Skeletal muscle myostatin gene expression and sarcopenia in overweight and obese middle-aged and older adults

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Abstract

Background—Myostatin (MSTN) is a key negative regulator of muscle mass in humans and animals, having direct and indirect influences on molecular regulators of atrophy and hypertrophy, thus potentially impacting fitness and physical function. We have shown that myostatin is elevated in conditions of chronic disability (e.g. paretic limb of stroke). Our hypothesis is that myostatin would be elevated in older adults with sarcopenia. The purpose of this study was to examine the role of skeletal muscle myostatin in sarcopenia.

Methods—Sixty-four overweight to obese aged 45–81 years underwent a maximal aerobic capacity (VO$_2$max) test, dual-energy X-ray absorptiometry (DXA) scan to determine appendicular lean tissue (ALM), and vastus lateralis muscle biopsy to determine myostatin mRNA expression by quantitative real time PCR (Q-RT-PCR). Rates of sarcopenia were determined using (ALM/BMI), and sarcopenia was defined as <0.789 in men and <0.512 in women. Subjects had low fitness (VO$_2$max: 22.7 ± 0.7 mL/kg/min) and on average 40.9 ± 1% body fat.

Results—The prevalence of sarcopenia in this cohort was 16%. BMI, % body fat, and fat mass were higher in adults with sarcopenia than those without sarcopenia (all P < 0.001). Myostatin mRNA expression was lower in those without sarcopenia than those with sarcopenia (P < 0.05) and higher in men than women (P < 0.001). Myostatin expression was associated with BMI (r = 0.36, P < 0.01) and mid-thigh intramuscular fat (r = 0.29, P < 0.05).

Conclusion—Given that myostatin is important in muscle atrophy, fat accumulation, and sarcopenia, further work could address its implication in other aging cohorts of disability and chronic disease.

Keywords

Sarcopenia; Myostatin; Skeletal muscle atrophy; Fat accumulation; Obesity; Aging
Introduction

Sarcopenia, the age-related loss in skeletal muscle mass and strength, is a leading contributor to the development of frailty. Sarcopenic obesity is described as the combination of loss of muscle mass, function and elevated body mass index and also increases the risk for metabolic disturbances. Over 15 years ago, Baumgartner and colleagues reported that adults with sarcopenic obesity are two to three times more likely to report an onset of Instrumental Activities of Daily Living (IADL) disability than adults with normal body composition, lean sarcopenic, and obese nonsarcopenic adults, indicating an independent association of sarcopenic obesity and the development of disability. Furthermore, this >2.5 relative risk for incident disability in sarcopenic obese adults occurs after adjustment for confounding factors of age, sex, physical activity level, length of follow-up, and prevalent morbidity. Although several other groups have nicely reviewed sarcopenia obesity and potential biological pathways that could explain its pathology, the skeletal muscle mechanisms linking sarcopenia with obesity in adults is not completely understood.

Myostatin is a member of the transforming growth factor beta family of secreted growth factors and a significant regulator of skeletal muscle development and size. In fact, anti-myostatin antibodies are potential therapeutic options for sarcopenia. Myokines, including myostatin, play a role in the pathogenesis of sarcopenic obesity. Furthermore, ectopic fat deposition, obesity, and aging are implicated in impaired skeletal muscle protein synthesis leading to muscle lipid accumulation.

We have shown greater muscle atrophy, increased intramuscular fat, and higher skeletal muscle myostatin in conditions of chronic disability (e.g. paretic limb of stroke). We hypothesized that skeletal muscle myostatin would be elevated in adults with sarcopenia. Therefore, the aim of the study was to compare skeletal muscle myostatin expression in middle-aged and older overweight and obese adults with and without sarcopenia.

Methods

Participants

Male and female adults aged 45–80 years from the Baltimore/Washington area were eligible to participate if generally healthy, weight stable (<2.0 kg weight change in past year), sedentary (<20 min of aerobic exercise 2× per week), overweight or obese (BMI 29–50 kg/m²), and without the presence of heart disease, diabetes, cancer, anaemia, dementia, untreated dyslipidaemia, or other unstable or chronic diseases affecting the liver, lungs, or kidneys. Women were included only if they had undergone menopause for at least 1 year. Potential participants were screened and underwent a physical examination including a comprehensive past medical history, fasting blood profile, and a graded exercise treadmill test as part of another longitudinal investigation. Each study participant provided written consent. All methods and procedures were approved by the Institutional Review Board at University of Maryland School of Medicine and the VA Research and Development Committee.
Tests

Height (cm) and weight (kg) were measured to calculate body mass index (BMI). Body fat mass, lean mass of the arms and legs (to calculate appendicular lean mass) were measured by dual-energy X-ray absorptiometry (Lunar iDXA, GE Healthcare). Sarcopenia was defined as appendicular lean mass/height$^2$, $\text{ALM/ht}^2 < 7.26 \text{ kg/m}^2$ in men and $< 5.45 \text{ kg/m}^2$ in women.\textsuperscript{10} Measurements of grip strength and gait speed were not part of the original longitudinal study goals and so were not available for use in the definition of sarcopenia. Computed tomography (CT) scans (Siemens Somatom Sensation 64 Scanner, Fairfield, CT) at L$^4$-L$^5$ region was used to determine visceral adipose tissue area, subcutaneous adipose tissue area, and a scan at the mid-thigh to quantify muscle area, subcutaneous fat area, and intramuscular adipose tissue (IMAT). Scans were analysed using Medical Image Processing, Analysis and Visualization, v.7.0.0. Ten individuals were missing measurement of visceral fat and 25 were missing thigh scans for measurement of IMAT. $\text{VO}_2\text{max}$ was measured using a continuous treadmill test protocol. Vastus lateralis percutaneous needle muscle biopsies were taken from each participant under local anaesthesia using a 5 mm Bergstrom needle (Stille, Solna, Sweden). Muscle was immediately freeze-clamped and stored at $-80^\circ\text{C}$. Approximately 50–80 mg of muscle was used for RNA isolation and myostatin gene expression. Total RNA extraction, cDNA synthesis and quantitative real-time PCR (qPCR) were done by our standard laboratory methods.\textsuperscript{17} To be included in this research question, each participant had to have sufficient muscle sample for myostatin analysis. All testing and lab analysis were blinded as to sarcopenia status of either the participant and the study samples to eliminate any bias.

Statistical analyses

Descriptive means were calculated on 14 variables. One-way ANOVAs were used to test differences between groups. Pearson correlations and partial correlations were used to assess relationships between key variables. Statistical significance was set at a two-tailed $P < 0.05$. Data were analysed using SPSS Statistics 24 (SPSS Inc., Chicago); results are expressed as mean ± SEM.

Results

Study sample

Descriptive characteristics of the participants ($n = 64$) are provided in Table 1. They were 78% Caucasian and 22% African American ($n = 14$) and included 31 men (48%) and 33 women (52%). On average, participants had low fitness levels with a wide range of total body fat.

Subject characteristics between sarcopenic ($n = 10$) and non-sarcopenic ($n = 54$) adults are presented in Table 2. The prevalence of sarcopenia in this cohort was 16%. Age, height, $\text{VO}_2\text{max}$, or FFM did not differ between groups defined by sarcopenia status. Adults with sarcopenia had higher body weight ($P < 0.05$), BMI ($P < 0.05$), per cent body fat ($P < 0.05$), and body fat mass ($P < 0.01$). Myostatin mRNA expression was lower (30.4%) in those without sarcopenia than those with sarcopenia ($P < 0.05$, Figure 1). Myostatin expression was higher in men than women in the total group ($83.4 ± 5.7$ vs. $54.3 ± 4.2$ AU, $P < 0.0001$).
It was also higher in men (n = 6) than women (n = 4) with sarcopenia (120.6 ± 13.7 vs. 48.8 ± 11.6 AU, P < 0.01) and men (n = 25) than women (n = 29) without sarcopenia (74.4 ± 4.9 vs. 55.1 ± 4.5 AU, P < 0.01).

**Relationships**

Skeletal muscle myostatin expression was associated with mid-thigh IMAT (r = 0.29, P < 0.05), mid-thigh muscle area (r = 0.43, P < 0.05), BMI (r = 0.36, P < 0.01), and WHR (r = 0.43, P < 0.01) but not related to age, VO\(_2\)max, total body fat mass, or visceral fat in the total group. Skeletal muscle myostatin mRNA level tends to be positively correlated with fat mass in both men and women (men: r = 0.33, P = 0.07; women: r = 0.32, P = 0.07) and was positively correlated with subcutaneous abdominal fat in men (r = 0.40, P < 0.05) but not in women (r = 0.30, P = 0.11). Myostatin levels were not related to mid-thigh muscle area in either men or women alone.

**Discussion**

Sarcopenia is an age-related condition of loss of skeletal muscle mass, physical function, and strength. Participants in our study were overweight and obese who were on average 60 years of age. The mechanisms linking sarcopenia with obesity in adults are still not completely clear. Our data show that myostatin mRNA expression was significantly higher in adults with sarcopenia than those without sarcopenia. Myostatin as a myokine is expressed in skeletal muscle and is a negative regulator of skeletal muscle growth in animals and human and is associated with body fatness and aerobic capacity. Our data suggest that myostatin levels in the skeletal muscle might be one of the drivers for sarcopenia.

Muscle and fat are the two major metabolic tissues secreting cytokines, myokines, and adipokines that regulate metabolism and body composition. Myostatin is encoded by the myostatin gene in humans and is a myokine produced and released by myocytes that acts on muscle cells’ autocrine function to inhibit muscle cell growth and differentiation. The full length gene encodes a preproprotein that has 375 amino acids. This preproprotein is inactive until a protease cleaves the NH2-terminal, or “pro-domain” portion of the molecule, forming the active COOH-terminal dimer with a total molecular weight of 25.0 kDa. Each monomer of the protein dimer has 109 amino acid residues. Thus, myostatin detected in skeletal muscle by the biopsy technique can provide insight into muscle loss.

There are several lines of evidence to support that myostatin levels would differ between those with and without sarcopenia. Myostatin acts in two ways: by inhibiting either muscle differentiation or Akt-induced protein synthesis. When myostatin binds to the activin type II receptor, it results in a recruitment of either coreceptor Alk-3 or Alk-4. This coreceptor then initiates a cell signalling cascade in the muscle including the activation of transcription factors, SMAD2 and SMAD3. These factors then induce myostatin-specific gene regulation. When applied to myoblasts, myostatin inhibits their differentiation into mature muscle fibres. Myostatin also inhibits Akt, a kinase that causes muscle hypertrophy through the activation of protein synthesis. Additional studies could examine muscle transcription factors or Akt levels and their relationship to sarcopenia in adults with obesity.
Sarcopenia can affect up to one-fourth of older adults\textsuperscript{24} whereas sarcopenic obesity using muscle mass in the definition of sarcopenia has a lower prevalence ranging from 4% to 12%.\textsuperscript{25-27} Thus, the 16% prevalence observed in our study population is slightly higher than that previously reported. Moreover, our participants with sarcopenia would be considered sarcopenic obesity. Specifically, all the men were obese by BMI and the two women whose BMI were below the obesity cut-point (e.g. 28.6 and 29.9 kg/m\textsuperscript{2}), and body fat percentage were high (51% and 45%, respectively). Obesity is associated with metabolic disorders and implies a connection between muscle and adipose tissue. Both aging and an unhealthy lifestyle such as physical inactivity and over nutrition contribute to the development of sarcopenic obesity.\textsuperscript{28} When people age, the capability for protein synthesis decreases while protein degradation increases.\textsuperscript{28-30} The proliferation and self-renewal of satellite cells as well as the differentiation of myocytes decreases, causing a loss in muscle mass and decline in physical function. In adults with obesity, mitochondrial biogenesis tends to decrease, causing an increase in oxidative stress and inflammation.\textsuperscript{28,31} Myostatin has also been found to be expressed in fat tissues and plays a key role in adipogenesis and potential control of body fat mass.\textsuperscript{32} One study shows that myostatin is associated with abdominal obesity in women with polycystic ovary syndrome.\textsuperscript{33} In a pre-clinical study that compared the myostatin-null mice with wild-type littersmates, the former had a significant reduction in fat accumulation with increasing age.\textsuperscript{34} Myostatin deletion or inhibition leads to a decrease in fat mass in mice.\textsuperscript{32} Further, there is no effect on body composition on the inhibition of myostatin signals within adipose tissue in mice fed a standard diet or a high-fat diet whereas in contrast, inhibition of myostatin signalling in skeletal muscle results in an increase in lean mass, a decrease in fat mass, and improved glucose metabolism on standard and high-fat diets, and resistance to diet-induced obesity.\textsuperscript{35} In a pre-clinical aging model, protein synthesis is reduced in diet-induced obesity and the lipid redistribution in lean tissues or ectopic fat reduces protein turnover.\textsuperscript{12} Our data would support these pre-clinical studies and aligns with our finding that ectopic fat deposition in the muscle was associated with skeletal muscle myostatin. In addition, our findings that myostatin levels and BMI and waist-hip ratio are positively associated suggest that skeletal muscle myostatin may be important in the development of both central obesity and IMAT. The higher myostatin levels in men than women likely drive its relationship with mid-thigh muscle area in the total group. In myostatin-knockout mice, muscle hypertrophy is augmented and in androgen receptor-knockout mice, skeletal muscle growth is inhibited.\textsuperscript{36} Thus, the higher myostatin expression observed in men could speculatively reduce the actions of androgens including the regulation of muscle mass, strength, and fibre-type distribution. Nevertheless, myostatin is also related to central obesity (subcutaneous abdominal fat) in men demonstrating the role of this myokine in adiposity.

Plasma or serum myostatin has been suggested to be implicated in the muscle-wasting in sarcopenia.\textsuperscript{37} Strengths of this study include the conduct of skeletal muscle biopsies and measurement of myostatin in the skeletal muscle, careful characterization of the study sample with fitness and body composition measures, sample size, and novelty of the question. Our study may not be generalizable to older adults with chronic disease or multi-morbidities. Study limitations are the inability to discern sex differences in sarcopenia and myostatin and the lack of measurement of grip strength or gait speed to use in other
definitions of sarcopenia, and additional factors in skeletal muscle that could contribute to the sarcopenia of loss of muscle mass and function.

**Conclusion**

Skeletal muscle myostatin appears to be important in muscle atrophy, fat accumulation and sarcopenic obesity. Further work could address its implication in other aging cohorts of disability and chronic disease.

**Acknowledgements**

This research was supported funds from a Senior Research Career Scientist Award (ASR) from the United States Department of Veterans Affairs (VA) Rehabilitation R&D (Rehab RD) Service, VA Merit Award 1153486 (ASR) Clinical Service R&D, VA Medical Center Baltimore Geriatric Research, Education and Clinical Center (GRECC), National Institutes of Health Grant P30-AG028747, and P30 DK072488.

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Figure 1.
Skeletal muscle MSTN level between sarcopenia and non-sarcopenia. *P < 0.05.
Table 1

| Variables                              | Mean ± SEM | Range     |
|----------------------------------------|------------|-----------|
| Age (yrs)                              | 61 ± 1     | 45–81     |
| Weight (kg)                            | 94.3 ± 2.2 | 63.6–142.2|
| Height (m)                             | 1.70 ± 0.01| 1.53–1.88 |
| BMI (kg/m²)                            | 32.7 ± 0.6 | 24.9–46.1 |
| VO₂ max (mL/kg/min)                    | 22.7 ± 0.7 | 8.9–36.4  |
| VO₂ max (L/min)                        | 2.2 ± 0.1  | 0.8–3.7   |
| Body fat (%)                           | 40.9 ± 1   | 24.5–59.3 |
| Fat mass (kg)                          | 38.9 ± 1.4 | 20.0–78.3 |
| Fat-free mass (kg)                     | 55.9 ± 1.5 | 38.2–75.2 |
| Appendicular lean mass (kg)            | 24.2 ± 0.7 | 14.57–34.1|
| Mid-thigh muscle area (cm²)            | 91.2 ± 5.1 | 47.8–169.6|
| Mid-thigh subcutaneous fat area (cm²)  | 118.6 ± 9.8| 36.2–313.9|
| Mid-thigh intramuscular fat area (cm²) | 27.3 ± 1.8 | 12.0–56.1 |
| Visceral fat area (cm²)                | 185.2 ± 10.7| 66.4–393.8|
| Subcutaneous abdominal fat area (cm²)  | 414.2 ± 17.2| 216.2–684 |
| Myostatin gene expression (AU)         | 68.4 ± 3.9 | 9.4–172.5 |

Mean ± SEM.
| Variables          | Sarcopenia (n = 10) | Non-sarcopenic (n = 54) |
|-------------------|--------------------|------------------------|
| Age (years)       | 60 ± 2             | 62 ± 1                 |
| Weight (kg)       | 110.4 ± 6.9*       | 91.3 ± 2.1             |
| Height (m)        | 1.71 ± 0.02        | 1.69 ± 0.01            |
| BMI (kg/m²)       | 37.6 ± 1.9 **      | 31.8 ± 0.6             |
| VO₂max (mL/kg/min)| 20.9 ± 1.1         | 22.8 ± 0.8             |
| Body fat (%)      | 46.1 ± 2.1*        | 40.0 ± 1.1             |
| Fat mass (kg)     | 50.5 ± 4.0 **      | 36.8 ± 1.3             |
| Fat-free mass (kg)| 59.1 ± 4.2         | 55.3 ± 1.6             |

Mean ± SEM. Sarcopenia versus non-Sarcopenia.

* P < 0.05.

** P < 0.001.