Self-Diffusion of Rodlike Viruses through Smectic Layers

M. Paul Lettinga and Eric Grelet
1IFF, Institut Weiche Materie, Forschungszentrum Jülich, D-52425 Jülich, Germany
2Centre de Recherche Paul Pascal, CNRS-Université Bordeaux 1, 115 Avenue Schweitzer, 33600 Pessac, France

We report the direct visualization at the scale of single particles of mass transport between smectic layers, also called permeation, in a suspension of rodlike viruses. Self-diffusion takes place preferentially in the direction normal to the smectic layers, and occurs by quasiquantized steps of one rod length. The diffusion rate corresponds with the rate calculated from the diffusion in the nematic state with a lamellar periodic ordering potential that is obtained experimentally.

DOI: 10.1103/PhysRevLett.99.197802
PACS numbers: 61.30.-v, 82.70.Dd, 87.15.Vv

Since the pioneering work of Onsager on the entropy driven phase transition to a liquid crystalline state [1], the structure and the phase behavior of complex fluids containing anisotropic particles with hard core interactions has been a subject of considerable interest, both theoretically [2] and experimentally [3]. Understanding the particle mobility in the different liquid crystalline phases is more recent [4]. In experiments various methods have been applied to obtain the ensemble averaged self-diffusion coefficients in thermotropic [5] and amphiphilic [6] liquid crystals, block copolymer [7] and colloidal systems [8]. Only a few studies have been done where dynamical phenomena are probed at the scale of a single anisotropic particle: the Brownian motion of an isolated colloidal ellipsoid in confined geometry [9] and the self-diffusion in a nematic phase formed by rodlike viruses [10] represent two recent examples. In the latter case, the diffusion parallel (\(D_\parallel\)) and perpendicular (\(D_\perp\)) to the average rod orientation (the director) has been measured, showing an increase of the ratio \(D_\parallel/D_\perp\) with particle concentration.

Knowledge of the dynamics at the single-particle level is fundamental for understanding the physics of mesophases with spatial order like the smectic (lamellar) phase of rodlike particles. In this mesophase the particle density is periodic in one dimension parallel to the long axis of the rods, while the interparticle correlations perpendicular to this axis are short-ranged (fluidlike order). For parallel diffusion to take place, the rods need to jump between adjacent smectic layers, overcoming an energy barrier related to the smectic order parameter [11]. This process of interlayer diffusion, or permeation, was first predicted by Helfrich [12]. In this Letter, we use video fluorescence microscopy to monitor the dynamics of individual labeled colloidal rods in the background of a smectic mesophase formed by identical but unlabeled rods. In this way we directly observe permeation of single rods in adjacent layers. As in the nematic phase, self-diffusion in a smectic phase is anisotropic: the diffusion through the smectic layers is shown here to be much faster than the diffusion within each liquidlike layer, i.e., \(D_\parallel/D_\perp \gg 1\), in contrast to thermotropic systems. Moreover, since the individual rod positions within the layer are monitored, the potential barrier for permeation is straightforwardly determined for the first time. The permeation can then be described in terms of Brownian particles diffusing in a one-dimensional periodic symmetric potential.

The system of rods used in this work consists of filamentous bacteriophages \(fd\), which are semirigid polyelectrolytes with a contour length of 0.88 \(\mu m\), a diameter of 6.6 \(nm\), and a persistence length of 2.2 \(\mu m\) [13]. Suspensions of \(fd\) rods in aqueous solution form several lyotropic liquid crystalline phases, in particular, the chiral nematic (cholesteric) phase and the smectic phase [14]. The existence of a smectic phase in suspensions of hard rods is an evidence of the high monodispersity and therefore of the model system character of such filamentous viruses [15, 16]. The colloidal scale of the \(fd\) bacteriophage facilitates the imaging of individual rods by fluorescence microscopy, as well as smectic layers by differential interference contrast (DIC) microscopy [14]. Figure 1(a) shows a sequence of images of a single region [17] where both techniques are combined. A comparison of the images shows that some rods jump between two layers while others remain within a given layer. The trajectory of one of the rods is plotted in Fig. 1(b) in the direction parallel (\(z\)) and perpendicular (\(\chi\)) to the director. This figure summarizes the key observation of this Letter: the diffusion throughout the smectic layers takes place in quasiquantized steps of one rod length; i.e., the mass transport between the layers is a discontinuous process. Moreover, it shows that the diffusion within the smectic layer is extremely slow [18].

The “hopping-type” diffusion is the consequence of the underlying ordering potential of the smectic phase and the vacancies available in adjacent layers. A phenomenological expression for permeation has been derived by coupling the displacement of a segment of a smectic layer \(u\) to the compressibility modulus \(\tilde{B}\) via the permeation parameter \(\lambda_b\) [11]:

\[
\frac{\partial u}{\partial t} = \lambda_b \tilde{B} \frac{\partial^2 u}{\partial z^2}.
\]

On a single-particle level, the fundamental solution of this
diffusion equation is the self–van Hove function [19], which is the probability for a displacement $z$ during a time $t$:

$$G(z, t) = \frac{1}{N} \sum_{i=1}^{N} \delta[z + z_i(0) - z_i(t)].$$  \hspace{1cm} (2)

Since single particles are experimentally identified, the self–van Hove function can be directly obtained from the histogram of particle positions after a time $t$, as plotted in Fig. 2 for low ($I = 20$ mM) and high ionic ($I = 110$ mM) strengths. For a fluid made of Brownian particles, a smooth Gaussian distribution that smears out over time is expected for the self–van Hove function. However at low ionic strength, $G(z, t)$ shows distinct peaks exactly at integer multiples of the particle length [and therefore of the layer thickness, see Fig. 2(a)] as expected from visual observation (Fig. 1). At high ionic strength the curves are smoother [Fig. 2(b)], but in all cases the experimental self–van Hove function is not Gaussian at any time. This implies that the permeation parameter, can be directly obtained.

The energy landscape can be determined experimentally from the distribution of particle positions with respect to the middle of a layer parallel to the director. To this end, time windows are selected where the particle remains for ten frames or more within the same layer. The distribution of particles within a single layer is then obtained by addition of all particle positions relative to the average position of particles for all selected time windows. The resulting distributions are plotted in Fig. 3(a) for the two ionic strengths. To obtain the total particle distribution for the full smectic phase, the distributions of particles in a single layer [Fig. 3(a)] is added periodically to itself at all integer numbers of layer spacing $L$ [Fig. 1(a)]. The smectic ordering potential is then deduced from the Boltzmann factor $P(z) \sim e^{-U_{layer}(z)/k_B T}$ for the probability of finding a particle at position $z$, as shown in Fig. 3(b). Both potentials can be best fitted with a sinusoidal $U_{layer}(z) = U_0 \sin(2\pi z/L)$, giving an amplitude of $U_0 = 1.36k_B T$ at low ionic strength and $U_0 = 1.66k_B T$ at high ionic strength. The difference between the two amplitudes explains the fact that for $I = 20$ mM the self–van Hove function exhibits discrete peaks, while for $I = 110$ mM the potential barrier is small enough to exhibit a monotonic behavior of the probability density function. The reason for the more pronounced potential at low ionic strength might be that electrostatic interactions between rods are more long ranged; i.e., particles are more strongly correlated so that it is more difficult to create a void between them. The fact that the potential can be fitted by a sinusoidal is remarkable by itself. Indeed, the use of such a potential is very common due to its simplicity [20], but this ordering potential has never been directly observed until now. Moreover the height of the potential, i.e., the smectic order parameter, can be directly obtained.
The overall mean square displacement (MSD) of rods parallel and perpendicular to the director of the smectic and nematic phase is plotted in Fig. 4 for both ionic strengths. The time evolution of the MSD given by \( \langle \Delta r^2(t) \rangle \sim t^\gamma \) provides the diffusion exponent \( \gamma \); \( \gamma < 1 \) is characteristic of a subdiffusive behavior, while \( \gamma > 1 \) is referred to as superdiffusion. The parallel motion is close to be diffusive \( (\gamma \approx 1) \) in both the nematic \( (\gamma = 0.97) \) and smectic \( (\gamma = 0.94) \) phases for \( I = 110 \) mM and in the nematic phase for \( I = 20 \) mM \( (\gamma = 0.95) \). Only the parallel motion in the smectic phase for low ionic strength, i.e., where the discrete peaks in the self–van Hove function are observed, is significantly subdiffusive: \( \gamma = 0.81 \). The perpendicular motion is in all cases strongly subdiffusive: for \( I = 110 \) mM, \( \gamma \) reduces from 0.63 before to 0.56 after the nematic-smectic \( (N\text{-Sm}) \) transition and for \( I = 20 \) mM it reduces from 0.68 to 0.46. Anomalous subdiffusive behavior has often been observed in systems where diffusion takes place by steps, e.g., in the case of release from a surrounding cage [21]. This “cage escape” might be at the origin of the observed subdiffusive behavior for both parallel and perpendicular diffusion. For parallel diffusion the cage is formed by the energy barrier imposed by the smectic layers, as shown by smaller \( \gamma \) for higher ordering potential. Perpendicular diffusion at high volume fractions is only possible through a reptation-like motion along the long axis to escape the local excluded volume, as observed for polymers for which typically \( \gamma = 0.5 \) [22]. This excluded volume is huge, even for rods at high orientational order, due to the large rod aspect ratio of \( \approx 130 \). In addition, perpendicular diffusion in the smectic phase is hindered due to the ordering potential, which couples this diffusion to the permeation and which thus explains the decrease of \( \gamma \) from the nematic to the smectic phases. For subdiffusive systems, a non-Gaussian distribution of the probability density functions has been observed as in Fig. 2 [21], even though these two features are not a priori correlated. Note also that boundary effects might influence the probability density [23].

The anisotropy in the total diffusion, \( D_\parallel/D_\perp \), which is about 20 in the nematic phase [10], increases in the smectic phase as a result of the pronounced subdiffusivity of the perpendicular motion (decrease of \( \gamma \)). These observations show an opposite trend as compared to thermotropic liquid crystals [4,5], where usually \( D_\parallel/D_\perp \) evolves from being larger than 1 at temperatures close to the \( N\text{-Sm} \) transition.
temperature to being smaller than 1 at lower temperatures [24]. Therefore the diffusion in the smectic phase can be effectively considered as a one-dimensional diffusion of a Brownian particle in a periodic potential in the high friction limit. A general expression for such a diffusion process is given by [25]

\[ D_\parallel = \frac{D_0}{\langle e^{-U_{\text{layer}}(z)/k_B T}\rangle} \]

(3)

The brackets indicate averaging over one period of the ordering potential. The diffusion coefficient in the smectic phase can then be calculated taking \( D_0 \) as the diffusion coefficient in the nematic phase close to the N-Sm transition, and using \( U_{\text{layer}} \) as obtained from the fit of the potentials plotted in Fig. 3: the diffusion coefficient decreases by a factor 0.84 at \( I = 20 \) mM. Indeed the MSD in the smectic phase is obtained from the MSD in the nematic phase, using these factors for both ionic strengths (see Fig. 4), although at \( I = 20 \) mM some deviation appears due to the subdiffusivity in the MSD. Thus, we have shown how the mobility of rods decreases after the N-Sm transition, contrary to the isotropic-nematic transition where the global mobility increases due to entropic gain [1,10]. It seems therefore to indicate that fd virus suspensions do not behave as a system of rigid hard rods for high concentration in agreement with a recent work [16]. Moreover, the very slow diffusion within the layers suggests that the smectic phase of semiflexible colloidal rods consists of layers of glasslike, rather than fluidlike, particles.

In conclusion, we have for the first time visualized the process of permeation in the smectic phase at the scale of single particles for a system of charged rods. This allowed us to give a full and coherent description of the diffusion process without any assumptions on the system. The diffusion is strongly anisotropic in the direction normal to the smectic layers and quasidiscontinuous due to the presence of the layers. The parallel diffusion rate complies with the rate in the nematic phase, taking into account the ordering potential, which is obtained directly from our measurements.

We thank Jan Dhont for fruitful discussions. This project was supported by the European network of excellence SoftComp. MPL acknowledges also the SFB TR6 for financial support.

[1] L. Onsager, Ann. N.Y. Acad. Sci. 51, 627 (1949).
[2] D. Frenkel et al., Nature (London) 332, 822 (1988); G. J. Vroege and H. N. W. Lekkerkerker, Rep. Prog. Phys. 55, 1241 (1992).
[3] X. Wen et al., Phys. Rev. Lett. 63, 2760 (1989); H. Maeda and Y. Maeda, Phys. Rev. Lett. 90, 018303 (2003); Z. Dogic and S. Fraden, Curr. Opin. Colloid Interface Sci. 11, 47 (2006).
[4] S. Hess et al., Mol. Phys. 74, 765 (1991); H. Löwen, Phys. Rev. E 59, 1989 (1999); R. L. B. Selinger, Phys. Rev. E 65, 051702 (2002); M. A. Bates and G. R. Luckhurst, J. Chem. Phys. 120, 394 (2004); M. Cifelli et al., J. Chem. Phys. 125, 164912 (2006).
[5] G. J. Krüger, Phys. Rep. 82, 229 (1982); S. V. Dvinskikh et al., Phys. Rev. E 65, 061701 (2002); J. Xu et al., Phys. Rev. E 72, 051703 (2005).
[6] R. Blinc et al., Phys. Rev. Lett. 25, 1327 (1970); S. Gaemers and A. Bax, J. Am. Chem. Soc. 123, 12343 (2001); P. L. Hubbard et al., Langmuir 21, 4340 (2005); Y. Gambin et al., Phys. Rev. Lett. 94, 110602 (2005).
[7] G. H. Fredrickson and F. S. Bates, Annu. Rev. Mater. Sci. 26, 501 (1996).
[8] Z. Bu et al., Macromolecules 27, 6871 (1994); M. P. B. van Bruggen et al., Phys. Rev. E 58, 7668 (1998); R. Kush and P. S. Russo, Macromolecules 35, 8659 (2002).
[9] Y. Han et al., Science 314, 626 (2006).
[10] M. P. Lettinga et al., Europhys. Lett. 71, 692 (2005).
[11] P. G. de Gennes and J. Prost, The Physics of Liquid Crystals (Clarendon, Oxford, 1993).
[12] W. Helfrich, Phys. Rev. Lett. 23, 372 (1969).
[13] fd was grown using the XLI-Blue strain of E. coli as the host bacteria and purified following standard biological protocols [14]. To vary the ionic strength, viruses have been dialyzed against a 20 mM TRIS-HCl buffer at pH = 8.2 with an adjusted amount of NaCl. About one fd out of 10^3 has been labeled with the dye Alexa-488 (Invitrogen).
[14] Z. Dogic and S. Fraden, in Soft Matter, edited by G. Gompper and M. Schick (Wiley-VCH, Weinheim, 2006), Vol. 2.
[15] M. A. Bates and D. Frenkel, J. Chem. Phys. 109, 6193 (1998).
[16] K. R. Purdy and S. Fraden, Phys. Rev. E 76, 011705 (2007).
[17] The time resolution of the used CCD camera (Hamamatsu C9100) was between 10 and 120 ms, depending on the region of interest.
[18] To quantify this hopping-type diffusion, the jumps have been identified with a home-written algorithm based on the criterion that the first \( N \) positions in time should deviate in one direction from the previous average, and that the distance between two jumps should be close to the particle length.
[19] J. P. Hansen and I. R. McDonald, Theory of Simple Liquids (Academic, London, 1986).
[20] B. Mulder, Phys. Rev. A 35, 3095 (1987); A. Stroobants et al., Phys. Rev. Lett. 57, 1452 (1986).
[21] E. R. Weeks and D. A Weitz, Phys. Rev. Lett. 89, 095704 (2002); Chem. Phys. 284, 361 (2002).
[22] T. C. B. McLeish, Adv. Phys. 51, 1379 (2002).
[23] Although data acquisitions were always taken in the middle of the sample which is rather thin (about 10 \( \mu m \)), the diffusion has been observed to slow down close to the cell walls.
[24] The interpretation of such behaviors is ambiguous due to the different temperature dependence of the viscosity parallel and perpendicular to the director.
[25] R. Festa and E. Galleani d’Aglionio, Physica (Amsterdam) 90A, 229 (1978).