Antioxidant capacity of cinnamon extract for palm oil stability

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Abstract

Background: Spices and their bioactive components are more promising attractions for their inclusion in diet-based regimes to improve human health. These are sources of natural antioxidants and play an important role in the chemoprevention of diseases and aging. The aim of the current study was to explore the antioxidant potential of cinnamon; a widely used spice throughout the world.

Methods: The current research was aimed to investigate the antioxidant potential of cinnamon extract. For the purpose, cinnamon sticks were procured from local super market, while palm oil was obtained from local oil industry. The resultant extract was analyzed for its antioxidant activity through total phenolic content (TPC), free radical scavenging activity (DPPH assay), and total antioxidant activity was measured by ferric reducing antioxidant power (FRAP) test. The shelf life of palm oil was checked by adding cinnamon extract in oil at different levels i.e. 0.05, 0.10, 0.15, 0.20 and 0.25%, to compare the antioxidant potential of the extract whereas, T⁰ acted as control and T BHA @ 0.1% was used as synthetic antioxidant in the oil samples. The oil samples were analyzed for rancidity check during storage (after every seven days for a storage period of four weeks).

Results: The results indicated that total phenolic contents (TPC); 1,1-diphenyl-2-picrylhydrazyl (DPPH) and ferric reducing antioxidant power (FRAP) values of cinnamon extract were as 355.01 ± 8.34 gallic acid equivalent per gram (mg GAE/g), 90.18 ± 2.12 (%) and 132.82 ± 3.12 (μmol/g), respectively. The oxidative parameters for treatments i.e., T⁰, T BHA, T₁, T₂, T₃, T₄, T₅ were recorded as peroxide value (2.61 ± 0.07, 2.42 ± 0.08, 2.57 ± 0.05, 2.56 ± 0.03, 2.54 ± 0.02, 2.54 ± 0.01, 2.46 ± 0.06 meq/kg, respectively), free fatty acids (0.601 ± 0.05, 0.522 ± 0.02, 0.580 ± 0.07, 0.572 ± 0.03, 0.56 ± 0.07, 0.552 ± 0.03, 0.536 ± 0.05%, respectively), TBA value (22.4 ± 1.45, 20.1 ± 0.73, 21.8 ± 0.42, 21.2 ± 1.56, 20.7 ± 0.48, 20.5 ± 0.59, 20.2 ± 0.91 μg/kg, respectively) and iodine value (52.82 ± 2.12, 52.71 ± 2.38, 52.68 ± 2.96, 52.97 ± 2.14, 52.93 ± 2.12, 53.15 ± 2.38, 52.71 ± 2.96, respectively). Overall, the statistical analysis indicated that all parameters regarding oil stability i.e., peroxide value (PV), free fatty acid content, (FFA) thiobarbituric acid (TBA) value and iodine value (IV) were significant with respect to treatments and storage.

Conclusion: From the present study, it can be concluded that the cinnamon extract proved effective in reducing the lipid oxidation of palm oil and it can be successfully used in place of synthetic antioxidants in food preparations.

Keywords: Cinnamon extract, Antioxidants, Palm oil, Oxidation, Oil stability

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**Background**

Novel health care strategies of the millennium have illuminated functional foods as one of the promising therapeutic tool to curb various lifestyle related maladies. In the recent era health enhancing properties of specific foods have been documented with antioxidant and anti-inflammatory properties [1]. Spices and their bioactive components are more promising attractions for their inclusion in diet-based regimes to improve human health. According to the current nutritional guidelines, diet and health interplay has encouraged the consumers to choose food with some additional health benefits beyond basic nutrition [2]. These are sources of natural antioxidants that play key role in prevention of aging and diseases [3, 4]. Amongst various plants investigated, one showing enormous potential is the cinnamon belonging to family Lauraceae. Cinnamon is gained from the tree bark of cinnamomum genus that is a tropical perennial plant. It has two leading varieties *zeylanicum Cinnamomum (CZ)* and *cassia (CC) cinnamon* [5]. The bioactive compounds of essential oils found from *CZ* bark are trans-cinnamaldehyde, eugenol and linalool, which are representative of almost 82.5% of total compounds [6]. It possesses antimicrobial, antifungal, antioxidant, and anti-diabetic activity [7–9]. Moreover, it also has been used as anti-inflammatory, antitermitic, nematocidal, mosquito larvicial, insecticidal, antimycotic, and anticancer agent [10–12].

Oxidation reactions generates free radicals that start off chain reactions. This spontaneous process of oxidation causes rancidity and food spoilage. Moreover, oxidative stress leads to many lethal diseases in humans like cancer [13]. Currently synthetic antioxidants are used in the food and pharmaceutical industries to extend product shelf life [14]. However, there is increasing interest to explore natural sources of antioxidants to minimize the damage caused by synthetic ones. Phytonutrients are increasingly known for antioxidant activity [15, 16]. Cinnamon spice is one of the sources of effective antioxidants and enhances the efficacy of other important antioxidants. It contains several potent antioxidants and is most important spice used for curtailing oxidative stress. The antioxidant potential of cinnamon is attributed to array of flavonoid compounds that it contains. The essential oils present in cinnamon, including cinnamaldehyde, eugenol, and linalool, has been investigated in reference to peroxyxynitrite induced nitration and lipid peroxidation [17].

Palm oil is an important part of human and animal diet and plays key role in food processing industry. Its use in the commercial food industry is widespread owing to low cost and frying stability. One of the important quality parameters in oil refining industry is low level of free fatty acid (FFA) content and oxidative products [18]. However most of the times palm oil for domestic use is processed through low grade mills which enhances the FFA content and ultimately leads to oxidation during storage. In order to shield the palm oil from oxidative damage during extended storage and transportation, stabilization of the oil with antioxidants is undoubtedly indispensable to preserve the good oxidative status of the palm oil [19]. The developing countries are spending huge amounts on the import of edible oils from other developed countries and the cinnamon extract can be used to preserve the quality of this product. The present study was aimed to explore the potential of natural antioxidant and its comparison with synthetic one to enhance the storage stability of palm oil. The specific objectives of the current study were as (1) probe the antioxidant potential of cinnamon extract (2) exploration the potential of the natural antioxidants in comparison to synthetic ones, and (3) improving the storage stability of palm oil.

**Methods**

**Purchase of raw material**

Cinnamon sticks were procured from Metro cash and carry, Faisalabad. They were dried in an oven at 55 °C till the constant moisture content of 8.5%, crushed using electrical grinder, refined & bleached. Deodorized (RBD) palm oil was obtained from United Ghee Industries Ltd., Faisalabad. All chemicals were of analytical grade and were purchased from Sigma-Aldrich (St. Louis, USA); Merck (Darmstadt, Germany) and Fluka (Buchs, Switzerland). The powdered sample were subjected to solvent extraction using ethanol.

**Solvent extraction of bioactive compounds**

Solvent extraction was done following the method used by Al-Juhaimi and Ghafoor [20] with modifications using ethanol as solvent. The powdered cinnamon sample (10 g) was extracted (50 mL), for 1 h under agitation at room temperature (25 °C) and was filtered through vacuum filtration assembly. The residues were extracted again in ethanol (50 mL), filtered and stored in a freezer at –18 °C till for further use.

**Determination of antioxidant potential of the extract**

**Total phenolic content (TPC)**

Total phenolic contents of cinnamon extract were determined spectrophotometrically following Folin-Ciocalteau method described previously, [21] with minor modification. The appropriate dilution of extract 200 μL oxidized with 1 mL of Folin-Ciocalteau reagent, and then the reaction mixture was neutralized with saturated 2 mL of 7.5% sodium carbonate (w/v). The final mixture volume was maintained up to 7 mL with deionized water. After incubation for 2 h in dark at room temperature, absorbance of the resulting blue color was measured at 765 nm using UV-Visible spectrophotometer. Gallic acid was
used as a standard for the calibration curve. The phenolic content was determined as gallic acid equivalents using the linear equation based on the calibration curve.

**Free radical scavenging activity (DPPH assay)**
The free radical scavenging activity of the extracts was determined by using 1, 1-diphenyl-2-picrylhydrazyl or DPPH [22]. The scavenging ability (SA) was calculated as follows:

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SA\% = \frac{\Delta A_{517 \text{ control}} - \Delta A_{517 \text{ sample}}}{\Delta A_{517 \text{ Control}}} \times 100
\]

**Estimation of total antioxidant activity (FRAP test)**
Total antioxidant activity was measured by ferric reducing antioxidant power (FRAP) assay of Benzie and Strain [23].

**Addition of extract**
The shelf life of palm oil was checked by adding cinnamon extract in oil at different levels, i.e., 0.05, 0.10, 0.15, 0.20 and 0.25%, to compare the antioxidant potential of the extract, whereas, T0 acted as control and TBHA @ 0.1% was used as synthetic antioxidant in the oil samples.

**Oil stability analysis**
To check the antioxidant potential of the extract on the oil stability, following chemical analysis were performed. Peroxide value was measured through procedure No. Cd 8–53 [24]. Free fatty acid content of oil samples was measured by titrating the oil samples dissolved alcohol against sodium hydroxide (NaOH) to a specific end point by procedures described in AOCS method No. Ca 5a-40 [24]. The TBA-reactive substances (TBARS) were determined as described by [25]. Iodine value (IV) of the oil samples was determined by [24] method.

**Statistical analysis**
All determinations were carried out in triplicate and data was reported as mean ± standard deviation. Data obtained for the different parameters was analyzed using the analysis of variance (ANOVA) technique and the least significance difference (LSD) test was used to compare the means, according to the procedure described by Steel et al. [26].

**Results**

**Antioxidant potential of cinnamon extract**
The antioxidant activity of cinnamon extract was investigated using total phenolic content (TPC), free radical scavenging activity (DPPH assay), and ferric reducing antioxidant power (FRAP) test and the values were recorded as 355.01 ± 8.34 mg GAE/g, 90.18 ± 2.12 (%) and 132.82 ± 3.12 (μmol/g), respectively.

**Palm oil stability tests**
In the current study the mean squares relating to PV, FFA, TBA and IV of palm oil associated with treatments and storage have been discussed in Table 1. After the addition of cinnamon extracts RBD, Palm oil was stored at room temperature for 28 days. The oil stability tests were performed weekly to assess the effectiveness of cinnamon extract against oxidation damage or rancidity. The peroxide value of treatments for palm oil i.e., T0, TBHA, T1, T2, T3, T4, T5 was recorded as 2.61 ± 0.07, 2.42 ± 0.08, 2.57 ± 0.05, 2.56 ± 0.03, 2.54 ± 0.02, 2.54 ± 0.01, 2.46 ± 0.06 (Table 2). However, storage caused a significant increase in peroxide value from 0 to 28th day as 2.36 ± 0.03, 2.46 ± 0.08, 2.54 ± 0.06, 2.61 ± 0.04 and 2.68 ± 0.03. The results in the Table 3 revealed that there was a significant difference among treatments on free fatty acid contents of palm oil as the values for T0, TBHA, T1, T2, T3, T4, T5 were observed as 0.601 ± 0.05, 0.522 ± 0.02, 0.580 ± 0.07, 0.572 ± 0.03, 0.56 ± 0.007, 0.552 ± 0.03, 0.536 ± 0.05. The free fatty acid content decreased as the concentration of extract was increased. The effect was more pronounced for the synthetic antioxidant BHA, yet cinnamon extract at 0.25% had almost similar effect (Table 3). Similarly, the effect of storage was also significant as the value of free fatty acids increased as the storage period moved from 0 to 28 days. The values for different storage periods i.e., 0, 7, 14, 21 and 28 days were noticed as 0.491 ± 0.02, 0.521 ± 0.05, 0.555 ± 0.07, 0.580 ± 0.06, 0.654 ± 0.06.

The palm oil was also analysed periodically for TBA value during 28 days storage. The mean values obtained with respect to TBA value in fresh oil samples was 18.6 ± 0.73, whilst at 7, 14, 21 and 28 days the value increased gradually to 19.4 ± 1.45, 20.6 ± 1.56, 21.8 ± 1.56 and 24.7 ± 0.98. The means pertaining to treatments were observed as 22.4 ± 1.45, 20.1 ± 0.73, 21.8 ± 0.42, 21.2 ± 1.56, 20.7 ± 0.48, 20.5 ± 0.59, 20.2 ± 0.91, for T0, TBHA, T1, T2, T3, T4, T5, respectively (Table 4). Mean squares for iodine value of palm oil showed non-significant effect of treatments on this trait. However, during storage this trait behaved differently. As depicted in Table 5, the

**Table 1** Mean squares for PV, FFA, TBA and IV of palm oil during storage

| SOV                        | df | PV    | FFA   | TBA   | IV    |
|----------------------------|----|-------|-------|-------|-------|
| Treatments                 | 6  | 0.06773<sup>a</sup> | 0.01079<sup>a</sup> | 10.610<sup>a</sup> | 0.4468<sup>a</sup> |
| Storage (days)             | 4  | 0.33322<sup>b</sup> | 0.08145<sup>b</sup> | 118.649<sup>b</sup> | 15.9591<sup>b</sup> |
| Treatments x storage       | 24 | 0.00357<sup>NS</sup> | 0.00160<sup>NS</sup> | 2.421<sup>NS</sup> | 0.1246<sup>NS</sup> |
| Error                      | 70 | 0.00045<sup>b</sup> | 0.00018<sup>b</sup> | 0.313 | 1.9391 |

<sup>NS</sup> Non-significant, <sup>PV</sup> peroxide value, <sup>FFA</sup> free fatty acids, <sup>TBA</sup> thiobarbituric acid, <sup>IV</sup> iodine value

<sup>a</sup>Significant
<sup>b</sup>Highly significant
(p < 0.05)
means for iodine value of respective treatments were 52.82 ± 2.12, 52.71 ± 2.38, 52.68 ± 2.96, 52.97 ± 2.14, 52.93 ± 2.12, 53.15 ± 2.38 and 52.71 ± 2.96 respectively, for T0, TBHA, T1, T2, T3, T4 and T5. Concerning the effect of storage, the data exhibited differences among storage days showing a little decline from 0 to 28th day. The values were 53.80 ± 2.12, 53.73 ± 2.96, 52.62 ± 2.63, 52.21 ± 2.12 and 51.90 ± 2.21 for 0, 7, 14, 21 and 28 days, correspondingly.

**Discussion**

The reactive oxygen species may cause deterioration of lipid base foods during processing and storage. They produce off flavors and odors consequently reducing the shelf life and marketability of foods. In the past, synthetic antioxidants were extensively used to prevent lipid oxidation, however recently their use in food is governed by strict regulations owing to their detrimental effects on human health. Today consumers are more conscious about their health and demand food that is more like fresh, resultantly this leads to search for antioxidants from natural sources. In this connection, spices are lavish source of polyphenolic compounds that possess strong antioxidant capacity and can be potentially used to substitute the synthetic antioxidants from food products and confer some additional health effects. The results of the present study regarding TPC, DPPH and FRAP assay were in accordance with the finding of

| Treatments | Storage (days) | Means |
|------------|----------------|-------|
|            | 0 | 7 | 14 | 21 | 28 |
| T0         | 2.36 ± 0.06 | 2.55 ± 0.06 | 2.63 ± 0.08 | 2.69 ± 0.04 | 2.83 ± 0.03 | 2.61 ± 0.07<sup>a</sup> |
| TBHA       | 2.33 ± 0.03 | 2.34 ± 0.04 | 2.41 ± 0.02 | 2.47 ± 0.06 | 2.55 ± 0.03 | 2.42 ± 0.08<sup>d</sup> |
| T1         | 2.36 ± 0.06 | 2.51 ± 0.06 | 2.59 ± 0.03 | 2.66 ± 0.08 | 2.73 ± 0.06 | 2.57 ± 0.05<sup>a</sup><sup>b</sup> |
| T2         | 2.36 ± 0.05 | 2.50 ± 0.02 | 2.58 ± 0.06 | 2.66 ± 0.06 | 2.71 ± 0.02 | 2.56 ± 0.03<sup>b</sup> |
| T3         | 2.37 ± 0.03 | 2.49 ± 0.05 | 2.57 ± 0.04 | 2.65 ± 0.08 | 2.70 ± 0.08 | 2.54 ± 0.02<sup>b</sup> |
| T4         | 2.38 ± 0.06 | 2.48 ± 0.03 | 2.55 ± 0.06 | 2.62 ± 0.06 | 2.66 ± 0.07 | 2.54 ± 0.01<sup>b</sup> |
| T5         | 2.35 ± 0.04 | 2.38 ± 0.07 | 2.46 ± 0.03 | 2.52 ± 0.07 | 2.59 ± 0.08 | 2.46 ± 0.06<sup>c</sup> |
| Means      | 2.36 ± 0.03<sup>a</sup> | 2.46 ± 0.08<sup>d</sup> | 2.54 ± 0.06<sup>c</sup> | 2.61 ± 0.04<sup>b</sup> | 2.68 ± 0.03<sup>a</sup> | – |

**Table 3**

Effect of treatments and storage on free fatty acids content (%) of palm oil

| Treatments | Storage (days) | Means |
|------------|----------------|-------|
|            | 0 | 7 | 14 | 21 | 28 |
| T0         | 0.497 ± 0.03 | 0.548 ± 0.02 | 0.599 ± 0.04 | 0.640 ± 0.05 | 0.721 ± 0.03 | 0.601 ± 0.05<sup>a</sup> |
| TBHA       | 0.498 ± 0.05 | 0.508 ± 0.01 | 0.518 ± 0.05 | 0.528 ± 0.03 | 0.559 ± 0.01 | 0.522 ± 0.02<sup>g</sup> |
| T1         | 0.477 ± 0.02 | 0.538 ± 0.04 | 0.588 ± 0.03 | 0.609 ± 0.05 | 0.690 ± 0.02 | 0.580 ± 0.07<sup>d</sup> |
| T2         | 0.487 ± 0.04 | 0.527 ± 0.01 | 0.568 ± 0.01 | 0.598 ± 0.03 | 0.679 ± 0.03 | 0.572 ± 0.03<sup>b</sup> |
| T3         | 0.483 ± 0.01 | 0.513 ± 0.03 | 0.553 ± 0.03 | 0.583 ± 0.01 | 0.666 ± 0.03 | 0.56 ± 0.07<sup>c</sup> |
| T4         | 0.503 ± 0.03 | 0.513 ± 0.06 | 0.533 ± 0.02 | 0.563 ± 0.02 | 0.646 ± 0.02 | 0.552 ± 0.03<sup>c</sup> |
| T5         | 0.493 ± 0.03 | 0.503 ± 0.05 | 0.523 ± 0.03 | 0.543 ± 0.03 | 0.616 ± 0.04 | 0.536 ± 0.05<sup>d</sup> |
| Means      | 0.491 ± 0.02<sup>d</sup> | 0.521 ± 0.05<sup>d</sup> | 0.555 ± 0.07<sup>c</sup> | 0.580 ± 0.06<sup>b</sup> | 0.654 ± 0.06<sup>a</sup> | – |

Means carrying same letters do not differ significantly (<i>p</i> < 0.05)
Thakur et al. [27]. They probed the antioxidant and antibacterial and activity of anti-inflammatory activity of ethanolic extract of cinnamon, turmeric and ginger. They observed that turmeric and ginger ethanolic extracts were effective against all tested bacteria. Furthermore, the FRAP test proved that the antioxidant potential of ginger was highest among all tested spices whilst the turmeric possess the lowest score. The antioxidant potential of Cinnamon is also good and the value of FRAP assay for cinnamon was 0.40 mM/100 g. They concluded that cinnamon extract has good antioxidant potential. Similar results were also recorded by Jayaprakasha et al. [17]. They studied that cinnamon extract also revealed the highest value for water extract followed by methanol, acetone and ethyl acetate. They concluded that cinnamon has good antioxidant activity and can be used as natural antioxidant in food products. Additionally, research conducted by Wang et al. [28], employed 45 essential oils to determine the antioxidant activity via total phenolics content and DPPH free radical scavenging activity. They noticed best antioxidant activity for leaf of cinnamon and clove bud oils. They narrated that ½ milliliter of these two oils has shown 96.74% and 96.12% DPPH ability, correspondingly. Next, for total phenolic content (mg/g of GAE) cinnamon, clove and thyme essential oils 1 mg/mL of each exhibited 420, 480, and 270, respectively. The present results are also in harmony with the findings of Su et al. [29].

Table 4 Effect of treatments and storage on thiobarbituric acid content (ug/kg) of palm oil

| Treatments | Storage (days) | Means |
|------------|---------------|-------|
|            | 0  | 7  | 14 | 21 | 28 |       |
| T1         | 18.6 ± 0.54   | 20.3 ± 0.48   | 22.1 ± 0.59  | 23.9 ± 0.42 | 26.9 ± 0.73 | 22.4 ± 1.45a |
| T2         | 18.8 ± 0.73   | 19.2 ± 0.98   | 20.0 ± 0.42  | 20.7 ± 1.56 | 21.7 ± 0.54 | 20.1 ± 0.73f |
| T3         | 18.5 ± 0.68   | 20.0 ± 1.56   | 21.4 ± 0.49  | 22.8 ± 0.67 | 26.3 ± 0.91 | 21.8 ± 0.42b |
| T4         | 18.3 ± 1.34   | 19.1 ± 0.59   | 20.5 ± 0.91  | 22.3 ± 1.45 | 26.1 ± 1.45 | 21.2 ± 1.56c |
| T5         | 18.3 ± 0.98   | 18.8 ± 1.56   | 20.1 ± 1.45  | 21.3 ± 0.91 | 24.2 ± 0.42 | 20.7 ± 0.48d |
| T6         | 18.6 ± 1.45   | 19.0 ± 0.54   | 20.0 ± 0.59  | 20.9 ± 0.54 | 24.2 ± 1.56 | 20.5 ± 0.59a |
| T7         | 18.9 ± 0.91   | 19.4 ± 1.45   | 20.5 ± 0.73  | 20.5 ± 0.91 | 22.3 ± 1.45 | 20.2 ± 0.91f |
| Means      | 18.6 ± 0.73a  | 19.4 ± 1.45b  | 20.6 ± 1.56c | 21.8 ± 1.56d | 24.7 ± 0.98e | – |

To = control  
TBHA = synthetic antioxidant (butylated hydroxyl anisole)  
T1 = cinnamon extract 0.5%  
T2 = cinnamon extract 1%  
T3 = cinnamon extract 1.5%  
T4 = cinnamon extract 2%  
T5 = cinnamon extract 2.5%  
Means carrying same letters do not differ significantly (p < 0.05)

Table 5 Effect of treatments and storage on Iodine value of palm oil

| Treatments | Storage (days) | Means |
|------------|---------------|-------|
|            | 0  | 7  | 14 | 21 | 28 |       |
| T1         | 53.84 ± 2.63  | 53.68 ± 2.12 | 52.65 ± 2.96 | 52.09 ± 2.38 | 51.85 ± 2.14 | 52.82 ± 2.12a |
| T2         | 53.80 ± 2.96  | 53.55 ± 2.63 | 52.42 ± 2.38 | 52.05 ± 2.63 | 51.73 ± 2.96 | 52.71 ± 2.38b |
| T3         | 53.77 ± 2.12  | 53.53 ± 2.96 | 52.40 ± 2.38 | 52.02 ± 2.14 | 51.71 ± 2.63 | 52.68 ± 2.96b |
| T4         | 54.06 ± 2.38  | 53.82 ± 2.14 | 52.68 ± 2.12 | 52.30 ± 2.96 | 51.98 ± 2.12 | 52.97 ± 2.14d |
| T5         | 53.06 ± 2.96  | 54.03 ± 2.63 | 52.89 ± 2.63 | 52.51 ± 2.96 | 52.19 ± 2.63 | 52.93 ± 2.12b |
| T6         | 54.25 ± 2.12  | 54.00 ± 2.38 | 52.86 ± 2.12 | 52.48 ± 2.63 | 52.16 ± 2.14 | 53.15 ± 2.38a |
| T7         | 53.81 ± 2.14  | 53.53 ± 2.96 | 52.45 ± 2.14 | 52.04 ± 2.96 | 51.70 ± 2.96 | 52.71 ± 2.96b |
| Means      | 53.80 ± 2.12a | 53.73 ± 2.96a | 52.62 ± 2.63b | 52.21 ± 2.12b | 51.90 ± 2.38e | – |

To = control  
TBHA = synthetic antioxidant (butylated hydroxyl anisole)  
T1 = cinnamon extract 0.5%  
T2 = cinnamon extract 1%  
T3 = cinnamon extract 1.5%  
T4 = cinnamon extract 2%  
T5 = cinnamon extract 2.5%  
Means carrying same letters do not differ significantly (p < 0.05)
They evaluated different spices for their radical scavenging capacity against DPPH, cation 2, 2-azinobis (3-ethylbenzothiazoline- 6-sulfonic acid) (ABTS +), peroxyl oxygen radical absorbance capacity (ORAC) and hydroxyl radicals. They noticed that the extracts of all tested spices including cinnamon exhibited appreciable antioxidant capacities. They found that the acetone extract of cinnamon showed the maximum ABTS + scavenging ability and deduced that the spices especially cinnamon could be employed as a source of natural antioxidant for food products to improve human health and nutrition. The findings from the present study are also in line with the observations of Asimi et al. [30]. They conducted a study to evaluate the antioxidant activity and antimicrobial activities of five Indian spices including Cinnamon, Turmeric, Ginger, Cumin and Garlic. The antioxidant activity in decreasing order was observed as for DPPH Cumin > Garlic > Cinnamon > Turmeric > Ginger, for FRAP as Garlic > Cumin > Turmeric > Ginger > Cinnamon and for TPC as Turmeric > Cinnamon > Garlic > Cumin > Ginger. Hence their findings proved the antioxidant capacity of cinnamon and other spices. Storage conditions of refined deodorized palm oil (RDPO) have an effect on the quality and safety of the products [31, 32]. These results are in agreement with the previous findings of Umerie et al. [33]. They explored the efficacy of the use of Ficus exasperata leaves to improve the quality and stability of palm oil. They concluded that use of Ficus leaves enhanced the storage stability of palm oil. Liang et al. [34] determined the effect of synthetic and natural antioxidants on the oxidative stability of palm-based diesel and noticed that both antioxidants exposed beneficial effects in preventing the oxidation of palm diesel. Zhang et al. [35] explored the effect of natural and synthetic antioxidants on shelf life stability of sunflower oil. They investigated the peroxide values, p-anisidine value, free fatty acids, thiobarbituric acid-reactive substances and concluded that the natural antioxidants are better in reducing the oxidation damage to the oil as compared to synthetic one. The results of current research are in accordance with the findings of Zia-ur-Rehman et al. [36]. They explored the potential of natural antioxidants from potato peels in improving the oxidative stability of soybean oil and inferred that potato peel extract exhibited strong antioxidant capacity which is comparable to synthetic antioxidants BHA and BHT. Aleman et al. [37] concluded that garlic extract is a potent antioxidant and can be employed to reduce lipid oxidation in food products.

Both natural and synthetic antioxidants may inhibit or postpone the process of fat oxidation. An antioxidant can play important role in the prevention of oxidation as by acting as reducing agent, free radical scavenger, chelator and as singlet oxygen scavenger [38]. A number of different antioxidants are being registered as synthetic antioxidants yet few are acceptable as food additive by regulation due to toxicity and other allied harmful effects. Natural antioxidants are readily acceptable to modern consumers as being ideal food additives with good free radical scavenging activity moreover they are safer and healthier as compared to synthetic ones [24]. Cinnamon spice is one of the sources of effective antioxidants and enhances the efficacy of other important antioxidant molecules. It contains several potent antioxidants and is most important spice used for curtailing oxidative stress [39]. The antioxidant potential of cinnamon is attributed to array of flavonoid compounds that it contains. The essential oils present in cinnamon, including cinnamaldehyde, eugenol, and linalool, has been investigated in reference to peroxynitrite induced nitration and lipid peroxidation. Overall, research has proved that cinnamon exhibit higher antioxidant activity as compared to that of other dessert spices [18]. Recently, public’s fear about human health issues caused by food additives has aroused food scientists to keenness of trailing natural antioxidants from several sources. Until now the considerations on this matter is that natural antioxidants are predominantly plant phenolics arising different parts of plants. Some of the phenolic antioxidants from the plant source comprise flavonoids, coumarin, tocopherols, carotenoids, organic acids and cinnamic acid derivatives. The cinnamon extract scavenges harmful free radicals, which are implicated in common cancers and other degenerative diseases including poor brain activity [40]. Recent studies have observed the role of synthetic antioxidants as carcinogens [41]. Hence the search of effective, non-toxic, natural compounds with antioxidative ability has been strengthened in recent years [42].

Essential oils and eugenol from cinnamon bark has exhibited very strong in vitro antioxidant activities, reducing nitrotyrosine generation thus decreasing the peroxynitrite prompted lipid oxidation [12]. The volatile oils obtained from cinnamon has revealed 55.9% and 66.9% antioxidant potential at 100 ppm and 200 ppm, accordingly [43]. Additionally, cinnamon bark extract was observed to be powerful in scavenging free radicals and ABTS radical cations, DPPH, hydroxyl & superoxides were also chelated by these compounds [44]. The current study supported by previous observations has proved that cinnamon extract has antioxidant activity related to its content of phenolic compounds and other antioxidants. It can be deduced that cinnamon can be employed as a source of natural antioxidant for food products to improve human health and nutrition.

Limitations
Despite the proven antioxidant potential of the cinnamon extract further research is suggested to underpin
Conclusions
The edible oils can be degraded through oxidation and decomposition of oxidation products, resulting in generation of alcohols, ketone, aldehyde, acids and hydrocarbons. They can badly affect the appearance, quality and palatability of a food product by creating obnoxious off-flavors and off-odors in food and hereafter affect safety, nutritional value, healthfulness, flavor, color and texture of different foods. These undesirable episodes of oxidation process can be inhibited by various ways including reduction of excess oxygen, and addition of oxidation inhibitors or antioxidants. There is growing trend for the use of natural antioxidants, for food ingredients preservation in recent years due to toxicity problems associated with synthetic antioxidants. The main purpose of the present research was to probe the antioxidant capacity of cinnamon spice. Moreover, the reducing ability of cinnamon extract on lipid oxidation was investigated after addition of extract in palm oil at different concentration levels along with synthetic antioxidant and control for comparison purposes. From the present study, it can be concluded that cinnamon extract has strong antioxidant potential and it can be successfully used in place of synthetic antioxidants in food preparations to improve human health and nutrition by reducing the production of hazardous oxidative derivateis and free fatty acids.

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Availability of data and materials
The dataset supporting the conclusions of this article is included within the article.

Authors’ contributions
The contribution of each author for this paper was as follows, ZS, HS and MTN designed the experimental study. AY and MA provide technical assistance. MI helped for drafting the manuscript. It is evident that all authors read and approved the final manuscript.

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The authors declare that they have no competing interests.

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References
1. Akusu MO, Achinewhu SC, Mitchell J. Quality attributes and storage stability of locally and mechanically extracted crude palm oils in selected communities in rivers and Bayelsa states, Nigeria. Plant Foods Hum Nutr. 2000;55:119–26.
2. Aleman M, Nuchi CD, Bou R, Tres B, Polo J, Guardiola F, Codony R. Effectiveness of antioxidants in preventing oxidation of palm oil enriched with heine iron: a model for iron fortification in baked products. Eur J Lipid Sci Technol. 2010;112:761–9.
3. Al-Juhaimi F, Ghafor K. Total phenols and antioxidant activities of leaf and stem extracts from coriander, black pepper and parsley grown in Saudi Arabia. Pak J Bot. 2011;43:2357–7.
4. Amin KA, El-Twab TMA. Oxidative markers, nitric oxide and homocysteine alteration in hypercholesterolemic rats: role of atorvastatin and cinnamon. Int J Clin Exp Med. 2009;2:254–65.
5. AOCS. Official methods and recommended practices of the American oil chemists’ society. 5th ed. Champaign: AOCS; 2004.
6. Asimi A, Sahu NP, Palc AK. Antioxidant activity and antimicrobial property of some Indian spices. Int J Scient Res Pub. 2013;3:3–4.
7. Benzie FF, Strain JJ. Ferric reducing antioxidant power assay: direct measure of Total antioxidant activity of biological fluids and modified version for simultaneous measurement of Total antioxidant power and ascorbic acid concentration. Method Enzymol. 1999;299:15–23.
8. Bessonov V, Zaltaeva LV, Stepanova LI, Balkov KG. Oxidative and hydrolytic deterioration of palm oil and fat products based on it under various conditions of storage and transportation. Vopr Pitan. 2012;4:18–23.
9. Bruni R, Muzzoli M, Ballero M, Loi MCF, G. Tocopherols, fatty acids and sterols in seeds of four Sardinian wild Euphorbia species. Fitoferapia. 2004;75:50–61.
10. Butt MS, Sultan MT, Butt MS, Iqbal J. Garlic: nature’s protection against physiological threats. Crit Rev Food Sci Nutr. 2009;49:538–51.
11. Cheng SS, Liu JY, Huang CG, Hsu YR, Chen WJ, Chang ST. Insecticidal activities of leaf essential oils from Cinnamomum oshomephileum against three mosquito species. Bioresour Technol. 2009;100:457–64.
12. Chericoni S, Prieto JA, Iacopi P, Cioni P, Morelli I. In vitro activity of the essential oil of cinnamomum zeylanicum and eugenol in peroxynitriuseduced oxidative processes. J Agric Food Chem. 2005;53:4762–5.
13. Dhulasavant V, Shinde S, Pawar M, Naikwade NS. Antihyperlipidemic activity of Cinnamomum tamala Nées. On high cholesterol diet induced hyperlipidemia. Int J Pharm Tech Res. 2010;2:2517–21.
14. Gende LB, Flores I, Fritz R, Eguaras MJ. Antimicrobial activity of cinnamon (Cinnamomum zeylanicum) essential oil and its main components against paenibacillus larvae from argentine. Bulle Insectol. 2008;61:1–4.
15. Geoffrey PW. Dietary supplements and functional foods. New York: Wiley-Blackwell; 2006.
16. Guerra FQS, Mendes JM, de Sousa JP, Morais-Braga MF, Santos BHC, Coutinho HDM, Lima EDO. Increasing antiboic activity against a multidrug-resistant acinetobacter spp by essential oils of citrus lemon and cinnamomum zeylanicum. Nat Prod Res. 2012;26:2235–8.
17. Iqbal S, Banger MI, Faoaqo A. Antioxidant properties and components of some commercially available varieties of rice bran in Pakistan. Food Chem. 2005;93:265–72.
18. Kusum R, Bommayara H, Fayaz Pasha P, Ramachandran HD. Palm oil and rice bran oil current status and future prospects. Int J Plant Physiol Biochem. 2011;3:125–32.
19. Matthäus B. Use of palm oil for frying in comparison with other high-stability oils. Eur J Lipid Sci Technol. 2007;109(4):400.
20. Jayaprakash GK, Negi PS, Jena BS, LIM R. Antioxidant and antimutagenic activities of cinnamomum zeylanicum fruit extracts. Food Compost Anal. 2007;20:330–6.

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21. Lee SK, Mbwanzo ZH, Chung HS, Luyengi L, Games EJC, Mehta RG. Evaluation of the antioxidant potential of natural products. Combinatorial Chemistry & High Chem. High T Scr. 1998;1:35–46.

22. Liang YC, May CY, Foon CS, Ngan MA, Hock CC, Basiron Y. The effect of natural and synthetic antioxidants on the oxidative stability of palm diesel. Fuel. 2006;85:867–70.

23. Mathew S, Abraham TE. Studies on the antioxidant activities of cinnamon (Cinnamomum verum) bark extracts, through various in vitro models. Food Chem. 2006;94:520–8.

24. Meghwal M, Goswami TK. Chemical composition, nutritional, medicinal and functional properties of black pepper, a review. Open Access Sci Rep. 2012;1:1–5.

25. Menoyo D, López-Bote CJ, Bautista JM, Obach A. Herring vs. anchovy fish oils in salmon feeding. Aquat Living Resour. 2002;15:217–23.

26. Pandey A, Pandey KG, Gupta RK, Rizvi SI. Ferric reducing antiradical and β-carotene bleaching activities of nicotinic acid and picolinic acid biocjugates of curcumin. Nat Prod Commun. 2011;6:1877–80.

27. Pushpangadan P, Dan VM, Ijimu TP, George V. Food, nutrition and beverage. Indian J Tardit Knowl. 2012;11:26–34.

28. Ramalho VC, Jorge N. Antioxidants used in oils, fats and fatty foods. Química Nova. 2006;29:755–60.

29. Steel RGD, Torrie JH, Dickey D. Principles and procedures of statistics - a biometrical approach. 3rd ed. New York: McGraw Hill Book Co Inc; 1997.

30. Su L, Yin JJ, Charles D, Zhou K, Moore J, Yu L. Total phenolic contents, chelating capacities, and radical-scavenging properties of black peppercorn, nutmeg, rosehip, cinnamon and oregano leaf. Food Chem. 2007;100:990–7.

31. Thakur R, Yadav K, Khadka KB. Study of antioxidant, antibacterial and anti-inflammatory activity of cinnamon (Cinnamomum tamala), ginger (Zingiber officinale) and turmeric (Curcuma longa). Am J Life Sci. 2013;1:273–7.

32. Ullah J, Hamayoun M, Ahmad T, Ayub M, Zarafullah M. Effect of light, natural and synthetic antioxidants on stability of edible oil and fats. Asian J Plant Sci. 2003;2:1192–4.

33. Umerie SC, Ogbuagu AS, Ogbuagu JO. Stabilization of palm oils by using Ficus exasperata leaves in local processing methods. Bioreos Technol. 2004;94:307–10.

34. Wang HF, Wang YK, Yih KH. DPPH free-radical scavenging ability, total phenolic content, and chemical composition analysis of forty-five kinds of essential oils. Int J Cosm Sci. 2008;59:6509–22.

35. Wang SY, Chen PF, Chang ST. Antifungal activities of essential oils and their constituents from indigenous cinnamon (Cinnamomum osmophloeum) leaves against wood decay fungi. Bioreos Technol. 2005;96:813–8.

36. Zhang YY, Lei Z, Yuanang C, Xiaqiang W, Fuji LF. Oxidative stability of sunflower oil supplemented with carnosic acid compared with synthetic antioxidants during accelerated storage. Food Chem. 2010;118:566–62.

37. Zia-ur-Rehman Habib F, Shah WH. Utilization of potato peels extract as a natural antioxidant in soy bean oil. Food Chem. 2004;85:215–20.

38. Ebrahimabadi AH, Delfar-Bidogoli Z, Mazocchi A, Kashf FJ, Batooli H. Essential oil composition, antioxidant and antimicrobial activity of the leaves and flowers of Chaerophyllum macropodum Boiss. Food Control. 2010;21:1173–8.

39. Jayaprakasha GK, Ohnishi-Kameyama M, Ono H, Yoshida M, Rao LJ. Phenolic constituents in the fruits of cinnamomum zeylanicum and their antioxidant activity. J Agric Food Chem. 2006;54:1672–9.

40. Reddy V, Utrooj A, Kumar A. Evaluation of antioxidant activity of some plant extracts and their application in biscuits. Food Chem. 2005;90:317–21.

41. Gohari Ardabili A, Farhoosh R, Haddad Khodaparast MH. Frying stability of canola oil in presence of pumpkin seed and olive oils. Eur J Lipid Sci Technol. 2010;112:871–7.

42. Gupta VK, Sharma SK. Plants as natural antioxidants. Nat Prod Radiance. 2006;5(4):326–34.

43. Jayaprakasha GK, Jagan MRL, Sakariah KK. Volatile constituents from cinnamomum zeylanicum fruit stalks and their antioxidant activities. J Agric Food Chem. 2005;51:4344–8.

44. Mathew S, Abraham TE. Studies on the antioxidant activities of cinnamon (Cinnamomum verum) bark extracts, through various in vitro models. Food Chem. 2004;94:520–8.