Article

Real-Time Detection of the Nutritional Compounds in Green ‘Ratuni UNPAD’ Cayenne Pepper

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Abstract: The detection of nutritional compounds is usually performed through laboratory analysis, which requires extensive planning, time, cost, and effort. Alternatively, visible/near-infrared (Vis/NIR) spectroscopy can be used to determine the presence of nutritional compounds in real-time. This study aimed to investigate the potential application of the Vis/NIR spectroscopy in the quantitative detection of nutritional compounds in green ‘Ratuni UNPAD’ cayenne pepper. Prior to the model development, 80 samples were prepared for the calibration set, while another 40 samples were provided for the prediction set. Subsequently, the parameters used to calculate the model accuracy included the coefficient of correlation in calibration set ($R_{cal}$), coefficient of correlation in prediction set ($R_{pred}$), root mean square error of calibration set (RMSEC), root mean square error of prediction set (RMSEP) and the ratio of prediction to deviation (RPD). The experimental results involving the total carotenoids showed good model indicators with $R_{cal}$, $R_{pred}$, RMSEC, RMSEP, and RPD at 0.94, 0.89, 1.29, 1.75, and 2.21, respectively. Additionally, the analysis of the water content indicated $R_{cal}$, $R_{pred}$, RMSEC, RMSEP, and RPD values of 0.86, 0.85, 0.59, 0.61, and 1.90, respectively, while that of capsaicin had $R_{cal}$, $R_{pred}$, RMSEC, RMSEP, and RPD values of 0.89, 0.90, 117.82, 115.62, and 2.29, respectively. The results showed that Vis/NIR spectroscopy can be used to detect the nutritional compounds in green ‘Ratuni UNPAD’ cayenne pepper based on total carotenoids, water content, and capsaicin parameters.

Keywords: chemometrics; fruit quality; multivariate analysis; rapid detection; Vis–NIR spectroscopy

1. Introduction

Cayenne pepper (Capsicum frustencens L.) is one of the world’s most cultivated horticultural crops. ‘Ratuni UNPAD’ is one of the varieties that has been grown extensively in West Java, Indonesia due to its high yield potential and suitability for planting in the medium-high plains. The potential varies from 7–10 tons/hectare. Moreover, cayenne pepper is also nutrient dense. Generally, some of the ingredients associated with the cayenne pepper include color pigments, water content, and capsaicin [1–4].

The color pigments present in cayenne pepper contain carotenoids, such as lutein, β-carotene, β-cryptoxanthin, zeaxanthin, violaxanthin, capsanthin, and capsorubin, which affect the appearance of the fruit [5] and have a positive relationship with the color of the fruit peel [6]. Additionally, the water content can also affect the appearance of the fruit since it determines the texture, and freshness.

Cayenne pepper, like other commodities in the Capsicum group, contains a unique compound called capsaicin, which gives it a spicy flavor. According to Cheema and Pant [7], capsaicin levels are affected by variety, climate, geographical location, maturity level, and post-harvest processing methods. Capsaicin is the most abundant compound in the capsaicinoid group, accounting for approximately 49–68% of total capsaicinoid content [8].
The various methods used to quantify the water content, carotenoid, and capsaicin in cayenne pepper include gravimetry, spectrophotometry, and chromatography due to their high precision [9,10]. However, the disadvantages associated with these methods include the need for expertise in laboratory analysis, sample preparation, time, cost, the use of chemicals, and the uncertainty of the destruction measurement results in real-time. Practically, it will be challenging to ascertain the nutritional content within a short time frame in the industrial world. Consequently, a technology is required for measuring nutritional compounds present in cayenne pepper in a real-time manner, which provides fast, chemical-free, and accurate determination regarding the chemical content of samples [11,12].

Visible/near-infrared (Vis/NIR) spectroscopy is a recent technology that can be used to predict the chemical content of fruit with speed and accuracy. The quantitative nutritional content can be determined in real-time using this method. Vis/NIR spectroscopy has been extensively used to monitor the quality of various vegetables and fruits for example in the Cucurbitaceae family [13–15], tomato [16], apple [17], banana [18], sapodilla [19], and mandarin orange [20]. Kusumiyati et al. [21] conducted a study on cayenne pepper and predicted the quality attributes of intact and powdered ‘Manik’ and ‘Domba’ cayenne peppers. However, the built calibration model only considers antioxidant property parameters. To the best of our knowledge, there has never been a real-time detection of green ‘Ratuni UNPAD’ cayenne pepper on various nutritional compounds. This calibration model is designed to be more efficient, representative, fast, and accurate in prediction. The application of this technology to the ‘Ratuni UNPAD’ cayenne pepper allows for grading based on its nutritional compounds. This will replace the conventional grading method based solely on the appearance of the sample. The hypothesis is that Vis/NIR spectroscopy guarantees precision and reliability in predicting nutritional compounds in green ‘Ratuni UNPAD’ cayenne pepper. Consequently, this study aimed to test the reliability of Vis/NIR spectroscopy, as well as regression and spectra correction methods to predict water content, total carotenoids, and capsaicin present in the green ‘Ratuni UNPAD’ cayenne pepper.

2. Materials and Methods

2.1. Cayenne Pepper Samples

The samples were harvested at the green stage based on visual appearance. There were 120 samples of green ‘Ratuni UNPAD’ cayenne pepper, all fresh and powdered, with approximately 30 g of cayenne pepper in each. The samples were split into two groups, with 80 representing the calibration set and 40 representing the prediction set. The study was conducted at Horticulture Laboratory, Faculty of Agriculture, Universitas Padjadjaran, Indonesia.

2.2. Chemical Analysis Methods

Various methods were employed to conduct the water content, total carotenoids, and capsaicin analysis. The gravimetric method was used to measure the water content [13]. For total carotenoid quantification, approximately 0.05 g of powdered sample was weighed into a centrifuge tube, followed by 10 mL of acetone. The sample was then centrifuged at 4000 rpm for 10 minutes. The working standard was prepared to obtain 0.5; 1; 2; 4; 8; 16; 32 ppm of β-carotene. Total carotenoids were analyzed at a wavelength of 450 nm using UV–visible spectrophotometer (Shimadzu, UV mini-1240, Tokyo, Japan) [22]. Lastly, the capsaicin was evaluated using high-performance liquid chromatography (HPLC) (Shimadzu, LC 20AT Prominence, Tokyo, Japan). The HPLC was equipped with C18 column (particle size 5 µm; 4.6 mm × 150 mm) and UV–visible detector. The standard solution of capsaicin was made at 2, 4, 8, 16, 32, and 64 ppm. The detection of capsaicin was set at 222 nm. The sample preparation and extraction for capsaicin analysis refer to the procedures described by González-Zamora et al. [23].
2.3. Measurement of Vis/NIR Spectra

The Vis/NIR spectrometer (NirVana AG410, Integrated Spectronics Pty, Ltd., North Ryde, Australia) with a wavelength range of 381–1065 nm at 3 nm intervals was utilized for spectra acquisition. Each sample was placed into a Petri dish and exposed to radiation five times [21]. The irradiated parts of each sample included the top, bottom, right, left, and center surfaces. Afterward, the original spectra were generated from the absorbance spectra and averaged for each sample.

2.4. Data Analysis

The Unscrambler X 10.4 (Camo AS, Oslo, Norway) software was used for the data analysis. The regression methods employed were the partial least squares regression (PLSR) and principal component regression (PCR). PLSR and PCR are the most commonly used linear regression methods for the development of the Vis/NIR calibration model. The similarity between the two methods is that they transform independent variables into principal components (PCs). However, in PCR, PCs are formed only by involving the independent variables, while in PLSR, PCs are determined by independent and dependent variables. The goal of PLSR is to maximize the covariance of both variables [24,25]. Additionally, the standard normal variate (SNV) and first derivative Savitzky–Golay (dg1) spectra correction methods were applied. The parameters associated with the model accuracy were evaluated by the coefficient of correlation in the calibration set (R\textsuperscript{cal}), root mean square error of the calibration set (RMSEC), coefficient of correlation in the prediction set (R\textsuperscript{pred}), root mean square error of the prediction set (RMSEP), and the ratio of prediction to deviation (RPD). The developed model is expected to have low RMSEC and RMSEP, and also have high R\textsuperscript{cal}, R\textsuperscript{pred}, and RPD [26,27].

3. Results and Discussion

3.1. Analysis of Vis/NIR Spectra

The absorbance spectra of green ‘Ratuni UNPAD’ cayenne pepper are shown in Figure 1. Several peaks and valleys have been identified to provide relevant information and are closely related to the chemical content of the sample. The wavelengths used ranged from 381–1065 nm, and the absorption peaks indicated that these wavelengths were related, and they became the primary focus of biochemical information [28]. The green ‘Ratuni UNPAD’ cayenne pepper displayed absorption peaks approximately at 465 nm, 498 nm, 639 nm, 642 nm, and 915 nm, and they were found to be closely related to color pigments and water. The wavelengths ranging from 465 nm to 642 nm were associated with various color pigments such as carotenoids, xanthophylls, and chlorophyll [29]. Furthermore, there was a peak of approximately 915 nm that correlated with the water content. The previous study reported the wavelength of 760–970 nm was discovered to be the second overtones of O-H bands that correspond to water absorption [30].

3.2. Prediction Results of Nutritional Compounds

The nutritional compounds data collected from laboratory analysis were displayed in Table 1. The data were then used for model development and evaluation. The calibration model was achieved through regression analysis involving spectra data of samples and actual data of nutritional compounds (calibration). The developed calibration model was then applied to predict the unknown samples (prediction). The best calibration model was selected based on the consistency of the high R\textsuperscript{cal}, R\textsuperscript{pred}, and RPD, as well as the low RMSEC/RMSEP [31]. The presence of noise in the spectra could reduce the model accuracy, which led to the application of various spectra correction methods. The application of PLSR + original resulted in the best model for total carotenoids with R\textsuperscript{cal}, R\textsuperscript{pred}, RMSEC, RMSEP, and RPD having values of 0.94, 0.89, 1.29, 1.75, and 2.21, respectively (Table 2). These results indicate that the total carotenoids can be predicted with a high degree of accuracy and reliability for further prediction [32,33]. This analysis is consistent with the use of Vis/NIR spectroscopy on total banana fruit carotenoids, which yielded
$R^2_{\text{cal}}, R^2_{\text{pred}}, \text{RMSEC}, \text{RMSEP}, \text{and RPD values of 0.95, 0.96, 18.89, 28.70, and 3.35, respectively [34]. This discovery supported the hypothesis that Vis/NIR spectroscopy can predict total carotenoids in cayenne pepper, particularly the ‘Ratuni UNPAD’ variety, which was determined statistically through the multivariate analysis.}

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**Figure 1.** Original spectra for green ‘Ratuni UNPAD’ cayenne pepper.

**Table 1.** Summary of laboratory analysis of water content, total carotenoids, and capsaicin in calibration set and prediction set.

| Nutritional Compounds       | Calibration Set | Prediction Set |          |          |          |          |
|-----------------------------|-----------------|----------------|----------|----------|----------|----------|
|                             | Range       | Mean     | Range   | Mean     |          |          |
| Water content (%)           | 72.26–76.87  | 74.52    | 72.47–76.86 | 74.57    |
| Total carotenoids (mg/100 g) | 7.79–25.69  | 16.65    | 9.24–22.73 | 16.93    |
| Capsaicin (mg/100 g)        | 208.57–1424.72 | 752.26 | 240.35–1328.36 | 762.95   |

**Table 2.** Summary of PLSR and PCR of water content, total carotenoids, and capsaicin in calibration set and prediction set.

| Nutritional Compounds | Regression Methods | Spectra | PC | $R_{\text{cal}}$ | RMSEC | $R_{\text{pred}}$ | RMSEP | RPD |
|-----------------------|--------------------|---------|----|-----------------|-------|-----------------|-------|-----|
| Water content         | PLSR               | Original | 7  | 0.86            | 0.59  | 0.85            | 0.61  | 1.90 |
|                       |                    | dg1     | 7  | 0.83            | 0.64  | 0.76            | 0.75  | 1.57 |
|                       |                    | SNV     | 7  | 0.82            | 0.65  | 0.81            | 0.68  | 1.72 |
|                       | PCR                | Original | 11 | 0.86            | 0.59  | 0.83            | 0.64  | 1.81 |
|                       |                    | dg1     | 7  | 0.76            | 0.75  | 0.80            | 0.69  | 1.69 |
|                       |                    | SNV     | 8  | 0.81            | 0.68  | 0.81            | 0.67  | 1.74 |
| Total carotenoids     | PLSR               | Original | 14 | 0.94            | 1.29  | 0.89            | 1.75  | 2.21 |
|                       |                    | dg1     | 15 | 0.95            | 1.18  | 0.86            | 1.97  | 1.96 |
|                       |                    | SNV     | 14 | 0.93            | 1.47  | 0.82            | 2.17  | 1.78 |
|                       | PCR                | Original | 20 | 0.93            | 1.43  | 0.87            | 1.91  | 2.02 |
|                       |                    | dg1     | 20 | 0.91            | 1.65  | 0.78            | 2.40  | 1.61 |
|                       |                    | SNV     | 20 | 0.90            | 1.67  | 0.83            | 2.11  | 1.83 |
Table 2. Cont.

| Nutritional Compounds | Regression Methods | Spectra | PC | R<sub>cal</sub> | RMSEC | R<sub>pred</sub> | RMSEP | RPD |
|-----------------------|--------------------|---------|----|----------------|--------|-------------|--------|-----|
| Capsaicin             | PLSR               | Original| 8  | 0.89           | 117.82 | 0.90        | 115.62 | 2.29|
|                       |                    | dg1     | 9  | 0.89           | 119.29 | 0.64        | 199.98 | 1.32|
|                       |                    | SNV     | 9  | 0.89           | 120.60 | 0.74        | 176.03 | 1.50|
|                       | PCR                | Original| 11 | 0.89           | 118.26 | 0.90        | 115.92 | 2.28|
|                       |                    | dg1     | 14 | 0.87           | 130.14 | 0.57        | 215.19 | 1.23|
|                       |                    | SNV     | 13 | 0.88           | 124.40 | 0.69        | 188.14 | 1.41|

PC: principal component, R<sub>cal</sub>: coefficient of correlation in calibration set, RMSEC: root mean square error of calibration set, R<sub>pred</sub>: coefficient of correlation in prediction set, RMSEP: root mean square error of prediction set, RPD: the ratio of prediction to deviation.

The model development and evaluation for water content and capsaicin also showed sufficient results. Furthermore, the best water content results were obtained using PLSR + original, with R<sub>cal</sub>, RMSEC, R<sub>pred</sub>, RMSEP, and RPD with their values given as 0.86, 0.59, 0.85, 0.61, and 1.90, respectively. The Vis/NIR prediction of the water content of pomelo fruit was reported to have similar accuracy, with the R<sup>2</sup><sub>cal</sub>, R<sup>2</sup><sub>pred</sub>, RMSEC, and RMSEP having accuracies of 0.89, 0.71, 0.03, and 0.04, respectively [35]. The accuracy of the capsaicin parameter with PLSR + original was the highest, with R<sub>cal</sub>, R<sub>pred</sub>, RMSEC, RMSEP, and RPD having accuracies of 0.89, 0.90, 117.82, 115.62, and 2.29, respectively. The results of this study produced higher accuracies than those obtained by Johnson et al. [36] in Habanero chili, which gave the capsaicin accuracy results of R<sup>2</sup><sub>cal</sub>, RMSEC, R<sub>pred</sub>, RMSEP, and RPD to be 0.19, 0.589, 0.15, 0.611, and 1.08, respectively. However, Lim et al. [37] obtained better results on capsaicinoids in Korean red pepper with R<sup>2</sup><sub>cal</sub>, R<sup>2</sup><sub>pred</sub>, RMSEC, and RMSEP having values of 0.87, 0.84, 12.18, and 13.63, respectively. Different accuracy in the prediction of nutritional compounds can occur due to various factors, for example, the variability of data, the number of samples, type of samples, samples condition, instrumentation, model development, etc.

Figure 2 depicts the visualization of the data distribution for the calibration and prediction sets. The two sets were selected after comparing the results of the best regression and spectra correction methods for each quality attribute. Subsequently, satisfactory results were obtained from all quality attributes, with R<sub>cal</sub> and R<sub>pred</sub> having values greater than or equal to 0.86 and 0.85, respectively. The model obtained met the criteria for being designated as a robust model for predicting total carotenoids, as well as the water content and capsaicin. Furthermore, RPD values obtained for water content, total carotenoids, and capsaicin were 1.90, 2.21, and 2.29, respectively. The RPD below 1.5 indicate poor prediction and unsuitability, while those between 1.5 and 2 show that the model can distinguish between low and high response variables. Additionally, RPD values between 2 and 2.5 suggest the possibility of making coarse quantitative predictions, while a value between 2.5 and 3 or above indicates good and outstanding prediction accuracy [32,38].

3.3. Wavelength Selection by Regression Coefficient

Figure 3 describes the regression coefficients on model development of water content, total carotenoids, and capsaicin. The water absorption was found at wavelengths of 561, 621, 651, 906, and 999 nm (Figure 3a). Our findings regarding the water absorption peaks were comparable to those of strawberries at various stages of ripeness [39]. In this study, we employed a total of 230 wavelengths; however, the peaks that contributed to the prediction of total carotenoids were 402, 429, 483, 633, 645, 669, 750, 891, and 930 nm (Figure 3b). This discovery is consistent with studies arranged by Walsh et al. [29] and Ruiz et al. [40], which stated that the carotenoid content has wavelengths ranging from 420–503 nm and 940–1200 nm. Zhang et al. [30] and Jamshidi et al. [41], also reported that the absorption around 550 nm and 650 nm were associated with pigments interpretation.
Figure 2. Scatter plots and regression line of calibration set (green) and prediction set (orange) of water content (a,b), total carotenoids (c,d), and capsaicin (e,f) in green ‘Ratuni UNPAD’ cayenne pepper.
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Figure 3.

Figure 3. Regression coefficients: (a) water content, (b) total carotenoids, and (c) capsaicin of green ‘Ratuni UNPAD’ cayenne pepper.

Furthermore, this study revealed information about the wavelength that contributed to the capsaicin. Figure 3c represents wavelengths of 468, 519, 633, 657, 714, 912, and 1005 nm at peaks for capsaicin. The previous study addressed that absorption of the capsaicinoids content was around 450–950 nm [37]. This finding verifies that Vis/NIR spectroscopy has peaks that can predict water content, total carotenoids, and capsaicin of green ‘Ratuni UNPAD’ cayenne pepper. In this study, reliable and potent nutritional compounds were detected in real-time and with high accuracy. However, there has not been any other study on Vis/NIR technology that has developed a predictive potential for various quality attributes of green ‘Ratuni UNPAD’ cayenne pepper besides this study. The application of Vis/NIR spectroscopy has the advantage of being practical, efficient, and real-time. Overall, the hypothesis in this study demonstrates that Vis/NIR spectroscopy can be trusted to predict nutritional compounds in green ‘Ratuni UNPAD’ cayenne pepper.
4. Conclusions

The Vis/NIR spectroscopy has the potential to predict the nutritional compounds of green ‘Ratuni UNPAD’ cayenne pepper in real-time. The overall accuracy of the carotenoids obtained for \( R^2_{cal} \), \( R^2_{pred} \), RMSEC, RMSEP, and RPD were 0.94, 0.89, 1.29, 1.75, and 2.21, respectively. Furthermore, the \( R^2_{cal} \), \( R^2_{pred} \), RMSEC, RMSEP, and RPD values for water content and capsaicin were 0.86, 0.85, 0.59, 0.61, and 1.90, and of 0.89, 0.90, 117.82, 115.62, and 2.29, respectively. The finding of this study also confirmed the expectations that Vis/NIR spectroscopy is reliable in detecting the total carotenoids, water content, and capsaicin of green ‘Ratuni UNPAD’ cayenne pepper. The application of Vis/NIR spectroscopy as a real-time and accurate quality prediction technology needs to be explored further because it has many advantages over other measurement methods in terms of practicality, labor, time, and cost. In terms of cost–benefit, this technique can reduce cost for quality control when used by cayenne pepper producers who need daily quality control for large samples.

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