as cases and fifty men with normal semen analysis as controls were recruited. Detailed history was taken and general and systemic examinations were carried out. OP pesticides exposure was determined by assessment of pseudocholinesterase and acetylcholinesterase levels and urinary OP pesticides metabolites dialkyl phosphate (DAP) consisting of dimethyl phosphate (DMP), diethyl thiophosphate (DETP), and diethyl dithiophosphate (DEDTP).

Results: Cases had statistically significantly lower levels of pseudocholinesterase (5792.07 ± 1969.89 vs. 10267.01 ± 3258.58 IU/L) (P=0.006) and acetylcholinesterase [102.90 (45.88-262.74) vs. 570.31 (200.24-975.30) IU/L] (P=0.001) as compared to controls. Cases had a statistically significantly higher percentage of urinary DAP positivity as compared to controls (80 vs. 38%, P<0.0001). Hence, cases had a significantly higher percentage of OP pesticides exposure as compared to controls (20 vs. 4%, P=0.015). OP-exposed cases had significantly higher urinary DETP and DEDTP levels as compared to OP non-exposed cases. Also, urinary DETP and DEDTP levels were significantly negatively associated with sperm concentration, motility, and normal morphology among OP-exposed cases.

Conclusion: Southern Indian men with idiopathic abnormal semen analysis had a significantly higher percentage of OP pesticides exposure as compared to men with a normal semen analysis.

Keywords: Acetylcholinesterase, Male Infertility, Pseudocholinesterase, Organophosphate Pesticides

Citation: Manikandan I, Bora S, Adole PS, Thyagaraju C, Nachiappa Ganesh R. Assessment of organophosphate pesticides exposure in men with idiopathic abnormal semen analysis: a cross-sectional pilot study. Int J Fertil Steril. 2021; 15(3): 219-225. doi: 10.22074/IJFS.2020.134650. This open-access article has been published under the terms of the Creative Commons Attribution Non-Commercial 3.0 (CC BY-NC 3.0).

Introduction

Infertility as one of the major public health problems is defined as “a failure in achieving a clinical pregnancy after 12 or more months of regular unprotected sexual intercourse” as per the World Health Organization (WHO) (1). According to the WHO, 45-52.6 million married couples were suffering from infertility worldwide in 2010 (2). The prevalence of infertility among Indians was ranging from 3.9 to 16.8% as estimated by the WHO (3). As per the report of a multicentric study by the WHO, 20% of cases of infertility were due to male factors, 38% due to female factors, 27% to both partners, and 15% cases of infertility were idiopathic. In India, nearly 50% of cases of infertility were due to the reproduction anomaly or disorders in males and in 25% of cases, no detectable causes were found and it was considered idiopathic (4). Male infertility is rising in society and its causes are multifactorial. Many studies have shown a declining trend in the semen quality and sperm count among the population (5, 6). A study conducted in the Indian population over the past 37 years has shown a decline in sperm count and motility, and altered sperm morphology with time (7). No
clear cause has been found for the decline of semen quality, but it might be due to environmental, dietary, or other unknown causes (5).

Organophosphate (OP) pesticides are synthetic chemicals used worldwide for controlling domestic and agricultural pests. Pesticides used to control pests and weed on crops are registered under the central Malaria board and registration committee (CIBRC), which comes under the Ministry of Agriculture and Farmer welfare. Section-3 of the Insecticides Act, 1968 has registered around 30 OP pesticides, which are in use in India (8). OP pesticides like monocrotophos, phorate, quinalphos, malathion, chlorpyrifos, diazinon, methyl parathion, ethion, and so on, used extensively in India, were already banned or severely restricted in the USA and Europe. The OP pesticides are associated with severe toxicity, contributing to more than 80% of pesticides-related hospitalization in India (9). OP pesticides cause phosphorylation of acetylcholinesterase resulting in acetylcholine accumulation in synapses. OP pesticides affect reproduction function by reducing acetylcholinesterase activity in the brain, and influencing gonads. OP pesticides like parathion and methyl parathion have a structure similar to hormones like estrogen, thus altering genes expression by interacting with hormone receptors. OP pesticides alter the hypothalamic-pituitary (HPO), pituitary-thyroid, and pituitary-adrenal axes and serum prolactin levels. OP pesticides affect spermatogenesis by damaging the Sertoli and Leydig cells and increasing their apoptosis (10). A toxicological study demonstrated that OP pesticides cause low sperm concentration by affecting germ cell proliferation and damaging the seminiferous epithelium (11). Also, OP pesticides disturb sperm motility by disturbing its tail assembly proteins or ATP synthesis (12). Concerning the association between semen parameters and OP pesticides exposure among agricultural workers, pesticide sprayers, and workers in pesticides manufacturing industries, several studies concluded that there was a decrease in sperm concentration, motility, viability, and normal morphology due to OP pesticides exposure (13-18). There has been contamination of agricultural soil, sediment, and water by various OP pesticides throughout India (9). Hence, subtle OP pesticides exposure is occurring among human beings through food, water, air, tainted breast milk, playing in the field, or skin contact.

Most of the studies were done on high-risk populations to find out potential associations between OP pesticides exposure and alteration in semen parameters. However, there is limited data available in the literature to say that environmental OP pesticides exposure associates with abnormal semen parameters among the general population. Therefore, the present study aimed to assess the environmental OP pesticides exposure among Southern Indian men from Pondicherry and surrounding districts of Tamil Nadu, like Tindivanam, Villianur, Chennai, and Villupuram with idiopathic abnormal semen analysis by measuring pseudocholinesterase and acetylcholinesterase levels and urinary OP pesticides metabolites. The objectives of the study were to compare environmental OP pesticides exposure between men with and without idiopathic abnormal semen analysis and to determine possible sources of OP pesticides exposure by comparing percentages of farmers, rural population, smokers, undergraduates, lower socioeconomic status, vegetarians, people using underground water source, and alcoholics.

Materials and Methods

Study design and population

This cross-sectional pilot study was conducted in the Jawaharlal Institute of Postgraduate Medical Education and Research (JIPMER) hospital, Puducherry, 605 006 from January 2018 to July 2019 after obtaining the approvals from Institute Research Council and Institute Human Ethics Committee (JIP/IEC/2017/0351 dated November 27, 2017). All Southern Indian men, 25 to 45 years old, from Pondicherry and surrounding districts of Tamil Nadu, like Tindivanam, Villianur, Chennai, and Villupuram, who was attending the JIPMER Infertility Clinic, Pondicherry for the inability of their spouse to conceive after 1 year of unprotected sexual intercourse, were recruited. Written informed consent (both in English and Tamil) after explaining the purpose and the procedure of the study, was obtained from all the participants. After obtaining the consent, detailed history of the patient was taken including his occupation, location, source of water, history of any medication, or surgery, and other demographic factors. General and systemic examinations were carried out and the scheduled date for semen analysis was given. On the scheduled day, a semen sample, 10 ml of urine, and 5 ml of blood were collected under sterile conditions. Fifty men who had abnormal semen analysis (sperm count ≤15 million/ml, sperm motility ≤40% and sperm morphology ≤4% normal) as per WHO criteria 2010 (4) with no identifiable pathology, were recruited as cases. Cases with underlying pathology such as varicocele, history of diabetes, cardiovascular or thyroid disorders, tuberculosis, testicular carcinoma, obstruction or congenital bilateral absence of vas deferens, or use of lipid-lowering drugs, were excluded. Fifty men with normal semen analysis were recruited as controls. So, it was a pilot study with 50 cases and 50 controls, which was accepted by the institute research council.

Criteria for analysis of anthropometry, alcohol use, smoking, and socioeconomic status

The height, weight, body mass index (BMI), and waist circumference were measured by the same observer. The nearest half-kilogram for body weight and half-centimeter for height were recorded. The waist circumference was determined by measuring the shortest point below the lowermost rib cage margin and the iliac crest and was recorded to the nearest half-centimeter. BMI was calculated as weight (kg) divided by the square of height (m). Being alcoholic was defined by the consumption of at least two drinks per day. A standard drink was equal to either...
Ten milliliters of urine were collected and stored at -80°C until further analysis. All urine samples were thawed and mixed by a vortex. Cleaning of urine sample and derivatization of alkyl phosphate were done as per Hemakanthi De Alwis et al. (24). A Trace GC Ultra equipped with A/1 3000 Auto-Injector (Rodano, Italy) and ITQ 900 mass spectrometer from Thermo Scientific (Austin, USA) was used for analysis with a constant flow rate of 1.2 ml/minutes and Helium as a carrier gas. One microliter of a sample from the low volume insert was injected in splitless mode onto a Thermo Electron Corporation (Rodana, Italy) TR-5MS ([5%-phenyl]-polysilphenylene-siloxane) TRACE GC capillary column (30 m, 0.25 mm, 0.25 μm) using the autosampler. The GCMS protocol followed was as per Hemakanthi De Alwis et al. (24). All the standards for dimethyl phosphate (DMP), diethyl thiophosphate (DETP), diethyl dithiophosphate (DEDTP), Sulfotep, an internal standard and derivatization agent 2,4,5,6-Pentafluorobenzyl bromide were purchased from Sigma-Aldrich with a purity of ≥90%. The mass spectra of the pentafluorobenzyl esters of DMP, DETP, DEDTP, and Sulfotep was determined. The analysis was done on selective ion monitoring (SIM) mode. The retention time (RT), linearity, the limit of detection (LOD), and limit of quantification (LOQ) for DMP, DETP, DEDTP, and Sulfotep were detected. The data obtained were transferred to X-Calibur files and manually evaluated. The peaks of the samples processed were recognized using the RTs and confirmed by comparison with the analyte/Sulfotep ratio for the two ions of the analyte. Results were reported utilizing creatinine adjustment.

**Organophosphate pesticides exposure criteria**

Both the presence of DAP in urine and inhibition of pseudocholinesterase were mandatory for the patients to be labeled as OP pesticides exposed. Patients with DMP, DETP, and/or DEDTP detected in urine above the LOD labeled as DAP positive. Proudfoot formula was used as a basis for the determination of inhibition of pseudocholinesterase. We considered 4621-11500 IU/L as normal level (≥50%), 2311-4620 IU/L as mild inhibition (20-50%), 460-2310 IU/L as moderate inhibition (10-20%) and less than 460 IU/L as severe inhibition (less than 10%) (14).

**Statistical analysis**

The normality of data was assessed by the Kolmogorov-Smirnov test. The distribution of categorical data such as socio-demographic status, occupation, being vegetarian, being smoker, being alcoholic, and people using underground water, are expressed as percentages. The continuous data such as semen parameters, pseudocholinesterase and acetylcholinesterase levels, and OP metabolites in urine are expressed as mean with standard deviation or median (interquartile range). Creatinine was analyzed in urine and all OP metabolites values were adjusted for creatinine. All OP metabolites concentrations were log-transformed for statistical analysis. Descriptive statistics for OP metabolites among exposed and non-exposed included the percent above the LOD, mean and standard deviation, geometric mean and standard deviation, ranges, and calculation of the 25th, 75th, and 90th percentile. OP metabolites concentration below the LOD was assigned a value equal to the LOD/√2 (25). Binary logistic regression was done to estimate relative odds of urinary OP metabolites among exposed and non-
exposed groups after adjustment to age, the number of married years, height, weight, waist circumference, percentage of undergraduates, lower socioeconomic status, vegetarians, primary infertility and alcoholics. Spearman’s or Pearson’s correlation was assessed between semen parameters and urinary OP metabolites in the OP-exposed group. Normally distributed variables were compared using the student’s t test. Non-parametric parameters were compared by the Kruskal-Wallis H test. Statistical analyses were done using SPSS 10 software at a significance level of 5% and P<0.05 was considered significant.

**Results**

General characteristics were compared between cases and controls (Table 1). Cases had a significantly higher percentage of farmers as compared to controls (44 vs. 18%, P=0.009). Similarly, cases had a significantly higher percentage of rural population as compared to controls (60 vs. 38%, P=0.045). Contradictorily, a high percentage of smokers was found among controls as compared to cases (28 vs. 10%, P=0.022). However, no significant difference was found between cases and controls in age, the number of married years, height, weight, waist circumference, percentage of undergraduates, lower socioeconomic status, being vegetarian, using underground water, primary infertility, and being alcoholic. Pseudocholinesterase levels (5792.07 ± 1969.89 vs. 10267.01 ± 3258.58 IU/L, P=0.006) and acetylcholinesterase levels [102.90 (45.88-262.74) vs. 570.31 (200.24-975.30) IU/L, P=0.001] were statistically significantly lower among cases as compared to controls.

Forty out of 50 cases had urinary DAP positivity. Pseudocholinesterase (4917.65 ± 900.54 vs. 5496.97 ± 1515.90 IU/L, P=0.007) and acetylcholinesterase [88.00 (45.62-249.47) vs. 197.64 (54.57-262.74) IU/L, P=0.001] levels were significantly lower among 40 cases with urinary DAP positivity as compared to 10 cases with urinary DAP negativity. Out of the 40 cases with urinary DAP positivity, 10 (25%) cases had mild inhibition (4417 ± 200 IU/L) and 30 (75%) cases had normal pseudocholinesterase levels. Nineteen out of 50 controls had urinary DAP positivity. Out of the 19 controls with urinary DAP positivity, 2 (10.6%) men had mild inhibition and 17 (89.4%) men had normal pseudocholinesterase levels. However, all controls with DAP negativity had normal pseudocholinesterase levels.

Ten out of 50 cases had both inhibitions of pseudocholinesterase and urinary DAP positivity, hence they were labeled as OP pesticides-exposed. Two out of 50 controls had both inhibitions of pseudocholinesterase and urinary DAP positivity, hence they were labeled as OP pesticides-exposed. Cases had a significantly higher percentage of OP pesticides exposure in comparison with controls (20 vs. 4%, P=0.015). Also, cases with OP pesticides exposure had significantly higher urinary DETP and DEDTP levels as compared to cases without OP pesticides exposure (Table 2). Binary logistics regression showed that OP-exposed cases had significantly higher urinary DETP (OR=1.12, 95% CI=1.01-1.26), DEDTP (OR=1.27, 95% CI=1.02-1.45) and DAP (OR=1.33, 95% CI=1.13-1.66) levels as compared to non-exposed cases after adjustment to age, the number of married years, height, weight, waist circumference, percentage of undergraduates, lower socioeconomic status, vegetarians, people using underground water, primary infertility and alcoholics (Table 3). Correlation analysis among OP-exposed cases showed that urinary DAP levels were significantly negatively associated with sperm concentration (P=0.001, r=-0.634), motility (P=0.001, r=-0.523) and normal morphology (P=0.001, r=-0.721).

### Table 1: Comparison of general characteristics between cases and controls

| Parameters                        | Men with idiopathic abnormal semen analysis, cases (n=50) | Men with normal semen analysis, controls (n=50) | P value* |
|-----------------------------------|----------------------------------------------------------|-------------------------------------------------|----------|
| Age (Y)                           | 34.94 ± 5.23                                            | 34.20 ± 6.61                                    | 0.479    |
| Duration of marriage (Y)          | 5.49 ± 3.74                                             | 5.04 ± 2.53                                     | 0.889    |
| Height (cm)                       | 169.64 ± 12.21                                          | 168.76 ± 4.74                                   | 0.636    |
| BMI (Kg/m²)                       | 24.98 ± 4.44                                            | 24.65 ± 3.79                                    | 0.692    |
| Waist circumference (cm)          | 92.28 ± 16.03                                           | 86.90 ± 17.44                                   | 0.112    |
| Undergraduates (%)                | 41 (82)                                                 | 37 (74)                                         | 0.334    |
| Farmers (%)                       | 22 (44)                                                 | 9 (18)                                          | 0.009    |
| Lower socioeconomic status (%)    | 17 (34)                                                 | 8 (16)                                          | 0.068    |
| Rural population (%)              | 30 (60)                                                 | 19 (38)                                         | 0.045    |
| Vegetarians (%)                   | 7 (14)                                                  | 7 (14)                                          | 1.000    |
| Underground water source (%)      | 46 (92)                                                 | 42 (84)                                         | 0.218    |
| Primary infertility (%)           | 48 (96)                                                 | 45 (90)                                         | 0.436    |
| Smokers (%)                       | 5 (10)                                                  | 14 (28)                                         | 0.022    |
| Alcoholics (%)                    | 13 (26)                                                 | 12 (24)                                         | 0.579    |
| Semen volume (ml)                 | 2.0 (1.5-2.5)                                           | 2.0 (1.5-2.5)                                   | 0.939    |
| Sperm concentration (million/ml)  | 11.20 (3.00-46.25)                                      | 85.90 (65.45-121.90)                            | 0.001    |
| Sperm number (million/jaculate)   | 26.95 (10.25-112.50)                                    | 127.75 (93.75-219.68)                           | 0.001    |
| Sperm motility (%)                | 8 (2-12.80)                                             | 53 (47-61)                                      | 0.001    |
| Sperm morphology (%)              | 3 (2-4)                                                 | 19.5 (17-23)                                    | 0.001    |
| Pseudocholinesterase (IU/L)       | 5792.07 ± 1969.89                                       | 10267.01 ± 3258.58                              | 0.006    |
| Acetylcholinesterase (IU/L)       | 102.90 (45.88-262.74)                                    | 570.31 (200.24-975.30)                          | 0.001    |
| DAP positivity (%)                | 40 (80)                                                 | 19 (38)                                         | <0.0001  |

Data are presented as mean ± SD, median (interquartile range) or percentage (%). BMI: Body mass index, DAP: Dialkyl phosphate, and *: Independent sample t test/ Kruskal-Wallis H test.
To find out the possible source of OP pesticides exposure, the general characteristics between OP-exposed cases and non-exposed cases were compared. Percentages of farmers and residing in a rural area were significantly higher in OP-exposed cases as compared to non-exposed cases. However, there was no significant difference in age, BMI, or waist circumference as well as percentages of men with undergraduate education, lower socioeconomic status, being vegetarian, using underground water, being smokers, and being alcoholics among OP-exposed cases as compared to non-exposed cases.

### Discussion

Our study reports that Southern Indian men with idiopathic abnormal semen analysis had a significantly higher percentage of OP pesticides exposure as compared to men with a normal semen analysis. Also, we found a significant correlation between urinary OP metabolites and semen parameters among OP-exposed cases.

As there is rampant use of OP pesticides in agriculture, their residues can be found in cooked meals, water, wine, fruit juices, refreshments, and so on. Also, washing and peeling cannot remove the OP residues completely (26, 27). Chronic, low-dose exposure to OP pesticides was found to be associated with neurodevelopmental problems in children, Parkinson’s disease, metabolic syndrome, obesity, diabetes, reduced semen quality, reduced gestational age, reduced birth weight, and so on (28, 29). OP pesticides were found to affect the sperm quality directly or indirectly resulting in infertility and reproduction problems in the agricultural workers. OP pesticides act as endocrine-disrupting chemicals, alter the HPO axis, and impair spermatogenesis by damaging the Sertoli and Leydig cells (30). The general population is exposed to OP pesticides mainly through diet, inhalation of air, dermal absorption, and unintentional ingestion (31, 32).

Comparing general characteristics, we noticed that men with idiopathic abnormal semen analysis were mostly farmers and from the rural area as compared to men with a normal semen analysis. Our observations were consistent with those reported by Miranda-Contreras et al. (14) who concluded that sperm count, motility, and membrane integrity among Venezuelan farmworkers were affected by occupational pesticides exposure. Also, Katole and Saoji (33) have reported a lower prevalence of primary infertility among urban populations. Dutta and Bahadur (34) showed that pseudocholinesterase and acetylcholinesterase levels were decreased among occupationally-exposed tea garden workers of the Northern part of West Bengal, India, similar to our observations. Education has an important role in maintaining personal hygiene, prevention of sexually transmitted disease, and understanding the effect of alcohol and smoking on sperm count. Our study has not found any difference between cases and controls in education as the two groups have the same percentage of educated participants.

Many studies have used the measurement of urinary DAP as a tool for determining OP pesticides exposure (13-18). As Yucra et al. (16) showed that occupation exposure of OP pesticides cannot be decided solely by OP metabolites measurement in urine, we have included both the determination of DAP in urine and measurement of pseudocholinesterase levels for labeling patient as OP-exposed. Hence, we may conclude that men with idiopathic abnormal semen analysis had high baseline exposure to OP pesticides. Li and Kannan (35) established the baseline levels of exposure to OP and pyrethroid pesticides among the population of several Asian countries. They concluded that India has the second-highest sum concentration of 11 pesticides in urine, next to Vietnam. Also, they found that daily intake of chlorpyrifos and parathion was high among the Indian population as compared to the population from other Asian countries. We got higher urinary levels of

### Table 2: OP pesticides metabolites levels between OP exposed and non-exposed cases

| OP metabolites (selected ions) | LOD*/LOQ | Mean (SD) | GM (GSD) | Median | 25th per | 75th per | 90th per | Range |
|-------------------------------|----------|-----------|----------|--------|----------|----------|----------|-------|
| DMP (306, 307)                | 10/33.4  | 15.50 (11.43) | 14.41 (89.66) | 15.65 | 8.32 | 18.87 | 22.34 | 7.09-55.98 |
| OP exposed                    | 14.52 (6.87) | 13.67 (76.67) | 14.42 | 7.98 | 9.78 | 12.98 | 7.09-39.17 |
| DEDTP (274, 350)             | 0.14/0.457 | 46.78 (39.78) | 32.45 (7.98) | 35.26 | 11.24 | 55.67 | 88.65 | 0.14-146.67 |
| OP exposed                    | 26.56 (22.34) | 11.46 (6.54) | 15.50 | 8.45 | 38.87 | 67.54 | 0.14-80.67 |
| DEDTP (366, 185)             | 3.06/10.20 | 55.23 (46.32) | 42.87 (9.45) | 46.11 | 14.34 | 55.98 | 75.98 | 3.06-97.56 |
| OP exposed                    | 35.34 (30.21) | 19.45 (8.67) | 27.02 | 10.09 | 54.56 | 66.98 | 3.06-76.07 |

*OP: Organophosphate, SD: Standard deviation, GM: Geometric mean, GSD: Geometric standard deviation, DMP: Dimethyl phosphate, DETP: Diethyl thiophosphate, DEDTP: Diethyl dithiophosphate, LOD: Limit of detection, LOQ: Limit of quantification, and ; Samples below the LOD were defined as LOD/2.

### Table 3: Relative odd of OP metabolites after adjustment among OP exposed and non-exposed cases

| OP metabolites | OR | CI | P value |
|----------------|----|----|---------|
| DMP            | 0.65 | 0.23-1.43 | 0.363 |
| DETP           | 1.12 | 1.01-1.26 | 0.004 |
| DEDTP          | 1.27 | 1.02-1.45 | 0.004 |
| DAP            | 1.33 | 1.13-1.66 | 0.001 |

*OP: Organophosphate, DAP: Dialkyl phosphate, DMP: Dimethyl phosphate, DETP: Diethyl thiophosphate, DEDTP: Diethyl dithiophosphate, OR: Odd ratio, and CI: Confidence interval.*
DEDTP and DETP in cases as compared to controls and these levels were significantly negatively associated with sperm concentrations, motility, and normal morphology. Perry et al. (10) concluded that men with lower semen quality had higher urinary DMP levels as compared to men with normal semen quality. Muñoz-Quezada et al. (36) concluded that urinary DAP levels were high in Chilean school children due to the presence of chlorpyrifos and phosmet residues in fruits.

There is a rising trend of male infertility among the population and for most of them, no detectable cause has been found. Hence, it has become the burning question and need of the hour to address what are the possible reasons for the decline in semen parameters? Because there is extensive use of OP pesticides in agriculture, its contamination in the food chain and its effect on sperm parameters, can sustain and a low dose of OP pesticides exposure be one of the causes for the decline of semen parameters among the Southern Indian population? In our study, we found that men with abnormal semen analysis had significantly higher OP pesticides exposure as compared to men with a normal semen analysis. OP-exposed men were farmers and from the rural population where they might be daily exposed to OP pesticides through food, water, and air, affecting their sperm parameters.

There were certain limitations in this study: i. We estimated acetylcholinesterase activity in serum instead of RBC. ii. There were six DAPs: DMP, DMT, DMDTP, DEP, DETP, and DEDTP. Out of 6 metabolites, we estimated only 3 DAPs i.e. DMP, DETP, DEDTP due to lack of availability of remaining standards. iii. History of time of recent exposure was not known in our study. Hence, the impact of exposure on the spermatogenesis cycle was not estimated and there was not much information on the chemical insult window period in humans. iv. The seasonal variation of OP pesticides exposure was not considered in our study. v. Urinary DAP can be derived from pre-formed metabolites in the environment. vi. We have estimated semen analysis on one occasion. We were unable to repeat semen analysis hence characterization was not confirmed. vii. We didn’t estimate the hormonal changes in our study population. viii. This observational study has various unmeasured confounders like an instrumental variable, design, and so on. Due to time constraints, we have not addressed these confounders.

Though this is a pilot study, it explained a strong association between unintentional OP exposure and semen parameters. Hence, OP exposure status can be included as one of the investigations during the workup of men with an abnormal semen analysis. However, a future study including a larger sample size, more DAP metabolites, collection of more detailed information on demographic and socioeconomic parameters will be required to support our claim.

Conclusion
As OP pesticides exposure can occur through inhalation, ingestion, and so on, their subtle and chronic exposure is affecting various organs of the human body. The current study showed the effect of OP pesticides on semen parameters and concluded that men with idiopathic abnormal semen analysis had significantly higher OP pesticides exposure as compared to men with normal semen analysis. OP-exposed cases had higher urinary OP metabolites levels and more inhibition of pseudocholinesterase and acetylcholinesterase as compared to non-exposed cases pointing towards a severe degree of OP pesticides exposure. A higher percentage of OP-exposed men were farmers and from the rural area.

Acknowledgements
We gratefully acknowledge the participations of all cases and controls and Mrs. Durga, a laboratory technician, for her technical support during this study. This work was performed in partial fulfillment of the requirement for a Master’s Degree in the Faculty of Biochemistry by Dr. Induja Manikandan at JIPMER, Pondicherry. This study was supported by an Intramural research grant from JIPMER, Puducherry, India (JIP/INTRAMURAL/PHS1/2018-19 dated November 11, 2018). There is no conflict of interest in this study.

Authors’ Contributions
I.M.; Sample collections, sample processing and analysis, data acquisition, preparation of Excel files, and statistical analysis. S.B.; Validation of method, sample processing and analysis, data acquisition, and statistical analysis. P.S.A.; Conceptualization, formal analysis, funding acquisition, project administration, resources, software, supervision, validation, drafting, and final approval of the manuscript. C.T., R.N.G.; Conceptualization, selection of cases and controls, project administration, supervision, and final approval of the manuscript. All authors read and approved the final manuscript.

References
1. Zegers-Hochschild F, Adamson GD, de Mouzon J, Ishihara O, Mansour R, Nygren K, et al. International Committee for Monitoring Assisted Reproductive Technology (ICMART) and the World Health Organization (WHO) revised glossary of ART terminology, 2009. Fertil Steril. 2009; 92: 1520-1524.
2. Elhussein OG, Ahmed MA, Suliman SO, Yahya LI, Adam I. Epidemiology of infertility and characteristics of infertile couples requesting assisted reproduction in a low-source setting in Africa, Sudan. Fertil Res Pract. 2019; 5: 7.
3. Adamson PC, Krupp K, Freeman AH, Klausner JD, Reingold AL, Madhivanan P. Prevalence & correlates of primary infertility among young women in Mysore, India. Indian J Med Res. 2011; 134(4): 440-446.
4. Kumar N, Singh AK. Trends of male factor infertility, an important cause of infertility: A review of literature. J Hum Reprod Sci. 2015; 8(4): 191-196.
5. Levine H, Jørgensen N, Martino-Andrade A, Mendiola J, Weksler- Derri D, Måndls I, et al. Temporal trends in sperm count: a systemic review and meta-regression analysis. Hum Reprod Update. 2017; 23(6): 646-659.
6. Sengupta P, Dutta S, Krajewska-Kulak E. The disappearing sperms: analysis of reports published between 1980 and 2015. Am J Mens Health. 2017; 11(4): 1279-1304.
7. Mishra P, Negi MPS, Srivastava M, Singh K, Rajender S. Decline in seminal quality in Indian men over the last 37 years. Reprod Biol}
Bonovosin T, Utyasheva L, Knipe D, Gunnell D, Eddleston M. Suicide by pesticide poisoning in India: a review of pesticide regulation and their impact on suicide trends. BMC Public Health. 2020; 20(1): 251.

Kumar S, Kaushik G, Villaarreal-Chiu JF. Scenario of organophosphate pollution and toxicity in India: a review. Environ Sci Pollut Res Int. 2016; 23(10): 9480-9491.

Perry MJ, Venners SA, Chen X, Liu X, Tang G, Xing H, et al. Organophosphorus pesticide exposures and sperm quality. Reprod Toxicol. 2011; 31(1): 75-79.

Rodríguez H, Bustos-Obregon E. An in vitro model to evaluate the effect of an organophosphor acyloxidase on cell proliferation in mouse seminiferous tubules. Andrologia. 2000; 32(1): 1-5.

Contreras HR, Bustos-Obregon E. Morphological alterations in mouse testis by a single dose of malathion. J Exp Zool. 1999; 284(3): 355-359.

Ghafoori-Khosrowshahi A, Ranjbar A, Mousavi L, Nili-Ahmadabadi H, Ghaffari F, Zeinvand-Lorestani H, et al. Chronic exposure to organophosphate pesticides as an important challenge in promoting reproductive health: a comparative study. J Educ Health Promot. 2019; 8: 149.

Miranda-Contreras L, Cruz I, Osuna JA, Gómez-Pérez R, Berrueta L, Salmen S, et al. Effects of occupational exposure to pesticides on semen quality of workers in an agricultural community of Merida state, Venezuela. Invest Clin. 2015; 56(2): 123-136.

Cremonese C, Piccoli C, Pasqualotto F, Clapauch R, Kolpmann RJ, Kolpmann S, et al. Occupational exposure to pesticides, reproductive hormones levels and sperm quality in young Brazilian men. Reprod Toxicol. 2017; 67: 174-185.

Yucra S, Gasco M, Rubio J, Gonzales GF. Sperm quality in Peruvian pesticide applicators: association between urinary organophosphate metabolites and semen parameters. Environ Health. 2008; 7: 59.

Jamal F, Haque QS, Singh S, Rastogi SK. The influence of organophosphate and carbamate on sperm chromatin and reproductive hormones among pesticide sprayers. Toxicol Ind Health. 2016; 32(8): 1527-1536.

Daoud S, Sellami A, Bouassida M, Kebaili S, Ammar Keskes L, Rebei T, et al. Routine assessment of occupational exposure and its relation to semen quality in infertile men: a cross-sectional study. Turk J Med Sci. 2017; 47(3): 902-907.

Lou F, Cariona G, Vitsale P, Masseroni L, Fino MG, Maggi M. Current smoking is associated with lower seminal vesicles and ejaculate volume, despite higher testosterone levels, in male subjects of infertile couples. Hum Reprod. 2015; 30(3): 590-602.

Matsumoto S, Beeson WL, Shavlik DJ, Siapco G, Jaoeido-Stieg I, Fraser G, et al. Association between vegetarian diet and cardiovascular risk factors in non-Hispanic white participants of the Adventist Health Study-2. J Nutr Sci. 2019; 8: e6.

Dalvi TM, Khairnar MR, Kalghatgi SR. An Update of B.G. Prasad and Kuppswamy socioeconomic status classification scale for Indian population. Indian J Pediatr. 2020; 87(7): 567-568.

World Health Organization. WHO laboratory manual for the examination of human semen and sperm-cervical mucus interaction. 4th ed. Cambridge: Cambridge University Press; 1999.

Ellman GL, Courtney KD, Andres V Jr, Feather-Stone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem Pharmacol. 1961; 7: 88-95.

Hemakanthi De Alwis GK, Needham LL, Barr DB. Determination of dialkyl phosphate metabolites of organophosphorous pesticides in human urine by automated solid-phase extraction, derivatization and gas chromatography mass spectrometry. J Anal Toxicol. 2008; 32(9): 721-727.

Horung RW, Reed LD. Estimation of average concentration in the presence of nondetectable values. Appl Occup Environ Hyg. 1990; 5(1): 46-51.

Nicolopoulos-Stamatel P, Maipas S, Kotampasi C, Stamatel P, Hens L. Chemical pesticides and human health: The urgent need for a new concept in agriculture. Front Public Health. 2016; 4: 148.

Nishant N, Upadhya R. Presence of pesticide residue in vegetable crops: a review. Agricultural Review. 2016; 37(3): 173-185.

Mehpour O, Karrari P, Zamani N, Tsatsakis AM, Abdollahi M. Occupational exposure to pesticides and consequences on male semen and fertility: a review. Toxicol Lett. 2014; 230(2): 146-156.

Kaur K, Kaur R. Occupational pesticide exposure, impaired DNA repair, and diseases. Indian J Occup Environ Med. 2018; 22(2): 74-81.

Rehman S, Usman Z, Rehman S, AlDraihem M, Rehman N, Rehman I, et al. Endocrine disrupting chemicals and impact on male reproductive health. Transl Androl Urol. 2018; 7(3): 490-503.

Gracia-Lor E, Castiglioni S, Bade P, Castignanò E, González-Maríño I, Hapesi E, et al. Measuring biomarkers in wastewater as a new source of epidemiological information: current state and future perspectives. Environ Int. 2017; 99: 131-150.

Glörennec P, Serrano T, Fravallo M, Warembourg C, Monfort C, Cardier S, et al. Determinants of children’s exposure to pyrethroid insecticides in western France. Environ Int. 2017; 104: 76-82.

Katole A, Saoji AV. Prevalence of primary infertility and its associated risk factors in urban population of central India: a community-based cross-sectional study. Indian J Community Med. 2019; 44(4): 337-341.

Dutta S, Bahadur M. Effect of pesticide exposure on the cholinesterase activity of the occupationally exposed tea garden workers of northern part of West Bengal, India. Biomarkers. 2019; 24(4): 317-324.

Li AJ, Kannan K. Urinary concentrations and profiles of organophosphate and pyrethroid pesticide metabolites and phenoxyacid herbicides in populations in eight countries. Environ Int. 2018; 121( Pt 2): 1148-1154.

Muñoz-Quezada MT, Iglesias V, Lucero B, Steenland K, Barr DB, Levy K, et al. Predictors of exposure to organophosphate pesticides in schoolchildren in the Province of Talca, Chile. Environ Int. 2012; 47: 28-36.