Evaluation of *Zapoteca tetragona* forage as alternative protein source in ruminants’ feeding

Hadriana Bansi,1 Elizabeth Wina,2 Procula Rudolf Mattaputy,1 Vito Laudadio,2 Vincenzo Tufarelli3
1Assessment Institute for Agricultural Technology, Makassar, Indonesia
2Indonesian Research Institute for Animal Production, Bogor, Indonesia
3Dipartimento dell’Emergenza e dei Trapianti di Organi, Università di Bari Aldo Moro, Valenzano, Italy

Abstract

The aim of this study was to determine the nutritional characteristics of *Zapoteca tetragona* (Willd.) H. Hern to assess the suitability of this plant for ruminant nutrition. The nutritional evaluation consisted of in vitro and in vivo trials. Secondary compounds including total phenols, condensed tannin and non-protein amino acids (NPAA) were determined. Two stage in vitro digestibility was conducted using substrates with increasing levels of *Z. tetragona* replacing elephant grass (*Pennisetum purpureum*) as control feed. The inclusion of 30% *Z. tetragona* was compared to 100% elephant grass by in vitro gas production technique and in vivo digestibility trial using sheep. Forage from *Z. tetragona* was appreciably high in crude protein (CP) and lower in neutral detergent fibre. Moreover, it was rich in Ca and P. Total phenols, condensed tannin and NPAA contents were very low. In vitro gas production technique showed that after 48 h incubation, the gas produced from *Z. tetragona* was higher than elephant grass (P<0.05). Increasing level of *Z. tetragona* led to better dry matter (DM) and CP digestibility compared to elephant grass. In vivo trial showed no difference in DM intake between the two tested feed, however higher CP intake was reported when sheep fed *Z. tetragona* as well for CP digestibility and N retention (P<0.05). It can be concluded that *Z. tetragona* has a strong potential as forage crop with valuable nutritional quality. Moreover, *Z. tetragona* could represent an alternative feedstuff to conventional forage and a promising substitute fodder in tropical ecosystem.

Introduction

There is currently an increasing interest in the production of alternative forage crops for ruminant feeding. This reflects both the potentially lower production costs per unit of energy associated with some alternative forage crops and the ability of some of these crops to increase total dry matter (DM) intake and improve animal productions (Khan and Habib, 2012; Tufarelli et al., 2012). Leaves of browse species are potential source of nutrients that could be used to improve the production of ruminants consuming tropical pastures of low nutritive value (Barakat et al., 2013). Tree leaves are richer in crude protein (CP), minerals and digestible nutrients than grasses (Olahadehan, 2013).

*Zapoteca tetragona* (Willd.) H. Hern. is a tropical leguminous shrub widely cultivated in the forest buffers area of Indonesia. This shrub originally came from Latin or Central America and brought to Indonesia by the forestry people. Previously, *Z. tetragona* was called white Calliandra since the flower has white colour, if compared to the red Calliandra (*Calliandra calothyrsus*) having red flower. *Z. tetragona* is a fast growing species and regrows well after several cuttings. Compared to other tropical shrub species, such as *Acacia angustissima*, *Calliandra calothyrsus* and *Leucaena diversifolia*, the DM biomass production of *Z. tetragona* is higher (Sajimin and Purwarianti, 2006). The protein content of *Z. tetragona* is quite high and it represents a valuable protein source compared to other tropical shrubs (Laudadio et al., 2009; Cazzato et al. 2013). However, to date the information of *Z. tetragona* forage as alternative feed for ruminants are limited, and little attention has been given to the nutritional evaluation of *Z. tetragona* in ruminant feeding. Moreover, since the *Z. tetragona* leaves contain several secondary compounds which may influence the animal production, it is necessary to assess the nutritional value of *Z. tetragona* through not only chemical analyses but also by in vitro techniques and in vivo trials.

Materials and methods

Study area and sampling

*Z. tetragona* and elephant grass (*Pennisetum purpureum*) were locally produced from mature crops after harvesting of plants in the research facility of the Indonesian Research Institute for Animal Production, Bogor, Indonesia (7°5’S latitude, 107°40’E longitude). The crops were harvested, wilted, and transported to the research facility. The feeding trial was conducted at the animal research facilities of Indonesian Research Institute for Animal Production, Bogor, Indonesia.

In vivo digestibility trial

A metabolism trial was conducted lasted for 21 d in metabolism cages using eight healthy sheep of 24±2.2 kg live body weight (BW) with facility of quantitative collection of faeces and urine separately. Each animal received the two diets in different periods, following a Latin square design. Each period consisted of 14 days of adaptation to diet in individual stalls with a concrete floor, two days of adaptation to faecal collection bags applied on bucks, and five days of faeces collection (Givens et al., 2000). Daily feed intake was also recorded. The diets consisted of a mixture of elephant grass and *Z. tetragona* in the ratio of 70:30 w/w and a control diet of 100% elephant grass, both diets at a level of 4% BW per day. The proximate composition of elephant grass and *Z. tetragona* was reported in Table 1. Samples of feed offered, oris, faeces and urine voided were collected every morning. A 0.1 portion of faeces and urine, respectively, were pooled
over the 5-day collection period and finally sub-sample to obtain representative samples for analysis. Rumen liquor (50 mL) was taken from each sheep in the morning before feeding, using an oesophageal tube under mild vacuum from the reticulum near the reticulor- omasal orifice, and filtered through two layers of cheesecloth. An aliquot of the filtered rumen liquor was collected for protozoal and bacterial counts (Ogimoto and Imai, 1981), and another aliquot of filtered rumen liquor was collected for the determination of the ammonia concentration (Conway and Byrne, 1933).

**Two-stage in vitro digestibility**

In order to determine the best level of *Z. tetragona*, different percentages (15, 30, 45 and 100%, respectively) were tested by **in vitro** fermentation assay. The Tilley and Terry (1963) rumen fluid/pepsin two-stage **in vitro** technique was used to estimate the **in vitro** DM digestibility and CP digestibility. Results were compared with control incubations (i.e., samples without *Z. tetragona*) using elephant grass. Rumen fluid was collected from sheep via rumen canulae before the morning meal was pooled, placed in a prewarmed (39°C) vacuum flask and transported immediately to the laboratory. The rumen fluid was diluted (1:2 v/v) with a culture medium containing macro- and micromineral solutions, resazurin and bicarbonate buffer solution prepared as described by Menke and Steingass (1988). The medium was kept at 39°C and saturated with CO2. Samples (0.5 g) of each forage (control or with one of the *Z. tetragona* levels) were weighed in triplicate and placed in bottles, to which were added 40 mL of rumen fluid/artificial saliva solution. The bottles were kept for 24 h of incubation and jointly with the levels of forage chemical components.

**Chemical analysis**

Samples of *Z. tetragona*, elephant grass and dry faeces were ground in a hammer mill with a 1 mm screen and analysed in triplicate for DM, ash (967.05), CP (Kjeldahl N×6.25; 990.03), ether extract (945.16), Ca and P according to AOAC (2000). The neutral detergent fibre (NDF), acid detergent fibre (ADF) and lignin were analysed according to Mertens (2002), AOAC (2000; 973.187), and Van Soest et al. (1991), respectively, using the sequential procedure and the filter bag system (Ankom Technology, Macedon, NY, USA). The NDF and ADF fractions include residual ash. Total phenols and condensed tannin analysis were carried out following the methods described by FAO/IAEA (2000) and non-protein amino acids (NPAAs) according to the AOAC (2000). Rumen ammonia was analysed by the Kjeldahl method (990.03; AOAC, 2000).

**Statistical analysis**

Data on intake, digestibility, N retention and ruminal parameters in sheep were analysed for the fixed effect of diet and random effects of feeding period and sheep, using PROC MIXED procedure of the SAS (SAS, 2004). The DM and CP digestibility data were compared using *t*-test (Steel et al., 1997). Similar design was used also for the two-stage **in vitro** digestibility and **in vitro** gas production.

### Table 1. Proximate chemical composition of forage from *Z. tetragona* and elephant grass.

| Item                  | *Z. tetragona* | Elephant grass |
|-----------------------|----------------|----------------|
| DM                    | 28.67          | 14.34          |
| CP                    | 23.49          | 17.46          |
| NDF                   | 27.60          | 56.20          |
| ADF                   | 17.62          | 30.28          |
| Lignin                | 3.47           | 3.34           |
| Ash                   | 8.44           | 15.80          |
| Ca                    | 1.71           | 0.52           |
| P                     | 0.33           | 0.25           |
| Total phenols         | 3.31           | nd             |
| Condensed tannin      | 0.73           | nd             |
| NPAAs                 | 0.60           | nd             |

DM, dry matter; CP, crude protein; NDF, neutral detergent fibre; ADF, acid detergent fibre; Na, sodium; P, phosphorus; NPAAs, non-protein amino acids; nd, not detected.
supplied by *Z. tetragona* forage was efficiently utilised and resulted in a greater N retention as also recently found by Khan and Habib (2012) using peanut (*Arachis hypogaea*) forage.

Apparent *in vivo* digestibility of DM and CP showed that in sheep fed *Z. tetragona* the CP digestibility was improved (P=0.003), whereas DM digestibility was unaffected by dietary treatment. Our results are in line with previous findings of Khamseekhiew et al. (1991) in another tropical leguminous species *Giricidia sepium*, as protein source replacing elephant grass, indicated that total DM and CP digestibility using 40% *Giricidia* in ruminant ration were higher compared to control ration. Further, the CP digestibility of *Z. tetragona* was also higher than another tropical leguminous forage as red calliandra (*Calliandra calothyrsus*) (Palmer and Jones, 2000). Therefore, we can state that *Z. tetragona* resulted a species highly degradable in the rumen.

Ruminal ammonia concentration, and total bacteria and protozoa counts were unaffected by dietary treatments, resulting similar in sheep fed elephant grass or *Z. tetragona*. These positive findings could be relate to the secondary compounds in *Ztetragona* that not showed any negative effect on total bacteria or protozoa in the sheep rumen. This also explains that the high total bacteria and protozoa will provide an high degradation rate in the rumen.

Data on the effect of substituting elephant grass with different levels of *Z. tetragona* on two-stage *in vitro* DM and CP digestibility are reported in Table 3. Digestibility of DM was higher (P<0.05) for *Z. tetragona* at all levels of inclusion compared to the control elephant grass. Similarly, the CP digestibility was higher (P<0.05) for *Z. tetragona*, in particular at 15% of inclusion when incubated using rumen liquor and pepsin. The improved digestibility in the *Z. tetragona* as compared to elephant grass is consistent with the previous studies involving Guinea grass with *Ficus religiosa* (Bamikole, 2003) or *Pterocarpus erinaceus* combined with Gamba grass (*Andropogon gayanus*; Oladahunsi, 2013). It seems that *Z. tetragona* resulted in improved rumen ecology and a faster degradation of the diets compared to the elephant grass. The higher nutrient digestibility of our alternative could be attributed to lower fibre contents of the *Z. tetragona* than elephant grass. The lower fibre of *Z. tetragona* could result in fast degradation. Moreover, legumes are generally retained in the rumen for a shorter time than grasses (Khan et al., 2012).

The *in vitro* gas production technique trial showed that starting from the first 3 h of incubation, whereas the gas production from the mixture of elephant grass- *Z. tetragona* was higher than elephant grass. The lower fibre of *Z. tetragona* could result in fast degradation. Moreover, legumes are generally retained in the rumen for a shorter time than grasses (Khan et al., 2012).

Figure 1. Means of volume of gas produced during 48 h of incubation of 100% elephant grass (■) and 30% of *Z. tetragona* (●).

Table 2. Dry matter and crude protein intake, *in vivo* digestibility, N retention and rumen parameters in sheep fed elephant grass (100%) and *Z. tetragona* (30%).

| Parameters                  | Elephant grass | Z. tetragona | SEM | P     |
|-----------------------------|----------------|--------------|-----|-------|
| Intake                      |                |              |     |       |
| DM, g/day                   | 682.7          | 795.2        | 78.92 | 0.090 |
| DM, g/kg BW<sup>23</sup>    | 64.70          | 70.36        | 5.912 | 0.227 |
| CP, g/day                   | 60.54          | 122.63       | 6.03  | <0.001|
| Digestibility, %            |                |              |     |       |
| DM                          | 56.58          | 62.23        | 1.379 | 0.670 |
| CP                          | 56.53          | 71.45        | 4.321 | 0.003 |
| N retention, g/day          | 2.34           | 4.45         | 0.898 | <0.001|
| Rumen parameters            |                |              |     |       |
| Ammonia, mM                 | 14.55          | 13.75        | 2.909 | 0.810 |
| Total bacteria, n/mL        | 1.72×10<sup>9</sup> | 6.90×10<sup>9</sup> | 1.113 | 0.051 |
| Total protozoa, n/mL        | 2.07×10<sup>9</sup> | 1.04×10<sup>9</sup> | 3.825 | 0.082 |

DM, dry matter; CP, crude protein.

Table 3. *In vitro* dry matter and crude protein digestibility of elephant grass and *Z. tetragona* at different inclusion levels.

| Item                        | Rumen liquor | Rumen liquor + pepsin | Rumen liquor | Rumen liquor + pepsin |
|-----------------------------|--------------|-----------------------|--------------|-----------------------|
| Elephant grass              |              |                       |              |                       |
| *Z. tetragona*              |              |                       |              |                       |
| 15%                         | 61.79<sup>a</sup> | 68.72<sup>a</sup> | 68.97<sup>c</sup> | 87.61<sup>b</sup>   |
| 30%                         | 72.08<sup>a</sup> | 73.84<sup>a</sup> | 82.92<sup>a</sup> | 92.38<sup>b</sup>   |
| 45%                         | 70.04<sup>c</sup> | 75.71<sup>b</sup> | 82.84<sup>a</sup> | 89.64<sup>a</sup>   |
| 100%                        | 71.55<sup>b</sup> | 73.21<sup>b</sup> | 77.86<sup>a</sup> | 88.16<sup>a</sup>   |
| SEM                         | 2.025        | 1.708                 | 2.841        | 1.952                 |
| P                           | 0.039        | 0.042                 | 0.027        | 0.031                 |

Means within a column with different superscript letters differ significantly (P<0.05).
bation that the gas produced by 30% *Z. tetragona* was higher than 100% of elephant grass, and after 48 h of incubation the total gas production was 83.35 mL for 30% *Z. tetragona* and 72.2 mL for elephant grass, respectively (Figure 1). The higher volume of gas produced when *Z. tetragona* was included reflects the higher ruminal degradability of this tropical leguminous shrub. Moreover, this increase suggests that more soluble carbohydrates and proteins were made available to microorganisms as supplementation increased (Arhab et al. 2009). This may also explain the increase in cumulative gas production of *Z. tetragona* at 48 h as previously reported by Chenost et al. (2001).

**Conclusions**

Forage from *Z. tetragona* was significantly high in CP and low in NDF compared with conventional tropical forages. Due to the high nutritional value, the use this tropical shrub improved intake, digestibility and N retention in sheep. These findings show that including *Z. tetragona* in ruminant diet can sustain animal production and minimise the production losses during the feed shortage periods in tropical areas.

**References**

AOAC, 2000. Official methods of analysis. 17th ed., Association of Official Analytical Chemists, Arlington, VA, USA.

Arhab, R., Macheboeuf, D., Aggoun, M., Bousseboua, H., Vialva, D., Besle, J.M., 2009. Effect of polyethylene glycol on in vitro gas production and digestibility of tannin containing feedstuffs from North African Arid Zone. Trop. Subtrop. Agroecosyst. 10:475-486.

Bamikole, M.A., 2003. Macro-minerals bioavailability study in goats fed forages of nitrogen fertilized Guinea grass and Guinea grass-Verano stylo mixture. Available from: http://www.lrrd.org/lrrd15/12/bami1512.htm

Barakat, N.A.M., Laudadio, V., Cazzato, E., Tufarelli, V., 2013. Potential contribution of retama raetam (Forssk.) Webb & Berthel as a forage shrub in Sinai, Egypt. Ard Land Res. Manag. 27:257-271.

Cazzato, E., Tufarelli, V., Laudadio, V., Stellacci, A.M., Selvaggi, M., Leoni, B., Troccoli, C., 2013. Forage yield and quality of emmer (Triticum dicoccum Schübler) and spelt (Triticum spelta L.) as affected by harvest period and nitrogen fertilization. Acta Agr. Scand. B-S. P. 63:571-578.

Chenost, M., Aufrere, J., Macheboeuf, D., 2001. The gas-test technique as a tool for predicting the energetic value of forage plants. Anim. Res. 50:349-364.

Conway, E.J., Byrne, A., 1933. An absorption apparatus for the micro-determination of certain volatile substances. The micro-determination of ammonia. Biochem. J. 27:419-429.

FAO-IAEA, 2000. Quantification of tannins in tree foliage. FAO-IAEA ed., Vienna, Austria.

Givens, D.I., Owen, E., Axford, R.E., Omed, H.M., 2000. Forage evaluation in ruminant nutrition. CABI Publishing, Wallingford, UK.

Khamseekhoew, B., Liang, J.B., Wong, C.C., Jalani, Z.A., 1991. Ruminal and intestinal digestibility of some tropical legume forages. Asian Austral. J. Anim. 14:321-325.

Khan, N.A., Habib, G., 2012. Assessment of Grewia oppositifolia leaves as crude protein supplement to low-quality forage diets of sheep. Trop. Anim. Health Pro. 44:1375-1381.

Laudadio, V., Tufarelli, V., Dario, M., Hammadi, M., S€eddk, M.M., Lacalandra, G.M., Dario, C., 2009. A survey of chemical and nutritional characteristics of halophytes plants used by camels in Southern Tunisia. Trop. Anim. Health Pro. 41:209-215.

Menke, K.H., Steingass, H., 1988. Estimation of the energetic feed value from chemical analysis and in vitro gas production using rumen fluid. Anim. Res. Dev. 28:47-55.

Mertens, D.R., 2002. Gravimetrical determination of anylase-treated neutral detergent fiber in feeds with reflushing in beakers or crucibles: collaborative study. J. AOAC Int. 85:1217-1240.

Ogimoto, K., Imai, S., 1981. Atlas of rumen microbiology. Japan Scientific Society Press, Tokyo, Japan.

Olafadehan, O.A., 2013. Feeding value of Pterocarpus erinaceus for growing goats. Anim. Feed Sci. Tech. 185:259-268.

Sajim, Y.C., Purwantari, N.D., 2006. Forage production of some legume tree. pp 952-957 in Proc. Nat. Conf. Anim. Vet. Technol., Puslitbang Peternakan, Bogor, Indonesia.

SAS, 2004. SAS/STAT 9.13 user’s guide. SAS Inst. Inc., Cary, NC, USA.

Steel, R.G.D., Torrie, J.H., Dicky, D.A., 1997. Principles and procedures of statistics: a biometrical approach. WCB/McGraw-Hill, New York, USA.

Tilley, J.M.A., Terry, R.A., 1963. A two-stage technique for the in vitro digestion of forage crops. J. Brit. Grassland Soc. 18:104-111.

Tufarelli, V., Khan, R.U., Laudadio, V., 2012. Evaluating the suitability of field beans as a substitute for soybean meal in early-lactating dairy cow: production and metabolic responses. Anim. Sci. J. 83:136-140.

Van Soest, P.J., Robertson, J.B., Lewis, B.A., 1991. Methods for dietary fiber, neutral detergent fiber, and non-starch polysaccharides in relation to animal nutrition. J. Dairy Sci. 74:3583-3592.

Waghorn, G., 2008. Beneficial and detrimental effects of dietary condensed tannins for sustainable sheep and goat production. Progress and challenges. Anim. Feed Sci. Tech. 147:116-139.