Metal halide perovskite toxicity effects on Arabidopsis thaliana plants are caused by iodide ions

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Highlights
Natural Trichodesmium colonies collect and maintain dust within their colony core

Using synthetic particles we tested if colonies select the particles they collect

Colonies selectively collect and retain nutrient-rich over nutrient-free particles

Selective collection of particles optimize their nutrient-acquisition from dust

Hutter et al., iScience 25, 103583 January 21, 2022 © 2021 The Author(s). https://doi.org/10.1016/j.isci.2021.103583
Metal halide perovskite toxicity effects on Arabidopsis thaliana plants are caused by iodide ions

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SUMMARY

Highly efficient solar cells containing lead halide perovskites are expected to revolutionize sustainable energy production in the coming years. Perovskites are generally assumed to be toxic because of the lead (Pb), but experimental evidence to support this prediction is scarce. We tested the toxicity of the perovskite MAPbI3 (MA = CH3NH3) and several precursors in Arabidopsis thaliana plants. Both MAPbI3 and the precursor MAI hamper plant growth at concentrations above 5 mM. Lead-based precursors without iodide are only toxic above 500 μM. Iodine accumulation in Arabidopsis correlates with growth inhibition at much lower concentrations than lead. This reveals that perovskite toxicity at low concentrations is caused by iodide ions specifically, instead of lead. We calculate that toxicity thresholds for iodide, but not lead, are likely to be reached in soils upon perovskite leakage. This work stresses the importance to further understand and predict harmful effects of iodide-containing perovskites in the environment.

INTRODUCTION

Sunlight provides enough energy to fulfill the global demand, making solar cells the most promising route toward a sustainable economy. Combining solar panels with agriculture, named agri(photo)voltaics, maximizes land use, while making optimum use of the sunlight for both crops and power generation (Figure 1A). (Adeh et al., 2019) Solar cells based on both silicon and lead halide perovskites (LHPs) are considered the next-generation commercial solar cells, with potential efficiencies up to >45% (Futscher and Ehrler, 2016; Leijtens et al., 2018). However, placing such next-generation solar cells on e.g., agricultural lands raises questions about the safety of LHPs for the environment. That is, if unintentional leakage releases Pb2+ ions in the soil, this may be harmful either to plants themselves or to humans and livestock through consumption of contaminated crops (Pourrut et al., 2011). A few studies have specifically tested the toxicity of the LHP MAPbI3 (MA = CH3NH3) and its supposedly less toxic tin-based counterpart on both plants and animals (Hailegnaw et al., 2015; Babayigit et al., 2016a, 2016b; Li et al., 2020). These studies focused on the heavy metals, but the presence of halides also raises environmental concerns (Medrano-Macias et al., 2016; Incrocci et al., 2019), which has not been studied in plants to date.

In this work, we find that the iodide in MAPbI3 causes greater harm to plants than the lead. Model species Arabidopsis thaliana was used to study the effect of MAPbI3 and different perovskite precursors on the growth and development of plants. Unlike previous work on perovskite toxicity (Babayigit et al., 2016a; Li et al., 2020), we avoid acidification effects by buffering the growth media to maintain a fixed pH, so we could selectively study the effect of the perovskite and its precursors. We used controlled growth settings with a high-resolution range of salt concentrations to define the exact toxicity thresholds. We find that the presence of MAPbI3 in the growth medium starts to affect plant performance at 5 mM and becomes significant at 10 mM. This concentration (10 mM) exceeds, according to our calculations, for a practical situation where one perovskite solar cell leaks into a similar soil area of 25 cm deep. In contrast to previous works, we conclude from experiments comparing the precursors MAI and PbI2 to MABr and PbBr2, that the iodide is responsible for inhibiting Arabidopsis development, before toxicity effects of lead appear. These results show that a more rigorous assessment on the potential harmfulness of LHPs is needed and stress the importance of developing strategies to avoid halides from being released into the environment.
RESULTS AND DISCUSSION

Perovskites are fabricated from a lead halide salt, such as PbI₂, and an organic halide salt, such as MAI (Figure 1A):

\[
\text{MAI} + \text{PbI}_2 \rightarrow \text{MAPbI}_3
\]

Vice versa, decomposition of MAPbI₃ leads to the formation of its precursors PbI₂ and MAI. To assess the toxicity of LHPs, we germinated seeds of \textit{Arabidopsis thaliana} (ecotype Columbia-0) on media containing different concentrations of MAPbI₃, and several precursor salts. The media were buffered at a pH of 5.8, to avoid acidification effects (Babayigit et al., 2016a). Although seed germination is not affected, both MAPbI₃ and PbI₂ significantly inhibit plant growth at the seedling stage (depicted as rosette diameter) at concentrations >10⁻⁷ M (Figures 1B and 1C). Growth inhibition stagnated from 50⁻⁷ M, suggesting that no additional toxicity occurs beyond this concentration. To specify the level of lead perovskite toxicity, we grew plants at a range of MAPbI₃ concentrations around 10⁻⁷ M. As shown in Figure 1D, growth is significantly inhibited by concentrations >5⁻⁷ M. In addition, plants appeared bleached when treated with higher concentrations of MAPbI₃ or PbI₂ (Figure 1B), which is supported by reduced chlorophyll levels in plants treated with over 10⁻⁷ M of MAPbI₃ (Figure 1E).

The Pb²⁺ oxidation state is toxic to plants (Pourrut et al., 2011; Babayigit et al., 2016b; Gupta et al., 2020). However, at concentrations for which we observed toxicity for both MAPbI₃ and PbI₂ (Figure 1C), lead nitrate (Pb(NO₃)₂) and another lead halide precursor (PbBr₂) did not affect plant growth (Figures 2A and 2B). We found these lead-containing salts to significantly impede \textit{Arabidopsis} growth at concentrations >750⁻⁷ M (Figure 2C). This effect could be directly attributed to the lead, as MABr did not affect plant growth at these concentrations. Interestingly, even though growth was inhibited at high concentrations, Pb(NO₃)₂ and PbBr₂ did not cause plant bleaching similar to MAPbI₃ and PbI₂ (compare Figures 1B and 2C). The observation that MAPbI₃ and PbI₂ hamper \textit{Arabidopsis} growth at one order of magnitude lower concentrations (Figure 1) suggests that the iodide is at least in part responsible for this hampered growth. We repeated the growth experiments using the iodide precursor MAI and compared this to its bromide equivalent MABr (Figure 2D). As before, we found a significant inhibition of rosette size at 50⁻⁷ M of MAI. In contrast, \textit{Arabidopsis} growth is not affected by any concentration of MABr up to 100⁻⁷ M (Figures 2C and 2D).

Toxicity of MAPbI₃ starts at 10⁻⁷ M, and around 50⁻⁷ M for MAI (Figure 2D), which we attribute to the three-fold higher concentration of available iodide ions in the MAPbI₃ treatment. The hypothesis that the iodide is responsible for the toxicity at low MAPbI₃ concentrations is further confirmed by the absence of toxicity of MABr and PbBr₂ in the same concentration ranges (Figure 2C).

Even though iodide is considered a (micro-) nutrient (Kiferle et al., 2021), the toxicity effects of iodide at higher concentrations are poorly understood. To confirm that iodide ions build up in the plants to reach a toxic level, we measured iodine content in \textit{Arabidopsis} plants grown on different concentrations of MAPbI₃ (Figure 3A). Iodine levels increased significantly in plants with increasing MAPbI₃ concentrations. When comparing this to rosette size, we can conclude that in-plant levels >15 ng iodine/mg fresh weight caused growth inhibition in \textit{Arabidopsis}. Similar to what was shown before, low concentrations of iodide correlated with slightly induced \textit{Arabidopsis} growth (although not significant, see 1⁻¹⁷ M in Figures 1C and 1D), most probably because of its nutritional value (Kiferle et al., 2021). This trend for iodide-mediated mild growth stimulation at low concentrations and toxicity at higher concentrations is not an
Arabidopsis-specific phenotype, as it was seen before in strawberry, tomato, and various vegetable crops (Hong et al., 2009; Landini et al., 2011; Li et al., 2017).

Next, we measured Pb levels in Arabidopsis plants grown on 50 μM MAPbI3, or high (>500 μM) levels of MABr and PbBr2, see Figure 3B. As expected, Pb levels in control and MABr treated plants were extremely low or were not detectable. Plants treated with 50 μM MAPbI3 showed some accumulation, although not significant. The rosette size data shows that growth is strongly impaired in these plants when compared to the control treatment. However, similar Pb levels (approx. 10 ng Pb/mg fresh weight) were detected in plants treated with 500 or 750 μM PbBr2, but do not inhibit rosette size as severely in those cases. Only after applying 1000 μM PbBr2, plant growth is strongly repressed. These data support that increasing iodine levels in Arabidopsis plants treated with the LHP MAPbI3 are the main cause of toxicity and growth inhibition over increasing Pb levels. Our conclusion that the low toxicity threshold of MAPbI3 is caused by the presence of iodide, rather than lead, stresses the importance of further investigating the environmental effects of using iodide salts in solar panels.

Likely, the extent of toxicity of these halide salts in the environment will depend on the plant species, soil type, and depth to which the salts penetrate the soil in case of leakage. We estimate that a solar panel with a 400 nm thick perovskite layer contains 0.26 μmol of Pb2⁺ and 0.79 μmol of iodide (I⁻) per cm². If the full panel would leak and be homogeneously distributed over a water column of the same area and 10 cm deep, this would yield concentrations of 26 μM (<5 mgkg⁻¹ soil) Pb2⁺ and 79 μM iodide. Hence, the expected maximum concentration of lead in case of leakage is much lower than the concentration (>750 μM) at which it is toxic to the plant, and far below quality standards that range from 100 to 530 mgkg⁻¹ depending on the country (Li et al., 2020). The concentration of iodide (79 μM), on the other hand, exceeds...
the toxicity threshold of 50 μM, so that the release of iodide into the environment upon leakage of a perovskite solar panel may affect plant growth and fitness.

To conclude, although the perovskite community so far has mainly discussed the potential toxicity of lead (Hailegnaw et al., 2015; Babayigit et al., 2016b; Slavney et al., 2016; Li et al., 2020), we found that iodide is toxic to Arabidopsis plants at one order of magnitude lower concentrations. We further find that lead is only significantly toxic at levels far above those expected for a solar module failure. Our observations stress the importance of getting a more complete picture on the potential harmfulness of solar panels that contain LHPs, especially if these are placed on agricultural lands, and to develop strategies to prevent the release of halides into the environment. The latter should primarily be realized by careful encapsulation of the solar cells to prevent leakage, or in case of unforeseen calamities, by considering halide-tolerant plants that accumulate leaked halides in a phytoremediation approach to clean the soil (Incrocci et al., 2019).

Limitations of the study

In this study we have grown Arabidopsis seedlings in vitro, to be able to precisely apply treatments with perovskite salts. This helped to find a clear threshold for toxicity, especially iodide. In addition, we used a buffered solution to exclude effects from perovskite-induced changes in pH. However, in natural environments, where perovskites might leak from solar panels into the soil, toxicity of these compounds not only depends on their concentrations, but also on the soil type, presence of microbiome, and other environmental factors such as water availability, pH, and the plant species. Additional experiments in natural settings would be required to test perovskite toxicity levels in natural conditions and for different plant species.

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SUPPLEMENTAL INFORMATION

Supplemental information can be found online at https://doi.org/10.1016/j.isci.2021.103583.

ACKNOWLEDGMENTS

We thank Marielle Schreuder for technical assistance. The Dutch Research Council (NWO) is acknowledged for funding: OCENW.XS.033 to CMMG.

AUTHOR CONTRIBUTIONS

Conceptualization: E.M.H, C.T., B.E. and C.M.M.G.; Investigation: E.M.H., R.S. and C.M.M.G.; Data analysis: E.M.H, R.S. and C.M.M.G.; Writing – Original draft: E.M.H, C.T., B.E. and C.M.M.G.; Writing – Review & Editing: E.M.H. and C.M.M.G.

DECLARATION OF INTERESTS

The authors declare no competing interests.

Received: January 6, 2021
Revised: October 29, 2021
Accepted: December 6, 2021
Published: January 21, 2022

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STAR★METHODS

KEY RESOURCES TABLE

| REAGENT or RESOURCE | SOURCE | IDENTIFIER |
|---------------------|--------|------------|
| Chemicals, peptides, and recombinant proteins | N/A | N/A |
| Commercial bleach | N/A | N/A |
| SDS | VWR life science | Cat#0227 |
| Murashige & Skoog inc. vitamins | Duchefa Biochemie | Cat#M0222.0050 |
| Daishin agar | Duchefa Biochemie | Cat#D1004.1000 |
| Potassium hydroxide (KOH) | Duchefa Biochemie | Cat#P0517.1000 |
| Dimethyl sulfoxide (DMSO) | Sigma-Aldrich (now Merck) | Cat#41640 |
| Lead iodide (PbI₂) | TCI chemicals | Cat#L0279 |
| Methylammonium iodide (MAI, methylamine hydroiodide) | TCI chemicals | Cat#M2556 |
| Methylammonium bromide (MABr, methylamine hydrobromide) | TCI chemicals | Cat#M2589 |
| Lead bromide (PbBr₂) | Sigma-Aldrich (now Merck) | Cat#915696 |
| Lead nitrate (Pb(NO₃)₂) | Sigma-Aldrich (now Merck) | Cat#467790 |

Experimental models: Organisms/strains

Arabidopsis thaliana ecotype Col-0 | N/A | N/A |

Software and algorithms

Fiji programme | Schindelin et al. 2012 | https://imagej.net/software/fiji/ |
MVApp | Julkowska et al. 2019 | https://mvapp.kaust.edu.sa |

RESOURCE AVAILABILITY

Lead contact

Further information and requests for resources and reagents should be directed to and will be fulfilled by the lead contact, Charlotte M. M. Gommers (charlotte.gommers@wur.nl).

Materials availability

This study did not generate new unique reagents.

Data and code availability

● Data: The data reported in this paper will be shared by the lead contact upon request.
● Code: This paper does not report original code.
● Any additional information required to reanalyze the data reported in this paper is available from the lead contact upon request.

METHOD DETAILS

Plant material and growth conditions

Arabidopsis thaliana ecotype Columbia-0 was used for all experiments. Seeds were surface-sterilized (20% bleach, 0.5%SDS), rinsed with sterile water and sown on half-strength Murashige & Skoog medium including vitamins (Duchefa Biochemie), containing 0.1% MES monohydrate buffer (Duchefa Biochemie) and 1% v/w Daishin agar (Duchefa Biochemie). Salts were added from a 1 mM stock in the desired concentration before autoclaving. The pH of all media was set at 5.8 using 0.1 N KOH to prevent any harmful acidification effects as described by Babayigit et al. (Babayigit et al., 2016a) Seeds were sown on petri dishes (9 cm diameter) containing 20 mL medium, stratified (4°C, dark) for 4 days to synchronize germination, and afterwards placed in a climate chamber (16 hours light period, 22°C, photosynthetic active radiation 150 µmol photons/m²/s).
**Perovskite solutions**

MAPbI₃ was prepared by mixing and grinding the dry precursor powders in a stoichiometric ratio (0.4 millimole of MAI and 0.4 millimole of PbI₂), using a mortar and pestle until a black powder was obtained. This was one under airtight conditions, inside a glovebox. Consequently, deionized water was added to the powder. On the addition of water, the powder immediately turned yellow, indicating decomposition of the perovskite into MAI (high solubility in water) and PbI₂ (yellow, poor solubility in water). In addition, the separate precursors PbI₂, PbBr₂, MAI and MABr were dissolved in water at concentrations of 1 mM, as well as Pb(NO₃)₂ for reference experiments. The solutions for the experiments were prepared by dilution of the above stock solution. Concentrations of all salts used in this study, expressed in µM and g L⁻¹ are given in Table S1.

**Growth assays and chlorophyll quantification**

Pictures of the plants were taken after 18 days of growth (2 petri dishes per treatment, 21 plants per petri dish). Maximum rosette diameter of all plants was analyzed using FIJI software (Schindelin et al., 2012). From each petri dish, three plants were harvested, weighed, and used for chlorophyll quantification: plants were shaken in 1 mL DMSO at 65°C in darkness for 1 hour, cooled and kept at room temperature for 30 minutes. Absorption was measured at 664 and 647 nm using a SpectraMax® Plus 384 Microplate Reader (Molecular Devices). Concentration of chlorophyll A and B was calculated by: [ChlA] (mg/L) = 12.25*(A664) – 2.55*(A647); [ChlB] (mg/L) = 20.31*(A647) – 4.91*(A664) (Porra et al., 1989). Values were converted into mg per mg fresh weight.

**Element analysis**

18-day old plants were harvested, pooled (2 plants per sample) in 50ml PE tubes and kept at -80°C until sent for analysis. Iodine (I) and lead (Pb) quantification by ICP-MS was performed by UT2A (Ultra Traces Analyses Aquitaine) at Pau University in France.

**Estimated concentrations in the soil**

Assuming a density of 4.09 g cm⁻³ for MAPbI₃ (Li et al., 2020) and a thickness of 400 nm (4x10⁻⁵ cm), a solar panel contains 1.6x10⁻⁴ g perovskite per cm². With molar weight fractions of 0.05 for MA⁺, 0.33 for Pb²⁺ and 0.61 for I⁻, this translates to 8.5x10⁻⁶ g MA⁺, 5.47x10⁻⁵ g Pb²⁺ and 1 x 10⁻⁴ g I⁻ per cm² solar panel. If a full solar panel would break and leak into an equivalent area, the concentration in the soil will depend on the depth of leakage. For example, homogeneous distribution of the perovskite over the first 10 cm soil leads to concentrations of 8.5x10⁻⁷ g/cm³ MA⁺, 5.47x10⁻⁶ g/cm³ Pb²⁺ and 1 x 10⁻⁵ g/cm³ I⁻. Dividing these numbers by the respective molar weights, and considering that 1 cm³ = 1 mL, yields molarities of 26 µM MA⁺, 26 µM Pb²⁺ and 79 µM I⁻. If the soil has a density between 1 and 2 g/cm³, the concentrations are between 5 and 2.7 mg kg⁻¹ for Pb²⁺ and between 10 and 5 mg kg⁻¹ for the I⁻.

**Statistical analysis**

Statistical analyses were performed using the MVApp. (Jukowska et al., 2019). Levene’s test was used to test for equal variances. Multivariate comparisons were made with ANOVA, followed by a post-hoc Tukey test.