Dioxin and Related Compound Detection: Perspectives for Optical Monitoring

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Received: 20 April 2019; Accepted: 28 May 2019; Published: 30 May 2019

Abstract: Dioxins and related compounds are environmental xenobiotics that are dangerous to human life, due to the accumulation and persistence in the environment and in the food chain. Cancer, reproductive and developmental issues, and damage to the immune system and endocrine system are only a few examples of the impact of such substances in everyday life. For these reasons, it is fundamental to detect and monitor these molecules in biological samples. The consolidated technique for analytical evaluation is gas chromatography combined with high-resolution mass spectrometry. Nowadays, the development of mid-infrared optical components like broadband laser sources, optical frequency combs, high performance Fourier-transform infrared spectroscopy, and plasmonic sensors open the way to new techniques for detection and real time monitoring of these organic pollutants in gaseous or liquid phase, with sufficient sensitivity and selectivity, and in short time periods. In this review, we report the latest techniques for the detection of dioxins, furans and related compounds based on optical and spectroscopic methods, looking at future perspectives.

Keywords: dioxins; 2,3,7,8-TCDD; PCBs; POPs; optical detection; environmental xenobiotics; dioxin bio-monitoring; dioxin detection

1. Introduction

Dioxins are among the most toxic and persistent organic pollutants (POPs). They are principally released into the atmosphere as undesired products of various combustion and industrial processes [1] such as incineration of municipal and medical solid waste, backyard waste burning, paper and wood pulp chlorine bleaching, coal fired power plants, and also from natural processes like forest fires. Furthermore, these pollutants also occur as contaminants in several pesticides, herbicides, and fungicides [2].

When accidently released in the various environmental matrices, like soil and water, dioxins accumulate in plants and animal tissues, until they reach human tissues where accumulate at higher and higher concentrations, mainly in the fatty tissues, through food chain biomagnifications processes.

These halogenated organic POPs mainly belong to three families of molecules, i.e., polychlorinated dibenzo-p-dioxins (PCDDs), polychlorinated dibenzofurans (PCDFs) and polychlorinated biphenyls (PCBs); their molecular structures are presented in Figure 1.

All these compounds together with Polychlorinated Diphenyl Ethers (PCDEs) belong to the main family of Polyhalogenated Aromatic Compounds (PHAs). PCBs and PCDEs are industrial compounds or by-products, mainly present as impurities in chlorophenol preparations. These two classes of
POPs have been detected in the environment and especially in chemical-waste dumpsites. PCBs, in particular, are synthetic PHAs used as industrial reagents in the past. Because they persist for long times in both the environmental matrices and in tissues of living organisms, their production and utilization were discontinued.

**Figure 1.** General molecular structures of chlorinated dioxins (PCDDs), furans (PCDFs) and biphenyls (PCBs).

PCDDs and PCDFs have two benzene rings connected by one (furans) or two (dioxins) oxygen atoms. The two benzene rings can bind from one to eight chlorine atoms, generating a wide family of congeners (75 PCDD congeners and 135 PCDF congeners) whose molecular reactivity toward cellular targets can change dramatically, determining different levels of toxicity. Within these 210 congeners, the 2,3,7,8-tetrachlorinated dibenzo dioxins (TCDD) species have been classified as the most toxic. Toxicity drastically decreases when non-lateral chlorines are present or when lateral chlorines are removed from the two aromatic rings [3]. Laterally chlorinated dioxins and especially TCDD show the highest affinity toward the AhR (Aryl hydrocarbon Receptor) receptor, a signal transducer protein which is responsible of the biological effects cascade following the xenobiotic–receptor interaction [4–6].

There are seven laterally chlorinated PCDDs and ten laterally chlorinated PCDFs. The toxicity equivalent factor (TEF) [7] expresses the toxicity of dioxins, furans and PCBs in terms of the most toxic congeners, i.e., the 2,3,7,8-TCDD and 1,2,3,7,8-pentachlorodibenzo-p-dioxin, whose TEF value has been set to 1. It must be taken into account that dioxins are extremely toxic molecules; indeed, they can cause cancer, reproductive and developmental aberrations, immune system damages, and can deeply interfere with the endocrine system [8–11].

In this regard, it is important to emphasize that the half-life of TCDD in humans is very long; it has been estimated to be in the range between 7.1 [12] and 11.3 years [13].

Similar to dioxins and furans, the number of chlorine atoms (from 1 to 10) and their positions in PCBs give rise to 209 congeners (see Figure 1).

In particular, ortho-PCBs are characterized by two chlorine atoms in ortho positions, while mono-ortho-PCBs (i.e., PCBs 105, 114, 118, 123, 156, 157, 167, and 189) are characterized by one chlorine atom in ortho position. On the other hand, in non-ortho-PCBs (i.e., PCBs 77, 81, 126, and 169) non-ortho positions are occupied by chlorine atoms. Both non-ortho and mono-ortho PCBs have coplanar structures that make them more toxic since they bind with higher affinity the AhR receptor. For this reason non-ortho and mono-ortho PCBs have been classified with different values of TEF [14], likewise dioxins and furans.
In particular, PCB 77 (3,3′,4,4′ tetrachlorobiphenyl) is able, like TCDD, to bind strongly to the AhR receptor leading to the activation of the expression of genes involved in polycyclic aromatic hydrocarbon metabolism and detoxification [6,15]. Prolonged exposure to PCBs leads to immune system suppression increasing the risk of diseases developing. Both non-ortho and mono-ortho PCBs congeners are cancer promoters and enhance the effects of other carcinogens. PCBs are endocrine disruptors, altering thyroid and reproductive function in males and females. These endocrine alterations increase the risk of developing of other cardiovascular diseases and diabetes [16].

Due to their high toxicity and environmental persistence, many efforts for real-time monitoring of these organic pollutants from the principal emission sources have been recently realized [17–21]. Unfortunately, the objective of constant monitoring of the potential sources of emission and the possibility of recognizing each single congener in a complex mixture have not yet been fulfilled.

Moreover, it is also very important to detect dioxins and related compounds in environmental matrices such as soil, water, air and so on. PHAs are ubiquitous in the environment, they have been found in aqueous (mainly surface water, plant oils and petroleum products), air and solid samples such as sediments, soils and wastewater sludge. In this regard, very often it is of fundamental importance to isolate PHAs not only from environmental matrices, but also from biological tissues and fluids, by extraction and clean-up procedures. The clean-up step allows eliminating the matrix effects and pre-concentrates the analytes for instrumental quantitative detection. In the last years several techniques were developed and implemented for environmental samples clean-up, employing different kind of adsorbents materials such as silica gel and alumina, mesoporous organic silica, mesoporous silica nanoparticles, different nanoparticles and nanotubes functionalized at their surface, and molecularly imprinted polymers for more efficient adsorption capacity [22,23].

Then, the analytes must be extracted by using a suitable solvent, in order to proceed with the appropriate instrumental detection.

In the last years, several instrumental automated extraction techniques for rapid sample processing have been developed. Some of these techniques are supercritical fluid extraction (SFE), microwave-assisted extraction (MAE), pressurized liquid extraction [24], and solid-phase extraction (SPE) [25–27].

Due to their high resolution, high sensitivity and selectivity liquid and gas chromatographies (LC and GC) are the most commonly employed analytical methods for the qualitative and quantitative determination of POPs in environmental and biological samples. In particular, High Performance Liquid Chromatography (HPLC) coupled with UV/fluorescence detection, and Gas Chromatography coupled to Mass Spectrometry (GC/MS) has shown to be the best technique amongst others. GC/MS is a reference analytical method because provides the advantages of congener identification using both retention time and mass spectrum [28–30]. Furthermore, GC/MS provides the required high sensitivity for quantification in the Selected Ion Monitoring (SIM) mode.

Even though Liquid Chromatography coupled to Mass Spectrometry (LC/MS) offers similar advantages, it does not supply a specific interface appropriate for the separation of all the PHAs at the same time [28,31].

The possibility to use new generation materials for the separation and clean-up and to automate instrumental extraction of pollutants from environmental and biologic samples will allow new optical detection methods to become more selective and sensitive and to achieve real time and fast monitoring of chlorinated organic compounds.

2. Optical Monitoring of Dioxins and Related Compounds, a Screening of Old and New Mid-IR Light Sources and Techniques

Optical monitoring offers many advantages with respect to the cited techniques like chromatography or high-resolution mass spectrometry in terms of costs, destructive sampling and fast detection. Optical techniques can be divided into two main classes: (1) those which employ wide
wavelength range sources and (2) those which use narrow wavelength range sources. Let us discuss
them separately.

2.1. Wide Wavelength Range Sources

The use of Mid-IR radiation to detect PCDD/Fs and related compounds allows discriminating
between congeners, since their own fingerprints in this spectral region characterize these molecules.

The most frequent use of the absorption spectroscopy in Mid-IR lies in the identification of
substances through their characteristic molecular vibrations. Furans and dioxins, and in general,
molecules with a similar structure, are not in gaseous phase at room temperature, and high temperatures
(more than 550 °C) are needed to perform gaseous spectroscopy. However, in order to work at room
temperature, it is possible to carry out spectroscopy in liquid phase, dissolving the target molecules in
a Mid-IR transparent solution [18,19]. In both gaseous and liquid spectroscopy, the detected absorption
peaks are tens of cm⁻¹ wide. In order to detect these bands, techniques and components spanning a
wide frequency range are needed. In this framework, a consolidated technique is Fourier-Transform
Infrared Spectroscopy (FT-IR) [32]. In this scheme, sketched in Figure 2, using a broadband source,
typically a lamp, an interferometric pattern is recorded from the light interacting with the sample.
By using the Fourier Transform approach, it is possible to reconstruct the absorption spectrum of the
substance under analysis.

**Figure 2.** Principles of the Fourier-Transform Infrared Spectroscopy (FT-IR).

The resolution of commercial instruments is better than 1 cm⁻¹. This technique has been used for
measuring dioxins, furans and related chlorinated organic molecules. In Figures 3 and 4, we report
the infrared spectra of 13 of the 17 most toxic congeners of dioxins dissolved in carbon tetrachloride
(CCl4) [19]. In Tables 1 and 2, the characteristic vibrational frequencies of PCDDs and PCDFs, obtained
by FT-IR in condensed phase [19] and other techniques, i.e., the GC/MI/FT-IR [33] and Vapor-Phase
FT-IR [34], are presented and compared. In Tables 1 and 2, each frequency range is characterized by
different IR active modes that represent the fingerprints of each single PCDD/Fs congener.

These vibrational frequencies are influenced by chlorine substitution patterns and thus allow
us to discriminate between different molecules and congeners, and also to analyze mixtures of these
compounds [19].

Although this method is robust and consolidated, FT-IR performance is limited by the brilliance
of the light source used.
**Figure 3.** FT-IR spectra of PCDDs and dibenzo-p-dioxin (D-p-D). Reprinted with permission from [19], copyright 2014.
Figure 4. FT-IR spectra of PCDFs and dibenzofuran (DBF). Reprinted with permission from [19], copyright 2014.
Table 1. Principal FT-IR bands of PCDDs. Reprinted with permission from [19], copyright 2014.

| PCDD Congener | Mode | Frequency Range (1020–1205 and 1265–1510 cm⁻¹) GC/MC/FT-IR [33] | Frequency Range (1020–1205 and 1265–1510 cm⁻¹) Vapor-Phase FT-IR [34] | Frequency Range (1020–1205 and 1265–1510 cm⁻¹) Condensed-Phase FT-IR [19] |
|---------------|------|--------------------------------------------------|--------------------------------------------------|--------------------------------------------------|
| dibenzo-p-dioxin | C=C aromatic ring skeletal vibrations | 1489 | 1489 | 1489 |
| | C-O-C asymmetric stretch | 1390 | 1390 | 1390 |
| | C-C-C tri-ring bending | 1290 | 1287 | 1295 |
| | Ring breathing | 1117 | 1117 | 1197 |
| 2,3,7,8-TCDD | C=C aromatic ring skeletal vibrations | 1495 | 1495 | 1495 |
| | C-O-C asymmetric stretch | 1391 | 1384 | 1391 |
| | C-C-C tri-ring bending | 1176 | 1173 | 1197 |
| | Ring breathing | 1114 | 1114 | 1197 |
| 1,2,3,7,8-PeCDD | C=C aromatic ring skeletal vibrations | 1493 | 1493 | 1493 |
| | C-O-C asymmetric stretch | 1399 | 1394 | 1399 |
| | C-C-C tri-ring bending | 1319 | 1314 | 1319 |
| | Ring breathing | 1113 | 1108 | 1113 |
| 1,2,3,7,8,9-HxCDD | C=C aromatic ring skeletal vibrations | 1480 | 1480 | 1480 |
| | C-O-C asymmetric stretch | 1333 | 1328 | 1333 |
| | C-C-C tri-ring bending | 1147 | 1147 | 1147 |
### Table 1. Cont.

| PCDD Congener | Mode | Frequency Range (1020–1205 and 1265–1510 cm\(^{-1}\)) | Frequency Range (1020–1205 and 1265–1510 cm\(^{-1}\)) | Frequency Range (1020–1205 and 1265–1510 cm\(^{-1}\)) |
|---------------|------|--------------------------------------------------|--------------------------------------------------|--------------------------------------------------|
|               |      | GC/MI/FT-IR \[33\] | Vapor-Phase FT-IR \[34\] | Condensed-Phase FT-IR \[19\] |
| 1,2,3,6,7,8-HxCDD | C=C aromatic ring skeletal vibrations | 1481 | 1455 | 1400 | 1392 | 1395 |
|               | C-O-C asymmetric stretch | 1333 | 1328 | 1318 | 1310 | 1312 |
|               | C-C-C tri-ring bending | 1147 | 1150 | 1143 | 1142 | 1140 |
| 1,2,3,4,6,7,8-HpCDD | C=C aromatic ring skeletal vibrations | 1480 | 1457 | 1451 | 1452 | 1447 |
|               | C-O-C asymmetric stretch | 1334 | 1329 | 1303 | 1302 | 1301 |
|               | C-C-C tri-ring bending | 1148 | 1153 | 1144 | 1143 | 1142 |
| OCDD | C=C aromatic ring skeletal vibrations | 1436 | 1424 | 1429 | 1419 | 1402 |
|               | | | | | | 1399 |

### Table 2. Principal FT-IR bands of PCDFs. Reprinted with permission from \[19\], copyright 2014.

| PCDF Congener | Mode | Frequency Range (1020–1205 and 1265–1510 cm\(^{-1}\)) | Frequency Range (1020–1205 and 1265–1510 cm\(^{-1}\)) |
|---------------|------|--------------------------------------------------|--------------------------------------------------|
|               |      | GC/MI/FT-IR \[33\] | Condensed-Phase FT-IR \[19\] |
| dibenzofuran | C=C aromatic ring skeletal vibrations | 1473 | 1465 | 1454 |
|               | C-O-C asymmetric stretch | 1320 | 1283 | 1289 |
|               | C-C-C tri-ring bending | 1189 | 1196 | 1193 |
|               | Ring breathing | 1112 | 1110 | 1102 |
| 2,3,7,8-TCDF | C=C aromatic ring skeletal vibrations | 1443 | 1439 | 1402 | 1398 | 1395 |
|               | C-O-C asymmetric stretch | 1289 | 1287 | 1308 | 1307 | 1305 |
|               | C-H in plane deformations | 1276 | 1292 | 1295 | 1297 | 1299 |
|               | C-C-C tri-ring bending | 1191 | 1198 | 1195 | 1192 | 1190 |
|               | Ring breathing | 1109 | 1107 | 1104 | 1102 | 1100 | 1099 | 1096 | 1093 | 1090 | 1087 | 1084 | 1081 | 1078 | 1075 | 1072 | 1069 | 1066 | 1063 | 1060 | 1057 | 1054 | 1051 | 1048 | 1045 | 1042 | 1039 | 1036 | 1033 | 1030 | 1027 | 1024 | 1021 | 1018 | 1015 | 1012 | 1009 | 1006 | 1003 | 1000 | 997 | 994 | 991 | 988 | 985 | 982 | 979 | 976 | 973 | 970 | 967 | 964 | 961 | 958 | 955 | 952 | 949 | 946 | 943 | 940 | 937 | 934 | 931 | 928 | 925 | 922 | 919 | 916 | 913 | 910 | 907 | 904 | 901 | 898 | 895 | 892 | 889 | 886 | 883 | 880 | 877 | 874 | 871 | 868 | 865 | 862 | 859 | 856 | 853 | 850 | 847 | 844 | 841 | 838 | 835 | 832 | 829 | 826 | 823 | 820 | 817 | 814 | 811 | 808 | 805 | 802 | 799 | 796 | 793 | 790 | 787 | 784 | 781 | 778 | 775 | 772 | 769 | 766 | 763 | 760 | 757 | 754 | 751 | 748 | 745 | 742 | 739 | 736 | 733 | 730 | 727 | 724 | 721 | 718 | 715 | 712 | 709 | 706 | 703 | 700 | 697 | 694 | 691 | 688 | 685 | 682 | 679 | 676 | 673 | 670 | 667 | 664 | 661 | 658 | 655 | 652 | 649 | 646 | 643 | 640 | 637 | 634 | 631 | 628 | 625 | 622 | 619 | 616 | 613 | 610 | 607 | 604 | 601 | 598 | 595 | 592 | 589 | 586 | 583 | 580 | 577 | 574 | 571 | 568 | 565 | 562 | 559 | 556 | 553 | 550 | 547 | 544 | 541 | 538 | 535 | 532 | 529 | 526 | 523 | 520 | 517 | 514 | 511 | 508 | 505 | 502 | 499 | 496 | 493 | 490 | 487 | 484 | 481 | 478 | 475 | 472 | 469 | 466 | 463 | 460 | 457 | 454 | 451 | 448 | 445 | 442 | 439 | 436 | 433 | 430 | 427 | 424 | 421 | 418 | 415 | 412 | 409 | 406 | 403 | 399 | 396 | 393 | 390 | 387 | 384 | 381 | 378 | 375 | 372 | 369 | 366 | 363 | 360 | 357 | 354 | 351 | 348 | 345 | 342 | 339 | 336 | 333 | 330 | 327 | 324 | 321 | 318 | 315 | 312 | 309 | 306 | 303 | 299 | 296 | 293 | 290 | 287 | 284 | 281 | 278 | 275 | 272 | 269 | 266 | 263 | 260 | 257 | 254 | 251 | 248 | 245 | 242 | 239 | 236 | 233 | 230 | 227 | 224 | 221 | 218 | 215 | 212 | 209 | 206 | 203 | 200 | 197 | 194 | 191 | 188 | 185 | 182 | 179 | 176 | 173 | 170 | 167 | 164 | 161 | 158 | 155 | 152 | 149 | 146 | 143 | 140 | 137 | 134 | 131 | 128 | 125 | 122 | 119 | 116 | 113 | 110 | 107 | 104 | 101 | 98 | 95 | 92 | 89 | 86 | 83 | 80 | 77 | 74 | 71 | 68 | 65 | 62 | 59 | 56 | 53 | 50 | 47 | 44 | 41 | 38 | 35 | 32 | 29 | 26 | 23 | 20 | 17 | 14 | 11 | 8 | 5 | 2 | 1 | 80 | 77 | 74 | 71 | 68 | 65 | 62 | 59 | 56 | 53 | 50 | 47 | 44 | 41 | 38 | 35 | 32 | 29 | 26 | 23 | 20 | 17 | 14 | 11 | 8 | 5 | 2 | 1 |
Table 2. Cont.

| PCDF Congener  | Mode                                      | Frequency Range (1020–1205 and 1265–1510 cm\(^{-1}\)) | Frequency Range (1020–1205 and 1265–1510 cm\(^{-1}\)) |
|----------------|-------------------------------------------|------------------------------------------------------|------------------------------------------------------|
|                | C=C aromatic ring skeletal vibrations      | 1460 1455 1423                                       | 1427 1423                                             |
| 1,2,3,7,8-PeCDF| C-O-C asymmetric stretch                   | 1382 1284                                           | 1356 1301 1283                                       |
|                | C-C-C tri-ring bending                     | 1138                                                | 1136                                                 |
|                | Ring breathing                             | 1103                                                | 1100                                                 |
|                | C=C aromatic ring skeletal vibrations      | 1455 1429 1408                                       | 1451 1425 1404                                       |
| 2,3,4,7,8-PeCDF| C-O-C asymmetric stretch                   | 1384 1381 1287                                       | 1379 1374 1363                                       |
|                | C-O-C asymmetric stretch                   | 1162                                                | 1191 1159                                            |
|                | C=C aromatic ring skeletal vibrations      | 1415 1403                                           | 1452 1412                                             |
| 1,2,3,4,7,8-HxCDF| C-O-C asymmetric stretch                  | 1361                                                | 1362 1357 1345                                       |
|                | C-H in plane deformations                  | 1277                                                | 1276                                                 |
|                | Ring breathing                             | 1108                                                | 1105                                                 |
|                | C=C aromatic ring skeletal vibrations      | 1294                                                | 1296                                                 |
| 1,2,3,7,8,9-HxCDF| C-H in plane deformations                 | 1273                                                | 1272                                                 |
|                | C-C-C tri-ring bending                     | 1146 1138                                           | 1143 1136                                            |
|                | Ring breathing                             | 1289                                                |                                                      |
|                | C=C aromatic ring skeletal vibrations      | 1387 1369 1361                                      | 1384 1366 1361                                       |
| 1,2,3,7,8,9-HpCDF| C-O-C asymmetric stretch                  | 1294                                                | 1296                                                 |
|                | C-H in plane deformations                  | 1273                                                | 1272                                                 |
|                | C-C-C tri-ring bending                     | 1146 1138                                           | 1143 1136                                            |
|                | Ring breathing                             | 1381 1371 1352 1347                                 | 1376 1365 1356 1346 1282                             |
|                | C-O-C asymmetric stretch                   | 1269                                                | 1269 1266                                            |
|                | C-H in plane deformations                  | 1182 1142                                           | 1181 1141                                            |
|                | C-C-C tri-ring bending                     | 1084                                                |                                                      |
In the last decades the development of powerful infrared sources like Interband Cascade Lasers (ICL), Quantum Cascade Laser (QCL), QCL combs, optical frequency combs, Non-Linear sources [35–41] paved the way for the improvement of the performances of classical techniques, or innovative detection scheme. In particular, these new sources could be very suitable for detecting complex organic molecules (as dioxins, furans, hydrocarbons, proteins) which present absorption spectra in the mid-infrared with absorption profiles width of ten or more wavenumbers. For simplicity, in the following, we refer to molecules with these characteristics as “complex molecules” [42].

Typically, narrow laser sources are used for spectroscopy of simple molecules in gas phase, like CO2, H2S, CO, HF with a large variety of experimental setups [43–49].

In the last ten years, the availability of broadband tunable QCLs in the range 4–12 \( \mu \)m enabled the development of optical schemes for measuring complex molecules. These sources, based on a moving external grating, which closes the laser cavity, allow wavelength selection and high gain for the active media on a very large frequency range as sketched in Figure 5.

![EXTERNAL-CAVITY QCL](image)

**Figure 5.** Spectroscopic set-up based on a broadband tunable QCL. By moving a grating, it is possible to select the operation wavelength on a very large frequency range.

The application of such schemes spans from health to security, from imaging to environmental monitoring [50–55].

Using a couple of broadly tunable external cavity QCLs (EC-QCLs), the detection of furans and dioxins dissolved in CCl\(_{4}\) was demonstrated [18] in the range 1205–1310 cm\(^{-1}\) and 1335–1590 cm\(^{-1}\). The measurement time spanned from few minutes to 15 minutes. Nowadays, the technological development of this kind of source allows the same measurement using only one laser frame equipped with 3 or 4 active media [56] with tuning performances up to 1000 cm\(^{-1}\), reaching speeds up to...
thousands cm$^{-1}$/s in particular cases. This enables very fast measurements on very large frequency ranges, with frequency resolution strongly dependent on the acquisition system. Generally, a fast scan corresponds to low frequency resolution. When using such systems for liquid spectroscopy, the accuracy of measurements depends on many parameters, and above all on the sample itself. For example in the biomedical field, Lendl and co-workers successfully used EC-QCLs for detection of glucose and lactate in aqueous phase, showing the usability of such laser sources for in-vivo and in-vitro bio fluid solutions [51,57,58].

EC-QCLs have already been extensively used in biological and fundamental chemistry studies involving peptide, breath test, kinetics reactions and DNA research [59–61].

In addiction such lasers, due to their high optical power, could be used for infrared imaging with tens of different approaches: Chemical Imaging, Photo-thermal imaging, holography, microscopy, etc. [62–66].

However, EC-QCLs cannot be used when it is necessary to observe and resolve fine absorption structures, narrower than the laser line (which is typically at least tens of MHz), or when the scan rate is still too slow compared to the experimental time scale.

A different approach in broadband spectroscopy is provided by Optical Frequency Combs (OFCs) laser systems. Two main features of such lasers are the characteristic equally spaced comb spectrum and the very narrow linewidth. Indeed they are largely used for measuring absorptions of simple molecules in the visible and near infrared using different schemes: Comb assisted Spectroscopy, Direct Comb Spectroscopy, Dual Comb Spectroscopy [35,67–71].

In direct comb spectroscopy, as shown in Figure 6, each tooth interacts with the sample and the absorption is measured tooth by tooth, using a dispersive system on a single detector or a 1-D or 2-D detector arrays. This technique enables the reconstruction of fine structure absorption profiles with large accuracy. However, to apply this approach to organic molecules, the comb spectrum has to be translated in the Mid-IR via Non-linear processes.

Recently, Dual Comb Spectroscopy is proving to be a very powerful technique. It is based, as sketched in Figure 7, on the detection of the interference between two combs with slightly different tooth spacing in the frequency-domain. The detected heterodyne beat signal consists of a comb in the Radio-Frequency (RF) domain, so that the absorption signal is translated from the optical frequencies to the “more easy to measure” RF spectrum. From the Fourier transform of the acquired signal it is possible to reconstruct entire absorption bands, like in FT spectroscopy but without moving components, with the frequency resolution and accuracy of OFCs. Until recently, this type of technique was limited to the spectral range covered by OFCs, typically in the Near IR. However, the invention of QCL-combs pushed the development of such technique also in the mid-IR with some commercial
instrument available on the market. The strong point of Dual Comb Spectroscopy is the combination of accuracy, very fast measurement time, and high brilliance of the source [70]. This technique was successfully demonstrated in a protein reaction study in the region around 8 µm [72], thus paving the way for the non-invasive analysis in biological context.

Figure 7. Principles of Dual-Comb Spectroscopy. Adapted with permission from [70], copyright 2014.

In Table 3, we report a summary of the described optical detection techniques in the 1000–1500 cm\(^{-1}\) region. The first two techniques have already been used to analyze PCDD/Fs [18], while the last two techniques have the potentiality to be used for detecting dioxins and furans, but they have not yet been applied for this purpose. Their potentiality is demonstrated by the fact that these techniques were used to measure organic molecules [56,72] with similar absorption characteristics. The most performing technique is the dual comb spectroscopy based on QCL combs. In this case, the price to pay is a limited spectral coverage and a very high cost. The cheapest instrument is the consolidated FT-IR spectrometer, but the low optical power represents a limit for analysis in thick samples or cells (hundreds microns). The EC-QCLs, due to their high power, are suitable for imaging, spectroscopy in longer measurement cells (few millimeters in liquid phase), and for slow measurements.

Table 3. Summary of the different optical techniques described in Section 2.1.

| Technique                        | Spectral Range cm\(^{-1}\) | Molecule                        | Measurement Time | Frequency Accuracy (cm\(^{-1}\)) | Optical Power/Wavelength | Cost  | Refs |
|----------------------------------|----------------------------|---------------------------------|------------------|-------------------------------|--------------------------|-------|------|
| FT-IR                            | 1020–1510                  | dioxins and furans              | <60 s            | 0.9                           | <1 µW                    | Low   | [19] |
| ECDL-QCL                         | 1205–1310 and 1335–1590    | dioxins and furans              | 4 min (first range); 14 min (second range) | 0.1                        | Tens/hundreds of mW     | Medium | [18] |
| Rapid scan QCL                   | 860–1100                   | ethylene, propene, 1-butene, 2-butene, 1,3-butadiene, methanol | down to 3 ms      | 0.35                          | ≤1 W                     | High   | [56] |
| Dual Comb based IR spectrometer  | 1180–1250                  | Protein in Halobacterium salinarum | 1 µs             | 0.3 *                        | Typically between 1 µW and 1 mW | High   | [72] |

* The target of this paper was fast reaction, so for different experiments the accuracy could reach the order of 3 \( \times 10^{-4} \) cm\(^{-1}\) (10 MHz).

2.2. Narrow Wavelength Range Sources

Mid-IR Laser technology is nowadays gaining interest due to new device development. However, sometimes chemical analysis and biological assays need a fast response, not necessarily quantitative
but at least indicative of the presence of a toxic molecule. Therefore, a class of sensors characterized by
cost-effectiveness, ready for real-time and on-site analysis is highly desirable. In many cases, sensors
might be used for the determination of dioxins and dioxin like-PCBs throughout the environment and
the food chain, involving various types of specimens such as water, air, soil, food, animal tissues and
so on. So far, we compared different techniques based on wide spectrum sources. However, there are
other classes of optical sensors based on lasers with limited tunability (a few nm at most) and possibly
cheaper. One of them takes advantage of plasmonic resonances, and is able to detect complex organic
molecules [73–78]. These sensors would deserve a broader discussion, so we refer also to other review
articles and books present in the literature [73,79].

Basically, the interaction between the target molecule and the structure (nanoparticles,
nanomaterial, fiber sensors), provided with plasmonic resonances, produces a relevant effect in
absorption, fluorescence enhancement, or frequency shifting of a laser beam. In the latter case, when
the molecule is on the plasmonic structure, the change of physical conditions at the interface produces
some changes in the plasmon coupling, namely, an easily detectable resonance shift. In this way it is
possible to obtain a very sensitive sensor, tailored onto a specific target molecule or molecular class.
In most cases, the plasmonic sensor needs to be functionalized to enable the sticking of the molecule to
the structure, so making the detection possible. A scheme of a kind of plasmonic sensor is reported in
Figure 8.

Figure 8. Sketch of a plasmonic sensor. In this particular case the light is used to “read” the sensor.
Molecule “detection” is performed by means of the interaction between the sample and a material
supporting the surface plasmons.

In the literature there are some examples of plasmonic sensors for measuring dioxins, dioxin
precursors, polychlorinated biphenyls and atrazine [80,81]. The strong point of these sensors is the
very high selectivity and sensitivity [81], down to concentrations of the order of ng/mL. In some cases,
single molecule detection could be achieved. It is fundamental specifying that this kind of plasmonic
sensor detects the molecule sticking on the surface, so the chemical selectivity is due essentially to
the selectivity of the functionalized material. For example, if this material is able to capture all the
molecules of dioxin and furan families, the sensor could detect also a single molecule but it would
not discriminate between the most toxic 2,3,7,8 TCDD and the non-toxic DBF. For evident reasons,
plasmonic sensors could suffer polluting phenomena, so they may have to be cleaned or replaced.
This could not be a real problem, as using some classes of plasmonic structure or functionalizing
materials, plasmonic sensors could be very cheap.

Similar mechanisms of operation are common to other kind of sensors based on Surface-Enhanced
Raman Scattering (SERS), [82–87], fluorescence quenching and enhancement [88] and surface photo
voltage [89]. Raman spectroscopy is a well-established technique, based on the inelastic scattering of
light. Starting from a single-frequency laser excitation it is possible to get from the target molecule one
or more emissions at lower (Stokes) or higher (anti-Stokes) frequencies. The frequency differences,
with respect to the pumping laser, are typical of each target, so allowing selectivity. Usually the Raman signal deriving from a single layer of molecules is very low. When plasmon resonance is involved, the effect rises by a factor up to 108, and this allows single molecule detection. For many years, the poor reproducibility of the metal coatings needed for SERS limited the applications of this technique. Nevertheless, since a couple of decades ago, this technological issue has been overcome and applications became possible. In the literature, it is possible to find examples of detection of dioxins [82] and of other molecules, like pesticides [90].

SERS is one of the most employed spectroscopic techniques for ultrasensitive detection in chemistry and biology. One of the most important requirements in this field is the design of advanced materials capable of generating high-quality SERS signals. Zhu et al. [85] used Ag nanoplate-assembled films as SERS substrates for the rapid detection of PCB-77. The authors obtained a detection limit of about $10^{-7}$ M (mol/L) and found a linear dependence between the logarithmic concentrations of PCB-77 and the fingerprint peaks intensities. The authors used the same Ag nanoplate-assembled also to distinguish the characteristic peaks of different PCBs congeners in mixed solutions. Abalde-Cela and co-workers [82] obtained the SERS spectrum of 2-benzoyldibenzo-p-dioxin at a concentration lower than $10^{-8}$ M (mol/L), a detection level that matches current immunological methods. They used a SERS substrate based on thin films made by exponential layer-by-layer, infiltrated with silver nanoparticles. The authors found a characteristic vibrational pattern characterized by bands at 1392 (ring stretching) and 930 cm$^{-1}$ (ring breathing), and smaller intensity bands at 1641 (CO stretching), 1605 and 1445 (ring stretching), 1296 (CO stretching), 852 (CH wagging), and 791 (ring deformation) cm$^{-1}$.

These sensors represent a rapid, direct, and ultrasensitive detection method for environmental pollutants from different matrices.

Wang and co-workers [88] reported on the use of a sensitive and selective fluorescent membrane for rapid detection of trace 2,2',4,5,5'-pentachlorinated biphenyl (PCB101). The fluorophore phenyl isothiocyanate (PICT) was immobilized onto porous anodic aluminium oxide membrane. The fluorescence of this membrane is enhanced after adding the analyte PCB101 into the membrane, due to the interaction between the fluorophore PICT and the analyte PCB101. The fluorescence intensity increased with the PCB101 concentration in the low range below 1 ppm, and the membrane showed good selectivity.

3. Conclusions and Future Perspectives

Following these premises, optical monitoring of these organic molecules in liquid phase with high time resolution, in vitro (with minimal sample consumption) or even in vivo, is not so far from being achieved.

Nowadays, the development of photonics has already provided us with some tools, i.e., laser sources such as OFC, ICL, QCL, ICL- and QCL-comb, non-linear sources. Furthermore, other new mid-IR components such as waveguides, micro fluidic/waveguides, detectors [91,92] are under development and implementation, thus paving the way toward compact instruments for in vivo measurements of dioxins and related toxic compounds.

On the other hand, plasmonic sensors represent a compact alternative to Mid-IR sensors for non-invasive analysis or in-vivo measures, although their selectivity is strictly dependent on the material used to selectively capture the molecule of interest, and they could suffer surface pollution with a consequent performance reduction.

Author Contributions: B.P., M.S.d.C., S.V. and F.D. shared the tasks of conceptualization, investigation, formal analysis, writing (both original draft, and review and editing), and visualization, according to their specific skills.

Funding: This research received funding from Regione Toscana, in the frame of “POR CREO FESR 2007-2013”, Project SIMPAS (Innovative Measurement Systems for the Protection of Environment and Health) grant number 51018.

Conflicts of Interest: The authors declare no conflict of interest.
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