We could not find any evidence that neutrophils in patients with stable CAD were primed, i.e. more prone to increase either the expression or the amount of high-affinity CD11b/CD18-integrins in CAD patients compared to controls, a marker of chronic disease, an atherogenic factor or a consequence of the drug treatment.

...showed an impaired activation status. It remains to be elucidated whether the neutrophil dysfunction in CAD is mainly...
changes of the integrin, thereby exposing an epitope that enables ligand binding, the so called I-domain [17].

In the present study, we hypothesized that neutrophils in patients with stable CAD were in a primed or pre-activated state compared to neutrophils in healthy individuals. The neutrophil activation status, involving the expression, affinity state and signalling capacity of β2-integrins during basal and stimulatory conditions as well as the innate ROS production, was investigated in a paired patient-control design.

RESULTS

Clinical and laboratory characteristics of subjects

Clinical and laboratory characteristics of patients and controls are given in Table 1. The use of medication differed significantly between the two groups. A small number of controls received treatment with low-dose aspirin, beta-blockers, angiotensin-converting enzyme inhibitors or statins due to the presence of risk factors, such as hypertension and hyperlipidemia. There were no differences in the amount of C-reactive protein (CRP) or leukocyte blood count between patients and controls.

Table 1. Clinical and laboratory characteristics of patients and controls.

|                         | Patients | Controls | p values |
|-------------------------|----------|----------|----------|
| Total no.               | 30       | 30       | ns       |
| Female/Male (no.)       | 5/25     | 5/25     | ns       |
| Age (years)             | 63 (59–67) | 63 (60–68) | ns       |
| Smokers n (%)           | 4 (13)   | 4 (13)   | ns       |
| Blood pressure (mm Hg)  |          |          |          |
| Systolic                | 135 (120–146) | 141 (130–152) | ns       |
| Diastolic               | 83 (78–91)  | 84 (78–92)  | ns       |
| Body mass index (kg/m²) | 27 (24–29)  | 25 (24–29)  | ns       |
| Medication n (%)        | 28 (93)   | 3 (10)    | <0.001   |
| ACE-I/ARB 2)            | 16 (52)   | 2 (7)     | <0.001   |
| Statin                  | 30 (100)  | 3 (10)    | <0.001   |
| Low-dose Aspirin        | 30 (100)  | 1 (3)     | <0.001   |
| Laboratory variables 3) |          |          |          |
| CRP 4) (mg/mL)          | 0.9 (0.3–1.8) | 1.2 (0.4–2.2) | ns       |
| Blood glucose (mmol/l)  | 5.1 (4.4–5.9) | 5.8 (5.2–6.5) | <0.01   |
| Lipids (mmol/l)         |          |          |          |
| Total cholesterol       | 4.5 (4.1–5.0) | 5.2 (5.0–6.0) | <0.01   |
| Low-density lipoprotein (LDL) cholesterol | 2.2 (1.9–2.8) | 2.9 (2.5–3.4) | <0.05 |
| High-density lipoprotein (HDL) cholesterol | 1.4 (1.2–1.6) | 1.5 (1.3–2.1) | ns       |
| Triglycerides           | 1.5 (1.0–1.9) | 1.2 (0.9–1.8) | ns       |
| Blood count (cells/µL)  | 6.1 (5.3–6.8) | 5.6 (4.8–6.7) | ns       |
| Neutrophils             | 3.0 (2.6–3.5) | 2.7 (2.3–3.9) | ns       |

1) Data are given as median (inter-quartile range).
2) ACE-I, angiotensin converting enzyme inhibitor; ARB, angiotensin receptor blocker.
3) Measured at the accredited clinical chemistry laboratory at Linköping University Hospital, Sweden.
4) CRP, C-reactive protein.
5) doi:10.1371/journal.pone.0001056.t001

Baseline expression and high-affinity state of β2-integrins on neutrophils

Using flow cytometry, the mean fluorescence intensity (MFI) values revealed that patients with stable CAD and control subjects expressed the same amount of β2-integrins on their neutrophils (349±80 and 337±104, respectively; mean MFI±SD; Fig. 1A.C). Also, no differences in the amount of β2-integrins exposing the high-affinity epitope could be detected when comparing neutrophils from patients and controls (19±8 and 24±12, respectively; Fig. 1B,D).

β2-integrin up-regulation on neutrophils upon IL-8 or LTB4 stimulation

The pro-inflammatory chemoattractants IL-8 and LTB4 were used to study up-regulation of β2-integrins on neutrophils. In both patients and healthy individuals, stimulation with IL-8 induced an up-regulation of β2-integrins (148%±30 and 152%±37, respectively; Fig. 2A,C), and an even stronger up-regulation was induced by LTB4 (172%±37 and 180%±36, respectively; Fig. 2B,D) compared to unstimulated cells. No significant difference was detected between patients and controls in regard to the total expression of β2-integrins (Fig. 2C and D) or in the increased percentage of up-regulated β2-integrins upon chemoattractant stimulation (data not shown).

The expression of high-affinity state β2-integrins on neutrophils upon IL-8 or LTB4 stimulation

The inside-out regulation of the high-affinity β2-integrins was evaluated in neutrophils stimulated with IL-8 or LTB4. Stimulation with IL-8 (Fig. 3A–B) and LTB4 (Fig. 3C–D) induced a prominent increase in the amount of CD11b exposing the high-affinity epitope compared to unstimulated neutrophils in both patients and controls (230%±81 and 225%±76; 308%±102 and 316%±95, respectively). No significant difference in the total number of high-affinity β2-integrins (Fig. 3B and D) or in the increased percentage of high-affinity β2-integrins (data not shown) was detected between neutrophils from patients and controls upon chemoattractant stimulation.

β2-integrin signalling capacity in neutrophils

The signalling capacity of β2-integrins on neutrophils was determined by measuring ROS production in response to C3bi-opsonized yeast particles. We found that neutrophils from the patients produced significantly lower levels of ROS compared to their individually matched controls (with a mean reduction of 30%±17; Fig. 3E–F). In support of previous studies on neutrophils from healthy blood donors [18], the production of ROS elicited by C3bi-opsonized yeast particles in neutrophils isolated from patients was mainly intracellular (data not shown).

ROS production in neutrophils upon IL-8 or LTB4 stimulation

To investigate if neutrophils from patients with stable CAD displayed a more primed character, the total ROS production (i.e. intracellularly produced plus extracellularly released) was measured in response to IL-8 and LTB4. Both IL-8 and LTB4 elicited mainly extracellular ROS production (data not shown). The ROS responses were of the same magnitude in neutrophils from patients as in neutrophils from controls (Fig. 4A–D). However, PMA (i.e. a non-receptor mediated response) elicited significantly lower production of ROS in neutrophils from patients compared to neutrophils from individually matched controls, with a mean reduction of 21%±12 (Fig. 4E–F).
Figure 1. Baseline expression of CD18 and high-affinity state CD11b on neutrophils in patients with stable CAD.Whole blood from patients (P) and individually matched control persons (C) were incubated at 37°C with (A,C) the MHM23-antibody or with (B,D) the CBRM1/5-antibody for 5 and 10 min, respectively, whereafter the samples were incubated on ice. Contaminating erythrocytes were removed by lysis, and antibody bound to CD18 or to CD11b-integrins displaying the high-affinity epitope was detected by FACS analysis. The cells were gated to identify the neutrophil population, and 10,000 cells were analyzed in each sample. Data are shown as one representative histogram plot (A,B) or as median of the mean fluorescence intensity (MFI) values (C,D) from 30 (A,C) and 25 (B,D) experiments per cohort done in triplicate. Mean value is indicated as a black hyphen. Wilcoxon signed-ranks test was used to determine significance between each patient and its individually matched control.
doi:10.1371/journal.pone.0001056.g001
Figure 2. IL-8 and LTB$_4$ induced expression of CD18 on neutrophils from patients with stable CAD. Whole blood from patients (P) and individually matched control persons (C) were stimulated for 10 min at 37°C with (A,C) IL-8 (10 ng/mL) or (B,D) LTB$_4$ (100 nM), and the samples were co-incubated with the MHM23-antibody during the last 5 min of stimulation. The stimulation was stopped by incubation the samples on ice, and contaminating erythrocytes were removed by lysis. Antibody bound to CD18 was detected by FACS analysis. The cells were gated to identify the neutrophil population, and 10 000 cells were counted in each sample. Data are shown as one representative histogram plot (A,B) or as median of the mean fluorescence intensity (MFI) values (C,D) from 22 (A,C) and 30 (B,D) experiments per cohort done in triplicate. Mean value is indicated as a black hyphen. Wilcoxon signed-ranks test was used to determine significance between each patient and its individually matched control.

doi:10.1371/journal.pone.0001056.g002
DISCUSSION

The present study investigated the activation status of neutrophils in patients with stable CAD by determining the cells adhesive property and their ability to produce ROS. We found that the baseline expression of \( \beta^2 \)-integrins did not differ between neutrophils in patients and healthy controls; neither did the amount of \( \beta^2 \)-integrins exposing the high-affinity epitope. The results are in line with previous reports showing similar levels of \( \beta^2 \)-integrins on neutrophils in patients with stable CAD as in healthy controls [6,7,10,11]. However, the number of integrins on the neutrophil surface does not reflect the functional state of the integrin. In order to enable cellular adhesion, a conformational change of the \( \beta^2 \)-integrin is needed to increase its affinity [19].

Our results, demonstrating similar levels of high-affinity CD11b-integrin in patients with stable CAD as in healthy controls, provide new knowledge and extend thereby data from previous reports [6,7]; also contradicting an \textit{in vivo} neutrophil activation in stable CAD. IL-8 and LTB4 are both potent leukocyte chemoattractants, known to promote several inflammatory disorders, including atherosclerosis. Serum levels of IL-8 are elevated in patients with stable CAD [20], and in healthy individuals elevated levels of IL-8 are associated with an increased risk of future CAD [21]. As expected, the expression and affinity of \( \beta^2 \)-integrins, as well as the ROS production were increased in neutrophils upon stimulation with IL-8 or LTB4. However, these stimuli-induced responses of neutrophils in patients were comparable to the ones detected in healthy individuals. Data from previous studies have suggested

Figure 3. IL-8 and LTB4-induced expression of high-affinity state CD11b on neutrophils from patients with stable CAD. (A–D) Whole blood from patients (P) and individually matched control persons (C) were incubated for 1 min with CBRM1/5 at 37 °C, before stimulated for 10 min with (A–B) IL-8 (10 ng/mL) or (C–D) LTB4 (100 nM) at 37 °C. The stimulation was stopped by incubating the samples on ice, and contaminating erythrocytes were removed by lysis. Antibody recognizing integrins displaying the high-affinity epitope was detected by FACS analysis. The cells were gated to identify the neutrophil population, and 10 000 cells were counted in each sample. Data are shown as one representative histogram plot (A,C) or as median of the mean fluorescence intensity (MFI) values (B,D) from 20 (A–B) and 25 (C–D) experiments per cohort done in triplicate. (E–F) Isolated neutrophils from patients and control persons were stimulated with C3bi-opsonized yeast particles (5×10⁶/mL), and the production of ROS was measured continuously for 60 min at 37 °C. Data are expressed as counts per min (cpm) and shown as one representative histogram plot (E) or as total ROS production peak value from 10 experiments per cohort (F). Mean value is indicated as a black hyphen. Wilcoxon signed-ranks test was used to determine significance between each patient and its individually matched control (** represents significant difference; \( p < 0.01 \)).

doi:10.1371/journal.pone.0001056.g003
that neutrophils in patients with stable CAD or in individuals at high risk for vascular events possess a primed character compared to neutrophils from healthy individuals, i.e. the cells are "hyperactive" ex vivo as assessed by increased chemotactic activity and increased production of ROS, LTB₄ and IL-8 [6,8,9]. However, our data could not support the hypothesis that neutrophils in patients with stable CAD are more prone to activation.

In a recently published study, a different approach was used to study neutrophil activation. In a small group of CAD patients, a skin blister model was applied in order to describe transmigration of neutrophils at different stages of interstitial inflammation [10]. In vivo transmigrated neutrophils in patients showed an increased capacity to release gelatine-associated lipocalin and matrix metalloproteinase-9 in a milieu of intermediate, but not intense, inflammation. The authors thereby proposed that the dysfunctional neutrophil response in CAD patients was a consequence of a primed state of circulating neutrophils. It is possible that the stimulation with IL-8 and LTB₄ represented a too intense inflammatory state in which a primed character of the neutrophils was abolished by e.g. receptor desensitization [22,23]. Arguing against this is that IL-8 and LTB₄ are intermediate chemoattractants, known to induce more moderate cellular responses [24]. In addition, it could be argued that the stimulation with IL-8 and LTB₄ is a more clinically relevant model with regard to atherosclerosis than the skin blister model [10].

An aberrant neutrophil activation status in atherosclerosis, not only involving priming but also refractivity to further stimulation, has been discussed. Paulsson and co-workers [10] found that in vivo transmigrated neutrophils in CAD patients had a reduced capacity to up-regulate CD11b and to produce H₂O₂ compared to neutrophils in healthy controls. In the present study, we show that neutrophils in patients with stable CAD possessed a functional inside-out regulation in relation to β₂-integrin expression and affinity. However, upon stimulation with C3bi-opsonised yeast, which binds and activates CD11b/CD18 [25], the β₂-integrin signalling was significantly decreased in neutrophils from CAD patients compared to controls. This reduced response to C3bi-opsonised yeast might also reflect the adhesive property of the β₂-integrin [26], but regardless of whether the observed down-regulated signalling capacity was due to a dysfunctional signalling capacity per se or resulted from decreased adhesion, the results indicate that inside-out signalling regulating CD11b/CD18-

Figure 4. IL-8 and LTB₄ induced ROS production in neutrophils from patients with stable CAD. The production of ROS in response to (A–B) IL-8 (100 ng/mL), (C–D) LTB₄ (100 nM), or (E–F) PMA (50 nM), was measured in isolated neutrophils from patients (black dashed line and black dot, respectively) and individually matched controls (grey line and grey dot, respectively). Data are expressed as counts per min (cpm) and shown as one representative histogram plot (A,C,E) or as total ROS production peak value from 9–10 experiments per cohort (B,D,F). Mean value is indicated as a black hyphen. Wilcoxon signed-ranks test was used to determine significance between each patient and its individually matched control (** represents significant difference; p<0.01).

doi:10.1371/journal.pone.0001056.g004
integran avidity was down-regulated in patients with stable CAD. In addition, the non-receptor-mediated ROS production by PMA was significantly decreased in patients, suggesting that the optimal capacity to generate ROS was deficient in patients. Altogether, our data point towards a dysfunctional activation status of neutrophils in CAD patients, involving cellular processes mediated by β2-integrins.

In contrast to our previous data [11], the CAD patients in the present study were without signs of systemic inflammatory activity, as assessed by CRP and neutrophil counts. One plausible explanation is that each patient in the present study was treated with both low-dose aspirin and statin. Aspirin and statin are both well-known CRP-lowering agents (reviewed in: [27]). A primed neutrophil response has also been shown to attenuate upon therapy with these drugs. By affecting protein prenylation, statin modulates a variety of signalling molecules; many of which are involved in regulating adhesion (reviewed in: [28]). In high-risk individuals, a 4-week treatment with statin could be to reduce the synthesis of IL-8 in neutrophils and decrease the neutrophil migration and chemotaxis [9]. Aspirin influence a number of mechanisms and signalling pathways involved in neutrophil activation, for instance attenuating the expression of IL-8 [29] and the LTB4-induced migration of neutrophils [8]. Aspirin-triggered lipoxins attenuates the expression of β2-integrins [30], decreases the LTB4-induced adhesion-state of neutrophils by affecting the avidity of the β2-integrin [31], and inhibits neutrophil adhesion to activated endothelial cells [32].

To conclude, the present study does not give any support for a role of neutrophil priming in stable CAD. The neutrophils in patients with stable CAD were not more activated in vitro than were cells in healthy controls, neither were the neutrophils in patients more prone to activation in vivo. Rather, the neutrophils in patients displayed a decreased β2-integrin signalling and/or adhesion capacity and a deficient intrinsic capacity to generate ROS. Although the results indicate that the drug therapy of today is efficient in maintaining the neutrophils in a non-adhesive manner; an outcome that most certainly has a positive impact on the prognosis, the clinical relevance of neutrophil dysfunction in CAD remains to be elucidated. Is it mainly a marker of chronic inflammatory disease or a factor with impact on the progress of CAD?

MATERIALS AND METHODS

Subjects

Thirty patients with angiographically verified stable CAD were recruited from the Department of Cardiology, Heart Centre, Linkoping University Hospital, Sweden. The patients had effort-related angina in accordance with the Canadian Cardiovascular Society functional classes I and II without any worsening of symptoms the latest 6 months. Patients were excluded if they were >65 years old, had severe heart failure, immunologic disorders, neoplasm disease, evidence of acute or recent (<2 months) infection, recent major trauma, surgery or revascularization procedure, or treatment with immunosuppressive or anti-inflammatory agents (except low-dose aspirin). Each patient was matched, regarding age and sex, with a clinically healthy control randomly selected from a population register representing the hospital recruitment area. The control subjects were anamnestically healthy and with normal routine laboratory tests. All patients and control persons gave informed oral consent. The appointed ethics committee at Linkoping University approved the research protocol, and the study was conducted in accordance with the ethical guidelines of Declaration of Helsinki.

Collection of blood

For each experimental set-up, venous peripheral blood was drawn from one patient and its matched control in the morning after a 12 h fast, and collected in heparin-containing vacutainer tubes. The samples were blinded during all experiments.

Flow cytometry

Immmuno-labelling was performed, as previously described [31], to investigate changes in the total expression of β2-integrin and in the expression of β2-integrins displaying the high-affinity epitope. In short, whole blood (50 µl/sample) was pre-heated at 37°C. In order to study the high-affinity state of the β2-integrin, a monoclonal FITC-conjugated mouse anti-human CD11b antibody, recognizing the ligand binding domain (i.e., the I-domain [33]; CBRM1/5; Biosite) was used. This antibody was given 1 min before stimulation with Interleukin-8 (IL-8; Sigma-Aldrich) or and Leukotriene B4 (LTB4; Larodan Fine Chemicals AB). In samples used for detecting the plasma membrane expression of CD18, the monoclonal FITC-conjugated mouse anti-human CD18 antibody (MHM23; DakoCytomation) was added 5 min after stimulation. For all samples, the stimulation was stopped by incubating the samples on ice. Contaminating erythrocytes were removed by ice-cold cell lysis (NH4Cl 150 mM, KHCO3 10 mM, EDTA 100 µM) for 5 min at 15°C. The cells were collected and fixed in ice-cold 0.1% paraformaldehyde (PFA). Mean fluorescence values from 10 000 cells/sample were determined by flow cytometry using a Becton Dickinson FACSCalibur (Becton Dickinson, San Jose, CA/USA). Cell populations were identified by plotting FSC versus SSC excluding cell debris, followed by gating and further analysis of the neutrophil (granulocyte) population (Fig. 5). Auto-fluorescence of unstained cells was routinely analyzed. Unspecific binding was determined using a FITC-conjugated isotypic mouse anti-human IgG1 (DakoCytomation).

Preparation of neutrophils

Neutrophils were prepared by density gradient centrifugation, as previously described [31]. In short, freshly drawn heparinized...
blood was layered on top of Lymphoprep™ and Polymorph-
prep™ (Axis-Shield AS), and centrifuged in a swing-out centrifuge
(400 xg, 30 min, RT). The granulocyte band was collected, washed and resuspended in a small volume of PBS (pH 7.3; made-in-house), and concentrated by a short spin. Contaminating erythrocytes were removed by hypotonic lysis, and the cells were washed, resuspended in Krebs-Ringer glucose buffer (KRG; 10 mM glucose, 1 mM Ca²⁺, and 1.2 mM Mg²⁺; pH 7.3; made-in-house) without Ca²⁺, and kept on ice.

Oxygenation of yeast particles
Serum opsonized yeast particles were prepared, as previously described [34], using a six-channel Biolumat LB9505 (Berthold Co., Wildbad, Germany). Neutrophils (5 x 10⁶/mL) were diluted in KRG in polypropylene tubes to a total volume of 1 mL (400 µL of 10 mM glucose, 1 mM Ca²⁺, 1.2 mM Mg²⁺; pH 7.3; made-in-house), and concentrated by a short spin. Contaminating erythrocytes were removed by hypotonic lysis, and the cells were washed, resuspended in Krebs-Ringer glucose buffer (KRG; 10 mM glucose, 1 mM Ca²⁺, and 1.2 mM Mg²⁺; pH 7.3; made-in-house) without Ca²⁺, and kept on ice.

Determination of ROS production
Chemiluminescence was used to determine the ROS production of isolated neutrophils from 10 patients with stable CAD and 10 control subjects, respectively. The measurements were performed, as previously described [33], using a six-channel Biolumat LB9505 (Berthold Co., Wildbad, Germany). Neutrophils (5 x 10⁶/mL) were diluted in KRG in polypropylene tubes to a total volume of 1 mL (400 µL of 10 mM glucose, 1 mM Ca²⁺, 1.2 mM Mg²⁺; pH 7.3; made-in-house), and concentrated by a short spin. Contaminating erythrocytes were removed by hypotonic lysis, and the cells were washed, resuspended in a small volume of PBS (pH 7.3; made-in-house), and concentrated by a short spin. Contaminating erythrocytes were removed by hypotonic lysis, and the cells were washed, resuspended in Krebs-Ringer glucose buffer (KRG; 10 mM glucose, 1 mM Ca²⁺, and 1.2 mM Mg²⁺; pH 7.3; made-in-house) without Ca²⁺, and kept on ice.

Chemiluminescence was used to determine the ROS production of isolated neutrophils from 10 patients with stable CAD and 10 control subjects, respectively. The measurements were performed, as previously described [33], using a six-channel Biolumat LB9505 (Berthold Co., Wildbad, Germany). Neutrophils (5 x 10⁶/mL) were diluted in KRG in polypropylene tubes to a total volume of 1 mL (400 µL of 10 mM glucose, 1 mM Ca²⁺, 1.2 mM Mg²⁺; pH 7.3; made-in-house), and concentrated by a short spin. Contaminating erythrocytes were removed by hypotonic lysis, and the cells were washed, resuspended in a small volume of PBS (pH 7.3; made-in-house), and concentrated by a short spin. Contaminating erythrocytes were removed by hypotonic lysis, and the cells were washed, resuspended in Krebs-Ringer glucose buffer (KRG; 10 mM glucose, 1 mM Ca²⁺, and 1.2 mM Mg²⁺; pH 7.3; made-in-house) without Ca²⁺, and kept on ice.

REFERENCES

1. Ott I, Neumann FJ, Gawaz M, Schmitt M, Schomig A (1996) Increased neutrophil-platelet adhesion in patients with unstable angina. Circulation 94: 1239–1246.
2. Biffon A, Biasucci LM, Liao Z, D’Onofrio G, Crea F, et al. (2002) Widespread coronary inflammation in unstable angina. N Engl J Med 347: 5–12.
3. Patel PB, Pflü SE, Cleman MW, Brennan JJ, Howes G, et al. (2004) Comparison of coronary artery specific leukocyte-platelet conjugate formation in unstable versus stable angina pectoris. Am J Cardiol 93: 410–413.
4. Naruo T, Ueda M, Hase K, van der Wal AC, van der Loos CM, et al. (2002) Neutrophil infiltration of culprit lesions in acute coronary syndromes. Circulation 106: 2984–2990.
5. Madjid M, Awan I, Willerson JT, Casscells SW (2004) Leukocyte count and ROS production were analysed by Wilcoxon signed rank test to compare levels between a patient and its paired control. A p<0.05 was considered significant.

ACKNOWLEDGMENTS

We would like to thank Ms Mona Borjeson at the Department of Cardiology for preparing blood samples from all study subjects, and Ms Johanna Wernersson for excellent technical assistance. Dr. Tony Forslund is acknowledged for advice regarding flow cytometry analyses performed in whole blood.

Author Contributions

Conceived and designed the experiments: ES LJ. Performed the experiments: IB VP. Analyzed the data: ES IB VP JN HL. Contributed reagents/materials/analysis tools: ES LJ. Wrote the paper: ES LJ. Other: Supervised some of the experiments: VP. Took part in the preparation of the manuscript: VP IB. Responsible for visits of patients and control persons, including anamnesis and physical examination: JN. Performed statistical analyses: JN VP. Designed and supervised the ROS experiments: HL. Responsible for the clinical part of the study: LJ. Initiator and coordinator of the study: ES.

wikipedia.org
associated with interaction between formyl peptide receptors and the cytoskeleton. J Biol Chem 271: 15267–15271.

24. Heit B, Tavener S, Raharjo E, Kubes P (2002) An intracellular signaling hierarchy determines direction of migration in opposing chemotactic gradients. J Cell Biol 159: 91–102.

25. Beller DI, Springer TA, Schreiber RD (1982) Anti-Mac-1 selectively inhibits the mouse and human type three complement receptor. J Exp Med 156: 1000–1009.

26. Cougoule C, Wiedemann A, Lim J, Caron E (2004) Phagocytosis, an alternative model system for the study of cell adhesion. Semin Cell Dev Biol 15: 679–689.

27. Prasad K (2006) C-reactive protein (CRP)-lowering agents. Cardiovasc Drug Rev 24: 33–50.

28. Greenwood J, Steinman L, Zamvil SS (2006) Statin therapy and autoimmune disease: from protein prenylation to immunomodulation. Nat Rev Immunol 6: 330–370.

29. Yang YY, Hu CJ, Chang SM, Tai TY, Leu SJ (2004) Aspirin inhibits monocyte chemoattractant protein-1 and interleukin-8 expression in TNF-alpha stimulated human umbilical vein endothelial cells. Atherosclerosis 174: 207–213.

30. Papayianni A, Serhan CN, Brady HR (1996) Lipoxin A4 and B4 inhibit leukotriene-stimulated interactions of human neutrophils and endothelial cells. J Immunol 156: 2264–2272.

31. Pancha V, Wijdenes J, Winberg ME, Rasmussen B, Li J, et al. (2004) Differential inside-out activation of beta2-integrins by leukotriene B4 and FMLP in human neutrophils. Exp Cell Res 300: 308–319.

32. Fiorucci S, Distretti E, Mencarelli A, Morelli A, Laufor SA, et al. (2003) Evidence that 5-lipoxygenase and acetylated cyclooxygenase 2-derived eicosanoids regulate leukocyte-endothelial adherence in response to aspirin. Br J Pharmacol 139: 1351–1359.

33. Diamond MS, Springer TA (1993) A subpopulation of Mac-1 (CD11b/CD18) molecules mediates neutrophil adhesion to ICAM-1 and fibrinogen. J Cell Biol 120: 545–556.

34. Hed J, Stendahl O (1982) Differences in the ingestion mechanisms of IgG and C3b particles in phagocytosis by neutrophils. Immunology 43: 727–736.

35. Wilson A, Landquist H, Gustafsson M, Stendahl O (1996) Killing of phagocytosed Staphylococcus aureus by human neutrophils requires intracellular free calcium. J Leukoc Biol 59: 902–907.